



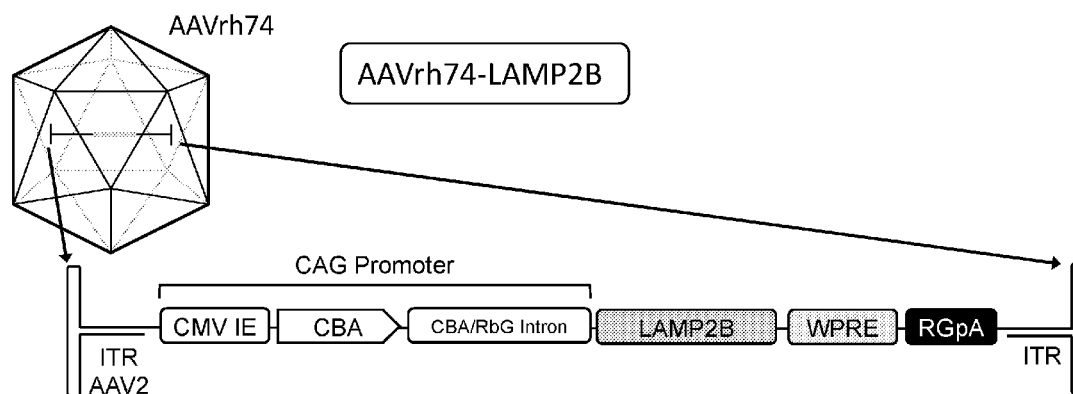
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(54) **Title:** GENE THERAPY VECTORS FOR TREATMENT OF DANON DISEASE



**FIG. 1**

(57) **Abstract:** The present disclosure provides gene therapy vectors comprising a polynucleotide sequence encoding a LAMP-2 polypeptide, methods of use thereof, pharmaceutical compositions, and more. In particular, the disclosure provides recombinant AAV vectors having AAVrh74 serotype expressing LAMP-2A, LAMP-2B, or LAMP-2C for use as a therapeutic in, for example, Danon Disease.



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## **GENE THERAPY VECTORS FOR TREATMENT OF DANON DISEASE**

### **RELATED APPLICATIONS**

This application claims priority to U.S. Provisional Patent Appl. No. 62/934,928, filed November 13, 2019, and U.S. Provisional Patent Appl. No. 62/804,521, filed February 12, 2019, each of which is incorporated herein by reference in its entirety.

### **SEQUENCE LISTING**

This application is being filed electronically via EFS-Web and includes an electronically submitted sequence listing in .txt format. The .txt file contains a sequence listing entitled "ROPA\_013\_02WO\_ST25.txt" created on February 11, 2020 and having a size of ~61 kilobytes. The sequence listing contained in this .txt file is part of the specification and is incorporated herein by reference in its entirety.

### **FIELD OF INVENTION**

The invention relates generally to gene therapy for diseases associated with mutations in lysosome-associated membrane protein 2 (LAMP-2, also known as CD107b).

### **BACKGROUND**

Lysosome-associated membrane protein 2 (LAMP-2, also known as CD107b) is a gene that encodes a lysosome-associated membrane glycoprotein. Alternative splicing of the gene produces three isoforms: LAMP-2A, LAMP-2B, and LAMP-2C. Loss-of-function mutations in LAMP-2 are associated with human diseases, including Danon disease, a familial cardiomyopathy associated with impaired autophagy. Danon disease is a rare but serious cardiac and skeletal myopathy leading to substantial morbidity and early mortality due to arrhythmia and cardiomyopathy. The X-linked nature of inheritance accounts for reported differences in phenotypic severity between men and women. Boucek et al. *Genetics in Medicine* 13:563-568 (2011). The disease is now understood to be caused by a primary deficiency in lysosome-associated membrane protein-2 (LAMP-2), which functions as a lysosomal membrane receptor in chaperone-mediated autophagy. Nishino et al. *Nature* 406:906-910 (2000).

## SUMMARY OF THE INVENTION

The present disclosure provides such gene therapy vectors related to LAMP2, methods of use thereof, pharmaceutical compositions, and more. Although clinical use of adeno-associated virus (AAV) vectors is known, the selection of preferred serotype(s) of AAV for gene therapy remains challenging and unpredictable.

The present disclosure provides improved gene therapy vectors comprising a polynucleotide sequence encoding a LAMP-2 polypeptide, methods of use thereof, pharmaceutical compositions, and more. In particular, the disclosure provides recombinant AAV vectors having AAVrh74 serotype expressing LAMP-2A, LAMP-2B, or LAMP-2C for use as a therapeutic in, for example, Danon disease.

Other features and advantages of the invention will be apparent from and encompassed by the following detailed description and claims.

## BRIEF DESCRIPTION OF THE DRAWINGS

**FIG. 1** depicts an embodiment of an AAV vector having AAVrh74 serotype.

**FIG. 2** shows a bar graph of vector DNA quantification in organs most affected in Danon disease by qPCR.

**FIG. 3A** shows a bar graph of vector DNA quantification in regions of the heart by qPCR.

**FIG. 3B** shows a bar graph of vector DNA quantification in muscles by qPCR.

**FIG. 4** shows a bar graph of mRNA quantification in organs most affected by Danon disease by RT-qPCR.

**FIG. 5A** shows a bar graph of mRNA quantification in regions of the heart by RT-qPCR.

**FIG. 5B** shows a bar graph of mRNA quantification in muscles by RT-qPCR.

**FIG. 6A** shows a micrograph of semi-quantitative mRNA analysis by RNAscope in an untreated left ventricle.

**FIG. 6B** shows micrographs of semi-quantitative mRNA analysis by RNAscope in treated left ventricles.

**FIG. 7A** shows a micrograph of semi-quantitative mRNA analysis by RNAscope in an untreated quadriceps.

**FIG. 7B** shows micrographs of semi-quantitative mRNA analysis by RNAscope in treated quadriceps.

**FIG. 8** shows micrographs of semi-quantitative mRNA analysis by RNAscope in treated gastrocnemius.

**FIG. 9** shows a bar graph of protein quantification in tissues most affected in Danon disease by ELISA.

**FIG. 10A** shows a bar graph of protein quantification in regions of the heart by ELISA.

**FIG. 10B** shows a bar graph of protein quantification in muscles by ELISA.

**FIGS. 11A-11D** show line graphs of clinical pathology measurement in NHP serum over course of study. Clinical pathology levels were assessed as changes in (**FIG. 11A**) alanine aminotransferase, ALT; (**FIG. 11B**) aspartate aminotransferase, AST; (**FIG. 11C**) white blood cells, WBC; and (**FIG. 11D**) neutrophils over the study duration.

## DETAILED DESCRIPTION OF THE INVENTION

The present disclosure provides AAVrh74-based gene therapy vectors that employ optimized expression cassettes to deliver a polynucleotide encoding one of the Lysosome-associated membrane protein 2 (LAMP2) proteins, also known as CD107b. Generally, the LAMP2 is a human LAMP2, though expression of any mammalian LAMP-2 is envisioned. The native *LAMP2* gene encodes by alternative splicing three variants: LAMP-2A, LAMP-2B, and LAMP-2C. LAMP-2B is associated with Danon disease. Although the disclosure concerns primarily Danon disease, LAMP2 is implicated in various other disease, including cancer. The disclosed vectors may be used to treat any of these diseases.

The disclosure further relates to AAVrh74 capsids or capsids having substantial homology to the AAVrh74 capsid and retaining the function of the AAVrh74 capsid. The disclosure provides the sequences listed in **Table 1**. Table 1 further provides polynucleotide sequences used in various embodiments. The sequences are not intended to limit the invention, as substitution or modification of these sequences with different promoters, enhancer, or other genetic elements is contemplated.

**Table 1: Sequences**

SEQ ID NO:	Type	Description
1	nucleotide	AAVrh74 capsid coding sequence
2	protein	AAVrh74 VP1
3	protein	AAVrh74 VP2
4	protein	AAVrh74 VP3
5	protein	LAMP-2B (wild-type)
6	nucleotide	LAMP-2B coding sequence (wild-type)
7	nucleotide	LAMP-2B engineered coding sequence
8	nucleotide	LAMP-2B engineered coding sequence
9	nucleotide	LAMP-2B engineered coding sequence
10	nucleotide	Engineered expression cassette
11	nucleotide	Engineered expression cassette
12	nucleotide	Engineered expression cassette
13	nucleotide	AAV inverted terminal repeat
14	nucleotide	AAV inverted terminal repeat
15	protein	LAMP-2A protein sequence
16	protein	LAMP-2B protein sequence
17	protein	LAMP-2C protein sequence
18	nucleotide	CAG promoter
19	nucleotide	WPRE
20	nucleotide	Kozak sequence
21	nucleotide	Kozak sequence
22	nucleotide	Kozak sequence
23	nucleotide	Kozak sequence
24	nucleotide	Kozak sequence
25	nucleotide	Kozak sequence
26	nucleotide	polyadenylation signal (full length)
27	nucleotide	bGH polyadenylation signal (bGHpA)
28	nucleotide	SV40 early/late polyadenylation signal
29	nucleotide	human growth hormone (HGH) polyadenylation signal

The disclosure provides recombinant adeno-associated virus (rAAV) gene therapy vectors. As used herein, an “rAAV gene therapy vector” refers to a complete virus including nucleic acid and protein components, including capsid proteins. In some embodiments, the capsid protein is encoded by a polynucleotide supplied on a plasmid *in trans* to the transfer plasmid. The polynucleotide sequence of wild-type AAVrh74 *cap* is as follows:

AAVrh74 capsid coding sequence (SEQ ID NO: 1)

ATGGCTGCCGATGGTTATCTTCCAGATTGGCTCGAGGACAACCTCTCTGAGGGCA  
TTCGCGAGTGGTGGGACCTGAAACCTGGAGCCCCGAAACCCAAAGCCAACCAGC  
AAAAGCAGGACAACGGCCGGGGTCTGGTGCTTCCTGGCTACAAGTACCTCGGAC  
CCTTCAACGGACTCGACAAGGGGGAGCCCGTCAACGCGGCGGACGCAGCGGCCC  
TCGAGCACGACAAGGCCTACGACCAGCAGCTCCAAGCGGGTGACAATCCGTACC  
TGCGGTATAATCACGCCGACGCCGAGTTTCAGGAGCGTCTGCAAGAAGATACTG  
CTTTTGGGGGCAACCTCGGGCGCGCAGTCTTCCAGGCCAAAAGCGGGTTCTCG  
AACCTCTGGGCCTGGTTGAATCGCCGGTTAAGACGGCTCCTGGAAAGAAGAGAC  
CGGTAGAGCCATCACCCAGCGCTCTCCAGACTCCTCTACGGGCATCGGCAAGA  
AAGGCCAGCAGCCC GCAAAAAGAGACTCAATTTTGGGCAGACTGGCGACTCAG  
AGTCAGTCCCCGACCCTCAACCAATCGGAGAACCACCAGCAGGCCCTCTGGTCT  
GGGATCTGGTACAATGGCTGCAGGCGGTGGCGCTCCAATGGCAGACAATAACGA  
AGGCGCCGACGGAGTGGGTAGTTCTCAGGAAATTGGCATTGCGATTCCACATG  
GCTGGGCGACAGAGTCATCACACCAGCACCCGCACCTGGGCCCTGCCACCTA  
CAACAACCACCTCTACAAGCAAATCTCCAACGGGACCTCGGGAGGAAGCACCAA  
CGACAACACCTACTTCGGCTACAGCACCCCTGGGGGTATTTTGACTTCAACAGA  
TTCCACTGCCACTTTTACCACGTGACTGGCAGCGACTCATCAACAACA ACTGGG  
GATTCCGGCCCAAGAGGCTCAACTTCAAGCTCTTCAACATCCAAGTCAAGGAGG  
TCACGCAGAATGAAGGCACCAAGACCATCGCCAATAACCTTACCAGCACGATTC  
AGGTCTTTACGGACTCGGAATACCAGCTCCCGTACGTGCTCGGCTCGGCGCACCA  
GGGCTGCCTGCCTCCGTTCCCGGCGGACGTCTTCATGATTCTCAGTACGGGTAC  
CTGACTCTGAACAATGGCAGTCAGGCTGTGGGCCGGTCGTCCTTCTACTGCCTGG  
AGTACTTTCCTTCTCAAATGCTGAGAACGGGCAACA ACTTTGAATTCAGCTACAA  
CTTCGAGGACGTGCCCTTCCACAGCAGCTACGCGCACAGCCAGAGCCTGGACCG  
GCTGATGAACCCTCTCATCGACCAGTACTTGTACTACCTGTCCCGGACTCAAAGC  
ACGGGCGGTACTGCAGGAACTCAGCAGTTGCTATTTTCTCAGGCCGGGCCTAACA  
ACATGTCGGCTCAGGCCAAGA ACTGGCTACCCGGTCCCTGCTACCGGCAGCAAC

GCGTCTCCACGACACTGTCGCAGAACAACAACAGCAACTTTGCCTGGACGGGTG  
 CCACCAAGTATCATCTGAATGGCAGAGACTCTCTGGTGAATCCTGGCGTTGCCAT  
 GGCTACCCACAAGGACGACGAAGAGCGATTTTTTCCATCCAGCGGAGTCTTAAT  
 GTTTGGGAAACAGGGAGCTGGAAAAGACAACGTGGACTATAGCAGCGTGATGCT  
 AACCAGCGAGGAAGAAATAAAGACCACCAACCCAGTGGCCACAGAACAGTACG  
 GCGTGGTGGCCGATAACCTGCAACAGCAAACGCCGCTCCTATTGTAGGGGCCG  
 TCAATAGTCAAGGAGCCTTACCTGGCATGGTGTGGCAGAACCGGGACGTGTACC  
 TGCAGGGTCCCATCTGGGCCAAGATTCTCATAACGGACGGCAACTTTCATCCCTC  
 GCCGCTGATGGGAGGCTTTGGACTGAAGCATCCGCCTCCTCAGATCCTGATTA  
 AACACACCTGTTCCCGCGGATCCTCCGACCACCTTCAATCAGGCCAAGCTGGCTT  
 CTTTCATCACGCAGTACAGTACCGGCCAGGTCAGCGTGGAGATCGAGTGGGAGC  
 TGCAGAAGGAGAACAGCAAACGCTGGAACCCAGAGATTCAGTACACTTCCA  
 ACTACAAATCTACAAATGTGGACTTTGCTGTCAATACTGAGGGTACTTATTCCGA  
 GCCTCGCCCCATTGGCACCCGTTACCTCACCCGTAATCTGTAA

The disclosure further provides protein sequences for AAVrh74 VP1, VP2, and VP3, including SEQ ID NOs: 2-4, and homologs or functional variants thereof.

AAVrh74 VP1 (SEQ ID NO: 2)

MAAGGGAPMADNNEGADGVGSSSGNWHCDSTWLGDRVITTSTRTWALPTYNNHL  
 YKQISNGTSGGSTNDNTYFGYSTPWGYFDNRFHCHFSPRDWQRLINNNWGFRPKR  
 LNFKLFNIQVKEVTQNEGTKTIANNLTSTIQVFTDSEYQLPYVLGSAHQGCLPPFPAD  
 VFMIPQYGYLTLNNGSQAVGRSSFYCLEYFPSQMLRTGNNFEFSYNFEDVPFHSSYA  
 HSQSLDRLMNPLIDQYLYYLSRTQSTGGTAGTQQLFSQAGPNNMSAQAKNWLPGP  
 CYRQQRVSTTLSQNNNSNFAWTGATKYHLNGRDSLVPNGVAMATHKDDEERFFPS  
 SGVLMFGKQGAGKDNVDYSSVMLTSEEEIKTTNPVATEQYGVVADNLQQQNAAPI  
 VGAVNSQGALPGMVWQNRDVYLQGPWAKIPHTDGNFHPSPLMGGFGLKHPPPQIL  
 IKNTPVPADPPTTFNQAKLASFITQYSTGQVSVEIEWELQKENSKRWNPEIQYTSNYY  
 KSTNVDFAVNTEGTYSEPRPIGTRYLTRNL

AAVrh74 VP2 (SEQ ID NO: 3)

STIQVFTDSEYQLPYVLGSAHQGCLPPFPADVFMIPQYGYLTLNNGSQAVGRSSFYCL  
 EYFPSQMLRTGNNFEFSYNFEDVPFHSSYAHSQSLDRLMNPLIDQYLYYLSRTQSTG

GTAGTQQLLFSQAGPNNMSAQAKNWLPGPCYRQQRVSTTLSQNNNSNFAWTGATK  
 YHLNGRDSL VNPGVAMATHKDDDEERFFPSSGVLMFGKQGAGKDNVDYSSVMLTSE  
 EEIKTTNPVATEQYGVVADNLQQQNAAPIVGAVNSQGALPGMVWQNRDVYLQGP  
 I WAKIPHTDGNFHPSPMLGGFGLKHPPPQILIKNTPVPADPPTTFNQAKLASFITQYSTG  
 QVSVEIEWELQKENS KRWNPEIQYTSNYYKSTNVDFAVNTEGTYSEPRPIGTRYLTR  
 NL

AAVrh74 VP3 (SEQ ID NO: 4)

RTGNNFEFSYNFEDVPFHSSYAHSQSLDRLMNPLIDQYLYLSRTQSTGGTAGTQQL  
 LFSQAGPNNMSAQAKNWLPGPCYRQQRVSTTLSQNNNSNFAWTGATKYHLNGRDS  
 LVNPGVAMATHKDDDEERFFPSSGVLMFGKQGAGKDNVDYSSVMLTSEEEIKTTNPV  
 ATEQYGVVADNLQQQNAAPIVGAVNSQGALPGMVWQNRDVYLQGP I WAKIPHTD  
 GNFHPSPMLGGFGLKHPPPQILIKNTPVPADPPTTFNQAKLASFITQYSTGQVSVEIEW  
 ELQKENS KRWNPEIQYTSNYYKSTNVDFAVNTEGTYSEPRPIGTRYLTRNL

In certain cases, the AAVrh74 capsid comprises the amino acid sequence set forth in SEQ ID NO: 2. In some embodiments, the rAAV vector comprises a polypeptide that comprises, or consists essentially of, or yet further consists of a sequence, *e.g.*, at least 65%, at least 70%, at least 75%, at least 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, or 89%, more typically 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more identical to amino acid sequence of AAVrh74 VP1 which is set forth in SEQ ID NO: 2. In some embodiments, the rAAV vector comprises a polypeptide that comprises, or consists essentially of, or yet further consists of a sequence, *e.g.*, at least 65%, at least 70%, at least 75%, at least 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, or 89%, more typically 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more identical to amino acid sequence of AAVrh74 VP2 which is set forth in SEQ ID NO: 3. In some embodiments, the rAAV vector comprises a polypeptide that comprises, or consists essentially of, or yet further consists of a sequence, *e.g.*, at least 65%, at least 70%, at least 75%, at least 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, or 89%, more typically 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more identical to amino acid sequence of AAVrh74 VP3 which is set forth in SEQ ID NO: 4.

The wild-type polypeptide sequence of LAMP-2B (SEQ ID NO: 5) and the wild-type polynucleotide sequence of LAMP-2B (SEQ ID NO: 6) are, respectively:

MVCFRLFPVPGSGLVLVCLVLGAVRSYALELNLTDSENATCLYAKWQMNFT  
 VRYETTNKTYKTVTISDHGTVTYNGSICGDDQNGPKIAVQFGPGFSWIANFTK  
 AASTYSIDSVSFSYNTGDNTTFPDAEDKGILTVDELLAIRIPLNDFRCNSLSTL  
 EKNDVVQHYWDVLVQAFVQNGTVSTNEFLCDKDKTSTVAPTIHTTVPSPTTT  
 PTPKEKPEAGTYSVNNNGNDTCLLATMGLQLNITQDKVASVININPNTTHSTGS  
 CRSHTALLRLNSSTIKYLDFVFAVKNENRFYLKEVNISMVYLVNGSVFSIANN  
 LSYWDAPLGSSYMCNKEQTVSVSGAFQINTFDLRVQPFNVTQGKYSTAQEC  
 LDDDTILIPVIVGAGLSGLIIVIVIAVYVIGRRKSYAGYQT (SEQ ID NO: 5); and

ATGGTGTGCTTCCGCCTCTTCCCGGTTCCGGGCTCAGGGCTCGTTCTGGTCTGCCT  
 AGTCCTGGGAGCTGTGCGGTCTTATGCATTGGAACCTAATTTGACAGATTCAGAA  
 AATGCCACTTGCCTTTATGCAAAATGGCAGATGAATTCACAGTTCGCTATGAAA  
 CTACAAATAAACTTATAAACTGTAACCATTTAGACCATGGCACTGTGACATA  
 TAATGGAAGCATTGTGGGGATGATCAGAATGGTCCCAAATAGCAGTGCAGTT  
 CGGACCTGGCTTTTCTGGATTGCGAATTTTACCAAGGCAGCATCTACTTATCA  
 ATTGACAGCGTCTCATTTCCTACAACACTGGTGATAACACAACATTTCTGATG  
 CTGAAGATAAAGGAATTCTTACTGTTGATGAACTTTTGGCCATCAGAATTCCATT  
 GAATGACCTTTTTAGATGCAATAGTTTATCAACTTTGGAAAAGAATGATGTTGTC  
 CAACACTACTGGGATGTTCTTGTACAAGCTTTTGTCCAAAATGGCACAGTGAGCA  
 CAAATGAGTTCCTGTGTGATAAAGACAAAACCTCAACAGTGGCACCCACCATAC  
 ACACCACTGTGCCATCTCCTACTACAACACCTACTCCAAAGGAAAAACCAGAAG  
 CTGGAACCTATTCAGTTAATAATGGCAATGATACTTGTCTGCTGGCTACCATGGG  
 GCTGCAGCTGAACATCACTCAGGATAAGGTTGCTTCAGTTATTAACATCAACCCC  
 AATACAACCTCACTCCACAGGCAGCTGCCGTTCTCACACTGCTCTACTTAGACTCA  
 ATAGCAGCACCATTAAGTATCTAGACTTTGTCTTTGCTGTGAAAAATGAAAACCG  
 ATTTTATCTGAAGGAAGTGAACATCAGCATGTATTTGGTTAATGGCTCCGTTTTT  
 AGCATTGCAAATAACAATCTCAGCTACTGGGATGCCCCCTGGGAAGTTCTTATA  
 TGTGCAACAAAGAGCAGACTGTTTCAGTGTCTGGAGCATTTCAGATAAATACCTT  
 TGATCTAAGGGTTCAGCCTTTCAATGTGACACAAGGAAAGTATTCTACAGCCCAA  
 GAGTGTTTCGCTGGATGATGACACCATTCTAATCCCAATTATAGTTGGTGCTGGTC  
 TTTACAGGCTTGATTATCGTTATAGTGATTGCTTACGTAATTGGCAGAAGAAAAAG  
 TTATGCTGGATATCAGACTCTGTAA (SEQ ID NO:6).

In an embodiment, the transgene shares at least 95% identity to the polynucleotide sequence of SEQ ID NO: 5. In an embodiment, the transgene shares at least 99% identity to the polynucleotide sequence of SEQ ID NO: 5. In an embodiment, the transgene comprises the polynucleotide sequence of SEQ ID NO: 5. In embodiment, the transgene shares at least 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or complete identity to SEQ ID NO:5.

In an embodiment, the transgene encodes a polypeptide that shares at least 95% identity to the amino acid sequence of SEQ ID NO: 6. In an embodiment, the transgene encodes a polypeptide shares at least 99% identity to the amino acid sequence of SEQ ID NO: 6. In an embodiment, the polypeptide encoded by the transgene comprises the amino acid sequence of SEQ ID NO: 6. In embodiment, the polypeptide encoded by the transgene shares at least 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or complete identity to SEQ ID NO:6.

