



US00PP32459P3

(12) **United States Plant Patent**  
**Mehlenbacher et al.**

(10) **Patent No.:** **US PP32,459 P3**

(45) **Date of Patent:** **Nov. 17, 2020**

(54) **CORYLUS PLANT NAMED ‘POLLYO’**

(50) Latin Name: ***Corylus avellana* cultivar**  
Varietal Denomination: **PollyO**

(71) Applicant: **Oregon State University**, Corvallis,  
OR (US)

(72) Inventors: **Shawn A. Mehlenbacher**, Corvallis,  
OR (US); **David C. Smith**, Corvallis,  
OR (US); **Rebecca L. McCluskey**,  
Corvallis, OR (US)

(73) Assignee: **Oregon State University**, Corvallis,  
OR (US)

(\* ) Notice: Subject to any disclaimer, the term of this  
patent is extended or adjusted under 35  
U.S.C. 154(b) by 120 days.

(21) Appl. No.: **15/998,094**

(22) Filed: **Jun. 29, 2018**

(65) **Prior Publication Data**

US 2020/0008334 P1 Jan. 2, 2020

(51) **Int. Cl.**  
**A01H 5/08** (2018.01)  
**A01H 6/00** (2018.01)

(52) **U.S. Cl.**  
USPC ..... **Plt./152**  
CPC ..... **A01H 6/00** (2018.05); **A01H 5/08**  
(2013.01)

(58) **Field of Classification Search**

USPC ..... Plt./152  
CPC ..... A01H 5/00; A01H 5/08; A01H 5/0825  
See application file for complete search history.

(56) **References Cited**

PUBLICATIONS

[https://www.capitalpress.com/state/oregon/new-hazelnut-variety-nanned-for-industry-official/article\\_076b181c-449a-56cb-b0f8-60645024787f.html](https://www.capitalpress.com/state/oregon/new-hazelnut-variety-nanned-for-industry-official/article_076b181c-449a-56cb-b0f8-60645024787f.html); Jan. 22, 2018; 3 pages.\*

\* cited by examiner

*Primary Examiner* — Kent L Bell

(74) *Attorney, Agent, or Firm* — Klarquist Sparkman,  
LLP

(57) **ABSTRACT**

A new and distinct *Corylus* plant named ‘PollyO’ is characterized by high nut yield, early nut maturity, small round nuts and kernels, high kernel percentage, good kernel blanching and excellent flavor. The tree is vigorous and has a desirable growth habit. ‘PollyO’ is resistant to bud mite (primarily *Phytoptus avellanae* Nal.). DNA markers and field exposure indicate that it is highly resistant to eastern filbert blight (EFB) caused by *Anisogramma anomala* (Peck) E. Müller.

**12 Drawing Sheets**

**Specification includes a Sequence Listing.**

**1**

**ACKNOWLEDGMENT OF GOVERNMENT  
SUPPORT**

This invention was made with government support under  
USDA/ARS 58-5358-4-025, USDA-NIFA-SCRI 2009-  
51181-06028, USDA-NIFA-AFRI 2014-67013-22421, and  
USDA-NIFA-SCRI 2016-04991 awarded by the United  
States Department of Agriculture. The government has cer-  
tain rights in the invention.

**INCORPORATION OF ELECTRONIC  
SEQUENCE LISTING**

The nucleic and amino acid sequences listed in the  
accompanying sequence listing are shown using standard  
letter abbreviations for nucleotide bases, and three letter  
code for amino acids, as defined in 37 C.F.R. 1.822. The  
Sequence Listing is submitted as an ASCII text file, created  
on Aug. 6, 2018, 8 KB, which is incorporated by reference  
herein.

**BACKGROUND**

The present Invention relates to a new and distinct culti-  
var of *Corylus* plant, botanically known as *Corylus avellana*,  
and hereinafter referred to by the name ‘PollyO’.

**2**

The new *Corylus* resulted from a controlled cross of OSU  
684.104×OSU 669.104 made in 2001 (FIG. 1); the progeny  
was designated 01021. OSU 684.104 is from a cross of ‘Birk  
5-39’ (unpatented) and ‘Mortarella’ (unpatented). ‘Birk  
5-39’, an OSU selection growing in a small plot near Canby,  
Oreg., is from a cross of OSU 14.084 (‘Barcelona’×‘Davi-  
ana’, both unpatented) and ‘Negret’ (unpatented). OSU  
669.104 is a full sib of the pollinizer ‘Zeta’ (unpatented).  
OSU 669.104 carries a dominant allele for a very high level  
of resistance to eastern filbert blight (EFB) from the grower  
selection ‘Zimmerman’ (unpatented). Microsatellite marker  
analysis indicates that ‘Zimmerman’ is from a cross of  
‘Barcelona’ and the EFB-resistant pollinizer ‘Gasaway’  
(Gökirmak et al., Characterization of European hazelnut  
(*Corylus avellana*) cultivars using SSR markers. Genetic  
Resources and Crop Evolution 56:147-172, 2009) (un-  
patented). The pedigree of ‘PollyO’ includes germplasm  
from Spain (‘Barcelona’, ‘Casina’, and ‘Negret’, all  
unpatented), Italy (‘Mortarella’ and ‘Montebello’, both  
unpatented) and England (‘Daviana’).

Hybrid seeds from the controlled cross were harvested in  
August 2001, stratified, and the resulting seedlings grown in  
a glasshouse during the summer of 2002. 99 seedlings from  
this cross were planted in the field in Corvallis, Oreg. in  
October 2002. The designation OSU 1108.001 indicates the  
row and tree location of the original seedling. Nuts were first

observed on the original seedling in September 2006. The nuts were harvested from the original seedling tree and evaluated over four years (2006-2009).

'PollyO' was propagated by tie-off layerage of the suckers in the summer beginning in 2008. The rooted layers from the first propagation (2008) were lined out in a nursery row the year after layerage (2009), and used to plant a replicated yield trial the following spring (2010). The first trial included four trees of each variety. EFB-susceptible selections and check cultivars 'Barcelona', 'Clark' and 'Sacajawea' (unpatented) (Mehlenbacher et al., 2008. 'Sacajawea' hazelnut. *HortScience* 43:255-257) were planted on the same date in a trial adjacent to the first trial. Fungicides were applied to the trial of susceptible genotypes to reduce the incidence of EFB. The rooted layers from the second propagation (2009) were lined out in a nursery row in 2010 and used to plant a second trial in 2011. The second trial had seven blocks. Both trials were planted as randomized complete block designs with a single tree of each genotype in each block. EFB-resistant cultivars 'Jefferson' (unpatented) (Mehlenbacher et al., 2011. 'Jefferson' hazelnut. *HortScience* 46:662-664), 'Yamhill' (unpatented) (Mehlenbacher et al., 2009. 'Yamhill' hazelnut. *HortScience* 44:845-847) and 'Gamma' (unpatented) (Mehlenbacher and Smith. 2004. Hazelnut pollenizers 'Gamma', 'Delta', 'Epsilon' and 'Zeta'. *HortScience* 39:1498-1499) served as checks in the first trial, while 'Jefferson' and 'Yamhill' were the checks in the second trial. Both trials included additional EFB-resistant selections.

The new cultivar was asexually reproduced by tie-off layerage of the suckers in the summer of 2008 and 2009 in Corvallis, Oreg. It was moved with a tree spade to the layer beds in spring, 2010. Of the 12 layers harvested in 2008, three were well-rooted, seven had a modest number of roots, and two had no roots. The size (caliper) was rated as medium. Layers harvested in 2009 showed improved rooting. Further propagation by layerage of about 28 suckers per year was performed in 2011-14, of which 39% were rated as well-rooted, 26% with fair rooting, 26% with poor rooting and 9% with no roots. In general, the layers rooted sufficiently well and size (caliper) was medium to small. The unique features of this new *Corylus* are stable and reproduced true-to-type in successive generations of asexual reproduction.

#### SUMMARY

The following traits have been observed and are determined to be the unique characteristics of 'PollyO'. 'PollyO' is well-suited to the blanched kernel market for use in chocolate products and baked goods. 'PollyO' combines high nut yield, early nut maturity, small round nuts and kernels, high kernel percentage, good kernel blanching and excellent flavor. The tree is vigorous and has a desirable growth habit. 'PollyO' is resistant to bud mite (primarily *Phytoptus avellanae* Nal.). DNA markers and field exposure indicate that it is highly resistant to eastern filbert blight (EFB) caused by *Anisogramma anomala* (Peck) E. Müller. The resistance is conferred by a dominant allele from 'Zimmerman', which microsatellite markers indicate is from a cross of 'Barcelona'×'Gasaway' (Gökirmak et al., Characterization of European hazelnut (*Corylus avellana*) cultivars using SSR markers. Genetic Resources and Crop Evolution 56:147-172, 2009). EFB is now present throughout the Willamette Valley where 99% of the U.S. hazelnut crop

is grown. Pruning to remove cankers and fungicide applications are currently used to manage the disease in orchards of 'Barcelona' and other susceptible cultivars. 'PollyO' is suitable for planting in areas with high disease pressure, as are previous releases 'Yamhill' (2008, unpatented), 'Jefferson' (2009, unpatented), 'Dorris' (2012, US Plant Patent No. PP25,022) (Mehlenbacher et al., 2013. 'Dorris' hazelnut. *HortScience* 48:796-799), 'Wepster' (2013, US Plant Patent No. PP27,141) (Mehlenbacher et al., 2014. 'Wepster' hazelnut. *HortScience* 49:346-349) and 'McDonald' (2014, US Plant Patent No. PP28,200) (Mehlenbacher et al., 2016. 'McDonald' hazelnut. *HortScience* 51:757-760).

'PollyO' has incompatibility alleles S<sub>2</sub> and S<sub>10</sub>. Both alleles are expressed in the stigmas, but only S<sub>10</sub> is expressed in the pollen because of dominance. Female receptivity is early, with 'Wepster' and 'McDonald', while pollen is shed by 'PollyO' about ten days later than the other two. Pollen germination is high and similar to 'Wepster' and 'McDonald'. Recommended pollinizers include 'Wepster' (S<sub>1</sub> S<sub>2</sub>), 'McDonald' (S<sub>2</sub> S<sub>15</sub>) and 'York' (S<sub>2</sub> S<sub>21</sub>) (US Plant Patent No. PP24,972).

'PollyO' can serve as a companion for 'Wepster' and 'McDonald', and an alternative to 'Yamhill'. Many growers find 'Yamhill' trees difficult to manage because of low vigor, and the kernels do not blanch well. 'Wepster' and 'McDonald' are cross-compatible with 'PollyO' in all directions, giving growers additional options in orchard design. All three cultivars ('Wepster', 'McDonald' and 'PollyO') could be planted together with each serving as a pollinizer for the others.

Comparisons in two replicated trials conducted in Corvallis, Oreg., plants of 'PollyO' differed from plants of other cultivars and selections of *Corylus avellana* known to the Inventors primarily in nut yield, time of nut maturity, nut size, and kernel percentage (ratio of kernel weight to nut weight). For example:

Nut yield (kg per tree): First trial: 'PollyO' (29.39), 'Jefferson' (24.26), and 'Yamhill' (26.47); second trial 'PollyO' (23.46), 'Jefferson' (16.91), and 'Yamhill' (21.69);

Nut maturity: 'PollyO' nuts mature 10-14 days earlier than 'Barcelona', a few days earlier than 'Yamhill';

Nut size (L×W×D): 'PollyO' (19.5 mm×19.8 mm×17.2 mm), 'McDonald' (18.4 mm×18.9 mm×17.9 mm), 'Wepster' (18.3 mm×19 mm×16.6 mm) (see also FIG. 14); and

Kernel Percentage: 'PollyO' (47.9%), 'McDonald' (51.5%), 'Wepster' (43.9%), 'Felix' (50.1%), 'Dorris' (43.0%), and 'Barcelona' (43.0%).

#### BRIEF DESCRIPTION OF THE DRAWINGS

The accompanying colored photographs illustrate the overall appearance of the new cultivar, showing the colors as true as it is reasonably possible to obtain in colored reproductions of this type. Foliage colors in the photographs may differ slightly from the color values cited in the detailed botanical description which accurately describe the colors of the new *Corylus*.

