



(11) **EP 1 338 198 B1**

(12) **EUROPEAN PATENT SPECIFICATION**

(45) Date of publication and mention
of the grant of the patent:
28.11.2007 Bulletin 2007/48

(51) Int Cl.:
A01K 67/027 ^(2006.01) **G01N 33/50** ^(2006.01)
G01N 33/15 ^(2006.01) **A61K 39/395** ^(2006.01)
C12N 15/12 ^(2006.01) **C12N 5/16** ^(2006.01)

(21) Application number: **01978918.9**

(86) International application number:
PCT/JP2001/009401

(22) Date of filing: **25.10.2001**

(87) International publication number:
WO 2002/043477 (06.06.2002 Gazette 2002/23)

(54) **METHOD OF CONSTRUCTING MOUSE SUITABLE FOR THE TAKE, DIFFERENTIATION AND PROLIFERATION OF HETEROGENOUS CELLS, MOUSE CONSTRUCTED BY THIS METHOD AND USE OF THE MOUSE**

VERFAHREN ZUR HERSTELLUNG EINER MAUS, DIE FÜR DIE AUFNAHME, DIFFERENZIERUNG UND PROLIFERATION VON HETEROGENEN ZELLEN GEEIGNET IST, DIE HIERDURCH HERGESTELLTE MAUS UND IHRE VERWENDUNG

METHODE D'OBTENTION D'UNE SOURIS ADAPTEE A LA PRISE, A LA DIFFERENCIATION ET A LA PROLIFERATION DE CELLULES HETEROGENES, SOURIS OBTENUE SELON CETTE METHODE ET SON UTILISATION

(84) Designated Contracting States:
**AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU
MC NL PT SE TR**

(30) Priority: **01.12.2000 JP 2000367296**

(43) Date of publication of application:
27.08.2003 Bulletin 2003/35

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Description

TECHNICAL FIELD

5 **[0001]** The present invention relates to a method of producing an excellent mouse for engraftment of heterologous cells, a mouse produced by this method and use of the mouse.

BACKGROUND ART

10 **[0002]** Laboratory animals to which heterologous cells including human cells are engrafted are very important for analysis of onset mechanisms of various diseases and drug developments for the treatments or preventions thereof, and development of animals as receptors therefor is one of major themes in laboratory animal sciences. In particular, in recent years, treatments etc. (known as regenerative medicine) in which tissues or cells differentiated from stem cells are transplanted have received world-wide attention, and therefore these animals are of increasing importance.

15 **[0003]** The inventors of the present invention have continued to develop and improve these laboratory animals. In particular, they made improvements or the like on a nude mouse or a SCID mouse, and they have already filed a patent application (Japanese Patent Application Laying-Open (kokai) No. 9-94040) concerning an immunodeficient mouse etc. produced for this purpose. Above all, an NOD/Shi-scid mouse and an NOD/LtSz-scid mouse which exhibit multifunctional immunodeficiency (functional deficiency of T cells and B cells, decline of macrophage function, reduction of complement activity, reduction of natural killer (NK) activity etc.) are the most noteworthy as laboratory animals suitable for engraftment of heterologous cells. Since it became clear that they could be used for various types of research including stem cell differentiation and proliferation, the range of applications in which they are used has increased to the present level.

20 **[0004]** However, human cells are engrafted to the NOD/Shi-scid mouse at a high ratio, but it is recognized that the engraftment capacity is substantially varied.

25 **[0005]** In order to enhance the engraftment capacity of the NOD/Shi-scid mouse, it has already been revealed that reduction of NK activity in the mouse by administering anti IL-2R β chain antibodies (TM β 1), anti-asialo-GM1 antibodies or the like is important. (Koyanagi, Y. et al., 1997. "Primary human immunodeficiency virus type 1 viremia and central nervous system invasion in a novel hu-PBL-immunodeficient mouse strain." J Virol 71:2417; Koyanagi, Y. et al., 1997. "High levels of viremia in hu-PBL-NOD-scid mice with HIV-1 infection." Leukemia 11 Suppl. 3:109; Yoshino H, et al., 30 2000. "Natural killer cell depletion by anti-asialo GM1 antiserum treatment enhances human hematopoietic stem cell engraftment in NOD/Shi-scid mice." Bone Marrow Transplant 26:1211-6. However, these antibodies are very expensive, and it is recognized that their efficacies vary between individuals. Further, when anti-asialo GM1 antibodies are used, the administration thereof should be conducted with the frequency of every eleventh day during the experiment period, and thus a degree of complexity is attached.

35 **[0006]** Therefore, Dr.Shultz,L.D. et al. of The Jackson Laboratory in the United States produced an NOD/LtSz-scid, β 2m null(β 2m (null) NOD/SCID)mouse (Kollet O, Peled A, Byk T et al., beta2 microglobulin-deficient (β 2m(null))NOD/SCID mice are excellent recipients for studying human stem cell function. Blood 2000;95(10):3102-5) by crossing an NOD/LtSz-scid mouse having high engraftment capacity of human cells with a β 2m KO mouse from which NK activity has been depleted.

40 **[0007]** With respect to the NOD/LtSz-scid, β 2m null mouse, T cells, B cells and natural killer (NK) cells are depleted, and the function of macrophages and complements is reduced. However, other cells (e.g. dendritic cells) and factors (e.g. IFN γ) are also involved in the rejection of transplanted heterologous cells or tissues.

45 **[0008]** Document WO99/60846 is concerned with an immunocompromised mouse. Specifically the immunocompromised mouse described in this document contains T-cells and B-cells, has reduced macrophage function and reduced natural killer cell activity and it's capacity to tolerate any grafted cells is enhanced by reducing the activity of dendritic cells by administering an antibody directed to the interleukin 2-receptor λ -chain.

[0009] Accordingly, a mouse which, compared to the NOD/LtSz-scid, β 2m null mouse, has no variation in heterologous cell engraftment capacity, requires no antibodies, and has excellent heterologous cell engraftment is desirable.

50 DISCLOSURE OF THE INVENTION

[0010] Thus, the object of the present invention is to solve the above problems and to provide a method of producing a mouse having excellent heterologous cell engraftment capacity and a mouse produced by the same method.

55 **[0011]** The present inventors have made intensive studies to solve the above problems. As a result, they have obtained the findings that a mouse which has no variation in engraftment capacity of heterologous cells and requires no antibodies (that is, is suitable for the engraftment of heterologous cells) can be obtained by backcrossing an NOD/Shi mouse with a C.B-17-scid mouse, and further backcrossing the thus obtained mouse with an interleukin 2-receptor γ -chain (IL-2R γ) gene-knockout mouse. The present invention has been accomplished based on the above findings.

[0012] Namely, the present invention is as follows.

(1) A NOG (NOD/Shi-scid, IL-2R γ KO) mouse having excellent engraftment capacity of heterologous cells, wherein both of functional T-cells and functional B-cells are deleted, macrophage function is reduced, NK cells or NK activity are eliminated, dendritic cell function is reduced.

(2) The NOG mouse described in (1), wherein transplanted human stem cells efficiently differentiate and proliferate without being eliminated, with the proviso that the human stem cells are not human embryonic stem cells.

(3) A stem cell assay method comprising transplanting human stem cells to the mouse described in any of (1) or (2), sacrificing the mouse, collecting the human stem cells from the mouse and analyzing the differentiated and proliferated cells from the transplanted human stem cells, with the proviso that the human stem cells are not human embryonic stem cells.

(4) The stem cell assay method described in (3), comprising analyzing the differentiation and proliferation of T-cells and B-cells.

(5) A method of proliferating human stem cells comprising:

transplanting and proliferating the human stem cells to the mouse described in any of (1) or (2); sacrificing the mouse;

collecting the human stem cells from bone marrow of the mouse; and

repeatedly transplanting the collected cells to the mouse described in any of (1) or (2) with the proviso that the human stem cells are not human embryonic stem cells.

(6) The method of proliferating human stem cells described in (5), wherein the frequency of repeating is at least three times.

(7) The method described in (5) or (6), wherein the human stem cells have foreign genes introduced therein.

(8) A human tumor model mouse wherein the mouse is a mouse described in any of (1) or (2) and retains human tumor cells.

(9) The human tumor model mouse described in (8), wherein the human tumor cells are derived from HTLV-1 leukemia.

(10) The human tumor model mouse described in (8) or (9), wherein the mouse has the human tumor cells at an auricle thereof.

(11) A method of producing a human tumor model mouse comprising transplanting human tumor cells to the mouse described in any of (1) or (2), and sacrificing said mouse prior to said analysis.

(12) The method described in (8), wherein the human tumor cells are derived from HTLV-1 leukemia.

(13) The method described in (11) or (12), wherein the human tumor cells are transplanted at an auricle of the mouse.

(14) A virus-infected model mouse wherein the mouse is a mouse described in any of (1) or (2) and retains T cells infected with a T-tropic (T-cell affinity) virus as well as a macrophage-tropic virus.

(15) The virus-infected model mouse described in (14), wherein the virus is HIV.

(16) The virus-infected model mouse described in (14), wherein the virus is HTLV-1.

[0013] Hereinafter, general embodiments of the present invention will be described.

1. Production of a NOD/Shi-sci, IL - 2R γ KO (NOG) mouse of the present invention

[0014] A method of producing a NOD/Shi-sci, IL - 2R γ KO (NOG) mouse suitable for the engraftment of heterologous cells is characterized in backcrossing a mouse B with a mouse A, as described below.

A: a mouse obtained by backcrossing a C.B-17-scid mouse with an NOD/Shi mouse; and

B: an interleukin 2-receptor γ -chain gene knockout mouse.

[0015] Here, examples of the heterologous cells include cells or tissues derived from mammals such as humans, mice, rats etc., particularly human stem cells, lymphocytes or tumor cells etc. of humans, but not limited cells thereto, with the proviso that the human stem cells are not human embryonic stem.

[0016] With respect to the mouse A, backcrossing the C.B-17-scid mouse with the NOD/Shi mouse is done in accordance with methods well-known to a person skilled in the art, for example, backcrossing by Cross Intercross method (Inbred Strains in Biomedical Research, M.F.W.Festing, 1979, ISBN 0-333-23809-5, The Macmillan Press, London and Basingstoke). The C.B-17-scid mouse is crossed with the NOD/Shi mouse, and the obtained F1 mice are further crossed with each other. Then, the immunoglobulin amount in blood serum of the thus obtained F2 mice is measured for selecting a mouse, from which immunoglobulin cannot be detected. The selected mouse is again crossed with a NOD/Shi mouse.

Repeating this process (Cross Intercross method) 9 times or more enables the accomplishment of the backcrossing.

[0017] A NOD/Shi mouse and a C.B-17-scid mouse are both commercially available from CLEA JAPAN, INC. Further, examples of mice obtained by crossing these mice with each other include a NOD/Shi-scid mouse (also called as a NOD-scid mouse) (Japanese Patent Application Laying-Open (kokai) No. 9-94040) which the present inventors have already established. This mouse is purchased from CLEA JAPAN, INC., and can be used directly as the mouse A. In addition, the present inventors possess, other than the ones mentioned above, NOD/Shi mice and NOD/Shi-scid mice, which can be split up and provided whenever the need arises.

[0018] Moreover, with respect to the mouse B, knockout of an interleukin 2-receptor γ -chain (IL-2R γ) gene is carried out in accordance with methods well known to a person skilled in the art, for example, a homologous recombination method using mouse ES cells (Capecchi, M. R., Altering the genome by homologous recombination, Science, (1989) 244, 1288-1292). After substituting a specific mouse-derived gene by a homologous gene including a gene resistant to a drug, for example neomycin etc. at ES cell stage, the ES cells are inserted into a fertilized egg, thereby accomplishing the gene-knockout.

[0019] Specifically, for example, gene clones containing a mouse IL-2R γ are isolated, from a genome library of 129/SV mouse, using a human IL-2R γ cDNA as a probe. Using a fragment of 8.6 kb containing the full length of IL-2R γ among the clones, a targeting vector is prepared. That is, PMC1-neo poly A which expresses a neomycin resistant gene, is inserted between exons 7 and 8 of IL-2R in the fragment, and also a diphtheria toxin-A gene is placed at 3' side 1 kb away from exon 8. Next, the vector is made linear, and introduced into 1×10^7 of E14 ES cells by electroporation. Thereafter, ES clones which bring about homologous recombination in the culture solution including G418, are selected (confirmed by PCR or Southern method), and after injecting the ES clones into blastocysts of C57BL/6 mice, they are transplanted into the uterus of foster parent mice. Chimeric mice born from the foster parent mice are further crossed with C57BL/6 mice, thereby obtaining IL-2R γ KO hetero mice wherein knockout is transduced to germ cells.

[0020] Alternatively, pre-established interleukin-2 receptor γ chain gene (IL-2R γ) knockout mouse strain may directly be obtained for use from suppliers, and examples of the mouse strains include interleukin-2 receptor γ chain (IL-2R γ) knockout mice (Ohbo K, Suda T, Hashiyama M et al., Modulation of hematopoiesis in mice with a truncated mutant of the interleukin-2 receptor gamma chain. Blood 1996;87(3):956-67)) which was produced from IL-2R γ KO mouse strains [Prof. Kazuo Sugamura, Department of Microbiology and Immunology, Tohoku University School of Medicine]. Incidentally, IL-2R γ KO mice are presently stored in the embryo preservation bank of the applicants (Central Institute for Experimental Animals) at the request of Prof. Sugamura, a producer of the mouse strain, and whenever the need arises they can be provided as frozen embryos or as thaw-reconstruction mice.

[0021] Further, backcrossing the mouse B with the mouse A can be carried out, in similar fashion as described above, according to conventional methods well-known to a person skilled in the art. For example, in accordance with the above backcrossing, that is, a NOD/Shi-scid mouse is crossed with an IL-2R γ KO mouse, and the obtained F1 mouse is backcrossed with an NOD/Shi-scid mouse, thereby accomplishing the backcross.

[0022] The mouse of the present invention is referred to as a NOG mouse (NOG mouse; NOD/Shi-scid, γ c null mouse; NOD/Shi-scid, IL-2R γ chain-/-mouse; NOD/Shi-scid, IL-2R(γ c)^{null} mouse etc.)

[0023] The mouse of the present invention is a severe immunodeficient mouse in which has both of functional T-cells and functional B-cells are deleted, macrophage function are reduced, and NK cells or NK activity are eliminated. Therefore, when heterologous cells (e.g. human peripheral blood mononuclear leukocytes) are introduced into the mouse of the present invention, much higher ratios of engraftment and proliferation are observed even in comparison with conventional immunodeficient mice which are subjected to anti NK antibody treatment. (See Example 1 described below) Further, dendritic cells of the mouse of the present invention are also functionally incompetent, and the production of cytokine is remarkably reduced. Thus, the mouse of the present invention has the most excellent engraftment capacity of heterologous cells as compared with conventional immunodeficient mice, and it is considered effective for analyses of various introduced heterologous cells (including stem cells, differentiated cells and cancer cells) which are engrafted in this mouse. Additionally, it is possible to use the mouse in order to establish a pathologic model mouse and produce a human antibody for HIV, HTLV-1 or cancer.

[0024] Hereinafter, the applications of the mouse of the present invention will be described. Usually, the mouse to be used is preferably 8 to 12 weeks old, but not limited thereto.

2. Establishment of human stem cell assay system using the mouse of the present invention

[0025] Using the mouse of the present invention, it is possible to establish a human stem cell assay system for examining factors and mechanisms which are engaged in differentiation and proliferation of human stem cells. Also, it is possible to research various therapeutic products using the human stem cell assay system.

[0026] Introducing human stem cells into the mouse of the present invention enables the establishment of the human stem cell assay system. Here, stem cells include, in addition to hematopoietic stem cells, stem cells not derived from the hematopoietic system, such as neural stem cells etc. Human stem cells are identified by the existence of a cell

surface marker which relates to a specific epitope site identified by an antibody, and for example, they can be isolated as CD34 positive cells from e.g. human bone marrow, umbilical cord blood, peripheral blood etc.

[0027] Stem cells are suspended in a solution such as physiological saline, phosphate buffered physiological saline etc., which exerts no influence on cells and living organisms, and 1×10^4 to 1×10^6 of cells are intravenously administered into the mouse, thereby carrying out the transplantation.

[0028] Cells are collected, several weeks after transplanting, from each organs such as peripheral blood, the spleen, bone marrow, the thymus etc. of the mouse to which the cells have been transplanted. The surface antigens of these cells are examined using e.g. FACS (Fluorescence-activated cell sorter), and thereby the differentiations of the transplanted cells are examined. In this case, examples of cell surface antigen markers to be used as index include: CD34 which relates to stem cells; CD3, CD4, CD8 etc. which relate to T-cells; CD10, CD19, CD20 etc. which relate to B-cells; CD5 etc. which relate to B1a cells; CD33 etc. which relate to myeloid cells; CD11c etc. which relate to dendritic cells; CD45 etc. which relate to the whole leukocytes; CD 11 a, CD 11b etc. which relate to macrophages; CD56 etc. which relate to NK cells; CD38 etc. which relate to plasma cells; CD41 etc. which relate to platelets; and glycophorin A etc. which relate to erythrocytes. According to need, various related markers can be selected.

[0029] The production of cytokines such as interferon, interleukin, TNFa etc. in the collected cells, is measured by ELISA etc., and thereby the differentiations of the stem cells are examined.

[0030] Further, it is possible to conduct successive transplantations of human stem cells using the mouse of the present invention. That is, true self-replicable human stem cells can be obtained. Specifically, human stem cells are transplanted in the mouse of the present invention, after several weeks undifferentiated human stem cells are collected from bone marrow of the mouse, and further the collected stem cells are transplanted in the mouse of the present invention. By repeating the transplantation and collection, human stem cells which are free from other cells and have high purity can be obtained in large quantities. By the successive transplantations, human stem cells with at least 99% or more purity, preferably 99.7% or more purity can be obtained. Conventional mice allow up to secondary transplantation, though the mouse of the present invention enables more than two successive transplantations.

[0031] For the treatment of leukemia or the like, human stem cells obtained using the mouse of the present invention can be transplanted to humans. Also, the mouse is usable for gene therapy targeting human stem cells by introducing a foreign gene into human stem cells and transplanting them to the mouse of the present invention for proliferation. With the aid of virus vectors such as lentivirus vectors, retrovirus vectors, adenovirus vector, and adeno-associated virus vector, a gene can be introduced into stem cells. Examples of the gene to be used here include an ADA gene for adenosine deaminase deficiency (ADA) patients. After these genes are introduced into human stem cells, the cells are transplanted to the mouse of the present invention for proliferation and purification and then administered to patients, thereby enabling gene therapy.

3. Production of human antibodies using the mouse of the present invention

[0032] Using the mouse of the present invention, established human cell lines which produce human antibodies, and human antibodies can be obtained. The above-described human stem cells are transplanted to the mouse of the present invention, and cells responsible for immunity such as T-cells or B-cells are differentiated and proliferated. Alternatively, cells responsible for immunity such as human T-cells or B-cells are transplanted to the mouse and engrafted in the mouse body, and thereby obtained is a mouse having the cells responsible for human immunity and capable of producing human antibodies. When human stem cells are transplanted, the differentiation and proliferation of T-cells and B-cells are realized in 6 to 8 weeks, enabling the production of human antibodies.

[0033] By administering antigens to the mouse having human T-cells and B-cells engrafted thereto and held, it is possible to obtain human antibodies against the antigens and cells capable of producing the antibodies. The administration of the antigens to the mouse of the present invention may be carried out by the same method as is conventionally used for immunizing a mouse.

[0034] Human antibody producing cells can be collected from each organ of the mouse, especially the spleen, lymph nodes etc. When the ratio of the human antibody producing cells is high, the collected cells can directly be used for establishing a cell line. However, when the ratio is low, if necessary, purification may be carried out by e.g. the affinity column method using anti human B-cell antibodies. Further, it is desirable to eliminate mixed-in mouse cells by e.g. cytotoxic method using anti mouse antibodies and complements.

[0035] The thus obtained human antibody producing cells are made into a established cell line by a transformation method using Epstein-Barr virus (EBV), a cell fusion method wherein the cells are fused with suitable proliferation viable cells, or the like. Then, obtainable is a human antibody-producing established cell line capable of multiple passages while producing antibodies.

4. Production of a pathologic model mouse with tumor

[0036] In the mouse of the present invention, human tumors can be engrafted and proliferated, and an animal model of a human tumor can be obtained by transplanting tumor cells to the mouse of the present invention. For example, the administration of the human tumor cells causes the proliferation thereof inside the mouse body, and thus a mouse having a human tumor can be obtained. Examples of the cells to be used in this case include subcultured lines of human tumor in a conventional nude mouse, and cell lines derived from HTLV-1 leukemia such as ED-40515(-), MT-1 and TL-Oml. In addition, human tumor tissues are chopped into pieces having a size of several mm, and these cancer tissue pieces may directly be transplanted and engrafted to the mouse of the present invention. In this case, the site of the mouse to which tumor cells or tissues are transplanted is not limited, but in the case of cells, they may be transplanted intraperitoneally, intravenously or subcutaneously to the mouse and in the case of the tissues, they may be transplanted subcutaneously to the mouse. Any subcutaneous site of the mouse may be acceptable such as subcutaneous gluteal region, but it is desirable to transplant them subcutaneously at an auricle or a dorsal region because the tumor can be checked without incision. Further, in order to obtain results which reflect clinical effects of an anti cancer agent, it is desirable to transplant them at the identical site as that for clinical test (in the case of colon cancer cells, the cells are to be transplanted to the colon). When the cells are transplanted, a tumor is formed within several weeks to several months. Specifically, when HTLV-1 cells are transplanted subcutaneously at a posterior auricle, a tumor is formed in 2 weeks, thereby enabling expeditious production of a practical tumor model mouse.

