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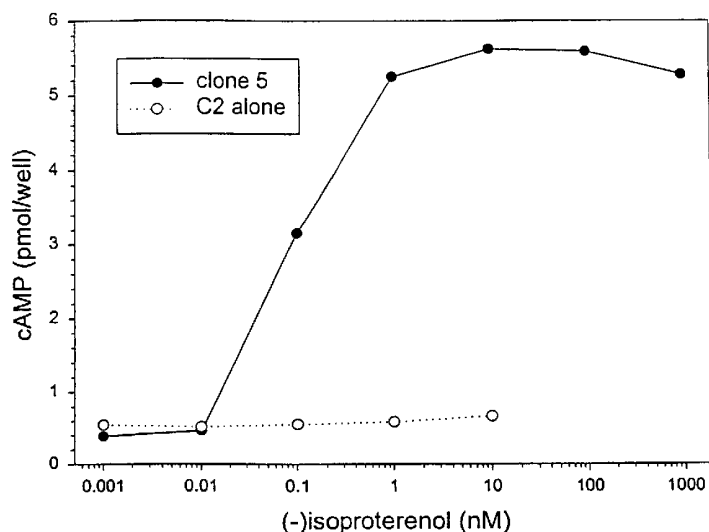
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(54) Title: IMPROVED SYSTEMS FOR SENSITIVE DETECTION OF G-PROTEIN COUPLED RECEPTOR AND ORPHAN RECEPTOR FUNCTION USING REPORTER ENZYME MUTANT COMPLEMENTATION

Agonist Stimulated cAMP Response in C2 Cells Expressing $\beta 2AR-\beta gal\Delta\alpha$



(57) Abstract: Methods for detecting G-protein coupled receptor (GPCR) activity; methods for assaying GPCR activity; and methods for screening for GPCR ligands, G-protein-coupled receptor kinase (GRK) activity, and compounds that interact with components of the GPCR regulatory process are described. Included are methods for expanding ICAST technologies for assaying GPCR activity with applications for ligand fishing, and agonist or antagonist screening. These methods include: engineering seronine/threonine phosphorylation sites into known or orphan GPCR open reading frames in order to increase the affinity of arrestin for the activated form of the GPCR or to increase the reside time of arrestin on the activated GPCR; engineering mutant arrestin proteins

that bind to activated GPCRs in the absence of G-protein coupled receptor kinases which may be limiting; and engineering mutant super arrestin proteins that have an increased affinity for activated GPCRs with or without phosphorylation. These methods are intended to increase the robustness of the GPCR/ICAST technology in situations in which G-protein coupled receptor kinases are absent or limiting, or in which the GPCR is not efficiently down-regulated or is rapidly resensitized (thus having a labile interaction with arrestin). Included are also more specific methods for using ICAST complementary enzyme fragments to monitor GPCR homo- and hetero- dimerization with applications for drug lead discovery and ligand and function discovery for orphan GPCRs.

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TITLE OF THE INVENTION**IMPROVED SYSTEMS FOR SENSITIVE DETECTION OF G-PROTEIN
COUPLED RECEPTOR AND ORPHAN RECEPTOR FUNCTION
USING REPORTER ENZYME MUTANT COMPLEMENTATION****BACKGROUND OF THE INVENTION**

This application is a continuation-in-part of U.S. Application Serial No. 09/654,499, filed September 1, 2000, which claims the benefit from Provisional Application Serial No. 60/180,669, filed February 7, 2000. The entirety of U.S. Application Serial No. 09/654,499 and Provisional Application Serial No. 60/180,669 are incorporated herein by reference.

Field of the Invention

The present invention relates to methods of detecting G-protein-coupled receptor (GPCR) activity, and provides methods of assaying GPCR activity, methods for screening for GPCR ligands, agonists and/or antagonists, methods for screening natural and surrogate ligands for orphan GPCRs, and methods for screening compounds that interact with components of the GPCR regulatory process.

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Background of the Technology

The actions of many extracellular signals are mediated by the interaction of G-protein-coupled receptors (GPCRs) and guanine nucleotide-binding regulatory proteins (G-proteins). G-protein-mediated signaling systems have been identified in many divergent organisms, such as mammals and yeast. The GPCRs represent a

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large super family of proteins which have divergent amino acid sequences, but share common structural features, in particular, the presence of seven transmembrane helical domains. GPCRs respond to, among other extracellular signals, neurotransmitters, hormones, odorants and light. Individual GPCR types activate a particular signal transduction pathway; at least ten different signal transduction pathways are known to be activated via GPCRs. For example, the beta 2-adrenergic receptor (β 2AR) is a prototype mammalian GPCR. In response to agonist binding, β 2AR receptors activate a G-protein (Gs) which in turn stimulates adenylate cyclase activity and results in increased cyclic adenosine monophosphate (cAMP) production in the cell.

The signaling pathway and final cellular response that result from GPCR stimulation depends on the specific class of G-protein with which the particular receptor is coupled (Hamm, "The Many Faces of G-Protein Signaling." J. Biol. Chem., 273:669-672 (1998)). For instance, coupling to the Gs class of G-proteins stimulates cAMP production and activation of the Protein Kinase A and C pathways, whereas coupling to the Gi class of G-proteins down regulates cAMP. Other second messenger systems such as calcium, phospholipase C, and phosphatidylinositol 3 may also be utilized. As a consequence, GPCR signaling events have predominantly been measured via quantification of these second messenger products.

The decrease of a response to a persistent stimulus is a widespread biological phenomenon. Signaling by diverse GPCRs is believed to be terminated by a uniform two-step mechanism. Activated receptor is first phosphorylated by a

GPCR kinase (GRK). An arrestin protein binds to the activated and phosphorylated receptor, thus blocking G-protein interaction. This process is commonly referred to as desensitization, a general mechanism that has been demonstrated in a variety of functionally diverse GPCRs. Arrestin also plays a part in regulating GPCR internalization and resensitization, processes that are heterogenous among different GPCRs (Oakley, et al., J. Biol. Chem., 274:32248-32257 (1999)). The interaction between an arrestin and GPCR in processes of internalization and resensitization is dictated by the specific sequence motif in the carboxyl terminus of a given GPCR. Only a subset of GPCRs, which possess clusters of three serine or threonine residues at the carboxyl termini, were found to co-traffick with the arrestins into the endocytic vesicles after ligand stimulation. The number of receptor kinases and arrestins involved in desensitization of GPCRs is rather limited.

A common feature of GPCR physiology is desensitization and recycling of the receptor through the processes of receptor phosphorylation, endocytosis and dephosphorylation (Ferguson, et al., "G-protein-coupled receptor regulation: role of G-protein-coupled receptor kinases and arrestins." Can. J. Physiol. Pharmacol., 74:1095-1110 (1996)). Ligand-occupied GPCRs can be phosphorylated by two families of serine/threonine kinases, the G-protein-coupled receptor kinases (GRKs) and the second messenger-dependent protein kinases such as protein kinase A and protein kinase C. Phosphorylation by either class of kinases serves to down-regulate the receptor by uncoupling it from its corresponding G-protein. GRK-phosphorylation also serves to down-regulate the receptor by recruitment of a

class of proteins known as the arrestins that bind the cytoplasmic domain of the receptor and promote clustering of the receptor into endocytic vesicles. Once the receptor is endocytosed, it will either be degraded in lysosomes or dephosphorylated and recycled back to the plasma membrane as a fully-functional
5 receptor.

Binding of an arrestin protein to an activated receptor has been documented as a common phenomenon of a variety of GPCRs ranging from rhodopsin to β 2AR to the neurotensin receptor (Barak, et al., "A β -arrestin/Green Fluorescent Fusion Protein Biosensor for Detecting G-Protein-Coupled Receptor Activation," J. Biol.
10 Chem., 272:27497-500 (1997)). Consequently, monitoring arrestin interaction with a specific GPCR can be utilized as a generic tool for measuring GPCR activation. Similarly, a single G-protein and GRK also partner with a variety of receptors (Hamm, et al. (1998) and Pitcher et al., "G-Protein-Coupled Receptor Kinases,"
15 Annu. Rev. Biochem., 67:653-92 (1998)), such that these protein/protein interactions may also be monitored to determine receptor activity.

Many therapeutic drugs in use today target GPCRs, as they regulate vital physiological responses, including vasodilation, heart rate, bronchodilation, endocrine secretion and gut peristalsis. See, e.g., Lefkowitz et al., Annu. Rev. Biochem., 52:159 (1983). Some of these drugs mimic the ligand for this receptor.
20 Other drugs act to antagonize the receptor in cases when disease arises from spontaneous activity of the receptor.

Efforts such as the Human Genome Project are identifying new GPCRs ("orphan" receptors) whose physiological roles and ligands are unknown. It is estimated that several thousand GPCRs exist in the human genome.

Various approaches have been used to monitor intracellular activity in response to a stimulant, e.g., enzyme-linked immunosorbent assay (ELISA); Fluorescence Imaging Plate Reader assay (FLIPR™, Molecular Devices Corp., Sunnyvale, CA); EVOscreen™, EVOTEC™, Evotec Biosystems GmbH, Hamburg, Germany; and techniques developed by CELLOMICS™, Cellomics, Inc., Pittsburgh, PA.

10 Germino et al., "Screening for *in vivo* protein-protein interactions." Proc. Natl. Acad. Sci., 90(3):933-937 (1993), discloses an *in vivo* approach for the isolation of proteins interacting with a protein of interest.

Phizicky et al., "Protein-protein interactions: methods for detection and analysis." Microbiol. Rev., 59(1): 94-123 (1995), discloses a review of
15 biochemical, molecular biological and genetic methods used to study protein-protein interactions.

Offermanns et al., " $G_{\alpha_{15}}$ and $G_{\alpha_{16}}$ Couple a Wide Variety of Receptors to Phospholipase C." J. Biol. Chem., 270(25):15175-15180 (1995), discloses that $G_{\alpha_{15}}$ and $G_{\alpha_{16}}$ can be activated by a wide variety of G-protein-coupled receptors.
20 The selective coupling of an activated receptor to a distinct pattern of G-proteins is regarded as an important requirement to achieve accurate signal transduction. Id.

Barak et al., "A β -arrestin/Green Fluorescent Protein Biosensor for Detecting G Protein-coupled Receptor Activation." J. Biol. Chem., 272(44):27497-

27500 (1997) and U.S. Patents Nos. 5,891,646 and 6,110,693 disclose the use of a β -arrestin/green fluorescent fusion protein (GFP) for imaging protein translocation upon stimulation of GPCR with optical devices.

Each of the references described above has drawbacks. For example,

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- The prior art methodologies require over-expression of the proteins, which could cause artifact and tip the balance of cellular regulatory machineries.
 - The prior art visualization or imaging assays are low throughput and lack thorough quantification. Therefore, they are not suitable for
- 10 high throughput pharmacological and kinetic assays.

In addition, many of the prior art assays require isolation of the GPCR rather than observation of the GPCR in a cell. There thus exists a need for improved methods for monitoring GPCR function.

15 **SUMMARY OF THE INVENTION**

The present invention provides modifications to the disclosure in U.S. Application Serial No. 09/654,499. In particular, the present invention is directed to modifications of the below aspects of the invention to further enhance assay sensitivity. The modifications include the use of genetically modified arrestins that

20 exhibit enhanced binding to activated GPCR regardless of whether the GPCR is phosphorylated or non-phosphorylated; the use of a serine/threonine cluster strategy to facilitate screening assays for orphan receptors that do not possess this

structural motif on their own; and the use of a combination of the above modifications to achieve even more enhanced detection.

A first aspect of the present invention is a method that monitors GPCR function proximally at the site of receptor activation, thus providing more information for drug discovery purposes due to fewer competing mechanisms. 5
Activation of the GPCR is measured by a read-out for interaction of the receptor with a regulatory component such as arrestin, G-protein, GRK or other kinases, the binding of which to the receptor is dependent upon agonist occupation of the receptor. The present invention involves the detection of protein/protein 10
interaction by complementation of mutant reporter enzymes.

Binding of arrestin to activated GPCR is a common process in the first step of desensitization that has been demonstrated for most, if not all, GPCRs studied so far. Measurement of GPCR interaction with arrestin via mutant enzyme complementation (*i.e.*, ICAST) provides a more generic assay technology 15
applicable for a wide variety of GPCRs and orphan receptors.

A further aspect of the present invention is a method of assessing GPCR pathway activity under test conditions by providing a test cell that expresses a GPCR, *e.g.*, muscarinic, adrenergic, dopamine, angiotensin or endothelin, as a fusion protein to a mutant reporter enzyme and interacting a protein in the GPCR 20
pathway, *e.g.*, G-protein, arrestin or GRK, as a fusion protein with a complementing mutant reporter enzyme. When test cells are exposed to a known agonist to the target GPCR under test conditions, activation of the GPCR will be

monitored by complementation of the reporter enzyme. Increased reporter enzyme activity reflects interaction of the GPCR with its interacting protein partner.

A further aspect of the present invention is a method of assessing GPCR pathway activity in the presence of a test arrestin, e.g., β -arrestin.

5 A further aspect of the present invention is a method of assessing GPCR pathway activity in the presence of a test G-protein.

A further aspect of the present invention is a method of assessing GPCR pathway activity upon exposure of the test cell to a test ligand.

A further aspect of the present invention is a method of assessing GPCR
10 activity upon co-expression in the test cell of a second receptor. The second receptor could be the same GPCR or orphan receptor (i.e., homo-dimerization), a different GPCR or orphan receptor (i.e., hetero-dimerization) or could be a receptor of another type.

A further aspect of the present invention is a method for screening for a
15 ligand or agonist to an orphan GPCR. The ligand or agonist could be contained in natural or synthetic libraries or mixtures or could be a physical stimulus. A test cell is provided that expresses the orphan GPCR as a fusion protein with a mutant reporter enzyme, e.g., a β -galactosidase mutant, and, for example, an arrestin or mutant form of arrestin as a fusion protein with a complementing mutant reporter
20 enzyme, e.g., another β -galactosidase mutant. The interaction of the arrestin with the orphan GPCR upon receptor activation is measured by enzymatic activity of the complemented reporter enzyme. The test cell is exposed to a test compound, and an increase in reporter enzyme activity indicates the presence of a ligand or agonist.

A further aspect of the present invention is a method for screening a protein of interest, for example, an arrestin protein (or mutant form of the arrestin protein) for the ability to bind to a phosphorylated, or activated, GPCR. A test cell is provided that expresses a GPCR as a fusion protein with a mutant reporter enzyme, e.g., a β -galactosidase mutant, and contains arrestin (or a mutant form of arrestin) as a fusion protein with a complementing mutant reporter enzyme, e.g., another β -galactosidase mutant. The interaction of arrestin with the GPCR upon receptor activation is measured by enzymatic activity of the complemented reporter enzyme. The test cell is exposed to a known GPCR agonist and then reporter enzyme activity is detected. Increased reporter enzyme activity indicates that the β -arrestin molecule can bind to phosphorylated, or activated, GPCR in the test cell.

A further aspect of the present invention is a method to screen for an agonist to a specific GPCR. The agonist could be contained in natural or synthetic libraries or could be a physical stimulus. A test cell is provided that expresses a GPCR as a fusion protein with a mutant reporter enzyme, e.g., a β -galactosidase mutant, and, for example, an arrestin as a fusion protein with a complementing mutant reporter enzyme, e.g., another β -galactosidase mutant. The interaction of arrestin with the GPCR upon receptor activation is measured by enzymatic activity of the complemented reporter enzyme. The test cell is exposed to a test compound, and an increase in reporter enzyme activity indicates the presence of an agonist. The test cell may express a known GPCR or a variety of known GPCRs, or may express an unknown GPCR or a variety of unknown GPCRs. The GPCR may be, for example, an odorant GPCR or a β AR GPCR.

A further aspect of the present invention is a method for screening a test compound for GPCR antagonist activity. A test cell is provided that expresses a GPCR as a fusion protein with a mutant reporter enzyme, e.g., a β -galactosidase mutant, and, for example, an arrestin as a fusion protein with a complementing mutant reporter enzyme, e.g., another β -galactosidase mutant. The interaction of arrestin with the GPCR upon receptor activation is measured by enzymatic activity of the complemented reporter enzyme. The test cell is exposed to a test compound, and an increase in reporter enzyme activity indicates the presence of an agonist. The cell is exposed to a test compound and to a GPCR agonist, and reporter enzyme activity is detected. When exposure to the agonist occurs at the same time as or subsequent to exposure to the test compound, a decrease in reporter enzyme activity after exposure to the test compound indicates that the test compound has antagonist activity to the GPCR.

A further aspect of the present invention is a method of screening a sample solution for the presence of an agonist, antagonist or ligand to a GPCR. A test cell is provided that expresses GPCR as a fusion protein with a mutant reporter enzyme, e.g., a β -galactosidase mutant, and contains, for example, a β -arrestin as a fusion protein with a complementing reporter, e.g., another β -galactosidase mutant. The test cell is exposed to a sample solution, and reporter enzyme activity is assessed. Changed reporter enzyme activity after exposure to the sample solution indicates the sample solution contains an agonist, antagonist or ligand for a GPCR expressed in the cell.

A further aspect of the present invention is a method of screening a cell for the presence of a GPCR. According to this aspect, an arrestin fusion protein with a mutant reporter enzyme and a GPCR downstream signaling fusion protein with a mutant reporter enzyme are employed to detect GPCR action. A modification of this aspect of the invention can be employed to provide a method of screening a plurality of cells for those cells which contain a GPCR. According to this aspect, a plurality of cells containing a conjugate comprising a β -arrestin protein as a fusion protein with a reporter enzyme are provided; the plurality of cells are exposed to a GPCR agonist; and activity of reporter enzyme activity is detected. An increase in reporter enzymatic activity after exposure to the GPCR agonist indicates β -arrestin protein binding to a GPCR, thereby indicating that the cell contains a GPCR responsive to the GPCR agonist.

A further aspect of the invention is a method for mapping GPCR-mediated signaling pathways. For instance, the system could be utilized to monitor interaction of c-src with β -arrestin-1 upon GPCR activation. Additionally, the system could be used to monitor protein/protein interactions involved in cross-talk between GPCR signaling pathways and other pathways such as that of the receptor tyrosine kinases or Ras/Raf. According to this aspect, a test cell is provided that expresses a GPCR or other related protein with a mutant reporter enzyme, e.g., a β -galactosidase mutant, and contains a protein from another pathway as a fusion protein with a complementing mutant reporter enzyme, e.g., another β -galactosidase mutant. Increased reporter enzymatic activity indicates protein/protein interaction.

A further aspect of the invention is a method for monitoring homo- or hetero- dimerization of GPCRs upon agonist or antagonist stimulation. Increasing evidence indicates that GPCR dimerization is important for biological activity (AbdAlla, et al., "AT1-receptor heterodimers show enhanced G-protein activation and altered receptor sequestration." *Nature*, 407:94-98 (2000); Bockaert, et al., "Molecular tinkering of G protein-coupled receptors: an evolutionary success." *EMBO J.* 18:1723-29 (1999)). Jordan, et al., "G-protein-coupled receptor heterodimerization modulates receptor function." *Nature*, 399:697-700 (1999), demonstrated that two non-functional opioid receptors, κ and δ , heterodimerize to form a functional receptor. Gordon et al., "Dopamine D2 receptor dimers and receptor blocking peptides." *Bioch. Biophys. Res. Commun.* 227:200-204 (1996), showed different pharmacological properties associated with the monomeric and dimeric forms of Dopamine receptor D2. The D2 receptors exist either as monomers that are selective targets for spiperone or as dimer forms that are targets for nemonapride. Herbert, et al., "A peptide derived from a β 2-adrenergic receptor transmembrane domain inhibits both receptor dimerization and activation." *J.B.C.* 271:16384-92 (1996), demonstrated that the agonist stimulation was found to stabilize the dimeric state of the receptor, whereas inverse agonists favored the monomeric form. Indeed, the same study showed that a peptide corresponding to the sixth transmembrane domain of the β 2-adrenergic receptor inhibited both receptor dimerization and activation. Further, Angers et al., Detection of beta-2-adrenergic receptor dimerization in living cells using bioluminescence resonance energy transfer, *Proc. Natl. Acad. Sci. USA*, 97(7):3684-3689, discloses the use of

β 2-adrenergic receptor fusion proteins (i.e., β 2-adrenergic receptor fused to luciferase and β 2-adrenergic receptor fused to an enhanced red-shifted green fluorescent protein) to study β 2-adrenergic receptor dimerization.

GPCR dimerization in the context of cellular physiology and
5 pharmacology can be monitored in accordance with the invention. For example, β -galactosidase complementation can be measured in test cells that co-express GPCR fusion proteins of β -galactosidase mutant enzymes, e.g., GPCR₁ $\Delta\alpha$ and GPCR₂ $\Delta\omega$ (FIGURE 27). According to this aspect, the interconversion between monomeric to dimeric forms of the GPCRs or orphan receptors can be measured by mutant
10 reporter enzyme complementation. FIGURE 27 illustrates a test cell co-expressing GPCR or an orphan receptor as a fusion protein with $\Delta\alpha$ form of β -galactosidase mutant (e.g., GPCR₁ $\Delta\alpha$), and the same GPCR or orphan receptor as a fusion protein with $\Delta\omega$ form of β -galactosidase mutant (e.g., GPCR₁ $\Delta\omega$). Formation of the GPCR homodimer is reflected by formation of an active enzyme, which can be
15 measured by enzyme activity assays, such as the Gal-Screen™ assay. Similarly, hetero-dimerization between two distinct GPCRs, or two distinct orphan receptors, or between one known GPCR and one orphan receptor can be analyzed in test cells co-expressing two fusion proteins, e.g., GPCR₁ $\Delta\alpha$ and GPCR₂ $\Delta\omega$. The increased β -galactosidase activity indicates that the two receptors can form a heterodimer.

20 A further aspect of the invention is a method of monitoring the interconversion between the monomeric and dimeric form of GPCRs under the influence of agonist or antagonist treatment. The test receptor(s) can be between the same GPCR or orphan receptor (homodimer), or between two distinct GPCRs

or orphan receptors (heterodimer). The increased β -galactosidase activity after treatment with a compound means that the compound binds to and/or stabilizes the dimeric form of the receptor. The decreased β -galactosidase activity after treatment with a compound means that the compound binds to and/or stabilizes the
5 monomeric form of the receptor.

A further aspect of the invention is a method of screening a cell for the presence of a GPCR responsive to a GPCR agonist. A cell is provided that contains protein partners that interact downstream in the GPCR's pathway. The protein partners are expressed as fusion proteins to the mutant, complementing
10 enzyme and are used to monitor activation of the GPCR. The cell is exposed to a GPCR agonist and then enzymatic activity of the reporter enzyme is detected. Increased reporter enzyme activity indicates that the cell contains a GPCR responsive to the agonist.

The present invention involves the use of a combination of proprietary
15 technologies (including ICASTTM, Intercistronic Complementation Analysis Screening Technology, Gal-ScreenTM, etc.) to monitor protein/protein interactions in GPCR signaling. As disclosed in U.S. Application Serial No. 09/654,499, the method of the invention in part involves using ICASTTM, which in turn involves the use of two inactive β -galactosidase mutants, each of which is fused with one of
20 two interacting target protein pairs, such as a GPCR and an arrestin. The formation of an active β -galactosidase complex is driven by interaction of the target proteins. In this system, β -galactosidase activity can be detected using, e.g., the Gal-ScreenTM assay system, wherein direct cell lysis is combined with rapid

ultrasensitive chemiluminescent detection of β -galactosidase reporter enzyme.

This system uses, e.g., a Galacton-*Star*® chemiluminescent substrate for measurement in a luminometer as a read out of GPCR activity.

FIGURE 23 is a schematic depicting the use of the complementation
5 technology in the method of the present invention. FIGURE 23 shows two inactive
 β -galactosidase mutants that become active when they are forced together by
specific interactions between the fusion partners of an arrestin molecule and an
activated GPCR or orphan receptor. This assay technology will be especially
useful in high throughput screening assays for ligand fishing for orphan receptors, a
10 process called de-orphaning. As illustrated in FIGURE 28, a β -galactosidase
fusion protein of an orphan receptor (e.g., GPCR_{orphan} $\Delta\alpha$) is co-expressed in the test
cell with a fusion protein of β -arrestin (e.g., β -Arr $\Delta\omega$). When the test cell is
subjected to compounds, which could be natural or synthetic, the increased β -
galactosidase activity means the compound is either a natural or surrogate ligand
15 for this GPCR. The same assay system can be used to find drug leads for the new
GPCRs. The increased β -galactosidase activity in the test cell after treatment
indicates the agonist activity of the compound. The decreased β -galactosidase
activity in the test cell indicates antagonist activity or inverse agonist activity of the
compound. In addition, the method of the invention could be used to monitor
20 GPCR-mediated signaling pathways via other downstream signaling components
such as G-proteins, GRKs or the proto-oncogene c-Src.

The invention is achieved in part by using ICAST™ protein/protein
interaction screening to map signaling pathways. This technology is applicable to

a variety of known and unknown GPCRs with diverse functions. They include, but are not limited to, the following sub-families of GPCRs:

(a) receptors that bind to amine-like ligands-Acetylcholine muscarinic receptor (M1 to M5), alpha and beta Adrenoceptors, Dopamine receptors (D1, D2, 5 D3 and D4), Histamine receptors (H1 and H2), Octopamine receptor and Serotonin receptors (5HT1, 5HT2, 5HT4, 5HT5, 5HT6, 5HT7);

(b) receptors that bind to a peptide ligand-Angiotensin receptor, Bombesin receptor, Bradykinin receptor, C-C chemokine receptors (CCR1 to CCR8, and CCR10), C-X-C type Chemokine receptors (CXC-R5), Cholecystokinin type A 10 receptor, CCK type receptors, Endothelin receptor, Neurotesin receptor, FMLP-related receptors, Somatostatin receptors (type 1 to type 5) and Opioid receptors (type D, K, M, X);

(c) receptors that bind to hormone proteins-Follic stimulating hormone receptor, Thyrotrophin receptor and Lutropin-choriogonadotropic hormone 15 receptor;

(d) receptors that bind to neurotransmitters-substance P receptor, Substance K receptor and neuropeptide Y receptor;

(e) Olfactory receptors-Olfactory type 1 to type 11, Gustatory and odorant receptors;

(f) Prostanoid receptors-Prostaglandin E2 (EP1 to EP4 subtypes), 20 Prostacyclin and Thromboxane;

(g) receptors that bind to metabotropic substances-Metabotropic glutamate group I to group III receptors;

(h) receptors that respond to physical stimuli, such as light, or to chemical stimuli, such as taste and smell; and

(i) orphan GPCRs-the natural ligand to the receptor is undefined.

Use of the ICAST™ technology in combination with the invention
5 provides many benefits to the GPCR screening process, including the ability to monitor protein interactions in any sub-cellular compartment-membrane, cytosol and nucleus; the ability to achieve a more physiologically relevant model without requiring protein overexpression; and the ability to achieve a functional assay for receptor binding allowing high information content.

10

BRIEF DESCRIPTION OF THE DRAWINGS

FIGURE 1. Cellular expression levels of β 2 adrenergic receptor (β 2AR) and β -arrestin-2 (β Arr2) in C2 clones. Quantification of β -galactosidase (β -gal) fusion protein was performed using antibodies against β -gal and purified β -gal
15 protein in a titration curve by a standardized ELISA assay. Figure 1A shows expression levels of β 2AR- β gal $\Delta\alpha$ clones (in expression vector pICAST ALC). Figure 1B shows expression levels of β Arr2- β gal $\Delta\omega$ in expression vector pICAST OMC4 for clones 9-3, -7, -9, -10, -19 and -24, or in expression vector pICAST OMN4 for clones 12-4, -9, -16, -18, -22 and -24.

20 FIGURE 2. Receptor β 2AR activation was measured by agonist-stimulated cAMP production. C2 cells expressing pICAST ALC β 2AR (clone 5) or parental cells were treated with increasing concentrations of (-)-isoproterenol and 0.1mM

IBMX. The quantification of cAMP level was expressed as pmol/well.

FIGURE 3. Interaction of activated receptor β 2AR and arrestin can be measured by β -galactosidase complementation. Figure 3A shows a time course of β -galactosidase activity in response to agonist (-)isoproterenol stimulation in C2
5 expressing β 2AR- β gal $\Delta\alpha$ (β 2AR alone, in expression vector pICAST ALC), or a pool of doubly transduced C2 co-expressing β 2AR- β gal $\Delta\alpha$ and β Arr2- β gal $\Delta\omega$ (in expression vectors pICAST ALC and pICAST OMC and clones isolated from the same pod (43-1, 43-2, 43-7 and 43-8)). Figure 3B shows a time course of β -galactosidase activity in response to agonist (-)isoproterenol stimulation in C2 cells
10 expressing β 2AR- β gal $\Delta\alpha$ alone (in expression vector pICAST ALC) and C2 clones co-expressing β 2AR- β gal $\Delta\alpha$ and β Arr1- β gal $\Delta\omega$ (in expression vectors ICAS ALC and pICAST OMC).

FIGURE 4. Agonist dose response for interaction of β 2AR and arrestin can be measured by β -galactosidase complementation. Figure 4A shows a dose
15 response to agonists (-)isoproterenol and procaterol in C2 cells co-expressing β 2AR- β gal $\Delta\alpha$ and β Arr2- β gal $\Delta\omega$ fusion constructs. Figure 4B shows a dose response to agonists (-)isoproterenol and procaterol in C2 cells co-expressing β 2AR- β gal $\Delta\alpha$ and β Arr1- β gal $\Delta\omega$ fusion constructs.

FIGURE 5. Antagonist mediated inhibition of receptor activity can be
20 measured by β -galactosidase complementation in cells co-expressing β 2AR- β gal $\Delta\alpha$ and β Arr- β gal $\Delta\omega$. Figure 5A shows specific inhibition with adrenergic

antagonists ICI-118,551 and propranolol of β -galactosidase activity in C2 clones co-expressing β 2AR- β gal $\Delta\alpha$ and β Arr2- β gal $\Delta\omega$ fusion constructs after incubation with agonist (-)isoproterenol. Figure 5B shows specific inhibition of β -galactosidase activity with adrenergic antagonists ICI-118,551 and propranolol in C2 clones co-expressing β 2AR- β gal $\Delta\alpha$ and β Arr1- β gal $\Delta\omega$ fusion constructs in the presence of agonist (-)isoproterenol.

FIGURE 6. C2 cells expressing adenosine receptor A2a show cAMP induction in response to agonist (CGS-21680) treatment. C2 parental cells and C2 cells co-expressing A2aR- β gal $\Delta\alpha$ and β Arr1- β gal $\Delta\omega$ as a pool or as selected clones (47-2 and 47-13) were measured for agonist-induced cAMP response (pmol/well).

FIGURE 7. Agonist stimulated cAMP response in C2 cells co-expressing Dopamine receptor D1 (D1- β gal $\Delta\alpha$) and β -arrestin-2 (β Arr2- β gal $\Delta\omega$). The clone expressing β Arr2- β gal $\Delta\omega$ (Arr2 alone) was used as a negative control in the assay. Cells expressing D1- β gal $\Delta\alpha$ in addition to β Arr2- β gal $\Delta\omega$ responded agonist treatment (3-hydroxytyramine hydrochloride at 3 μ M). D1(PIC2) or D1(PIC3) designate D1 in expression vector pICAST ALC2 or pICAST ALC4, respectively.

FIGURE 8. Variety of mammalian cell lines can be used to generate stable cells for monitoring GPCR and arrestin interactions. FIGURE 8A, FIGURE 8B and FIGURE 8C show the examples of HEK 293, CHO and CHW cell lines co-expressing adrenergic receptor β 2AR and arrestin fusion proteins of β -

galactosidase mutants. The β -galactosidase activity was used to monitor agonist-induced interaction of β 2AR and arrestin proteins.

FIGURE 9. Beta-gal complementation can be used to monitor β 2 adrenergic receptor homo-dimerization. FIGURE 9A shows β -galactosidase activity in HEK 293 clones co-expressing β 2AR- β gal $\Delta\alpha$ and β 2AR- β gal $\Delta\omega$.
5 FIGURE 9B shows a cAMP response to agonist (-)isoproterenol in HEK 293 clones co-expressing β 2AR- β gal $\Delta\alpha$ and β 2AR- β gal $\Delta\omega$. HEK293 parental cells were included in the assays as negative controls.

FIGURE 10A. pICAST ALC: Vector for expression of β -gal $\Delta\alpha$ as a C-terminal fusion to the target protein. This construct contains the following
10 features: MCS, multiple cloning site for cloning the target protein in frame with the β -gal $\Delta\alpha$; GS Linker, (GGGGS) n ; NeoR, neomycin resistance gene; IRES, internal ribosome entry site; ColE1ori, origin of replication for growth in *E. coli*; 5'MoMuLV LTR and 3'MoMuLV LTR, viral promoter and polyadenylation
15 signals from the Moloney Murine leukemia virus.

FIGURE 10B. Nucleotide sequence for pICAST ALC.

FIGURE 11A. pICAST ALN: Vector for expression of β -gal $\Delta\alpha$ as an N-terminal fusion to the target protein. This construct contains the following
features: MCS, multiple cloning site for cloning the target protein in frame with the
20 β -gal $\Delta\alpha$; GS Linker, (GGGGS) n ; NeoR, neomycin resistance gene; IRES, internal ribosome entry site; ColE1ori, origin of replication for growth in *E. coli*;

5'MoMuLV LTR and 3'MoMuLV LTR, viral promoter and polyadenylation signals from the Moloney Murine leukemia virus.

FIGURE 11B. Nucleotide sequence for pICAST ALN.

FIGURE 12A. pICAST OMC: Vector for expression of β -gal $\Delta\omega$ as a C-terminal fusion to the target protein. This construct contains the following features: MCS, multiple cloning site for cloning the target protein in frame with the β -gal $\Delta\omega$; GS Linker, (GGGGS) $_n$; Hygro, hygromycin resistance gene; IRES, internal ribosome entry site; ColE1ori, origin of replication for growth in *E. coli*; 5'MoMuLV LTR and 3'MoMuLV LTR, viral promoter and polyadenylation signals from the Moloney Murine leukemia virus.

FIGURE 12B. Nucleotide sequence for pICAST OMC.

FIGURE 13A. pICAST OMN: Vector for expression of β -gal $\Delta\omega$ as an N-terminal fusion to the target protein. This construct contains the following features: MCS, multiple cloning site for cloning the target protein in frame with the β -gal $\Delta\omega$; GS Linker, (GGGGS) $_n$; Hygro, hygromycin resistance gene; IRES, internal ribosome entry site; ColE1ori, origin of replication for growth in *E. coli*; 5'MoMuLV LTR and 3'MoMuLV LTR, viral promoter and polyadenylation signals from the Moloney Murine leukemia virus.

FIGURE 13B. Nucleotide sequence for pICAST OMN.

FIGURE 14. pICAST ALC β Arr2: Vector for expression of β -gal $\Delta\alpha$ as a C-terminal fusion to β -arrestin-2. The coding sequence of human β -arrestin-2 (Genebank Accession Number: NM_004313) was cloned in frame to β -gal $\Delta\alpha$ in a

pICAST ALC vector.

FIGURE 15. pICAST OMC β Arr2: Vector for expression of β -gal $\Delta\omega$ as a C-terminal fusion to β -arrestin-2. The coding sequence of human β -arrestin-2 (Genebank Accession Number: NM_004313) was cloned in frame to β -gal $\Delta\omega$ in a pICAST OMC vector.

FIGURE 16. pICAST ALC β Arr1: Vector for expression of β -gal $\Delta\alpha$ as a C-terminal fusion to β -arrestin-1. The coding sequence of human β -arrestin-1 (Genebank Accession Number: NM_004041) was cloned in frame to β -gal $\Delta\alpha$ in a pICAST ALC vector.

FIGURE 17. pICAST OMC β Arr1: Vector for expression of β -gal $\Delta\omega$ as a C-terminal fusion to β -arrestin-1. The coding sequence of human β -arrestin-1 (Genebank Accession Number: NM_004041) was cloned in frame to β -gal $\Delta\omega$ in a pICAST OMC vector.

FIGURE 18. pICAST ALC β 2AR: Vector for expression of β -gal $\Delta\alpha$ as a C-terminal fusion to β 2 Adrenergic Receptor. The coding sequence of human β 2 Adrenergic Receptor (Genebank Accession Number: NM_000024) was cloned in frame to β -gal $\Delta\alpha$ in a pICAST ALC vector.

FIGURE 19. pICAST OMC β 2AR: Vector for expression of β -gal $\Delta\omega$ as a C-terminal fusion β 2 Adrenergic Receptor. The coding sequence of human β 2 Adrenergic Receptor (Genebank Accession Number: NM_000024) was cloned in frame to β -gal $\Delta\omega$ in a pICAST OMC vector.

FIGURE 20. pICAST ALC A2aR: Vector for expression of β -gal $\Delta\alpha$ as a C-terminal fusion to Adenosine 2a Receptor. The coding sequence of human Adenosine 2a Receptor (Genebank Accession Number: NM_000675) was cloned in frame to β -gal $\Delta\alpha$ in a pICAST ALC vector.

5 FIGURE 21. pICAST OMC A2aR: Vector for expression of β -gal $\Delta\omega$ as a C-terminal fusion to Adenosine 2a Receptor. The coding sequence of human Adenosine 2a Receptor (Genebank Accession Number: NM_000675) was cloned in frame to β -gal $\Delta\omega$ in a pICAST OMC vector.

FIGURE 22. pICAST ALC D1: Vector for expression of β -gal $\Delta\alpha$ as a C-
10 terminal fusion to Dopamine D1 Receptor. The coding sequence of human Dopamine D1 Receptor (Genebank Accession Number: X58987) was cloned in frame to β -gal $\Delta\alpha$ in a pICAST ALC vector.

