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(54) Title: IMMUNE RESPONSE INDUCER

(57) **Abrégé/Abstract:**

An immunity-inducing agent comprising as an effective ingredient a specific polypeptide is disclosed. These polypeptides were isolated, by the SEREX method using a cDNA library derived from canine testis and serum from a cancer-bearing dog, as a polypeptide which binds to an antibody existing specifically in serum derived from a cancer-bearing living body. The polypeptides can induce immunity in a living body and cause regression of a tumor in a cancer-bearing living body. Therefore, these polypeptides are especially effective as a therapeutic and/or prophylactic agent for a cancer(s).

ABSTRACT

An immunity-inducing agent comprising as an effective ingredient a specific polypeptide is disclosed. These polypeptides were isolated, by the SEREX method using a cDNA library derived from canine testis and serum from a cancer-bearing dog, as a polypeptide which binds to an antibody existing specifically in serum derived from a cancer-bearing living body. The polypeptides can induce immunity in a living body and cause regression of a tumor in a cancer-bearing living body. Therefore, these polypeptides are especially effective as a therapeutic and/or prophylactic agent for a cancer(s).

DESCRIPTION

Immune Response Inducer

TECHNICAL FIELD

[0001]

5 The present invention relates to a novel immunity-inducing agent useful as a therapeutic and/or prophylactic agent for a cancer(s).

BACKGROUND ART

[0002]

10 Cancers are the commonest cause for death among all of the causes for death, and the therapies therefor are mainly surgical treatment in combination with radiotherapy and chemotherapy. In spite of the developments of new surgical methods and discovery of new anti-cancer agents in recent years, treatment results of cancers are not improved very much at present except for some cancers. In recent years, by virtue of the development in molecular biology and cancer immunology, cancer antigens recognized by cytotoxic T cells reactive with cancers, as well as the genes encoding the cancer antigens, were identified, and expectations for antigen-specific immunotherapies have been raised (see Non-patent Literature 1). In immunotherapy, to reduce side effects, it is necessary that the peptide or protein recognized as the antigen exist hardly in normal cells and exist specifically in cancer cells. In 1991, Boon *et al.* of Ludwig Institute in Belgium isolated human melanoma antigen MAGE 1 recognized by CD8-positive T cells by a cDNA-expression cloning method using an autologous cancer cell line and cancer-reactive T cells (see Non-patent Literature 2). Thereafter, the SEREX (serological identifications of antigens by recombinant expression cloning) method, wherein tumor antigens recognized by antibodies produced in the living body of a cancer patient in response to the cancer of the patient himself are identified by application of a gene expression cloning method, was reported (Non-patent Literature 3; Patent

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Literature 1), and various cancer antigens have been isolated (see Non-patent Literatures 4 to 9). Using a part thereof as targets, clinical tests for cancer immunotherapy have started.

[0003]

5 On the other hand, as in human, a number of tumors such as mammary gland tumor and squamous cell carcinoma are known in dogs and cats, and they rank high also in the statistics of diseases in dogs and cats. However, at present, no therapeutic, prophylactic or diagnostic agents exist which are effective for cancers in dogs and cats. Most of tumors in dogs and cats are realized by owners only after
10 they advance to grow bigger, and in many cases, it is already too late to visit a hospital to receive surgical excision of the tumor or administration of a human drug (an anticancer preparation or the like), so that those dogs and cats die shortly after the treatment. Under such circumstances, if therapeutic agents, prophylactic agents and diagnostic agents for cancers effective for dogs and cats become available, their uses
15 for canine cancers are expected to be developed.

[0004]

Patent Literature 1: US 5698396 B

Non-patent Literature 1: Tsuyoshi Akiyoshi, Cancer and Chemotherapy, 24, 551-519 (1997)

20 Non-patent Literature 2: Bruggen P. et al., Science, 254:1643-1647 (1991)

Non-patent Literature 3: Proc. Natl. Acad. Sci. USA, 92:11810-11813 (1995)

Non-patent Literature 4: Int. J. Cancer, 72:965-971 (1997)

Non-patent Literature 5: Cancer Res., 58:1034-1041 (1998)

Non-patent Literature 6: Int. J. Cancer, 29:652-658 (1998)

25 Non-patent Literature 7: Int. J. Oncol., 14:703-708 (1999)

Non-patent Literature 8: Cancer Res., 56:4766-4772 (1996)

Non-patent Literature 9: Hum. Mol. Genet 6:33-39, 1997

Non-patent Literature 10: Naokazu Inoue, Ryo Yamaguchi and Masahito Ikawa,
Protein, Nucleic Acid and Enzyme, Vol. 50, No. 11, 1405-1412

Non-patent Literature 11: J Cell Sci. 115:1825-35

Non-patent Literature 12: Blood. 95:1788-96

5 Non-patent Literature 13: Mol Endocrinol. 9:243-54 (1995)

Non-patent Literature 14: J Cell Biol. 145 : 83-98 (1999)

DISCLOSURE OF THE INVENTION

PROBLEMS TO BE SOLVED BY THE INVENTION

[0005]

10 An object of the present invention is to provide a novel immunity-inducing
agent which is useful as a therapeutic and/or prophylactic agent for a cancer(s)

MEANS FOR SOLVING THE PROBLEMS

[0006]

The present inventors intensively studied to obtain a cDNA encoding a
15 protein which binds to an antibody existing in serum derived from a cancer-bearing
living body by the SEREX method using a cDNA library derived from canine testis
and serum of a cancer-bearing dog, which cDNA was used to prepare a polypeptide
having the amino acid sequence shown in SEQ ID NO:2, a canine calmegin protein
having the amino acid sequence shown in SEQ ID NO:16, a canine centrosomal
20 protein (which may be hereinafter abbreviated as CEP) having the amino acid
sequence shown in SEQ ID NO:26, and the canine thyroid hormone receptor
interactor 11 (which may be hereinafter described as "TRIP11") having the amino
acid sequence shown in SEQ ID NO:39. Further, based on a registered canine gene
having a high homology to the canine CEP of the above-described SEQ ID NO:26, a
25 canine CEP having the amino acid sequence shown in SEQ ID NO:28 was prepared.
Further, based on a human gene homologous to the obtained gene, a polypeptide
having the amino acid sequence shown in SEQ ID NO:4, a human calmegin protein

having the amino acid sequence shown in SEQ ID NO:18, a human CEP having the amino acid sequence shown in SEQ ID NO:30, and a human TRIP11 having the amino acid sequence shown in SEQ ID NO:41 were prepared. The inventors then discovered that these polypeptides can induce immunocytes in a living body and cause regression of an already occurred tumor when administered to the living body, thereby completing the present invention.

[0007]

That is, the present invention provides an immunity-inducing agent comprising as an effective ingredient any one of the polypeptides (a) to (c) below, the polypeptide having an immunity-inducing activity, or as an effective ingredient a recombinant vector which comprises a polynucleotide encoding the polypeptide and is capable of expressing the polypeptide *in vivo*: (a) a polypeptide consisting of not less than 7 consecutive amino acids of the amino acid sequence shown in SEQ ID NO:2, 4, 16, 18, 26, 28, 30, 39 or 41 in SEQUENCE LISTING; (b) a polypeptide having a homology of not less than 80% to the polypeptide (a) and consisting of not less than 7 amino acids; and (c) a polypeptide comprising the polypeptide (a) or (b) as a partial sequence thereof. The present invention also provides a method for inducing immunity, the method comprising administering to an individual an effective amount of any one of the above-described polypeptides (a) to (c), the polypeptide having an immunity-inducing activity, or an effective amount of a recombinant vector which comprises a polynucleotide encoding the polypeptide and is capable of expressing the polypeptide *in vivo*. The present invention further provides a method for treating antigen-presenting cells, the method comprising bringing any one of the above-described polypeptides (a) to (c), the polypeptide having an immunity-inducing activity, into contact with antigen-presenting cells. The present invention further provides use of any one of the above-described polypeptides (a) to (c), the polypeptide having an immunity-inducing activity, or a

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recombinant vector which comprises a polynucleotide encoding the polypeptide and is capable of expressing the polypeptide *in vivo*, for production of an immunity-inducing agent.

[0007A]

The present invention as claimed relates to:

5 - an immunity-inducing agent to treat and/or prevent a calmegin-expressing cancer(s), said agent comprising as an effective ingredient any one of the polypeptides (a) to (c) below, said polypeptide having an immunity-inducing activity: (a) a polypeptide having the amino acid sequence shown in SEQ ID NO: 16 or 18 in SEQUENCE LISTING; (b) a polypeptide having a sequence identity of not less than 89% to the full length of the polypeptide (a); and (c) a
10 polypeptide comprising the polypeptide (a) or (b) as a partial sequence;

- a method for preparing antigen-presenting cells *in vitro*, said method comprising bringing a polypeptide having the amino acid sequence shown in SEQ ID NO: 16 or 18 in SEQUENCE LISTING into contact with dendritic cells or B cells which have MHC class I molecules; and

15 - use of any one of the polypeptides (a) to (c) below, said polypeptide having an immunity-inducing activity for production of an immunity-inducing agent to treat and/or prevent a calmegin-expressing cancer(s): (a) a polypeptide having the amino acid sequence shown in SEQ ID NO: 16 or 18 in SEQUENCE LISTING; (b) a polypeptide having a sequence identity of not less than 89% to the full length of the polypeptide (a); and (c) a polypeptide comprising the
20 polypeptide (a) or (b) as a partial sequence.

