NON-NATURAL AMINO ACIDS AND NEUROTENSIN ANALOGUES

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ABSTRACT

This invention relates to methods of synthesis of amino acid analogs contained within peptide sequences that is adapted for preparation of peptides containing analogs of basic amino acids lysine and arginine.
Figure 1
Figure 3A

1. Et₃N, t-BuCOCl
2. n-BuLi, Xps

Br(CH₂)ₙCO₂H → Br(CH₂)ₙNCOO

H₂O₂, LiOH
(when R=H)

Br(CH₂)ₙCO₂H

1. KHMDS
2. RI

Br(CH₂)ₙNCOO

1. Requisite amine nucleophile

Z'(CH₂)ₙCO₂H

Z = NR₁R₂
R=H or an organic substituent
Figure 3B

R=H or an organic substituent
Figure 4

1. Et₃N, t-BuCOCl
2. n-BuLi, XPS

H₂O₂, LiOH

R=H or an organic substituent
Figure 7

1. PyBOP, HOBT, DIPEA
2. Wang-resin bound peptide leader sequence
3. TFA, Scavengers
4. RP-HPLC

KH28: n=5, R=H; R^4=NH_2
KH29: n=3, R=H; R^4=NH(CH_3)
KH30: n=3, R=H; R^4=N(CH_3)_3^+ 

n=5, R=H organic substituent; R^4=NH_2 
n=3, R=H organic substituent; R^4=NH(CH_3) 
n=3, R=H organic substituent; R^4=N(CH_3)_3^+
FIG. 8A

FIG. 8B
FIG. 10A

FIG. 10B
**FIG. 11A**

![Graph showing change in body temperature (°C) over time (min) for different doses of substance.](image)

**FIG. 11B**

![Graph showing change in body temperature (°C) over log dose (mg/kg) with ED50 = 0.943 mg/kg.](image)
FIG. 12
**Figure 13**

![Graph showing total activity over time with different doses of drugs](image)

- 0.5 mg/Kg
- 1 mg/Kg
- 3 mg/Kg
- 5 mg/Kg
- 10 mg/Kg
- Saline Control

**Time (min)**

0 60 120 180 240

**Figure 14**

![Graph showing total activity over time with different doses of drugs](image)

- 10 mg/Kg
- 20 mg/Kg
- 30 mg/Kg
- Saline Control

**Time (min)**

0 60 120 180 240
Figure 15

![Graph showing % PPI for different doses of ABS201.]

ABS201 Dose (Mg/Kg P.O.)

Figure 16

![Graph showing time (sec) for different treatments: saline, KH28, haloperidol.]

Treatment
Figure 17

![Graph showing change in body temperature over time for different conditions.]

- Saline
- Day One
- Day Five

Time (min)

Figure 18

![Graph showing total activity over time for different conditions.]

- Control
- Day One
- Day Five

ABS48 or saline

δ-AMPHETAMINE (1 mg/kg)

Time (min)
Figure 19

\[
\text{L=[U}^{14}\text{C]proline} \xrightarrow{1. \text{Na}_2\text{CO}_3, 2. \text{FMOC-OSu}} \text{Fmoc-Proline-OH}
\]
Figure 26

\[ R = \text{Arg-Pro-Tyr-isoleu-Leu-COOH} \]
\[ R' = \text{Arg-Pro-Tyr-tertleu-Leu-COOH} \]
Figure 27

ABS801 \( R = \text{Me} \)
ABS802 \( R = \text{Et} \)
ABS803 \( R_2 = \text{Me}, R_3 = \text{H} \)
ABS804 \( R_2 = \text{Me}, R_3 = \text{Me} \)
ABS805 \( R_2 = \text{Et}, R_3 = \text{H} \)
ABS806 \( R_2 = \text{Et}, R_3 = \text{Me} \)

\[ R = \text{Arg-Pro-Tyr-tertleu-Leu-COOH} \]
Figure 28

A.

![Graph A](image)

B.

![Graph B](image)

C.

![Graph C](image)

D.

![Graph D](image)

E.

![Graph E](image)
Figure 29

A.

B.
As 120'15010 22 Time(min)

A. 

Maximal Positive Effect

ABS 212 (10 mg/kg)
Morphine (5 mg/kg)
Saline

Time (min)

B. 

ABS 212 0.625 mg/kg
ABS 212 1:25 mg/kg
ABS 212 2.5 mg/kg
ABS 212 5 mg/kg
ABS 212 10 mg/kg

MPE

Time (min)

C. 

ABS 212 80 mg/Kg
ABS 212 60 mg/Kg
ABS 212 40 mg/Kg
ABS 212 20 mg/Kg

MPE

Time (min)
Figure 31

A.

B.

C.
Figure 32

A.

![Graph A](image1)

B.

![Graph B](image2)
Figure 33

Argolyn Biosciences, Inc. TY-06-161
Chung Model
IP Injection

- ■ ABS-212.2 TFA 10mg/kg n=5
- □ Saline n=5

Threshold / Grams

Time / Min.
Figure 34

A.

B.
NON-NATURAL AMINO ACIDS AND NEUROTENSIN ANALOGUES


BACKGROUND OF THE INVENTION

[0002] The influence of some non-natural amino acids have on the structural and biological activity of peptides has been briefly studied. For example, Moore et al. (Can. J. Biochem. 1978, 56, 315) disclosed the effect of the basic amino acid side chain length and the penultimate residue on the hydrolysis of benzoylpeptides by carboxypeptidase B1 (CPB). Non-natural amino acids including homolysine and homoarginine were incorporated into small peptide chains, and the kinetic parameters were determined for the CPB catalyzed hydrolysis of the peptide. Also, Lindeberg et al. (Int. J. Peptide Protein Res. 1977, 10, 240) disclosed the synthesis of 1-deamino-4-L-valine-8-DL-homolysine-vasopressin and protected 1-deamino-4-L-valine-8-D-lysine-vasopressin in which non-natural amino acids were incorporated. The non-natural amino acids were formed by addition of a methylene group to lysine and arginine to generate the non-natural amino acids homolysine and homoarginine, respectively. The study revealed that peptides with homolysine and homoarginine reduced the antidiuretic activity of the peptides.

[0003] Naturally occurring endogenous peptides are ideal drug candidates lead by virtue of their myriad activities in promoting and regulating biological processes. Inherent in the chemistry and biology of peptides, however, are several factors that also make them poor drug candidates. Peptidases most often exert localized effects and are rapidly degraded within the body. In addition, most peptides are unable to cross biological membranes, including the small intestine and blood brain barrier (BBB). Finally, peptides often bind to more than one receptor or receptor subtype, thus rarely showing the selectivity required of a viable drug candidate. Therefore, for a peptide to become a viable drug candidate, improvements in blood stability, receptor selectivity, and barrier crossing should be made without eliminating inherent binding affinity.

[0004] Numerous strategies have been developed as methods for improving peptide stability, including N- and C-terminal modifications to prevent exopeptidase activity, amide backbone modifications, and the introduction of conformational constraints to disguise peptides from peptidase degradation. Other therapeutic compounds employ a prodrug moiety intended to modify its overall hydrophobicity, which can result in the compound crossing biological membranes. In this case, the compound is cleaved into its active component by endogenous enzymes. While each of these strategies has been used to improve peptides as drug candidates, a universal solution for creating stable, receptor-selective peptides that cross biological barriers has not been discovered.

[0005] Consequently, there is a need in the art for non-natural amino acids and for peptides incorporating such acids to achieve superior effects, such as, for example, improved diagnostic or disease-fighting activity. Thus, the non-natural amino acid concept could be applied to development of new peptide pharmaceuticals. One example of such a development is the application to neuropeptides such as neurotensin. Neurotensin (NT) is a 13-amino acid residue peptide found primarily in the brain. It has multifunctional activities manifested through binding and activation of two neurotensin receptors (NTRs), NTR-1 and NTR-2 (see Carraway & Leeman, J. of Biol. Chem. 248: 6854 (1973)). Although full activity of NT resides in its C-terminal six amino acid sequence (NH2-Arg(6)-Arg(9)-Pro(10)-Tyr(11)-Ile(12)-Leu(13)), COOH, designated NT[8-13], see Carraway & Leeman, J. Biol. Chem. 250: 1907 (1975)), the C-terminal six amino acid sequence does not have activity when administered IP or orally due to its instability in blood and inability to cross the blood brain and/or gut barrier.

SUMMARY OF CERTAIN EMBODIMENTS OF THE INVENTION

[0006] The present invention concerns alpha-desamino amino acid compounds (desamino amino acid compounds) that are capable of carrying positively charged side chains, their synthesis, their application as substitutes for natural amino acid moieties of biologically active peptides and the resulting peptides as well. In particular, alpha-desamino arginine, lysine and ornithine as well as their substituted and derivatized side chain analogs constitute preferred embodiments of the invention. These desamino amino acid compounds can be substituted for arginine and/or lysine moieties in any known, biologically active peptide such that the substituted peptide will be truncated at the substitution position. Alternatively, these desamino amino acid compounds can be coupled to the amino group of the C-terminus of any known biologically active peptide to produce an extended peptide. The truncated and extended peptides have significant biological selectivity and biological half lives owing to their resistance toward amino peptidase degradation.

[0007] In a first aspect, the invention relates to a non-natural desamino amino acid compound having Formula I:

\[
\begin{align*}
\text{H} & \longrightarrow \text{\text{N}(R_{\text{H}})(R_{\text{I}})}_{m} \\
\text{HOOC} & \longrightarrow \text{R} \\
\text{H} &
\end{align*}
\]

[0008] wherein

[0009] n is an integer of from 0 to 5, preferably, 2 to 5;

[0010] m is zero or an integer of 1;

[0011] R is H or an organic substituent such as a straight or branched chain alkyl group of C1-C6, or an aromatic group of C6-C18 or a corresponding substituted aromatic group with one or two substituents selected from halogen, alkyl, alkoxy, carboxy, amido or alkyl in any combination, or a heteroaromatic group of C2-C18 and one or two heteroatoms selected from oxygen, sulfur and nitrogen in any combination or a corre-
sponding substituted heteraromatic group with one or two substituents selected from halogen, alkyloxy, carboxy, amide or alkyl in any combination;

[0012] R₁, R₂ and R₃ are, independently, hydrogen or branched or straight chain alkyl, alkenyl or alkynyl of C₁₋₅ or an aromatic group of C₅₋₁₈ or a corresponding substituted aromatic group with one or two substituents selected from halogen, alkyloxy, carboxy, amide or alkyl in any combination, or a heteroaromatic group of C₅₋₁₈ and one or two heteroatoms selected from oxygen, sulfur and nitrogen in any combination or a corresponding substituted heteraromatic group with one or two substituents selected from halogen, alkyloxy, carboxy, amide or alkyl in any combination and with the proviso that a maximum of two of R₁, R₂ and R₃ may be selected to be the aromatic, substituted aromatic, heteroaromatic or substituted heteroaromatic group, and provided that when m is 0 or 1 and n is 0 to 5, R₁, R₂ and R₃ are not all H;

[0013] Cα is a carbon atom having either R or S stereochemistry if the substituent at R is an organic substituent;

[0014] or an ester, amide, alkyl amide or metal cation or ammonium salt of the carboxylic acid group thereof, or an organic or inorganic acid salt of the amine group thereof, or any combination thereof.

[0015] In a second aspect, the invention relates to a non-natural desamino amino acid compound of the formula II:

(II)

wherein

[0016] n is an integer of from 0 to 6, preferably, 2 to 5;

[0017] when dashed line a is not present, X and Y are independently, hydrogen or lower branched or straight chain alkyl, alkenyl or alkynyl of C₁₋₅;

[0018] when dashed line a is present, X-Y is (CH₂)ₙ, wherein z is an integer of from 1-8, preferably, 2 to 4;

[0019] when dashed line a is present, X-Y is (CH₂)ₙ, wherein z is an integer of from 1-8, preferably, 2 to 4;

[0020] R is H or an organic substituent such as a straight or branched chain alkyl group of C₁₋₅, or an aromatic group of C₅₋₁₈ or a corresponding substituted aromatic group with one or two substituents selected from halogen, alkyloxy, carboxy, amide or alkyl in any combination, or a heteroaromatic group of C₅₋₁₈ and one or two heteroatoms selected from oxygen, sulfur and nitrogen in any combination or a corresponding substituted heteraromatic group with one or two substituents selected from halogen, alkyloxy, carboxy, amide or alkyl in any combination;

[0021] R is hydrogen or lower branched or straight chain alkyl, alkenyl or alkynyl of C₁₋₅ or an aromatic group of C₅₋₁₈ or a corresponding substituted aromatic group with one or two substituents selected from halogen, alkyloxy, carboxy, amide or alkyl in any combination, or a heteroaromatic group of C₅₋₁₈ and one or two heteroatoms selected from oxygen, sulfur and nitrogen in any combination or a corresponding substituted heteraromatic group with one or two substituents selected from halogen, alkyloxy, carboxy, amide or alkyl in any combination;

[0022] Cα is a carbon atom and the stereochemistry at Cα is either R or S if the substituent at R is an organic substituent;

[0023] or an ester, amide, alkyl amide or metal cation or ammonium salt of the carboxylic acid group thereof, or an organic or inorganic acid salt of the amine group thereof, or any combination thereof.

[0024] A third aspect of the present invention relates to a non-natural desamino amino acid compound of the formula III:

(III)

wherein

[0025] n is an integer of from 0 to 5, preferably, 2 to 5;

[0026] X-Y is (CH₂)ₙ, wherein z is an integer of from 0 to 6, preferably, 2 to 4;

[0027] X-Y is (CH₂)ₙ, wherein z is an integer of from 0 to 6, preferably, 2 to 4;

[0028] R is H or an organic substituent such as a straight or branched chain alkyl group of C₁₋₅, or an aromatic group of C₅₋₁₈ or a corresponding substituted aromatic group with one or two substituents selected from halogen, alkyloxy, carboxy, amide or alkyl in any combination, or a heteroaromatic group of C₅₋₁₈ and one or two heteroatoms selected from oxygen, sulfur and nitrogen in any combination or a corresponding substituted heteraromatic group with one or two substituents selected from halogen, alkyloxy, carboxy, amide or alkyl in any combination;

[0029] R₁₀ and R₁₁ are, independently, hydrogen or lower branched or straight chain alkyl, alkenyl or alkynyl of C₁₋₅, or an aromatic group of C₅₋₁₈ or a corresponding substituted aromatic group with one or two substituents selected from halogen, alkyloxy, carboxy, amide or alkyl in any combination and with the proviso that a maximum of two of R₁₀ and R₁₁ may be selected to be the aromatic, substituted aromatic, heteroaromatic or substituted heteroaromatic group, and provided that when m is 0 or 1 and n is 0 to 5, R₁₀ and R₁₁ are not all H;

[0030] Cα is a carbon atom and the stereochemistry at Cα is either R or S if the substituent at R is an organic substituent;

[0031] or an ester, amide, alkyl amide or metal cation or ammonium salt of the carboxylic acid group thereof, or an organic or inorganic acid salt of the amine group thereof, or any combination thereof.

[0032] A fourth aspect of the invention relates to a non-natural desamino amino acid compound of the formula IV:

(IV)
heterocyclic group with one or two substituents selected from halogen, alkoxy, carboxy, amide or alkyl in any combination and with the proviso that a maximum of two of \( R_{12}, R_{13}, \) and \( R_{14} \) may be selected to be the aromatic, substituted aromatic, heterocyclic or substituted heterocyclic group; and

\[ \text{wherein} \]
\[ n \text{ is an integer of from 0 to 5, preferably, 2 to 4;} \]
\[ \text{R is H or an organic substituent such as a straight or branched chain alkyl group of } C_1-C_9, \text{ or an aromatic group of } C_6-C_{18}, \text{ or a corresponding substituted aromatic group with one or two substituents selected from halogen, alkoxy, carboxy, amide or alkyl in any combination, or a heterocyclic group of } C_6-C_{18} \text{ and one or two heteroatoms selected from oxygen, sulfur and nitrogen in any combination or a corresponding substituted heterocyclic group with one or two substituents selected from halogen, alkoxy, carboxy, amide or alkyl in any combination;} \]
\[ \text{R}_9, \text{R}_{10} \text{ and } R_{11} \text{ are, independently, hydrogen or lower branched or straight chain alkyl, alkenyl or alkynyl of } C_1-C_9, \text{ or an aromatic group of } C_6-C_{18}, \text{ or a corresponding substituted aromatic group with one or two substituents selected from halogen, alkoxy, carboxy, amide or alkyl in any combination, or a heterocyclic group of } C_6-C_{18} \text{ and one or two heteroatoms selected from oxygen, sulfur and nitrogen in any combination or a corresponding substituted heterocyclic group with one or two substituents selected from halogen, alkoxy, carboxy, amide or alkyl in any combination and with the proviso that a maximum of two of } \text{R}_9, \text{R}_{10} \text{ and } R_{11} \text{ may be selected to be the aromatic, substituted aromatic, heterocyclic or substituted heterocyclic group; and} \]
\[ \text{C}_6 \text{ is a carbon atom and the stereochemistry at } \text{C}_6 \text{ is either R or S if the substituent at } R \text{ is an organic substituent;} \]
\[ \text{or an ester, amide, alkyl amide or metal cation or ammonium salt of the carboxylic acid group thereof, or an organic or inorganic acid salt of the amine group thereof, or any combination thereof.} \]

A fifth aspect of the invention relates to a non-natural desamino amino acid compound of the formula V:

\[ \text{wherein} \]
\[ n \text{ is an integer of from 0 to 5, preferably, 2 to 4;} \]
\[ \text{R is H or an organic substituent such as a straight or branched chain alkyl group of } C_1-C_9, \text{ or an aromatic group of } C_6-C_{18}, \text{ or a corresponding substituted aromatic group with one or two substituents selected from halogen, alkoxy, carboxy, amide or alkyl in any combination, or a heterocyclic group of } C_6-C_{18} \text{ and one or two heteroatoms selected from oxygen, sulfur and nitrogen in any combination or a corresponding substituted heterocyclic group with one or two substituents selected from halogen, alkoxy, carboxy, amide or alkyl in any combination;} \]

\[ \text{R}_{12}, R_{13}, \text{ and } R_{14} \text{ are, independently, hydrogen or lower branched or straight chain alkyl, alkenyl or alkynyl of } C_1-C_9, \text{ or an aromatic group of } C_6-C_{18}, \text{ or a corresponding substituted aromatic group with one or two substituents selected from halogen, alkoxy, carboxy, amide or alkyl in any combination, or a heterocyclic group of } C_6-C_{18} \text{ and one or two heteroatoms selected from oxygen, sulfur and nitrogen in any combination or a corresponding substituted heterocyclic group with one or two substituents selected from halogen, alkoxy, carboxy, amide or alkyl in any combination;} \]

\[ \text{C}_6 \text{ is a carbon atom and the stereochemistry at } \text{C}_6 \text{ is either R or S if the substituent at } R \text{ is an organic substituent;} \]

\[ \text{or an ester, amide, alkyl amide or metal cation or ammonium salt of the carboxylic acid group thereof, or an organic or inorganic acid salt of the amine group thereof, or any combination thereof.} \]
will be exchanged for a desamino amino acid moiety, thus providing the truncated peptide. Several known biologically active peptides are penultimately formed as pro-peptides with an arginine or lysine moiety at the pro-peptide or precursor cleavage position, or are formed as final peptides containing an arginine or lysine moiety at a position that can be cleaved to provide an active truncated peptide. Trypsin is an enzyme specific for such cleavage points. Examples include growth hormone-like peptide, neurotensin, proinsulin, and thrombin. The truncated versions of these examples with a desamino amino acid compound substituted for the arginine or lysine moiety provide selective, long-lasting biological activity.

A further aspect of the invention includes pharmaceutical and cosmetic compositions of the desamino amino acid compound, of the extended or truncated peptide, and combinations thereof. Unit dosage forms and biologically effective formulations of the pharmaceutical compositions are included. The cosmetic formulations include appropriate oil, creme, wax or aqueous base cosmetic carriers.

Yet another aspect of the invention includes methods of screening, diagnosis and treatment using the desamino amino acid compounds of the invention and/or the addition or truncation of peptides.

One embodiment of the invention is a truncated neurotensin peptide having a desamino amino acid as its N-terminus amino acid moiety.

An embodiment of the invention also is directed to processes and intermediates disclosed and claimed herein that are useful for preparing compounds of the inventions, such as compounds of Formula I, II, III, IV and/or V and peptides that contain such compounds. One class of such intermediates includes the N-protected or carboxyl protected or N- and carboxyl protected compounds of Formulas I, II, III, IV and V. These protected intermediates are described in detail in the following sections of the application. Another class of such intermediates includes the carboxylate salts of the compounds of Formulas I, II, III, IV and V, the organic or inorganic acid amine salts of those compounds and the double salts (carboxylate, amine salts).

An embodiment of the present invention concerns a method of preparing a compound of Formula (XI):

or a salt, hydrate, solvate, stereoisomer, or tautomer thereof,

wherein X is OH, O(C₆-C₁₀ alkyl), O(C₆-C₁₀ aryl), NR₂, an aminoacid residue or a peptide residue, or wherein X is O, NR, an aminoacid residue, or a peptide residue, respectively covalently bonded to a polymer;

Z is H, NR₂, alkyl, aryl, aralkyl, heterocyclyl, heteroaryl, heterocyclylalkyl, heteroaryalkyl, cycloalkyl, cycloalkylalkyl, an aminoacid residue, or a peptide residue; or is NR, an aminoacid residue, or a peptide residue, respectively covalently bonded to a polymer;

provided that X and Z are not both bonded to polymers; and provided that at least one of X or Z comprises an amino acid residue or a peptide residue, or an aminoacid residue or a peptide residue respectively covalently bonded to a polymer;

R is independently at each occurrence H, alkyl, aryl, heterocyclyl, heteroaryl, a nitrogen protecting group, or a nitrogen capping group, or two R groups together with the nitrogen atom to which they are bound form a 5-7 membered heterocyclic ring that can further comprise 1-3 additional heteroatoms N, O or S;

L is a saturated or unsaturated, linear, branched or cyclic hydrocarbyl or a combination thereof, comprising n carbon atoms, wherein any carbon atom can be substituted or unsubstituted, wherein 0-3 heteroatoms O, S, or NR can be substituted for a carbon atom of L;

n is 1 to about 10,

R¹, R² and R³ are independently hydrogen, alkyl, aryl, aralkyl, cycloalkyl, heterocyclyl, or heteroaryl, wherein one of R¹, R² or R³ may be absent, wherein any alkyl can contain 0-3 heteroatoms comprising N, O, or S, wherein any alkyl, aryl, cycloalkyl, heterocyclyl, or heteroaryl may be unsubstituted or substituted;

the method comprising contacting a compound of Formula (XII):

wherein Y is a leaving group;

with an amine of the formula R'R'RN, in a solvent for a time period and at a temperature sufficient for the formation of the compound of Formula (XI).

Another embodiment of the present invention concerns a method for the preparation of a compound of Formula (XIII):

or a salt, hydrate, solvate, stereoisomer, or tautomer thereof,

wherein X is OH, O(C₆-C₁₀ alkyl), O(C₆-C₁₀ aryl), NR₂, an aminoacid residue or a peptide residue, or wherein X is O, NR, an aminoacid residue, or a peptide residue, respectively covalently bonded to a polymer;

Z is H, NR₂, alkyl, aryl, aralkyl, heterocyclyl, heteroaryl, heterocyclylalkyl, heteroaryalkyl, cycloalkyl, cycloalkylalkyl, an aminoacid residue, or a peptide residue; or is NR, an aminoacid residue, or a peptide residue, respectively covalently bonded to a polymer;
[0070] provided that X and Z are not both bonded to polymers; and provided that at least one of X or Z comprises an amino acid residue or a peptide residue, or an amino acid residue or a peptide residue respectively covalently bonded to a polymer;

[0071] R is independently at each occurrence H, alkyl, aryl, heterocyclyl, heteroarylyl, a nitrogen protecting group, or a nitrogen capping group, or two R groups together with the nitrogen atom to which they are bound form a 5-7 membered heterocyclic ring that can further comprise 1-3 additional heteroatoms N, O, or S;

[0072] L is a saturated or unsaturated, linear, branched or cyclic hydrocarbyl or a combination thereof, comprising n carbon atoms, wherein any carbon atom can be substituted or unsubstituted, wherein 0-3 heteroatoms O, S, or N can be substituted for a carbon atom of L;

[0073] n is 1 to about 10;

[0074] R² and R⁵ are independently at each occurrence H, alkyl, cycloalkyl, heterocyclyl, aryl or heteroaryl, wherein any alkyl, cycloalkyl, aryl, heterocyclyl or heteroaryl residue may be unsubstituted or substituted; provided that R⁴ or R⁵ is not a Boc group;

[0075] the method comprising contacting a compound of Formula (XV):

\[
\text{NHR}^1 X Z \text{O}
\]

[0076] wherein R¹ is hydrogen, alkyl, aryl, aralkyl, cycloalkyl, heterocycles, or heteroaryl; wherein any alkyl can further contain 0-3 heteroatoms comprising N, O, or S, wherein any alkyl, aryl, cycloalkyl, heterocyclyl, or heteroaryl may be unsubstituted or substituted;

[0077] with a compound of Formula (XIV):

\[
\text{SR}^6 R^4 R^5 N R^1
\]

[0078] wherein R⁶ is substituted or unsubstituted alkyl or aryl, or a salt thereof;

[0079] in a solvent for a time period and at a temperature sufficient for the formation of the compound of Formula (XIII);

[0080] Yet another embodiment of the present invention concerns a method of preparation of a compound of formula (XIII):

\[
\text{R}^1 Y Z X \text{O}
\]
wherein \( Y \) is a leaving group;

the method comprising contacting the compound of formula (XII) with an amine of the formula \( R'\text{NH}_2 \), in a solvent for a time period and at a temperature sufficient for the formation of the compound of Formula (XV):

![Diagram](image)

then, contacting the compound of formula (XV) with a compound of Formula (XIV):

![Diagram](image)

wherein \( R^4 \) is substituted or unsubstituted alkyl or aryl, or a salt thereof;

in a solvent for a time period and at a temperature sufficient for the formation of the compound of Formula (XIII).

**BRIEF DESCRIPTION OF THE DRAWINGS**

[0096] FIG. 1. Structure comparisons of NT(8-13), an intermediate NT analogue, and ABS201.

[0097] FIG. 2. Representative Examples of Compounds of Formulas I-V.

[0098] FIG. 3A-3C. Scheme for Synthesis of Compounds of Formulas I-V.

[0099] FIG. 4. Asymmetric synthesis of \( \alpha \)-bromo acids.

[0100] FIG. 5. Synthesis of ethylene-bridged (N\(^\text{2} \) to N\(^\text{w} \)) arginine analogues.


[0102] FIG. 7. Peptide synthesis of representative peptides of the invention.

[0103] FIG. 8A-8C. Comparisons of induced hypothermia by \( \alpha \)-methyl NT(8-13) analogues.

[0104] FIG. 9. Hypothermic effects of ABS201 after IP (solid symbols) and oral administration (open symbols) at 2 mg/kg and 20 mg/kg, respectively.

[0105] FIG. 10A-10B. Comparison of hypothermic effects after IP and oral administration for KH29 (10A) and KH30 (10B).

[0106] FIG. 11A-11B. Dose-response curves for ABS201 after IP administration.

[0107] FIG. 12. Dose-dependent hypothermic response to ABS201 after oral administration.

[0108] FIG. 13. Attenuation of \( \text{d-amp} \) induced hyperactivity after IP administration of ABS201.


[0110] FIG. 15. Reversal of PPI induced in Brattleboro rats by ABS201 administered orally.


[0112] FIG. 17. Hypothermic effects of chronic administration of ABS201 after daily dose of 5 mg/kg ABS201.

[0113] FIG. 18. Effect of repeated daily administration of ABS201 on \( \text{d-amp} \) induced hyperlocomotion after daily doses of 5 mg/kg ABS201.


[0115] FIG. 20. Blood concentration-time curve in male SD rats following intravenous administration of ABS201. TFA at a dose of 5 mg/kg Free Base (n=3).

[0116] FIG. 21. Blood concentration-time curve in male SD rats following oral administration of ABS201.TFA at a dose of 50 mg/kg Free Base (n=3).

[0117] FIG. 22. Plasma and brain concentration-time curve in male SD rats following intravenous administration of ABS201.TFA at a dose of 100 mg/kg free base (n=3).

[0118] FIG. 23. Comparison of the sites of absorbion of ABS 201 in the GI tract.

[0119] FIG. 24. Antinociceptive Activity of ABS201 (5 mg/kg) and Morphine (5 mg/kg).

[0120] FIG. 25. Evaluation of ABS tolerance in rats dosed daily with 5 mg/kg ABS201 for 5 days as determined by MEPE.

[0121] FIG. 26. The structures of NT[8-13] and peptides ABS1, ABS13, ABS201, ABS15, ABS16, ABS 17 and ABS19, which exhibit enhanced NTR-2 to NTR-1 binding affinities.

[0122] FIG. 27. Structures of peptides designed to be better NTR-2 selective ligands.

[0123] FIG. 28. Analgesic effects of (A) ABS201, (B) ABS205, (C) ABS210, (D) ABS212, and (E) ABS205 administered by gavage at a dose of 20 mg/kg.

[0124] FIG. 29. Analgesic effect of (A) ABS232 and (B) ABS239 administered by gavage at a dose of 20 mg/kg.

[0125] FIG. 30. Analgesic effects of ABS212 determined using the hotplate assay. Comparison of the analgesic effects of ABS212 (10 mg/kg) and morphine (5 mg/kg) (A); dose response data for ABS212 administered by I.P. injection (B) or orally (C).

[0126] FIG. 31. Analgesic effects of ABS212 determined using the formalin assay. Comparison of the analgesic effects of ABS212 (10 mg/kg) and morphine (5 mg/kg) (A); dose response data for ABS212 administered by I.P. injection (B) or orally (C).

[0127] FIG. 32. Dose response data for ABS212 administered I.P.(A) and orally (B) as determined in the tail flick assay.

[0128] FIG. 33. Effects of ABS212 administered by I.P. injection determined using the Chung model of neuropathic pain.

[0129] FIG. 34. Concentration-Time curve in male SD rats following intravenous administration of ABS212.2HCl at a dose of 5 mg/kg free base (A) and oral administration of ABS212.2HCl at a dose of 50 mg/kg free base (B), (n=3).

