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(56) Related Art
MORGAN ET AL: "Characterization and optimization of RGD-containing silk blends to support osteoblastic differentiation", BIOMATERIALS, vol. 29, no. 16, 5 March 2008, pages 2556-2563
ANNA RISING ET AL: "Spider silk proteins: recent advances in recombinant production, structure-function relationships and biomedical applications", CMLS CELLULAR AND MOLECULAR LIFE SCIENCES, vol. 68, no. 2, 29 July 2010, pages 169-184
D. Huemmerich et al. "Primary Structure Elements of Spider Dragline Silks and Their Contribution to Protein Solubility", Biochemistry, Vol 43, pages 13604-13612, (2004)
J. Nilsson et al. "Affinity Fusion Strategies for Detection, Purification, and Immobilization of Recombinant Proteins", Protein Expression and Purification, Vol 11, No 1, pages 1-16, (October 1997)

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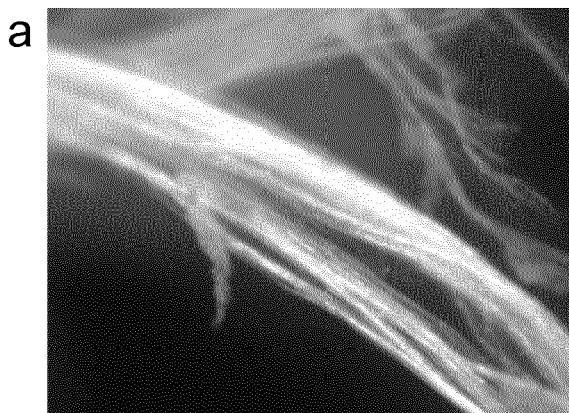


Fig 3

(57) Abstract: A recombinant fusion protein comprising the moieties Band CTis provided. B is a non-spidroin moiety which provides the capacity of selective interaction with an organic target. CTis a moiety of from 70 to 120 amino acid residues and is derived from the C-terminal fragment of a spider silk protein. The fusion protein is not comprising any moiety derived from the repetitive fragment of a spider silk protein.

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SPIDER SILK FUSION PROTEIN STRUCTURES WITHOUT REPETITIVE FRAGMENT FOR BINDING TO AN ORGANIC TARGET

Technical field of the invention

The present invention relates to the field of recombinant fusion proteins, and more specifically to novel fusion proteins comprising moieties derived from spider silk proteins (spidroins). The present invention provides methods for providing a protein structure which is a polymer comprising a recombinant fusion protein, which is comprising moieties derived from spidroins. There is also provided novel protein structures for binding to an organic target.

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Background to the invention

In applied protein chemistry, it is a common problem how to formulate or present a biologically active peptide or protein to the relevant site of activity, typically an organic target, such as a nucleic acid, a protein, a complex of proteins, or a complex of a protein(s) and/or lipids and/or carbohydrates and/or a nucleic acid(s). The simplest solution is simply to provide an aqueous solution of the biologically active peptide or protein. Many applications do however require some further means to achieve the desired goal. For instance, the peptides/proteins may be associated with a lipid mixture or chemically immobilized to a support structure.

Applications for peptides/proteins immobilized to a support structure include preparative and analytical separation procedures, such as bioprocesses, chromatography, cell capture and culture, active filters, and diagnostics. Structures based on extracellular matrix proteins, e.g. collagen, are disclosed in EP 704 532 and EP 985 732.

It has also been suggested to use spider silk proteins in a supporting structure. Spider silks are nature's high-performance polymers, obtaining extraordinary toughness and extensibility due to a combination of strength and elasticity. Spiders have up to seven different glands which produce a

variety of silk types with different mechanical properties and functions. Dragline silk, produced by the major ampullate gland, is the toughest fiber. It consists of two main polypeptides, mostly referred to as major ampullate spidroin (MaSp) 1 and 2, but e.g. as ADF-3 and ADF-4 in *Araneus diadematus*. These proteins have molecular masses in the range of 200-720 kDa. Spider dragline silk proteins, or MaSp, have a tripartite composition; a non-repetitive N-terminal domain, a central repetitive region comprised of many iterated poly-Ala/Gly segments, and a non-repetitive C-terminal domain. It is generally believed that the repetitive region forms intermolecular contacts in the silk fibers, while the precise functions of the terminal domains are less clear. It is also believed that in association with fiber formation, the repetitive region undergoes a structural conversion from random coil and α -helical conformation to β -sheet structure. The C-terminal region of spidroins is generally conserved between spider species and silk types.

15 WO 07/078239 and Stark, M. et al., *Biomacromolecules* 8: 1695-1701, (2007) disclose a miniature spider silk protein consisting of a repetitive fragment with a high content of Ala and Gly and a C-terminal fragment of a protein, as well as soluble fusion proteins comprising the spider silk protein. Fibers of the spider silk protein are obtained spontaneously upon liberation of 20 the spider silk protein from its fusion partner.

Rising, A. et al., *CMLS* 68(2): 169-184 (2010) reviews advances in the production of spider silk proteins. Fibres are created from a miniaturized spidroin construct consisting of four repeats of a segment derived from the repetitive region and the non-repetitive C-terminal domain of a spider silk 25 protein.

US 2009/0263430 discloses chemical coupling of the enzyme β -galactosidase to films of a miniature spider silk protein. However, chemical coupling may require conditions which are unfavourable for protein stability and/or function. Proteins containing multiple repeats of a segment derived 30 from the repetitive region of spider silk proteins have been designed to include a RGD cell-binding segment (Bini, E et al., *Biomacromolecules* 7:3139-3145 (2006); Morgan et al., *Biomaterials* 29(16): 2556-2563 (2008)) and/or a R5 peptide (Wong Po Foo, C et al., *Proc Natl Acad Sci* 103 (25):

9428-9433 (2006)) or other protein segments involved in mineralization (Huang, J *et al.*, Biomaterials 28: 2358-2367 (2007); WO 2006/076711). In these prior art documents, films are formed by solubilizing the fusion proteins in the denaturing organic solvent hexafluoroisopropanol (HFIP) and drying.

US 2005/261479 A1 discloses a method of for purification of recombinant silk proteins consisting of a repetitive fragment and an affinity tag, involving magnetic affinity separation of individual silk proteins from complex mixtures without formation of silk protein fibers or other polymer structures.

Known supporting structures and associated techniques have certain drawbacks with regard to e.g. economy, efficiency, stability, regenerating capacity, bioactivity and biocompatibility.

Summary of the Invention

According to one aspect of the present invention provides a recombinant fusion protein comprising the moieties B and CT, wherein: B is a non-spidroin moiety which provides the capacity of selective interaction with an organic target, wherein the B moiety is a protein or polypeptide fragment comprising more than 30 amino acid residues; and wherein the B moiety has less than 30% identity to any of SEQ ID NOS: 6-10; and CT is a moiety of from 70 to 120 amino acid residues and is derived from the C-terminal fragment of a spider silk protein has at least 80% identity to any one of SEQ ID NO: 7 and SEQ ID NOS: 14-44; wherein the CT moiety provides the capacity of forming a polymer; with the proviso that the fusion protein is not comprising any moiety derived from the repetitive fragment of a spider silk protein.

It is an aspect of the present invention to provide novel recombinant fusion proteins. In particular, it is an aspect of the present invention to provide novel recombinant fusion proteins which are capable of selective interaction with an organic target.

It is an aspect of the present invention to provide a novel protein structure that is capable of selective interaction with an organic target.

It is also an aspect of the present invention to provide a protein structure that is capable of selective interaction with an organic target,

wherein the structure is formed without use of harsh solvents which may have

an unpredictable effect on the secondary structure or activity of the protein and/or remain in the protein structure.

It is one aspect of the present invention to provide a stable protein structure that is capable of selective interaction with an organic target, which protein structure can readily be regenerated after use, e.g. with chemical treatment.

It is another aspect of the present invention to provide a stable protein structure that is biocompatible and suitable for cell culture and as an implant.

It is yet another aspect of the invention to provide a protein structure with a high density of evenly spaced functionalities that are capable of selective interaction with an organic target.

It is a further aspect of the invention to provide a protein structure which maintains its selective binding ability upon storage at +4°C or at room temperature for months.

It is also an aspect of the invention to provide a protein structure which is autoclavable, i.e. maintains its selective binding ability after sterilizing heat treatment.

For these and other aspects that will be evident from the following disclosure, the present invention provides according to a first aspect a fusion protein and a protein structure consisting of polymers comprising as a repeating structural unit the fusion protein as set out in the claims.

According to a related aspect, the present invention provides an isolated nucleic acid encoding the fusion protein and a method of producing the fusion protein as set out in the claims.

The present invention provides according to another aspect a method for providing a protein structure as set out in the claims.

The present invention provides according to a further aspect an affinity medium as set out in the claims.

The present invention provides according to one aspect a cell scaffold material as set

out in the claims. According to a related aspect, the present invention also provides a combination of cells and a cell scaffold material according to the claims.

The present invention provides according to an aspect novel uses of a protein structure and a fusion protein as set out in the claims.

The present invention provides according to another aspect a method for separation of an organic target from a sample as set out in the claims.

The present invention provides according to a further aspect a method for immobilization and optionally cultivation of cells as set out in the claims.

Brief description of the drawings

Fig. 1 shows a sequence alignment of spidroin C-terminal domains.

Fig. 2 shows a sequence alignment of spidroin N-terminal domains.

Fig. 3 shows microscopic pictures of silk fused ABD in fiber format.

5 Fig. 4 shows SDS-PAGE gels of fractions of human blood plasma eluted from ABD-NTCT and ABD-CT films.

Fig. 5 shows microscopic pictures of silk fused M4 fragments in fiber and foam format.

Fig. 6 shows fluorescence microscopy pictures of Atto-565-biotin 10 bound to M4-NTCT and M4-CT films.

Fig. 7 shows microscopic pictures of silk fused antibody fragments in foam format.

Fig. 8 shows an antigen binding analysis of pure and silk fused antibody fragments.

15 Fig. 9 shows a macroscopic foam of silk fused xylanase.

List of appended sequencesSEQ ID NO

1	4Rep
2	4RepCT
3	NT4Rep
4	NT5Rep
5	NT4RepCTHis
6	NT
7	CT
8	consensus NT sequence
9	consensus CT sequence
10	repetitive sequence from <i>Euprosthenops australis</i> MaSp1
11	consensus G segment sequence 1
12	consensus G segment sequence 2
13	consensus G segment sequence 3
14	CT <i>Euprosthenops</i> sp MaSp1
15	CT <i>Euprosthenops australis</i> MaSp1

SEQ ID NO

16 CT *Argiope trifasciata* MaSp1
17 CT *Cyrtophora moluccensis* Sp1
18 CT *Latrodectus geometricus* MaSp1
19 CT *Latrodectus hesperus* MaSp1
20 CT *Macrothele holsti* Sp1
21 CT *Nephila clavipes* MaSp1
22 CT *Nephila pilipes* MaSp1
23 CT *Nephila madagascariensis* MaSp1
24 CT *Nephila senegalensis* MaSp1
25 CT *Octonoba varians* Sp1
26 CT *Psechrus sinensis* Sp1
27 CT *Tetragnatha kauaiensis* MaSp1
28 CT *Tetragnatha versicolor* MaSp1
29 CT *Araneus bicentenarius* Sp2
30 CT *Argiope amoena* MaSp2
31 CT *Argiope aurantia* MaSp2
32 CT *Argiope trifasciata* MaSp2
33 CT *Gasteracantha mammosa* MaSp2
34 CT *Latrodectus geometricus* MaSp2
35 CT *Latrodectus hesperus* MaSp2
36 CT *Nephila clavipes* MaSp2
37 CT *Nephila madagascariensis* MaSp2
38 CT *Nephila senegalensis* MaSp2
39 CT *Dolomedes tenebrosus* Fb1
40 CT *Dolomedes tenebrosus* Fb2
41 CT *Araneus diadematus* ADF-1
42 CT *Araneus diadematus* ADF-2
43 CT *Araneus diadematus* ADF-3
44 CT *Araneus diadematus* ADF-4
45 NT *Euprosthenops australis* MaSp1
46 NT *Latrodectus geometricus* MaSp1
47 NT *Latrodectus hesperus* MaSp1

SEQ ID NO

48 NT *Nephila clavipes* MaSp1
49 NT *Argiope trifasciata* MaSp2
50 NT *Latrodectus geometricus* MaSp2
51 NT *Latrodectus hesperus* MaSp2
52 NT *Nephila inaurata madagascariensis* MaSp2
53 NT *Nephila clavipes* MaSp2
54 NT *Argiope bruennichi* cylindriform spidroin 1
55 NT *Nephila clavata* cylindriform spidroin 1
56 NT *Latrodectus hesperus* tubuliform spidroin
57 NT *Nephila clavipes* flagelliform silk protein
58 NT *Nephila inaurata madagascariensis* flagelliform silk protein
59 His₆NT-CT
60 His₆NTNT-CT
61 His₆Z-CT
62 His₆CT-Z
63 His₆Z-NTCT
64 His₆NTCT-Z
65 His₆Z-NTNTCT
66 His₆NTNTCT-Z
67 His₆-ABD-NTCT (DNA)
68 His₆-ABD-NTCT
69 His₆-ABD-CT (DNA)
70 His₆-ABD-CT
71 His₆-M4-NTCT (DNA)
72 His₆-M4-NTCT
73 His₆-M4-CT (DNA)
74 His₆-M4-CT
75 His₆-scFv1-NTCT (DNA)
76 His₆-scFv1-NTCT
77 His₆-scFv1-CT (DNA)
78 His₆-scFv1-CT
79 His₆Xyl-NTCT (DNA)

SEQ ID NO

80	His ₆ Xyl-NTCT
81	His ₆ Xyl-CT (DNA)
82	His ₆ Xyl-CT
83	His ₆ EGF-NTCT (DNA)
84	His ₆ EGF-NTCT
85	His ₆ EGF-CT (DNA)
86	His ₆ EGF-CT

Detailed description of the invention

The present invention is generally based on the insight that solid protein structures capable of selective interaction with an organic target can 5 be prepared in the form of polymers of a recombinant fusion protein as a repeating structural unit. The fusion protein is comprising at least one non-spidroin moiety, preferably of more than 30 amino acid residues, that is capable of selective interaction with the organic target, and moieties corresponding to at least the C-terminal fragment of a spider silk protein, with 10 the proviso that the fusion protein is not comprising any moiety derived from the repetitive fragment of a spider silk protein.

Surprisingly, the moiety derived from the spider silk protein can be induced to rearrange structurally and as a result form polymeric, solid structures, while the non-spidroin moiety is not structurally rearranged but 15 maintains its desirable structure and function, i.e. capability of selective interaction with the organic target. The protein structures can be obtained without a chemical coupling step or a denaturing method step, which facilitates the procedure and improves the chances of obtaining a fusion protein with maintained functionality of its moieties, in particular when the 20 functions are dependent on the secondary structure of the moieties. The formation of these fusion protein polymers can be tightly controlled, and this insight has been developed into further novel protein structures, methods of producing the protein structures and uses of the protein structures in various applications and methods.

The fusion protein according to the invention thus harbors both the desired selective interaction activity and an internal solid support activity that is employed in the protein structure under physiological conditions. It must be considered as surprising that the binding activity of the fusion protein is

5 maintained although the non-spidroin moiety is covalently attached to the spidroin moiety when the latter is structurally rearranged to form polymeric, solid structures. In fact, the heat and/or chemical stability and/or binding activity of the moiety providing the selective interaction activity may be increased when integrated in a fusion protein structure according to the

10 invention. The protein structure also provides a high and predictable density of the selective interaction activity towards an organic target. Losses of valuable protein moieties with selective interaction activity are minimized, since all expressed protein moieties are associated with the solid support.

The polymers which are formed from the fusion proteins according to

15 the invention are solid structures and are useful for their physical properties, especially the useful combination of high strength, elasticity and light weight. A particularly useful feature is that the spidroin-derived moieties of the fusion protein are biochemically robust and suitable for regeneration, e.g. with acid, base or chaotropic agents, and suitable for heat sterilization, e.g. autoclaving

20 at 120°C for 20 min. The polymers are also useful for their ability to support cell adherence and growth. The properties derived from dragline silk are attractive in development of new materials for medical or technical purposes. In particular, protein structures according to the invention are useful in preparative and analytical separation procedures, such as chromatography,

25 cell capture, selection and culture, active filters, and diagnostics. Protein structures according to the invention are also useful in medical devices, such as implants and medical products, such as wound closure systems, band-aids, sutures, wound dressings, and scaffolds for cell immobilization, cell culture, tissue engineering and guided cell regeneration.

30 The present invention provides a recombinant fusion protein that is capable of selective interaction with an organic target, which fusion protein is comprising the moieties **B** and **CT**, and optionally **NT**. The present invention also provides a protein structure that is capable of selective interaction with

an organic target, wherein said protein structure is a polymer comprising, and optionally consisting of, the recombinant fusion protein according to the invention, i.e. comprising, and optionally consisting of, the moieties **B** and **CT**, and optionally **NT**. The fusion protein according to the invention is not

5 comprising any moiety derived from the repetitive (**REP**) fragment of a spider silk protein.

Although the **CT** and the optional **NT** moieties of the fusion proteins in the examples by necessity relate to specific proteins, e.g. proteins derived from major spidroin 1 (MaSp1) from *Euprosthenops australis*, it is considered

10 that the present disclosure is applicable to any structurally similar moieties for the purpose of producing fusion protein structures according to the invention. Furthermore, although the **B** moiety which provides the selective interaction activity of the fusion proteins in the examples by necessity relate to specific protein moieties, e.g. moieties derived from protein A, protein G and

15 streptavidin, it is considered that the present disclosure is applicable to any structurally and/or functionally similar **B** moiety for the purpose of producing fusion protein structures according to the invention, capable of selective interaction with an organic target.

Specific fusion proteins according to the invention are defined by the

20 formulas **B_x-CT-B_z**,, wherein x and z are integers from 0 to 5; and x+z ≥ 1, optionally further containing one or more **NT** moieties, e.g. 1-2 **NT** moieties, at either end of the fusion protein or between any two protein moieties in the fusion protein, e.g. **NT-B_x-CT-B_z**, **B_x-NT-CT-B_z**, **B_x-CT-NT-B_z**, **B_x-CT-B_z-NT**, **NT-NT-B_x-CT-B_z** or **B_x-CT-B_z-NT-NT**,. If x+z > 1, i.e. if there are two or more

25 **B** moieties, they may be identical or different. The two or more **B** moieties may have capacity of selective interaction with the same organic target or with different organic targets. It is preferred that the two or more **B** moieties are substantially identical, each having capacity of selective interaction with the same organic target. Alternatively, it is preferred that the two or more **B**

30 moieties are not identical, and that they together provide the capacity of selective interaction with the organic target.

In preferred fusion proteins according to the invention, x and z are integers from 0 to 2, preferably from 0 to 1. In certain preferred fusion proteins

according to the invention, either x or z are 0, i.e. the fusion proteins are defined by the formulas **B_x-CT**, and **CT-B_z**, wherein either x or z is an integer from 1 to 5, and optionally containing 1-2 **NT** moieties.. In preferred fusion proteins according to the invention, x and z are integers from 0 to 1; and x+z = 1. Thus, certain preferred fusion proteins according to the invention are defined by the formulas **B-CT**, and **CT- B**, optionally containing 1-2 **NT** moieties. In preferred fusion proteins according to the invention, the optional **NT** moiety is missing.

The term "fusion protein" implies here a protein that is made by expression from a recombinant nucleic acid, i.e. DNA or RNA that is created artificially by combining two or more nucleic acid sequences that would not normally occur together (genetic engineering). The fusion proteins according to the invention are recombinant proteins, and they are therefore not identical to naturally occurring proteins. In particular, wildtype spidroins are not fusion proteins according to the invention, because they are not expressed from a recombinant nucleic acid as set out above. The combined nucleic acid sequences encode different proteins, partial proteins or polypeptides with certain functional properties. The resulting fusion protein, or recombinant fusion protein, is a single protein with functional properties derived from each of the original proteins, partial proteins or polypeptides. Furthermore, the fusion protein according to the invention and the corresponding genes are chimeric, i.e. the protein/gene moieties are derived from at least two different species. The **CT** moiety and the optional **NT** moiety are derived from a spider silk protein. For avoidance of doubt, the **B** moiety according to the invention is a non-spidroin protein or polypeptide, i.e. it is not derived from a spider silk protein. In particular, the **B** moiety according to the invention is not derived from the C-terminal, repetitive or N-terminal fragments of a spider silk protein.

The fusion protein typically consists of from 170 to 2000 amino acid residues, such as from 170 to 1000 amino acid residues, such as from 170 to 600 amino acid residues, preferably from 170 to 500 amino acid residues, such as from 170 to 400 amino acid residues. The small size is advantageous because longer proteins containing spider silk protein fragments may form amorphous aggregates, which require use of harsh solvents for solubilisation

and polymerisation. The recombinant fusion protein may contain more than 2000 residues, in particular in cases where the spider silk protein more than one **B** moiety and/or when it contains a **NT** moiety, e.g. 1-2 **NT** moieties.

The terms "spidroins" and "spider silk proteins" are used

5 interchangeably throughout the description and encompass all known spider silk proteins, including major ampullate spider silk proteins which typically are abbreviated "MaSp", or "ADF" in the case of *Araneus diadematus*. These major ampullate spider silk proteins are generally of two types, 1 and 2. These terms furthermore include non-natural proteins with a high degree of

10 identity and/or similarity to the known spider silk proteins.

Consequently, the term "non-spidroin" implies proteins that are not derived from a spider silk protein, i.e. with a low (or no) degree of identity and/or similarity to spider silk proteins.

The protein structure according to the invention is capable of selective

15 interaction with an organic target. This capacity resides in the fusion protein according to the invention, and more specifically in the **B** moiety of the fusion protein. Any interactions of the **CT** moiety and the optional **NT** moiety with organic molecules are not encompassed by the term "capable of selective interaction with an organic target". For avoidance of doubt, the term "capable

20 of selective interaction with an organic target" does not encompass dimerization, oligomerization or polymerization of the fusion proteins according to the invention that rely on interactions involving the **CT** moiety and/or the optional **NT** moiety.

The term "organic target" encompasses all chemical molecules

25 containing carbon with the exception of what is traditionally considered inorganic molecules by the skilled person, e.g. carbonates, simple oxides of carbon, cyanides, diamond and graphite. For avoidance of doubt, inorganic molecules, salts and ions, such as silica and calcium chloride, are not organic. The organic target may be a complex containing or consisting of

30 organic molecules, e.g. a receptor complex on a cell surface. The organic target may be a monomer, dimer, oligomer or polymer of one or more organic molecule types, which may be held together by covalent bonds or other types of association. It may of course also simply be a single organic molecule.

Preferred organic targets according to the invention include, but are not limited to, nucleic acids, proteins and polypeptides, lipids and carbohydrates, as well as combinations thereof. Further preferred organic targets according to the invention include, but are not limited to, immunoglobulins, molecules 5 comprising immunoglobulin or derivatives thereof, albumin, molecules comprising albumin or derivatives thereof, biotin, molecules comprising biotin or derivatives or analogues thereof, and biological disease markers, e.g. from blood, serum, urine, saliva or other samples from body tissues.

In the context of the present invention, "specific" or "selective" 10 interaction of a ligand, e.g. a **B** moiety of the fusion protein according to the invention with its target means that the interaction is such that a distinction between specific and non-specific, or between selective and non-selective, interaction becomes meaningful. The interaction between two proteins is sometimes measured by the dissociation constant. The dissociation constant 15 describes the strength of binding (or affinity) between two molecules.

Typically the dissociation constant between an antibody and its antigen is from 10^{-7} to 10^{-11} M. However, high specificity does not necessarily require high affinity. Molecules with low affinity (in the molar range) for its counterpart have been shown to be as specific as molecules with much higher affinity. In 20 the case of the present invention, a specific or selective interaction refers to the extent to which a particular method can be used to determine the presence and/or amount of a specific protein, the target protein or a fragment thereof, under given conditions in the presence of other proteins in a sample of a naturally occurring or processed biological or biochemical fluid. In other 25 words, specificity or selectivity is the capacity to distinguish between related proteins. Specific and selective are sometimes used interchangeably in the present description.

The fusion protein according to the invention may also contain one or 30 more linker peptides. The linker peptide(s) may be arranged between any moieties of the fusion protein, e.g. between the **CT** and **NT** moieties, between two **B** moieties, and between **B** and **CT** moieties, or may be arranged at either terminal end of the fusion protein. If the fusion protein contains two or more **B** moieties, the linker peptide(s) may also be arranged in between two

B moieties. The linker(s) may provide a spacer between the functional units of the fusion protein, but may also constitute a handle for identification and purification of the fusion protein, e.g. a His and/or a Trx tag. If the fusion protein contains two or more linker peptides for identification and purification of the fusion protein, it is preferred that they are separated by a spacer sequence, e.g. His₆-spacer-His₆-. The linker may also constitute a signal peptide, such as a signal recognition particle, which directs the fusion protein to the membrane and/or causes secretion of the fusion protein from the host cell into the surrounding medium. The fusion protein may also include a cleavage site in its amino acid sequence, which allows for cleavage and removal of the linker(s) and/or other relevant moieties, typically the **B** moiety or moieties. Various cleavage sites are known to the person skilled in the art, e.g. cleavage sites for chemical agents, such as CNBr after Met residues and hydroxylamine between Asn-Gly residues, cleavage sites for proteases, such as thrombin or protease 3C, and self-splicing sequences, such as intein self-splicing sequences.

The **CT** and **B** moieties are linked directly or indirectly to one another. A direct linkage implies a direct covalent binding between the moieties without intervening sequences, such as linkers. An indirect linkage also implies that the moieties are linked by covalent bonds, but that there are intervening sequences, such as linkers and/or one or more further moieties, e.g. 1-2 **NT** moieties.

The **B** moiety or moieties may be arranged internally or at either end of the fusion protein, i.e. C-terminally arranged or N-terminally arranged. It is preferred that the **B** moiety or moieties are arranged at the N-terminal end of the fusion protein. If the fusion protein contains one or more linker peptide(s) for identification and purification of the fusion protein, e.g. a His or Trx tag(s), it is preferred that it is arranged at the N-terminal end of the fusion protein.

A preferred fusion protein has the form of an N-terminally arranged **B** moiety, coupled by a linker peptide of 1-30 amino acid residues, such as 1-10 amino acid residues, to a C-terminally arranged **CT** moiety. The linker peptide may contain a cleavage site. Optionally, the fusion protein has an N-terminal

or C-terminal linker peptide, which may contain a purification tag, such as a His tag, and a cleavage site.

Another preferred fusion protein has the form of an N-terminally arranged **B** moiety coupled directly to a C-terminally arranged **CT** moiety.

5 Optionally, the fusion protein has an N-terminal or C-terminal linker peptide, which may contain a purification tag, such as a His tag, and a cleavage site.

The protein structure according to the invention is a polymer comprising as a repeating structural unit recombinant fusion proteins according to the invention, which implies that it contains an ordered plurality 10 of fusion proteins according to the invention, typically well above 100 fusion protein units, e.g. 1000 fusion protein units or more. Optionally, the polymer may comprise as a further repeating structural unit complementary proteins without a **B** moiety, preferably proteins derived from spider silk. This may be advantageous if the **B** moiety of the fusion protein is large and/or bulky.

15 These complementary proteins typically comprise, and may even consist of, a **CT** moiety, and optionally a further **NT** moiety, e.g. 1-2 **NT** moieties. Preferred complementary proteins according to the invention can have any of the structures set out herein with a deleted **B** moiety. It is preferred that the complementary fusion protein is substantially identical to the fusion protein 20 with a deleted **B** moiety. However, it is preferred that the protein structure according to the invention is a polymer consisting of recombinant fusion proteins according to the invention as a repeating structural unit, i.e. that the protein structure according to the invention is a polymer of the recombinant fusion protein according to the invention.

25 The magnitude of fusion units in the polymer implies that the protein structure obtains a significant size. In a preferred embodiment, the protein structure has a size of at least 0.1 µm in at least two dimensions. Thus, the term "protein structure" as used herein relates to fusion protein polymers having a thickness of at least 0.1 µm, preferably macroscopic polymers that 30 are visible to the human eye, i.e. having a thickness of at least 1 µm. The term "protein structure" does not encompass unstructured aggregates or precipitates. While monomers of the fusion protein are water soluble, it is understood that the protein structures according to the invention are solid

structures, i.e. not soluble in water. The protein structures are polymers comprising as a repeating structural unit monomers of the recombinant fusion proteins according to the invention.

It is preferable that the protein structure according to the invention is in
5 a physical form selected from the group consisting of fiber, film, foam, net,
mesh, sphere and capsule.

It is preferable that the protein structure according to the invention is a
fiber or film with a thickness of at least 1 nm or at least 0.1 μ m, preferably at
least 1 μ m. It is preferred that the fiber or film has a thickness in the range of
10 1 nm -400 m, such as 1-400 μ m, and preferably 60-120 μ m. It is preferred
that fibers have a length in the range of 0.5-300 cm, preferably 1-100 cm.
Other preferred ranges are 0.5-30 cm and 1-20 cm. The fiber has the capacity
to remain intact during physical manipulation, i.e. can be used for spinning,
weaving, twisting, crocheting and similar procedures. The film is
15 advantageous in that it is coherent and adheres to solid structures, e.g. the
plastics in microtiter plates. This property of the film facilitates washing and
regeneration procedures and is very useful for separation purposes. A
particularly useful protein structure is a film or a fiber wherein the **B** moiety is
the Z domain derived from staphylococcal protein A or a protein fragment
20 having at least 70% identity thereto.

It is also preferred that the protein structure according to the invention
has a tensile strength above 1 MPa, preferably above 2 MPa, more preferably
10 MPa or higher. It is preferred that the protein structure according to the
invention has a tensile strength above 100 MPa, more preferably 200 MPa or
25 higher.

The term "% identity", as used throughout the specification and the
appended claims, is calculated as follows. The query sequence is aligned to
the target sequence using the CLUSTAL W algorithm (Thompson, J.D.,
Higgins, D.G. and Gibson, T.J., Nucleic Acids Research, 22: 4673-4680
30 (1994)). A comparison is made over the window corresponding to the shortest
of the aligned sequences. The amino acid residues at each position are
compared, and the percentage of positions in the query sequence that have
identical correspondences in the target sequence is reported as % identity.

The term "% similarity", as used throughout the specification and the appended claims, is calculated as described for "% identity", with the exception that the hydrophobic residues Ala, Val, Phe, Pro, Leu, Ile, Trp, Met and Cys are similar; the basic residues Lys, Arg and His are similar; the acidic residues Glu and Asp are similar; and the hydrophilic, uncharged residues Gln, Asn, Ser, Thr and Tyr are similar. The remaining natural amino acid Gly is not similar to any other amino acid in this context.

Throughout this description, alternative embodiments according to the invention fulfill, instead of the specified percentage of identity, the corresponding percentage of similarity. Other alternative embodiments fulfill the specified percentage of identity as well as another, higher percentage of similarity, selected from the group of preferred percentages of identity for each sequence. For example, a sequence may be 70% similar to another sequence; or it may be 70% identical to another sequence; or it may be 70% identical and 90% similar to another sequence.

The **CT** moiety is a protein fragment containing from 70 to 120 amino acid residues and is derived from the C-terminal fragment of a spider silk protein. The expression "derived from" implies in the context of the **CT** moiety according to the invention that it has a high degree of similarity to the C-terminal amino acid sequence of spider silk proteins. As shown in Fig 1, this amino acid sequence is well conserved among various species and spider silk proteins, including MaSp1 and MaSp2. A consensus sequence of the C-terminal regions of MaSp1 and MaSp2 is provided as SEQ ID NO: 9. In Fig 1, the following MaSp proteins are aligned, denoted with GenBank accession entries where applicable (SEQ ID NOS: 14-44):

TABLE 1 - Spidroin **CT** moieties

<u>Species and spidroin protein</u>	<u>Entry</u>
<i>Euprosthenops</i> sp MaSp1 (Pouchkina-Stantcheva, NN & McQueen-Mason, SJ, <i>ibid</i>)	Cthyb_Esp
<i>Euprosthenops australis</i> MaSp1	CTnat_Eau
<i>Argiope trifasciata</i> MaSp1	AF350266_At1

<u>Species and spidroin protein</u>	<u>Entry</u>
<i>Cyrtophora moluccensis</i> Sp1	AY666062_Cm1
<i>Latrodectus geometricus</i> MaSp1	AF350273_Lg1
<i>Latrodectus hesperus</i> MaSp1	AY953074_Lh1
<i>Macrothele holsti</i> Sp1	AY666068_Mh1
<i>Nephila clavipes</i> MaSp1	U20329_Nc1
<i>Nephila pilipes</i> MaSp1	AY666076_Np1
<i>Nephila madagascariensis</i> MaSp1	AF350277_Nm1
<i>Nephila senegalensis</i> MaSp1	AF350279_Ns1
<i>Octonoba varians</i> Sp1	AY666057_Ov1
<i>Psechrus sinensis</i> Sp1	AY666064_Ps1
<i>Tetragnatha kauaiensis</i> MaSp1	AF350285_Tk1
<i>Tetragnatha versicolor</i> MaSp1	AF350286_Tv1
<i>Araneus bidentatus</i> Sp2	ABU20328_Ab2
<i>Argiope amoena</i> MaSp2	AY365016_Aam2
<i>Argiope aurantia</i> MaSp2	AF350263_Aau2
<i>Argiope trifasciata</i> MaSp2	AF350267_At2
<i>Gasteracantha mammosa</i> MaSp2	AF350272_Gm2
<i>Latrodectus geometricus</i> MaSp2	AF350275_Lg2
<i>Latrodectus hesperus</i> MaSp2	AY953075_Lh2
<i>Nephila clavipes</i> MaSp2	AY654293_Nc2
<i>Nephila madagascariensis</i> MaSp2	AF350278_Nm2
<i>Nephila senegalensis</i> MaSp2	AF350280_Ns2
<i>Dolomedes tenebrosus</i> Fb1	AF350269_DtFb1
<i>Dolomedes tenebrosus</i> Fb2	AF350270_DtFb2
<i>Araneus diadematus</i> ADF-1	U47853_ADF1
<i>Araneus diadematus</i> ADF-2	U47854_ADF2
<i>Araneus diadematus</i> ADF-3	U47855_ADF3
<i>Araneus diadematus</i> ADF-4	U47856_ADF4

It is not critical which specific **CT** moiety is present in spider silk proteins according to the invention, as long as the **CT** moiety is not entirely missing. Thus, the **CT** moiety according to the invention can be selected from 5 any of the amino acid sequences shown in Fig 1 and Table 1 (SEQ ID NOS:

14-44) or sequences with a high degree of similarity. A wide variety of C-terminal sequences can be used in the spider silk protein according to the invention.

The sequence of the **CT** moiety according to the invention has at least 5 50% identity, preferably at least 60%, more preferably at least 65% identity, or even at least 70% identity, to the consensus amino acid sequence SEQ ID NO: 9, which is based on the amino acid sequences of Fig 1 (SEQ ID NOS: 14-44).

A representative **CT** moiety according to the invention is the 10 *Euprosthenops australis* sequence SEQ ID NO: 7. Thus, according to a preferred aspect of the invention, the **CT** moiety has at least 80%, preferably at least 90%, such as at least 95%, identity to SEQ ID NO: 7 or any individual amino acid sequence of Fig 1 and Table 1 (SEQ ID NOS: 14-44). In preferred aspects of the invention, the **CT** moiety is identical to SEQ ID NO: 7 or any 15 individual amino acid sequence of Fig 1 and Table 1.

The **CT** moiety typically consists of from 70 to 120 amino acid residues. It is preferred that the **CT** moiety contains at least 70, or more than 80, preferably more than 90, amino acid residues. It is also preferred that the **CT** moiety contains at most 120, or less than 110 amino acid residues. A 20 typical **CT** moiety contains approximately 100 amino acid residues.