[0037] Moreover, when a tumor is transplanted to the mouse of the present invention, metastases of tumor cells such as leukemic changes are observed and the mouse is usable as a model animal for tumor metastasis.

[0038] Using the thus obtained human tumor model mouse, screening of an anti cancer agent, antimetastatic drug etc. can be performed. As a method therefor, a candidate agent is administered to a mouse having a tumor formed, by a suitable method e.g. oral, transdermal administration or the like. Then, observation on the size of the tumor, the size and number of metastatic focuses, the viability of the mouse etc., allows judgment on the effect of the drug.

5. Production of a viral infectious disease model mouse

[0039] Use of the mouse of the present invention enables the obtainment of a viral infectious disease model mouse. Namely, by transplanting to the mouse of the present invention cells which may be infected with a human virus, and infecting the cells with the virus, or transplanting cells infected with a virus, it is possible to obtain a viral infectious disease model animal, which has virus-infected cells engrafted and held.

[0040] In a conventional mouse, only an M-tropic virus which infects macrophages can proliferate, but the proliferation of T-tropic viruses such as HIV, HTLV-1, which infects T-cells becomes possible using the mouse of the present invention.

[0041] For example, 1×10^7 to 1×10^8 of human peripheral blood mononuclear leukocytes are intraperitoneally administered to the mouse of the present invention, and after several days some hundreds to thousands of TCID₅₀ of HIV were inoculated, thereby obtaining a HIV-infected model mouse having human cells infected with HIV. The HIV infection can be detected through the expression of HIV antigens such as p24 positive cells as an index.

[0042] Instead of HIV, the inoculation of HTLV-1 enables the obtainment of an HTLV-1 infected model mouse.

[0043] Use of the animal model for disease obtained according to the present invention, allows in vivo research on proliferation mechanisms of HIV, HTLV-1 etc., further development of therapies for virus infections, screening of therapeutic products for virus infection, or the like.

6. Production of a mouse having enhanced engraftment capacity of heterologous cells using a NOG mouse

[0044] Use of the NOG mouse of the present invention enables the production of a mouse having more enhanced heterologous cell engraftment. For example, such a mouse can be obtained by backcrossing a mouse wherein a gene relating to the mouse immune system is knocked out with the NOG mouse of the present invention. Examples of the genes relating to the immune system include cytokine receptor gene, cytokine gene etc.

[0045] Further, by introduction of human cytokine gene, which relates to the differentiation and proliferation of human cells, or the like (e.g. hGM-CSF or hSCF etc.), it is possible to produce a mouse having more enhanced heterologous cell engraftment. For instance, in accordance with a method of Pro.Natl.Acad.Sci.USA 77:7380-7384, 1980, or the like, the above gene is inserted into a pronuclear fertilized egg of the mouse, and an individual having this introduced gene incorporated therein is selected, thereby producing a mouse which expresses a human cytokine gene etc. Then, this mouse and the NOG mouse of the present invention were crossed with each other, thereby producing a mouse having enhanced heterologous cell engraftment.

BRIEF DESCRIPTION OF THE DRAWINGS

[0046]

- 5 FIG. 1 shows an outline of backcrossing for producing a NOG mouse.
 FIG. 2 shows time-course changes of human CD45 positive cells and human CD41 positive cells, after the introduction thereof, in peripheral blood of the NOG mouse to which CD34 positive cells are transplanted.
 FIG. 3 shows the ratio of CD45 positive cells in the bone marrow and spleen of the NOG mouse to which CD34 positive cells are transplanted.
 10 FIG. 4 shows the ratios of CD45 positive cells in peripheral blood of mice in a comparative test between the NOG mouse and a $\beta 2$ microglobulin deficient NOD-SCID mouse (NOD/LtSz-scid, $\beta 2m$ null mouse).
 FIG. 5 shows FACS patterns of NK cells and dendritic cells in spleen cells obtained from each mouse strain.
 FIG. 6 shows the results of ELISA for detecting the production amount of cytokine under the stimulation of *Listeria monocytogenes* antigens in spleen cells obtained from each mouse strain.
 15 FIG. 7 shows the removal of NK activity in the NOG mouse and the NOD/LtSz-scid, $\beta 2m$ null mouse.
 FIG. 8 shows the results of FACS wherein bone marrow cells from primary, secondary and tertiary mice to which human CD34 positive cells have been transplanted, are stained with human CD45.
 FIG. 9 shows engraftment and differentiation of human cells in the thymus of the NOG mouse to which human CD34 positive cells have been introduced.
 20 FIG. 10 shows engraftment and differentiation of human cells in the thymus of the NOG mouse to which human CD34 positive cells have been introduced.
 FIG. 11 shows engraftment and differentiation of human cells in the spleen of the NOG mouse to which human CD34 positive cells have been introduced.
 FIG. 12 shows engraftment and differentiation of human cells in the spleen of the NOG mouse to which human CD34 positive cells have been introduced.
 25 FIG. 13 shows engraftment and differentiation of human cells in peripheral blood of the NOG mouse to which human CD34 positive cells have been introduced.
 FIG. 14 shows engraftment and differentiation of human cells in bone marrow of the NOG mouse to which human CD34 positive cells have been introduced.
 30 FIG. 15 shows the production ability of human antibodies in the NOG mouse.
 FIG. 16 shows the tumor formations after transplanting LM-2-JCK to each mouse strain.

BEST MODE FOR CARRYING OUT THE INVENTION

- 35 **[0047]** Next, the present invention will be described in detail by referring to Examples.
[0048] Example 1 Production of an immunodeficient mouse (NOG mouse) with the deletion of NK activity and declined dendritic cell function, examination of the heterologous cell engraftment in the mouse, and establishment of an assay system of human stem cells using the mouse
 40 (1) Production of an immunodeficient mouse (NOG mouse) with the elimination of NK activity and reduced dendritic cell function
[0049] In order to obtain multifunctional immunodeficient mice with depleted NK activity, interleukin-2 receptor γ chain knockout mice (IL-2R γ KO mice) (8 week-old) which were transferred from Prof. Kazuo Sugamura (Department of Microbiology and Immunology, Tohoku University, School of Medicine) were backcrossed with NOD-Shi-scid mice (8 week-old) which were kept in the Central Institute for Experimental Animals (also available from CLEA JAPAN, INC.), and thereby F1 mice having an IL-2R γ mutant gene introduced thereinto were produced. The introduction of the mutant IL-2R γ chain gene in the F1 mice was confirmed by PCR amplification and detection of the gene. Specifically, first, DNAs were extracted by a DNA automatic extractor (MagExtractor manufactured by TOYOBO) from 100 μ L of blood taken
 45 from ocular fundus of the F1 mice. PCR buffer solution containing 23.5 μ L of 1.5mM MgCl₂, 0.4mM dNTP and two sets of 25pmol primers (the following primers PI and PIII were used for the determination of wild type, and a set of the following primers PI and PII were used for the determination of mutant type) was added to 1.5 μ L of this DNA, and PCR was conducted under the following amplification conditions for the determination of whether IL-2R γ chain genes were wild type or mutant type. (primers)
 50 PI : 5'-CTGCTCAGAAATGATGCCTCCAATTCC-3' (SEQ ID NO:1)
 PII : 5'-CCTGCGTGCAATCCATCTTGTTCAAT-3' (SEQ ID NO:2)
 PIII : 5'-GATCCAGATTGCCAAGGTGAGTAG-3' (SEQ ID NO:3)
 55

(PCR amplification conditions)

[0050] The conditions were heating at 94°C for 5 minutes; 30 to 35 cycles of 1 minute at 94°C, 1 minute at 55°C, and 1 minute at 72°C; and thereafter heating at 72°C for 10 minutes.

[0051] The PCR products obtained by the above PCR were subjected to electrophoresis in 2% agarose gel, and measured according to the size of the coloring band detected after ethidium bromide stain. The sizes of the bands, about 660 bp for wild type and about 350 bp for mutant type, were observed.

(Backcrossing)

[0052] Next, the F1 mice having the mutant IL-2R γ gene introduced thereinto were crossed with NOD/Shi-scid mice, thereby obtaining F2 mice. Further, by detecting the introduction of the mutant IL-2R γ chain gene into the F2 mice in the same manner as above, and detecting immunoglobulins in serum by an immunodiffusion method, mouse individuals which have the mutant IL-2R γ chain gene and has a homozygous scid gene were selected. Thereafter, the mouse individuals were crossed with NOD/Shi-scid mice, and among the born mice, mice having mutant IL-2R γ chain gene were further crossed with NOD/Shi-scid mice.

[0053] The above backcross was repeated at least 9 times, thereby producing NOG (NOG) mice (FIG. 1 shows the outline). Here, since the IL-2R γ chain gene exists on an X chain chromosome, it is effective to use male IL-2R γ KO mice.

(2) Examination on engraftment capacity of heterologous cells in NOG mice

[0054] Next, using the NOG mice obtained by the above crossing and conventional immunodeficient mice, NOD/Shi-scid mice, examinations were made on the level of impact that anti-NK antibody treatment has on engraftment of heterologous cells in these mice.

① (anti-IL-2 receptor β chain monoclonal antibody)

[0055] Anti-IL-2 receptor β chain monoclonal antibodies (clone TM β 1) were produced from hybridomas produced and provided by Prof. Masayuki Miyasaka, School of Medicine, Osaka University (Tanaka T, Tsudo M, Karasuyama H et al., A novel monoclonal antibody against murine IL-2Receptor beta-chain. Characterization on of receptor expression in normal lymphoid cells and EL-4 cells. J Immunol 1991;147(7):2222-8). In particular, the hybridomas were intraperitoneally administered to BALB/cA-nu mice and collected from ascites after several weeks.

[0056] One mg per mouse of the antibodies was intraperitoneally administered to five NOD/Shi-scid mice (8 to 12 week-old) and three NOG mice (8 to 12 week-old). Further, as controls with no administration of the antibodies, physiological saline was administered to four NOG mice (8 to 12 week-old).

[0057] Moreover, human peripheral blood lymphocytes were collected by use of density gradient centrifugation using Lymphoprep, from blood taken from volunteers.

[0058] 1×10^7 of the obtained human peripheral blood mononuclear leukocytes were intraperitoneally administered to the above mice on the second day after the administration of TM β 1.

[0059] The mice were sacrificed 2 weeks after the administration of human peripheral blood lymphocytes, and the ascites containing the whole peritoneal exudates cells were fully washed with RPMI-1640 culture medium and thereby collected. Out of all the peritoneal cells, all the peritoneal exudates cells were counted by flow cytometry, and depending on their amounts, the engraftment and proliferation of human cells into mice were determined. The same operations were conducted for control mice with no administration of the antibodies. Their results are shown in Table 1.

Table 1

Impact of anti-NK antibodies (TM β 1) on engraftment and proliferation of human peripheral blood mononuclear leukocytes in NOD/Shi-scid mice and NOG mice						
Mouse strain	TM β 1 treatment	Number of mice	Number of collected cells ($\times 10^6$, distribution)	Human cells %		
				HLA+ (distribution)	hCD4+	hCD8+
NOD/Shi-scid	+	5	4.8 (2.77-6.8)	41.1 (4.2-65)	ND	ND

(continued)

Impact of anti-NK antibodies (TM β 1) on engraftment and proliferation of human peripheral blood mononuclear leukocytes in NOD/Shi-scid mice and NOG mice						
Mouse strain	TM β 1 treatment	Number of mice	Number of collected cells ($\times 10^6$, distribution)	Human cells %		
				HLA+ (distribution)	hCD4+	hCD8+
NOG	+	3	11.1 (8.3-15.0)	61.9 (47.4-74.6)	19.8	28.2
	-	4	11.2 (7.9-20)	63.3 (51.4-69.7)	34.3	25.9

[0060] It is clear from the results shown in Table 1 that extremely high ratios of human cells were differentiated, engrafted and proliferated, even without the treatment of the antibodies, in NOG mice to which human peripheral blood mononuclear leukocytes were introduced, as compared with conventional TM β 1 treated mice.

② (anti asialo-GM1 antibody)

[0061] Next, using NOG mice and NOD/Shi-scid mice, examinations were made on the level of impact, which would be given by anti asialo-GM1 antibodies (AGM1) (rabbit) (Wako Pure Chemical Industries, Ltd., 014-09801), on engraftment capacity of cells derived from human umbilical cord blood in these mice.

[0062] First, 2.4 Gy of X ray were irradiated on NOG mice (8 to 12 week-old) and NOD/Shi-scid mice (8 to 12 week-old) which were numbered as indicated in Table 2. Then, 20 μ L per mouse of anti asialo-GM1 antibodies diluted with 400 μ L PBS were administered to these mice just before the administration of the cells derived from human umbilical cord blood. Also, physiological saline was intraperitoneally administered to control mice (No. 5, 6, 10, 11, 15, 16 mice in Table 2) without administration of the antibodies.

[0063] With respect to human umbilical cord blood CD34 positive cells, mononuclear leukocytes were collected by Ficoll-Hypaque gradient centrifugation from umbilical cord blood taken from volunteers from whom approval was each obtained in advance. Further, CD34 positive cells were isolated by Dynabeads M-450 CD34 and DETACHaBEAD CD34 (DynaL As, Oslo, Norway). (Ueda T, Tsuji K, Yoshino H et al., Expansion of human NOD/SCID -repopulating cells by stem cell factor, Flk2/Flt3 ligand, thrombopoietin, IL-6, and soluble IL-6 receptor. J Clin Invest 2000;105(7):1013-21)

[0064] 1×10^5 of the obtained CD34 positive cells derived from human umbilical cord blood were introduced through tail veins of the mice immediately after the administration of anti asialo-GM1 antibodies.

[0065] The mice were sacrificed 4 weeks after the administration of CD34 positive cells derived from human umbilical cord blood, their peripheral blood was collected, and human cells (CD45⁺) in mononuclear leukocytes and human platelets (CD41⁺) in the whole blood were counted by flow cytometry. Depending on their amounts, the engraftment capacity and proliferation of human cells into mice were determined. The same operations were conducted for control mice without administration of the antibodies. Their results are shown in Table 2.

55 50 45 40 35 30 25 20 15 10 5

Table 2 Differentiation and proliferation of introduced human umbilical cord blood CD34⁺ in NOG mice

				4 weeks after introduction							
Mouse No.	Mouse strain	AGM1 treatment	No. of introduced CD34 ⁺ cells	Days after transplant	No. of leukocytes (10 ² /ul)	No. of erythrocytes (10 ⁴ /ul)	No. of platelets (10 ⁴ /ul)	hCD45 ⁺ (%)	hCD45 ⁺ (/ul)	hCD41 ⁺ (%)	hCD41 ⁺ (/ul)
1	NOD/Shi-scid	+	100000	31 (4wk)	12	782	134	0.19	2	0.001	20
2	NOD/Shi-scid	+	100000	31(4wk)	12	809	147	0.94	11	0.022	318
3	NOD/Shi-scid	-	100000	31(4wk)	10	758	104	0.48	5	0.004	41
4	NOD/Shi-scid	-	100000	31(4wk)	14	773	109	3.24	45	0.166	1814
5	NOG	-	100000	31(4wk)	12	713	89.3	5.84	70	0.255	2274
6	NOG	-	100000	31(4wk)	17	749	108	9.29	158	0.324	3498
7	NOD/Shi-scid	+	100000	29(4wk)	10	734	139	1.82	18	0.056	772
8	NOD/Shi-scid	+	100000	29(4wk)	10	787	118	1.51	15	0.094	1105
9	NOD/Shi-scid	-	100000	29(4wk)	7	722	132	0.97	7	0.059	776
10	NOG	-	100000	29(4wk)	36	602	218.1	1.84	66	0.170	3709
11	NOG	-	100000	29(4wk)	10	725	111	9.28	93	0.518	5748
12	NOD/Shi-scid	+	100000	28(4wk)	7	696	92.4	1.74	12	0.007	63
13	NOD/Shi-scid	+	100000	28(4wk)	10	787	96	0.40	4	0.008	73
14	NOD/Shi-scid	-	100000	28(4wk)	10	710	82.6	0.41	4	0.021	173
15	NOG	-	100000	28(4wk)	2	684	103	8.52	17	0.101	1040
16	NOG	-	100000	28(4wk)	2	773	86.5	14.98	30	0.196	1692

(continued)

				4 weeks after introduction							
Mouse No.	Mouse strain	AGM1 treatment	No. of introduced CD34 ⁺ cells	Days after transplant	No. of leukocytes (10 ² /ul)	No. of erythrocytes (10 ⁴ /ul)	No. of platelets (10 ⁴ /ul)	hCD45 ⁺ (%)	hCD45 ⁺ (/ul)	hCD41 ⁺ (%)	hCD41 ⁺ (/ul)
Average	NOD/Shi-scid (AGM1+)							1.10	10.52	0.03	391.74
	NOD/Shi-scid (AGM1-)							1.27	15.25	0.06	701.14
	NOG							7.11	61.99	0.22	2565.96

[0066] It is clear from the results shown in Table 2 that extremely high ratios of human cells were differentiated, engrafted and proliferated, even without the treatment of the antibodies, in NOG mice to which CD34 positive cells derived from human umbilical cord blood were introduced, as compared with conventional mice which were treated with anti asialo-GM1 antibody.

(3) Establishment of assay system of human stem cells using NOG mice

[0067] Mice to be used here were NOD/Shi-scid mice and NOG mice of the present invention. 1×10^5 of CD34 positive cells taken from human umbilical cord blood (CB) were transplanted to the above mice which had received 2.4 Gy of radiation.

[0068] Human cells in peripheral blood, bone marrow, spleens, and thymuses were analyzed by FACS. The ratios of CD45 positive cells in peripheral blood 8 weeks after the transplant were 37% for NOG mice and 7% for NOD/Shi-scid mice. In bone marrow, the ratios were 65% and 20%, respectively. In addition to a high ratio of chimerism, it was found that differentiations occurred into various cell lines including B and T lymphocytes in peripheral blood, bone marrow, spleens, and thymuses of the NOG mice 3 month after the transplant. It was observed that human CD33⁺ bone marrow cells, CD19⁺B cells, CD3⁺T cells, CD56⁺NK cells, CD41a⁺ huge nucleoplasm, and glycophorin A⁺ erythrocytes were present in bone marrow. Further, it is interesting to note that though there were observed a small number of human T cells in the thymuses, CD4⁺/CD8⁻ T cells, CD4⁻/CD8⁺ T cells, and CD4⁺/CD8⁺ T cells were observed in the spleens. This indicates that human hematopoietic stem cells are differentiated and proliferated into mature T cells at sites apart from the thymus.

[0069] Umbilical cord blood was obtained at birth of a healthy newborn from a normal pregnant mother from whom approval was obtained in advance. The umbilical cord blood was heparinized and preserved, and treated by the operation described below within 24 hours after collecting. Then, it was used for a transplant test.

[0070] The purification of CD34 positive cells was conducted as follows. The heparinized umbilical cord blood was diluted two times with a buffer solution prepared by mixing phosphate buffered saline with 5% fetal bovine serum, and thereafter mononuclear leukocytes were isolated using Ficoll. From the isolated mononuclear leukocytes, CD34 positive cells were purified by use of Dynabeads™ M-450 CD34 available from Dynal Biotech, Ltd. Although the method therefor is as instructed by Dynal Biotech, Ltd., it is summarized as follows. Using phosphate buffered physiological saline containing 2% bovine serum albumin and sodium citrate, the concentration of the mononuclear leukocytes was adjusted so as to be 4×10^7 /mL. 100 μ L of well-suspended Dynabeads CD34 was added per 1 mL of mononuclear leukocyte suspension, and they were blended and reacted with each other over ice for 30 minutes. Beads which formed rosettes with cells were collected using a magnet. Subsequently, to the Beads forming rosettes with cells, a required amount of Detachabead™ CD34 was added and reacted therewith, blending at 37°C for 15 minutes. As CD34 positive cells were liberated from the Beads, a magnet was used to remove only the Beads, thereby obtaining CD34 positive cells.

[0071] For transplanting, 8 to 12 week-old NOD/Shi-scid mice, NOG mice, and NOD/LtSz-scid, β 2m null mice, all raised in an SPF (specific pathogen free) environment were used. These mice were raised in the animal experiment facility of Kyoto University, School of Medicine under the regulations of the facility.

[0072] In a comparative test for the former 2 types of mice, the mice were twice irradiated with 1.2Gy gamma ray (gamma cell ¹³⁷Cs), and 100,000 of CD34 positive cells per mouse were transplanted through the tail vein. For the NOD/Shi-scid mice, 200 μ g of anti asialo GM1 antibodies (Wako Pure Chemical Industries, Ltd.) were intraperitoneally administered immediately before the transplant and every eleventh day after the transplant.

[0073] In a comparative test between NOG mice and NOD/LtSz-scid, β 2m null mice, the mice were twice irradiated with 1.2Gy gamma ray in the same way as above, and thereafter 40,000 or 10,000 of CD34 positive cells were transplanted through the tail vein.

[0074] After transplantation, 732 mg/L of neomycin sulfate were added to the mice's drinking water for protection against infection.