FIG. 1 provides the pedigree of the 'PollyO' (OSU 1108.001) hazelnut tree. The pedigree of the female parent OSU 684.104 and the male parent OSU 669.104 are shown.

FIG. 2 shows a 'PollyO' (OSU 1108.001) hazelnut tree at the end of the seventh growing season. The pole height is 10 feet.

FIG. 3 shows nuts, blanched kernels and raw kernels of ‘Barcelona’ (left) and ‘PollyO’ (OSU 1108.001) (right).

FIG. 4 shows nuts and husks of ‘PollyO’ hazelnut.

FIGS. 5 and 6 show husks and nuts of ‘Barcelona’ (left) and ‘PollyO’ (OSU 1108.001) (right).

FIG. 7 is a graph showing the cumulative percentage of nuts harvested from the ground on four dates in fall 2016. Nuts were harvested by hand, dried and weighed. Nuts were harvested in the first trial planted in 2010. Nut maturity of ‘PollyO’ (OSU 1108.001) is a few days earlier than ‘Yamhill’ or 10-14 days before ‘Barcelona’.

FIG. 8 is a graph showing the estimated percentage of nuts on the ground for eight hazelnut cultivars on seven dates in fall 2017. Reading on the 60% drop line from left to right, the cultivars from earliest to latest nut maturity are: ‘McDonald’, ‘Sacajawea’ (unpatented), ‘PollyO’ (OSU 1108.001), ‘Yamhill’ (unpatented), ‘York’, ‘Wepster’, ‘Dorris’ and ‘Jefferson’. ‘McDonald’, ‘Sacajawea’, ‘York’, ‘Wepster’ and ‘Dorris’ were in the guard row planted at the same time as the trial.

FIG. 9 is a graph showing the cumulative percentage of nuts harvested (based on weight) from the ground for four hazelnut cultivars on five dates in fall 2017. ‘McDonald’ trees were in the guard row planted at the same time as the trial. Nuts of ‘PollyO’ (OSU 1108.001) mature after those of ‘McDonald’ but before those of ‘Yamhill’.

FIG. 10 shows the time of female receptivity (bottom, red) and pollen shed (top, green) of ‘PollyO’ (OSU 1108.001) and other hazelnut cultivars in a trial planted in 2011 (December 2014-March 2015).

FIG. 11 shows the time of female receptivity (bottom, red) and pollen shed (top, green) of ‘PollyO’ (OSU 1108.001) and other hazelnut cultivars in a trial planted in 2011 (December 2015-March 2016).

FIG. 12 shows the time of female receptivity (bottom, red) and pollen shed (top, green) of ‘PollyO’ (OSU 1108.001) and other hazelnut cultivars in a trial planted in 2011 (December 2016-March 2017).

FIG. 13 shows the time of female receptivity (bottom, red) and pollen shed (top, green) of ‘PollyO’ (OSU 1108.001) and other hazelnut cultivars in a trial planted in 2011 (December 2017-March 2018).

FIG. 14 shows nuts, blanched kernels and raw kernels of ‘Wepster’, ‘McDonald’, and ‘PollyO’ (OSU 1108.001).

#### SEQUENCE LISTING

The nucleic acid sequences listed in the accompanying sequence listing are shown using standard letter abbreviations for nucleotide bases as defined in 37 C.F.R. 1.822. Only one strand of each nucleic acid sequence is shown, but the complementary strand is understood as included by any reference to the displayed strand. In the accompanying sequence listing:

SEQ ID NOS: 1-40 are primer sequences that can be used for genetic fingerprinting.

#### DETAILED DESCRIPTION

The cultivar ‘PollyO’ has not been observed under all possible environmental conditions. The phenotype may vary somewhat with variations in environment such as temperature and light intensity, without, however, any variance in genotype. The aforementioned photographs and following observations and measurements describe trees grown in Corvallis, Oreg. under commercial practice outdoors in the

field during the fall, winter and spring. Plants used for the photographs and description were propagated by tie-off layerage and growing on their own roots, and seven or eight years old. In the following description, color references are made to The Royal Horticultural Society Colour Chart, 1966 Edition, except where general terms of ordinary dictionary significance are used. The list of UPOV descriptors are from the Mar. 28, 1979 Hazelnut guidelines from UPOV, unless otherwise noted.

Botanical Classification:

*Corylus avellana* cultivar ‘PollyO’.

Parentage:

Female, or seed, parent: *Corylus avellana* cultivar OSU 684.104

Male, or pollen, parent: *Corylus avellana* cultivar OSU 669.104

Comparison to Parents:

‘PollyO’ differs from both parents for alleles at the incompatibility alleles. ‘PollyO’ differs from its female parent OSU 684.104 in that it is resistant to eastern filbert blight. ‘PollyO’ nuts mature 17 days before those of its male parent OSU 669.104.

Propagation (Type Rooted Suckers):

Time to initiate roots: About 30 days at 20° C.

Time to produce a rooted young plant: About six months at 22° C.

Root description. Fine to thick; freely branching; creamy white in color.

Propagation (Type Whip Grafting):

Time to budbreak on the scions: About 14 days at 25° C.

Time to produce a grafted plant: About six months at 25° C.

Plant Description:

General appearance: natural habit is a perennial shrub, but in commercial orchards, is a single trunk tree. Upright-spreading plant habit.

Growth and branching habit: Freely branching; about 15 lateral branches develop per plant. Pinching, that is, removal of the terminal apices, enhances branching with lateral branches potentially forming at every node.

Vigor: Strong vigor

Growth habit. semi-erect

Size: Plant height is about 5 meters; plant diameter or spread is about 5 meters.

Trunk diameter at the end of the seventh leaf, averaged over two trials=12.14 cm.

Trunk color: 177D.

Trunk texture: smooth.

Lateral Branch Description:

Length: About 45.6 cm.

Diameter: About 4.4 mm.

Internode length: About 3.4 mm.

Texture: Smooth, glabrous.

Strength: Strong.

Color, immature: 139D.

Color, mature: 177D.

Foliage Description:

Arrangement: Alternate, simple.

Length: About 13.3 cm.

Width: About 11.8 cm.

Shape: Oblong to ovate.

Apex: Obtuse to acute.

Base: Cordate.  
 Margin: Serrate.  
 Texture, upper and lower surfaces: Slightly pubescent.  
 Venation pattern: Pinnate.  
 Color: Developing foliage, upper surface 141C, lower surfaces: 139C. Fully expanded foliage, upper surface: Spring, summer, and fall 146A; Fully expanded foliage, lower surface: Spring, summer, and fall, 146B. Venation, upper surface: Spring, summer, and fall, 146A. Venation, lower surface: Spring, summer, and fall, 146D.  
 Leaf Bud Description:  
 Shape: ovoid  
 Time of leaf budbreak: descriptor=6 (medium to late).  
 Color: 143C  
 Length is 6.4 mm, and width is 4.0 mm.  
 Petiole Description:  
 Length: About 16.3 mm.  
 Diameter: About 1.6 mm.  
 Texture, upper and lower surfaces: Pubescent.  
 Color, upper surface: Spring, summer, and fall, 146C.  
 Color, lower surface: Spring, summer, and fall, 146D.  
 Flower Description:  
 Male inflorescences are catkins, color prior to elongation 138B.  
 Catkin length: 20.1 mm  
 Catkin width (diameter): 5.4 mm  
 Female inflorescence style color 047B (with reference to the Voss and Hale, HortScience 33:13-17, 1998).  
 Time of female flowering: descriptor=6 (medium to late)  
 Time of female flowering compared to male flowering: protogyny (descriptor=2, same time).  
 Involucre constriction: absent  
 Involucre length: 1.25 times length of nut.  
 Size of indentation: descriptor=5 (medium)  
 Strength of serration of indentation: descriptor=5 (medium)  
 Pubescence on husk: descriptor=9 (present)  
 Thickness of callus at base: descriptor=7 (thick)  
 Jointing of bracts: descriptor=1 (absent)  
 Female inflorescences are modified leaf buds, with a color of 143C.  
 Nut Description:  
 Length: About 19.5 mm.  
 Width: About 19.8 mm.  
 Depth: About 17.2 mm.  
 Nut shape: Round. Nut shape index  $[(\text{Width}+\text{Depth})/2*\text{Length}]=0.950$ . Nut compression index  $(\text{Width}/\text{Depth})=1.15$ .  
 Nut shell color: 164A.  
 Nut shell texture: smooth.  
 Nut weight: About 2.88 grams.  
 Kernel weight: About 1.38 grams.  
 Kernel percentage (kernel weight/nut weight): About 47.9%.  
 Kernel length: from base to apex 19.2 mm.  
 Kernel width: at widest point 19.3 mm.  
 Kernel depth: perpendicular to width 17.4 mm.  
 Kernel texture: smooth.  
 Kernel pellicle color: brown 165C.  
 Number of fruits per cluster: 4.  
 Number of stripes on shell: descriptor=3 (few).  
 Prominence of fruit apex: slightly prominent, descriptor=3.  
 Size of fruit pistil scar on shell: small, descriptor=3.  
 Hairiness of top of fruit: weak to medium.

Curvature of nut basal scar: plane.  
 Double kernels: absent.  
 Kernel shape: globular.  
 Lateral groove in kernel: absent.  
 Skin of kernel: descriptor=5 (medium corky).  
 Nut yield (kg per tree): total 29.39 kg per tree in first trial, 23.46 kg per tree in second trial.  
 Storability of fruits: excellent, similar to OSU releases and check cultivars  
 Disease/Pest Resistance:  
 Plants of ‘PollyO’ are highly resistant to eastern filbert blight caused by the fungus *Anisogramma anomala* (Peck) E. Müller, although a few small cankers may develop under high disease pressure. Plants of ‘PollyO’ are resistant to bud mites (*Phytoptus avellanae* Nal.), while plants of ‘Tonda Gentile delle Langhe’ are highly susceptible, and plants of ‘Barcelona’ are highly resistant.  
 Temperature Tolerance:  
 Plants of ‘PollyO’ have been observed to tolerate temperatures from -21 to 38° C. in the field in Corvallis, Ore.  
 Comparative Data:  
 Tree Size, Growth Habit, Yield, and Yield Efficiency.  
 Tree sizes in the two trials were estimated by measuring trunk circumference 30 cm above the soil line, at the end of the 7<sup>th</sup> growing season (December 2016 and November 2017, respectively). Trunk cross-sectional area (TCA) was calculated from trunk circumference. In the first trial, TCA of ‘PollyO’ (OSU 1108.001) was 118 cm<sup>2</sup> or 126% of ‘Jefferson’ and 91% of ‘Barcelona’ in the adjacent trial of EFB-susceptible selections. In the second trial, TCA of ‘PollyO’ (OSU 1108.001) was 117.6 cm<sup>2</sup>, or 146% of ‘Jefferson’. In previous trials, TCA of ‘Jefferson’ has been about 65-70% of the vigorous standard ‘Barcelona’, a size that pleases Oregon growers. Trees of ‘PollyO’ (OSU 1108.001) are vigorous, and their globose growth habit (FIG. 2) should be grower-friendly (easy to manage).  
 In the first trial, mean total nut yield (2013-16) of ‘PollyO’ (OSU 1108.001) was 29.4 kg compared to 24.3 kg for ‘Jefferson’ (Tables 1-2). Yield efficiency, the ratio of yield to TCA, adjusts yield for differences in tree size. Yield efficiency of ‘PollyO’ (OSU 1108.001) was 0.251 kg/cm<sup>2</sup> which is not significantly different from ‘Jefferson’ (0.261 kg/cm<sup>2</sup>). In the second trial, mean total nut yield was 23.46 kg for ‘PollyO’ (OSU 1108.001) which is significantly higher than the 16.91 kg for ‘Jefferson’ (Tables 3-4). Yield efficiency was 0.201 kg/cm<sup>2</sup> for ‘PollyO’ (OSU 1108.001) which is not significantly different from ‘Jefferson’ (0.210 kg/cm<sup>2</sup>).

TABLE 1

Nut yield of ‘PollyO’ (OSU 1108.001) in comparison with other hazelnut cultivars and selections in the first trial, planted in 2010. Randomized complete block design with four blocks and a single tree of each selection in each block.