The optional **NT** moiety is a protein fragment containing from 100 to 160 amino acid residues and is derived from the N-terminal fragment of a spider silk protein. The expression "derived from" implies in the context of the 25 **NT** moiety according to the invention that it has a high degree of similarity to the N-terminal amino acid sequence of spider silk proteins. As shown in Fig 2, this amino acid sequence is well conserved among various species and spider silk proteins, including MaSp1 and MaSp2. In Fig 2, the following spidroin **NT** moieties are aligned, denoted with GenBank accession entries 30 where applicable (SEQ ID NOS: 45-58):

TABLE 2 - Spidroin **NT** moieties

<u>Code</u>	<u>Species and spidroin protein</u>	<u>GenBank acc. no.</u>
Ea MaSp1	Euprosthenops australis MaSp 1	AM259067
Lg MaSp1	Latrodectus geometricus MaSp 1	ABY67420

<u>Code</u>	<u>Species and spidroin protein</u>	<u>GenBank acc. no.</u>
Lh MaSp1	<i>Latrodectus hesperus</i> MaSp 1	ABY67414
Nc MaSp1	<i>Nephila clavipes</i> MaSp 1	ACF19411
At MaSp2	<i>Argiope trifasciata</i> MaSp 2	AAZ15371
Lg MaSp2	<i>Latrodectus geometricus</i> MaSp 2	ABY67417
Lh MaSp2	<i>Latrodectus hesperus</i> MaSp 2	ABR68855
Nim MaSp2	<i>Nephila inaurata madagascariensis</i> MaSp 2	AAZ15322
Nc MaSp2	<i>Nephila clavipes</i> MaSp 2	ACF19413
Ab CySp1	<i>Argiope bruennichi</i> cylindriform spidroin 1	BAE86855
Ncl CySp1	<i>Nephila clavata</i> cylindriform spidroin 1	BAE54451
Lh TuSp1	<i>Latrodectus hesperus</i> tubuliform spidroin	ABD24296
Nc Flag	<i>Nephila clavipes</i> flagelliform silk protein	AF027972
Nim Flag	<i>Nephila inaurata madagascariensis</i> flagelliform silk protein	AF218623 (translated)

Only the part corresponding to the N-terminal moiety is shown for each sequence, omitting the signal peptide. Nc flag and Nim flag are translated and edited according to Rising A. et al. *Biomacromolecules* 7, 3120-3124 (2006)).

5 It is not critical which specific **NT** moiety is present in spider silk proteins according to the invention. Thus, the **NT** moiety according to the invention can be selected from any of the amino acid sequences shown in Fig 2 and Table 2 (SEQ ID NOS: 45-58) or sequences with a high degree of similarity. A wide variety of N-terminal sequences can be used in the spider 10 silk protein according to the invention. Based on the homologous sequences of Fig 2, the following sequence constitutes a consensus **NT** amino acid sequence:

15 QANTPWSSPNLADAFINSF(M/L)SA(A/I)SSSGAFSADQLDDMSTIG(D/N/Q)T LMSAMD(N/S/K)MGRSG(K/R)STKSKLQALNMAFASSMAEIAAESGG(G/Q) ANEV (SEQ ID NO: 8).

The sequence of the **NT** moiety according to the invention has at least 50% identity, preferably at least 60% identity, to the consensus amino acid sequence SEQ ID NO: 8, which is based on the amino acid sequences of Fig 20 2. In a preferred embodiment, the sequence of the **NT** moiety according to the invention has at least 65% identity, preferably at least 70% identity, to the consensus amino acid sequence SEQ ID NO: 8. In preferred embodiments,

the **NT** moiety according to the invention has furthermore 70%, preferably 80%, similarity to the consensus amino acid sequence SEQ ID NO: 8.

A representative **NT** moiety according to the invention is the *Euprosthenops australis* sequence SEQ ID NO: 6. According to a preferred 5 embodiment of the invention, the **NT** moiety has at least 80% identity to SEQ ID NO: 6 or any individual amino acid sequence in Fig 2 (SEQ ID NOS: 45-58). In preferred embodiments of the invention, the **NT** moiety has at least 90%, such as at least 95% identity, to SEQ ID NO: 6 or any individual amino acid sequence in Fig 2. In preferred embodiments of the invention, the **NT** 10 moiety is identical to SEQ ID NO: 6 or any individual amino acid sequence in Fig 2, in particular to Ea MaSp1 (SEQ ID NO: 45).

The **NT** moiety contains from 100 to 160 amino acid residues. It is preferred that the **NT** moiety contains at least 100, or more than 110, 15 preferably more than 120, amino acid residues. It is also preferred that the **NT** moiety contains at most 160, or less than 140 amino acid residues. A typical **NT** moiety contains approximately 130-140 amino acid residues.

The fusion protein is not comprising any moiety derived from the repetitive fragment of a spider silk protein. A typical moiety that is derived 20 from the repetitive fragment of a spider silk protein and thus void in the present fusion protein is a **REP** moiety, i.e. a protein fragment containing from 70 to 300 amino acid residues that is derived from the repetitive fragment of a spider silk protein. In fusion proteins lacking a REP moiety, non-specific binding of the B moiety to other molecules than its antigen target has 25 advantageously been observed to decrease even further. It is also particularly surprising that solid structures are formed spontaneously from fusion proteins lacking a REP moiety.

The **REP** moiety has a repetitive character, alternating between alanine-rich stretches and glycine-rich stretches. The **REP** moiety generally 30 contains more than 70, such as more than 140, and less than 300, preferably less than 240, such as less than 200, amino acid residues, and can itself be divided into several **L** (linker) segments, **A** (alanine-rich) segments and **G** (glycine-rich) segments, as will be explained in more detail below. Typically, said linker segments, which are optional, are located at the **REP** moiety 35 terminals, while the remaining segments are in turn alanine-rich and glycine-

rich. Thus, the **REP** moiety can generally have either of the following structures, wherein n is an integer:

L(AG)_nL, such as LA₁G₁A₂G₂A₃G₃A₄G₄A₅G₅L;

L(AG)_nAL, such as LA₁G₁A₂G₂A₃G₃A₄G₄A₅G₅A₆L;

5 L(GA)_nL, such as LG₁A₁G₂A₂G₃A₃G₄A₄G₅A₅L; or

L(GA)_nGL, such as LG₁A₁G₂A₂G₃A₃G₄A₄G₅A₅G₆L.

It follows that it is not critical whether an alanine-rich or a glycine-rich segment is adjacent to the N-terminal or C-terminal linker segments. It is preferred that n is an integer from 2 to 10, preferably from 2 to 8, preferably

10 from 4 to 8, more preferred from 4 to 6, i.e. n=4, n=5 or n=6.

The alanine content of the **REP** moiety is typically above 20%, preferably above 25%, more preferably above 30%, and below 50%, preferably below 40%, more preferably below 35%.

Now turning to the segments that constitute the **REP** moiety, it shall be
15 emphasized that each segment is individual, i.e. any two **A** segments, any two **G** segments or any two **L** segments of a specific **REP** moiety may be identical or may not be identical. Thus, it is not a general feature that each type of segment is identical within a specific **REP** moiety. Rather, the following disclosure provides the skilled person with guidelines how to identify
20 a **REP** moiety which is thereby considered to be derived from the repetitive fragment of a spider silk protein, and which does not constitute a part of a functional fusion protein according to the invention.

Each individual **A** segment is an amino acid sequence having from 8 to 18 amino acid residues. A vast majority of these amino acid residues are
25 alanine residues. More specifically, from 0 to 3 of the amino acid residues are not alanine residues, and the remaining amino acid residues are alanine residues. Thus, all amino acid residues in each individual **A** segment are alanine residues, with no exception or the exception of one, two or three amino acid residues, which can be any amino acid. The alanine-replacing
30 amino acid(s) is (are) natural amino acids, preferably individually selected from the group of serine, glutamic acid, cysteine and glycine, more preferably serine. Of course, it is possible that one or more of the **A** segments are all-alanine segments, while the remaining **A** segments contain 1-3 non-alanine residues, such as serine, glutamic acid, cysteine or glycine.

Each individual **A** segment typically has at least 80%, preferably at least 90%, more preferably 95%, most preferably 100% identity to an amino acid sequence selected from the group of amino acid residues 7-19, 43-56, 71-83, 107-120, 135-147, 171-183, 198-211, 235-248, 266-279, 294-306, 5 330-342, 357-370, 394-406, 421-434, 458-470, 489-502, 517-529, 553-566, 581-594, 618-630, 648-661, 676-688, 712-725, 740-752, 776-789, 804-816, 840-853, 868-880, 904-917, 932-945, 969-981, 999-1013, 1028-1042 and 1060-1073 of SEQ ID NO: 10. Each sequence of this group corresponds to a segment of the naturally occurring sequence of *Euprosthenops australis*

10 10 MaSp1 protein, which is deduced from cloning of the corresponding cDNA, see WO 2007/078239. Alternatively, each individual **A** segment has at least 80%, preferably at least 90%, more preferably 95%, most preferably 100% identity to an amino acid sequence selected from the group of amino acid residues 143-152, 174-186, 204-218, 233-247 and 265-278 of SEQ ID NO: 3.

15 15 Furthermore, it has been concluded from experimental data that each individual **G** segment is an amino acid sequence of from 12 to 30 amino acid residues. It is preferred that each individual **G** segment consists of from 14 to 23 amino acid residues. At least 40% of the amino acid residues of each **G** segment are glycine residues. Typically the glycine content of each individual 20 20 **G** segment is in the range of 40-60%.

Each individual **G** segment typically has at least 80%, preferably at least 90%, more preferably 95%, most preferably 100% identity to an amino acid sequence selected from the group of amino acid residues 20-42, 57-70, 84-106, 121-134, 148-170, 184-197, 212-234, 249-265, 280-293, 307-329, 25 343-356, 371-393, 407-420, 435-457, 471-488, 503-516, 530-552, 567-580, 595-617, 631-647, 662-675, 689-711, 726-739, 753-775, 790-803, 817-839, 854-867, 881-903, 918-931, 946-968, 982-998, 1014-1027, 1043-1059 and 1074-1092 of SEQ ID NO: 10. Each sequence of this group corresponds to a segment of the naturally occurring sequence of *Euprosthenops australis*

30 30 MaSp1 protein, which is deduced from cloning of the corresponding cDNA, see WO 2007/078239. Alternatively, each individual **G** segment has at least 80%, preferably at least 90%, more preferably 95%, most preferably 100% identity to an amino acid sequence selected from the group of amino acid

residues 153-173, 187-203, 219-232, 248-264 and 279-296 of SEQ ID NO: 3.

Each sequence of this group corresponds to a segment of expressed, non-natural spider silk proteins, which proteins have capacity to form silk structures under appropriate conditions. Thus, each individual **G** segment

5 may be identical to an amino acid sequence selected from the above-mentioned amino acid segments.

There are the three subtypes of the **G** segment. This classification is based upon careful analysis of the *Euprosthenops australis* MaSp1 protein sequence (WO 2007/078239), and the information has been employed and 10 verified in the construction of novel, non-natural spider silk proteins.

The first subtype of the **G** segment is represented by the amino acid one letter consensus sequence GQG(G/S)QGG(Q/Y)GG (L/Q)GQGGYGGQGA GSS (SEQ ID NO: 11). This first, and generally the longest, **G** segment subtype typically contains 23 amino acid residues, but may contain as little as 15 17 amino acid residues, and lacks charged residues or contain one charged residue. Thus, this first **G** segment subtype typically contains 17-23 amino acid residues, but it is contemplated that it may contain as few as 12 or as many as 30 amino acid residues. Representative **G** segments of this first subtype are amino acid residues 20-42, 84-106, 148-170, 212-234, 307-329, 20 371-393, 435-457, 530-552, 595-617, 689-711, 753-775, 817-839, 881-903, 946-968, 1043-1059 and 1074-1092 of SEQ ID NO: 10. In certain embodiments, the first two amino acid residues of each **G** segment of this first subtype are not -Gln-Gln-.

The second subtype of the **G** segment is represented by the amino acid one letter consensus sequence GQGGQGQG(G/R)Y GQG(A/S)G(S/G) (SEQ ID NO: 12). This second, generally mid-sized, **G** segment subtype typically contains 17 amino acid residues and lacks charged residues or contain one charged residue. This second **G** segment subtype typically contains 14-20 amino acid residues, but it is contemplated that it may contain 30 as few as 12 or as many as 30 amino acid residues. Representative **G** segments of this second subtype are amino acid residues 249-265, 471-488, 631-647 and 982-998 of SEQ ID NO: 10; and amino acid residues 187-203 of SEQ ID NO: 3.

The third subtype of the **G** segment is represented by the amino acid one letter consensus sequence G(R/Q)GQG(G/R)YGQQ (A/S/V)GGN (SEQ ID NO: 13). This third **G** segment subtype typically contains 14 amino acid residues, and is generally the shortest of the **G** segment subtypes. This third 5 **G** segment subtype typically contains 12-17 amino acid residues, but it is contemplated that it may contain as many as 23 amino acid residues. Representative **G** segments of this third subtype are amino acid residues 57-70, 121-134, 184-197, 280-293, 343-356, 407-420, 503-516, 567-580, 662-675, 726-739, 790-803, 854-867, 918-931, 1014-1027 of SEQ ID NO: 10; and 10 amino acid residues 219-232 of SEQ ID NO: 3.

Thus, in preferred embodiments, each individual **G** segment has at least 80%, preferably 90%, more preferably 95%, identity to an amino acid sequence selected from SEQ ID NO: 11, SEQ ID NO: 12 and SEQ ID NO: 13.

In a preferred embodiment of the alternating sequence of **A** and **G** 15 segments of the **REP** moiety, every second **G** segment is of the first subtype, while the remaining **G** segments are of the third subtype, e.g. ...**A**₁**G**_{short}**A**₂**G**_{long}**A**₃**G**_{short}**A**₄**G**_{long}**A**₅**G**_{short}... In another preferred embodiment of the **REP** moiety, one **G** segment of the second subtype interrupts the **G** segment regularity via an insertion, e.g. 20 ...**A**₁**G**_{short}**A**₂**G**_{long}**A**₃**G**_{mid}**A**₄**G**_{short}**A**₅**G**_{long}...

Each individual **L** segment represents an optional linker amino acid sequence, which may contain from 0 to 20 amino acid residues, such as from 0 to 10 amino acid residues. There are also linker amino acid sequences present in the repetitive part (SEQ ID NO: 10) of the deduced amino acid 25 sequence of the MaSp1 protein from *Euprosthenops australis*. In particular, the amino acid sequence of a linker segment may resemble any of the described **A** or **G** segments, but usually not sufficiently to meet their criteria as defined herein.

Representative **L** segments are amino acid residues 1-6 and 1093-30 1110 of SEQ ID NO: 10; and amino acid residues 138-142 of SEQ ID NO: 3, but the skilled person in the art will readily recognize that there are many suitable alternative amino acid sequences for these segments. In one embodiment of the **REP** moiety, one of the **L** segments contains 0 amino

acids, i.e. one of the **L** segments is void. In another embodiment of the **REP** moiety, both **L** segments contain 0 amino acids, i.e. both **L** segments are void. Thus, these embodiments of the **REP** moieties may be schematically represented as follows: $(AG)_nL$, $(AG)_nAL$, $(GA)_nL$, $(GA)_nGL$; $L(AG)_n$,
5 $L(AG)_nA$, $L(GA)_n$, $L(GA)_nG$; and $(AG)_n$, $(AG)_nA$, $(GA)_n$, $(GA)_nG$.

The fusion protein is not comprising any moiety derived from the repetitive fragment of a spider silk protein. i.e. it has a low (or no) degree of identity and/or similarity to repetitive spider silk protein fragments. The sequence of the fusion protein according to the invention preferably has less
10 than 30% identity, such as less than 20% identity, preferably less than 10% identity, to any of the repetitive spidroin amino acid sequences disclosed herein, and specifically to any of SEQ ID NO: 10-13.

The **B** moiety is a protein or polypeptide fragment comprising more
15 than 30 amino acid residues. The **B** moiety is preferably comprising more than 50 amino acid residues, such as more than 100 amino acid residues. The **B** moiety is preferably comprising less than 1000 amino acid residues, such as less than 400 amino acid residues, more preferably less than 300 amino acid residues. It is capable of selective interaction with the organic
20 target, and it is the **B** moiety in the fusion protein which provides the capacity of selective interaction with the organic target.

The **B** moiety is a non-spidroin moiety. This implies that it is not derived from a spider silk protein, i.e. it has a low (or no) degree of identity and/or similarity to spider silk proteins. The sequence of the **B** moiety
25 according to the invention preferably has less than 30% identity, such as less than 20% identity, preferably less than 10% identity, to any of the spidroin amino acid sequences disclosed herein, and specifically to any of SEQ ID NO: 6-10.

It is regarded as within the capabilities of those of ordinary skill in the
30 art to select the **B** moiety. Nevertheless, examples of affinity ligands that may prove useful as **B** moieties, as well as examples of formats and conditions for detection and/or quantification, are given below for the sake of illustration.

The biomolecular diversity needed for selection of affinity ligands may be generated by combinatorial engineering of one of a plurality of possible scaffold molecules, and specific and/or selective affinity ligands are then selected using a suitable selection platform. Non-limiting examples of such structures, useful for generating affinity ligands against the organic target, are staphylococcal protein A and domains thereof and derivatives of these domains, such as the Z domain (Nord K *et al.* (1997) *Nat. Biotechnol.* 15:772-777); lipocalins (Beste G *et al.* (1999) *Proc. Natl. Acad. Sci. U.S.A.* 96:1898-1903); ankyrin repeat domains (Binz HK *et al.* (2003) *J. Mol. Biol.* 332:489-503); cellulose binding domains (CBD) (Smith GP *et al.* (1998) *J. Mol. Biol.* 277:317-332; Lehtio J *et al.* (2000) *Proteins* 41:316-322); γ crystallines (Fiedler U and Rudolph R, WO01/04144); green fluorescent protein (GFP) (Peelle B *et al.* (2001) *Chem. Biol.* 8:521-534); human cytotoxic T lymphocyte-associated antigen 4 (CTLA-4) (Hufton SE *et al.* (2000) *FEBS Lett.* 475:225-231; Irving RA *et al.* (2001) *J. Immunol. Meth.* 248:31-45); protease inhibitors, such as Knottin proteins (Wentzel A *et al.* (2001) *J. Bacteriol.* 183:7273-7284; Baggio R *et al.* (2002) *J. Mol. Recognit.* 15:126-134) and Kunitz domains (Roberts BL *et al.* (1992) *Gene* 121:9-15; Dennis MS and Lazarus RA (1994) *J. Biol. Chem.* 269:22137-22144); PDZ domains (Schneider S *et al.* (1999) *Nat. Biotechnol.* 17:170-175); peptide aptamers, such as thioredoxin (Lu Z *et al.* (1995) *Biotechnology* 13:366-372; Klevenz B *et al.* (2002) *Cell. Mol. Life Sci.* 59:1993-1998); staphylococcal nuclease (Norman TC *et al.* (1999) *Science* 285:591-595); tendamistats (McConell SJ and Hoess RH (1995) *J. Mol. Biol.* 250:460-479; Li R *et al.* (2003) *Protein Eng.* 16:65-72); trinectins based on the fibronectin type III domain (Koide A *et al.* (1998) *J. Mol. Biol.* 284:1141-1151; Xu L *et al.* (2002) *Chem. Biol.* 9:933-942); zinc fingers (Bianchi E *et al.* (1995) *J. Mol. Biol.* 247:154-160; Klug A (1999) *J. Mol. Biol.* 293:215-218; Segal DJ *et al.* (2003) *Biochemistry* 42:2137-2148); adnectin; anticalin; DARPin; affilin and avimer.

30 The above-mentioned examples include scaffold proteins presenting a single randomized loop used for the generation of novel binding specificities, protein scaffolds with a rigid secondary structure where side chains protruding from the protein surface are randomized for the generation of novel binding

specificities, and scaffolds exhibiting a non-contiguous hyper-variable loop region used for the generation of novel binding specificities.

Oligonucleotides may also be used as affinity ligands. Single stranded nucleic acids, called aptamers or decoys, fold into well-defined three-dimensional structures and bind to their target with high affinity and specificity. (Ellington AD and Szostak JW (1990) *Nature* 346:818-822; Brody EN and Gold L (2000) *J. Biotechnol.* 74:5-13; Mayer G and Jenne A (2004) *BioDrugs* 18:351-359). The oligonucleotide ligands can be either RNA or DNA and can bind to a wide range of target molecule classes.

For selection of the desired affinity ligand from a pool of variants of any of the scaffold structures mentioned above, a number of selection platforms are available for the isolation of a specific novel ligand against a target protein of choice. Selection platforms include, but are not limited to, phage display (Smith GP (1985) *Science* 228:1315-1317), ribosome display (Hanes J and Plückthun A (1997) *Proc. Natl. Acad. Sci. U.S.A.* 94:4937-4942), yeast two-hybrid system (Fields S and Song O (1989) *Nature* 340:245-246), yeast display (Gai SA and Wittrup KD (2007) *Curr Opin Struct Biol* 17:467-473), mRNA display (Roberts RW and Szostak JW (1997) *Proc. Natl. Acad. Sci. U.S.A.* 94:12297-12302), bacterial display (Daugherty PS (2007) *Curr Opin Struct Biol* 17:474-480, Kronqvist N *et al.* (2008) *Protein Eng Des Sel* 1-9, Harvey BR *et al.* (2004) *PNAS* 101(25):913-9198), microbead display (Nord O *et al.* (2003) *J Biotechnol* 106:1-13, WO01/05808), SELEX (System Evolution of Ligands by Exponential Enrichment) (Tuerk C and Gold L (1990) *Science* 249:505-510) and protein fragment complementation assays (PCA) (Remy I and Michnick SW (1999) *Proc. Natl. Acad. Sci. U.S.A.* 96:5394-5399). A preferred group of **B** moieties with affinity for immunoglobulins, albumin or other organic targets are bacterial receptin domains or derivatives thereof.

A group of preferred **B** moieties are capable of selective interaction with immunoglobulins and molecules comprising immunoglobulin or derivatives thereof, e.g. the fragment crystallisable (Fc) region of IgG. A preferred group of immunoglobulin subclasses are the subclasses that are recognized by the Z domain derived from staphylococcal protein A, i.e. IgG1, IgG2, IgG4, IgA and IgM from human, all Ig subclases from rabbit and cow,

IgG1 and IgG2 from guinea pig, and IgG1, IgG2a, IgG2b, IgG3 and IgM from mouse (see Hober, S. *et al.*, *J. Chromatogr B*. 848:40-47 (2007)), more preferably the immunoglobulin subclasses IgG1, IgG2, IgG4, IgA and IgM from human. The Z domain is an engineered version of the immunoglobulin G (IgG) binding domain B of staphylococcal protein A, and is a 58 amino acid long triple-helix motif that binds the Fc region of IgG. Another preferred group of immunoglobulin subclasses are the subclasses that are recognized by the C2 domain streptococcal protein G; i.e. all human subclasses of IgG, including IgG3, and IgG from several animals, including mouse, rabbit and sheep.

One group of preferred **B** moieties are selected from the group consisting of the Z domain derived from staphylococcal protein A, staphylococcal protein A and domains thereof, preferably the E, D, A, B and C domains, streptococcal protein G and domains thereof, preferably the C1, C2 and C3 domains; and protein fragments having at least 70% identity, such as at least 80% identity, or at least 90% identity, to any of these amino acid sequences. Preferably, the **B** moiety is selected from the group consisting of the Z domain derived from staphylococcal protein A, the B domain of staphylococcal protein A, and the C2 domain of streptococcal protein G; and protein fragments having at least 70% identity, such as at least 80% identity, or at least 90% identity, to any of these amino acid sequences. Preferably, the **B** moiety is selected from the group consisting of the Z domain derived from staphylococcal protein A and protein fragments having at least 70% identity, such as at least 80% identity, or at least 90% identity, to this amino acid sequence. It is preferred that the **B** moiety is selected from the group consisting of the Z domain derived from staphylococcal protein A and the C2 domain of streptococcal protein G. A preferred group of **B** moieties with affinity for immunoglobulins are bacterial receptin domains or derivatives thereof.

Another group of preferred **B** moieties are capable of selective interaction with albumin and molecules comprising albumin or derivatives thereof. A preferred group of **B** moieties with affinity for albumin are bacterial receptin domains or derivatives thereof. Preferred **B** moieties are selected

from streptococcal protein G, the albumin-binding domain of streptococcal protein G, GA modules from *Finegoldia magna*; and protein fragments having at least 70% identity, such as at least 80% identity, or at least 90% identity, to any of these amino acid sequences. Preferably, the **B** moiety is selected from

5 the albumin-binding domain of streptococcal protein G and protein fragments having at least 70% identity, such as at least 80% identity, or at least 90% identity, thereto. It is preferred that the **B** moiety is the albumin-binding domain of streptococcal protein G.

A further group of preferred **B** moieties are capable of selective

10 interaction with biotin and molecules comprising biotin or derivatives or analogues thereof. Preferred **B** moieties are selected from the group consisting of streptavidin, monomeric streptavidin (M4); and protein fragments having at least 70% identity, such as at least 80% identity, or at least 90% identity to any of these amino acid sequences. It is preferred that the **B**

15 moiety is monomeric streptavidin (M4).

Another group of preferred **B** moieties are enzymes, capable of selective interaction with substrates for an enzymatically catalyzed reaction. Preferred enzyme **B** moieties include xylanase and lysozyme.

A further group of preferred **B** moieties are growth factors, capable of

20 stimulating cell growth. Preferred growth factor **B** moieties include epidermal growth factor (EGF), in particular human EGF, fibroblast growth factor 2 (FGF2), nerve growth factor 1 (NGF1) and stromal cell-derived factor 1 (SDF1). Specific fusion proteins and protein structures according to the invention are provided in the Examples. These preferred fusion proteins form

25 the group consisting of SEQ ID NOS 61-66, 68, 70, 72, 74, 76, 80, 82, 84 and 86. Further preferred fusion proteins are having at least 80%, preferably at least 90%, more preferably at least 95%, identity to any of these sequences.

The present invention further provides isolated nucleic acids encoding a fusion protein according to the invention. In particular, specific nucleic acids

30 are provided in the Examples and the appended sequence listing. Further preferred nucleic acids encode fusion proteins having at least 80%, preferably at least 90%, more preferably at least 95%, identity to any of SEQ ID NOS 61-66, 68, 70, 72, 74, 76, 80, 82, 84 and 86.

The nucleic acids according to the invention are useful for producing the fusion proteins according to the invention. The present invention provides a method of producing a fusion protein. The first step involves expressing in a suitable host a fusion protein according to the invention. Suitable hosts are

5 well known to a person skilled in the art and include e.g. bacteria and eukaryotic cells, such as yeast, insect cell lines and mammalian cell lines. Typically, this step involves expression of a nucleic acid molecule which encodes the fusion protein in *E. coli*.

The second method step involves obtaining a mixture containing the

10 fusion protein. The mixture may for instance be obtained by lysing or mechanically disrupting the host cells. The mixture may also be obtained by collecting the cell culture medium, if the fusion protein is secreted by the host cell. The thus obtained protein can be isolated using standard procedures. If desired, this mixture can be subjected to centrifugation, and the appropriate

15 fraction (precipitate or supernatant) be collected. The mixture containing the fusion protein can also be subjected to gel filtration, chromatography, e.g. anion exchange chromatography, dialysis, phase separation or filtration to cause separation. Optionally, lipopolysaccharides and other pyrogens are actively removed at this stage. If desired, linker peptides may be removed by

20 cleavage in this step.

Proteins structures, or formats, according to the invention are assembled spontaneously from the fusion proteins according to the invention under suitable conditions, and the assembly into polymers is promoted by the presence of shearing forces and/or an interface between two different phases

25 e.g. between a solid and a liquid phase, between air and a liquid phase or at a hydrophobic/hydrophilic interface, e.g. a mineral oil-water interface. The presence of the resulting interface stimulates polymerization at the interface or in the region surrounding the interface, which region extends into the liquid medium, such that said polymerizing initiates at said interface or in said

30 interface region. Various protein structures can be produced by adapting the conditions during the assembly. For instance, if the assembly is allowed to occur in a container that is gently wagged from side to side, a fiber is formed at the air-water interface. If the mixture is allowed to stand still, a film is

formed at the air-water interface. If the mixture is evaporated, a film is formed at the bottom of the container. If oil is added on top of the aqueous mixture, a film is formed at the oil-water interface, either if allowed to stand still or if wagged. If the mixture is foamed, e.g. by bubbling of air or whipping, the foam

5 is stable and solidifies if allowed to dry.

The present invention thus provides a method for providing a protein structure displaying a binding activity towards an organic target. In the first method step, there is provided a recombinant fusion protein according to the invention. The fusion protein may e.g. be provided by expressing it in a

10 suitable host from a nucleic acid according to the invention. In the second method step, the fusion protein is subjected to conditions to achieve formation of a polymer comprising the recombinant fusion protein. Notably, although the spontaneously assembled protein structures can be solubilized in hexafluoroisopropanol, the solubilized fusion proteins are then not able to

15 spontaneously reassemble into e.g. fibers.

The protein structure is useful as part of an affinity medium for immobilization of an organic target, wherein the **B** moiety is capable of selective interaction with the organic target. A sample, e.g. a biological sample, may be applied to a fusion protein or a protein structure according to

20 the invention which is capable of binding to an organic target present in the biological sample, and the fusion protein or protein structure is then useful for separation of the organic target from the sample. A biological sample, such as blood, serum or plasma which has been removed from a subject may be subjected to detection, separation and/or quantification of the organic target.

25 The present invention thus provides a method for separation of an organic target from a sample. A sample, e.g. a biological sample such as blood, serum or plasma, containing the organic target is provided. The biological sample may be an earlier obtained sample. If using an earlier obtained sample in a method, no steps of the method are practiced on the

30 human or animal body.

An affinity medium according to the invention is provided, comprising a fusion protein or a protein structure according to the invention. In certain embodiments, the affinity medium is consisting of the fusion protein or protein

structure according to the invention. The affinity medium is capable of selective interaction with the organic target by means of the **B** moiety in the fusion protein according to the invention. The affinity medium is contacted with the sample under suitable conditions to achieve binding between the

5 affinity medium and the organic target. Non-bound sample is removed under suitable conditions to maintain selective binding between the affinity medium and the organic target. This method results in an organic target immobilized to the affinity medium, and specifically to the fusion protein, according to the invention.

10 In a preferred method according to the invention, the fusion protein in the affinity medium is present as a protein structure according to the invention when contacting the affinity medium with the sample to achieve binding between the affinity medium and the organic target.

15 A particularly useful protein structure in this respect is a film or a fiber wherein the **B** moiety is the Z domain derived from staphylococcal protein A or a protein fragment having at least 70% identity, such as at least 80% identity, or at least 90% identity, thereto. The film is advantageous in that it adheres to solid supports, e.g. the plastics in microtiter plates. This property of the film facilitates washing and regeneration procedures and is very useful

20 for separation purposes.

It has surprisingly been observed that the alkali stability of the Z domain may even be enhanced when being part of a fusion protein according to the invention in a protein structure according to the invention. This property may be very useful for washing and regeneration purposes, e.g. allowing for

25 high concentrations of NaOH, such as 0.1 M, 0.5 M, 1 M or even above 1 M, e.g. 2 M, and/or for high concentrations of urea, e.g. 6-8 M. The chemical stability may also be useful to allow for repeated cycles of use of the Z domain for affinity purification. This alkali stability may be further increased by utilizing a stabilized mutant of the Z domain. Furthermore, it has

30 advantageously been shown that the fusion proteins according to the invention, including the Z domain, are heat stable. This allows for sterilization by heat with maintained solid protein format/structure as well as binding ability.

A known problem with traditional affinity matrices with Z domains is leakage of the Z domain from the affinity matrix. Due to the stable incorporation of the Z domain by a peptide bond into the fusion protein of the invention, it is contemplated that the undesirable leakage of the Z domain

5 from the protein structures according to the invention is low or absent.

Another advantage of the fusion proteins according to the invention is that the resulting protein structure has a high density of Z domains (or other **B** moieties). It is contemplated that this high density provides a high binding capacity. Altogether, these properties of the fusions proteins are very 10 attractive for various **B** moieties, and in particular for affinity purification using protein Z with good production economy. These properties are also useful in other formats than in traditional gel bead affinity columns, e.g. in filter-like formats.

15 The immobilized organic target is capable of selective interaction with a second organic target. The method is then further comprising the step of contacting said affinity medium and the immobilized organic target with a second organic target, which is capable of selective interaction with the first organic target, under suitable conditions to achieve binding between the first and second organic targets.

20 The immobilized organic target is detectable and/or quantifiable. The detection and/or quantification of the organic target may be accomplished in any way known to the skilled person for detection and/or quantification of binding reagents in assays based on various biological or non-biological interactions. The organic targets may be labeled themselves with various

25 markers or may in turn be detected by secondary, labeled affinity ligands to allow detection, visualization and/or quantification. This can be accomplished using any one or more of a multitude of labels, which can be conjugated to the organic target or to any secondary affinity ligand, using any one or more of a multitude of techniques known to the skilled person, and not as such

30 involving any undue experimentation. Non-limiting examples of labels that can be conjugated to organic targets and/or secondary affinity ligands include fluorescent dyes or metals (e.g., fluorescein, rhodamine, phycoerythrin, fluorescamine), chromophoric dyes (e.g., rhodopsin), chemiluminescent

compounds (e.g., luminal, imidazole) and bioluminescent proteins (e.g., luciferin, luciferase), haptens (e.g., biotin). A variety of other useful fluorophores and chromophores are described in Stryer L (1968) *Science* 162:526-533 and Brand L and Gohlke JR (1972) *Annu. Rev. Biochem.* 41:843-868. Organic targets and/or secondary affinity ligands can also be labeled with enzymes (e.g., horseradish peroxidase, alkaline phosphatase, beta-lactamase), radioisotopes (e.g., ³H, ¹⁴C, ³²P, ³⁵S or ¹²⁵I) and particles (e.g., gold). In the context of the present disclosure, “particles” refer to particles, such as metal particles, suitable for labeling of molecules. Further, the affinity ligands may also be labeled with fluorescent semiconductor nanocrystals (quantum dots). Quantum dots have superior quantum yield and are more photostable compared to organic fluorophores and are therefore more easily detected (Chan *et al.* (2002) *Curr Opin Biotech.* 13: 40-46). The different types of labels can be conjugated to an organic target or a secondary affinity ligand using various chemistries, e.g., the amine reaction or the thiol reaction. However, other reactive groups than amines and thiols can be used, e.g., aldehydes, carboxylic acids and glutamine.

If the detection and/or quantification involves exposure to a second organic target or secondary affinity ligand, the affinity medium is washed once again with buffers to remove unbound secondary affinity ligands. As an example, the secondary affinity ligand may be an antibody or a fragment or a derivative thereof. Thereafter, organic targets may be detected and/or quantified with conventional methods. The binding properties for a secondary affinity ligand may vary, but those skilled in the art should be able to determine operative and optimal assay conditions for each determination by routine experimentation.

The detection, localization and/or quantification of a labeled molecule may involve visualizing techniques, such as light microscopy or immunofluorescence microscopy. Other methods may involve the detection via flow cytometry or luminometry. The method of visualization of labels may include, but is not restricted to, fluorometric, luminometric and/or enzymatic techniques. Fluorescence is detected and/or quantified by exposing fluorescent labels to light of a specific wavelength and thereafter detecting

and/or quantifying the emitted light in a specific wavelength region. The presence of a luminescently tagged molecule may be detected and/or quantified by luminescence developed during a chemical reaction. Detection of an enzymatic reaction is due to a color shift in the sample arising from 5 chemical reaction. Those of skill in the art are aware that a variety of different protocols can be modified in order for proper detection and/or quantification.