**DETAILED DESCRIPTION OF THE INVENTION**

**Definitions**

[0130] As used in the specification and the appended claims, the singular forms "a," "an" and "the" include plural referents unless the context clearly dictates otherwise.
Variables, such as $R_1$, $R_2$, $n$, $z$, $X$, $Y$, $C_n$, and $C_P$, throughout the application are the same variables as defined herein unless stated to the contrary.

The term “alkyl” as used herein refers to a branched or unbranched saturated hydrocarbon group of 1 to 24 carbon atoms, such as methyl, ethyl, n-propyl, isopropyl, n-butyl, isobutyl, t-butyl, pentyl, hexyl, heptyl, octyl, decyl, tetradecyl, hexadecyl, eicosyl, tetracontyl and the like. Preferred alkyl groups herein contain from 1 to 6 carbon atoms.

The term “alkenyl” as used herein refers to a hydrocarbon group of 2 to 24 carbon atoms, with preferred groups within this class containing 2 to 6 carbon atoms, and structural formula containing a carbon-carbon double bond.

The term “alkynyl” as used herein refers to a hydrocarbon group of 2 to 24 carbon atoms, with preferred groups within this class containing 2 to 6 carbon atoms, and a structural formula containing a carbon-carbon triple bond.

As used herein, especially in reference to alkyl, alkenyl and alkynyl, unless defined otherwise, the term “lower” refers to a moiety having from 1 to 6 carbon atoms, preferably 1 to 4 carbon atoms, more preferably 1 to 2 carbon atoms.

The term “alkylating agent” as provided herein is a compound with the structural formula $RX$, where $R$ is an alkyl, alkenyl or alkynyl group as previously described, and $X$, which is preferably a halide such as chloride, bromide or iodide.

As used herein, the term “non-natural amino acid” refers to an organic compound that is a congener of a natural amino acid in that it has a structure similar to a natural amino acid so that it mimics the structure and reactivity of a natural amino acid. The non-natural amino acid as defined herein generally increases or enhances the properties of a peptide (e.g., selectivity, stability) when the non-natural amino acid is either substituted for a natural amino acid unit of a peptide or otherwise incorporated into a peptide.

As used herein, the term “peptide” refers to a class of compounds composed of amino acids chemically bound together. In general, the amino acids are chemically bound together via amide linkages (—CONH—); however, the amino acids may be bound together by other chemical bonds known in the art. For example, the amino acids may be bound by amine linkages. Peptide as used herein includes oligomers of amino acids and small and large peptides, including polypeptides.

As used herein, the term “activity” refers to a biological activity.

As used herein, the term “pharmacological activity” refers to the inherent physical properties of a peptide or polypeptide. These properties include but are not limited to half-life, solubility, and stability and other pharmacokinetic properties.

The term “organic acid salt” as used herein relates to the salt form of an amino group with an alkyl or aryl $C_1$-$C_3$ carboxylic, sulfonic, or phosphoric acid.

The term “inorganic acid salt” as used herein refers to the salt form of an amino group with a mineral acid such as hydrochloric, sulfuric, sulfonic, phosphoric, nitric, nitrous, or hydrobromic acid.

The term “aromatic of $C_6$ to $C_{18}$” as used herein refers to an aromatic hydrocarbon such as phenyl, naphthyl, anthracenyl, or an arylalkyl hydrocarbon such as benzyl, phenethyl or naphthylmethyl.

The term “heteroaromatic of $C_4$ to $C_{18}$ and of one or two heteroatoms selected from oxygen, sulfur and nitrogen in any combination” as used herein refers to a heteroaromatic hydrocarbon containing one or two heteroatoms or an alkyl heteroaromatic hydrocarbon such as thiethyl, furyl, pyrrolyl, azathienyl, azafuryl, pyridinyl, thiopyridinyl, pyrazinyl, methylenpyridinyl, ethylenpyridinyl, methylenpyrrolyl and the like.


Detailed Description

The present invention is directed to certain desaminoo amino acid compounds, their incorporation as extenders or as congeners in known biologically active peptides, and the use of the compounds and peptides in medical diagnosis, treatment and screening. Several aspects of the invention concern the mimics of the desaminoo amino acid compounds for the natural amino acids arginine and/or lysine. By their use as congeners for these natural amino acid moieties in known biologically active peptides, truncated versions of the peptides can be prepared in which the biological activity is more selective and longer lasting than that of the known peptide. By their use as extenders, their position as the N-terminal moiety adduct of a known biologically active peptide will also provide longer lasting biological activity than that of the known peptide.

An example of the use of the desaminoo amino acid compounds of the invention in a truncated peptide is provided by neurotensin. Neurotensin (NT) is a 13 amino acid peptide having neurological properties. Its cleavage at AA7 to produce truncated neurotensin (8-13) provides a peptide having selective biological activity. According to the invention, conversion of the AA8 arginine to a desaminoo amino acid moiety results in a peptide also having significant and selective biological activity. The examples of NT and the converted versions are shown in FIG. 1.

The biologically active peptides of the present invention have the desaminoo amino acid moiety as their N-terminal moiety. These peptides have known amino acid sequences of biologically active amino acids wherein the desaminoo amino acid is either covalently linked through an amide bond with the N-terminal amine group of the known peptide (extended peptide) or is substituted for its corre-
sponding congener moiety (analogous natural amino acid moiety) within the peptide (truncated version). In another alternative, the peptide becomes truncated at the position of substitution so that the desamino amino acid moiety becomes the new N-terminus and the amino acid residues upstream of this position are no longer part of the sequence (truncated peptide). The extended and truncated peptides can have longer lifetimes in vivo and can have biological activities like those of the natural peptides except that the activities will be more selective.

One aspect of the desamino amino acid compounds of the invention is provided by Formula I given above. Preferred embodiments of Formula I include those wherein:

- **R₁, R₂, and R₃** are independently, hydrogen or lower branched or straight chain alky of C₁-C₄, more preferably hydrogen or methyl, provided that when m is 0 or 1 and n is 0 to 5, R₁, R₂, and R₃ are not all H. In another embodiment, n is 4. In yet another embodiment, R is an H, methyl, ethyl or propyl. Selected preferred embodiments include those wherein R is H, methyl, ethyl, propyl or butyl and:

- **(0151) a) n is 4, m is 0, R₁ is hydrogen, R₂ is methyl, the compound of formula I is an acid, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0152) b) n is 4, m is 1, R₁, and R₃ are methyl, R₂ is hydrogen or methyl, the compound of formula I is an acid, the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0153) c) n is 4, m is 1, R₁ is methyl, R₂, and R₃ are hydrogen, the compound of formula I is an acid, the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0154) d) n is 4, m is 1, R₂, and R₃ are hydrogen, the compound of formula I is an acid, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0155) e) n is 3, m is 0, R₂ and R₃ are methyl, the compound of formula I is an acid, the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0156) f) n is 3, m is 0, R₂ and R₃ are ethyl, the compound of formula I is an acid, the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0157) g) n is 3, m is 0, R₂ and R₃ are propyl, the compound of formula I is an acid, the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0158) h) n is 3, m is 0, R₂ and R₃ are butyl, the compound of formula I is an acid, the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0159) i) n is 2, m is 0, R₂ and R₃ are methyl, the compound of formula I is an acid, the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0160) j) n is 2, m is 0, R₂ and R₃ are ethyl, the compound of formula I is an acid, the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0161) k) n is 2, m is 0, R₂ and R₃ are propyl, the compound of formula I is an acid, the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0162) l) n is 2, m is 0, R₂ and R₃ are butyl, the compound of formula I is an acid, the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

Also preferred are the esters or salts of any of the foregoing preferred embodiments.

Another aspect of the desamino amino acid compounds of the invention is illustrated by Formula II given above. Preferred embodiments of Formula II include those wherein:

- **(0165) a) n is 3, dashed line a is not present, the compound of formula II is an acid, R₄ is hydrogen, X is hydrogen, Y is methyl, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0166) b) n is 3, dashed line a is not present, the compound of formula II is an acid, R₄ is methyl, X is hydrogen, Y is methyl, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0167) c) n is 3, dashed line a is present, the compound of formula II is an acid, z is 2, R₅ is hydrogen, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0168) d) n is 3, dashed line a is present, the compound of formula II is an acid, z is 2, R₅ is hydrogen, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0169) e) n is 3, dashed line a is not present, the compound of formula II is an acid, R₄ is hydrogen, X is methyl, Y is hydrogen, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0170) f) n is 3, dashed line a is not present, the compound of formula II is an acid, R₄ is hydrogen, X is ethyl, Y is hydrogen, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0171) g) n is 2, dashed line a is not present, the compound of formula II is an acid, R₄ is hydrogen, X is hydrogen, Y is methyl, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0172) h) n is 2, dashed line a is not present, the compound of formula II is an acid, R₄ is methyl, X is hydrogen, Y is propyl, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0173) i) n is 4, dashed line a is present, the compound of formula II is an acid, z is 2, R₅ is hydrogen, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0174) j) n is 3, dashed line a is present, the compound of formula II is an acid, z is 2, R₅ is methyl, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0175) k) n is 2, dashed line a is present, the compound of formula II is an acid, z is 3, R₅ is methyl, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0176) l) n is 3, dashed line a is not present, the compound of formula II is an acid, R₄ is methyl, X is hydrogen, Y is ethyl, and the stereochemistry at C₃₀ is R or S if the substituent at R is methyl, ethyl, propyl or butyl;**

- **(0177) Also preferred are the esters or salts of any of the foregoing preferred embodiments a-l.
A third aspect of the desamino amino acid compounds of the invention is illustrated by Formula III. Preferred embodiments of Formula III include those wherein R₉ and R₈ are independently, hydrogen or lower alkyl or straight chain alkyl of C₁-C₈, preferably hydrogen or methyl, even more preferably all are hydrogen. In another embodiment, z is 2 or 3, preferably 3. In a preferred embodiment, n is 3. Additional preferred embodiments include those wherein R is H, methyl, ethyl, propyl or butyl and:

- a) n is 3, z is 2, R₈ and R₉ are hydrogen, the compound of formula III is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- b) n is 3, z is 3, R and R₉ are hydrogen, the compound of formula III is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- c) n is 2, z is 2, R₈ and R₉ are hydrogen, the compound of formula III is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- d) n is 4, z is 2, R₈ and R₉ are hydrogen, the compound of formula III is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- e) n is 2, z is 3, R₈ and R₉ are hydrogen, the compound of formula III is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- f) n is 4, z is 3, R₈ and R₉ are hydrogen, the compound of formula III is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- g) n is 2, z is 2, R₈ and R₉ are methyl, the compound of formula III is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- h) n is 4, z is 2, R₈ and R₉ are methyl, the compound of formula III is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- i) n is 2, z is 3, R₈ and R₉ are methyl, the compound of formula III is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- j) n is 4, z is 3, R₈ and R₉ are methyl, the compound of formula III is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;

Also preferred are the esters or salts of the preferred foregoing embodiments a-j.

A fourth aspect of the invention is provided by the desamino amino acid compounds of Formula IV. Preferred embodiments of the compounds of Formula IV include those wherein R₉, R₁₀ and R₁₁ are, independently, hydrogen or lower alkyl or straight chain alkyl of C₁-C₈, preferably hydrogen, methyl or ethyl. In another embodiment, R₁₀ is methyl. In yet another preferred embodiment, R₈ is hydrogen, R₉ is methyl, R₁₁ is hydrogen, and n is 3. Additional preferred embodiments include those wherein R is H, methyl, ethyl, propyl or butyl and:

- a) n is 3, R₈ and R₁₁ are hydrogen, R₉ is methyl, the compound of formula IV is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- b) n is 3, R₈ is hydrogen, R₁₀ and R₁₁ are methyl, the compound of formula IV is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- c) n is 3, R₈ is hydrogen, R₁₀ is methyl, R₁₁ is ethyl, the compound of formula IV is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- d) n is 2, R₈ and R₁₁ are hydrogen, R₁₀ is methyl, the compound of formula IV is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- e) n is 2, R₈ is hydrogen, R₁₀ and R₁₁ are methyl, the compound of formula IV is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- f) n is 4, R₈ is hydrogen, R₁₀ is methyl, R₁₁ is ethyl, the compound of formula IV is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- g) n is 2, R₈ is hydrogen, R₁₀ and R₁₁ are methyl, the compound of formula IV is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- h) n is 4, R₈ is hydrogen, R₁₀ is methyl, R₁₁ is ethyl, the compound of formula IV is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- i) n is 2, R₈ is hydrogen, R₁₀ and R₁₁ are methyl, the compound of formula IV is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
- j) n is 4, R₈ is hydrogen, R₁₀ is methyl, R₁₁ is ethyl, the compound of formula IV is an acid, and the stereochemistry at Cα is R or S if the substituent at R is methyl, ethyl, propyl or butyl;
the invention. Certain embodiments provide protected intermediates and protected non-natural amino acids of the invention, wherein the side chain amino group is protected by a protecting group that prevents undesired reaction of the amino group and is removable by a chemical method that does not also cause amide group cleavage. Certain embodiments provide protected intermediates and protected non-natural amino acids of the invention, wherein the side chain carboxyl group is protected by a protecting group that prevents undesired reaction of the carboxyl group and is removable by a chemical method that does not also cause carboxyl group cleavage. In certain embodiments, the protecting group is t-butyloxycarbonyl (BOC) or fluorenlymethoxy carbonyl (FOMOC). In certain embodiments, the protecting group is BOC, FOMOC, Alloc (allyloxycarbonyl), CBZ (benzoxycarbonyl), PbI(2,2,4,6,7-pentamethyldihydrobenzofuran-5-sulfonyl), NO2 (nitro), Pmc (2,2,5,7,8-pentamethylchroman-6-sulfonyl), Mtr (4-methoxy-2,3,6-trimethylbenzenesulfonyl), or Tos (tosyl).

[0208] In one embodiment, the structures of the non-natural desaminio amino acids of formulas I-V are similar to those of the naturally occurring amino acids lysine, arginine as well as the naturally occurring glutamate biosynthesis intermediate, ornithine. In preferred embodiments, the compounds of the invention differ from the corresponding natural amino acids due, inter alia, a longer or shorter methylene bridge between the (i) carboxyl terminus, which forms the N-terminus bond with the adjacent amino acid unit in a peptide, (ii) the presence of a —H or an alkyl group in place of the alpha amino group, and (iii) the organo group substitution of the amine side chain. Preferably, the extended bridge of the invention compared to the natural amino acid bridge is one carbon length longer or shorter (i.e., the homo- or des-forms).

In other preferred embodiments, the compounds of the invention have, inter alia, longer, shorter, or equivalent methylene bridge lengths and have substitutions at various moieties, form different moieties, or link moieties to form ring structures, compared to the comparable natural amino acid.

[0209] Each of the compounds of the invention can be prepared as the acid, amide, salt or ester. In water, the non-natural amino acids of the present invention will be charged; however, in cell membranes and other non-polar regions of the cell, the non-natural amino acids may not be charged. In one embodiment, the ester group of the non-natural amino acids of the present invention is methyl, ethyl, t-butyl, benzyl or allyl. In another embodiment, the counter-ion for the salts of the non-natural amino acids is sodium, potassium, ammonium and tetra-alkyl ammonium.

[0210] Some embodiments of the invention provide semisynthetic peptides comprising a non-natural amino acid compound of the invention. In some embodiments, the semisynthetic peptide comprises a non-natural amino acid compound as its N-terminus moiety. In some embodiments, the semisynthetic peptide comprises a non-natural amino acid compound as the N-terminal moiety of a semisynthetic peptide of neurotensin (8-13). In one embodiment, the semisynthetic peptide is ABS201, ABS202, ABS205, ABS207, ABS208, ABS210, ABS211, ABS212, ABS220, ABS225, ABS226, ABS227, ABS228, ABS230, ABS232, ABS234 or ABS239. In some embodiments, the semisynthetic peptide has an extended half-life in vivo as compared to a peptide having the same sequence as the semisynthetic peptide that does not comprise the non-natural amino acid compound substituted as its N-terminus moiety. In some embodiments, the semisynthetic peptide has analgesic and/or antipsychotic activity. Examples of semisynthetic peptides having antipsychotic and/or analgesic activity are shown in the tables below.

**TABLE 1**

| Peptides | N-terminus | Sequence* | Structure*
<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>ABS201</td>
<td>CH3</td>
<td>L-Mys-Arg-Pro-Tyr-Tleu-Leu</td>
<td>FIG. 7</td>
</tr>
<tr>
<td>ABS202</td>
<td>CH3</td>
<td>7-Arg-Pro-Tyrl-Leu-Leu</td>
<td>FIG. 7</td>
</tr>
<tr>
<td>ABS203</td>
<td>CH3</td>
<td>9-Arg-Pro-Tyre-Leu-Leu</td>
<td>FIG. 7</td>
</tr>
<tr>
<td>ABS204</td>
<td>N3</td>
<td>1-L-Mys-Arg-Pro-Tyr-Tleu-Leu</td>
<td>Scheme 1</td>
</tr>
<tr>
<td>ABS205</td>
<td>N3</td>
<td>13-Arg-Pro-Tyr-Leu-Leu</td>
<td>Scheme 1</td>
</tr>
<tr>
<td>ABS206</td>
<td>N3</td>
<td>7-Arg-Pro-Tyr-Leu-Leu</td>
<td>Scheme 1</td>
</tr>
<tr>
<td>ABS207</td>
<td>N3</td>
<td>9-Arg-Pro-Tyr-Leu-Leu</td>
<td>Scheme 1</td>
</tr>
<tr>
<td>ABS208</td>
<td>CH3</td>
<td>L-Mys-Arg-Pro-Tyr-Tleu-Leu</td>
<td>Scheme 1</td>
</tr>
<tr>
<td>ABS209</td>
<td>CH3</td>
<td>28-Arg-Pro-Tyr-Leu-Leu</td>
<td>Scheme 2</td>
</tr>
<tr>
<td>ABS210</td>
<td>CH3</td>
<td>29-Arg-Pro-Tyr-Leu-Leu</td>
<td>Scheme 2</td>
</tr>
</tbody>
</table>

*The "L-Leu" or "T-Leu" in the sequence indicates a non-natural amino acid moiety, the structure of which is shown in FIG. 7 or structural Scheme 1 or 2.

**TABLE 2**

| Peptide | N-terminus | Sequence* | Structure*
<table>
<thead>
<tr>
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<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>ABS21</td>
<td>NH3</td>
<td>X-Arg-Pro-Tyr-Isoleu-Leu</td>
<td>FIG. 26</td>
</tr>
<tr>
<td>ABS23</td>
<td>NH3</td>
<td>X-Arg-Pro-Tyr-Isoleu-Leu</td>
<td>FIG. 26</td>
</tr>
<tr>
<td>ABS24</td>
<td>CH3</td>
<td>X-Arg-Pro-Tyr-Isoleu-Leu</td>
<td>FIG. 26</td>
</tr>
<tr>
<td>ABS25</td>
<td>CH3</td>
<td>X-Arg-Pro-Tyr-Isoleu-Leu</td>
<td>FIG. 26</td>
</tr>
<tr>
<td>ABS26</td>
<td>CH3</td>
<td>4-Arg-Pro-Tyr-Isoleu-Leu</td>
<td>FIG. 26</td>
</tr>
<tr>
<td>ABS27</td>
<td>CH3</td>
<td>5-Arg-Pro-Tyr-Isoleu-Leu</td>
<td>FIG. 26</td>
</tr>
<tr>
<td>ABS28</td>
<td>CH3</td>
<td>6-Arg-Pro-Tyr-Isoleu-Leu</td>
<td>FIG. 26</td>
</tr>
<tr>
<td>ABS31</td>
<td>CH3</td>
<td>Orn-Age-Pro-Tyr-Isoleu-Leu</td>
<td>Scheme 3</td>
</tr>
<tr>
<td>ABS32</td>
<td>CH3</td>
<td>7-Arg-Pro-Tyr-Isoleu-Leu</td>
<td>Scheme 3</td>
</tr>
<tr>
<td>ABS34</td>
<td>CH3</td>
<td>9-Arg-Pro-Tyr-Isoleu-Leu</td>
<td>Scheme 3</td>
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<tr>
<td>ABS35</td>
<td>CH3</td>
<td>10-Arg-Pro-Tyr-Isoleu-Leu</td>
<td>Scheme 3</td>
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<td>ABS36</td>
<td>CH3</td>
<td>11-Arg-Pro-Tyr-Isoleu-Leu</td>
<td>Scheme 3</td>
</tr>
<tr>
<td>ABS37</td>
<td>CH3</td>
<td>12-Arg-Pro-Tyr-Isoleu-Leu</td>
<td>Scheme 3</td>
</tr>
<tr>
<td>ABS38</td>
<td>CH3</td>
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*The letter "X" in the sequence indicates a non-natural amino acid moiety, the structure of which is shown in FIG. 26.

**TABLE 3**

| Peptides | N-terminus | Sequence* | Structure*
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<td>CH3</td>
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<td>ABS239</td>
<td>CH3</td>
<td>35-Arg-Pro-Tyr-Tleu-Leu</td>
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TABLE 3-continued

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<th>Peptides</th>
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<tbody>
<tr>
<td>ABS229</td>
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</tr>
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<td>Scheme 3</td>
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</table>

*The "L-Leu" or "N-Leu" means tert-Leu. The number or term "L-Lys" in the sequence indicates a non-natural amino acid moiety, the structure of which is shown in Scheme 3.

[0219] Certain embodiments of the present invention provide a method to lower the body temperature of a patient, comprising administering to the patient an effective amount of a composition of the invention so as to lower the body temperature of the patient.

[0220] Certain embodiments of the present invention provide a method to treat a patient with psychosis, comprising administering to the patient an effective amount of a peptide of the invention so as to treat the psychosis.

[0221] Certain embodiments of the present invention provide a method to treat a patient with psychosis, comprising administering to the patient an effective amount of a composition of the invention so as to treat the psychosis.

[0222] Certain embodiments of the present invention provide a method to treat cancer, comprising administering to a patient an effective amount of a peptide of any of the invention so as to treat the cancer.

[0223] Certain embodiments of the present invention provide a method to treat cancer, comprising administering to a patient an effective amount of a composition of the invention so as to treat the cancer.

[0224] Certain embodiments of the present invention provide a method to treat pain, comprising administering to a patient an effective amount of a peptide of the invention so as to treat the pain.

[0225] Certain embodiments of the present invention provide a method to treat pain, comprising administering to a patient an effective amount of a composition of the invention so as to treat the pain.

[0226] Certain embodiments of the present invention provide a method for screening a peptide containing a non-natural amino acid compound for an activity, comprising the steps of: a) measuring a biological activity of a first peptide having a known amino acid sequence; and b) measuring the same biological activity of a semisynthetic peptide of any of the invention wherein the semisynthetic peptide has the same sequence as the first peptide except for the non-natural amino acid compound, or is a truncated version of the first peptide except for the non-natural amino acid compound.

[0227] Certain embodiments of the present invention provide a method of treating a patient with a disease that is affected by administration to the patient of a known first peptide, comprising administering to the patient a semisynthetic peptide of the invention wherein the semisynthetic peptide has the same sequence as the first peptide except for the non-natural amino acid compound, or is a truncated version of the first peptide except for the non-natural amino acid compound.

[0228] Certain embodiments of the present invention provide a method of increasing the ability of a known first peptide to cross a biological barrier of a subject, comprising substituting a semisynthetic peptide of the invention wherein the semisynthetic peptide has the same sequence as the first peptide except for the non-natural amino acid compound, or is a truncated version of the first peptide except for the non-
natural amino acid compound. In certain embodiments, the barrier comprises the blood brain barrier, a cell membrane, intestinal epithelium, skin, or blood-ocular.

[0229] Certain embodiments of the present invention provide a method of increasing the selectivity of a known peptide, comprising substituting for the known peptide a semisynthetic peptide of the invention wherein the semisynthetic peptide has the same sequence as the first peptide except for the non-natural amino acid compound, or is a truncated version of the first peptide except for the non-natural amino acid compound.

[0230] Certain embodiments of the present invention provide a method of increasing the resistance of a known peptide to digestion by a peptidase, comprising substituting for the known peptide a semisynthetic peptide of the invention wherein the semisynthetic peptide has the same sequence as the first peptide except for the non-natural amino acid compound, or is a truncated version of the first peptide except for the non-natural amino acid compound.

[0231] Certain embodiments of the present invention provide a method of treating a patient with a disease that is affected by administration to the patient of a known first peptide that crosses a body barrier, comprising administering to the patient a semisynthetic peptide of the invention wherein the semisynthetic peptide has the same sequence as the first peptide except for the non-natural amino acid compound, or is a truncated version of the first peptide except for the non-natural amino acid compound.

[0232] Certain embodiments of the present invention provide a method of preparing a semisynthetic peptide with an extended half-life in vivo comprising substituting for a known peptide a semisynthetic peptide of the invention wherein the semisynthetic peptide has the same sequence as the first peptide except for the non-natural amino acid compound, or is a truncated version of the first peptide except for the non-natural amino acid compound.

[0234] The compound designators ABS201, ABS48, K1H48, and peptide 28, as used herein, unless otherwise indicated, represent the same compound.

Preparation of the Desamino Amino Acid Compounds

[0235] The preparation of the desamino amino acid compounds of the invention follows the overall synthetic scheme depicted in FIG. 3. The first step in this process is the production of omega halogen carboxylic acids having a methylene unit chain length corresponding to n of formulas I through V. In the following discussion, and in FIG. 3, this intermediate is designated as compound 27. Following the production of compound 27, its ω-halo group can be easily displaced with excess nucleophilic agent to produce the desamino amino acid compounds of Formulas I-V.

[0236] When R—H, the omega halogen compound can be used directly in succeeding steps. When R is an organic substituent, the R group is added through the following synthesis. When R is an organic substituent, the reaction conditions for production of compound 27 involve protection of the carboxyl group of an omega carboxylic acid by formation of an acyl oxazoline. The acyl oxazoline is converted to an enolate and the enolate is combined with an alkylating agent such as alkyl iodide or alkyl mesylate to form compound 27. Use of a large excess of the alkylating agent and long reaction times promote significant yields of compound 27.

[0237] As shown in the synthetic scheme of FIG. 3, the omega halo carboxylic acid compounds 25 can be converted to any of the side chain modifications by coupling the appropriate side chain moiety and the omega halo group of compound 25. Appropriate protection of the carboxyl group is also advantageously employed. The conditions for these reactions, and the appropriate alkylating and substituting agents follow the teaching set forth in "Advanced Organic Chemistry", 4th Edition, J. March, Wiley InterScience, New York, N.Y. 1992, the entire disclosure of which is incorporated herein by reference.

[0238] In particular, to prepare the compounds of Formula I, (see synthetic schemes of FIGS. 3 and 4), the omega halo carboxylic acid compound 27 can be combined with the appropriate amine nucleophile such as ammonia, a primary amine or a secondary amine. The formulas of the amine nucleophile correspond to the side chain moiety of Formula I. The reaction conditions will follow those appropriate for amine nucleophilic substitution as are disclosed in "Advanced Organic Chemistry" cited above and incorporated herein as if fully repeated. These compounds can be used directly in the following peptide synthesis provided that the side chain amine group is appropriately protected or otherwise inhibited from carboxyl condensation.

[0239] Similarly, to prepare the compounds of Formula II (see the synthetic schemes of FIGS. 3 and 5) the omega halo compound 27 may first be protected at the carboxyl position and then can be reacted sequentially with a diamine and cyanogen bromide. Deprotection and purification will afford the desamino amino acid compounds of Formula II. These compounds can be used directly in peptide synthesis with appropriate side chain protection.

[0240] The compounds of Formulas III and IV (see the synthetic schemes of FIGS. 3 and 6) can also be prepared by addition of the side chain moiety to the omega halo carboxylic acid compounds 27. In this instance, protection of the carboxyl group is unnecessary. Preparation of the appropriate thiourea compound can be accomplished by addition of an alkylating agent such as an alkyl, ankenyl or alkynyl halide to thiourea, N-substituted thiourea or N,N-disubstituted thiourea (commercially available). The nucleophilic substitution of the resulting appropriate thiourea compound at the omega halo position of compound 27 under basic conditions provides the desamino amino acid compounds of Formula IV. Similarly, addition of the appropriate cyclic thiourea compounds to the omega halo compound 27 under basic conditions provides the desamino amino acid compounds of Formula III. The appropriate cyclic thiourea compounds can be prepared by combining an alkylating agent such as an alkyl, ankenyl or alkynyl halide with the corresponding unsubstituted, N-substituted or N,N-disubstituted cyclic diazathione (commercially available). The crude reaction products can be purified by known methods such as ion-exchange chromatography to yield the desamino amino acid compounds of Formulas III and IV which can be used directly in peptide synthesis with appropriate side chain protection.
The compounds of formula V can be prepared by first protection of the carboxyl group as an acyl oxazolone. Subsequent to formation of the protected carboxylic acid, an organic substituent at the alpha carbon (R) can be added if desired following the procedure given above. In the next step, the omega halo group can be reacted with cyanide to form an omega cyano protected carboxylic acid. This intermediate can be reacted with an alkali metal organo amine (XNR, R, R, R) to form the amidine anion. The amidine anion may be quenched with acid, water or an alkyl halide to form either the unsubstituted imine moiety of the amidine or the alkyl substituted imine moiety of the amidine, respectively. This amidine protected carboxylic acid may then be employed in the synthetic scheme described above for production of the desired peptide.

Before their use in peptide synthesis, the side chains of the compounds of Formulas I-V may either be appropriately protected or determined to be sufficiently hindered that they will not enter into a peptide condensation reaction. For example, if the side chain of the compounds of Formula I is a primary amine group, it may be appropriately protected according to the teaching of the art associated with peptide synthesis. See, for example, the review of amine protecting groups provided in “Compendium of Organic Synthetic Methods,” &S Harrison, Wiley Interscience, New York, N.Y., 1971, the disclosure of which is incorporated herein by reference. In these instances, an appropriate protecting group can be t-butoxy carbonyl having the acronym BOC or fluorinylmethoxycarbonyl having the acronym FMOC. The BOC and FMOC protecting groups can be removed by mild treatment with acid, such as aqueous trifluoroacetic acid, and base, such as piperidine, respectively.