FIGURE 23. A schematic depicting use of the complementation
technology in the method of the invention. FIGURE 23 shows two inactive
15 mutant reporter enzymes that become active when the corresponding fusion partners, GPCR and β -arrestin interact.

FIGURE 24. Vector for expression of a GPCR with inserted seronine/threonine amino acid sequences as a fusion with β -gal $\Delta\alpha$. The open reading frame of a known or orphan GPCR is engineered to contain additional
20 seronine/threonine sequences, such as SSS (seronine, seronine, seronine), within the C-terminal tail. The engineered GPCR is cloned in frame with β -gal $\Delta\alpha$ in a pICAST ALC vector. The pICAST ALC vector contains the following features:

MCS, multiple cloning site for cloning the target protein in frame with the β -gal $\Delta\alpha$;
GS Linker, (GGGGS) n ; NeoR, neomycin resistance gene; IRES, internal ribosome
entry site; ColE1ori, origin of replication for growth in *E. coli*; 5'MoMuLV LTR
and 3'MoMuLV LTR, viral promoter and polyadenylation signals from the
5 Moloney Murine leukemia virus.

FIGURE 25. Vector for expression of mutant (R170E) β -arrestin2 as a
fusion with β -gal $\Delta\omega$. The open reading frame of β -arrestin2 is engineered to
contain a point mutation that converts arginine 170 to a glutamate. The mutant β -
arrestin2 is cloned in frame with β -gal $\Delta\omega$ in a pICAST OMC vector. The pICAST
10 OMC vector contains the following features: MCS, multiple cloning site for
cloning the target protein in frame with the β -gal $\Delta\alpha$; GS Linker, (GGGGS) n ;
Hygro, hygromycin resistance gene; IRES, internal ribosome entry site; ColE1ori,
origin of replication for growth in *E. coli*; 5'MoMuLV LTR and 3'MoMuLV LTR,
viral promoter and polyadenylation signals from the Moloney Murine leukemia
15 virus.

FIGURE 26. Phosphorylation insensitive Mutant R170E β -Arrestin2 $\Delta\omega$
binds to β 2AR $\Delta\alpha$ in Response to Agonist Activation. A parental β 2AR $\Delta\alpha$ C2 cell
line was transduced with the Mutant R170E β -Arrestin2 $\Delta\omega$ construct. Clonal
populations co-expressing the two constructions were plated at 10,000 cells/well in
20 96 well plates and treated with 10 μ M (-)isoproterenol, 0.3mM ascorbic acid for the
indicated time period. β -galactosidase activity was measured by addition of Tropix
Gal-ScreenTM assay system substrate (Applied Biosystems) and luminescence was
measured using a Tropix TR717TM luminometer (Applied Biosystems). Treatments

were performed in triplicate. For comparison, a clonal cell line (43-8) co-expressing $\beta 2AR\Delta\alpha$ and wild-type β -Arrestin2 $\Delta\omega$ was also plated at 10,000 cells/well and given the same agonist treatment regimen. Minutes of (-)isoproterenol treatment is shown on the X-axis and β -galactosidase activity indicated by relative light units (RLU) is shown on the Y-axis.

FIGURE 27. GPCR dimerization measured by β -galactosidase complementation. A schematic depicting the utilization of the invention for monitoring GPCR homo- or hetero- dimerization. One GPCR is fused to one complement enzyme fragment, while the second GPCR is fused to the second complement enzyme fragment. Interaction of the two GPCRs is monitored by complementation of the enzyme fragments to produce an active enzyme complex (i.e., β -galactosidase activity). GPCR homo- or hetero- dimerization can be monitored in the absence or presence of ligand, agonists, inverse agonists or antagonists.

FIGURE 28. Ligand fishing for orphan receptors by β -galactosidase mutant complementation in ICAST™ system. A schematic depicting the utilization of the invention for ligand fishing and agonist/antagonist screening for orphan GPCRs. As an example, a test cell expressing two β -gal fusion proteins, $GPCR_{\text{orphan}}\Delta\alpha$ and Arrestin- $\Delta\omega$, is subjected to treatments with samples from natural or synthetic compound libraries, or from tissue extracts, or from conditioned media of cultured cells. An increased β -gal activity after treatment indicates the activation of the orphan receptor by a ligand in the testing sample. The readout of increased β -gal activity reflects the interaction of an activated

GPCR orphan receptor with a β -arrestin. Therefore, a cognate or a surrogate ligand for the testing receptor is identified.

DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS

5 The present invention provides a method to interrogate GPCR function and pathways. The G-protein-coupled superfamily continues to expand rapidly as new receptors are discovered through automated sequencing of cDNA libraries or genomic DNA. It is estimated that several thousand GPCRs may exist in the human genome. Only a portion have been cloned and even fewer have been
10 associated with ligands. The means by which these, or newly discovered orphan receptors, will be associated with their cognate ligands and physiological functions represents a major challenge to biological and biomedical research. The identification of an orphan receptor generally requires an individualized assay and a guess as to its function. The present invention involves the interrogation of
15 GPCR function by monitoring the activation of the receptor using activation dependent protein-protein interactions between the test GPCR or orphan receptor and a β -arrestin. The specific protein-protein interactions are measured using the mutant enzyme complementation technology disclosed herein. This assay system eliminates the prerequisite guessing because it can be performed with and without
20 prior knowledge of other signaling events. It is sensitive, rapid and easily performed and is applicable to nearly all GPCRs because the majority of these receptors desensitize by a common mechanism.

The present invention provides a complete assay system for monitoring

protein-protein interactions in GPCR pathways. The invention employs the complementation technology, ICAST™ (Intercistronic Complementation Analysis Screening Technology as disclosed in pending U.S. patent application serial no. 053,614, filed April 1, 1998, the entire contents of which are incorporated herein

5 by reference). The ICAST™ technology involves the use of two mutant forms of a reporter enzyme fused to proteins of interest. When the proteins of interest do not interact, the reporter enzyme remains inactive. When the proteins of interest do interact, the reporter enzyme mutants come together and form an active enzyme.

According to an embodiment of the invention, the activity of β -galactosidase may

10 be detected with the Gal-Screen™ assay system developed by Advanced Discovery Sciences™, which involves the use of Galacton-*Star*®, an ultrasensitive chemiluminescent substrate. The Gal-Screen™ assay system and the Galacton-*Star*® chemiluminescent substrate are disclosed in U.S. Patent Nos. 5,851,771; 5,538,847; 5,326,882; 5,145,772; 4,978,614; and 4,931,569, the contents of which

15 are incorporated herein by reference in their entirety. The invention provides an array of assays, including GPCR binding assays, that can be achieved directly within the cellular environment in a rapid, non-radioactive assay format. The methods of the invention are an advancement over the invention disclosed in U.S. Patent Nos. 5,891,646 and 6,110,693 and the method disclosed in Angers et al.,

20 supra., which rely on microscopic imaging or spectrometry of GPCR components as fusion with Green-fluorescent-protein. The imaging technique disclosed in U.S. Patent Nos. 5,891,646 and 6,110,693 and spectrometry-based technique in Angers et al. are limited by low-throughput and lack of thorough quantification.

The assay system of the invention combined with Advanced Discovery Sciences™ technologies provide highly sensitive cell-based methods for interrogating GPCR pathways which are amenable to high-throughput screening (HTS). Among some of the technologies developed by Advanced Discovery Sciences™ that may be used with the present invention are the Gal-Screen™ assay system (discussed above) and the cAMP-Screen™ immunoassay system. The cAMP-Screen™ immunoassay system provides ultrasensitive determination of cAMP levels in cell lysates. The cAMP-Screen™ assay utilizes the high-sensitivity chemiluminescent alkaline phosphatase (AP) substrate CSPD® (disodium 3-(4-methoxy Spiro {1,2-dioxetane-3,2'-(5'-chloro) tricyclo 3.3.1.1.^{3,7}} decan-4-yl phenyl phosphate) with Sapphire-II™ luminescence enhancer.

Unlike yeast-based-two-hybrid assays used to monitor protein/protein interactions in high-throughput assays, the present invention (1) is applicable to a variety of cells including mammalian cells, plant cells, protozoa cells such as *E. coli* and cells of invertebrate origin such as yeast, slime mold (*Dictyostelium*) and insects; (2) detects interactions at the membrane at the site of the receptor target or in the cytosol at the site of downstream target proteins rather than a limited cellular localization, i.e., nucleus; and (3) does not rely on indirect read-outs such as transcriptional activation. The present invention thus provides assays with greater physiological relevance and fewer false positives.

The present inventors have developed modifications to the embodiment disclosed in U.S. patent application serial no. 053,614 described above in order to enhance the sensitivity of the inventive GPCR assay. According to an

embodiment, the invention incorporates the use of serine/threonine clusters to enhance and prolong the interaction of GPCR with arrestin in order to make the detection more robust. The clusters can be utilized for orphan receptors or known GPCRs, which do not have this sequence motif. By adding this sequence to the C-terminal tail of the receptor, the activation of the receptor can be detected more readily by readouts of arrestin binding to GPCR, i.e., β -galactosidase complementation from fusion proteins of target proteins with β -galactosidase mutants.

According to another embodiment, the invention incorporates the use of arrestin point mutations to bypass the requirement of phosphorylation, by the action of specific GRK, on the C-terminal tail or intracellular loops of GPCR upon activation. The applications include i) wherein the cognate GRK for a particular GPCR or orphan receptor is unknown; and ii) wherein the specific GRK for the receptor of interest (or under test) may not be present or may have low activity in the host cell that is used for receptor activation assay.

According to another embodiment, the invention incorporates the use of a super arrestin to increase the binding efficiency of arrestin to an activated GPCR and to stabilize the GPCR/arrestin complex during GPCR desensitization. This application can be used to increase the robustness of ICAST/GPCR applications in cases where the GPCR is normally resensitized rapidly post desensitization.

Each of these methodologies is discussed below.

The invention will now be described in the following non-limiting examples.

EXAMPLE:

According to an embodiment of the invention, GPCR activation is measured through monitoring the binding of arrestin to ligand-activated GPCR. In this assay system, a GPCR, e.g., β -adrenergic receptor (β 2AR), and an arrestin, e.g., β -arrestin, are co-expressed in the same cell as fusion proteins with mutant forms of a reporter enzyme, e.g., β -galactosidase (β -gal). As illustrated in Figure 23, the β 2AR is expressed as a fusion protein with $\Delta\alpha$ form of β -gal mutant (β 2AR $\Delta\alpha$) and the β -arrestin as a fusion protein with the $\Delta\omega$ form of β -gal mutant (β -Arr $\Delta\omega$). The two fusion proteins, which at first exist in a resting (or un-

stimulated) cell in separate compartments, i.e., the membrane for GPCR and the cytosol for arrestin, cannot form an active β -galactosidase enzyme. When such a cell is treated with an agonist or a ligand, the ligand-occupied and activated receptor becomes a high affinity binding site for arrestin. The interaction between an activated GPCR, β 2AR $\Delta\alpha$, and arrestin, β -Arr $\Delta\omega$, drives the β -gal mutant complementation. The enzyme activity can be measured by using an enzyme substrate, which upon cleavage releases a product measurable by colorimetry, fluorescence, or chemiluminescence (e.g., the Gal-Screen™ assay system).

Experiment protocol-

1. In the first step, the expression vectors for β 2AR $\Delta\alpha$ and β Arr2 $\Delta\omega$ were engineered in selectable retroviral vectors pICAST ALC, as described in Figure 18 and pICAST OMC, as described in Figure 15.

2. In the second step, the two expression constructs were transduced into either C2C12 myoblast cells, or other mammalian cell lines, such as COS-7, CHO, A431, HEK 293, and CHW. Following selection with antibiotic drugs, stable clones expressing both fusion proteins at appropriate levels were selected.

5 3. In the last step, the cells expressing both $\beta 2AR\Delta\alpha$ and $\beta Arr2\Delta\omega$ were tested for response by agonist/ligand stimulated β -galactosidase activity. Triplicate samples of cells were plated at 10,000 cells in 100 microliter volume into a well of 96-well culture plate. Cells were cultured for 24 hours before assay. For agonist assay (Figures 3 and 4), cells were treated with variable concentrations of agonist, 10 for example, (-) isoproterenol, procaterol, dobutamine, terbutaline or L-L-phenylephrine for 60 min at 37° C. The induced β -galactosidase activity was measured by addition of Tropix Gal-Screen™ assay system substrate (Applied Biosystems) and luminescence measured in a Tropix TR717™ luminometer (Applied Biosystems). For antagonist assay (Figure 5), cells were pre-incubated for 15 10 min in fresh medium without serum in the presence of ICI-118,551 or propranolol followed by addition of 10 micro molar (-) isoproterenol.

Serine/Threonine Cluster Strategy

Background

20 Based on structure-function relationship studies on β -arrestins, a large region within the amino-terminal half of β -arrestins (termed the activation-recognition domain) recognizes the agonist-activated state of GPCRs. This region of β -arrestin also contains a small positively charged domain (approximately 20

amino acids with net charge +7) called the phosphorylation-recognition domain, which appears to interact with the GRK-phosphorylated carboxyl termini of GPCRs.

GPCRs can be divided into two classes based on their affinities for β -arrestins. Oakley et al., "Association of β -Arrestin with G Protein-Coupled Receptors During Clathrin-Mediated Endocytosis Dictates the Profile of Receptor Resensitization." *J. Biol. Chem.*, 274(45):32248-32257 (1999). The molecular determinants underlying this classification appear to reside in specific serine or threonine residues located in the carboxyl-terminal tail of the receptor. The receptor class that contains serine/threonine clusters (defined as serine or threonine residues occupying three consecutive or three out of four positions) in the carboxyl-termini binds β -arrestin with high affinity upon activation and phosphorylation and remains bound with β -arrestin even after receptor internalization, whereas the receptor class that contains only scattered serine and threonine residues in the carboxy-terminal tail binds β -arrestins with less affinity and disassociates from the β -arrestin upon internalization. Several known GPCRs, such as vasopressin V2 receptor (Oakley, et al.), neurotensin receptor 1 and angiotensin II receptor type 1A (Zhang, et al., "Cellular Trafficking of G Protein-Coupled Receptor/ β -Arrestin Endocytic Complexes." *J. Biol. Chem.*, 274(16):10999-11006 (1999)), which possess one or more of such serine/threonine clusters in their carboxyl-termini, were shown to bind β -arrestins with high affinity.

EXAMPLE

According to an embodiment of the invention, a serine/threonine cluster strategy is used to facilitate screening assays for orphan receptors that do not possess this structural motif of their own. The orphan receptors are easily classified by sequence alignment. Orphan receptors lacking the serine/threonine clusters are each cloned into an expression vector that is modified to introduce one or more serine/threonine cluster(s) to the carboxyl-terminal tail of the receptor (FIGURE 24). The serine/threonine clusters enhance the receptor activation dependent interaction between the activated and phosphorylated receptor (negative charges) and β -arrestin (positive charges in the phosphorylation-recognition domain) through strong ionic interactions, thus prolonging interaction between the receptor and arrestin. The modification of the orphan receptor tail thus makes detection of receptor activation more robust.

Experiment protocol -

1. In a first step, the open-reading-frame (ORF) of an orphan receptor, which lacks the serine/threonine clusters, is cloned into a modified expression vector such as pICAST ALC described in Figure 10A. The modified pICAST ALC includes coding sequences for one or more sets of serine/threonine clusters (for example, SSS or SST) located downstream from the insert of the ORF of an orphan receptor (FIGURE 24).

2. In a second step, chimeric orphan receptor, $ORF_{\text{orphan R}}-(SSS)_n-\Delta\alpha$, is co-

expressed in a mammalian cell with a β -arrestin chimera, such as β Arr2 $\Delta\omega$ described in Figure 15.

3. In a third step, the cell is treated with an agonist or a ligand and the activated receptor with phosphorylated serine cluster(s) binds the β -arrestin with high affinity producing strong signals in readouts of β -gal complementation.

This assay, which provides a means for sensitive measurement of functional activation of the orphan receptors, can be used to screen for natural or surrogate ligands for orphan receptors, a process called de-orphaning or target discovery for new GPCRs (FIGURE 28). Furthermore, this assay is also useful in screening for potential agonists and antagonists for lead discovery of GPCRs.

Enhanced Binding of Arrestin in the Presence and in the Absence of GPCR

Phosphorylation

Background

Six different classes of G-protein coupled receptor kinases (GRKs) have been identified and each of these has been reported to be expressed as multiple splice variants. Krupnick et al., "The role of receptor kinases and arrestins in G protein-coupled receptor regulation." *Ann. Rev. Pharmacol. Toxicol.*, 38:289-319 (1998). Although many cell lines express a variety of GRKs, the specific GRK required for phosphorylation of a given GPCR may not always be present in the cell line used for recombinant GPCR and arrestin expression. This is particularly an issue for applications using orphan receptors, in which case the cognate GRK will likely be unknown. In other cases, the cell line used for recombinant

expression work may have the required GRK, but may express the GRK at low levels. In order to bypass such caveats, genetically modified arrestins that bind specifically to activated GPCRs, but without the requirement of GRK phosphorylation are employed.

5 Mutagenesis studies on arrestins demonstrate that point mutations in the phosphorylation-recognition domain, particularly mutations converting Arg175 (of visual arrestin) to an oppositely charged residue such as glutamate (R175E mutation), result in an arrestin which specifically binds to activated GPCRs, but does so without the requirement for phosphorylation.

10 Numerous observations have led to the hypothesis that arrestin exists in an inactive state that has a low affinity for GPCRs. Once a GPCR is both activated and phosphorylated, the phosphorylated region of the GPCR C-terminus interacts with the phosphorylation-recognition domain of arrestin causing the arrestin to change conformations allowing the activation-recognition region to be exposed for
15 binding to the activated/ phosphorylated receptor. Vishnivetskiy et al., "How does arrestin respond to the phosphorylated state of rhodopsin?" J. Biol. Chem., 274(17):11451-11454 (1999); Gurevich et al., "Arrestin interactions with G protein-coupled receptors. Direct binding studies of wild-type and mutant arrestins with rhodopsin, beta 2-adrenergic and m2 muscarinic cholinergic receptors." J.
20 Biol. Chem., 270(2):720-731, (1995); Gurevich et al., "Mechanism of phosphorylation-recognition by visual arrestin and the transition of arrestin into a high affinity binding site." Mol. Pharmacol., 51(1):161-169 (1997); Kovoor et al., "Targeted construction of phosphorylation-independent beta-arrestin mutants with

constitutive activity in cells.” J. Biol. Chem., 274(11):6831-6834 (1999). In summary, binding studies of single mutation, double mutation, deletion, and chimerical arrestins with inactive, inactive and phosphorylated, activated but not phosphorylated, or activated and phosphorylated visual or non-visual GPCRs all support this model.

EXAMPLE

A phosphorylation insensitive mutant of arrestin fused to mutant reporter protein can be produced that will bind to activated GPCRs in a phosphorylation independent manner. As proof of concept, a point mutation for β -arrestin2, R170E β -arrestin2, has been produced and its interaction with β 2AR has been analyzed in accordance with the invention.

Experimental protocol:

- 1) In the first step, β -arrestin2 was mutated such that Arg170 was converted to Glu. This mutation is equivalent to the R175E mutation of visual arrestin. The mutant β -arrestin2 open reading frame was cloned in frame with $\Delta\omega$ - β -galactosidase in the pICAST OMC expression vector to produce a modified expression vector R170E β -arrestin2 (FIGURE 25).
- 2) In the second step, the R170E β -arrestin2 expression construct was transduced into a C2C12 myoblast cell line that had been engineered to express β 2AR as a fusion to $\Delta\alpha$ - β -galactosidase as described in Figure 18 of U.S. Application Serial No. 09/654,499. Following selection with antibiotic drugs, a

population of clones expressing both fusion proteins was obtained.

- 3) In the last step, this population of cells expressing both R170E β -arrestin2 $\Delta\omega$ and β 2AR $\Delta\alpha$ were tested for response by agonist/ligand stimulated β -galactosidase activity as demonstrated in FIGURE 26. The C2C12 clone 43-8 co-
5 expressing β 2AR $\Delta\alpha$ and wild-type β -arrestin2 $\Delta\omega$ (FIGURE 26) was used as reference control. Triplicate samples of cells were plated at 10,000 cells in 100 microliter volume into wells of a 96-well culture plate. Cells were cultured for 24 hours before assay. For agonist assay as in FIGURE 26, cells were treated with 10 μ m (-)isoproterenol stabilized with 0.3mM ascorbic acid 37° C for 0, 5, 10, 15,
10 30, 45 or 60 minutes. The induced β -galactosidase activity was measured by addition of Tropix Gal-Screen™ assay system substrate (Applied Biosystems) and luminescence measured in a Tropix TR717™ luminometer (Applied Biosystems). As shown in Figure 26, the mutant arrestin interacts with β 2AR in an agonist-dependent manner and was comparable with that of wild-type arrestin.
- 15 4) To expand the application of phosphorylation-insensitive arrestin, cell lines such as C2C12, CHO or HEK 293, are developed that express the R170E β -arrestin2 $\Delta\omega$ construction. These cell lines can be used to transduce orphan or known GPCRs as fusions with $\Delta\alpha$ - β -galactosidase in order to develop cell lines for agonist and antagonist screening and

Development of Super Arrestins:

Background

Attenuation of GPCR signaling by the arrestin pathway serves to ensure that a cell or organism does not over-react to a stimulus. At the same time, the arrestin pathway often serves to recycle the GPCR such that it can be temporarily
5 inactivated but then quickly resensitized to allow for sensitivity to new stimuli. The down-regulation process involves phosphorylation of the receptor, binding to arrestin and endocytosis. Following endocytosis of the desensitized receptor, the receptor is either degraded in lysosomes or resensitized and sent back to the
10 membrane. Resensitization involves release of arrestin from the receptor, dephosphorylation and cycling back to the membrane. The actual route a GPCR follows upon activation depends on its biological function and the needs of the organism. Because of these diverse pathways that may be required of the down-regulation pathway, arrestin affinities for activated GPCRs vary from receptor to
15 receptor. It would thus be very advantageous to engineer super arrestins that have a higher affinity and avidity for activated GPCRs than what nature has provided.

Although mutational, deletion and chimerical studies of arrestins have focused on understanding regulatory switches in the molecule that respond to GPCR phosphorylation states, several of these altered recombinant forms of
20 arrestin have resulted in molecules with enhanced binding to activated, phosphorylated GPCRs. Conversion of Arg175 to histidine, tyrosine, phenylalanine or threonine results in significantly higher amounts of binding to phosphorylated, activated rhodopsin than wild-type arrestin or R175E arrestin,

although these mutations result in less binding to activated, non-phosphorylated receptor. Gurevich et al. (1997). In addition, conversion of Valine 170 to alanine increased the constitutive affect of the R175E mutation, but also nearly doubled the amount of interaction of wild-type arrestin with activated, phosphorylated
5 rhodopsin. Gurevich et al. (1997).

Truncation of β -arrestin1 at amino acid 382 has been reported to enhance binding of both R169E (equivalent to arrestin R175E) and wild-type β -arrestin1 to activated or activated and phosphorylated receptor, respectively. Kovoor et al. Chimerical arrestins in which functional regions of visual arrestin were swapped
10 with those of β -arrestin1 have been reported to be altered in binding affinity to activated, phosphorylated GPCRs. Gurevich et al. (1995). Several of these chimeras, such as β -arrestin1 containing the visual arrestin extreme N-terminus, show increased specific binding to phosphorylated activated GPCRs compared to wild-type β -arrestin1 (Gurevich et al. (1995)). Modifications that enhance arrestin
15 affinity for the activated GPCR such as described above, whether phosphorylated or non-phosphorylated, could also enhance signal to noise of β -galactosidase activity since the arrestin/GPCR complex is stabilized and/or more long-lived. The use of mutant arrestins with higher activated-GPCR affinity would improve the inventive technology for GPCR targets, without compromising receptor/ligand
20 biology.

In addition, this “super arrestin” approach can be combined with the use of arrestin point mutations to provide a stronger signal to noise with or without GRK requirements.

EXAMPLE

An arrestin mutant fused to mutant reporter protein can be produced to enhance binding of the arrestin to an activated GPCR to enhance sensitivity of detection.

5 Experiment protocol -

- 1) In the first step, mutant β -arrestin2 constructions will be generated which include R170E/T/Y/or H, V165A, substitution of a.a. 1-43 with a.a. 1-47 of visual arrestin, or deletion of the C-terminal and combinations of these alterations. The mutant β -arrestin2 open reading frames will be cloned in frame with $\Delta\omega$ - β -galactosidase in the pICAST OMC expression vector similar to cloning of the
10 R170E β -arrestin2 mutation shown in FIGURE 25.
- 2) In the second step, mutant expression constructs will be transduced into a C2C12 myoblast cell line that has been engineered to express β 2AR as a fusion to $\Delta\alpha$ - β -galactosidase. Following selection with antibiotic drugs, a population of
15 clones expressing both fusion proteins will be obtained. Wild type and R170E β -arrestin2 constructions will be transduced to generate control, reference clonal populations.
- 3) In the third step, populations of cells expressing both β -arrestin2 $\Delta\omega$ (mutant or wild type) and β 2AR $\Delta\alpha$ will be tested for response by agonist/ligand stimulated
20 β -galactosidase activity.
- 4) In the next step, mutant (super) β -arrestin2 $\Delta\omega$ constructions that show a significantly higher signal to noise ratio in the agonist assay compared with wild-type β -arrestin2 $\Delta\omega$ will be chosen. These constructions will be used to develop

stable cell lines expressing the “super” β -arrestin2 $\Delta\omega$ that can be used for transducing in known or orphan GPCRs. Use of a super β -arrestin2 $\Delta\omega$ could increase the signal to noise of ICAST/GPCR applications allowing improved screening capabilities for lead and ligand discovery.

5 Super Arrestin is used to increase the binding efficiency of arrestin to an activated GPCR and to stabilize the GPCR/arrestin complex during GPCR desensitization. This application can be used to increase the robustness of ICAST/GPCR applications in cases where the GPCR is normally resensitized rapidly post desensitization.

10 The assays of this invention, and their application and preparation have been described both generically, and by specific example. The examples are not intended as limiting. Other substituent identities, characteristics and assays will occur to those of ordinary skill in the art, without the exercise of inventive faculty. Such modifications remain within the scope of the invention, unless excluded by
15 the express recitation of the claims advanced below.

WHAT IS CLAIMED IS:

1. A method of assessing the effect of a test condition on G-protein-coupled receptor (GPCR) pathway activity, comprising:

a) providing a cell that expresses a GPCR as a fusion protein to one mutant
5 form of reporter enzyme and an interacting protein partner as a fusion to another mutant form of enzyme,

wherein said cell also expresses an arrestin, wherein said arrestin is modified to enhance binding of said arrestin to said GPCR, wherein said enhanced binding between said arrestin and said GPCR increases sensitivity of detection of
10 said effect of said test condition;

b) exposing the cell to a ligand for said GPCR under said test condition; and

c) monitoring activation of said GPCR by complementation of said reporter enzyme;

wherein increased reporter enzyme activity in the cell compared to that
15 which occurs in the absence of said test condition indicates increased GPCR interaction with its interacting protein partner compared to that which occurs in the absence of said test condition, and decreased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates
20 decreased GPCR interaction with its interacting protein partner compared to that which occurs in the absence of said test condition.

2. A method of assessing the effect of a test condition on G-protein-coupled receptor (GPCR) pathway activity, comprising:

a) providing a cell that expresses a GPCR as a fusion protein to one mutant

form of reporter enzyme and an interacting protein partner as a fusion to another mutant form of enzyme;

wherein said GPCR fusion protein is modified to include one or more sets of serine/threonine clusters, wherein said one or more sets of serine/threonine clusters enhance binding of said GPCR to arrestin, wherein said enhanced binding between said GPCR and said arrestin increases sensitivity of detection of said effect of said test condition;

b) exposing the cell to a ligand for said GPCR under said test condition; and
c) monitoring activation of said GPCR by complementation of said reporter enzyme;

wherein increased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates increased GPCR interaction with said interacting protein partner compared to that which occurs in the absence of said test condition, and decreased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates decreased GPCR interaction with interacting protein partner compared to that which occurs in the absence of said test condition.

3. A DNA molecule comprising a sequence encoding a biologically active hybrid GPCR, wherein said hybrid GPCR comprises a GPCR as a fusion protein to one mutant form of reporter enzyme and wherein said hybrid GPCR is modified to include one or more sets of serine/threonine clusters, wherein said one or more sets of serine/threonine clusters enhance binding of said hybrid GPCR to arrestin.

4. A DNA construct capable of directing the expression of a biologically

active hybrid GPCR in a cell, comprising the following operatively linked elements:

a promoter; and

a DNA molecule comprising a sequence encoding a biologically active hybrid GPCR, wherein said hybrid GPCR comprises a GPCR as a fusion protein to one mutant form of reporter enzyme and wherein said hybrid GPCR is modified to include one or more sets of serine/threonine clusters, wherein said one or more sets of serine/threonine clusters enhance binding of said hybrid GPCR to arrestin.

5. A cell transformed with a DNA construct capable of expressing a biologically active hybrid GPCR in a cell, comprising the following operatively linked elements:

a promoter; and

a DNA molecule comprising a sequence encoding a biologically active hybrid GPCR, wherein said hybrid GPCR comprises a GPCR as a fusion protein to one mutant form of reporter enzyme and wherein said hybrid GPCR is modified to include one or more sets of serine/threonine clusters, wherein said one or more sets of serine/threonine clusters enhance binding of said hybrid GPCR to arrestin.

6. A DNA molecule comprising a sequence encoding a biologically active hybrid arrestin, wherein said hybrid arrestin comprises an arrestin as a fusion to one mutant form of reporter enzyme and wherein said hybrid arrestin is modified to enhance binding of said arrestin to GPCR.

7. A DNA construct capable of directing the expression of a biologically active hybrid arrestin in a cell, comprising the following operatively linked

elements:

a promoter; and

a DNA molecule comprising a sequence encoding a biologically active hybrid arrestin, wherein said hybrid arrestin comprises an arrestin as a fusion to one mutant form of reporter enzyme and wherein said hybrid arrestin is modified to enhance binding of said arrestin to GPCR.

8. A cell transformed with a DNA construct capable of expressing a biologically active hybrid arrestin in a cell, comprising the following operatively linked elements:

10 a promoter; and

a DNA molecule comprising a sequence encoding a biologically active hybrid arrestin, wherein said hybrid arrestin comprises an arrestin as a fusion to one mutant form of reporter enzyme and wherein said hybrid arrestin is modified to enhance binding of said arrestin to GPCR.

15 9. A method of assessing the effect of a test condition on G-protein-coupled receptor (GPCR) pathway activity, comprising:

a) providing a cell that expresses a GPCR as a fusion protein to one mutant form of reporter enzyme and an interacting protein partner as a fusion to another mutant form of enzyme,

20 wherein said cell also expresses an arrestin, wherein said arrestin is modified by introducing a point mutation in a phosphorylation-recognition domain to remove a requirement for phosphorylation of said GPCR for arrestin binding to permit binding of said arrestin to said GPCR in said cell regardless of whether said

GPCR is phosphorylated,

b) exposing the cell to a ligand for said GPCR under said test condition; and

c) monitoring activation of said GPCR by complementation of said reporter enzyme;

5 wherein increased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates increased GPCR interaction with its interacting protein partner compared to that which occurs in the absence of said test condition, and decreased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates
10 decreased GPCR interaction with its interacting protein partner compared to that which occurs in the absence of said test condition.

10. The method of Claim 9, wherein said arrestin is mutated to increase a property selected from affinity and avidity for activated, non-phosphorylated GPCR.

15 11. The method of Claim 10, wherein said arrestin is β -arrestin2 and wherein said β -arrestin2 is mutated to convert Arg169 to an oppositely charged residue.

12. The method of Claim 11, wherein said oppositely charged residue is selected from the group consisting of histidine, tyrosine, phenylalanine and
20 threonine.

13. The method of Claim 9, wherein said arrestin is mutated to increase a property selected from affinity and avidity for activated and phosphorylated GPCR.

14. A method of assessing the effect of a test condition on G-protein-

coupled receptor (GPCR) pathway activity, comprising:

a) providing a cell that expresses a GPCR as a fusion protein to one mutant form of reporter enzyme and an interacting protein partner as a fusion to another mutant form of enzyme;

5 wherein said GPCR fusion protein is modified to include one or more sets of serine/threonine clusters, said one or more serine/threonine clusters defined as serine or threonine residues occupying three consecutive or three out of four positions in a carboxyl-termini of said GPCR, wherein said one or more sets of serine/threonine clusters enhance binding of said GPCR to arrestin, wherein said
10 enhanced binding between said GPCR and said arrestin increases sensitivity of detection of said effect of said test condition;

b) exposing the cell to a ligand for said GPCR under said test condition; and

c) monitoring activation of said GPCR by complementation of said reporter enzyme;

15 wherein increased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates increased GPCR interaction with said interacting protein partner compared to that which occurs in the absence of said test condition, and decreased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates
20 decreased GPCR interaction with interacting protein partner compared to that which occurs in the absence of said test condition.

15. The method of Claim 1, wherein said modified arrestin exhibits enhanced binding to activated, phosphorylated GPCR.

25. The method of Claim 14, wherein said modified arrestin comprises conversion of Arg170 to an amino acid selected from the group consisting of histidine, tyrosine, phenylalanine and threonine.

