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DESCRIPTION

FIELD OF THE INVENTION

[0001] The present invention provides variant subtilisins and compositions comprising at least one variant subtilisin set forth herein, as well as methods for using these variants and compositions.

BACKGROUND OF THE INVENTION

[0002] Serine proteases are a subgroup of carbonyl hydrolases comprising a diverse class of enzymes having a wide range of specificities and biological functions. Much research has been conducted on the subtilisins, due largely to their usefulness in cleaning and feed applications. Additional work has been focused on the adverse environmental conditions (e.g., exposure to oxidative agents, chelating agents, extremes of temperature and/or pH) which can adversely impact the functionality of these enzymes in various applications. Nonetheless, there remains a need in the art for enzyme systems that are able to resist these adverse conditions and retain or have improved activity over those currently known in the art.

SUMMARY OF THE INVENTION

[0003] The invention provides a variant of subtilisin BPN[®] which differs from the sequence of BPN[®] subtilisin of SEQ ID NO:2 only by one of the following sets of modifications:

G097A/G128S/Y217Q,

G097A/G128S/P129D/Y217Q,

G097A/G128S/P129V/A134T/Y217Q,

G097A/G128S/P129V/Y217Q,

V068I/G097A/G128S/P129D/Y217Q,

G097A/S101D/G128S/S204F/Y217Q/S236F/T254P,

G097A/S101D/G128S/Y217Q,

G097A/S101D/G128S/P129D/Y217Q,

G097A/Z09901S/A114S/G128S/Y217Q,

G097A/G128S/P129T/Y217Q,

G097A/G128S/P129C/Y217Q,

G097A/G128S/P129Q/Y217Q,

G097A/G128S/P129E/Y217Q,

G097A/G128S/P129A/Y217Q,

G097A/G128S/P129K/Y217Q,

G097A/G128S/P129K/G31S/Y217Q,

G097A/G128S/P129R/Y217Q,

G097A/G128S/P129V/Y217Q,

G097A/G128S/P129Y/Y217Q,

G097A/G128S/P129L/Y217Q, or

G097A/G128S/P129W/Y217Q.

[0004] The present invention also provides a method of cleaning, said method comprising the steps of:

1. a) contacting a surface and/or an article comprising a fabric with at least one variant of subtilisin BPN' according to the invention; and
2. b) optionally washing and/or rinsing said surface or article.

[0005] The present invention also provides animal feeds comprising at least one subtilisin variant provided herein, as well as food processing compositions comprising at least one subtilisin variant provided herein.

BRIEF DESCRIPTION OF THE DRAWINGS

[0006]

FIG. 1 shows a diagram illustrating the method used for creating in-frame deletions and insertions. Library was designed to have 33% of clones with substitutions; 33% of clones with insertions and 33% of clones with deletions.

FIG. 2A Plasmid map of pAC-FNA10. The plasmid elements are as follows: pUB110 = DNA fragment from plasmid pUB110 (McKenzie et al., Plasmid 15:93-103 [1986], pBR322 = DNA fragment from plasmid pBR322 (Bolivar et al., Gene 2:95-113 [1977]), pC194 = DNA fragment from plasmid pC194 (Horinouchi and Weisblum, J. Bacteriol 150:815-825 [1982]).

FIG. 2B provides a map of pHPLT-FNA.

[0007] IG. 2C provides a map of pHPLT-BPN. Vector pHPLT contains the LAT promoter (PLAT), a sequence encoding the LAT signal peptide (preLAT). The origin of replication and neoR are derived from plasmid pUB 110.

DESCRIPTION OF THE INVENTION

[0008] The present invention provides variant subtilisins and compositions comprising at least one variant subtilisin set forth herein, as well as methods for using these variants and compositions.

[0009] Several suitable methods are known in the art for generating modified polynucleotide sequences of the present invention, including but not limited to site-saturation mutagenesis, scanning mutagenesis, insertional mutagenesis, deletion mutagenesis, random mutagenesis, site-directed mutagenesis, and directed-evolution, as well as various other recombinatorial approaches. The commonly used methods include DNA shuffling (See, Stemmer, Proc Natl Acad Sci U S A. 25:10747-51 [1994]), methods based on non-homologous recombination of genes (e.g. ITCHY) (Ostermeier et al., Bioorg Med Chem. 7:2139-44 [1999]), SCRACHY (Lutz et al. Proc Natl Acad Sci USA. 98:11248-53 [2001]), SHIPREC (Sieber et al., Nat Biotechnol., 19:456-60 [2001]), and NRR (Bittker et al., Nat Biotechnol., 20:1024-9 [2001]; Bittker et al., Proc Natl Acad Sci. USA. 101:7011-6 [2004]), and methods that rely on the use of oligonucleotides to insert random and targeted mutations, deletions and/or insertions (Ness et al., Nat Biotechnol. 20:1251-5 [2002]; Coco et al., Nat Biotechnol., 20:1246-50 [2002]; Zha et al., Chembiochem. 3:34-9 [2003], Glaser et al., J Immunol., 149:3903-13 [1992], Sondek and Shortle, Proc Natl Acad Sci USA 89:3581-5 [1992], Yáñez et al., Nucleic Acids Res., 32:e158 [2004], Osuna et al., Nucleic Acids Res., 32:e136 [2004], Gaytán et al., Nucleic Acids Res., 29:E9 [2001], and Gaytán et al., Nucleic Acids Res., 30:e84 [2002]).

[0010] In some embodiments, the full-length parent polynucleotide is ligated into an appropriate expression plasmid, and the following mutagenesis method is used to facilitate the construction of the modified protease of the present invention, although other methods may be used. The method is based on that described by Pisarchik *et al.* (Pisarchik et al., Prot. Eng. Des. Select., 20:257-265 [2007]). In some embodiments, an added advantage is provided, in that the restriction enzyme used herein cuts outside its recognition sequence, which allows digestion of practically any nucleotide sequence and precludes formation of a restriction site scar. First, as described herein, a naturally-occurring gene encoding the full-length protease is obtained and

sequenced and scanned for one or more points at which it is desired to make a mutation (deletion, insertion, substitution, or a combination thereof) at one or more amino acids. Mutation of the gene in order to change its sequence to conform to the desired sequence is accomplished by primer extension in accord with generally known methods. Fragments to the left and to the right of the desired point(s) of mutation are amplified by PCR and to include the *Eam*1104 restriction site. The left and right fragments are digested with *Eam*1104I to generate a plurality of fragments having complementary three base overhangs, which are then pooled and ligated to generate a library of modified sequences containing one or more mutations. This method avoids the occurrence of frame-shift mutations. In addition, this method simplifies the mutagenesis process because all of the oligonucleotides can be synthesized so as to have the same restriction site, and no synthetic linkers are necessary to create the restriction sites as is required by some other methods

Compositions

[0011] The present invention provides variant proteases and compositions comprising at least one of the variant subtilisins set forth herein. The variant proteases may be employed in various commercial applications where degradation or synthesis of polypeptides are desired, including cleaning compositions, as well as feed components, textile processing, leather finishing, grain processing, meat processing, food processing, preparation of protein hydrolysates, digestive aids, microbicidal compositions, bacteriostatic compositions, fungistatic compositions, and personal care products (*e.g.*, oral care, hair care, and/or skin care).

[0012] The variant proteases have comparable or improved wash performance, as compared to presently used subtilisin proteases.

Definitions

[0013] Unless otherwise indicated, the practice of the present invention involves conventional techniques commonly used in molecular biology, protein engineering, microbiology, and recombinant DNA, which are within the skill of the art. Such techniques are known to those of skill in the art and are described in numerous texts and reference works (*See e.g.*, Sambrook et al., "Molecular Cloning: A Laboratory Manual", Second Edition (Cold Spring Harbor), [1989]); and Ausubel et al., "Current Protocols in Molecular Biology" [1987]). All patents, patent applications, articles and publications mentioned herein, both *supra* and *infra*, are hereby expressly incorporated herein by reference.

[0014] Unless defined otherwise herein, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention pertains. There are various dictionaries available and known to those in the art that provide definitions of these terms. Although any methods and materials similar or equivalent to those described herein find use in the practice of the present invention, the preferred methods and materials are described herein. Accordingly, the terms defined immediately below are more fully described by reference to the Specification as a whole. Also, as used herein, the singular "a", "an" and "the" includes the plural reference unless the context clearly indicates otherwise. Numeric ranges are inclusive of the numbers defining the range. Unless otherwise indicated, nucleic acids are written left to right in 5' to 3' orientation; amino acid sequences are written left to right in amino to carboxy orientation, respectively. It is to be understood that this invention is not limited to the particular methodology, protocols, and reagents described, as these may vary, depending upon the context they are used by those of skill in the art.

[0015] The practice of the present invention employs, unless otherwise indicated, conventional techniques of protein purification, Molecular biology, microbiology, recombinant DNA techniques and protein sequencing, all of which are within the skill of those in the art.

[0016] Furthermore, the headings provided herein are not limitations of the various aspects or embodiments of the invention which can be had by reference to the specification as a whole. Accordingly, the terms defined immediately below are more fully defined by reference to the specification as a whole. Nonetheless, in order to facilitate understanding of the invention, a number of terms are defined below.

[0017] As used herein, the terms "protease," and "proteolytic activity" refer to a protein or peptide exhibiting the ability to hydrolyze peptides or substrates having peptide linkages. Many well known procedures exist for measuring proteolytic activity (Kalisz, "Microbial Proteinases," In: Fiechter (ed.), *Advances in Biochemical Engineering/Biotechnology*, [1988]). For example, proteolytic activity may be ascertained by comparative assays which analyze the respective protease's ability to hydrolyze a commercial substrate. Exemplary substrates useful in the analysis of protease or proteolytic activity, include, but are not limited to

di-methyl casein (Sigma C-9801), bovine collagen (Sigma C-9879), bovine elastin (Sigma E-1625), and bovine keratin (ICN Biomedical 902111). Colorimetric assays utilizing these substrates are well known in the art (See e.g., WO 99/34011; and U.S. Pat. No. 6,376,450). The pNA assay (See e.g., Del Mar et al., Anal. Biochem., 99:316-320 [1979]) also finds use in determining the active enzyme concentration for fractions collected during gradient elution. This assay measures the rate at which p-nitroaniline is released as the enzyme hydrolyzes the soluble synthetic substrate, succinyl-alanine-alanine-proline-phenylalanine-p-nitroanilide (sAAPF-pNA). The rate of production of yellow color from the hydrolysis reaction is measured at 410 nm on a spectrophotometer and is proportional to the active enzyme concentration. In addition, absorbance measurements at 280 nm can be used to determine the total protein concentration. The active enzyme/total-protein ratio gives the enzyme purity.

[0018] As used herein, "the genus *Bacillus*" includes all species within the genus "*Bacillus*," as known to those of skill in the art, including but not limited to *B. subtilis*, *B. licheniformis*, *B. lentus*, *B. brevis*, *B. stearothermophilus*, *B. alkalophilus*, *B. amyloliquefaciens*, *B. clausii*, *B. halodurans*, *B. megaterium*, *B. coagulans*, *B. circulans*, *B. lautus*, and *B. thuringiensis*. It is recognized that the genus *Bacillus* continues to undergo taxonomical reorganization. Thus, it is intended that the genus include species that have been reclassified, including but not limited to such organisms as *B. stearothermophilus*, which is now named "*Geobacillus stearothermophilus*." The production of resistant endospores in the presence of oxygen is considered the defining feature of the genus *Bacillus*, although this characteristic also applies to the recently named *Alicyclobacillus*, *Amphibacillus*, *Aneurinibacillus*, *Anoxybacillus*, *Brevibacillus*, *Filobacillus*, *Gracilibacillus*, *Halobacillus*, *Paenibacillus*, *Salibacillus*, *Thermobacillus*, *Ureibacillus*, and *Virgibacillus*.

[0019] The terms "polynucleotide" and "nucleic acid", used interchangeably herein, refer to a polymeric form of nucleotides of any length, either ribonucleotides or deoxyribonucleotides. These terms include, but are not limited to, a single-, double- or triple-stranded DNA, genomic DNA, cDNA, RNA, DNA-RNA hybrid, or a polymer comprising purine and pyrimidine bases, or other natural, chemically, biochemically modified, non-natural or derivatized nucleotide bases. The following are non-limiting examples of polynucleotides: genes, gene fragments, chromosomal fragments, ESTs, exons, introns, mRNA, tRNA, rRNA, ribozymes, cDNA, recombinant polynucleotides, branched polynucleotides, plasmids, vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes, and primers. In some embodiments, polynucleotides comprise modified nucleotides, such as methylated nucleotides and nucleotide analogs, uracyl, other sugars and linking groups such as fluororibose and thioate, and nucleotide branches. In alternative embodiments, the sequence of nucleotides is interrupted by non-nucleotide components.

[0020] As used herein, the terms "DNA construct" and "transforming DNA" are used interchangeably to refer to DNA used to introduce sequences into a host cell or organism. The DNA may be generated *in vitro* by PCR or any other suitable technique(s) known to those in the art. In particularly preferred embodiments, the DNA construct comprises a sequence of interest (e.g., as an incoming sequence). In some embodiments, the sequence is operably linked to additional elements such as control elements (e.g., promoters, etc.). The DNA construct may further comprise a selectable marker. It may further comprise an incoming sequence flanked by homology boxes. In a further embodiment, the transforming DNA comprises other non-homologous sequences, added to the ends (e.g., stuffer sequences or flanks). In some embodiments, the ends of the incoming sequence are closed such that the transforming DNA forms a closed circle. The transforming sequences may be wild-type, mutant or modified. In some embodiments, the DNA construct comprises sequences homologous to the host cell chromosome. In other embodiments, the DNA construct comprises non-homologous sequences. Once the DNA construct is assembled *in vitro* it may be used to: 1) insert heterologous sequences into a desired target sequence of a host cell, and/or 2) mutagenize a region of the host cell chromosome (i.e., replace an endogenous sequence with a heterologous sequence), 3) delete target genes, and/or 4) introduce a replicating plasmid into the host.

[0021] As used herein, the terms "expression cassette" and "expression vector" refer to nucleic acid constructs generated recombinantly or synthetically, with a series of specified nucleic acid elements that permit transcription of a particular nucleic acid in a target cell. The recombinant expression cassette can be incorporated into a plasmid, chromosome, mitochondrial DNA, plastid DNA, virus, or nucleic acid fragment. Typically, the recombinant expression cassette portion of an expression vector includes, among other sequences, a nucleic acid sequence to be transcribed and a promoter. In preferred embodiments, expression vectors have the ability to incorporate and express heterologous DNA fragments in a host cell. Many prokaryotic and eukaryotic expression vectors are commercially available. Selection of appropriate expression vectors is within the knowledge of those of skill in the art. The term "expression cassette" is used interchangeably herein with "DNA construct," and their grammatical equivalents. Selection of appropriate expression vectors is within the knowledge of those of skill in the art.

[0022] As used herein, the term "vector" refers to a polynucleotide construct designed to introduce nucleic acids into one or more cell types. Vectors include cloning vectors, expression vectors, shuttle vectors, plasmids, cassettes and the like. In some embodiments, the polynucleotide construct comprises a DNA sequence encoding the protease (e.g., precursor or mature protease) that is operably linked to a suitable prosequence (e.g., secretory, etc.) capable of effecting the expression of the DNA in a suitable host.

[0023] As used herein, the term "plasmid" refers to a circular double-stranded (ds) DNA construct used as a cloning vector, and which forms an extrachromosomal self-replicating genetic element in some eukaryotes or prokaryotes, or integrates into the host chromosome.

[0024] As used herein in the context of introducing a nucleic acid sequence into a cell, the term "introduced" refers to any method suitable for transferring the nucleic acid sequence into the cell. Such methods for introduction include but are not limited to protoplast fusion, transfection, transformation, conjugation, and transduction (*See e.g.*, Ferrari et al., "Genetics," in Hardwood et al, (eds.), *Bacillus*, Plenum Publishing Corp., pages 57-72, [1989]).

[0025] As used herein, the terms "transformed" and "stably transformed" refers to a cell that has a non-native (heterologous) polynucleotide sequence integrated into its genome or as an episomal plasmid that is maintained for at least two generations.

[0026] A nucleic acid is "operably linked" when it is placed into a functional relationship with another nucleic acid sequence. For example, DNA encoding a secretory leader (*i.e.*, a signal peptide), is operably linked to DNA for a polypeptide if it is expressed as a preprotein that participates in the secretion of the polypeptide; a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the sequence; or a ribosome binding site is operably linked to a coding sequence if it is positioned so as to facilitate translation. Generally, "operably linked" means that the DNA sequences being linked are contiguous, and, in the case of a secretory leader, contiguous and in reading phase. However, enhancers do not have to be contiguous. Linking is accomplished by ligation at convenient restriction sites. If such sites do not exist, the synthetic oligonucleotide adaptors or linkers are used in accordance with conventional practice.

[0027] As used herein the term "gene" refers to a polynucleotide (*e.g.*, a DNA segment), that encodes a polypeptide and includes regions preceding and following the coding regions as well as intervening sequences (introns) between individual coding segments (exons).

[0028] As used herein, "homologous genes" refers to a pair of genes from different, but usually related species, which correspond to each other and which are identical or very similar to each other. The term encompasses genes that are separated by speciation (*i.e.*, the development of new species) (*e.g.*, orthologous genes), as well as genes that have been separated by genetic duplication (*e.g.*, paralogous genes).

[0029] As used herein, "ortholog" and "orthologous genes" refer to genes in different species that have evolved from a common ancestral gene (*i.e.*, a homologous gene) by speciation. Typically, orthologs retain the same function during the course of evolution. Identification of orthologs finds use in the reliable prediction of gene function in newly sequenced genomes.

[0030] As used herein, "paralog" and "paralogous genes" refer to genes that are related by duplication within a genome. While orthologs retain the same function through the course of evolution, paralogs evolve new functions, even though some functions are often related to the original one. Examples of paralogous genes include, but are not limited to genes encoding trypsin, chymotrypsin, elastase, and thrombin, which are all serine proteinases and occur together within the same species.

[0031] As used herein, proteins are defined as having a common "fold" if they have the same major secondary structures in the same arrangement and with the same topological connections. Different proteins with the same fold often have peripheral elements of secondary structure and turn regions that differ in size and conformation. In some cases, these differing peripheral regions may comprise half the structure. Proteins placed together in the same fold category do not necessarily have a common evolutionary origin (*e.g.*, structural similarities arising from the physics and chemistry of proteins favoring certain packing arrangements and chain topologies).

[0032] As used herein, "homology" refers to sequence similarity or identity, with identity being preferred. This homology is determined using standard techniques known in the art (*See e.g.*, Smith and Waterman, *Adv. Appl. Math.*, 2:482 [1981]; Needleman and Wunsch, *J. Mol. Biol.*, 48:443 [1970]; Pearson and Lipman, *Proc. Natl. Acad. Sci. USA* 85:2444 [1988]; programs such as GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package (Genetics Computer Group, Madison, WI); and Devereux et al., *Nucl. Acid Res.*, 12:387-395 [1984]).

[0033] As used herein, an "analogous sequence" is one wherein the function of the gene is essentially the same as the gene based on the wild-type subtilisin protease. Additionally, analogous genes include at least about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 95%, about 97%, about 98%, about 99%, or about 100% sequence identity with the sequence of the wild-type subtilisin protease. Alternately, analogous sequences

have an alignment of between about 70 to about 100% of the genes found in the *B. amyloliquefaciens* subtilisin protease region. In additional embodiments more than one of the above properties applies to the sequence. Analogous sequences are determined by known methods of sequence alignment. A commonly used alignment method is BLAST, although as indicated above and below, there are other methods that also find use in aligning sequences.

[0034] One example of a useful algorithm is PILEUP. PILEUP creates a multiple sequence alignment from a group of related sequences using progressive, pair-wise alignments. It can also plot a tree showing the clustering relationships used to create the alignment. PILEUP uses a simplification of the progressive alignment method of Feng and Doolittle (Feng and Doolittle, *J. Mol. Evol.*, 35:351-360 [1987]). The method is similar to that described by Higgins and Sharp (Higgins and Sharp, *CABIOS* 5:151-153 [1989]). Useful PILEUP parameters including a default gap weight of 3.00, a default gap length weight of 0.10, and weighted end gaps.

[0035] Another example of a useful algorithm is the BLAST algorithm, described by Altschul *et al.*, (Altschul *et al.*, *J. Mol. Biol.*, 215:403-410, [1990]; and Karlin *et al.*, *Proc. Natl. Acad. Sci. USA* 90:5873-5787 [1993]). A particularly useful BLAST program is the WU-BLAST-2 program (See, Altschul *et al.*, *Meth. Enzymol.*, 266:460-480 [1996]). WU-BLAST-2 uses several search parameters, most of which are set to the default values. The adjustable parameters are set with the following values: overlap span = 1, overlap fraction = 0.125, word threshold (T) = 11. The HSP S and HSP S2 parameters are dynamic values and are established by the program itself depending upon the composition of the particular sequence and composition of the particular database against which the sequence of interest is being searched. However, the values may be adjusted to increase sensitivity. A % amino acid sequence identity value is determined by the number of matching identical residues divided by the total number of residues of the "longer" sequence in the aligned region. The "longer" sequence is the one having the most actual residues in the aligned region (gaps introduced by WU-Blast-2 to maximize the alignment score are ignored).

[0036] Thus, "percent (%) nucleic acid sequence identity" is defined as the percentage of nucleotide residues in a candidate sequence that are identical with the nucleotide residues of the starting sequence (*i.e.*, the sequence of interest). A preferred method utilizes the BLASTN module of WU-BLAST-2 set to the default parameters, with overlap span and overlap fraction set to 1 and 0.125, respectively.

[0037] As used herein, "recombinant" includes reference to a cell or vector, that has been modified by the introduction of a heterologous nucleic acid sequence or that the cell is derived from a cell so modified. Thus, for example, recombinant cells express genes that are not found in identical form within the native (non-recombinant) form of the cell or express native genes that are otherwise abnormally expressed, under expressed or not expressed at all as a result of deliberate human intervention. "Recombination," "recombining," and generating a "recombined" nucleic acid are generally the assembly of two or more nucleic acid fragments wherein the assembly gives rise to a chimeric gene.

[0038] As used herein, the terms "amplification" and "gene amplification" refer to a process by which specific DNA sequences are disproportionately replicated such that the amplified gene becomes present in a higher copy number than was initially present in the genome. In some embodiments, selection of cells by growth in the presence of a drug (*e.g.*, an inhibitor of an inhibitable enzyme) results in the amplification of either the endogenous gene encoding the gene product required for growth in the presence of the drug or by amplification of exogenous (*i.e.*, input) sequences encoding this gene product, or both.

[0039] "Amplification" is a special case of nucleic acid replication involving template specificity. It is to be contrasted with non-specific template replication (*i.e.*, replication that is template-dependent but not dependent on a specific template). Template specificity is here distinguished from fidelity of replication (*i.e.*, synthesis of the proper polynucleotide sequence) and nucleotide (ribo- or deoxyribo-) specificity. Template specificity is frequently described in terms of "target" specificity. Target sequences are "targets" in the sense that they are sought to be sorted out from other nucleic acid. Amplification techniques have been designed primarily for this sorting out.

[0040] As used herein, the term "primer" refers to an oligonucleotide, whether occurring naturally as in a purified restriction digest or produced synthetically, which is capable of acting as a point of initiation of synthesis when placed under conditions in which synthesis of a primer extension product which is complementary to a nucleic acid strand is induced, (*i.e.*, in the presence of nucleotides and an inducing agent such as DNA polymerase and at a suitable temperature and pH). The primer is preferably single stranded for maximum efficiency in amplification, but may alternatively be double stranded. If double stranded, the primer is first treated to separate its strands before being used to prepare extension products. Preferably, the primer is an oligodeoxyribonucleotide. The primer must be sufficiently long to prime the synthesis of extension products in the presence of the inducing agent. The exact lengths of the primers will depend on many factors, including temperature, source of primer and the use of the method.

[0041] As used herein, the term "probe" refers to an oligonucleotide (*i.e.*, a sequence of nucleotides), whether occurring naturally as in a purified restriction digest or produced synthetically, recombinantly or by PCR amplification, which is capable of hybridizing to another oligonucleotide of interest. A probe may be single-stranded or double-stranded. Probes are useful in the detection, identification and isolation of particular gene sequences. It is contemplated that any probe used in the present invention will be labeled with any "reporter molecule," so that is detectable in any detection system, including, but not limited to enzyme (*e.g.*, ELISA, as well as enzyme-based histochemical assays), fluorescent, radioactive, and luminescent systems. It is not intended that the present invention be limited to any particular detection system or label.

[0042] As used herein, the term "target," when used in reference to the polymerase chain reaction, refers to the region of nucleic acid bounded by the primers used for polymerase chain reaction. Thus, the "target" is sought to be sorted out from other nucleic acid sequences. A "segment" is defined as a region of nucleic acid within the target sequence.

[0043] As used herein, the term "polymerase chain reaction" ("PCR") refers to the methods of U.S. Patent Nos. 4,683,195 4,683,202, and 4,965,188, which include methods for increasing the concentration of a segment of a target sequence in a mixture of genomic DNA without cloning or purification. This process for amplifying the target sequence consists of introducing a large excess of two oligonucleotide primers to the DNA mixture containing the desired target sequence, followed by a precise sequence of thermal cycling in the presence of a DNA polymerase. The two primers are complementary to their respective strands of the double stranded target sequence. To effect amplification, the mixture is denatured and the primers then annealed to their complementary sequences within the target molecule. Following annealing, the primers are extended with a polymerase so as to form a new pair of complementary strands. The steps of denaturation, primer annealing and polymerase extension can be repeated many times (*i.e.*, denaturation, annealing and extension constitute one "cycle"; there can be numerous "cycles") to obtain a high concentration of an amplified segment of the desired target sequence. The length of the amplified segment of the desired target sequence is determined by the relative positions of the primers with respect to each other, and therefore, this length is a controllable parameter. By virtue of the repeating aspect of the process, the method is referred to as the "polymerase chain reaction" (hereinafter "PCR"). Because the desired amplified segments of the target sequence become the predominant sequences (in terms of concentration) in the mixture, they are said to be "PCR amplified".

[0044] As used herein, the term "amplification reagents" refers to those reagents (deoxyribonucleotide triphosphates, buffer, etc.), needed for amplification except for primers, nucleic acid template and the amplification enzyme. Typically, amplification reagents along with other reaction components are placed and contained in a reaction vessel (test tube, microwell, etc.).

[0045] As used herein, the term "RT-PCR" refers to the replication and amplification of RNA sequences. In this method, reverse transcription is coupled to PCR, most often using a one enzyme procedure in which a thermostable polymerase is employed, as described in U.S. Patent No. 5,322,770. In RT-PCR, the RNA template is converted to cDNA due to the reverse transcriptase activity of the polymerase, and then amplified using the polymerizing activity of the polymerase (*i.e.*, as in other PCR methods).

[0046] As used herein, the terms "restriction endonucleases" and "restriction enzymes" refer to bacterial enzymes, each of which cut double-stranded DNA at or near a specific nucleotide sequence.

[0047] A "restriction site" refers to a nucleotide sequence recognized and cleaved by a given restriction endonuclease and is frequently the site for insertion of DNA fragments. In certain embodiments of the invention restriction sites are engineered into the selective marker and into 5' and 3' ends of the DNA construct.

[0048] "Homologous recombination" means the exchange of DNA fragments between two DNA molecules or paired chromosomes at the site of identical or nearly identical nucleotide sequences. In a preferred embodiment, chromosomal integration is homologous recombination.

[0049] As used herein "Amino acid" refers to peptide or protein sequences or portions thereof. The terms "protein," "peptide," and "polypeptide" are used interchangeably.