Disclosed herein are modifications to the gene sequence of LAMP-2B including: codon-optimization, CpG depletion, removal of cryptic splice sites, and reduction of alternative open-reading frames (ORFs). In embodiments, the disclosure provides a transgene encoding an isoform of lysosome-associated membrane protein 2 (LAMP-2) or a functional variant thereof. In embodiments, the transgene shares at least 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or complete identity to a sequence selected from SEQ ID NO: 7-9. The disclosure provides at least three variant gene sequences for LAMP-2B (SEQ ID NO: 7-9):

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ATGGTCTGCTTCAGACTGTTCCCTGTCCCTGGATCTGGTCTGGTGCTTGTGTGCTT
GGTGCTGGGTGCTGTGAGATCCTATGCCCTTGAGCTGAACCTGACTGACTCAGAA
AATGCCACTTGCCTGTATGCCAAGTGGCAGATGAACTTCACTGTGAGATATGAGA
CTACCAACAAGACCTACAAGACTGTGACCATCTCAGACCATGGCACTGTCACCTA
CAATGGATCAATCTGTGGTGATGATCAGAATGGCCCAAAGATAGCAGTGCAGTT
TGGGCCCGGTTTTTCTGGATTGCTAACTTCACCAAGGCAGCCTCCACCTACAGC
ATTGACTCAGTCAGCTTCACTGCTACAACACTGGGGATAACACCACCTTCCCTGACG
CAGAGGACAAGGGAATCCTTACTGTGGACGAACTCCTGGCAATCAGAATCCCCC
TTAACGACCTGTTTCAGATGCAACTCCCTTTCAACCCTTGAAAAGAATGATGTGGT
GCAACACTATTGGGACGTCCTGGTGCAAGCCTTTGTGCAGAATGGGACAGTGAG
TACCAACGAGTTCCTCTGTGACAAGGACAAGACCAGCACTGTGGCCCCCACTATC
CACACCACTGTGCCCAGCCCTACCACTACCCCCACCCCTAAAGAGAAGCCAGAA

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GCTGGAACCTACTCAGTCAACAATGGAAATGACACATGCCTCCTTGCCACCATGG  
GACTGCAGCTGAACATCACTCAGGACAAGGTGGCCTCAGTGATTAACATCAACC  
CTAACACCACTCATAGCACTGGGAGCTGCAGATCACATACAGCTCTGCTGAGGCT  
CAACTCCTCCACCATCAAGTACCTGGACTTTGTGTTTGCTGTGAAGAATGAGAAC  
AGGTTCTACCTCAAGGAAGTGAACATTTCCATGTACCTGGTCAATGGTTCAGTGT  
TCTCTATTGCCAACAACAATCTGAGCTACTGGGATGCACCCCTGGGATCCTCCTA  
CATGTGCAACAAGGAGCAGACTGTGAGTGTGTCAGGTGCTTTTCAGATCAACACT  
TTTGACCTGAGGGTGCAGCCCTTCAATGTGACTCAGGGAAAGTACTCCACTGCAC  
AAGAGTGTTCCCTGGATGATGACACTATCCTCATCCCCATTATTGTGGGAGCTGG  
ACTGTCAGGATTGATTATAGTGATTGTGATTGCTTATGTGATTGGAAGGAGAAAG  
AGCTATGCTGGCTACCAGACCCTGTAA (SEQ ID NO: 7);

ATGGTGTGCTTTAGACTGTTTCCTGTGCCTGGTTCAGGGCTGGTCCTGGTCTGTCT  
GGTGCTGGGGGCTGTCAGAAGCTATGCCTTGGAGCTGAACCTCACTGATAGTGA  
AAATGCCACTTGTCTGTATGCTAAGTGGCAGATGAACTTCACTGTGAGATATGAA  
ACCACCAACAAGACTTACAAAACAGTGACCATCTCAGATCATGGAACTGTGACC  
TACAACGGCAGCATTGTGGAGACGACCAGAACGGACCAAAAATCGCTGTCCAA  
TTTGGGCCTGGATTCTCCTGGATTGCCAATTTCACTAAAGCTGCCTCCACATATTC  
AATTGACTCAGTGTCTTCTCCTACAACACTGGGGACAACACTACTTTCCCTGAT  
GCTGAAGATAAGGGAATCTTGACAGTGGATGAGCTGCTGGCTATCAGGATCCCT  
TTGAATGACCTGTTTAGGTGTAATTCAGTACTGAGCACTCTGGAGAAGAACGACGTGG  
TGCAGCACTACTGGGACGTGCTGGTGCAGGCCTTTGTGCAGAACGGCACTGTGTC  
CACCAACGAATTCCTGTGTGATAAGGACAAAACCTTCCACTGTGGCACCTACAATT  
CACACTACTGTGCCTTCACCTACCACCACTCCAACCTCAAAGGAAAAGCCTGAAG  
CAGGAACCTACTCTGTGAACAATGGCAATGATACCTGTCTGTTGGCCACCATGGG  
CCTCCAACCTGAACATTACTCAGGACAAGGTGGCCTCAGTGATTAACATTAACCCC  
AACACTACCACTCCACTGGCAGCTGTAGATCACACACAGCCTTGCTCAGACTGA  
ATAGCAGCACCATCAAGTATTTGGATTTTGTGTTTGCAGTGAAGAATGAAAACAG  
GTTCTACCTGAAGGAAGTCAACATCTCAATGTACCTGGTGAACGGCTCAGTGTTT  
AGCATTGCCAACAACAACCTCTCCTATTGGGACGCTCCACTGGGGAGCAGCTAC  
ATGTGTAACAAGGAACAGACTGTGTCAGTGTGTCAGGAGCCTTCCAGATTAACACC  
TTTGATCTGAGGGTCCAACCCTTTAATGTCACTCAAGGAAAGTATAGCACTGCCC  
AGGAGTGCTCCCTGGATGATGACACCATTCTGATTCCAATCATTGTGGGTGCAGG

ACTTTCTGGGCTTATTATTGTGATTGTGATTGCCTATGTGATTGGCAGAAGGAAA  
TCCTATGCAGGGTACCAAACCTCTGTAA (SEQ ID NO: 8); or

ATGGTCTGTTTTAGGCTGTTCCCTGTCCCTGGTTCAGGACTGGTCTTAGTGTGTCT  
GGTGCTTGGAGCTGTCAGAAGCTATGCCCTGGAGCTGAACCTGACTGACTCAGA  
AAATGCCACTTGCCTGTATGCCAAGTGGCAGATGAACTTCACTGTCAGATATGAA  
ACCACCAACAAGACCTATAAGACTGTGACCATCTCAGACCATGGCACTGTGACTT  
ACAATGGGTCAATTTGTGGAGATGACCAGAATGGCCCTAAGATAGCTGTCCAGT  
TTGGTCCAGGATTCAGCTGGATTGCCAACTTCACCAAGGCAGCCAGCACCTACAG  
CATTGACTCTGTGTCCTTCTCCTACAACACAGGAGACAACACCACTTTCCCTGAT  
GCAGAGGACAAAGGTATCCTGACTGTGGATGAGTTGCTGGCAATCAGGATCCCA  
CTGAACGATCTGTTTCAGGTGCAACTCACTGTCCACTCTGGAAAAGAATGATGTGG  
TGCAGCACTATTGGGATGTGCTAGTCCAGGCCTTTGTCCAGAATGGGACTGTGTC  
AACTAATGAGTTCCTGTGTGACAAGGACAAGACAAGCACTGTAGCCCCACTAT  
CCATACCACAGTACCTAGCCCCACCACTACTCCAACCCCCAAGGAGAAGCCTGA  
GGCTGGCACCTACTCAGTGAACAATGGGAATGACACCTGTTTGCTGGCCACTATG  
GGACTCCAACCTGAACATCACCCAGGACAAAGTGGCCTCTGTGATCAATATCAAT  
CCCCAACACCACCCACAGCACTGGGTCCCTGCAGAAGCCACACTGCCCTCCTGAGG  
CTCAACTCATCAACTATCAAGTACTTGGATTTTGTGTTTGCAGTGAAGAATGAGA  
ACAGATTCTACCTCAAAGAGGTCAACATTTCAATGTACCTGGTGAATGGGAGTGT  
GTTCTCCATTGCTAACAACAACCTGAGCTACTGGGATGCCCTCTGGGCTCCTCA  
TACATGTGCAACAAGGAACAGACTGTGAGTGTGTCAGGGGCCCTTCCAGATCAAC  
ACTTTTGACCTGAGAGTGCAGCCCTTAATGTGACACAGGGAAAGTACAGCACT  
GCTCAGGAGTGCAGCCTGGATGATGACACTATCCTGATCCCTATCATTGTGGGGG  
CAGGCCTGTCTGGACTCATTATTGTGATTGTGATTGCCTATGTGATAGGGAGAAG  
GAAGTCTTATGCTGGATAACCAGACCCTGTAA (SEQ ID NO: 9).

In an embodiment, the transgene shares at least 95% identity to a sequence selected from SEQ ID NO: 7-9. In an embodiment, the transgene shares at least 99% identity to a sequence selected from SEQ ID NO: 7-9. In an embodiment, the transgene comprises a sequence selected from SEQ ID NO: 7-9. In an embodiment, the transgene shares at least 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or complete identity to SEQ ID NO: 7. In an embodiment, the transgene shares at least 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or complete identity to SEQ ID NO: 8. In an embodiment, the transgene shares at least 85%, 86%, 87%, 88%, 89%,

90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or complete identity to SEQ ID NO: 9.

In some cases, the transgene has a polynucleotide sequence that is different from the polynucleotide sequence of a reference sequence, *e.g.*, a “native” or “wild-type” LAMP-2B sequence. In some embodiments, the transgene shares at most 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, or 95% identity with a reference sequence. In some embodiments, the reference sequence is SEQ ID NO: 6. For example, SEQ ID NO: 7 shares 78.5% identity to SEQ ID NO: 6.

In some embodiments, the transgene is similar to or identical to a subsequence of any one of SEQ ID NOs: 5 or 7-9. In some embodiments, the transgene comprises a subsequence of any one of SEQ ID NOs: 5 or 7-9. In various embodiments, the subsequence may comprise any set of consecutive nucleotides (nt) in the full sequence having a length of at least about 50 nt, at least about 100 nt, at least about 150 nt, at least about 250 nt, at least about 200 nt, at least about 350 nt, at least about 450 nt, at least about 400 nt, at least about 450 nt, at least about 550 nt, at least about 600 nt, at least about 650 nt, at least about 600 nt, at least about 650 nt, at least about 700 nt, at least about 750 nt, at least about 800 nt, at least about 850 nt, at least about 900 nt, at least about 950 nt, at least about 1000 nt, at least about 1050 nt, at least about 1100 nt, at least about 1150 nt, or at least about 1200 nt.

In some embodiments, the transgene encodes a polypeptide similar to or identical to a subsequence of any one of SEQ ID NOs: 6 or 16-18. In some embodiments, the transgene encodes a polypeptide comprises a subsequence of any one of SEQ ID NOs: 6 or 16-18. In some embodiments, the subsequence may comprises any set of consecutive amino acids (aa) in the full sequence having a length of at least about 20 aa, at least about 30 aa, at least about 50 aa, at least about 70 aa, at least about 80 aa, at least about 100 aa, at least about 120 aa, at least about 130 aa, at least about 150 aa, at least about 170 aa, at least about 180 aa, at least about 200 aa, at least about 220 aa, at least about 230 aa, at least about 250 aa, at least about 270 aa, at least about 280 aa, at least about 300 aa, at least about 320 aa, at least about 330 aa, at least about 350 aa, at least about 370 aa, at least about 380 aa, or at least about 400 aa.

In some embodiments, the transgene encodes a LAMP-2 polypeptide comprising an N-terminal truncation 1 to 10 amino acids (aa), 1 to 20 aa, 1 to 30 aa, 1 to 40 aa, or 1 to 50 aa, or an N-terminal truncation 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20,

21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 48, 50, or more aa; and/or a C-terminal truncation 1 to 10 amino acids (aa), 1 to 20 aa, 1 to 30 aa, 1 to 40 aa, or 1 to 50 aa, or a C-terminal truncation 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 48, 50, or more aa.

In some embodiments, the subsequence of the LAMP2 polypeptide comprises a functional variant of LAMP-2A, LAMP-2B, or LAMP-2C. As used herein, a “functional variant” refers to polypeptide sharing sequence similarity to a reference LAMP-2A, LAMP-2B, or LAMP-2C and having at least one biological property of LAMP-2A, LAMP-2B, or LAMP-2C. The biological property may include the ability to specifically interact with one or more binding partners, the ability to bind an anti-LAMP2 antibody, and/or the ability to complement a defect in LAMP2 activity in a cell, tissue, and/or organism.

In some embodiments, the subsequence of the LAMP2 polypeptide comprises a functional fragment of LAMP-2A, LAMP-2B, or LAMP-2C. As used herein, a “functional fragment” refers to polypeptide sharing sequence similarity to a subsequence of a reference LAMP-2A, LAMP-2B, or LAMP-2C and having at least one biological property of LAMP-2A, LAMP-2B, or LAMP-2C. The biological property may include the ability to specifically interact with one or more binding partners, the ability to bind an anti-LAMP2 antibody, and/or the ability to complement a defect in LAMP2 activity in a cell, tissue, and/or organism.

In an embodiment, the transgene is codon-optimized for expression in a human host cell. In an embodiment, the transgene coding sequence is modified, or “codon optimized” to enhance expression by replacing infrequently represented codons with more frequently represented codons. The coding sequence is the portion of the mRNA sequence that encodes the amino acids for translation. During translation, each of 61 trinucleotide codons are translated to one of 20 amino acids, leading to a degeneracy, or redundancy, in the genetic code. However, different cell types, and different animal species, utilize tRNAs (each bearing an anticodon) coding for the same amino acids at different frequencies. When a gene sequence contains codons that are infrequently represented by the corresponding tRNA, the ribosome translation machinery may slow, impeding efficient translation. Expression can be improved via “codon optimization” for a particular species, where the coding sequence is altered to encode the same protein sequence, but utilizing codons that are highly represented,

and/or utilized by highly expressed human proteins (Cid-Arregui et al., 2003; J. Virol. 77: 4928).

In some embodiments, the coding sequence of the transgene is modified to replace codons infrequently expressed in mammal or in primates with codons frequently expressed in primates. For example, in some embodiments, the transgene encodes a polypeptide having at least 85% sequence identity to a reference polypeptide (*e.g.* wild-type LAMP-2B; SEQ ID NO: 16)—for example, at least 90% sequence identity, at least 95% sequence identity, at least 98% identity, or at least 99% identity to the reference polypeptide—wherein at least one codon of the coding sequence has a higher tRNA frequency in humans than the corresponding codon in the sequence disclosed above or herein.

In an embodiment, the transgene comprises fewer alternative open reading frames than SEQ ID: 6. In an embodiment, the transgene is modified to enhance expression by termination or removal of open reading frames (ORFs) that do not encode the desired transgene. An open reading frame (ORF) is the nucleic acid sequence that follows a start codon and does not contain a stop codon. ORFs may be in the forward or reverse orientation, and may be “in frame” or “out of frame” compared with the gene of interest. Such open reading frames have the potential to be expressed in an expression cassette alongside the gene of interest, and could lead to undesired adverse effects. In some embodiments the transgene has been modified to remove open reading frames by further altering codon usage. This is done by eliminating one or more start codons (ATG, TTG, CTG) and/or introducing one or more stop codons (TAG, TAA, or TGA) in reverse orientation or out-of-frame to the desired ORF, while preserving the encoded amino acid sequence and, optionally, maintaining highly utilized codons in the gene of interest (*i.e.*, avoiding codons with frequency < 20%).

In variations of the present disclosure, the transgene coding sequence may be optimized by either of codon optimization and removal of non-transgene ORFs or using both techniques. In some cases, one removes or minimizes non-transgene ORFs after codon optimization in order to remove ORFs introduced during codon optimization.

In an embodiment, the transgene contains fewer CpG sites than SEQ ID: 6. Without being bound by theory, it is believed that the presence of CpG sites in a polynucleotide sequence is associated with the undesirable immunological responses of the host against a viral vector comprising the polynucleotide sequence. In some embodiments, the transgene is

designed to reduce the number of CpG sites. Exemplary methods are provided in U.S. Patent Application Publication No. US20020065236A1.

In an embodiment, the transgene contains fewer cryptic splice sites than SEQ ID: 6. For the optimization, GeneArt® software may be used, *e.g.*, to increase the GC content and/or remove cryptic splice sites in order to avoid transcriptional silencing and, therefore, increase transgene expression. Alternatively, any optimization method known in the art may be used. Removal of cryptic splice sites is described, for example, in International Patent Application Publication No. WO2004015106A1.

Also disclosed herein are expression cassettes and gene therapy vectors encoding LAMP-2B, *e.g.*, a codon-optimized LAMP-2B sequence disclosed herein, comprising: a consensus optimal Kozak sequence, a full-length polyadenylation (polyA) sequence (or substitution of full-length polyA for a truncated polyA), and minimal or no upstream (*i.e.* 5') start codons (*i.e.* ATG sites).

In some cases, the expression cassette contains two or more of a first inverted terminal repeat, an enhancer/promoter region, a consensus optimal Kozak sequence, a transgene (*e.g.*, a transgene encoding a LAMP-2B disclosed herein), a 3' untranslated region including a full-length polyA sequence, and a second inverted terminal repeat.

In an embodiment, the expression cassette comprises a Kozak sequence operatively linked to the transgene. In an embodiment, the Kozak sequence is a consensus optimal Kozak sequence comprising or consisting of SEQ ID NO: 20.

GCCGCCACCATGG (SEQ ID NO: 20)

In various embodiments, the expression cassette comprises an alternative Kozak sequence operatively linked to the transgene. In an embodiment, the Kozak sequence is an alternative Kozak sequence comprising or consisting of any one of SEQ ID NOs. 21-25.

(gcc)gccRccAUGG (SEQ ID NO: 21)

AGNNAUGN (SEQ ID NO: 22)

ANNAUGG (SEQ ID NO: 23)

ACCAUGG (SEQ ID NO: 24)

GACACCAUGG (SEQ ID NO: 25)

In SEQ ID NO: 21, a lower-case letter denotes the most common base at a position where the base can nevertheless vary; an upper-case letter indicates a highly conserved base; 'R' indicates adenine or guanine. In SEQ ID NO: 21, the sequence in parentheses (gcc) is optional. IN SEQ ID NOs: 22-23, 'N' denotes any base.

A variety of sequences can be used in place of this consensus optimal Kozak sequence as the translation-initiation site and it is within the skill of those in the art to identify and test other sequences. *See* Kozak M. An analysis of vertebrate mRNA sequences: intimations of translational control. *J. Cell Biol.* 115 (4): 887–903 (1991).

In an embodiment, the expression cassette comprises a full-length polyA sequence operatively linked to the transgene. In an embodiment, the full-length polyA sequence comprises SEQ ID NO: 26.

TGGCTAATAAAGGAAATTTATTTTCATTGCAATAGTGTGTTGGAATTTTTTGTGTC  
 TCTCACTCGGAAGGACATATGGGAGGGCAAATCATTAAAACATCAGAATGAGT  
 ATTTGGTTTAGAGTTTGGCAACATATGCCCATATGCTGGCTGCCATGAACAAAGG  
 TTGGCTATAAAGAGGTCATCAGTATATGAAACAGCCCCCTGCTGTCCATTCCTTA  
 TTCCATAGAAAAGCCTTGACTTGAGGTTAGATTTTTTTTATATTTTGTGTTTGTGTT  
 ATTTTTTTCTTTAACATCCCTAAAATTTTCCTTACATGTTTTACTAGCCAGATTTTT  
 CCTCCTCCTGACTACTCCAGTCATAGCTGTCCCTCTTCTCTTATGGAGATC  
 (SEQ ID NO: 26)

Various alternative polyA sequences may be used in expression cassettes of the present disclosure, including without limitation, bovine growth hormone polyadenylation signal (bGHpA) (SEQ ID NO: 27), the SV40 early/late polyadenylation signal (SEQ ID NO: 28), and human growth hormone (HGH) polyadenylation signal (SEQ ID NO: 29).

TCGACTGTGCCTTCTAGTTGCCAGCCATCTGTTGTTTGGCCCTCCCCCGTGCCTTC  
 CTTGACCCTGGAAGGTGCCACTCCACTGTCCTTTCCTAATAAAAATGAGGAAATT  
 GCATCGCATTGTCTGAGTAGGTGTCATTCTATTCTGGGGGGTGGGGTGGGGCAGG  
 ACAGCAAGGGGGAGGATTGGGAGGACAATAGCAGGCATGCTGGGGATGCGGTG  
 GGCTCTATGGCTTCTG (SEQ ID NO: 27)

CAGACATGATAAGATACATTGATGAGTTTGGACAAACCACAACACTAGAATGCAGT  
 GAAAAAATGCTTTATTTGTGAAATTTGTGATGCTATTGCTTTATTTGTAACCATT  
 ATAAGCTGCAATAAACAAGTTAACAACAACAATTGCATTCATTTTATGTTTCAGG

TTCAGGGGGAGATGTGGGAGGTTTTTTAAAGCAAGTAAAACCTCTACAAATGTG  
GTA (SEQ ID NO: 28)

CTGCCCGGGTGGCATCCCTGTGACCCCTCCCCAGTGCCTCTCCTGGCCCTGGAAG  
TTGCCACTCCAGTGCCCACCAGCCTTGTCCTAATAAAATTAAGTTGCATCATTTTG  
TCTGACTAGGTGTCCTTCTATAATATTATGGGGTGGAGGGGGGTGGTATGGAGCA  
AGGGGCCCAAGTTGGGAAGAAACCTGTAGGGCCTGC (SEQ ID NO: 29)

In some embodiments, the expression cassette comprises an active fragment of a polyA sequence. In particular embodiments, the active fragment of the polyA sequence comprises or consists of less than 20 base pair (bp), less than 50 bp, less than 100 bp, or less than 150 bp, *e.g.*, of any of the polyA sequences disclosed herein.

In some cases, expression of the transgene is increased by ensuring that the expression cassette does not contain competing ORFs. In an embodiment, the expression cassette comprises no start codon within 20, 30, 40, 50, 60, 70, 80, 90, 100, 200, or 300 base pairs 5' of the start codon of the transgene. In an embodiment, the expression cassette comprises no start codon 5' of the start codon of the transgene. In some embodiments, the expression cassette comprises no alternative transcripts. In some embodiments, the expression cassette comprises no alternative transcripts, except small transcripts, *e.g.* 300 base pairs or less.

In an embodiment, the expression cassette comprises operatively linked, in the 5' to 3' direction, a first inverted terminal repeat, an enhancer/promoter region, introns, a consensus optimal Kozak sequence, the transgene, a 3' untranslated region including a full-length polyA sequence, and a second inverted terminal repeat, where the expression cassette comprises no start codon 5' to the start codon of the transgene.

In an embodiment, the enhancer/promoter region comprises, in the 5' to 3' direction: a CMV IE Enhancer and a Chicken Beta-Actin Promoter. In an embodiment, the enhancer/promoter region comprises a CAG promoter. As used herein "CAG promoter" refers to a polynucleotide sequence comprising a CMV early enhancer element, a chicken beta-actin promoter, the first exon and first intron of the chicken beta-actin gene, and a splice acceptor from the rabbit beta-globin gene.

In an embodiment, the expression cassette shares at least 95% identity to a sequence selected from SEQ ID NOs: 10-12. In an embodiment, the expression cassette shares complete identity to a sequence selected from SEQ ID NOs: 10-12, or shares at least 80%, at least 85%, at least 90%, at least 95%, at least 98%, or at least 99% identity to a sequence

selected from SEQ ID NOs: 10-12. In certain embodiments, the expression cassette comprises one or more modifications as compared to a sequence selected from SEQ ID NOs: 10-12. In particular embodiments, the one or more modifications comprises one or more of: removal of one or more (*e.g.*, all) upstream ATG sequences, replacement of the Kozak sequence with an optimized consensus Kozak sequence or another Kozak sequence, including but not limited to any of those disclosed herein, and/or replacement of the polyadenylation sequence with a full-length polyadenylation sequence or another polyadenylation sequence, including but not limited to any of those disclosed herein. An illustrative configuration of genetic elements within these exemplary expression cassettes is depicted in **FIG. 1**.

