

(72) TSAO, YOW-MIN D., US

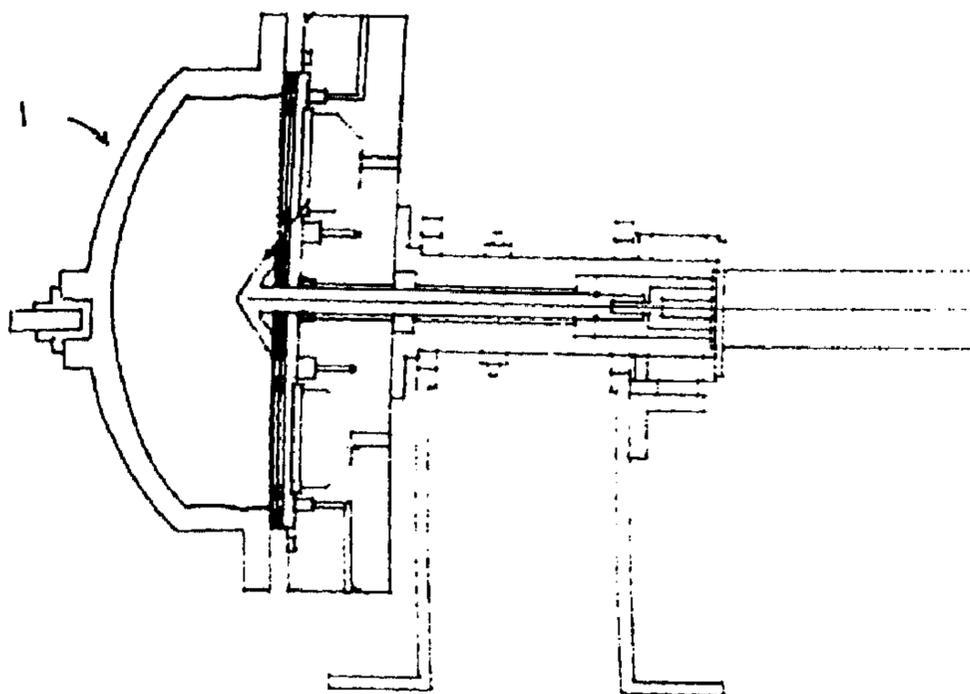
(71) WYLE LABORATORIES, INC., LIFE SCIENCES, US

(51) Int.Cl.⁶ C12N 5/02, C12M 3/02

(30) 1998/06/29 (09/106,987) US

(54) **BIOREACTEUR ET PROCÉDES DE CULTURE DE CELLULES
UTILISANT CE BIOREACTEUR**

(54) **BIOREACTOR AND CELL CULTURING PROCESSES USING
THE BIOREACTOR**



(57) L'invention concerne un bioréacteur et un procédé de culture de cellules utilisant ce bioréacteur. Ce réacteur et ce procédé ont été développés pour améliorer la production de cellules et leur croissance et pour réduire le cisaillement agissant sur les cellules et les agrégats multicellulaires. Le bioréacteur comprend un récipient de culture en forme de dôme (1) qui possède des murs délimitant un volume interne (2), un orifice en pointe (5), un bord inférieur circulaire (3) et un axe de symétrie perpendiculaire au bord inférieur circulaire (3) ainsi qu'une membrane perméable aux gaz (6) intégrée de manière étanche au bord inférieur circulaire (3) du récipient de culture (1). Le bioréacteur peut en outre contenir une base rotative (7) intégrée de manière étanche à la membrane perméable aux gaz (6), ladite base rotative (7) étant fixée de manière à pouvoir tourner autour de l'axe de symétrie du récipient de culture en forme de dôme (1) placé sur un axe sensiblement horizontal.

(57) A bioreactor and cell culturing method using the bioreactor have been developed to enhance cell production and growth and decrease the shear forces on the cells and multicellular aggregates. The bioreactor contains a dome-shaped culture vessel (1) having walls defining an interior volume (2), an apex (5), a bottom circular edge (3), and an axis of symmetry perpendicular to the bottom circular edge (3); and a gas-permeable membrane (6) fluidtightly integrated with the bottom circular edge (3) of the culture vessel (1). The bioreactor may further contain a rotating base (7) fluidtightly integrated with the gas-permeable membrane (6), wherein the rotating base (7) is fixed to rotate about the axis of symmetry of the dome-shaped culture vessel (1) set on a substantially horizontal axis.