[0050] As used herein, "protein of interest" and "polypeptide of interest" refer to a protein/polypeptide that is desired and/or being assessed. In some embodiments, the protein of interest is expressed intracellularly, while in other embodiments, it is a secreted polypeptide. In particularly preferred embodiments, these enzymes include the serine proteases of the present invention. In some embodiments, the protein of interest is a secreted polypeptide which is fused to a signal peptide (*i.e.*, an amino-terminal extension on a protein to be secreted). Nearly all secreted proteins use an amino-terminal protein extension which plays a crucial role in the targeting to and translocation of precursor proteins across the membrane. This extension is proteolytically removed by a signal peptidase during or immediately following membrane transfer.

[0051] A polynucleotide is said to "encode" an RNA or a polypeptide if, in its native state or when manipulated by methods known to those of skill in the art, it can be transcribed and/or translated to produce the RNA, the polypeptide or a fragment thereof. The anti-sense strand of such a nucleic acid is also said to encode the sequences. As is known in the art, a DNA can be transcribed by an RNA polymerase to produce RNA, but an RNA can be reverse transcribed by reverse transcriptase to produce a DNA. Thus a DNA can encode a RNA and vice versa.

[0052] "Host strain" or "host cell" refers to a suitable host for an expression vector comprising DNA according to the present invention.

[0053] An enzyme is "overexpressed" in a host cell if the enzyme is expressed in the cell at a higher level than the level at which it is expressed in a corresponding wild-type cell.

[0054] The terms "protein" and "polypeptide" are used interchangeably herein. The 3-letter code for amino acids as defined in conformity with the IUPAC-IUB Joint Commission on Biochemical Nomenclature (JCBN) is used throughout this disclosure. It is also understood that a polypeptide may be coded for by more than one nucleotide sequence due to the degeneracy of the genetic code.

[0055] A "prosequence" is an amino acid sequence between the signal sequence and mature protease that is necessary for the secretion of the protease. Cleavage of the pro sequence results in a mature active protease.

[0056] The term "signal sequence" or "signal peptide" refers to any sequence of nucleotides and/or amino acids which may participate in the secretion of the mature or precursor forms of the protein. This definition of signal sequence is a functional one, meant to include all those amino acid sequences encoded by the N-terminal portion of the protein gene, which participate in the effectuation of the secretion of protein. They are often, but not universally, bound to the N-terminal portion of a protein or to the N-terminal portion of a precursor protein. The signal sequence may be endogenous or exogenous. The signal sequence may be that normally associated with the protein (e.g., protease), or may be from a gene encoding another secreted protein. One exemplary exogenous signal sequence comprises the first seven amino acid residues of the signal sequence from *Bacillus subtilis* subtilisin fused to the remainder of the signal sequence of the subtilisin from *Bacillus lentus* (ATCC 21536).

[0057] The term "hybrid signal sequence" refers to signal sequences in which part of sequence is obtained from the expression host fused to the signal sequence of the gene to be expressed. In some embodiments, synthetic sequences are utilized.

[0058] The term "mature" form of a protein or peptide refers to the final functional form of the protein or peptide. For example, the mature form of the BPN' subtilisin protease has the amino acid sequence of SEQ ID NO:2, as set forth below:

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AQSVPYGVSIKAPALISQGYTGSNVKVAVIDSGIDSSIIPIIKVAGGASMVPSETNP
FQDNNSHGTHVAGTVAAALNNSIGVLGVAPSASLYAVKVLGADGSGQYSWIINGIEW
AIANNMDVINMSLGGPSGAALKAAVDKAVASGVVVVAAAAGNEGTSGSSSTVGYT
GKYPSVIAVGAVDSSNQASFSVGPGLDVMAPGVSIQSTLPGNKYGAYNGTSMASP
HVAGAAALILSKHPNWTNTQVRSSLENTITTKLGDSFYFGKGLINVQAAAQ (SEQ ID
NO:2)
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[0059] The term "precursor" form of a protein or peptide refers to a mature form of the protein having a prosequence operably linked to the amino or carboxyl terminus of the protein. The precursor may also have a "signal" sequence operably linked, to the amino terminus of the prosequence. The precursor may also have additional polynucleotides that are involved in post-translational activity (e.g., polynucleotides cleaved therefrom to leave the mature form of a protein or peptide).

[0060] "Naturally occurring enzyme" refers to an enzyme having the unmodified amino acid sequence identical to that found in nature. Naturally occurring enzymes include native enzymes, those enzymes naturally expressed or found in the particular microorganism.

[0061] The terms "derived from" and "obtained from" refer to not only a protease produced or producible by a strain of the organism in question, but also a protease encoded by a DNA sequence isolated from such strain and produced in a host organism containing such DNA sequence. Additionally, the term refers to a protease which is encoded by a DNA sequence of synthetic and/or cDNA origin and which has the identifying characteristics of the protease in question..

[0062] The term "identical" in the context of two nucleic acids or polypeptide sequences refers to the residues in the two sequences that are the same when aligned for maximum correspondence, as measured using one of the following sequence

comparison or analysis algorithms.

[0063] The term "optimal alignment" refers to the alignment giving the highest percent identity score.

[0064] "Percent sequence identity," "percent amino acid sequence identity," "percent gene sequence identity," and/or "percent nucleic acid/polynucleotide sequence identity," with respect to two amino acid, polynucleotide and/or gene sequences (as appropriate), refer to the percentage of residues that are identical in the two sequences when the sequences are optimally aligned. Thus, 80% amino acid sequence identity means that 80% of the amino acids in two optimally aligned polypeptide sequences are identical.

[0065] The phrase "substantially identical" in the context of two nucleic acids or polypeptides thus refers to a polynucleotide or polypeptide that comprising at least about 70% sequence identity, preferably at least about 75%, preferably at least about 80%, preferably at least about 85%, preferably at least about 90%, preferably at least about 95%, preferably at least about 97%, preferably at least about 98%, and preferably at least about 99% sequence identity as compared to a reference sequence using the programs or algorithms (e.g., BLAST, ALIGN, CLUSTAL) using standard parameters. One indication that two polypeptides are substantially identical is that the first polypeptide is immunologically cross-reactive with the second polypeptide. Typically, polypeptides that differ by conservative amino acid substitutions are immunologically cross-reactive. Thus, a polypeptide is substantially identical to a second polypeptide, for example, where the two peptides differ only by a conservative substitution. Another indication that two nucleic acid sequences are substantially identical is that the two molecules hybridize to each other under stringent conditions (e.g., within a range of medium to high stringency).

[0066] The phrase "equivalent," in this context, refers to serine proteases enzymes that are encoded by a polynucleotide capable of hybridizing to the polynucleotide having the sequence as shown in SEQ ID NO: 1, under conditions of medium to maximum stringency. For example, being equivalent means that an equivalent mature serine protease comprises at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, and/or at least about 99% sequence identity to the mature subtilisin protease having the amino acid sequence of SEQ ID NO:2.

[0067] The term "isolated" or "purified" refers to a material that is removed from its original environment (e.g., the natural environment if it is naturally occurring). For example, the material is said to be "purified" when it is present in a particular composition in a higher or lower concentration than exists in a naturally occurring or wild type organism or in combination with components not normally present upon expression from a naturally occurring or wild type organism. For example, a naturally-occurring polynucleotide or polypeptide present in a living animal is not isolated, but the same polynucleotide or polypeptide, separated from some or all of the coexisting materials in the natural system, is isolated. In some embodiments, such polynucleotides are part of a vector, and/or such polynucleotides or polypeptides are part of a composition, and still be isolated in that such vector or composition is not part of its natural environment. In preferred embodiments, a nucleic acid or protein is said to be purified, for example, if it gives rise to essentially one band in an electrophoretic gel or blot.

[0068] The term "isolated", when used in reference to a DNA sequence, refers to a DNA sequence that has been removed from its natural genetic milieu and is thus free of other extraneous or unwanted coding sequences, and is in a form suitable for use within genetically engineered protein production systems. Such isolated molecules are those that are separated from their natural environment and include cDNA and genomic clones. Isolated DNA molecules of the present invention are free of other genes with which they are ordinarily associated, but may include naturally occurring 5' and 3' untranslated regions such as promoters and terminators. The identification of associated regions will be evident to one of ordinary skill in the art (See e.g., Dynan and Tijan, Nature 316:774-78 [1985]). The term "an isolated DNA sequence" is alternatively referred to as "a cloned DNA sequence".

[0069] The term "isolated," when used in reference to a protein, refers to a protein that is found in a condition other than its native environment. In a preferred form, the isolated protein is substantially free of other proteins, particularly other homologous proteins. An isolated protein is more than about 10% pure, preferably more than about 20% pure, and even more preferably more than about 30% pure, as determined by SDS-PAGE. Further aspects of the invention encompass the protein in a highly purified form (i.e., more than about 40% pure, more than about 60% pure, more than about 80% pure, more than about 90% pure, more than about 95% pure, more than about 97% pure, and even more than about 99% pure), as determined by SDS-PAGE.

[0070] As used herein, the term, "combinatorial mutagenesis" refers to methods in which libraries of variants of a starting sequence are generated. In these libraries, the variants contain one or several mutations chosen from a predefined set of mutations. In addition, the methods provide means to introduce random mutations which were not members of the predefined set of mutations. In some embodiments, the methods include those set forth in U.S. Patent Appln. Ser. No. 09/699,250, filed October

26, 2000. In alternative embodiments, combinatorial mutagenesis methods encompass commercially available kits (e.g., QuikChange® Multisite, Stratagene, San Diego, CA).

[0071] As used herein, the term "library of mutants" refers to a population of cells which are identical in most of their genome but include different homologues of one or more genes. Such libraries can be used, for example, to identify genes or operons with improved traits.

[0072] As used herein, the term "starting gene" refers to a gene of interest that encodes a protein of interest that is to be improved and/or changed using the present invention.

[0073] As used herein, the term "multiple sequence alignment" ("MSA") refers to the sequences of multiple homologs of a starting gene that are aligned using an algorithm (e.g., Clustal W).

[0074] As used herein, the terms "consensus sequence" and "canonical sequence" refer to an archetypical amino acid sequence against which all variants of a particular protein or sequence of interest are compared. The terms also refer to a sequence that sets forth the nucleotides that are most often present in a DNA sequence of interest. For each position of a gene, the consensus sequence gives the amino acid that is most abundant in that position in the MSA.

[0075] As used herein, the term "consensus mutation" refers to a difference in the sequence of a starting gene and a consensus sequence. Consensus mutations are identified by comparing the sequences of the starting gene and the consensus sequence resulting from an MSA. In some embodiments, consensus mutations are introduced into the starting gene such that it becomes more similar to the consensus sequence. Consensus mutations also include amino acid changes that change an amino acid in a starting gene to an amino acid that is more frequently found in an MSA at that position relative to the frequency of that amino acid in the starting gene. Thus, the term consensus mutation comprises all single amino acid changes that replace an amino acid of the starting gene with an amino acid that is more abundant than the amino acid in the MSA.

[0076] As used herein, the term "enhanced combinatorial consensus mutagenesis library" refers to a CCM library that is designed and constructed based on screening and/or sequencing results from an earlier round of CCM mutagenesis and screening. In some embodiments, the enhanced CCM library is based on the sequence of an initial hit resulting from an earlier round of CCM. In additional embodiments, the enhanced CCM is designed such that mutations that were frequently observed in initial hits from earlier rounds of mutagenesis and screening are favored. In some preferred embodiments, this is accomplished by omitting primers that encode performance-reducing mutations or by increasing the concentration of primers that encode performance-enhancing mutations relative to other primers that were used in earlier CCM libraries.

[0077] As used herein, the term "performance-reducing mutations" refer to mutations in the combinatorial consensus mutagenesis library that are less frequently found in hits resulting from screening as compared to an unscreened combinatorial consensus mutagenesis library. In preferred embodiments, the screening process removes and/or reduces the abundance of variants that contain "performance-reducing mutations."

[0078] As used herein, the term "functional assay" refers to an assay that provides an indication of a protein's activity. In particularly preferred embodiments, the term refers to assay systems in which a protein is analyzed for its ability to function in its usual capacity. For example, in the case of enzymes, a functional assay involves determining the effectiveness of the enzyme in catalyzing a reaction.

[0079] As used herein, the term "target property" refers to the property of the starting gene that is to be altered. It is not intended that the present invention be limited to any particular target property. However, in some preferred embodiments, the target property is the stability of a gene product (e.g., resistance to denaturation, proteolysis or other degradative factors), while in other embodiments, the level of production in a production host is altered. Indeed, it is contemplated that any property of a starting gene will find use in the present invention.

[0080] The term "property" or grammatical equivalents thereof in the context of a nucleic acid, as used herein, refer to any characteristic or attribute of a nucleic acid that can be selected or detected. These properties include, but are not limited to, a property affecting binding to a polypeptide, a property conferred on a cell comprising a particular nucleic acid, a property affecting gene transcription (e.g., promoter strength, promoter recognition, promoter regulation, enhancer function), a property affecting RNA processing (e.g., RNA splicing, RNA stability, RNA conformation, and post-transcriptional modification), a property affecting translation (e.g., level, regulation, binding of mRNA to ribosomal proteins, post-translational modification). For example, a binding site for a transcription factor, polymerase, regulatory factor, etc., of a nucleic acid may be altered to produce desired characteristics or to identify undesirable characteristics.

[0081] The term "property" or grammatical equivalents thereof in the context of a polypeptide (including proteins), as used herein, refer to any characteristic or attribute of a polypeptide that can be selected or detected. These properties include, but are not limited to oxidative stability, substrate specificity, catalytic activity, thermal stability, alkaline stability, pH activity profile, resistance to proteolytic degradation, K_M , k_{cat} , k_{cat}/K_M ratio, protein folding, inducing an immune response, ability to bind to a ligand, ability to bind to a receptor, ability to be secreted, ability to be displayed on the surface of a cell, ability to oligomerize, ability to signal, ability to stimulate cell proliferation, ability to inhibit cell proliferation, ability to induce apoptosis, ability to be modified by phosphorylation or glycosylation, and/or ability to treat disease, etc.

[0082] The terms "modified sequence" and "modified genes" are used interchangeably herein to refer to a sequence that includes a deletion, insertion or interruption of naturally occurring nucleic acid sequence. In some preferred embodiments, the expression product of the modified sequence is a truncated protein (e.g., if the modification is a deletion or interruption of the sequence). In some particularly preferred embodiments, the truncated protein retains biological activity. In alternative embodiments, the expression product of the modified sequence is an elongated protein (e.g., modifications comprising an insertion into the nucleic acid sequence). In some embodiments, an insertion leads to a truncated protein (e.g., when the insertion results in the formation of a stop codon). Thus, an insertion may result in either a truncated protein or an elongated protein as an expression product.

[0083] As used herein, the terms "mutant sequence" and "mutant gene" are used interchangeably and refer to a sequence that has an alteration in at least one codon occurring in a host cell's wild-type sequence. The expression product of the mutant sequence is a protein with an altered amino acid sequence relative to the wild-type. The expression product may have an altered functional capacity (e.g., enhanced enzymatic activity).

[0084] The terms "mutagenic primer" or "mutagenic oligonucleotide" (used interchangeably herein) are intended to refer to oligonucleotide compositions which correspond to a portion of the template sequence and which are capable of hybridizing thereto. With respect to mutagenic primers, the primer will not precisely match the template nucleic acid, the mismatch or mismatches in the primer being used to introduce the desired mutation into the nucleic acid library. As used herein, "non-mutagenic primer" or "non-mutagenic oligonucleotide" refers to oligonucleotide compositions which will match precisely to the template nucleic acid. In one embodiment of the invention, only mutagenic primers are used. In another preferred embodiment of the invention, the primers are designed so that for at least one region at which a mutagenic primer has been included, there is also non-mutagenic primer included in the oligonucleotide mixture. By adding a mixture of mutagenic primers and non-mutagenic primers corresponding to at least one of the mutagenic primers, it is possible to produce a resulting nucleic acid library in which a variety of combinatorial mutational patterns are presented. For example, if it is desired that some of the members of the mutant nucleic acid library retain their precursor sequence at certain positions while other members are mutant at such sites, the non-mutagenic primers provide the ability to obtain a specific level of non-mutant members within the nucleic acid library for a given residue. The methods of the invention employ mutagenic and non-mutagenic oligonucleotides which are generally between 10-50 bases in length, more preferably about 15-45 bases in length. However, it may be necessary to use primers that are either shorter than 10 bases or longer than 50 bases to obtain the mutagenesis result desired. With respect to corresponding mutagenic and non-mutagenic primers, it is not necessary that the corresponding oligonucleotides be of identical length, but only that there is overlap in the region corresponding to the mutation to be added. Primers may be added in a pre-defined ratio according to the present invention. For example, if it is desired that the resulting library have a significant level of a certain specific mutation and a lesser amount of a different mutation at the same or different site, by adjusting the amount of primer added, it is possible to produce the desired biased library. Alternatively, by adding lesser or greater amounts of non-mutagenic primers, it is possible to adjust the frequency with which the corresponding mutation(s) are produced in the mutant nucleic acid library.

[0085] As used herein, the phrase "contiguous mutations" refers to mutations which are presented within the same oligonucleotide primer. For example, contiguous mutations may be adjacent or nearby each other, however, they will be introduced into the resulting mutant template nucleic acids by the same primer.

[0086] The terms "wild-type sequence," or "wild-type gene" are used interchangeably herein, to refer to a sequence that is native or naturally occurring in a host cell. In some embodiments, the wild-type sequence refers to a sequence of interest that is the starting point of a protein engineering project (i.e., the "parent" sequence). The wild-type sequence may encode either a homologous or heterologous protein. A homologous protein is one the host cell would produce without intervention. A heterologous protein is one that the host cell would not produce but for the intervention.

[0087] As used herein, the terms "protease variant," "subtilisin variant," "subtilisin protease variant," are used in reference to

proteases that are similar to a wild-type subtilisin or a parent protease (e.g., subtilisin) used as a starting point in protein engineering. These variants are similar to the wild-type or other parent in their function, but have mutations in their amino acid sequence that make them different in sequence from the wild-type or parent protease (e.g., subtilisin).

[0088] Unless otherwise indicated, the amino acid position numbers refer to those assigned to the mature *Bacillus amyloliquefaciens* subtilisin sequence of SEQ ID NO:2.

[0089] As used herein, the terms "modification" and "mutation" refers to any change or alteration in an amino acid sequence. It is intended that the term encompass substitutions, deletions, insertions, and/or replacement of amino acid side chains in an amino acid sequence of interest (e.g., a subtilisin sequence). It is also intended that the term encompass chemical modification of an amino acid sequence of interest (e.g., a subtilisin sequence).

[0090] The term "oxidation stable" refers to proteases of the present invention that retain a specified amount of enzymatic activity over a given period of time under conditions prevailing during the proteolytic, hydrolyzing, cleaning or other process of the invention, for example while exposed to or contacted with bleaching agents or oxidizing agents. In some embodiments, the proteases retain at least about 50%, about 60%, about 70%, about 75%, about 80%, about 85%, about 90%, about 92%, about 95%, about 96%, about 97%, about 98%, or about 99% proteolytic activity after contact with a bleaching or oxidizing agent over a given time period, for example, at least about 1 minute, about 3 minutes, about 5 minutes, about 8 minutes, about 12 minutes, about 16 minutes, about 20 minutes, etc. In some embodiments, the stability is measured as described in the Examples.

[0091] The term "chelator stable" refers to proteases of the present invention that retain a specified amount of enzymatic activity over a given period of time under conditions prevailing during the proteolytic, hydrolyzing, cleaning or other process of the invention, for example while exposed to or contacted with chelating agents. In some embodiments, the proteases retain at least about 50%, about 60%, about 70%, about 75%, about 80%, about 85%, about 90%, about 92%, about 95%, about 96%, about 97%, about 98%, or about 99% proteolytic activity after contact with a chelating agent over a given time period, for example, at least about 10 minutes, about 20 minutes, about 40 minutes, about 60 minutes, about 100 minutes, etc. In some embodiments, the chelator stability is measured as described in the Examples.

[0092] The terms "thermally stable" and "thermostable" refer to proteases of the present invention that retain a specified amount of enzymatic activity after exposure to identified temperatures over a given period of time under conditions prevailing during the proteolytic, hydrolyzing, cleaning or other process of the invention, for example while exposed altered temperatures. Altered temperatures includes increased or decreased temperatures. In some embodiments, the proteases retain at least about 50%, about 60%, about 70%, about 75%, about 80%, about 85%, about 90%, about 92%, about 95%, about 96%, about 97%, about 98%, or about 99% proteolytic activity after exposure to altered temperatures over a given time period, for example, at least about 60 minutes, about 120 minutes, about 180 minutes, about 240 minutes, about 300 minutes, etc. In some embodiments, the thermostability is determined as described in the Examples.

[0093] The term "enhanced stability" in the context of an oxidation, chelator, thermal and/or pH stable protease refers to a higher retained proteolytic activity over time as compared to other serine proteases (e.g., subtilisin proteases) and/or wild-type enzymes.

[0094] The term "diminished stability" in the context of an oxidation, chelator, thermal and/or pH stable protease refers to a lower retained proteolytic activity over time as compared to other serine proteases (e.g., subtilisin proteases) and/or wild-type enzymes.

[0095] The term "cleaning activity" refers to the cleaning performance achieved by the protease under conditions prevailing during the proteolytic, hydrolyzing, cleaning or other process of the invention. In some embodiments, cleaning performance is determined by the application of various cleaning assays concerning enzyme sensitive stains, for example grass, blood, milk, or egg protein as determined by various chromatographic, spectrophotometric or other quantitative methodologies after subsection of the stains to standard wash conditions. Exemplary assays include, but are not limited to those described in WO 99/34011, and U.S. Pat. 6,605,458, as well as those methods included in the Examples.

[0096] The term "cleaning effective amount" of a protease refers to the quantity of protease described hereinbefore that achieves a desired level of enzymatic activity in a specific cleaning composition. Such effective amounts are readily ascertained by one of ordinary skill in the art and are based on many factors, such as the particular protease used, the cleaning application, the specific composition of the cleaning composition, and whether a liquid or dry (e.g., granular, bar) composition is required, etc.

[0097] The term "cleaning adjunct materials," as used herein, means any liquid, solid or gaseous material selected for the particular type of cleaning composition desired and the form of the product (e.g., liquid, granule, powder, bar, paste, spray, tablet, gel; or foam composition), which materials are also preferably compatible with the protease enzyme used in the composition. In

some embodiments, granular compositions are in "compact" form, while in other embodiments, the liquid compositions are in a "concentrated" form.

[0098] The term "enhanced performance" in the context of cleaning activity refers to an increased or greater cleaning activity of certain enzyme sensitive stains such as egg, milk, grass or blood, as determined by usual evaluation after a standard wash cycle and/or multiple wash cycles.

[0099] The term "diminished performance" in the context of cleaning activity refers to an decreased or lesser cleaning activity of certain enzyme sensitive stains such as egg, milk, grass or blood, as determined by usual evaluation after a standard wash cycle.

[0100] The term "comparative performance" in the context of cleaning activity refers to at least about 60%, at least about 70%, at least about 80% at least about 90%, or at least about 95% of the cleaning activity of a comparative subtilisin protease (*e.g.*, commercially available proteases), including but not limited to OPTIMASE™ protease (Genencor), PURAFECT™ protease products (Genencor), SAVINASE™ protease (Novozymes), BPN'-variants (*See e.g.*, U.S. Pat. No. Re 34,606), RELASE™, DURAZYME™, EVERLASE™, KANNASE™ protease (Novozymes), MAXACAL™, MAXAPEM™, PROPERASE™ proteases (Genencor; *See also*, U.S. Pat. No. Re 34,606, and U.S. Pat. Nos. 5,700,676; 5,955,340; 6,312,936; and 6,482,628), and *B. lentus* variant protease products (*e.g.*, those described in WO 92/21760, WO 95/23221 and/or WO 97/07770).

[0101] Cleaning performance can be determined by comparing the proteases of the present invention with those subtilisin proteases in various cleaning assays concerning enzyme sensitive stains such as grass, blood or milk as determined by usual spectrophotometric or analytical methodologies after standard wash cycle conditions.

[0102] As used herein, "cleaning compositions" and "cleaning formulations" refer to compositions that find use in the removal of undesired compounds from items to be cleaned, such as fabric, dishes, contact lenses, other solid substrates, hair (shampoos), skin (soaps and creams), teeth (mouthwashes, toothpastes) etc. The term encompasses any materials/compounds selected for the particular type of cleaning composition desired and the form of the product (*e.g.*, liquid, gel, granule, or spray composition), as long as the composition is compatible with the perhydrolase and other enzyme(s) used in the composition. The specific selection of cleaning composition materials are readily made by considering the surface, item or fabric to be cleaned, and the desired form of the composition for the cleaning conditions during use.

[0103] The terms further refer to any composition that is suited for cleaning, bleaching, disinfecting, and/or sterilizing any object and/or surface. It is intended that the terms include, but are not limited to detergent compositions (*e.g.*, liquid and/or solid laundry detergents and fine fabric detergents; hard surface cleaning formulations, such as for glass, wood, ceramic and metal counter tops and windows; carpet cleaners; oven cleaners; fabric fresheners; fabric softeners; and textile and laundry pre-spotters, as well as dish detergents).

[0104] Indeed, the term "cleaning composition" as used herein, includes unless otherwise indicated, granular or powder-form all-purpose or heavy-duty washing agents, especially cleaning detergents; liquid, gel or paste-form all-purpose washing agents, especially the so-called heavy-duty liquid (HDL) types; liquid fine-fabric detergents; hand dishwashing agents or light duty dishwashing agents, especially those of the high-foaming type; machine dishwashing agents, including the various tablet, granular, liquid and rinse-aid types for household and institutional use; liquid cleaning and disinfecting agents, including antibacterial hand-wash types, cleaning bars, mouthwashes, denture cleaners, car or carpet shampoos, bathroom cleaners; hair shampoos and hair-rinses; shower gels and foam baths and metal cleaners; as well as cleaning auxiliaries such as bleach additives and "stain-stick" or pre-treat types.

[0105] As used herein, the terms "detergent composition" and "detergent formulation" are used in reference to mixtures which are intended for use in a wash medium for the cleaning of soiled objects. In some preferred embodiments, the term is used in reference to laundering fabrics and/or garments (*e.g.*, "laundry detergents"). In alternative embodiments, the term refers to other detergents, such as those used to clean dishes, cutlery, etc. (*e.g.*, "dishwashing detergents"). It is not intended that the present invention be limited to any particular detergent formulation or composition. Indeed, it is intended that in addition to perhydrolase, the term encompasses detergents that contain surfactants, transferase(s), hydrolytic enzymes, oxido reductases, builders, bleaching agents, bleach activators, bluing agents and fluorescent dyes, caking inhibitors, masking agents, enzyme activators, antioxidants, and solubilizers.

[0106] As used herein, "fabric cleaning compositions" include hand and machine laundry detergent compositions including laundry additive compositions and compositions suitable for use in the soaking and/or pretreatment of stained fabrics (*e.g.*, clothes, linens, and other textile materials).

[0107] As used herein, "non-fabric cleaning compositions" include non-textile (*i.e.*, fabric) surface cleaning compositions, including but not limited to dishwashing detergent compositions, oral cleaning compositions, denture cleaning compositions, and personal cleansing compositions.

[0108] The "compact" form of the cleaning compositions herein is best reflected by density and, in terms of composition, by the amount of inorganic filler salt. Inorganic filler salts are conventional ingredients of detergent compositions in powder form. In conventional detergent compositions, the filler salts are present in substantial amounts, typically about 17 to about 35% by weight of the total composition. In contrast, in compact compositions, the filler salt is present in amounts not exceeding about 15% of the total composition. In some embodiments, the filler salt is present in amounts that do not exceed about 10%, or more preferably, about 5%, by weight of the composition. In some embodiments, the inorganic filler salts are selected from the alkali and alkaline-earth-metal salts of sulfates and chlorides. A preferred filler salt is sodium sulfate.

[0109] As used herein, the term "surfactant" refers to any compound generally recognized in the art as having surface active qualities. Surfactants generally include anionic, cationic, nonionic, and zwitterionic compounds, which are further described, herein.