CTGCGCGCTCGCTCGCTCACTGAGGCCGCCCGGGCAAAGCCCGGGCGTCGGGCG  
ACCTTTGGTCGCCCCGGCCTCAGTGAGCGAGCGAGCGCGCAGAGAGGGAGTGGCC  
AACTCCATCACTAGGGGTTCCCTGTAGTTAATGATTAACCCGCCATGCTACTTAT  
CTACCAGGGTAATGGGGATCCTCTAGAACTATAGCTAGTCGACATTGATTATTGA  
CTAGTTATTAATAGTAATCAATTACGGGGTCATTAGTTCATAGCCCATATATGGA  
GTTCCGCGTTACATAACTTACGGTAAATGGCCCGCCTGGCTGACCGCCCAACGAC  
CCCCGCCATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGA  
CTTTCATTGACGTCAATGGGTGGAGTATTTACGGTAAACTGCCCACTTGGCAGT  
ACATCAAGTGTATCATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAA  
TGGCCCGCCTGGCATTATGCCCAGTACATGACCTTATGGGACTTTCCTACTTGGC  
AGTACATCTACGTATTAGTCATCGCTATTACCATGGTCGAGGTGAGCCCCACGTT  
CTGCTTCACTCTCCCCATCTCCCCCCCCCTCCCCACCCCAATTTTGTATTTATTTAT  
TTTTTAATTATTTTGTGCAGCGATGGGGGCGGGGGGGGGGGGGGGGGGGCGCGCGCC  
AGGCGGGGCGGGGCGGGGCGAGGGGCGGGGCGGGGCGAGGCGGAGAGGTGCG  
GCGGCAGCCAATCAGAGCGGCGCGCTCCGAAAGTTTCCTTTTATGGCGAGGCGG  
CGGCGGCGGCGGCCCTATAAAAAGCGAAGCGCGCGGGCGGGGAGTCGCTGC  
GCGCTGCCTTCGCCCCGTGCCCGCTCCGCCGCCGCTCGCGCCGCCCGCCCCGG  
CTCTGACTGACCGGTTACTCCACAGGTGAGCGGGCGGGACGGCCCTTCTCCTC  
CGGGCTGTAATTAGCGCTTGGTTTAATGACGGCTTGTTCCTTTCTGTGGCTGCGT  
GAAAGCCTTGAGGGGCTCCGGGAGGGCCCTTGTGCGGGGGGAGCGGCTCGGGG  
GGTGCCTGCGTGTGTGTGTGCGTGGGGAGCGCCGCGTGC GGCTCCGCGCTGCC  
GGCGGCTGTGAGCGCTGCGGGCGCGGCGCGGGGCTTTGTGCGCTCCGCAGTGTG  
CGCGAGGGGAGCGCGGCCGGGGGCGGTGCCCGCGGTGCGGGGGGGGCTGCGA  
GGGGAACAAAGGCTGCGTGC GGGGTGTGTGCGTGGGGGGGTGAGCAGGGGGTG

TGGGCGCGTCGGTCGGGCTGCAACCCCCCTGCACCCCCCTCCCCGAGTTGCTGA  
GCACGGCCCCGGCTTCGGGTGCGGGGCTCCGTACGGGGCGTGGCGCGGGGCTCGC  
CGTGCCGGGCGGGGGGTGGCGGCAGGTGGGGGTGCCGGGCGGGGCGGGGCCGC  
CTCGGGCCGGGGAGGGCTCGGGGGAGGGGCGCGGGCGGCCCCCGGAGCGCCGGC  
GGCTGTCGAGGCGCGGCGAGCCGCAGCCATTGCCTTTTATGGTAATCGTGCGAG  
AGGGCGCAGGGACTTCCTTTGTCCCAAATCTGTGCGGAGCCGAAATCTGGGAGG  
CGCCGCCGCACCCCCTCTAGCGGGCGCGGGGCGAAGCGGTGCGGCGCCGGCAGG  
AAGGAAATGGGCGGGGAGGGCCTTCGTGCGTCGCCGCGCCCGCTCCCCTTCTC  
CCTCTCCAGCCTCGGGGCTGTCCGCGGGGGGACGGCTGCCTTCGGGGGGGACGG  
GGCAGGGCGGGGTTCGGCTTCTGGCGTGTGACCGGCGGCTCTAGAGCCTCTGCTA  
ACCATGTTTCATGCCTTCTTCTTTTTCTACAGCTCCTGGGCAACGTGCTGGTTATT  
GTGCTGTCTCATCATTTTGGCAAAGAATTCGAGCGGCCGCCAGCCGCCACCATGG  
TCTGCTTCAGACTGTTCCCTGTCCCTGGATCTGGTCTGGTGCTTGTGTGCTTGGTG  
CTGGGTGCTGTGAGATCCTATGCCCTTGAGCTGAACCTGACTGACTCAGAAAATG  
CCACTTGCCTGTATGCCAAGTGGCAGATGAACTTCACTGTGAGATATGAGACTAC  
CAACAAGACCTACAAGACTGTGACCATCTCAGACCATGGCACTGTCACCTACAA  
TGGATCAATCTGTGGTGATGATCAGAATGGCCCAAAGATAGCAGTGCAGTTTGG  
GCCCGGTTTTTCTGGATTGCTAACTTCACCAAGGCAGCCTCCACCTACAGCATT  
GACTCAGTCAGCTTCAGCTACAACACTGGGGATAACACCACCTTCCCTGACGCAG  
AGGACAAGGGAATCCTTACTGTGGACGAACTCCTGGCAATCAGAATCCCCCTTA  
ACGACCTGTTTCAGATGCAACTCCCTTTCAACCCTTGAAAAGAATGATGTGGTGCA  
ACACTATTGGGACGTCCCTGGTGCAAGCCTTTGTGCAGAATGGGACAGTGAGTAC  
CAACGAGTTCCTCTGTGACAAGGACAAGACCAGCACTGTGGCCCCACTATCCA  
CACCCTGTGCCAGCCCTACCACTACCCCCACCCCTAAAGAGAAGCCAGAAGC  
TGGAACCTACTCAGTCAACAATGGAAATGACACATGCCTCCTTGCCACCATGGG  
ACTGCAGCTGAACATCACTCAGGACAAGGTGGCCTCAGTGATTAACATCAACCC  
TAACACCACTCATAGCACTGGGAGCTGCAGATCACATACAGCTCTGCTGAGGCTC  
AACTCCTCCACCATCAAGTACCTGGACTTTGTGTTTGCTGTGAAGAATGAGAACA  
GGTTCTACCTCAAGGAAGTGAACATTTCCATGTACCTGGTCAATGGTTCAGTGTT  
CTCTATTGCCAACAACAATCTGAGCTACTGGGATGCACCCCTGGGATCCTCCTAC  
ATGTGCAACAAGGAGCAGACTGTGAGTGTGTCAGGTGCTTTTCAGATCAACACTT  
TTGACCTGAGGGTGCAGCCCTTCAATGTGACTCAGGGAAAGTACTCCACTGCACA  
AGAGTGTTCCCTTGGATGATGACACTATCCTCATCCCCATTATTGTGGGAGCTGGA  
CTGTCAGGATTGATTATAGTGATTGTGATTGCTTATGTGATTGGAAGGAGAAAGA

GCTATGCTGGCTACCAGACCCTGTAAAAGGGCGAATTCCAGCACACGCGTCCTA  
GGAGCTCGAGTACTACTGGCGGCCGTTACTAGTGGATCCGCGGTACAAGTAAGC  
ATGCAAGCTTCGAGGACGGGGTGAACCTACGCCTGAATCAAGCTTATCGATAAAT  
TCGAGCATCTTACCGCCATTTATTCCCATATTTGTTCTGTTTTTCTTGATTTGGGTA  
TACATTTAAATGTTAATAAAAACAAAATGGTGGGGCAATCATTTACATTTTTAGGG  
ATATGTAATTACTAGTTCAGGTGTATTGCCACAAGACAAACATGTTAAGAACTT  
TCCCGTTATTTACGCTCTGTTCCCTGTTAATCAACCTCTGGATTACAAAATTTGTGA  
AAGATTGACTGATATTCTTAACTATGTTGCTCCTTTTACGCTGTGTGGATATGCTG  
CTTTAATGCCTCTGTATCATGCTATTGCTTCCCGTACGGCTTTCGTTTTCTCCTCCT  
TGTATAAATCCTGGTTGCTGTCTCTTTATGAGGAGTTGTGGCCCGTTGTCCGTCAA  
CGTGGCGTGGTGTGCTCTGTGTTTGCTGACGCAACCCCCACTGGCTGGGGCATTG  
CCACCACCTGTCAACTCCTTTCTGGGACTTTCGCTTTCCCCCTCCCGATCGCCACG  
GCAGAACTCATCGCCGCCTGCCTTGCCCGCTGCTGGACAGGGGCTAGGTTGCTGG  
GCACTGATAATTCCGTGGTGTGTCGGGGAAGGGCCTCGATACCGTCGATATCGA  
TCCTGGCTAATAAAGGAAATTTATTTTCATTGCAATAGTGTGTTGGAATTTTTTGT  
GTCTCTCACTCGGAAGGACATATGGGAGGGCAAATCATTTAAAACATCAGAATG  
AGTATTTGGTTTAGAGTTTGGCAACATATGCCCATATGCTGGCTGCCATGAACAA  
AGGTTGGCTATAAAGAGGTCATCAGTATATGAAACAGCCCCCTGCTGTCCATTCC  
TTATTCATAGAAAAGCCTTGACTTGAGGTTAGATTTTTTTTTATATTTTGT  
GTTATTTTTTTCTTTAACATCCCTAAAATTTTCCTTACATGTTTTACTAGCCAGATT  
TTTCTCCTCTCCTGACTACTCCAGTCATAGCTGTCCCTCTTCTCTTATGGAGAT  
CGAAGCAATTCGTTGATCTGAATTCGACCACCATAATAGATCTCCCATTACCC  
TGGTAGATAAGTAGCATGGCGGGTTAATCATTAACACTACAAGGAACCCCTAGTGA  
TGGAGTTGGCCACTCCCTCTCTGCGCGCTCGCTCGCTCACTGAGGCCGGGCGACC  
AAAGGTCGCCCAGCCCCGGGCTTTGCCCGGGCGGCCTCAGTGAGCGAGCGAGC  
GCGCAG (SEQ ID NO: 10)

CTGCGCGCTCGCTCGCTCACTGAGGCCGCCGGGCAAAGCCCCGGGCGTCGGGCG  
ACCTTTGGTCGCCC GGCTCAGTGAGCGAGCGAGCGCGCAGAGGGGAGTGGCC  
AACTCCATCACTAGGGGTTCCCTGTAGTTAATGATTAACCCGCCATGCTACTTAT  
CTACCAGGGTAATGGGGATCCTCTAGA ACTATAGCTAGTCGACATTGATTATTGA  
CTAGTTATTAATAGTAATCAATTACGGGGTCATTAGTTCATAGCCCATATATGGA  
GTTCCGCGTTACATAACTTACGGTAAATGGCCCCGCTGGCTGACCGCCCAACGAC  
CCCCGCCCATGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGA

CTTTCCATTGACGTCAATGGGTGGAGTATTTACGGTAAACTGCCCACTTGGCAGT  
ACATCAAGTGTATCATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAA  
TGGCCCGCCTGGCATTATGCCCAGTACATGACCTTATGGGACTTTCCTACTTGGC  
AGTACATCTACGTATTAGTCATCGCTATTACCATGGTCGAGGTGAGCCCCACGTT  
CTGCTTCACTCTCCCCATCTCCCCCCCCCTCCCCACCCCCAATTTTGTATTTATTTAT  
TTTTTAATTATTTTGTGCAGCGATGGGGGCGGGGGGGGGGGGGGGGGGGCGCGCGCC  
AGGCGGGGCGGGGCGGGGCGAGGGGCGGGGCGGGGCGAGGCGGAGAGGTGCG  
GCGGCAGCCAATCAGAGCGGCGCGCTCCGAAAGTTTCCTTTTATGGCGAGGCGG  
CGGCGGCGGCGGCCCTATAAAAAGCGAAGCGCGCGGCGGGCGGGAGTCGCTGC  
GCGCTGCCTTCGCCCCGTGCCCGCTCCGCCGCCGCTCGCGCCGCCCGCCCCGG  
CTCTGACTGACCGGTTACTCCCACAGGTGAGCGGGCGGGACGGCCCTTCTCCTC  
CGGGCTGTAATTAGCGCTTGGTTTAATGACGGCTTGTTCCTTTCTGTGGCTGCGT  
GAAAGCCTTGAGGGGCTCCGGGAGGGCCCTTTGTGCGGGGGGAGCGGCTCGGGG  
GGTGCCTGCGTGTGTGTGTGCGTGGGGAGCGCCGCGTGC GGCTCCGCGCTGCC  
GGCGGCTGTGAGCGCTGCGGGCGCGGCGCGGGGCTTTGTGCGCTCCGCAGTGTG  
CGCGAGGGGAGCGCGGCCGGGGGCGGTGCCCGCGGTGCGGGGGGGGCTGCGA  
GGGGAACAAAGGCTGCGTGC GGGGTGTGTGCGTGGGGGGGTGAGCAGGGGGTG  
TGGGCGCGTCGGTCGGGCTGCAACCCCCCTGCACCCCCCTCCCCGAGTTGCTGA  
GCACGGCCCGGCTTCGGGTGCGGGGCTCCGTACGGGGCGTGGCGCGGGGCTCGC  
CGTGCCGGGCGGGGGGTGGCGGCAGGTGGGGGTGCCGGGCGGGGCGGGGCCGC  
CTCGGGCCGGGGAGGGCTCGGGGGAGGGGCGCGGCGGCCCCCCGGAGCGCCGGC  
GGCTGTCGAGGCGCGGCGAGCCGCAGCCATTGCCTTTTATGGTAATCGTGCGAG  
AGGGCGCAGGGACTTCCTTTGTCCCAAATCTGTGCGGAGCCGAAATCTGGGAGG  
CGCCGCCGCACCCCCCTTAGCGGGCGCGGGGCGAAGCGGTGCGGCGCCGGCAGG  
AAGGAAATGGGCGGGGAGGGCCTTCGTGCGTCGCCGCGCCGCGCTCCCCTTCTC  
CCTCTCCAGCCTCGGGGCTGTCCGCGGGGGGACGGCTGCCTTCGGGGGGGACGG  
GGCAGGGGCGGGGTTCGGCTTCTGGCGTGTGACCGGCGGCTCTAGAGCCTCTGCTA  
ACCATGTTTCATGCCTTCTTCTTTTTCTACAGCTCCTGGGCAACGTGCTGGTTATT  
GTGCTGTCTCATCATTTTGGCAAAGAATTCGAGCGGCCGCCAGCCGCCACCATGG  
TGTGCTTTAGACTGTTTCCTGTGCCTGGTTCAGGGCTGGTCTGCTGTCTGGTG  
CTGGGGGCTGTCAGAAGCTATGCCTTGGAGCTGAACCTCACTGATAGTGAAAAT  
GCCACTTGTCTGTATGCTAAGTGGCAGATGAACTTCACTGTGAGATATGAAACCA  
CCAACAAGACTTACAAAACAGTGACCATCTCAGATCATGGAAGTGTGACCTACA  
ACGGCAGCATTTGTGGAGACGACCAGAACGGACCAAAAATCGCTGTCCAATTTG

GGCCTGGATTCTCCTGGATTGCCAATTTCACTAAAGCTGCCTCCACATATTCAATT  
GACTCAGTGTCTTCTCCTACAACACTGGGGACAACACTACTTTCCCTGATGCTG  
AAGATAAGGGAATCTTGACAGTGGATGAGCTGCTGGCTATCAGGATCCCTTTGA  
ATGACCTGTTTAGGTGTAATTCCTGAGCACTCTGGAGAAGAACGACGTGGTGC  
AGCACTACTGGGACGTGCTGGTGCAGGCCTTTGTGCAGAACGGCACTGTGTCCAC  
CAACGAATTCCTGTGTGATAAGGACAAAACCTTCCACTGTGGCACCTACAATTCAC  
ACTACTGTGCCTTCACCTACCACCACTCCAACCTCAAAGGAAAAGCCTGAAGCA  
GGAACCTACTCTGTGAACAATGGCAATGATACCTGTCTGTTGGCCACCATGGGCC  
TCCAACCTGAACATTACTCAGGACAAGGTGGCCTCAGTGATTAACATTAACCCCAA  
CACTACCCACTCCACTGGCAGCTGTAGATCACACACAGCCTTGCTCAGACTGAAT  
AGCAGCACCATCAAGTATTTGGATTTTGTGTTTGCAGTGAAGAATGAAAACAGGT  
TCTACCTGAAGGAAGTCAACATCTCAATGTACCTGGTGAACGGCTCAGTGTTTCAG  
CATTGCCAACAAACCTCTCCTATTGGGACGCTCCACTGGGGAGCAGCTACATG  
TGTAACAAGGAACAGACTGTGTCAGTGTGAGGAGCCTTCCAGATTAACACCTTTG  
ATCTGAGGGTCCAACCCTTTAATGTCCTCAAGGAAAGTATAGCACTGCCCAGG  
AGTGCTCCCTGGATGATGACACCATTCTGATTCCAATCATTGTGGGTGCAGGACT  
TTCTGGGCTTATTATTGTGATTGTGATTGCCTATGTGATTGGCAGAAGGAAATCCT  
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GCTCGAGTACTACTGGCGGCCGTTACTAGTGGATCCGCGGTACAAGTAAGCATG  
CAAGCTTCGAGGACGGGGTGAACCTACGCTGAATCAAGCTTATCGATAAATTCG  
AGCATCTTACCGCCATTTATTCCCATATTTGTTCTGTTTTTCTTGATTTGGGTATAC  
ATTTAAATGTTAATAAAAACAAAATGGTGGGGCAATCATTACATTTTTAGGGATA  
TGTAATTACTAGTTCAGGTGTATTGCCACAAGACAAACATGTTAAGAACTTTCC  
CGTTATTTACGCTCTGTTCCCTGTTAATCAACCTCTGGATTACAAAATTTGTGAAAG  
ATTGACTGATATTCTTAACCTATGTTGCTCCTTTTACGCTGTGTGGATATGCTGCTT  
TAATGCCTCTGTATCATGCTATTGCTTCCCGTACGGCTTTCGTTTTCTCCTCCTTGT  
ATAAATCCTGGTTGCTGTCTCTTTATGAGGAGTTGTGGCCCGTTGTCCGTCAACGT  
GGCGTGGTGTGCTCTGTGTTTGCTGACGCAACCCCCACTGGCTGGGGCATTGCCA  
CCACCTGTCAACTCCTTTCTGGGACTTTCGCTTTCCCCCTCCCGATCGCCACGGCA  
GAACTCATCGCCGCTGCCTTGCCCGCTGCTGGACAGGGGCTAGGTTGCTGGGGCA  
CTGATAATTCCGTGGTGTGTCGGGGAAGGGCCTCGATACCGTCGATATCGATCC  
TGGCTAATAAAGGAAATTTATTTTCATTGCAATAGTGTGTTGGAATTTTTTGTGTC  
TCTCACTCGGAAGGACATATGGGAGGGCAAATCATTAAAACATCAGAATGAGT  
ATTTGGTTTAGAGTTTGGCAACATATGCCCATATGCTGGCTGCCATGAACAAAGG

TTGGCTATAAAGAGGTCATCAGTATATGAAACAGCCCCCTGCTGTCCATTCCTTA  
TTCATAGAAAAGCCTTGACTTGAGGTTAGATTTTTTTTATATTTTGTGTTTGTGTT  
ATTTTTTTCTTTAACATCCCTAAAATTTTCCTTACATGTTTTACTAGCCAGATTTTT  
CCTCCTCTCCTGACTACTCCCAGTCATAGCTGTCCCTCTTCTCTTATGGAGATCGA  
AGCAATTCGTTGATCTGAATTCGACCACCATAATAGATCTCCCATTACCCTGG  
TAGATAAGTAGCATGGCGGGTTAATCATTAACTACAAGGAACCCCTAGTGATGG  
AGTTGGCCACTCCCTCTCTGCGCGCTCGCTCGCTCACTGAGGCCGGGCGACCAA  
GGTCGCCCCGACGCCCGGGCTTTGCCCGGGCGGCCTCAGTGAGCGAGCGAGCGCG  
CAG (SEQ ID NO: 11)

CTGCGCGCTCGCTCGCTCACTGAGGCCGCCCGGGCAAAGCCCCGGGCGTCGGGCG  
ACCTTTGGTCGCCCCGGCCTCAGTGAGCGAGCGAGCGCGCAGAGAGGGAGTGGCC  
AACTCCATCACTAGGGGTTCCCTGTAGTTAATGATTAACCCGCCATGCTACTTAT  
CTACCAGGGTAATGGGGATCCTCTAGAACTATAGCTAGTCGACATTGATTATTGA  
CTAGTTATTAATAGTAATCAATTACGGGGTCATTAGTTCATAGCCCATATATGGA  
GTTCCGCGTTACATAACTTACGGTAAATGGCCCCGCCTGGCTGACCGCCCAACGAC  
CCCCGCCATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGA  
CTTTCCATTGACGTCAATGGGTGGAGTATTTACGGTAAACTGCCCACTTGGCAGT  
ACATCAAGTGTATCATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAA  
TGGCCCGCCTGGCATTATGCCCAGTACATGACCTTATGGGACTTTCCTACTTGGC  
AGTACATCTACGTATTAGTCATCGCTATTACCATGGTCGAGGTGAGCCCCACGTT  
CTGCTTCACTCTCCCCATCTCCCCCCCCCTCCCCACCCCAATTTTGTATTTATTTAT  
TTTTTAATTATTTTGTGCAGCGATGGGGGCGGGGGGGGGGGGGGGGGGGCGCGCGCC  
AGGCGGGGCGGGGCGGGGCGAGGGGCGGGGCGGGGCGAGGCGGAGAGGTGCG  
GCGGCAGCCAATCAGAGCGGCGCGCTCCGAAAGTTTCCTTTTATGGCGAGGCGG  
CGGCGGCGGCGGCCCTATAAAAAGCGAAGCGCGCGGGCGGGGAGTCGCTGC  
GCGCTGCCTTCGCCCCGTGCCCGCTCCGCCGCCGCTCGCGCCGCCCGCCCCGG  
CTCTGACTGACCGGTTACTCCACAGGTGAGCGGGCGGGACGGCCCTTCTCCTC  
CGGGCTGTAATTAGCGCTTGGTTTAATGACGGCTTGTCTTTCTGTGGCTGCGT  
GAAAGCCTTGAGGGGCTCCGGGAGGGCCCTTTGTGCGGGGGGAGCGGCTCGGGG  
GGTGCCTGCGTGTGTGTGTGCGTGGGGAGCGCCGCGTGC GGCTCCGCGCTGCC  
GGCGGCTGTGAGCGCTGCGGGCGCGGCGCGGGGCTTTGTGCGCTCCGCAGTGTG  
CGCGAGGGGAGCGCGGCCGGGGGCGGTGCCCGCGGTGCGGGGGGGGCTGCGA  
GGGGAACAAAGGCTGCGTGC GGGGTGTGTGCGTGGGGGGGTGAGCAGGGGGTG

TGGGCGCGTCGGTCGGGCTGCAACCCCCCTGCACCCCCCTCCCCGAGTTGCTGA  
GCACGGCCCGGCTTCGGGTGCGGGGCTCCGTACGGGGCGTGGCGCGGGGCTCGC  
CGTGCCGGGCGGGGGGTGGCGGCAGGTGGGGGTGCCGGGCGGGGCGGGGCCGC  
CTCGGGCCGGGGAGGGCTCGGGGGAGGGGCGCGGGCGGCCCCCGGAGCGCCGGC  
GGCTGTTCGAGGCGCGGCGAGCCGCAGCCATTGCCTTTTATGGTAATCGTGCGAG  
AGGGCGCAGGGACTTCCTTTGTCCAAATCTGTGCGGAGCCGAAATCTGGGAGG  
CGCCGCCGCACCCCCTCTAGCGGGCGCGGGGCGAAGCGGTGCGGCGCCGGCAGG  
AAGGAAATGGGCGGGGAGGGCCTTCGTGCGTCGCCGCGCCCGCCGTCCCCTTCTC  
CCTCTCCAGCCTCGGGGCTGTCCGCGGGGGGACGGCTGCCTTCGGGGGGGACGG  
GGCAGGGCGGGGTTCGGCTTCTGGCGTGTGACCGGCGGCTCTAGAGCCTCTGCTA  
ACCATGTTTCATGCCTTCTTCTTTTTCTACAGCTCCTGGGCAACGTGCTGGTTATT  
GTGCTGTCTCATCATTTTGGCAAAGAATTCGAGCGGCCGCCAGCCGCCACCATGG  
TCTGTTTTAGGCTGTTCCCTGTCCCTGGTTCAGGACTGGTCTTAGTGTGTCTGGTG  
CTTGAGCTGTCAGAAGCTATGCCCTGGAGCTGAACCTGACTGACTCAGAAAAT  
GCCACTTGCCTGTATGCCAAGTGGCAGATGAACTTCACTGTCAGATATGAAACCA  
CCAACAAGACCTATAAGACTGTGACCATCTCAGACCATGGCACTGTGACTTACA  
ATGGGTCAATTTGTGGAGATGACCAGAATGGCCCTAAGATAGCTGTCCAGTTTGG  
TCCAGGATTCAGCTGGATTGCCAACTTCACCAAGGCAGCCAGCACCTACAGCATT  
GACTCTGTGTCCTTCTCCTACAACACAGGAGACAACACCACTTTCCCTGATGCAG  
AGGACAAAGGTATCCTGACTGTGGATGAGTTGCTGGCAATCAGGATCCCCTGA  
ACGATCTGTTTCAGGTGCAACTCACTGTCCACTCTGGAAAAGAATGATGTGGTGCA  
GCACTATTGGGATGTGCTAGTCCAGGCCTTTGTCCAGAATGGGACTGTGTCAACT  
AATGAGTTCCTGTGTGACAAGGACAAGACAAGCACTGTAGCCCCACTATCCAT  
ACCACAGTACCTAGCCCCACCACTACTCCAACCCCCAAGGAGAAGCCTGAGGCT  
GGCACCTACTCAGTGAACAATGGGAATGACACCTGTTTGCTGGCCACTATGGGA  
CTCCAACCTGAACATCACCCAGGACAAAGTGGCCTCTGTGATCAATATCAATCCCA  
ACACCACCCACAGCACTGGGTCCTGCAGAAGCCACACTGCCCTCCTGAGGCTCA  
ACTCATCAACTATCAAGTACTTGGATTTTGTGTTTGCAGTGAAGAATGAGAACAG  
ATTCTACCTCAAAGAGGTCAACATTTCAATGTACCTGGTGAATGGGAGTGTGTTC  
TCCATTGCTAACAACAACCTGAGCTACTGGGATGCCCTCTGGGCTCCTCATA  
TGTGCAACAAGGAACAGACTGTGAGTGTGTCAGGGGCCTTCCAGATCAACACTT  
TTGACCTGAGAGTGCAGCCCTTTAATGTGACACAGGGAAAGTACAGCACTGCTC  
AGGAGTGCAGCCTGGATGATGACACTATCCTGATCCCTATCATTGTGGGGGCAG  
GCCTGTCTGGACTCATTATTGTGATTGTGATTGCCTATGTGATAGGGAGAAGGAA

