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- (71) Applicant (*for all designated States except US*): **NATIONAL RESEARCH COUNCIL OF CANADA** [CA/CA]; Corporate Services, Intellectual Property Services, 1200 Montreal Road, Building M-58, Room EG-12, Ottawa, Ontario K1A 0R6 (CA).
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- (72) Inventors; and
- (75) Inventors/Applicants (*for US only*): **IWAKI, Hiroaki** [JP/CA]; 1625 West De Maisonneuve Boulevard, Suite 1508, Montreal, Québec H3H 2N4 (CA). **HASEGAWA, Yoshie** [JP/JP]; 8-45 Gojo-cho, Higashi-Osaka, Higashi-Osaka, Osaka 579-8042 (JP). **LAU, Peter, C., K.** [CA/CA]; 30 Piper's Crescent, Kirkland, Québec H9H 3J4 (CA).
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(54) Title: CLONING, SEQUENCING AND EXPRESSION OF A COMAMONAS CYCLOPENTANONE 1,2-MONOOXYGENASE-ENCODING GENE IN *ESCHERICHIA COLI*

(57) Abstract: Cyclopentanone 1,2-monooxygenase (CPMO) from *Comamonas* (previously *Pseudomonas*) sp. strain NCIMB 9872 carries out the second step of a degradation pathway that allows the bacterium to use cyclopentanol as a sole carbon source for growth. In the present invention there is reported the localization of the CPMO-encoding gene (*cpnB*) on a 4.3-kb *SphI* fragment, the determination of its sequence. The 550-amino acid CPMO polypeptide (*M_r*, 62,111) encoded by the gene was found to have 36.5% identity with the sequence of cyclohexanone 1,2-monooxygenase (CHMO) of *Acinetobacter* sp. strain NCIMB 9871. The 62-kDa CPMO was expressed in *E. coli* as an IPTG-inducible protein.



WO 02/06452 A2

CLONING, SEQUENCING AND EXPRESSION OF A COMAMONAS
CYCLOPENTANONE 1,2-MONOOXYGENASE-ENCODING GENE IN
ESCHERICHIA COLI

TECHNICAL FIELD

5 The present invention relates to an isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, and expression vector and a transformed cell containing the isolates DNA.

BACKGROUND OF THE INVENTION

10 *Comamonas* (previously *Pseudomonas*) sp. NCIMB 9872 was one of the few microorganisms that have been characterized to produce a Baeyer-Villiger monooxygenase (BVMO; Griffin, M., et al., *Biochem. J.* **129**:595-603, 1972; Griffin, M., et al., *Eur. J. Biochem.* **63**:199-209, 1976; and Willetts, A., *Trends in Biotech.* **15**:55-62, 1997; for a recent review). BVMOs are flavoproteins that
15 mimic the classical Baeyer-Villiger organic chemical reaction which is a peracid-catalyzed oxidation of a ketone to an ester or lactone. The use of enzyme substitutes for the production of lactones in high yield and optical purity is an attractive feature in current trends of research and development toward replacing chemical methods with biological alternatives (Stinson, S.C., *Chem.*
20 *Eng. News*, 83-104, 1998). To date, the best characterized BVMO enzyme is that of cyclohexanone monooxygenase (CHMO) produced by *Acinetobacter* sp. NCIMB 9871 (Stewart, J.D., *Curr. Org. Chem.* **2**:195-216, 1998; Willetts, A., *Trends in Biotech.* **15**:55-62, 1997). This is also the only BVMO whose gene has been cloned and sequenced (Chen, et al., *J. Bacteriol.* **170**:781-789, 1988).
25 Recently, this valuable resource was used to engineer a "designer yeast" in a whole-cell approach to effect a variety of asymmetric Baeyer-Villiger oxidations (Stewart, J.D., et al., *J. Am. Chem. Soc.* **120**:3541-3548, 1998).

 It would be highly desirable to be provided with a new CPMO having an increased enzymatic activity for growing cells in a medium containing
30 cyclopentanol or cyclopentanone as sole carbon source.

SUMMARY OF THE INVENTION

One aim of the present invention is to provide a new CPMO having an increased enzymatic activity for growing cells in a medium containing cyclopentanol or cyclopentanone as sole carbon source.

5 In accordance with the present invention there is provided an isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, the isolated DNA being characterized by the ability to hybridize specifically with the complement of the DNA represented in SEQ ID NO:8 under stringent hybridization conditions.

10 Also in accordance with the present invention, there is provided an isolated DNA, wherein it codes for a cyclopentanone monooxygenase (CPMO), and contains:

- (1) the nucleic acid sequence of SEQ ID NO:8;
- (2) a sequence corresponding to said nucleic acid sequence in
15 the scope of the degeneration of the genetic code; or
- (3) a sequence hybridizing under stringent conditions with the sequence from (1) or (2), and still coding for cyclopentanone monooxygenase (CPMO).

20 Still in accordance with the present invention, there is provided an isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, said isolated DNA having SEQ ID NO:8.

The present invention further provides an isolated DNA expression vector encoding an enzymatically active cyclopentanone monooxygenase (CPMO)
25 comprising a DNA characterized by a sequence as set forth in SEQ ID NO:8, or a portion thereof, said portion encoding said CPMO, in expressible form.

In accordance with the present invention, there is also provided a recombinant vector comprising the isolated DNA as described above, wherein the isolated DNA encodes cyclopentanone monooxygenase.

In a preferred embodiment of the present invention, the isolated DNA has a nucleic acid sequence of SEQ ID NO:8 or which, due to the degeneracy of the genetic code, is a functional equivalent thereof.

Also in accordance with the present invention, there is provided a
5 recombinant vector containing one or more copies of a recombinant DNA described above.

The recombinant vector may be a prokaryotic vector. The recombinant vector may also be a plasmid.

Therefore, in accordance with the present invention, there is also
10 provided a biologically functional plasmid or viral DNA vector, which contains a DNA as described above.

The present invention also provide a host cell comprising a recombinant vector as described above.

Accordingly, there is also provided a cell transformed with a heterologous
15 DNA expression construct encoding an enzymatically active cyclopentanone monooxygenase (CPMO) comprising a DNA characterized by a sequence as set forth in SEQ ID NO:8, or a portion thereof, said portion encoding said CPMO, in expressible form.