EFFECT OF THE INVENTION

[0008]

By the present invention, a novel immunity-inducing agent useful as a therapeutic and/or prophylactic agent for a cancer(s) was provided. As indicated in the Examples below, the polypeptide
25 used in the present invention can induce immunocytes in a cancer-bearing dog and also can cause reduction or regression of an already occurred tumor when administered to a cancer-bearing dog. Therefore, the polypeptide is useful for therapy and prophylaxis of a cancer(s).

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BRIEF DESCRIPTION OF THE DRAWINGS

[0009]

Fig. 1 shows the expression pattern of the gene identified in Example A-1 in normal tissues and tumor cell lines. Reference numeral 1: the expression pattern of the identified gene; Reference numeral 2: the expression pattern of the *GAPDH* gene.

Fig. 2 shows the detection by Coomassie staining of the canine-derived protein produced in *E. coli* and purified in Example A, which protein was identified in the present invention. Reference numeral 3: the band for the canine-derived protein of the present invention.

Fig. 3 shows the expression pattern of the calmegin gene identified in the present invention in normal tissues and tumor cell lines. Reference numeral 1: the expression pattern of the calmegin gene; Reference numeral 2: the expression pattern of the *GAPDH* gene.

Fig. 4 shows the detection by Coomassie staining of the canine calmegin protein, which is an example of the polypeptide used in the present invention, produced in *E. coli* and purified in Example B. Reference numeral 3: the band for

the canine calmeglin protein.

Fig. 5 shows the expression pattern of the gene encoding the CEP protein in normal tissues and tumor cell lines. Reference numeral 1: the expression pattern of the gene encoding the CEP protein; Reference numeral 2: the expression pattern of the *GAPDH* gene.

Fig. 6 shows the detection by Coomassie staining of the canine CEP of SEQ ID NO:26, which is an example of the polypeptide used in the present invention, produced in *E. coli* and purified in Example C. Reference numeral 3: the band for the canine CEP protein.

Fig. 7 shows the expression pattern of the gene encoding the TRIP11 protein in normal tissues and tumor cell lines. Reference numeral 1: the expression pattern of the gene encoding the TRIP11 protein; Reference numeral 2: the expression pattern of the *GAPDH* gene.

Fig. 8 shows the detection by Coomassie staining of the canine TRIP11 protein, which is one of the polypeptides used in the present invention, produced in *E. coli* and purified in Example D. Reference numeral 3: the band for the canine TRIP11 protein.

BEST MODE FOR CARRYING OUT THE INVENTION

[0010]

The polypeptides contained in the immunity-inducing agents of the present invention as effective ingredients are as follows. It should be noted that the term "polypeptide" in the present invention means a molecule formed by peptide bonding of a plurality of amino acids, and includes not only polypeptide molecules having large numbers of amino acids constituting them, but also low molecular weight molecules having small numbers of amino acids (oligopeptides) and full-length proteins. Thus, in the the present invention, proteins consisting of the full length of SEQ ID NO:2, 4, 16, 18, 26, 28, 30, 39 or 41 are also included in "polypeptide".

(a) A polypeptide which consists of not less than 7 consecutive amino acids of a polypeptide having the amino acid sequence shown in SEQ ID NO:2, 4, 16, 18, 26, 28, 30, 39 or 41 in SEQUENCE LISTING and has an immunity-inducing activity.

5 (b) A polypeptide which has a homology of not less than 80% to the polypeptide (a), consists of not less than 7 amino acids, and has an immunity-inducing activity.

(c) A polypeptide which comprises the polypeptide (a) or (b) as a partial sequence thereof and has an immunity-inducing activity.

10 [0011]

It should be noted that the term "having the amino acid sequence" in the present invention means that amino acid residues are aligned in that order.

Accordingly, for example, "a polypeptide having the amino acid sequence shown in SEQ ID NO:2" means a polypeptide having a size of 306 amino acid residues, whose amino acid sequence is Met Ala Ala Leu ...(snip)... Ile Thr Ser Pro as shown in SEQ ID NO:2. Further, "a polypeptide having the amino acid sequence shown in SEQ ID NO:2" may be abbreviated as "a polypeptide of SEQ ID NO:2". This also applies to the term "having the base sequence".

[0012]

20 As used herein, the term "immunity-inducing activity" means an ability to induce immunocytes which secrete cytokines such as interferon in a living body. Whether or not a polypeptide has an immunity-inducing activity can be confirmed using, for example, the known ELISPOT assay. More particularly, for example, as described in the Examples below, cells such as peripheral blood mononuclear cells are obtained from a living body to which a polypeptide whose immunity-inducing activity is to be evaluated was administered, which cells are then cocultivated with
25 the polypeptide, followed by measuring the amount of a cytokine produced by the

cells using a specific antibody, thereby measuring the number of immunocytes in the cells, which enables evaluation of the immunity-inducing activity. Further, as described in the Examples below, a recombinant polypeptide prepared based on the amino acid sequence of SEQ ID NO:2, 4, 16, 18, 26, 28, 30, 39 or 41 can cause regression of a tumor by its immunity-inducing activity when administered to a cancer-bearing living body. Therefore, the above-described immunity-inducing activity can be evaluated also as the ability to inhibit the growth of cancer cells expressing the polypeptide of SEQ ID NO:2, 4, 16, 18, 26, 28, 30, 39 or 41 or to cause reduction or disappearance of a cancer tissue (tumor) (hereinafter referred to as "anti-tumor activity"). The anti-tumor activity of a polypeptide can be confirmed by, for example, observation of whether or not the tumor is reduced when the polypeptide was administered to a cancer-bearing living body, as more particularly described in the Examples below. Further, the anti-tumor activity of a polypeptide can be evaluated also by observation of whether or not T cells stimulated with the polypeptide (that is, T cells brought into contact with antigen-presenting cells which present the polypeptide) show a cytotoxic activity against tumor cells *in vitro*. The contact between T cells and antigen-presenting cells can be carried out by cocultivation of the both in a liquid medium, as mentioned below. Measurement of the cytotoxic activity can be carried out by, for example, a known method called ⁵¹Cr release assay described in Int.J.Cancer,58:p317,1994. In cases where a polypeptide is used for therapy and/or prophylaxis of a cancer(s), the evaluation of the immunity-inducing activity is preferably carried out using the anti-tumor activity as an index, although the index is not restricted.

[0013]

The amino acid sequence shown in SEQ ID NO:2 in SEQUENCE LISTING is the amino acid sequence of the polypeptide with unknown function isolated as a polypeptide which binds to an antibody existing specifically in serum derived from a

cancer-bearing dog, which isolation was carried out by the SEREX method using a canine testis-derived cDNA library and serum of a cancer-bearing dog (see Example A-1). It is registered in the NCBI database under Accession No. XP_535343 (protein) and Accession No. XM_535343 (coding gene), but its function has not been reported. Further, the amino acid sequence shown in SEQ ID NO:4 is an amino acid sequence of a human homologous factor of the polypeptide of SEQ ID NO:2 isolated as described above. This human homologous factor is also a protein whose function is unknown, which is registered in the NCBI database under Accession No. NP_689873 (protein) and Accession No. NM_152660 (coding gene). The homology between them is 93% in terms of base sequence and 99% in terms of amino acid sequence.

[0014]

The respective amino acid sequences shown in SEQ ID NOs:16 and 18 are those of the calmegin protein isolated as a polypeptide and a human homologous factor thereof, which polypeptide binds to an antibody existing specifically in serum derived from a cancer-bearing dog, which isolation was carried out by the SEREX method using a canine testis-derived cDNA library and serum of a cancer-bearing dog (see Example B-1). Calmegin was identified as a protein which is expressed specifically at the time of differentiation of a spermatid, and it has a chaperone activity *in vitro*. Since it is expressed only in testis and disappears in a mature sperm, calmegin is considered to have a function to fold proteins involved in differentiation of spermatid (Non-patent Literature 10, Naokazu Inoue, Ryo Yamaguchi and Masahito Ikawa, Protein, Nucleic Acid and Enzyme, Vol. 50, No. 11, 1405-1412). However, there has been no report showing that the protein is expressed in a cancer and useful for therapy or prophylaxis thereof. The homology between the canine calmegin gene and the human calmegin gene is 90% in terms of base sequence and 89% in terms of amino acid sequence.

[0015]

The respective amino acid sequences shown in SEQ ID NOs:26, 28 and 30 are those of the CEP isolated as a polypeptide, a canine factor having a high homology to the polypeptide and a human homologous factor of the polypeptide, which polypeptide binds to an antibody existing specifically in serum derived from a cancer-bearing dog, which isolation was carried out by the SEREX method using a canine testis-derived cDNA library and serum of a cancer-bearing dog (see Example C-1). CEP is a protein which is required by the centrosome to control microtubules and also involved in maturation of the centrosome. It is known that chromosomal translocation frequently occurs in some of myeloproliferative disorders, and since the *CEP* gene exists at the point where the translocation occurs, CEP is considered to have a certain relationship with the disorders. However, there has been no report showing that the protein is expressed in a cancer and useful for therapy or prophylaxis thereof (Non-patent Literature 11: J Cell Sci. 115:1825-35; Non-patent Literature 12: Blood. 95:1788-96). The homology between the canine *CEP* gene encoding the CEP of SEQ ID NO:26 and the human *CEP* gene is 87% in terms of base sequence and 84% in terms of amino acid sequence.