EXPERIMENTAL

[0110] The present invention is described in further detail in the following Examples which are not in any way intended to limit the scope of the invention as claimed. The attached Figures are meant to be considered as integral parts of the specification and description of the invention. The following Examples are offered to illustrate, but not to limit the claimed invention

[0111] In the experimental disclosure which follows, the following abbreviations apply: PI or Pi (Performance Index), ppm (parts per million); M (molar); mM (millimolar); μ M (micromolar); nM (nanomolar); mol (moles); mmol (millimoles); μ mol (micromoles); nmol (nanomoles); gm (grams); mg (milligrams); μ g (micrograms); pg (picograms); L (liters); ml and mL (milliliters); μ l and μ L (microliters); cm (centimeters); mm (millimeters); μ m (micrometers); nm (nanometers); U (units); V (volts); MW (molecular weight); sec (seconds); min(s) (minute/minutes); h(s) and hr(s) (hour/hours); °C. (degrees Centigrade); QS (quantity sufficient); ND (not done); NA (not applicable); rpm (revolutions per minute); H₂O (water); dH₂O (deionized water); HCl (hydrochloric acid); aa (amino acid); bp (base pair); kb (kilobase pair); kD (kilodaltons); cDNA (copy or complementary DNA); DNA (deoxyribonucleic acid); ssDNA (single stranded DNA); dsDNA (double stranded DNA); dNTP (deoxyribonucleotide triphosphate); RNA (ribonucleic acid); MgCl₂ (magnesium chloride); NaCl (sodium chloride); w/v (weight to volume); v/v (volume to volume); g (gravity); OD (optical density); Dulbecco's phosphate buffered solution (DPBS); SOC (2% Bacto-Tryptone, 0.5% Bacto Yeast Extract, 10 mM NaCl, 2.5 mM KCl); Terrific Broth (TB; 12 g/l Bacto Tryptone, 24 g/l glycerol, 2.31 g/l KH₂PO₄, and 12.54 g/l K₂HPO₄); OD₂₈₀ (optical density at 280 nm); OD₆₀₀ (optical density at 600 nm); A₄₀₅ (absorbance at 405 nm); V_{max} (the maximum initial velocity of an enzyme catalyzed reaction); PAGE (polyacrylamide gel electrophoresis); PBS (phosphate buffered saline [150 mM NaCl, 10 mM sodium phosphate buffer, pH 7.2]); PBST (PBS+0.25% TWEEN® 20); PEG (polyethylene glycol); PCR (polymerase chain reaction); RT-PCR (reverse transcription PCR); SDS (sodium dodecyl sulfate); Tris (tris(hydroxymethyl)aminomethane); HEPES (N-[2-Hydroxyethyl]piperazine-N-[2-ethanesulfonic acid]); HBS (HEPES buffered saline); Tris-HCl (tris[Hydroxymethyl]aminomethane-hydrochloride); Tricine (N-[tris-(hydroxymethyl)-methyl]-glycine); CHES (2-(N-cyclo-hexylamino) ethane-sulfonic acid); TAPS (3-[[tris-(hydroxymethyl)-methyl]-amino]-propanesulfonic acid); CAPS (3-(cyclo-hexylamino)-propane-sulfonic acid); DMSO (dimethyl sulfoxide); DTT (1,4-dithio-DL-threitol); SA (sinapinic acid (s,5-dimethoxy-4-hydroxy cinnamic acid); TCA (trichloroacetic acid); Glut and GSH (reduced glutathione); GSSG (oxidized glutathione); TCEP (Tris[2-carboxyethyl] phosphine); Ci (Curies); mCi (milliCuries); μ Ci (microCuries); HPLC (high pressure liquid chromatography); RP-HPLC (reverse phase high pressure liquid chromatography); TLC (thin layer chromatography); MALDI-TOF (matrix-assisted laser desorption/ionization--time of flight); Ts (tosyl); Bn (benzyl); Ph (phenyl); Ms (mesyl); Et (ethyl), Me (methyl); *Taq* (*Thermus aquaticus* DNA polymerase); Klenow (DNA polymerase I large (Klenow) fragment); EGTA (ethylene glycol-bis(β -aminoethyl ether) N, N, N', N'-tetraacetic acid); EDTA (ethylenediaminetetracetic acid); bla (β -lactamase or ampicillin-resistance gene); HDL (high density liquid); MJ Research (MJ Research, Reno, NV); Baseclear (Baseclear BV, Inc., Leiden, the Netherlands); PerSeptive (PerSeptive Biosystems, Framingham, MA); ThermoFinnigan (ThermoFinnigan, San Jose, CA); Argo (Argo BioAnalytica, Morris Plains, NJ); Seitz EKS (SeitzSchenk Filtersystems GmbH, Bad Kreuznach, Germany); Pall (Pall Corp., East Hills, NY); Spectrum (Spectrum Laboratories, Dominguez Rancho, CA); Molecular Structure (Molecular Structure Corp., Woodlands, TX); Accelrys (Accelrys, Inc., San Diego, CA); Chemical Computing (Chemical Computing Corp., Montreal, Canada); New Brunswick (New Brunswick Scientific, Co., Edison, NJ); CFT (Center for Test Materials, Vlaardingen, the Netherlands); Test Fabrics (Test Fabrics, Inc., West Pittston, PA); Procter & Gamble (Procter & Gamble, Inc., Cincinnati, OH); Epicentre (Epicentre Biotechnologies, Madison, WI); GE Healthcare (GE Healthcare, Chalfont St. Giles, United Kingdom); OXOID (Oxoid, Basingstoke, Hampshire, UK);

Megazyme (Megazyme International Ireland Ltd., Bray Business Park, Bray, Co., Wicklow, Ireland); Finnzymes (Finnzymes Oy, Espoo, Finland); Kelco (CP Kelco, Wilmington, DE); Corning (Corning Life Sciences, Corning, NY); (NEN (NEN Life Science Products, Boston, MA); Pharma AS (Pharma AS, Oslo, Norway); Dynal (Dynal, Oslo, Norway); Bio-Synthesis (Bio-Synthesis, Lewisville, TX); ATCC (American Type Culture Collection, Rockville, MD); Gibco/BRL (Gibco/BRL, Grand Island, NY); Sigma (Sigma Chemical Co., St. Louis, MO); Pharmacia (Pharmacia Biotech, Piscataway, NJ); NCBI (National Center for Biotechnology Information); Applied Biosystems (Applied Biosystems, Foster City, CA); BD Biosciences and/or Clontech (BD Biosciences CLONTECH Laboratories, Palo Alto, CA); Operon Technologies (Operon Technologies, Inc., Alameda, CA); MWG Biotech (MWG Biotech, High Point, NC); Oligos Etc (Oligos Etc. Inc, Wilsonville, OR); Bachem (Bachem Bioscience, Inc., King of Prussia, PA); Difco (Difco Laboratories, Detroit, MI); Mediatech (Mediatech, Herndon, VA); Santa Cruz (Santa Cruz Biotechnology, Inc., Santa Cruz, CA); Oxoid (Oxoid Inc., Ogdensburg, NY); Worthington (Worthington Biochemical Corp., Freehold, NJ); GIBCO BRL or Gibco BRL (Life Technologies, Inc., Gaithersburg, MD); Millipore (Millipore, Billerica, MA); Bio-Rad (Bio-Rad, Hercules, CA); Invitrogen (Invitrogen Corp., San Diego, CA); New England Biolabs and NEB (New England Biolabs, Ipswich, MA); Sigma (Sigma Chemical Co., St. Louis, MO); Pierce (Pierce Biotechnology, Rockford, IL); Takara (Takara Bio Inc. Otsu, Japan); Roche (Hoffmann-La Roche, Basel, Switzerland); EM Science (EM Science, Gibbstown, NJ); Qiagen (Qiagen, Inc., Valencia, CA); Biodesign (Biodesign Intl., Saco, Maine); Aptagen (Aptagen, Inc., Herndon, VA); Sorvall (Sorvall brand, from Kendro Laboratory Products, Asheville, NC); United States Testing (United States Testing Co., Hoboken, NJ); Molecular Devices (Molecular Devices, Corp., Sunnyvale, CA); R&D Systems (R&D Systems, Minneapolis, MN); Stratagene (Stratagene Cloning Systems, La Jolla, CA); Marsh (Marsh Biosciences, Rochester, NY); Geneart (Geneart GmbH, Regensburg, Germany); DNA2.0 (DNA2.0, Menlo Park, CA); Gene Oracle (Gene Oracle, Mountain View, CA); Zymo Research (Zymo Research, Orange, CA); Bio-Tek (Bio-Tek Instruments, Winooski, VT); Biacore (Biacore, Inc., Piscataway, NJ); PeproTech (PeproTech, Rocky Hill, NJ); SynPep (SynPep, Dublin, CA); New Objective (New Objective brand; Scientific Instrument Services, Inc., Ringoes, NJ); Waters (Waters, Inc., Milford, MA); Matrix Science (Matrix Science, Boston, MA); Dionex (Dionex, Corp., Sunnyvale, CA); Monsanto (Monsanto Co., St. Louis, MO); Wintershall (Wintershall AG, Kassel, Germany); BASF (BASF Co., Florham Park, NJ); Huntsman (Huntsman Petrochemical Corp., Salt Lake City, UT); Enichem (Enichem Iberica, Barcelona, Spain); Fluka Chemie AG (Fluka Chemie AG, Buchs, Switzerland); Gist-Brocades (Gist-Brocades, NV, Delft, the Netherlands); Dow Corning (Dow Corning Corp., Midland, MI); and Microsoft (Microsoft, Inc., Redmond, WA).

EXAMPLE 1

Assays

[0112] In the following Examples, various assays were used as set forth below for ease in reading. Any deviations from the protocols provided below are indicated in the Examples. In the present invention, the subtilisin numbering corresponds to that of the mature BPN' sequence, as set forth below:

AQSVPYGVSGIKAPALHSQGYTGSNVKAVIDSGIDSSHPDLKVAGGASMVPSETNPFQDNN
SHGTHVAGTVAALNNSIGVLGVAPSASLYAVKVLGADGSGQYSWIINGIEWAIAANNMDVIN
MSLGGPSGSAALKAADVKAASGVVVVAAAGNEGTSAGSSSTVGYPGKYPSVIAVGAVDSSN
QRASFSSVGPGLDVMAPGVSIQSILPGNKYGAYNGTSMASPIVAGAAALILSKIPNW'NTQ
VRSSLENTTITKLGDSTFYTGKGLINVQAAAQ (SEQ ID NO:1)

A. TCA Assay for Protein Content Determination in 96-well Microtiter Plates

[0113] For BPN' (e.g., reference protease) and BPN' variants, this assay was started using filtered culture supernatant from microliter plates grown 3-4 days at 33 °C with shaking at 230 rpm and humidified aeration. A fresh 96-well flat bottom microtiter plate (MTP) was used for the assay. First, 100 µL/well of 0.25 N HCl was placed in each well. Then, 50 µL of filtered culture broth was added. The light scattering/absorbance at 405 nm (use 5 sec mixing mode in the plate reader) was then determined, in order to provide the "blank" reading. For the test, 100 µL/well of 15% (w/v) trichloroacetic acid (TCA) was placed in the plates and incubated between 5 and 30 min at room temperature. The light scattering/absorbance at 405 nm (use 5 sec mixing mode in the plate reader) was then determined.

[0114] The equipment used was a Biomek FX Robot (Beckman Coulter) and a SpectraMAX (type 340; Molecular Devices) MTP Reader; the MTP's were from Costar (type 9017). The equipment used was a Biomek FX Robot (Beckman Coulter) and a SpectraMAX type 340 (Molecular Devices) MTP Reader; and the MTPs were type 9017 (Costar).

[0115] The calculations were performed by subtracting the blank (no TCA) from the test reading with TCA to provide a relative measure of the protein content in the samples. If desired, a standard curve can be created by calibrating the TCA readings with AAPF assays of clones with known conversion factors. However, the TCA results are linear with respect to protein concentration from 50 to 500 protein per ml (ppm) and can thus be plotted directly against enzyme performance for the purpose of choosing good-performing variants. The turbidity/light scatter increase in the samples correlates to the total amount of precipitable protein in the culture supernatant.

B. AAPF Protease Assay in 96-well Microtiter Plates

[0116] In order to determine the protease activity of the proteases and variants thereof of the present invention, the hydrolysis of N-succinyl-L-alanyl-L-alanyl-L-prolyl-L-phenyl-p-nitroanilide (suc-AAPF-pNA) was measured. The reagent solutions used were: 100 mM Tris/HCl, pH 8.6, containing 0.005% TWEEN®-80 (Tris dilution buffer); 100 mM Tris buffer, pH 8.6, containing 10 mM CaCl₂ and 0.005% TWEEN®-80 (Tris/Ca buffer); and 160 mM suc-AAPF-pNA in DMSO (suc-AAPF-pNA stock solution) (Sigma: S-7388). To prepare a suc-AAPF-pNA working solution, 1 ml suc-AAPF-pNA stock solution was added to 100 ml Tris/Ca buffer and mixed well for at least 10 seconds. The assay was performed by adding 10 µl of diluted protease solution to each well, immediately followed by the addition of 190 µl 1 mg/ml suc-AAPF-pNA working solution. The solutions were mixed for 5 sec., and the absorbance change in kinetic mode (20 readings in 5 minutes) was read at 410 nm in an MTP reader, at 25°C. The protease activity was expressed as AU (activity = $\Delta\text{OD} \cdot \text{min}^{-1} \text{ ml}^{-1}$).

C. Surfactant and Chelant Stability Assays

[0117] LAS and LAS/EDTA stability was measured after incubation of the test protease in the presence of LAS and LAS/EDTA respectively, as a function of residual activity determined using the AAPF assay.

LAS Stability Method

Reagents:

[0118]

Dodecylbenzenesulfonate, Sodium salt (=LAS): Sigma D-2525

TWEEN®-80: Sigma P-8074

TRIS buffer (free acid): Sigma T-1378); 6.35 g is dissolved in about 960 ml water; pH is adjusted to 8.2 with 4N HCl. Final concentration of TRIS is 52.5 mM.

LAS stock solution: Prepare a 10.5 % LAS solution in MQ water (=10.5 g per 100 ml MQ)

TRIS buffer-100 mM / pH 8.6 (100mM Tris/0.005% Tween®-80)

TRIS-Ca buffer, pH 8.6 (100mM Tris/10mM CaCl₂/0.005% Tween-®80)

Hardware:

[0119]

Flat bottom MTPs (Costar No. 9017)

Biomek FX

ASYS Multipipettor

Spectramax MTP Reader

iEMS Incubator/Shaker

Innova 4330 Incubator/Shaker

Biohit multichannel pipette

BMG ThermoStar Shaker

[0120] A 0.063% LAS solution was prepared in 52.5 mM Tris buffer pH 8.2. The suc-AAPF-pNA working solution was prepared by adding 1 ml of 100 mg/ml suc-AAPF-pNA stock solution (in DMSO) to 100 ml (100 mM) TRIS buffer, pH 8.6. To dilute the supernatants, flat-bottomed plates were filled with dilution buffer and an aliquot of the supernatant was added and mixed well. The dilution ratio depended on the concentration of the protease controls in the growth plates (AAPF activity). The desired protein concentration was 80 ppm.

[0121] Ten µl of the diluted supernatant were added to 190 µl 0.063% LAS buffer/well. The MTP was covered with tape, shaken for a few seconds and placed in an incubator (Innova 4230) at 45°C for for 60 minutes at 200 rpm agitation. The initial activity ($t=10$ minutes) was determined after 10 minutes of incubation by transferring 10 µl of the mixture in each well to a fresh MTP containing 190µl suc-AAPF-pNA work solution. These solutions were mixed well and the AAPF activity was measured using a MTP Reader (20 readings in 5 minutes and 25°C).

[0122] The final activity ($t=60$ minutes) was determined by removing another 10 µl of solution from the incubating plate after 60 minutes of incubation. The AAPF activity was then determined as described above. The stability of the samples was determined by calculating the ration of the residual and initial AAPF activity as follows:

Residual Activity (%) = $[t=60 \text{ value}] * 100 / [t=10 \text{ value}]$

LAS/EDTA Stability Method

[0123] The stability of protease variants in the presence of a representative anionic surfactant (LAS=linear alkylbene sulfonate, sodium dodecylbenzenesulfonate-DOBS) and di-sodium EDTA was measured after incubation under defined conditions and the residual activity was determined using the AAPF assay. The reagents used were dodecylbenzene sulfonate, sodium salt (DOBS, Sigma No. D-2525), TWEEN®-80 (Sigma No. P-8074), di-sodium EDTA (Siegfried Handel No. 164599-02), HEPES (Sigma No. H-7523), unstress buffer: 50 mM HEPES (11.9 g/l) + 0.005% TWEEN®-80, pH 8.0, Stress buffer: 50 mM HEPES (11.9 g/l), 0.1% (w/v) DOBS (1 g/l), 10 mM EDTA (3.36 g/l), pH 8.0, reference protease and protease variant culture supernatants, containing 200 - 400 µg/ml protein. The equipment used was V- or U-bottom MTP as dilution plates (Greiner 651101 and 650161 respectively), F-bottom MTP (Corning 9017) for unstress and LAS/EDTA buffer as well as for suc-AAPF-pNA plates, Biomek FX (Beckman Coulter), Spectramax Plus 384 MTP Reader (Molecular Devices), iEMS Incubator/Shaker (1 mm amplitude) from Thermo Electron Corporation, sealing tape: Nunc (236366)

[0124] The iEMS incubator/shaker (Thermo/Labsystems) was set at 29°C. Culture supernatants were diluted into plates containing unstress buffer to a concentration of ~ 25 ppm (master dilution plate). 20 µl of sample from the master dilution plate was added to plates containing 180 µl unstress buffer to give a final incubation concentration of 2.5 ppm. The contents were mixed and kept at room temperature and a AAPF assay was performed on this plate. 20 µl of sample from the master dilution plate was also added to plates containing 180 µl stress buffer (50 mM HEPES (11.9 g/l), 0.1% (w/v) DOBS (1 g/l), 10 mM EDTA (3.36 g/l), pH 8.0). The solutions were mixed and immediately placed in 29°C iEMS shaker for 30 min at 400 rpm. Following 30 minutes of incubation, a AAPF assay was performed on the stress plate. The stability of the samples was determined by calculating the ration of the residual and initial AAPF activity as follows: Residual Activity (%) = $[mOD.min-1 \text{ stressed}] * 100 / [mOD.min-1 \text{ unstressed}]$.

D. Cleaning Performance Assays

[0125] The stain removal performance of the protease variants was determined in commercially available detergents. Heat inactivation of commercial detergent formulas serves to destroy the enzymatic activity of any protein components while retaining the properties of non-enzymatic components. Thus this method was suitable for preparing commercially purchased detergents for use in testing the enzyme variants of the present invention.

Microswatches:

[0126] Microswatches of ¼" circular diameter were obtained from CFT. Single Microswatches or two Microswatches were placed vertically in each well of a 96-well MTP to expose the whole surface area (*i.e.*, not flat on the bottom of the well).

BMI Microswatch Assay

[0127] Microswatches containing blood milk and ink (BMI) of 0.25 inch circular diameter were obtained from CFT. Before cutting of the swatches, the fabric (EMPA 116) was washed with water. One microswatch was vertically placed in each well of a 96-well microliter plate in order to expose the whole surface area (*i.e.*, not flat on the bottom of the well). The desired detergent solution was prepared as described herein. After equilibrating the Thermomixer at 25°C, 190 µl of detergent solution was added to each well of the MTP, containing microswatches. To this mixture, 10 µl of the diluted enzyme solution was added so that the final enzyme concentration was 1 µg/ml (determined from BCA assay). The MTP was sealed with tape and placed in the incubator for 30 minutes, with agitation at 1400 rpm. Following incubation under the appropriate conditions, 100 µl of the solution from each well was transferred into a fresh MTP. The new MTP containing 100 µl of solution/well was read at 405 nm using a MTP SpectraMax reader. Blank controls, as well as a control containing two Microswatches and detergent but no enzyme were also included.

"Pre-washed" Swatch

[0128] This type of microswatch was pre-washed in deionised water for 20 minutes at ambient temperature. After the pre-washing step, the swatches were put on top of paper towels to dry. The air-dried swatches were then punched using a ¼" circular die on an expulsion press. Finally two microswatches were put into each well of a 96-well MTP vertically to expose the whole surface area (*i.e.* not flat on the bottom of the well).

Detergents

[0129] For North American (NA) and Western European (WE) heavy duty liquid laundry (HDL) detergents, heat inactivation was performed by placing pre-weighed liquid detergent (in a glass bottle) in a water bath at 95°C for 2 hours. The detergents were purchased from local supermarket stores. Both un-heated and heated detergents were assayed within 5 minutes of dissolving the detergent to accurately determine percentage deactivated. Enzyme activity was tested by the AAPF assay.

[0130] For testing of enzyme activity in heat-inactivated detergents, working solutions of detergents were made from the heat inactivated stocks. Appropriate amounts of water hardness (6 gpg or 12 gpg) and buffer were added to the detergent solutions to match the desired conditions. The solutions were mixed by vortexing or inverting the bottles.

Enzymes and Equipment

[0131] Samples of reference serine proteases variants thereof were obtained from filtered culture broth of cultures grown in MTP plates. The equipment used was a Biomek FX Robot (Beckman Coulter), a SpectraMAX MTP Reader (type 340; Molecular Devices), an iEMS incubator/shaker (Thermo/Labsystems); F-bottom MTPs (Costar type 9017 used for reading reaction plates after incubation); and V-bottom MTPs (Greiner 651101 used for pre-dilution of supernatant). In this assay, the proteases hydrolyze the substrate and liberate pigment and insoluble particles from the substrate. Thus the rate of turbidity is a measure of enzyme activity.

[0132] The stain removal performance of reference serine proteases and variants therefrom on microswatches was determined

on a MTP scale in commercially available heat-inactivated detergent. The reagents used were: 5 mM HEPES, pH 8.0 or 5 mM MOPS, pH 7 buffer, 3:1 Ca: Mg for medium water hardness. (CaCl₂: MgCl₂•6H₂O); 15000 grains per gallon (gpg) stock diluted to 6 gpg, 2 BMI (blood/milk/ink) swatches per plate: EMPA-116 BMI cotton swatches processed by CFT: pre-rinsed and punched 2 swatches per well, and heat inactivated TIDE® 2X Cold off-the-shelf detergent in which lack of protease activity was confirmed.

Table 1-2 Working Detergent Solutions

Detergent	Temp (C)	Detergent g/L	pH	Buffer	Gpg	Protease
TIDE® 2X Cold	16	0.98	8	5mM HEPES	6	BPN'
TIDE® 2X Cold	32	0.98	8	5mM HEPES	6	BPN'
TIDE® 2X Cold	16	0.98	7	5mM MOPS	6	BPN'

[0133] The incubator was set at the desired temperature (16°C or 32°C). 10 µL samples from the master dilution plate of ~10 ppm enzyme was added to BMI 2-swatch plates with 190 µL working detergent solutions listed above. The volume was adjusted to give final concentration of 0.5 ppm for variants in the assay plates. The plates were immediately transferred to iEMS incubators and incubated for 30 minutes with 1400 rpm shaking at given temperature. Following incubation, 100 µL of supernatant was transferred into a new 96-well plate and the absorbance was measured in MTP Reader at 405nm and/or 600nm. Control wells, containing 1 or 2 Microswatches and detergent without the addition of protease samples were also included in the test. The measurement at 405 nm provides a higher value and tracks pigment removal, while the measurement at 600 nm tracks turbidity and cleaning.

Calculation of the Stain Removal Activity for All Microswatch Assay Methods:

[0134] The absorbance value obtained was corrected for the blank value (substrate without enzyme), providing a measure of hydrolytic activity. For each sample (variant) the performance index was calculated. The performance index compares the performance of the variant (actual value) and the standard enzyme (theoretical value) at the same protein concentration. In addition, the theoretical values can be calculated, using the parameters of the Langmuir equation of the standard enzyme.

E. Relative Specific Activity of Proteases and Variants Thereof

[0135] In order to discriminate the protease variants, the relative specific activity was calculated using suc-AAPF-pNA as a substrate, which enabled the comparison and ranking of the variants versus the wild-type or standard protease. The specific activity on the suc-AAPF-pNA substrate was determined by dividing the proteolytic activity by the measured TCA-values of each sample, using the assays described above. Using these values, the relative specific activity was calculated (specific activity of variant/specific activity of reference protease).

F. Performance Index

[0136] The performance index compares the performance of the variant (actual value) and the standard or reference protease (theoretical value) at the same protein concentration. In addition, the theoretical values can be calculated, using the parameters of the Langmuir equation of the standard protease. A performance index (PI) that is greater than 1 (PI>1) identifies a better variant as compared to the standard (e.g., wild-type), while a PI of 1 (PI=1) identifies a variant that performs the same as the standard, and a PI that is less than 1 (PI<1) identifies a variant that performs worse than the standard. Thus, the PI identifies winners, as well as variants that are less desirable for use under certain circumstances.

EXAMPLE 2

Targeted ISD (Insertion Substitution Deletion) Library Construction

[0137] A PCR-based method was used to create a library of modified *B. amyloliquefaciens* subtilisin BPN'-Y217L (commercially

available as PURAFECT® PRIME subtilisin) polynucleotides containing in-frame insertions, deletions and/or substitutions in two regions of BPN'-Y217L (positions 63-77 and 92-132 of the mature region) as shown in Figure 1 (See, Pisarchik et al., Prot. Eng. Des. Select., 20:257-265 [2007]). Two sets of oligonucleotides that evenly cover the targeted regions of the BPN'-Y217L gene of a full-length protein of 392 amino acids, in both forward and reverse direction were used to amplify the 5' and 3' segments of the portion of the BPN'-Y217L gene. The coding region of the BPN'-Y217L mature protease contains the *KpnI* and *XhoI* restriction sites for cloning purposes:

gaattcatctcaaaaatgggtctactaaaatatttccatctattataaataattcacagaatagcttttaagtaagctactctgaatttttta
aaaggagagggttaagagtgagaagcaaaaattgtgagagcaaaaattgtgagcatttctgttcttagcgttaattctttacgatggc
gttcggcagcacatccagcgcgcaggctgcagggaatcaaacggggaagaaatatttctcgggttttaacagacaatgagcacatgagc
gcccgttaaggaagaagacgtcatttctgaaaaggcgggaaagtgcaaaagcaattcaaatatctagacgcagctagcgcctacattaaacgaaaa
agccttaaaaagaattgaaaaagaccgcacgtccttacgttgaagaaatcacctagcacacgcgtacgcgcagtcctgtccatattggccta
tcacaaattaaagccctctctctcactctcaaggctacacgggttcaaatgttaaaagtacgggttatcgacagcgggtatcgtattcttctcat
ccagatcttaaaagtcaggcgggagccagcatggttctctgaaacaaatcttccagacaacactctcacggaaacacacgttctgtg
gtacgtttgcgctcttaataactcaatcggtgtattaggcgttgcgcaagcgcatcactttacgtgttaaaagttctcgccgcgacggtt
ccggccaatcacgtggtatcattaaacggaatcgagtggtggcgatgcgaacaaatattggacgttattaaatgagcctcgccggaccgtccg
gttctgctctttaaagcggcagtgataaagccgttgatccggcgtctgtagctgttcggcagcggcaacgaagcactctcggcag
ctcaagcacagtggtgctaccctgttaataacccttctgtcattgtaggtggcgtgtcgacagcagcaaccaagagcatcttctcaage
gtaggacctgagctgagtgatgcacctggcgtatctatccaaagcagcgttcttgaaacaaatagcgcggttgaaacggtacatcaa
tggcatctcgacgttgcggagcggcgttctgattcttcttaagcaccggaactggacaacacactcaagtcgcagctctctagaaaaa
accactcaaaacttgggtgattctctactatggaaaaggctgatcaatgtacagcggcagctcagtaaaactcagataaaaaaccg
gccctggcccccgcgggtttttat (SEQ ID NO:3).