Cellular Expression of β_2 AR- β gal $\Delta\alpha$ Fusion Protein in C2 Clones
 (measured by anti- β -gal ELISA)

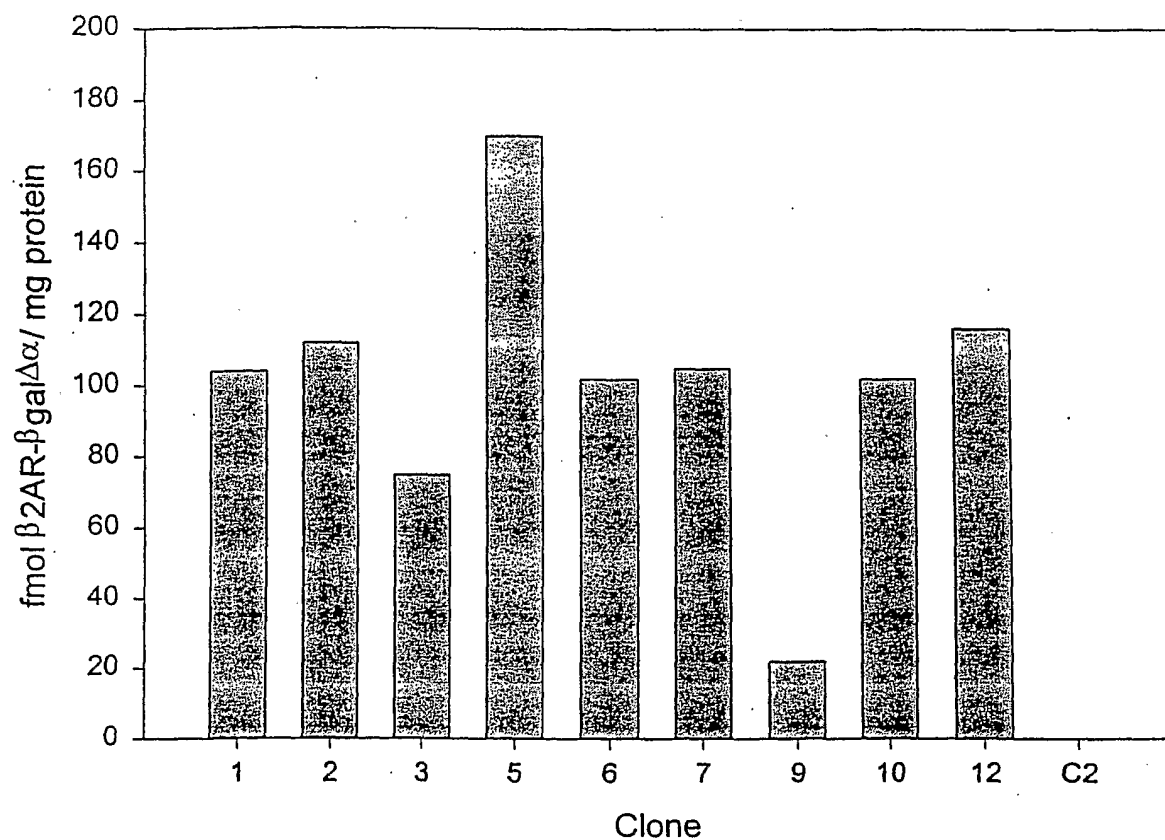


FIGURE 1A

Cellular expression of β Arr2- β gal $\Delta\omega$ fusion protein in C2 clones
(measured by anti- β gal ELISA)

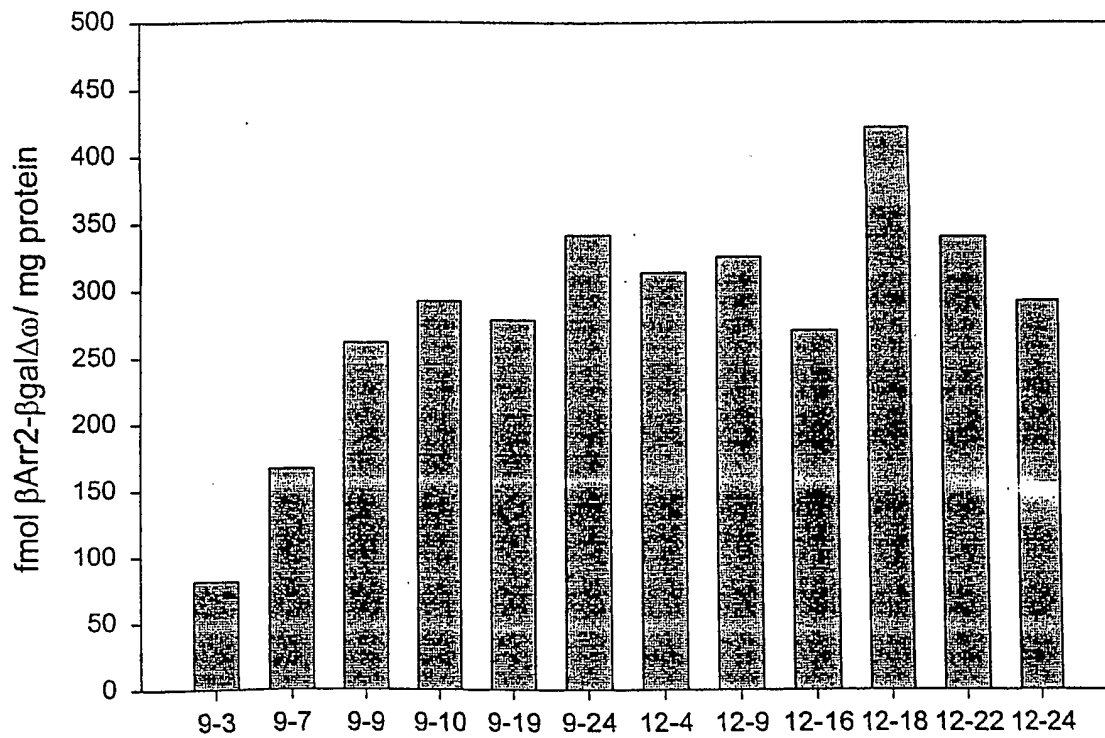


FIGURE 1B

Agonist Stimulated cAMP Response in C2 Cells Expressing β 2AR- β gal $\Delta\alpha$

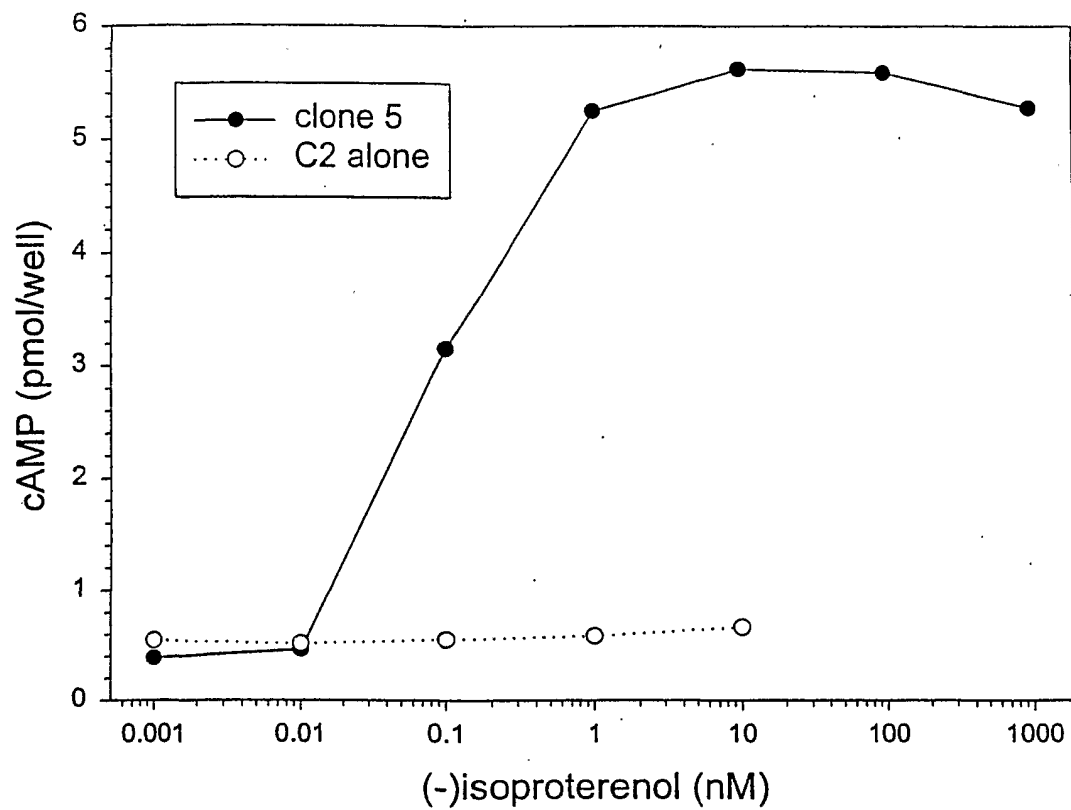


FIGURE 2

β -galactosidase Complementation as a Measurement for β 2AR- β gal $\Delta\alpha$ interacting with β Arrestin2- β gal $\Delta\omega$ upon agonist Stimulation

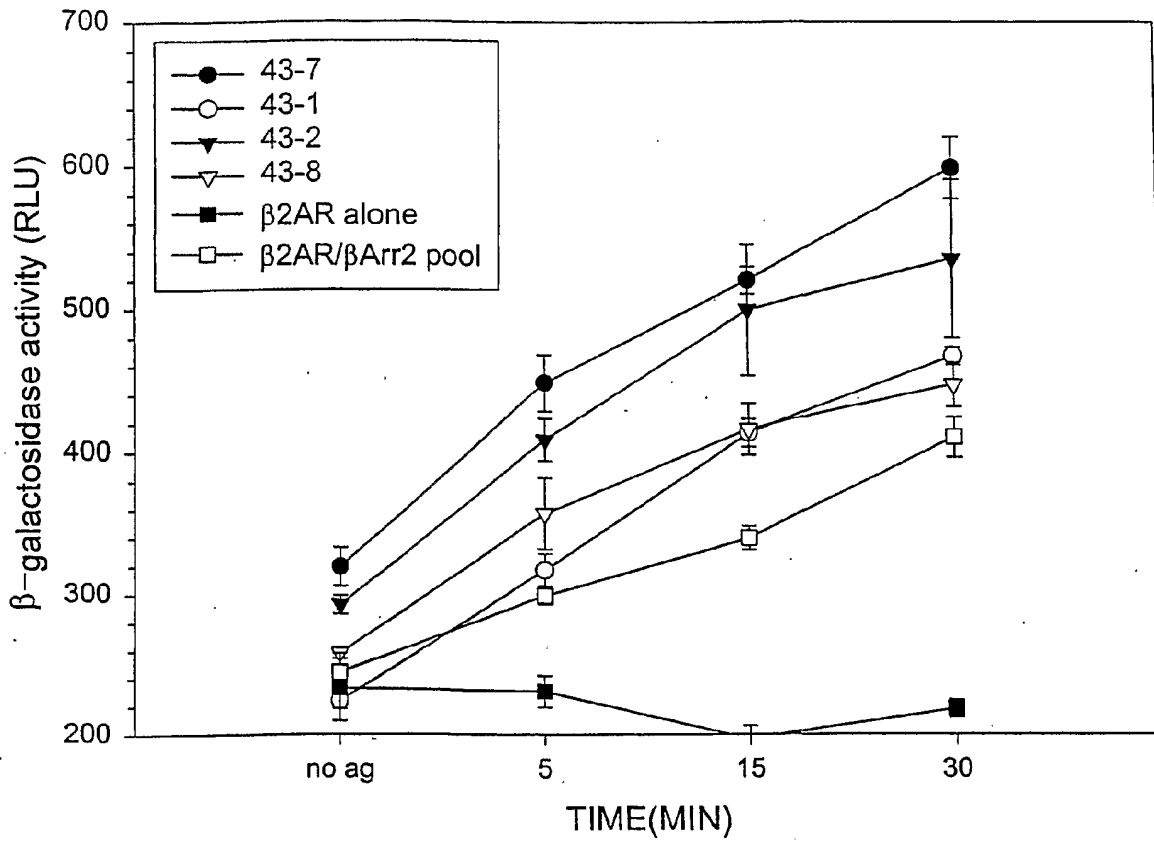


FIGURE 3A

β -galactosidase Complementation as a Measurement for β 2AR- β gal $\Delta\alpha$ Interaction with β Arrestin1- β gal $\Delta\omega$ upon Agonist Stimulation

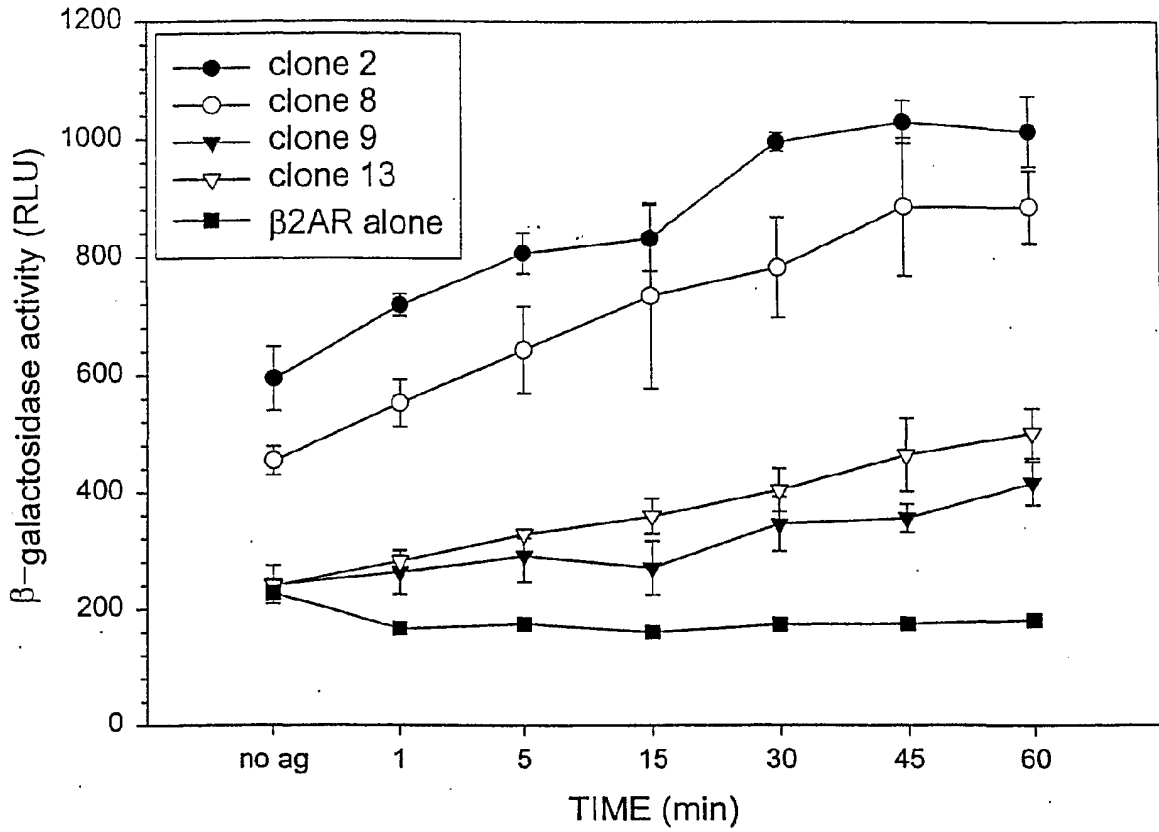


FIGURE 3B

β -galactosidase Activity in Response to Agonist in C2 Cells
Coexpressing β 2AR- β gal $\Delta\alpha$ and β Arrestin2- β gal $\Delta\omega$ Fusion Proteins

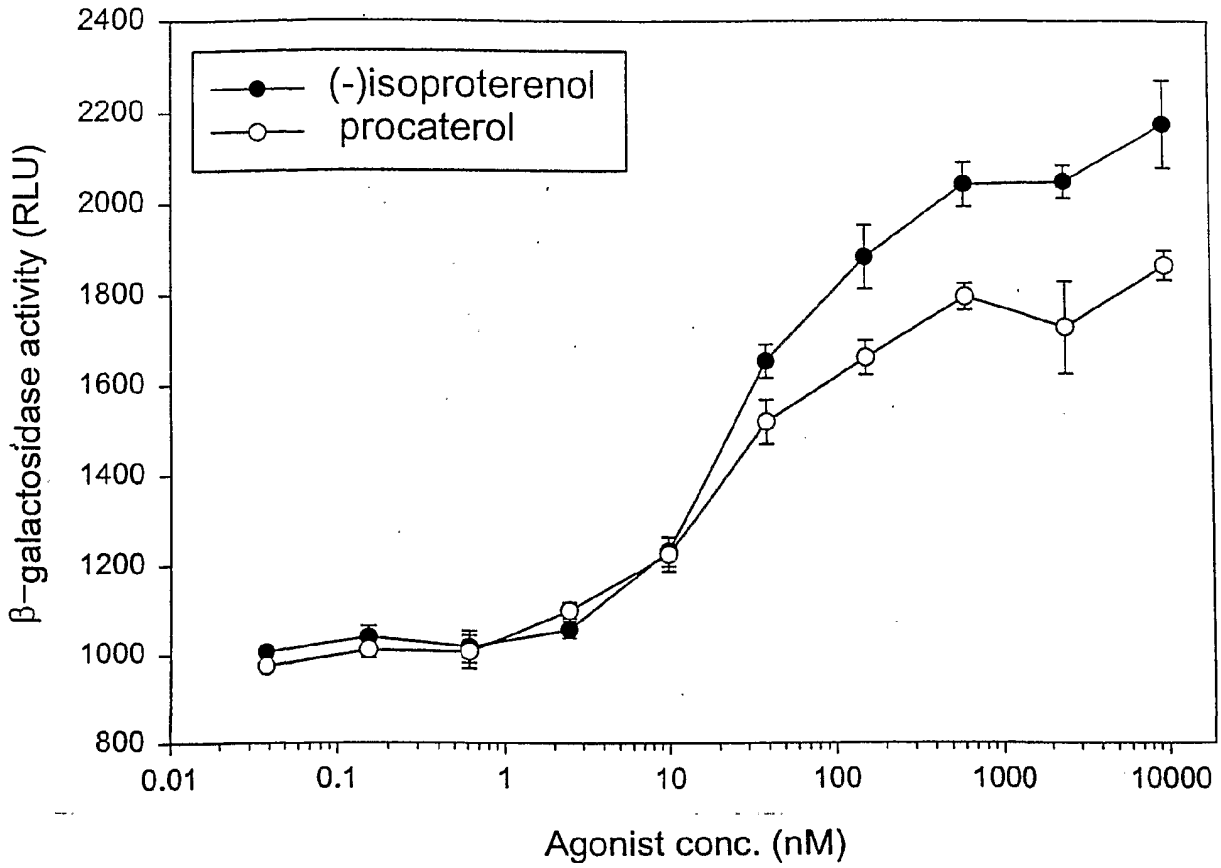


FIGURE 4A

β -galactosidase Activity in Response to Agonist in C2 Cells
Coexpressing β 2AR- β gal $\Delta\alpha$ and β Arrestin1- β gal $\Delta\omega$ Fusion Proteins

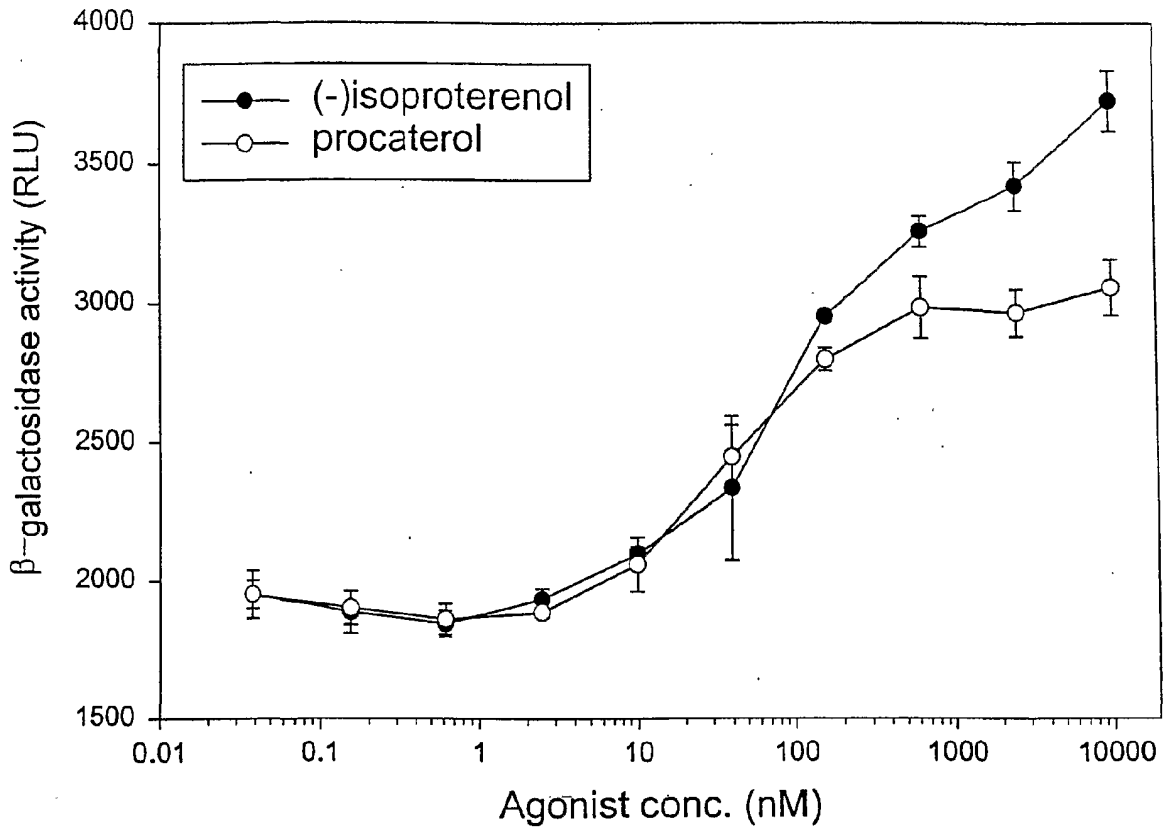


FIGURE 4B

Inhibition of β -galactosidase activity in C2 Cells Coexpressing β 2AR- β gal $\Delta\alpha$ and β Arrestin2- β gal $\Delta\omega$ Fusion Proteins

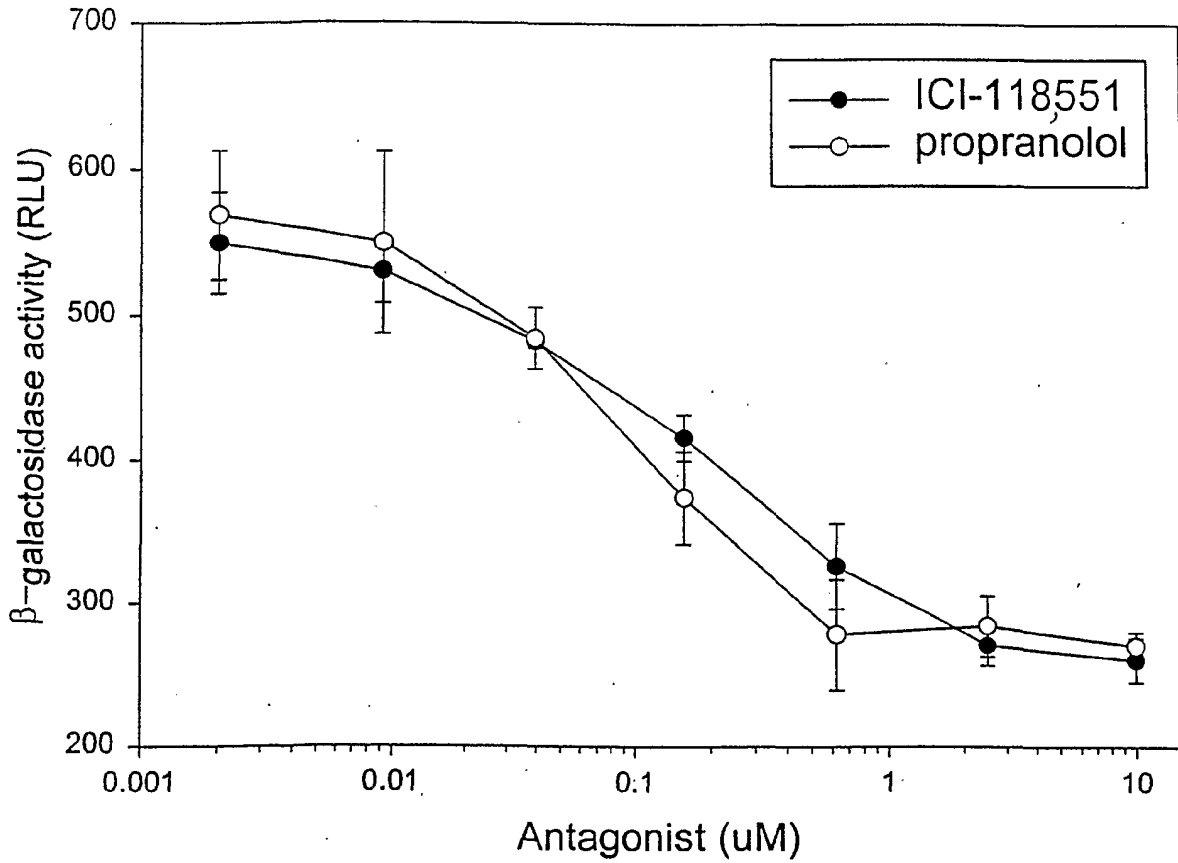


FIGURE 5A

Antagonist Inhibition of β -galactosidase Activity in C2 Cells
 Coexpressing β 2AR- β gal $\Delta\alpha$ and β Arrestin1- β gal $\Delta\omega$ Fusion Proteins

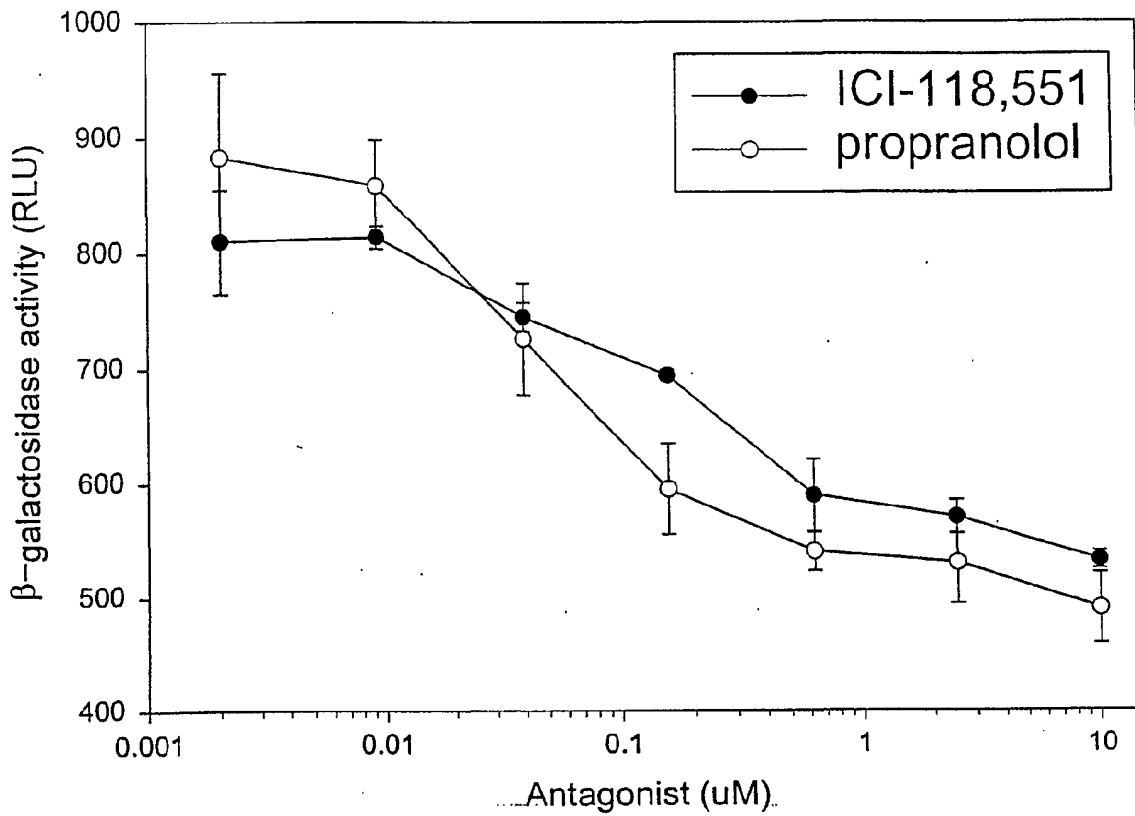


Figure 5B

Agonist Stimulated cAMP Response in Clones or Pools of C2 Cells Coexpressing A2aR- β gal $\Delta\alpha$ and β Arrestin1- β gal $\Delta\omega$ Fusion Proteins

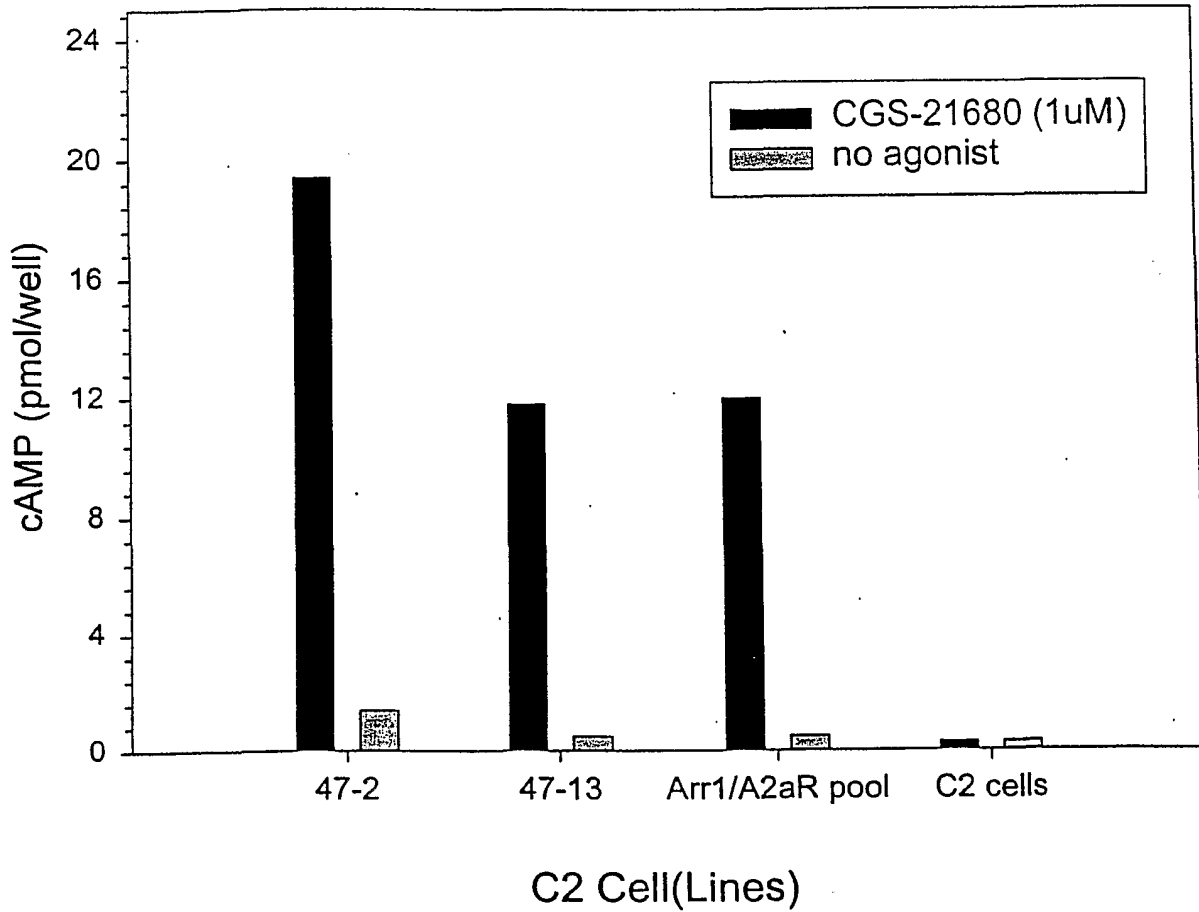


FIGURE 6

Agonist Stimulated cAMP Response in Clones or Pools of C2 Cells Expressing D1-βgalΔα and βArrestin2-βgalΔω Fusion Proteins

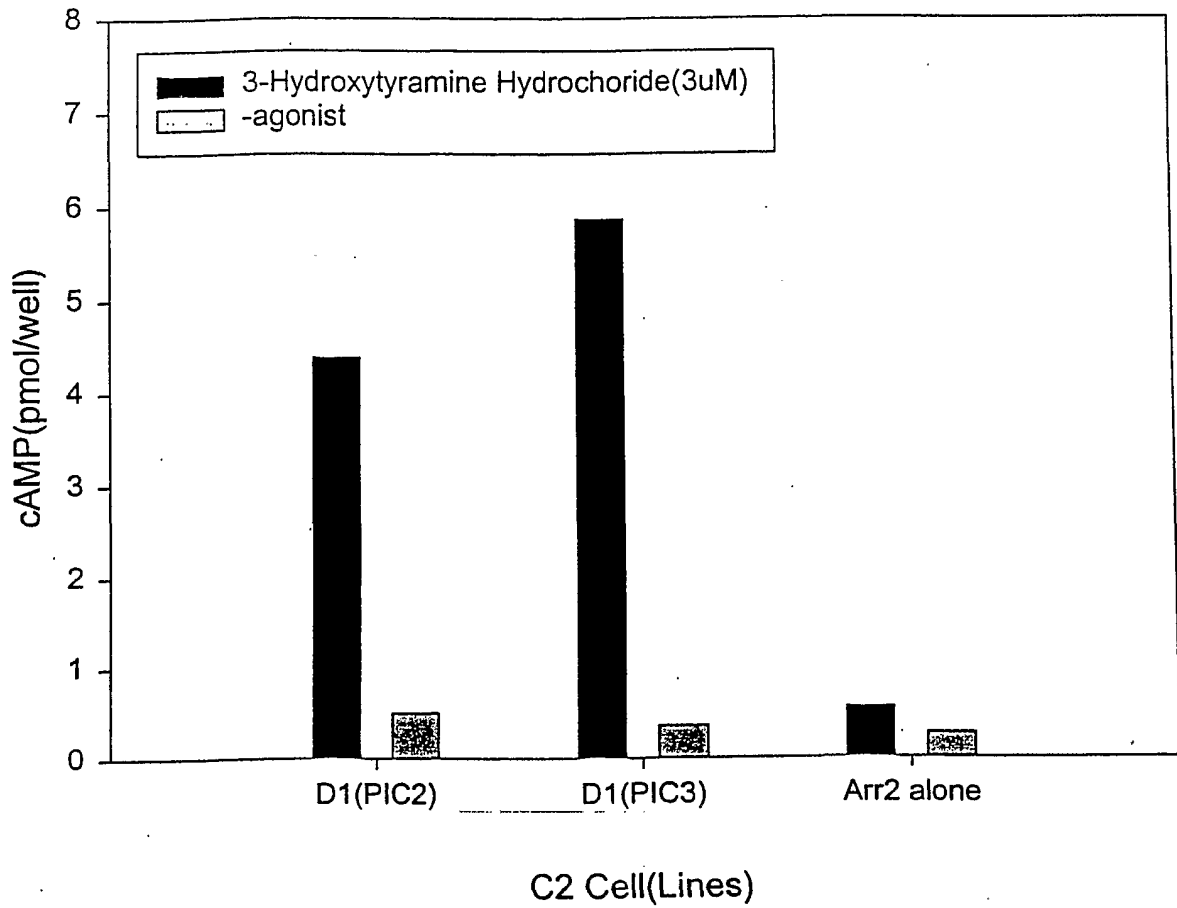


FIGURE 7

β_2 AR- β gal $\Delta\omega$ and β arr2- β gal $\Delta\alpha$ Interaction in HEK293 Clones in Response to Isoproterenol Treatment (1 μ M)

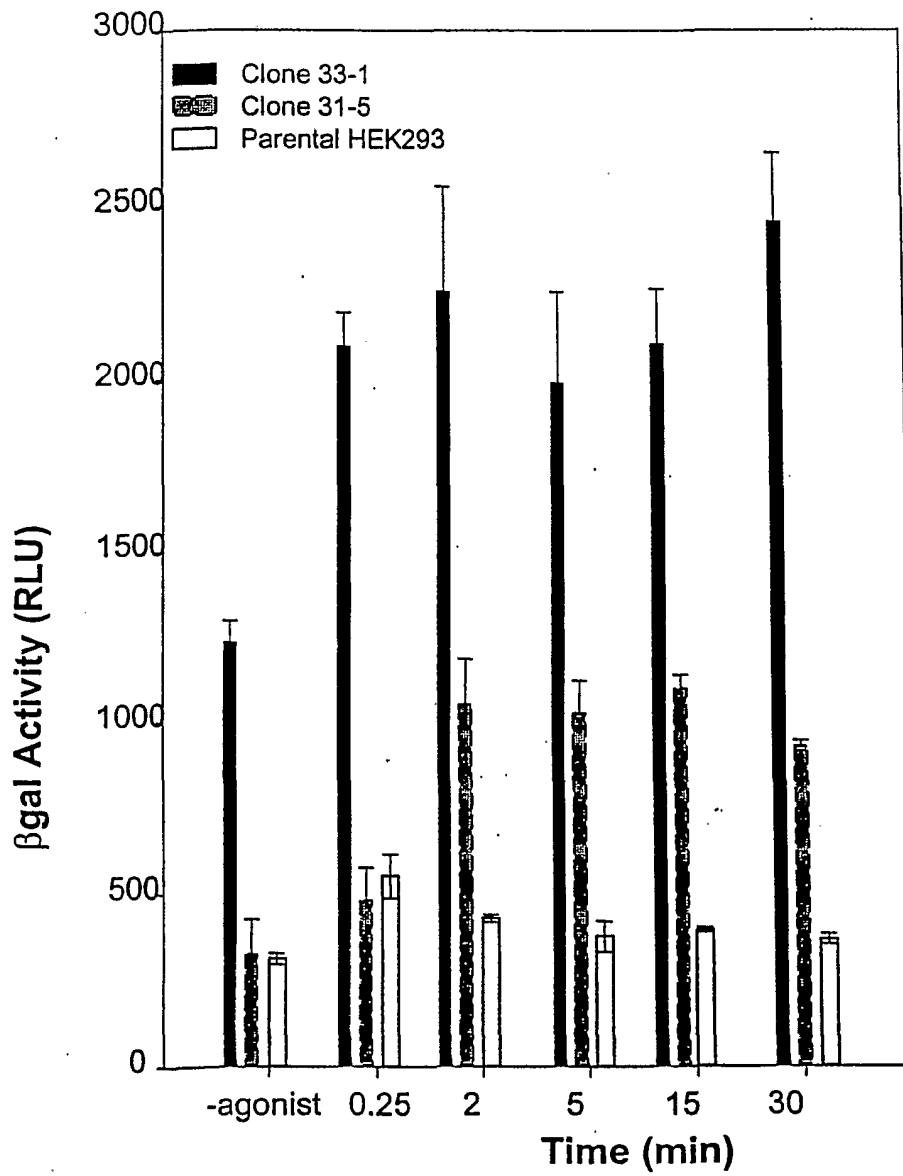


FIGURE 8A

β 2AR- β gal $\Delta\alpha$ and β Arr1- β gal $\Delta\omega$ Interaction in a CHO Pool in Response to Isoproterenol Treatment(10uM)

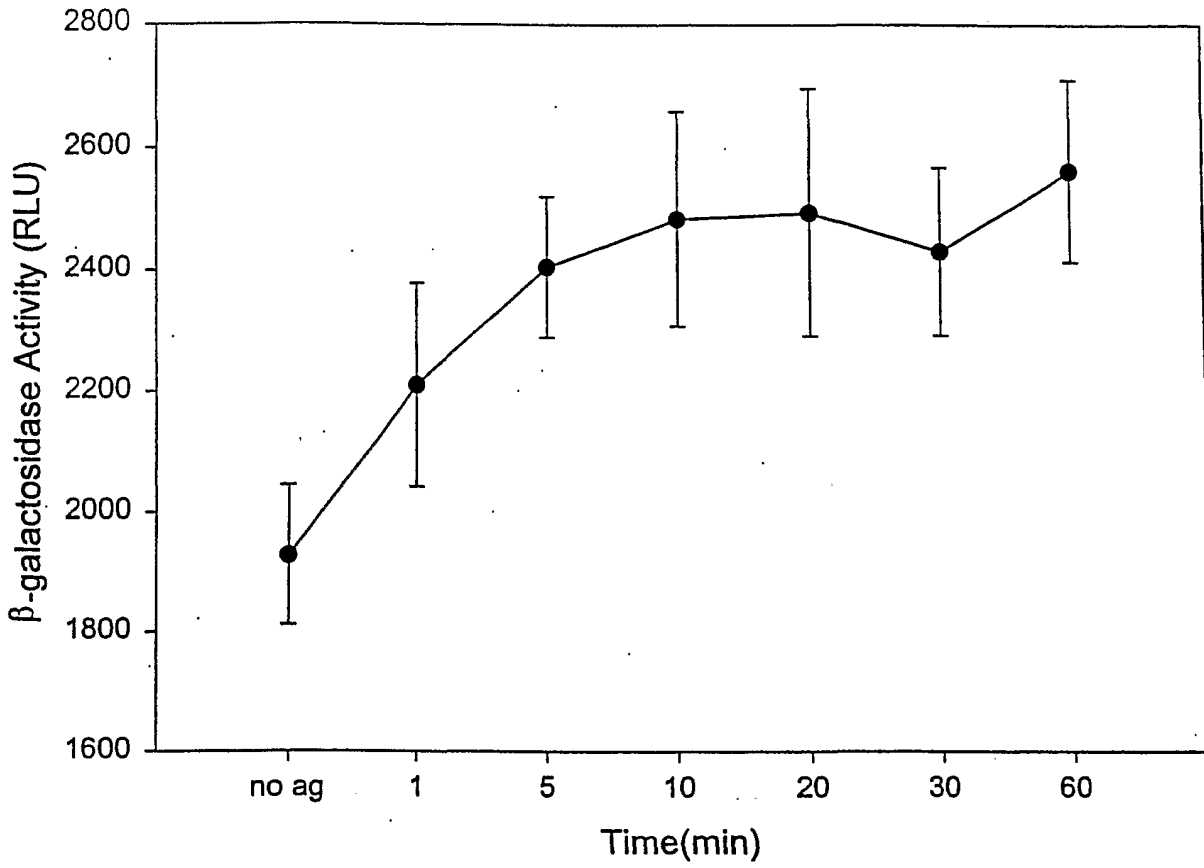


FIGURE 8B

β 2AR- β gal $\Delta\alpha$ and β Arr2- β gal $\Delta\omega$ Interaction in CHW Clone in Response to Isoproterenol Treatment (10 μ M)

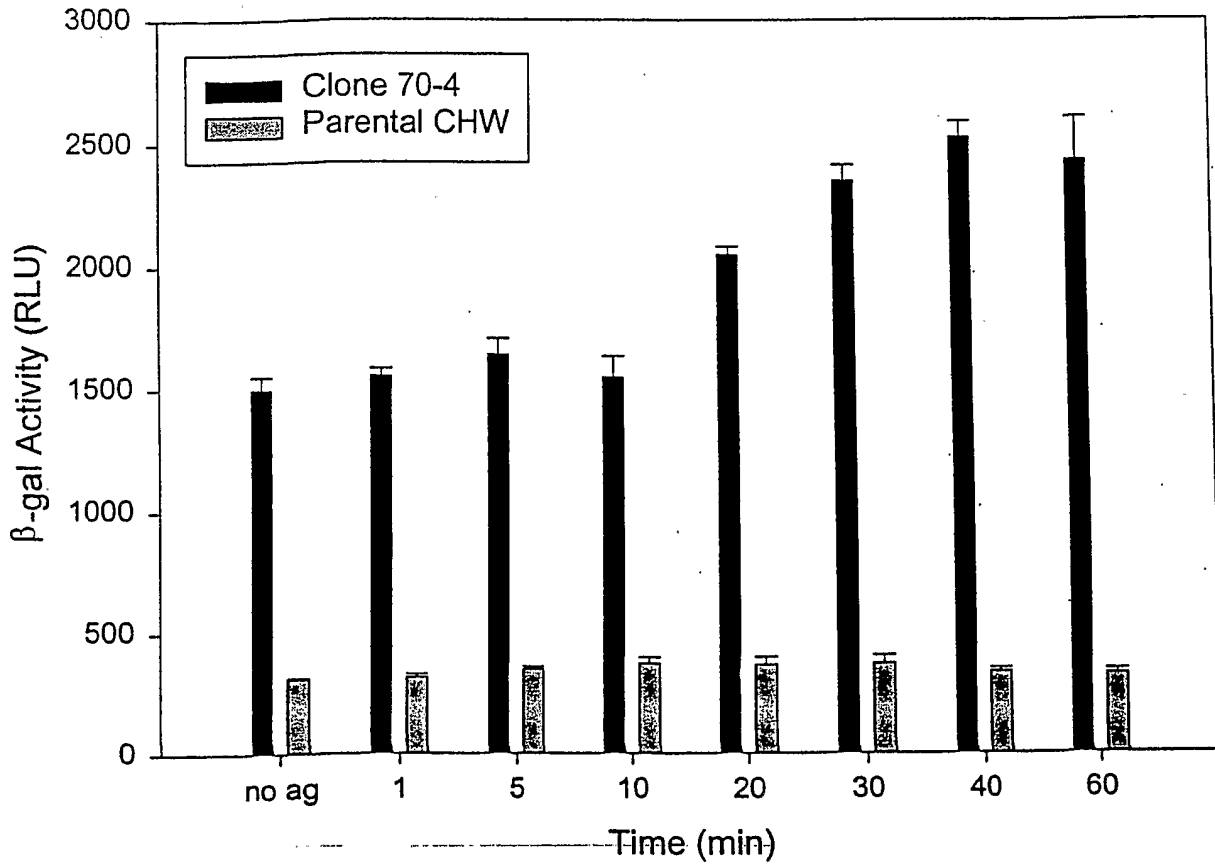


FIGURE 8C

β -galactosidase Complementation as a Measurement for Adrenergic Receptor Homodimerization in HEK 293 Cells Coexpressing β 2AR- β gal $\Delta\alpha$ and β 2AR- β gal $\Delta\omega$.