Selection	CvNo	Nut Yield per tree (kg)				
		2013	2014	2015	2016	Total
EFB-resistant selections						
1069.092	1	1.06	2.76	3.54	7.93	15.29
1072.105	2	0.98	3.14	4.65	8.39	17.14
1081.101	3	1.03	4.02	4.23	7.45	16.73
1084.075	4	1.07	2.27	4.58	5.81	13.73
1085.066	5	2.57	3.97	6.24	8.47	21.25

TABLE 1-continued

Nut yield of 'PollyO' (OSU 1108.001) in comparison with other hazelnut cultivars and selections in the first trial, planted in 2010. Randomized complete block design with four blocks and a single tree of each selection in each block.

Selection	CvNo	Nut Yield per tree (kg)				
		2013	2014	2015	2016	Total
1086.145	6	1.45	4.97	5.77	10.64	22.82
1102.032	7	1.02	3.86	3.83	8.29	17.00
1102.069	8	1.50	3.48	4.84	7.48	17.30
1108.001	9	2.69	7.21	8.07	11.42	29.39
1119.081	10	2.59	4.88	5.46	7.84	20.77
1130.006	11	1.59	4.44	4.36	8.22	18.61
1130.009	12	0.87	1.76	3.20	4.62	10.44
1131.072	13	2.39	4.70	3.60	10.05	20.74
Gamma	14	0.82	3.45	3.89	7.10	15.26
Jefferson	15	3.44	5.37	4.59	10.87	24.26
Yamhill	16	1.93	5.80	6.85	11.89	26.47
LSD (0.05)		0.56	0.97	1.13	2.55	3.83

TABLE 2

Trunk cross-sectional area (TCA), yield efficiency, and relative tree size of 'PollyO' (OSU 1108.001) in comparison with other hazelnut cultivars and selections in the first trial, planted in 2010. Randomized complete block design with four blocks and a single tree of each selection in each block.

Selection	CvNo	TCA (cm <sup>2</sup> )	Yield Efficiency (kg/cm <sup>2</sup> )	Relative tree size (%)	
				To Barcelona	To Jefferson
EFB-resistant selections					
1069.092	1	143.76	0.107	110	154
1072.105	2	131.08	0.130	101	141
1081.101	3	85.81	0.194	66	92
1084.075	4	82.73	0.166	64	89
1085.066	5	87.55	0.248	67	94
1086.145	6	122.81	0.186	94	132
1102.032	7	100.57	0.169	77	108
1102.069	8	90.98	0.190	70	98
1108.001	9	117.78	0.251	91	126
1119.081	10	64.16	0.327	49	69
1130.006	11	129.45	0.147	100	139
1130.009	12	88.23	0.119	68	95
1131.072	13	110.57	0.188	85	119
Gamma	14	108.68	0.141	84	117
Jefferson	15	93.15	0.261	72	100
Yamhill	16	79.32	0.334	61	85
LSD (0.05)		16.00	0.044		

TABLE 3

Nut yield of 'PollyO' (OSU 1108.001) in comparison with other hazelnut cultivars and selections in the second trial, planted in 2011. Randomized complete block design with seven blocks and a single tree of each selection in each block.

Selection	n	Nut yield (kg/tree)				
		2013	2014	2015	2016	2017
OSU 992.022	7	0.22	1.89	3.18	7.43	5.32
OSU 1014.058	7	0.56	2.35	3.21	5.49	5.60
OSU 1018.001	7	0.23	1.10	2.65	5.82	5.48
OSU 1086.053	7	0.38	1.59	2.59	6.57	4.39
OSU 1086.145	7	0.17	2.37	3.77	8.22	6.39
OSU 1102.069	7	0.33	0.99	2.98	4.35	3.70
OSU 1108.001	7	0.18	2.24	4.48	9.99	6.57

TABLE 3-continued

Nut yield of 'PollyO' (OSU 1108.001) in comparison with other hazelnut cultivars and selections in the second trial, planted in 2011. Randomized complete block design with seven blocks and a single tree of each selection in each block.

Selection	n	Nut yield (kg/tree)				
		2013	2014	2015	2016	2017
OSU 1108.042	7	0.19	1.57	4.12	8.73	4.73
OSU 1130.006	6	0.18	0.49	1.68	4.66	4.70
OSU 1130.009	7	0.18	0.79	1.28	2.87	3.04
OSU 1131.072	7	0.34	2.24	3.00	6.36	5.87
OSU 1136.051	7	0.67	2.42	2.78	5.28	4.02
OSU 1152.063	7	0.17	1.97	3.71	5.38	5.37
OSU 1153.052	6	0.21	0.88	2.94	7.03	4.96
OSU 1154.027	7	0.13	2.21	4.62	6.58	6.95
Jefferson	7	0.40	2.59	3.01	5.94	4.98
Yamhill	7	0.43	2.44	4.19	8.14	6.50
LSD(.05)		0.14	0.43	0.66	1.10	0.97

TABLE 4

TCA and yield efficiency of 'PollyO' (OSU 1108.001) in comparison with other hazelnut cultivars and selections in the second trial, planted in 2011. Randomized complete block design with seven blocks and a single tree of each selection in each block.

Selection	n	TCA (cm <sup>2</sup> ) <sup>z</sup>	Total field-run yield (kg/tree) <sup>y</sup>		Yield efficiency (total kg/cm <sup>2</sup> )	
			Nut	Kernel	Nut	Kernel
EFB-resistant selections						
OSU 992.022	7	105.4	18.04	8.32	.171	.079
OSU 1014.058	7	93.9	17.21	7.90	.186	.085
OSU 1018.001	7	80.6	15.27	7.11	.190	.088
OSU 1086.053	7	105.4	15.52	6.76	.148	.066
OSU 1086.145	7	130.6	20.91	9.72	.160	.075
OSU 1102.069	7	78.5	12.34	5.85	.156	.074
OSU 1108.001	7	117.6	23.46	11.26	.201	.097
OSU 1108.042	7	108.6	19.34	8.51	.179	.079
OSU 1130.006	6	107.5	11.71	5.36	.110	.050
OSU 1130.009	7	73.9	8.15	3.74	.110	.050
OSU 1131.072	7	121.5	17.80	8.31	.147	.069
OSU 1136.051	7	70.1	15.18	6.60	.217	.095
OSU 1152.063	7	103.4	16.60	7.60	.162	.077
OSU 1153.052	6	106.3	16.02	7.46	.151	.070
OSU 1154.027	7	111.3	20.49	9.47	.184	.084
Jefferson	7	80.3	16.91	7.53	.210	.094
Yamhill	7	84.4	21.69	9.89	.258	.118
LSD (.05)		11.0	2.10	0.98	.021	.009

<sup>z</sup>TCA = Trunk cross-sectional area calculated from trunk circumference measured 30 cm above the soil line in November 2017;

<sup>y</sup>Cumulative kernel yield and yield efficiency are based on field-run samples, with defects not removed.

Nut and Kernel Defects.

Samples of 100 nuts were cracked from each tree in each year. No problems were noted in either trial, although many differences among selections in the trials lack statistical significance because of the small number of replications. In both trials (Tables 3-5), 'PollyO' (OSU 1108.001) showed a high percentage of good nuts, and low percentages of blanks, brown stain, poorly filled nuts, twins, and kernels with black tips. In the first trial, the frequency of moldy kernels (2.56%) was similar to 'Yamhill' and lower than 'Jefferson'. In the second trial, the frequency of moldy kernels (3.4%) was similar to 'Jefferson' (4.3%). Percent kernel (the ratio of kernel weight to nut weight) for 'PollyO' (OSU 1108.001) was 47.9% in the first trial based on well-filled nuts, and 46.9% in the second trial based on

field-run nuts. In the same trials, the percent kernel for ‘Jefferson’ was 45.7% and 44.4%, respectively. Yields of kernels per acre would be high for ‘PollyO’ (OSU 1108.001). In the two trials, nut yields of ‘PollyO’ (OSU 1108.001) were consistently good and the nuts consistently well-filled. In contrast, ‘Yamhill’ trees occasionally set very heavy nut crops, and the nuts can be so poorly filled that the kernels are not marketable. This was the case for ‘Yamhill’ in the final year of the first trial, when 27% of the nuts were poorly filled.

TABLE 5

Nut weight, kernel weight, and percent kernel of ‘PollyO’ (OSU 1108.001) in comparison with other hazelnut cultivars and selections in the first trial, planted in 2010. Randomized complete block design with four blocks and a single tree of each selection in each block.

Selection	CvNo	Nut Weight (g)	Kernel Weight (g)	Percent Kernel
1069.092	1	3.20	1.47	45.93
1072.105	2	2.73	1.22	44.65
1081.101	3	3.01	1.34	44.58
1084.075	4	2.53	1.22	48.42
1085.066	5	3.40	1.45	42.70
1086.145	6	2.60	1.26	48.60
1102.032	7	3.08	1.46	47.48
1102.069	8	2.94	1.46	49.64
1108.001	9	2.88	1.38	47.90
1119.081	10	3.09	1.37	44.36
1130.006	11	2.56	1.19	46.57
1130.009	12	2.57	1.24	48.00
1131.072	13	2.99	1.43	47.73
Gamma	14	2.48	1.29	51.90
Jefferson	15	3.65	1.67	45.70
Yamhill	16	2.32	1.10	47.11
LSD (0.05)		0.16	0.07	0.94

TABLE 6

Ratings for fiber, blanching (pellicle removal), and bud mite susceptibility of ‘PollyO’(OSU 1108.001) in comparison with other hazelnut cultivars and selection sin the first trial, planted in 2010. Randomized complete block design with four blocks and a single tree of each selection in each block.

Selection	CvNo	Fiber Rating (1-4)	Blanching Rating (1-7)	Bud Mite Rating (1-5)
1069.092	1	2.66	5.59	1.13
1072.105	2	1.38	4.56	2.42
1081.101	3	2.69	4.34	2.49
1084.075	4	2.47	4.28	1.02
1085.066	5	1.44	2.97	1.94
1086.145	6	1.61	4.03	2.35
1102.032	7	2.39	4.96	1.91
1102.069	8	1.25	4.31	1.04
1108.001	9	3.28	3.66	1.15
1119.081	10	2.72	3.28	1.97
1130.006	11	2.97	4.00	2.88
1130.009	12	2.88	3.69	2.25
1131.072	13	3.06	3.94	1.56
Gamma	14	3.09	6.28	2.96
Jefferson	15	3.00	4.34	1.34
Yamhill	16	1.25	5.00	1.10
LSD (0.05)		0.26	0.29	0.26

Fiber amount on the pellicle rated from 1 (no fiber) to 4 (heavy fiber). Blanching after roasting at 275 F. for 15 min rated from 1 (complete pellicle removal) to 7 (no pellicle removal). Bud mite: incidence of blasted bud rated in 5 years in December on a scale of 1.

TABLE 7

Frequency of good nuts, and of nut and kernel defects in ‘PollyO’ (OSU 1108.001) and other hazelnut cultivars and selections in the first trial, planted in 2010.

Selection	CvNo	Frequency (%)			
		Good	Blank	Brown Stain	Moldy
1069.092	1	81.62	3.26	0.56	4.12
1072.105	2	83.62	2.32	2.06	4.32
1081.101	3	80.18	7.18	0.18	6.44
1084.075	4	80.88	3.06	0.18	6.18
1085.066	5	78.50	3.38	0.32	4.94
1086.145	6	73.86	9.00	0.30	9.02
1102.032	7	83.02	4.14	0.36	4.08
1102.069	8	74.44	7.50	1.00	6.26
1108.001	9	83.38	4.00	0.06	2.56
1119.081	10	75.32	4.26	0.18	6.12
1130.006	11	84.32	5.06	0.44	4.38
1130.009	12	80.32	8.32	0.26	5.12
1131.072	13	78.68	7.38	1.26	3.44
Gamma	14	75.38	7.62	1.26	5.38
Jefferson	15	73.68	4.00	0.18	6.12
Yamhill	16	74.18	2.18	0.26	2.68
LSD (0.05)		6.80	2.75	1.18	2.38

TABLE 8

Frequency of nut and kernel defects in ‘PollyO’ (OSU 1108.001) and other hazelnut cultivars and selections in the first trial, planted in 2010.