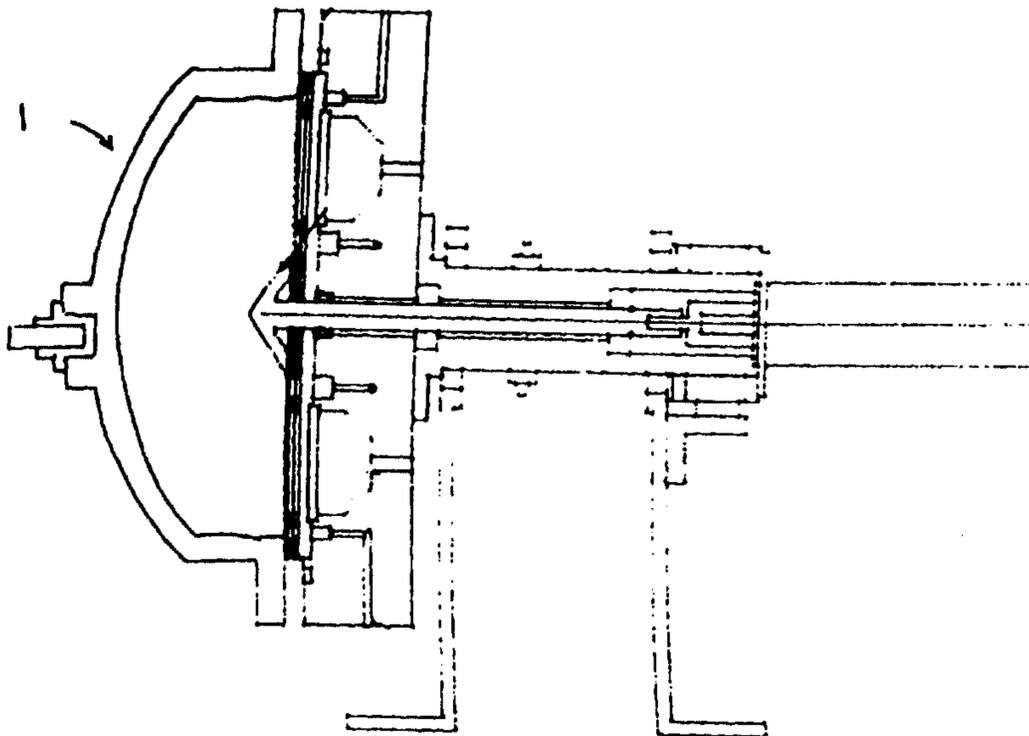
PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<p>(51) International Patent Classification ⁶ : C12N 5/02, C12M 3/02</p>	A1	<p>(11) International Publication Number: WO 00/00586</p> <p>(43) International Publication Date: 6 January 2000 (06.01.00)</p>
<p>(21) International Application Number: PCT/US99/14972</p> <p>(22) International Filing Date: 28 June 1999 (28.06.99)</p> <p>(30) Priority Data: 09/106,987 29 June 1998 (29.06.98) US</p> <p>(71) Applicant: WYLE LABORATORIES, INC., LIFE SCIENCES [US/US]; Suite 120, 1290 Hercules Drive, Houston, TX 77058 (US).</p> <p>(72) Inventor: TSAO, Yow-Min, D.; 4910 Red Lantern Drive, Friendswood, TX 77546 (US).</p> <p>(74) Agent: WAACK, Janelle, D.; Arnold, White & Durkee, P.O. Box 4433, Houston, TX 77210 (US).</p>	<p>(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).</p> <p>Published <i>With international search report.</i></p>	

(54) Title: BIOREACTOR AND CELL CULTURING PROCESSES USING THE BIOREACTOR



(57) Abstract

A bioreactor and cell culturing method using the bioreactor have been developed to enhance cell production and growth and decrease the shear forces on the cells and multicellular aggregates. The bioreactor contains a dome-shaped culture vessel (1) having walls defining an interior volume (2), an apex (5), a bottom circular edge (3), and an axis of symmetry perpendicular to the bottom circular edge (3); and a gas-permeable membrane (6) fluidly integrated with the bottom circular edge (3) of the culture vessel (1). The bioreactor may further contain a rotating base (7) fluidly integrated with the gas-permeable membrane (6), wherein the rotating base (7) is fixed to rotate about the axis of symmetry of the dome-shaped culture vessel (1) set on a substantially horizontal axis.

5

BIOREACTOR AND CELL CULTURING PROCESSES USING THE BIOREACTOR

The invention relates to a bioreactor and a method for culturing cells using the
10 bioreactor. More particularly, the invention is directed to a bioreactor having a dome-
shaped culture vessel that rotates about its axis of symmetry set on a substantially
horizontal plane and a method for culturing cells in that bioreactor.

Many academic and industrial processes rely on *in vitro* culturing to generate
greater amounts of cells of interest. In these processes, a sample of cells is generally
15 placed in a vessel, provided with nutrients, and agitated. After a sufficient period of time
to allow production and growth of new cells, the cultured cells are removed from the
vessel and purified. For example, small-scale cell culturing has traditionally been
performed in shallow shaker flasks. The cells and nutrient media are combined in the
flask and a mechanical or magnetic stirring mechanism circulates the mixture. This
20 circulation is important to prevent the cells from settling to the bottom of the culture
vessel due to gravity and to ensure sufficient nutrient transfer to and waste bioproducts
from the growing cells.