One available method for detection and/or quantification of the organic target is by linking it or the secondary affinity ligand to an enzyme that can then later be detected and/or quantified in an enzyme immunoassay (such as 10 an EIA or ELISA). Such techniques are well established, and their realization does not present any undue difficulties to the skilled person. In such methods, the biological sample is brought into contact with a protein structure according to the invention which binds to the organic target, which is then detected and/or quantified with an enzymatically labeled secondary affinity ligand. 15 Following this, an appropriate substrate is brought to react in appropriate buffers with the enzymatic label to produce a chemical moiety, which for example is detected and/or quantified using a spectrophotometer, fluorometer, luminometer or by visual means.

The organic target or the secondary affinity ligands can be labeled with 20 radioisotopes to enable detection and/or quantification. Non-limiting examples of appropriate radiolabels in the present disclosure are ^3H , ^{14}C , ^{32}P , ^{35}S or ^{125}I . The specific activity of the labeled affinity ligand is dependent upon the half-life of the radiolabel, isotopic purity, and how the label has been incorporated into the affinity ligand. Affinity ligands are preferably labeled 25 using well-known techniques (Wensel TG and Meares CF (1983) in: *Radioimmunoimaging and Radioimmunotherapy* (Burchiel SW and Rhodes BA eds.) Elsevier, New York, pp 185-196). A thus radiolabeled affinity ligand can be used to visualize the organic target by detection of radioactivity. Radionuclear scanning can be performed with e.g. a gamma camera, 30 magnetic resonance spectroscopy, emission tomography, gamma/beta counters, scintillation counters and radiographies.

Thus, the sample may be applied to the protein structure for detection, separation and/or quantification of the organic target. This procedure enables

not only detection of the organic target, but may in addition show the distribution and relative level of expression thereof. Optionally, the organic target may be released from the affinity medium and collected. Thus, the use may comprise affinity purification on an affinity medium onto which the

5 organic target has been immobilized. The protein structure may for example be arranged in a column or in well plates (such as 96 well plates), or on magnetic beads, agarose beads or sepharose beads. Further, the use may comprise use of the protein structures on a soluble matrix, for example using a dextran matrix, or use in a surface plasmon resonance instrument, such as

10 a Biacore™ instrument, wherein the analysis may for example comprise monitoring the affinity for the immobilized organic target or a number of potential affinity ligands.

The protein structures according to the invention can be washed and regenerated with various cleaning agents, including acid, base and chaotropic agents. Particularly useful cleaning agents include NaOH, such as 0.1, 0.5 or 1 M NaOH, and urea, such as 6-8 M urea, Since the protein structures according to the invention are surprisingly resistant to chemical treatment and/or sterilizing heat treatment, the methods according to the invention involving use of the protein structures may comprise a final step of

15 regenerating the protein structure. The methods preferably comprise a final step of regenerating the affinity medium by chemical treatment and/or sterilizing heat treatment. It is preferred that the chemical treatment comprises treatment with NaOH, such as 0.1, 0.5 or 1 M NaOH, and/or urea, such as 6-8 M urea,

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25 Fusion proteins according to the invention can be also be allowed to bind to an organic target in solution, i.e. prior to allowing the fusion protein to polymerize and form a protein structure, such as a film, a foam or a fibre. Both the spidroin-derived moieties (e.g. **CT**) as such and the corresponding fusion proteins incorporating a **B** moiety polymerise into solid structures even

30 in the presence of contaminating proteins, without appreciable incorporation of contaminants into the material, and the functional (**B**) moieties retain their expected binding properties. It is therefore contemplated that the binding properties of the **B** moiety can be used to capture compounds or cells from

the surrounding solution and incorporate the captured compounds or cells into or on a protein structure according to the invention.

Thus, in another preferred method according to the invention, the fusion protein in the affinity medium is present in solution when contacting the 5 affinity medium with the sample to achieve binding between the affinity medium and the organic target. The complex of fusion protein bound to the organic target is then allowed to form a fusion protein structure according to the invention.

This method may be particularly useful when the purpose is to “fish 10 out” specific molecules or cells from a solution, e.g. to obtain target molecules from the media in large scale eukaryotic cell production systems when the target proteins are secreted. Since the binding of target molecules and formation of solid structures by the spidroin-derived moieties can take place at physiological conditions and since the spidroin-derived moieties are 15 cyocompatible, the method can be applied repeatedly to an ongoing production process.

The protein structure according to the invention is also useful in separation, immobilization and/or cultivation of cells. A particularly useful protein structure in this respect is a film, a fiber or a foam. The film is 20 advantageous in that it adheres to solid structures, e.g. the plastics in microtiter plates. This property of the film facilitates washing and regeneration procedures and is very useful for selective detection and separation purposes.

The present invention thus provides a cell scaffold material for 25 cultivation of cells having an organic target that is present on the cell surface. The cell scaffold material is comprising a protein structure according to the invention. In certain embodiments, the cell scaffold material is consisting of the protein structure according to the invention.

It has been found by the present inventors that a cell scaffold material 30 comprising a polymer comprising, and optionally consisting of, the fusion protein according to the invention provides a beneficial environment for the cultivation of cells, and preferably eukaryotic cells, in a variety of different settings. Furthermore, this environment enables the establishment of cultures

of cells that are otherwise very difficult, very costly or even impossible to culture in a laboratory, and for the establishment of cell-containing materials useful for tissue engineering and/or transplantation.

The invention also provides a combination of cells, preferably 5 eukaryotic cells, and the cell scaffold material according to the invention. Such a combination according to the invention may be presented in a variety of different formats, and tailored to suit the needs of a specific situation. It is contemplated, for example, that the inventive combination may be useful as a cell-containing implant for the replacement of cells in damaged or diseased 10 tissue.

The cell scaffold material can be utilized to capture cells either directly or indirectly. In direct capture, the **B** moiety is capable of selective interaction with an organic target that is present on the cell surface. Alternatively, the **B** moiety is capable of selective interaction with and is bound to an intermediate 15 organic target, and that intermediate organic target is capable of selective interaction with an organic target that is present on the cell surface. Thus, in indirect capture, the cell scaffold material is further comprising an intermediate organic target, and the **B** moiety is capable of selective interaction with and is bound to said intermediate organic target. The 20 intermediate organic target, in turn, is capable of selective interaction with the organic target that is present on the cell surface.

In one embodiment of the cell scaffold materials as disclosed herein, the fusion protein is further comprises an oligopeptide cell-binding motif. In connection with the cultivation of certain cells in certain situations, the 25 presence of oligopeptide cell-binding motifs has been observed to improve or maintain cell viability, and the inclusion of such a motif into the cell scaffold material as a part of the spider silk protein is thought to provide additional benefits. The cell-binding motif is an oligopeptide coupled to the rest of the fusion protein via at least one peptide bond. For example, it may be coupled 30 to the N-terminal or the C-terminal of the rest of the fusion protein, or at any position within the amino acid sequence of the rest of the spider silk protein. With regard to the selection of oligopeptidic cell-binding motifs, the skilled person is aware of several alternatives. The coupling of an oligopeptide cell-

binding motif to the rest of the spider silk protein is readily accomplished by the skilled person using standard genetic engineering or chemical coupling techniques. Thus, in some embodiments, the cell-binding motif is introduced via genetic engineering, i.e. forming part of a genetic fusion between a nucleic acid encoding a fusion protein and the cell-binding motif. As an additional beneficial characteristic of such embodiments, the cell-binding motif will be present in a 1:1 ratio to the monomers of fusion protein in the polymer making up the cell scaffold material.

The polymer in the cell scaffold material used in the methods or combination described herein may adopt a variety of physical forms, and use of a specific physical form may offer additional advantages in different specific situations. For example, in an embodiment of the methods or combination, said cell scaffold material is in a physical form selected from the group consisting of film, foam, capsules, fiber and fiber-mesh.

15 The present invention accordingly provides a method for immobilization of cells. A sample e.g. a biological sample such as blood, comprising cells of interest is provided. The biological sample may be an earlier obtained sample. If using an earlier obtained sample in a method, no steps of the method are practiced on the human or animal body.

20 The sample is applied to a cell scaffold material according to the invention under suitable conditions to allow selective interaction between the cell scaffold material and an organic target that is present on the surface of the cells of interest. The cells are allowed to immobilize to said cell scaffold material by binding between the organic target on the cell surface and said cell scaffold material. Non-bound sample is removed under suitable conditions to maintain selective binding between the cell scaffold material and the organic target. This method results in cells exhibiting the organic target being immobilized to the cell scaffold material, and specifically to the protein structure, according to the invention.

25 As set out above, the cell scaffold material can be utilized to capture cells either directly or indirectly. In direct capture, the **B** moiety is capable of selective interaction with an organic target that is present on the cell surface. Alternatively, the **B** moiety is capable of selective interaction with and is

bound to an intermediate organic target, and that intermediate organic target is capable of selective interaction with an organic target that is present on the cell surface. Thus, in indirect capture, the cell scaffold material is further comprising an intermediate organic target, and the **B** moiety is capable of 5 selective interaction with and is bound to said intermediate organic target. The intermediate organic target, in turn, is capable of selective interaction with the organic target that is present on the cell surface.

Regardless of capture method, the captured cells may be released from the fusion protein by cleavage of the fusion protein to release the moiety 10 involved in cell capture from the cell scaffold material. As mentioned hereinabove, the fusion protein may include a cleavage site in its amino acid sequence, which allows for cleavage and removal of the relevant moiety, typically the **B** moiety or a cell-binding motif. Various cleavage sites are known to the person skilled in the art, e.g. cleavage sites for chemical agents, 15 such as CNBr after Met residues and hydroxylamine between Asn-Gly residues, cleavage sites for proteases, such as thrombin or protease 3C, and self-splicing sequences, such as intein self-splicing sequences.

The present invention also provides a method for cultivation of cells. Cells of interest are immobilized to the cell scaffold material using the method 20 disclosed hereinabove. The combination of the cell scaffold material and the immobilized cells are maintained under conditions suitable for cell culture.

In the context of the present invention, the terms "cultivation" of cells, "cell culture" etc are to be interpreted broadly, such that they encompass for 25 example situations in which cells divide and/or proliferate, situations in which cells are maintained in a differentiated state with retention of at least one functional characteristic exhibited by the cell type when present in its natural environment, and situations in which stem cells are maintained in an undifferentiated state.

30 According to another aspect, the present invention provides a novel recombinant protein comprising a **CT** moiety and at least one **NT** moiety, with the proviso that the protein is not comprising any moiety derived from the repetitive fragment of a spider silk protein. In a preferred embodiment, the protein is comprising 1-2 **NT** moieties.

In one preferred embodiment, the recombinant protein is consisting of a **CT** moiety and at least one **NT** moiety, such as 1-2 **NT** moieties. The protein may schematically be written as **NT-CT**, **CT-NT**, **NTNT-CT**, **CT-NTNT** or **NT-CT-NT**, and preferably **NT-CT** or **NTNT-CT**.

5 A preferred recombinant protein according to the invention is selected from the group consisting of SEQ ID NOS: 59-60 ; and proteins having at least 80%, preferably at least 90%, more preferably at least 95% identity to any of these sequences. The present invention further provides an isolated nucleic acid encoding a recombinant protein according to the invention.

10 This protein is void of the repetitive fragment of a spider silk protein, and it is therefore surprising that it is still capable of forming solid protein structures, e.g. fibers and films. An advantage with the present recombinant protein is that it can be produced with higher yield than CT alone or the corresponding proteins containing REP but lacking NT, see e.g. Examples 1-15 2.

15 The present invention provides a protein structure which is a polymer comprising as a repeating structural unit a recombinant protein according to this aspect. The protein structure preferably has a size of at least 0.1 µm in at least two dimensions. The protein structure is preferably in a physical form selected from the group consisting of fiber, film, foam, net, mesh, sphere and capsule.

20 Since this recombinant protein is not comprising any moiety derived from the repetitive fragment of a spider silk protein, it has a low (or no) degree of identity and/or similarity to repetitive spider silk protein fragments. The sequence of the protein according to the invention preferably has less than 30% identity, such as less than 20% identity, preferably less than 10% identity, to any of the repetitive spidroin amino acid sequences disclosed herein, and specifically to any of SEQ ID NO: 10-13.

25 The present invention will in the following be further illustrated by the following non-limiting examples.

30

ExamplesExample 1 - Cloning, expression and fiber formation of NT-CT

5 To investigate if CT covalently linked to NT can form fibers, a NT-CT fusion protein (a **NT** moiety and a **CT** moiety) was produced and purified.

Cloning

10 Genes encoding the His₆NT-CT fusion protein (SEQ ID NO: 59) were constructed. The vectors were transformed into chemocompetent *Escherichia coli* (*E. coli*) BL21 (DE3) cells that were allowed to grow onto agar plates supplemented with kanamycin (70 µg/ml). Colonies were thereafter picked and PCR screened for correct insert and subsequently also sequenced to confirm the DNA sequence.

15 *Production*

E. coli BL21 (DE3) cells possessing the pT7His₆NTCT vector were grown in Luria-Bertani medium (6 litre in total) supplemented with kanamycin (70 µg/ml) to an OD₆₀₀ value of 1-1.5 in 30°C, followed by induction of expression with 300 µM IPTG (isopropyl β-D-1-thiogalactopyranoside) and 20 further incubation in 20°C for approximately 2 h. Next, the cells were harvested by a 20 min centrifugation at 4 700 rpm, and the resulting cell pellets were dissolved in 20 mM Tris (pH 8.0).

Purification

25 Cell pellets dissolved in 20 mM Tris (pH 8.0) were supplemented with lysozyme and DNase I in order to lyse the bacterial cells, whereupon the cell lysates were recovered after 15 000 rpm of centrifugation for 30 min. Next, the recovered cell lysates were divided and loaded onto a total of four Chelating Sepharose Fast Flow Zn²⁺ columns, keeping the protein bound to 30 the column matrix via the His₆ tag. After washing, bound proteins were eluted with 20 mM Tris/300 mM imidazole (pH 8.0). The yield of NTCT was typically higher than that of CT or Rep4CT. Next, the pooled eluate liquid was dialysed against 5 litres of 20 mM Tris (pH 8.0) over night, concentrated to 1 mg/ml

and finally allowed to form fibers or films. Fibers were faster formed if pH was decreased to below pH 6.4.

5 The fact that macroscopic fibers of His₆NTCT could be obtained
demonstrates that CT retains its fiber forming properties when fused to NT.

Example 2 - Cloning, expression and fiber formation of NTNT-CT

To investigate if CT can form fibers although covalently linked to NTNT, a NTNT-CT fusion protein (a **NTNT** moiety and a **CT** moiety) was
10 produced and purified.

Cloning

Genes encoding the His₆NTNT-CT fusion protein (SEQ ID NO: 60) were constructed. The vectors were transformed into chemocompetent *E. coli*
15 BL21 (DE3) cells that were allowed to grow onto agar plates supplemented with kanamycin (70 µg/ml). Colonies were thereafter picked and PCR screened for correct insert and subsequently also sequenced to confirm the DNA sequence.

20 *Production*

E. coli BL21 (DE3) cells possessing the pT7His₆NTNTCT vector were grown in Luria-Bertani medium (6 litre in total) supplemented with kanamycin (70 µg/ml) to an OD₆₀₀ value of 1-1.5 in 30°C, followed by induction of expression with 300 µM IPTG and further incubation in 20°C for
25 approximately 2 h. Next, the cells were harvested by a 20 min centrifugation at 4 700 rpm, and the resulting cell pellets were dissolved in 20 mM Tris (pH 8.0).

Purification

30 Cell pellets dissolved in 20 mM Tris (pH 8.0) were supplemented with lysozyme and DNase I in order to lyse the bacterial cells, whereupon the cell lysates were recovered after 15 000 rpm of centrifugation for 30 min. Next, the recovered cell lysates were divided and loaded onto a total of four

Chelating Sepharose Fast Flow Zn²⁺ columns, keeping the protein bound to the column matrix via the His₆ tag. After washing, bound proteins were eluted with 20 mM Tris/300 mM imidazole (pH 8.0). The yield of NTNTCT was typically higher than that of CT or Rep4CT. Next, the pooled eluate liquid was 5 dialysed against 5 litres of 20 mM Tris (pH 8.0) over night, concentrated to 1 mg/ml and finally allowed to form fibers or films. Fibers were faster formed if pH was decreased to below pH 6.4.

10 The fact that macroscopic fibers of His₆NTNTCT could be obtained demonstrates that CT retains its fiber forming properties when fused to NTNT.

Example 3 - Cloning, expression and fiber formation of an IgG-binding CT fusion protein

15 To prove the fusion protein concept, a CT protein (a **CT** moiety) is produced in fusion with the Z protein domain (a **B** moiety). The Z domain is an engineered version of the immunoglobulin G (IgG) binding domain B of staphylococcal protein A, and is a 58 amino acid long triple-helix motif that binds the fragment crystallisable (F_c) region of IgG. The aim is to investigate whether it is possible to produce structures, such as fibers, films and 20 membranes, from a fusion protein consisting of the Z domain fused to CT and still retain the IgG-binding ability of domain Z, as well as the structure forming properties of CT. In order to do so a fusion protein consisting of the Z domain N-terminally or C-terminally to CT is cloned.

25 *Cloning*

Genes encoding the His₆ZCT and His₆CTZ fusion proteins (SEQ ID NOS: 61-62) are constructed and transformed into chemocompetent *E. coli* BL21 (DE3) cells that are allowed to grow onto agar plates supplemented with kanamycin (70 µg/ml). Colonies are thereafter picked and PCR screened for 30 correct insert and subsequently also sequenced to confirm the correct DNA sequence.

Production

E. coli BL21 (DE3) cells possessing the pT7His₆ZCT or pT7His₆CTZ vector are grown in Luria-Bertani medium (6 litre in total) supplemented with kanamycin (70 µg/ml) to an OD₆₀₀ value of 1-1.5 in 30°C, followed by

5 induction of His₆ZCT or His₆CTZ expression with 300 µM IPTG and further incubation in 20°C for approximately 2 h. Next, the cells are harvested by a 20 min centrifugation at 4 700 rpm, and the resulting cell pellets are dissolved in 20 mM Tris (pH 8.0).

10 *Purification*

Cell pellets dissolved in 20 mM Tris (pH 8.0) are supplemented with lysozyme and DNase I in order to lyse the bacterial cells, whereupon the cell lysates are recovered after 15 000 rpm of centrifugation for 30 min. Next, the recovered cell lysates are divided and loaded onto a total of four Chelating

15 Sepharose Fast Flow Zn²⁺ columns, keeping the His₆ZCT protein bound to the column matrix via the His₆ tag. After washing, bound proteins are eluted with 20 mM Tris/300 mM imidazole (pH 8.0). The pooled eluate fractions are dialysed against 5 litres of 20 mM Tris (pH 8.0) over night, concentrated to 1 mg/ml and finally allowed to form fibers.

20

Analysis

To explore the capacity of the **B** moiety in a fusion protein structure of selective interaction with an organic target, the ability of domain Z in the fusion protein to bind IgG is studied. Fibers and films of this fusion protein are

25 used for binding of purified IgG and IgG from serum, followed by elution and subsequent analysis on SDS-PAGE, where IgG under non-reducing conditions appears as a ~146 kDa band.

Example 4 - Cloning, expression and fiber formation of an IgG-binding NTCT

30 fusion protein

To prove the fusion protein concept, a NT-CT protein (a **NT** and a **CT** moiety) is produced in fusion with the Z protein domain (a **B** moiety). The aim is to investigate whether it is possible to produce structures, such as fibers,

films and membranes, from a fusion protein consisting of the Z domain fused to NTCT and still retain the IgG-binding ability of domain Z, as well as the structure forming properties of CT. In order to do so a fusion protein consisting of the Z domain N-terminally and C-terminally to NTCT is cloned.

5

Cloning

Genes encoding the His₆ZNTCT and His₆NTCTZ fusion proteins (SEQ ID NOS: 63-64) are constructed and transformed into chemocompetent *E. coli* BL21 (DE3) cells that are allowed to grow onto agar plates supplemented with 10 kanamycin (70 µg/ml). Colonies are thereafter picked and PCR screened for correct insert and subsequently also sequenced to confirm the correct DNA sequence.

Production

15 *E. coli* BL21 (DE3) cells possessing the pT7His₆ZNTCT and pT7His₆NTCTZ vector are grown in Luria-Bertani medium (6 litre in total) supplemented with kanamycin (70 µg/ml) to an OD₆₀₀ value of 1-1.5 in 30°C, followed by induction of expression with 300 µM IPTG and further incubation in 20°C for approximately 2 h. Next, the cells are harvested by a 20 min 20 centrifugation at 4 700 rpm, and the resulting cell pellets are dissolved in 20 mM Tris (pH 8.0).

Purification

Cell pellets dissolved in 20 mM Tris (pH 8.0) are supplemented with 25 lysozyme and DNase I in order to lyse the bacterial cells, whereupon the cell lysates are recovered after 15 000 rpm of centrifugation for 30 min. Next, the recovered cell lysates are divided and loaded onto a total of four Chelating Sepharose Fast Flow Zn²⁺ columns, keeping the His₆ZNTCT or His₆NTCTZ protein bound to the column matrix via the His₆ tag. After washing, bound 30 proteins are eluted with 20 mM Tris/300 mM imidazole (pH 8.0). The pooled eluate fractions are dialysed against 5 litres of 20 mM Tris (pH 8.0) over night, concentrated to 1 mg/ml and finally allowed to form fibers.

Analysis

To explore the capacity of the **B** moiety in a fusion protein structure of selective interaction with an organic target, the ability of domain Z in the fusion protein to bind IgG is studied. Fibers and films of this fusion protein are 5 used for binding of purified IgG and IgG from serum, followed by elution and subsequent analysis on SDS-PAGE, where IgG under non-reducing conditions appears as a ~146 kDa band.

Example 5 - Cloning, expression and fiber formation of an IgG-binding10 NTNTCT fusion protein

To prove the fusion protein concept, a NTNT-CT protein (a **NTNT** and a **CT** moiety) is produced in fusion with the Z protein domain (a **B** moiety). The aim is to investigate whether it is possible to produce structures, such as fibers, films and membranes, from a fusion protein consisting of the Z domain 15 fused to NTNTCT and still retain the IgG-binding ability of domain Z, as well as the structure forming properties of CT. In order to do so a fusion protein consisting of the Z domain N-terminally and C-terminally to NTNTCT is cloned.

20 Cloning

Genes encoding the His₆ZNTNTCT and His₆NTNTCTZ fusion proteins (SEQ ID NOS: 65-66) are constructed and transformed into chemocompetent *E. coli* BL21 (DE3) cells that are allowed to grow onto agar plates supplemented with kanamycin (70 µg/ml). Colonies are thereafter picked and 25 PCR screened for correct insert and subsequently also sequenced to confirm the correct DNA sequence.

Production

E. coli BL21 (DE3) cells possessing the pT7His₆ZNTNTCT and 30 pT7His₆NTNTCTZ vector are grown in Luria-Bertani medium (6 litre in total) supplemented with kanamycin (70 µg/ml) to an OD₆₀₀ value of 1-1.5 in 30°C, followed by induction of expression with 300 µM IPTG and further incubation in 20°C for approximately 2 h. Next, the cells are harvested by a 20 min

centrifugation at 4 700 rpm, and the resulting cell pellets are dissolved in 20 mM Tris (pH 8.0).

Purification

5 Cell pellets dissolved in 20 mM Tris (pH 8.0) are supplemented with lysozyme and DNase I in order to lyse the bacterial cells, whereupon the cell lysates are recovered after 15 000 rpm of centrifugation for 30 min. Next, the recovered cell lysates are divided and loaded onto a total of four Chelating Sepharose Fast Flow Zn²⁺ columns, keeping the His₆ZNTNTCT or

10 His₆NTNTCTZ protein bound to the column matrix via the His₆ tag. After washing, bound proteins are eluted with 20 mM Tris/300 mM imidazole (pH 8.0). The pooled eluate fractions are dialysed against 5 litres of 20 mM Tris (pH 8.0) over night, concentrated to 1 mg/ml and finally allowed to form fibers.

15 *Analysis*

To explore the capacity of the **B** moiety in a fusion protein structure of selective interaction with an organic target, the ability of domain Z in the fusion protein to bind IgG is studied. Fibers and films of this fusion protein are used for binding of purified IgG and IgG from serum, followed by elution and

20 subsequent analysis on SDS-PAGE, where IgG under non-reducing conditions appears as a ~146 kDa band.

Example 6 - Cloning, expression and formation of solid structures of ABD-NTCT and ABD-CT fusion proteins

25 NTCT and CT were produced in fusion with the albumin binding domain (ABD) derived from streptococcal protein G. ABD is a 5 kDa triple-helix motif that binds to albumin. The aim was to investigate whether it is possible to produce structures, such as fibers and films, from the fusion proteins consisting of the ABD domain fused to NTCT (denoted His₆-ABD-

30 NTCT, SEQ ID NO: 68) and to CT (denoted His₆-ABD-CT, SEQ ID NO: 70) respectively, and still retain the albumin binding ability of ABD domain as well as the structure forming properties of NTCT and CT. In order to do so, two

fusion proteins consisting of the ABD domain fused N-terminally to NTCT and to CT were cloned.

Cloning

5 A gene (SEQ ID NO: 69) encoding the His₆-ABD-CT fusion protein (SEQ ID NO: 70) was constructed as follows. Primers were designed in order to generate PCR fragments of domain ABD from a vector containing such an ABD sequence. Also, the primers contained recognition sites for the restriction endonucleases *NdeI* and *EcoRI*. The resulting PCR products were
10 then treated with the restriction endonucleases *NdeI* and *EcoRI*, as was the target vector (denoted pAff8His₆TrxHis₆CT, harbouring a kanamycin resistance gene). Upon restriction cleavage of the target vector, the His₆TrxHis₆ part was cleaved off. Cleaved PCR fragments and target vector were joined together with the aid of a T4 DNA Ligase, whereupon the
15 resulting correctly ligated vector (pT7His₆-ABD-CT) was transformed into chemocompetent *E. coli* BL21 (DE3) cells that were allowed to grow onto agar plates supplemented with kanamycin (50 µg /ml). Colonies were thereafter picked and screened for correct insert and subsequently sequenced to confirm the DNA sequence of the inserted ABD into the target
20 vector.

Cloning of a gene (SEQ ID NO: 67) encoding the His₆-ABD-NTCT fusion protein (SEQ ID NO: 68) was constructed in the same way as described for His₆-ABD-CT, but the target vector here was denoted by pT7His₆scFv1-NTCT, where the pT7His₆scFv1 part was cleaved off upon
25 treatment with *NdeI* and *EcoRI*. The correctly ligated vector is denoted as pT7His₆ABD-NTCT.

Production

E. coli BL21 (DE3) cells possessing the pT7His₆-ABD-CT vector were
30 grown in Luria-Bertani medium (3 liters in total) supplemented with kanamycin (50 µg/ml) to an OD₆₀₀ value of 1-1.5 in 30°C, followed by induction of pT7His₆-ABD-CT expression with 300 µM IPTG and further incubation at 14°C for approximately 17 h. Next, the cells were harvested by a 20 min

centrifugation at 4 700 rpm, and the resulting cell pellet was dissolved in 20 mM Tris (pH 8.0).

Production of His₆ABD-NTCT was performed in the same way as described for His₆ABD-CT.

5

Purification

The cell pellet dissolved in 20 mM Tris (pH 8.0) was supplemented with lysozyme and DNase I in order to lyse the bacterial cells, followed by the addition of NaCl and imidazole to a final concentration of 200 mM and 10 mM, 10 respectively. After 30 min of centrifugation (15 000 rpm) the cell lysate was recovered. Next, the recovered cell lysate was loaded onto a Chelating Sepharose Fast Flow Zn²⁺ column, keeping the His₆-ABD-CT protein bound to the column matrix via the His₆ tag. After washing, bound proteins were eluted with 20 mM Tris/200 mM imidazole (pH 8.0)/300 mM NaCl. The eluate 15 contained 28.8 mg of His₆-ABD-CT protein according to an A₂₈₀ measurement. Next, the eluted protein was dialyzed against 3 liters of 20 mM Tris (pH 8.0) over night and thereafter concentrated to 1.48 mg/ml, yielding a final amount of 6.216 mg His₆-ABD-CT fusion protein.

The same purification procedure was carried out for His₆-ABD-NTCT. 20 The eluate concentration of His₆ABD-NTCT was 1.76 mg/ml, and a final amount of 35.2 mg of fusion protein was obtained.

Film and fiber formation

Films of His₆-ABD-CT were casted in 96-well plates (Tissue culture 25 plate, Suspension cells, 83.1835.500, Sarstedt) from 15 µl of 1 mg/ml soluble fusion protein per film. The films were then allowed to solidify over night (20°C, 35% relative humidity). The same procedure was followed for casting films of His₆-ABD-NTCT from 15 µl of 1 mg/ml protein solution.

Fibers were also made for both His₆ABD-CT and His₆ABD-NTCT from 30 1.76 and 1.06 mg/ml of soluble fusion protein, respectively. Fig. 3a shows a microscopic fiber picture of a His₆-ABD-NTCT fusion protein (SEQ ID NO: 68), while Fig 3b shows a microscopic fiber picture of His₆-ABD-CT fusion protein (SEQ ID NO: 70). The fact that macroscopic fibers of both His₆ABD-

CT and His₆ABD-NTCT could be obtained although CT or NTCT has been fused to another protein, i.e. the 46 amino acid long ABD domain, demonstrates that CT and NTCT retain their structural forming properties despite being fused to the ABD domain.

5

Analysis

To evaluate the ability of ABD-NTCT films to bind albumin, human blood plasma was used as albumin source. Four films of ABD-NTCT and ABD-CT were pre-wetted with 150 µl of 1×PBS followed by incubation of 100 10 µl of human blood plasma (1:5 dilution) for 30 min at room temperature. After washing three times with 200 µl 1×PBS, bound albumin was eluted in 50 µl by lowering the pH to approximately 2.7 with elution buffer (i.e. 0.5 M acetic acid, 1 M urea, 100 mM NaCl), after which the eluted fractions were analyzed by non-reducing SDS-PAGE. Films of NTCT and CT were used as control 15 material, and were treated in the same way.

Fig. 4 shows a non-reducing SDS-PAGE gel of eluted fractions after binding of albumin from human blood plasma to ABD-NTCT and ABD-CT films. The gel was loaded according to:

- (1-4) Quadruplicates of ABD-NTCT films, 14 µl loaded;
- 20 (5-8) Quadruplicates of ABD-CT films, 14 µl loaded;
- (9) Protein ladder;
- (10-13) Quadruplicates of NTCT (control) films;
- (14-16) Triplicates of CT (control) films;
- (17) Human blood plasma (1:50), 8 µl loaded.

25 All films of ABD-NTCT and ABD-CT have bound albumin from human blood plasma (Fig. 4, lanes 1-8). As only one single albumin band (~65 kDa) appears in the eluted fraction of the ABD-NTCT and ABD-CT films, they seem to not bind anything else unspecifically from the human blood plasma. Control films of NTCT and CT do not show any albumin in the eluted fractions (Fig. 4, 30 lanes 10-16). It is concluded that the ABD domain is functional in the macroscopic solid structures of fusion proteins with NTCT and CT.

Example 7 - Cloning, expression and formation of solid structures of M4-NTCT and M4-CT fusion proteins

Monomeric streptavidin (M4) domain was produced in fusion with NTCT and CT protein, respectively. The M4 domain is a mutated version of 5 the tetrameric streptavidin, and is a 159 amino acid long protein domain that binds non-covalently to biotin. Our aim was to investigate whether it is possible to produce structures, such as films, foams and fibers from the fusion proteins consisting of the M4 domain fused to NTCT (denoted His₆-M4-NTCT, SEQ ID NO: 72) and to CT (denoted His₆-M4-CT, SEQ ID NO: 74), 10 respectively, and still retain the biotin-binding ability of M4 domain as well as the structure forming properties of NTCT and CT. In order to do so, two fusion proteins consisting of the M4 domain fused N-terminally to NTCT and to CT were cloned.

15 *Cloning*

A gene (SEQ ID NO: 73) encoding the His₆-M4-CT fusion protein (SEQ ID NO: 74) was constructed as set out in Example 6, but the primers were designed in order to generate PCR fragments of domain M4 from a vector containing such a M4 sequence. The target vector was denoted 20 pAff8His₆TrxHis₆CT, where the His₆TrxHis₆ part was cleaved off upon treatment with *NdeI* and *EcoRI*. The correctly ligated vector is denoted pT7His₆M4-CT.

Cloning of a gene (SEQ ID NO: 71) encoding the His₆-M4-NTCT fusion protein (SEQ ID NO: 72) was constructed in the same way as described for 25 His₆-M4-CT, but the target vector here was denoted by pT7His₆scFv1-NTCT, where the pT7His₆scFv1 part was cleaved off upon treatment with *NdeI* and *EcoRI*. The correctly ligated vector is denoted as pT7His₆M4-NTCT.

Production

30 Production of His₆M4-CT and His₆M4-NTCT was performed in the same way as described in Example 6.

Purification

Purification of His₆M4-CT and His₆M4-NTCT was performed in the same way as described in Example 6.

5 The eluate contained 3.6 mg of His₆M4-CT protein. After protein concentration to 1.39 mg/ml, a final amount of 0.834 mg His₆M4-CT fusion protein was obtained.

The eluate content of His₆M4-NTCT protein was 3.2 mg. After protein concentration to 1.14 mg/ml, a final amount of 1.368 mg His₆M4-NTCT fusion protein was obtained.

10

Film, foam and fiber formation

Films of His₆M4-CT and His₆M4-NTCT were casted as as described in Example 6. Fiber was made for His₆-M4-NTCT from 1.14 mg/ml of soluble fusion protein (Fig. 5a). Foam was made for His₆-M4-CT from 30 μ l of 1.39 mg/ml of soluble fusion protein (Fig. 5b). The fact that films, fiber and foam of His₆-M4-NTCT and His₆-M4-CT could be obtained although NTCT or CT has been fused to another protein, i.e. the 159 amino acids long M4 domain, demonstrates that NTCT and CT retain their structural forming properties despite being fused to the M4 domain.

20

Analysis

Spotted films of His₆-M4-NTCT and His₆-M4-CT contain 0.34 nmoles and 0.50 nmoles of target protein molecules, respectively. In order to evaluate the biotin binding ability of the two silk fused M4 constructs, two films for each 25 construct were selected and analyzed by incubating with an equal amount of Atto-565-biotin as compared to the amount of target protein molecules present in the films. Then labeled biotin was removed and the films were washed three times with 100 μ l of 1 \times PBS. Finally, 100 μ l of 1 \times PBS was added to the films before fluorescence microscope analysis using an inverted 30 Nikon Eclipse Ti instrument (excitation at 563 nm, emission at 592 nm). Films of NTCT (0.55 nmoles) and CT (1.28 nmoles) were used as control material, and were treated in the same way.

M4-NTCT and M4-CT films contain monomeric streptavidin (M4), which has an inherent affinity of binding to biotin. As labeled biotin was used in the analysis, binding of biotin to films could be detected by fluorescence microscopy at the wavelengths indicated above. Fig. 6 shows fluorescence 5 microscopy pictures at 2× magnification for detection of Atto-565-biotin bound to M4-NTCT and M4-CT films. A: M4-NTCT; B: NTCT; C: M4-CT and D: CT. Fluorescence can be seen only from M4-NTCT and M4-CT films in Fig. 6 (A, C) but not from the control films (B, D). This confirms that the biotin binding ability of M4 is retained despite being fused to NTCT and CT.

10

Example 8 - Cloning, expression and formation of solid structures of scFv1-NTCT and scFv1-CT fusion proteins

NTCT and CT were produced in fusion with an engineered antibody fragment named single chain fragment variable (scFv1). scFv1 is a 27-kDa 15 monovalent, engineered antibody fragment that recognizes the antigens specific for an autoimmune disease, Systemic Lupus Erythematosus (SLE). Our aim was to investigate whether it is possible to produce structures, such as fibers, foams and films, from the fusion proteins consisting of the scFv1 protein domain fused to NTCT (denoted His₆-scFv1-NTCT, SEQ ID NO: 76) 20 and to CT (denoted His₆-scFv1-CT, SEQ ID NO: 78), respectively, and still retain the antigen detection ability of scFv1 domain as well as the structure forming properties of NTCT and CT. In order to do so, two fusion proteins consisting of the scFv1 domain fused N-terminally to NTCT and to CT were cloned.