[0075] The mice were ether-anaesthetized at moments as indicated, and then peripheral blood was collected from the orbital venous plexus for measuring the positive ratio of human hemocytes by use of flow cytometry. Although the flow cytometry was conducted in accordance with a widely-used general method, it is summarized as follows. The collected blood was instantly blended well with EDTA-2Na and allowed to stand at room temperature until analysis. An optimum amount of antibodies was added to and reacted with 50 to 100 μ L of the whole blood at 4°C for 30 minutes. Thereafter, hemolysis and immobilization were effected using FACS solution (Becton Dickinson and Co.), and the ratio was measured using FACS Calibur (Becton Dickinson and Co.). In the case of bone marrow and spleen, the mice were sacrificed with cervical vertebra dislocation and femora and spleens were eviscerated. Then, bone marrow and splenic cells were each liberated in culture solution containing 5% fetal bovine serum, and thereafter as cell suspensions, they were treated in the same manner as peripheral blood and analyzed by flow cytometry.

[0076] Antibodies to be used for the analysis were FITC conjugated anti human CD45 antibodies, PE conjugated anti human CD10 antibodies, PE conjugated anti human CD33 antibodies, PE conjugated anti human CD3 antibodies, PE

conjugated anti human CD34 antibodies, PE conjugated anti human CD41 antibodies (Beckton, Dickinson and Co.), PC5 conjugated anti human CD38 antibodies, PC5 conjugated anti human CD56 antibodies, PC5 conjugated anti human CD19 antibodies (Immunotech), APC conjugated anti mouse CD45 antibodies, and FITC conjugated anti mouse CD41 antibodies (BD Pharmingen).

[0077] With respect to human CD45 positive cells and human CD41 positive cells in mouse peripheral blood, changes in the ratios (%) and the absolute numbers (/μL) were investigated every fourth week until the 12th week (FIG. 2A and FIG. 2B). As shown in FIGS. 2A and 2B, significantly high ratios and absolute numbers were observed for NOG mice. The mice were sacrificed from 12 weeks onward after the transplant, and when the ratios of human CD45 positive cells in bone marrow and spleens were investigated in the same way as above, the NOG mice had very high positive ratios. (FIG. 3)

[0078] In the comparative test between NOG mice and NOD/LtSz-scid, β2m null mice, the positive ratios of human CD45 positive cells in mouse peripheral blood were investigated (FIG. 4). As shown in FIG. 4, the NOG mice had a higher positive ratio of human cells and exhibited a better engraftment capacity of human cells than NOD/LtSz-scid, β2m null mice.

Example 2 Examinations on functional incompetence of dendritic cells of NOG mice in NK activity and cytokine production

[0079] Three female NOG mice (10 to 12 week-old) of the present invention, four female and four male NOD/Shi-scid mice (10 to 12 week-old) obtained by backcrossing C.B-17-scid mice with NOD/Shi mice, and two female and two male NOD/LtSz-scid, β2m null mice (10 to 12 week-old) (or referred to as null β 2m(null)NOD/LtSz-SCID, β2m microglobulin deficient NOD/SCID mice; produced by Dr. Shultz, L.D. et al. of The Jackson Laboratory (Kolle O, Peled A, Byk T et al., beta2 microglobulin-deficient (β2m(null))NOD/SHI-SCID mice are excellent recipients for studying human stem cell function. Blood 2000 ; 95(10):3102-5.) were used for the following examinations.

[0080] After the NOD/Shi-scid mice were treated with anti asialo GM1 antibodies (αAGM), spleen cells were collected. The obtained cells were divided into two groups, and CD11c antigen positive cells (regarded as dendritic cells) were removed from one of them using a magnetic cell sorter (MACS). Likewise, splenic cells were collected from non-treated NOD/LtSz-scid, β2m null mice and NOG mice. A small amount of these cells was taken from each of them and provided for FACS analysis. Further, these four kinds of cells were divided into three groups: group I: NOG mice (3 mice); group II: αAGM non-treated NOD/Shi-scid mice (2 males), αAGM treated NOD/Shi-scid mice (3 mice), and NOD/Shi-scid mice (3 mice) which were treated with αAGM and further from which CD11c were removed; and group III: NOD/LtSz-scid, β2m null mice (4 mice). They were adjusted to have a cell concentration of 1×10^7 /ml in RPMI-1640, and 100 μl of each of them were dispensed into a well, stimulated by an equivalent amount of *Listeria monocytogenes* (LM) antigens, and cultured (using Triplicate). Since LM is 10^{11} Units/mL, it was diluted for use so as to be 2×10^7 Units/mL with RPMI-1640. In this case, specimens with no LM addition were used as a control. After 24 hours, the supernatants were collected, and the amounts of cytokine such as IFN-γ were determined by ELISA.

[0081] Subclasses (in particular, CD11c, CD11b) in spleen cells were analyzed by FACS.

[0082] FITC-labeled CD3 and biotiline-labeled Pan NK(DX5) were used as a T-cell marker and an NK cell marker, respectively. FITC-labeled anti-CD11c and PE-labeled anti-CD 11b were used as a macrophage and dendritic cell marker.

[0083] Further, NK activity in NOG, C.B-17-scid, NOD/Shi-scid, and NOD/LtSz-scid, β2m null mice (10 to 12 week-old) was examined by cytotoxicity test targeting ^{51}Cr -labeled NK susceptible YAC-1 cells. Namely, splenic cells were isolated from the mice and mixed and cultured with ^{51}Cr -labeled YAC-1 cells at various ratios therebetween. After culturing at 37°C for 4 hours under 5% CO₂, the radioactivity in supernatants was measured by a liquid scintillation counter. The NK activity is indicated according to the following calculation method.

$$\% \text{ specific cytotoxicity} = \frac{(\text{specific radioactivity} - \text{background radioactivity})}{(\text{maximum radioactivity} - \text{background radioactivity})} \times 100$$

[0084] FIGS. 5A and 5B show FACS patterns of the spleen cells obtained from each mouse strain.

[0085] NK cells were detected from NOD/Shi-scid and NOD/LtSz-scid, β2m null mice, though NK cells were not detected at all from NOD/Shi-scid mice [not] treated with anti asialo GM1 antibody and NOG mice. CD11c positive cells which were used as a dendritic cell marker were detected from all the mice at extremely high ratios. It was confirmed that CD11c positive cells were almost completely removed from the splenic cells of the NOD/Shi-scid mice [not] treated with anti asialo GM1 antibody by magnetic beads.

[0086] FIGS. 6A, 6B and 6C show detection results by ELISA on the production amount of cytokine in the above splenic cells under the LM stimulation. The following was found: although IFN γ production was observed in NOD/Shi-

scid mice not treated with anti asialo GM1 antibody and NOD/LtSz-scid, β 2m null mice, there was no detection from NOG mice; and, as with NOG mice, by removing CD11c positive cells from the NOD/Shi-scid mice [not] treated with anti asialo GM1 antibody, IFN γ production was not observed.

[0087] FIG. 7 shows NK activity of the splenic cells obtained from each mouse strain.

[0088] In comparison with C.B-17-scid mice, reduction of NK activity in NOD/Shi-scid mice was observed, but absolutely no NK activity was observed in NOG mice and NOD/LtSz-scid, β 2m null mice.

[0089] In view of the foregoing, it became clear that NOG mice and NOD/LtSz-scid, β 2m null mice completely lost their NK activity. However, according to the result of FACS analysis, it became clear that the NOG mice lost NK cells, though the NOD/LtSz-scid, β 2m null mice had NK cells present therein but lost NK activity.

[0090] Further, it was revealed that the functional incompetence of the dendritic cells was a cause for the cytokine production decline in splenic cells which was observed in the NOG mice. On the other hand, it was indicated that the NOD/LtSz-scid, β 2m null mice had almost the same patterns of FACS and cytokine production as the anti asialo GM 1 antibody[-non-]treated NOD/Shi-scid mice. It was obvious that the NOD/LtSz-scid, β 2m null mice lost NK cells and their dendritic cells were normal.

Example 3 Successive transplant of human stem cells using NOG mice

[0091] In order to examine self-replicability of gene-introduced human stem cells, umbilical cord blood stem cells were transplanted to NOG mice, and it was examined whether or not secondary or tertiary transplant is possible.

[0092] Umbilical cord blood was obtained from pregnant women, with their consent and approval, for use in tests. After mononuclear cells were isolated from umbilical cord blood using Ficoll-Hypaque (Lymphoprep, 1.077 ± 0.001 g/ml; Nycomed, Oslo, Norway), CD34 positive cells were purified using a CD34 positive separation column (MACS, Miltenyi Biotec, Glodbach, Germany). The CD34 positive cells had a purity of $96 \pm 3\%$.

[0093] Lentivirus vector pCS-CG was a vector which enables a GFP gene to be expressed by a CMP promoter, and it was provided by Dr. Miyoshi (Immunology, Medical Branch, University of Tsukuba). After umbilical cord blood CD34 positive cells were cultured for 24 hours in serum-free medium StemPro TM-34SFM (Gibco BRL) containing 50 ng/mL of each TPO, SCF and Flk-2/Flt-3 ligand (FL), they were subjected to five-hour infection of recombinant lentivirus CS-CG by MOI30 (Kawada H et al., Exp. Hematol. 27:904-915, 1999). Thereafter, cells were washed and subjected to 5-day extracorporeal amplification culture in the same serum-free medium as above in the presence of murine bone marrow stroma cells HESS-5 (Oki M et al., Exp. Hematol. In press. 2001).

[0094] After 7 week-old NOD/Shi-scid mice were subjected to 3Gy radiation, 3×10^5 of gene-introduced cells which were amplified by culture, were introduced through the tail vein. After 6 weeks, surface antigens of murine bone marrow cells were analyzed, and 1×10^7 of cells were introduced into irradiated NOG mice (secondary transplant). After 6 weeks, the same analysis was conducted, and 1×10^7 of cells were introduced into irradiated NOG mice (tertiary transplant). Further, after 6 weeks, the same analysis was conducted.

[0095] The analysis of surface antigens was conducted using FACS Calibur (Becton, Dickinson and Co.). The antibodies used here were FITC-labeled anti human CD45 (T-200), PE-labeled anti human CD2 (39C1.5), CD3(UCHT1), CD4(SK3), CD14(LeuM3), CD19(4G7), CD20(2H7), CD33(WM53), CD41(P2), CD56(N901), and glycophorin A (KC16), and all of them were purchased from Becton, Dickinson and Co.

[0096] The gene introduction efficiency into CD34 positive cells was $40 \pm 5\%$ (n=5). FIG. 8 shows the results of FACS wherein mouse bone marrow cells of primary, secondary, and tertiary transplant mice were stained with human CD45. It was observed that gene-introduced human cells existed in all the mice.

[0097] By using NOG mice of the present invention as a host, the inventors became the first in the world to successfully accomplish the introduction of a gene capable of being expressed in hematopoietic stem cells which can be transplanted to up to a tertiary host. Use of conventional NOD-scid mice enabled up to secondary transplant (Guenechea G et al., Nature Immunol, 2, 75-82, 2001), and beyond this it was shown that the present mice were more sensitive and useful as an assay system for hematopoietic stem cells.

Example 4 Production of complete human antibodies using NOG mice

[0098] After NOG mice (8week-old) and NOD/Shi-scid mice (8 week-old) were irradiated with 2.5Gy and 3.5Gy, respectively, they were subjected to intravenous transplant of 1×10^6 of CD34⁺ cells which were purified with magnet beads from human umbilical cord blood (provided by Tokai University, Cell Transplant Research Center, with the consent and approval of the pregnant women). Specifically, MNC was isolated from umbilical cord blood using Ficoll, and then CD34⁺ cells were isolated by MACS immunomagnetic separation system (Miltenyi Biotec, Glodach, Germany). After confirming that the CD34⁺ cells had 97% or more purity by FACS, they were used for transplant.

[0099] Blood, having been chronologically taken from orbits, and after MNC fraction was obtained by Ficoll, was stained with fluorescence-labeled CD45, CD19, and CD3 antibodies (Becton Dickinson, San Jose, CA). The reconstruc-

tion of human lymphocytes, and the ratios of B-cells and T-cells were confirmed with CD45, and fluorescence-labeled CD19 and CD3, respectively, by FACS.

[0100] Moreover, thymuses and spleens were extracted from these mice, cells were prepared therefrom, the cell numbers were counted, and they were stained with various fluorescence antibodies (Becton Dickinson, San Jose, CA). Then, they were analyzed by FACS in the same manner as the above reconstruction confirmation.

[0101] From 6 or 8 weeks after the transplant, antigens had been administered to these mice. As an antigen, DNP-KLH was used. The immunization was performed intraperitoneally with 100 μ g/mouse of DNP-KLH together with an adjuvant of aluminum hydroxide (ALUM). The same immunization was performed every two weeks and serum was taken for measuring antibody titer by ELISA.

[0102] At this juncture, ELISA was performed as follows.

[0103] 96-well plate was coated with anti human IGs (ICN, Aurora, Ohio) or DNP-KLH, and blocking and washing were performed with 3% BSA. Diluted anti serum was added thereto. After the reaction at room temperature for 2 hours, washing was performed. Then, biotinylated anti human IgM or anti human IgG monoclonal antibodies (Pharmingen, San Diego, CA) were added. After the reaction at 37°C for 2 hours, washing was performed. Then, avidinylated peroxidase was added for 1 hour reaction at room temperature. The plate was washed and TMB peroxidase EIA substrate kit solution (Bio-Rad Laboratories, USA) was added for 30-minute reaction at room temperature. Then, the reaction was inhibited with 10% HC1, and the absorbance was measured at 450nm. The Ig concentration was calculated in accordance with the standard curve.

(1) Efficiency of transplanting and reconstituting CD34⁺ cells derived from umbilical cord blood

[0104] With respect to the ratio of human CD45⁺ cells in the NOG mice after transplanting the umbilical cord blood cells, the ratio of human CD45 in peripheral blood was gradually decreased from 4 weeks onward. However, from 12 weeks onward, suddenly the ratio of increase of T-cells became high, and along with this, it was observed that the ratio of human CD45 was also increased in some mice (Table 3). Increases of the CD45⁺ ratio, caused by such T-cell increase, had never been observed in peripheral blood of the existing NOD/Shi-scid mouse. These NOG mice had never been affected with GVHD until 14th week.

Table 3 Reconstitution of human cells in NOG mice: peripheral blood

SCT Gp.	Source	CD34 ⁺ cells	4 weeks			6 weeks			8 weeks			10 weeks			12 weeks			remarks
			%CD45	%CD19	%CD3	%CD45	%CD19	%CD3	%CD45	%CD19	%CD3	%CD45	%CD19	%CD3	%CD45	%CD19	%CD3	
2nd SCT	CB2	freeze thawing	47.3	45.7	1.3	N.A.	N.A.	N.A.	27.5	42.7	7.2	19.8	27.9	4.9				
	PBSC1	freeze thawing	79.5	0.7	13.4	72.5	22.9	47.7	13.3	14.3	65.7							Analyzed because of death at 8-week
	PBSC2	freeze thawing	30.8	3.0	0.1	62.0	63.2	3.8	68.5	93.6	2.1	13.7	64.5	23.9	10.3	48.4	32.6	
	PBSC3	freeze thawing	26.9	3.7	0.0	N.A.	N.A.	N.A.	43.5	62.0	7.6	28.4	34.1	11.1	22.0	35.5	14.0	
3rd SCT	CB	fresh	4.8	5.7	0.1	82.2	95.9	0.0	12.4	79.1	1.0	21.8	71.7	10.1				
	BM	freeze thawing	1.7	0.0	0.0	8.2	48.2	0.0	13.5	49.8	0.0	7.0	73.7	0.0				mixed
	PBSC2	freeze thawing	1.4	2.0	0.0	51.6	91.7	0.0	25.6	91.4	0.0	25.8	85.6	0.1				
	PBSC3	freeze	2.1	1.3	0.2	24.1	88.4	0.0	45.3	90.2	0.0	14.3	82.5	0.0				

[illegible]

(2) T-cell differentiation in NOG mice

A. T-cell differentiation in the thymus

5 **[0105]** FIGS. 9A, 9B, 10A and 10B show T-cell differentiation patterns in thymuses. As shown in the figures, CD3 positive cells were differentiated in the thymuses of these mice, and it was indicated that these cells contained CD4/CD8 DN (Double negative), DP (Double positive), and SP (Single positive) as well as those cells in a normal thymus or detected by hu/m-RTOC, and a part of them were mature to be CD1a-low T-cells in 11th or 13th week after transplanting. The number of the thymic cells was 1 to 2×10^6 and this was about one-hundredth of the number of thymic cells in a normal mouse. This phenomenon was observed also in NOD/Shi-scid mice, but its frequency was remarkably low, such as 8/28 (28.6%). As opposed to this, NOG mice had a frequency of 7/7 (100%).

B. T-cells at peripheries

15 **[0106]** FIGS. 11A, 11B, 12A, 12B, 13A, and 13B show further analysis on T-cells in splenic cells or peripheral blood. CD3 positive cells were observed even at the peripheries of the mice from which T-cell differentiation was observed in their thymuses. These are a group of CD4/CD8 positive cells, and this indicated that there was the possibility that they included cells differentiated in thymuses.

20 (3) B-cell differentiation in NOG mice

A. Subset differentiation of B-cells

25 **[0107]** Also, FIGS. 14A and 14B show the expression ratios of CD5 by B-cells among bone marrow cells. In bone marrow, the differentiation of CD5 positive cell, so-called B1a cells, was not facilitated, and groups of CD19 positive or IgM positive cells were dominant. Further, cells in a group in which IgM is highly positive expressed CD20. No abnormality was detected on the differentiation of human B-cells in the bone marrow of these mice. This phenomenon is also observed in B-cell differentiation in NOD/Shi-scid mice, and thus it is considered a common quality.

30 B. B-cell at peripheries

[0108] FIGS. 13A and 13B show FACS patterns of B-cells at their peripheries. The ratios of B-cells were almost the same as those in NOD/Shi-scid mice. However, in spleens, CD5 positive cells, so-called B1a cells which was a subgroup of B-cells were dominant, and a large difference was detected from differentiation patterns in bone marrow. On the other hand, IgM positive CD5 negative cell groups were also detected at a ratio of about 20% and thus this proved that cells, other than B1a cells, were also detected.

(4) Antibody production ability in NOG mice

40 **[0109]** DNP-KLH was administered as an antigen to these mice, and IgM and IgG antibody production amounts were chronologically measured. As a result, antigen non-specific IgM and IgG antigen specific IgM were detected by repeating the administration three times. Although T-cells were detected in thymuses and peripheries, antigen specific IgG production was not detected. (FIGS. 15A, 15B and 15C) FIGS. 15A, 15B and 15C also show the results of mice having transplanted thereto CD34⁺ cells derived from bone marrow (BM) (FIG. 15B) and peripheral blood (PBSC) (FIG. 15C). In bone marrow and peripheral blood, there was observed a tendency of lower productions of specific IgM and non-specific IgG as compared with the production in umbilical cord blood.

[0110] The above results indicate that the present mouse differentiates human T-cells and B-cells and is valuable for use in human antibody production systems.

50 Example 5 Neoplasm proliferation system

[0111] NOD/Shi-scid mice, NOG mice, BALB/cAJcl-nu mice (purchased from CLEA), and C.B-17/lcr-scid mice (purchased from CLEA) were used. All the used mice were 5 weeks old or older.

55 **[0112]** As cells to be transplanted to the mice, transplant human tumor cell line, LM-2-JCK was used. LM-2-JCK was a cell line which was established from lymphoblast lymphoma of a 13-year-old female patient and maintained by successive heterografts to nude mice. It has been reported that though LM-2JCK expresses T-cell antigen CD4 and CD5, it does not express other cell antigens including antibodies.

[0113] A solid tumor which was subcutaneously passaged 12 times in nude mice was used for heterograft assay. The

tumor was chopped into pieces in F-10 nutrition supplementary medium (GIBCO BRL) by scissors and fully dispersed by a pipette, and then cell suspension was prepared by passing through a nylon mesh. The concentration of viable cells in the suspension was calculated by trypan blue stain (GIBCO BRL). After centrifugation, tumor cells were dispersed at concentrations of 1×10^7 and 1×10^6 viable cells/ml into physiological saline. The tumor cell dispersion liquid was subcutaneously injected, in an amount of 0.1mL, at both flanks of mice by using 1 ml syringe equipped with 25 gage needle. After the injection of the tumor cells, the size of the tumor and the body weight were measured at intervals of one week. On the 21st day after the 1×10^6 cells were transplanted, when the tumor became large, the mice were sacrificed and the weights and sizes of the tumors were measured.

[0114] The difference in transplantability of LM-2-JCK among the mice having different backgrounds is shown in Table 4.

Table 4 Heterotransplantation of LM-2JCK to mice having different genetic backgrounds

Mouse strain	No. of cells grafted	Observation period	No. of tumor-engrafted mice/No. of transplanted mice	Weight of tumor a)
NOG	10^6	21 days	10/10 (100%)	3.97 ± 2.10 b)
NOD/Shi-scid	10^6	21 days	10/10 (100%)	1.34 ± 0.77
C.B-17/lcr-scid	10^6	21 days	8/10 (80%)	1.21 ± 1.06
NOG	10^5	< 9 weeks	8/8 (100%)c)	n.t.
NOD/Shi-scid	10^5	< 9 weeks	5/10 (50%)	n.t.
C.B-17/lcr-scid	10^5	< 9 weeks	0/10 (0%)	n.t.
a) The weight of tumor was measured 21 days after the transplanting and indicated with average $\pm$ SD(g). b) As to NOD/Shi-scid and C.B-17/lcr-scid, when $p < 0.05$ (t-test), the significant difference is observed. c) As to NOD/Shi-scid ($p < 0.05$) and C.B-17/lcr-scid ($p < 0.01$), the significant difference is observed. (χ^2 test) n.t.: not tested				

[0115] In the case of the transplant of 1×10^6 cells, all the tumors were engrafted in NOG mice and NOD/Shi-scid mice. In contrast, the engraftment ratio for C.B-17/lcr-scid was 80%.