These mass transfer requirements are important for all forms of culturing cells,
for example, in the form of a single cells, multicellular aggregates or cells attached to a
25 substrate. In many culture systems, particularly mammalian culture systems, the
formation of multicellular aggregates is important to accumulate large biomasses.
Mammalian cells are also often cultured in an attached state to mimic the *in vivo*
environment. When shear forces become too high, these forces can deform and damage
individual cells, impede cell growth, prevent or limit the aggregation of cells, or pull
30 apart aggregates and tear the walls of adjacent cells.

Several research groups have studied the effects of shear stress on cell cultures.
Stathopoulos et al., *Biotechnol. Bioeng.* 27:1021-1026 (1985), studied shear stress effects
on human embryonic kidney cells *in vitro* and reported that forces of 0.65 N/m² had
significant effects on cell morphology and forces higher than 2.6 N/m² caused marked
35 reduction in cell viability. Croughan et al., *Biotechnol. Bioeng.*, 29:130-141 (1987), and
Croughan et al., *Biotechnol. Bioeng.* 33:731-744 (1989), evaluated growth of FS-4 (human

5 diploid fibroblasts) microcarrier cultures at various impeller rotation rates in spinner vessels equipped with magnetic stir bars. Croughan et al. (1987) suggests that this analysis would not apply to vessels having different geometries.

A variety of geometries have been proposed for cell culture vessels. Tsao et al., "Fluid Dynamics Within a Rotating Bioreactor in Space and Earth Environments," *Journal of Spacecraft and Rockets* 31:937-943 (1994), and Tsao et al., "Mass Transfer Characteristics of NASA Bioreactors by Numerical Simulation," *Advances in Heat and Mass Transfer in Biotechnology* (Clegg, S., ed.) HTD-Vol. 355:69-73 (1997), disclose a bioreactor having a culturing vessel with two concentric, independently rotating cylinders. U.S. Patent Nos. 5,026,650 (Schwartz., issued June 25, 1991); 5,155,034 (Wolf et al., issued October 13, 1992); and 5,153,133 (Schwartz et al., issued October 6, 1992) disclose a bioreactor having a horizontally disposed cylindrical culture vessel that rotates about its longitudinal axis. The vessel contains a coaxially-disposed, oxygen-permeable membrane and access ports to inject or withdraw nutrient media.

U.S. Patent No. 5,151,368 (Brimhall et al., issued September 29, 1992) describes a dual-axis continuous flow bioreactor for handling high solids loaded materials such as coal and mineral ores. The bioreactor is mounted to a horizontal axle, corresponding to the bioreactor's longitudinal axis, and moves in a circular path about its vertical shaft while simultaneously rotating about its horizontal axis by an interlocking set of bevel ring gears. The vessel contains several conduits for introducing and withdrawing media and gases.

U.S. Patent No. 5,565,361 (Mutsakis et al., issued October 15, 1996) describes a bioreactor containing a porous, fibrous sheet material that acts as a motionless mixing element and as a substrate for attaching the cultivating cells. U.S. Patent No. 5,622,857 (Goffe, issued April 22, 1997) describes a bundled hollow fiber bioreactor and its use to prepare eucaryotic cells. The flow of oxygenated nutrient medium is maintained at a sufficient pressure to prevent the formation of bubbles that may block flow of media through the fibers. U.S. Patent No. 5,688,687 (Palsson et al., issued November 18, 1997) describes a bioreactor for mammalian cell growth having a circular cell growth chamber between a planar cell bed and a gas permeable membrane. Media inlet and outlets are arranged in a concentric circular arrangement and provide for the delivery of nutrients to the cells. U.S. Patent No. 5,605,835 (Hu et al., issued February 25, 1997) describes a

5 bioreactor that immobilizes animal cells in an insoluble, biocompatible matrix. Selective membranes allow for the separation of cell nutrients and cell wastes while collecting the desired cell products within the cell chamber.

There is a need for improved bioreactors and cell culturing methods that enhance the production and growth of cells without creating shear forces that damage cultivating
10 cells or multicellular aggregates.

This invention relates to a bioreactor and a cell culturing method that use the bioreactor. More particularly, the invention is directed to a bioreactor having a dome-shaped culture vessel that rotates about its axis of symmetry set on a substantially horizontal plane and a method for culturing cells in that bioreactor.

15 The inventive bioreactor comprises a dome-shaped culture chamber having walls defining an interior volume and a circular ridge at the bottom of the chamber. The circular ridge of the culture chamber is fluidtightly integrated with a gas permeable membrane. The membrane side of the culture chamber may be fluidtightly integrated with a rotating base in an orientation where the axis of symmetry of the dome-shaped
20 culture chamber is substantially horizontal. The dome-shaped design and horizontal orientation provide a low shear environment and mass transfer that is favorable for culturing cells, particularly mammalian cells that form multicellular aggregates for the production of large biomasses.