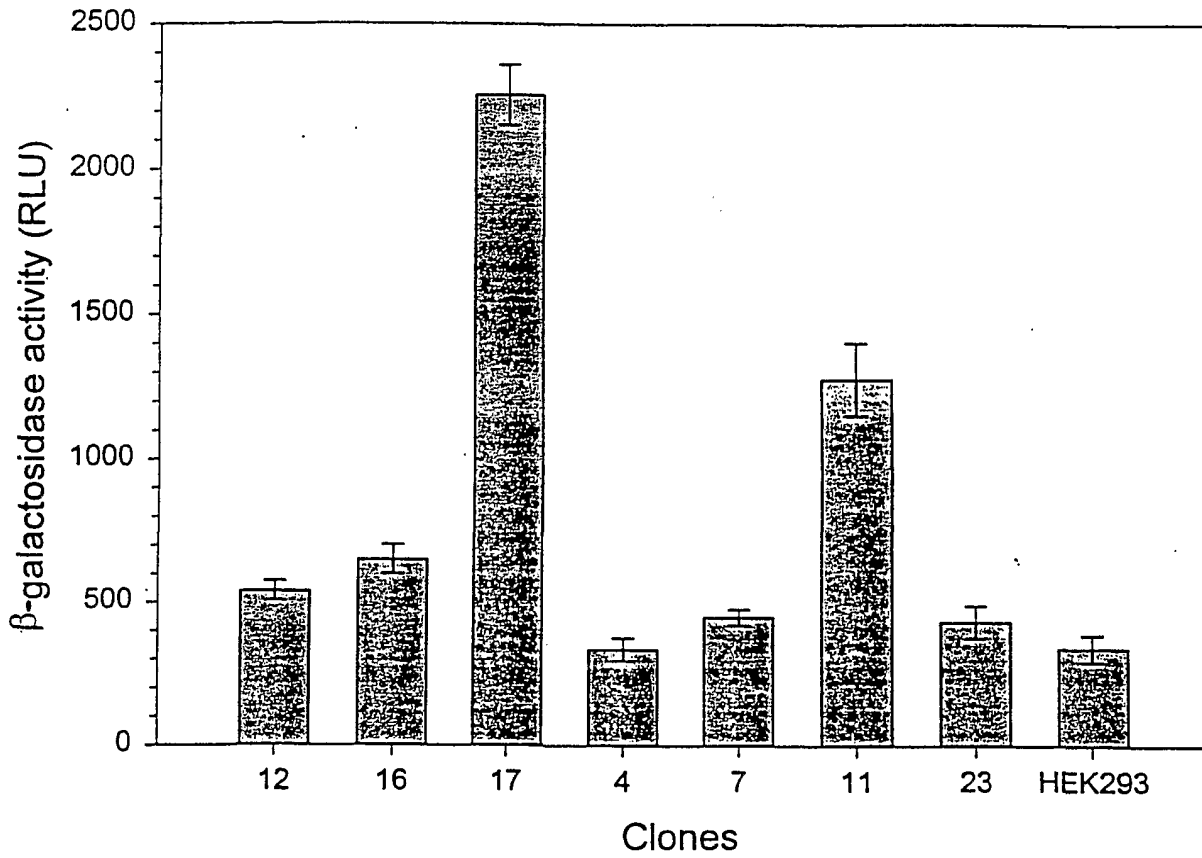


FIGURE 9A

Agonist Stimulated cAMP Response in HEK 293 Cells
Coexpressing β 2AR- β gal $\Delta\alpha$ and β 2AR- β gal $\Delta\omega$

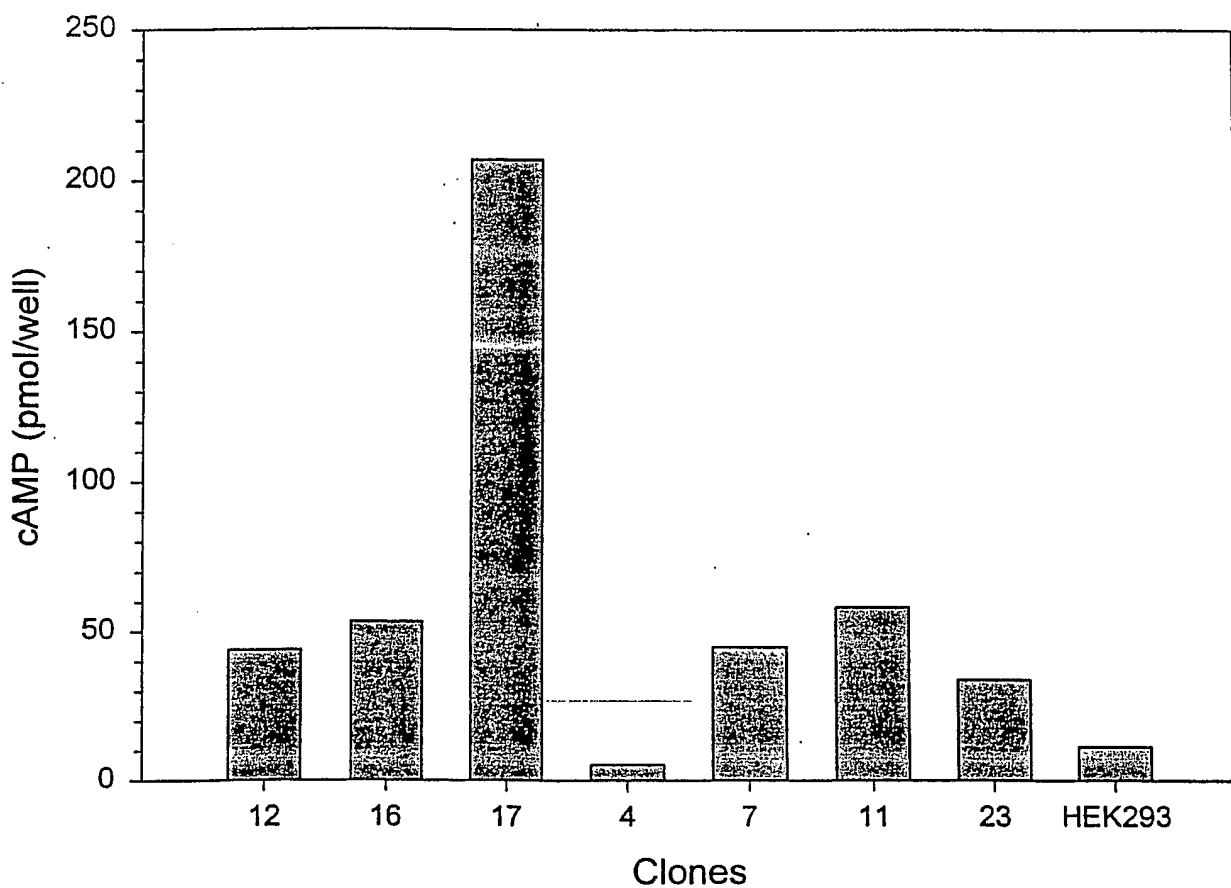


FIGURE 9B

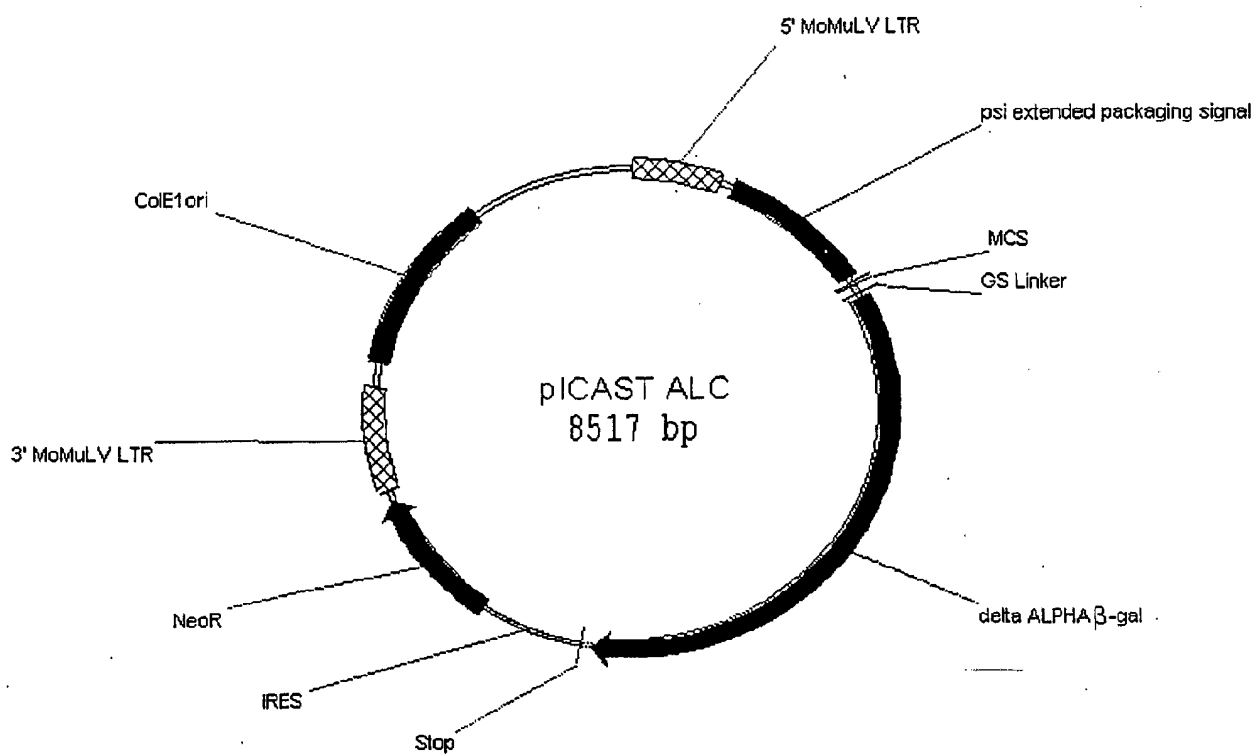


Figure 10A

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1  CTGCAGCCTG AATATGGGCC AAACAGGATA TCTGTGGTAA GCAGTTCCTG
   GACGTCGGAC TTATACCCGG TTTGTCCTAT AGACACCATT CGTCAAGGAC
-----
51  CCCC GGCTCA GGGCCAAGAA CAGATGGAAC AGCTGAATAT GGGCCAAACA
   GGGGCCGAGT CCCGGTTCTT GTCTACCTTG TCGACTTATA CCCGGTTTGT
-----
101 GGATATCTGT GGTAAGCAGT TCCTGCCCCG GCTCAGGGCC AAGAACAGAT
   CCTATAGACA CCATTCGTCA AGGACGGGGC CGAGTCCCGG TTCTTGTCTA
-----
151 GGTCCCCAGA TGCGGTCCAG CCCTCAGCAG TTTCTAGAGA ACCATCAGAT
   CCAGGGGTCT ACGCCAGGTC GGGAGTCGTC AAAGATCTCT TGGTAGTCTA
-----
201 GTTTCAGGGG TGCCCAAGG ACCTGAAATG ACCCTGTGCC TTATTGAAAC
   CAAAGGTCCC ACGGGGTTC TGGACTTTAC TGGGACACGG AATAAACTTG
-----
251 TAACCAATCA GTTCGCTTCT CGCTTCTGTT CGCGCGCTTC TGCTCCCCGA
   ATTGGTTAGT CAAGCGAAGA GCGAAGACAA GCGCGCGAAG ACGAGGGGCT
-----
301 GCTCAATAAA AGAGCCACA ACCCCTCACT CGGGGCGCCA GTCCTCCGAT
   CGAGTTATTT TCTCGGGTGT TGGGGAGTGA GCCCGCGGT CAGGAGGCTA
-----
351 TGACTGAGTC GCCCGGTAC CCGTGTATCC AATAAACCTT CTGTCAGTTG
   ACTGACTCAG CCGGCCCATG GGCACATAGG TTATTGGGA GAACGTCAAC
-----
401 CATCCGACTT GTGGTCTCGC TGTTCCCTGG GAGGGTCTCC TCTGAGTGAT
   GTAGGCTGAA CACCAGAGCG ACAAGGAACC CTCCAGAGG AGACTCACTA
-----
451 TGACTACCCG TCAGCGGGGG TCTTTCATTT GGGGGCTCGT CCGGGATCGG
   ACTGATGGGC AGTCGCCCCC AGAAAGTAAA CCCCCGAGCA GGCCTTAGCC
-----
501 GAGACCCCTG CCCAGGGACC ACCGACCCAC CACCGGGAGG CAAGCTGGCC
   CTCTGGGGAC GGTTCCTTGG TGGCTGGGTG GTGGCCCTCC GTTCGACCGG
-----
551 AGCAACTTAT CTGTGTCTGT CCGATTGTCT AGTGTCTATG ACTGATTTTA
   TCGTTGAATA GACACAGACA GGCTAACAGA TCACAGATAC TGACTAAAT
-----
601 TGCGCCTGCG TCGGTACTAG TTAGCTAACT AGCTCTGTAT CTGGCGGACC
   ACGCGGACGC AGCCATGATC AATCGATTGA TCGAGACATA GACCGCCTGG
-----
651 CGTGGTGGAA CTGACGAGTT CTGAACACCC GGCCGCAACC CTGGGAGACG
   GCACCACCTT GACTGCTCAA GACTTGTGGG CCGCGTGGG GACCCTCTGC
-----
701 TCCCAGGGAC TTTGGGGGCC GTTTTTGTGG CCCGACCTGA GGAAGGGAGT
   AGGGTCCCTG AAACCCCGG CAAAACACC GGGCTGGACT CCTTCCCTCA
-----
751 CGATGTGGAA TCCGACCCCG TCAGGATATG TGGTCTGGT AGGAGACGAG
   GCTACACCTT AGGCTGGGGC AGTCCTATAC ACCAAGACCA TCCTCTGCTC
-----
801 AACCTAAAAC AGTTCCCGCC TCCGTCTGAA TTTTGTCTTT CGGTTTGGAA
   TTGGATTTTG TCAAGGGCGG AGGCAGACTT AAAAACGAAA GCCAAACCTT
-----
851 CCGAAGCCGC GCGTCTGTG TGCTGCAGCA TCGTCTGTG TTGTCTCTGT
   GGCTTCGGCG CGCAGAACAG ACGACGTCGT AGCAAGACAC AACAGAGACA
-----
901 CTGACTGTGT TTCTGTATTT GTCTGAAAAT TAGGGCCAGA CTGTTACCAC
   GACTGACACA AAGACATAAA CAGACTTTTA ATCCCGGTCT GACAATGGTG
-----

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FIGURE 10B

951 TCCCTTAAGT TTGACCTTAG GTAAC TGAA AGATGTGGAG CCGCTGGCTC
 AGGGAATTCA AACTGGAATC CATTGACCTT TCTACAGCTC GCCGAGCGAG

1001 ACAACCAGTC GGTAGATGTC AAGAAGAGAC GTTGGGTTAC CTTCTGCTCT
 TGTGGTTCAG CCATCTACAG TTCTTCTCTG CAACCCAATG GAAGACGAGA

1051 GCAGAAATGGC CAACCTTTAA CGTCGGATGG CCGCGAGACG GCACCTTTAA
 CGTCTTACCG GTTGGAAATT GCAGCCTACC GCGCTCTGTC CGTGGAAATT

1101 CCGAGACCTC ATCACCCAGG TTAAGATCAA GGTCTTTTCA CCTGGCCCGC
 GGCTCTGGAG TAGTGGGTCC AATTCTAGTT CCAGAAAAGT GGACCGGGCG

1151 ATGGACACCC AGACCAGGTC CCCTACATCG TGACCTGGGA AGCCTTGGCT
 TACCTGTGGG TCTGGTCCAG GGGATGTAGC ACTGGACCCCT TCGGAACCGA

1201 TTTGACCCCC CTCCTGGGT CAAGCCCTTT GTACACCCTA AGCCTCCGCC
 AAACCTGGGGG GAGGGACCCA GTTCGGGAAA CATGTGGGAT TCGGAGGCGG

1251 TCCTCTTCCT CCATCCGCC CGTCTCTCCC CCTTGAACCT CCTCGTTCGA
 AGGAGAAGGA GGTAGGCGGG GCAGAGAGGG GGAACCTGGA GGAGCAAGCT

1301 CCCCGCCTCG ATCCTCCCTT TATCCAGCCC TCACTCCTTC TCTAGGCGCC
 GGGGCGGAGC TAGGAGGGAA ATAGGTGCGG AGTGAGGAAG AGATCCGCGG

1351 GGCCGCTCTA GCCATTAAT ACGACTCACT ATAGGGCGAT TCGAATCAGG
 CCGGCGAGAT CGGTAATTA TGCTGAGTGA TATCCCGCTA AGCTTAGTCC

1401 CTTGGCGCG CCGGATCCTT AATTAAGCGC AATTGGGAGG TGGCGGTAGC
 GGAACCGCGC GGCCTAGGAA TTAATTGCGG TTAACCCCTC ACCGCCATCG

+2 M G V I T D S L A V V A R T D
]

1451 CTCGAGATGG GCGTGATTAC GGATTCAC TGCCGTCGTGG CCCGCACCGA
 GAGCTCTACC CGCACTAATG CCTAAGTGAC CCGCAGCACC GGGCGTGGCT

+2 R P S Q Q L R S L N G E W R F A

1501 TCGCCCTTCC CAACAGTTAC GCAGCCTGAA TGGCGAATGG CGCTTTGCCT
 AGCGGAAGG GTTGTCAATG CGTCGGACTT ACCGCTTACC GCGAAACGGA

+2 W F P A P E A V P E S W L E C D L

1551 GGTTCCCGC ACCAGAAGCG GTGCCGAAA GCTGGCTGGA GTGCGATCTT
 CCAAAGGCCG TGGTCTTCGC CACGGCCTTT CGACCGACCT CACGCTAGAA

+2 P E A D T V V V P S N W Q M H G Y

1601 CTTGAGGCGG ATACTGTGCT CGTCCCCTCA AACTGGCAGA TGCACGGTTA
 GGACTCCGGC TATGACAGCA GCAGGGGAGT TTGACCGTCT ACGTGCCAAT

+2 D A P I Y T N V T Y P I T V N P

1651 CGATGCGCCC ATCTACACCA ACGTGACCTA TCCCATACG GTCAATCCGC
 GCTACGCGGG TAGATGTGGT TGCACGGAT AGGGTAATGC CAGTTAGCG

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+2 P F V P T E N P T G C Y S L T F N
-----
1701 CGTTTGTTC CACGGAGAAT CCGACGGGT GTTACTCGCT CACATTTAAT
    GCAAACAAGG GTGCCTCTTA GGCTGCCCAA CAATGAGCGA GTGTAAATTA
-----
+2 V D E S W L Q E G Q T R I I F D G
-----
1751 GTTGATGAAA GCTGGCTACA GGAAGGCCAG ACGCGAATTA TTTTGTATGG
    CAACTACTTT CGACCGATGT CCTTCCGGTC TGCCTTAAT AAAAATACC
-----
+2 V N S A F H L W C N G R W V G Y
-----
1801 CGTTAACTCG GCGTTTCATC TGTGGTGCAA CGGCGCTGG GTCGGTTACG
    GCAATTGAGC CGCAAAGTAG ACACCACGTT GCCCGCGACC CAGCCAATGC
-----
+2 G Q D S R L P S E F D L S A F L R
-----
1851 GCCAGGACAG TCGTTTGGCG TCTGAATTTG ACCTGAGCGC ATTTTACGC
    CGGTCCTGTC AGCAAACGGC AGACTTAAAC TGGACTCGCG TAAAAATGCG
-----
+2 A G E N R L A V M V L R W S D G S
-----
1901 GCCGGAGAAA ACCGCCTCGC GGTGATGGTG CTGCGCTGGA GTGACGGCAG
    CGGCCTCTTT TGGCGGAGCG CCACTACCAC GACGCGACCT CACTGCCGTC
-----
+2 Y L E D Q D M W R M S G I F R D
-----
1951 TTATCTGGAA GATCAGGATA TGTGGCGGAT GAGCGGCATT TTCCGTGACG
    AATAGACCTT CTAGTCCTAT ACACCGCCTA CTCGCCGTAA AAGGCACTGC
-----
+2 V S L L H K P T T Q I S D F H V A
-----
2001 TCTCGTTGCT GCATAAACCG ACTACACAAA TCAGCGATTT CCATGTTGCC
    AGAGCAACGA CGTATTTGGC TGATGTGTTT AGTCGCTAAA GGTACAACGG
-----
+2 T R F N D D F S R A V L E A E V Q
-----
2051 ACTCGCTTTA ATGATGATTT CAGCCGCGCT GTACTGGAGG CTGAAGTTCA
    TGAGCGAAAT TACTACTAAA GTCGGCGCGA CATGACCTCC GACTTCAAGT
-----
+2 M C G E L R D Y L R V T V S L W
-----
2101 GATGTGCGGC GAGTTGCGTG ACTACCTACG GGTAACAGTT TCTTTATGGC
    CTACACGCCG CTCAACGCAC TGATGGATGC CCATTGTCAA AGAAATACCG
-----
+2 Q G E T Q V A S G T A P F G G E I
-----
2151 AGGGTGA AAC GCAGTTCGCC AGCGGCACCG CGCCTTCGG CGGTGAAATT
    TCCCACTTTG CGTCCAGCGG TCGCCGTGGC GCGGAAAGCC GCCACTTTAA
-----
+2 I D E R G G Y A D R V T L R L N V
-----
2201 ATCGATGAGC GTGGTGGTTA TGCCGATCGC GTCACACTAC GTCTGAACGT
    TAGCTACTCG CACCACCAAT ACGGCTAGCG CAGTGTGATG CAGACTTGCA
-----
+2 E N P K L W S A E I P N L Y R A
-----
2251 CGAAAACCCG AACTGTGGA GCGCCGAAAT CCCGAATCTC TATCGTGGCG
    GCTTTTGGGC TTTGACACCT CGCGGCTTTA GGGCTTAGAG ATAGCACGCC
-----

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+2 V V E L H T A D G T L I E A E A C
-----
2301 TGGTTGAACT GCACACCGCC GACGGCACGC TGATTGAAGC AGAAGCCTGC
    ACCAACTTGA CGTGTGGCGG CTGCCGTGCG ACTAACTTCG TCTTCGGACG
-----
+2 D V G F R E V R I E N G L L L L N
-----
2351 GATGTCGGTT TCCGCGAGGT GCGGATTGAA AATGGTCTGC TGCTGCTGAA
    CTACAGCCAA AGGCGCTCCA CGCCTACTT TTACCAGACG ACGACGACTT
-----
+2 G K P L L I R G V N R H E H H P
-----
2401 CGGCAAGCCG TTGCTGATTC GAGGCGTTAA CCGTCACGAG CATCATCCTC
    GCCGTTCCGC AAGACTAAG CTCCGCAATT GGCAGTGCTC GTAGTAGGAG
-----
+2 L H G Q V M D E Q T M V Q D I L L
-----
2451 TGCATGGTCA GGTCATGGAT GAGCAGACGA TGGTGCAGGA TATCCTGCTG
    ACGTACCAGT CCAGTACCTA CTCGTCTGCT ACCACGTCCT ATAGGACGAC
-----
+2 M K Q N N F N A V R C S H Y P N H
-----
2501 ATGAAGCAGA ACAACTTTAA CGCCGTGCGC TGTTGCGATT ATCCGAACCA
    TACTTCGTCT TGTGAAATT GCGGCACGCG ACAAGCGTAA TAGGCTTGGT
-----
+2 P L W Y T L C D R Y G L Y V V D
-----
2551 TCCGCTGTGG TACACGCTGT GCGACCCTA CGGCCTGTAT GTGGTGGATG
    AGGCGACACC ATGTGCGACA CGCTGGCGAT GCCGGACATA CACCACCTAC
-----
+2 E A N I E T H G M V P M N R L T D
-----
2601 AAGCCAATAT TGAAACCCAC GGCATGGTGC CAATGAATCG TCTGACCGAT
    TTCGGTTATA ACTTTGGGTG CCGTACCACG GTTACTTAGC AGACTGGCTA
-----
+2 D P R W L P A M S E R V T R M V Q
-----
2651 GATCCGCGCT GGCTACCGGC GATGAGCGAA CGCGTAACGC GAATGGTGCA
    CTAGGCGCGA CCGATGGCCG CTA CTCTGCTT GCGCATTGCG CTTACCACGT
-----
+2 R D R N H P S V I I W S L G N E
-----
2701 GCGCGATCGT AATCACCCGA GTGTGATCAT CTGGTCGCTG GGGAAATGAAT
    CGCGCTAGCA TTAGTGGGCT CACACTAGTA GACCAGCGAC CCCTTACTTA
-----
+2 S G H G A N H D A L Y R W I K S V
-----
2751 CAGGCCACGG CGCTAATCAC GACGCGCTGT ATCGCTGGAT CAAATCTGTC
    GTCCGGTGCC GCGATTAGTG CTGCGCGACA TAGCGACCTA GTTTAGACAG
-----
+2 D P S R P V Q Y E G G G A D T T A
-----
2801 GATCCTTCCC GCCCGTGCA GTATGAAGGC GCGGAGCCG ACACCAGGC
    CTAGGAAGGG CGGGCCACGT CATACTTCCG CCGCCTCGGC TGTGGTGCCG
-----
+2 T D I I C P M Y A R V D E D Q P
-----
2851 CACCGATATT ATTTGCCCGA TGTACGCGCG CGTGGATGAA GACCAGCCCT
    GTGGCTATAA TAAACGGGCT ACATGCGCGC GCACCTACTT CTGGTCGGGA
-----

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+2 F P A V P K W S I K K W L S L P G
-----
2901 TCCCGGCTGT GCCGAAATGG TCCATCAAAA AATGGCTTTC GCTACCTGGA
    AGGGCCGACA CGGCTTTACC AGGTAGTTTT TTACCGAAAG CGATGGACCT
-----
+2 E T R P L I L C E Y A H A M G N S
-----
2951 GAGACGCGCC CGCTGATCCT TTGCGAATAC GCCCAGCGCA TGGGTAACAG
    CTCTGCGCGG GCGACTAGGA AACGCTTATG CCGGTGCGCT ACCCATTGTC
-----
+2 L G G F A K Y W Q A F R Q Y P R
-----
3001 TCTTGGCGGT TTCGCTAAAT ACTGGCAGGC GTTTCGTCAG TATCCCCGT
    AGAACCGCCA AAGCGATTTA TGACCGTCCG CAAAGCAGTC ATAGGGGCAA
-----
+2 L Q G G F V W D W V D Q S L I K Y
-----
3051 TACAGGGCGG CTTCGTCTGG GACTGGGTGG ATCAGTCGCT GATTAAATAT
    ATGTCCC GCCA GAAGCAGACC CTGACCCACC TAGTCAGCGA CTAATTTATA
-----
+2 D E N G N P W S A Y G G D F G D T
-----
3101 GATGAAAACG GCAACCCGTG GTCGGCTTAC GCGCGTGATT TTGGCGATAC
    CTACTTTTGC CGTTGGGCAC CAGCCGAATG CCGCCACTAA AACCGCTATG
-----
+2 P N D R Q F C M N G L V F A D R
-----
3151 GCCGAACGAT CGCCAGTTCT GTATGAACGG TCTGGTCTTT GCCGACCGCA
    CCGCTTGCTA GCGGTCAAGA CATACTTGCC AGACCAGAAA CCGCTGGCGT
-----
+2 T P H P A L T E A K H Q Q Q F F Q
-----
3201 CGCCGCATCC AGCGTGACG GAAGCAAAC ACCAGCAGCA GTTTTCCAG
    GCGGCGTAGG TCGCGACTGC CTTCGTTTTG TGGTCGTCGT CAAAAGGTC
-----
+2 F R L S G Q T I E V T S E Y L F R
-----
3251 TTCCGTTTTAT CCGGGCAAAC CATCGAAGTG ACCAGCGAAT ACCTGTTCCG
    AAGGCAAATA GGCCCGTTG GTAGCTTAC TGGTCGCTTA TGGACAAGGC
-----
+2 H S D N E L L H W M V A L D G K
-----
3301 TCATAGCGAT AACGAGCTCC TGCCTGGAT GGTGGCGCTG GATGGTAAGC
    AGTATCGCTA TTGCTCGAGG ACGTGACCTA CCACCGCGAC CTACCATTCC
-----
+2 P L A S G E V P L D V A P Q G K Q
-----
3351 CGCTGGCAAG CCGTGAAGTG CCTCTGGATG TCGCTCCACA AGGTAAACAG
    GCGACCGTTC GCCACTTCAC GGAGACCTAC AGCGAGGTGT TCCATTGTC
-----
+2 L I E L P E L P Q P E S A G Q L W
-----
3401 TTGATTGAAC TGCCTGAACT ACCGCAGCCG GAGAGCGCCG GGCAACTCTG
    AACTAACTTG ACGGACTTGA TGGCGTCGGC CTCTCGCGGC CCGTTGAGAC
-----
+2 L T V R V V Q P N A T A W S E A
-----
3451 GCTCACAGTA CGCGTAGTGC AACCGAACGC GACCGCATGG TCAGAAGCCG
    CGAGTGTTCAT GCGCATCAG TTGGCTTGCG CTGGCGTACC AGTCTTCGGC
    
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+2 G H I S A W Q Q W R L A E N L S V
-----
3501 GGCACATCAG CGCCTGGCAG CAGTGGCGTC TGGCGGAAAA CCTCAGTGTG
    CCGTGTAGTC GCGGACCGTC GTCACCGCAG ACCGCCTTTT GGAGTCACAC
-----
+2 T L P A A S H A I P H L T T S E M
-----
3551 ACGCTCCCCG CCGCGTCCCA CGCCATCCCG CATCTGACCA CCAGCGAAAT
    TGGAGAGGGG GCGCGAGGGT GCGGTAGGGC GTAGACTGGT GGTGCGTTTA
-----
+2 D F C I E L G N K R W Q F N R Q
-----
3601 GGATTTTTGC ATCAGACTGG GTAATAAGCG TTGGCAATTT AACCGCCAGT
    CCTAAAAACG TAGCTCGACC CATTATTCGC AACCGTTAAA TTGGCGGTCA
-----
+2 S G F L S Q M W I G D K K Q L L T
-----
3651 CAGGCTTTCT TTCACAGATG TGGATTGGCG ATAAAAACA ACTGCTGACG
    GTCCGAAAGA AAGTGTCTAC ACCTAACCGC TATTTTTTGT TGACGACTGC
-----
+2 P L R D Q F T R A P L D N D I G V
-----
3701 CCGCTGCGCG ATCAGTTCAC CCGTGCACCG CTGGATAACG ACATTGGCGT
    GCGCAGCGCG TAGTCAAGTG GGCACGTGGC GACCTATTGC TGTAACCGCA
-----
+2 S E A T R I D P N A W V E R W K
-----
3751 AAGTGAAGCG ACCCGCATTG ACCCTAACGC CTGGGTCGAA CGCTGGAAGG
    TTCACTTCGC TGGGCGTAAC TGGGATTGCG GACCCAGCTT GCGACCTTC
-----
+2 A A G H Y Q A E A A L L Q C T A D
-----
3801 CCGCGGGCCA TTACCAGGCC GAAGCAGCGT TGTTGCAGTG CACGGCAGAT
    GCGCGCCGGT AATGGTCCGG CTTGTCGCA ACAACGTCAC GTGCCGTCTA
-----
+2 T L A D A V L I T T A H A W Q H Q
-----
3851 ACACTTGCTG ATGCGGTGCT GATTACGACC GCTCACGCGT GGCAGCATCA
    TGTGAACGAC TAGCCACGA CTAATGCTGG CGAGTGCACA CCGTCGTAGT
-----
+2 G K T L F I S R K T Y R I D G S
-----
3901 GGGGAAAACC TTATTTATCA GCGGAAAAC CTACCGGATT GATGGTAGTG
    CCCCTTTTGG AATAAATAGT CCGCCTTTTG GATGGCCTAA CTACCATCAC
-----
+2 G Q M A I T V D V E V A S D T P H
-----
3951 GTCAAATGGC GATTACCGTT GATGTTGAAG TGGCGAGCGA TACACCGCAT
    CAGTTTACCG CTAATGGCAA CTACAACCTC ACCGCTCGCT ATGTGGCGTA
-----
+2 P A R I G L N C Q L A Q V A E R V
-----
4001 CCGGCGCGGA TTGGCCTGAA CTGCCAGCTG GCGCAGGTAG CAGAGCGGGT
    GGCCGCGCCT AACCGGACTT GACGGTCGAC CGCGTCCATC GTCTCGCCCA
-----
+2 N W L G L G P Q E N Y P D R L T
-----
4051 AAAGTGGCTC GGATTAGGGC CGCAAGAAAA CTATCCGGAC CGCCTTACTG
    TTTGACCGAG CCTAATCCCG GCGTTCCTTT GATAGGGCTG GCGGAATGAC
-----

