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(54) Title: SINGLE B-CELL CULTIVATION METHOD

(57) Abstract: Herein is reported a method for obtaining a B-cell comprising the following steps a) labeling B-cells, b) depositing the labeled B-cells as single cells, c) co-cultivating the single cell deposited B-cells with feeder cells, d) selecting a B-cell proliferating and secreting IgG in step c) and thereby obtaining a B-cell. The labeling can be of IgG⁺CD19⁺-B-cells, IgG⁺CD38⁺-B-cells, IgG⁺CD268⁺-B-cells, IgG⁺CD138⁺-B-cells, CD27⁺CD138⁺-B-cells or CD3⁺CD27⁺-B-cells. The method can comprise the step of incubating said B-cells at 37 °C for one hour in EL-4 B5 medium prior to the depositing step. The method can also comprise the step of centrifuging said single cell deposited B-cells prior to the co-cultivation. In the co-cultivation a feeder mix comprising interleukin-1beta, and tumor necrosis factor alpha and *Staphylococcus aureus* strain Cowans cells or BAFF or interleukin-2 and/or interleukin-10 and/or interleukin-6 and/or interleukin-4 can be used.

Single B-cell cultivation method

Herein is reported a method for obtaining the amino acid sequence of at least the variable domains of a monoclonal antibody secreted by a single B-cell that has been obtained from a population of B-cells from an experimental animal by single cell deposition and co-cultivation with feeder cells in the presence of a feeder mix.

5 Background of the Invention

For obtaining cells secreting monoclonal antibodies the hybridoma technology developed by Kohler and Milstein is widely used. But in the hybridoma technology only a fraction of the B-cells obtained from an immunized experimental animal can be fused and propagated. The source of the B-cells is generally an organ 10 of an immunized experimental animal such as the spleen.

Zubler et al. started in 1984 to develop a different approach for obtaining cells secreting monoclonal antibodies (see e.g. Eur. J. Immunol. 14 (1984) 357-63, J. Exp. Med. 160 (1984) 1170-1183). Therein the B-cells are obtained from the blood of the immunized experimental animal and co-cultivated with murine EL-4 B5 15 feeder cells in the presence of a cytokine comprising feeder mix. With this methodology up to 50 ng/ml antibody can be obtained after 10-12 days of co-cultivation.

Weitkamp, J-H., et al., (J. Immunol. Meth. 275 (2003) 223-237) report the 20 generation of recombinant human monoclonal antibodies to rotavirus from single antigen-specific B-cells selected with fluorescent virus-like particles. A method of producing a plurality of isolated antibodies to a plurality of cognate antigens is reported in US 2006/0051348. In WO 2008/144763 and WO 2008/045140 antibodies to IL-6 and uses thereof and a culture method for obtaining a clonal 25 population of antigen-specific B cells are reported, respectively. A culture method for obtaining a clonal population of antigen-specific B-cells is reported in US 2007/0269868. Masri et al. (in Mol. Immunol. 44 (2007) 2101-2106) report the cloning and expression in E.coli of a functional Fab fragment obtained from single human lymphocyte against anthrax toxin. A method for preparing immunoglobulin libraries is reported in WO 2007/ 031550.

Summary of the Invention

Herein is reported a method for the isolation of a B-cell from a population of B-cells that has special properties. First already within four weeks after the first immunization of an experimental animal the induced antibody producing cells can be isolated and the binding specificity of the antibodies can be determined. Second it is possible to enhance the number and/or the quality (e.g. the antibody production/secretion capacity) of antibody producing cells by any one of the following steps: i) a pre-incubation step, and/or ii) a centrifugation step, and/or iii) a panning step. Third, the feeder mix used for the co-cultivation of B-cells and feeder cells can be improved by the addition of IL-21, or IL-6, or SAC, or BAFF.

Thus, herein is reported as an aspect a method for selecting a B-cell comprising the following steps:

- a) optionally labeling the B-cells of a population of B-cells,
- b) individually co-cultivating each B-cell of a population of B-cells, which have been deposited as single cell, with feeder cells,
- c) selecting a B-cell clone proliferating and secreting antibody in step b).

Herein is reported further as an aspect a method for obtaining a B-cell clone comprising the following steps:

- a) obtaining B-cells from an experimental animal,
- b) labeling the B-cells,
- c) depositing the labeled B-cells as single cells,
- d) individually co-cultivating the single cell deposited B-cells with feeder cells,
- e) selecting a B-cell clone proliferating and secreting antibody in step d) and thereby obtaining a B-cell clone.

Herein is reported as another aspect a method for producing an antibody specifically binding to a target antigen comprising the following steps

- a) optionally labeling the cells of a population of B-cells with at least one fluorescence dye,
- b) cultivating each B-cell of a population of B-cells, which has been deposited as single cell in individual containers, in the presence of

feeder cells and a feeder mix, to obtain individual B-cell clones and cultivation supernatants,

- c) selecting a B-cell clone producing an antibody specifically binding to a target antigen,
- 5 d) cultivating a cell, which contains a nucleic acid that encodes the antibody specifically binding to the target antigen, which is produced by the B-cell clone selected in step c), or a humanized variant thereof, and recovering the antibody from the cell or the cultivation supernatant and thereby producing the antibody.

10 In one embodiment the method comprises one or more of the following steps:

after step c): c1) determining the nucleic acid sequence encoding the variable light chain domain and the variable heavy chain domain of the antibody by a reverse transcriptase PCR,

15 after step c1): c2) transfecting a cell with a nucleic acid comprising the nucleic acid sequence encoding the antibody variable light chain domain and the variable heavy chain domain.

Herein is also reported as an aspect a method for producing an antibody comprising the following steps

- a) providing a population of (mature) B-cells (obtained from the blood of an experimental animal),
- 20 b) labeling the cells of the population of B-cells with at least one fluorescence dye (in one embodiment with one to three, or two to three fluorescence dyes),
- c) depositing single cells of the labeled population of B-cells in individual containers (in one embodiment is the container a well of a multi well plate),
- 25 d) cultivating the deposited individual B-cells in the presence of feeder cells and a feeder mix (in one embodiment the feeder cells are EL-4 B5 cells, in one embodiment the feeder mix is natural TSN, in one embodiment the feeder mix is a synthetic feeder mix),
- e) determining the binding specificity of the antibodies secreted in the cultivation medium of the individual B-cells,
- 30 f) determining the amino acid sequence of the variable light and heavy chain domain of specifically binding antibodies by a reverse

transcriptase PCR and nucleotide sequencing, and thereby obtaining a monoclonal antibody variable light and heavy chain domain encoding nucleic acid,

- 5 g) introducing the monoclonal antibody variable light and heavy chain variable domain encoding nucleic acid in an expression cassette for the expression of an antibody,
- h) introducing the nucleic acid in a cell,
- i) cultivating the cell and recovering the antibody from the cell or the cell culture supernatant and thereby producing an antibody.

10 In one embodiment of all aspects as reported herein the method comprises the step of incubating the population of B-cells in the co-cultivation medium prior to single cell depositing. In one embodiment the incubating is at about 37 °C. In one embodiment the incubating is for 0.5 to two hours. In a specific embodiment the incubating is for about one hour. In one embodiment the incubating is at about 15 37 °C for about one hour.

20 In one embodiment of all aspects as reported herein the method comprises the step of centrifuging the single cell deposited B-cells prior to the co-cultivation. In one embodiment the centrifuging is for about 1 min. to about 30 min. In a specific embodiment the centrifuging is for about 5 min. In one embodiment the centrifuging is at about 100 x g to about 1,000 x g. In a specific embodiment the centrifuging is at about 300 x g. In one embodiment the centrifuging is for about 5 min. at about 300 x g.

25 In one embodiment of all aspects as reported herein the method comprises immediately prior to the labeling step the following step: panning the B-cells with immobilized antigen.

In one embodiment of all aspects as reported herein the population of B-cells is obtained from the blood of an animal by a density gradient centrifugation.

30 In one embodiment of all aspects as reported herein the population of B-cells is obtained from the blood of an experimental animal after 4 days after the immunization. In another embodiment the population of B-cells is obtained from the blood of an experimental animal of from 4 days to at least 9 days after immunization. In a further embodiment the population of B-cells is obtained from the blood of an experimental animal of from 4 days to 9 days after immunization.

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In one embodiment of all aspects as reported herein the population of B-cells is isolated by density gradient centrifugation.

In one embodiment of all aspects as reported herein the B-cells are mature B-cells.

In one embodiment of all aspects as reported herein the labeling is with one to three fluorescence dyes. In a specific embodiment the labeling is with two or three fluorescence dyes.

In one embodiment of all aspects as reported herein the labeling of the B-cells results in labeling of 0.1 % to 2.5 % of the cells of the total B-cell population.

In one embodiment of all aspects as reported herein the B-cells are mouse B-cells, or hamster B-cells, or rabbit B-cells.

In one embodiment of all aspects as reported herein the single cell depositing is in the wells of a multi well plate.

In one embodiment of all aspects as reported herein the feeder cells are murine EL-4 B5 cells.

In one embodiment of all aspects as reported herein the antibody is a monoclonal antibody.

In one embodiment of all aspects as reported herein the labeling is of IgG⁺CD19⁺-B-cells, IgG⁺CD38⁺-B-cells, IgG⁺CD268⁺-B-cells, IgG⁻CD138⁺-B-cells, CD27⁺CD138⁺-B-cells, or CD3⁻CD27⁺-B-cells.

In one embodiment of all aspects as reported herein the B-cells are of mouse origin and the labeling is of IgG⁺CD19⁺-B-cells, and/or IgG⁻CD138⁺-B-cells.

In one embodiment of all aspects as reported herein the B-cells are of hamster origin and the labeling is of IgG⁺IgM⁻-B-cells.

In one embodiment of all aspects as reported herein the B-cells are of rabbit origin and the labeling is of IgG⁺-B-cells and/or CD138⁺-B-cells, or CD138⁺IgG⁺-B-cells and/or IgG⁺IgM⁻-B-cells.

In one embodiment of all aspects as reported herein the co-cultivating is in an RPMI 1640 medium supplemented with 10 % (v/v) FCS, 1 % (w/v) of a 200 mM glutamine solution that comprises penicillin and streptomycin, 2 % (v/v) of a

100 mM sodium pyruvate solution, and 1 % (v/v) of a 1 M 2-(4-(2-hydroxyethyl)-1-piperazine)-ethane sulfonic acid (HEPES) buffer. In another embodiment the co-cultivating medium further comprises 0.05 mM beta-mercaptoethanol.