The method according to the invention comprises providing the inventive
25 bioreactor, adding cells and appropriate nutrients to the interior volume of the dome-shaped culture chamber, and rotating the base at a rate that generates suitable mass transfer and low shear stress.

In a preferred embodiment, the bioreactor comprises additional features that permit efficient addition of cell samples and removal of cell product, efficient addition
30 and removal of nutrients and gases, easy assembly of the bioreactor components, and a flexible design to allow for a wide variety of cell cultures and culturing conditions.

The following figures form part of the specification and are included to further demonstrate certain aspects of the present invention. The invention may be better understood by reference to one or more of these drawings in combination with the
35 detailed description of specific embodiments presented herein.

5 FIG. 1 is a schematic diagram a bioreactor having a dome-shaped culture vessel.

FIG. 2 is a schematic drawing of a bioreactor having a culture vessel with an alternative dome-shaped design.

FIG. 3 is a schematic drawing of a bioreactor having a culture vessel with another alternative dome-shaped design.

10 The bioreactor according to the invention has a dome-shaped culture vessel that rotates about its axis of symmetry set on a substantially horizontal plane. The circular ridge of the culture chamber is fluidtightly integrated with a gas-permeable membrane. The membrane-side of the culture chamber may be fluidtightly integrated with a rotating
15 base in an orientation where the axis of symmetry of the dome-shaped culture chamber is substantially horizontal. The dome-shaped design and horizontal orientation provide a low shear environment and mass transfer that is favorable for culturing cells, particularly mammalian cells that form multicellular aggregates for the production of large biomasses.

For purposes of this invention, "dome-shaped," in the context of describing the
20 culture vessel, means having the shape of a dome or cone, or a shape that substantially resembles a dome or cone. That is, the bottom of the vessel has a circular ridge, the vessel's axis of symmetry is perpendicular to the circular ridge and the walls of the vessel extend to an apex. The walls of the vessel leading to the apex are, for at least a portion, concave; the walls can have a partial non-curved portion. The apex can be a
25 single point at the peak of the curved walls or can be a level surface at the peak of the curved walls. Several variations of the dome-shaped culture vessel are exemplified in FIGS. 1-3.

The term "substantially horizontal," in the context of describing the orientation of the axis of symmetry of the dome-shaped culture chamber, means that the axis is no
30 more than 20 degrees from the horizontal plane. In a further preferred embodiment, the axis of symmetry is set no more than 10 degrees from the horizontal plane and, ideally, is in a horizontal plane.

With reference to FIG. 1, the preferred embodiments of the bioreactor are described as follows. The inventive bioreactor comprises a dome-shaped culture chamber
35 1 having an interior volume 2, a bottom circular ridge 3, walls having at least a partially curved surface 4 and an apex 5. The circular ridge of the culture chamber is fluidtightly

5 integrated with a gas-permeable membrane 6. The membrane-side of the culture chamber is fluidtightly integrated with a rotating base 7 in an orientation where the axis of symmetry of the dome-shaped culture chamber 8 is substantially horizontal.

The dome-shaped culture chamber can have a variety of shapes, as exemplified by the alternative dome-shapes depicted in FIGS. 1, 2, and 3. The culture chamber can be made of any material that is compatible with the culturing system, including glass, plastic, or stainless steel. Preferably, the vessel comprises a material that can be sterilized by ethylene oxide, gamma irradiation, or autoclaving, or, alternatively, is disposable.

The interior volume 2 of the culture vessel can vary greatly, depending upon the types of cells, amount of cells to be produced and available laboratory space. In a preferred embodiment, the volume of the substantially dome-shaped vessel is between about 10 mL and about 10 L. Small and medium scale laboratory cultures can preferably be performed in vessels of 100 mL, 250 mL, and 500 mL volumes. Larger preparative scale cultures can preferably be performed in vessels of 1 L, 5 L, and 10 L volumes.

20 The gas permeable material 3 can generally be any material compatible with the culturing system, *i.e.*, to allow flow of appropriate gases and to restrict flow of cells and liquid nutrients across the membrane. For example, the gas permeable membrane can comprise polytetrafluoroethylene, polyethylene, or porous hydrophobic TEFLON® membrane. The gas permeable membrane may additionally allow flow of liquids, nutrients, and metabolites across the membrane.

The bioreactor can further comprise an access port or a plurality of access ports to the culture vessel 9, 10, 11. The access ports allow transfer of materials and gases into and out of the bioreactor. Access ports can lead to the surface of the gas-permeable membrane 10 or can lead directly into the culture vessel 9, 11. The access ports leading directly into the culture vessel can go through the gas permeable membrane 9 or can go through the wall or apex of the culture vessel 11. The access ports can be any type of port used with culture or reaction vessels, including valves and membranes that can be penetrated by tubing, syringe, pipette or other sampling device.

