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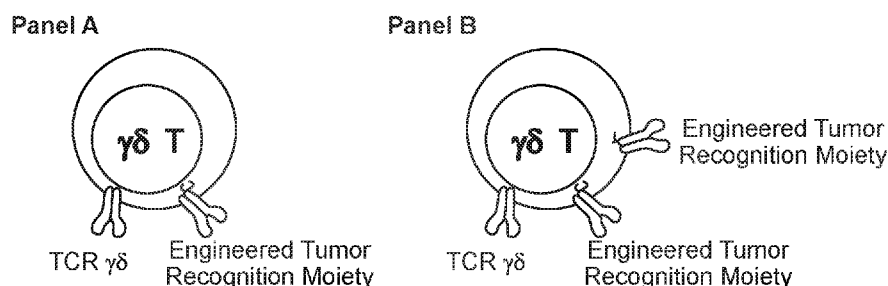
(54) Title: METHODS FOR SELECTIVE EXPANSION OF $\gamma\delta$ T-CELL POPULATIONS AND COMPOSITIONS THEREOF

FIG. 1

(57) Abstract: The present invention relates to methods for the selective expansion of $\gamma\delta$ T-cell population(s), compositions and admixtures thereof and methods for using the same as a therapeutic. Non-engineered and engineered, enriched $\gamma\delta$ T-cell populations of the disclosure are useful in the treatment of various cancers, infectious diseases, and immune disorders.



METHODS FOR SELECTIVE EXPANSION OF $\gamma\delta$ T-CELL POPULATIONS AND COMPOSITIONS THEREOF

CROSS-REFERENCE TO RELATED APPLICATION

5 This application claims priority under Section 119(e) and the benefit of U.S. Provisional Application Ser. No. 62/335,572, filed May 12, 2016, the entire disclosure of which is hereby incorporated by reference in the entirety and for all purposes.

BACKGROUND

10 Antigen recognition by T lymphocytes may be achieved by highly diverse heterodimeric receptors, the T-cell receptors (TCRs). Approximately 95% of human T-cells in blood and lymphoid organs express a heterodimeric $\alpha\beta$ TCR receptor ($\alpha\beta$ T-cell lineage). Approximately 5% of human T-cells in the blood and lymphoid organs express heterodimeric $\gamma\delta$ TCR receptor ($\gamma\delta$ T-cell lineage). These T-cell subsets may be referred to as ' $\alpha\beta$ ' and ' $\gamma\delta$ ' T-cells, respectively. $\alpha\beta$ and $\gamma\delta$ T-cells are
15 different in function. Activation of $\alpha\beta$ T-cells then occurs when an antigen presenting cell (APC) presents an antigen in the context of class I/II MHC. In contrast to $\alpha\beta$ T-cells, $\gamma\delta$ T-cells can recognize an antigen independent of MHC restriction. In addition, $\gamma\delta$ T-cells combine both innate and adoptive immune recognition and responses.

$\gamma\delta$ T cells utilize a distinct set of somatically rearranged variable (V), diversity (D), joining
20 (J), and constant (C) genes. $\gamma\delta$ T cells contain fewer V, D, and J segments than $\alpha\beta$ T cells. Although the number of germline $V\gamma$ and $V\delta$ genes is more limited than the repertoire of $V\alpha$ and $V\beta$ TCR genes, more extensive junctional diversification processes during TCR γ and δ chain rearrangement leads to a potential larger $\gamma\delta$ TCRs repertoire than that of $\alpha\beta$ TCRs (Carding and Egan, Nat Rev Immunol (2002) 2:336).

25 Human $\gamma\delta$ T-cells use 3 main $V\delta$ ($V\delta 1$, $V\delta 2$, $V\delta 3$) and at most six $V\gamma$ region genes to make their TCRs (Hayday AC., Annu Rev Immunol. 2000;18, 975-1026). Two main $V\delta$ subsets are $V\delta 1$ and $V\delta 2$ $\gamma\delta$ T cells. $V\delta 1$ T cells with different $V\gamma$ predominate in the intraepithelial subset of mucosal $\gamma\delta$ T cells where the TCRs appear to recognize stress molecules on epithelial cells (Beagley KW, Husband AJ. Crit Rev Immunol. 1998;18(3):237-254). $V\delta 2$ T cells that generally coexpress
30 $V\gamma 9$ are abundant in the peripheral blood and lymphatic system.

The ability of $\gamma\delta$ T-cells to recognize an antigen on diseased cells directly and to exhibit inherent ability to kill tumor cells renders $\gamma\delta$ T-cells an attractive therapeutic tool. Adoptive transfer of V γ 9V δ 2 T cells has yielded limited objective clinical responses for investigational treatment of cancer (Kondo et al, Cytotherapy, 10:842- 856, 2008; Lang et al, Cancer Immunology, Immunotherapy: CII, 60: 1447-1460, 2011; Nagamine et al , 2009; Nicol et al, British Journal of Cancer, 105:778-786, 2011; Wilhelm et al, Blood. 2003 Jul 1;102(1):200-6), indicating the need to isolate and test clinically new $\gamma\delta$ T-cell populations.

The ability to selectively expand $\gamma\delta$ T-cell subset populations having potent anti-tumor activity with improved purity and in clinically-relevant levels is highly desirable. Although antibodies and cytokine cocktails have been used to propagate a more diverse set of $\gamma\delta$ T cells, activation of specific $\gamma\delta$ T-cell subsets to sufficient purity and clinically-relevant levels, was not achieved (Dokouhaki et al, 2010; Kang et al, 2009; Lopez et al, 2000; Kress, 2006). Therefore, clinically-relevant methods of expanding specific $\gamma\delta$ T cell subsets *ex vivo*, and the cells produced thereby, are greatly needed.

SEQUENCE LISTING

The instant application contains a Sequence Listing which has been submitted in ASCII format via EFS-Web and is hereby incorporated by reference in its entirety. Said ASCII copy, created on March 3, 2016 is named 47165-701.201-SL.txt and is 4,366 Kilobytes in size.

SUMMARY OF THE INVENTION

The present invention, in one aspect, provides *ex vivo* methods for producing enriched $\gamma\delta$ T-cell populations. The enriched $\gamma\delta$ T-cell populations can be produced from isolated mixed cell populations by a method comprising contacting the mixed cell population, or a purified fraction thereof, with one or more agents:

- i) which selectively expand δ 1 T-cells by binding to an epitope specific of a δ 1 TCR,
- ii) which selectively expand δ 2 T-cells by binding to an epitope specific of a δ 2 TCR,
- iii) which selectively expand δ 1 and δ 4 T-cells by binding to an epitope specific of a δ 1 TCR and a δ 4 TCR, or

iv) which selectively expand $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T-cells by binding to an epitope specific of a $\delta 1$ TCR, a $\delta 3$ TCR, a $\delta 4$ TCR, and a $\delta 5$ TCR, to provide an enriched $\gamma\delta$ T cell population. In preferred embodiments, the enriched $\gamma\delta$ T cell population comprises a clinically-relevant level of $\gamma\delta$ T cells.

5 In another aspect, the invention provides *ex vivo* methods for producing an enriched $\gamma\delta$ T-cell population from an isolated mixed cell population, comprising directly contacting the mixed cell population with one or more agents:

i) which selectively expand $\delta 1$ T-cells by binding to an epitope specific of a $\delta 1$ TCR,

ii) which selectively expand $\delta 2$ T-cells by binding to an epitope specific of a $\delta 2$ TCR,

10 iii) which selectively expand $\delta 1$ and $\delta 4$ T-cells by binding to an epitope specific of a $\delta 1$ TCR and a $\delta 4$ TCR, or

iv) which selectively expand $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T-cells by binding to an epitope specific of a $\delta 1$ TCR, a $\delta 3$ TCR, a $\delta 4$ TCR, and a $\delta 5$ TCR, to provide a clinically-relevant level of an enriched $\gamma\delta$ T cell population.

15 In certain embodiments, the clinically-relevant level comprises more than about, or at least about, 10^8 $\gamma\delta$ T cells, 10^9 $\gamma\delta$ T cells, 10^{10} $\gamma\delta$ T cells, 10^{11} $\gamma\delta$ T cells, or 10^{12} $\gamma\delta$ T cells (*e.g.*, from about 10^8 to about 10^{12}). In one embodiment, the isolated mixed cell population is derived from a single donor and the method provides a clinical relevant level of an enriched $\gamma\delta$ T cell population expanded from a single donor, *e.g.*, from a single sample from the single donor or 2 or more samples
20 from the single donor. In other embodiments, the isolated mixed cell population is derived from more than one or multiple donors. In some embodiments, following the first enrichment step of the invention, the enriched $\gamma\delta$ T-cell population comprises clinically-relevant levels of $\gamma\delta$ T-cell subsets of $>10^8$. In other embodiments, following the second, third, fourth, fifth, etc. enrichment step of the invention, the enriched $\gamma\delta$ T-cell population comprises clinically-relevant levels of $\gamma\delta$ T-cell subsets
25 of $>10^8$.

In certain embodiments, the agent which selectively expands $\delta 1$ T-cells is selected from an agent which binds to the same epitope as an antibody selected from TS-1 and TS8.2. In some embodiments, the agent which selectively expands $\delta 1$ T-cells is an agent that binds a different epitope than the epitope bound by TS-1 and/or TS8.2 antibody. In some embodiments, the agent
30 which selectively expands $\delta 1$ T-cells is an agent that binds an epitope that does not overlap with the

epitope bound by, or does not compete with, TS-1 or TS8.2 antibody. In some embodiments, the agent which selectively expands $\delta 1$ T-cells is selected from an agent which specifically binds to an epitope comprising a $\delta 1$ variable region. In other embodiments, the agent binds to a $\delta 1$ TCR variable region comprising an amino acid sequence of a consensus sequence of **FIG. 24**. In yet
5 other embodiments, the agent binds to an epitope comprising residues Arg71 or Asp72 of the $\delta 1$ variable region, and/or Lys120 of a J1 or J2 region. In some embodiments, the agent has reduced binding to a mutant $\delta 1$ TCR polypeptide comprising a mutation at K120 of $\delta 1$ J1 or $\delta 1$ J2, e.g. K120A, K120G, K120P, K120V, K120E, K120D, or K120S.

In certain embodiments, the agent which selectively expands $\delta 1$ T-cells is selected from an
10 agent which binds to the same epitope as antibody R9.12. In some embodiments, the agent which selectively expands $\delta 1$ T-cells is an agent that binds a different epitope than the epitope bound by antibody R9.12. In some embodiments, the agent which selectively expands $\delta 1$ T-cells is an agent that binds an epitope that does not overlap with the epitope bound by, or does not compete with, antibody R9.12.

15 In certain embodiments, the agent that selectively expands $\delta 1$ T-cells is an agent that selectively binds to and/or expands $\delta 1$ and $\delta 4$ T-cells, or $\delta 1$, $\delta 3$, $\delta 4$ and $\delta 5$ T cells. In certain embodiments, the agent that selectively expands $\delta 1$ T-cells is an agent that comprises the complementarity determining regions (CDRs) of an antibody selected from the group consisting of $\delta 1$ -05, $\delta 1$ -08, $\delta 1$ -18, $\delta 1$ -22, $\delta 1$ -23, $\delta 1$ -26, $\delta 1$ -35, $\delta 1$ -37, $\delta 1$ -39, $\delta 1$ -113, $\delta 1$ -143, $\delta 1$ -149, $\delta 1$ -155, $\delta 1$ -
20 182, $\delta 1$ -183, $\delta 1$ -191, $\delta 1$ -192, $\delta 1$ -195, $\delta 1$ -197, $\delta 1$ -199, $\delta 1$ -201, $\delta 1$ -203, $\delta 1$ -239, $\delta 1$ -253, $\delta 1$ -257, $\delta 1$ -278, $\delta 1$ -282, and $\delta 1$ -285. In certain embodiments, the agent that selectively expands $\delta 1$ T-cells is an agent that comprises the variable regions of an antibody selected from the group consisting of $\delta 1$ -05, $\delta 1$ -08, $\delta 1$ -18, $\delta 1$ -22, $\delta 1$ -23, $\delta 1$ -26, $\delta 1$ -35, $\delta 1$ -37, $\delta 1$ -39, $\delta 1$ -113, $\delta 1$ -143, $\delta 1$ -149, $\delta 1$ -155, $\delta 1$ -182, $\delta 1$ -183, $\delta 1$ -191, $\delta 1$ -192, $\delta 1$ -195, $\delta 1$ -197, $\delta 1$ -199, $\delta 1$ -201, $\delta 1$ -203, $\delta 1$ -239, $\delta 1$ -253, $\delta 1$ -257, $\delta 1$ -278,
25 $\delta 1$ -282, and $\delta 1$ -285. In certain embodiments, the agent that selectively expands $\delta 1$ T-cells is an agent that binds the same epitope, or essentially the same epitope, as, or competes with, an antibody selected from the group consisting of $\delta 1$ -05, $\delta 1$ -08, $\delta 1$ -18, $\delta 1$ -22, $\delta 1$ -23, $\delta 1$ -26, $\delta 1$ -35, $\delta 1$ -37, $\delta 1$ -39, $\delta 1$ -113, $\delta 1$ -143, $\delta 1$ -149, $\delta 1$ -155, $\delta 1$ -182, $\delta 1$ -183, $\delta 1$ -191, $\delta 1$ -192, $\delta 1$ -195, $\delta 1$ -197, $\delta 1$ -199, $\delta 1$ -201, $\delta 1$ -203, $\delta 1$ -239, $\delta 1$ -253, $\delta 1$ -257, $\delta 1$ -278, $\delta 1$ -282, and $\delta 1$ -285. In certain embodiments, the agent

that selectively expands $\delta 1$ T-cells is an antibody selected from the group consisting of $\delta 1$ -05, $\delta 1$ -08, $\delta 1$ -18, $\delta 1$ -22, $\delta 1$ -23, $\delta 1$ -26, $\delta 1$ -35, $\delta 1$ -37, $\delta 1$ -39, $\delta 1$ -113, $\delta 1$ -143, $\delta 1$ -149, $\delta 1$ -155, $\delta 1$ -182, $\delta 1$ -183, $\delta 1$ -191, $\delta 1$ -192, $\delta 1$ -195, $\delta 1$ -197, $\delta 1$ -199, $\delta 1$ -201, $\delta 1$ -203, $\delta 1$ -239, $\delta 1$ -253, $\delta 1$ -257, $\delta 1$ -278, $\delta 1$ -282, and $\delta 1$ -285. In certain embodiments, the agent that selectively expands $\delta 1$ T-cells is an antibody that binds a Bin 1 $\delta 1$ epitope, a Bin 1b $\delta 1$ epitope, a Bin 2 $\delta 1$ epitope, a Bin 2b $\delta 1$ epitope, a Bin 2c $\delta 1$ epitope, a Bin 3 $\delta 1$ epitope, a Bin 4 $\delta 1$ epitope, a Bin 5 $\delta 1$ epitope, a Bin 6 $\delta 1$ epitope, a Bin 7 $\delta 1$ epitope, a Bin 8 $\delta 1$ epitope, or a Bin 9 $\delta 1$ epitope.

In some embodiments, the methods of the invention provide enriched $\gamma\delta$ T-cell population(s) comprising greater than 60%, 70%, 80% or 90% $\delta 1$ cells from an isolated mixed cell population comprising T lymphocytes. In some embodiments, the methods of the invention provide enriched $\gamma\delta$ T-cell population(s) comprising greater than 60%, 70%, 80% or 90% $\delta 1$ cells from an isolated, e.g., mixed, cell population comprising T lymphocytes, before a step of purifying an expanded $\gamma\delta$ T-cell population, e.g., by positive selection of $\gamma\delta$ T-cells (e.g., positive selection of $\delta 1$ T cells, $\delta 1$ and $\delta 3$ $\gamma\delta$ T cells; $\delta 1$ and $\delta 4$ $\gamma\delta$ T cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells (e.g., $\gamma\delta$ T cells of the indicated δ -subtype(s)); and/or positive selection of $\delta 2$ T cells) from the enriched $\gamma\delta$ T-cell population(s), or by depletion of non- $\gamma\delta$ T-cells (e.g., non- $\delta 1$ T cells such as $\alpha\beta$ T cells) from the enriched $\gamma\delta$ T-cell population(s).

In some embodiments, the methods of the present invention provide a $\gamma\delta$ T-cell population comprising $\delta 1$ $\gamma\delta$ T cells and $\delta 2$ $\gamma\delta$ T cells, wherein the population is greater than 60%, 70%, 80% or 90% $\delta 1$ $\gamma\delta$ T cells. In some embodiments, the methods of the present invention provide a $\gamma\delta$ T-cell population comprising $\delta 1$ $\gamma\delta$ T cells; $\delta 2$ $\gamma\delta$ T cells; $\delta 1$ and $\delta 4$ $\gamma\delta$ T cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ $\gamma\delta$ T cells, wherein (i) the population is greater than 60%, 70%, 80% or 90% $\delta 1$ $\gamma\delta$ T cells; (ii) the population is greater than 60%, 70%, 80% or 90% $\delta 1$ and $\delta 3$ $\gamma\delta$ T cells; (iii) the population is greater than 60%, 70%, 80% or 90% $\delta 1$ and $\delta 4$ $\gamma\delta$ T cells; or (iv) the population is greater than 60%, 70%, 80% or 90% $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ $\gamma\delta$ T cells. In some embodiments, the methods of the present invention provide a composition comprising a population of $\gamma\delta$ T cells (e.g., $\delta 1$ $\gamma\delta$ T cells) that is free of, or contains less than about 2%, 1%, 0.5%, 0.4%, 0.1%, 0.05%, or 0.01%, $\alpha\beta$ T cells.

In certain embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises about or less than about 30%, 25%, 20%, 15%, 10%, 5%, 4%, 3%, 2%, 1.5%, 1.2%, 1%, 0.9%, 0.8%, 0.7%, 0.6%, 0.5%, 0.4%, 0.3%, 0.2%, or 0.1%

$\gamma\delta$ T cells (e.g., $\delta 1$, $\delta 2$, $\delta 1$ and $\delta 2$, $\delta 1$ and $\delta 3$, or $\delta 1$ and $\delta 4$ $\gamma\delta$ T cells, or a combination thereof). In certain embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises from about 0.5% to about 5% $\gamma\delta$ T cells (e.g., $\delta 1$, $\delta 2$, $\delta 3$, $\delta 4$, or $\delta 8$ $\gamma\delta$ T cells, or a combination thereof).

5 In certain embodiments, the isolated mixed cell population is from a donor or multiple donors having a larger than median level of circulating or sample- (e.g., tumor- or epithelial sample-) infiltrating $\gamma\delta$ T-cells. Thus, for example, in some embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises from about 0.5% to about 10% $\gamma\delta$ T cells (e.g., $\delta 1$, $\delta 2$, $\delta 3$, $\delta 4$, or $\delta 8$ $\gamma\delta$ T cells, or a combination thereof). As another
10 example, in some embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises about from about 10% to about 30% $\gamma\delta$ T cells (e.g., $\delta 1$, $\delta 2$, $\delta 3$, $\delta 4$, or $\delta 8$ $\gamma\delta$ T cells, or a combination thereof). In some embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises about or less than about 1.5% $\gamma\delta$ T cells, preferably less than about 1.2% $\gamma\delta$ T cells, more
15 preferably less than about 1% $\gamma\delta$ T cells, yet more preferably less than about 0.5% $\gamma\delta$ T cells, e.g., from about 0.1% to about 0.4% $\gamma\delta$ T cells.

In certain embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises about 80%, 70%, 60%, 50%, 40%, 30%, or 20% T lymphocytes and less than about 30%, 25%, 20%, 15%, 10%, 5%, 4%, 3%, 2%, 1.5%, 1.2%, 1%,
20 0.9%, 0.8%, 0.7%, 0.6%, 0.5%, 0.4%, 0.3%, 0.2%, or 0.1% $\gamma\delta$ T cells (e.g., $\delta 1$, $\delta 2$, $\delta 1$ and $\delta 2$, $\delta 1$ and $\delta 3$, or $\delta 1$ and $\delta 4$ $\gamma\delta$ T cells, or a combination thereof). In certain embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises about 80%, 70%, 60%, 50%, 40%, 30%, or 20% T lymphocytes and from about 0.5% to about 5% $\gamma\delta$ T cells (e.g., $\delta 1$, $\delta 2$, $\delta 3$, $\delta 4$, or $\delta 8$ $\gamma\delta$ T cells, or a combination thereof).

25 In certain embodiments, the isolated mixed cell population is from a donor or multiple donors having a larger than median level of circulating or sample- (e.g., tumor- or epithelial sample-) infiltrating $\gamma\delta$ T-cells. Thus, for example, in some embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises about 80%, 70%, 60%, 50%, 40%, 30%, or 20% T lymphocytes and from about 0.5% to about 10% $\gamma\delta$ T cells (e.g., $\delta 1$,
30 $\delta 2$, $\delta 3$, $\delta 4$, or $\delta 8$ $\gamma\delta$ T cells, or a combination thereof). As another example, in some embodiments,

the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises about 80%, 70%, 60%, 50%, 40%, 30%, or 20% T lymphocytes and from about 10% to about 30% $\gamma\delta$ T cells (e.g., $\delta 1$, $\delta 2$, $\delta 3$, $\delta 4$, or $\delta 8$ $\gamma\delta$ T cells, or a combination thereof). In some embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises about 80%, 70%, 60%, 50%, 40%, 30%, or 20% T lymphocytes and about or less than about 1.5% $\gamma\delta$ T cells, preferably less than about 1.2% $\gamma\delta$ T cells, more preferably less than about 1% $\gamma\delta$ T cells, yet more preferably less than about 0.5% $\gamma\delta$ T cells, e.g., from about 0.1% to about 0.4% $\gamma\delta$ T cells.

In some embodiments, the isolated mixed cell population is selected from a peripheral blood sample (e.g., whole blood, PBMCs, or PBLs), a leukapheresis sample, a cord blood sample, a tumor sample, or a tissue sample (e.g., an epithelial sample). In some embodiments, the isolated mixed cell population is derived from a single donor. In other embodiments, the isolated mixed cell population is derived from more than one or multiple donors. In certain embodiments, the methods of the invention provide enriched $\gamma\delta$ T-cell population(s) comprising polyclonal $\gamma\delta$ TCR diversity.

In other embodiments, the agent which selectively expands $\delta 2$ T-cells is selected from an agent which binds to the same epitope as an antibody selected from B6 and 15D. In yet other embodiments, the agent which selectively expands $\delta 2$ T-cells is selected from an agent which binds to a different epitope than an antibody selected from B6 and 15D. In still other embodiments, the agent which selectively expands $\delta 2$ T-cells is selected from an agent which binds to an epitope that does not overlap with an epitope bound by, or compete with, a B6 or 15D antibody. In one embodiment, the agent which selectively expands $\delta 2$ T-cells is selected from an agent which specifically binds to an epitope comprising a $\delta 2$ variable region. In a specific embodiment, the agent has reduced binding to a mutant $\delta 2$ TCR polypeptide comprising a mutation at G35 of the $\delta 2$ variable region.

In certain embodiments, the agent that selectively expands $\delta 2$ T-cells is an agent that comprises the complementarity determining regions (CDRs) of an antibody selected from the group consisting of $\delta 2$ -14, $\delta 2$ -17, $\delta 2$ -22, $\delta 2$ -30, $\delta 2$ -31, $\delta 2$ -32, $\delta 2$ -33, $\delta 2$ -35, $\delta 2$ -36, and $\delta 2$ -37 (or $\delta 2$ -14, $\delta 2$ -17, $\delta 2$ -30, $\delta 2$ -31, $\delta 2$ -32, $\delta 2$ -33, $\delta 2$ -35, $\delta 2$ -36, and $\delta 2$ -37). In certain embodiments, the agent that selectively expands $\delta 2$ T-cells is an agent that comprises the variable regions of an antibody selected

from the group consisting of $\delta 2$ -14, $\delta 2$ -17, $\delta 2$ -22, $\delta 2$ -30, $\delta 2$ -31, $\delta 2$ -32, $\delta 2$ -33, $\delta 2$ -35, $\delta 2$ -36, and $\delta 2$ -37 (or $\delta 2$ -14, $\delta 2$ -17, $\delta 2$ -30, $\delta 2$ -31, $\delta 2$ -32, $\delta 2$ -33, $\delta 2$ -35, $\delta 2$ -36, and $\delta 2$ -37).

In certain embodiments, the agent that selectively expands $\delta 2$ T-cells is an agent that binds the same, or essentially the same, epitope as, or competes with, an antibody selected from the group consisting of $\delta 2$ -14, $\delta 2$ -17, $\delta 2$ -22, $\delta 2$ -30, $\delta 2$ -31, $\delta 2$ -32, $\delta 2$ -33, $\delta 2$ -35, $\delta 2$ -36, and $\delta 2$ -37 (or $\delta 2$ -14, $\delta 2$ -17, $\delta 2$ -30, $\delta 2$ -31, $\delta 2$ -32, $\delta 2$ -33, $\delta 2$ -35, $\delta 2$ -36, and $\delta 2$ -37). In certain embodiments, the agent that selectively expands $\delta 2$ T-cells is an antibody selected from the group consisting of $\delta 2$ -14, $\delta 2$ -17, $\delta 2$ -22, $\delta 2$ -30, $\delta 2$ -31, $\delta 2$ -32, $\delta 2$ -33, $\delta 2$ -35, $\delta 2$ -36, and $\delta 2$ -37 (or $\delta 2$ -14, $\delta 2$ -17, $\delta 2$ -30, $\delta 2$ -31, $\delta 2$ -32, $\delta 2$ -33, $\delta 2$ -35, $\delta 2$ -36, and $\delta 2$ -37). In certain embodiments, the agent that selectively expands $\delta 2$ T-cells is an antibody that binds a Bin 1 $\delta 2$ epitope, a Bin 2 $\delta 2$ epitope, a Bin 3 $\delta 2$ epitope, or a Bin 4 $\delta 2$ epitope.

In some embodiments, the methods of the invention provide enriched $\gamma\delta$ T-cell population(s) comprising greater than 60%, 70%, 80% or 90% $\delta 2$ cells from an isolated mixed cell population comprising T lymphocytes. In some embodiments, the methods of the invention provide enriched $\gamma\delta$ T-cell population(s) comprising greater than 60%, 70%, 80% or 90% $\delta 2$ cells from an isolated, e.g., mixed, cell population comprising T lymphocytes, before a step of purifying an expanded $\gamma\delta$ T-cell population, e.g., by positive selection of $\gamma\delta$ T-cells (e.g., positive selection of $\delta 2$ T cells) from the enriched $\gamma\delta$ T-cell population(s), or by depletion of non- $\gamma\delta$ T-cells (e.g., depletion of non- $\delta 2$ T cells or depletion of $\alpha\beta$ T cells) from the enriched $\gamma\delta$ T-cell population(s). In some embodiments, the methods of the present invention provide a $\gamma\delta$ T-cell population comprising $\delta 2$ $\gamma\delta$ T cells and $\delta 1$ $\gamma\delta$ T cells, wherein the population is greater than 60%, 70%, 80% or 90% $\delta 2$ $\gamma\delta$ T cells. In some embodiments, the methods of the present invention provide a composition comprising a population of $\delta 2$ $\gamma\delta$ T cells that is free of, or contains less than about 2%, 1%, 0.5%, 0.4%, 0.1%, 0.05%, or 0.01%, $\alpha\beta$ T cells.

In certain embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises about 80%, 70%, 60%, 50%, 40%, 30%, or 20% T lymphocytes and less than about 60%, 50%, 40%, 30%, 25%, 20%, 15%, 10%, 5%, 4%, 3%, 2%, 1%, 0.9%, 0.8%, 0.7%, 0.6%, 0.5%, 0.4%, 0.3%, 0.2%, 0.1%, 0.05%, or 0.02% $\delta 2$ cells. In some embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein is selected from a peripheral blood sample (e.g., PBMC or PBL), a

leukapheresis sample, a cord blood sample, a tumor, or a tissue. In certain embodiments, the methods of the invention provide an enriched $\gamma\delta$ T-cell population(s) comprising polyclonal TCR diversity.

In certain embodiments, the enriched $\gamma\delta$ T-cell population is not expanded by, or not
5 expanded in the presence of, an antigen presenting cell (APC), an artificial antigen presenting cell (aAPC), an irradiated population of antigen presenting cells (e.g., irradiated PBMCs, an irradiated immortalized cell line, or irradiated aAPCs), an aminophosphonate, an aminophosphate, a bisphosphonate, or a combination thereof. In certain embodiments, the enriched $\gamma\delta$ T-cell population is not expanded by, or not expanded in the presence of, irradiated PBMCs. In certain
10 embodiments, the enriched $\gamma\delta$ T-cell population is not expanded by, or not expanded in the presence of, irradiated aAPCs (e.g., engineered K562, RPMI8226, T2, or JVM-3). In certain embodiments, the enriched $\gamma\delta$ T-cell population is not expanded by, or not expanded in the presence of, an irradiated population of cells of an immortalized cell line. In preferred embodiments, said one or more agents which selectively expand $\delta 1$ T-cells, $\delta 2$ T-cells, $\delta 1$ T-cells and $\delta 4$ T-cells, or $\delta 1$, $\delta 3$, $\delta 4$,
15 and $\delta 5$ T cells (e.g., $\gamma\delta$ T cells) are antibodies.

In some embodiments, said one or more agents which selectively expand $\delta 1$ T-cells, $\delta 2$ T-cells, $\delta 1$ T-cells and $\delta 4$ T-cells, or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells (e.g., $\gamma\delta$ T cells of the indicated δ -subtype(s)) are immobilized on a surface. In some embodiments, said one or more agents which selectively expand $\delta 1$ T-cells, $\delta 2$ T-cells, $\delta 1$ T-cells and $\delta 4$ T-cells, or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells (e.g.,
20 $\gamma\delta$ T cells of the indicated δ -subtype(s)) are immobilized on a surface of an antigen presenting cell (e.g., an aAPC) in a first and/or second or subsequent expansion. The agents immobilized on the surface of an antigen presenting cell can, e.g., be bound to an Fc receptor expressed on the surface of the APC or expressed on the surface of the APC.

In some embodiments, the methods of the present invention are performed using a culture
25 medium supplemented with IL-2, IL-4, IL-7, IL-9, IL-12, IL-15, IL-18, IL-19, IL-21, IL-23, IL-33, , IFN γ , granulocyte-macrophage colony stimulating factor (GM-CSF), granulocyte colony stimulating factor (G-CSF), a lectin (e.g., PHA-E, PHA-L, or ConA), or a combination of two or more, or all thereof. In some embodiments, the methods of the present invention are performed using a culture medium that is not supplemented with IL-21. In some embodiments, the methods of the present
30 invention are performed with a first $\gamma\delta$ T-cell expansion using a culture medium supplemented with

IL-2, IL-4, IL-7, IL-9, IL-12, IL-15, IL-18, IL-19, IL-21, IL-23, IL-33, IFN γ , granulocyte-macrophage colony stimulating factor (GM-CSF), granulocyte colony stimulating factor (G-CSF), a lectin (e.g., PHA-E, PHA-L, or ConA), or a combination of two or more, or all thereof. In some embodiments, the methods of the present invention are performed with a first $\gamma\delta$ T-cell expansion
5 using a culture medium that is not supplemented with IL-21. In some embodiments, the methods of the present invention are performed with a second $\gamma\delta$ T-cell expansion using a culture medium supplemented with IL-2, IL-4, IL-7, IL-9, IL-12, IL-15, IL-18, IL-19, IL-21, IL-23, IL-33, IFN γ , granulocyte-macrophage colony stimulating factor (GM-CSF), granulocyte colony stimulating factor (G-CSF), a lectin (e.g., PHA-E, PHA-L, or ConA), or a combination of two or more, or all thereof.

10 In some embodiments, the methods of the present invention are performed with a second $\gamma\delta$ T-cell expansion using a culture medium that is not supplemented with IL-21.

In some embodiments, the methods of the present invention are performed with a first $\gamma\delta$ T-cell expansion comprising any of the $\gamma\delta$ T-cell expansion methods or compositions described herein, and then a second $\gamma\delta$ T-cell expansion using a culture medium that is free of the one or more agents
15 which selectively expand $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 3$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells (*e.g.*, $\gamma\delta$ T cells of the indicated δ -subtype(s)). In some embodiments, the methods of the present invention are performed with a first $\gamma\delta$ T-cell expansion comprising any of the $\gamma\delta$ T-cell expansion methods or compositions (*e.g.*, activation agents) described herein, which method includes contacting an engineered or non-engineered $\gamma\delta$ T-cell, a population of engineered or
20 non-engineered $\gamma\delta$ T-cells, and/or an isolated mixed population of cells with an immobilized agent that selectively expands $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells (*e.g.*, $\gamma\delta$ T cells of the indicated δ -subtype(s)), and then a second $\gamma\delta$ T-cell expansion using a culture medium that: i) is free of the one or more agents which selectively expand $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells; ii) contains an (*e.g.*, immobilized)
25 structurally different agent that selectively expands $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells; iii) contains a non-immobilized (soluble) agent that selectively expands $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells; or iv) contains an agent that expands T cells (*e.g.*, expands $\alpha\beta$ and $\gamma\delta$ T cells) or selectively expands $\gamma\delta$ T cells as compared to $\alpha\beta$ T cells.

In some cases, the agent that expands T cells is an agent that binds CD3, such as an anti-CD3 antibody (*e.g.*, OKT3). In some cases, the agent that selectively expands $\gamma\delta$ T cells is an agent that binds a γ -chain constant region or a δ -chain constant region, or an agent that binds $\gamma\delta$ TCR, such as an antibody that binds $\gamma\delta$ TCR (*e.g.*, IMMU510). In some cases, the $\gamma\delta$ T-cell population is enriched
5 by positive selection between the first cell expansion and the second cell expansion, and/or after the second cell expansion. In some cases, the $\gamma\delta$ T-cell population is enriched by depletion of $\alpha\beta$ T cells between the first cell expansion and the second cell expansion, and/or after the second cell expansion.

In certain embodiments of the methods of the present invention, an, *e.g.* enriched, $\gamma\delta$ T-cell population is expanded in a first, second, or subsequent expansion in a culture medium containing an
10 antigen presenting cell (APC). In some embodiments, the culture medium contains one or more agents that expand T cells, expand $\gamma\delta$ T-cells, selectively expand $\gamma\delta$ T-cells, or selectively expands $\delta 1$ T-cells, $\delta 2$ T-cells, $\delta 1$ T-cells and $\delta 3$ T-cells, or $\delta 1$ T-cells and $\delta 4$ T-cells (*e.g.*, an antibody that binds a constant or variable region of a $\gamma\delta$ TCR, an antibody that binds CD3, and/or an aminophosphonate). In certain embodiments, the enriched $\gamma\delta$ T-cell population is expanded in a
15 culture medium containing irradiated PBMCs. In certain embodiments, the enriched $\gamma\delta$ T-cell population is expanded in a culture medium containing irradiated artificial APCs (aAPCs). In certain embodiments, the enriched $\gamma\delta$ T-cell population is expanded in a culture medium containing irradiated population of cells of an immortalized cell line (*e.g.*, K562 APCs). In some cases, the APCs do not express, or exhibit reduced expression of, HLA class I, HLA class II, HLA class II invariant chain, and/or HLA-DM. In some cases, the APCs express adhesion or co-stimulatory
20 molecules such as intercellular adhesion molecule-1, CD11a, CD18, CD54, CD80, CD86, 4-1BBL, OX-40L, CD70, or one or more $\gamma\delta$ T cell activating agents anchored in the membrane, and/or leukocyte function-associated antigen-3. In some cases, the APCs express an Fc receptor, such as an Fc receptor that is specific for an isotype of an activation agent used in a $\gamma\delta$ T-cell expansion method
25 described herein. In some cases, the APCs express one or more Fc receptors selected from the group consisting of CD64, CD32A, CD32B, CD32C, CD16A, CD16B, FcRn, TRIM21, or CD307, or an engineered variant thereof having a higher affinity or altered specificity.

In some embodiments, the culture medium containing an antigen presenting cell (APC), an artificial antigen presenting cell (aAPC), or an irradiated population of antigen presenting cells (*e.g.*,
30 PBMCs, an immortalized cell line, or aAPCs) further contains one or more agents that selectively

expand $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 3$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells. In certain embodiments, the, e.g., irradiated, PBMCs, APCs, aAPCs, or population of cells of an immortalized cell line express on their cell surface the one or more agents that selectively expand $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells. In some preferred embodiments, said one or more agents which selectively expand $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells are antibodies. In some embodiments, said one or more agents that selectively expand $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 4$; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T-cells are antibodies expressed on the surface of the APC or bound to an Fc-receptor (e.g., Fc γ receptor) expressed on the surface of the APC.

In some embodiments, the invention provides *ex vivo* methods for producing an enriched $\gamma\delta$ T-cell population from an isolated, e.g., mixed, cell population, comprising (i) directly contacting the, e.g., mixed, cell population with one or more agents which (a) expand $\gamma\delta$ T-cells (e.g., by binding to a $\gamma\delta$ TCR), or (b) selectively expand:

- $\delta 1$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR;
- $\delta 2$ T-cells by binding to an activating epitope specific of a $\delta 2$ TCR;
- $\delta 1$ T-cells and $\delta 4$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR and a $\delta 4$ TCR; or

- $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells by binding to an activating epitope specific of a $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ TCR in a first $\gamma\delta$ T-cell expansion, thereby producing a first enriched $\gamma\delta$ T-cell population, and then

(ii) directly contacting at least a portion of the first enriched $\gamma\delta$ T-cell population with antigen presenting cells (APCs), optionally in the presence of one or more soluble or immobilized agents that (a) expand T cells, (b) expand $\gamma\delta$ T-cells (e.g., by binding to a $\gamma\delta$ TCR), or (c) selectively expands:

- $\delta 1$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR;
- $\delta 2$ T-cells by binding to an activating epitope specific of a $\delta 2$ TCR;
- $\delta 1$ T-cells and $\delta 4$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR and a $\delta 4$ TCR; or

- $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells by binding to an activating epitope specific of a $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ TCR in a second $\gamma\delta$ T-cell expansion, thereby producing a second enriched $\gamma\delta$ T-cell population,

wherein the second enriched $\gamma\delta$ T-cell population comprises a clinically relevant number (e.g., $> 10^8$) of $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells.

In some cases, the clinically relevant number (e.g., $> 10^8$) of the $\gamma\delta$ T-cells is obtained from a single donor, e.g., from a single sample or 2 or more samples from the donor. In some cases, the clinically relevant number (e.g., $> 10^8$) of the $\gamma\delta$ T-cells is obtained from a single donor, e.g., from a single sample or 2 or more samples from the donor, within less than 30 days of expansion, preferably less than 21 days of expansion, more preferably within 19 days of expansion.

In some cases, the $\gamma\delta$ T-cell population is enriched by positive selection between (i) and (ii), or after (ii). In some cases, the $\gamma\delta$ T-cell population is enriched by depletion of $\alpha\beta$ T cells between (i) and (ii), or after (ii). In some cases, the agent that expands T cells is an agent that binds CD3, such as an anti-CD3 antibody. In some cases, the agent that selectively expands $\gamma\delta$ T cells is an agent that binds a γ -chain constant region or a δ -chain constant region, or an agent that binds $\gamma\delta$ TCR, such as an antibody that binds $\gamma\delta$ TCR (e.g., IMMU510).

In certain embodiments of the methods of the present invention, an enriched $\gamma\delta$ T-cell population is produced by (i) contacting an isolated, e.g., mixed, cell population with one or more first agents which (a) expand $\gamma\delta$ T-cells (e.g., by binding to a $\gamma\delta$ TCR) or (b) selectively expands:

- $\delta 1$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR;
- $\delta 2$ T-cells by binding to an activating epitope specific of a $\delta 2$ TCR;
- $\delta 1$ T-cells and $\delta 4$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR and a $\delta 4$

TCR; or

- $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells by binding to an activating epitope specific of a $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ TCR to provide a first enriched $\gamma\delta$ T cell population in a first $\gamma\delta$ T-cell expansion; and then (ii) in a second $\gamma\delta$ T-cell expansion, contacting the first enriched $\gamma\delta$ T cell population, or a portion thereof, with one or more second agents that (a) expand T cells, (b) expand $\gamma\delta$ T-cells (e.g., by binding to a $\gamma\delta$ TCR), or (c) selectively expand:

- $\delta 1$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR;
- $\delta 2$ T-cells by binding to an activating epitope specific of a $\delta 2$ TCR;
- $\delta 1$ T-cells and $\delta 4$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR and a $\delta 4$

TCR; or

- $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells by binding to an activating epitope specific of a $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ TCR, wherein the one or more second agents are structurally different from the one or more first agents, thereby providing a second enriched $\gamma\delta$ T cell population. In some cases, the one or more second agents bind a different $\gamma\delta$ TCR epitope as compared to the one or more first agents.

5 In some cases, the $\gamma\delta$ T-cell population is enriched by positive selection between (i) and (ii), or after (ii). In some cases, the $\gamma\delta$ T-cell population is enriched by depletion of $\alpha\beta$ T cells between (i) and (ii), or after (ii). In some cases, the agent that expands T cells is an agent that binds CD3, such as an anti-CD3 antibody. In some cases, the agent that selectively expands $\gamma\delta$ T cells is an agent that binds a γ -chain constant region or a δ -chain constant region, or an agent that binds $\gamma\delta$ TCR, such as
10 an antibody that binds $\gamma\delta$ TCR (*e.g.*, IMMU510).

In some cases, the first $\gamma\delta$ T-cell expansion is performed in a culture medium that contains an antigen presenting cell (APC). In some cases, the second $\gamma\delta$ T-cell expansion is performed in a culture medium that contains an antigen presenting cell (APC). In some cases, the first $\gamma\delta$ T-cell expansion is performed in a culture medium that does not contain an antigen presenting cell (APC).
15 In some cases, the first $\gamma\delta$ T-cell expansion is performed in a culture medium that does not contain an antigen presenting cell (APC), and the second $\gamma\delta$ T-cell expansion is performed in a culture medium that does contain an antigen presenting cell (APC). In some cases, the first and second $\gamma\delta$ T-cell expansion is performed in a culture medium that does contain an antigen presenting cell (APC). In some cases, the first and second $\gamma\delta$ T-cell expansion is performed in a culture medium
20 that does contain an aminophosphate, an aminophosphonate, a bisphosphonate, or a combination thereof.

In another embodiment, the enriched $\gamma\delta$ T-cell population(s) of the invention may be further formulated for administration to a subject. The enriched $\gamma\delta$ T-cell population(s) of the invention includes a therapeutically effective amount of $\gamma\delta$ T-cells. In certain embodiments, the $\gamma\delta$ T-cell
25 population(s) are engineered to stably express one or more structurally distinct tumor recognition moieties. In certain embodiments, the engineered $\gamma\delta$ T-cell(s) are expanded and/or further expanded as described herein.

In some of the foregoing aspects, embodiments, cases, and examples the one or more agents stimulates an expansion of the $\gamma\delta$ T-cell at a mean rate of 1 cell division in less than 30 hours, *e.g.*,

from greater than about 17 to less than about 30 hours. In some embodiments, said mean rate of division is for 0-4, 0-5, 0-7 contiguous days of $\gamma\delta$ T-cell expansion, 0-13 contiguous days of $\gamma\delta$ T-cell expansion, 0-19 contiguous days of $\gamma\delta$ T-cell expansion, 0-21 contiguous days of $\gamma\delta$ T-cell expansion, or for at least 3, 4, 5, 6, 7, 10, 13, 19, or 21 contiguous days of $\gamma\delta$ T-cell expansion. In

5 other embodiments, the one or more agents stimulates an expansion of the $\gamma\delta$ T-cell at a mean rate of 1 cell division in less than 24 hours, e.g., from greater than about 17 to less than about 24 hours. In other embodiments, the one or more agents stimulates an expansion of the $\gamma\delta$ T-cell at a mean rate of 1 cell division in less than 18 hours or about 18 hours.

It will be appreciated that one or more of the foregoing aspects, embodiments, cases, and
10 examples can be readily adapted for $\gamma\delta$ T-cell expansion (e.g., selective $\gamma\delta$ T-cell expansion) from a substantially homogenous population of cells, such as after establishing a $\gamma\delta$ T-cell engineered clone or cell line, population containing one or more (e.g., structurally different) engineered $\gamma\delta$ T-cells, or after one or more steps of negative or positive selection of a population of cells containing $\gamma\delta$ T-cells. Thus, one or more of the foregoing methods, or a combination thereof, can be used to expand
15 engineered $\gamma\delta$ T-cells by contacting an engineered $\gamma\delta$ T-cell or population thereof with any one or more of the activation agents described herein, including one or more of the antibodies described herein in soluble or immobilized form (e.g., immobilized on the surface of an APC).

As such, in some embodiments, the isolated mixed cell population is substituted in a method or composition provided herein, with a $\gamma\delta$ T-cell engineered clone or cell line, or a population
20 containing one or more (e.g., structurally different) engineered $\gamma\delta$ T-cells.

In other aspects, the invention provides selectively expanded $\gamma\delta$ T-cell population(s), wherein greater than 60%, 70%, 80%, or 90% of the expanded $\gamma\delta$ T-cells are $\delta 1$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$ T-cells, $\delta 3$ T-cells, $\delta 4$ T-cells, and $\delta 5$ T-cells. In certain embodiment, the selectively expanded $\gamma\delta$ T-cell population(s) comprise $\delta 1$ T-cells and $\delta 2$ T-cells, wherein greater than 60%, 70%,
25 80%, or 90% of the expanded $\gamma\delta$ T-cells are $\delta 1$ T-cells $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$ T-cells, $\delta 3$ T-cells, $\delta 4$ T-cells, and $\delta 5$ T-cells. In certain embodiments, the selectively expanded $\gamma\delta$ T-cell population(s) have not been purified by positive selection of $\gamma\delta$ T-cells (e.g., positive selection of $\delta 1$ T cells, $\delta 1$ T-cells and $\delta 3$ T-cells, or $\delta 1$ T-cells and $\delta 4$ T-cells) from an enriched $\gamma\delta$ T-cell population,

or by depletion of non- $\gamma\delta$ T-cells (e.g., depletion of non- $\delta 1$ T cells such as $\alpha\beta$ T cells) from the enriched $\gamma\delta$ T-cell population.

In certain embodiments, the selectively expanded $\gamma\delta$ T-cell population(s) are expanded directly from an isolated mixed cell population comprising T lymphocytes. In certain embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises about 80%, 70%, 60%, 50%, 40%, 30%, 20% T lymphocytes and less than about 30%, 25%, 20%, 15%, 10%, 5%, 4%, 3%, 2%, 1.5%, 1.2%, 1%, 0.9%, 0.8%, 0.7%, 0.6%, 0.5%, 0.4%, 0.3%, 0.2%, or 0.1% $\delta 1$ cells. In certain embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises less than about 30%, 25%, 20%, 15%, 10%, 5%, 4%, 3%, 2%, 1.5%, 1.2%, 1%, 0.9%, 0.8%, 0.7%, 0.6%, 0.5%, 0.4%, 0.3%, 0.2%, or 0.1% $\delta 1$ cells. In some embodiments, the isolated mixed cell population is selected from a peripheral blood sample, a cord blood sample, a tumor, epithelial tissue, or a biopsy of skin, liver, or other tissue. In some embodiments, the isolated mixed cell population is derived from a single donor. In other embodiments, the isolated mixed cell population is derived from more than one or multiple donors. In one embodiment, the expanded $\gamma\delta$ T-cell population(s) are derived from tumor infiltrating lymphocytes, which may be isolated, for example, from cancers of colon adenocarcinoma metastasis, liver, ovary, head and neck, or kidney.

In some embodiments, the selectively expanded $\gamma\delta$ T-cell population(s) of the invention comprise greater than 60% or 70% the $\delta 1$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$ T-cells, $\delta 3$ T-cells, $\delta 4$ T-cells, and $\delta 5$ T-cells which express the naïve or T central memory (TCM) phenotype CD45RA+/CD27+ and/or CD45RA-/CD27+, respectively. In some embodiments, the expanded $\gamma\delta$ T-cell population comprises polyclonal $\gamma\delta$ TCR diversity.

In other aspects, the invention provides selectively expanded $\gamma\delta$ T-cell population(s), wherein greater than 80% or 90% of the expanded $\gamma\delta$ T-cells are $\delta 2$ T-cells. In certain embodiments, the selectively expanded $\gamma\delta$ T-cell population(s) have not been purified by positive selection of $\gamma\delta$ T-cells (e.g., positive selection of $\delta 2$ T cells) from an enriched $\gamma\delta$ T-cell population, or by depletion of non- $\gamma\delta$ T-cells (e.g., depletion of non- $\delta 2$ T cells such as depletion of $\alpha\beta$ T cells) from the enriched $\gamma\delta$ T-cell population. In certain embodiments, the selectively expanded $\gamma\delta$ T-cell population(s) are expanded directly from an isolated mixed cell population comprising T lymphocytes. In certain embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cell population(s) are

expanded comprises about 80%, 70%, 60%, 50%, 40%, 30%, or 20% T lymphocytes and less than about 60%, 50%, 40%, 30%, 25%, 20%, 15%, 10%, 5%, 4%, 3%, 2%, 1.5%, 1.2%, 1%, 0.9%, 0.8%, 0.7%, 0.6%, 0.5%, 0.4%, 0.3%, 0.2%, or 0.1% $\delta 2$ cells. In certain embodiments, the isolated mixed cell population from which the $\gamma\delta$ T-cells are expanded in a method described herein comprises less than about 30%, 25%, 20%, 15%, 10%, 5%, 4%, 3%, 2%, 1.5%, 1.2%, 1%, 0.9%, 0.8%, 0.7%, 0.6%, 0.5%, 0.4%, 0.3%, 0.2%, or 0.1% $\delta 2$ cells. In some embodiments, the isolated mixed cell population is selected from a peripheral blood sample, a cord blood sample or a tumor. In one embodiment, the expanded $\gamma\delta$ T-cell population(s) are derived from tumor infiltrating lymphocytes, which may be isolated, for example, from cancers of colon adenocarcinoma metastasis, liver, ovary, head and neck, or kidney.

In other aspects, the invention provides selectively expanded $\gamma\delta$ T-cell population(s), wherein 10-90% of the expanded $\gamma\delta$ T-cells are $\delta 1$ T-cells and 90-10% of the expanded $\gamma\delta$ T-cells are $\delta 2$ T-cells.

In certain embodiments, said populations are expanded together, *e.g.*, by contacting a mixed cell population with an agent which selectively expands $\delta 1$ cells; $\delta 1$ T-cells and $\delta 3$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$ T-cells, $\delta 3$ T-cells, $\delta 4$ T-cells, and $\delta 5$ T-cells, and with an agent that selectively expands $\delta 2$ cells, either simultaneously or in separate contacting steps. In other embodiments, the populations are admixtures of $\delta 1$ and $\delta 2$ T-cell populations which have been separately and selectively expanded from an isolated mixed cell population. In all embodiments, the expanded $\gamma\delta$ T-cell population(s) or admixtures thereof of the invention may further be formulated for administration to a subject.

In certain embodiments, a $\gamma\delta$ T-cell or $\gamma\delta$ T-cell population(s) of the invention are engineered to stably express one or more structurally distinct tumor recognition moieties encoded by expression cassettes. In one embodiment, the $\gamma\delta$ T-cell is engineered to stably express two or more structurally distinct tumor recognition moieties encoded by expression cassettes. In some embodiments, the two or more structurally distinct tumor recognition moieties recognize different epitopes of the same antigen or different epitopes of different antigens. The tumor recognition moiety may be selected from the group consisting of $\alpha\beta$ TCR, $\gamma\delta$ TCR, a fragment of an $\alpha\beta$ TCR or $\gamma\delta$ TCR, a chimeric antigen receptor (CAR), whole antibody or their antigen-binding fragment, single-chain variable fragment (scFv), a heavy chain or a light chain single domain antibody (sdAb), a Fab, a $F(ab)_2$, or

any combination thereof that binds to: (i) a cell surface tumor antigen or (ii) a peptide derived from a tumor antigen expressed on the cell surface as a complex with MHC (peptide-MHC complex). In certain embodiments, the tumor recognition moiety is a $\gamma\delta$ TCR of a tumor infiltrating lymphocyte that recognizes a tumor-specific antigen in a non-MHC restricted manner.

5 In another aspect, the invention provides expanded $\gamma\delta$ T-cell population(s) obtained according to the methods of the invention. In some embodiments, the $\gamma\delta$ T-cell population is expanded *ex vivo* without antigen stimulation by an antigen presenting cell or an aminophosphonate. In certain embodiments, $\gamma\delta$ T-cell population is activated *ex vivo* with a monoclonal antibody, antibody fragment, or a fusion protein. The $\gamma\delta$ T-cell population(s) of the invention may also be
10 formulated for administration to a subject without co-administration with IL-2. Alternatively, the $\gamma\delta$ T-cell population(s) of the invention may also be formulated for administration to a subject with co-administration with IL-2.

In other aspects, the invention provides methods for treating a cancer in a subject in need thereof, comprising administering a therapeutically effective amount of an expanded $\gamma\delta$ T-cell
15 population according to the invention. In one embodiment, the expanded $\gamma\delta$ T-cell population is allogeneic with respect to the MHC loci of the subject. In certain embodiments, the expanded $\gamma\delta$ T-cell population is engineered to stably express one or more, or two or more, tumor recognition moieties encoded by one or more expression cassettes. In further embodiments, the expanded $\gamma\delta$ T-cell population stably expresses at least two structurally different recognition moieties and each
20 structurally different recognition moiety recognizes (i) different epitopes of the same antigen, or (ii) different epitopes of different antigens. The tumor recognition moiety may be selected from the group consisting of $\alpha\beta$ TCR, $\gamma\delta$ TCR, a chimeric antigen receptor (CAR), including whole antibody or their antigen-binding fragment, single-chain variable fragment (scFv), a heavy chain or a light chain single domain antibody (sdAb), a Fab, a F(ab)₂, or any combination thereof that binds to : (i) a
25 cell surface tumor antigen or (ii) a peptide derived from a tumor antigen expressed on the cell surface as a complex with MHC (peptide-MHC complex).

In another aspect, the invention provides engineered $\gamma\delta$ T-cells, wherein the engineered $\gamma\delta$ T-cell is engineered to express a tumor recognition moiety, wherein the tumor recognition moiety recognizes a tumor antigen, and/or wherein the engineered $\gamma\delta$ T-cell expands *in vitro* in the presence
30 of (*e.g.*, is in contact with *in vitro*) one or more agents which bind the same epitope as TS-1, TS8.2,

B6, or 15D antibodies. In another aspect, the invention provides engineered $\gamma\delta$ T-cells, wherein the engineered $\gamma\delta$ T-cell is engineered to express a tumor recognition moiety, wherein the tumor recognition moiety recognizes a tumor antigen, and/or wherein the engineered $\gamma\delta$ T-cell expands *in vitro* in the presence of (*e.g.*, is in contact with *in vitro*) one or more agents which bind a different epitope than, or an epitope that does not overlap with, the epitope bound by TS-1, TS8.2, B6, or 15D antibodies. In some cases, the engineered $\gamma\delta$ T-cell expands *in vitro* in the presence of (*e.g.*, is in contact with *in vitro*) one or more agents that do not compete with binding of TS-1, TS8.2, B6, or 15D to a $\gamma\delta$ T-cell. In one embodiment, the agent is any one or more of the soluble or immobilized activation agents described herein. In one embodiment, the engineered $\gamma\delta$ T-cell is a $\delta 1$, $\delta 2$, $\delta 3$, or $\delta 4$ engineered T-cell.

In certain embodiments, the engineered $\gamma\delta$ T-cell is further engineered to lack gene expression from at least one HLA loci. In one embodiment, the one or more agents stimulates an expansion of the engineered $\gamma\delta$ T-cell at a mean rate of 1 cell division in less than 30 hours, *e.g.*, from greater than about 17 to less than about 30 hours. In some embodiments, said mean rate of division is for 0-4, 0-5, 0-7 contiguous days of $\gamma\delta$ T-cell expansion, 0-13 contiguous days of $\gamma\delta$ T-cell expansion, 0-19 contiguous days of $\gamma\delta$ T-cell expansion, 0-21 contiguous days of $\gamma\delta$ T-cell expansion, or for at least 3, 4, 5, 6, 7, 10, 13, 19, or 21 contiguous days of $\gamma\delta$ T-cell expansion. In other embodiments, the one or more agents stimulates an expansion of the engineered $\gamma\delta$ T-cell at a mean rate of 1 cell division in less than 24 hours, *e.g.*, from greater than about 17 to less than about 24 hours. In other embodiments, the one or more agents stimulates an expansion of the engineered $\gamma\delta$ T-cell at a mean rate of 1 cell division in less than 18 hours or about 18 hours. In certain embodiments, the tumor recognition moiety is derived from a tumor infiltrating lymphocyte.

In another aspect, the present invention provides an expanded population of $\gamma\delta$ T-cells, wherein the $\gamma\delta$ T-cell population comprises anti-tumor cytotoxicity. In some cases, the $\gamma\delta$ T-cell population comprises anti-tumor cytotoxicity that is independent of NKp30 activity, NKp44 activity, and/or NKp46 activity. In some cases, the $\gamma\delta$ T-cell population comprises anti-tumor cytotoxicity, wherein the anti-tumor cytotoxicity consists of, or consists essentially of, anti-tumor activity that is independent of NKp30 activity, NKp44 activity, and/or NKp46 activity. In some cases, the $\gamma\delta$ T-cell population comprises anti-tumor cytotoxicity, wherein at least 50%, 60%, 75%, 80%, 90%, or 99%

of the anti-tumor cytotoxicity is independent of NKp30 activity, NKp44 activity, and/or NKp46 activity.

In some cases, the $\gamma\delta$ T-cell population is a population of engineered $\gamma\delta$ T-cells. In some cases, the $\gamma\delta$ T-cell population is a population of non-engineered $\gamma\delta$ T-cells. In some cases, the $\gamma\delta$ T-cell population does not comprise NKp30 activity-dependent anti-tumor cytotoxicity, NKp44 activity-dependent anti-tumor cytotoxicity, and/or NKp46 activity-dependent anti-tumor cytotoxicity. In some cases, the $\gamma\delta$ T-cell population does not comprise NKp30 activity-dependent anti-tumor cytotoxicity. In some cases, the $\gamma\delta$ T-cell population does not comprise NKp44 activity-dependent anti-tumor cytotoxicity. In some cases, the $\gamma\delta$ T-cell population is an engineered population of $\gamma\delta$ T-cells that comprise an anti-CD20 chimeric antigen receptor (CAR). In some cases, less than 90%, 80%, 75%, 70%, 60%, 50%, 40%, 30%, 20%, or 10% of the $\gamma\delta$ T-cell cells in the population express a detectable level of NKp30, NKp44, and/or NKp46.

INCORPORATION BY REFERENCE

All publications, patents, and patent applications mentioned in this specification are herein incorporated by reference to the same extent as if each individual publication, patent, or patent application was specifically and individually indicated to be incorporated by reference.

BRIEF DESCRIPTION OF THE DRAWINGS

The novel features of the invention are set forth with particularity in the appended claims. A better understanding of the features and advantages of the present invention will be obtained by reference to the following detailed description that sets forth illustrative embodiments, in which the principles of the invention are utilized, and the accompanying drawings (also “figure” and “FIG.” herein), of which:

FIG. 1 schematically illustrates engineered $\gamma\delta$ T-cells. **Panel A** illustrates an engineered $\gamma\delta$ T-cell expressing one tumor recognition moiety. **Panel B** illustrates an engineered $\gamma\delta$ T-cell expressing two structurally distinct tumor recognition moieties.

FIG. 2 schematically illustrates a method for treating a subject.

FIG. 3 schematically illustrates a method for administering a population of engineered $\gamma\delta$ T-cells to a subject.

FIG. 4 depicts a graph illustrating growth of $\gamma\delta 1$ and $\gamma\delta 2$ lymphocytes isolated from colon adenocarcinoma metastasis to liver (TIL 1) and renal tumors (TIL 2) and have been shown to express CCR4 and CCR7.

FIG. 5 depicts a graph illustrating $\gamma\delta$ T-cells growth in serum-containing and serum-free media.

FIG. 6 depicts a graph illustrating anti- $\gamma\delta$ TCR antibody blocking experiments with 5A6.E9, B1, TS8.2, 15D, B3, B6, TS-1, $\gamma 3.20$, IMMU510, or 11F2.

FIG. 7 depicts a graph illustrating anti- $\gamma\delta$ TCR antibody blocking experiments with 5A6.E9, B1, TS8.2, 15D, B3, B6, TS-1, $\gamma 3.20$, IMMU510, or 11F2.

FIG. 8 depicts a competition experiment with anti-TCR V $\delta 1$ TS-1 antibody.

FIG. 9 depicts a competition experiment with anti-TCR V $\delta 1$ TS8.2 antibody.

FIG. 10 depicts a graph illustrating activation and expansion of $\delta 1$ T-cells from PBMC.

FIG. 11 depicts a graph illustrating activation and expansion of $\delta 2$ T-cells from PBMC.

FIG. 12 depicts a graph illustrating fold expansion of $\delta 1$ T-cells from PBMC.

FIG. 13 depicts a graph illustrating fold expansion of $\delta 2$ T-cells from PBMC.

FIG. 14 depicts the human V $\delta 1$ amino acid sequence. CDR1, CDR3, and CDR3 regions are underlined.

FIG. 15 depicts the human V $\delta 2$ amino acid sequence. CDR1, CDR3, and CDR3 regions are underlined.

FIG. 16 depicts activation of PBMCs with TS-1 or TS8.2 antibodies resulting in significant and specific expansion of V $\delta 1^+$ T cells. **(A)** MAbs were directly coated (1 $\mu\text{g/mL}$) (TS1, TS8.2, B6, and 15D) or captured (0.1 $\mu\text{g/mL}$) with goat-anti-mouse Fc (5 $\mu\text{g/mL}$) (TS1Fc, TS8.2Fc, B6Fc and 15DFc) in 24-well plates. PBMCs were plated at 10^6 cells/mL in RPMI with 10% FBS and 100 IU/mL IL-2. On day 7, cells were transferred to new plates without antibodies and further expanded until day 14. Culture media was replenished every 2-3 days. Data depicts expansion of V $\delta 1^+$ cells over 14 days; **(B)** Same culture as A, showing percentage of V $\delta 1^+$ cells on day 14 and day 0 (d0); **(C)** V $\delta 1^+$ cells were expanded from isolated PBMCs from a different donor as follows. MAbs were directly coated (1 $\mu\text{g/mL}$) (TS1, TS8.2, B6 and 15D) or captured (0.1 $\mu\text{g/mL}$) with goat-anti-mouse Fc (5 $\mu\text{g/mL}$) (TS1Fc, TS8.2Fc, B6Fc and 15DFc) in 24-well plates. PBMCs f were plated at 10^6 cells/mL in RPMI with 10% FBS and 100 IU/mL IL-2. Cells were transferred to new plate without

antibodies on day 7, adjusted to 10^6 cells/mL with fresh media. Culture media was replenished every 2-3 days and adjusted to 10^6 cells/mL. Data depicts V δ 1+ cells expansion over 14 days. **(D)** PBMCs were plated at 10^6 cells/mL in RPMI with 10% FBS and 100 IU/mL IL-2. On day 7, cells were transferred to a new plate without antibodies, adjusted to 10^6 cells/mL and further expanded until day 23. Culture media was replenished every 2-3 days.

FIG. 17 depicts activation of PBMCs with B6 and 15D antibodies result in significant specific activation and expansion of V δ 2+ T cells. **(A)** MAbs were directly coated (1 μ g/mL) (TS1, TS8.2, B6, and 15D) or captured (0.1 μ g/mL) with goat-anti-mouse Fc (5 μ g/mL) (TS1Fc, TS8.2Fc, B6Fc and 15DFc) in 24-well plates. PBMCs were plated at 10^6 cells/mL in RPMI with 10% FBS and 100 IU/mL IL-2. On day 7, cells were transferred to new plates without antibodies and further expanded until day 14. Culture media was replenished every 2-3 days. Data depicts V δ 2+ T-cells expansion over 14 days; **(B)** Same culture as A, showing percentage of V δ 2+ cells on day 14 and day 0 (d0); **(C)** MAbs were directly coated (1 μ g/mL) (15D and pan- $\gamma\delta$ TCR Mab Immu510) in 24-well plates. PBMCs from a different donor were plated at 10^6 cells/mL in RPMI with 10% FBS and 100 IU/mL IL-2. Cells were transferred to new plates without antibodies on day 7. Culture was replenished every 2-3 days and adjusted to 10^6 cells/mL with fresh media. Data depicts V δ 2+ T-cells expansion over 14 days; **(D)** Same culture as C, showing percentage of V δ 2+ T-cells on day 14 and day 0 (d0). **(E)** MAbs were directly coated (1 μ g/mL) (TS1, TS8.2, B6, and 15D) or captured (0.1 μ g/mL) with goat-anti-mouse Fc (5 μ g/mL) (TS1Fc, TS8.2Fc, B6Fc and 15DFc) in 24-well plates. PBMCs from another donor were plated at 10^6 cells/mL in RPMI with 10% FBS and 100 IU/mL IL-2. Cells were transferred to new plates without antibodies on day 7, adjusted to 10^6 cells/mL with fresh media. Culture media was replenished every 2-3 days and adjusted to 10^6 cells/mL. Data depicts V δ 2+ T-cells expansion over 14 days.

FIG. 18 depicts significant reduction in percentage of $\alpha\beta$ T cells in PBMC cultured with $\gamma\delta$ -specific antibodies TS1, TS8.2, B6, and 15D. **(A)** MAbs were directly coated (1 μ g/mL) (TS1, TS8.2 B6 and 15D) or captured (0.1 μ g/mL) with goat-anti-mouse Fc (5 μ g/mL) (TS1Fc, TS8.2Fc, B6Fc and 15DFc) in 24-well plates. PBMCs were plated at 10^6 cells/mL in RPMI with 10% FBS and 100 IU/mL IL-2. On day 7, cells were transferred to new plates without antibodies and further expanded until day 14. Culture media was replenished every 2-3 days. Data depicts $\alpha\beta$ T-cells expansion over 14 days; **(B)** Same culture as A, showing percentage of $\alpha\beta$ T-cells on day 14 and day

0 (d0).

FIG. 19 depicts activation of $\gamma\delta$ TCR results in reduction of $\gamma\delta$ T-cell doubling time

MAbs were directly coated (1 $\mu\text{g/mL}$) (TS1, TS8.2, B6 and 15D) or captured (0.1 $\mu\text{g/mL}$) with goat-anti-mouse Fc (5 $\mu\text{g/mL}$) (TS1Fc, TS8.2Fc, B6Fc and 15DFc) in 24-well plates. PBMCs were plated at 10^6 cells/mL in RPMI with 10% FBS and 100 IU/mL IL-2. Cells were transferred to new plate without antibodies on day 7, adjusted to 10^6 cells/mL with fresh media. Culture was replenished every 2-3 days and adjusted to 10^6 cells/mL with fresh media. Doubling times for V δ 1, V δ 2 and $\alpha\beta$ T-cells are shown in (A), (B) and (C), respectively.

FIG. 20 depicts activation of V δ 1 by antibodies TS8.2 and TS1 results in predominantly naïve and central memory phenotypes. Antibodies were directly coated at 1 $\mu\text{g/mL}$ in 24 well plates (TS8.2, R9.12 and pan- $\gamma\delta$ Immu510). PBMCs were plated at 10^6 /mL in RPMI with 10% FBS and 100 IU/mL IL-2. Media was replenished every 2-3 days until day 23, except on day15 when cultures were diluted 1:2 in fresh media. Cells phenotypes on days 14 and 23 are determined by CD45RA and CD27 expression using flow cytometry analysis.

FIG. 21 depicts activation of PBMC by TS8.2 and MICA results in enhanced expansion of V δ 1 T-cells but not $\alpha\beta$ T-cells. Antibody TS8.2 (1 $\mu\text{g/mL}$) or TS8.2 with MICA-Fc (1 and 5 $\mu\text{g/mL}$) were directly coated in 24 well plates. PBMCs were plated at 10^6 /mL in RPMI with 10% FBS and 100 IU/mL IL-2. Media is replenished every 2-3 days. (A) V δ 1 T-cells expansion; (B) $\alpha\beta$ T-cells expansion.

FIG. 22 depicts activation of umbilical cord blood mononuclear cells with δ 1 and δ 2 specific antibodies result in significant and specific expansion of V δ 1⁺ and V δ 2⁺ T-cells. MAbs TS8.2, R9.12, B6 and 15D were directly coated at 1 $\mu\text{g/mL}$ in 24 well plates. Umbilical cord blood mononuclear cells were activated at 10^6 /mL in RPMI with 10% FBS and 100 IU/mL IL-2. V δ 1 and V δ 2 T-cells expansion are shown in (A) and (B), respectively.

FIG. 23 depicts binding to soluble TCR generated from pairing of V δ 1J1, V δ 1J2 and V δ 1J3 chain with γ 8 chain. Data show that TS1 and TS8.2 recognize the soluble TCR generated from V δ 1J1 and V δ 1J2 chains but failed to bind to the V δ 1J3 chain, indicating that the J1 and J2 gene segments are critical for TS1 and TS8.2 binding. R9.12 that bind to V δ 1, and the pan antibody Immuno510, which binds to δ constant region, are not affected by the specific J region.

FIG. 24 depicts sequence alignment of human V δ 1J1, J2 and J3 regions and mutations made

to the V δ 1J1 chain at the selected positions. BE-13 refers to δ 1J region from a T cell leukemia cell line expressing δ 1 γ 8 TCR (DSMZ accession #ACC 396). Replacement of a single amino acid residue at position Lys120 by Thr or Ala in the V δ 1J1 and V δ 1J2 region completely abolished binding of TS-1 and TS8.2 MAbs indicating that this amino acid in the V δ 1J1 and V δ 1J2 region is
 5 contributes to the binding of TS-1 and TS8.2. Replacement of a single amino acid residue at position Thr120 by Lys in the V δ 1J3 region resulted in a gain of binding of TS-1 and TS8.2 MAbs further indicating that this amino acid in the V δ 1J1 and V δ 1J2 region contributes to the binding of TS-1 and TS8.2.

FIG. 25 depicts that point mutation in V δ 1J1 from Lys120 to Thr or Ala resulted in the loss
 10 of binding by TS-1 and TS8.2 MAbs.

FIG. 26 depicts of human V δ 1 protein sequence and the six mutated human V δ 1 sequences based on the differences between human V δ 1 and the bovine V δ 1 amino acid sequence (GenBank:AFP25162.1). Mutation are shown in bold.

FIG. 27 depicts loss of binding of TS-1 and TS8.2 antibodies to the human V δ 1 mutated
 15 chains.

FIG. 28 depicts binding of the B6 MAb to different V δ 2/ γ chain pairings (γ 3, γ 8, γ 9).

FIG. 29 depicts the protein sequence alignment of human (IMGT Human TRDV2) and Rhesus monkey (GenBank: AY190028.1) V δ 2 variable regions and the changes made in V δ 2 CDR1 (G35S), CDR2 (D65G) and CDR3 (C104S).

FIG. 30 depicts loss of binding of 15D to V δ 2 (G35S) mutated at CDR1.

FIG. 31 depicts characterization of highly enriched V δ 1+, V δ 2+ and $\alpha\beta$ T cell cultures using TS8.2, B6 and IP26 antibodies for δ 1, δ 2 and $\alpha\beta$, respectively. The populations derived from PBMC were expanded with a combination of TS-1 and TS8.2 to expand δ 1 cells, or 15D and B6 to expand δ 2 cells. The expanded culture was depleted of $\alpha\beta$ T cells using IP26 MicroBeads. Positively
 25 selected $\alpha\beta$ T cells were also collected ($\alpha\beta$ enriched).

FIG. 32 depicts cytotoxicity of V δ 1+, V δ 2+ populations toward solid tumor (BxPC3, SKMEL5) and plasmacytoma (RPMI8226) cell lines.

FIG. 33 depicts heavy-chain framework and complementarity determining region amino acid sequences of δ 1-specific MAbs.

FIG. 34 depicts light-chain framework and complementarity determining region amino acid

sequences of the δ 1-specific MAbs described in **FIG. 33**.

FIG. 35 depicts heavy-chain framework and complementarity determining region amino acid sequences of δ 2-specific MAbs.

FIG. 36 depicts light-chain framework and complementarity determining region amino acid sequences of δ 2-specific MAbs.

FIG. 37 depicts cross-reactivity of the indicated δ 1-specific antibodies with other $\gamma\delta$ TCRs as determined by ELISA.

FIG. 38 depicts an alignment of V δ 1 deduced amino acid sequence between human and dolphin.

FIG. 39 depicts the percentage of proliferating V δ 1 cells (A) or V δ 2 cells (B) obtained by a $\gamma\delta$ T cell expansion method described herein.

FIG. 40 depicts the fold expansion of V δ 1 cells (A) or V δ 2 cells (B) from an isolated mixed cell population obtained by a $\gamma\delta$ T cell expansion method described herein.

FIG. 41 depicts phenotype data from an isolated mixed cell population obtained by a $\gamma\delta$ T cell expansion method described herein. Cell phenotypes are determined by CD45RA and CD27 expression using flow cytometry analysis.

FIG. 42 illustrates δ 1 T cells activated and expanded according to $\gamma\delta$ T cell expansion methods described herein can show robust cytotoxicity against tumor cell lines.

FIG. 43 illustrates V δ 1 fold expansion and purity on day 19 achieved by a method of the present invention.

FIG. 44 illustrates V δ 1 fold expansion and accumulative doubling time on day 19 achieved by a method of the present invention.

FIG. 45 illustrates V δ 1 fold expansion and accumulative doubling time from day 14 to day 19 achieved by a method of the present invention.

FIG. 46 illustrates expression of an anti-CD20 chimeric antigen receptor (CAR) on δ 1 T cells. Expanded cells were stained with V δ 1 specific antibody (R9.12) and anti-Rituximab FITC conjugate on Day 7 post-transduction with CAR retroviral construct. Up to 35% of V δ 1 cells were CD20 CAR+.

FIG. 47A-B depicts epitope binding specificity data for δ -1 specific $\gamma\delta$ T cell activators.

FIG. 48 depicts epitope binding specificity data for δ -2 specific $\gamma\delta$ T cell activators.

DETAILED DESCRIPTION OF THE INVENTION

While various embodiments of the invention have been shown and described herein, it will be obvious to those skilled in the art that such embodiments are provided by way of example only.

5 Numerous variations, changes, and substitutions may occur to those skilled in the art without departing from the invention. It should be understood that various alternatives to the embodiments of the invention described herein may be employed.

DEFINITIONS

10 Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which the inventions described herein belong. For purposes of interpreting this specification, the following definitions will apply and whenever appropriate, terms used in the singular will also include the plural and vice versa. In the event that any definition set forth conflicts with any document incorporated herein by reference, the definition set forth below shall control.

15 The term “ $\gamma\delta$ T-cells (gamma delta T-cells)” as used herein refers to a subset of T-cells that express a distinct T-cell receptor (TCR), $\gamma\delta$ TCR, on their surface, composed of one γ -chain and one δ -chain. The term “ $\gamma\delta$ T-cells” specifically includes all subsets of $\gamma\delta$ T-cells, including, without limitation, V δ 1 and V δ 2, V δ 3 $\gamma\delta$ T cells, as well as naïve, effector memory, central memory, and terminally differentiated $\gamma\delta$ T-cells. As a further example, the term “ $\gamma\delta$ T-cells” includes V δ 4, V δ 5,
20 V δ 7, and V δ 8 $\gamma\delta$ T cells, as well as V γ 2, V γ 3, V γ 5, V γ 8, V γ 9, V γ 10, and V γ 11 $\gamma\delta$ T cells.

As used herein, the term “T lymphocyte” or “T cell” refers to an immune cell expressing CD3 (CD3+) and a T Cell Receptor (TCR+). T cells play a central role in cell-mediated immunity.

As used herein, the term “TCR” or “T cell receptor” refers to a dimeric heterologous cell surface signaling protein forming an alpha-beta or gamma-delta receptor. $\alpha\beta$ TCR recognize an
25 antigen presented by an MHC molecule, whereas $\gamma\delta$ TCR recognize an antigen independently of MHC presentation.

The term “MHC” (major histocompatibility complex) refers to a subset of genes that encodes cell-surface antigen-presenting proteins. In humans, these genes are referred to as human leukocyte antigen (HLA) genes. Herein, the abbreviations MHC or HLA are used interchangeably.

30 As used herein, the term “peripheral blood lymphocyte(s)” or “PBL(s)” is used in the

broadest sense and refers to white blood cell(s) comprising T cells and B cells of a range of differentiation and functional stages, plasma cells, monocytes, macrophages, natural killer cells, basocytes, eosinophils, etc. The range of T lymphocytes in peripheral blood is about 20-80%.

As used herein, the term "cell population" refers to a number of cells obtained by isolation
5 directly from a suitable source, usually from a mammal. The isolated cell population may be subsequently cultured *in vitro*. Those of ordinary skill in the art will appreciate that various methods for isolating and culturing cell populations for use with the present invention and various numbers of cells in a cell population that are suitable for use in the present invention. A cell population may be, for example, a mixed heterogeneous cell population derived from a peripheral blood sample, a cord
10 blood sample, a tumor, a stem cell precursor, a tumor biopsy, a tissue, a lymph, or from epithelial sites of a subject directly contacting the external milieu, or derived from stem precursor cells. Alternatively, the mixed cell population may be derived from *in vitro* cultures of mammalian cells, established from a peripheral blood sample, a cord blood sample, a tumor, a stem cell precursor, a tumor biopsy, a tissue, a lymph, or from epithelial sites of a subject directly contacting the external
15 milieu, or derived from stem precursor cells.