5 In one embodiment of all aspects as reported herein the co-cultivating of the B-cells is with feeder cells and a feeder mix. In one embodiment the feeder mix is a natural thymocyte cultivation supernatant (TSN) or a synthetic feeder mix.

10 In one specific embodiment the feeder mix is a synthetic feeder mix. In one embodiment the synthetic feeder mix comprises interleukin-1 beta and tumor necrosis factor alpha. In one embodiment the synthetic feeder mix comprises interleukin-2 (IL-2) and/or interleukin-10 (IL-10). In one embodiment the synthetic feeder mix further comprises *Staphylococcus aureus* strain Cowans cells (SAC). In one embodiment the synthetic feeder mix comprises interleukin-21 (IL-21). In one embodiment the synthetic feeder mix comprises B-cell activation factor of the tumor necrosis factor family (BAFF). In one embodiment the synthetic feeder mix comprises interleukin-6 (IL-6). In one embodiment the synthetic feeder mix comprises interleukin-4 (IL-4).

15

In one embodiment the co-cultivating is in the presence of a thymocyte cultivation supernatant as feeder mix. In a specific embodiment the thymocyte cultivation supernatant is obtained from thymocytes of the thymus gland of a young animal.

20 In one embodiment the method for obtaining a B-cell clone further comprises the step of

25 f) determining the amino acid sequence of the variable light and heavy chain domain of the antibody produced by the selected B-cell clone of step e) by a reverse transcriptase PCR and nucleotide sequencing, and thereby obtaining a monoclonal antibody amino acid variable domain sequence.

In one embodiment the experimental animal is selected from mouse, hamster, and rabbit.

30 **Detailed Description of the Invention**

The method reported herein allows for a rapid characterization of the binding specificity of monoclonal antibodies obtained from individual B-cell clones, i.e.

within four weeks after the first immunization of the experimental animal the induced antibody producing cells can be isolated and the binding specificity of the antibodies produced therefrom can be determined, whereby at least 4 different experiments can be performed due to the antibody amount/concentration in the B-cell co-cultivation supernatant.

Immunization:

Often non-human animals, such as mice, rabbits, hamster and rats, are used as animal model for evaluating antibody based therapies. Therefore, it is often required to provide cross-reactive antibodies binding to the non-human animal antigen as well as to the human antigen. The method as reported herein can be used to provide cross-reactive antibodies. In the method as reported herein B-cells obtained from e.g. mouse, hamster and rabbit can be used. In one embodiment the mouse is an NMRI-mouse or a balb/c-mouse. In another embodiment the hamster is selected from Armenian hamster (*Cricetulus migratorius*), Chinese hamster (*Cricetulus griseus*), and Syrian hamster (*Mesocricetus auratus*). In a specific embodiment the hamster is the Armenia hamster. In one embodiment the rabbit is selected from New Zealand White (NZW) rabbits, Zimmermann-rabbits (ZIKA), Alicia-mutant strain rabbits, basilea mutant strain rabbits, transgenic rabbits with a human immunoglobulin locus, rbIgM knock-out rabbits, and cross-breeding thereof.

In one embodiment the experimental animals, e.g. mice, hamster and rabbits, chosen for immunization are not older than 12 weeks.

Source and isolation of B-cells:

The blood of an experimental animal provides a high diversity of antibody producing B-cells. The therefrom obtained B-cells secrete antibodies that have almost no identical or overlapping amino acid sequences within the CDRs, thus, show a high diversity.

In one embodiment the B-cells of an experimental animal, e.g. from the blood, are obtained of from 4 days after immunization until at least 9 days after immunization or the most recent boost. This time span allows for a high flexibility in the method as reported herein. In this time span it is likely that the B-cells providing for the most affine antibodies migrate from spleen to blood (see e.g. Paus, D., et al., JEM

203 (2006) 1081–1091; Smith, K.G.S., et al., The EMBO J. 16 (1997) 2996–3006; Wrammert, J., et al., Nature 453 (2008) 667-672).

B-cells from the blood of an experimental animal may be obtained with any method known to a person skilled in the art. For example, density gradient centrifugation (DGC) or red blood cell lysis (lysis) can be used. Density gradient centrifugation compared to hypotonic lysis provides for a higher overall yield, i.e. number of B-cell clones. Additionally from the cells obtained by density gradient centrifugation a larger number of cells divides and grows in the co-cultivation step. Also the concentration of secreted antibody is higher compared to cells obtained with a different method. Therefore, in one embodiment the providing of a population of B-cells is by density gradient centrifugation.

Table 1: Number of IgG producing wells/cell clones when the cells are obtained by density gradient centrifugation (DGC) or hypotonic lysis of erythrocytes.

	mouse, DGC	mouse, lysis	hamster, DGC	hamster, lysis
number of isolated cells [$\times 10^6$]	1.7 ± 0.2 (n= 2)	1.6 ± 0.1 (n= 2)	2.1 ± 0.2 (n= 2)	0.9 ± 0.1 (n= 2)
IgG ⁺ -wells [%]	22	12	7	6

15 Selection steps prior to co-cultivation:

B-cells producing antibodies that specifically bind an antigen can be enriched from peripheral blood mononuclear cells (PBMCs). Thus, in one embodiment of all methods as reported herein the B-cell population is enriched from peripheral blood mononuclear cells (PBMCs).

20 The term “specifically binding” and grammatical equivalents thereof denote that the antibody binds to its target with a dissociation constant (Kd) of 10^{-7} M or less, in one embodiment of from 10^{-8} M to 10^{-13} M, in a further embodiment of from 10^{-9} M to 10^{-13} M. The term is further used to indicate that the antibody does not specifically bind to other biomolecules present, i.e. it binds to other biomolecules with a dissociation constant (Kd) of 10^{-6} M or more, in one embodiment of from 10^{-6} M to 1 M.

In one embodiment of all methods as reported herein the PBMCs are depleted of macrophages. This is advantageous as outlined below, e.g. as in one embodiment for B-cells of rabbit origin, for the co-cultivation step.

Macrophages can be depleted from PBMCs by adhesion to the surface of the cell culture plate (see preincubation step).

In one embodiment of the methods as reported herein the cells are from a protein-immunized animal and are depleted of macrophages prior to the labeling.

It has been found that incubating the population of B-cells in co-cultivation medium prior to the single cell depositing increases the total number of antibody secreting cells obtained after the single cell depositing compared to a single cell depositing directly after the isolation and optional enrichment of the population of B-cells from the blood of an experimental animal (example rabbit, see Tables 2a and 2b). Specifically the incubating is at about 37 °C for about one hour in EL-4 B5 medium, e.g. using a cell culture incubator.

Table 2a: IgG positive wells/cell clones with and without one hour incubation in EL-4 B5 medium prior to single cell depositing of all cells (rb=rabbit).

rbIgG ELISA	fresh PBMCs (Ø 100-20 cells)	PBLs after incubation* (Ø 50-10 cells)
rbIgG ⁺ wells [n]	40	108
rbIgG ⁺ wells [% total wells]	28	75

* depleted of
macrophages and
monocytes

Table 2b: IgG positive wells/cell clones with and without one hour incubation in EL-4 B5 medium prior to single cell depositing of B-cells.

rbIgG ELISA	single B-cells from fresh PBMCs	single B-cells from blood, 1 h incubated	single B-cells from fresh spleen cells	single B-cells from spleen, 1 h incubated
rbIgG ⁺ wells [n]	2	55	6	52
rbIgG ⁺ wells [% of total wells]	2	33	7	31

In one embodiment of the methods as reported herein the cells are obtained from a protein-immunized animal and depleted of macrophages.

5 Cells not producing an antibody binding the antigen or, likewise, cells producing an antibody binding to the antigen can be reduced or enriched, respectively, by using a panning approach. Therein a binding partner is presented attached to a surface and cells binding thereto are selectively enriched in the cell population in case the bound cells are processed further, or reduced in the cell population in case 10 the cells remaining in solution are processed further.

Table 3: Enrichment of B-cells secreting an antigen-specific antibody by panning with the respective antigen.

protein antigen	without panning	with panning using the antigen
total wells [n]	4284	2113
antigen specific IgG ⁺ wells [n]	235	419
antigen specific IgG ⁺ wells [% total wells]	5	20
small molecule antigen	without panning	with panning using the small molecule
total wells [n]	336	336
small molecule IgG ⁺ wells [n]	2	115
small molecule IgG ⁺ wells [% total wells]	1	34

The method as reported herein comprises in one embodiment prior to the single cell depositing a selecting step in which B-cells producing specific and/or non-cross-reactive antibodies are selected based on cell surface markers and fluorescence activated cell sorting/gating. In one embodiment mature B-cells are sorted/enriched/selected. For selection of B-cells from different experimental animal species different cell surface markers can be used. It has been found that many of the available cell surface markers, either individually or in combination, do not provide for a suitable labeling.

With the labeling of non-target cell populations and non-specifically binding lymphocytes it is possible to selectively deplete these cells. In this depletion step only a non total depletion can be achieved. Albeit the depletion is not quantitative it provides for an advantage in the succeeding fluorescence labeling of the remaining cells as the number of interfering cells can be reduced or even minimized. By a single cell depositing of mature B-cells (memory B-cells, affinity matured plasmablasts and plasma cells) by fluorescence activated cell sorting using the labeling as outlined below a higher number of IgG⁺-wells/cell clones can be obtained in the co-cultivation step.

The term "labeling" denotes the presence or absence of a surface marker which can be determined by the addition of a specifically binding and labeled anti-surface marker antibody. Thus, the presence of a surface marker is determined e.g. in the case of a fluorescence label by the occurrence of a fluorescence whereas the absence of a surface marker is determined by the absence of a fluorescence after incubation with the respective specifically binding and labeled anti-surface marker antibody.

Different cell populations can be labeled by using different surface markers such as CD3⁺-cells (T-cells), CD19⁺-cells (B-cells), IgM⁺-cells (mature naive B-cells), IgG⁺-cells (mature B-cells), CD38⁺-cells (e.g. plasmablasts), and IgG⁺CD38⁺-cells (pre-plasma cells).

As reported herein an immuno-fluorescence labeling for selection of mature IgG⁺-B-cells, such as memory B-cells, plasmablasts, and plasma cells, has been developed. For a selection or enrichment of B-cells the cells are either single labeled, or double labeled, or triple labeled. Also required is a labeling that results in about 0.1 % to 2.5 % of labeled cells of the total cell population. In one embodiment B-cells are deposited as single cells selected by the labeling of surface

molecules present on 0.1 % to 2.5 % of the B-cells in the population, in another embodiment on 0.3 % to 1.5 % of the B-cells of the population, in a further embodiment on 0.5 % to 1 % of the B-cells of the population.