35 The bioreactor can further comprise a agitator within the culture vessel 12. This agitator can be any type of device that provides additional agitation to the culture mixture, including a magnetic spinner or a mechanically driven propeller 12.

5 The invention is further directed to a method for culturing cells. The method according to the invention comprises providing the inventive bioreactor, adding cells and appropriate nutrients to the interior volume of the dome-shaped culture chamber, and rotating the base at a rate that generates suitable mass transfer and low shear stress. Where the mass transfer of the culture solution is provided by both the rotation of the culture chamber and by spinning an agitator in the culture vessel, the combined effects of these means must be weighed in determining the respective rotation rates.

10 The culturing of cells is preferably performed under conditions that optimize mass transfer and create low shear forces. Preferably, the culturing cells are subjected to average shear forces of less than about 1 dyne/cm², more preferably less than about 0.75 dyne/cm², and most preferably less than about 0.5 dyne/cm².

The bioreactor and culturing method according to the invention can be use for culturing any type of cell. In a preferred embodiment, the cells are eucaryotic cells, preferably mammalian cells. The cells can be adherent cells or non-adherent cells, single cells or multicellular aggregates. Adherent cells are attached to a substrate, such as microcarriers, fibrous supports or other cells.

20 The culturing method further comprise adding nutrients that enhance cell production and growth and removing waste products from the culturing process. Preferably, the gases are transferred across the gas permeable membrane 3. Alternatively, gases, nutrients or cell samples can be extracted from a port extending through the gas permeable membrane 9 or other access ports in the culture vessel wall or at its apex 11. For example, gas bubbles can be removed or culture fluid samples taken from an access port at the culture vessel apex to avoid disrupting the operation of the bioreactor. The culture vessel can be temporarily tilted to position the apex of the culture vessel so that bubbles rise to the apex access port.

EXAMPLE

30 A bioreactor can be designed with the following dimensions. The outer dome-shaped cell culture vessel has a radius of $r_o = 11$ cm, radius of curvature $r = 8.6$ cm and rotating at w_o rpm, while the inner spinner has a radius of $r_i = 3$ cm, height $h = 1$ cm and rotates in the opposite direction at w_i rpm. The rotating base has a perfluorocarbon chamber of 60 mL volume for oxygenation, nutrient inlet and outlet ports. The dome-shaped culture vessel of 125 mL volume is filled with culture medium into which

5 aggregates of cells and microcarrier beads were introduced. The rates of rotation of the vessel and spinner are limited to a range in which the radial, circumferential, and axial velocity were large enough to suspend particles and provide adequate mixing, but small enough to prevent turbulence. Relative rotation rates may be determined and optimized by simulating flow fields with software such as FLUENT or FIDAP (both are
10 commercially available from Fluent, Inc., Lebanon, NH). The unique advantage of this rotational perfused hemispherical bioreactor is that the fluid shear force created by the differential rate of rotation of the dome and spinner would be more uniform and controllable.

Table 1: Alternative Bioreactor Dimensions

Design	Inner Spinner rpm	Outer Dome rpm	Differential Rate ^a (rpm)	Outer Dome Radius	Inner Spinner Radius (cm)	Dome Height (cm)
1	2	-2	4	3.6	2.7	3.0
2	5	-2	7	5.5	1.5	2.0
3	5	-2	7	11	3	1.0

15

^a The difference between the inner spinner and outer dome rotating rate.

The rates of rotation of the concave shaped vessel and spinner were limited to a range in which the radial, circumferential, and axial velocity were large enough to suspend particles and provide adequate mixing, but small enough to prevent turbulence.
20 The unique advantage of this bioreactor was that the fluid shear force created by the differential rate of rotation of the dome and spinner would be more uniform and controllable.

All of the disclosed and claimed compositions, methods, and apparatus can be made and executed without undue experimentation in light of the present disclosure.
25 While the compositions and methods of this invention have been described in terms of preferred embodiments, it will be apparent to those of skill in the art that variations can be applied to the compositions, methods, and apparatus and in the steps or in the sequence of steps of the methods described herein without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain

WO 00/00586

PCT/US99/14972

- 5 agents which are both chemically and physiologically related can be substituted for the agents described herein while the same or similar results would be achieved. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention.