[0016]

The respective amino acid sequences shown in SEQ ID NOs:39 and 41 are those of the TRIP11 protein isolated as a polypeptide and a human homologous factor thereof, which polypeptide binds to an antibody existing specifically in serum derived from a cancer-bearing dog, which isolation was carried out by the SEREX method using a canine testis-derived cDNA library and serum of a cancer-bearing dog (see Example D-1). TRIP11 (thyroid hormone receptor interactor 11) was first identified as a factor which interacts with the thyroid hormone receptor β , and its binding to Golgi bodies and microtubules also became evident, so that TRIP11 is considered to play a role in maintaining the shapes of these organelles. However,

there has been no report showing that the protein is expressed in a cancer and useful for therapy or prophylaxis thereof (Non-patent Literature 13, Mol Endocrinol. 9:243-54 (1995); Non-patent Literature 14, J Cell Biol. 145: 83-98 (1999)). The homology between the canine *TRIP11* gene and the human *TRIP11* gene is 88% in terms of base sequence and 86% in terms of amino acid sequence.

[0017]

The polypeptide (a) consists of not less than 7 consecutive, preferably not less than 9 consecutive amino acids of a polypeptide having the amino acid sequence shown in SEQ ID NO:2, 4, 16, 18, 26, 28, 30, 39 or 41, and has an immunity-inducing activity. The polypeptide especially preferably has the amino acid sequence shown in SEQ ID NO:2, 4, 16, 18, 26, 28, 30, 39 or 41. As known in the art, a polypeptide consists of not less than about 7 amino acid residues can exert its antigenicity. Thus, a polypeptide consists of not less than 7 consecutive amino acid residues of the amino acid sequence shown in SEQ ID NO:2, 4, 16, 18, 26, 28, 30, 39 or 41 can have an immunity-inducing activity, so that it can be used for preparation of the immunity-inducing agent of the present invention. However, in view of the fact that antibodies produced against antigenic substances in a living body are polyclonal antibodies, it is thought that an antigenic substance composed of larger number of amino acid residues can induce more types of antibodies which can recognize various sites on the antigenic substance, thereby attaining higher immunity-inducing activity. Therefore, in order to increase the immunity-inducing activity, in the case of SEQ ID NO:2 or 4, the number of the amino acid residues may be preferably not less than 30, more preferably not less than 100, still more preferably not less than 200, still more preferably not less than 250. In the case of SEQ ID NO:16 or 18, the number of the amino acid residues may be preferably not less than 30, more preferably not less than 100, still more preferably not less than 200, still more preferably not less than 400, still more preferably not less than 550. In the

case of SEQ ID NO:26, 28 or 30, the number of the amino acid residues may be preferably not less than 30, more preferably not less than 100, still more preferably not less than 300, still more preferably not less than 600, still more preferably not less than 1000, still more preferably not less than 1500, still more preferably not less than 2000. In the case of SEQ ID NO:39 or 41, the number of the amino acid residues may be preferably not less than 30, more preferably not less than 100, still more preferably not less than 300, still more preferably not less than 600, still more preferably not less than 1000, still more preferably not less than 1500.

[0018]

As a principle of immune induction by administration of a cancer antigenic polypeptide, the following process is known: the polypeptide is incorporated into an antigen-presenting cell and then degraded into smaller fragments by peptidases in the cell, followed by presentation of the fragments on the surface of the cell. The fragments are then recognized by a cytotoxic T cell or the like, which selectively kills cells presenting the antigen.

[0019]

The size of the polypeptide presented on the surface of the antigen-presenting cell is relatively small and about 7 to 30 amino acids. Therefore, from the view point of presenting thereof on the surface of the antigen-presenting cell, a polypeptide consisting of about 7 to 30, preferably about 9 to 30 consecutive amino acids of the amino acid sequence shown in SEQ ID NO:2, 4, 16, 18, 26, 28, 30, 39 or 41 is sufficient as the above-described polypeptide (a). In some cases, these relatively small polypeptides are presented directly on the surface of the antigen-presenting cells without incorporation thereof into the antigen-presenting cells.

[0020]

However, as described above, since a polypeptide incorporated into an antigen-presenting cell is cleaved at random sites by peptidases in the cell to yield

various polypeptide fragments, which are then presented on the surface of the antigen-presenting cell, administration of a large polypeptide such as the entire region of SEQ ID NO:2, 4, 16, 18, 26, 28, 30, 39 or 41 inevitably causes production of polypeptide fragments by degradation thereof in the antigen-presenting cell, which fragments are effective for immune induction via the antigen-presenting cell.

Therefore, for immune induction via antigen-presenting cells, a large polypeptide can also be preferably used. In the case of SEQ ID NO:2 or 4, the number of the amino acids may be preferably not less than 30, more preferably not less than 100, still more preferably not less than 200, still more preferably not less than 250. In the case of SEQ ID NO:16 or 18, the number of the amino acids may be preferably not less than 30, more preferably not less than 100, still more preferably not less than 200, still more preferably not less than 400, still more preferably not less than 550. In the case of SEQ ID NO:26, 28 or 30, the number of the amino acids may be preferably not less than 30, more preferably not less than 100, still more preferably not less than 300, still more preferably not less than 600, still more preferably not less than 1000, still more preferably not less than 1500, still more preferably not less than 2000. In the case of SEQ ID NO:39 or 41, the number of the amino acids may be preferably not less than 30, more preferably not less than 100, still more preferably not less than 300, still more preferably not less than 600, still more preferably not less than 1000, still more preferably not less than 1500.

[0021]

The above-described polypeptide (b) is the same polypeptide as the above-described polypeptide (a) except that a small number of amino acid residues are substituted, deleted and/or inserted, which has a homology of not less than 80%, preferably not less than 90%, more preferably not less than 95%, still more preferably not less than 98% to the original sequence, and has an immunity-inducing activity. It is well known in the art that, in general, there are cases where a protein antigen

retains substantially the same antigenicity as the original even if the amino acid sequence of the protein is modified such that a small number of amino acids are substituted, deleted and/or inserted. Therefore, since the above-described polypeptide (b) may also exert an immunity-inducing activity, it can be used for preparation of the immunity-inducing agent of the present invention. Further, the above-described polypeptide (b) is also preferably the same polypeptide as one having the amino acid sequence shown in SEQ ID NO:2, 4, 16, 18, 26, 28, 30, 39 or 41 except that one or several amino acid residues are substituted, deleted and/or inserted.

10 [0022]

As used herein, the term "homology" of amino acid sequences means a value expressed in percentage which is calculated by aligning two amino acid sequences to be compared such that the number of matched amino acid residues is the maximum, and dividing the number of the matched amino acid residues by the number of the total amino acid residues. When the above-described alignment is carried out, a gap(s) is/are inserted into one or both of the two sequences to be compared as required. Such alignment of sequences can be carried out using a well-known program such as BLAST, FASTA or CLUSTAL W. When a gap(s) is/are inserted, the above-described number of the total amino acid residues is calculated by counting one gap as one amino acid residue. When the thus counted numbers of the total amino acid residues are different between the two sequences to be compared, the homology (%) is calculated by dividing the number of matched amino acid residues by the number of the total amino acid residues in the longer sequence.

20 [0023]

25 The 20 types of amino acids constituting the naturally occurring proteins may be classified into groups each of which has similar properties, for example, into neutral amino acids with side chains having low polarity (Gly, Ile, Val, Leu, Ala, Met,

Pro), neutral amino acids having hydrophilic side chains (Asn, Gln, Thr, Ser, Tyr, Cys), acidic amino acids (Asp, Glu), basic amino acids (Arg, Lys, His) and aromatic amino acids (Phe, Tyr, Trp). It is known that, in most cases, substitutions of amino acids within the same group do not change the properties of the polypeptide.

5 Therefore, in cases where amino acid residue(s) in the above described polypeptide (a) in the present invention is/are substituted, the probability that the immunity-inducing activity can be maintained may be made high by conducting the substitution(s) within the same group.

[0024]

10 The above-described polypeptide (c) comprises the above-described polypeptide (a) or (b) as a partial sequence and has an immunity-inducing activity. That is, the polypeptide (c) has another/other amino acid(s) or polypeptide(s) added at one or both ends of the polypeptide (a) or (b), and has an immunity-inducing activity. Such a polypeptide can also be used for preparation of the immunity-
15 inducing agent of the present invention.

[0025]

For example, the above-described polypeptides can be synthesized by a chemical synthesis method such as the Fmoc method (fluorenylmethylcarbonyl method) or the tBoc method (*t*-butyloxycarbonyl method). Further, they can be
20 synthesized by conventional methods using various commercially available peptide synthesizers. Further, the polypeptide of interest can be obtained by a known genetic engineering method wherein a polynucleotide encoding the above-described polypeptide is prepared and incorporated into an expression vector, which is then introduced into a host cell, in which the polypeptide is produced.