Selection	CvNo	Frequency (%)			
		Shrivels	Poor Fill	Twins	Black Tips
1069.092	1	2.18	6.44	1.76	0.38
1072.105	2	0.56	7.00	0.12	0.38
1081.101	3	0.82	3.18	1.18	1.06
1084.075	4	0.68	8.82	1.18	0.26
1085.066	5	1.06	11.32	0.38	0.88
1086.145	6	1.30	4.94	0.94	1.62
1102.032	7	0.88	6.00	2.00	0.16
1102.069	8	1.18	6.80	4.00	0.18
1108.001	9	1.50	8.62	0.12	0.06
1119.081	10	3.06	11.32	0.76	0.12
1130.006	11	0.68	4.06	0.12	1.12
1130.009	12	0.38	5.68	0.26	0.26
1131.072	13	1.82	7.12	1.18	0.26
Gamma	14	0.88	8.32	0.12	0.26
Jefferson	15	0.56	13.26	0.88	2.12
Yamhill	16	3.88	16.76	0.06	0.12
LSD (0.05)		1.83	5.12	0.88	0.66

TABLE 9

Nut weight, kernel weight, and percent kernel in ‘PollyO’ (OSU 1108.001) and other hazelnut cultivars and selections in the second trial, planted in 2011.

Selection <sup>2</sup>	n	Frequency (%)		
		Nut weight (g)	Kernel weight (g)	Percent Kernel
OSU 992.022	7	2.57	1.18	46.0
OSU 1014.058	7	2.31	1.05	45.6
OSU 1018.001	7	2.35	1.07	45.4
OSU 1086.053	7	2.58	1.11	43.2
OSU 1086.145	7	2.38	1.12	46.7
OSU 1102.069	7	2.80	1.32	47.0
OSU 1108.001	7	2.75	1.29	46.9
OSU 1108.042	7	2.57	1.12	43.5
OSU 1130.006	6	2.39	1.08	45.0

TABLE 9-continued

Nut weight, kernel weight, and percent kernel in 'PollyO' (OSU 1108.001) and other hazelnut cultivars and selections in the second trial, planted in 2011.				
Selection <sup>z</sup>	n	Frequency (%)		
		Nut weight (g)	Kernel weight (g)	Percent Kernel
OSU 1130.009	7	2.40	1.08	45.2
OSU 1131.072	7	2.86	1.31	45.7
OSU 1136.051	7	2.69	1.16	43.2
OSU 1152.063	7	2.38	1.10	46.1
OSU 1153.052	6	2.68	1.22	45.8
OSU 1154.027	7	2.34	1.06	45.4
Jefferson	7	3.45	1.53	44.4
Yamhill	7	2.28	1.04	45.8
LSD(.05)		0.15	0.03	0.8

<sup>z</sup>Data from seven replications of each selection (2013-17) unless otherwise noted, with a single tree of each selection in each block.

TABLE 10

Frequency of good nuts and of nut and kernel defects in 'PollyO' (OSU 1108.001) and other hazelnut cultivars and selections in the second trial, planted in 2011.				
Selection <sup>z</sup>	n	Blanch	Good Nuts	Blanks
				(Frequency %)
OSU 992.022	7	4.1	89.3	4.8
OSU 1014.058	7	4.7	96.0	1.2
OSU 1018.001	7	3.9	90.6	4.5
OSU 1086.053	7	3.7	89.1	6.1
OSU 1086.145	7	4.4	87.8	6.0
OSU 1102.069	7	4.1	85.9	4.1
OSU 1108.001	7	3.5	93.0	2.5
OSU 1108.042	7	4.6	91.8	3.3
OSU 1130.006	6	4.5	91.6	3.5
OSU 1130.009	7	3.9	89.8	5.1
OSU 1131.072	7	4.2	90.5	3.8
OSU 1136.051	7	3.4	86.5	3.6
OSU 1152.063	7	4.2	92.7	4.7
OSU 1153.052	6	3.9	89.9	2.9
OSU 1154.027	7	4.9	95.0	2.1
Jefferson	7	4.6	89.0	3.9
Yamhill	7	5.1	95.6	1.6
LSD (.05)		0.3	2.2	1.4

<sup>z</sup>Data from seven replications of each selection (2013-17) unless otherwise noted, with a single tree of each selection in each block.

TABLE 11

Frequency of nut and kernel defects in 'PollyO' (OSU 1108.001) and other hazelnut cultivars and selections in the second trial, planted in 2011.							
Selection <sup>z</sup>	n	Frequency (%)					
		Brown Stain	Mold	Shrivel	Poor Fill	Twin	Black Tip
OSU 992.022	7	0.1	4.4	0.7	0.4	0.2	0.2
OSU 1014.058	7	0.0	1.9	0.4	0.1	0.2	0.2
OSU 1018.001	7	0.1	3.3	0.1	0.2	0.9	0.3
OSU 1086.053	7	0.0	4.3	0.2	0.2	0.2	0.1
OSU 1086.145	7	0.0	3.6	0.8	0.6	0.6	0.7
OSU 1102.069	7	0.3	4.1	0.7	0.4	3.6	0.8
OSU 1108.001	7	0.1	3.4	0.3	0.3	0.1	0.4
OSU 1108.042	7	0.1	2.9	0.5	0.9	0.3	0.2
OSU 1130.006	6	0.1	2.6	0.2	0.3	0.1	1.7
OSU 1130.009	7	0.2	4.0	0.4	0.2	0.0	0.2
OSU 1131.072	7	0.2	1.9	0.8	1.0	1.7	0.1
OSU 1136.051	7	0.3	4.5	2.6	0.4	1.4	0.4
OSU 1152.063	7	0.7	0.7	0.1	0.6	0.1	0.4

TABLE 11-continued

Frequency of nut and kernel defects in 'PollyO' (OSU 1108.001) and other hazelnut cultivars and selections in the second trial, planted in 2011.							
Selection <sup>z</sup>	n	Frequency (%)					
		Brown Stain	Mold	Shrivel	Poor Fill	Twin	Black Tip
OSU 1153.052	6	0.4	5.2	0.5	0.3	0.2	0.8
OSU 1154.027	7	0.1	1.5	0.5	0.1	0.5	0.2
Jefferson	7	0.1	4.3	0.4	0.0	0.6	1.7
Yamhill	7	0.1	1.7	0.7	0.1	0.1	0.1
LSD (.05)		0.3	1.1	0.5	0.3	0.4	0.5

<sup>z</sup>Data from seven replications of each selection (2013-17) unless otherwise noted, with a single tree of each selection in each block.

Nut and Kernel Characteristics.

'PollyO' can be used for the blanched kernel market as a companion for 'Wepster' and 'McDonald', and an alternative to 'Yamhill'. The nut shape is nearly round and lends itself well to sizing and cracking (FIG. 3). The shells are thin and easy to crack, and most kernels remain whole when the shell is broken. The kernel size of 'PollyO' is small, similar to 'McDonald' but larger than 'Wepster' and 'Yamhill'. The amount of fiber on the pellicle was rated on a scale of 1 (no fiber) to 4 (heavy fiber). Raw kernels have fibrous pellicles (mean rating 3.3) and thus are not particularly attractive. Pellicle removal (blanching) is rated on a scale of 1 (complete pellicle removal) to 7 (no pellicle removal), with ratings <4.0 being desired. Most of the pellicle on 'PollyO' kernels is removed from the kernels with dry heat in the blanching process (ratings=3.7 and 3.5 in the first and second trials, respectively) which is better than 'Yamhill' (ratings=5.0 and 5.1) and 'Jefferson' (ratings 4.3 and 4.6). Blanching removes about half of the pellicle from 'Barcelona' kernels (FIG. 3). Kernel texture, flavor and aroma were evaluated in September 2016 or 2017. Tasters consider kernel quality to be excellent, and well-suited for use in baked goods and chocolate products.

Nut Maturity Date.

The nuts of 'PollyO' are borne in clusters of 3-4 in husks about 25% longer than the nuts (FIG. 4). The husks are slit down the side, and flare open as they dry at maturity (FIG. 6). About 92% of the nuts fall free of the husk at maturity (range 85-97%). The other 8% of the nuts would come out of the husks as they moved through the harvester. When mature, the shells are light tan in color. Nuts in the first trial were hand-harvested and notes recorded at the time of harvest. Notes included the date, the percentage of nuts on the ground (as opposed to in the tree), and the percentage of the nuts on the ground that were free of the husk (as opposed to inside the husk). More than 2000 trees are harvested by hand every fall, visiting a block once every 7 to 10 days, and harvesting a tree if >70% of the nuts are on the ground. Thus the percentages in the first trial are estimates. The harvest dates and percentage down indicate that maturity of 'PollyO' is 10-14 days earlier than 'Barcelona', allowing harvest before the start of the rainy season. To more precisely estimate nut maturity, four trees each of 'PollyO', 'Jefferson' and 'Yamhill' in the first trial and four trees of 'Barcelona' in adjacent rows were harvested by hand at weekly intervals in the final year of the trial (2016). The maturity curves from the 2016 harvest (FIG. 7) show that nuts of 'PollyO' drop a few days earlier than 'Yamhill' and 10-14 days sooner than 'Barcelona'. Average harvest dates compared to 'Barcelona' are about 14 days early for 'McDon-

ald', 10 days early for 'Yamhill' and 7 days early for 'Wepster'. 'Jefferson' nuts mature with 'Barcelona' or up to three days later. Nuts in the second trial were annually raked and fed through a Mave harvester. Prior to harvest, the percentage of nuts on the ground was estimated weekly in the final year of the trial, beginning on September 15 (FIG. 8). The nut drop estimate curves indicate that nut maturity of 'PollyO' is between 'McDonald' and 'Yamhill'. Plantings of 'PollyO' with 'McDonald' and 'Wepster' with additional 'York' pollinizer trees would allow harvest to begin two weeks earlier than 'Barcelona'. Nuts were harvested from four cultivars in the second trial on five dates (FIG. 9), and confirmed the relative maturities. Early harvest before the rainy season begins results in lower drying costs and higher nut quality.

Incompatibility and Pollinizers.

Trees of 'PollyO' set a moderate to high amount of catkins that shed copious amounts of pollen in early mid-season, with 'McDonald', 'Wepster', 'York' and 'Yamhill'. Pollen has been collected and used in several controlled pollinations, and both quantity and viability appear to be very good. Pollen of 'PollyO' has germinated well on agar plates (85% in 2014, 81% in 2015 and 48% in 2017), in contrast to 'Barcelona' (36%, 36% and 40%, respectively). 'PollyO' has incompatibility alleles S<sub>2</sub> and S<sub>10</sub> as determined by fluorescence microscopy. Both alleles are expressed in the females, but only S<sub>10</sub> is expressed in the pollen because of dominance. By convention, alleles expressed in the pollen are underlined.

Time of pollen shed and female receptivity were recorded weekly from mid-December to mid-March and are shown for four years (2014-2018) (FIGS. 10-13). Female inflorescences of 'PollyO' emerged in early mid-season and were fully receptive in mid-January, overlapping the period of receptivity of 'McDonald', 'Wepster', 'Yamhill' and 'York'. Time of pollen shed peaks in the first half of February, about ten days later than the other four cultivars. There would be good overlap between female receptivity and pollen shed for any combination of these five cultivars, which are also cross-compatible in all combinations and directions. Many orchard designs are possible using 'PollyO' (S<sub>2</sub> S<sub>10</sub>), 'McDonald' (S<sub>2</sub> S<sub>15</sub>) and 'Wepster' (S<sub>1</sub> S<sub>2</sub>). Inclusion of a few trees of the pollinizer 'York' (S<sub>2</sub> S<sub>21</sub>) is also recommended. 'Yamhill' (S<sub>8</sub> S<sub>26</sub>) is also a suitable pollinizer for 'PollyO' but its tree is lower in vigor and its kernels do not blanch well. Pollen of 'Jefferson' (S<sub>1</sub> S<sub>3</sub>) is also compatible on females of 'PollyO', but 'Jefferson' females emerge very late in the season, and 'Jefferson' has large nuts for the in-shell market whereas all of the others are for the kernel market. Pollen of 'Dorris' (S<sub>1</sub> S<sub>12</sub>), 'Sacajawea' (S<sub>1</sub> S<sub>22</sub>) and 'Barcelona' (S<sub>1</sub> S<sub>2</sub>) is also compatible on females of 'PollyO'; of these only 'Dorris' is highly resistant to EFB. Pollen of 'Felix' (S<sub>15</sub> S<sub>21</sub>) and 'Theta' (S<sub>5</sub> S<sub>15</sub>) is also compatible but they shed pollen too late to be effective pollinizers for 'PollyO'. Pollen of 'Tonda di Giffoni' (S<sub>2</sub> S<sub>23</sub>) (unpatented) is incompatible because it expresses S<sub>2</sub>. Inclusion of 'McDonald' and 'Wepster' as companion cultivars in the orchard results in no loss in total nut yield. Alternative orchard designs include plantings of different cross-compatible cultivars in adjacent rows. The inclusion of pollinizers with a high level of EFB resistance would eliminate the need for fungicide control in the entire orchard.