25

Cloning

A gene (SEQ ID NO: 77) encoding the His₆-scFv1-CT fusion protein (SEQ ID NO: 78) was constructed as set out in Example 6, but the primers were designed in order to generate PCR fragments of domain scFv1 from a 30 vector containing such a scFv1 sequence. The target vector was denoted pAff8His₆TrxHis₆CT, where the His₆TrxHis₆ part was cleaved off upon treatment with *NdeI* and *EcoRI*. The correctly ligated vector is denoted pT7His₆scFv1-CT.

Cloning of a gene (SEQ ID NO: 75) encoding the His₆-scFv1-NTCT fusion protein (SEQ ID NO: 76) was constructed in the same way as described for His₆-scFv1-CT, but the primers used for the amplification of NTCT contained sites for the restriction endonucleases EcoRI and HindIII and 5 the target vector here was denoted by T7His₆scFv1-RepCT, where the RepCT part was cleaved off upon treatment with EcoRI and HindIII. The correctly ligated vector is denoted as pT7His₆scFv1-NTCT.

Production

10 Production of His₆-scFv1-CT and His₆-scFv1-NTCT was performed in the same way as described in Example 6, except for that production of pT7His₆-scFv1-NTCT was performed in a culture media total volume of 6 liters.

15 *Purification*

Purification of His₆-scFv1-CT and pHis₆-scFv1-NTCT was performed in the same way as described in Example 6.

20 The eluate contained 0.93 mg of His₆-scFv1-CT protein. After protein concentration to 0.87 mg/ml, a final amount of 0.348 mg His₆-scFv1-CT fusion protein was obtained.

The eluate content of His₆-scFv1-NTCT protein was 4.86 mg. After protein concentration to 2.14 mg/ml, a final amount of 2.57 mg His₆-scFv1-NTCT fusion protein was obtained.

25 *Film, foam and fiber formation*

Films of His₆-scFv1-CT were spotted onto microarray slides (plastic MaxiSorp, Nunc) from 1 μ l of 5 μ M soluble fusion protein per film. The films were then allowed to solidify over night in a climate controlled room. The same procedure was followed for casting films of His₆-scFv1-NTCT from 1 μ l 30 of 5 μ M protein solution.

Fiber was made for His₆-scFv1-NTCT from 0.49 mg/ml (data not shown) and foams were made for both His₆-scFv1-NTCT and His₆-scFv1-CT from 30 μ l of 0.22 and 0.38 mg/ml of soluble fusion protein, respectively (Fig-

7a and 7b). The fact that macroscopic fiber and foam for His₆-scFv1-NTCT and His₆-scFv1-CT respectively, could be obtained although NTCT or CT has been fused to another protein, i.e. the 263 amino acids long scFv1 domain, demonstrates that NTCT and CT still retains there structure forming
5 properties despite fused to the scFv1 domain.

Analysis

Pure antibody (scFv1, control) and silk fused antibody (scFv1-NTCT) were spotted in the microarray format manually by adding 1 μ L of 5 μ M
10 protein solution onto clear and black polymer MaxiSorp microarray slides (NUNC, 25x76 mm) resulting in 135 pmoles of pure antibody (scFv1) and 274 pmoles of silk fused antibody (scFv1-NTCT) in the spotted films, respectively. After spotting the proteins in film format, the films were dried overnight in a climate controlled room. The arrays were then blocked by applying 200 μ l of
15 sample buffer (1% (w/v) fat-free milk powder and 1% (v/v) Tween-20 in PBS) for 90 min and then washed three times by applying 200-300 μ l of wash buffer (0.05% (v/v) Tween-20 in PBS). All incubations were performed at room temperature on gentle agitation. Next, 100-200 μ l of biotinylated antigen sample (10 nM) diluted in sample buffer was applied and incubated for 1h.
20 The arrays were then washed three times by applying 200-300 μ l of wash buffer and to detect the bound antigens, 100-200 μ l of Alexa-647-labeled streptavidin (1 μ g/ml) diluted in sample buffer, was applied onto the arrays and incubated for 1 h. Finally, the arrays were washed three times with 200-300 μ l of wash buffer and dried under a stream of nitrogen gas. The arrays
25 were then scanned using a confocal microarray fluorescence scanner (ScanArray Express, Perkin-Elmer Life & Analytical Sciences). The ScanArray Express software V2.0 (Perkin-Elmer Life & Analytical Sciences) was used to quantify the intensity of each spot. The same analysis procedure was carried out for analyzing His₆-scFv1-CT fusion protein.
30 In order to detect the low abundant serum proteins which can be of potential biomarkers, scFv1 was fused to N-terminal of NTCT or CT giving rise to His₆-scFv1-NTCT and His₆-scFv1-CT, respectively. Pure antibody (control) and silk fused antibody fragments were spotted onto the microarray

slide and their antigen binding capacity was analyzed using biotinylated antigen sample. Alexa-647-labeled streptavidin was then used to detect the bound antigens. Fig. 8 shows an antigen binding analysis of pure (control) and silk fused antibody fragments. Intensity of the spots was measured at 5 5090 detection intensity. The analysis showed that the antigen recognition of silk fused antibody (His₆-scFv1-NTCT) fragment was increased by 25 times compared to the scFv1 control alone, and no sign of cross reactivity with other antigens was observed for His₆-scFv1-NTCT.

10 Example 9 - Cloning, expression and formation of solid structures of xylanase-NTCT and xylanase-CT fusion proteins

To prove the concept of fusing a protein with enzymatic activity to NTCT and CT, the enzyme xylanase A from *Bacillus subtilis* was produced in fusion with NTCT and CT, respectively. Xylanase A (endo-1,4-beta-xylanase 15 A) is 185 amino acids long (without signal peptide) and belongs to the glycosyl hydrolase 11 (cellulose G) family. The enzymatic function of xylanase A is to cleave beta-1,4-glycosidic linkages of xylan, the main constituent of hemicellulose in plant cell walls. Our aim was to investigate whether it is possible to produce structures, such as fibers, foams and films, 20 from a fusion protein consisting of xylanase A fused to NTCT (denoted Xyl-NTCT, SEQ ID NO: 80) and to CT (denoted Xyl-CT, SEQ ID NO: 82), respectively, and still retain the enzymatic ability of xylanase as well as the structure forming properties of CT. In order to do so two fusion proteins were cloned consisting of xylanase N-terminally to 1) NTCT and 2) CT.

25

Cloning

A gene (SEQ ID NO: 81) encoding the His₆Xyl-CT fusion protein (SEQ ID NO: 82) was constructed as set out in Example 6, but the primers were designed in order to generate PCR fragments of the xylanase domain from a vector 30 containing such a xylanase sequence. The target vector was denoted pAff8His₆TrxHis₆CT, where the His₆TrxHis₆ part was cleaved off upon treatment with *NdeI* and *EcoRI*. The correctly ligated vector is denoted pT7His₆Xyl-CT.

Cloning of a gene (SEQ ID NO: 79) encoding the His₆Xyl-NTCT fusion protein (SEQ ID NO: 80) was constructed in the same way as described for His₆Xyl-CT, but the target vector here was denoted by pT7His₆scFv1-NTCT, where the pT7His₆scFv1 part was cleaved off upon treatment with *NdeI* and 5 *EcoRI*. The correctly ligated vector is denoted as pT7His₆Xyl-NTCT.

Production

Production of His₆Xyl-CT and His₆Xyl-NTCT was performed in the same way as described in Example 6.

10

Purification

Purification of His₆Xyl-CT and His₆Xyl-NTCT was performed in the same way as described in Example 6.

15 The eluate contained 3.6 mg of His₆Xyl-CT protein. After protein concentration to 2.1 mg/ml, a final amount of 2 mg His₆Xyl-CT fusion protein was obtained.

The eluate content of His₆Xyl-NTCT protein was 4.3 mg. After protein concentration to 0.65 mg/ml, a final amount of 0.3 mg His₆Xyl-NTCT fusion protein was obtained.

20

Film, foam and fiber formation

Films of His₆Xyl-CT were casted in 96-well plates (Tissue culture plate, Suspension cells, 83.1835.500, Sarstedt). Each film was made from 15 µl of 1.0 mg/ml soluble His₆Xyl-CT at both pH 8 and pH 6. The films were then 25 allowed to solidify over night (20°C, 35% relative humidity). The same procedure was followed for casting of His₆Xyl-NTCT films, each film casted from 15 µl of 0.41 mg/ml soluble His₆Xyl-NTCT (both at pH 8 and pH 6).

30 Foam was made from soluble His₆Xyl-CT (1-2 mg/ml) by introducing air (by pipetting) into 40 µl of the protein solution, followed by overnight drying at room temperature. The appearance of the formed foam of His₆Xyl-CT (Fig. 9), demonstrates that CT retains its structural forming properties despite being fused to the Xyl domain.

Fig. 9 shows a macroscopic foam of the fusion protein His₆Xyl-CT (SEQ ID NO: 82), made from soluble fusion protein. The appearance of the foam demonstrates that the spider silk CT domain has retained its structural forming properties even though produced in fusion with the enzyme xylanase 5 (Xyl).

Analysis of enzymatic activity of xylanase fused to NTCT or CT

Xylanase is an enzyme that cleaves the beta-1,4-glycosidic linkage between two xylose residues. To test the enzymatic ability in films of xylanase 10 (Xyl) fused to NTCT or CT, each film of His₆Xyl-CT and His₆Xyl-NTCT is incubated with 90 µl of McIlvaine buffer (pH 6.0). After a 10 min preincubation at 50°C of all films, 10 µl of 40 mM PNX (p-nitrophenyl-xylopyranoside) substrate is added, followed by an additional incubation at 50°C for at least 10 min. Then, 100 µl of stop solution (0.5 M Na₂CO₃) is added to each film, 15 followed by absorbance measurements at 410 nm to identify the product from the enzymatic reaction. Films of NTCT and CT, casted according to the same procedure previously stated, are included as controls.

Example 10 - Cloning, expression and formation of solid structures of
20 xylanase-NTCT and xylanase-CT fusion proteins

To prove the concept of fusing a peptide with cell stimulating effect to NTCT and CT, the human epidermal growth factor (EGF) was produced in fusion with NTCT and CT, respectively. EGF is a 53 amino acid residues long growth factor with high affinity for the Epidermal Growth Factor Receptor 25 (EGFR) found on the cell surface of many cell types, e.g. keratinocytes. Upon binding to EGFR, protein-tyrosin kinase activity is stimulated, resulting in a variety of biochemical changes in the cell, triggering cell growth and proliferation. Our aim was to investigate whether it is possible to produce structures, such as fibers and films, from a fusion protein consisting of EGF 30 fused to NTCT (denoted EGF-NTCT, SEQ ID NO: 84) and to CT (denoted EGF-CT, SEQ ID NO: 86), respectively, and still retain the cell stimulating effect of EGF as well as the structure forming properties of CT. In order to do

so two fusion proteins were cloned consisting of EGF N-terminally to 1) NTCT and 2) CT.

Cloning

5 A gene (SEQ ID NO: 85) encoding the His₆EGF-CT (SEQ ID NO: 86) fusion protein was constructed as set out in Example 6, but the primers were designed in order to generate PCR fragments of EGF from a vector containing such an EGF sequence. The target vector was denoted pAff8His₆TrxHis₆CT, where the His₆TrxHis₆ part was cleaved off upon 10 treatment with *NdeI* and *EcoRI*. The correctly ligated vector is denoted pT7His₆EGF-CT.

Cloning of a gene (SEQ ID NO: 83) encoding the His₆EGF-NTCT fusion protein (SEQ ID NO: 84) was constructed in the same way as described for His₆EGF-CT, but the target vector here was denoted by 15 pT7His₆scFv1-NTCT, where the pT7His₆scFv1 part was cleaved off upon treatment with *NdeI* and *EcoRI*. The correctly ligated vector is denoted as pT7His₆EGF-NTCT.

Production

20 Production of His₆EGF-CT and His₆EGF-NTCT is performed in the same way as described in Example 6.

Purification

Purification of His₆EGF-CT and His₆EGF-NTCT is performed in the 25 same way as described in Example 6.

Film and fiber formation

Films and fibers are made from both His₆EGF-CT and His₆EGF-NTCT as described in Example 6.

30

Analysis of the cell stimulating ability of EGF fused to NTCT or CT

The cell stimulating ability towards keratinocytes of EGF when fused to CT (denoted His₆EGF-CT) or NTCT (denoted His₆EGF-NTCT) is investigated.

For this purpose, films of the fusion proteins His₆EGF-CT and His₆EGF-NTCT, respectively are used. Normal human epidermal keratinocytes (primary cells) are seeded onto matrices at a density of 3500 or 7000 cells/cm² in cell culture medium (KGM-GOLD, Lonza) with or without 5 recombinant human EGF. The medium is exchanged every second day. Viable and dead cells are stained with Live/Dead assay (Molecular probes) after 24, 48, 72 and 96 h. Micrographs are taken in an Inverted Fluorescent microscope (Nikon Eclipse Ti) at 10× magnification.

CLAIMS:

1. A recombinant fusion protein comprising the moieties **B** and **CT**, wherein:

B is a non-spidroin moiety which provides the capacity of selective interaction with an organic target, wherein the **B** moiety is a protein or polypeptide fragment comprising more than 30 amino acid residues; and wherein the **B** moiety has less than 30% identity to any of SEQ ID NOS: 6-10; and

CT is a moiety of from 70 to 120 amino acid residues and has at least 80% identity to any one of SEQ ID NO: 7 and SEQ ID NOS: 14-44; wherein the **CT** moiety provides the capacity of forming a polymer;

with the proviso that the fusion protein is not comprising any moiety derived from the repetitive fragment of a spider silk protein.

2. A recombinant fusion protein according to claim 1, wherein the organic target is selected from the group consisting of immunoglobulins and molecules comprising immunoglobulin or derivatives thereof.

3. A recombinant fusion protein according to claim 2, wherein the immunoglobulins are selected from the immunoglobulin subclasses IgG1, IgG2, IgG4, IgA and IGM from human.

4. A recombinant fusion protein according to claim 2 or 3, wherein the **B** moiety is selected from the group consisting of the Z domain derived from staphylococcal protein A and the E, D, A, B and C domains thereof, streptococcal protein G and the C1, C2 and C3 domains thereof; and protein fragments having at least 70% identity to any of these amino acid sequences.

5. A recombinant fusion protein according to claim 4, wherein the **B** moiety is selected from the group consisting of the Z domain derived from staphylococcal protein A, the B domain of staphylococcal protein A, and the C2 domain of streptococcal protein G; and protein fragments having at least 70% identity to any of these amino acid sequences.

6. A recombinant fusion protein according to claim 1, wherein the organic target is selected from the group consisting of albumin and molecules comprising albumin or derivatives thereof.

7. A recombinant fusion protein according to claim 6, wherein the **B** moiety is selected from streptococcal protein G, the albumin-binding domain of streptococcal protein G, GA

modules from *Finegoldia magna*; and protein fragments having at least 70% identity to any of these amino acid sequences.

8. A recombinant fusion protein according to claim 1, wherein the organic target is selected from the group consisting of biotin and molecules comprising biotin or derivatives or analogues thereof.

9. A recombinant fusion protein according to claim 8, wherein the **B** moiety is selected from the group consisting of streptavidin, monomeric streptavidin (M4); and protein fragments having at least 70% identity to any of these amino acid sequences.

10. A recombinant fusion protein according to any preceding claim, selected from the group of proteins defined by the formula **B_x-CT-B_z**,

wherein x and z are integers from 0 to 5;

and x+z \geq 1.

11. A recombinant fusion protein according to any previous claim, selected from the group consisting of SEQ ID NOS: 61-66; and proteins having at least 80% identity to any of these sequences.

12. An isolated nucleic acid encoding a fusion protein according to any one of claims 1 to 11.

13. A protein structure capable of selective interaction with an organic target, wherein said protein structure is a polymer comprising as a repeating structural unit a recombinant fusion protein according to any one of claims 1 to 11, wherein the **B** moiety provides the capacity of selective interaction with the organic target.

14. A protein structure according to claim 13, wherein said protein structure is in a physical form selected from the group consisting of fiber, film, foam, net, mesh, sphere and capsule.

15. Use of a protein structure according to any one of claims 13 or 14 in cultivation of cells.

16. A method of producing a fusion protein, comprising the following steps:

a) expressing in a suitable host a fusion protein according to any one of claims 1 to 11; and

b) obtaining a mixture containing the fusion protein, and optionally isolating the fusion protein.

17. A method for providing a protein structure according to any one of claims 12 to 14, displaying a binding activity towards an organic target, comprising the steps of:

(a) providing a recombinant fusion protein according to any one of claims 1 to 11;

(b) subjecting the fusion protein to conditions to achieve formation of a polymer comprising the recombinant fusion protein.

Spiber Technologies AB

Patent Attorneys for the Applicant/Nominated Person

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CThyb_Esp	SRLSSPEASS RVSSAVSNLV SSG-PTNSAA LSSTISNVVS QIGASNPGLS
CTnat_Eau	SRLSSPSAVS RVSSAVSSLV SNG-QVNMAA LPNIISNISS SVSASAPGAS
AF350266_At1	SRLSSPGAAS RVSSAVTSLV SSGGPTNSAA LSNTISNVVS QISSSNPGLS
AY666062_Cm1	SHLSSPEASS RVSSAVSNLV SSG-STNSAA LPNTISNVVS QISSSNPGLS
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U47855_ADF3	SRLSSPAASS RVSSAVSSLV SSG-PTKHAA LSNTISSVVS QVSASNPGLS
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CTnat_Eau	GCEVIVQALL EVITALVQIV SSSSVGYINP SAVNQITNVV ANAMAQVMG- -
AF350266_At1	GCDVLVQALL EIVSALVHIL GSANIGQVNS SGVGRSASIV GQSINQAFS- -
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Consensus

GCDVLVQALL EVVSALVHIL GSSSIGQVNY GSAGQATQIV GQSVAQALGE F

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Lg	MaSp1	QANTPWSSKANADAFINSFISSAQNTGSFSQDQMDMSLIGNTLMAMDNMG--GRITPSKLQALDMAFA
Lh	MaSp1	QANTPWSSKANADAFINSFISSAQNTGSFSQDQMDMSLIGNTLMAMDNMG--GRITPSKLQALDMAFA
NC	MaSp1	-QNTPWSSTELADEFINAFMNEAGRIGAFTADQDDMSTIGDTIKTAMDKMARSNKSSKGKLQALNMAFA
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Lg	MaSp2	---LRWSSKDNADRFINAFIQLQAAASNQGAFSSDQVDDMSVIGNTLMTAMDNMG--GRITPSKLQALDMAFA
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NC	MaSp2	QARSPWSDTATADAFIQLNFIQVSGSGAFTSDQDDMSTIGDTIMSDQDQLDDMSTVGDTIMSAMDKMARSNKSSQHKLQALNMAFA
Ab	CySp1	AVPSVFPSSSPNLASGFLQCLTFIGNSPAFPTQEQQDLDIAQVILNAVSSNTGATASAR--AQALSTALA
Nc1	CySp1	PVPSVFPSSSPSLASGFLGCLTTCGIGLSPAFPFQEQQDLDLAKVILSAVTSNTDTKSAR--AQALSTALA
Lh	TuSp1	ASVNIFNSPNAATSFNLNCRSNIESSPAFFPQEQADLDSIAEVILSDVSS-VNTASSAT--SLALSTALA
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N1m	flag	IVNSPESNPNNTAEAFARSFVSNIVSSGEFGAQGAEDFDDIIQSLIQAQ-SMGKGRHDTKAKAKAMQVALA
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N1m	MaSp2	SSMAEIAAVEQGGMSMMAVKTNAIVDGLNSAFYMTTGAANPQFVNEMRSLISMISSAANEV
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Ab	CySp1	SSL DLLI SE SGGSSY QTOQI ISAL TNILS DCF VTTT G SNN PAFT VASV K SLL GVL S QSS S NYE
Nc1	CySp1	SSLAE LLV TE SAE EDIDNQVVALSTIILS QCF VETT G S P N PAFT VASV K SLL GVL S QSS S NYE
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Fig 2

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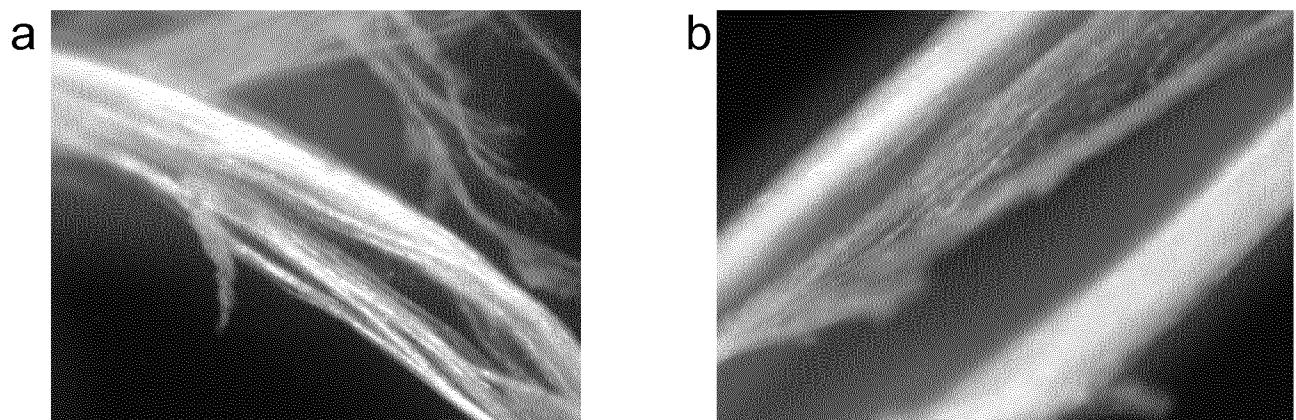


Fig 3

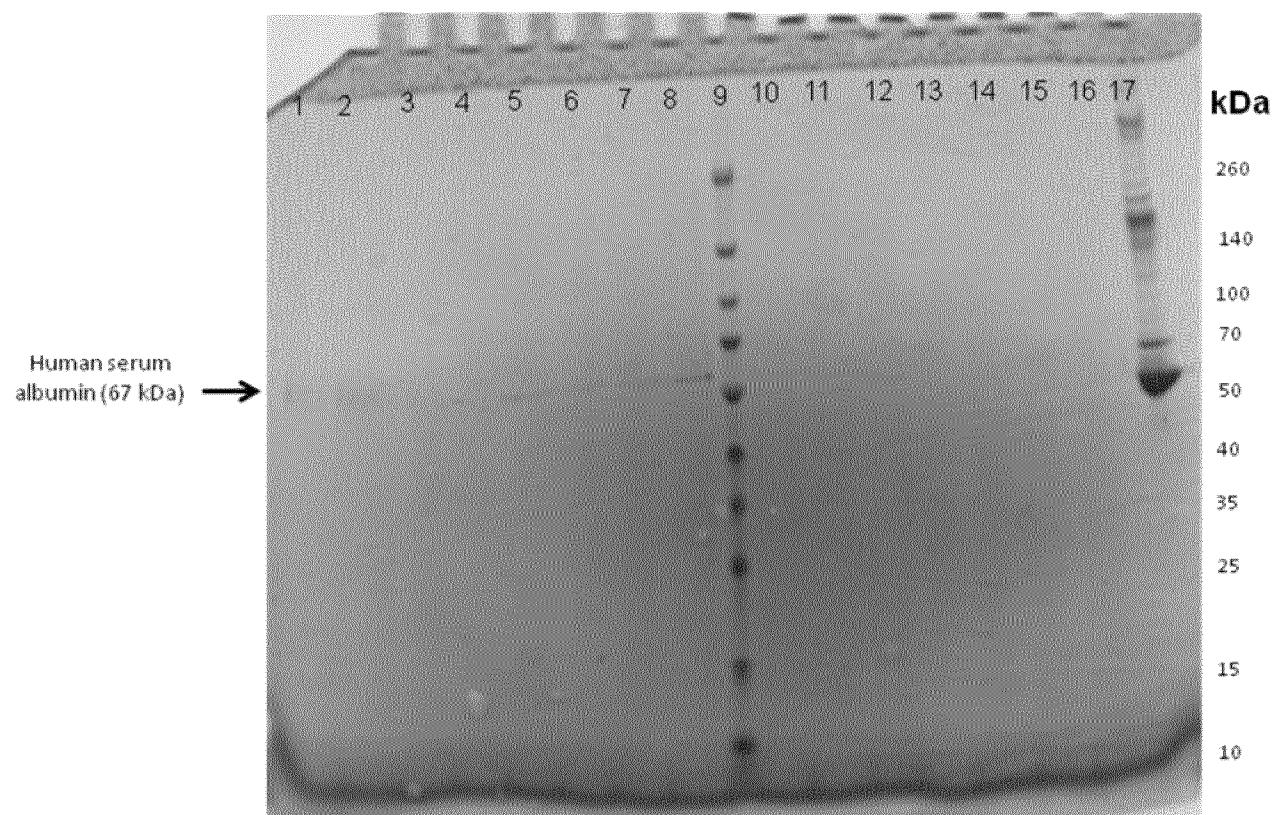


Fig 4

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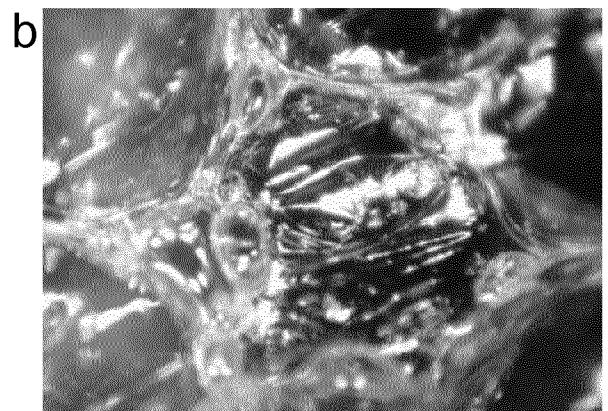
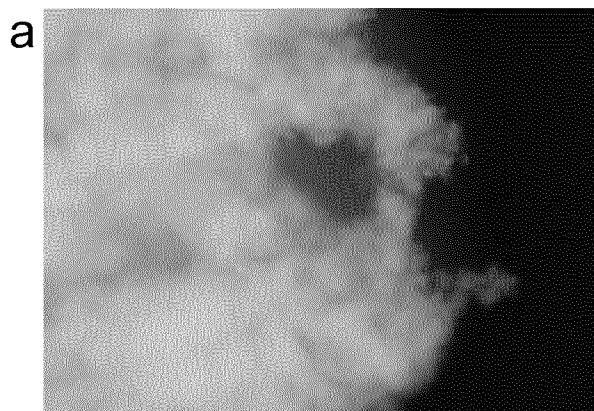


Fig 5

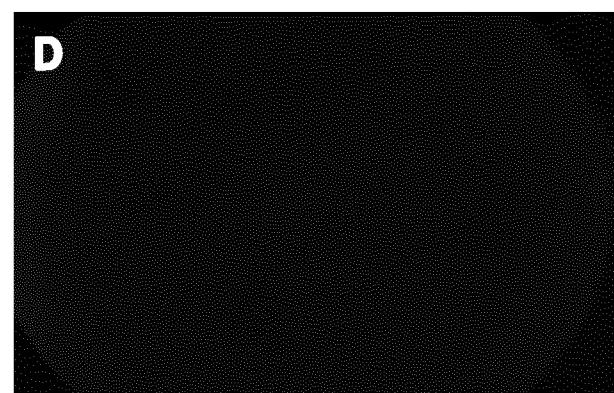
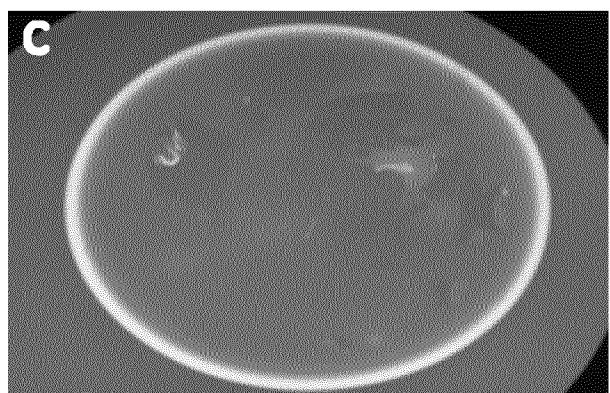
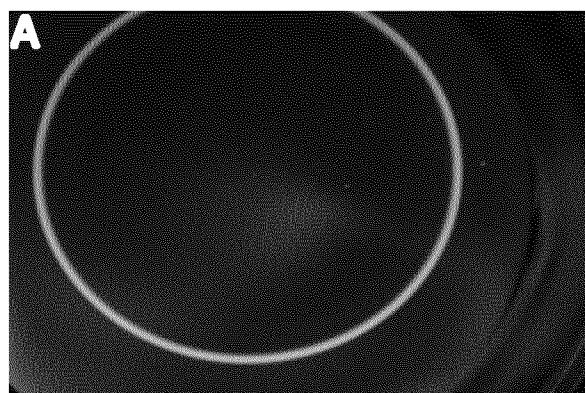


Fig 6

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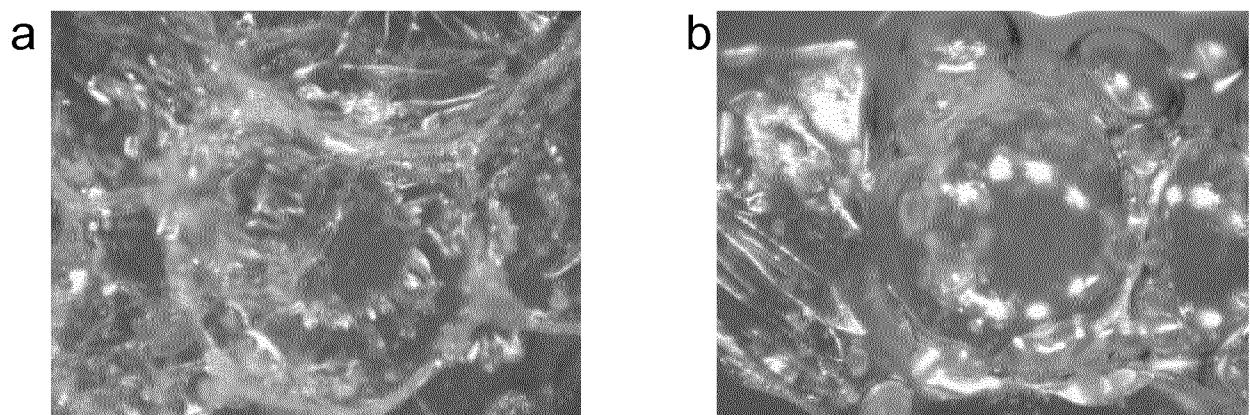


Fig 7

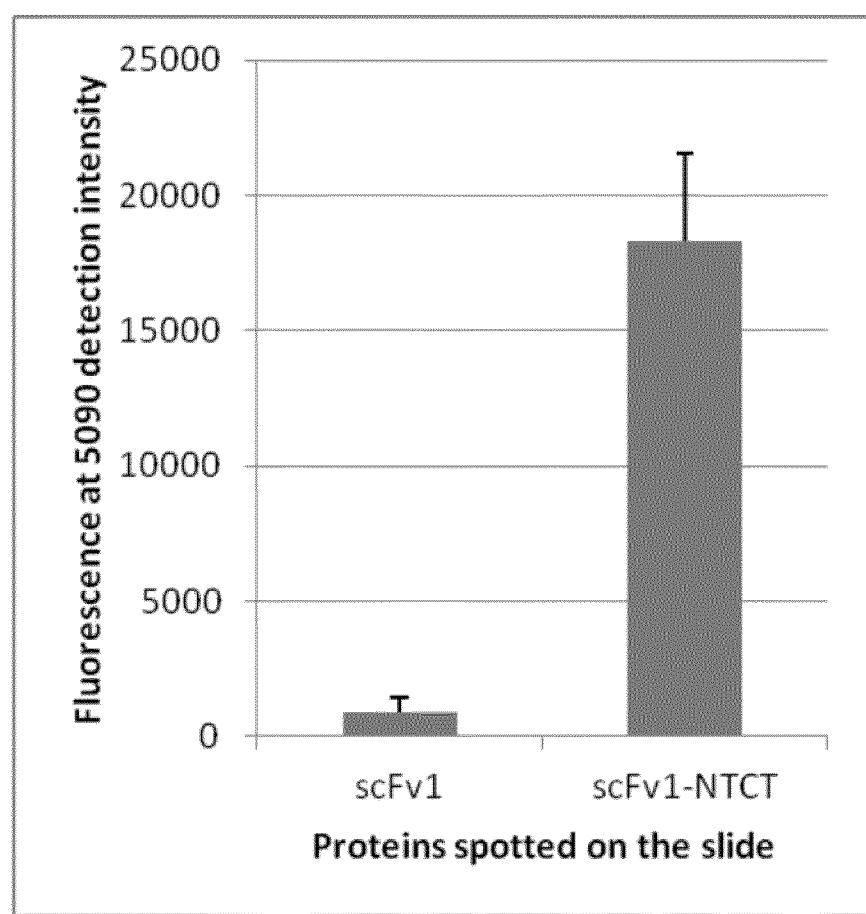


Fig 8

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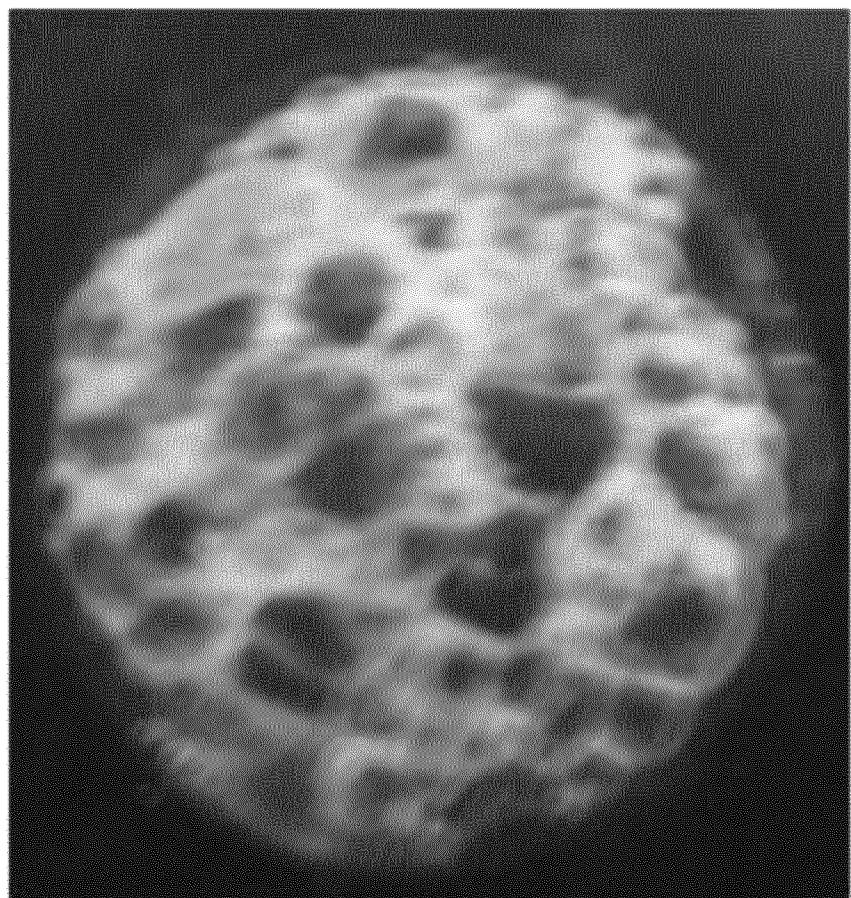