An "enriched" cell population or preparation refers to a cell population derived from a starting mixed cell population that contains a greater percentage of a specific cell type than the percentage of that cell type in the starting population. For example, a starting mixed cell population can be enriched for a specific $\gamma\delta$ T-cell population. In one embodiment, the enriched $\gamma\delta$ T-cell
20 population contains a greater percentage of $\delta 1$ cells than the percentage of that cell type in the starting population. As another example, an enriched $\gamma\delta$ T-cell population can contain a greater percentage of both $\delta 1$ cells and a greater percentage of $\delta 3$ cells than the percentage of that cell type in the starting population. As yet another example, an enriched $\gamma\delta$ T-cell population can contain a greater percentage of both $\delta 1$ cells and a greater percentage of $\delta 4$ cells than the percentage of that
25 cell type in the starting population. As yet another example, an enriched $\gamma\delta$ T-cell population can contain a greater percentage of $\delta 1$ T cells, $\delta 3$ T cells, $\delta 4$ T cells, and $\delta 5$ T cells than the percentage of that cell type in the starting population. In another embodiment, the enriched $\gamma\delta$ T-cell population contains a greater percentage of $\delta 2$ cells than the percentage of that cell type in the starting population. In yet another embodiment, the enriched $\gamma\delta$ T-cell population contains a greater
30 percentage of both $\delta 1$ cells and $\delta 2$ cells than the percentage of that cell type in the starting

population. In all embodiments, the enriched $\gamma\delta$ T-cell population contains a lesser percentage of $\alpha\beta$ T-cell populations.

By "expanded" as used herein is meant that the number of the desired or target cell type (*e.g.*, $\delta 1$ and/or $\delta 2$ T-cells) in the enriched preparation is higher than the number in the initial or starting cell population. By "selectively expand" is meant that the target cell type (*e.g.*, $\delta 1$ or $\delta 2$ T-cells) are preferentially expanded over other non-target cell types, *e.g.*, $\alpha\beta$ T-cells or NK cells. In certain embodiments, the activating agents of the invention selectively expand, *e.g.*, engineered or non-engineered, $\delta 1$ T-cells without significant expansion of $\delta 2$ T-cells. In other embodiments, the activating agents of the invention selectively expand, *e.g.*, engineered or non-engineered, $\delta 2$ T-cells without significant expansion of $\delta 1$ T-cells. In certain embodiments, the activating agents of the invention selectively expand, *e.g.*, engineered or non-engineered, $\delta 1$ and $\delta 3$ T-cells without significant expansion of $\delta 2$ T-cells. In certain embodiments, the activating agents of the invention selectively expand, *e.g.*, engineered or non-engineered, $\delta 1$ and $\delta 4$ T-cells without significant expansion of $\delta 2$ T-cells. In certain embodiments, the activating agents of the invention selectively expand, *e.g.*, engineered or non-engineered, $\delta 1$, $\delta 3$, $\delta 4$ and $\delta 5$ T-cells without significant expansion of $\delta 2$ T-cells. In this context, the term "without significant expansion of" means that the preferentially expanded cell population are expanded at least 10-fold, preferably 100-fold, and more preferably 1,000-fold more than the reference cell population.

The term "admixture" as used herein refers to a combination of two or more isolated, enriched cell populations derived from a mixed, heterogeneous cell population. According to certain embodiments, the cell populations of the present invention are isolated $\gamma\delta$ T cell populations.

The term "isolated," as applied to a cell population, refers to a cell population, isolated from the human or animal body, which is substantially free of one or more cell populations that are associated with said cell population *in vivo* or *in vitro*.

The term "contacting" in the context of a cell population, as used here refers to incubation of an isolated cell population with a reagent, such as, for example, an antibody, cytokine, ligand, mitogen, or co-stimulatory molecule that can be linked either to beads or to cells. The antibody or cytokine can be in a soluble form, or it can be immobilized. In one embodiment, the immobilized antibody or cytokine is tightly bound or covalently linked to a bead or plate. In one embodiment, the antibody is immobilized on Fc-coated wells. In desirable embodiments, the contact occurs *ex vivo*

(e.g., in vitro), or in vivo.

As used herein, the term "antibody" refers to immunoglobulin molecules and immunologically active portions of immunoglobulin (Ig) molecules, *i.e.*, molecules that contain an antigen binding site that specifically binds (immunoreacts with) an antigen. By "specifically bind" or "immunoreacts with" or "directed against" is meant that the antibody reacts with one or more antigenic determinants of the desired antigen and does not react with other polypeptides or binds at much lower affinity ($K_D > 10^{-6}$ molar). Antibodies include, but are not limited to, polyclonal, monoclonal, chimeric, sdAb (heavy or light single domain antibody), single chain, F_{ab} , F_{ab}' and $F_{(ab)2}$ fragments, scFvs, diabodies, minibodies, nanobodies, and F_{ab} expression library.

The term "chimeric antigen receptors (CARs)," as used herein, may refer to artificial T-cell receptors, T-bodies, single-chain immunoreceptors, chimeric T-cell receptors, or chimeric immunoreceptors, for example, and encompass engineered receptors that graft an artificial specificity onto a particular immune effector cell. CARs may be employed to impart the specificity of a monoclonal antibody onto a T cell, thereby allowing a large number of specific T cells to be generated, for example, for use in adoptive cell therapy. In specific embodiments, CARs direct specificity of the cell to a tumor associated antigen, for example. In some embodiments, CARs comprise an intracellular activation domain (allowing the T cell to activate upon engagement of targeting moiety with target cell, such as a target tumor cell), a transmembrane domain, and an extracellular domain that may vary in length and comprises a disease- or disorder-associated, *e.g.*, a tumor-antigen binding region. In particular aspects, CARs comprise fusions of single-chain variable fragments (scFv) derived from monoclonal antibodies, fused to CD3-zeta a transmembrane domain and endodomain. The specificity of other CAR designs may be derived from ligands of receptors (e.g., peptides) or from pattern-recognition receptors, such as Dectins. In certain cases, the spacing of the antigen-recognition domain can be modified to reduce activation-induced cell death. In certain cases, CARs comprise domains for additional co-stimulatory signaling, such as CD3-zeta, FcR, CD27, CD28, CD137, DAP 10/12, and/or OX40, ICOS, TLRs, etc. In some cases, molecules can be co-expressed with the CAR, including co-stimulatory molecules, reporter genes for imaging (e.g., for positron emission tomography), gene products that conditionally ablate the T cells upon addition of a pro-drug, homing receptors, chemokines, chemokine receptors, cytokines, and cytokine receptors.

The basic antibody structural unit is known to comprise a tetramer. Each tetramer is composed of two identical pairs of polypeptide chains, each pair having one "light" (about 25 kDa) and one "heavy" chain (about 50-70 kDa). The amino-terminal portion of each chain includes a variable region of about 100 to 110 or more amino acids primarily responsible for antigen
5 recognition. The carboxy-terminal portion of each chain defines a constant region primarily responsible for effector function. In general, antibody molecules obtained from humans relate to any of the classes IgG, IgM, IgA, IgE and IgD, which differ from one another by the nature of the heavy chain present in the molecule. Certain classes have subclasses as well, such as IgG₁, IgG₂, and others. Furthermore, in humans, the light chain may be a kappa chain or a lambda chain.

10 The term "Fab" refers to an antibody fragment that consists of an entire L chain (V_L and C_L) along with the variable region domain of the H chain (V_H), and the first constant domain of one heavy chain (CH1). Papain digestion of an intact antibody can be used to produce two Fab fragments, each of which contains a single antigen-binding site. Typically, the L chain and H chain fragment of the Fab produced by papain digestion are linked by an interchain disulfide bond.

15 The term "Fc" refers to an antibody fragment that comprises the carboxy-terminal portions of both H chains (CH2 and CH3) and a portion of the hinge region held together by disulfide bonds. The effector functions of antibodies are determined by sequences in the Fc region; this region is also the part recognized by Fc receptors (FcR) found on certain types of cells. One Fc fragment can be obtained by papain digestion of an intact antibody.

20 The term "F(ab')₂" refers to an antibody fragment produced by pepsin digestion of an intact antibody. F(ab')₂ fragments contain two Fab fragments and a portion of the hinge region held together by disulfide bonds. F(ab')₂ fragments have divalent antigen-binding activity and are capable of cross-linking antigen.

25 The term Fab' refers to an antibody fragment that is the product of reduction of an F(ab')₂ fragment. Fab' fragments differ from Fab fragments by having a few additional residues at the carboxy terminus of the CH1 domain including one or more cysteines from the antibody hinge region. Fab'-SH is the designation herein for Fab' in which the cysteine residue(s) of the constant domains bear a free thiol group.

30 The term "Fv" refers to an antibody fragment that consists of a dimer of one heavy-chain variable region and one light-chain variable region domain in tight, non-covalent association. From

the folding of these two domains emanate six hypervariable loops (3 loops each from the H and L chain) that contribute the amino acid residues for antigen binding and confer antigen binding specificity to the antibody. However, even a single variable domain (or half of an Fv comprising only three CDRs specific for an antigen) has the ability to recognize and bind antigen, although often
5 at a lower affinity than the entire binding site.

The term “single-chain Fv” also abbreviated as “sFv” or “scFv” refer to antibody fragments that comprise the VH and VL antibody domains connected into a single polypeptide chain. Typically, the scFv polypeptide further comprises a polypeptide linker between the VH and VL domains, which enables the scFv to form the desired structure for antigen binding. For a review of scFv, see, *e.g.*,
10 Pluckthun, The Pharmacology of Monoclonal Antibodies, vol. 113, Rosenberg and Moore eds., Springer-Verlag, New York, pp. 269-315 (1994); and Malmberg *et al.*, J. Immunol. Methods 183:7-13, 1995.

The expression “linear antibody” is used to refer to a polypeptide comprising a pair of tandem V_H - C_H1 segments (V_H - C_H1 - V_H - C_H1) which form a pair of antigen binding regions. Linear
15 antibodies can be bispecific or monospecific and are described, for example, by Zapata *et al.*, *Protein Eng.* 8(10):1057-1062 (1995).

The term “variable” refers to the fact that certain portions of the variable domains differ extensively in sequence among antibodies and are used in the binding and specificity of each particular antibody for its particular antigen. However, the variability is not evenly distributed
20 throughout the variable domains of antibodies. It is concentrated in three segments called hypervariable regions both in the light chain and the heavy chain variable domains. The more highly conserved portions of variable domains are called the framework regions (FRs). The variable domains of native heavy and light chains each comprise four FRs, largely adopting a beta-sheet configuration, connected by three hypervariable regions, which form loops connecting, and in some
25 cases forming part of, the beta-sheet structure. The hypervariable regions in each chain are held together in close proximity by the FRs and, with the hypervariable regions from the other chain, contribute to the formation of the antigen-binding site of antibodies (see Kabat et al (1991) Sequences of Proteins of Immunological Interest, 5th Ed. Public Health Service, National Institutes of Health, Bethesda, Md.). The constant domains are not involved directly in binding an antibody to
30 an antigen, but exhibit various effector functions, such as participation of the antibody in antibody

dependent cellular cytotoxicity (ADCC).

The term "antigen-binding site" or "binding portion" refers to the part of the immunoglobulin molecule that participates in antigen binding. The antigen binding site is formed by amino acid residues of the N-terminal variable ("V") regions of the heavy ("H") and light ("L") chains. Three highly divergent stretches within the V regions of the heavy and light chains, referred to as "hypervariable regions," are interposed between more conserved flanking stretches known as "framework regions," or "FRs". Thus, the term "FR" refers to amino acid sequences which are naturally found between, and adjacent to, hypervariable regions in immunoglobulins. In an antibody molecule, the three hypervariable regions of a light chain and the three hypervariable regions of a heavy chain are disposed relative to each other in three dimensional space to form an antigen-binding surface. The antigen-binding surface is complementary to the three-dimensional surface of a bound antigen, and the three hypervariable regions of each of the heavy and light chains are referred to as "complementarity-determining regions," or "CDRs." The assignment of amino acids to each domain is in accordance with the definitions of Kabat Sequences of Proteins of Immunological Interest (National Institutes of Health, Bethesda, Md. (1987 and 1991)), or Chothia & Lesk J. Mol. Biol. 196:901-917 (1987), Chothia *et al.* Nature 342:878-883 (1989).

The term "hypervariable region," "HVR," or "HV," refers to the regions of an antibody variable domain which are hypervariable in sequence and/or form structurally defined loops. Generally, antibodies comprise six HVRs; three in the VH (H1, H2, H3), and three in the VL (L1, L2, L3). In native antibodies, H3 and L3 display the most diversity of the six HVRs, and H3 in particular is believed to play a unique role in conferring fine specificity to antibodies. See, *e.g.*, Xu *et al.*, *Immunity* 13:37-45 (2000); Johnson and Wu, in *Methods in Molecular Biology* 248:1-25 (Lo, ed., Human Press, Totowa, N.J., 2003). Indeed, naturally occurring camelid antibodies consisting of a heavy chain only are functional and stable in the absence of light chain. See, *e.g.*, Hamers-Casterman *et al.*, *Nature* 363:446-448 (1993); Sheriff *et al.*, *Nature Struct. Biol.* 3:733-736 (1996).

"Framework regions" (FR) are those variable domain residues other than the CDR residues. Each variable domain typically has four FRs identified as FR1, FR2, FR3, and FR4. If the CDRs are defined according to Kabat, the light chain FR residues are positioned at about residues 1-23 (LCFR1), 35-49 (LCFR2), 57-88 (LCFR3), and 98-107 (LCFR4) and the heavy chain FR residues are positioned about at residues 1-30 (HCFR1), 36-49 (HCFR2), 66-94 (HCFR3), and 103-113

(HCFR4) in the heavy chain residues. If the CDRs comprise amino acid residues from hypervariable loops, the light chain FR residues are positioned about at residues 1-25 (LCFR1), 33-49 (LCFR2), 53-90 (LCFR3), and 97-107 (LCFR4) in the light chain and the heavy chain FR residues are positioned about at residues 1-25 (HCFR1), 33-52 (HCFR2), 56-95 (HCFR3), and 102-113 (HCFR4) in the heavy chain residues. In some instances, when the CDR comprises amino acids from both a CDR as defined by Kabat and those of a hypervariable loop, the FR residues will be adjusted accordingly. For example, when CDRH1 includes amino acids H26-H35, the heavy chain FR1 residues are at positions 1-25 and the FR2 residues are at positions 36-49.

A "human consensus framework" is a framework that represents the most commonly occurring amino acid residues in a selection of human immunoglobulin VL or VH framework sequences. Generally, the selection of human immunoglobulin VL or VH sequences is from a subgroup of variable domain sequences. Generally, the subgroup of sequences is a subgroup as in Kabat. In certain instances, for the VL, the subgroup is subgroup kappa I as in Kabat. In certain instances, for the VH, the subgroup is subgroup III as in Kabat.

As used herein, the "Kd" or "Kd value" refers to a dissociation constant measured by using surface plasmon resonance assays, for example, using a BIAcore.TM.-2000 or a BIAcore.TM.-3000 (BIAcore, Inc., Piscataway, N.J.) at 25 °C. with CM5 chips immobilized with antigen or antibody at about 10 response units (RU). For divalent or other multivalent antibodies, typically the antibody is immobilized to avoid avidity-induced interference with measurement of the dissociation constant. For further details see, *e.g.*, Chen *et al.*, *J. Mol. Biol.* 293:865-881 (1999).

The term "epitope" includes any protein determinant, lipid or carbohydrate determinant capable of specific binding to an immunoglobulin or T-cell receptor. Epitopic determinants usually consist of active surface groupings of molecules such as amino acids, lipids or sugar side chains and usually have specific three dimensional structural characteristics, as well as specific charge characteristics. An antibody is said to specifically bind an antigen when the equilibrium dissociation constant (K_D) is range $10^{-6} - 10^{-12}$ M. An "activating epitope" is capable of activation of the specific $\gamma\delta$ T-cell population upon binding.

An antibody binds "essentially the same epitope" as a reference antibody, when the two antibodies recognize identical or sterically overlapping epitopes. The most widely used and rapid methods for determining whether two epitopes bind to identical or sterically overlapping epitopes

are competition assays, which can be configured in a number of different formats, using either labeled antigen or labeled antibody. In some embodiments, the antigen is immobilized on a 96-well plate, and the ability of unlabeled antibodies to block the binding of labeled antibodies is measured using radioactive or enzyme labels. Alternatively, the competition studies, using labelled and
5 unlabeled antibodies, are performed using flow cytometry on antigen-expressing cells.

“Epitope mapping” is the process of identifying the binding sites, or epitopes, of antibodies on their target antigens. Antibody epitopes may be linear epitopes or conformational epitopes. Linear epitopes are formed by a continuous sequence of amino acids in a protein. Conformational epitopes are formed of amino acids that are discontinuous in the protein sequence, but which are
10 brought together upon folding of the protein into its three-dimensional structure.

“Epitope binning”, as defined herein, is the process of grouping antibodies based on the epitopes they recognize. More particularly, epitope binning comprises methods and systems for discriminating the epitope recognition properties of different antibodies, combined with computational processes for clustering antibodies based on their epitope recognition properties and
15 identifying antibodies having distinct binding specificities.

An “agent” or “compound” according to the present invention comprises small molecules, polypeptides, proteins, antibodies or antibody fragments. Small molecules, in the context of the present invention, mean in one embodiment chemicals with molecular weight smaller than 1000 Daltons, particularly smaller than 800 Daltons, more particularly smaller than 500 Daltons. The term
20 “therapeutic agent” refers to an agent that has biological activity. The term “anti-cancer agent” refers to an agent that has biological activity against cancer cells.

As used herein, the term “cell culture” refers to any *in vitro* culture of cells. Included within this term are continuous cell lines (e.g., with an immortal phenotype), primary cell cultures, finite cell lines (e.g., non-transformed cells), and any other cell population maintained in vitro, including
25 stem cells, blood cells, embryonic cord blood cells, tumor cells, transduced cells, etc.

The terms “treat” or “treatment” refer to both therapeutic treatment and prophylactic or preventative measures, wherein the object is to prevent or slow down (lessen) an undesired physiological change or disorder. Beneficial or desired clinical results include, but are not limited to, alleviation of symptoms, diminishment of extent of disease (e.g., decrease of tumor size, tumor
30 burden, or tumor distribution), stabilized (i.e., not worsening) state of disease, delay or slowing of

disease progression, amelioration or palliation of the disease state, and remission (whether partial or total), whether detectable or undetectable. "Treatment" can also mean prolonging survival, as compared to expected survival if not receiving treatment. Those in need of treatment include those already with the condition or disorder as well as those prone to have the condition or disorder or
5 those in which the condition or disorder is to be prevented.

Administration "in combination with" one or more further therapeutic agents includes simultaneous (concurrent) and consecutive administration in any order.

The term "identical," as used herein, refers to two or more sequences or subsequences that are the same. In addition, the term "substantially identical," as used herein, refers to two or more
10 sequences which have a percentage of sequential units which are the same when compared and aligned for maximum correspondence over a comparison window, or designated region as measured using comparison algorithms or by manual alignment and visual inspection. By way of example only, two or more sequences may be "substantially identical" if the sequential units are about 60% identical, about 65% identical, about 70% identical, about 75% identical, about 80% identical, about
15 85% identical, about 90% identical, or about 95% identical over a specified region. Such percentages to describe the "percent identity" of two or more sequences. The identity of a sequence can exist over a region that is at least about 75-100 sequential units in length, over a region that is about 50 sequential units in length, or, where not specified, across the entire sequence. This definition also refers to the complement of a test sequence. In addition, by way of example only, two or more
20 polynucleotide sequences are identical when the nucleic acid residues are the same, while two or more polynucleotide sequences are "substantially identical" if the nucleic acid residues are about 60% identical, about 65% identical, about 70% identical, about 75% identical, about 80% identical, about 85% identical, about 90% identical, or about 95% identical over a specified region. The identity can exist over a region that is at least about 75 to about 100 nucleic acids in length, over a
25 region that is about 50 nucleic acids in length, or, where not specified, across the entire sequence of a polynucleotide sequence.

The term "pharmaceutically acceptable", as used herein, refers to a material, including but not limited, to a salt, carrier or diluent, which does not abrogate the biological activity or properties of the compound, and is relatively nontoxic, i.e., the material may be administered to an individual
30 without causing undesirable biological effects or interacting in a deleterious manner with any of the

components of the composition in which it is contained.

The term “subject,” or “patient,” as used herein, refers to a vertebrate. In certain embodiments, the vertebrate is a mammal. Mammals include, but are not limited to, humans, non-human primates, farm animals (such as cows), sport animals, and pets (such as cats, dogs, and horses). In certain embodiments, a mammal is a human.

The term “therapeutically effective amount,” as used herein, refers to the amount of a composition containing the expanded cell populations and/or admixtures of the present invention administered to a subject, *e.g.*, a human patient, already suffering from a disease, condition or disorder, sufficient to cure or at least partially arrest, or relieve to some extent one or more of the symptoms of the disease, disorder or condition being treated. The effectiveness of such compositions depend conditions including, but not limited to, the severity and course of the disease, disorder or condition, previous therapy, the patient’s health status and response to the drugs, and the judgment of the treating physician. By way of example only, therapeutically effective amounts may be determined by routine experimentation, including but not limited to a dose escalation clinical trial.

The term antigen presenting cell (APC) refers to a wild-type APC, or an engineered or artificial antigen presenting cell (aAPC). APCs can be provided as an irradiated population of APCs. APCs can be provided from a immortalized cell line (*e.g.*, K562 or an engineered aAPC derived from an immortalized cell line) or as a fraction of cells from a donor (*e.g.*, PBMCs).

As used herein, the terms “structurally different” and “structurally distinct,” in reference to a protein or polypeptide fragment thereof, or an epitope, refer to a covalent (*i.e.*, structural) difference between at least two different proteins, polypeptide fragments thereof, or epitopes. For example, two structurally different proteins (*e.g.*, antibodies) can refer to two proteins that have different primary amino acid sequences. In some cases, structurally different activating agents bind structurally different epitopes, such as epitopes having a different primary amino acid sequence.

As used herein, the term “anti-tumor cytotoxicity” that is “independent of” a specified receptor activity (*e.g.*, NKp30 activity, NKp44 activity, and/or NKp46 activity), refers to anti-tumor cytotoxicity that is exhibited whether or not the specified receptor or specified combination of receptors is expressed by the cell or functional. As such, a $\gamma\delta$ T-cell that exhibits anti-tumor cytotoxicity that is independent of NKp30 activity, NKp44 activity, and/or NKp46 activity can also exhibit NKp30 activity-dependent anti-tumor cytotoxicity, NKp44 activity -dependent anti-tumor

cytotoxicity, and/or NKp46 activity-dependent anti-tumor cytotoxicity.

As used herein, the terms “NKp30 activity-dependent anti-tumor cytotoxicity,” “NKp44 activity-dependent anti-tumor cytotoxicity,” and “NKp46 activity-dependent anti-tumor cytotoxicity” refer to anti-tumor cytotoxicity that requires functional expression of the specified receptor. The presence or absence of such receptor dependent anti-tumor cytotoxicity can be determined by performing standard *in vitro* cytotoxicity assays, such as performed in Example 48, in the presence or absence of an antagonist to the specified receptor. For example, presence or absence of NKp30 activity-dependent anti-tumor cytotoxicity can be determined by comparing the results of an *in vitro* cytotoxicity assays as described in Example 48 in the presence of an anti-NKp30 antagonist to the results obtained in the absence of an anti-NKp30 antagonist.

As used herein, a $\gamma\delta$ T-cell population that comprises anti-tumor cytotoxicity, wherein at least a specified “%” of the anti-tumor cytotoxicity is “independent of” a specified receptor activity (e.g., NKp30 activity, NKp44 activity, and/or NKp46 activity), refers to a cell where blocking specified receptor reduces measured anti-tumor cytotoxicity by no more than the numerical % value. Thus, a $\gamma\delta$ T-cell population that comprises anti-tumor cytotoxicity, wherein at least 50% of the anti-tumor cytotoxicity is independent of NKp30 activity would exhibit a reduction of 50% or less of *in vitro* anti-tumor cytotoxicity in the presence of an NKp30 antagonist as compared to in the absence of the NKp30 antagonist.

Overview

In humans, $\gamma\delta$ T-cell(s) are a subset of T-cells that provide a link between the innate and adaptive immune responses. These cells undergo V-(D)-J segment rearrangement to generate antigen-specific $\gamma\delta$ T-cell receptors ($\gamma\delta$ TCRs), and $\gamma\delta$ T-cell(s) and can be directly activated via the recognition of an antigen by either the $\gamma\delta$ TCR or other, non-TCR proteins, acting independently or together to activate $\gamma\delta$ T-cell effector functions. $\gamma\delta$ T-cells represent a small fraction of the overall T-cell population in mammals, approximately 1-5% of the T-cells in peripheral blood and lymphoid organs, and they appear to reside primarily in epithelial cell-rich compartments like skin, liver, digestive, respiratory, and reproductive tracks. Unlike $\alpha\beta$ TCRs, which recognize antigens bound to major histocompatibility complex molecules (MHC), $\gamma\delta$ TCRs can directly recognize bacterial antigens, viral antigens, stress antigens expressed by diseased cells, and tumor antigens in the form

of intact proteins or non-peptide compounds.

TS-1, TS8.2, B6, and 15D can activate $\gamma\delta$ T cells. Without being bound by theory, different levels of activation and expansion of cultures originating from different donors may be due to the donor $\gamma\delta$ variable TCR repertoire and the specificity of the antibody binding epitope. It has been discovered that not every agent which binds to specific $\gamma\delta$ T-cell subsets is capable of activating the specific $\gamma\delta$ T-cell and particularly activating the specific $\gamma\delta$ T-cell population to clinically-relevant levels, *i.e.*, $>10^8$ target $\gamma\delta$ T cells in an enriched culture. Similarly, not every binding epitope of a $\gamma\delta$ T-cell population is an activating epitope, *i.e.*, capable of activation of the specific $\gamma\delta$ T-cell population upon binding.

The inventors of the present invention have identified specific $\gamma\delta$ variable TCR binding regions associated with potent activation of specific $\gamma\delta$ T cell subtypes thus enabling the specific activation of $\gamma\delta$ T cell subtypes which produces clinically relevant levels of highly enriched $\gamma\delta$ T-cell populations with increased purity, and admixtures thereof, that can be administered to patients. Novel activating ligands, including antibodies, which specifically bind the activating epitopes capable of inducing enhanced activation and expansion of $\gamma\delta$ T cell subtypes are also contemplated and further described herein.

In some cases, the production of clinically relevant levels (*i.e.*, $>10^8$) of $\gamma\delta$ T cells using the methods of the present invention can be obtained with relatively small volumes of culture medium. For example, in some embodiments, the clinically relevant levels of $\gamma\delta$ T cells can be obtained from expansion of a population of cells (e.g., an isolated mixed population of cells), in a final culture volume of approximately 25 L; 20 L; 10 L; 5 L; 3 L; 2 L; 1,5000 mL; 1,000 mL; 500 mL; 200 mL; 150 mL; 100 mL, or less (e.g., from about 10 mL to about 100 mL, from about 100 mL to about 500 mL, from about 500 mL to about 5,000 mL, or from about 5 L to about 25 L). As another example, in some embodiments, the clinically relevant levels of $\gamma\delta$ T cells can be obtained under conditions such that the total volume of culture media used in expanding an isolated mixed population of cells obtained from a donor, or multiple donors, is less than about 50 L, 25L, 20L, 10 L, 5L, 1 L, or 750 mL (e.g., from about 750 mL to less than about 50 L, from about 100 mL to about 750 mL, from about 750 mL to about 5 L, from about 1 L to about 10 L, from about 10L to about 50 L, or from about 10 L to about 25 L).

Described herein are methods for the selective activation and expansion of $\gamma\delta$ T-cell subtypes

directly from isolated mixed cell populations, *e.g.*, without prior depletion of non-target cell types, providing clinically-relevant levels of enriched $\gamma\delta$ T cell population(s) having cytotoxic properties. Activating $\gamma\delta$ variable TCR epitopes of specific $\gamma\delta$ cell population(s) are also described. The present invention also provides methods of treatment with compositions comprising the enriched $\gamma\delta$ T-cell population(s) of the invention.

Described herein are methods of producing or providing clinically relevant levels ($>10^8$) of engineered or non-engineered $\gamma\delta$ T-cells, including one or more specific subsets of $\gamma\delta$ T-cells. Such methods can be used to produce such clinically relevant levels from a single donor, including from a single sample of a single donor. Moreover, such methods can be used to produce significantly greater than 10^8 engineered or non-engineered $\gamma\delta$ T-cells. For example, in some embodiments about, or at least about, 10^9 , 10^{10} , 10^{11} , or 10^{12} engineered or non-engineered $\gamma\delta$ T-cells, including one or more specific subsets of $\gamma\delta$ T-cells, can be produced in the methods described herein. In some cases, such population sizes can be achieved in as few as 19-30 days and/or with a total volume of culture media used of less than about 1L.

Isolation of $\gamma\delta$ T-cells

In some aspects, the instant invention provides *ex vivo* methods for expansion of engineered or non-engineered $\gamma\delta$ T-cells. In some cases, the method employ one or more (*e.g.*, first and/or second) expansion steps that do not include a cytokine that favors expansion of a specific population of $\gamma\delta$ T-cells, such as IL-4, IL-2, or IL-15, or a combination thereof. In some embodiments, the instant invention provides *ex vivo* methods for producing enriched $\gamma\delta$ T-cell populations from isolated mixed cell populations, comprising contacting the mixed cell population with one or more agents which selectively expand $\delta 1$ T-cells; $\delta 1$ T-cells and $\delta 3$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells by binding to an epitope specific of a $\delta 1$ TCR; a $\delta 1$ and $\delta 4$ TCR; or a $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ TCR respectively to provide an enriched $\gamma\delta$ T cell population. In other aspects, the instant invention provides *ex vivo* methods for producing enriched $\gamma\delta$ T-cell populations from isolated mixed cell populations, comprising contacting the mixed cell population with one or more agents which selectively expand $\delta 2$ T-cells by binding to an epitope specific of a $\delta 2$ TCR to provide an enriched $\gamma\delta$ T cell population.

In other aspects, the present disclosure provides methods for the genetic engineering of $\gamma\delta$ T-cells that have been isolated from a subject. Methods of enrichment, activation, expansion, or

genetic engineering can be performed singly or in combination, in any order. In one embodiment, $\gamma\delta$ T-cells can be isolated, genetically engineered, and then activated and expanded. In a preferred embodiment, $\gamma\delta$ T-cells can be isolated, activated and expanded, and then optionally genetically engineered. In some embodiments, such activated and expanded and then genetically engineered $\gamma\delta$ T-cells can be further activated and/or expanded.

An, *e.g.*, non-engineered, $\gamma\delta$ T-cell population can be expanded from a complex sample of a subject. A complex sample can be a peripheral blood sample (*e.g.*, PBLs or PBMCs), a leukapheresis sample, a cord blood sample, a tumor, a stem cell precursor, a tumor biopsy, a tissue, a lymph, or from epithelial sites of a subject directly contacting the external milieu, or derived from stem precursor cells. In some cases, the present disclosure provides methods for selective expansion of $V\delta 1^+$ cells, $V\delta 2^+$ cells, $V\delta 1^+$ cells and $V\delta 3^+$ cells, $V\delta 1^+$ cells and $V\delta 4^+$ cells, $V\delta 1^+$ cells, $V\delta 3^+$ cells, $V\delta 4^+$ cells, and $V\delta 5^+$ cells, or any combination thereof.

Peripheral blood mononuclear cells can be collected from a subject, for example, with an apheresis machine, including the Ficoll-Paque™ PLUS (GE Healthcare) system, or another suitable device/system. $\gamma\delta$ T-cell(s), or a desired subpopulation of $\gamma\delta$ T-cell(s), can be purified from the collected sample with, for example, flow cytometry techniques. Cord blood cells can also be obtained from cord blood during the birth of a subject. See WO 2016/081518, incorporated by reference herein in its entirety for all purposes including but not limited to methods and compositions for PBMC isolation, $\gamma\delta$ T cell activation, and making and using $\gamma\delta$ T cell activation agents.

A $\gamma\delta$ T-cell may be expanded from an isolated complex sample or mixed cell population that is cultured *in vitro* by contacting the mixed cell population with one or more agents which expand $\gamma\delta$ T-cell by specifically binding to an epitope of a $\gamma\delta$ TCR to provide an enriched $\gamma\delta$ T-cell population, *e.g.*, in a first enrichment step. In some embodiments, $\gamma\delta$ T cells comprised in a whole PBMC population, without prior depletion of one or more specific cell populations such as one or more or all of the following non- $\gamma\delta$ T cell monocytes: $\alpha\beta$ T-cells, B-cells, and NK cells, can be activated and expanded, resulting in an enriched $\gamma\delta$ T-cell population. In some aspects, activation and expansion of $\gamma\delta$ T-cell are performed without the presence of native or engineered APCs. In some aspects, isolation and expansion of $\gamma\delta$ T cells from tumor specimens can be performed using immobilized $\gamma\delta$ T cell mitogens, including antibodies specific to activating epitopes of a $\gamma\delta$ TCR, and other

activating agents, including lectins, which bind the activating epitopes of a $\gamma\delta$ TCR provided herein.

In certain embodiments, the isolated mixed cell population is contacted with one or more agents which expand $\gamma\delta$ T-cells for about, or at least about, 2 days, about 3 days, about 4 days, about 5 days, about 6 days, about 7 days, about 8 days, about 9 days, about 10 days, about 11 days, about 12 days, about 13 days, about 14 days, about 15 days, about 17 days, about 19 days, about 21 days, about 25 days, about 29 days, about 30 days, or any range therein. For example, the isolated mixed cell population is contacted with one or more agents which expand $\gamma\delta$ T-cells for about 1 to about 4 days, about 2 to about 4 days, about 2 to about 5 days, about 3 to about 5 days, about 5 to about 21 days, about 5 to about 19 days, about 5 to about 15 days, about 5 to about 10 days, or about 5 to about 7 days, to provide a first enriched $\gamma\delta$ T-cell population. As another example, the isolated mixed cell population is contacted with one or more agents which expand $\gamma\delta$ T-cells for about 7 to about 21 days, about 7 to about 19 days, about 7 to about 23 days, or about 7 to about 15 days to provide a first enriched $\gamma\delta$ T-cell population.

In some cases, a purification or isolation step is performed between the first and second expansion steps. In some cases, the isolation step includes removal of one or more activating agents. In some cases, the isolation step includes specific isolation of $\gamma\delta$ T-cells, or a subtype thereof. In some cases, one or more (*e.g.*, all) activating agents (*e.g.*, all activating agents that are not common components of cell culture media such as serum components and/or IL-2)) are removed between first and second expansion steps, but $\gamma\delta$ T-cells are not specifically isolated from other cell types ($\alpha\beta$ T-cells).

In some embodiments, following the activation and expansion of $\gamma\delta$ T cells using activating agents which bind to an activating epitope of a $\gamma\delta$ TCR, in a first enrichment step, and optionally a second enrichment step, the, *e.g.*, first, enriched $\gamma\delta$ T cell population(s) of the invention may be further enriched or purified using techniques known in the art to obtain a second or further enriched $\gamma\delta$ T cell population(s) in a second, third, fourth, fifth, etc. enrichment step. For example, the, *e.g.*, first, enriched $\gamma\delta$ T cell population(s) may be depleted of $\alpha\beta$ T-cells, B-cells and NK cells. Positive and/or negative selection of cell surface markers expressed on the collected $\gamma\delta$ T-cell(s) can be used to directly isolate a $\gamma\delta$ T-cell, or a population of $\gamma\delta$ T-cell(s) expressing similar cell surface markers from the, *e.g.*, first, enriched $\gamma\delta$ T-cell population(s). For instance, a $\gamma\delta$ T-cell can be isolated from an enriched $\gamma\delta$ T-cell population (*e.g.*, after a first and/or second step of expansion) based on positive

or negative expression of markers such as CD2, CD3, CD4, CD8, CD24, CD25, CD44, Kit, TCR α , TCR β , TCR γ (including one or more TCR γ sub-types), TCR δ (including one or more TCR δ sub-types), NKG2D, CD70, CD27, CD28, CD30, CD16, OX40, CD46, CD161, CCR7, CCR4, NKp30, NKp44, NKp46, DNAM-1, CD242, JAML, and other suitable cell surface markers.

5 In some embodiments, after a first step of expansion (*e.g.*, after an isolation step performed subsequent to the first step of expansion), the expanded cells are, optionally diluted, and cultured in a second step of expansion. In preferred embodiments, the second step of expansion is performed under conditions in which culture media is replenished about every 1-2, 1-3, 1-4, 1-5, 2-5, 2-4, or 2-3 days in a second expansion step. In some embodiments, the second step of expansion is performed
10 under conditions in which the cells are diluted or adjusted to a density that supports further $\gamma\delta$ T-cell expansion 1, 2, 3, 4, 5, 6, or more times. In some cases, the cell density adjustment is performed contemporaneously with (*i.e.*, on the same day as, or at the same time as) replenishment of culture media. For example, cell density can be adjusted every 1-2, 1-3, 1-4, 1-5, 2-5, 2-4, or 2-3 days in a second expansion step. Typical cell densities that support further $\gamma\delta$ T-cell expansion include, but
15 are not limited to, about 1×10^5 , 2×10^5 , 3×10^5 , 4×10^5 , 5×10^5 , 6×10^5 , 7×10^5 , 8×10^5 , 9×10^5 , 1×10^6 , 2×10^6 , 3×10^6 , 4×10^6 , 5×10^6 cells/mL, 10×10^6 cells/mL, 15×10^6 cells/mL, 20×10^6 cells/mL, or 30×10^6 cells/mL of culture.

In some embodiments, cell density is adjusted to a density of from about 0.5×10^6 to about 1×10^6 cells/mL, from about 0.5×10^6 to about 1.5×10^6 cells/mL, from about 0.5×10^6 to about 2×10^6 cells/mL, from about 0.75×10^6 to about 1×10^6 cells/mL, from about 0.75×10^6 to about 1.5×10^6 cells/mL, from about 0.75×10^6 to about 2×10^6 cells/mL, from about 1×10^6 to about 2×10^6 cells/mL, or from about 1×10^6 to about 1.5×10^6 cells/mL, from about 1×10^6 to about 2×10^6 cells/mL, from about 1×10^6 to about 3×10^6 cells/mL, from about 1×10^6 to about 4×10^6 cells/mL, from about 1×10^6 to about 5×10^6 cells/mL, from about 1×10^6 to about 10×10^6 cells/mL, from
25 about 1×10^6 to about 15×10^6 cells/mL, from about 1×10^6 to about 20×10^6 cells/mL, or from about 1×10^6 to about 30×10^6 cells/mL.

In some embodiments, the second step of expansion is performed under conditions in which the cells are monitored and maintained at a predetermined cell density (or density interval) and/or maintained in culture medium having a predetermined glucose content. For example, the cells can
30 be maintained at a viable cell density of from about 0.5×10^6 to about 1×10^6 cells/mL, from about

0.5 x 10⁶ to about 1.5 x 10⁶ cells/mL, from about 0.5 x 10⁶ to about 2 x 10⁶ cells/mL, from about 0.75 x 10⁶ to about 1 x 10⁶ cells/mL, from about 0.75 x 10⁶ to about 1.5 x 10⁶ cells/mL, from about 0.75 x 10⁶ to about 2 x 10⁶ cells/mL, from about 1 x 10⁶ to about 2 x 10⁶ cells/mL, or from about 1 x 10⁶ to about 1.5 x 10⁶ cells/mL, from about 1 x 10⁶ to about 3 x 10⁶ cells/mL, from about 1 x 10⁶ to about 4 x 10⁶ cells/mL, from about 1 x 10⁶ to about 5 x 10⁶ cells/mL, from about 1 x 10⁶ to about 10 x 10⁶ cells/mL, from about 1 x 10⁶ to about 15 x 10⁶ cells/mL, from about 1 x 10⁶ to about 20 x 10⁶ cells/mL, from about 1 x 10⁶ to about 30 x 10⁶ cells/mL.

In some cases, the cells can be maintained at a higher concentration for at least a portion of the expansion. For example, for a first portion of a first or second expansion, cells viability may be enhanced at a higher cell concentration. As another example, for a final portion of a first or second expansion culture volume may be most efficiently utilized at a higher cell concentration. Thus, in some embodiments, cells can be maintained at a viable cell density of from about 1 x 10⁶ cells/mL to about 20 x 10⁶ cells/mL for at least a portion of a first or second expansion culture or all of a first or second expansion culture.

As another example, the cells can be maintained in culture medium having a glucose content of from about 0.5 g/L to about 1 g/L, from about 0.5 g/L to about 1.5 g/L, from about 0.5 g/L to about 2 g/L, from about 0.75 g/L to about 1 g/L, from about 0.75 g/L to about 1.5 g/L, from about 0.75 g/L to about 2 g/L, from about 1 g/L to about 1.5 g/L, from about 1 g/L to about 2 g/L, from 1 g/L to 3 g/L, or from 1 g/L to 4 g/L.. In some embodiments, the cells can be maintained in culture medium having a glucose content of about 1.25 g/L. In some cases, such as where a high cell density culture is maintained, cells can be maintained in culture medium having a glucose content of about 1 g/L to about 5 g/L, from about 1 g/L to about 4 g/L, from about 2 g/L to about 5 g/L, or from about 2 g/L to about 4 g/L.

Typically glucose content is maintained by addition of fresh serum containing or serum free culture medium to the culture. In some embodiments, the cells can be maintained at a predetermined viable cell density interval and in a culture medium having a predetermined glucose content interval, *e.g.*, by monitoring each parameter and adding fresh media to maintain the parameters within the predetermined limits. In some embodiments, glucose content is maintained by adding fresh serum containing or serum free culture medium in the culture while removing spent medium in a perfusion bioreactor while retaining the cells inside. In some embodiments, additional parameters including,

without limitation, one or more of: pH, partial pressure of O₂, O₂ saturation, partial pressure of CO₂, CO₂ saturation, lactate, glutamine, glutamate, ammonium, sodium, potassium, and calcium, are monitored and/or maintained during a $\gamma\delta$ T-cell expansion (*e.g.*, selective $\gamma\delta$ T-cell expansion) or during a first or second step of $\gamma\delta$ T-cell expansion (*e.g.*, selective $\gamma\delta$ T-cell expansion) described
 5 herein.

A $\gamma\delta$ T-cell subtype may be selectively expanded from an isolated complex sample or mixed cell population that is cultured *in vitro* by contacting the mixed cell population with one or more agents which:

- i) selectively expand $\delta 1$ T-cells by specifically binding to an epitope of a $\delta 1$ TCR,
- 10 ii) selectively expand $\delta 2$ T-cells by specifically binding to an epitope of a $\delta 2$ TCR,
- iii) selectively expand $\delta 1$ and $\delta 4$ T cells by specifically binding to an epitope of a $\delta 1$ and a $\delta 4$ TCR; or
- iv) selectively expand $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T cells by specifically binding to an epitope of a $\delta 1$, $\delta 3$, $\delta 4$, and a $\delta 5$ TCR,

15 to provide an enriched $\gamma\delta$ T-cell population, *e.g.*, in a first enrichment step. In some cases, the one or more agents specifically bind to a $\delta 1J1$, $\delta 1J2$, or $\delta 1J3$ TCR, or two thereof, or all thereof. In some embodiments, $\gamma\delta$ cells in a whole PBMC population, without prior depletion of specific cell populations such as monocytes, $\alpha\beta$ T-cells, B-cells, and NK cells, can be activated and expanded, resulting in an enriched $\gamma\delta$ T-cell population. In some aspects, activation and expansion of $\gamma\delta$ T-cell
 20 are performed without the presence of native or engineered APCs. In some aspects, isolation and expansion of $\gamma\delta$ T cells from tumor specimens can be performed using immobilized $\gamma\delta$ T cell mitogens, including antibodies specific to activating epitopes specific of a $\delta 1$ TCR; a $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ TCR, a $\delta 1$ and $\delta 4$ TCR; or a $\delta 2$ TCR, and other activating agents, including lectins, which bind the activating epitopes specific of a $\delta 1$ TCR; a $\delta 1$, $\delta 3$, $\delta 4$ and $\delta 5$ TCR; a $\delta 1$ and $\delta 4$ TCR; or a $\delta 2$
 25 TCR provided herein.

In certain embodiments, the isolated mixed cell population is contacted with one or more agents which selectively expand $\delta 1$, $\delta 1$ and $\delta 4$, $\delta 2$, or $\delta 1$ and $\delta 2$ T-cells for about 5 days, 6 days, about 7 days, about 8 days, about 9 days, about 10 days, about 11 days, about 12 days, about 13 days, about 14 days, about 15 days, or any range therein. For example, the isolated mixed cell population
 30 is contacted with one or more agents which selectively expand $\delta 1$ or $\delta 2$ T-cells for about 1 to about 3

days, about 1 to about 4 days, about 1 to about 5 days, about 2 to about 3 days, about 2 to about 4 days, about 2 to about 5 days, about 3 to about 4 days, about 3 to about 5 days, about 4 to about 5 days, about 5 to about 15 days, or about 5 to about 7 days, to provide a first enriched $\gamma\delta$ T-cell population. In some embodiments selectively expanded $\delta 1$, $\delta 1$ and $\delta 3$, $\delta 1$ and $\delta 4$, $\delta 2$, or $\delta 1$ and $\delta 2$ T-cells are further expanded in a second step of expansion as described herein.

In certain embodiments, the starting isolated mixed cell population, *e.g.*, peripheral blood sample, comprises T lymphocytes in the range of about 20-80%. In certain embodiments, the percent of residual $\alpha\beta$ T cells and NK cells in enriched $\gamma\delta$ T-cell population(s) of the invention is about, or less than about, 2.5% and 1 %, respectively. In certain embodiments, the percent of residual $\alpha\beta$ T cells or NK cells in enriched $\gamma\delta$ T-cell population(s) of the invention is about, or less than about, 1%, 0.5%, 0.4%, 0.2%, 0.1%, or 0.01%. In certain embodiments, the percent of residual $\alpha\beta$ T cells in enriched $\gamma\delta$ T-cell population(s) of the invention is about, or less than about, 0.4%, 0.2%, 0.1%, or 0.01% (*e.g.*, after a step of positive selection for $\gamma\delta$ T-cells or a sub-type thereof or after depletion of $\alpha\beta$ T cells). In some embodiments, $\alpha\beta$ T cells are depleted, but NK cells are not depleted before or after a first and/or second $\gamma\delta$ T-cell expansion. In certain aspects, the isolated mixed cell population is derived from a single donor. In other aspects, the isolated mixed cell population is derived from more than one donor or multiple donors (*e.g.*, 2, 3, 4, 5, or from 2-5, 2-10, or 5-10 donors, or more).

As such, in some embodiments, the methods of the present invention can provide a clinically relevant number ($>10^8$, $>10^9$, $>10^{10}$, $>10^{11}$, or $>10^{12}$, or from about 10^8 to about 10^{12}) of expanded $\gamma\delta$ T-cells from as few as one donor. In some cases, the methods of the present invention can provide a clinically relevant number ($>10^8$, $>10^9$, $>10^{10}$, $>10^{11}$, or $>10^{12}$, or from about 10^8 to about 10^{12}) of expanded $\gamma\delta$ T-cells within less than 19 or 21 days from the time of obtaining a donor sample.

Following the specific activation and expansion of the specific $\gamma\delta$ T cell subsets using activating agents which bind to an activating epitope specific of a $\delta 1$, a $\delta 1$ and $\delta 3$ TCR, a $\delta 1$ and $\delta 4$ TCR, or a $\delta 2$ TCR, in a first enrichment step, the first enriched $\gamma\delta$ T cell population(s) of the invention may be further enriched or purified using techniques known in the art to obtain a second or further enriched $\gamma\delta$ T cell population(s) in a second, third, fourth, fifth, etc. enrichment step. For example, the first enriched $\gamma\delta$ T cell population(s) may be depleted of $\alpha\beta$ T-cells, B-cells and NK cells. Positive and/or negative selection of cell surface markers expressed on the collected $\gamma\delta$ T-

cell(s) can be used to directly isolate a $\gamma\delta$ T-cell, or a population of $\gamma\delta$ T-cell(s) expressing similar cell surface markers from the first enriched $\gamma\delta$ T-cell population(s). For instance, a $\gamma\delta$ T-cell can be isolated from a first enriched $\gamma\delta$ T-cell population based on positive or negative expression of markers such as CD2, CD3, CD4, CD8, CD24, CD25, CD44, Kit, TCR α , TCR β , TCR γ (or one or more subtypes thereof), TCR δ (or one or more subtypes thereof), NKG2D, CD70, CD27, CD28, CD30, CD16, OX40, CD46, CD161, CCR7, CCR4, DNAM-1, JAML, and other suitable cell surface markers.

In some embodiments, following the first $\gamma\delta$ T-cell expansion, first enrichment step, second $\gamma\delta$ T-cell expansion, and/or second enrichment step, of the invention, the enriched $\gamma\delta$ T-cell population comprises clinically-relevant levels of $\gamma\delta$ T-cell subsets of $>10^8$ cells, e.g., in a culture volume of less than 10 mL, 25 mL, 50 mL, 100 mL, 150 mL, 200 mL, 500 mL, 750 mL, 1 L, 2 L, 3 L, 4 L, 5 L, 10 L, 20 L, or 25 L.. For example, the methods of the present invention can provide clinically-relevant levels of $\gamma\delta$ T-cell subsets of $>10^8$ cells in a expansion culture having a volume of from 10-100 mL; from 25-100 mL; from 50-100 mL; from 75-100mL; from 10-150 mL; from 25-150 mL; from 50-150 mL; from 75-150 mL; from 100-150 mL; from 10-200 mL; from 25-200 mL; from 50-200 mL; from 75-200 mL, from 100-200 mL; from 10-250 mL; from 25-250 mL; from 50-250 mL; from 75-250 mL, from 100-250 mL; from 150-250 mL; from 5-1,000 mL; from 10-1,000 mL, or from 100-1,000 mL; from 150-1,000 mL; from 200-1,000 mL; from 250-1,000 mL, 400 mL to 1L, 1 L to 2 L, 2 L to 5 L, 2 L to 10 L, 4 L to 10 L, 4 L to 15 L, 4 L to 20 L, or 4 L to 25 L. In other embodiments, following the second, third, fourth, fifth, etc. enrichment step of the invention, the enriched $\gamma\delta$ T-cell population comprises clinically-relevant levels of $\gamma\delta$ T-cell subsets of $>10^8$.

In some embodiments, $\gamma\delta$ T-cell(s) can rapidly expand in response to contact with one or more antigens. Some $\gamma\delta$ T-cell(s), such as V γ 9V δ 2⁺ $\gamma\delta$ T-cell(s) rapidly expand *in vitro* in response to contact with some antigens, like prenyl-pyrophosphates, alkyl amines, and metabolites or microbial extracts during tissue culture. In addition, some wild-type $\gamma\delta$ T-cell(s), such as V γ 2V δ 2⁺ $\gamma\delta$ T-cell(s) rapidly expand *in vivo* in humans in response to certain types of vaccination(s). Stimulated $\gamma\delta$ T-cells can exhibit numerous antigen-presentation, co-stimulation, and adhesion molecules that can facilitate the isolation of a $\gamma\delta$ T-cell(s) from a complex sample. A $\gamma\delta$ T-cell(s) within a complex sample can be stimulated *in vitro* with at least one antigen for 1 day, 2 days, 3 days, 4 days, 5 days, 6 days, 7 days, about 5-15 days, 5-10 days, or 5-7 days, or another suitable period of

time, *e.g.*, in combination with, before, or after expansion with a selective $\gamma\delta$ T-cell expansion agent described herein such as an antibody or an immobilized antibody. Stimulation of the $\gamma\delta$ T-cell with a suitable antigen can expand the $\gamma\delta$ T-cell population *in vitro*.

Non-limiting examples of antigens that may be used to stimulate the expansion of $\gamma\delta$ T-cell(s) from a complex sample *in vitro* include, prenyl-pyrophosphates, such as isopentenyl pyrophosphate (IPP), alkyl-amines, metabolites of human microbial pathogens, metabolites of commensal bacteria, -methyl-3-butenyl-1-pyrophosphate (2M3B1PP), (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate (HMB-PP), ethyl pyrophosphate (EPP), farnesyl pyrophosphate (FPP), dimethylallyl phosphate (DMAP), dimethylallyl pyrophosphate (DMAPP), ethyl-adenosine triphosphate (EPPPA), geranyl pyrophosphate (GPP), geranylgeranyl pyrophosphate (GGPP), isopentenyl-adenosine triphosphate (IPPPA), monoethyl phosphate (MEP), monoethyl pyrophosphate (MEPP), 3-formyl-1-butyl-pyrophosphate (TUBAg 1), X-pyrophosphate (TUBAg 2), 3-formyl-1-butyl-uridine triphosphate (TUBAg 3), 3-formyl-1-butyl-deoxythymidine triphosphate (TUBAg 4), monoethyl alkylamines, allyl pyrophosphate, crotyl pyrophosphate, dimethylallyl- γ -uridine triphosphate, crotyl- γ -uridine triphosphate, allyl- γ -uridine triphosphate, ethylamine, isobutylamine, sec-butylamine, iso-amylamine and nitrogen containing bisphosphonates.

Activation and expansion of $\gamma\delta$ T-cells can be performed using activation and co-stimulatory agents described herein to trigger specific $\gamma\delta$ T-cell proliferation and persistent populations. In some embodiments, activation and expansion of $\gamma\delta$ T-cells from different cultures can achieve distinct clonal or mixed polyclonal population subsets. In some embodiments, different agonist agents can be used to identify agents that provide specific $\gamma\delta$ activating signals. In one aspect, agents that provide specific $\gamma\delta$ activating signals can be different monoclonal antibodies (MAbs) directed against the $\gamma\delta$ TCRs.

In one aspect, the MAbs can bind to different epitopes on the constant or variable regions of γ TCR and/or δ TCR. In one aspect, the MAbs can include $\gamma\delta$ TCR pan MAbs. In one aspect, the $\gamma\delta$ TCR pan MAbs may recognize domains shared by different γ and δ TCRs on either the γ or δ chain or both, including $\delta 1$ and $\delta 2$ cell populations. In one aspect, the antibodies may be 5A6.E9 (Thermo scientific), B1 (Biolegend), IMMU510 and/or 11F2 (11F2) (Beckman Coulter). In one aspect, the MAbs can be directed to specific domains unique to the variable regions of the γ chain (7A5 Mab, directed to like V $\gamma 9$ TCR (Thermo Scientific #TCR1720)), or domains on V $\delta 1$ variable region (Mab

TS8.2 (Thermo scientific #TCR1730; MAb TS-1 (ThermoFisher #TCR 1055), MAb R9.12 (Beckman Coulter #IM1761)), or V δ 2 chain (MAb 15D (Thermo Scientific #TCR1732 or Life technologies #TCR2732) B6 (Biolegend #331402), one of the δ 1-# antibodies described in **FIGs.** 33-34, or one of the δ 2-# antibodies described in **FIGs.** 35-36.

5 In some embodiments, antibodies against different domains of the $\gamma\delta$ TCR (pan antibodies and antibodies recognizing specific variable region epitopes on subset populations) can be combined to evaluate their ability to enhanced activation of $\gamma\delta$ T cells. In some embodiments, $\gamma\delta$ T-cells activators can include $\gamma\delta$ TCR-binding agents such as MICA, an agonist antibody to NKG2D, an, *e.g.*, Fc tag, fusion protein of MICA, ULBP1, or ULBP3 (R&D systems Minneapolis, MN) ULBP2, 10 or ULBP6 (Sino Biological Beijing, China). In some embodiments, companion co-stimulatory agents to assist in triggering specific $\gamma\delta$ T cell proliferation without induction of cell anergy and apoptosis can be identified. These co-stimulatory agents can include ligands to receptors expressed on $\gamma\delta$ cells, such as ligand(s) to one or more of the following: NKG2D, CD161, CD70, JAML, DNAX, CD81 accessory molecule-1 (DNAM-1) ICOS, CD27, CD196, CD137, CD30, HVEM, 15 SLAM, CD122, DAP, and CD28. In some aspects, co-stimulatory agents can be antibodies specific to unique epitopes on CD2 and CD3 molecules. CD2 and CD3 can have different conformation structures when expressed on $\alpha\beta$ or $\gamma\delta$ T-cells (s), and in some cases, specific antibodies to CD3 and CD2 can lead to selective activation of $\gamma\delta$ T-cells.

A population of $\gamma\delta$ T-cell(s) may be expanded *ex vivo* prior to engineering of the $\gamma\delta$ T-cell(s).

20 Non-limiting example of reagents that can be used to facilitate the expansion of a $\gamma\delta$ T-cell population *in vitro* include anti-CD3 or anti-CD2, anti-CD27, anti-CD30, anti-CD70, anti-OX40 antibodies, IL-2, IL-4, IL-7, IL-9, IL-12, IL-15, IL-18, IL-19, IL-21, IL 23, IL-33, IFN γ , granulocyte-macrophage colony stimulating factor (GM-CSF), granulocyte colony stimulating factor (G-CSF), CD70 (CD27 ligand), concavalin A (ConA), pokeweed (PWM), protein peanut agglutinin 25 (PNA), soybean agglutinin (SBA), Les Culinaris Agglutinin (LCA), Pisum Sativum Agglutinin (PSA), Helix pomatia agglutinin (HPA), Vicia graminea Lectin (VGA), Phaseolus Vulgaris Erythroagglutinin (PHA-E), Phaseolus Vulgaris Leucoagglutinin (PHA-L), Sambucus Nigra Lectin (SNA, EBL), Maackia Amurensis, Lectin II (MAL II), Sophora Japonica Agglutinin (SJA), Dolichos Biflorus Agglutinin (DBA), Lens Culinaris Agglutinin (LCA), Wisteria Floribunda Lectin (WFA, 30 WFL) or another suitable mitogen capable of stimulating T-cell proliferation.

Genetic engineering of the $\gamma\delta$ T-cell(s) may comprise stably integrating a construct expressing a tumor recognition moiety, such as an $\alpha\beta$ TCR, a $\gamma\delta$ TCR, a CAR encoding an antibody, an antigen binding fragment thereof, or a lymphocyte activation domain into the genome of the isolated $\gamma\delta$ T-cell(s), a cytokine (*e.g.*, IL-15, IL-12, IL-2, IL-7, IL-21, IL-18, IL-19, IL-33, IL-4, IL-9, IL-23, or IL1 β) to enhance T-cell proliferation, survival, and function *ex vivo* and *in vivo*. Genetic engineering of the isolated $\gamma\delta$ T-cell may also comprise deleting or disrupting gene expression from one or more endogenous genes in the genome the isolated $\gamma\delta$ T-cell, such as the MHC locus (loci).

Ex-vivo Expansion of $\gamma\delta$ T-cells

In other aspects, the present disclosure provides methods for the *ex vivo* expansion of a population of non-engineered and engineered $\gamma\delta$ T-cells for adoptive transfer therapy. A non-engineered or engineered $\gamma\delta$ T-cell of the disclosure may be expanded *ex vivo*. A non-engineered or engineered $\gamma\delta$ T-cell of the disclosure can be expanded *in vitro* without activation by APCs, or without co-culture with APCs and/or aminophosphonates. Additionally, or alternatively, a non-engineered or engineered $\gamma\delta$ T-cell of the disclosure can be expanded *in vitro* with at least one expansion step that includes activation by or co-culture with APCs and/or with one or more aminophosphonates.

In some embodiments, a non-engineered or engineered $\gamma\delta$ T-cell of the disclosure can be expanded *in vitro* without activation by APC in a first $\gamma\delta$ T-cell expansion, and then expanded *in vitro* with activation by APC in a second $\gamma\delta$ T-cell expansion. In some cases, the first $\gamma\delta$ T-cell expansion includes contacting the $\gamma\delta$ T-cells with one or more agents which (a) expand $\gamma\delta$ T-cells, or (b) selectively expand $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 3$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR; a $\delta 2$ TCR; a $\delta 1$ and $\delta 4$ TCR; or a $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ TCR respectively.

In some cases, the second $\gamma\delta$ T-cell expansion is performed in a culture medium that is free of the one or more agents used in the first $\gamma\delta$ T-cell expansion. In some cases, the second $\gamma\delta$ T-cell expansion is performed in a culture medium that contains one or more second agents that (a) expand T cells, (b) expand $\gamma\delta$ T-cells, or (c) selectively expand $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 3$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T-cells by binding to an activating epitope

specific of a $\delta 1$ TCR; a $\delta 2$ TCR; a $\delta 1$ and $\delta 4$ TCR; or a $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ TCR respectively.

In some cases, the second agents are different (e.g., have a different primary amino acid sequence and/or bind a structurally different $\gamma\delta$ TCR epitope) as compared to the agents used in the first $\gamma\delta$ T-cell expansion. In some cases, the second agents bind an overlapping $\gamma\delta$ TCR epitope, the same $\gamma\delta$ TCR epitope, or can compete for binding to $\gamma\delta$ TCR with the agents used in the first $\gamma\delta$ T-cell expansion. In some cases, the second agents are expressed on the cell surface of an APC. In some cases, the second agents are bound to the surface of an APC, e.g., by a binding interaction between a constant region of the second agent and an Fc-receptor on the surface of the APC. In some cases, the second agents are soluble. In some cases, the second $\gamma\delta$ T-cell expansion is performed in a culture medium containing soluble second agents and APCs, optionally wherein the APC express on their cell surface or bind to their cell surface an agent that expands or selectively expands a $\gamma\delta$ T cell population.

In some cases, the first $\gamma\delta$ T-cell expansion is performed without an APC, and the second $\gamma\delta$ T-cell expansion is performed with an APC. In some cases, the second $\gamma\delta$ T-cell expansion is performed with an APC and one or more second agents that (a) expand T cells, (b) expand $\gamma\delta$ T-cells, or (c) selectively expand $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 3$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR; a $\delta 2$ TCR; a $\delta 1$ and $\delta 4$ TCR; or a $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ TCR respectively.

One of skill in the art will appreciate that, in certain embodiments, the methods of the second expansion step described herein can be performed as a first expansion step and methods of the first step described herein can be performed as a second expansion step. As an example, and without limitation, in some embodiments, a mixed population of cells (e.g., PBMC) can be expanded by contacting with an APC in a first step, and then expanded in the absence of an APC, e.g., by contacting the expanded population from the first expansion step with an immobilized agent that selectively expands $\delta 1$ T-cells; $\delta 2$ T-cells; $\delta 1$ T-cells and $\delta 3$ T-cells; $\delta 1$ T-cells and $\delta 4$ T-cells; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR; a $\delta 2$ TCR; a $\delta 1$ and $\delta 4$ TCR; or a $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ TCR respectively.

A method of the invention can expand various $\gamma\delta$ T-cell(s) populations, such as a $V\gamma 1^+$, a $V\gamma 2^+$, or $V\gamma 3^+$ $\gamma\delta$ T-cell population. In some cases, a method of the invention can expand a $V\delta 1^+$ T-

cell population; a $V\delta 1^+$ and a $V\delta 3^+$ T-cell population; a $V\delta 1^+$ and a $V\delta 4^+$ T-cell population; a $V\delta 1^+$ and a $V\delta 2^+$ T-cell population; or a $V\delta 1^+$, $V\delta 3^+$, $V\delta 4^+$, and a $V\delta 5^+$ T-cell population.

In some instances, a $\gamma\delta$ T-cell population can be expanded *in vitro* in fewer than 36 days, fewer than 35 days, fewer than 34 days, fewer than 33 days, fewer than 32 days, fewer than 31 days, fewer than 30 days, fewer than 29 days, fewer than 28 days, fewer than 27 days, fewer than 26 days, fewer than 25 days, fewer than 24 days, fewer than 23 days, fewer than 22 days, fewer than 21 days, fewer than 20 days, fewer than 19 days, fewer than 18 days, fewer than 17 days, fewer than 16 days, fewer than 15 days, fewer than 14 days, fewer than 13 days, fewer than 12 days, fewer than 11 days, fewer than 10 days, fewer than 9 days, fewer than 8 days, fewer than 7 days, fewer than 6 days, fewer than 5 days, fewer than 4 days, or fewer than 3 days.

In some aspects, provided are methods for selectively expanding various $\gamma\delta$ T-cells, including engineered and non-engineered $\gamma\delta$ T-cells by contacting the $\gamma\delta$ T-cells from the mixed cell population with an activation agent. In some cases, the activation or activating agent binds to a specific epitope on a cell-surface receptor of a $\gamma\delta$ T-cell. The activation agent can be an antibody, such as a monoclonal antibody. The activation agent can specifically activate the growth of one or more types of $\gamma\delta$ T-cells, such as $\delta 1$, $\delta 2$, $\delta 1$ and $\delta 3$, or $\delta 1$ and $\delta 4$ cell populations. In some embodiments the activation agent specifically activates the growth of $\delta 1$ cell populations to provide an enriched $\delta 1$ T-cell population. In other cases, the activation agent specifically activates the growth of $\delta 2$ cell populations to provide an enriched $\delta 2$ T-cell population.

An activation agent may stimulate the expansion of engineered and non-engineered $\gamma\delta$ T-cells at a fast rate of growth. For instance, an agent that stimulates an expansion of the $\gamma\delta$ T-cell population at a mean rate of 1 cell division in less than 30 hours, 1 cell division in less than 29 hours, 1 cell division in less than 28 hours, 1 cell division in less than 27 hours, 1 cell division in less than 26 hours, 1 cell division in less than 25 hours, 1 cell division in less than 24 hours, 1 cell division in less than 23 hours, 1 cell division in less than 22 hours, 1 cell division in less than 21 hours, 1 cell division in less than 20 hours, 1 cell division in less than 19 hours, 1 cell division in less than 18 hours, 1 cell division in less than 17 hours, 1 cell division in less than 16 hours, 1 cell division in less than 15 hours, 1 cell division in less than 14 hours, 1 cell division in less than 13 hours, 1 cell division in less than 12 hours, 1 cell division in less than 11 hours, 1 cell division in less than 10 hours, 1 cell division in less than 9 hours, 1 cell division in less than 8 hours, 1 cell division in less

than 7 hours, 1 cell division in less than 6 hours, 1 cell division in less than 5 hours, 1 cell division in less than 4 hours, 1 cell division in less than 3 hours, 1 cell division in less than 2 hours.

In some cases, an activation agent may stimulate the expansion of engineered and non-engineered $\gamma\delta$ T-cells at a mean rate of about 1 division per about 4 hours, a mean rate of about 1
5 division per about 5 hours, a mean rate of about 1 division per about 6 hours, a mean rate of about 1
division per about 7 hours, a mean rate of about 1 division per about 8 hours, a mean rate of about 1
division per about 9 hours, a mean rate of about 1 division per about 10 hours, a mean rate of about 1
division per about 11 hours, a mean rate of about 1 division per about 12 hours, a mean rate of about
10 about 1 division per about 13 hours, a mean rate of about 1 division per about 14 hours, a mean rate of
about 1 division per about 15 hours, a mean rate of about 1 division per about 16 hours, a mean rate
of about 1 division per about 17 hours, a mean rate of about 1 division per about 18 hours, a mean
rate of about 1 division per about 19 hours, a mean rate of about 1 division per about 20 hours, a
mean rate of about 1 division per about 21 hours, a rate of about 1 division per about 22 hours, a rate
of about 1 division per about 23 hours, a mean rate of about 1 division per about 24 hours, a mean
15 rate of about 1 division per about 25 hours, a mean rate of about 1 division per about 26 hours, a
mean rate of about 1 division per about 27 hours, a rate of about 1 division per about 28 hours, a rate
of about 1 division per about 29 hours, a mean rate of about 1 division per about 30 hours, a mean
rate of about 1 division per about 31 hours, a mean rate of about 1 division per about 32 hours, a
mean rate of about 1 division per about 33 hours, a rate of about 1 division per about 34 hours, a rate
20 of about 1 division per about 35 hours, a mean rate of about 1 division per about 36 hours.

In some cases, an activation agent may stimulate the rapid expansion of engineered and/or
non-engineered $\gamma\delta$ T-cells in a $\gamma\delta$ T-cell expansion culture, wherein the rapid expansion is at any one
of the foregoing mean rates of cell division and is maintained for between about 1 contiguous day
and about 19 contiguous days, between about 1 contiguous day and about 14 contiguous days,
25 between about 1 contiguous day and about 7 contiguous days, between about 1 contiguous day and
about 5 contiguous days, between about 2 contiguous days and about 19 contiguous days, between
about 2 contiguous days and about 14 contiguous days, between about 2 contiguous days and about
7 contiguous days, between about 2 contiguous days and about 5 contiguous days, between about 4
contiguous days and about 19 contiguous days, between about 4 contiguous days and about 14
30 contiguous days, between about 4 contiguous days and about 7 contiguous days, or between about 4

contiguous days and about 5 contiguous days.

In some cases, an activation agent may stimulate the expansion of engineered and/or non-engineered $\gamma\delta$ T-cells in a $\gamma\delta$ T-cell expansion culture that has been maintained for between about 2 and about 7 contiguous days, or between about 2 and about 5 contiguous days, at a mean rate of about 1 division per about 12 hours (e.g., 10-12 hours), a mean rate of about 1 division per about 13 hours (e.g., 10-13 hours), a mean rate of about 1 division per about 14 hours (e.g., 10-14 hours), a mean rate of about 1 division per about 15 hours (e.g., 10-15 hours), a mean rate of about 1 division per about 16 hours (e.g., 10-16 hours), a mean rate of about 1 division per about 17 hours (e.g., 10-17 hours or 12-17 hours), a mean rate of about 1 division per about 18 hours (e.g., 10-18 hours or 12-18 hours), a mean rate of about 1 division per about 19 hours (e.g., 10-19 hours or 12-19 hours), a mean rate of about 1 division per about 20 hours (e.g., 12-20 hours, 16-20 hours or 18-20 hours), a mean rate of about 1 division per about 21 hours (e.g., 12-21 hours, 16-21 hours or 18-21 hours), a rate of about 1 division per about 22 hours (e.g., 12-22 hours, 16-22 hours or 18-22 hours), a rate of about 1 division per about 23 hours or less (e.g., 12-23 hours, 16-23 hours or 18-23 hours), a mean rate of about 1 division per about 24 hours (e.g., 12-24 hours, 16-24 hours or 18-24 hours).

In some cases, an activation agent may stimulate the expansion of engineered and/or non-engineered $\gamma\delta$ T-cells in a $\gamma\delta$ T-cell expansion culture that has been maintained for between about 2 and about 7 contiguous days, or between about 2 and about 5 contiguous days at a mean rate of about 1 division per about 25 hours (e.g., 12-25 hours, 16-25 hours 18-25 hours, or 20-25 hours), a mean rate of about 1 division per about 26 hours (e.g., 12-26 hours, 16-26 hours 18-26 hours, or 20-26 hours), a mean rate of about 1 division per about 27 hours (e.g., 12-27 hours, 16-27 hours 18-27 hours, or 20-27 hours), a rate of about 1 division per about 28 hours (e.g., 12-28 hours, 16-28 hours 18-28 hours, 20-28 hours, or 22-28 hours), a rate of about 1 division per about 29 hours (e.g., 16-29 hours 18-29 hours, 20-29 hours, or 22-29 hours), a mean rate of about 1 division per about 30 hours (e.g., 16-30 hours 18-30 hours, 20-30 hours, or 22-30 hours), a mean rate of about 1 division per about 31 hours (e.g., 16-31 hours 18-31 hours, 20-31 hours, 22-31 hours, or 24-31 hours), a mean rate of about 1 division per about 32 hours (e.g., 18-32 hours, 20-32 hours, 22-32 hours, or 24-32 hours), a mean rate of about 1 division per about 33 hours (e.g., 18-33 hours, 20-33 hours, 22-33 hours, or 24-33 hours), a rate of about 1 division per about 34 hours (e.g., 18-34 hours, 20-34 hours, 22-34 hours, or 24-34 hours), a rate of about 1 division per about 35 hours (e.g., 18-35 hours, 20-35

hours, 22-35 hours, or 24-35 hours), a mean rate of about 1 division per about 36 hours (e.g., 18-36 hours, 20-36 hours, 22-36 hours, or 24-36 hours).

In some cases, an activation agent may stimulate the expansion of engineered and/or non-engineered $\gamma\delta$ T-cells in a $\gamma\delta$ T-cell expansion culture that has been maintained for at least 14
5 contiguous days at a mean rate of about 1 division per about 12 hours (e.g., 10-12 hours), a mean rate of about 1 division per about 13 hours (e.g., 10-13 hours), a mean rate of about 1 division per about 14 hours (e.g., 10-14 hours), a mean rate of about 1 division per about 15 hours (e.g., 10-15 hours), a mean rate of about 1 division per about 16 hours (e.g., 10-16 hours), a mean rate of about 1 division per about 17 hours (e.g., 10-17 hours or 12-17 hours), a mean rate of about 1 division per
10 about 18 hours (e.g., 10-18 hours or 12-18 hours), a mean rate of about 1 division per about 19 hours (e.g., 10-19 hours or 12-19 hours), a mean rate of about 1 division per about 20 hours (e.g., 12-20 hours, 16-20 hours or 18-20 hours), a mean rate of about 1 division per about 21 hours (e.g., 12-21 hours, 16-21 hours or 18-21 hours), a rate of about 1 division per about 22 hours (e.g., 12-22 hours, 16-22 hours or 18-22 hours), a rate of about 1 division per about 23 hours or less (e.g., 12-23 hours, 16-23 hours or 18-23 hours), a mean rate of about 1 division per about 24 hours (e.g., 12-24 hours, 16-24 hours or 18-24 hours).
15

In some cases, an activation agent may stimulate the expansion of engineered and/or non-engineered $\gamma\delta$ T-cells in a $\gamma\delta$ T-cell expansion culture that has been maintained for at least 14 contiguous days at a mean rate of about 1 division per about 25 hours (e.g., 12-25 hours, 16-25 hours
20 18-25 hours, or 20-25 hours), a mean rate of about 1 division per about 26 hours (e.g., 12-26 hours, 16-26 hours 18-26 hours, or 20-26 hours), a mean rate of about 1 division per about 27 hours (e.g., 12-27 hours, 16-27 hours 18-27 hours, or 20-27 hours), a rate of about 1 division per about 28 hours (e.g., 12-28 hours, 16-28 hours 18-28 hours, 20-28 hours, or 22-28 hours), a rate of about 1 division per about 29 hours (e.g., 16-29 hours 18-29 hours, 20-29 hours, or 22-29 hours), a mean rate of
25 about 1 division per about 30 hours (e.g., 16-30 hours 18-30 hours, 20-30 hours, or 22-30 hours), a mean rate of about 1 division per about 31 hours (e.g., 16-31 hours 18-31 hours, 20-31 hours, 22-31 hours, or 24-31 hours), a mean rate of about 1 division per about 32 hours (e.g., 18-32 hours, 20-32 hours, 22-32 hours, or 24-32 hours), a mean rate of about 1 division per about 33 hours (e.g., 18-33 hours, 20-33 hours, 22-33 hours, or 24-33 hours), a rate of about 1 division per about 34 hours (e.g.,
30 18-34 hours, 20-34 hours, 22-34 hours, or 24-34 hours), a rate of about 1 division per about 35 hours

(e.g., 18-35 hours, 20-35 hours, 22-35 hours, or 24-35 hours), a mean rate of about 1 division per about 36 hours (e.g., 18-36 hours, 20-36 hours, 22-36 hours, or 24-36 hours).

In some cases, an activation agent may stimulate the expansion of engineered and/or non-engineered $\gamma\delta$ T-cells in a $\gamma\delta$ T-cell expansion culture that has been maintained for at least 19
5 contiguous days at a mean rate of about 1 division per about 12 hours (e.g., 10-12 hours), a mean rate of about 1 division per about 13 hours (e.g., 10-13 hours), a mean rate of about 1 division per about 14 hours (e.g., 10-14 hours), a mean rate of about 1 division per about 15 hours (e.g., 10-15 hours), a mean rate of about 1 division per about 16 hours (e.g., 10-16 hours), a mean rate of about 1 division per about 17 hours (e.g., 10-17 hours or 12-17 hours), a mean rate of about 1 division per
10 about 18 hours (e.g., 10-18 hours or 12-18 hours), a mean rate of about 1 division per about 19 hours (e.g., 10-19 hours or 12-19 hours), a mean rate of about 1 division per about 20 hours (e.g., 12-20 hours, 16-20 hours or 18-20 hours), a mean rate of about 1 division per about 21 hours (e.g., 12-21 hours, 16-21 hours or 18-21 hours), a rate of about 1 division per about 22 hours (e.g., 12-22 hours, 16-22 hours or 18-22 hours), a rate of about 1 division per about 23 hours or less (e.g., 12-23 hours, 16-23 hours or 18-23 hours), a mean rate of about 1 division per about 24 hours (e.g., 12-24 hours, 16-24 hours or 18-24 hours).
15

In some cases, an activation agent may stimulate the expansion of engineered and/or non-engineered $\gamma\delta$ T-cells in a $\gamma\delta$ T-cell expansion culture that has been maintained for at least 19
contiguous days at a mean rate of about 1 division per about 25 hours (e.g., 12-25 hours, 16-25 hours
20 18-25 hours, or 20-25 hours), a mean rate of about 1 division per about 26 hours (e.g., 12-26 hours, 16-26 hours 18-26 hours, or 20-26 hours), a mean rate of about 1 division per about 27 hours (e.g., 12-27 hours, 16-27 hours 18-27 hours, or 20-27 hours), a rate of about 1 division per about 28 hours (e.g., 12-28 hours, 16-28 hours 18-28 hours, 20-28 hours, or 22-28 hours), a rate of about 1 division per about 29 hours (e.g., 16-29 hours 18-29 hours, 20-29 hours, or 22-29 hours), a mean rate of
25 about 1 division per about 30 hours (e.g., 16-30 hours 18-30 hours, 20-30 hours, or 22-30 hours), a mean rate of about 1 division per about 31 hours (e.g., 16-31 hours 18-31 hours, 20-31 hours, 22-31 hours, or 24-31 hours), a mean rate of about 1 division per about 32 hours (e.g., 18-32 hours, 20-32 hours, 22-32 hours, or 24-32 hours), a mean rate of about 1 division per about 33 hours (e.g., 18-33 hours, 20-33 hours, 22-33 hours, or 24-33 hours), a rate of about 1 division per about 34 hours (e.g.,
30 18-34 hours, 20-34 hours, 22-34 hours, or 24-34 hours), a rate of about 1 division per about 35 hours

(e.g., 18-35 hours, 20-35 hours, 22-35 hours, or 24-35 hours), a mean rate of about 1 division per about 36 hours (e.g., 18-36 hours, 20-36 hours, 22-36 hours, or 24-36 hours).