Pests and Diseases.

Based on DNA marker data, 'PollyO', like 'McDonald', 'Wepster' and 'Yamhill', has a very high level of resistance

to EFB conferred by a dominant allele from 'Gasaway' through the grower selection 'Zimmerman', so fungicide applications are not needed. RAPD markers 152-800 and 268-580 that flank the resistance allele in 'Gasaway' are present in 'PollyO', and these markers are transmitted to its seedlings. Additional RAPD markers linked to resistance are also present. No cankers have been observed on the 11 trees of 'PollyO' in the yield trials, while several cankers have been noted on adjacent trees of susceptible genotypes. During the trials, a nearby heavily infected commercial orchard provided high EFB disease pressure on all plantings. 'PollyO' and many other selections were propagated by tie-off layerage of the suckers in 2013. Eight trees of 'PollyO' were potted and grown in the lathhouse in the summer of 2014 and exposed to EFB under a structure topped with diseased branches in spring 2015. Unfortunately, inspection in December 2016 showed that very few trees became infected, even the highly susceptible check 'Ennis' (unpatented). It seems that the diseased branches were of low-quality inoculum due to freeze injury, as no useful data were provided by this exposure.

Susceptibility to bacterial blight caused by *Xanthomonas campestris* pv. *corylina* has not been quantified, but no trees in the two trials were affected.

Susceptibility to big bud mite (primarily *Phytoptus avelanae* Nal.) was rated in the first trial (Tables 5-6) after leaf fall once per year for five years (December 2012-2016). The scale was from 1 (no blasted buds) to 5 (many blasted buds), with chemical control considered unnecessary for cultivars with a mean rating <3.3. The average ratings indicate high resistance for 'PollyO' (1.15) and 'Jefferson' (1.34), and an intermediate rating for 'Gamma' (2.96). Blasted buds are very rare on 'PollyO', so chemical applications should not be necessary to control bud mite.

Microsatellite Marker Analysis:

Twenty microsatellite (simple sequence repeat) markers were used. PCR products were multiplexed post-PCR and sized using capillary electrophoresis (Tables 12-13, and see for example Bassil et al., *Acta Horticulturae* 686:105-110, 2005; Gökirmak et al., *Genetic Resources and Crop Evolution* 56:147-172, 2009; Gürcan and Mehlenbacher. *Molecular Breeding* 26:551-559, 2010; Gürcan et al. *Tree Genetics and Genomes* 6:513-531, 2010).

TABLE 12

Primers for the microsatellite marker loci used to fingerprint 'PollyO' and other hazelnut cultivars.					
Set	Lo- cus	Repeat Motif	Allele sizes	Primers (5'-3') (SEQ ID NO:)	
3	A613	(TC) <sub>13</sub> (CA) <sub>12</sub>	149- 177	Ned- CACACGCCT TGTCACCTC TT(1)	R- CCCCTTT CACATGT TTGCTT (2)
2	A614	(TC) <sub>17</sub> (CA) <sub>10</sub> NNN(CA) <sub>6</sub>	125- 156	Hex- TGGCAGAGC TTTGTACGC TT(3)	R- GCAGTG GAGGAT TGCTGAC T(4)

TABLE 12-continued

Primers for the microsatellite marker loci used to fingerprint 'PollyO' and other hazelnut cultivars.				
Lo-Set	Repeat Motif	Allele sizes	Primers (5'-3') (SEQ ID NO:)	
3	A616 (AC) <sub>11</sub>	136-162	Fam-CACTCATAC CGCAAACCTC CA(5)	R-ATGGCTT TTGCTTC GTTTTG (6)
1	A640 (CT) <sub>15</sub> (CA) <sub>13</sub>	354-378	F-TGCCTCTGC AGTTAGTCA TCAAATGTA GG(7)	Fam-CGCCAT ATAATTG GGATGC TTGTTG (8)
3	B617 (GA) <sub>15</sub>	280-298	Fam-TCCGTGTTG AGTATGGAC GA(9)	R-TGTTTTT GGTGGA GCGATG (10)
2	B619 (TC) <sub>21</sub>	146-180	Fam-AGTCGGCTC CCCTTTTCT C(11)	R-GCGATCT GACCTC ATTTTTG (12)
4	B634 (AG) <sub>15</sub>	218-238	Hex-CCTGCATCC AGGACTCAT TA(13)	R-GTGCAG AGGTTG CACTCA AA(14)
4	B657 (AG) <sub>15</sub>	210-228	Ned-GAGAGTGCG TCTTCCTCT GG(15)	R-AGCCTC ACCTCCA ACGAAC (16)
3	B671 (AG) <sub>6</sub> NN (GA) <sub>17</sub>	221-249	Hex-TTGCCAGTG CATACTCTG ATG(17)	R-ACCAGC TCTGGGC TTAACAC (18)
2	B709 (GA) <sub>21</sub>	219-233	Ned-CCAAGCACG AATGAACCTC AA(19)	R-GCGGGT TCTCGTT GTACACT (20)
1	B733 (TC) <sub>15</sub>	161-183	Ned-CACCTCTT CACCACCTC AT(21)	R-CATCCCC TGTTGGA GTTTTT (22)
2	B749 (TC) <sub>12</sub>	200-210	Hex-GGCTGACAA CACAGCAGA AA(23)	R-TCGGCTA GGGTTA GGGTTTT (24)
4	B751 (GA) <sub>15</sub>	141-153	Fam-AGCTGGTTC TTCGACATT CC(25)	R-AAACTC AAATAA AACCCCT GCTC(26)

TABLE 12-continued

Primers for the microsatellite marker loci used to fingerprint 'PollyO' and other hazelnut cultivars.				
Lo-Set	Repeat Motif	Allele sizes	Primers (5'-3') (SEQ ID NO:)	
1	B774 (AG) <sub>15</sub>	195-213	Ned-GTTTTGCGA GCTCATTGT CA(27)	R-TGTGTGT GGTCTGT AGGCAC T(28)
3	C115 (TAA) <sub>5</sub> (GAA) <sub>12</sub>	167-225	Fam-CATTTTCCG CAGATAATA CAGG(29)	R-GTTTCCA GATCTGC CTCCATA TAAT(30)
3	KG807 (TAAA) <sub>2</sub> AA (TAAA) <sub>2</sub> A (TAAA) <sub>2</sub>	226-248	F-AAGCAAGAA AGGGATGGT (31)	Fam-CTTACAG ATAAAT GGCTCA AA(32)
1	KG809 (AGG) <sub>6</sub>	333-345	F-GGAAGGTGA GAGAAATCA AGT(33)	Hex-AGGCAT CAGTTCA TCCAA (34)
2	KG811 (GA) <sub>17</sub>	240-278	F-GAACAACTG AAGACAGCA AAG(35)	Ned-AAGGCG GCACTC GCTCAC (36)
4	KG827 (CT) <sub>13</sub> AA (CA) <sub>7</sub>	264-282	Fam-AGAACTCCG ACTAATAAT CCTAACCTT TGC(37)	R-GAGGGA GCAAGT CAAAGT TGAGAA GAAA (38)
2	KG830 (CT) <sub>14</sub> GTATT (CA) <sub>8</sub>	279-311	Ned-TGGAGGAAG TTTTGAATG GTAGTAGAG GA(39)	R-AAAGCA ACTCATA GCTGAA GTCCAAT C(40)A

45 Set = Multiplex set, with samples mixed after PCR but before submission for sizing by capillary electrophoresis; Allele size = range of sizes; Primers, forward (F, listed first) and reverse (R, second) (Hex and Fam are dyes).

TABLE 13

Annealing temperatures for the microsatellite marker loci used to fingerprint 'PollyO' and other hazelnut cultivars.										
Set	(° C.)	n	He	Ho	PIC	r	LG	Locus	Reference	T <sub>m</sub>
										3
2	60	14	0.85	0.85	0.84	0.00	65,6R	A614	Gurcan et al., 2010	
3	60	13	0.85	0.85	0.83	0.00	8R	A616	Gurcan et al., 2010	
1	67	11	0.80	0.73	0.77	0.04	10R	A640	Gurcan et al., 2010	
3	60	9	0.80	0.78	0.78	0.01	85,8R	B617	Gurcan et al., 2010	
2	60	14	0.88	0.88	0.87	0.00	35,3R	B619	Gurcan et al., 2010	

TABLE 13-continued

Annealing temperatures for the microsatellite marker loci used to fingerprint 'PollyO' and other hazelnut cultivars.									
Set	Tm (° C.)	n	He	Ho	PIC	r	LG	Locus	Reference
4	60	9	0.76	0.76	0.73	0.00	4R	B634	Gurcan et al., 2010
4	60	8	0.84	0.98	0.82	-0.08	11S,11	B657	Gurcan et al., 2010
3	60	13	0.86	0.88	0.84	-0.01	95,9R	B671	Gurcan et al., 2010
2	60	8	0.74	0.76	0.70	-0.01	55,5R	B709	Gurcan et al., 2010
1	60	8	0.68	0.68	0.63	0.00	75,2R	B733	Gurcan et al., 2010
2	60	6	0.60	0.64	0.51	-0.03	1R	B749	Gurcan et al., 2010
4	60	7	0.80	0.78	0.77	0.01	75,2R	B751	Gurcan et al., 2010
1	60	8	0.80	0.80	0.77	0.00	55,5R	B774	Gurcan et al., 2010
3	60	10	0.84	0.90	0.82	0.035	45,4R	C115	B assil et al., 2005b; Gokirmak et al., 2009
3	54	4	0.67	0.78	0.60	-0.07	11	KG807	Gurcan and Mehlenbacher, 2010
1	55	5	0.66	0.64	0.60	0.01	4	KG809	Gurcan and Mehlenbacher, 2010
2	58	12	0.83	0.82	0.81	0.01	2	KG811	Gurcan and Mehlenbacher, 2010
4	67	9	0.78	0.84	0.75	-0.04	9	KG827	Gurcan and Mehlenbacher, 2010
2	67	9	0.79	0.78	0.76	0.00	9	KG830	Gurcan and Mehlenbacher, 2010

Tm = Annealing temperature ; n = Number of alleles; He = expected heterozygosity; Ho = observed heterozygosity; PIC = Polymorphism information content; r = frequency of null alleles; LG = linkage group (S is susceptible female parent 'OSU 252.146'; R is resistant parent 'OSU 414.062'); Reference is the journal article where additional details were published.

Table 14 shows allele sizes at 20 microsatellite markers for 'PollyO', its parents 'OSU684.104' and 'OSU 669.104', cultivars and selections that appear in the pedigree of 'PollyO' ('Mortarella' to 'Montebello'), and four kernel cultivars ('Yamhill', 'Tonda Pacifica', 'McDonald' and 'Wepster'). 'PollyO' is easily distinguished from all others shown in Table 13. 'PollyO' is different from its female parent 'OSU 684.104' at 11 of the 20 marker loci. 'PollyO' is different from its male parent 'OSU 669.104' at 14 of the 20 marker loci. 'PollyO' is the same as both parents at 3 loci. 'PollyO' is the same as one parent at 9 loci. 'PollyO' is different from both parents at 8 loci. Allele A614-125, Allele A616-144, Allele B617-285, Alleles B709-223 and B709-227, and Allele KG807-238 observed in 'PollyO' are not found any of 'Yamhill', 'Tonda Pacifica', 'McDonald', and 'Wepster'.