Fig 9

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Ala Ala Ala Ala Ala Ala Ala Gly Gly Gln Gly Gly Gln Gly Gly
35 40 45

Gly Gly Tyr Gly Gln Gly Ser Gly Gly Ser Ala Ala Ala Ala Ala Ala
50 55 60

Ala Ala Ala Ala Ala Ala Ala Gly Arg Gly Gln Gly Gly Tyr
65 70 75 80

Gly Gln Gly Ser Gly Gly Asn Ala Ala Ala Ala Ala Ala Ala Ala
85 90 95

Ala Ala Ala Ala Ala Gly Gln Gly Gly Gln Gly Gly Tyr Gly Arg Gln
100 105 110

Ser Gln Gly Ala Gly Ser Ala Ala Ala Ala Ala Ala Ala Ala Ala
115 120 125

Ala Ala Ala Ala Gly Ser Gly Gln Gly Gly Tyr Gly Gln Gly Gln Gly
130 135 140

Gly Tyr Gly Gln Ser
145

<210> 2
<211> 265
<212> PRT
<213> Euprosthenops australis

eol f-seql . txt

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<220>
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<222> (168)..(265)
<223> CT fragment

<400> 2

Gly Ser Gly Asn Ser Gly Ile Gln Gly Gln Gly Gly Tyr Gly Gly Leu
1 5 10 15

Gly Gln Gly Gly Tyr Gly Gln Gly Ala Gly Ser Ser Ala Ala Ala Ala Ala
20 25 30

Ala Ala Ala Ala Ala Ala Ala Gly Gly Gln Gly Gly Gln Gly Gly Gln
35 40 45

Gly Gly Tyr Gly Gln Gly Ser Gly Gly Ser Ala Ala Ala Ala Ala Ala
50 55 60

Ala Ala Ala Ala Ala Ala Ala Gly Arg Gly Gln Gly Gly Tyr
65 70 75 80

Gly Gln Gly Ser Gly Gly Asn Ala Ala Ala Ala Ala Ala Ala Ala
85 90 95

Ala Ala Ala Ala Ala Gly Gln Gly Gly Gln Gly Gly Tyr Gly Arg Gln
100 105 110

Ser Gln Gly Ala Gly Ser Ala Ala Ala Ala Ala Ala Ala Ala
115 120 125

Ala Ala Ala Ala Gly Ser Gly Gln Gly Gly Tyr Gly Gln Gly Gln Gly
130 135 140

Gly Tyr Gly Gln Ser Ser Ala Ser Ala Ser Ala Ala Ser Ala
145 150 155 160

Ser Thr Val Ala Asn Ser Val Ser Arg Leu Ser Ser Pro Ser Ala Val
165 170 175

Ser Arg Val Ser Ser Ala Val Ser Ser Leu Val Ser Asn Gly Gln Val
180 185 190

Asn Met Ala Ala Leu Pro Asn Ile Ile Ser Asn Ile Ser Ser Ser Val
195 200 205

Ser Ala Ser Ala Pro Gly Ala Ser Gly Cys Glu Val Ile Val Gln Ala
210 215 220

eol f-seql . txt

Leu Leu Glu Val Ile Thr Ala Leu Val Glu Ile Val Ser Ser Ser Ser
225 230 235 240

Val Gly Tyr Ile Asn Pro Ser Ala Val Asn Glu Ile Thr Asn Val Val
245 250 255

Ala Asn Ala Met Ala Glu Val Met Gly
260 265

<210> 3
<211> 296
<212> PRT
<213> Euprosthenops australis

<220>
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<222> (1)..(137)
<223> NT fragment

<220>
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<222> (138)..(296)
<223> REP fragment

<400> 3

Gly Ser Gly Asn Ser His Thr Thr Pro Trp Thr Asn Pro Gly Leu Ala
1 5 10 15

Glu Asn Phe Met Asn Ser Phe Met Glu Gly Leu Ser Ser Met Pro Gly
20 25 30

Phe Thr Ala Ser Glu Leu Asp Asp Met Ser Thr Ile Ala Glu Ser Met
35 40 45

Val Glu Ser Ile Glu Ser Leu Ala Ala Glu Gly Arg Thr Ser Pro Asn
50 55 60

Lys Leu Glu Ala Leu Asn Met Ala Phe Ala Ser Ser Met Ala Glu Ile
65 70 75 80

Ala Ala Ser Glu Glu Gly Gly Ser Leu Ser Thr Lys Thr Ser Ser
85 90 95

Ile Ala Ser Ala Met Ser Asn Ala Phe Leu Glu Thr Thr Gly Val Val
100 105 110

Asn Glu Pro Phe Ile Asn Glu Ile Thr Glu Leu Val Ser Met Phe Ala
115 120 125

Glu Ala Gly Met Asn Asp Val Ser Ala Ser Ala Ser Ala Glu Ala Ser
130 135 140

Ala Ala Ala Ser Ala Gly Ala Ala Ser Gly Glu Gly Gly Tyr Gly Gly
Page 3

eol f-seql . txt

145

150

155

160

Leu Gl y Gl n Gl y Gl y Tyr Gl y Gl n Gl y Al a Gl y Ser Ser Al a Al a Al a
165 170 175

Al a Gl y Gl y Gl n Gl y Gl y Gl n Gl y
180 185 190

Gl n Gl y Gl y Tyr Gl y Gl n Gl y Ser Gl y Gl y Ser Al a Al a Al a Al a Al a
195 200 205

Al a Gl y Arg Gl y Gl n Gl y Gl y
210 215 220

Tyr Gl y Gl n Gl y Ser Gl y Gl y Asn Al a
225 230 235 240

Al a Gl y Gl n Gl y Gl y Gl n Gl y Gl y Tyr Gl y
245 250 255

Arg Gl n Ser Gl n Gl y Al a Gl y Ser Al a
260 265 270

Al a Al a Al a Al a Al a Al a Gl y Ser Gl y Gl n Gl y Gl y Gl y Tyr Gl y Gl y Gl n
275 280 285

Gl y Gl n Gl y Gl y Tyr Gl y Gl n Ser
290 295

<210> 4
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<213> Euprosthenops australis

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<222> (138)..(340)
<223> REP fragment

<400> 4

Gl y Ser Gl y Asn Ser His Thr Thr Pro Trp Thr Asn Pro Gl y Leu Al a
1 5 10 15

Gl u Asn Phe Met Asn Ser Phe Met Gl n Gl y Leu Ser Ser Met Pro Gl y
20 25 30

Phe Thr Al a Ser Gl n Leu Asp Asp Met Ser Thr Ile Al a Gl n Ser Met
35 40 45

eol f-seql . txt

Val Glu Ser Ile Glu Ser Leu Ala Ala Glu Gly Arg Thr Ser Pro Asn
50 55 60

Lys Leu Glu Ala Leu Asn Met Ala Phe Ala Ser Ser Met Ala Glu Ile
65 70 75 80

Ala Ala Ser Glu Glu Gly Gly Ser Leu Ser Thr Lys Thr Ser Ser
85 90 95

Ile Ala Ser Ala Met Ser Asn Ala Phe Leu Glu Thr Thr Gly Val Val
100 105 110

Asn Glu Pro Phe Ile Asn Glu Ile Thr Glu Leu Val Ser Met Phe Ala
115 120 125

Glu Ala Gly Met Asn Asp Val Ser Ala Ser Ala Ser Ala Gly Ala Ser
130 135 140

Ala Ala Ala Ser Ala Gly Ala Pro Gly Tyr Ser Pro Ala Pro Ser Tyr
145 150 155 160

Ser Ser Gly Gly Tyr Ala Ser Ser Ala Ala Ser Ala Ala Ala Ala
165 170 175

Gly Glu Glu Gly Pro Gly Gly Tyr Gly Pro Ala Pro Asn Glu Gly Ala
180 185 190

Ser Ser Ala Ala Ala Ala Ala Gly Ser Gly Glu Glu Pro Ser Gly
195 200 205

Pro Tyr Gly Thr Ser Tyr Glu Ile Ser Thr Glu Tyr Thr Glu Thr Thr
210 215 220

Thr Ser Glu Gly Glu Gly Tyr Gly Ser Ser Ser Ala Gly Ala Ala Ala
225 230 235 240

Ala Gly Ala Ala Gly Ala Gly Glu Gly Gly Tyr Gly Glu Gly Glu
245 250 255

Gly Gly Tyr Gly Glu Gly Ala Gly Glu Ala Ala Ala Ala Ala Ala
260 265 270

Ala Ala Ala Ala Ala Ala Ala Gly Glu Gly Gly Glu Gly Glu Gly
275 280 285

Gly Tyr Gly Glu Gly Glu Glu Gly Glu Gly Glu Gly Glu Gly Glu
290 295 300

Gly Tyr Gly Glu Gly Glu Ala Gly Ser Ser Ala Ala Ala Ala Ala
305 310 315 320

eol f-seql . txt

Ala Ala Ala Ala Ala Ala Gly Arg Gly Gln Gly Tyr Gly Pro
325 330 335

Gly Ser Gly Gly
340

<210> 5
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<213> Euprosthenops australis

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<222> (1)..(136)
<223> NT fragment

<220>
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<222> (137)..(313)
<223> REP fragment

<220>
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<222> (314)..(411)
<223> CT fragment

<220>
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<222> (412)..(424)
<223> His tag

<400> 5

Met Lys Ala Ser His Thr Thr Pro Trp Thr Asn Pro Gly Leu Ala Glu
1 5 10 15

Asn Phe Met Asn Ser Phe Met Gln Gly Leu Ser Ser Met Pro Gly Phe
20 25 30

Thr Ala Ser Gln Leu Asp Asp Met Ser Thr Ile Ala Gln Ser Met Val
35 40 45

Gln Ser Ile Gln Ser Leu Ala Ala Gln Gly Arg Thr Ser Pro Asn Lys
50 55 60

Leu Gln Ala Leu Asn Met Ala Phe Ala Ser Ser Met Ala Glu Ile Ala
65 70 75 80

Ala Ser Glu Glu Gly Gly Ser Leu Ser Thr Lys Thr Ser Ser Ile
85 90 95

Ala Ser Ala Met Ser Asn Ala Phe Leu Gln Thr Thr Gly Val Val Asn
100 105 110

Gln Pro Phe Ile Asn Glu Ile Thr Gln Leu Val Ser Met Phe Ala Gln
115 120 125

eol f-seql . txt

Ala Glu Met Asn Asp Val Ser Ala Ser Ala Ser Ala Glu Ala Ser Ala
130 135 140

Ala Ala Ser Ala Glu Ala Ala Ser Glu Gln Glu Glu Tyr Glu Glu Leu
145 150 155 160

Gly Gln Gly Gly Tyr Gly Gln Gly Ala Gly Ser Ser Ala Ala Ala Ala
165 170 175

Ala Ala Ala Ala Ala Ala Ala Gly Gly Gln Gly Gly Gln Gly Gln
180 185 190

Gly Gly Tyr Gly Gln Gly Ser Gly Gly Ser Ala Ala Ala Ala Ala Ala
195 200 205

Ala Ala Ala Ala Ala Ala Ala Gly Arg Gly Gln Gly Gly Tyr
210 215 220

Gly Gln Gly Ser Gly Gly Asn Ala Ala Ala Ala Ala Ala Ala Ala Ala
225 230 235 240

Ala Ala Ala Ala Ala Ala Gly Gln Gly Gly Gln Gly Gly Tyr Gly Arg
245 250 255

Gln Ser Gln Gly Ala Gly Ser Ala Ala Ala Ala Ala Ala Ala Ala
260 265 270

Ala Ala Ala Ala Ala Gly Ser Gly Gln Gly Gly Tyr Gly Gly Gln Gly
275 280 285

Gln Gly Gly Tyr Gly Gln Ser Ser Ala Ser Ala Ser Ala Ala Ala Ser
290 295 300

Ala Ala Ser Thr Val Ala Asn Ser Val Ser Arg Leu Ser Ser Pro Ser
305 310 315 320

Ala Val Ser Arg Val Ser Ser Ala Val Ser Ser Leu Val Ser Asn Gly
325 330 335

Gln Val Asn Met Ala Ala Leu Pro Asn Ile Ile Ser Asn Ile Ser Ser
340 345 350

Ser Val Ser Ala Ser Ala Pro Gly Ala Ser Gly Cys Glu Val Ile Val
355 360 365

Gln Ala Leu Leu Glu Val Ile Thr Ala Leu Val Gln Ile Val Ser Ser
370 375 380

Ser Ser Val Gly Tyr Ile Asn Pro Ser Ala Val Asn Gln Ile Thr Asn
385 390 395 400

eol f-seql . txt

Val Val Ala Asn Ala Met Ala Glu Val Met Gly Lys Leu Ala Ala Ala
405 410 415

Leu Glu His His His His His
420

<210> 6
<211> 137
<212> PRT
<213> Euprosthenops australis

<220>
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<222> (6)..(6)
<223> deletion (del taHis)

<400> 6

Gly Ser Gly Asn Ser His Thr Thr Pro Trp Thr Asn Pro Gly Leu Ala
1 5 10 15

Gl u Asn Phe Met Asn Ser Phe Met Glu Gly Leu Ser Ser Met Pro Gly
20 25 30

Phe Thr Ala Ser Glu Leu Asp Asp Met Ser Thr Ile Ala Glu Ser Met
35 40 45

Val Glu Ser Ile Glu Ser Leu Ala Ala Glu Gly Arg Thr Ser Pro Asn
50 55 60

Lys Leu Glu Ala Leu Asn Met Ala Phe Ala Ser Ser Met Ala Glu Ile
65 70 75 80

Ala Ala Ser Glu Glu Gly Gly Ser Leu Ser Thr Lys Thr Ser Ser
85 90 95

Ile Ala Ser Ala Met Ser Asn Ala Phe Leu Glu Thr Thr Gly Val Val
100 105 110

Asn Glu Pro Phe Ile Asn Glu Ile Thr Glu Leu Val Ser Met Phe Ala
115 120 125

Glu Ala Gly Met Asn Asp Val Ser Ala
130 135

<210> 7
<211> 98
<212> PRT
<213> Euprosthenops australis

<400> 7

Ser Arg Leu Ser Ser Pro Ser Ala Val Ser Arg Val Ser Ser Ala Val
1 5 10 15

eol f-seql . txt

Ser Ser Leu Val Ser Asn Gl y Gl n Val Asn Met Al a Al a Leu Pro Asn
20 25 30

Ile Ile Ser Asn Ile Ser Ser Val Ser Al a Ser Al a Pro Gl y Al a
35 40 45

Ser Gl y Cys Gl u Val Ile Val Gl n Al a Leu Leu Gl u Val Ile Thr Al a
50 55 60

Leu Val Gl n Ile Val Ser Ser Ser Val Gl y Tyr Ile Asn Pro Ser
65 70 75 80

Al a Val Asn Gl n Ile Thr Asn Val Val Al a Asn Al a Met Al a Gl n Val
85 90 95

Met Gl y

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<211> 131
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<213> Artificial Sequence

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<223> Consensus sequence derived from spidroin NT fragments

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<223> Leu

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<223> Asn

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<222> (42)..(42)
<223> Gl n

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<223> Ser

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<222> (50)..(50)
<223> Lys

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<222> (56)..(56)
<223> Arg

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<222> (84)..(84)
<223> Leu

eol f-seql . txt

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<223> Asn

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<223> Leu

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<222> (124).. (124)
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Gl n Al a Asn Thr Pro Trp Ser Ser Pro Asn Leu Al a Asp Al a Phe Ile
1 5 10 15

Asn Ser Phe Met Ser Al a Al a Ser Ser Ser Gly Al a Phe Ser Al a Asp
20 25 30

Gl n Leu Asp Asp Met Ser Thr Ile Gly Asp Thr Leu Met Ser Al a Met
35 40 45

Asp Asn Met Gly Arg Ser Gly Lys Ser Thr Lys Ser Lys Leu Gl n Al a
50 55 60

Leu Asn Met Al a Phe Al a Ser Ser Met Al a Gl u Ile Al a Al a Al a Gl u
65 70 75 80

Ser Gl y Gl y Gl y Ser Val Gl y Val Lys Thr Asn Al a Ile Ser Asp Al a
85 90 95

Leu Ser Ser Al a Phe Tyr Gl n Thr Thr Gl y Ser Val Asn Pro Gl n Phe
100 105 110

Val Asn Gl u Ile Arg Ser Leu Ile Gl y Met Phe Al a Gl n Al a Ser Al a
115 120 125

Asn Gl u Val
130

<210> 9
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<212> PRT
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<220>
<223> Consensus sequence derived from known MaSp1 and MaSp2 proteins

eol f-seql . txt

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<222> (1)..(71)
<223> Sequence length present in known species variants

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<223> Glu

<400> 9

Ser Arg Leu Ser Ser Pro Gln Ala Ser Ser Arg Val Ser Ser Ala Val
1 5 10 15

Ser Asn Leu Val Ser Ser Gly Pro Thr Asn Ser Ala Ala Leu Ser Asn
20 25 30

Thr Ile Ser Asn Val Val Ser Gln Ile Ser Ala Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Val Leu Val Gln Ala Leu Leu Glu Val Val Ser Ala
50 55 60

Leu Val His Ile Leu Gly Ser Ser Ser Ile Gly Gln Val Asn Tyr Gly
65 70 75 80

Ser Ala Gly Gln Ala Thr Gln Ile Val Gly Gln Ser Val Ala Gln Ala
85 90 95

Leu Gly Glu Phe
100

<210> 10
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eol f-seql . txt

<222> (84)..(106)

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eol f-seql . txt

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eol f-seql . txt

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eol f-seql . txt

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<220>

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<222> (982)..(998)

<220>

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<222> (999)..(1013)

<220>

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Gl n Gl y Al a Gl y Gl y Asn Al a 15

Al a Al a Al a Gl y Gl n Gl y Gl n Gl y Gl y Tyr Gl y Gl y Leu Gl y Gl n 20 25 30

Gl y Gl y Tyr Gl y Gl n Gl y Al a Gl y Ser Ser Al a 35 40 45

Al a Al a Al a Al a Al a Al a Gl y Arg Gl y Gl n Gl y Gl y Tyr Gl y 50 55 60

Gl n Gl y Ser Gl y Gl y Asn Al a 65 70 75 80

eol f-seql . txt

Ala Ala Ser Gly Glu Gly Glu Gly Glu Gly Glu Gly Glu Gly Glu
85 90 95

Gly Gly Tyr Gly Glu Gly Ala Gly Ser Ser Ala Ala Ala Ala Ala
100 105 110

Ala Ala Ala Ala Ala Ala Ala Gly Glu Gly Glu Gly Arg Tyr Gly
115 120 125

Gly Gly Ala Gly Gly Asn Ala Ala Ala Ala Ala Ala Ala Ala Ala
130 135 140

Ala Ala Ala Gly Glu Gly Glu Gly Glu Gly Glu Gly Glu Leu Gly Glu
145 150 155 160

Gly Gly Tyr Gly Glu Gly Ala Gly Ser Ser Ala Ala Ala Ala Ala
165 170 175

Ser Ala Ala Ala Ala Ala Ala Gly Arg Gly Glu Gly Glu Tyr Gly Glu
180 185 190

Gly Ala Gly Gly Asn Ala Ala Ala Ala Ala Ala Ala Ala Ala
195 200 205

Ala Ala Ala Gly Glu Gly Glu Gly Glu Gly Tyr Gly Glu Leu Gly Glu
210 215 220

Gly Gly Tyr Gly Glu Gly Ala Gly Ser Ser Ala Ala Ala Ala Ala
225 230 235 240

Ala Ala Ala Ala Ala Ala Ala Gly Glu Gly Glu Gly Glu Gly Glu
245 250 255

Arg Tyr Gly Glu Gly Ala Gly Ser Ser Ala Ala Ala Ala Ala
260 265 270

Ala Ala Ala Ala Ala Ala Ala Gly Glu Gly Glu Gly Glu Gly Glu
275 280 285

Gly Ala Gly Gly Asn Ala Ala Ala Ala Ala Ala Ala Ala
290 295 300

Ala Ala Gly Glu Gly Glu Gly Glu Gly Glu Gly Leu Gly Glu Gly
305 310 315 320

Gly Tyr Gly Glu Gly Ala Gly Ser Ser Ala Ala Ala Ala Ala
325 330 335

Ala Ala Ala Ala Ala Ala Gly Arg Gly Glu Gly Glu Tyr Gly Glu Gly
340 345 350

eol f-seql . txt

Ala Gly Gly Asn Ala Ala Ala Ala Ala Ala Ala Ala Ala Glu Ala Ala
355 360 365

Ala Ala Gly Gly Glu Gly Gly Tyr Gly Leu Gly Glu Gly
370 375 380

Gly Tyr Gly Glu Gly Ala Gly Ser Ser Ala Ala Ala Ala Ala Ala
385 390 395 400

Ala Ala Ala Ala Ala Ala Gly Arg Gly Glu Gly Gly Tyr Gly Glu Gly
405 410 415

Ala Gly Gly Asn Ala
420 425 430

Ala Ala Gly Gly Glu Gly Gly Tyr Gly Leu Gly Glu Gly
435 440 445

Gly Tyr Gly Glu Gly Ala Gly Ser Ser Ala Ala Ala Ala Ala Ala
450 455 460

Ala Ala Ala Ala Ala Ala Gly Glu Gly Glu Gly Glu Gly Glu Gly Arg
465 470 475 480

Tyr Gly Glu Gly Ala Gly Ser Ser Ala Ala Ala Ala Ala Ala
485 490 495

Ala Ala Ala Ala Ala Ala Gly Arg Gly Glu Gly Glu Gly Glu Gly
500 505 510

Ser Gly Gly Asn Ala
515 520 525

Ser Gly Glu Gly Ser Glu Gly Gly Glu Gly Glu Gly Glu Gly
530 535 540

Tyr Gly Glu Gly Ala Gly Ser Ser Ala Ala Ala Ala Ala
545 550 555 560

Ala Ala Ala Ala Ala Ser Gly Arg Gly Glu Gly Glu Gly Tyr Gly Glu Gly
565 570 575

Ala Gly Gly Asn Ala Ala Ala Ala Ala Ala Ala Ala
580 585 590

Ala Ala Gly Glu Gly Glu Gly Gly Tyr Gly Leu Gly Glu Gly
595 600 605

Gly Tyr Gly Glu Gly Ala Gly Ser Ser Ala Ala Ala Ala Ala
610 615 620

eol f-seql . txt

Ala Ala Ala Ala Ala Gly 630 Glu Glu Glu Glu Glu Glu Tyr 640
625
Gly Glu Glu Glu Glu Ser Ser Ala Ala Ala Ala Ala Ala Ala Ala Ala 655
645 650
Ala Ala Ala Ala Ala Gly Arg 665 Glu Glu Glu Tyr Glu Glu Ser 670
660
Gly Glu Asn Ala Ser 685
675
Gly Glu Glu Glu Glu Glu 695 Glu Glu Glu Glu Glu Glu Glu Glu Glu Tyr 700
690
Gly Glu Glu Glu Glu Ser 710 Ser Ala Ala Ala Ala Ala Ala Ala Ala Ala 720
705
Ala Ala Ala Ala Ala Gly 730 Glu Glu Glu Glu Glu Tyr Glu Glu Ala 735
725
Gly Glu Asn Ala 750
740
Gly Glu Glu Glu Glu Glu 760 Glu Glu Glu Leu Glu Glu Glu Glu Tyr 765
755
Gly Glu Glu Glu Glu Ser 775 Ala 780
770
Ala Ala Ala Ala Ala Gly 790 Arg Glu Glu Glu Glu Tyr Glu Glu Val 800
785
Gly Glu Asn Ala 815
805
Gly Glu Glu Glu Glu Glu 820 Glu Glu Glu Leu Glu Glu Glu Glu Tyr 830
825
Gly Glu Glu Glu Glu Ser 840 Ser Ala Ala Ala Ala Ala Ala Ala Ala 845
835
Ala Ala Ala Ala Ala Gly 855 Arg Glu Glu Glu Glu Tyr Glu Glu Glu Ser 880
850
Gly Glu Asn Ala Ser 880
865
870
Gly Glu Glu Glu Ser 885 Glu Glu Glu Glu Glu Glu Glu Glu Glu Tyr 895
890

eol f-seql . txt

Gly Glu Glu Gly Ala Gly Ser Ser Ala Ala Ala Ala Ala Ala Ala Ala Ala
900 905 910

Ala Ala Ala Ala Ala Ser Gly Arg Gly Glu Glu Gly Tyr Gly Glu Glu Gly Ala
915 920 925

Gly Glu Asn Ala
930 935 940

Ala Gly Glu Gly Gly Glu Gly Tyr Gly Gly Leu Gly Glu Glu Gly
945 950 955 960

Tyr Gly Glu Gly Ala Gly Ser Ser Ala Ala Ala Ala Ala Ala Ala Ala
965 970 975

Ala Ala Ala Ala Ala Gly Gly Glu Glu Gly Glu Glu Glu Gly Tyr Gly
980 985 990

Glu Glu Ser Gly Gly Ser Ala
995 1000 1005

Ala Ala Ala Ala Ala Gly Arg Gly Glu Glu Gly Tyr Gly Glu Glu
1010 1015 1020

Ser Gly Gly Asn Ala
1025 1030 1035

Ala Ala Ala Ala Ala Gly Glu Gly Glu Glu Gly Tyr Gly Arg Glu
1040 1045 1050

Ser Glu Gly Ala Gly Ser Ala Ala Ala Ala Ala Ala Ala
1055 1060 1065

Ala Ala Ala Ala Ala Ala Gly Ser Gly Glu Glu Gly Tyr Gly Gly Glu
1070 1075 1080

Gly Glu Glu Gly Gly Tyr Gly Glu Ser Ser Ala Ser Ala Ser
1085 1090 1095

Ala Ser Ala Ala Ser Thr Val Ala Asn Ser Val Ser
1100 1105 1110

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australis MaSp1

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eol f-seql . txt

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<223> Ser

<220>
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<223> Tyr

<220>
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<223> Gl n

<400> 11

Gl y Gl n Gl y Gl y Gl n Gl y Gl n Gl y Gl y Leu Gl y Gl n Gl y Gl y Tyr
1 5 10 15

Gl y Gl n Gl y Al a Gl y Ser Ser
20

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<220>
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australis MaSp1

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<222> (9)..(9)
<223> Arg

<220>
<221> VARI ANT
<222> (14)..(14)
<223> Ser

<220>
<221> VARI ANT
<222> (16)..(16)
<223> Gl y

<400> 12

Gl y Gl n Gl y Gl y Gl n Gl y Gl n Gl y Gl y Tyr Gl y Gl n Gl y Al a Gl y Ser
1 5 10 15

Ser

<210> 13
<211> 14
<212> PRT
<213> Artificial Sequence

<220>
<223> Consensus sequence derived from internal repeats of Euprosthenops
australis MaSp1

eol f-seql . txt

<220>
<221> VARI ANT
<222> (2)..(2)
<223> Gl n

<220>
<221> VARI ANT
<222> (6)..(6)
<223> Arg

<220>
<221> VARI ANT
<222> (11)..(11)
<223> Ser

<220>
<221> VARI ANT
<222> (11)..(11)
<223> Val

<400> 13

Gl y Arg Gl y Gl n Gl y Gl y Tyr Gl y Gl n Gl y Al a Gl y Gl y Asn
1 5 10

<210> 14
<211> 100
<212> PRT
<213> Euprosthenops sp

<400> 14

Ser Arg Leu Ser Ser Pro Gl u Al a Ser Ser Arg Val Ser Ser Al a Val
1 5 10 15

Ser Asn Leu Val Ser Ser Gl y Pro Thr Asn Ser Al a Al a Leu Ser Ser
20 25 30

Thr Ile Ser Asn Val Val Ser Gl n Ile Gl y Al a Ser Asn Pro Gl y Leu
35 40 45

Ser Gl y Cys Asp Val Leu Val Gl n Al a Leu Leu Gl u Val Val Ser Al a
50 55 60

Leu Ile His Ile Leu Gl y Ser Ser Ser Ile Gl y Gl n Val Asn Tyr Gl y
65 70 75 80

Ser Al a Gl y Gl n Al a Thr Gl n Leu Val Gl y Gl n Ser Val Tyr Gl n Al a
85 90 95

Leu Gl y Gl u Phe
100

<210> 15
<211> 98
<212> PRT
<213> Euprosthenops australis

<400> 15

eol f-seql . txt

Ser Arg Leu Ser Ser Pro Ser Ala Val Ser Arg Val Ser Ser Ala Val
1 5 10 15

Ser Ser Leu Val Ser Asn Gly Glu Val Asn Met Ala Ala Leu Pro Asn
20 25 30

Ile Ile Ser Asn Ile Ser Ser Val Ser Ala Ser Ala Pro Gly Ala
35 40 45

Ser Gly Cys Glu Val Ile Val Glu Ala Leu Leu Glu Val Ile Thr Ala
50 55 60

Leu Val Glu Ile Val Ser Ser Ser Val Gly Tyr Ile Asn Pro Ser
65 70 75 80

Ala Val Asn Glu Ile Thr Asn Val Val Ala Asn Ala Met Ala Glu Val
85 90 95

Met Gly

<210> 16

<211> 99

<212> PRT

<213> Argiope trifasciata

<400> 16

Ser Arg Leu Ser Ser Pro Gly Ala Ala Ser Arg Val Ser Ser Ala Val
1 5 10 15

Thr Ser Leu Val Ser Ser Gly Gly Pro Thr Asn Ser Ala Ala Leu Ser
20 25 30

Asn Thr Ile Ser Asn Val Val Ser Glu Ile Ser Ser Ser Asn Pro Gly
35 40 45

Leu Ser Gly Cys Asp Val Leu Val Glu Ala Leu Leu Glu Ile Val Ser
50 55 60

Ala Leu Val His Ile Leu Gly Ser Ala Asn Ile Gly Glu Val Asn Ser
65 70 75 80

Ser Gly Val Gly Arg Ser Ala Ser Ile Val Gly Glu Ser Ile Asn Glu
85 90 95

Ala Phe Ser

<210> 17

<211> 89

<212> PRT

<213> Cyrtophora moluccensis

eol f-seql . txt

<400> 17

Ser His Leu Ser Ser Pro Glu Ala Ser Ser Arg Val Ser Ser Ala Val
1 5 10 15

Ser Asn Leu Val Ser Ser Gly Ser Thr Asn Ser Ala Ala Leu Pro Asn
20 25 30

Thr Ile Ser Asn Val Val Ser Gln Ile Ser Ser Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Val Leu Val Gln Ala Leu Leu Glu Val Val Ser Ala
50 55 60

Leu Ile His Ile Leu Gly Ser Ser Ser Ile Gly Gln Val Asn Tyr Gly
65 70 75 80

Ser Ala Gly Gln Ala Thr Gln Ile Val
85

<210> 18

<211> 98

<212> PRT

<213> *Latrodectus geometricus*

<400> 18

Ser Ala Leu Ala Ala Pro Ala Thr Ser Ala Arg Ile Ser Ser His Ala
1 5 10 15

Ser Thr Leu Leu Ser Asn Gly Pro Thr Asn Pro Ala Ser Ile Ser Asn
20 25 30

Val Ile Ser Asn Ala Val Ser Gln Ile Ser Ser Ser Asn Pro Gly Ala
35 40 45

Ser Ser Cys Asp Val Leu Val Gln Ala Leu Leu Glu Leu Val Thr Ala
50 55 60

Leu Leu Thr Ile Ile Gly Ser Ser Asn Val Gly Asn Val Asn Tyr Asp
65 70 75 80

Ser Ser Gly Gln Tyr Ala Gln Val Val Ser Gln Ser Val Gln Asn Ala
85 90 95

Phe Val

<210> 19

<211> 98

<212> PRT

<213> *Latrodectus hesperus*

<400> 19

eof f-seql.txt

Ser Ala Leu Ser Ala Pro Ala Thr Ser Ala Arg Ile Ser Ser His Ala
1 5 10 15

Ser Ala Leu Leu Ser Ser Gly Pro Thr Asn Pro Ala Ser Ile Ser Asn
20 25 30

Val Ile Ser Asn Ala Val Ser Gln Ile Ser Ser Ser Asn Pro Gly Ala
35 40 45

Ser Ala Cys Asp Val Leu Val Gln Ala Leu Leu Glu Leu Val Thr Ala
50 55 60

Leu Leu Thr Ile Ile Gly Ser Ser Asn Ile Gly Ser Val Asn Tyr Asp
65 70 75 80

Ser Ser Gly Gln Tyr Ala Gln Val Val Thr Gln Ser Val Gln Asn Val
85 90 95

Phe Gly

<210> 20

<211> 93

<212> PRT

<213> Macrothel e hol sti

<400> 20

Ser His Leu Ser Ser Pro Glu Ala Ser Ser Arg Val Ser Ser Ala Val
1 5 10 15

Ser Asn Leu Val Ser Gly Gly Ser Thr Asn Ser Ala Ala Leu Pro Asn
20 25 30

Thr Ile Ser Asn Val Val Ser Gln Ile Ser Ser Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Val Leu Val Gln Ala Leu Leu Glu Val Val Ser Ala
50 55 60

Leu Ile His Ile Leu Gly Ser Ser Ser Ile Gly Gln Val Asp Tyr Gly
65 70 75 80

Ser Ala Gly Gln Ala Thr Gln Ile Val Gly Gln Ser Ala
85 90

<210> 21

<211> 98

<212> PRT

<213> Nephila clavipes

<400> 21

Ser Arg Leu Ser Ser Pro Gln Ala Ser Ser Arg Val Ser Ser Ala Val
1 5 10 15

eol f-seql . txt

Ser Asn Leu Val Al a Ser Gl y Pro Thr Asn Ser Al a Al a Leu Ser Ser
20 25 30

Thr Ile Ser Asn Val Val Ser Gl n Ile Gl y Al a Ser Asn Pro Gl y Leu
35 40 45

Ser Gl y Cys Asp Val Leu Ile Gl n Al a Leu Leu Gl u Val Val Ser Al a
50 55 60

Leu Ile Gl n Ile Leu Gl y Ser Ser Ser Ile Gl y Gl n Val Asn Tyr Gl y
65 70 75 80

Ser Al a Gl y Gl n Al a Thr Gl n Ile Val Gl y Gl n Ser Val Tyr Gl n Al a
85 90 95

Leu Gl y

<210> 22

<211> 89

<212> PRT

<213> Nephi la pilipes

<400> 22

Ser Arg Leu Ser Ser Pro Gl u Al a Ser Ser Arg Val Ser Ser Al a Val
1 5 10 15

Ser Asn Leu Val Ser Ser Gl y Pro Thr Asn Ser Al a Al a Leu Ser Asn
20 25 30

Thr Ile Ser Asn Val Val Ser Gl n Ile Ser Ser Ser Asn Pro Gl y Leu
35 40 45

Ser Gl y Cys Asp Val Leu Val Gl n Al a Leu Leu Gl u Val Val Ser Al a
50 55 60

Leu Ile His Ile Leu Gl y Ser Ser Ser Ile Gl y Gl n Val Asn Tyr Gl y
65 70 75 80

Ser Al a Gl y Gl n Al a Thr Gl n Ile Val
85

<210> 23

<211> 87

<212> PRT

<213> Nephi la madagascariensis

<400> 23

Ser Arg Leu Ser Ser Pro Gl n Al a Ser Ser Arg Val Ser Ser Al a Val
1 5 10 15

eol f-seql . txt

Ser Asn Leu Val Ala Ser Gly Pro Thr Asn Ser Ala Ala Leu Ser Ser
20 25 30

Thr Ile Ser Asn Ala Val Ser Glu Ile Gly Ala Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Val Leu Ile Glu Ala Leu Leu Glu Val Val Ser Ala
50 55 60