An activation agent may stimulate the expansion of sub-populations of engineered or non-engineered $\gamma\delta$ T-cells at different rates of growth. For instance, an agent may stimulate the growth of a $\delta 1$ cell population at a faster rate such that over a period of time from 1 day to 90 days of culture (e.g., about 1 day to about 19, 21, or 23 days of culture) the expansion results in greater than about 10-fold, 100-fold, 200-fold, 300-fold, 400-fold, 500-fold, 600-fold, 700-fold, 800-fold, 900-fold, 1,000-fold, 10,000-fold, 20,000-fold, 30,000-fold, 50,000-fold, 70,000-fold, 100,000-fold or 1,000,000-fold expansion over another $\gamma\delta$ T-cell population, such as a $\delta 2$ or $\delta 3$ population; over a starting number of $\gamma\delta$ T-cells before the expansion; over a starting number of $\gamma\delta 1$ T-cells before the expansion; or over an $\alpha\beta$ T cell population in the culture.

In other cases, the agent may stimulate the growth of a $\delta 1$ and $\delta 4$ population at faster rates such that over a period of time from 1 day to 90 days of culture (e.g., about 1 day to about 19, 21, or 23 days of culture) the expansion results in greater than 10-fold, 100-fold, 200-fold, 300-fold, 400-fold, 500-fold, 600-fold, 700-fold, 800-fold, 900-fold, 1,000-fold, 10,000-fold, 20,000-fold, 30,000-fold, 50,000-fold, 70,000-fold, 100,000-fold or 1,000,000-fold expansion over a $\delta 2$ T-cell population; over another $\gamma\delta$ T-cell sub-population; over a starting number of $\gamma\delta$ T-cells before the expansion; over a starting number of $\gamma\delta 1$ T-cells before the expansion; over a starting number of $\gamma\delta 1$ and $\gamma\delta 3$ T-cells before the expansion; or over an $\alpha\beta$ T cell population in the culture.

In other cases, the agent may stimulate the growth of a $\delta 1$ and $\delta 4$ population at faster rates such that over a period of time from 1 day to 90 days of culture (e.g., about 1 day to about 19, 21, or 23 days of culture) the expansion results in greater than 10-fold, 100-fold, 200-fold, 300-fold, 400-fold, 500-fold, 600-fold, 700-fold, 800-fold, 900-fold, 1,000-fold, 10,000-fold, 20,000-fold, 30,000-fold, 50,000-fold, 70,000-fold, 100,000-fold or 1,000,000-fold expansion over a $\delta 2$ T-cell population; over another $\gamma\delta$ T-cell sub-population; over a starting number of $\gamma\delta$ T-cells before the expansion; over a starting number of $\gamma\delta 1$ T-cells before the expansion; over a starting number of $\gamma\delta 1$ and $\gamma\delta 4$ T-cells before the expansion; or over an $\alpha\beta$ T cell population in the culture.

In other cases, the agent may stimulate the growth of a $\delta 1$, $\delta 3$, $\delta 4$ and $\delta 5$ population at faster rates such that over a period of time from 1 day to 90 days of culture (e.g., about 1 day to about 19, 21, or 23 days of culture) the expansion results in greater than 10-fold, 100-fold, 200-fold, 300-fold,

400-fold, 500-fold, 600-fold, 700-fold, 800-fold, 900-fold, 1,000-fold, 10,000-fold, 20,000-fold, 30,000-fold, 50,000-fold, 70,000-fold, 100,000-fold or 1,000,000-fold expansion over a $\delta 2$ T-cell population; over another $\gamma\delta$ T-cell sub-population; over a starting number of $\gamma\delta$ T-cells before the expansion; over a starting number of $\gamma\delta 1$ T-cells before the expansion; over a starting number of $\gamma\delta 1$ and $\gamma\delta 3$ T-cells before the expansion; over a starting number of $\gamma\delta 1$, $\gamma\delta 3$, $\gamma\delta 4$, and $\gamma\delta 5$ T-cells before the expansion; or over an $\alpha\beta$ T cell population in the culture.

In other cases, the agent may stimulate the growth of a $\delta 2$ population at faster rates such that over a period of time from 1 day to 90 days of culture (*e.g.*, about 1 day to about 19, 21, or 23 days of culture) the expansion results in greater than 10-fold, 100-fold, 200-fold, 300-fold, 400-fold, 500-fold, 600-fold, 700-fold, 800-fold, 900-fold, 1,000-fold, 10,000-fold, 20,000-fold, 30,000-fold, 50,000-fold, 70,000-fold, 100,000-fold or 1,000,000-fold expansion over a $\delta 1$ T-cell population; over a $\delta 3$ T-cell population; over another $\gamma\delta$ T-cell sub-population; over a starting number of $\gamma\delta$ T-cells before the expansion, over a starting number of $\gamma\delta 2$ T-cells before the expansion, or over $\alpha\beta$ T-cells.

In some aspects, the disclosure provides an engineered or non-engineered $\gamma\delta$ T-cell population, in contact with an agent that stimulates an expansion of the $\gamma\delta$ T-cell population at a rapid rate, such as a rate of about 1 cell division per 30 hours or faster. In some cases, the agent selectively stimulates the proliferation of either $\delta 1$; $\delta 2$; $\delta 1$ and $\delta 4$; or $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ T-cells. A $\gamma\delta$ T-cell population can comprise an amount of non-engineered $\gamma\delta$ T-cells and an amount of engineered $\gamma\delta$ T-cells. In some cases, the $\gamma\delta$ T-cell population comprises different percentages of $\delta 1$, $\delta 2$, $\delta 3$, and $\delta 4$ T-cells. An engineered or non-engineered $\gamma\delta$ T-cell population can comprise, for example, fewer than 90% $\delta 1$ T-cells, fewer than 80% $\delta 1$ T-cells, fewer than 70% $\delta 1$ T-cells, fewer than 60% $\delta 1$ T-cells, fewer than 50% $\delta 1$ T-cells, fewer than 40% $\delta 1$ T-cells, fewer than 30% $\delta 1$ T-cells, fewer than 20% $\delta 1$ T-cells, fewer than 10% $\delta 1$ T-cells, or fewer than 5% $\delta 1$ T-cells.

Alternatively, an engineered or non-engineered $\gamma\delta$ T-cell population can comprise greater than 5% $\delta 1$ T-cells, greater than 10% $\delta 1$ T-cells, greater than 20% $\delta 1$ T-cells, greater than 30% $\delta 1$ T-cells, greater than 40% $\delta 1$ T-cells, greater than 50% $\delta 1$ T-cells, greater than 60% $\delta 1$ T-cells, greater than 70% $\delta 1$ T-cells, greater than 80% $\delta 1$ T-cells, or greater than 90% $\delta 1$ T-cells. In some cases, the agent is one of the selective expansion agents described herein. In some cases, the agent is

immobilized on a surface such as a cell culture surface, or a surface of an APC (*e.g.*, expressed on the surface of the APC or bound to an Fc receptor expressed on the surface of the APC).

An engineered or non-engineered $\gamma\delta$ T-cell population can comprise, for example, fewer than 90% $\delta 2$ T-cells, fewer than 80% $\delta 2$ T-cells, fewer than 70% $\delta 2$ T-cells, fewer than 60% $\delta 2$ T-cells, fewer than 50% $\delta 2$ T-cells, fewer than 40% $\delta 2$ T-cells, fewer than 30% $\delta 2$ T-cells, fewer than 20% $\delta 2$ T-cells, fewer than 10% $\delta 2$ T-cells, or fewer than 5% $\delta 2$ T-cells. Alternatively, an engineered or non-engineered $\gamma\delta$ T-cell population can comprise greater than 5% $\delta 2$ T-cells, greater than 10% $\delta 2$ T-cells, greater than 20% $\delta 2$ T-cells, greater than 30% $\delta 2$ T-cells, greater than 40% $\delta 2$ T-cells, greater than 50% $\delta 2$ T-cells, greater than 60% $\delta 2$ T-cells, greater than 70% $\delta 2$ T-cells, greater than 80% $\delta 2$ T-cells, or greater than 90% $\delta 2$ T-cells.

An engineered or non-engineered $\gamma\delta$ T-cell population can comprise, for example, fewer than 90% $\delta 1$ and $\delta 4$ T-cells, fewer than 80% $\delta 1$ and $\delta 4$ T-cells, fewer than 70% $\delta 1$ and $\delta 4$ T-cells, fewer than 60% $\delta 1$ and $\delta 4$ T-cells, fewer than 50% $\delta 1$ and $\delta 4$ T-cells, fewer than 40% $\delta 1$ and $\delta 4$ T-cells, fewer than 30% $\delta 1$ and $\delta 4$ T-cells, fewer than 20% $\delta 1$ and $\delta 4$ T-cells, fewer than 10% $\delta 1$ and $\delta 4$ T-cells, or fewer than 5% $\delta 1$ and $\delta 4$ T-cells. Alternatively, an engineered or non-engineered $\gamma\delta$ T-cell population can comprise greater than 5% $\delta 1$ and $\delta 4$ T-cells, greater than 10% $\delta 1$ and $\delta 4$ T-cells, greater than 20% $\delta 1$ and $\delta 4$ T-cells, greater than 30% $\delta 1$ and $\delta 4$ T-cells, greater than 40% $\delta 1$ and $\delta 4$ T-cells, greater than 50% $\delta 1$ and $\delta 4$ T-cells, greater than 60% $\delta 1$ and $\delta 4$ T-cells, greater than 70% $\delta 1$ and $\delta 4$ T-cells, greater than 80% $\delta 1$ and $\delta 4$ T-cells, or greater than 90% $\delta 1$ and $\delta 4$ T-cells.

An engineered or non-engineered $\gamma\delta$ T-cell population can comprise, for example, fewer than 90% $\delta 4$ T-cells, fewer than 80% $\delta 4$ T-cells, fewer than 70% $\delta 4$ T-cells, fewer than 60% $\delta 4$ T-cells, fewer than 50% $\delta 4$ T-cells, fewer than 40% $\delta 4$ T-cells, fewer than 30% $\delta 4$ T-cells, fewer than 20% $\delta 4$ T-cells, fewer than 10% $\delta 4$ T-cells, or fewer than 5% $\delta 4$ T-cells. Alternatively, an engineered or non-engineered $\gamma\delta$ T-cell population can comprise greater than 5% $\delta 1$ and $\delta 4$ T-cells, greater than 10% $\delta 1$ and $\delta 4$ T-cells, greater than 20% $\delta 1$ and $\delta 4$ T-cells, greater than 30% $\delta 1$ and $\delta 4$ T-cells, greater than 40% $\delta 1$ and $\delta 4$ T-cells, greater than 50% $\delta 1$ and $\delta 4$ T-cells, greater than 60% $\delta 1$ and $\delta 4$ T-cells, greater than 70% $\delta 1$ and $\delta 4$ T-cells, greater than 80% $\delta 1$ and $\delta 4$ T-cells, or greater than 90% $\delta 1$ and $\delta 4$ T-cells. An engineered or non-engineered $\gamma\delta$ T-cell population can comprise, for example, fewer than 90% $\delta 1$ and $\delta 4$ T-cells, fewer than 80% $\delta 1$ and $\delta 4$ T-cells, fewer than 70% $\delta 1$ and $\delta 4$ T-

cells, fewer than 60% $\delta 1$ and $\delta 4$ T-cells, fewer than 50% $\delta 1$ and $\delta 4$ T-cells, fewer than 40% $\delta 1$ and $\delta 4$ T-cells, fewer than 30% $\delta 1$ and $\delta 4$ T-cells, fewer than 20% $\delta 1$ and $\delta 4$ T-cells, fewer than 10% $\delta 1$ and $\delta 4$ T-cells, or fewer than 5% $\delta 1$ and $\delta 4$ T-cells.

In certain embodiments, the present invention provides admixtures of expanded $\gamma\delta$ T-cell populations comprising 10-90% $\delta 1$ T-cells and 90-10% $\delta 2$ T-cells. In certain embodiments, the present invention provides admixtures of expanded $\gamma\delta$ T-cell populations comprising 10-90% $\delta 1$ and $\delta 3$ T-cells and 90-10% $\delta 2$ T-cells. In certain embodiments, the present invention provides admixtures of expanded $\gamma\delta$ T-cell populations comprising 10-90% $\delta 1$ and $\delta 4$ T-cells and 90-10% $\delta 2$ T-cells. In certain embodiments, the present invention provides admixtures of expanded $\gamma\delta$ T-cell populations comprising 10-90% $\delta 1$, $\delta 3$, $\delta 4$ and $\delta 5$ T-cells and 90-10% $\delta 2$ T-cells.

One or more activation agent can contact the $\gamma\delta$ T-cells (for example an activator $\gamma\delta$ T cell innate receptor) and thereafter a costimulatory molecule can contact the $\gamma\delta$ T-cells to provide further stimulation and to expand the $\gamma\delta$ T-cells. In some embodiments, the activation agent and/or costimulatory agent can be lectins of plant and non-plant origin, monoclonal antibodies that activate $\gamma\delta$ T-cells, and other non-lectin/ non- antibody agents. In other cases, the plant lectin can be concanavalin A (ConA) although other plant lectins such as may be used. Other examples of lectins include protein peanut agglutinin (PNA), soybean agglutinin (SBA), les culinaris agglutinin (LCA), pisum sativum agglutinin (PSA), Helix pomatia agglutinin (HPA), Vicia graminea Lectin (VGA), Phaseolus Vulgaris Erythroagglutinin (PHA-E), Phaseolus Vulgaris Leucoagglutinin (PHA-L), Sambucus Nigra Lectin (SNA, EBL), Maackia Amurensis, Lectin II (MAL II), Sophora Japonica Agglutinin (SJA), Dolichos Biflorus Agglutinin (DBA), Lens Culinaris Agglutinin (LCA), Wisteria Floribunda Lectin (WFA, WFL).

Non-limiting examples of activating agents and costimulatory molecules include any one or more antibodies selective for a δ or γ -chain or subtypes thereof described herein, antibodies such as 5A6.E9, B1, TS8.2, 15D, B6, B3, TS-1, $\gamma 3.20$, 7A5, IMMU510, R9.12, 11F2, or a combination thereof. Other examples of activating agents and costimulatory molecules include zoledronate, phorbol 12-myristate-13-acetate (TPA), mezerein, staphylococcal enterotoxin A (SEA), streptococcal protein A, or a combination thereof.

In other cases, the activation agent and/or costimulatory agent can be, antibodies or ligands to α TCR, β TCR, γ TCR, δ TCR, CD277, CD28, CD46, CD81, CTLA4, ICOS, PD-1, CD30, NKG2D,

NKG2A, HVEM, 4-1 BB (CD137), OX40 (CD134), CD70, CD80, CD86, DAP, CD122, GITR, FcεRIγ, CD1, CD16, CD161, DNAX, accessory molecule-1 (DNAM-1), one or more NCRs (*e.g.*, Nkp30, Nkp44, Nkp46), SLAM, Coxsackie virus and adenovirus receptor or a combination thereof.

Engineered $\gamma\delta$ T cells

Engineered $\gamma\delta$ T-cells (see, *e.g.*, Fig. 1) may be generated with various methods known in the art. An engineered $\gamma\delta$ T-cell may be designed to stably express a particular tumor recognition moiety. A polynucleotide encoding an expression cassette that comprises a tumor recognition, or another type of recognition moiety, can be stably introduced into the $\gamma\delta$ T-cell by a

transposon/transposase system or a viral-based gene transfer system, such as a lentiviral or a retroviral system, or another suitable method, such as transfection, electroporation, transduction, lipofection, calcium phosphate (CaPO₄), nanoengineered substances, such as Ormosil, viral delivery methods, including adenoviruses, retroviruses, lentiviruses, adeno-associated viruses, or another suitable method. An antigen specific TCR, either $\alpha\beta$ or $\gamma\delta$, can be introduced into the engineered $\gamma\delta$

T-cell by stably inserting a polynucleotide comprising a genetic code for the antigen specific TCR into the genome of the $\gamma\delta$ T-cell. A polynucleotide encoding a CAR with a tumor recognition moiety may be introduced into the engineered $\gamma\delta$ T-cell by stably inserting the polynucleotide into the genome of the $\gamma\delta$ T-cell. In some cases, the engineered tumor recognition moiety is an engineered T-cell receptor, and the expression cassette incorporated into the genome of an

engineered $\gamma\delta$ T-cell comprises a polynucleotide encoding an engineered TCR α (TCR alpha) gene, an engineered TCR β (TCR beta) gene, an TCR δ (TCR delta) gene, or an engineered TCR γ (TCR gamma) gene. In some cases, the expression cassette incorporated into the genome of the

engineered $\gamma\delta$ T-cell comprises a polynucleotide encoding an antibody fragment or an antigen binding portion thereof. In some cases, the antibody fragment or antigen binding fragment thereof is

a polynucleotide encoding a whole antibody, an antibody fragment, a single-chain variable fragment (scFv), a single domain antibody (sdAb), a Fab, F(ab)₂, an Fc, the light or heavy chains on an antibody, the variable or the constant region of an antibody, or any combination thereof that binds to a cell surface tumor antigen as part of the Chimeric Antigen Receptor (CAR) construct, or a bi-specific construct, comprising a CAR and a T-cell receptor (TCR), or CARs with antibodies directed to different antigens. In some cases, the polynucleotide is derived from a human or from another

species. An antibody fragment or antigen binding fragment polynucleotide that is derived from a non-human species can be modified to increase their similarity to antibody variants produced naturally in humans, and an antibody fragment or antigen binding fragment can be partially or fully humanized. An antibody fragment or antigen binding fragment polynucleotide can also be chimeric,
5 for example a mouse-human antibody chimera. An engineered $\gamma\delta$ T-cell that expresses a CAR can also be engineered to express a ligand to the antigen recognized by the tumor recognition moiety.

Various techniques known in the art can be used to introduce a cloned, or synthetically engineered, nucleic acid comprising the genetic code for a tumor recognition moiety into a specific location within the genome of an engineered $\gamma\delta$ T-cell. The RNA-guided Cas9 nuclease from the
10 microbial clustered regularly interspaced short palindromic repeats (CRISPR) system, zinc finger nucleases (ZFNs), transcription activator-like effector nucleases (TALENs), and meganuclease technologies, as described, respectively by WO201409370, WO2003087341, WO2014134412, and WO2011090804, each of which is incorporated by reference herein in its entirety, can be used to provide efficient genome engineering in $\gamma\delta$ T-cell(s). The technologies described herein can also be
15 used to insert the expression cassette into a genomic location that simultaneously provides a knock-out of one gene and a knock-in of another gene. For example, a polynucleotide comprising an expression cassette of the disclosure can be inserted into a genomic region that encodes for an MHC gene. Such engineering can simultaneously provide the knock-in of one or more genes, e.g. the genes comprised in the expression cassette, and a knock-out of another gene, e.g. an MHC locus.

20 In one case, a Sleeping Beauty transposon that includes a nucleic acid coding for the tumor recognition moiety is introduced into the cell $\gamma\delta$ T-cell that is being engineered. A mutant Sleeping Beauty transposase that provides for enhanced integration as compared to the wild-type Sleeping Beauty, such as the transposase described in US 7,985,739, which is incorporated by reference herein in its entirety, may be used to introduce a polynucleotide in the engineered $\gamma\delta$ T-cell.

25 In some cases, a viral method is used to introduce a polynucleotide comprising a tumor recognition moiety into the genome of an engineered $\gamma\delta$ T-cell. A number of viral methods have been used for human gene therapy, such as the methods described in WO 1993020221, which is incorporated herein in its entirety. Non-limiting examples of viral methods that can be used to engineer a $\gamma\delta$ T-cell include retroviral, adenoviral, lentiviral, herpes simplex virus, vaccinia virus,
30 pox virus, or adeno-virus associated viral methods.

A polynucleotide containing the genetic code for a tumor recognition moiety may comprise mutations or other transgenes that affect the growth, proliferation, activation status of the engineered $\gamma\delta$ T-cell or an antigen specific to tumor cells such as testis-specific cancer antigens. A $\gamma\delta$ T-cell of the disclosure may be engineered to express a polynucleotide comprising an activation domain that is linked to the antigen recognition moiety, such as a molecule in TCR-CD3 complex or a co-stimulatory factor. An engineered $\gamma\delta$ T-cell can express an intracellular signaling domain that is a T-lymphocyte activation domain. The $\gamma\delta$ T-cell may be engineered to express an intracellular activation domain gene or an intracellular signaling domain. The intracellular signaling domain gene, may be, for example CD3 ζ , CD28, CD2, ICOS, JAML, CD27, CD30, OX40, NKG2D, CD4, OX40/CD134, 4-1BB/CD137, Fc ϵ RI γ , IL-2RB/CD 122, IL- 2RG/CD132, DAP molecules, CD70, cytokine receptor, CD40, or any combination thereof. In some cases, the engineered $\gamma\delta$ T-cell is also engineered to express a cytokine, an antigen, a cellular receptor, or other immunomodulatory molecule.

The appropriate tumor recognition moiety to be expressed by the engineered $\gamma\delta$ T-cell can be selected based on the disease to be treated. For example, in some cases a tumor recognition moiety is a TCR. In some cases, a tumor recognition moiety is a receptor to a ligand that is expressed on a cancer cell. Non-limiting examples of suitable receptors include NKG2D, NKG2A, NKG2C, NKG2F, LIT1, AICL, CD26, NKRP1, CD244 (2B4), DNAM-1, NKp30, NKp44, NKp46, and NKp80. In some cases, a tumor recognition moiety can include a ligand, *e.g.* IL-13 ligand, or a ligand mimetic to the tumor antigen, such as the IL-13 mimetic to IL13R.

A $\gamma\delta$ T-cell may be engineered to express a chimeric tumor recognition moiety comprising a ligand binding domain derived from NKG2D, NKG2A, NKG2C, NKG2F, LIT1, AICL, CD26, NKRP1, CD244 (2B4), DNAM-1, or an anti-tumor antibody such as anti-Her2neu or anti-EGFR and a signaling domain obtained from CD3- ζ , Dap 10, Dap 12, CD28, 41BB, and CD40L. In some examples, the chimeric receptor binds MICA, MICB, Her2neu, EGFR, EGFRvIII, mesothelin, CD38, CD20, CD19, BCMA, PSA, RON, CD30, CD22, CD37, CD38, CD56, CD33, CD138, CD123, CD79b, CD70, CD75, CA6, GD2, alphafetoprotein (AFP), CS1, carcinoembryonic antigen (CEA), CEACAM5, CA-125, MUC-16, 5T4, NaPi2b, ROR1, ROR2, PLIF, Her2/Neu, EGFRvIII, GPMNB, LIV-1, glycolipidF77, fibroblast activation protein (FAP), PSMA, STEAP-1, STEAP-2, c-Met, CSPG4, CD44v6, PVRL-4, VEGFR2, C4.4a, PSCA, folate binding protein/receptor, SLC44A4,

Cripto, CTAG1B, AXL, IL-13R α 2, IL-3R, EPHA3, SLTRK6, gp100, MART1, Tyrosinase, SSX2, SSX4, NYESO-1, epithelial tumor antigen (ETA), MAGEA family genes (such as MAGEA3, MAGEA4), KKL1, mutated ras (H, N, K), BRAf, p53, β -catenin, EGFR790, MHC class I chain-related molecule A (MICA), or MHC class I chain-related molecule B (MICB), or one or more
 5 antigens of HPV, CMV, or EBV.

In some cases, the tumor recognition moiety targets an MHC class I molecule (HLA-A, HLA-B, or HLA-C) in complex with a tumor-associated peptide. Methods and compositions for generating and using tumor recognition moieties that target a tumor-associated peptide in complex with a MHC class I molecule include those described in Weidanz *et al.*, Int. Rev. Immunol. 30:328-
 10 40, 2011; Scheinberg et al, Oncotarget. 4(5):647-8, 2013; Cheever et al, Clin. Cancer Res. 15(17):5323-37, 2009; Dohan & Reiter Expert Rev Mol Med. 14:e6, 2012; Dao *et al.*, Sci Transl Med. 2013 Mar 13;5(176):176ra33; U.S. 9,540,448; and WO 2017/011804. In some embodiments, the targeted tumor-associated peptide of the peptide MHC complex is a peptide of Wilms' tumor protein 1 (WT1), human telomerase reverse transcriptase (hTERT), survivin, mouse double minute 2
 15 homolog (MDM2), cytochrome P450 (CYP1B), KRAS, or BRAF.

Two or more tumor recognition moieties may be expressed in the $\gamma\delta$ T-cell from genetically different, substantially different, or substantially identical, $\alpha\beta$ TCR polynucleotides stably expressed from the engineered $\gamma\delta$ T-cell or from genetically distinct $\alpha\beta$ TCR polynucleotides stably incorporated in the engineered $\gamma\delta$ T-cell. In the case of genetically distinct $\alpha\beta$ TCR(s), $\alpha\beta$ TCR(s)
 20 recognizing different antigens associated with the same condition may be utilized. In one preferred embodiment, a $\gamma\delta$ T-cell is engineered to express different TCRs, from human or mouse origin, from one or more expression cassettes that recognize the same antigen in the context of different MHC haplotypes. In another preferred embodiment, a $\gamma\delta$ T-cell is engineered to express one TCR and two or more antibodies directed to the same or different peptides from a given antigen complexed with
 25 different MHC haplotypes. In some cases, expression of a single TCR by an engineered $\gamma\delta$ T-cell facilitates proper TCR pairing. An engineered $\gamma\delta$ T-cell that expresses different TCRs can provide a universal allogeneic engineered $\gamma\delta$ T-cell. In a second preferred embodiment, a $\gamma\delta$ T-cell is engineered to express one or more different antibodies directed to peptide-MHC complexes, each directed to the same or different peptide complexed with the same or different MHC haplotypes. In
 30 some cases, a tumor recognition moiety can be an antibody that binds to peptide-MHC complexes.

A $\gamma\delta$ T-cell can be engineered to express TCRs from one or more expression cassettes that recognize the same antigen in the context of different MHC haplotypes. In some cases, an engineered $\gamma\delta$ T-cell is designed to express a single TCR, or a TCR in combination with a CAR to minimize the likelihood of TCR mispairing within the engineered cell. The tumor recognition
5 moieties expressed from two or more expression cassettes preferably have different polynucleotide sequences, and encode tumor recognition moieties that recognize different epitopes of the same target, *e.g.*, in the context of different HLA haplotypes. An engineered $\gamma\delta$ T-cell that expresses such different TCRs or CARs can provide a universal allogeneic engineered $\gamma\delta$ T-cell.

In some cases, a $\gamma\delta$ T-cell is engineered to express one or more tumor recognition moieties.
10 Two or more tumor recognition moieties may be expressed from genetically identical, or substantially identical, antigen-specific chimeric (CAR) polynucleotides engineered in the $\gamma\delta$ T-cell. Two or more tumor recognition moieties may be expressed from genetically distinct CAR polynucleotides engineered in the $\gamma\delta$ T-cell. The genetically distinct CAR(s) may be designed to recognize different antigens associated with the same condition.

15 A $\gamma\delta$ T-cell may alternatively be bi-specific. A bi-specific engineered $\gamma\delta$ T-cell can express two or more tumor recognition moieties. A bi-specific engineered $\gamma\delta$ T-cell can express both TCR and CAR tumor recognition moieties. A bi-specific engineered $\gamma\delta$ T-cell can be designed to recognize different antigens associated with the same condition. An engineered $\gamma\delta$ T-cell can express two or more CAR/TCR(s) bi-specific polynucleotides that recognize an identical or
20 substantially identical antigen. An engineered $\gamma\delta$ T-cell can express two or more CAR/TCR(s) bi-specific constructs that recognize distinct antigens. In some cases, a bi-specific construct of the disclosure binds to an activating and an inactivating domain of a target cell, thereby providing increased target specificity. The $\gamma\delta$ T-cell may be engineered to express at least 1 tumor recognition moiety, at least 2 tumor recognition moieties, at least 3 tumor recognition moieties, at least 4 tumor
25 recognition moieties, at least 5 tumor recognition moieties, at least 6 tumor recognition moieties, at least 7 tumor recognition moieties, at least 8 tumor recognition moieties, at least 9 tumor recognition moieties, at least 10 tumor recognition moieties, at least 11 tumor recognition moieties, at least 12 tumor recognition moieties, or another suitable number of tumor recognition moieties.

Proper TCR function may be enhanced by two functioning ζ (zeta) proteins comprising
30 ITAM motifs. Proper TCR function may also be enhanced by expression of $\alpha\beta$ or $\gamma\delta$ activation

domains, such as CD3 ζ , CD28, CD2, CTLA4, ICOS, JAML, PD-1, CD27, CD30, 41-BB, OX40, NKG2D, HVEM, CD46, CD4, Fc ϵ RI γ , IL-2RB/CD122, IL-2RG/CD132, DAP molecules, and CD70. The expressed polynucleotide may include the genetic code for a tumor recognition moiety, a linker moiety, and an activation domain. Translation of the polynucleotide by the engineered $\gamma\delta$ T-cell
5 may provide a tumor recognition moiety and an activation domain linked by a protein linker. Often, the linker comprises amino acids that do not obstruct the folding of the tumor recognition moiety and the activation domain. A linker molecule can be at least about 5 amino acids, about 6 amino acids, about 7 amino acids, about 8 amino acids, about 9 amino acids, about 10 amino acids, about 11
10 amino acids, about 12 amino acids, about 13 amino acids, about 14 amino acids, about 15 amino acids, about 16 amino acids, about 17 amino acids, about 18 amino acids, about 19 amino acids, or about 20 amino acids in length. In some cases, at least 50%, at least 70% or at least 90% of the amino acids in the linker are serine or glycine.

In some cases, an activation domain can comprise one or more mutations. Suitable mutations may be, for example, mutations that render an activation domain constitutively active. Altering the
15 identity of one or more nucleic acids changes the amino acid sequence of the translated amino acid. A nucleic acid mutation can be made such that the encoded amino acid is modified to a polar, non-polar, basic or acidic amino acid. A nucleic acid mutation can be made such that the tumor recognition moiety is optimized to recognize an epitope from a tumor. The engineered tumor recognition moiety, an engineered activation domain, or another engineered component of a $\gamma\delta$ T-
20 cell may include more than 1 amino acid mutation, 2 amino acid mutations, 3 amino acid mutations, 4 amino acid mutations, 5 amino acid mutations, 6 amino acid mutations, 7 amino acid mutations, 8 amino acid mutations, 9 amino acid mutations, 10 amino acid mutations, 11 amino acid mutations, 12 amino acid mutations, 13 amino acid mutations, 14 amino acid mutations, 15 amino acid
25 acid mutations, 16 amino acid mutations, 17 amino acid mutations, 18 amino acid mutations, 19 amino acid mutations, 20 amino acid mutations, 21 amino acid mutations, 22 amino acid mutations, 23 amino acid mutations, 24 amino acid mutations, 25 amino acid mutations, 26 amino acid mutations, 27 amino acid mutations, 28 amino acid mutations, 29 amino acid mutations, 30 amino acid
30 amino acid mutations, 31 amino acid mutations, 32 amino acid mutations, 33 amino acid mutations, 34 amino acid mutations, 35 amino acid mutations, 36 amino acid mutations, 37 amino acid mutations, 38 amino acid mutations, 39 amino acid mutations, 40 amino acid mutations, 41 amino acid mutations,

42 amino acid mutations, 43 amino acid mutations, 44 amino acid mutations, 45 amino acid mutations, 46 amino acid mutations, 47 amino acid mutations, 48 amino acid mutations, 49 amino acid mutations, or 50 amino acid mutations.

In some cases, a $\gamma\delta$ T-cell of the disclosure does not express one or more MHC molecules.

5 Deletion of one or more MHC loci in an engineered $\gamma\delta$ T-cell can decrease the likelihood that the engineered $\gamma\delta$ T-cell will be recognized by the host immune system. The human Major Histocompatibility Complex (MHC) loci, known as the human leukocyte antigen (HLA) system, comprises a large gene family that is expressed in antigen presenting cells, including $\gamma\delta$ T-cells. The HLA-A, HLA-B, and HLA-C molecules function to present intracellular peptides as antigens to
10 antigen presenting cells. The HLA-DP, HLA-DM, HLA-DOA, HLA-DOB, HLA-DQ, and HLA-DR molecules function to present extracellular peptides as antigens to antigen presenting cells. Some alleles of the HLA genes have been associated with GVHD, autoimmune disorders, and cancer. An engineered $\gamma\delta$ T-cell described herein can be further engineered to lack, or to disrupt gene expression of one or more HLA genes. An engineered $\gamma\delta$ T-cell described herein can be further
15 engineered to lack, or to disrupt gene expression of one or more components of the MHC complex, such as complete deletion of one or more of the MHC genes, deletion of specific exons, or deletion of the β_2 microglobulin (B2m). Genetic excision or genetic disruption of at least one HLA gene can provides a clinically therapeutic $\gamma\delta$ T-cell that can be administered to a subject with any HLA haplotype without causing host-versus-graft disease. An engineered $\gamma\delta$ T-cell as described herein
20 can be a universal donor for a human subject with any HLA haplotype.

A $\gamma\delta$ T-cell can be engineered to lack one or various HLA locus (loci). An engineered $\gamma\delta$ T-cell can be engineered to lack an HLA-A allele, an HLA-B allele, an HLA-C allele, an HLA-DR allele, an HLA-DQ allele, or an HLA-DP allele. In some cases, an HLA allele is associated with a human condition, such as an auto-immune condition. For instance, the HLA-B27 allele has been
25 associated with arthritis and uveitis, the HLA-DR2 allele has been associated with systemic lupus erythematosus, and multiple sclerosis, the HLA-DR3 allele has been associated with 21-hydroxylase deficiency, the HLA-DR4 has been associated with rheumatoid arthritis and type 1 diabetes. An engineered $\gamma\delta$ T-cell that lacks, for example, the HLA-B27 allele can be administered to a subject afflicted with arthritis without being readily recognized the immune system of the subject. In some
30 cases, deletion of one or more HLA loci provides an engineered $\gamma\delta$ T-cell that is a universal donor

for any subject with any HLA haplotype.

In some cases, engineering a $\gamma\delta$ T-cell requires the deletion of a portion of the $\gamma\delta$ T-cell genome. In some cases, the deleted portion of the genome comprises a portion of the MHC locus (loci). In some instances, the engineered $\gamma\delta$ T-cell is derived from a wild-type human $\gamma\delta$ T-cell, and the MHC locus is an HLA locus. In some cases, the deleted a portion of the genome comprises a portion of a gene corresponding to a protein in the MHC complex. In some cases, the deleted portion of the genome comprises the $\beta 2$ microglobulin gene. In some instances, the deleted portion of the genome comprises an immune checkpoint gene, such as PD-1, CTLA-4, LAG3, ICOS, BTLA, KIR, TIM3, A2aR, B7-H3, B7-H4, and CECAM-1. In some cases, an engineered $\gamma\delta$ T-cell can be designed to express an activation domain that enhances T-cell activation and cytotoxicity. Non-limiting examples of activation domains that can be expressed by an engineered $\gamma\delta$ T-cell include: CD2, ICOS, 4-1 BB (CD137), OX40 (CD134), CD27, CD70, CD80, CD86, DAP molecules, CD122, GITR, Fc ϵ RI γ .

Any portion of the genome of an engineered $\gamma\delta$ T-cell can be deleted to disrupt the expression of an endogenous $\gamma\delta$ T-cell gene. Non-limiting examples of genomic regions that can be deleted or disrupted in the genome of an $\gamma\delta$ T-cell include a promoter, an activator, an enhancer, an exon, an intron, a non-coding RNA, a micro-RNA, a small-nuclear RNA, variable number tandem repeats (VNTRs), short tandem repeat (STRs), SNP patterns, hypervariable regions, minisatellites, dinucleotide repeats, trinucleotide repeats, tetranucleotide repeats, or simple sequence repeats. In some cases, the deleted a portion of the genome ranges between 1 nucleic acid to about 10 nucleic acids, 1 nucleic acid to about 100 nucleic acids, 1 nucleic acid to about 1,000 nucleic acids, 1 nucleic acid to about 10,000 nucleic acids, 1 nucleic acid to about 100,000 nucleic acids, 1 nucleic acid to about 1,000,000 nucleic acids, or other suitable range.

HLA gene expression in an engineered $\gamma\delta$ T-cell can also be disrupted with various techniques known in the art. In some cases, large loci gene editing technologies are used to excise a gene from the engineered $\gamma\delta$ T-cell genome, or to disrupt gene expression of at least one HLA locus in the engineered $\gamma\delta$ T-cell. Non-limiting examples of gene editing technologies that can be used to edit a desired locus on a genome of an engineered $\gamma\delta$ T-cell include Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR)-Cas, zinc finger nucleases (ZFNs), Transcription activator-like effector nucleases (TALENs), and meganuclease technologies, as described, respectively by

WO201409370, WO2003087341, WO2014134412, and WO 2011090804, and each of which is incorporated by reference herein in its entirety.

A $\gamma\delta$ T-cell may be engineered from an isolated non-engineered $\gamma\delta$ T-cell that already expresses a tumor recognition moiety. The engineered $\gamma\delta$ T-cell can retain a tumor cell recognition moiety that is endogenously expressed by the isolated wild-type $\gamma\delta$ T-cell, *e.g.*, isolated from tumor infiltrating lymphocytes of a tumor sample. In some cases, the engineered $\gamma\delta$ T-cell tumor cell recognition moiety replaces the wild-type $\gamma\delta$ TCR.

A $\gamma\delta$ T-cell can be engineered to express one or more homing molecules, such as a lymphocyte homing molecule. Homing molecules can be, for instance, lymphocyte homing receptors or cell adhesion molecules. A homing molecule can help an engineered $\gamma\delta$ T-cell to migrate and infiltrate a solid tumor, including a targeted solid tumor upon administration of the engineered $\gamma\delta$ T-cell to the subject. Non-limiting examples of homing receptors include members of the CCR family, *e.g.*: CCR2, CCR4, CCR7, CCR8, CCR9, CCR10, CLA, CD44, CD103, CD62L, E-selectin, P-selectin, L-selectin, integrins, such as VLA-4 and LFA-1. Non-limiting examples of cell adhesion molecules include ICAM, N-CAM, VCAM, PE-CAM, L1-CAM, Nectins (PVRL1, PVRL2, PVRL3), LFA-1, integrin $\alpha\text{X}\beta\text{2}$, $\alpha\text{v}\beta\text{7}$, macrophage-1 antigen, CLA-4, glycoprotein IIb/IIIa. Additional examples of cell adhesion molecules include calcium dependent molecules, such as T-cadherin, and antibodies to matrix metalloproteinases (MMPs) such as MMP9 or MMP2.

The steps involved in T-cell maturation, activation, proliferation, and function may be regulated through co-stimulatory and inhibitory signals through immune checkpoint proteins. Immune checkpoints are co-stimulatory and inhibitory elements intrinsic to the immune system. Immune checkpoints aid in maintaining self-tolerance and modulating the duration and amplitude of physiological immune responses to prevent injury to tissues when the immune system responds to disease conditions, such as cell transformation or infection. The equilibrium between the co-stimulatory and inhibitory signals used to control the immune response from either $\gamma\delta$ and $\alpha\beta$ T-cells can be modulated by immune checkpoint proteins. Immune checkpoint proteins, such as PD1 and CTLA4 are present on the surface of T-cells and can be used to turn an immune response “on” or “off.” Tumors can dysregulate checkpoint protein function as an immune-resistance mechanism, particularly against T-cells that are specific for tumor antigens. An engineered $\gamma\delta$ T-cell of the disclosure can be further engineered to lack one or more immune checkpoint locus (loci), such as

PD-1, CTLA-4, LAG3, ICOS, BTLA, KIR, TIM3, A2aR, CEACAM1, B7-H3, and B7-H4.

Alternatively, the expression of an endogenous immune check point gene in an engineered $\gamma\delta$ T-cell of the disclosure can be disrupted with gene editing technologies.

Immunological checkpoints can be molecules that regulate inhibitory signaling pathways (exemplified by CTLA4, PD1, and LAG3) or molecules that regulate stimulatory signaling pathways (exemplified by ICOS) in an engineered $\gamma\delta$ T-cell of the disclosure. Several proteins in the extended immunoglobulin superfamily can be ligands for immunological checkpoints. Non-limiting examples of immune checkpoint ligand proteins include B7-H4, ICOSL, PD-L1, PD-L2, MegaCD40L, MegaOX40L, and CD137L. In some cases, immune checkpoint ligand proteins are antigens expressed by a tumor. In some cases, the immune checkpoint gene is a CTLA-4 gene. In some cases, the immune checkpoint gene is a PD-1 gene.

PD1 is an inhibitory receptor belonging to the CD28/CTLA4 family and is expressed on activated T lymphocytes, B cells, monocytes, DCs, and T-regs. There are two known ligands for PD1, PD-L1 and PD-L2, which are expressed on T cells, APCs, and malignant cells function to suppress self-reactive lymphocytes and to inhibit the effector function of TAA-specific cytotoxic T lymphocytes (CTLs). Accordingly, an engineered $\gamma\delta$ T-cell that lacks PD1 can retain its cytotoxic activity regardless of expression of PD-L1 and PD-L2 by tumor cells. In some cases, an engineered $\gamma\delta$ T-cell of the disclosure lacks the gene locus for the PD-1 gene. In some cases, expression of the PD-1 gene in an engineered $\gamma\delta$ T-cell is disrupted by gene editing technologies.

CTLA4 (cytotoxic T-lymphocyte antigen 4) is also known as CD152 (Cluster of differentiation 152). CTLA4 shares sequence homology and ligands (CD80/B7-1 and CD86/B7-2) with the costimulatory molecule CD28, but differs by delivering inhibitory signals to T-cells expressing CTLA4 as a receptor. CTLA4 has a much higher overall affinity for both ligands and can out-compete CD28 for binding when ligand densities are limiting. CTLA4 is often expressed on the surface of CD8⁺ effector T-cells, and plays a functional role in the initial activation stages of both naive and memory T-cells. CTLA4 counteracts the activity of CD28 via increased affinity for CD80 and CD86 during the early stages of T-cell activation. The major functions of CTLA4 include down-modulation of helper T-cells and enhancement of regulatory T-cell immunosuppressive activity. In some instances, an engineered $\gamma\delta$ T-cell of the disclosure lacks the CTLA4 gene. In some cases, expression of the CTLA4 gene in an engineered $\gamma\delta$ T-cell is disrupted by gene editing

technologies.

LAG3 (Lymphocyte-activation gene 3) is expressed on activated antigen-specific cytotoxic T-cells, and can enhance the function of regulatory T-cells and independently inhibit CD8⁺ effector T-cell activity. LAG3 is a CD-4-like negative regulatory protein with a high affinity binding to MHC Class II proteins, which are upregulated on some epithelial cancers, leading to tolerance of T cell proliferation and homeostasis. Reduction of the LAG-3/Class II interaction using a LAG-3-IG fusion protein may enhance antitumor immune responses. In some cases, an engineered $\gamma\delta$ T-cell of the disclosure lacks the gene locus for the LAG3 gene. In some instances, expression of the LAG3 gene in an engineered $\gamma\delta$ T-cell is disrupted by gene editing technologies.

Phenotype of Non-Engineered and Engineered $\gamma\delta$ T-cells

An engineered $\gamma\delta$ T-cell may home to a specific physical location in a subject's body. Migration and homing of engineered $\gamma\delta$ T cells, can be dependent on the combined expression and actions of specific chemokines and/or adhesion molecules. Homing of engineered $\gamma\delta$ T cells can be controlled by the interactions between chemokines and their receptors. For example, cytokines including but not limited to CXCR3 (whose ligands are represented by IP-10/CXCL10 and 6Ckine/SLC/CCL21) CCR4⁺ CXCR5⁺ (receptor for RANTES, MIP-1 α , MIP-1 β), CCR6⁺ and CCR7 may affect homing of engineered $\gamma\delta$ T cells. In some cases, an engineered $\gamma\delta$ T-cell may home to sites of inflammation and injury, and to diseased cells to perform repair functions. In some cases, an engineered $\gamma\delta$ T-cell can home to a cancer. In some cases, an engineered $\gamma\delta$ T-cell may home to a thymus, a bone marrow, a skin, a larynx, a trachea, pleurae, a lung, an esophagus, an abdomen, a stomach, a small intestine, a large intestine, a liver, a pancreas, a kidney, a urethra, a bladder, a testis, a prostate, a ductus deferens, an ovary, an ureter, a mammary gland, a parathyroid gland, a spleen or another site in a subject's body. An engineered $\gamma\delta$ T-cell can express one or more homing moieties, such as particular TCR allele and/or a lymphocyte homing molecule.

An engineered $\gamma\delta$ T-cell may have a particular phenotype and a phenotype can be described in terms of cell-surface marker expression. Various types of $\gamma\delta$ T-cells can be engineered as described herein. In preferred embodiments, the engineered $\gamma\delta$ T-cell is derived from a human, but the engineered $\gamma\delta$ T-cell may also be derived from a different source, such as a mammal or a synthetic cell.

The immunophenotype of the expanded cell populations may be determined using markers including but not limited to CD27, CD45RA, CD45RO, CCR7 and CD62L (Klebanoff et al., Immunol Rev.211: 214 2006). CD45RA is expressed on naïve T lymphocytes, replaced by CD45RO upon antigen encounter, but re-expressed in late effector cells (Michie et al., Nature 360, 264 - 265 (1992); CD62L is a cell adhesion molecule that acts as a homing molecule to enter secondary lymphoid tissues and is lost after T-cell activation, when T-cells acquire effector functions (Sallusto et al., Nature. 401:708 (1999);. CD27 is costimulation markers that are lost during T-cell differentiations (Appay et al., Nat Med.8:379 (2002); Klebanoff et al., Immunol Rev.211: 214 2006).

Antigens

The invention disclosed herein provides an engineered $\gamma\delta$ T-cell that expresses an antigen recognition moiety, wherein the antigen recognition moiety recognizes a disease-specific epitope. An antigen may be a molecule that provokes an immune response. This immune response may involve either antibody production, the activation of specific immunologically-competent cells, or both. An antigen may be, for example, a peptide, a protein, a hapten, a lipid, a carbohydrate, bacteria, a pathogen, or a virus. An antigen may be a tumor antigen. A tumor epitope may be presented by the MHC I or MHC II complexes on the surface of tumor cells. An epitope can be the portion of the antigen that is expressed on the cell surface and recognized by the tumor recognition moiety.

Non-limiting examples of antigens recognized by an engineered $\gamma\delta$ T-cell include CD19, CD20, CD30, CD22, CD37, CD38, CD56, CD33, CD138, CD123, CD79b, CD70, CD75, CA6, GD2, alphafetoprotein (AFP), carcinoembryonic antigen (CEA), RON, CEACAM5, CA-125, MUC-16, 5T4, NaPi2b, ROR1, ROR2, PLIF, Her2/Neu, EGFRvIII, GPMNB, LIV-1, glycolipidF77, fibroblast activation protein (FAP), PSMA, STEAP-1, STEAP-2, mesothelin, c-Met, CSPG4, PVRL-4, VEGFR2, PSCA, CLEC12a, L1CAM, GPC2, GPC3, folate binding protein/receptor, SLC44A4, Cripto, CTAG1B, AXL, IL-13R, IL-3R α 2, SLTRK6, gp100, MART1, Tyrosinase, SSX2, SSX4, NYESO-1, WT-1, PRAME, epithelial tumor antigen (ETA), MAGEA family genes (such as MAGEA3, MAGEA4), KKLC1, mutated ras, $\square\square$ af, p53, MHC class I chain-related molecule A (MICA), or MHC class I chain-related molecule B (MICB), or one or more antigens of HPV, CMV, or EBV.

An antigen can be expressed in the intracellular or the extracellular compartment of a cell and an engineered $\gamma\delta$ T-cell can recognize an intracellular or an extracellular tumor antigen. In some cases, an $\alpha\beta$ TCR in the engineered $\gamma\delta$ T-cell recognizes a peptide derived from either an intracellular or an extracellular tumor antigen. For example, an antigen may be a protein intracellularly or extracellularly produced by a cell infected with a virus, such as an HIV, an EBV, a CMV, or an HPV protein. An antigen may also be a protein intracellularly or extracellularly expressed by a cancerous cell.

An antigen recognition moiety may recognize an antigen from a cell in distress, such as a cancerous cell or a cell that has been infected with a virus. For instance, the human MHC class I chain-related genes (MICA and MICB) are located within the HLA class I region of chromosome 6. MICA and MICB proteins are considered to be markers of “stress” in the human epithelia, and act as ligands for cells expressing a common natural killer-cell receptor (NKG2D). As stress markers, MICA and MICB can be highly expressed from cancerous cells. An engineered $\gamma\delta$ T-cell can recognize a MICA or a MICB tumor epitope.

A tumor recognition moiety may be engineered to recognize an antigen with certain avidity. For instance, a tumor recognition moiety encoded by a TCR or CAR construct may recognize an antigen with a dissociation constant of at least at least 10 fM, at least 100 fM, at least 1 picomolar (pM), at least 10 pM, at least 20 pM, at least 30 pM, at least 40 pM, at least 50 pM, at least 60 pM, at least 7 pM, at least 80 pM, at least 90 pM, at least 100 pM, at least 200 pM, at least 300 pM, at least 400 pM, at least 500 pM, at least 600 pM, at least 700 pM, at least 800 pM, at least 900 pM, at least 1 nanomolar (nM), at least 2 nM, at least 3 nM, at least 4 nM, at least 5 nM, at least 6 nM, at least 7 nM, at least 8 nM, at least 9 nM, at least 10 nM, at least 20 nM, at least 30 nM, at least 40 nM, at least 50 nM, at least 60 nM, at least 70 nM, at least 80 nM, at least 90 nM, at least 100 nM, at least 200 nM, at least 300 nM, at least 400 nM, at least 500 nM, at least 600 nM, at least 700 nM, at least 800 nM, at least 900 nM, at least 1 μ M, at least 2 μ M, at least 3 μ M, at least 4 μ M, at least 5 μ M, at least 6 μ M, at least 7 μ M, at least 8 μ M, at least 9 μ M, at least 10 μ M, at least 20 μ M, at least 30 μ M, at least 40 μ M, at least 50 μ M, at least 60 μ M, at least 70 μ M, at least 80 μ M, at least 90 μ M, or at least 100 μ M.

In some instances, a tumor recognition moiety may be engineered to recognize an antigen with a dissociation constant of at most 10 fM, at most 100 fM, at most 1 picomolar (pM), at most 10

pM, at most 20 pM, at most 30 pM, at most 40 pM, at most 50 pM, at most 60 pM, at most 7 pM, at most 80 pM, at most 90 pM, at most 100 pM, at most 200 pM, at most 300 pM, at most 400 pM, at most 500 pM, at most 600 pM, at most 700 pM, at most 800 pM, at most 900 pM, at most 1 nanomolar (nM), at most 2 nM, at most 3 nM, at most 4 nM, at most 5 nM, at most 6 nM, at most 7
 5 nM, at most 8 nM, at most 9 nM, at most 10 nM, at most 20 nM, at most 30 nM, at most 40 nM, at most 50 nm, at most 60 nM, at most 70 nM, at most 80 nM, at most 90 nM, at most 100 nM, at most 200 nM, at most 300 nM, at most 400 nM, at most 500 nM, at most 600 nM, at most 700 nM, at most 800 nM, at most 900 nM, at most 1 μ M, at most 2 μ M, at most 3 μ M, at most 4 μ M, at most 5 μ M, at most 6 μ M, at most 7 μ M, at most 8 μ M, at most 9 μ M, at most 10 μ M, at most 20 μ M, at
 10 most 30 μ M, at most 40 μ M, at most 50 μ M, at most 60 μ M, at most 70 μ M, at most 80 μ M, at most 90 μ M, or at most 100 μ M.

Novel Activating Agents

The inventors of the present invention have identified activation agents that bind specific
 15 subtypes of $\gamma\delta$ TCRs and thereby activate specific populations of $\gamma\delta$ T cells. In one aspect, the invention provides novel activating agents which bind the novel activating epitopes identified and described herein in Examples 14, and 39, and FIGS 23-30, and 33-36. Activation agents include, but are not limited to MAbs TS8.2, TS-1, 15D, B6, R9.12, δ 1-05, δ 1-08, δ 1-18, δ 1-22, δ 1-23, δ 1-26, δ 1-35, δ 1-37, δ 1-39, δ 1-113, δ 1-143, δ 1-149, δ 1-155, δ 1-182, δ 1-183, δ 1-191, δ 1-192, δ 1-195, δ 1-197,
 20 δ 1-199, δ 1-201, δ 1-203, δ 1-239, δ 1-253, δ 1-257, δ 1-278, δ 1-282, δ 1-285, δ 2-14, δ 2-17, δ 2-22, δ 2-30, δ 2-31, δ 2-32, δ 2-33, δ 2-35, δ 2-36, and δ 2-37.

These activation agents further include, but are not limited to activation agents that bind the same epitope or compete with one or more MAbs selected from the group consisting of TS8.2, TS-1, 15D, B6, R9.12, δ 1-05, δ 1-08, δ 1-18, δ 1-22, δ 1-23, δ 1-26, δ 1-35, δ 1-37, δ 1-39, δ 1-113, δ 1-143, δ 1-
 25 149, δ 1-155, δ 1-182, δ 1-183, δ 1-191, δ 1-192, δ 1-195, δ 1-197, δ 1-199, δ 1-201, δ 1-203, δ 1-239, δ 1-253, δ 1-257, δ 1-278, δ 1-282, δ 1-285, δ 2-14, δ 2-17, δ 2-22, δ 2-30, δ 2-31, δ 2-32, δ 2-33, δ 2-35, δ 2-36, and δ 2-37.

These activation agents further include, but are not limited to activation agents that contain the complementarity determining regions (CDRs) and/or variable regions of a MAb selected from
 30 the group consisting of TS8.2, TS-1, 15D, B6, R9.12, δ 1-05, δ 1-08, δ 1-18, δ 1-22, δ 1-23, δ 1-26, δ 1-

35, δ 1-37, δ 1-39, δ 1-113, δ 1-143, δ 1-149, δ 1-155, δ 1-182, δ 1-183, δ 1-191, δ 1-192, δ 1-195, δ 1-197, δ 1-199, δ 1-201, δ 1-203, δ 1-239, δ 1-253, δ 1-257, δ 1-278, δ 1-282, δ 1-285, δ 2-14, δ 2-17, δ 2-22, δ 2-30, δ 2-31, δ 2-32, δ 2-33, δ 2-35, δ 2-36, and δ 2-37.

The present invention also provides a nucleic acid encoding an activation agent that: (i) contains the complementarity determining regions (CDRs) and/or variable regions of; (ii) binds the same epitope as or competes with; or (iii) is a MAb selected from the group consisting of TS8.2, TS-1, 15D, B6, R9.12, δ 1-05, δ 1-08, δ 1-18, δ 1-22, δ 1-23, δ 1-26, δ 1-35, δ 1-37, δ 1-39, δ 1-113, δ 1-143, δ 1-149, δ 1-155, δ 1-182, δ 1-183, δ 1-191, δ 1-192, δ 1-195, δ 1-197, δ 1-199, δ 1-201, δ 1-203, δ 1-239, δ 1-253, δ 1-257, δ 1-278, δ 1-282, δ 1-285, δ 2-14, δ 2-17, δ 2-22, δ 2-30, δ 2-31, δ 2-32, δ 2-33, δ 2-35, δ 2-36, and δ 2-37. In some cases, the nucleic acid is in a host cell (*e.g.*, a heterologous host cell). In some cases, the nucleic acid is operably linked to a heterologous promoter or operably linked to a nucleic acid encoding a heterologous polypeptide. As used herein, the term “heterologous” refers to two components that do not naturally exist together in nature.

In certain embodiments the activating agent (*e.g.*, antibody) expands or activates one or more $\gamma\delta$ T-cell populations (*e.g.*, δ 1 T-cells or δ 2 T-cells). In certain embodiments, the activating agent selectively activates δ 1 and δ 3 T-cells. In certain embodiments, the activating agent selectively activates δ 1 and δ 4 T-cells. In certain embodiments, the activating agent selectively activates δ 1 T-cells. In certain embodiments, the activating agent selectively activates δ 1, δ 3, δ 5, and δ 5 T-cells. In certain embodiments, the activating agent selectively activates δ 2 T-cells.

The present invention also provides methods of producing one or more of the foregoing activation agents. For example, a host cell containing a nucleic acid encoding an activation agent can be cultured to produce one or more of the foregoing activation agents.

APCs

Also described herein are APCs for expansion of engineered or non-engineered $\gamma\delta$ T-cells, such as one or more subpopulations of $\gamma\delta$ T-cells. In some embodiments, described herein is an APC that contains a heterologous nucleic acid encoding one or more of the foregoing activation agents. In some embodiments, described herein is an APC that expresses one or more of the foregoing activation agents on a cell surface. In some embodiments, described herein is an APC that expresses one or more Fc receptors on its cell surface, wherein the Fc receptor(s) are in contact with

and/or bound to one or more of the foregoing activation agents.

In some cases, the APCs (*e.g.*, APCs having one or more of the foregoing activation agents expressed, on or bound to an Fc receptor expressed on, the cell surface) do not express, or exhibit reduced expression of, HLA class I, HLA class I, invariant chain, and/or HLA-DM. In some cases, the APCs express adhesion molecules such as intercellular adhesion molecule-1, CD11a, CD18, CD54, and/or leukocyte function-associated antigen-3. In some cases, the APCs express an Fc receptor, such as an Fc receptor that is specific for an isotype of an activation agent used in a $\gamma\delta$ T-cell expansion method described herein. In some cases, the APCs express one or more Fc receptors selected from the group consisting of CD64, CD32A, CD32B, CD32C, CD16A, CD16B, FcRn, TRIM21, or CD307, or an engineered variant thereof having a higher affinity or altered specificity.

Also described herein cultures comprising one or more of the foregoing APCs. The culture can further contain expanded or unexpanded, engineered or non-engineered, $\gamma\delta$ T-cells. The culture can additionally or alternatively contain a selective or non-selective $\gamma\delta$ T-cell activation agent including any one of the $\gamma\delta$ T-cell activation agents described herein. In some cases, the culture does not contain IL-21. In some cases, the culture does not contain IL-4, IL-2, or IL-15, or a combination thereof. In some cases, the culture does not contain a cytokine that selectively expands a sub-population of $\gamma\delta$ T cells.

Epitope Identification

The inventors of the present invention have identified binding regions within the epitope of $\gamma\delta$ TCR activating MAbs TS8.2, TS-1, 15D, B6, R9.12, δ 1-05, δ 1-08, δ 1-18, δ 1-22, δ 1-23, δ 1-26, δ 1-35, δ 1-37, δ 1-39, δ 1-113, δ 1-143, δ 1-149, δ 1-155, δ 1-182, δ 1-183, δ 1-191, δ 1-192, δ 1-195, δ 1-197, δ 1-199, δ 1-201, δ 1-203, δ 1-239, δ 1-253, δ 1-257, δ 1-278, δ 1-282, δ 1-285, δ 2-14, δ 2-17, δ 2-22, δ 2-30, δ 2-31, δ 2-32, δ 2-33, δ 2-35, δ 2-36, and δ 2-37. Exemplary epitopes include, but are not limited to, the epitopes of $\gamma\delta$ TCR activating MAbs TS8.2, TS-1, 15D, B6, R9.12, δ 1-05, δ 1-08, δ 1-18, δ 1-22, δ 1-23, δ 1-26, δ 1-35, δ 1-37, δ 1-39, δ 1-113, δ 1-143, δ 1-149, δ 1-155, δ 1-182, δ 1-183, δ 1-191, δ 1-192, δ 1-195, δ 1-197, δ 1-199, δ 1-201, δ 1-203, δ 1-239, δ 1-253, δ 1-257, δ 1-278, δ 1-282, δ 1-285, δ 2-14, δ 2-17, δ 2-22, δ 2-30, δ 2-31, δ 2-32, δ 2-33, δ 2-35, δ 2-36, and δ 2-37. In some embodiments, the epitope is an epitope specifically bound by one or more of $\gamma\delta$ TCR activating MAbs TS8.2, TS-1, 15D, B6, R9.12, δ 1-05, δ 1-08, δ 1-22, δ 1-26, δ 1-35, δ 1-37, δ 1-39, δ 1-113, δ 1-

143, δ 1-149, δ 1-155, δ 1-182, δ 1-183, δ 1-191, δ 1-192, δ 1-195, δ 1-197, δ 1-199, δ 1-201, δ 1-203, δ 1-253, δ 1-257, δ 1-278, δ 1-282, δ 1-285, δ 2-14, δ 2-17, δ 2-30, δ 2-31, δ 2-32, δ 2-33, δ 2-35, δ 2-36, and δ 2-37.

In one aspect, the disclosure provides a method for identifying the epitope of an agent that stimulates the expansion of engineered and non-engineered $\gamma\delta$ T-cells at a fast rate of growth. An epitope can include at least 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 amino acids in a unique spatial conformation. Epitopes can be formed both from contiguous amino acids or noncontiguous amino acids juxtaposed by tertiary folding of a protein. Epitopes formed from contiguous amino acids can be typically retained on exposure to denaturing solvents whereas epitopes formed by tertiary folding can be typically lost on treatment with denaturing solvents.

Epitope mapping can be performed to identify the linear or non-linear, discontinuous amino acid sequence(s), i.e. the epitope, that is (e.g., specifically) recognized by an activating agent of interest, such as the TS8.2, 15D, B6, TS-1, and R9.12, antibodies. A general approach for epitope mapping can require the expression of the full-length polypeptide sequence that is recognized by an antibody or ligand of interest, as well as various fragments, i.e., truncated forms of the polypeptide sequence, generally in a heterologous expression system. These various recombinant polypeptide sequences or fragments thereof (e.g., fused with an N-terminal protein (e.g., GFP)) can then be used to determine if the antibody or ligand of interest is capable of binding to one or more of the truncated forms of the polypeptide sequence.

In some embodiments, the recombinant polypeptide sequences are chimeras containing joined fragments of two or more homologous parental polypeptides, wherein at least one parental polypeptide binds to the activating agent of interest and at least one parental polypeptide does not bind to the activating agent of interest. For example, segments of a human δ 1 chain can gene be joined with segments of a homologous dolphin δ chain gene, and tested for the ability to generate a chimeric TCR in a recombinant expression system. Chimeric TCR genes that form a TCR, as e.g., indicated by detection with a pan- $\gamma\delta$ -TCR antibody on a cell surface, can then be tested for binding to the activation agent of interest. As another example, segments of a human δ 2 chain can gene be joined with segments of a homologous macaque δ chain gene, and tested for the ability to generate a chimeric TCR in a recombinant expression system. Chimeric TCR genes that form a TCR, as e.g., indicated by detection with a pan- $\gamma\delta$ -TCR antibody on a cell surface, can then be tested for binding

to the activation agent of interest.

Through the use of reiterative truncation and the generation of recombinant polypeptide sequences with overlapping amino acid regions, it is possible to identify the region of the polypeptide sequence that is recognized by the antibody of interest (see, e.g., Epitope Mapping
5 Protocols in Methods in Molecular Biology, Vol. 66, Glenn E. Morris, Ed (1996)). The methods rely on the ability of an agent such as an antibody of interest to bind to sequences that have been recreated from epitope libraries, such as epitope libraries derived from, synthetic peptide arrays on membrane supports, combinatorial phage display peptide libraries. The epitope libraries then provide a range of possibilities that are screened against an antibody. Additionally, site specific mutagenesis,
10 or random Ala scan, targeting one or more residues of an epitope can be pursued to confirm the identity of an epitope.

A library of epitopes can be created by synthetically designing various possible recombinations of a $\gamma\delta$ T-cell receptor ($\gamma\delta$ TCR) as cDNA constructs and expressing them in a suitable system. For instance, a plurality of V δ 1 gene segments differing in their J δ region can be
15 synthetically designed, including J δ 1, J δ 2 and J δ 3 gene segments. Alternatively, V δ 2J δ 1 and V δ 3J δ 1 chains can also be ordered as synthetic genes and cloned into suitable vectors. A plurality of synthetically cloned δ TCR chains, such as V δ 1J δ 1, V δ 1J δ 2, V δ 1J δ 3, V δ 1J δ 4, V δ 2 and V δ 3, chains can be co-transfected into a host system with synthetically cloned γ TCR chains such as V γ 2, V γ 3, V γ 4, V γ 5, V γ 8, V γ 9 and V γ 10 synthetically designed gene segments. In other cases, δ TCR chains,
20 such as V δ 1J δ 1, V δ 1J δ 2, V δ 1J δ 3, V δ 1J δ 4, V δ 2 and V δ 3, chains can be amplified out of Total RNA extracted from human PBMCs or $\gamma\delta$ T-cells isolated from human normal and malignant tissue.

The host system can be any suitable expression system such as 293 cells, insect cells, or a suitable in-vitro translation system. The plurality of various possible recombinations of synthetically designed $\gamma\delta$ T-cell segments transfected into a host system can provide, for instance, more than 25,
25 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, or 90 possible pairing combinations of $\gamma\delta$ TCRs. The binding of an agent to one of the epitopes in the previously described library can be detected by contacting a labeled antibody, such as TS8.2, 15D, B6, TS-1, and R9.12, with an epitope of the library and detecting a signal from the label.

For epitope mapping, computational algorithms have also been developed which have been
30 shown to map conformational discontinuous epitopes. Conformational epitopes can be identified by

determining spatial conformation of amino acids with methods that include, e.g., x-ray crystallography and 2-dimensional nuclear magnetic resonance. Some epitope mapping methods, such as, x-ray analyses of crystals of antigen:antibody complexes can provide atomic resolution of the epitope. In other cases, computational combinatorial methods for epitope mapping can be employed to model a potential epitope based on the sequence of the antibody, such as TS-1 antibody or TS8.2 antibody. In such cases, the antigen binding portion of the antibody is sequenced, and computation models are used to reconstruct and predict a potential binding site of the antibody.

In some cases the disclosure provides a method of determining an epitope of a $\gamma\delta$ T-cell receptor, comprising: (a) preparing a library of epitopes from the $\gamma\delta$ T-cell receptor; (b) contacting the library of epitopes with an antibody; and (b) identifying the amino acid sequence of at least one epitope in the library of epitopes that is bound by the antibody. In some cases, the antibody is selected from the group consisting of, TS8.2, 15D, B6, TS-1, and R9.12, antibodies. In one instance, the antibody is attached to a solid support. The library of epitopes can comprise sequences that correspond to continuous and discontinuous epitopes of a T-cell receptor, such as a γ TCR or a δ TCR. In some cases, the library of epitopes comprises fragments from a $\gamma\delta$ T-cell receptor ranging from about 10 amino acids to about 30 amino acids in length, from about 10 amino acids to about 20 amino acids in length, or from about 5 amino acids to about 12 amino acids in length. In some cases, the antibody is labeled and the label is a radioactive molecule, a luminescent molecule, a fluorescent molecule, an enzyme, or biotin.

δ 1 Epitope Bins

In some embodiments, the epitope, that is (e.g., specifically) recognized by an activating agent of interest is an epitope comprised of amino acids 47-70 of human V δ 1 (SKEMIFLIRQGSDEQNA) and J1 (TDKLIFGKGTRVTVEP) or J2 (LTAQLFFGKGTLIVEP), wherein the activating agent does not bind an epitope containing a K120T mutation in J1 or J2. This epitope is referred to herein as a Bin 1 δ 1 epitope. Exemplary activating agents that bind a Bin 1 δ 1 epitope include but are not limited to TS-1; and δ 1-18. A human V δ 1 J region can vary due to (D)(J) recombination, an exemplary V δ 1 J region of a δ 1 chain of a $\gamma\delta$ TCR having the Bin 1 δ 1 epitope is SKEMIFLIRQGSDEQNAKSGRYSVNFKKAASVALTISALQLEDSAKYFCALGTGVRGLQD TDKLIFGKGTRVTVEP; and another exemplary V δ 1 J region Bin 1 δ 1 epitope of a δ 1 chain of a $\gamma\delta$ TCR is

SKEMIFLIRQGSDEQNAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGEAPSAWGK
HLTAQLFFGKGTQLIVEP.

In some cases, activating agents that bind a Bin 1 $\delta 1$ epitope bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

5 AQKVTQAQSSVSMPPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQDKLIFGKGTRVTVE
P; but do not bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

10 AQKVTQAQSSVSMPPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDSWDTRQMFFGTGIK
LFVEP.

The activating agents that bind a Bin 1 $\delta 1$ epitope can also bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

15 AQKVTQAQSSVSMPPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGEAPSAWGKHLTAQLFFGKGTQLI
VEP.

In some embodiments, the epitope, that is (*e.g.*, specifically) recognized by an activating agent of interest is an epitope comprised of amino acids 47-70 of human V $\delta 1$ (SKEMIFLIRQGSDEQNA) and J1 (TDKLIFGKGTRVTVEP), wherein the activating agent does not bind an epitope containing a K120T mutation in J1. This epitope is referred to herein as a Bin 1b $\delta 1$ epitope.
20 Exemplary activating agents that bind a Bin 1b $\delta 1$ epitope include but are not limited to $\delta 1$ -37. An exemplary V $\delta 1$ J region of a $\delta 1$ chain of a $\gamma\delta$ TCR having the Bin 1b $\delta 1$ epitope is
SKEMIFLIRQGSDEQNAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQD
TDKLIFGKGTRVTVEP.

25 In some cases, activating agents that bind a Bin 1b $\delta 1$ epitope bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

 AQKVTQAQSSVSMPPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQDKLIFGKGTRVTVE
P; but do not bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

30 AQKVTQAQSSVSMPPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDSWDTRQMFFGTGIK

LFVEP.

The activating agents that bind a Bin 1b $\delta 1$ epitope also do not bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

AQKVTQAQSSVSMPIVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
 5 NAKS.GRYSVNFKKAAKSVALTISALQLEDSAKYFCALGEAPSAWGKHLTAQLFFGKGQTQLI
 VEP.

In some embodiments, the epitope that is (*e.g.*, specifically) recognized by an activating agent of interest is an epitope comprised of amino acids 11-21 of human V $\delta 1$ (VSMPVRKAVTL). This epitope is referred to herein as a Bin 2 $\delta 1$ epitope. Exemplary activating agents that bind a Bin
 10 2 $\delta 1$ epitope include but are not limited to and $\delta 1$ -285.

In some cases, activating agents that bind a Bin 2 $\delta 1$ epitope bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

AQKVTQAQSSVSMPIVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
 NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQTDKLIFGKGTRVTVE
 15 P; but do not bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

AQKVTQVQRAMSSQLGEAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
 NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQTDKLIFGKGTRVTVE
 PRSQPHTKPSVFVMKNGTNVACLVEF.

In some embodiments, the epitope that is (*e.g.*, specifically) recognized by an activating
 20 agent of interest is an epitope comprised of amino acids 11-21 of human V $\delta 1$ (VSMPVRKAVTL), wherein the activating agent that binds this epitope does not bind an epitope containing a mutation of R16 in V $\delta 1$, such as an R16N mutation. This epitope is referred to herein as a Bin 2b $\delta 1$ epitope. Exemplary activating agents that bind a Bin 2b $\delta 1$ epitope include but are not limited to R9.12.

In some cases, activating agents that bind a Bin 2b $\delta 1$ epitope bind a $\delta 1$ chain of a $\gamma\delta$ TCR
 25 having the sequence of:

AQKVTQAQSSVSMPIVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
 NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQTDKLIFGKGTRVTVE
 P; but do not bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

AQKVTQAQSSVSMPIVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
 30 NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQTDKLIFGKGTRVTVE

P; and/or do not bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

AQKVTQVQRAMSSQLGEAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQTDKLIFGKGTRVTVE
PRSQPHTKPSVFVMKNGTNVACLVKEF.

5 In some embodiments, the epitope that is (*e.g.*, specifically) recognized by an activating agent of interest is an epitope comprised of amino acids 11-21 of human V $\delta 1$ (VSMPVRKAVTL), wherein the activating agent that binds this epitope also binds (cross-reacts) with $\delta 3$, $\delta 4$, and $\delta 5$ $\gamma\delta$ TCRs. This epitope is referred to herein as a Bin 2c $\delta 1$ epitope. Exemplary activating agents that bind a Bin 2c $\delta 1$ epitope include but are not limited to $\delta 1$ -39.

10 In some cases, activating agents that bind a Bin 2c $\delta 1$ epitope bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQTDKLIFGKGTRVTVE
P; but do not bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

15 AQKVTQVQRAMSSQLGEAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQTDKLIFGKGTRVTVE
PRSQPHTKPSVFVMKNGTNVACLVKEF.

In some embodiments, the epitope that is (*e.g.*, specifically) recognized by an activating agent of interest is an epitope comprised of amino acids 80-95 of human V $\delta 1$ (FKKAAKSVALTISALQ) or 70 to 95 of human V $\delta 1$ (AKSGRYSVNFKKAAKSVALTISALQ).
20 This epitope is referred to herein as a Bin 3 $\delta 1$ epitope. Exemplary activating agents that bind a Bin 3 $\delta 1$ epitope include but are not limited to $\delta 1$ -08; and $\delta 1$ -23.

In some cases, activating agents that bind a Bin 3 $\delta 1$ epitope bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

25 AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQTDKLIFGKGTRVTVE
P

but do not bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

30 AQKVTQVQRAMSSQLGEAVTLSCQYETSLSWYDIFWYKQLPSGEMTFLIHQISSDQ
NAKNGRYSVNFQERHKFISLTISALQLEDSAKYFCALGTGVRGLQDQTDKLIFGKGTRVTVEP

RSQPHTKPSVFVMKNGTNAVCLVKEFYPKD.

In some embodiments, the epitope that is (*e.g.*, specifically) recognized by an activating agent of interest is an epitope comprised of amino acids 1-11 of human V δ 1 (AQKVTQAQSSV) and J1 or J2. This epitope is referred to herein as a Bin 4 δ 1 epitope. In some cases, the Bin 4 δ 1 epitope binding activating agent does not bind an epitope containing a K120T mutation in J1 or J2. Exemplary activating agents that bind a Bin 4 δ 1 epitope include but are not limited to δ 1-35; and δ 1-203.

In some cases, activating agents that bind a Bin 4 δ 1 epitope bind a δ 1 chain of a $\gamma\delta$ TCR having the sequence of:

AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQTDKLIFGKGTRVTVE
P; but do not bind a δ 1 chain of a $\gamma\delta$ TCR having the sequence of:

AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDSWDTRQMFFGTGIK
LFVEP.

In some cases, activating agents that bind a Bin 4 δ 1 epitope bind a δ 1 chain of a $\gamma\delta$ TCR having the sequence of:

AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGEAPSAWGKHLTAQLFFGKGQTQLI
VEP.

In some embodiments, the epitope that is (*e.g.*, specifically) recognized by an activating agent of interest is an epitope comprised of amino acids 28-47 of human V δ 1

(SWWSYYIFWYKQLPS) and J1. This epitope is referred to herein as a Bin 5 δ 1 epitope.

Exemplary activating agents that bind a Bin 5 δ 1 epitope include but are not limited to δ 1-113; δ 1-155; δ 1-183; δ 1-191; δ 1-278; and δ 1-282. An exemplary V δ 1 J1 region of a δ 1 chain of a $\gamma\delta$ TCR having the Bin 5 δ 1 epitope is

SWWSYYIFWYKQLPSKEMIFLIRQGSDEQNAKSGRYSVNFKKAAKSVALTISALQLEDSAK
YFCALGTGVRGLQDQTDKLIFGKGTRVTVEP.

In some cases, activating agents that bind a Bin 5 δ 1 epitope bind a δ 1 chain of a $\gamma\delta$ TCR having the sequence of:

AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQTDKLIFGKGTRVTVE
P; but do not bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
5 NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGEAPSAWGKHLTAQLFFGKGQTQLI
VEP.

In some embodiments, the epitope that is (*e.g.*, specifically) recognized by an activating
agent of interest is an epitope comprised of amino acids 21-28 of human V $\delta 1$ (LNCLYETS) and J1.
This epitope is referred to herein as a Bin 6 $\delta 1$ epitope. Exemplary activating agents that bind a Bin
10 6 $\delta 1$ epitope include but are not limited to TS 8.2 and $\delta 1$ -143. An exemplary V $\delta 1$ J1 region of a $\delta 1$
chain of a $\gamma\delta$ TCR having the Bin 6 $\delta 1$ epitope is
LNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQNAKSGRYSVNFKKAAKSVALTISAL
QLEDSAKYFCALGTGVRGLQDQTDKLIFGKGTRVTVEP.

In some cases, activating agents that bind a Bin 6 $\delta 1$ epitope bind a $\delta 1$ chain of a $\gamma\delta$ TCR
15 having the sequence of:

AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQTDKLIFGKGTRVTVE
P; but do not bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
20 NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDSWDTRQMFFGTGIK
LFVEP.

In some embodiments, the epitope that is (*e.g.*, specifically) recognized by an activating
agent of interest is an epitope comprised of amino acids 47-70 of human V $\delta 1$
(SKEMIFLIRQGSDEQNA) and J1 or J2. This epitope is referred to herein as a Bin 7 $\delta 1$ epitope.
25 Exemplary activating agents that bind a Bin 7 $\delta 1$ epitope include but are not limited to $\delta 1$ -149; $\delta 1$ -
253, and $\delta 1$ -257. An exemplary V $\delta 1$ J region of a $\delta 1$ chain of a $\gamma\delta$ TCR having the Bin 7 $\delta 1$ epitope
is
SKEMIFLIRQGSDEQNAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQD
TDKLIFGKGTRVTVEP; and another exemplary V $\delta 1$ J region Bin 7 $\delta 1$ epitope of a $\delta 1$ chain of a $\gamma\delta$
30 TCR is

SKEMIFLIRQGSDEQNAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGEAPSAWGK
HLTAQLFFGKGTQLIVEP.