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+2 A A C F D R W D L P L S D M Y T P
-----
4101 CCGCCTGTTT TGACCCTGG GATCTGCCAT TGTCAGACAT GTATACCCCG
    GCGGACAAA ACTGGCGACC CTAGACGGTA ACAGTCTGTA CATATGGGGC
-----
+2 Y V F P S E N G L R C G T R E L N
-----
4151 TACGTCTTCC CGAGCGAAAA CCGTCTGCGC TGCGGGACGC GCGAATTGAA
    ATGCAGAAGG GCTCGCTTTT GCCAGACGCG ACGCCCTGCG CGCTTAACTT
-----
+2 Y G P H Q W R G D F Q F N I S R
-----
4201 TTATGGCCCA CACCAGTGGC CCGGCGACTT CCACTTCAAC ATCAGCCCGT
    AATACCGGGT GTGGTCACCG CGCCGCTGAA GGTCAGTTG TAGTCGGCGA
-----
+2 Y S Q Q Q L M E T S H R H L L H A
-----
4251 ACAGTCAACA GCAACTGATG GAAACCAGCC ATCGCCATCT GCTGCACGCG
    TGTCAGTTGT CGTTGACTAC CTTTGGTGGG TAGCGGTAGA CGACGTGCGC
-----
+2 E E G T W L N I D G F H M G I G G
-----
4301 GAAGAAGGCA CATGGCTGAA TATCGACGGT TTCCATATGG GGATTGGTGG
    CTTCTCCCGT GTACCGACTT ATAGCTGCCA AAGGTATACC CCTAACCCACC
-----
+2 D D S W S P S V S A E F Q L S A
-----
4351 CGACGACTCC TGGAGCCCGT CAGTATCGGC GGAATTECAG CTGAGCGCCG
    GCTGCTGAGG ACCTCGGGCA GTCATAGCCG CCTTAAGGTC GACTCGCGGC
-----
+2 G R Y H Y Q L V W C Q K R S D Y K
-----
4401 GTCGCTACCA TTACCAGTTG GTCTGGTGTG AAAAAAGATC TGAATAAAA
    CAGCGATGGT AATGGTCAAC CAGACCACAG TTTTCTAG ACTGATATT
-----
+2 D E D L D H H H H H H R
-----
4451 GATGAGGACC TCGACCATCA TCATCATCAT CACCGGTAAT AATAGGTAGA
    CTACTCCTGG AGCTGGTAGT AGTAGTAGTA GTGGCATT AATCCATCT
-----
4501 TAAGTGACTG ATTAGATGCA TTGATCCCTC GACCAATTCC GGTATTATTC
    ATTCACTGAC TAATCTACGT AACTAGGGAG CTGGTTAAGG CCAATAAAAG
-----
4551 CACCATATTG CCGTCTTTTG GCAATGTGAG GGCCCGGAAA CCTGGCCCTG
    GTGGTATAAC GGCAGAAAAC CGTTACACTC CCGGGCCTTT GGACCGGGAC
-----
4601 TCTTCTTGAC GAGCATTCTT AGGGGTCTTT CCCCTCTCGC CAAAGGAATG
    AGAAGAAGTG CTCGTAAGGA TCCCAGAAA GGGGAGAGCG GTTTCCTTAC
-----
4651 CAAGGTCTGT TGAATGTCGT GAAGGAAGCA GTTCCTCTGG AAGCTTCTTG
    GTCCAGACA ACTTACAGCA CTCCTTCGT CAAGGAGACC TTCGAAGAAC
-----
4701 AAGACAAACA ACGTCTGTAG CGACCCTTTG CAGGCAGCGG AACCCCCAC
    TTCTGTTTGT TGCAGACATC GCTGGGAAAC GTCCGTCGCC TTGGGGGGTG
-----
4751 CTGGCGACAG GTGCCTCTGC GGCCAAAAGC CACGTGTATA AGATACACCT
    GACCGCTGTC CACGGAGACG CCGGTTTTCG GTGCACATAT TCTATGTGGA
    
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4801 GCAAAGGCGG CACAACCCCA GTGCCACGTT GTGAGTTGGA TAGTTGTGGA
CGTTTCCGCC GTGTTGGGGT CACGGTGCAA CACTCAACCT ATCAACACCT

4851 AAGAGTCAAA TGGCTCTCCT CAAGCGTATT CAACAAGGGG CTGAAGGATG
TTCTCAGTTT ACCGAGAGGA GTTCGCATAA GTTGTTCGCC GACTTCTCTAC

4901 CCCAGAAGGT ACCCCATTGT ATGGGATCTG ATCTGGGGCC TCGGTGCACA
GGGTCTTCCA TGGGGTAACA TACCCTAGAC TAGACCCCGG AGCCACGTGT

4951 TGCTTTACAT GTGTTTAGTC GAGGTTAAAA AACGTCTAGG CCCCCGAAC
ACGAAATGTA CACAAATCAG CTCCAATTTT TTGCAGATCC GGGGGGCTTG

5001 CACGGGGACG TGGTFTTCTT TTGAAAAACA CGATGATAAT ACCATGATTG
GTGCCCTGC ACCAAAAGGA AACTTTTTGT GCTACTATTA TGGTACTAAC

5051 AACAAAGATGG ATTGCACGCA GGTTCTCCGG CCGCTTGGGT GGAGAGGCTA
TTGTTCTACC TAACGTGCGT CCAAGAGGCC GCGGAACCCA CCTCTCCGAT

5101 TTCGGCTATG ACTGGGCACA ACAGACAATC GGCTGCTCTG ATGCCGCCGT
AAGCCGATAC TGACCCGTGT TGTCTGTAG CCGACGAGAC TACGGCGGCA

5151 GTTCCGGCTG TCAGCGCAGG GCGGCCCGGT TCTTTTTGTC AAGACCGACC
CAAGGCCGAC AGTCGCGTCC CCGCGGGCCA AGAAAAACAG TTCTGGCTGG

5201 TGTCCGGTGC CCTGAATGAA CTGCAGGACG AGGCAGCGCG GCTATCGTGG
ACAGGCCACG GGACTTACTT GACGTCCTGC TCCGTGCGCG CGATAGCACC

5251 CTGGCCACGA CGGGCGTTC CTGCGCAGCT GTGCTCGACG TTGTCACTGA
GACCGGTGCT GCCCGCAAGG AACGCGTCGA CACGAGCTGC AACAGTGACT

5301 AGCGGGAAGG GACTGGCTGC TATTGGGCGA AGTGCCGGGG CAGGATCTCC
TCGCCCTTCC CTGACCGACG ATAACCCGCT TCACGGCCCC GTCCCTAGAGG

5351 TGTCATCTCA CCTTGCTCCT GCCGAGAAAG TATCCATCAT GGCTGATGCA
ACAGTAGAGT GGAACGAGGA CGGCTCTTTC ATAGGTAGTA CCGACTACGT

5401 ATGCCGCGGC TGCATACGCT TGATCCGGCT ACCTGCCCAT TCGACCACCA
TACGCCCGCG ACGTATGCGA ACTAGGCCGA TGGACGGGTA AGCTGGTGGT

5451 AGCGAAACAT CGCATCGAGC GAGCACGTAC TCGGATGGAA GCCGGTCTTG
TCGCTTTGTA GCGTAGCTCG CTCGTGCATG AGCCTACCTT CGGCCAGAAC

5501 TCGATCAGGA TGATCTGGAC GAAGAGCATC AGGGGCTCGC GCCAGCCGAA
AGCTAGTCCT ACTAGACCTG CTTCTCCTAG TCCCCGAGCG CGGTCCGCTT

5551 CTGTTGCGCA GGCTCAAGGC GCGCATGCCC GACGGCGAGG ATCTCGTCGT
GACAAGCGGT CCGAGTCCG CCGGTACGGG CTGCCGCTCC TAGAGCAGCA

5601 GACCCATGGC GATGCCTGCT TGCCGAATAT CATGGTGCAA AATGGCCGCT
CTGGGTACCG CTACGACGCA ACGGCTTATA GTACCACCTT TTACCGGCCA

5651 TTTCTGGATT CATCGACTGT GGCCGGCTGG GTGTGGCGGA CCGCTATCAG
AAAGACCTAA GTAGCTGACA CCGGCCGACC CACACCCTT GCGGATAGTC

5701 GACATAGCGT TGGCTACCCG TGATATTGCT GAAGAGCTTG GCGCGAATG
CTGTATCGCA ACCGATGGGC ACTATAACGA CTTCTCGAAC CGCCGCTTAC

5751 GGCTGACCGC TTCCTCGTGC TTTACGGTAT CGCCGCTCCC GATTGCGAGC
 CCGACTGGCG AAGGAGCAGC AAATGCCATA GCGGCGAGGG CTAAGCGTCG

5801 GCATCGCCTT CTATCGCCTT CTTGACGAGT TCTTCTGAGC GGGACTCTGG
 CGTAGCGGAA GATAGCGGAA GAAGTGTCTA AGAAGACTCG CCCTGAGACC

5851 GGTTCGCATC GATAAAATAA AAGATTTTAT TTAGTCTCCA GAAAAGGGG
 CCAAGCGTAG CTATTTTATT TTCTAAAATA AATCAGAGGT CTTTTTCCCC

5901 GGAATGAAAG ACCCCACCTG TAGGTTTGGC AAGCTAGCTT AAGTAACGCC
 CCTTACTTTT TGGGGTGGAC ATCCAAACCG TCGATCGAA TTCATTGCGG

5951 ATTTTGCAAG GCATGGAAAA ATACATAACT GAGAATAGAG AAGTTCAGAT
 TAAAACGTTT CGTACCTTTT TATGTATTGA CTCTTATCTC TTCAAGTCTA

6001 CAAGGTCAGG AACAGATGGA ACAGCTGAAT ATGGGCCAAA CAGGATATCT
 GTTCCAGTCC TTGTCTACCT TGTCGACTTA TACCCGGTTF GTCCTATAGA

6051 GTGGTAAGCA GTTCTGCCCC CGGCTCAGGG CCAAGAACAG ATGGAACAGC
 CACCATTTCG CAAGGACGGG GCCGAGTCCC GGTCTTTGTC TACCTTGTCG

6101 TGAATATGGG CCAAACAGGA TATCTGTGGT AAGCAGTTC TGCCCCGGCT
 ACTTATACCC GSTTTGTCTT ATAGACACCA TTCGTCAAGG ACGGGGCCGA

6151 CAGGGCCAAG AACAGATGGT CCCCAGATGC GGTCCAGCCC TCAGCAGTTT
 GTCCCGGTTT TTGTCTACCA GGGGTCTACG CCAGGTCGGG AGTCGTCAAA

6201 CTAGAGAACC ATCAGATGTT TCCAGGGTGC CCCAAGGACC TGAAATGACC
 GATCTCTTGG TAGTCTACAA AGGTCCACG GGGTTCCTGG ACTTTACTGG

6251 CTGTGCCTTA TTGAACTAA CCAATCAGTT CGCTTCTCGC TTCTGTTTCG
 GACACGGAAT AAACCTGATT GGTTAGTCAA GCGAAGAGCG AAGACAAGCG

6301 GCGCTTCTGC TCCCCGAGCT CAATAAAGA GCCACAACC CCTCACTCGG
 CGCGAAGACG AGGGGCTCGA GTTATTTTCT CGGGTGTTGG GGAGTGAGCC

6351 GGCGCCAGTC TCCGATTGA CTGAGTCGCC CGGGTACCCG TGTATCCAAT
 CCGCGGTCAG GAGGCTAACT GACTCAGCGG GCCCATGGGC ACATAGGTTA

6401 AAACCCTCTT GCAGTTGCAT CCGACTTGTG GTCTCGCTGT TCCTTGGGAG
 TTTGGGAGAA CGTCAACGTA GGCTGAACAC CAGAGCGACA AGGAACCCTC

6451 GGTCTCCTCT GAGTGATTGA CTACCCGTCA GCGGGGTCT TTCATTCTAG
 CCAGAGGAGA CTCACTAACT GATGGGCAGT CGCCCCAGA AAGTAAGTAC

6501 CAGCATGTAT CAAAATTAAT TTGGTTTTTT TTCTTAAGTA TTTACATTA
 GTCGTACATA GTTTTAATTA AACCAGAAAA AAGAATTCAT AAATGTAATT

6551 ATGGCCATAG TTGCATTAAT GAATCGGCCA ACGCGCGGGG AGAGGCGGTT
 TACCGGTATC AACGTAATTA CTTAGCCGGT TGCGCGCCCC TCTCCGCCAA

6601 TGCGTATTGG CGCTCTTCCG CTTCTCGCT CACTGACTCG CTGCGCTCGG
 ACGCATAACC GCGAGAAGGC GAAGGAGCGA GTGACTGAGC GACGCGAGCC

6651 TCGTTCGGCT GCGGCGAGCG GTATCAGCTC ACTCAAAGGC GGTAATACGG
 AGCAAGCCGA CGCCGCTCGC CATAGTCGAG TGAGTTTCCG CCATTATGCC

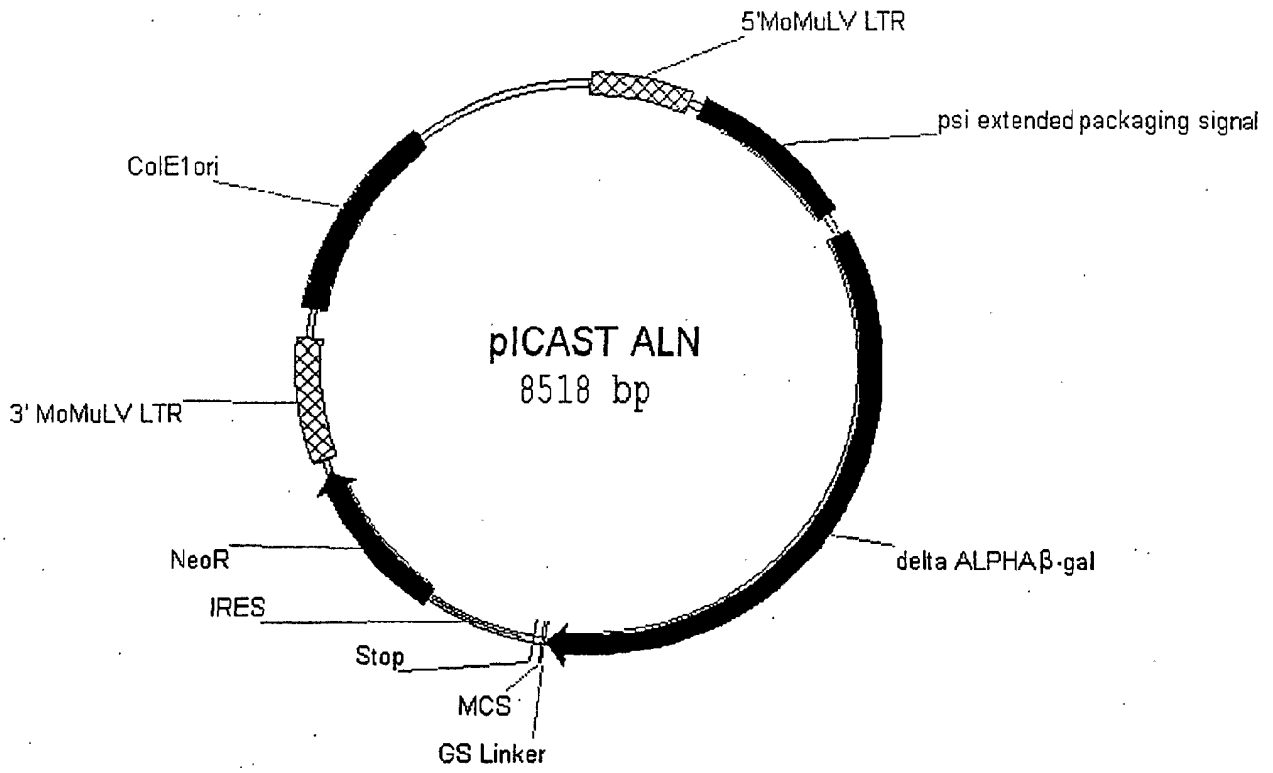


Figure 11A

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1  CTGCAGCCTG AATATGGGCC AAACAGGATA TCTGTGGTAA GCAGTTCCTG
   GACGTCGGAC TTATACCCGG TTTGTCCTAT AGACACCATT CGTCAAGGAC
-----
51  CCCC GGCTCA GGGCCAAGAA CAGATGGAAC AGCTGAATAT GGGCCAAACA
   GGGGCCGAGT CCCGGTTCTT GTCTACCTTG TCGACTTATA CCCGGTTTGT
-----
101 GGATATCTGT GGTAAGCAGT TCCTGCCCCG GCTCAGGGCC AAGAACAGAT
   CCTATAGACA CCATTCTGCA AGGACGGGGC CGAGTCCCGG TTCTTGTCTA
-----
151 GGTCCCCAGA TGCGGTCCAG CCCTCAGCAG TTCTAGAGA ACCATCAGAT
   CCAGGGTCT ACGCCAGGTC GGGAGTCGTC AAAGATCTCT TGGTAGTCTA
-----
201 GTTCCAGGG TGCCCAAGG ACCTGAAATG ACCCTGTGCC TTATTGAAAC
   CAAAGGTCCC ACGGGGTTCC TGGACTTAC TGGGACACGG AATAAACTTG
-----
251 TAACCAATCA GTTCGCTTCT CGCTTCTGTT CGCGCGCTTC TGCTCCCCGA
   ATTGGTTAGT CAAGCGAAGA GCGAAGACAA GCGCGGAAG ACGAGGGGCT
-----
301 GCTCAATAAA AGAGCCCACA ACCCTCACT CGGGGCGCCA GTCCTCCGAT
   CGAGTTATTT TCTCGGGTGT TGGGGAGTGA GCCCCGCGGT CAGGAGGCTA
-----
351 TGA CTGAGTC GCCCGGGTAC CCGTGTATCC AATAAACCCCT CTG CAGTTG
   ACTGACTCAG CGGGCCCATG GGCACATAGG TTATTTGGGA GAACGTCAAC
-----
401 CATCCGACTT GTGGTCTCGC TGTTCCCTGG GAGGGTCTCC TCTGAGTGAT
   GTAGGCTGAA CACCAGAGCG ACAAGGAACC CTCCAGAGG AGACTCACTA
-----
451 TGA CTACCCG TCAGCGGGGG TCTTTCATTT GGGGGCTCGT CCGGGATCGG
   ACTGATGGGC AGTCGCCCCC AGAAAGTAAA CCCCAGACA GGCCCTAGCC
-----
501 GAGACCCCTG CCCAGGGACC ACCGACCCAC CACCGGGAGG CAAGCTGGCC
   CTCTGGGGAC GGGTCCCTGG TGGCTGGGTG GTGGCCCTCC GTTCGACCGG
-----
-551 AGCAACTTAT CTGTGTCTGT CEGATTGTCT AGTGTCTATG ACTGATTTTA
   TCGTTGAATA GACACAGACA GGCTAACAGA TCACAGATC TGACTAAAAT
-----
601 TCGCCTGCG TCGGTACTAG TTAGCTAACT AGCTCTGTAT CTGGCGGACC
   ACGCGGACGC AGCCATGATC AATCGATTGA TCGAGACATA GACCCCTGG
-----
651 CGTGGTGGAA CTGACGAGTT CTGAACACCC GGCCGCAACC CTGGGAGACG
   GCACCACCTT GACTGTCAA GACTTGTGGG CCGGCGTTGG GACCCTCTGC
-----
701 TCCAGGGAC TTTGGGGGCC GTTTTGTGG CCCGACCTGA GGAAGGGAGT
   AGGGTCCCTG AAACCCCGG CAAAACACC GGGCTGGACT CCTTCCCTCA
-----
751 CGATGTGGAA TCCGACCCCG TCAGGATATG TGGTTCTGGT AGGAGACGAG
   GCTACACCTT AGGCTGGGGC AGTCCTATAC ACCAAGACCA TCCTCTGCTC
-----
801 AACCTAAAC AGTTCCCGCC TCCGTCTGAA TTTTGTCTT CGGTTTGGAA
   TTGGATTTTG TCAAGGGCGG AGGCAGACTT AAAAACGAAA GCCAAACCTT
-----
851 CCGAAGCCGC GCGTCTGTG TGCTGCAGCA TCGTTCTGTG TTGTCTCTGT
   GGCTTCGGCG CGCAGAACAG ACGACGTCGT AGCAAGACAC AACAGAGACA
-----
901 CTGACTGTGT TTCTGTATTT GTCTGAAAAT TAGGGCCAGA CTGTTACCAC
   GACTGACACA AAGACATAAA CAGACTTTTA ATCCCGTCT GACATGGTG
-----

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FIGURE 11B

951 TCCCTTAAGT TTGACCTTAG GTAAGTGGAA AGATGTCGAG CCGCTCGCTC
 AGGGAATTCA AACTGGAATC CATTGACCTT TCTACAGCTC GCCGAGCGAG

1001 ACAACCAGTC GGTAGATGTC AAGAAGAGAC GTTGGGTTAC CTTCTGCTCT
 TGTGGTTCAG CCATCTACAG TTCTTCTCTG CAACCCAAIG GAAGACGAGA

1051 GCAGAATGGC CAACCTTTAA CGTCGGATGG CCGCGAGACG GCACCTTTAA
 CGTCTTACCG GTTGAAATT GCAGCCTACC GCGCTCTGC CGTGGAAATT

1101 CCGAGACCTC ATCACCAGG TTAAGATCAA GGTCTTTTCA CCTGGCCCGC
 GGCTCTGGAG TAGTGGGTCC AATTCTAGTT CCAGAAAAGT GGACCGGGCG

1151 ATGGACACCC AGACCAGGTC CCCTACATCG TGACCTGGGA AGCCTTGGCT
 TACCTGTGGG TCTGGTCCAG GGGATGTAGC ACTGGACCCT TCGGAACCGA

1201 TTTGACCCCC CTCCCTGGGT CAAGCCCTTT GTACACCCTA AGCCTCCGCC
 AACTGGGGG GAGGACCCA GTTCGGGAAA CATGTGGGAT TCGGAGCGG

1251 TCCTCTTCTT CCATCCGCC CGTCTCTCCC CTTGAACCT CCTCGTTCGA
 AGGAGAAGGA GGTAGCGGG GCAGAGAGG GGAAGTTGGA GGAGCAAGCT

1301 CCCCOCCTCG ATCCTCCCTT TATCCAGCCC TCACTCCTTC TCTAGGCGCC
 GGGGCGGAGC TAGGAGGGAA ATAGGTCGGG AGTGAGGAAG AGATCCGCGG

1351 GGCCGCTCTA GCCCATTAAAT ACGACTCACT ATAGGGCGAT TCGAACACCA
 CCGCGGAGAT CGGGTAATTA TGCTGAGTGA TATCCCGCTA AGCTTGTGGT

1401 TGCACCATCA TCATCATCAC GTCGACTATA AAGATGAGGA CCTCGAGATG
 ACCTGGTAGT AGTAGTAGTG CAGCTGATAT TTCTACTCCT GGAGCTCTAC

1451 GCGGTGATTA CGGATTCACT GCGGTCGTC GCGGCACCG ATCGCCCTTC
 CCGCACTAAT GCCTAAGTGA CCGGCAGCAC CCGGCGTGGC TAGCGGGAAG

1501 CCAACAGTTA CCGAGCCTGA ATGGCGAATG GCGCTTTGCC TGGTTCCGG
 GGTGTCAAT GCGTCGGACT TACCGCTTAC CCGGAAACGG ACCAAAGGCC

1551 CACCAGAAGC GGTGCCGAA AGCTGGCTGG AGTGCGATCT TCCTGAGGCC
 GTGGTCTTCG CCACGGCCTT TCGACCGACC TCACGCTAGA AGGACTCCGG

1601 GATACTGTGC TCGTCCCCTC AACTGGCAG ATGCACGGTT ACGATGCGCC
 CTATGACAGC AGCAGGGGAG TTTGACCGTC TACGTGCCAA TGCTACGCGG

1651 CATCTACACC AACGTGACCT ATCCATTAC GGTCAATCCG CCGTTTGTTC
 GTAGATGTGG TTGCACTGGA TAGGGTAATG CCAAGTAGGC GGCAACAAG

1701 CCACGGAGAA TCCGACGGGT TGTTACTCGC TCACATTTAA TGTTGATGAA
 GGTGCCTCTT AGGCTGCCCA ACAATGAGCG AGTGTAAATT ACAACTACTT

1751 AGCTGGCTAC AGGAAGGCCA GACGCGAAPT ATTTTGTATG GCGTTAACTC
 TCGACCGATG TCCTCCGGT CTGCGCTTAA TAAAACTAC CGCAATTGAG

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1851 GTCGTTTGCC GTCTGAATTT GACCTGAGCG CATTTTACG CGCCGGAGAA
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1951 AGATCAGGAT ATGTGGCGGA TGAGCGGCAT TTTCCGTGAC GTCTCGTTGC
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2001 TGCATAAACC GACTACACAA ATCAGCGATT TCCATGTTGC CACTCGCTTT
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2101 CGAGTTGCGT GACTACCTAC GGGTAACAGT TTCTTTATGG CAGGGTGAAA
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2151 CGCAGGTCGC CAGCGGCACC GCGCCTTTCG GCGGTGAAAT TATCGATGAG
 GCGTCCAGCG GTGCGCGTGG CCGGAAAGC CGCCACTTTA ATAGCTACTC

2201 CGTGGTGGTT ATGCCGATCG CGTCACACTA CGTCTGAACG TCGAAAACCC
 GCACCACCAA TACGGCTAGC GCAGTGTGAT GCAGACTTGC AGCTTTTGGG

2251 GAAACTGTGG ACGCCCGAAA TCCCGAATCT CTATCGTGCG GTGGTTGAAC
 CTTTGACACC TCGCGGCTTT AGGGCTTAGA GATAGCACGC CACCACTTG

2301 TGCACACCGC CGACGGCAGC CTGATTGAAG CAGAAGCCTG CGATGTCCGT
 ACGTGTGGCG GCTGCCGTGC GACTAACTTC GTCTTCGGAC GCTACAGCCA

2351 TTCCGCGAGG TCGGGATTGA AAATGGTCTG CTGCTGCTGA ACGGCAAGCC
 AAGGCCTCC ACGCCTAACT TTTACCAGAC GACGACGACT TGCCGTTCCG

2401 GTTGCTGATT CGAGGCGTTA ACCGTCACGA GCATCATCCT CTGCATGGTC
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2451 AGGTCATGGA TGAGCAGACG ATGGTGCAGG ATATCCTGCT GATGAAGCAG
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2501 AACAACTTTA ACGCCGTGCG CTGTTCEGAT TATCCGAACC ATCCGCTGTG
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ATTGCTCGAG GACGTGACCT ACCACCGCGA CCTACCATTG GCGGACCGTT

3351 GCGGTGAAGT GCCTCTGGAT GTCGCTCCAC AAGGTAAACA GTTGATTGAA
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3401 CTGCCTGAAC TACCGCAGCC GGAGAGCGCC GGGCAACTCT GGCTCACAGT
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TGCGCATCAC GTTGGCTTGC GCTGGCGTAC CAGTCTTCGG CCCGTGTAGT

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3851 GATGCGGTGC TGATTACGAC CGCTCACGCG TGGCAGCATC AGGGGAAAAAC
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4151 CCGAGCGAAA ACGGTCTGCG CTGCGGGACG CGCGAATTGA ATTATGGCCC
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4301 ACATGGCTGA ATATCGACGG TTTCCATATG GGGATTGGTG GCGACGACTC
TGTACCGACT TATAGCTGCC AAAGGTATAC CCCTAACCCAC CGCTGCTGAG

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4401 ATTACCAGTT GGTCTGGTGT CAAAAAGAT CTGGAGGTGG TGGCAGCAGG
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CAGAAGAACT GCTCGTAAGG ATCCCCAGAA AGGGGAGAGC GGTTTCTTA

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5751  GGGCTGACCG  CTTCTCGTG  CTTTACGGTA  TCGCCGCTCC  CGATTCCGAG
      CCCGACTGGC  GAAGGAGCAC  GAAATGCCAT  AGCGGCGAGG  GCTAAGCGTC
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5801  CGCATCGCCT  TCTATCGCCT  TCTTGACGAG  TTCTTCTGAG  CGGGACTCTG
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6601  TTGCGTATTG  GCGCTCTTCC  GCTTCTCGC  TCACTGACTC  GCTGCGCTCG
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6751  GCCAGCAAAA  GGCCAGGAAC  CGTAAAAAGG  CCGCGTTGCT  GCGTTTTTTC
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7551  ATGCTTAATC  AGTGAGGCAC  CTATCTCAGC  GATCTGTCTA  TTTGTTTCA
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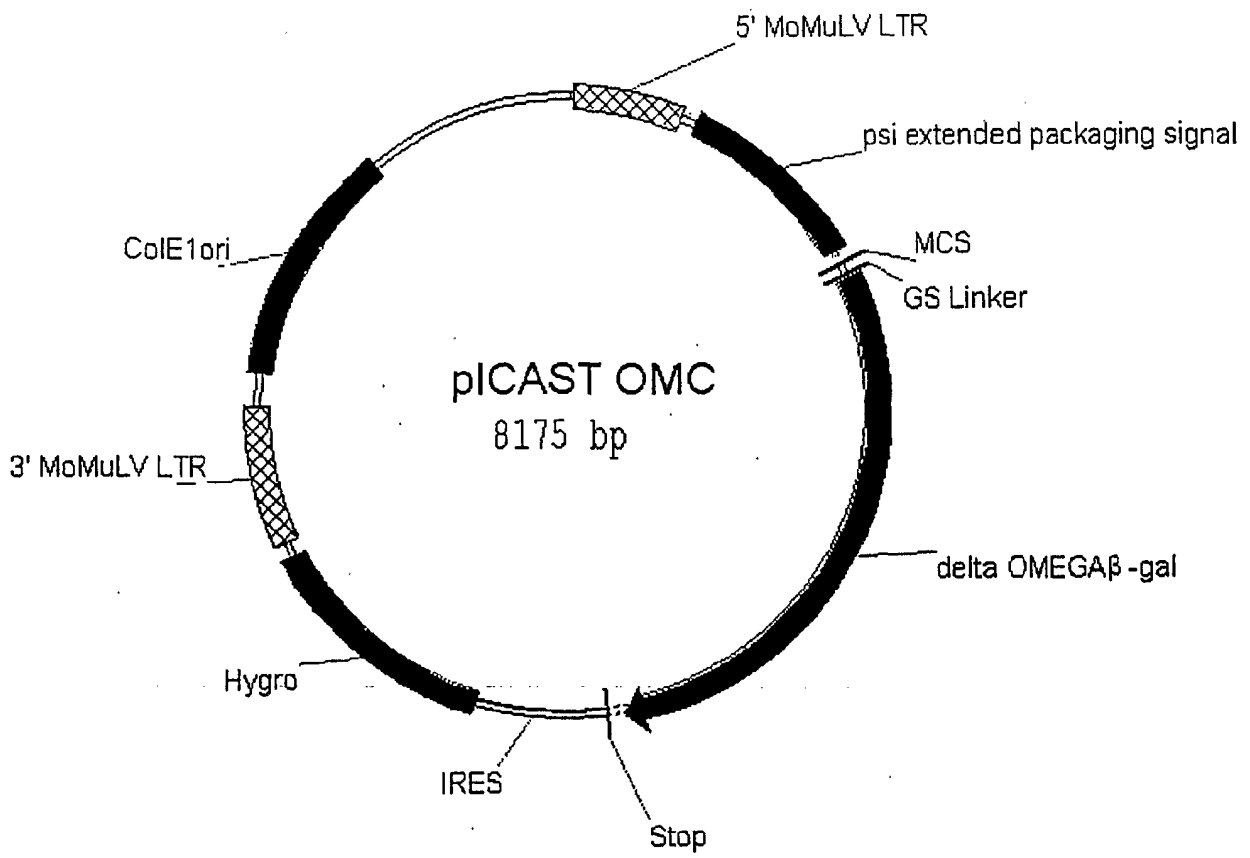


Figure 12A

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   CCTATAGACA CCATTCGTCA AGGACGGGGC CGAGTCCCGG TTCTTGTCTA
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151 GGTCCCCAGA TGCGGTCCAG CCCTCAGCAG TTTCTAGAGA ACCATCAGAT
   CCAGGGGTCT ACGCCAGGTC GGGAGTCGTC AAAGATCTCT TGGTAGTCTA
-----
201 GTTTCCAGGG TGCCCCAAGG ACCTGAAATG ACCCTGTGCC TTATTTGAAC
   CAAAGGTCCC ACGGGTTC ACAGGTTTAC TGGGACACGG AATAAACTTG
-----
251 TAACCAATCA GTTCGCTTCT CGTTCTGTT CGCGCGCTC TGCTCCCCGA
   ATTGTTAGT CAAGCGAAGA GCGAAGACAA GCGCGCAAG ACGAGGGGCT
-----
301 GCTCAATAAA AGAGCCACA ACCCCTCACT CGGGGCGCCA GTCCTCCGAT
   CGAGTTATTT TCTCGGGTGT TGGGGAGTGA GCCCGCGGT CAGGAGGCTA
-----
351 TGA CTGAGTGC GCCGGGTAC CGGTGTATCC AATAAACCTT CTTGCACTG
   ACTGACTCAG CCGGCCCATG GGCACATAGG TTATTTGGGA GAACGTCAAC
-----
401 CATCCGACTT GTGGTCTCGC TGTTCCTTGG GAGGGTCTCC TCTGAGTGAT
   GTAGGCTGAA CACCAGAGCG ACAAGGAACC CTCCAGAGG AGACTACTA
-----
451 TGA CTACCCG TCAGCGGGG TCTTTCATTT GGGGCTCCT CCGGGATCGG
   ACTGATGGC AGTCGCCCC AGAAAGTAAA CCCCAGGCA GGCCCTAGCC
-----
501 GAGACCCCTG CCCAGGGACC ACCGACCCAC CACCGGGAGG CAAGCTGGCC
   CTCTGGGGAC GGGTCCCTGG TGGCTGGGTG GTGGCCCTCC GTTCGACCGG
-----
551 AGCAACTTAT CTGTGTCTGT CCGATTGTCT AGTGCTATG ACTGATTTTA
   TCGTTGAATA GACACAGACA GGCTAACAGA TCACAGATC TGAATAAAT
-----
601 TGCGCCTGCG TCGGTACTAG TTAGCTAACT AGCTCTGTAT CTGGCGGACC
   ACGCGGACGC AGCCATGATC AATCGATTGA TCGAGACATA GACCGCCTGG
-----
651 CGTGGTGGAA CTGACGAGTT CTGAACACCC GGCCGCAACC CTGGGAGACG
   GCACCACCTT GACTGCTCAA GACTTGTGGG CCGGCGTTGG GACCCTCTGC
-----
701 TCCCAGGGAC TTTGGGGGCC GTTTTGTGG CCCGACCTGA GGAAGGGAGT
   AGGGTCCCTG AAACCCCGG CAAAACACC GGGCTGGACT CCTTCCCTCA
-----
751 CGATGTGGAA TCCGACCCCG TCAGGATATG TGGTCTCGT AGGAGACGAG
   GCTACACCTT AGGCTGGGGC AGTCTATAC ACCAAGACCA TCCTCTGCTC
-----
801 AACCTAAAAC AGTTCGCCG TCCGTCTGAA TTTTGTCTT CGGTTTGGAA
   TTGGATTTT TCAAGGGCGG AGGCAGACTT AAAACGAAA GCCAACCTT
-----
851 CCGAAGCCGC GCGTCTGTG TGCTGCAGCA TCGTCTGTG TTGTCTCTGT
   GGCTTCGGCG CGCAGAACAG ACGACGTCGT AGCAAGACAC AACAGAGACA
-----
901 CTGACTGTGT TTCTGTATTT GTCTGAAAT TAGGGCCAGA CTGTTACCAC
   GACTGACACA AAGACATAAA CAGACTTTTA ATCCCGGTCT GACAATGGTG
-----