Alternatively, the omega halo carboxylic acid compound 27 can be coupled with the penultimate peptide to form an omega halo acyl moiety at the N-terminus of the penultimate peptide. Because the omega halo carboxylic acid compound 27 does not contain an amino moiety on its side chain, protection and spurious peptide formation are of less concern. In this alternative, the amino moiety can undergo nucleophilic reaction with the omega halo group of the acylated penultimate peptide as described above for formation of the compounds of Formulas I-V. The desired peptide having a residue of a compound of Formula I-V at its N-terminus is produced. In this alternative, appropriate protection of carboxyl and amino side chains and appropriate protection of the C-terminus may be employed to prevent undesirable reactions of these groups.

Peptide Synthesis and Purification

The invention includes the truncated and extended peptides which contain as their N-terminus moiety the residue of the compound of Formula I, II, III, IV or V. These peptides can be synthesized by the Merrifield solid phase method, which is an established method for preparing peptides to those skilled in the art. See R. B. Merrifield, Science, 232, 341-347 (1986), the disclosure of which is incorporated herein by reference for an explanation of, and conditions for, the Merrifield solid phase peptide synthesis. Alternatively, the peptide minus the N-terminal amino acid unit, or penultimate peptide, can be expressed recombinantly by known biological methods and the desamino amino acid compounds of Formula I-V can be added as the N-terminus by enzymatic condensation using an aminopeptidase. See “Enzyme Structure and Mechanism,” Alan Fersht, W. H. Freeman, New York, N.Y. (1985), the disclosure of which is incorporated herein by reference, for an explanation of, and conditions for, recombinant expression of peptides. The Formulas I-V compounds can be appropriately protected at the side chain amino group with standard protecting groups. In a preferred embodiment, the protecting groups are BOC and/or FMOC.

Briefly, for a solid phase synthesis, the penultimate peptide can be produced in bulk and then coupled to any of the Formula I-V compounds using the protection and coupling techniques of the Merrifield solid phase synthesis. Starting with an appropriate anchor resin designed for amino group exposure, the carboxy terminus amino acid unit of the peptide having an amino protecting group such as an FMOC group is anchored to the resin through a selectively cleavable carboxyl coupling link. The amino group of the anchored carboxy terminus unit is then deprotected and the additional amino protected amino acid units are then sequentially coupled in proper sequence. Each coupling step will involve deprotection of the protected amino group of the anchored peptide chain followed by peptide condensation between that unprotected amino group and the carboxyl group of the next amino acid unit. The condensation can be facilely obtained by carbodiimide coupling, by Schotten Bauman reaction or by activated acyl group condensation. These condensation reactions are described in “Advanced Organic Chemistry”, cited above. Protection of amine and carboxyl side chains using appropriate protecting groups that differ from the protecting groups of the alpha amino group entering into the peptide condensation will enable selective peptide condensation of the sequential amino acid units. Selection of appropriate protection groups and conditions for solid phase peptide synthesis are described in the Merrifield reference, cited above.

The penultimate peptides may also be produced by recombinant expression. This biological method involves re-engineering a microbe to express the penultimate peptide. A DNA segment encoding the penultimate peptide sequence can be inserted in proper reading from into a plasmid or other vector capable of causing microbial expression of the DNA. The vector will also contain appropriate control, promoter and selection DNA segments. Upon insertion into a microbe such as E. coli or B. subtilis, the microbe mixture can be selected for appropriate transfection by treatment with the corresponding selection agent. Typically the agent will be an antibiotic and the vector will contain a sequence encoding the corresponding detoxifying enzyme for the antibiotic. Chloranaphenicol and penicillin are two of such agents. Culturing the transfected microbe and harvesting the expressed peptide as either secreted material of the culture medium or by lysing the microbe cells will provide the crude penultimate peptide. The penultimate peptide may be purified by known techniques such as lyophilization, chromatography and the like. These recombinant techniques for peptide expression are fully set forth in “Cold Spring Harbor—Current Protocols in Molecular Biology,” Wiley Interscience, Cold Spring Harbor (2003), the disclosure of which is incorporated herein by reference.

An example of the solid phase peptide production is provided by the development of a set of neurotensin (8-13) compounds (NT peptide). These compounds incorporate the desamino amino acid compounds of Formulas I-V as their N-termini. They are a novel class of antipsychotic drugs, the biological study and background of which are described in the sections below.
NT Peptide Synthesis—General

[0248] The penultimate sequence of the peptide, NT(9-13), can be synthesized in bulk using p-alkoxybenzyl alcohol solid phase methodology (65) and stored in the fully protected form.

[0249] Starting Materials. P-alkoxybenzyl alcohol resin-bound Nα-Fmoc-Leucine, Nα-Fmoc-isoleucine, Nα-Fmoc-tert-Leucine, Nα-Fmoc-(But)-tyrosine, Nα-Fmoc-(Boc)-tryptophan, Nα-Fmoc-proline, and Nα-Fmoc-(Pbf)-arginine were purchased from Advanced Chemtech (Louisville, Ky.). PyBOP® was purchased from Novabiochem (San Diego, Calif.). N-hydroxybenzozoricole (HOBr), anhydrous N,N-dimethylformamide (DMF), N,N-dimethylpropylethylamine (DIPBA, trisopropylsilane (TIS), and trifluoroacetic acid (TFA) were purchased from Aldrich (Milwaukee, Wis.). Non-natural amino acid analogues were as prepared. Abbreviations: Fmoc, fluorenlymethoxycarbonyl; NH₂, ammonia; NH₂CH₂Cl, methylvamine; NH(CH₂)₂Cl, dimethylamine; N(CH₃)₃, trimethylamine; EtOH, ethanol.

[0250] Briefly, resin-bound Nα-Fmoc-leucine can be swollen in DMF prior to Fmoc cleavage with piperidine (20% in DMF). The piperidine solution can be removed with vacuum filtration and the resin-bound amino acid washed with DMF and CH₂Cl₂. Amino acids (4 eq) can be activated in DMF with HOBr (4 eq), PyBOP (4 eq), and DIPBA (10 eq) and added directly to the peptide reaction vessel. The amino acid couplings can be conducted for approximately 6 hr, the resin washed with DMF and CH₂Cl₂, and monitored with the Kaiser test (66) for the presence of free amines. Residues can be recoupled when necessary.

[0251] This procedure was repeated with subsequent amino acids to give the penultimate peptide sequence. Aliquots of the resin-bound pentamer can then be coupled as described above with the Formula I-V compounds to give the desired peptides. Acid-catalyzed deprotection can be performed with a TFA solution containing appropriate scavengers and crude peptides can be precipitated in ice-cold ether.

Peptide Purification—General

[0252] Reverse phase high pressure liquid chromatography can be used to purify the foregoing crude peptides. For example, a Waters dual pump system in combination with a Waters C18 radial compression column can be used for this purpose. Effluent can be monitored by UV absorbance at 280 nm.

Screening of Non-Natural Amino Acid-containing Peptides

[0253] The invention provides a method for screening a peptide for an activity or pharmacological activity. The method includes the steps of a) measuring an activity or pharmacological activity of a peptide having a selected natural amino acid sequence, and b) measuring the same activity or pharmacological activity of an extended or truncated peptide based upon the same amino acid sequence as the foregoing peptide wherein the N-terminus is a non-natural amino acid having the formula I-V, described above; and c) comparing the measured activity or pharmacological activity of the peptides from steps a) and b) to determine whether the peptide of step b) has the desired activity or pharmacological activity.

[0254] The activities for which the present invention screens can include any activity associated with a biologically active peptide or peptidomimetic. The following is a partial list of the many activities that can be determined in the present screening method:


2. Enzyme inhibition: A compendium of examples of specific screens for measuring these activities can be found in: H. Zoller “Handbook of Enzyme Inhibitors”, 2nd Ed. VCH Weinheim, FRG, 1989, and references therein.


7. Pharmacokinetic properties: The pharmacological activities assayed in the screening method include half-life, solubility, or stability, among others. For example, methods of analysis and measurement of pharmacokinetic properties can be found in: J.-P. Labaume “Handbook of Pharmacokinetics: Toxicity Assessment of Chemicals”, Ellis Horwood Ltd., Chichester, 1989, and references therein.

In the screening method, the peptide of step a) can consist of natural amino acids. Alternatively, the peptide of step a) can contain mostly natural amino acids, but also contain one or a small number of non-natural amino acids. Such a peptide is considered to consist essentially of natural amino acids.

In the screening method, the peptide of step b) will be the truncated or extended peptide of the invention as described above. In one embodiment, the structures of the non-natural amino acids of formulas I-V will be similar to those of the naturally occurring amino acids, lysine and arginine.

Thus, in the screening method contemplated herein, any extended or truncated peptide can be compared to any peptide having the same downstream sequence and having a
known activity or pharmacological activity to determine whether or not the extended or truncated peptide has the same or similar activity or pharmacological activity at the same or different level. Depending on the specifics of how the measuring step is carried out, the present screening method can also be used to detect an activity or pharmacological activity exhibited by the extended or truncated peptide. Also, the screening method can be used to detect and measure qualitative and quantitative differences in the same or similar activity or pharmacological activity.

Thus, the methods of the present invention provide evaluation of the alteration of activity of the extended or truncated peptide. Typically, the hydrophobicity of the peptide is increased, which can result indirectly in increased binding activity when the desamino amino acid moiety is involved in binding (e.g., receptor-ligand binding, enzyme- cofactor binding, enzyme-substrate binding) and since binding strength is correlated with activity, a peptide with a higher potency (higher measured activity level) can result.

Furthermore, the desamino amino acids of the present invention also can enhance or increase the pharmacological activity of a peptide. For example, because the desamino amino acids are more hydrophobic (i.e., more lipophilic), a peptide containing a non-natural amino acid is more able to pass a body barrier (e.g., blood brain, blood osular, skin, intestinal epithelium). Additionally, because the desamino amino acids impart increased selectivity and stability to a peptide, the pharmacological activity can also be screened when compared to other peptides.

Treatment

The invention further relates to a method of treating or preventing in a subject a male condition comprising administering to the subject an extended or truncated peptide having as its N-terminus an amino acid having the formula I-V. The basis peptide from which the extended or truncated peptide is formed will have or will be believed to have a biochemical, physiological, pharmacological or biological relationship with the malecondition to be treated or prevented. The male condition may be a disease, biological or organic dysfunction or an undesirable biological condition that is not ordinarily regarded as a disease or dysfunction, such as not limited to cosmetic malconditions such as skin blotches, acne, and the like. The subject may be a medical or veterinary patient including mammals such as humans, and non-humans mammals such as dogs, cats, cows, sheep, pigs as well as avian.

In the method of the present invention, the conditions that can be treated or prevented and the peptides that can be used are numerous. For example, a neurotransin peptide analogue that incorporates a non-natural amino acid of the invention can be synthesized. Such a synthetic NT peptide of the invention can be used to treat obesity, diabetes, sexual dysfunction, Parkinson’s disease, atherosclerosis, insulin resistance, impaired glucose tolerance, hypercholesterolemia, or hypertriglycerideremia. A synthetic NT peptide of the invention can also be used to induce vasodilation, muscle relaxation, or to decrease appetite.

In addition, a synthetic NT peptide of the invention can be used to treat hyperthermia; hypothermia; gastrointestinal ulcers; substance abuse; depression; Alzheimer’s disease; tardive dyskinesia; panic attack; gastrointestinal reflux disorder; irritable bowel syndrome; diarrhea; chonic; dyspepsia; pancreatitis; esophagitis; gastroparesis; neurological diseases such as schizophrenia, psychoses, anxiety, manic depression, delirium dementia, severe mental retardation, and dyskinesias such as Huntington’s disease and Tourette’s syndrome; fungal and viral infections, including HIV-1 and HIV-2 infections; pain (i.e., an analgesic); cancer (including gastrointestinal tumors); anorexia; bulimia; asthma; Parkinson’s disease; acute heart failure; hypotension; hypertension; urinary retention; osteoporosis; angina pectoris; myocardial infarction; allergies; inflammation; or benign prostatic hypertrophy.

The methods of treatment of the present invention can also include combination therapy where other pharmacologically active compounds useful for the treatment of obesity or other diseases are used in combination with a synthetic NT peptide of the invention.

It is known that obese patients have higher incidences of certain diseases such as atherosclerosis, hypercholesterolemia, hypertriglycerideremia, hypertension, sexual dysfunction (including erectile dysfunction), insulin resistance, impaired glucose tolerance, diabetes, [particularly non-insulin dependent diabetes mellitus (NIDDM or type 2 diabetes)] and the diseases associated with diabetes such as nephropathy, neuropathy, retinopathy, cardiomypathy, cata reacts, and polycystic ovary syndrome. These diseases can be treated indirectly by treating obesity using a synthetic NT peptide of the invention or directly by treating the specific disease itself using a synthetic NT peptide of the invention. These diseases can be treated in the absence of obesity using a synthetic NT peptide of the invention.

In one embodiment of the invention, an obese patient or a patient at risk of becoming obese can be administered a combination of: 1) a synthetic NT peptide of the invention; and 2) an additional compound useful to treat obesity, diabetes [including (NIDDM) and the conditions and/or diseases associated with diabetes, such as nephropathy, neuropathy, retinopathy, cardiomypathy, cata reacts, and polycystic ovary syndrome], atherosclerosis, hypercholesterolemia, hypertriglycerideremia, sexual dysfunction (including erectile dysfunction), insulin resistance, or impaired glucose tolerance, or combinations of compounds useful to treat these diseases.

Examples of classes of compounds that can be used to treat obesity include the active compound(s) in appetite suppressants such as Adipex®, Bontril®, Desoxyn Grunmet®, Fastin®, Ionamin®, and Meridia®, and lipase inhibitors such as Xenical®. Additional anti-obesity agents that can be used in combination with a synthetic NT peptide of the invention include a β3-adrenergetic receptor agonist, a cholecystokinin-A agonist, a monoamine reuptake inhibitor, a sympathomimetic agent, a serotoninergic agent, a dopamine agonist, a melanocyte-stimulating hormone receptor agonist or mimetic, a melanocyte-stimulating hormone receptor analog, a cannabinoid receptor antagonist, a melanin concentrating hormone antagonist, leptin, a leptin analog, a leptin receptor agonist, a galanin antagonist, a bombesin agonist, a neuropeptide-Y antagonist (including NPY-1 and NPY-5), a thyromimetic agent, dehydroepiandrosterone or an analog thereof, a glucocorticoid receptor agonist or antagonist, an orexin receptor antagonist, a uroctin binding protein antagonist, a glucagon-like peptide-1 receptor agonist, and a ciliary neurotrophic factor.

In another aspect of the invention, a synthetic NT peptide of the invention can be administered in combination with a compound that is known to treat hypertension. Examples of classes of compounds that can be used to treat
hypertension include calcium blockers, ACE inhibitors, diuretics, angiotensin II receptor blockers, beta-blockers, and alpha-adrenergic blockers. In addition, combinations of compounds in the above-referenced classes have been used to treat hypertension. Some examples of specific compounds that can be used in combination with a synthetic NT peptide of the invention include quinapril; amiodipine, including the besylate salt; nifedipine; doxazosin, including the mesylate salt; and prazosin, including the hydrochloride salt.

[0275] In another aspect, a synthetic NT peptide of the invention can be used in combination with compounds useful for the treatment of diabetes, including impaired glucose tolerance, insulin resistance, insulin-dependent diabetes mellitus (Type 1) and non-insulin dependent diabetes mellitus (NIDDM or Type 2). Also intended to be encompassed in the treatment of diabetes are the diabetic complications, such as neuropathy, nephropathy, cardiomyopathy or cataracts.

[0276] A synthetic NT peptide of the invention can also be used in combination with an aldose reductase inhibitor. Aldose reductase inhibitors constitute a class of compounds that have become widely known for their utility in treating conditions arising from complications of diabetes, such as diabetic neuropathy and nephropathy. Such compounds are well known to those skilled in the art and are readily identified by standard biological tests.

[0277] A synthetic NT peptide of the invention can also be used in combination with a sorbitol dehydrogenase inhibitor. Sorbitol dehydrogenase inhibitors lower fructose levels and have been used to treat or prevent diabetic complications such as neuropathy, retinopathy, nephropathy, cardiomyopathy, microangiopathy, and macroangiopathy.

[0278] A synthetic NT peptide of the invention can also be used in combination with a glucocorticoid receptor antagonist. The glucocorticoid receptor (GR) is present in glucocorticoid responsive cells where it resides in the cytosol in an inactive state until it is stimulated by an agonist. Upon stimulation the glucocorticoid receptor translocates to the cell nucleus where it specifically interacts with DNA and/or protein(s) and regulates transcription in a glucocorticoid responsive manner. GR antagonists can be used in the treatment of diseases associated with an excess or a deficiency of glucocorticoids in the body. As such, they may be used to treat the following: obesity, diabetes, cardiovascular disease, hypertension, Syndrome X, depression, anxiety, gout, hypercholesterolemia, hyperlipidemia, hyperlipoproteinemia, and the like. A variety of lipase inhibitors are known to one of ordinary skill in the art.

[0279] In addition, a synthetic NT peptide of the invention can be administered in combination with other pharmaceutical agents such as cholesterol biosynthesis inhibitors and cholesterol absorption inhibitors, especially HMG-CoA reductase inhibitors and HMG-CoA synthase inhibitors, HMG-CoA reductase and synthase gene expression inhibitors, CETP inhibitors, bile acid sequesterants, statins, ACAT inhibitors, squalene synthetase inhibitors, anti-oxidants and niacin. A synthetic NT peptide of the invention may also be administered in combination with naturally occurring compounds that act to lower plasma cholesterol levels. These naturally occurring compounds are commonly called nutraceuticals and include, for example, garlic extract, Benecon®, and niacin.

[0280] It is also contemplated that a synthetic NT peptide of the invention be administered with a lipase inhibitor and/or a glucosidase inhibitor, which are typically used in the treatment of conditions resulting from the presence of excess triglycerides, free fatty acids, cholesterol, cholesterol esters or glucose including, inter alia, obesity, hyperlipidemia, hyperlipoproteinemia, Syndrome X, and the like.

[0281] Any lipase inhibitor or glucosidase inhibitor may be used in a combination with a synthetic NT peptide of the invention. Preferred lipase inhibitors comprise gastric or pancreatic lipase inhibitors such as orlistat. Preferred glucosidase inhibitors comprise amylose inhibitors. A lipase inhibitor is a compound that inhibits the metabolic cleavage of dietary triglycerides into free fatty acids and monoglycerides. Under normal physiological conditions, lipolysis occurs via a two-step process that involves acylation of an activated serine moiety of the lipase enzyme. This leads to the production of a fatty acid-lipase hemicentral intermediate, which is then cleaved to release a diglyceride. Following further decylation, the lipase-fatty acid intermediate is cleaved, resulting in free lipase, a monoglyceride and a fatty acid. The resultant free fatty acids and monoglycerides are incorporated into bile acid-phospholipid micelles, which are subsequently absorbed at the level of the brush border of the small intestine. The micelles eventually enter the peripheral circulation as chylomicrons. Accordingly, compounds, including lipase inhibitors that selectively limit or inhibit the absorption of ingested fat precursors are useful in the treatment of conditions including obesity, hyperlipidemia, hyperlipoproteinemia, Syndrome X, and the like. A variety of lipase inhibitors are known to one of ordinary skill in the art.

[0282] Any glucosidase inhibitor may be used in combination with a synthetic NT peptide of the invention, however, a generally preferred glucosidase inhibitor is an amylase inhibitor. An amylase inhibitor is a glucosidase inhibitor that inhibits the enzymatic degradation of starch or glycogen into maltose. The inhibition of such enzymatic degradation is beneficial in reducing amounts of bioavailable sugars, including glucose and maltose, and the concomitant deleterious conditions resulting therefrom. Glucosidase and amylase inhibitors are known to one of ordinary skill in the art.

[0283] In addition, the present invention includes the use of a synthetic NT peptide of the invention in combination with apo B secretion/ MTP inhibitors. A variety of apo B secretion/ MTP inhibitors are known to one of ordinary skill in the art. A synthetic NT peptide of the invention also can be used in combination with one or more additional compounds that are neurotransmitter receptor ligands.

[0284] Non-natural amino acids of the invention can also be incorporated into other peptides to treat maladies as set out below.

[0285] Peptides for triggering B and T cell activity can be used to treat autoimmune disease, including uveitis, collagen-induced, adjuvant and rheumatoid arthritis, thyroiditis, myasthenia gravis, multiple sclerosis and diabetes. Examples of these peptides are interleukins (referred to in Allitsky, WE; Schuler, M; Peschel, C.; Huber C.; Interleukins. Clinical pharmacology and therapeutic use. Drugs. 48(5):667-77,

[0286] Enkephalin and analogs, agonists and antagonists can be used to treat AIDS, ARC, and cancer, pain modulation, Huntington's, Parkinson's diseases.

[0287] LHRH and analogs, agonists and antagonists can be used to treat prostatic tumors and reproductive physiopathology, including breast cancer, and infertility.

[0288] Peptides and peptidomimetics that target crucial enzymes, oncogenes or oncogenic products, tumor-suppressor genes and their products, growth factors and their corresponding receptors can be used to treat cancer. Examples of these peptides are described in Unger, C. Current concepts of treatment in medical oncology: new antinecancer drugs. Journal of Cancer Research & Clinical Oncology. 122(4):189-98, 1996.

[0289] Neuropeptide Y and other pancreatic polypeptides, and analogs, agonists and antagonists can be used to treat stress, anxiety, depression and associated vasocostrictive activities.

[0290] Glucore-incretins, including gastric inhibitory polypeptide, glucell-dependent insulinotropic polypeptide, PACAP/Glucagon and glucagon-like polypeptide-1 and 2 and analogs, agonists and antagonists can be used to treat Type II diabetic hyperglycemia.

[0291] Atrial natriuretic factor and analogs, agonists and antagonists can be used to treat congestive heart failure.

[0292] Integrin and analogs, agonists and antagonists can be used to treat osteoporosis, scar formation, bone synthesis, inhibition of vascular occlusion, and inhibition of tumor invasion and metastasis.

[0293] Glucagon, glucagon-like peptide 1, PACAP/Glucagon, and analogs, agonists and antagonists can be used to treat diabetes cardiovascular emergencies.

[0294] Antithrombotic peptides and analogs, agonists and antagonists can be used to treat cardiovascular and cerebrovascular diseases. Examples of these peptides RGD, D-Phe-Pro-Arg and others named are described in Ojima I.; Chakravarty S.; Dong Q. Antithrombotic agents: from RGD to peptide mimetics. Bioorganic & Medicinal Chemistry. 3(4):337-60, 1995.

[0295] Cytokines interleukins and analogs, agonists and antagonists can be used to treat inflammatory disease, immune response dysfunction, hematopoiesis, mycosis fungoides, aplastic anemia, thrombocytopenia, and malignant melanoma. Examples of these interleukins are referenced in Aulitzky et al. and Peters et al.

[0296] Endothelin and analogs, agonists and antagonists can be used to treat arterial hypertension, myocardial infarction, congestive heart failure, atherosclerosis, shock conditions, renal failure, asthma and vasospasm Natriuretic hormones and analogs, agonists and antagonists can be used to treat cardiovascular disease and acute renal failure. Examples of these peptides are named and described in Espiner, E. A.; Richards, A. M.; Yandle, T. G.; Nicholls, M. G.; Natriuretic hormones. Endocrinology & Metabolism Clinics of North America. 24(3):481-509, 1995.

[0297] Peptides that activate or inhibit tyrosine kinase, or bind to TK-activating or inhibiting peptides and analogs, agonists and antagonists can be used to treat chronic myelogenous and acute lymphocytic leukemias, breast and ovarian cancers and other tyrosine kinase associated diseases. Examples of these peptides are described in Smithgall, TE.; SH2 and SH3 domains: potential targets for anti-cancer drug design. Journal of Pharmacological & Toxicological Methods. 34(3):125-32, 1995.

[0298] Renin inhibitors analogs, agonists and antagonists can be used to treat cardiovascular disease, including hypertension and congestive heart failure. Examples of these peptides are described in Rosenberg, S. H.; Renin inhibition. Cardiovascular Drugs & Therapy. 9(5):645-55, 1995.

[0299] Angiotensin-converting enzyme inhibitors, analogs, agonists and antagonists can be used to treat cardiovascular disease, including hypertension and congestive heart failure.

[0300] Peptides that activate or inhibit tyrosine phosphorylases can be used to treat cardiovascular diseases. Examples of these peptides are described in Srivastava, A. K.; Protein tyrosine phosphorylation in cardiovascular system. Molecular & Cellular Biochemistry. 149-150:87-94, 1995.

[0301] Peptide based antivirals can be used to treat viral diseases. Examples of these peptides are described in Toes, R. E.; Feltkamp, M. C.; Ressing, M. E.; Vierboom, M. P.; Blom, R. J.; Brandt, R. M.; Hartman, M.; Offringa, R.; Melief, C. J.; Kast, W. M.; Cellular immunity against DNA tumour viruses: possibilities for peptide-based vaccines and immune escape. Biochemical Society Transactions. 23(3):692-6, 1995.

[0302] Corticotropin releasing factor and peptide analogs, agonists and antagonists can be used to treat disease associated with high CRF, i.e Alzheimer's disease, anorexia nervosa, depressive disorders, arthritis, and multiple sclerosis.

[0303] Peptide agonists and antagonists of platelet-derived wound-healing formula (PDWHF) can be used as a therapy for donor tissue limitations and wound-healing constraints in surgery. Examples of these peptides are described in Rudkin, G. H.; Miller, T. A.; Growth factors in surgery. Plastic & Reconstructive Surgery. 97(2):469-76, 1996.

[0304] Fibronectin, fibropeptide inhibitors and analogs, agonists and antagonists can be used to treat metastasis (i.e. enzyme inhibition, tumor cell migration, invasion, and metastasis).

[0305] Chemokine (types of cytokine, including interleukin-6, RANTES, and monocyte chemotactic peptide) analogs, agonists and antagonists can be used to treat arthritis, hyperesensitivity, angiogenesis, renal disease, gliomerulonephritis, inflammation, and hematopoesis.

[0306] Neutral endopeptidase inhibitors and analogs, agonists and antagonists can be used to treat hypertension and inflammation. Examples of these peptides are described in Gregoire, J. R.; Sheps, S. G.; Newer antihypertensive drugs. Current Opinion in Cardiology. 10(5):445-9, 1995.

[0307] Substance P and analogs, agonists and antagonists can be used to treat immune system dysfunction, pain transmission/perception and in autonomic reflexes and behaviors.

[0308] Alpha-melanocyte-stimulating hormone and analogs, agonists and antagonists can be used to treat AIDS, rheumatoid arthritis, and myocardial infarction.

[0309] Bradykinin (BK) and analogs, agonists and antagonists can be used to treat inflammatory diseases (edema, etc), asthma, allergic reactions (rhinitis, etc), anesthetic uses, and septic shock.

[0310] Secretin can be used to treat cardiovascular emergencies.

[0311] GnRH and analogs, agonists and antagonists can be used to treat hormone-dependent breast and prostate tumors.

[0312] Somatostatin and analogs, agonists and antagonists can be used to treat gut neuroendocrine tumors.
Gastrin, gastrin releasing peptide and analogs, agonists and antagonists can be used as an adjuvant to chemotherapy or surgery in small cell lung cancer and other malignancies, or to treat allergic respiratory diseases, asthma and allergic rhinitis.


Defensins, corticotropins, dermaseptins, manganins, and other antibiotic (antibacterial and antimiobacterial) peptides and analogs, agonists and antagonists can be used to treat infections, tissue inflammation and endocrine regulation.

Vasopressin and analogs, agonists and antagonists can be used to treat neurological disorders, stress and diabetes insipidus.

Oxytocin and analogs, agonists and antagonists can be used to treat neurological disorders and to induce labor.

ACTH-related peptides and analogs, agonists and antagonists can be used as neurotrophic, neuroprotective, and peripheral demyelinating neuropathy agents.

Amyloid-beta peptide and analogs, agonists and antagonists can be used to treat Alzheimer’s disease.

Epidermal growth factor, receptor, and agonists, antagonists and agonists can be used to treat neuropathic enterocolitis, Zollinger-Ellison syndrome, gastrointestinal ulceration, colitis, and congenital microvillus atrophycarcinoma.

Leukocyte adhesion molecules and their ligands, and agonists and antagonists can be used to treat atherosclerosis, inflammation. Examples of these peptides are described in Barker, J.N.; Adhesion molecules in cutaneous inflammation. Ciba Foundation Symposium. 189:91-101.

Major histocompatibility complex (MHC) binding peptides and analogs, agonists and antagonists can be used to treat autoimmune, immunodysfunctional, immuno modulatory diseases and as well as used for their corresponding therapies. Examples of these peptides are described in Appelria, E.; Padlan, E.A.; Hunt, D.F. Analysis of the structure of naturally processed peptides bound by class I and class II major histocompatibility complex molecules. EXS. 73:105-19, 1995.

Corticotropin releasing factor can be used to treat neurological disorders.

Neurotrophins (including brain-derived neurotrophic factor (BDNF), nerve growth factor, and neurotrophin 3) and analogs, agonists and antagonists can be used to treat neurological disorders.


Peptide immunogens for prevention of HIV-1 and HTLV-I retroviral infections can be used to treat AIDS. Examples of these peptides are described in Hart, M. K.; Palkar, T. J.; Haynes, B. F. Design of experimental synthetic peptide immunogens for prevention of HIV-1 and HTLV-I retroviral infections. Pharmaceutical Biotechnology. 6:821-45, 1995.

Galantin and analogs, agonists and antagonists can be used to treat Alzheimer’s disease, depression, eating disorders, chronic pain, prevention of ischemic damage, and growth hormone modulation.

Tachykinins (neurokinin A and neurokinin B) and analogs, agonists and antagonists can be used to treat pain transmission/perception and in autonomic reflexes and behaviors.

RGD containing peptides can be used to treat various diseases involved with cell adhesion, antithrombotics, and acute renal failure.

Osteogenic growth peptide and analogs, agonists and antagonists can be used as treatment of systemic bone loss. Examples of these peptides are described in Bab IA. Regulatory role of osteogenic growth peptide in proliferation, osteogenesis, and hemopoiesis. Clinical Orthopaedics & Related Research. (313):64-8, 1995.

Parathyroid hormone, parathyroid hormone related peptide and analogs, agonists and antagonists can be used to treat diseases affecting calcium homeostasis (hypocalcemia), bone metabolism, vascular disease, and atherosclerosis.

Kallidin and analogs, agonists and antagonists can be used to treat tissue injury or inflammation and pain signaling pathological conditions of the CNS.