[0116] The tumor engraftment ratios in these strains were lower in the case of the transplant of 1×10^5 cells, and particularly tumor proliferation was not observed in any of the C.B-17/lcr-scid mice.

[0117] In the case of the transplant of 1×10^6 cells, the growth curves of the tumors are shown in FIG. 16.

[0118] The tumor proliferation in NOG mice exceeded that of the other 2 strains.

[0119] On the 21st day, the NOG mice had 2.89 and 3.97 times larger average tumor volume than the NOD/Shi-scid mice and C.B-17/lcr-scid mice, respectively, had on the same day, and significant differences were observed by Student t-test. ($p < 0.001$)

[0120] No significant difference was observed in the average volume on the 21st day between tumors engrafted in NOD/Shi-scid mice and tumors engrafted in C.B-17/lcr-scid mice.

Example 6 Establishment of HIV-infection model system using NOG mice

[0121] Establishment of HIV-infection model system using NOG mice was examined by use of various HIV lines. Typical examples are shown below.

[0122] NOG mice were used in this example. For comparison, NOD/Shi-scid mice were also used, which were produced before by backcrossing both mutants of C.B-17-scid mice and NOD/Shi mice with each other.

[0123] HIV-1 used herein was a JRFL virus which was isolated from frontal lobe tissue of a patient with AIDS-associated encephalopathy and had infectivity on a DNA-cloned macrophage and T-cells. Further, a GFP-HIV-1 in which a JRFL virus and a GFP gene were incorporated at env V3 region and downstream of gp41, respectively was used.

(1) Production of mice (hu-PBL-NOG mice) to which human mononuclear lymphocytes are transplanted

[0124] 1×10^7 per mouse of peripheral blood human lymphocytes (PBL), provided by a normal person with consent and approval for test use, were intraperitoneally inoculated directly into NOG mice. For comparison, the PBL from the

EP 1 338 198 B1

same donor was intraperitoneally inoculated into NOD/Shi-scid mice having a normal IL-2R γ chain.

(2) Infection of mice with HIV-1

5 **[0125]** On the 6th day after the PBL inoculation, 1,000 TCID₅₀ of viruses were intraperitoneally inoculated into mice. The mice were sacrificed 2 weeks after the virus infection and ascites including abdominal exudates cells were collected. The number of human cells in the ascites was calculated by FACS. Further, spleens were extracted and fixed by paraformaldehyde, and then made into paraffin-embedded sections.

10 (3) Pathological analysis

[0126] The spleen tissue sections were subjected to HE-stain, and further to immunostaining using antibodies against human CD3, CD4, CD8, and HIV-1p24 antigens. For immunohistologic stain, in addition to conventionally reported ABC method, EPOS method and Envision method were employed.

15 **[0127]** The engraftment of human cells in hu-PBL-NOG mice was evaluated.

[0128] A series of examination results are shown in Table 5.

Table 5

HIV-1 infection test on human PBMC grafted-NOG mice female without TM β 1 non existent PBL NOG mice					
PBL transplantation total PBL 160mL 4.37×10^8 cells 2×10^7 cells per mouse Donor A 14 NOG mice (6 males, 8 females)					
Infection NOG mice					
	JRFL	M-tropic	3 males		
	NL4-3	T-tropic	3 males		
	JRCSF	M-tropic	3 females		
	NLCSFV3EGFP	M-tropic	2 females		
	MOCK		3 females		
13th day after infection					
	Cell number $\times 10^4$ cells	%HLA	%CD4	%CD8	p24
1. Mock ♀					
Ascites	75	67.7	21.4	56.4	
Spleen	510	71.8	13.8	45.5	
PBL	430	77.9	16.8	49.9	0
2. Mock ♀					
Ascites	130	67.6	14.8	60.5	
Spleen	698	63.3	12.7	49.2	
PBL	86	66.6	11.6	55.5	0
3. Mock ♀					
Ascites	110	62.3	36.4	44.2	
Spleen	434	61.3	20.9	45.2	
PBL	93	34.7	20.6	44.7	0

EP 1 338 198 B1

(continued)

4. JRFL ♂					
Ascites	152	75.5	1.7	53.9	
Spleen	335	54.6	2.8	64.6	
PBL	210	42.8	2.1	77.3	3,090
5. JRFL ♂					
Ascites	220	75.8	1.8	57	
Spleen	12	75.8	1.65	79.6	
PBL	870	81.1	1.8	77.9	5,445
6. JRFL ♂					
Ascites	178	36.3	1.79	54.1	
Spleen	418	65.3	1.1	74.4	
PBL	65	33.1	1.72	75.6	1,961
HIV-1 infection to human PBMC grafted-NOG mice (2)					
	Cell number × 10 ⁴ cells	%HLA	%CD4	%CD8	p24
7. NL4-3 ♂small spleen					
Ascites	120	7.6	9.74	74.1	
Spleen	14	no FACS			
PBL	non-existent				
8. NL4-3 ♂					
Ascites	84	75.5	1.43	45.7	
Spleen	465	53	1.64	60.0	
PBL	26	11.3	4.59	49.3	118
9. NL4-3 ♂					
Ascites	110	8.1	9.33	80.6	
Spleen	17	14.8	19.1	23.5	
PBL	3	0.6	13.0	27.8	0
for three male NL4-3, their ears were excised.					
10. JRCSF ♀					
Ascites	320	29.9	0.77	93.7	
Spleen	230	67	1.85	74.9	
PBL	49	33.7	0.48	71.7	3,132
11. JRCSF ♀					
Ascites	63	68.5	3.18	41.6	
Spleen	390	68.7	3.58	85.8	
PBL	121	2.9	4.5	31.5	4,180
12. JRCSF ♀ poor construction					
Ascites	110	15.2	23.0	62.4	

(continued)

12. JRCSF ♀ poor construction					
Spleen	380	14.5	2.98	89.8	
PBL	110	44.7	2.06	89.4	864
13. NLCSFV3EGFP ♀ GFP+ of spleen and PBL are 0.1 % or less					
Ascites	43	72.5	2.26	69.8	
Spleen	250	50.8	0.77	67.7	
PBL	124	76.5	5.07	63.3	271
14. NLCSFV3EGFP ♀ GFP+ of ascites, spleen and PBL are 0.2 to 0.3% or less					
Ascites	110	44.3	6.36	76.3	
Spleen	560	2.7	23.3	44.4	
PBL	127	3.4	5.26	84.2	114

[0129] CD4 positive cells were specifically killed by all the HIV viruses. Conventionally, it has been reported that only M-tropic viruses can proliferate in mice and CD4 positive cells are killed, but T-tropic viruses can proliferate in NOG mice.

[0130] It was found that human CD4 positive T-cells and CD8 positive T-cells were each engrafted around central arteries of spleens of the mice to which human peripheral blood was transplanted. With respect to the number of human CD3 positive cells, NOG mice had obviously a larger number of human cells than NOD/Shi-scid mice. According to the measurement by FACS, there were 1×10^7 or more human CD4 positive cells on average in ascites.

[0131] On the other hand, CD4 positive cells were hardly detected from NOD/Shi-scid mice and almost all the detected cells were CD8 positive cells. In the example previously described, the antibodies (TM β -1) (0.5 mg per mouse) which were against mouse IL-2 receptor β chain and suppressed the differentiation of NK cells were intraperitoneally injected into the NOD/Shi-scid mice 3 days before the inoculation of human PBL. However, this treatment was not conducted in this example. It has been already confirmed in other tests that antibody-treated mice would have many CD4 and CD8 positive human cells engrafted to them. Still though the number of human cells from NOG mice was apparently 3 to 4 times more than the number of cells from NOD/Shi-scid mice that had been subjected to the antibody treatment.

[0132] In the HIV-infected mice, p24 positive cells which were HIV antigens were detected around spleen arteries. Further, in spleens and ascites of the HIV-infected mice which express GFP, cells with GFP expression were clearly detected. The number of virus-infected cells of NOG mice was apparently larger than that of NOD/Shi-scid mice.

Example 7 Establishment of HTLV-1 infection model system using NOG mice

[0133] 2×10^7 cells per mouse of MT-2 cells which were cell lines derived from HTLV-1 leukemia, were transplanted to NOG mice.

[0134] In the 4th week after the transplant, the number of mice having lymphomas formed therein (Table 6) was counted, and it was checked whether or not the lymphomas mice were infected with a HTLV gene by Southern blotting and PCR. As a result of pathological search, dermal lymphoma and posterior intraperitoneally bilateral lymphomas are observed. The mice having a large tumor had metastasis in lymph nodes around stomach and had cancer cells infiltrated in a pleura thereof. Further, HTLV positive cells increased in peripheral blood. This indicated a typical onset of leukemia. Moreover, lung-interstitial invasion was also observed. All of these cells expressed Tax protein, but there was no intracerebral invasion.

Table 6

Transplantation of HTLV-infected cells to NOG mice 2×10^7 cells per mouse				
	No. of mice	TM β -1	Cell treatment by gamma ray	No. of mice having lymphoma (after 1 month)
NOD/Shi-scid mice	5	-	-	0
NOG mice	40	-	-	33(82.5%)
NOG mice	4	-	20,000 rad	0
NOG mice (no cell transplanted)	2	-	-	0

Example 8 Examination of leukemic changes by introducing neoplastic cells and transplantability of HTLV-1 positive cells using NOG mice

[0135] Transplant tests of various HTLV-1 cell lines and tumor formation tests were performed on NOG mice and the usefulness of the mice was examined.

[0136] Using 3 samples of HTLV-1 leukemia derived cell lines [L=Leukemic cell lines: ED-40515(-), MT-1 and TL-Oml], 6 samples of HTLV-1 infected cell lines [IT=infected transformed cell lines: SLB-1, M8116, HUT-102, MT-2, MT-4 and TY9-31MT], 10^{7-8} cells/0.5mL were transplanted to NOG mice and NOD/Shi-scid mice subcutaneously at a left posterior auricle thereof, and to some of them subcutaneously at a gluteal region thereof. Then, the tumor size in the mice, tissue image, and the mice with, and those without, leukemia were chronologically compared for examination, and the relations between these and transcription factors such as NFkB or fluctuation of Tax protein were also analyzed.

[0137] Within 2 weeks, a tumor with a size of 24x17x11 mm was formed in all cases of HTLV-1 leukemia derived cell lines [L] and exceptionally in SLB-1 of HTLV-1 infected cell line [IT]. In contrast, in 5 out of 6 cases of IT, a tumor with a size of at most 10x10x7 mm was formed, and in the case of M8116 or TY9-31MT, there was almost no tumor formation. (Table 7)

Table 7 Development of HTLV-1 infected-cells and leukemia cells in NOG mice and their features in vitro and in vivo

Cell line	Cell type	No. of mice	In vitro			Cells inoculated ($\times 10^7$)	Route and site of inoculation	In vivo	
			proliferation pattern	IL-2 dependence	Tax expression			period (days)	size of tumor (mm)
ED-40516(-)	L	1	separately	-	-	7.5	I.m Buttock	16	23 $\times$ 18 $\times$ 12
		7	separately	-	-	7.5	s.c post-auricular	15-16	25 $\times$ 18 $\times$ 12
		3	separately	-	-	4	s.c post-auricular	16	25 $\times$ 18 $\times$ 12
		3	separately	-	-	1	s.c post-auricular	15	22 $\times$ 17 $\times$ 10
SLB-1	IT	1	clustered	-	+	7.5	I.m Buttock	15	20 $\times$ 10 $\times$ 18
		7	clustered	-	+	7.5	s.c post-auricular	15-16	20 $\times$ 10 $\times$ 18
		3	clustered	-	+	5	s.c post-auricular	15	21 $\times$ 12 $\times$ 17
MT-1	L	1	separately	-	-	7.5	I.m Buttock	20	25 $\times$ 15 $\times$ 10
		1	separately	-	-	7.5	s.c post-auricular	20	25 $\times$ 15 $\times$ 10
		3	separately	-	-	4	s.c post-auricular	24	23 $\times$ 16 $\times$ 12
TI-oml	L	1	separately	-	-	7.5	I.m Buttock	20	25x 15x 10
		1	separately	-	-	7.5	s.c post-auricular	20	25x 15x 10
		3	separately	-	-	4	s.c post-auricular	20	22x23x8
M8116	IT	1	clustered	-	+	7.5	I.m Buttock	18	sesame
		2	clustered	-	+	7.5	s.c post-auricular	18	sesame

(continued)

Cell line	Cell type	No. of mice	In vitro			Cells inoculated (×10 ⁷)	Route and site of inoculation	In vivo	
			proliferation pattern	IL-2 dependence	Tax expression			period (days)	size of tumor (mm)
HUT-102	IT	1	clustered	-	+	7.5	I.m Buttock	18	10x10x7
		2	clustered	-	+	7.5	s.c post-auricular	18	10x10x7
MT-2	IT	1	clustered	-	+	7.5	I.m Buttock	18	10x10x7
		2	clustered	-	+	7.5	s.c post-auricular	18	10x10x7
MT-4	IT	1	clustered	-	+	7.5	I.m Buttock	18	5×5×3
		2	clustered	-	+	7.5	s.c post-auricular	18	5×5×3
TY9-31MT-2	IT	1	clustered	-	+	7.5	I.m Buttock	18	sesame
		2	clustered	-	+	7.5	s.c post-auricular	18	sesame

L: leukemia cell line IT: infected-tranformant cell line

EP 1 338 198 B1

[0138] The case of this tumorigenic cell line showed that the degree of the emergence of leukemic cells in peripheral blood and the invasion of organs were relatively high, but some cases of nontumorigenic cell lines had higher degrees thereof (e.g. HUT-102, MT-2 etc.). Thus, there were not necessarily correlations between tumorigenicity and the degrees of the emergence of leukemic cells in peripheral blood and the invasion on organs. (Table 8)

Table 8

Infiltration of leukemia cells into various organs of NOG mice									
Cell strain	Tumor	peripheral blood (PB)	bone marrow (BM)	liver	spleen	lung	brain	kidney	heart
Ed-40515 (-)	+++	++	-	+	+	+-	Spongy +-	++	-
SLB-1	+++	+++	+	++ +	+	Granular +++	Spongy ++	+	+
MT-1	+++	++ +	-	+	+	+	Spongy +-	-	-
TI-oml	+++	++ +	-	Gaint +++	+	+-	-	-	-
M8116	+	+	-	-	-	-	-	-	-
Hut-102	++	++	+++	Nodule +++	+++	Pneumonia +	Spongy +-	-	-
MT-2	++	++	+ -	+ -	++	+-	-	+	-
MT-4	++	+	+-	-	++	-	-	-	-
TY9-31MT-2	+-	-	-	-	-	-	-	-	-

[0139] 10^7 cells of tumorigenic ED-40515(-) were transplanted subcutaneously at a left posterior auricle of NOG mice and NOD/Shi-scid mice and they were compared in tumorigenicity in the second week. A tumor with a size of 22x17x10 mm was formed in the NOG mice, and in contrast a tumor was hardly formed at all in NOD/Shi-scid mice. This revealed that NOG mice were prone to tumor formation. (Table 9)

Table 9

Comparison of tumor formation between NOG mice and NOD/Shi-scid mice to which ED-40515(-) cell type was transplanted				
Mouse strain	No. of mice	Cells inoculated ($\times 10^7$)	Survival period (days)	Tumor
NOG mouse	3	1	15	+++
NOD/Shi-scid mouse	3	1	15	-
+++ tumor large enough to visually observe - tumor which cannot visually be observed				

[0140] For the purpose of investigating whether or not the NOG mice are prone to tumor formation in B-cell type, as well as in T-cell type, 7×10^7 of BJAB cells of which only EBEB (EBV-Encoded Small RNA) and a vector were transformed, were transplanted subcutaneously at a left posterior auricle of NOG mice. In the 3rd week, they were compared in tumorigenicity. A significantly large tumor (26x18x7/13x18x3 mm) was formed in the mice with BJAB-EBEB compared to the mice with BJAB-VECTOR. The tumor size thereof was comparable to that in the case of ED-40515(-) in the above example. (Table 10)

EP 1 338 198 B1

Table 10

Proliferation of BJAB-EBER cell line and BJAB-VECTOR cell line in NOG mice							
Cell line	Gene introduction	No. of mice	Cells inoculated ($\times 10^7$)	Route and site of inoculation	Survival period (days)	Tumor size (mm)	Weight of tumor
BJAB-EBER	gene introduced	3	7	s.c.post-auricular	21	26 $\times$ 18 $\times$ 7	2.86g
BJAB-VECTOR	gene not introduced	3	7	s.c.post-auricular	21	13 $\times$ 18 $\times$ 3	0.73g

[0141] Assuming that effective tumor formation by 10^7 of ED-40515(-) in NOG mice was attributable to active neo-genesis of tumoral vessels via CXCR4 of tumor cells, it has been expected that tumor formation would be inhibited as long as there was continuous administration of KRH-1636, a competitive agent of SDF-1 which was a ligand thereof in endothelial cells. However, in the case of ED-40515(-) and SLB-1 (histologically angiotropic) accompanied by bleeding, the mice formed tumors with sizes of 25 $\times$ 18 $\times$ 12 mm and 20 $\times$ 10 $\times$ 18 mm equivalent to those of non-treated group, even though KRH-1636 was intraperitoneally administered everyday. (Table 11)

Table 11

Effect of CXDR4 antagonists on In Vivo Growth and Proliferation of HTLV-1 infected cell lines in NOG mice							
Cell line	group	No. of mice	Cells inoculated ($\times 10^7$)	Route of drug administration	Survival period (days)	Tumor size (mm)	Remarks
ED-40515 (-)	Drug	3	7.5	intraperitoneally KRH-1636 0.14mg/mouse	15	25 $\times$ 18 $\times$ 12	progressive large tumor
	Control	3	7.5	intraperitoneally medium 0.2ml/ mouse	15	25 $\times$ 18 $\times$ 12	progressive large tumor
SLB-1	Drug	3	7.5	intraperitoneally KRH-1636 0.14mg/mouse	15	20 $\times$ 10 $\times$ 18	progressive large tumor with hemorrhage
	Control	3	7.5	intraperitoneally medium 0.2ml/ mouse	15	20 $\times$ 10 $\times$ 18	progressive large tumor with hemorrhage

[0142] Using Cytospin specimens of in vitro culture cells and frozen section specimens of in vivo tumor formative cells, both from ED-40515(-) and SLB-1, the immunostaining manners of CD4, CD8, CD3, CXCR4, CCR5 and SDF-1 were comparatively tested by enzyme antibody technique. All the cases of CD4, CD3, CXCR4 and SDF-1 were almost equally positive, and all the cases of CCR5 were negative. CD8 was negative in vitro, but positive in vivo. (Table 12)

Table 12

In vitro and in vivo examination of HTLV-1 infected cell line-transplanted NOG mice by FACS, WB, EMSA and immunohistochemistry						
Immunohistochemistry						
	CD4	CD8	CD3	CXCR4	CCR5	SDF-1
In-vitro						
ED-40515(-)	++	-	+	+	-	++

EP 1 338 198 B1

(continued)

In vitro and in vivo examination of HTLV-1 infected cell line-transplanted NOG mice by FACS, WB, EMSA and immunohistochemistry						
Immunohistochemistry						
	CD4	CD8	CD3	CXCR4	CCR5	SDF-1
SLB-1	++	-	+	+	-	++
In-vivo						
ED-40515(-)	+++	+	+	+	-	+++
SLB-1	+++	+	+	+	-	+++
+++ strongly positive ++ positive + weakly positive - negative						

[0143] Using in vitro culture cells and in vivo tumor formative cells of ED-40515(-), Western blotting analyses on Tax, CXCR4, OX40 and OX40L were performed. All the cases of Tax were negative. However, all the cases of CXCR4, OX40 and OX40L were positive, and there was no significant difference in their strengths. (Table 13)

Table 13

In vitro and in vivo examination of HTLV-1 infected cell line-transplanted NOG mice by FACS, WB, EMSA and immunohistochemistry								
WB	In-vitro				In-vivo			
	TAX	CXCR4	OX40	OX40L	TAX	CXCR4	OX40	OX40L
ED-40515 (-)	-	++	+	+	-	+	+	+
SLB-1	++	++	+++	-				

[0144] Using in vitro culture cells and in vivo tumor formative cells of ED-40515(-), transcription factor activity of NFkB was examined by electrophoretic mobility shift assay (EMSA), though no difference therebetween was observed. (Table 14)

Table 14

In vitro and in vivo examination of HTLV-1 infected cell line-transplanted NOG mice by FACS, WB, EMSA and immunohistochemistry			
EMSA	In-vitro		In-vivo
	NFkB		NFkB
ED-40515 (-)	+++		+++
SLB-1	+++		

[0145] The following points were recognized in the system wherein the transplant was conducted subcutaneously at posterior auricles of the NOG mice.

1) It was certainly confirmed that large tumors were formed in the NOG mice within very short periods, like 15 days after innoculation, which had not been expected. Conversely, NOD/Shi-scid mice (IL-2R γ chain^{+/+}) did not form any tumors.

2) It was revealed that the NOG mice genetically delete NK cells, and therefore they did not require the pre-treatment against anti NK cells using monoclonal antibodies etc., which is essential in the case of C.B-17/Icr-scid or NOD/Shi-scid mice. (IL-2R γ chain^{+/+})

3) The transplant was performed subcutaneously at a posterior auricle, that is, selecting a site which has an anatomically lower number of NK cells than an intraperitoneal site, and this allows a tumor to be easily formed. As a result, the size of the tumor was easily observed by appearance without need for incision.