GTCTTATGCTGGATACCAGACCCTGTAAAAGGGCGAATTCCAGCACACGCGTCCT  
 AGGAGCTCGAGTACTACTGGCGGCCGTTACTAGTGGATCCGCGGTACAAGTAAG  
 CATGCAAGCTTCGAGGACGGGGTGAACACTACGCCTGAATCAAGCTTATCGATAAA  
 TTCGAGCATCTTACCGCCATTTATTCCCATATTTGTTCTGTTTTTCTTGATTTGGGT  
 ATACATTTAAATGTTAATAAAAACAAAATGGTGGGGCAATCATTTACATTTTTAGG  
 GATATGTAATTACTAGTTCAGGTGTATTGCCACAAGACAAACATGTTAAGAACT  
 TTCCCGTTATTTACGCTCTGTTCCCTGTTAATCAACCTCTGGATTACAAAATTTGTG  
 AAAGATTGACTGATATTTCTTAACTATGTTGCTCCTTTTACGCTGTGTGGATATGCT  
 GCTTTAATGCCTCTGTATCATGCTATTGCTTCCCGTACGGCTTTCGTTTTCTCCTCC  
 TTGTATAAATCCTGGTTGCTGTCTCTTTATGAGGAGTTGTGGCCCGTTGTCCGTCA  
 ACGTGGCGTGGTGTGCTCTGTGTTTGCTGACGCAACCCCCACTGGCTGGGGCATT  
 GCCACCACCTGTCAACTCCTTTCTGGGACTTTCGCTTTCCCCCTCCCGATCGCCAC  
 GGCAGAACTCATCGCCGCTGCCTTGCCCGCTGCTGGACAGGGGCTAGGTTGCTG  
 GGCAGTATAATTCCGTGGTGTGTCGGGGAAGGGCCTCGATACCGTCGATATCG  
 ATCCTGGCTAATAAAGGAAATTTATTTTCATTGCAATAGTGTGTTGGAATTTTTTG  
 TGTCTCTCACTCGGAAGGACATATGGGAGGGCAAATCATTTAAAACATCAGAAT  
 GAGTATTTGGTTTAGAGTTTGGCAACATATGCCCATATGCTGGCTGCCATGAACA  
 AAGGTTGGCTATAAAGAGGTCATCAGTATATGAAACAGCCCCCTGCTGTCCATTC  
 CTTATTCATAGAAAAGCCTTGACTTGAGGTTAGATTTTTTTTTATATTTTGTTTTGT  
 GTTATTTTTTTCTTTAACATCCCTAAAATTTTCCTTACATGTTTTACTAGCCAGATT  
 TTTCTCCTCTCCTGACTACTCCAGTCATAGCTGTCCCTCTTCTCTTATGGAGAT  
 CGAAGCAATTCGTTGATCTGAATTCGACCACCATAATAGATCTCCCATACCC  
 TGGTAGATAAGTAGCATGGCGGGTTAATCATTAACAAGGAACCCCTAGTGA  
 TGGAGTTGGCCACTCCCTCTCTGCGCGCTCGCTCGCTCACTGAGGCCGGGCGACC  
 AAAGGTCGCCCAGCCCCGGGCTTTGCCCGGGCGGCCTCAGTGAGCGAGCGAGC  
 GCGCAG (SEQ ID NO: 12).

In an embodiment, the vector is an adeno-associated virus (AAV) vector. In an embodiment, the expression cassette comprises ITR sequences selected from SEQ ID NOs: 13 and 14.

CTGCGCGCTCGCTCGCTCACTGAGGCCGCCCGGGCAAAGCCCCGGGCGTCGGGCG  
 ACCTTTGGTCGCCCCGGCCTCAGTGAGCGAGCGAGCGCGCAGAGAGGGAGTGGCC  
 AACTCCATCACTAGGGGTTCCCT (SEQ ID NO: 13)

AGGAACCCCTAGTGATGGAGTTGGCCACTCCCTCTCTGCGCGCTCGCTCGCTCAC  
 TGAGGCCGGGCGACCAAAGGTCGCCCCGACGCCCGGGCTTTGCCCGGGCGGCCTC  
 AGTGAGCGAGCGAGCGCGCAG (SEQ ID NO: 14)

In related embodiments, the disclosure provides gene therapy vectors comprising an expression cassette disclosed herein. Generally, the gene therapy vectors described herein comprise an expression cassette comprising a polynucleotide encoding one or more isoforms of lysosome-associated membrane protein 2 (LAMP-2), that allows for the expression of LAMP-2 to partially or wholly rectify deficient LAMP-2 protein expression levels and/or autophagic flux in a subject in need thereof (*e.g.*, a subject having Danon disease or another disorder characterized by deficient autophagic flux at least in part due to deficient LAMP-2 expression).

LAMP-2A protein sequence

MVCFRLFPVPGSGLVLVCLVLGAVRSYALELNLTDSENATCLYAKWQMNFTVRYET  
 TNKTYKTVTISDHGTVTYNGSICGDDQNGPKIAVQFGPGFSWIANFTKAASTYSIDSV  
 SFSYNTGDNTTFPDAEDKGILTVDELLAIRIPLNDFRCNSLSTLEKNDVVQHYWDVL  
 VQAFVQNGTVSTNEFLCDKDKTSTVAPTIHTTVPSPTTTPKPEAGTYSVNNGN  
 DTCLLATMGLQLNITQDKVASVININPNTTHSTGSCRSHALLRLNSSTIKYLDFVFA  
 VKNENRFYLKEVNISMVYLVNGSVFSIANNLSYWDAPLGSSYMCNKEQTVSVSGAF  
 QINTFDLRVQPFNVTQGKYSTAQDCSADDDNFLVPIAVGAALAGVLILVLLAYFIGL  
 KHHHAGYEQF (SEQ ID NO: 15)

LAMP-2B protein sequence

MVCFRLFPVPGSGLVLVCLVLGAVRSYALELNLTDSENATCLYAKWQMNFTVRYET  
 TNKTYKTVTISDHGTVTYNGSICGDDQNGPKIAVQFGPGFSWIANFTKAASTYSIDSV  
 SFSYNTGDNTTFPDAEDKGILTVDELLAIRIPLNDFRCNSLSTLEKNDVVQHYWDVL  
 VQAFVQNGTVSTNEFLCDKDKTSTVAPTIHTTVPSPTTTPKPEAGTYSVNNGN  
 DTCLLATMGLQLNITQDKVASVININPNTTHSTGSCRSHALLRLNSSTIKYLDFVFA  
 VKNENRFYLKEVNISMVYLVNGSVFSIANNLSYWDAPLGSSYMCNKEQTVSVSGAF  
 QINTFDLRVQPFNVTQGKYSTAQECSLDDDTILIPPIVAGLSGLIIVIVIAAYVIGRRKSY  
 AGYQTL (SEQ ID NO: 16)

LAMP-2C protein sequence

MVCFRLFPVPGSGLVLVCLVLGAVRSYALELNLTDSENATCLYAKWQMNFTVRYET  
 TNKTYKTVTISDHGTVTYNGSICGDDQNGPKIAVQFGPGFSWIANFTKAASTYSIDSV

SFSYNTGDNTTFPDAEDKGILTVDELLAIRIPLNDLFR CNSLSTLEKNDVVQHYWDVL  
VQAFVQNGTVSTNEFLCDKDKTSTVAPTIHTTVPSPTTTPKPEAGTYSVNNGN  
DTCLLATMGLQLNITQDKVASVININPNTTHSTGSCRSHALLRLNSSTIKYLDFVFA  
VKNENRFYLKEVNISMVYLVNGSVFSIANNLSYWDAPLGSSYMCNKEQTVSVSGAF  
QINTFDLRVQPFNVQTQGYSTAECSADSDLNFLIPVAVGVALGFLIIVVFISYMIGRR  
KSRTGYQSV (SEQ ID NO: 17)

In particular embodiments, the expression cassette comprises a polynucleotide sequence encoding LAMP-2 disclosed herein, *e.g.*, SEQ ID NOs: 15-17 or a sequence having at least 90%, at least 95%, at least 98%, or at least 99% identity to any of SEQ ID NOs: 15-17. The gene therapy vectors can be viral or non-viral vectors. Illustrative non-viral vectors include, *e.g.*, naked DNA, cationic liposome complexes, cationic polymer complexes, cationic liposome-polymer complexes, and exosomes. Examples of viral vector include, but are not limited, to adenoviral, retroviral, lentiviral, herpesvirus and adeno-associated virus (AAV) vectors.

In some embodiments, the expression cassette comprising a polynucleotide sequence encoding one or more, two or more, or all three of SEQ ID NOs: 15-17. In some embodiments, the polynucleotide sequence comprising the native introns of the LAMP-2 gene, enabling expression of more than one isoform in the same cell using one vector. In some embodiments, artificial introns, splice acceptors, and/or splice donors are used to optimize the length of the polynucleotide and/or optimize the ratio of isoforms expressed by the polynucleotide encoding two or more, or all three of SEQ ID NOs: 15-17.

In some embodiments, the expression cassette, AAV capsid gene, and/or helper genes are delivered to cells using transduction, transfection, electroporation, lipofection, and any other methods known in the art. In some embodiments, the expression cassette, AAV capsid gene, and/or helper genes are delivered in a liposome or a lipid nanoparticle (LNP). The expression cassette, AAV capsid gene, and/or helper genes may be provided as DNA, *e.g.* on one or more plasmids, bacmids, or other DNA molecules. In some embodiments, expression cassette, AAV capsid gene, and/or helper genes are delivered as RNA molecules. In some embodiments, the RNA molecules comprise one or more mRNA molecules, *e.g.*, one or more *in vitro* transcribed mRNA molecules. In some embodiments, the mRNA molecules are modified mRNA molecules. Illustrative modifications include lock nucleic acids, phosphothiolate linkages, and modified nucleosides (*e.g.* pseudouridine, 5-methylcytosine, or 5-methylcytidine). In some embodiments, the modified mRNA comprises a cap, *e.g.* an

ARCA cap. The expression cassette, AAV capsid gene, and/or helper genes may be delivered *in vitro* or *in vivo*. In some embodiments, the AAV capsid gene comprises one or more of an AAV9 capsid gene and an AAVrh74 capsid gene.

Gene delivery viral vectors useful in the practice of the present invention can be constructed utilizing methodologies well known in the art of molecular biology. Typically, viral vectors carrying transgenes are assembled from polynucleotides encoding the transgene, suitable regulatory elements and elements necessary for production of viral proteins, which mediate cell transduction. Such recombinant viruses may be produced by techniques known in the art, *e.g.*, by transfecting packaging cells or by transient transfection with helper plasmids or viruses. Typical examples of virus packaging cells include but are not limited to HeLa cells, SF9 cells (optionally with a baculovirus helper vector), 293 cells, etc. A Herpesvirus-based system can be used to produce AAV vectors, as described in US20170218395A1. Detailed protocols for producing such replication-defective recombinant viruses may be found for instance in W095/14785, W096/22378, U.S. Pat. No. 5,882,877, U.S. Pat. No. 6,013,516, U.S. Pat. No. 4,861,719, U.S. Pat. No. 5,278,056 and W094/19478, the complete contents of each of which is hereby incorporated by reference.

AAV is a 4.7 kb, single stranded DNA virus. Recombinant vectors based on AAV are associated with excellent clinical safety, since wild-type AAV is nonpathogenic and has no etiologic association with any known diseases. In addition, AAV offers the capability for highly efficient gene delivery and sustained transgene expression in numerous tissues. By an “AAV vector” is meant a vector derived from an adeno-associated virus serotype, including without limitation, AAV1, AAV2, AAV3, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, AAV10, AAVrh.10, AAVrh74, *etc.* AAV vectors can have one or more of the AAV wild-type genes deleted in whole or part, *e.g.*, the rep and/or cap genes, but retain functional flanking inverted terminal repeat (ITR) sequences. Functional ITR sequences are necessary for the rescue, replication and packaging of the AAV virion. Thus, an AAV vector is defined herein to include at least those sequences required in cis for replication and packaging (*e.g.*, functional ITRs) of the virus. The ITRs need not be the wild-type nucleotide sequences, and may be altered, *e.g.* by the insertion, deletion or substitution of nucleotides, as long as the sequences provide for functional rescue, replication and packaging. AAV vectors may comprise other modifications, including but not limited to one or more modified capsid proteins (*e.g.*, VP1, VP2 and/or VP3). For example, a capsid protein may be modified to alter tropism and/or reduce immunogenicity. AAV expression vectors are constructed using

known techniques to at least provide as operatively linked components in the direction of transcription, control elements including a transcriptional initiation region, the DNA of interest (*i.e.* the LAMP-2 gene) and a transcriptional termination region.

Adeno-associated virus (AAV) is single stranded DNA virus. The AAV genome is built of single-stranded deoxyribonucleic acid (ssDNA), either positive- or negative-sensed, which is about 4.7 kilobase long. The genome comprises inverted terminal repeats (ITRs) at both ends of the DNA strand, and two open reading frames (ORFs): *rep* and *cap*. The first, *rep*, is composed of four overlapping genes encoding Rep proteins required for the AAV life cycle, and the second, *cap*, encodes three capsid proteins: VP1, VP2 and VP3. The *cap* gene is expressed as a messenger RNA (mRNA) from the p40 promoter of AAV. The mRNA is alternatively spliced into 2.3 kb and 2.6 kb transcripts, with the 2.3 kb transcript being more abundant. VP1 is expressed only from the 2.6 kb transcript and the VP1 protein is 87 kilodaltons (kDa) in molecular weight. VP2 is expressed from an open reading frame that begins with an ACG codon, rather than a canonical AUG codon, due to the presence of an optimal Kozak sequence for translation initiation. VP2 is 72 kDa. VP3, only 62 kDa, is expressed from the ATG sequence presence in the 2.3 kb transcript, as well as the 2.6 kb transcript. The relative abundances of VP1:VP2:VP3 are 1:1:10. VP1, VP2, and VP3 interact together to form a capsid of an icosahedral symmetry.

Recombinant vectors based on AAV are associated with excellent clinical safety, since wild-type AAV is nonpathogenic and has no etiologic association with any known diseases. In addition, AAV offers the capability for highly efficient gene delivery and sustained transgene expression in numerous tissues. Various serotypes of AAV are known, including, AAV1, AAV2, AAV3, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, AAV10, AAVrh.10, AAVrh74, *etc.*. AAV vectors can have one or more of the AAV wild-type genes deleted in whole or part, *e.g.*, the *rep* and/or *cap* genes, but retain functional flanking inverted terminal repeat (ITR) sequences. The serotype of a recombinant AAV vector is determined by its capsid. International Patent Publication No. WO2003042397A2 discloses various capsid sequences including those of AAV1, AAV2, AAV3, AAV8, AAV9, and AAVrh10. International Patent Publication No. WO2013078316A1 discloses the polypeptide sequence of the VP1 from AAVrh74. Numerous diverse naturally occurring or genetically modified AAV capsid sequences are known in the art.

The present disclosure also provides pharmaceutical compositions comprising an expression cassette or vector (*e.g.*, gene therapy vector) disclosed herein and one or more

pharmaceutically acceptable carriers, diluents or excipients. In particular embodiments, the pharmaceutical composition comprises an AAV vector comprising an expression cassette disclosed herein, *e.g.*, wherein the expression cassette comprises a codon-optimized transgene encoding LAMP-2B, *e.g.*, any of SEQ ID NOs: 7-9. Provided are pharmaceutical compositions, *e.g.*, for use in preventing or treating a disorder characterized by deficient autophagic flux (*e.g.*, Danon disease) which comprises a therapeutically effective amount of a vector that comprises a nucleic acid sequence of a polynucleotide that encodes one or more isoforms of LAMP-2.

The pharmaceutical compositions that contain the expression cassette or vector may be in any form that is suitable for the selected mode of administration, for example, for intraventricular, intramyocardial, intracoronary, intravenous, intra-arterial, intra-renal, intraurethral, epidural or intramuscular administration. The gene therapy vector comprising a polynucleotide encoding one or more LAMP-2 isoforms can be administered, as sole active agent, or in combination with other active agents, in a unit administration form, as a mixture with conventional pharmaceutical supports, to animals and human beings. In some embodiments, the pharmaceutical composition comprises cells transduced *ex vivo* with any of the gene therapy vectors of the disclosure.

In various embodiments, the pharmaceutical compositions contain vehicles (*e.g.*, carriers, diluents and excipients) that are pharmaceutically acceptable for a formulation capable of being injected. These may be in particular isotonic, sterile, saline solutions (monosodium or disodium phosphate, sodium, potassium, calcium or magnesium chloride and the like or mixtures of such salts). Illustrative pharmaceutical forms suitable for injectable use include, *e.g.*, sterile aqueous solutions or dispersions; formulations including sesame oil, peanut oil or aqueous propylene glycol; and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions.

In another aspect, the disclosure provides methods of preventing, mitigating, ameliorating, reducing, inhibiting, eliminating and/or reversing one or more symptoms of Danon disease or another autophagy disorder in a subject in need thereof, comprising administering to the subject a gene therapy vector of the disclosure. The term “Danon disease” refers to an X-linked dominant skeletal and cardiac muscle disorder with multisystem clinical manifestations. Danon disease mutations lead to an absence of lysosome-associated membrane protein 2 (LAMP-2) protein expression. Major clinical features include skeletal and cardiac myopathy, cardiac conduction abnormalities, cognitive

difficulties, and retinal disease. Men are typically affected earlier and more severely than women.

In an embodiment, the vector is administered via a route selected from the group consisting of parenteral, intravenous, intra-arterial, intracardiac, intracoronary, intramyocardial, intrarenal, intraurethral, epidural, and intramuscular. In an embodiment, the vector is administered multiple times. In an embodiment, the vector is administered by intramuscular injection of the vector. In an embodiment, the vector is administered by injection of the vector into skeletal muscle. In an embodiment, the expression cassette comprises a muscle-specific promoter, optionally a muscle creatine kinase (MCK) promoter or a MCK/SV40 hybrid promoter as described in Takeshita et al. Muscle creatine kinase/SV40 hybrid promoter for muscle-targeted long-term transgene expression. *Int J Mol Med.* 2007 Feb;19(2):309-15. In an embodiment, the vector is administered by intracardiac injection.

In an embodiment, the disclosure provides a method of treating a disease or disorder, optionally Danon disease, in a subject in need thereof, comprising contacting cells with a gene therapy vector according to the present disclosure and administering the cells to the subject. In an embodiment, the cells are stem cells, optionally pluripotent stem cells. In an embodiment, the stem cells are capable of differentiation into cardiac tissue. In an embodiment, the stem cells are capable of differentiation into muscle tissue, *e.g.*, cardiac muscle tissue and/or skeletal muscle tissue. In an embodiment, the stem cells are autologous. In an embodiment, the stem cells are induced pluripotent stem cells (iPSCs).

In an embodiment, the autophagy disorder is selected from the group consisting of end-stage heart failure, myocardial infarction, drug toxicities, diabetes, end-stage renal failure, and aging. In an embodiment, the subject is a mammal, *e.g.*, a human. In an embodiment, the subject is exhibiting symptoms of Danon disease or another autophagy disorder. In an embodiment, the subject has been identified as having reduced or non-detectable LAMP-2 expression. In an embodiment, the subject has been identified as having a mutated LAMP-2 gene.

Subjects/patients amenable to treatment using the methods described herein include individuals at risk of a disease or disorder characterized by insufficient autophagic flux (*e.g.*, Danon disease as well as other known disorders of autophagy including, but not limited to, systolic and diastolic heart failure, myocardial infarction, drug toxicities (for example,

anthracyclines, chloroquine, and its derivatives), diabetes, end-stage renal disease, and aging) but not showing symptoms, as well as subjects presently showing symptoms. Such subject may have been identified as having a mutated LAMP-2 gene or as having reduced or non-detectable levels of LAMP-2 expression.

In some embodiments, the subject is exhibiting symptoms of a disease or disorder characterized by insufficient autophagic flux (*e.g.*, Danon disease as well as other known disorders of autophagy including, but not limited to, systolic and diastolic heart failure, myocardial infarction, drug toxicities, diabetes, end-stage renal disease, and aging). The symptoms may be actively manifesting, or may be suppressed or controlled (*e.g.*, by medication) or in remission. The subject may or may not have been diagnosed with the disorder, *e.g.*, by a qualified physician.

### ***Definitions***

The terms “lysosome-associated membrane protein 2” and “LAMP-2” interchangeably refer to nucleic acids and polypeptide polymorphic variants, alleles, mutants, and interspecies homologs that: (1) have an amino acid sequence that has greater than about 90% amino acid sequence identity, for example, 91 %, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% or greater amino acid sequence identity, preferably over a region of at least about 25, 50, 100, 200, 300, 400, or more amino acids, or over the full-length, to an amino acid sequence encoded by a LAMP-2 nucleic acid (*see, e.g.*, GenBank Accession Nos. NM\_002294.2 (isoform A), NM\_013995.2 (isoform B), NM\_001122606.1 (isoform C)) or to an amino acid sequence of a LAMP-2 polypeptide (*see e.g.*, GenBank Accession Nos. NP\_002285.1 (isoform A), NP\_054701.1 (isoform B), NP\_001116078.1 (isoform C)); (2) bind to antibodies, *e.g.*, polyclonal antibodies, raised against an immunogen comprising an amino acid sequence of a LAMP-2 polypeptide (*e.g.*, LAMP-2 polypeptides described herein); or an amino acid sequence encoded by a LAMP-2 nucleic acid (*e.g.*, LAMP-2 polynucleotides described herein), and conservatively modified variants thereof; (3) specifically hybridize under stringent hybridization conditions to an anti-sense strand corresponding to a nucleic acid sequence encoding a LAMP-2 protein, and conservatively modified variants thereof; (4) have a nucleic acid sequence that has greater than about 90%, preferably greater than about 91 %, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or higher nucleotide sequence identity, preferably over a region of at least about 25, 50, 100, 200, 500, 1000, 2000 or more nucleotides, or over the full-length, to a LAMP-2 nucleic acid (*e.g.*,

LAMP-2 polynucleotides, as described herein, and LAMP-2 polynucleotides that encode LAMP-2 polypeptides, as described herein).

The terms “identical” or percent “identity,” in the context of two or more nucleic acids or polypeptide sequences, refer to two or more sequences or subsequences that are the same or have a specified percentage of amino acid residues or nucleotides that are the same (*i.e.*, share at least about 80% identity, for example, at least about 85%, 86%, 87%, 88%, 89%, 90%, 91 %, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% identity over a specified region to a reference sequence, *e.g.*, LAMP-2 polynucleotide or polypeptide sequence as described herein, when compared and aligned for maximum correspondence over a comparison window, or designated region as measured using one of the following sequence comparison algorithms or by manual alignment and visual inspection. Such sequences are then said to be “substantially identical.” This definition also refers to the complement of a test sequence. Preferably, the identity exists over a region that is at least about 25 amino acids or nucleotides in length, for example, over a region that is 50, 100, 200, 300, 400 amino acids or nucleotides in length, or over the full-length of a reference sequence.

For sequence comparison, typically one sequence acts as a reference sequence, to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are entered into a computer, subsequence coordinates are designated, if necessary, and sequence algorithm program parameters are designated. Default program parameters can be used, or alternative parameters can be designated. The sequence comparison algorithm then calculates the percent sequence identities for the test sequences relative to the reference sequence, based on the program parameters. For sequence comparison of nucleic acids and proteins to LAMP-2 nucleic acids and proteins, the BLAST and BLAST 2.0 algorithms and the default parameters are used.