In some cases, activating agents that bind a Bin 7 $\delta 1$ epitope bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

5 A Q K V T Q A Q S S V S M P V R K A V T L N C L Y E T S W W S Y Y I F W Y K Q L P S K E M I F L I R Q G S D E Q
N A K S G R Y S V N F K K A A K S V A L T I S A L Q L E D S A K Y F C A L G T G V R G L Q D T D K L I F G K G T R V T V E
P; but do not bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

10 A Q K V T Q A Q S S V S M P V R K A V T L N C L Y E T S W W S Y Y I F W Y K Q L P S K E M I F L I R Q G S D E Q
N A K S G R Y S V N F K K A A K S V A L T I S A L Q L E D S A K Y F C A L G T G V R G L Q D S W D T R Q M F F G T G I K
L F V E P.

In some embodiments, the epitope that is (*e.g.*, specifically) recognized by an activating agent of interest is an epitope comprised of amino acids 70-80 of human V $\delta 1$ (AKSGRYSVNF) and J1 or J2. This epitope is referred to herein as a Bin 8 $\delta 1$ epitope. Exemplary activating agents that bind a Bin 8 $\delta 1$ epitope include but are not limited to $\delta 1$ -192. An exemplary V $\delta 1$ J region of a $\delta 1$ chain of a $\gamma\delta$ TCR having the Bin 8 $\delta 1$ epitope is
15 A K S G R Y S V N F K K A A K S V A L T I S A L Q L E D S A K Y F C A L G T G V R G L Q D T D K L I F G K G T R V T V E P;
and another exemplary V $\delta 1$ J region Bin 8 $\delta 1$ epitope of a $\delta 1$ chain of a $\gamma\delta$ TCR is
A K S G R Y S V N F K K A A K S V A L T I S A L Q L E D S A K Y F C A L G E A P S A W G K H L T A Q L F F G K G T Q L I V
E P.

20 In some cases, activating agents that bind a Bin 8 $\delta 1$ epitope bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

A Q K V T Q A Q S S V S M P V R K A V T L N C L Y E T S W W S Y Y I F W Y K Q L P S K E M I F L I R Q G S D E Q
N A K S G R Y S V N F K K A A K S V A L T I S A L Q L E D S A K Y F C A L G T G V R G L Q D T D K L I F G K G T R V T V E
P; but do not bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

25 A Q K V T Q A Q S S V S M P V R K A V T L N C L Y E T S W W S Y Y I F W Y K Q L P S K E M I F L I R Q G S D E Q
N A K S G R Y S V N F K K A A K S V A L T I S A L Q L E D S A K Y F C A L G T G V R G L Q D S W D T R Q M F F G T G I K
L F V E P.

In some embodiments, the epitope that is (*e.g.*, specifically) recognized by an activating agent of interest is an epitope comprised of amino acids 80-95 of human V $\delta 1$
30 (FKKAAKSVALTISALQ). This epitope is referred to herein as a Bin 9 $\delta 1$ epitope. Exemplary

activating agents that bind a Bin 9 $\delta 1$ epitope include but are not limited to $\delta 1$ -201.

In some cases, activating agents that bind a Bin 9 $\delta 1$ epitope bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of:

AQKVTQAQSSVSMVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
 5 NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQDKLIFGKGTRVTVE
 P; but do not bind a $\delta 1$ chain of a $\gamma\delta$ TCR having the sequence of

AQKVTQVQRAMSSQLGEAVTLSCQYETSLSWYDIFWYKQLPSGEMTFLIHQISSDQ
 NAKNGRYSVNFQERHKFISLTISALQLEDSAKYFCALGTGVRGLQDQDKLIFGKGTRVTVEP.

10 The $\delta 1$ -specific antibodies described herein selectively bind $\delta 1$ -containing $\gamma\delta$ -TCRs over $\delta 2$ -containing $\gamma\delta$ -TCRs. As such, the foregoing $\delta 1$ -specific antibodies do not bind, *e.g.*, the sequence of and/or a $\gamma\delta$ -TCR comprising the sequence of:

AIELVPEHQTPVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGP
 GFKDNFQGDIDIAKNLAVLKILAPSERDEGSYYCACPGLGGPPDKLIFGKGTRVTVEP.

$\delta 2$ Epitope Bins

15 In some embodiments, the epitope, that is (*e.g.*, specifically) recognized by an activating agent of interest is an epitope comprised of amino acids 83-94 of human V $\delta 2$ (AKNLAVLKILAP). This epitope is referred to herein as a Bin 1 $\delta 2$ epitope. In some cases, the activating agent that binds the Bin 1 $\delta 2$ epitope does not bind an epitope containing a mutation of K90 in V $\delta 2$, such as an K90N mutation. Exemplary activating agents that bind a Bin 1 $\delta 2$ epitope include but are not limited to $\delta 2$ -
 20 17, and B6.

In some cases, activating agents that bind a Bin 1 $\delta 2$ epitope bind a $\delta 2$ chain of a $\gamma\delta$ TCR having the sequence of

AIELVPEHQTPVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGP
 GFKDNFQGDIDIAKNLAVLKILAPSERDEGSYYCACPGLGGPPDKLIFGKGTRVTVEP; but do
 25 not bind a $\delta 2$ chain of a $\gamma\delta$ TCR having the sequence of:

AIELVPEHQTPVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGP
 GFKDNFQGDIDFLNNQAVLNILEASERDEGSYYCACPGLGGPPDKLIFGKGTRVTVEP.

30 In some embodiments, the epitope, that is (*e.g.*, specifically) recognized by an activating agent of interest is an epitope comprised of amino acids 28-38 of human V $\delta 2$ (EAIGNYY). This epitope is referred to herein as a Bin 2 $\delta 2$ epitope. In some cases, the activating agent that binds the

Bin 2 $\delta 2$ epitope does not bind an epitope containing a mutation of G35 in V $\delta 2$, such as an G35S mutation. Exemplary activating agents that bind a Bin 2 $\delta 2$ epitope include but are not limited to 15D.

In some cases, activating agents that bind a Bin 2 $\delta 2$ epitope bind a $\delta 2$ chain of a $\gamma\delta$ TCR having the sequence of:

AIELVPEHQTVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGP
GFK.DNFQGDIDIAKNLAVLKILAPSERDEGSYYCACDPLGGPPDKLIFGKGTRVTVEP; but
do not bind a $\delta 2$ chain of a $\gamma\delta$ TCR having the sequence of:

AIELVPEHQTVPSIGVPATLRCSMKGDSISNYYTFWYRRTPGNTMTLIYREGGTYG
PGFEDNLQGEIDFLNNQAVLNILEASERDEGSYYCACDPLGGPPDKLIFGKGTRVTVEP.

In some embodiments, the epitope, that is (*e.g.*, specifically) recognized by an activating agent of interest is an epitope comprised of amino acids 72-83 of human V $\delta 2$ (KDNFQGDIDIA). This epitope is referred to herein as a Bin 3 $\delta 2$ epitope. Exemplary activating agents that bind a Bin 3 $\delta 2$ epitope include but are not limited to $\delta 2$ -32.

In some cases, activating agents that bind a Bin 3 $\delta 2$ epitope bind a $\delta 2$ chain of a $\gamma\delta$ TCR having the sequence of:

AIELVPEHQTVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGP
GFK.DNFQGDIDIAKNLAVLKILAPSERDEGSYYCACDPLGGPPDKLIFGKGTRVTVEP; but
do not bind a $\delta 2$ chain of a $\gamma\delta$ TCR having the sequence of:

AIELVPEHQTVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGP
GFEDNLQGEIDFLNNQAVLNILEASERDEGSYYCACDPLGGPPDKLIFGKGTRVTVEP.

In some embodiments, the epitope, that is (*e.g.*, specifically) recognized by an activating agent of interest is an epitope comprised of amino acids 1-27 of human V $\delta 2$ (AIELVPEHQTVPSIGVPATLRCSMKG). This epitope is referred to herein as a Bin 4 $\delta 2$ epitope. Exemplary activating agents that bind a Bin 4 $\delta 2$ epitope include but are not limited to $\delta 2$ -14; $\delta 2$ -22; $\delta 2$ -30; $\delta 2$ -31; $\delta 2$ -36; and $\delta 2$ -37.

In some cases, activating agents that bind a Bin 4 $\delta 2$ epitope bind a $\delta 2$ chain of a $\gamma\delta$ TCR having the sequence of:

AIELVPEHQTVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGP
GFK.DNFQGDIDIAKNLAVLKILAPSERDEGSYYCACDPLGGPPDKLIFGKGTRVTVEP; but

do not bind a $\delta 2$ chain of a $\gamma\delta$ TCR having the sequence of:

AVTLVPQNQARSVSVGESVTLRCSMKGDSISNYITFWYRRTPGNTMTLIYREGGTY
GPGFEDNLQGEIDFLNNQAVLNILEASERDEGSYYCACDPLGGPPDKLIFGKGTRVTVEP.

5 The $\delta 2$ -specific antibodies described herein selectively bind $\delta 2$ -containing $\gamma\delta$ -TCRs over $\delta 1$ -containing $\gamma\delta$ -TCRs. As such, the foregoing $\delta 2$ -specific antibodies do not bind, *e.g.*, the sequence of and/or a $\gamma\delta$ -TCR comprising the sequence of:

AQKVTQAQSSVSMPVRKA VTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDQDKLIFGKGTRVTVE
P.

10 Generally, the $\delta 1$ - and $\delta 2$ -specific antibodies described herein recognize a conformational epitope in the context of a $\gamma\delta$ -TCR. In some cases, the $\delta 1$ - and $\delta 2$ -specific antibodies described herein are specific for one or more pairs of $\gamma\delta 1$ - or $\gamma\delta 2$ -TCRs respectively. For example, in some cases, the $\delta 1$ -specific antibodies described herein are specific for a $\gamma\delta 1$ -TCR. In some cases, the $\delta 1$ -specific antibodies described herein bind $\gamma\delta 1$ -TCR but not $\gamma 9\delta 1$ -TCR.

15

Methods of Treatment

Pharmaceutical compositions containing a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, as described herein may be administered for prophylactic and/or therapeutic treatments. In therapeutic applications, the
20 compositions can be administered to a subject already suffering from a disease or condition in an amount sufficient to cure or at least partially arrest the symptoms of the disease or condition. A non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, can also be administered to lessen a likelihood of developing, contracting, or worsening a condition. Effective amounts of a population of a non-engineered, enriched $\gamma\delta$ T-cell
25 population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, for therapeutic use can vary based on the severity and course of the disease or condition, previous therapy, the subject's health status, weight, and/or response to the drugs, and/or the judgment of the treating physician.

A non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell
30 population, and /or admixtures thereof, of the disclosure can be used to treat a subject in need of

treatment for a condition. Examples of conditions include cancer, infectious disease, autoimmune disorder and sepsis. Subjects can be humans, non-human primates such as chimpanzees, and other apes and monkey species; farm animals such as cattle, horses, sheep, goats, swine; domestic animals such as rabbits, dogs, and cats; laboratory animals including rodents, such as rats, mice and guinea pigs, and the like. A subject can be of any age. Subjects can be, for example, elderly adults, adults, adolescents, pre-adolescents, children, toddlers, infants.

A method of treating a condition (e.g., ailment) in a subject with an enriched $\gamma\delta$ T-cell population of the instant invention may comprise administering to the subject a therapeutically-effective amount of a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof. An enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the disclosure may be administered at various regimens (e.g., timing, concentration, dosage, spacing between treatment, and/or formulation). A subject can also be preconditioned with, for example, chemotherapy, radiation, or a combination of both, prior to receiving a an enriched $\gamma\delta$ T-cell population and/or admixtures thereof, of the disclosure. As part of a treatment, a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, may be administered to a subject at a first regimen and the subject may be monitored to determine whether the treatment at the first regimen meets a given level of therapeutic efficacy. In some cases, the engineered $\gamma\delta$ T-cell or another engineered $\gamma\delta$ T-cell may be administered to the subject at a second regimen. **FIG. 2** schematically illustrates a method for treating a subject. In a first operation **201**, at least one engineered $\gamma\delta$ T-cell is administered to a subject that has or is suspected of having a given condition (e.g., cancer). The engineered $\gamma\delta$ T-cell may be administered at a first regimen. In a second operation **202**, the subject may be monitored, for example by a healthcare provider (e.g., treating physician or nurse). In some examples, the subject is monitored to determine or gauge an efficacy of the engineered $\gamma\delta$ T-cell in treating the condition of the subject. In some situations, the subject may also be monitored to determine the *in vivo* expansion of a $\gamma\delta$ T-cell population in the subject. Next, in a third operation **203**, at least one other engineered $\gamma\delta$ T-cell is administered to the subject at a second regimen. The second regimen may be the same as the first regimen or different than the first regimen. In some situations, the third operation **203** is not performed, for example, if the administration of the engineered $\gamma\delta$ T-cell in the first operation **201** is found to be effective (e.g., a single round of administration may be sufficient to

treat the condition). Due to their allogeneic and universal donor characteristics, a population of engineered $\gamma\delta$ T-cells may be administrated to various subjects, with different MHC haplotypes. An engineered $\gamma\delta$ T-cell may be frozen or cryopreserved prior to being administered to a subject.

A enriched population of $\gamma\delta$ T-cells (*i.e.*, engineered or non-engineered) and/or admixtures thereof, may also be frozen or cryopreserved prior to being administered to a subject. In certain embodiments, a population of engineered, enriched $\gamma\delta$ T-cells can comprise two or more cells that express identical, different, or a combination of identical and different tumor recognition moieties. For instance, a population of engineered, enriched $\gamma\delta$ T-cells can comprises several distinct engineered $\gamma\delta$ T-cells that are designed to recognize different antigens, or different epitopes of the same antigen. For example, human cells afflicted with melanoma can express the NY-ESO-1 oncogene. Infected cells within the human can process the NY-ESO-1 oncoprotein into smaller fragments and present various portions of the NY-ESO-1 protein for antigen recognition. A population of engineered, enriched $\gamma\delta$ T-cells can comprise various engineered $\gamma\delta$ T-cells that express different tumor recognition moieties designed to recognize different portions of the NY-ESO-1 protein. **FIG. 3** schematically illustrates a method for treating a subject with a population of engineered $\gamma\delta$ T-cells that recognizes different epitopes of the melanoma antigen NY-ESO-1. In a first operation **301**, a population of engineered $\gamma\delta$ T-cells that recognize different epitopes of the same antigen is selected. For example, the population of engineered $\gamma\delta$ T-cells may comprise two or more cells that expressing different tumor recognition moieties that recognize different portions of the NY-ESO-1protein. In a second operation **302**, The population of engineered $\gamma\delta$ T-cells may be administered at a first regimen. In a second operation **303**, the subject may be monitored, for example by a healthcare provider (e.g., treating physician or nurse).

An enriched $\gamma\delta$ T-cell population, *i.e.*, non-engineered or engineered, and/or admixtures thereof, of the disclosure may be used to treat various conditions. In some cases, a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the disclosure may be used to treat a cancer, including solid tumors and hematologic malignancies. Non-limiting examples of cancers include: acute lymphoblastic leukemia, acute myeloid leukemia, adrenocortical carcinoma, AIDS-related cancers, AIDS-related lymphoma, anal cancer, appendix cancer, astrocytomas, neuroblastoma, basal cell carcinoma, bile duct cancer, bladder cancer, bone cancers, brain tumors, such as cerebellar astrocytoma, cerebral

astrocytoma/malignant glioma, ependymoma, medulloblastoma, supratentorial primitive neuroectodermal tumors, visual pathway and hypothalamic glioma, breast cancer, bronchial adenomas, Burkitt lymphoma, carcinoma of unknown primary origin, central nervous system lymphoma, cerebellar astrocytoma, cervical cancer, childhood cancers, chronic lymphocytic leukemia, chronic myelogenous leukemia, chronic myeloproliferative disorders, colon cancer, cutaneous T-cell lymphoma, desmoplastic small round cell tumor, endometrial cancer, ependymoma, esophageal cancer, Ewing's sarcoma, germ cell tumors, gallbladder cancer, gastric cancer, gastrointestinal carcinoid tumor, gastrointestinal stromal tumor, gliomas, hairy cell leukemia, head and neck cancer, heart cancer, hepatocellular (liver) cancer, Hodgkin lymphoma, Hypopharyngeal cancer, intraocular melanoma, islet cell carcinoma, Kaposi sarcoma, kidney cancer, laryngeal cancer, lip and oral cavity cancer, liposarcoma, liver cancer, lung cancers, such as non-small cell and small cell lung cancer, lymphomas, leukemias, macroglobulinemia, malignant fibrous histiocytoma of bone/osteosarcoma, medulloblastoma, melanomas, mesothelioma, metastatic squamous neck cancer with occult primary, mouth cancer, multiple endocrine neoplasia syndrome, myelodysplastic syndromes, myeloid leukemia, nasal cavity and paranasal sinus cancer, nasopharyngeal carcinoma, neuroblastoma, non-Hodgkin lymphoma, non-small cell lung cancer, oral cancer, oropharyngeal cancer, osteosarcoma/malignant fibrous histiocytoma of bone, ovarian cancer, ovarian epithelial cancer, ovarian germ cell tumor, pancreatic cancer, pancreatic cancer islet cell, paranasal sinus and nasal cavity cancer, parathyroid cancer, penile cancer, pharyngeal cancer, pheochromocytoma, pineal astrocytoma, pineal germinoma, pituitary adenoma, pleuropulmonary blastoma, plasma cell neoplasia, primary central nervous system lymphoma, prostate cancer, rectal cancer, renal cell carcinoma, renal pelvis and ureter transitional cell cancer, retinoblastoma, rhabdomyosarcoma, salivary gland cancer, sarcomas, skin cancers, skin carcinoma merkel cell, small intestine cancer, soft tissue sarcoma, squamous cell carcinoma, stomach cancer, T-cell lymphoma, throat cancer, thymoma, thymic carcinoma, thyroid cancer, trophoblastic tumor (gestational), cancers of unknown primary site, urethral cancer, uterine sarcoma, vaginal cancer, vulvar cancer, Waldenström macroglobulinemia, and Wilms tumor.

In some cases, a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the disclosure may be used to treat an infectious disease. An infectious disease may be caused, for example, by a pathogenic bacterium or by a virus.

Various pathogenic proteins, nucleic acids, lipids, or fragments thereof can be expressed by a diseased cell. An antigen presenting cell can internalize such pathogenic molecules, for instance with phagocytosis or by receptor-mediated endocytosis, and display a fragment of the antigen bound to an appropriate MHC molecule. For instance, various 9 mer fragments of a pathogenic protein may be displayed by an APC. Engineered, enriched $\gamma\delta$ T-cell populations of the disclosure may be designed to recognize various antigens and antigen fragments of a pathogenic bacterium or a virus. Non-limiting examples of pathogenic bacteria can be found in the: a) *Bordetella* genus, such as *Bordetella pertussis* species; b) *Borrelia* genus, such *Borrelia burgdorferi* species; c) *Brucella* genus, such as *Brucella abortus*, *Brucella canis*, *Brucella meliteris*, and/or *Brucella suis* species; d) *Campylobacter* genus, such as *Campylobacter jejuni* species; e) *Chlamydia* and *Chlamydophila* genres, such as *Chlamydia pneumonia*, *Chlamydia trachomatis*, and/or *Chlamydophila psittaci* species; f) *Clostridium* genus, such as *Clostridium botulinum*, *Clostridium difficile*, *Clostridium perfringens*, *Clostridium tetani* species; g) *Corynebacterium* genus, such as *Corynebacterium diphtheria* species; h) *Enterococcus* genus, such as *Enterococcus faecalis*, and/or *Enterococcus faecium* species; i) *Escherichia* genus, such as *Escherichia coli* species; j) *Francisella* genus, such as *Francisella tularensis* species; k) *Haemophilus* genus, such as *Haemophilus influenza* species; l) *Helicobacter* genus, such as *Helicobacter pylori* species; m) *Legionella* genus, such as *Legionella pneumophila* species; n) *Leptospira* genus, such as *Leptospira interrogans* species; o) *Listeria* genus, such as *Listeria monocytogenes* species; p) *Mycobacterium* genus, such as *Mycobacterium leprae*, *mycobacterium tuberculosis*, and/or *mycobacterium ulcerans* species; q) *Mycoplasma* genus, such as *Mycoplasma pneumonia* species; r) *Neisseria* genus, such as *Neisseria gonorrhoeae* and/or *Neisseria meningitidis* species; s) *Pseudomonas* genus, such as *Pseudomonas aeruginosa* species; t) *Rickettsia* genus, such as *Rickettsia rickettsii* species; u) *Salmonella* genus, such as *Salmonella typhi* and/or *Salmonella typhimurium* species; v) *Shigella* genus, such as *Shigella sonnei* species; w) *Staphylococcus* genus, such as *Staphylococcus aureus*, *Staphylococcus epidermidis*, and/or *Staphylococcus saprophyticus* species; x) *Streptococcus* genus, such as *Streptococcus agalactiae*, *Streptococcus pneumonia*, and/or *Streptococcus pyogenes* species; y) *Treponema* genus, such as *Treponema pallidum* species; z) *Vibrio* genus, such as *Vibrio cholera*; and/or aa) *Yersinia* genus, such as *Yersinia pestis* species.

In some cases, a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$

T-cell population, and/or admixtures thereof, of the disclosure may be used to treat an infectious disease, an infectious disease may be caused a virus. Non-limiting examples of viruses can be found in the following families of viruses and are illustrated with exemplary species: a) Adenoviridae family, such as Adenovirus species; b) Herpesviridae family, such as Herpes simplex type 1, Herpes simplex type 2, Varicella-zoster virus, Epstein-barr virus, Human cytomegalovirus, Human herpesvirus type 8 species; c) Papillomaviridae family, such as Human papillomavirus species; d) Polyomaviridae family, such as BK virus, JC virus species; e) Poxviridae family, such as Smallpox species; f) Hepadnaviridae family, such as Hepatitis B virus species; g) Parvoviridae family, such as Human bocavirus, Parvovirus B19 species; h) Astroviridae family, such as Human astrovirus species; i) Caliciviridae family, such as Norwalk virus species; j) Flaviviridae family, such as Hepatitis C virus (HCV), yellow fever virus, dengue virus, West Nile virus species; k) Togaviridae family, such as Rubella virus species; l) Hepeviridae family, such as Hepatitis E virus species; m) Retroviridae family, such as Human immunodeficiency virus (HIV) species; n) Orthomyxoviridaw family, such as Influenza virus species; o) Arenaviridae family, such as Guanarito virus, Junin virus, Lassa virus, Machupo virus, and/or Sabiá virus species; p) Bunyaviridae family, such as Crimean-Congo hemorrhagic fever virus species; q) Filoviridae family, such as Ebola virus and/or Marburg virus species; Paramyxoviridae family, such as Measles virus, Mumps virus, Parainfluenza virus, Respiratory syncytial virus, Human metapneumovirus, Hendra virus and/or Nipah virus species; r) Rhabdoviridae genus, such as Rabies virus species; s) Reoviridae family, such as Rotavirus, Orbivirus, Coltivirus and/or Banna virus species. In some examples, a virus is unassigned to a viral family, such as Hepatitis D.

In some cases, a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the disclosure may be used to treat an immune disease, such as an autoimmune disease. Inflammatory diseases, including autoimmune diseases are also a class of diseases associated with B- cell disorders. Examples of immune diseases or conditions, including autoimmune conditions, include: rheumatoid arthritis, rheumatic fever, multiple sclerosis, experimental autoimmune encephalomyelitis, psoriasis, uveitis, diabetes mellitus, systemic lupus erythematosus (SLE), lupus nephritis, eczema, scleroderma, polymyositis/scleroderma, polymyositis/dermatomyositis, uncerative proctitis, severe combined immunodeficiency (SCID), DiGeorge syndrome, ataxia-telangiectasia, seasonal allergies, perennial allergies, food allergies,

anaphylaxis, mastocytosis, allergic rhinitis, atopic dermatitis, Parkinson's, Alzheimer's, hypersplenism, leukocyte adhesion deficiency, X-linked lymphoproliferative disease, X-linked agammaglobulinemia, selective immunoglobulin A deficiency, hyper IgM syndrome, HIV, autoimmune lymphoproliferative syndrome, Wiskott-Aldrich syndrome, chronic granulomatous disease, common variable immunodeficiency (CVID), hyperimmunoglobulin E syndrome, Hashimoto's thyroiditis, acute idiopathic thrombocytopenic purpura, chronic idiopathic thrombocytopenia purpura, dermatomyositis, Sydenham's chorea, myasthenia gravis, polyglandular syndromes, bullous pemphigoid, Henoch-Schonlein purpura, poststreptococcal nephritis, erythema nodosum, erythema multiforme, glomerulonephropathy, Takayasu's arteritis, Addison's disease, sarcoidosis, ulcerative colitis, polyarteritis nodosa, ankylosing spondylitis, Goodpasture's syndrome, thromboangitis obliterans, Sjogren's syndrome, primary biliary cirrhosis, Hashimoto's thyroiditis, thyrotoxicosis, chronic active hepatitis, polychondritis, pemphigus vulgaris, Wegener's granulomatosis, membranous nephropathy, amyotrophic lateral sclerosis, tabes dorsalis, giant cell arteritis, polymyalgia, pernicious anemia, rapidly progressive glomerulonephritis, psoriasis, fibrosing alveolitis, and cancer.

Treatment with a $\gamma\delta$ T-cell population, and/or admixtures thereof, of the disclosure may be provided to the subject before, during, and after the clinical onset of the condition. Treatment may be provided to the subject after 1 day, 1 week, 6 months, 12 months, or 2 years after clinical onset of the disease. Treatment may be provided to the subject for more than 1 day, 1 week, 1 month, 6 months, 12 months, 2 years, 3 years, 4 years, 5 years, 6 years, 7 years, 8 years, 9 years, 10 years or more after clinical onset of disease. Treatment may be provided to the subject for less than 1 day, 1 week, 1 month, 6 months, 12 months, or 2 years after clinical onset of the disease. Treatment may also include treating a human in a clinical trial. A treatment can comprise administering to a subject a pharmaceutical composition comprising a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixture thereof, of the disclosure.

In some cases, administration of a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixture thereof, of the disclosure to a subject modulates the activity of endogenous lymphocytes in a subject's body. In some cases, administration of the non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, to a subject provides an antigen to an endogenous T-cell

and may boost an immune response. In some cases, the memory T-cell is a CD4⁺ T-cell. In some cases, the memory T-cell is a CD8⁺ T-cell. In some cases, administration of the non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, to a subject activates the cytotoxicity of another immune cell. In some cases, the other
5 immune cell is a CD8⁺ T-cell. In some cases, the other immune cell is a Natural Killer T-cell. In some cases, administration of the non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, to a subject suppress a regulatory T-cell. In some cases, the regulatory T-cell is a Fox3⁺ Treg cell. In some cases, the regulatory T-cell is a Fox3⁻ Treg cell. Non-limiting examples of cells whose activity can be modulated by a non-
10 engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the disclosure include: hematopoietic stem cells; B cells; CD4; CD8; red blood cells; white blood cells; dendritic cells, including dendritic antigen presenting cells; leukocytes; macrophages; memory B cells; memory T-cells; monocytes; natural killer cells; neutrophil granulocytes; T-helper cells; and T-killer cells.

15 During most bone marrow transplants, a combination of cyclophosphamide with total body irradiation is conventionally employed to prevent rejection of the hematopoietic stem cells (HSC) in the transplant by the subject's immune system. In some cases, incubation of donor bone marrow with interleukin-2 (IL-2) *ex vivo* is performed to enhance the generation of killer lymphocytes in the donor marrow. Interleukin-2 (IL-2) is a cytokine that is necessary for the growth, proliferation, and
20 differentiation of wild-type lymphocytes. Current studies of the adoptive transfer of $\gamma\delta$ T-cells into humans may require the co-administration of $\gamma\delta$ T-cells and interleukin-2. However, both low- and high- dosages of IL-2 can have highly toxic side effects. IL-2 toxicity can manifest in multiple organs/systems, most significantly the heart, lungs, kidneys, and central nervous system. In some cases, the disclosure provides a method for administering a non-engineered, enriched $\gamma\delta$ T-cell
25 population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, to a subject without the co-administration of a cytokine, such as IL-2, IL-15, IL-12, or IL-21. In some cases, a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, can be administered to a subject without co-administration with IL-2. In some cases, a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population,
30 and/or admixtures thereof, is administered to a subject during a procedure, such as a bone marrow

transplant without the co-administration of IL-2.

Methods of Administration

One or multiple non-engineered, enriched $\gamma\delta$ T-cell population, engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the invention can be administered to a subject in any order or simultaneously. If simultaneously, the multiple non-engineered, enriched $\gamma\delta$ T-cell population, engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the invention can be provided in a single, unified form, such as an intravenous injection, or in multiple forms, for example, as multiple intravenous infusions, s.c. injections or pills. The non-engineered, enriched $\gamma\delta$ T-cell population, engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the invention can be packed together or separately, in a single package or in a plurality of packages. One or all of the non-engineered, enriched $\gamma\delta$ T-cell population, engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the invention can be given in multiple doses. If not simultaneous, the timing between the multiple doses may vary to as much as about a week, a month, two months, three months, four months, five months, six months, or about a year. In some cases, a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the invention can expand within a subject's body, *in vivo*, after administration to a subject. Non-engineered, enriched $\gamma\delta$ T-cell population, engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, can be frozen to provide cells for multiple treatments with the same cell preparation. Non-engineered, enriched $\gamma\delta$ T-cell population, engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the disclosure, and pharmaceutical compositions comprising the same, can be packaged as a kit. A kit may include instructions (e.g., written instructions) on the use of the non-engineered, enriched $\gamma\delta$ T-cell population, the engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, and compositions comprising the same.

In some cases, a method of treating a cancer comprises administering to a subject a therapeutically-effective amount of a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, wherein the administration treats the cancer. In some embodiments the therapeutically-effective amount of the non-engineered, enriched $\gamma\delta$ T-cell population, the engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, is administered for at least about 10 seconds, 30 seconds, 1 minute, 10 minutes, 30 minutes, 1 hour, 2

hours, 3 hours, 4 hours, 5 hours, 6 hours, 12 hours, 24 hours, 2 days, 3 days, 4 days, 5 days, 6 days, 1 week, 2 weeks, 3 weeks, 1 month, 2 months, 3 months, 4 months, 5 months, 6 months, or 1 year. In some embodiments the therapeutically-effective amount of the non-engineered, enriched $\gamma\delta$ T-cell population, the engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, is administered for at least one week. In some embodiments the therapeutically-effective amount of the non-engineered, enriched $\gamma\delta$ T-cell population, the engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, is administered for at least two weeks.

A non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, described herein can be administered before, during, or after the occurrence of a disease or condition, and the timing of administering a pharmaceutical composition containing a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, can vary. For example, the non-engineered, enriched $\gamma\delta$ T-cell population, the engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, can be used as a prophylactic and can be administered continuously to subjects with a propensity to conditions or diseases in order to lessen a likelihood of the occurrence of the disease or condition. The non-engineered, enriched $\gamma\delta$ T-cell population, the engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, can be administered to a subject during or as soon as possible after the onset of the symptoms. The administration of the non-engineered, enriched $\gamma\delta$ T-cell population, the engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, can be initiated immediately within the onset of symptoms, within the first 3 hours of the onset of the symptoms, within the first 6 hours of the onset of the symptoms, within the first 24 hours of the onset of the symptoms, within 48 hours of the onset of the symptoms, or within any period of time from the onset of symptoms. The initial administration can be via any route practical, such as by any route described herein using any formulation described herein. In some examples, the administration of a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the disclosure is an intravenous administration. One or multiple dosages of a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, can be administered as soon as is practicable after the onset of a cancer, an infectious disease, an immune disease, sepsis, or with a bone marrow transplant, and for a length of time necessary for the treatment of the immune disease, such as, for example, from about 24 hours to about 48 hours, from

about 48 hours to about 1 week, from about 1 week to about 2 weeks, from about 2 weeks to about 1 month, from about 1 month to about 3 months. For the treatment of cancer, one or multiple dosages of a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, can be administered years after onset of the cancer and before or after other treatments. In some examples, a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, can be administered for at least about 10 minutes, 30 minutes, 1 hour, 2 hours, 3 hours, 4 hours, 5 hours, 6 hours, 12 hours, 24 hours, at least 48 hours, at least 72 hours, at least 96 hours, at least 1 week, at least 2 weeks, at least 3 weeks, at least 4 weeks, at least 1 month, at least 2 months, at least 3 months, at least 4 months, at least 5 months, at least 6 months, at least 7 months, at least 8 months, at least 9 months, at least 10 months, at least 11 months, at least 12 months, at least 1 year, at least 2 years at least 3 years, at least 4 years, or at least 5 years. The length of treatment can vary for each subject.

Dosages

A non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, as disclosed herein may be formulated in unit dosage forms suitable for single administration of precise dosages. In some cases, the unit dosage forms comprise additional lymphocytes. In unit dosage form, the formulation is divided into unit doses containing appropriate quantities of one or more compounds. The unit dosage can be in the form of a package containing discrete quantities of the formulation. Non-limiting examples are packaged tablets or capsules, and powders in vials or ampoules. Aqueous suspension compositions can be packaged in single-dose non-reclosable containers. Multiple-dose reclosable containers can be used, for example, in combination with a preservative or without a preservative. In some examples, the pharmaceutical composition does not comprise a preservative. Formulations for parenteral injection can be presented in unit dosage form, for example, in ampoules, or in multi-dose containers with a preservative.

A non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, as described herein may be present in a composition in an amount of at least 5 cells, at least 10 cells, at least 20 cells, at least 30 cells, at least 40 cells, at least 50 cells, at least 60 cells, at least 70 cells, at least 80 cells, at least 90 cells, at least 100 cells, at least

200 cells, at least 300 cells, at least 400 cells, at least 500 cells, at least 600 cells, at least 700 cells, at least 800 cells, at least 900 cells, at least 1×10^3 cells, at least 2×10^3 cells, at least 3×10^3 cells, at least 4×10^3 cells, at least 5×10^3 cells, at least 6×10^3 cells, at least 7×10^3 cells, at least 8×10^3 cells, at least 9×10^3 cells, at least 1×10^4 cells, at least 2×10^4 cells, at least 3×10^4 cells, at least 4×10^4 cells, at least 5×10^4 cells, at least 6×10^4 cells, at least 7×10^4 cells, at least 8×10^4 cells, at least 9×10^4 cells, at least 1×10^5 cells, at least 2×10^5 cells, at least 3×10^5 cells, at least 4×10^5 cells, at least 5×10^5 cells, at least 6×10^5 cells, at least 7×10^5 cells, at least 8×10^5 cells, at least 9×10^5 cells, at least 1×10^6 cells, at least 2×10^6 cells, at least 3×10^6 cells, at least 4×10^6 cells, at least 5×10^6 cells, at least 6×10^6 cells, at least 7×10^6 cells, at least 8×10^6 cells, at least 9×10^6 cells, at least 1×10^7 cells, at least 2×10^7 cells, at least 3×10^7 cells, at least 4×10^7 cells, at least 5×10^7 cells, at least 6×10^7 cells, at least 7×10^7 cells, at least 8×10^7 cells, at least 9×10^7 cells, at least 1×10^8 cells, at least 2×10^8 cells, at least 3×10^8 cells, at least 4×10^8 cells, at least 5×10^8 cells, at least 6×10^8 cells, at least 7×10^8 cells, at least 8×10^8 cells, at least 9×10^8 cells, at least 1×10^9 cells, or more.

15 The therapeutically effective dose of a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the invention can be from about 1 cell to about 10 cells, from about 1 cell to about 100 cells, from about 1 cell to about 10 cells, from about 1 cell to about 20 cells, from about 1 cell to about 30 cells, from about 1 cell to about 40 cells, from about 1 cell to about 50 cells, from about 1 cell to about 60 cells, from about 1 cell about 20 70 cells, from about 1 cell to about 80 cells, from about 1 cell to about 90 cells, from about 1 cell to about 100 cells, from about 1 cell to about 1×10^3 cells, from about 1 cell to about 2×10^3 cells, from about 1 cell to about 3×10^3 cells, from about 1 cell to about 4×10^3 cells, from about 1 cell to about 5×10^3 cells, from about 1 cell to about 6×10^3 cells, from about 1 cell to about 7×10^3 cells, from about 1 cell to about 8×10^3 cells, from about 1 cell to about 9×10^3 cells, from about 1 cell to about 1×10^4 cells, from about 1 cell to about 2×10^4 cells, from about 1 cell to about 3×10^4 cells, from about 1 cell to about 4×10^4 cells, from about 1 cell to about 5×10^4 cells, from about 1 cell to about 6×10^4 cells, from about 1 cell to about 7×10^4 cells, from about 1 cell to about 8×10^4 cells, from about 1 cell to about 9×10^4 cells, from about 1 cell to about 1×10^5 cells, from about 1 cell to about 2×10^5 cells, from about 1 cell to about 3×10^5 cells, from about 1 cell to about 4×10^5 cells, 30 from about 1 cell to about 5×10^5 cells, from about 1 cell to about 6×10^5 cells, from about 1 cell to

about 7×10^5 cells, from about 1 cell to about 8×10^5 cells, from about 1 cell to about 9×10^5 cells, from about 1 cell to about 1×10^6 cells, from about 1 cell to about 2×10^6 cells, from about 1 cell to about 3×10^6 cells, from about 1 cell to about 4×10^6 cells, from about 1 cell to about 5×10^6 cells, from about 1 cell to about 6×10^6 cells, from about 1 cell to about 7×10^6 cells, from about 1 cell to about 8×10^6 cells, from about 1 cell to about 9×10^6 cells, from about 1 cell to about 1×10^7 cells, from about 1 cell to about 2×10^7 cells, from about 1 cell to about 3×10^7 cells, from about 1 cell to about 4×10^7 cells, from about 1 cell to about 5×10^7 cells, from about 1 cell to about 6×10^7 cells, from about 1 cell to about 7×10^7 cells, from about 1 cell to about 8×10^7 cells, from about 1 cell to about 9×10^7 cells, from about 1 cell to about 1×10^8 cells, from about 1 cell to about 2×10^8 cells, from about 1 cell to about 3×10^8 cells, from about 1 cell to about 4×10^8 cells, from about 1 cell to about 5×10^8 cells, from about 1 cell to about 6×10^8 cells, from about 1 cell to about 7×10^8 cells, from about 1 cell to about 8×10^8 cells, from about 1 cell to about 9×10^8 cells, or from about 1 cell to about 1×10^9 cells.

In some cases, the therapeutically effective dose of a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the invention can be from about 1×10^3 cells to about 2×10^3 cells, from about 1×10^3 cells to about 3×10^3 cells, from about 1×10^3 cells to about 4×10^3 cells, from about 1×10^3 cells to about 5×10^3 cells, from about 1×10^3 cells to about 6×10^3 cells, from about 1×10^3 cells to about 7×10^3 cells, from about 1×10^3 cells to about 8×10^3 cells, from about 1×10^3 cells to about 9×10^3 cells, from about 1×10^3 cells to about 1×10^4 cells, from about 1×10^3 cells to about 2×10^4 cells, from about 1×10^3 cells to about 3×10^4 cells, from about 1×10^3 cells to about 4×10^4 cells, from about 1×10^3 cells to about 5×10^4 cells, from about 1×10^3 cells to about 6×10^4 cells, from about 1×10^3 cells to about 7×10^4 cells, from about 1×10^3 cells to about 8×10^4 cells, from about 1×10^3 cells to about 9×10^4 cells, from about 1×10^3 cells to about 1×10^5 cells, from about 1×10^3 cells to about 2×10^5 cells, from about 1×10^3 cells to about 3×10^5 cells, from about 1×10^3 cells to about 4×10^5 cells, from about 1×10^3 cells to about 5×10^5 cells, from about 1×10^3 cells to about 6×10^5 cells, from about 1×10^3 cells to about 7×10^5 cells, from about 1×10^3 cells to about 8×10^5 cells, from about 1×10^3 cells to about 9×10^5 cells, from about 1×10^3 cells to about 1×10^6 cells, from about 1×10^3 cells to about 2×10^6 cells, from about 1×10^3 cells to about 3×10^6 cells, from about 1×10^3 cells to about 4×10^6 cells, from about 1×10^3 cells to about 5×10^6 cells, from about 1×10^3 cells to about

6 x 10⁶ cells, from about 1 x 10³ cells to about 7 x 10⁶ cells, from about 1 x 10³ cells to about 8 x 10⁶ cells, from about 1 x 10³ cells to about 9 x 10⁶ cells, from about 1 x 10³ cells to about 1 x 10⁷ cells, from about 1 x 10³ cells to about 2 x 10⁷ cells, from about 1 x 10³ cells to about 3 x 10⁷ cells, from about 1 x 10³ cells to about 4 x 10⁷ cells, from about 1 x 10³ cells to about 5 x 10⁷ cells, from
5 about 1 x 10³ cells to about 6 x 10⁷ cells, from about 1 x 10³ cells to about 7 x 10⁷ cells, from about 1 x 10³ cells to about 8 x 10⁷ cells, from about 1 x 10³ cells to about 9 x 10⁷ cells, from about 1 x 10³ cells to about 1 x 10⁸ cells, from about 1 x 10³ cells to about 2 x 10⁸ cells, from about 1 x 10³ cells to about 3 x 10⁸ cells, from about 1 x 10³ cells to about 4 x 10⁸ cells, from about 1 x 10³ cells to about 5 x 10⁸ cells, from about 1 x 10³ cells to about 6 x 10⁸ cells, from about 1 x 10³ cells to about 7 x 10⁸ cells,
10 cells, from about 1 x 10³ cells to about 8 x 10⁸ cells, from about 1 x 10³ cells to about 9 x 10⁸ cells, or from about 1 x 10³ cells to about 1 x 10⁹ cells.

In some cases, the therapeutically effective dose of a non-engineered, enriched $\gamma\delta$ T-cell population, an engineered, enriched $\gamma\delta$ T-cell population, and/or admixtures thereof, of the invention can be from about 1 x 10⁶ cells to about 2 x 10⁶ cells, from about 1 x 10⁶ cells to about 3 x 10⁶ cells,
15 from about 1 x 10⁶ cells to about 4 x 10⁶ cells, from about 1 x 10⁶ cells to about 5 x 10⁶ cells, from about 1 x 10⁶ cells to about 6 x 10⁶ cells, from about 1 x 10⁶ cells to about 7 x 10⁶ cells, from about 1 x 10⁶ cells to about 8 x 10⁶ cells, from about 1 x 10⁶ cells to about 9 x 10⁶ cells, from about 1 x 10⁶ cells to about 1 x 10⁷ cells, from about 1 x 10⁶ cells to about 2 x 10⁷ cells, from about 1 x 10⁶ cells to about 3 x 10⁷ cells, from about 1 x 10⁶ cells to about 4 x 10⁷ cells, from about 1 x 10⁶ cells to about 5 x 10⁷ cells, from about 1 x 10⁶ cells to about 6 x 10⁷ cells, from about 1 x 10⁶ cells to about 7 x 10⁷ cells,
20 cells, from about 1 x 10⁶ cells to about 8 x 10⁷ cells, from about 1 x 10⁶ cells to about 9 x 10⁷ cells, from about 1 x 10⁶ cells to about 1 x 10⁸ cells, from about 1 x 10⁶ cells to about 2 x 10⁸ cells, from about 1 x 10⁶ cells to about 3 x 10⁸ cells, from about 1 x 10⁶ cells to about 4 x 10⁸ cells, from about 1 x 10⁶ cells to about 5 x 10⁸ cells, from about 1 x 10⁶ cells to about 6 x 10⁸ cells, from about 1 x 10⁶ cells to about 7 x 10⁸ cells, from about 1 x 10⁶ cells to about 8 x 10⁸ cells, from about 1 x 10⁶ cells to about 9 x 10⁸ cells, from about 1 x 10⁶ cells to about 1 x 10⁹ cells, from about 1 x 10⁶ cells to about 2 x 10⁹ cells, from about 1 x 10⁶ cells to about 3 x 10⁹ cells, from about 1 x 10⁶ cells to about 4 x 10⁹ cells, from about 1 x 10⁶ cells to about 5 x 10⁹ cells, from about 1 x 10⁶ cells to about 6 x 10⁹ cells, from about 1 x 10⁶ cells to about 7 x 10⁹ cells, from about 1 x 10⁶ cells to about 8 x 10⁹ cells, from
25 about 1 x 10⁶ cells to about 9 x 10⁹ cells, from about 1 x 10⁷ cells to about 1 x 10⁹ cells, from about 1
30 about 1 x 10⁶ cells to about 9 x 10⁹ cells, from about 1 x 10⁷ cells to about 1 x 10⁹ cells, from about 1

x 10^7 cells to about 2×10^9 cells, from about 1×10^7 cells to about 3×10^9 cells, from about 1×10^7 cells to about 4×10^9 cells, from about 1×10^7 cells to about 5×10^9 cells, from about 1×10^7 cells to about 6×10^9 cells, from about 1×10^7 cells to about 7×10^9 cells, from about 1×10^7 cells to about 8×10^9 cells, from about 1×10^7 cells to about 9×10^9 cells, from about 1×10^8 cells to about 1×10^9 cells, from about 1×10^8 cells to about 2×10^9 cells, from about 1×10^8 cells to about 3×10^9 cells, from about 1×10^8 cells to about 4×10^9 cells, from about 1×10^8 cells to about 5×10^9 cells, from about 1×10^8 cells to about 6×10^9 cells, from about 1×10^8 cells to about 7×10^9 cells, from about 1×10^8 cells to about 8×10^9 cells, from about 1×10^8 cells to about 9×10^9 cells, or from about 1×10^9 cells to about 1×10^{10} cells.

Preservation

In some embodiments, enriched $\gamma\delta$ T-cell populations, and/or admixtures thereof, may be formulated in freezing media and placed in cryogenic storage units such as liquid nitrogen freezers (-195°C) or ultra-low temperature freezers (-65°C , -80°C or -120°C) for long-term storage of at least about 1 month, 2 months, 3 months, 4 months, 5 months, 6 months, 1 year, 2 years, 3 years, or at least 5 years. The freeze media can contain dimethyl sulfoxide (DMSO), and/or sodium chloride (NaCl), and/or dextrose, and/or dextran sulfate and/or hydroxyethyl starch (HES) with physiological pH buffering agents to maintain pH between about 6.0 to about 6.5, about 6.5 to about 7.0, about 7.0 to about 7.5, about 7.5 to about 8.0 or about 6.5 to about 7.5. The cryopreserved $\gamma\delta$ T-cells can be thawed and further processed by stimulation with antibodies, proteins, peptides, and/or cytokines as described herein. The cryopreserved $\gamma\delta$ T-cells can be thawed and genetically modified with viral vectors (including retroviral and lentiviral vectors) or non-viral means (including RNA, DNA, and proteins) as described herein. Alternatively, non-engineered $\gamma\delta$ T-cells can be expanded by the methods described herein, genetically modified, and cryopreserved.

Thus, genetically engineered and/or non-engineered $\gamma\delta$ T-cells can be further cryopreserved to generate cell banks in quantities of at least about 1, 5, 10, 100, 150, 200, 500 vials at about at least 10^1 , 10^2 , 10^3 , 10^4 , 10^5 , 10^6 , 10^7 , 10^8 , 10^9 , or at least about 10^{10} cells per mL in freeze media. The cryopreserved cell banks may retain their functionality and can be thawed and further stimulated and expanded. In some aspects, thawed cells can be stimulated and expanded in suitable closed vessels such as cell culture bags and/or bioreactors to generate quantities of cells as

allogeneic cell product. Cryopreserved $\gamma\delta$ T-cells can maintain their biological functions for at least about 6 months, 7 months, 8 months, 9 months, 10 months, 11 months, 12 months, 13 months, 15 months, 18 months, 20 months, 24 months, 30 months, 36 months, 40 months, 50 months, or at least about 60 months under cryogenic storage condition. In some aspects, no preservatives are used in the formulation. The cryopreserved $\gamma\delta$ T-cells can be thawed and infused into multiple patients as allogeneic off-the-shelf cell product.

All publications and patents mentioned herein are incorporated herein by reference in their entirety for the purpose of describing and disclosing, for example, the constructs and methodologies that are described in the publications, which might be used in connection with the presently described inventions. The publications discussed herein are provided solely for their disclosure prior to the filing date of the present application. Nothing herein is to be construed as an admission that the inventors described herein are not entitled to antedate such disclosure by virtue of prior invention or for any other reason.

The present invention is explained in more detail by the following non-limiting examples.

EXAMPLES

Example 1. Primary Cell Isolation, Digestion and Culture

Primary human peripheral blood mononuclear cells (PBMCs) are collected from healthy donors using an apheresis machine. The PBMCs are purified with the Ficoll-Paque™ PLUS (GE Healthcare Bio-Sciences AB, Uppsala, Sweden) system or similar systems. The cells are then re-suspended in an appropriate growth medium.

Alternatively, primary human cells are collected from peripheral blood, cord blood, bone marrow, healthy tissues, or tissues afflicted with a disease, such as cancerous tissues.

Tissue from Healthy Donors

Fresh tissues from healthy donors are received from The Cooperative Human Tissue Network (CHTN) and are transported to the laboratory in RPMI-1640 media. Tissues are sliced into 1–3 mm³ fragments with a scalpel. 2-5 fragments/well placed in a 24-well plate (Costar) in 2 mL RPMI-1640 supplemented with GlutaMAX, 25 mM HEPES pH 7.2, 100 U/ml penicillin, 100 U/ml streptomycin and 10% Human AB Serum and 100 IU/ml of rhIL-2 or digested as described below.

The plates are incubated in a humidified incubator at 37⁰ C, with 5% CO₂ in air. Cultures are inspected every other day to monitor proliferation of lymphocytes. Half of the medium is replaced in all wells every 7 days after culture initiation. Lymphocytes are collected when a dense lymphocytic carpet covers the surrounding the fragments or lymphocyte population derived from digested tissues
5 reached the proper concentration, as described below.

Tissue Enzymatic Digestion

Fresh tissue samples from healthy donors were received from The Cooperative Human Tissue Network (CHTN) and were transported to the laboratory in RPMI-1640 media. Lymphocytes
10 were isolated by enzymatic digestion using two enzyme blend Liberase™ DL Research Grade (Sigma Aldrich Co., St. Louis, MO), or Liberase™ TM Research Grade (Sigma Aldrich Co., St. Louis, MO) or the Miltenyi tumor dissociation kit (130-095-929) with the gentleMACS Dissociator.

The tissues were cut into 2-3mm³ fragments and digested for 1 hour at 37° C and 5% CO₂. The digested cell suspension was passed through a 40-micron filter, spun down and washed with
15 RPMI-1640 medium. Cells were counted and re-suspended in RPMI medium (GIBCO BRL), supplemented with 10% Human AB serum(Corning) and 100 IU / ml of rhIL-2. The collected cell population was seeded at 0.5 to 1 X 10⁶ cells/ml in 24-well tissue culture plates. Cells were split into RPMI-IL2 containing medium when cells exceeded concentration of 1.5 X 10⁶ cells/ml.

Culture of Tumor Specimens

Fresh tumor specimens from patients with primary and metastatic cancers, including those of the colon, breast, ovary, kidney, head and neck, oral cavity, pancreas and liver, were received from The Cooperative Human Tissue Network (CHTN) and were transported to the laboratory in RPMI media. Tumor specimens were sliced into 1–3 mm³ fragments with a scalpel. 2-5 fragments/well
25 were placed in a 24-well plate (Costar) in 2ml RPMI-1640 supplemented with GlutaMAX, 25 mM HEPES pH 7.2, 100 U/ml penicillin, 100 U/ml streptomycin and 10% Human AB serum and 100 IU / ml of rhIL-2. The plates were incubated in a humidified incubator at 37⁰ C, with 5% CO₂ in air. Cultures were inspected every other day to monitor proliferation of lymphocytes. Half of the medium was replaced in all wells every 7 days after culture initiation. Lymphocytes were collected
30 when a dense lymphocytic carpet covers the surrounding the fragments.

Fresh Tumor Specimens Enzymatic Digestion

Fresh tumor specimens from patients with primary and metastatic cancers, including those of the colon, breast, ovary, kidney, head and neck, oral cavity, pancreas and liver, were received from
5 The Cooperative Human Tissue Network (CHTN) and were transported to the laboratory in RPMI media. Lymphocytes were isolated by enzymatic digestion using enzyme blend Liberase™ DL Research Grade (Sigma Aldrich Co., St. Louis, MO), Liberase™ TM Research Grade (Sigma Aldrich Co., St. Louis, MO), or the Miltenyi tumor dissociation kit (130-095-929) with the gentleMACS Dissociator. The tissues were cut into 2–3mm³ fragments and digested for 1 hour at 37⁰
10 C with 5% CO₂ in air. The digested cell suspension was passed through a 40-micron filter, spun down and washed with RPMI-1640 medium. Cells were counted and re-suspended in RPMI medium (GIBCO BRL), supplemented with 10% Human AB Serum (Corning) and 100 IU/ml of rhIL-2. The collected cell population was seeded at 0.5 to 1 X 10⁶ cells/ml in 24-well tissue culture plates. Cells were split into RPMI-IL2 containing medium when cells exceeded concentration of 1.5 X 10⁶
15 cells/ml.

Culture of Primary Cells in Exemplary Serum Supplemented Media

PBMC populations were generated by separation from buffy coats derived from healthy donors, using Ficoll-Paque™ PLUS (GE Healthcare Bio-Sciences PA, USA). PBMCs were cultured
20 at 1 X 10⁶ cells/mL in 24-well tissue culture plate in RPMI-1640 (Corning CellGro) supplemented with 10% fetal Bovine serum (Gibco), 2 mmol/L L-glutamine, 100 U/mL penicillin, 100 U/mL streptomycin, and 100 IU rhIL-2/ml.

Similar culture conditions can be used to grow primary human cells collected from peripheral blood, cord blood, bone marrow, healthy tissues, or tissues afflicted with a disease, such as the
25 cancerous tissues previously described.

Culture of Primary Cells in Exemplary Serum-Free Media

PBMC populations were generated by separation from buffy coats derived from healthy donors, using Ficoll-Paque™ PLUS (GE Healthcare Bio-Sciences PA, USA). PBMCs were cultured
30 at 1 X 10⁶ cells/mL in 24-well tissue culture plate in CTS serum-free media with CTS-OpTmizer

supplement.

Similar culture conditions can be used to grow primary human cells collected from peripheral blood, cord blood, bone marrow, healthy tissues, or tissues afflicted with a disease, such as the cancerous tissues previously described.

5

Example 2: Depletion of Adherent Monocytes and Macrophages and CD4+ and CD8+ $\alpha\beta$ T cells

PMBCs are collected with apheresis methods as previously described and red blood cells are removed by hypotonic treatment or density separation using Ficoll gradient centrifugation. The red blood cell-free PBMCs are incubated in large-scale tissue culture vessels such as 10-stack or 40-stack Cell Factory (Nunc), roller bottles (Nunc). Adherent populations, comprising macrophages and monocytes, typically remain bound to the surface of the cell culture vessels. The cell population grown in suspension population is enriched in $\gamma\delta$ T-cells. Approximately 10^8 , 10^9 or 10^{10} PBMC are incubated with anti-human CD4 and anti-human CD8 coated iron-containing microbeads (e.g. Miltenyi Biotec Microbeads). The incubated cell population flows pass a magnetic field in which the CD4⁺ and CD8⁺ T-cells are retained. The “flow-through” cell population is enriched for $\gamma\delta$ T-Cells.

Example 3: Depletion of Monocytes and Macrophages

PMBCs are collected with apheresis methods as previously described and red blood cells are removed by hypotonic treatment or density separation using Ficoll gradient centrifugation. The red blood cell-free PBMCs are incubated in large-scale tissue culture vessels such as 10-stack or 40-stack Cell Factory (Nunc), roller bottles (Nunc). Monocytes and macrophages are removed by flowing the red blood cell-removed PBMC over a packed glass wool column. The “flow-through” cell population is enriched in $\gamma\delta$ T-cells for further processing.

Example 4: Enrichment of $\gamma\delta$ T-Cells

PMBCs are collected with apheresis methods as previously described and red blood cells are removed by hypotonic treatment or density separation using Ficoll gradient centrifugation. The red blood cell-free PBMCs are incubated in large-scale tissue culture vessels such as 10-stack or 40-

stack Cell Factory (Nunc), roller bottles (Nunc). Monocytes and macrophages are depleted either with the methods described in Example 2 or in Example 3. Approximately 10^8 , 10^9 or 10^{10} PBMC are incubated with CD4 and CD8 coated iron-containing microbeads (e.g. Miltenyi Biotech Microbeads). The incubated cell population flows pass a magnetic field in which the anti human
 5 CD4⁺ and anti human CD8⁺ T-cells are retained. The “flow-through” cell population is enriched for $\gamma\delta$ T-cells. Undesired cells, such as NK, $\gamma\delta$ T-cells, B cells, monocytes and macrophages are removed by immunomagnetic beads separation (e.g. Miltenyi Biotech AutoMACS system) using a cocktail of antibodies directed against the undesired cell types.

Alternatively, undesired cell types are removed by using mouse anti human $\alpha\beta$ TCR (IP26,
 10 BW242/412) antibody, one or more other antibody directed against surface receptors on NK, $\alpha\beta$ T cells, B cells, monocytes or macrophages and attached to Anti-Mouse IgG MicroBeads (Miltenyi 130-048-401), or one or more tetrameric antibody complex or bi-specific antibody directed against surface receptors on NK, $\alpha\beta$ T cells, B cells, monocytes or macrophages, or combinations thereof.

15 Isolation of $\gamma\delta$ T-Cells from Primary Cells with Antibodies

In one example, native $\gamma\delta$ T-cells are isolated from primary cultures with flow cytometry sorting, based on the positive (i.e. $\gamma\delta$ TCR) or negative (i.e. $\alpha\beta$ TCR, CD4, CD8, CD56), expression of cell surface markers.

20 **Example 5. Isolation of $\gamma\delta$ T-Cells from Primary Tumors**

$\gamma\delta$ T-cells were isolated according to the methods detailed in Example 1. Briefly, freshly harvested tumor specimens were obtained from NCI Cooperative Human Tissue Network (CHTN). Colon adenocarcinoma metastasis to liver (TIL 1) and renal tumor (TIL 2) were shipped in RPMI-1640 media. The tumor tissues were minced into small pieces of 2mm^3 in size using a flat blade
 25 followed by digestion with 2mL Liberase enzyme cocktail (Sigma Chemical Co., St. Louis, MO) in RPMI and 3000 units of DNase as described. After digestion tumors were filtered through sterile gauze 40-micron nylon mesh, and washed twice in RPMI-1640. Cells were counted and plated in 24 well plate at $1 \times 10^6/\text{ml}$ in RPMI-1640 containing 10% human AB serum supplemented with L-glutamine, and 100 U/mL of rhIL-2. Tumor infiltrating lymphocytes were collected after 6 days in
 30 culture. The presence of $\gamma\delta$ T lymphocytes was analyzed by flow cytometry anti- $\delta 1$ TCR (FITC

conjugated anti V δ 1TS8.2, (Thermo Fisher) and anti -V δ 2 B6 (Biolegend). Data was analyzed with FlowJo software.

FIG. 4 depicts a graph illustrating growth of $\gamma\delta$ 1 and $\gamma\delta$ 2 lymphocytes isolated from colon adenocarcinoma metastasis to liver (TIL 1) and renal tumors (TIL 2). These lymphocytes have been shown to express CCR4 and CCR7 (data not shown). As illustrated by **FIG. 4**, the V δ 1 subset was the predominant population isolated from both types of tumors.

Example 6. Stimulation and Expansion of $\gamma\delta$ T-Cells

$\gamma\delta$ T-Cells are stimulated and expanded in serum-free media such as Ex-Vivo 10 (Lonza 04-380Q), Ex-Vivo 15 (Lonza 04-744Q), Ex-Vivo 20 (Lonza 04-448Q), AIMV media (ThermoFisher Scientific 12055091), Optimizer CTS (ThermoFisher Scientific A1048501), containing cytokines (IL-2, IL-4, IL-7, IL-15, IL-12, IL-21, IL-23 or IL-33), growth factors (insulin and transferrin, insulin-like growth factors), albumin, lipids (cholesterol, lipid solutions, lipid pre-cursors), vitamins, copper, iron, selenium, protein hydrolysate, essential amino acids, non-essential amino acids, and shear protectant (Pluronic F-68).

The serum-free media described in the examples herein can also be supplemented with additives to support high cell density $\gamma\delta$ T-cell growth between 10^5 to 2×10^7 cells/mL in suspension culture (e.g. WAVE bioreactor) while maintaining biological functionality of the $\gamma\delta$ T-cell.

Additional additives that provided robust $\gamma\delta$ T-cell growth include, for example, Calcium Chloride, Anhydrous, Calcium Nitrate, Cupric Sulfate, Pentahydrate, Ferric Citrate, Ferric Nitrate, Ferrous Sulfate, Zinc Sulfate, and/or Putrescine.

Trace metals were provided in the serum free media to provide low levels of elemental components to replace serum, including Ammonium Paramolybdate, Vanadium, Manganese, Nickel, Sodium Selenite, Sodium Metasilicate, Nonahydrate, Stannous Chloride, Aluminum Chloride, Barium Acetate, Cadmium Chloride, Chromic Chloride, Cobalt, Germanium Dioxide, Potassium Bromide, Potassium Iodide, Rubidium Chloride, Silver Nitrate, Sodium Fluoride, and/or Zirconyl Chloride.

Other components added to cell culture media that support robust growth of $\gamma\delta$ T-cells are, for example, Adenosine, Guanosine, Cytidine, Uridine, Betaine, Taurine, Folinic acid, Ethanolamine,

Linoleic Acid, Oleic Acid Hydrocortisone, pyruvate, plant hydrolysates, yeast hydrolysates, and/or beta-mercaptoethanol.

Vitamins added to promote robust $\gamma\delta$ T cell growth include: Biotin (B7), D-Calcium Pantothenate (B5), Choline Chloride, Cyanocobalamin (B12), Folic Acid (B9), i-Inositol (myo-Inositol), Niacinamide(B3), Pyridoxal, Monohydrochloride, Pyridoxine, Monohydrochloride (B6), Riboflavin (B2), Thiamine, and/or Monohydrochloride (B1).

Example 7: Characterization of Expanded $\gamma\delta$ T-cells: Immunophenotype

Expanded T-cell populations may be characterized, for example, by FACS staining for cell surface markers that distinguish between the different populations. The cells were washed once in HEPES buffered saline solution (HBSS) containing 2% fetal bovine serum, incubated with appropriate amounts of MAbs at 4⁰ C for 30 minutes and rewashed in HBSS. Briefly, 1 x 10⁶ cells are stained in 100 μ l volume of FACS staining medium (FSM; HBSS containing 2% fetal bovine serum) containing fluoresoithiocyanate (FITC) or phycoerythrin (PE) conjugated MAbs directed against CD2, CD3 (BioLegend, clone OKT3), CD4 (BioLegend clone OKT4), CD7, CD8 (BioLegend, clone RPAT8), CD11a, CD16, CD18, CD19, CD27, CD28, CD38, CD45RA, CD56, CD57, CD69, CD71, CD95, CD107, ICAM-1, MICA/B, NKG2D DR5, CCR1, CCR2, CCR3, CCR4, CCR5 CCR6, CCR7, CCR10, CXCR1, CXCR2, CXCR3, CXCR5, CXCR5, CXCR6, CXCR7, IL-2R, IL-7R, Ki67, L-selectin, VLA-4, JAML, PD1, PDL1, CTLA-4, O $\times$ 40, TCR V δ 1 (ThermoFisher Scientific, clone TS8.2), or TCR V δ 2 (BioLegend, clone B6).

In addition to surface markers, cytokine secretion, intracellular cytokines and/or membrane associated cytokines including, for example, TNF- α IFN- γ , GM-CSF, IL-1, IL-2, IL-4, IL-6, IL-7, IL-10, IL-17, or IL-21 may be characterized according to methods known in the art.

In certain examples herein, live cells were determined by absence or low incorporation of zombie violet (BioLegend) amine dye. Fluorescence Minus One (FMO) controls are used to define positive and negative gate boundaries of the surface expression of each antigen. Stained cells are collected on a Sony SH800 cytometer and data analyzed using FlowJo v10.1. Flow cytometry data are visualized as dot plots.

Example 8. δ 2 T-Cell Expansion in Serum-Containing and Serum-Free Media

The growth and rate of expansion of different $\delta 2$ T-cells was evaluated in serum-containing media (R2:RPMI + 10% FBS), and serum-free media (AIMV + bovine albumin; CTS serum-free Supplement). **FIG. 5** depicts a graph illustrating the growth of $\delta 2$ T-cells. All media used in the current experiment contained 100 IU/mL IL-2, 2 mM glutamine and 1x penicillin/streptomycin. In addition, cells were stimulated with Zoledronic acid at 1, 5, and 20 μ M on day 0. Media was replenished every 2-3 days without further addition of Zoledronic acid. The total number $\delta 2$ T-cells expanded from 10^6 PBMC and the fold expansion of each treatment after a time period of 13 days are shown in **FIG. 5**. These results indicate that $\delta 2$ T-cells can be expanded in serum-free media.

Example 9. Anti- $\gamma\delta$ TCR Antibody Blocking and Competition Assays

FIG. 6 and **FIG. 7** depict graphs illustrating anti- $\gamma\delta$ TCR antibody blocking experiments and **FIG. 8** and **FIG. 9** depict antibody competition assays. **FIG. 6** and **FIG. 7** illustrate the results of blocking experiments where PBMCs were pre-incubated with various antibodies, namely, 5A6.E9, B1, TS8.2, 15D, B3, B6, TS-1, $\gamma 3.20$, IMMU510, or 11F2. The cells were subsequently washed and stained with the secondary TS8.2-FITC ($\delta 1$ -specific) or B6-PE ($\delta 2$ -specific) antibodies. PBMCs samples were analyzed by flow cytometry. Reduction in geometric mean fluoresce intensity (gMFI) was used to assess degree of blocking. The level of inhibition is illustrated against TS8.2-FITC (**FIG. 6**) and against B6-PE (**FIG. 7**).

Competition studies between MAbs TS-1 and TS8.2 and antibodies 5A6.E9, B1, IMMU510, R9.12-2, or 11F2 for binding on $\gamma\delta 1$ TCR-expressing cell line BE13 were performed by incubating 1×10^5 cells with 1, 2 or 10 μ g of unlabeled competing antibodies (IgG1, 5A6.E9, B1, TS8.2, TS-1, R9.12, IMMU510, or 11F2) and 0.2 μ g of FITC conjugated anti-V $\delta 1$ TCR clone TS8.2 (**FIG. 8**) or anti-V $\delta 1$ TCR clone TS-1 (**FIG. 9**) simultaneously on ice for 30 minutes. % Competition is calculated by change in geometric mean fluorescence divided by the maximum change in geometric mean fluorescence. As depicted in **FIG. 9**, TS-1 antibody competed with TS8.2 binding to the cells as effectively as the TS8.2 antibody itself. None of the other antibodies tested were able to compete with TS8.2 binding. TS8.2 antibody competed with TS-1 binding to the cells but not as effectively as the TS-1 itself. Some level of competition with TS-1 binding was also observed with 11F2 antibody. These results indicate that both TS-1 and TS8.2 antibodies bind to $\gamma\delta 1$ but likely not at the same epitope.

Example 10. Enzymatic Digestion of Tumor Specimens and $\gamma\delta$ T-cell Expansion with Antibodies to Specific $\gamma\delta$ Epitopes

24 well plates were coated with 0.5-1 μg anti- $\gamma\delta$ TCR antibodies. Cells isolated from digested tumor tissues as described in example 4 were counted and seeded on antibody coated wells at 0.5-1 X 10⁶ cell/ml in RPMI 1640 medium supplemented with 10% human AB serum and rhIL-2 (100 IU/mL). The cultures were incubated at 37⁰ C, 5% CO₂ for 7-21 days.

Example 11. Enzymatic Digestion of Tumor Specimens and $\gamma\delta$ T-cell Expansion with Antibodies to Specific $\gamma\delta$ Epitopes

Cells isolated from digested tumor tissues as described in example 1 in section “Fresh Tumor Specimens Enzymatic Digestion” were counted and seeded on antibody coated wells of a 3D cell culture plate (Corning® Costar® Ultra-Low attachment multi well plates) at 0.5-1 X 10⁶ cell/ml in RPMI 1640 medium supplemented with 10% human AB serum and rhIL-2 (100 IU/mL). The cultures were incubated at 37⁰ C, 5% CO₂ for 7-21 days.

Example 12. Activation and Expansion of $\gamma\delta$ T-Cells from PBMC

Activating agents were tested either as soluble agents, or agents immobilized on the culture wells. Soluble antigens and antibodies were added at a final concentration of 0.1-5 $\mu\text{g}/\text{ml}$ to human PBMCs cultured in 24 well plate at a cell density of 1 X 10⁶ cell/ml. Alternatively, the same anti $\gamma\delta$ TCR antibodies were immobilized by coating wells of 24-well culture plates. Anti $\gamma\delta$ TCR antibodies were added at 0.1-10 $\mu\text{g}/\text{ml}$ concentration. Wells were washed twice with PBS, then PBMCs were transferred to the plates and cultured in either RPMI-1640, AIM-V or CTS-OpTmizer media as described above. Cultured media was supplemented with 100 IU/mL of rhIL-2.

In a specific example, one million PBMCs/ml from donor B3 were stimulated on Day-0 with various antibodies immobilized at 0.5, 1, and 2 μg per well in 24-well plates. The antibodies tested were Mouse IgG1 Isotype control clone MG1-45 (Bio Legend), UCHT-1, 5A6.E9, B1, TS8.2, 15D, B6, B3, TS-1, γ 3.20, 7A5, and Zoledronate. **FIG. 10** and **FIG. 11** depict graphs illustrating activation and expansion of δ 1 and δ 2 T-cells respectively from PBMC. Cells were activated and expanded in media containing RPMI with 10% FBS, 100 IU/mL rhIL-2, glutamine and 1x penicillin streptomycin. On Day-7 after the initial stimulation, cells were passaged in fresh media and placed

in a newly coated 24-well plate with the same antibodies at the same concentrations. Media in the re-stimulated cultures were replenished every 2-3 days until Day 13 and analyzed by flow cytometry.

FIG. 12 illustrates the total number of $\delta 1$ T-cells, and **FIG. 13** illustrates the total number of $\delta 2$ T-cells after 13 days of growth and expansion. The total number of $\gamma\delta$ T-cells was calculated by multiplying the percentage $\delta 1$ and $\delta 2$ T-cells (as determined by flow cytometry using TS8.2-FITC and B6-PE, respectively) by the total viable cell number and subtracting the negative control values from $\delta 1$ and $\delta 2$ T-cell cells activated by the non-specific mouse IgG MG1-45 (BioLegend 401403).

Activation for cell expansion was obtained only when the antibodies were immobilized on the culture plates, with no detected expansion when these antibodies were added to the culture in soluble form, including in whole PBMC cell population (data not shown). Pan $\gamma\delta$ TCR MAbs 5A6.E9 and B1 and activated the growth of both $\delta 1$ and $\delta 2$ cell populations. MAbs 15D and B6 induced selective growth of $\delta 2$ cell population. MAbs TS8.2 and TS-1 induced selective growth of $\delta 1$ cell population. Of interest is that although MAbs TS-1 and TS8.2 compete with each other in binding to cell surface TCR, TS-1 induced proliferation was 3-fold higher. Similarly, different extent of $\delta 2$ cell population proliferation induction was detected between antibodies B1, 5A6.E9, 15D, B6, and B3. This data indicates that unique epitopes are required to trigger a specific and robust expansion of $\gamma\delta$ cell populations.

Example 13: Activation and Expansion of Specific $\gamma\delta$ T Cell Populations from PBMCs

Activation and Expansion of V $\delta 1$ T-Cells from PBMCs

Activating antibodies (TS1, TS8.2, R9.12, B6 and 15D) were immobilized directly to plastic 24-well plates at 0.25 or 1 $\mu\text{g/mL}$ (data shown for 0.25 $\mu\text{g/mL}$) or captured at 0.05 or 0.1 mg/mL (data shown for 0.05 $\mu\text{g/mL}$) by plate-bound goat anti-mouse IgGFc (5 $\mu\text{g/mL}$) in 24-well plates. Human PBMCs were activated at 1×10^6 cell/ml in media containing RPMI with 10% FBS, 100 IU/mL rhIL-2. Media in the cultures were replenished every 2-3 days. On day-7 cells were transferred into new plates without activating antibodies and further expanded by replenishing fresh media every 2-3 days for a total of 14 or 23 days. Cell count and flow cytometry analysis were conducted on day 0, 7, 14 and 23 to determine percentage and number of V $\delta 1$ and V $\delta 2$ T-cells. Fold expansion is defined as the number of the $\gamma\delta$ T-cells divided by the number the same $\gamma\delta$ T-cell type

in the starting PBMCs plated in each well. **FIG. 16** depicts graphs illustrating activation and expansion of V δ 1 T-cells from PBMC from different donors. MAbs TS-1, R9.12 and TS8.2 induced significant selective growth and enrichment of V δ 1 T-cell population. V δ 1 T-cells were expanded from 400 to 14,500 fold (**FIG. 16A, C**) and enriched from 1.2% up to 87.0% of the T lymphocytes population, which comprised about 95% of the total cell population (**FIG. 16B**) in 14 days. V δ 1 T-cells were expanded by 26,330 fold (**FIG. 16D**) in 23 days.

Activation and Expansion of V δ 2 T-Cells from PBMCs

Activating antibodies (TS1, TS8.2, R9.12, B6 and 15D) were immobilized directly to plastic 24-well plates at 0.25 or 1 μ g/mL (data shown for 0.25 μ g/mL) or captured at 0.05 or 0.1 μ g/mL (data shown for 0.05 μ g/mL) by plate-bound goat anti-mouse IgGFc (5 μ g/mL). Human PBMCs were activated at 1×10^6 cell/ml in media containing RPMI with 10% FBS, 100 IU/mL rhIL-2. Media in the cultures were replenished every 2-3 days. On day-7 cells were transferred into new plates without activating antibodies and further expanded by replenishing fresh media every 2-3 days for a total of 14 days. **FIG. 17** illustrate the fold expansion and percentage of V δ 2 T-cells after 14 days from different donors. MAbs B6 and 15D induced selective expansion of V δ 2 T-cell population from 70 fold to 4,740 fold (**FIG. 17A, 17C and 17E**) over 14-days and enriched δ 2 populations up to 80% of the T lymphocytes population, which comprised about 95% of the total cell population (**FIG. 17B and 17D**).

Reduction in the percentage of $\alpha\beta$ T-cells in $\gamma\delta$ TCR activated PBMC

Immobilized activating agents were tested in cell culture wells. MAbs (TS1, TS8.2, B6 and 15D) were directly coated (1 μ g/mL) or captured (0.1 μ g/mL) with goat-anti-mouse Fc (5 μ g/mL) in 24-well plates. Human PBMC were activated at 1×10^6 cell/ml in media containing RPMI with 10% FBS, 100 IU/mL rhIL-2. Media in the cultures were replenished every 2-3 days. On day-7, cells were transferred into new plates without antibodies and further expanded by replenishing media every 2-3 days until day 14 and analyzed by flow cytometry. **FIG. 18** illustrates the fold expansion and percentage of $\alpha\beta$ T-cells after 14 days. PBMCs cultured with MAbs TS1, TS8.2, B6 and 15D led to significant reduction in $\alpha\beta$ T-cells from 92% to as low as 9% over the 14-day culture period.

Activation with $\delta 1$ and $\delta 2$ specific antibodies reduces the population doubling time for $\delta 1$ and $\delta 2$ T-cells

MAbs were directly coated (1 $\mu\text{g/mL}$) or captured (0.1 $\mu\text{g/mL}$) with goat-anti-mouse Fc (5 $\mu\text{g/mL}$) in 24-well plates. PBMCs were plated at 10^6 cells/mL in RPMI with 10% FBS and 100 IU/mL IL-2. Cells were transferred to new plate without antibodies on day 7, adjusted to 10^6 cells/mL with fresh media. Culture was transferred to a new plate without antibodies and adjusted to 10^6 cells/mL on day 7 and replenished with fresh media every 2-3 days. The results are depicted in Fig. 19. V $\delta 1$ T-cells population doubling time was reduced from 37 hours in the absence of activating antibody to 24 hours in the presence of immobilized TS1 antibody. V $\delta 2$ T-cells population doubling time was reduced from 125 hours in the absence of activating antibody to 27.5 hours when activated with immobilized 15D antibody. Alpha beta T-cells population doubling time was not affected significantly by $\gamma\delta$ TCR antibody activation.

Activation using TS8.2 and TS1 antibodies results in predominately naïve and central memory phenotypes

Antibodies TS8, R9.12 and Immuno510 were immobilized directly to plastic wells at a concentration of 1.0 $\mu\text{g/mL}$. Human PBMCs were activated at 1×10^6 cell/ml and cultured in media containing RPMI with 10% FBS, 100 IU/mL rhIL-2. Media in the cultures were replenished every 2-3 days until Day 23 and analyzed by flow cytometry on days 0, 14 and 23. Fig. 20 depicts phenotype of V $\delta 1$ T-cells on days 14 and 23 as determined by CD45RA and CD27 expression analysis by flow cytometry. Phenotypes of the $\gamma\delta$ cells were defined as naïve (CD45RA+/CD27+), central memory (TCM, CD45RA-/CD27+), effector memory (TEM, CD45RA-/CD27-) and effector memory expressing (TEMRA, CD45RA+/CD27-). Activation with TS8.2 and R9.12 maintained the V $\delta 1$ T-cell phenotype over the 23-day expansion period with 43% to 63% naïve populations and 23% to 27% central memory populations. In contrast, activation with pan- $\gamma\delta$ antibody Immuno510 resulted in the reduction in the naïve V $\delta 1$ -T cells from 47% to between 8% and 33%.

Specific enhanced expansion of V $\delta 1$ T-cell population by MICA-Fc

Human PBMCs were activated with plate-bound TS8.2 antibody (1 $\mu\text{g/mL}$) or plate-bound TS8.2 and MICA-Fc fusion protein (1 and 5 $\mu\text{g/mL}$, respectively). Cultures were expanded in RPMI

media containing 10% FBS, 100 IU/mL rhIL-2. **Fig. 21** depicts cell expansion after 10 days. Combination of TS8.2 with MICA-Fc enhanced the expansion of V δ 1 T-cell population by 2 fold over TS8.2-only control (A) with no effect on $\alpha\beta$ T-cell population (B).

