



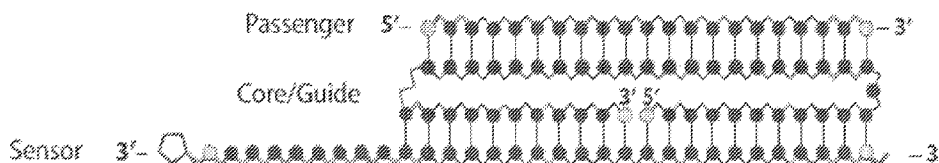
(12) **DEMANDE DE BREVET CANADIEN
CANADIAN PATENT APPLICATION**

(13) **A1**

(86) Date de dépôt PCT/PCT Filing Date: 2022/07/05
 (87) Date publication PCT/PCT Publication Date: 2023/01/12
 (85) Entrée phase nationale/National Entry: 2024/01/04
 (86) N° demande PCT/PCT Application No.: US 2022/073426
 (87) N° publication PCT/PCT Publication No.: 2023/283546
 (30) Priorité/Priority: 2021/07/06 (US63/218,833)

(51) Cl.Int./Int.Cl. *C12N 15/113* (2010.01),
C07H 21/00 (2006.01)
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(54) Titre : COMPLEXES D'ACIDES NUCLEIQUES A ACTIVATION CONDITIONNELLE
 (54) Title: CONDITIONALLY ACTIVATABLE NUCLEIC ACID COMPLEXES



- | | | | | | |
|--|---|--|------------------------------|--|---------------------|
| | Delivery ligand, fluorophore, or exonuclease | | Phosphorothioate | | Locked nucleic acid |
| | blocker attached to suitable connector | | C ₃ connector | | 2'-OMe |
| | Eg: fatty acid, Cy3, or inverted dT attached to a tri-ethylene glycol | | Backbone sites for screening | | 2'-F |
| | | | | | Bases for screening |

FIG. 3

(57) **Abrégé/Abstract:**

Provided herein include conditionally activatable small interfering RNA (siRNA) complexes, components, compositions, and related methods and systems. The siRNA complex can be conditionally activated upon a complementary binding to an input nucleic acid strand (e.g. a mRNA of a biomarker gene specific to a target cell) through a sequence in a sensor nucleic acid strand of the nucleic acid complex. The activated nucleic acid complex can release a potent RNAi duplex formed by a core nucleic acid strand and a passenger nucleic acid strand, which can specifically inhibit a target RNA.

Date Submitted: 2024/01/04

CA App. No.: 3224942

Abstract:

Provided herein include conditionally activatable small interfering RNA (siRNA) complexes, components, compositions, and related methods and systems. The siRNA complex can be conditionally activated upon a complementary binding to an input nucleic acid strand (e.g. a mRNA of a biomarker gene specific to a target cell) through a sequence in a sensor nucleic acid strand of the nucleic acid complex. The activated nucleic acid complex can release a potent RNAi duplex formed by a core nucleic acid strand and a passenger nucleic acid strand, which can specifically inhibit a target RNA.

CONDITIONALLY ACTIVATABLE NUCLEIC ACID COMPLEXES

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application claims the benefit under 35 U.S.C. §119(e) to U.S. Provisional Patent Application No. 63/218,833 filed on July 6, 2021, the content of which is incorporated herein by reference in its entirety for all purposes.

REFERENCE TO SEQUENCE LISTING

[0002] The present application is being filed along with a Sequence Listing in electronic format. The Sequence Listing is provided as a file entitled 75EN-329791-WO, created July 3, 2022, which is 200 kilobytes in size. The information in the electronic format of the Sequence Listing is incorporated herein by reference in its entirety.

BACKGROUND

Field

[0003] The present disclosure relates generally to the field of nucleic acids, for example, conditionally activatable small interfering RNA complexes.

Description of the Related Art

[0004] Despite emerging developments in the field of dynamic nucleic acid nanotechnology and biomolecular computing, there is still a challenge to develop targeted RNAi therapy that can use nucleic acid logic switches to sense RNA transcripts (such as mRNAs and miRNAs), thereby restricting RNA interfering (RNAi) therapy to specific populations of disease-related cells. In particular, there is a need to develop targeted and conditionally activated RNAi therapy with improved drug potency, sensitivity, and stability, low design complexity, and low dosage requirement.

SUMMARY

[0005] Disclosed herein include a nucleic acid complexes, comprising: a first nucleic acid strand comprising 20-70 linked nucleosides; a second nucleic acid strand binding to a central region of the first nucleic acid strand to form a first nucleic acid duplex; and a third nucleic acid strand binding to a 5' region and a 3' region of the first nucleic acid strand to form a second nucleic acid duplex, wherein the third nucleic acid strand comprises a overhang, wherein the overhang is not complementary to the first nucleic acid strand and is capable of binding to an input nucleic acid strand to cause the displacement of the third nucleic acid strand from the first nucleic acid strand. In some embodiments, the central region of the first nucleic acid strand

comprises a sequence complementary to a target RNA, wherein the sequence is 10-35 nucleosides in length.

[0006] The sequence complementary to the target RNA can be, for example, 10-21 nucleotides in length. In some embodiments, the second nucleic acid strand binds to 19-25 linked nucleotides in the central region of the first nucleic acid strand to form the first nucleic acid duplex. In some embodiments, the first nucleic acid duplex, the nucleic acid complex, or both, does not comprise a Dicer cleavage site.

[0007] The central region of the first nucleic acid strand can be, for example, linked to the 5' region of the first nucleic acid strand via a 5' connector. The central region of the first nucleic acid strand can be, for example, linked to the 3' region of the first nucleic acid strand via a 3' connector. In some embodiments, the 5' connector, the 3' connector, or both comprise a C3 3-carbon linker, a nucleotide, a modified nucleotide, or an exonuclease cleavage-resistant moiety, or a combination thereof. In some embodiments, the modified nucleotide is a 2'-O-methyl nucleotide or a 2'-F nucleotide. In some embodiments, the 5' connector comprises, or is, a C3 3-carbon linker, 2'-O-methyl nucleotide, 2'-F nucleotide, a nucleotide with a phosphodiester 5' and 3' connection cleavable by an exonuclease when in a single stranded form, or a combination thereof. The 3' connector can be, for example, a C3 3-carbon linker. In some embodiments, the 3' connector comprises a C3 3-carbon linker, a nucleotide, a modified nucleotide, an exonuclease cleavage-resistant moiety when in a single stranded form, or a combination thereof. In some embodiments, the 3' connector comprises, or is, a 2'-O-methyl nucleotide, and wherein the 2'-O-methyl nucleotide is optionally 2'-O-methyladenosine, 2'-O-methylguanosine, 2'-O-methyluridine, or 2'-O-methylcytidine.

[0008] In some embodiments, the second nucleic strand is fully complementary to the central region of the first nucleic acid strand, thereby forming blunt ends at the 5' and 3' termini of the second nucleic acid strand in the first nucleic acid duplex. In some embodiments, the second nucleic acid strand does not have an overhang at 3' terminus, or 5' terminus, or both in the first nucleic acid duplex. In some embodiments, the second nucleic acid strand has a 3' overhang, a 5' overhang, or both in the first nucleic acid duplex. In some embodiments, the second nucleic acid strand has a 3' overhang and the 3' overhang is one to five nucleosides in length. In some embodiments, the 5' terminus of the central region of the first nucleic acid strand, the 3' terminus of the central region of the first nucleic acid strand, or both, comprises at least one phosphorothioate internucleoside linkage. In some embodiments, each of the 5' terminus of the central region of the first nucleic acid strand and the 3' terminus of the central region of the first nucleic acid strand independently comprises one or more phosphorothioate internucleoside linkages. In some embodiments, the central region of the first nucleic acid strand

does not comprise phosphorothioate internucleoside linkages except for the internucleoside linkage(s) between two or three nucleosides at the 5' terminus, 3' terminus, or both, of the central region. In some embodiments, at least 80%, at least 85%, at least 90%, or at least 95% of the nucleosides of one or more of (1) the central region of the first nucleic acid strand, (2) the 5' region of the first nucleic strand, and (3) the 3' region of the first nucleic strand are chemically modified. In some embodiments, at least 80%, at least 85%, at least 90%, or at least 95% of the nucleosides of one or more of the first nucleic acid strand, the second nucleic strand and the third nucleic strand are chemically modified. In some embodiments, at least 80%, at least 85%, at least 90%, at least 95%, or all of the nucleosides of the nucleic acid complex are chemically modified. The chemical modifications can be, for example, to resist nuclease degradation, to increase melting temperature (T_m), or both, of the nucleic acid complex.

[0009] In some embodiments, at least 90%, at least 95%, or all of the nucleotides of the nucleic acid complex are non-DNA and non-RNA nucleotides. In some embodiments, at most 5%, at most 10%, or at most 15% of the nucleosides of the second nucleic strand are LNA. In some embodiments, about 10%-50% of the bases have a 2'-4' bridging modifications. In some embodiments, about 10%-50% of the bases are locked locked nucleic acid (LNA) or analogues thereof. In some embodiments, about 10%-50% of the bases comprises 2'-O-methyl modification, 2'-F modification, or both. In some embodiments, less than 5%, less than 10%, less than 25%, less than 50% of the internucleoside linkages in the first nucleic acid strand are phosphorothioate internucleoside linkages. In some embodiments, the first nucleic acid strand does not comprise phosphorothioate internucleoside linkages. In some embodiments, the internucleoside linkages between (1) the one to three nucleotides adjacent to the 3' of the 5' connector, and/or (2) the one or two nucleotides adjacent to the 5' of the 3' connector, and/or (3) the one to three nucleotides adjacent to the 3' of the 3' connector, are phosphorothioate internucleoside linkages

[0010] The input nucleic acid strand can be a RNA. In some embodiments, the target RNA is a cellular RNA transcript. In some embodiments, the target RNA is an mRNA, an miRNA, a non-coding RNA, a viral RNA transcript, or a combination thereof.

[0011] In some embodiments, the overhang of the second nucleic acid strand is capable of binding to the input nucleic acid strand to form a toehold, thereby causing the displacement of the second nucleic acid strand from the first nucleic acid strand. In some embodiments, the overhang of the second nucleic acid strand is 5 to 20 nucleosides in length, and optionally 9 nucleotides in length. In some embodiments, all internucleoside linkages of the overhang of the third nucleic acid strand are phosphorothioate internucleoside linkages. In some embodiments, the 5' terminus, the 3' terminus, or both of the third nucleic acid strand comprises

a terminal moiety. In some embodiments, the terminal moiety comprises a ligand, a fluorophore, a exonuclease, a fatty acid, a Cy3, an inverted dT attached to a tri-ethylene glycol, or a combination thereof.

[0012] Provided herein include a method of modulating a target RNA, wherein the method includes: contacting a cell comprising a target RNA with any one or more of the nucleic acid complex disclosed herein, wherein an input strand binds to the overhang of the third nucleic acid strand to cause displacement of the third nucleic acid strand from the first nucleic acid strand to release the sequence complementary to the target RNA into the cell, thereby modulating the target RNA.

[0013] Contacting the cell with the nucleic acid complex can be performed *in vitro*, *in vivo*, *ex vivo*, or a combination thereof. In some embodiments, contacting the cell with the nucleic acid complex occurs in the body of a subject. In some embodiments, the cell is a disease cell, and optionally the cell is a cancer cell. In some embodiments, the cell is a neuron.

[0014] Also provided herein includes a method of treating a disease or a condition, wherein the method includes administering any one or more of the nucleic acid complex disclosed herein to a subject in need thereof, wherein the input strand binds to the overhang of the third nucleic acid strand to cause displacement of the third nucleic acid strand from the first nucleic acid strand to release the sequence complementary to a target RNA, thereby reducing the activity of the target RNA or protein expression from the target RNA in the subject to treat the disease or condition. In some embodiments, the disease or condition is a central nervous system (CNS) disease or disorder or cancer. In some embodiments, the target RNA is a mRNA or a miRNA. In some embodiments, the nucleic acid complex is administered to a subject via a lipid-mediated delivery system, optionally via liposomes, nanoparticles, or micelles. In some embodiments, the nucleic acid complex is administered to a subject via nanoparticles, inorganic nanoparticles, nucleic acid lipid particles, polymeric nanoparticles, lipidoid nanoparticles (LNPs), chitosan and inulin nanoparticles, cyclodextrins nanoparticles, carbon nanotubes, liposomes, micellar structures, capsids, polymers, polymer matrices, hydrogels, dendrimers, nucleic acid nanostructure, exosomes, GalNAc-conjugated melittin-like peptides, or combinations thereof. In some embodiments, the nucleic acid complex is administered to a subject in need thereof via a subcutaneous injection. In some embodiments, the nucleic acid complex of any one of claims 1 to 40 is administered to a subject in need thereof via an intravenous injection. In some embodiments, the nucleic acid complex is administered to a subject in need thereof at a concentration about 0.1-10 nM, optionally about 0.1-1.0 nM.

[0015] Details of one or more implementations of the subject matter described in this specification are set forth in the accompanying drawings and the description below. Other

features, aspects, and advantages will become apparent from the description, the drawings, and the claims. Neither this summary nor the following detailed description purports to define or limit the scope of the inventive subject matter.

BRIEF DESCRIPTION OF THE DRAWINGS

[0016] **FIG. 1** illustrates a schematic representation of two non-limiting exemplary nucleic acid complex constructs from Design 1 and Design 2.

[0017] **FIG. 2** illustrates a schematic representation of a non-limiting exemplary nucleic acid complex with component strands (a sensor nucleic acid strand, a core nucleic acid strand and a passenger nucleic acid strand) and chemical modification patterns.

[0018] **FIG. 3** illustrates a schematic representation of a non-limiting exemplary nucleic acid complex construct with regions for screening highlighted in yellow.

[0019] **FIG. 4A** is a schematic diagram showing the activation of a nucleic acid complex in targeted cells following the base-pairing of the sensor strand to a RNA marker. **FIG. 4B** is a schematic diagram showing the formation of an active RNAi duplex following the displacement of a sensor nucleic acid strand from a core nucleic acid strand and the degradation of the core nucleic acid strand overhangs.

[0020] **FIG. 5A** and **FIG. 5B** show sequence diagrams of two non-limiting exemplary nucleic acid complex constructs having the same passenger strand but different core strand. Passenger strand v3p1 and passenger strand 1: SEQ ID NO: 2; Core strand v3c1: SEQ ID NOs: 3-5 connected by a C3 spacer. Core strand v3c5: SEQ ID NO: 11.

[0021] **FIG. 6** show sequence diagrams of two positive control constructs. HTT Guide 1: SEQ ID NO: 21; HTT Pass 1: SEQ ID NO: 22; HTT Guide 2: SEQ ID NO: 23; HTT Pass 2: SEQ ID NO: 24.

[0022] **FIG. 7** shows various siRNA complex variants with different passenger strand (V3P1, V3P2, V3P3, V3P5, V3P5, V3P6, V3P7, V3P8, and V3P9) assembled with an exemplary core strand (v3c1 which include two C3 linkers) shown in **FIG. 5** and used in target protein expression shown in **FIG. 8**.

[0023] **FIG. 8** shows a graphic representation of the target protein expression data generated using the siRNA complex design variants shown in **FIG. 7**.

[0024] **FIG. 9** shows various siRNA complex variants with different passenger strand (V3P1, V3P2, V3P3, V3P5, V3P5, V3P6, V3P7, V3P8, and V3P9) assembled with an exemplary core strand (v3c5 which does not include a C3 linker) shown in **FIG. 5**.

[0025] **FIG. 10** shows a graphic representation of the target protein expression data generated using the siRNA complex variants shown in **FIG. 9**.

[0026] FIG. 11A and FIG. 11B show sequence diagrams of various exemplary nucleic acid complex constructs each having the same passenger strand (Passenger strand 1) and the same sensor strand (Mir23 Sensor 1) but a different core strand (Core strand v3c1, Core strand v3c2, Core strand v3c3, Core strand v3c4, Core strand v3c5, and Core strand v3c6, which are referred to as C1, C2, C3, C4, C5, C6, respectively, in FIGS. 13-14 and description thereof). The sequences shown in FIGS. 11A and 11B are listed in Table 1.

[0027] FIG. 12 shows non-denaturing polyacrylamide gel (PAGE) of various nucleic acid complex constructs.

[0028] FIG. 13 shows the RNAi activity of two-stranded assemblies each having the same passenger strand v3p1 and a different core strand (C1, C2, C3, C4, C5, and C6) at different concentrations.

[0029] FIG. 14 shows the RNAi activity of three-stranded assemblies each having the same passenger strand v3p1, the same sensor strand (Mir23 sensor 1), and a different core strand (C1, C2, C3, C4, C5, and C6) at three different concentrations.

[0030] FIG. 15 shows sequence diagrams of a non-limiting exemplary nucleic acid complex construct disclosed herein (top: V3C3a) and a partially modified nucleic acid complex (bottom: G1C1S1). The sequences shown in FIG. 15 are listed in Table 2.

[0031] FIG. 16 shows the RNAi activity of the exemplary two-stranded nucleic acid complex constructs (V3C3a siRNA) and three-stranded nucleic acid complex constructs (V3C3a and V3C3b) in comparison with the partially modified two-stranded construct (G1C1 siRNA) and the partially modified three-stranded constructs (G1C1S1) shown in FIG. 15 at three different concentrations.

[0032] Throughout the drawings, reference numbers may be re-used to indicate correspondence between referenced elements. The drawings are provided to illustrate example embodiments described herein and are not intended to limit the scope of the disclosure.

DETAILED DESCRIPTION

[0033] In the following detailed description, reference is made to the accompanying drawings, which form a part hereof. In the drawings, similar symbols typically identify similar components, unless context dictates otherwise. The illustrative embodiments described in the detailed description, drawings, and claims are not meant to be limiting. Other embodiments may be utilized, and other changes may be made, without departing from the spirit or scope of the subject matter presented herein. It will be readily understood that the aspects of the present disclosure, as generally described herein, and illustrated in the Figures, can be arranged, substituted, combined, separated, and designed in a wide variety of different configurations, all

of which are explicitly contemplated herein and made part of the disclosure herein.

[0034] All patents, published patent applications, other publications, and sequences from GenBank, and other databases referred to herein are incorporated by reference in their entirety with respect to the related technology.

[0035] RNA interference (RNAi) is an intrinsic cellular mechanism conserved in most eukaryotes, that helps to regulate the expression of genes critical to cell fate determination, differentiation, survival and defense from viral infection. Researchers have exploited this natural mechanism by designing synthetic double-stranded RNA for sequence-specific gene silencing. Emerging developments in the field of dynamic nucleic acid nanotechnology and biomolecular computing also offer a conceptual approach to design programmable RNAi agents. However, challenges still remain in developing targeted RNAi therapy that can use nucleic acid logic switches to sense RNA transcripts (such as mRNAs and miRNAs) in order to restrict RNA silencing to specific populations of disease-related cells and spare normal tissues from toxic side effects. Significant challenges include poorly suppressed background drug activity, weak activated state drug potency, input and output sequence overlap, high design complexity, short lifetimes (< 24 hours) and high required device concentrations (> 10 nM).

[0036] Provided herein include conditionally activatable small interfering RNA (siRNA) complexes, components, compositions, and related methods and systems. The conditionally activatable siRNA complex can switch from an inactivated state to an activated state when triggered by a complementary binding of an input nucleic acid strand (e.g. a disease biomarker gene specific to disease-related cells) to the siRNA complex, thereby activating the RNA interference activity of the siRNA complex to target a specific target RNA (e.g. a RNA to be silenced). The nucleic acid complexes herein described can mediate conditionally activated RNA interference activity to silence target RNA in specific populations of disease-related cells with improved potency at a low concentration as well as improved specificity that can reduce off-target effects.

[0037] Disclosed herein includes a nucleic acid complex. The nucleic acid complex comprises a first nucleic acid strand (e.g. core nucleic acid strand), a second nucleic acid strand (e.g. passenger nucleic acid strand) binding to a central region of the first nucleic acid strand to form a first nucleic acid duplex (e.g. RNAi duplex), and a third nucleic acid strand (e.g. sensor nucleic acid strand) binding to a 5' region and a 3' region of the core nucleic acid strand to form a second nucleic acid duplex (e.g. sensor duplex). The sensor nucleic acid strand comprises an overhang, wherein the overhang is not complementary to the first nucleic acid strand and is capable of binding to an input nucleic acid strand to cause the displacement of the sensor nucleic acid strand from the core nucleic acid strand. The central region of the core nucleic acid strand

comprises a sequence complementary to a target RNA. The sequence can be 10-35 nucleosides in length. The first nucleic acid strand (e.g., core nucleic acid strand) can comprise 20-70 linked nucleosides

[0038] Disclosed herein also includes a method of modulating a target RNA. The method comprises contacting a cell comprising a target RNA with the nucleic acid complex herein described. Upon detection of an input nucleic acid strand, the input nucleic acid strand can bind to the overhang of the sensor nucleic acid strand to cause displacement of the sensor nucleic acid strand from the core nucleic acid strand to release the sequence complementary to the target RNA into the cell, thereby modulating the target RNA.

[0039] Disclosed herein also includes a method of treating a disease or a condition. The method comprises administering the nucleic acid complex herein described to a subject in need thereof. Upon detection of an input nucleic acid strand, the input strand can bind to the overhang of the sensor nucleic acid strand to cause displacement of the sensor nucleic acid strand from the core nucleic acid strand to release the sequence complementary to a target RNA, thereby reducing the activity of the target RNA or protein expression from the target RNA in the subject to treat the disease or condition.

Definitions

[0040] Unless defined otherwise, technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which the present disclosure belongs. *See, e.g.*, Singleton et al., Dictionary of Microbiology and Molecular Biology 2nd ed., J. Wiley & Sons (New York, NY 1994); Sambrook et al., Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press (Cold Spring Harbor, NY 1989). For purposes of the present disclosure, the following terms are defined below.

[0041] As used herein, the term “nucleoside” refers to a molecule having a purine or pyrimidine base covalently linked to a ribose or deoxyribose sugar. Exemplary nucleosides include adenosine, guanosine, cytidine, uridine and thymidine.

[0042] The term “nucleotide” refers to a nucleoside having one or more phosphate groups joined in ester linkages to the sugar moiety. Exemplary nucleotides include nucleoside monophosphates, diphosphates and triphosphates.

[0043] The terms “polynucleotide” and “nucleic acid molecule” are used interchangeably herein and refer to a polymer of nucleotides joined together by a phosphodiester linkage between 5' and 3' carbon atoms.

[0044] The term “RNA” or “RNA molecule” or “ribonucleic acid molecule” refers to a polymer of ribonucleotides. The term “DNA” or “DNA molecule” or “deoxyribonucleic acid molecule” refers to a polymer of deoxyribonucleotides. DNA and RNA can be synthesized

naturally (e.g., by DNA replication or transcription of DNA, respectively). RNA can be post-transcriptionally modified. DNA and RNA can also be chemically synthesized. DNA and RNA can be single-stranded or multi-stranded (e.g., double-stranded or triple-stranded). “mRNA” or “messenger RNA” is single-stranded RNA molecule that is complementary to one of the DNA strands of a gene. “miRNA” or “microRNA” is a small single-stranded non-coding RNA molecule that functions in RNA silencing and post-transcriptional regulation of gene expression.

[0045] The term “RNA analog” refers to an polynucleotide having at least one altered or modified nucleotide as compared to a corresponding unaltered or unmodified RNA. The nucleotide can retain the same or similar nature or function as the corresponding unaltered or unmodified RNA such as forming base pairs.

[0046] A single-stranded polynucleotide has a 5' terminus or 5' end and a 3' terminus or 3' end. The terms “5' end” “5' terminus” and “3' end” “3' terminus” of a single-stranded polynucleotide indicate the terminal residues of the single-stranded polynucleotide and are distinguished based on the nature of the free group on each extremity. The 5'-terminus of a single-stranded polynucleotide designates the terminal residue of the single-stranded polynucleotide that has the fifth carbon in the sugar-ring of the deoxyribose or ribose at its terminus (5' terminus). The 3'-terminus of a single-stranded polynucleotide designates the residue terminating at the hydroxyl group of the third carbon in the sugar-ring of the nucleotide or nucleoside at its terminus (3' terminus). The 5' terminus and 3' terminus in various cases can be modified chemically or biologically e.g. by the addition of functional groups or other compounds as will be understood by the skilled person.

[0047] As used herein, the terms “complementary binding” and “bind complementarily” mean that two single strands are base paired to each other to form nucleic acid duplex or double-stranded nucleic acid. The term “base pair” as used herein indicates formation of hydrogen bonds between base pairs on opposite complementary polynucleotide strands or sequences following the Watson-Crick base pairing rule. For example, in the canonical Watson-Crick DNA base pairing, adenine (A) forms a base pair with thymine (T) and guanine (G) forms a base pair with cytosine (C). In RNA base pairing, adenine (A) forms a base pair with uracil (U) and guanine (G) forms a base pair with cytosine (C). A certain percentage of mismatches between the two single strands are allowed as long as a stable double-stranded duplex can be formed. In some embodiments, the two strands that bind complementarily can have a mismatches can be, about, be at most, or be at most about 1%, 2%, 3%, 4%, 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, or 50%.

[0048] As used herein, the terms “RNA interference” “RNA interfering” or “RNAi” refer to a selective intracellular degradation of RNA. RNAi can occur in cells naturally to remove foreign RNAs (e.g., viral RNAs). Natural RNAi proceeds via fragments cleaved from free dsRNA which direct the degradative mechanism to other similar RNA sequences. Alternatively, RNAi can be initiated by the hand of man, for example, to silence the expression of target genes.

[0049] As used herein, the terms “small interfering RNA” and “siRNA” refers to an RNA or RNA analog capable of reducing or inhibiting expression of a gene or a target gene when the siRNA is activated in the same cell as the target gene. The siRNA used herein can comprise naturally occurring nucleic acid bases and/or chemically modified nucleic acid bases (RNA analogs).

Nucleic acid complexes

[0050] Provided herein include a nucleic acid complex that can be conditionally activated upon a complementary binding to an input nucleic acid strand (e.g. a mRNA of a disease biomarker gene specific to a target cell (e.g., disease-related cells)) through a sequence in a sensor nucleic acid strand of the nucleic acid complex. The activated nucleic acid complex can release a potent RNAi duplex formed by a core nucleic acid strand and a passenger nucleic acid strand, which can specifically inhibit or silence a target RNA. The target RNA can have a sequence independent from the input nucleic acid strand. The target RNA can be from a gene that is different from the gene that the input nucleic acid strand is from. The target RNA can be from a gene that is the same as the gene that the input nucleic acid strand is from.

[0051] **FIG. 1** illustrates a schematic representation of two non-limiting exemplary nucleic acid complex constructs. In some embodiments, the nucleic acid complexes described herein comprise a core nucleic acid strand (e.g. a first nucleic acid strand), a passenger nucleic acid strand (e.g. a second nucleic acid strand), and a sensor nucleic acid strand (e.g. a third nucleic acid strand) as shown in a non-limiting embodiment of **FIG. 2**. These three strands can base-pair with one another to form, for example, a RNAi duplex (e.g. a first nucleic acid duplex) and a sensor duplex (e.g. a second nucleic acid duplex). One or more of the core nucleic acid strand, the passenger nucleic acid strand, and the sensor nucleic acid strand can be RNA analogs comprising modified nucleotides.