The cell may be a prokaryotic cell or it may be *E. coli*.

20 Still in accordance with the present invention, there is also provided a purified cyclopentanone monooxygenase (CPMO) having :

- a) an amino acid sequence as set forth in SEQ ID NO:5;
- b) an amino acid sequence encoded by a nucleic acid sequence as set forth in SEQ ID NO:8; or
- 25 c) an amino acid sequence encoded by a nucleic acid sequence hybridizing to a nucleic acid sequence complementary to the nucleic acid sequence of step b) above under stringent conditions, said amino acid sequence encoded in step c) having a same activity as the amino acid sequence in a).

The present invention also provides a recombinant cyclopentanone monooxygenase (CPMO) having an enzymatic activity superior to the one from a native *Pseudomonas*, and more preferably twice superior.

5 The recombinant cyclopentanone monooxygenase (CPMO) may be prepared from *Comamonas* sp. NCIMB 9872. The recombinant cyclopentanone monooxygenase (CPMO) has preferably a sequence as set forth in SEQ ID NO:5.

10 A method for growing cells *in vitro* in presence of cyclopentanol or cyclopentanone as sole source of carbon, said method comprising the steps of:

- a) transforming a cell with the expression construct described above; and
 - b) growing the cell of step a) under suitable conditions in a medium containing cyclopentanol or cyclopentanone as a sole source of carbon.
- 15

To increase this gene potential, according to the invention it is reported herein the cloning of a cyclopentanone monooxygenase (CPMO)-encoding gene (*cpnB*) from *Comamonas* (*Pseudomonas*) sp. NCIMB 9872, the determination of its DNA and surrounding sequence and expression of CPMO activity and protein in *E. coli*.

20

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 illustrates the first two steps of cyclopentanol degradation by *Pseudomonas* sp. NCIMB 9872;

25 Fig. 2 illustrates the genetic organization in *Comamonas* sp. NCIMB 9872 in the *SphI* fragment containing cyclopentanone monooxygenase-encoding gene (*cpnB*) and additional open reading frames;

Fig. 3 illustrates an alignment of the amino acid sequence of the CPMO of *Comamonas* sp. NCIMB 9872 with that of CHMO from *Acinetobacter* sp.

NCIMB 9871 and a steroid monooxygenase (STMO) from *Rhodococcus rhodochromus*;

Fig. 4 illustrates a SDS-PAGE of crude extracts from *E. coli* (pCMP201); and

5 Fig. 5 illustrates CPMO-encoding gene, designated cpnB.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

Cloning of the *Comamonas* sp. NCIMB 9872 CPMO-encoding gene

Pseudomonas sp. NCIMB 9872 (henceforth strain 9872) identified as a *Comamonas* by 16S rDNA sequencing in this study, was purchased from the
10 National Collections of Industrial and Marine Bacteria Ltd (NCIMB, Aberdeen, Scotland) and grown at 30°C in Luria-Bertani (LB) broth (Sambrook, J., et al., Molecular cloning: a laboratory manual, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N. Y, 1989), or mineral salt medium (MSM), pH 7.0, containing 2 ml of cyclopentanone. The MSM recipe contains per liter: 1.0 g of
15 NH₄NO₃, 1.5 g of KH₂PO₄, 1.5 g of Na₂HPO₄, 0.2 g MgSO₄·7H₂O, 0.01 g of CaCl₂·2H₂O, 0.005 of FeSO₄·7H₂O, 0.002 g of MnSO₄·4H₂O and 0.1 g of yeast extract. Agar was added to 1.5% for plates. Genomic DNA of strain 9872 was prepared by the Marmur method (Marmur, J., *J. Mol. Biol.* **3**: 208-218, 1961). At first, a Southern hybridization of DNA digested with *Bam*HI was carried out
20 using the *Acinetobacter* NCIMB 9871 CHMO-containing gene as probe. Since there was no positive result (hybridization conditions carried out at 65°C) the CPMO protein was purified in order to obtain an N-terminal amino acid sequence. The purification of CPMO protein from cyclopentanone-grown cells was according to Griffin and Turgill (Griffin, M., et al., *Eur. J. Biochem.* **63**:199-
25 209, 1976). Using an automated protein sequencer (Perkin-Elmer model 477) a 40-residue amino-terminal sequence of the purified CPMO was obtained (Fig. 2). This sequence, longer by 11 amino acids, is in perfect agreement with that reported previously from the same organism (Willetts, A., *Trends in Biotech.* **15**:55-62, 1997). Two degenerate oligodeoxynucleotide primers (5'-
30 ACIACIATGA CIACNATGAC-3' (SEQ ID NO:1) and 5'-ARRTGRTAIA

RYTGRTA-3' (SEQ ID NO:2), corresponding to amino acids 2-8 and 35-40, respectively) were synthesized to amplify a 116-bp product from total DNA prepared from strain 9872. The PCR amplification was performed in a Perkin Elmer-Model 2400 Thermal Cycler™ and the amplification conditions were 94°C for 1 min, 50°C for 1 min and 72°C for 1 min for 30 cycles. The amplified product was cloned directly in the pXcmkn12 vector (Cha, J., et al., *Gene* **136**, 369-370, 1993), transformed in *E. coli* JM109 and the resulting plasmid was designated pCMP10. Before using the amplified product as a gene probe its nucleotide sequence was confirmed. Nucleotide sequencing was determined by the Taq DyeDeoxy terminator cycle sequencing kit and the ABI Prism 310 Genetic Analyzer (Perkin Elmer). Plasmid isolation was performed by the method of Birnboim and Doly (Birnboim, H. C., and J. Doly, *DNA. Nucleic Acids Res.* **7**:1513-1523, 1979).