5 **Specific and significant activation of δ 1 and δ 2 T-cells from umbilical cord blood**

Mononuclear cells from human umbilical cord blood were isolated by density gradient centrifugation using Ficoll and activated with antibodies TS8.2, R9.12, B6 and 15D in media containing RPMI, 10% FBS, 100 IU/mL rhIL-2. **Fig. 22** shows that V δ 1 T-cells were expanded by up to 152 fold using δ 1-specific antibodies and V δ 2 T-cells were expanded by up to 93 fold using
10 δ 2-specific antibodies over 7 days.

Example 14. Epitope mapping of activating $\gamma\delta$ TCR MAb

Epitope binding domains of $\gamma\delta$ TCR activating antibodies were determined. The binding epitope of $\gamma\delta$ TCR δ 1 specific activating MAb TS8.2, TS1 and the δ 2 specific activating MAb 15D
15 and B6 was determined in ELISA binding assays using different combinations of wild type and mutated $\gamma\delta$ TCR paired chains. TS1, TS8.2 and R9.12 are δ 1 specific MAb that bind the surface of human T-leukemia cell line BE13 expressing δ 1 TCR as detected by flow cytometry (data not shown). The BE13 TCR chains δ 1J1 and γ 8J2 were cloned and used for epitope mapping of δ 1 specific MAb.

20 Soluble TCR constructs containing the extracellular V, diversity (D), joining (J), and C regions domains of δ and γ chains were cloned and fused to the hinge region, CH2 and CH3 domains of human IgG1 heavy chain Fusion $\gamma\delta$ TCR-FC proteins expressed by transient transfection of 293 cells.

For the mapping of δ 1 specific MAb, different δ 1 chains were expressed to include V δ 1J1,
25 V δ 1J2 and V δ 1J3. Additional V δ 1J1 chains mutated at the V and J regions were generated. All of the different δ 1 chains were paired with the γ 8 TCR chain cloned out the BE13 cell line, the binding of δ 1 specific MAb were not affected by the pairing with γ chains.

The different $\gamma\delta$ TCRs were co- transfected into 293 cells and were able to assemble correctly and to form disulfide-linked, heterodimers secreted as correctly paired receptor chains.

The correct folding of the TCR $\gamma\delta$ -FC heterodimers was confirmed by an ELISA binding assay using the anti-TCR pan $\gamma\delta$ antibodies including B1.1 (Biolegend #331202), 5E6.E9 (ThermoFisher Scientific #TCR1061), 11F2 (Beckman Coulter #340884) and IMM510 (Beckman Coulter #IM1349) antibodies which recognize all $\gamma\delta$ T cell receptors.

5 In all cases binding of all $\gamma\delta$ TCR MAbs was restricted to the $\gamma\delta$ TCR heterodimers indicating that the pairs of the $\delta 1$ and $\delta 2$ chains with γ chain is necessary for the proper binding structure.

Transient Transfection of 293 Cells and Production of Chimeric Proteins. 239 cells were grown in
10 Dulbecco's modified Eagle medium (DMEM) supplemented with 10% fetal calf serum. Adherent 293 cell were plated on 24 well plates the day before transfection, at 1×10^5 cells/well, and incubated at 37°C (5% CO₂) overnight. One μ g of each TCR γ -Fc and TCR δ -FC plasmids were combined. Plasmid DNA and fectin293 transfection reagent (ThermoFisher Scientific # 12347019) were diluted in serum-free Opti-MEM media for 5 minutes, before complexing at room temperature for 20-30
15 minutes. The entire volume of transfection mixture was added drop wise to prepared cells. Transfected 293 cells were incubated for 48–72 hours, at 37°C (5% CO₂). Cell supernatants were collected at 48 hours post transfection and tested for binding to selected anti human $\gamma\delta$ TCR monoclonal antibodies by ELISA assay.

20 Sandwich ELISA Assay. Cell culture supernatants were tested by enzyme-linked immunosorbent assay (ELISA). Assay plates (Greiner Bio-One high binding microplates) were incubated overnight at 4°C with 100 μ l of 1 μ g/ml Goat Anti-Human IgG, Fc γ fragment specific antibody (Jackson ImmunoResearch # 109-005-008). Plates were blocked for 1 h at room temperature with 200 μ l/well of blocking buffer (PBS containing 3% BSA). Supernatants containing soluble $\gamma\delta$ TCR were added in
25 PBS-Tween (Tween at 0.5 ml/L in PBS) and incubated at room temperature for 1 h, followed by washing and incubation with selected mouse anti human $\gamma\delta$ TCR specific mabs. Binding of $\gamma\delta$ TCR mAbs to the set of soluble TCRs was detected using HRP conjugated Goat anti mouse FC specific (Jackson ImmunoResearch Laboratories; Peroxidase-AffiniPure Goat Anti-Mouse IgG, Fc γ Fragment Specific # 115-035-008) diluted to 1: 10,000 in blocking buffer for one hour. The plate

was then washed and developed with TMB reagent. Color changes on the plates were measured on a Victor X3 plate reader (Perkin Elmer) at wavelength 450 nm.

As shown in **FIG. 23**, binding of TS1 and TS8.2 to soluble TCRs was detected when the $\delta 1$ chain include V $\delta 1J1$ and V $\delta 1J2$ sequences but not to the V $\delta 1J3$ chain, indicating that the binding of TS1 and TS8.2 involved critical residues in the delta J1 and delta J2 region, that are missing in the delta J3. Data indicates that TS1 and TS8.2 bind $\delta 1J1$ and $\delta 1J2$ TCR, and R9.12 binds $\delta 1J1$ $\delta 1J2$ and $\delta 1J3$ TCR, and that J1 and J2 segments contain sequences that are critical for TS1 and TS8.2 binding.

To identify the critical amino acid(s) in the J1/J2 regions necessary for the TS8.2 and TS1 binding, sequence alignment of human delta J region pointed to 4 possible residues 9 J1/J2 residues Leu116, Lys120 and Thr122 that are shared between J1 and J2 but different in the J3 sequence.

The $\delta J1$ sequence was modified according to the J3 sequence at the selected positions; Leu116 was modified to Met, Lys 120 was modified to Thr and Ala, and Thr122 was replaced by Ile. Change of Lys120 by Thr abolished the binding of both TS8.2 and TS1 but not binding of R9.12. As shown in FIGS. 24 and 25, the Lysine residue at position 120 at the delta J1 and delta J2 is critical for TS-1 and TS8.2 antibody binding.

To further identify V $\delta 1$ sequences important for TS8.2 and TS1 binding, additional mutated V $\delta 1$ TCR constructs were made. As shown in **Fig 26**, six mutated V $\delta 1$ chains were constructed. The mutations were designed based on the differences in amino acid sequences between the highly homologous bovine V $\delta 1S1$ and the human V $\delta 1$ (68% identity). Mutated V $\delta 1$ chains were generated using synthetic genes (Integrated DNA Technologies). Soluble TCRS were expressed as V $\delta 1$ chains paired with human $\gamma 8$ TCR Fc chain.

As shown in **FIG 27**, TS-1 and TS8.2 antibodies failed to bind to the two of the mutated V $\delta 1$ chains, V $\delta 1$ Mut1 and V $\delta 1$ Mut6. The V $\delta 1$ Mut1 displays 9 residue changes in the stretch of residues between the region of Gly57 to Ala88 of the variable region of human $\delta 1$ chain, and V $\delta 1$ Mut6 displays a change in two residues at position 71 and 72 (numbering as shown in **FIG. 14**). The binding of V $\delta 1$ MAb R9.12 and pan δ chain constant MAb Immuno510 were not affected by any of the changes to the V $\delta 1$. Therefore, the V $\delta 1$ residues at position Arg71 and Asp72 together with Lys120 at The J1/J2 region are critical for binding of both Ts-1 and TS8.2 MABs.

Taken together, the data indicate that the critical binding and activating epitope of both TS-1 and TS8.2 is located within the V and J1/J2 regions of the $\delta 1$ chain.

Epitope Mapping of Vδ2 specific activating MAbs 15D and B6

Specific binding of MAbs B6 and 15D to the δ2 T cell population activated by zoledronic acid was detected by flow cytometry, while no binding was detected to the BE13 cell line expressing δ1TCR (data not shown). Competition binding assay data also showed that B6 and 15D do not compete with each other and recognize different epitopes.

γδ TCR chain combinations. To identify the binding epitope for 15D and B6 MAbs, different γδ2 TCR constructs were generated. Vδ2 chain was co-transfected with γ3, γ4, γ8, γ9J1 and γ9JP FC fusion chains. The correct folding of the secreted δ2 TCR heterodimer was confirmed by an ELISA binding assay using the anti-TCR PAN γδ antibody IMMU510 (Backman Coulter #IM1349) and with the γ9 specific MAb the 7A5 (ThermoFisher Scientific #TCR1720).

Binding of B6 was affected by the γchain selected for pairing with the δ2 chain. Binding of B6 was detected when the δ2 chain was paired with the γ9 chain regardless of the γ9 J sequence. The binding to the γ3δ2 TCR pair was greatly reduced, and the binding to the δ2 chain paired with γ8 was undetectable. As shown in **FIG. 28**, the most stabilized binding of the B6 MAb is to the δ2γ9 TCR. Both γ3 and γ8 belong to gamma family 1 and share high sequence homology (75%) with each other, γ9 a member of the gamma 2 family and shares low sequence homology with γ3 and γ8 (~20%). High binding affinity of B6 involves residues on both δ2 and γ9 chains. Positive control MAbs were IMMU510 a pan γδTCR that binds the constant region of the δchain and recognize all γδTCRs, and 7A5 MAb a γ9 specific MAb. Binding of the B6 MAb was detected to the δ2γ9 TCR. B3 is a Vγ9 antibody.

Recently, it was shown that human and Rhesus γδ2 T cells show a high degree of germline sequence homology of the TCR gene segments and the diversity of the CDR3 regions (Daubenberger CA, et al., J Immunol 2001; 167:6421–30). To identify critical sequences that are required for the binding of 15D, specific point mutations were introduced into the CDRs domain of human Vδ2 chain, and were replaced by the rhesus monkey sequence at positions Gly35 (Vδ2 CDR1) Asp65 (Vδ2 CDR2) and Cys104 (Vδ2 CDR3), by site directed mutagenesis. **FIG 29** shows the protein sequence alignment of human (IMGT Human TRDV2) and Rhesus monkey (GenBank:

AY190028.1) V δ 2 variable regions. The changes were made in CDR1 (G35S), CDR2 (D65G) and the CDR3 C104S (numbering as shown in **FIG. 15**). The replacement of a single residue at the CDR1 of human V δ 2 from Gly to Ser led to complete loss of binding activity by 15D. As shown in **FIG 30**, Gly35 was identified as a critical residue for the binding of 15D. Changes in CDR1, CDR2 and CDR3 of V δ 2 did not affect the binding of B6, IMMU510 a $\gamma\delta$ TCR pan MAb and 7A5 a γ 9 specific Mab.

Example 15. Epitope Mapping of Novel Modulators of $\gamma\delta$ T-cell Activation and Expansion

CD2 epitopes:

The CD2 molecule consists of two extracellular immunoglobulin superfamily domains Ig-like V-type domain and Ig-like C-type domain, on which three major immunogenic regions have been described (Davis et al., Immunol Today 1996; 17: 177–187). Regions 1 and 2 are located in the first domain and region 3 is located in the second domain. MAbs that recognize region 1 bind both resting and activated T-cells and can strongly inhibit the binding of CD58 to CD2. Monoclonal antibodies that recognize region 2 have similar binding properties but are not effective at blocking CD58 binding. Monoclonal antibodies that recognize region 3 recognize CD2 only on activated T-cells and do not block CD58 binding.

Mapping of the binding epitope of CD2 agonist MAbs is done by competition assays against CD58, the natural ligand of CD2 and activation assays. In addition, site directed mutagenesis of critical CD2 binding residues is that leads to loss of binding of CD58, is tested for MAbs binding. Examples of mutations include mutagenesis of the CD2 sequence at position 67 (K to R), 70 (Q to K), 110 (Y to D) and at position 111 (D to H). To design constructs for the mapping of the binding epitope of CD2 a sequence alignment of human and mouse CD2 using VectorNTI (Thermo Fisher) are performed. The alignments show a 50% sequence homology between human and mouse CD2 . MAbs that bind the extracellular domain of human CD2 are not expected to cross react with the mouse CD2. To identify the binding epitopes, we generate domain swapping of mouse/human chimeric CD2 constructs. In such constructs, the human N-terminal Ig-like V-type domain (residues 25-128) is replaced with mouse residues (23-121) in expression vector expressing the extra cellular domain of CD2. Chimeric cDNA species are transiently transfected into CHO or 293 cells. Human wildtype CD2 and human mouse chimeric constructs are expressed on the surface of CHO cells.

Chimeric CD2 Cell-surface expression is analyzed by FACScanto. Binding or loss of binding to chimeric molecule is detected by flow cytometry and validated by activation assays.

NKG2D epitopes:

Epitope mapping of activating MAb against NKG2D is tested by the ability of anti-hNKG2D antibody to reduce or block NKG2D interactions with one or more of its ligands, (MICA/MICB) or by competition with an antibody known to block hNKG2D ligand interaction. Agonist MAb is identified by its ability to reduce NKG2D-mediated activation of NK or T-cells. This can be evaluated by typical cytotoxicity assays, addition of ligand-blocking antibodies, or by blocking $\gamma\delta$ T-cells mediated killing of MICA-expressing tumor cells in a dose-dependent fashion.

CD27 epitopes:

Epitope mapping of agonist antibodies against CD27 is done by functional assay and their ability to block the interaction between human CD27 and its natural ligand human CD70. CD27 expressing cells are incubated for 30 min with 5 μ g of MAb at 4 °C, after which 1 μ g of biotinylated CD70 is added to the tube. The mixture is incubated for another 15 min at 4 °C before washing. Next, cells are incubated for 30 min with a mixture of streptavidin-PE (for detection of CD70 ligand binding) followed by several washing steps and analysis using FACS. Blockage of CD70 binding indicates that the MAb and CD70 share the same epitope.

Example 16: Characterization of TCR V δ Repertoire of a Population of Expanded $\gamma\delta$ T-cells

Clonal diversity of expanded $\gamma\delta$ T-cells from individual donors is assessed with polymerase chain reaction (PCR). The clonal diversity of freshly isolated PBMC from healthy donors is compared to the clonal diversity of cultures that are stimulated for 7 and 13 days with different activation agents described herein, including anti-TCR specific antibodies, ligand, and lectins.

RNA from the aforementioned $\gamma\delta$ T-cells is extracted, reverse transcribed into cDNA, and amplified by PCR using V δ 1, V δ 2, and V δ 3 -specific primer pairs. V δ 1 forward primer (5'ATGCTGTICTCCAGCCTGCTGTGTGTATTT 3'; SEQ ID NO: 1) V δ 2 forward primer (5'ATGCAGAGGATCTCCTCCCTCATCCATCT 3'; SEQ ID NO: 2) and V δ 3 forward primer (5'ATGATTCTTACTGTGGGCTTTAGCTTTTTG 3'; SEQ ID NO: 3) were used in combination

with A C δ reverse primer (5'CTTGGGGTAGAATTCCTTCACCAGACAAGC3'; SEQ ID NO: 4) to amplify 500 bp DNA fragments.

The γ chain was amplified in separated RT-PCR reaction using V γ specific primers to amplify V γ 2, V γ 3, V γ 4, and V γ 5 (5' GGTGCTTCTAGCTTTCCTGTCTCCTGC3' (SEQ ID NO: 10)), V γ 8 (5'ATGCTGTTGGCTCTAGCTCTGCTTCTA3' (SEQ ID NO: 11)) and V γ 9 (5'ATGCTGTCACTGCTCCACACATCAACG3' (SEQ ID NO: 12)) V γ primers are paired with the C γ reverse primer (5'GGAAGAAAAATAGTGGGCTTGGGGGAA3' (SEQ ID NO: 13)). The γ reverse primer based on consensus sequence of the C γ 1 and C γ 2. PCR fragments are cloned and the nucleotide sequence of 80 independent V γ inserts are obtained. The V δ , D δ and J δ junctional sequences are analyzed, together with CDR3 sequence alignment.

Example 17: MABs and Related Targeting Constructs for Engineering of $\gamma\delta$ T-Cells

The human CD20 gene is obtained from a cDNA clone (Origene Technologies, Inc., 6 Taft Court, Suite 100, Rockville, Md. 20850). The CD20 gene is amplified using the forward primer (5' ATGACAACACCCAGAAATTCAGTAAATGG3' (SEQ ID NO: 14)) and the reverse primer (5' TCAAGGAGAGCTGTCATTTTCTATTGGTG3' (SEQ ID NO: 15)) the amplified CD20 cDNA is incorporated into pCDNA3.4 (Thermo Fisher Scientific) as a high expression vector for mammalian cells, and transfected into CHO cells as the host cells. Recombinant CHO cells (CD20/CHO cells) expressing CD20 molecules at a high level on their cell surfaces are identified by FACS analysis.

The CD20/CHO cells are used to immunize BALB/C mice or transgenic mice engineered to produce fully human antibodies as described in Jakobovits and Bornstein (Jakobovits Curr. Opin. Biotechnol. 1995 6:561-6; Bornstein et al., Invest New Drugs. 2010 28:561-74.). Antibodies with high affinity and specificity towards human CD20 are screened by FACS assays. Mouse antibodies are humanized by CDR grafting (Kim and Hong Methods Mol Biol. 2012;907:237-45. Human single domain CD20 antibodies are also generated from mice or rats engineered to produce human heavy chain single domain antibodies (Janssens et al., PNAS 2006 vol. 103:15130).

The gene coding for the high affinity/specificity CD20 antibody described above is cloned into the MSGV1 retroviral vector backbone or the pCAG lentiviral vector. VH and VL domains are cloned either from the mouse hybridoma expressing the selected MAb, or from humanized antibody chains. $\gamma\delta$ T-Cells are engineered with the CD20 MABs described herein.

Example 18: TCR Constructs for Engineering of $\gamma\delta$ T-Cells

Constructs expressing TCRs comprising sequences of highly reactive $\alpha\beta$ TCR chains are isolated from T-lymphocytes expressing NY-ESO-1-MHC specific peptide complexes. The NY-ESO-1-MHC specific peptide complexes induce potent *in vitro* and *in vivo* anti-tumor activity
 5 against various NYESO-1-expressing tumors.

The highly reactive $\alpha\beta$ TCR chains are isolated from melanoma, sarcoma patients or from mice bearing patient-derived xenografts derived from human melanoma or sarcoma tumors. Alternatively, the TCR is derived from mice altered to have a humanized immune system that expresses NY-ESO-1 peptides or NYESO-1-peptide complexes (*See* Gonzales et al., Immunol Res.
 10 2013 57: 326-334; Boucherma et al., J Immuno. 2013. 191; 583-593; Liang-PingL. Et al., Nature Med. 2010, 16:1029-1035). T-cells that recognize epitopes of the NY-ESO-1-in the context of the dominant class I alleles HLA-A*02 (for example, peptide SLLMWITQC Residues 157-167(SEQ ID NO: 16)) and dominant HLA-A*01-associated peptide are identified.

Sequences of TCR α and TCR β transcripts are generated by reverse transcription-polymerase
 15 chain reaction (RT-PCR) using the One step RT-PCR kit (Qiagen Hilden Germany) according to the manufacturer's suggestions. First strand cDNA is generated from RNA isolated from the reactive T-cell. Total RNA is extracted with TRIzol Total RNA Isolation Reagent (Invitrogen Life Technologies) from CTL clones. Amplification of TCR α and β chains is done by a set of degenerate primers that can bind to highly conserved region of the TCR α and β chain V regions
 20 centered around the tryptophan-tyrosine residues at amino acid positions 34 and 35 (Kabat numbering) are used in combination with the α and β constant region reverse primers (Moonka and Loh Journal of Immunological Methods 169 (1994) 41-51). Amplified PCR fragments are gel purified and directly sequenced. Sequence information is used to design PCR primers suitable for cloning of the individual full-length cDNAs.

25 TCRs genes are cloned and inserted into MSGV-based retroviral vectors and full-length cDNAs amplified from selected T cells. Retroviral vectors encoding both α and β chains of wild-type NY-ESO-1-reactive human TCR are constructed using a MSGV1 backbone. Linking of the TCR α and TCR β chains is done via an internal ribosome entry site (IRES) element in one construct, or by separation using a cleavable picorovirus peptide sequence.

Retroviral supernatants are generated by co-transfecting 293 cells that stably expressed MMLV gag and pol proteins with each MSGV1 TCR vector and a vector encoding the endogenous virus retroviral envelope protein using Lipofectamine 2000 (Invitrogen) as described previously or by electroporation using the Nucleofector (Lonza). Supernatants are collected at day 2 and 3 post-transfection and were diluted 1:1 with fresh DMEM containing 10% FCS. Engineered $\gamma\delta$ T-cells are capable of properly express the genes encoding the TCR α and β chains without any further manipulation.

Example 19: Engineering $\gamma\delta$ T-Cells with an $\alpha\beta$ TCR Construct

A polynucleotide comprising an $\alpha\beta$ TCR (tumor recognition moiety) is cloned from T-cells selected to be specific to the desired antigen using standard techniques. Isolated endogenous wild-type $\gamma\delta$ T-cells are grown with methods described in previous examples to at least 6×10^6 cells prior to infection with the retrovirus or a lentivirus comprising an expression cassette encoding the tumor recognition moiety. A standard protocol for viral infection can be used to introduce the vector system into the wild-type $\gamma\delta$ T-cell. Expression of the selection marker is used to select cells that have been successfully transfected.

The expression of the engineered $\alpha\beta$ TCR can be evaluated by flow cytometry and/or by quantitative QRT-PCR and by functional assays with target cells for cytotoxicity and cytokine secretion. The expression of the engineered activation domain can also be evaluated by flow cytometry and/or by quantitative qRT-PCR. The number of engineered $\gamma\delta$ T-cells expressing a cell surface marker of interest is determined by flow cytometry. The engineered $\gamma\delta$ T-cell is further engineered with a suitable methodology described herein, such as the CRISPR-Cas, talen, meganucleases, zinc finger, or sleeping beauty transposon technologies to delete an exon associated with an HLA gene or a $\beta 2M$ gene.

Example 20: Engineering $\gamma\delta$ T-Cells with CAR and TCR Constructs

$\gamma\delta$ T-cells are transduced with retro- or lenti- viral based vectors to express targeting moieties that can direct the engineered $\gamma\delta$ T-cells to specifically recognize tumor cells and get activated to kill it. The transduced targeting moieties include MAbs directed against tumor-specific surface proteins

or tumor-specific intracellular peptides. $\gamma\delta$ T-cells are also engineered with high affinity TCRs directed to peptide-MHC complexes.

Alternatively, cells can be engineered via transduction with non-viral vectors.

Example 21: Engineering of Isolated $\gamma\delta$ T-Cells with Targeting Moieties

CAR construct design comprises of different main functional domains, a target moiety that recognizes a protein or a MHC associated peptide of interest displayed on a tumor cell, a short spacer that connects the extracellular receptor targeting element to the transmembrane domain, which transverses the cell membrane and connects to the intracellular activation signaling domain.

The target moiety receptor expressed on the surface of a $\gamma\delta$ T-cell will be designed to specifically bind to a target protein that is expressed on a cancer cell. Tumor recognition moieties will be designed against targets including CD19, CD20, CD22, CD37, CD38, CD56, CD33, CD30, CD138, CD123, CD79b, CD70, CD75, CA6, GD2, alphafetoprotein (AFP), carcinoembryonic antigen (CEA), CEACAM5, CA-125, MUC-16, 5T4, NaPi2b, ROR1, ROR2, 5T4, PLIF, Her2/Neu, EGFRvIII, GPMNB, LIV-1, glycolipidF77, fibroblast activation protein (FAP), PSMA, STEAP-1, STEAP-2, mesothelin, c-Met, CSPG4, PVRL-4, VEGFR2, PSCA, folate binding protein/receptor, SLC44A4, Cripto, CTAG1B, AXL, IL-13R α 2, IL-3R, and SLTRK6.

The target moiety receptor can be derived from a portion of an antibody specific to tumor glycoprotein expressed on the surface of tumor cells, or alternatively the engineered receptor can be derived from a TCR receptor with a known specificity or an antibody recognizing a specific peptide sequence derived from intracellular tumor specific antigen presented on the on the surface in association with MHC complex including gp100, MART1, Tyrosinase, SSX2, SSX4, NYESO-1, epithelial tumor antigen (ETA), MAGEA family genes (such as MAGEA3, MAGEA4), KKLC1, mutated N, and K and H ras, BRAF, p53, MHC class I chain-related molecule A (MICA), MHC class I chain-related molecule B (MICB), or one or more antigens of HPV, EBV, or CMV. Such high affinity T-cell receptor like tumor recognition moieties will recognize peptide-MHC complex with a high degree of specificity.

Example 22: Targeting Moiety Constructs with a Spacer and a Transmembrane

Domain

Different spacers will be engineered into a tumor recognition moiety construct in order to optimize the potency of an engineered T-cell towards a cancer cell. The size of each spacer will vary according to the size of the target protein, the epitope recognized by the receptor, the size of the engineered tumor recognition moiety and the affinity of the receptor. Spacers that can accommodate conformational changes including sequences of human IgG, CD8a and CD4 hinge region.

Spacers that are tested consist of Gly, Ser, and Thr amino acids used at different lengths (from 19 to 9 residues) in order to provide chimeric receptor with improved binding affinity properties. The hinge and transmembrane portions of each construct is derived from the CD8a sequence (residues 117 to 178 of human CD8a) or alternatively the human IgG1 hinge-Fc cDNA with CD28 transmembrane domain (residues 153-179).

Example 23: Targeting Moiety Constructs with Co-Stimulatory Domains

Different co-stimulatory domains will be engineered into a construct comprising a tumor recognition moiety. A co-stimulatory domain comprising a CD28, 4-1BB, CD2, CD27, NKG2D, DAP10, DAP12, CD161, CD30, JAML, TLRs, CD244 or CD100 costimulatory signaling domain is engineered into a $\gamma\delta$ T-cell to mimic a “second signal” that amplifies the activation through the chimeric receptor, leading to a more robust signal to multiply and kill the cancer cell.

The cytoplasmic region is derived from the endodomains of $\alpha\beta$ and/or $\gamma\delta$ T-cell co-stimulatory molecules including: CD28 (residues 180-220), CD137 (residues 214-255), ICOS (residues 165-199) CD27 (residues 213-260) NKG2D (Residues 1-51) , JAML (residues 297 - 394) CD2 (residues 236 - 351), CD30 (residues 408 - 595) OX40 (Residues 1 - 23), HVEM (residues 224 - 283), or CD46 molecules. The optimal constructs are selected based on the degree of activation of the engineered $\gamma\delta$ T-cell populations for induced cell cytotoxicity and based on the degree of cytokine secretion *in-vitro* and *in-vivo*.

Example 24: Targeting Moieties Comprising CD3 ζ Activating Domain

The intracellular CD3 ζ (residues 52-164) containing three ITAM domains (ITAM1: APAYQQGQNQLYNELNLGRREEYDVLDKR, (SEQ ID NO: 5); ITAM 2: PQRRKNPQEGLYNELQKDKMAEAYSEIGM, (SEQ ID NO: 6); and ITAM3: ERRRGKGHDGLYQGLSTATKDTYDALHMQ, (SEQ ID NO: 7) was cloned.

The intracellular domain of TCR ζ was amplified using primers 5' AGAGTGAAGTTCAGCAGGAGCGCA -3' (SEQ ID NO: 8) and the reverse primer 5' CTCGAGTGGCTGTTAGCCAGA-3' (SEQ ID NO: 9).

The CAR constructs are generated by multistep overlap extension PCR. The products were fused in a separate PCR reaction driven by primers tailed with the Platinum Taq DNA Polymerase High Fidelity kit (Invitrogen), using the Overlap extension polymerase chain reaction protocol. The DNA encoding the full-length construct was ligated into MSGV1 Retroviral Vector. The construct provides a CAR targeting moiety comprising a CD3 ζ activating domain.

Example 25: Engineering $\gamma\delta$ T-Cells with More Than One Recognition Moiety

$\gamma\delta$ T-cells are transduced with more than one construct comprising a tumor recognition moiety, including a TCR and a MAb directed to the same intracellular tumor-specific protein. Each construct is selected to recognize a specific peptide in the context of different MHC haplotypes, such as A2 and A1, antibodies directed to different targets expressed on the same tumor cells or antibodies directed to different epitopes on the same target.

Example 26. In vitro Expansion of Engineered $\gamma\delta$ T-Cells

Engineered $\gamma\delta$ T-cells are grown and expanded with an appropriate tissue culture media, such as the tissue culture media described in previous examples. The Engineered $\gamma\delta$ T-cells are grown exponentially to about 1×10^6 in a 5% CO₂ incubator at 37⁰ C with or without stimulation by an external antigen and without co-culture with APCs or aminophosphates.

Example 27: Functional Characterization of Cytokines Released by Activated Engineered and non-Engineered $\gamma\delta$ T-cells

The expression of IFN- γ , TNF- α (R&D Systems), IL-1 β , IL-2, (Biosource International), IL-12 (Diacclone Research), IL-6, and IL-18 will be measured using commercial enzyme-linked immunosorbent assay (ELISA) kits. The enzyme-linked immunosorbent assays will be performed according to the manufacturers' instructions. The amount of cytokine will be measured at different time points (from 24 to 72 hours) in a polystyrene 96-well plate (Maxisorb, Nunc) coated with a monoclonal mouse IgG1 against the human cytokine, at a concentration of 1 μ g/ml in 0.05 M

sodium bicarbonate buffer overnight at 4⁰ C. After washing with PBS containing 0.05% Tween 20 the plate will be blocked with 3% bovine serum albumin (BSA, wt/vol, Sigma) in PBST for 1 h at 37⁰ C. A standard (recombinant human Cytokine from R&D) and supernatant from $\gamma\delta$ cultures samples will be added and the plate will be incubated at RT for 2 h. Detection is performed with a
5 matched antibody pairs in relation to recombinant human cytokine standards.

Example 28. Identifying Co-Stimulatory Agents

The ability of different co-stimulatory agents to support activation, expansion and viability of engineered and non-engineered $\gamma\delta$ T-cells is tested by adding co-stimulatory agents to whole PBMCs
10 or enriched engineered and non-engineered $\gamma\delta$ T-cell populations. Co-stimulatory agents are added in a soluble or immobilized form to different activating agents, including anti $\gamma\delta$ TCR specific MAbs. Human PBMCs purified from buffy coats of healthy donors as described in previous examples or lymphocytes isolated from tissues are plated at 2×10^6 in 1 mL of complete RPMI-1640 media supplemented with 100 IU/mL rhIL-2 in 24-well flat-bottom tissue with 2-10 μ g of anti $\gamma\delta$ TCR
15 antibody in the presence or absence of soluble or immobilized agonistic antibodies to CD2, CD27, CD28, CD30, CD137, ICOS, CD161, CD122, CD244, and NKG2D, or stimulating ligands including, CD70-FC (ligand to CD27) MICA, MICB and ULBP (ligands to NKG2D), 4-1BB (ligand to CD137), and Pilar 9 (ligand to CD161).

Example 29. Cytokine Support Activation

The ability of different cytokines to support activation, expansion and viability of engineered and non-engineered $\gamma\delta$ T-cells is tested by adding cytokines to whole PBMCs or enriched engineered and non-engineered $\gamma\delta$ T-cell populations. To test cytokine activation support, various cytokines are individually added to separate cell cultures every 3 days at 100 IU/mL. The cytokines that are tested
25 include IL-2, IL-7, IL-12, IL-15, IL-33, IL-21, IL-18, IL-19, IL-4, IL-9, IL-23, IFN- γ , and IL1 β . After the end of a select time period, a sample of cells is harvested and the composition of the cell population, i.e., percentages of $\gamma\delta$ T-cells, $\alpha\beta$ T-cells, B-cells, and NK cells is determined by flow cytometry.

Cells are kept in culture and expansion of select populations is tested at day 14 and day 21.

Example 30: Activation-derived V δ 1+, V δ 2+ T cell populations are cytotoxic to tumor cells

To determine the cytotoxicity of selectively activated, enriched δ 1 and δ 2 T-cell populations of the invention, PBMCs were isolated from buffy coat obtained from normal donors using Ficoll
 5 paque PLUS (GE Healthcare #17-1440-02) density gradient centrifugation and plated in 24-well plates at 1×10^6 cells/mL in R2 medium (10% fetal bovine serum in RPMI1640 supplemented with penicillin/streptomycin, L-glutamine and 100 IU/mL of human recombinant IL2).

On Day 0, cells were plated on wells coated with either 1 μ g/ml of δ 1 specific MAbs (TS1, TS8.2) or δ 2 specific MAbs (15D, B6) to activate $\gamma\delta$ cells within the PBMC population. 50%
 10 medium exchange was performed on day 3 and 5. On Day 7, activated cells were moved to 6-well culture dishes without any MAb coating, expanded and maintained at $\sim 1 \times 10^6$ cells/mL until day 14. To further the V δ 1+ and V δ 2+ activated T cell cultures, $\alpha\beta$ T cells were positively selected and depleted from δ 1 and δ 2 activated cultures using IP26 monoclonal antibody (Biolegend #306702) bound to anti-Mouse IgG MicroBeads (Miltenyi # 130-048-401). This led to highly enriched V δ 1+
 15 and V δ 2+ T cell populations, as shown in **FIG. 31**. Positively selected $\alpha\beta$ T cells were also collected ($\alpha\beta$ enriched). These enriched V δ 1+, V δ 2+ and $\alpha\beta$ T cell cultures were maintained in R2 medium until Day 22.

On day 22, $\gamma\delta$ and $\alpha\beta$ T cell composition of enriched cultures was determined by flow cytometry surface TCR staining using TS8.2, B6 and IP26 antibodies for δ 1, δ 2 and $\alpha\beta$, respectively.
 20 The highly enriched cultures contained 92%, 98% and 99% of V δ 1+ (TS8.2+), V δ 2+ (B6+) and $\alpha\beta$ (IP26+) T cells respectively.

In vitro cytotoxicity assay with V δ 1+, V δ 2+ and $\alpha\beta$ T cell populations

To assess cytotoxicity of the V δ 1+, V δ 2+ and $\alpha\beta$ T cell cultures, effector cells were
 25 incubated at a 10:1 ratio with target cell lines for 6 hours. Target cell lines tested in this assay included solid tumor cell lines BxPc3 (ATCC® CRL-1687™) pancreatic adenocarcinoma and SK-MEL-5 (ATCC® HTB-70™) human melanoma as well as hematological B tumor cell line RPMI 8226 (ATCC® CCL-155™). Cell death was measured using CytoTox Glo reagent (Promega Cat #). % Specific lysis was calculated using the following formula:

$$\% \text{ Specific Lysis} = \left(\frac{(T_{Exp}) - (T_{spon} + E_{spon})}{T_{100\%} - T_{spon}} \right) * 100$$

Highly enriched Vδ1+, Vδ2+ and αβ T cell cultures were tested for killing of BxPC3, SKMEL5 and RPMI8226 cell lines. As shown in **FIG. 32**, Vδ1+ T cells show killing of the epithelial tumor cell lines BxPC3 and SKMEL5 as well as the plasmacytoma cell line RPMI8226. (FIG 32A) while Vδ2+ T cells show potency mostly towards RPMI8226 and were less effective against the solid tumor cell lines. αβ T cells do not show any cytotoxic activity.

Example 31: Anti-Tumor Activity of Expanded γδ T-cells in-vitro

The cytotoxic activity against various tumor cell lines and primary tumor cells are tested at effector to target ratio between 1:1 to 40:1. Lysis of tumor cell lines and primary tumor cells are measured by detecting the release of intracellular enzymes lactate dehydrogenase. The percentage of γδ T-cells expressing engineered tumor recognition moieties in the culture is measured by flow cytometry, ELISA and/or ELISPOT assays.

Example 32: In vivo Anti-Tumor Activity

Cohorts of immune-deficient mice engrafted with human tumor xenografts, or huPBMC-NOG (Taconic) mice, or mice with humanized immune system as described above, are injected subcutaneously or orthotopically with cells derived from patient-derived tumor or tumor cell lines, including cancers of the colon, breast, ovary, kidney, head and neck, prostate, bladder, oral cavity, pancreas and liver, and allowed to reach an average size of 50-100mm³. Enriched or isolated γδ-T cells, either naïve or engineered, are injected intravenously into mice or directly into the tumor at a range of doses. Tumor regression is defined as a reduction in tumor volume after γδ-T cell dosing, and compared to untreated and standard of care for the specific indication. In some experiments naïve or engineered γδ T-cells are labeled with GFP or luciferase and injected to tumor-bearing mice to follow their persistence and homing. At the end of the study, tumors are harvested and GFP positive cells were analyzed by flow cytometry and immunohistochemistry.

Example 33. Polyclonal TCR diversity of Vδ1 and Vδ2 populations derived from antibody specific activation

TCR chains were amplified and cloned out of TS8.2 and B6 activated populations. PBMCs cultures were harvested after 14 days of specific activation with $\delta 1$ specific activating mAbs TS8.2 and the $\delta 2$ specific MAb B6. PBMCs cultures were lysed in 350 μ l RLT buffer (Qiagen,), Total RNA was purified from 5×10^5 cell using the Qiagen RNeasy Mini Kit (Qiagen, catalog number 74106), following the manufacturer's instructions. In brief, 350 μ l of 70% Ethanol was added to the cell lysate to provide ideal binding conditions. The lysate was then loaded onto the RNeasy silica membrane spin column. Contaminants were washed away. Concentrated RNA was eluted in 50 μ l water.

One step RT PCR: Full length delta chains were amplified using the 5' $\delta 1$ (5' GCCCAGAAGGTTACTCAAGCCAGTC3') and 5' $\delta 2$ (5' GCCATTGAGTTGGTGCCTGAACACC 3') primers, in combination with 3' human C δ (5' TTACAAGAAAAATAACTTGGCAGTCAAGAGAAA 3') primer. A 800 bp PCR fragments of the full length delta chain were cloned into the pCI expression vector (Promega catalog number E1841). Similarly, full length gamma chains were amplified using the 5' primers $\gamma 2, \gamma 3, \gamma 4$ (5' TCTTCCAACCTTGAAGGGAGAACGAAGTC 3') $\gamma 5$ (5' TCTTCCAACCTTGAAGGGGGAACGA 3'), $\gamma 8$ (5' TCTTCCAACCTTGAAGGGAGAACAAAGTC 3') $\gamma 9$ (5' GCAGGTCACCTAGAGCAACCTCAAATTTCC 3') in combination with a single 3' Cgamma primer (5' GGAAGAAAAATAGTGGGCTTGGGGGAA 3')

The delta and gamma transcripts were amplified from 100 ng total RNA using reverse transcriptase polymerase chain reaction (RT-PCR). A total of five RT-PCR reactions were run for each sample activated with the TS8.2 MAb ($\delta 1, \gamma 2-\gamma 4, \gamma 5, \gamma 8$ and $\gamma 9$) and two RT-PCR reactions were run for each sample activated with the B6 MAb ($\delta 2$, and $\gamma 9$). The QIAGEN One Step RT-PCR kit was used for amplification, (Qiagen, Inc.). This kit provides a blend of Sensiscript and Omniscript Reverse Transcriptases, HotStarTaq DNA Polymerase, dNTP mix, buffer and Q-Solution, a novel additive that enables efficient amplification of "difficult" (e.g., GC-rich) templates. Reaction mixtures were prepared that included 5 μ l of RNA, 0.5 of 100 μ M of either delta or gamma chains primers (custom synthesized by IDT), 5 μ l of 5 $\times$ RT-PCR buffer, 1 μ l dNTPs, 1 μ l of enzyme mix containing reverse transcriptase and DNA polymerase, and 0.4 μ l of ribonuclease inhibitor RNasin (1 unit). The reaction mixture contains all of the reagents required for both reverse

transcription and PCR. The thermal cycler program was RT step 50° C. for 30 minutes 95° C. for 15 minutes followed by 30 cycles of (95° C. for 30 seconds, 48° C. for 30 seconds, 72° C. for 1.0 minutes). There was then a final incubation at 72° C. for 10 minutes. The extracted PCR products were directly cloned into pCI expression vector. Nucleotide sequences were analyzed using IMGT
 5 to identify germline V, D and J gene members with the highest sequence homology.

Examples of sequences of V δ 1 chains activated by TS8.2 CDR3 (junctional diversity is indicated by bold text):

TS8.2 activated clone1 V δ 1, J1

10 AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQNAKS
 GRYSVNFKKAAKSVALTISALQLEDSAKYF**CALGPVVIPKGKLSFGKGTRVTVEP**

TS8.2 activated clone2 V δ 1, D3, J1

15 AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQNAKS.
 GRYSVNFKKAAKSVALTISALQLEDSAKYF**CALGELCLGDTYTDKLIFGKGTRVTVEP**

TS8.82 activated clone 3 V δ 1, D2+D3, J1

20 AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQNAKS
 GRYSVNFKKAAKSVALTISALQLEDSAKYF**CALGDPKVYWGCTDKLIFGKGTRVTVEP**

TS8.2 activated clone 4 V δ 1, D3, J1

AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQDAKS
 GRYSVNFKKAAKSVALTISALQLEDSAKYF**CALGTGVRGLQDADKLIFGKGTRVTVEP**

TS8.2 activated clone 5 V δ 1 D1+D3, J1

25 AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQNAKS
 GRYSVNFKKAAKSVALTISALQLEDSAKYF**CALLLGDTSFYTDKLIFGKGTRVTVEP**

TS8.2 activated clone 6 V δ 1 D1+D3, J1

30 AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQNAKS
 GRYSVNFKKAAKSVALTISALQLEDSAKYF**CAALLPFLPSDWGIPVTDKLIFGKGTRVTVEP**

TS8.2 activated clone 7 V δ 1, D2 and D3, J1

35 AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQNAKS
 GRYSVNFKKAAKSVALTISALQLEDSAKYF**CALGTGVRGLQDQTDKLIFGTGTRVTVEP**

TS8.2 activated clone 8 V δ 1, D2 and D3, J1

40 AQKVTQAQSSVSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQNAKS
 GRYSVNFKKAAKSVALTISALQLEDSAKYF**CALGGLSSLDLGDNDNHYTDKLIFGKGTRVTVEP**

TS8.2 activated clone 9 Vδ1, D2 and D3, J1

AQKVTQAQSSVSM PVRKA VTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQNAKS
GRYSVNFKKAAKSVALTISALQLEDSAKYFCALGHPRSLMGVYTDKLI FGKGTRVTVEP

5 **Examples of Gamma chains activated by TS8.2 Mab:**B8 TS8.82 activated Clone-1 γ4, J1/J2

SSNLEGR TKSVIRQTGSSAEITCDLAEGSTGYIHWY LHQEGKAPQRLLYYDSYTSSVVLES GI
SPGKYDTYGSTRKNLRILRNLIENDSGVYYCATWDDGKKLFGSGTTLVVT

10

B8 TS8.82 activated Clone-2 γ9, JP1

AGHLEQPQISSTKTLSKTARLECVVSGITISATSVYWYRERPGEVIQFLVSISYDGTVRKESGI
PSGKFEVD RIPETSTSTLTIHNVEKQDIATYYCALWDDTRLGKKIKVFAPGTKLIIT

15 TS8.2 activated Clone-3 γ4, J1

SSNLEGR TKSVIRQTGSSAEITCDLAEGSTGYIHWY LHQEGKAPQRLLYYDSYTSSVVLES GI
SPGKYDTYGSTRKNLRMILRNLIENDSGVYYCATWSDKKLFGSGTTLVVT

20 TS8.2 activated Clone-4 γ9, J1/J2

AGHLEQPQISSTKTLSKTARLECVVSGITISATSVYWYRERPGEVIQFLVSISYDGTVRKESGI
PSGKFEVD RIPETSTSTLTIHNVEKQDIATYYCALWIHKKLFGSGTTLVVT

25 TS8.2 activated Clone 5 γ2, J1

SSNLEGR TKSVIRQTGSSAEITCDLAEGSNGYIHWY LHQEGKTPQRLQYYDSYNSKVVLES
GVSPGKYTYASTRNRLRLILRNLIENDSGVYYCATWDCHYKKLFGSGTTLVVT

30 TS8.2 activated Clone 6 γ3, JP2

SSNLEGR TKSVTRQTGSSAEITCDLTVTNTFYIHWY LHQEGKAPQRLLYYDVSTARDVLES G
LSPGKYTYHTPRRWSWILRLQNLIENDSGVYYCATWDRRWIKTFAGTKLIVTSP

30

TS8.2 activated Clone 7 γ2, JP1

SSNLEGR TKSVIRQTGSSAEITCDLAEGSNGYIHWY LHQEGKAPQRLQYYDSYNSKVVLES
GVSPGKYTYASTRNRLRLILRNLIENDSGVYYCATWDGLDATCGVDTTGWFKIFAEGT
KLIVTSP

35

TS8.2 activated Clone 8 γ4, J1/J2

SSNLEGR TKSVTRPTGSSAVITCDLPVENAVYTHWY LHQEGKAPRRLLYYDSYTSSVVLES
GISPGKYDTYGSTRKNLRMILRNLIENDSGVYYCATKKLFGSGTTLVVT

40 TS8.2 activated Clone 9 γ9, JP1

AGHLEQPQISSTKTLSKTARLECVVSGITISATSVYWYRERPGEVIQFLVSISYDGTVRKESGI
PSGKFEVD RIPETSTSTLTIHNVEKQDIATYYCALWEIASQLGKKIKVFGPGTKLIIT

TS8.2 activated Clone 10 γ3, J1

SSNLGGRTKSVTRQTGSSAEITCDLTVTNTFYIHWYHQQEGKAPQRLLYYDVSTARDVLES
GLSPGKYYTHTPRRWSWILRLQNLIENDSGVYYCATWDRYYYKKLFGSGTTLVVT

TS8.2 activated Clone 11 γ 3, JP2

5 SSNLEGRTKSVTRQTGSSAEITCDLTVTNTFYIHWYHQQEGKAPQRLLYYDVSTARDVLES
LSPGKYYTHTPRRWSWILRLQNLIENDSGVYYPNSSDWIKTFAKGTCLIVTSP

TS8.2 activated Clone 12 γ 4, J2

10 SSNLEGGTKSVTRPTRSSAEITCDLAERNTFYIHWYHQQEGKAPQRLQYYDSYTSSVVLES
ISPGKYDTYGSTRKNLRMLRNLIENDSGVYYCATRDVPNYYKKLFGSGTTLVVT

TS8.2 activated Clone 13 γ 2, JP1

15 SSNLEGRTKSVIRQTGSSAEITCDLAEGSNGYIHWYHQQEGKAPQRLQYYDSYNSKVVLES
GVSPGKYYTYASTRNNLRILRNLIENDSGVYYCATWDGRVSYTTGWFKIFAEGTKLIVTS
P

TS8.2 activated Clone 14 γ 4, J1/J2

SSNLEGRTKSVIRQTGSSAEITCDLAEGSTGYIHWYHQQEGKAPQRLLYYDSYTSSVVLES
SPGKYDTYGSTRKNLRMLRNLIENDSGVYYCATWDKGRKLFGSGTTLVVT

TS8.2 activated Clone 15 γ 9, J1

20 AGHLEQPQISSTKTLSTARLECVVSGITISATSVYWYRERPGEVIQFLVSISYDGTVRKESGI
PSGKFEVDRIPESTSTLTIHNVEKQDIATYYCALWETHYKKLFGSGTTLVVT

TS8.2 activated Clone 16 γ 2, J1

25 SSNLEGRTKSVIRQTGSSAEITCDLAEGSNGYIHWYHQQEGKAPQRLQYYDSYNSKVVLES
GVSPGKYYTYASTRNNLRILRNLIENDSGVYYCATWDGTRYKKLFGSGTTLVVT

TS8.2 activated Clone 17 γ 4, JP1

30 SSNLEGRTKSVIRQTGSSAEITCDLAEGSTGYIHWYHQQEGKAPQRLLYYDSYTSSVVLES
SPGKYDTYGSTRKNLRMLRNLIENDSGVYYCATWGTTGWFKIFAEGTKLIVTSP

B8(Buffy coat 8)- B6 activated clones:

B6 activated Clone δ 2, D3, J1

35 AIELVPEHQTPVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGPGFKD
NFQGDIDIAKNLAVLKILAPSERDEGSYYCACDKVLGVPTASYTDNKLIFGKGTRVTVEP

B6 activated Clone δ 2, D3, J1

40 AIELVPEHQTPVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMSFIYREKDIYGPGFKD
NFQGDIDIAKNLAVLKILAPSERDEGSYYCACDTVGLPYDKLIFGKGTRVTVEP

B6 activated Clone δ 2, D2+D3, J1

45 AIELVPEHQTPVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGPGFKD
NFQGDIDIAKNLAVLKILAPSERDEGSYYCACDILTVLGDNRDKNLIFGKGTRVTVEP

B6 activated Clone $\delta 2$, D3,J1

AIELVPEHQTPVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGPFGKD
 NFQGDIDIAKNLAVLKILAPSERDEGSYYC**ACDVVGEGGADKLIFGKGTRVTVEP**

5 B6 activated Clone $\delta 2$, D3,J1

AIELVPEHQTPVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGPFGKD
 NFQGDIDIAKNLAVLKILAPSERDEGSYYC**ACDTVGGGEYTDKLIFGKGTRVTVEP**

10 B6 activated Clone $\delta 2$, D3,J1

AIELVPEHQTPVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGPFGKD
 NFQGDIDIAKNLAVLKILAPSERDEGSYYC**ACDTVGTGDIRTYTDKLIFGKGTRVTVEP**

15 B6 activated Clone $\delta 2$, D2+D3,J1

AIELVPEHQTPVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGPFGKD
 NFQGDIDIAKNLAVLKILAPSERDEGSYYC**ACDSLTTGGSLTDKLIFGKGTRVTVEP**

20 B6 activated Clone $\delta 2$, D3,J3

AIELVPEHQTPVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGPFGKD
 NFQGDIDIAKNLAVLKILPPSERDEGSYYC**ACDTGGYSSWDTRQMFFGTGIKLFVEP**

25 B6 activated Clone $\delta 2$, D2+D3, J1

AIELVPEHQTPVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGPFGKD
 NFQGDIDIAKNLAVLKILAPSERDEGSYYC**ACDPLKTLGTYTDKLIFGKGTRVTVEP**

30 B6 activated Clone $\delta 2$, D1+D3, J1

AIELVPEHQTPVPSIGVPATLRCSMKGEAIGNYYINWYRKTQGNTMTFIYREKDIYGPFGKD
 NFQGDIDIAKNLAVLKILAPSERDEGSYYC**ACDAVIAGGSFTDKLIFGKGTRVTVEP**

B6 activated Gamma chains:35 B6 activated clone $\gamma 9$, JP1

AGHLEQPQISSTKTLTKARLECVVSGITISATSVYWYRERPGEVIQFLVSISYDGTVRKESGI
 PSGKFEVDRIPESTSTLTIHNVEKQDIATYY**CALWEDQELGKKIKVFGPGTKLIIT**

40 B6 activated clone $\gamma 9$, JP1

AGHLEQPQISSTKTLTKARLECVVSGITISATSVYWYRERPGEVIQFLVSISYDGTVRKESGI
 PSGKFEVDRIPESTSTLTIHNVEKQDIATYY**CALWAYPELGKKIKVFGPGTKLIIT**

45 B6 activated clone $\gamma 9$, JP1

AGHLEQPQISSTKTLTKARLECAVSGITISATSVYWYRERPGEVIQFLVSISYDGTVRKESGI
 PSGKFEVDRIPESTSTLTIHNVEKQDIATYY**CALWEVQELGKKIKVFGPGTKLIIT**

45 B6 activated clone $\gamma 9$, JP1

AGHLEQPQISSTKTLTKARLECVVSGITISATSVCWYRERPGEVIQFLVSISYDGTVRKESGI
 PSGKFEVDRIPESTSTLTIHNVEKQDIATYY**CALWEVLQELGKKIKVFGPGTKLIIT**

B6 activated clone $\gamma 9$, JP1

AGHLEQ PQISSTKTL SKTARLECVVSGITISATSVYWYRERPGEVIQFLVSISYDGTVRKESGI
PSGKFEVD RIPETSTSTLTIHNVKKQDIATYYCALWEVRELGKKIKVFGPGTKLIIT

5

B6 activated clone $\gamma 9$,

JP1AGHLEQ PQISSTKTL SKTARLECVVSGITISATSVYWYRERPGEVIQFLVSISYDGTVRKE
SGIPSGKFEVD RIPETSTSTLTIHNVEKQDIATYYCALWRELGKKIKVFGPGTKLIIT

B6 activated clone $\gamma 9$,

10 JP1AGHLEQ PQISSTKTL SKTARLECVVSGITISATSVYWYRERPGEVIQFLVSISYDGTVRKE
SGIPSGKFEVD RIPETSTSTLTIHNVEKQDIATYYCALWEAQELGKKIKVFGPGTKLIIT

B6 activated clone $\gamma 9$,

15 JP1AGHLEQ PQISSTKTL SKTARLECVVSGITISATSVYWYRERPGEVIQFLVSISYDGTVRKE
SGIPSGKFEVD RIPETSTSTLTIHNVEKQDIATYYCALWETELGKKIKVFGPGTKLIIT

B6 activated clone $\gamma 9$,

20 JP1AGHLEQ PQISSTKTL SKTARLECVVSGITISATSVYWYRERPGEVIQFLVSISYDGTVRKE
SGIPSGKFEVD RIPETSTSTLTIHNVEKQDIATYYCALWEEVELGKKIKVFGPGTKLIIT

B6 activated clone $\gamma 9$, JP1

AGHLEQ PQISSTKTL SKTARLECVVSGITISATSVYWYRERPGEVIQFPVSISYDGTVRKESGI
PSGKFEVD RIPETSTSTLTIHNVEKQDIATYYCALWEPPQSLGKKIKVFGPGTKLIIT

25

**Example 34. Cryopreservation of $\gamma\delta$ T-Cells in Freeze Media to Generate Cell Banks
for Further Processing**

Non-modified $\gamma\delta$ T-Cells will be formulated in freezing media and placed in cryogenic storage units such as liquid nitrogen freezers (-195°C) or ultra-low temperature freezers (-65°C , -80°C or -120°C) for long-term storage. The freeze media will contain dimethyl sulfoxide (DMSO), sodium chloride (NaCl), dextrose, dextran sulfate or hydroxyethyl starch (HES) with physiological pH buffering agents ranging between 6.5 and 7.5.

The cryopreserved $\gamma\delta$ T-cells will be thawed and further processed by stimulation with antibodies, proteins, peptides, and cytokines. The cryopreserved $\gamma\delta$ T-Cells will be thawed and genetically modified as previously described in this application. The engineered $\gamma\delta$ T-cells will be further cryopreserved to generate cell banks in quantities of 10, 100, 200 vials at 10^6 to 10^8 cells per mL in freeze media. The non-engineered and/or engineered $\gamma\delta$ T-cells will be further cryopreserved to generate cell banks in quantities of 10, 100, 200, 500, and 1,000 vials or bags at 10^6 to 10^8 cells per mL in freeze media.

Example 35. Alternative Cryopreservation of $\gamma\delta$ T-Cells in Freeze Media to Generate Cell Banks for Further Processing

Other cryoprotectants are added to the cryopreservation support described in the previous example to provide nutritional support and biophysical cell protection against lysis during the freezing and thawing process. These include D-glucose, mannitol, sucrose, adenine, guanosine, recombinant human albumin, citrate, ant-coagulant, Benzonase, DNase, propylene glycol, ethylene glycol, 2-methyl-2,4-pentanediol. Additional additives designed to provide buffering capacity for high cell density frozen product include inorganic phosphate, sodium bicarbonate and HEPES.

Example 36. Cryopreservation of $\gamma\delta$ T-Cells in Freeze Media to Generate Cell Banks for Further Processing

Initial freezing of $\gamma\delta$ T-cells are performed in a temperature controlled ramp designed to achieve a freezing rate of between -0.1°C to -5°C per minute using a controlled rate freezer (e.g. CryoMed Controlled Rate Freezer) or a mechanical -70°C freezer with appropriately insulated racking system to deliver the desired freeze rate. The frozen cells are placed in -70°C freezers for short-term storage of up to 30 to 60 days. The frozen cells are placed in liquid N_2 storage tanks for longer-term storage of up to 12, 24, 36 and 48 months while maintaining $\gamma\delta$ T-cell number and cell functions without deterioration as measured by methods described in earlier sections.

The cryopreserved cells described in this example will be thawed and further stimulated and expanded in suitable closed vessels such as cell culture bags and/or bioreactors to generate suitable quantities of engineered $\gamma\delta$ T-cells for administration to a subject.

Example 37. Formulation of $\gamma\delta$ T-Cells for Direct Infusion into Patients

Engineered $\gamma\delta$ T-cells are concentrated by centrifugation and/or membrane dia-filtration to between 5×10^6 cells/mL and 10^8 cells/mL in physiological buffer containing cryo-protective excipients and placed in cryogenic storage units such as liquid nitrogen, or ultra-low temperature freezers for long-term storage.

Example 38. Treatment of a Human Subject Afflicted with Cancer

Fresh or frozen engineered and/or non-engineered $\gamma\delta$ T-cells are thawed at bedside and intravenously infused into human subjects. About 1 cell per kilogram to about 1×10^{10} cells per kilogram engineered and/or non-engineered $\gamma\delta$ T-cells are infused into the human subject over a 30-60 minute period of time. The engineered $\gamma\delta$ T-cells are administered with or without the aid of the co-stimulatory cytokine IL-2 or other cytokines. Optionally, the procedure is repeated. *In vivo* expansion of the $\gamma\delta$ T-cells in the subject is measured by flow cytometry.

Example 39. Generation of specific $\gamma\delta$ T cell activators

$\gamma\delta$ T cell activators in the form of murine antibodies were produced by immunization of recombinant soluble human $\gamma\delta$ TCR. $\delta 1$ -specific activators were generated by immunization using $\gamma 8\delta 1$ - TCR- FC, and $\delta 2$ -specific activators were generated by immunization using $\gamma 9\delta 2$ - TCR- FC, both constructs comprising the mature ECD of the $\gamma\delta$ TCR chains fused to human IgG1FC. Three strains of mice (Balb/c, CD-1 and FVB) were inoculated with human recombinant $\gamma\delta$ TCR to provide hybridomas that secrete high affinity, murine monoclonal antibody activators.

The $\gamma\delta$ TCR-Fc fusion constructs were generated by PCR amplification. $\gamma 8\delta 1$ TCR chains were amplified from RNA isolated from the BE13 (DSMZ #ACC 396), a T cell leukemia cell line expressing $\gamma 8\delta 1$ TCR. $\gamma 9\delta 2$ TCR chains were amplified from RNA isolated from Zoledronic acid activated human PBMCs. Soluble $\delta 1$ TCR-FC and $\delta 2$ TCR-FC were purified from the supernatant of transiently transfected HEK 293 cells. 10 μ g of soluble TCR was emulsified with an equal volume of TITERMAX® Gold (Sigma Aldrich) or Imject Alum Adjuvant (Thermo Fisher) and used for the immunization of each mouse. The resulting emulsions were then injected into six mice (2 each: Balb/c, CD-1 and FVB) via the footpad route.

Solid-phase ELISA assays were used to screen mouse sera for mouse IgG antibodies specific for human $\gamma\delta$ TCR. Briefly, 96 well plates (VWR International, Cat. #610744) were coated with recombinant $\gamma\delta$ TCR- at 1 μ g/ml in ELISA coating buffer overnight. After washing with PBS containing 0.02% (v/v) Tween 20, the wells were blocked with 3% (w/v) BSA in PBS, 200 μ L/well for 1 hour at room temperature (RT). Mouse serum was titrated (1:100, 1:200, 1:400, and 1:800) and added to the $\gamma\delta$ TCR coated plates at 50 μ L/well and incubated at RT for 1 hour. The plates are washed and then incubated with 50 μ L/well HRP-labeled goat anti-mouse IgG diluted 1:10,000 in 3% BSA-PBS or 2% FCS in PBS for 1 hour at RT. Again the plates were washed and 40 μ L/well of

a TMB substrate solution (Thermo Scientific 34028) was added for 15 minutes at RT. After developing, an equal volume of 2N H₂SO₄ was added to stop substrate development and the plates were analyzed by spectrophotometer at OD 450.

5 Sera-positive immunized mice were sacrificed and draining lymph nodes (popliteal and inguinal) were dissected out and used as a source for antibody producing cells. A single cell suspension of B cells (766×10⁶ cells) was fused with non-secreting P3x63Ag8.653 myeloma cells (ATCC #CRL-1580) at a ratio of 1:1 by electrofusion. Electrofusion was performed using the BTX Hybrimmune™ System, (BTX Harvard Apparatus) as per the manufacturer's directions. After the fusion, the cells were resuspended in hybridoma selection medium supplemented with Azaserine
10 (Sigma #A9666), high glucose DMEM medium with sodium pyruvate (Cellgro cat#15-017-CM) containing 15% Fetal Clone I serum (Hyclone), 10% BM Condimed (Roche Applied Sciences), 4 mM L-glutamine, 100 IU Penicillin-Streptomycin and 50 μM 2-mercaptoethanol and then plated in three T150 flasks in 50 mL selection medium per flask. The flasks were then placed in a humidified 37° C. incubator containing 5% CO₂ and 95% air for 6-7 days.

15 After six days of growth the library consisting of the cells grown in bulk in the T150 flask was plated at 1 cell per well in Falcon 384 well flat-bottom plates using the Aria II cell sorter. The selected hybridomas were then grown in 90 μL of culture medium containing 15% Fetal Clone I serum (Hyclone), 10% BM-Condimed (Roche Applied Sciences), 1 mM sodium pyruvate, 4 mM L-glutamine, 100 IU Penicillin-Streptomycin, 50 μM 2-mercaptoethanol, and 100 μM hypoxanthine.
20 Any remaining unused hybridoma library cells were frozen for future library testing. After ten days of growth supernatants from each well of the plated cells were assayed for antibodies reactive for δ1TCR by ELISA and FACS assays.

For screening by FACS- hybridoma wells secreting murine immunoglobulin were screened for human δ1/TCR specificity using a flow cytometry based assay. BE13 cells expressing δ1γ8TCR,
25 or Zoledronic acid treated human PBMCs expressing δ2γ9TCR were incubated for 60 minutes on ice with 25μL hybridoma supernatant. Cells were washed with PBS, 2% FCS, twice and then incubated with 50 μL of a goat-anti-mouse IgG Fc fragment specific secondary conjugated to PE diluted 1:300 in PBS/2% FCS. After 15 minutes of incubation, cells were washed twice with PBS/2% FCS and re-suspended in PBS/2% FCS with DAPI and analyzed by flow cytometry using a FACS Canto per the
30 manufacturer's instructions. Wells containing immunoglobulin that preferentially bound the BE13

cells were expanded and further tested for $\delta 1$ specificity by ELISA assay. Wells containing immunoglobulin that preferentially bound the Zoledronic acid activated PBMCs cells were expanded and further tested for $\delta 2$ specificity by ELISA assay.

High bond ELISA 96 well plates were coated with soluble $\gamma\delta$ TCR proteins generated by
 5 paring the following $\gamma\delta$ TCR chains: $\gamma 8\delta 1$, $\gamma 8\delta 2$, $\gamma 9\delta 1$ and $\gamma 9\delta 2$. Soluble TCRs were diluted and used at 1 μ g/ml in sodium carbonate buffer overnight at 4° C. The plates were washed and blocked with 3% BSA in PBS/Tween for one hour at 37° C. and used immediately or kept at 4° C. Undiluted hybridoma supernatants were incubated on the plates for one hour at RT. The plates were washed and probed with HRP labeled goat anti-mouse IgG diluted 1:10,000 in 3% BSA-PBS for one hour at
 10 RT. The plates were then incubated with substrate solution as described above and read at OD 450. Wells containing immunoglobulin that preferentially bound human $\delta 1$ TCR chain as determined by a signal above background, were transferred and expanded.

The resulting $\delta 1$ and $\delta 2$ TCR specific clonal hybridomas were cryopreserved in CS-10 freezing medium (Biolife Solutions) and stored in liquid nitrogen.

Example 40. Sequencing of $\delta 1$ - and $\delta 2$ -specific Activators

Exemplary distinct monoclonal antibodies that specifically bind, activate, and expand $\delta 1$ T cells were selected for sequencing and further analysis. As shown in the tables below, sequence analysis of the heavy chain variable regions (**FIG. 33**) and light chain variable regions (**FIG. 34**) of
 20 selected $\delta 1$ - $\gamma\delta$ TCR specific monoclonal antibodies generated in Example 39 confirmed novel complementarity determining regions and display of novel VDJ arrangements. The complementarity determining regions set forth in **FIGs. 33-34**, are as defined by Kabat. Exemplary distinct monoclonal antibodies that specifically bind, activate, and expand $\delta 2$ T cells were separately selected for sequencing and further analysis. As shown in the tables below, sequence analysis of the
 25 heavy chain variable regions (**FIG. 35**) and light chain variable regions (**FIG. 36**) of selected $\delta 2$ - $\gamma\delta$ TCR specific monoclonal antibodies generated in Example 39 confirmed novel complementarity determining regions and display of novel VDJ arrangements. The complementarity determining regions set forth in **FIGs. 35-36**, are as defined by Kabat.

Total RNA was extracted from selected hybridoma cells using the RNeasy isolation kit
 30 (RNeasy Mini Kit Qiagen # 74106). 10^4 hybridoma cells were lysed in 350 μ l RLT Buffer, an equal

volume of 70% ethanol was added, and the sample was loaded to RNeasy Mini spin column. Column was washed twice and RNA was eluted by 100 µl of RNase-free water loaded directly to the spin column membrane. The quality of the RNA preparations was determined by fractionating 3 µL in a 1% agarose gel before being stored at -80° C. until used.

5 The variable region of the Ig heavy chain of each hybridoma was amplified using a 5' primer mix comprising thirty-two mouse specific leader sequence primers, designed to target the complete mouse VH repertoire, in combination with a 3' mouse C γ primer specific for all mouse Ig isotypes. A 400 bp PCR fragment of the VH was sequenced from both ends using the same PCR primers. Similarly, a mix of thirty-two 5' V κ leader sequence primers designed to amplify each of the V κ
10 mouse families combined with a single reverse primer specific to the mouse kappa constant region were used to amplify and sequence the kappa light chain. The VH and VL transcripts were amplified from 100 ng total RNA using reverse transcriptase polymerase chain reaction (RT-PCR).

A total of eight RT-PCR reactions were run for each hybridoma: four for the V κ light chain and four for the V γ heavy chain. The One Step OneStep Ahead RT-PCR kit was used for
15 amplification (Qiagen # 220213). Reaction mixtures were prepared that included 5 µL of RNA, 0.5 of 100 µM of either heavy chain or κ light chain primers (custom synthesized by IDT), 12.5 µL of master mix with DNA polymerases, buffers, dNTPs, 1 µL of enzyme mix containing reverse transcriptase and DNA polymerase, and 0.4 µL of ribonuclease inhibitor RNasin (1 unit). The reaction mixture contains all of the reagents required for both reverse transcription and PCR. The
20 thermal cycler program was set for an RT step 50° C. for 10 minutes, 95° C. for 5 minutes, followed by 30 cycles of PCR (95° C. for 30 seconds, 58° C. for 30 seconds, 72° C. for one minute). There was then a final incubation at 72° C. for 10 minutes.

PCR products were prepared for direct DNA sequencing, with the QIAquick™ PCR Purification Kit (Qiagen) according to the manufacturer's protocol. The DNA was eluted from the
25 spin column using 50 µL of sterile water and then sequenced directly from both strands using specific V region primers. Nucleotide sequences were analyzed using VBase2 to identify germline V, D and J gene members with the highest sequence homology.

FIG. 33 depicts the contiguous amino acid sequences of heavy chain variable regions from δ 1-specific antibodies. **FIG. 34** depicts the corresponding contiguous amino acid sequences of the

light chain variable regions. Taken together **FIGs. 33-34** provide the annotated sequences of the identified operable anti $\delta 1$ TCR antibodies.

FIG. 35 depicts the contiguous amino acid sequences of heavy chain variable regions from $\delta 2$ -specific antibodies. **FIG. 36** depicts the corresponding contiguous amino acid sequences of the light chain variable regions. Taken together **FIGs. 35-36** provide the annotated sequences of the identified operable anti $\delta 2$ TCR antibodies.

Example 41. Cross-reactivity of $\delta 1$ -specific Activators

The third gamma delta population is V $\delta 3$ T cells, which take up about 0.2% of circulating T cells.

The V $\delta 3$ T cells are rare in blood but are rich in liver and in patients with leukemias and some chronic viral infections. Other minor subsets include V $\delta 4$, V $\delta 5$, V $\delta 6$, V $\delta 7$, and V $\delta 8$ $\gamma\delta$ T cells, are composed of VJ α genes spliced to the C δ respectively, encoding distinct TCR alpha and delta proteins.

Cross reactivity and the ability of $\delta 1$ -specific activators cross react with other $\gamma\delta$ T cell populations was tested by binding assay to detect binding of $\delta 1$ -specific specific activators to soluble TCRs composed of 8 different delta chains (V $\delta 1$, V $\delta 2$, V $\delta 3$, V $\delta 4$, V $\delta 5$, V $\delta 6$, V $\delta 7$, and V $\delta 8$). All eight delta chains were cloned as a C-terminal Fc-fusion and co-transfected with a γ -FC chain into 293 cells soluble TCRs-FC were secreted into the medium.

A day before the transfection 293 adherent cells 1×10^5 cells/ well were plated in 12 well plate.

On the day of transfection, 0.5 μ g of each plasmid (delta+ gamma chains) were co-transfected using 293fectinTM Transfection Reagent (Thermo Fisher #12347019) in serum free media.

Secreted TCR was collected 3 days post transfection for binding analysis in ELISA assay.

ELISA binding assay.

Soluble TCRs-FC proteins were captured to the surface of ELISA plate coated with 1 μ g/mL goat anti human FC (Goat Anti-Human IgG, Fc γ Fragment Specific- Jackson ImmunoResearch # 109-005-098). Binding was perform overnight at 4° C. Binding of $\delta 1$ -specific specific activators to soluble $\gamma\delta$ TCRs captured on plates was tested by detection with HRP conjugated secondary antibody: Goat anti mouse FC specific (Jackson ImmunoResearch Laboratories, Inc. Peroxidase-

AffiniPure Goat Anti-Mouse IgG, Fc γ Fragment Specific # 115-035-008) diluted to 1: 10,000 in blocking buffer, followed by adding TMB substrate per well. The results are shown in **Fig. 37**.

The ability to cross react and activate other subsets of $\gamma\delta$ T cells in addition to the $\delta 1$ T cells may be of importance as clonally expanded TRDV4 and TRDV8 T cells were shown to contribute to the immune response directed against AML, while oligoclonal TRDV5 and TRDV6 might occur in patients who undergo relapse. (Jin et al. Journal of Hematology & Oncology (2016) 9:126).

Example 42. Activation and Proliferation Assay

$\delta 1$ and $\delta 2$ TCR specific antibodies were tested for induction of activation and proliferation of $\gamma\delta$ T cells. Briefly, 48-well plates (Corning) were coated with goat-anti-mouse Fc- γ polyclonal antibodies at 5 $\mu\text{g/mL}$ in 50 mM NaHCO_3 or PBS overnight. Wells were washed with PBS and further blocked with 1% BSA in PBS for 1 hr at 37C and $\gamma\delta$ -TCR specific antibodies were added at 1 $\mu\text{g/mL}$ in 0.1% BSA for 2 hrs. For cell labeling, after overnight rest in complete medium fresh or frozen human PBMCs were washed and reconstituted in 5 μM CFSE (PBS) and incubated for 5 min in the dark at room temperature. Cells were washed with FBS containing media and reconstituted in RPMI-1640 culture media containing 10% FBS, 2mM Glutamine, 25mM HEPES and 200 IU/mL of rhIL-2 at $10^6/\text{mL}$. CFSE-labeled PBMCs were plated at $0.5 \times 10^6/\text{well}$ and cultured for 5 days.

After 5 days, cells were harvested and stained with specific surface markers to pan- $\gamma\delta$, V $\delta 1$, V $\delta 2$ and $\alpha\beta$ TCR and cell proliferation was assessed by progressive changes in CFSE fluorescence. Percentage of cells that divided during the 5 days as a result of activation is shown in **Fig. 39** and served to rank antibodies for potency of activation.

Example 43. One-step expansion of V $\delta 1$ and V $\delta 2$ T-cells from human PBMCs using specific MAbs

Selected activating $\delta 1$ -TCR and $\delta 2$ -TCR antibodies (D1-08, D1-35, D1-39, D2-14, D2-17, D2-22, D2-30, D2-32, D2-33, D2-33, D2-35, D2-36, D2-37) and OKT3 were captured at 1 $\mu\text{g/mL}$ in 2% FBS (PBS) by pre-coated goat-anti-mouse Fc γ antibodies in 24-well plates as in example 2 above. Human PBMCs from several donors having initial $\delta 1$ populations between 0.2-0.28% of total cells and V $\delta 2$ population of ~ 0.2 -4% of total cells were plated at $10^6/\text{mL}/\text{well}$ in RPMI-1640 media containing 10% FBS, 2mM Glutamine, 100 IU/mL of rhIL-2. Cells in culture were fed every 2-3

days by media replenishment. On day 7 cells were harvested and re-plated at 10^6 /mL into new plates without activating antibodies and further expanded by continued feeding and re-plating on Day 14 to 10^6 /mL cell density. Cell analysis (counts and flow cytometry) was performed on Day 21 to determine purity and fold expansion of $\delta 1$ and $\delta 2$ cells. **Fig. 40** shows fold expansion of $\delta 1$ (A) and $\delta 2$ (B) cells in PBMC cultures. V $\delta 1$ and V $\delta 2$ MAbs induced expansion of $\delta 1$ and $\delta 2$ T cells in 21 days up to ~12000 fold for V $\delta 1$ and up to 6700 fold for V $\delta 2$ T cells.

Phenotype of expanded $\delta 1$ cells on Day 21 was evaluated by CD27 surface marker expression. **Fig. 41** shows that phenotype of V $\delta 1$ T-cells activated with of MAbs on Day 21 is predominantly (>50%) CD27+CD45RA+ (naïve) and CD27+CD45RA- (central memory).

Example 44. One-step expansion of $\delta 1$ -T cells from umbilical cord blood

Mononuclear cells from human umbilical cord blood containing 0.4% of V $\delta 1$ cells were isolated by Ficoll density gradient and activated with captured $\delta 1$ TCR specific antibodies as described in example 43. As illustrated in the table below, V $\delta 1$ T-cells expanded by up to 1503 fold in 21 days after activation with $\delta 1$ specific MAbs.

D1 Antibody	Fold expansion, Day 21
TS8.2	1062
$\delta 1$ -08	1104
$\delta 1$ -22	1503
$\delta 1$ -26	557
$\delta 1$ -35	918
$\delta 1$ -37	1017
D1-39	1039
Immuno510	460

Example 45. Two-step expansion of V $\delta 1$ T cells using feeder cells

In this example, human PBMCs were activated and $\delta 1$ cells were expanded for 14 days following protocol described in Example 4. Briefly, V $\delta 1$ antibodies TS-1, D1-08, D1-37, D1-39 were captured in 24-well plates via coated goat-anti-mouse Fc γ polyclonal antibodies and cells were activated and expanded as described. On Day 14 cell culture were harvested and $\delta 1$ T cells were

enriched by depletion of $\alpha\beta$ T cells using biotinylated anti-TCR $\alpha\beta$ antibody and magnetic microbeads (Miltenyi Biotec). Enriched V δ 1 cells were subjected to a second step activation/expansion using irradiated PBMC feeder cells. Briefly, 130,000 total enriched cells were cultured in 20 mL of RPMI-1640 medium containing 10% FBS, 2 mM Glutamine, 100 IU/mL IL-2 and 30 ng/mL D1-35 MAb in the presence of irradiated human allogeneic PBMCs at a ratio of 200:1. The cultures were placed vertically in T25 flask. On Day 18 cells were fed by replacing 70% of the media. From then on, cells were fed every 2-3 days by exchanging 50% of the media and adjusting viable cell density to 1×10^6 /mL and harvested on Day 28. As illustrated in the table below, δ 1 cells were expanded up to about 28,000-fold in a 2-step expansion procedure.

1st Step activation, MAb	2nd Step Activation after $\alpha\beta$ T cell depletion/MAb	V δ 1 fold expansion
δ 1-37	iPBMCs/D1-35	11573
δ 1-08	iPBMCs/D1-35	27965
δ 1-39	iPBMCs/D1-35	15769

Example 46. Two-step expansion of V δ 1 T cells using feeder cells, without $\alpha\beta$ T cell depletion

In this example, human PBMCs with starting V δ 1 population of 0.2% were activated and δ 1 cells were expanded for 7 days following a protocol described in Example 6. Briefly, V δ 1 antibodies TS-1, D1-08, D1-37, D1-39 were captured in 24-well plates via coated goat-anti-mouse Fc γ polyclonal antibodies and cells were activated and expanded as described. On Day 7 cell cultures were harvested and expanded cells were subjected to a second step activation/expansion using irradiated feeder cells. Briefly, 130,000 total cells were cultured in 20 mL of RPMI1640 medium containing 10% FBS, 2 mM Glutamine, 100 IU IL-2 and 30 ng/mL D1-35 MAb in the presence of irradiated human allogeneic PBMCs at a ratio of 200:1. The cultures were placed vertically in T25 flasks. On Day 12 cells were fed by replacing 70% of the media. From then on, cells were fed every 2-3 days by exchanging 50% of the media and adjusting viable cell density to 10^6 /mL until harvest on Day 21. As illustrated in the table below, δ 1 cells were expanded up to about 114,000-fold in a 2-step expansion procedure.