25 [0026]

The polynucleotide encoding the above-described polypeptide can be easily prepared by a known genetic engineering method or a conventional method using a

commercially available nucleic acid synthesizer. For example, DNA having the base sequence of SEQ ID NO:1, 15, 25, 27 or 38 can be prepared by carrying out PCR using the chromosomal DNA or a cDNA library of a dog as a template and using a pair of primers designed such that the primers can amplify the base sequence described in SEQ ID NO:1, 15, 25, 27 or 38, respectively. DNA having the base sequence of SEQ ID NO:3, 17, 29 or 40 can be prepared similarly by using as the above-described template the human chromosomal DNA or a cDNA library.

Conditions for the PCR reaction can be selected as appropriate, and examples of the conditions include, but are not limited to, those wherein a cycle comprising the reaction steps of 94°C for 30 seconds (denaturing), 55°C for 30 seconds to 1 minute (annealing), and 72°C for 2 minutes (extension) is repeated, for example, 30 times, followed by allowing the reaction to proceed at 72°C for 7 minutes. Further, a

desired DNA can be isolated by preparing an appropriate probe or primer based on the information of the base sequence and the amino acid sequence shown in SEQ ID NOs:1 to 4, 15 to 18, 25 to 30, 38 to 41 in SEQUENCE LISTING of the present specification and then using the probe or primer for screening of a cDNA library from a dog or a human. The cDNA library is preferably prepared from cells, an organ or a tissue expressing the protein of SEQ ID NO:2, 4, 16, 18, 26, 28, 30, 39 or 41. Operations such as the above-described preparation of a probe or a primer,

construction of a cDNA library, screening of a cDNA library and cloning of a gene of interest are known to those skilled in the art, and can be carried out according to, for example, Molecular Cloning, 2nd Ed. or Current Protocols in Molecular Biology.

From the thus obtained DNA, DNA encoding the above-described polypeptide (a) can be obtained. Further, since codons encoding each amino acid are known, the base sequence of a polynucleotide encoding a specific amino acid sequence can be easily specified. Therefore, the base sequences of polynucleotides encoding the above-described polypeptide (b) and polypeptide (c) can also be easily specified, so

that such polynucleotides can also be easily synthesized using a commercially available nucleic acid synthesizer according to a conventional method.

[0027]

The above-described host cells are not restricted as long as they can express
5 the above-described polypeptide, and examples thereof include, but are not limited to, prokaryotic cells such as *E. coli*; and eukaryotic cells such as mammalian cultured cells including monkey kidney cells COS 1 and Chinese hamster ovary cells CHO, budding yeast, fission yeast, silkworm cells, and *Xenopus laevis* egg cells.

[0028]

10 In cases where prokaryotic cells are used as the host cells, an expression vector having the origin that enables its replication in a prokaryotic cell, a promoter, a ribosome binding site, a DNA cloning site, a terminator and the like is used as the expression vector. Examples of the expression vector for *E. coli* include the pUC system, pBluescript II, pET expression system and pGEX expression system. By
15 incorporating DNA encoding the above-described polypeptide into such an expression vector and transforming prokaryotic host cells with the vector, followed by culturing the obtained transformant, the polypeptide encoded by the above-described DNA can be expressed in the prokaryotic host cells. In this case, the polypeptide can also be expressed as a fusion protein with another protein.

20 [0029]

In cases where eukaryotic cells are used as the host cells, an expression vector for eukaryotic cells having a promoter, splicing site, poly(A) addition site and the like is used as the expression vector. Examples of such an expression vector include
25 pKA1, pCDM8, pSVK3, pMSG, pSVL, pBK-CMV, pBK-RSV, the EBV vector, pRS, pcDNA3, pMSG and pYES2. In the same manner as described above, by incorporating DNA encoding the above-described polypeptide into such an expression vector and transforming eukaryotic host cells with the vector, followed by

culturing the obtained transformant, the polypeptide encoded by the above-described DNA can be expressed in the eukaryotic host cells. In cases where pIND/V5-His, pFLAG-CMV-2, pEGFP-N1 or pEGFP-C1 was used as the expression vector, the above-described polypeptide can be expressed as a fusion protein having various added tags such as His tag, FLAG tag, myc tag, HA tag or GFP.

[0030]

Introduction of the expression vector to the host cells can be carried out using a well-known method such as electroporation, the calcium phosphate method, the liposome method or the DEAE dextran method.

[0031]

Isolation and purification of a polypeptide of interest from the host cells can be carried out by a combination of known separation operations. Examples of the operations include, but are not limited to, treatment by a denaturant such as urea or by a surfactant; ultrasonication treatment; enzyme digestion; salting-out and solvent fractional precipitation; dialysis; centrifugation; ultrafiltration; gel filtration; SDS-PAGE; isoelectric focusing; ion-exchange chromatography; hydrophobic chromatography; affinity chromatography; and reversed-phase chromatography.

[0032]

The polypeptides obtained by the above method include, as mentioned above, those in the form of a fusion protein with another arbitrary protein. Examples thereof include fusion proteins with glutathion S-transferase (GST) and with a His tag. Such a polypeptide in the form of a fusion protein is also included within the scope of the present invention as the above-described polypeptide (c). Further, in some cases, a polypeptide expressed in a transformed cell is modified in various ways in the cell after translation thereof. Such a polypeptide having a post-translational modification is also included within the scope of the present invention as long as it has an immunity-inducing activity. Examples of such a post-

translational modification include elimination of N-terminus methionine, N-terminus acetylation, glycosylation, limited degradation by an intracellular protease, myristoylation, isoprenylation and phosphorylation.

[0033]

5 As described concretely in the following Examples, the above-described polypeptide having an immunity-inducing activity can cause regression of an already occurred tumor when administered to a cancer-bearing living body. Therefore, the immunity-inducing agent of the present invention can be used as a therapeutic and/or prophylactic agent for a cancer(s). In this case, cancers to be treated are those

10 expressing the gene encoding the polypeptide of SEQ ID NO:2 or 4, and examples thereof include, but are not limited to, brain tumor; squamous cell carcinomas of head, neck, lung, uterus and esophagus; melanoma; adenocarcinomas of lung, breast and uterus; renal cancer; malignant mixed tumor; hepatocellular carcinoma; basal cell carcinoma; acanthomatous epulis; intraoral tumor; perianal adenocarcinoma;

15 anal sac tumor; anal sac apocrine carcinoma; Sertoli cell tumor; vulva cancer; sebaceous adenocarcinoma; sebaceous epithelioma; sebaceous adenoma; sweat gland carcinoma; intranasal adenocarcinoma; nasal adenocarcinoma; thyroid cancer; colon cancer; bronchial adenocarcinoma; adenocarcinoma; ductal carcinoma; mammary adenocarcinoma; combined mammary adenocarcinoma; mammary gland malignant

20 mixed tumor; intraductal papillary adenocarcinoma; fibrosarcoma; hemangiopericytoma; osteosarcoma; chondrosarcoma; soft tissue sarcoma; histiocytic sarcoma; myxosarcoma; undifferentiated sarcoma; lung cancer; mastocytoma; cutaneous leiomyoma; intra-abdominal leiomyoma; leiomyoma;

25 chronic lymphocytic leukemia; lymphoma; gastrointestinal lymphoma; digestive organ lymphoma; small cell or medium cell lymphoma; adrenomedullary tumor; granulosa cell tumor; pheochromocytoma; bladder cancer (transitional cell carcinoma); suppurative inflammation; intra-abdominal liver tumor; liver cancer;

plasmacytoma; malignant hemangiopericytoma; angiosarcoma; anal sac
adenocarcinoma; oral cancer; metastatic malignant melanoma; amelanotic malignant
melanoma; cutaneous malignant melanoma; malignant myoepithelioma; malignant
seminoma; seminoma; adenocarcinoma of the large intestine; gastric
5 adenocarcinoma; low-grade sebaceous carcinoma; ceruminous adenocarcinoma;
apocrine carcinoma; poorly differentiated apocrine sweat gland carcinoma; malignant
fibrous histiocytoma; multiple myeloma; mesenchymal malignant tumor;
liposarcoma; osteosarcoma; sarcoma of unknown origin; soft part sarcoma (spindle
cell tumor); poorly differentiated sarcoma; synovial sarcoma; angiosarcoma;
10 metastatic malignant epithelioma; tubular mammary adenocarcinoma; mammary
ductal carcinoma; inflammatory breast cancer; germinoma; leukemia; invasive
trichoepithelioma; medium cell lymphoma; multicentric lymphoma; osteosarcoma
(mammary gland); mastocytoma (Patnaik II type); mastocytoma (Grade II); and
leiomyosarcoma. The animals to be treated are mammals, especially preferably
15 humans, dogs and cats.