T cell receptor peptide vaccines and analogs, agonists and antagonists can be used in immunotherapy. Examples of these peptides are described in Bostoff, S W; T cell receptor peptide vaccines as immunotherapy. Agents & Actions—Supplements. 47:53-8, 1995.

Platelet-derived growth factor (PDGF) and analogs, agonists and antagonists can be used to treat non-neoplastic hyperproliferative disorders, therapy for donor tissue limitations and wound-healing constraints in surgery.

Amylin, calcitonin gene related peptides (CGRP) and analogs, agonists and antagonists can be used to treat insulin-dependent diabetes.

Vasoactive intestinal polypeptide and analogs, agonists and antagonists can be used to treat allergic respiratory diseases, asthma and allergic rhinitis, and nervous control of reproductive functions.

Growth hormone-releasing hormone and analogs, agonists and antagonists can be used to treat growth hormone deficiency and immunomodulation.

HIV protease inhibiting peptides can be used to treat AIDS. Examples of these peptides are described in Bugelski,

[0339] Thymopoietin active fragment peptides and analogs, agonists and antagonists can be used to treat rheumatoid arthritis and virus infections.

[0340] Cecropins and analogs, agonists and antagonists can be used as antibacterials.

[0341] Thyroid releasing hormone and analogs, agonists and antagonists can be used to treat spinal cord injury and shock.

[0342] Erythropoietin and analogs, agonists and antagonists can be used to treat anemia.

[0343] Fibroblast growth factor (FGF), receptor and analogs, agonists and antagonists can be as stimulation of bone formation, as well as used as a treatment for Kaposi's sarcoma, neuron regeneration, prostate growth, tumor growth inhibition, and angiogenesis.

[0344] Stem cell factor and analogs, agonists and antagonists can be used to treat anemias.

[0345] GP120, GP160, CD4 fragment peptides and analogs, agonists and antagonists can be used to treat AIDS.

[0346] Insulin-like growth factor, receptor, and analogs, agonists and antagonists can be used to treat breast and other cancers, non-insulin-dependen diabetest mellitus, cell proliferation, apoptosis, hematoipoiesis, AIDS, growth disorders, osteoporosis, and insulin resistance.

[0347] Colony stimulating factors (granulocyte-macrophage colony-stimulating factor, granulocyte colony-stimulating factor, and macrophage colony-stimulating factor, and analogs, agonists and antagonists can be used to treat anemias.

[0348] Kentsin and analogs, agonists and antagonists can be used for immunomodulation.


[0350] Tuftsin and analogs, agonists and antagonists can be used for immunomodulation.

[0351] Prolactin and analogs, agonists and antagonists can be used to treat rheumatic diseases, systemic lupus erythematosus, and hyperprolactemia.

[0352] Angiotensin II and receptor(s) and analogs, agonists and antagonists can be used to treat hypertension, hemodynamic regulation, neurological disorders, diabetic nephropathies, aortoarteries induced RVH, hyperaldosteronism, heavy metal induced cardiovascular effects, diabetes mellitus and thyroid dysfunction.

[0353] Dynorphin and analogs, agonists and antagonists can be used to treat neurological disorders, pain management, algesia, spinal cord injury and epilepsy.

[0354] Calcitonin, and analogs, agonists and antagonists can be used to treat neurological disorders, immune system dysfunction, calcium homeostasis, and osteoporosis.

[0355] Pituiary adenylate cyclase activating polypeptide can play a role in growth, signal transduction vasoactivity roles, exact role in diseases not determined yet.

[0356] Cholecystokinin and analogs, agonists and antagonists can be used to treat feeding disorders, panic disorders, and anti-opiod properties.

[0357] Pepstatin and analogs, agonists and antagonists can be used as peptic and H1 protease inhibitors (AIDS).

[0358] Bestatin and analogs, agonists and antagonists can be used to treat muscular dystrophy, anticeancer, antileukemia, immune response modulator, and acute non-lymphocytic leukemia.

[0359] Leupeptin and analogs, agonists and antagonists can be used as a protease inhibitor, exact role in diseases not determined yet.

[0360] Luteinizing hormone and releasing hormone and analogs, agonists and antagonists can be used as an infertility mal contraceptive.

[0361] Neotensin and analogs, agonists and antagonists can be used, e.g., as antipsychotic, analgesic, anti-cancer, and/or neuroprotective agents, e.g., for treating stroke victims, e.g., by inducing hypothermia so as to provide neuromutation.

[0362] Motilin and analogs, agonists and antagonists can be used for the control of gastric emptying.

[0363] Insulin and analogs, agonists and antagonists can be used to treat diabetes.

[0364] Transforming growth factor (TGF) and analogs, agonists and antagonists can be used for cell proliferation and differentiation, cancer treatment, immunoregulation, therapy for donor tissue limitations, and wound-healing constraints in surgery.

[0365] Bone morphogenetic proteins (BMPs) and analogs, agonists and antagonists can be used as therapy for donor tissue limitations, osteogenesis, and wound-healing constraints in surgery.


[0367] Glucagon, glucagon-like peptide 1 and analogs, agonists and antagonists can be used to treat diabetes cardiovascular emergencies.


[0369] Endorphins and analogs, agonists and antagonists can be used to treat neurological disorders, alleviating pain, treatment of opioid abuse, obesity, and diabetes. Examples of these peptides are named and described in Dallayeun, J. F.;

Miscellaneous opioid peptides, including (but not limited to) adrenal peptide E, alpha casein fragment, beta casomorphin, dermorphin, kyotorphin, metophamide neuropetide FF (NPPFF), melanocyte inhibiting factor, and analogues, agonists and antagonists can be used to treat neurological disorders, alleviating pain, as well as for the treatment of opioid abuse.

Vasotocin and analogues, agonists and antagonists can be used for clinical uses to be determined.

Protein kinase C and inhibitors and analogues, agonists and antagonists can be used to treat cancer, apoptosis, smooth muscle function, and Alzheimer's disease. Examples of these peptides are named and described in Philip, P. A.; Harris, A. I.; Potential for protein kinase C inhibitors in cancer therapy. Cancer Treatment & Research. 78:3-27, 1995.

Amyloid, amyloid fibrin, fragments and analogues, agonists and antagonists can be used to treat neurodegenerative diseases and diabetes.

Calpain and other calmodulin-inhibitory proteins and analogues, agonists and antagonists can be used to treat neurodegenerative disorders, cerebral ischaemia, catatras, myocardial ischaemia, muscular dystrophy and platelet aggregation.

Charybdotoxin, Apamin and analogues, agonists and antagonists can be used for treatment of neurodegenerative diseases and pain and cerebral ischemia.

Phospholipase A2 and receptor inhibiting/activating peptides and analogues, agonists and antagonists can be used to treat acute pancreatitis, pancreatic cancer, abdominal trauma, and inflammation, e.g., sepsis, infections, acute pancreatitis, various forms of arthritis, cancer, complications of pregnancy, and postoperative states.

Potassium channel activating and inhibiting proteins and analogues, agonists and antagonists can be used to treat various diseases. Examples of these peptides are described in Edwards, G.; Weston, A. H. Pharmacology of the potassium channel openers. Cardiovascular Drugs & Therapy. 9 Suppl 2:185-93, March 1995.

IgG activators, inhibitors and analogues, agonists and antagonists can be used to treat autoimmune diseases and immune dysfunctions. Examples of these peptides are described in Mouton L.; Kaveri, S. V.; Spalter, S. H.; Lacrazo-Desmares, S.; Lefmme, C.; Desai, R.; Kazatchkine, M. D; Mechanisms of action of intravenous immune globulin in immune-mediated diseases. Clinical & Experimental Immunology. 104 Suppl 1:3-9, 1996.

Endotoxin and inhibitors and analogues, agonists and antagonists can be used for decreasing cardiac output, systemic hypotension, decreased blood flow and O2 delivery to tissues, intense pulmonary vasoconstriction and hypertension, bronchoconstriction, increased permeability, pulmonary oedema, ventilation-to-perfusion inequalities, hypoxaemia, and haemoconcentration. Examples of these peptides are named and described in Burrell, R; Human responses to bacterial endotoxin. Circulatory Shock. 43(3):137-53, July 1994.


The Use of Peptides Containing Desamino Amino Acid(s) to Pass a Body Barrier

The invention relates to a method of increasing the ability of a peptide to cross a body barrier of a subject by use of the extended or truncated peptide having as its N-terminus a residue of the compound formula I-V.

The invention further relates to a method of treating or preventing in a subject a disease or condition treated or prevented by the administration of an extended or truncated peptide, whereby the extended or truncated peptide crosses the body barrier in higher amounts than the peptide having no non-natural amino acid.

The invention also relates to a method of treating or preventing in a subject a disease or condition of the brain treated or prevented by the administration of an extended or truncated peptide.

The use of peptides as therapeutic agents is limited by their inability to cross body barriers. The phrase “body barrier” is defined herein as a cellular membrane or other structure that functions to prevent free (e.g., diffusional) passage of certain molecules. The use of an extended or truncated peptide of the invention facilitates the passage of the resultant peptide through a variety of body barriers. Examples of body barriers include, but are not limited to, the blood brain barrier,
a cell membrane, intestinal epithelium, skin cell, or the bloodocular. In a preferred embodiment, the body barrier is the blood brain barrier.

Selectivity and Stability of Peptides Containing Non-Natural Amino Acid(s)

[0386] Certain embodiments of the invention relate to a method of increasing the selectivity of a chosen peptide through use of an extended or truncated peptide based upon the sequence of the chosen peptide as described above.

[0387] Enhancing the selectivity of a drug to a biological target is of great importance. In one embodiment, a peptide containing arginine and/or lysine can be converted according to the invention into an extended or truncated peptide in order to increase the selectivity of the peptide. In another embodiment, any of the non-natural amino acids disclosed herein can be used to increase the selectivity of a peptide.

Pharmaceutical Composition

[0388] The peptides of the invention can be used in any therapeutic procedure available to one of skill in the art to treat any disease or physiological problem with which the corresponding known peptide is associated.

[0389] The peptides of the invention can be formulated as pharmaceutical compositions and administered to a mammalian host, such as a human patient, in a variety of dosage forms adapted to the chosen route of administration, i.e., orally or parenterally, by intravenous, intramuscular, topical or subcutaneous routes.

[0390] Thus, the peptides may be systemically administered, for example, intravenously or intraperitoneally by infusion or injection. Solutions of the peptide or peptide conjugate can be prepared in water, optionally mixed with a non-toxic surfactant. Dispersions can also be prepared in gelatin, liquid polyethylene glycol, triacetin, and mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations contain a preservative to prevent the growth of microorganisms.

[0391] The pharmaceutical dosage forms suitable for injection or infusion can include sterile aqueous solutions or dispersions or sterile powders comprising the active ingredient(s) that are adapted for the extemporaneous preparation of sterile injectable or infusible solutions or dispersions, optionally encapsulated in liposomes. In all cases, the ultimate dosage form must be sterile, fluid and stable under the conditions of manufacture and storage. The liquid carrier or vehicle can be a solvent or liquid dispersion medium comprising, for example, water, ethanol, a polyol (for example, glycerol, propylene glycol, liquid polyethylene glycol, and the like), vegetable oils, non-toxic glycerol esters, and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the formation of liposomes, by the maintenance of the required particle size in the case of dispersions or by the use of surfactants. The prevention of the action of microorganisms can be brought about by various antibacterial and antifungal agents, for example, parabens, chlorbutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars, buffers or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

[0392] Sterile injectable solutions are prepared by incorporating the peptide or peptide conjugate in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filter sterilization. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods for preparation of such powders are vacuum drying and freeze-drying techniques, which yield a powder of the active ingredient plus any additional desired ingredient present in the previously sterile-filtered solutions.

[0393] In some instances, the peptides of the invention can also be administered orally, in combination with a pharmaceutically acceptable vehicle such as an inert diluent or an absorbable edible carrier. The peptides may be enclosed in hard or soft shell gelatin capsules, may be compressed into tablets, or may be incorporated directly with the food of the patient's diet. For oral therapeutic administration, the peptide or peptide conjugate may be combined with one or more excipients and used in the form of ingestible tablets, buccal tablets, troches, capsules, elixirs, suspensions, syrups, wafers, and the like. Such compositions and preparations should contain at least 0.1% of active compound. The percentage of the compositions and preparations may, of course, be varied and may conveniently be between about 2 to about 60% of the weight of a given unit dosage form. The amount of peptide in such therapeutically useful compositions is such that an effective dosage level will be obtained.

[0394] The tablets, troches, pills, capsules, and the like may also contain the following: binders such as gum tragacanth, acacia, corn starch or gelatin; excipients such as dicalcium phosphate; a disintegrating agent such as corn starch, potato starch, alginic acid and the like; a lubricant such as magnesium stearate; and a sweetening agent such as sucrose, fructose, lactose or aspartame or a flavoring agent such as peppermint, oil of wintergreen, or cherry flavoring may be added. When the unit dosage form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier, such as a vegetable oil or a polyethylene glycol. Various other materials may be present as coatings or to otherwise modify the physical form of the solid unit dosage form. For instance, tablets, pills, or capsules may be coated with gelatin, wax, shellac or sugar and the like. A syrup or elixir may contain the active compound, sucrose or fructose as a sweetening agent, methyl and propylparabens as preservatives, a dye and flavoring such as cherry or orange flavor. Of course, any material used in preparing any unit dosage form should be pharmaceutically acceptable and substantially non-toxic in the amounts employed. In addition, the peptides of the invention may be incorporated into sustained-release preparations and devices.

[0395] Useful solid carriers include finely divided solids such as talc, clay, microcrystalline cellulose, silica, alumina and the like. Useful liquid carriers include water, alcohol or glycols or water-alcohol/glycol blends, in which the present compounds can be dissolved or dispersed at effective levels, optionally with the aid of non-toxic surfactants. Adjuvants such as fragrances and additional antimicrobial agents can be added to optimize the properties for a given use.

[0396] Thickeners such as synthetic polymers, fatty acids, fatty acid salts and esters, fatty alcohols, modified celluloses or modified mineral materials can also be employed with liquid carriers to form spreadable pastes, gels, ointments, soaps, and the like, for application directly to the skin of the user.
Useful dosages of the peptides of the invention can be determined by correlating their in vitro activity, and in vivo activity in animal models described herein.

The therapeutically effective amount of peptide of the invention necessarily varies with the subject and the disease or physiological problem to be treated and correlates with the effective amounts of the corresponding known peptide. For example, a therapeutic amount between 30 to 112,000 μg per kg of body weight can be effective for intravenous administration. As one skilled in the art would recognize, the amount can be varied depending on the method of administration. The amount of the peptide of the invention, required for use in treatment will also vary with the route of administration, but also the nature of the condition being treated and the age and condition of the patient and will be ultimately at the discretion of the attendant physician or clinician.

The compound can conveniently be administered in unit dosage form; for example, containing 1 to 1000 mg, conveniently 10 to 750 mg, most conveniently, 20 to 500 mg of peptide per unit dosage form.

Ideally, the peptide should be administered to achieve peak plasma concentrations of from about 0.1 to about 75 μM, preferably, about 1 to 50 μM, most preferably, about 2 to about 30 μM. This may be achieved, for example, by the intravenous injection of a 0.05 to 5% solution of the peptide, optionally in saline, or orally administered as a bolus containing about 1-100 mg of the peptide. Desirable blood levels may be maintained by continuous infusion to provide about 0.01-5.0 mg/kg/hr or by intermittent infusions containing about 0.4-15 mg/kg of the active ingredient(s).

The desired dose may conveniently be presented in a single dose, as divided doses, or as a continuous infusion. The desired dose may also be administered at appropriate intervals, for example, as two, three, four or more sub-doses per day.

Cosmetic Formulation

An important role for makeup cosmetic is “beautification” or making the appearance more beautiful. Often which role involves correction of skin roughness, blemishes and color as well as vitality.

A cosmetic composition of the present invention contains the typical and common base carriers as well as a desamin amino acid compound of the invention. Usually the compound of the invention will be in the form of an ester, amide or salt for this purpose. Generally, the cosmetic base will depend upon the kind of make-up being formulated: face creme, face powder, pancake make-up, skin creme, lip stick, rouge and the like. These bases will contain appropriate, nontoxic colorants, emulsuants, oils, waxes, solvents, emulsifiers, fatty acids, alcohols or esters, gums, inorganic inert builders and the like.

For example, the gums, may include various known polysaccharide compounds, for example, cellulose, hemicellulose, gum arabic, tragacanth gum, tamarind gum, pectin, starch, mannann, guar gum, locust bean gum, quinice seed gum, algicin acid, carrageenan, agar, xanthane gum, dextran, pul lulan, chitin, chitosan, hyaluronic acid, chondroitin sulfuric acid, etc., derivatives of polysaccharide compounds, for example, carboxymethylated derivatives, sulfate derivatives, phosphated derivatives, methylated derivatives, ethylated derivatives, addition derivatives of alkylene oxide such as ethylene oxide or propylene oxide, acylated derivatives, carionated derivatives, low molecular weight derivatives, and other polysaccharide derivatives may be mentioned.

Another component which may be included in the external composition of the present invention is a powder component. Powder components based on inorganic components such as talc, kaolin, mica, sericite, dolomite, phlogopite, synthetic mica, lepidolite, biotite, lithia mica, vermiculite, magnesium carbonate, calcium carbonate, aluminum silicate, barium silicate, calcium silicate, magnesium silicate, strontium silicate, metal salts of tungstic acid, magnesium, silica, zeolite, barium sulfate, sintered calcium sulfate (sintered gypsum), calcium phosphate, fluorapatite, hydroxyapatite, ceramic powder, metal soap (zinc myristate, calcium palmitate, ammonium stearate), boronitrile, etc.; and organic powder components such as polyamide resin powder (nylon powder), polyethylene powder, polymethyl methacrylate powder, polystyrene powder, copolymer resin powder of styrene and acrylic acid, benzoguanamine resin powder, silicone resin powder, silicone rubber powder, silicone resin covered rubber powder, polyethylene tetrfluoride powder, cellulose powder, etc. may be mentioned.

Further, powder components obtained by treating the surfaces of these powder components by a silicone compound, fluorine-modified silicone compound, fluorine compound, higher aliphatic acid, higher alcohol, aliphatic acid ester, metal soap, alkyl phosphate, etc. may be formulated into the external composition of the present invention depending upon the need.

The known dyes or pigments, may be used. For example, inorganic white pigments such as titanium dioxide, zinc oxide; inorganic red pigments such as iron oxide (bengala), iron titinate, inorganic brown pigments such as gamma-iron oxide; inorganic yellow pigments such as yellow iron oxide, yellow earth; inorganic black pigments such as black iron oxide, carbon black, lower titanium oxide, and; inorganic violet pigments such as manganese violet, cobalt violet; organic green pigments such as chromium oxide, chromium hydroxide, cobalt titinate; blue pigments such as prussian blue, ultramarine; pearl pigments such as titanium oxide coated mica, titanium oxide coated bismuth oxychloride, titanium oxide coated talc, colored titanium oxide coated mica, bismuth oxychloride, fish scales; metal powder pigments such as aluminum powder, copper powder, organic pigments of zirconium, barium or aluminum lakes etc. such as Lithol rubine R (Red No. 201), Lithol rubine BCA (Red No. 204), Lithol red (Red No. 205), Deep maroon (Red No. 220), Heliolone pink CN (Red No. 226), Permatone Red (Red No. 228), Permanent red FSR (Red No. 405), Permanent orange (Orange No. 203), Benzidine Orange (Orange No. 204), Benzidine yellow G (Yellow No. 205), Hanzu yellow (Yellow No. 401), Blue No. 404, and other organic pigments; Erythrosine (Red No. 3), Phloxine B (Red No. 104), Acid red (Red No. 106), Fast acid magenta (Red No. 227), Eosine YS (Red No. 230), Violamine R (Red No. 401), Oil red XO (Red No. 505), Orange II (Orange No. 205), Tartrazine (Yellow No. 4), Sunset yellow FCF (Yellow No. 5), Uranine (Yellow No. 202), Quinoline yellow (Yellow No. 203), Fast green FCF (Green No. 3), Brilliant blue FCF (Blue No. 1) may be mentioned.

The cosmetic composition of the present invention may be formulated with a liquid. As the liquid, it is possible to select a volatile component ordinarily used in external compositions such as cosmetics. Specifically, it is possible to mention, for example, volatile silicone oil, water, or a lower
alcohol (or mixtures of the same). These volatile components may be suitably selected depending upon the specific form of the external composition of the present invention (for example, the later mentioned “roughness correcting composition” or “makeup composition” etc.) or type of carrier (for example, oil base or emulsion base etc.). By formulating these volatile components, it is possible to adjust the viscosity of the product at the time of use of the external composition of the present invention and adjust the thickness of coating of the external composition on the skin.

[0409] As the volatile silicone oil, it is possible to use a volatile silicone oil which is used in the field of cosmetics and other external compositions. It is not particularly limited. Specifically, for example, a low boiling point linear silicone oil such as hexamethyl disiloxane, octamethyl trisiloxane, decamethyl cyclopentasiloxane, dodecamethyl cyclohexasiloxane, and tetradecamethyl cycloheptasiloxane; a low boiling point cyclic silicone oil such as octamethyl cycloetetrasiloxane, decamethyl cyclopentasiloxane, dodecamethyl cyclohexasiloxane, and tetradecamethyl cycloheptasiloxane; etc. may be mentioned.

[0410] The external composition of the present invention may contain, depending upon the need, the following components as auxiliary components to an extent not detracting from the desired effect of the present invention.

[0411] For example, as the oil component, hydrocarbon oils such as liquid paraffin, isoiliquid paraffin, squalane, oils and fats such as olive oil, palm oil, coconut oil, macadamia nut oil, jojoba oil; higher alcohols such as isostearl alcohol; ester oils such as higher aliphatic oils and isopropyl myristate, etc. may be formulated in the external composition of the present invention. Among these oil components, in particular, formulating a polar oil in the external composition of the present invention enables improvement of the stability with the elapse of time.

[0412] Further, a benzophenone derivative, para-amino benzoate derivative, para-methoxysuccinate derivative, salicylate derivative, and other UV absorbers; humectants, blood circulation promoters, refrigerants, antiperspirants, bactericides, skin activators, anti-inflammatory agents, vitamins, antioxidants, antioxidant adjuvants, preservatives, flavors and fragrances, etc. may be blended in the external composition of the present invention.

[0413] The cosmetic formulation of the present invention may be produced in an appropriate medium including but not limited to a paste, a powder, a cake, a creme, an oil, a lotion, a grease, a wax or similar cosmetic bases. The process to produce involves combining the cosmetic ingredients and desaminino amino acid compound of any of formulas I-V preferably as the ester, amide or salt. The combination is mixed, kneaded, rolled, ground, heated or otherwise treated to form a substantially homogeneous mass or mixture for use. These steps can be accomplished by use of a kneader, grind wheel, rollers, mixer, heat exchanger, extruder and the like.

[0414] As explained above, the invention is exemplified by modification of the natural peptide neurotensin. In the following section, the background, modification and biological activity of neurotensin and the corresponding peptides of the invention are discussed.

Neurotensin Structure and Biology

[0415] Neurotensin (NT) was first isolated from bovine hypothalami as a hypotensive peptide by Caraway and Leeman in 1973. Since then, NT has been shown to have numerous distinct physiological effects in the central nervous system (CNS) and the periphery. Hypothermia, antinociception, attenuation of d-amphetamine-induced hyperlocomotion, and potentiation of barbiturate-induced sedation are promoted by direct injection of NT into the brain. Peripherally, NT acts as a hormone to induce hypotension and decrease gastric acid secretion. Structurally, NT is a linear tripeptide with the following sequence: pGlu-Leu-Tyr-Glu-Asn-Lys-Pro-Arg-Pro-Tyr-Ile-Leu-OH. Early in the history of NT research, it was shown that the C-terminal hexapeptide fragment, Arg⁵-Arg⁴-Pro¹⁰-Tyr¹¹-Ile¹²-Leu¹³ [NT(8-13)], was equipotent at producing the physiological effects of NT in vitro and in vivo.

[0416] Tanaka and colleagues first identified an NT receptor (NTR₁) from rat brain in 1990. Since then, human NTR₁ has been successfully cloned and expressed. Both are classic G-protein coupled receptors containing seven transmembrane (7TM) domains and share 84% homology. Second messenger systems, including cGMP production, calcium mobilization, and phosphatidylinositol turnover, are triggered upon NTR₁ activation. The mRNA for NTR₁ is expressed in both rat brain and human intestine. A second NT receptor (NTR₂) with a substantially lower affinity for NT than NTR₁ (Kᵣ=2.5 and 0.5 nM respectively), also has been identified in rat and human brain (23-25). NTR₂ is also a 7TM G-protein coupled receptor, yet has a shorter N-terminal extracellular tail and a longer third intracytoplasmic loop compared to NTR₁. A third receptor (NTR₃) was cloned from a human brain cDNA library and found to be identical to the previously cloned gp95/ sortilin. NTR₃ is a non-G-protein coupled sorting protein having only a single transmembrane region.

NT as an Endogenous Neuroleptic

[0417] Several distinct lines of evidence implicate NT in the pathophysiology of schizophrenia. Advances in the dopamine theory of schizophrenia support that a flaw in the convergence of various neural circuits on the mesolimbic dopamine system is responsible for the development of the disorder. The anatomical positioning of the NT system is such that it interacts with the glutaminergic, dopaminergic, GABAergic, and serotonergic systems within the brain. In particular the NT and dopamine systems are closely related within the nucleus accumbens, the area of the brain believed to be responsible for delusions and hallucinations. NTR₁ receptors are dense in the ventral tegmental area, a brain region closely associated with the neuronal systems described above. Almost 90% of NT receptors are located on dopaminergic neurons and over 80% of dopamine neurons in the brain express NTR₁. Co-localization of the NT system with brain regions implicated in schizophrenia also implies its involvement.

Neurotensin and Its Biological Activity

[0418] Since NT was hypothesized as an “endogenous neuroleptic” and NT(8-13) was identified as its active fragment, efforts have been made to develop NT(8-13) derivatives as potential antipsychotics. Two groups in particular, the Eisai pharmaceutical company (Tokyo, Japan) and the Richelson research group (Mayo Clinic, Jacksonville, Fla.) have prepared numerous derivatives of NT(8-13) analogues that showed promise as antipsychotic drugs. In particular, amino
acid substitutions at Arg<sup>8</sup>, Arg<sup>9</sup>, Tyr<sup>11</sup>, and Ile<sup>12</sup> have yielded several analogues that are centrally active after peripheral administration.

**[0419]** An Eisai compound (the Eisai hexapeptide) was the first NT(8-13) analogue that elicited behavioral effects after peripheral administration. However, the various modifications incorporated in this peptide resulted in a 700-fold loss of binding affinity at NTR<sub>2</sub>. In addition, this analogue was not able to elicit central activity after oral administration.

**[0420]** More recently, NT69L has been developed by the Richelot group as an NT(8-13) analogue that maintains nanomolar binding affinity at NTR<sub>2</sub> (K<sub>d</sub> = 1.55 nM) (55) and exhibits a pronounced hypothermic effect after a 1 mg/kg injection (~5°C at 90 min PI) (41). NT69L also attenuates hyperactivity induced by both cocaine and d-amphetamine.

However, tolerance to its hypothermic effect and to its suppression of d-amphetamine-induced hyperactivity was observed after chronic administration of the compound. As with the Eisai hexapeptide, NT69L produced only a slight hypothermic response after oral administration.

Methods of Preparation of Peptide Analogs

**[0421]** An embodiment of the present invention concerns a method of preparation of a compound of Formula (XI):

\[
\text{(XI)}
\]

**[0422]** or a salt, hydrate, solvate, stereoisomer, or tautomer thereof, wherein X is OH, O(C<sub>2</sub>-C<sub>6</sub> alkyl), O(C<sub>6</sub>-C<sub>10</sub> aryl), NR<sub>2</sub>, an amino acid residue or a peptide residue, or wherein X is O, NR, an amino acid residue, or a peptide residue, respectively covalently bonded to a polymer;

**[0423]** Z is H, NR<sub>2</sub>, alkyl, aryl, aralkyl, heterocyclyl, heteroaryl, heterocyclylalkyl, heterocyclylalkyl, cycloalkyl, cycloalkylalkyl, an amino acid residue, or a peptide residue, or is NR, an amino acid residue, or a peptide residue, respectively covalently bonded to a polymer;

**[0424]** provided that X and Z are not both bonded to polymers; and provided that at least one of X or Z comprises an amino acid residue or a peptide residue, or an amino acid residue or a peptide residue respectively covalently bonded to a polymer;

**[0425]** R is independently at each occurrence H, alkyl, aryl, heterocyclyl, heteroaryl, a nitrogen protecting group, or a nitrogen capping group, or two R groups together with the nitrogen atom to which they are bound form a 5-7 membered heterocyclic ring that can further comprise 1-3 additional heteroatoms N, O or S;

**[0426]** L is a saturated or unsaturated, linear, branched or cyclic hydrocarbyl or a combination thereof, comprising n carbon atoms, wherein any carbon atom can be substituted or unsubstituted, wherein 0-3 heteroatoms O, S, or NR can be substituted for a carbon atom of L;

**[0427]** n is 1 to about 10;

**[0428]** R<sup>1</sup>, R<sup>2</sup> and R<sup>3</sup> are independently hydrogen, alkyl, aryl, aralkyl, cycloalkyl, heterocyclyl, or heteroaryl, wherein one of R<sup>1</sup>, R<sup>2</sup> or R<sup>3</sup> may be absent, wherein any alkyl can contain 0-3 heteroatoms comprising N, O, or S, wherein any alkyl, aryl, cycloalkyl, heterocyclyl, or heteroaryl may be unsubstituted or substituted;

**[0429]** the method comprising contacting a compound of Formula (XII):

\[
\text{(XII)}
\]

**[0430]** wherein Y is a leaving group;

**[0431]** with an amine of the formula R<sup>4</sup>R<sup>5</sup>R<sup>6</sup>N, in a solvent for a time period and at a temperature sufficient for the formation of the compound of Formula (XI);

**[0432]** An embodiment of the invention also concerns a method for the preparation of a compound of Formula (XIII):

\[
\text{(XIII)}
\]

**[0433]** or a salt, hydrate, solvate, stereoisomer, or tautomer thereof,

**[0434]** wherein X is OH, O(C<sub>2</sub>-C<sub>6</sub> alkyl), O(C<sub>6</sub>-C<sub>10</sub> aryl), NR<sub>2</sub>, an amino acid residue or a peptide residue, or wherein X is O, NR, an amino acid residue, or a peptide residue, respectively covalently bonded to a polymer;

**[0435]** Z is H, NR<sub>2</sub>, alkyl, aryl, aralkyl, heterocyclyl, heteroaryl, heterocyclylalkyl, heterocyclylalkyl, cycloalkyl, cycloalkylalkyl, an amino acid residue, or a peptide residue, or is NR, an amino acid residue, or a peptide residue, respectively covalently bonded to a polymer;

**[0436]** provided that X and Z are not both bonded to polymers; and provided that at least one of X or Z comprises an amino acid residue or a peptide residue, or an amino acid residue or a peptide residue respectively covalently bonded to a polymer;

**[0437]** R is independently at each occurrence H, alkyl, aryl, heterocyclyl, heteroaryl, a nitrogen protecting group, or a nitrogen capping group, or a nitrogen capping group, or two R groups together with the nitrogen atom to which they are bound form a 5-7 membered heterocyclic ring that can further comprise 1-3 additional heteroatoms N, O or S;

**[0438]** L is a saturated or unsaturated, linear, branched or cyclic hydrocarbyl or a combination thereof, comprising n carbon atoms, wherein any carbon atom can be substituted or unsubstituted, wherein 0-3 heteroatoms O, S, or NR can be substituted for a carbon atom of L;

**[0439]** n is 1 to about 10;

**[0440]** R<sup>4</sup> and R<sup>5</sup> are independently at each occurrence H, alkyl, cycloalkyl, heterocyclyl, aryl or heteroaryl, wherein
any alkyl, cycloalkyl, aryl, heterocyclic or heteroaryl residue may be unsubstituted or substituted; provided that R or R is not a Boc group;

[0441] the method comprising contacting a compound of Formula (XV):

![Chemical structure](image1)

wherein R is hydrogen, alkyl, aryl, aralkyl, cycloalkyl, heterocycles, or heteroaryl; wherein any alkyl can further contain 0-3 heteroatoms comprising N, O, or S, wherein any alkyl, aryl, cycloalkyl, heterocyclic, or heteroaryl may be unsubstituted or substituted;

[0442] with a compound of Formula (XIV):

![Chemical structure](image2)

wherein R is substituted or unsubstituted alkyl or aryl, or a salt thereof;

[0444] in a solvent for a time period and at a temperature sufficient for the formation of the compound of Formula (XIII).