In Fig. 2, Orf1 is most likely a transcriptional activator of the NtrC-type (Morett, E., L. Segovia, *J. Bacteriol.* **175**:6067-6074, 1993). The amino acid sequence of ORF1 (C-terminal 391-amino acids) showed 38-40% identity to equivalent regions of proteins such as NTRC_ECOLI (Nitrogen regulation protein NR(I) from *E. coli*; Miranda, et al., *The complete nucleotide sequence of the glnALG operon of Escherichia coli K12* **15**:2757-2770, 1987), ACOR_ALCEU (Acetoin catabolism regulatory protein from *Ralstonia eutropha*; Kruger, N., et al., *J. Bacteriol.* **179**:4391-4400, 1992). The amino acid sequence of ORF2, showing similarity to enzymes of the short-chain alcohol dehydrogenase family (Jornvall, H., et al., *Biochemistry* **34**: 6003-6013, 1995), is most homologous (45-46% identity) to a putative oxidoreductase CY39.16C of *Mycobacterium tuberculosis* (Swiss Prot sp:Q10855) and fadG3 of *M. tuberculosis* (GenBank accession number Z74025). For *Pseudomonas* sp. strain HI-201 the *lacZ*-Km^r cassette from pKOK6.1 (Kokotek, W., et al., *Gene* **84**: 467-471, 1989) was inserted into *cpnB* at the *Nsi*I site.

In Fig. 2, the following terms are defined as follows: t_{fd}, transcriptional termination sequence of phage fd; Km^r, kanamycin resistance gene, *lacZ*, gene encoding b-galactosidase. Genes and markers are indicated with arrows.

To clone the CPMO-containing gene, the DNA insert from pCMP10 was amplified, labeled by the digoxigenin-11-UTP system according to manufacturer's instructions (Boehringer Mannheim GmbH) and used to probe a Southern hybridization of strain 9872 genomic DNA digested with various restriction enzymes (*Bam*HI, *Eco*RI, *Hind*III, *Kpn*I, *Nhe*I, *Pst*I, *Sal*I, *Sph*I and *Xba*I). As a result, a single hybridizing band of ca 4.3-kb *Sph*I fragment was obtained. Conditions of hybridization were as before. Subsequently, a purified 4.0- to 4.5-kb size fraction of *Sph*I-cut total DNA separated on a 0.8 % agarose gel was ligated to *E. coli* plasmid pUC18, which had been linearized and dephosphorylated. A clone containing the 4.3-kb insert was screened by colony hybridization using the PCR product as a probe; this recombinant plasmid was designated pCMP200.

DNA sequence of the CPMO-encoding gene (*cpnB*) and the flanking region

Nucleotide sequencing of the CPMO-encoding gene was initiated by using a primer designed from the sequence of the PCR product cloned in pCMP10 and further extended using oligonucleotides derived from the new sequence. Both DNA strands of the *Sph*I fragment were sequenced and found to consist of 4281 base pairs (bp). The sequence was analyzed by GENETYX-Mac (Software Development Co., Ltd. Chiba, Japan) and the BLAST program (Altschul, S. F., et al., *Nucleic Acids Res.* **25**:3389-3402, 1997). As a result three open reading frames (ORFs) arranged in the same direction were predicted (Fig. 2). The nucleotide sequence of the 1650-bp ORF encoding CPMO is preceded by a partial ORF1 (1173-bp) coding for the C-terminus of an NtrC-type transcriptional activator (Miranda, et al., *The complete nucleotide sequence of the glnALG operon of Escherichia coli K12* **15**:2757-2770, 1987) and by a complete ORF2 (750-bp) coding for a homolog of the short-chain dehydrogenases/reductases (Jornvall, H., et al., *Biochemistry* **34**: 6003-6013, 1995). The two intergenic regions are 244-bp and 32-bp, respectively. The CPMO-encoding gene is referred to *cpnB* (cyclopentanone and *B* designates the second step of the degradation pathway, see Fig. 1) hereafter. In Fig. 5, the CPMO-encoding gene starts at nucleotide position 1822 and ends 3471 that does not include the stop codon. Accordingly, the boundary of *cpnA* is 3507-

4256. The partial open reading frame preceding *cpnB* is from 1-1174.

Fig. 1 has been adapted from Griffin, M., et al. (Griffin, M., et al., *Biochem. J.* **129**:595-603, 1972). The designated genes are: *cpnA* encoding cyclopentanol dehydrogenase; *cpnB* encoding cyclopentanone 1,2-
5 monooxygenase (CPMO). An alternative name for 5-valerolactone is 5-pentanolide. Subsequent reaction steps are the formation of 5-hydroxyvalerate, 5-oxovalerate, glutarate and finally acetyl CoA.

The amino acid sequence of the CPMO enzyme consists of 550 residues (Fig. 3). This sequence shows 36.5% identity and an additional 13.6% amino
10 acid similarity with the 543-residue CHMO of *Acinetobacter* sp. strain NCIMB 9871. An equally related protein (549 amino acids; 37.3% identity and 12.4% similarity) is the putative steroid monooxygenase (STMO) of *Rhodococcus rhodochrous* (Morii, S., et al., GenBank accession number AB010439, 1998). The latter enzyme carries out the oxidation of progesterone to produce
15 testosterone acetate. A CLUSTAL alignment of these three sequences gave 24.6% positional identity (Fig. 3).

In Fig. 3, asterisks indicate identical amino acids, dots indicated similar amino acids and dashes indicate gaps introduced to maximize the alignment. The amino-terminal peptide sequence confirmed by Edman degradation is
20 underlined. The locations of the consensus FAD fingerprint sequences as described by Eppink et al. (Eppink, et al., *Prot. Sci.* **6**:2454-2458, 1997) are as indicated. The conserved GD motif found in flavoprotein hydroxylases as a second FAD fingerprint is also indicated. Not shown is the DG motif of flavoprotein hydroxylases which has the sequence of chhhssDGxcSxhR. Lower
25 case letters identify certain residues types: h, hydrophobic residues, s, small residues, c, charged residues, and x, any residues. Note that a DG doublet is present in CPMO and STMO sequence.