[0052] The term “nucleic acid duplex” as used herein refers to two single-stranded polynucleotides bound to each other through complementarily binding. The nucleic acid duplex can form a helical structure, such as a double-stranded RNA molecule, which is maintained largely by non-covalent bonding of base pairs between the two single-stranded polynucleotides

and by base stacking interactions.

[0053] The core nucleic acid strand can comprise a 5' region, a 3' region, and a central region between the 5' region and the 3' region. The central region of the core nucleic acid strand can be linked to the 5' region and/or the 3' region of the core nucleic acid strand via a connector. In some embodiments, the central region of the core nucleic acid strand is linked to the 5' region of the core nucleic acid strand via a 5' connector. In some embodiments, the central region of the core nucleic acid strand is linked to the 3' region of the core nucleic acid strand via a 3' connector. The central region of the core nucleic acid strand is complementarily bound to the passenger nucleic acid strand to form a RNAi duplex (e.g. a first nucleic acid duplex). Not the entire sequence of the core nucleic acid strand is complementarily bound to the passenger nucleic acid strand. For example, the 5' region and the 3' region of the core nucleic acid strand is not complementarily bound to the passenger nucleic acid strand.

[0054] The central region of the core nucleic acid strand can comprise a sequence complementary to a target nucleic acid (e.g. a RNA to be silenced). The core nucleic acid strand of the nucleic acid complex therefore acts as a guide strand (antisense strand) and is used to base pair with a target RNA. The passenger nucleic acid strand can therefore comprise a sequence homologous to the same target nucleic acid.

[0055] Upon activation of the nucleic acid complex (e.g. binding to an input nucleic acid strand), the released RNAi duplex can complementarily bind a target nucleic acid through the binding between the target nucleic acid and the central region of the core nucleic acid strand. In some embodiments, the sequence complementary to a target RNA in the core nucleic acid strand can be about 10-35 nucleosides in length. In some embodiments, the core nucleic acid strand comprises 20-70 linked nucleosides.

[0056] The sensor nucleic acid strand is complementarily bound to the 5' region and the 3' region of the core nucleic acid strand to form a sensor duplex (e.g. a second nucleic acid duplex). The sensor nucleic acid strand does not bind to the central region of the core nucleic acid strand nor the passenger nucleic acid strand.

[0057] The sensor nucleic acid strand can comprise an overhang. The term "overhang" as used herein refers to a stretch of unpaired nucleotides that protrudes at one of the ends of a double-stranded polynucleotide (e.g. a duplex). An overhang can be on either strand of the polynucleotide and can be included at either the 3' terminus of the strand (3' overhang) or at the 5' terminus of the strand (5' overhang). The overhang can be at the 3' terminus of the sensor nucleic acid strand. The overhang of the sensor nucleic acid strand does not bind to any region of the core nucleic acid strand.

[0058] The sensor nucleic acid strand can comprise a sequence capable of binding to

an input nucleic acid strand (e.g. a mRNA of a disease biomarker gene specific to a target cell, including a disease-related cell). Upon activation, the binding of the sensor nucleic acid strand to the input nucleic acid strand can cause displacement and subsequent release of the sensor nucleic acid strand from the core nucleic acid strand, thereby releasing the potent RNAi duplex and switching on the RNA interfering activity of the RNAi duplex. In the absence of an input nucleic acid strand or a detectable amount of the input nucleic acid strand, the nucleic acid complex herein described remains in an inactivated state (switched off) and the displacement of the sensor nucleic acid strand from the core nucleic acid strand does not take place. Therefore, the input nucleic acid strand can act as a trigger to activate (switch on) the RNA interfering activity of the nucleic acid complex (e.g. RNAi duplex).

[0059] The length of the RNAi duplex of the nucleic acid complex herein described can vary in different embodiments. In some embodiments, the length of the RNAi duplex can be 10-35 nucleotides. For example, the length of the RNAi duplex can be, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, a range of any two of these values, nucleotides. In some embodiments, the length of the RNAi duplex can be 19-25 nucleotides. In some embodiments, the length of the RNAi duplex can be 17-22 nucleotides. In some embodiments, the length of the RNAi duplex is about 21 nucleotides.

[0060] The length of the sensor duplex of the nucleic acid complex herein described can vary in different embodiments. In some embodiments, the length of the sensor duplex can be 10-35 nucleotides. For example, the length of the sensor duplex can be, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, a range of any two of these values, nucleotides.

[0061] In some embodiments, the nucleic acid complexes herein described do not have a dicer cleavage site, and therefore the RNAi interference mediated by the nucleic acid complexes can bypass Dicer-mediated cleavage.

[0062] As will be apparent to a skilled artisan, Dicer is an endoribonuclease in the RNase III family that can initiate the RNAi pathway by cleaving double-stranded RNA (dsRNA) molecule into short fragments of dsRNAs about 20-25 nucleotides in length.

[0063] In some embodiments, the nucleic acid complexes herein described differentiate from the conditionally activated small interfering RNAs (Cond-siRNAs) disclosed in the international application published as WO 2020/033938 in that the nucleic acid complexes herein described can bypass the Dicer processing. Cond-siRNAs have previously demonstrated *in vitro* control over RNAi activity. However, Cond-siRNAs require Dicer cleavage at specific sites (see, for example, Design 1 in FIG. 1) for RNAi activation. Furthermore, the suppression of OFF-state RNAi activity was achieved via steric hindrance of Dicer binding and activity.

[0064] In some embodiments, the nucleic acid complexes disclosed herein have structural features that discourage the Dicer binding. In some embodiments, the RNAi duplex does not create a Dicer substrate. For example, the RNAi duplex formed by the passenger nucleic acid strand and the central region of the core nucleic acid strand do not have a 3' and/or 5' overhang, but instead forming a blunt end that can render the passenger nucleic acid strand unfavorable for Dicer binding. In some embodiments, the passenger nucleic acid strand has about 17-22 nucleotides in length (e.g., 21 nucleotides), making it short enough to bypass Dicer cleavage. In some embodiments, the passenger nucleic acid strand does not have G/C rich bases to the 3' and/or 5' end of the passenger nucleic acid strand. In some embodiments, the passenger nucleic acid strand are attached to a terminal moiety to avoid Dicer binding.

[0065] In some embodiments of the nucleic acid complex disclosed herein, extensive or full chemical modification is introduced to one or more strands of the nucleic acid complex to improve *in vivo* potency, duration of drug activity, and suppression of background RNAi activity. Dicer cleavage would be impeded by full chemical modification of the siRNA domain and the sensor domain, as chemical modifications such as 2'-O-methyl bases and phosphorothioate backbones are known to impede endonuclease activity. In order to allow full chemical modification of the siRNA domain, it is advantageous to change the geometry and the functions of the core and passenger strands on the modified nucleic acid complex construct versus the original Cond-siRNA.

[0066] In some embodiments, the siRNA domain duplex in the modified nucleic acid complex disclosed herein is shorten such that the siRNA domain of the nucleic acid complex does not need to undergo endonuclease cleavage by Dicer in order to have active RNAi activity. In some embodiments, the siRNA domain duplex of a modified nucleic acid complex is about 21 base pairs compared to the 23 base-pair siRNA domain duplex in the Cond-siRNA construct.

[0067] To allow the sensor to still block RNAi activity in the OFF state, the guide strand for the siRNA (the strand antisense to the mRNA targeted for RNAi knockdown) is incorporated into the core strand, while the strand homologous to the mRNA target is now the passenger strand. Accordingly, the modified nucleic acid complex cannot have RNAi activity against the mRNA target until the 5' overhang on the siRNA domain is removed by exonuclease or endonuclease activity. When the nucleic acid complex is in the OFF state, the sensor duplex protects the overhang from degradation, ensuring that the RNAi activity stays OFF. When the sensor strand is removed by base-pairing to an RNA marker, the 5' and 3' overhangs on the siRNA domain become exposed, allowing them to be degraded by cellular nucleases. In order to prevent exonucleases from degrading the released siRNA, in some embodiments 5' and 3' exonuclease blocking domains are incorporated in the core strand (see, for example, FIG. 2).

[0068] The nucleic acid complexes herein described can be synthesized using standard methods for oligonucleotide synthesis well-known in the art including, for example, Oligonucleotide Synthesis by Herdewijin, Piet (2005) and Modified oligonucleotides: Synthesis and Strategy for Users, by Verma and Eckstein, *Annu Rev. Biochem.* (1998): 67:99-134, the contents of which are incorporated herein by reference in their entirety. The synthesized nucleic acid complexes can be allowed to form its secondary structure under a desirable physiological condition as will be apparent to a skilled artisan. The formed secondary structure can be tested using standard methods known in the art such as chemical mapping, NMR, or computational simulations. The nucleic acid complex construct can be further modified, according to the test result, by introducing or removing chemical modifications or mismatches, as necessary, until the desired structure is obtained.

[0069] Suitable software suites can be used to aid in the design and analysis of nucleic acid structures. For example, Nupack can be used to check the formation of the duplexes and to rank the thermodynamic stability of the duplexes. Oligonucleotide design tools can be used to optimize the placement of LNA modifications. Any of the regions of one or more of the strands in a nucleic acid complex herein described can be screened for an input nucleic acid sequence, a target nucleic acid sequence and/or chemical modifications herein described. For example, **FIG. 3** illustrates a schematic representation of a non-limiting exemplary nucleic acid complex construct, highlighting in yellow the terminal bases that can be screened for chemical modifications such as LNA placements and other nucleotide analogs herein described .

RNA interference (RNAi)

[0070] Described herein are nucleic acid complexes that can be conditionally activated to switch from an assembled, inactivated state to an activated state to act on (e.g. degrade or inhibit) a specific target nucleic acid in response to the detection of an input nucleic acid (e.g. a nucleic acid sequence specific to a target cell, including a disease-related cell) having a sequence complementary to a sequence in the sensor nucleic acid strand of a nucleic acid complex. In the assembled, inactivated configuration, the sensor nucleic acid strand of the nucleic acid complex inhibits enzymatic processing of the RNAi duplex, thereby keeping RNAi activity switched off. In the event that an input nucleic acid strand complementary to the sensor nucleic acid strand of a nucleic acid complex is present, the input nucleic acid strand can activate the nucleic acid complex by inducing separation of the sensor nucleic acid strand from the core nucleic acid strand via toehold mediated strand displacement. Displacement can start from a toehold formed at the 3' or 5' terminus of the sensor nucleic acid strand (e.g. a toehold formed at the 3' terminus of the sensor nucleic acid strand) through a complementary binding

between the input nucleic acid strand and an overhang of the sensor nucleic acid strand. **FIG. 4A** is a schematic diagram showing the activation of a nucleic acid complex in targeted cells following the base-pairing of the sensor strand to a RNA marker. **[0064]** After removal of the sensor nucleic acid strand, the 3' and 5' region of the core nucleic acid strand become 3' and 5' overhangs that can be degraded by nucleases (e.g. exonuclease). This degradation stops at the 3' end and 5' end of the RNAi duplex due to the presence of chemically modified nucleotides and/or exonuclease cleavage-resistance moieties, thus rendering an active RNAi duplex for further endonuclease processing if needed and RNA-induced silencing complex (RISC) loading. RISC is a multiprotein complex that incorporates one strand of a siRNA or miRNA and uses the siRNA or miRNA as a template for recognizing complementary target nucleic acid. Once a target nucleic acid is identified, RISC activates RNase (e.g. Argonaute) and inhibits the target nucleic acid by cleavage. In some embodiments, Dicer is not required for loading the RNAi duplex into RISC. **FIG. 4B** is a schematic diagram showing the formation of an active RNAi duplex following the displacement of a sensor nucleic acid strand from a core nucleic acid strand and the degradation of the core nucleic acid strand overhangs.

[0071] The passenger nucleic acid strand is then discarded, while the core nucleic acid strand (the central region of the core nucleic acid strand) is incorporated into RICS. The core nucleic acid strand of the nucleic acid complex disclosed herein acts as a guide strand (antisense strand) and is used to base pair with a target RNA. The passenger nucleic acid strand acts as a protecting strand prior to the loading of the core nucleic acid strand into RICS. RICS uses the incorporated core nucleic acid strand as a template for recognizing a target RNA that has complementary sequence to the core nucleic acid strand, particularly the central region of the core nucleic acid strand. Upon binding to the target RNA, the catalytic component of RICS, Argonaute, is activated which can degrade the bound target RNA. The target RNA can be degraded or the translation of the target RNA can be inhibited.

[0072] Upon activation, the nucleic acid complex herein described can inhibit a target nucleic acid in target cells, therefore resulting in a reduction or loss of expression of the target nucleic acid in the target cells. The target cells are cells associated or related to a disease or disorder. The term "associated to" "related to" as used herein refers to a relation between the cells and the disease or condition such that the occurrence of a disease or condition is accompanied by the occurrence of the target cells, which includes but is not limited to a cause-effect relation and sign/symptoms-disease relation. The target cells used herein typically have a detectable expression of an input nucleic acid.

[0073] In some embodiments, the expression of a target nucleic acid in target cells is inhibited about, at least, at least about, 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%,

20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%, or a number or a range between any of these values.

[0074] As used herein, inhibition of gene expression refers to the absence or observable decrease in the level of protein and/or mRNA product from a target gene in target cells. The degree of inhibition can be evaluated by examination of the expression level of the target gene as demonstrated in the examples.

[0075] Gene expression and/or the inhibition of target gene expression can be determined by use of a reporter or drug resistance gene whose protein product is easily assayed. Exemplary reporter genes include, but are not limited to, acetohydroxyacid synthase (AHAS), alkaline phosphatase (AP), beta galactosidase (LacZ), beta glucuronidase (GUS), chloramphenicol acetyltransferase (CAT), green fluorescent protein (GFP), horseradish peroxidase (HRP), luciferase (Luc), nopaline synthase (NOS), octopine synthase (OCS), and derivatives thereof. Multiple selectable markers are available that confer resistance to ampicillin, bleomycin, chloramphenicol, gentamycin, hygromycin, kanamycin, lincomycin, methotrexate, phosphinothricin, puromycin, and tetracyclin. Quantitation of the amount of gene expression allows one to determine a degree of inhibition as compared to cells not treated with the nucleic acid complexes or treated with a negative or positive control. Various biochemical techniques may be employed as will be apparent to a skilled artisan such as RNA solution hybridization, nuclease protection, Northern hybridization, reverse transcription, gene expression monitoring with a microarray, antibody binding, enzyme linked immunosorbent assay (ELISA), Western blotting, radioimmunoassay (RIA), other immunoassays, and fluorescence activated cell analysis (FACS).

[0076] In some embodiments, the nucleic acid complexes disclosed herein exhibit improved switching performance and reduced off-target effects. The nucleic acid complexes disclosed herein can have a reduced unwanted RNAi activity when the nucleic acid complexes are in an inactivated state (switched off) and an enhanced RNAi activity when the nucleic acid complexes are activated upon detection of an input nucleic acid strand.

[0077] In some embodiments, the expression of a target nucleic acid in non-target cells (e.g. cells not having an input nucleic acid strand) is inhibited about, at most, or at most about 1%, 2%, 3%, 4%, 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%,

34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, or a number or a range between any of these values. Non-target cells can comprise cells of the subject other than target cells.

[0078] In some embodiments, the nucleic acid complexes herein described have an enhanced potency, thus capable of evoking an RNAi activity at low concentrations. Nonspecific, off-target effects and toxicity (e.g. undesired proinflammatory responses) can be minimized by using low concentrations of the nucleic acid complexes.

[0079] The concentration of the nucleic acid complexes disclosed herein can vary in different embodiments. In some embodiments, the nucleic acid complexes disclosed herein can be provided at a concentration of, about, at most, or at most about, 0.001 nM, 0.01 nM, 0.02 nM, 0.03 nM, 0.04 nM, 0.05 nM, 0.06 nM, 0.07 nM, 0.08 nM, 0.09 nM, 0.1 nM, 0.2 nM, 0.3 nM, 0.4 nM, 0.5 nM, 0.6 nM, 0.7 nM, 0.8 nM, 0.9 nM, 1.0 nM, 1.5 nM, 2.0 nM, 2.5 nM, 3.0 nM, 3.5 nM, 4.0 nM, 4.5 nM, 5.0 nM, 5.5 nM, 6.0 nM, 6.5 nM, 7.0 nM, 7.5 nM, 8.0 nM, 8.5 nM, 9.0 nM, 9.5 nM, 10 nM, 11 nM, 12 nM, 13 nM, 14 nM, 15 nM, 16 nM, 17 nM, 18 nM, 19 nM, 20 nM, 30 nM, 40 nM, 50 nM, or a number or a range between any two of these values. For example, the nucleic acid complexes disclosed herein can be provided at a concentration between about 0.1-10 nM, preferably between about 0.1-1 nM. In some embodiments, the nucleic acid complex herein disclosed has a transfection concentration at about 0.1 nM or lower.

[0080] The nucleic acid complex herein described can allow lasting and consistently potent inhibition effects at low concentrations. For example, the nucleic acid complex can remain active for an extended period of time such as at least 12 hours, at least 24 hours, at least 36 hours, at least 48 hours, at least 60 hours, at least 72 hours, at least 84 hours, or at least 96 hours. In some embodiments, the nucleic acid complex can remain active for up to 30 days, up to 60 days, or up to 90 days.

Chemical modification

[0081] The nucleic acid strands (the core nucleic acid strand, the passenger nucleic acid strand, and/or the sensor nucleic acid strand) comprised in the nucleic acid complexes herein described can be a non-standard, modified nucleic acid strand comprising non-standard, modified nucleotides (nucleotide analog) or non-standard, modified nucleosides (nucleoside analog). The term “nucleotide analog” or “modified nucleotide” refers to a non-standard nucleotide comprising one or more modifications (e.g. chemical modifications), including non-naturally occurring ribonucleotides or deoxyribonucleotides. The term “nucleoside analog” or “modified nucleoside” refers to a non-standard nucleoside comprising one or more modification (e.g. chemical modification), including non-naturally occurring nucleosides other than cytidine,

uridine, adenosine, guanosine, and thymidine. The modified nucleoside can be a modified nucleotide without a phosphate group. The chemical modifications can include replacement of one or more atoms or moieties with a different atom or a different moiety or functional group (e.g. methyl group or hydroxyl group).

[0082] The modifications are introduced to alter certain chemical properties of the nucleotide/nucleoside such as to increase thermodynamic stability, to increase resistance to nuclease degradation (e.g. exonuclease resistant), and/or to increase binding specificity and minimize off-target effects. For example, thermodynamic stability can be determined based on measurement of melting temperature T_m . A higher T_m can be associated with a more thermodynamically stable chemical entity.

[0083] In some embodiments, the modification can render one or more of the nucleic acid strands in the nucleic acid complex to resist exonuclease degradation/cleavage. The term “exonuclease” as used herein, indicates a type of enzyme that works by cleaving nucleotides one at a time from the end (exo) of a polynucleotide chain. A hydrolyzing reaction that breaks phosphodiester bonds at either the 3' or the 5' end occurs. A 3' and 5' exonuclease can degrade RNA and DNA in cells, and can degrade RNA and DNA in the interstitial space between cells and in plasma, with a high efficiency and a fast kinetic rate. A close relative is the endonuclease, which cleaves phosphodiester bonds in the middle (endo) of a polynucleotide chain. 3' and 5' exonuclease and exonucleolytic complexes can degrade RNA and DNA in cells, and can degrade RNA and DNA in the interstitial space between cells and in plasma. The term “exoribonuclease” as used herein, refers to exonuclease ribonucleases, which are enzymes that degrade RNA by removing terminal nucleotides from either the 5' end or the 3' end of the RNA molecule. Enzymes that remove nucleotides from the 5' end are called 5'-3' exoribonucleases, and enzymes that remove nucleotides from the 3' end are called 3'-5' exoribonucleases.

[0084] The modification can comprise phosphonate modification, ribose modification (in the sugar portion), and/or base modification. Preferred modified nucleotides used herein include sugar- and/or backbone-modified ribonucleotides.

[0085] In some embodiments, the modified nucleotide can comprise modifications to the sugar portion of the nucleotides. For example, the 2' OH-group of a nucleotide can be replaced by a group selected from H, OR, R, F, Cl, Br, I, SH, SR, NH₂, NHR, NR₂, COOR, or OR, wherein R is substituted or unsubstituted C₁-C₆ alkyl, alkenyl, alkynyl, aryl, etc. In some embodiments, the 2' OH-group of a nucleotide or nucleoside is replaced by 2' O-methyl group and the modified nucleotide or nucleoside is a 2'-O-methyl nucleotide or 2'-O-methyl nucleoside (2'-OMe). The 2'-O-methyl nucleotide or 2'-O-methyl nucleoside can be 2'-O-methyladenosine, 2'-O-methylguanosine, 2'-O-methyluridine, or 2'-O-methylcytidine. In some

embodiments, the 2' OH-group of a nucleotide is replaced by fluorine (F), and the modified nucleotide or nucleoside is a 2'-F nucleotide or 2'-F nucleoside (2'-deoxy-2'-fluoro or 2'-F). The 2'-F nucleotide or 2'-F nucleoside can be 2'-F-adenosine, 2'-F-guanosine, 2'-F-uridine, or 2'-F-cytidine. The modifications can also include other modifications such as nucleoside analog phosphoramidites. In some embodiments, glycol nucleic acids can be used.

[0086] In some embodiments, the modified nucleotide can comprise a modification in the phosphate group of the nucleotide, e.g. by substituting one or more of the oxygens of the phosphate group with sulfur or a methyl group. In some embodiments, one or more of the nonbridging oxygens of the phosphate group of a nucleotide is replaced by a sulfur.

[0087] In some embodiments, the nucleic acid strands herein described comprise one or more non-standard internucleoside linkage that is not a phosphodiester linkage. In some embodiments, the nucleic acid strands herein described comprise one or more phosphorothioate internucleoside linkages. The term "phosphorothioate linkage" (PS) as used herein, indicates a bond between nucleotides in which one of the nonbridging oxygens is replaced by a sulfur. In some embodiments, both nonbridging oxygens may be replaced by a sulfur (PS2). In some embodiments, one of the nonbridging oxygens may be replaced by a methyl group. The term "phosphodiester linkage" as described herein indicates the normal sugar phosphate backbone linkage in DNA and RNA wherein a phosphate bridges the two sugars. In some embodiments, the introduction of one or more phosphorothioate linkage in the core nucleic acid strand, the passenger nucleic acid strand, and/or the sensor nucleic acid strand can endow the modified nucleotides with increased resistance to nucleases (e.g. endonucleases and/or exonucleases).

[0088] In some embodiments, the modified nucleotide can comprise modifications to or substitution of the base portion of a nucleotide. For example, uridine and cytidine residues can be substituted with pseudouridine, 2-thiouridine, N6-methyladenosine, 5-methylcytidine or other base analogs of uridine and cytidine residues. Adenosine can comprise modifications to Hoogsteen (e.g. 7-triazolo-8-aza-7-deazaadenosines) and/or Watson-Crick face of adenosine (e.g. N2-alkyl-2-aminopurines). Examples of adenosine analogs also include Hoogsteen or Watson-Crick face-localized N-ethylpiperidine triazole-modified adenosine analogs, N-ethylpiperidine 7-EAA triazole (e.g. 7-EAA, 7-ethynyl-8-aza-7-deazaadenosine) and other adenosine analogs identifiable to a person skilled in the art. Cytosine may be substituted with any suitable cytosine analogs identifiable to a person skilled in the art. For example, cytosine can be substituted with 6'-phenylpyrrolocytosine (PhpC) which has shown comparable base pairing fidelity, thermal stability and high fluorescence.

[0089] One or more nucleotides in the nucleic acid complex disclosed herein can be substituted with a universal base. The term "universal base" refers to nucleotide analogs that

form base pairs with each of the natural nucleotides with little discrimination between them. Examples of universal bases include, but are not limited to, C-phenyl, C-naphthyl and other aromatic derivatives, inosine, azole carboxamides, and nitroazole derivatives such as 3-nitropyrrole, 4-nitroindole, 5-nitroindole, and 6-nitroindole as known in the art (*see e.g.*, Loakes, 2001, *Nucleic Acids Research*, 29, 2437-2447).

[0090] In some embodiments, base modification disclosed herein can reduce innate immune recognition while making the nucleic acid complex more resistant to nucleases. Examples of base modifications that can be used in the nucleic acid complex disclosed herein are also described, for example, in Hu et al. (*Signal Transduction and targeted Therapy* 5: 101 (2020)), the content of which is incorporated by reference in its entirety

[0091] In some embodiments, the nucleic acid strands (the core nucleic acid strand, the passenger nucleic acid strand, and/or the sensor nucleic acid strand) comprised in the nucleic acid complexes herein described can comprise one or more locked nucleic acids or analogs thereof. Exemplary locked nucleic acid analogs include, for example, their corresponding locked analog phosphoramidites and other derivatives apparent to a skilled artisan.

[0092] As used herein, the term “locked nucleic acids” (LNA) indicates a modified RNA nucleotide. The ribose moiety of an LNA nucleotide is modified with an extra bridge connecting the 2' and 4' carbons (a 2'-O, 4'-C methylene bridge). The bridge “locks” the ribose in the 3'-endo structural conformation and restricts the flexibility of the ribofuranose ring, thereby locking the structure into a rigid bicyclic formation. LNA nucleotides can be mixed with DNA or RNA bases in the oligonucleotide whenever desired. The incorporation of LNA into the nucleic acid complexes disclosed herein can increase the thermal stability (e.g. melting temperature), hybridization specificity of oligonucleotides as well as accuracies in allelic discrimination. LNA oligonucleotides display hybridization affinity toward complementary single-stranded RNA and complementary single- or double-stranded DNA. Additional information about LNA can be found, for example, at www.sigmaaldrich.com/technical-documents/articles/biology/locked-nucleic-acids-faq.html. In some embodiments, glycol nucleic acids can be used.

[0093] The nucleic acid strands (the core nucleic acid strand, the passenger nucleic acid strand, and/or the sensor nucleic acid strand) comprised in the nucleic acid complexes herein described can comprise other chemically modified nucleotide or nucleoside with 2'-4' bridging modifications. A 2'-4' bridging modification refers to the introduction of a bridge connecting the 2' and 4' carbons of a nucleotide. The bridge can be a 2'-O, 4'-C methylene bridge (e.g. in LNA). The bridge can also be a 2'-O, 4'-C ethylene bridge (e.g. in ethylene-bridged nucleic acids (ENA)) or any other chemical linkage identifiable known in the art.

[0094] The introduction of LNA, analogues thereof, or other chemically modified nucleotides with 2'-4' bridging modifications in the nucleic acid complex herein described can enhance hybridization stability as well as mismatch discrimination. For example, a nucleic acid complex comprising a sensor nucleic acid strand with LNA, analogues thereof, or other chemically modified nucleotides with 2'-4' bridging modifications can have an enhanced sensitivity to distinguish between matched and mismatched input nucleic acid strand (e.g. in the complementary binding between an input nucleic acid strand and a sensor nucleic acid strand).