Leu Ile His Ile Leu Gly Ser Ser Ser Ile Gly Glu Val Asn Tyr Gly
65 70 75 80

Ser Ala Gly Glu Ala Thr Glu
85

<210> 24

<211> 87

<212> PRT

<213> Nephi la senegalensis

<400> 24

Ser Arg Leu Ser Ser Pro Glu Ala Ser Ser Arg Val Ser Ser Ala Val
1 5 10 15

Ser Asn Leu Val Ser Ser Gly Pro Thr Asn Ser Ala Ala Leu Ser Ser
20 25 30

Thr Ile Ser Asn Val Val Ser Glu Ile Gly Ala Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Val Leu Ile Glu Ala Leu Leu Glu Val Val Ser Ala
50 55 60

Leu Val His Ile Leu Gly Ser Ser Ser Ile Gly Glu Val Asn Tyr Gly
65 70 75 80

Ser Ala Gly Glu Ala Thr Glu
85

<210> 25

<211> 89

<212> PRT

<213> Octonoba varians

<400> 25

Ser Arg Leu Ser Ser Pro Glu Ala Ser Ser Arg Val Ser Ser Ala Val
1 5 10 15

Ser Asn Leu Val Ser Ser Gly Pro Thr Asn Ser Ala Ala Leu Ser Asn
20 25 30

Thr Ile Ser Asn Val Val Ser Glu Ile Ser Ser Ser Asn Pro Gly Leu
35 40 45

eol f-seql . txt

Ser Gl y Cys Asp Val Leu Val Gl n Al a Leu Leu Gl u Val Val Ser Al a
50 55 60

Pro Ile His Ile Leu Gl y Ser Ser Ser Ile Gl y Gl n Val Asn Tyr Gl y
65 70 75 80

Ser Al a Gl y Gl n Al a Thr Gl n Ile Val
85

<210> 26
<211> 89
<212> PRT
<213> Psechrus si nensis

<400> 26

Ser Arg Leu Ser Ser Pro Gl u Al a Ser Ser Arg Val Ser Ser Al a Val
1 5 10 15

Ser Asn Leu Val Ser Ser Gl y Pro Thr Asn Ser Al a Al a Leu Pro Asn
20 25 30

Thr Ile Ser Asn Val Val Ser Gl n Ile Ser Ser Ser Asn Pro Gl y Leu
35 40 45

Ser Gl y Cys Asp Val Leu Val Gl n Al a Leu Leu Gl u Val Val Ser Al a
50 55 60

Leu Ile His Ile Leu Gl y Ser Ser Ser Ile Gl y Gl n Val Asn Tyr Gl y
65 70 75 80

Ser Al a Gl y Gl n Al a Thr Gl n Ile Val
85

<210> 27
<211> 88
<212> PRT
<213> Tetragnatha kauai ensis

<400> 27

Ser Leu Leu Ser Ser Pro Al a Ser Asn Al a Arg Ile Ser Ser Al a Val
1 5 10 15

Ser Al a Leu Al a Ser Gl y Al a Al a Ser Gl y Pro Gl y Tyr Leu Ser Ser
20 25 30

Val Ile Ser Asn Val Val Ser Gl n Val Ser Ser Asn Ser Gl y Gl y Leu
35 40 45

Val Gl y Cys Asp Thr Leu Val Gl n Al a Leu Leu Gl u Al a Al a Al a Al a
50 55 60

eof f-seql.txt

Leu Val His Val Leu Ala Ser Ser Ser Gly Gly Gln Val Asn Leu Asn
65 70 75 80

Thr Ala Gly Tyr Thr Ser Gln Leu
85

<210> 28

<211> 88

<212> PRT

<213> *Tetragnatha versicolor*

<400> 28

Ser Arg Leu Ser Ser Pro Ala Ser Asn Ala Arg Ile Ser Ser Ala Val
1 5 10 15

Ser Ala Leu Ala Ser Gly Gly Ala Ser Ser Pro Gly Tyr Leu Ser Ser
20 25 30

Ile Ile Ser Asn Val Val Ser Gln Val Ser Ser Asn Asn Asp Gly Leu
35 40 45

Ser Gly Cys Asp Thr Val Val Gln Ala Leu Leu Glu Val Ala Ala Ala
50 55 60

Leu Val His Val Leu Ala Ser Ser Asn Ile Gly Gln Val Asn Leu Asn
65 70 75 80

Thr Ala Gly Tyr Thr Ser Gln Leu
85

<210> 29

<211> 89

<212> PRT

<213> *Araneus bicentenarius*

<400> 29

Ser Arg Leu Ser Ser Ser Ala Ala Ser Ser Arg Val Ser Ser Ala Val
1 5 10 15

Ser Ser Leu Val Ser Ser Gly Pro Thr Thr Pro Ala Ala Leu Ser Asn
20 25 30

Thr Ile Ser Ser Ala Val Ser Gln Ile Ser Ala Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Val Leu Val Gln Ala Leu Leu Glu Val Val Ser Ala
50 55 60

Leu Val His Ile Leu Gly Ser Ser Ser Val Gly Gln Ile Asn Tyr Gly
65 70 75 80

Ala Ser Ala Gln Tyr Ala Gln Met Val
85

eol f-seql . txt

<210> 30

<211> 97

<212> PRT

<213> Argi ope amoena

<400> 30

Arg Leu Ser Ser Pro Gln Ala Ser Ser Arg Val Ser Ser Ala Val Ser
1 5 10 15

Thr Leu Val Ser Ser Gly Pro Thr Asn Pro Ala Ser Leu Ser Asn Ala
20 25 30

Ile Gly Ser Val Val Ser Gln Val Ser Ala Ser Asn Pro Gly Leu Pro
35 40 45

Ser Cys Asp Val Leu Val Gln Ala Leu Leu Glu Ile Val Ser Ala Leu
50 55 60

Val His Ile Leu Gly Ser Ser Ser Ile Gly Gln Ile Asn Tyr Ser Ala
65 70 75 80

Ser Ser Gln Tyr Ala Arg Leu Val Gly Gln Ser Ile Ala Gln Ala Leu
85 90 95

Gly

<210> 31

<211> 82

<212> PRT

<213> Argi ope auranti a

<400> 31

Ser Arg Leu Ser Ser Pro Gln Ala Ser Ser Arg Val Ser Ser Ala Val
1 5 10 15

Ser Thr Leu Val Ser Ser Gly Pro Thr Asn Pro Ala Ala Leu Ser Asn
20 25 30

Ala Ile Ser Ser Val Val Ser Gln Val Ser Ala Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Val Leu Val Gln Ala Leu Leu Glu Leu Val Ser Ala
50 55 60

Leu Val His Ile Leu Gly Ser Ser Ser Ile Gly Gln Ile Asn Tyr Ala
65 70 75 80

Ala Ser

eol f-seql . txt

<210> 32
<211> 98
<212> PRT
<213> Argi ope tri fasci ata

<400> 32

Ser Arg Leu Ser Ser Pro Gln Ala Ser Ser Arg Val Ser Ser Ala Val
1 5 10 15

Ser Thr Leu Val Ser Ser Gly Pro Thr Asn Pro Ala Ser Leu Ser Asn
20 25 30

Ala Ile Ser Ser Val Val Ser Gln Val Ser Ser Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Val Leu Val Gln Ala Leu Leu Glu Ile Val Ser Ala
50 55 60

Leu Val His Ile Leu Gly Ser Ser Ser Ile Gly Gln Ile Asn Tyr Ala
65 70 75 80

Ala Ser Ser Gln Tyr Ala Gln Leu Val Gly Gln Ser Leu Thr Gln Ala
85 90 95

Leu Gly

<210> 33
<211> 89
<212> PRT
<213> Gasteracantha mammosa

<400> 33

Ser Arg Leu Ser Ser Pro Gln Ala Gly Ala Arg Val Ser Ser Ala Val
1 5 10 15

Ser Ala Leu Val Ala Ser Gly Pro Thr Ser Pro Ala Ala Val Ser Ser
20 25 30

Ala Ile Ser Asn Val Ala Ser Gln Ile Ser Ala Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Val Leu Val Gln Ala Leu Leu Glu Ile Val Ser Ala
50 55 60

Leu Val Ser Ile Leu Ser Ser Ala Ser Ile Gly Gln Ile Asn Tyr Gly
65 70 75 80

Ala Ser Gly Gln Tyr Ala Ala Met Ile
85

<210> 34
<211> 90

eol f-seql . txt

<212> PRT

<213> *Latroductus geometricus*

<400> 34

Ser Ala Leu Ser Ser Pro Thr Thr His Ala Arg Ile Ser Ser His Ala
1 5 10 15

Ser Thr Leu Leu Ser Ser Gly Pro Thr Asn Ser Ala Ala Ile Ser Asn
20 25 30

Val Ile Ser Asn Ala Val Ser Gln Val Ser Ala Ser Asn Pro Gly Ser
35 40 45

Ser Ser Cys Asp Val Leu Val Gln Ala Leu Leu Glu Leu Ile Thr Ala
50 55 60

Leu Ile Ser Ile Val Asp Ser Ser Asn Ile Gly Gln Val Asn Tyr Gly
65 70 75 80

Ser Ser Gly Gln Tyr Ala Gln Met Val Gly
85 90

<210> 35

<211> 98

<212> PRT

<213> *Latroductus hesperus*

<400> 35

Ser Ala Leu Ser Ser Pro Thr Thr His Ala Arg Ile Ser Ser His Ala
1 5 10 15

Ser Thr Leu Leu Ser Ser Gly Pro Thr Asn Ala Ala Ala Leu Ser Asn
20 25 30

Val Ile Ser Asn Ala Val Ser Gln Val Ser Ala Ser Asn Pro Gly Ser
35 40 45

Ser Ser Cys Asp Val Leu Val Gln Ala Leu Leu Glu Ile Ile Thr Ala
50 55 60

Leu Ile Ser Ile Leu Asp Ser Ser Ser Val Gly Gln Val Asn Tyr Gly
65 70 75 80

Ser Ser Gly Gln Tyr Ala Gln Ile Val Gly Gln Ser Met Gln Gln Ala
85 90 95

Met Gly

<210> 36

<211> 97

<212> PRT

<213> *Nephila clavipes*

eol f-seql . txt

<400> 36

Ser Arg Leu Al a Ser Pro Asp Ser Gly Al a Arg Val Al a Ser Al a Val
1 5 10 15

Ser Asn Leu Val Ser Ser Gly Pro Thr Ser Ser Al a Al a Leu Ser Ser
20 25 30

Val Ile Ser Asn Al a Val Ser Gln Ile Gly Al a Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Val Leu Ile Gln Al a Leu Leu Glu Ile Val Ser Al a
50 55 60

Cys Val Thr Ile Leu Ser Ser Ser Ser Ile Gly Gln Val Asn Tyr Gly
65 70 75 80

Al a Al a Ser Gln Phe Al a Gln Val Val Gly Gln Ser Val Leu Ser Al a
85 90 95

Phe

<210> 37

<211> 82

<212> PRT

<213> Nephi la madagascari ensis

<400> 37

Ser Arg Leu Al a Ser Pro Asp Ser Gly Al a Arg Val Al a Ser Al a Val
1 5 10 15

Ser Asn Leu Val Ser Ser Gly Pro Thr Ser Ser Al a Al a Leu Ser Ser
20 25 30

Val Ile Ser Asn Al a Val Ser Gln Ile Gly Al a Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Val Leu Ile Gln Al a Leu Leu Glu Ile Val Ser Al a
50 55 60

Cys Val Thr Ile Leu Ser Ser Ser Ser Ile Gly Gln Val Asn Tyr Gly
65 70 75 80

Al a Al a

<210> 38

<211> 82

<212> PRT

<213> Nephi la senegal ensis

eol f-seql . txt

<220>
<221> mi sc_feature
<222> (35)..(35)
<223> Xaa can be any naturally occurring amino acid

<220>
<221> mi sc_feature
<222> (56)..(56)
<223> Xaa can be any naturally occurring amino acid

<400> 38

Ser Arg Leu Ala Ser Pro Asp Ser Gly Ala Arg Val Ala Ser Ala Val
1 5 10 15

Ser Asn Leu Val Ser Ser Gly Pro Thr Ser Ser Ala Ala Leu Ser Ser
20 25 30

Val Ile Xaa Asn Ala Val Ser Gln Ile Gly Ala Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Val Leu Ile Xaa Ala Leu Leu Glu Ile Val Ser Ala
50 55 60

Cys Val Thr Ile Leu Ser Ser Ser Ser Ile Gly Gln Val Asn Tyr Gly
65 70 75 80

Ala Ala

<210> 39
<211> 71
<212> PRT
<213> Dolomedes tenebrosus

<400> 39

Ser Arg Leu Ser Ser Pro Glu Ala Ala Ser Arg Val Ser Ser Ala Val
1 5 10 15

Ser Ser Leu Val Ser Asn Gly Gln Val Asn Val Asp Ala Leu Pro Ser
20 25 30

Ile Ile Ser Asn Leu Ser Ser Ser Ile Ser Ala Ser Ala Thr Thr Ala
35 40 45

Ser Asp Cys Glu Val Leu Val Gln Val Leu Leu Glu Val Val Ser Ala
50 55 60

Leu Val Gln Ile Val Cys Ser
65 70

<210> 40
<211> 97
<212> PRT
<213> Dolomedes tenebrosus

eol f-seql . txt

<400> 40

Ser Arg Leu Ser Ser Pro Glu Ala Ala Ser Arg Val Ser Ser Ala Ala Val
1 5 10 15

Ser Ser Leu Val Ser Asn Gly Glu Val Asn Val Ala Ala Leu Pro Ser
20 25 30

Ile Ile Ser Ser Leu Ser Ser Ser Ile Ser Ala Ser Ser Thr Ala Ala
35 40 45

Ser Asp Cys Glu Val Leu Val Glu Val Leu Leu Glu Ile Val Ser Ala
50 55 60

Leu Val Glu Ile Val Ser Ser Ala Asn Val Gly Tyr Ile Asn Pro Glu
65 70 75 80

Ala Ser Gly Ser Leu Asn Ala Val Gly Ser Ala Leu Ala Ala Ala Met
85 90 95

Gly

<210> 41

<211> 93

<212> PRT

<213> Araneus diadematus

<400> 41

Asn Arg Leu Ser Ser Ala Gly Ala Ala Ser Arg Val Ser Ser Asn Val
1 5 10 15

Ala Ala Ile Ala Ser Ala Gly Ala Ala Leu Pro Asn Val Ile Ser
20 25 30

Asn Ile Tyr Ser Gly Val Leu Ser Ser Gly Val Ser Ser Ser Glu Ala
35 40 45

Leu Ile Glu Ala Leu Leu Glu Val Ile Ser Ala Leu Ile His Val Leu
50 55 60

Gly Ser Ala Ser Ile Gly Asn Val Ser Ser Val Gly Val Asn Ser Ala
65 70 75 80

Leu Asn Ala Val Glu Asn Ala Val Gly Ala Tyr Ala Gly
85 90

<210> 42

<211> 98

<212> PRT

<213> Araneus diadematus

<400> 42

eol f-seql . txt

Ser Arg Leu Ser Ser Pro Ser Ala Ala Ala Arg Val Ser Ser Ala Val
1 5 10 15

Ser Leu Val Ser Asn Gly Gly Pro Thr Ser Pro Ala Ala Leu Ser Ser
20 25 30

Ser Ile Ser Asn Val Val Ser Gln Ile Ser Ala Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Ile Leu Val Gln Ala Leu Leu Glu Ile Ile Ser Ala
50 55 60

Leu Val His Ile Leu Gly Ser Ala Asn Ile Gly Pro Val Asn Ser Ser
65 70 75 80

Ser Ala Gly Gln Ser Ala Ser Ile Val Gly Gln Ser Val Tyr Arg Ala
85 90 95

Leu Ser

<210> 43
<211> 98
<212> PRT
<213> Araneus diadematus

<400> 43

Ser Arg Leu Ser Ser Pro Ala Ala Ser Ser Arg Val Ser Ser Ala Val
1 5 10 15

Ser Ser Leu Val Ser Ser Gly Pro Thr Lys His Ala Ala Leu Ser Asn
20 25 30

Thr Ile Ser Ser Val Val Ser Gln Val Ser Ala Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Val Leu Val Gln Ala Leu Leu Glu Val Val Ser Ala
50 55 60

Leu Val Ser Ile Leu Gly Ser Ser Ser Ile Gly Gln Ile Asn Tyr Gly
65 70 75 80

Ala Ser Ala Gln Tyr Thr Gln Met Val Gly Gln Ser Val Ala Gln Ala
85 90 95

Leu Ala

<210> 44
<211> 94
<212> PRT
<213> Araneus diadematus

eof f-seql . txt

<400> 44

Ser Val Tyr Leu Arg Leu Glu Pro Arg Leu Glu Val Ser Ser Ala Val
1 5 10 15

Ser Ser Leu Val Ser Ser Gly Pro Thr Asn Gly Ala Ala Val Ser Gly
20 25 30

Ala Leu Asn Ser Leu Val Ser Glu Ile Ser Ala Ser Asn Pro Gly Leu
35 40 45

Ser Gly Cys Asp Ala Leu Val Glu Ala Leu Leu Glu Leu Val Ser Ala
50 55 60

Leu Val Ala Ile Leu Ser Ser Ala Ser Ile Gly Glu Val Asn Val Ser
65 70 75 80

Ser Val Ser Glu Ser Thr Glu Met Ile Ser Glu Ala Leu Ser
85 90

<210> 45

<211> 131

<212> PRT

<213> Euprosthenops australis

<400> 45

Ser His Thr Thr Pro Trp Thr Asn Pro Gly Leu Ala Glu Asn Phe Met
1 5 10 15

Asn Ser Phe Met Glu Gly Leu Ser Ser Met Pro Gly Phe Thr Ala Ser
20 25 30

Glu Leu Asp Asp Met Ser Thr Ile Ala Glu Ser Met Val Glu Ser Ile
35 40 45

Glu Ser Leu Ala Ala Glu Glu Arg Thr Ser Pro Asn Lys Leu Glu Ala
50 55 60

Leu Asn Met Ala Phe Ala Ser Ser Met Ala Glu Ile Ala Ala Ser Glu
65 70 75 80

Glu Gly Gly Ser Leu Ser Thr Lys Thr Ser Ser Ile Ala Ser Ala
85 90 95

Met Ser Asn Ala Phe Leu Glu Thr Thr Glu Val Val Asn Glu Pro Phe
100 105 110

Ile Asn Glu Ile Thr Glu Leu Val Ser Met Phe Ala Glu Ala Glu Met
115 120 125

Asn Asp Val
130

eol f-seql . txt

<210> 46
<211> 127
<212> PRT
<213> *Latroductus geometricus*

<400> 46

Gln Ala Asn Thr Pro Trp Ser Ser Lys Gln Asn Ala Asp Ala Phe Ile
1 5 10 15

Ser Ala Phe Met Thr Ala Ala Ser Gln Ser Gly Ala Phe Ser Ser Asp
20 25 30

Gln Ile Asp Asp Met Ser Val Ile Ser Asn Thr Leu Met Ala Ala Met
35 40 45

Asp Asn Met Gly Gly Arg Ile Thr Pro Ser Lys Leu Gln Ala Leu Asp
50 55 60

Met Ala Phe Ala Ser Ser Val Ala Glu Ile Ala Ala Val Glu Gly Gln
65 70 75 80

Asn Ile Gly Val Thr Thr Asn Ala Ile Ser Asp Ala Leu Thr Ser Ala
85 90 95

Phe Tyr Gln Thr Thr Gly Val Val Asn Asn Lys Phe Ile Ser Glu Ile
100 105 110

Arg Ser Leu Ile Asn Met Phe Ala Gln Ala Ser Ala Asn Asp Val
115 120 125

<210> 47
<211> 127
<212> PRT
<213> *Latroductus hesperus*

<400> 47

Gln Ala Asn Thr Pro Trp Ser Ser Lys Ala Asn Ala Asp Ala Phe Ile
1 5 10 15

Asn Ser Phe Ile Ser Ala Ala Ser Asn Thr Gly Ser Phe Ser Gln Asp
20 25 30

Gln Met Glu Asp Met Ser Leu Ile Gly Asn Thr Leu Met Ala Ala Met
35 40 45

Asp Asn Met Gly Gly Arg Ile Thr Pro Ser Lys Leu Gln Ala Leu Asp
50 55 60

Met Ala Phe Ala Ser Ser Val Ala Glu Ile Ala Ala Ser Glu Gly Gln
65 70 75 80

Asp Leu Gly Val Thr Thr Asn Ala Ile Ala Asp Ala Leu Thr Ser Ala

eol f-seql . txt

85

90

95

Phe Tyr Glu Thr Thr Gly Val Val Asn Ser Arg Phe Ile Ser Glu Ile
 100 105 110

Arg Ser Leu Ile Gly Met Phe Ala Glu Ala Ser Ala Asn Asp Val
 115 120 125

<210> 48
 <211> 130
 <212> PRT
 <213> Nephi la clavi pes

<400> 48

Gl n Asn Thr Pro Trp Ser Ser Thr Glu Leu Ala Asp Ala Phe Ile Asn
 1 5 10 15

Ala Phe Met Asn Glu Ala Gly Arg Thr Gly Ala Phe Thr Ala Asp Glu
 20 25 30

Leu Asp Asp Met Ser Thr Ile Gly Asp Thr Ile Lys Thr Ala Met Asp
 35 40 45

Lys Met Ala Arg Ser Asn Lys Ser Ser Lys Gly Lys Leu Glu Ala Leu
 50 55 60

Asn Met Ala Phe Ala Ser Ser Met Ala Glu Ile Ala Ala Val Glu Glu
 65 70 75 80

Gly Gly Leu Ser Val Asp Ala Lys Thr Asn Ala Ile Ala Asp Ser Leu
 85 90 95

Asn Ser Ala Phe Tyr Glu Thr Thr Gly Ala Ala Asn Pro Glu Phe Val
 100 105 110

Asn Glu Ile Arg Ser Leu Ile Asn Met Phe Ala Glu Ser Ser Ala Asn
 115 120 125

Gl u Val
 130

<210> 49
 <211> 131
 <212> PRT
 <213> Argiope trifasciata

<400> 49

Gl n Gl y Ala Thr Pro Trp Glu Asn Ser Glu Leu Ala Glu Ser Phe Ile
 1 5 10 15

Ser Arg Phe Leu Arg Phe Ile Gly Glu Ser Gl y Ala Phe Ser Pro Asn
 20 25 30

eol f-seql . txt

Gl n Leu Asp Asp Met Ser Ser Ile Gl y Asp Thr Leu Lys Thr Al a Ile
35 40 45

Gl u Lys Met Al a Gl n Ser Arg Lys Ser Ser Lys Ser Lys Leu Gl n Al a
50 55 60

Leu Asn Met Al a Phe Al a Ser Ser Met Al a Gl u Ile Al a Val Al a Gl u
65 70 75 80

Gl n Gl y Gl y Leu Ser Leu Gl u Al a Lys Thr Asn Al a Ile Al a Ser Al a
85 90 95

Leu Ser Al a Al a Phe Leu Gl u Thr Thr Gl y Tyr Val Asn Gl n Gl n Phe
100 105 110

Val Asn Gl u Ile Lys Thr Leu Ile Phe Met Ile Al a Gl n Al a Ser Ser
115 120 125

Asn Gl u Ile
130

<210> 50

<211> 124

<212> PRT

<213> *Latrodectus geometricus*

<400> 50

Leu Arg Trp Ser Ser Lys Asp Asn Al a Asp Arg Phe Ile Asn Al a Phe
1 5 10 15

Leu Gl n Al a Al a Ser Asn Ser Gl y Al a Phe Ser Ser Asp Gl n Val Asp
20 25 30

Asp Met Ser Val Ile Gl y Asn Thr Leu Met Thr Al a Met Asp Asn Met
35 40 45

Gl y Gl y Arg Ile Thr Pro Ser Lys Leu Gl n Al a Leu Asp Met Al a Phe
50 55 60

Al a Ser Ser Val Al a Gl u Ile Al a Val Al a Asp Gl y Gl n Asn Val Gl y
65 70 75 80

Gl y Al a Thr Asn Al a Ile Ser Asn Al a Leu Arg Ser Al a Phe Tyr Gl n
85 90 95

Thr Thr Gl y Val Val Asn Asn Gl n Phe Ile Ser Gl u Ile Ser Asn Leu
100 105 110

Ile Asn Met Phe Al a Gl n Val Ser Al a Asn Gl u Val
115 120

eol f-seql . txt

<210> 51
<211> 127
<212> PRT
<213> *Latrodectus hesperus*

<400> 51

Gln Ala Asn Thr Pro Trp Ser Ser Lys Glu Asn Ala Asp Ala Phe Ile
1 5 10 15

Gly Ala Phe Met Asn Ala Ala Ser Gln Ser Gly Ala Phe Ser Ser Asp
20 25 30

Gln Ile Asp Asp Met Ser Val Ile Ser Asn Thr Leu Met Ala Ala Met
35 40 45

Asp Asn Met Gly Gly Arg Ile Thr Gln Ser Lys Leu Gln Ala Leu Asp
50 55 60

Met Ala Phe Ala Ser Ser Val Ala Glu Ile Ala Val Ala Asp Gly Gln
65 70 75 80

Asn Val Gly Ala Ala Thr Asn Ala Ile Ser Asp Ala Leu Arg Ser Ala
85 90 95

Phe Tyr Gln Thr Thr Gly Val Val Asn Asn Gln Phe Ile Thr Gly Ile
100 105 110

Ser Ser Leu Ile Gly Met Phe Ala Gln Val Ser Gly Asn Glu Val
115 120 125

<210> 52
<211> 131
<212> PRT
<213> *Nephila iaurata madagascariensis*

<400> 52

Gln Ala Asn Thr Pro Trp Ser Asp Thr Ala Thr Ala Asp Ala Phe Ile
1 5 10 15

Gln Asn Phe Leu Gly Ala Val Ser Gly Ser Gly Ala Phe Thr Pro Asp
20 25 30

Gln Leu Asp Asp Met Ser Thr Val Gly Asp Thr Ile Met Ser Ala Met
35 40 45

Asp Lys Met Ala Arg Ser Asn Lys Ser Ser Lys Ser Lys Leu Gln Ala
50 55 60

Leu Asn Met Ala Phe Ala Ser Ser Met Ala Glu Ile Ala Ala Val Glu
65 70 75 80

Gln Gly Gly Gln Ser Met Asp Val Lys Thr Asn Ala Ile Ala Asn Ala
85 90 95

eol f-seql . txt

Leu Asp Ser Al a Phe Tyr Met Thr Thr Gly Ser Thr Asn Gl n Gl n Phe
100 105 110

Val Asn Gl u Met Arg Ser Leu Ile Asn Met Leu Ser Al a Al a Al a Val
115 120 125

Asn Gl u Val
130

<210> 53
<211> 131
<212> PRT
<213> Nephi la clavi pes

<400> 53

Gl n Al a Arg Ser Pro Trp Ser Asp Thr Al a Thr Al a Asp Al a Phe Ile
1 5 10 15

Gl n Asn Phe Leu Al a Al a Val Ser Gly Ser Gly Al a Phe Thr Ser Asp
20 25 30

Gl n Leu Asp Asp Met Ser Thr Ile Gly Asp Thr Ile Met Ser Al a Met
35 40 45

Asp Lys Met Al a Arg Ser Asn Lys Ser Ser Gl n His Lys Leu Gl n Al a
50 55 60

Leu Asn Met Al a Phe Al a Ser Ser Met Al a Gl u Ile Al a Al a Val Gl u
65 70 75 80

Gl n Gl y Gl y Met Ser Met Al a Val Lys Thr Asn Al a Ile Val Asp Gl y
85 90 95

Leu Asn Ser Al a Phe Tyr Met Thr Thr Gly Al a Al a Asn Pro Gl n Phe
100 105 110

Val Asn Gl u Met Arg Ser Leu Ile Ser Met Ile Ser Al a Al a Ser Al a
115 120 125

Asn Gl u Val
130

<210> 54
<211> 129
<212> PRT
<213> Argi ope bruenni chi

<400> 54

Al a Val Pro Ser Val Phe Ser Ser Pro Asn Leu Al a Ser Gl y Phe Leu
1 5 10 15

eof f-seql.txt

Gln Cys Leu Thr Phe Gly Ile Gly Asn Ser Pro Ala Phe Pro Thr Gln
20 25 30

Gl u Gl n Gl n Asp Leu Asp Ala Ile Ala Gl n Val Ile Leu Asn Ala Val
35 40 45

Ser Ser Asn Thr Gly Ala Thr Ala Ser Ala Arg Ala Gl n Ala Leu Ser
50 55 60

Thr Ala Leu Ala Ser Ser Leu Thr Asp Leu Leu Ile Ala Gl u Ser Ala
65 70 75 80

Gl u Ser Asn Tyr Ser Asn Gl n Leu Ser Gl u Leu Thr Gly Ile Leu Ser
85 90 95

Asp Cys Phe Ile Gl n Thr Thr Gly Ser Asp Asn Pro Ala Phe Val Ser
100 105 110

Arg Ile Gl n Ser Leu Ile Ser Val Leu Ser Gl n Asn Ala Asp Thr Asn
115 120 125

Ile

<210> 55

<211> 129

<212> PRT

<213> Nephi la clavata

<400> 55

Pro Val Pro Ser Val Phe Ser Ser Pro Ser Leu Ala Ser Gl y Phe Leu
1 5 10 15

Gl y Cys Leu Thr Thr Gl y Ile Gl y Leu Ser Pro Ala Phe Pro Phe Gl n
20 25 30

Gl u Gl n Gl n Asp Leu Asp Asp Leu Ala Lys Val Ile Leu Ser Ala Val
35 40 45

Thr Ser Asn Thr Asp Thr Ser Lys Ser Ala Arg Ala Gl n Ala Leu Ser
50 55 60

Thr Ala Leu Ala Ser Ser Leu Ala Asp Leu Leu Ile Ser Gl u Ser Ser
65 70 75 80

Gl y Ser Ser Tyr Gl n Thr Gl n Ile Ser Ala Leu Thr Asn Ile Leu Ser
85 90 95

Asp Cys Phe Val Thr Thr Thr Gl y Ser Asn Asn Pro Ala Phe Val Ser
100 105 110

Arg Val Gl n Thr Leu Ile Gl y Val Leu Ser Gl n Ser Ser Ser Asn Ala

115 120 eol f-seql . txt 125

Ile

<210> 56
<211> 128
<212> PRT
<213> Latrodectus hesperus

<400> 56

Ala Ser Val Asn Ile Phe Asn Ser Pro Asn Ala Ala Thr Ser Phe Leu
1 5 10 15

Asn Cys Leu Arg Ser Asn Ile Glu Ser Ser Pro Ala Phe Pro Phe Glu
20 25 30

Gl u Gl n Ala Asp Leu Asp Ser Ile Ala Gl u Val Ile Leu Ser Asp Val
35 40 45

Ser Ser Val Asn Thr Ala Ser Ser Ala Thr Ser Leu Ala Leu Ser Thr
50 55 60

Ala Leu Ala Ser Ser Leu Ala Gl u Leu Leu Val Thr Gl u Ser Ala Gl u
65 70 75 80

Gl u Asp Ile Asp Asn Glu Val Val Ala Leu Ser Thr Ile Leu Ser Glu
85 90 95

Cys Phe Val Glu Thr Thr Gly Ser Pro Asn Pro Ala Phe Val Ala Ser
100 105 110

Val Lys Ser Leu Leu Glu Val Leu Ser Glu Ser Ala Ser Asn Tyr Glu
115 120 125

<210> 57
<211> 130
<212> PRT
<213> Nephi la clavi pes

<400> 57

Ile Ala Asn Ser Pro Phe Ser Asn Pro Asn Thr Ala Glu Ala Phe Ala
1 5 10 15

Arg Ser Phe Val Ser Asn Ile Val Ser Ser Gly Glu Phe Glu Ala Glu
20 25 30

Gly Ala Glu Asp Phe Asp Asp Ile Ile Glu Ser Leu Ile Glu Ala Glu
35 40 45

Ser Met Gly Lys Gly Arg His Asp Thr Lys Ala Lys Ala Lys Ala Met
50 55 60

eol f-seql . txt

Gl n Val Al a Leu Al a Ser Ser Ile Al a Gl u Leu Val Ile Al a Gl u Ser
65 70 75 80

Ser Gl y Gl y Asp Val Gl n Arg Lys Thr Asn Val Ile Ser Asn Al a Leu
85 90 95

Arg Asn Al a Leu Met Ser Thr Thr Gl y Ser Pro Asn Gl u Gl u Phe Val
100 105 110

Hi s Gl u Val Gl n Asp Leu Ile Gl n Met Leu Ser Gl n Gl u Gl n Ile Asn
115 120 125

Gl u Val
130

<210> 58

<211> 130

<212> PRT

<213> Nephi la i naurata madagascari ensi s

<400> 58

Ile Val Asn Ser Pro Phe Ser Asn Pro Asn Thr Al a Gl u Al a Phe Al a
1 5 10 15

Arg Ser Phe Val Ser Asn Val Val Ser Ser Gl y Gl u Phe Gl y Al a Gl n
20 25 30

Gl y Al a Gl u Asp Phe Asp Asp Ile Ile Gl n Ser Leu Ile Gl n Al a Gl n
35 40 45

Ser Met Gl y Lys Gl y Arg Hi s Asp Thr Lys Al a Lys Al a Lys Al a Met
50 55 60

Gl n Val Al a Leu Al a Ser Ser Ile Al a Gl u Leu Val Ile Al a Gl u Ser
65 70 75 80

Ser Gl y Gl y Asp Val Gl n Arg Lys Thr Asn Val Ile Ser Asn Al a Leu
85 90 95

Arg Asn Al a Leu Met Ser Thr Thr Gl y Ser Pro Asn Gl u Gl u Phe Val
100 105 110

Hi s Gl u Val Gl n Asp Leu Ile Gl n Met Leu Ser Gl n Gl u Gl n Ile Asn
115 120 125

Gl u Val
130

<210> 59

<211> 283

<212> PRT

<213> Euprosthenops austral i s

eol f-seql . txt

<400> 59

His His His His His Met Ser His Thr Thr Pro Trp Thr Asn Pro
1 5 10 15

Gly Leu Ala Glu Asn Phe Met Asn Ser Phe Met Gln Gly Leu Ser Ser
20 25 30

Met Pro Gly Phe Thr Ala Ser Gln Leu Asp Asp Met Ser Thr Ile Ala
35 40 45

Gln Ser Met Val Gln Ser Ile Gln Ser Leu Ala Ala Gln Gly Arg Thr
50 55 60

Ser Pro Asn Lys Leu Gln Ala Leu Asn Met Ala Phe Ala Ser Ser Met
65 70 75 80

Ala Glu Ile Ala Ala Ser Glu Glu Gly Gly Ser Leu Ser Thr Lys
85 90 95

Thr Ser Ser Ile Ala Ser Ala Met Ser Asn Ala Phe Leu Gln Thr Thr
100 105 110

Gly Val Val Asn Gln Pro Phe Ile Asn Glu Ile Thr Gln Leu Val Ser
115 120 125

Met Phe Ala Gln Ala Gly Met Asn Asp Val Ser Ala Ser Ala Ser Ala
130 135 140

Gly Ala Ser Ala Ala Ala Ser Ala Gly Ala Gly Ser Gly Asn Ser Gly
145 150 155 160

Ile Gln Gly Tyr Gly Gln Ser Ser Ala Ser Ala Ser Ala Ala Ala Ser
165 170 175

Ala Ala Ser Thr Val Ala Asn Ser Val Ser Arg Leu Ser Ser Pro Ser
180 185 190

Ala Val Ser Arg Val Ser Ser Ala Val Ser Ser Leu Val Ser Asn Gly
195 200 205

Gln Val Asn Met Ala Ala Leu Pro Asn Ile Ile Ser Asn Ile Ser Ser
210 215 220

Ser Val Ser Ala Ser Ala Pro Gly Ala Ser Gly Cys Glu Val Ile Val
225 230 235 240

Gln Ala Leu Leu Glu Val Ile Thr Ala Leu Val Gln Ile Val Ser Ser
245 250 255

Ser Ser Val Gly Tyr Ile Asn Pro Ser Ala Val Asn Gln Ile Thr Asn
Page 45

260

eol f-seql . txt
265

270

Val Val Al a Asn Al a Met Al a Gl n Val Met Gl y
275 280

<210> 60
<211> 425
<212> PRT
<213> Euprosthenops australis

<400> 60

Met Gl y His His His His His Gl y Gl y Gl y Ser Gl y Gl y Gl y
1 5 10 15

Gl y Ser His His His His His Met Ser His Thr Thr Pro Trp Thr
20 25 30

Asn Pro Gl y Leu Al a Gl u Asn Phe Met Asn Ser Phe Met Gl n Gl y Leu
35 40 45

Ser Ser Met Pro Gl y Phe Thr Al a Ser Gl n Leu Asp Asp Met Ser Thr
50 55 60

Ile Al a Gl n Ser Met Val Gl n Ser Ile Gl n Ser Leu Al a Al a Gl n Gl y
65 70 75 80