![Chemical structure](image3)

or a salt, hydrate, solvate, stereoisomer, or tautomer thereof;

[0447] wherein X is OH, O(C_1-C_6 alkyl), O(C_6-C_{10} aryl), NR_3, an amino acid residue or a peptide residue, or wherein X is O, NR, an amino acid residue, or a peptide residue, respectively covalently bonded to a polymer;

[0449] Z is H, NR_2, alkyl, aryl, aralkyl, heterocyclic, heteroaryl, heterocyclylalkyl, heteroarylylalkyl, cycloalkyl, cycloalkylalkyl, an amino acid residue, or a peptide residue, or is NR, an amionic acid residue, or a peptide residue, respectively covalently bonded to a polymer;

[0450] provided that X and Z are not both bonded to polymers; and provided that at least one of X or Z comprises an amino acid residue or a peptide residue, or an amino acid residue or a peptide residue respectively covalently bonded to a polymer;

[0451] R at each occurrence is independently H, alkyl, aryl, heterocyclic, heteroaryl, a nitrogen protecting group, or a nitrogen capping group, or two R groups together with the nitrogen atom to which they are bound form a 5-7 membered heterocyclic ring that can further comprise additional heteroatoms N, O or S;

[0452] L is a saturated or unsaturated, linear, branched or cyclic hydrocarbyl or a combination thereof, comprising n carbon atoms, wherein any carbon atom can be substituted or unsubstituted, wherein 0-3 heteroatoms O, S, or NR can be substituted for a carbon atom of L;

[0453] n is 1 to about 10;

[0454] R is hydrogen, alkyl, aryl, aralkyl, cycloalkyl, heterocycles, or heteroaryl; wherein any alkyl can contain 0-3 heteroatoms comprising N, O, or S, wherein any alkyl, aryl, cycloalkyl, heterocyclic, or heteroaryl may be unsubstituted or substituted;

[0455] R and R are at each occurrence independently H, alkyl, cycloalkyl, heterocyclic, aryl or heteroaryl, wherein any alkyl, cycloalkyl, aryl heterocyclic or heteroaryl residue may be unsubstituted or substituted; provided that R or R is not a Boc group;

[0456] from a compound of formula (XII):

![Chemical structure](image4)

wherein Y is a leaving group;

[0457] wherein Z and Z are at each occurrence independently H, alkyl, cycloalkyl, heterocyclic, aryl or heteroaryl, wherein any alkyl, cycloalkyl, aryl heterocyclic or heteroaryl residue may be unsubstituted or substituted; provided that R or R is not a Boc group;

[0458] the method comprising contacting the compound of formula (XII) with an amine of the formula R'NH_2, in a solvent for a time period and at a temperature sufficient for the formation of the compound of Formula (XV):

![Chemical structure](image5)

then, contacting the compound of formula (XV) with a compound of Formula (XIV):

![Chemical structure](image6)

wherein R is substituted or unsubstituted alkyl or aryl, or a salt thereof;
[0461] in a solvent for a time period and at a temperature sufficient for the formation of the compound of Formula (XIII).

[0462] In the method of preparation of a compound of formula (XI) from a compound of formula (XII), of the method of preparation of a compound of formula (XV) from a compound of formula (XII) as an intermediate in the method of preparation of a compound of formula (XIII), the compound of formula (XII) comprises Y being a leaving group, which can include Y=halo, for example, Y=chloro, bromo or iodo, as well as Y=a sulfonate ester, for example alkyloxy sulfonil (e.g., methylsulfonyl), haloalkyloxy sulfonil (e.g., trihalomethylsulfonyl) or arylsulfonyl (e.g., p-toluenesulfonyl). Y, as a leaving group, undergoes nucleophile displacement by NR'R'R'2 (in the preparation of formula (XII)), or by R'NH2 (in the preparation of formula (XV)), at the carbon center to which Y is attached.

[0463] As is known in the art, certain groups are better leaving groups or have better “leaving ability”, so it is within the knowledge of a skilled artisan to select without undue experimentation an appropriate leaving group based upon the degree of reactivity of the reagent amine. When an amine that is more sterically hindered, or electronically less nucleophile (e.g., more electron-withdrawn) is used, a leaving group Y of greater leaving ability will be selected in order to achieve a reasonable reaction rate such that the product is obtained in good yield in a relatively short time period and with minimal side reactions. For example, an amine like trimethylamine is both highly nucleophile and unhindered sterically. Therefore a poorer leaving group Y such as chloro may be adequate for the practice of the method of the invention. Conversely, a less reactive amine, for example an aniline derivative, may require the selection of a better leaving group Y, such as triflate or bromo, by the skilled artisan.

[0464] The compound of formula (XII) can be a dipeptide, a polypeptide, a free peptide or an immobilized polypeptide, a peptide analog (for example, having a hydrogen atom or an alkyl group other than a methyl group in place of the ω-alkyl group), containing a precursor sidechain —(CH2)nX, wherein n is 1 to about 10. A compound of formula (XII), for example, a compound including the types of sidechains found on ribosomal aminoacyls, can have those sidechains in blocked form (e.g., amines substituted with Boc groups, Cbz groups, and the like) when the reaction with the reagent amine (NR'R'R'2 or R'NH2) is carried out, or, alternatively, the sidechains can all be unblocked as long as there is no sidechain that can react with the reagent amine at a higher rate than does the precursor sidechain —(CH2)nX. When selecting a sidechain blocking group as are well known in the art for use with the assorted sidechain functionalities as are found in natural aminoacyls (carboxylylates, phenols, thiols, carboxamides, and the like), the sidechain should not be protected in such a way as to activate it for nucleophilic displacement by the reagent amine. Such protecting groups are well known in the art, and a suitable blocking group can be selected without undue experimentation by the skilled artisan. However, none of the sidechains found in natural (ribosomal) aminoacyls interfere with the reaction of the invention, therefore sidechain protected is unneeded for the purpose of carrying out the reaction of the invention.

[0465] In the case where unnatural aminoacyl residues make up a peptide of formula (XII), any sidechain functionalities that are prone to nucleophilic displacement by the amine NR'R'R'2 should be suitably protected unless it is the intention that those functionalities undergo the displacement reaction. For example, if there are two hydroxyl-containing sidechain functionalities in a peptide of formula (XII), and it is only desired that one of the hydroxyl-containing sidechains should be converted to a compound of formula (XI), an amine, for instance via conversion to a sulfonate ester followed by displacement with the reagent amine, the other hydroxyl-containing sidechain should be suitably protected prior to carrying out the method of the invention. The selection of blocking groups when needed to protect sidechain functionalities is readily made by a person of ordinary skill without undue experimentation based on well-known blocking groups for the kinds of sidechain functionalities that may be encountered.

[0466] The substituents of the sidechain amino group in the compound of formula (XI) R1, R2 and R3, are independently hydrogen, alkyl, aryl, cycloalkyl, heterocyclyl, or heteroaryl, wherein one of R1, R2 or R3 may be absent, wherein any alkyl can contain 0-3 heteroatoms comprising N, O, or S, and wherein any alkyl, aryl, cycloalkyl, heterocyclyl, or heteroaryl may be unsubstituted or substituted. For example, compounds of formula (I) that may be prepared by a method of the invention include N-methyllysine, N,N-dimethyllysine, and the quaternary ammonium methosalt of N,N-dimethyllysine. Other N-alkyllysine derivatives such as N-benzyllysine, N-picolyllysine, and the like, can be also prepared. By N-alkyl or N-aryls lysine derivatives is meant herein lysine derivatives bearing the substituent on the sidechain (α) amino group.

[0467] X can be OH, O(CH2)4alkyl, O(CH2)4C8H18, NR2, an aminoacid residue or a peptide residue, or wherein X is O, NR, an aminoacid residue, or a peptide residue, respectively covalently bonded to a polymer; and Z can be H, NR2, alkyl, aryl, aralkyl, heterocyclyl, heteroaryl, heterocyclylalkyl, heteroarylalkyl, cycloalkyl, cycloalkylalkyl, an aminoacid residue, or a peptide residue, or is N, an aminoacid residue, or a peptide residue, respectively covalently bonded to a polymer; provided that when Z is NH, X is an amino acid residue or a peptide residue, or is an aminoacid residue or a peptide residue respectively covalently bonded to a polymer. For example X can be hydrogen or alkyl, Z and X cannot both be bonded to a polymer; furthermore, at least one of Z and X must comprise an aminoacid residue, a peptide residue, or an aminoacid residue or a peptide residue respectively bonded to a polymer.

[0468] The ratio of the reagent amine to the substrate precursor peptide can vary; preferably an excess of reagent amine is used. Molar ratios can vary between about 1:1 up to about 100:1 of the reagent amine to the precursor peptide. Higher ratios of reagent amine are advantageously used to suppress possible unwanted side reactions, for example, if a precursor group —(CH2)nY is present in the same molecule with another nucleophilic sidechain group, such as a thiol (cysteine), an amine (lysine), or a non-natural group of high nucleophilicity, use of a large excess of reagent amine is favored in reducing the amount of intra- or intermolecular coupling. Larger excesses of the reagent amine also serve to increase the reaction rate and to drive the reaction to completion due to the bimolecular kinetics of the nucleophilic displacement reaction.

[0469] The conversion of a compound of formula (XII) to a compound of formula (XII) or a compound of formula (XV) can be carried out in any suitable solvent. Examples are water, methanol, ethanol, or a mixture thereof. Other solvents, such
as dipolar aprotic solvents that are capable of dissolving the substrates and which may enhance the rate of nucleophilic displacement of Y by the reagent amine can also be used. Examples are DMSO, DMF, DMAc, NMP, and the like. When the peptidyl reagents have sufficient solubility in non-polar solvents, suitable nonpolar solvents such as chloroform and dichloromethane can also be used.

[0470] The reaction time and temperature can be varied depending upon the specific reaction in question to provide a reasonable yield of the desired product within a relatively short period of time. For example, the reaction can be carried out at about 20-80 °C, for a time period of about 1 hr to about 7 days. If necessary, reaction temperatures can be as high as about 100 °C or 120 °C using a solvent with a reflux temperature that exceeds the given temperature.

[0471] In solution phase reactions, the progress of the reaction towards completion is readily monitored by a person of ordinary skill in the art using techniques such as high performance liquid chromatography (HPLC), by which means the disappearance of precursor and the appearance of product are readily viewed. In solid phase reactions, for the monitoring of reaction progress it may be necessary to remove small aliquots of the polymeric material and cleave the product from them to ascertain the degree of reaction using HPLC. Other methods such as nuclear magnetic resonance (NMR) may also be used to follow the reaction progress. It is within the knowledge of a person of ordinary skill in the art to determine suitable reaction conditions for a particular conversion using standard analytical techniques that are available.

[0472] The conversion of a compound of formula (XV) to a compound of formula (XIII) via contact of the compound of formula (XV) with a compound of formula (XIV) takes place by addition of the NH₂ group of the compound of formula (XV) to the compound of formula (XIV) with elimination of the SR₂ group. The compound of formula (XV) can be a dipeptide, a polypeptide, a free peptide or an immobilized polypeptide, a peptide analog (for example, having a hydrogen atom or an alkyl group such as a methyl group in place of the c-amino group), all containing a sidechain amino group bearing at least a single hydrogen atom. This amino group bears an R group, wherein R is hydrogen, alkyl, aryl, aralkyl, cycloalkyl, heterocycles, or heteroaryl; wherein any alkyl can further contain 0-3 heteroatoms comprising N, O, or S, and wherein any alkyl, aryl, cycloalkyl, heterocyclyl, or heteroaryl may be unsubstituted or substituted.

[0473] A compound of formula (XIV), for example, which can be a compound including the types of sidechains found on ribosomal aminoacids, can have those sidechains in blocked form (e.g., amines substituted with Boc groups, Z groups, and the like) when the reaction with the compound of formula (XIV) is carried out, or, alternatively, the sidechains can all be unblocked as long as there is no sidechain that can react with a compound of formula (XIV) at a higher rate than does the precursor sidechain bearing the NH₂ group. When selecting a sidechain blocking group as are well known in the art for use with the asserted sidechain functionalities as are found in natural aminoacids (carboxylates, phenols, thiols, carboxamides, and the like), the sidechain should not be protected in such a way as to activate it for nucleophilic attack on a compound of formula (XIV). Such protecting groups are well known in the art, and a suitable blocking group can be selected without undue experimentation by the skilled artisan. For example, carboxylates and phenols can respectively be blocked as esters and ethers, such as benzyl ethers. However, none of the sidechains found in natural (ribosomal) aminoacids, except for amino groups undergo reaction with the thiouronium compound, therefore sidechain protected is generally unnecessary for the purpose of carrying out the reaction of the invention. In the case where unnatural aminoacid residues make up a compound of formula (XV), any sidechain functionalities that are prone to nucleophilic addition to the compound of formula (XIV) should be suitably protected unless it is the intention that those functionalities undergo the reaction. For example, if there are two amine-containing sidechain functionalities in a compound of formula (XV), and it is only desired that one of the amine-containing sidechains should be allowed to react with the S-substituted thiouronium compound (XIV) the other amine-containing sidechain should be suitably protected prior to carrying out the method of the invention. For example, a Boc or Cbz group can be used. The selection of blocking groups when needed to protect sidechain functionalities can readily be made by a person of ordinary skill without undue experimentation based on well-known blocking groups for the kinds of sidechain functionalities that may be encountered.

[0474] The compound of formula (XIV) is an S-substituted thiourea or a thiouronium salt thereof. Preferably the compound of formula (XIV) is a thiouronium salt, for example a hydroxide (Η) salt. The sulfur atom of the reagent bears a substituent R which can be substituted or unsubstituted alkyl or aryl. For example, R can be a methyl group. It is within the knowledge of a person of ordinary skill to modify R to confer a greater or a lesser degree of reactivity upon the thiouronium compound of formula (XIV). For example, to increase the reactivity of the compound of formula (XIV), for example when the compound of formula (XV) bears a relatively unreactive NH₂ group, such as where R is electron-withdrawing, with the compound of formula (XIV), a trifluoromethyl group could replace the methyl group as R. This kind of substitution is well-known in the art to increase the reactivity of a thiouronium by increasing the electrophilicity of the central carbon atom. Alternatively R can be an substituted or unsubstituted aryl group, for example a phenyl group. To increase the reactivity of the thiouronium reagent of formula (XIV), it would be within the knowledge of a person of ordinary skill to substitute the phenyl group with an electron-withdrawing group such as a nitro group in order to increase the reactivity of the thiouronium compound.

[0475] The compound of formula (XIV) can also be substituted on its nitrogen atoms with substituents R³ and R⁴ which are at each occurrence independently H, alkyl, cycloalkyl, heterocyclic aryl or heteroary1, wherein any alkyl, cycloalkyl, aryl, heterocyclyl or heteroaryl residue may be unsubstituted or substituted. It is understood that when one R³ group is a hydrogen atom, the tautomeric nature of guanido groups makes R³ and R⁴ interchangeable through a proton shift process.

[0476] X can be OH, O(C₁₋₄ alkyl), O(C₆₋₁₀ ary1), NR₂, an amionic residue or a peptide residue, or wherein X is O, NR, an aminoacid residue, or a peptide residue, respectively covalently bonded to a polymer; and Z can be H, NR₂, alkyl, aryl, aralkyl, heterocyclic, heteroaryl, heterocyclylalkyl, heteroarylkyl, cycloalkyl, cycloalkylalkyl, an aminoacid residue, or a peptide residue, or is NR, an aminoacid residue, or a peptide residue, respectively covalently bonded to a polymer; provided that when Z is NR, X is an amino acid residue or a peptide residue, or is an aminoacid residue or a peptide residue respectively covalently bonded to a polymer. For
example, Z can be H or alkyl, more specifically methyl. When Z is a nitrogen, X comprises at least a single amino acid residue, which may or may not be covalently bonded to a polymer.

[0477] The compound of formula (XIV) offers a major advantage over art reagents in that a greater number and variety of substituents can be disposed on the final arginine analog of formula (XIII) than was previously possible in a single step. Using a method of the invention, an arginine analog bearing a substituted guanido group can be synthesized with as many as four substituents included, one substituent each on two of the three guanido nitrogen atoms and two substituents on the third guanido nitrogen atom.

[0478] The reaction can have molar ratios of reagents between about 1:1 compound of formula (XV) and thiouronium compound of formula (IV) ranging up to about a 1:10 ratio. Use of some excess of the thiouronium compound can be used to drive the reaction to completion and to increase the rate of the reaction, as well as in reducing unwanted side reactions.

[0479] The reaction between a compound of formula (XV) and a compound of formula (XIV) can be carried out in any of a variety of solvents, including water or organic solvents. For example, the reaction can be carried out in water, acetonitrile, methanol, ethanol, tetrahydrofuran, N,N-dimethylformamide, or a mixture thereof. Other suitable solvents will be known to a person of ordinary skill.

[0480] The reaction can be carried out at any suitable temperature; for example, the reaction can be carried out within a time period of hours to days at a reaction temperature of about 20-80°C. The time period of the reaction can be as short as a few hours, for example about four hours, or can be in the order of days, for example up to about seven days. If necessary, reaction temperatures can be as high as about 100°C or 120°C using a solvent with a reflux temperature that exceeds the given temperature.

[0481] In a method of the invention for preparation of a compound of formula (XIII) from a compound of formula (XII), the two reactions described herein can be combined to provide a single process for carrying out the transformation of an amino acid residue with a sidechain —(CH₂)ₙ₋₁—Y to an amino acid residue bearing a substituted guanido group, with the provision that the intermediate amino acid residue bearing an amine sidechain must bear at least a single hydrogen atom on the sidechain amino group to allow for condensation with the thiouronium reagent. It may be advantageous under certain circumstances to carry out the two synthetic conversions sequentially, for example, to obtain a product with a modified arginine analog residue in a molecule that also contains a lysine or lysine analog residue. If one were to synthesize the precursor peptide with the residues that were to be modified arginine residues present with the —(CH₂)ₙ₋₁—Y sidechain, and the lysine residues present as protected lysine, a high degree of selectively could be obtained wherein no inadvertent conversion of lysine residues to substituted arginine residues took place.

[0482] There are numerous peptides for which use of a method of the invention could provide a target compound, including an analog, of interest. For example, the peptide neurotensin is a highly cationic peptide of medicinal interest, as is described in PCT Application Publication No. WO2006/009302, which is incorporated herein by reference. This same PCT Published Application also discusses the C-terminal region of neurotensin, amino acids number 8-13 as a structural lead for the preparation of peptide derivatives with analgesic activity. For example, the molecule wherein the N-terminal arginine of the NT(8-13) fragment is converted to a desamino alkyl derivative, i.e., wherein the N-terminal amino group is replaced by a methyl group, possesses desirable bioactivity and the preparation and biological testing of a series of analogs, such as can be prepared by a method of the current invention, is of interest in the field.

[0483] While the invention has been described and exemplified in sufficient detail for those skilled in this art to make and use it, various alternatives, modifications, and improvements will be apparent to those skilled in the art without departing from the spirit and scope of the claims.

[0484] All documents referred to herein are incorporated by reference in their entireties.

EXAMPLES

Example 1

Summary of Neurological Effects of the NT Peptides of the Invention

[0485] The N-terminal alpha methyl, alpha desamino homolysyl and orinthyl analogues of NT(8-13) prepared according to the invention (see the foregoing general discussion and the Examples) were synthesized and screened for activity in numerous behavioral assays predictive of antipsychotic potential. These peptides induced hypothermia in a dose-dependent fashion after oral administration. In addition, oral administration of the peptides significantly reduced d-amphetamine induced hyperlocomotion, a measure of the therapeutic efficacy of current or potential APDs. The low dose of peptide (10 mg/kg) that elicits a significant response after oral administration in these assays is significant. The peptides also demonstrate an ability to maintain efficacy after repeated administration. In fact they demonstrate an ability to increase maximal hypothermic response over time, implying that repeated administration may actually improve their CNS activity. Thus, the NT peptides of the invention are shown to have biological activity like that of the known naturally occurring peptide NT and are more selective. Details of these effects are as follows.

[0486] Hypothermia as a Preliminary Screen of CNS Activity. NT induces hypothermia when directly administered into the CNS. As a result, induced hypothermia can be used to determine the ability of NT(8-13) peptides of the invention to cross the BBB after peripheral administration and indirectly to determine their in vivo CNS activity. The hypothermic effect of NT can be attributed to its actions at NTR₁, the NTR most often implicated in the pathophysiology of schizophrenia. An NT(8-13) peptide that induced hypothermia after IP injection is thus shown to be an antipsychotic agent. A significant hypothermic effect would demonstrate that the peptide showed marked improvements in blood stability and membrane crossing.

[0487] IP injection is the standard route of administration to determine the extent of BBB crossing of neurotensin analogues. The methods and protocols are provided in the Examples section. IV administration results in a dose that is completely available to the systemic circulation. By contrast, an IP injection is a more rigorous test of stability because the peptide is exposed to first pass metabolism in the liver.

[0488] The hypothermic effects of peptides 28-30, after a 5 mg/kg IP injection, are given in Table 4. Each peptide exhibited a significant effect over a 5 hr time course. The hypother-
Ermic results for these three peptides demonstrate that the substitution of an alpha alkyl group in place of the N-terminal amine group (i.e., the α-methyl group) does not abolish the in vivo activity of the NT(8-13) peptide. For use as an antischizophrenic pharmaceutical, the ability of these peptides to elicit CNS activity after oral administration was evaluated.

<table>
<thead>
<tr>
<th>TABLE 4</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Amino acid sequence of NT(8-13) analogues</strong></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Peptide</th>
<th>N-terminus</th>
<th>6</th>
<th>9</th>
<th>10</th>
<th>11</th>
<th>12</th>
<th>13</th>
<th>C-terminus</th>
</tr>
</thead>
<tbody>
<tr>
<td>KH28</td>
<td>CH3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>L-Hlys L-Arg L-Pro L-Tyr L-Leu L-Leu COOH</td>
</tr>
<tr>
<td>(ABS201)</td>
<td>CH3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>L-Hlys L-Arg L-Pro L-Tyr L-Leu L-Leu COOH</td>
</tr>
<tr>
<td>KH29</td>
<td>CH3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>L-Arg L-Pro L-Tyr L-Leu L-Leu COOH</td>
</tr>
<tr>
<td>KH30</td>
<td>CH3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>L-Arg L-Pro L-Tyr L-Leu L-Leu COOH</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>TABLE 5</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Hypothermic response to IP administration of NT(8-13) and NT(8-13) analogues</strong></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Peptide</th>
<th>$t_{max}$ (min)$^b$</th>
<th>$\Delta$ in BT ($^{\circ}$ C)$^c$</th>
</tr>
</thead>
<tbody>
<tr>
<td>NT (8-13)</td>
<td>90</td>
<td>-0.45 ± 0.17$^e$</td>
</tr>
<tr>
<td>ABS201</td>
<td>150</td>
<td>-2.51 ± 0.17</td>
</tr>
<tr>
<td>KH29</td>
<td>150</td>
<td>-3.75 ± 0.24</td>
</tr>
<tr>
<td>KH30</td>
<td>300</td>
<td>-3.84 ± 0.20</td>
</tr>
</tbody>
</table>

$^a$IP dose was 5 mg/kg for all peptides.
$^b$t$_{max}$ (min) = Time to maximal temperature decrease.
$^c$Δ in BT ($^{\circ}$ C) = Decrease in body temperature measured at $t_{max}$
$^d$N = 5 for all peptides.

Oral Administration. A goal in the development of NT(8-13) peptides as antischizophrenic pharmaceuticals is to determine their ability to exhibit CNS activity after oral administration. The known NT peptides, NT69L and the Eisai hexapeptide, fail in this respect to elicit central activity when given orally. Accordingly, the N-terminal methyl peptides 28-30 were tested for their ability to induce hypothermia after oral administration.

ABS201, an example of a peptide of the invention, demonstrated maximal hypothermic responses greater than 2$^{\circ}$C. (Table 6) and its maximal hypothermic effect was equal to its hypothermic effect after IP dosing (Fig. 9), resulting in an approximate oral bioavailability of 25%. While peptides 29 and 30 also were orally active, their ratio of oral activity to IP activity was not as balanced as that of ABS201. The oral activity of ABS201 was an important factor to support it as a lead NT(8-13) analogue for further evaluation of antipsychotic potential.

<table>
<thead>
<tr>
<th>TABLE 6</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Hypothermic response to chronic IP administration of ABS201</strong></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Peptide</th>
<th>$t_{max}$ (min)$^b$</th>
<th>$\Delta$ in BT ($^{\circ}$ C)$^c$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Saline$^d$</td>
<td>180</td>
<td>0.70 ± 0.20</td>
</tr>
<tr>
<td>Day 1</td>
<td>120</td>
<td>2.72 ± 0.24</td>
</tr>
<tr>
<td>Day 2</td>
<td>90</td>
<td>2.85 ± 0.26</td>
</tr>
</tbody>
</table>

$^a$IP dose was 5 mg/kg for all days.
$^b$t$_{max}$ (min) = Time to maximal temperature decrease.
$^c$Δ in BT ($^{\circ}$ C) = Decrease in body temperature measured at $t_{max}$
$^d$N = 5 for all days.

Comparison of the maximal hypothermic effects of peptide ABS201 after IP administration.

<table>
<thead>
<tr>
<th>Dose</th>
<th>Peptide</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.1 mg/kg</td>
<td>-1.14 ± 0.21</td>
</tr>
<tr>
<td>0.5 mg/kg</td>
<td>-1.92 ± 0.12</td>
</tr>
<tr>
<td>1.0 mg/kg</td>
<td>-2.63 ± 0.21</td>
</tr>
<tr>
<td>5.0 mg/kg</td>
<td>-3.61 ± 0.22</td>
</tr>
</tbody>
</table>

$^a$Change in body temperature ($^{\circ}$ C) is the maximal decrease recorded for each individual dose.

Schizophrenia Investigation. The blockade of locomotion caused by d-amphetamine, a "DA agonist," has become the standard measure of therapeutic efficacy of current or potential drug candidates for treatment of schizophrenia. This model operates on the assumption that the direct stimulation of DA receptors within the mesolimbic DA system is responsible for the locomotor response.

Catalepsy, commonly defined as a state of tonic immobility in rodents, is regarded as analogous to (extrapyramidal side effects) EPSEs in humans. Consequently, catalepsy is a side effect to be avoided in a successful drug candidate. Concurrently, the degree to which a drug candidate causes catalepsy in rats may also be used as a predictor for the probable occurrence of EPSEs associated with that particular candidate.
Hypothermic Analysis. To examine the antipsychotic properties of ABS201, a dose-response curve for hypothermic induction after IP administration was generated. In addition, the hypothermic effects elicited by oral administration of ABS201 (10 mg/kg-30 mg/kg) were determined. The ability of ABS201 to reduce d-amphetamine induced hyperlocomotion after both IP and oral administration also was measured. To assess the effects on CNS activity caused by prolonged treatment of ABS201, hypothermia and attenuation of d-amphetamine induced hyperlocomotion were measured after repeated daily dosing. Finally, the bar test was utilized to measure catalepsy as a predictor of EPSs in humans.

The dose-response curve for ABS201 after IP injection over a concentration range of 0.1-10 mg/kg (FIG. 11) gave some conflicting results. First, the maximal effect elicited at 5 mg/kg (-3.61±0.22°C at 150 min PI) was a full degree greater than the maximal effect seen after the preliminary screen (-2.51±0.17°C at 150 min PI). This discrepancy is most likely due to environmental factors (air temperature, rectal probes, rat size, etc.) that can affect the rats’ response. Most importantly, ABS201 continued to elicit a significant CNS effect irrespective of these differences. The ED50 value for ABS201, 0.943 mg/kg, compares favorably with other NT(8-13) analogues with CNS activity (41, 60).