4) Although Uchiyama et al. reported that MT-1, T-2 and TL-Oml did not form any tumors, in the present case relatively small tumors were formed in them. (Imada K, Takaori-Kondo A, Akagi T, Shimotohno K, Sugamura K, Hattori T, Yamabe H, Okuma M, Uchiyama T: Tumorigenicity of human T-cell leukemia virus type I-infected cell lines in severe combined immunodeficient mice and characterization of the cells proliferating in vivo. Blood 86: 2350-7., 1995, Uchiyama, T: Human T cell leukemia virus type I (HTLV-I) and human diseases. Annu Rev Immunol 15:15-37, 1997)

[0146] Therefore, the system wherein the transplant is carried out subcutaneously at posterior auricles of the NOG mice, is an innovative tumor transplant system. Further, it was revealed that it could work in the same manner on B-cell tumors as well as on T-cell type tumors. Namely, these indicate also that this system is a valuable transplant system for the transplant of cancer cells or human normal lymphoid tissues.

[0147] In addition, when there is no laboratory animal model or it is difficult, even if there is, to put one into practical use, a human disease model can be established by transplanting human cells or tissues to this NOG mouse, thereby enabling disease researches, for which there has been no other choice but to be dependent on in vitro tests, with a test system which comes close to a limitless human in vivo test. Thus, this mouse is considered to make great contributions to the elucidation of mechanisms of disease onset, development of therapies etc. For example, irrespective of potent HAART therapy against HIV-1 infection, the rebound or mutant virus emergence of viremia is of primary importance. The reservoir of infectious viruses thereof is known to be FDC (follicular dendritic cells) of lymphatic follicle, and for researches on it, it is indispensable to establish a humanized model mouse to which lymphoid tissues including human lymphatic follicle are transplanted. For this, the present NOG mouse is useful.

[0148] Furthermore, with respect to a cause to make this system prone to form a tumor, examinations were made on what factor is increased or activated in transplanted cells, as compared with in vitro culture cells. All the cell lines used in the above examples were IL-2 independent, IL-2 producing, further Tax and CXCR4-SDF-1 system were revealed not to be directly associated therewith. However, within the search the present inventors have made, there was no factor except the above mentioned which the inventors recognized indicated a significant difference in comparison between in vitro culture cells and in vivo tumor-forming cells.

INDUSTRIAL APPLICABILITY

[0149] The present invention provides a method of producing NOG mice more suitable for engraftment of heterologous cells, particularly human cells, and a mouse produced by the method, as compared with an NOD/Shi-scid mouse and an NOD/LtSz-scid, $\beta 2m$ null mouse both conventionally known as an immunodeficient mouse. Transplanting human stem cells to the thus obtained mice enables a human stem cell assay system to be established. Further, transplanting human cells responsible for immunity enables human antibodies to be produced using the mouse of the present invention. Furthermore, a human tumor model mouse can be produced by transplanting and engrafting human tumors to the mouse, and the mouse can be used for therapies for tumors, screening of therapeutic agents and the like. Moreover, it is possible to produce an HIV or HTLV-1 infected model mouse to which an HIV or HTLV1 infected human lymphocyte is engrafted, and researches can be made on in vivo proliferation mechanism of HIV-1, HTLV-1 etc. Also, it is possible to conduct development of a therapy for virus infection, screening of a therapeutic agent for virus infection or the like.

SEQUENCE LISTING

[0150]

<110> Central Institute for Experimental Animals.

<120> METHOD OF PRODUCING A MOUSE SUITABLE FOR ENGRAFTMENT, DIFFERENTIATION AND PROLIFERATION OF HETEROLOGOUS CELLS, MOUSE PRODUCED BY THIS METHOD AND USE OF THE MOUSE

<130>

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<170> Patent In Ver. 2.0

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 <213> Artificial Sequence

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 <223> Description of Artificial Sequence: Primer PIII.

30

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35

Claims

1. A NOD/Shi-scid, IL-2R- γ KO(NOG) mouse, wherein both of functional T-cells and functional B-cells are deleted, macrophage function is reduced, NK cells or NK activity are eliminated and dendritic cell function is reduced.
2. The NOG mouse according to claim 1, wherein transplanted human stem cells efficiently differentiate and proliferate without being eliminated, with the proviso that the human stem cells are not human embryonic stem cells.
3. The NOG mouse according to claim 2, wherein the human stem cells are human hematopoietic stem cells.
4. A stem cell assay method comprising transplanting human stem cells to the mouse according to any of claims 1 to 3, sacrificing the mouse, collecting the human stem cells from the mouse and analyzing the differentiated and proliferated cells from the transplanted human stem cells, with the proviso that the human stem cells are not human embryonic stem cells.
5. The stem cell assay method according to Claim 4, comprising analyzing the differentiation and proliferation of T-cells and B-cells.
6. A method of proliferating human stem cells comprising:
 - transplanting and proliferating the human stem cells in the mouse according to any one of claims 1 to 3;
 - sacrificing the mouse;
 - collecting human stem cells from bone marrow of the mouse; and
 - repeatedly transplanting the human stem cells to the mouse according to claims 1 to 3, with the proviso that

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the human stem cells are not human embryonic stem cells.

- 5
7. The method of proliferating human stem cells according to claim 6, wherein the frequency of repeating is at least three times.
8. The method according to claim 6 or 7, wherein the human stem cell has a foreign gene introduced therein.
9. A human tumor model mouse, wherein the mouse is a mouse according to any one of claims 1 to 3 and retains a human tumor cell.
- 10
10. The human tumor model mouse according to Claim 9, wherein the human tumor cell is derived from HTLV-1 leukemia.
11. The human tumor model mouse according to claim 9 or 10, wherein the mouse has the human tumor cell at an auricle thereof.
- 15
12. A method of producing a human tumor model mouse comprising transplanting human tumor cells to the mouse according to any one of claims 1 to 3 and sacrificing said mouse prior to analysis.
13. The method according to claim 12, wherein the human tumor cells are derived from HTLV-1 leukemia.
- 20
14. The method according to claim 12 or 13, wherein the human tumor cells are transplanted at an auricle of the mouse.
15. A virus-infected model mouse wherein the mouse is a mouse according to any one of claims 1 to 3 and retains a T-cell infected with a T-tropic virus as well as a macrophage-tropic virus.
- 25
16. The virus-infected model mouse according to claim 15, wherein the virus is HIV.
17. The virus-infected model mouse according to claim 15, wherein the virus is HTLV-1.
- 30

Patentansprüche

- 35
1. NOD/Shi-scid,IL-2R γ KO(NOG)-Maus, bei der sowohl die funktionsfähigen T-Zellen als auch die funktionsfähigen B-Zellen deletiert sind, die Makrophagenfunktion reduziert ist, die NK-Zellen oder die NK-Aktivität eliminiert sind und die Funktion der dendritischen Zellen reduziert ist.
2. NOG-Maus nach Anspruch 1, bei der transplantierte humane Stammzellen sich effizient differenzieren und proliferieren, ohne eliminiert zu werden, mit der Maßgabe, daß die humanen Stammzellen keine humanen embryonalen Stammzellen sind.
- 40
3. NOG-Maus nach Anspruch 2, wobei die humanen Stammzellen humane hämatopoietische Stammzellen sind.
4. Stammzelltestverfahren, bei dem man der Maus nach einem der Ansprüche 1 bis 3 humane Stammzellen transplantiert, die Maus tötet, die humanen Stammzellen aus der Maus sammelt und die differenzierten und proliferierten Zellen von den transplantierten humanen Stammzellen analysiert, mit der Maßgabe, daß die humanen Stammzellen keine humanen embryonalen Stammzellen sind.
- 45
5. Stammzelltestverfahren nach Anspruch 4, bei dem man die Differenzierung und Proliferation von T-Zellen und B-Zellen analysiert.
- 50
6. Verfahren zum Proliferieren von humanen Stammzellen, bei dem man:
- der Maus nach einem der Ansprüche 1 bis 3 die humanen Stammzellen transplantiert und dort proliferiert, die Maus tötet,
- 55
- die humanen Stammzellen aus dem Knochenmark der Maus sammelt und wiederholt der Maus nach einem der Ansprüche 1 bis 3 die humanen Stammzellen transplantiert, mit der Maßgabe, daß die humanen Stammzellen keine humanen embryonalen Stammzellen sind.

EP 1 338 198 B1

7. Verfahren zum Proliferieren von humanen Stammzellen nach Anspruch 6, wobei die Häufigkeit der Wiederholung weniger als dreimal beträgt.
8. Verfahren nach Anspruch 6 oder 7, wobei die humane Stammzelle ein darin eingebrachtes Fremdgen aufweist.
9. Modellmaus für einen humanen Tumor, wobei die Maus eine Maus nach einem der Ansprüche 1 bis 3 ist und eine humane Tumorzelle enthält.
10. Modellmaus für einen humanen Tumor nach Anspruch 9, wobei die humane Tumorzelle von HTLV-1-Leukämie abgeleitet ist.
11. Modellmaus für einen humanen Tumor nach Anspruch 9 oder 10, wobei die Maus die humane Tumorzelle an einer Auricula davon aufweist.
12. Verfahren zur Herstellung einer Modellmaus für einen humanen Tumor, bei dem man der Maus nach einem der Ansprüche 1 bis 3 humane Tumorzellen transplantiert und diese Maus vor der Analyse tötet.
13. Verfahren nach Anspruch 12, wobei die humanen Tumorzellen von HTLV-1-Leukämie abgeleitet sind.
14. Verfahren nach einem der Ansprüche 12 oder 13, bei dem die humanen Tumorzellen an eine Auricula der Maus transplantiert werden.
15. Virusinfizierte Modellmaus, wobei die Maus eine Maus nach einem der Ansprüche 1 bis 3 ist und eine sowohl mit einem T-tropistischen Virus als auch mit einem makrophagentropistischen Virus infizierte T-Zelle enthält.
16. Virusinfizierte Modellmaus nach Anspruch 15, wobei der Virus HIV ist.
17. Virusinfizierte Modellmaus nach Anspruch 15, wobei der Virus HTLV-1 ist.

Revendications

1. Souris NOD/Shi-scid, IL-2R γ KO (NOG) dans laquelle à la fois les lymphocytes T fonctionnels et les lymphocytes B fonctionnels sont supprimés, une fonction macrophage est réduite, des cellules NK ou l'activité NK sont éliminées et une fonction des cellules dendritiques est réduite.
2. Souris NOG selon la revendication 1, dans laquelle des cellules souche humaines transplantées se différencient de façon efficace et prolifèrent sans être éliminées, à condition que les cellules souche humaines ne soient pas des cellules souche embryonnaires humaines.
3. Souris NOG selon la revendication 2, dans laquelle les cellules souche humaines sont des cellules souche hématopoïétiques humaines.
4. Procédé d'analyse des cellules souche comprenant la transplantation de cellules souche humaines chez la souris selon l'une quelconque des revendications 1 à 3, le sacrifice de la souris, la récupération des cellules souche humaines provenant de cette souris et l'analyse des cellules ayant proliféré et s'étant différenciées par rapport aux cellules souche humaines transplantées, à condition que les cellules souche humaines ne soient pas des cellules souche embryonnaires humaines.
5. Procédé d'analyse des cellules souche selon la revendication 4, comprenant l'analyse de la différenciation et de la prolifération des lymphocytes T et des lymphocytes B.
6. Procédé de prolifération des cellules souche humaines comprenant :
 - la transplantation et la prolifération des cellules souche humaines chez la souris, selon l'une quelconque des revendications 1 à 3 ;
 - le sacrifice de la souris ;
 - la récupération des cellules souche humaines à partir de la moelle osseuse de la souris ; et

EP 1 338 198 B1

la transplantation répétitive des cellules souche humaines chez la souris selon les revendications 1 à 3, à condition que les cellules souche humaines ne soient pas des cellules souche embryonnaires humaines.

- 5 **7.** Procédé de prolifération des cellules souche humaines selon la revendication 6, dans lequel la fréquence de répétition est d'au moins trois fois.
- 8.** Procédé selon la revendication 6 ou 7, dans lequel la cellule souche humaine présente un gène étranger introduit en son sein.
- 10 **9.** Souris modèle portant une tumeur humaine, la souris étant une souris selon l'une quelconque des revendications 1 à 3 et retenant une cellule tumorale humaine.
- 10.** Souris modèle portant une tumeur humaine selon la revendication 9, dans laquelle la cellule tumorale humaine est dérivée de la leucémie HTLV-1.
- 15 **11.** Souris modèle portant une tumeur humaine selon la revendication 9 ou 10, la souris présentant la cellule tumorale humaine au niveau de l'auricule.
- 12.** Procédé de production d'une souris modèle portant une tumeur humaine comprenant la transplantation de cellules tumorales humaines chez la souris selon l'une quelconque des revendications 1 à 3 et le sacrifice de ladite souris avant analyse.
- 20 **13.** Procédé selon la revendication 12, dans lequel les cellules tumorales humaines sont dérivées de la leucémie HTLV-1.
- 14.** Procédé selon la revendication 12 ou 13, dans lequel les cellules tumorales humaines sont transplantées au niveau de l'auricule de la souris.
- 25 **15.** Souris modèle infectée par un virus, la souris étant une souris selon l'une quelconque des revendications 1 à 3 et retenant un lymphocyte T infecté par un virus T-tropique ainsi que par un virus tropique par macrophage.
- 30 **16.** Souris modèle infectée par un virus selon la revendication 15, dans laquelle le virus est le VIH.
- 17.** Souris modèle infectée par un virus selon la revendication 15, dans laquelle le virus est HTLV-1.

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FIG.1

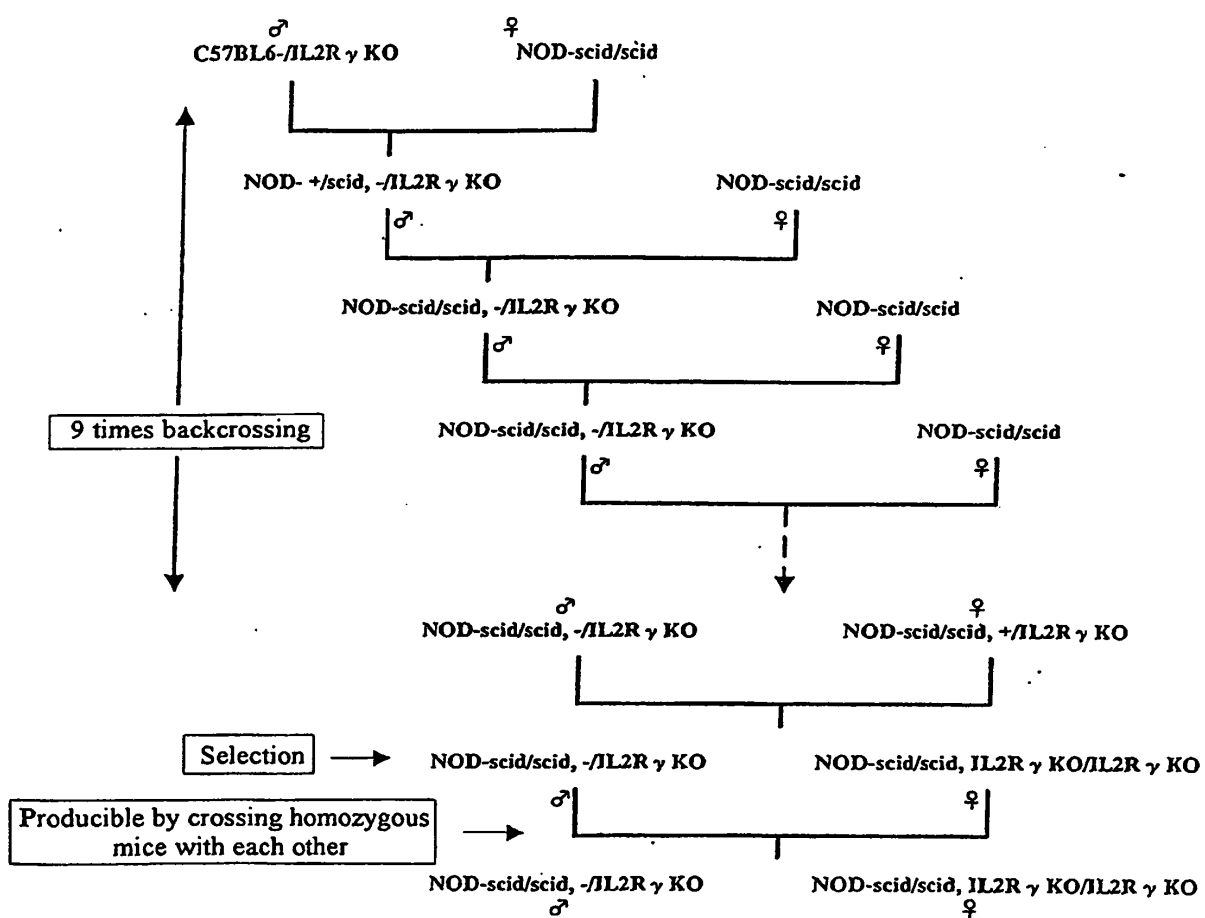


FIG.2A

Posttrans- -plantation	mouse	hCD45 ⁺ (%)	hCD45 ⁺ (μ l)	hCD41 ⁺ (%)	hCD41 ⁺ (μ l)
4th week	I 8 mice Mean	1.345	12.503	0.037	387.087
	SE	0.281	2.683	0.013	144.057
	II 8 mice Mean	8.585	79.116	0.384	3096.365
	SE	1.668	21.701	0.138	647.664
8th week	I 8 mice Mean	6.623	81.513	0.086	590.814
	SE	3.097	33.896	0.053	349.314
	II 8 mice Mean	37.152	533.456	0.380	2422.976
	SE	7.324	99.283	0.112	547.911
12th week	I 7 mice Mean	5.860	61.999	2.064	1510.084
	SE	2.395	22.302	1.308	769.132
	II 6 mice Mean	24.086	530.097	2.586	2227.427
	SE	5.789	281.806	0.600	466.970
Mouse I: NOD/Shi-scid + antiAGN1					
Mouse II: NOG					