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FIGURE 12B


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951  TCCCTTAAGT TTGACCTTAG GTAACCTGGAA AGATGTCGAG CGGCTCGCTC
    AGGGAATCA AACTGGAATC CATTGACCTT TCTACAGCTC GCCGAGCGAG
-----
1001  ACAACCAGTC GGTAGATGTC AAGAAGAGAC GTTGGGTTAC CTTCTGCTCT
    TGTGTGTCAG CCATCTACAG TTCTTCTCTG CAACCCAATG GAAGACGAGA
-----
1051  GCAGAATGGC CAACCTTTAA CGTCGGATGG CCGCGAGACG GCACCTTTAA
    CGTCTTACCG GTTGGAATTT GCAGCCTACC GCGCTCTGTC CGTGGAATTT
-----
1101  CCGAGACCTC ATCACCAGG TTAAGATCAA GGTCTTTTCA CCTGGCCCGC
    GGCTCTGGAG TAGTGGGTCC AATTCTAGTT CCAGAAAAGT GGACCGGGCG
-----
1151  ATGGACACCC AGACCAGGTC CCCTACATCG TGACCTGGGA AGCCTTGGCT
    TACCTGTGGG TCTGGTCCAG GGGATGTAGC ACTGGACCCT TCGGAACCGA
-----
1201  TTTGACCCCC CTCCTGGGT CAAGCCCTTT GTACACCCTA AGCCTCCGCC
    AAACCTGGGGG GAGGGACCCA GTTCGGGAAA CATGTGGGAT TCGGAGGCGG
-----
1251  TCCTCTTCCT CCATCCGCC CGTCTCTCCC CCTTGAACCT CCTCGTTCGA
    AGGAGAAGGA GGTAGGCGGG GCAGAGAGGG GGAACCTTGA GGAGCAAGCT
-----
1301  CCCCGCCTCG ATCCTCCCTT TATCCAGCCC TCACTCCTTC TCTAGGCGCC
    GGGGCGGAGC TAGGAGGGAA ATAGGTCGGG AGTGAGGAAG AGATCCGCGG
-----
1351  GGCCGCTCTA GCCCATTAA TCGACTCACT ATAGGGCGAT TCGAATCAGG
    CCGGCGAGAT CGGTAATTA TGCTGAGTGA TATCCCGCTA AGCTTAGTCC
-----
1401  CCTTGGCGCG CCGGATCCTT AATTAAGCGC AATTGGGAGG TGGCGGTAGC
    GGAACCGCGC GCCTAGGAA TTAATTCGCG TTAACCCCTC ACCGCCATCG
-----
1451  CTCGAGATGG GCGTGATTAC GGATTCACTG GCCGTCGTTT TACAACGTCG
    GAGCTCTACC CGCACTAATG CCTAAGTGAC CGGCAGCAA ATGTTGCAGC
-----
1501  TGACTGGGAA AACCTGGCG TTACCCAACT TAATCGCCTT GCAGCACATC
    ACTGACCCTT TTGGGACCGC AATGGGTTGA ATTAGCGGAA CGTCTGTAG
-----
1551  CCCCTTTCGC CAGCTGGCGT AATAGCGAAG AGGCCCGCAC CGATCGCCCT
    GGGGAAAGCG GTCGACCGCA TTATCGCTTC TCCGGGCGTG GCTAGCGGGA
-----
1601  TCCCAACAGT TACGAGCCT GAATGGCGAA TGGCGCTTTG CCTGGTTTCC
    AGGGTTGTCA ATGCGTCGGA CTTACCGCTT ACCGCGAAAC GGACCAAAGG
-----
1651  GGCACCAGAA GCGGTGCCGG AAAGCTGGCT GGAGTGGCAT CTTCTGAGG
    CCGTGGTCTT CGCCACGGCC TTTCGACCGA CCTCAGCTA GAAGGACTCC
-----
1701  CCGATACTGT CGTCGTCCCC TCAAACCTGGC AGATGCAOAG TTACGATGCG
    GGCTATGACA GCAGCAGGGG AGTTTGACCG TCTACGTGCC AATGCTACGC
-----
1751  CCCATCTACA CCAACGTGAC CTATCCCATC ACGGTCAATC CGCCGTTTGT
    GGGTAGATGT GGTGCACTG GATAGGGTAA TGCCAGTTAG GCGGCAACA
-----
1801  TCCCACGGAG AATCCGACGG GTTGTACTC GCTCACATTT AATGTTGATG
    AGGGTGCCTC TTAGGCTGCC CAACAATGAG CGAGTGAAA TTACAACCTA
-----
1851  AAAGCTGGCT ACAGGAAGGC CAGACGCGAA TTATTTTGA TGGCGTTAAC
    TTTCGACCGA TGCTTCCG GTCTGCGCTT AATAAAAAC ACCGCAATTG
-----

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1901 TCGGCGTTTC ATCTGTGGTG CAACGGGCGC TGGGTCGGTT ACGGCCAGGA
 AGCCGCAAAAG TAGACACCAC GTTGCCCGCG ACCCAGCCAA TGCCGGTCCT

1951 CAGTCGTTTG CCGTCTGAAT TTGACCTGAG CGCATTTTAA CGCGCCGAG
 GTCAGCAAAC GGCAGACTTA AACTGGACTC GCGTAAAAAT GCGCGGCCTC

2001 AAAACCGCCT CGCGGTGATG GTGCTGCGCT GGAGTGACGG CAGTTATCTG
 TTTTGGCGGA GCGCCACTAC CACGACGCGA CCTCACTGCC GTCAATAGAC

2051 GAAGATCAGG ATATGTGGCG GATGAGCGGC ATTTTCCGTG ACGTCTCGTT
 CTTCTAGTCC TATACACCGC CTA CTACTCGCCG TAAAAGGCAC TGCAGAGCAA

2101 GCTGCATAAA CCGACTACAC AAATCAGCGA TTTCCATGTT GCCACTCGCT
 CGACGTATTT GGCTGATGTG TTTAGTCGCT AAAGGTACAA CCGTGAGCGA

2151 TTAATGATGA TTTAGCCGCG GCTGFACTGG AGGCTGAAGT TCAGATGTGC
 AATTACTACT AAAGTCGGCG CGACATGACC TCCGACTTCA AGTCTACAGC

2201 GGGGAGTTGC GTGACTACCT ACGGGTAACA GTTTCCTTAT GGCAGGGTGA
 CCGCTCAACG CACTGATGGA TGCCATTGT CAAAGAAATA CCGTCCCCT

2251 AACGCAGGTC GCCAGCGGCA CCGCGCCTTT CGGCGGTGAA ATTATCGATG
 TTGCGTCCAG CGGTCGCCGT GGCGCGGAAA GCCGCCACTT TAATAGCTAC

2301 AGCGTGGTGG TTATGCCGAT CGCGTCACAC TACGTCTGAA CGTCGAAAAC
 TCGCACCACC AATACGGCTA GCGCAGTGTG ATGCAGACTT GCAGCTTTTG

2351 CCGAAACTGT GGAGCGCCGA AATCCCGAAT CTCTATCGTG CCGTGGTTGA
 GGCTTTGACA CCTCGCGGCT TTAGGGCTTA GAGATAGCAC GCCACCACT

2401 ACTGCACACC GCCGACGGCA CGCTGATTGA AGCAGAAGCC TGCGATGTGC
 TGACGTGTGG CGGCTGCCGT GCGACTAACT TCGTCTTCGG ACGCTACAGC

2451 GTTCCCGCA GGTGCGGATT GAAAATGGTC TGCTGCTGCT GAACGGCAAG
 CAAAGGCGCT CCACGCCTAA CTTTTACCAG ACGACGACGA CTTGCCGTTT

2501 CCGTTGCTGA TTCGAGGCGT TAACCGTCAC GAGCATCATC CTCTGCATGG
 GGCAACGACT AAGCTCCGCA ATTGGCAGTG CTCGTAGTAG GAGACGTACC

2551 TCAGGTCAAG GATGAGCAGA CGATGGTGCA GGATATCCTG CTGATGAAGC
 AGTCCAGTAC CTACTCGTCT GCTACCACGT CCTATAGGAC GACTACTTCG

2601 AGAACAACCT TAACGCCGTG CGCTGTTTCG ATTATCCGAA CCATCCGCTG
 TCTTGTTGAA ATTGCCGCAC GCGACAAGCG TAATAGGCTT GGTAGGCGAC

2651 TGGTACACGC TGTGCGACCG CTACGGCCTG TATGTGGTGG ATGAAGCCAA
 ACCATGTGCG ACACGCTGGC GATGCCGGAC ATACACCACC TACTTCGTT

2701 TATTGAAACC CACGGCATGG TGCCAATGAA TCGTCTGACC GATGATCCGC
 ATAACCTTGG GTGCCGTACC ACGGTTACTT AGCAGACTGG CTACTAGGCG

2751 GCTGGCTACC GGCATGAGC GAACGCGTAA CGCGAATGGT GCAGCGCGAT
 CGACCGATGG CCGCTACTCG CTTGCGCATT GCGCTTACCA CGTCGCGCTA

2801 CGTAATCACC CGAGTGTGAT CATCTGGTCG CTGGGGAATG AATCAGGCCA
 GCATTAGTGG GCTCACACTA GTAGACCAGC GACCCCTTAC TTAGTCCGGT

```

2851  CGGCGCTAAT  CACGACGCGC  TGTATCGCTG  GATCAAATCT  GTCGATCCTT
      GCCGCGATTA  GTGCTGCGCG  ACATAGCGAC  CTAGTTTAGA  CAGCTAGGAA
-----
2901  CCCGCCCGGT  GCAGTATGAA  GGCGGCGGAG  CCGACACCAC  GGCCACCGAT
      GGGCGGGCCA  CGTCATACTT  CCGCCGCCCTC  GGCTGTGGTG  CCGGTGGCTA
-----
2951  ATTATTTGCC  CGATGTACGC  GCGCGTGGAT  GAAGACCAGC  CCTTCCCGGC
      TAATAAACGG  GCTACATGCG  CGCGCACCTA  CTTCTGGTCG  GGAAGGGCCG
-----
3001  TGTGCCGAAA  TGGTCCATCA  AAAAATGGCT  TTCGCTACCT  GGAGAGACGC
      ACACGGCTTT  ACCAGGTAGT  TTTTACCGA  AAGCGATGGA  CCTCTCTGCG
-----
3051  GCCCCGTGAT  CCTTTGCGAA  TACGCCACGC  CGATGGGTAA  CAGTCTTGGC
      CGGGCGACTA  GGAACCGCTT  ATGCGGGTGC  GCTACCCATT  GTCAGAACCG
-----
3101  GGTTCGCTA  AATACTGGCA  GGCGTTTCGT  CAGTATCCCC  GTTTACAGGG
      CCAAAGCGAT  TTATGACCGT  CCGCAAAGCA  GTCATAGGGG  CAAATGTCCC
-----
3151  CGGCTTCGTC  TGGGACTGGG  TGGATCAGTC  GCTGATTAAT  TATGATGAAA
      GCCGAAGCAG  ACCCTGACCC  ACCTAGTCAG  CGACTAATTT  ATACTACTTT
-----
3201  ACGGCAACCC  GTGGTCGGCT  TACGGCGGTG  ATTTGGGCGA  TACGCCGAAC
      TGCCGTTGGG  CACCAGCCGA  ATGCCGCCAC  TAAAACCGCT  ATGCGGCTTG
-----
3251  GATCGCCAGT  TCTGTATGAA  CGGTCTGGTC  TTTGCCGACC  GCACGCCGCA
      CTAGCGGTCA  AGACATACTT  GCCAGACCAG  AAACGGCTGG  CGTGCGGCGT
-----
3301  TCCAGCGCTG  ACGGAAGCAA  AACACCAGCA  GCAGTTTTTC  CAGTTCCGTT
      AGGTCCGGAC  TGCCTTCGTT  TTGTGGTGGT  CGTCAAAAAG  GTC AAGGCAA
-----
3351  TATCCGGGCA  AACCATCGAA  GTGACCAGCG  AATACCTGTT  CCGTCATAGC
      ATAGGCCCGT  TTGGTAGCTT  CACTGGTTCG  TTATGGACAA  GGCAGTATCG
-----
3401  GATAACGAGC  TCCTGCACTG  GATGGTGGCG  CTGGATGGTA  AGCCGCTGGC
      CTATTGCTCG  AGGACGTGAC  CTACCACCGC  GACCTACCAT  TCGGCGACCG
-----
3451  AAGCGGTGAA  GTGCCTCTGG  ATGTCGCTCC  ACAAGGTAAA  CAGTTGATTG
      TTCGCCACTT  CACGGAGACC  TACAGCGAGG  TGTTCATTTT  GTC AACTAAC
-----
3501  AACTGCCTGA  ACTACCGCAG  CCGGAGAGCG  CCGGGCAACT  CTGGCTCACA
      TTGACGGACT  TGATGGCGTC  GGCCTCTCGC  GGCCCGTTGA  GACCGAGTGT
-----
3551  GTACGCGTAG  TGCAACCGAA  CGCGACCGCA  TGGTCAGAAG  CCGGGCACAT
      CATGCGCATC  ACGTTGGCTT  GCGCTGGCGT  ACCAGTCTTC  GGCCCGTGTA
-----
3601  CAGCGCCTGG  CAGCAGTGGC  GTCTGGCGGA  AAACCTCAGT  GTGACGCTCC
      GTCGCGGACC  GTCGTCACCG  CAGACCGCCT  TTTGGAGTCA  CACTGCGAGG
-----
3651  CCGCCGCGTC  CCACGCCATC  CCGCATCTGA  CCACCAGCGA  AATGGATTTT
      GGGCGCGCAG  GGTGCGGTAG  GGCCTAGACT  GGTGGTTCGT  TTACCTAAAA
-----
3701  TGCATCGAGC  TGGTAATAAA  GCGTTGGCAA  TTTAACCGCC  AGTCAGGCTT
      ACGTAGCTCG  ACCCATTATT  CGCAACCGTT  AAATTGGCGG  TCAGTCCGAA
-----
3751  TCTTTCACAG  ATGTGGATTG  GCGATAAAAA  ACAACTGCTG  ACGCCGCTGC
      AGAAAGTGTC  TACACCTAAC  CGCTATTTTT  TGTGACGAC  TCGGCGGACG
-----

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3801 GCGATCAGTT CACCCGTGTC GATAGATCTG AACAGAAACT CATTTCGGAA
 CGCTAGTCAA GTGGGCACAG CTATCTAGAC TTGTCTTTGA GTAAAGGCTT

3851 GAAGACCTAG TCGACCATCA TCATCATCAT CACCGGTAAT AATAGGTAGA
 CTTCTGGATC AGCTGGTAGT AGTAGTAGTA GTGGCCATTA TTATCCATCT

3901 TAAGTGACTG ATTAGATGCA TTTGACTAG ATCCCTCGAC CAATTCCGGT
 ATTCACTGAC TAATCTACGT AAAGCTGATC TAGGGAGCTG GTTAAGGCCA

3951 TATTTTCCAC CATATTGCCG TCTTTTGGCA ATGTGAGGGC CCGGAAACCT
 ATAAAAGGTG GTATAACGGC AGAAAACCGT TACACTCCCG GGCCTTTGGA

4001 GGCCCTGTCT TCTFGACGAG CATTCTAGG GGTCTTTCCC CTCTCGCAA
 CCGGGACAGA AGAACTGCTC GTAAGGATCC CCAGAAAGGG GAGAGCGGTT

4051 AGGAATGCAA GGTCTGTTGA ATGTCGTGAA GGAAGCAGTT CCTCTGGAAG
 TCCTTACGTT CCAGACAAC TACAGCACTT CCTTCGTCAA GGAGACCTTC

4101 CTTCTTGAAG ACAAACAACG TCTGTAGCGA CCCTTTGCAG GCAGCGGAAC
 GAAGAACTTC TGTTTGTTGC AGACATCGCT GGGAAACGTC CGTCGCCTTG

4151 CCCCCACCTG GCGACAGGTG CCTCTGCGGC CAAAAGCCAC GTGTATAAGA
 GGGGGTGGAC CGCTGTCCAC GGAGACGCCG GTTTTCGGTG CACATATTCT

4201 TACACCTGCA AAGGCGGCAC AACCCAGTG CCACGTTGTG AGTTGGATAG
 ATGTGGACGT TTCCGCCGTG TTGGGGTCAC GGTGCAACAC TCAACCTATC

4251 TTGTGGAAAG AGTCAAATGG CTCTCCTCAA GCGTATTCAA CAAGGGGCTG
 AACACCTTTC TCAGTTTACC GAGAGGAGTT CGCATAAGTT GTTCCCCGAC

4301 AAGGATGCCC AGAAGGTACC CCATTGTATG GGATCTGATC TGGGGCCTCG
 TTCCTACGGG TCTTCCATGG GGTACATAC CCTAGACTAG ACCCCGGAGC

4351 GTGCACATGC TTTACATGTG TTTAGTCGAG GTTAAAAAAC GTCTAGGCCC
 CACGTGTACG AAATGTACAC AAATCAGCTC CAATTTTTTG CAGATCCGGG

4401 CCCGAACCAC GGGGACGTGG TTTTCCTTTG AAAAACACGA TGATAATACC
 GGGCTTGGTG CCCCTGCACC AAAAGGAAAC TTTTTGTGCT ACTATTATGG

4451 ATGAAAAAGC CTGAACTCAC CGCGACGTCT GTCGAGAAGT TTCTGATCGA
 TACTTTTTTC GACTTGAGTG GCGCTGCAGA CAGCTCTTCA AAGACTAGCT

4501 AAAGTTGAC AGCGTCTCCG ACCTGATGCA GCTCTCGGAG GGCGAAGAAT
 TTTCAAGCTG TCGCAGAGGC TGGACTACGT CGAGAGCCTC CCGCTTCTTA

4551 CTCGTGCTTT CAGCTTCGAT GTAGGAGGGC GTGGATATGT CCTGCGGGTA
 GAGCACGAAA GTCGAAGCTA CATCCTCCCG CACCTATACA GGACGCCCAT

4601 AATAGCTGCG CCGATGTTTT CTACAAAGAT CGTTATGTTT ATCGGCACTT
 TTATCGACGC GGCTACCAA GATGTTTCTA GCAATACAAA TAGCCGTGAA

4651 TGCATCGGCC GCGCTCCCGA TTCCGGAAGT GCTTGACATT GGGGAATTA
 ACGTAGCCGG CGCGAGGGCT AAGGCCTTCA CGAACTGTAA CCCCTTAAAT

4701 GCGAGAGCCT GACCTATTGC ATCTCCCGCC GTGCACAGGG TGTCACGTTG
 CGCTCTCGGA CTGGATAACG TAGAGGGCGG CACGTGTCCC ACAGTGCAAC

4751 CAAGACCTGC CTGAAACCGA AC'TGCCCGCT GTTCTGCAGC CGGTCCGGGA
 GTTCTGGACG GACTTTGGCT TGACGGGCGA CAAGACGTGC GCCAGCGCCT

4801 GGCCATGGAT GCGATCGCTG CGGCCGATCT TAGCCAGACG AGCGGGTTCG
 CCGGTACCTA CGCTAGCGAC GCCGGCTAGA ATCGGTCTGC TCGCCCAAGC

4851 GCCCATTCCG ACCGCAAGGA ATCGGTCAAT ACACTACATG GCGTGATTTT
 CGGGTAAGCC TGGCGTTCCT TAGCCAGTTA TGTGATGTAC CGCACTAAG

4901 ATATGCGCGA TTGCTGATCC CCATGTGTAT CACTGGCAAA CTGTGATGGA
 TATACGCGCT AACGACTAGG GGTACACATA GTGACCGTTT GACTACTACCT

4951 CGACACCGTC AGTGCCTCCG TCGCGCAGGC TCTCGATGAG CTGATGCTTT
 GCTGTGGCAG TCACGCAGGC AGCGGCTCCG AGAGCTACTC GACTACGAAA

5001 GGCCCGAGGA CTGCCCCGAA GTCCCGCACC TCGTGCACGC GGATTTCGGC
 CCCGGCTCCT GACGGGGCTT CAGGCCGTGG AGCACGTGCG CCTAAAGCCG

5051 TCCAACAATG TCCTGACGGA CAATGGCCGC ATAACAGCGG TCATTGACTG
 AGGTTGTTAC AGGACTGCCT GTTACCGGGG TATTGTGCGC AGTAACTGAC

5101 GAGCGAGGCG ATGTTCCGGG ATTCCCAATA CGAGGTGCGC AACATCTTCT
 CTCGCTCCGC TACAAGCCCC TAAGGTTAT GCTCCAGCGG TTGTAGAAGA

5151 TCTGGAGGCC GTGGTTGGCT TGTATGGAGC AGCAGACGCG CTACTIONGAG
 AGACCTCCGG CACCAACCGA ACATACCTCG TCGTCTGCGC GATGAAGCTC

5201 CGGAGGCATC CGGAGCTTGC AGGATCGCCG CGGCTCCGGG CGTATATGCT
 GCCTCCGTAG GCCTCGAACG TCCTAGCGGC GCCGAGGCC GCATATACGA

5251 CCGCATTGGT CTGACCAAC TCTATCAGAG CTGTTGAC GGCAATTCG
 GCGTAACCA GAACTGTTG AGATAGTCTC GAACCAACTG CCGTTAAAGC

5301 ATGATGCAGC TTGGGCGCAG GGTGATGCG ACGCAATCGT CCGATCCGGA
 TACTACGTGC AACCCGCGTC CCAGCTACGC TCGGTTAGCA GGCTAGGCCT

5351 GCCGGGACTG TCGGGCGTAC ACAAATCGCC CGCAGAAGCG CGGCCGTCTG
 CGGCCCTGAC AGCCCGCATG TGTTTAGCGG GCGTCTTCGC GCCGGCAGAC

5401 GACCGATGGC TGTGTAGAAG TACTCGCCGA TAGTGAAAC CGACGCCCA
 CTGGCTACCG ACACATCTTC ATGAGCGGCT ATCACCTTTG GCTGCGGGGT

5451 GCACTCGTCC GAGGGCAAAG GAATAGAGTA GATGCCGACC GGGATCTATC
 CGTGAGCAGG CTCCGTTTC CTTATCTCAT CTACGGCTGG CCTAGATAG

5501 GATAAATAA AAGATTTTAT TTAGTCTCCA GAAAAAGGGG GGAATGAAAG
 CTATTTTATT TTCTAAAATA AATCAGAGGT CTTTTTCCCC CCTTACTTTC

5551 ACCCCACCTG TAGGTTTGGC AAGCTAGCTT AAGTAACGCC ATTTTGCAAG
 TGGGGTGGAC ATCCAACCG TTCGATCGAA TTCATTGCGG TAAAACGTTT

5601 GCATGGAAAA ATACATAACT GAGAATAGAG AAGTTCAGAT CAAGGTCAGG
 CGTACCTTTT TATGTATTGA CTCTTATCTC TTCAAGTCTA GTTCCAGTCC

5651 AACAGATGGA ACAGCTGAAT ATGGGCCAAA CAGGATATCT GTGGTAAGCA
 TTGTCTACCT TGTCGACTTA TACCCGGTTT GTCCTATAGA CACCATTCTG

5701 GTTCTGCC CGGCTCAGGG CCAAGAACAG ATGGAACAGC TGAATATGGG
CAAGGACGGG GCCGAGTCCC GGTTCTTGTC TACCTTGTCG ACTTATACCC

5751 CCAAACAGGA TATCTGTGGT AAGCAGTTCC TGCCCCGGCT CAGGGCCAAG
GGTTTGICCT ATAGACACCA TTCGTCAAGG ACGGGGCCGA GTCCCCGGTC

5801 AACAGATGGT CCCCAGATGC GGTCCAGCCC TCAGCAGTTT CTAGAGAACC
TTGTCTACCA GGGGTCTACG CCAGGTCGGG AGTCGTCAA GATCTCTTGG

5851 ATCAGATGTT TCCAGGGTGC CCCAAGGACC TGAAATGACC CTGTGCCTTA
TAGTCTACAA AGGTCCCACG GGGTTCCTGG ACTTTACTGG GACACGGAAT

5901 TTTGAACTAA CCAATCAGTT CGCTTCTCGC TTCTGTTGGC GCGTCTCTGC
AAACTTGATT GGTTAGTCAA GCGAAGAGCG AAGACAAGCG CGCGAAGACG

5951 TCCCCGAGCT CAATAAAGA GCCCACAACC CCTCACTCGG GCGCCAGTC
AGGGGCTCGA GTTATTTTCT CCGGTGTTGG GGAGTGAGCC CCGCGGTCAG

6001 CTCCGATTGA CTGAGTCGCC CGGGTACCGG TGTATCCAAT AAACCCTCTT
GAGGCTAACT GACTCAGCGG GCCCATGGGC ACATAGGTTA TTTGGGAGAA

6051 GCAGTTGCAT CCGACTTGTG GTCTCGCTGT TCCTGGGAG GGTCTCCTCT
CGTCAACGTA GGCTGAACAC CAGAGCGACA AGGAACCCTC CCAGAGGAGA

6101 GAGTGATTGA CTACCCGTCG CCGGGGGTCT TTCATTATG CAGCATGTAT
CTCACTAACT GATGGGCAGT CGCCCCAGA AAGTAAGTAC GTCGTACATA

6151 CAAAATTAAT TTGGTTTTTT TTCTTAAGTA TTTACATTAA ATGGCCATAG
GTTTTAATTA AACCAAAAAA AAGAATTCAT AAATGTAATT TACCGGTATC

6201 TTGCATTAAT GAATCGGCCA ACGCGCGGGG AGAGGCGGTT TGCGTATTGG
AACGTAATTA CTTAGCCGGT TGCGCGCCCC TCTCCGCCAA ACGCATAACC

6251 CGCTTTCCG CTTCCTCGCT CACTGACTCG CTGCGCTCGG TCGTTCGGCT
GCGAGAAGGC GAAGGAGCGA GTGACTGAGC GACGCGAGCC AGCAAGCCGA

6301 GCGGCGAGCG GTATCAGCTC ACTCAAAGGC GGTAAATACGG TTATCCACAG
CGCCGCTCGC CATAGTCGAG TGAGTTCCG CCATTATGCC AATAGGTGTC

6351 AATCAGGGGA TAACGCAGGA AAGAACATGT GAGCAAAGG CCAGCAAAG
TTAGTCCCCT ATGCGTCCTT TTCTTGTAACA CTCGTTTTCC GGTCTGTTTC

6401 GCCAGGAACC GTAAAAAGGC CGCGTTGCTG GCGTTTTTCC ATAGGCTCCG
CGGTCTTGG CATTTTTCCG GCGCAACGAC CGCAAAGG TATCCGAGGC

6451 CCCCCCTGAC GAGCATCACA AAAATCGACG CTCAAGTCAG AGGTGGCGAA
GGGGGACTG CTCGTAGTGT TTTTAGCTGC GAGTTCAGTC TCCACCGCTT

6501 ACCCGACAGG ACTATAAAGA TACCAGGCGT TTCCCCTGG AAGCTCCCTC
TGGGCTGTCC TGATATTTCT ATGGTCCGCA AAGGGGGACC TTCGAGGGAG

6551 GTGCGCTCTC CTGTTCCGAC CCTGCGCTT ACCGGATACC TGTCCGCCTT
CACGCGAGAG GACAAGGCTG GGACGGCGAA TGGCCTATGG ACAGGCGGAA

6601 TCTCCCTCG GGAAGCGTGG CGCTTTCTCA TAGCTCACGC TGTAGGTATC
AGAGGGAAGC CCTTCGCACC GCGAAAGAGT ATCGAGTGCG ACATCCATAG

6651 TCAGTTCGGT GTAGGTCGTT CGCTCCAAGC TGGGCTGIGT GCACGAACCC
 AGTCAAGCCA CATCCAGCAA GCGAGGTTCC ACCCGACACA CGTGCTTGGG

6701 CCGTTCAGC CCGACCGTG CGCCTTATCC GGTAACATC GTCTTGAGTC
 GGGCAAGTCG GGCTGGCGAC GCGGAATAGG CCATTGATAG CAGAACTCAG

6751 CAACCCGGTA AGACACGACT TATCGCCACT GGCAGCAGCC ACTGGTAACA
 GTTGGGCCAT TCTGTGCTGA ATAGCGGTGA CCGTCGTCGG TGACCATTGT

6801 GGATTAGCAG AGCGAGGTAT GTAGGCGGTG CTACAGAGTT CTTGAAGTGG
 CCTAATCGTC TCGCTCCATA CATCCGCCAC GATGTCTCAA GAACTTCACC

6851 TGGCCTAACT ACGGCTACAC TAGAAGAACA GTATTTGGTA TCTGCGCTCT
 ACCGGATTGA TGCCGATGTG ATCTTCTTGT CATAAACCAT AGACGGGAGA

6901 GCTGAAGCCA GTTACCTTCG GAAAAGAGT TGGTAGCTCT TGATCCGGCA
 CGACTTCGGT CAATGGAAGC CTTTTTCTCA ACCATCGAGA ACTAGGCCGT

6951 AACAAACCAC CGCTGGTAGC GGTGGTTTTT TTGTTTGCAA GCAGCAGATT
 TTGTTTGGTG GCGACCATCG CCACCAAAAA AACAAACGTT CGTCGTCTAA

7001 ACGCGCAGAA AAAAAGGATC TCAAGAAGAT CCTTTGATCT TTTCTACGGG
 TGC GCGTCTT TTTTCCCTAG AGTTCTTCTA GGAACTAGA AAAGATGCCC

7051 GTCTGACGCT CAGTGAACG AAAACTCAG TTAAGGGATT TTGGTCATGA
 CAGACTGCCA GTCACCTTGC TTTTGAGTGC AATCCCTAA AACCAGTACT

7101 GATTATCAA AAGGATCTTC ACCTAGATCC TTTTAAATTA AAAATGAAGT
 CTAATAGTTT TTCCTAGAAG TGGATCTAGG AAAATTTAAT TTTTACTTCA

7151 TTGCGGCCGC AAATCAATCT AAAGTATATA TGAGTAACT TGGTCTGACA
 AACGCCGGCG TTTAGTTAGA TTTTATATAT ACTCATTTGA ACCAGACTGT

7201 GTTACCAATG CTTAATCAGT GAGGCACCTA TCTCAGCGAT CTGTCTATTT
 CAATGGTTAC GAATTAGTCA CTCCGTGGAT AGAGTCGCTA GACAGATAAA

7251 CGTTCATCCA TAGTGCCTG ACTCCCGTGC GTGTAGATAA CTACGATACG
 GCAAGTAGGT ATCAACGGAC TGAGGGGCAG CACATCTATT GATGCTATGC

7301 GGAGGGCTTA CCATCTGGCC CCACTGCTGC AATGATACCG CGAGACCCAC
 CCTCCCGAAT GTTAGACCGG GGTCACGACG TTAATATGGC GCTCTGGGTG

7351 GCTCACCGGC TCCAGATTTA TCAGCAATAA ACCAGCCAGC CGGAAGGGCC
 CGAGTGGCCG AGGTCTAAAT AGTCGTTATT TGGTCGGTCG GCCTTCCCGG

7401 GAGCGCAGAA GTGGTCTGC AACTTTATCC GCCTCCATCC AGTCTATTAA
 CTCGCTCTT CACCAGGACG TTGAAATAGG CGGAGGTAGG TCAGATAATT

7451 TTGTTGCCGG GAAGCTAGAG TAAGTAGTTC GCCAGTTAAT AGTTTGCGCA
 AACAAACGGC CTTCGATCTC ATTCATCAAG CCGTCAATTA TCAAACGCGT

7501 ACGTTGTTGC CATTGCTACA GGCATCGTGG TGTACGCTC GTCGTTTGGT
 TGCAACAACG GTAACGATGT CCGTAGCACC ACAGTGCAG CAGCAAACCA

7551 ATGGCTTCAT TCAGCTCCGG TTCCAACGA TCAAGGCGAG TTACATGATC
 TACCGAAGTA AGTCGAGGCC AAGGTTGCT AGTTCCGCTC AATGTACTAG

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7601  CCCCATGTTG TGCAAAAAG CGGTTAGCTC CTTCGGTCCT CCGATCGTTG
      GGGGTACAAC ACGTTTTTTC GCCAATCGAG GAAGCCAGGA GGCTAGCAAC
-----
7651  TCAGAAGTAA GTTGGCCGCA GTGTTATCAC TCATGGTTAT GGCAGCACTG
      AGTCTTCATT CAACCGGCGT CACAATAGTG AGTACCAATA CCGTCGTGAC
-----
7701  CATAATTCTC TTACTGTCAT GCCATCCGTA AGATGCTTTT CTGTGACTGG
      GTATTAAGAG AATGACAGTA CGGTAGGCAT TCTACGAAAA GACACTGACC
-----
7751  TGAGTACTCA ACCAAGTCAT TCTGAGAATA GTGTATGCGG CGACCGAGTT
      ACTCATGAGT TGGTTCAGTA AGACTCTTAT CACATACGCC GCTGGCTCAA
-----
7801  GCTCTTGCCC GGCCTCAATA CGGGATAATA CCGCGCCACA TAGCAGAACT
      CGAGAACGGG CCGCAGTTAT GCCCTATTAT GGC CGGCTGT ATCGTCTTGA
-----
7851  TTRAAAGTGC TCATCATTGG AAAACGTTCT TCGGGGCGAA AACTCTCAAG
      AATTTTCACG AGTAGTAACC TTTTGCAAGA AGCCCCGCTT TTGAGAGTTC
-----
7901  GATCTTACCG CTGTTGAGAT CCAGTTCGAT GTAACCCACT CGTGCACCCA
      CTAGAATGGC GACAACCTA GGTCAAGCTA CATTGGGTGA GCACGTGGGT
-----
7951  ACTGATCTTC AGCATCTTTT ACTTTCACCA GCGTTTCTGG GTGAGCAAAA
      TGA CTAGAAG TCGTAGAAAA TGAAGTGGT CGCAAAGACC CACTCGTTTT
-----
8001  ACAGGAAGGC AAAATGCCGC AAAAAAGGGA ATAAGGGCGA CACGGAAATG
      TGTCTTCCG TTTTACGGCG TTTTTCCCT TATTCGCGCT GTGCCTTAC
-----
8051  TTGAATACTC ATACTCTTCC TTTTCAATA TTATTGAAGC ATTTATCAGG
      AACTTATGAG TATGAGAAGG AAAAAGTTAT AATAACTTCG TAAATAGTCC
-----
8101  GTTATTGTCT CATGAGCGGA TACATATTG AATGTATTTA GAAAAATAAA
      CAATAACAGA GTACTCGCCT ATGTATAAAC TTACATAAAT CTTTTTATT
-----
8151  CAAATAGGGG TTCCGCGCAC ATTTC
      GTTTATCCCC AAGGCGCGTG TAAAG
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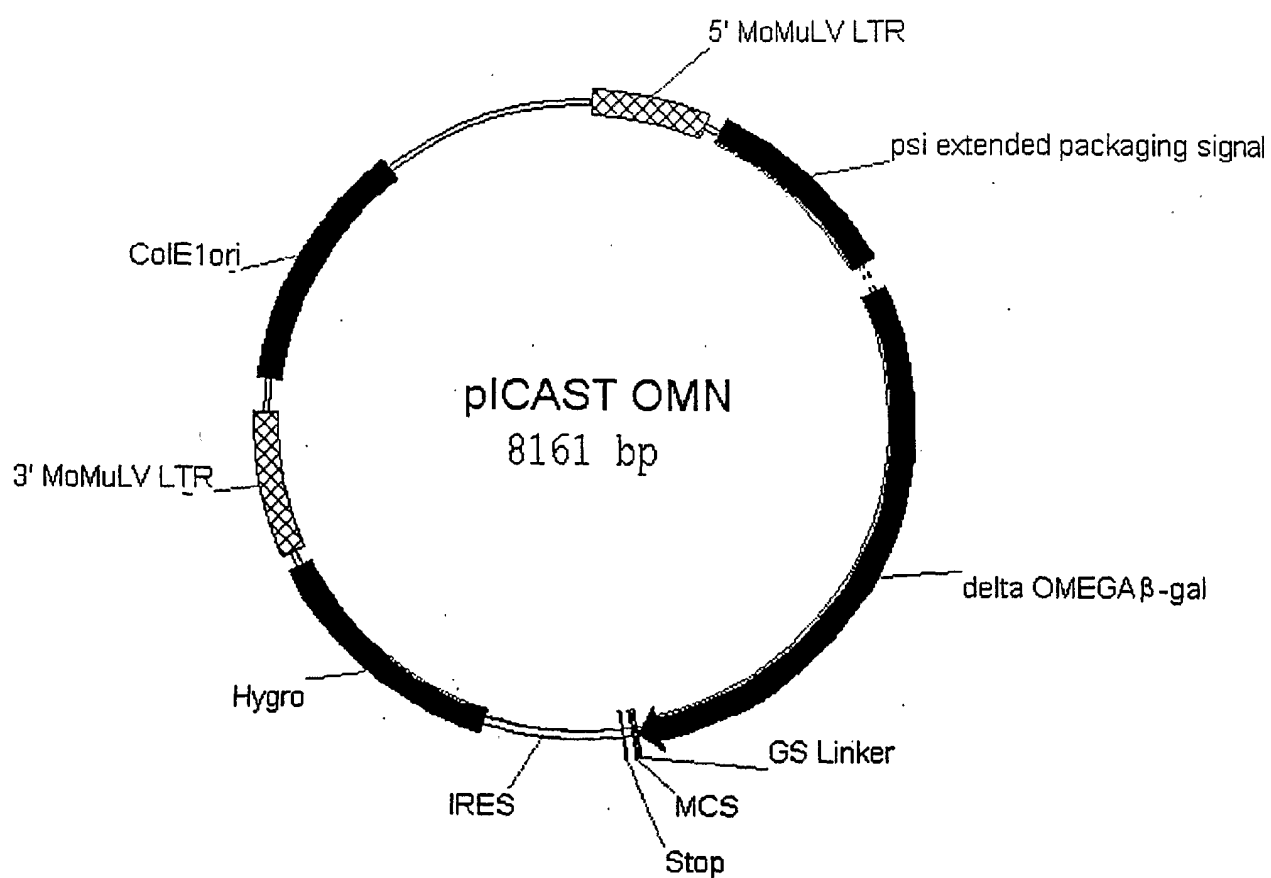



Figure 13A

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1  CTGCAGCCTG AATATGGGCC AAACAGGATA TCTGTGGTAA GCAGTTCCTG
   GACGTCCGGAC TTATACCCGG TTTGTCCTAT AGACACCATT CGTCAAGGAC
-----
51  CCCC GGCTCA GGGCCAAGAA CAGATGGAAC AGCTGAATAT GGGCCAAACA
   GGGGCCGAGT CCCGGTTCTT GTCTACCTTG TCGACTTATA CCCGGTTTGT
-----
101 GGATATCTGT GGTAA GCAGT TCCTGCCCCG GCTCAGGGCC AAGAACAGAT
   CCTATAGACA CCATTTCGTCA AGGACGGGGC CGAGTCCCGG TTCTTGTCTA
-----
151 GGTCCCCAGA TGGGGTCCAG CCCTCAGCAG TTTCTAGAGA ACCATCAGAT
   CCAGGGGTCT ACGCCAGGTC GGGAGTCGTC AAAGATCTCT TGGTAGTCTA
-----
201 GTTTCAGGG TGGCCCAAGG ACCTGAAATG ACCCTGTGCC TTATTTGAAC
   CAAAGGTCCC ACGGGGTTC TGGACTTTAC TGGGACACGG AATAAACTTG
-----
251 TAACCAATCA GTTCGCTTCT CGCTTCTGTT CGCGCGCTTC TGCTCCCCGA
   ATTGGTTAGT CAAGCGAAGA GCGAAGACAA GCGCGCGAAG ACGAGGGGCT
-----
301 GCTCAATAAA AGAGCCCACA ACCCCTCACT CGGGGCGCCA GTCCTCCGAT
   CGAGTTATTT TCTCGGGTGT TGGGGAGTGA GCCCCGCGGT CAGGAGGCTA
-----
351 TGACTGAGTC GCCCGGGTAC CCGTGTATCC AATAAACCCCT CTTGCAGTTG
   ACTGACTCAG CCGGCCCATG GGCACATAGG TTATTTGGGA GAACGTCAAC
-----
401 CATCCGACTT GTGGTCTCGC TGTTCCCTGG GAGGGTCTCC TCTGAGTGAT
   GTAGGCTGAA CACCAGAGCG ACAAGGAACC CTCCAGAGG AGACTCACTA
-----
451 TGACTACCCG TCAGCGGGGG TCTTTCATTT GGGGGCTCGT CCGGGATCGG
   ACTGATGGGC AGTCGCCCCC AGAAAGTAAA CCCCCGAGCA GGCCCTAGCC
-----
501 GAGACCCCTG CCCAGGGACC ACCGACCCAC CACCGGGAGG CAAGCTGGCC
   CTCTGGGGAC GGGTCCCTGG TGGCTGGGTG GTGGCCCTCC GTTCGACCGG
-----
551 AGCAACTTAT CTGTGCTGT CCGATTGTCT AGTGTCTATG ACTGATTTA
   TCGTTGAATA GACACAGACA GGCTAACAGA TCACAGATAC TGACTAAAAT
-----
601 TGGCCTGCG TCGGTACTAG TTAGCTRACT AGCTCTGTAT CTGGCGGACC
   ACGCGGACGC AGCCATGATC AATCGATTGA TCGAGACATA GACCGCTGG
-----
651 CGTGGTGGAA CTGACGAGTT CTGAACACCC GGCCGCAACC CTGGGAGACG
   GCACCACCTT GACTGCTCAA GACTTGTGGG CCGGCGTTGG GACCCTCTGC
-----
701 TCCCAGGGAC TTTGGGGGCC GTTTTTGTGG CCCGACCTGA GGAAGGGAGT
   AGGGTCCCTG AAACCCCGG CAAAACACC GGGCTGGACT CCTTCCCTCA
-----
751 CGATGIGGAA TCCGACCCCG TCAGGATATG TGGTCTGGT AGGAGACGAG
   GCTACACCTT AGGCTGGGGC AGTCCTATAC ACCAAGACCA TCCTCTGCTC
-----
801 AACCTAAAAC AGTCCCGCC TCCGTCTGAA TTTTGTCTT CGGTTTGGAA
   TTGGATTTG TCAAGGGCGG AGGCAGACTT AAAACGAAA GCCAACCTT
-----
851 CCGAAGCCGC GCGTCTTGTG TGCTGCAGCA TCGTCTGTG TTGTCTCTGT
   GGCTTCGGCG CCGAGAACAG ACGACGTCGT AGCAAGACAC AACAGAGACA
-----
901 CTGACTGTGT TTCTGTATTT GTCTGAAAAT TAGGGCCAGA CTGTTACCAC
   GACTGACACA AAGACATAAA CAGACTTTTA ATCCCGGTCT GACAATGGTG
-----

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FIGURE 13B

951 TCCCTTAAGT TTGACCTTAG G'AACTGGAA AGATGTCGAG CGGCTCGCTC
 AGGGAATTCA AACTGGAATC CATTGACCTT TCTACAGCTC GCCGAGCGAG

1001 ACAACCAGTC GGTAGATGTC AAGAAGAGAC GTTGGGTAC CTTCTGCTCT
 TGTTGGTCAG CCATCTACAG TTCTTCTCTG CAACCCAATG GAAGACGAGA

1051 GCAGAATGGC CAACCTTTAA CGTCGGATGG CCGCGAGACG GCACCTTTAA
 CGTCTTACCG GTTGGAAATT GCAGCCTACC GGGCTCTGC CGTGGAAATT

1101 CCGAGACCTC ATCACCAGG TTAAGATCAA GGTCTTTTCA CCTGGCCCGC
 GGCTCTGGAG TAGTGGGTCC AATTCTAGTT CCAGAAAAGT GGACCGGGCG

1151 ATGGACACCC AGACCAGGTC CCTACATCG TGACCTGGGA AGCCTTGGCT
 TACCTGTGGG TCTGGTCCAG GGGATGTAGC ACTGGACCCT TCGGAACCGA

1201 TTTGACCCCC CTCCCTGGGT CAAGCCCTTT GTACACCCTA AGCCTCCGCC
 AAACCTGGGG GAGGGACCCA GTTCGGGAAA CATGTGGGAT TCGGAGGCGG

1251 TCCTCTTCTT CCATCCGCC CGTCTCTCCC CCTTGAACCT CCTCGTTCGA
 AGGAGAAGGA GGTAGCGGG GCAGAGAGGG GGAACCTTGA GGAGCAAGCT

1301 CCGCGCTCG ATCCTCCCTT TATCCAGCCC TCACTCCTTC TCTAGGCGCC
 GGGCGGAGC TAGGAGGGAA ATAGGTCGGG AGTGAGGAAG AGATCCGCGG

1351 GGCCGCTCTA GCCCATTAAT ACGACTCACT ATAGGGCGAT TCGAACACCA
 CCGGGGAGAT CGGGTAATTA TGCTGAGTGA TATCCGCTA AGCTTGTGGT

1401 TGCACCATCA TCATCATCAC GTCGACGAAC AGAAACTCAT TTCCGAAGAA
 ACGTGGTAGT AGTAGTAGTG CAGCTGCTTG TCTTTGAGTA AAGGCTTCTT

1451 GACCTACTCG AGATGGGCGT GATTACGGAT TCACTGGCCG TCGTTTTACA
 CTGGATGAGC TCTACCCGCA CTAATGCCTA AGTGACCGGC AGCAAAATGT

1501 ACGTCGTGAC TGGGAAAACC CTGGCGTTAC CCAACTTAAT CGCCTTGCAG
 TGCAGCACTG ACCCTTTTGG GACCGCAATG GGTTGAATTA GCGGAACGTC

1551 CACATCCCC TTTCCGCCAGC TGCGTAATA GCGAAGAGGC CCGCACCGAT
 GTGTAGGGGG AAAGCGGTGC ACCGCATTAT CGCTTCTCCG GCGGTGGCTA

1601 CGCCCTTCCC AACAGTTACG CAGCCTGAAT GCGGAATGGC GCTTTGCCTG
 GCGGGAAGGG TTGTCAATGC GTCGGACTTA CCGCTTACCG CGAAACGGAC

1651 GTTCCGGCA CCAGAAGCGG TGCCGAAAG CTGGCTGGAG TGCGATCTTC
 CAAAGGCCGT GGTCTTCGCC ACGCCTTTC GACCGACCTC ACGCTAGAAG

1701 CTGAGGCCGA TACTGTCGTC GTCCCTCAA ACTGGCAGAT GCACGGTTAC
 GACTCCGGCT ATGACAGCAG CAGGGGAGTT TGACCGTCTA CGTGCCAATG

1751 GATGCGCCCA TCTACACCAA CGTGACCTAT CCCATTACGG TCAATCCGCC
 CTACGCGGGT AGATGTGGTT GCACTGGATA GGGTAATGCC AGTTAGGCGG

1801 GTTGTTCCTT ACGGAGAATC CGACGGGTTG TTA CTGCTC ACATTTAATG
 CAAACAAGGG TGCTCTTAG GTCGCCAAC AATGAGCGAG TGTAARTAC

1851 TTGATGAAAG CTGGCTACAG GAAGGCCAGA CGCGAATTAT TTTTGATGGC
 AACTACTTTC GACCGATGTC CTTCCGGTCT GCGCTTAATA AAACTACCG

1901	GTAACTCGG CAATTGAGCC	CGTTTCATCT GCAAAGTAGA	GTGGTGCAAC CACCACGTTG	GGGCGCTGGG CCCGCGACCC	TCGGTTACGG AGCCAATGCC
1951	CCAGGACAGT GGTCCGTGCA	CGTTTGCCGT GCAAACGGCA	CTGAATTTGA GACTTAAACT	CCTGAGCGCA GGACTCGCGT	TTTTTACGGG AAAAATGCGC
2001	CCGGAGAAAA GGCCTCTTTT	CCGCCTCGCG GGCGGAGCGC	GTGATGGTGC CACTACCACG	TGCGCTGGAG ACGCGACCTC	TGACGGCAGT ACTGCCGTCA
2051	TATCTGGAAG ATAGACCTTC	ATCAGGATAT TAGTCCTATA	GTGGCGGATG CACCGCCTAC	AGCGGCATTT TCGCCGTAAA	TCCGTGACGT AGGCACTGCA
2101	CTCGTTGCTG GAGCAACGAC	CATAAACCGA GTATTTGGCT	CTACACAAAT GATGTGTTTA	CAGCGATTTT GTCGCTAAAG	CATGTTGCCA GTACAACGGT
2151	CTCGCTTTAA GAGCGAAATT	TGATGATTTT ACTACTAAAG	AGCCGCGCTG TCGGCGCGAC	TACTGGAGGC ATGACCTCCG	TGAAGTTCAG ACTTCAAGTC
2201	ATGTGCGGCG TACACGCCGC	AGTTGCGTGA TCAACGCACT	CTACCTACGG GATGGATGCC	GTAACAGTTT CATTGTCAA	CTTTATGGCA GAAATACCGT
2251	GGGTGAAACG CCCCTTTTGC	CAGGTCGCCA GTCCAGCGGT	GCGGCACCGC CGCCGTGGCG	GCCTTTCGGC CGGAAAGCCG	GGTGAATTA CCACTTTAAT
2301	TCGATGAGCG AGCTACTCGC	TGGTGGTTAT ACCACCAATA	GCCGATCGCG CGGCTAGCGC	TCACACTACG AGTGTGATGC	TCTGAACGTC AGACTTGCAG
2351	GAAAACCCGA CTTTTGGGCT	AACTGTGGAG TTGACACCTC	CGCCGAAATC GCGGCTTAG	CCGAATCTCT GGCTTAGAGA	ATCGTGCGGT TAGCACGCCA
2401	GGTTGAACTG CCAACTTGAC	CACACCGCCG GTGTGGCGCG	ACGGCACGCT TGCCGTGCGA	GATTGAAGCA CTAACTTCGT	GAAGCCTGCG CTTCGGACGC
2451	ATGTGCGTTT TACAGCCAAA	CCGCGAGGTG GGCGCTCCAC	CGGATTGAAA GCCTAACTTT	ATGGTCTGCT TACCAGACGA	GCTGCTGAAC CGACGACTTG
2501	GGCAAGCCGT CCGTTTCGGCA	TGCTGATTCG ACGACTAAGC	AGGCGTTAAC TCCGCAATTG	CGTCACGAGC GCAGTGCTCG	ATCATCCTCT TAGTAGGAGA
2551	GCATGGTCTG CGTACCAGTC	GTCATGGATG CAGTACCTAC	AGCAGACGAT TCGTCTGCTA	GGTGCAGGAT CCACGTCCTA	ATCCTGCTGA TAGGACGACT
2601	TGAAGCAGAA ACTTCGTCTT	CAACTTTAAC GTTGAAATTG	GCCGTGCGCT CGGCACGCGA	GTTCCGATTA CAAGCGTAAT	TCCGAACCAT AGGCTTGGTA
2651	CCGCTGTGGT GGCGACACCA	ACACGCTGTG TGTGCGACAC	CGACCGCTAC GCTGGCGATG	GGCCTGTATG CCGGACATAC	TGGTGGATGA ACCACCTACT
2701	AGCCAATATT TCGGTTATAA	GAAACCCACG CTTTGGGTGC	GCATGGTGCC CGTACCACGG	AATGAATCGT TACTTAGCA	CTGACCGATG GACTGGCTAC
2751	ATCCGCGCTG TAGGCGCGAC	GCTACCGGCG CGATGGCCGC	ATGAGCGAAC TACTCGCTTG	GCGTAACGCG CGCATTGCGC	AATGGTGCAG TTACCACGTC
2801	CGCGATCGTA GCGCTAGCAT	ATCACCCGAG TAGTGGGCTC	TGTGATCATC ACACTAGTAG	TGGTCGCTGG ACCAGCGACC	GGAATGAATC CCTTACTTAG