[0034]

The administration route of the immunity-inducing agent of the present
invention to a living body may be either oral administration or parenteral
administration, and is preferably parenteral administration such as intramuscular
20 administration, subcutaneous administration, intravenous administration or
intraarterial administration. In cases where the immunity-inducing agent is used for
therapy of a cancer, it may be administered to a regional lymph node in the vicinity of
the tumor to be treated, as described in the Examples below, in order to enhance its
anticancer activity. The dose may be any dose as long as the dose is effective for
25 immune induction, and in cases where the agent is used for therapy and/or
prophylaxis of a cancer, the dose may be one effective for therapy and/or prophylaxis
of the cancer. The dose effective for therapy and/or prophylaxis of a cancer is

appropriately selected depending on the size of the tumor, the symptom and the like, and usually, 0.0001 μg to 1000 μg , preferably 0.001 μg to 1000 μg of the agent in terms of the effective ingredient may be administered once or in several times per day per animal to be treated. The agent is preferably administered in several times,
5 every several days to several months. As concretely shown in the Examples below, the immunity-inducing agent of the present invention can cause regression of an already occurred tumor. Therefore, since the agent can exert its anticancer activity also against a small number of cancer cells in the early stage, development or recurrence of the cancer can be prevented by using the agent before development of a
10 cancer or after therapy for a cancer. That is, the immunity-inducing agent of the present invention is effective for both therapy and prophylaxis of a cancer.

[0035]

The immunity-inducing agent of the present invention may contain only a polypeptide or may be formulated by mixing as appropriate with an additive such as a
15 pharmaceutically acceptable carrier, diluent or vehicle suitable for each administration mode. Formulation methods and additives which may be used are well-known in the field of formulation of pharmaceuticals, and any of the methods and additives may be used. Specific examples of the additive include, but are not limited to, diluents such as physiological buffer solutions; vehicles such as sucrose,
20 lactose, corn starch, calcium phosphate, sorbitol and glycine; binders such as syrup, gelatin, gum arabic, sorbitol, polyvinyl chloride and tragacanth; and lubricants such as magnesium stearate, polyethylene glycol, talc and silica. Examples of the formulation include oral preparations such as tablets, capsules, granules, powders and syrups; and parenteral preparations such as inhalants, injection solutions,
25 suppositories and solutions. These formulations may be prepared by commonly known production methods.

[0036]

The immunity-inducing agent of the present invention may be used in combination with an immunoenhancer capable of enhancing the immune response in a living body. The immunoenhancer may be contained in the immunity-inducing agent of the present invention or administered as a separate composition to a patient
5 in combination with the immunity-inducing agent of the present invention.

[0037]

Examples of the above-described immunoenhancer include adjuvants. Adjuvants can enhance the immune response by providing a reservoir of antigen (extracellularly or within macrophages), activating macrophages and stimulating
10 specific sets of lymphocytes, thereby enhancing the anticancer activity. Therefore, especially in cases where the immunity-inducing agent of the present invention is used for therapy and/or prophylaxis of a cancer, the immunity-inducing agent preferably comprises an adjuvant, in addition to the above-described polypeptide as an effective ingredient. Many types of adjuvants are well-known in the art, and any
15 of these adjuvants may be used. Specific examples of the adjuvants include MPL (SmithKline Beecham) and homologues of *Salmonella minnesota* Re 595 lipopolysaccharide obtained after purification and acid hydrolysis of the lipopolysaccharide; QS21 (SmithKline Beecham), pure QA-21 saponin purified from extract of *Quillja saponaria*; DQS21 described in WO96/33739 (SmithKline
20 Beecham); QS-7, QS-17, QS-18 and QS-L1 (So and 10 others, "Molecules and cells", 1997, Vol. 7, p. 178-186); Freund's incomplete adjuvant; Freund's complete adjuvant; vitamin E; Montanide; alum; CpG oligonucleotides (see, for example, Kreig and 7 others, "Nature", Vol. 374, p. 546-549); poly-I:C and derivatives thereof (e.g., poly ICLC); and various water in oil emulsions prepared from biodegradable oils such as
25 squalene and/or tocopherol. Among these, Freund's incomplete adjuvant; Montanide; poly-I:C and derivatives thereof, and CpG oligonucleotides are preferred. The mixing ratio between the above-described adjuvant and polypeptide is typically

about 1:10 to 10:1, preferably about 1:5 to 5:1, more preferably about 1:1. However, the adjuvant is not limited to the above-described examples, and adjuvants known in the art other than the above-described ones (for example, see Goding, "Monoclonal Antibodies: Principles and Practice", 2nd edition, 1986) may be used when the
5 immunity-inducing agent of the present invention is administered. Preparation methods for mixtures or emulsions of a polypeptide and an adjuvant are well-known to those skilled in the art of vaccination.

[0038]

Further, in addition to the above-described adjuvants, factors that stimulate
10 the immune response of the subject may be used as the above-described immunoenhancer. For example, various cytokines having a property to stimulate lymphocytes and/or antigen-presenting cells may be used as the immunoenhancer in combination with the immunity-inducing agent of the present invention. A number of such cytokines capable of enhancing the immune response are known to those
15 skilled in the art, and examples thereof include, but are not limited to, interleukin-12 (IL-12), GM-CSF, IL-18, interferon- α , interferon- β , interferon- ω , interferon- γ , and Flt3 ligand, which have been shown to promote the prophylactic action of vaccines. Such factors may also be used as the above-described immunoenhancer, and can be contained in the immunity-inducing agent of the present invention, or can be
20 prepared as a separate composition to be administered to a patient in combination with the immunity-inducing agent of the present invention.

[0039]

Further, by bringing the above-described polypeptide into contact with antigen-presenting cells *in vitro*, the antigen-presenting cells can be made to present
25 the polypeptide. That is, the above-described polypeptides (a) to (c) can be used as agents for treating antigen-presenting cells. As the antigen-presenting cells, dendritic cells or B cells, which have MHC class I molecules, may preferably be

employed. Various MHC class I molecules have been identified and well-known. MHC molecules in human are called HLA. Examples of HLA class I molecules include HLA-A, HLA-B and HLA-C, more specifically, HLA-A1, HLA-A0201, HLA-A0204, HLA-A0205, HLA-A0206, HLA-A0207, HLA-A11, HLA-A24, HLA-A31, HLA-A6801, HLA-B7, HLA-B8, HLA-B2705, HLA-B37, HLA-Cw0401 and HLA-Cw0602.

[0040]

The dendritic cells or B cells having MHC class I molecules can be prepared from peripheral blood by a well-known method. For example, tumor-specific dendritic cells can be induced by inducing dendritic cells from bone marrow, umbilical cord blood or patient's peripheral blood using granulocyte-macrophage colony-stimulating factor (GM-CSF) and IL-3 (or IL-4), and then adding a tumor-related peptide to the culture system. By administering an effective amount of such dendritic cells, a response desired for therapy of a cancer can be induced. As the cells to be used, bone marrow or umbilical cord blood donated by a healthy individual, or bone marrow, peripheral blood or the like from the patient himself may be used. When autologous cells of the patient are used, high safety can be attained and serious side effects are expected to be avoided. The peripheral blood or bone marrow may be a fresh sample, cold-stored sample or frozen sample. As for the peripheral blood, whole blood may be cultured or the leukocyte components alone may be separated and cultured, and the latter is efficient and thus preferred. Further, among the leukocyte components, mononuclear cells may be separated. In cases where the cells are originated from bone marrow or umbilical cord blood, the whole cells constituting the bone marrow may be cultured, or mononuclear cells may be separated therefrom and cultured. Peripheral blood, the leukocyte components thereof and bone marrow cells contain mononuclear cells, hematopoietic stem cells and immature dendritic cells, from which dendritic cells are originated, and also

CD4-positive cells and the like. As for the cytokine to be used, the production method thereof is not restricted and naturally-occurring or recombinant cytokine or the like may be employed as long as its safety and physiological activity have been confirmed. Preferably, a preparation with assured quality for medical use is used in
5 a minimum necessary amount. The concentration of the cytokine(s) to be added is not restricted as long as the dendritic cells are induced, and usually, the total concentration of the cytokine(s) is preferably about 10 to 1000 ng/mL, more preferably about 20 to 500 ng/mL. The culture may be carried out using a well-known medium usually used for the culture of leukocytes. The culturing
10 temperature is not restricted as long as the proliferation of the leukocytes is attained, and about 37°C which is the body temperature of human is most preferred. The atmospheric environment during the culturing is not restricted as long as the proliferation of the leukocytes is attained, and to flow 5% CO₂ is preferred. The culturing period is not restricted as long as the necessary number of the cells is
15 induced, and is usually 3 days to 2 weeks. As for the apparatuses used for separation and culturing of the cells, appropriate apparatuses, preferably those whose safety when applied to medical uses have been confirmed, and whose operations are stable and simple, may be employed. Particularly, as for the cell-culturing apparatus, not only the general vessels such as a Petri dish, flask and bottle, but also a
20 layer type vessel, multistage vessel, roller bottle, spinner type bottle, bag type culturing vessel, hollow fiber column and the like may be used.

[0041]

Bringing the above-described peptide of the present invention into contact with the antigen presenting cells *in vitro* may be carried out by a well-known method.
25 For example, it may be carried out by culturing the antigen-presenting cells in a culture medium containing the above-described polypeptide. The concentration of the peptide in the medium is not restricted, and usually about 1 µg/ml to 100 µg/ml,

preferably about 5 $\mu\text{g/ml}$ to 20 $\mu\text{g/ml}$. The cell density during the culturing is not restricted and is usually about 10^3 cells/ml to 10^7 cells/ml, preferably about 5×10^4 cells/ml to 5×10^6 cells/ml. The culturing may be carried out according to a conventional method, and is preferably carried out at 37°C under atmosphere of 5% CO_2 . The maximum length of the peptide which can be presented on the surface of the antigen-presenting cells is usually about 30 amino acid residues. Therefore, in cases where the antigen-presenting cells are brought into contact with the polypeptide *in vitro*, the polypeptide may be prepared such that its length is not more than about 30 amino acid residues.