[0095] One or more of the nucleic acid strands of the nucleic acid complex can comprise a chemical moiety linked to the 3' and/or 5' terminus of the strand. The terminal moiety can include one or more any suitable terminal linkers or modifications. For example, the terminal moiety can include a linker to link the oligonucleotide with another molecule or a particular surface (biotins, amino-modifiers, alkynes, thiol modifiers, azide, N-Hydroxysuccinimide, and cholesterol), a dye (e.g. fluorophore or a dark quencher), a fluorine modified ribose, a space (e.g. C3 spacer, Spacer 9, Spacer 18, dSpacer, tri-ethylene glycol spacer, hexa-ethylene glycol spacer), moieties and chemical modification involved in click chemistry (e.g. alkyne and azide moieties), and any linkers or terminal modifications that can be used to attach the 3' and 5' end to other chemical moieties such as antibodies, gold or other metallic nanoparticles, polymeric nanoparticles, dendrimer nanoparticles, small molecules, single chain or branched fatty acids, peptides, proteins, aptamers, and other nucleic acid strands and nucleic acid nanostructures. The terminal moiety can serve as a label capable of detection or a blocker to protect a single-stranded nucleic acid from nuclease degradation. Additional linkers and terminal modification that can be attached to the terminus of the sensor nucleic acid strand are described in www.idtdna.com/pages/products/custom-dna-rna/oligo-modifications and www.glenresearch.com/browse/labels-and-modifiers, the contents of which are incorporated herein by reference in their entirety.

[0096] Additional modifications to the nucleotides and/or nucleosides can also be introduced to one or more strands of the nucleic acid complex herein described, such as modifications described in Hu et al. (Signal Transduction and targeted Therapy 5: 101 (2020)), the content of which is incorporated by reference in its entirety.

Ribose modification

[0097] The percentage of the modified nucleosides of the nucleic acid complex can vary in different embodiments. In some embodiments, the percentage of the modified nucleosides of the nucleic acid complex herein described can be, be about, be at least, or be at least about 80%, 85%, 90%, or 95%. For example, percentage of the modified nucleosides of the nucleic acid complex herein described can be, be about, be at least, or be at least about 80%,

81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%, or a number or a range between any two of these values. In some embodiments, at least 90%, 91%, 92%, 93%, 94%, 95% , or a number or a range between any two of these values of the nucleotides of the nucleic acid complex are modified (e.g. are non-DNA and non-RNA). In some embodiments, all of the nucleotides of the nucleic acid complex are modified (e.g. are non-DNA and non-RNA).

[0098] The percentage of the modified nucleosides in one or more strands of the nucleic acid complex can vary in different embodiments. In some embodiments, the percentage of the modified nucleosides in a core nucleic acid strand herein described can be, be about, be at least, or be at least about 80%, 85%, 90%, or 95%. For example, the percentage of the modified nucleosides in a core nucleic acid strand herein described can be, be about, be at least, or be at least about 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%, or a number or a range between any two of these values. In some embodiments, all of the nucleosides of a core nucleic acid strand are chemically modified.

[0099] The percentage of the modified nucleosides in the central region of a core nucleic acid strand herein described can be, be about, be at least, or be at least about 80%, 85%, 90%, or 95%. For example, the percentage of the modified nucleosides in the central region of a core nucleic acid strand herein described can be, be about, be at least, or be at least about 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%, or a number or a range between any two of these values.

[0100] The percentage of the modified nucleosides in the 5' region of a core nucleic acid strand herein described can be, be about, be at least, or be at least about 80%, 85%, 90%, or 95%. For example, the percentage of the modified nucleosides in the 5' region of a core nucleic acid strand herein described can be, be about, be at least, or be at least about 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%, or a number or a range between any two of these values. In some embodiments, all of the nucleosides of the 5' region of a core nucleic acid strand are chemically modified.

[0101] The percentage of the modified nucleosides in the 3' region of a core nucleic acid strand herein described can be, be about, be at least, or be at least about 80%, 85%, 90%, or 95%. For example, the percentage of the modified nucleosides in the 3' region of a core nucleic acid strand herein described can be, be about, be at least, or be at least about 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%, or a number or a range between any two of these values. In some embodiments, all of the nucleosides of the 3' region of a core nucleic acid strand are chemically modified.

[0102] The percentage of the modified nucleosides in a passenger nucleic acid strand herein described can be, be about, be at least, or be at least about 80%, 85%, 90%, or 95%. For example, the percentage of the modified nucleosides in a passenger nucleic acid strand herein described can be, be about, be at least, or be at least about 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%, or a number or a range between any two of these values. In some embodiments, all of the nucleosides of a passenger nucleic acid strand are chemically modified.

[0103] The percentage of the modified nucleosides in a sensor nucleic acid strand herein described can be, be about, be at least, or be at least about 80%, 85%, 90%, or 95%. In some embodiments, the percentage of the modified nucleosides in a sensor nucleic acid strand herein described can be, be about, be at least, or be at least about 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%, or a number or a range between any two of these values. In some embodiments, all of the nucleosides of a sensor nucleic acid strand are chemically modified.

[0104] The modified nucleosides in one or more of the core nucleic acid strand, the passenger nucleic acid strand, and the sensor nucleic acid strand can comprise 2'-O-methyl nucleoside and/or 2'-F nucleoside.

[0105] In some embodiments, the percentage of 2'-O-methyl nucleoside and/or 2'-F nucleoside in the nucleic acid complex herein described can be, be about, be at least, be at least about, be at most, or be at most about 10%-50%. For example, the percentage of 2'-O-methyl nucleoside and/or 2'-F nucleoside in the nucleic acid complex herein described can be, be about, be at least, be at least about, be at most, or be at most about 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, or a number or a range between any two of these values.

[0106] In some embodiments, the percentage of 2'-O-methyl nucleoside and/or 2'-F nucleoside in a core nucleic acid strand herein described can be, be about, be at least, be at least about, be at most, or be at most about 10%-50%. For example, the percentage of 2'-O-methyl nucleoside and/or 2'-F nucleoside in a core nucleic acid strand herein described can be, be about, be at least, be at least about, be at most, or be at most about 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, or a number or a range between any two of these values.

[0107] In some embodiments, the percentage of 2'-O-methyl nucleoside and/or 2'-F nucleoside in a passenger nucleic acid strand herein described can be, be about, be at least, be at

least about, be at most, or be at most about 10%-50%. For example, the percentage of 2'-O-methyl nucleoside and/or 2'-F nucleoside in a passenger nucleic acid strand herein described can be, be about, be at least, be at least about, be at most, or be at most about 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, or a number or a range between any two of these values.

[0108] In some embodiments, the percentage of 2'-O-methyl nucleoside and/or 2'-F nucleoside in a sensor nucleic acid strand herein described can be, be about, be at least, be at least about, be at most, or be at most about 10%-50%. For example, the percentage of 2'-O-methyl nucleoside and/or 2'-F nucleoside in a sensor nucleic acid strand herein described can be, be about, be at least, be at least about, be at most, or be at most about 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, or a number or a range between any two of these values.

Phosphate modification

[0109] The percentage of phosphate modification to the nucleotides in the nucleic acid complex described herein can vary in different embodiments. In some embodiments, the phosphate modification comprises or is a phosphorothioate internucleoside linkage. In some embodiments, the percentage of phosphorothioate internucleoside linkages in a core nucleic acid strand is less than 5%, less than 10%, less than 25%, less than 50%, or a number or a range between any two of these values. For example, percentage of phosphorothioate internucleoside linkages in a core nucleic acid strand is about, less than, or less than about 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50% or a number or a range between any two of these values. In some embodiments, the core nucleic acid strand does not comprise a phosphorothioate internucleoside linkage modification.

[0110] In some embodiments, the percentage of phosphodiester internucleoside linkages in a core nucleic acid strand is about, at least, or at least about 50%, 80% or 95%, or a number or a range between any two of these values. For example, percentage of phosphodiester internucleoside linkages in a core nucleic acid strand is about, at least, or at least about 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or a number or a range between any two of these values. In some embodiments, all the

internucleoside linkages in the core nucleic acid strand are phosphodiester internucleoside linkage.

[0111] In some embodiments, the 5' terminus of the central region of the core nucleic acid strand comprises at least one phosphorothioate internucleoside linkage (e.g. one, two or three phosphorothioate internucleoside linkage). In some embodiments, the 3' terminus of the central region of the core nucleic acid strand comprises at least one phosphorothioate internucleoside linkage (e.g. one, two or three phosphorothioate internucleoside linkage). In some embodiments, each of the 5' terminus of the central region of the core nucleic acid strand and the 3' terminus of the central region of the core nucleic acid strand independently comprises one or more phosphorothioate internucleoside linkages (e.g. one, two or three phosphorothioate internucleoside linkage). In some embodiments, the central region of the core nucleic acid strand does not comprise phosphorothioate internucleoside linkages except for the phosphorothioate internucleoside linkage(s) between two or three nucleosides at the 5' terminus, 3' terminus, or both, of the central region.

[0112] In some embodiments, the internucleoside linkages between the one to three nucleotides (e.g. one, two, or three nucleotides) adjacent to the 3' of the 5' connector of the core nucleic acid strand are phosphorothioate internucleoside linkages. In some embodiments, the internucleoside linkages between the one or two nucleotides adjacent to the 5' of the 3' connector of the core nucleic acid strand are phosphorothioate internucleoside linkages. In some embodiments, the internucleoside linkages between the one to three nucleotides (e.g. one, two, or three nucleotides) adjacent to the 3' of the 3' connector of the core nucleic acid strand are phosphorothioate internucleoside linkages. In some embodiments, the 3' region of the core nucleic acid strand does not comprise phosphorothioate internucleoside linkages except for the phosphorothioate internucleoside linkage(s) between the one to three nucleotides (e.g. one, two, or three nucleotides) adjacent to the 3' of the 3' connector of the core nucleic acid strand. In some embodiments, the 5' region of the core nucleic acid strand does not comprise phosphorothioate internucleoside linkages.

[0113] In some embodiments, the passenger nucleic acid strand comprises one or more phosphorothioate internucleoside linkage. The percentage of phosphorothioate internucleoside linkages in a passenger nucleic acid strand is less than 5%, less than 10%, less than 25%, less than 50%, or a number or a range between any two of these values. For example, percentage of phosphorothioate internucleoside linkages in a passenger nucleic acid strand is about, less than, or less than about 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%,

48%, 49%, 50%, or a number or a range between any two of these values.

[0114] In some embodiments, the 5' terminus of the passenger nucleic acid strand comprises at least one phosphorothioate internucleoside linkage (e.g. one, two, or three phosphorothioate internucleoside linkage). In some embodiments, the 3' terminus of the passenger nucleic acid strand comprises at least one phosphorothioate internucleoside linkage (e.g. one, two, or three phosphorothioate internucleoside linkage). In some embodiments, the passenger nucleic acid strand does not comprise phosphorothioate internucleoside linkages except for the phosphorothioate internucleoside linkage(s) between the last two, three, or four nucleosides at the 5' terminus, 3' terminus, or both, of the passenger nucleic acid strand. In some embodiments, the passenger nucleic acid strand does not comprise phosphorothioate internucleoside linkages except for the phosphorothioate internucleoside linkage(s) between the last two to three nucleosides at the 5' terminus and the last two to three nucleosides at 3' terminus of the passenger nucleic acid strand.

[0115] In some embodiments, the sensor nucleic acid strand comprises one or more phosphorothioate internucleoside linkage. The percentage of phosphorothioate internucleoside linkages in a sensor nucleic acid strand can be less than 5%, less than 10%, less than 25%, less than 50%, or a number or a range between any two of these values. For example, percentage of phosphorothioate internucleoside linkages in a sensor nucleic acid strand is about, less than, or less than about 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50% or a number or a range between any two of these values.

[0116] In some embodiments, the 5' terminus of the sensor nucleic acid strand comprises at least one phosphorothioate internucleoside linkage (e.g. one, two or three phosphorothioate internucleoside linkage). In some embodiments, the 3' terminus of the sensor nucleic acid strand comprises at least one phosphorothioate internucleoside linkage (e.g. one to twenty phosphorothioate internucleoside linkage). In some embodiments, each of the 5' terminus of the sensor nucleic acid strand and the 3' terminus of the sensor nucleic acid strand independently comprises one or more phosphorothioate internucleoside linkages (e.g. one, two or three at the 5' terminus or one to twenty at the 3' terminus). In some embodiments, the sensor nucleic acid strand does not comprise phosphorothioate internucleoside linkages except for the phosphorothioate internucleoside linkage(s) at the 5' terminus, 3' terminus, or both, of the passenger nucleic acid strand. In some embodiments, the phosphorothioate internucleoside linkages at the 3' terminus of the passenger nucleic acid strand are in the singled-stranded overhang of the passenger nucleic acid strand.

LNA, analogues thereof and 2'-4' bridging modification

[0117] The percentage of the LNA or analogues thereof of the nucleic acid complex can vary in different embodiments. In some embodiments, the percentage of the LNA or analogues thereof of the nucleic acid complex herein described can be about 10%-50%. For example, the percentage of the LNA or analogues thereof of the nucleic acid complex herein described can be about, at most, at most about 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, or a number or a range between any two of these values.

[0118] The percentage of the LNA or analogues thereof in one or more strands of the nucleic acid complex can vary in different embodiments. In some embodiments, the percentage of the LNA or analogues thereof in a core nucleic acid strand herein described can be, be about, be at most, or be at most about 5%, 10%, or 15%. For example, the percentage of the LNA or analogues thereof of a core nucleic acid strand herein described can be, be about, be at most, or be at most about 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, or a number or a range between any two of these values.

[0119] In some embodiments, the percentage of the LNA or analogues thereof in a passenger nucleic acid strand herein described can be, be about, be at most, or be at most about 5%, 10%, or 15%. For example, the percentage of the LNA or analogues thereof of a passenger nucleic acid strand herein described can be, be about, be at most, or be at most about 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, or a number or a range between any two of these values. In some embodiments, a percentage of the LNA or analogues thereof in a passenger nucleic acid strand herein described greater than 5%, greater than 10%, or greater than 15% can decrease the RNAi activity of the nucleic acid complex (see **Example 1**).

[0120] In some embodiments, the percentage of the LNA or analogues thereof in a sensor nucleic acid strand herein described can be, be about, be at least, be at least about, be at most, or be at most about 10%-50%. For example, the percentage of the LNA or analogues thereof of a sensor nucleic acid strand herein described can be, be about, be at least, be at least about, be at most, or be at most about 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, or a number or a range between any two of these values.

[0121] The percentage of 2'-4' bridging modification of the nucleic acid complex can vary in different embodiments. In some embodiments, the percentage of the 2'-4' bridging modification of the nucleic acid complex herein described can be about 10%-50%. For example,

the percentage of the 2'-4' bridging modification of the nucleic acid complex herein described can be about, at most, at most about 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50% or a number or a range between any two of these values.

Core Strand

[0122] The core nucleic acid strand of the nucleic acid complex described herein can comprise a 5' region, a 3' region, and a central region between the 5' region and the 3' region. Each of the 5' region, the 3' region, and the central region can be directly adjacent to each other, that is no nucleotide between the two adjacent regions. In some embodiments, the 3' end of the 5' region can be 1, 2, 3, 4, 5, 8, 9, 10, 11, 12, 13, 14, 15, 20, or a number or a range between any two of these values, nucleotides away from the 5' end of the central region. In some embodiments, the 5' end of the 3' region can be 1, 2, 3, 4, 5, 8, 9, 10, 11, 12, 13, 14, 15, 20, or a number or a range between any two of these values, nucleotides away from 3' of the central region.

[0123] The length of the core nucleic acid strand can vary in different embodiments. In some embodiments, the core nucleic acid strand comprises 20-70 linked nucleosides. For example, the core nucleic acid strand can comprise 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, or 70 linked nucleosides.

[0124] The length of the central region of the core nucleic acid strand can vary in different embodiments. In some embodiments, the central region of the core nucleic acid strand comprises 10-35 linked nucleosides. For example, the central region of the core nucleic acid strand can comprise 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, or 35 linked nucleosides.

[0125] The 3' region and the 5' region of the core nucleic acid strand can have a same length or a different length. The length of the 3' region and the 5' region of the core nucleic acid strand can vary in different embodiments. In some embodiments, the length of the 3' region and the 5' region of the core nucleic acid strand comprises 2-33 linked nucleosides. For example, the 3' region and the 5' region of the core nucleic acid strand can comprise 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, or 33 linked nucleosides. In some embodiments, the 3' region and the 5' region each comprises 11 linked nucleotides that base-pair with a sensor strand to form a sensor duplex.

[0126] The central region of the core nucleic acid strand comprises a sequence

complementary to a target RNA. The length of the sequence complementary to a target RNA can vary in different embodiments. In some embodiments, the sequence complementary to a target RNA is 10-21 nucleotides in length. For example, the sequence complementary to a target RNA is 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, or 21 nucleotides in length. In some embodiments, a core nucleic acid strand comprises a central region of 21 nucleotides in length that base-pairs with a passenger strand to form a siRNA duplex.

[0127] The central region of the core nucleic acid strand comprises a sequence complementary to a passenger nucleic acid strand. The length of the sequence complementary to a passenger nucleic acid strand can vary in different embodiments. In some embodiments, the sequence complementary to a passenger nucleic acid strand is 19-25 nucleotides in length. For example, the sequence complementary to a passenger nucleic acid strand is 19, 20, 21, 22, 23, 24, or 25 nucleotides in length.

[0128] In some embodiments, the central region of the core nucleic acid strand is linked to the 5' region and the 3' region of the core nucleic acid strand via a connector. For example, the central region of the core nucleic acid strand is linked to the 5' region of the core nucleic acid strand via a 5' connector. In some embodiments, the central region of the core nucleic acid strand is linked to the 3' region of the core nucleic acid strand via a 3' connector.

[0129] The 5' connector and/or 3' connector can comprise a three-carbon linker (C_3 linker), a nucleotide, any modified nucleotide described herein, or any moiety that can resist exonuclease cleavage when the core nucleic acid strand is single-stranded (e.g. after displacement of the sensor nucleic acid strand from the core nucleic acid strand). For example, the 5' connector and/or the 3' connector can comprise a 2'-F nucleotide such as 2'-F-adenosine, 2'-F-guanosine, 2'-F-uridine, or 2'-F-cytidine. The 5' connector and/or the 3' connector can comprise a 2'-O-methyl nucleotide such as 2'-O-methyladenosine, 2'-O-methylguanosine, 2'-O-methyluridine, or 2'-O-methylcytidine. The 5' connector and/or the 3' connector can comprise a naturally occurring nucleotide such as cytidine, uridine, adenosine, or guanosine. The 5' connector and/or the 3' connector of the core nucleic acid strand can comprise a phosphodiester linkage (phosphodiester 5' and 3' connection) cleavable by an exonuclease when in a single-stranded form. The 5' connector and/or the 3' connector of the core nucleic acid strand can comprise any suitable moiety that can resist exonuclease cleavage when in single-stranded form.

[0130] In some embodiments, the 5' connector can comprise or is, a C_3 3-carbon linker, a nucleotide, a modified nucleotide (2'-O-methyl nucleotide, 2'-F nucleotide), a nucleotide with a phosphodiester 5' and 3' connection cleavable by an exonuclease when in a single stranded form, or a combination thereof. In some embodiments, the 5' connector can comprise or is a 2'-O-methyl nucleotide such as 2'-O-methyladenosine, 2'-O-methylguanosine,

2'-O-methyluridine, or 2'-O-methylcytidine.

[0131] In some embodiments, the 3' connector comprises or is, a C₃ 3-carbon linker, a nucleotide, a modified nucleotide, an exonuclease cleavage-resistant moiety when in a single stranded form, or a combination thereof. In some embodiments, the 3' connector can comprise or is a 2'-O-methyl nucleotide such as 2'-O-methyladenosine, 2'-O-methylguanosine, 2'-O-methyluridine, or 2'-O-methylcytidine.

[0132] In some embodiments, the 3' connector comprises or is a 2'-O-methyl nucleotide such as 2'-O-methyladenosine, 2'-O-methylguanosine, 2'-O-methyluridine, or 2'-O-methylcytidine and the 5' connector comprises or is a 2'-O-methyl nucleotide such as 2'-O-methyladenosine, 2'-O-methylguanosine, 2'-O-methyluridine, or 2'-O-methylcytidine

[0133] In some embodiments, the 5' connector of the core nucleic acid strand does not comprise or is not a C₃ 3-carbon linker. In some embodiments, the 3' connector of the core nucleic acid strand comprises or is a C₃ 3-carbon linker. In some embodiments, it is advantageous to not have a C₃ 3-carbon linker as the 5' connector. In some embodiments, it is advantageous to have a C₃ 3-carbon linker as the 3' connector. In some embodiments, the 5' connector of the core nucleic acid strand does not comprise or is not a C₃ 3-carbon linker, while the 3' connector of the core nucleic acid strand comprises or is a C₃ 3-carbon linker.

[0134] In some embodiments, a nucleic acid complex not having a C₃ 3-carbon linker as the 5' connector exhibit higher RNA interfering activity (*see Examples 1-2*). The nucleic acid complex can have a modified nucleotide or a nucleotide as the 5' connector. The nucleic acid complex can have no 5' connector. The nucleic acid complex can have a C₃ 3-carbon linker, a modified nucleotide, or a nucleotide as the 3' connector. The nucleic acid complex can have no 3' connector. In some embodiments, not having a C₃ 3-carbon linker as the 5' connector increases RNA interfering activity of the nucleic acid complex by at least about 2-fold, 3-fold, 4-fold, 5-fold, 6-fold, 7-fold, 8-fold, 9-fold, 10-fold, 20-fold, 30-fold, 40-fold, 50-fold, 60-fold, 70-fold, 80-fold, 90-fold, 100-fold, or a number or a range between any of these value, greater than nucleic acid complexes having a C₃ 3-carbon linker as the 5' connector.

[0135] In some embodiments, a nucleic acid complex having a C₃ 3-carbon linker as the 3' connector exhibit higher RNA interfering activity (*see Examples 1-2*). The nucleic acid complex can have a modified nucleotide or a nucleotide as the 5' connector. The nucleic acid complex can have no 5' connector. The nucleic acid complex does not have a C₃ 3-carbon linker as the 5' connector. In some embodiments, having a C₃ 3-carbon linker as the 3' connector increases RNA interfering activity of the nucleic acid complex by at least about 2-fold, 3-fold, 4-fold, 5-fold, 6-fold, 7-fold, 8-fold, 9-fold, 10-fold, 15-fold, 20-fold, or a number or a range between any of these value, greater than nucleic acid complexes having a modified nucleotide

(e.g. 2'-O-methyl nucleotide) as the 3' connector. In some embodiments, having a C₃ 3-carbon linker as the 3' connector increases RNA interfering activity of the nucleic acid complex by at least about 2-fold, 3-fold, 4-fold, 5-fold, 6-fold, 7-fold, 8-fold, 9-fold, 10-fold, 15-fold, 20-fold, or a number or a range between any of these value, greater than nucleic acid complexes having no 3' connector.

[0136] In some embodiments, the core nucleic acid strand do not comprise a 5' connector and/or a 3' connector. Instead, the central region of the core nucleic acid strand is linked the 3' region and/or the 5' region via a standard phosphodiester linkage. In some embodiments, the central region of the core nucleic acid strand is linked to the 5' region of the core nucleic acid strand via a phosphodiester linkage. In some embodiments, the central region of the core nucleic acid strand is linked to the 3' region of the core nucleic acid strand via a phosphodiester linkage. In some embodiments, the central region of the core nucleic acid strand is linked to the 3' region of the core nucleic acid strand via a phosphodiester linkage, while the central region of the core nucleic acid strand is linked to the 5' region of the core nucleic acid strand via a 2'-O-methyl nucleotide such as 2'-O-methyladenosine, 2'-O-methylguanosine, 2'-O-methyluridine, or 2'-O-methylcytidine. In some embodiments, the central region of the core nucleic acid strand is linked to the 5' region of the core nucleic acid strand via a phosphodiester linkage, while the central region of the core nucleic acid strand is linked to the 3' region of the core nucleic acid strand via a 2'-O-methyl nucleotide such as 2'-O-methyladenosine, 2'-O-methylguanosine, 2'-O-methyluridine, or 2'-O-methylcytidine. In some embodiments, the central region of the core nucleic acid strand is linked to the 3' region and the 5' region of the core nucleic acid strand both via a phosphodiester linkage.

[0137] In an exemplary embodiment, a chemically modified core strand can comprise a 3' region and a 5' region each having 11 linked nucleotides in length that base pair with a sensor strand to form a sensor duplex and a central region having 21 nucleotides in length that base pair with a passenger strand to form a siRNA duplex (see, for example, FIG. 2). The 5' linker connecting the sensor duplex and siRNA duplex can consist of a 2'-O-methyl modified base with phosphodiester backbone connections and the 3' linker can be a C₃ spacer. The core strand can further comprise a 5' exonuclease blocking domain consisting of three chemically modified bases connected by two phosphorothioate backbone linkages and a 3' exonuclease blocking domain consisting of three chemically modified bases connected by the 3' linker and a phosphorothioate backbone linkage. All the nucleotides in the core strand are chemically modified, either by 2-O-methylation or 2'-F modification. For example, as shown in FIG. 2, bases 2, 6, 14 and 16 from the 5' end of the central region are 2'-F modified while the rest of the core strand is 2'-O-methylated.

[0138] In some embodiments, not having a 5' connector and/or a 3' connector increases RNA interfering activity of the nucleic acid complex by at least about 2-fold, 3-fold, 4-fold, 5-fold, 6-fold, 7-fold, 8-fold, 9-fold, 10-fold, 20-fold, 30-fold, 40-fold, 50-fold, 60-fold, 70-fold, 80-fold, 90-fold, 100-fold or a number or a range between any of these value, greater than nucleic acid complexes having a C₃ 3-carbon linker as the 5' connector.

Passenger Nucleic Acid Strand

[0139] The passenger nucleic acid strand of the nucleic acid complex described herein is complementary bound to the central region of the core nucleic acid strand to form a RNAi duplex (e.g. a first nucleic acid duplex). Since the central region of the core nucleic acid strand is complementary to a target nucleic acid strand, the passenger nucleic strand of the nucleic acid complex can comprise a sequence homologous to the target nucleic acid strand.

[0140] As used herein, the term “homologous” or “homology” refers to sequence identity between at least two sequences. The term “sequence identity” or “identity” in the context of two nucleic acid or polypeptide sequences makes reference to the nucleotide bases or residues in the two sequences that are the same when aligned for maximum correspondence over a specified comparison window.

[0141] In some embodiments, the sequence identity between a passenger nucleic acid strand and a target nucleic acid or a portion thereof can be, be about, be at least, or be at least about 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 100%, or a number or a range between any two of these values. The passenger nucleic acid strand of a nucleic acid complex can have a sequence substantially identical, e.g. at least 80%, 90%, or 100%, to a target nucleic acid or a portion thereof.

[0142] The length of the passenger nucleic acid strand can vary in different embodiments. In some embodiments, the passenger nucleic acid strand comprises 10-35 linked nucleosides. For example, the core nucleic acid strand can comprise 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, or 35 linked nucleosides. In some embodiments, the passenger nucleic acid strand comprises 17-21 linked nucleosides.

[0143] In some embodiments, the passenger nucleic acid strand has a 3' overhang, a 5' overhang, or both in the RNAi duplex. In some embodiments, the passenger nucleic acid strand has a 3' overhang, and the 3' overhang is one to five nucleosides in length.

[0144] In some embodiments, the overhang of the passenger nucleic acid strand is capable of binding to the input nucleic acid strand to form a toehold, thereby initiating a toehold mediated strand displacement and causing the displacement of the passenger nucleic acid strand

from the core nucleic acid strand.