5

CLAIMS

1. A bioreactor comprising:
a dome-shaped culture vessel having walls defining an interior volume, an apex, a bottom circular edge, and an axis of symmetry perpendicular to the bottom circular edge and running through the apex; and
10 a gas-permeable membrane fluidtightly integrated with the bottom circular edge of the culture vessel, wherein the culture vessel is adapted to rotate about its axis of symmetry.
2. The bioreactor of claim 1, further comprising a rotating base fluidtightly integrated with the gas-permeable membrane, wherein the rotating base is fixed to rotate
15 about the axis of symmetry of the dome-shaped culture vessel set on a substantially horizontal axis.
3. The bioreactor of claim 1, wherein the rotating base is attached to a horizontally disposed axle that is rotated by a motor.
4. The bioreactor of claim 1, wherein the rotating base comprises a plurality of
20 access ports for transporting materials to and from the culture chamber.
5. The bioreactor of claim 1, wherein the culture vessel comprises an access port at its apex.
6. The bioreactor of claim 1, wherein the culture vessel contains an agitator.
7. The bioreactor of claim 6, wherein the agitator is a magnetic spinner or a
25 mechanically driven propeller.
8. The bioreactor of claim 1, wherein the gas-permeable membrane comprises polytetrafluoroethylene, polyethylene, or porous hydrophobic TEFLON.
9. The bioreactor of claim 1, wherein the volume of the culture vessel between about 10 mL and about 10 L.
- 30 10. A method for culturing cells comprising:
 - (a) providing a bioreactor, wherein the bioreactor comprises a dome-shaped culture vessel having walls defining an interior volume, an apex, a bottom circular edge, and a axis of symmetry perpendicular to the bottom circular edge and running through the apex; and a gas-permeable membrane fluidtightly integrated with the bottom circular
35 edge of the culture vessel; wherein the culture vessel is adapted to rotate about its axis of symmetry;

- 5 (b) adding cells of interest to the culture vessel;
 (c) adding culture medium to the culture vessel;
 (d) rotating the rotating base at a rate sufficient to provide sufficient mass
transfer and low shear forces on the culturing cells.

11. The method of claim 10, wherein:
10 the bioreactor further comprises a rotating base fluidtightly integrated with the gas-
permeable membrane; and
the rotating base is fixed to rotate about the axis of symmetry of the dome-shaped culture
vessel set on a substantially horizontal axis.

12. The method of claim 10, wherein the average shear force on the culturing cells is
15 less than about 1 dyne/cm².

13. The method of claim 10, wherein the cells are eucaryotic cells.

14. The method of claim 13, wherein the eucaryotic cells are mammalian cells.

15. The method of claim 14, wherein the culturing cells are in the form of
multicellular aggregates.

20 16. The method of claim 10, further comprising adding oxygen to the culture vessel
through the gas permeable membrane.

17. The method of claim 10, further comprising providing a substrate to the culture
vessel for the attachment of culturing cells.