TABLE 14

Allele sizes at 20 microsatellite markers					
Marker	'PollyO'	'OSU 684.104'	'OSU 669.104'	'Mortarella'	
5	A613	153/167	153/167	153/153	151/153
	A614	125/158	125/132	132/158	132/135
	A616	144/152	150/152	144/150	150/158
	A640	354/354	354/374	354/354	354/374
	B617	285/295	285/293	289/295	293/295
10	B619	158/166	158/178	166/176	158/172
	B634	228/236	236/236	222/228	228/236
	B657	218/218	218/218	218/218	218/224
	B671	229/249	225/249	229/237	225/243
	B709	223/227	223/229	227/229	221/229
	B733	175/181	175/175	175/181	175/175
15	B749	216/216	216/216	214/216	214/216
	B751	144/154	144/154	154/154	144/154
	B774	209/209	209/215	209/215	209/220
	C115	194/215	194/215	174/215	182/215
	KG807	238/252	238/252	238/252	230/234
	KG809	342/342	342/342	339/342	342/342
	KG811	261/267	261/267	261/267	261/267
20	KG827	272/284	272/282	272/284	282/282
	KG830	291/295	291/295	295/295	291/307
Marker	'OSU 14.084'	'Negret'	'Barcelona'	'Daviana'	'OSU 342.019'
25	A613	153/167	153/159	153/161	167/179
	A614	125/148	132/132	125/132	148/150
	A616	144/152	152/160	144/152	150/152
	A640	354/354	354/368	354/374	354/354
	B617	289/295	285/291	285/289	289/295
	B619	172/178	166/170	158/172	168/178
30	B634	228/236	228/234	228/228	228/236
	B657	218/226	218/222	218/222	218/226
	B671	229/249	229/237	225/229	239/249
	B709	229/235	223/227	227/235	229/229
	B733	173/181	167/175	173/175	173/181
	B749	216/216	216/216	216/216	216/216
35	B751	144/152	152/154	144/154	144/152
	B774	213/213	209/215	209/213	213/215
	C115	174/194	182/215	174/194	174/194
	KG807	252/252	238/252	238/252	238/252
	KG809	339/342	339/342	339/339	339/342
	KG811	261/267	259/267	261/267	255/261
40	KG827	272/284	272/284	282/284	272/272
	KG830	295/295	295/303	291/295	289/295
Marker	'Zimmerman'	'Gasaway'	'Casina'	'Montebello'	
	A613	153/163	161/163	151/153	151/153
	A614	132/158	143/158	125/139	125/132
45	A616	144/150	150/150	144/152	152/160
	A640	354/368	362/368	354/372	362/374
	B617	289/295	291/295	285/295	285/293
	B619	172/176	172/176	150/158	160/166
	B634	222/228	222/234	228/232	232/236
	B657	218/224	224/228	216/218	218/226
50	B671	229/237	237/249	229/249	225/243
	B709	229/235	229/229	227/233	229/235
	B733	175/175	175/175	175/175	175/185
	B749	216/216	214/216	214/216	216/216
	B751	144/154	144/144	152/154	154/154
	B774	213/215	209/215	209/215	209/213
55	C115	174/215	215/218	174/197	174/197
	KG807	238/252	242/252	238/252	238/242
	KG809	339/339	339/348	339/339	339/348
	KG811	261/267	257/261	251/267	251/267
	KG827	272/284	272/282	272/284	282/282
	KG830	295/305	291/305	295/303	291/295
Marker	'Yamhill'	'Tonda Pacifica'	'McDonald'	'Wepster'	
	A613	153/163	159/169	153/169	159/167
	A614	132/158	135/150	135/158	135/158
	A616	150/150	150/160	150/160	152/160
65	A640	354/368	368/374	362/368	368/374

TABLE 14-continued

Allele sizes at 20 microsatellite markers				
B617	289/295	293/295	293/295	293/295
B619	158/172	166/172	158/172	166/172
B634	236/236	228/228	222/228	228/228
B657	218/228	210/226	210/218	226/226
B671	225/243	229/239	229/237	239/249
B709	229/229	229/235	229/229	229/235
B733	181/185	173/175	173/175	173/175
B749	216/216	214/216	214/216	214/216
B751	152/152	144/154	144/144	144/144

TABLE 14-continued

Allele sizes at 20 microsatellite markers				
B774	209/217	209/213	209/220	209/213
C115	197/215	174/182	174/197	182/194
KG807	230/252	228/252	252/252	252/252
KG809	348/348	339/342	339/339	342/342
KG811	251/261	245/257	245/267	257/257
KG827	282/282	270/284	272/284	270/282
KG830	291/295	291/295	291/295	295/305

## SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 40

<210> SEQ ID NO 1  
 <211> LENGTH: 20  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 1

cacacgcctt gtcactcttt 20

<210> SEQ ID NO 2  
 <211> LENGTH: 20  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 2

cccctttcac atgtttgctt 20

<210> SEQ ID NO 3  
 <211> LENGTH: 20  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 3

tggcagagct ttgtcagctt 20

<210> SEQ ID NO 4  
 <211> LENGTH: 20  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 4

gcagtgagg attgctgact 20

<210> SEQ ID NO 5  
 <211> LENGTH: 20  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 5

cactcatacc gcaaaactcca 20

---

-continued

---

<210> SEQ ID NO 6  
<211> LENGTH: 20  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 6  
  
atggcttttg cttcgttttg 20

<210> SEQ ID NO 7  
<211> LENGTH: 29  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 7  
  
tgccctctgca gttagtcac aaatgtagg 29

<210> SEQ ID NO 8  
<211> LENGTH: 25  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 8  
  
cgccatataa ttgggatgct tgttg 25

<210> SEQ ID NO 9  
<211> LENGTH: 20  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 9  
  
tccgtggttga gtagggacga 20

<210> SEQ ID NO 10  
<211> LENGTH: 19  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 10  
  
tgtttttggt ggagcgatg 19

<210> SEQ ID NO 11  
<211> LENGTH: 19  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 11  
  
agtcggctcc ccttttctc 19

<210> SEQ ID NO 12  
<211> LENGTH: 20  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:

-continued

---

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 12

gcgatctgac ctcatttttg 20

<210> SEQ ID NO 13  
 <211> LENGTH: 20  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 13

cctgcatcca ggactcatta 20

<210> SEQ ID NO 14  
 <211> LENGTH: 20  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 14

gtgcagaggt tgcactcaaa 20

<210> SEQ ID NO 15  
 <211> LENGTH: 20  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 15

gagagtgcgt cttcctctgg 20

<210> SEQ ID NO 16  
 <211> LENGTH: 19  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 16

agcctcacct ccaacgaac 19

<210> SEQ ID NO 17  
 <211> LENGTH: 21  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 17

ttgccagtgc atactctgat g 21

<210> SEQ ID NO 18  
 <211> LENGTH: 20  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 18

accagctctg ggcttaacac 20

---

-continued

---

<210> SEQ ID NO 19  
<211> LENGTH: 20  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 19  
  
ccaagcacga atgaactcaa 20

<210> SEQ ID NO 20  
<211> LENGTH: 20  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 20  
  
gcggttctc gttgtacact 20

<210> SEQ ID NO 21  
<211> LENGTH: 20  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 21  
  
caccctcttc accacctcat 20

<210> SEQ ID NO 22  
<211> LENGTH: 20  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 22  
  
catcccctgt tggagttttc 20

<210> SEQ ID NO 23  
<211> LENGTH: 20  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 23  
  
ggctgacaac acagcagaaa 20

<210> SEQ ID NO 24  
<211> LENGTH: 20  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 24  
  
tcggctaggg ttagggtttt 20

<210> SEQ ID NO 25  
<211> LENGTH: 23  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:

-continued

---

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 25

aaactcaaat aaaaccctg ctc 23

<210> SEQ ID NO 26  
 <211> LENGTH: 23  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 26

aaactcaaat aaaaccctg ctc 23

<210> SEQ ID NO 27  
 <211> LENGTH: 20  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 27

gttttgagag ctccattgtca 20

<210> SEQ ID NO 28  
 <211> LENGTH: 21  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 28

tgtgtgtggt ctgtaggcac t 21

<210> SEQ ID NO 29  
 <211> LENGTH: 22  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 29

cattttccgc agataatata gg 22

<210> SEQ ID NO 30  
 <211> LENGTH: 25  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 30

gtttccagat ctgcctccat ataat 25

<210> SEQ ID NO 31  
 <211> LENGTH: 18  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 31

aagcaagaaa gggatggt 18

-continued

---

<210> SEQ ID NO 32  
<211> LENGTH: 21  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 32  
  
cttacagata aatggctcaa a 21

<210> SEQ ID NO 33  
<211> LENGTH: 21  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 33  
  
ggaaggtgag agaaatcaag t 21

<210> SEQ ID NO 34  
<211> LENGTH: 18  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 34  
  
aggcatcagt tcatccaa 18

<210> SEQ ID NO 35  
<211> LENGTH: 21  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 35  
  
gaacaactga agacagcaaa g 21

<210> SEQ ID NO 36  
<211> LENGTH: 18  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 36  
  
aaggcggcac tcgctcac 18

<210> SEQ ID NO 37  
<211> LENGTH: 30  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 37  
  
agaactccga ctaataatcc taacccttgc 30

<210> SEQ ID NO 38  
<211> LENGTH: 28  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:

-continued

---

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 38

gagggagcaa gtcaaagttg agaagaaa 28

<210> SEQ ID NO 39

<211> LENGTH: 29

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 39

tggaggaagt tttgaatggt agtagagga 29

<210> SEQ ID NO 40

<211> LENGTH: 27

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 40

aaagcaactc atagctgaag tccaatc 27

---

We claim:

1. A new and distinct cultivar of *Corylus* plant as illustrated and described. 30

\* \* \* \* \*

FIG. 1

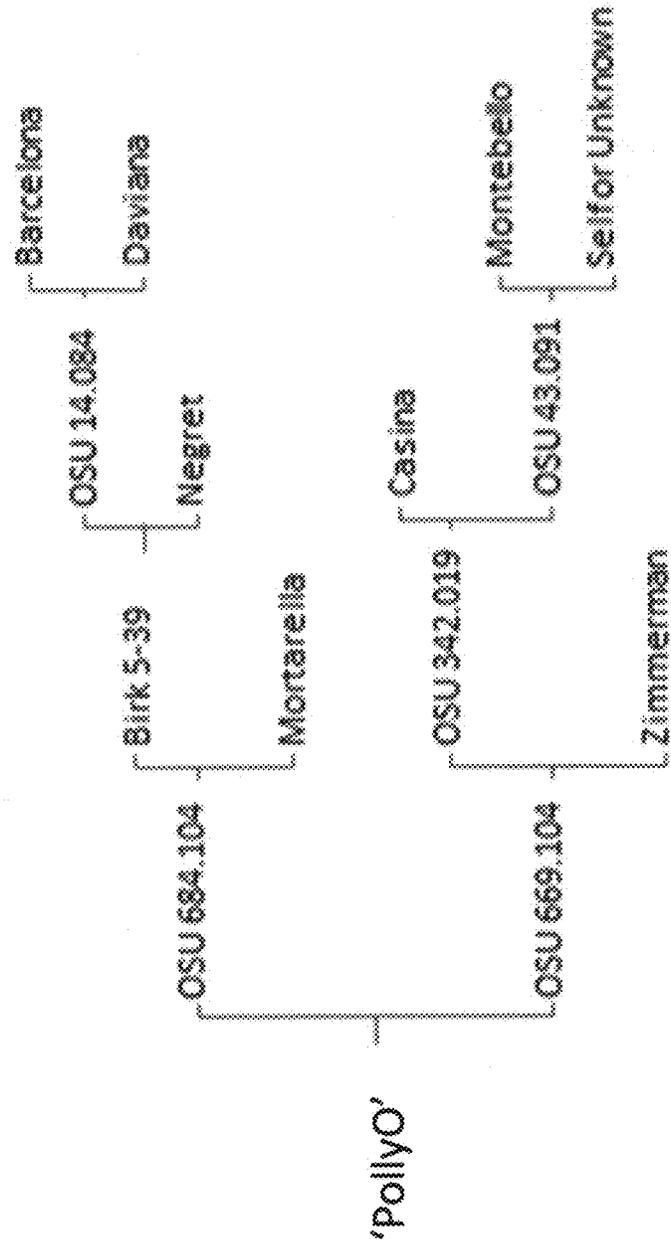




FIG. 2

FIG. 4

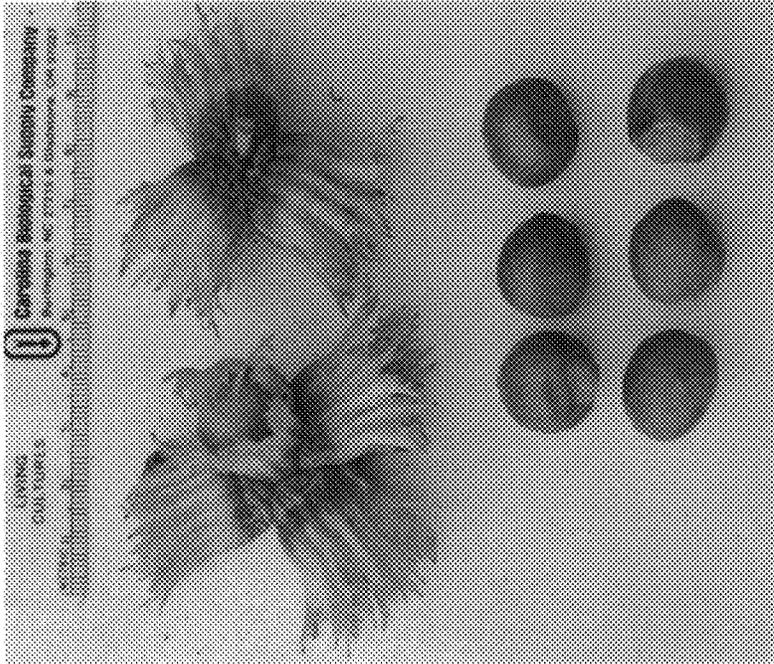


FIG. 3



FIG. 6

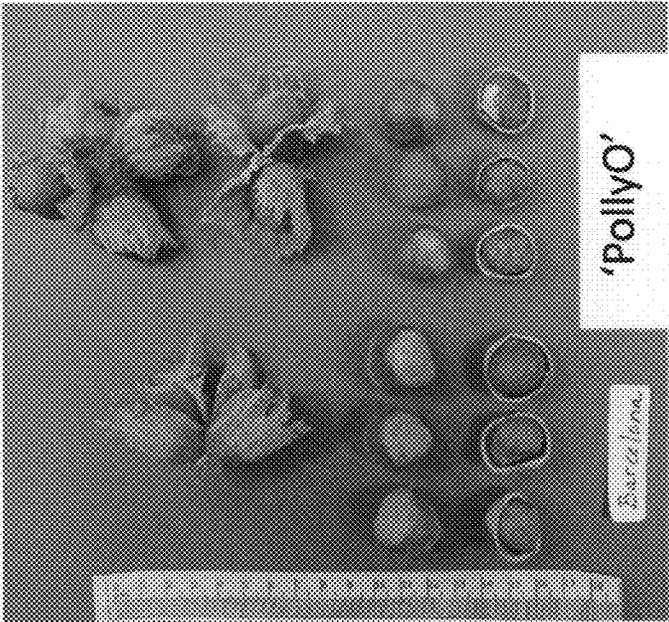


FIG. 5

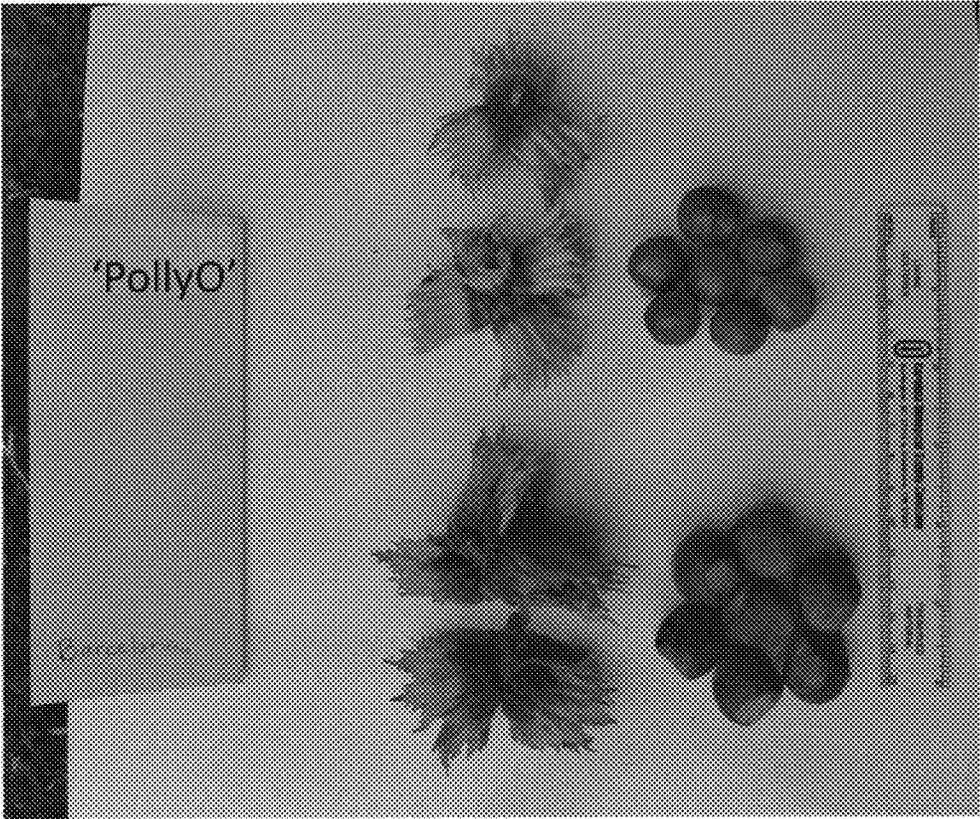


FIG. 7

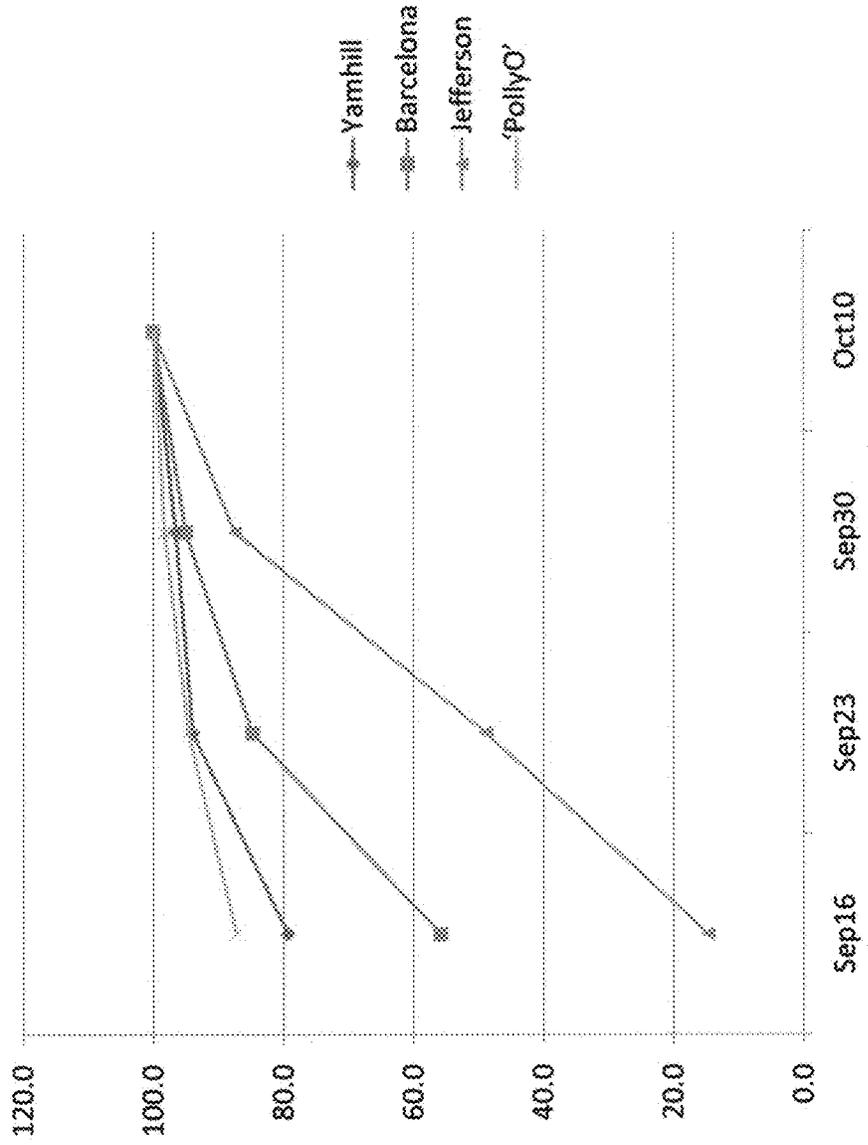


FIG. 8

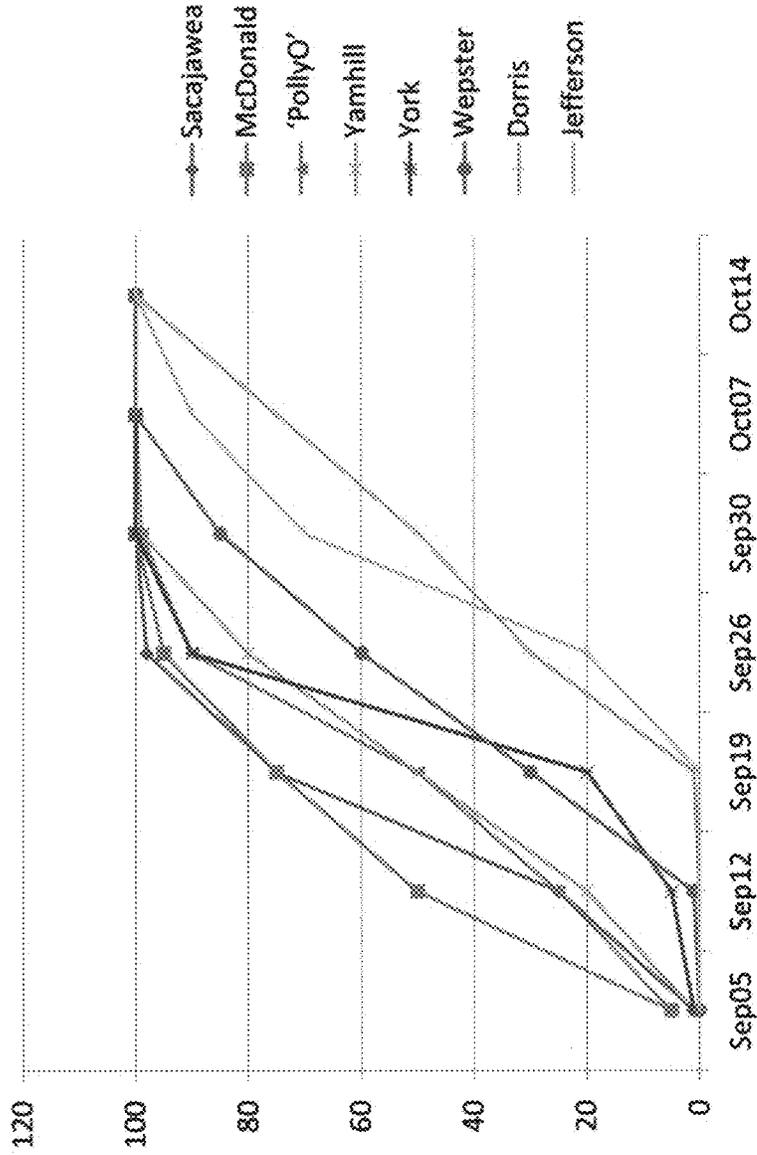


FIG. 9

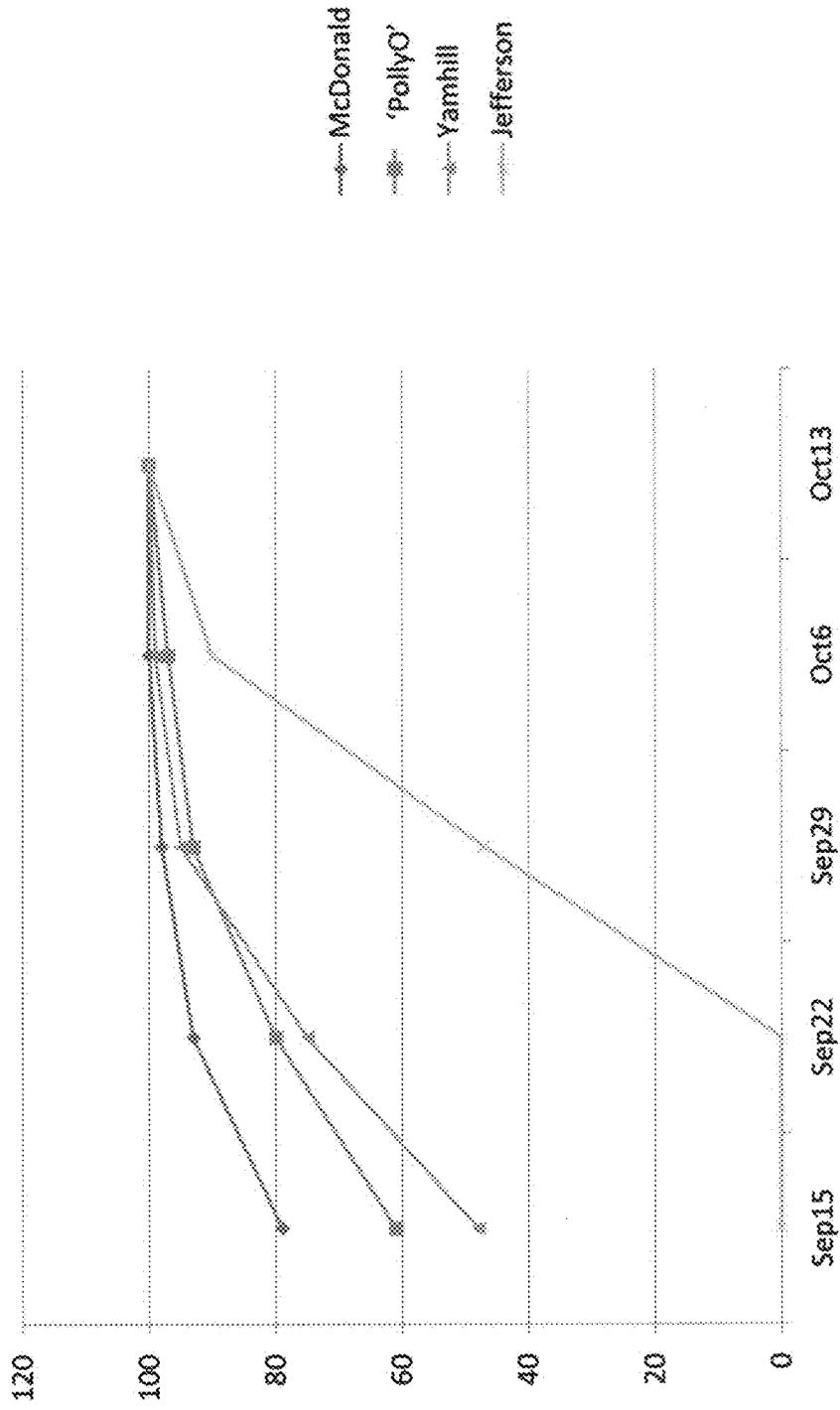


FIG. 10

Phenology Chart 1. Time of female receptivity (bottom red) and pollen shed (top green) from Dec 2014-Mar 2015 of selections in the second trial planted in 2011, including its guard row.

Genotype	Dec			Jan			Feb			March								
	15-Dec	22-Dec	30-Dec	4-Jan	11-Jan	19-Jan	26-Jan	30-Jan	5-Feb	10-Feb	15-Feb	19-Feb	25-Feb	3-Mar	7-Mar	14-Mar	18-Mar	27-Mar
1108.001 (2, 10)	Rd			Rd+2	1+2 Rd+2S+2	1st 5	70 100-20	100-20	5-Feb 10-Feb	20 70	100-10	100-30	gr tp	BB				
Wepster 1, 2	Rd	Rd+3	Rd+2f	1st Rd+2f	5 Rd+3S 1+3S	70 100-20	100-20	100-20	gr tp					BB				
McDonald 2, 15	Rd+2			Rd+2	Rd+2 1+3S 1+3S+2	70 100-20	100-20	100-20	gr tp					BB				
York 2, 21	Rd			Rd+2f	1st 40 70	40 75	95-20	95-20	gr tp					BB				
Yamhill 3, 26	Rd few	Rd+1		1+3 1+3S+2	1+3 1+3S+2	100-70	100-40	100-70	gr tp					BB				
Jefferson 1, 3				Rd	10 50	100	100	100										

Flower stages: RD = Red dot; 1 = styles protruding ~1-3mm, straight; FS = First appearance of 'spiders' or flowers with reflexed styles, and S+3 = majority of flowers are in 'spider' or fully reflexed stage.  
 Green = Pollen shed (a minus sign in front of the number indicates the percent of catkins that have already shed out).  
 Red = Female flowers; red, blk (black), dk (dark, not quite red, not yet black).  
 BB = leaf budbreak

FIG. 11

Phenology Chart 2. Time of female receptivity (bottom red) and pollen shed (top green) from Dec 2015-Mar 2016 of selections in the second trial planted in 2011, including its guard row.

	Dec			Jan				Feb				March						
	15-Dec	22-Dec	30-Dec	4-Jan	6-Jan	15-Jan	20-Jan	25-Jan	30-Jan	5-Feb	10-Feb	15-Feb	20-Feb	25-Feb	3-Mar	7-Mar	11-Mar	12-Mar
Grandtype 1108.001				1st Rd				1st Rd										