A notable sequence motif present in CPMO and related proteins is the FAD-binding fingerprint (GXGXXG) that is similar to those found in flavoprotein
30 hydroxylases (Eppink, et al., *Prot. Sci.* **6**:2454-2458, 1997). Flavoprotein hydroxylases (eg. phenol hydroxylase, the structure is now known; Enroth, C.,

et al., *Structure* **6**:605-617, 1998) are monooxygenases that catalyze the insertion of one atom of molecular oxygen into the substrate using NAD(P)H as electron donor. These proteins possess a conserved "Asp-Gly (DG)" motif for both FAD and NAD(P)H binding in between two fingerprint motifs for the FAD binding (fingerprint 1: GXGXXG; fingerprint 2: Gly-Asp [GD] motif). Sequence motifs in CPMO, STMO and CHMO differ from those in flavoprotein hydroxylases by having a repeated GXGXXG motif (amino acids 24 to 33 and 193 to 202 in CPMO numbering). The possibility that the second FAD fingerprint in CPMO and related proteins fulfils a dual role of FAD and NADPH binding awaits structural determination of a representative member of this family of proteins. It is reasonable to assume that a different mechanism in catalysis is reflected in the motifs seen in the two classes of proteins.

Expression of *cpnB* gene in *E. coli*

Two primers of the following sequence were synthesized to amplify the *cpnB* gene and the resultant 1.7-kb DNA fragment was cloned in the pSD80 plasmid to yield pCMP201. Plasmid pSD80 is a third generation derivative of the commercially available pKK223-3 vector (Pharmacia) that contains a *tac* promoter upstream of the multiple cloning site (MCS), an *unc* terminator sequence downstream of the MCS, and *lacI^q* elsewhere on the plasmid (Smith, S.P., et al., *Biochemistry* **35**:8805-8814, 1996). The primers were: 5'-AAAAGGCCTG AACTTCAATT ATTTAGGAGA C-3' (SEQ ID NO:3) and 5'-AAAACTGCAG GAGTTGCACA ACAGAGTCTT AG-3' with built-in *StuI* and *PstI* restriction sites (underlined), respectively, to facilitate cloning at the compatible sites (*SmaI* and *PstI*) of the pSD80 vector. Vent DNA polymerase (New England BioLabs, Beverly, MA) was used and the amplification conditions were 94°C for 1 min, 50°C for 1 min, and 72°C for 1 min, for 30 cycles. The amplified DNA fragment was purified from an agarose gel and digested with *StuI* and *PstI*. One of the resulting recombinant plasmids was designated pCMP201. By DNA sequencing it was established that no mutation had been introduced in the *cpnB* gene during PCR amplification.

Fig. 4 shows the production of a 60-kDa protein in a Coomassie blue-stained SDS-polyacrylamide gel of the crude protein extract prepared from *E. coli* JM109 (pCMP201) cells that were induced by 0.1 mM isopropyl-beta-D-thiogalactopyranoside (IPTG). The cells were induced at an absorbance (A₆₀₀) of 0.4 to 0.5 and the induction period was up to 4 hr. The observed molecular mass was in agreement with the predicted size of the 62-kDa CPMO. In the absence of IPTG, this protein band was not produced. Also, the CPMO enzyme activity was observed only in those cells grown in the presence of IPTG. CPMO activity was assayed at 25°C by measuring a decrease in absorbance at 340 nm in 50 mM phosphate buffer (pH 7.8) containing 1 µmol of cyclopentanone, 0.2 µmol of NADPH, and the crude enzyme extract prepared from *E. coli* JM109 (pCMP201). These cells were cultivated in 100 ml of LB medium containing 100 µg/ml of ampicillin at 25°C. The IPTG-induced cells were harvested by centrifugation, washed in 50 mM phosphate buffer (pH 7.2), resuspended in 1/20 volume of same buffer, and sonicated by four-20 sec bursts with a Braun-Sonifier™ 250 apparatus. After centrifugation for 30 min at 18,000 x g and at 4°C, the supernatant was used for determination of enzyme activity. One unit (U) of activity is defined as the amount of enzyme required to convert 1 µmol of substrate in 1 min. Protein concentration was determined by the method of Bradford (Bradford, M. M., *Anal. Biochem.* **72**: 248-254, 1976). As a result the specific activity of the CPMO enzyme was found to be 0.28 U/mg. The specific activity of CPMO in the native *Pseudomonas* was reported to be 0.11 U/mg (Griffin, M., et al., *Biochem. J.* **129**:595-603, 1972).

In Fig. 4, lane 1 has been loaded with extracts of IPTG-induced *E. coli* and lane 2 has been loaded with extracts of *E. coli* in absence of IPTG. M means molecular weight markers as indicated in kilo daltons. The arrow indicates the production of the desired 60-kDa protein.

Inactivation of *cpnB* gene

Pseudomonas sp. strain HI-201 was constructed by chromosomal inactivation of the *cpnB* gene using a *lacZ*-Km^r cassette from the mobilizable pKOK6.1 vector (Kokotek, W., et al., *Gene* **84**: 467-471, 1989). In pKOK6.1 the

lacZ gene is promoterless and in addition to Km^r it is ampicillin resistant (Ap^r). The *lacZ*-Km^r cassette was excised as a *Pst*I-fragment and inserted into the *Nsi*I site within the *cpnB* gene in pCMP200, yielding pCMP202. Electroporation of this plasmid into 9872 cells was carried out in the Gene Pulser™ (BioRads) and the parameters of electroporation were 2.5 kV, 25 uF and 200 ohm. The cells were initially washed with 1mM HEPES buffer and resuspended in 1mM HEPES containing 10% glycerol. Km^r colonies were selected on LB plates containing Km (250 µg/ml). To select for double crossover mutants, a second screening on LB plates containing Ap (300 µg /ml) was carried out. The inactivation of *cpnB* (Fig. 2), was confirmed by PCR. The resulting mutant HI-201 was found not to be able to grow on cyclopentanol or cyclopentanone as a sole carbon and energy source. This result indicated that *cpnB* is essential for the degradation of cyclopentanol and it appeared that there was only one copy of the *cpnB* gene in strain 9872.

As expected of a flavoprotein the amino acid sequence of CPMO contains motifs of FAD fingerprints similar to those found in flavoprotein hydroxylases.

Nucleotide sequence accession number

The DNA sequence of the 4,281-bp *Sph*I fragment has been submitted to DDBJ and assigned accession number AB022102. The release of this data awaits the inventors' authorization.