[0145] In some embodiments, the overhang of the passenger nucleic acid strand is 5 to 20 nucleosides in length. For example, the overhang of the passenger nucleic acid strand can be 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 nucleosides in length. In some embodiments, the overhang of the passenger nucleic acid strand is 9 nucleosides in length.

[0146] In some embodiments, one or more internucleoside linkages of the overhang of the passenger nucleic acid strand are phosphorothioate internucleoside linkage which can protect the overhang from degradation. In some embodiments, all internucleoside linkages of the overhang of the passenger nucleic acid strand can be phosphorothioate internucleoside linkage.

[0147] In some embodiments, the passenger nucleic acid strand does not have a 3' overhang, a 5' overhang, or both in the RNAi duplex. In some embodiments, having a blunt end with no overhang can render the passenger nucleic acid strand unfavorable for Dicer binding, thereby bypassing the Dicer-mediated cleavage.

[0148] In an exemplary embodiment, a passenger nucleic acid strand is fully chemically modified and comprises 21 nucleotides that base pair with the central region of a core strand to form a siRNA complex (see, for example, FIG. 2). To provide resistance to nuclease activity, the first and last two backbone linkage on the 5' and 3' end of the passenger strand are phosphorothioate. Bases 9, 10 and 11 from the 5' end of the passenger strand are 2'-F modified to reduce unintended RNAi activity stemming from incorporation of the passenger strand into RISC. LNA modification(s) can be added (e.g., at base 2 from the 5' end of the passenger strand shown in FIG. 4) to enhance thermodynamic stability of the siRNA duplex, which can improve RNAi switching.

Sensor Nucleic Acid Strand

[0149] The sensor nucleic acid strand of the nucleic acid complex described herein comprises a region complementary bound to the 5' region and the 3' region of the core nucleic acid strand to form a sensor duplex (e.g. a second nucleic acid duplex). The length of the region complementary bound to the 5' region and the 3' region of the core nucleic acid strand can vary in different embodiments. In some embodiments, the region complementary bound to the 5' region and the 3' region of the core nucleic acid strand comprises 10-35 linked nucleosides. For example, the region in the sensor nucleic strand complementary bound to the 5' region and the 3' region of the core nucleic acid strand can comprise 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, or 35 linked nucleosides.

[0150] The sensor nucleic acid strand can comprise an overhang. The overhang can be at the 3' end or 5' end, or both, of the sensor nucleic acid strand. The overhang is not

complementary to the core nucleic acid strand and is capable of binding to an input nucleic acid strand, thereby initiating a toehold mediated strand displacement and causing the displacement of the passenger nucleic acid strand from the core nucleic acid strand.

[0151] The length of the overhang in the sensor nucleic acid strand can vary in different embodiments. In some embodiments, the length of the overhang can be 5-20 linked nucleotides. For example, the length of the overhang in the sensor nucleic acid strand can comprise 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 nucleotides in length. In some embodiments, the overhang of the sensor nucleic acid strand is 12 nucleotides in length.

[0152] The overhang of the sensor nucleic acid strand can comprise nucleotide modification introduced to improve the base-pairing affinity, nuclease resistance of the single-stranded overhang, and thermodynamic stability to avoid spurious exonuclease induced activation of the strand. Exemplary modifications include, but not limited to, 2'-O-methyl modification, 2'-Fluoro modifications, phosphorothioate internucleoside linkages, inclusions of LNA, and the like that are identifiable by a skilled person. In some embodiments, at least 50% of the internucleoside linkages in the overhang of the sensor nucleic acid strand are phosphorothioate internucleoside linkages. For example, at least 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or a number or a range between any two values, of the internucleoside linkages in the overhang of the sensor nucleic acid strand are phosphorothioate internucleoside linkages. In some embodiments, all internucleoside linkages in the overhang of the sensor nucleic acid strand are phosphorothioate internucleoside linkages.

[0153] In some embodiments, the 5' terminus and/or the 3' terminus of the sensor nucleic acid strand can comprise a terminal moiety. Any suitable terminal moiety described herein can be used. In some embodiments, the terminal moiety can include a tri- or hexaethylene glycol spacer, a C3 spacer, an inverted dT, an amine linker, a ligand (e.g. a targeting ligand), a fluorophore, an exonuclease, a fatty acid, a Cy3, an inverted dT attached to a triethylene glycol, or a combination thereof.

[0154] In an exemplary embodiment, a sensor strand is fully chemically modified and comprises 31 nucleotides (see, for example, FIG. 2). The first 22 nucleotides from the 5' end of the sensor strand are fully base-paired with the 3' and 5' regions of a core strand to form a sensor duplex and the last 9 nucleotides from the 5' end are single-stranded to form a sensor toehold (overhang). The sensor toehold at the 3' end comprises phosphorothioate backbone linkages between adjacent nucleotides to reduce nuclease activity and to increase protein

binding for self-delivery. To further reduce nuclease activity, the first two bases from the 5' end of the sensor strand are also connected via phosphorothioate backbone linkages. A ligand, such as C16 palmitic acid, can be attached to the sensor strand to improve self-delivery. The ligand can be attached to any nucleotide on the sensor strand. The 5' end of the sensor strand can also be modified with a tri-ethylene glycol spacer to improve resistance to nuclease activity.

[0155] The sequence of the sensor nucleic acid strand can be designed to sense an input nucleic acid strand or a portion thereof. For example, from the sequence of an input biomarker, a list of all possible sensor segments which are antisense to the input strand can be generated. The sensor sequences for uniqueness in the transcriptome of the target animal can be ranked using NCBI BLAST. For human cancer cell lines, sequences can be checked against human transcript and genomic collection using the BLASTn algorithm. In some embodiments, sensor segments that have more than 17 bases of sequence complementarity and complete overhang complementarity to known or predicted RNA transcripts may be eliminated. Examples of design features of the sensor nucleic acid strand are disclosed in, for example, WO 2020/033938, the contents of which are incorporated herein by reference.

Input Nucleic Acid Strand

[0156] The input nucleic acid strand described herein acts as a trigger to activate (switch on) the RNA interfering activity of the nucleic acid complex (e.g. RNAi duplex) upon binding to a sequence of the sensor nucleic acid in the nucleic acid complex.

[0157] The input nucleic acid strand comprises a sequence complementary to a sequence in the sensor nucleic acid of the nucleic acid complex. The complementary binding between the input nucleic acid strand and the sensor nucleic acid strand (e.g. an overhang) causes displacement of the sensor nucleic acid strand from the core nucleic acid strand, thereby activating the RNA interfering activity of the RNAi duplex formed by the passenger nucleic acid strand and the central region of the core nucleic acid strand.

[0158] The input nucleic acid strand can be cellular RNA transcripts that are present at relatively high expression levels in a set of target cells (e.g. cancer cells) and at a relatively low level of expression in a set of non-target cells (e.g. normal cells). In some embodiments, the nucleic acid complex herein described is activated (switched on) in target cells. While in the non-target cells, the nucleic acid complex remains inactivated (switched off).

[0159] In the target cells, the input nucleic acid strand can be expressed at a level of, about, at least, or at least about 2-fold, 5-fold, 10-fold, 20-fold, 30-fold, 40-fold, 50-fold, 60-fold, 70-fold, 80-fold, 90-fold, or 100-fold higher than in the non-target cells.

[0160] In the target cells, the input nucleic acid strand can be expressed at a level of, about, at least, at least about 50, 100, 200, 300, 400, 500, 600, 700, 800, 900, 1000 transcripts. In some embodiments, in the non-target cells, the input nucleic acid strand is expressed at a level of less than 50, less than 40, less than 30, less than 20, or less than 10 transcripts. Preferably, the non-target cells have no detectable expression of the input nucleic acid strand.

[0161] The input nucleic acid strand can comprise an mRNA, an miRNA, or a non-coding RNA such as a long non-coding RNA, an RNA fragment, or an RNA transcript of a virus. In some embodiments, the input nucleic acid strand is an RNA transcript that is expressed in a set of cells that are causing the progression of a disease and are therefore targeted for RNAi therapy. The non-target cells are usually a set of cells where silencing of a target RNA can cause side effects that are not beneficial for therapy. For treating a disease or a condition where the input RNA is overexpressed in target cells, the nucleic acid complex can be designed such that the sensor nucleic acid strand comprises a sequence complementary to the input RNA sequence. Upon administration of the nucleic acid complex, the binding of sensor nucleic acid strand to the input RNA induces the dissociation of the RNAi duplex from the sensor duplex in target cells thereby to activate the RNAi targeting the disease or condition.

[0162] In some embodiments, the input nucleic acid strand comprises a biomarker. The term "biomarker" refers to a nucleic acid sequence (DNA or RNA) that is an indicator of a disease or disorder, a susceptibility to a disease or disorder, and/or of response to therapeutic or other intervention. A biomarker can reflect an expression, function or regulation of a gene. The input nucleic acid strand can comprise any disease biomarker known in the art.

[0163] The input nucleic acid strand can be a mRNA, including a cell type or cell state specific mRNA. Examples of a cell type or cell-state specific mRNA include, but are not limited to, C3, GFAP, NPPA, CSF1R, SLC1A2, PLP1, and MBP mRNA. In some embodiments, the input nucleic acid is a microRNA (also known as miRNA), including but is not limited to, hsa-mir-23a-3p, hsa-mir-124-3p, and hsa-mir-29b-3p. In some embodiments, the input nucleic acid strand is a non-coding RNA, for example MALAT1 (metastasis associated lung adenocarcinoma transcript 1, also known as NEAT2 (noncoding nuclear-enriched abundant transcript 2)).

Target RNA

[0164] The central region of the core nucleic acid strand comprises a sequence complementary to a target RNA in order to direct target-specific RNA interference. In some embodiments, the target RNA is a cellular RNA transcript. The target RNA can be an mRNA, an miRNA, a non-coding RNA, a viral RNA transcript, or a combination thereof.

[0165] As used herein, a “target RNA” refers to a RNA whose expression is to be selectively inhibited or silenced through RNA interference. A target RNA can be a target gene comprising any cellular gene or gene fragment whose expression or activity is associated with a disease, a disorder or a condition. A target RNA can also be a foreign or exogenous RNA or RNA fragment whose expression or activity is associated with a disease, a disorder or a certain condition (e.g. a viral RNA transcript or a pro-viral gene).

[0166] In some embodiments, the target RNA can comprise an oncogene, a cytokinin gene, an idioype protein gene (Id protein gene), a prion gene, a gene that expresses a protein that induces angiogenesis, an adhesion molecule, a cell surface receptor, a gene of a protein involved in a metastasizing and/or invasive process, a gene of a proteinase, a gene of a protein that regulates apoptosis and the cell cycle, a gene that expresses the EGF receptor, a multi-drug resistance 1 gene (MDR1), a gene of a human papilloma virus, a hepatitis C virus, or a human immunodeficiency virus, a gene involved in cardiac hypertrophy, or a fragment thereof.

[0167] The target RNA can comprise a gene encoding for a protein involved in apoptosis. Exemplary target RNA genes include, but are not limited to, bcl-2, p53, caspases, cytotoxic cytokines such as TNF- α or Fas ligand, and a number of other genes known in the art as capable of mediating apoptosis. The target RNA can comprise a gene involved in cell growth. Exemplary target RNA genes include, but not limited to, oncogenes (e.g., genes encoding for ABLI, BCLI, BCL2, BCL6, CBFA2, CBL, CSFIR, ERBA, ERBB, EBRB2, ETSI, ETSI, ETV6, FGR, FOS, FYN, HCR, HRAS, JUN, KRAS, LCK, LYN, MDM2, MLL, MYB, MYC, MYCLI, MYCN, NRAS, PIM I, PML, RET, SRC, TALI, TCL3, and YES), as well as genes encoding for tumor suppressor proteins (e.g., APC, BRCA1, BRCA2, MADH4, MCC, NF I, NF2, RB I, TP53, and WTI). The target RNA can comprise a human major histocompatibility complex (MHC) gene or a fragment thereof. Exemplary MHC genes include MHC class I genes such as genes in the HLA-A, HLA-B or HLA-C subregions for class I cc chain genes, or β 2-microglobulin and MHC class II genes such as any of the genes of the DP, DQ and DR subregions of class II α chain and β chain genes (i.e. DP α , DP β , DQ α , DQ β , DR α , and DR β).

[0168] In some embodiments, the target RNA can comprise a gene encoding for a pathogen-associated protein. Pathogen associated protein include, but are not limited to, a viral protein involved in immunosuppression of the host, replication of the pathogen, transmission of the pathogen, or maintenance of the infection, or a host protein which facilitates entry of the pathogen into the host, drug metabolism by the pathogen or host, replication or integration of the pathogen's genome, establishment or spread of infection in the host, or assembly of the next generation of pathogen. In some embodiments, the pathogen can be a virus, such as a herpesvirus (e.g., herpes simplex, varicella-zoster virus, Epstein-Barr virus, cytomegalovirus

(CMV)), hepatitis C, HIV, JC virus), a bacteria or a yeast.

[0169] The target RNA comprises a gene associated with a disease or a condition of the central nervous system (CNS). Exemplary genes associated with a CNS disease or a condition include, but are not limited to, APP, MAPT, SOD1, BACE1, CASP3, TGM2, NFE2L3, TARDBP, ADRB1, CAMK2A, CBLN1, CDK5R1, GABRA1, MAPK10, NOS1, NPTX2, NRG1, NTS, PDCD2, PDE4D, PENK, SYT1, TTR, FUS, LRDD, CYBA, ATF3, ATF6, CASP2, CASP1, CASP7, CASP8, CASP9, HRK, C1QBP, BNIP3, MAPK8, MAPK14, Rac1, GSK3B, P2RX7, TRPM2, PARG, CD38, STEAP4, BMP2, GJA1, TYROBP, CTGF, ANXA2, RHOA, DUOX1, RTP801, RTP801L, NOX4, NOX1, NOX2 (gp91pho, CYBB), NOX5, DUOX2, NOXO1, NOXO2 (p47phox, NCF1), NOXA1, NOXA2 (p67phox, NCF2), p53 (TP53), HTRA2, KEAP1, SHC1, ZNHIT1, LGALS3, HI95, SOX9, ASPP1, ASPP2, CTSD, CAPNS1, FAS and FASLG, NOGO and NOGO-R; TLR1, TLR2, TLR3, TLR4, TLR6, TLR7, TLR8, TLR9, IL1bR, MYD88, TICAM, TIRAP, and HSP47.

Pharmaceutical compositions and methods of administration

Compositions

[0170] Also provided herein include pharmaceutical compositions comprising the nucleic acid complex as herein described, in combination with one or more compatible and pharmaceutically acceptable carriers.

[0171] The nucleic acid complex herein described can be suitably formulated and introduced into cell environment by any means that allows for a sufficient portion of the constructs to enter the cells to induce gene silencing, if it occurs.

[0172] The nucleic acid complex can be admixed, encapsulated, conjugated, or associated with other molecules, molecule structures, mixtures of compounds or agent, or other formulations for assistance in uptake, distribution, and/or absorption during delivery.

[0173] The phrase “pharmaceutically acceptable” is employed herein to refer to those agents, materials, compositions, and/or dosage forms which are, within the scope of sound medical judgment, suitable for use in contact with the tissues of human beings and animals without excessive toxicity, irritation, allergic response, or other problem or complication, commensurate with a reasonable benefit/risk ratio.

[0174] The phrase “pharmaceutically acceptable carrier” as used herein means a pharmaceutically-acceptable material, composition or vehicle, such as a liquid or solid filler, diluent, excipient, solvent or encapsulating material, involved in carrying or transporting the subject chemical from one organ, or portion of the body, to another organ, or portion of the body. Each carrier must be “acceptable” in the sense of being compatible with the other

ingredients of the formulation and not injurious to the subject. Some examples of materials which can serve as pharmaceutically-acceptable carriers include: (1) sugars, such as lactose, glucose and sucrose; (2) starches, such as corn starch and potato starch; (3) cellulose, and its derivatives, such as sodium carboxymethyl cellulose, ethyl cellulose and cellulose acetate; (4) powdered tragacanth; (5) malt; (6) gelatin; (7) talc; (8) excipients, such as cocoa butter and suppository waxes; (9) oils, such as peanut oil, cottonseed oil, safflower oil, sesame oil, olive oil, corn oil and soybean oil; (10) glycols, such as propylene glycol; (11) polyols, such as glycerin, sorbitol, mannitol and polyethylene glycol; (12) esters, such as ethyl oleate and ethyl laurate; (13) agar; (14) buffering agents, such as magnesium hydroxide and aluminum hydroxide; (15) alginic acid; (16) pyrogen-free water; (17) isotonic saline; (18) Ringer's solution; (19) ethyl alcohol; (20) phosphate buffer solutions; and (21) other non-toxic compatible substances employed in pharmaceutical formulations.

[0175] In some embodiments, pharmaceutically acceptable carrier comprise a pharmaceutical acceptable salt. As used herein, a “pharmaceutical acceptable salt” includes a salt of an acid form of one of the components of the compositions herein described. These include organic or inorganic acid salts of the amines. Preferred acid salts are the hydrochlorides, acetates, salicylates, nitrates and phosphates. Other suitable pharmaceutically acceptable salts are well known to those skilled in the art and include basic salts of a variety of inorganic and organic acids.

[0176] In some embodiments, pharmaceutically acceptable salts to be used with the nucleic acid complex herein described include but are not limited to (1) salts formed with cations such as sodium, potassium, ammonium, magnesium, calcium, polyamines such as spermine and spermidine; (2) acid addition salts formed with inorganic acids, for example hydrochloric acid, hydrobromic acid, sulfuric acid, phosphoric acid, nitric acid and the like; (3) salts formed with organic acids such as, for example, acetic acid, oxalic acid, tartaric acid, succinic acid, maleic acid, fumaric acid, gluconic acid, citric acid, malic acid, ascorbic acid, benzoic acid, tannic acid, palmitic acid, alginic acid, polyglutamic acid, naphthalenesulfonic acid, methanesulfonic acid, p-toluenesulfonic acid, naphthalene disulfonic acid, polygalacturonic acid, and the like; and (4) salts formed from elemental anions such as chlorine, bromine, and iodine.

Delivery vesicles

[0177] Various delivery systems can be employed for delivering the nucleic acid complex herein described such as antibody conjugates, micelles, natural polysaccharides, peptides, synthetic cationic polymers, microparticles, lipid-based nanovectors among others.

[0178] Delivery systems and the related excipients used for delivery of the nucleic acid complex herein described can vary in different embodiments. Delivery systems can be selected based on the mode of administration utilized, types of formulations, target sites, and types of diseases or disorders to be treated to facilitate tissue penetration, cellular uptake and to prevent extravasation and endosomal escape.

[0179] In some embodiments, the nucleic acid complex can be formulated with one or more polymers to form a supramolecular complex containing the nucleic acid complex and a multi-dimensional polymer network. The polymer can be linear or branched. The supramolecular complex can take any suitable form, and preferably, is in the form of particles.

[0180] The nucleic acid complex can be delivered via a lipid-mediated delivery system. In some embodiments, the nucleic acid complex can be encapsulated or associated with liposomes. For example, the nucleic acid complex can be condensed with a polycationic condensing agent, suspended in a low-ionic strength aqueous medium and cationic liposomes formed of a cationic vesicle-forming lipid.

[0181] As used herein, the term “liposomes” refers to lipid vesicles having an outer lipid shell, typically formed on one or more lipid bilayers, encapsulating an aqueous interior. In some embodiments, the liposomes are cationic liposomes composed of between about 20-80 mole percent of a cationic vesicle-forming lipid, with the remaining neutral vesicle-forming lipids and/or other components. As used herein, “vesicle-forming lipid” refers to any amphipathic lipid having hydrophobic and polar head group moieties and which by itself can form spontaneously into bilayer vesicles in water (e.g. phospholipids). A preferred vesicle-forming lipid is a diacyl-chain lipid, such as a phospholipid, whose acyl chains are typically between about 14-22 carbon atoms in length, and have varying degrees of unsaturation.

[0182] A cationic vesicle-forming lipid is a vesicle-forming lipid whose polar head group with a net positive charge, at the operational pH, e.g., pH 4-9. Examples include phospholipids (e.g., phosphatidylethanolamine), glycolipids (e.g., cerebrosides and gangliosides having a cationic polar head-group), cholesterol amine and related cationic sterols (e.g., 1,2-diolelyloxy-3-(trimethylammonio) propane (DOTAP), N-[1-(2,3,-ditetradecyloxy)propyl]-N,N-dimethyl-N-hydroxyethylammonium bromide (DMRIE), N-[1-(2,3,-dioleyloxy)propyl]-N,N-dimethyl-N-hydroxy ethylammonium bromide (DORIE), N-[1-(2,3-dioleyloxy)propyl]-N,N,N-trimethylammonium chloride (DOTMA), 3β [N—(N',N'-dimethylaminoethane) carbamoyl] cholesterol (DC-Choi), and dimethyldioctadecylammonium (DDAB)).

[0183] A neutral vesicle-forming lipid is a vesicle-forming lipid having no net charge or including a small percentage of lipids having a negative charge in the polar head group. Examples of vesicle-forming lipids include phospholipids, such as phosphatidylcholine (PC),

phosphatidyl ethanolamine (PE), phosphatidylinositol (PI), and sphingomyelin (SM), and cholesterol, cholesterol derivatives, and other uncharged sterols.

[0184] In some embodiments, the delivery systems used herein include, but are not limited to, nanoparticles (NPs), inorganic nanoparticles (e.g. silica NPs, gold NPs, Qdots, superparamagnetic iron oxide NPs, paramagnetic lanthanide ions) and other nanomaterials, nucleic acid lipid particles, polymeric nanoparticles, lipidoid nanoparticles (LNPs), chitosan and inulin nanoparticles, cyclodextrins nanoparticles, carbon nanotubes, liposomes, micellar structures, capsids, polymers (e.g. polyethylenimine, anionic polymers), polymer matrices, hydrogels, dendrimers (e.g. poly-propylenimine (PPI) and poly-amidoamine (PAMAM)), nucleic acid nanostructure, exosomes, and GalNAc-conjugated melittin-like peptides (NAG-MLPs). In some embodiments, the nucleic acid complex can be formulated in buffer solutions such as phosphate buffered saline solutions.

[0185] In some embodiments, the nucleic acid complex herein described is delivered via lipidoid nanoparticles (LNPs). LNPs can comprise ionizable LNPs, cationic LNPs, and/or neutral LNPs. Ionizable LNPs are nearly uncharged during circulation but become protonated in a low pH environment, e.g., in the endosomes and lysosomes. Cationic LNPs exhibit a constitutive positive charge in blood circulation and in endosomes or lysosomes. Neutral LNPs are neutral, uncharged during circulation and in endosomes or lysosomes.

[0186] The nucleic acid complex herein described can be provided naked or conjugated to a ligand. Naked siRNA refer to a system that contains no delivery system that is associated with the siRNA either covalently or noncovalently. When delivered in naked form, the naked siRNAs can be locally injected to a target site such as specific organs that are relatively closed off and contain few nucleases (e.g. eye).

[0187] In some embodiments, the nucleic acid complex herein described can be attached to (e.g. fused or conjugated) a ligand to form ligand-siRNA conjugates that can transport siRNA to desired tissues and cells by specific recognition and interactions between the ligand and the surface receptor of the cells or tissues. Common targeting ligands include carbohydrate, aptamers, antibodies or antibody fragments, peptides (e.g., cell-penetrating peptides, endosomolytic peptides), and small molecules (e.g., N-Acetylgalactosamine (GalNAc)), and others as will be apparent to a skilled artisan.

[0188] The nucleic acid complex can be conjugated to an aptamer. The term “aptamers” as used here refers to oligonucleotide or peptide molecules that bind a specific target with high affinity and specificity. In particular, nucleic acid aptamers can comprise, for example, nucleic acid species that have been engineered through repeated rounds of in vitro selection or equivalently, SELEX (systematic evolution of ligands by exponential enrichment) to bind to

various molecular targets such as small molecules, proteins, nucleic acids, and even cells, tissues and organisms. Peptide aptamers are peptides that are designed to specifically bind to and interfere with protein-protein interactions inside cells. In particular, peptide aptamers can be derived, for example, according to a selection strategy that is derived from the yeast two-hybrid (Y2H) system. Aptamers are useful in biotechnological and therapeutic applications as they offer molecular recognition properties that rival that of the antibodies.

[0189] In some embodiments, the nucleic acid complex is conjugated to a small molecule. The term “small molecule” as used herein indicates an organic compound that is of synthetic or biological origin and that, although may include monomers and/or primary metabolites, is not a polymer. In some embodiments, small molecules can comprise molecules that are not protein or nucleic acids, which play a biological role that is endogenous (e.g., inhibition or activation of a target) or exogenous (e.g., cell signaling), which are used as a tool in molecular biology, or which are suitable as drugs in medicine. Small molecules can also have no relationship to natural biological molecules. Typically, small molecules have a molar mass lower than 1 kg/mol. Exemplary small molecules include secondary metabolites (e.g., actinomycin-D), certain antiviral drugs (such as amantadine and rimantadine), teratogens and carcinogens (such as phorbol 12-myristate 13-acetate), natural products (such as penicillin, morphine and paclitaxel) and additional molecules identifiable by a skilled artisan. In some embodiments, the nucleic acid complex herein described is conjugated to GalNAc.

[0190] Examples of ligands suitable for use in targeting the nucleic acid complex to specific cell types include, but are not limited to, folate capable of binding to folate receptor of epithelial carcinomas and bone marrow stem cells, water soluble vitamins capable of binding to vitamin receptors of various cells, pyridoxyl phosphate capable of binding to CD4⁺ lymphocytes, apolipoproteins capable of binding to LDL of liver hepatocytes and vascular endothelial cells, insulin capable of binding to insulin receptor, transferrin capable of binding to transferrin receptor of endothelial cells, galactose capable of binding to asialoglycoprotein receptor of liver hepatocytes, sialyl-Lewis_x capable of binding to E, P selectin of activated endothelial cells, Mac-1 capable of binding to L selectin of neutrophils and leukocytes, VEGF capable of binding to Flk-1,2 of tumor epithelial cells, basic FGF capable of binding to FGF receptor of tumor epithelial cells, EFG capable of binding to EFG receptor of epithelial cells, VCAM-1 capable of binding to $\alpha_4\beta_1$ integrin of vascular endothelial cells, ICAM-1 capable of binding to $\alpha_L\beta_2$ integrin of vascular endothelial cells, PECAM-1/CD31 capable of binding to $\alpha_v\beta_3$ integrin of vascular endothelial cells and activated platelets, osteopontin capable of binding to $\alpha_v\beta_1$ integrin and $\alpha_v\beta_5$ integrin of endothelial cells and smooth muscle cells in atherosclerotic plaques, RGD sequences capable of binding to $\alpha_v\beta_3$ integrin of tumor endothelial cells and

vascular smooth muscle cells, or HIV GP 120/41 or GP120 capable of binding to CD4 or CD4⁺ lymphocytes, and others identifiable to a skilled artisan.

[0191] In some embodiments, the delivery of the nucleic acid complex herein described is such that at least 30%, 40%, 50%, 60%, 70%, 80%, 90% or more of the target cells incorporate the nucleic acid complex. In some embodiments, about 0.1-10 nM nucleic acid complex is delivered to the target cells.