ABS201 also induced hypothermia in a dose-dependent fashion after oral administration (FIG. 12). A significant hypothermic effect was demonstrated at 10 mg/kg, the lowest dose tested (-1.02±0.10°C at 150 min PI). The generation of an ED50 value for the oral administration of ABS201 was impractical due to the inordinate amount of peptide necessary to produce a complete dose-response curve. Previous NT(8-13) analogues that have been under development as antipsychotic compounds have contained a Trp1 substitution. Evidence from the studies presented herein supports the theory that this modification abolishes the oral activity of a NT analogue. Further studies are necessary to determine what specific role Trp1 plays in the oral bioavailability of NT(8-13) analogues.

The blockade of locomotion caused by d-amphetamine, a “DA agonist”, has become the standard measure of therapeutically efficacious or potential antipsychotic drug candidates, and NT(8-13) analogues currently under investigation as candidates have demonstrated the ability to decrease d-amphetamine induced hyperactivity in a dose-dependent fashion. Sound- and light-attenuated locomotor cages are used to measure the ability of potential candidates to decrease d-amphetamine-induced hyperactivity.

The effects of ABS201 on d-amphetamine induced hyperactivity at varying doses were also examined. ABS201 significantly reduced hyperlocomotion for all doses tested (doses of 3 mg/kg and 10 mg/kg not shown). Another hallmark of current APDs is the ability to reduce spontaneous locomotor activity. All ABS201 dose groups responded significantly lower than saline during the drug phase, indicating the ability of ABS201 to reduce spontaneous activity.

The ability to attenuate d-amphetamine induced hyperlocomotion after oral administration is also demonstrated by ABS201. During the drug phase, only the 10 and 30 mg/kg doses reduced spontaneous locomotor activity. The lack of significance seen with the 20 mg/kg dose is most likely an anomaly resulting from slight variation in response for this group of rats. However, no main effect of DOSE was detected during the baseline phase, indicating that there is not a significant difference in baseline activity across the different dosing groups.

ABS201 maintained a significant CNS effect after repeated daily dosing (Table 6) and over the 5-day period the absolute hypothermic response increased. A comparison of the induced hypothermia of ABS201 on days 1 and 5 was made. On day 5, the maximal hypothermic response was achieved faster (90 min) compared to day 1 (120 min). In contrast to day 1, on day 5 the maximal hypothermic effect was not maintained for an extended period, implying that while repeated dosing does not decrease the maximal effect, it may reduce the duration of the hypothermic effect. Repeated daily dosing had no effect on the ability of ABS201 to attenuate d-amphetamine induced hyperlocomotion. Both the acute and chronic dosing groups produced a reduction in hyperactivity that was significant for almost two hours after amphetamine administration. Of note, chronic administration of ABS201 did abolish its inhibitory effect on spontaneous locomotor activity.

Catalepsy Analysis. In laboratory tests, catalepsy is characterized by the inability of an animal to correct its position after placement in an unnatural posture. Catalepsy tests can be greatly influenced by a number of variables. These include stress-induced inhibition of catalepsy caused by a new environment and the contribution of learned “pseudo-catalepsy” that can result upon repeated measures with the same animal. To circumvent these potential confounding factors, tests are performed on an animal only once in a quiet, controlled environment.

Neither ABS201 (5 mg/kg) nor saline caused catalepsy after peripheral administration. Haloperidol, a typical antipsychotic drug known to produce a fully cataleptic response in rats, induced catalepsy that lasted for greater than 30 sec. These results demonstrate that ABS201 does not induce catalepsy after peripheral administration, a hallmark of current clinically effective candidates.

Bioavailability Study with CACO-2 Cells. Caco-2 cells, derived from a human colorectal carcinoma, spontaneously differentiate into polarized cells that exhibit well-developed microvilli and brush-border enzymes. These features make the cells an excellent model of the human small intestine. A strong correlation between uptake of a compound in the Caco-2 cell model and oral bioavailability of the compound has been identified.

ABS201 is stable in rat serum for greater than 24 hours, however, its stability in cells has not been determined. Consequently, determination of the ability of intact peptide to enter the Caco-2 cells in the uptake experiments will show oral bioavailability and cellular stability. Reverse phase HPLC is an ideal method to analyze the solubilized cell components for ABS201 and ABS201 degradation products. This analysis will show oral availability and cellular stability. Fractions can be collected at determined intervals and counted for radioactivity via LSC. By establishing the ABS201 elution time via a standard gradient, direct comparisons can be made to the contents of Caco-2 cells after uptake experiments.

To verify that intact peptide is entering the Caco-2 cells and in a preliminary attempt to assess the stability of the peptides in cell culture, an RP-HPLC assay to analyze ABS201 after cellular uptake can be carried out. After a 2 min incubation, intact ABS201 likely can be identified within the cells using HPLC techniques. These studies will demonstrate that
ABS201 can be extensively taken up by the Caco-2 cells thus showing its oral bioavailability.

Example 2

Compound Synthesis

The following examples and protocols are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how the compounds claimed herein are made and evaluated, and are intended to be purely exemplary of the invention and are not intended to limit the scope of what the inventors regard as their invention. Efforts have been made to ensure accuracy with respect to numbers (e.g., amounts, temperature, etc.) but some errors and deviations should be accounted for. Unless indicated otherwise, parts are by weight, temperature is in °C and is at room temperature, and pressure is at or near atmospheric.

Starting Materials. Solvents are from Fisher Scientific (Pittsburgh, Pa.) and reagents from Aldrich (Milwaukee, Wis.) unless otherwise noted.

Abbreviations. Trisyl-N₃, 2,4,6-trisopropynylbenzene-n-sulfonyl azide; Et₃N, triethylamine; t-BuOCOCl, trimethylchloroacetate; n-BuLi, n-butyllithium; H₂, hydrogen gas; Pd–C, palladium on activated carbon; Xps, (2S)-4-aminobenzyl-2-oxazolidinone; KHMDS, potassium bis(trimethylsilyl)amide; CH₃I, methyl iodide; H₂O₂, hydrogen peroxide; LiOH, lithium hydroxide; THF, tetrahydrofuran; CH₂Cl₂, dichloromethane; MgSO₄, magnesium sulfate; Hex, hexane; EtOAc, ethyl acetate; NaHCO₃, sodium bicarbonate; HCl, hydrochloric acid; N₂, nitrogen; H₂O, distilled water.

Protocol was as described above for 24a. Additional silica gel purification eluting with 100% CHCl₃ gave pure 24b in 10% yield. The H NMR (400 MHz, CDCl₃) δ 7.36-7.19 (m, 5H), 4.72-4.65 (m, 1H), 4.25-4.16 (d, J=4.2 Hz, 2H), 3.77-3.67 (m, 1H), 3.46-3.36 (t, J=7.6 Hz, 2H), 3.29-3.22 (dd, J=4.0, 14.0 Hz, 1H), 2.82-2.74 (dd, J=9.0, 14.0 Hz, 1H), 1.92-1.74 (m, 3H), 1.50-1.42 (m, 3H), 1.25-1.21 (d, J=7.2 Hz, 3H). The C NMR (100 MHz, CDCl₃) δ 176.9, 152.3, 135.3, 129.6, 129.1, 66.4, 55.6, 38.1, 37.8, 34.1, 32.8, 32.5, 26.1, 18.7.

Example 3

Alpha methyl, Alpha desamino, omega N-substituted homolysyl and ornithyl (8) neurotensin (8-13)

Alpha methyl, Alpha desamino omega N-substituted homolysyl and ornithyl (8) neurotensin (8-13) were synthesized (Fig. 7). The α-methyl) bromo acids, 27a and c, were coupled to the resin-bound peptide as outlined in the general section. The solid state coupling was conducted as follows.

Resin bound N alpha Fmoc leucine was swelled in DMF prior to Fmoc cleavage with piperidine (20% in DMF). The piperidine solution was removed with vacuum filtration and the resin-bound amino acid washed with DMS and methyl
ethylene chloride (5x each). Amino acids (4 eq) were activated in DMF with HOBT (4 eq) PyBOP (4 eq) and DIPEA (10 eq) and added directly to the peptide reaction vessel. Amino acids were coupled for 6 hours, the resins was washed with DMF and methylene chloride and monitored with a Kaiser test for the presence of free amines. Residues were recoupled when necessary. This procedure was repeated with subsequent amino acids to give the penultimate peptide sequence (pentamer).

[0516] Aliquots of the resin-bound pentamer were then coupled with the appropriate omega bromo carboxylic acid as described above to give the N-omega bromo acyl pentamers. The N-omega acyl pentamers were then reacted with ammonia, dimethyl amine or trimethyl amine as described in these Examples to produce the desired peptides of the invention. Acid catalyzed deprotection of the side chain protecting groups was performed with a TFA solution containing appropriate scavengers.

[0517] RP-HPLC purification using a linear gradient of 15% to 75% B over 55 min at a constant flow rate of 4 mL/min afforded pure omega bromo peptides 53 and 54. These bromo peptides were reacted at 40° C. for 12 hr with 150 eq of ammonium hydroxide (29% in H2O), dimethylaniline (40% in H2O), or trimethylamine (40% in H2O) in ethanol (EtOH). Solvents were removed in vacuo and crude peptides were taken up in mobile phase and purified with a linear gradient of 2% to 50% B over 65 min at a constant flow rate of 4 mL/min.

[0518] Peptides were characterized and assessed for purity via MALDI-TOFMS on a Voyager DE-STR System 4117 mass spectrometer (Applied Biosystems, Foster City, Calif.). Peptides were used at greater than 95% purity in vivo.

Example 4

Identification of Antipsychotic NT Peptide Analogs

[0519] Structures of Compounds. The peptide analogs evaluated contain one non-natural amino acid (Scheme 1) or desaminoacid (Scheme 2).

Scheme 1. Non-natural amino acids used in generating NT analogs.

Scheme 2. Desamino amino acids used in generating NT analogs.
Benchmarking Compounds. From an initial screen of about 50 compounds, those listed in Table 8 were selected for further testing and development as antipsychotic compounds. These compounds are based on the neurtensin fragment NT(8-13). These compounds possess useful characteristics as each: bound as competitive agonists in vitro to NTR-1, the brain neurtensin receptor involved in schizophrenia, demonstrated central activity when injected IP in rat using hypothermia as a surrogate for central activity, which also occurs through NTR-1 binding and elicited the appropriate activities in rat behavioral models of schizophrenia.

![Image of neurtensin structure](image)

### Table 8-continued

<table>
<thead>
<tr>
<th>Peptide</th>
<th>Structure</th>
</tr>
</thead>
<tbody>
<tr>
<td>NT</td>
<td>NH₂-Arg-Arg-Pro-Tyr-Ile-Leu-COOH (8-13)</td>
</tr>
<tr>
<td>ABS13</td>
<td>H₁₇-L-homolysine-Arg-Pro-Tyr-tertLeu-Leu-COOH</td>
</tr>
<tr>
<td>ABS41</td>
<td>H₁₇-13-Arg-Pro-Tyr-tertLeu-Leu-COOH</td>
</tr>
<tr>
<td>ABS44</td>
<td>H₁₇-7-Arg-Pro-Tyr-tertLeu-Leu-COOH</td>
</tr>
<tr>
<td>ABS46</td>
<td>H₁₇-9-Arg-Pro-Tyr-tertLeu-Leu-COOH</td>
</tr>
<tr>
<td>ABS201</td>
<td>43-Arg-Pro-Tyr-tertLeu-Leu-COOH</td>
</tr>
</tbody>
</table>

### Example 5

**Protocols for Neurotesting of ABS201**

**General Animal Protocols.** Male Sprague Dawley Rats (250-350 g) or Brattleboro rats (270-310 g) were obtained from Harlan (Indianapolis, Ind.) and housed in an AAALAC-approved colony room maintained at a constant temperature and humidity. Lighting was controlled on a 12 hr light:dark cycle with lights on at 0700 hr. Animals were housed two per cage and fed laboratory chow and water ad libitum. All experiments were performed during the light cycle.

**Animal Restraint.** Rats were restrained in Plas-Labs® plastic cages fitted with wooden dowels to restrict movement. Rectal temperature probes (Physitemp®, RET-2, Clifton, N.J.), lubricated with mineral oil, were inserted into the rectum of each animal. Probes were connected to a microprobe thermometer (Physitemp®, BAI-12) in conjunction...
with a thermocouple selector (Physitemp®, SWT-5). Rats were allowed to acclimate to the cages for 1 hr prior to IP injection.

Peptide Preparation. Each peptide tested was dissolved in 0.9% NaCl to the appropriate final concentration for IP injection (10, 30, or 100 mg NT8-13 or equimolar equivalent of analog in 1 ml 0.9% NaCl). For oral administration, the test article was administered directly into the animals’ stomach by gavage. Controls involved the administration of 0.9% NaCl. Thus, peptides were dissolved in saline, and following the equilibration period, rats were given an IP injection of peptide (5 mg/kg) or saline (1 ml/kg).

Initial temperature values were the average temperatures of the rats immediately before and after the injection. Subsequent measurements were taken every 30 min for 5 hr. Statistical analysis was performed using a one-way analysis of variance (ANOVA) followed by Tukey’s post hoc test for multiple comparisons using GraphPad Prism® to measure significance. Results were considered significant for p<0.05.

Hypothermia. Hypothermia induction is an easy and direct measure of neurotensin derivatives entering the brain and binding to NTR-1, the receptor involved in psychosis. For these experiments, animals were housed in individual Plexiglass cages without bedding at 25°C. For each rat, a thermistor probe was placed 4.5 cm into the rectum; baseline temperatures were recorded for 1 hr to allow habituation to the cages prior to injection of compound.

Dose-response curves for hypothermic induction. All animal restraint and hypothermia protocols were as described above. Variable slope dose-response curve and ED₅₀ value was generated using GraphPad Prism®.

D- Amphetamine induced hyperlocomotion. Experimentally naive Sprague-Dawley rats were housed as described above. Rats were handled for three days prior to testing to minimize experimenter induced hyperlocomotion on test day. For experiments, sound- and light-attenuated automated photocell beam activity chambers (AccuScan Instruments, Inc., Columbus, Ohio) were used to measure locomotion. Cages were connected to a VersaMax Analyzer (AccuScan) in conjunction with an IBM computer using VersaMax 1.80-0146 software (AccuScan) to record vertical and horizontal activity. Total activity values recorded were the sum of vertical and horizontal activity.

Each animal’s activity was measured by how many times per minute the animal “trips” a photobeam in a computer-controlled photocell activity chamber (San Diego Instruments, San Diego, Calif.). The chambers were arranged in a four-level rack in a quiet room with ambient light and temperature approximately the same as in the separate housing room. The experiment was done over 6 days, with 1 run each day in mid-afternoon. On each day, 6 naïve rats were assigned to each treatment condition. Doses were assigned so that each was assessed in 12 chambers that were distributed over the apparatus, which controls for variations in lighting, etc. of the different chambers.

Rats were placed in the activity chambers for 30 minutes to habituate and establish baseline activity levels. Then, rats were removed and given an IP or oral dose of peptide (N=7) or saline (N=8) and returned to the chamber to establish the peptide’s effect on spontaneous activity levels. About 30 minutes after administration, rats were removed and given an IP injection of d-amphetamine (1 mg/kg) or vehicle and returned to the chamber for a further 1 hr to assess the effect of the peptide on induced hyperlocomotion.

Chronic Testing Protocols. For chronic hypothermia testing, rats were given an IP injection of ABS201 (5 mg/kg) or saline once daily for five consecutive days. Induced hypothermia was monitored and tested for significance as described above. To assess the ability of ABS201 to decrease d-amphetamine induced hyperactivity after repeated administration, rats were divided into three dosing groups: chronic, acute, and control (N=7 for all groups). On days 1-4, the chronic group received an IP injection of ABS201 (5 mg/kg) while the acute and control groups received saline. On test day, day five, chronic and acute animals received ABS201 (5 mg/kg) while control animals received saline. The test protocol for day five was as described above.

Catalepsy Assessment. ABS201 (5 mg/kg) was dissolved in saline (1 ml/kg). Rats were given an IP injection of peptide (5 mg/kg), saline, or haloperidol (1 mg/kg). After 3 hours, catalepsy was measured using the horizontal bar test. Briefly, the front paws were placed directly on a horizontal bar 5 mm in diameter placed 7.5 cm above the cage floor. The rat was held in this position for 3 sec and then released. The time from release until the paws return to the cage floor was measured and recorded. A cut-off time of 30 sec was observed; this indicated a fully cataleptic animal as cataleptic animals are frozen to the bar. Normal animals withdraw virtually immediately. Measures were repeated every 30 min for 4 hr. Data are mean±SEM (p<0.01).

Example 6

Results of Neurotesting of ABS201

Hypothermic induction by ABS201 through IP and PO dosing. ABS201 has an ED₅₀ of about 1 mg/kg when administered IP, and 10 mg/kg administered orally (hypothermia data not shown, see amphetamine-induced hyperlocomotion results below). As seen in FIG. 9, ABS201 actually induces a quicker hypothermic effect when dosed orally versus IP at 2x the approximate ED₅₀ for each compound.

Dose-response curves. The dose-response curve for ABS201 after IP injection over a concentration range of 0.1-10.0 mg/kg is shown in FIG. 11. The calculated ED₅₀ value is 0.943 mg/kg. The hypothermic response to the oral administration of ABS201 over a concentration range of 10.0-30.0 mg/kg is shown in FIG. 12.

Attenuation of d-amphetamine induced hyperlocomotion after IP and oral dosing. FIG. 13 illustrates that ABS201 delivered IP inhibits amphetamine-induced hyperlocomotion with an approximate ED₅₀ of about 10 mg/kg. FIG. 14 illustrates that ABS201 delivered by oral dosing induced hyperlocomotion with an approximate ED₅₀ of about 10 mg/kg. Larger doses were not attempted in the PO administration experiments due to lack of sufficient material; however, the IP data clearly illustrate that the hyperlocomotion effect can be fully reversed.

Separate two-factor ANOVAs for Dose × TIME were performed for each different time-phase of the assay. Phases for the IP dosing and oral dosing experiments consisted of habituation (time points 10-60), drug (time points 70-120), and amphetamine (time points 130-240).

For IP dosing experiments, during the habituation phase, there was a main effect of TIME [F(5,185)=264.335 (p<0.001)], indicating a gradual decrease in activity levels over time regardless of dose. Tukey’s post-hoc tests, collapsed over dose, indicated activity levels for time points 10-30 were significantly higher than time points 40-60 (p<0.
001). These results are attributed to the initial spontaneous exploratory activity associated with the novel environment. During the drug phase, there was a main effect of TIME [F(5, 185) = 12.336 (p < 0.001)] and a main effect of DOSE [F(5, 37) = 11.775 (p < 0.001)]. The main effect of TIME resulted from a decrease in activity for all doses relative to the first time point (70 min). Tukey’s post-hoc tests, collapsed over time, indicated that all doses responded significantly different from saline (p < 0.001). During the amphetamine phase, there was DOSE × TIME interaction [F(5, 407) = 4.474 (p < 0.001)]. Tukey’s post-hoc tests revealed that all ABS201 dose groups demonstrated reduced locomotor activity for time points 130-200 as compared to saline (p < 0.05).

[0537] For oral dosing experiments, during the habituation phase, there was a main effect of TIME [F(5, 120) = 201.797 (p < 0.001)], indicating a gradual decrease in activity levels over time, regardless of dose. Tukey’s post-hoc tests, collapsed over dose, indicated activity levels significantly lower at each time point. During the drug phase, there was DOSE × TIME interaction [F(15, 120) = 11.584 (p < 0.037)]. Tukey’s post-hoc tests, collapsed over time, indicated that only the 10 mg/kg and 30 mg/kg dose groups responded significantly different from saline (p < 0.01). During the amphetamine phase, there was DOSE × TIME interaction [F(11, 264) = 35.616 (p < 0.001)]. Tukey’s post-hoc tests revealed that all dose groups demonstrated reduced locomotor activity for time points 140-180 as compared to saline (p < 0.05). In addition, the 20 mg/kg and 30 mg/kg dose groups responded significantly lower than the saline group at time points 190-200.

[0538] Effects of chronic ABS201 administration of hypothermic induction. ABS201 maintained a significant CNS effect after repeated daily dosing (Table 6) and over the 5-day period the absolute hypothermic response increased.

[0539] Effects of repeated ABS201 dosing on d-amphetamine induced hyperactivity. Separate two-factor ANOVAs for GROUP × TIME were performed for each different phase of the experiment. Phases were consistent with those described above. During the habituation phase, there was a main effect of TIME [F(5, 90) = 146.164 (p < 0.001)], indicating that there was a gradual decrease in activity levels over time, regardless of group. Tukey’s post-hoc tests, collapsed over dose, indicated activity levels for time points 10-20 were significantly higher than time points 30-60 (p < 0.001) for each dose level. These results are attributed to the habituation of the rats to a novel environment over time. During the drug phase, there was a main effect of TIME [F(5, 90) = 13.512 (p < 0.001)] and a main effect of GROUP [F(2, 18) = 4.37 (p < 0.028)]. The main effect of TIME resulted from a decrease in activity for all doses relative to the first time point (70 min). Tukey’s post-hoc tests, collapsed over time, indicated that only the 10 mg/kg dose group responded significantly different from saline during the drug phase (p < 0.05). During the amphetamine phase, there was GROUP × TIME interaction [F(22,198) = 4.069 (p < 0.001)]. Tukey’s post-hoc tests revealed that both the acute and chronic groups demonstrated reduced locomotor activity for time points 140-220 as compared to saline (p < 0.05).

[0540] Reversal of Prepulse Inhibition. Prepulse inhibition (PPI) of acoustic startle is decreased in unmedicated schizophrenia patients and similar deficits can be induced in rats through pharmacological, environmental, or neuroanatomical manipulations. Recently, it was reported that Brattleboro (BB) rats, a Long Evans (LE) strain with a single gene mutation, have inherent deficits in PPI homologous to those observed in schizophrenia patients (see Feifel et al. Neuropsychopharmacology 29:731 (2004)). Atypical-type antipsychotics are active in this model. As seen in FIG. 15, orally dosed ABS201 reverses PPI in Brattleboro rats.

[0541] Cataplectic assessment. Neither ABS201 (5 mg/kg) nor saline caused cataplexy after peripheral administration (N=5). Haloperidol, a typical APD known to produce a fully cataplectic response in rats, induced cataplexy that lasted for greater than 30 sec. See FIG. 16.

[0542] Dose Tolerance. The animals were I.P. dosed with ABS201 for 5 days straight at 5x the ED50, using both induction of hypothermia and inhibition of amphetamine-induced locomotion as the monitoring methods using protocols described above. Dose tolerance is not observed with either compound (see FIGS. 17 and 18).

[0543] Summary of the Behavioral Effects of ABS201. The “gold standard” animal model for evaluating a molecule with antipsychotic potential is inhibition of amphetamine-induced hyperlocomotion. ABS201 is active in a dose-dependent fashion whether I.P. or orally injected (FIGS. 13 and 14). The action of ABS201 is apparent following both IV and PO administration; dose-dependent; and long acting (observable 1 hr post administration and apparent for at least one additional hour).

[0544] The effects of ABS201 and haloperidol on catalepsy was examined in rats. Rats (N=5) were given an IP injection of ABS201 (5 mg/kg) or haloperidol (1 mg/kg). After 2 h, catalepsy was measured using the horizontal bar test. Data are mean ± SEM (p < 0.01). ABS201 does not induce a cataleptic state in rats (FIG. 16), is antinociceptive, and tolerance to multiple dosings does not occur either with monitoring hypothermia or inhibition of amphetamine-induced hyperlocomotion (FIGS. 17 and 18). Thus, ABS201 reliably induces hypothermia in rodents following both IV and PO administration. The action of ABS201 is dose-dependent; and long acting, being observable for a period of 3-4 hours following administration. The doses producing hypothermia are similar, if not identical, to those which reverse d-amphetamine responses.

[0545] The oral and intravenous dosing of 3 male and 3 female rats each receiving 50 mg/kg (IV) or 250 mg/kg (PO) of ABS201 HCl administered in neutral physiological saline was observed. During the 2 and 24 hour post-dosing clinical observation periods, measurements of core body temperatures were taken.

[0546] The following was observed during the period immediately during and following intravenous administration of ABS201; during the dosing period (slow push; >1 min<2 min via the tail vein) the animals receiving ABS201 HCl became noticeably sedate in the body restraining cages; upon removal from the restraining cages, animals were obviously sedated, lacked spontaneous menthathop locomotor activity, assumed a curled position upon handling, and exhibited a greatly impaired, or loss of, the righting reflex; notwithstanding, ptosis was absent; there was no evidence of flaccid paralysis although muscle tone was substantially reduced; pupil reflex was present; the hind limb pinch response was impaired or absent; there was no evidence of parasympathetic responses, e.g., spontaneous urination, defecation, salivation and lacrimation were absent; there was no evidence of acute sympathetic responses, e.g., piloerection; there was no evidence of seizures, either tonic or clonic. The acute effects were short lived with the righting reflex returning by the end of the complete dosing period (approximately 30 min). At the two hour post-dosing observation period, all animals
appeared grossly normal although marked hypothermia was present. At the 24 hour post-dosing observation period, all animals appeared grossly normal; hypothermia was absent. Animals administered ABS201 HCl orally appeared grossly normal at all time points. Thus, the evidence indicates that the acute behavioral appearance and response(s) of animals following the intravenous administration of ABS201 HCl (50 mg/kg) is directly attributable to a rapid and marked central nervous system effect.

Example 7

Oral Bioavailability Studies of ABS201

**[0547]** Synthesis of Fmoc-Proline-OH* for Generation of Radioactive ABS201 (FIG. 19). L-Proline (20.7 mg, 0.18 mmol) (Advanced Chemtech) was dissolved in 450 µL of a 10% Na2CO3 solution to which 5 mL of EtOH-H2O (2:98) containing 250 µCi of L-[U-14C] egrada (Moravek, Brea, Calif.) was added. Fmoc-N-hydroxyxysuccinimide (Fmoc-Osu) (100 mg, 1.5 eq) in 3 mL dimethoxyethane (DME) was added dropwise to the stirring amino acid solution. The reaction was allowed to stir for 12 hr at room temperature and the DME was removed in vacuo. The remaining aqueous solution was diluted with 10 mL H2O and extracted with saturated N-butanol (4×10 mL). The butanol extracts were combined and concentrated to give a pale oil. Residual Fmoc-Osu was removed on silica gel eluting with MeOH:CH2Cl2 (50:50). Crude Fmoc-Proline-OH* was used without further purification in peptide synthesis.

**[0548]** Study of ABS201 Oral Bioavailability Using Caco-2 Cell Model. Caco-2 cells, derived from a human colorectal carcinoma, spontaneously differentiate into polarized cells that exhibit well-developed microvilli and brush-border enzymes (78). These features make the cells an excellent model of the human small intestine. A strong correlation between uptake in the Caco-2 cell model and oral bioavailability has been identified (79). Studies that focused on the transport of peptides across Caco-2 cells have identified solute-solvent hydrogen bonds as a major determining factor in the permeability of the peptide. The non-natural amino acid technology is designed to reduce solute-solvent interactions, in particular, water solvation that occurs through hydrogen bonding, hence the current modifications should confer enhanced intestinal absorption in Caco-2 cells. The studies described below are designed to evaluate the potential oral bioavailability of the NT(8-13) analogues and the mechanisms of transport responsible for their uptake.

**[0549]** ABS201 is a lead compound for the development of NT(8-13) analogues as novel APDs. ABS201 can therefore function as a prototype for evaluating the cellular uptake of the NT(8-13) analogues. Liquid scintillation counting (LSC) is the preferred method of analysis for these assays, as extraction of the peptide from the cell monolayer is not required and dissolved cell components can be directly analyzed without an extraction protocol that can be ineffectual, resulting in inconsistent analysis. L-[U-14C] egrada was used as the radiolabel for these studies. Proline is easily protected at the α-amine for peptide synthesis with the base-labile Fmoc moiety. In addition, Pro[15] has not been identified as a major site of cleavage of NT(8-13). NT(8-13) analogues that show antagonistic potential have not included Pro[15] modifications.

**[0550]** To examine the occurrence and mechanism of cellular uptake of the NT(8-13) analogues, Caco-2 cells, a well-established model of the intestinal epithelium can be utilized. These studies were designed to provide insight into the potential for oral activity of the peptide analogues. As described above, the NT(8-13) analogues elicit CNS activity after oral administration. They are the first analogues of NT(8-13) to exhibit oral activity, and these preliminary studies should provide information that aids in the development of future peptide analogues with enhanced oral activity.

**[0551]** The concentration of ABS201 used for these uptake studies, 200 µM, can be chosen for two distinct reasons. The concentration of a 20 mg/kg dose of peptide, delivered in saline (1 mL/kg), is 24 mM. As gavage dosing insures direct administration into the stomach, a concentration only slightly below 24 mM should be seen by the small intestine. Therefore, the concentration added to the Caco-2 cells is well below that theoretically seen in vivo. In addition, the standard circulating blood volume in the rat is 64 mL/kg (82). After gavage dosing, the concentration of a 20 mg/kg dose of peptide circulating throughout the entire rat is 377 µM. For these reasons, 200 µM is determined to be a physiologically relevant concentration to study ABS201 uptake in vitro.

Example 8

Preclinical Studies of ABS201

**[0552]** Receptor screening. Three separate concentrations of ABS201 (10⁻⁷, 10⁻⁵ M) were screened individually against the following 16 receptors: adrenergic (α1a, α2, β), dopamine, histamine (H1, H2, H3), muscarinic (central, peripheral), nicotinic, opioid (nonselective), orphelin, serotonin (transporter, nonselective), monoamine oxidase (A, B). No displacement of the receptors’ endogenous substrate were observed. Hence ABS201 does not appear to bind with any of these receptors. In contrast, the ABS201 has nM affinity for the target receptor (NTR).”

**[0553]** Liver Receptor Screening (Stanford Research Institute Study B213-06). ABS201 (10⁻⁶, 10⁻⁵ M) was screened against the following liver CYPs: CYP1A2, CYP2B6, CYP2C9, CYP2C19, CYP2D6, CYP3A4.  No inhibition of substrate binding at any concentration of ABS201 to these enzymes was observed, indicating a lack of potential for drug-drug interactions.

**[0554]** Blood Distribution and Metabolite Identification. ABS201 was added to freshly isolated whole rat blood to a concentration of 100 µg/mL, allowed to partition, and the cellular fraction was removed by centrifugation. ABS201 at this concentration distributes almost evenly between the cellular and serum fractions. No metabolites of ABS201 were been detected, consistent with previous experiments in which a very long serum/plasma half-life was demonstrated.