5 The IgG⁺-B-cells within the PBMC population 0.5 % - 1 % can be doubly labeled as IgG⁺CD19⁺-cells, IgG⁺CD38⁺-cells, and IgG⁺CD268⁺-cells. Thus, in one embodiment of all methods as reported herein IgG⁺CD19⁺-B-cells, IgG⁺CD38⁺-B-cells, or IgG⁺CD268⁺-B-cells are deposited as single cells.

10 Of IgG⁻-B-cells within the PBMC population 0.5 % - 1 % can be doubly labeled as IgG⁻CD138⁺-cells. Thus, in one embodiment of all methods as reported herein IgG⁻CD138⁺-B-cells are deposited as single cells.

15 The labeling of CD27⁺CD138⁺-cells or CD3⁻CD27⁺-cells results in about 1.5 % of the cells of the cell population to be labeled, respectively. Thus, in one embodiment of all methods as reported herein CD27⁺CD138⁺-B-cells or CD3⁻CD27⁺-B-cells are deposited as single cells.

20 15 Of IgG⁺-hamster-B-cells within the PBMC population 0.6 % ± 0.1 % can be doubly labeled as IgG⁺IgM⁻-hamster-B-cells. Thus, in one embodiment of all methods as reported herein IgG⁺IgM⁻-hamster-B-cells are deposited as single cells.

25 In one embodiment IgG⁻CD138⁺-B-cells are deposited as single cells from the B-cells obtained from an immunized animal. In one embodiment of all methods as reported herein IgG⁺CD19⁺-B-cells are deposited as single cells from the B-cells obtained from a non-immunized animal. In another embodiment of all methods as reported herein IgG⁺IgM⁻-B-cells are deposited as single cells from the B-cells obtained from a non-immunized or immunized animal. In one embodiment of all methods as reported herein IgG⁺CD19⁺-murine-B-cells are deposited as single cells.

30 25 This selection step results in an improved or even the highest yield of IgG⁺-wells in the succeeding co-cultivation step. In another embodiment of all methods as reported herein IgG⁻CD138⁺-murine-B-cells are deposited as single cells. Therewith cells producing the highest amount of B-cell clones in the first place and secondly the highest concentration of IgG are selected (see Table 5). In another embodiment of all methods as reported herein IgG⁺CD19⁺-murine-B-cells and IgG⁻CD138⁺-murine-B-cells are deposited as single cells. In one specific embodiment the method is with the proviso that if the cells are of rabbit origin the labeling is not of IgG⁺-B-cells and/or CD138⁺-B-cells.

IgG⁺-murine-B-cells can be labeled with the anti-mouse-IgG-antibody 227 (Ab 227), IgG⁺-hamster-B-cells can be labeled with the anti-hamster-IgG-antibody 213 (AB 213) and/or anti-hamster-IgG-antibody 225 (AB 225), and rabbit B-cells can be labeled with the anti-IgG-antibody 184 (see Table 4).

5 **Table 4:** Immunofluorescence labeling of B-cells – the table present the average labeled fraction of the population of murine B-cells (A-E), hamster B-cells (F-H) and rabbit B-cells (I-J).

	Single IgG labeling	IgG+CD19 labeling	IgG+IgM labeling
A	IgG ⁺ AB 185 PE 17 % ± 3 % n=4	-	IgG ⁺ IgM ⁺ AB 185 PE, AB 219 APC 12 % n=1
B	IgG ⁺ AB 215 APC 12 % ± 3 % n=5	IgG ⁺ CD19 ⁺ AB 215 APC, AB 218 PE 11 % n=1	IgG ⁺ IgM ⁺ AB 215 APC, AB 200 PE 14 % n=1
C	IgG ⁺ AB 217 FITC 17 % ± 4 % n=7	IgG ⁺ CD19 ⁺ AB 217 FITC, AB 218 PE 10 % n=1	IgG ⁺ IgM ⁺ AB 217 FITC, AB 200 PE 19 % n=1
D	IgG ⁺ AB 222 FITC 18 % ± 2 % n=3	IgG ⁺ CD19 ⁺ AB 222 FITC, AB 218 PE 15 % n=1	IgG ⁺ IgM ⁺ AB 222 FITC, AB 200 PE 14 % n=1
E	IgG ⁺ AB 227 FITC 0.8 % ± 0.3 % n=13	IgG ⁺ CD19 ⁺ AB 227 FITC, AB 218 PE 0.5 % n=1	IgG ⁺ IgM ⁺ AB 227 FITC, AB 200 PE 0.2 % n=1
F	IgG ⁺ AB 212 FITC 43 % ± 6 % n=7	no B-cell marker known	IgG ⁺ IgM ⁺ AB 212 FITC, AB 223 APC 43 % n=1
G	IgG ⁺ AB 213 APC 0.9 % ± 0.4 % n=27	no B-cell marker known	IgG ⁺ IgM ⁺ AB 213 APC, AB 224 FITC 0.07 % n=1
H	IgG ⁺ AB 225 PE 17 % ± 3 % n=5	no B-cell marker known	IgG ⁺ IgM ⁺ AB 225 PE, AB 224 FITC 0.7 % n=1
I	IgG ⁺ AB 120 PE > 10 %	-	-
J	IgG ⁺ AB 184 FITC 0.3 – 2 %	-	-

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	AB 120 - goat anti-rabbit IgG-antibody	Southern Biotech	4030-09
	AB 184 - goat anti-rabbit IgG Fc-antibody	AbDSerotech	STAR121F
	AB 185 - goat anti-mouse IgG-antibody	Caltag	M35004-3
	AB 200 - goat anti-mouse IgM-antibody	Invitrogen	M31504
5	AB 212 - goat anti-hamster IgG-antibody	AbDSerotech	STAR79F
	AB 213 - mouse anti-hamster IgG-antibody	Becton Dickinson	554010
	AB 215 - goat anti-mouse IgG-antibody	Sigma	B 0529
	AB 217 - goat anti-mouse IgG-antibody	AbDSerotech	STAR120F
	AB 218 - rat anti-mouse CD19-antibody	Abcam	ab22480
10	AB 219 - goat anti-mouse IgM-antibody	Rockland	710-1607
	AB 222 - goat anti-mouse IgG-antibody	Abcam	ab7064
	AB 223 - mouse anti-hamster IgM-antibody	Becton Dickinson	554035
	AB 224 - mouse anti-hamster IgM-antibody	Becton Dickinson	554033
	AB 225 - mouse anti-hamster IgG-antibody	Becton Dickinson	554056
15	AB 227 - goat anti-mouse IgG-antibody	Sigma	F 8264

PE: Phycoerythrin

APC: Allophycocyanin

FITC: Fluorescein isothiocyanate

It has to be pointed out that not all commercially available antibodies can be used for the labeling due to their low or non existing specificity.

Murine-B-cells can be labeled with the anti-IgG-antibody 227, hamster-B-cells can be labeled with the anti-IgG-antibody 213.

IgG⁺CD19⁺-murine-B-cells can be labeled with antibody 227 and antibody 218,

IgG⁺IgM⁻-murine-B-cells can be labeled with antibody 227 and antibody 219,

IgG⁺IgM⁻-hamster-B-cells can be labeled with antibody 213 and antibody 224,

IgG⁺-rabbit-B-cells can be labeled with antibody 184,

IgG⁺IgM⁻-rabbit-B-cells can be labeled with antibody 184 and antibody 254 and SA 263,

IgG⁺CD138⁺-rabbit-B-cells can be labeled with antibody 259 and antibody 256.

Murine B-cells can be labeled with the anti-CD27 antibody 235 or 236 (AB 235, AB 236), the anti-CD38 antibody 192 (AB 192), the anti-CD138 antibody 233 (AB 233) and the anti-CD268 antibody 246 (AB 246).

Table 5: Immuno fluorescence labeling for the determination of mature mouse- (A-J), hamster- (K) and rabbit (L-N)-B-cells.

labeling	Immuno fluorescence labeling for sorting of B-cells	Percentage of all viable cells %
A	IgG ⁺ CD19 ⁺ - AB 227 FITC, AB 218 PE	0.5 ± 0.2 n=14
B	IgG ⁺ CD38 ⁺ - AB 227 FITC, AB 192 PE	0.8 ± 0.5 n= 9
C	IgG ⁺ CD138 ⁺ - AB 227 FITC, AB 233 PE	0.06 ± 0.07 n= 6
D	IgG ⁻ CD138 ⁺ - AB 227 FITC, AB 233 PE	0.6 ± 0.5 n=6
E	IgG ⁺ CD27 ⁺ - AB 227 FITC, AB 235 PE	0.1 ± 0.1 n= 8
F	CD27 ⁺ CD138 ⁺ - AB 236 A647, AB 233 PE	1.5 ± 0.5 n= 2
G	CD27 ⁺ IgG ⁺ CD3 ⁻ - AB 235 PE, AB 227 FITC, AB 241 A647	0.10 ± 0.04 n= 3
H	CD3 ⁻ CD27 ⁺ - AB 189 FITC, AB 235 PE	1.33 n= 1
I	IgG ⁺ CD268 ⁺ - AB 227 FITC, AB 246 A647	0.8 n= 1
J	CD38 ⁺ CD3 ⁻ - AB 192 PE, AB 189 FITC	12 ± 7 n= 2
K	IgG ⁺ IgM ⁻ - AB 213 A647, AB 224 FITC	0.6 ± 0.1 n= 15
L	IgG ⁺ - AB 184 FITC	0.6 ± 0.2, n= 5
M	IgG ⁺ IgM ⁻ - AB 184 FITC, AB 254 Biotin, SA 263 PE	0.4 ± 0.2, n=2
N	IgG ⁺ CD138 ⁺ - AB 259, AB 256 PE	0.3 ± 0.1, n= 5

	AB 184 - goat anti-rabbit IgG-antibody	AbD Serotec	STAR121F
	AB 189 - hamster anti-mouse CD3-antibody	Becton Dickinson	553062
5	AB 192 - rat anti-mouse CD38-antibody	Becton Dickinson	553764
	AB 213 - mouse anti-hamster IgG-antibody	Becton Dickinson	554010
	AB 218 - rat anti-mouse CD19-antibody	Abcam	ab22480
	AB 224 - mouse anti-hamster IgM-antibody	Becton Dickinson	554033
	AB 227 - goat anti-mouse IgG-antibody	Sigma	F 8264
10	AB 233 - rat anti-mouse CD138-antibody	Becton Dickinson	553714
	AB 235 - hamster anti-mouse CD27-antibody	Becton Dickinson	558754
	AB 236 - hamster anti-mouse CD27-antibody	Becton Dickinson	558753
	AB 241 - hamster anti-mouse CD3-antibody	Becton Dickinson	553060
	AB 246 - rat anti-mouse BAFF-R-antibody	eBioscience	51-5943
15	AB 254 - mouse anti-rabbit IgM-antibody	Becton Dickinson	custom made
	AB 256 – goat anti-rat IgG-antibody	Southern Biotech	3030-09
	AB 259 – rat anti-rabbit CD138-antibody	Roche Glycart AG	
	SA 263 – Streptavidin	Invitrogen	S866

A647: Alexa Fluor® 647

FITC: Fluorescein isothiocyanate

In one embodiment the methods comprise the step of depleting the B-cell population of macrophages and enriching of B-cells of the B-cell population secreting antibody specifically binding a target antigen.