1st Step	2nd Step	V δ 1 fold expansion
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activation, MAb	Activation/MAb	
TS-1	iPBMCs/D1-35	82298
D1-37	iPBMCs/D1-35	113764
D1-08	iPBMCs/D1-35	55348
D1-39	iPBMCs/D1-35	81391

Example 47. Two-step expansion of V δ 1 T cells using feeder cells with $\alpha\beta$ T cell depletion

In this example, human PBMCs with starting V δ 1 population of 0.36% were activated and V δ 1 cells were expanded for 14 days following protocol described in Example 4. On Day 14 cell culture was harvested and δ 1 T cells were enriched by depletion of $\alpha\beta$ T cells using biotinylated anti-TCR $\alpha\beta$ antibody and magnetic microbeads (Miltenyi Biotec). Enriched V δ 1 cells were subjected to a second step activation/expansion using irradiated feeder cells. Briefly, enriched cells were cultured in 20 mL of RPMI-1640 medium containing 10% FBS, 2 mM Glutamine, 100 IU/mL IL-2 and 30 ng/mL D1-35 MAb in the presence of irradiated K562 cells at ratios of 49:1, 9:1 and 2:1. The cultures at the specified ratios (25×10^6 each total live cells) were placed in 6-well G-Rex flask (Wilson Wolf). On Day 18 cells were fed by replacing 75% of the media and harvested on Day 21. As illustrated in the table below, δ 1 cells were expanded up to about 31,000-fold in a 2-step expansion procedure.

1st Step activation, MAb	2nd Step Activation/MAb	V δ 1 fold expansion,
D1-35	iK562/D1-35 at 49:1	31121
D1-35	iK562/D1-35 at 9:1	18142
D1-35	iK562/D1-35 at 2:1	6346

Example 48. δ 1 T cells activated and expanded with V δ 1 MAbs show robust cytotoxicity against tumor cell lines

Human PBMCs were activated with immobilized D1-08 MAb for 7 days and cultures were expanded by feeding every 2 days in static conditions. At the end of expansion, δ 1 T cells were enriched by depletion of $\alpha\beta$ T cells using anti-TCR $\alpha\beta$ antibody and magnetic microbeads (Miltenyi).

Purified $\delta 1$ cells were used in cytotoxicity assay against Jurkat (leukemia) and Colo205 (colon cancer) cell lines. Briefly, expanded $\delta 1$ T cells were incubated with CFSE labeled target cells at 10:1, 5:1, 2:1, 1:1 and 1:5 effector:target cell ratio for 4 hours in 96-well round bottom plates (suspension) in complete media containing 100 IU/mL IL-2. At the end of incubation, cell suspension was stained with Annexin-V-APC conjugate and Zombie Violet viability dye and analyzed by flow cytometry to detect dead/dying cells. As shown in **Fig. 42**, V $\delta 1$ cells demonstrated cytotoxicity against both tumor cell lines at various ratios in 4hr assay up to 45%.

Example 49. $\delta 1$ T cells activated and expanded with a 2-step procedure including a second step with shaking

Isolation of Peripheral Blood Mononuclear Cells

Fresh blood product collected from an apheresis machine is diluted 1:3 with PBS without Ca^{2+} and Mg^{2+} . Diluted blood product is then purified using the Ficoll-Paque™ PLUS (GE Healthcare) system using the Sepmate (STEMCELL). The enriched lymphocyte layer is then enriched washed twice, first with PBS without Ca^{2+} and Mg^{2+} , then with 0.25% human serum albumin in PBS without Ca^{2+} and Mg^{2+} . Finally, purified blood product is rested overnight in fetal bovine serum containing and serum free medium on ultra-low attachment 24-well plate, 6-well plate, and T-75 flasks at 1×10^7 cells/mL.

T-cell Activation

The day before T-cell activation, the appropriate cell culture vessels, including tissue culture treated 24-well plate, 6-well plate, T75 flask, or gas permeable bags are coated with anti-Fc antibody. After initial coating, the vessels are stored at 2-8 °C in the dark overnight. After initial anti-Fc coating, the vessel is blocked with 5% fetal bovine serum in PBS without Ca^{2+} and Mg^{2+} , and incubated for 1-hour at 37 °C CO_2 incubator. After the 1-hour incubation, the vessel is washed once with PBS without Ca^{2+} and Mg^{2+} , then the vessel is captured with V $\delta 1$ specific monoclonal antibodies. Then the vessel is incubated at room temperature for up to 4 hours with and without shaking. At the end of the capture, the vessel is washed twice with PBS without Ca^{2+} and Mg^{2+} , and is ready for T-cell activation.

Overnight rested PBMC is diluted to 0.5×10^6 to 2×10^6 cells/cm² using either fetal bovine serum containing and serum free medium. Then the diluted PBMC is added to the V $\delta 1$ specific

monoclonal antibodies immobilized vessel. PBMC is maintained statically between 3 to 7 days. Fresh cell culture medium is added daily starting on day 3 to maintain sufficient nutrient level such as glucose and glutamine, and diluting the accumulation of toxic byproducts such as lactate and ammonium. At the end of the initial activation, cells are harvested by mechanical or chemical methods. Cells are then transferred to new flasks for subsequent expansion.

T-cell Expansion

After initial T-cell activation, cell culture is maintained in 37 °C CO₂ incubator for up to 21 days. Initially, activated cell culture is diluted to 1e6 cells/mL in either fetal bovine serum containing and serum free medium. The T-cells can be cultivated in static T-flasks with ultra-low attachment or tissue culture treated surfaces, including T-25, T-75, T-150 and T-225 flasks, or in standard non-baffled or baffled shake flasks, including 125 mL, 250 mL, 500 mL, 1000 mL, 2000 mL, and 3000 mL total vessel volumes. In static T-flask, the working volume to surface area ratio is maintained at 0.5 mL/cm² or below to ensure sufficient culture oxygenation. In shake flask, the working volume to total vessel volume ratio is maintain at 1 to 2 or below on orbital shaking platforms to ensure sufficient mixing and aeration.

Viable cell culture density (VCD), metabolite, pH and gas measurements are taken daily. Metabolite measurements are taken of glucose, lactate, glutamine, glutamate, ammonium, sodium ion, potassium ion, and calcium ion. pH and gas measurements including partial pressure of oxygen, CO₂, and pH are also taken. Oxygen, CO₂, and pH measurement are taken with temperature compensation. Feeding based on VCD or metabolite can be both used to maintain cell cultures. For VCD-based feeding strategy, if VCD increased to 1.5 x 10⁶ cells/mL or above, culture is diluted to 1 x 10⁶ cells/mL using fresh medium. For metabolite-based feeding strategy, fresh medium is added when glucose level is below 1g/L to increase to 1.5g/L.

Cell Harvest and Banking

Cell viability of above 90% is always observed at the end of the 19 day or 21-day expansion. Post cell expansion, cell culture is washed and resuspended in cell banking medium. Cell is then aliquoted at more than 20 x 10⁶ cells/mL cell density. Cryopreservation in the vapor phase of the liquid nitrogen is proceeded by control freezing using control rate freezer or freezing container.

Example 49a

In this example, Donor A fresh blood product, which had a starting V δ 1 composition of 0.4%, was purified by Ficoll and rested overnight in ultra-low attachment T75 flask. After activation with immobilized δ 1 specific monoclonal antibodies δ 1-35 and δ 1-08 from day 0 to day 5, the cultures were transferred to shake flasks. From day 5 to day 19, the cultures were sampled daily, and nutrient and waste concentrations were maintained by VCD-based feeding strategies using fresh serum medium. The V δ 1 fold expansion and purity for δ 1-35 and δ 1-08 activated cultures are illustrated in **Fig. 43**.

Example 49b

In this example, Donor B fresh blood product, which had a starting V δ 1 composition of 0.1%, was purified by Ficoll and rested overnight in ultra-low attachment T75 flask. After activation with immobilized δ 1 specific monoclonal antibody δ 1-08 from day 0 to day 5, the cultures were transferred to shake flasks. From day 5 to day 19, the cultures are sampled daily, and nutrient and waste concentrations were maintained by VCD-based feeding strategies using either fresh serum-containing medium or fresh serum free medium. The V δ 1 fold expansion and doubling time (DT) (accumulative from day 0 to day 19) in serum containing and serum free medium expanded cultures are illustrated in **Fig. 44**.

Example 49c

In this example, Donor C fresh blood product, which had a starting V δ 1 composition of 0.5%, was purified by Ficoll and rested overnight in ultra-low attachment T75 flask. After activation with immobilized δ 1 specific monoclonal antibody δ 1-08 from day 0 to day 7 (49c.1) and from day 0 to day 5, the cultures were transferred to shake flasks. From day 7 to day 19, the cultures were sampled daily, and nutrient and waste concentrations were maintained by glucose-based feeding strategies using fresh serum medium. And for the other two cultures, from day 7 to day 19, the cultures were sampled daily, and nutrient and waste concentrations were maintained by either VCD based feeding or glucose-based feeding strategies using fresh serum medium. The V δ 1 fold expansion and doubling time (DT) (accumulative from day 14 to day 19) are illustrated in **Fig. 45**.

Fig. 45 section 49c.1 illustrates V δ 1 fold expansion and DT (accumulative from day 14 to day 19) from culture that was 7 day activated and then maintained using glucose-based feeding strategy from day 7 to day 19. Fig. 45 section 49c.2 illustrates V δ 1 fold expansion and DT

(accumulative from day 14 to day 19) 5 day activated culture that was then maintained using VCD-based feeding strategy from day 7 to day 19. Fig. 45 section 49c.3 illustrates V δ 1 fold expansion and DT (accumulative from day 14 to day 19) 5 day activated culture that was then maintained using VCD-based feeding strategy from day 7 to day 19.

5

Example 50. Transduction of V δ 1 T cells with retroviral chimeric antigen receptor construct

Peripheral blood mononuclear cells from a healthy donor were stimulated with an immobilized anti-V δ 1 specific monoclonal antibody (δ 1-08) and cultured for 19 days in media supplemented with rhIL-2 as described in example 49a. Briefly, isolated PBMCs were activated in 24-well plates with D1-08 V δ 1 MAb captured via pre-coated goat-anti-mouse Fc γ antibodies. Cells were plated at 1×10^6 /mL/well in RPMI1640 media containing 10% FBS, 25 mM HEPES, 2 mM Glutamine and 100 IU/mL IL-2. After five days cells were transferred to shake flasks and expanded as described in example 49a. On day 19 cells were harvested, washed and re-stimulated with PHA (2 μ g/mL) for 2 days followed by transduction with a retroviral vector encoding an anti-CD20 (Rituximab) derived chimeric antigen receptor (scFv linked to CD28-CD3 ζ). Briefly, wells of 24-well plate were pre-coated with retronectin (TaKaRa Bio) and supernatant containing retrovirus (1 mL) was incubated for 4 hours at 32C to allow adhesion of viral particles, after which the supernatant was removed (retronectin plate). Expanded δ 1 T cells were washed, resuspended in 1 mL of viral supernatant supplemented with 100 IU/mL of IL-2 at 0.5×10^6 cells/mL and plated into retronectin plate. Plate was centrifuged at 1800 rpm for 10 min and placed into 37C incubator. Cells were cultured for an additional 7 days with feeding every 2 days and then analyzed by flow cytometry for percent transduction using fluorescent rat anti-Rituximab idiotypic antibody (FITC conjugate) that recognizes the CD20-specific CAR expressed on the T-cell surface. More than 34% of the viable V δ 1⁺ cells expressed the CAR construct as shown in Fig. 46.

Example 51. Epitope Mapping of antibodies to $\gamma\delta$ T cell TCRs

The aim was to map the epitope specificities of δ 1-specific activators. All δ 1-specific activators are directed against the δ 1 chain of the $\gamma\delta$ TCR. Mapping was done by using recombinant chimeric molecules of human and dolphin δ 1 TCR chain. Despite of a high amino acid identity

30

between the human and dolphin $\delta 1$ chain, $\delta 1$ specific activators did not bind dolphin $\delta 1$ chain. Therefore, we have utilized the similarities in sequence and conformation between human V $\delta 1$ and dolphin V $\delta 1$ (65% sequence similarity) to construct recombinant chimeric molecules in which we replaced different segments of the human residues with their homologous counterpart from the dolphin $\delta 1$ TCR chain.

All mAbs were confirmed to bind specifically to the variable region of the $\delta 1$ chain of $\gamma\delta$ TCR by Flow cytometry analysis. The $\delta 1\gamma 8$ TCR was expressed as soluble FC fusion protein in a correctly folded conformation, determined by binding of all panel of anti $\delta 1$ mouse monoclonal antibodies (mAbs). These mAbs were shown to target conformationally-dependent epitopes of the $\gamma\delta$ TCR.

In-order to express discrete epitopes of human $\delta 1$ TCR, a set of 7 recombinant chimeric $\delta 1$ chains were cloned to produce human/dolphin $\delta 1$ -FC TCR chains. Dolphin residues comprised amino acids 1-11, 1-21, 1-28, 1-47, 1-70, 1-80 and 1-95 respectively, according to IMGT unique numbering for T cell receptor variable domains (Lefranc et al., Developmental and Comparative Immunology 27 (2003) 55–77), as illustrated in **Fig. 38**.

These $\delta 1$ specific binding domains were identified by ELISA binding assay and detection of loss of binding without prior knowledge of potential contact residues. By overlaying key dolphin residues that impacted the binding of selected $\delta 1$ activators we have identified 8 possible binding bins within the V and J region of the $\delta 1$ TCR chain.

The expression vector used in this study is pCI-Neo Mammalian Expression Vector (Promega # E1841) Expression of soluble TCRs was derived from the CMV promoter. The mouse IgH signal peptide, was used for exporting the soluble TCR proteins to the medium, followed by human FC domain for capture of soluble TCR to ELISA plate.

The ECD of human $\delta 1$ chain was amplified from $\gamma\delta 1$ BE-13 T cell line. The dolphin V $\delta 1$ sequence was ordered as a synthetic gene base on the published sequence (*Tursiops truncatus* mRNA for T cell receptor delta chain (TRD gene), isolate 5R1D80) (GenBank: LN610748.1). Using these cDNAs as templates new chimeric molecules were cloned from synthetic fragments (IDT DNA) to be cloned into the 5' AgeI site at the signal peptide and 3' E.coRI at the delta constant region. Six constructs consisting of dolphin/human $\delta 1$ chains were cloned and co transfected with a $\gamma 8$ -FC chain to express soluble $\delta 1\gamma 8$ TCR proteins in 293 cells.

HEK-293 cells were cultured in 90-mm cell culture plates in a DMEM medium (Thermo Fisher Scientific) with 10% fetal calf serum (HyClone™ FetalClone™ I Serum (U.S.) - GE Healthcare Life Sciences). For every construct 1×10^6 HEK-293 cells were transfected with 10 µg of plasmid using 293 Fectin as transfection reagent. Supernatants were harvested 72 hours post
 5 transfection

δ1 specific activators- monoclonal antibodies differed in their binding pattern to the chimeras and a distinct binding pattern could be identified for each monoclonal antibody, as tested in ELISA assay.

Polystyrene microtiter plates (Greiner bio-one # 655061) were coated overnight at room
 10 temperature with 1 µg/well of Goat anti human FC specific antibody (Jackson ImmunoResearch # 109-005-098) in ELISA coating buffer (50 mM Na₂CO₃, 0.05% NaH₃, pH 9.6). The plates were washed three times with washing buffer (PBS 0.05% (v/v) Tween 20) and then blocked for 1 h at room temperature (RT) with 100 µl/well of ELISA blocking buffer (3%BSA in PBST). Binding of soluble TCRs to blocked plates was done by adding 100µl of each supernatant and incubation for 1
 15 hour at Room temperature. Each antibody was diluted to 1 µg/ml in ELISA blocking buffer, and tested for binding to each γδ TCR chimeric molecule.

Detection of binding was done by adding 100 µl/well of HRP conjugated secondary antibody: Goat anti mouse FC specific (Jackson ImmunoResearch Laboratories, Inc. Peroxidase-AffiniPure Goat Anti-Mouse IgG, Fcγ Fragment Specific # 115-035-008) diluted to 1:10,000 in blocking buffer.
 20 The amount of bound antibodies was detected by the use of TMB substrate (Thermo Fisher #34029)Color development was measured spectrophotometrically at 450nm after 10 minutes. All assays were done in duplicate.

Sequence of all constructs

δ1-D3-J1 FC- Fully human TCR

25 gcccagaaggttactcaagcccagtcacgatccatgccagtgaggaaagcagtcaccctgaactgcctgtatgaaacaagttggt
 ggtcatattatatttttggtacaagcaactcccagcaaagagatgatttccttattgccagggttctgatgaacagaatgaaaaagtggcgcta
 ttctgtcaactcaagaaagcagcgaaatccgtcgccttaaccatttcagccttacagctagaagattcagcaaagtactttgtgctcttgggacgg
 ggggtgaggggactccaggacaccgataaactcatctttggaaaaggaaccgtgtgactgtggaaccaa

AQKVTQAQSSVSM PVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
 NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDTDKLIFGKGTRVTVE
 P

5 Dolphin TRD isolate 5R1D8

gcccagaaagtactcaagtcagcgagccatgtccagtcagctaggggagcggtcaccttgagctgtcagtatgaaacaagcttg
 agctgggtacgatatttttgggtataagcagcttcccagtgagagatgactttccttattcatcagatttcttgacaaaaatgcaaagaatggccgct
 attctgtaaaactttcaggaaagacataaattcatcagcctcaccatttcagccttactgggtgaagattctgcaaactacttctgtgctctccgggagc
 gcgtgtccacgctggtttattcaatacgcgcaataagccactgctattcggcaaaggaacctatctgaacgttgaaacaa

10

AQKVTQVQGRAMSSQLGEAVTLSCQYETSLSWYDIFWYKQLPSGEMTFLIHQISSDQ
 NAKN.GRYSVNFQERHKFISLTISALLVEDSANYFCALRERVVHVLFNTRNKPLLFGKGT
 Y LNVEP

15 Set of dolphin/human chimera -soluble TCRs were cloned -Human sequence in blue

Dolphin/Human 1-11

gcccagaaagtactcaagtcagcgagccatgtccatgccagtgaggaaagcagtcaccctgaactgcctgtatgaaacaagttggt
 ggtcatattatatttttgggtacaagcaactcccagcaaagagatgattttccttattcgcagggttctgatgaacagaatgaaaaagtggctgcta
 20 ttctgtcaacttaagaaagcagcgaaatccgtcgccttaaccatttcagccttacagctagaagattcagcaaagtactttgtgctcttgggacgg
 ggggtgaggggactccaggacaccgataaactcatctttggaaaaggaacccgtgtgactgtggaacaa

AQKVTQVQGRAMSMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDE
 QNAKS.GRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDTDKLIFGKGTRVT
 25 VEP

Dolphin/Human 1-21

gcccagaaagtactcaagtcagcgagccatgtccagtcagctaggggagcggtcaccttgaaactgcctgtatgaaacaagttggt
 ggtcatattatatttttgggtacaagcaactcccagcaaagagatgattttccttattcgcagggttctgatgaacagaatgaaaaagtggctgcta

ttctgtcaacttcaagaaagcagcgaaatccgtcgccttaaccatttcagccttacagctagaagattcagcaaagtacttttgctcttgggacgg
gggtgaggggactccaggacaccgataaactcatctttggaaaaggaacccgtgtgactgtggaaccaa

AQKVTQVQRAMSSQLGEAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
5 NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDTDKLIFGKGTRVTVE
P

Dolphin/Human 1-28

gcccagaaagtactcaagtccagcgagccatgtccagtcagctaggggaggcgggtcaccttgagctgtcagtatgaaacaagctgg
10 tggatcatattatatttttggtacaagcaacttcccagcaaagagatgatttcttattcgccagggttctgatgaacagaatgcaaaaagtggctgct
attctgtcaacttcaagaaagcagcgaaatccgtcgccttaaccatttcagccttacagctagaagattcagcaaagtacttttgctcttgggacg
ggggtgaggggactccaggacaccgataaactcatctttggaaaaggaacccgtgtgactgtggaaccaa

AQKVTQVQRAMSSQLGEAVTLSCQYETSWWSYYIFWYKQLPSKEMIFLIRQGSDEQ
15 NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDTDKLIFGKGTRVTVE
P

Dolphin/Human 1-47

gcccagaaagtactcaagtccagcgagccatgtccagtcagctaggggaggcgggtcaccttgagctgtcagtatgaaacaagcttg
20 agctggtacgatatttttggtataagcagcttcccagtaaagagatgatttcttattcgccagggttctgatgaacagaatgcaaaaagtggctgc
tattctgtcaacttcaagaaagcagcgaaatccgtcgccttaaccatttcagccttacagctagaagattcagcaaagtacttttgctcttgggacg
ggggtgaggggactccaggacaccgataaactcatctttggaaaaggaacccgtgtgactgtggaaccaa

AQKVTQVQRAMSSQLGEAVTLSCQYETSLSWYDIFWYKQLPSKEMIFLIRQGSDEQ
NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDTDKLIFGKGTRVTVE
25 P

Dolphin/Human 1-70

gcccagaaagtactcaagtccagcgagccatgtccagtcagctaggggaggcgggtcaccttgagctgtcagtatgaaacaagcttg
agctggtacgatatttttggtataagcagcttcccagtgagagatgacttcttattcatcagatttctgatgaacagaatgcaaaaagtggctgct

attctgtcaacttcaagaaagcagcgaaatccgtcgccttaaccatttcagccttacagctagaagattcagcaaagtactttgtgctcttgggacg
ggggtgaggggactccaggacaccgataaactcatctttggaaaaggaacccgtgtgactgtggaaccaa

AQKVTQVQRAMSSQLGEAVTLSCQYETSLSWYDIFWYKQLPSGEMTFLIHQISDEQ
5 NAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDTDKLIFGKGTRVTVE
P

Dolphin/human 1-80

gcccagaaagtactcaagtccagcgagccatgtccagtcagctaggggagggcggtcaccttgagctgtcagtatgaaacaagcttg
10 agctggtacgatatttttggataagcagcttcccagtgagagatgactttccttattcatcagatttcttctgacaaaaatgcaaagaatggccgct
attctgtaaactttaagaaagcagcgaaatccgtcgccttaaccatttcagccttacagctagaagattcagcaaagtactttgtgctcttgggacgg
gggtgaggggactccaggacaccgataaactcatctttggaaaaggaacccgtgtgactgtggaaccaa

AQKVTQVQRAMSSQLGEAVTLSCQYETSLSWYDIFWYKQLPSGEMTFLIHQISSDQ
15 NAKNGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGLQDTDKLIFGKGTRVTVE
P

Dolphin/human 1-95

gcccagaaagtactcaagtccagcgagccatgtccagtcagctaggggagggcggtcaccttgagctgtcagtatgaaacaagcttg
20 agctggtacgatatttttggataagcagcttcccagtgagagatgactttccttattcatcagatttcttctgacaaaaatgcaaagaatggccgct
attctgtaaactttcaggaaagacataaattcatcagcctcaccatttcagccttacagctagaagattcagcaaagtactttgtgctcttgggacgg
gggtgaggggactccaggacaccgataaactcatctttggaaaaggaacccgtgtgactgtggaaccaa

AQKVTQVQRAMSSQLGEAVTLSCQYETSLSWYDIFWYKQLPSGEMTFLIHQISSDQ
NAKNGRYSVNFQERHKFISLTISALQLEDSAKYFCALGTGVRGLQDTDKLIFGKGTRVTVEP
25

RESULTS

Functional TCR genes are assembled from separate V, D and J region segments by genetic recombination. When the TCR delta chain gene is assembled a D gene segment is juxtaposed to a J segment and this is followed by rearrangement of a V segment to the assembled downstream DJ
30 segment. The immunoglobulin-like fold of the TCR chains positions three loops or complementarity

determining regions (CDRs) from each chain in close proximity to each other, creating a binding face that will contact antigen. Two of these CDR loops, namely 1 and 2, are encoded by the V gene segment and have only the diversity provided by the V δ 1 region gene segments. The third CDR loop is created by the juxtaposition of V(D)J segments and provides much more diversity, a result of the ability of each V segment to rearrange to any (D)J segment compounded by the fact that the joining of the coding sequences is imprecise. Nucleotides may be added or deleted at each of these junctions. Epitope binding specificity is shown by ELISA (FIGs. 47A-B).

We have identified activators that bind different epitopes on the framework region of the δ 1 chain of the $\gamma\delta$ TCR molecule. One group of monoclonal antibodies is reactive with δ 1 variable region of the delta chain of the TCR. In particular, the invention provides monoclonal antibodies, such as d1-08, d1-39, d1-192, d1-201, d1-285 which bind the δ 1 chain variable region, regardless of the junctional diversity (juxtaposition of all $\gamma\delta$ TCR compositions (D)J1, V(D)J2 and V(D)J3). These monoclonal antibodies are reactive with the “variable region” of the delta TCR and therefore are monoclonal antibodies reactive with an epitope of the V region.

Other δ 1 activators specifically recognize the δ 1 chain variable region, and the CDR3 loop created by the restricted juxtaposition of V(D)J1 segments. These monoclonal antibodies are reactive with the “variable region” of the TCR in combination epitope of the V-D or V-D-J regions: d1-37, d1-113, d1-155, d1-182, d1-183, d1-191, d1-278 and d1-282.

Other δ 1 activators specifically recognize the δ 1 chain variable region, and the CDR3 loop created by the juxtaposition of V(D)J1 and V(D)J1but not V(D)J3 segments: d1-35, d1-143, d1-149, d1-203.

Antibodies specific for δ 1 were grouped into the following epitope Bins:

Bin1: VDJ Junction- JH1/JH2 : TS-1; and δ 1-18

Bin 1b: VDJ Junction- J1, not J3, loss of binding for K120T/A mutant: δ 1-37

Bin2: (aa 11-21): δ 1-285

Bin2b: (aa 11-21), loss of binding for R16N mutant: R9.12

Bin2c: (aa 11-21), cross reacts with δ 3, δ 4, and δ 5 $\gamma\delta$ TCRs: δ 1-39

Bin3: (aa 80-95 or 70-95): δ 1-08; δ 1-23

Bin4: (aa 1-11), loss of binding for K120T/A mutant: δ 1-35; δ 1-203

Bin5: (aa 28-47+ J1 region): δ 1-113; δ 1-155; δ 1-183; δ 1-191; δ 1-278; δ 1-282

Bin6: (aa 21-28+ J1 region): TS8.2; δ 1-143

Bin7: (aa 47-70+ J1/J2 region): δ 1-149; δ 1-253, and δ 1-257

Bin8: (aa 70-80) + J1/J2: δ 1-192

Bin9: (aa 80-95): δ 1-201.

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Similar to the d1 epitope mapping: based on Linguiti G. et al., 2016. Genomic and expression analyses of *Tursiops truncatus* T cell receptor gamma (TRG) and alpha/delta (TRA/TRD) loci reveal a similar basic public repertoire in dolphin and human. BMC Genomics. A set of 5 recombinant chimeric δ 2 chains were cloned to produce human/dolphin δ 2-FC TCR chains. Dolphin residues comprised amino acids 28-94, 44-94, 72-94 and, 82-94 respectively, according to IMGT unique numbering for T cell receptor variable domains (Lefranc et al., Developmental and Comparative Immunology 27 (2003) 55–77), as illustrated in **Fig.15**.

Transfection of δ 2/ γ 9-FC constructs

15 Human TRGV9-JP-FC

GCAGGTCACCTAGAGCAACCTCAAATTTCCAGTACTAAAACGCTGTCAAAAACAGCCCGCCTGGAATGTGTGGTGTCTGGAATAACAAT
TTCTGCAACATCTGTATATTGGTATCGAGAGAGACCTGGTGAAGTCATACAGTTCCTGGTGTCCATTTCATATGACGGCACTGTCAGAA
AGGAATCCGGCATTCCGTCAGGCAAAATTTGAGGTGGATAGGATACCTGAAACGCTCTACATCCACTCTCACCATTCCAAATGTAGAGAAA
CAGGACATAGCTACCTACTACTGTGCCTTGTGGGAGGTAAAGCAAGAGTTGGGCCAAAAAATCAAGGTATTTGGTCCCGGAACAAAGCT
TATCATTACAG

20

AGHLEQPQISSTKTLTKTARLECVVSGITISATSVYWRERPEVIGFLVVISYDGTVRKESGIPSGKFEVDRIPESTSTLTITHNVEK
QDIATYYCALWEVKQELGKRKIKVFGPGTKLIIT

25 Human V δ 21J1-FC

GCCATTGAGTTGGTGCCTGAACACCAACAGTGCCTGTGTCAATAGGGGTCCCTGCCACCCCTCAGGTGCTCCATGAAAGG
AGAAGCGATCGGTAACCTACTATATCAACTGGTACAGGAAGACCCAAAGGTAACACAATGACTTTTCATATACCGAGAAAAGG
ACATCTATGGCCCTGGTTTCAAGACAATTTCCAAGGTGACATTGATATTGCAAAGAACCTGGCTGTACTTAAGATACTT
GCACCATCAGAGAGAGATGAAGGGTCTTACTACTGTGCCTGTGACCCTCTTGGCGGACCCCCGATAAACTCATCTTTGG
AAAAGGAACCCGTGTGACTGTGGAACCAA

30

AIELVPEHQTVFVSIQVPATLRCSMKGEAIGNYYINWYRKTQGNMTTFIYREKDIYGPGEKDNFQGDIDIAKNLAVLKIL
APSERDEGSYYCACDPLGGPPDKLIFGKGTTRVTVEP

35 Dolphin V δ 2J1-FC

GCTGTACGTTGGTGCCTCAAACCAAGCAAGGAGTGTGTCTGTGGGGGAATCTGTCAACCCTCAGGTGCTCCATGAAAGG
AGACTCCATCAGTAACCTATTATACCTTCTGGTACAGGAGAACACCGGGTAACACAATGACTCTCATATACCGAGAAAGGGG
GCACATATGGCCCTGGTTTCAAGACAACCTCCAAGGTGAAATTGATTTTTTAAACAACCAGGCTGTGCTGAATATCCTG
GAGGCATCAGAGAGAGATGAAGGATCTTACTACTGTGCCTGTGACCCTCTTGGCGGACCCCCCGATAAACTCATCTTTGG
AAAA
GGAACCCGTGTGACTGTGGAACCAA

40

AVTLVPQNQARSVSVGESVTLRCSMKGDSISNYTTFWYRRTPGNTMTLIYREGGTYGPGFEDNLQGEIDFLNNQAVLNIL
EASERDEGSYYCACDPLGGPPDKLIFGKGTTRVTVEP

Human/Dolphin Vδ2J1-FC (28-94)

GCCATTGAGTTGGTGCCTGAACACCAAACAGTGCCTGTGTCAATAGGGGTCCCTGCCACCCTCAGGTGCTCCATGAAAGG
AGACTCCATCAGTAACTATTATACCTTCTGGTACAGGAGAACACCGGGTAACACAATGACTCTCATATACCGAGAAGGGG
5 GCACATATGGCCCTGGTTTTCGAAGACAACCTCCAAGGTGAAATTGATTTTTTAAACAACCAGGCTGTGCTGAATATCCTG
GAGGCATCAGAGAGAGATGAAGGATCTTACTACTGTGCCTGTGACCCTCTTGGCGGACCCCCCGATAAACTCATCTTTGG
AAAA
GGAACCCGTGTGACTGTGGAACCAA

10 AIELVPEHQTVFVSI GVPATLRCSMKGDSISNYTTFWYRRTPGNTMTLIYREGGTYGPGFEDNLQGEIDFLNNQAVLNIL
EASERDEGSYYCADCPLGGPPDKLI FGKGTRVTVEP

Human/Dolphin Vδ2J1-FC (44-94)

GCCATTGAGTTGGTGCCTGAACACCAAACAGTGCCTGTGTCAATAGGGGTCCCTGCCACCCTCAGGTGCTCCATGAAAGG
15 AGAAGCGATCGGTAACACTACTATATCAACTGGTACAGGAGAACACCGGGTAACACAATGACTCTCATATACCGAGAAGGGG
GCACATATGGCCCTGGTTTTCGAAGACAACCTCCAAGGTGAAATTGATTTTTTAAACAACCAGGCTGTGCTGAATATCCTG
GAGGCATCAGAGAGAGATGAAGGATCTTACTACTGTGCCTGTGACCCTCTTGGCGGACCCCCCGATAAACTCATCTTTGG
AAAAGGAACCCGTGTGACTGTGGAACCAA

20 AIELVPEHQTVFVSI GVPATLRCSMKGEAIGNYYINWYRRTPGNTMTLIYREGGTYGPGFEDNLQGEIDFLNNQAVLNIL
EASERDEGSYYCADCPLGGPPDKLI FGKGTRVTVEP

Human/ Dolphin Vδ2J1-FC (72-94)

GCCATTGAGTTGGTGCCTGAACACCAAACAGTGCCTGTGTCAATAGGGGTCCCTGCCACCCTCAGGTGCTCCATGAAAGG
25 AGAAGCGATCGGTAACACTACTATATCAACTGGTACAGGAAGACCCAAGGTAACACAATGACTTTTCATATACCGAGAAAAGG
ACATCTATGGCCCTGGTTTTCGAAGACAACCTCCAAGGTGAAATTGATTTTTTAAACAACCAGGCTGTGCTGAATATCCTG
GAGGCATCAGAGAGAGATGAAGGATCTTACTACTGTGCCTGTGACCCTCTTGGCGGACCCCCCGATAAACTCATCTTTGG
AAAAGGAACCCGTGTGACTGTGGAACCAA

30 AIELVPEHQTVFVSI GVPATLRCSMKGEAIGNYYINWYRKTQGNMTMTFIYREKDIYGPFGFEDNLQGEIDFLNNQAVLNIL
EASERDEGSYYCADCPLGGPPDKLI FGKGTRVTVEP

Human Dolphin Vδ2J1-FC (82-94)

GCCATTGAGTTGGTGCCTGAACACCAAACAGTGCCTGTGTCAATAGGGGTCCCTGCCACCCTCAGGTGCTCCATGAAAGG
35 AGAAGCGATCGGTAACACTACTATATCAACTGGTACAGGAAGACCCAAGGTAACACAATGACTTTTCATATACCGAGAAAAGG
ACATCTATGGCCCTGGTTTCAAAGACAATTTCCAAGGTGACATTGATTTTTTAAACAACCAGGCTGTGCTGAATATCCTG
GAGGCATCAGAGAGAGATGAAGGATCTTACTACTGTGCCTGTGACCCTCTTGGCGGACCCCCCGATAAACTCATCTTTGG
AAAAGGAACCCGTGTGACTGTGGAACCAA

40 AIELVPEHQTVFVSI GVPATLRCSMKGEAIGNYYINWYRKTQGNMTMTFIYREKDIYGPFGKDNFQGDIDFLNNQAVLNIL
EASERDEGSYYCADCPLGGPPDKLI FGKGTRVTVEP

Human Vδ21J1-FC (CDR1-S)

GCCATTGAGTTGGTGCCTGAACACCAAACAGTGCCTGTGTCAATAGGGGTCCCTGCCACCCTCAGGTGCTCCATGAAAGG
45 AGAAGCGATCAGTAACTACTATATCAACTGGTACAGGAAGACCCAAGGTAACACAATGACTTTTCATATACCGAGAAAAGG
ACATCTATGGCCCTGGTTTCAAAGACAATTTCCAAGGTGACATTGATATTGCAAAGAACCTGGCTGTACTTAAGATACTT
GCACCATCAGAGAGAGATGAAGGGTCTTACTACTGTGCCTGTGACCCTCTTGGCGGACCCCCCGATAAACTCATCTTTGG
AAAAGGAACCCGTGTGACTGTGGAACCAA

50 AIELVPEHQTVFVSI GVPATLRCSMKGEAISNYINWYRKTQGNMTMTFIYREKDIYGPFGKDNFQGDIDIAKNLAVLKIL
APSERDEGSYYCADCPLGGPPDKLI FGKGTRVTVEP

Human V δ 21J1-FC (CDR2-G)

GCCATTGAGTTGGTGCCTGAACACCAAACAGTGCCTGTGTCAATAGGGGTCCCTGCCACCCCTCAGGTGCTCCATGAAAGG
 AGAAGCGATCGGTAACACTACTATATCAACTGGTACAGGAAGACCCAAGGTAACACAATGACTTTCATATACCGAGAAAAGG
 GCATCTATGGCCCTGGTTTCAAAGACAATTTCCAAGGTGACATTGATATTGCAAAGAACCTGGCTGTACTTAAGATACTT
 5 GCACCATCAGAGAGAGATGAAGGGTCTTACTACTGTGCCTGTGACCCTCTTGGCGGACCCCCCGATAAACTCATCTTTGG
 AAAAGGAACCCGTGTGACTGTGGAACCAA

AIELVPEHQTVPVSIGVPATLRCSMKGEAIGNYYINWYRKQTQNTMTFIYREKDIYGPFGKDNFQGDIDIAKNLAVLKIL
 APSEDEGSYYCADDPLGGPPDKLIFGKGRVTVEP

Human V δ 21J1-FC (CDR3-S)

GCCATTGAGTTGGTGCCTGAACACCAAACAGTGCCTGTGTCAATAGGGGTCCCTGCCACCCCTCAGGTGCTCCATGAAAGG
 AGAAGCGATCGGTAACACTACTATATCAACTGGTACAGGAAGACCCAAGGTAACACAATGACTTTCATATACCGAGAAAAGG
 ACATCTATGGCCCTGGTTTCAAAGACAATTTCCAAGGTGACATTGATATTGCAAAGAACCTGGCTGTACTTAAGATACTT
 15 GCACCATCAGAGAGAGATGAAGGGTCTTACTACTGTGCCTCTGACCCTCTTGGCGGACCCCCCGATAAACTCATCTTTGG
 AAAAGGAACCCGTGTGACTGTGGAACCAA

AIELVPEHQTVPVSIGVPATLRCSMKGEAIGNYYINWYRKQTQNTMTFIYREKDIYGPFGKDNFQGDIDIAKNLAVLKIL
 APSEDEGSYYCADDPLGGPPDKLIFGKGRVTVEP

ELISA results are shown in **Fig. 48**, identifying the following epitope bins:

Bin	1	2	3	4
Residues	83-94	28-38	72-83	1-27
mAbs	δ 2-17	15D	δ 2-32	δ 2-14
	B6			δ 2-22
				δ 2-30
				δ 2-31
				δ 2-36
				δ 2-37

* * *

While preferred embodiments of the present invention have been shown and described herein,
 it will be obvious to those skilled in the art that such embodiments are provided by way of example
 only. Numerous variations, changes, and substitutions will now occur to those skilled in the art
 without departing from the invention. It should be understood that various alternatives to the
 embodiments of the invention described herein may be employed in practicing the invention. It is

intended that the following claims define the scope of the invention and that methods and structures within the scope of these claims and their equivalents be covered thereby.

CLAIMS**WHAT IS CLAIMED IS:**

1. An *ex vivo* method for producing an enriched $\gamma\delta$ T-cell population from an isolated mixed cell population, comprising directly contacting the mixed cell population with one or more agents
5 which selectively expand $\delta 1$ T-cells or $\delta 2$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR or a $\delta 2$ TCR to provide clinically-relevant levels of an enriched $\gamma\delta$ T cell population.
2. The method of claim 1, wherein an agent which selectively expands $\delta 1$ T-cells is selected from an agent which binds to the same epitope as an antibody selected from TS-1 and TS8.2.
10
3. The method of claim 1, wherein an agent which selectively expands $\delta 1$ T-cells is selected from an agent which specifically binds to an epitope comprising a $\delta 1$ variable region.
4. The method of claim 1, wherein the agent binds to a $\delta 1$ TCR variable region comprising an
15 amino acid sequence of the consensus sequence in FIG. 24.
5. The method of claim 1, wherein the agent binds to an epitope comprising residues Arg71, Asp72 and Lys120 of the $\delta 1$ variable region.
- 20 6. The method of claim 4, wherein the agent has reduced binding to a mutant $\delta 1$ TCR polypeptide comprising a mutation at K120 at the delta J1 and delta J2.
7. The method of claim 1, wherein the percentage of $\delta 1$ T cells in the enriched $\gamma\delta$ T cell population is greater than 60% of $\delta 1$ T-cells.
25
8. The method of claim 1, wherein an agent which selectively expands $\delta 2$ T-cells is selected from an agent which binds to the same epitope as an antibody selected from B6 and 15D.
9. The method of Claim 1, wherein an agent which selectively expands $\delta 2$ T-cells is selected
30 from an agent which specifically binds to an epitope comprising a $\delta 2$ variable region.

10. The method of claim 9, wherein the agent has reduced binding to a mutant $\delta 2$ TCR polypeptide comprising a mutation at G35 of the $\delta 2$ variable region.

5 11. The method of claim 7, wherein the percentage of $\delta 2$ T cells in the enriched $\gamma\delta$ T cell population is greater than 80% of $\delta 2$ T-cells.

12. The method of any one of the preceding claims, wherein the enriched $\gamma\delta$ T-cell population is not expanded by an antigen presenting cell or an aminophosphate.

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13. The method of any one of the preceding claims, wherein said one or more agents which selectively expand $\delta 1$ T-cells or $\delta 2$ T-cells are antibodies.

14. The method of any one of the preceding claims, wherein said one or more agents which
15 selectively expand $\delta 1$ T-cells or $\delta 2$ T-cells are immobilized on a surface.

15. The method of any one of the preceding claims, wherein said isolated mixed cell population is selected from a peripheral blood sample, a cord blood sample or a tumor.

20 16. The method of any one of the preceding claims, wherein said enriched $\gamma\delta$ T-cell population comprise polyclonal TCR diversity.

17. The method of any one of the preceding claims, wherein the enriched $\gamma\delta$ T-cell population is further formulated for administration to a subject.

25

18. The method of any one of the preceding claims, wherein the enriched $\gamma\delta$ T-cell population includes a therapeutically effective amount of $\gamma\delta$ T-cells.

19. The method of any one of the preceding claims, wherein said $\gamma\delta$ T-cell population is further
30 engineered to stably express one or more tumor recognition moieties.

20. An expanded $\gamma\delta$ T-cell population, wherein greater than 60%, 70%, 80%, or 90% of the expanded $\gamma\delta$ T-cells are $\delta 1$ T-cells.

21. The expanded $\gamma\delta$ T-cell population of claim 20 wherein greater than 60% or 70% the $\delta 1$ T-cells express the phenotype CD45RA+/CD27+ and/or CD45RA-/CD27+.

22. The expanded $\gamma\delta$ T-cell population of any one of the preceding claims, wherein said expanded $\gamma\delta$ T-cell population comprise polyclonal TCR diversity.

23. An expanded $\gamma\delta$ T cell population, wherein greater than 80% or 90% of the expanded $\gamma\delta$ T-cells are $\delta 2$ T-cells.

24. An expanded $\gamma\delta$ T-cell population, wherein 10-90% of the expanded $\gamma\delta$ T-cells are $\delta 1$ T-cells and 90-10% of the expanded $\gamma\delta$ T-cells are $\delta 2$ T-cells.

25. The expanded $\gamma\delta$ T-cell population according to any one of claims 20-24, which is derived from tumor infiltrating lymphocytes.

26. The expanded $\gamma\delta$ T-cell population according to claim 25, which is derived from tumor infiltrating lymphocytes isolated from colon adenocarcinoma metastasis to liver, ovarian tumors, head and neck tumor, or renal tumors.

27. The expanded $\gamma\delta$ T-cell population according to any one of claims 20-26, which is further formulated for administration to a subject.

28. The expanded $\gamma\delta$ T-cell population according to any one of claims 20-26, which includes a therapeutically effective amount of $\gamma\delta$ T-cells.

29. The expanded $\gamma\delta$ T-cell population according to any one of claims 20-26, wherein said $\gamma\delta$ T-cell population is engineered to stably express one or more tumor recognition moieties encoded by expression cassettes.

30. The expanded $\gamma\delta$ T-cell population according to Claim 29, wherein said $\gamma\delta$ T-cell is engineered to stably express two or more tumor recognition moieties encoded by expression cassettes.

31. The expanded $\gamma\delta$ T-cell population according to Claim 30, wherein said two or more tumor recognition moieties are different, wherein each different recognition moiety recognizes different epitopes of the same antigen or different epitopes of different antigens.

32. The expanded $\gamma\delta$ T-cell population according to any one of claims 29-31, wherein said tumor recognition moiety is selected from the group consisting of $\alpha\beta$ TCR, $\gamma\delta$ TCR, a chimeric antigen receptor (CAR), whole antibody or their antigen-binding fragment, single-chain variable fragment (scFv), a heavy chain or a light chain single domain antibody (sdAb), a Fab, a F(ab)₂, or any combination thereof that binds to: (i) a cell surface tumor antigen or (ii) a peptide derived from a tumor antigen expressed on the cell surface as a complex with MHC (peptide-MHC complex).

33. The expanded $\gamma\delta$ T-cell population according to claim 32, wherein said tumor recognition moiety is a $\gamma\delta$ TCR of a tumor infiltrating lymphocyte and which recognizes a tumor-specific antigen in a non-MHC restricted manner.

34. An expanded $\gamma\delta$ T-cell population obtained according to the method of any one of claims 1-19.

35. The expanded $\gamma\delta$ T-cell population according to any one of claims 20-26, wherein the $\gamma\delta$ T-cell population is expanded *ex vivo* without antigen stimulation by an antigen presenting cell or an aminophosphate.

36. The expanded $\gamma\delta$ T-cell population according to any one of claims 20-26, wherein the $\gamma\delta$ T-cell population is activated *ex vivo* with a monoclonal antibody or a fusion protein.

37. The expanded $\gamma\delta$ T-cell population according to any one of claims 20-26, wherein the $\gamma\delta$ T-cell population is formulated for administration to a subject without co-administration with IL-2.

38. A method for treating a cancer in a subject in need thereof, comprising administering a therapeutically effective amount of an expanded $\gamma\delta$ T-cell population according to any one of the preceding claims.

39. The method of Claim 38, wherein said expanded $\gamma\delta$ T-cell population is allogeneic with respect to the MHC loci of the subject.

40. The method of Claim 38, wherein said expanded $\gamma\delta$ T-cell population is engineered to stably express one or more tumor recognition moieties encoded by expression cassettes.

41. The method of Claim 38, wherein said expanded $\gamma\delta$ T-cell population stably express at least two different recognition moieties and each different recognition moiety recognizes (i) different epitopes of the same antigen, or (ii) different epitopes of different antigens.

42. The method of Claim 38, wherein said tumor recognition moiety is selected from the group consisting of $\alpha\beta$ TCR, $\gamma\delta$ TCR, a chimeric antigen receptor (CAR), whole antibody or their antigen-binding fragment, single-chain variable fragment (scFv), a heavy chain or a light chain single domain antibody (sdAb), a Fab, a F(ab)₂, or any combination thereof that binds to : (i) a cell surface tumor antigen or (ii) a peptide derived from a tumor antigen expressed on the cell surface as a complex with MHC (peptide-MHC complex).

43. An engineered $\gamma\delta$ T-cell, wherein the engineered $\gamma\delta$ T-cell is engineered to express a tumor recognition moiety, wherein the tumor recognition moiety recognizes a tumor antigen, wherein the

engineered $\gamma\delta$ T-cell expands *in vitro* in the presence of one or more agents which bind the same epitope as TS.1, TS8.2, B6, or 15D antibodies.

44. The engineered $\gamma\delta$ T-cell of claim 43, wherein the engineered $\gamma\delta$ T-cell is a $\delta 1$, $\delta 2$, or $\delta 3$ engineered T-cell.

45. The engineered $\gamma\delta$ T-cell of claim 43, wherein said engineered $\gamma\delta$ T-cell is further engineered to lack gene expression from at least one HLA loci.

46. The engineered $\gamma\delta$ T-cell of claim 43, wherein said one or more agents stimulates an expansion of the engineered $\gamma\delta$ T-cell at a mean rate of 1 cell division in less than 18 hours.

47. The engineered $\gamma\delta$ T-cell of claim 43, wherein said tumor recognition moiety is derived from a tumor infiltrating lymphocyte.

48. The method of claim 1, wherein the method comprises directly contacting the mixed cell population with one or more agents which selectively expand $\delta 1$ T-cells, wherein an agent which selectively expands $\delta 1$ T-cells is selected from an agent which binds to the same, or essentially the same, epitope as, or competes with, an antibody selected from the group consisting of $\delta 1$ -05, $\delta 1$ -08, $\delta 1$ -18, $\delta 1$ -22, $\delta 1$ -23, $\delta 1$ -26, $\delta 1$ -35, $\delta 1$ -37, $\delta 1$ -39, $\delta 1$ -113, $\delta 1$ -143, $\delta 1$ -149, $\delta 1$ -155, $\delta 1$ -182, $\delta 1$ -183, $\delta 1$ -191, $\delta 1$ -192, $\delta 1$ -195, $\delta 1$ -197, $\delta 1$ -199, $\delta 1$ -201, $\delta 1$ -203, $\delta 1$ -239, $\delta 1$ -253, $\delta 1$ -257, $\delta 1$ -278, $\delta 1$ -282, $\delta 1$ -285, and optionally also selected from the group consisting of TS8.2, TS-1, and R9.12.

49. The method of claim 1, wherein the method comprises directly contacting the mixed cell population with one or more agents which selectively expand $\delta 2$ T-cells, wherein an agent which selectively expands $\delta 2$ T-cells is selected from an agent which binds to the same, or essentially the same, epitope as, or competes with, an antibody selected from the group consisting of, $\delta 2$ -14, $\delta 2$ -17, $\delta 2$ -22, $\delta 2$ -30, $\delta 2$ -31, $\delta 2$ -32, $\delta 2$ -33, $\delta 2$ -35, $\delta 2$ -36, and $\delta 2$ -37, and optionally further selected from 15D and B6.

50. The method of claim 1, wherein the method comprises directly contacting the mixed cell population with one or more agents which selectively expand $\delta 1$ T-cells, $\delta 3$ T-cells, $\delta 4$ T cells, and $\delta 5$ T cells.

5 51. The method of claim 50, wherein an agent which selectively expands $\delta 1$ T-cells, $\delta 3$ T-cells, $\delta 4$ T cells, and $\delta 5$ T cells is selected from an agent which binds to the same epitope as an antibody selected from the group consisting of $\delta 1$ -39.

10 52. The method of claim 1, wherein the method comprises directly contacting the mixed cell population with one or more agents which selectively expand $\delta 1$ T-cells and $\delta 4$ T-cells.

53. The method of claim 52, wherein an agent which selectively expands $\delta 1$ T-cells and $\delta 4$ T-cells is selected from an agent which binds to the same epitope as an antibody selected from the group consisting of $\delta 1$ -18, and $\delta 1$ -35.

15

54. An *ex vivo* method for producing an enriched $\gamma\delta$ T-cell population from an isolated mixed cell population, comprising:

i) directly contacting the mixed cell population with one or more activating agents which:

20 - selectively expand $\delta 1$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR;

- selectively expand $\delta 1$ T-cells, $\delta 3$ T-cells, $\delta 4$ T-cells, and $\delta 5$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR, a $\delta 3$ TCR, a $\delta 4$ TCR, and a $\delta 5$ TCR;

- selectively expand $\delta 1$ T-cells and $\delta 4$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR and a $\delta 4$ TCR; or

25 - selectively expand $\delta 2$ T-cells by binding to an activating epitope specific of a $\delta 2$ TCR in a first $\gamma\delta$ T-cell expansion, thereby producing a first enriched $\gamma\delta$ T-cell population; and then

ii) directly contacting at least a portion of the first enriched $\gamma\delta$ T-cell population with antigen presenting cells (APCs) in a second $\gamma\delta$ T-cell expansion, thereby producing a second enriched $\gamma\delta$ T-cell population,
wherein the second enriched $\gamma\delta$ T-cell population comprises a clinically relevant number of $\gamma\delta$ T-cells.

55. The method of claim 54, wherein the second $\gamma\delta$ T-cell expansion further comprises directly contacting at least a portion of the first enriched $\gamma\delta$ T-cell population with one or more second activating agents which:

- selectively expand $\delta 1$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR;
- selectively expand $\delta 1$ T-cells, $\delta 3$ T-cells, $\delta 4$ T-cells, and $\delta 5$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR, a $\delta 3$ TCR, a $\delta 4$ TCR and a $\delta 5$ TCR;
- selectively expand $\delta 1$ T-cells and $\delta 4$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR and a $\delta 4$ TCR; or
- selectively expand $\delta 2$ T-cells by binding to an activating epitope specific of a $\delta 2$ TCR.

56. The method of claim 55, wherein one or more of the second activating agents are structurally different from one or more of the activating agents of the first $\gamma\delta$ T-cell expansion.

57. The method of claim 56, wherein the second activation agents are structurally different from the first activating agents of the first $\gamma\delta$ T-cell expansion.

58. The method of any one of claims 55-57, wherein the one or more activating agents of i) are agents that selectively expand $\delta 1$ T-cells, $\delta 1$ T-cells, $\delta 3$ T-cells, $\delta 4$ T-cells, and $\delta 5$ T-cells; or $\delta 1$ T-cells and $\delta 4$ T-cells, and are selected from agents that bind to the same epitope as an antibody selected from the group consisting of $\delta 1$ -05, $\delta 1$ -08, $\delta 1$ -18, $\delta 1$ -22, $\delta 1$ -23, $\delta 1$ -26, $\delta 1$ -35, $\delta 1$ -37, $\delta 1$ -39, $\delta 1$ -113, $\delta 1$ -143, $\delta 1$ -149, $\delta 1$ -155, $\delta 1$ -182, $\delta 1$ -183, $\delta 1$ -191, $\delta 1$ -192, $\delta 1$ -195, $\delta 1$ -197, $\delta 1$ -199, $\delta 1$ -201, $\delta 1$ -203, $\delta 1$ -239, $\delta 1$ -253, $\delta 1$ -257, $\delta 1$ -278, $\delta 1$ -282, $\delta 1$ -285, and optionally further selected from TS-1, R9.12, and TS8.2.

59. The method of claim 58, wherein the second $\gamma\delta$ T-cell expansion further comprises directly contacting at least a portion of the first enriched $\gamma\delta$ T-cell population with one or more second activating agents which selectively expand:

- $\delta 1$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR;

5 - $\delta 1$ T-cells, $\delta 3$ T-cells, $\delta 4$ T-cells, and $\delta 5$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR, a $\delta 3$ TCR, a $\delta 4$ TCR, and a $\delta 5$ TCR; or

- $\delta 1$ T-cells and $\delta 4$ T-cells, by binding to an activating epitope specific of a $\delta 1$ TCR and a $\delta 4$ TCR, wherein the second activating agents bind to the same epitope as an antibody selected from the group consisting of $\delta 1$ -05, $\delta 1$ -08, $\delta 1$ -18, $\delta 1$ -22, $\delta 1$ -23, $\delta 1$ -26, $\delta 1$ -35, $\delta 1$ -37, $\delta 1$ -39, $\delta 1$ -113, $\delta 1$ -143,
10 $\delta 1$ -149, $\delta 1$ -155, $\delta 1$ -182, $\delta 1$ -183, $\delta 1$ -191, $\delta 1$ -192, $\delta 1$ -195, $\delta 1$ -197, $\delta 1$ -199, $\delta 1$ -201, $\delta 1$ -203, $\delta 1$ -239, $\delta 1$ -253, $\delta 1$ -257, $\delta 1$ -278, $\delta 1$ -282, $\delta 1$ -285, and optionally further selected from TS-1, R9.12, and TS8.2.

60. The method of any one of claims 55-57, wherein the one or more activating agents of i) are
15 agents that selectively expand $\delta 2$ T-cells and selected from agents that bind to the same epitope as an antibody selected from the group consisting of $\delta 2$ -14, $\delta 2$ -17, $\delta 2$ -22, $\delta 2$ -30, $\delta 2$ -31, $\delta 2$ -32, $\delta 2$ -33, $\delta 2$ -35, $\delta 2$ -36, and $\delta 2$ -37, and optionally further selected from B6, and 15D.

61. The method of claim 60, wherein the second $\gamma\delta$ T-cell expansion further comprises directly
20 contacting at least a portion of the first enriched $\gamma\delta$ T-cell population with one or more second activating agents which selectively expand $\delta 2$ T-cells by binding to an activating epitope specific of a $\delta 2$ TCR, wherein the second activating agents bind to the same epitope as an antibody selected from the group consisting of $\delta 2$ -14, $\delta 2$ -17, $\delta 2$ -22, $\delta 2$ -30, $\delta 2$ -31, $\delta 2$ -32, $\delta 2$ -33, $\delta 2$ -35, $\delta 2$ -36, and $\delta 2$ -37, and optionally further selected from B6, and 15D.

25 62. An *ex vivo* method for producing an enriched $\gamma\delta$ T-cell population from an isolated mixed cell population, comprising:

i) directly contacting the mixed cell population with one or more first agents which selectively expand $\delta 1$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR or $\delta 2$ T-cells

by binding to an activating epitope specific of a $\delta 2$ TCR in a first $\gamma\delta$ T-cell expansion, thereby producing a first enriched $\gamma\delta$ T-cell population; and then

ii) directly contacting at least a portion of the first enriched $\gamma\delta$ T-cell population with one or more second agents that (a) expand $\gamma\delta$ T-cells, or (b) selectively expand:

- 5 - $\delta 1$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR; or
- $\delta 2$ T-cells by binding to an activating epitope specific of a $\delta 2$ TCR;
- $\delta 1$ T cells, $\delta 3$ T cells, $\delta 4$ T cells, and $\delta 5$ T cells by binding to an activating epitope specific of a $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ TCR; or
- $\delta 1$ T cells and $\delta 4$ T cells by binding to an activating epitope specific of a $\delta 1$ and $\delta 4$ TCR in
- 10 a second $\gamma\delta$ T-cell expansion, thereby producing a second enriched $\gamma\delta$ T-cell population.

63. The method of claim 62, wherein one or more of the second activating agents are structurally different from one or more of the activating agents of the first $\gamma\delta$ T-cell expansion.

15 64. The method of claim 63, wherein the second activation agents are structurally different from the first activating agents of the first $\gamma\delta$ T-cell expansion.

65. The method of any one of claims 62-64, wherein the one or more first agents are agents that selectively expand $\delta 1$ T-cells; $\delta 1$ T-cells, $\delta 3$ T-cells, $\delta 4$ T-cells, and $\delta 5$ T-cells; or $\delta 1$ T-cells and $\delta 4$

20 T-cells, and are selected from agents that bind to the same epitope as an antibody selected from the group consisting of $\delta 1$ -05, $\delta 1$ -08, $\delta 1$ -18, $\delta 1$ -22, $\delta 1$ -23, $\delta 1$ -26, $\delta 1$ -35, $\delta 1$ -37, $\delta 1$ -39, $\delta 1$ -113, $\delta 1$ -143, $\delta 1$ -149, $\delta 1$ -155, $\delta 1$ -182, $\delta 1$ -183, $\delta 1$ -191, $\delta 1$ -192, $\delta 1$ -195, $\delta 1$ -197, $\delta 1$ -199, $\delta 1$ -201, $\delta 1$ -203, $\delta 1$ -239, $\delta 1$ -253, $\delta 1$ -257, $\delta 1$ -278, $\delta 1$ -282, $\delta 1$ -285, and optionally further selected from TS-1, R9.12, and TS8.2.

25 66. The method of claim 65, wherein the one or more second agents are agents that selectively expand $\delta 1$ T-cells; $\delta 1$ T-cells, $\delta 3$ T-cells, $\delta 4$ T-cells, and $\delta 5$ T-cells; or $\delta 1$ T-cells and $\delta 4$ T-cells, and are selected from agents that bind to the same epitope as an antibody selected from the group consisting of $\delta 1$ -05, $\delta 1$ -08, $\delta 1$ -18, $\delta 1$ -22, $\delta 1$ -23, $\delta 1$ -26, $\delta 1$ -35, $\delta 1$ -37, $\delta 1$ -39, $\delta 1$ -113, $\delta 1$ -143, $\delta 1$ -149,

δ 1-155, δ 1-182, δ 1-183, δ 1-191, δ 1-192, δ 1-195, δ 1-197, δ 1-199, δ 1-201, δ 1-203, δ 1-239, δ 1-253, δ 1-257, δ 1-278, δ 1-282, δ 1-285, and optionally further selected from TS-1, R9.12, and TS8.2.

5 67. The method of any one of claims 62-64, wherein the one or more first agents are agents that selectively expand δ 2 T-cells and are selected from agents that bind to the same epitope as an antibody selected from the group consisting of δ 2-14, δ 2-17, δ 2-22, δ 2-30, δ 2-31, δ 2-32, δ 2-33, δ 2-35, δ 2-36, and δ 2-37, and optionally further selected from B6, and 15D.

10 68. The method of claim 67, wherein the one or more second agents are agents that selectively expand δ 2 T-cells and are selected from agents that bind to the same epitope as an antibody selected from the group consisting of δ 2-14, δ 2-17, δ 2-22, δ 2-30, δ 2-31, δ 2-32, δ 2-33, δ 2-35, δ 2-36, and δ 2-37, and optionally further selected from B6, and 15D.

15 69. The method of any one of claims 1, 48, 50-59, 62-65, or 66, wherein the percentage of δ 1 T cells in the enriched $\gamma\delta$ T cell population is greater than 60% δ 1 T-cells.

20 70. The method of claim 69, wherein the agents that selectively expand δ 1 T-cells; δ 1 T-cells, δ 3 T-cells, δ 4 T-cells, and δ 5 T-cells; or δ 1 T-cells and δ 4 T-cells are agents that do not compete with TS-1 or TS8.2 antibody, or do not compete with TS-1, TS8.2, or R9.12 antibody.

25 71. The method of claim 70, wherein the agents that selectively expand δ 1 T-cells; δ 1 T-cells, δ 3 T-cells, δ 4 T-cells, and δ 5 T-cells; or δ 1 T-cells and δ 4 T-cells are agents that bind a δ 1 TCR Bin 1 δ 1 epitope, Bin 1b δ 1 epitope, Bin 2 δ 1 epitope, Bin 2b δ 1 epitope, Bin 2c δ 1 epitope, Bin 3 δ 1 epitope, Bin 4 δ 1 epitope, Bin 5 δ 1 epitope, Bin 6 δ 1 epitope, Bin 7 δ 1 epitope, Bin 8 δ 1 epitope, or Bin 9 δ 1 epitope of a human δ 1 TCR.

72. The method of any one of claims 1, 49, 54-57, 60-64, 67, or 68, wherein the percentage of δ 2 T cells in the enriched $\gamma\delta$ T cell population is greater than 60% δ 2 T-cells.

73. The method of claim 72, wherein the agents that selectively expand $\delta 2$ T-cells are agents that do not compete with B6 or 15D antibody.

74. The method of claim 73, wherein the agents that selectively expand $\delta 2$ T-cells are agents that bind a $\delta 2$ TCR epitope within residues 72-83, or 1-27 of a human $\delta 2$ TCR.

75. The method of claim 72, wherein the agents that selectively expand $\delta 2$ T-cells are agents that bind a $\delta 2$ TCR Bin 1 $\delta 2$ epitope, Bin 2 $\delta 2$ epitope, Bin 3 $\delta 2$ epitope, or Bin 4 $\delta 2$ epitope of a human $\delta 2$ TCR.

76. The method of any one of claims 1-19, or 48-75, wherein the method comprises serum-free culture conditions and/or suspension cell culture conditions.

76. The method of any one of claims 1-19, or 48-75, wherein the method comprises serum-free culture conditions.

77. The method of any one of claims 1-19, or 48-76, wherein the method achieves at least about 10^8 $\gamma\delta$ T cells expanded from the isolated mixed population of cells within less than 90 days, less than 60 days, less than 30 days, less than 21 days, or less than 19 days from:

- initiation of a first expansion,
- a step of providing a donor sample, or
- a first step of directly contacting the mixed cell population with one or more activation agents.

78. An *ex vivo* method for producing an expanded population of engineered $\gamma\delta$ T-cells, comprising directly contacting one or more engineered $\gamma\delta$ T-cells with one or more agents which selectively expand:

- $\delta 1$ T-cells by binding to an activating epitope specific of a $\delta 1$ TCR;
- $\delta 2$ T-cells by binding to an activating epitope specific of a $\delta 2$ TCR;

- δ 1 T cells, δ 3 T cells, δ 4 T cells, and δ 5 T cells by binding to an activating epitope specific of a δ 1, δ 3, δ 4, and δ 5 TCR; or

- δ 1 T cells and δ 4 T cells by binding to an activating epitope specific of a δ 1 and δ 4 TCR to provide clinically-relevant levels of engineered $\gamma\delta$ T-cells.

5

79. An expanded $\gamma\delta$ T-cell population obtained according to the method of any one of claims 1-19, or 48-78.

80. A method for treating a cancer, an infectious disease, or an inflammatory disease in a subject in need thereof, comprising administering a therapeutically effective amount of an expanded $\gamma\delta$ T-cell population according to claim 79.

81. An antibody that binds to the same, or essentially the same, epitope as, or competes with, an antibody selected from the group consisting of δ 1-05, δ 1-08, δ 1-18, δ 1-22, δ 1-23, δ 1-26, δ 1-35, δ 1-37, δ 1-39, δ 1-113, δ 1-143, δ 1-149, δ 1-155, δ 1-182, δ 1-183, δ 1-191, δ 1-192, δ 1-195, δ 1-197, δ 1-199, δ 1-201, δ 1-203, δ 1-239, δ 1-253, δ 1-257, δ 1-278, δ 1-282, δ 1-285, δ 2-14, δ 2-17, δ 2-22, δ 2-30, δ 2-31, δ 2-32, δ 2-33, δ 2-35, δ 2-36, and δ 2-37.

82. An antibody that binds to a Bin 1 δ 2 epitope, Bin 2 δ 2 epitope, Bin 3 δ 2 epitope, or Bin 4 δ 2 epitope of a human δ 2 TCR; or a Bin 1 δ 1 epitope, Bin 1b δ 1 epitope, Bin 2 δ 1 epitope, Bin 2b δ 1 epitope, Bin 2c δ 1 epitope, Bin 3 δ 1 epitope, Bin 4 δ 1 epitope, Bin 5 δ 1 epitope, Bin 6 δ 1 epitope, Bin 7 δ 1 epitope, Bin 8 δ 1 epitope, or Bin 9 δ 1 epitope of a human δ 1 TCR.

83. The antibody of claim 81 or 82, wherein the antibody is not TS8.2, TS-1, or R9.12.

25

84. The antibody of claim 81 or 82, wherein the antibody comprises the complementary determining regions of any one of the antibodies selected from the group consisting of δ 1-05, δ 1-08, δ 1-18, δ 1-22, δ 1-23, δ 1-26, δ 1-35, δ 1-37, δ 1-39, δ 1-113, δ 1-143, δ 1-149, δ 1-155, δ 1-182, δ 1-183, δ 1-191, δ 1-192, δ 1-195, δ 1-197, δ 1-199, δ 1-201, δ 1-203, δ 1-239, δ 1-253, δ 1-257, δ 1-278, δ 1-282, δ 1-285, δ 2-14, δ 2-17, δ 2-22, δ 2-30, δ 2-31, δ 2-32, δ 2-33, δ 2-35, δ 2-36, and δ 2-37.

30

85. The antibody of claim 81 or 82, wherein the antibody specifically activates $\gamma\delta 1$ T cells as compared to $\gamma\delta 2$ T cells and as compared to $\alpha\beta$ T cells in a mixed cell population containing $\gamma\delta 1$ T cells, $\gamma\delta 2$ T cells, and $\alpha\beta$ T cells.

5

86. The antibody of claim 81 or 82, wherein the antibody specifically activates $\gamma\delta 2$ T cells as compared to $\gamma\delta 1$ T cells and as compared to $\alpha\beta$ T cells in a mixed cell population containing $\gamma\delta 1$ T cells, $\gamma\delta 2$ T cells, and $\alpha\beta$ T cells.

10 87. A nucleic acid encoding an antibody that binds an epitope specific of a $\delta 1$ TCR; a $\delta 2$ TCR; a $\delta 1$ and $\delta 4$ TCR; or a $\delta 1$, $\delta 3$, $\delta 4$, and $\delta 5$ TCR, wherein the nucleic acid is operably linked to a heterologous promoter.

88. The nucleic acid of claim 87, wherein:

15 (i) the nucleic acid encodes an antibody that binds the same epitope, or essentially the same epitope, as, or competes with, an antibody selected from the group consisting of $\delta 1$ -05, $\delta 1$ -08, $\delta 1$ -18, $\delta 1$ -22, $\delta 1$ -23, $\delta 1$ -26, $\delta 1$ -35, $\delta 1$ -37, $\delta 1$ -39, $\delta 1$ -113, $\delta 1$ -143, $\delta 1$ -149, $\delta 1$ -155, $\delta 1$ -182, $\delta 1$ -183, $\delta 1$ -191, $\delta 1$ -192, $\delta 1$ -195, $\delta 1$ -197, $\delta 1$ -199, $\delta 1$ -201, $\delta 1$ -203, $\delta 1$ -239, $\delta 1$ -253, $\delta 1$ -257, $\delta 1$ -278, $\delta 1$ -282, $\delta 1$ -285, and optionally further selected from TS-1, R9.12, and TS8.2; or

20 (ii) the nucleic acid encodes an antibody that binds the same epitope, or essentially the same epitope, as, or competes with, an antibody selected from the group consisting of $\delta 2$ -14, $\delta 2$ -17, $\delta 2$ -22, $\delta 2$ -30, $\delta 2$ -31, $\delta 2$ -32, $\delta 2$ -33, $\delta 2$ -35, $\delta 2$ -36, and $\delta 2$ -37, and optionally further selected from B6, and 15D.

25 89. The nucleic acid of claim 87, wherein:

(i) the nucleic acid encodes an antibody that comprises the CDRs of an antibody selected from the group consisting of $\delta 1$ -05, $\delta 1$ -08, $\delta 1$ -18, $\delta 1$ -22, $\delta 1$ -23, $\delta 1$ -26, $\delta 1$ -35, $\delta 1$ -37, $\delta 1$ -39, $\delta 1$ -113, $\delta 1$ -143, $\delta 1$ -149, $\delta 1$ -155, $\delta 1$ -182, $\delta 1$ -183, $\delta 1$ -191, $\delta 1$ -192, $\delta 1$ -195, $\delta 1$ -197, $\delta 1$ -199, $\delta 1$ -201, $\delta 1$ -203, $\delta 1$ -239, $\delta 1$ -253, $\delta 1$ -257, $\delta 1$ -278, $\delta 1$ -282, $\delta 1$ -285, and optionally further selected from TS-1, R9.12, and TS8.2; or

30

(ii) the nucleic acid encodes an antibody that comprises the CDRs of an antibody selected from the group consisting of δ 2-14, δ 2-17, δ 2-22, δ 2-30, δ 2-31, δ 2-32, δ 2-33, δ 2-35, δ 2-36, and δ 2-37, and optionally further selected from B6, and 15D.

5 90. A host cell that comprises a nucleic acid according to any one of claims 87-89, and/or an antibody according to any one of claims 81-86.

91. The host cell of claim 90, wherein the nucleic acid encodes a membrane anchor or transmembrane domain fused to the antibody, wherein the antibody is presented on an extracellular
10 surface of the host cell.

92. An antigen presenting cell that comprises a heterologous nucleic acid, wherein the heterologous nucleic acid encodes a polypeptide, wherein the polypeptide is a cell surface receptor that is in contact with an antibody, wherein the antibody binds an epitope specific of a δ 1 TCR; a δ 2 TCR; a
15 δ 1 and δ 4 TCR; or a δ 1, δ 3, δ 4, and δ 5 TCR.

93. The antigen presenting cell of claim 92, wherein the cell surface receptor is in contact with an Fc region of the antibody.

20 94. The antigen presenting cell of claim 92 or 93, wherein the antibody is bound to a δ -chain of a $\gamma\delta$ TCR expressed on a surface of a $\gamma\delta$ TCR.

95. An expanded population of $\gamma\delta$ T-cells, wherein the $\gamma\delta$ T-cell population comprises anti-tumor cytotoxicity that is independent of NKp30 activity, NKp44 activity, and/or NKp46 activity.

25 96. The expanded population of $\gamma\delta$ T-cells of claim 95, wherein the $\gamma\delta$ T-cell population is a population of engineered $\gamma\delta$ T-cells.

97. The expanded population of $\gamma\delta$ T-cells of claim 95, wherein the $\gamma\delta$ T-cell population is a
30 population of non-engineered $\gamma\delta$ T-cells.

98. The expanded population of $\gamma\delta$ T-cells of claim 95, 96, or 97, wherein the $\gamma\delta$ T-cell population does not comprise NKp30 activity-dependent anti-tumor cytotoxicity, NKp44 activity-dependent anti-tumor cytotoxicity, and/or NKp46 activity-dependent anti-tumor cytotoxicity.

5

99. The expanded population of $\gamma\delta$ T-cells of claim 95, 96, or 97, wherein the $\gamma\delta$ T-cell population does not comprise NKp30 activity-dependent anti-tumor cytotoxicity.

99. The expanded population of $\gamma\delta$ T-cells of claim 95, 96, or 97, wherein the $\gamma\delta$ T-cell population does not comprise NKp44 activity-dependent anti-tumor cytotoxicity.

10

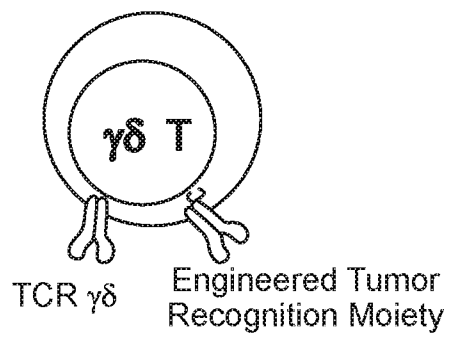
101. The expanded population of $\gamma\delta$ T-cells of any one of claims 95, 96, 98, 99, or 100, wherein the $\gamma\delta$ T-cell population is an engineered population of $\gamma\delta$ T-cells that comprise an anti-CD20 chimeric antigen receptor (CAR).

15

102. The expanded population of $\gamma\delta$ T-cells of any one of claims 95-101, wherein less than 40% of the cells express a detectable level of NKp30, NKp44, and/or NKp46.