[0138] The amino acid sequence of the BPN'-Y217L precursor protein is provided below. In this sequence, bold indicates the mature BPN'-Y217L protease:

MRSKKLWISLLFALALIFIMAFGSTSSAQAGKSNGEKKYIVGFKQIMSTMSAAKKKDVISE
 KGGKVQKQFKYVDAASATLNEKAVKELKDPVSVAYVEEDHVAHAYAQSVPYGVSVQIKAP
 ALIISQGYTGSNVKVAVIDSGIDSSHIDPKVAGGASMVPSSETNPFQDNNSTIGTIIIVAGTVA
 ALNNSIGVLGVAPSASLYAVKVLGADGSGQYSWIINGIEWAIAINNMMDVINMSLGGPSGS
 AALKAADVKAVASGVVVVAAAGNEGTSOSSSTVGYPGKYPSVIAVGAVDSSNQRAFSFSS
 VGPGLDVMAPGVSIQSTLPGNKYGALNGTSMASPHVAGAAALILSKHPNWTNTQVRSSSL
 ENTTTKLGDSEFYFGKGLINVQAAAQ (SEQ ID NO:4).

[0139] The amino acid sequence of the mature BPN'-Y217L protease was used as the basis for making the variant libraries described herein:

AQSVYPYGVSVQIKAPALHSQGYTGSNVKVAVIDSGIDSSHIDPKVAGGASMVPSSETNPFQD
 NNSIIGTIIIVAGTVAALNNSIGVLGVAPSASLYAVKVLGADGSGQYSWIINGIEWAIAINNM
 DVINMSLGGPSGSAAALKAADVKAVASGVVVVAAAGNEGTSOSSSTVGYPGKYPSVIAVGA
 AVDSSNQRAFSFSSVGPGLDVMAPGVSIQSTLPGNKYGALNGTSMASPHVAGAAALILSK
 HPNWTNTQVRSSLENTTKLGDSEFYFGKGLINVQAAAQ (SEQ ID NO:5).

[0140] Each oligonucleotide contained about 20-24 nucleotides necessary for annealing to the template, targeted codons (with or without degeneracy) and the *Eam1104I* recognition sequence. Every following primer was moved 3 bp downstream until the end of the targeted region was reached. Two PCR reactions (5' and 3' segments) contained either the 5' forward or the 3' reverse gene sequence flanking oligonucleotides each in combination with the corresponding oppositely priming oligonucleotides. The 5' fragments were amplified from pAC-FNA10 plasmid (See, Figure 2A) using a single forward primer (P3203, TGGATCAGTTTGCTGTTTGCT; SEQ ID NO:6) and reverse primers P4385-P4441 (See, Table 2-1) each containing the *Eam1104I* restriction site. The 3' fragments were amplified using a single reverse primer (P3435, ATGTATCAAGATAAGAAAGACAAG; SEQ ID NO:7) and forward primers P4328-P4384 (See, Table 2-1) each containing an *Eam1104I* restriction site.

Table 2-1. Primers Used for BPN'-Y217L Library Construction

SEQ ID NO.	PRIMER NAME	PRIMER SEQUENCE
8	P4328	AAACTCTTCANDTNDTCACGGAACCTCACGTTGCCGGGCACA
9	P4329	AAACTCTTCANDTNDTGGAACCTCACGTTGCCGGGCACAGTT
10	P4330	AAACTCTTCANDTNDTACTCACGTTGCCGGGCACAGTTG
11	P4331	AAACTCTTCANDTNDTCACGTTGCCGGGCACAGTTGCGGCT
12	P4332	AAACTCTTCANDTNDTGTGTCGGGCACAGTTGCGGCTCTT
13	P4333	AAACTCTTCANDTNDTGCCGGGCACAGTTGCGGCTCTTAAT
14	P4334	AAACTCTTCANDTNDTCGGGCACAGTTGCGGCTCTTAATAAC
15	P4335	AAACTCTTCANDTNDTACAGTTGCGGCTCTTAATAACTCA
16	P4336	AAACTCTTCANDTNDTGTGTCGGCTCTTAATAACTCAATC

Table 2-1. Primers Used for BPN'-Y217L Library Construction

SEQ ID NO.	PRIMER NAME	PRIMER SEQUENCE
17	P4337	AAACTCTTCANDTNDTGC GGCTCTTAATAACTCAATCGGT
18	P4338	AAACTCTTCANDTNDTGC TCTTAATAACTCAATCGGTGTA
19	P4339	AAACTCTTCANDTNDCTTAATAACTCAATCGGTGTATTA
20	P4340	AAACTCTTCANDTNDTAATAACTCAATCGGTGTATTAG
21	P4341	AAACTCTTCANDTNDTAACTCAATCGGTGTATTAGGCGTT
22	P4342	AAACTCTTCANDTNDTTCAATCGGTGTATTAGGCGTTG
23	P4343	AAACTCTTCANDTTCAATCGGTGTATTAGGCGTTG
24	P4344	AAACTCTTCANDTNDTAAAGTTCTCGGTGCTGACGGTTC
25	P4345	AAACTCTTCANDTNDTGTCTCGGTGCTGACGGTTC
26	P4346	AAACTCTTCANDTNDTCTCGGTGCTGACGGTTCGGCCAA
27	P4347	AAACTCTTCANDTNDTGGTGCTGACGGTTCGGCCAATAC
28	P4348	AAACTCTTCANDTNDTGCTGACGGTTCGGCCAATACA
29	P4349	AAACTCTTCANDTNDTGACGGTTCGGCCAATACAGCTG
30	P4350	AAACTCTTCANDTNDTGGTTCCGGCCAATACAGCTGGATC
31	P4351	AAACTCTTCANDTNDTTCCGGCCAATACAGCTGGATCATT
32	P4352	AAACTCTTCANDTNDTGCCAATACAGCTGGATCATTAAAC
33	P4353	AAACTCTTCANDTNDTCAATACAGCTGGATCATTAAACGGA
34	P4354	AAACTCTTCANDTNDTTACAGCTGGATCATTAAACGGAATC
35	P4355	AAACTCTTCANDTNDTAGCTGGATCATTAAACGGAATCGAG
36	P4356	AAACTCTTCANDTNDTTGGATCATTAAACGGAATCGAGTG
37	P4357	AAACTCTTCANDTNDTATCATTAAACGGAATCGAGTG
38	P4358	AAACTCTTCANDTNDTATTAACGGAATCGAGTGGGCGATC
39	P4359	AAACTCTTCANDTNDTAACGGAATCGAGTGGGCGATCGCA
40	P4360	AAACTCTTCANDTNDTGGAATCGAGTGGGCGATCGCAAAC
41	P4361	AAACTCTTCANDTNDTATCGAGTGGGCGATCGCAAACAAT
42	P4362	AAACTCTTCANDTNDTGAGTGGGCGATCGCAAACAATATG
43	P4363	AAACTCTTCANDTNDTTGGGCGATCGCAAACAATATGGAC
44	P4364	AAACTCTTCANDTNDTGCGATCGCAAACAATATGGACGTT
45	P4365	AAACTCTTCANDTNDTATCGCAAACAATATGGACGTTATT
46	P4366	AAACTCTTCANDTNDTGCAAACAATATGGACGTTATTAAC
47	P4367	AAACTCTTCANDTNDTAACAATATGGACGTTATTAACATG
48	P4368	AAACTCTTCANDTNDTAATATGGACGTTATTAACATGAG
49	P4369	AAACTCTTCANDTNDTATGGACGTTATTAACATGAGCCTC
50	P4370	AAACTCTTCANDTNDTGACGTTATTAACATGAGCCTC
51	P4371	AAACTCTTCANDTNDTGTTATTAACATGAGCCTCGGCGGA
52	P4372	AAACTCTTCANDTNDTATTAACATGAGCCTCGGCGGACCT
53	P4373	AAACTCTTCANDTNDTAACATGAGCCTCGGCGGACCTTCT
54	P4374	AAACTCTTCANDTNDTATGAGCCTCGGCGGACCTTCTGGT
55	P4375	AAACTCTTCANDTNDTAGCCTCGGCGGACCTTCTGGTTCT
56	P4376	AAACTCTTCANDTNDTCTCGGCGGACCTTCTGGTTCTGCT
57	P4377	AAACTCTTCANDTNDTGCGGACCTTCTGGTTCTGCTGCT
58	P4378	AAACTCTTCANDTNDTGACCTTCTGGTTCTGCTGCTTA

Table 2-1. Primers Used for BPN'-Y217L Library Construction

SEQ ID NO.	PRIMER NAME	PRIMER SEQUENCE
59	P4379	AAACTCTTCANDTNDTCCTTCTGGTTCTGCTGCTTTAAAA
60	P4380	AAACTCTTCANDTNDTTCCTGGTTCTGCTGCTTTAAAAAG
61	P4381	AAACTCTTCANDTNDTGGTTCTGCTGCTTTAAAAAGCGGCA
62	P4382	AAACTCTTCANDTNDTTCCTGCTGCTTTAAAAAGCGGCAGTT
63	P4383	AAACTCTTCANDTNDTGCTGCTTTAAAAAGCGGCAGTTGAT
64	P4384	AAACTCTTCANDTGCTGCTTTAAAAAGCGGCAGTTGAT
65	P4385	AAACTCTTCAAHNGTTGCTTGGAAAGGATTTGTTTC
66	P4386	AAACTCTTCAAHNAHNGTTGCTTGGAAAGGATTTGTTTC
67	P4387	AAACTCTTCAAHNAHNGTTGTTGCTTGGAAAGGATTTGT
68	P4388	AAACTCTTCAAHNAHNAAGTTGTTGCTTGGAAAGGATT
69	P4389	AAACTCTTCAAHNAHNGTGAGAGTTGTTGCTTGGAAAGG
70	P4390	AAACTCTTCAAHNAHNTCCGTGAGAGTTGTTGCTTGGAA
71	P4391	AAACTCTTCAAHNAHNAAGTTCCGTGAGAGTTGTTGCTTG
72	P4392	AAACTCTTCAAHNAHNGTGAGTTCCGTGAGAGTTGTTGTC
73	P4393	AAACTCTTCAAHNAHNAACGTGAGTTCCGTGAGAGTTGTT
74	P4394	AAACTCTTCAAHNAHNGGCAACGTGAGTTCCGTGAGAGTT
75	P4395	AAACTCTTCAAHNAHNGCCGGCAACGTGAGTTCCGTGAGA
76	P4396	AAACTCTTCAAHNAHNTGTGCCGGCAACGTGAGTTCCGTG
77	P4397	AAACTCTTCAAHNAHNAAGTGTGCCGGCAACGTGAGTTCC
78	P4398	AAACTCTTCAAHNAHNCGCAACTGTGCCGGCAACGTGAGT
79	P4399	AAACTCTTCAAHNAHNAAGCCGCAACTGTGCCGGCAACGTG
80	P4400	AAACTCTTCAAHNAHNAAGAGCCGCAACTGTGCCGGCAAC
81	P4401	AAACTCTTCAAHNGTAAAGTGATGCGCTTGGCGCAAC
82	P4402	AAACTCTTCAAHNAHNGTAAAGTGATGCGCTTGGCGCAAC
83	P4403	AAACTCTTCAAHNAHNAAGCGTAAAGTGATGCGCTTG
84	P4404	AAACTCTTCAAHNAHNTACAGCGTAAAGTGATGCGCTTG
85	P4405	AAACTCTTCAAHNAHNTTTTACAGCGTAAAGTGATGCGCT
86	P4406	AAACTCTTCAAHNAHNAAGTTTACAGCGTAAAGTGATG
87	P4407	AAACTCTTCAAHNAHNGAGAACTTTTACAGCGTAAAGTGA
88	P4408	AAACTCTTCAAHNAHNAACCGAGAACTTTTACAGCGTAAAG
89	P4409	AAACTCTTCAAHNAHNAAGCACCAGAACTTTTACAGCGTA
90	P4410	AAACTCTTCAAHNAHNGTCAGCACCAGAACTTTTACAG
91	P4411	AAACTCTTCAAHNAHNAACCGTCAGCACCAGAACTTTTAC
92	P4412	AAACTCTTCAAHNAHNGGAACCGTCAGCACCAGAACTTT
93	P4413	AAACTCTTCAAHNAHNGCCGAACCGTCAGCACCAGAAAC
94	P4414	AAACTCTTCAAHNAHNTTGCCGAACCGTCAGCACCAGAG
95	P4415	AAACTCTTCAAHNAHNGTATTGGCCGAACCGTCAGCAC
96	P4416	AAACTCTTCAAHNAHNGCTGTATTGGCCGAACCGTCAG
97	P4417	AAACTCTTCAAHNAHNCAGCTGTATTGGCCGAACCGTC
98	P4418	AAACTCTTCAAHNAHNGATCCAGCTGTATTGGCCGAAC
99	P4419	AAACTCTTCAAHNAHNAATGATCCAGCTGTATTGGCCGA
100	P4420	AAACTCTTCAAHNAHNGTAAATGATCCAGCTGTATTG

Table 2-1. Primers Used for BPN'-Y217L Library Construction

SEQ ID NO.	PRIMER NAME	PRIMER SEQUENCE
101	P4421	AAACTCTTCAAHNAHNTCCGTTAATGATCCAGCTGTATTG
102	P4422	AAACTCTTCAAHNAHNGATTCCGTTAATGATCCAGCTGTA
103	P4423	AAACTCTTCAAHNAHNCTCGATTCCGTTAATGATCCAGCT
104	P4424	AAACTCTTCAAHNAHNCCACTCGATTCCGTTAATGATCCA
105	P4425	AAACTCTTCAAHNAHNCGCCACTCGATTCCGTTAATGAT
106	P4426	AAACTCTTCAAHNAHNGATCGCCACTCGATTCCGTTAAT
107	P4427	AAACTCTTCAAHNAHNTGCGATCGCCACTCGATTCCGTT
108	P4428	AAACTCTTCAAHNAHNGTTTGCGATCGCCACTCGATTCC
109	P4429	AAACTCTTCAAHNAHNATTGTTTGCGATCGCCACTCGAT
110	P4430	AAACTCTTCAAHNAHNCATATTGTTTGCGATCGCCACTC
111	P4431	AAACTCTTCAAHNAHNGTCCATATTGTTTGCGATCGCCCA
112	P4432	AAACTCTTCAAHNAHNAACGTCCATATTGTTTGCGATCGC
113	P4433	AAACTCTTCAAHNAHNAATAACGTCCATATTGTTTGCGAT
114	P4434	AAACTCTTCAAHNAHNGTTAATAACGTCCATATTGTTTGC
115	P4435	AAACTCTTCAAHNAHNCATGTTAATAACGTCCATATTGTT
116	P4436	AAACTCTTCAAHNAHNGTCTCATGTTAATAACGTCCATATT
117	P4437	AAACTCTTCAAHNAHNGAGGCTCATGTTAATAACGTCCAT
118	P4438	AAACTCTTCAAHNAHNGCCGAGGCTCATGTTAATAACGTC
119	P4439	AAACTCTTCAAHNAHNTCCGCCGAGGCTCATGTTAATAAC
120	P4440	AAACTCTTCAAHNAHNAGGTCCGCCGAGGCTCATGTTAAT
121	P4441	AAACTCTTCAAHNAHNAGAAGGTCCGCCGAGGCTCATGTT

[0141] Each amplification reaction contained 30pmol of each oligonucleotide and 100 ng of pAC-FNA10 template DNA. Amplifications were carried out using Vent DNA polymerase (New England Biolabs). The PCR mix (20 ul) was initially heated at 95°C for 2.5 min followed by 30 cycles of denaturation at 94°C for 15s, annealing at 55°C for 15s and extension at 72°C for 40s. Following amplification, left and right fragments generated by the PCR reactions were gel-purified and mixed (about 200 ng of each fragment). Every mix contained three left fragments targeting three adjacent codons and three right fragments targeting same codons. These mixes were digested with *Eam*1104I, ligated with T4 DNA ligase and amplified by flanking primers (P4299 CGTTGAAGAAGATCACGTAGCA; SEQ ID NO:122, and P3246 TTTATTTTATAAACTCATTCCTGAT; SEQ ID NO:123). The resulting fragments were digested with *Mlu*I and *Xho*I, and cloned into the *Mlu*I/*Xho*I sites in the *Bacillus subtilis* expression plasmid pHPLT- FNA (Figure 2B). The BPN'-Y217L expression cassette from the pHPLT vector used has the polynucleotide sequence shown below.

[0142] The polynucleotide sequence of the expression cassette from the pHPLT vector (Plat promoter-pre-pro-BPN'-Y217L - terminator) (SEQ ID NO. 124) is shown below:

GCTTTTCTTTTGAAGAAAATATAGGGAAAATGGTACTTGTAAAAATTCGGAATATTTA
TACAATATCATAIGTTTCACATTGAAAGGGGAGGAAAATCGTGAACAACAAAAACGGC
TTTAGTCTAGCAAAAGGAGAGGGTAAAGAGTGAGAAGCAAAAAATTGTGGATCAGTTTG
CTGTTTGCTTTAGCGTTAATCTTTACGATGGCGTTTCGGCAGCACATCCTCTGCCAGGCGG
CAGGGAAATCAAACGGGAAAAGAAATATATTGTCGGGTTTAAACAGACAATGAGCAC
GATGAGCGCCGCTAAGAAGAAAGATGTCTATTCTGAAAAAGCGGAAAAGTGCAAAAG
CAATTCAAATATGTAGACGACGCTTCAGCTACATTAAACGAAAAAGCTGTAAAAAGAAAT
GAAAAAGACCGGAGCGTTCGCTTACGTTGAAGAAGATCAGTACGACACGCGTACGCGC
AGTCCGTGCTTACGGCTATCACAAATTAAGCCCTGCTCTGCACTCTCAAGGCTACA
CIGGATCAAAATGTAAAGTAGCGGTTATCGACAGCGGTATCGATCTCTCTCATCTGATT
TAAAGGTAGCAGGCGGAGCCAGCATGGTTCTTCTGAAACAAATCTTTCCAAGACAAC
AACTCTACGGAACTACGTTGCCGGCACAGTTGCGGCTCTTAATAACTCAATCGGTGTA
TTAGGCGTTGCGCAAGCGCATCACTTACGCTGTAAGTTCTCGGTGCTGACGGTTCC
GGCCAATACAGCTGGATCATTAAACGGAATCGAGTGGGCGATCGCAACAATATGGACGT
TATTAACATGAGCTCGGCGGACCTTCTGGTTCTGCTGCTTTAAAGCGGCAAGTTGATAA
AGCCGTTGCATCCGGCTCGTAGTCTGTCGGCAGCCGTAACGAAGGCACTTCCGGCA

GCTCAAGCACAGTGGGCTACCCCTGGTAAATACCCCTTCTGTCATTGCAGTAGGGCGTGTG
 ACAGCAGCAACCAAGAGCATCTTTCTCAAGCGTAGGACCTGAGCTTGATGTCATGGCA
 CCTGGCGTATCTATCCAAAGCACGCTTCTGGAACAAATACGGCGCGTTGAACGGTAC
 ATCAATGGCATCTCCGACGTTGGCCGAGCGGCTGCTTTGATTCTTTCTAAGCACCCGAA
 CTGGACAAACACTCAAGTCCGACGAGTTAGAAAACACCACTACAAAACCTGGTGATT
 CTTTCTACTATGGAAAAGGGCTGATCAACGTACAGGCGGCAGCTCAGTAAACTCGAGAG
 AGGACGGATTTCCTGAAGGAAATCCGTTTTTTATTTTAAGCTTG (SEQ ID NO:124)

[0143] Ligation mixtures were amplified using rolling circle amplification according to manufacturer's recommendation (Epicentre). One ul of the ligation mixture was mixed with 5 ul of the sample buffer, heated to 95°C for 3 min and cooled on ice. Next, 5 ul of the reaction buffer and 0.2 ul of the enzyme were added to each tube, followed by incubation at 30°C for 10 hours. Products of the rolling circle amplification were diluted 100 times and used to transform *Bacillus subtilis*. *Bacillus subtilis* cells (genotype: $\Delta aprE$, $\Delta nprE$, $\Delta spoII E$, $amyE::xyIRPxylAcomK-phleo$) made competent by the induction of the *comK* gene under control of a xylose inducible promoter (See e.g., Hahn et al., Mol Microbiol, 21:763-775 [1996]) were transformed with 45 libraries containing DNA sequences encoding BPN'-Y217L protease with mutated regions. The cells were grown in 1 ml of Luria Broth (LB) at 37°C for 1 hour and then plated on LB plates containing 1.6% skim milk and 10 mg/l neomycin, and were incubated overnight at 37°C. At least 2000 clones from each of the 45 libraries were screened for producing halos. When plated on skim milk plates, 0.05% to 20% of clones generated halos. A limited number of the clones generating halos were sequenced. These clones varied in the ratio of insertions and deletions, but none of them had frame shift mutations. Clones generating halos were further screened for AAPF activity using a 96-well plate assay as described in Example 1. The BPN'-Y217L variant proteins were produced by growing the *B. subtilis* transformants in 96 well microtiter plates as described earlier. Protein concentration of culture supernatants was determined by TCA precipitation as described in Example 1.

EXAMPLE 3

Stain Removal Performance of BPN'-Y217L Variants Generated by Targeted ISD

[0144] Results of experiments conducted to determine stain removal activity (microswatch assay to determine stain removal performance in laundry applications using EMPA 116 swatches (BMI stain, CFT) at pH 8/16°C and protein determination by TCA precipitation (tests of properties of interest) of BPN'-Y217L variants generated as described above are shown in Table 3-1. The results were obtained using the methods described in Example 1, with the following modifications for the stain removal performance assay. The test detergent used was heat inactivated Tide 2X detergent (Procter & Gamble).

[0145] Heat inactivation of commercial detergent formulas serves to destroy the endogenous enzymatic activity of any protein components while retaining the properties of nonenzymatic components. Heat inactivation of the detergents was performed by placing pre-weighed amounts of liquid detergent (in a glass bottle) in a water bath at 95°C for 2 hours. The detergent was purchased from local supermarket stores. Both unheated and heated detergents were assayed within 5 minutes of dissolving the detergent to accurately determine percentage deactivated. Enzyme activity was tested by AAPF assay. As described throughout functionality of BPN'-Y217L variants was quantified as a performance index ("Pi" or "PI"), which is the ratio of performance of a variant to parent protein BPN'-Y217L. BPN'-Y217L variants showing a PI value greater or equal than 0.5 for BMI stain removal performance and/or TCA precipitation showed improved cleaning benefits and/or expression.

[0146] Mutations are named by the one letter code for the parent amino acid, followed by a three digit position number and then the one letter code for the variant amino acid. For example, mutating glycine (G) at position 87 to serine (S) is represented as "G087S." Multiple mutations are indicated by inserting a "/" between the mutations. Mutations at positions 87 and 90 are represented as "G087S/A090Y." For deletions, the one letter code "Z" is used. For an insertion relative to the parent sequence, the one letter code "Z" is on the left side of the position number. For a deletion, the one letter code "Z" is on the right side of the position number. For insertions, the position number is the position number before the inserted amino acid(s), plus 0.01 for each amino acid. For example, an insertion of three amino acids alanine (A), serine (S) and tyrosine (Y) between position 87 and 88 is shown as "Z087.01A/Z087.02S/Z087.03Y." Thus, combining all the mutations above plus a deletion at position 100 is: "G087S/Z087.01A/Z087.02S/Z087.03Y/A090Y/A100Z."

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
A001T/L096V/G097S/A098D/D099I	0.45	1.1
A001V/G097S/A098R/D099G	0.66	0.74

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
S003F	1.08	0.89
V004M/A098D/D099G	0.69	0.97
V004M/D099R/G100S/Z100.01L	0.8	0.75
V004M/G127C/G128S/P129R	0.92	0.52
V004M/M119C/D120S	0.68	1
V008I	0.97	0.985
V008I/A098G/D099R	0.67	0.66
V008I/A098N	0.52	1.14
V008I/M119C/D120G	0.53	0.99
Q010L/G097R/A098C/D099S	0.27	0.52
Q010R	0.91	0.86
K012N/A098R/D099V	0.46	0.8
K012R/G097D/A098H/D099G	0.86	0.96
A013T/S101R/Q103S	0.52	0.68
A013V/G097N/A098H/D099G/G160S	0.46	0.74
A013V/G097S/Z097.01R/A098R	0.46	0.64
A013V/G100R/S101G	0.43	0.79
A013V/N118G/M119I	0.5	0.85
A013V/P129Z/S130H/S132G	0.47	0.58
P014L/N062D/S101R/Q103S	0.41	1.02
P014S/A098S/D099G	0.56	0.99
A015T/V068N/A069G	0.52	0.38
L016M/G128R/P129R/Z129.01G	0.5	0.28
H017Q/G127R/P129G	0.54	0.43
H017R	0.5	1.025
H017Y/A098R/D099I	0.42	0.56
H017Y/A098R/D099Z/S101G	0.6	0.47
H017Y/P129R	0.685	0.85
H017Y/S130Y/G131S/S132L	0.38	0.58
Q019H/G128R/Z128.01R/P129D	0.4	0.78
Q019H/Z128.01G/Z128.02G	0.56	0.29
G020D/G097R/A098G/D099C	0.39	0.8
G020D/P129Z/S130G/G131R/S132C	0.37	0.51
N025Y/S101L/Q103N	1.05	1.13
A029V/G097R/A098S	0.44	0.94
S033G	0.9	1.05
S033I/T066G/H067R/V068D/A216E	0.88	1.01
S038F/A098R/D099S	0.98	0.6
V044I/G097C/A098D/D099G	0.42	0.9
V044I/G097R/A098R	0.83	0.87
V044I/G100S/S101G	0.36	1.09
A045V/G097C/A098R/D099S	0.41	0.88

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
A048V/G097L/A098G/D099S	0.42	1
A048V/G100R/S101G	0.65	1.085
A048V/G128S/P129L/Z129.01G	0.45	0.68
M050I/A098G	0.2	0.54
S053F/S130I/S132G	0.65	0.54
S053T/D099H/S101Z	0.66	0.74
E054D/A098R/D099G	1.05	0.76
T055I/S101R/Q103R	0.58	0.51
T055S/T066I/Z066.01G/Z066.02G	0.33	0.7
Q059H/D099S/S101G	0.36	0.79
Q059H/M119S/D120R	0.37	0.52
Q059K/M119V/V121C	0.23	0.58
Q059L/P129G	0.69	0.87
D060N	0.47	0.9
N061D/G128R/P129V/Z129.01G	0.59	0.43
S063T/G097D/A098R/D099S	0.97	0.95
G065R/Z101.01R/Z101.02G	0.57	1.035
T066S/H067S	0.51	0.61
V068C/A069G	0.42	1.22
V068C/A069G/H238Q	0.32	0.62
V068G/A069G	0.36	0.71
V068G/A069G/V150I	0.33	0.59
V068G/A069S	0.34	0.63
V068I/A069G/A144V	0.46	0.98
V068I/G097D/A098G/D099Z/G100R	0.41	0.58
V068L/A069G	0.445	0.905
V068L/A069S	0.47	0.83
V068S/A069G	0.48	1.04
V068S/A069G/A116T	0.4	1.05
A069G	0.7	1.02
A069G/T071G	0.23	0.84
A069G/T071I	0.79	1
L075G/N076G/N077Z	0.29	0.77
L075I	0.78	0.94
L075R/N076G/N077Z	0.34	0.99
N076D/D099R/Z100.01G	0.28	0.63
I079F/A098G	1.16	1.16
V081A/S101R/Q103V	0.4	0.81
V081I/G097R/A098G/D099C	0.36	0.54
V084D/G097S/A098G/D099N	0.42	0.63
V084I/G127C/P129G	1.28	1.01
V084I/G127L	0.88	0.72