A “comparison window”, as used herein, includes reference to a segment of any one of the number of contiguous positions selected from the group consisting of from 20 to 600, usually about 50 to about 200, more usually about 100 to about 150 in which a sequence may be compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned. Methods of alignment of sequences for comparison are well-known in the art. Optimal alignment of sequences for comparison can be conducted, *e.g.*, by the local homology algorithm of Smith & Waterman, *Adv. Appl. Math.* 2:482 (1981), by the homology alignment algorithm of Needleman & Wunsch, *J. Mol. Biol.* 48:443 (1970), by the search for similarity method of Pearson & Lipman, *Proc. Nat’l. Acad. Sci. USA*

85:2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI), or by manual alignment and visual inspection (see, *e.g.*, Ausubel et al., eds., *Current Protocols in Molecular Biology* (1995 supplement)). Examples of algorithms that are suitable for determining percent sequence identity and sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul et al., *J. Mol. Biol.* 215:403-410 (1990) and Altschul et al., *Nucleic Acids Res.* 25:3389-3402 (1977), respectively. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (on the worldwide web at [ncbi.nlm.nih.gov/](http://ncbi.nlm.nih.gov/)).

An indication that two nucleic acid sequences or polypeptides are substantially identical is that the polypeptide encoded by the first nucleic acid is immunologically cross reactive with the antibodies raised against the polypeptide encoded by the second nucleic acid, as described below. Thus, a polypeptide is typically substantially identical to a second polypeptide, for example, where the two peptides differ only by conservative substitutions. Another indication that two nucleic acid sequences are substantially identical is that the two molecules or their complements hybridize to each other under stringent conditions. Yet another indication that two nucleic acid sequences are substantially identical is that the same primers can be used to amplify the sequence.

As used herein, “administering” refers to local and systemic administration, *e.g.*, including enteral, parenteral, pulmonary, and topical/transdermal administration. Routes of administration for compounds (*e.g.*, polynucleotide encoding one or more LAMP- 2 isoforms) that find use in the methods described herein include, *e.g.*, oral (per os (P.O.)) administration, nasal or inhalation administration, administration as a suppository, topical contact, transdermal delivery (*e.g.*, via a transdermal patch), intrathecal (IT) administration, intravenous (“iv”) administration, intraperitoneal (“ip”) administration, intramuscular (“im”) administration, intralesional administration, or subcutaneous (“sc”) administration, or the implantation of a slow-release device *e.g.*, a mini-osmotic pump, a depot formulation, etc. , to a subject. Administration can be by any route including parenteral and transmucosal (*e.g.*, oral, nasal, vaginal, rectal, or transdermal). Parenteral administration includes, *e.g.*, intravenous, intramuscular, intraarterial, intrarenal, intraurethral, intracardiac, intracoronary, intramyocardial, intradermal, epidural, subcutaneous, intraperitoneal, intraventricular, ionophoretic and intracranial. Other modes of delivery include, but are not limited to, the use

of liposomal formulations, intravenous infusion, transdermal patches, etc. In some embodiments, the dose of rAAV gene therapy vector administered is about  $1\text{E}+11$  vector genomes (vg)/kg to about  $1\text{E}+12$  vg/kg, about  $1\text{E}+12$  vg/kg to about  $2\text{E}+12$  vg/kg, about  $2\text{E}+12$  vg/kg to about  $3\text{E}+12$  vg/kg, about  $3\text{E}+12$  vg/kg to about  $3\text{E}+13$  vg/kg, or about  $3\text{E}+13$  vg/kg to about  $3\text{E}+14$  vg/kg. In some embodiments, the dose of rAAV gene therapy vector administered is about  $3\text{E}+12$  vg/kg to about  $3\text{E}+14$  vg/kg.

The terms “systemic administration” and “systemically administered” refer to a method of administering a compound or composition to a mammal so that the compound or composition is delivered to sites in the body, including the targeted site of pharmaceutical action, via the circulatory system. Systemic administration includes, but is not limited to, oral, intranasal, rectal and parenteral (*e.g.*, other than through the alimentary tract, such as intramuscular, intravenous, intra-arterial, transdermal and subcutaneous) administration.

The term “co-administering” or “concurrent administration”, when used, for example with respect to the compounds (*e.g.*, LAMP-2 polynucleotides) and/or analogs thereof and another active agent, refers to administration of the compound and/or analogs and the active agent such that both can simultaneously achieve a physiological effect. The two agents, however, need not be administered together. In certain embodiments, administration of one agent can precede administration of the other. Simultaneous physiological effect need not necessarily require presence of both agents in the circulation at the same time. However, in certain embodiments, co-administering typically results in both agents being simultaneously present in the body (*e.g.*, in the plasma) at a significant fraction (*e.g.*, 20% or greater, *e.g.*, 30% or 40% or greater, *e.g.*, 50% or 60% or greater, *e.g.*, 70% or 80% or 90% or greater) of their maximum serum concentration for any given dose.

The term “effective amount” or “pharmaceutically effective amount” refer to the amount and/or dosage, and/or dosage regime of one or more compounds (*e.g.*, gene therapy vectors) necessary to bring about the desired result *e.g.*, increased expression of one or more LAMP-2 isoforms in an amount sufficient to reduce the ultimate severity of a disease characterized by impaired or deficient autophagy (*e.g.*, Danon disease). In some embodiments, the effective amount is about  $1\text{E}+11$  vg/kg to about  $1\text{E}+12$  vg/kg, about  $1\text{E}+12$  vg/kg to about  $2\text{E}+12$  vg/kg, about  $2\text{E}+12$  vg/kg to about  $3\text{E}+12$  vg/kg, about  $3\text{E}+12$  vg/kg to about  $3\text{E}+13$  vg/kg, or about  $3\text{E}+13$  vg/kg to about  $3\text{E}+14$  vg/kg of rAAV gene therapy vector. In some embodiments, the effective amount is about  $3\text{E}+12$  vg/kg to about  $3\text{E}+14$  vg/kg of rAAV gene therapy vector.

The phrase “cause to be administered” refers to the actions taken by a medical professional (*e.g.*, a physician), or a person controlling medical care of a subject, that control and/or permit the administration of the agent(s)/compound(s) at issue to the subject. Causing to be administered can involve diagnosis and/or determination of an appropriate therapeutic or prophylactic regimen, and/or prescribing particular agent(s)/compounds for a subject. Such prescribing can include, for example, drafting a prescription form, annotating a medical record, and the like.

As used herein, the terms “treating” and “treatment” refer to delaying the onset of, retarding or reversing the progress of, reducing the severity of, or alleviating or preventing either the disease or condition to which the term applies, or one or more symptoms of such disease or condition. The terms “treating” and “treatment” also include preventing, mitigating, ameliorating, reducing, inhibiting, eliminating and/or reversing one or more symptoms of the disease or condition.

The term “mitigating” refers to reduction or elimination of one or more symptoms of that pathology or disease, and/or a reduction in the rate or delay of onset or severity of one or more symptoms of that pathology or disease, and/or the prevention of that pathology or disease. In certain embodiments, the reduction or elimination of one or more symptoms of pathology or disease can include, *e.g.*, measurable and sustained increase in the expression levels of one or more isoforms of LAMP-2.

As used herein, the phrase “consisting essentially of” refers to the genera or species of active pharmaceutical agents recited in a method or composition, and further can include other agents that, on their own do not have substantial activity for the recited indication or purpose.

The terms “subject,” “individual,” and “patient” interchangeably refer to a mammal, preferably a human or a non-human primate, but also domesticated mammals (*e.g.*, canine or feline), laboratory mammals (*e.g.*, mouse, rat, rabbit, hamster, guinea pig) and agricultural mammals (*e.g.*, equine, bovine, porcine, ovine). In various embodiments, the subject can be a human (*e.g.*, adult male, adult female, adolescent male, adolescent female, male child, female child).

The terms “gene transfer” or “gene delivery” refer to methods or systems for reliably inserting foreign DNA into host cells. Such methods can result in transient expression of non-integrated transferred DNA, extrachromosomal replication and expression of transferred

replicons (*e.g.* episomes), or integration of transferred genetic material into the genomic DNA of host cells.

The term “vector” is used herein to refer to a nucleic acid molecule capable of transferring or transporting another nucleic acid molecule. The transferred nucleic acid is generally linked to, *e.g.*, inserted into, the vector nucleic acid molecule. A vector may include sequences that direct autonomous replication or reverse transcription in a cell, or may include sequences sufficient to allow integration into host cell DNA. “vectors” include gene therapy vectors. As used herein, the term “gene therapy vector” refers to a vector capable of use in performing gene therapy, *e.g.*, delivering a polynucleotide sequence encoding a therapeutic polypeptide to a subject. Gene therapy vectors may comprise a nucleic acid molecule (“transgene”) encoding a therapeutically active polypeptide, *e.g.*, a LAMP-2B or other gene useful for replacement gene therapy when introduced into a subject. Useful vectors include, but are not limited to, viral vectors.

As used herein, the term “expression cassette” refers to a DNA segment that is capable in an appropriate setting of driving the expression of a polynucleotide (a “transgene”) encoding a therapeutically active polypeptide (*e.g.*, LAMP-2B) that is incorporated in said expression cassette. When introduced into a host cell, an expression cassette *inter alia* is capable of directing the cell’s machinery to transcribe the transgene into RNA, which is then usually further processed and finally translated into the therapeutically active polypeptide. The expression cassette can be comprised in a gene therapy vector. Generally, the term expression cassette excludes polynucleotide sequences 5’ to the 5’ ITR and 3’ to the 3’ ITR.

All patents, patent publications, and other publications referenced and identified in the present specification are individually and expressly incorporated herein by reference in their entirety for all purposes.

## EXAMPLES

### ***EXAMPLE 1: Pre-Clinical and Clinical Evaluation of AAVrh74-LAMP-2B***

A plasmid vector including the gene expression cassette as depicted in **FIG. 1** is generated. The transgene is modified to encode LAMP2B-HA-FLAG, so that the protein may be detected using either anti-HA or anti-FLAG antibodies. AAVrh74-LAMP2B viral vector is generated using a three-plasmid, helper virus-free system to generate recombinant AAV particles containing serotype rh74 capsid proteins and viral genomes that have AAV2 ITRs

flanking a human LAMP-2B expression cassette. The viral vector is tested in non-human primates.

Pharmacology and toxicology studies are conducted in LAMP-2B<sup>-/-</sup> and wild-type mice. Based on the preclinical safety and efficacy data observed in mice and non-human primate studies, clinical studies in patients with Danon disease are performed.

***EXAMPLE 2: DNA, RNA, and Protein Expression in Non-Human Primates Following Intravenous Administration of  $1 \times 10^{13}$  vg/kg AAV9.LAMP2B and AAVrh74.LAMP2B***

Non-human primate studies of AAV9 versus AAVrh74 vectors were performed in paired male and female African Green Monkeys (AGM). Subjects received either AAV9.LAMP2B-HA-Flag or AAVrh74.LAMP2B-HA-Flag. “AAV9.LAMP2B-HA-Flag” is an AAV9 serotype adeno-associated virus vector encoding LAMP2B C-terminally fused to an HA-Flag tag. “AAVrh74.LAMP2B-HA-Flag” is an AAVrh74 serotype adeno-associated virus vector encoding LAMP2B C-terminally fused to an HA-Flag tag. One subject was given vehicle control. The vectors were administered by intravenous injection of 2 mL of  $1.85 \times 10^{13}$  vector genomes (vg)/mL as determined by quantitative polymerase chain reaction (qPCR) using a plasmid containing the WPRE sequence to generate a reference curve. This injection achieved the target dose of vector, which was about  $1.0 \times 10^{13}$  vg/kg. Due to lower body weight, female subjects received about  $1.2 \times 10^{13}$  vg/kg of their respective vectors. This experiment is summarized in **Table 2**.

**Table 2**

<b>Animal</b>	<b>Sex</b>	<b>Treatment</b>	<b>Weight (kg)</b>	<b>Dose (vg/kg)</b>
B059	M	Vehicle buffer	3.88	n/a
A991	F	AAV9	2.97	$1.2 \times 10^{13}$
A602	M	AAV9	3.56	$1.0 \times 10^{13}$
A710	F	AAVrh74	3.27	$1.2 \times 10^{13}$
A981	M	AAVrh74	3.69	$1.0 \times 10^{13}$

Subjects were humanely sacrificed two months after injection and tissues were collected for DNA, RNA, and protein analysis. The following tissues were examined: heart

(left atrium, right atrium, left ventricle and right ventricle); skeletal muscle (quadricep and gastrocnemius); liver (left, right, middle and quadrate lobes); brain (frontal lobe, parietal lobe, temporal lobe, occipital lobe, cortex, hippocampus, medulla, and cerebellum); and gonads.

### ***Vector DNA – Quantitative PCR***

DNA was extracted from frozen tissues using Qiagen DNeasy® kit. DNA purity (A260/A280) and concentration were evaluated on a NanoDrop One™ spectrophotometer (Thermo). Quantitative PCR (qPCR) was performed on 20 ng DNA using TaqMan Universal Master Mix II (Thermo, 4440038) on a real-time PCR system (QuantStudio5, Thermo) using the following primers/probes:

WPRE (cassette):

Forward primer: 5'-ATCATGCTATTGCTTCCCGTA-3' (SEQ ID NO:30)

Reverse primer: 5'-GGGCCACAACCTCATAAAA-3' (SEQ ID NO:31)

Probe: 5'-CCTCCTTGTATAAATCCTGGTTGCTGTCT-3' (SEQ ID NO:32)

### **RNaseP (housekeeping gene, Thermo)**

A standard curve was generated using plasmid DNA containing the WPRE sequence. **FIG. 2** shows a bar graph of vector DNA quantification in organs most affected in Danon disease by qPCR.

### ***LAMP2B mRNA – Quantitative RT-PCR***

RNA was extracted from heart and muscle tissues using the RNeasy Fibrous Tissue kit (Qiagen), and from liver and brain using RNeasy Lipid Tissue kit (Qiagen). Purity (A260/A280) and concentration were determined on a NanoDrop One spectrophotometer. RNA was converted to cDNA using the Superscript IV VILO master mix (Thermo). qPCR was performed on 10 ng of RNA in TaqMan Universal Master Mix II (Thermo) on a real-time PCR system (QuantStudio5, Thermo) using the following primers/probes:

WPRE (cassette):

Forward primer: 5'-ATCATGCTATTGCTTCCCGTA-3' (SEQ ID NO:33)

Reverse primer: 5'-GGGCCACAACCTCATAAAA-3' (SEQ ID NO:34)

Probe: 5'-CCTCCTTGTATAAATCCTGGTTGCTGTCT-3' (SEQ ID NO:35)

HPRT-1 (Housekeeping gene, Thermo)

A standard curve was generated using plasmid DNA containing the WPRE sequence. **FIG. 3A** shows a bar graph of vector DNA quantification in regions of the heart by qPCR. **FIG. 3B** shows a bar graph of vector DNA quantification in muscles by qPCR. **FIG. 4** shows a bar graph of mRNA quantification in organs most affected by Danon disease by RT-qPCR. **FIG. 5A** shows a bar graph of mRNA quantification in regions of the heart by RT-qPCR. **FIG. 5B** shows a bar graph of mRNA quantification in muscles by RT-qPCR.

*LAMP2B mRNA - RNAscope*

5mm tissue cubes fixed in 10% neutral buffered formalin, embedded in paraffin and sectioned. Transgene mRNA was detected using WPRE-O3 ZZ probe (ACD) with RNAscope 2.5 LS RED. Semi-quantitative visual assessment of one section from each tissue was performed with cells with  $\geq 1$  dot per cell considered positive. The percentage of cells positive were binned into five categories: 0%, 1-25%, 26-50%, 51-75% or 100%.

**FIG. 6A** shows a micrograph of semi-quantitative mRNA analysis by RNAscope in an untreated left ventricle. **FIG. 6B** shows micrographs of semi-quantitative mRNA analysis by RNAscope in treated left ventricles. **FIG. 7A** shows a micrograph of semi-quantitative mRNA analysis by RNAscope in an untreated quadricep. **FIG. 7B** shows micrographs of semi-quantitative mRNA analysis by RNAscope in treated quadriceps. **FIG. 8** shows micrographs of semi-quantitative mRNA analysis by RNAscope in treated gastrocnemius. Results are summarized for vehicle control (**Table 3A**), AAV9 (**Table 3A**), and AAVrh74 (**Table 3C**).

**Table 3A**

<b>Animal</b>	<b>Tissue</b>	<b>Location</b>	<b>Treatment</b>	<b>% Expressing Cells</b>
B059	Heart	left ventricle	Untreated, vehicle	0%
B059	Muscle	Quadricep	Untreated, vehicle	0%
B059	Liver	Left Lobe	Untreated, vehicle	0%

**Table 3B**

<b>Animal</b>	<b>Tissue</b>	<b>Location</b>	<b>Treatment</b>	<b>% Expressing Cells</b>
A991	Heart	left ventricle	Treated, AAV9	26-50%
A991	Heart	right ventricle	Treated, AAV9	26-50%
A991	Heart	left atrium	Treated, AAV9	1-25%
A991	Heart	right atrium	Treated, AAV9	26-50%
A991	Muscle	quadricep	Treated, AAV9	0%
A991	Muscle	gastrocnemius	Treated, AAV9	0%
A991	Liver	left lobe	Treated, AAV9	26-50%
A991	Liver	right lobe	Treated, AAV9	51-75%
A602	Heart	left ventricle	Treated, AAV9	1-25%
A602	Heart	right ventricle	Treated, AAV9	1-25%
A602	Heart	left atrium	Treated, AAV9	26-50%
A602	Heart	right atrium	Treated, AAV9	1-25%
A602	Muscle	quadricep	Treated, AAV9	1-25%
A602	Muscle	gastrocnemius	Treated, AAV9	1-25%
A602	Liver	left lobe	Treated, AAV9	51-75%
A602	Liver	right lobe	Treated, AAV9	51-75%

**Table 3C**

<b>Animal</b>	<b>Tissue</b>	<b>Location</b>	<b>Treatment</b>	<b>% Expressing Cells</b>
A710	Heart	left ventricle	Treated, AAVrh.74	1-25%
A710	Heart	right ventricle	Treated, AAVrh.74	1-25%
A710	Heart	left atrium	Treated, AAVrh.74	1-25%
A710	Heart	right atrium	Treated, AAVrh.74	1-25%
A710	Muscle	quadricep	Treated, AAVrh.74	0%
A710	Muscle	gastrocnemius	Treated, AAVrh.74	1-25%
A710	Liver	left lobe	Treated, AAVrh.74	51-75%
A710	Liver	right lobe	Treated, AAVrh.74	51-75%
A981	Heart	left ventricle	Treated, AAVrh.74	26-50%

Animal	Tissue	Location	Treatment	% Expressing Cells
A981	Heart	right ventricle	Treated, AAVrh.74	26-50%
A981	Heart	left atrium	Treated, AAVrh.74	0%
A981	Heart	right atrium	Treated, AAVrh.74	26-50%
A981	Muscle	quadricep	Treated, AAVrh.74	0%
A981	Muscle	gastrocnemius	Treated, AAVrh.74	0%
A981	Liver	left lobe	Treated, AAVrh.74	51-75%
A981	Liver	right lobe	Treated, AAVrh.74	26-51%

### *LAMP2B protein - ELISA*

Approximately 125 mg of tissue was homogenized in 500  $\mu$ L of lysis buffer using 0.9-2.00 mm stainless steel beads (Next Advance) and a Next Advance Bullet Blender 24. The lysis buffer contains 300mM NaCl, 20mM EDTA, 100mM Tris pH 8.0, 2% NP-40 and 0.2% SDS with Complete™ EDTA-free protease inhibitor and PhosSTOP™ phosphatase inhibitor. Total protein was assessed by BCA (Thermo). 100 mg of total protein was loaded per well. A standard curve was constructed using purified human LAMP2 protein (Origene). ELISA was performed with a mouse monoclonal antibody (H4B4, Novus Biologicals) as the capture antibody, a goat polyclonal antibody (R&D Systems) as the detection antibody, HRP-linked antibody: Donkey anti-goat (Millipore) as the secondary antibody. Plates were developed with TMB (Thermo) and quantified on a spectrophotometer (Spectramax M5c).

**FIG. 9** shows a bar graph of protein quantification in tissues most affected in Danon disease by ELISA. **FIG. 10A** shows a bar graph of protein quantification in regions of the heart by ELISA. **FIG. 10B** shows a bar graph of protein quantification in muscles by ELISA.

### *Clinical Pathology*

Pathological effects of the vectors were assessed. **FIGS. 11A-11D** show line graphs of clinical pathology measurement in NHP serum over course of study. Clinical pathology levels were assessed as changes in (**FIG. 11A**) alanine aminotransferase, ALT; (**FIG. 11B**) aspartate aminotransferase, AST; (**FIG. 11C**) white blood cells, WBC; and (**FIG. 11D**) neutrophils over the study duration. B059 is the vehicle control. A991 and A602 are AAV9-treated animals. A710 and A981 are AAVrh74-treated animals.

## Conclusions

AAV-based gene therapy using a LAMP2B transgene was well tolerated in non-human primates at vector dose  $1.0 \times 10^{13}$  vg/kg. This result is an important and unexpected result because experiments with AAV-based gene therapy for some other transgenes have demonstrated pathological effects at doses equal to or lower than  $1.0 \times 10^{13}$  vg/kg. Furthermore, both AAV9 and AAVrh74 were well tolerated. Elevated levels of certain markers were observed in A602 and A710 animals at day 21, but these outliers may be due to experimental error or pathology that was self-resolving.

Both vectors localized to and transduce target tissues for treatment of Danon disease (heart and muscle), but not as much in the brain or gonads. Expression in the gonads would be undesirable for safety reasons. As expected significant amounts of vector accumulate in the liver, which is desirable because liver is a tissue affected by Danon disease. Vector is present in each quadrant of the heart and in both quadriceps and gastrocnemius muscles. This is a desirable result for treatment of Danon disease.

Localization of a serotype of AAV vector (*e.g.* AAV9) is not predictive of localization of others (*e.g.* AAVrh74). This experiment demonstrates that AAVrh74 achieves desirable localization for treatment of Danon disease, or other diseases with etiology linked to heart and muscle tissues. Both LAMP2B transgene mRNA and protein are expressed in the same sets of tissues in both AAV9 and AAVrh74 groups. Expression is comparable between vector serotypes. The number of animals in the study is too few to discern statistically significant trends in expression levels between AAV9 and AAVrh74. RNAscope suggests that a similar fraction of cells are infected in heart, muscle, and liver tissues in AAV9 and AAVrh74 groups.

These data demonstrate that AAVrh74 may be used as a vector to deliver LAMP2B to tissues relevant to treatment of Danon disease. AAVrh74 was non-inferior to AAV9 in these experiments.

## CLAIMS

What is claimed is:

1. A recombinant adeno-associated virus (rAAV) gene therapy vector, comprising a polynucleotide comprising a 5' ITR, an expression cassette, and a 3' ITR; and a capsid protein,  
  
wherein the expression cassette comprises a transgene encoding a lysosome-associated membrane protein 2 (LAMP-2) or a functional variant thereof,  
  
wherein the expression cassette is flanked by the 5' ITR and the 3' ITR, and  
  
wherein the capsid protein comprises an AAVrh.74 capsid protein or a functional variant thereof.
2. The rAAV gene therapy vector of claim 1, wherein the LAMP-2 is selected from LAMP-2A, LAMP-2B and LAMP-2C.
3. The rAAV gene therapy vector of claim 1, wherein the capsid protein has at least 95% sequence identity to an amino acid sequence selected from SEQ ID NOs: 2-4.
4. The rAAV gene therapy vector of claim 3, wherein the capsid protein shares at least 95% sequence identity to SEQ ID NOs: 2.
5. The rAAV gene therapy vector of claim 4, wherein the capsid protein shares at least 97% sequence identity to SEQ ID NOs: 2.
6. The rAAV gene therapy vector of claim 5, wherein the capsid protein shares at least 99% sequence identity to SEQ ID NOs: 2.
7. The rAAV gene therapy vector of any one of claims 1-6, wherein the capsid protein is an AAVrh.74 capsid protein.
8. The rAAV gene therapy vector of any one of claims 1-7, wherein the 5' ITR and the 3' ITR are each respectively the 5' ITR of AAV2 and the 3' ITR of AAV2, or variants thereof.
9. The rAAV gene therapy vector of claim 8, wherein the 5' ITR shares at least 98% identity to SEQ ID NO: 13 and the 3' ITR shares at least 98% identity to SEQ ID NO: 14.