18. The method of claim 17, wherein the substrate comprises microcarrier beads.

25

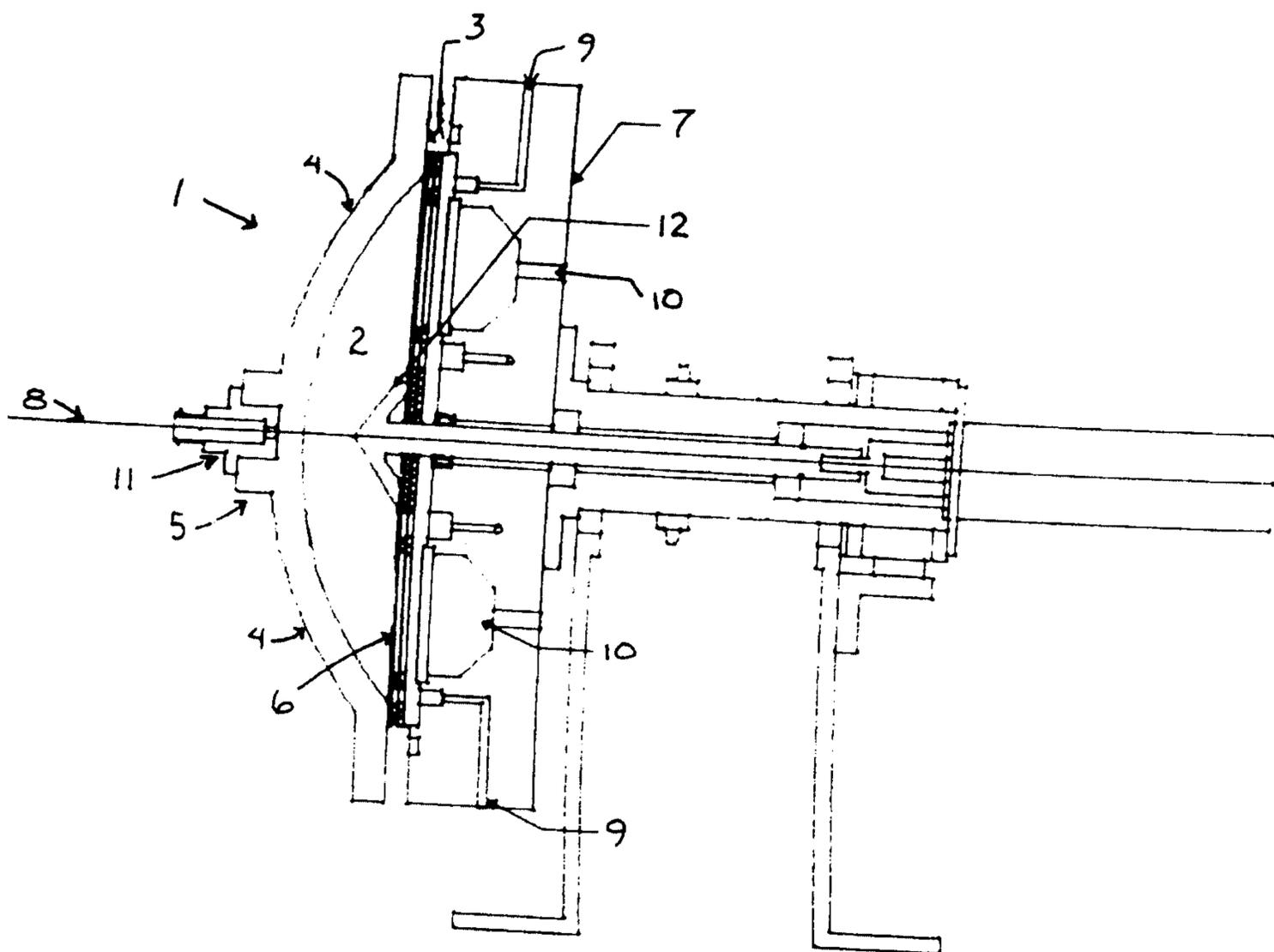