**[0555]** Maximum Tolerated Dose. ABS201 was administered to rats at IV doses up to 100 mg/kg and oral doses up to 500 mg/kg. No adverse effects of the compound (body weight loss, mortality, abnormal clinical evaluations panel) were seen out to 48 hr post administration. These experiments thus define lower limits for the MTD of ABS201 at 100 times the ED₅₀ for the compound in antipsychosis and other tests reflective of brain activity.

**[0556]** Genetic Toxicology. The micronucleus study, mouse lymphoma study and Ames study were conducted. The micronucleus study evaluated the synthetic NT peptide for its potential to cause genetic damage as manifested by induced micronucleated polychromatic erythrocytes in mouse bone marrow cells. The mouse lymphoma study evaluated the synthetic NT peptide for its potential to cause mutations at the
thymidine kinase locus of L5178Y TK- mouse lymphoma cells, both unactivated and under S9 metabolic activation conditions. The Ames study evaluated the synthetic NT peptide for its potential to cause mutations in the histidine operon of Salmonella typhimurium strains TA98, TA100, TA1535 and TA1537 and the tryptophan operon of Escherichia coli strain WP2 uvrA, both unactivated and under S9 metabolic activation conditions. Experimental protocols are as follows.

In the micromolecule study, 90 CD-1 mice used in the study consisted of 10/sex/group in Groups 1-4 (control, low-, mid-, and high-dose) and 5/sex in the positive control group. Test peptide as administered orally, the positive control, Cyclophosphamide at 80 mg/kg, as administered orally. Approximately 24 and 48 hours after dosing, mice were killed and bone marrow obtained from both femurs. A slide of a bone marrow suspension was prepared, stained with Wright-Giemsa stain, dried, and scored blind. The number of polychromatic erythrocytes (PCE) among the total erythrocytes (PCE + normochromatc erythrocytes (NCE)) were determined for each animal by counting at least 200 erythrocytes. The number of micronucleated polycromatic erythrocytes (MPCe) then were scored for 2000 PCE per animal. A positive result was obtained if there is a positive dose-response trend or statistically significant increase in the number of MPCe at one or more dose levels compared to controls.

In the mouse lymphoma study, the standard rat liver S-9 was prepared by inducing male Sprague-Dawley rats with Aroclor-1254 or Phenobarbital and/or β-naphthoflavone. Positive controls included lycanthone methanesulfonate (HYC), which induces mutations at the TK locus without metabolic activation, and 7,12-Dimethylbenz[a]anthracene (DMBA), which induces mutations at the TK locus with metabolic activation. A range finding test was performed to identify the ABS201 concentrations that produce 0-100% toxicity. The positive controls (HYC and DMBA) and five concentrations of ABS201 were used in the assay with and without metabolic activation. Cells were exposed to each ABS dose, and the supernatant obtained, and incubated for 20 and 44 hours. After a 2-day expression period the cultures were cloned with the restrictive agent trifluorothymidine (allows the growth of TK-/-cells only) or vehicle control. The mutation frequency and induced mutation frequency were calculated. A positive response was obtained if at least one culture had a MF that was two times or greater than the average MF of the corresponding solvent control cultures and the response was dose dependent. A confirmatory assay was performed without S-9 activation to confirm Mutation Assay results.

In the Ames study, the synthetic NT peptide analogue was evaluated for its potential to cause mutations in the histidine operon of Salmonella typhimurium strains TA98, TA100, TA1535, TA1537 and the tryptophan operon of Escherichia coli strain WP2 uvrA. In brief, the standard rat liver S-9 was prepared by inducing male Sprague-Dawley rats with Aroclor-1254 or Phenobarbital and/or β-naphthoflavone. Bacterial strains used were Salmonella typhimurium strains TA98, TA100, TA1535, TA1537 and Escherichia coli strain WP2 uvrA. The positive controls included 2-AA, 2-NF, 9-AA, NaAz, and MMS. A solubility/miscibility assay was conducted to determine the maximum achievable concentration of peptide analogue in the selected solvent (water, DMSO, acetone, or ethanol). A range finding test was performed with and without S-9 activation using tester strains TA100 and WP2 uvrA only to identify ABS201 concentrations that produce 0-100% toxicity. The mutation assay was performed using the four Salmonella typhimurium strains (TA98, TA100, TA1535, TA1537) and the Escherichia coli strain WP2 uvrA strain using the plate incorporation method of treatment. A response was considered positive if either strain TA98 or TA 100 has a dose that produces a mean reversion frequency greater than or equal to two times the mean reversion frequency of the corresponding solvent control plates or if either strain TA 1535, TA 1537 or WP2 uvrA has a dose producing a three-fold or greater increase in the mean reversion frequency compared to the solvent control frequency.

Initial data indicate that ABS201 is inactive in all three assays.

hERG Assay. The objective of the hERG study is to assess the effects of the test article on the rapidly activating inward rectifying potassium current (I_Ko) conducted by hERG (human ether-a-go-go related gene) channels stably expressed in a HEK293 cell line. The method used is as follows. HEK293 cells, stably transfected with hERG cDNA, were used in the study. All experiments were performed at near-physiological temperature (35°± 2° C). A positive control (60 mM terfenadine) was applied to two cells (n=2). Cells were transferred to the recording chamber and superfused with HB-PS solution. The recording chamber and bath solution were maintained at a temperature of 35± 2° C, using a combination of in-line solution pre-heater, chamber floor heater, and feedback temperature controller. Bath temperature were measured using a thermistor probe.

Patch pipettes were made from glass capillary tubing using a P-97 micropipette puller (Sutter Instruments, Calif.). A commercial patch clamp amplifier was used for whole cell recordings. Before digitization, current records were low-pass filtered at one-fifth of the sampling frequency.

Cells stably expressing hERG were held at −80 mV. Onset and steady state block of hERG current due to ABS201 were measured using a pulse pattern with fixed amplitudes (conditioning prepulse: +20 mV for 1 sec; repolarizing test ramp to −80 mV (−0.5 V/s) repeated at 5 s intervals. Each recording ended with a final application of a supramaximal concentration of the reference substance (E-4031, 500 nM), to assess the contribution of endogenous currents. The remaining unblocked current was subtracted off-line digitally from the data to determine the potency of ABS201 for hERG inhibition.

Initial results indicate that ABS201 is inactive in the hERG study.

Pharmacokinetics and Brain Distribution of ABS201. ABS201 was administered in one IV dose (5 mg/kg) or oral dose (50 mg/kg) to rats. At selected time points, blood was removed or brain harvested and the concentration of the peptide analog determined using LC/MS/MS. It was demonstrated that ABS201 could be detected in the blood and brain up to 120 minutes after both IV and oral administration. The amount in the brain was sufficient to saturate NTR-1 to produce the observed behavioral effects.

ABSO201 was cleared from whole blood in two phases with an initial phase and a second phase where compound was measurable at low levels up to 120 minutes following IV administration of 5 mg/kg; the levels of ABS201 in whole blood were below the LLOD at all times following oral administration of the compound (50 mg/kg); the levels of ABS201 in brain were below the LLOD at all times following
IV administration; and measurable quantities of ABS201 were detected in brains of 2 of 3 animals 15 minutes post oral administration of 50 mg/kg.

Following IV injection, the mean value of systemic clearance was 2.45 L/hr/kg, which corresponded to 74.02% of rat hepatic blood flow. The mean values of half-life and Vc were 0.29 hr and 0.87 L/kg. ABS201 distributes well into the tissues. The volume of distribution at terminal phase was 0.87 ± 0.27 L/kg, which was similar to the total body water (0.67 L/kg) in the rats. The bioavailability of ABS201 following oral administration was 0.07% when in 0.9% saline solution. The mean value of half-life was 0.99 hr.

For pharmacokinetics studies of ABS201 in the brain, plasma samples and brain samples were collected from SD rats at four time points following intravenous infusion of test article ABS201.2HCl. These samples were then used for the determination of plasma and brain drug levels by LC/MS/MS for estimating BBB penetration and pharmacokinetic parameters. Time-course results are shown in FIG. 22. Following a 15 min intravenous infusion of ABS201.2HCl at 100 mg/kg, the value of systemic clearance was 22.77 L/hr/kg, which corresponded to 6.88 fold of rat hepatic blood flow (3.31 L/hr/kg). The value of half-life (TV) for ABS201.2HCl was 0.49 hr. The mean value of Cmax (at 25 minute) following intravenous infusion at a dose of 100 mg/kg was 5459.78 µg/L. The value of AUC(TV) was 4391.09 hr*µg/L.

ABS201.2HCl distributes well into the tissues. The volume of distribution at terminal phase was 16.10 L/kg, which was greater than the total body water (0.67 L/kg) in the rat.

Following a 15 min intravenous infusion of ABS201.2HCl at 100 mg/kg, the values of Cmax (at 45 minute) and AUC(TV) in brain were 32.93 ng/g and 60.96 hr*ng/g, respectively. The value of half-life (TV) for ABS201.2HCl was 4.85 hr. The brain-to-plasma ratio from 25 min to 135 min following intravenous infusion ranged from 0.0015 to 0.1453. These results indicate that ABS201 is selectively taken up into the brain, as it has a significantly longer half-life than in the blood. The brain half-life is in order with the half-lives of the behavioral effect seen for ABS201.

The plasma and brain pharmacokinetics of ABS201 was also studied following IV administration of 1 mg/kg and, oral administration of 30 mg/kg of ABS201 to non fasted rats. The results of this study indicated that: ABS201 was rapidly cleared from plasma following IV administration with t1/2 of about 5 minutes; the levels of the compound decreased below the LLOD by 45 min; the levels of ABS201 were below the LLOD at all times following oral administration; and the levels of ABS201 in brain were below the LLOD at all times following both IV and PO administration.

In vitro metabolism and compartmentalization of ABS201. To evaluate the distribution of ABS201 in blood, the extent of protein binding in plasma, and to gain a preliminary assessment of the metabolism of ABS201 in blood and plasma. The results of this study demonstrated: little or no binding of ABS201 to plasma proteins following a 5 or 30 min incubation at 37°C; no evidence of metabolism of ABS201 in whole blood or plasma following a 5 or 30 min incubation at 37°C; and a rapid distribution of about 33 percent of ABS201 into blood cellular elements when whole blood was incubated with ABS201 for 5 or 30 min at 37°C.

Evaluation of the Site of Uptake of ABS201 from PO Administration Brain. With the goal of ultimately increasing the overall oral bioavailability of ABS201 through formulation, the site of uptake in the gut was evaluated. The objective of this study was to collect plasma samples from SD rats at various time points following intra-stomach, intra-duodenum, intra-jejunum, and intra-colon administration of test article ABS201.2HCl in the in situ looped models. These samples were used for the determination of plasma drug by LC/MS/MS for estimating pharmacokinetic parameters and segmental absorption. The bioavailability of intra-stomach, intra-duodenum, intra-jejunum, intra-colon administration relative to intravenous administration and gavage was calculated. As can be seen in FIG. 23, ABS201 is absorbed almost entirely in the intestine, with intra-duodenum exhibiting the maximal amount of uptake. Virtually no ABS201 is taken up in the stomach.

Summary. Substitution of the N-terminal α-amino group of homolyzyne (Scheme 1) of ABS13 with the methyldesamino derivative 43 (Scheme 2) to produce ABS201 resulted in a molecule possessing important characteristics of a potential antipsychotic. In particular, ABS201 is highly selective for NTR-1 and exhibits effective antipsychotic activity in key rat models of psychosis when administered either IP or orally. ABS201 exhibited a 300% increase in central activity when administered orally, and achieved a more rapid response with oral versus IV injection (these unique attributes are attributable to the desamino modification). ABS201 does not cause catalepsy or induce drug resistance. Initial rat toxicity and mutagenesis experiments have been negative.

The pharmacokinetics, compartmentalization, and possible metabolism of ABS201 were evaluated in vitro and in vivo. The results suggest little or no metabolism, and complex pharmacokinetics which indicate that the compound initially undergoes a rapid clearance from blood, followed by a longer lasting, deep compartment phenomenon.

Also, the pharmacodynamic response of ABS201 is long acting (2-4 hr) following both IV and PO administration; the acute effects of IV ABS201 are mediated via central nervous system; the compound does not appear to be metabolized, blood. The brain half-life is in order with the half-lives of the behavioral effect seen for ABS201. ABS201 partitions between the aqueous and cellular phases of blood in vitro; the PK profile of ABS201 in whole blood is consistent with a two phase clearance process; and the pharmacodynamic response which has been observed is likewise consistent with a two phase clearance process.

ABS201 is preferentially taken up into the brain, presumably through an active transport process, as it is lost from the brain at about 10% the rate that it is lost from the blood. Given the long apparent half life of ABS201 in rat brain (detectable to 6 hours post dosing), and evidence that the drug forms a depot in blood elements (i.e., a slow release delivery system for the drug) the compounds of the invention, such as the semisynthetic peptide ABS201, could be admistered on a once or twice daily basis.

Example 9

Biological Improvements Resulting from Substitution of —H for —CH3 in Bioactive Peptides

A series of amino acid analogues of Arg and Lys have been created. See U.S. Pat. Nos. 6,043,218; 6,358,822; 6,566,330; 6,783,946; 6,858,396. When substituted for Arg8
in NT(8-13) a series of compounds have been generated that possess both antipsychotic and analgesic activity. Examples of these compounds can be found in Table 10 below (ABS202, ABS203, ABS204, ABS206). In addition to the non-natural side chains, it was demonstrated that substitution of a —CH₃ for the N-terminal —NH₂ resulted in the most active compounds for both indications. See PCT/US2005/021580, filed on Jun. 17, 2005, and U.S. Provisional Patent Application Ser. No. 60/591,333, filed on Jun. 17, 2004.

A further set of compounds have been generated in which a terminal —H was substituted for the —CH₃ to evaluate the role of the —CH₃ in the biological activity of these compounds (ABS226, ABS227, ABS228, ABS230, Table 10). The structures of compounds ABS202 and ABS203 are shown in Table 8. Compounds ABS226 and ABS227 differ from ABS202 and ABS203, respectively, in that the N-terminal —CH₃ groups of ABS202 and ABS203 are replaced by —H groups in ABS226 and ABS227. The structure of ABS204 is 30-Arg-Pro-Tyr-Ile-tertLeu-COOH, in which 30 refers to the non-natural residues shown in Scheme 2. The structure of ABS206 is N-Me-Lys-Arg-Pro-Tyr-Ile-tertLeu-COOH. Compounds ABS228 and ABS229 differ from compounds ABS204 and ABS206, respectively, in that the N-terminal —CH₃ groups of ABS204 and ABS206 are replaced by —H groups in ABS228 and ABS230.

Each of compound ABS226, ABS227, ABS228 and ABS230 was evaluated against its parent compound ABS202, ABS203, ABS204, ABS206, respectively, for antipsychotic and analgesic activity in appropriate rat models. The former activity is most easily monitored by measuring the extent of induced hypothermia (core body temperature loss), which is a measure of NTR-1 binding and agonism and thus antipsychotic activity. Hypothermia is followed routinely over time through insertion of a thermometer probe hooked up to a digital readout box into the anus of the rats. Analgesia is monitored by the hotplate assay, in which hotplate measurements are taken on a walled-in metal surface that is maintained at 52°C. The latency between the time the rat was placed on the surface and when it licks its back paw is measured. If the animal doesn’t lick its hind paw in 30 seconds, it is removed and a maximum positive effect (or MPE) of 100% is recorded. Intermediate responses achieve MPEs between 0-100%, with saline controls defined 0% latency.

**Example 10**

Assays for Analgesic Activity of NT Derivatives

**[0583]** Animals. Experimentally naïve male Sprague-Dawley rats from Charles River Laboratories, Raleigh, N.C. (230-260 g) or Harlan, Pratville Ala. (240-280 g), were maintained 4 per cage in animal quarters on a regular light/dark cycle (lights on 0600-1800) with ad libitum food and water for approximately 1 week before the experiment. The range of body weights on the 2 days of testing was 230-260 grams. The experiment was done on one day, with animals allowed to recover for three days before experiments are repeated.

**[0584]** Peptide Preparation. Test articles are prepared at the desired concentration in saline for either IP injection or gavage administration with N=6 for each dose and vehicle. Morphine (5 mg/kg) is used as a control for certain protocols. Specifically, test peptide was suspended in 0.9% NaCl and injected I.P. in a volume of 1 mL/kg (rat weights, 230-260 g) to give a dose of 10 mg/kg. Apomorphine HCI (Sigma-Aldrich, St. Louis, Mo.) was dissolved in distilled water at a concentration of 5 mg/mL within 40 minutes of administration for injection at 1 mL/kg S.C. to give a dose of 5 mg/kg. When complete ED₅₀ is determined, dosing range of the peptide will be 10, 3, 1, 0.3, and 0.1 of the estimated ED₅₀, covering two log orders of range.

**[0585]** The Hotplate Assay. The hotplate assay, a standard rat model for chronic pain (see LeBars et al., Pharmacol. Rev. 53: 597-652 (2001) and Chapman et al., Pain 22: 1-31 (2002)), was performed as follows. A hotplate maintained at 52°C to 53°C, was enclosed on four sides in a plexiglass chamber that extended 6 in. above the plate. Animals were placed on the plate, and the latency between the time a rat was placed on the

<table>
<thead>
<tr>
<th>Compound</th>
<th>N-terminus</th>
<th>Position</th>
<th>Hypothermia (°C)</th>
<th>Hypothermia (tmax, min)</th>
<th>% MPE</th>
<th>% MPE (tmax, min)</th>
</tr>
</thead>
<tbody>
<tr>
<td>ABS202</td>
<td>—CH₃</td>
<td>llys-Me</td>
<td>-4.00</td>
<td>150</td>
<td>100</td>
<td>45</td>
</tr>
<tr>
<td>ABS226</td>
<td>—H</td>
<td>llys-Me</td>
<td>-4.25</td>
<td>150</td>
<td>40</td>
<td>45</td>
</tr>
<tr>
<td>ABS203</td>
<td>—CH₃</td>
<td>llys-Me2</td>
<td>-1.0</td>
<td>150</td>
<td>61</td>
<td>60</td>
</tr>
<tr>
<td>ABS227</td>
<td>—H</td>
<td>llys-Me2</td>
<td>-1.7</td>
<td>135</td>
<td>29</td>
<td>60</td>
</tr>
<tr>
<td>ABS204</td>
<td>—CH₃</td>
<td>llys-Me3</td>
<td>-1.6</td>
<td>150</td>
<td>50</td>
<td>45</td>
</tr>
<tr>
<td>ABS228</td>
<td>—H</td>
<td>llys-Me3</td>
<td>-2.0</td>
<td>120</td>
<td>60</td>
<td>75</td>
</tr>
<tr>
<td>ABS206</td>
<td>—CH₃</td>
<td>lys-Me</td>
<td>-1.8</td>
<td>180</td>
<td>56</td>
<td>60</td>
</tr>
<tr>
<td>ABS230</td>
<td>—H</td>
<td>lys-Me</td>
<td>-3.8</td>
<td>180</td>
<td>100</td>
<td>30</td>
</tr>
</tbody>
</table>

1*All compounds had identical 9-13 residues of -Arg⁴⁰-Tyr⁴¹-Tyr⁴²-tertLeu⁴³-Lys⁴⁴.*

2*Indicates a particular patented non-natural analogue of Arg or Lys.

As seen in Table 10, substitution of —H for —CH₃ has an effect in each case. ABS226 has about the same antipsychotic potential as ABS202, but its much smaller analgesic activity is resulting from poorer NTR-2 binding. Thus the compound is more selective for NTR-1 and a better antipsychotic candidate. A similar effect is seen with ABS203 and ABS227. ABS228 has a better analgesic activity than ABS224, hence it is more selective for NTR-2. ABS226 is more active in both potential indications.
plate for 30 seconds without responding. Thus, % MPE are calculated using the following equation: % MPE = ((post-drug latency - pre-drug latency) / cut-off pre-drug latency) × 100%. 

N = 6 or greater for all experiments.

[0586] Values for each set of experiments will be submitted to analysis of variance, with dose (vehicle and 5 levels) as the factor. Following significant effects in analysis of variance, each dose will be compared to vehicle by the Newman-Keuls test. An effect will be considered significant if p < 0.05.

[0587] The Tail Flick Assay. Each of 6 rats per experiment were placed in a Plexiglas restraint box. A standard laboratory hot water bath was maintained at 49° C. The distal 3 cm of the rat’s tail was submerged into the water, and the time from insertion to tail flick or removal of tail from the water was recorded. Animals are removed immediately at the cutoff latency of 10 seconds. MPEs were calculated, with a mean MPE of 100% corresponding to the rat’s tail remaining in the water bath for 10 seconds. This was repeated 3 times. The tail was dried and allowed to rest for 20 seconds between trials. The rat was then given the appropriate amount of peptide and the experiment repeated at 30 and 60 minutes. Treatment conditions were distributed over the study in a counterbalanced design to control for such factors as calendar period and order of dosing. N = 6 or greater for all experiments.

[0588] Formalin Assay. Six naive rats were assigned to a single treatment condition, given either a test peptide as detailed above, or saline (control) and placed into Plexiglas observation cages (43 cm×24 cm×20 cm) without bedding material and their behavior monitored for 15 min. Animals were scored every minute on the following scale: 0 – forepaw flat on floor, 1 – forepaw just touching floor, 2 – forepaw held up off of floor, and 3 – forepaw licked or bitten. Following this baseline determination, animals were injected with a 5% solution of formaldehyde (50 mL of 5% formaldehyde in sterile saline prepared in 1 mL syringes with 26G/8 needle) subcutaneously into the right forepaw. The animals were immediately be placed into the observation cages, and their behavior recorded over the following 30 minutes. After each experiment, rats were euthanized with CO2 followed by cervical dislocation. An average Pain Rating over time versus the baseline and formalin-only controls for each peptide tested were plotted, and statistical analysis were performed using a one-way analysis of variance (ANOVA). Differences were considered statistically significant at the 5% confidence interval. N = 6 or greater for all experiments. A biphasic behavioral response has been established, with an initial phase (approximately 3 minutes after injection) reflecting the initial pain stimulus followed by a secondary phase after about 30 minutes reflecting sensitization due to instigation of inflammatory processes.

[0589] Chung assay. To assess tactile thresholds, rats are placed in a clear plastic, wire mesh-bottomed cage, divided into individual compartments. Animals are allowed to acclimate and then baseline thresholds are assessed prior to drug treatment. To assess the 50% mechanical threshold for paw withdrawal, von Frey hairs are applied to the plantar mid-end paw, avoiding the tori (footpads). The eight von Frey hairs used are designated by log10 (10 force required to bend hair, mg) and range from 0.4-15.1 grams (6’s 3.61-5.10). Each hair is pressed perpendicularly against the paw with sufficient force to cause slight bending, and held for approximately 6-8 seconds. A positive response is noted if the paw is sharply withdrawn. Flinching immediately upon removal of the hair is also considered a positive response. Absence of a response (“-“) is cause to present the next consecutive stronger stimulus; a positive response (“+”) is cause to present the next weaker stimulus. Stimuli are presented successively until either six data points are collected, or the maximum or minimum stimulus is reached. If a minimum stimulus is reached and positive responses still occurred, the threshold is assigned an arbitrary minimum value of 0.25 grams; if a maximum stimulus is presented and no response occurred, a maximum threshold value of 15 grams is assigned. If a change in response occurs, either G to “+” or “-” to “-”, causing a change in the direction of stimulus presentation from descending to ascending or vice-versa, four additional data points are collected subsequent to the change. The resulting pattern of responses are tabulated and the 50% response threshold computed using the formula: log (threshold, mg×10) = Xf/k + d; where:

[0590] Xf = value of the last von Frey hair applied;

[0591] k = correction factor based on pattern of responses (from calibration table);

[0592] d = mean distance in log units between stimuli.

Based on observations on normal, un-operated rats and sham-operated rats, the cutoff of a 15.1-g hair is selected as the upper limit for testing.

Example 11

ABS201 as a Potential Analgesic

[0593] The antipsychotic and hypothermic effects of NT and derivatives are mediated through NTR-1 agonism while analgesic effects are mediated through NTR-2 agonism. To be a viable analog, NT derivative should in vitro bind with high affinity to NTR-2 and in vivo be orally available and stable. ABS201 fulfills all of these requirements—having sub-micromolar NTR-2 affinity prolonged plasma stability and orally bioavailability as shown in Table 11.

<table>
<thead>
<tr>
<th>TABLE 11</th>
</tr>
</thead>
<tbody>
<tr>
<td>Indication</td>
</tr>
<tr>
<td>NTR-1 receptor binding (ED50, nM)</td>
</tr>
<tr>
<td>Functional</td>
</tr>
<tr>
<td>NTR-1 Agonist</td>
</tr>
<tr>
<td>NTR-2 receptor binding (ED50, nM)</td>
</tr>
<tr>
<td>Functional</td>
</tr>
<tr>
<td>NTR-2 Agonist</td>
</tr>
<tr>
<td>Plasma stability T1/2 (min)</td>
</tr>
<tr>
<td>Inhibits locomotor activity</td>
</tr>
<tr>
<td>Hypersalivation Inhibition</td>
</tr>
<tr>
<td>Catalepsy</td>
</tr>
<tr>
<td>Toxicity</td>
</tr>
<tr>
<td>Dose Tolerance</td>
</tr>
</tbody>
</table>

1Competitive NTR binding affinities versus radiolabeled NT[8-13][103]
2Induction of cAMP and inositol phosphate production (Orwig, Ph.D. dissertation, 2005).
3Quantitative MALDI-MS assay developed in our laboratory [53].
4Behavioral assay methods as described in [103].
5Single acute dose at 100 × ED50 administered IP [unpublished]
610 × ED50 administered daily over 5 days. [103].

[0594] ABS201 also exhibits analgesic activity as indicated by % MPE in the hotplate assay comparable in extent and duration to morphine (Fm = 24). Of interest, however, are differences in the animals’ affect after administration of these
peptides. Rats given morphine looked “drugged”, becoming more rigid and completely unresponsive to environmental stimuli, while animals given ABS201 appeared simply to be relaxed. The rats do not become tolerant to the analgesic activity of ABS201 as rats dosed for five days straight show statistically identical % MPEs as when initially dosed (FIG. 25). This is especially noteworthy as the rats are not experimentally naive by the end of the dose time course, which can result in them learning to avoid the hotplate and mask the potential for analgesia. Finally, the analgesic activity of ABS201 is not linked to its hypothermic effect since the two do not show parallel kinetics—specifically the analgesic effect wears off while the hypothermic effect is still strong. This clearly indicates that the effects are mediated by different receptors. Overall, these data indicate the usefulness of ABS201 as a potential analgesic and antipsychotic.

Example 12  
Analgesic Peptides Having Side Chain Guanidinium Groups

[0595] By virtue of the fact that ABS201 binds both NTR-1 and -2 with high affinity, there is a chance that effects associated with NTR-1 binding may make the peptide less desirable as an analgesic. These effects include induction of systemic hypothermia in rats, which is actually a very small effect in humans (D. Feifel, personal communication), and antipsychotic potency, which may not be undesirable. A peptide highly selective for NTR-2 that maintains all of the favorable attributes of ABS201 would be a “cleaner” analgesic candidate.

[0596] In the process of developing an NTR-2 selective ligand, the peptides shown in FIG. 26 were evaluated. These peptides exhibited enhanced NTR-2 to NTR-1 binding affinities, and the relative selectivity for the two receptors are provided in Table 12.

<table>
<thead>
<tr>
<th>Peptide</th>
<th>NTR-1 (nM)</th>
<th>NTR-2 (nM)</th>
<th>NTR-1/NTR-2 selectivity</th>
</tr>
</thead>
<tbody>
<tr>
<td>NT[8-13]</td>
<td>0.21 ± 0.01</td>
<td>4.1 ± 0.01</td>
<td>19.5</td>
</tr>
<tr>
<td>A8S1</td>
<td>0.38 ± 0.07</td>
<td>0.7 ± 0.02</td>
<td>1.8</td>
</tr>
<tr>
<td>A8S13</td>
<td>5.7 ± 0.37</td>
<td>11.1 ± 0.1</td>
<td>1.9</td>
</tr>
<tr>
<td>A8S201</td>
<td>11.2 ± 0.3</td>
<td>n/d</td>
<td>n/d</td>
</tr>
<tr>
<td>A8S15</td>
<td>0.4 ± 0.1</td>
<td>0.9 ± 0.2</td>
<td>2.5</td>
</tr>
<tr>
<td>A8S16</td>
<td>0.3 ± 0.05</td>
<td>0.67 ± 0.07</td>
<td>2.2</td>
</tr>
<tr>
<td>A8S17</td>
<td>0.25 ± 0.07</td>
<td>0.3 ± 0.1</td>
<td>1.2</td>
</tr>
<tr>
<td>A8S19</td>
<td>1.2 ± 0.7</td>
<td>0.23 ± 0.05</td>
<td>0.19</td>
</tr>
</tbody>
</table>

*Competitive

[0597] Since NT[8-13] has a 20-fold preference for NTR-1 binding over NTR-2 (Table 12), it is less useful as an NTR-2 selective ligand. However, the data for peptides A8S1, A8S13, and A8S201 indicate that there is a structure-binding effect in varying the ε-amino group. Changing the polar —NH$_2$ (which is protonated at physiological pH significantly adding to its hydrophobicity) of A8S1 to nonpolar, uncharged —N$_3$ (A8S13) resulted in about a 10-fold improvement of NTR-2 to NTR-1 selectivity. In addition, as the arginine side chains get larger and more nonpolar, significantly better binding to NTR-2 resulted. The differences in the arginine sidechain binding site of NTR-2 is illustrated in the progression of A8S15, A8S16, A8S17, A8S19. A8S19 has a 5-fold selectivity for NTR-2, and an overall selectivity improvement of about 100-fold over NT[8-13]. In addition, significant loss of receptor binding affinity is tolerated in this system as A8S201, which binds 50-fold less effectively than its parent, still exhibited saturable effects in the rat schizophrenic models when given orally. A8S201 is about three times as physiologically active as A8S13 in the rat behavioral models of schizophrenia when administered orally, clearly defining this as the best substitution for the ε-amino group. Based on this rationale, peptides designed to be better NTR-2 selective ligands are created and their structures are presented in FIG. 27. All of these peptides contain R=Arg-Pro-Tyr-tert-leu-CONH$_2$ and the N-terminal methyl group of A8S201, which are required for the potential of oral activity. These peptides also incorporate added steric bulk around the guanidinium group, elements that demonstratively improve NTR-2 versus NTR-1 selectivity versus NT[8-13] (as does the N-terminal methyl group). Desirable analgesic peptides are those which (1) exhibit analgesic activity comparable to or better than A8S201 in the hotplate, tail flick and formalin assays when IP injected, (2) exhibit activity when orally administered, (3) have desirable NTR-1 and NTR-2 binding activities, and (4) exhibit plasma stability.