10 [0042]

By culturing the antigen-presenting cells in the coexistence of the above-described polypeptide, the polypeptide is incorporated into MHC molecules of the antigen-presenting cells and presented on the surface of the antigen-presenting cells. Therefore, using the above-described polypeptide, isolated antigen-presenting cells containing the complex between the polypeptide and the MHC molecule can be prepared. Such antigen-presenting cells can present the polypeptide against T cells *in vivo* or *in vitro*, and induce, and allow proliferation of, cytotoxic T cells specific to the polypeptide.

[0043]

20 By bringing the antigen-presenting cells prepared as described above having the complex between the above-described polypeptide and the MHC molecule into contact with T cells *in vitro*, cytotoxic T cells specific to the polypeptide can be induced and allowed to proliferate. This may be carried out by cocultivating the above-described antigen-presenting cells and T cells in a liquid medium. For
25 example, it may be attained by suspending the antigen-presenting cells in a liquid medium, placing the suspension in vessels such as wells of a microplate, adding thereto T cells and then culturing the cells. The mixing ratio of the antigen-

presenting cells to the T cells in the cocultivation is not restricted, and is usually about 1:1 to 1:100, preferably about 1:5 to 1:20 in terms of the number of cells. The density of the antigen-presenting cells suspended in the liquid medium is not restricted, and is usually about 100 to 10,000,000 cells/ml, preferably about 10,000 to 1,000,000 cells/ml. The cocultivation is preferably carried out at 37°C under atmosphere of 5% CO₂ in accordance with a conventional method. The culturing time is not restricted, and is usually 2 days to 3 weeks, preferably about 4 days to 2 weeks. The cocultivation is preferably carried out in the presence of one or more interleukins such as IL-2, IL-6, IL-7 and IL-12. In this case, the concentration of IL-2 and IL-7 is usually about 5 U/ml to 20 U/ml, the concentration of IL-6 is usually about 500 U/ml to 2000 U/ml, and the concentration of IL-12 is usually about 5 ng/ml to 20 ng/ml, but the concentrations of the interleukins are not restricted thereto. The above-described cocultivation may be repeated once to several times adding fresh antigen-presenting cells. For example, the operation of discarding the culture supernatant after the cocultivation and adding a fresh suspension of antigen-presenting cells to further conduct the cocultivation may be repeated once to several times. The conditions of the each cocultivation may be the same as described above.

[0044]

By the above-described cocultivation, cytotoxic T cells specific to the polypeptide are induced and allowed to proliferate. Thus, using the above-described polypeptide, isolated T cells can be prepared which selectively bind the complex between the polypeptide and the MHC molecule.

[0045]

As described in the Examples below, the genes encoding the polypeptides of SEQ ID NOs:2, 16, 26, 28 and 39 and SEQ ID NOs:4, 18, 30 and 41 are expressed specifically in cancer cells and testis of dogs and humans, respectively. Thus, in cancer cells, significantly higher numbers of the polypeptides of SEQ ID NOs:2, 16,

26, 28 and 39 or SEQ ID NOs:4, 18, 30 and 41 exist than in normal cells. When cytotoxic T cells prepared as described above are administered to a living body while a part of the polypeptides existing in cancer cells are presented by MHC molecules on the surfaces of the cancer cells, the cytotoxic T cells can damage the cancer cells using the presented polypeptides as markers. Since antigen-presenting cells presenting the above-described polypeptides can induce, and allow proliferation of, cytotoxic T cells specific to the polypeptides also *in vivo*, cancer cells can be damaged also by administering the antigen-presenting cells to a living body. That is, the above-described cytotoxic T cells and the above-described antigen-presenting cells prepared using the above-described polypeptide are also effective as therapeutic and/or prophylactic agents for a cancer(s).

[0046]

In cases where the above-described isolated antigen-presenting cells or isolated T cells are administered to a living body, these are preferably prepared by treating antigen presenting cells or T cells collected from the patient to be treated with the polypeptide (a) to (c) as described above in order to avoid the immune response in the living body that attacks these cells as foreign bodies.

[0047]

The therapeutic and/or prophylactic agent for a cancer(s) comprising as an effective ingredient the antigen-presenting cells or T cells is preferably administered via a parenteral administration route such as intravenous or intraarterial administration. The dose is appropriately selected depending on the symptom, the purpose of administration and the like, and is usually 1 cell to 10,000,000,000,000 cells, preferably 1,000,000 cells to 1,000,000,000 cells, which dose is preferably administered once per several days to once per several months. The formulation may be, for example, the cells suspended in physiological buffered saline, and the formulation may be used in combination with another/other anticancer preparation(s)

and/or cytokine(s). Further, one or more additives well-known in the field of formulation of pharmaceuticals may also be added.

[0048]

Also by expression of the polynucleotide encoding the above-described polypeptide (a) to (c) in the body of the animal to be treated, antibody production and cytotoxic T cells can be induced in the living body, and an effect comparable to the administration of a polypeptide can be obtained. That is, the immunity-inducing agent of the present invention may be one comprising as an effective ingredient a recombinant vector having a polynucleotide encoding the above-described polynucleotide (a) to (c), which recombinant vector is capable of expressing the polypeptide in a living body. Such a recombinant vector capable of expressing an antigenic polypeptide is also called gene vaccine. The vector used for production of a gene vaccine is not restricted as long as it is a vector capable of expressing a polypeptide in cells of the animal to be treated (preferably in a mammalian cell), and may be either a plasmid vector or a virus vector, and any known vector in the field of gene vaccines may be used. The polynucleotide such as DNA or RNA encoding the above-described polypeptide can be easily prepared, as mentioned above, by a conventional method. Incorporation of the polynucleotide into a vector can be carried out using a method well-known to those skilled in the art.

20 [0049]

The administration route of the gene vaccine is preferably a parenteral route such as intramuscular, subcutaneous, intravenous or intraarterial administration, and the dose may be appropriately selected depending on the type of the antigen and the like, and usually about 0.1 μg to 100 mg, preferably about 1 μg to 10 mg in terms of the weight of the gene vaccine per 1 kg of body weight.

25 [0050]

Methods using a virus vector include those wherein a polynucleotide

encoding the above-described polypeptide is incorporated into an RNA virus or DNA virus such as retrovirus, adenovirus, adeno-associated virus, herpes virus, vaccinia virus, pox virus, poliovirus or Sindbis virus, and then the animal to be treated is infected by the resulting virus. Among these methods, those using retrovirus, adenovirus, adeno-associated virus, vaccinia virus or the like are especially preferred.

[0051]

Other methods include a method wherein an expression plasmid is directly intramuscularly administered (DNA vaccine method), the liposome method, lipofectin method, microinjection method, calcium phosphate method, electroporation method and the like, and the DNA vaccine method and liposome method are especially preferred.

[0052]

Methods for actually making the gene encoding the above-described polypeptide of the present invention act as a pharmaceutical include the *in vivo* method wherein the gene is directly introduced into the body, and the *ex vivo* method wherein a kind of cells are collected from the animal to be treated, the gene is introduced into the cells *ex vivo*, and then the cells are returned to the body (Nikkei Science, 1994, April, p. 20-45; The Pharmaceutical Monthly, 1994, Vol. 36, No. 1, p. 23-48; Experimental Medicine, Extra Edition, 1994, Vol.12, No. 15; and references cited in these papers and the like). The *in vivo* method is more preferred.

[0053]

In cases where the gene is administered by the *in vivo* method, the gene may be administered through an appropriate administration route depending on the disease to be treated, symptom and so on. It may be administered, for example, by intravenous, intraarterial, subcutaneous, intramuscular administration or the like. In cases where the gene is administered by the *in vivo* method, the gene may be formulated into a preparation such as a solution, and in general, it is formulated into

an injection solution or the like containing the DNA encoding the above-described peptide of the present invention as an effective ingredient. A commonly used carrier(s) may be added as required. In the case of a liposome or membrane fusion liposome (Sendai virus (HVJ)-liposome or the like) containing the DNA, the liposome may be formulated into a liposome preparation such as a suspension, frozen preparation or centrifugally concentrated frozen preparation.

[0054]

In the present invention, "the base sequence shown in SEQ ID NO:1" includes not only the base sequence expressly written in SEQ ID NO:1, but also the sequence complementary thereto. Thus, "a polynucleotide having the base sequence shown in SEQ ID NO:1" includes a single-stranded polynucleotide having the base sequence expressly written in SEQ ID NO:1, a single-stranded polynucleotide having the base sequence complementary thereto, and a double-stranded polynucleotide composed of these single strand polynucleotides. When the polynucleotide encoding the polypeptide used in the present invention is prepared, any one of these base sequences should be appropriately selected, and those skilled in the art can easily carry out the selection.

EXAMPLES

[0055]

The present invention will now be described more concretely by way of Examples.

[0056]

Example A-1: Acquisition of Novel Cancer Antigen Protein by SEREX Method

(1) Preparation of cDNA Library

Total RNA was prepared from testis tissue of a healthy dog by the Acid guanidium-Phenol-Chloroform method, and poly(A) RNA was purified using Oligotex-dT30 mRNA purification Kit (manufactured by Takara Shuzo Co., Ltd.) in

accordance with the protocol attached to the kit.