10. The rAAV gene therapy vector of any one of claims 1-9, wherein the transgene is codon-optimized for expression in a human host cell.
11. The rAAV gene therapy vector of any one of claims 1-10, wherein the expression cassette contains fewer CpG sites than SEQ ID: 6.
12. The rAAV gene therapy vector of any one of claims 1-11, wherein the expression cassette contains fewer cryptic splice sites than SEQ ID: 6.
13. The rAAV gene therapy vector of any one of claims 1-12, wherein the expression cassette encodes fewer alternative open reading frames than SEQ ID: 6.
14. The rAAV gene therapy vector of any one of claims 1 to 13, wherein the transgene shares at least 95% identity to a sequence selected from SEQ ID NO: 7-9.
15. The rAAV gene therapy vector of claim 14, wherein the transgene shares at least 99% identity to a sequence selected from SEQ ID NO: 7-9.
16. The rAAV gene therapy vector of claim 15, wherein the transgene comprises a sequence selected from SEQ ID NO: 7-9.
17. The rAAV gene therapy vector of any one of claims 1 to 16, where the expression cassette comprises a consensus optimal Kozak sequence operatively linked to the transgene, wherein the consensus optimal Kozak sequence comprises SEQ ID NO: 20.
18. The rAAV gene therapy vector of any one of claims 1 to 17, where the expression cassette comprises a full-length polyA sequence operatively linked to the transgene, wherein the full-length polyA sequence comprises SEQ ID NO: 26.
19. The rAAV gene therapy vector of any one of claims 1 to 18, where the expression cassette comprises no start codon 5' to the start codon of the transgene.
20. The rAAV gene therapy vector of any one of claims 1 to 19, wherein the expression cassette comprises operatively linked, in the 5' to 3' direction, a first inverse terminal repeat, an enhancer/promoter region, an intron, a consensus optimal Kozak sequence, the transgene, a 3' untranslated region including a full-length polyA sequence, and a second inverse terminal repeat, where the expression cassette comprises no start codon 5' to the start codon of the transgene.

21. The rAAV gene therapy vector of claim 20, wherein the enhancer/promoter region comprises in the 5' to 3' direction a CMV IE Enhancer and a Chicken Beta-Actin Promoter, and optionally wherein the enhancer/promoter region further comprises a first exon and first intron of a chicken beta-actin gene and a splice acceptor of a rabbit beta-globin gene.
22. The rAAV gene therapy vector of claim 20, wherein the enhancer/promoter region comprises a tissues-specific promoter capable of mediating increased expression in cardiac tissue and/or skeletal muscle tissue compared to liver tissue.
23. The rAAV gene therapy vector of any one of claims 1 to 21, wherein the expression cassette shares at least 95% identity to a sequence selected from SEQ ID NOs: 10-12.
24. The rAAV gene therapy vector of claim 23, wherein the expression cassette comprises a sequence selected from SEQ ID NOs: 10-12.
25. A pharmaceutical composition comprising the rAAV gene therapy vector of any one of claims 1 to 24.
26. A method of treating or preventing Danon disease or another autophagy disorder in a subject in need thereof, comprising administering to the subject the rAAV gene therapy vector of any one of claims 1 to 24 or the pharmaceutical composition of claim 25.
27. The method of claim 26, wherein the rAAV gene therapy vector or pharmaceutical composition is administered via a route selected from the group consisting of intravenous, intra-arterial, intracardiac, intracoronary, intramyocardial, intrarenal, intraurethral, epidural, and intramuscular.
28. The method of claim 26 or claim 27, wherein the autophagy disorder is selected from the group consisting of end-stage heart failure, myocardial infarction, drug toxicities, diabetes, end-stage renal failure, and aging.
29. The method of any one of claims 26 to 28, wherein the subject is a human.
30. The method of any one of claims 26 to 29, wherein the subject is exhibiting symptoms of Danon disease or another autophagy disorder.
31. The method of any one of claims 26 to 30, wherein the subject has been identified as having reduced or non-detectable expression of endogenous LAMP-2.

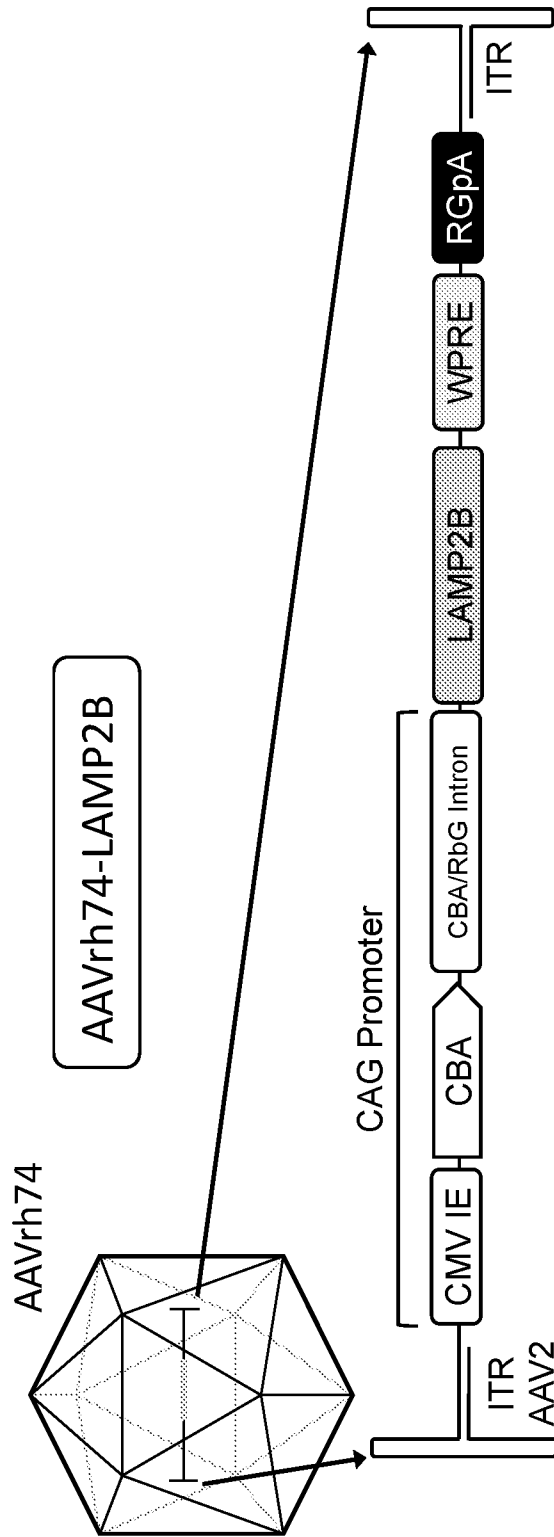
32. The method of any one of claims 26 to 31, wherein the subject has been identified as having a mutated LAMP-2 gene.
33. The method of any one of claims 26 to 32, wherein the rAAV gene therapy vector is administered at a dose of about  $3 \times 10^{12}$  vg/kg to about  $3 \times 10^{14}$  vg/kg.
34. The method of any one of claims 26 to 33, wherein the rAAV gene therapy vector is administered at a dose of about  $3 \times 10^{12}$  vg/kg to about  $1.2 \times 10^{13}$  vg/kg.
35. The method of any one of claims 26 to 33, wherein the rAAV gene therapy vector is administered at a dose of about  $1.0 \times 10^{13}$  vg/kg.
36. The method of any one of claims 26 to 35, wherein the dose of rAAV gene therapy vector does not cause clinical pathology when administered, optionally when administered at a dose of about  $1.0 \times 10^{13}$  vg/kg.
37. The method of any one of claims 26 to 36, wherein administration of the rAAV gene therapy vector transduces one or more of heart, muscle, and liver.
38. The method of any one of claims 26 to 37, wherein administration of the rAAV gene therapy vector causes LAMP2B mRNA expression in one or more of heart, muscle, and liver.
39. The method of any one of claims 26 to 38, wherein administration of the rAAV gene therapy vector causes LAMP2B protein expression in one or more of heart, muscle, and liver.
40. The method of any one of claims 26 to 39, wherein administration of the rAAV gene therapy vector causes infection with the rAAV gene therapy vector of at least about 10%, at least about 20%, or at least about 30% of cells in one or more of heart, muscle, and liver.
41. The method of any one of claims 26 to 40, wherein administration of the rAAV gene therapy vector causes transduction of the rAAV gene therapy vector in gonads at less 0.1 vector genomes (vg) per diploid genome.
42. The method of any one of claims 26 to 41, wherein administration of the rAAV gene therapy vector causes LAMP2B mRNA expression in gonads at less than  $2 \times 10^4$  mRNA copies per  $\mu$ g total RNA.
43. The method of any one of claims 26 to 42, wherein administration of the rAAV gene therapy vector causes no LAMP2B protein expression in brain and/or gonads.

44. The method of any one of claims 26 to 43, wherein administration of the rAAV gene therapy vector transduces and/or causes transgene expression at about the same level as an AAV9 gene therapy vector having the same expression cassette.
45. A method of delivering a LAMP-2 polynucleotide encoding a LAMP-2 protein to a cell, comprising contacting the cell with the rAAV gene therapy vector of any one of claims 1 to 24 or the pharmaceutical composition of claim 25, wherein the cell is optionally selected from a heart cell, a lung cell, and/or a muscle cell.
46. A method of transducing cells, comprising contacting the cells with the rAAV gene therapy vector of any one of claims 1 to 24 or the pharmaceutical composition of claim 25, wherein the cell is optionally selected from a heart cell, a lung cell, and/or a muscle cell.
47. A method of delivering a LAMP-2 polynucleotide encoding a LAMP-2 protein to a tissue and/or expressing a LAMP-2 protein in a tissue, comprising contacting the tissue with the rAAV gene therapy vector of any one of claims 1 to 24 or the pharmaceutical composition of claim 25, wherein the tissue is optionally selected from heart tissue, lung tissue, and/or muscle tissue.
48. A method of delivering a LAMP-2 polynucleotide encoding a LAMP-2 protein to a subject and/or expressing a LAMP-2 protein in a subject, comprising administering to the subject the rAAV gene therapy vector of any one of claims 1 to 24 or the pharmaceutical composition of claim 25.
49. The method of claim 48, wherein the rAAV gene therapy vector or pharmaceutical composition is administered via a route selected from the group consisting of intravenous, intra-arterial, intracardiac, intracoronary, intramyocardial, intrarenal, intraurethral, epidural, and intramuscular.
50. The method of claim 48 or claim 49, wherein the subject suffers from or is at risk for an autophagy disorder selected from the group consisting of Danon disease, end-stage heart failure, myocardial infarction, drug toxicities, diabetes, end-stage renal failure, and aging.
51. The method of any one of claims 48 to 50, wherein the subject is a human.
52. The method of any one of claims 48 to 51, wherein the subject is exhibiting symptoms of the autophagy disorder.

53. The method of any one of claims 48 to 52, wherein the subject has been identified as having reduced or non-detectable expression of endogenous LAMP-2.
54. The method of any one of claims 48 to 53, wherein the subject has been identified as having a mutated LAMP-2 gene.
55. The method of any one of claims 48 to 54, wherein the rAAV gene therapy vector is administered at a dose of about  $3 \times 10^{12}$  vg/kg to about  $3 \times 10^{14}$  vg/kg.
56. The method of any one of claims 48 to 55, wherein the rAAV gene therapy vector is administered at a dose of about  $3 \times 10^{12}$  vg/kg to about  $1.2 \times 10^{13}$  vg/kg.
57. The method of any one of claims 48 to 56, wherein the rAAV gene therapy vector is administered at a dose of about  $1.0 \times 10^{13}$  vg/kg.
58. The method of any one of claims 48 to 57, wherein the dose of rAAV gene therapy vector does not cause clinical pathology when administered, optionally when administered at a dose of about  $1.0 \times 10^{13}$  vg/kg.
59. The method of any one of claims 48 to 58, wherein administration of the rAAV gene therapy vector transduces one or more of heart, muscle, and liver.
60. The method of any one of claims 48 to 59, wherein administration of the rAAV gene therapy vector causes LAMP2B mRNA expression in one or more of heart, muscle, and liver.
61. The method of any one of claims 48 to 60, wherein administration of the rAAV gene therapy vector causes LAMP2B protein expression in one or more of heart, muscle, and liver.
62. The method of any one of claims 48 to 61, wherein administration of the rAAV gene therapy vector causes infection with the rAAV gene therapy vector of at least about 10%, at least about 20%, or at least about 30% of cells in one or more of heart, muscle, and liver.
63. The method of any one of claims 48 to 62, wherein administration of the rAAV gene therapy vector causes transduction of the rAAV gene therapy vector in gonads at less 0.1 vector genomes (vg) per diploid genome.
64. The method of any one of claims 48 to 63, wherein administration of the rAAV gene therapy vector causes LAMP2B mRNA expression in gonads at less than  $2 \times 10^4$  mRNA copies per  $\mu\text{g}$  total RNA.

65. The method of any one of claims 48 to 64, wherein administration of the rAAV gene therapy vector causes no LAMP2B protein expression in brain and/or gonads.

66. The method of any one of claims 48 to 65, wherein administration of the rAAV gene therapy vector transduces and/or causes transgene expression at about the same level as an AAV9 gene therapy vector having the same expression cassette.



**FIG. 1**

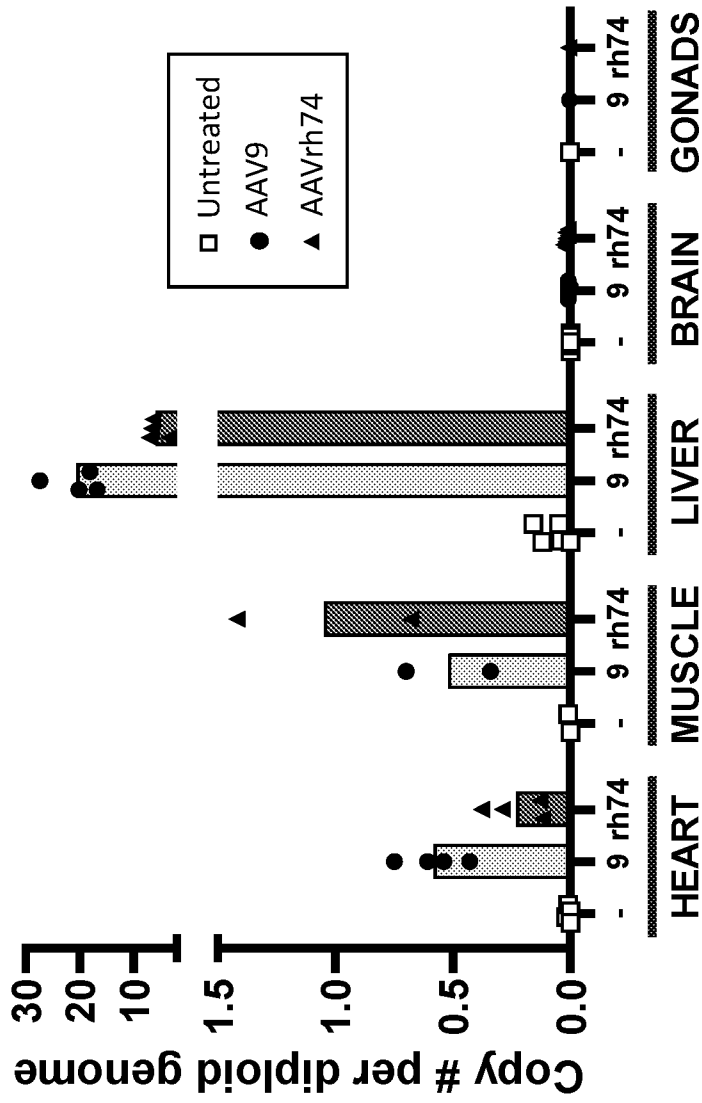


FIG. 2

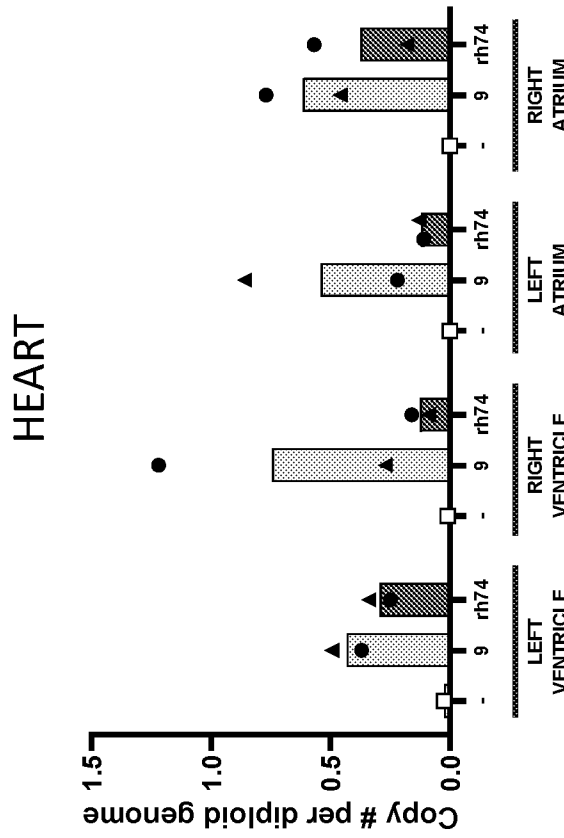
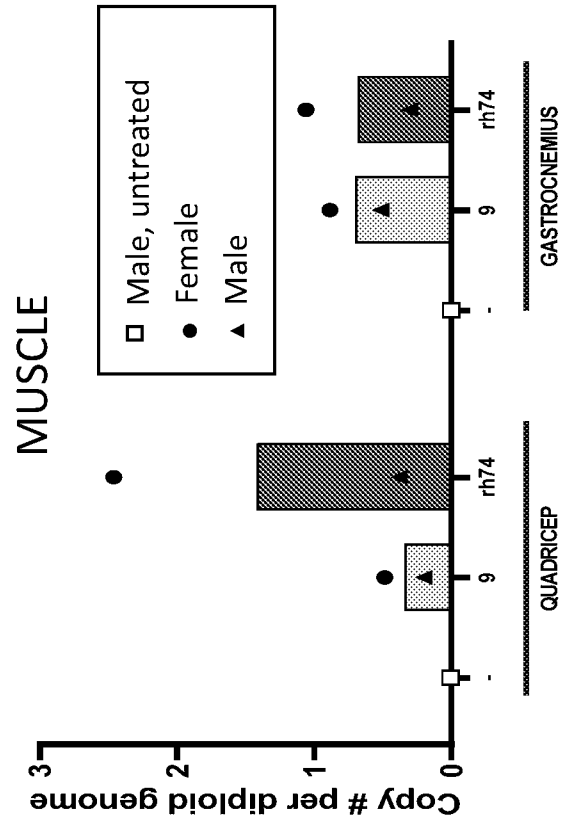


FIG. 3B

FIG. 3A

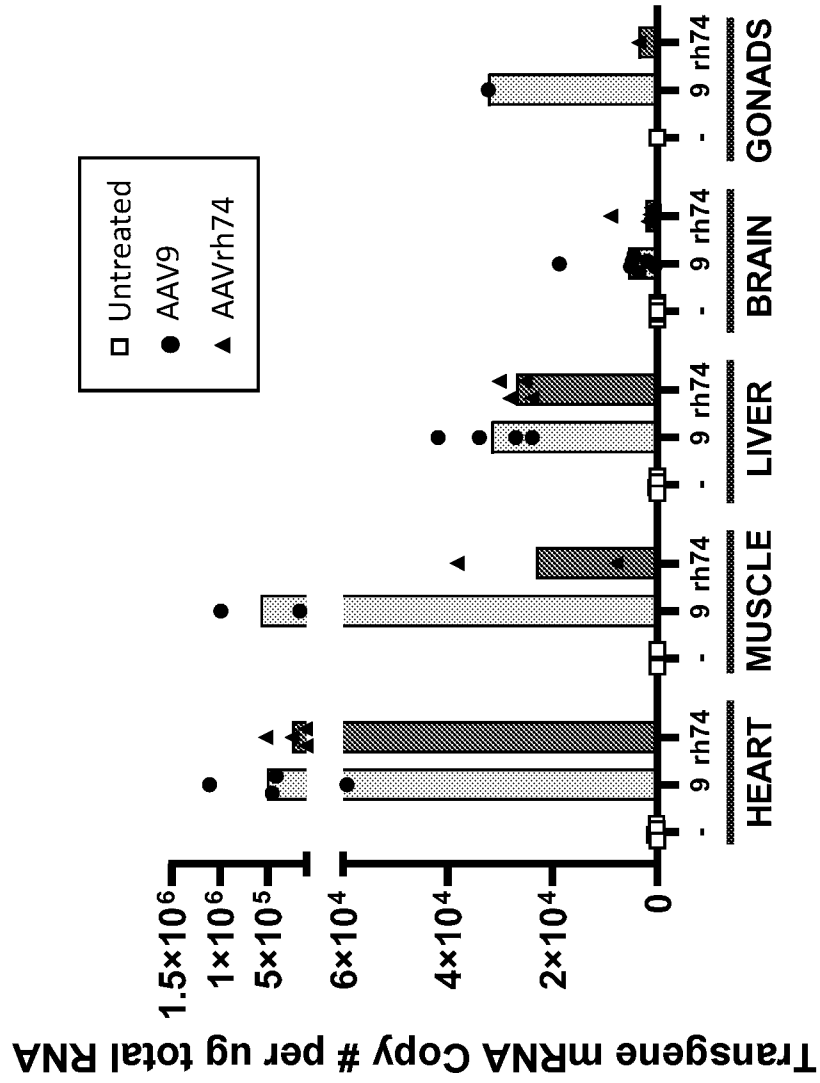


FIG. 4

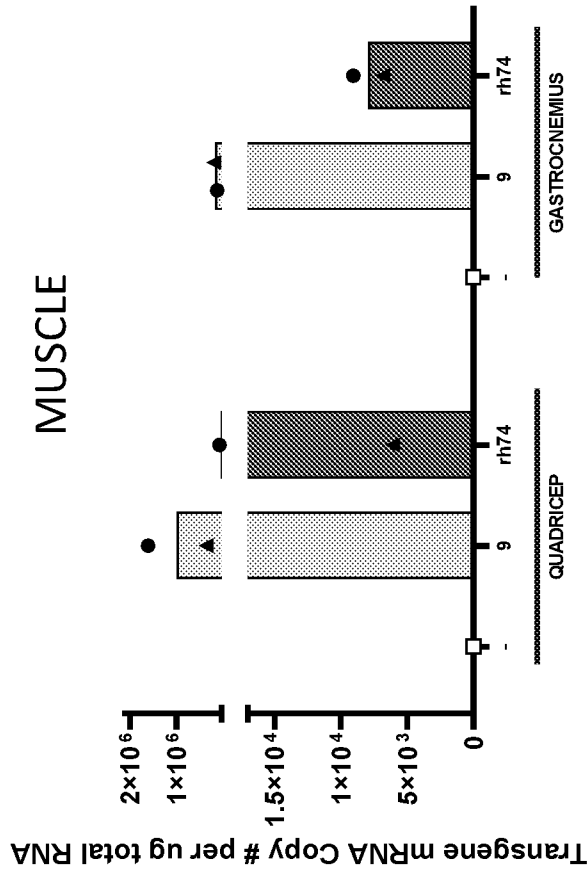


FIG. 5B

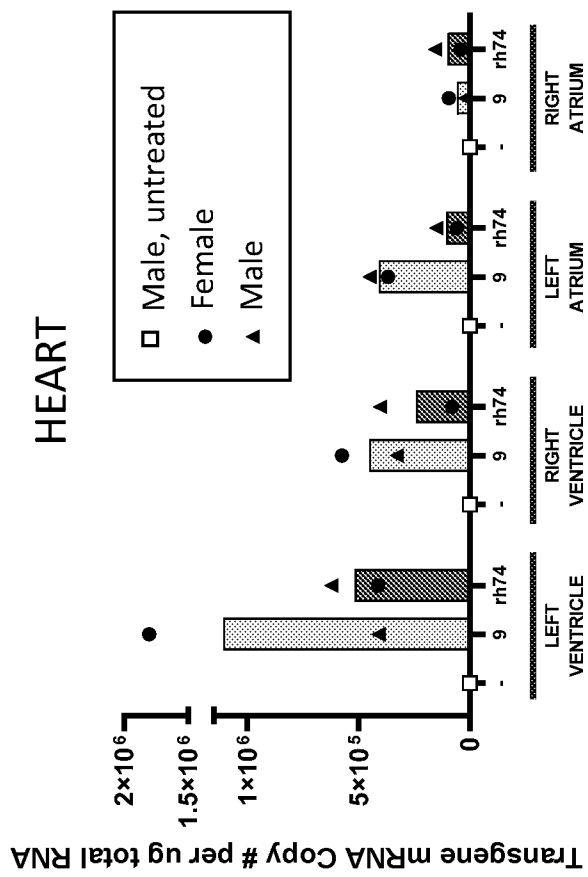
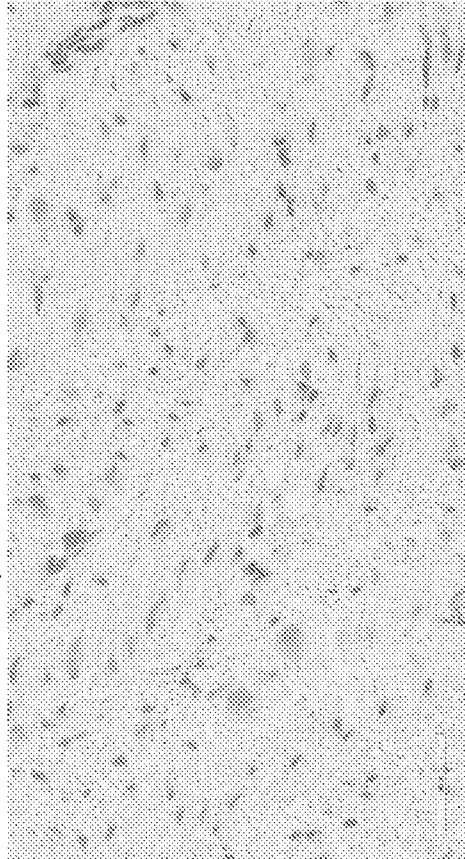