FIG.2B

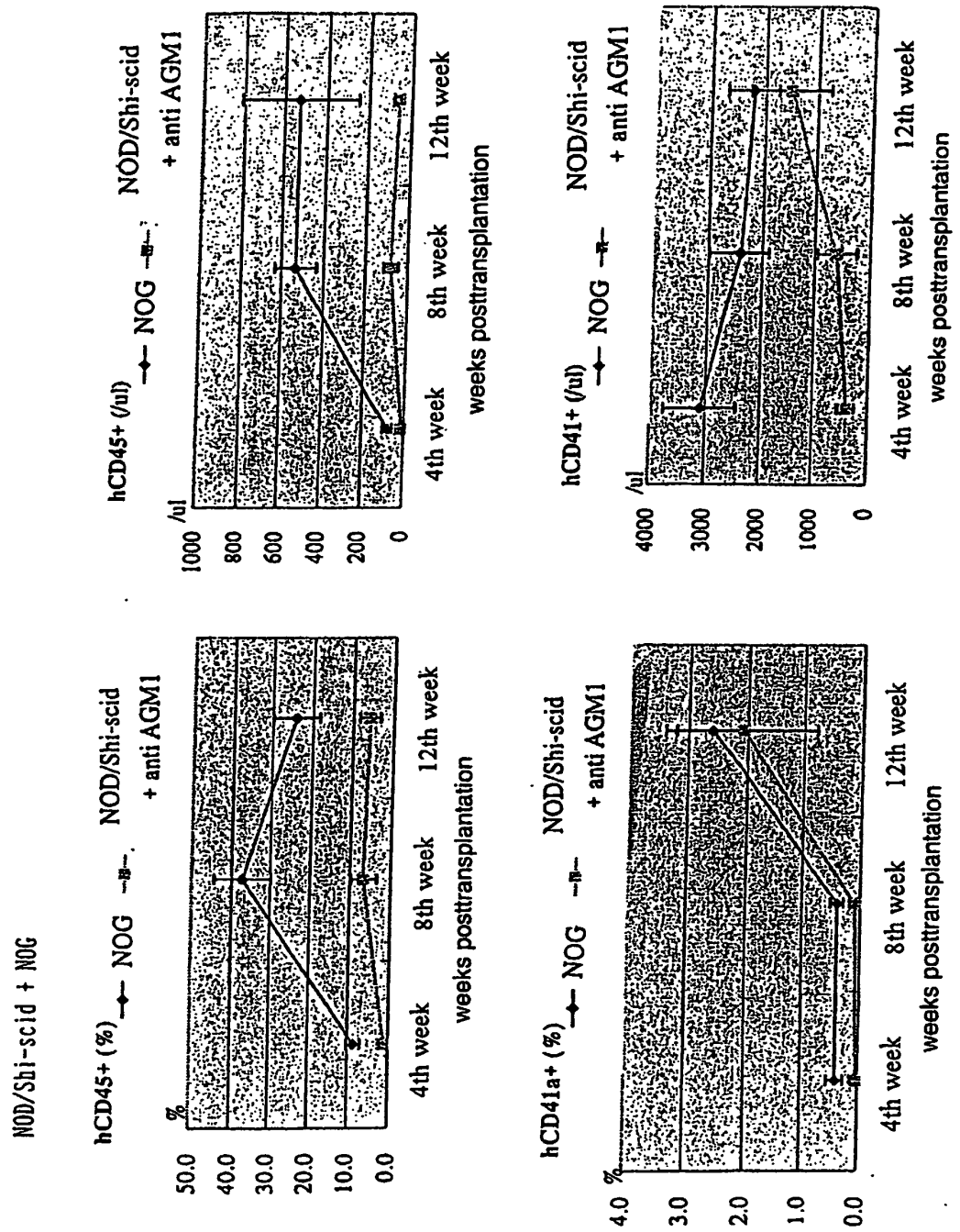


FIG.3



FIG.4

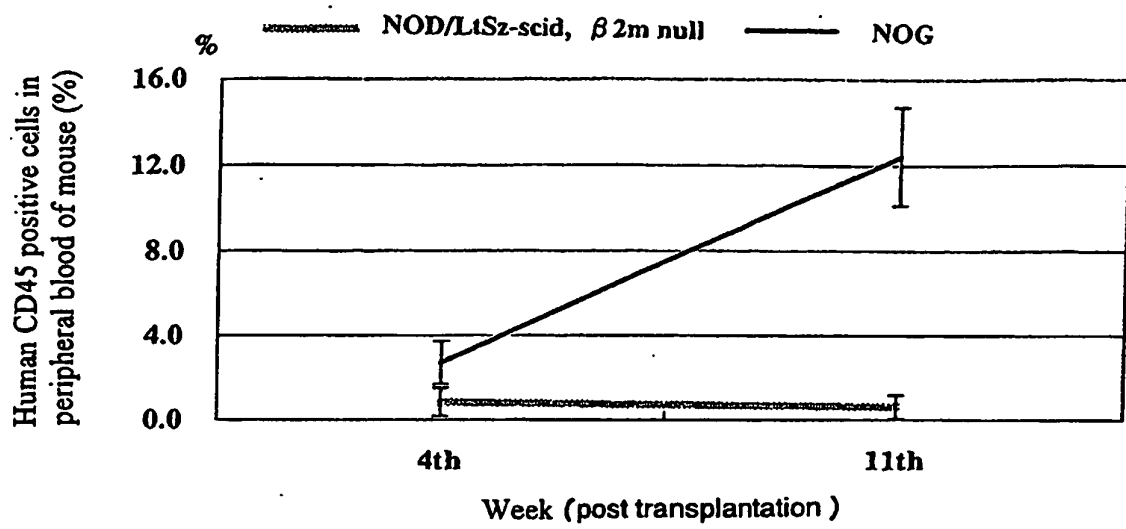


FIG.5A

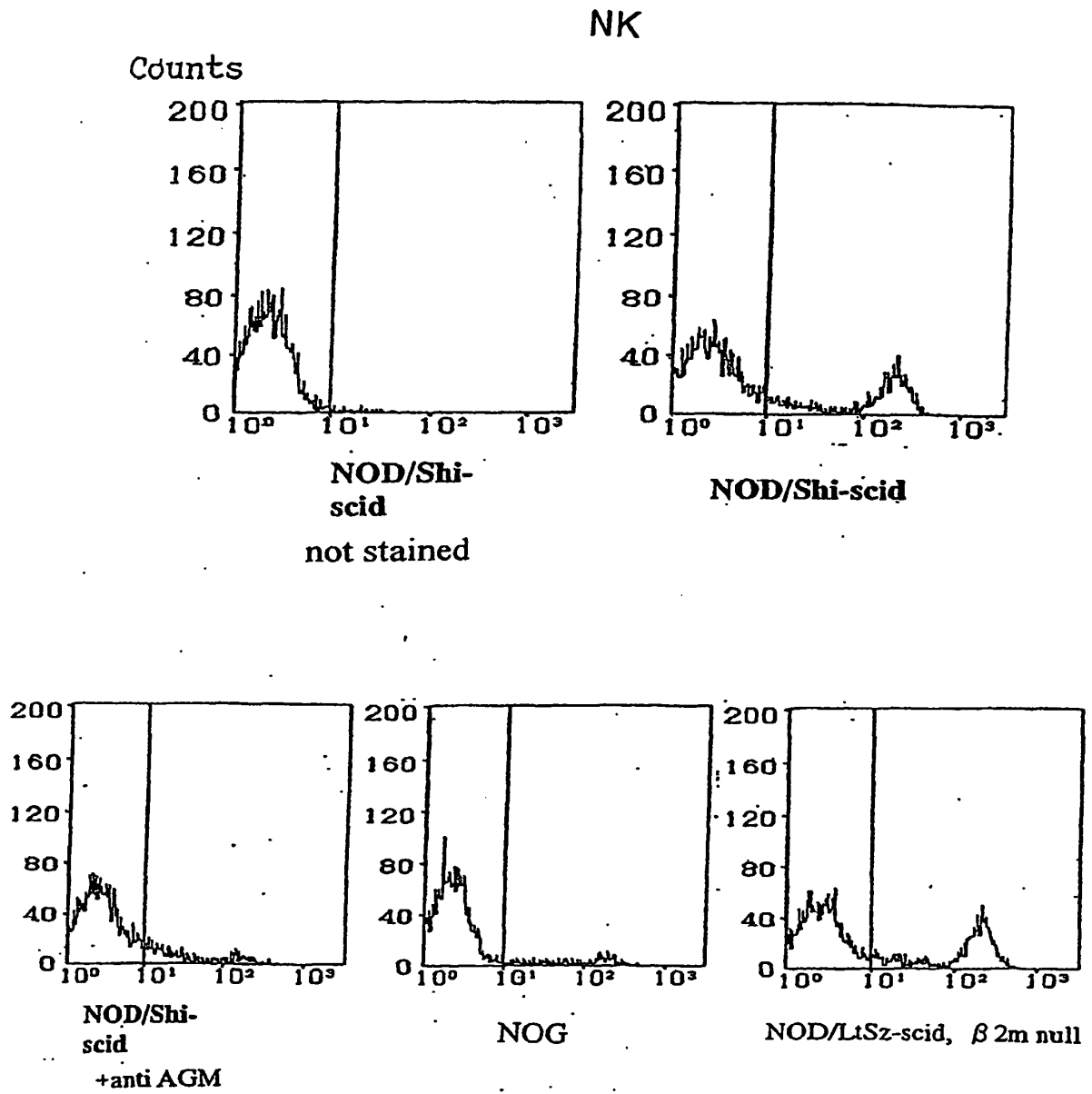


FIG.5B

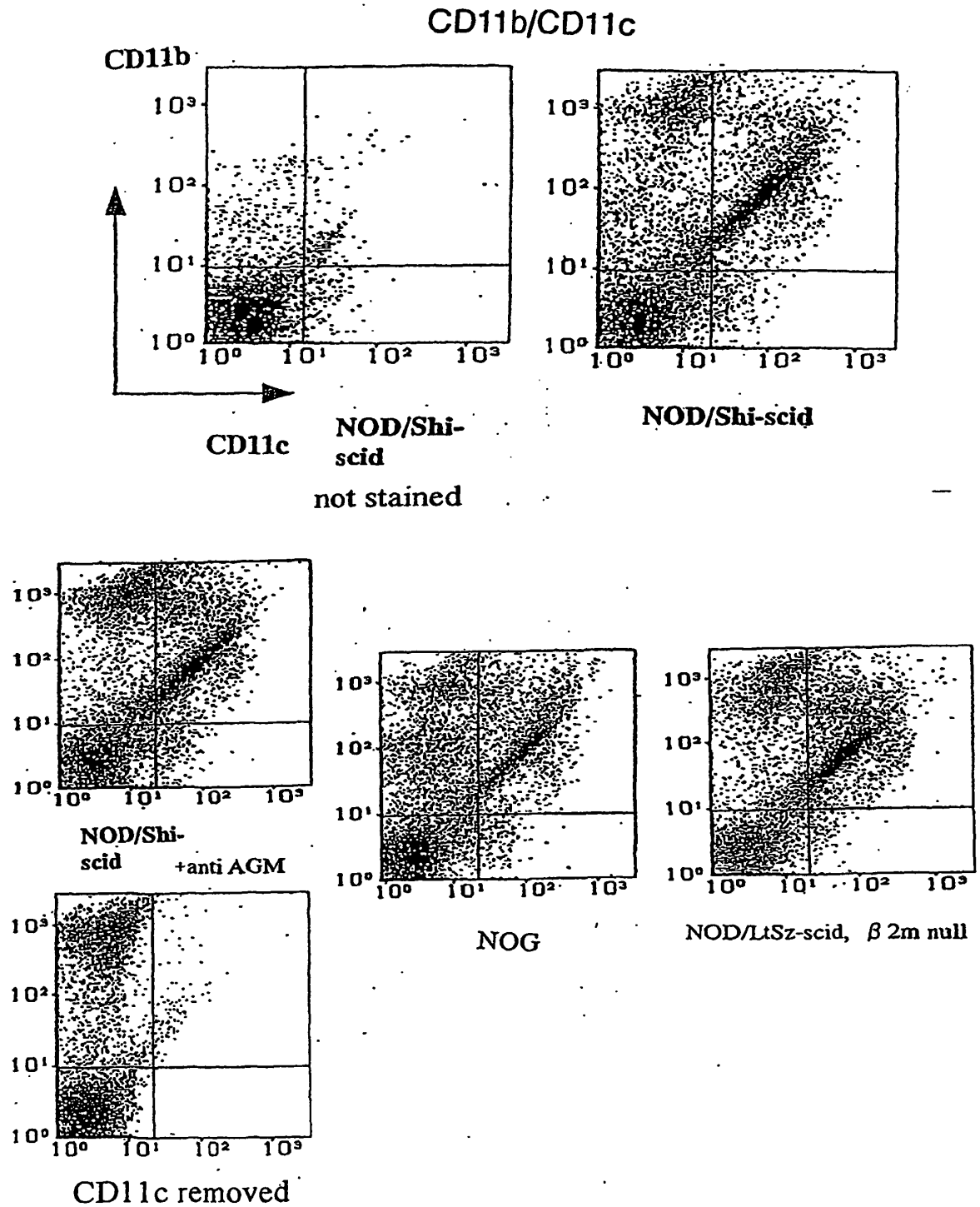


FIG.6A

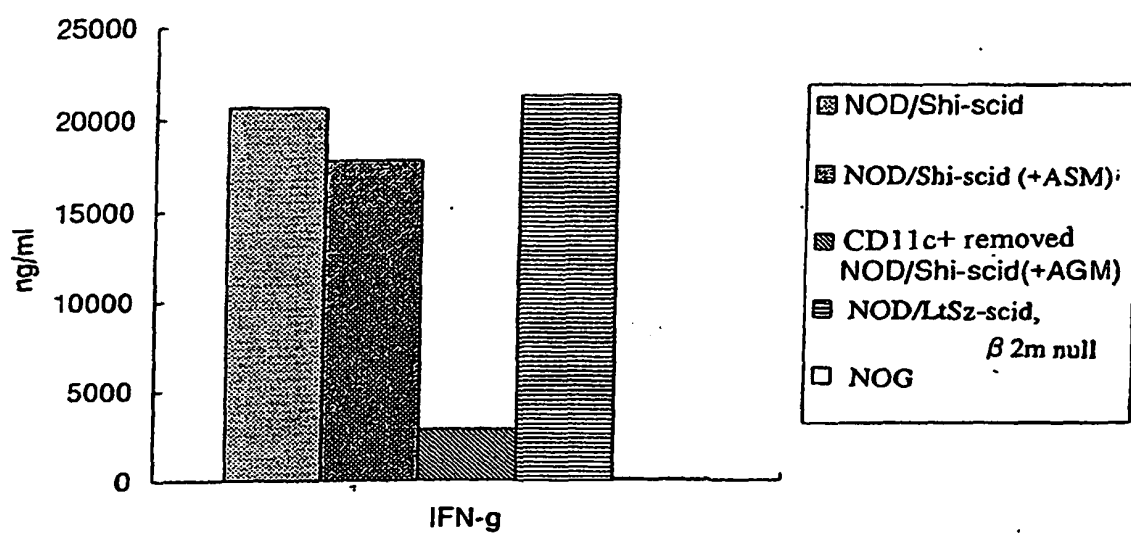


FIG.6B

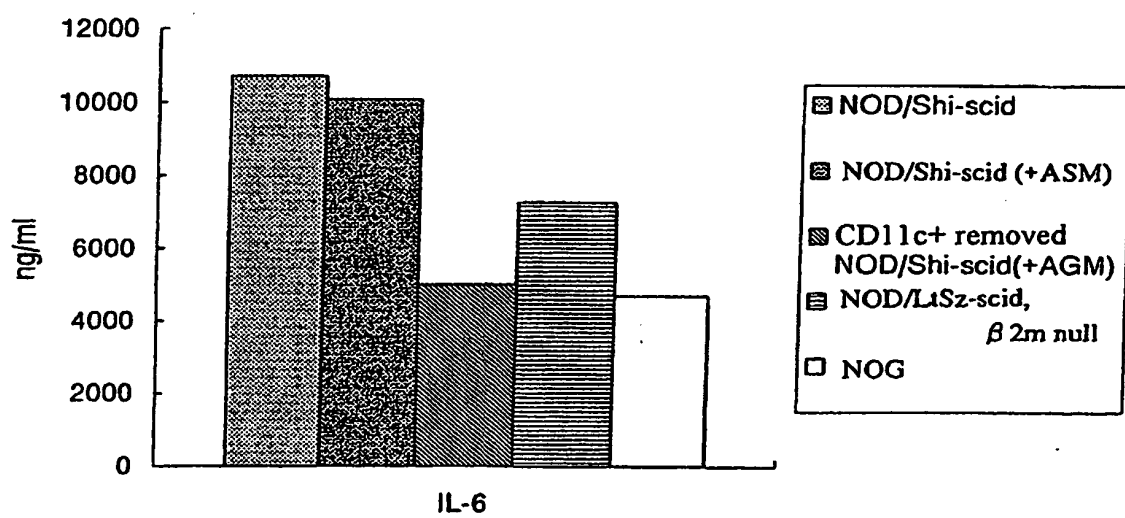


FIG.6C

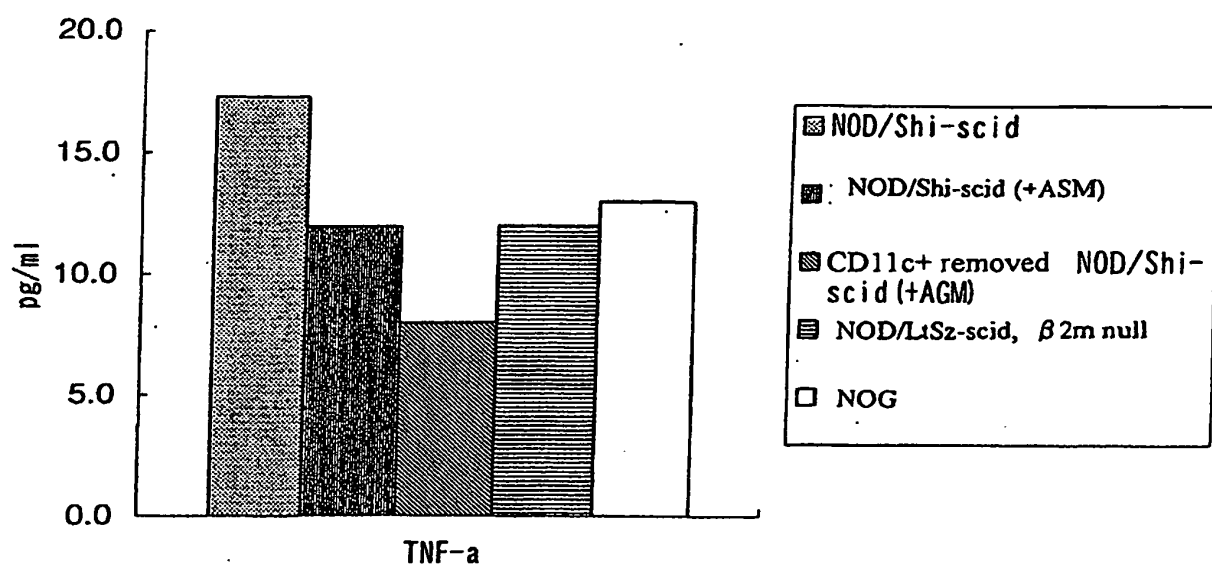


FIG. 7

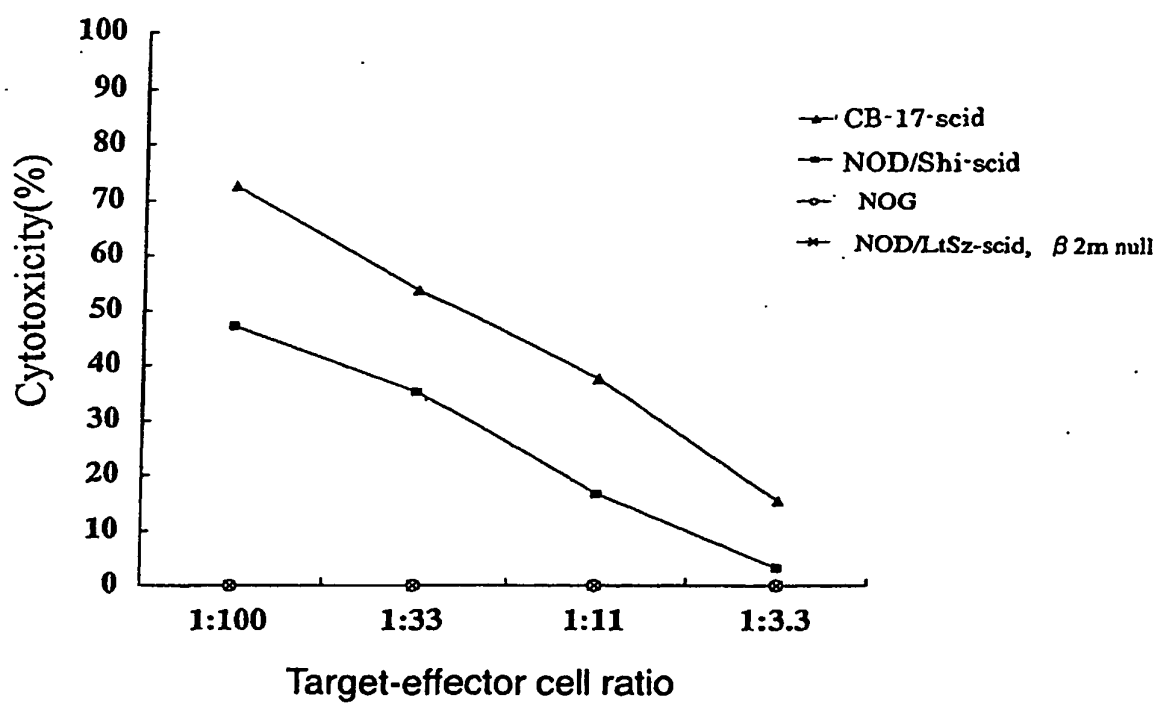


FIG.8

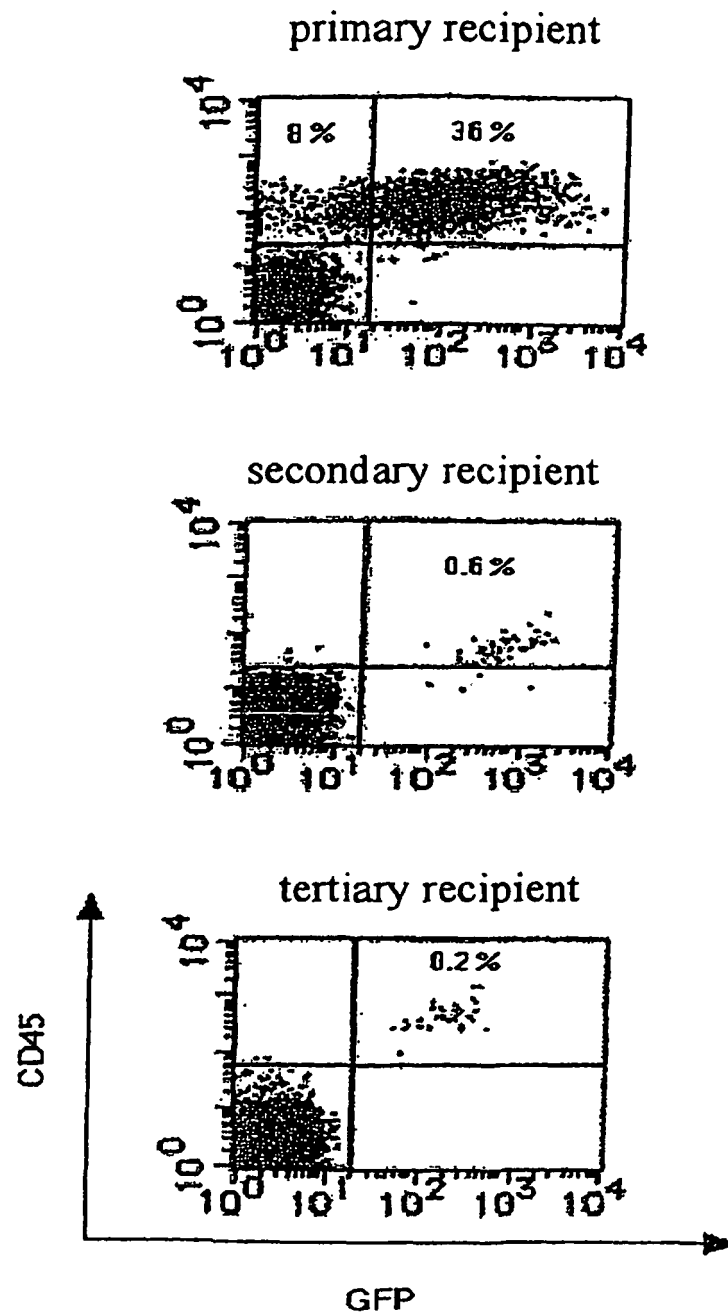


FIG. 9A Flowcytometric Analysis of thymic cells

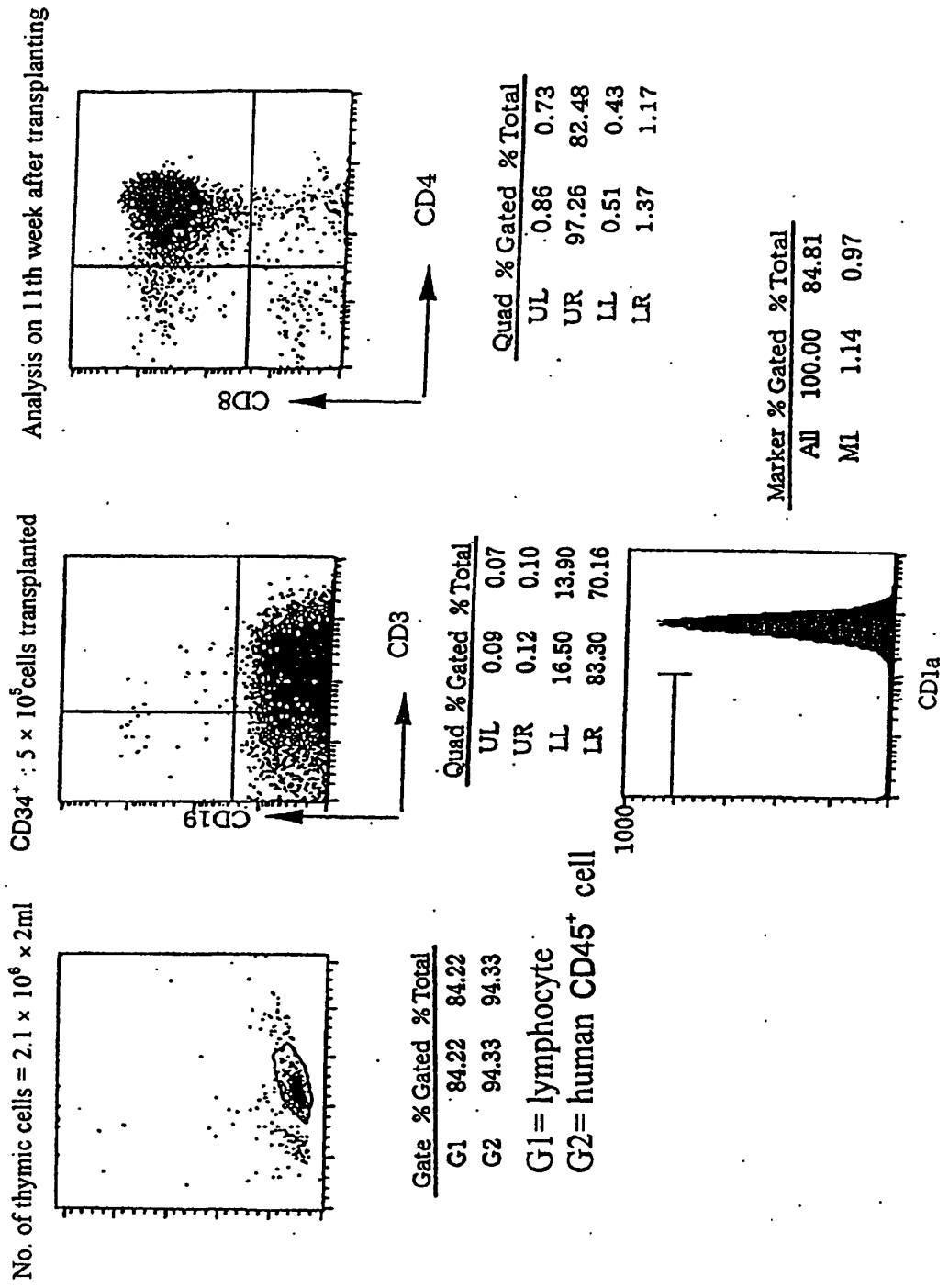


FIG.9B