[0057]

Using the obtained mRNA (5 μ g), a dog testis cDNA phage library was synthesized. Preparation of the cDNA phage library was carried out using cDNA
5 Synthesis Kit, ZAP-cDNA Synthesis Kit, and ZAP-cDNA Gigapack III Gold Cloning Kit (manufactured by STRATAGENE) in accordance with the protocols attached to the kits. The size of the prepared cDNA phage library was 1.3×10^6 pfu/ml.

[0058]

(2) Screening of cDNA Library with Serum

10 Using the dog testis-derived cDNA phage library prepared as described above, immunoscreening was carried out. More particularly, host *E. coli* cells (XL1-Blue MRF') were infected with the library such that 2,340 clones should appear on an NZY agarose plate having the size of $\Phi 90 \times 15$ mm, and cultured at 42°C for 3 to 4 hours to allow the phage to form plaques. The plate was covered with nitrocellulose
15 membrane (Hybond C Extra: manufactured by GE Healthcare Bio-Science) impregnated with IPTG (isopropyl- β -D-thiogalactoside) at 37°C for 4 hours to induce and express proteins, which were thus transferred to the membrane. Subsequently, the membrane was recovered and soaked in TBS (10 mM Tris-HCl, 150 mM NaCl; pH 7.5) containing 0.5% non-fat dry milk, followed by shaking it at 4°C overnight to
20 suppress non-specific reactions. This filter was allowed to react with 500-fold diluted canine patient serum at room temperature for 2 to 3 hours.

[0059]

As the above-described canine patient serum, serum collected from canine patients suffering from squamous cell carcinoma was used. The serum was stored
25 at -80°C and pretreated immediately before use. The method of the pretreatment of the serum was as follows. That is, host *E. coli* cells (XL1-Blue MRF') were infected with λ ZAP Express phage to which no foreign gene was inserted, and then cultured

on NZY plate medium at 37°C overnight. Subsequently, the buffer of 0.2 M NaHCO₃, pH 8.3 containing 0.5 M NaCl was added to the plate, and the plate was left to stand at 4°C for 15 hours, followed by collecting the supernatant as an *E. coli*/phage extract. Thereafter, the collected *E. coli*/phage extract was allowed to flow through an NHS column (manufactured by GE Healthcare Bio-Science) to immobilize proteins derived from the *E. coli*/phage thereon. The serum from the canine patients was allowed to flow through and react with this protein-immobilized column to remove antibodies adsorbed on *E. coli* and/or the phage. The serum fraction that passed through the column was 500-fold diluted with TBS containing 0.5% non-fat dry milk, and the resulting diluent was used as the material for the immunoscreening.

[0060]

The membrane on which the thus treated serum and the above-described fusion protein were blotted was washed 4 times with TBS-T (0.05% Tween 20/TBS), and allowed to react with goat anti-dog IgG (Goat anti Dog IgG-h+I HRP conjugated: manufactured by BETHYL Laboratories) 5000-fold diluted with TBS containing 0.5% non-fat dry milk as a secondary antibody at room temperature for 1 hour, followed by detection by the enzyme coloring reaction using the NBT/BCIP reaction solution (manufactured by Roche). Colonies at positions where a positive coloring reaction was observed were recovered from the NZY agarose plate having the size of $\Phi 90 \times 15$ mm, and dissolved in 500 μ l of SM buffer (100 mM NaCl, 10 mM MgClSO₄, 50 mM Tris-HCl, 0.01% gelatin; pH 7.5). The screening was repeated as a second and third screening in the same manner as described above until a single coloring reaction-positive colony was obtained, thereby isolating one positive clone after screening of 30,940 phage clones reactive with IgG in the serum.

[0061]

(3) Homology Search of Isolated Antigen Gene

To subject the single positive clone isolated by the above-described method to a base sequence analysis, an operation of conversion of the phage vector to a plasmid vector was carried out. More particularly, 200 μ l of a solution prepared to contain a host *E. coli* (XL1-Blue MRF') such that the absorbance OD₆₀₀ should be 1.0 was
5 mixed with 100 μ l of a purified phage solution and further with 1 μ l of ExAssist helper phage (manufactured by STRATAGENE), and the reaction was allowed to proceed at 37°C for 15 minutes. To the reaction mixture, 3 ml of LB medium was added, and the mixture was cultured at 37°C for 2.5 to 3 hours, followed by immediate incubation in a water bath at 70°C for 20 minutes. The mixture was then
10 centrifuged at 4°C at 1000 \times g for 15 minutes, and the supernatant was recovered as a phagemid solution. Subsequently, 200 μ l of a solution prepared to contain a phagemid host *E. coli* (SOLR) such that the absorbance OD₆₀₀ should be 1.0 was mixed with 10 μ l of a purified phage solution, and the reaction was allowed to proceed at 37°C for 15 minutes. Thereafter, 50 μ l of the reaction mixture was
15 plated on ampicillin (final concentration: 50 μ g/ml)-containing LB agar medium, and cultured at 37°C overnight. A single colony of transformed SOLR was recovered and cultured in ampicillin (final concentration: 50 μ g/ml)-containing LB medium at 37°C, followed by purification of plasmid DNA having an insert of interest using QIAGEN plasmid Miniprep Kit (manufactured by Qiagen).

20 [0062]

The purified plasmid was subjected to an analysis of the entire sequence of the insert by the primer walking method using the T3 primer described in SEQ ID NO:5 and the T7 primer described in SEQ ID NO:6. By this sequence analysis, the gene sequence described in SEQ ID NO:1 was obtained. Using the base sequence
25 and the amino acid sequence of this gene, homology search against known genes was carried out using a homology search program BLAST (<http://www.ncbi.nlm.nih.gov/BLAST/>). As a result, it was revealed that the

obtained gene is the gene (Accession No. XM_535343) encoding a protein (Accession No. XP_535343) whose function is unknown. The human homologous factor of this gene was the gene (Accession No. NM_152660) encoding a protein (Accession No. NP_689873) whose function is also unknown (homology: base sequence, 93%; amino acid sequence, 99%). The base sequence of the human homologous factor is shown in SEQ ID NO:3, and the amino acid sequence thereof is shown in SEQ ID NO:4.

[0063]

(4) Analysis of Expression in Each Tissue

The expression of the gene, which was obtained by the above-described method, in normal tissues and various cell lines of dog and human were investigated by the RT-PCR (Reverse Transcription-PCR) method. The reverse transcription reaction was carried out as follows. That is, total RNA was extracted from 50 to 100 mg of each tissue or 5 to 10×10^6 cells of each cell line using TRIZOL reagent (manufactured by Invitrogen) in accordance with the protocol attached to the kit. Using this total RNA, cDNA was synthesized by Superscript First-Strand Synthesis System for RT-PCR (manufactured by Invitrogen) in accordance with the protocol attached to the kit. As the cDNAs from human normal tissues (brain, hippocampus, testis, colon and placenta), Gene Pool cDNA (manufactured by Invitrogen), QUICK-Clone cDNA (manufactured by CLONTECH) and Large-Insert cDNA Library (manufactured by CLONTECH) were used. The PCR reactions were carried out as follows using primers (described in SEQ ID NOs:7 and 8) specific to the obtained canine gene and its human homologous gene. That is, respective reagents and the attached buffer were mixed such that the mixture should contain 0.25 μ l of the sample prepared by the reverse transcription reaction, 2 μ M each of the above primers, 0.2 mM each of dNTP and 0.65 U of ExTaq polymerase (manufactured by Takara Shuzo Co., Ltd.) in a total volume of 25 μ l, and the reaction was carried out

with 30 cycles of 94°C for 30 seconds, 55°C for 30 seconds and 72°C for 1 minute using Thermal Cycler (manufactured by BIO RAD). The gene-specific primers having the base sequences shown in the above-described SEQ ID NOs:7 and 8 were those which amplify the regions of the 87th to 606th bases of the base sequence of SEQ ID NO:1 and the 173rd to 695th bases of the base sequence of SEQ ID NO:3, and can be used for investigation of the expression of both the canine gene and its human homologous gene. As a control for comparison, primers (described in SEQ ID NOs:9 and 10) specific to GAPDH were used simultaneously. As a result, as shown in Fig. 1, strong expression of the obtained canine gene was observed in testis among the normal dog tissues, and on the other hand, strong expression was observed in the canine breast cancer cell line. Expression of the human homologous gene was confirmed, as is the case with the canine gene, only in testis among the human normal tissues, but the expression was detected in brain tumor, leukemia, breast cancer and lung cancer cells among human cancer cell lines. Thus, the human homologous gene was also confirmed to be specifically expressed in testis and cancer cells.

[0064]

In Fig. 1, reference numeral 1 in the ordinate indicates the expression pattern of the above identified gene, and reference numeral 2 indicates the expression pattern of the *GAPDH* gene as a control for comparison.