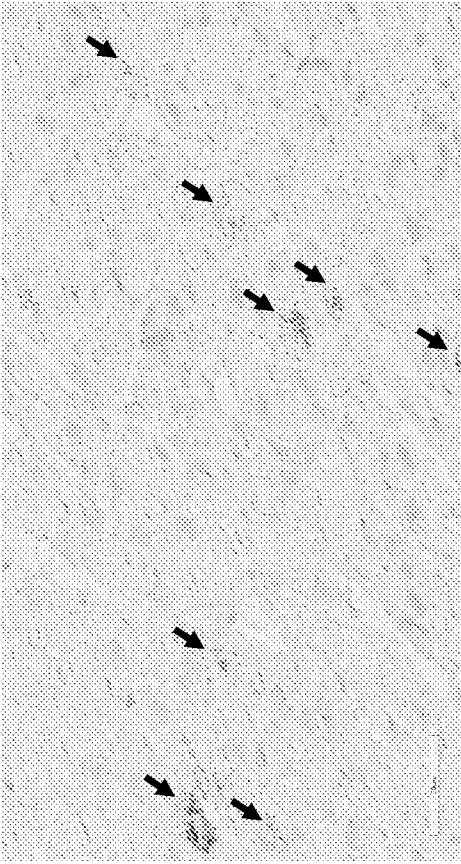
FIG. 5A

B059: Male, Untreated – 0%

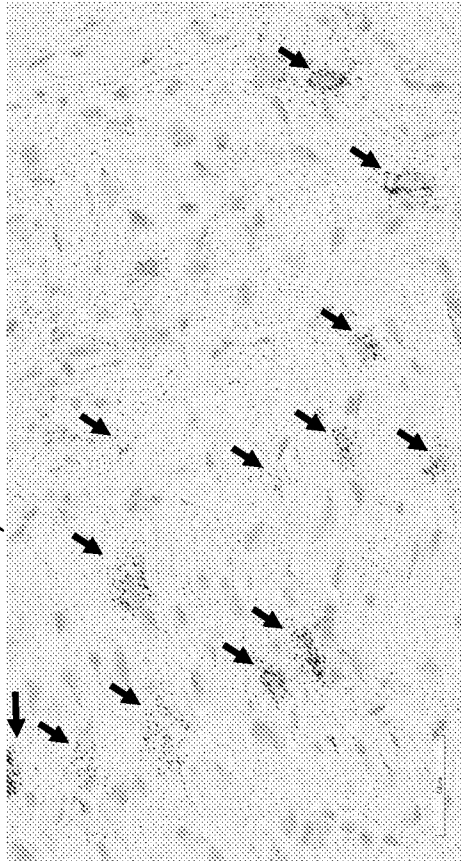


**FIG. 6A**

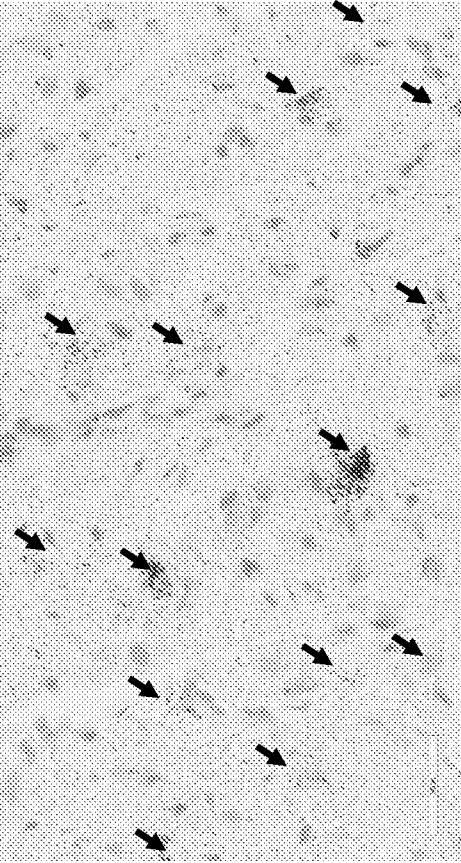
A710: Female, AAVrh.74 – 1-25%



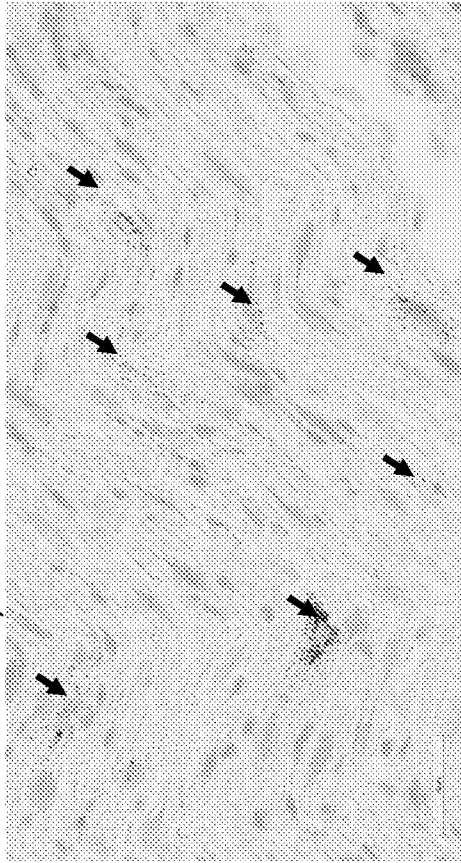
A981: Male, AAVrh.74 – 26-50%



A991: Female, AAV9 – 26-50%



A602: Male, AAV9 – 1-25%



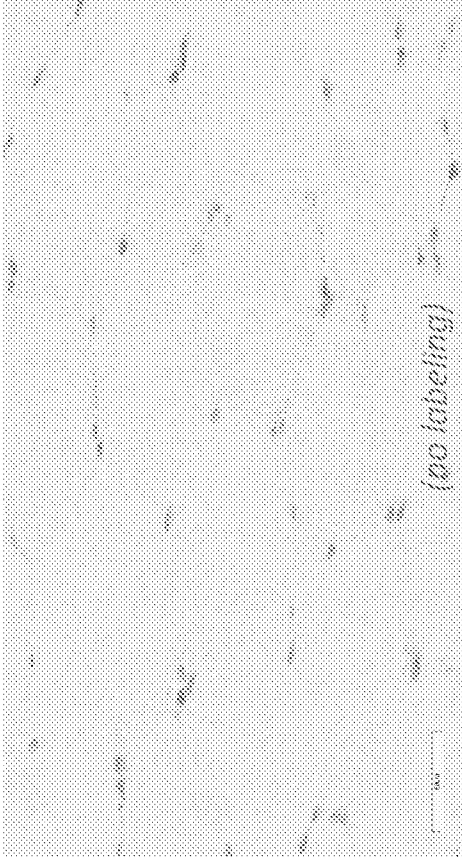
**FIG. 6B**

B059: Male, Untreated – 0%



**FIG. 7A**

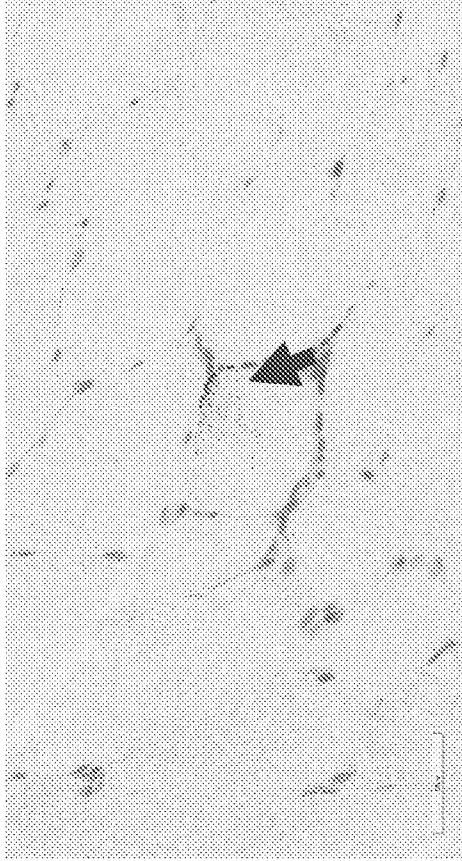
A710: Female, AAVrh74



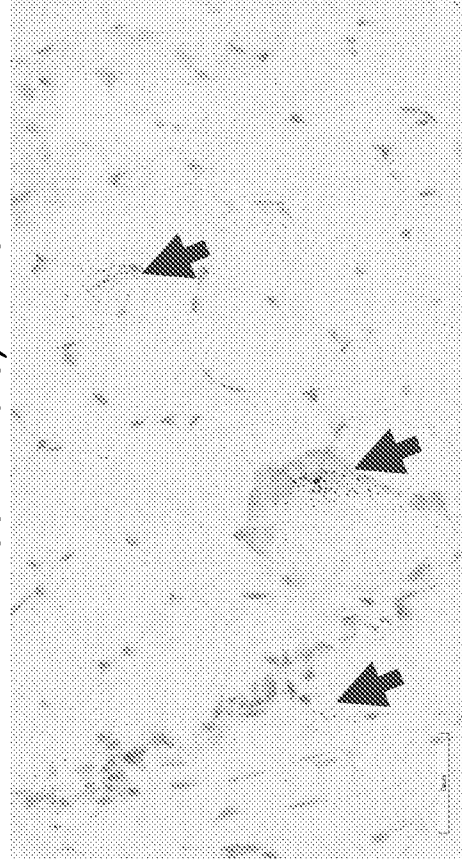
A981: Male, AAVrh74



A991: Female, AAV9



A602: Male, AAV9



**FIG. 7B**

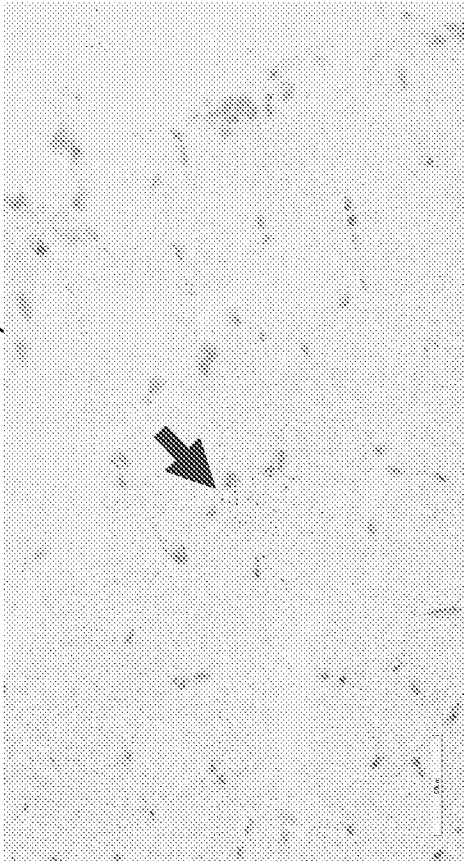
A710: Female, AAVrh74



A981: Male, AAVrh74



A991: Female, AAV9



A602: Male, AAV9



**FIG. 8**

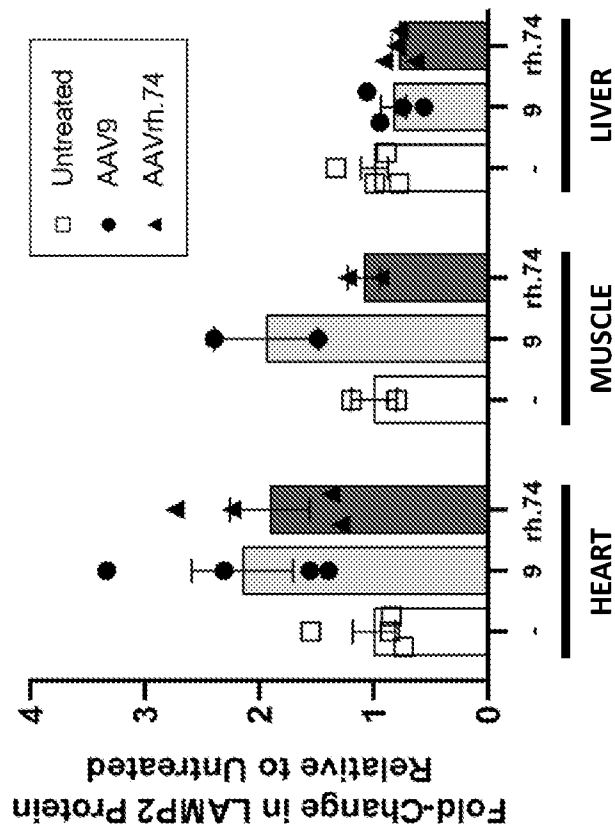


FIG. 9

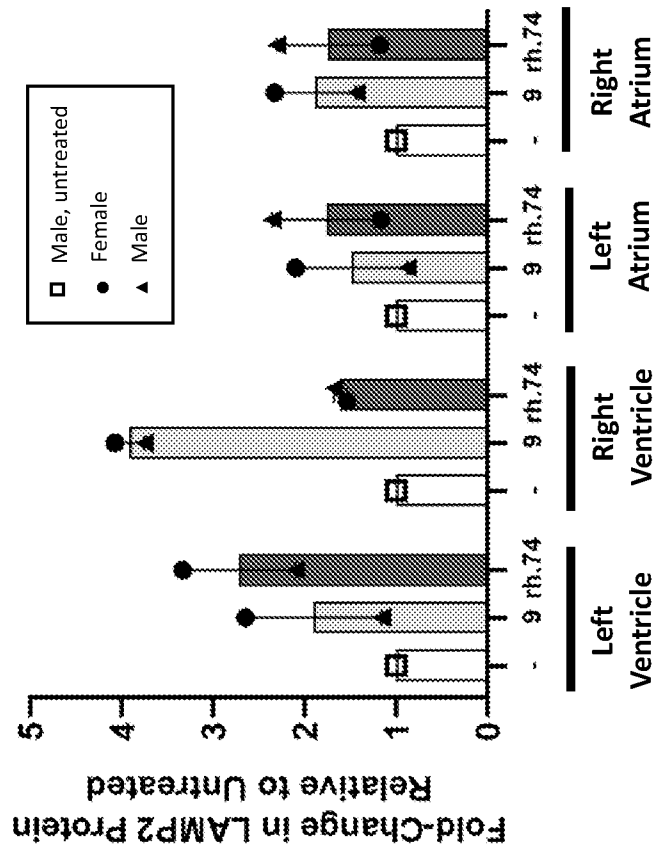
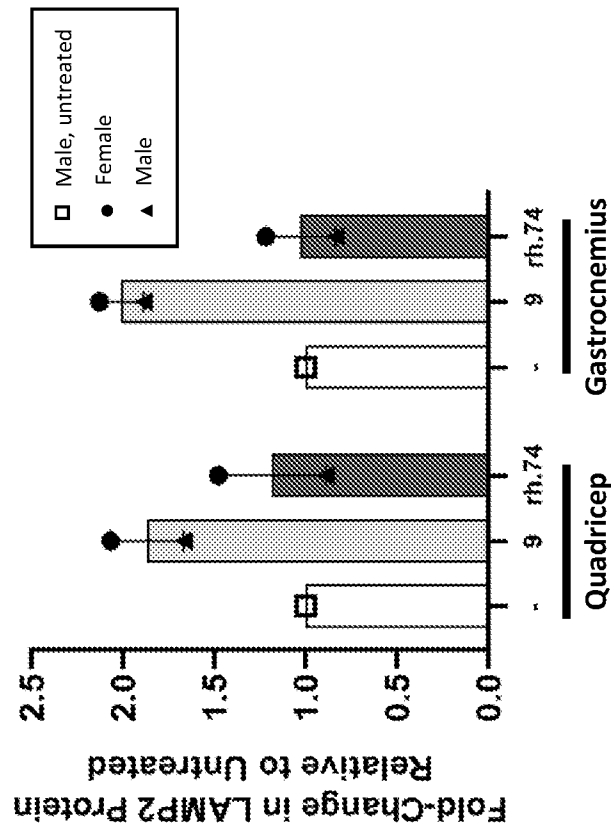


FIG. 10A



**FIG. 10B**

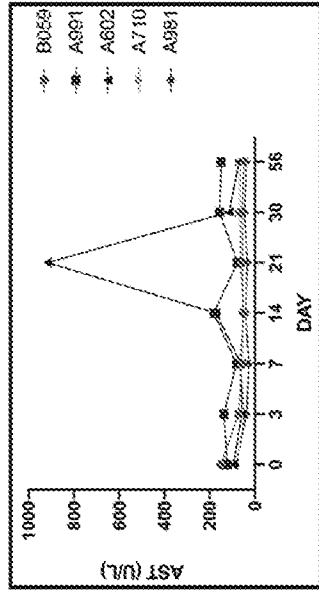


FIG. 11B

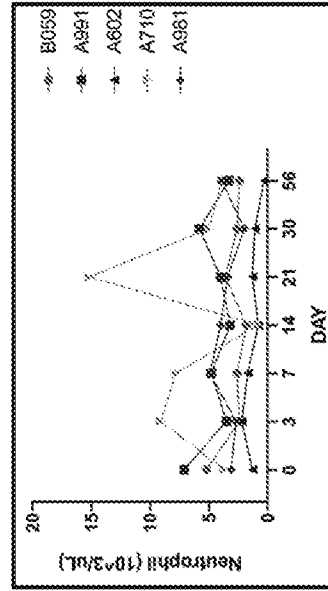


FIG. 11D

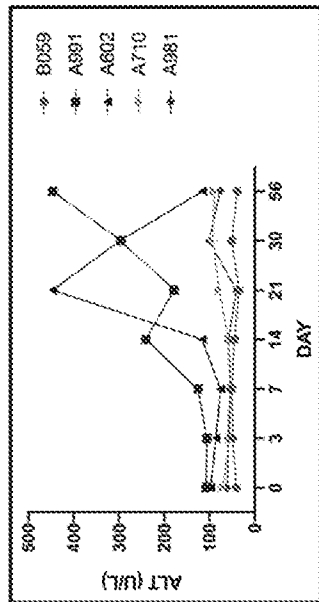


FIG. 11A

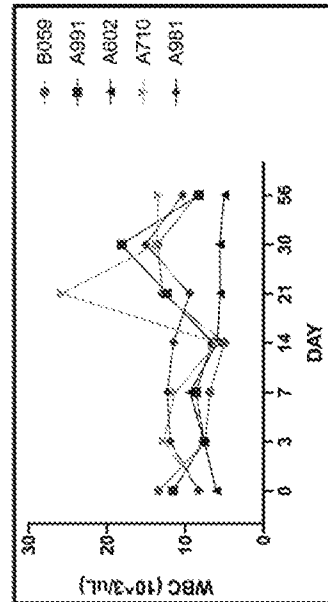


FIG. 11C

SEQUENCE LISTING

<110> Rocket Pharmaceuticals  
 Keravala, Annahita  
 Prabhakar, Raj  
 Shah, Gaurav  
 Wong, Roderick  
 Yalamanchi, Naveen  
 Pratumswan, Piratip

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<151> 2019-02-12

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Tyr Phe Asp Phe Asn Arg Phe His Cys His Phe Ser Pro Arg Asp Trp  
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Cys Leu Pro Pro Phe Pro Ala Asp Val Phe Met Ile Pro Gln Tyr Gly  
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Asn Pro Val Ala Thr Glu Gln Tyr Gly Val Val Ala Asp Asn Leu Gln  
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Gln Gln Asn Ala Ala Pro Ile Val Gly Ala Val Asn Ser Gln Gly Ala  
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Lys Leu Ala Ser Phe Ile Thr Gln Tyr Ser Thr Gly Gln Val Ser Val  
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485 490 495

Glu Ile Gln Tyr Thr Ser Asn Tyr Tyr Lys Ser Thr Asn Val Asp Phe  
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Val Pro Phe His Ser Ser Tyr Ala His Ser Gln Ser Leu Asp Arg Leu  
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Cys Tyr Arg Gln Gln Arg Val Ser Thr Thr Leu Ser Gln Asn Asn Asn  
145 150 155 160

Ser Asn Phe Ala Trp Thr Gly Ala Thr Lys Tyr His Leu Asn Gly Arg  
165 170 175

Asp Ser Leu Val Asn Pro Gly Val Ala Met Ala Thr His Lys Asp Asp  
180 185 190

Glu Glu Arg Phe Phe Pro Ser Ser Gly Val Leu Met Phe Gly Lys Gln  
195 200 205

Gly Ala Gly Lys Asp Asn Val Asp Tyr Ser Ser Val Met Leu Thr Ser  
210 215 220

Glu Glu Glu Ile Lys Thr Thr Asn Pro Val Ala Thr Glu Gln Tyr Gly  
225 230 235 240

Val Val Ala Asp Asn Leu Gln Gln Gln Asn Ala Ala Pro Ile Val Gly  
245 250 255

Ala Val Asn Ser Gln Gly Ala Leu Pro Gly Met Val Trp Gln Asn Arg  
260 265 270

Asp Val Tyr Leu Gln Gly Pro Ile Trp Ala Lys Ile Pro His Thr Asp

275

280

285

Gly Asn Phe His Pro Ser Pro Leu Met Gly Gly Phe Gly Leu Lys His  
290 295 300

Pro Pro Pro Gln Ile Leu Ile Lys Asn Thr Pro Val Pro Ala Asp Pro  
305 310 315 320

Pro Thr Thr Phe Asn Gln Ala Lys Leu Ala Ser Phe Ile Thr Gln Tyr  
325 330 335

Ser Thr Gly Gln Val Ser Val Glu Ile Glu Trp Glu Leu Gln Lys Glu  
340 345 350

Asn Ser Lys Arg Trp Asn Pro Glu Ile Gln Tyr Thr Ser Asn Tyr Tyr  
355 360 365

Lys Ser Thr Asn Val Asp Phe Ala Val Asn Thr Glu Gly Thr Tyr Ser  
370 375 380

Glu Pro Arg Pro Ile Gly Thr Arg Tyr Leu Thr Arg Asn Leu  
385 390 395

<210> 4  
<211> 332  
<212> PRT  
<213> Non-human primate adeno-associated virus

<400> 4

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Phe His Ser Ser Tyr Ala His Ser Gln Ser Leu Asp Arg Leu Met Asn  
20 25 30

Pro Leu Ile Asp Gln Tyr Leu Tyr Tyr Leu Ser Arg Thr Gln Ser Thr  
35 40 45

Gly Gly Thr Ala Gly Thr Gln Gln Leu Leu Phe Ser Gln Ala Gly Pro  
50 55 60

Asn Asn Met Ser Ala Gln Ala Lys Asn Trp Leu Pro Gly Pro Cys Tyr  
65 70 75 80

Arg Gln Gln Arg Val Ser Thr Thr Leu Ser Gln Asn Asn Asn Ser Asn  
85 90 95

Phe Ala Trp Thr Gly Ala Thr Lys Tyr His Leu Asn Gly Arg Asp Ser  
100 105 110

Leu Val Asn Pro Gly Val Ala Met Ala Thr His Lys Asp Asp Glu Glu  
115 120 125

Arg Phe Phe Pro Ser Ser Gly Val Leu Met Phe Gly Lys Gln Gly Ala  
130 135 140

Gly Lys Asp Asn Val Asp Tyr Ser Ser Val Met Leu Thr Ser Glu Glu  
145 150 155 160

Glu Ile Lys Thr Thr Asn Pro Val Ala Thr Glu Gln Tyr Gly Val Val  
165 170 175

Ala Asp Asn Leu Gln Gln Gln Asn Ala Ala Pro Ile Val Gly Ala Val  
180 185 190

Asn Ser Gln Gly Ala Leu Pro Gly Met Val Trp Gln Asn Arg Asp Val  
195 200 205

Tyr Leu Gln Gly Pro Ile Trp Ala Lys Ile Pro His Thr Asp Gly Asn  
210 215 220

Phe His Pro Ser Pro Leu Met Gly Gly Phe Gly Leu Lys His Pro Pro  
225 230 235 240