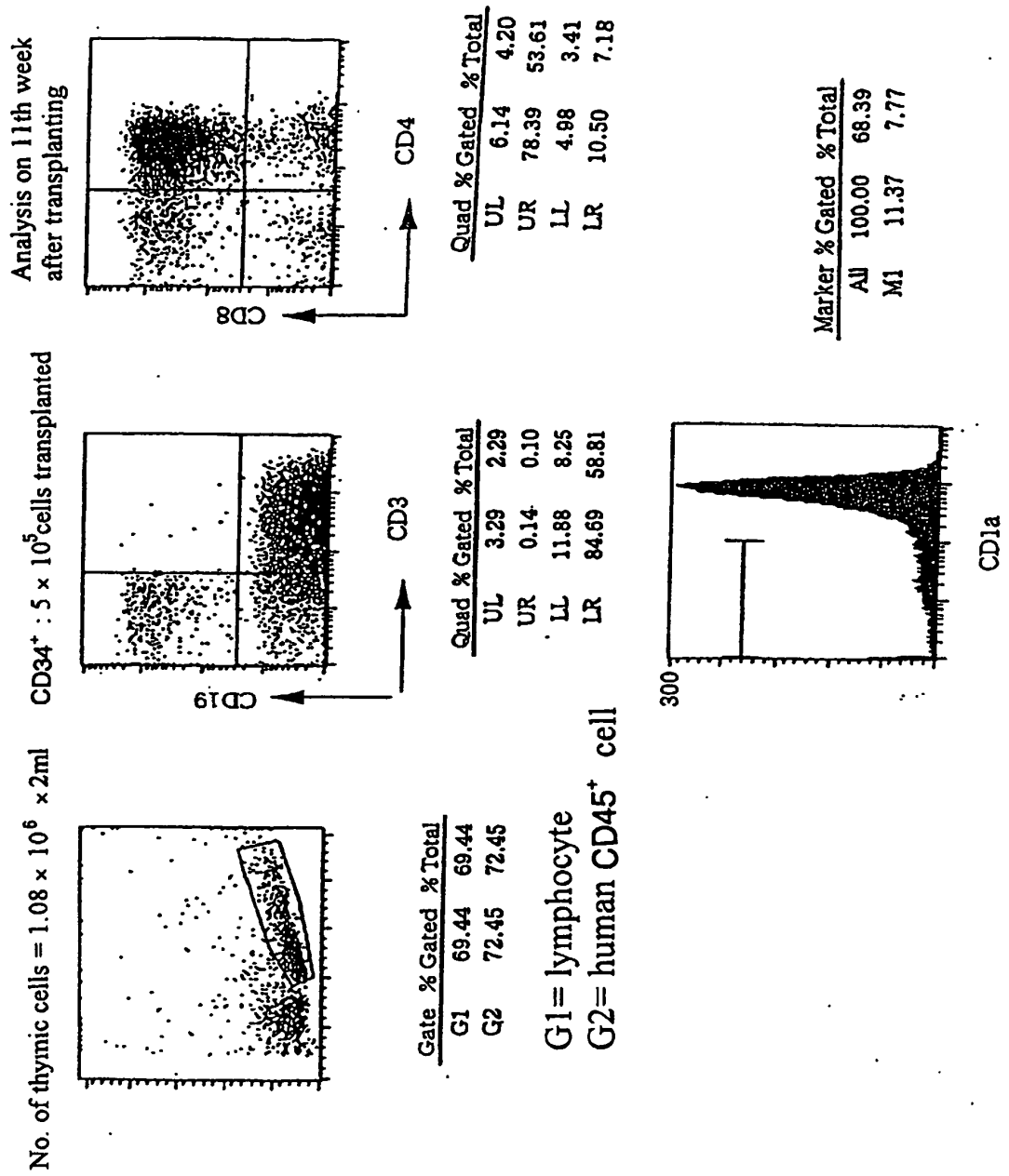


FIG.10A

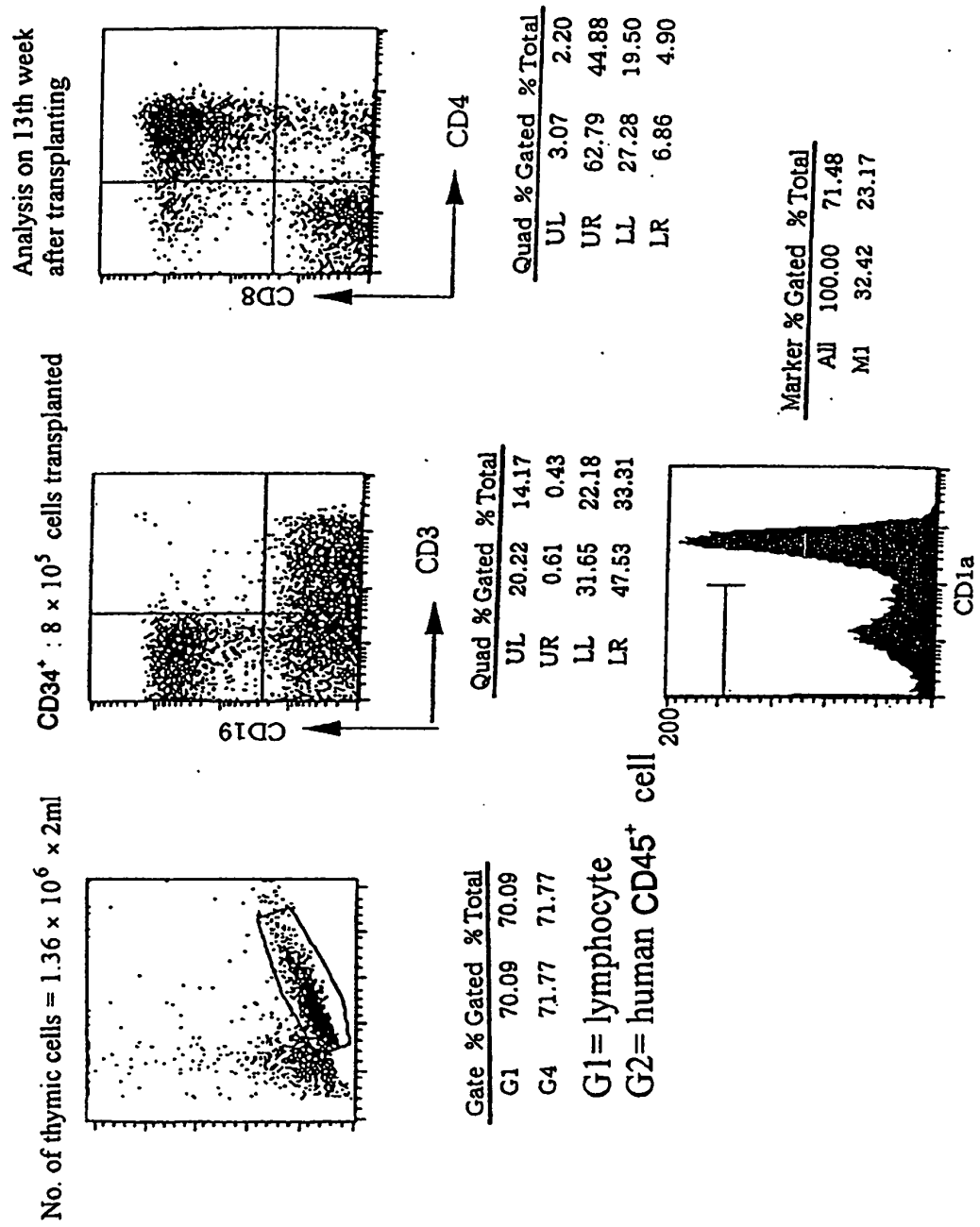


FIG.10B

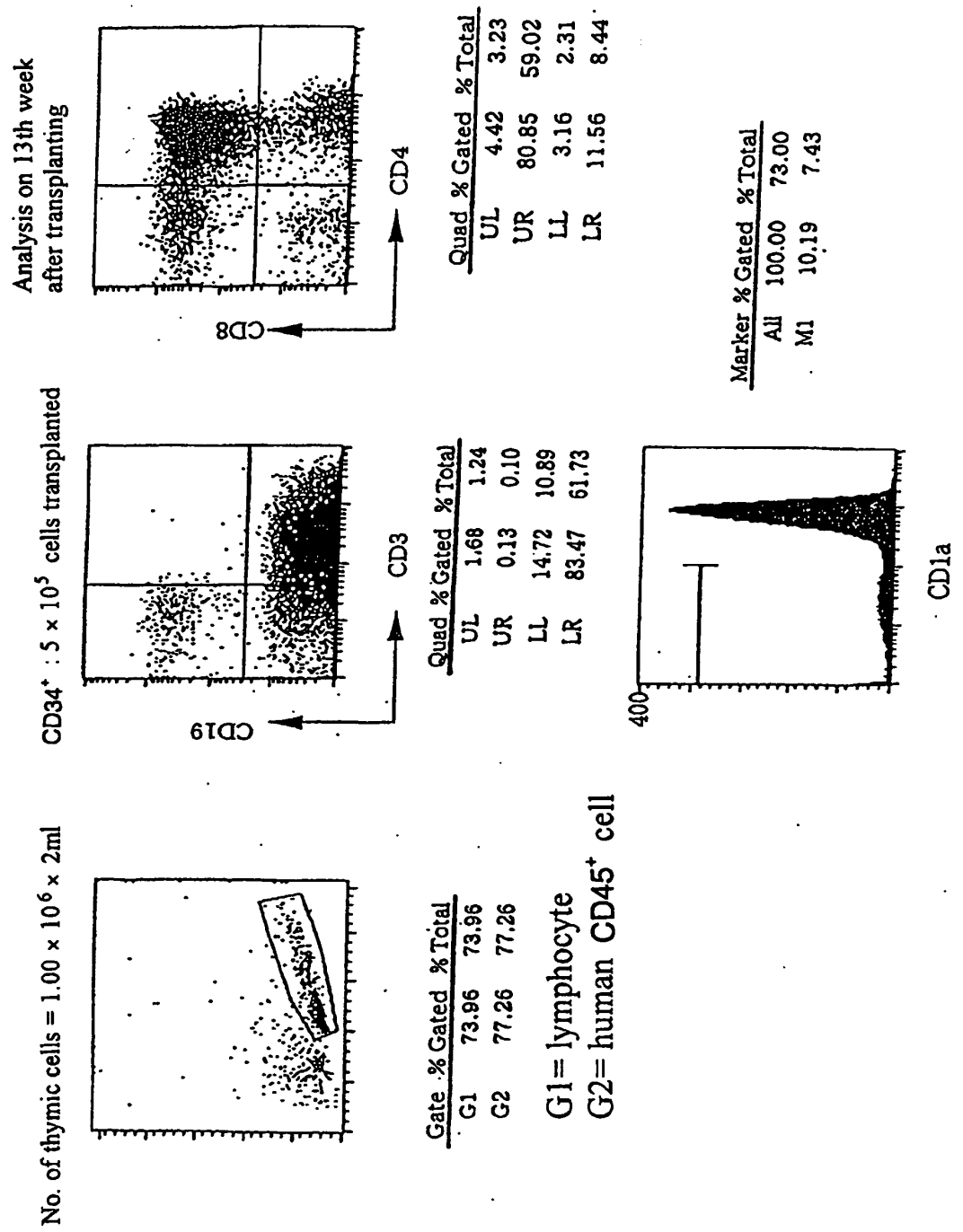
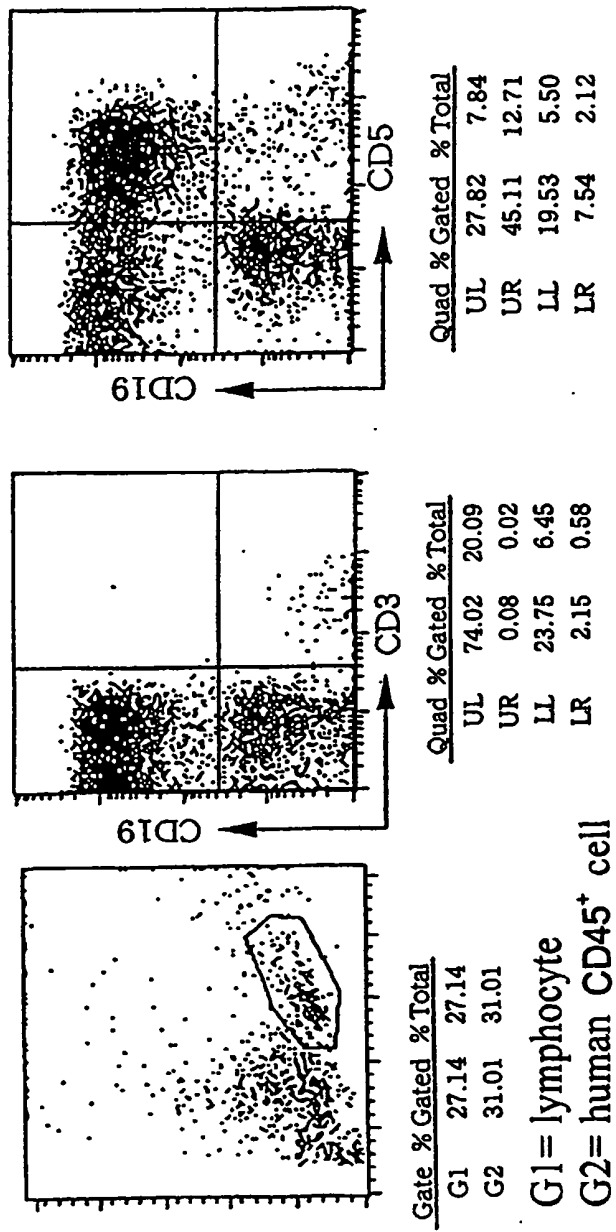


FIG.11A

Flowcytometric Analysis of spleen cells

No. of spleen cells = $5.02 \times 10^6 \times 5\text{ml} \times 2$



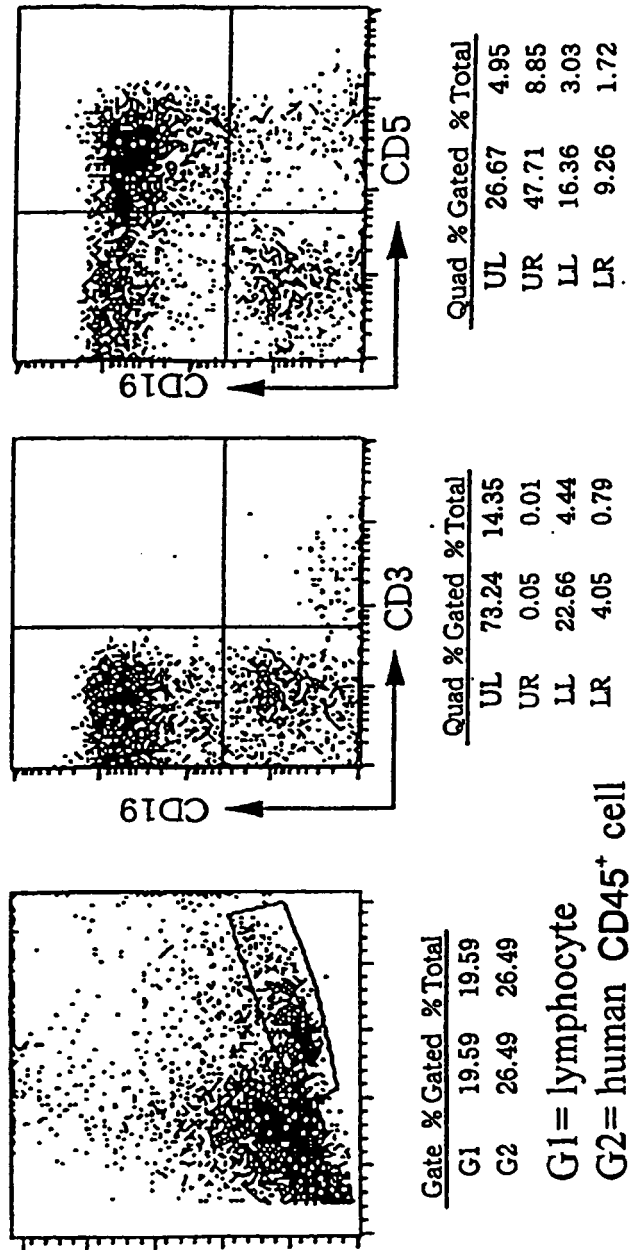
CD34⁺ : 5×10^5 cells transplanted

Analysis on 11th week
after transplanting

FIG.11B

Flowcytometric Analysis of thymic cells

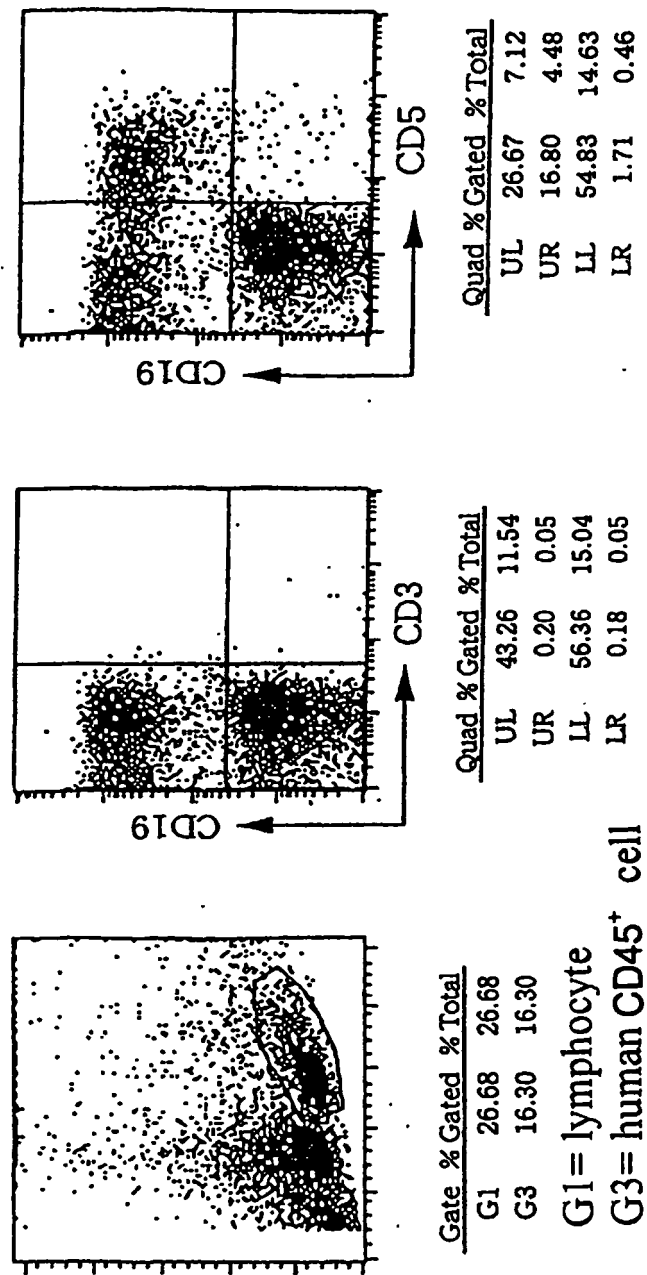
No. of spleen cells = $2.48 \times 10^6 \times 5\text{ml} \times 2$