Formulations

[0192] Any suitable pharmaceutical formulations can be employed. In some embodiments, the pharmaceutical compositions of the present disclosure may be specially formulated for administration in solid or liquid form, including those adapted for the following: (1) oral administration, for example, drenches (aqueous or non-aqueous solutions or suspensions), tablets, boluses, powders, granules, pastes; (2) parenteral administration, for example, by subcutaneous, intramuscular or intravenous injection as, for example, a sterile solution or suspension; (3) topical application, for example, as a cream, ointment or spray applied to the skin; (4) intravaginally or intrarectally, for example, as a pessary, cream or foam; or (5) aerosol, for example, as an aqueous aerosol, liposomal preparation or solid particles containing the hydrogel composition. The pharmaceutical compositions can comprise one or more pharmaceutically-acceptable carriers.

[0193] Formulations useful in the methods of the present disclosure include those suitable for oral, nasal, topical (including buccal and sublingual), rectal, vaginal, aerosol and/or parenteral administration. The formulations may conveniently be presented in unit dosage form and may be prepared by any methods well known in the art of pharmacy. The amount of active ingredient which can be combined with a carrier material to produce a single dosage form will vary depending upon the host being treated, the particular mode of administration. The amount of active ingredient, which can be combined with a carrier material to produce a single dosage form will generally be that amount of the RNAi constructs which produces a therapeutic effect. Generally, out of one hundred percent, this amount will range from about 1% to about 99% of active ingredient, preferably from about 5% to about 70%, most preferably from about 10% to about 30%.

[0194] Formulations suitable for oral administration may be in the form of capsules, cachets, pills, tablets, lozenges (using a flavored basis, usually sucrose and acacia or tragacanth), powders, granules, or as a solution or a suspension in an aqueous or non-aqueous liquid, or as an oil-in-water or water-in-oil liquid emulsion, or as an elixir or syrup, or as pastilles (using an inert base, such as gelatin and glycerin, or sucrose and acacia) and/or as mouth washes and the like, each containing a predetermined amount of a respiration uncoupling agent as an active

ingredient. A nucleic acid complex composition may also be administered as a bolus, electuary or paste.

[0195] In solid dosage forms for oral administration (capsules, tablets, pills, dragees, powders, granules and the like), the active ingredient is mixed with one or more pharmaceutically-acceptable carriers, such as sodium citrate or dicalcium phosphate, and/or any of the following: (1) fillers or extenders, such as starches, lactose, sucrose, glucose, mannitol, and/or silicic acid; (2) binders, such as, for example, carboxymethylcellulose, alginates, gelatin, polyvinyl pyrrolidone, sucrose and/or acacia; (3) humectants, such as glycerol; (4) disintegrating agents, such as agar-agar, calcium carbonate, potato or tapioca starch, alginic acid, certain silicates, and sodium carbonate; (5) solution retarding agents, such as paraffin; (6) absorption accelerators, such as quaternary ammonium compounds; (7) wetting agents, such as, for example, acetyl alcohol and glycerol monostearate; (8) absorbents, such as kaolin and bentonite clay; (9) lubricants, such as talc, calcium stearate, magnesium stearate, solid polyethylene glycols, sodium lauryl sulfate, and mixtures thereof; and (10) coloring agents. In the case of capsules, tablets and pills, the pharmaceutical compositions may also comprise buffering agents. Solid compositions of a similar type may also be employed as fillers in soft and hard-filled gelatin capsules using such excipients as lactose or milk sugars, as well as high molecular weight polyethylene glycols and the like.

[0196] A tablet may be made by compression or molding, optionally with one or more accessory ingredients. Compressed tablets may be prepared using binder (for example, gelatin or hydroxypropylmethyl cellulose), lubricant, inert diluent, preservative, disintegrant (for example, sodium starch glycolate or cross-linked sodium carboxymethyl cellulose), surface-active or dispersing agent. Molded tablets may be made by molding in a suitable machine a mixture of the powdered peptide or peptidomimetic moistened with an inert liquid diluent.

[0197] Actual dosage levels of the active ingredients in the pharmaceutical compositions of the present disclosure may be determined by the methods of the present invention so as to obtain an amount of the active ingredient, which is effective to achieve the desired therapeutic response for a particular subject, composition, and mode of administration, without being toxic to the subject.

[0198] Tablets, and other solid dosage forms, such as dragees, capsules, pills and granules, may optionally be scored or prepared with coatings and shells, such as enteric coatings and other coatings well known in the pharmaceutical-formulating art. They may also be formulated so as to provide slow or controlled release of the active ingredient therein using, for example, hydroxypropylmethyl cellulose in varying proportions to provide the desired release profile, other polymer matrices, liposomes and/or microspheres. They may be sterilized by, for

example, filtration through a bacteria-retaining filter, or by incorporating sterilizing agents in the form of sterile solid compositions, which can be dissolved in sterile water, or some other sterile injectable medium immediately before use. These compositions may also optionally contain opacifying agents and may be of a composition that they release the active ingredient(s) only, or preferentially, in a certain portion of the gastrointestinal tract, optionally, in a delayed manner. Examples of embedding compositions, which can be used include polymeric substances and waxes. The active ingredient can also be in micro-encapsulated form, if appropriate, with one or more of the above-described excipients.

[0199] Liquid dosage forms for oral administration include pharmaceutically acceptable emulsions, microemulsions, solutions, suspensions, syrups and elixirs. In addition to the active ingredient, the liquid dosage forms may contain inert diluents commonly used in the art, such as, for example, water or other solvents, solubilizing agents and emulsifiers, such as ethyl alcohol, isopropyl alcohol, ethyl carbonate, ethyl acetate, benzyl alcohol, benzyl benzoate, propylene glycol, 1,3-butylene glycol, oils (in particular, cottonseed, groundnut, corn, germ, olive, castor and sesame oils), glycerol, tetrahydrofuryl alcohol, polyethylene glycols and fatty acid esters of sorbitan, and mixtures thereof.

[0200] Besides inert diluents, the oral compositions can also include adjuvants such as wetting agents, emulsifying and suspending agents, sweetening, flavoring, coloring, perfuming and preservative agents.

[0201] Suspensions, in addition to the active agent may contain suspending agents as, for example, ethoxylated isostearyl alcohols, polyoxyethylene sorbitol and sorbitan esters, microcrystalline cellulose, aluminum metahydroxide, bentonite, agar-agar and tragacanth, and mixtures thereof.

[0202] Formulations for rectal or vaginal administration may be presented as a suppository, which may be prepared by mixing one or more respiration uncoupling agents with one or more suitable nonirritating excipients or carriers comprising, for example, cocoa butter, polyethylene glycol, a suppository wax or a salicylate, and which is solid at room temperature, but liquid at body temperature and, therefore, will melt in the rectum or vaginal cavity and release the active agent.

[0203] Formulations which are suitable for vaginal administration also include pessaries, tampons, creams, gels, pastes, foams or spray formulations containing such carriers as are known in the art to be appropriate.

[0204] Dosage forms for the topical or transdermal administration of hydrogel compositions include powders, sprays, ointments, pastes, creams, lotions, gels, solutions, patches and inhalants. The active component may be mixed under sterile conditions with a

pharmaceutically-acceptable carrier, and with any preservatives, buffers, or propellants which may be required.

[0205] The ointments, pastes, creams and gels may contain, in addition to a respiration uncoupling agent, excipients, such as animal and vegetable fats, oils, waxes, paraffins, starch, tragacanth, cellulose derivatives, polyethylene glycols, silicones, bentonites, silicic acid, talc and zinc oxide, or mixtures thereof.

[0206] Ophthalmic formulations, eye ointments, powders, solutions (e.g. eye drops) and the like, are also contemplated as being within the scope of the present disclosure.

[0207] Examples of suitable aqueous and nonaqueous carriers which may be employed in the pharmaceutical compositions of the invention include water, ethanol, polyols (such as glycerol, propylene glycol, polyethylene glycol, and the like), and suitable mixtures thereof, vegetable oils, such as olive oil, and injectable organic esters, such as ethyl oleate. Proper fluidity can be maintained, for example, by the use of coating materials, such as lecithin, by the maintenance of the required particle size in the case of dispersions, and by the use of surfactants.

[0208] These compositions may also contain adjuvants such as preservatives, wetting agents, emulsifying agents and dispersing agents. Prevention of the action of microorganisms may be ensured by the inclusion of various antibacterial and antifungal agents, for example, paraben, chlorobutanol, phenol sorbic acid, and the like. It may also be desirable to include isotonic agents, such as sugars, sodium chloride, and the like into the compositions. In addition, prolonged absorption of the injectable pharmaceutical form may be brought about by the inclusion of agents which delay absorption such as aluminum monostearate and gelatin.

[0209] The pharmaceutical compositions herein described comprise a therapeutically-effective amount of the nucleic acid complexes.

[0210] The phrase “therapeutically-effective amount” as used herein means that amount of nucleic acid complex disclosed herein which is effective for producing some desired therapeutic effect, e.g., cancer treatment, at a reasonable benefit/risk ratio. The therapeutically-effective amount also varies depending on the structure of the constructs, the route of administration utilized, the target sites, and the specific diseases or disorders to be treated as will be understood to a person skilled in the art. For example, if a given clinical treatment is considered effective when there is at least a 20% reduction in a measurable parameter associated with a disease or disorder, a therapeutically-effective amount of the constructs for the treatment of that disease or disorder is the amount necessary to achieve at least a 20% reduction in that measurable parameter.

[0211] In some embodiments, the pharmaceutical composition herein described comprises the nucleic acid complex in a suitable dosage sufficient to inhibit expression of the target gene in a subject (e.g. animal or human) being treated. In some embodiments, a suitable dosage of the nucleic acid complex is in the range of 0.001 to 0.25 milligrams per kilogram body weight of the subject per day, or in the range of 0.01 to 20 micrograms per kilogram body weight per day, or in the range of 0.01 to 10 micrograms per kilogram body weight per day, or in the range of 0.10 to 5 micrograms per kilogram body weight per day, or in the range of 0.1 to 2.5 micrograms per kilogram body weight per day. The pharmaceutical compositions comprising the nucleic acid complex can be administered once daily, twice daily, three times daily or as needed or prescribed by a physician. The pharmaceutical composition herein described can also be provided in dosage units comprising two, three, four, five, six or more sub-doses administered at appropriate intervals throughout the day. The dosage unit can also be compounded for a single dose (e.g. using sustained or controlled release formulation) which can be sustainably released over several days in a controlled manner.

[0212] As will be apparent to a skilled person, a suitable dosage unit of the pharmaceutical composition herein described can be estimated from data obtained from cell culture assays and further determined from data obtained in animal studies. For example, toxicity and therapeutic efficacy of the pharmaceutical compositions described herein can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., for determining the LD50 (the dose lethal to 50% of the population) and the ED50 (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD50/ED50. Compositions that exhibit large therapeutic indices are preferred. Suitable dosages of the compositions in combination with particular delivery systems can be selected in order to minimize toxicity, such as to minimize potential damage to untargeted cells and to reduce side effects.

Administration

[0213] As will be apparent to a skilled artisan, the nucleic acid complexes herein described and compositions thereof can be administered to a subject using any suitable administration routes. The nucleic acid complexes and compositions thereof can be administered to a target site locally or systematically.

[0214] The wording “local administration” or “topic administration” as used herein indicates any route of administration by which a composition is brought in contact with the body of the individual, so that the resulting composition location in the body is topic (limited to a specific tissue, organ or other body part where the imaging is desired). Exemplary local administration routes include injection into a particular tissue by a needle, gavage into the

gastrointestinal tract, and spreading a solution containing hydrogel composition on a skin surface.

[0215] The wording “systemic administration” as used herein indicates any route of administration by which a nucleic acid complex composition is brought in contact with the body of the individual, so that the resulting composition location in the body is systemic (i.e. non limited to a specific tissue, organ or other body part where the imaging is desired). Systemic administration includes enteral and parenteral administration. Enteral administration is a systemic route of administration where the substance is given via the digestive tract, and includes but is not limited to oral administration, administration by gastric feeding tube, administration by duodenal feeding tube, gastrostomy, enteral nutrition, and rectal administration. Parenteral administration is a systemic route of administration where the substance is given by route other than the digestive tract and includes but is not limited to intravenous administration, intra-arterial administration, intramuscular administration, subcutaneous administration, intradermal administration, intraperitoneal administration, and intravesical infusion.

[0216] In some embodiments, the methods of administration can comprise aerosol delivery, nasal delivery, vaginal delivery, rectal delivery, buccal delivery, ocular delivery, local delivery, topical delivery, intracisternal delivery, intraperitoneal delivery, oral delivery, intramuscular injection, intravenous (IV) injection, subcutaneous (SC) injection, intranodal injection, intratumoral injection, intraperitoneal injection, and/or intradermal injection, or any combination thereof. The administration can also be site-specific injection (e.g. in the eye or the cerebral spinal fluid).

[0217] In some embodiments, the administration can be Ex vivo transduction, cell injection, subcutaneous injection, intravenous injection, intrathecal delivery, intracerebroventricular injection, intradermal injection, intravitreal delivery, intratumoral delivery, or topical application (e.g. topical eye drop).

[0218] The methods of administration depends on the target site, the type of cells/tissues to be targeted at, and how the constructs are formulated. In some embodiments, lipid formulations can be administered to animals such as by intravenous, intramuscular, or intraperitoneal injection, or orally or by inhalation or other methods as known in the art.

[0219] In some embodiments, the administration can be SC injection into the adipose tissue below the epidermis and dermis. In some embodiments, SC administration can be associated with ligand-conjugated nucleic acid complex herein described. In some embodiments, SC administration can render a slower release rate of the drugs into the systemic circulation and an entering into the lymphatic system, giving more time for recycling of cellular receptors that

mediate uptake. In some embodiments, SC administration can be faster and easier to administer, reducing treatment burden.

[0220] IV administration can, for example, be associated with nanoparticle and lipid nanoparticle formulated nucleic acid complex herein described. In some embodiments, IV administration can avoid first-pass metabolism in the liver and affords quick access to target tissue through the systemic circulation.

Target sites

[0221] The compositions herein described can be administered to any suitable target site. Target sites can be *in vitro*, *in vivo* or *ex vivo*. Exemplary target sites can include cells grown in an *in vitro* culture, including, primary mammalian, cells, immortalized cell lines, tumor cells, stem cells, and the like. Additional exemplary target sites include cells, tissues and organs in an *ex vivo* culture and cells, tissues, organs, or organs systems *in vivo* in a subject, for example, lungs, brain, kidney, liver, heart, the central nervous system, the peripheral nervous system, the gastrointestinal system, the circulatory system, the immune system, the skeletal system, the sensory system, within a body of an individual and additional environments identifiable by a skilled person.

[0222] The target site can comprise a site of disease or disorder or can be proximate to a site of a disease or disorder. The location of the one or more sites of a disease or disorder can be predetermined. The location of the one or more sites of a disease or disorder can be determined during the method (e.g., by an imaging-based method such as ultrasound or MRI). The target site can comprise a tissue, such as, for example, adrenal gland tissue, appendix tissue, bladder tissue, bone, bowel tissue, brain tissue, breast tissue, bronchi, coronal tissue, ear tissue, esophagus tissue, eye tissue, gall bladder tissue, genital tissue, heart tissue, hypothalamus tissue, kidney tissue, large intestine tissue, intestinal tissue, larynx tissue, liver tissue, lung tissue, lymph nodes, mouth tissue, nose tissue, pancreatic tissue, parathyroid gland tissue, pituitary gland tissue, prostate tissue, rectal tissue, salivary gland tissue, skeletal muscle tissue, skin tissue, small intestine tissue, spinal cord, spleen tissue, stomach tissue, thymus gland tissue, trachea tissue, thyroid tissue, ureter tissue, urethra tissue, soft and connective tissue, peritoneal tissue, blood vessel tissue and/or fat tissue. The tissue can be inflamed tissue. The tissue can comprise (i) grade I, grade II, grade III or grade IV cancerous tissue; (ii) metastatic cancerous tissue; (iii) mixed grade cancerous tissue; (iv) a sub-grade cancerous tissue; (v) healthy or normal tissue; and/or (vi) cancerous or abnormal tissue. In some embodiments, upon administration, the nucleic acid complex and a composition thereof accumulates in vasculature of cancerous tissue. In some embodiments, the target site can comprise a solid tumor.

[0223] Target sites where the nucleic acid complex or compositions thereof can be administered can vary in different embodiments depending on the mode of administration utilized and the types of diseases or disorders to be treated. In some embodiments, the target sites can be related to ocular tissues, respiratory system, muscle, liver, central nerve system, solid tumors, hematopoietic system, skin, eye, placenta, bone, or other target sites in an individual as will be apparent to a skilled artisan.

[0224] The term “individual” or “subject” or “patient” as used herein in the context of imaging includes an animal and in particular higher animals and in particular vertebrates such as mammals and more particularly human beings.

[0225] In some embodiments, the ratio of the concentration of the nucleic acid complex at the subject’s target site to the concentration of the nucleic acid complex outside the target site (e.g. in subject’s blood circulation, serum, or plasma) can vary. In some embodiments, the ratio of the concentration of the nucleic acid complex at the subject’s target site to the concentration of the nucleic acid complex outside the target site (e.g. in subject’s blood circulation, serum, or plasma) can be, or be about, be at least, be at least about, be at most, or be at most about, 1:1, 1.1:1, 1.2:1, 1.3:1, 1.4:1, 1.5:1, 1.6:1, 1.7:1, 1.8:1, 1.9:1, 2:1, 2.5:1, 3:1, 4:1, 5:1, 6:1, 7:1, 8:1, 9:1, 10:1, 11:1, 12:1, 13:1, 14:1, 15:1, 16:1, 17:1, 18:1, 19:1, 20:1, 21:1, 22:1, 23:1, 24:1, 25:1, 26:1, 27:1, 28:1, 29:1, 30:1, 31:1, 32:1, 33:1, 34:1, 35:1, 36:1, 37:1, 38:1, 39:1, 40:1, 41:1, 42:1, 43:1, 44:1, 45:1, 46:1, 47:1, 48:1, 49:1, 50:1, 51:1, 52:1, 53:1, 54:1, 55:1, 56:1, 57:1, 58:1, 59:1, 60:1, 61:1, 62:1, 63:1, 64:1, 65:1, 66:1, 67:1, 68:1, 69:1, 70:1, 71:1, 72:1, 73:1, 74:1, 75:1, 76:1, 77:1, 78:1, 79:1, 80:1, 81:1, 82:1, 83:1, 84:1, 85:1, 86:1, 87:1, 88:1, 89:1, 90:1, 91:1, 92:1, 93:1, 94:1, 95:1, 96:1, 97:1, 98:1, 99:1, 100:1, 200:1, 300:1, 400:1, 500:1, 600:1, 700:1, 800:1, 900:1, 1000:1, 2000:1, 3000:1, 4000:1, 5000:1, 6000:1, 7000:1, 8000:1, 9000:1, 10000:1, or a number or a range between any two of the values.

[0226] The target site can comprise target cells. The target cells can be tumor cells (e.g. solid tumor cells). In some embodiments, the administration of the nucleic acid complex and/or compositions herein described to a target site of the subject results in the death of at least about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 95%, about 100%, or a number or a range between any two of these values, of the target cells. The ratio of target cell death to non-target cell death after administration of the nucleic acid complex and/or compositions can be at least about 2:1. In some embodiments, the ratio of target cell death to non-target cell death after administration of the nucleic acid complex and/or compositions can be, or be about, or be at least, or be at least about, or be at most, or be at most about, 1:1, 1.1:1, 1.2:1, 1.3:1, 1.4:1, 1.5:1, 1.6:1, 1.7:1, 1.8:1,

1.9:1, 2:1, 2.5:1, 3:1, 4:1, 5:1, 6:1, 7:1, 8:1, 9:1, 10:1, 11:1, 12:1, 13:1, 14:1, 15:1, 16:1, 17:1, 18:1, 19:1, 20:1, 21:1, 22:1, 23:1, 24:1, 25:1, 26:1, 27:1, 28:1, 29:1, 30:1, 31:1, 32:1, 33:1, 34:1, 35:1, 36:1, 37:1, 38:1, 39:1, 40:1, 41:1, 42:1, 43:1, 44:1, 45:1, 46:1, 47:1, 48:1, 49:1, 50:1, 51:1, 52:1, 53:1, 54:1, 55:1, 56:1, 57:1, 58:1, 59:1, 60:1, 61:1, 62:1, 63:1, 64:1, 65:1, 66:1, 67:1, 68:1, 69:1, 70:1, 71:1, 72:1, 73:1, 74:1, 75:1, 76:1, 77:1, 78:1, 79:1, 80:1, 81:1, 82:1, 83:1, 84:1, 85:1, 86:1, 87:1, 88:1, 89:1, 90:1, 91:1, 92:1, 93:1, 94:1, 95:1, 96:1, 97:1, 98:1, 99:1, 100:1, 200:1, 300:1, 400:1, 500:1, 600:1, 700:1, 800:1, 900:1, 1000:1, 2000:1, 3000:1, 4000:1, 5000:1, 6000:1, 7000:1, 8000:1, 9000:1, 10000:1, or a number or a range between any two of the values.

Methods of Modulating a Target RNA

[0227] Also provided herein is a method of modulating a target RNA using the nucleic acid complex or a composition thereof herein described. The method can comprise contacting a cell comprising a target RNA with the nucleic acid complex herein describe. Upon detection of an input nucleic acid strand, an input strand can bind to the overhang of the sensor nucleic acid strand to cause displacement of the sensor nucleic acid strand from the core nucleic acid strand to release the sequence complementary to the target RNA into the cell, thereby modulating the target RNA.

[0228] Contacting the cells with the nucleic acid complex can be performed with cells *in vitro*, *in vivo* or *ex vivo*. For example, the cells can be cells grown in an *in vitro* culture, including, primary mammalian, cells, immortalized cell lines, tumor cells, stem cells, and the like. The cells can comprise cells, tissues and organs in an *ex vivo* culture and cells, tissues, organs, or organs systems *in vivo* in a subject, for example, lungs, brain, kidney, liver, heart, the central nervous system, the peripheral nervous system, the gastrointestinal system, the circulatory system, the immune system, the skeletal system, the sensory system, within a body of an individual and additional environments identifiable by a skilled person. The cell can be a disease cell or a cell of disorder. The cell can be a cancer cell. Contacting the cell with the nucleic acid complex can occur can also occur *in vitro*, *ex vivo*, or *in vivo* e.g., in the body of a subject.

Methods of Treating a Disease or Disorder

[0229] Also provided herein is a method of treating a disease or a condition using the nucleic acid complex or a composition thereof herein described. The method can comprise administering the nucleic acid complex described herein to a subject in need thereof. Upon detection of an input nucleic acid strand, the input nucleic acid strand can bind to the overhang of the sensor nucleic acid strand to cause displacement of the sensor nucleic acid strand from the core nucleic acid strand to release the sequence complementary to a target RNA, thereby

reducing the activity of the target RNA or protein expression from the target RNA in the subject to treat the disease or condition.

[0230] The term “condition” as used herein indicates a physical status of the body of an individual (as a whole or as one or more of its parts), that does not conform to a standard physical status associated with a state of complete physical, mental and social well-being for the individual. Conditions herein described include but are not limited disorders and diseases wherein the term “disorder” indicates a condition of the living individual that is associated to a functional abnormality of the body or of any of its parts, and the term “disease” indicates a condition of the living individual that impairs normal functioning of the body or of any of its parts and is typically manifested by distinguishing signs and symptoms.

[0231] As used herein, the term “treatment” “treat” refers to an intervention made in response to a disease, disorder or physiological condition manifested by a patient. The aim of treatment may include, but is not limited to, one or more of the alleviation or prevention of symptoms, slowing or stopping the progression or worsening of a disease, disorder, or condition and the remission of the disease, disorder or condition. The term “treat” and “treatment” includes, for example, therapeutic treatments, prophylactic treatments, and applications in which one reduces the risk that a subject will develop a disorder or other risk factor. Treatment does not require the complete curing of a disorder and encompasses embodiments in which one reduces symptoms or underlying risk factors. In some embodiments, “treatment” refers to both therapeutic treatment and prophylactic or preventative measures. Those in need of treatment include those already affected by a disease or disorder or undesired physiological condition as well as those in which the disease or disorder or undesired physiological condition is to be prevented. As used herein, the term “prevention” refers to any activity that reduces the burden of the individual later expressing those symptoms. This can take place at primary, secondary and/or tertiary prevention levels, wherein: a) primary prevention avoids the development of symptoms/disorder/condition; b) secondary prevention activities are aimed at early stages of the condition/disorder/symptom treatment, thereby increasing opportunities for interventions to prevent progression of the condition/disorder/symptom and emergence of symptoms; and c) tertiary prevention reduces the negative impact of an already established condition/disorder/symptom by, for example, restoring function and/or reducing any condition/disorder/symptom or related complications. The term “prevent” does not require the 100% elimination of the possibility of an event. Rather, it denotes that the likelihood of the occurrence of the event has been reduced in the presence of the compound or method.

[0232] Various diseases and disorders can be treated with the nucleic acid complex compositions provided herein. Diseases and disorders disclosed herein include, but are not

limited to, HIV infection with lymphoma, hemophilia A, hemophilia B, hypercholesterolemia, atherosclerotic cardiovascular disease, renal impairment, chronic hepatitis B, acute intermittent porphyria, atypical hemolytic uraemic syndrome, primary hyperoxaluria, hereditary transthyretin amyloidosis (hATTR), α 1-antitrypsin deficiency liver disease, hepatitis B, sickle cell disease, primary hyperoxaluria, ewing sarcoma, advanced gynecological cancer, stage III/IV ovarian cancer, pancreatic cancer, advanced solid tumors, hepatocellular carcinoma/liver cancer, lymphoma and leukemias, heart disease, heart failure, keloids, hypertrophic cicatrix, relapsed or refractory B cell lymphoma, hypertrophic scar, age-related macular degeneration, retinal scarring, cardia surgery, cardiac hypertrophy, non-arteritic anterior ischaemic optic neuropathy, alport syndrome, HIV infections/AIDS, pancreatic ductal adenocarcinoma/pancreatic cancer, dry eye disease, and various solid tumors.

[0233] In some embodiments, the disease or disorder can be a cancer. The cancer can be a solid tumor, a liquid tumor, or a combination thereof. The nucleic acid complex herein described or a composition thereof can be administered to the cells, tissues and/or organs comprising a tumor using any suitable administration route. For example, the nucleic acid complex or a composition thereof can be administered to the cells, tissues and/or organs comprising a tumor via subcutaneous injection or intratumoral delivery.

[0234] The cancer can be selected from the group consisting of colon cancer, rectal cancer, renal-cell carcinoma, liver cancer, non-small cell carcinoma of the lung, cancer of the small intestine, cancer of the esophagus, melanoma, bone cancer, pancreatic cancer, skin cancer, cancer of the head or neck, cutaneous or intraocular malignant melanoma, uterine cancer, ovarian cancer, rectal cancer, cancer of the anal region, stomach cancer, testicular cancer, uterine cancer, carcinoma of the fallopian tubes, carcinoma of the endometrium, carcinoma of the cervix, carcinoma of the vagina, carcinoma of the vulva, Hodgkin's Disease, non-Hodgkin lymphoma, cancer of the endocrine system, cancer of the thyroid gland, cancer of the parathyroid gland, cancer of the adrenal gland, sarcoma of soft tissue, cancer of the urethra, cancer of the penis, solid tumors of childhood, cancer of the bladder, cancer of the kidney or ureter, carcinoma of the renal pelvis, neoplasm of the central nervous system (CNS), primary CNS lymphoma, tumor angiogenesis, spinal axis tumor, brain stem glioma, pituitary adenoma, Kaposi's sarcoma, epidermoid cancer, squamous cell cancer, T-cell lymphoma, environmentally induced cancers, combinations of said cancers, and metastatic lesions of said cancers.