Pro Gln Ile Leu Ile Lys Asn Thr Pro Val Pro Ala Asp Pro Pro Thr  
245 250 255

Thr Phe Asn Gln Ala Lys Leu Ala Ser Phe Ile Thr Gln Tyr Ser Thr  
260 265 270

Gly Gln Val Ser Val Glu Ile Glu Trp Glu Leu Gln Lys Glu Asn Ser  
275 280 285

Lys Arg Trp Asn Pro Glu Ile Gln Tyr Thr Ser Asn Tyr Tyr Lys Ser  
290 295 300

Thr Asn Val Asp Phe Ala Val Asn Thr Glu Gly Thr Tyr Ser Glu Pro  
305 310 315 320

Arg Pro Ile Gly Thr Arg Tyr Leu Thr Arg Asn Leu  
325 330

<210> 5  
<211> 409  
<212> PRT  
<213> Homo sapiens

<400> 5

Met Val Cys Phe Arg Leu Phe Pro Val Pro Gly Ser Gly Leu Val Leu  
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Val Cys Leu Val Leu Gly Ala Val Arg Ser Tyr Ala Leu Glu Leu Asn  
20 25 30

Leu Thr Asp Ser Glu Asn Ala Thr Cys Leu Tyr Ala Lys Trp Gln Met  
35 40 45

Asn Phe Thr Val Arg Tyr Glu Thr Thr Asn Lys Thr Tyr Lys Thr Val  
50 55 60

Thr Ile Ser Asp His Gly Thr Val Thr Tyr Asn Gly Ser Ile Cys Gly  
65 70 75 80

Asp Asp Gln Asn Gly Pro Lys Ile Ala Val Gln Phe Gly Pro Gly Phe  
85 90 95

Ser Trp Ile Ala Asn Phe Thr Lys Ala Ala Ser Thr Tyr Ser Ile Asp  
100 105 110

Ser Val Ser Phe Ser Tyr Asn Thr Gly Asp Asn Thr Thr Phe Pro Asp  
115 120 125

Ala Glu Asp Lys Gly Ile Leu Thr Val Asp Glu Leu Leu Ala Ile Arg

130

135

140

Ile Pro Leu Asn Asp Leu Phe Arg Cys Asn Ser Leu Ser Thr Leu Glu  
145 150 155 160

Lys Asn Asp Val Val Gln His Tyr Trp Asp Val Leu Val Gln Ala Phe  
165 170 175

Val Gln Asn Gly Thr Val Ser Thr Asn Glu Phe Leu Cys Asp Lys Asp  
180 185 190

Lys Thr Ser Thr Val Ala Pro Thr Ile His Thr Thr Val Pro Ser Pro  
195 200 205

Thr Thr Thr Pro Thr Pro Lys Glu Lys Pro Glu Ala Gly Thr Tyr Ser  
210 215 220

Val Asn Asn Gly Asn Asp Thr Cys Leu Leu Ala Thr Met Gly Leu Gln  
225 230 235 240

Leu Asn Ile Thr Gln Asp Lys Val Ala Ser Val Ile Asn Ile Asn Pro  
245 250 255

Asn Thr Thr His Ser Thr Gly Ser Cys Arg Ser His Thr Ala Leu Leu  
260 265 270

Arg Leu Asn Ser Ser Thr Ile Lys Tyr Leu Asp Phe Val Phe Ala Val  
275 280 285

Lys Asn Glu Asn Arg Phe Tyr Leu Lys Glu Val Asn Ile Ser Met Tyr  
290 295 300

Leu Val Asn Gly Ser Val Phe Ser Ile Ala Asn Asn Asn Leu Ser Tyr  
305 310 315 320

Trp Asp Ala Pro Leu Gly Ser Ser Tyr Met Cys Asn Lys Glu Gln Thr  
325 330 335

Val Ser Val Ser Gly Ala Phe Gln Ile Asn Thr Phe Asp Leu Arg Val  
340 345 350

Gln Pro Phe Asn Val Thr Gln Gly Lys Tyr Ser Thr Ala Gln Glu Cys  
355 360 365

Ser Leu Asp Asp Asp Thr Ile Leu Ile Pro Ile Ile Val Gly Ala Gly  
370 375 380

Leu Ser Gly Leu Ile Ile Val Ile Val Ile Ala Tyr Val Ile Gly Arg  
385 390 395 400

Arg Lys Ser Tyr Ala Gly Tyr Gln Thr  
405

<210> 6

<211> 1233

<212> DNA

<213> Homo sapiens

<400> 6

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tgcctttatg caaaatggca gatgaatttc acagttcgct atgaaactac aaataaaact 180  
tataaaactg taaccatttc agaccatggc actgtgacat ataatggaag catttgtggg 240  
gatgatcaga atggtcccaa aatagcagtg cagttcggac ctggcttttc ctggattgcg 300  
aathttacca aggcagcatc tacttattca attgacagcg tctcattttc ctacaacact 360  
ggtgataaca caacatttcc tgatgctgaa gataaaggaa ttcttactgt tgatgaactt 420  
ttggccatca gaattccatt gaatgacctt tttagatgca atagtttatc aactttggaa 480  
aagaatgatg ttgtccaaca ctactgggat gttcttgtac aagcttttgt ccaaaatggc 540  
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ggaacctatt cagttaataa tggcaatgat acttgtctgc tggctaccat ggggctgcag 720  
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tgggatgccc ccctgggaag ttcttatatg tgcaacaaag agcagactgt ttcagtgtct	1020
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aagtattcta cagcccaaga gtgttcgctg gatgatgaca ccattctaata cccaattata	1140
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agaaaaagt atgctggata tcagactctg taa	1233

<210> 7  
 <211> 1233  
 <212> DNA  
 <213> Artificial Sequence

<220>  
 <223> Made in Lab - Engineered Lamp-2B coding sequence

<400> 7	
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tgcctgtatg ccaagtggca gatgaacttc actgtgagat atgagactac caacaagacc	180
tacaagactg tgaccatctc agaccatggc actgtcacct acaatggatc aatctgtggt	240
gatgatcaga atggcccaaa gatagcagtg cagtttgggc ccggtttttc ctggattgct	300
aacttcacca aggcagcctc cacctacagc attgactcag tcagcttcag ctacaacact	360
ggggataaca ccaccttccc tgacgcagag gacaaggga tccttactgt ggacgaactc	420
ctggcaatca gaatccccct taacgacctg ttcagatgca actccccttc aacccttgaa	480
aagaatgatg tggtgcaaca ctattgggac gtcctgggtgc aagcctttgt gcagaatggg	540
acagtgagta ccaacgagtt cctctgtgac aaggacaaga ccagcactgt ggccccact	600
atccacacca ctgtgcccag ccctaccact acccccaccc ctaaagagaa gccagaagct	660
ggaacctact cagtcaaaa tggaatgac acatgcctcc ttgccaccat gggactgcag	720
ctgaacatca ctcaggaaa ggtggcctca gtgattaaca tcaaccctaa caccactcat	780
agcactggga gctgcagatc acatacagct ctgctgaggc tcaactcctc caccatcaag	840
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aagtactcca ctgcacaaga gtgttccttg gatgatgaca ctatcctcat cccattatt 1140  
gtgggagctg gactgtcagg attgattata gtgattgtga ttgcttatgt gattggaagg 1200  
agaaagagct atgctggcta ccagaccctg taa 1233

<210> 8  
<211> 1233  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Made in Lab - Engineered Lamp-2B coding sequence

<400> 8  
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ctgggggctg tcagaagcta tgccttggag ctgaacctca ctgatagtga aatgccact 120  
tgtctgtatg ctaagtggca gatgaacttc actgtgagat atgaaaccac caacaagact 180  
tacaaaacag tgaccatctc agatcatgga actgtgacct acaacggcag catttgtgga 240  
gacgaccaga acggaccaa aatcgctgtc caatttgggc ctggattctc ctggattgcc 300  
aatttacta aagctgcctc cacatattca attgactcag tgtccttctc ctacaacact 360  
ggggacaaca ctactttccc tgatgctgaa gataagggaa tcttgacagt ggatgagctg 420  
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aagaacgacg tggcgcagca ctactgggac gtgctgggtc aggcctttgt gcagaacggc 540  
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ggaacctact ctgtgaacaa tggcaatgat acctgtctgt tggccaccat gggcctccaa 720  
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aagtatagca ctgcccagga gtgctccctg gatgatgaca ccattctgat tccaatcatt 1140  
gtgggtgcag gactttctgg gcttattatt gtgattgtga ttgcctatgt gattggcaga 1200  
aggaaatcct atgcagggtta ccaaactctg taa 1233

<210> 9  
<211> 1233  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Made in Lab - Engineered Lamp-2B coding sequence

<400> 9  
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cttggagctg tcagaagcta tgcctggag ctgaacctga ctgactcaga aatgcccact 120  
tgcctgtatg ccaagtggca gatgaacttc actgtcagat atgaaaccac caacaagacc 180  
tataagactg tgaccatctc agaccatggc actgtgactt acaatgggtc aatttgtgga 240  
gatgaccaga atggccctaa gatagctgtc cagtttggtc caggattcag ctggattgcc 300  
aattcacca aggagccag cacctacagc attgactctg tgtccttctc ctacaacaca 360  
ggagacaaca ccactttccc tgatgcagag gacaaaggta tcctgactgt ggatgagttg 420  
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aagaatgatg tgggtgcagca ctattgggat gtgctagtcc aggcctttgt ccagaatggg 540  
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aagtacagca ctgctcagga gtgcagcctg gatgatgaca ctatcctgat ccctatcatt 1140

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<210> 10  
<211> 4549  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Made in Lab - Engineered expression cassette

<400> 10  
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aggggttcct tgtagttaat gattaaccg ccatgctact tatctaccag ggtaatgggg 180  
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aactgcccac ttggcagtac atcaagtgta tcatatgcca agtacgccc ctattgacgt 480  
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tacttggcag tacatctacg tattagtcac cgctattacc atggtcgagg tgagccccac 600  
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<210> 11  
<211> 4549  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Made in Lab - Engineered expression cassette

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<212> DNA  
<213> Artificial Sequence

<220>  
<223> Made in Lab - Engineered expression cassette

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<213> Adeno-associated viurs

<400> 13  
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aggggttcct 130

<210> 14  
<211> 130  
<212> DNA  
<213> Adeno-associated virus

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<210> 15  
<211> 410  
<212> PRT  
<213> Homo sapiens

<400> 15

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Leu Thr Asp Ser Glu Asn Ala Thr Cys Leu Tyr Ala Lys Trp Gln Met  
35 40 45

Asn Phe Thr Val Arg Tyr Glu Thr Thr Asn Lys Thr Tyr Lys Thr Val  
50 55 60

Thr Ile Ser Asp His Gly Thr Val Thr Tyr Asn Gly Ser Ile Cys Gly  
65 70 75 80

Asp Asp Gln Asn Gly Pro Lys Ile Ala Val Gln Phe Gly Pro Gly Phe  
85 90 95

Ser Trp Ile Ala Asn Phe Thr Lys Ala Ala Ser Thr Tyr Ser Ile Asp  
100 105 110

Ser Val Ser Phe Ser Tyr Asn Thr Gly Asp Asn Thr Thr Phe Pro Asp  
115 120 125

Ala Glu Asp Lys Gly Ile Leu Thr Val Asp Glu Leu Leu Ala Ile Arg  
130 135 140

Ile Pro Leu Asn Asp Leu Phe Arg Cys Asn Ser Leu Ser Thr Leu Glu  
145 150 155 160

Lys Asn Asp Val Val Gln His Tyr Trp Asp Val Leu Val Gln Ala Phe  
165 170 175

Val Gln Asn Gly Thr Val Ser Thr Asn Glu Phe Leu Cys Asp Lys Asp  
180 185 190

Lys Thr Ser Thr Val Ala Pro Thr Ile His Thr Thr Val Pro Ser Pro  
195 200 205

Thr Thr Thr Pro Thr Pro Lys Glu Lys Pro Glu Ala Gly Thr Tyr Ser  
210 215 220

Val Asn Asn Gly Asn Asp Thr Cys Leu Leu Ala Thr Met Gly Leu Gln  
225 230 235 240

Leu Asn Ile Thr Gln Asp Lys Val Ala Ser Val Ile Asn Ile Asn Pro  
245 250 255

Asn Thr Thr His Ser Thr Gly Ser Cys Arg Ser His Thr Ala Leu Leu  
260 265 270

Arg Leu Asn Ser Ser Thr Ile Lys Tyr Leu Asp Phe Val Phe Ala Val

275

280

285

Lys Asn Glu Asn Arg Phe Tyr Leu Lys Glu Val Asn Ile Ser Met Tyr  
290 295 300

Leu Val Asn Gly Ser Val Phe Ser Ile Ala Asn Asn Asn Leu Ser Tyr  
305 310 315 320

Trp Asp Ala Pro Leu Gly Ser Ser Tyr Met Cys Asn Lys Glu Gln Thr  
325 330 335

Val Ser Val Ser Gly Ala Phe Gln Ile Asn Thr Phe Asp Leu Arg Val  
340 345 350

Gln Pro Phe Asn Val Thr Gln Gly Lys Tyr Ser Thr Ala Gln Asp Cys  
355 360 365

Ser Ala Asp Asp Asp Asn Phe Leu Val Pro Ile Ala Val Gly Ala Ala  
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Lys His His His Ala Gly Tyr Glu Gln Phe  
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<210> 16  
<211> 410  
<212> PRT  
<213> Homo sapiens

<400> 16

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Val Cys Leu Val Leu Gly Ala Val Arg Ser Tyr Ala Leu Glu Leu Asn  
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Leu Thr Asp Ser Glu Asn Ala Thr Cys Leu Tyr Ala Lys Trp Gln Met  
35 40 45

Asn Phe Thr Val Arg Tyr Glu Thr Thr Asn Lys Thr Tyr Lys Thr Val  
50 55 60

Thr Ile Ser Asp His Gly Thr Val Thr Tyr Asn Gly Ser Ile Cys Gly  
65 70 75 80

Asp Asp Gln Asn Gly Pro Lys Ile Ala Val Gln Phe Gly Pro Gly Phe  
85 90 95

Ser Trp Ile Ala Asn Phe Thr Lys Ala Ala Ser Thr Tyr Ser Ile Asp  
100 105 110

Ser Val Ser Phe Ser Tyr Asn Thr Gly Asp Asn Thr Thr Phe Pro Asp  
115 120 125

Ala Glu Asp Lys Gly Ile Leu Thr Val Asp Glu Leu Leu Ala Ile Arg  
130 135 140

Ile Pro Leu Asn Asp Leu Phe Arg Cys Asn Ser Leu Ser Thr Leu Glu  
145 150 155 160

Lys Asn Asp Val Val Gln His Tyr Trp Asp Val Leu Val Gln Ala Phe  
165 170 175

Val Gln Asn Gly Thr Val Ser Thr Asn Glu Phe Leu Cys Asp Lys Asp  
180 185 190

Lys Thr Ser Thr Val Ala Pro Thr Ile His Thr Thr Val Pro Ser Pro  
195 200 205

Thr Thr Thr Pro Thr Pro Lys Glu Lys Pro Glu Ala Gly Thr Tyr Ser  
210 215 220

Val Asn Asn Gly Asn Asp Thr Cys Leu Leu Ala Thr Met Gly Leu Gln  
225 230 235 240

Leu Asn Ile Thr Gln Asp Lys Val Ala Ser Val Ile Asn Ile Asn Pro  
245 250 255

Asn Thr Thr His Ser Thr Gly Ser Cys Arg Ser His Thr Ala Leu Leu  
260 265 270

Arg Leu Asn Ser Ser Thr Ile Lys Tyr Leu Asp Phe Val Phe Ala Val  
275 280 285

Lys Asn Glu Asn Arg Phe Tyr Leu Lys Glu Val Asn Ile Ser Met Tyr  
290 295 300

Leu Val Asn Gly Ser Val Phe Ser Ile Ala Asn Asn Asn Leu Ser Tyr  
305 310 315 320

Trp Asp Ala Pro Leu Gly Ser Ser Tyr Met Cys Asn Lys Glu Gln Thr  
325 330 335

Val Ser Val Ser Gly Ala Phe Gln Ile Asn Thr Phe Asp Leu Arg Val  
340 345 350

Gln Pro Phe Asn Val Thr Gln Gly Lys Tyr Ser Thr Ala Gln Glu Cys  
355 360 365

Ser Leu Asp Asp Asp Thr Ile Leu Ile Pro Ile Ile Val Gly Ala Gly  
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385 390 395 400

Arg Lys Ser Tyr Ala Gly Tyr Gln Thr Leu  
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<210> 17  
<211> 411  
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<213> Homo sapiens

<400> 17

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Leu Thr Asp Ser Glu Asn Ala Thr Cys Leu Tyr Ala Lys Trp Gln Met

35

40

45

Asn Phe Thr Val Arg Tyr Glu Thr Thr Asn Lys Thr Tyr Lys Thr Val  
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Thr Ile Ser Asp His Gly Thr Val Thr Tyr Asn Gly Ser Ile Cys Gly  
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Asp Asp Gln Asn Gly Pro Lys Ile Ala Val Gln Phe Gly Pro Gly Phe  
85 90 95

Ser Trp Ile Ala Asn Phe Thr Lys Ala Ala Ser Thr Tyr Ser Ile Asp  
100 105 110

Ser Val Ser Phe Ser Tyr Asn Thr Gly Asp Asn Thr Thr Phe Pro Asp  
115 120 125

Ala Glu Asp Lys Gly Ile Leu Thr Val Asp Glu Leu Leu Ala Ile Arg  
130 135 140

Ile Pro Leu Asn Asp Leu Phe Arg Cys Asn Ser Leu Ser Thr Leu Glu  
145 150 155 160

Lys Asn Asp Val Val Gln His Tyr Trp Asp Val Leu Val Gln Ala Phe  
165 170 175

Val Gln Asn Gly Thr Val Ser Thr Asn Glu Phe Leu Cys Asp Lys Asp  
180 185 190

Lys Thr Ser Thr Val Ala Pro Thr Ile His Thr Thr Val Pro Ser Pro  
195 200 205

Thr Thr Thr Pro Thr Pro Lys Glu Lys Pro Glu Ala Gly Thr Tyr Ser  
210 215 220

Val Asn Asn Gly Asn Asp Thr Cys Leu Leu Ala Thr Met Gly Leu Gln  
225 230 235 240

Leu Asn Ile Thr Gln Asp Lys Val Ala Ser Val Ile Asn Ile Asn Pro  
245 250 255

Asn Thr Thr His Ser Thr Gly Ser Cys Arg Ser His Thr Ala Leu Leu  
260 265 270

Arg Leu Asn Ser Ser Thr Ile Lys Tyr Leu Asp Phe Val Phe Ala Val  
275 280 285

Lys Asn Glu Asn Arg Phe Tyr Leu Lys Glu Val Asn Ile Ser Met Tyr  
290 295 300

Leu Val Asn Gly Ser Val Phe Ser Ile Ala Asn Asn Asn Leu Ser Tyr  
305 310 315 320

Trp Asp Ala Pro Leu Gly Ser Ser Tyr Met Cys Asn Lys Glu Gln Thr  
325 330 335

Val Ser Val Ser Gly Ala Phe Gln Ile Asn Thr Phe Asp Leu Arg Val  
340 345 350

Gln Pro Phe Asn Val Thr Gln Gly Lys Tyr Ser Thr Ala Glu Glu Cys  
355 360 365

Ser Ala Asp Ser Asp Leu Asn Phe Leu Ile Pro Val Ala Val Gly Val  
370 375 380

Ala Leu Gly Phe Leu Ile Ile Val Val Phe Ile Ser Tyr Met Ile Gly  
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Arg Arg Lys Ser Arg Thr Gly Tyr Gln Ser Val  
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<211> 1730

<212> DNA

<213> Artificial Sequence

<220>

<223> Made in Lab - promoter sequence

<400> 18

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cgcccaacga cccccgcca ttgacgtcaa taatgacgta tgttcccata gtaacgccaa	180
tagggacttt ccattgacgt caatgggtgg agtattttacg gtaaactgcc cacttggcag	240
tacatcaagt gtatcatatg ccaagtacgc cccctattga cgtcaatgac ggtaaattggc	300
ccgcctggca ttatgcccag tacatgacct tatgggactt tcctacttgg cagtacatct	360
acgtattagt catcgctatt accatggctg aggtgagccc cacgttctgc ttcactctcc	420
ccatctcccc cccctcccca cccccaattt tgtattttatt tatttttttaa ttattttgtg	480
cagcgatggg ggcggggggg gggggggggc gcgcgccagg cggggcgggg cggggcgagg	540
ggcggggcgg ggcgaggcgg agaggtgcgg cggcagccaa tcagagcggc gcgctccgaa	600
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gggagggcct tcgtgcgtcg ccgcgccgcc gtccccttct ccctctccag cctcggggct	1560
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<210> 19  
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<212> DNA  
<213> Woodchuck hepatitis virus

<400> 19  
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aattactagt tcaggtgtat tgccacaaga caaacatggt aagaaacttt cccgttattt 180  
acgctctggt cctgttaatc aacctctgga ttacaaaatt tgtgaaagat tgactgatat 240  
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tgctattgct tcccgtacgg ctttcgtttt ctctccttg tataaatcct ggttgctgtc 360  
tctttatgag gagttgtggc ccgttgccg tcaacgtggc gtgggtgtgct ctgtgtttgc 420  
tgacgcaacc cccactggct ggggcattgc caccacctgt caactccttt ctgggacttt 480  
cgctttcccc ctcccgatcg ccacggcaga actcatgcc gcctgccttg cccgctgctg 540  
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<210> 20  
<211> 13  
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<213> Artificial Sequence

<220>  
<223> Kozak consensus sequence

<400> 20  
gccgccacca tgg 13

<210> 21  
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<212> RNA  
<213> Artificial Sequence

<220>  
<223> Kozak consensus sequence

<400> 21  
gccgccrcca ugg 13

<210> 22  
<211> 8

<212> RNA  
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<220>  
<221> misc\_feature  
<222> (3)..(4)  
<223> n is a, c, g, or u

<220>  
<221> misc\_feature  
<222> (8)..(8)  
<223> n is a, c, g, or u

<400> 22  
agnnaugn

8

<210> 23  
<211> 7  
<212> RNA  
<213> Artificial Sequence

<220>  
<223> Kozak consensus sequence

<220>  
<221> misc\_feature  
<222> (2)..(3)  
<223> n is a, c, g, or u

<400> 23  
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7

<210> 24  
<211> 7  
<212> RNA  
<213> Artificial Sequence

<220>  
<223> Kozak consensus sequence

<400> 24  
accaugg

7

<210> 25  
<211> 10  
<212> RNA

<213> Artificial Sequence

<220>

<223> Kozak consensus sequence

<400> 25

gacaccaugg 10

<210> 26

<211> 387

<212> DNA

<213> Mus musculus

<400> 26

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actcgggaagg acatatggga gggcaaatca tttaaaacat cagaatgagt atttggttta 120

gagtttggca acatatgccc atatgctggc tgccatgaac aaaggttggc tataaagagg 180

tcatcagtat atgaaacagc cccctgctgt ccattcctta ttccatagaa aagccttgac 240

ttgaggttag atTTTTTTta tattttgttt tgtgttattt ttttctttaa catcccataa 300

atTTTcctta catgTTTTac tagccagatt tttcctcctc tcctgactac tcccagtcac 360

agctgtccct cttctcttat ggagatc 387

<210> 27

<211> 235

<212> DNA

<213> Bos taurus

<400> 27

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accctggaag gtgccactcc cactgtcctt tcctaataaa atgaggaaat tgcacgcat 120

tgtctgagta ggtgtcattc tattctgggg ggtgggggtgg ggcaggacag caagggggag 180

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<210> 28

<211> 222

<212> DNA

<213> Simian viurs 40

<400> 28

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aatgctttat ttgtgaaatt tgtgatgcta ttgctttatt tgtaaccatt ataagctgca 120

ataaacaagt taacaacaac aattgcattc attttatgtt tcaggttcag ggggagatgt 180  
gggaggtttt ttaaagcaag taaaacctct acaaatgtgg ta 222

<210> 29  
<211> 202  
<212> DNA  
<213> Homo sapiens

<400> 29  
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gtgtccttct ataatattat ggggtggagg ggggtggat ggagcaagg gccaagttg 180  
ggaagaaacc tgtaggcct gc 202

<210> 30  
<211> 21  
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<213> Artificial Sequence

<220>  
<223> Primer

<400> 30  
atcatgctat tgcttcccgt a 21

<210> 31  
<211> 20  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Primer

<400> 31  
gggccacaac tcctcataaa 20

<210> 32  
<211> 29  
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<220>  
<223> Primer

<400> 32

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<210> 33  
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<220>  
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gggccacaac tcctcataaa 20

<210> 35  
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<220>  
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<400> 35  
cctccttgta taaatcctgg ttgctgtct 29