CD34⁺ : 5×10^5 cells transplanted Analysis on 11th week after transplanting

FIG.12A

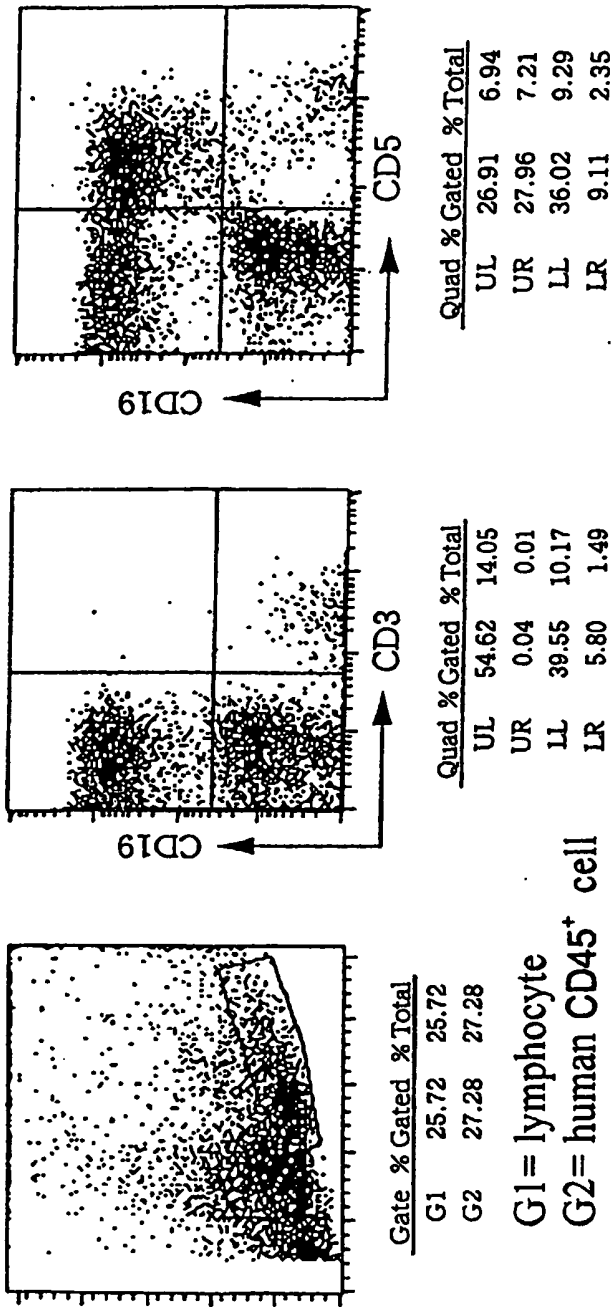
No. of spleen cells = $2.52 \times 10^6 \times 5\text{ml} \times 2$



CD34⁺ : 8×10^5 cells transplanted Analysis on 13th week after transplanting

FIG.12B

No. of spleen cells = $2.7 \times 10^6 \times 10\text{ml} \times 2$



CD34⁺ : 5×10^5 cells transplanted Analysis on 13th week after transplanting

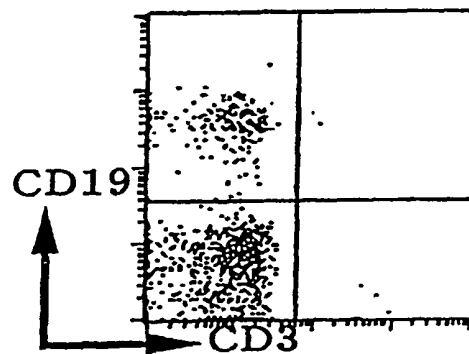
FIG.13A

Flowcytometric Analysis of peripheral blood

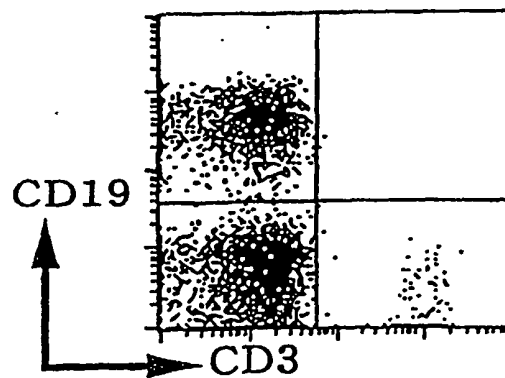
G1= lymphocyte

G2= human CD45⁺ cell

11th week after transplanting CD34



Quad	% Gated	% Total
UL	21.46	0.22
UR	0.10	0.00
LL	77.97	0.81
LR	0.48	0.01



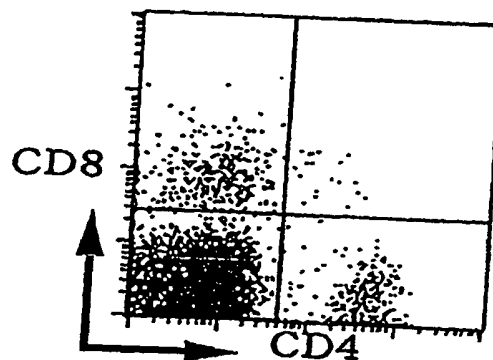
Quad	% Gated	% Total
UL	36.19	1.75
UR	0.06	0.00
LL	60.66	2.93
LR	3.09	0.15

FIG.13B

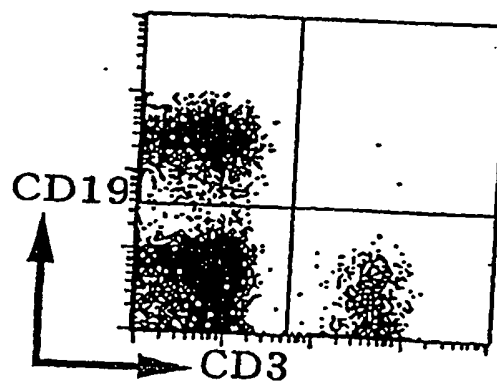
Flowcytometric Analysis of peripheral blood

G1= lymphocyte
G2= human CD45⁺ cell

13th week after transplanting CD34



Quad	% Gated	% Total
UL	3.77	0.67
UR	0.18	0.03
LL	90.34	16.12
LR	5.72	1.02

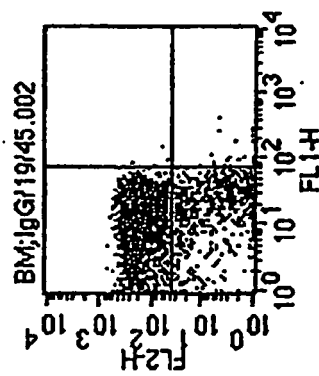


Quad	% Gated	% Total
UL	31.41	3.40
UR	0.02	0.00
LL	55.11	5.97
LR	13.46	1.46

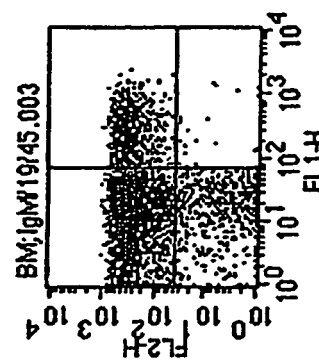
FIG.14A

Flowcytometric Analysis of bone marrow cells

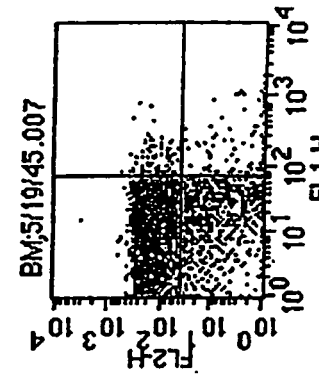
EP 1 338 198 B1



Quad Events % Gated % Total			
UL	7516	78.55	15.03
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LL	1998	20.88	4.00
LR	50	0.52	0.10

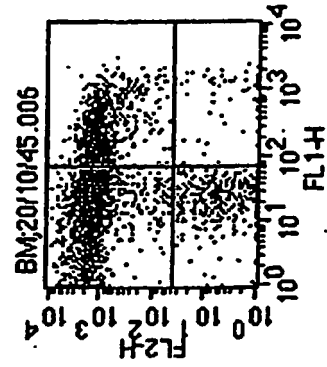


Quad Events % Gated % Total			
UL	6210	66.47	12.42
UR	1567	16.77	3.13
LL	1503	16.09	3.01
LR	62	0.66	0.12

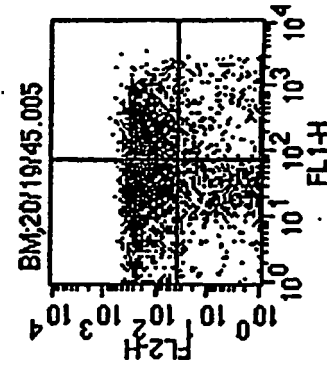


Quad Events % Gated % Total			
UL	7174	75.35	14.35
UR	204	2.14	0.41
LL	1960	20.59	3.92
LR	183	1.92	0.37

FIG. 14B



Quad	Events	% Gated	% Total
UL	5640	62.88	11.28
UR	2379	26.52	4.76
LL	813	9.06	1.63
LR	137	1.53	0.27



Quad	Events	% Gated	% Total
UL	4347	47.17	8.69
UR	2683	29.12	5.37
LL	1545	16.77	3.09
LR	640	6.95	1.28

FIG.15A

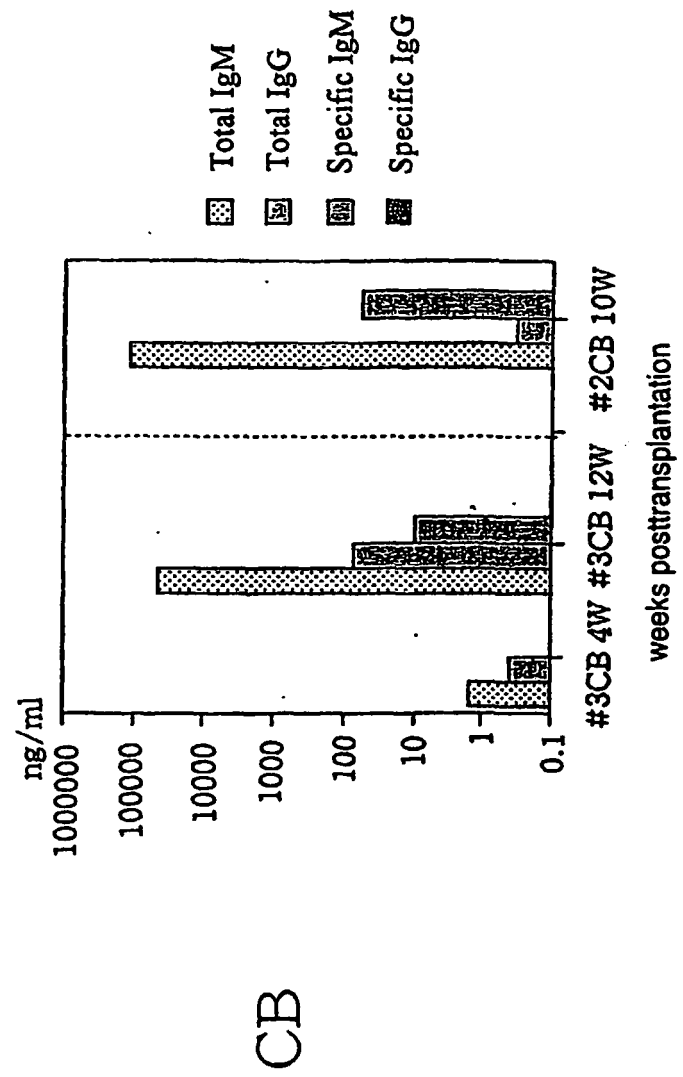


FIG. 15B

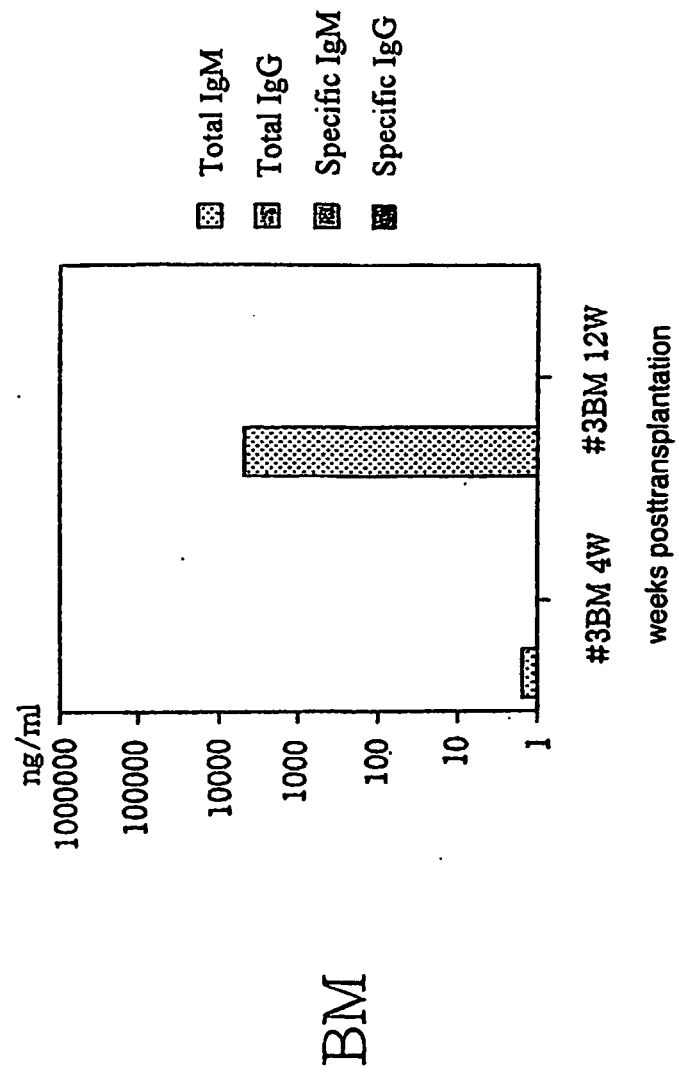


FIG.15C

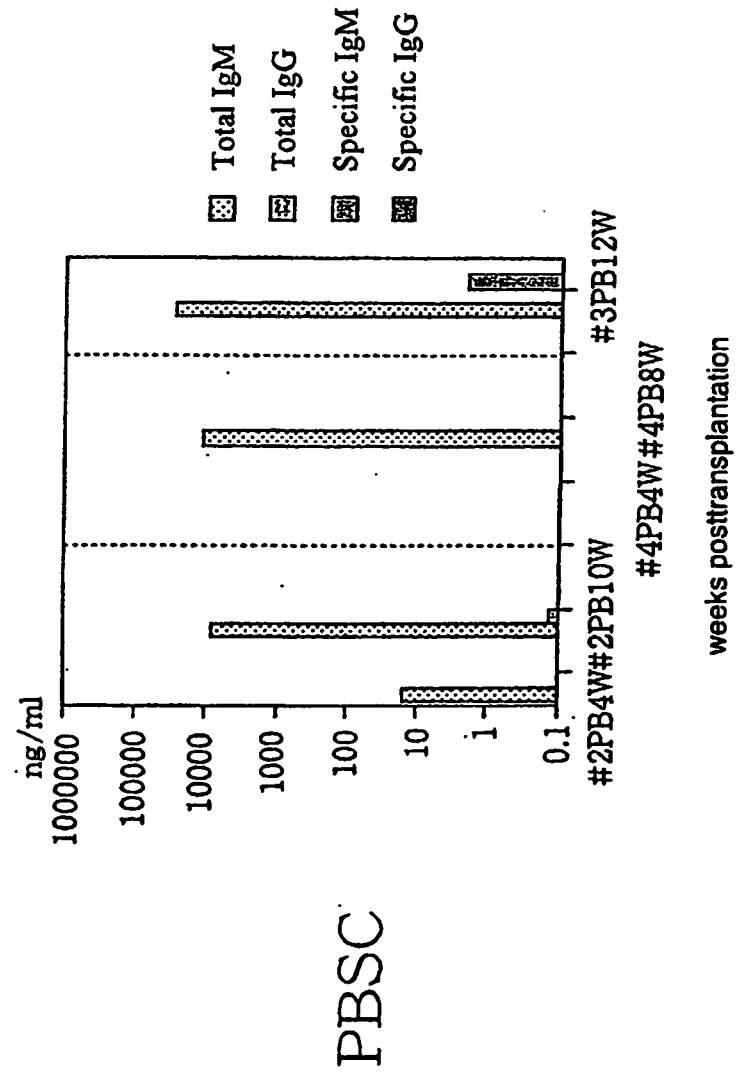
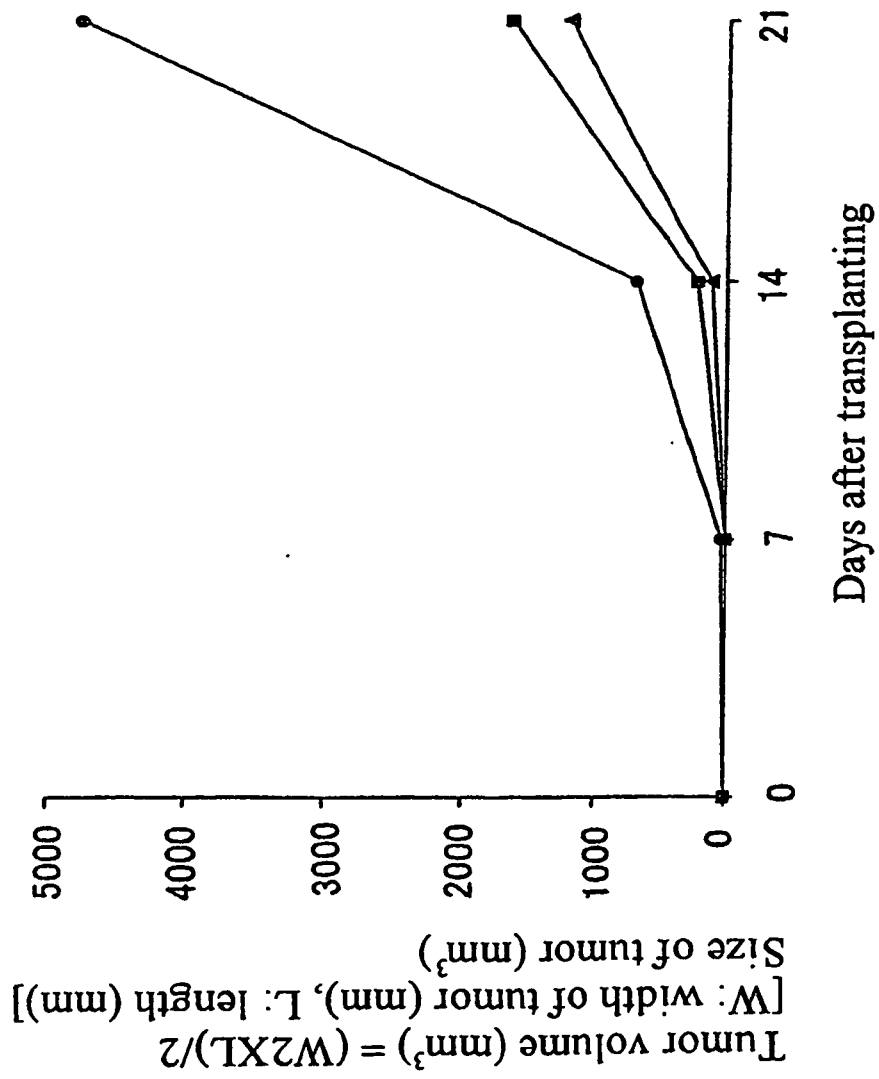


FIG.16



NOG : ●, NOD/Shi-scid : ■, C.B-17/Icr-scid : ▲.

REFERENCES CITED IN THE DESCRIPTION

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