5 Single cell depositing:

The method as reported herein comprises the step of depositing the B-cells of a B-cell population as single cells. In one embodiment of all methods as reported herein the depositing as single cells is by fluorescence activated cell sorting (FACS). The labeling required for the FACS single cell depositing can be carried 10 out as reported in the previous section.

In one embodiment of all methods as reported herein specifically labeled B-cells are deposited as single cells. In a further embodiment of all methods as reported herein the labeling is a labeling of cell surface markers with fluorescence labeled 15 antibodies. In another embodiment the methods as reported herein provide for monoclonal antibodies. In one embodiment of all methods as reported herein mature B-cells are deposited as single cells.

It has also been found that an additional centrifugation step after the single cell depositing and prior to the co-cultivation provides for an increased number of 20 antibody secreting cells and increases the amount of the secreted IgG (example experimental animal with human immunoglobulin locus, see Table 6).

Table 6: IgG positive wells/cell clones with and without centrifugation step after single cell depositing.

huCk ELISA	with centrifugation step	without centrifugation step
huCk ⁺ wells [n]	9	1
huCk ⁺ wells [% total wells]	13	1
huCk conc. of all huCk ⁺ wells [average ng/ml]	76.4	9.7

In one embodiment of all methods as reported herein the method comprises the step 25 of centrifuging the single deposited cells prior to the co-cultivation. In one specific embodiment the centrifuging is for 5 min. at 300 x g.

Co-cultivation:

The co-cultivation step with feeder cells can be preceded and also succeeded by a number of additional steps.

In one embodiment of all methods as reported herein the single deposited B-cells are co-cultivated with feeder cells in the presence of a feeder mix. In a specific embodiment the B-cells are co-cultivated with murine EL-4 B5 feeder cells. By suitable immuno fluorescence labeling as outlined above an increase in the yield in the co-cultivation step (number of IgG⁺-wells/cell clones as well as IgG-concentration) and also an enrichment or isolation of mature IgG⁺-B-cell from

10 PBMCs can be achieved.

With the single cell depositing of IgG⁺CD19⁺- and/or IgG⁺CD38⁺-B-cells from freshly isolated PBMCs the highest number of IgG⁺-wells/cell clones can be obtained. With the single cell depositing of IgG⁺CD19⁺-, IgG⁺CD38⁺- and/or IgG⁻CD138⁺-B-cells after the depletion of macrophages or KLH-specific cells (keyhole 15 limpet haemocyanine) good results can be obtained. With the single cell depositing of IgG⁺CD19⁺-, IgG⁺CD38⁺- and/or IgG⁻CD138⁺-B-cells after the depletion of antigen-specific B-cells improved results can be obtained. Thus, in one embodiment of all methods as reported herein IgG⁺CD19⁺-, IgG⁺CD38⁺- and/or IgG⁻CD138⁺-B-cells are deposited as single cells.

20 It has been found that a single cell depositing based on a labeling as outlined above results in the highest fraction of IgG⁺-wells/cell clones and in the wells/cell clones with the highest IgG-concentration in the supernatant. Thus, in one embodiment of all methods as reported herein IgG⁺CD19⁺- and/or IgG⁻CD138⁺-murine-B-cells are deposited as single cells. In one embodiment of all methods as reported herein IgG⁺IgM⁻-hamster-B-cells are deposited as single cells. In one embodiment of all methods as reported herein IgG⁺-, and/or IgG⁺CD138⁺-, and/or CD138⁺- and/or IgG⁺IgM⁻-rabbit-B-cells are deposited as single cells.

25

Table 7: Yield in the co-cultivation depending on the immuno fluorescence labeling.

labeling		n _{total wells} isol/depl/enr	IgG ⁺ -wells of n _{total} wells (%)			average IgG- concentration (ng/ml)		
			isol.	depl.	enr.	isol.	depl.	enr.
mouse	IgG ⁺ CD19 ⁺	356/ 356/324	45	50	37	68	46	42
	IgG ⁺	-/144/144	-	32	7	-	34	31
	IgG ⁺ CD38 ⁺	72/190/190	36	41	43	37	26	27
	IgG ⁺ CD138 ⁺	72/72/72	3	13	12	22	59	43
	IgG ⁻ CD138 ⁺	36/108/48	19	52	37	55	31	51
	IgG ⁺ CD27 ⁺	64/64/64	4	28	20	102	54	32
	CD27 ⁺ CD138 ⁺	-/32/-	-	6	-	-	135	-
	CD27 ⁺ IgG ⁺ CD3 ⁻	72/72/72	14	0	14	4	0	0
	CD3 ⁻ CD27 ⁺	-/32/-	-	13	-	-	29	-
hamster	IgG ⁺ CD268 ⁺	-/72/-	-	35	-	-	93	-
	IgG ⁺ IgM ⁻	-/216/216	-	17	22	-	78	93
	IgG ⁺	-/216/216	-	10	35	1	71	64
rabbit	IgG ⁺	-/1512/1307	-	33	28	-	59	60
	IgG ⁺ IgM ⁻	-/76/-	-	29	-	-	5	-
	CD138 ⁺	-/2016/-	-	14	-	-	16	-
	IgG ⁺ CD138 ⁺	-/168/-	-	37	-	-	64	-

For murine B-cells with the single cell depositing of IgG⁺CD19⁺-cells after each enrichment (enr.) and/or depletion (depl.) step the highest number of IgG⁺-wells/cell clones after co-cultivation can be obtained. Alternatively, with the single cell depositing of IgG⁻CD138⁺-cells wells/cell clones with the best IgG-concentration in the supernatant can be obtained. The single cell depositing of IgG⁻CD138⁺-cells can be used for B-cells from immunized animals. The single cell depositing of IgG⁺CD19⁺-cells can be used for B-cells from non-immunized animals. The single cell depositing of IgG⁺IgM⁻-cells can be used for hamster-B-cells of immunized and non-immunized animals. The single cell depositing of IgG⁺- and/or IgG⁺CD138⁺- and/or CD138⁺- and/or IgG⁺IgM⁻-B-cells can be used for rabbit-B-cells.

The immuno fluorescence labeling used for B-cells obtained from the blood of an experimental animal can also be used for the labeling of B-cells obtained from the spleen and other immunological organs of an experimental animal, such as mouse,

5 For murine B-cells with the single cell depositing of IgG⁺CD19⁺-cells after each enrichment (enr.) and/or depletion (depl.) step the highest number of IgG⁺-wells/cell clones after co-cultivation can be obtained. Alternatively, with the single cell depositing of IgG⁻CD138⁺-cells wells/cell clones with the best IgG-concentration in the supernatant can be obtained. The single cell depositing of IgG⁻CD138⁺-cells can be used for B-cells from immunized animals. The single cell depositing of IgG⁺CD19⁺-cells can be used for B-cells from non-immunized animals. The single cell depositing of IgG⁺IgM⁻-cells can be used for hamster-B-cells of immunized and non-immunized animals. The single cell depositing of IgG⁺- and/or IgG⁺CD138⁺- and/or CD138⁺- and/or IgG⁺IgM⁻-B-cells can be used for rabbit-B-cells.

10 The immuno fluorescence labeling used for B-cells obtained from the blood of an experimental animal can also be used for the labeling of B-cells obtained from the spleen and other immunological organs of an experimental animal, such as mouse,

15

hamster and rabbit. For mouse B-cells the fraction of IgG⁺-B-cells from spleen was about 0.8 % compared to 0.4 % for IgG⁺CD19⁺-cells. For hamster B-cells the respective numbers are 1.9 % and 0.5 % IgG⁺IgM⁻-cells. For rabbit-blood derived B-cells 0.2 % of IgG⁺-cells were found after depletion of macrophages. Peyer'sche plaques from rabbit showed 0.4 % of IgG⁺-cells and spleen showed 0.3 % of IgG⁺-cells after depletion of macrophages.

With the methods as reported herein after about seven (7) days, i.e. after 5, 6, 7, or 8 days, especially after 7 or 8 days, of co-cultivation antibody concentrations of from about 30 ng/ml up to 15 µg/ml or more can be obtained (average value about 500 ng/ml). With the thereby provided amount of antibody a high number of different analyses can be performed in order to characterize the antibody, e.g. regarding binding specificity, in more detail. With the improved characterization of the antibody at this early stage in the screening / selection process it is possible to reduce the number of required nucleic acid isolations and sequencing reactions that have to be performed. Additionally the B-cell clone provides an amount of mRNA encoding monoclonal light and heavy chain variable region allowing the use of degenerated PCR primer and obviates the requirement of highly specific primer. Also the required number of PCR cycles is reduced. Thus, in one embodiment the reverse transcriptase PCR is with degenerated PCR primer for the light and heavy chain variable domain.

In one embodiment of all methods as reported herein the feeder mix is a thymocyte cultivation supernatant. In a specific embodiment the thymocyte cultivation supernatant is obtained from the thymocytes of the thymus gland of the respective young animal. It is especially suited to use the thymus gland of young animals compared to the isolation of thymocytes from the blood adult animals. The term "young animal" denotes an animal before sexual maturity occurs. A young hamster, for example, is of an age of less than 6 weeks, especially less than 4 weeks. A young mouse, for example, is of an age of less than 8 weeks, especially less than 5 weeks.