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
A085V/A098G	0.75	1.09
A085V/G097R/A098S/D099N	0.65	0.75
A085V/G128S/P129R/Z129.01G	0.61	0.47
A085V/N218I	0.435	0.845
A085V/S101G	0.59	0.84
A085V/S204P	0.43	0.78
A085V/Z099.01H/S101R	0.46	0.6
P086S/G131D/S132G	0.25	0.64
P086T/S101D/Q103S/V147I	0.35	1.27
A088S/S101R/Q103V	0.63	0.88
A088T	0.855	1.01
A088T/A098R/D099R	0.73	0.36
A088T/D099G/Z100.01G	0.21	0.75
A088T/G127N/P129R	0.99	0.8
A088T/G127S/P129R/Q206R/A273S	1.22	0.67
A088T/P129S/G146D	0.74	1.16
A088T/S101G	1.13	1.08
A088T/S130R/G131H/S132N	1.24	0.78
A088V/G100Z/S101C/Q103R	0.52	0.46
A088V/P129Z	0.61	1.05
A092G	0.61	1.15
A092G/S249R	0.69	1.07
A092G/V093C	0.2	0.855
A092G/V093I	0.5	1.15
A092G/V093L	0.275	0.75
A092L/V093G/K094Z	0.54	0.96
A092S/K094V	0.47	1.13
A092S/V093C	0.72	1.06
A092S/V093G	0.21	0.79
A092S/V093S	0.33	0.67
A092V/V093L	0.34	0.695
V093I/G128D/P129R	0.6	1.22
V093I/P129Z	0.6	0.92
V093R/Z093.01G/K094R	0.28	0.88
K094E/V095D	0.87	0.95
K094G	0.33	0.56
K094S/V148I	0.3	0.85
L096I/N118R/M119C/D120G/A151V	0.29	0.58
Z096.01D/A098R	0.66	1.2
Z096.01H/A098G	0.32	1.16
Z096.01N/A098H	0.65	1.05
Z096.01R/A098C	0.47	0.95

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
Z096.01R/A098G	0.39	0.82
Z096.01R/A098G/V192I	0.4	0.77
Z096.01R/A098R	0.60	0.70
Z096.01R/A098R/V192A	0.61	0.68
Z096.01R/A098S	0.47	0.92
Z096.01R/A098S/G157S	0.3	0.88
Z096.01R/Z096.02G	0.52	0.67
Z096.01R/Z096.02G/V147I	0.41	0.96
Z096.01S/A098G	0.44	1.12
Z096.01S/A098N	0.47	1.04
G097C/A098D/D099S	0.675	1.09
G097C/A098G/D099G	0.42	0.86
G097C/A098G/D099N	0.82	1.07
G097C/A098G/D099S	0.45	0.95
G097C/A098G/D099S/G100Z	0.32	0.64
G097C/A098G/D099V	0.43	0.88
G097C/A098N/D099S	0.45	0.94
G097C/A098R	1.24	1.11
G097C/A098R/D099G	0.40	0.84
G097C/A098R/D099H	0.45	0.64
G097C/A098R/D099H	0.46	0.78
G097C/A098R/D099N	0.79	0.87
G097C/A098R/D099R	0.4	0.55
G097C/A098R/D099R/N109D/A114S	0.56	0.5
G097C/A098S/D099H/A151V	0.32	0.65
G097C/A098V/D099S	0.2	0.81
G097C/Z097.01G/A098H/T253I	0.32	0.58
G097C/Z097.01S/A098G	0.45	1.04
G097D/A098C	1	1.03
G097D/A098G/D099G	0.89	0.985
G097D/A098G/D099R	1.00	0.83
G097D/A098H/D099R	1.24	0.92
G097D/A098H/D099V	0.95	0.86
G097D/A098R/D099G	0.955	0.96
G097D/A098R/D099R	1.29	0.79
G097D/A098R/D099S	1.11	0.87
G097D/A098R/D099Z	0.255	1.09
G097D/G100C/S101R	1.15	0.86
G097D/G100R/S101G	0.47	0.85
G097H/A098C/D099G	0.34	0.76
G097H/A098D/D099N	0.5	1.15
G097H/A098G	0.67	1.1

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
G097H/A098G/D099G	0.38	0.78
G097H/A098G/D099R	0.42	0.73
G097H/A098G/D099Z	0.39	1.02
G097H/A098G/D099Z/A151V	0.37	0.52
G097H/A098G/D099Z/G100R	0.55	0.49
G097H/A098G/D099Z/G100V	0.2	0.87
G097H/A098G/D099Z/S101N/G166S	0.41	0.68
G097H/A098R	0.60	0.97
G097H/A098R/D099G	0.46	0.67
G097H/A098R/D099L	0.31	0.5
G097H/A098R/D099N	0.48	0.74
G097H/A098S/D099G	0.76	0.84
G097H/A098S/D099I	0.35	0.55
G097H/A098S/D099L	0.36	0.83
G097H/A098S/D099S/K237E	0.53	0.88
G097L/A098C/D099S	0.26	0.68
G097L/A098G/D099S	0.33	0.77
G097L/A098G/D099S/G 100Z	0.28	0.53
G097L/A098G/D099Z	0.29	0.56
G097L/A098H	0.71	1.1
G097L/A098R/D099N	0.5	0.78
G097L/A098V	0.42	1.11
G097N/A098G/D099C	0.74	0.99
G097N/A098G/D099R	0.82	0.88
G097N/A098G/D099R/V270I	0.74	0.88
G097N/A098G/D099S	0.45	0.7
G097N/A098G/D099V	0.47	0.9
G097N/A098G/D099V/A151V	0.3	1.07
G097N/A098G/D099V/S101F	0.54	0.82
G097N/A098H/D099N	0.79	0.75
G097N/A098R/D099R	0.69	0.25
G097N/A098R/D099S	0.94	0.73
G097N/A098R/D099V	0.59	0.675
G097N/A098S/D099S	0.72	0.91
G097R/A098C	1.04	0.98
G097R/A098C/D099S	0.28	0.735
G097R/A098F/A200V	0.305	0.74
G097R/A098G	0.97	0.985
G097R/A098G/D099C	0.31	0.68
G097R/A098G/D099G	0.33	0.60
G097R/A098G/D099G/A273T	0.48	0.85
G097R/A098G/D099N	0.64	0.82

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
G097R/A098G/D099S	0.31	0.66
G097R/A098G/D099S/A133D	0.26	0.61
G097R/A098G/D099V	0.275	0.55
G097R/A098H/D099C/S182N	0.38	0.75
G097R/A098H/D099G	0.47	0.695
G097R/A098N/D099G	0.32	1.05
G097R/A098N/D099Z	0.52	0.72
G097R/A098R	1.06	0.83
G097R/A098R/D099C	0.50	0.53
G097R/A098R/D099G	0.54	0.51
G097R/A098R/D099S	0.45	0.55
G097R/A098S/D099C	0.39	0.66
G097R/A098S/D099G	0.51	0.59
G097R/A098S/D099L	0.2	0.51
G097R/A098S/D099R	0.45	0.52
G097R/A098S/D099S	0.33	0.81
G097R/A098S/D099V	0.32	0.61
G097R/A098V/D099G	0.45	0.68
G097R/Z097.01H/A098C	0.56	0.81
G097R/Z097.01R/A098G	0.5	0.74
G097R/Z097.01R/A098G/V147I	0.56	0.74
G097R/Z097.01R/A098S	0.4	0.59
G097R/Z097.01R	0.39	0.6
G097R/Z097.01S/A098G	0.57	0.95
G097R/Z097.01S/A098R	0.53	0.7
G097S/A098C	0.87	1.06
G097S/A098C/D099G	0.47	1.09
G097S/A098C/D099H	0.62	1.02
G097S/A098C/D099L	0.45	0.96
G097S/A098C/D099R	0.575	0.79
G097S/A098C/D099S	0.71	1.15
G097S/A098C/D099Z	0.2	0.81
G097S/A098G/D099F	0.28	0.63
G097S/A098G/D099G	0.43	0.90
G097S/A098G/D099G	0.43	0.91
G097S/A098G/D099S	0.56	0.95
G097S/A098G/D099Z	0.3	0.94
G097S/A098G/D099Z/G100R	0.3	0.52
G097S/A098H/D099C	0.46	0.84
G097S/A098H/D099G	0.42	0.97
G097S/A098N/D099G	0.53	1.14
G097S/A098R	1.06	0.94

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
G097S/A098R/D099C	0.45	0.7
G097S/A098R/D099G	0.63	0.76
G097S/A098R/D099L	0.55	0.715
G097S/A098R/D099V	0.27	0.63
G097S/A098R/D099Y	0.44	0.74
G097S/A098S/D099G	0.65	0.945
G097S/A098S/D099G/A133V/A153V/S236Y	0.48	0.69
G097S/A098S/D099G/A153V	0.46	0.82
G097S/A098S/D099Z	0.875	0.76
G097S/Z097.01D/A098G/A144T/Q271H	0.46	1.04
G097S/Z097.01G/A098V	0.6	1.02
G097S/Z097.01S/A098G	0.6	1.12
G097S/Z097.01S/A098G/A273T	0.41	1.2
G097S/Z097.01Y	0.33	1.04
G097V/A098R	0.41	0.74
G097V/A098R/D099G	0.46	0.56
G097V/A098S/D099S	0.24	0.84
G097Y/A098H/D099N	0.3	0.56
G097Z/A098S/D099H	0.31	0.77
Z097.01D/A098R/D099G	0.39	1
Z097.01D/A098S/D099L	0.36	0.59
Z097.01D/A098V/D099G	0.345	0.805
Z097.01G	0.54	1.12
Z097.01G/A098G	0.47	1.02
Z097.01G/A098G/D099S	0.21	1.03
Z097.01G/A098R	0.73	0.86
Z097.01G/A098R/D099C	0.43	0.73
Z097.01G/A098R/D099G	0.39	0.64
Z097.01G/A098R/D099S	0.48	0.63
A098C	0.97	0.91
A098C/D099G	0.57	0.98
A098C/D099G/A116T	0.48	1.03
A098C/D099H	0.68	0.78
A098C/D099L	0.4	0.64
A098C/D099L/K213R	0.36	0.85
A098C/D099L	0.39	0.95
A098C/D099N	0.66	0.96
A098C/D099R	0.62	0.80
A098C/D099S	0.69	0.98
A098C/D099S/P194L	0.47	0.79
A098D/D099C	0.59	0.91
A098D/D099G	0.59	1.09

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
A098D/D099G/G100L	0.65	1.06
A098D/D099H/G100S	0.3	0.91
A098D/D099H/H238Y	0.38	1.07
A098D/D099R	0.69	0.72
A098D/D099R/G100D	0.28	0.91
A098D/D099R/G160S	0.6	0.78
A098F/D099S/G100L	0.3	0.52
A098G	0.91	1.01
A098G/D099C	0.67	0.77
A098G/D099G	0.55	0.85
A098G/D099G/A228T	0.58	0.9
A098G/D099G/G100H	0.58	0.38
A098G/D099G/N243D	0.27	0.71
A098G/D099G/S101P	0.55	0.57
A098G/D099G/T244S	0.51	0.895
A098G/D099G	0.49	0.85
A098G/D099H	0.44	0.89
A098G/D099L	0.4	0.75
A098G/D099R	0.56	0.70
A098G/D099R/A116T	0.6	0.68
A098G/D099R/G100D	0.28	0.62
A098G/D099R/V148I	0.79	0.61
A098G/D099S	0.58	1.01
A098G/D099S/L267M	0.48	0.89
A098G/D099V	0.32	0.93
A098G/D099Z	0.25	0.69
A098H	1.04	0.96
A098H/D099C	0.46	0.79
A098H/D099G	0.82	0.89
A098H/D099G/A216T	0.72	0.76
A098H/D099G/G100D	0.52	1.09
A098H/D099G/G100N/L267M	0.35	1.02
A098H/D099G/G100S	0.355	0.9
A098H/D099G/V143I	0.41	0.84
A098H/D099R	1.07	0.62
A098H/D099R/G100S	0.37	0.54
A098H/D099S/G100C	0.33	0.67
A098H/D099S/G100N	0.43	1.05
A098H/D099Y	0.32	0.81
A098I/D099G	0.54	0.97
A098I/D099S	0.47	0.91
A098L/D099G	0.44	1.15

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
A098L/D099S	0.5	0.91
A098N	1.05	1
A098N/D099G	1.14	1.01
A098N/D099L	0.32	0.71
A098N/D099S	0.74	1
A098R/A144T	0.81	0.91
A098R/A274V	1.11	0.88
A098R/D099C	0.56	0.82
A098R/D099G	0.94	0.70
A098R/D099G/G100C	0.35	0.68
A098R/D099G/G100D	0.83	0.92
A098R/D099G/G100L/S 145T	0.68	0.84
A098R/D099G/G100N	0.66	0.74
A098R/D099G/G100R	0.57	0.25
A098R/D099G/G100S	0.55	0.82
A098R/D099G/V150I	0.84	0.68
A098R/D099H	0.85	0.59
A098R/D099I	0.39	0.665
A098R/D099L	0.54	0.62
A098R/D099N	1	0.895
A098R/D099N/G100L	0.57	0.79
A098R/D099R	0.72	0.32
A098R/D099R/G100D	0.41	0.53
A098R/D099R/S101Z	0.61	0.23
A098R/D099R/S130F	0.23	0.51
A098R/D099R	0.64	0.51
A098R/D099S	0.75	0.80
A098R/D099S/G100L/A153V	0.55	0.79
A098R/D099S	0.87	0.81
A098R/D099V	0.505	0.775
A098R/D099Z/S101G	0.395	0.61
A098R/G100C/A137T	0.64	0.96
A098R/G100S	0.95	1
A098S/D099C	0.655	0.865
A098S/D099G	0.89	0.91
A098S/D099G/G100C	0.35	0.86
A098S/D099G/G100D	0.26	1.26
A098S/D099G/G100R/L250I	0.58	0.55
A098S/D099G/G100S	0.48	0.98
A098S/D099G/Z099.01R	0.25	0.52
A098S/D099H	0.6	0.95
A098S/D099H/M124V	0.53	0.95

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
A098S/D099H/Z099.01R/G169A/V180A	0.63	0.62
A098S/D099R	0.80	0.78
A098S/D099R/G100L	0.71	0.67
A098S/D099R/G154S	0.41	0.61
A098S/D099R/G166S	1.16	0.75
A098S/D099R/Z100.01G	0.47	0.66
A098S/D099S	0.82	0.97
A098S/D099S/Z099.01R	0.38	0.66
A098S/D099V	0.41	1.03
A098T/P129Z/S130R/S132G	0.33	0.53
A098T/S101G	0.785	0.925
A098V/D099G	0.965	0.96
A098V/D099G/G100D	0.5	0.98
A098V/D099G/G100D/A153V	0.41	0.62
A098V/D099G/G100S	0.66	0.83
A098Y/D099R/G100L	0.31	0.54
A098Y/D099R/G100Z/S101Z/G102D	0.98	0.66
A098Z/S101R	0.56	0.62
Z098.01N	0.5	1.31
Z098.01R	0.54	0.97
Z098.01R/G100R	0.55	0.51
D099C/K136N	0.3	0.8
D099C/S101D	0.73	1.08
D099C/S101R	0.655	0.67
D099C/S101Z	0.40	0.61
D099C/Z099.01S	0.3	0.88
D099C/Z100.01G	0.36	0.56
D099F/S101Z	0.35	0.51
D099G/G100C	0.5	0.58
D099G/G100D/S101R	0.54	0.93
D099G/G100H	0.62	0.71
D099G/G100R/S101V	0.45	0.58
D099G/G100S/S101R	0.55	0.79
D099G/S101G	0.67	0.83
D099G/S101H	0.79	0.95
D099G/S101V	0.85	0.95
D099G/S101Z	0.56	0.81
D099G/S101Z/N269D	0.45	0.81
D099G/Z099.01N	0.33	1.21
D099G/Z099.01R/G100S	0.26	0.59
D099G/Z099.01S	0.21	1.17
D099G/Z100.01G	0.26	0.71

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
D099G/Z100.01G/S183N/Q275K	0.35	0.71
D099H/S101Z	0.49	0.74
D099H/S101Z/V150I	0.56	0.59
D099H/Z100.01G/A273V	1.91	0.38
D099H/Z100.01G/Z100.02G	0.2	0.84
D099I/Z099.01 S	0.48	0.805
D099L/N118D	0.5	0.82
D099L/S101G	0.23	0.91
D099L/S101V	0.46	0.88
D099L/Z100.01G	0.26	0.51
D099N/A116T	1	0.92
D099N/G100R/S101V	0.525	0.725
D099N/S101G	0.57	0.95
D099N/S101H	0.98	0.89
D099N/S101Z	0.48	0.94
D099N/Z099.01S	0.69	1
D099N/Z100.01G	0.42	0.94
D099R	0.7	0.71
D099R/S101D	0.78	1
D099R/S101G	0.595	0.58
D099R/S101V	0.75	0.66
D099R/S101Z	0.48	0.52
D099R/S101Z/A133T	0.35	0.52
D099R/Z099.01 N	0.28	1.06
D099R/Z099.01S	0.42	0.71
D099R/Z100.01G	0.43	0.66
D099R/Z100.01N	0.44	0.5
D099S/S101G	0.53	0.98
D099S/S101I	0.69	0.925
D099S/S101Z	0.71	0.77
D099S/Z099.01H	0.62	0.67
D099S/Z099.01L	0.35	0.59
D099S/Z099.01V	0.5	0.35
D099S/Z100.01G	0.47	0.85
D099S/Z100.01G/Z100.02G	0.21	1.07
D099V/S101D	0.54	1.24
D099V/S101V	0.33	0.825
D099V/S101Z	0.36	0.51
Z099.01H/S101L	0.56	0.58
Z099.01H/Z099.02R	0.5	0.44
Z099.01N/S101R	0.46	1.09
Z099.01S	0.48	1.17

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
Z099.01S/S101V	0.51	0.91
Z099.01S/Z099.02G/A228T	0.61	0.24
G100C	0.85	0.98
G100D	0.54	1.18
G100D/S101G	0.54	0.68
G100D/S101R	0.74	1.13
G100H/G102V	0.26	1.04
G100H/S101G	0.35	0.91
G100H/S101R	0.74	0.945
G100L/S101G	0.775	1.135
G100L/S101R	1.24	1.01
G100N/S101D	0.5	0.86
G100N/S101L	0.93	1.075
G100N/S101R	1.27	0.99
G100R	0.64	0.84
G100R/L267R/I268V	0.7	1.01
G100R/S101G	0.46	0.78
G100R/S101G/G160S/S182N	0.5	0.69
G100R/S101G/Q103Z/Y104Z/S161N	0.33	0.73
G100R/S101G/S163N	0.55	0.54
G100R/S101N	0.78	0.89
G100R/S101R	0.60	0.87
G100R/S101R/Q103Z/V270I	0.58	1
G100R/S101Z/G102C/Q103D	0.5	0.26
G100S	0.59	1.14
G100S/S101C	0.69	0.96
G100S/S101G	0.47	1.02
G100S/S101R	0.75	0.99
G100S/S101R	0.94	1.01
G100S/S101V	0.53	1.13
G100S/S101V/A231T	0.51	0.46
Z100.01G	0.43	0.76
Z100.01L	0.43	1.05
Z100.01N	0.43	0.67
Z100.01R	0.47	0.76
Z100.01R/S101G	0.45	0.95
S101C	1.48	0.89
S101C/Q103I	0.68	0.91
S101C/Q103S	1.34	0.97
S101D	1.055	1.125
S101D/Q103C	0.85	1.15
S101D/Q103D	0.94	1.1

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
S101D/Q103F	1.04	1.1
S101D/Q103H	0.89	1.24
S101D/Q103H	0.68	1.09
S101D/Q103R	1.06	0.87
S101D/Q103R/H238Y	0.69	1.11
S101D/Q103S	0.63	1.14
S101D/Q103V	0.41	1.09
S101F/Q103G	0.32	0.75
S101G	0.83	0.99
S101G/A151V	0.41	1.23
S101G/A179V	0.29	0.58
S101G/G102S	0.42	1.13
S101G/Q103G	0.38	0.59
S101G/Q103L	0.74	1.01
S101G/Q103N	0.75	1.14
S101G/Q103R	0.71	0.83
S101G/Q103R/A179T	0.32	0.52
S101G/Q103R/G157D	0.59	0.81
S101G/Q103S	0.57	1.12
S101G/Q103S/L233S	0.8	0.97
S101G/Q103S/L233S	0.45	1.13
S101G/Q245H	0.56	0.93
S101G/S249N	0.77	1.115
S101G/V139I	0.85	1.01
S101G/Z101.01R	0.435	0.965
S101G/Z101.01R/A200T	0.52	0.26
S101G/Z102.01G	0.465	0.69
S101H	0.92	1.03
S101H/G102S	0.68	1.23
S101H/Q103D	0.985	1.185
S101H/Q103G	0.38	0.84
S101H/Q103G/E251Q	0.37	0.7
S101H/Q103H	1.07	0.93
S101H/Q103I	0.715	1.035
S101H/Q103R	0.73	0.82
S101H/Q103R/A274Z/Q275Z	0.67	0.68
S101H/Q103S	0.76	1.07
S101H/Q103V	0.67	1.09
S101I	0.78	0.95
S101I/Q103R	0.81	0.91
S101L	1.09	1.04
S101L/G102S/Q103R	0.87	1.07

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
S101L/Q103D	1.03	1.07
S101L/Q103G	0.33	0.9
S101L/Q103R	0.61	0.75
S101L/Q103R/A138T	0.43	0.73
S101L/Q103R/V121W/K213E	0.81	0.95
S101L/Q103S	1.04	1.04
S101L/Q103V	0.66	1.05
S101L/Z101.01R	0.5	0.99
S101N/Q103R	0.94	0.925
S101N/Z102.01G	0.49	0.65
S101R	0.85	0.90
S101R/A151V	0.4	0.66
S101R/A273V	0.78	0.71
S101R/E156G	1.1	0.66
S101R/G102L/Q103G	0.95	1.04
S101R/G102S	1.58	1.1
S101R/G102S/Q103R	0.81	0.91
S101R/G102S/Q103V	0.54	0.89
S101R/G131D	1.45	0.79
S101R/Q103C	1.05	0.89
S101R/Q103D	0.75	1.01
S101R/Q103D/S248R	0.89	0.75
S101R/Q103G	0.49	0.84
S101R/Q103G/A116T	0.52	0.97
S101R/Q103G/V147I/A153V	0.445	1.075
S101R/Q103G	0.49	0.7
S101R/Q103L	0.80	0.63
S101R/Q103N	1.00	0.89
S101R/Q103R	0.74	0.56
S101R/Q103R/S161N	1.35	0.63
S101R/Q103R/V148I	0.58	0.44
S101R/Q103S	0.93	0.85
S101R/Q103S/L126F	1.23	0.84
S101R/Q103V	0.74	0.83
S101V	0.915	1.05
S101V/Q103G	0.46	0.99
S101V/Q103H	0.75	0.98
S101V/Q103N/Z275.01K	1.17	1.13
S101V/Q103R	0.62	1.03
S101V/Q103V	0.58	1.09
S101Y/G102S/Q103R	0.83	0.94
S101Y/Q103L	0.54	0.75

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
S101Y/Q103S	0.54	0.93
S101Y/Q103V	0.58	0.95
S101Z	0.34	0.95
S101Z/G102I/Q103H/Y104G/L135F	0.32	0.7
S101Z/G102V/Q103R/Y104L	0.37	0.61
S101Z/Q103D	0.52	0.29
S101Z/Q103F	0.45	1.07
S101Z/Q103L	0.41	1
S101Z/Q103R	0.49	0.76
G102C/Q103L/Y104S	0.96	1.15
G102R/Q103C/Y104C/V192I	0.33	1.26
G102R/Q103G/Y104R	0.25	0.69
G102R/Q103R/Z103.01G	0.655	0.885
G102Z/Q103S	0.33	0.61
Z102.01G/Q103R	0.41	1.02
Z102.01G/Q103R/L267V	0.31	0.64
Z102.01G/Z102.02R/Z102.03G	0.53	0.67
Q103D	1.04	1.21
Q103G	0.335	1.04
Q103H/Y104N/Q275Z	0.56	0.81
Q103N/E156D	0.92	1.07
Q103R	0.75	0.82
Q103R/I122L/N123S/M124V	0.68	0.65
Q103R/Y104F	0.7	0.795
Q103S	1.35	0.86
Q103V	0.61	0.955
Q103Y/S182N	0.87	0.77
S105G/P129S/S130H/G131H	1.09	0.78
W106C/A116D/N117S/N118C	0.47	0.73
N109S/M119C/D120G/V121L	0.28	0.97
A116G/N117Z/N118R/M119C	0.37	0.88
A116H/N117G/N118R	0.39	0.94
N117C/N118G/M119S	0.44	1.05
N117C/N118G/M119S/A216V	0.33	0.87
N117G/N118G/M119C	0.43	0.94
N117G/N118R/M119C	0.29	1.03
N117H/N118C/M119C	0.45	0.57
N117H/N118D/M119S	0.63	0.88
N117H/N118G/M119C	0.97	0.83
N117H/N118G/M119L	0.59	0.86
N117H/N118G/M119S	0.72	0.89
N117H/N118G/M119V	0.62	0.95

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
N117H/N118H/M119V	1.07	0.96
N117H/N118R/M119C	0.98	0.75
N117H/N118R/M119S	0.53	0.77
N117H/N118S/M119C	0.71	0.84
N117H/N118S/M119S	0.4	0.985
N117H/N118S/M119V	0.64	0.89
N117I/N118G/M119V	0.38	1.01
N117L/N118G/M119V	0.38	0.77
N117R/N118G/M119C	0.63	0.80
N117R/N118G/M119V270I	0.46	0.88
N117R/N118G/M119S	0.375	0.79
N117R/N118H/M119V	0.66	0.91
N117R/N118R/M119C	0.755	0.755
N117R/N118R/M119I	0.67	0.71
N117R/N118R/M119S	0.28	0.605
N117R/N118R/M119V	0.80	0.73
N117R/N118S/M119I	0.38	0.75
N117R/N118S/M119L	0.23	0.72
N117S/N118G/M119C	0.595	1.085
N117S/N118G/M119S	0.3	1.1
N117S/N118H/M119V	0.49	1.01
N117S/N118R/M119V	0.58	0.93
N117V/N118G/M119C	0.47	0.99
N117V/N118H/M119C	0.39	0.71
N117Y/N118G/M119S	0.26	0.74
N117Y/N118G/M119V	0.475	1
Z117.01G/N118R/M119C	0.29	0.65
N118C/M119V/D120S	0.27	0.82
N118C/Z118.01G/M119C	0.485	0.86
N118D/M119C/D120L	0.43	0.98
N118D/M119S/D120R	0.24	0.88
N118G/M119C/D120G	0.63	1.02
N118G/M119C/D120L	0.5	0.99
N118G/M119/D120R	0.65	0.94
N118G/M119N	0.28	0.73
N118G/M119S/D120G	0.29	0.69
N118G/M119S/D120R	0.38	0.83
N118G/M119V/D120G	0.49	0.94
N118G/M119V/D120S/S182N	0.52	0.99
N118H/M119C/D120R	0.88	0.87
N118H/M119/D120G	0.54	1.05
N118H/M119N/V148I	0.38	0.63