1 / 55

Panel A



Panel B

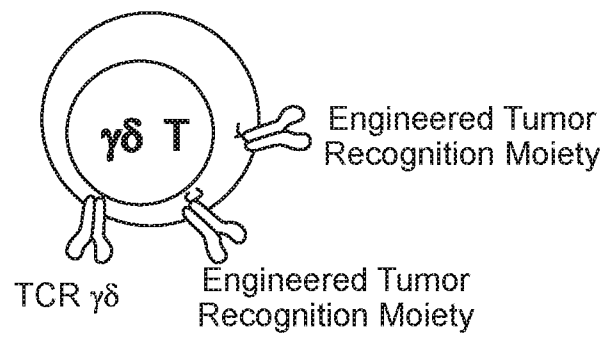


FIG. 1

2 / 55

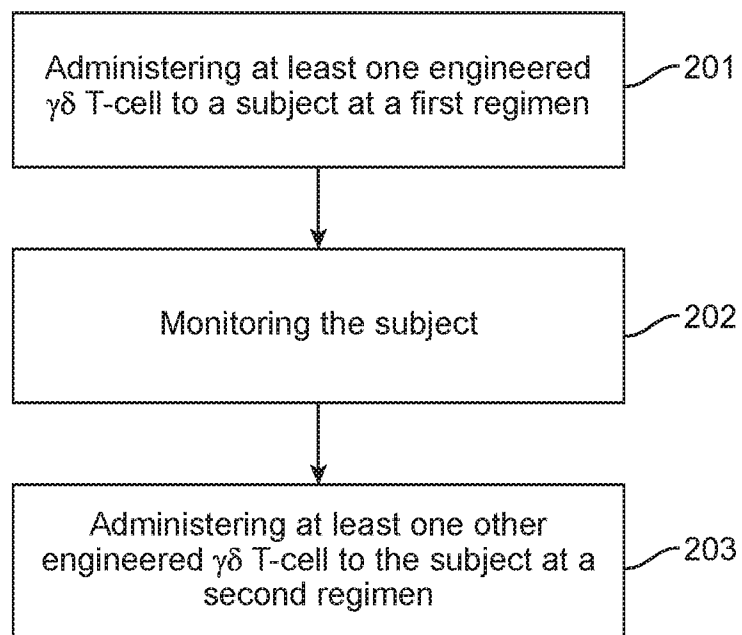


FIG. 2

3 / 55

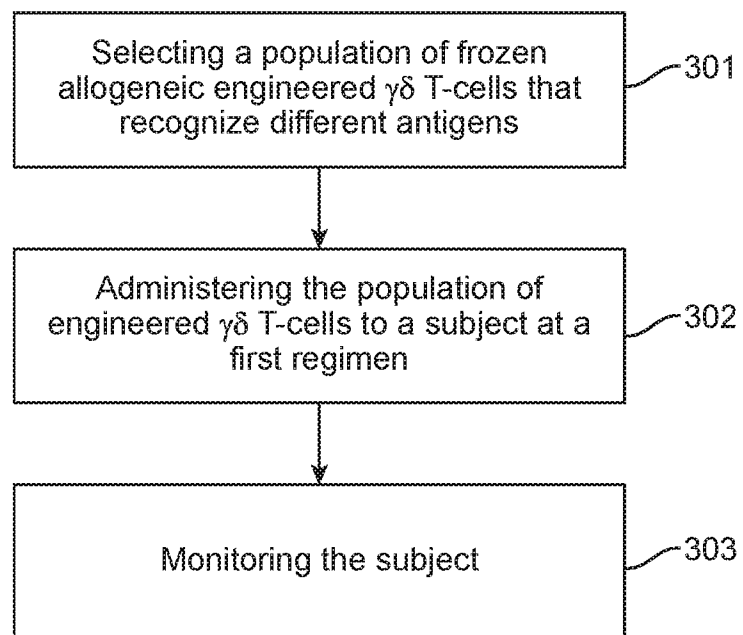


FIG. 3

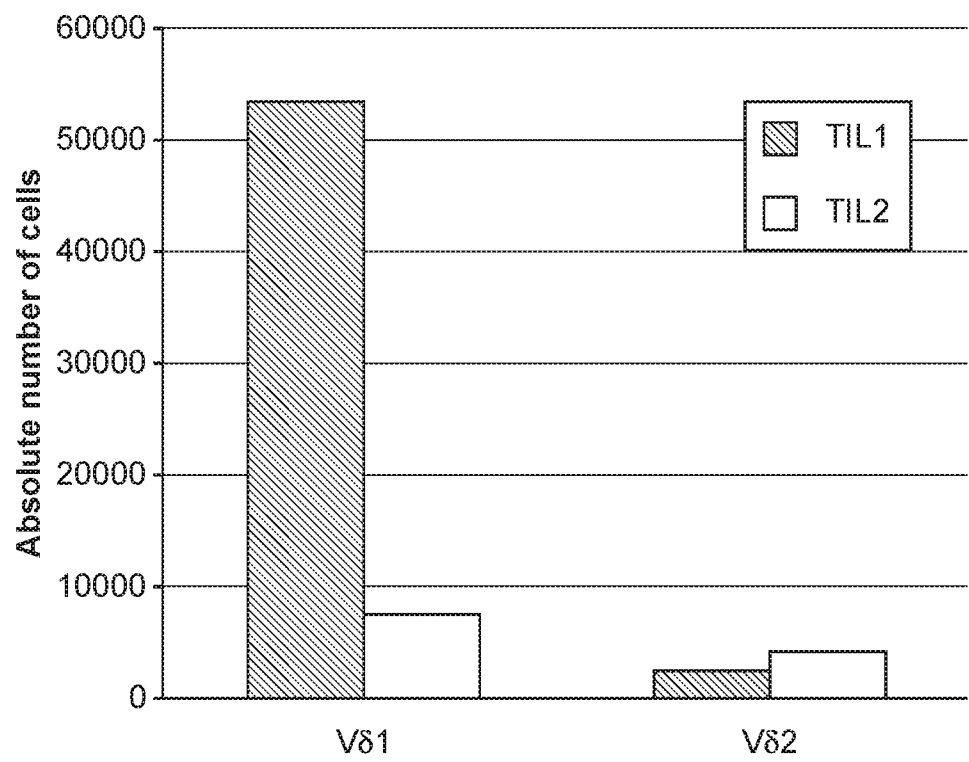


FIG. 4

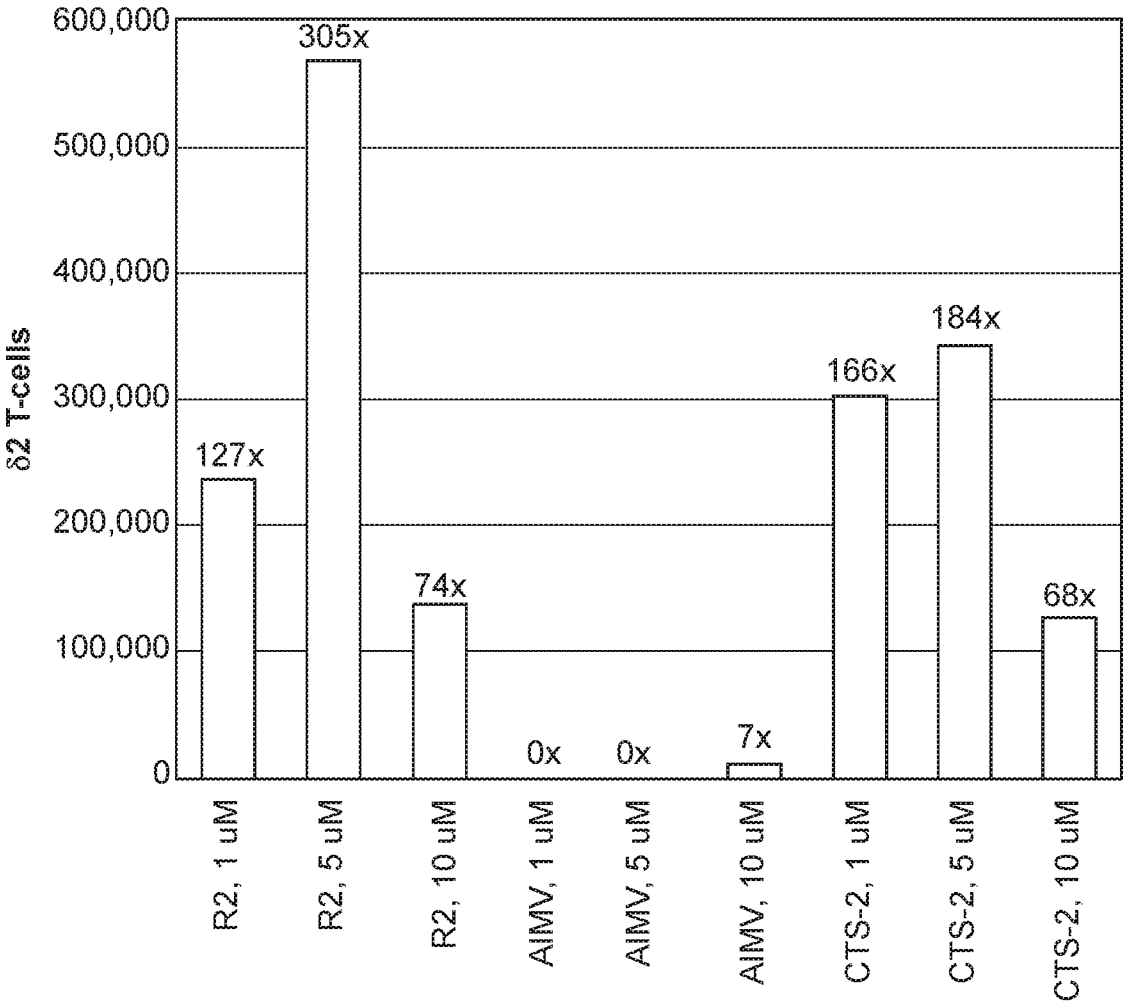


FIG. 5

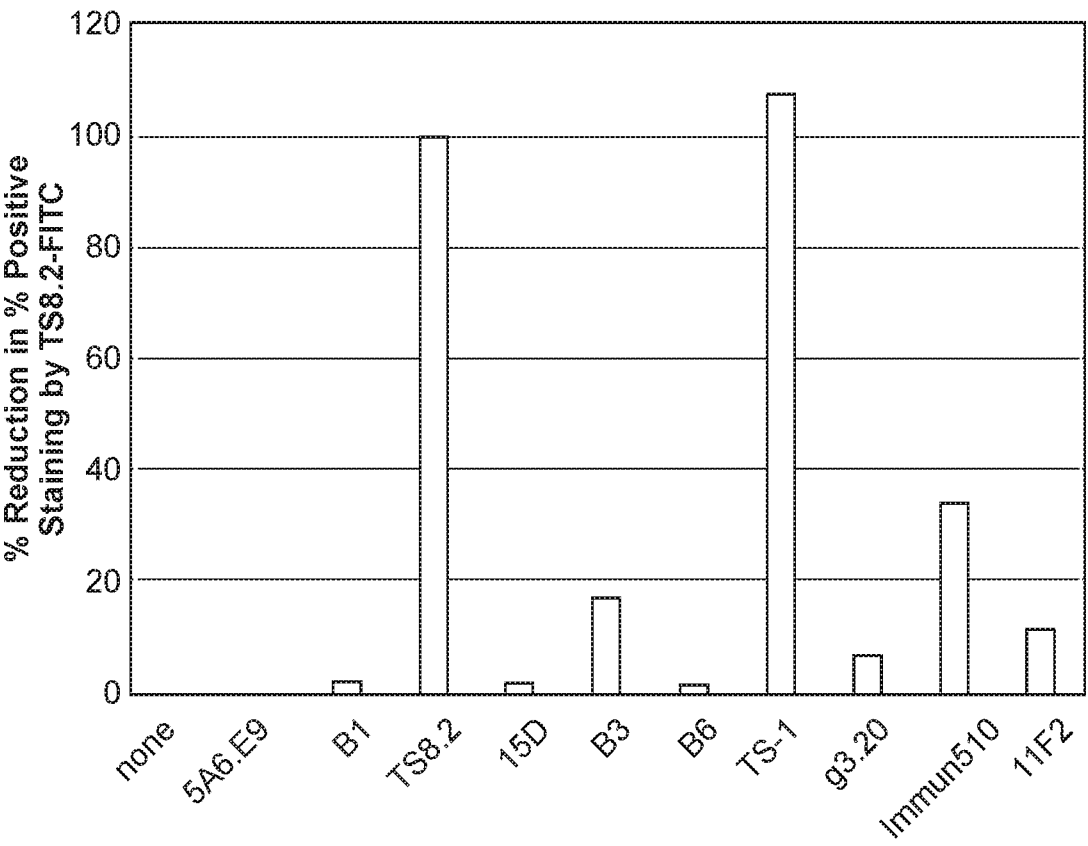


FIG. 6

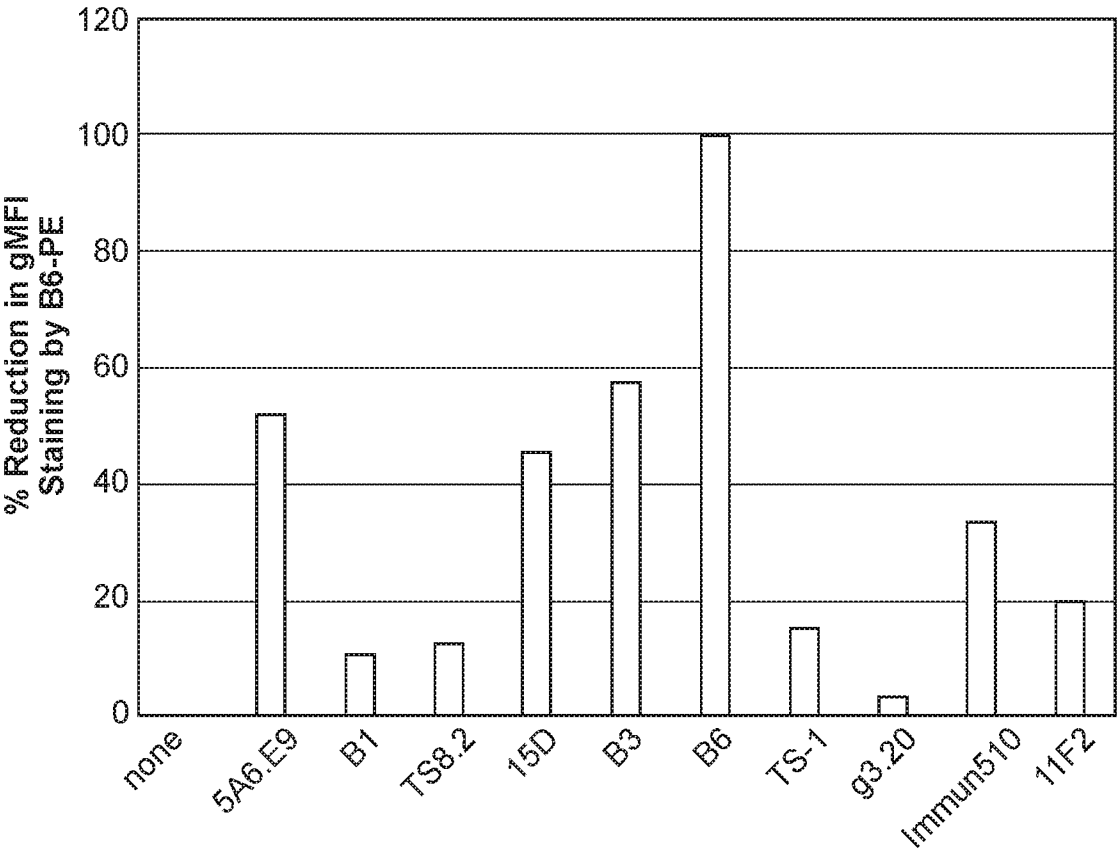


FIG. 7

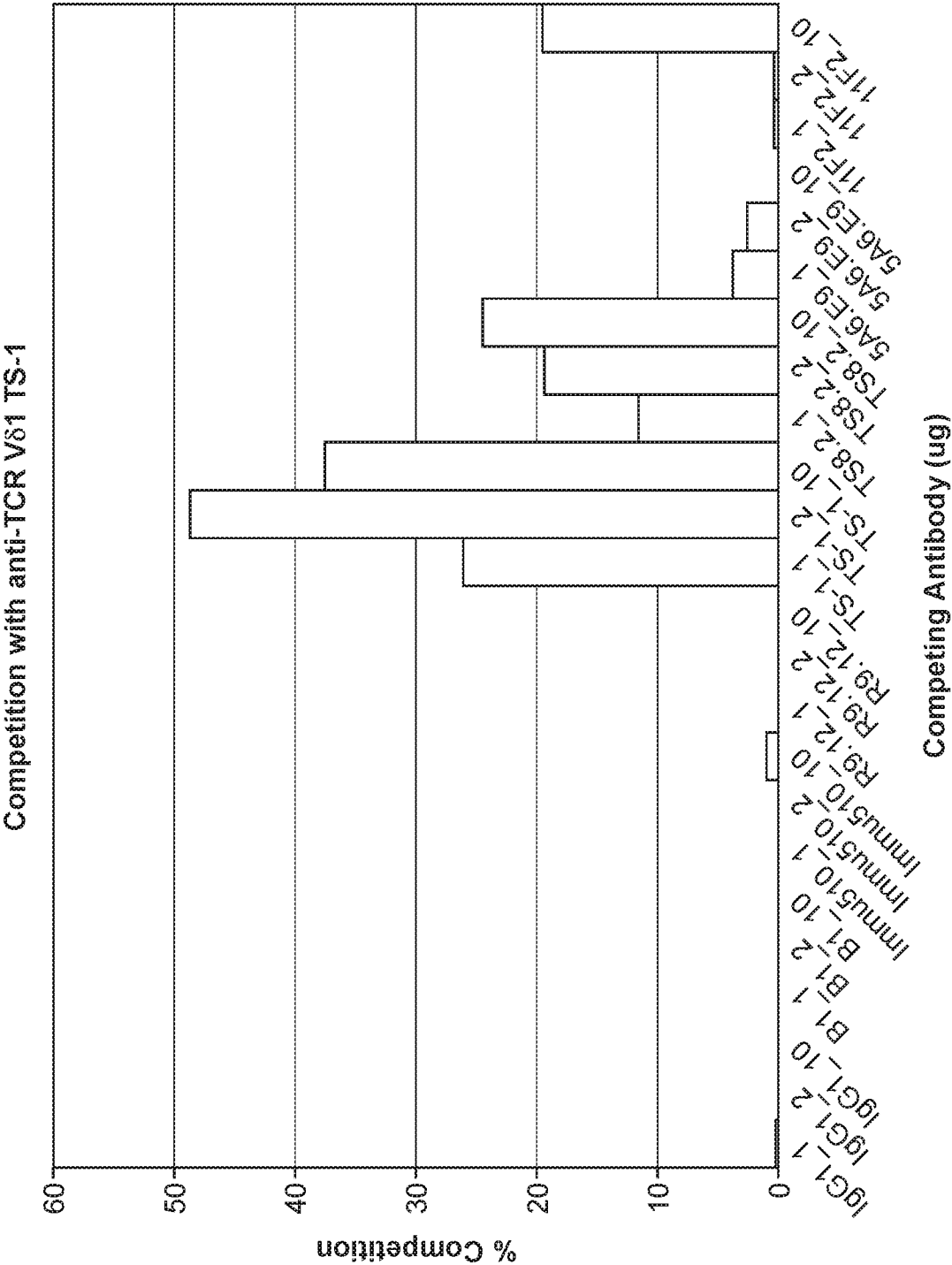


FIG. 8

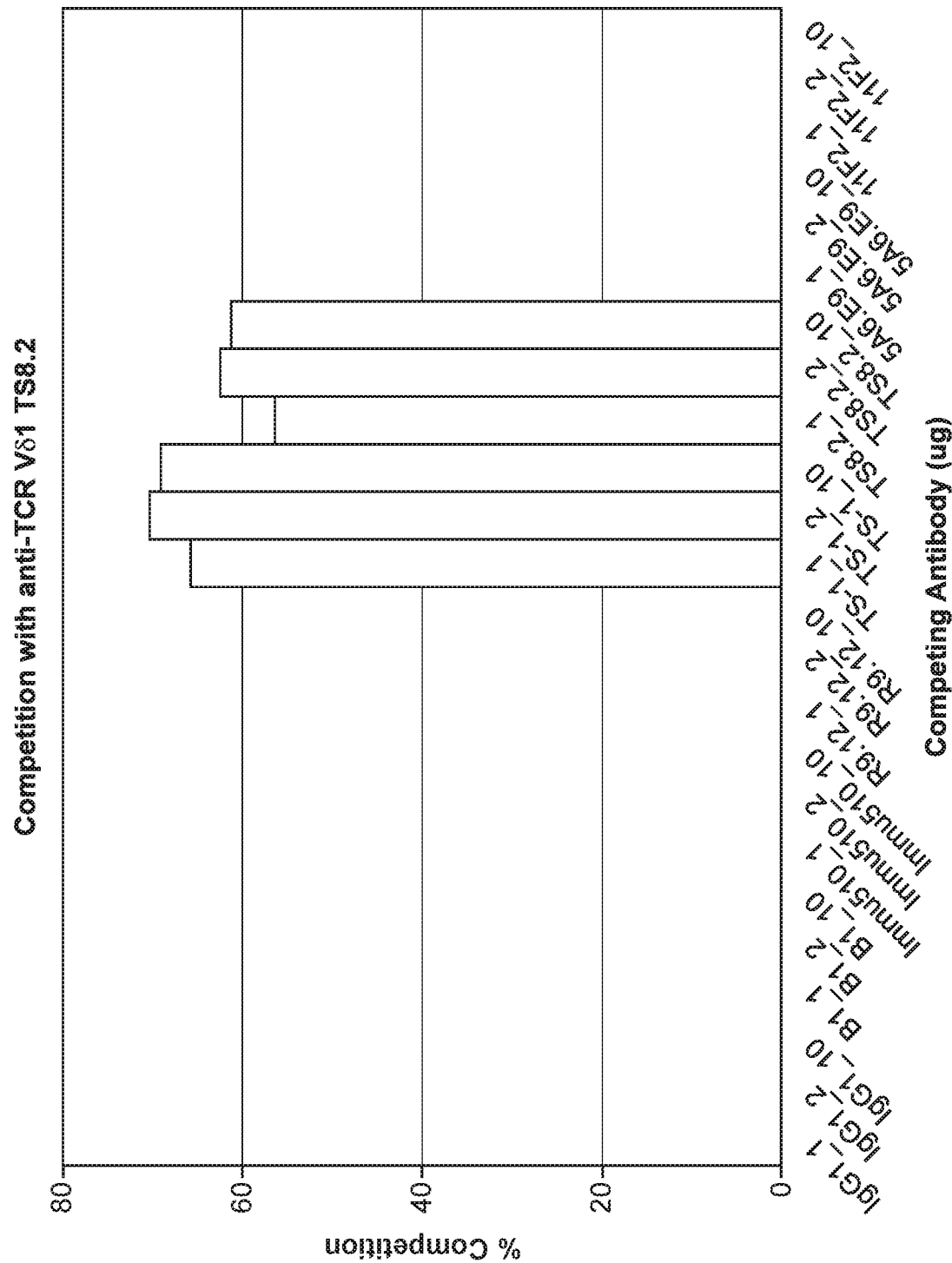
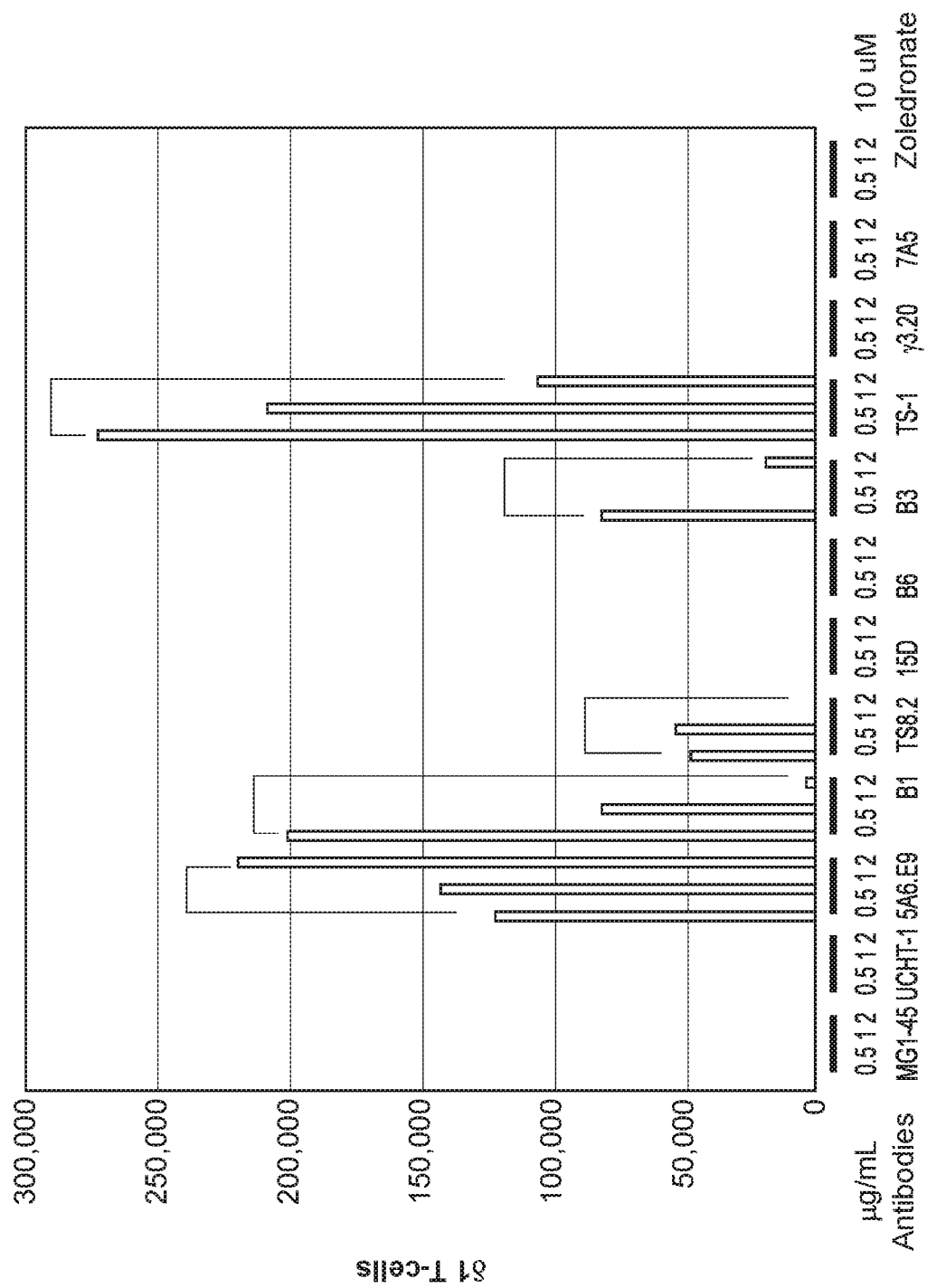
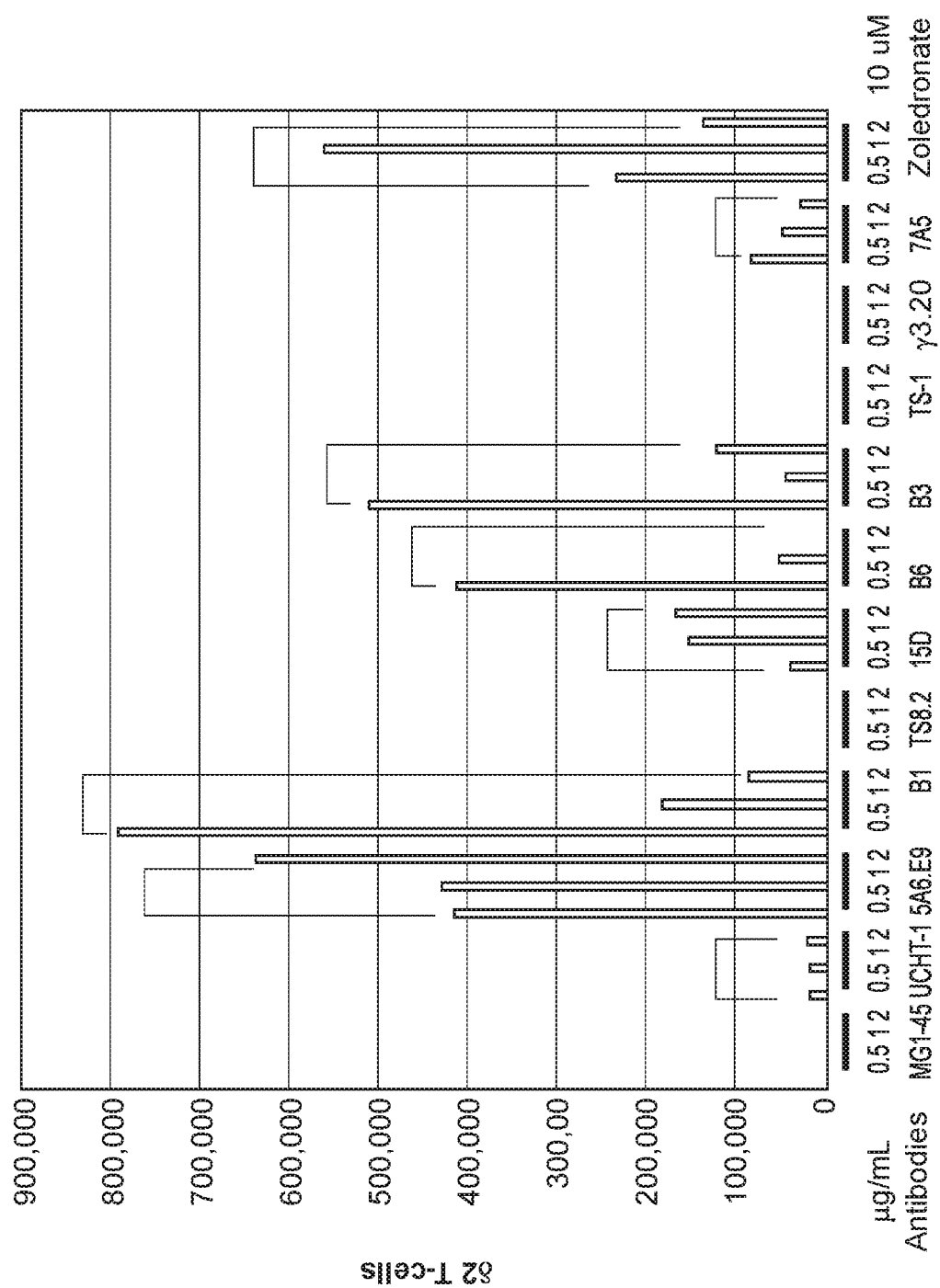


FIG. 9



10
G
E



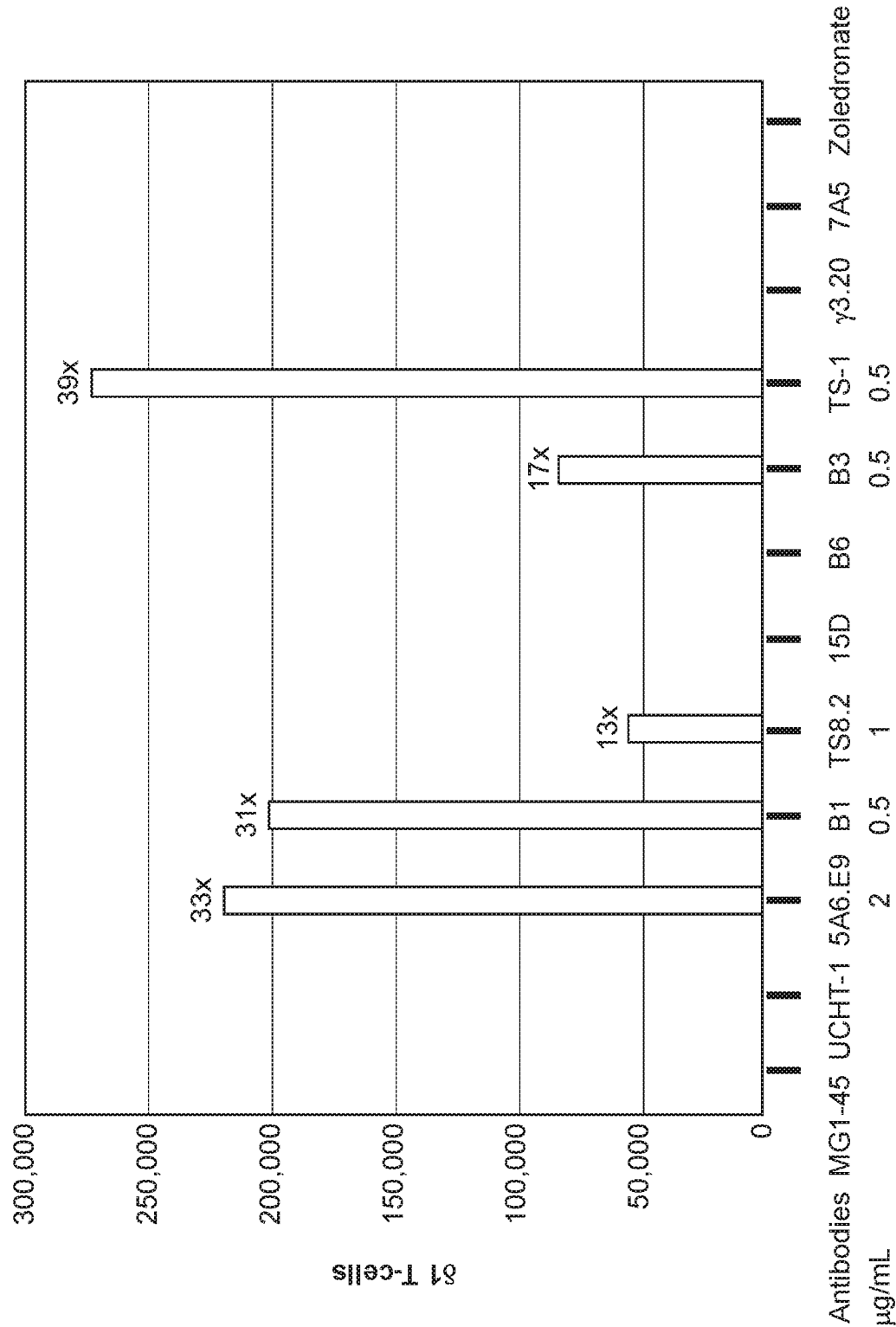


FIG. 12

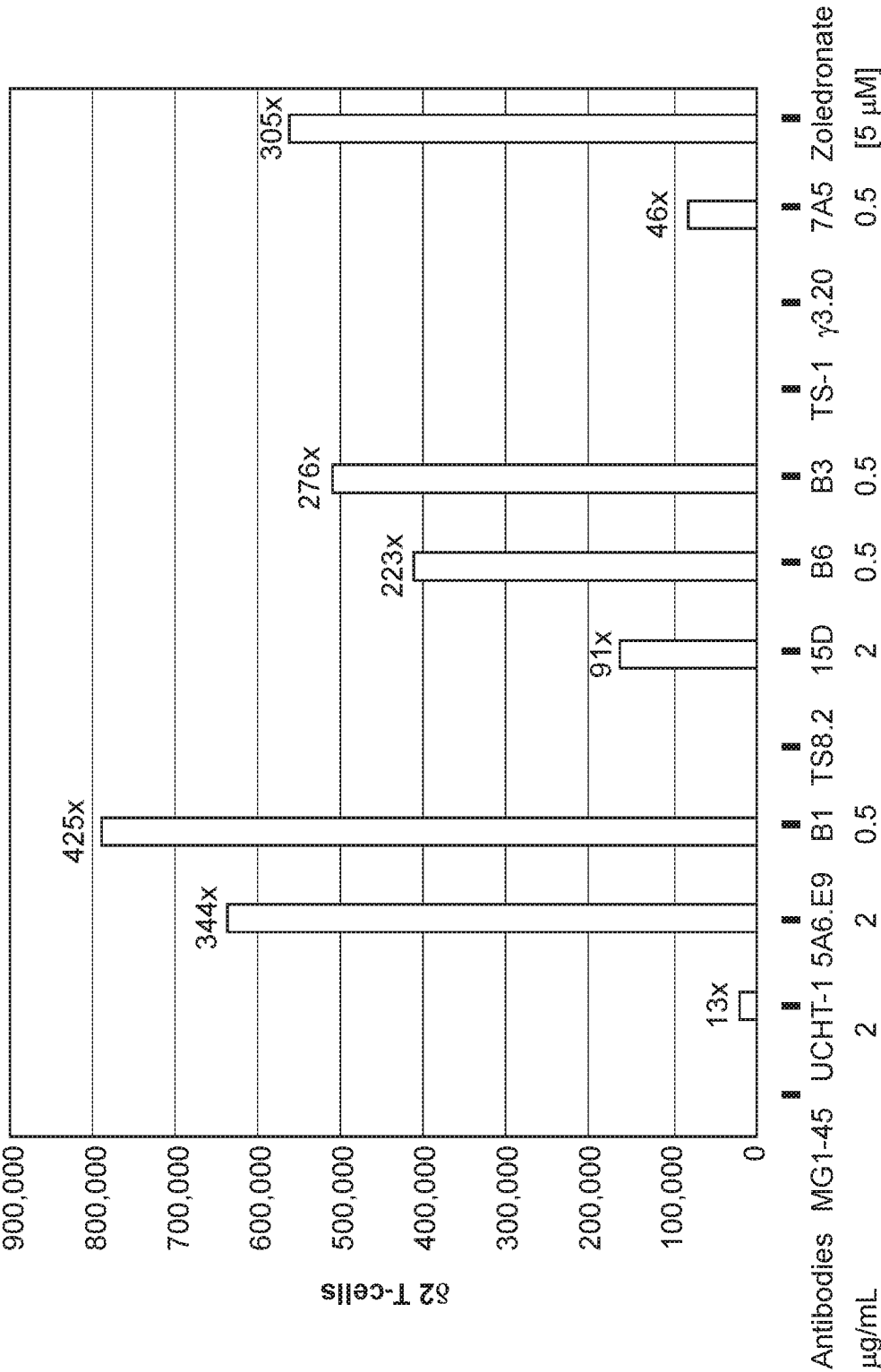


FIG. 13

V delta 1

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20
A Q K V T Q A Q S S S V S M P V R K A V T
GCC CAG AAG GTT ACT CAA GCC CAG TCA TCA GTA TCC ATG CCA GTG AGG AAA GCA GTC ACC

CDR1-IMGT
21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40
L N C L Y E T S W W S Y Y
CTG AAC TGC CTG TAT GAA ACA AGT TGG TGG TCA TAT TAT ATT TTT

CDR2-
41 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60
W Y K Q L P S K E M I F L I R Q G S
TGG TAC AAG CAA CTT CCC AGC AGC AAA GAG ATG ATT TTC CTT ATT CGC CAG GGT TCT

IMGT

61 62 63 64 65 66 67 68 69 70 71 72 73 74 75 76 77 78 79 80
D E Q N A K S G R Y S V N F
... .. . GAT GAA CAG AAT GCA AAA AGT ... GGT CGC TAT TCT GTC AAC TTC
81 82 83 84 85 86 87 88 89 90 91 92 93 94 95 96 97 98 99 100
K K A A K S V A L T I S A L Q L E D S A
AAG AAA GCA GCG AAA TCC GTC GCC TTA ACC ATT TCA GCC TTA CAG CTA GAA GAT TCA GCA

CDR3-IMGT
101 102 103 104 105 106 107 108
K Y F C A L G E
AAG TAC TTT TGT GCT CTT GGG GAA CT

FIG. 14

V delta 2																															
1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20												
A	I	E	L	V	P	E	H	Q	T	V	P	V	S	I	G	V	P	A	T												
GCC ATT GAG TTG GTG CCT GAA CAC CAA ACA GTG CCT GTG TCA ATA GGG GTC CCT GCC ACC																															
I																															
A																															

CDR1-IMGT																															
21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40												
L	R	C	S	M	K	G	E	A	I					G	N	Y	Y	I	N												
CTC AGG TGC TCC ATG AAA GGA GAA GCG ATC GGT AAC TAC TAT ATC AAC																															
CDR2-																															
41	42	43	44	45	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60												
W	Y	R	K	T	Q	G	N	T	I	T	F	I	Y	R	E	K	D														
TGG TAC AGG AAG ACC CAA GGT AAC ACA ATC ACT TTC ATA TAC CGA GAA AAG GAC																															
M																															

G																															
IMGT																															
61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77	78	79	80												
					I	Y	G	P	G	F	K		D	N	F	Q	G	D	I												
.... ATC TAT GGC CCT GGT TTC AAA ... GAC AAT TTC CAA GGT GAC ATT																															
81	82	83	84	85	86	87	88	89	90	91	92	93	94	95	96	97	98	99	100												
D	I	A	K	N	L	A	V	L	K	I	L	A	P	S	E	R	D	E	G												
GAT ATT GCA AAG AAC CTG GCT GTA CTT AAG ATA CTT GCA CCA TCA GAG AGA GAT GAA GGG																															
CDR3-IMGT																															
101	102	103	104	105	106	107	108																								
S	Y	Y	C	A	C	D	T																								
TCT TAC TAC TGT GCC TGT GAC ACC																															

FIG 15

FIG. 15

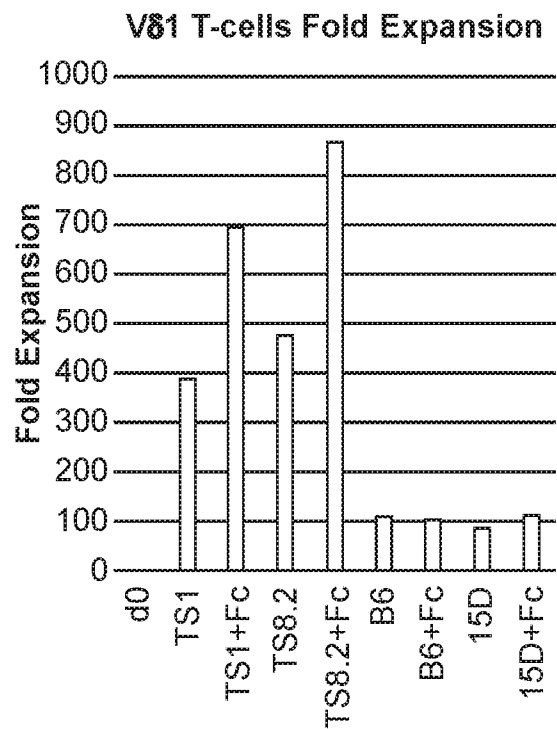


FIG. 16A

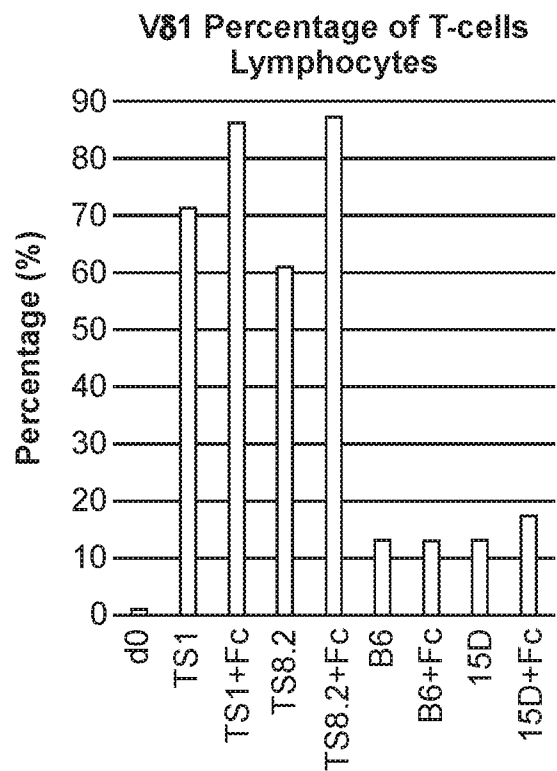


FIG. 16B

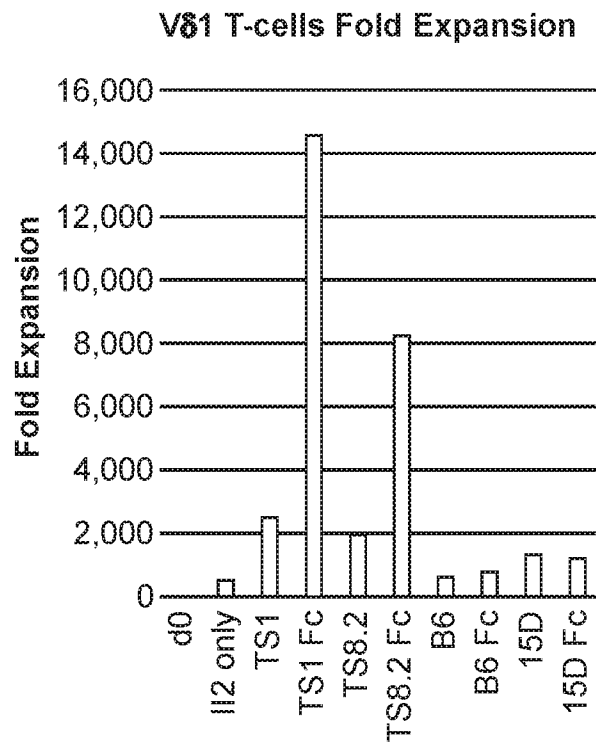


FIG. 16C

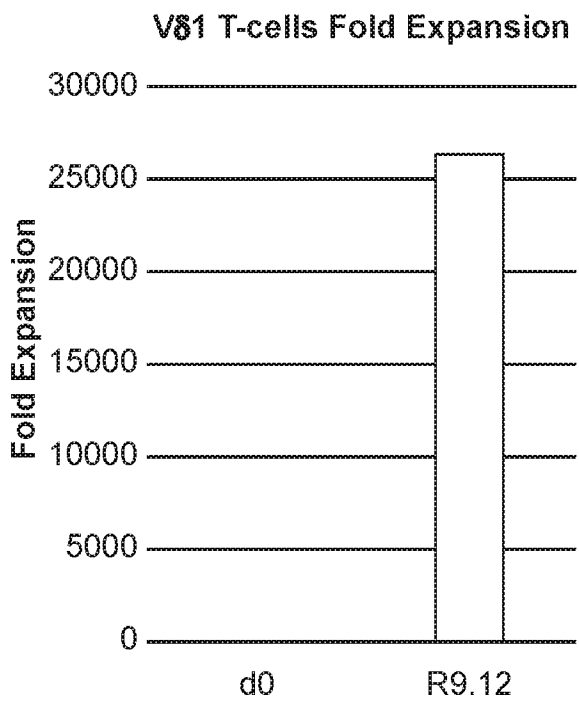


FIG. 16D

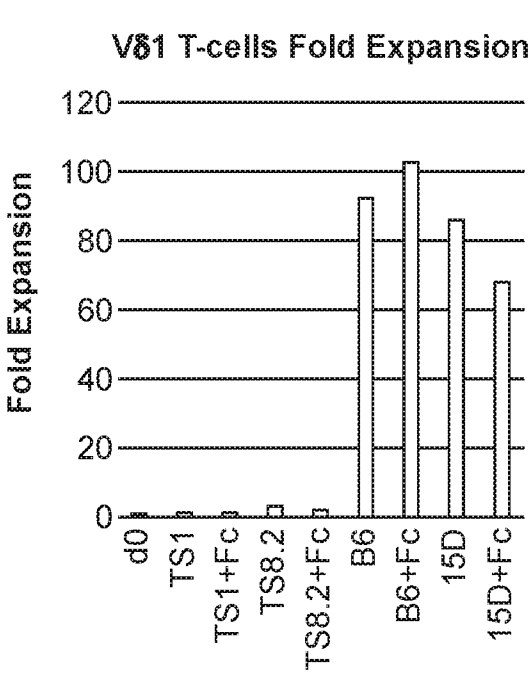


FIG. 17A

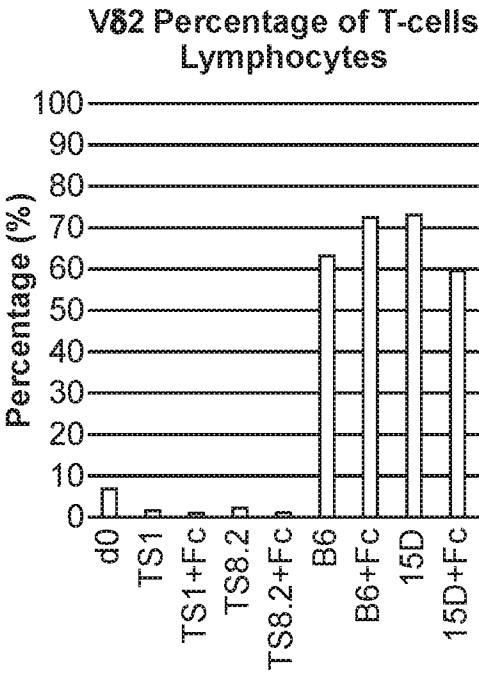


FIG. 17B

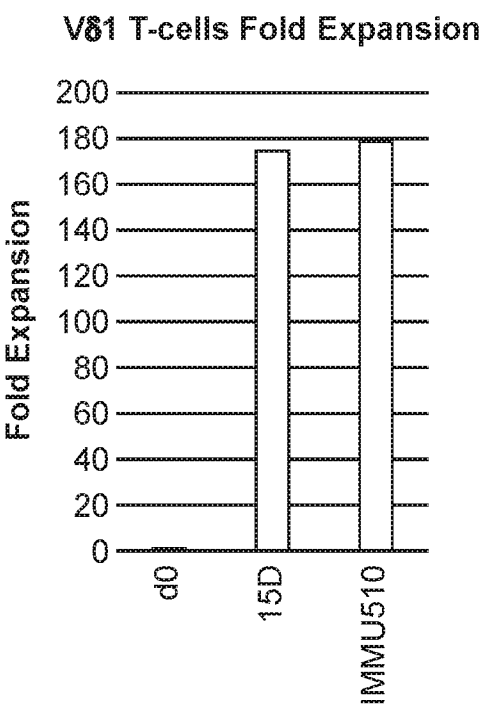


FIG. 17C

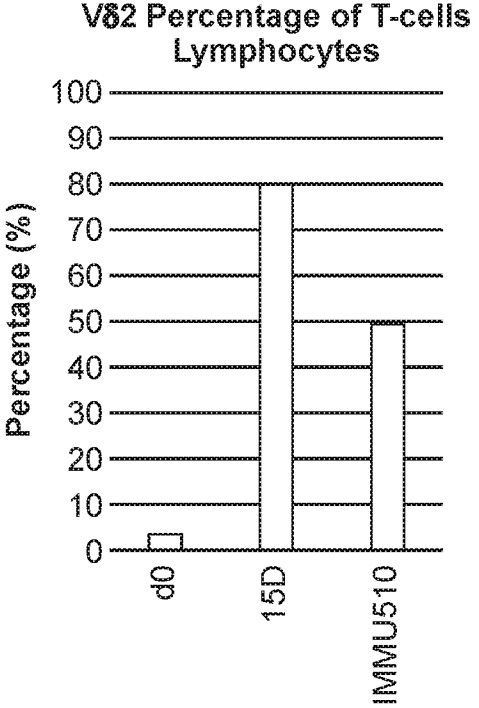


FIG. 17D

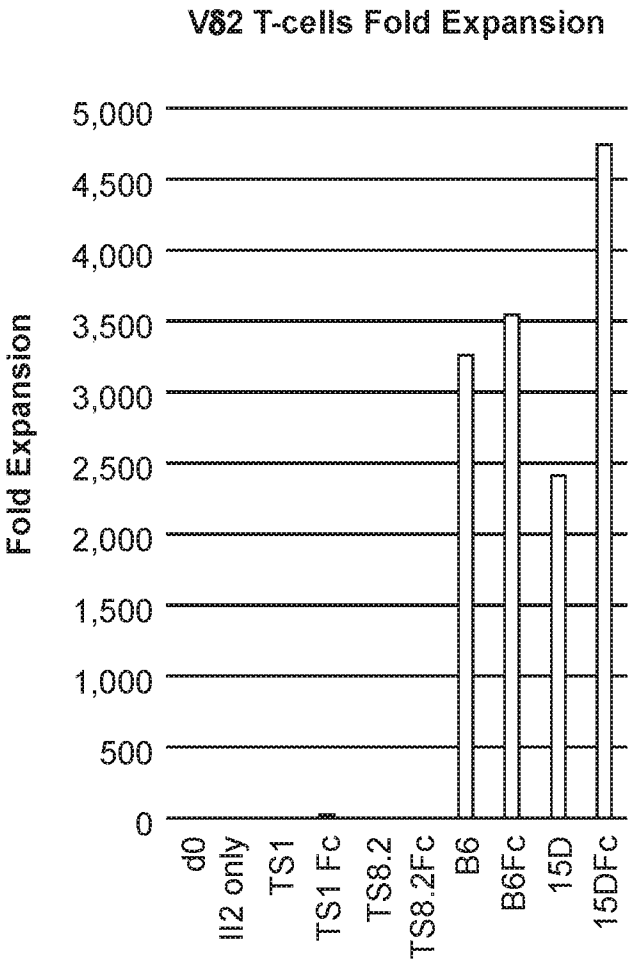


FIG. 17E

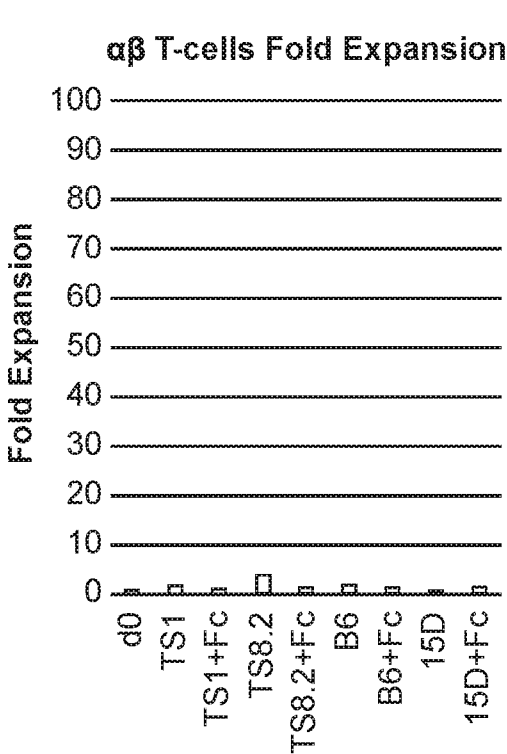


FIG. 18A

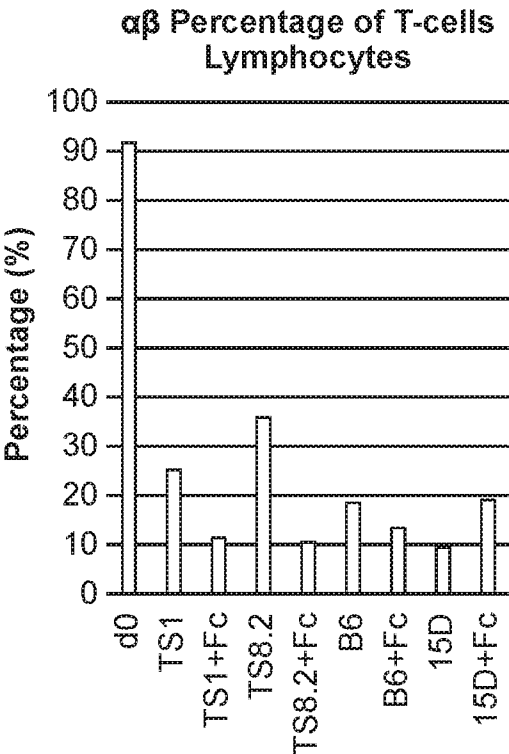


FIG. 18B

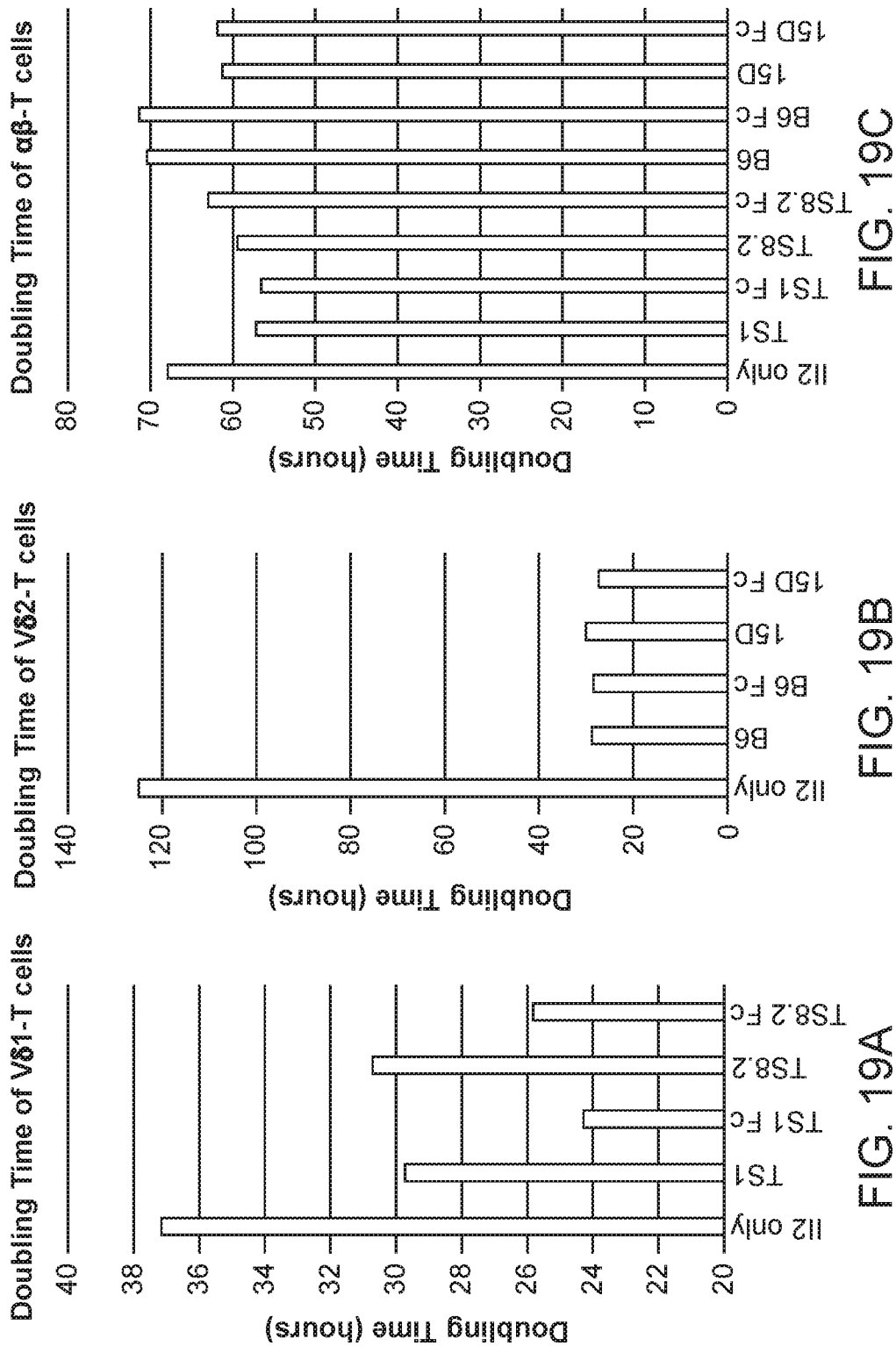


FIG. 19C

FIG. 19B

FIG. 19A

21 / 55

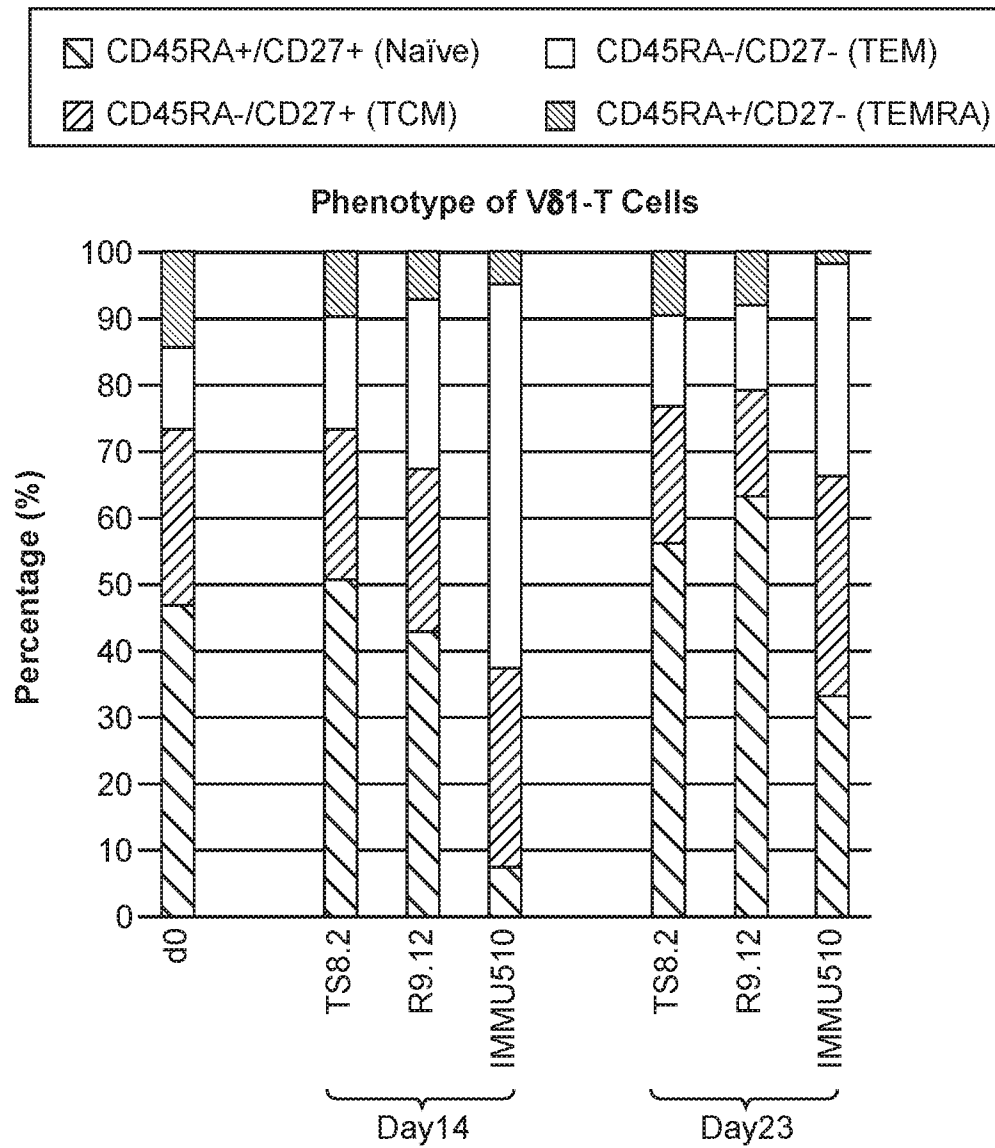


FIG. 20

22 / 55

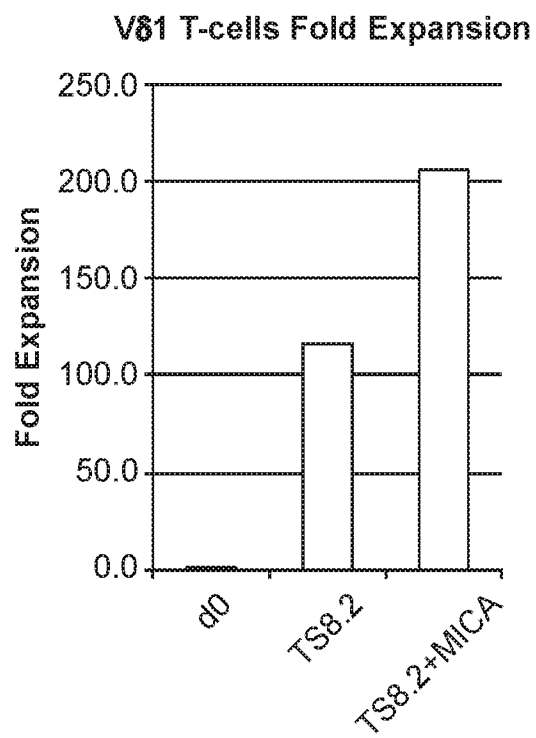


FIG. 21A

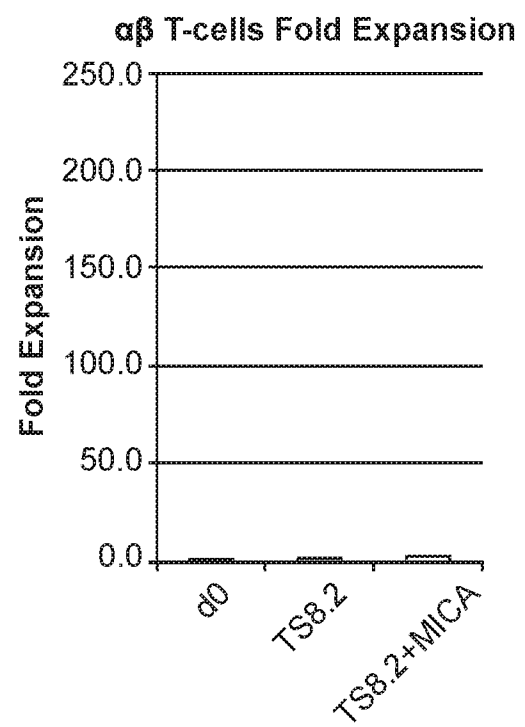


FIG. 21B

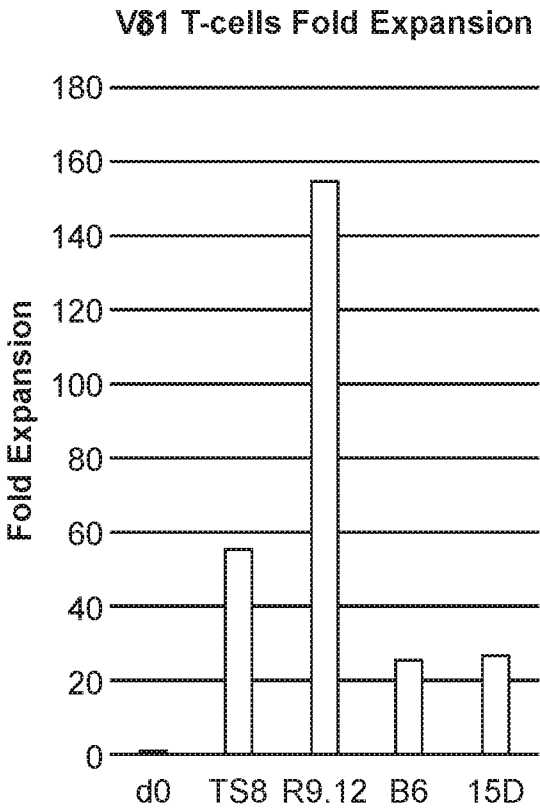


FIG. 22A

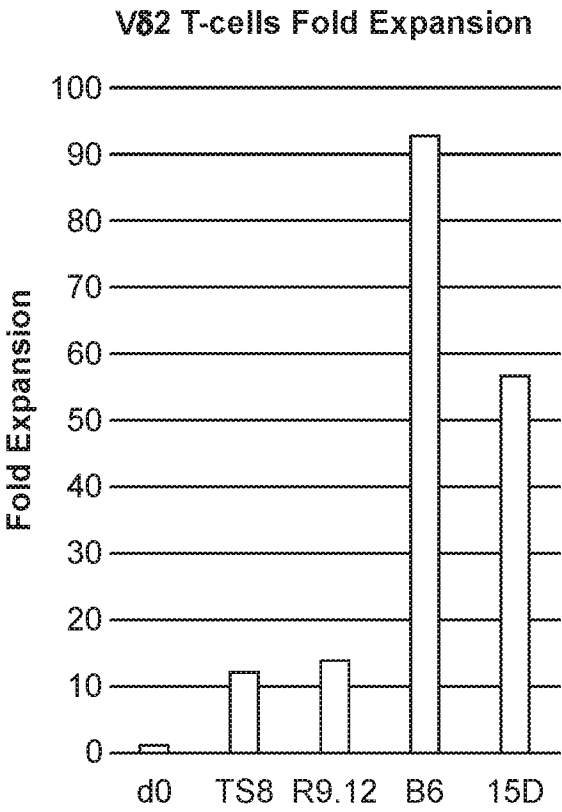


FIG. 22B

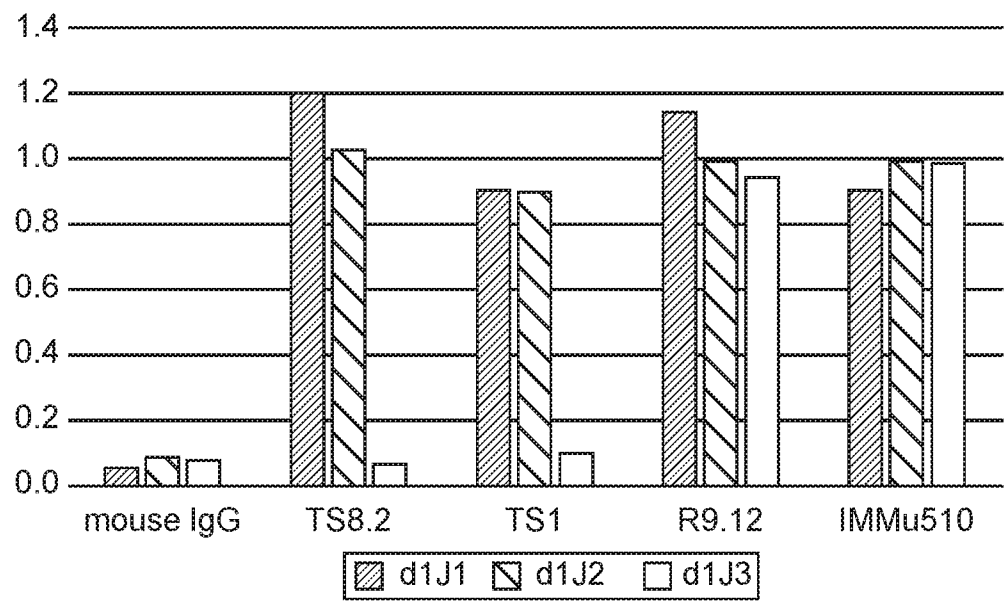


FIG. 23

V81	N-D-N	J	TS1 TS8.2 R9.12
BE-13	AKYFCALG--TGVRGLQD---TDKLIFC	KCTRVTVTEPRSQPHT	+ + +
δ1J1	AKYFCALLPFLPSDWGIP--VTDKLIFFC	KCTRVTVTEPRSQPHT	+ + +
δ1J2	AKYFCALG-EAPSAWKGH--LTAQLFFC	KGTQLIVEPRSQPHT	+ + +
δ1J3	AKYFCALG-EAPSAWKGKHSWDTRQMFFC	TGIKLFVEPRSQPHT	- - +
Site Directed Mutagenesis			
BE-13	AKYFCALG--TGVRGLQD---TDKLIFC	TCTRVTVTEPRSQPHT	- - +
δ1J3	AKYFCALG-EAPSAWKGKHSWDTRQMFFC	KGIKLFVEPRSQPHT	+ + +

FIG. 24

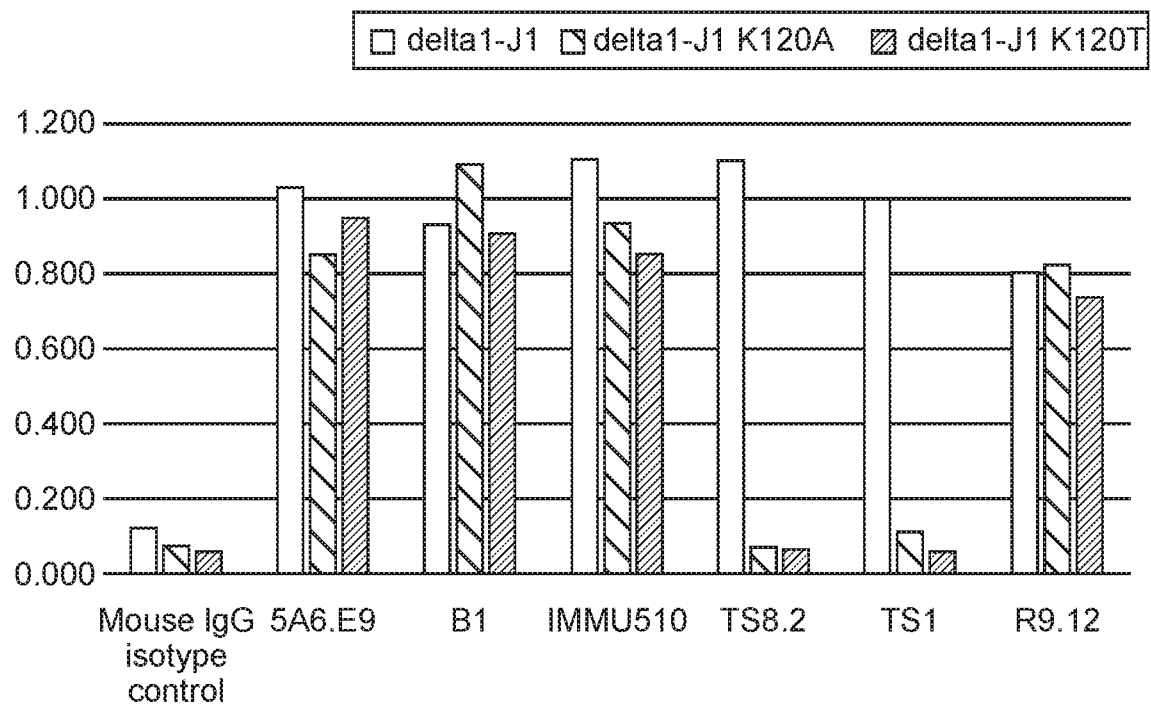


FIG. 25

Human Vδ1	AQKVTAQSSVMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIR
Human Vδ1 Mut1	AQKVTAQSSVMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIR
Human Vδ1 Mut2	AQKVTAQSSVMPVRKAVTLNCLYETSWWSYYIFWYKQLPS GQMTY LLIR
Human Vδ1 Mut3	AQKVTAQSSVMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIR
Human Vδ1 Mut4	AQKVTAQ QPDV SMVPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIR
Human Vδ1 Mut5	AQKVTAQSSVMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIR
Human Vδ1 Mut6	AQKVTAQSSVMPVRKAVTLNCLYETSWWSYYIFWYKQLPSKEMIFLIR
Human Vδ1	QGSDEQNAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGL
Human Vδ1 Mut1	Q YSD DGNARD GRYSVNF QKAQKSIN LTISALQLEDSAKYFCALGTGVRGL
Human Vδ1 Mut2	QGSDEQNAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGL
Human Vδ1 Mut3	QGSDEQNAKSGRYSVNFKKAAKSVALTISAL E LEDS AM YFCVLTGTGVRGL
Human Vδ1 Mut4	QGSDEQNAKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGL
Human Vδ1 Mut5	QGS D DGN AKSGRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGL
Human Vδ1 Mut6	QGSDEQ NARD GRYSVNFKKAAKSVALTISALQLEDSAKYFCALGTGVRGL

FIG. 26

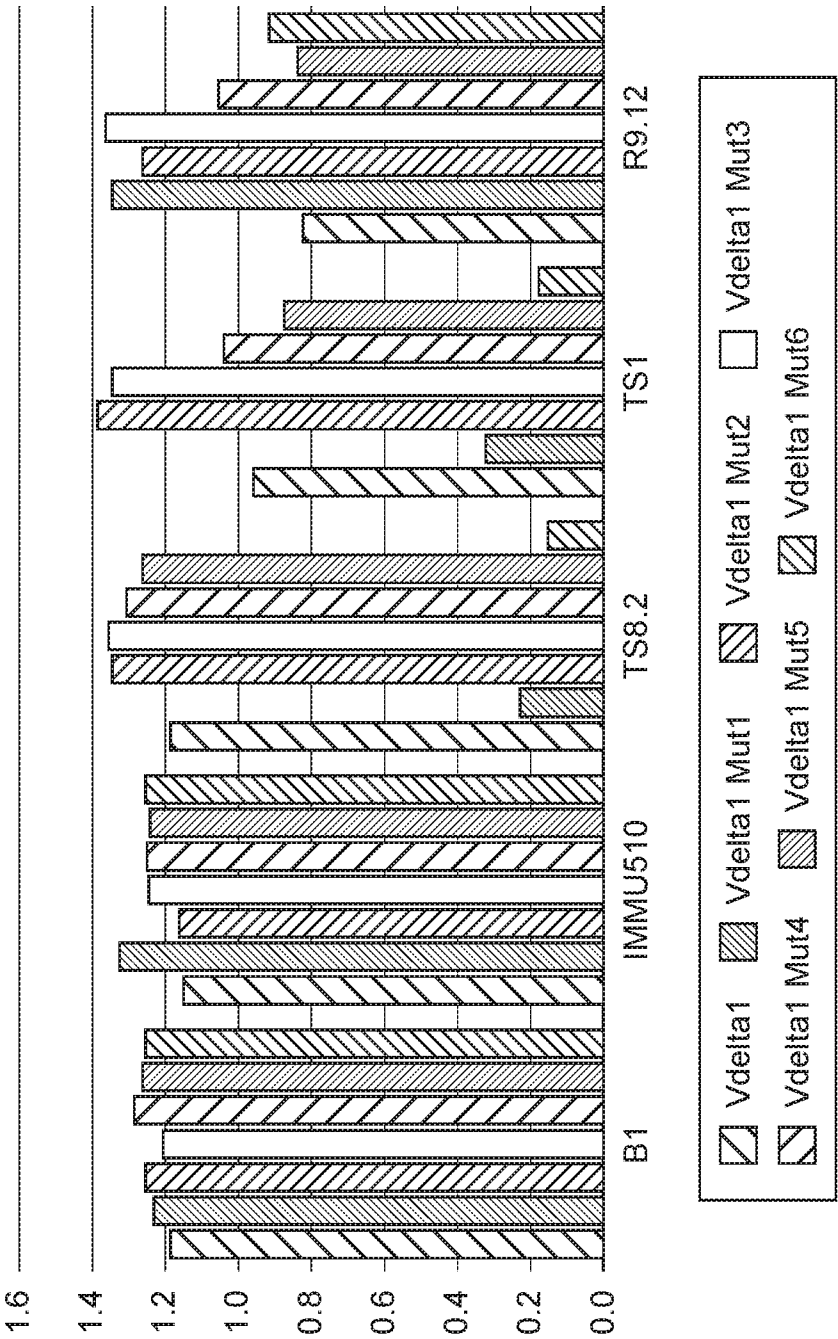


FIG. 27

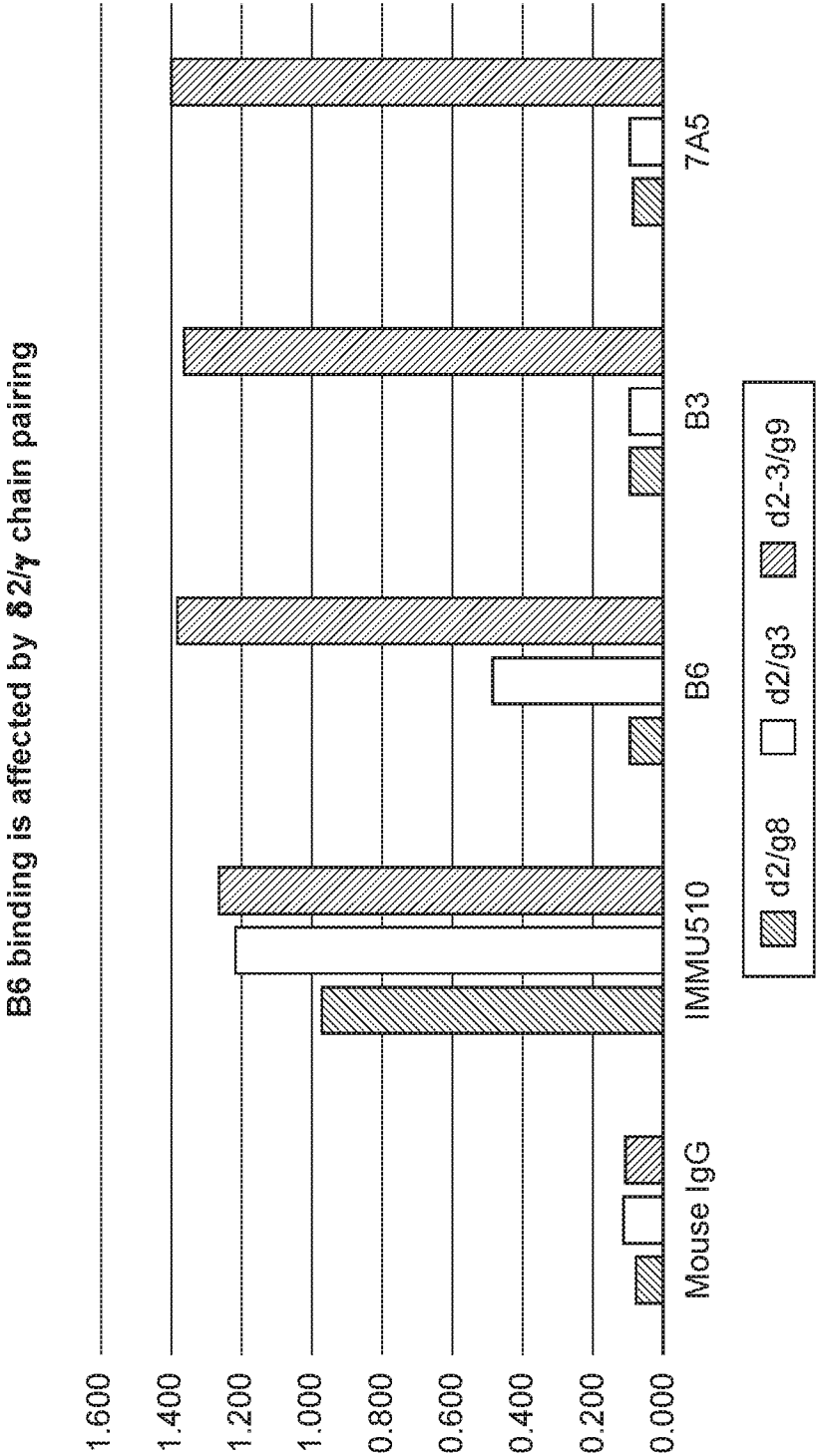


FIG. 28

Human TRDV2	v82CDR1									
Macaca mulatta	A	I	E	L	V	P	E	H	Q	T
	V	S	I	G	V	P	A	T	I	R
	C	S	M	K	G	E	A	I	S	N
	Y	I	N	W	Y	R	K	T	Q	G
	N	T	I	T	E	I	Y	R		
	A	V	E	L	V	P	E	H	Q	T
	V	S	I	G	V	P	A	T	I	R
	C	S	M	K	G	E	A	I	S	N
	Y	I	N	W	Y	R	K	T	Q	G
	N	T	M	T	E	I	Y	R		
	v82CDR2									
Human TRDV2	E	K	D	I	Y	G	P	G	F	K
Macaca mulatta	E	K	G	I	Y	G	P	G	F	K
	D	N	F	Q	G	D	I	D	I	A
	D	I	A	K	N	L	A	V	L	K
	I	A	V	L	K	I	L	A	P	S
	E	R	D	E	G	S	Y	Y	C	A
	D	I	T	E	E	N	Q	A	V	L
	A	V	L	K	I	L	A	P	S	E
	R	D	E	G	S	Y	Y	C	A	S
	D	R								
	v82CDR3									
Human TRDV2										
Macaca mulatta										