Due to the origin of the feeder mix, which is derived from the supernatant of cultivated thymocytes (thymocyte cultivation supernatant – TSN), considerable batch to batch variations occur. In order to overcome this variability a synthetic feeder mix consisting of synthetic components has been developed. A feeder mix consisting of IL-1 β (interleukin-1 beta), TNF α (tumor necrosis factor alpha), IL-2

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(interleukin-2) and IL-10 (interleukin-10) is known from Tucci, A., et al., J. Immunol. 148 (1992) 2778-2784.

It is reported herein a synthetic feeder mix for the co-cultivation of single deposited B-cells and feeder cells. Also reported herein are B-cell-species-specific additives for the synthetic feeder mix for increasing the amount of secreted antibody by the respective B-cell clone. Concomitantly highly producing cells contain more mRNA which in turn facilitates the reverse transcription and sequencing of the encoding nucleic acid, e.g. with a redundant, non-specific primer set.

By the addition of SAC (Staphylococcus aureus strain Cowans cells, a single SAC lot was used) the number of antibody secreting B-cells and the average IgG-concentration in the supernatant after co-cultivation can be increased. It has been found that for the addition of SAC in the co-cultivation a concentration range can be defined as reduced as well as increased concentrations of SAC reduce the amount of secreted antibody.

Table 8a: Results of a huCk ELISA (huCk = human C kappa) or rbIgG ELISA of cell culture supernatants of B-cells obtained from an experimental animal with human IgG locus or a wildtype rabbit (NZW) co-cultivated with EL-4 B5 feeder cells and TSN as feeder mix with or without added SAC.

	TSN	TSN + SAC		
huCk ⁺ wells [n]	7	45		
huCk ⁺ wells [% total wells]	5	31		
huCk conc. of all huCk ⁺ wells [Ø ng/ml]	89.1	41.0		
	SAC 1:5000	SAC 1:10000	SAC 1:20000	SAC 1:40000
rbIgG ⁺ wells [n]	13	15	27	30
rbIgG ⁺ wells [% total wells]	15	18	32	36
rbIgG conc. of all rbIgG ⁺ wells [Ø ng/ml]	149.0	159.1	233.7	197.2

	w/o	SAC 1:20000	SAC 1:50000	SAC 1:100000	SAC 1:150000
rbIgG ⁺ wells [n]	12	75	93	92	72
rbIgG ⁺ wells [% total wells]	5	30	37	37	29
rbIgG conc. of all rbIgG ⁺ wells [Ø ng/ml]	199	665	742	774	668

It can be seen that a SAC ratio of from 1:20000 to 1:150000 provides for an increased number of IgG⁺-wells/cell clones, whereby the ratio of from 1:50000 to 1:100000 shows the highest numbers. In one embodiment the amount of SAC added to the cultivation medium is determined by providing a dilution series and determining the dilution at which the added SAC provides for the highest number of IgG positive wells/cell clones.

5

It has been observed that by the addition of SAC to the feeder-mix the co-cultivation of B-cells was surprisingly changed in such a way that only single deposited B-cells have a benefit in growth, whereas B-cell growth was inhibited when using a PBL (e.g. B cells and endogenous T cells) mixture for co-cultivation.

10

Table 8b: Results of a huCk ELISA or rbIgG ELISA of cell culture supernatants of PBLs and single deposited B-cells co-cultivated with EL-4 B5 feeder cells and TSN as feeder mix with added SAC.

rbIgG ELISA	PBLs* (30 cells)	single deposited rbIgG ⁺ -B-cell
rbIgG ⁺ wells [n]	8	104
rbIgG ⁺ wells [% total wells]	6	58
rbIgG conc. of all huCk ⁺ wells [average ng/ml]	55.0	129.2

* depleted of macrophages

Further data obtained with different feeder mixes is presented in the following Tables 9 and 10.

15

In one embodiment of all methods as reported herein the synthetic feeder mix for the co-cultivation of B-cells comprises IL-1 β , TNF α , IL-2, IL-10 and IL-21 (interleukin-21). In one embodiment of all methods as reported herein the synthetic feeder mix for the co-cultivation of B-cells comprises IL-1 β , TNF α , IL-2, IL-10

and SAC. In one specific embodiment IL-1 β , TNF α , IL-2, IL-10 and IL-21 are recombinant murine IL-1 β , murine TNF α , murine IL-2, murine IL-10, and murine IL-21.

In one embodiment of all methods as reported herein the synthetic feeder mix for the co-cultivation of murine B-cells comprises IL-1 β , IL-2, IL-10, TNF- α and BAFF. In one specific embodiment BAFF is added at a concentration of 5 ng/ml.

In one embodiment of all methods as reported herein the synthetic feeder mix for the co-cultivation of hamster B-cells comprises IL-1 β , IL-2, IL-10, TNF- α , IL-6 and SAC. In one specific embodiment IL-6 is added at a concentration of 10 ng/ml.

In one specific embodiment SAC is added at a 1:75,000 ratio.

Table 9: Results of an rbIgG ELISA of cell culture supernatants of rabbit B-cells co-cultivated with EL-4 B5 feeder cells and different synthetic feeder mixes comprising recombinant murine substances in different combinations.

	rabbit TSN, SAC	IL-6, IL- 1β, TNFα, IL-2, IL- 10	IL-6, TNFα, IL-2, IL-10	IL-6, IL-1β, IL-2, IL-10	IL-6, IL-1β, TNFα, IL-2	IL-6, IL-1β, TNFα, IL-10	IL-1β, TNFα, IL-2, IL-10
rbIgG ⁺ wells [n]	37	24	12	16	18	23	24
rbIgG ⁺ wells [% total wells]	51	33	17	22	25	32	33
rbIgG conc. of all rbIgG ⁺ wells [\emptyset ng/ml]	196.0	289.9	32.4	75.7	166.4	134.4	203.6

Table 10: IgG⁺-wells of cell culture supernatants of rabbit B-cells co-cultivated with EL-4 B5 feeder cells and TSN or a feeder mix comprising recombinant murine substances and SAC (rb = rabbit, m = mouse).

rbIgG ⁺ wells [n]	TSN + SAC	IL-1 β , TNF α , IL-2, IL-10 + SAC
pure	64	55
+ mIL21	22	25
+ mIL10	78	61
+ mIL21+ mIL10	57	93
rbIgG ⁺ wells [% total wells]		
pure	25	22
+ mIL21	9	10
+ mIL10	31	24
+ mIL21+ mIL10	23	37
rbIgG conc. of all rbIgG+ wells [\emptyset ng/ml]		
pure	312.3	662.3
+ mIL21	263.7	541.1
+ mIL10	553.0	522.3
+ mIL21+ mIL10	422.6	307.5

5 A co-cultivation of feeder cells and murine B-cells without IL-2, without IL-10, as well as without IL-2 and IL-10 results in an increase in the yield of IgG⁺-wells albeit the IgG-concentration is reduced. Without TNF α the IgG-concentration is also reduced. Without IL-1 β no IgG can be found in the supernatant.

10 A co-cultivation of hamster B-cells without IL-2 or without IL-10, respectively, results in IgG⁺-wells with detectable IgG-concentration. In contrast thereto in a co-cultivation without IL-2 and IL-10 almost no B-cell growth can be detected. In the absence of TNF- α or IL-1 β no IgG-secretion can be determined.

15 In the presence of EL-4 B5 feeder cells at least IL-1 β and TNF α are required for the co-cultivation of mouse, hamster and rabbit B-cells. IL-2 and IL-10 can be omitted for the co-cultivation of murine cells. Hamster B-cells can be cultivated in the absence of either IL-2 or IL-10. Rabbit B-cells can be cultivated in the absence of either IL-2 or IL-10 or IL-6.

For murine and hamster B-cells the addition of IL-4 to the feeder mix increases the number of IgG⁺-wells/cell clones as well as the IgG-concentration in the supernatant. Thus, in one embodiment of all methods as reported herein the feeder mix for the co-cultivation of murine- or hamster-B-cells comprises IL-4.

5 The addition of IL-6 to the feeder mix for the co-cultivation of murine-B-cells or hamster-B-cells results in an increased number of IgG⁺-wells/cell clones or increased IgG-concentration, respectively. Thus, in one embodiment of all methods as reported herein the feeder mix for the co-cultivation of murine-B-cells or hamster-B-cells comprises IL-6. In one specific embodiment the IL-6 is added at a 10 concentration of 50 ng/ml. In one specific embodiment IL-6 is added at a concentration of 10 ng/ml, if high IgG-concentration is required. In one specific embodiment the addition of IL-6 is after three days of co-cultivation of the selected B-cells and EL-4 B5 cells.

15 One aspect as reported herein is a synthetic feeder mix for the co-cultivation of B-cells and feeder cells that comprises IL-1 β , TNF α , IL-10, and one or more selected from IL-21, SAC, BAFF, IL-2, IL-4, and IL-6.

One aspect as reported herein is a synthetic feeder mix for the co-cultivation of B-cells and feeder cells that comprises IL-1 β , TNF α , IL-2, IL-10 and SAC.

20 One aspect as reported herein is a synthetic feeder mix for the co-cultivation of murine B-cells and feeder cells that is consisting of IL-1 β , TNF α , and optionally comprises IL-21, and/or SAC, and/or BAFF, and/or IL-6.

One aspect as reported herein is a synthetic feeder mix for the co-cultivation of murine B-cells and feeder cells that comprises IL-1 β , IL-2, IL-10, TNF- α and BAFF.

25 One aspect as reported herein is a synthetic feeder mix for the co-cultivation of murine or hamster B-cells and feeder cells that comprises IL-1 β , TNF α , IL-2, IL-10 and IL-6

30 One aspect as reported herein is a synthetic feeder mix for the co-cultivation of hamster B-cells and feeder cells that is consisting of IL-1 β , TNF α , and IL-2 or IL-10, and optionally comprises IL-21, and/or SAC, and/or BAFF.

One aspect as reported herein is a synthetic feeder mix for the co-cultivation of hamster B-cells and feeder cells comprises IL-1 β , IL-2, IL-10, TNF- α , IL-6 and SAC.

One aspect as reported herein is a synthetic feeder mix for the co-cultivation of 5 rabbit B-cells and feeder cells that comprises IL-1 β , TNF α , IL-10, and IL-6.

One aspect as reported herein is a synthetic feeder mix for the co-cultivation of rabbit B-cells and feeder cells that comprises IL-1 β , TNF α , IL-10, IL-6 or IL-2, and SAC

In one specific embodiment IL-1 β , TNF α , IL-2, IL-10 and IL-21 are recombinant 10 murine IL-1 β , murine TNF α , murine IL-2, murine IL-10, and murine IL-21.

In one specific embodiment BAFF is added at a concentration of 5 ng/ml.