While the invention has been described in connection with specific embodiments thereof, it will be understood that it is capable of further modifications and this application is intended to cover any variations, uses, or adaptations of the invention following, in general, the principles of the invention and including such departures from the present disclosure as come within known or customary practice within the art to which the invention pertains and as may be applied to the essential features hereinbefore set forth, and as follows in the scope of the appended claims.

WHAT IS CLAIMED IS:

1. An isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, the isolated DNA being characterized by the ability to hybridize specifically with the complement of the DNA represented in SEQ ID NO:8 under stringent hybridization conditions.
2. An isolated DNA, wherein it codes for a cyclopentanone monooxygenase (CPMO), and contains:
 - (a) the nucleic acid sequence of SEQ ID NO:8;
 - (b) a sequence corresponding to said nucleic acid sequence in the scope of the degeneration of the genetic code; or
 - (c) a sequence hybridizing under stringent conditions with the sequence from (1) or (2), and still coding for cyclopentanone monooxygenase (CPMO).
3. An isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, said isolated DNA having SEQ ID NO:8.
4. An isolated DNA expression vector encoding an enzymatically active cyclopentanone monooxygenase (CPMO) comprising a DNA characterized by a sequence as set forth in SEQ ID NO:8, or a portion thereof, said portion encoding said CPMO, in expressible form.
5. A recombinant vector comprising the isolated DNA of any one of claims 1 to 3, wherein the isolated DNA encodes cyclopentanone monooxygenase.
6. The recombinant vector of claim 5, wherein the isolated DNA has a nucleic acid sequence of SEQ ID NO:8 or which, due to the degeneracy of the genetic code, is a functional equivalent thereof.
7. A recombinant vector, wherein it contains one or more copies of a recombinant DNA according to claim 1, 2 or 3.
8. A recombinant vector according to claim 4, 5, 6 or 7, wherein it is a prokaryotic vector.

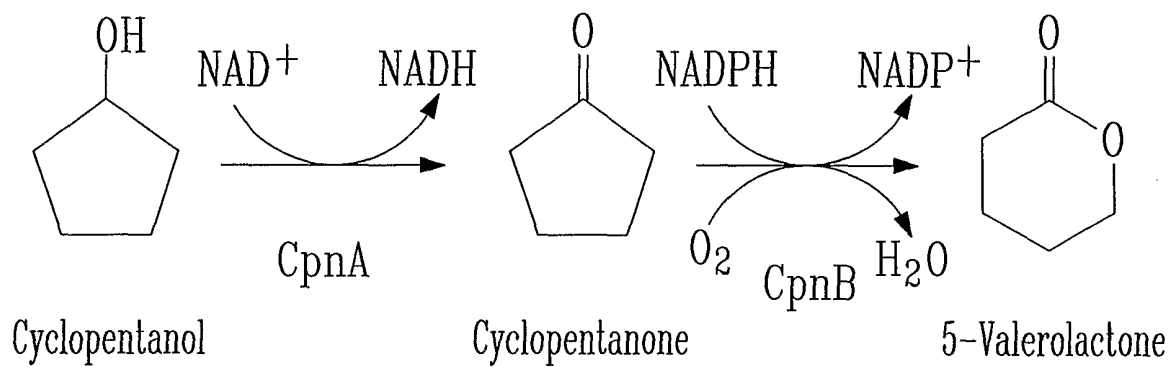
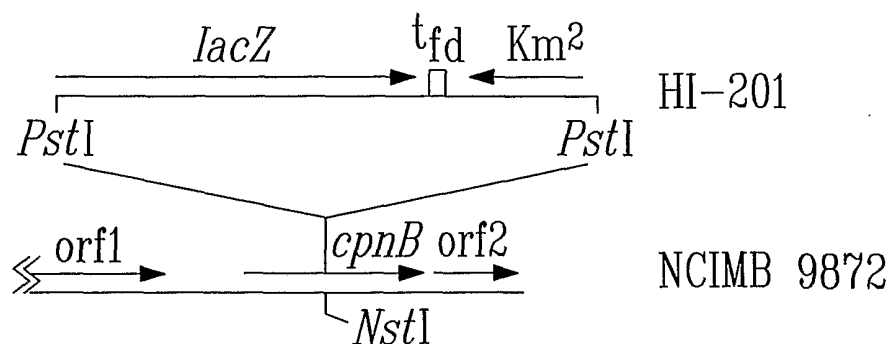
9. A recombinant vector according to claim 4, 5, 6 or 7, wherein it is a plasmid.
10. Biologically functional plasmid or viral DNA vector, which contains a DNA as defined in claim 1, 2 or 3.
11. A host cell comprising a recombinant vector of any one of claims 4 to 9.
12. A cell transformed with a heterologous DNA expression construct encoding an enzymatically active cyclopentanone monooxygenase (CPMO) comprising a DNA characterized by a sequence as set forth in SEQ ID NO:8, or a portion thereof, said portion encoding said CPMO, in expressible form.
13. A cell as defined in claim 12, which is a prokaryotic cell.
14. A cell as defined in claim 12, which is *E. coli*.
15. A purified cyclopentanone monooxygenase (CPMO) having :
 - (a) an amino acid sequence as set forth in SEQ ID NO:5;
 - (b) an amino acid sequence encoded by a nucleic acid sequence as set forth in SEQ ID NO:8; or
 - (c) an amino acid sequence encoded by a nucleic acid sequence hybridizing to a nucleic acid sequence complementary to the nucleic acid sequence of step b) above under stringent conditions, said amino acid sequence encoded in step c) having a same activity as the amino acid sequence in a).
16. A recombinant cyclopentanone monooxygenase (CPMO) having an enzymatic activity superior to the one naturally occurring.
17. The recombinant cyclopentanone monooxygenase (CPMO) of claim 16, wherein said CPMO is prepared from *Comamonas* sp. NCIMB 9872.
18. The recombinant cyclopentanone monooxygenase (CPMO) of claim 16, wherein said CPMO has a sequence as set forth in SEQ ID NO:5.

19. The recombinant cyclopentanone monooxygenase (CPMO) of claim 16, wherein said CPMO has an enzymatic activity twice superior to that of a CPMO from a native *Pseudomonas*.