Example 13  
Analgesic NT Peptide Analogues

[0598] The process of identification of A8S201 enabled definition of the key structural parameters necessary for biological stability and blood brain barrier crossing. Accordingly, a group of neurotensin (NT)[8-13] derivatives that incorporate structural elements that may enhance IP or oral bioavailability were created. Within these parameters, other proprietary structural changes were incorporated that can affect various pharmacological and behavioral parameters relevant to the potential development as therapeutic agents. In sum, peptides with potential for (1) enhanced binding affinities for their receptors, e.g. enhanced binding and selectivity (e.g. NT-derivatives with selectivity for brain NTR-2 have analgesic activity, while those with selectivity for NTR-1 have antipsychotic activity); (2) enhanced biological barrier crossings; (3) enhanced stability; and (4) feasible synthetic costs were identified.

[0599] Specifically, chemical changes to the established NT[8-13] derivative included incorporation of an non-natural amino acid residue at position 8. The new peptides were designed to retain the structural elements of the antipsychotic lead A8S201 that are necessary for oral activity while varying the Arg(8) residue and moiety at the N-terminus. Peptides with enhanced analgesic activity, i.e. predicted to have NTR-2 selectivity, were screened via IP injection in the rat hotplate model of analgesia as described above. Initial screening experiments of five peptides using the three different assays for analgesic activity described above indicated that the assays yielded comparable relative activity, with the hotplate assay being the most sensitive. The IP dose chosen was designed to produce a maximal response from active compounds as measured by the % MPE (Maximal Positive Effect). An MPE of 100% indicates that the rat did not feel pain for the extent of time on the hotplate, using a cutoff of 30 seconds. Tmax indicates the time after dosing in which a maximum response was observed. The tmx for all compounds were recorded as this gave an indication of the duration of each compound’s analgesic effect. Compounds that exhibited a % MPE of 100% were evaluated with oral dosing.
Table 13 provides the sequences of the peptides tested, selectivity for the NTR-1 or NTR-2 as indicated by ability to induce hypothermia, and the results of the Hotplate assay. Compounds with an asterisk exhibited significant activity in the hotplate assay.

<table>
<thead>
<tr>
<th>Peptide* N-terminus</th>
<th>Peptide Sequence</th>
<th>Hypo (I.P.)</th>
<th>Tmax (oral)</th>
<th>MPE (I.P.)</th>
<th>Tmax (oral)</th>
</tr>
</thead>
<tbody>
<tr>
<td>ABS201* CH₃</td>
<td>hLys-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-5.20</td>
<td>150.00</td>
<td>-2.50</td>
<td>100</td>
</tr>
<tr>
<td>ABS202* CH₃</td>
<td>1-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-4.00</td>
<td>150.00</td>
<td>-2.15</td>
<td>100</td>
</tr>
<tr>
<td>ABS203 CH₃</td>
<td>2-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.00</td>
<td>150.00</td>
<td>ND</td>
<td>60</td>
</tr>
<tr>
<td>ABS204 CH₃</td>
<td>3-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.60</td>
<td>150.00</td>
<td>ND</td>
<td>19</td>
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<tr>
<td>ABS205* CH₃</td>
<td>Lys-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-4.00</td>
<td>195.00</td>
<td>-2.50</td>
<td>86</td>
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<tr>
<td>ABS206 CH₃</td>
<td>4-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.80</td>
<td>180.00</td>
<td>ND</td>
<td>55</td>
</tr>
<tr>
<td>ABS207* CH₃</td>
<td>5-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-2.90</td>
<td>165.00</td>
<td>-1.50</td>
<td>84</td>
</tr>
<tr>
<td>ABS208* CH₃</td>
<td>6-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-2.80</td>
<td>135.00</td>
<td>-0.20</td>
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<td>ABS209 CH₃</td>
<td>Orn-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-2.55</td>
<td>195.00</td>
<td>ND</td>
<td>89</td>
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<td>ABS210* CH₃</td>
<td>7-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.18</td>
<td>180.00</td>
<td>-2.13</td>
<td>82</td>
</tr>
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<td>ABS211* CH₃</td>
<td>8-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-4.20</td>
<td>210.00</td>
<td>-1.15</td>
<td>100</td>
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<tr>
<td>ABS212* CH₃</td>
<td>9-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-4.20</td>
<td>240.00</td>
<td>-2.45</td>
<td>100</td>
</tr>
<tr>
<td>ABS213 CH₃</td>
<td>Arg-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-2.65</td>
<td>180.00</td>
<td>ND</td>
<td>48</td>
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<tr>
<td>ABS214 CH₃</td>
<td>10-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-4.20</td>
<td>195.00</td>
<td>-2.40</td>
<td>100</td>
</tr>
<tr>
<td>ABS215 CH₃</td>
<td>11-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-2.45</td>
<td>165.00</td>
<td>ND</td>
<td>63</td>
</tr>
<tr>
<td>ABS216 CH₃</td>
<td>12-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-2.83</td>
<td>195.00</td>
<td>ND</td>
<td>74</td>
</tr>
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<td>ABS217 CH₃</td>
<td>13-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.80</td>
<td>90.00</td>
<td>ND</td>
<td>8</td>
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<td>ABS218 CH₃</td>
<td>15-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.88</td>
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<td>ABS220 CH₃</td>
<td>14-Arg-Pro-Tyr-t-Leu-Leu</td>
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<td>-1.30</td>
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<td>ABS221 CH₃</td>
<td>16-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.80</td>
<td>165.00</td>
<td>ND</td>
<td>76</td>
</tr>
<tr>
<td>ABS224 CH₃</td>
<td>alkene-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.30</td>
<td>195.00</td>
<td>ND</td>
<td>65</td>
</tr>
<tr>
<td>ABS501 CH₃</td>
<td>15-Arg-Pro-Tyr-Ile-Leu</td>
<td>-4.20</td>
<td>100.00</td>
<td>ND</td>
<td>51</td>
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<tr>
<td>ABS700 H</td>
<td>Boc-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-2.06</td>
<td>90.00</td>
<td>ND</td>
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<tr>
<td>ABS225* H</td>
<td>hLys-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-3.00</td>
<td>180.00</td>
<td>-1.20</td>
<td>100</td>
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<tr>
<td>ABS226 H</td>
<td>1-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-3.55</td>
<td>150.00</td>
<td>-0.48</td>
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<td>ABS227 H</td>
<td>2-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.70</td>
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<tr>
<td>ABS228 H</td>
<td>3-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.98</td>
<td>120.00</td>
<td>ND</td>
<td>55</td>
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<tr>
<td>ABS229 H</td>
<td>Lys-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.65</td>
<td>105.00</td>
<td>ND</td>
<td>48</td>
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<tr>
<td>ABS230* H</td>
<td>4-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-3.75</td>
<td>180.00</td>
<td>-1.10</td>
<td>100</td>
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<tr>
<td>ABS231 H</td>
<td>5-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-2.39</td>
<td>125.00</td>
<td>-1.63</td>
<td>100</td>
</tr>
<tr>
<td>ABS232* H</td>
<td>6-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-3.87</td>
<td>240.00</td>
<td>-0.38</td>
<td>84</td>
</tr>
<tr>
<td>ABS233 H</td>
<td>Orn-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.40</td>
<td>180.00</td>
<td>ND</td>
<td>51</td>
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TABLE 13 - continued

<table>
<thead>
<tr>
<th>Peptide N-terminus</th>
<th>Peptide Sequence</th>
<th>Hypo (I.P.)</th>
<th>Hypo (oral)</th>
<th>MPE (I.P.)</th>
<th>MPE (oral)</th>
</tr>
</thead>
<tbody>
<tr>
<td>ABS234 H</td>
<td>7-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-4.18 240.00</td>
<td>-0.93</td>
<td>100</td>
<td>15-120</td>
</tr>
<tr>
<td>ABS235 H</td>
<td>8-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-2.58 165.00</td>
<td>-2.18</td>
<td>100</td>
<td>30-75</td>
</tr>
<tr>
<td>ABS236 H</td>
<td>9-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-2.37 180.00</td>
<td>ND</td>
<td>100</td>
<td>30-60</td>
</tr>
<tr>
<td>ABS237 H</td>
<td>Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.85 165.00</td>
<td>ND</td>
<td>50</td>
<td>30</td>
</tr>
<tr>
<td>ABS238 H</td>
<td>10-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-3.80 165.00</td>
<td>-2.08</td>
<td>100</td>
<td>30, 60-75</td>
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<tr>
<td>ABS239 H</td>
<td>11-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-3.53 165.00</td>
<td>-0.78</td>
<td>92</td>
<td>15</td>
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<tr>
<td>ABS240 H</td>
<td>12-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-2.55 165.00</td>
<td>ND</td>
<td>37</td>
<td>45</td>
</tr>
<tr>
<td>ABS241 H</td>
<td>13-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-3.35 135.00</td>
<td>-0.75</td>
<td>95</td>
<td>60</td>
</tr>
<tr>
<td>ABS242 H</td>
<td>14-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-2.65 135.00</td>
<td>ND</td>
<td>76</td>
<td>30</td>
</tr>
<tr>
<td>ABS243 H</td>
<td>15-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-2.38 135.00</td>
<td>ND</td>
<td>79</td>
<td>60</td>
</tr>
<tr>
<td>ABS244 H</td>
<td>16-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-4.25 165.00</td>
<td>-1.50</td>
<td>82</td>
<td>45</td>
</tr>
<tr>
<td>ABS245 H</td>
<td>17-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-3.13 165.00</td>
<td>-1.27</td>
<td>50</td>
<td>45</td>
</tr>
<tr>
<td>ABS246 H</td>
<td>18-Arg-Pro-Tyr-t-Leu-Leu</td>
<td>-1.68 165.00</td>
<td>ND</td>
<td>80</td>
<td>45</td>
</tr>
</tbody>
</table>

1Hot plate measurements were taken on a walled-in metal surface maintained at 52°C. The latency between the time the rat was placed on the surface and when it licks its back paw is measured. Animals are removed immediately at the cutoff latency of 30 sec, which corresponds to a %MPE of 100%.
2Side chain residue at 8-position as defined in Scheme 3.
3Group substituted for the α-amino group.
4%MPE and Tmax of %MPE for compound administered IP at 10 mg/kg.
5%MPE and Tmax of %MPE for compound administered orally at 20 mg/kg.
ND = not determined, IP = in progress

Scheme 3 - Structures of Non-natural Amino Acids

1 n = 3; R = NHMe 4 n = 2; R = NHMe 7 n = 1; R = NHMe
2 n = 3; R = NMMe 5 n = 2; R = NMMe 8 n = 1; R = NMMe
3 n = 3; R = NMMe 6 n = 2; R = NMMe 9 n = 1; R = NMMe
10 R' = Me; R'' = H 11 R' = Me; R'' = Me
12 R' = Et; R'' = H 13 R' = Et; R'' = Et
14 R' = Et; R'' = Me
Initial screens of all peptides included (1) evaluating their analgesic ability when dosed IP or orally (indicative of NTR-2 agonism) and (2) evaluating their ability to induce hypothermia when IP, a secondary effect of NTR-1 agonist activity. Peptides that exhibited significant analgesic activity, that is, greater than a set threshold level, when IP dosed were evaluated with oral dosing. The peptides that have high analgesic and low hypothermia-inducing activity when dosed orally, indicating high selectivity for NTR-2, were preferred. However, peptides with high hypothermia activity may show significantly greater oral bioavailability and were also preferred as they were better analgesic candidates, particularly if side effects of NTR-1 binding were minor. A third parameter was that IP analogues may have different utilities, and potencies, than orally administered peptides. Defined dose-response curves were also considered in selecting an analgesic candidate. Accordingly, the different parameters were weighted, and peptides that were not as “good” at others under the above analysis while showing overall similar profiles were not selected. A secondary screening that involved evaluating analgesia when dosed orally was performed, and the best peptides were subjected to a full dose-response analysis.

Based on the above criteria, peptides ABS201, ABS202, ABS205, ABS207, ABS208, ABS210, ABS211, ABS212, ABS220, ABS225, ABS230, ABS232, ABS234, and ABS239 were identified as preferred analgesic peptides. The structures for these peptides are summarized in the following table.

### TABLE 14

<table>
<thead>
<tr>
<th>Peptide (Ri)</th>
<th>N-terminus</th>
<th>C-terminus*</th>
</tr>
</thead>
<tbody>
<tr>
<td>ABS201</td>
<td>CH₃</td>
<td>hLys-Arg-Pro-Tyr-tLeu-Leu-COOH</td>
</tr>
<tr>
<td>ABS202</td>
<td>CH₃</td>
<td>4-Arg-Pro-Tyr-tLeu-Leu-COOH</td>
</tr>
<tr>
<td>ABS205</td>
<td>CH₃</td>
<td>5-Arg-Pro-Tyr-tLeu-Leu-COOH</td>
</tr>
<tr>
<td>ABS207</td>
<td>CH₃</td>
<td>6-Arg-Pro-Tyr-tLeu-Leu-COOH</td>
</tr>
<tr>
<td>ABS208</td>
<td>CH₃</td>
<td>7-Arg-Pro-Tyr-tLeu-Leu-COOH</td>
</tr>
<tr>
<td>ABS210</td>
<td>CH₃</td>
<td>8-Arg-Pro-Tyr-tLeu-Leu-COOH</td>
</tr>
<tr>
<td>ABS211</td>
<td>CH₃</td>
<td>9-Arg-Pro-Tyr-tLeu-Leu-COOH</td>
</tr>
<tr>
<td>ABS212</td>
<td>CH₃</td>
<td>11-Arg-Pro-Tyr-tLeu-Leu-COOH</td>
</tr>
<tr>
<td>ABS220</td>
<td>CH₃</td>
<td>15-Arg-Pro-Tyr-tLeu-Leu-COOH</td>
</tr>
</tbody>
</table>

*See Scheme 3 above for the structure of the N-terminal moiety.

Example 14

Evaluation of the Analgesic Effects of Selected NTR-analogues

The analgesic effects of ABS201, ABS205, ABS210, ABS212, and ABS220 administered by gavage at a dose of 20 mg/kg are shown in FIG. 28. These results indicate that ABS201, ABS205, ABS210 and ABS220 were as much or more active than ABS212 when given orally. The analgesic effects of ABS232 and ABS239 as determined using the hotplate assay are illustrated in FIGS. 29 A and B.

When administered by I.P. injection, however, the most active peptide was ABS212. The analgesic property of ABS212 was compared with that of morphine using the hotplate, tail flick and formalin assays. Results obtained from the hotplate assay and the formalin assays are shown in FIGS. 30 A and B, respectively, while results obtained from the tail flick assay is summarized in Table 15 below.

ABS212 at an approximately equal molar ratio exhibits analgesic activity (% MPE) in the hotplate assay comparable or better in extent and duration to morphine (FIG. 30A). Of interest, however, are differences in the animals' affect after administration of these peptides. Rats given morphine looked “drugged,” becoming more rigid and completely unresponsive to environmental stimuli, while animals give ABS212 appeared simply to be relaxed. In addition, the rats do not become tolerant to the analgesic activity of ABS212 as rats dosed for five days straight show statistically identical % MPEs as when initially dosed (data not shown). This is especially noteworthy as the rats are not experimentally naïve by the end of the dose time course, which can result in them learning to avoid the hotplate and mask the potential for analgesia. FIGS. 30B and C show the dose response data for ABS212 when administered IP or orally, respectively. Approximate ED₅₀'s for IP and oral doses are 2.5-5 mg/kg and 30-40 mg/kg respectively.

In the formalin assay, ABS212 provides a full analgesic effect for the time course of the experiment (FIG. 31A). As also seen in the hotplate assay, morphine at approximately equimolar concentrations, loses activity after 30 minutes. ABS212, however, has a more pronounced analgesic effect when administered by I.P. injection. FIGS. 31 B and C show the dose responses determined using the formalin assay for I.P. and oral administration, respectively. Approximate IP and oral ED₅₀'s are in the same range as the assays performed above. The analgesic activity ABS212 appears to be highly selective for NTR-2 as very little hypothermia (a secondary
effect of NTR-1 binding) is elicited from this peptide at therapeutic analgesic doses. (In vitro receptor binding studies are in progress).

**Example 16**

Synthesis of 5-oxo-Pro-His-Trp-Ser-Tyr-LEU-2HCL was 0.11±0.08% with choice of 1 mg/kg group as IV dose.

**Example 17**

Synthesis of butyryl-His-DPhe-(N"-Et)-Arg-Trp-Sar-NH₂ 3 TFA (4)

**List of Documents**

**[0606]** In the tail flick assay, ABS212 exhibits statistically significant analgesia at 30 minutes, with an apparent greater effect at 60 minutes, when compared to saline (see Table 15). ABS212 statistically compares to morphine.

**TABLE 15**

<table>
<thead>
<tr>
<th>Treatment</th>
<th>MPE (0 min)</th>
<th>MPE (30 min)</th>
<th>MPE (60 min)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Saline</td>
<td>-0.37 +/- 0.57</td>
<td>9.7 +/- 5.6</td>
<td>14.6 +/- 7.1</td>
</tr>
<tr>
<td>Morphine</td>
<td>0.03 +/- 0.05</td>
<td>63.7 +/- 41.9</td>
<td>67.5 +/- 25.8</td>
</tr>
<tr>
<td>ABS212</td>
<td>1.16 +/- 0.05</td>
<td>75.4 +/- 28.4</td>
<td>88.7 +/- 19.6</td>
</tr>
</tbody>
</table>

**[0607]** FIGS. 32A and B show the dose response data for ABS212 when administered I.P. and orally, respectively, in the tail flick assay. Approximate IP and oral ED₀₅₀ are in the same range as seen in the above assay. FIG. 33 demonstrate that ABS212 is highly active in the Chung model for neuropathic pain when administered by I.P. injection. In sum, ABS212 is a highly effective analgesic in the four key rat models for the spectrum of pain states: acute (hotplate, tail flick), chronic (formalin) and neuropathic (Chung).

**Example 15**

Preclinical Characterization of ABS212

**[0608]** To evaluate the pharmacokinetics of ABS212, plasma samples were collected from Sprague-Dawley (SD) rats at various time points following intravenous and oral administration of test article ABS212. These samples were used for the determination of plasma parent drug concentrations by LC/MS/MS for estimating pharmacokinetic parameters and oral bioavailability. The concentration versus time curve for IV and oral administration of ABS212 are shown in FIGS. 34 A and B.

**[0609]** Following IV bolus injection of ABS-212.2HCL at 5, 2.5 and 1 mg/kg, the mean±SD values of systemic clearance were 0.40±0.05, 0.40±0.17 and 1.07±0.15 L/hr/kg, which corresponded to 12.08%, 12.08% and 32.33% of rat hepatic blood flow (3.31 L/hr/kg), respectively. The mean±SD values of half-life (T₁/₂) for ABS212.2HCL were 0.29±0.13, 0.28±0.11 and 0.28±0.08 hr. ABS-212.2HCL dose not distributes well into the tissues. The volume of distribution at terminal phase following IV administration at nominal doses of 5, 2.5 and 1 mg/kg were 0.16±0.06, 0.15±0.01 and 0.43±0.15 L/kg, respectively, which were less than the total body water (0.67 L/kg) in the rats. The dose-normalized AUC₀→∞ was 2.68±2.92:1 at respective doses of 5, 2.5, and 1 mg/kg, while the dose-normalized Cₘₚ was 2.68±2.46:1 at doses of 5, 2.5, and 1 mg/kg, respectively, suggesting there was a saturable elimination when given high dosage.

**[0610]** Following oral administration of ABS-212.2HCL at a dose of 50 mg/kg, the mean±SD values of Cmax and Tₘₚ for ABS-212.2HCL were 27.08±11.55 µg/L and 0.14±0.05 hr, respectively; the mean±SD values of AUC₀→∞ and half-life (T₁/₂) were 53.06±39.65 hr·µg/L and 1.42±0.87 hr, respectively. The mean±SD value of bioavailability for ABS-212.2HCL was 0.11±0.08% with choice of 1 mg/kg group as IV dose data.


agonist, but not haloperidol: a potential predictive model for novel antipsychotic effects. 


[0741] Additional advantages of the invention in part will be obvious from the description, or may be learned by practice of the invention. The advantages of the invention will be realized and attained by means of the elements and combinations particularly pointed out in the appended claims.

[0742] Throughout this application, where publications are referenced, the disclosures of these publications in their entirety are hereby incorporated by reference into this application in order to more fully describe the state of the art to which this invention pertains.

wherein Y is a leaving group; with an amine of the formula R'R'R'N, in a solvent for a time period and at a temperature sufficient for the formation of the compound of Formula (XI). 2. A method for the preparation of a compound of Formula (XIII):

![Diagram](XIII)

or a salt, hydrate, solvate, stereoisomer, or tautomer thereof, wherein X is OH, O(C₁₋₆ alkyl), O(C₆₋₁₀ aryl), NR₂, an amino acid residue or a peptide residue, or wherein X is O, NR, an amino acid residue, or a peptide residue, respectively covalently bonded to a polymer;

wherein R¹, R², and R³ are independently hydrogen, alkyl, aryl, alkylalkoxy, hydroxyalkyl, or heteroaryl, wherein one of R¹, R² or R³ may be absent, wherein any alkyl can contain 0-3 heteroatoms comprising N, O, or S, wherein any alkyl, alkylalkyl, hydroxyalkyl, or heteroaryl may be unsubstituted or substituted; the method comprising contacting a compound of Formula (XII):

![Diagram](XII)

with an amine of the formula R¹'R²'R³'R₄'R₅', in a solvent for a time period and at a temperature sufficient for the formation of the compound of Formula (XII).
O, NR, an amino acid residue, or a peptide residue, respectively covalently bonded to a polymer;

Z is H, NR₃, alkyl, aryl, alkenyl, heterocyclyl, heteroaryl, heterocyclylalkyl, heteroarylalkyl, cycloalkyl, cycloalkylalkyl, an amino acid residue, or a peptide residue, or is NR, an amino acid residue, or a peptide residue, respectively covalently bonded to a polymer;

provided that X and Z are not both bonded to polymers; and provided that at least one of X or Z comprises an amino acid residue or a peptide residue, or an amino acid residue or a peptide residue respectively covalently bonded to a polymer;

R at each occurrence is independently H, alkyl, aryl, heterocyclyl, heteroaryl, a nitrogen protecting group, or a nitrogen capping group, or two R groups together with the nitrogen atom to which they are bound form a 5-7 membered heterocyclic ring that can further comprise 1-3 additional heteroatoms N, O, or S;

L is a saturated or unsaturated, linear, branched or cyclic hydrocarbyl or a combination thereof, comprising n carbon atoms, wherein any carbon atom can be substituted or unsubstituted, wherein 0-3 heteroatoms O, S, or NR can be substituted for a carbon atom of L;

n is 1 to about 10;

R¹ is hydrogen, alkyl, aryl, alkenyl, cycloalkyl, heterocycles, or heteroaryl; wherein any alkyl can contain 0-3 heteroatoms comprising N, O, or S; wherein any alkyl, aryl, cycloalkyl, heterocyclic, or heteroaryl may be unsubstituted or substituted;

R⁶ and R⁸ are at each occurrence independently H, alkyl, cycloalkyl, heterocyclyl, aryl or heteroaryl, wherein any alkyl, cycloalkyl, aryl, heterocyclic or heteroaryl residue may be unsubstituted or substituted; provided that R or R₅ is not a Boc group;

from a compound of formula (XII):

\[
(XII)
\]

wherein Y is a leaving group;

the method comprising contacting the compound of formula (XII) with an amine of the formula R¹NH₂, in a solvent for a time period and at a temperature sufficient for the formation of the compound of Formula (XV):

\[
(XV)
\]

then, contacting the compound of formula (XV) with a compound of Formula (XIV):

\[
(XIV)
\]

wherein R⁵ is substituted or unsubstituted alkyl or aryl, or a salt thereof;

in a solvent for a time period and at a temperature sufficient for the formation of the compound of Formula (XIII).

4. The method of claim 1 wherein Z is not methyl.

5. The method of claim 1 wherein Y is halo, alkyloxy, aralkyloxy, aryloxy or alkarylloxy, wherein the alkyl or aryl is unsubstituted or is substituted with electron-withdrawing groups.

6. The method of claim 1 wherein Z is H.

7. The method of claim 1 wherein Z is methyl.

8. The method of claim 1 wherein the compound of Formula (XI) comprises a lysine residue, a N-methyllysine residue, a N,N-dimethyllysine residue, or an N,N-dimethyllysine N-methyl salt residue.

9. The method of claim 1 wherein the compound of Formula (XI) is a neurotensin analog or an analog of a neurotensin fragment.

10. The method of claim 1 wherein X comprises a Pro-Tyr residue.

11. The method of claim 1 wherein Z comprises a PGlu-Leu-Tyr residue.

12. The method of claim 2 wherein R⁶ is methyl or ethyl.


15. A compound of Formula (XIV) comprising a salt thereof.

16. The compound of claim 15 wherein the salt is a hydroiodide salt.

17. The method of claim 1 wherein the solvent is water, methanol, or ethanol, or a mixture thereof.

18. The method of claim 1 wherein the solvent is DMSO, DMF, DMAC, or NMP, or a mixture thereof.

19. The method of claim 2 wherein the solvent is water, acetonitrile, methanol, ethanol, tetrahydrofuran, or N,N-dimethylformamide, or a mixture thereof.

20. The method of claim 1 wherein L is a linear or cycloalkylalkyl, saturated hydrocarbyl of 1 to about 10 carbon atoms.

21. A method of use of a peptide analog prepared according to claim 1 for treatment of a malcondition in a patient in need thereof, the method comprising administering the analog in a dosage, at a frequency, and over a period of time sufficient to provide a beneficial effect to the patient.

22. The method of claim 21 wherein the peptide analog is a neurotensin analog.

23. The method of claim 21 wherein the malcondition comprises pain.

24. The method of claim 21 wherein the malcondition comprises psychosis.

25. The method of claim 21 wherein the malcondition comprises obesity.
26. The method of claim 3 wherein Y is halo, alkylsulfonyl, arylsulfonyl or alkarylsulfonyl, wherein the alky or ary is unsubstituted or is substituted with electron-withdrawing groups.

27. The method of claim 4 wherein Y is halo, alkylsulfonyl, arylsulfonyl or alkarylsulfonyl, wherein the alky or ary is unsubstituted or is substituted with electron-withdrawing groups.

28. The method claim 2 wherein Z is H.

29. The method of claim 3 wherein Z is H.

30. The method of claim 4 wherein Z is H.

31. The method of claim 2 wherein Z is methyl.

32. The method of claim 3 wherein Z is methyl.

33. The method of claim 3 wherein the compound of Formula (XIII) comprises a lysine residue, a N-methyllysine residue, a N,N-dimethyllysine residue, or an N,N-dimethyllysine N-metho salt residue.

34. The method of claim 8 wherein Z is not methyl.

35. The method of claim 33 wherein Z is not methyl.

36. The method of claim 2 wherein the compound of Formula (XIII) is a neurotensin analog or an analog of a neurotensin fragment.

37. The method of claim 3 wherein the compound of Formula (XIII) is a neurotensin analog or an analog of a neurotensin fragment.

38. The method of claim 2 wherein the compound of Formula (XIII) is a neurotensin analog or an analog of a neurotensin fragment.

39. The method of claim 3 wherein the compound of Formula (XIII) is a neurotensin analog or an analog of a neurotensin fragment.

40. The method claim 4 wherein the compound of Formula (XI) is a neurotensin analog or an analog of a neurotensin fragment.

41. The method of claim 2 wherein X comprises a Pro-Tyr residue.

42. The method of claim 3 wherein X comprises a Pro-Tyr residue.

43. The method of claim 4 wherein X comprises a Pro-Tyr residue.

44. The method of claim 2 wherein Z comprises a PGLu-Leu-Tyr residue.

45. The method of claim 3 wherein Z comprises a PGLu-Leu-Tyr residue.

46. The method of claim 4 wherein Z comprises a PGLu-Leu-Tyr residue.

47. The method of claim 3 wherein R⁶ is methyl or ethyl.

48. A compound of Formula (XIII) prepared by the method of claim 3.

49. The method of claim 2 wherein the solvent is water, methanol, or ethanol, or a mixture thereof.

50. The method of claim 3 wherein the solvent is water, methanol, or ethanol, or a mixture thereof.

51. The method of claim 4 wherein the solvent is water, methanol, or ethanol, or a mixture thereof.

52. The method of claim 2 wherein the solvent is DMSO, DMF, DMAc, or NMP, or a mixture thereof.

53. The method of claim 3 wherein the solvent is DMSO, DMF, DMAc, or NMP, or a mixture thereof.

54. The method of claim 4 wherein the solvent is DMSO, DMF, DMAc, or NMP, or a mixture thereof.

55. The method of claim 2 wherein L is a linear or cycloalkylalkyl, saturated hydrocarbyl of 1 to about 10 carbon atoms.

56. The method of claim 3 wherein L is a linear or cycloalkylalkyl, saturated hydrocarbyl of 1 to about 10 carbon atoms.

57. A method of use of a peptide analog prepared according to claim 2 for treatment of a malcondition in a patient in need thereof, the method comprising administering the analog in a dosage, at a frequency, and over a period of time sufficient to provide a beneficial effect to the patient.

58. The method of claim 57 wherein the peptide analog is a neurotensin analog.

59. The method of claim 57 wherein the malcondition comprises pain.

60. The method of claim 57 wherein the malcondition comprises psychosis.

61. The method of claim 57 wherein the malcondition comprises obesity.

62. A method of use of a peptide analog prepared according to claim 3 for treatment of a malcondition in a patient in need thereof, the method comprising administering the analog in a dosage, at a frequency, and over a period of time sufficient to provide a beneficial effect to the patient.

63. The method of claim 62 wherein the peptide analog is a neurotensin analog.

64. The method of claim 62 wherein the malcondition comprises pain.

65. The method of claim 62 wherein the malcondition comprises psychosis.

66. The method of claim 62 wherein the malcondition comprises obesity.

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