In one specific embodiment IL-6 is added at a concentration of 10 ng/ml.

In one specific embodiment SAC is added at a 1:75,000 ratio.

In one specific embodiment and feeder cells are murine EL-4 B5 cells.

15 The addition of an inhibitor of a certain potassium channel (= PAP-1, 5-(4-phenoxy butoxy) psoralene) surprisingly increases the rbIgG secretion of B-cells in a concentration dependent manner without decreasing the number of B-cell clones. Usually a cytokine which induced rbIgG productivity can be correlated with a decrease of the overall number of B-cell clones. This was not the case with PAP-1.

20 **Table 11:** Results of an rbIgG ELISA of cell culture supernatants of B-cells co-cultivated with EL-4 B5 feeder cells in the presence of TSN and SAC (=w/o) and different concentrations of PAP-1. DMSO: solvent for PAP-1 (1 μ M).

	w/o	0.01 μM	0.1 μM	1 μM	10 μM	DMSO
rbIgG ⁺ wells [n]	53	72	69	93	80	76
rbIgG ⁺ wells [% total wells]	21	29	27	37	32	30
rbIgG conc. of all huCk ⁺ wells [average ng/ml]	195.8	289.0	452.9	579.5	890.7	225.3

With a TSN concentration of 7.5 % the highest IgG concentration in the supernatant can be obtained.

Table 12: Influence of TSN on co-cultivation. A TSN concentration of 7.5 % results in improved B-cell growth and productivity

	5% TSN	7.5% TSN	10% TSN
rbIgG ⁺ wells [n]	71	71	81
rbIgG ⁺ wells [% total wells]	28	28	32
rbIgG conc. of all rbIgG ⁺ wells [Ø ng/ml]	246	512	372

5 With a number of 30,000 feeder cells per well of a 96-well plate the highest number of IgG⁺-wells in combination with IgG concentration in the supernatant can be obtained. In one embodiment of all methods as reported herein the number of feeder cells per single deposited B-cell is about 30,000.

Table 13: Influence of the amount of EL-4 B5 feeder cells on co-cultivation.

	20000	22000	24000	30000	35000	40000
rbIgG ⁺ wells [n]	71	73	78	78	73	38
rbIgG ⁺ wells [% total wells]	28	29	31	31	29	15
rbIgG conc. of all rbIgG ⁺ wells [Ø ng/ml]	246	319	346	418	457	656

10 The co-cultivation is in one embodiment of all methods as reported herein in polystyrene multi well plates with wells with a round bottom. The working volume of the wells is in one embodiment of all methods as reported herein of 50 µl to 250 µl. In one specific embodiment the wells are coated at least partially with a non-fibrous substrate prepared from a blend of polymer plastic resin and 15 amphipathic molecules, wherein the amphipathic molecule comprises a hydrophilic moiety and a hydrophobic region, wherein the hydrophobic regions are anchored within the substrate and the hydrophilic moieties are exposed on the substrate. In one specific embodiment the amphipathic molecules are chosen from alkylamine ethoxylated, poly (ethylene imine), octyldecamine or mixtures thereof (see e.g. 20 EP 1 860 181).

Characterization of co-cultivated cells:

For the (qualitative and quantitative) determination of secreted IgG after the co-cultivation generally all methods known to a person of skill in the art such as an ELISA can be used. In one embodiment of all methods as reported herein an 5 ELISA is used. In one specific embodiment for the determination of IgG secreted by murine B-cells an ELISA with the anti-IgG antibodies AB 216 (capture antibody) and AB 215 (tracer antibody) is used. In one specific embodiment for the determination of IgG secreted by hamster B-cells an ELISA with the monoclonal antibodies AB 220 (capture antibody) and AB 213 (tracer antibody) is used.

10 Depending on the characterization results a B-cell clone can be obtained, i.e. selected. The term "clone" denotes a population of dividing and antibody secreting B-cells arising from/originating from a single B-cell. Thus, a B-cell clone produces a monoclonal antibody.

Isolation of mRNA, cloning and sequencing:

15 From the B-cells the total mRNA can be isolated and transcribed in cDNA. With specific primers the cognate VH- and VL-region encoding nucleic acid can be amplified. With the sequencing of the therewith obtained nucleic acid it was confirmed that the obtained antibodies are monoclonal antibodies in most cases (71-95 %). Also can be seen from the sequencing of the individual B-cells that 20 almost no identical sequences are obtained. Thus, the method provides for highly diverse antibodies binding to the same antigen.

The primers used for the amplification of the VH-encoding nucleic acid can be used for cDNA obtained from cells from the NMRI-mouse, the Armenian Hamster, the Balb/c-mouse as well as the Syrian hamster and the rabbit.

25 In one embodiment of all methods as reported herein the amino acid sequence is derived from the amplified VH-encoding nucleic acid and the exact start and end point is identified by locating the amino acid sequences of EVQL/QVQL to VSS (VH-region) and DIVM/DIQM to KLEIK (VL-region).

30 The term "antibody" denotes a protein consisting of one or more polypeptide chain(s) substantially encoded by immunoglobulin genes. The recognized immunoglobulin genes include the different constant region genes as well as the myriad immunoglobulin variable region genes. Immunoglobulins may exist in a

variety of formats, including, for example, Fv, Fab, and F(ab)₂ as well as single chains (scFv), diabodies, monovalent, bivalent, trivalent or tetravalent forms, and also as bispecific, trispecific or tetraspecific form (e.g. Huston, J.S., et al., Proc. Natl. Acad. Sci. USA 85 (1988) 5879-5883; Bird, R.E., et al., Science 242 (1988) 5 423-426; in general, Hood et al., Immunology, Benjamin N.Y., 2nd edition (1984); and Hunkapiller, T. and Hood, L., Nature 323 (1986) 15-16).

Also reported herein is a method for producing an antibody comprising the following steps:

- a) providing a population of (mature) B-cells (obtained from the blood of an experimental animal),
- b) staining the cells of the population of B-cells with at least one fluorescence dye (in one embodiment with one to three, or two to three fluorescence dyes),
- c) depositing single cells of the stained population of B-cells in individual containers (in one embodiment is the container a well of a multi well plate),
- d) cultivating the deposited individual B-cells in the presence of feeder cells and a feeder mix (in one embodiment the feeder cells are EL-4 B5 cells, in one embodiment the feeder mix is natural TSN, in one embodiment the feeder mix is a synthetic feeder mix),
- e) determining the binding specificity of the antibodies secreted in the cultivation of the individual B-cells,
- f) determining the amino acid sequence of the variable light and heavy chain domain of specifically binding antibodies by a reverse transcriptase PCR and nucleotide sequencing, and thereby obtaining a monoclonal antibody variable light and heavy chain domain encoding nucleic acid,
- g) introducing the monoclonal antibody light and heavy chain variable domain encoding nucleic acid in an expression cassette for the expression of an antibody,
- h) introducing the nucleic acid in a cell,
- i) cultivating the cell and recovering the antibody from the cell or the cell culture supernatant and thereby producing an antibody.

An "expression cassette" refers to a construct that contains the necessary regulatory elements, such as promoter and polyadenylation site, for expression of at least the contained nucleic acid in a cell.

The term "experimental animal" denotes a non-human mammal. In one embodiment the experimental animal is selected from rat, mouse, hamster, rabbit, non-human primates, sheep, dog, cow, chicken, amphibians, and reptiles.

The following examples are provided to aid the understanding of the present invention, the true scope of which is set forth in the appended claims. It is understood that modifications can be made in the procedures set forth without departing from the spirit of the invention.

Examples

Example 1

Media and buffers:

Blocking buffer for ELISA comprises 1X PBS and 1% BSA.

Coating buffer for ELISA comprises 4.29g Na₂CO₃* 10 H₂O and 2.93g NaHCO₃ add water to a final volume of 1 liter, pH 9.6 adjusted with 2 N HCl.

Ethanol-solution for RNA isolation comprises 70 % Ethanol or 80 % Ethanol.

FACS-buffer for immuno fluorescence staining comprises 1X PBS and 0.1 % BSA.

IMDM-buffer for ELISA comprises 1X PBS, 5 % IMDM and 0.5 % BSA.

Incubation buffer 1 for ELISA comprises 1X PBS, 0.5 % CroteinC.

Incubation buffer 2 for ELISA comprises 1X PBS, 0.5 % CroteinC and 0.02 % Tween 20.

Incubation buffer 3 for ELISA comprises 1X PBS, 0.1 % BSA.

Incubation buffer 4 for ELISA comprises 1X PBS, 0.5 % BSA, 0.05 % Tween, PBS (10X), 0.01 M KH₂PO₄, 0.1 M Na₂HPO₄, 1.37 M NaCl, 0.027 M KCl, pH 7.0.

PCR-buffer comprises 500 mM KCl, 15 mM MgCl₂, 100 mM Tris/HCl, pH 9.0.

Wash buffer 1 for ELISA comprises 1X PBS, 0.05 % Tween 20.

Wash buffer 2 for ELISA comprises 1X PBS, 0.1 % Tween 20.

Wash buffer 3 for ELISA comprises water, 0.9 % NaCl, 0.05 % Tween 20.

EL-4 B5 medium comprises RPMI 1640, 10 % FCS, 1 %

5 Glutamin/Penicillin/Streptomycin-Mix, 2 % 100 mM sodium pyruvate, 1 % 1 M HEPES buffer.

Example 2

Animal care and immunization

The experimental animals were held according to the German animal protection law (TierSCHG) as well as according to the respective European guidelines.

10 Mice and hamster were received at an age of from 6 to 8 weeks and were immunized prior to an age of 12 weeks. The antigen was at first applied together with complete Freud's adjuvant (CFA). Further applications were with incomplete Freud's adjuvant (IFA). The antigen containing emulsion was applied 15 subcutaneously whereby the emulsion comprised an amount of from 50 to 100 µg antigen depending on the weight of the receiving experimental animal.

20 NZW rabbits (Charles River Laboratories International, Inc.) were used for immunization. The antigen was solved in K₃PO₄ buffer pH 7.0 at a concentration of 1 mg/ml and mixed (1:1) with complete Freud's adjuvant (CFA) till generation of stabile emulsion. The rabbits received an intra dermal (i.d.) injection of 2 ml of emulsion followed by a second intra muscular (i.m.) and third subcutaneous (s.c.) injection each with 1 ml in one week interval. The fourth i.m. injection of 1 ml was performed two weeks later followed by two further s.c. injections of 1 ml in four weeks interval.