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
N118H/M119S/D120G	0.28	0.62
N118H/M119V/D120G	0.54	1
N118R/M119C	0.97	0.99
N118R/M119C/D120G	0.63	0.85
N118R/M119C/D120R	0.8	0.85
N118R/M119S	0.45	0.99
N118R/M119S/D120G	0.32	0.53
N118R/M119S/D120R	0.29	0.57
N118R/M119S/D120S	0.28	0.82
N118R/M119V/D120C	0.27	0.97
N118R/M119V/D120G	0.56	0.76
N118R/M119V/D120G/A232T	0.32	0.825
N118S/M119C/D120R	0.56	0.91
M119C/D120G	0.58	0.98
M119C/D120G/V121I	0.48	0.95
M119C/D120G/V121L	0.39	0.52
M119C/D120H	0.91	1.01
M119C/D120H/A187V	0.44	0.93
M119C/D120R	0.76	0.95
M119H/V121I	0.42	0.58
M119/D120C	0.44	0.88
M119/D120G	0.47	0.96
M119/D120R	0.67	0.86
M119L/D120G	0.39	0.51
M119S/D120R	0.28	0.97
M119S/D120S/V121I	0.46	0.72
M119S/V121I	0.44	1.15
M119V/D120C	0.27	0.93
M119V/D120G	0.43	1.06
M119V/D120R	0.63	0.86
M119V/D120S/V198L	0.5	0.93
D120G/I122C	0.27	0.83
D120G/I122C/L217S	0.34	0.56
D120G/I122G	0.54	0.23
D120G/I122V	0.5	1
D120H/I122L	0.66	1.03
D120H/V121C/I122V	0.31	0.74
D120N/V121L/I122V	0.32	0.93
D120R/A151V	0.37	0.71
D120R/I122V	0.63	0.97
D120R/V121I	0.72	1
D120R/V121I/I122C	0.39	0.74

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
D120R/V121W/I122C/A228T	0.29	0.98
D120S/I122C/A133S	0.31	1.1
V121C	0.63	0.86
V121C/I122C/N123C	0.54	0.2
V121C/I122G/N123V	0.56	0.6
V121C/I122S/N123V	0.56	0.57
V121C/I122V/N123S	0.52	0.59
V121W/I122C/N123C	0.53	0.6
V121W/I122C/V150I	0.8	1.13
V121W/I122G/N123C	0.65	1
V121W/I122S/N123C	0.64	1.36
V121W/I122S/N123I	0.95	0.83
V121W/I122V/N123S	0.82	0.98
V121L/I122S/N123V	0.53	0.9
V121L/N123C	1.02	1.16
I122C/N123C	0.6	0.6
I122C/N123S/M124L	0.98	1.015
I122D/N123V/M124L	0.55	0.63
I122G/N123C/M124L	0.96	0.96
I122G/N123G/Z123.01C	0.63	0.45
I122H/N123V/M124L	0.65	0.76
I122S/N123S	0.63	1.27
I122V/N123G/M124C/A138V	0.51	0.25
I122V/N123S/M124I	0.81	0.75
I122V/S183G	0.91	0.97
N123C/M124S	0.98	0.785
N123C/M124V	1.875	0.745
N123D	0.7	0.89
N123S/M124L	1.55	1.13
N123S/M124S	0.71	0.26
M124C	0.92	1.05
M124C/L126S	0.7	0.51
M124C/L126S/T244I	0.72	0.455
M124C/L126V	1.535	1.085
M124G	0.53	0.3
M124I	1.72	1.16
M124I/L126H	0.87	0.54
M124S	0.6	0.58
M124V	1.08	1.20
L126C	1.8	0.35
L126F/P129Z	1.07	1
L126F/P129Z/S182N	1.47	1.24

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
L126G	0.67	0.58
L126I/P129Z	0.53	0.71
L126R/G127R	0.54	0.47
L126S	1.08	0.94
L126Y	1.04	1.33
Z126.01S	0.67	0.59
G127C/G128S/P129R	0.77	0.48
G127C/P129G	1.53	0.72
G127C/P129G/A200T	0.56	0.4
G127C/P129R	1.785	0.825
G127C/P129R/A153T	0.76	0.48
G127D/P129G	0.91	0.73
G127D/P129S	0.71	0.69
G127H/P129G	1.65	0.71
G127H/P129R	1.1	0.77
G127L	1.14	0.83
G127L/P129R	1.02	0.80
G127N/P129C	1.05	0.88
G127N/P129G	1.1	0.91
G127N/P129G/A133T	0.87	1.04
G127N/P129G/A151V/K213N	0.37	0.72
G127N/P129R	1.10	0.77
G127N/P129S/A273V	1.07	0.9
G127N/P129V	1.16	0.92
G127R/A230V	0.5	0.58
G127R/P129D	1.01	0.8
G127R/P129D/L217S	0.76	0.78
G127R/P129G	0.57	0.48
G127R/P129R	0.85	0.56
G127R/P129S	0.725	0.725
G127S/P129D	0.67	1.255
G127S/P129F	0.94	0.93
G127S/P129G	1.29	0.99
G127S/P129G/G166D	0.81	0.98
G127S/P129G/I175T	0.33	0.77
G127S/P129R	1.47	0.86
G127S/P129S	1.06	1.09
G127S/P129V	1.16	1.15
G127V/P129G	1.59	0.84
G127V/P129R	1.01	0.89
Z127.01G/Z127.02H/P129D	0.58	0.65
Z127.01G/Z127.02H/P129G	0.55	0.57

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
Z127.01G/Z127.02H/P129R	0.52	0.61
Z127.01H/G128C/P129C/A230V	0.57	0.57
Z127.01H/P129G	0.59	0.36
Z127.01H/P129G/V148I	0.61	0.27
Z127.01H/P129R	0.57	0.44
Z127.01N/G128C/P129V	0.5	0.2
Z127.01N/G128S/P129S	0.585	1.19
Z127.01N/G128S/P129V	0.57	0.67
Z127.01N/P129R	0.59	1.04
Z127.01R/G128S/P129S	0.67	0.5
Z127.01R/P129G	0.485	0.475
Z127.01S/P129D	0.58	0.5
Z127.01S/P129G	0.65	0.67
Z127.01S/P129R	0.42	0.94
G128D/P129G	0.53	1.00
G128D/P129R/Z129.01R	0.7	0.33
G128D/P129S/G154S	0.5	0.54
G128H/P129C	0.58	0.89
G128H/P129H/Z129.01G	0.65	0.53
G128H/P129H/Z129.01R	0.595	0.44
G128H/P129R	0.78	0.97
G128H/P129R/Z129.01G	0.52	0.54
G128H/P129R/Z129.01R	0.57	0.36
G128H/P129S/Z129.01G	0.66	0.54
G128H/P129S/Z129.01R	0.47	0.88
G128H/P129S/Z129.01R/S248N	0.54	0.62
G128H/P129Y	0.44	1.22
G128H/Z128.01R/P129D	0.66	0.63
G128H/Z128.01R/P129S	0.46	0.755
G128H/Z128.01R/P129S/A144T	0.555	0.705
G128H/Z129.01R	0.46	0.76
G128N/P129R/Z129.01G	0.55	0.83
G128N/P129R/Z129.01R	0.5	0.93
G128N/P129S/Z129.01D	0.67	0.93
G128N/Z128.01R/P129S	0.58	1.01
G128R/P129G	0.54	0.57
G128R/P129H/Z129.01G	0.495	0.755
G128R/P129L/Z130.01S	0.58	0.25
G128R/P129R	0.45	0.53
G128R/P129R/Z129.01G	0.55	0.32
G128R/P129R/Z129.01G/A216V	0.64	0.26
G128R/P129S	0.46	0.78

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
G128R/P129S/Z129.01G	0.605	0.7
G128R/P129Z/S130R/S132G	0.43	0.51
G128R/Z128.01R/P129D	0.46	0.57
G128R/Z128.01R/P129F	0.52	0.28
G128S/P129C	0.845	1.09
G128S/P129C/Z129.01R	0.575	0.435
G128S/P129D	0.87	1.37
G128S/P129D/S248R	0.9	1.28
G128S/P129D/T244N	1.3	1.13
G128S/P129G	1.05	1.38
G128S/P129G/A187V	0.75	0.89
G128S/P129H/Z129.01R	0.44	0.92
G128S/P129RL209F	1.2	1.01
G128S/P129R/Z129.01G	0.8	0.76
G128S/P129S/Z129.01D	0.6	0.55
G128S/P129S/Z129.01R	0.59	0.84
G128S/P129V	0.735	1.145
G128Y/P129R/Z129.01G	0.55	0.4
Z128.01G	0.57	0.515
Z128.01G/P129D	0.39	1.04
Z128.01G/P129R	0.46	0.87
Z128.01G/P129S/A134T	0.55	0.62
Z128.01G/Z128.02G	0.59	0.27
Z128.01R/P129D	0.35	0.6
Z128.01R/Z128.02G/P129S	0.5	0.58
Z128.01R/Z128.02H/P129G/G160S	0.59	0.44
Z128.01R/Z128.02R/P129D	0.58	0.74
Z128.01Y/Z128.02H/P129R	0.57	0.34
P129C/S130C/S132Z	0.42	0.6
P129D/Z129.01G/Z129.02D/G160D	0.54	0.61
P129D/Z129.01G/Z129.02G	0.69	0.3
P129G	0.64	0.90
P129G/G131Z	0.57	1.19
P129G/S130G/Z131.01G	0.51	1.02
P129G/S130H/S132Z	0.37	1.17
P129G/S130R/S132Z	0.71	0.3
P129G/V270I	0.7	0.9
P129G/Z129.01G	0.45	0.84
P129G/Z129.01R/Z129.02G	0.69	0.82
P129H	0.79	1
P129H/G131Z	0.6	1.16
P129H/G131Z/S132C	0.52	0.59

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
P129H/G131Z/S132H	0.43	1.04
P129H/S132Z	0.41	0.51
P129H/Z130.01D/Z130.02S	0.44	1.02
P129L	1	1.22
P129R	0.80	0.87
P129R/G131Z/S132H	0.58	0.78
P129R/S130D/S132Z	0.745	1.04
P129R/S130N/G131Z	1.23	0.95
P129R/S132T	0.37	0.69
P129R/S132Z	0.395	0.745
P129R/Z129.01G	0.42	0.60
P129R/Z129.01G/Z129.02G	0.49	0.57
P129R/Z129.01G/Z129.02G/A144T	0.64	0.49
P129R/Z129.01G/Z129.02G/G146D	0.53	0.31
P129R/Z129.01G/Z129.02R	0.52	0.42
P129R/Z129.01G	0.46	0.65
P129R/Z129.01R/Z129.02G	0.58	0.28
P129S	0.92	1.00
P129S/G131Z	0.75	1.15
P129S/S130H/S132Z	0.52	1.19
P129S/Z129.01C	0.55	0.45
P129S/Z129.01R/Z129.02S	0.59	0.82
P129S/Z130.01N/Z130.02S	0.45	1.1
P129V	1.32	1.09
P129V/S132Z	0.37	0.59
P129Z	0.57	1.16
P129Z/G131R/S132C	0.38	0.51
P129Z/S130G	0.47	1.29
P129Z/S130G/G131H/S132H	1.08	1.18
P129Z/S130G/G131V	0.52	0.82
P129Z/S130H	0.56	1.17
P129Z/S130H/S132G	0.42	0.61
P129Z/S130H/S132N	1.91	0.62
P129Z/S130L	0.49	1.14
P129Z/S130R	0.93	1.00
P129Z/S130R/G131C	0.53	0.75
P129Z/S130R/G131C	0.61	0.89
P129Z/S130R/G131D/S132C	0.7	0.75
P129Z/S130R/G131H/A144E	0.86	0.88
P129Z/S130R/G131H/S132D	0.33	0.52
P129Z/S130R/G131R/S132D	0.37	0.64
P129Z/S130R/L267M	0.67	0.99

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
P129Z/S130R/S132C	0.72	0.46
P129Z/S130R/S132G	0.42	0.64
P129Z/S130V/S132G	0.37	0.81
P129Z/S130Z/G131N/S132G	0.5	0.9
S130C	1.25	0.86
S130D/G131C/S132D	0.71	0.81
S130D/G131R	0.77	0.87
S130D/G131R/S132D	0.52	0.9
S130D/G131Z	1.53	0.91
S130D/G131Z/S132H	0.68	0.82
S130F/G131N/S132V	0.59	0.64
S130F/G131S/S132V	0.48	0.59
S130F/S132V	0.29	0.53
S130G	0.8	1.1
S130G/G131D/S132C	1.13	0.81
S130G/G131H/S132D	0.51	0.88
S130G/G131S/S132I	0.4	0.67
S130G/S132G	0.635	0.845
S130H	1.01	0.92
S130H/G131C/S132G	0.94	0.98
S130H/G131H/S132R	0.53	0.41
S130H/G131R/S132C	0.74	0.86
S130H/G131S	1.08	0.75
S130H/G131S/S132C	1.11	0.78
S130H/S132Z	0.81	0.94
S130I/G131S/S132H	0.69	0.77
S130I/S132G	0.94	0.76
S130L	1.06	0.87
S130N/G131S/S132C	1.04	0.91
S130R/G131R/S132C	0.7	0.625
S130R/G131S/S132C/S161N	1.29	0.84
S130R/S132G	1.05	0.36
S130R/S132G/S183N	0.98	0.36
S130R/S132Z	0.48	0.69
S130V/G131D/S132I	0.98	1.23
S130V/G131H/S132C	1.34	0.91
S130V/S132N	0.74	0.91
S130V/S132Z	0.35	0.52
S130Y/G131H/S132C	1.14	0.87
S130Y/S132G	0.97	0.62
S130Z/G131H/S132H	0.78	0.88
G131H/S132L/W241R	0.35	0.61

Table 3-1. Mutations and Performance Index for TCA protein and BMI Activity for BPN'-Y217L Variants

Mutations	TCA PI	PI on BMI at pH 8 16°C
G131N/S132C	0.94	0.93
G131S/S132G	0.42	0.63
G131Y/S132G	0.33	0.615
S132H	0.65	0.88
S132Z	0.795	1.045
A134T	0.66	1.1
A142V	0.55	1.05
A142V/G211V	0.33	1.15
A144T	0.89	0.96
S161N	0.92	1.04
A200T	0.5	0.9
L217Y	1.14	0.95
A231T	0.69	0.95
W241R	0.54	0.95
S248G	0.96	0.95
S260F	0.81	1.04
F261L	0.58	0.86
L267G	1.06	1.08
A272S	1.21	0.92
Q275Z	0.79	0.98

Table 3-2 provides a list of the insertions that make up the combinations.

Table 3-2. Mutations

POS	Variant
66.01	Z066.01G
93.01	Z093.01G
96.01	Z096.01D
96.01	Z096.01H
96.01	Z096.01N
96.01	Z096.01R
96.01	Z096.01S
96.02	Z096.02G
97.01	Z097.01D
97.01	Z097.01G
97.01	Z097.01H
97.01	Z097.01R
97.01	Z097.01S
97.01	Z097.01Y
98.01	Z098.01N
98.01	Z098.01R
99.01	Z099.01H
99.01	Z099.01L
99.01	Z099.01N
99.01	Z099.01R

Table 3-2. Mutations	
POS	Variant
99.01	Z099.01S
99.01	Z099.01V
99.02	Z099.02G
99.02	Z099.02R
100.01	Z100.01G
100.01	Z100.01L
100.01	Z100.01N
100.01	Z100.01R
100.02	Z100.02G
101.01	Z101.01R
101.02	Z101.02G
102.01	Z102.01G
102.02	Z102.02R
102.03	Z102.03G
103.01	Z103.01G
117.01	P117.01G
118.01	Z118.01G
123.01	Z123.01C
126.01	Z126.01S
127.01	Z127.01G
127.01	Z127.01H
127.01	Z127.01N
127.01	Z127.01R
127.01	Z127.01S
127.02	Z127.02H
128.01	Z128.01G
128.01	Z128.01R
128.01	Z128.01Y
128.02	Z128.02G
128.02	Z128.02H
128.02	Z128.02R
129.01	Z129.01C
129.01	Z129.01D
129.01	Z129.01G
129.01	Z129.01R
129.02	Z129.02D
129.02	Z129.02G
129.02	Z129.02R
129.02	Z129.02S
130.01	Z130.01D
130.01	Z130.01N

Table 3-2. Mutations	
POS	Variant
130.01	Z130.01S
131.01	Z131.01G
275	Q275Z

EXAMPLE 4**Construction of Combinatorial Variants of BPN' 3**

[0147] Using BPN'G97A-G128A-Y217Q ("BPN'3") as a parent, combinatorial variants were created by a strategy that involved extension (fusion) PCR using four fragments of the gene. Fragment 1 was amplified by primers P4974 and P4977 (See, Table 4-1). Fragment 4 was amplified by primers 4978 and P4976 (Table 4-1). Both fragment 1 and 4 did not contain any mutations. Fragments 2 and 3 each were created as a set of 12 and 16 sequences respectively that contained the desired mutations, using either BPN'3 as template or in the absence of template using partially overlapping forward and reverse primers.

[0148] The set of Fragment 2 sequences were amplified by the following primers:

- 01: P4979 and P4980 (BPN'3 as a template)
- 02: P4981 and P4982 (no template)
- 03: P4983 and P4984 (no template)
- 04: P4985 and P4986 (no template)
- 05: P4987 and P4988 (no template)
- 06: P4989 and P4990 (no template)
- 07: P4991 and P4992 (no template)
- 08: P4993 and P4994 (no template)
- 09: P4995 and P4996 (no template)
- 10: P4997 and P4998 (no template)
- 11: P4999 and P5000 (no template)
- 12: P5001 and P5002 (no template)

The set of Fragment 3 sequences were amplified by the following primers:

- 01: P5003 and P5004 (BPN'3 as a template)
- 02: P5005 and P5006 (no template)
- 03: P5007 and P5008 (no template)
- 04: P5009 and P5010 (no template)
- 05: P5011 and P5012 (no template)
- 06: P5013 and P5014 (no template)
- 07: P5015 and P5016 (no template)
- 08: P5017 and P5018 (no template)

- 09: P5019 and P5020 (no template)
- 10: P5021 and P5022 (no template)
- 11: P5023 and P5024 (no template)
- 12: P5025 and P5026 (no template)
- 13: P5027 and P5028 (no template)
- 14: P5029 and P5030 (no template)
- 15: P5031 and P5032 (no template)
- 16: P5033 and P5034 (no template)

[0149] The primer sequences used in the construction of the combinatorial variants are listed in Table 4-1.

Table 4-1. Primer Sequences and Mutations Introduced in Combinatorial Variants of BPN'3		
SEQ ID NO:	Primer Name	Sequence
125	P4948	GTGTTTTTCTTGGAAATTGTGCTG
126	P4974	GCCTCACATTTGTGCCACCTA
127	P4975	CATATGAGTTATGCAGTTTGTAG
128	P4976	CCTCTCGGTTATGAGTTAGTTC
129	P4977	GTAGAGCGAAGCAGACGGGGCTA
130	P4978	CTTAAAGCAGCAGTTGATAAAGCT
131	P4979	CTTGGTGTAGCCCCGTCTGCTT
132	P4980	ATCCATGTTATTCGCGATGGCCCATT
133	P4981	CTTGGTGTAGCCCCGTCTGCTTCTGCTCTACGCCCTCTTGCTCTGC AGCAGACGGATCAGGCCAATACTCATGGATTATCAA
134	P4982	ATCCATGTTATTCGCGATGGCCCATTCTGATGCCGTTGATAATCC ATGAGTATTGGCCTGATCCGTCGCTGCAGAGCAAG
135	P4983	CTTGGTGTAGCCCCGTCTGCTTCTGCTCTACGCCGTTAAAGTTCT TGATGCACGTGACGGATCAGGCCAATACTCATGGAT
136	P4984	ATCCATGTTATTCGCGATGGCCCATTCTGATGCCGTTGATAATCC ATGAGTATTGGCCTGATCCGTCACGTGCATCAAGAA
137	P4985	CTTGGTGTAGCCCCGTCTGCTTCTGCTCTACGCCGTTAAAGTTCT TGCAGCAGACTCTGGATCAGGCCAATACTCATGGAT
138	P4986	ATCCAIGTATTCGCGATGGCCCATTCGATGCCGTTGATAATCC ATGAGTATTGGCCTGATCCAGAGTCTGCTGCAAGAA
139	P4987	CTTGGTGTAGCCCCGTCTGCTTCTGCTCTACGCCGTTAAAGTTCT TGCAGCAGACTCTTCAGGCCAATACTCATGGATTAT

Table 4-1. Primer Sequences and Mutations Introduced in Combinatorial Variants of BPN'3

SEQ ID NO:	Primer Name	Sequence
140	P4988	ATCCAIGTATATTCGCGATGGCCCATTCGATGCCGTTGATAAATCC ATGAGTATTTGGCCTGAAGAGTCTGCTGCAAGAACTT
141	P4989	CTTGGTGTAGCCCCGTCTGCTTCGCTCTACGCCGTTAAAGTTCT TGCAGCAGACGGAGATGGCCAATACTCATGGATTAT
142	P4990	ATCCATGTTATTTCGCGATGGCCCATTCGATGCCGTTGATAATCC ATGAGTATTTGGCCATCTCCGTCTGCTGCAAGAACTT
143	P4991	CTTGGTGTAGCCCCGTCTGCTTCGCTCTACGCCGTTAAAGTTCT TGATGCACGTGACTCTGGATCAGGCCAATACTCATG
144	P4992	ATCCAIGTATATTCGCGATGGCCCATTCGATGCCGTTGATAAATCC ATGAGTATTTGGCCTGATCCAGAGTCACGTGCATCAA
145	P4993	CTTGGTGTAGCCCCGTCTGCTTCGCTCTACGCCGTTAAAGTTCT TGATGCACGTGACTCTTCAGGCCAATACTCATGGAT
146	P4994	ATCCAIGTATATTCGCGATGGCCCATTCGATGCCGTTGATAAATCC ATGAGTATTTGGCCTGAAGAGTCACGTGCATCAAGAA
147	P4995	CTTGGTGTAGCCCCGTCTGCTTCGCTCTACGCCGTTAAAGTTCT TGATGCACGTGACGGAGATGGCCAATACTCATGGAT
148	P4996	ATCCAIGTATATTCGCGATGGCCCATTCGATGCCGTTGATAATCC ATGAGTATTTGGCCATCTCCGTACGTGCATCAAGAA
149	P4997	CTTGGTGTAGCCCCGTCTGCTTCGCTCTACGCCGTTAAAGTTCT TGCAGCAGACTCTTCTTCAGGCCAATACTCATGGAT
150	P4998	ATCCAIGTATATTCGCGATGGCCCATTCGATGCCGTTGATAATCC ATGAGTATTTGGCCTGAAGAAGAGTCTGCTGCAAGAA
151	P4999	CTTGGTGTAGCCCCGTCTGCTTCGCTCTACGCCGTTAAAGTTCT TGCAGCAGACTCTGGAGATGGCCAATACTCATGGAT
152	P5000	ATCCATGTTATTTCGCGATGGCCCATTCGATGCCGTTGATAATCC ATGAGTATTTGGCCATCTCCAGAGTCTGCTGCAAGAA
153	P5001	CTTGGTGTAGCCCCGTCTGCTTCGCTCTACGCCGTTAAAGTTCT TGCAGCAGACTCTGATGGCCAATACTCATGGATTAT
154	P5002	ATCCAIGTATATTCGCGATGGCCCATTCGATGCCGTTGATAAATCC ATGAGTATTTGGCCATCAGAGTCTGCTGCAAGAACTT

Table 4-1. Primer Sequences and Mutations Introduced in Combinatorial Variants of BPN'3

SEQ ID NO:	Primer Name	Sequence
155	P5003	ATCGAATGGGCCATCGCGAAT
156	P5004	TGCAACAGCTTTATCAACTGCT
157	P5005	ATCGAATGGGCCATCGCGAATAACATGGATGTATCTGGAATGA GCCTGGGAGCACCAAGCGGCAGT
158	P5006	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGCCGCACTGCCG CTTGGTGCTCCAGGCTCATTCAGAT
159	P5007	ATCGAATGGGCCATCGCGAATAACATGGATGTAATCAACATGA GCCTGGGATCTCCAAGCGGCAGT
160	P5008	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGCCGCACTGCCG CTTGGAGATCCAGGCTCATGTTGATT
161	P5009	ATCGAATGGGCCATCGCGAATAACATGGATGTAATCAACATGA GCCTGGGAGCAGTTAGCGGCAGT
162	P5010	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGCCGCACTGCCG CTAACTGCTCCAGGCTCATGTTGATT
163	P5011	ATCGAATGGGCCATCGCGAATAACATGGATGTAATCAACATGA GCCTGGGAGCAGATAGCGGCAGT
164	P5012	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGCCGCACTGCCG CTATCTGCTCCAGGCTCATGTTGATT
165	P5013	ATCGAATGGGCCATCGCGAATAACATGGATGTAATCAACATGA GCCTGGGAGCAAGCGGCAGTGC GGCA
166	P5014	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGCCGCACTGCCG CTTGCTCCAGGCTCATGTTGATTACA
167	P5015	ATCGAATGGGCCATCGCGAATAACATGGATGTAATCAACATGA GCCTGGGAGCAGGAGGCAGTGC GGCA
168	P5016	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGCCGCACTGCCCTC CTGCTCCAGGCATGTTGATTACA
169	P5017	ATCGAATGGGCCATCGCGAATAACATGGATGTAATCAACATGA GCC TGGGAGCACCAGTTGATAT
170	P5018	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGCCGCGATATCA ACTGGTGCTCCAGGCTCATGTTGATT

Table 4-1. Primer Sequences and Mutations Introduced in Combinatorial Variants of BPN'3

SEQ ID NO:	Primer Name	Sequence
171	P5019	ATCGAATGGGCCATCGCGAATAACATGGATGTAAATCAACATGA GCCTGGGAGCACCAAGCGGCAGT
172	P5020	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGTGCGCACTGCCG TTGGTGTCTCCAGGCTCATGTTGATT
173	P5021	ATCGAATGGGCCATCGCGAATAACATGGATGTATCTGGAATGA GCCTGGGATCTCCAAGCGGCAGT
174	P5022	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGCCGCACTGCCG CTTGGAGATCCCAGGCTCATTCAGAT
175	P5023	ATCGAATGGGCCATCGCGAATAACATGGATGTATCTGGAATGA GCCTGGGAGCAGTAAGCGGCAGT
176	P5024	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGCCGCACTGCCG CTTACTGCTCCCAGGCTCATTCAGAT
177	P5025	ATCGAATGGGCCATCGCGAATAACATGGATGTATCTGGAATGA GCCTGGGAGCAGATAGCGGCAGT
178	P5026	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGCCGCACTGCCG CTATCTGCTCCCAGGCTCATTCAGAT
179	P5027	ATCGAATGGGCCATCGCGAATAACATGGATGTATCTGGAATGA GCCTGGGAGCAAGCGGCAGTGCGGCA
180	P5028	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGCCGCACTGCCG CTTGTCTCCAGGCTCATTCAGATACATCCATGTT
181	P5029	ATCGAATGGGCCATCGCGAATAACATGGATGTAAATCAACATGA GCCTGGGATCTGTTAGCGGCAGT
182	P5030	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGCCGCACTGCCG CTAACAGATCCCAGGCTCATGTTGATT
183	P5031	ATCGAATGGGCCATCGCGAATAACATGGATGTAAATCAACATGA GCCTGGGATCTGATAGCGGCAGT
184	P5032	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGCCGCACTGCCG CTATCAGATCCCAGGCTCATGTTGATT
185	P5033	ATCGAATGGGCCATCGCGAATAACATGGATGTAAATCAACATGA GCCTGGGATCTAGCGGCAGTGCGGCA

Table 4-1. Primer Sequences and Mutations Introduced in Combinatorial Variants of BPN'3		
SEQ ID NO:	Primer Name	Sequence
186	P5034	TGCAACAGCTTTATCAACTGCTGCTTTAAGTGGCGCACTGCCG CTAGATCCCAGGCTCATGTGATT
187	P5035	TGATCCGTCTGAGCAGGGGCTTTAATTTGTGA
188	P5036	ATTAAAGCCCCCTGCTCAGACGGATCAGGCCAATACTC

[0150] Each amplification reaction contained 30 pmol of each oligonucleotide and 100 ng of the template DNA. Amplifications were carried out using Vent DNA polymerase (New England Biolabs). The PCR mix (20 μ l) was initially heated at 95°C for 2.5 min followed by 30 cycles of denaturation at 94°C for 15s, annealing at 55°C for 15s and extension at 72°C for 40s. Following amplification all the fragments were purified by the Gel Band Purification kit (Qiagen) and mixed to serve as templates for a fusion PCR using primers P4975 and P4948. The full-length DNA fragment from the fusion PCR reaction was gel-purified by the QIAGEN gel-band purification kit, digested by the BamHI and HindIII restriction enzymes and ligated in the *Bacillus* expression plasmid pHPLT-BPN partial opt (Figure 2C), which was cut with the same restriction enzymes.