FIG. 1

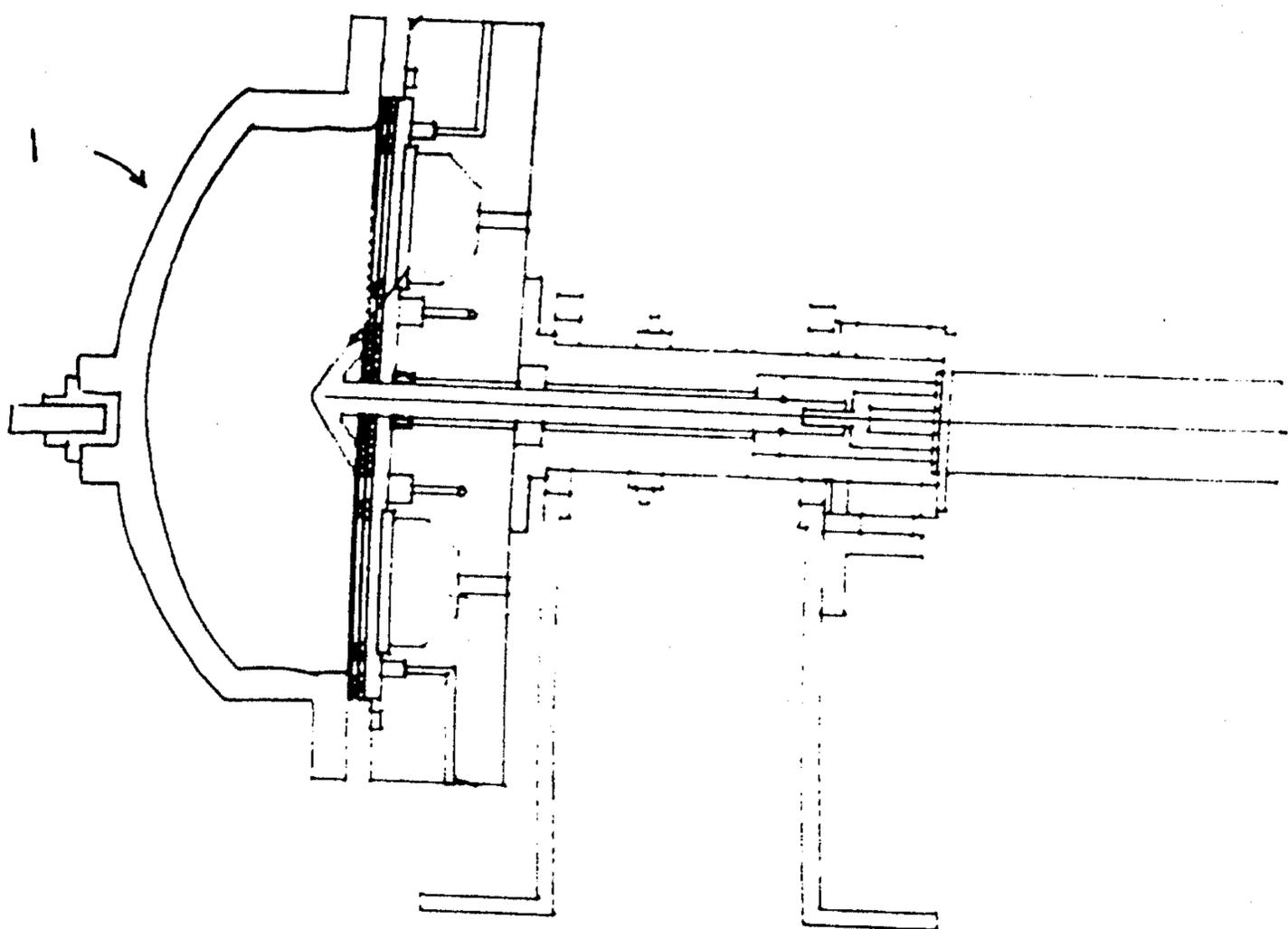


FIG. 2

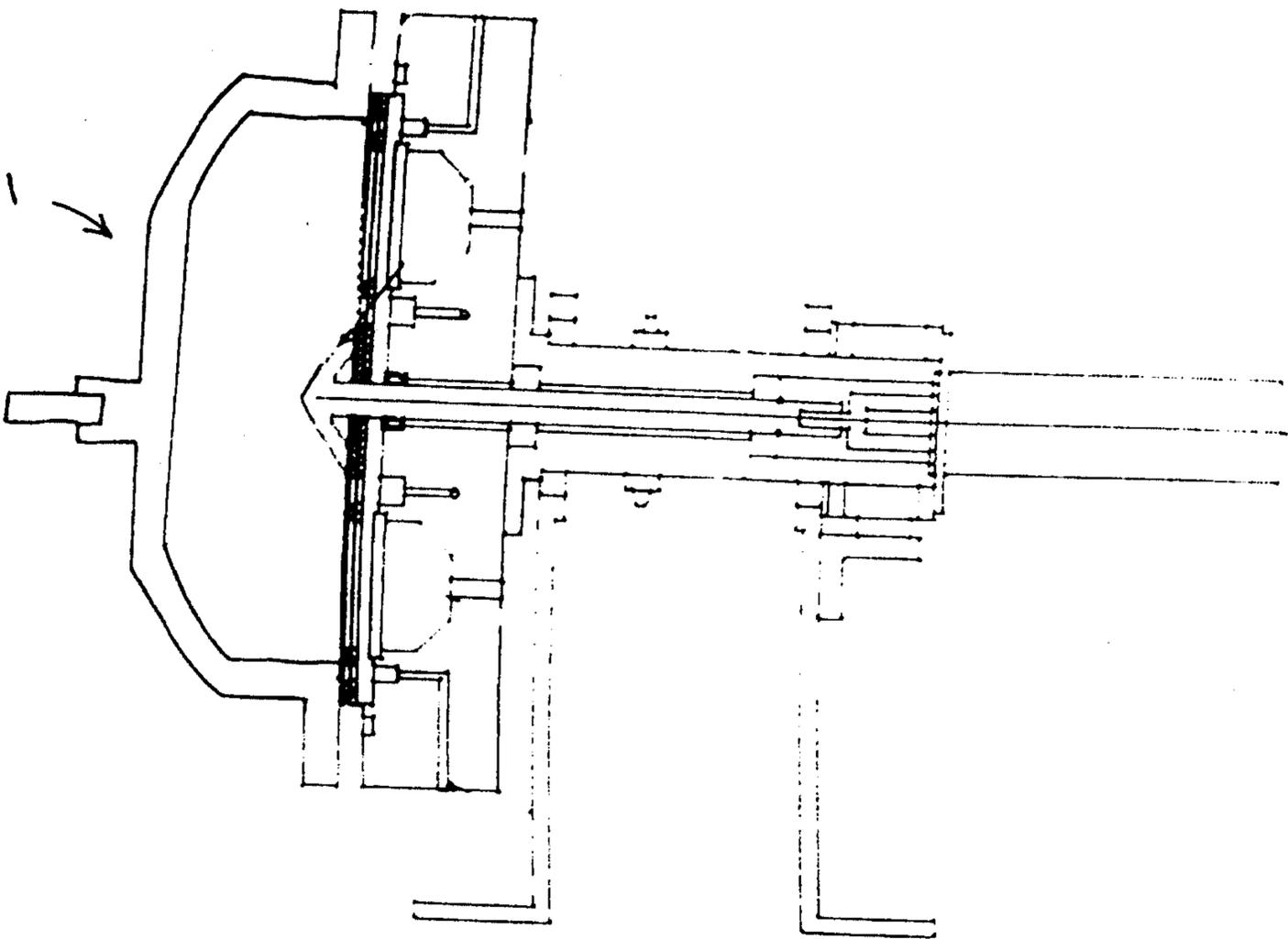


FIG. 3