25 During the immunization serum antibody titer was determined with an antigen specific assay. At an antibody titer with an IC₅₀ of 1:10000 the blood or the spleen of the immunized animal was removed. For reactivation of antigen specific B-cells 30 µg to 50 µg of the antigen was applied intravenously to the experimental animal three days prior to the removal of the blood or the spleen.

Example 3**Removal of organs, blood and macrophages**

Blood from mice and hamster was obtained by punctuation of the retrobulberic vein. Blood from rabbits was obtained by punctuation of the ear vein or, for larger volumes, of the ear artery. Whole blood (10 ml) was collected from rabbits 4-6 days after the third, fourth, fifth and sixth immunization and used for single cell sorting by FACS.

Macrophages were isolated from the obtained blood by attachment to cell culture plastic. From mice and hamsters, about 3×10^5 macrophages can be obtained from each animal by this method.

If a larger amount of mouse or hamster macrophages was required, peritoneal macrophages were isolated. For this the animals have to be at least 3 months of age. For the removal of peritoneal macrophages, animals were sacrificed and 5 ml of EL-4 B5 medium with a temperature of 37 °C was immediately injected into the peritoneal cavity. After kneading the animal's belly for 5 minutes, the solution containing the cells was removed.

Example 4**Cultivation of EL-4 B5 cells**

The frozen EL-4 B5 cells were thawed rapidly in a water bath at 37 °C and diluted with 10 ml EL-4 B5 medium. After centrifugation at 300 x g for 10 minutes the supernatant was discarded and the pellet resuspended in medium. After a further centrifugation step the supernatant was discarded again and the pellet was resuspended in 1 ml medium.

The EL-4 B5 cells were inoculated at a cell density of 3×10^4 cells/ml in 175 m² cultivation flasks. Cell density was determined every second day and adjusted to 3×10^4 cell/ml. The cells have a doubling time of approximately 12 hours and have to be cultivated at a cell density below 5×10^5 cell/ml because with higher cell density the stimulatory properties of the cells are lost.

When the total cell number was about 1.5×10^9 cells the medium was removed by centrifugation. Afterwards the cells were irradiated with 50 gray (5000 rad). After the determination of the viable cell number by trypan blue staining between 5×10^6 and 1×10^7 cells are aliquoted and frozen at -80 °C.

For co-cultivation the cells were thawed and washed twice with EL-4 B5 medium. For determination of the viable cell number the cell suspension is diluted 1:10 with 0.4 % (w/v) trypan blue solution and 10 µl of the mixture is transferred to a Neubauer counting chamber and cell number was counted.

5 **Example 5**

Density gradient centrifugation

The isolation of peripheral blood mononuclear cells (PBMCs) was effected by density gradient separation with Lympholyte® according to manufacturer's instructions A (Lympholyte®-mammal, cedarlane).

10 Withdrawn blood was diluted 2:1 with phosphate buffered saline (PBS). In a centrifuge vial the same volume of density separation medium was provided and the diluted blood is carefully added via the wall of the vial. The vial was centrifuged for 20 min. at 800 x g without braking. The lymphocytes were obtained from the white interim layer. The removed cells were supplemented with 10 ml
15 PBS and centrifuged at 800 x g for 10 min. The supernatant was discarded and the pellet was resuspended, washed, centrifuged. The final pellet was resuspended in PBS.

Example 6

Hypotonic lysis of red blood cells

20 For disruption of red blood cells by hypotonic lysis an ammonium chloride solution (BD Lyse™) was diluted 1:10 with water and added at a ratio of 1:16 to whole blood. For lysis of the red blood cells the mixture was incubated for 15 min. in the dark. For separation of cell debris from intact cells the solution was centrifuged for 10 min. at 800 x g. The supernatant was discarded, the pellet was resuspended in
25 PBS, washed again, centrifuged and the pellet was resuspended in PBS.

Example 7

Preparation of cells from inner organs of an experimental animal

For the preparation of spleen and thymus cells the respective organ was dissected in a Petri dish and the cells were taken up in PBS. For removal of remaining tissue the cell suspension was filtered through a 100 µm sieve. For obtaining lymphocytes from spleen cells density gradient centrifugation was employed. For thymus cells no further enrichment step was required.

Example 8**Depletion of macrophages**

Sterile 6-well plates (cell culture grade) were used to deplete macrophages and monocytes through unspecific adhesion. Wells were either coated with KLH (key hole limpet haemocyanine) or with streptavidin and the control peptides. Each well was filled with 3 ml to at maximum 4 ml medium and up to 6×10^6 peripheral blood mononuclear cells from the immunized rabbit and allowed to bind for 60 to 90 min. at 37 °C in the incubator. Thereafter the lymphocyte containing supernatant was transferred to a centrifugation vial and centrifuged at 800 x g for 10 min. The pellet was resuspended in PBS.

5 50 % of the cells in the supernatant were used for the panning step; the remaining 50 % of cells were directly subjected to immune fluorescence staining and single cell sorting.

Example 9**Depletion of KLH-specific B-cells**

15 Four milliliter of a solution containing keyhole limpet haemocyanine (KLH) was incubated with coating buffer at a concentration of 2 µg/ml in the wells of a multi well plate over night at room temperature. Prior to the depletion step the supernatant was removed and the wells were washed twice with PBS. Afterwards 20 the blood cells were adjusted to a cell density of 2×10^6 cells/ml and 3 ml are added to each well of a multi well plate. Afterwards the multi well plate was incubated for 60 to 90 min. at 37 °C. The supernatant was transferred to a centrifugation vial and the wells are washed twice with PBS and the supernatants are combined in the centrifugation vial. The cells were pelleted by centrifugation at 25 800 x g for 10 min. and the pellet was resuspended in PBS.

Example 10**Enrichment of antigen-specific B-cells**

30 The respective antigen was diluted with coating buffer to a final concentration of 2 µg/ml. 3 ml of this solution were added to the well of a 6-well multi well plate and incubated over night at room temperature. Prior to use the supernatant was removed and the wells were washed twice with PBS. The B-cell solution was adjusted to a concentration of 2×10^6 cells/ml and 3 ml are added to each well of a 6-well multi well plate. The plate was incubated for 60 to 90 min. at 37 °C. The supernatant was removed and the wells were washed two to four times with PBS.

For recovery of the antigen-specific B-cells 1 ml of a trypsin/EDTA-solution was added to the wells of the multi well plate and incubated for 10 to 15 min. at 37 °C. The incubation was stopped by addition of medium and the supernatant was transferred to a centrifugation vial. The wells were washed twice with PBS and the supernatants were combined with the other supernatants. The cells were pelleted by centrifugation for 10 min. at 800 x g. The pellet was resuspended in PBS.

Example 11

Co-cultivation of B-cells and EL-4 B5 cells

a) The co-cultivation was performed in 96-well multi well plates with round bottom. A basis solution comprising EL-4 B5 cells (1.6×10^6 cells / 15.2 ml) and cytokines in EL-4 B5 medium was prepared. 200 µl of the basis solution was added to each well of the multi well plate. To each well a single B-cell was added by fluorescence activated cell sorting. After the addition of the B-cells the plate was centrifuged for 5 min. at 300 x g. The plate is incubated for seven days at 37 °C.

b) Single sorted B cells were cultured in 96-well plates with 210 µl/well EL-4 B5 medium with Pansorbin Cells (1:20000) (Calbiochem (Merck), Darmstadt, Deutschland), 5 % rabbit thymocyte supernatant and gamma-irradiated EL-4-B5 murine thymoma cells (2×10^4 /well) for 7 days at 37 °C in an atmosphere of 5 % CO₂ in the incubator. B cell culture supernatants were removed for screening and the cells harvested immediately for variable region gene cloning or frozen at – 80 °C in 100 µl RLT buffer (Qiagen, Hilden, Germany).

Example 12

Cultivation of T-cells

The T-cells were isolated from the thymus of 3-4 week old mice and hamsters, or of 4-5 week old rabbits, respectively. The cells were centrifuged and immediately cultivated or frozen in aliquots of 3×10^7 cells. The thymocytes were seeded with a minimum cell density of 5×10^5 cells/ml of EL-4 B5 medium in 175 cm² culture flasks and incubated for 48 hours at 37 °C.

Example 13

Cultivation of macrophages

Macrophages were isolated from the peritoneal cavity of mice and hamsters, respectively, of an age of at least three months. Peritoneal macrophages from mice or hamsters, or blood mononuclear cells from rabbits were cultivated in EL-4 B5

medium at a cell density of at least 1×10^5 cells/ml in 175 cm² culture flasks for 1.5 hours at 37 °C. Afterwards the medium was removed and non-attached cells were removed from the attached macrophages by washing with warm EL-4 B5 medium, followed by cultivation for 48 hours in 35 ml medium.

5 **Example 14**

Co-cultivation of T-cells and macrophages

T-cells and macrophages were cultivated for 48 hours in separate flasks. Prior to combining both cell populations, the T-cells were centrifuged for 10 min. at 800 x g. The supernatant was discarded and the cell pellet was resuspended in 10 ml medium. The T-cells were adjusted to a minimal cell density of 5×10^5 cells/ml and 10 pg phorbol-12-myristate-13-acetate (PMA) and 5 ng or 50 ng Phytohemagglutinin M (PHA-M) per ml of medium were added. The cultivation medium was removed from macrophages and the T-cell suspension was added to the flasks containing macrophages. After 36 hours of co-cultivation, the cultivation medium was removed and was termed TSN solution. For removal of remaining cells the TSN solution was filtered through a 0.22 µm filter. The TSN solution was frozen at - 80 °C in aliquots of 4 ml.

10 **Example 15**

Immunofluorescence staining

20 Depending on the number of cells to be stained the cells were provided in 100 µl medium (less than 10^6 cells) or 200 µl medium (more than 10^6 cells), respectively. The fluorescent labeled antibody was diluted with 5 % serum of the experimental animal and FACS buffer to a final volume of 100 µl or 200 µl, respectively. The reaction mixture was incubated on a roller rack for 40 min. at 4 °C in the dark.

25 After the incubation the cells were washed twice at 300 x g for 5 min. The pellet was resuspended in 400 µl PBS and filtered through a 70 µm sieve. The filtered solution was transferred to a FACS-vial and directly before the FACS experiment dead cells were stained by addition of propidium iodide (6.25 µg/ml). If the labeled antibody was labeled with biotin the antibody was detected in a second step with streptavidin labeled Alexa Flour(R) 647 (antibody 197).