[0151] Ligation mixtures were amplified using rolling circle amplification according to manufacturer's recommendation (Epicentre). One μ l of the ligation mixture was mixed with 5 μ l of the sample buffer, heated to 95°C for 3 min and cooled on ice. Next, 5 μ l of the reaction buffer and 0.2 μ l of the enzyme was added to each tube, followed by incubation at 30°C for 10 hours. Products of the rolling circle amplification were diluted 100 times and 1 μ l of this dilution was used to transform *Bacillus subtilis* cells (genotype: $\Delta aprE$, $\Delta nprE$, $\Delta spoII E$, *nmvE::xyRPxylAcomK-phleo*) as described in Example 2. Transformations were plated on LB plates containing 1.6% skim milk and 10 ppm Neomycin and incubated overnight at 37°C. Only colonies with halos were picked and grown in microtiter plates for further analysis as described in Example 2. Protein concentration of culture supernatants was determined by TCA precipitation as described in Example 1.

Table 4.5: Combinatorial Libraries of BPN': G97A-G128A-Y217Q Parent	
Variant	Name
1	V093Z/K094S/V095C/L096S/G097A/G128A/Y217Q
2	Z096.01D/G097G/A098R/G128A/Y217Q
3	G097A/Z099.01S/G128A/Y217Q
4	G097A/G100S/G128A/Y217Q
5	G097A/S101D/G128A/Y217Q
6	Z096.01D/G097G/A098R/Z099.01S/G128A/Y217Q
7	Z096.01D/G097G/A098R/G100S/G128A/Y217Q
8	Z096.01D/G097G/A098R/S101D/G128A/Y217Q
9	G097A/Z099.01S/G100S/G128A/Y217Q
10	G097A/Z099.01S/S101D/G128A/Y217Q
11	G097A/G100S/S101D/G128A/Y217Q
12	G097A/I122S/N123G/G128A/Y217Q
13	V093Z/K094S/V095C/L096S/G097A/I122S/N123G/G128A/Y217Q
14	Z096.01D/G097G/A098R/I122S/N123G/G128A/Y217Q
15	G097A/Z099.01S/I122S/N123G/G128A/Y217Q
16	G097A/G100S/I122S/N123G/G128A/Y217Q
17	G097A/S101D/I122S/N123G/G128A/Y217Q
18	Z096.01D/G097G/A098R/Z099.01S/I122S/N123G/G128A/Y217Q
19	Z096.01D/G097G/A098R/G100S/I122S/N123G/G128A/Y217Q
20	Z096.01D/G097G/A098R/S101D/I122S/N123G/G128A/Y217Q
21	G097A/Z099.01S/G100S/I122S/N123G/G128A/Y217Q
22	G097A/Z099.01S/S101D/I122S/N123G/G128A/Y217Q

Table 4.5: Combinatorial Libraries of BPN': G97A-G128A-Y217Q Parent

Variant	Name
23	G097A/G100S/S101D/I122S/N123G/G128A/Y217Q
24	G097A/G128S/Y217Q
25	V093Z/K094S/V095C/L096S/G097A/G128S/Y217Q
26	Z096.01D/G097G/A098R/G128S/Y217Q
27	G097A/Z099.01S/G128S/Y217Q
28	G097A/G100S/G128S/Y217Q
29	G097A/S101D/G128S/Y217Q
30	Z096.01D/G097G/A098R/Z099.01S/G128S/Y217Q
31	Z096.01D/G097G/A098R/G100S/G128S/Y217Q
32	Z096.01D/G097G/A098R/S101D/G128S/Y217Q
33	G097A/Z099.01S/G100S/G128 S/Y217Q
34	G097A/Z099.01S/S101D/G128 S/Y217Q
35	G097A/G100S/S101D/G128S/Y217Q
36	G097A/G128A/P129V/Y217Q
37	V093Z/K094S/V095C/L096S/G097A/G128A/P129V/Y217Q
38	Z096.01D/G097G/A098R/G128A/P129V/Y217Q
39	G097A/Z099.01S/G128A/P129V/Y217Q
40	G097A/G100S/G128A/P129V/Y217Q
41	G097A/S101D/G128A/P129V/Y217Q
42	Z096.01D/G097G/A098R/Z099.01S/G128A/P129V/Y217Q
43	Z096.01D/G097G/A098R/G100S/G128A/P129V/Y217Q
44	Z096.01D/G097G/A098R/S101D/G128A/P129V/Y217Q
45	G097A/Z099.01S/G100S/G128A/P129V/Y217Q
46	G097A/Z099.01S/S101D/G128A/P129V/Y217Q
47	G097A/G100S/S101D/G128A/P129V/Y217Q
48	G097A/G128A/P129D/Y217Q
49	V093Z/K094S/V095C/L096S/G097A/G128A/P129D/Y217Q
50	Z096.01D/G097G/A098R/G128A/P129D/Y217Q
51	G097A/Z099.01S/G128A/P129D/Y217Q
52	G097A/G100S/G128A/P129D/Y217Q
53	G097A/S101D/G128A/P129D/Y217Q
54	Z096.01D/G097G/A098R/Z099.01S/G128A/P129D/Y217Q
55	Z096.01D/G097G/A098R/G100S/G128A/P129D/Y217Q
56	Z096.01D/G097G/A098R/S101D/G128A/P129D/Y217Q
57	G097A/Z099.01S/G100S/G128A/P129D/Y217Q
58	G097A/Z099.01S/S101D/G128A/P129D/Y217Q
59	G097A/G100S/S101D/G128A/P129D/Y217Q
60	G097A/G128A/P129Z/Y217Q
61	V093Z/K094S/V095C/L096S/G097A/G128A/P129Z/Y217Q
62	Z096.01D/G097G/A098R/G128A/P129Z/Y217Q
63	G097A/Z099.01S/G128A/P129Z/Y217Q

Table 4.5: Combinatorial Libraries of BPN': G97A-G128A-Y217Q Parent

Variant	Name
64	G097A/G100S/G128A/P129Z/Y217Q
65	G097A/S101D/G128A/P129Z/Y217Q
66	Z096.01D/G097G/A098R/Z099.01S/G128A/P129Z/Y217Q
67	Z096.01D/G097G/A098R/G100S/G128A/P129Z/Y217Q
68	Z096.01D/G097G/A098R/S101D/G128A/P129Z/Y217Q
69	G097A/Z099.01S/G100S/G128A/P129Z/Y217Q
70	G097A/Z099.01S/S101D/G128A/P129Z/Y217Q
71	G097A/G100S/S101D/G128A/P129Z/Y217Q
72	G097A/G128A/P129G/S130Z/Y217Q
73	V093Z/K094S/V095C/L096S/G097A/G128A/P129G/S 130Z/Y217Q
74	Z096.01D/G097G/A098R/G128A/P129G/S130Z/Y217Q
75	G097A/Z099.01S/G128A/P129G/S130Z/Y217Q
76	G097A/G100S/G128A/P129G/S130Z/Y217Q
77	G097A/S101D/G128A/P129G/S130Z/Y217Q
78	Z096.01D/G097G/A098R/Z099.01S/G128A/P129G/S130Z/Y217Q
79	Z096.01D/G097G/A098R/G100S/G128A/P129G/S130Z/Y217Q
80	Z096.01D/G097G/A098R/S101D/G128A/P129G/S130Z/Y217Q
81	G097A/Z099.01S/G100S/G128A/P129G/S130Z/Y217Q
82	G097A/Z099.01S/S101D/G128A/P129G/S130Z/Y217Q
83	G097AG100S/S101D/G128A/P129G/S130Z/Y217Q
84	G097A/G128A/S130V/G131D/S132I/Y217Q
85	V093Z/K094S/V095C/L096S/G097A/G128A/S130V/G131D/S132I/Y217Q
86	Z096.01D/G097G/A098R/G128A/S130V/G131D/S132I/Y217Q
87	G097A/Z099.01S/G128A/S130V/G131D/S132I/Y217Q
88	G097A/G100S/G128A/S130V/G131D/S132I/Y217Q
89	G097A/S101D/G128A/S130V/G131D/S132I/Y217Q
90	Z096.01D/G097G/A098R/Z099.01S/G128A/S130V/G131D/S132I/Y217Q
91	Z096.01D/G097G/A098R/G100S/G128A/S130V/G131D/S132I/Y217Q
92	Z096.01D/G097G/A098R/S101D/G128A/S130V/G131D/S132I/Y217Q
93	G097A/Z099.01S/G100S/G128A/S130V/G131D/S132I/Y217Q
94	G097A/Z099.01S/S101D/G128A/S130Y/G131D/S132I/Y217Q
95	G097A/G100S/S101D/G128A/S130V/G131D/S132I/Y217Q
96	G097A/G128A/A134T/Y217Q
97	V093Z/K094S/V095C/L096S/G097A/G128A/A134T/Y217Q
98	Z096.01D/G097G/A098R/G128A/A134T/Y217Q
99	G097A/Z099.01S/G128A/A134T/Y217Q
100	G097A/G100S/G128A/A134T/Y217Q
101	G097A/S101D/G128A/A134T/Y217Q
102	Z096.01D/G097G/A098R/Z099.01S/G128A/A134T/Y217Q
103	Z096.01D/G097G/A098R/G100S/G128A/A134T/Y217Q
104	Z096.01D/G097G/A098R/S101D/G128A/A134T/Y217Q
105	G097A/Z099.01S/G100S/G128A/A134T/Y217Q

Table 4.5: Combinatorial Libraries of BPN': G97A-G128A-Y217Q Parent

Variant	Name
106	G097A/Z099.01S/S101D/G128A/A134T/Y217Q
107	G097A/G100S/S101D/G128A/A134T/Y217Q
108	G097A/I122S/N123G/G128S/Y217Q
109	V093Z/K094S/V095C/L096S/G097A/I122S/N123G/G128S/Y217Q
110	Z096.01D/G097G/A098R/I122S/N123G/G128S/Y217Q
111	G097A/Z099.01S/I122S/N123G/G128 S/Y217Q
112	G097A/G100S/I122S/N123G/G128S/Y217Q
113	G097A/S101D/I122S/N123G/G128S/Y217Q
114	Z096.01D/G097G/A098R/Z099.01S/I122S/N123G/G128S/Y217Q
115	Z096.01D/G097G/A098R/G100S/I122S/N123G/G128S/Y217Q
116	Z096.01D/G097G/A098R/S101D/I122S/N123G/G128S/Y217Q
117	G097A/Z099.01S/G100S/I122S/N123G/G128S/Y217Q
118	G097A/Z099.01SS101D/I122S/N123G/G128S/Y217Q
119	G097A/G100S/S101D/I122S/N123G/G128S/Y217Q
120	G097A/I122S/N123G/G128A/P129V/Y217Q
121	V093Z/K094S/V095C/L096S/G097A/I122S/N123G/G128A/P129V/Y217Q
122	Z096.01D/G097G/A098R/I122S/N123G/G128A/P129V/Y217Q
123	G097A/Z099.01S/I122S/N123G/G128A/P129V/Y217Q
124	G097A/G100S/I122S/N123G/G128A/P129V/Y217Q
125	G097A/S101D/I122S/N123G/G128A/P129V/Y217Q
126	Z096.01D/G097G/A098R/Z099.01S/I122S/N123G/G128A/P129V/Y2170
127	Z096.01D/G097G/A098R/G100S/I122S/N123G/G128A/P129V/Y2170
128	Z096.01D/G097G/A098R/S101D/I122S/N123G/G128A/P129V/Y2170
129	G097A/Z099.01S/G100S/I122S/N123G/G128A/P129V/Y2170
130	G097A/Z099.01S/S101D/I122S/N123G/G128A/P129V/Y2170
131	G097A/G100S/S101D/I122S/N123G/G128A/P129V/Y217Q
132	G097A/I122S/N123G/G128A/P129D/Y217Q
133	V093Z/K094S/V095C/L096S/G097A/I122S/N123G/G128A/P129D/Y217Q
134	Z096.01D/G097G/A098R/I122S/N123G/G128A/P129D/Y217Q
135	G097A/Z099.01S/I122S/N123G/G128A/P129D/Y217Q
136	G097A/G100S/I122S/N123G/G128A/P129D/Y217Q
137	G097A/S101D/I122S/N123G/G128A/P129D/Y217Q
138	Z096.01D/G097G/A098R/Z099.01S/I122S/N123G/G128A/P129D/Y2170
139	Z096.01D/G097G/A098R/G100S/I122S/N123G/G128A/P129D/Y2170
140	Z096.01D/G097G/A098R/S101D/I122S/N123G/G128A/P129D/Y2170
141	G097A/Z099.01S/G100S/I122S/N123G/G128A/P129D/Y2170
142	G097A/Z099.01S/S101D/I122S/N123G/G128A/P129D/Y2170
143	G097A/G100S/S101D/I122S/N123G/G128A/P129D/Y217Q
144	G097A/I122S/N123G/G128A/P129Z/Y217Q
145	V093Z/K094S/V095C/L096S/G097A/I122S/N123G/G128A/P129Z/Y217Q
146	Z096.01D/G097G/A098R/I122S/N123G/G128A/P129Z/Y217Q
147	G097A/Z099.01S/I122S/N123G/G128A/P129Z/Y217Q

Table 4.5: Combinatorial Libraries of BPN': G97A-G128A-Y217Q Parent

Variant	Name
148	G097A/G100S/I122S/N123G/G128A/P129Z/Y217Q
149	G097A/S101D/I122S/N123G/G128A/P129Z/Y217Q
150	Z096.01D/G097G/A098R/Z099.01S/I122S/N123G/G128A/P129Z/Y217Q
151	Z096.01D/G097G/A098R/G100S/I122S/N123G/G128A/P129Z/Y217Q
152	Z096.01D/G097G/A098R/S101D/I122S/N123G/G128A/P129Z/Y217Q
153	G097A/Z099.01S/G100S/I122S/N123G/G128A/P129Z/Y217Q
154	G097A/Z099.01S/S101D/I122S/N123G/G128A/P129Z/Y217Q
155	G097A/G100S/S101D/I122S/N123G/G128A/P129Z/Y217Q
156	G097A/G128S/P129V/Y217Q
157	V093Z/K094S/V095C/L096S/G097A/G128S/P129V/Y217Q
158	Z096.01D/G097G/A098R/G128S/P129V/Y217Q
159	G097A/Z099.01S/G128S/P129V/Y217Q
160	G097A/G100S/G128S/P129V/Y217Q
161	G097A/S101D/G128S/P129V/Y217Q
162	Z096.01D/G097G/A098R/Z099.01S/G128S/P129V/Y217Q
163	Z096.01D/G097G/A098R/G100S/G128S/P129V/Y217Q
164	Z096.01D/G097G/A098R/S101D/G128S/P129V/Y217Q
165	G097A/Z099.01S/G100S/G128S/P129V/Y217Q
166	G097A/Z099.01S/S101D/G128S/P129V/Y217Q
167	G097A/G100S/S101D/G128S/P129V/Y217Q
168	G097A/G128S/P129D/Y217Q
169	V093Z/K094S/V095C/L096S/G097A/G128S/P129D/Y217Q
170	Z096.01D/G097G/A098R/G128S/P129D/Y217Q
171	G097A/Z099.01 S/G128 S/P129D/Y217Q
172	G097A/G100S/G128S/P129D/Y217Q
173	G097A/S101D/G128S/P129D/Y217Q
174	Z096.01D/G097G/A098R/Z099.01S/G128S/P129D/Y217Q
175	Z096.01D/G097G/A098R/G100S/G128S/P129D/Y217Q
176	Z096.01D/G097G/A098R/S101D/G128S/P129D/Y217Q
177	G097A/Z099.01S/G100S/G128S/P129D/Y217Q
178	G097A/Z099.01S/S101D/G128S/P129D/Y217Q
179	G097A/G100S/S101D/G128S/P129D/Y217Q
180	G097A/G128S/P129Z/Y217Q
181	V093Z/K094S/V095C/L096S/G097A/G128S/P129Z/Y217Q
182	Z096.01D/G097G/A098R/G128S/P129Z/Y217Q
183	G097A/Z099.01S/G128S/P129Z/Y217Q
184	G097A/G100S/G128S/P129Z/Y217Q
185	G097A/S101D/G128S/P129Z/Y217Q
186	Z096.01D/G097G/A098R/Z099.01S/G128S/P129Z/Y217Q
187	Z096.01D/G097G/A098R/G100S/G128S/P129Z/Y217Q
188	Z096.01D/G097G/A098R/S101D/G128S/P129Z/Y217Q
189	G097A/Z099.01S/G100S/G128S/P129Z/Y217Q

Table 4.5: Combinatorial Libraries of BPN': G97A-G128A-Y217Q Parent

Variant	Name
190	G097A/Z099.01S/S101D/G128S/P129Z/Y217Q
191	G097AG100S/S101D/G128S/P129Z/Y217Q

EXAMPLE 5**Stain Removal Performance of Combinatorial Variants of BPN' 3**

[0152] The stain removal activity of combinatorial variants of BPN' 3 generated as described in Example 4 was tested as described in Example 3. As described throughout, functionality of BPN' variants, was quantified as a performance index (P_i), which is the ratio of performance of a variant to parent protein BPN'3. BPN'3 variants showing a P_i value greater or equal than 0.5 for BMI stain removal performance and/or TCA precipitation showed improved cleaning benefits and/or expression. Performance indices less than or equal to 0.05 were fixed to 0.05 and indicated in bold italics in the table. Results are shown in Table 5-1.

Table 5-1: Stain Removal Performance and Expression of Combinatorial Variants of BPN'3

Variants	TCA	BMI pH 8 /16°C
V072G/G097A/G128A/Y217Q	0.93	1.08
G097A/G128S/Y217Q	1.16	1.06
G128A/Y217Q	1.07	1.05
G097A/G128S/P129D/Y217Q	0.85	1.03
G097A/G128A/A134T/Y217Q	0.56	1.03
M124V/L126A/Y217Q	1.05	1.03
G097A/G128A/P129D/Y217Q/A232P	0.46	1.01
G097A/G128A/P129D/Y217Q	0.57	1.00
G097A/G128S/P129V/A134T/Y217Q	0.29	0.95
G097A/G128S/P129V/Y217Q	0.31	0.95
V068I/G097A/G128S/P129D/Y217Q	0.41	0.85
G097A/S101D/G128S/S204F/Y217Q/S236F/T254P	0.41	0.79
G097A/S101D/G128S/Y217Q	0.43	0.76
G097A/G128A/P129V/Y217Q	0.30	0.70
G097A/S101D/G128A/Y217Q	0.50	0.64
G097A/S101D/G128S/P129D/Y217Q	0.35	0.63
G097A/S101D/G128A/A134T/Y217Q	0.40	0.58
G097A/Z099.01S/A114S/G128S/Y217Q	0.21	0.53
G097A/S101D/G128A/P194L/Y217Q	0.35	0.49
G097A/Z099.01S/G128S/Y217Q	0.39	0.38
G097A/S101D/G128A/P129D/Y217Q	0.36	0.36
Q019R/G097A/Z099.01S/G128S/P129D/Y217Q	0.33	0.33
G097A/G100S/G128S/Y217Q	0.46	0.25
G097A/S101D/G128S/P129V/Y217Q	0.39	0.22
G097A/Z099.01S/G128A/A134T/Y217Q	0.35	0.16
G097A/G100S/I107V/G128S/P129D/Y217Q	0.37	0.15
G097A/G100S/G128S/Y217Q/A230V	0.38	0.13
G097A/G100S/G128A/Y217Q/K237N	0.39	0.12

Table 5-1: Stain Removal Performance and Expression of Combinatorial Variants of BPN'3		
Variants	TCA	BMI pH 8 /16°C
G097A/Z099.01S/G128S/P129D/Y217Q	0.37	0.12
G097A/G100S/G128S/P129D/Y217Q	0.37	0.07
Z096.01D/G097A/A098R/G128A/Y217Q	0.36	0.05
G097A/Z099.01S/I122S/N123G/G128A/Y217Q	0.37	0.05
G097A/Z099.01S/I122S/N123G/G128A/Y217Q/T220A/A232T	0.38	0.05
P005S/G097A/Z099.01S/S101D/G128A/Y217Q	0.42	0.05
G097A/S101D/G128A/P129V/Y217Q	0.46	0.05
V093Z/K094S/V095C/L096S/G097A/G128A/Y217Q	0.50	0.05
G097A/G100S/S101D/G128A/Y217Q/A232P	0.51	0.05
Z096.01D/G097A/A098R/S101D/G128A/Y217Q	0.53	0.05
G097A/Z099.01S/G100S/G128A/Y217Q	0.53	0.05

EXAMPLE 6**Construction of Combinatorial Variants of BPN' 3 at Positions 128,129, and 130**

[0153] Combinatorial libraries focused around positions 128/129/130 in BPN'G97A-G128A-Y217Q (BPN'3) were created using site specific saturation mutagenesis and/or site specific insertion and/or deletion. The variants generated contain substitution, insertion, and deletion mutations. Libraries L1, L2 and L3 are based on WT position S130S and L4 is based on S130A. Four saturation libraries focused around positions 128/129 and 130 were constructed as shown in Table 6-1.

Table 6-1: Construction of Saturation Library at Positions 128, 129 and 130 in BPN'3						
Library	Position 128	Insertion	Position 129	Position 130	Short Name	Degenerate Primer Names
L1	NNS		NNS	S	XX	128/129 deg QC F/R
L2	deletion		NNS	S	-X	128/129 del QC F/R
L3	NNS	A	NNS	S	XAX	128 A 129 QC F/R
L4	deletion		NNS	A	-XA	128/129 del A QC F/R

[0154] Prior to library construction, two stop codons were introduced at positions 128 and 129 in the BPN'3' sequence to inactivate the protease. This was done to ensure a low background of the parent plasmid in the libraries. The parent plasmid (pHPLT-BPN'3) was methylated using two micrograms of plasmid DNA and methylase (New England Biolabs), according to the NEB protocol. Methylated DNA was then purified with DNA Clean and Concentrator kit from Zymo Research. Mutagenesis was performed using methods known in the art (See, Amin et al., Biotechniques 35: 1134-1140, [2003]) using a modified version of the QuikChange multi site-directed mutagenesis (QCMS) kit from Stratagene.

[0155] The mutagenesis reaction contained 60 ng template plasmid, 0.5 ul forward primer 128/129 Stop F (25 uM), 0.5 ul reverse primer 128/129 Stop R (25 uM), 1 ul dNTP's (QCMS kit), 2.5 ul 10x QCMS reaction buffer, 18.5 ul deionized water, and 1 ul of enzyme blend (QCMS kit), for a total volume of 25ul. For the mutagenesis reaction, the thermocycler program used was 1 cycle at 95° for 1 min., followed by 25 cycles of 95°C for 1 min., 55°C for 1 min., and 68°C for 11 minutes (MJ Research thermocycler). The template DNA was digested by DpnI (QCMS kit, Qiagen) and 1.5uL of the reaction was amplified by rolling circle amplification (RCA) using the Templiphi kit (Amersham) using manufacturer's protocol.

Table 6-2. Primers		
SEQ ID NO:	Primer Name	Primer Sequence

Table 6-2. Primers		
SEQ ID NO:	Primer Name	Primer Sequence
189	128/129 Stop F	/5Phos/CATGAGCCTGGGATAATGAA GCGGCAGTGCGGCACTTAAAGCAG
190	128/129 Stop R	/5Phos/GTGGCCGCACTGCCGCTTCAT TATCCCAGGCTCATGTTGATTACATC

[0156] One microliter of the amplified DNA was used to transform 100 μ L of competent *Bacillus subtilis* cells (genotype: $\Delta apr-E$, $\Delta nprE$, $\Delta spoII E$, *amyE::xyIRPxy/AcomK-phleo*). Aliquots of 20 μ L or 80 μ L of the transformation mixture were plated on Luria Agar plates supplemented with 10 μ g/ml neomycin + 1.6% skim milk (Teknova). After growth, 4 colonies not forming halos on the skim milk plates were picked to identify the plasmid with the correct sequence. The correct variant with the two stop codons at positions 128/129, called "pHPLT-BPN 128/129 Stop" was chosen as the parent plasmid for the construction of combinatorial libraries described in Table 6-1.

[0157] Plasmid pHPLT-BPN 128/129 Stop was methylated and used as template with the degenerate primer pairs shown in Table 6-3.

Table 6-3: NNS Mutagenesis Primer Sequences		
SEQ ID NO.	Primer Name	Sequence
191	128/129 deg QC F	/5Phos/CAT GAG CCT GGG ANN SNN SAG CGG CAG TGC GGC ACT TAA AGC AG
192	128/129 deg QC R	/5Phos/CCG CAC TGC CGC TSN NSN NTC CCA GGC TCA TGT TGA TTA CAT C
193	128/129 del QC F	/5Phos/CAT GAG CCT GGG ANN SAG CGG CAG TGC GGC ACT TAA AGC AG
194	128/129 del QC R	/5Phos/CCG CAC TGC CGC TSN NTC CCA GGC TCA TGT TGA TTA CAT C
195	128 A 129 QC F	/5Phos/CAT GAG CCT GGG ANN SGC GNN SAG CGG CAG TGC GGC ACT TAA AGC AG
196	128 A 129 QC R	/5Phos/CCG CAC TGC CGC TSN NCG CSN NTC CCA GGC TCA TGT TGA TTA CAT C
197	128/129 del A QC F	/5Phos/CAT GAG CCT GGG ANN SGC AGG CAG TGC GGC ACT TAA AGC AG
198	128/129 del A QC R	/5Phos/GTG CCG CAC TGC CTG CSN NTC CCA GGC TCA TGT TGA TTA CAT C

[0158] Overlapping forward and reverse primers (PAGE-purified) with the NNS codon(s) in the middle and 17 - 20 flanking bases were designed for each chosen library, and the sequences for each forward (F) and reverse (R) primer are shown in Table 6-3.

Quik-change mutagenesis and rolling circle amplification for the 4 reactions was performed as described above. Transformations were plated on Luria Agar plates supplemented with 10 µg/ml neomycin + 1.6% skim milk. All halo forming colonies were picked into two microtiter plates containing 125ul LB medium supplemented with 10ug/ml neomycin. The clones were sequenced by Quintara Biosciences. For protein expression, cultures were grown overnight in micro-filter plates (.22um, Millipore) containing 180ul of a enriched semi-defined media based on MOPs buffer, with urea as major nitrogen source, glucose as the main carbon source, 1% soytone for robust cell growth, and supplemented with 2.5ug/ml neomycin. Cultures were grown for 64 hours at 37C, 250rpm, and 70% humidity. Protein concentration of culture supernatants was determined by TCA precipitation as described in Example 1.

EXAMPLE 7

Stain Removal Performance of Combinatorial Variants of BPN' 3 at positions 128, 129, and 130

[0159] The stain removal activity of combinatorial variants of BPN' 3 at positions 128, 129, and 130 generated as described in Example 6 was tested as described in Example 3. As described throughout, functionality of BPN' variants, was quantified as a performance index (*P_i*), which is the ratio of performance of a variant to parent protein BPN'3. BPN'3 variants showing a *P_i* value greater or equal than 0.5 for BMI stain removal performance and/or TCA precipitation showed improved cleaning benefits and/or expression. Results are shown in Table 7-1.