Arg Thr Ser Pro Asn Lys Leu Gl n Al a Leu Asn Met Al a Phe Al a Ser
85 90 95

Ser Met Al a Gl u Ile Al a Al a Ser Gl u Gl u Gl y Gl y Ser Leu Ser
100 105 110

Thr Lys Thr Ser Ser Ile Al a Ser Al a Met Ser Asn Al a Phe Leu Gl n
115 120 125

Thr Thr Gl y Val Val Asn Gl n Pro Phe Ile Asn Gl u Ile Thr Gl n Leu
130 135 140

Val Ser Met Phe Al a Gl n Al a Gl y Met Asn Asp Gl y Gl y Gl y Thr Pro
145 150 155 160

Trp Thr Asn Pro Gl y Leu Al a Gl u Asn Phe Met Asn Ser Phe Met Gl n
165 170 175

Gl y Leu Ser Ser Met Pro Gl y Phe Thr Al a Ser Gl n Leu Asp Asp Met
180 185 190

Ser Thr Ile Al a Gl n Ser Met Val Gl n Ser Ile Gl n Ser Leu Al a Al a
195 200 205

Gl n Gl y Arg Thr Ser Pro Asn Lys Leu Gl n Al a Leu Asn Met Al a Phe
210 215 220

eol f-seql . txt

Ala Ser Ser Met Ala Glu Ile Ala Ala Ser Glu Glu Gly Gly Gly Ser
225 230 235 240

Leu Ser Thr Lys Thr Ser Ser Ile Ala Ser Ala Met Ser Asn Ala Phe
245 250 255

Leu Gln Thr Thr Gly Val Val Asn Gln Pro Phe Ile Asn Glu Ile Thr
260 265 270

Gln Leu Val Ser Met Phe Ala Gln Ala Gly Met Asn Asp Val Ser Ala
275 280 285

Leu Glu Ala Leu Phe Gln Gly Pro Asn Ser Gly Asn Ser Gly Ile Gln
290 295 300

Gly Tyr Gly Gln Ser Ser Ala Ser Ala Ser Ala Ala Ala Ser Ala Ala
305 310 315 320

Ser Thr Val Ala Asn Ser Val Ser Arg Leu Ser Ser Pro Ser Ala Val
325 330 335

Ser Arg Val Ser Ser Ala Val Ser Ser Leu Val Ser Asn Gly Gln Val
340 345 350

Asn Met Ala Ala Leu Pro Asn Ile Ile Ser Asn Ile Ser Ser Ser Val
355 360 365

Ser Ala Ser Ala Pro Gly Ala Ser Gly Cys Glu Val Ile Val Gln Ala
370 375 380

Leu Leu Glu Val Ile Thr Ala Leu Val Gln Ile Val Ser Ser Ser Ser
385 390 395 400

Val Gly Tyr Ile Asn Pro Ser Ala Val Asn Gln Ile Thr Asn Val Val
405 410 415

Ala Asn Ala Met Ala Gln Val Met Gly
420 425

<210> 61

<211> 206

<212> PRT

<213> Artificial Sequence

<220>

<223> Fusion protein

<400> 61

Met Gly Ser Ser Gly His His His His His His Met Val Asp Asn Lys
1 5 10 15

eof f-seql.txt

Phe Asn Lys Glu Gln Gln Asn Ala Phe Tyr Glu Ile Leu His Leu Pro
20 25 30

Asn Leu Asn Glu Glu Gln Arg Asn Ala Phe Ile Gln Ser Leu Lys Asp
35 40 45

Asp Pro Ser Gln Ser Ala Asn Leu Leu Ala Glu Ala Lys Lys Leu Asn
50 55 60

Asp Ala Gln Ala Pro Lys Leu Glu Ala Leu Phe Gln Gly Pro Ser Gly
65 70 75 80

Asn Ser Gly Ile Gln Gly Tyr Gly Gln Ser Ser Ala Ser Ala Ser Ala
85 90 95

Ala Ala Ser Ala Ala Ser Thr Val Ala Asn Ser Val Ser Arg Leu Ser
100 105 110

Ser Pro Ser Ala Val Ser Arg Val Ser Ser Ala Val Ser Ser Leu Val
115 120 125

Ser Asn Gly Gln Val Asn Met Ala Ala Leu Pro Asn Ile Ile Ser Asn
130 135 140

Ile Ser Ser Ser Val Ser Ala Ser Ala Pro Gly Ala Ser Gly Cys Glu
145 150 155 160

Val Ile Val Gln Ala Leu Leu Glu Val Ile Thr Ala Leu Val Gln Ile
165 170 175

Val Ser Ser Ser Val Gly Tyr Ile Asn Pro Ser Ala Val Asn Gln
180 185 190

Ile Thr Asn Val Val Ala Asn Ala Met Ala Gln Val Met Gly
195 200 205

<210> 62

<211> 206

<212> PRT

<213> Artificial Sequence

<220>

<223> Fusion protein

<400> 62

Met Gly Ser Ser Gly His His His His His Met Ser Gly Asn Ser
1 5 10 15

Gly Ile Gln Gly Tyr Gly Gln Ser Ser Ala Ser Ala Ser Ala Ala Ala
20 25 30

Ser Ala Ala Ser Thr Val Ala Asn Ser Val Ser Arg Leu Ser Ser Pro
35 40 45

eol f-seql . txt

Ser Ala Val Ser Arg Val Ser Ser Ala Val Ser Ser Leu Val Ser Asn
50 55 60

Gly Glu Val Asn Met Ala Ala Leu Pro Asn Ile Ile Ser Asn Ile Ser
65 70 75 80

Ser Ser Val Ser Ala Ser Ala Pro Gly Ala Ser Gly Cys Glu Val Ile
85 90 95

Val Glu Ala Leu Leu Glu Val Ile Thr Ala Leu Val Glu Ile Val Ser
100 105 110

Ser Ser Ser Val Gly Tyr Ile Asn Pro Ser Ala Val Asn Glu Ile Thr
115 120 125

Asn Val Val Ala Asn Ala Met Ala Glu Val Met Gly Leu Glu Ala Leu
130 135 140

Phe Glu Gly Pro Val Asp Asn Lys Phe Asn Lys Glu Glu Glu Asn Ala
145 150 155 160

Phe Tyr Glu Ile Leu His Leu Pro Asn Leu Asn Glu Glu Glu Arg Asn
165 170 175

Ala Phe Ile Glu Ser Leu Lys Asp Asp Pro Ser Glu Ser Ala Asn Leu
180 185 190

Leu Ala Glu Ala Lys Lys Leu Asn Asp Ala Glu Ala Pro Lys
195 200 205

<210> 63

<211> 354

<212> PRT

<213> Artificial Sequence

<220>

<223> Fusion protein

<400> 63

Met Gly Ser Ser Gly His His His His His Met Val Asp Asn Lys
1 5 10 15

Phe Asn Lys Glu Glu Glu Asn Ala Phe Tyr Glu Ile Leu His Leu Pro
20 25 30

Asn Leu Asn Glu Glu Glu Arg Asn Ala Phe Ile Glu Ser Leu Lys Asp
35 40 45

Asp Pro Ser Glu Ser Ala Asn Leu Leu Ala Glu Ala Lys Lys Leu Asn
50 55 60

eof f-seql.txt

Asp Ala Gln Ala Pro Lys Leu Glu Ala Leu Phe Gln Gly Pro Ser His
65 70 75 80

Thr Thr Pro Trp Thr Asn Pro Gly Leu Ala Glu Asn Phe Met Asn Ser
85 90 95

Phe Met Gln Gly Leu Ser Ser Met Pro Gly Phe Thr Ala Ser Gln Leu
100 105 110

Asp Asp Met Ser Thr Ile Ala Gln Ser Met Val Gln Ser Ile Gln Ser
115 120 125

Leu Ala Ala Gln Gly Arg Thr Ser Pro Asn Lys Leu Gln Ala Leu Asn
130 135 140

Met Ala Phe Ala Ser Ser Met Ala Glu Ile Ala Ala Ser Glu Glu Gly
145 150 155 160

Gly Gly Ser Leu Ser Thr Lys Thr Ser Ser Ile Ala Ser Ala Met Ser
165 170 175

Asn Ala Phe Leu Gln Thr Thr Gly Val Val Asn Gln Pro Phe Ile Asn
180 185 190

Gl u Ile Thr Gln Leu Val Ser Met Phe Ala Gln Ala Gly Met Asn Asp
195 200 205

Val Ser Ala Ser Ala Ser Ala Gly Ala Ser Ala Ala Ala Ser Ala Gly
210 215 220

Ala Gly Ser Gly Asn Ser Gly Ile Gln Gly Tyr Gly Gln Ser Ser Ala
225 230 235 240

Ser Ala Ser Ala Ala Ala Ser Ala Ala Ser Thr Val Ala Asn Ser Val
245 250 255

Ser Arg Leu Ser Ser Pro Ser Ala Val Ser Arg Val Ser Ser Ala Val
260 265 270

Ser Ser Leu Val Ser Asn Gly Gln Val Asn Met Ala Ala Leu Pro Asn
275 280 285

Ile Ile Ser Asn Ile Ser Ser Ser Val Ser Ala Ser Ala Pro Gly Ala
290 295 300

Ser Gly Cys Glu Val Ile Val Gln Ala Leu Leu Glu Val Ile Thr Ala
305 310 315 320

Leu Val Gln Ile Val Ser Ser Ser Ser Val Gly Tyr Ile Asn Pro Ser
325 330 335

Ala Val Asn Gln Ile Thr Asn Val Val Ala Asn Ala Met Ala Gln Val
 340 345 350

Met Gly

<210> 64
 <211> 348
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Fusion protein

<400> 64

His His His His His Met Ser His Thr Thr Pro Trp Thr Asn Pro
 1 5 10 15

Gly Leu Ala Glu Asn Phe Met Asn Ser Phe Met Gln Gly Leu Ser Ser
 20 25 30

Met Pro Gly Phe Thr Ala Ser Gln Leu Asp Asp Met Ser Thr Ile Ala
 35 40 45

Gln Ser Met Val Gln Ser Ile Gln Ser Leu Ala Ala Gln Gly Arg Thr
 50 55 60

Ser Pro Asn Lys Leu Gln Ala Leu Asn Met Ala Phe Ala Ser Ser Met
 65 70 75 80

Ala Glu Ile Ala Ala Ser Glu Glu Gly Gly Ser Leu Ser Thr Lys
 85 90 95

Thr Ser Ser Ile Ala Ser Ala Met Ser Asn Ala Phe Leu Gln Thr Thr
 100 105 110

Gly Val Val Asn Gln Pro Phe Ile Asn Glu Ile Thr Gln Leu Val Ser
 115 120 125

Met Phe Ala Gln Ala Gly Met Asn Asp Val Ser Ala Ser Ala Ser Ala
 130 135 140

Gly Ala Ser Ala Ala Ala Ser Ala Gly Ala Ser Gly Asn Ser Gly Ile
 145 150 155 160

Gln Gly Tyr Gly Gln Ser Ser Ala Ser Ala Ser Ala Ala Ala Ser Ala
 165 170 175

Ala Ser Thr Val Ala Asn Ser Val Ser Arg Leu Ser Ser Pro Ser Ala
 180 185 190

Val Ser Arg Val Ser Ser Ala Val Ser Ser Leu Val Ser Asn Gly Gln
 195 200 205

eol f-seql . txt

Val Asn Met Al a Al a Leu Pro Asn Ile Ile Ser Asn Ile Ser Ser Ser
210 215 220

Val Ser Al a Ser Al a Pro Gly Al a Ser Gly Cys Gl u Val Ile Val Gl n
225 230 235 240

Al a Leu Leu Gl u Val Ile Thr Al a Leu Val Gl n Ile Val Ser Ser Ser
245 250 255

Ser Val Gly Tyr Ile Asn Pro Ser Al a Val Asn Gl n Ile Thr Asn Val
260 265 270

Val Al a Asn Al a Met Al a Gl n Val Met Gl y Leu Gl u Al a Leu Phe Gl n
275 280 285

Gl y Pro Val Asp Asn Lys Phe Asn Lys Gl u Gl n Gl n Asn Al a Phe Tyr
290 295 300

Gl u Ile Leu His Leu Pro Asn Leu Asn Gl u Gl u Gl n Arg Asn Al a Phe
305 310 315 320

Ile Gl n Ser Leu Lys Asp Asp Pro Ser Gl n Ser Al a Asn Leu Leu Al a
325 330 335

Gl u Al a Lys Lys Leu Asn Asp Al a Gl n Al a Pro Lys
340 345

<210> 65
<211> 469
<212> PRT
<213> Artificial Sequence

<220>
<223> Fusion protein

<400> 65

Met Gl y Ser Ser Gl y His His His His His His Met Val Asp Asn Lys
1 5 10 15

Phe Asn Lys Gl u Gl n Gl n Asn Al a Phe Tyr Gl u Ile Leu His Leu Pro
20 25 30

Asn Leu Asn Gl u Gl u Gl n Arg Asn Al a Phe Ile Gl n Ser Leu Lys Asp
35 40 45

Asp Pro Ser Gl n Ser Al a Asn Leu Leu Al a Gl u Al a Lys Lys Leu Asn
50 55 60

Asp Al a Gl n Al a Pro Lys Leu Gl u Al a Leu Phe Gl n Gl y Pro Ser His
65 70 75 80

eof f-seql.txt

Thr Thr Pro Trp Thr Asn Pro Gly Leu Ala Glu Asn Phe Met Asn Ser
85 90 95

Phe Met Gln Gly Leu Ser Ser Met Pro Gly Phe Thr Ala Ser Gln Leu
100 105 110

Asp Asp Met Ser Thr Ile Ala Gln Ser Met Val Gln Ser Ile Gln Ser
115 120 125

Leu Ala Ala Gln Gly Arg Thr Ser Pro Asn Lys Leu Gln Ala Leu Asn
130 135 140

Met Ala Phe Ala Ser Ser Met Ala Glu Ile Ala Ala Ser Glu Glu Gly
145 150 155 160

Gly Gly Ser Leu Ser Thr Lys Thr Ser Ser Ile Ala Ser Ala Met Ser
165 170 175

Asn Ala Phe Leu Gln Thr Thr Gly Val Val Asn Gln Pro Phe Ile Asn
180 185 190

Gl u Ile Thr Gln Leu Val Ser Met Phe Ala Gln Ala Gly Met Asn Asp
195 200 205

Gly Gly Gly Thr Pro Trp Thr Asn Pro Gly Leu Ala Glu Asn Phe Met
210 215 220

Asn Ser Phe Met Gln Gly Leu Ser Ser Met Pro Gly Phe Thr Ala Ser
225 230 235 240

Gln Leu Asp Asp Met Ser Thr Ile Ala Gln Ser Met Val Gln Ser Ile
245 250 255

Gln Ser Leu Ala Ala Gln Gly Arg Thr Ser Pro Asn Lys Leu Gln Ala
260 265 270

Leu Asn Met Ala Phe Ala Ser Ser Met Ala Glu Ile Ala Ala Ser Glu
275 280 285

Gl u Gly Gly Ser Leu Ser Thr Lys Thr Ser Ser Ile Ala Ser Ala
290 295 300

Met Ser Asn Ala Phe Leu Gln Thr Thr Gly Val Val Asn Gln Pro Phe
305 310 315 320

Ile Asn Glu Ile Thr Gln Leu Val Ser Met Phe Ala Gln Ala Gly Met
325 330 335

Asn Asp Val Ser Gly Ser Gly Asn Ser Gly Ile Gln Gly Tyr Gly Gln
340 345 350

eol f-seql . txt

Ser Ser Ala Ser Ala Ser Ala Ala Ser Ala Ser Ala Ser Thr Val Ala
355 360 365

Asn Ser Val Ser Arg Leu Ser Ser Pro Ser Ala Val Ser Arg Val Ser
370 375 380

Ser Ala Val Ser Ser Leu Val Ser Asn Gly Gln Val Asn Met Ala Ala
385 390 395 400

Leu Pro Asn Ile Ile Ser Asn Ile Ser Ser Val Ser Ala Ser Ala
405 410 415

Pro Gly Ala Ser Gly Cys Glu Val Ile Val Gln Ala Leu Leu Glu Val
420 425 430

Ile Thr Ala Leu Val Gln Ile Val Ser Ser Ser Ser Val Gly Tyr Ile
435 440 445

Asn Pro Ser Ala Val Asn Gln Ile Thr Asn Val Val Ala Asn Ala Met
450 455 460

Ala Gln Val Met Gly
465

<210> 66

<211> 483

<212> PRT

<213> Artificial Sequence

<220>

<223> Fusion protein

<400> 66

Met Gly His His His His His Gly Gly Gly Gly Ser Gly Gly Gly
1 5 10 15

Gly Ser His His His His His Met Ser His Thr Thr Pro Trp Thr
20 25 30

Asn Pro Gly Leu Ala Glu Asn Phe Met Asn Ser Phe Met Gln Gly Leu
35 40 45

Ser Ser Met Pro Gly Phe Thr Ala Ser Gln Leu Asp Asp Met Ser Thr
50 55 60

Ile Ala Gln Ser Met Val Gln Ser Ile Gln Ser Leu Ala Ala Gln Gly
65 70 75 80

Arg Thr Ser Pro Asn Lys Leu Gln Ala Leu Asn Met Ala Phe Ala Ser
85 90 95

Ser Met Ala Glu Ile Ala Ala Ser Glu Glu Gly Gly Ser Leu Ser
100 105 110

eol f-seql . txt

Thr Lys Thr Ser Ser Ile Ala Ser Ala Met Ser Asn Ala Phe Leu Glu
115 120 125

Thr Thr Gly Val Val Asn Glu Pro Phe Ile Asn Glu Ile Thr Glu Leu
130 135 140

Val Ser Met Phe Ala Glu Ala Gly Met Asn Asp Gly Gly Gly Thr Pro
145 150 155 160

Trp Thr Asn Pro Gly Leu Ala Glu Asn Phe Met Asn Ser Phe Met Glu
165 170 175

Gly Leu Ser Ser Met Pro Gly Phe Thr Ala Ser Glu Leu Asp Asp Met
180 185 190

Ser Thr Ile Ala Glu Ser Met Val Glu Ser Ile Glu Ser Leu Ala Ala
195 200 205

Glu Glu Arg Thr Ser Pro Asn Lys Leu Glu Ala Leu Asn Met Ala Phe
210 215 220

Ala Ser Ser Met Ala Glu Ile Ala Ala Ser Glu Glu Gly Gly Ser
225 230 235 240

Leu Ser Thr Lys Thr Ser Ser Ile Ala Ser Ala Met Ser Asn Ala Phe
245 250 255

Leu Glu Thr Thr Gly Val Val Asn Glu Pro Phe Ile Asn Glu Ile Thr
260 265 270

Glu Leu Val Ser Met Phe Ala Glu Ala Gly Met Asn Asp Val Ser Ala
275 280 285

Leu Glu Ala Leu Phe Glu Glu Pro Asn Ser Glu Asn Ser Gly Ile Glu
290 295 300

Gly Tyr Gly Glu Ser Ser Ala Ser Ala Ser Ala Ala Ala Ser Ala Ala
305 310 315 320

Ser Thr Val Ala Asn Ser Val Ser Arg Leu Ser Ser Pro Ser Ala Val
325 330 335

Ser Arg Val Ser Ser Ala Val Ser Ser Leu Val Ser Asn Glu Glu Val
340 345 350

Asn Met Ala Ala Leu Pro Asn Ile Ile Ser Asn Ile Ser Ser Ser Val
355 360 365

Ser Ala Ser Ala Pro Gly Ala Ser Gly Cys Glu Val Ile Val Glu Ala
370 375 380

eol f-seql . txt

Leu Leu Glu Val Ile Thr Ala Leu Val Glu Ile Val Ser Ser Ser Ser
385 390 395 400

Val Glu Tyr Ile Asn Pro Ser Ala Val Asn Glu Ile Thr Asn Val Val
405 410 415

Ala Asn Ala Met Ala Glu Val Met Glu Val Asp Asn Lys Phe Asn Lys
420 425 430

Gl u Gl n Gl n Asn Ala Phe Tyr Gl u Ile Leu His Leu Pro Asn Leu Asn
435 440 445

Gl u Gl u Gl n Arg Asn Ala Phe Ile Glu Ser Leu Lys Asp Asp Pro Ser
450 455 460

Gl n Ser Ala Asn Leu Leu Ala Glu Ala Lys Lys Leu Asn Asp Ala Glu
465 470 475 480

Ala Pro Lys

<210> 67
<211> 963
<212> DNA
<213> Artificial Sequence

<220>
<223> Fusion protein

<220>
<221> misc_feature
<222> (37)..(174)
<223> ABD domain

<400> 67
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aaaactgtt aaggtgtaaa agcactgata gatgaaattt tagctgcatt acctggaaat 180
tcaagccata ccacccgtg gaccaacccg ggcctggcgaaaactttt gaacagcttt 240
atgcaggcc tgagcagcat gccggctt accgcgagcc agctggatga tatgagcacc 300
attgcgcaga gcatggcga gaggattcag agcctggcg cgcaggccg caccagcccg 360
aacaactgc aggcgctgaa catggcgaaa ggcgcgatca tggcgaaat tgcggcgagc 420
gaagaaggcg gcccgcgcct gagcacaaa accagcagca ttgcgagcgc gatgagcaac 480
gcgttctgc agaccacccg cgtggtaac cagccgttta ttaacgaaat tacccagctg 540
gtgagcatgt ttgcgcaggc gggcatgaac gatgtgagcg cggcaattc agggatccaa 600
gtttatggtc agatgtgc ttctgcttca gctgctgcgt cagctgctag tactgttagct 660
aattcggtga gtcgcctctc atgccttcc gcagtatctc gagttcttc agcagttct 720

eol f-seql . txt

agcttggttt caaatggtca agtgaatatg gcagcgttac ctaatatcat ttccaacatt 780
tcttcttctg tcagtgcac tgctcctgggt gcttctggat gtgaggtcat agtcaagct 840
ctactcgaag tcatcactgc tctgttcaa atcgtagtt cttctagtgt tggatataatt 900
aatccatctg ctgtgaacca aattactaat gttgttgcta atgccatggc tcaagtaatg 960
ggc 963

<210> 68
<211> 321
<212> PRT
<213> Artificial Sequence

<220>
<223> Fusion protein

<220>
<221> MI SC_FEATURE
<222> (13)..(58)
<223> ABD domain

<400> 68

Met Gly Ser Ser Gly His His His His His His Met Leu Ala Glu Ala
1 5 10 15

Lys Val Leu Ala Asn Arg Glu Leu Asp Lys Tyr Gly Val Ser Asp Tyr
20 25 30

Tyr Lys Asn Leu Ile Asn Asn Ala Lys Thr Val Glu Gly Val Lys Ala
35 40 45

Leu Ile Asp Glu Ile Leu Ala Ala Leu Pro Gly Asn Ser Ser His Thr
50 55 60

Thr Pro Trp Thr Asn Pro Gly Leu Ala Glu Asn Phe Met Asn Ser Phe
65 70 75 80

Met Gln Gly Leu Ser Ser Met Pro Gly Phe Thr Ala Ser Gln Leu Asp
85 90 95

Asp Met Ser Thr Ile Ala Gln Ser Met Val Gln Ser Ile Gln Ser Leu
100 105 110

Ala Ala Gln Gly Arg Thr Ser Pro Asn Lys Leu Gln Ala Leu Asn Met
115 120 125

Ala Phe Ala Ser Ser Met Ala Glu Ile Ala Ala Ser Glu Glu Gly Gly
130 135 140

Gly Ser Leu Ser Thr Lys Thr Ser Ser Ile Ala Ser Ala Met Ser Asn
145 150 155 160

eof f-seql.txt

Ala Phe Leu Glu Thr Thr Gly Val Val Asn Glu Pro Phe Ile Asn Glu
 165 170 175

Ile Thr Glu Leu Val Ser Met Phe Ala Glu Ala Gly Met Asn Asp Val
 180 185 190

Ser Ala Gly Asn Ser Gly Ile Glu Gly Tyr Gly Glu Ser Ser Ala Ser
 195 200 205

Ala Ser Ala Ala Ala Ser Ala Ala Ser Thr Val Ala Asn Ser Val Ser
 210 215 220

Arg Leu Ser Ser Pro Ser Ala Val Ser Arg Val Ser Ser Ala Val Ser
 225 230 235 240

Ser Leu Val Ser Asn Gly Glu Val Asn Met Ala Ala Leu Pro Asn Ile
 245 250 255

Ile Ser Asn Ile Ser Ser Ser Val Ser Ala Ser Ala Pro Gly Ala Ser
 260 265 270

Gly Cys Glu Val Ile Val Glu Ala Leu Leu Glu Val Ile Thr Ala Leu
 275 280 285

Val Glu Ile Val Ser Ser Ser Val Gly Tyr Ile Asn Pro Ser Ala
 290 295 300

Val Asn Glu Ile Thr Asn Val Val Ala Asn Ala Met Ala Glu Val Met
 305 310 315 320

Gly

<210> 69
 <211> 546
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Fusion protein

<220>
 <221> misc_feature
 <222> (28)..(165)
 <223> ABD domain

<400> 69
 atggggccatc atcatcatca tcatatgtta gctgaagcta aagtcttagc taacagagaa 60
 cttgacaaat atggagtaag tgactattac aagaacctaa tcaacaatgc caaaactgtt
 gaagggttaa aagcactgat agatgaaatt ttagctgcat tacctgggaa ttcaggatc
 caaggttatg gtcagagtag tgcttctgct tcagctgctg cgtcagctgc tagtactgta
 gctaattcgg tgagtcgcct ctcatgcct tccgcagtat ctcgagttc ttcagcagtt
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eol f-seql . txt

tctagcttgg tttcaaatgg tcaagtgaat atggcagcgt tacctaatac catttccaaac 360
atttcttctt ctgtcagtgc atctgctcct ggtgcttctg gatgtgaggt catagtgcaa 420
gctctactcg aagtcatcac tgctcttgc caaatcgtaa gttcttctag tgttggatat 480
attaatccat ctgctgtgaa ccaaattact aatgttggaa ctaatgccat ggctcaagta 540
atggc 546

<210> 70
<211> 182
<212> PRT
<213> Artificial Sequence

<220>
<223> Fusion protein

<220>
<221> MISC_FEATURE
<222> (10)..(55)
<223> ABD domain

<400> 70

Met Gly His His His His His Met Leu Ala Glu Ala Lys Val Leu
1 5 10 15

Ala Asn Arg Glu Leu Asp Lys Tyr Gly Val Ser Asp Tyr Tyr Lys Asn
20 25 30

Leu Ile Asn Asn Ala Lys Thr Val Glu Gly Val Lys Ala Leu Ile Asp
35 40 45

Glu Ile Leu Ala Ala Leu Pro Gly Asn Ser Gly Ile Gln Gly Tyr Gly
50 55 60

Gln Ser Ser Ala Ser Ala Ser Ala Ala Ser Ala Ala Ser Thr Val
65 70 75 80

Ala Asn Ser Val Ser Arg Leu Ser Ser Pro Ser Ala Val Ser Arg Val
85 90 95

Ser Ser Ala Val Ser Ser Leu Val Ser Asn Gly Gln Val Asn Met Ala
100 105 110

Ala Leu Pro Asn Ile Ile Ser Asn Ile Ser Ser Ser Val Ser Ala Ser
115 120 125

Ala Pro Gly Ala Ser Gly Cys Glu Val Ile Val Gln Ala Leu Leu Glu
130 135 140

Val Ile Thr Ala Leu Val Gln Ile Val Ser Ser Ser Ser Val Gly Tyr
145 150 155 160

Ile	Asn	Pro	Ser	Ala	Val	Asn	Gln	Ile	Thr	Asn	Val	Val	Ala	Asn	Ala
165						170							175		

Met	Ala	Gln	Val	Met	Gly
				180	

<210> 71

<211> 1302

<212> DNA

<213> Artificial Sequence

<220>

<223> Fusion protein

<220>

<221> misc_feature

<222> (37)..(513)

<223> M4 domain

<400> 71

atggcagca	cgccatca	tcatcatcat	catatggatc	cgagcaaaga	tagcaaagcg	60
caggtgagcg	cggcgaagc	ggcattacc	ggcacctgg	ataaccagct	ggcagcacc	120
tttattgtga	ccgcggcgc	gatggcg	ctgaccggca	cctatgaaag	cgcggggc	180
aacgcggaaa	gccgtatac	cctgaccggc	cgctatgata	gcgcgcggc	gaccgtggc	240
agcggcaccg	cgctggctg	gchggtggcg	tggaaaaaca	actatcgaa	cgcgcata	300
gcccacc	ggagcggcca	gtatgtggc	ggcgcgaag	cgcgcattaa	cacccagtgg	360
accctgacca	gcggcaccac	cgaagcgaac	gcgtggaaaa	gcaccctgc	cggccatgat	420
acctttacca	aagtgaaacc	gagcgcggcg	agcattgatg	cggcggggcgt	agcgggcgt	480
aacaacggca	acccgctgga	tgcgtgcag	cagggaaatt	caagccatac	cacccgtgg	540
accaacccgg	gcctggcgga	aaactttatg	aacagcttta	tgcagggcct	gagcagcat	600
ccggcctta	ccgcgagcca	gctggatgat	atgagcacca	ttgcgcagag	catggtgcag	660
agcattcaga	gcctggcg	gcagggccgc	accagcccga	acaaactgca	ggcgcgtgaac	720
atggcg	cgagcagcat	ggcgaaatt	gcggcgagcg	aagaaggcgg	cgccagcct	780
agcacaaaa	ccagcagcat	tgcgagcg	atgagcaac	cgttctgca	gaccaccggc	840
gtggtgaacc	agccgttat	taacgaaatt	acccagctgg	tgagcatgtt	tgcgcaggcg	900
ggcatgaac	atgtgagcg	ggcaattca	ggatccaag	gttatggtca	gagtagtgct	960
tctgcttcag	ctgctgc	agctgctgt	actgtagct	attcggtgag	tcgcctctca	1020
tcgccttcg	cagtatctcg	agtttctca	gcagttct	gcttggttc	aatggtcaa	1080
gtgaatatgg	cagcgttacc	taatatcatt	tccaaacattt	cttctctgt	cagtgcatt	1140
gctcctgg	cttctggat	tgaggtcata	gtgcaagctc	tactcgaagt	catcactgt	1200
cttggtaaa	tcgttagttc	ttcttagtgtt	ggatatatta	atccatctgc	tgtgaaccaa	1260
attactaatg	ttgttgctaa	tgccatggct	caagtaatgg	gc		1302

eol f-seql . txt

<210> 72
<211> 434
<212> PRT
<213> Artificial Sequence

<220>
<223> Fusion protein

<220>
<221> MI SC_FEATURE
<222> (13)..(171)
<223> M4 domain

<400> 72

Met Gly Ser Ser Gly His His His His His His Met Asp Pro Ser Lys
1 5 10 15

Asp Ser Lys Ala Glu Val Ser Ala Ala Glu Ala Gly Ile Thr Gly Thr
20 25 30

Trp Tyr Asn Glu Leu Gly Ser Thr Phe Ile Val Thr Ala Gly Ala Asp
35 40 45

Gly Ala Leu Thr Gly Thr Tyr Glu Ser Ala Val Gly Asn Ala Glu Ser
50 55 60

Arg Tyr Thr Leu Thr Gly Arg Tyr Asp Ser Ala Pro Ala Thr Asp Gly
65 70 75 80

Ser Gly Thr Ala Leu Gly Trp Arg Val Ala Trp Lys Asn Asn Tyr Arg
85 90 95

Asn Ala His Ser Ala Thr Thr Trp Ser Gly Glu Tyr Val Gly Gly Ala
100 105 110

Glu Ala Arg Ile Asn Thr Glu Trp Thr Leu Thr Ser Gly Thr Thr Glu
115 120 125

Ala Asn Ala Trp Lys Ser Thr Leu Arg Gly His Asp Thr Phe Thr Lys
130 135 140

Val Lys Pro Ser Ala Ala Ser Ile Asp Ala Ala Lys Lys Ala Gly Val
145 150 155 160

Asn Asn Gly Asn Pro Leu Asp Ala Val Glu Glu Gly Asn Ser Ser His
165 170 175

Thr Thr Pro Trp Thr Asn Pro Gly Leu Ala Glu Asn Phe Met Asn Ser
180 185 190

Phe Met Glu Gly Leu Ser Ser Met Pro Gly Phe Thr Ala Ser Glu Leu
195 200 205

eol f-seql . txt

Asp Asp Met Ser Thr Ile Ala Gln Ser Met Val Gln Ser Ile Gln Ser
210 215 220

Leu Ala Ala Gln Gly Arg Thr Ser Pro Asn Lys Leu Gln Ala Leu Asn
225 230 235 240

Met Ala Phe Ala Ser Ser Met Ala Glu Ile Ala Ala Ser Glu Glu Gly
245 250 255

Gly Gly Ser Leu Ser Thr Lys Thr Ser Ser Ile Ala Ser Ala Met Ser
260 265 270

Asn Ala Phe Leu Gln Thr Thr Gly Val Val Asn Gln Pro Phe Ile Asn
275 280 285

Gl u Ile Thr Gln Leu Val Ser Met Phe Ala Gln Ala Gly Met Asn Asp
290 295 300

Val Ser Ala Gly Asn Ser Gly Ile Gln Gly Tyr Gly Gln Ser Ser Ala
305 310 315 320

Ser Ala Ser Ala Ala Ser Ala Ala Ser Thr Val Ala Asn Ser Val
325 330 335

Ser Arg Leu Ser Ser Pro Ser Ala Val Ser Arg Val Ser Ser Ala Val
340 345 350

Ser Ser Leu Val Ser Asn Gly Gln Val Asn Met Ala Ala Leu Pro Asn
355 360 365

Ile Ile Ser Asn Ile Ser Ser Val Ser Ala Ser Ala Pro Gly Ala
370 375 380

Ser Gly Cys Glu Val Ile Val Gln Ala Leu Leu Glu Val Ile Thr Ala
385 390 395 400

Leu Val Gln Ile Val Ser Ser Ser Ser Val Gly Tyr Ile Asn Pro Ser
405 410 415

Ala Val Asn Gln Ile Thr Asn Val Val Ala Asn Ala Met Ala Gln Val
420 425 430

Met Gly

<210> 73
<211> 885
<212> DNA
<213> Artificial Sequence

<220>
<223> Fusion protein

eol f-seql . txt

<220>
 <221> mi sc_feature
 <222> (28)..(504)
 <223> M4 domain

<400> 73
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 gcggcggaaag cggcattac cggcacctgg tataaccagc tggcagcac ctattttgtg 120
 accgcggcgc cggatggcgc gctgaccggc acctatgaaa gcgcggtggg caacgcggaa 180
 agccgctata ccctgaccgg ccgtatgtat agcgcgccgg cgaccgatgg cagcggcacc 240
 ggcgtggct ggcgcgtggc gtggaaaaac aactatgca acgcgcatacg cgcgaccacc 300
 tggagcggcc agtatgtggg cggcggaa gcgcgcatta acacccagtg gaccctgacc 360
 agcggcacca ccgaagcgaa cgcgtggaaa agcaccctgc gcggccatga taccttacc 420
 aaagtgaaac cgagcgcggc gagcattgtat gcggcgaaaa aagcgggcgt gaacaacggc 480
 aacccgctgg atgcggtgca gcagggaaat tcagggatcc aagttatgg tcagagtagt 540
 gcttcgtctt cagctgctgc gtcagctgct agtactgttag ctaattcggt gagtcgcctc 600
 tcatcgccctt ccgcgtatc tcgagttct tcagcgttt ctatcggtt ttcaaattgg 660
 caagtgaata tggcagcggtt acctaatac atttccaaaca tttcttcttc tgtcagtgc 720
 tctgctcctg gtgcttcgtt atgtgaggc atagtgcag ctctactcga agtcatcact 780
 gctcttggc aaatcgtagt ttcttctgt gttggatata ttaatccatc tgctgtgaac 840
 caaattacta atgttgtgc taatgccatg gctcaagtaa tggc 885