(2, 10)																		
Wepster 1, 2	Rd+2	Rd+3						0	30	90-10	-95	done	gr tp					
								Rd+3	Rd+3/1	Rd+3	1-3/5		H-2/S+2	FS+3				
													Dark	Dark/Black				
McDonald 2, 15	Rd	Rd+3	Rd+3/1					50	95-40				gr tp					
								Rd+3/1	1-3	1-3/5			H-3/S+2	H-3/S+2				
													Red/Dk	Dark/Black				
York 2, 21	Rd	Rd+3						0	25	75	90-50							
								Rd+3	Rd+3/1	1-3			H-3/S	H-3/S				
Yamhill 8, 26	Rd	Rd+3	Rd+3/1															
Jefferson 1, 3								5	80	100-70								

Flower stages: RD = Red dot, 1 = styles protruding ~1.3mm, straight; FS = First appearance of 'spiders' or flowers with reflexed styles, and S+3 = majority of flowers are in 'spider' or fully reflexed stage.  
 Green = Pollen shed (a minus sign in front of the number indicates the percent of catkins that have already shed out).  
 Red = Female flowers; red, blk (black), dk (dark, not quite red, not yet black).  
 BB = Leaf budbreak

FIG. 12

Phenology Chart 3. Time of female receptivity (bottom red) and pollen shed (top green) from Dec. 2016-Mar 2017 of selections in the second that planted in 2011, including its guard row.

	Dec		Jan				Feb				March									
	5-Dec	11-Dec	15-Dec	22-Dec	30-Dec	4-Jan	8-Jan	15-Jan	20-Jan	25-Jan	30-Jan	5-Feb	10-Feb	15-Feb	19-Feb	25-Feb	3-Mar	7-Mar	11-Mar	18-Mar
Genotype																				
11081001																				
(2, 3)																				
Wespiter																				
(1, 2)																				
McDonald																				
(2, 3)																				
York																				
(2, 21)																				
Yamhill																				
(8, 26)																				
Jefferson																				
(1, 3)																				

Flower stages: RD = Red dot, l = styles protruding ~1-3mm, straight, FS = First appearance of 'spiders' on flowers with reflexed styles, and S+3 = majority of flowers are in 'spider' or fully reflexed stage.  
 Green = Pollen shed (a minus sign in front of the number indicates the percent of catkins that have already shed out).  
 Red = Female flowers, red, blk (black), dk (dark, not quite red, not yet black).  
 BB = Leaf budbreak

FIG. 13

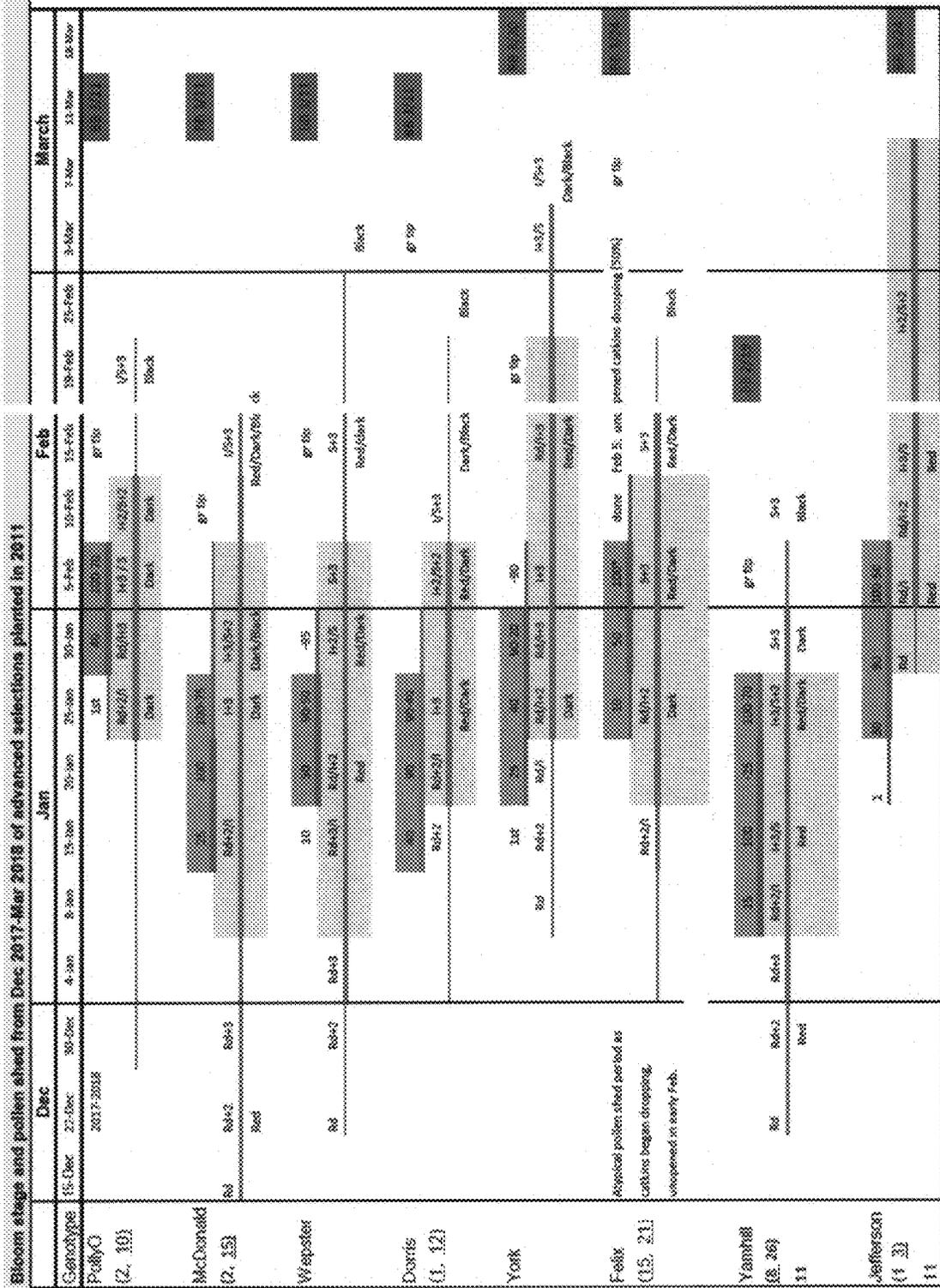


FIG. 14