[0235] The cancer can be a hematologic cancer chosen from one or more of chronic lymphocytic leukemia (CLL), acute leukemias, acute lymphoid leukemia (ALL), B-cell acute lymphoid leukemia (B-ALL), T-cell acute lymphoid leukemia (T-ALL), chronic myelogenous leukemia (CML), B cell prolymphocytic leukemia, blastic plasmacytoid dendritic cell neoplasm,

Burkitt's lymphoma, diffuse large B cell lymphoma, follicular lymphoma, hairy cell leukemia, small cell- or a large cell-follicular lymphoma, malignant lymphoproliferative conditions, MALT lymphoma, mantle cell lymphoma, marginal zone lymphoma, multiple myeloma, myelodysplasia and myelodysplastic syndrome, non-Hodgkin's lymphoma, Hodgkin's lymphoma, plasmablastic lymphoma, plasmacytoid dendritic cell neoplasm, Waldenstrom macroglobulinemia, or pre-leukemia.

[0236] Non-limiting examples of cancers that can be prevented and/or treated using the nucleic acid complexes and compositions disclosed herein include: renal cancer; kidney cancer; glioblastoma multiforme; metastatic breast cancer; breast carcinoma; breast sarcoma; neurofibroma; neurofibromatosis; pediatric tumors; neuroblastoma; malignant melanoma; carcinomas of the epidermis; leukemias such as but not limited to, acute leukemia, acute lymphocytic leukemia, acute myelocytic leukemias such as myeloblastic, promyelocytic, myelomonocytic, monocytic, erythroleukemia leukemias and myelodysplastic syndrome, chronic leukemias such as but not limited to, chronic myelocytic (granulocytic) leukemia, chronic lymphocytic leukemia, hairy cell leukemia; polycythemia vera; lymphomas such as but not limited to Hodgkin's disease, non-Hodgkin's disease; multiple myelomas such as but not limited to smoldering multiple myeloma, nonsecretory myeloma, osteosclerotic myeloma, plasma cell leukemia, solitary plasmacytoma and extramedullary plasmacytoma; Waldenstrom's macroglobulinemia; monoclonal gammopathy of undetermined significance; benign monoclonal gammopathy; heavy chain disease; bone cancer and connective tissue sarcomas such as but not limited to bone sarcoma, myeloma bone disease, multiple myeloma, cholesteatoma-induced bone osteosarcoma, Paget's disease of bone, osteosarcoma, chondrosarcoma, Ewing's sarcoma, malignant giant cell tumor, fibrosarcoma of bone, chordoma, periosteal sarcoma, soft-tissue sarcomas, angiosarcoma (hemangiosarcoma), fibrosarcoma, Kaposi's sarcoma, leiomyosarcoma, liposarcoma, lymphangioma sarcoma, neurilemmoma, rhabdomyosarcoma, and synovial sarcoma; brain tumors such as but not limited to, glioma, astrocytoma, brain stem glioma, ependymoma, oligodendroglioma, nonglial tumor, acoustic neurinoma, craniopharyngioma, medulloblastoma, meningioma, pineocytoma, pineoblastoma, and primary brain lymphoma; breast cancer including but not limited to adenocarcinoma, lobular (small cell) carcinoma, intraductal carcinoma, medullary breast cancer, mucinous breast cancer, tubular breast cancer, papillary breast cancer, Paget's disease (including juvenile Paget's disease) and inflammatory breast cancer; adrenal cancer such as but not limited to pheochromocytoma and adrenocortical carcinoma; thyroid cancer such as but not limited to papillary or follicular thyroid cancer, medullary thyroid cancer and anaplastic thyroid cancer; pancreatic cancer such as but not limited to, insulinoma, gastrinoma, glucagonoma, vipoma, somatostatin-secreting tumor, and carcinoid

or islet cell tumor; pituitary cancers such as but limited to Cushing's disease, prolactin-secreting tumor, acromegaly, and diabetes insipidus; eye cancers such as but not limited to ocular melanoma such as iris melanoma, choroidal melanoma, and ciliary body melanoma, and retinoblastoma; vaginal cancers such as squamous cell carcinoma, adenocarcinoma, and melanoma; vulvar cancer such as squamous cell carcinoma, melanoma, adenocarcinoma, basal cell carcinoma, sarcoma, and Paget's disease; cervical cancers such as but not limited to, squamous cell carcinoma, and adenocarcinoma; uterine cancers such as but not limited to endometrial carcinoma and uterine sarcoma; ovarian cancers such as but not limited to, ovarian epithelial carcinoma, borderline tumor, germ cell tumor, and stromal tumor; cervical carcinoma; esophageal cancers such as but not limited to, squamous cancer, adenocarcinoma, adenoid cystic carcinoma, mucoepidermoid carcinoma, adenosquamous carcinoma, sarcoma, melanoma, plasmacytoma, verrucous carcinoma, and oat cell (small cell) carcinoma; stomach cancers such as but not limited to, adenocarcinoma, fungating (polypoid), ulcerating, superficial spreading, diffusely spreading, malignant lymphoma, liposarcoma, fibrosarcoma, and carcinosarcoma; colon cancers; colorectal cancer, KRAS mutated colorectal cancer; colon carcinoma; rectal cancers; liver cancers such as but not limited to hepatocellular carcinoma and hepatoblastoma, gallbladder cancers such as adenocarcinoma; cholangiocarcinomas such as but not limited to papillary, nodular, and diffuse; lung cancers such as KRAS-mutated non-small cell lung cancer, non-small cell lung cancer, squamous cell carcinoma (epidermoid carcinoma), adenocarcinoma, large-cell carcinoma and small-cell lung cancer; lung carcinoma; testicular cancers such as but not limited to germinal tumor, seminoma, anaplastic, classic (typical), spermatocytic, nonseminoma, embryonal carcinoma, teratoma carcinoma, choriocarcinoma (yolk-sac tumor), prostate cancers such as but not limited to, androgen-independent prostate cancer, androgen-dependent prostate cancer, adenocarcinoma, leiomyosarcoma, and rhabdomyosarcoma; penile cancers; oral cancers such as but not limited to squamous cell carcinoma; basal cancers; salivary gland cancers such as but not limited to adenocarcinoma, mucoepidermoid carcinoma, and adenoidcystic carcinoma; pharynx cancers such as but not limited to squamous cell cancer, and verrucous; skin cancers such as but not limited to, basal cell carcinoma, squamous cell carcinoma and melanoma, superficial spreading melanoma, nodular melanoma, lentigo malignant melanoma, acral lentiginous melanoma; kidney cancers such as but not limited to renal cell cancer, adenocarcinoma, hypernephroma, fibrosarcoma, transitional cell cancer (renal pelvis and/or ureter); renal carcinoma; Wilms' tumor; and bladder cancers such as but not limited to transitional cell carcinoma, squamous cell cancer, adenocarcinoma, carcinosarcoma. In some embodiments, the cancer is myxosarcoma, osteogenic sarcoma, endotheliosarcoma, lymphangi endotheliosarcoma, mesothelioma, synovioma, hemangioblastoma, epithelial

carcinoma, cystadenocarcinoma, bronchogenic carcinoma, sweat gland carcinoma, sebaceous gland carcinoma, papillary carcinoma, or papillary adenocarcinomas.

[0237] In some embodiments, the disease or disorder can be a central nervous system (CNS) disease or condition. The nucleic acid complex herein described or a composition thereof can be administered to the cells, tissues and/or organs of the CNS using any suitable administration route. For example, the nucleic acid complex or a composition thereof can be administered to the cells, tissues and/or organs of the CNS of a subject via intrathecal injection, intracerebroventricular injection, or intracerebral injection to penetrate the blood-brain barrier. In some embodiments, the cell(s), tissue(s), and/or organ(s) of the CNS comprises damaged or inflamed cell(s), tissue(s), or organ(s). In some embodiments, the cells(s), tissue(s), and/or organ(s) of the CNS comprise the brain, the white matter, the gray matter, the brainstem, the cerebellum, the diencephalon, the cerebrum, the spinal cord, the cranial nerve, cell(s) of any of the preceding, tissue(s) of any of the preceding, or a combination thereof.

[0238] In some embodiments, the CNS disease is a movement disorder, a memory disorder, addiction, attention deficit/hyperactivity disorder (ADHD), autism, bipolar disorder, depression, encephalitis, epilepsy/seizure, migraine, multiple sclerosis, a neurodegenerative disorder, a psychiatric disease, a neuroinflammatory disease, Alzheimer's disease, Huntington's disease, Parkinson's disease, Tourette syndrome, dystonia, or a combination thereof. In some embodiments, the disease is a neuroinflammatory disease. For example, the neuroinflammatory disease is Parkinson's disease, Alzheimer's disease, multiple sclerosis, or a combination thereof.

Kits

[0239] Also provided herein include kits comprising one or more compositions described herein, in suitable packaging such as in a container, pack, or dispenser, and may further comprise written material that can include instructions for use, discussion of clinical studies, listing of side effects, and the like. Such kits can also include information, such as scientific literature references, package insert materials, clinical trial results, and/or summaries of these and the like, which indicate or establish the activities and/or advantages of the composition, and/or which describe dosing, administration, side effects, drug interactions, or other information useful to the health care provider. Such information can be based on the results of various studies, for example, studies using experimental animals involving in vivo models and studies based on human clinical trials. A kit can comprise one or more unit doses described herein. The compositions can be in the form of kits of parts. In a kit of parts, one or more components of the compositions disclosed herein are provided independent of one another (e.g., constructs, excipients, and/or diluents are provided as separate compositions) and are then

employed (e.g., by a user) to generate the compositions.

EXAMPLES

[0240] Some aspects of the embodiments discussed above are disclosed in further detail in the following examples, which are not in any way intended to limit the scope of the present disclosure.

Example 1

RNAi activity with or without a C3 linker

[0241] This example demonstrates the RNAi activity of various siRNA domain variants with or without a C3 linker as the 5' and the 3' connector.

[0242] The passenger and core strands of the new construct are assembled to form the siRNA domains of the new construct. The different variants of these siRNA domains are tested for RNAi activity.

[0243] To test the constructs, CASi siRNA segments were assembled by thermally annealing passenger and core strands in 1x phosphate buffer saline. The RNAi activities of the CASi siRNA segments were measured using dual luciferase assays. CASi siRNA segments were co-transfected into HCT 116 cells with dual luciferase vectors carrying the Huntingtin gene siRNA target sequence, using lipofectamine 2000. After 48 hours, cells were lysed and assayed for knockdown of the target gene by comparing the luminescence value of Renilla luciferase that carries the target sequence to Firefly luciferase that was used as a reference control. Methods and procedures of assembling CASi siRNA, cell transfection, and dual luciferase assays can be found in, for example, international application WO 2020/033938, the content of which is incorporated herein by reference in its entirety.

[0244] **FIG. 5A** and **FIG. 5B** show sequence diagrams of two exemplary nucleic acid complex constructs whose RNAi activities are determined in this example. Top nucleic acid complex construct comprises a core strand v3c1 base-paired to a passenger strand v3p1, in which a C3 linker is used as the 5' and the 3' connector. Bottom nucleic acid complex construct comprises a core strand v3c5 base-paired to the same passenger strand, in which no C3 linker is used as the 5' and the 3' connector. Instead, v3c5 core strand has a 3' mU connector and no connector at the 5' end.

[0245] **FIG. 6** show sequence diagrams of two positive control nucleic acid complex constructs (the passenger strand shown comprises the siRNA targeting the huntingtin gene (HTT)) used in the assay described in this example.

[0246] **FIG. 7** shows various siRNA variants with different passenger strand (V3P1, V3P2, V3P3, V3P5, V3P5, V3P6, V3P7, V3P8, and V3P9) assembled with v3c1 core strand

shown in **FIG. 5** and tested in this example. The v3c1 core strand has a C3 linker as the 5' and the 3' connector. The target protein expression was tested with the siRNA variants at three different concentrations: 10 nM, 1 nM, and 0.1 nM. **FIG. 8** shows a graphic representation of the target protein expression data for the siRNA variants shown in **FIG. 7**. Higher RNAi activity is suggested by lower expression of the target protein.

[0247] **FIG. 9** shows different siRNA variants with different passenger strand (V3P1, V3P2, V3P3, V3P5, V3P5, V3P6, V3P7, V3P8, and V3P9) assembled with a v3c5 core strand shown in **FIG. 5** and tested in this example. The v3c5 core strand does not have a C3 linker as the 5' and the 3' connector. Instead, v3c5 core strand has a 3' mU connector and no connector at the 5' end. The target protein expression was tested with the siRNA variants at three different concentrations: 10nM, 1.0nM, and 0.1 nM. **FIG. 10** shows a graphic representation of the target protein expression data for the siRNA variants shown in **FIG. 9**. Similar to **FIGS. 7-8**, higher RNAi activity is suggested by lower expression of the target protein.

[0248] These data indicate that a C3 linker as the 5' connector inhibits RNAi activity of the siRNA domain. A comparison of the target protein expression data among different passenger variants (V3P1, V3P2, V3P3, V3P5, V3P5, V3P6, V3P7, V3P8, and V3P9) indicates that extensive modification of the passenger strand with LNAs (e.g. HTT V3P8) can decrease RNAi activity.

Example 2

RNAi activity with different 5' and 3' connectors

[0249] In this example, different versions of the core strand were tested with the same sensor (Mir23 Sensor 1) and passenger strands (Passenger strand 1) to investigate the effects of different 5' and 3' connectors on the RNAi activity. RNAi activity was also evaluated between two-stranded constructs and three-stranded constructs.

[0250] Two-stranded constructs consist of the passenger strand base-paired to the core strand, forming an active siRNA domain. Three-stranded constructs consist of all three strands: the passenger strand, the core strand, and the sensor strand.

[0251] CASi siRNA segments (two-stranded constructs) and three-stranded constructs were assembled by thermally annealing passenger and core strands, or passenger, core and sensor strands in 1x phosphate buffer saline.

[0252] CASi siRNA segments or three-stranded constructs were co-transfected into HCT 116 cells at different concentrations (e.g., 0.1 nM, 1.0 nM and 10 nM) with dual luciferase vectors carrying the Huntingtin gene siRNA target sequence, using lipofectamine 2000. After 48 hours, cells were lysed and assayed for knockdown of the target gene by comparing the

luminescence value of Renilla luciferase that carries the target sequence to Firefly luciferase that was used as a reference control. Examples of methods and procedures of assembling CASi siRNA, cell transfection, and dual luciferase assays are described in, for example, international application WO 2020/033938.

[0253] FIG. 11A and FIG. 11B show sequence diagrams of various nucleic acid complexes disclosed herein each having the same passenger strand (Passenger strand 1) and the sensor strand (Mir23 Sensor 1) but a different core strand (Core strand v3c1, Core strand v3c2, Core strand v3c3, Core strand v3c4, Core strand v3c5, and Core strand v3c6), and particularly, a different 5' and 3' connector in the core strand. For example, CASi 1 has both a 5' C3 connector and a 3' C3 connector. CASi 2 has a 5' C3 connector and a 2-O-methylated U as the 3' linker. CASi 3 has a 2-O-methylated A as the 5' linker and a 3' C3 connector. CASi 4 has a 2-O-methylated A as the 5' linker and a 2-O-methylated U as the 3' linker. CASi 5 has a phosphodiester backbone linkage as the 5' linker and a 2-O-methylated U as the 3' linker. CASi 6 has a phosphodiester backbone linkage as the 5' linker and a phosphorothioate linkage as the 3' linker.

[0254] The sequences illustrated in FIGS. 11A and 11B are also provided in Table 1 below.

Table 1. Sequences of exemplary CASi strands.	
mir23 sensor	
mir23 Sensor 1	/5Sp9/*mC*+G*mA.+A.mG.mA.+A.mC.mG.+G.mA.+A.mA.mU.+C.mC.mC.+T.mG.mG.+C.mA*mA*+T*mG*mU*+G*+A*+T*/3CholTEG/ (SEQ ID NO: 1)
Passenger strand	
HTT V3P1	+T*+T*mA.+T.mA.mU.mC.mA.fG.mU.fA.fA.fA.mG.mA.mG.mA.mU.+T*mA*mA (SEQ ID NO: 2)
Core strands	
CASi 1: V3C1	mU.mC.mC.mG.mU.mU.mC.mU.mU.mC.mG./iSpC3/.mU*fU*mA.mA.mU.fC.mU.mC.mU.mU.mU.fA.mC.fU.mG.mA.mU.mA.mU.mA.mA./iSpC3/.mU*mG.mC.mC.mA.mG.mG.mG.mA.mU.mU (SEQ ID NO: 3, SEQ ID NO: 4, and SEQ ID NO: 5 separated by iSpC3)
CASi 2: V3C2	mU.mC.mC.mG.mU.mU.mC.mU.mU.mC.mG./iSpC3/.mU*fU*mA.mA.mU.fC.mU.mC.mU.mU.mU.fA.mC.fU.mG.mA.mU.mA.mU.mA.mA*mU*mU.mG.mC.mC.mA.mG.mG.mG.mA.mU.mU (SEQ ID NO: 6 and SEQ ID NO: 7 separated by iSpC3)
CASi 3: V3C3	mU.mC.mC.mG.mU.mU.mC.mU.mU.mC.mG.mA.mU*fU*mA.mA.mU.fC.mU.mC.mU.mU.mU.fA.mC.fU.mG.mA.mU.mA.mU.mA.mA./iSpC3/.mU*mG.mC.mC.mA.mG.mG.mG.mA.mU.mU (SEQ ID NO: 8 and SEQ ID NO: 9 separated by iSpC3)
CASi 4: V3C4	mU.mC.mC.mG.mU.mU.mC.mU.mU.mC.mG.mA.mU*fU*mA.mA.mU.fC.mU.mC.mU.mU.mU.fA.mC.fU.mG.mA.mU.mA.mU.mA.mA*mU*mU.mG.mC.mC.mA.mG.mG.mG.mA.mU.mU (SEQ ID NO: 10)

CASi 5: V3C5	mU.mC.mC.mG.mU.mU.mC.mU.mU.mC.mG.mU*fU*mA.mA.mU.fC.mU.mC.mU.mU.mU.fA.mC.fU.mG.mA.mU.mA.mU.mA.mA*mU*mU.mG.mC.mC.mA.mG.mG.mG.mA.mU.mU (SEQ ID NO: 11)
CASi 6: V3C6	mU.mC.mC.mG.mU.mU.mC.mU.mU.mC.mG.mU*fU*mA.mA.mU.fC.mU.mC.mU.mU.mU.fλ.mC.fU.mG.mλ.mU.mλ.mU.mλ.mλ*mU*mG.mC.mC.mA.mG.mG.mG.mA.mU.mU (SEQ ID NO: 12)
/Sp9/ = triethylene glycol spacer CholTEG = Cholesterol-TEG /iSpC3/= internal C3 spacer * = phosphorothioate backbone . = phosphodiester backbone mA, mG, mC, mU = 2'-O-methyl bases +A, +T, +C, +G = locked nucleic acid (LNA) bases fA, fU, fC, fG = 2'-fluoro bases NH2 = primary amine linker. rA, rU, rC, rG = RNA	

[0255] FIG. 12 shows non-denaturing polyacrylamide gel (PAGE) of various nucleic acid complex constructs, indicating all the complexes are assembled as desired. Lanes are as follows (from left to right): P1C1; P1C1S2; P1C2; P1C2S2; P1C3; P1C3S2; P1C4; P1C4S2; P1C5; P1C5S2; P1C6; P1C6S2; G1RC1; and G1RC1S2. P1 indicates the passenger strand 1.

[0256] FIG. 13 shows the RNAi activity of two-stranded assemblies each having the same passenger strand v3p1 and a different core strand (C1, C2, C3, C4, C5, and C6) at different concentrations. The sequences of the passenger strand and the core strand are shown in FIGS. 11A and 11B.

[0257] FIG. 14 shows the RNAi activity of three-stranded assemblies (CASi 1, CASi 2, CASi 3, CASi 4, CASi 5 and CASi 6) each having the same passenger strand v3p1, the same sensor strand (Mir23 sensor 1), and a different core strand (C1, C2, C3, C4, C5, and C6) in comparison to the two-stranded assemblies (siRNA 1: C1; siRNA 2: C2; siRNA 3: C3; siRNA 4: C4; siRNA 5: C5; siRNA 6: C6) from FIG. 13 at three different concentrations. The sequences of the passenger strand, the sensor strand, the core strand are shown in FIGS. 11A and 11B and Table 1.

[0258] These data indicate that assemblies, including two-stranded and three-stranded assemblies, with 5' mA connector and 3' C3 (3-carbon linker) connector (e.g., CASi 3 construct and siRNA 3 duplex) has the highest RNAi activity. The different RNAi activity between the siRNA 3 duplex and the CASi 3 construct also suggests good RNAi activity switching of CASi. Assemblies, including two-stranded and three-stranded assemblies, which do not have a 5' C3 connector (such as C3, C4, C5, C6) have a higher RNAi activity than assemblies having a 5' C3 connector (C1 and C2). Assemblies that do not have a 5' connector

(C5 and C6) have a lower RNAi activity than assemblies (C3 and C4) having a 5' connector (such as mA) but not a C3 linker. For the same core strand, the three-stranded assemblies are generally expected to have lower RNAi activity than two-stranded assemblies.

Example 3

RNAi activity of various RNA complex designs

[0259] In this example, experiments were carried out to compare the RNAi switching and RNAi activity of Design 1 shown in **FIG. 1**. and the RNA complex design disclosed herein (e.g., Design 2 shown in **FIG. 1**). V3C3a and V3C3b are the constructs in the form of Design 2. G1C1S1 is a construct in the form of the Design 1.

[0260] CASi siRNA segment (two-stranded constructs) and three-stranded constructs were assembled by thermally annealing passenger and core strands, or passenger, core and sensor strands in 1x phosphate buffer saline. The CASi siRNA segment (two-stranded constructs) and three-stranded constructs were co-transfected into HCT 116 cells using lipofectamine 2000. The HCT116 cells can express either an RNA biomarker that could activate the CASi sensor (e.g. NPPA gene sequence encoding atrial natriuretic peptide (ANP))(denoted as “Act” in **FIG. 16**) or a control nucleic acid strand that could not activate the CASi sensor (denoted as “Neg” in **FIG. 16**) using a short RNA transcript driven by a Pol III promoter. The HCT 116 cells also have a dual luciferase vector carrying the PPP3CA (calcineurin) gene siRNA target sequence. Calcineurin is a calcium and calmodulin dependent serine/threonine protein phosphatase, and has been identified as a key driver of cardiac hypertrophy. ANP has been used as diagnostic markers for cardiac hypertrophy. Therefore, the sensor strand of the three-stranded CASi siRNA constructs is designed to detect ANP mRNA while the siRNA domain (e.g. the passenger strand) is designed to inhibit calcineurin.

[0261] After 72 hours, cells were lysed and assayed for knockdown of the target gene (calcineurin) by comparing the luminescence value of Renilla luciferase (carrying the target sequence) to Firefly luciferase.

[0262] **FIG. 15** shows sequence diagrams of a nucleic acid complex including a core strand V3C3a (T2 CASi construct) in the form of Design 2 shown in **FIG. 1** and a nucleic acid complex (Cond-siRNA construct) in the form of Design 1 shown in **FIG. 1** (bottom: G1C1S1). The sequences of T2 CASi and Cond-siRNA strands are provided in Table 2.

Table 2: Sequences of T2 CASi of Design 2 and an Cond-siRNA of Design 1.	
Passenger for T2 ANP-calcineurin CASi	
Calc V3P3 passenger	/5Cy3/*mU*+A*mC.mA.mG.mG.fA.mA.fA.fA.fG.mC.mC.mA.mA.mA.mC.mA.mA*mC*mA (SEQ ID NO: 13)
Sensor strand for both T2 CASi and Cond-siRNA	
ANP	/5Sp9/.mC.+T.mU.mC.+A.mC.mC.+A.mC.+C.mU.mC.mU.+C.mA.m

Sensor 1	G.+T.mG.+G.mC.mA.+A.mU*mG*mC*+G*mA*+C*mC*+A*mA*/3TEGC hol/ (SEQ ID NO: 14)
Core strand for T2 ANP-calcineurin CASi	
Rat ANP V3C3a	mA.mG.mG.mU.mG.mG.mU.mG.mA.mA.mG.mA.mU*fG*mU.mU.mG.fU .mU.mU.mG.mG.mC.fU.mU.fU.mU.mC.mC.mU.mG.mU.mA*/iSpC3/ *mU*mU.mG.mC.mC.mA.mC.mU.mG.mA.mG (SEQ ID NO: 15 and SEQ ID NO: 16 separated by iSpC3)
PPP3CA guide strand for Cond-siRNA	
Calc G4	/5Cy3/.+C*+G.rA.rG.rU.rG.rU.rU.rG.rU.mU.rU.mG.mG.mC.r U.mU.rU.rU.rC.mC.mU.mG*mU*mU (SEQ ID NO: 17)
Core strand for Cond-siRNA	
ANP-Calc core strand	mA.mG.mG.mU.mG.mG.mU.mG.mA.mA.mG./iSpC3/.mC*+A*mG.rG. rA.rA.rA.rA.rG.rC.rC.rA.rA.rA.rC.rA.rA.rC.rA.rC.rU.rC *mG./iSpC3/.mA.mU.mU.mG.mC.mC.mA.mU.mU.mG.mA.mG (SEQ ID NO: 18, SEQ ID NO: 19 and SEQ ID NO: 20 separated by iSpC3)

[0263] FIG. 16 shows the RNAi activity of the modified two-stranded constructs (V3C3a siRNA) and three-stranded constructs (V3C3a and V3C3b) in comparison with the original two-stranded (G1C1 siRNA) and three-stranded constructs (G1C1S1) at three different concentrations.

[0264] These data indicate that the modified CASi constructs shows lower RNAi activity in the absence of the RNA biomarker (Neg) and higher RNAi activity in the presence of the RNAi biomarker (Act), thus indicating that the RNAi activity of the modified CASi constructs is switched OFF when the RNA biomarker is absent. The RNAi activity of the modified constructs (T2 CASi: V3C3a and V3C3b) was also significantly improved compared to the original design (Cond-siRNA construct G1C1S1). The modified CASi siRNA segments (two-stranded assemblies, e.g. V3C3a siRNA)) also show significantly improved RNAi activity compared to the original two-stranded design (G1C1 siRNA).

Terminology

[0265] In at least some of the previously described embodiments, one or more elements used in an embodiment can interchangeably be used in another embodiment unless such a replacement is not technically feasible. It will be appreciated by those skilled in the art that various other omissions, additions and modifications may be made to the methods and structures described above without departing from the scope of the claimed subject matter. All such modifications and changes are intended to fall within the scope of the subject matter, as defined by the appended claims.

[0266] With respect to the use of substantially any plural and/or singular terms herein, those having skill in the art can translate from the plural to the singular and/or from the

singular to the plural as is appropriate to the context and/or application. The various singular/plural permutations may be expressly set forth herein for sake of clarity. As used in this specification and the appended claims, the singular forms “a,” “an,” and “the” include plural references unless the context clearly dictates otherwise. Any reference to “or” herein is intended to encompass “and/or” unless otherwise stated.