Table 7-1: Stain Removal Performance and Expression of BPN' 3 Variants at Positions 128,129, and 130		
Variant	TCA	BMI
G097A/G128S/P129T/Y217Q	0.77	1.04
G097A/G128A/P129S/Y217Q	0.70	1.02
G097A/G128S/P129C/Y217Q	0.61	1.00
G097A/G128S/P129Q/Y217Q	0.96	1.00
G097A/G128S/P129E/Y217Q	0.99	0.99
G097A/P129S/Y217Q	0.87	0.98
G097A/G128S/P129A/Y217Q	0.65	0.98
G097A/G128S/P129K/Y217Q	0.90	0.96
G097A/P129N/Y217Q	0.80	0.96
G097A/P129E/Y217Q	1.01	0.95
G097A/G128S/P129K/G131S/Y217Q	0.95	0.93
G097A/G128S/P129R/Y217Q	0.83	0.92
G097A/G128S/P129V/Y217Q	0.48	0.90
G097A/G128S/P129Y/Y217Q	0.41	0.89
G097A/G128A/P129R/Y217Q	0.69	0.88
G097A/G128H/P129S/Y217Q	0.59	0.85
G097A/G128N/P129S/Y217Q/V270A	0.51	0.84
G097A/G128S/P129L/Y217Q	0.45	0.81
G097A/P129R/Y217Q	0.70	0.80
G097A/P129W/Y217Q	0.51	0.79
G097A/P129Z/S130A/Y217Q	0.51	0.78
G097A/G128H/P129R/Y217Q	0.58	0.76
G097A/G128H/P129T/Y217Q	0.57	0.73
G097A/P129G/Y217Q	0.55	0.70
G097A/G128N/P129A/Y217Q	0.42	0.70
G097A/G128H/Y217Q	0.58	0.68

Table 7-1: Stain Removal Performance and Expression of BPN' 3 Variants at Positions 128,129, and 130

Variant	TCA	BMI
G097A/G128S/P129W/Y217Q	0.36	0.63
G097A/P129Z/Y217Q	0.47	0.60
G097A/G128N/Z128.01A/P129S/Y217Q	0.39	0.60
G097A/G128N/Y217Q	0.47	0.59
G097A/G128A/P129L/Y217Q	0.39	0.58
G097A/G128T/P129N/Y217Q	0.39	0.56
G097A/G128H/P129A/Y217Q	0.50	0.53
G097A/G128M/P129R/Y217Q	0.53	0.53
G097A/G128Q/P129K/Y217Q	0.49	0.53
G097A/G128T/P129Q/Y217Q	0.42	0.47
G097A/G128T/P129H/Y217Q	0.41	0.46
G097A/G128T/P129R/Y217Q	0.40	0.44
G097A/G128D/P129E/Y217Q	0.46	0.43
G097A/G128T/P129T/Y217Q	0.37	0.42
G097A/G128T/P129A/Y217Q	0.39	0.38
G097A/G128Z/P129Z/Y217Q	0.45	0.36
G097A/G128S/Z128.01A/P129S/Y217Q	0.42	0.31
G097A/G128N/P129M/Y217Q	0.42	0.26
G097A/G128N/P129V/Y217Q	0.46	0.26
G097A/G128E/P129K/Y217Q	0.52	0.25
G097A/G128N/Z128.01A/P129F/Y217Q	0.30	0.22
G097A/G128C/P129R/Y217Q	0.77	0.21
G097A/G128T/P129M/Y217Q	0.34	0.20
G097A/G128R/Z128.01A/P129S/Y217Q	0.41	0.20
G097A/G128S/Z128.01A/P129A/Y217Q	0.33	0.15
G097A/G128D/Y217Q	0.41	0.15
G097A/G128T/P129Y/Y217Q	0.38	0.14
G097A/G128R/P129S/Y217Q	0.47	0.13
G097A/G128R/Z128.01A/P129T/Y217Q	0.42	0.12
G097A/G128R/Z128.01A/P129A/Y217Q	0.38	0.11
G097A/G128T/P129W/Y217Q	0.37	0.11
G097A/G128Y/Z128.01A/P129R/Y217Q	0.41	0.10
G097A/G128C/P129D/Y217Q	0.84	0.10
V084L/G097A/P129Z/Y217Q	0.46	0.10
G097A/G128R/P129D/Y217Q	0.51	0.09
G097A/G128A/Z128.01A/P129H/E195A/Y217Q	0.35	0.09
G097A/G128S/Z128.01A/P129Y/Y217Q	0.40	0.09
G097A/G128N/P129L/Y217Q	0.51	0.08
G097A/G128C/P129H/Y217Q	0.71	0.07
G097A/G128T/Z128.01A/P129T/Y217Q	0.38	0.05
G097A/G128C/Z128.01A/P129A/Y217Q	0.39	0.05
G097A/G128S/Z128.01A/P129I/Y217Q	0.41	0.05

Table 7-1: Stain Removal Performance and Expression of BPN' 3 Variants at Positions 128,129, and 130

Variant	TCA	BMI
G097A/G128S/Z128.01A/P129E/Y217Q/Q271H	0.41	0.05
G097A/G128S/Z128.01A/P129L/Y217Q	0.42	0.05
G097A/G128Y/Z128.01A/P129S/Y217Q	0.43	0.05
G097A/G128S/Z128.01A/P129M/R186H/Y217Q	0.44	0.05
G097A/G128E/Z128.01A/P129D/Y217Q	0.44	0.05
G097A/G128T/P129L/Y217Q	0.46	0.05
G097A/G128F/P129D/Y217Q	0.47	0.05
G097A/G128V/Z128.01A/P129R/Y217Q	0.47	0.05
G097A/G128L/Z128.01A/P129S/Y217Q	0.49	0.05
G097A/G128Y/P129T/Y217Q	0.51	0.05
G097A/G128C/P129Y/Y217Q	0.53	0.05
G097A/G128C/P129T/Y217Q	0.59	0.05
Parent: G097A/G128A/Y217Q	1.00	1.00

EXAMPLE 8**Construction of Combinatorial Variants of BPN'**

[0160] Combinatorial variants of BPN' were created by extension (fusion) PCR using the following BPN' mutants as parent molecules-- BPN' G97A/G128A/Y217Q, BPN' M124V/L126A/Y217Q, BPN' G128A/Y217Q, and BPN' N123G/Y217Q to create insertions or substitutions at positions 96, 98, 129 and/or 222. For each combinatorial mutant created, the parent DNA molecule, mutations introduced and PCR primers used are listed in the Table 8-1. To create each mutant, two fragments 1 and 2 were generated by the PCR primers (Table 8-2) in the 5' and 3' regions of the gene using the respective parent molecule listed in Table 8-1. These fragments were mixed and amplified by the flanking primers P4973 and P4950 (Table 8-2) to restore the full-length gene.

[0161] Each amplification reaction contained 30pmol of each PCR primer and 100 ng of the template DNA. Amplifications were carried out using Vent DNA polymerase (New England Biolabs). The PCR mix (20 ul) was initially heated at 95°C for 2.5 min followed by 30 cycles of denaturation at 94°C for 15s, annealing at 55°C for 15s and extension at 72°C for 40s. Following amplification, the 5' and 3' fragments were gel-purified by the QIAGEN gel-band purification kit, mixed and amplified by the flanking primers P4973 and P4950. The full-length DNA fragment produced was gel-purified by the QIAGEN gel-band purification kit, digested by the *Bam*HI and *Hind*III restriction enzymes and ligated with the pHPLT-BPN' vector containing the respective parent molecules, which were also cut with the same restriction enzymes. Ligation mixtures were amplified using rolling circle amplification according to manufacturer's recommendation (Epicentre). One microliter of the ligation mixture was mixed with 5 ul of the sample buffer, heated to 95°C for 3 min and cooled on ice. Next, 5 ul of the reaction buffer and 0.2 ul of the enzyme was added to each tube, followed by incubation at 30°C for 10 hours. Products of the rolling circle amplification were diluted 100 times and used to transform *Bacillus subtilis* cells (genotype: $\Delta npr-E$, $\Delta nprE$, $\Delta spoII E$, *amyE::xylRPxylAcomK-phleo*) as described in Example 2. Transformations were plated on LB plates containing 1.6% skim milk and 10 ppm Neomycin and incubated overnight at 37°C. Only colonies with halos were picked and grown in microtiter plates for further analysis as described in Example 2. Protein concentration of culture supernatants was determined by TCA precipitation as described in Example 1.

Table 8-1: Parent DNA Molecule, Mutations Introduced and PCR Primers for the Generation of Combinatorial Variants in BPN'

Variant	Parent Molecule/ DNA Template	Mutations Introduced	Primers Used for PCR Amplification	
			Fragment 1	Fragment 2
1	G97A/G128A/Y217Q	Z098.001S	P4947, P4952	P4951 , P4948
2	G97A/G128A/Y217Q	Z096.001D/A098R	P4947, P4956	P4955 , P4948
3	G97A/G128A/Y217Q	P129V	P4947, P4960	P4959 , P4948
4	M124V/L126A/Y217Q	Z098.001S	P4947, P4954	P4953 , P4948
5	M124V/L126A/Y217Q	Z096.001D/A098R	P4947, P4958	P4957 , P4948
6	M124V/L126A/Y217Q	P129V	P4947, P4962	P4961 , P4948
7	G128A/Y217Q	Z098.001S	P4947, P4954	P4953 , P4948
8	G128A/Y217Q	Z096.001D/A098R	P4947, P4958	P4957 , P4948
9	G128A/Y217Q	P129V	P4947, P4966	P4965 , P4948
10	N123G/Y217Q	Z098.001S	P4947, P4954	P4953 , P4948
11	N123G/Y217Q	Z096.001D/A098R	P4947, P4958	P4957 , P4948
12	N123G/Y217Q	P129V	P4947, P4964	P4963 , P4948
13	G97A/G128A/Y217Q	M222Q	P4947, P4968	P4967 , P4948

Table 8-2: PCR primer Sequences Used for the Creation of Combinatorial Variants of BPN'.

SEQ ID NO:	Primer Name	Sequence
199	P4947	CTTAGTCTAGCAAAAAGGAGAG
200	P4948	GTGTTTTCTTGGAATTGTGCTG
201	P4949	CGACTCGAGCCATCCAGATC
202	P4950	CTTGCTCCAAGCTTAAATAAAAA
203	P4951	AAAGTTCTTGCAGCAAGTGACGGATCAGGCCAATACTCA
204	P4952	GCCTGATCCGTCACCTTGCTGCAAGAACTTTAACGGCGTAG
205	P4953	TAAAGTTCTTGGCGCAAGTGACGGATCAGGCCAATACTCA
206	P4954	CCTGATCCGTCACCTTGCGCCAAGAACTTTAACGGCGTA
207	P4955	GTTAAAGTTCTTGATGCACGTGACGGATCAGGCCAATACTCA
208	P4956	GATCCGTCACGTGCATCAAGAACTTTAACGGCGTAGAGCGA
209	P4957	GTTAAAGTTCTTGATGGTCGTGACGGATCAGGCCAATACTCA
210	P4958	GATCCGTCACGACCATCAAGAACTTTAACGGCGTAGAGCGAA
211	P4959	TGAGCCTGGGAGCAGTAAGCGGCAGTGCGGCACTTAAA
212	P4960	GCACTGCCGCTTACTGCTCCCAGGCTCATGTTGATTAC
213	P4961	TTAGCGCAGGAGGAGTAAGCGGCAGTGCGGCACTTAAA
214	P4962	GCACTGCCGCTTACTCCTCCTGCGCTAACGTTGATTA
215	P4963	GAGCCTGGGAGGAGTAAGCGGCAGTGCGGCACTTAAA
216	P4964	TGCCGCACTGCCGCTTACTCCTCCCAGGCTCATGCCGATT
217	P4965	GAGCCTGGGAGCAGTAAGCGGCAGTGCGGCACTTAAA
218	P4966	GCACTGCCGCTTACTGCTCCCAGGCTCATGTTGATT
219	P4967	AACGGGACTTCCCAAGCCTCGCCGCATGTAGCTG
220	P4968	ACATGCGGCGAGGCTTGGGAAGTCCCGTTTTGCGCAC
221	P4973	AAAGGATCCTAATCGGCGCTTTTC

EXAMPLE 9

Stain Removal Performance of Combinatorial Variants of BPN'

[0162] The stain removal activity of combinatorial variants of BPN' generated as described in Example 8 was tested as described in Example 3. As described throughout, functionality of BPN' variants was quantified as a performance index (P_i), which is the ratio of performance of a variant to BPN'3 (BPN' G97A-G128A-Y217Q). Variants showing a P_i value greater or equal than 0.5 for BMI stain removal performance and/or TCA precipitation showed improved cleaning benefits and/or expression. Performance indices less than or equal to 0.05 were fixed to 0.05 and indicated in bold italics in the table. Results are shown in Table 9-1.

Table 9-1: Stain Removal Performance and Expression of BPN' Variants		
Variants	TCA	BMI pH 8/16°C
G097A/G128A/Y217Q/M222Q	0.56	0.88
M124V/L126A/P129V/Y217Q	0.69	0.79
G128A/P129V/Y217Q	0.36	0.75
G097A/G128A/P129V/Y217Q	0.27	0.75
N123G/P129V/Y217Q	0.47	0.37
V30I/Z096.01D/G097A/A098R/G128A/Y217Q	0.54	0.19
Z096.01D/A098R/G128A/Y217Q	0.47	0.19
Z096.01D/A098R/M124V/L126A/Y217Q	0.49	0.17
G097A/Z098.01S/G128A/Y217Q	0.71	0.17
Z096.01D/A098R/N123G/Y217Q	0.54	0.16
Z096.01D/G097A/A098R/G128A/Y217Q/S236F	0.78	0.12
Z098.01S/M124V/L126A/Y217Q	0.45	0.11
Z098.01S/G128A/Y217Q	0.56	0.07
Z096.01D/G097A/A098R/G128A/Y217Q	0.55	0.07
V093I/Z098.01S/M124V/L126A/Y217Q	0.64	0.06
A073V/Z098.01S/N123G/Y217Q	0.52	0.05
G065D/Z098.01S/G128A/Y217Q	0.66	0.05
I011L/Z098.01S/G128A/Y217Q	0.53	0.05
L082F/Z096.01D/A098R/N123G/Y217Q	0.41	0.05
Z098.01S/N123G/A138V/Y217Q	0.44	0.05
Z098.01S/N123G/Y217Q	0.52	0.05

SEQUENCE LISTING

[0163]

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<130> GRF/FP6850440

<140> Divisional application of 09759304.0

<141> 2009-06-03

<150> EP 09759304.0

<151> 2009-06-03

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Ser Met Val Pro Ser Glu Thr Asn Pro Phe Gln Asp Asn Asn Ser His
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Gly Thr His Val Ala Gly Thr Val Ala Ala Leu Asn Asn Ser Ile Gly
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Val Leu Gly Val Ala Pro Ser Ala Ser Leu Tyr Ala Val Lys Val Leu
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Ser Gly Val Val Val Val Ala Ala Ala Gly Asn Glu Gly Thr Ser Gly
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Ser Ser Ser Thr Val Gly Tyr Pro Gly Lys Tyr Pro Ser Val Ile Ala
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Gly Pro Glu Leu Asp Val Met Ala Pro Gly Val Ser Ile Gln Ser Thr
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Pro His Val Ala Gly Ala Ala Ala Leu Ile Leu Ser Lys His Pro Asn
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 <400> 37
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 <400> 54
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 <400> 55
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 <400> 56
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 <400> 57
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 <400> 58
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 <400> 61
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 <400> 62
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 <400> 63
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 <400> 66
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 <400> 68
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 <210> 69
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<212> DNA

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<223> Synthetic primer: P4953

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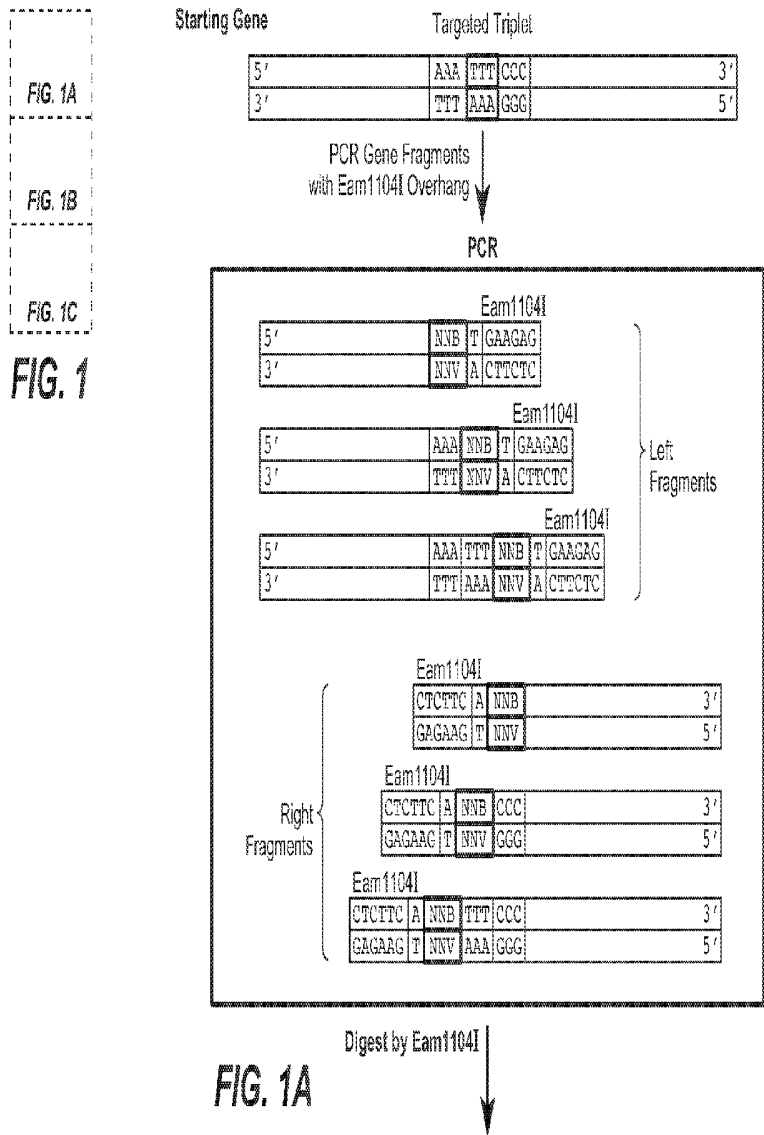
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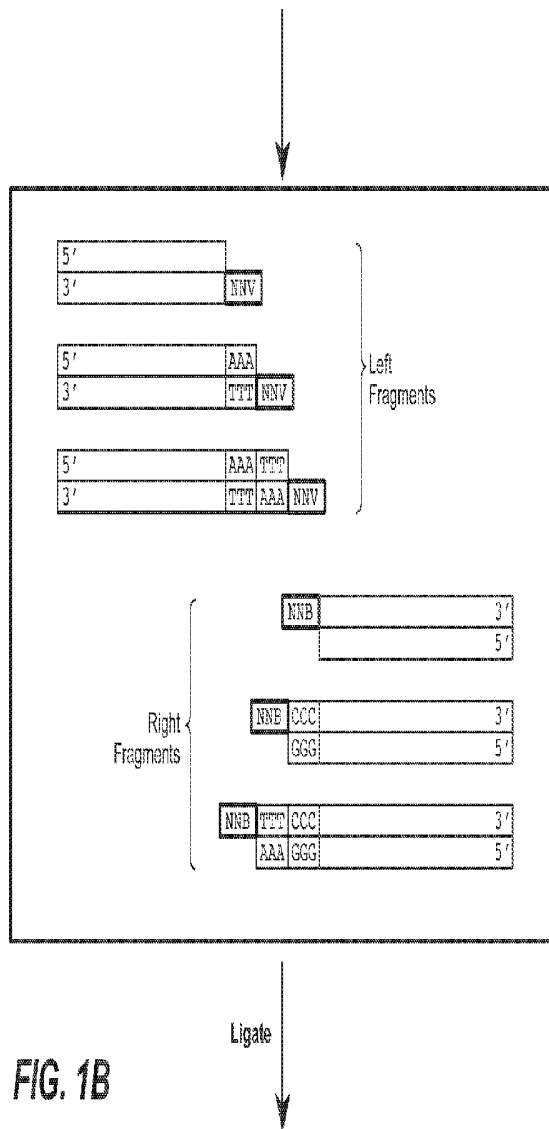
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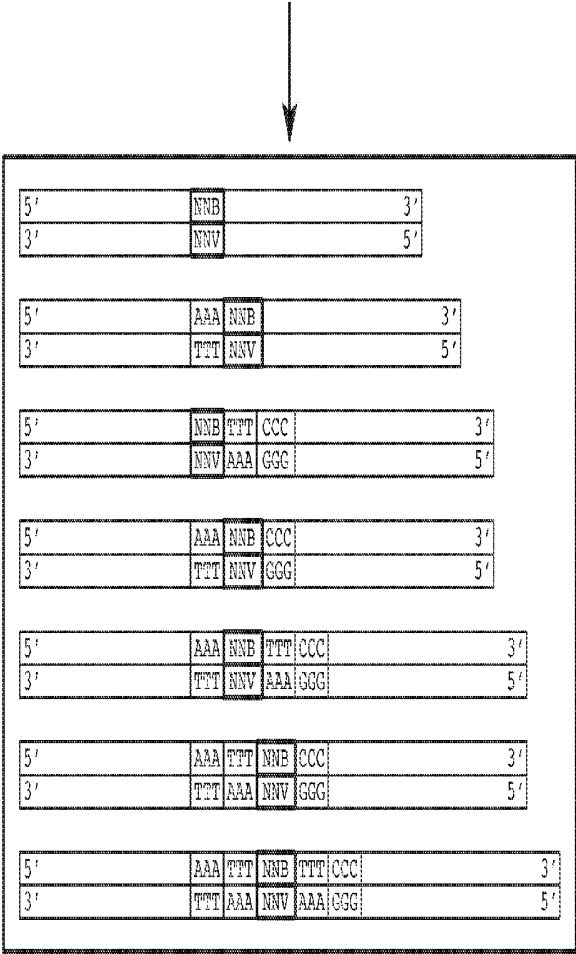
PATENTKRAV

1. Variant af subtilisin BPN', der kun adskiller sig fra sekvensen af BPN' subtilisin ifølge SEQ ID
- 5 NO: 2 ved ét af følgende ændringssæt:
 - G097A/G128S/Y217Q,
 - G097A/G128S/P129D/Y217Q,
 - G097A/G128S/P129V/A134T/Y217Q,
 - G097A/G128S/P129V/Y217Q,
 - 10 V068I/G097A/G128S/P129D/Y217Q,
 - G097A/S101D/G128S/S204F/Y217Q/S236F/T254P,
 - G097A/S101D/G128S/Y217Q,
 - G097A/S101D/G128S/P129D/Y217Q,
 - G097A/Z099.01S/A114S/G128S/Y217Q (hvor Z099.01S indikerer en indsætning af serin
 - 15 mellem positionerne 99 og 100),
 - G097A/G128S/P129T/Y217Q,
 - G097A/G128S/P129C/Y217Q,
 - G097A/G128S/P129Q/Y217Q,
 - G097A/G128S/P129E/Y217Q,
 - 20 G097A/G128S/P129A/Y217Q,
 - G097A/G128S/P129K/Y217Q,
 - G097A/G128S/P129K/G131S/Y217Q,
 - G097A/G128S/P129R/Y217Q,
 - G097A/G128S/P129V/Y217Q,
 - 25 G097A/G128S/P129Y/Y217Q,
 - G097A/G128S/P129L/Y217Q eller
 - G097A/G128S/P129W/Y217Q.
2. Fremgangsmåde til oprensning, hvilken fremgangsmåde omfatter følgende trin:
 - a) etablering af kontakt mellem en overflade og/eller en genstand omfattende et stof med
 - 30 mindst én variant af subtilisin BPN' ifølge krav 1; og
 - b) eventuelt vask og/eller skylning af overfladen eller genstanden.
3. Dyrefoder omfattende mindst én variant af subtilisin BPN' ifølge krav 1.
4. Foderbearbejdningssammensætning omfattende mindst én variant af subtilisin BPN' ifølge krav 1.

DRAWINGS

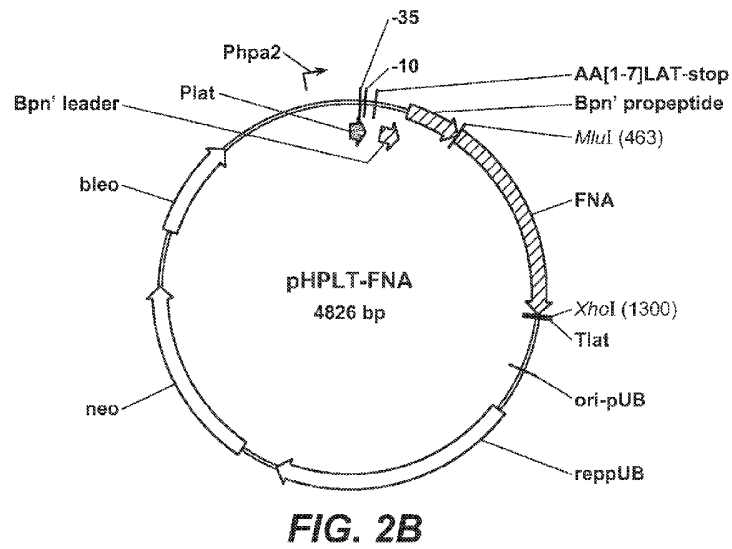
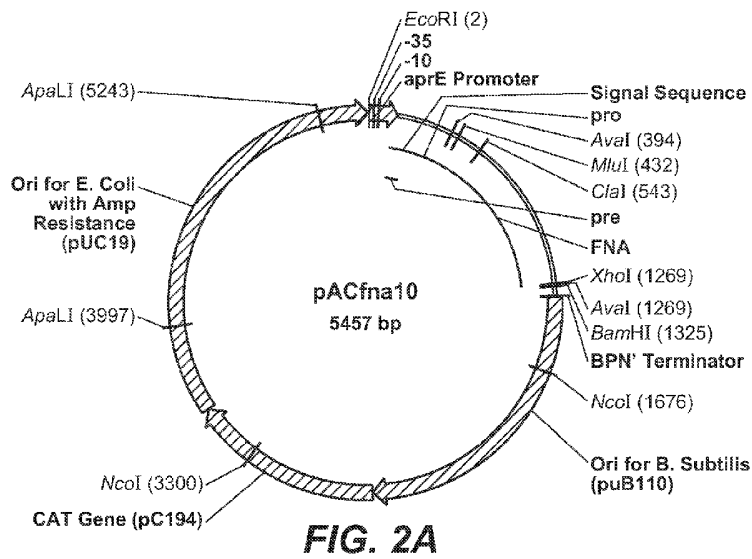


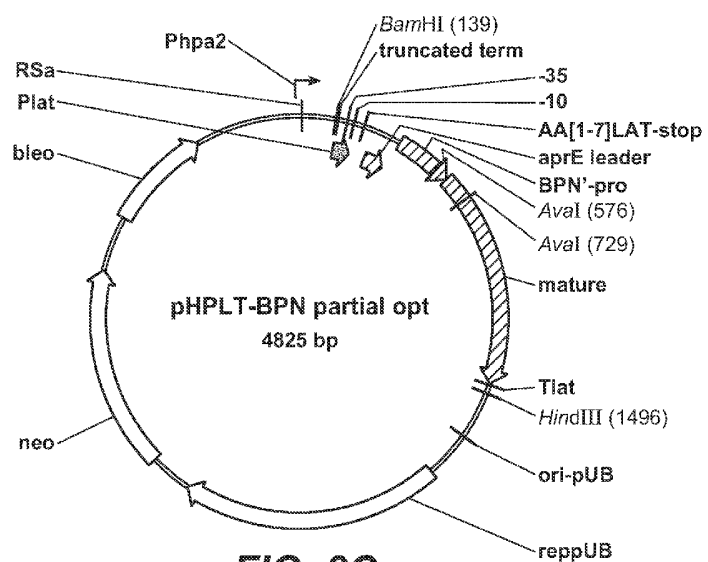




Examples of the Resulting Mutants

FIG. 1C



**FIG. 2C**