30 **Example 16**

Quantification of IgG

The 96-well multi well plate in which the co-cultivation was performed was centrifuged after seven days of co-cultivation at 300 x g for 5 min. 150 µl

supernatant was removed and diluted at a ratio of 2:1 with PBS in a second 96-well multi well plate.

The ELISA was performed as outlined in Example 17.

The antibody was used at a concentration of 50 ng/ml. If the OD was or exceeded 1 after an incubation time of 5 min. a dilution series of from 0.8 to 108 ng/ml IgG was tested.

Example 17

Detection of antigen-specific IgG

Antibodies produced by single deposited and co-cultivated B-cells or from B-cells obtained from an immunized experimental animal can be characterized with respect to specific antigen binding. The ELISA was performed at room temperature and the ELISA-solution was incubated between the individual steps on a shaker at 20 x g. In the first step the antigen was bound to the wells of a 96-well multi well plate. If the antigen was a protein it had been diluted in coating buffer and applied directly to the plate. Peptide antigens were bound via the specific binding pair biotin/streptavidin. The wells of the multi well plate can be already coated with soluble CroteinC (CrC) by the manufacturer. If not, the wells were incubated after the immobilization of the antigen with 200 µl blocking buffer. After the incubation with 100 µl antigen solution per well (pre-coated multi well plate) or 200 µl blocking buffer, respectively, non-bound antigen or blocking buffer was removed by washing with wash buffer. The diluted B-cell supernatants were added in a volume of 100 µl per well and incubated. After the incubation the wells were washed. Afterwards the detection antibody was added in a volume of 100 µl per well. The antibody can be either conjugated to horseradish peroxidase or labeled with biotin. The detection antibody was determined with a streptavidin-horseradish peroxidase conjugate. After the incubation the multi well plate was washed and afterwards 50 µl of a substrate solution containing 3,3',5,5' tetramethyl benzidine (TMB) were added per well and incubated for a period as given in Table X. The enzymatic reaction was stopped by the addition of 50 µl sulfuric acid and the optical density was determined at 450 nm and 680 nm with a photometer (Rainbow Thermo ELISA Reader) and the Xread plus-software.

Example 18**Isolation of ribonucleic acid (RNA)**

The cells from which the RNA had to be isolated were at first pelleted by centrifugation. The cell pellet was lysed by the addition of 100 µl RLT-buffer with 10 µl/ml beta-mercaptoethanol. The cells were resuspended by multiple mixing with a pipette. The solution was transferred to a well of a multi well plate. The plate was shortly shock at 200 x g and frozen at -20 °C.

The isolation of the RNA was performed with the RNeasy® Kit (Qiagen, Hilden, Germany) according to the manufacturer's instructions.

Example 19**Reverse transcription polymerase chain reaction**

The reverse transcription was carried out in a volume of 20 µl. For each reaction a control was performed with and without reverse transcriptase. Per reaction 1 µl dNTP (each at 10 mM), 0.4 µl oligo(dT)₁₂₋₁₈ (0.2 µg) and 0.6 µl random hexamer (0.03 µg) were pre-mixed and added to 8.5 µl RNA in H₂O. The reaction mixture was incubated for 5 min. at 65 °C and directly afterwards transferred to ice. Thereafter 2 µl RT-buffer (10 x), 4 µl MgCl₂ (25 mM), 2 µl DTT (0.1 M) and 1 µl RNase Out™ (40 units) were pre-mixed and added to the ice cold reaction mixture. After an incubation time of 2 min. at room temperature 0.5 µl Superscript™ II reverse transcriptase (25 units) were added. The reaction mixture was incubated for 10 min. at room temperature.

The translation was carried out for 50 min. at 42 °C. After the translation the reverse transcriptase was inactivated by incubation for 15 min. at 70 °C. The cDNA was stored at -20 °C.

Example 20**Polymerase chain reaction**

The polymerase chain reaction was carried out with the Taq PCR Core Kit (Qiagen, Hilden, Germany) according to the manufacturer's instructions. The PCR was carried out in a volume of 20 µl. The samples were transferred to the Mastercyler® at a temperature of 95°C.

Example 21**Sequencing**

All sequences were determined by SequiServe (Vaterstetten, Germany).

Example 22**5 Panning on antigen****a) Coating of plates**

Biotin/Streptavidin: Sterile streptavidin-coated 6-well plates (cell culture grade) were incubated with biotinylated antigen at a concentration of 0.5 - 2 µg/ml in PBS at room temperature for one hour. Plates were washed in sterile PBS three times before use.

Covalently bound protein: Cell culture 6-well plates were coated with 2 µg/ml protein in carbonate buffer (0.1 M sodium bicarbonate, 34 mM disodium hydrogen carbonate, pH 9.55) over night at 4 °C. Plates were washed in sterile PBS three times before use.

15 b) Panning of B-cells on peptides

6-well tissue culture plates coated with the respective antigen were seeded with up to 6×10^6 cells per 4 ml medium and allowed to bind for one hour at 37 °C in the incubator. Non-adherent cells were removed by carefully washing the wells 1-2 times with 1x PBS. The remaining sticky cells were detached by trypsin for 10 min. at 37 °C in the incubator and then washed twice in media. The cells were kept on ice until the immune fluorescence staining.

Patent Claims

1. A method for selecting a B-cell comprising the following steps:
 - a) co-cultivating each of the B-cells of a population of B-cells, which has been deposited as single cell, with feeder cells,
 - b) selecting a B-cell clone proliferating and secreting antibody in step a).
2. A method for producing an antibody binding to a target antigen comprising the following steps
 - a) co-cultivating each B-cell of a population of B-cells, which has been deposited as single cell in an individual container, in the presence of feeder cells and a feeder mix,
 - b) selecting a B-cell clone producing an antibody specifically binding to the target antigen,
 - c) cultivating a cell, which contains a nucleic acid that encodes the antibody produced by the B-cell clone selected in step b) or a humanized variant thereof, and recovering the antibody from the cell or the cultivation supernatant and thereby producing the antibody.
3. The method according to any one of the preceding claims further comprising the step of incubating the population of B-cells in the co-cultivation medium prior to single cell depositing.
- 20 4. The method according to claim 3, characterized in that the incubating is at about 37 °C for about one hour.
5. The method according to any one of the preceding claims further comprising the step of centrifuging the single cell deposited B-cells prior to the co-cultivation.
- 25 6. The method according to claim 5, characterized in that the centrifuging is for about 5 min. at about 300 x g.
7. The method according to any one of the preceding claims, characterized in that the population of B-cells is isolated by density gradient centrifugation.
- 30 8. The method according to any one of the preceding claims, characterized in that the B-cells are mature B-cells.

9. The method according to any one of the preceding claims, characterized in that the labeling is with two or three fluorescence dyes.
10. The method according to any one of the preceding claims, characterized in that 0.1 % to 2.5 % of the cells of the total B-cell population are labeled.
- 5 11. The method according to any one of the preceding claims, characterized in that the B-cells are mouse B-cells, or hamster B-cells, or rabbit B-cells.
12. The method according to any one of the preceding claims, characterized in that the feeder cells are murine EL-4 B5 cells.
13. The method according to any one of the preceding claims, characterized in 10 that the co-cultivating is in an RPMI 1640 medium supplemented with 10 % (v/v) FCS, 1 % (w/v) of a 200 mM glutamine solution that comprises penicillin and streptomycin, 2 % (v/v) of a 100 mM sodium pyruvate solution, and 1 % (v/v) of a 1 M 2-(4-(2-hydroxyethyl)-1-piperazine)-ethane sulfonic acid (HEPES) buffer.
- 15 14. The method according to any one of claims 1 to 10 and 12 to 13, characterized in that the B-cells are of mouse origin and the labeling is of IgG⁺CD19⁺-B-cells, and/or IgG⁻CD138⁺-B-cells.
15. The method according to any one of claims 1 to 10 and 12 to 13, characterized in that the B-cells are of hamster origin and the labeling is of 20 IgG⁺IgM⁻-B-cells.
16. The method according to any one of claims 1 to 10 and 12 to 13, characterized in that the B-cells are of rabbit origin and the labeling is of IgG⁺-B-cells and/or CD138⁺-B-cells, or CD138⁺IgG⁺-B-cells and/or IgG⁺IgM⁻-B-cells.
- 25 17. The method according to any one of the preceding claims, characterized in that the co-cultivating is in the presence of a synthetic feeder mix that comprises interleukin-1 beta and tumor necrosis factor alpha, and interleukin-2 and interleukin-10 and *Staphylococcus aureus* strain Cowans cells.
18. The method according to any one of claims 1 to 16, characterized in that the 30 co-cultivating is in the presence of a synthetic feeder mix that comprises

interleukin-1 beta and tumor necrosis factor alpha, and interleukin-2 and interleukin-10 and interleukin-21.

- 5 19. The method according to any one of claims 1 to 16, characterized in that the co-cultivating is in the presence of a synthetic feeder mix that comprises interleukin-1 beta and tumor necrosis factor alpha, and interleukin-2 and interleukin-10 and B-cell activation factor of the tumor necrosis factor family (BAFF).
- 10 20. The method according to any one of claims 1 to 16, characterized in that the co-cultivating is in the presence of thymocyte cultivation supernatant as feeder mix.
21. The method according to claim 20, characterized in that the thymocyte cultivation supernatant is obtained from thymocytes of the thymus gland of a young animal.

INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2011/058616

A. CLASSIFICATION OF SUBJECT MATTER
INV. G01N33/50
ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, BIOSIS, EMBASE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

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X	US 2007/269868 A1 (CARVALHO JENSEN ANNE E [US] ET AL) 22 November 2007 (2007-11-22)	1-8, 11-13, 17-21
Y	claims 1-9, 40, 59, 62	9, 10, 14-16
Y	----- MASRI ET AL: "Cloning and expression in E. coli of a functional Fab fragment obtained from single human lymphocyte against anthrax toxin", MOLECULAR IMMUNOLOGY, PERGAMON, GB LNKD-DOI:10.1016/J.MOLIMM.2006.09.007, vol. 44, no. 8, 1 December 2006 (2006-12-01), pages 2101-2106, XP005792748, ISSN: 0161-5890 abstract ----- - / --	9, 10, 14-16

Further documents are listed in the continuation of Box C.

See patent family annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance
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Date of the actual completion of the international search 31 August 2011	Date of mailing of the international search report 12/09/2011
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INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2011/058616

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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A	WO 91/16418 A1 (US HEALTH [US]) 31 October 1991 (1991-10-31) claim 1 -----	1-15
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INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No
PCT/EP2011/058616

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