20. A method for growing cells in vitro in presence of cyclopentanol or cyclopentanone as sole source of carbon, said method comprising the steps of:

- (a) transforming a cell with the expression construct of claim 4, 5, 6, 7, 8 or 9; and
- (b) growing the cell of step a) under suitable conditions in a medium containing cyclopentanol or cyclopentanone as a sole source of carbon.

1/10

FIG. 1FIG. 2

2/10

GxGxxG

CHMO 1:-----MSQKMDFDAIVIGGGFGGLYAVKKLRDELELVQAFDKATDVAGT 45

STMO 1:MNGQHPRSVVTAPDATTGTTSDVVVVGAGIAGLYAIHRFRSQ-GLTVRAFEAASGVGGV 59

CPMO 1:MTTFTMTTEQLGMNNSVNDKLDVLLIGAGFTGLYQLYHLRKL-GYKVHLVDAGADIGGI 59

. * ...* *. *** . ..* * . . . *

CHMO 46:WYWNRYPGALTDTEHLYCYSWDKELLQSLEIKKKYVQGPDVVKYLQQVAEKHDLKKSYQ 105

STMO 60:WYWNRYPGARCDVESIDYSYSFSPELEQEWNNSEKYATQPEILAYLEHVADRFDLRRDIR 119

CPMO 60:WHWNCYPGARVDTHCQIYQYSI-PELWQEFNWKELFPNWAQMREYFHFADKKLDLSKDIS 118

* . ** ***** * * * * * * . . . ** .