[0267] It will be understood by those within the art that, in general, terms used herein, and especially in the appended claims (*e.g.*, bodies of the appended claims) are generally intended as “open” terms (*e.g.*, the term “including” should be interpreted as “including but not limited to,” the term “having” should be interpreted as “having at least,” the term “includes” should be interpreted as “includes but is not limited to,” etc.). It will be further understood by those within the art that if a specific number of an introduced claim recitation is intended, such an intent will be explicitly recited in the claim, and in the absence of such recitation no such intent is present. For example, as an aid to understanding, the following appended claims may contain usage of the introductory phrases “at least one” and “one or more” to introduce claim recitations. However, the use of such phrases should not be construed to imply that the introduction of a claim recitation by the indefinite articles “a” or “an” limits any particular claim containing such introduced claim recitation to embodiments containing only one such recitation, even when the same claim includes the introductory phrases “one or more” or “at least one” and indefinite articles such as “a” or “an” (*e.g.*, “a” and/or “an” should be interpreted to mean “at least one” or “one or more”); the same holds true for the use of definite articles used to introduce claim recitations. In addition, even if a specific number of an introduced claim recitation is explicitly recited, those skilled in the art will recognize that such recitation should be interpreted to mean at least the recited number (*e.g.*, the bare recitation of “two recitations,” without other modifiers, means at least two recitations, or two or more recitations). Furthermore, in those instances where a convention analogous to “at least one of A, B, and C, etc.” is used, in general such a construction is intended in the sense one having skill in the art would understand the convention (*e.g.*, “a system having at least one of A, B, and C” would include but not be limited to systems that have A alone, B alone, C alone, A and B together, A and C together, B and C together, and/or A, B, and C together, etc.). In those instances where a convention analogous to “at least one of A, B, or C, etc.” is used, in general such a construction is intended in the sense one having skill in the art would understand the convention (*e.g.*, “a system having at least one of A, B, or C” would include but not be limited to systems that have A alone, B alone, C alone, A and B together, A and C together, B and C together, and/or A, B, and C together, etc.). It will be further understood by those within the art that virtually any disjunctive word and/or phrase presenting two or more alternative terms, whether in the description, claims, or drawings, should

be understood to contemplate the possibilities of including one of the terms, either of the terms, or both terms. For example, the phrase “A or B” will be understood to include the possibilities of “A” or “B” or “A and B.”

[0268] In addition, where features or aspects of the disclosure are described in terms of Markush groups, those skilled in the art will recognize that the disclosure is also thereby described in terms of any individual member or subgroup of members of the Markush group.

[0269] As will be understood by one skilled in the art, for any and all purposes, such as in terms of providing a written description, all ranges disclosed herein also encompass any and all possible sub-ranges and combinations of sub-ranges thereof. Any listed range can be easily recognized as sufficiently describing and enabling the same range being broken down into at least equal halves, thirds, quarters, fifths, tenths, etc. As a non-limiting example, each range discussed herein can be readily broken down into a lower third, middle third and upper third, etc. As will also be understood by one skilled in the art all language such as “up to,” “at least,” “greater than,” “less than,” and the like include the number recited and refer to ranges which can be subsequently broken down into sub-ranges as discussed above. Finally, as will be understood by one skilled in the art, a range includes each individual member. Thus, for example, a group having 1-3 articles refers to groups having 1, 2, or 3 articles. Similarly, a group having 1-5 articles refers to groups having 1, 2, 3, 4, or 5 articles, and so forth.

[0270] While various aspects and embodiments have been disclosed herein, other aspects and embodiments will be apparent to those skilled in the art. The various aspects and embodiments disclosed herein are for purposes of illustration and are not intended to be limiting, with the true scope and spirit being indicated by the following claims.

WHAT IS CLAIMED IS:

1. A nucleic acid complex, comprising:
 - a first nucleic acid strand comprising 20-70 linked nucleosides;
 - a second nucleic acid strand binding to a central region of the first nucleic acid strand to form a first nucleic acid duplex; and
 - a third nucleic acid strand binding to a 5' region and a 3' region of the first nucleic acid strand to form a second nucleic acid duplex, wherein the third nucleic acid strand comprises a overhang, wherein the overhang is not complementary to the first nucleic acid strand and is capable of binding to an input nucleic acid strand to cause the displacement of the third nucleic acid strand from the first nucleic acid strand,
 - wherein the central region of the first nucleic acid strand comprises a sequence complementary to a target RNA, wherein the sequence is 10-35 nucleosides in length.
2. The nucleic acid complex of claim 1, wherein the sequence complementary to the target RNA is 10-21 nucleotides in length.
3. The nucleic acid complex of any one of claims 1-2, wherein the second nucleic acid strand binds to 19-25 linked nucleotides in the central region of the first nucleic acid strand to form the first nucleic acid duplex.
4. The nucleic acid complex of any one of claims 1-3, wherein the first nucleic acid duplex does not comprise a Dicer cleavage site.
5. The nucleic acid complex of any one of claims 1-4, wherein the nucleic acid complex does not comprise a Dicer cleavage site.
6. The nucleic acid complex of any one of claims 1-5, wherein the central region of the first nucleic acid strand is linked to the 5' region of the first nucleic acid strand via a 5' connector.
7. The nucleic acid complex of any one of claims 1-6, wherein the central region of the first nucleic acid strand is linked to the 3' region of the first nucleic acid strand via a 3' connector.
8. The nucleic acid complex of any one of claims 1-7, wherein the 5' connector, the 3' connector, or both comprise a C₃ 3-carbon linker, a nucleotide, a modified nucleotide, or a exonuclease cleavage-resistant moiety, or a combination thereof.
9. The nucleic acid complex of claim 8, wherein the modified nucleotide is a 2'-O-methyl nucleotide or a 2'-F nucleotide.
10. The nucleic acid complex of any one of claims 1-7, wherein the 5' connector comprises, or is, a C₃ 3-carbon linker, 2'-O-methyl nucleotide, 2'-F nucleotide, a nucleotide

with a phosphodiester 5' and 3' connection cleavable by an exonuclease when in a single stranded form, or a combination thereof.

11. The nucleic acid complex of claim 10, wherein the 3' connector is a C₃ 3-carbon linker.

12. The nucleic acid complex of any one of claims 1-7, wherein the 3' connector comprises a C₃ 3-carbon linker, a nucleotide, a modified nucleotide, an exonuclease cleavage-resistant moiety when in a single stranded form, or a combination thereof.

13. The nucleic acid complex of claim 12, wherein the 3' connector comprises, or is, a 2'-O-methyl nucleotide, and wherein the 2'-O-methyl nucleotide is optionally 2'-O-methyladenosine, 2'-O-methylguanosine, 2'-O-methyluridine, or 2'-O-methylcytidine.

14. The nucleic acid complex of any one of claims 1-13, wherein the second nucleic strand is fully complementary to the central region of the first nucleic acid strand, thereby forming blunt ends at the 5' and 3' termini of the second nucleic acid strand in the first nucleic acid duplex.

15. The nucleic acid complex of any one of claims 1-13, wherein the second nucleic acid strand does not have an overhang at 3' terminus, or 5' terminus, or both in the first nucleic acid duplex.

16. The nucleic acid complex of any one of claims 1-13, wherein the second nucleic acid strand has a 3' overhang, a 5' overhang, or both in the first nucleic acid duplex.

17. The nucleic acid complex of claim 16, wherein the second nucleic acid strand has an 3' overhang and the 3' overhang is one to five nucleosides in length.

18. The nucleic acid complex of any one of claims 1-17, wherein the 5' terminus of the central region of the first nucleic acid strand, the 3' terminus of the central region of the first nucleic acid strand, or both, comprises at least one phosphorothioate internucleoside linkage.

19. The nucleic acid complex of any one of claims 1-17, wherein each of the 5' terminus of the central region of the first nucleic acid strand and the 3' terminus of the central region of the first nucleic acid strand independently comprises one or more phosphorothioate internucleoside linkages.

20. The nucleic acid complex of any one of claims 1-17, wherein the central region of the first nucleic acid strand does not comprise phosphorothioate internucleoside linkages except for the internucleoside linkage(s) between two or three nucleosides at the 5' terminus, 3' terminus, or both, of the central region.

21. The nucleic acid complex of any one of claims 1-20, wherein at least 80%, at least 85%, at least 90%, or at least 95% of the nucleosides of one or more of (1) the central

region of the first nucleic acid strand, (2) the 5' region of the first nucleic strand, and (3) the 3' region of the first nucleic strand are chemically modified.

22. The nucleic acid complex of any one of claims 1-20, wherein at least 80%, at least 85%, at least 90%, or at least 95% of the nucleosides of one or more of the first nucleic acid strand, the second nucleic strand and the third nucleic strand are chemically modified.

23. The nucleic acid complex of any one of claims 1-22, wherein at least 80%, at least 85%, at least 90%, at least 95%, or all of the nucleosides of the nucleic acid complex are chemically modified.

24. The nucleic acid complex of any one of claims 20-23, wherein the chemical modifications are to resist nuclease degradation, to increase melting temperature (T_m), or both, of the nucleic acid complex.

25. The nucleic acid complex of any one of claims 1-24, wherein at least 90%, at least 95%, or all of the nucleotides of the nucleic acid complex are non-DNA and non-RNA nucleotides.

26. The nucleic acid complex of any one of claims 1-25, wherein at most 5%, at most 10%, or at most 15% of the nucleosides of the second nucleic strand are LNA.

27. The nucleic acid complex of any one of claims 1-26, wherein about 10%-50% of the bases have a 2'-4' bridging modifications.

28. The nucleic acid complex of any one of claims 1-27, wherein about 10%-50% of the bases are locked locked nucleic acid (LNA) or analogues thereof.

29. The nucleic acid complex of any one of claims 1-28, wherein about 10%-50% of the bases comprises 2'-O-methyl modification, 2'-F modification, or both.

30. The nucleic acid complex of any one of claims 1-29, wherein less than 5%, less than 10%, less than 25%, less than 50% of the internucleoside linkages in the first nucleic acid strand are phosphorothioate internucleoside linkages.

31. The nucleic acid complex of any one of claims 1-29, wherein the first nucleic acid strand does not comprise phosphorothioate internucleoside linkages.

32. The nucleic acid complex of any one of claims 1-29, wherein the internucleoside linkages between (1) the one to three nucleotides adjacent to the 3' of the 5' connector, and/or (2) the one or two nucleotides adjacent to the 5' of the 3' connector, and/or (3) the one to three nucleotides adjacent to the 3' of the 3' connector, are phosphorothioate internucleoside linkages

33. The nucleic acid complex of any one of claims 1-32, wherein the input nucleic acid strand is a RNA.

34. The nucleic acid complex of claim 33, wherein the target RNA is an mRNA, an miRNA, a non-coding RNA, a viral RNA transcript, a cellular RNA transcript, or a combination thereof.

35. The nucleic acid complex of any one of claims 1-34, wherein the overhang of the second nucleic acid strand is capable of binding to the input nucleic acid strand to form a toehold, thereby causing the displacement of the second nucleic acid strand from the first nucleic acid strand.

36. The nucleic acid complex of any one of claims 1-35, wherein the overhang of the second nucleic acid strand is 5 to 20 nucleosides in length, and optionally 9 nucleotides in length.

37. The nucleic acid complex of any one of claims 1-36, wherein all internucleoside linkages of the overhang of the third nucleic acid strand are phosphorothioate internucleoside linkages.

38. The nucleic acid complex of any one of claims 1-37, wherein the 5' terminus, the 3' terminus, or both of the third nucleic acid strand comprises a terminal moiety; and optionally the terminal moiety comprises a ligand, a fluorophore, an exonuclease, a fatty acid, a Cy3, an inverted dT attached to a tri-ethylene glycol, or a combination thereof.

39. A method of modulating a target RNA, comprising:

contacting a cell comprising a target RNA with the nucleic acid complex of any one of claims 1 to 38, wherein an input strand binds to the overhang of the third nucleic acid strand to cause displacement of the third nucleic acid strand from the first nucleic acid strand to release the sequence complementary to the target RNA into the cell, thereby modulating the target RNA.

40. The method of claim 39, wherein contacting the cell with the nucleic acid complex is performed *in vitro*, *in vivo*, *ex vivo*, or a combination thereof.

41. The method of claim 39, wherein contacting the cell with the nucleic acid complex occurs in the body of a subject.

42. The method of any one of claims 39-41, wherein the cell is a disease cell, and optionally the cell is a cancer cell.

43. The method of any one of claims 39-41, wherein the cell is a neuron.

44. A method of treating a disease or a condition, comprising administering the nucleic acid complex of any one of claims 1 to 38 to a subject in need thereof, wherein the input strand binds to the overhang of the third nucleic acid strand to cause displacement of the third nucleic acid strand from the first nucleic acid strand to release the sequence complementary to a

target RNA, thereby reducing the activity of the target RNA or protein expression from the target RNA in the subject to treat the disease or condition.

45. The method of claim 0, wherein the disease or condition is a central nervous system (CNS) disease or disorder or cancer.

46. The method of any one of claims 39-45, wherein the target RNA is a mRNA or a miRNA.

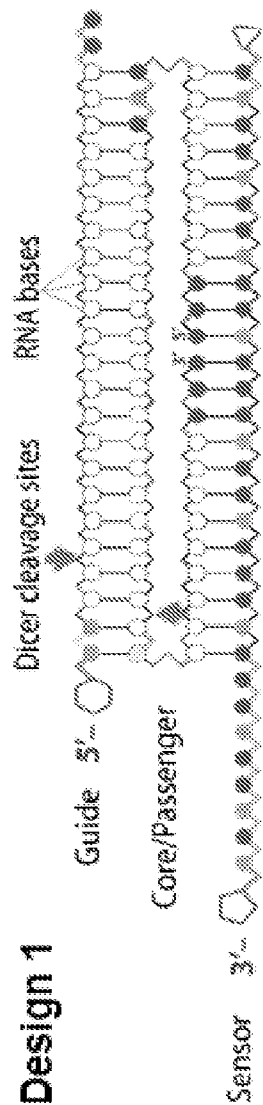
47. The method of any one of claims 39-46, wherein the nucleic acid complex is administered to a subject via a lipid-mediated delivery system, optionally via liposomes, nanoparticles, or micelles.

48. The method of any one of claims 39-46, wherein the nucleic acid complex is administered to a subject via nanoparticles, inorganic nanoparticles, nucleic acid lipid particles, polymeric nanoparticles, lipidoid nanoparticles (LNPs), chitosan and inulin nanoparticles, cyclodextrins nanoparticles, carbon nanotubes, liposomes, micellar structures, capsids, polymers, polymer matrices, hydrogels, dendrimers, nucleic acid nanostructure, exosomes, GalNAc-conjugated melittin-like peptides, or combinations thereof.

49. The method of any one of claims 39-48, wherein the nucleic acid complex is administered to a subject in need thereof via a subcutaneous injection or an intravenous injection.

50. The method of any one of claims 39-49, wherein the nucleic acid complex is administered to a subject in need thereof at a concentration about 0.1-10 nM, optionally about 0.1-1.0 nM.

Design 1



Design 2

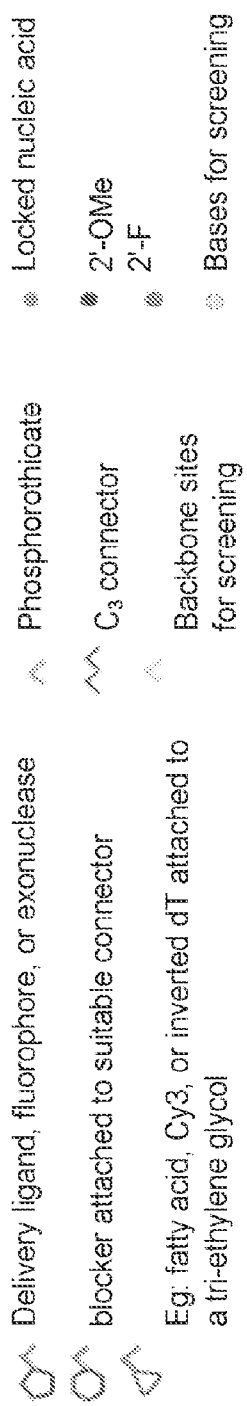
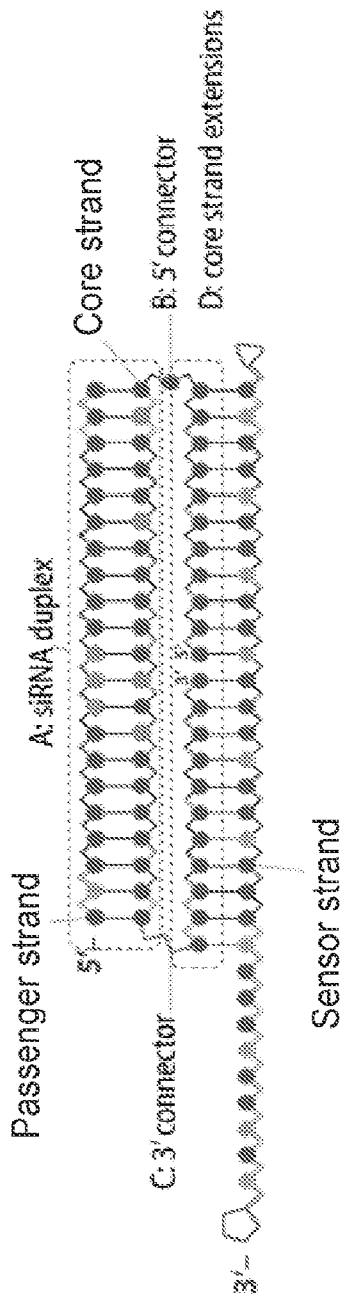


FIG. 1

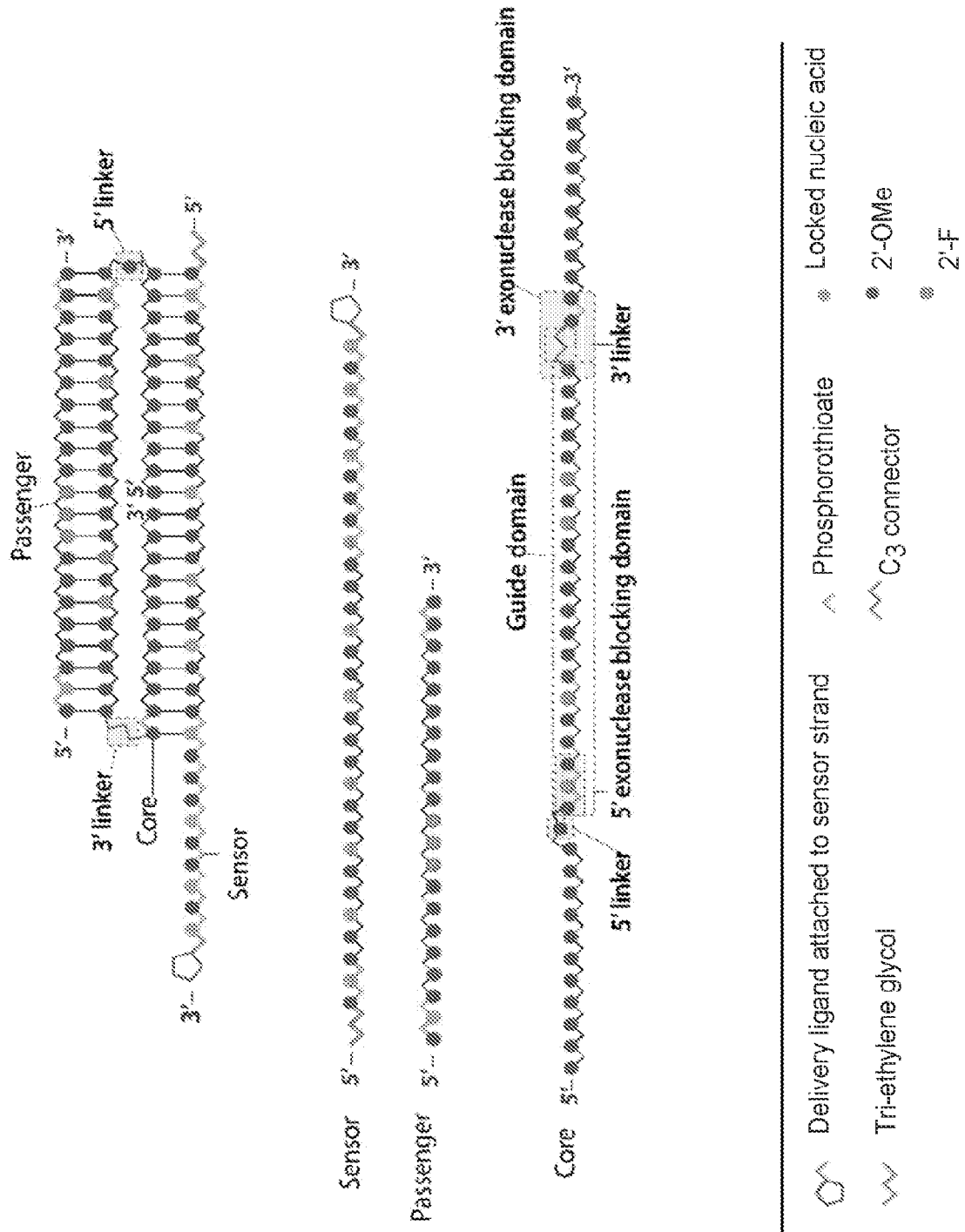
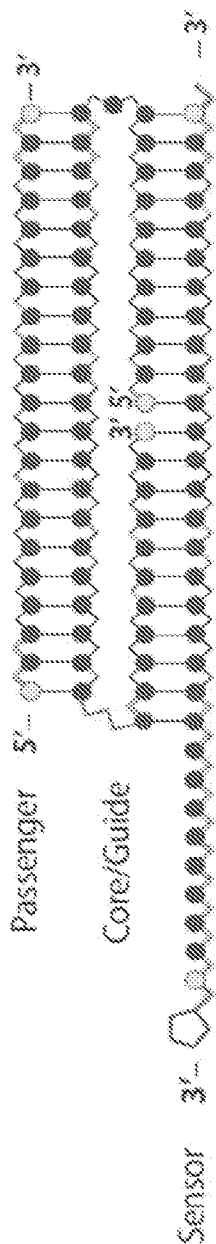


FIG. 2



- Delivery ligand, fluorophore, or exonuclease
- blocker attached to suitable connector
Eg: fatty acid, Cy3, or inverted dT attached to a tri-ethylene glycol
- Phosphorothioate
- C₃ connector
- Locked nucleic acid
- 2'-OMe
- 2'-F
- Backbone sites for screening
- Bases for screening