```

2851  AGGCCACGGC  GCTAATCAGC  ACGCGCTGTA  TCGCTGGATC  AAATCTGTGC
      TCCGGTGCCG  CGATTAGTGC  TGC GCGACAT  AGCGACCTAG  TTTAGACAGC
-----
2901  ATCCTTCCCG  CCCGGTGCAG  TATGAAGGCG  GCGGAGCCGA  CACCACGGCC
      TAGGAAGGGC  GGGCCACGTC  ATACTTCCGC  CGCCTCGGCT  GTGGTGCCGG
-----
2951  ACCGATATTA  TTTGCCCGAT  GTACGCGCGC  GTGGATGAAG  ACCAGCCCTT
      TGGCTATAAT  AAACGGGCTA  CATGCGCGCG  CACCTACTTC  TGGTCGGGAA
-----
3001  CCCGGCTGTG  CCGAAATGGT  CCATCAAAAA  ATGGCTTTCG  CTACCTGGAG
      GGGCCGACAC  GGCTTTACCA  GGTAGTTTTT  TACCGAAAGC  GATGGACCTC
-----
3051  AGACGCGCCC  GCTGATCCTT  TGCGAATACG  CCCACGCGAT  GGGTAACAGT
      TCTGCGCGGG  CGACTAGGAA  ACGCTTATGC  GGGTGCCTA  CCCATTGTCA
-----
3101  CTTGGCGGTT  TCGCTAAATA  CTGGCAGGCG  TTTCGTCAGT  ATCCCCGTTT
      GAACCGCCAA  AGCGATTTAT  GACCGTCCGC  AAAGCAGTCA  TAGGGGCAAA
-----
3151  ACAGGGCGGC  TTCGTCTGGG  ACTGGGTGGA  TCAGTCGCTG  ATTAAATATG
      TGTCCCGCCG  AAGCAGACCC  TGACCCACCT  AGTCAGCGAC  TAATTTATAC
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3201  ATGAAAACGG  CAACCCGTGG  TCGGCTTACG  GCGGTGATTT  TGGCGATACG
      TACTTTTGCC  GTTGGGCACC  AGCCGAATGC  CGCCACTAAA  ACCGCTATGC
-----
3251  CCGAACGATC  GCCAGTCTG  TATGAACGGT  CTGGTCTTTG  CCGACCGCAC
      GGCTTGCTAG  CGGTCAAGAC  ATACTTGCCA  GACCAGAAAC  GGCTGGCGTG
-----
3301  GCCGCATCCA  GCGCTGACGG  AAGCAAAACA  CCAGCAGCAG  TTTTTCAGT
      CGGCGTAGGT  CCGGACTGCC  TTCGTTTTGT  GGTCTGCTAT  AAAAAGGTCA
-----
3351  TCCGTTTATC  CGGGCAAACC  ATCGAAGTGA  CCAGCGAATA  CCTGTTCCGT
      AGGCAAATAG  GCCCGTTTGG  TAGCTTCACT  GGTCTGCTAT  GGACAAGGCA
-----
3401  CATAGCGATA  ACGAGCTCCT  GCACTGGATG  GTGGCGCTGG  ATGGTAAGCC
      GTATCGCTAT  TGCTCGAGGA  CGTGACCTAC  CACCGCGACC  TACCATTCCG
-----
3451  GCTGGCAAGC  GGTGAAGTGC  CTCTGGATGT  CGCTCCACAA  GGTAACAGT
      CGACCGTTTC  CCACCTCACG  GAGACCTACA  GCGAGGTGTT  CCATTTGTCA
-----
3501  TGATTGAACT  GCCTGAACTA  CCGCAGCCGG  AGAGCGCCGG  GCAACTCTGG
      ACTAACTTGA  CGGACTTGAT  GCGCTCGGCC  TCTCGCGGCC  CGTTGAGACC
-----
3551  CTCACAGTAC  GCGTAGTGCA  ACCGAACGCG  ACCGCATGGT  CAGAAGCCGG
      GAGTGTCAAT  CGCATCACGT  TGGCTTGCCG  TGGCGTACCA  GTCTTCGGCC
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3601  GCACATCAGC  GCCTGGCAGC  AGTGGCGTCT  GGCGGAAAAC  CTCAGTGTGA
      CGTGTAGTCG  CGGACCGTCG  TCACCGCAGA  CCGCCTTTTG  GAGTCACACT
-----
3651  CGCTCCCCGC  CGGTCCCAC  GCCATCCGCG  ATCTGACCAC  CAGCGAAATG
      GCGAGGGGCG  GCGCAGGGTG  CGGTAGGGCG  TAGACTGGTG  GTCGCTTTAC
-----
3701  GATTTTTGCA  TCGAGCTGGG  TAATAAGCGT  TGGCAATTTA  ACCGCCAGTC
      CTAAAAACGT  AGCTCGACCC  ATTATTGCA  ACCGTTAAAT  TGGCGGTCAG
-----
3751  AGGCTTTCTT  TCACAGATGT  GGATTGGCGA  TAAAAAACA  CTGCTGACGC
      TCCGAAAGAA  AGTGCTTACA  CCTAACCGCT  ATTTTTGTGT  GACGACTGCC
-----

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3801 CGCTGCGCGA TCAGTTCACC CGTGTCGATA GATCTGGAGG TGGTGGCAGC
 GCGACGCGCT AGTCAAGTGG GCACAGCTAT CTAGACCTCC ACCACCGTCG

3851 AGGCCTTGGC GCGCCGGATC CTTAATTAAC AATTGACCGG TAATAATAGG
 TCCGGAACCG CGCGGCCTAG GAATTAATTG TTAAGTGGCC ATTATTATCC

3901 TAGATAAGTG ACTGATTAGA TGCATTTCTGA CTAGATCCCT CGACCAATTC
 ATCTATTAC TGAATAATCT ACGTAAAGCT GATCTAGGGA GCTGGTTAAG

3951 CGGTTATTTT CCACCATATT GCCGTCTTTT GGCAATGTGA GGGCCCGGAA
 GCCAATAAAA GGTGGTATAA CGGCAGAAAA CCGTTACACT CCCGGGCCTT

4001 ACCTGGCCCT GTCTTCTTGA CGAGCATTCC TAGGGGTCTT TCCCCTCTCG
 TGGACCGGGA CAGAAGAACT GCTCGTAAGG ATCCCCAGAA AGGGGAGAGC

4051 CCAAAGGAAT GCAAGGTCTG TTGAATGTCG TGAAGGAAGC AGTTCCTCTG
 GGTTCCTTA CGTTCAGAC AACTTACAGC ACTTCCTTCG TCAAGGAGAC

4101 GAAGCTTCTT GAAGACAAAC AACGTCTGTA GCGACCCTTT GCAGGCAGCG
 CTTCGAAGAA CTTCTGTTTG TTGCAGACAT CGCTGGGAAA CGTCCGTCTG

4151 GAACCCCCCA CCTGGCGACA GGTGCCTCTG CGGCCAAAAG CCACGTGTAT
 CTTGGGGGGT GGACCGCTGT CCACGGAGAC GCCGGTTTTC GGTGCACATA

4201 AAGATACACC TGCAAAGGCG GCACAACCCC AGTGCCACGT TGTGAGTTGG
 TTCTATGGG ACGTTTCCGC CGTGTGGGGG TCACGGTGCA ACACTCAACC

4251 ATAGTTGTGG AAAGAGTCAA ATGGCTCTCC TCAAGCGTAT TCAACAAGGG
 TATCAACACC TTTCTCAGTT TACCGAGAGG AGTTCGCATA AGTTGTCC

4301 GCTGAAGGAT GCCCAGAAGG TACCCATTG TATGGGATCT GATCTGGGGC
 CGACTTCCTA CGGTCTTCC ATGGGGTAAC ATACCCTAGA CTAGACCCCG

4351 CTCGGTGCAC ATGCTTTACA TGTGTTTAGT CGAGGTTAAA AAACGTCTAG
 GAGCCACGTG TACGAAATGT ACACAAATCA GCTCCAATTT TTTGCAGATC

4401 GCCCCCCGAA CCACGGGGAC GTGGTTTTCC TTTGAAAAC ACGATGATAA
 CGGGGGGCTT GGTGCCCTG CACCAAAGG AACTTTTTG TGCTACTATT

4451 TACCATGAAA AAGCCTGAAC TCACCGCGAC GTCTGTCGAG AAGTTTCTGA
 ATGGTACTTT TTCGGACTTG AGTGGCGCTG CAGACAGCTC TTCAAAGACT

4501 TCGAAAAGTT CGACAGCGTC TCCGACCTGA TGCAGCTCTC GGAGGGCGAA
 AGCTTTTCAA GCTGTGCGAG AGGCTGGACT ACGTCGAGAG CCTCCCGCTT

4551 GAATCTCGTG CTTTCAGCTT CGATGTAGGA GGGCGTGGAT AIGTCCTGCG
 CTTAGAGCAC GAAAGTCGAA GCTACATCCT CCCGCACCTA TACAGGACGC

4601 GGTAATAGC TGCGCCGATG GTTCTACAA AGATCGTTAT GTTTATCGGC
 CCATTTATCG ACGCGGCTAC CAAAGATGTT TCTAGCAATA CAAATAGCCG

4651 ACTTTGCATC GGCCGCGCTC CCGATTCCGG AAGTGCTTGA CATTGGGGAA
 TGAAACGTAG CCGGCGCGAG GGCTAAGGCC TTCACGAACT GTAACCCCTT

4701 TTTAGCGAGA GCCTGACCTA TTGCATCTCC CGCCGTGCAC AGGGTGTAC
 AAATCGCTCT CGGACTGGAT AACGTAGAGG GCGGCACGTG TCCCACAGTG

4751 GTTGCAAGAC CTGCCTGAAA CCGAACTGCC CGCTGTTCTG CAGCCGGTCC
 CAACGTTCTG GACGGACTTT GGCTTGACGG GCGACAAGAC GTCGGCCAGC

4801 CGGAGGCCAT GGATGCGATC GCTGCGGCCG ATCTTAGCCA GACGAGCGGG
 GCCTCCGGTA CCTACGCTAG CGACGCCGGC TAGAATCGGT CTGCTCGCCC

4851 TTCGGCCCAT TCGGACCGCA AGGAATCGGT CAATACACTA CATGGCGTGA
 AAGCCGGGTA AGCCTGGCGT TCCTTAGCCA GTTATGTGAT GTACCGCACT

4901 TTTCATATGC GCGATTGCTG ATCCCCATGT GTATCACTGG CAAACTGTGA
 AAAGTATACG CGCTAACGAC TAGGGGTACA CATAGTGACC GTTGTGACACT

4951 TGGACGACAC CGTCAGTGCG TCCGTGCGGC AGGCTCTCGA TGAGCTGATG
 ACCTGCTGTG GCAGTCACGC AGGCAGCGCG TCCGAGAGCT ACTCGACTAC

5001 CTTGGGGCCG AGGACTGCCC CGAAGTCCGG CACCTCGTGC ACGCGGATTT
 GAAACCCGGC TCCTGACGGG GCTTCAGGCC GTGGAGCAGG TGCGCCTAAA

5051 CGGCTCCAAC AATGTCTCTGA CGGACAATGG CCGCATAACA GCGGTCATTG
 GCCGAGGTTG TTACAGGACT GCCTGTTACC GGCATATTGT CGCCAGTAAC

5101 ACTGGAGCGA GGGCATGTTT GGGGATTCCC AATACGAGGT CGCCAACATC
 TGACCTCGCT CCGCTACAAG CCCCTAAGGG TTATGCTCCA GCGGTTGTAG

5151 TTCTTCTGGA GGCCGTGGTT GGCTTGTATG GAGCAGCAGA CGCGCTACTT
 AAGAAGACCT CCGGCACCAA CCGAACATAC CTCGTCTGCT GCGCGATGAA

5201 CGAGCGGAGG CATCCGGAGC TTGCAGGATC GCCGCGGCTC CGGGCGTATA
 GCTCGCCTCC GTAGGCCTCG AACGTCTTAG CGGCGCCGAG GCCCGCATAT

5251 TGCTCCGCAT TGGTCTTGAC CAACTCTATC AGAGCTTGGT TGACGGCAAT
 ACGAGGCGTA ACCAGAACTG GTTGAGATAG TCTCGAACCA ACTGCCGTTA

5301 TTCGATGATG CAGCTTGGGC GCAGGGTCGA TCGCAGCAA TCGTCCGATC
 AAGCTACTAC GTCGAACCCG CGTCCCAGCT ACGCTGCGTT AGCAGGCTAG

5351 CGGAGCCGGG ACTGTGCGGC GTACACAAAT CGCCCGCAGA AGCGCGGCCG
 GCCTCGGCC TGACAGCCCG CATGTGTTTA GCGGGCGTCT TCGCGCCGGC

5401 TCTGGACCGA TGGCTGTGTA GAAGTACTCG CCGATAGTGG AAACCGACGC
 AGACCTGGCT ACCGACACAT CTTCATGAGC GGCTATCACC TTTGGCTGCG

5451 CCCAGCACTC GTCCGAGGGC AAAGGAATAG AGTAGATGCC GACCGGGATC
 GGTTCGTGAG CAGGCTCCCG TTTCCTTATC TCATCTACGG CTGGCCCTAG

5501 TATCGATAAA ATAAAAGATT TTATTTAGTC TCCAGAAAAA GGGGGGAATG
 ATAGCTATTT TATTTTCTAA AATAAATCAG AGGTCTTTTT CCCCCTTAC

5551 AAAGACCCCA CCTGTAGGTT TGGCAAGCTA GCTTAAGTAA CGCCATTTTG
 TTTCTGGGGT GGACATCCAA ACCGTTCGAT CGAATTCATT GCGGTAANAAC

5601 CAAGGCATGG AAAAATACAT AACTGAGAAT AGAGAAGTTC AGATCAAGGT
 GTTCCGTACC TTTTATGTA TTGACTCTTA TCTCTTCAAG TCTAGTTCCA

5651 CAGGAACAGA TGGAACAGCT GAATATGGGC CAAACAGGAT ATCTGTGGTA
 GTCCTGTGCT ACCTTGTCGA CTTATACCCG GTTTGTCCTA TAGACACCAT

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5701  AGCAGTTCCT  GCCCCGGCTC  AGGGCCAAGA  ACAGATGGAA  CAGCTGAATA
      TCGTCAAGGA  CGGGGCCGAG  TCCCGGTTCT  TGTCTACCTT  GTCGACTTAT
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5751  TGGGCCAAAC  AGGATATCTG  TGGTAAGCAG  TTCCTGCCCC  GGCTCAGGGC
      ACCCGGTTTG  TCCTATAGAC  ACCATTGCTC  AAGGACGGGG  CCGAGTCCCG
-----
5801  CAAGAACAGA  TGGTCCCAG  ATGCGGTCCA  GCCCTCAGCA  GTTCTAGAG
      GTTCTGTCT  ACCAGGGGTC  TACGCCAGGT  CGGGAGTCGT  CAAAGATCTC
-----
5851  AACCATCAGA  TGTTCAGG  GTGCCCAAG  GACCTGAAAT  GACCCTGTGC
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5901  CTTATTTGAA  CTAACCAATC  AGTTCGCTTC  TCGCTTCTGT  TCGCGCGCTT
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      GACGAGGGGC  TCGAGTTATT  TTCTCGGGTG  TTGGGGAGTG  AGCCCCGCGG
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6001  AGTCCTCCGA  TTGACTGAGT  CGCCCGGGTA  CCCGTGTATC  CAATAAACCC
      TCAGGAGGCT  AACTGACTCA  GCGGGCCCAT  GGGCACATAG  GTTATTTGGG
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6051  TCTTGCAATT  GCATCCGACT  TGTGGTCTCG  CTGTTCCTTG  GGAGGGTCTC
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-----
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      GAGACTCACT  AACTGATGGG  CAGTCGCCCC  CAGAAAGTAA  GTACGTCGTA
-----
6151  GTATCAAAAT  TAATTTGTT  TTTTTCTTA  AGTATTTACA  TTAATGGCC
      CATAGTTTTA  ATTAACCAA  AAAAAAGAA  TCATAAATGT  AATTTACCGG
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6201  ATAGTTGCAT  TAATGAATCG  GCCAACGCGC  GGGGAGAGGC  GGTTCGCTA
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6251  TTGGCGCTCT  TCCGCTTCT  CGCTCACTGA  CTCGCTGCGC  TCGGTGCTC
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6301  GGCTGCGGCG  AGCGGTATCA  GCTCACTCAA  AGGCGGTAAT  ACGGTTATCC
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6351  ACAGAATCAG  GGGATAACGC  AGGAAAGAAC  ATGTGAGCAA  AAGGCCAGCA
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6451  TCCGCCCCCC  TGACGAGCAT  CACAAAAATC  GACGCTCAAG  TCAGAGGTGG
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6501  CGAAACCGGA  CAGGACTATA  AAGATACCAG  GCGTTTCCCC  CTGGAAGCTC
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6551  CCTCGTGCGC  TCTCCTGTT  CGACCTGCC  GCTTACCGGA  TACCTGTCCG
      GGAGCACGCG  AGAGGACAAG  GCTGGGACGG  CGAATGGCCT  ATGGACAGGC
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6601  CCTTCTCCC  TTCGGGAAGC  GTGGCGCTTT  CTCATAGCTC  ACGTGTAGG
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7001 GATTACGCGC AGAAAAAAG GATCTCAAGA AGATCCTTTG ATCTTTTCTA
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7151 CAATCTAAAG TATATATGAG TAAACTTGGT CTGACAGTTA CCAATGCTTA
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7251 TGCCTGACTC CCCGTCGTGT AGATAACTAC GATACGGGAG GGCTTACCAT
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7451 CTAGAGTAAG TAGTTCGCCA GTTAATAGTT TGCGCAACGT FTTGCCATT
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7551 CTCCGGTTCC CAACGATCAA GGCAGTTAC ATGATCCCCC ATGTTGTGCA
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TTTTTCGCCA ATCGAGGAAG CCAGGAGGCT AGCAACAGTC TTCATTCAAC

7651 GCCGCAGTGT TATCACTCAT GGTATGGCA GCACTGCATA ATTCTCTTAC
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8101 AGCGGATACA TATTGAATG TATTTAGAAA AATAAACAAA TAGGGGTTCC
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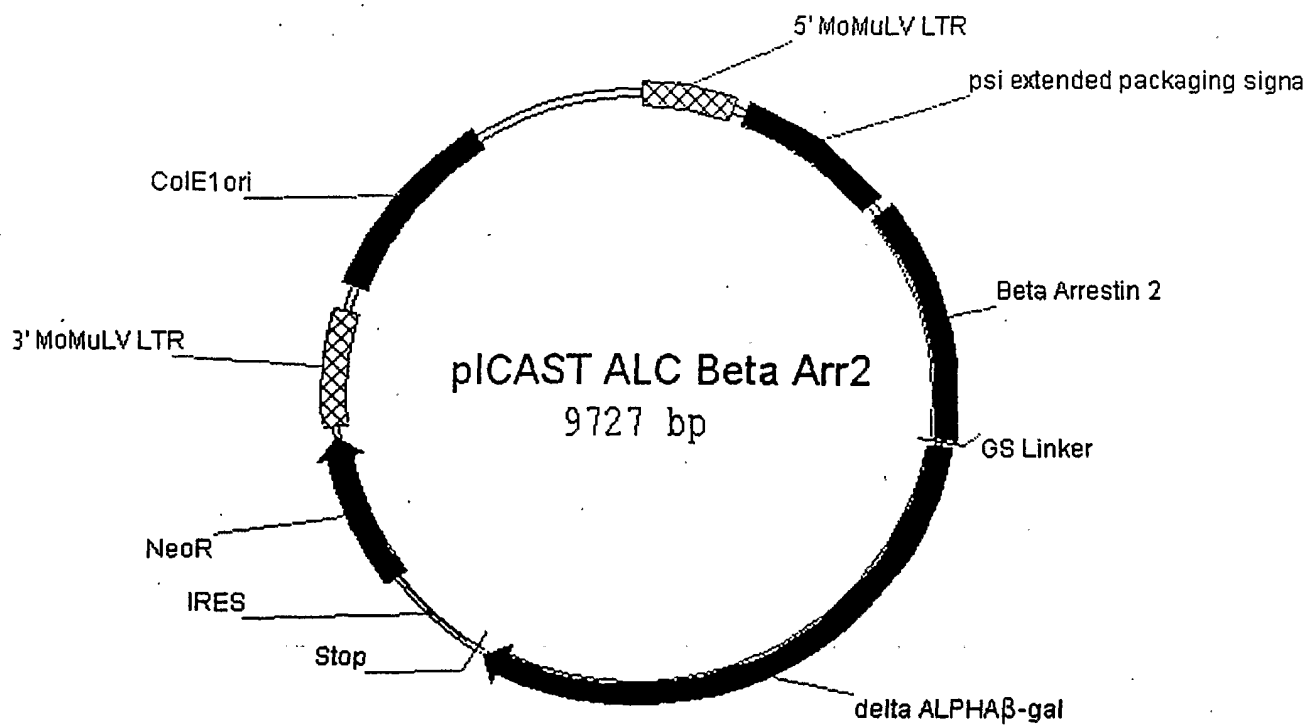


Figure 14

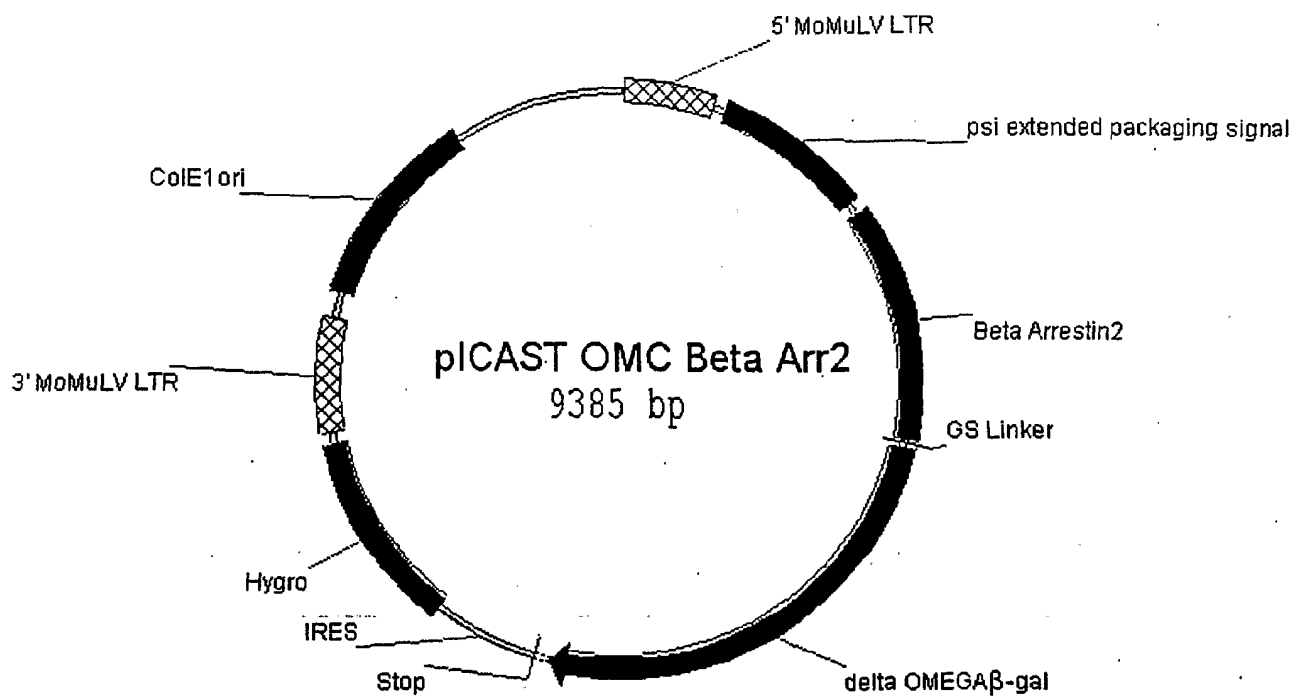


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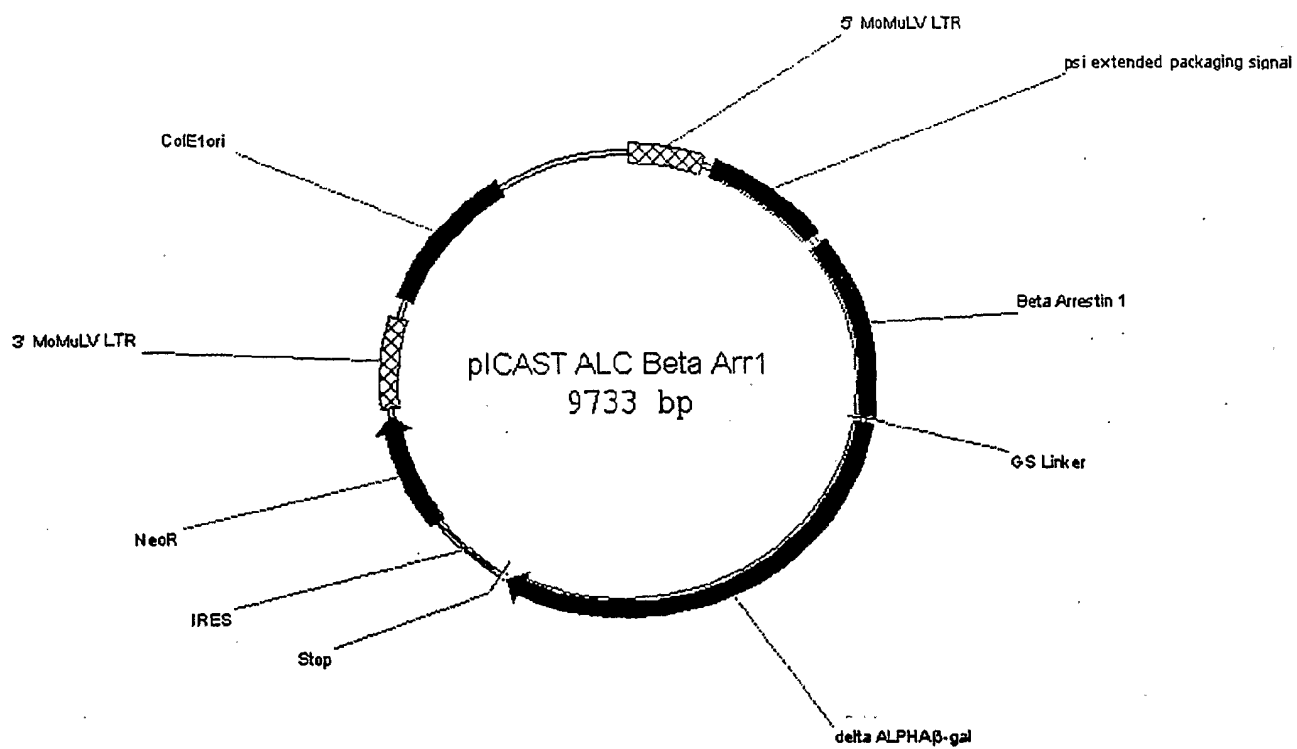