CHMO 106:FNTAVQSAHYNEADALWEVTTEYGDKYTARFLITALGLLSAPNLPNIKGINQFKGELHHT 165

STMO 120:FDTRVTSAVLDEEGLRWTVRTDRGDEV SARFLVVAAGPLSNANTPAFDGLDRFTGDIVHT 179

CPMO 119:FNTRVQSAVFDEGTREWTVRSIGHQPIQARFVIANLGFGASPSTPNVDGIETFKGQWYHT 178

* . * * * * . * * * . . . *** . . * . * * . * * . * *

GxGxxG

CHMO 166:SRWPDD-VSFEGRVGVIGTGSTGVQVITAVAPLAKHLTVFQRSAQYSVPIGNDPLSEED 224

STMO 180:ARWPHDGVDFGTGKRVGVIGTGSSGIQSIPIIAEQAEQLFVFQRSANY SIPAGNVPLDDAT 239

CPMO 179:ALWPQEGVNMAGKRVAIIGTGSSGVQVAQEAALDAKQVTVYQRTPNLALPMHQQLSAED 238

. ** * . ***** .*****.*.* * *...*.***. . .*. * . *

CHMO 225:VKKIKDNYDKSLGWCNMNSALAFALNESTVPAMSVSAEERKAVFEKAWQTGGGFRFMFETF 284

STMO 240:RAEQKANYAERRRLSRESGGSPHRPHPKSALEVSEEERRAVYEERWKLGG--VLFSKAF 297

CPMO 239:NLRMKPELPAAFERRGKCFAGFDFDFIAKNATELSAAERTEILEELWNAGG-FRYWLANF 297

* . . * . * * * . * . * * * *

FILE 3A

3/10

CHMO 285:GDIATNMEANIEAQNFIFKIAEIVKDPAPAIQKLMPQD----LYAKRPLCDSGYYNTFNR 340

STMO 298:PDQLTDPAAANDTARAFWEEKIRAVVDDPAVAELLTPKDH--AIGAKRIVTDSGYYETYNR 355

CPMO 298:QDYLFDDKANDYVVEFWRDKVRARIKDPKVAEKLAPMKKPHPYGAKRPSLEQWYYEIFNQ 357

* . ** * * . ** . * * *** . ** . *

CHMO 341.DNVRLEDVKANPIVEITENGVKLENGDFVELDMLICATGFDVAVDGNVVRMDIQKNGLAM 400

STMO 356:DNVELVDLRSTPIVGMDETGIIVT-TGAHYDLDMIVLATGFDAMTGSCLKLEIVGRGGRTL 414

CPMO 358:NNVTLVVDVNETPVLRITEKGIVT-AEGEAEFDLIVFATGFDVAVTGGITSIDFRNNQGQSF 416

. ** * * . * . . * * . . * *

CHMO 401:KDYWKEGPSSYMGVTVNNYPNMFVVLGPNGP--FTNLPPSIESQVEWISDTIQYTVENNV 458

STMO 415:KETWAAGPRTYLGLGIDGFPNFFNLTGPGSPSVLANMVLHSELHVDWVADAIAYLDARGA 474

CPMO 417:KDVWSDGIRTQLGVATAGFPNLLFGYGPQSPAGFCNGPSSAEYQGDLLIQLMNYLRDNNI 476

* . * * . . * *

CHMO 459:ESIEATKEAEEQWTQTCANIAEMTLFPKAQSWIFGANIPGKKNTVYFYLGLKEYRTCAS 518

STMO 475:AGIEGTPEAVADWVEECRNRAEASLLNSANSWYLGANIPGRPRVFMFPLGGFGVYREIIT 534

CPMO 477:SRIEAQSEAQEEWSKLIADFWDSSLFPRAKSWYQGSNIPGKKVESLNFPLGLPTYISKFN 536

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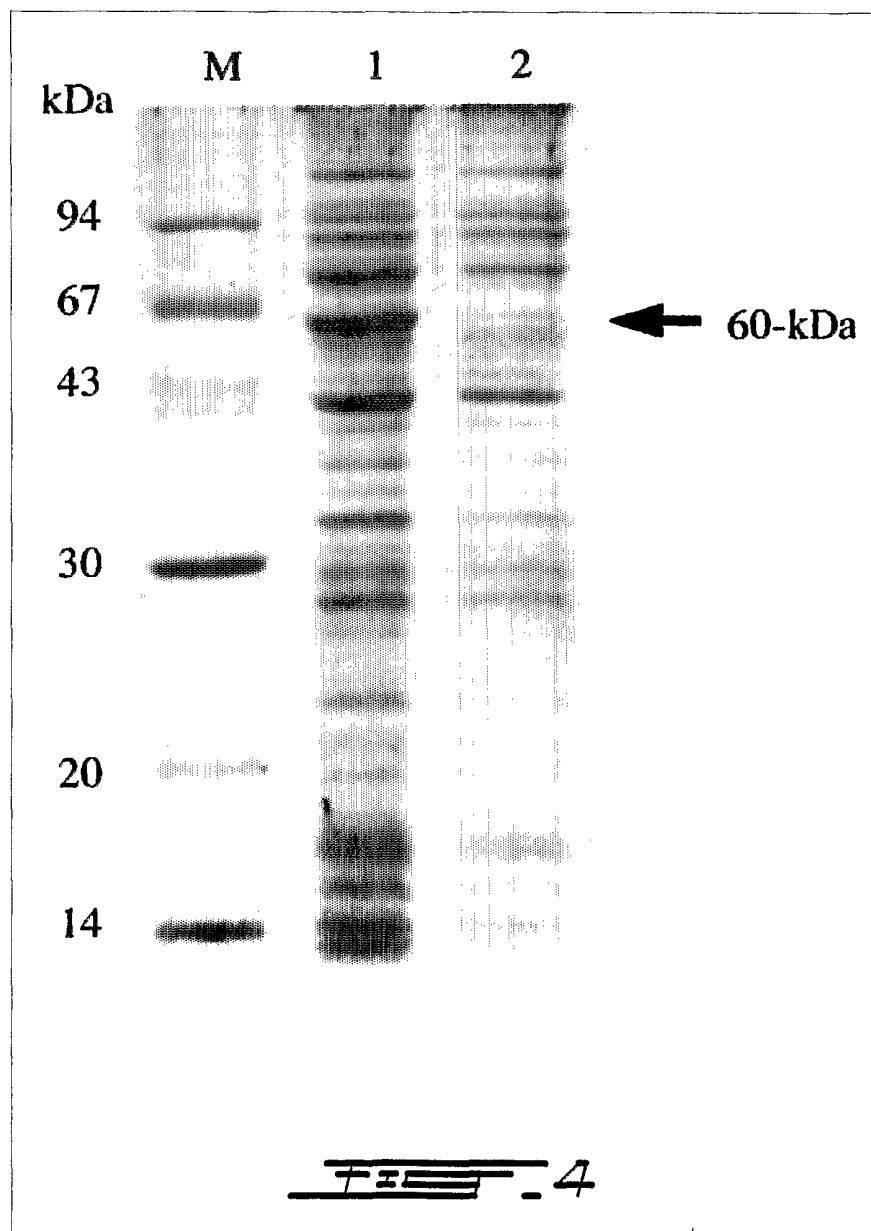
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CPMO 537:ESAEGYAGFSLAS----- 550 SEQ ID NO:5

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FILE - 3B

4/10



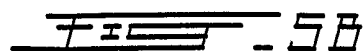
5/10

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61	CACCGACATCACATCGCCGGTCACCGGCGAGAACATTGGCAAGATCCTGCGCTTTTCGCC					
	GTGGCTGTAGTGTAGCGGCCAGTGGCCGCTCTTGTAACCGTTCTAGGACGCGAAAAGCGG					
121	GCTGAAGCCGCAGGGCTACGAGAGAATTTTTCAGTCCGGCCTGGCGGACTCGACCCAGCA					
	CGACTTCGGCGTCCCGATGCTCTCTTAAAAAGTCAGGCCGGACCGCTGAGCTGGGTGCT					
181	CACCCCCAGGATGGTGCCTTTTCGGACAGTTCCTGTCCTGTGATCCAGAGACCATCAACAC					
	GTGGGGGTCTACCACGCAAAGCCTGTCAAGGACAGGACACTAGGTCTCTGGTAGTTGTG					
241	CCTGGAGACATTGGAGCGGATCGCGCGCGCCGACGTCAATGTCTTGCTGCATGGAGAGAC					
	GGACCTCTGTAACCTCGCCTAGCGCGCGGGCTGCAGTTACAGAACGACGTACCTCTCTG					
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	CCCGTGCCCGTTTCTCGACTATCGAGCGGTGTAAGTACAGCGCTCGGCGGGCGCCCTACG					
361	GCCCTACCTCGCCATCAACTGCGGGGCAATCAGCTCGGAGTTGCTGGAGAGTACTTTTTT					
	CGGGATGGAGCGGTAGTTGACGCCCCGTTAGTCGAGCCTCAACGACCTCTCATGAAAAA					
421	TGGCTATGTGCGCGGAGCATTCTCCGGGGCAGATCCCAAGGGCCGCGCCGGCTACTTTGA					
	ACCGATACACGCGCCTCGTAAGAGGCCCCGTCTAGGGTTCCCGGCGCGGCCGATGAACT					
481	ATCGGTGGGCGAAGGCACCTTGTTTCTGGACGAGATCGGCGAGCTGCCCTTGGCCATGCA					
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601	GCAGCGCGCCTCGTGCCGCATTATCGCGGCCACGCACCGCAATCTGGAGGAATCATCGC					
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661	ACAGGGCCTGTTTCGCCAGGATCTCTACTACCGTCTCAAGATCGTTCAGAAAAGGCTCAA					
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FIG. 5A

6/10

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7/10

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2161 AGCAAGGACATCAGCTTCAACACCCGTGTGCAGTCGGCCGTCTTTGACGAAGGCACACGC
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8/10

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9/10

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10/10

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SEQUENCE LISTING

<110> NATIONAL RESEARCH COUNCIL OF CANADA
IWAKI, Hiroaki
HASEGAWA, Yoshie
LAU, Peter C.K.