<210> 74
 <211> 295
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Fusion protein

<220>
 <221> MI SC_FEATURE
 <222> (10)..(168)
 <223> M4 domain

<400> 74

Met Gly His His His His His Met Asp Pro Ser Lys Asp Ser Lys
 1 5 10 15

Ala Gln Val Ser Ala Ala Glu Ala Gly Ile Thr Gly Thr Trp Tyr Asn
 20 25 30

Gln Leu Gly Ser Thr Phe Ile Val Thr Ala Gly Ala Asp Gly Ala Leu
 35 40 45

Thr Gly Thr Tyr Glu Ser Ala Val Gly Asn Ala Glu Ser Arg Tyr Thr
 50 55 60

eol f-seql . txt

Leu Thr Gly Arg Tyr Asp Ser Ala Pro Ala Thr Asp Gly Ser Gly Thr
65 70 75 80

Ala Leu Gly Trp Arg Val Ala Trp Lys Asn Asn Tyr Arg Asn Ala His
85 90 95

Ser Ala Thr Thr Trp Ser Gly Glu Tyr Val Gly Gly Ala Glu Ala Arg
100 105 110

Ile Asn Thr Glu Trp Thr Leu Thr Ser Gly Thr Thr Glu Ala Asn Ala
115 120 125

Trp Lys Ser Thr Leu Arg Gly His Asp Thr Phe Thr Lys Val Lys Pro
130 135 140

Ser Ala Ala Ser Ile Asp Ala Ala Lys Lys Ala Gly Val Asn Asn Gly
145 150 155 160

Asn Pro Leu Asp Ala Val Glu Glu Gly Asn Ser Gly Ile Glu Gly Tyr
165 170 175

Gly Glu Ser Ser Ala Ser Ala Ser Ala Ala Ser Ala Ala Ser Thr
180 185 190

Val Ala Asn Ser Val Ser Arg Leu Ser Ser Pro Ser Ala Val Ser Arg
195 200 205

Val Ser Ser Ala Val Ser Ser Leu Val Ser Asn Gly Glu Val Asn Met
210 215 220

Ala Ala Leu Pro Asn Ile Ile Ser Asn Ile Ser Ser Ser Val Ser Ala
225 230 235 240

Ser Ala Pro Gly Ala Ser Gly Cys Glu Val Ile Val Glu Ala Leu Leu
245 250 255

Glu Val Ile Thr Ala Leu Val Glu Ile Val Ser Ser Ser Ser Val Gly
260 265 270

Tyr Ile Asn Pro Ser Ala Val Asn Glu Ile Thr Asn Val Val Ala Asn
275 280 285

Ala Met Ala Glu Val Met Gly
290 295

<210> 75
<211> 1614

<212> DNA
<213> Artificial Sequence

<220>

eol f-seql . txt

<223> Fusion protein

<220>
 <221> misc_feature
 <222> (37)..(825)
 <223> scFv1 domain

<400> 75
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 ggaggcttgg tacagcctgg ggggtccctg agactctcct gtgcagccctc tggattcacc 120
 ttcagtagtt atgaaatgaa ctgggtccgc caggctccag ggaaggggct ggagtgggtc 180
 tcaggcatta gtggtagtgg tggttcaca tactacgcag actccgtgaa gggccgattc 240
 accatctcca gagacaattc caagaacacg ctgtatctgc aaatgaacag cctgagagcc 300
 gaggacactg ccatgttatta ctgtgcgaga gaggggtacc aggtgcttt tgatatctgg 360
 ggccagggtta cactggtcac cgtgagcagc ggtggaggcg gttcaggcgg aggtggatcc 420
 ggcgggtggcg gatcgagtc tgtgctgact cagccaccct cagcgtctgg gaccccccggg 480
 cagagggtca ccatctcctg cactgggagc agctccaaca tcggggcagg ttatgtgta 540
 cactggtagtac agcagctccc aggaacggcc cccaaactcc tcatctatag taataatcag 600
 cggccctcag gggccctga ccgattctct ggctccaagt ctggcacccctc agcctccctg 660
 gccatcagtg ggctccggc cgaggatgag gctgattatt actgtgcagc atggatgac 720
 agcctgagtg gtccgccttg ggtttcggc ggaggaacca agctgacggt cctaggtgaa 780
 caaaaactca tctcagaaga ggatctgtct ggatcagcgg ctgcagggaa ttcaagccat 840
 accaccccggt ggaccaaccc gggcctggcg gaaaacttta tgaacagctt tatgcagggc 900
 ctgagcagca tgccgggctt taccgcgagc cagctggatg atatgagcac cattgcgcag 960
 agcatggtagc agagcattca gagcctggcg gcgcaggccc gcaccagccc gaacaaactg 1020
 cagggcgtga acatggcgaa tgcgagcagc atggcgaaa ttgcggcgag cgaagaaggc 1080
 ggcggcagcc tgagcaccaa aaccagcagc attgcgagcg cgatgagcaa cgcgttctg 1140
 cagaccaccc gctgtgtgaa ccagccgtt attaacgaaa ttacccagct ggtgagcatg 1200
 tttgcgcagg cgggcatgaa cgatgtgagc gcgggcaatt cagggatcca agttatggt 1260
 cagagtagtg cttctgcttc agctgctgcg tcagctgcta gtactgtagc taattcgggt 1320
 agtcgcctct catgccttc cgcagtatct cgagttctt cagcagttc tagcttgggt 1380
 tcaaatggtc aagtgaatat ggcagcgtta cctaataatca tttccaacat ttcttcttct 1440
 gtcagtgcatt ctgctcctgg tgcttctgga tgtgaggtca tagtgcagc tctactcgaa 1500
 gtcatcactg ctcttgttca aatcgtagt tcttcttagt ttggatataat taatccatct 1560
 gctgtgaacc aaattactaa tgtgttgct aatgcctatgg ctcaagtaat gggc 1614

<210> 76
 <211> 538
 <212> PRT
 <213> Artificial Sequence

eol f-seql . txt

<220>
<223> Fusion protein

<220>
<221> MISC_FEATURE
<222> (13)..(275)
<223> scFv1 domain

<400> 76

Met Gly Ser Ser Gly His His His His His His Met Glu Val Gln Leu
1 5 10 15

Leu Glu Ser Gly Gly Leu Val Gln Pro Gly Gly Ser Leu Arg Leu
20 25 30

Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser Ser Tyr Glu Met Asn Trp
35 40 45

Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val Ser Gly Ile Ser
50 55 60

Gly Ser Gly Gly Phe Thr Tyr Tyr Ala Asp Ser Val Lys Gly Arg Phe
65 70 75 80

Thr Ile Ser Arg Asp Asn Ser Lys Asn Thr Leu Tyr Leu Gln Met Asn
85 90 95

Ser Leu Arg Ala Glu Asp Thr Ala Met Tyr Tyr Cys Ala Arg Glu Gly
100 105 110

Tyr Gln Asp Ala Phe Asp Ile Trp Gly Gln Gly Thr Leu Val Thr Val
115 120 125

Ser Ser Gly Gly Gly Ser Gly Gly Gly Ser Gly Gly Gly Gly
130 135 140

Ser Gln Ser Val Leu Thr Gln Pro Pro Ser Ala Ser Gly Thr Pro Gly
145 150 155 160

Gln Arg Val Thr Ile Ser Cys Thr Gly Ser Ser Ser Asn Ile Gly Ala
165 170 175

Gly Tyr Asp Val His Trp Tyr Gln Gln Leu Pro Gly Thr Ala Pro Lys
180 185 190

Leu Leu Ile Tyr Ser Asn Asn Gln Arg Pro Ser Gly Val Pro Asp Arg
195 200 205

Phe Ser Gly Ser Lys Ser Gly Thr Ser Ala Ser Leu Ala Ile Ser Gly
210 215 220

eol f-seql . txt

Leu Arg Ser Glu Asp Glu Ala Asp Tyr Tyr Cys Ala Ala Trp Asp Asp
225 230 235 240

Ser Leu Ser Gly Pro Pro Trp Val Phe Gly Gly Gly Thr Lys Leu Thr
245 250 255

Val Leu Gly Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu Ser Gly Ser
260 265 270

Ala Ala Ala Gly Asn Ser Ser His Thr Thr Pro Trp Thr Asn Pro Gly
275 280 285

Leu Ala Glu Asn Phe Met Asn Ser Phe Met Gln Gly Leu Ser Ser Met
290 295 300

Pro Gly Phe Thr Ala Ser Gln Leu Asp Asp Met Ser Thr Ile Ala Gln
305 310 315 320

Ser Met Val Gln Ser Ile Gln Ser Leu Ala Ala Gln Gly Arg Thr Ser
325 330 335

Pro Asn Lys Leu Gln Ala Leu Asn Met Ala Phe Ala Ser Ser Met Ala
340 345 350

Gl u Ile Ala Ala Ser Glu Glu Gly Gly Ser Leu Ser Thr Lys Thr
355 360 365

Ser Ser Ile Ala Ser Ala Met Ser Asn Ala Phe Leu Gln Thr Thr Gly
370 375 380

Val Val Asn Gln Pro Phe Ile Asn Glu Ile Thr Gln Leu Val Ser Met
385 390 395 400

Phe Ala Gln Ala Gly Met Asn Asp Val Ser Ala Gly Asn Ser Gly Ile
405 410 415

Gln Gly Tyr Gly Gln Ser Ser Ala Ser Ala Ser Ala Ala Ala Ser Ala
420 425 430

Ala Ser Thr Val Ala Asn Ser Val Ser Arg Leu Ser Ser Pro Ser Ala
435 440 445

Val Ser Arg Val Ser Ser Ala Val Ser Ser Leu Val Ser Asn Gly Gln
450 455 460

Val Asn Met Ala Ala Leu Pro Asn Ile Ile Ser Asn Ile Ser Ser Ser
465 470 475 480

Val Ser Ala Ser Ala Pro Gly Ala Ser Gly Cys Glu Val Ile Val Gln
485 490 495

eol f-seql . txt

Ala Leu Leu Glu Val Ile Thr Ala Leu Val Gln Ile Val Ser Ser Ser
 500 505 510

Ser Val Gly Tyr Ile Asn Pro Ser Ala Val Asn Gln Ile Thr Asn Val
 515 520 525

Val Ala Asn Ala Met Ala Gln Val Met Gly
 530 535

<210> 77
 <211> 1197
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Fusion protein

<220>
 <221> misc_feature
 <222> (28)..(816)
 <223> scFv1 domain

<400> 77
 atgggccatc atcatcatca tcatatggag gtgcagctgt tggagtctgg gggaggcttg 60
 gtacagcctg ggggtccct gagactctcc tgtgcagcct ctggattcac cttcagtagt 120
 tatgaaatga actgggtccg ccaggctcca gggaaaggggc tggagtgggt ctcaggcatt 180
 agtggtagtg gtggttcac atactacgca gactccgtga agggccgatt caccatctcc 240
 agagacaatt ccaagaacac gctgtatctg caaatgaaca gcctgagagc cgaggacact 300
 gccatgtatt actgtgcgag agaggggtac caggatgctt ttgatatctg gggccagggt 360
 acactggtca ccgtgagcag cggtgaggc gttcaggcg gaggtggatc cggcggtggc 420
 ggatcgcagt ctgtgctgac tcagccaccc tcagcgtctg ggaccccccgg gcagagggtc 480
 accatctcct gcactggag cagctccaaac atcggggcag gttatgatgt acactggtat 540
 cagcagctcc caggaacggc ccccaaactc ctcatctata gtaataatca gcggccctca 600
 ggggtccctg accgattctc tggctccaaag tctggcacct cagcctccct ggccatcagt 660
 gggctccgt ccgaggatga ggctgattat tactgtcag catggatga cagcctgagt 720
 ggtccgcctt ggggttcgg cgaggaacc aagctgacgg tcctaggtga aaaaaactc 780
 atctcagaag aggatctgac tggatcagcg gctgcaggaa attcaggat ccaagttat 840
 ggtcagagta gtgcttcgtc ttcaagtcgt gcgtcagctg ctgtactgt agctaattcg 900
 gtgagtcgcc tctcatcgcc ttccgcagta tctcgatgtt cttcagcagt ttctagcttg 960
 gtttcaaattgtcaagtgtaa tatggcagcg ttacctaata tcatttccaa catttttct 1020
 tctgtcagtg catctgctcc tggtgcttct ggatgtgagg tcatagtgc agctctactc 1080
 gaagtcatca ctgctctgt tcaaattcggtt agttcttcta gtgtggata tattaaatcca 1140
 tctgctgtga accaaattac taatgttgtt gctaatgcca tggctcaagt aatggc 1197

eol f-seql . txt

<210> 78
<211> 399
<212> PRT
<213> Artificial Sequence

<220>
<223> Fusion protein

<220>
<221> MI SC_FEATURE
<222> (10)..(272)
<223> scFv1 domain

<400> 78

Met Gly His His His His His Met Glu Val Gln Leu Leu Glu Ser
1 5 10 15

Gly Gly Gly Leu Val Gln Pro Gly Gly Ser Leu Arg Leu Ser Cys Ala
20 25 30

Ala Ser Gly Phe Thr Phe Ser Ser Tyr Glu Met Asn Trp Val Arg Gln
35 40 45

Ala Pro Gly Lys Gly Leu Glu Trp Val Ser Gly Ile Ser Gly Ser Gly
50 55 60

Gly Phe Thr Tyr Tyr Ala Asp Ser Val Lys Gly Arg Phe Thr Ile Ser
65 70 75 80

Arg Asp Asn Ser Lys Asn Thr Leu Tyr Leu Gln Met Asn Ser Leu Arg
85 90 95

Ala Glu Asp Thr Ala Met Tyr Tyr Cys Ala Arg Glu Gly Tyr Gln Asp
100 105 110

Ala Phe Asp Ile Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser Gly
115 120 125

Gly Gly Gly Ser Gly Gly Ser Gly Gly Gly Ser Gln Ser
130 135 140

Val Leu Thr Gln Pro Pro Ser Ala Ser Gly Thr Pro Gly Gln Arg Val
145 150 155 160

Thr Ile Ser Cys Thr Gly Ser Ser Ser Asn Ile Gly Ala Gly Tyr Asp
165 170 175

Val His Trp Tyr Gln Gln Leu Pro Gly Thr Ala Pro Lys Leu Leu Ile
180 185 190

Tyr Ser Asn Asn Gln Arg Pro Ser Gly Val Pro Asp Arg Phe Ser Gly
195 200 205

eol f-seql.txt

Ser Lys Ser Gly Thr Ser Ala Ser Leu Ala Ile Ser Gly Leu Arg Ser
210 215 220

Glu Asp Glu Ala Asp Tyr Tyr Cys Ala Ala Trp Asp Asp Ser Leu Ser
225 230 235 240

Gly Pro Pro Trp Val Phe Gly Gly Thr Lys Leu Thr Val Leu Gly
245 250 255

Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu Ser Gly Ser Ala Ala Ala
260 265 270

Gly Asn Ser Gly Ile Gln Gly Tyr Gly Gln Ser Ser Ala Ser Ala Ser
275 280 285

Ala Ala Ala Ser Ala Ala Ser Thr Val Ala Asn Ser Val Ser Arg Leu
290 295 300

Ser Ser Pro Ser Ala Val Ser Arg Val Ser Ser Ala Val Ser Ser Leu
305 310 315 320

Val Ser Asn Gly Gln Val Asn Met Ala Ala Leu Pro Asn Ile Ile Ser
325 330 335

Asn Ile Ser Ser Val Ser Ala Ser 345 Ala Pro Gly Ala Ser Gly Cys
340 350

Glu Val Ile Val Gln Ala Leu Leu Glu Val Ile Thr Ala Leu Val Gln
355 360 365

Ile Val Ser Ser Ser Val Gly Tyr Ile Asn Pro Ser Ala Val Asn
370 375 380

Gln Ile Thr Asn Val Val Ala Asn Ala Met Ala Gln Val Met Gly
385 390 395

<210> 79

<211> 1380

<212> DNA

<213> Artificial Sequence

<220>

<223> Fusion protein

<220>

<221> misc_feature

<222> (37)..(591)

<223> xylanase domain

<400> 79

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60

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120

aattggtcta ataccggaaa tttgttgtt ggtaaagggtt ggactacagg ttcgcccattt

180

eol f-seql . txt

aggacgataa	actataatgc	cgaggttgg	gcccgaatg	gcaatggata	tttaacttta	240
tatggttgga	cgagatcacc	tctcatagaa	tattatgtag	tggattcatg	gggtacttat	300
agacctactg	gaacgtataa	aggtaactgta	aaaagtgtatg	ggggtacgta	tgacatatat	360
acaactacac	gttataacgc	accccccatt	gatggcgatc	gcactacttt	tacgcagttac	420
tggagtgttc	gccagtcgaa	gagaccaacc	ggaagcaacg	ctacaatcac	tttcagcaat	480
catgtgaacg	catggaagag	ccatggaatg	aatctggca	gtaattggc	ttaccaagtc	540
atggcgacag	aaggatatac	aagtagtgg	agttctaacg	taacagtgt	gccgaattca	600
agccatacca	ccccgtggac	caacccggc	ctggcgaaa	actttatgaa	cagctttatg	660
cagggcctga	gcagcatgcc	gggtttacc	gcgagccagc	tggatgat	gagcaccatt	720
gcfgcagagca	tggtgcagag	cattcagagc	ctggcggcgc	aggccgcac	cagccgaaac	780
aaactgcagg	cgctgaacat	ggcggttgcg	agcagcatgg	cgaaattgc	ggcgagcgaa	840
gaaggcggcg	gcagcctgag	caccaaaacc	agcagcattg	cgagcgcgat	gagcaacgcg	900
tttctgcaga	ccaccggcgt	ggtgaaccag	ccgttattta	acgaaattac	ccagctggtg	960
agcatgtttg	cgcaggcggg	catgaacgat	gtgagcgcgg	gcaattcagg	gatccaaggt	1020
tatggtcaga	gtagtgcctc	tgcttcagct	gctgcgtcag	ctgcttagtac	tgtagctaat	1080
tcggtgagtc	gcctctcatc	gcctccgca	gtatctcgag	tttcttcagc	agtttctagc	1140
ttgggttcaa	atggtcaagt	gaatatggca	gcgttaccta	atatcatttc	caacatttct	1200
tcttctgtca	gtgcacatgc	tcctggtgct	tctggatgt	aggtcatagt	gcaagctcta	1260
ctcgaagtca	tcactgctct	tgttcaaatac	gttagttctt	ctagtggtgg	atataattaaat	1320
ccatctgctg	tgaaccaaata	tactaatgtt	gttgctaatg	ccatggctca	agtaatgggc	1380

<210> 80
 <211> 460
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Fusion protein

<220>
 <221> MISC_FEATURE
 <222> (13)..(197)
 <223> xylanase domain

<400> 80

Met Gl y Ser Ser Gl y His His His His His His Met Al a Ser Thr Asp
 1 5 10 15

Tyr Trp Gl n Asn Trp Thr Asp Gl y Gl y Gl y Ile Val Asn Al a Val Asn
 20 25 30

Gl y Ser Gl y Gl y Asn Tyr Ser Val Asn Trp Ser Asn Thr Gl y Asn Phe
 35 40 45

eol f-seql . txt

Val Val Gly Lys Gly Trp Thr Thr Gly Ser Pro Phe Arg Thr Ile Asn
50 55 60

Tyr Asn Ala Gly Val Trp Ala Pro Asn Gly Asn Gly Tyr Leu Thr Leu
65 70 75 80

Tyr Gly Trp Thr Arg Ser Pro Leu Ile Glu Tyr Tyr Val Val Asp Ser
85 90 95

Trp Gly Thr Tyr Arg Pro Thr Gly Thr Tyr Lys Gly Thr Val Val Lys Ser
100 105 110

Asp Gly Gly Thr Tyr Asp Ile Tyr Thr Thr Arg Tyr Asn Ala Pro
115 120 125

Ser Ile Asp Gly Asp Arg Thr Thr Phe Thr Glu Tyr Trp Ser Val Arg
130 135 140

Gln Ser Lys Arg Pro Thr Gly Ser Asn Ala Thr Ile Thr Phe Ser Asn
145 150 155 160

His Val Asn Ala Trp Lys Ser His Gly Met Asn Leu Gly Ser Asn Trp
165 170 175

Ala Tyr Gln Val Met Ala Thr Glu Gly Tyr Gln Ser Ser Gly Ser Ser
180 185 190

Asn Val Thr Val Trp Pro Asn Ser Ser His Thr Thr Pro Trp Thr Asn
195 200 205

Pro Gly Leu Ala Glu Asn Phe Met Asn Ser Phe Met Gln Gly Leu Ser
210 215 220

Ser Met Pro Gly Phe Thr Ala Ser Gln Leu Asp Asp Met Ser Thr Ile
225 230 235 240

Ala Gln Ser Met Val Gln Ser Ile Gln Ser Leu Ala Ala Gln Gly Arg
245 250 255

Thr Ser Pro Asn Lys Leu Gln Ala Leu Asn Met Ala Phe Ala Ser Ser
260 265 270

Met Ala Glu Ile Ala Ala Ser Glu Glu Gly Gly Ser Leu Ser Thr
275 280 285

Lys Thr Ser Ser Ile Ala Ser Ala Met Ser Asn Ala Phe Leu Gln Thr
290 295 300

Thr Gly Val Val Asn Gln Pro Phe Ile Asn Glu Ile Thr Gln Leu Val
305 310 315 320

eol f-seql . txt

Ser Met Phe Al a Gl n Al a Gl y Met Asn Asp Val Ser Al a Gl y Asn Ser
325 330 335

Gl y Ile Gl n Gl y Tyr Gl y Gl n Ser Ser Al a Ser Al a Ser Al a Al a Al a
340 345 350

Ser Al a Al a Ser Thr Val Al a Asn Ser Val Ser Arg Leu Ser Ser Pro
355 360 365

Ser Al a Val Ser Arg Val Ser Ser Al a Val Ser Ser Leu Val Ser Asn
370 375 380

Gl y Gl n Val Asn Met Al a Al a Leu Pro Asn Ile Ile Ser Asn Ile Ser
385 390 395 400

Ser Ser Val Ser Al a Ser Al a Pro Gl y Al a Ser Gl y Cys Gl u Val Ile
405 410 415

Val Gl n Al a Leu Leu Gl u Val Ile Thr Al a Leu Val Gl n Ile Val Ser
420 425 430

Ser Ser Ser Val Gl y Tyr Ile Asn Pro Ser Al a Val Asn Gl n Ile Thr
435 440 445

Asn Val Val Al a Asn Al a Met Al a Gl n Val Met Gl y
450 455 460

<210> 81

<211> 963

<212> DNA

<213> Artificial Sequence

<220>

<223> Fusion protein

<220>

<221> misc_feature

<222> (28)..(582)

<223> xylanase domain

<400> 81

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aataccggaa attttgttgt tggtaaaggt tggactacag gttcgccatt taggacgata 180

aactataatg ccggagttt ggcgccaat ggcaatggat atttaacttt atatggttg 240

acgagatcac ctctcataga atattatgta gtggattcat ggggtactta tagacctact 300

ggaacgtata aaggtactgt aaaaagtgtat ggggtacgt atgacatata tacaactaca 360

cgttataacg caccttccat tcatggcgat cgcaactactt ttacgcagta ctggagtgtt 420

cgccagtcga agagaccaac cgaaagcaac gctacaatca ctttcagcaa tcatgtgaac 480

eol f-seql . txt

gcatggaaga	gccatggaat	aatctgggc	agtaattggg	cttaccaagt	catggcgaca	540
gaaggatatac	aaagtagtgg	aagttctaac	gtaacagtgt	ggccgaattc	agggatccaa	600
ggttatggtc	agagtagtgc	ttctgcttca	gctgctgcgt	cagctgctag	tactgttagct	660
aattcggtga	gtcgccctc	atgccttcc	gcagtatctc	gagtttcttc	agcagttct	720
agcttggttt	caaatggtca	agtgaatatg	gcagcgttac	ctaatatcat	ttccaacatt	780
tcttcttctg	tcagtgcac	tgctcctggt	gcttctggat	gtgaggtcat	agtgcagct	840
ctactcgaag	tcatcactgc	tctgttcaa	atcgtagtt	cttctagtgt	tggatataatt	900
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ggc						963

<210> 82
 <211> 321
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Fusion protein

<220>
 <221> MISC_FEATURE
 <222> (10)..(194)
 <223> xylanase domain

<400> 82

Met Gly His His His His His Met Ala Ser Thr Asp Tyr Trp Gln
 1 5 10 15

Asn Trp Thr Asp Gly Gly Ile Val Asn Ala Val Asn Gly Ser Gly
 20 25 30

Gly Asn Tyr Ser Val Asn Trp Ser Asn Thr Gly Asn Phe Val Val Gly
 35 40 45

Lys Gly Trp Thr Thr Gly Ser Pro Phe Arg Thr Ile Asn Tyr Asn Ala
 50 55 60

Gly Val Trp Ala Pro Asn Gly Asn Gly Tyr Leu Thr Leu Tyr Gly Trp
 65 70 75 80

Thr Arg Ser Pro Leu Ile Glu Tyr Tyr Val Val Asp Ser Trp Gly Thr
 85 90 95

Tyr Arg Pro Thr Gly Thr Tyr Lys Gly Thr Val Lys Ser Asp Gly Gly
 100 105 110

Thr Tyr Asp Ile Tyr Thr Thr Arg Tyr Asn Ala Pro Ser Ile Asp
 115 120 125

eol f-seql . txt

Gly Asp Arg Thr Thr Phe Thr Glu Tyr Trp Ser Val Arg Glu Ser Lys
130 135 140

Arg Pro Thr Gly Ser Asn Ala Thr Ile Thr Phe Ser Asn His Val Asn
145 150 155 160

Ala Trp Lys Ser His Gly Met Asn Leu Gly Ser Asn Trp Ala Tyr Glu
165 170 175

Val Met Ala Thr Glu Gly Tyr Glu Ser Ser Gly Ser Ser Asn Val Thr
180 185 190

Val Trp Pro Asn Ser Gly Ile Glu Gly Tyr Gly Glu Ser Ser Ala Ser
195 200 205

Ala Ser Ala Ala Ala Ser Ala Ala Ser Thr Val Ala Asn Ser Val Ser
210 215 220

Arg Leu Ser Ser Pro Ser Ala Val Ser Arg Val Ser Ser Ala Val Ser
225 230 235 240

Ser Leu Val Ser Asn Gly Glu Val Asn Met Ala Ala Leu Pro Asn Ile
245 250 255

Ile Ser Asn Ile Ser Ser Val Ser Ala Ser Ala Pro Gly Ala Ser
260 265 270

Gly Cys Glu Val Ile Val Glu Ala Leu Leu Glu Val Ile Thr Ala Leu
275 280 285

Val Glu Ile Val Ser Ser Ser Val Gly Tyr Ile Asn Pro Ser Ala
290 295 300

Val Asn Glu Ile Thr Asn Val Val Ala Asn Ala Met Ala Glu Val Met
305 310 315 320

Gly

<210> 83
<211> 984
<212> DNA
<213> Artificial Sequence

<220>
<223> Fusion protein

<220>
<221> misc_feature
<222> (37)..(195)
<223> hEGF sequence

<400> 83
atggcagca gcggccatca tcatcatcat catatgaact ccgactccga atgtccattg
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eol f-seql . txt

tcccacgacg	gttactgttt	gcacgacggt	gtttgtatgt	acatcgaagc	tttggacaag	120
tacgcttcta	actgtgttgt	tggttacatc	ggtgaaagat	gtcaatacag	agacttcaag	180
tggtggaaat	tgagaccgaa	ttcaagccat	accacccgt	ggaccaaccc	gggcctggcg	240
gaaaacttta	tgaacagctt	tatgcagggc	ctgagcagca	tgccgggctt	taccgcgagc	300
cagctggatg	atatgagcac	cattgcgcag	agcatggtgc	agagcattca	gagcctggcg	360
gcfgcaggcc	gcaccagccc	gaacaaactg	caggcgtga	acatggcgtt	tgcgagcagc	420
atggcggaaa	ttgcggcgag	cgaagaaggc	ggcggcagcc	tgagcaccaa	aaccagcagc	480
attgcgagcg	cgtatgagcaa	cgcgttctg	cagaccaccg	gcgtggtgaa	ccagccgttt	540
attaacgaaa	ttacccagct	ggtgagcatg	tttgcgcagg	cgggcatgaa	cgtatgtgagc	600
gcgggcaatt	cagggatcca	agtttatggt	cagagtagtg	cttctgcttc	agctgctgcg	660
tcagctgcta	gtactgttagc	taattcggtg	agtcgcctct	catgcgccttc	cgcagtatct	720
cagagtttctt	cagcagtttc	tagttggtt	tcaaatggtc	aagtgaatat	ggcagcgtta	780
cctaataatca	tttccaacat	ttttcttctt	gtcagtgcatt	ctgctcctgg	tgcttctgga	840
tgtgaggtca	tagtgcaga	tctactcgaa	gtcatcactg	ctcttggta	aatcgttagt	900
tcttcttagtg	ttggatata	taatccatct	gctgtgaacc	aaattactaa	tgttggct	960
aatgccatgg	ctcaagtaat	gggc				984

<210> 84

<211> 328

<212> PRT

<213> Artificial Sequence

<220>

<223> Fusion protein

<220>

<221> MISC_FEATURE

<222> (13)..(65)

<223> hEGF

<400> 84

Met Gl y Ser Ser Gl y His His His His His His Met Asn Ser Asp Ser
 1 5 10 15

Gl u Cys Pro Leu Ser His Asp Gl y Tyr Cys Leu His Asp Gl y Val Cys
 20 25 30

Met Tyr Ile Gl u Al a Leu Asp Lys Tyr Al a Cys Asn Cys Val Val Gl y
 35 40 45

Tyr Ile Gl y Gl u Arg Cys Gl n Tyr Arg Asp Phe Lys Trp Trp Gl u Leu
 50 55 60

Arg Pro Asn Ser Ser His Thr Thr Pro Trp Thr Asn Pro Gl y Leu Al a
 65 70 75 80

eol f-seql . txt

Gl u Asn Phe Met Asn Ser Phe Met Gl n Gl y Leu Ser Ser Met Pro Gl y
85 90 95

Phe Thr Al a Ser Gl n Leu Asp Asp Met Ser Thr Ile Al a Gl n Ser Met
100 105 110

Val Gl n Ser Ile Gl n Ser Leu Al a Al a Gl n Gl y Arg Thr Ser Pro Asn
115 120 125

Lys Leu Gl n Al a Leu Asn Met Al a Phe Al a Ser Ser Met Al a Gl u Ile
130 135 140

Al a Al a Ser Gl u Gl u Gl y Gl y Ser Leu Ser Thr Lys Thr Ser Ser
145 150 155 160

Ile Al a Ser Al a Met Ser Asn Al a Phe Leu Gl n Thr Thr Gl y Val Val
165 170 175

Asn Gl n Pro Phe Ile Asn Gl u Ile Thr Gl n Leu Val Ser Met Phe Al a
180 185 190

Gl n Al a Gl y Met Asn Asp Val Ser Al a Gl y Asn Ser Gl y Ile Gl n Gl y
195 200 205

Tyr Gl y Gl n Ser Ser Al a Ser Al a Ser Al a Al a Ser Al a Al a Ser
210 215 220

Thr Val Al a Asn Ser Val Ser Arg Leu Ser Ser Pro Ser Al a Val Ser
225 230 235 240

Arg Val Ser Ser Al a Val Ser Ser Leu Val Ser Asn Gl y Gl n Val Asn
245 250 255

Met Al a Al a Leu Pro Asn Ile Ile Ser Asn Ile Ser Ser Ser Val Ser
260 265 270

Al a Ser Al a Pro Gl y Al a Ser Gl y Cys Gl u Val Ile Val Gl n Al a Leu
275 280 285

Leu Gl u Val Ile Thr Al a Leu Val Gl n Ile Val Ser Ser Ser Ser Val
290 295 300

Gl y Tyr Ile Asn Pro Ser Al a Val Asn Gl n Ile Thr Asn Val Val Al a
305 310 315 320

Asn Al a Met Al a Gl n Val Met Gl y
325

<210> 85
<211> 567

eol f-seql . txt

<212> DNA

<213> Artificial Sequence

<220>

<223> Fusion protein

<220>

<221> misc_feature

<222> (28)..(186)

<223> hEGF

<400> 85

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aactgtgttgggttacat cggtaaaaga tgtcaataca gagacttcaa gtggggaa 180
ttgagaccga attcagggat ccaagggtat ggtcagagta gtgcttctgc ttcagctgct 240
gcgtcagctg ctagtactgt agctaattcg gtgagtcgcc tctcatcgcc ttccgcagta 300
tctcgagttt cttcagcagt ttctagcttgggttcaatgtgtcaagtgtaa tatggcagcg 360
ttacctaata tcatttccaa catttcttct tctgtcagtgtcatgtctcc tggtgcttct 420
ggatgtgagg tcatagtgcagctctactc gaagtcatca ctgcttctgt tcaaatcggtt 480
agttcttcta gtgttgata tattaatcca tctgctgtga accaaattac taatgttgg 540
gctaatgcca tggctcaagt aatggc 567

<210> 86

<211> 189

<212> PRT

<213> Artificial Sequence

<220>

<223> Fusion protein

<220>

<221> MI SC_FEATURE

<222> (10)..(62)

<223> hEGF

<400> 86

Met Gly His His His His His Met Asn Ser Asp Ser Glu Cys Pro
1 5 10 15

Leu Ser His Asp Gly Tyr Cys Leu His Asp Gly Val Cys Met Tyr Ile
20 25 30

Glu Ala Leu Asp Lys Tyr Ala Cys Asn Cys Val Val Glu Tyr Ile Gly
35 40 45

Glu Arg Cys Glu Tyr Arg Asp Phe Lys Trp Trp Glu Leu Arg Pro Asn
50 55 60

Ser Gly Ile Glu Glu Tyr Glu Glu Ser Ser Ala Ser Ala Ser Ala Ala
65 70 75 80

eol f-seql . txt

Ala Ser Ala Ala Ser Thr Val Ala Asn Ser Val Ser Arg Leu Ser Ser
85 90 95

Pro Ser Ala Val Ser Arg Val Ser Ser Ala Val Ser Ser Leu Val Ser
100 105 110

Asn Gly Gln Val Asn Met Ala Ala Leu Pro Asn Ile Ile Ser Asn Ile
115 120 125

Ser Ser Ser Val Ser Ala Ser Ala Pro Gly Ala Ser Gly Cys Glu Val
130 135 140

Ile Val Gln Ala Leu Leu Glu Val Ile Thr Ala Leu Val Gln Ile Val
145 150 155 160

Ser Ser Ser Ser Val Gly Tyr Ile Asn Pro Ser Ala Val Asn Gln Ile
165 170 175

Thr Asn Val Val Ala Asn Ala Met Ala Gln Val Met Gly
180 185