FIG. 3

Activation in targeted cells

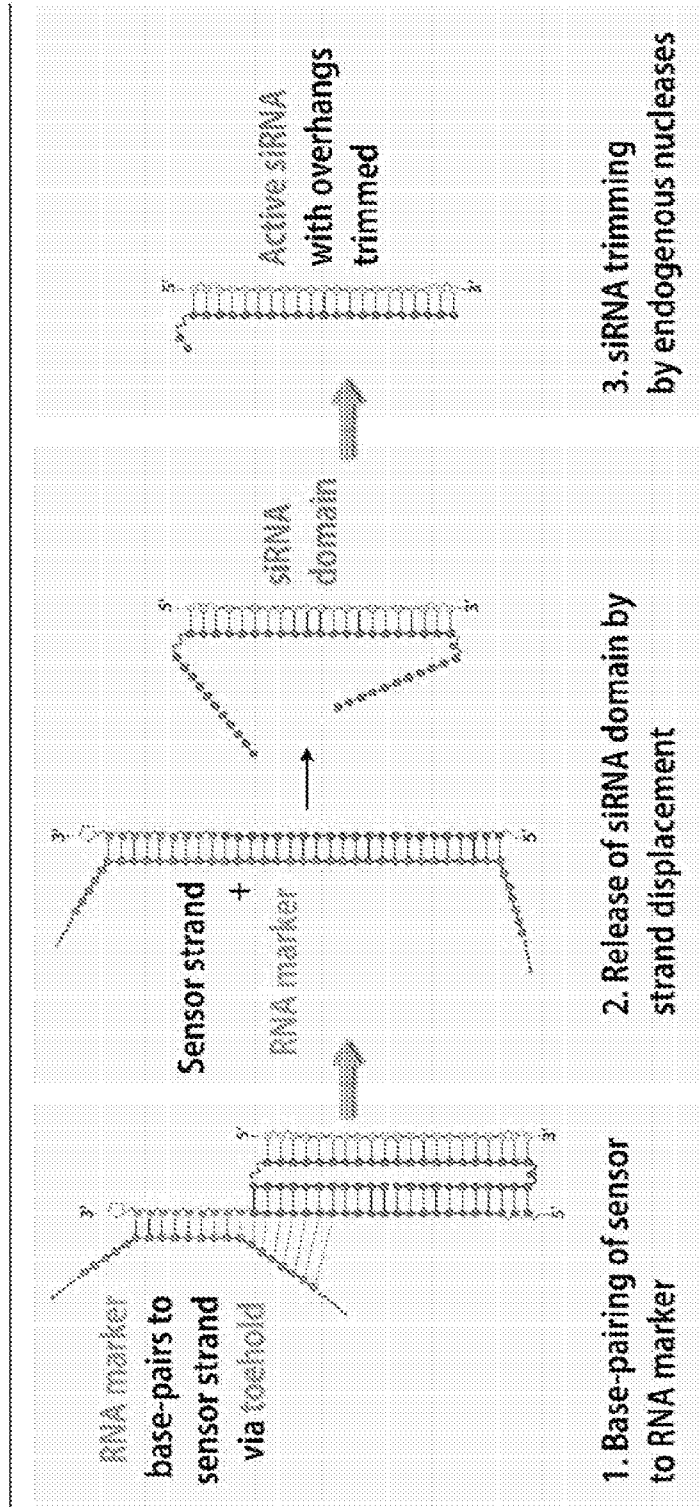


FIG. 4A

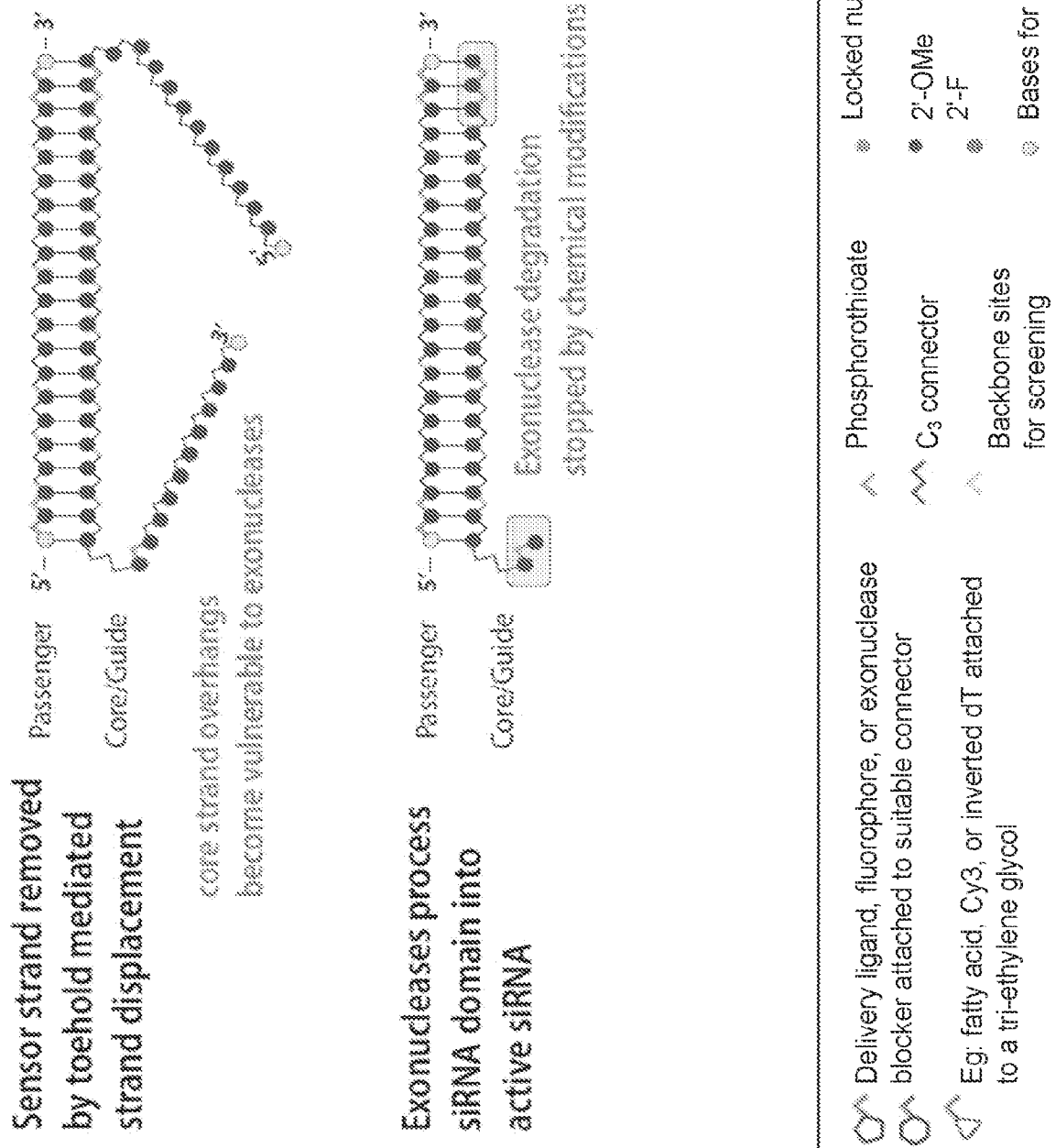


FIG. 4B

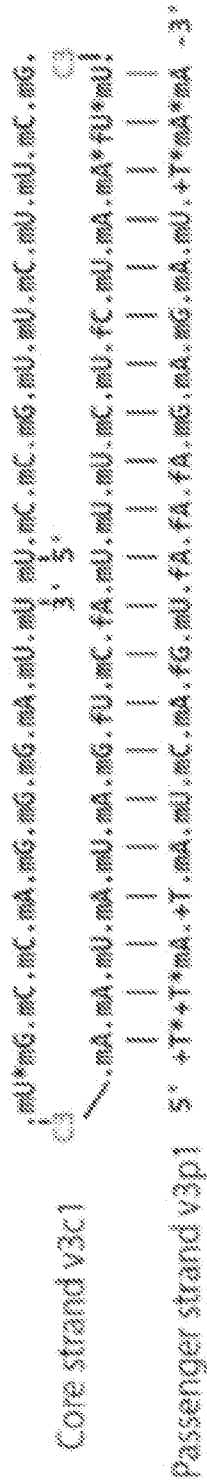


FIG. 5A

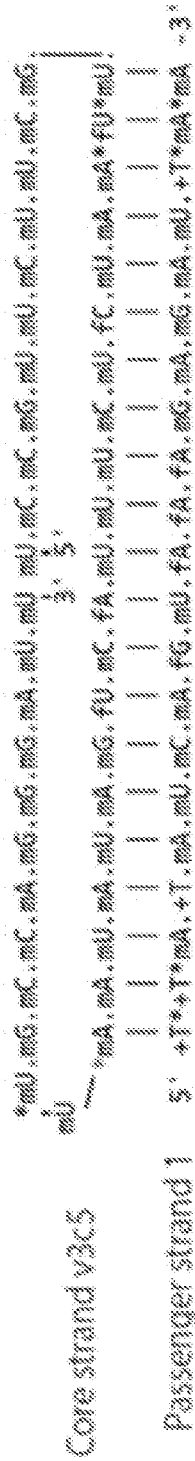


FIG. 5B

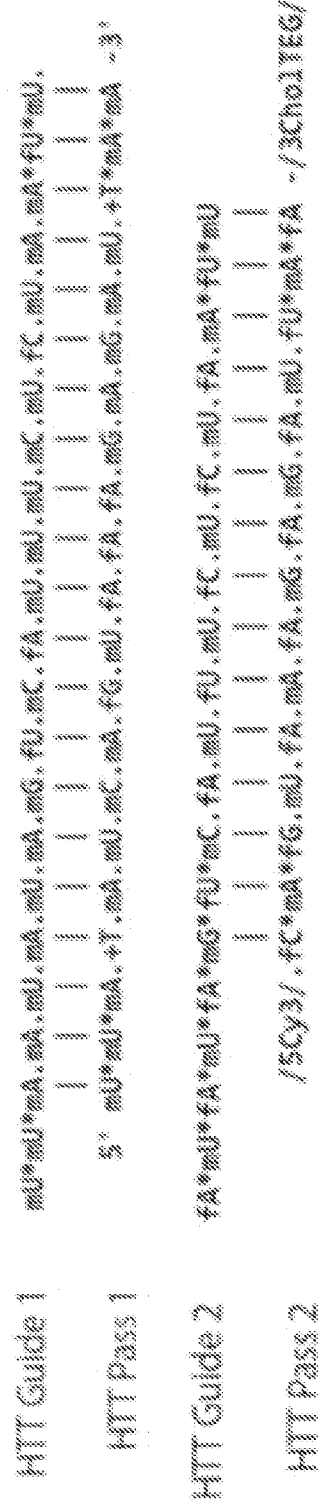


FIG. 6

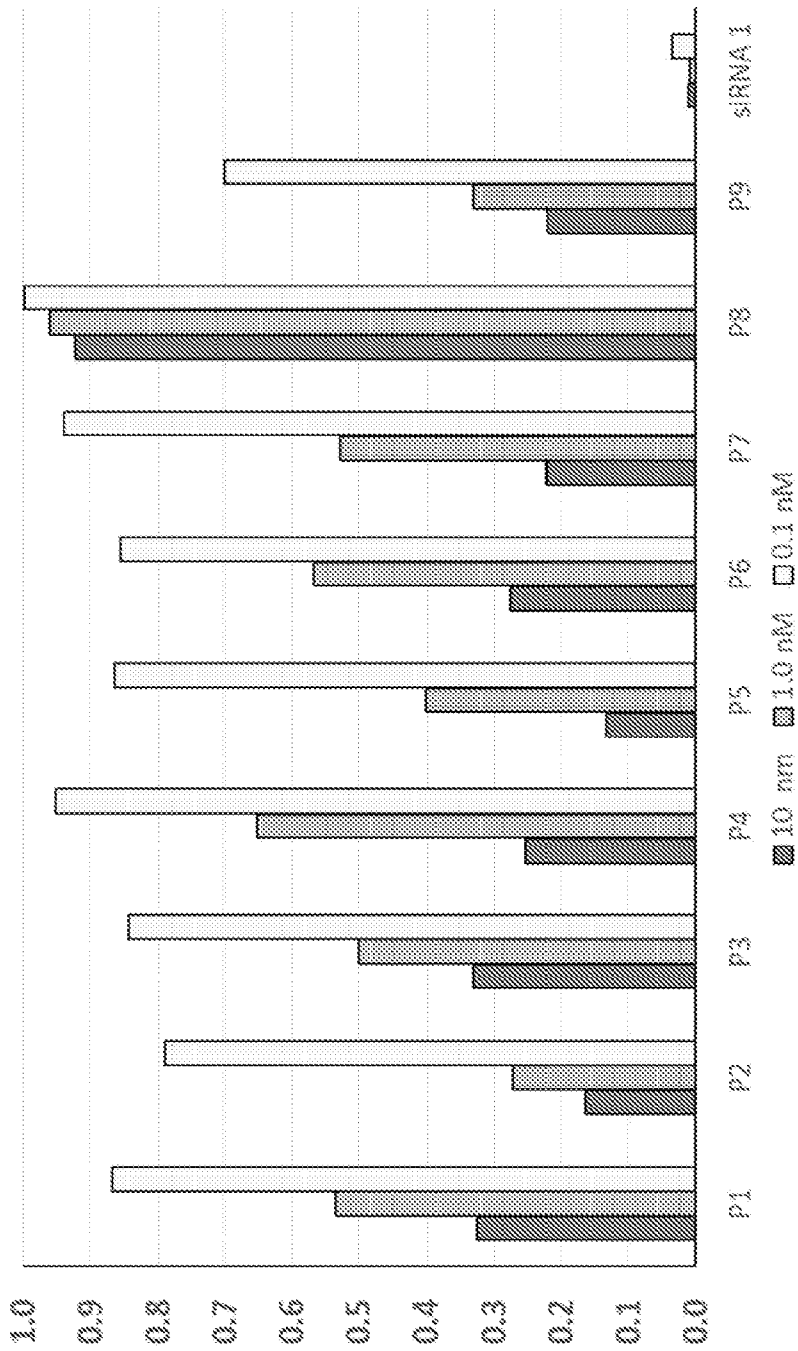


FIG. 8

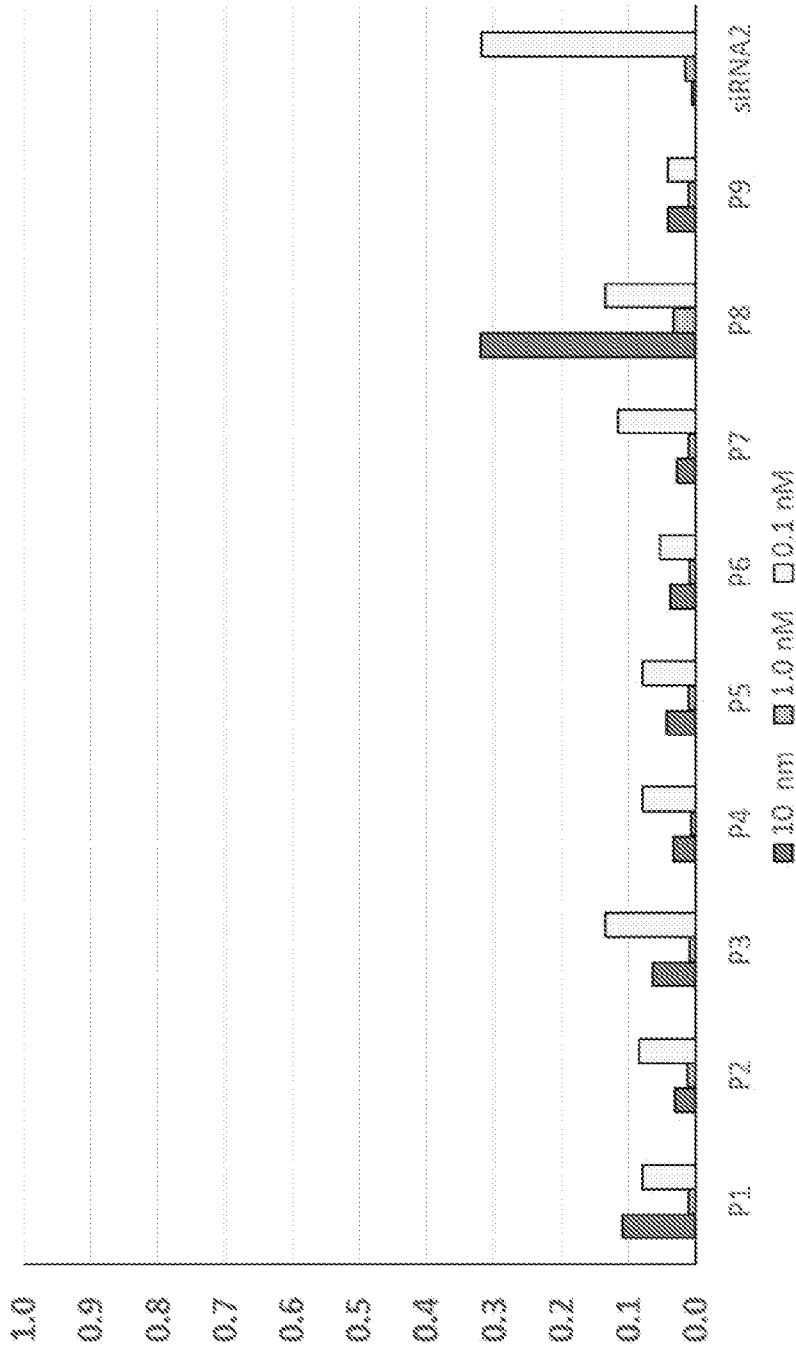
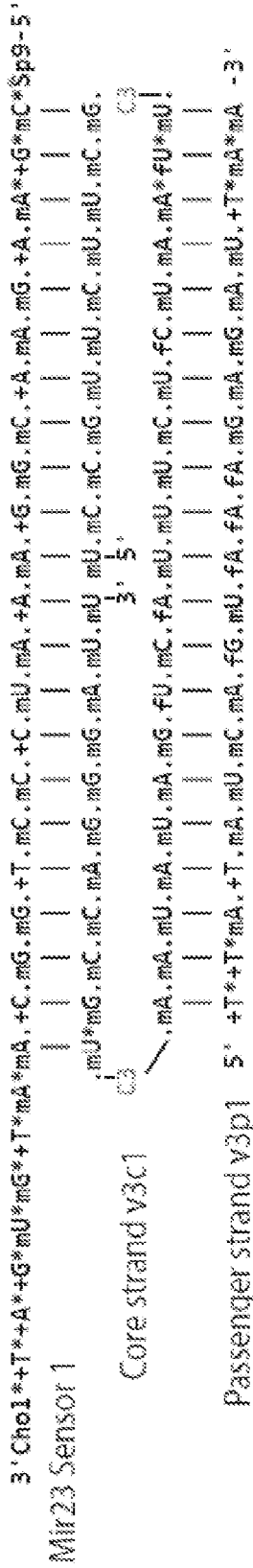
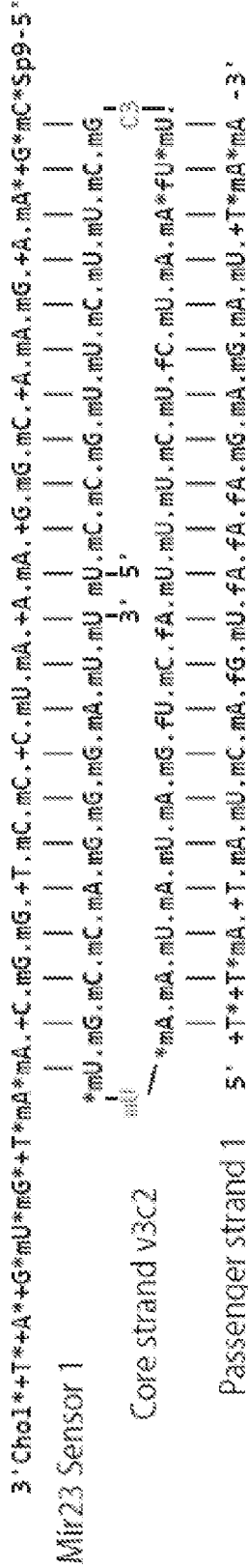


FIG. 10

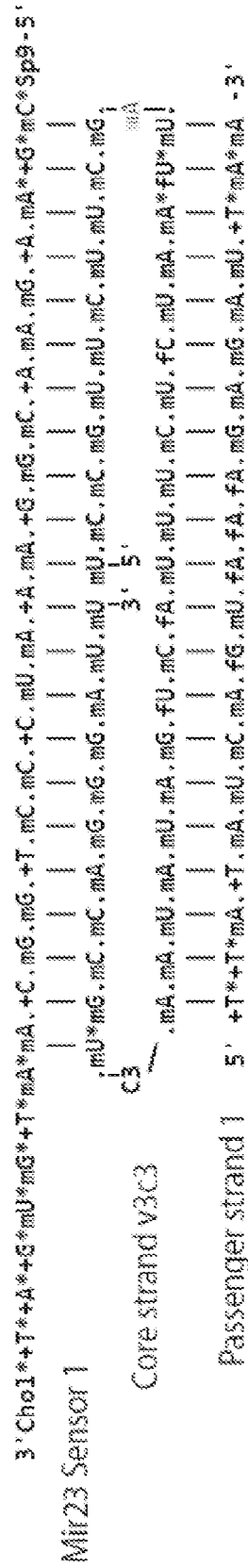
CASI 1



CASI 2



CASI 3



+A, +T, +C, +G = LNA; mA, mU, mC, mG = 2'-OMe; rA, rU, rC, rG = RNA; NH2 = primary amine linker

* = phosphorothioate; . = phosphodiester; C3 = C3 spacer; Sp9 = triethylene glycol; † = Dicer cleavage site

FIG. 11A

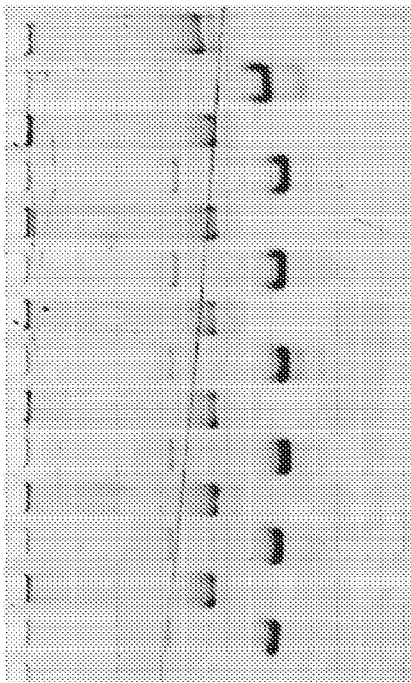


FIG. 12

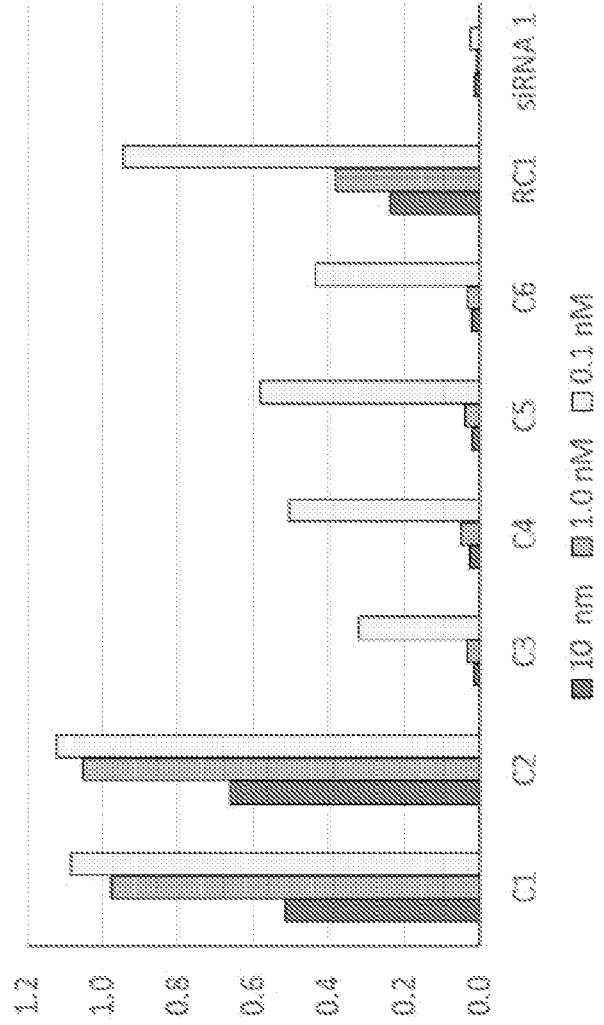


FIG. 13

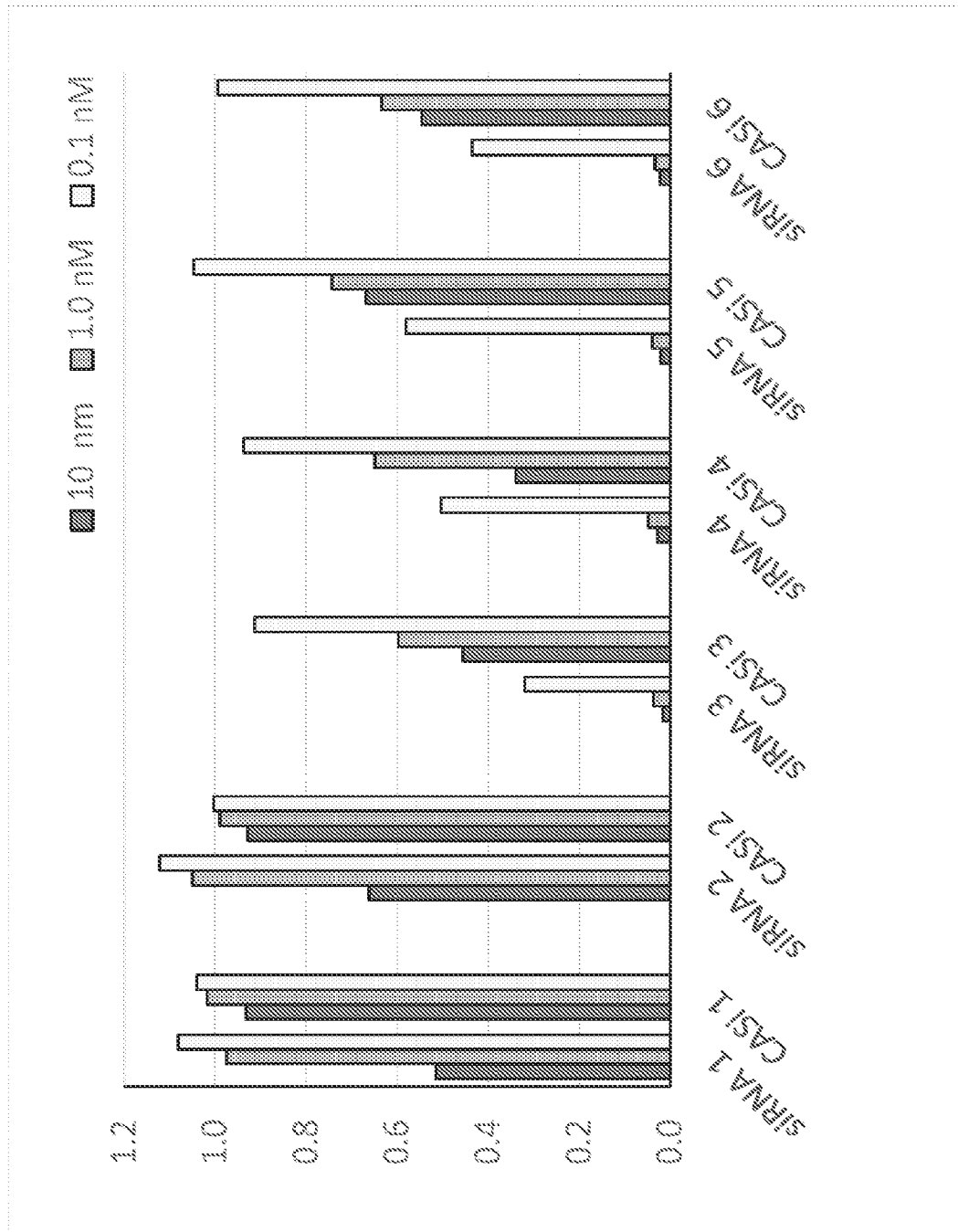


FIG. 14

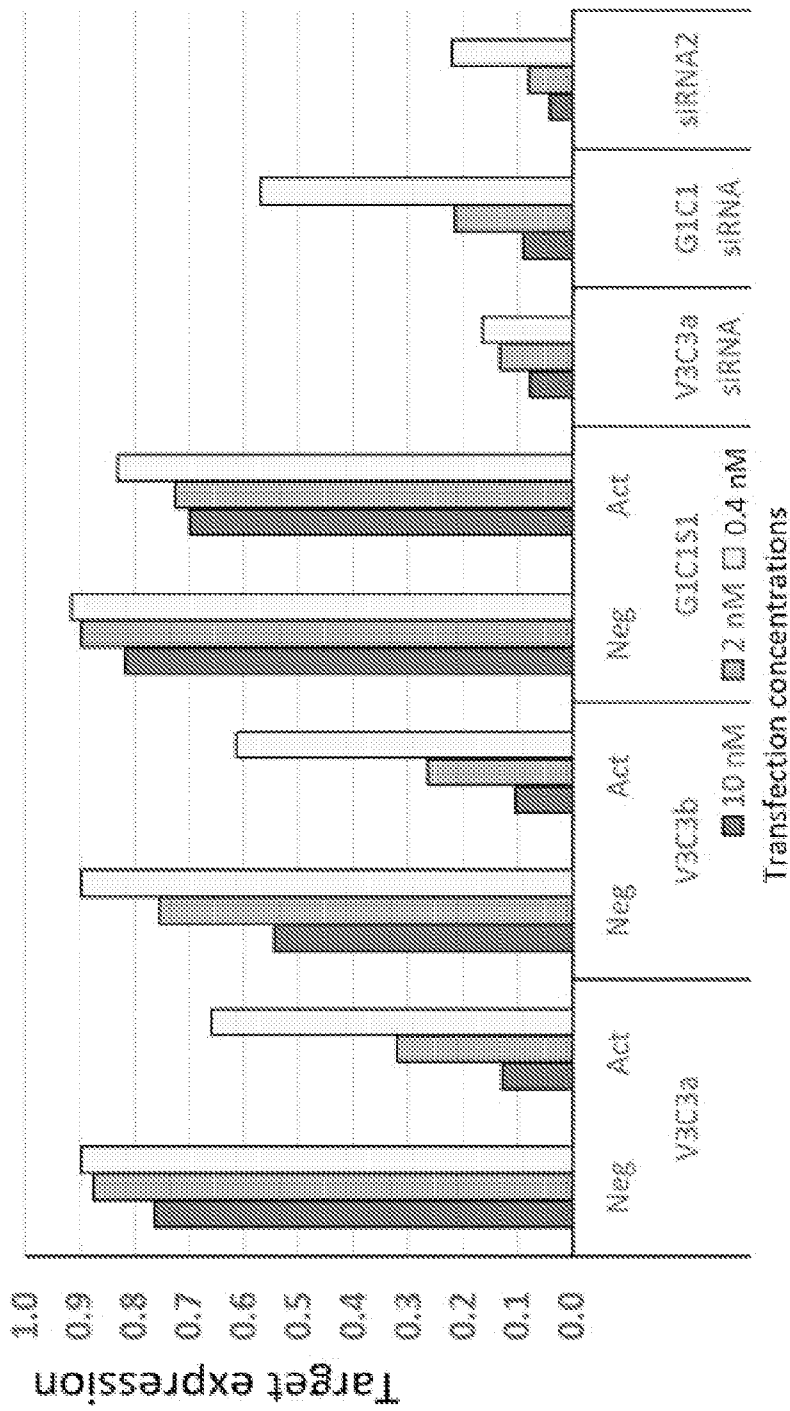


FIG. 16