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Comamonas Cyclopentanone 1,2-Monooxygenase-Encoding Gene in
Escherichia coli

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Leu Val Asp Ala Gly Ala Asp Ile Gly Gly Ile Trp His Trp Asn Cys
50 55 60
Tyr Pro Gly Ala Arg Val Asp Thr His Cys Gln Ile Tyr Gln Tyr Ser
65 70 75 80
Ile Pro Glu Leu Trp Gln Glu Phe Asn Trp Lys Glu Leu Phe Pro Asn
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Leu Ser Lys Asp Ile Ser Phe Asn Thr Arg Val Gln Ser Ala Val Phe
115 120 125
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145 150 155 160
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165 170 175
His Thr Ala Leu Trp Pro Gln Glu Gly Val Asn Met Ala Gly Lys Arg
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210 215 220

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Arg Ile Lys Asp Pro Lys Val Ala Glu Lys Leu Ala Pro Met Lys Lys
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Pro His Pro Tyr Gly Ala Lys Arg Pro Ser Leu Glu Gln Trp Tyr Tyr
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Glu Ile Phe Asn Gln Asn Asn Val Thr Leu Val Asp Val Asn Glu Thr
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 Asp Leu Arg Arg Asp Ile Arg Phe Asp Thr Arg Val Thr Ser Ala Val
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 Leu Asp Glu Glu Gly Leu Arg Trp Thr Val Arg Thr Asp Arg Gly Asp
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 Glu Val Ser Ala Arg Phe Leu Val Val Ala Ala Gly Pro Leu Ser Asn
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 Ala Asn Thr Pro Ala Phe Asp Gly Leu Asp Arg Phe Thr Gly Asp Ile
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 Val His Thr Ala Arg Trp Pro His Asp Gly Val Asp Phe Thr Gly Lys
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 Arg Val Gly Val Ile Gly Thr Gly Ser Ser Gly Ile Gln Ser Ile Pro
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 Ile Ile Ala Glu Gln Ala Glu Gln Leu Phe Val Phe Gln Arg Ser Ala
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 Asn Tyr Ser Ile Pro Ala Gly Asn Val Pro Leu Asp Asp Ala Thr Arg
 225 230 235 240
 Ala Glu Gln Lys Ala Asn Tyr Ala Glu Arg Arg Arg Leu Ser Arg Glu
 245 250 255
 Ser Gly Gly Gly Ser Pro His Arg Pro His Pro Lys Ser Ala Leu Glu
 260 265 270
 Val Ser Glu Glu Glu Arg Arg Ala Val Tyr Glu Glu Arg Trp Lys Leu
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 Gly Gly Val Leu Phe Ser Lys Ala Phe Pro Asp Gln Leu Thr Asp Pro
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 Ala Ala Asn Asp Thr Ala Arg Ala Phe Trp Glu Glu Lys Ile Arg Ala
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 Val Val Asp Asp Pro Ala Val Ala Glu Leu Leu Thr Pro Lys Asp His
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 Tyr Asn Arg Asp Asn Val Glu Leu Val Asp Leu Arg Ser Thr Pro Ile
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 Val Gly Met Asp Glu Thr Gly Ile Val Thr Thr Gly Ala His Tyr Asp
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 Leu Asp Met Ile Val Leu Ala Thr Gly Phe Asp Ala Met Thr Gly Ser
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 Thr Trp Ala Ala Gly Pro Arg Thr Tyr Leu Gly Leu Gly Ile Asp Gly
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 Pro Gly Arg Pro Arg Val Phe Met Pro Phe Leu Gly Gly Phe Gly Val
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Tyr Val Gln Gly Pro Asp Val Arg Lys Tyr Leu Gln Gln Val Ala Glu
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Lys His Asp Leu Lys Lys Ser Tyr Gln Phe Asn Thr Ala Val Gln Ser
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Ala His Tyr Asn Glu Ala Asp Ala Leu Trp Glu Val Thr Thr Glu Tyr
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Ser Ala Pro Asn Leu Pro Asn Ile Lys Gly Ile Asn Gln Phe Lys Gly
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Glu Leu His His Thr Ser Arg Trp Pro Asp Asp Val Ser Phe Glu Gly
      165           170           175
Lys Arg Val Gly Val Ile Gly Thr Gly Ser Thr Gly Val Gln Val Ile
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Thr Ala Val Ala Pro Leu Ala Lys His Leu Thr Val Phe Gln Arg Ser
      195           200           205
Ala Gln Tyr Ser Val Pro Ile Gly Asn Asp Pro Leu Ser Glu Glu Asp
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Val Lys Lys Ile Lys Asp Asn Tyr Asp Lys Ser Leu Gly Trp Cys Met
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Ser Val Ser Ala Glu Glu Arg Lys Ala Val Phe Glu Lys Ala Trp Gln
      260           265           270
Thr Gly Gly Gly Phe Arg Phe Met Phe Glu Thr Phe Gly Asp Ile Ala
      275           280           285
Thr Asn Met Glu Ala Asn Ile Glu Ala Gln Asn Phe Ile Lys Gly Lys
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Gln Asp Leu Tyr Ala Lys Arg Pro Leu Cys Asp Ser Gly Tyr Tyr Asn
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Thr Phe Asn Arg Asp Asn Val Arg Leu Glu Asp Val Lys Ala Asn Pro
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Val Glu Leu Asp Met Leu Ile Cys Ala Thr Gly Phe Asp Ala Val Asp
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Lys Asp Tyr Trp Lys Glu Gly Pro Ser Ser Tyr Met Gly Val Thr Val
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Lys Glu Ala Glu Glu Gln Trp Thr Gln Thr Cys Ala Asn Ile Ala Glu
465      470      475      480
Met Thr Leu Phe Pro Lys Ala Gln Ser Trp Ile Phe Gly Ala Asn Ile
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Pro Gly Lys Lys Asn Thr Val Tyr Phe Tyr Leu Gly Gly Leu Lys Glu
      500      505      510
Tyr Arg Thr Cys Ala Ser Asn Cys Lys Asn His Ala Tyr Glu Gly Phe
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Ile Gly Ala Gly Phe Thr Gly Leu Tyr Gln Leu Tyr His Leu Arg Lys	
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Leu Gly Tyr Lys Val His Leu Val Asp Ala Gly Ala Asp Ile Gly Gly	
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Ile Trp His Trp Asn Cys Tyr Pro Gly Ala Arg Val Asp Thr His Cys	
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Lys Glu Leu Phe Pro Asn Trp Ala Gln Met Arg Glu Tyr Phe His Phe	
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Lys Asn Ala Thr Glu Leu Ser Ala Ala Glu Arg Thr Glu Ile Leu Glu	
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