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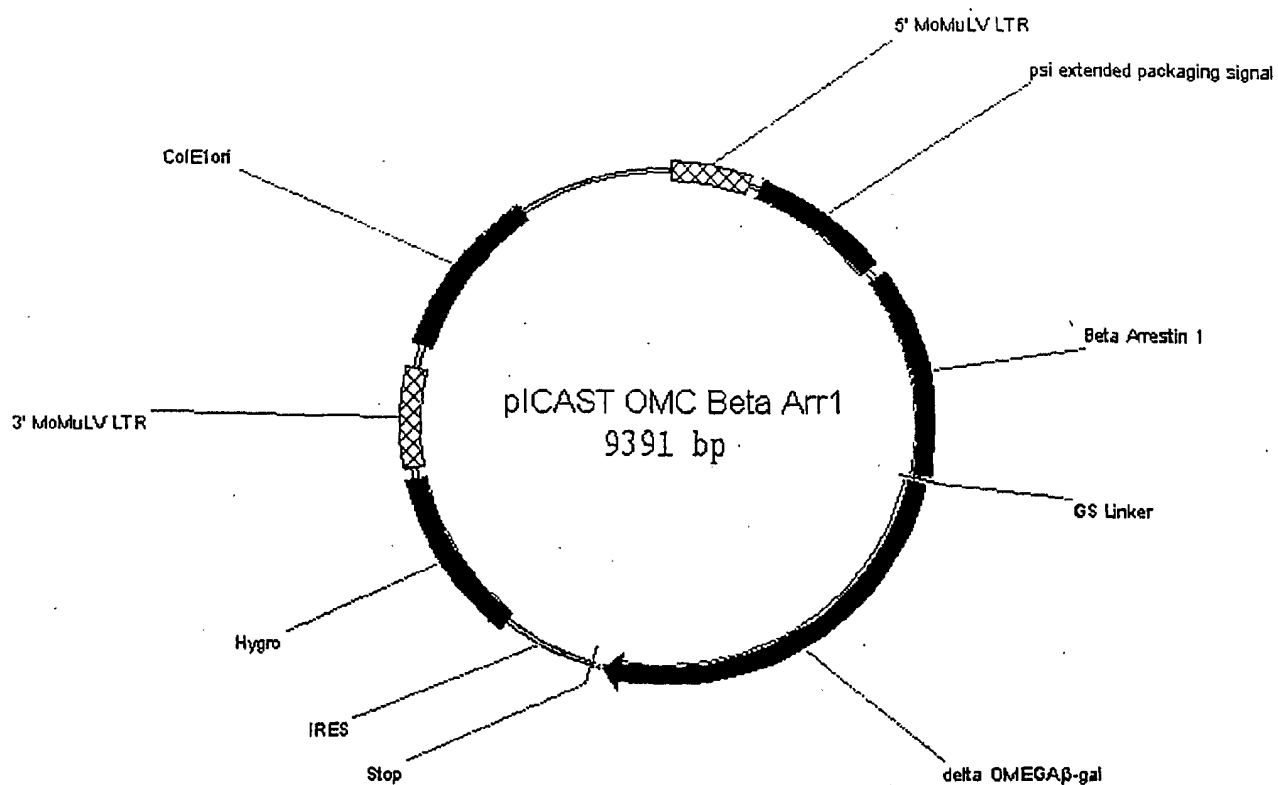


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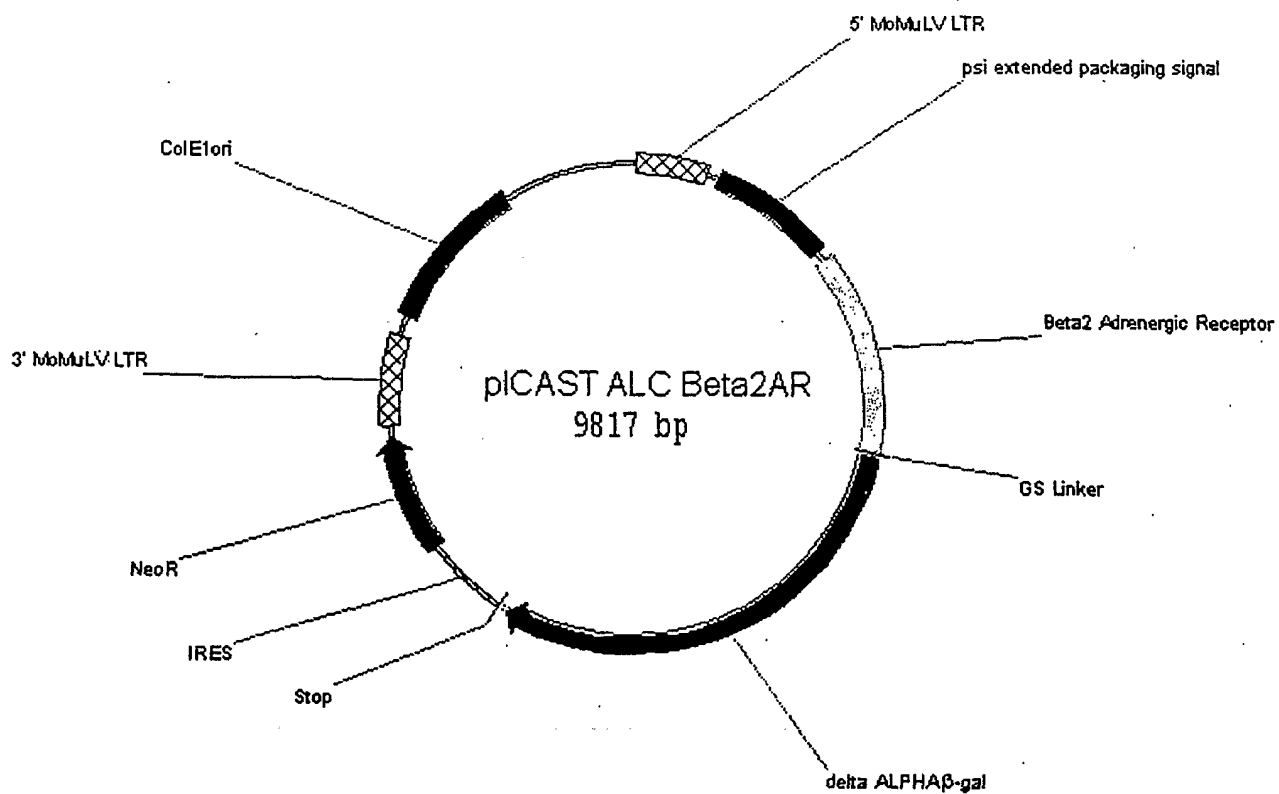


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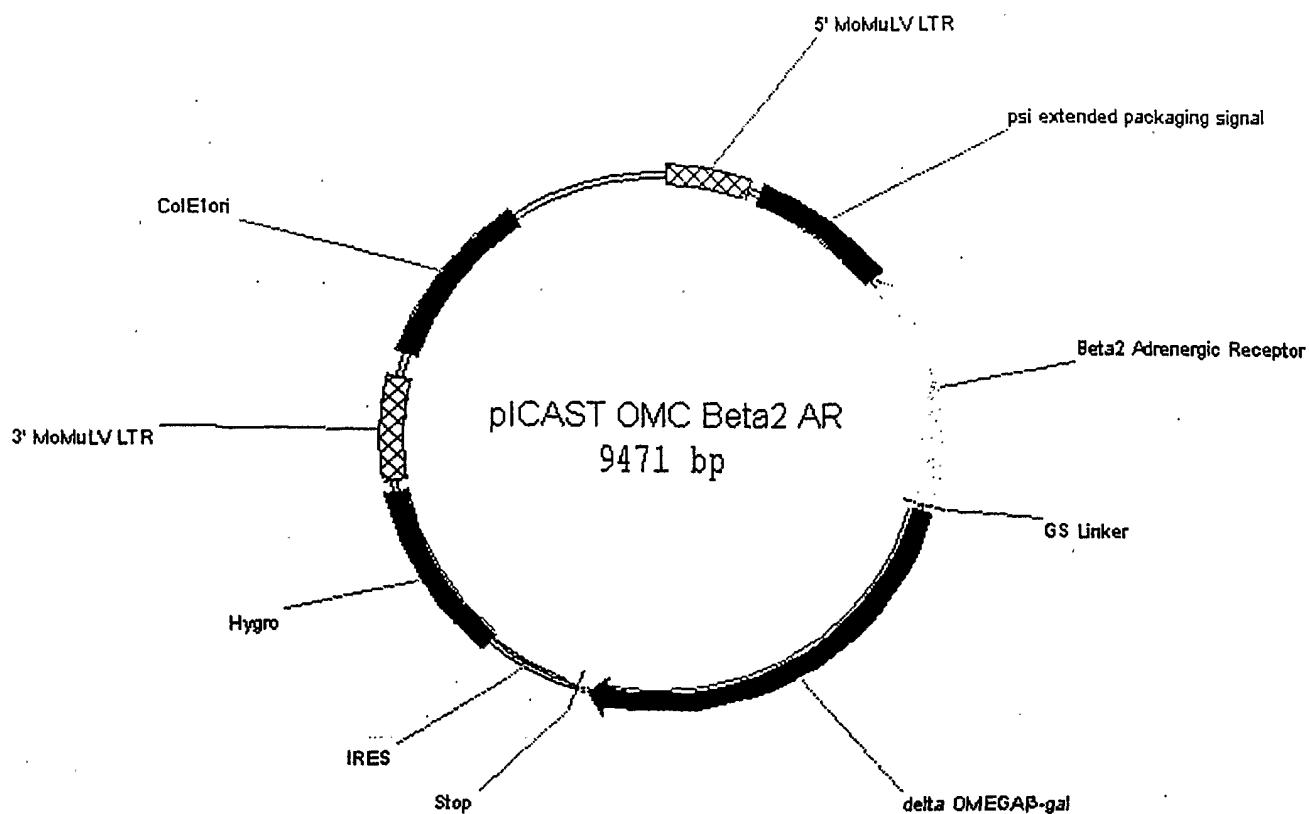


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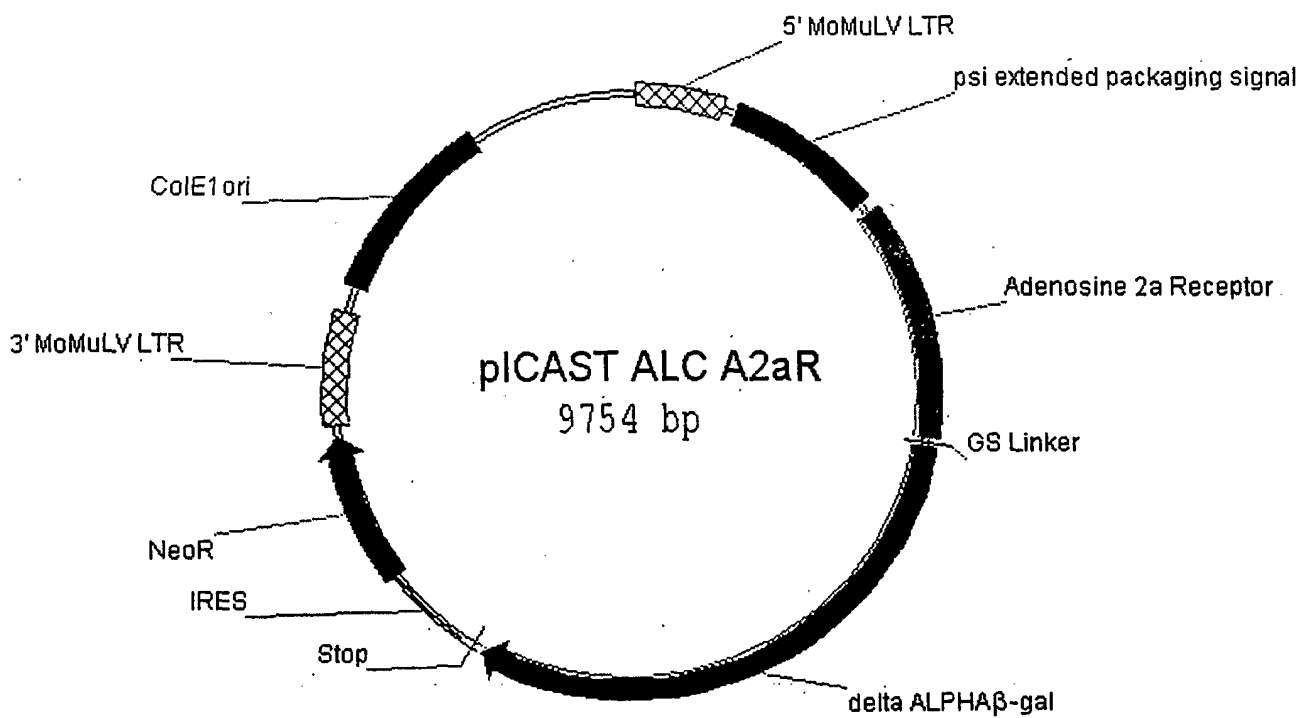


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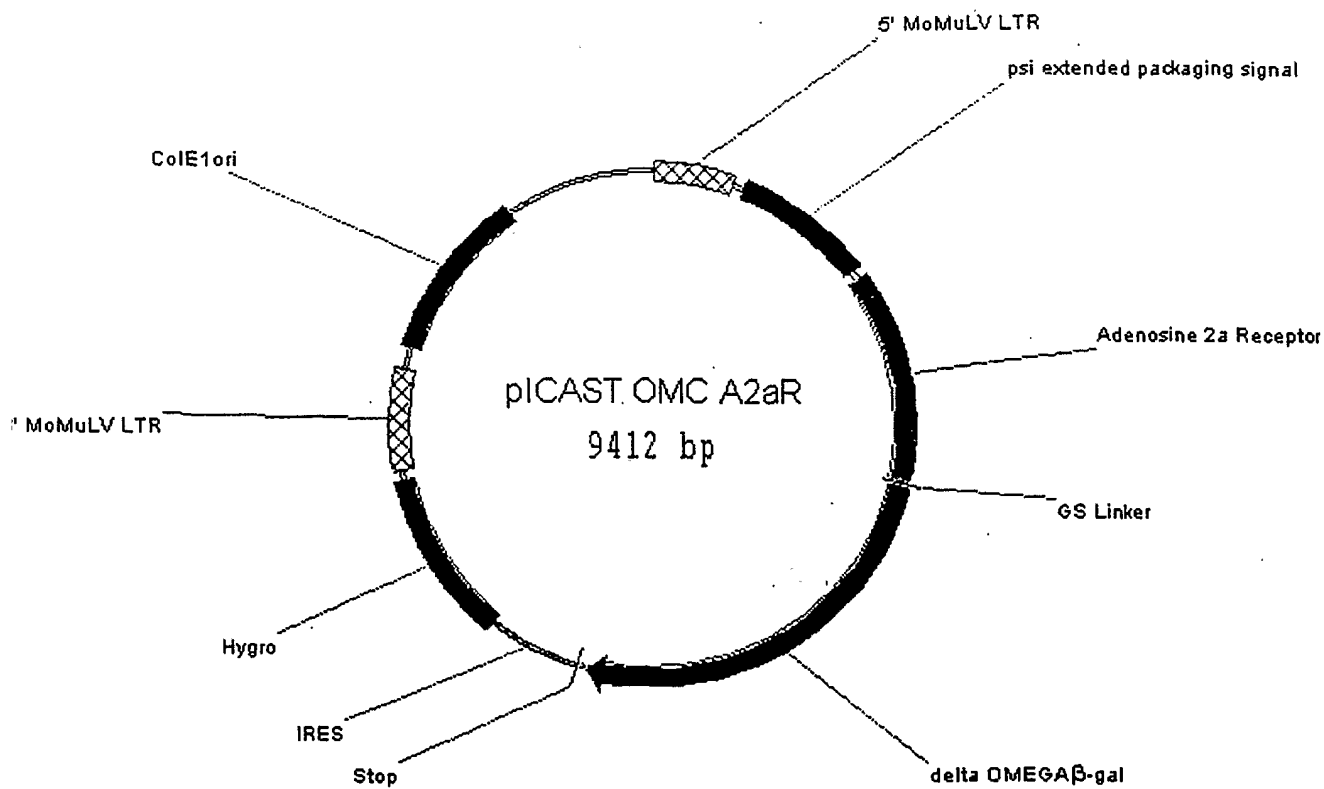


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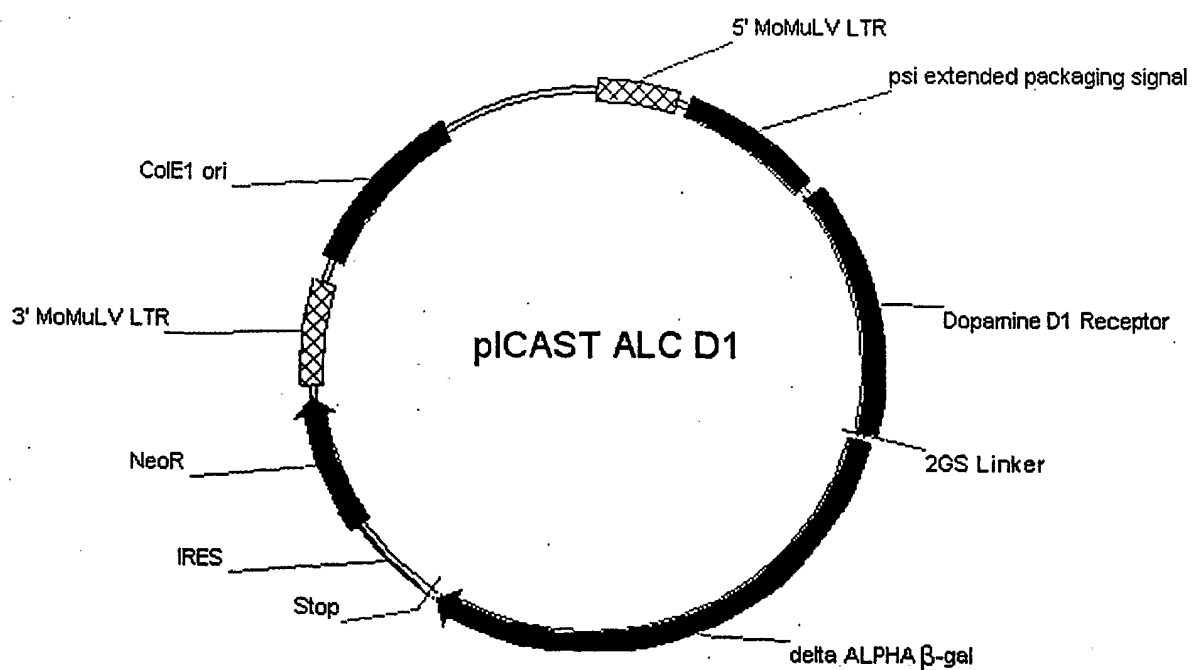


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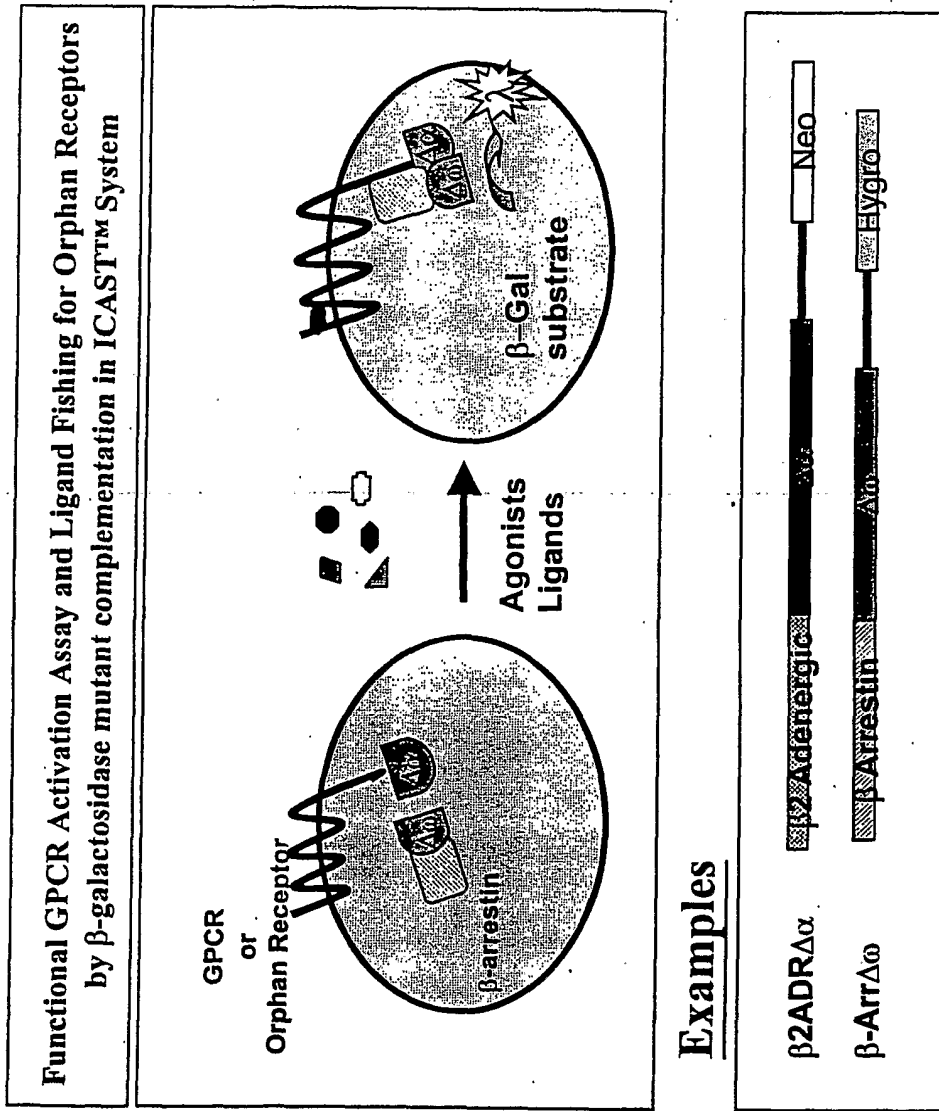
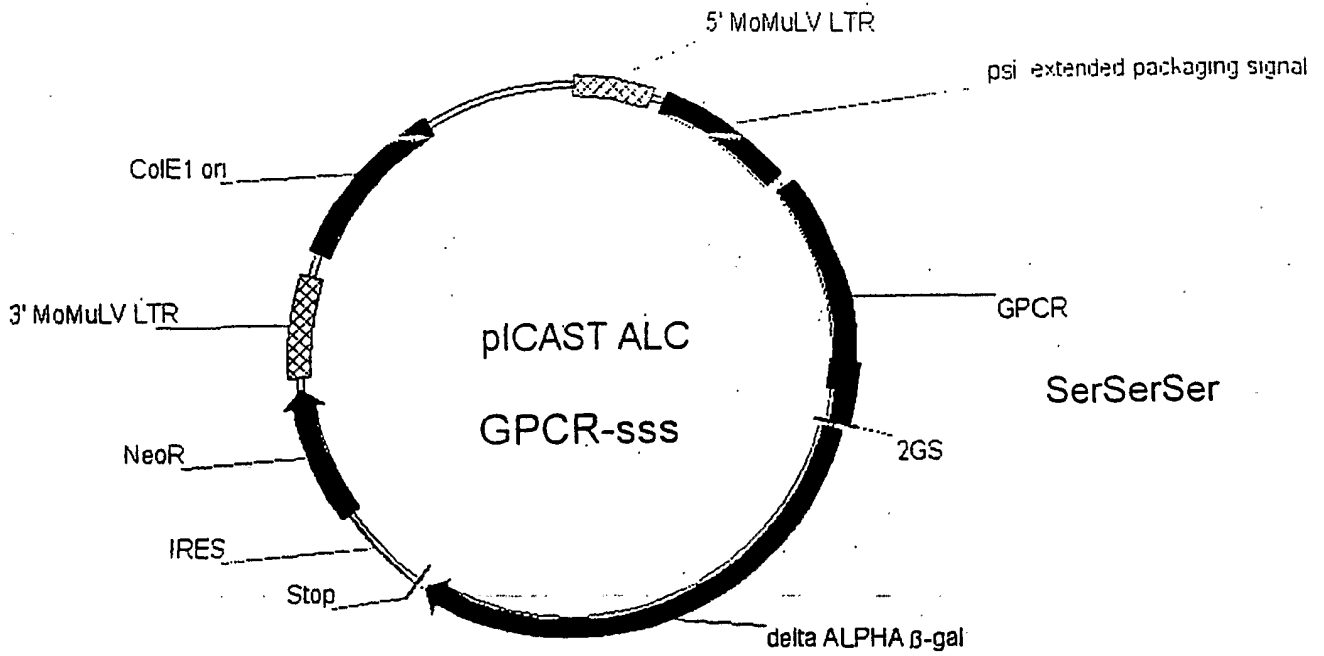
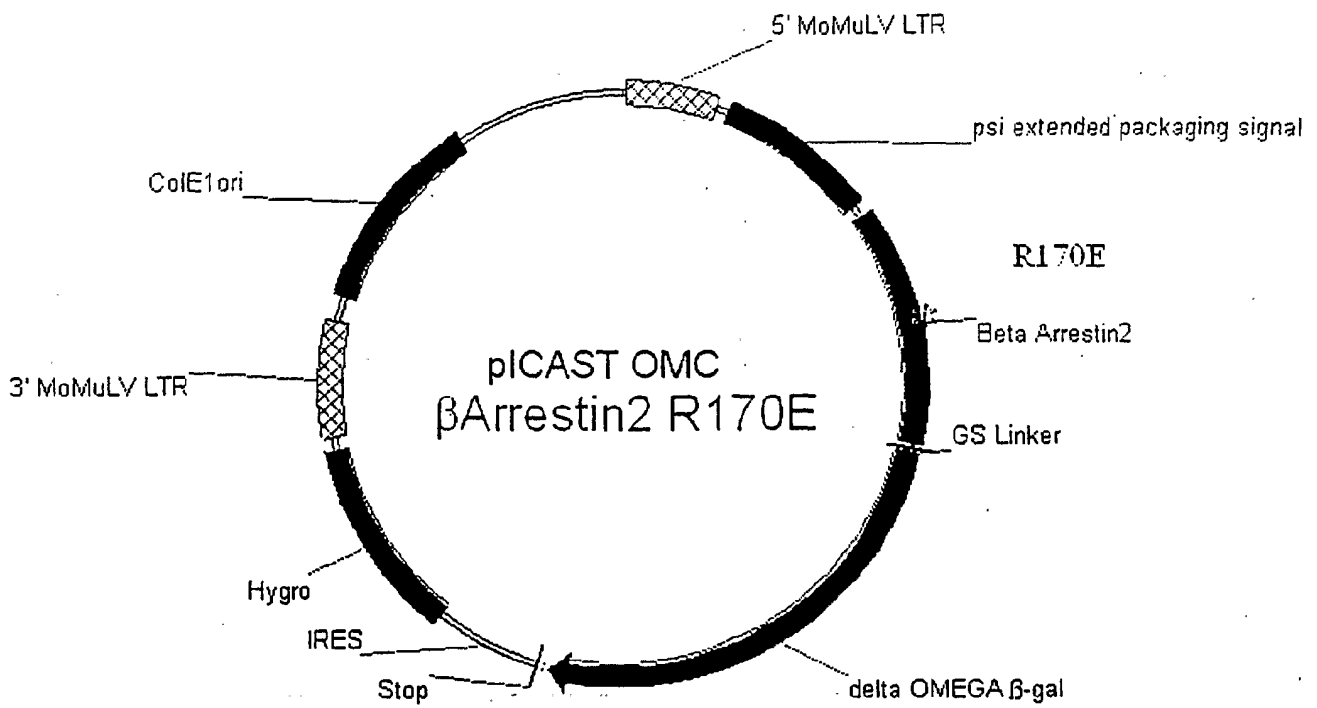


Figure 23



Vector for Expression of a GPCR with inserted Seronine/Threonine amino acid sequences as a fusion with β -gal $\Delta\alpha$.

FIGURE 24



Vector for Expression of mutant (R170E) β-arrestin2 as a fusion with β-gal Δω.

FIGURE 25

**Phosphorylation Insensitive Mutant R170E β -Arrestin2 $\Delta\omega$
Binds to β_2 AR $\Delta\alpha$ in Response to Agonist Activation**

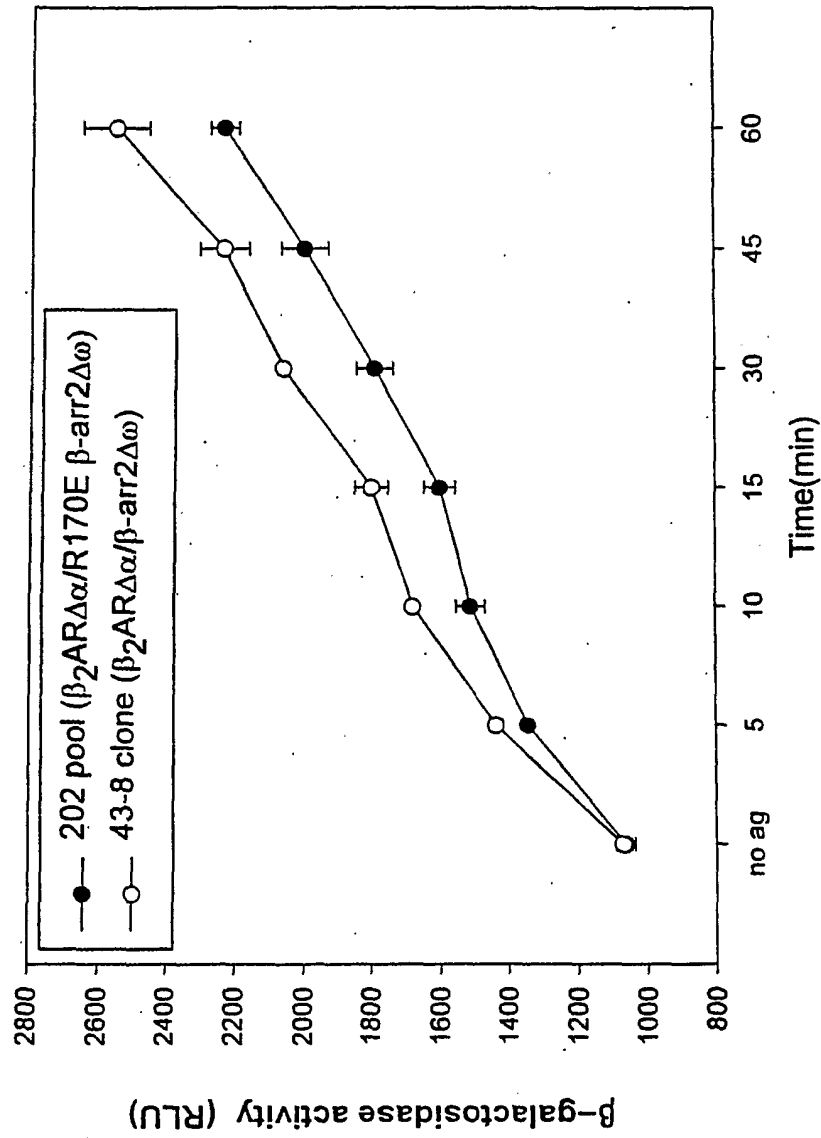
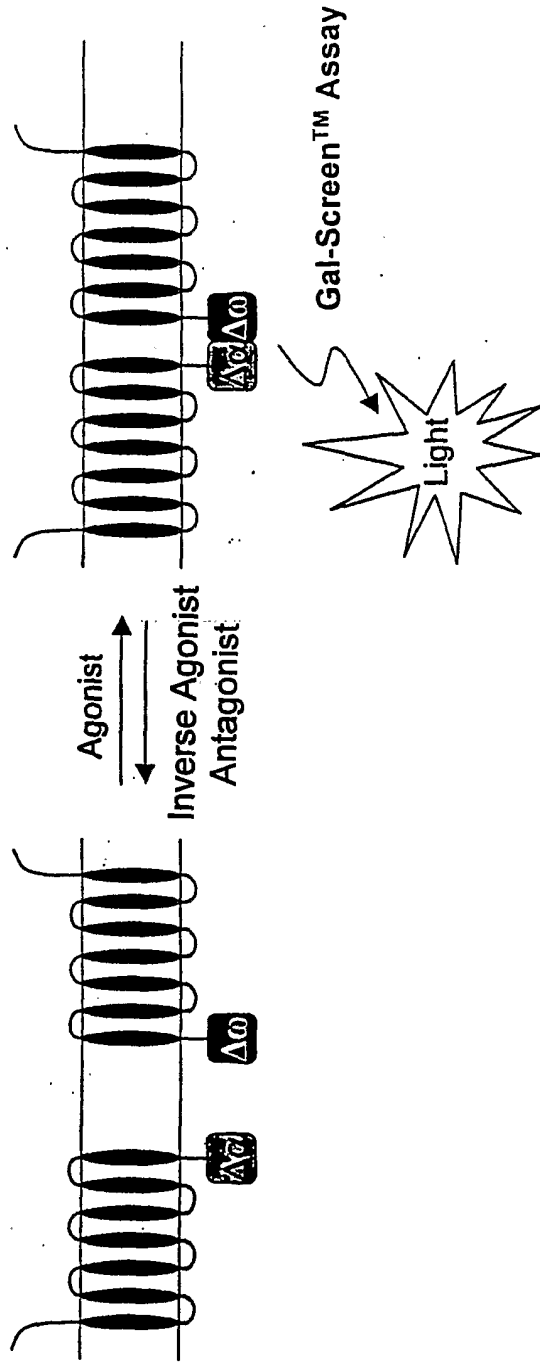


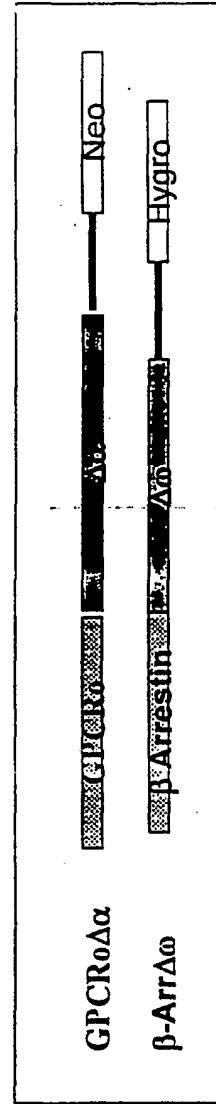
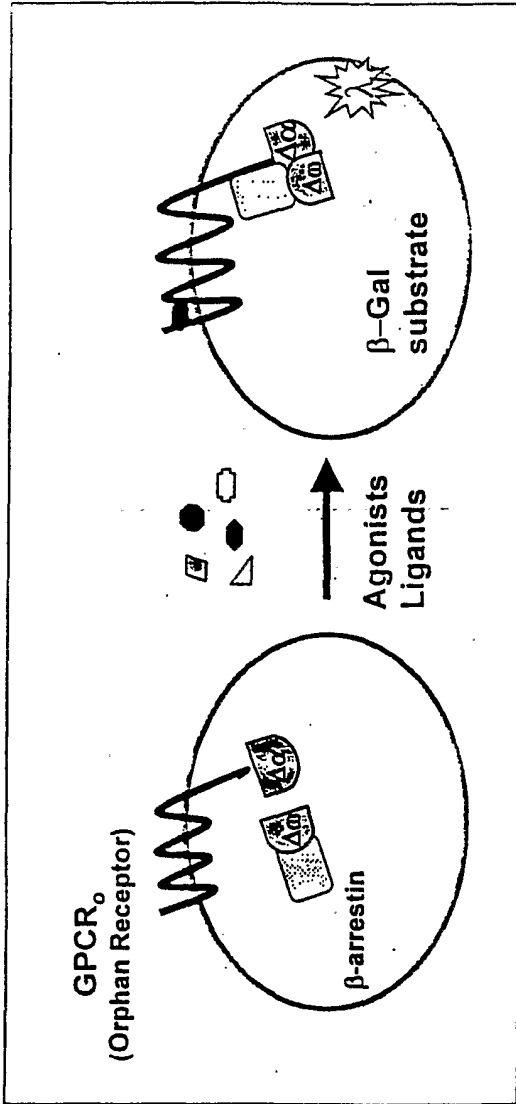
FIGURE 26



GPCR dimerization measured by β -gal complementation

FIGURE 27

Example-



Ligand Fishing for Orphan Receptors by β-galactosidase mutant complementation in ICASTM System

FIGURE 28