FIG. 29

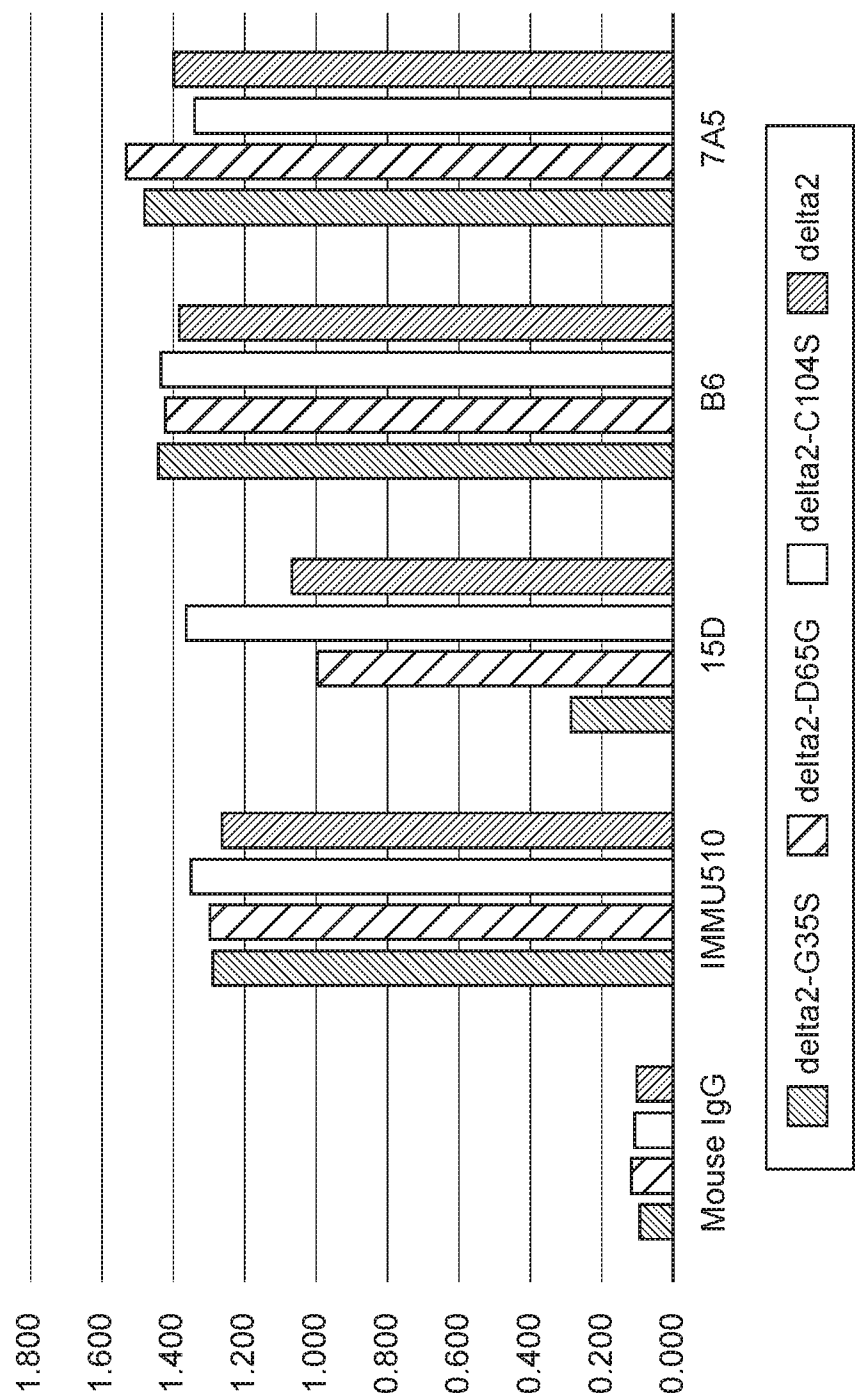


FIG. 30

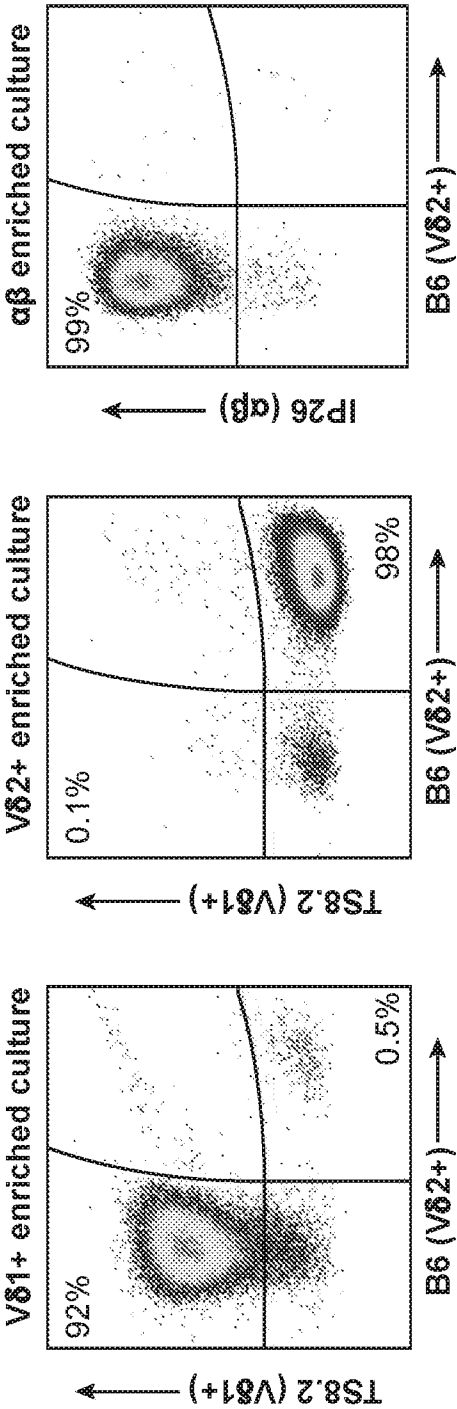


FIG. 31

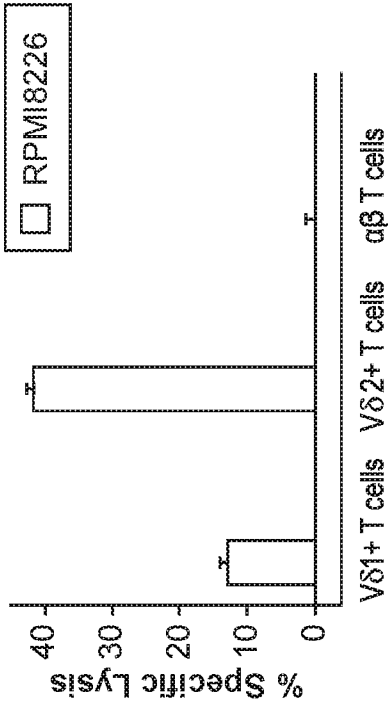


FIG. 32B

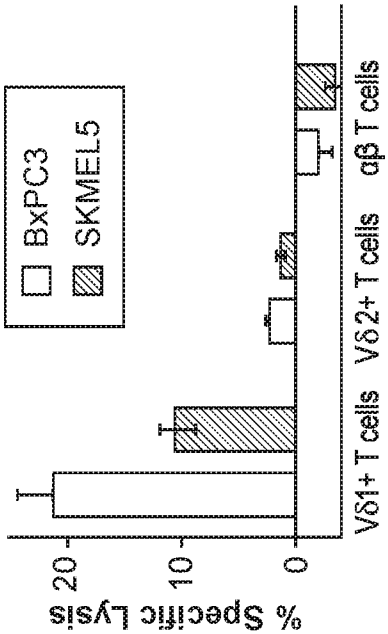


FIG. 32A

34 / 55

mAb	FW1	CDRH1	FW2	CDRH2
δ1-05	DVQLQESGPGMVKPSQSLSLTCTVTGYSIT	GGYDWH	WIRHFPGNKLEWMA	YISYSGSTDYNPSLKS
δ1-08	QVQLQQSGAELVRPGASVTLSCKASGYTFT	DYEVY	WVKQTPVHGLEWIG	AIDPETGRTAYNQKFKG
δ1-18	EVQLQQSGPELVKPGDSVKMSCKASGYTFT	DYYMD	WVKQSHGRSLEWIG	YIYPKNVGISYNQKFKG
δ1-22	QVQLQQSGPQLVKPGASVKLSCKASGYTFT	SYDIN	WVKQRPQGGLLEWIG	WIYPGDGTTDYNQKFKG
δ1-26	SDVQLQESGPGLVKPSQSLSVTCTVTGYSIT	SGYHWN	WIRQFPGNRLEWVG	YIHNSGSTNYNSFLKS
δ1-35	EVQLQQSGTIVLARPSSVKMSCKASGYTFT	TYWMH	WVKQRPQGGLDWIG	AIYPGNSDTNYNQKFRG
δ1-37	QVQLQQPGAELVRPGASVKLSCKAPGYTFT	SYWMN	WVKQRPEQGGLLEWIG	KIDPYDSETHYNQKFKD
δ1-39	QVQLQQPGADLVRPGTSVKLSCKASGYTFT	SYWMH	WVQRPQGGLLEWIG	VIDPDSYTNYNQKFKG
δ1-113	EVKLEESGGGLVQPGGSMKLSCAASGFTFS	DAWMD	WVRQSPEKGLEWVA	EIRAEANNHATYYAESVKG
δ1-143	QVQLQQPGAELVRPGASVKLSCKASGYAFT	DYWMN	WVKQRPQGGLLEWIG	TIDPDSYASYNQKFKG
δ1-149	QVQLQQSGAELVRPGASVKLSCKASDYKFT	DSEMY	WVKQTPVHGLEWIG	AIDPETGITAYNQKFKG
δ1-155	EVQLQQSGAELGRPGASVKLSCTTSGFNK	DDYMH	WVKQRPEQGGLLEWIG	WIDPENGDYASKFQG
δ1-182	QVQLQQSGAELVRPGASVTLSCKASGYKFI	DYEMH	WVKQTPVHGLEWVG	DLDPGTGVTAYNQKFKG
δ1-183	EVQLQMSGAELVRPGASVKLSCTASGFNIK	DDYMY	WVKQRPEQGGLLEWIG	WIDPENGDTEYASKFQG
δ1-191	EVQLQQSGAELVRPGASVKLSCTASDENIK	DDYMH	WVKQRPEQGGLLEWIG	WIDPENGETEYASKFQG
δ1-192	QIQLVQSGPELKKPGETVKISCKVSGDTFT	TYGMS	WVKQAPGKGLKWMG	WINTYSGVPTIADDFKG
δ1-195	EVKFEESGGGLVQPGGSMKLSCAASGFTFS	DAWMD	WVRQSPEKGLEWVA	EIRAEANNHATYYAESVKG
δ1-197	QVQLQQSGAELVRPGASVTLSCKASGYTFV	DYEMH	WVKQTPVHGLEWIG	AIDPETGITAYNQKFKG
δ1-199	EVQLQQSGAELVRPGASVKLSCTASGFNIK	DDYMS	WVKQRPEQGGLLEWIG	WIDPENGDTEYASKFQG
δ1-201	QVQLQQSGADLVRPGASVTLSCKASGYTFT	DYEMH	WVKQTPVHGLEWIG	AIDPETGITAYNQKFKG
δ1-203	SDVQLQESGPGLVKPSQSLSITCSVTGYSIT	SGYYWN	WIRQFPGNLEWVG	YISHDGSNNYNPALKN
δ1-239	QVQLQQSGAELVRPGASVKLSCKASDYKFT	DSEMY	WVKQTPVHGLEWIG	AIDPETGITAYNQKFKG
δ1-253	EVQLQQSGPELVKPGASVKMSCKASGYTFT	DYYMN	WVKQSHGKSLEWIG	HINPYNGGTSYNQKFKG
δ1-257	QVQLQQSGAELVRPGASVTLSCKASGYRFP	DYEMH	WVKQTPVHGLEWIG	AIDPETGRTAYNQKFRG
δ1-278	SDVQLQESGPGLVKPSQSLSITCSVTGYSIT	SDYYWN	WIRQFPGNKLEWVG	YITYDGSNNYNPSLKN
δ1-282	EVKLVESGGGLVQPGGSLKLSCATSGFTFS	DSYMY	WVRQTPEKRLWVA	YISYGGVNTYYPDTVRG
δ1-285	EVQLVESGGGLVKPGGSLKLSCAASGFTFS	DYGMH	WVRQAPEKGLEWVA	YISSGSRTIYYADTVKG

FIG. 33

35 / 55

FW3	CDRH3	FW4
RISVTHDTSKNLFFLNLTSVTTEDTATYYCAR	EGGRGFAY	WGQGTLLTVSA
KAILTTDKSSSTAYMALRSLTSEDSAVYYCAR	LKSGRIYGDLFAY	WGQGTLLTVSA
KATLTVDKSSSTAYMELHSLTSEDSAVYYCAR	SLLWDALDY	WGQGTSLTVSS
KATLTVDTSSSSAYMELHSLTSEDSAVYFCAR	MDDYDDGGAMDY	WGQGTSLTVSS
RISITRDTSKNQFFLQLNSVTTEDTATYYCVA	YYSNSREFWYAY	WGQGTLLTVSA
KAKLTAVTSASTAYMELSSLTNEEDSAVYYCTY	GYVVDYYAMDY	WGQGTSLTVSS
KAILTVDKSSSTAYMQLSSLTSEDSAVYYCAR	GGDNYDPFAY	WGQGTLLTVSA
KATLTVDTSSSTAYMQLSSLTSEDSAVYYCAR	SDDYDEGYFFDQ	WGQGTLLTVSA
RFTISRDDSKSRVFLQMNSLRAEDTGIYYCTG	LDYGSIGFAY	WGQGTLLTVSA
KATLTVDTSSNSAYMHLSSLTSEDSAVYFCAR	ESNDVCWYFDV	WGAGTTLTVSS
KATLTSDKSSSTAYMELRSLTSEDSAVYYCTR	AVPPWFAY	WGQGTLLTVSA
KATITADTSSNTAYLQLSSLTSEDTAVYYCNY	YGFDY	WGQGTTLTVSS
KAILTADKSSSTAYMELRSLTSEDSAVYYCTV	WSADF	WGQGTSLTVSS
KATITADTSSNTAYLQLSSLTSEDTAVYYCTY	YAMDY	WGQGTSLTVSS
KATLTADTSSNTAYLQLSSLTSEDTAVYYCTE	LGFDY	WGQGTTLTVSS
REAFSLETSASTAYLQINNKLKNETATYFCAR	SSYDYDDAMDY	WGQGTSLTVSS
RFTISRDDSKSRVFLQMNSLRAEDTGIYYCTG	LDYGSVGFAY	WGQGTLLTVSA
KATLTADKSSSTAYMELSSLTSEDSAVYYCIR	PRGGSHEFDY	WGQGTTLTVSS
KATITADTSSNTAYLRLSSLTSEDTAVYYCTE	LGFDY	WGQGTTLTVSS
KATLTADKSSSTAYMELSSLTSEDSAVYYCTR	PRGGSHEFDH	WGQGTPLTVSS
RISITRDTSKNQFFLKLNSVTTEDTGTYCAS	VYYGDYEVWYTY	WGQGTLLTVSA
KATLTSDKSSSTAYMELRSLTSEDSAVYYCTR	AVPPWFAY	WGQGTLLTVSA
KATLTVDKSSSTAYMQLNSLTSEDSAVYYCAR	NHIYYYDGGYFYYAMDY	WGQGTSLTVSS
KAKLTADKSSSTVYMELRSLTSEDSAVYYCTR	GYGIQFPY	WGQGTLLTVSA
RISITRDTSKNQFFLKLNSVTTEDTATYYCAR	DDGYFDY	WGQGTTLTVSS
RFTISRDNASTLYLQMSRLKSEDTAMYYCAS	RGYY	WGQGTSLTVSS
RFTISRDNANTLFLQMTSLRSEDTAMYYCAR	EGAYSSFYD	WGQGTTLTVSS

FIG. 33 (Cont.)

36 / 55

mAb	FW1	CDRL1	FW2
δ1-05	NIVLTQSPASLAVSLGQRATISC	RASESVDGYGNSFMH	WYQQKPGQPPKLLIY
δ1-08	QIVLTQSPALMSASPGEKVTMTC	SASSSVSYMY	WYQQKPRSSPKPWIY
δ1-18	DIQMTQSPASLSVSVGETVTITC	RASENIYSNLA	WYQQKQGKSPQLLVY
δ1-22	DVVMQTPTPLTSLVTVGQPASISC	KSSQSLLHSNGKTYLN	WLLQRPQGSPKLLIY
δ1-26	DIVMSQSPSSLAVSVGEKVTMSC	KSSQSLLYSNQKNSLA	WYQQKPGQSPKLLIY
δ1-35	DIVMTQSHKFMSTSVGDRVSITC	KASQDVSIDVA	WYQQKPGQSPKLLIY
δ1-37	DIQMTQSPASLSVSVGETVTITC	RASENIYSNLA	WYQQKQGKSPQLLVY
δ1-39	DVVMQTPTPLTSLVTVGQPASISC	KSSQSLLHSNGKTYLN	WLLQRPQGSPKLLIY
δ1-113	DVLMTQTPLSLFVSLGDQASISC	RSSQSVVHRNGNTFLE	WYLQKPGQSPKLLIY
δ1-143	DIQMTQTSSLSASLGDRVTISC	RASQDISNYLN	WYQQKPDGTVKLLIY
δ1-149	QIVLTQSPALMSASPGEKVTMTC	SASSSVSYMY	WYQQKPRSSPKPWIY
δ1-155	DVLMTQTPLSLFVSLGDQASISC	RSSQSIVHSDGNTFLO	WYLQKPGQSPKLLIY
δ1-182	SDVVRPTPLSLFVSLGDQASISC	RSSQSLVHSDGNTYLH	WYLQKPGQSPKLLIY
δ1-183	DVVMQTIPVSLPVTSLGDQASISC	RSSQSLLHSDGNTYLH	WYLQKPGQSPKLLIY
δ1-191	AVVMTQTPLSLFVSLGDQASISC	RSSQSLVHSDGNTYLH	WYLQKPGQSPKLLIY
δ1-192	DVQITQSPSYLAASPGETITINC	RTSKSISKYLA	WYQEKPGKTNKLLIY
δ1-195	DVIMTQTPLSLFVSLGDQASISC	RSSQSVVHRNGNTFLE	WYLQKPGQSPKLLIY
δ1-197	QIVLSQSPAILSASPGEKVTMTC	RASSSVNYMH	WYQQKPGSSPKPWIY
δ1-199	AVVMTQTPLSLFVSLGDQASISC	RSSQSLVHSDGNTYLH	WYLQKPGQSPKLLIY
δ1-201	QIVLSQSPAILSASPGEKVTMTC	RASSSVRYIH	WYQQKPGSSPKPWIY
δ1-203	DIVMSQSPSSLAVSVGEKVALNC	KSSQSLLYSINQKNYLA	WYQRNPGQSPKLLIY
δ1-239	QIVLTQSPALMSASPGEKVTMTC	SASSSVSYMY	WYQQKPRSSPKPWIY
δ1-253	NIVLTQSPASLAVSLGQRATISC	RPSESVDSYGNSFMH	WYQQKPGQPPKLLIY
δ1-257	QIVLSQSPAILSASPGEKVTMTC	RASSSVSYIH	WFQQKPGSSPKPWIY
δ1-278	QIVLSQSPAILSASPGEKVTMTC	RASSSVNYMH	WHQQKPGSSPKPRIY
δ1-282	DIVLTQSPSSLAVSLGQRATISC	KASQSVDDYDGDSYMN	WYQQKPGQPPKLLIY
δ1-285	DIVLTQSPASLAVSLGQRATISC	KASQSVDDYDGDSYMN	WYQQKPGQPPKLLIY

FIG. 34

37 / 55

CDRL2	FW3	CDRL3	FW4
LASNLES	GVPARFSGSGSRTDFTLTIDPVESDDAATYYC	QQNNEDPFT	FGSGTKLEIK
FTSNLAS	GVPARFSGSGSGTSYSLTISSMEAEDAATYYC	QQWSSNPLT	FGAGTKLELK
AATYLAD	GVPSRFSGSGSGTQYSLKINSLOSEDFGSYYC	QHFWDGIPYT	FGGGTKLEIK
LVSKLES	GVPDRFSGSGSGTDFTLKISRVEAEDLGVYYC	LQATHEPLT	CGAGTKLELK
WASTRES	GVPDRFTGSGSGTDFTLTISSVKAEDLAVYYC	QQDYSYPL	TFGGGTKLELK
SASYRYT	GVPDRFTGSGSGTDFTFTISNVOAEDLAVYYC	QQHYSIPCT	FGSGTKLEIK
GARNLAD	GVPSRFSGSGSGTQYSLKINSLOSEDFGSYFC	QHFWDTTFT	FGSGTKVEIK
LVSKVES	GVPDRFSGSGSGTDFTLKISRVEAEDLGLYYC	LQVTHEPLT	FGAGTKLELK
KVSNRFS	GVPDRFSGSGSGTDFTLKISRVEAEDLGLYYC	FOGSHVPYT	FGGGTKLEIKR
YTSRLHS	GVPSRFSGSGSGTDYSLTISNLEQEDIATYFC	QQGNTLRT	FGGGTKLEIK
LTSNLAA	GVPARFSGSGSGTSYSLTISSMEAEDAATYYC	QQWSGDPPT	FGGGTKLEIKR
KVSNRFS	GVPDRFSGSGSGTDFTLKISRVEAEDLGVYYC	FOGSHVPYT	FGGGTKLEIKR
KVSNRFS	GVPDRFSGSGSGTDFTLKISRVEAEDLGVYFC	SGSTHVPPWT	FGGGTKLEIK
KVSNRFS	GVPDRFSGSGSGTDFTLKISRVEAEDLGVYFC	SGFTHVPPWT	FGGGTKLEIK
KVSNRFS	GVPDRFSGSGSGTDFTLKISRVEAEDLGVYFC	SHSTHVPHYT	FGGGTKLEIKR
SGSTLQS	GIPSRFSGSGSGTDFTLTISLEPEDFAMYYC	QHHNEYPHYT	FGGCTRLEIKR
KVSNRFS	GVPDRFSGSGSGTDFTLKISRVEAEDLGVYYC	FOGSHVPYT	FGGGTKLEIKR
ATSNLAS	GVPARFSGSGSGTSYSLTISRVEAEDAATYYC	QQWSSHQPT	FGAGTKLELK
KVSNRFS	GVPDRFSGSGSGTDFTLKISRVEAEDLGVYFC	SHSTHVPHYT	FGGGTKLEIKR
ATSNLAS	GVPARFSGSGSGTSYSLTISRVEAEDAATYHC	QQWYSDTPT	FGAGTKLELK
WASTRES	GVPDRFTGSGSGTDFTLTISSVKTEDLAVYYC	QQYYSYPWT	FGGGTKLEIK
LTSNLAA	GVPARFSGSGSGTSYSLTISSMEAEDAATYYC	QQWSGDPPT	FGGGTKLEIKR
LASNLES	GVPARFSGSGSRTDLTLTIDPVEADDAATYYC	QQNNEDPWT	FGGGTK
GTSNLAS	GVPARFTGSGSGTSYSLTISRVEAEDAATYYC	QQWSSDSPPT	FGGGTKLEIK
GTSNLAS	GVPARFSGSGSGTSYSLTISRVEAEDAATYYC	QQWSSNPPT	FGAGTKLELK
GASNLES	GIPARFSGSGSGTDFTLNHPVEEEDAATYYC	QQSNEDPWT	FGGGTKLEIK
AASNLES	GIPARFSGSGSGTDFTLNHPVEEEDAATYYC	QQSNEDPWT	FGGGTKLEIK

FIG. 34 (Cont.)

38 / 55

mAb	FW1	CDRH1	FW2	CDRH2
δ2-14	EVQLQQSGPELVKPGASVKISCKASGYSFT	GYMN	WVKQSPEKSLEWIG	EINPSTGGTTYNQKFOA
δ2-17	SDVQLQESGPGLVTPSQSLSVTCTVTGYSIT	SGSYWN	WIRQFPGNKLEWMG	YIHNSGSTTYNPSLKS
δ2-30	QVQLQQSGAELVKPGASVKLSCKTSGYTFT	SYWIQ	WVKQRPGQGLGWIG	EIFPGTGTTYNEKFKG
δ2-31	QVQLLPQGAELVKPGASVKLSCKASGYSFT	NFWIN	WVKLRPGQGLEWIG	NIFPGSSSPNYNEKFKS
δ2-32	EVKLVESGGGLVQPGGSLSLSCAASGFTFT	DYYMS	WVRQPPGKALEWLG	FIRNKANGYTTESSASVKG
δ2-33	EVQLQQSGPELVKPGASVKISCKASGYSFT	GYMN	WVKQSPEKSLEWIG	EINPSTGGTTYNQKFOA
δ2-35	DVKLVESGGGLVQPGGSLKLSCAASGFTFS	SYYMS	WVRQTPEKRLEWVA	TISNSGGSTYYPDVSKG
δ2-36	QVQLLPQGAELVKPGASVKLSCKASGYTFT	NFWIN	WVKQRPGQGLEWIG	NIYPGSSSPNYNEKFKF
δ2-37	QVQLQQPGAELVKPGASVKLSCKASGYTFT	NHWIS	WVKQRPGQGLEWIG	NIFPGSSSPNYNEKFKS

FIG. 35

39 / 55



FW3	CDRH3	FW4
KATLTVDKSSSTAYMQLKSLTSEDSAVYYCSR	GEDDGYFPYSMDF	WGQGASVTVSS
RISITRDTSKNQFFLQLNSVTTEDTATYYCAR	STGPPPTY	WGQGLVTVSA
KATLTRDTSSSTAYMQLSSLTSEDSAVYFCAR	RGVFGNYAMDY	WGQGTSTVTVSS
KATLTVDISSTAYMQLSSLTSDGSAVYYCAR	YGSYGNWYFDV	WGAGTTVTVSS
RFTISRDNQSILYLQMNVLRAEDSATYYCAS	SKPGWPMDY	WGQGTSTVTVSS
KATLTVDKSSSTAYMQLKSLTSEDSAVYYCSR	GEDDGYFPYSMDF	WGQGASVTVSS
RFTISRDNAKNTLYLQISSLNSEDTAVYYCSR	EYDFDGEFFDY	WGQGTTLTVSS
KATLTVDISSTAYIQSSLPSDDSAVYYCAR	YGTFGNWYFDV	WGAGTTVTVSS
KATLTVDTSSSTAYMQLSSLTSDASAVYYCTR	WGNYGYYYAMDY	WGQGTSTVTVSS

FIG. 35 (Cont.)

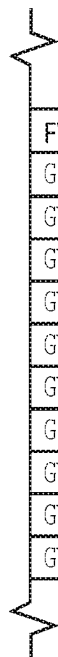
40 / 55



mAb	FW1	CDRL1	FW2	CDRL2
δ2-14	DIVLIQSPATLSVTPGDSVSLSC	RASQSINNNLH	WYQQKSHESPRLLIK	YVSQSIS
δ2-17	NIVLTQSPGSLAVSLGQRATISC	RASESDNYGNSEFMH	WYQQKPGQPPKLLIY	LASNLES
δ2-22	DIQMTQSPASLSVSVGETVTITC	RASENIYSNLA	WYQQKQKGKSPQLLVY	AATNLAG
δ2-30	DIQMTQSPASLSVSVGETVTITC	RASENIYSNLA	WYQQKQKGKSPQLLVY	AATNLAD
δ2-31	DLKMTQSPSSMYASLGERVTITC	KASQDINSFLS	WFQQIPGKSPKTLIY	RANRLVD
δ2-32	DIQMTQTSSFSVSLGDRVITIC	KASEDIYNRLA	WYQQKPGNAPRLIS	GATSLEA
δ2-33	DIVLIQSPATLSVTPGDSVSLSC	RASQSINNNLH	WYQQKSHESPRLLIK	YVSQSIS
δ2-35	DVVMQTPLSLPVSLGDQASISC	RSSQSLVHSNGNTYLH	WYLQKPGQSPKLLIY	KVSNRFS
δ2-36	DIKMTQSPSSMYASLGERVTITC	KASQDINNFLS	WFQQIPGKSPKTLIY	RANRLVVD
δ2-37	QIVLTQSPAISASLGEEITLTC	SASSRVNYMH	WYQQKSGTSPKLLIY	STSNLAS

FIG. 36

41 / 55



FW3	CDRL3	FW4
GIPSRFSGSGSGTDFTLSINSVETEDFGMYFC	QQNSNWPLT	FGAGTKLELK
GVPARFSGSGSRTDFTLTIDPVEADDAATYYC	QQNNEDPT	FGSGTKLEMK
GVPSRFSGSGSGTQYSLKINSLQSEDFGSYYC	QHFWGTPRT	FGGGTKLEIK
GVPSRFSGSGSGTQYSLKINSLQSEDFGSYYC	QHFGDTPYT	FGGGTKLEIKR
GVPSRFSGSGSGQDYSLTISSEYEDMGIYYC	LQSDEFPYT	IGGGTKLEIKR
GVPSRFSGSGSGNDYTLSTLSLQTEDVATYYC	QQYWYTPWT	FGGGTKLEIK
GIPSRFSGSGSGTDFTLSINSVETEDFGMYFC	QQNSNWPLT	FGAGTKLELK
GVPDFSGSGSGTDFTLKISRVEAEDLGVYFC	SQSRHVPYT	FGGGTKLEIKR
GVPSRFSGSGSGQDYSLTISSEYEDMGIYYC	LQSDEFPYT	IGGGTKLEIKR
GVPSRFSGSGSGTFYSLTISVEAEDAADYYC	HQWSSYPT	FGGGTKLEIK

FIG. 36 (Cont.)

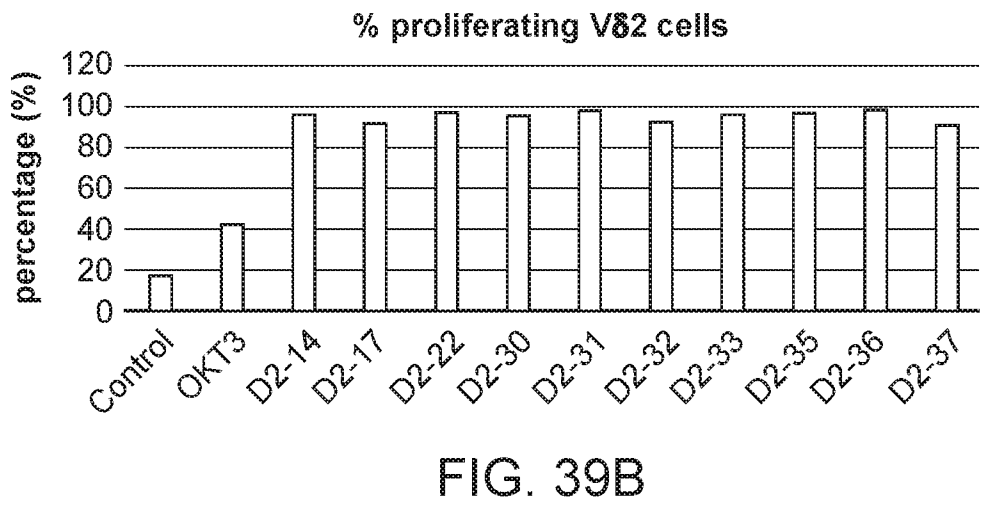
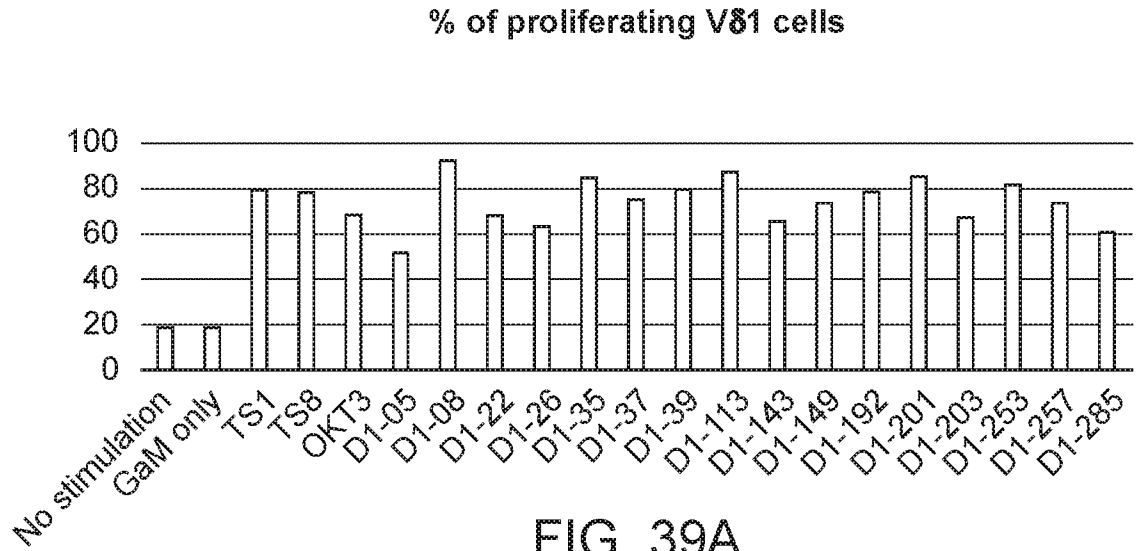
42 / 55

	$\delta 1\gamma 8$	$\delta 2\gamma 9$	$\delta 3\gamma 2$	$\delta 4\gamma 8$	$\delta 5\gamma 3$	$\delta 7\gamma 8$	$\delta 8\gamma 8$	cross reactivity
TS-1	1.754	0.080	0.061	0.064	0.059	0.067	0.066	$\delta 1, \delta 4$
	2.198	0.089	0.084	0.084	0.081	0.038	0.040	
d1-08	1.790	0.044	0.045	0.049	0.046	0.042	0.044	
	2.099	0.051	0.048	0.051	0.053	0.037	0.037	
d1-18	1.081	0.048	0.051	0.983	0.043	0.043	0.044	
	1.491	0.051	0.048	0.872	0.052	0.037	0.038	
d1-26	1.387	0.097	0.059	0.054	0.062	0.043	0.038	
	1.587	0.128	0.078	0.067	0.069	0.041	0.043	
d1-35	1.278	0.084	0.048	1.249	0.044	0.053	0.037	
	1.741	0.062	0.052	1.005	0.052	0.043	0.044	
d1-37	1.525	0.052	0.044	0.043	0.043	0.056	0.037	
	2.013	0.052	0.050	0.051	0.057	0.039	0.038	
d1-39	1.167	0.072	0.734	0.824	0.646	0.076	0.067	
	1.282	0.080	0.758	0.694	0.469	0.044	0.037	
B1	2.263	1.307	1.711	1.582	1.381	0.922	1.828	
$\gamma\delta$ pan	2.117	1.668	1.669	1.722	1.176	0.644	1.604	
d1-18 and d1-35 were found to cross reacts with $\delta 4$								
d1-39 cross reacts with $\delta 3, \delta 4$ and $\delta 5$ subsets								

FIG. 37

		CDR1	
Human Vd1	AQKVTQAQSSVSM	PVRKAVTLNCLYE	TSWWSYIFWYKQLPSKEMIFLIR
Dolphin Vd1	-----V-RAM-SQLGE	-----S-Q-----LSW-D-----G-T-----H	
	CDR2		
Human Vd1	QGS	DEQNAKSGRYSVNF	KKAAKSVALTISALQLEDSAKYFCALGE-----
Dolphin Vd1	-I-SD-----N-----	QERH-FIS-----LV-----N-----	TGVRGL

FIG. 38



45 / 55

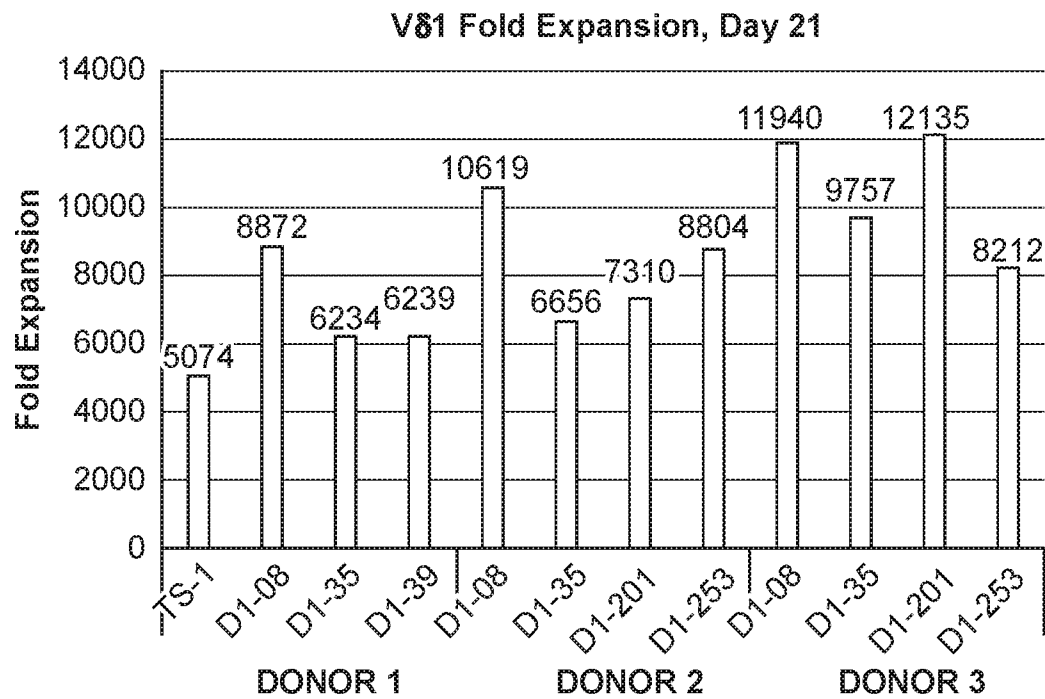


FIG. 40A

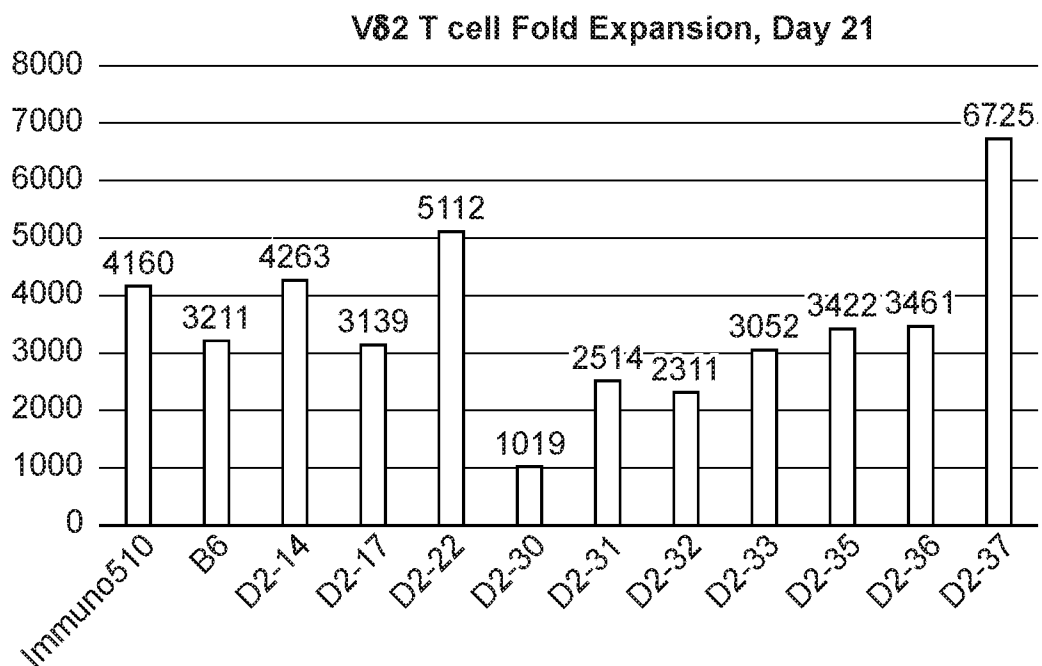


FIG. 40B

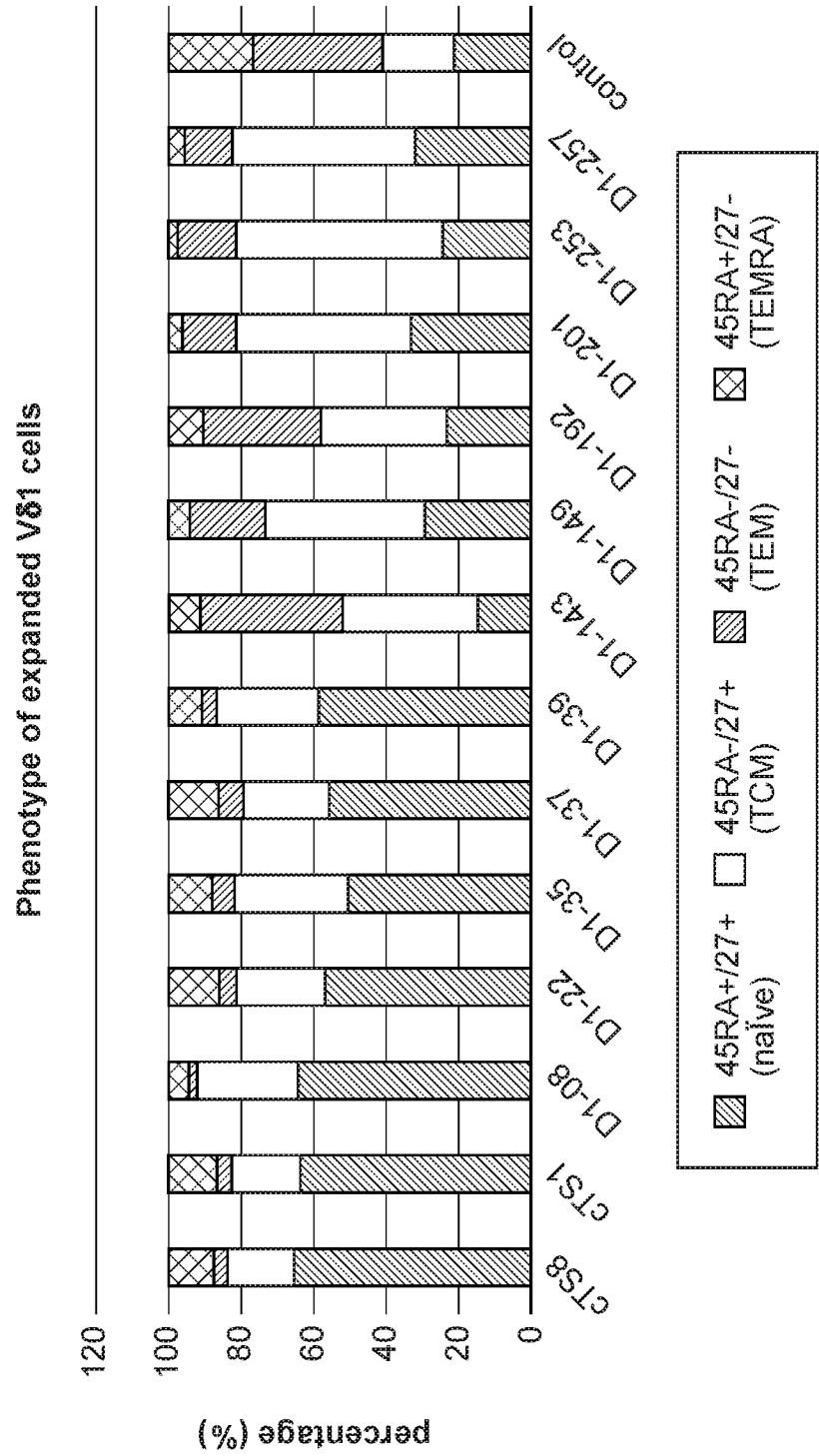


FIG. 41

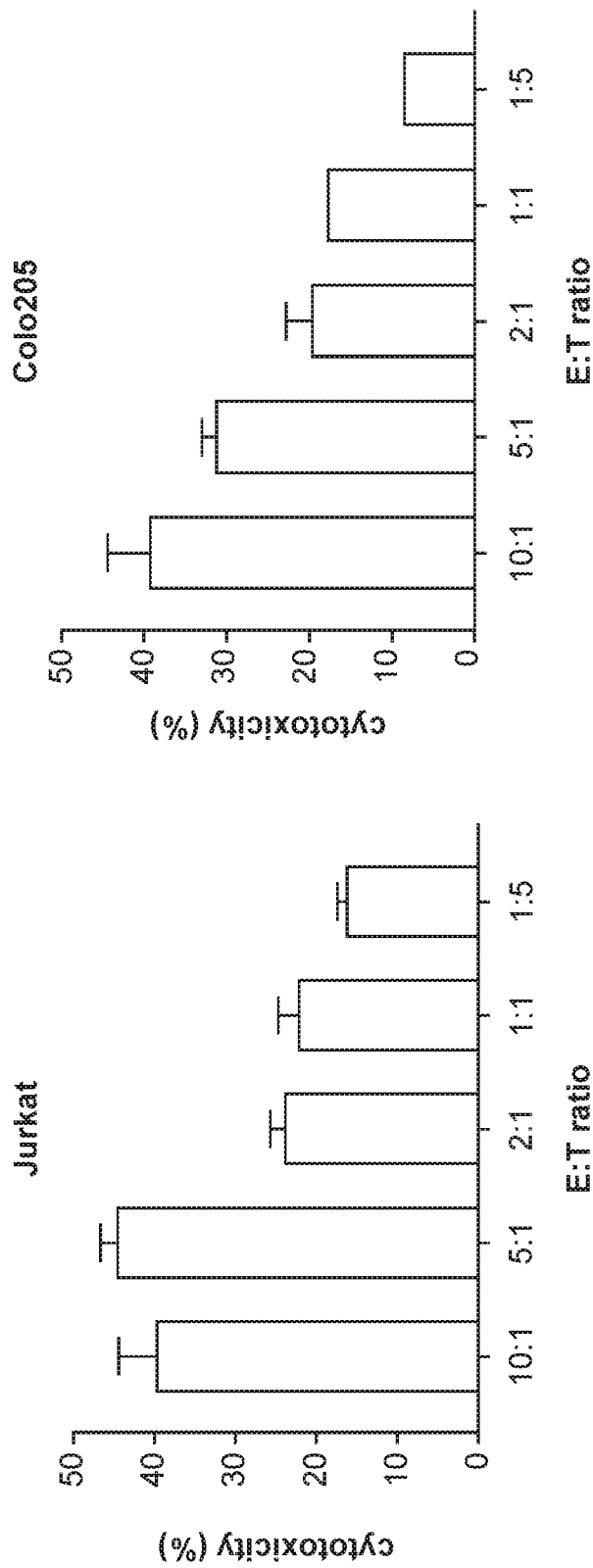


FIG. 42

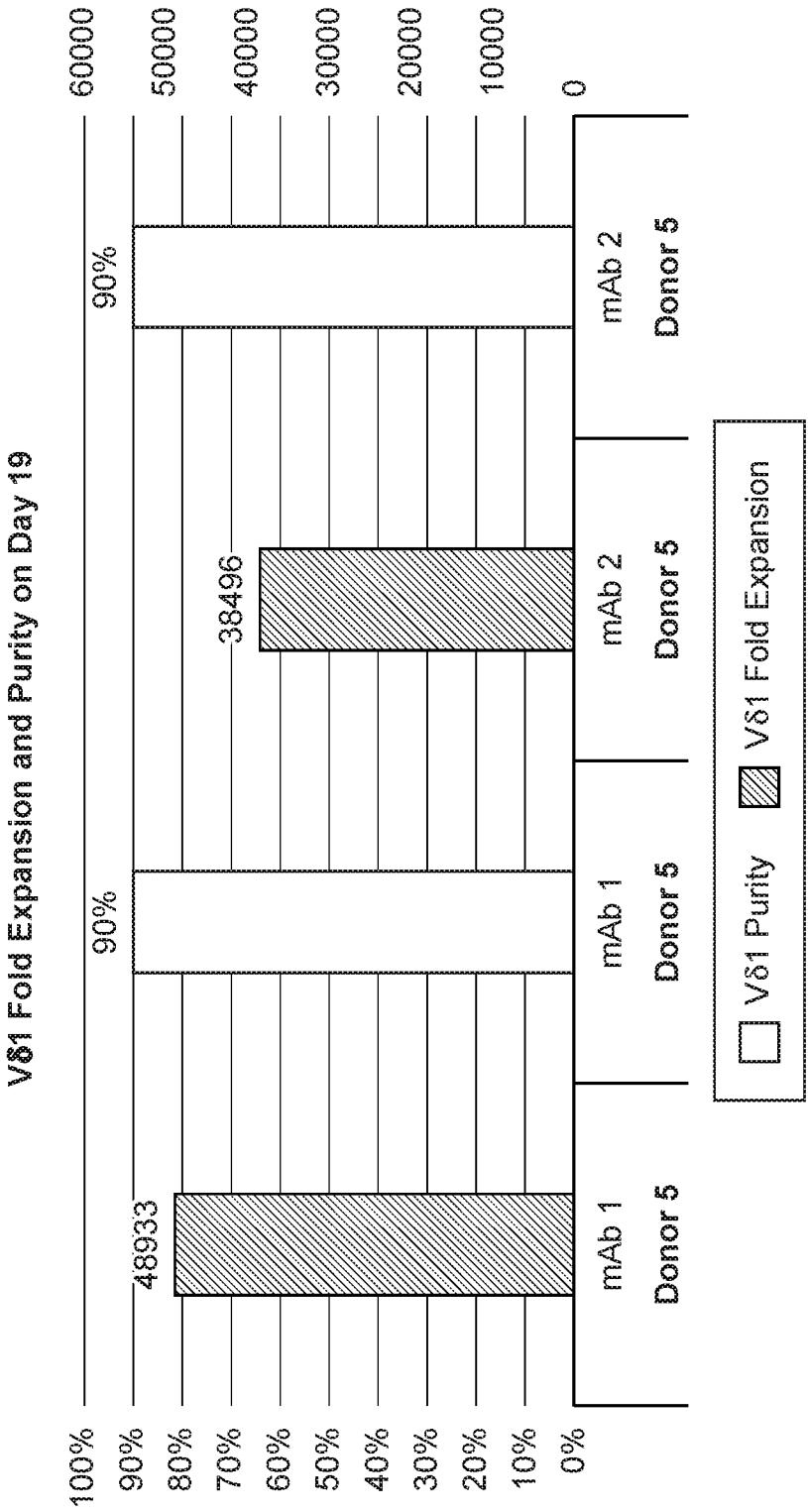


FIG. 43

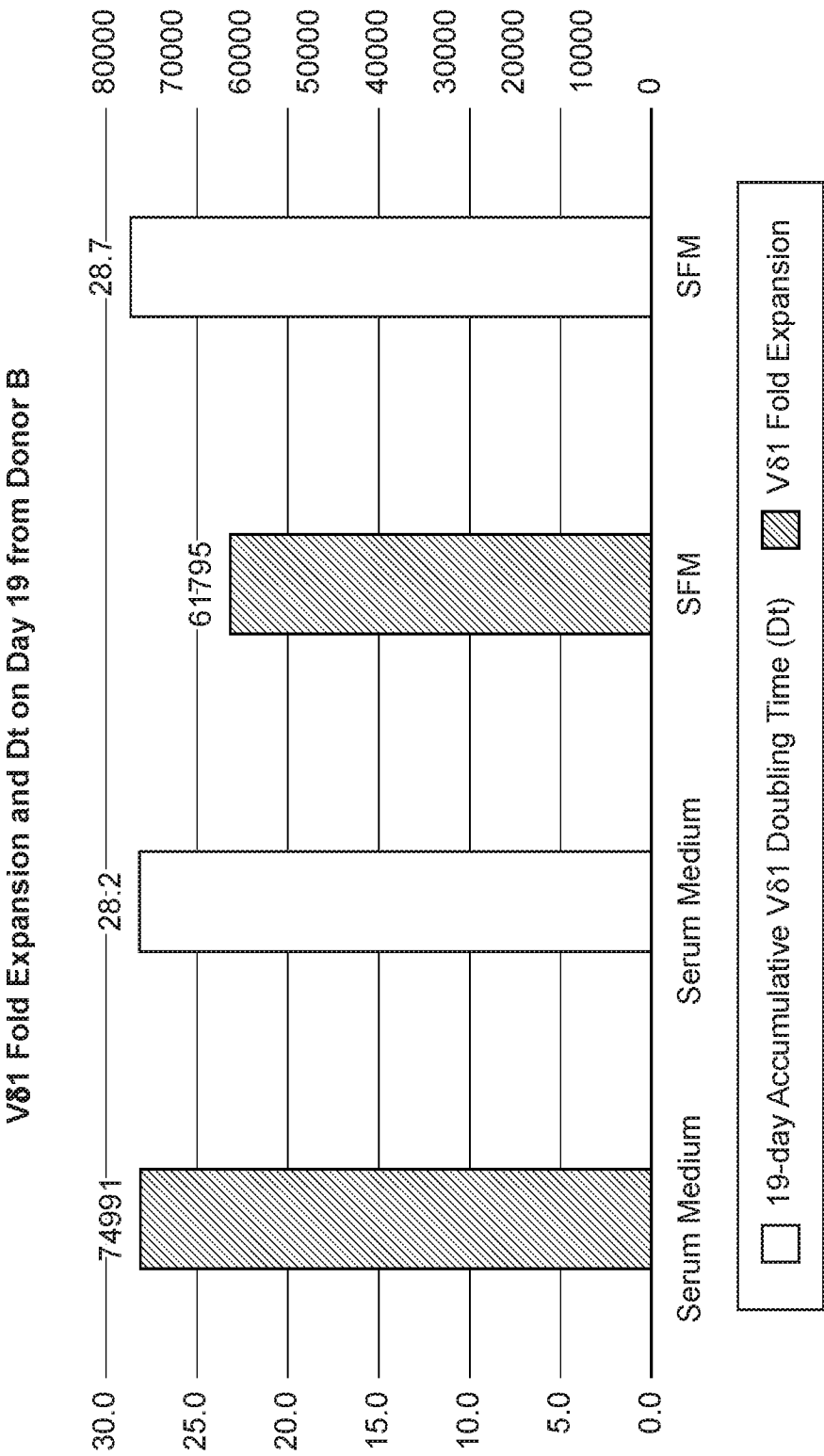


FIG. 44

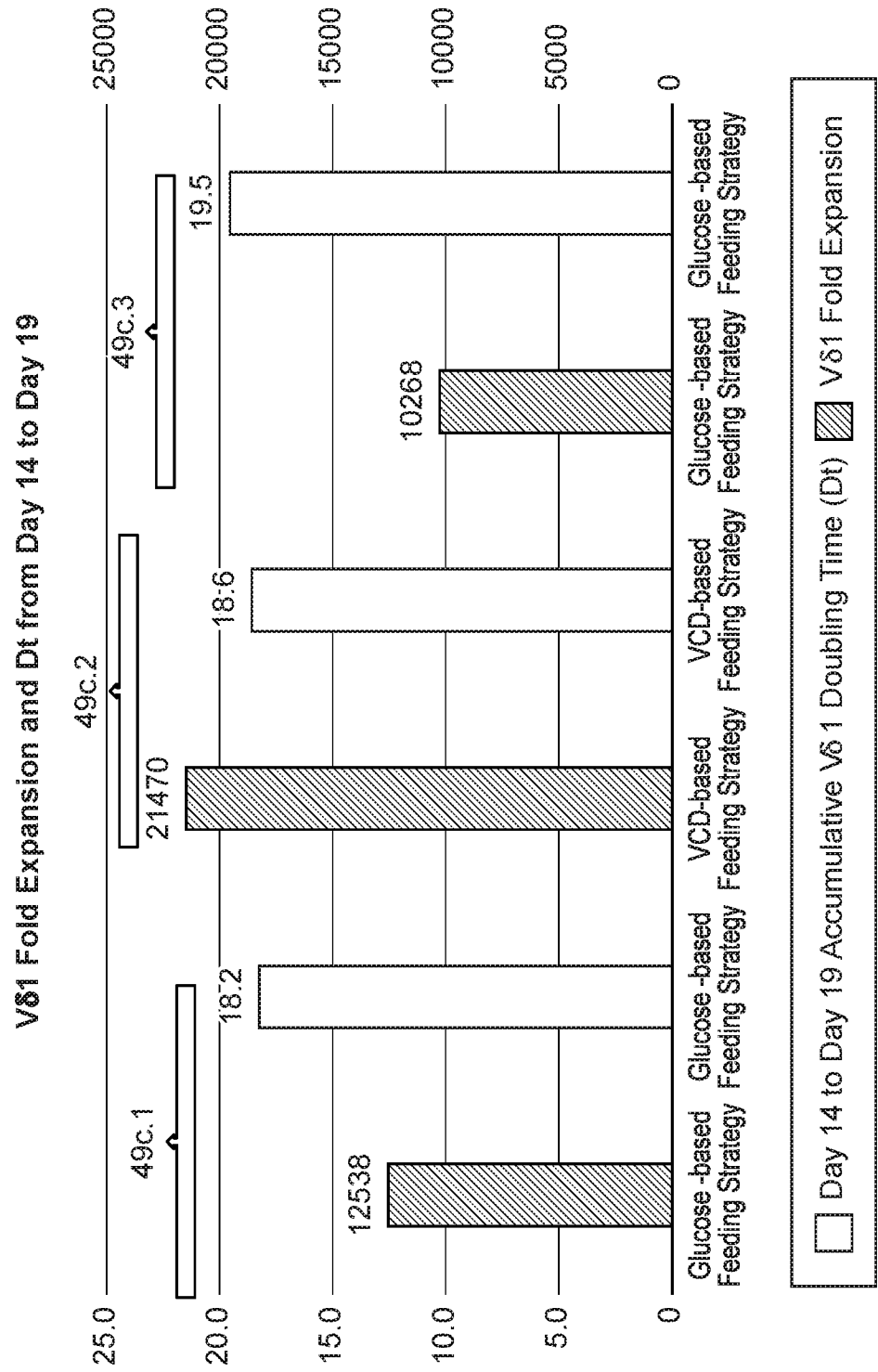


FIG. 45

51 / 55

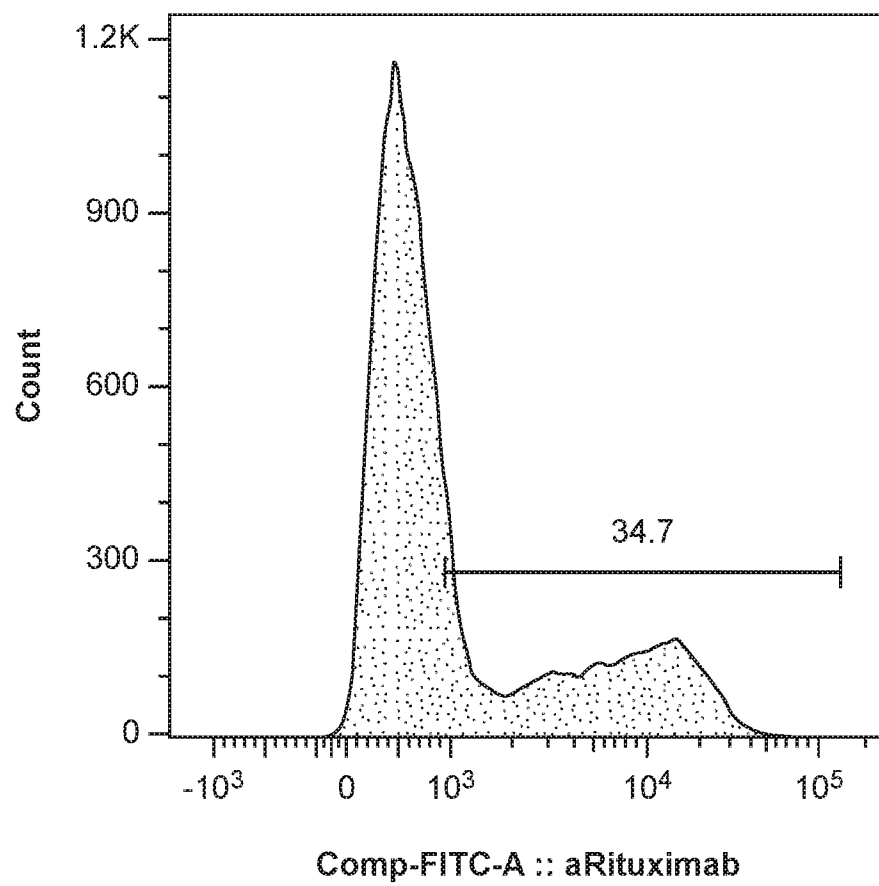


FIG. 46

52 / 55

	$\delta 1y8$	$\delta 2y9$	$\delta 3y2$	d1 Dolphin	$\delta 1-11$	$\delta 1-21$	$\delta 1-28$	$\delta 1-47$	$\delta 1-70$	$\delta 1-80$	$\delta 1-95$	J1	J2	J3	K120A	BIN
TS-1	1.46	0.07	0.07	0.09	1.27	2.17	1.88	1.77	0.54	0.64	0.50	1.14	0.77	0.05	0.07	47-70 J1, J2, K120 1
	2.27	0.09	0.09	0.14	1.74	2.29	1.95	2.06	0.95	0.91	0.82	1.08	0.74	0.05	0.06	
TS8.2	1.36	0.05	0.05	0.05	0.91	1.09	0.23	0.24	0.06	0.05	0.09	0.93	0.76	0.04	0.15	21-28 J1, J2 6
	2.04	0.04	0.04	0.06	1.02	1.92	0.39	0.33	0.10	0.07	0.17	1.00	0.77	0.04	0.20	
R9.12	1.76	0.06	0.06	0.06	1.19	0.06	0.06	0.06	0.09	0.05	0.07	0.95	0.94	0.94	0.82	11 to 21 Arg16 2b
	1.95	0.05	0.05	0.07	1.68	0.06	0.06	0.06	0.11	0.06	0.10	1.11	0.92	0.99	1.11	
d1-08	1.70	0.05	0.06	0.08	1.34	1.68	1.77	1.90	0.92	0.49	0.12	0.99	0.94	0.94	0.93	80 to 95 3
	2.08	0.05	0.06	0.07	1.65	1.91	2.03	2.05	1.63	0.72	0.15	1.20	1.18	0.95	1.23	
d1-35	1.49	0.07	0.06	0.09	0.06	0.06	0.07	0.06	0.07	0.07	0.11	0.86	0.96	0.04	0.49	1 to 11 J1, J2 4
	1.87	0.09	0.07	0.12	0.07	0.07	0.07	0.08	0.09	0.08	0.13	1.01	1.08	0.05	0.70	
d1-37	1.71	0.05	0.05	0.05	1.13	1.72	0.75	0.96	0.20	0.06	0.10	0.98	0.05	0.04	0.04	47-70 J1 K120 1b
	1.91	0.04	0.05	0.05	1.96	1.89	1.57	1.65	0.24	0.07	0.30	1.26	0.12	0.04	0.04	
d1-39	1.84	0.06	0.35	0.06	0.82	0.07	0.07	0.07	0.08	0.06	0.07	0.81	0.97	0.32	0.59	11 to 21 2c
	1.83	0.05	0.37	0.07	1.43	0.07	0.07	0.07	0.15	0.06	0.09	1.12	0.89	0.53	1.01	
d1-113	2.58	0.13	0.14	0.48	2.42	2.18	2.71	0.31	0.74	0.24	0.65	0.99	0.07	0.05	0.36	28-47 J1 5
	2.44	0.22	0.22	0.65	2.52	2.57	2.61	0.42	1.12	0.39	0.60	1.07	0.04	0.05	0.51	
d1-143	2.31	0.08	0.09	0.17	1.70	2.17	0.12	0.09	0.12	0.12	0.25	0.90	0.98	0.25	0.81	21 to 28 J1, J2 6
	2.29	0.12	0.12	0.30	2.29	2.25	0.17	0.12	0.24	0.18	0.30	1.04	1.05	0.37	0.95	
d1-149	1.84	0.06	0.07	0.08	1.57	2.14	2.08	1.28	0.16	0.06	0.07	0.59	0.64	0.10	0.51	47-70 J1, J2 7
	2.12	0.06	0.06	0.08	1.85	2.22	2.33	1.89	0.24	0.08	0.08	0.78	0.54	0.15	0.66	
d1-155	2.17	0.09	0.13	0.21	2.22	2.34	2.44	0.10	0.48	0.16	0.27	0.90	0.04	0.04	0.44	28-47 J1 5
	2.41	0.10	0.11	0.27	2.02	2.24	2.23	0.18	0.56	0.21	0.27	1.07	0.04	0.05	0.73	
d1-182	2.66	0.13	0.16	0.74	2.68	2.77	2.82	2.47	0.83	0.51	0.98	1.11	0.04	0.06	0.93	
	2.71	0.14	0.21	0.89	2.48	2.48	2.57	2.28	1.56	0.49	0.89	1.52	0.06	0.07	1.25	
d1-183	2.46	0.10	0.12	0.31	2.26	2.59	2.62	0.15	0.56	0.19	0.45	0.99	0.05	0.05	0.78	28-47 J1 5
	2.56	0.11	0.14	0.35	2.42	2.41	2.53	0.16	0.74	0.24	0.39	1.39	0.07	0.05	1.10	
d1-191	2.42	0.11	0.11	0.27	2.25	2.49	2.53	0.16	0.54	0.16	0.31	0.95	0.04	0.04	0.70	28-47 J1 5
	2.43	0.14	0.16	0.33	2.30	2.36	2.35	0.19	0.56	0.24	0.39	1.34	0.04	0.05	1.13	

FIG. 47A

53 / 55

d1-192	2.54	0.06	0.06	0.08	2.44	2.79	2.70	2.68	0.65	0.09	0.13	1.01	1.00	0.39	0.90	70-80 J1J2 + + + 8
	2.61	0.06	0.07	0.09	2.35	2.55	2.55	2.62	0.75	0.10	0.14	1.56	1.46	0.73	1.51	
d1-201	1.66	0.08	0.12	0.22	1.73	2.02	2.04	1.92	1.27	0.61	0.37	1.18	1.08	0.86	1.16	80-95 9
	2.14	0.14	0.27	0.48	1.92	1.93	2.13	2.07	1.89	1.10	0.63	1.31	1.30	1.12	1.29	
d1-203	1.80	0.04	0.04	0.19	0.05	0.06	0.06	0.06	0.05	0.05	0.05	0.58	0.33	0.04	0.18	1 to 11 J1J2 K120 4
	1.30	0.05	0.05	0.05	0.06	0.06	0.06	0.06	0.05	0.05	0.05	0.59	0.34	0.04	0.10	
d1-253	1.42	0.04	0.04	0.06	1.07	1.70	0.63	0.63	0.11	0.05	0.07	0.93	0.91	0.30	0.87	47-70 J1J2 7
	1.91	0.05	0.05	0.07	1.76	1.81	0.66	0.72	0.17	0.06	0.08	1.17	0.98	0.42	1.07	
d1-257	0.99	0.05	0.05	0.06	0.98	1.43	1.42	1.27	0.34	0.05	0.07	0.78	0.83	0.29	0.69	47-70 J1 + + + 7
	1.41	0.05	0.05	0.06	1.38	1.66	1.61	1.58	0.35	0.08	0.06	1.00	0.89	0.48	0.86	
d1-278	0.97	0.05	0.04	0.05	0.62	0.71	0.86	0.05	0.09	0.05	0.07	0.70	0.04	0.04	0.40	28-47 J1 5
	1.00	0.06	0.06	0.09	1.35	1.16	1.39	0.06	0.12	0.06	0.08	1.08	0.04	0.05	0.57	
d1-282	1.46	0.04	0.04	0.15	1.36	1.63	1.68	0.13	0.28	0.13	0.23	0.97	0.08	0.06	0.87	28-47 J1 5
	1.91	0.05	0.05	0.30	1.78	1.84	1.81	0.12	0.48	0.14	0.34	1.54	0.09	0.08	1.00	
d1-285	1.52	0.04	0.04	0.05	1.11	0.06	0.07	0.05	0.07	0.05	0.06	0.95	0.96	0.34	0.84	1 to 21 2
	1.97	0.05	0.05	0.07	1.57	0.06	0.06	0.06	0.09	0.05	0.06	1.31	0.99	0.52	1.21	
B1	1.38	1.15	0.94	0.89	1.25	1.48	1.70	1.59	0.92	0.84	0.74	0.97	0.97	0.98	0.95	
	1.88	1.75	1.55	0.81	1.64	1.66	1.70	1.57	1.42	1.21	1.18	1.51	1.28	1.02	1.42	
IMM510	1.59	1.31	1.09	0.56	1.27	1.61	1.72	1.63	0.86	0.90	0.91	1.03	0.99	0.92	0.81	
	1.53	1.43	1.41	0.69	1.59	2.00	1.80	1.87	1.00	0.96	0.88	1.30	1.15	0.88	1.32	

FIG. 47A (Cont.)

54 / 55

	Dolphin δ 1	Dolph-11	Dolph-21	Dolph-28	Dolph-47	Dolph-70	Dolph-80	Dolph-95	δ 1-Arg16	K120A	K120F	J2-M	Epitope
D1-113	0.053	1.444	1.384	1.556	0.081	0.060	0.053	0.054	1.574	1.634	0.059	0.051	28-47
D1-143	0.048	0.790	0.941	0.055	0.046	0.047	0.047	0.042	1.178	1.100	1.124	1.172	21-28
D1-155	0.053	0.933	1.201	1.282	0.051	0.050	0.050	0.049	1.107	1.093	0.053	0.050	28-47
D1-182	0.079	1.322	1.362	1.348	1.099	0.248	0.104	0.088	1.378	1.393	0.086	0.090	47-70
D1-183	0.048	1.077	1.012	1.154	0.046	0.048	0.048	0.046	1.202	1.060	0.088	0.065	28-47
D1-191	0.051	1.043	1.125	1.157	0.055	0.056	0.055	0.053	1.167	1.204	0.053	0.051	28-47
D1-192	0.049	0.926	1.195	1.020	0.973	0.199	0.092	0.047	1.173	1.207	1.157	0.995	47-70
D1-201	0.062	1.030	1.161	1.223	0.990	0.979	0.769	0.051	1.191	1.036	1.306	1.116	80-95
D1-203	0.053	0.046	0.051	0.061	0.072	0.058	0.055	0.050	0.972	0.267	0.241	0.491	1-11
D1-253	0.047	1.100	1.612	0.629	0.409	0.100	0.053	0.050	1.500	1.132	1.343	1.180	47-70
D1-257	0.053	0.871	1.359	1.181	0.812	0.115	0.075	0.055	1.252	0.928	1.179	0.882	47-70
D1-278	0.054	0.931	1.114	1.271	0.054	0.050	0.051	0.056	1.014	0.825	0.056	0.055	28-47
D1-282	0.116	1.354	1.460	1.575	0.105	0.099	0.114	0.104	1.599	1.370	0.173	0.220	28-47
D1-285	0.052	1.089	0.053	0.062	0.054	0.047	0.051	0.053	1.341	1.330	1.441	1.271	11 to 21
TS-1	0.111	1.036	1.642	0.910	0.561	0.081	0.103	0.329	1.649	0.492	0.658	1.002	

FIG. 47B

55 / 55

	Dolphin (1-94)	Dolphin (28-94)	Dolphin (44-94)	Dolphin (72-94)	Dolphin (83-94)	CDR 1(S)	CDR2(G)	CDR3(S)	Epitope	critical
Isotype	-	-	-	-	-	-	-	-		
δ 2-14	-	+	+	+	+	+	+	+	1-27	
δ 2-17	-	-	-	-	-	+	+	+	83-94	
δ 2-22	-	+	+	+	+	+	+	+	1-27	
δ 2-30	-	+	+	+	+	+	+	+	1-27	
δ 2-31	-	+	+	+	+	+	+	+	1-27	
δ 2-32	-	-	-	-	+	+	+	+	72-83	
δ 2-33	-	+	+	+	+	+	+	+	1-27	
δ 2-36	-	+	+	+	+	+	+	+	1-27	
δ 2-37	-	+	+	+	+	+	+	+	1-27	
B6	-	-	-	-	-	+	+	+	83-94	
15D	-	-	+	+	+	-	+	+	28-38	G35
7A5	+	+	+	+	+	+	+	+	γ 9 binder	
IMM510	+	+	+	+	+	+	+	+	γ 8 pan	
B1	+	+	+	+	+	+	+	+	γ 8 pan	

FIG. 48

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US17/32530

A. CLASSIFICATION OF SUBJECT MATTER

IPC - A61K 35/12, 35/17; C12N 5/0783 (2017.01)

CPC - A61K 35/12, 35/17, 39/0011; C12N 5/0636, 5/0638, 5/0783

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

See Search History document

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

See Search History document

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

See Search History document

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X ----- Y	US 2014/0141513 A1 (DE CARVALHO SILVA SANTOS et al.) May 22, 2014; abstract; paragraphs [0014], [0016]	95-96, 98/95-96, 99/95-96 --- 97, 98/97, 99/97
Y	US 2010/0272739 A1 (GELFAND et al.) October 28, 2010; paragraph [0062]	97, 98/97, 99/97
A	US 5,185,250 A (BRENNER et al.) February 9, 1993; abstract	1-3, 5, 7-11, 48-53, 87-89, 92-93, 94/92-93
A	US 2015/0259645 A1 (INSERM (INSTITUT NATAIONAL DE LA SANTÉ ET DE LA RECH MEDICALE) et al.) September 17, 2015; paragraph [0121]; claims 1, 4, 10	1-3, 5, 7-11, 20-21, 23-24, 43-57, 58/55-57, 59/58/55-57, 60/55-57, 61/60/55-57, 62-64, 65/62-64, 66/65/62-64, 67/62-64, 68/67/62-64, 78
A	US 2003/0138433 A1 (NEWELL et al.) July 24, 2003; paragraph [0027]	20-21, 23-24

☒ Further documents are listed in the continuation of Box C.☐ See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

09 August 2017 (09.08.2017)

Date of mailing of the international search report

28 AUG 2017

Name and mailing address of the ISA/

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Authorized officer

Shane Thomas

PCT Helpdesk: 571-272-4300

PCT OSP: 571-272-7774

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US17/32530

Box No. 1 Nucleotide and/or amino acid sequence(s) (Continuation of item 1.c of the first sheet)

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international search was carried out on the basis of a sequence listing:
- a. ☐ forming part of the international application as filed:
☐ in the form of an Annex C/ST.25 text file.
☐ on paper or in the form of an image file.
- b. ☐ furnished together with the international application under PCT Rule 13*ter*.1(a) for the purposes of international search only in the form of an Annex C/ST.25 text file.
- c. ☒ furnished subsequent to the international filing date for the purposes of international search only:
☒ in the form of an Annex C/ST.25 text file (Rule 13*ter*.1(a)).
☐ on paper or in the form of an image file (Rule 13*ter*.1(b) and Administrative Instructions, Section 713).
2. ☒ In addition, in the case that more than one version or copy of a sequence listing has been filed or furnished, the required statements that the information in the subsequent or additional copies is identical to that forming part of the application as filed or does not go beyond the application as filed, as appropriate, were furnished.
3. Additional comments:

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US17/32530

Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☒ Claims Nos.: 4, 6, 12-19, 22, 25-42, 69-77, 79-80, 90-91, 101-102
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying additional fees, this Authority did not invite payment of additional fees.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.
- ☐ The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.
- ☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US17/32530

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 2015/061694 A2 (BOARD OF REGENTS, THE UNIVERSITY OF TEXAS SYSTEM) April 30, 2015; (abstract; paragraphs [0012], [0013], [0018], [0021], [0023], [0061], [0062], [0067], [0081], [0082], [00108]; claim 6	43-47, 54-57, 58/55-57, 59/58/55-57, 60/55-57, 61/60/55-57, 62-64, 65/62-64, 66/65/62-64, 67/62-64, 68/67/62-64, 78, 81-89, 92-94
A	US 2012/0034221 A1 (BONVINI et al.) February 9, 2012; abstract; paragraphs [0023], [0060], [0079]	81-89