[0065]

Example A-2: Preparation of Novel Cancer Antigen Proteins

(1) Preparation of Recombinant Protein

Based on the gene of SEQ ID NO:1 obtained in Example A-1, a recombinant protein was prepared by the following method. Respective reagents and the attached buffer were mixed such that the mixture should contain 1 µl of the vector which was prepared from the phagemid solution obtained in Example A-1 and was

subjected to the sequence analysis, 0.4 μ M each of two kinds of primers having *Nde*I and *Xho*I restriction sites (described in SEQ ID NOs:11 and 12), 0.2 mM dNTP and 1.25 U of PrimeSTAR HS polymerase (manufactured by Takara Shuzo Co., Ltd.) in a total volume of 50 μ l, and PCR was carried out with 30 cycles of 98°C for 10
5 seconds, 55°C for 15 seconds and 72°C for 1 minute using Thermal Cycler (manufactured by BIO RAD). The above-described two kinds of primers were those which amplify the region encoding the entire amino acid sequence of SEQ ID NO:2. After the PCR, the amplified DNA was subjected to electrophoresis using 1% agarose gel, and a DNA fragment of about 930 bp was purified using QIAquick
10 Gel Extraction Kit (manufactured by QIAGEN).

[0066]

The purified DNA fragment was ligated into a cloning vector pCR-Blunt (manufactured by Invitrogen). *E. coli* was transformed with the resulting ligation product, and plasmids were recovered thereafter, followed by confirming, by
15 sequencing, that the amplified gene fragment matches the sequence of interest. The plasmid that matched the sequence of interest was treated with restriction enzymes *Nde*I and *Xho*I and purified using QIAquick Gel Extraction Kit, followed by inserting the gene sequence of interest into an expression vector for *E. coli*, pET16b (manufactured by Novagen) that had been treated with *Nde*I and *Xho*I. Usage of
20 this vector enables production of a His-tag fusion recombinant protein. *E. coli* for expression, BL21 (DE3), was transformed with this plasmid, and expression of the protein of interest was induced in *E. coli* with 1 mM IPTG.

[0067]

On the other hand, based on the gene of SEQ ID NO:3, a recombinant protein
25 of the human homologous gene was prepared by the following method. Respective reagents and the attached buffer were mixed such that the mixture should contain 1 μ l of the cDNA prepared in Example A-1 whose expression could be confirmed by

the RT-PCR method in various tissues/cells, 0.4 μ M each of two kinds of primers having *EcoRV* and *EcoRI* restriction sites (described in SEQ ID NOs:13 and 14), 0.2 mM dNTP and 1.25 U of PrimeSTAR HS polymerase (manufactured by Takara Shuzo Co., Ltd.) in a total volume of 50 μ l, and PCR was carried out with 30 cycles of 98°C for 10 seconds, 55°C for 15 seconds and 72°C for 1 minute using Thermal Cycler (manufactured by BIO RAD). The above-described two kinds of primers were those which amplify the region encoding the entire amino acid sequence of SEQ ID NO:4. After the PCR, the amplified DNA was subjected to electrophoresis using 1% agarose gel, and a DNA fragment of about 930 bp was purified using QIAquick Gel Extraction Kit (manufactured by QIAGEN).

[0068]

The purified DNA fragment was ligated into a cloning vector pCR-Blunt (manufactured by Invitrogen). *E. coli* was transformed with the resulting ligation product, and plasmids were recovered thereafter, followed by confirming, by sequencing, that the amplified gene fragment matches the sequence of interest. The plasmid that matched the sequence of interest was treated with restriction enzymes *EcoRV* and *EcoRI* and purified using QIAquick Gel Extraction Kit, followed by inserting the gene sequence of interest into an expression vector for *E. coli*, pET30a (manufactured by Novagen) that had been treated with *EcoRV* and *EcoRI*. Usage of this vector enables production of a His-tag fusion recombinant protein. *E. coli* for expression, BL21 (DE3), was transformed with this plasmid, and expression of the protein of interest was induced in *E. coli* with 1 mM IPTG.

[0069]

(2) Purification of Recombinant Protein

The above-obtained recombinant *E. coli* cells that expressed SEQ ID NO:1 and SEQ ID NO:3, respectively, were cultured in 100 μ g/ml ampicillin-containing LB medium at 37°C until the absorbance at 600 nm reached about 0.7, and then

isopropyl- β -D-1-thiogalactopyranoside was added thereto such that its final concentration should be 1 mM, followed by culturing them at 37°C for 4 hours. Subsequently, the cells were collected by centrifugation at 4,800 rpm for 10 minutes. The pellet of the cells was suspended in phosphate-buffered saline and further
5 subjected to centrifugation at 4,800 rpm for 10 minutes to wash the cells.

[0070]

The cells were suspended in 50 mM Tris-HCl buffer (pH 8.0) and subjected to sonication on ice. The sonicated solution of *E. coli* was centrifuged at 6,000 rpm for 20 minutes to obtain the supernatant as the soluble fraction and the precipitate as
10 the insoluble fraction.

[0071]

The insoluble fraction was suspended in 50 mM Tris-HCl buffer (pH 8.0) and centrifuged at 6,000 rpm for 15 minutes. This operation was repeated twice and an operation of removal of proteases was carried out.

15 [0072]

The residue was suspended in 6M guanidine hydrochloride, 0.15 M sodium chloride-containing 50 mM Tris-HCl buffer (pH 8.0), and the resulting suspension was left to stand at 4°C for 20 hours to denature proteins. Thereafter, the suspension was centrifuged at 6,000 rpm for 30 minutes, and the obtained soluble
20 fraction was placed in a nickel chelate column prepared by a conventional method (carrier: Chelating Sepharose (trademark) Fast Flow (GE Health Care); column volume: 5mL; equilibration buffer: 6M guanidine hydrochloride, 0.15 M sodium chloride-containing 50 mM Tris-HCl buffer (pH 8.0)), followed by leaving it to stand at 4°C overnight to allow adsorption to the nickel-chelated carrier. The supernatant
25 was recovered by centrifugation of this column carrier at 1,500 rpm for 5 minutes, and the column carrier was suspended in phosphate-buffered saline, followed by refilling the column with the resulting suspension.

[0073]

The fraction that was not adsorbed to the column was washed away with 10 column volumes of 0.5 M sodium chloride-containing 0.1 M acetate buffer (pH 4.0), and elution was immediately carried out with 0.5 M sodium chloride-containing 0.1 M acetate buffer (pH 3.0) to obtain a purified fraction, which was used as the material for administration tests thereafter. The proteins of interest in respective eluted fractions were confirmed by Coomassie staining carried out according to a conventional method. Among these, the canine protein of interest is shown in Fig. 2.

[0074]

The buffer contained in the purified preparation obtained by the above-described method was replaced with a reaction buffer (50mM Tris-HCl, 100mM NaCl, 5mM CaCl₂; pH8.0), and cleavage of His tag by Factor Xa protease and purification of the protein of interest were carried out, using FactorXa Cleavage Capture Kit (manufactured by Novagen), in accordance with the protocols attached to the kit. Subsequently, the buffer contained in 1.2 ml of the purified preparation obtained by the above-described method was replaced with physiological phosphate buffer (manufactured by Nissui Pharmaceutical) by ultrafiltration using NANOSEP 10K OMEGA (manufactured by PALL), and the resulting solution was filtered aseptically using HT Tuffryn Acrodisc 0.22 μm (manufactured by PALL) and used in the following experiments.

[0075]

Example A-3: Test of Administration of Recombinant Protein to Cancer-bearing Dogs

(1) Antitumor Assay

The anti-tumor effect of the two kinds of recombinant proteins which were purified as described above was assessed in two individuals of cancer-bearing dogs having epidermal tumor (2 individuals having mammary gland tumor).

[0076]

An equal amount of Freund's incomplete adjuvant (manufactured by Wako Pure Chemicals) was mixed with 100 μg (0.5 ml) of the recombinant polypeptides (derived from dog and human), respectively, to prepare two kinds of therapeutic agents for a cancer(s). Each of these agents was administered to a regional lymph node in the vicinity of the tumor a total of 3 times, by carrying out the subsequent administrations 3 days and 7 days after the first administration. As a result, the tumors with a size of about 25 mm^3 and 50 mm^3 at the time of administration of the therapeutic agents for a cancer(s) (derived from dog and human), respectively, were reduced in size to 20 mm^3 and 42 mm^3 , respectively, 10 days after the first administration; 13 mm^3 and 26 mm^3 , respectively, 20 days after the first administration; and to 5 mm^3 and 10 mm^3 , respectively, 30 days after the first administration.

[0077]

Further, to a canine patient suffering from malignant melanoma, a mixture of 100 μg (0.5 ml) of the above-described polypeptide derived from dog and 0.5 ml of Freund's incomplete adjuvant was administered intracutaneously at the periphery of the tumor a total of 3 times at the same intervals as described above. Further, concurrently with the respective administrations, 10 MU of "Intercat" which is a recombinant feline interferon was administered subcutaneously. As a result, the tumor with a size of about 142 mm^3 at the time of administration of the therapeutic agent for a cancer(s) completely regressed 29 days after the first administration.

[0078]

Further, to a canine patient suffering from nasal adenocarcinoma, a mixture of 100 μg (0.5 ml) of the above-described polypeptide derived from dog and 0.5 ml of Freund's incomplete adjuvant was administered in the same manner as described above a total of 3 times. Further, concurrently with the respective administrations,

100 µg of canine interleukin 12 was administered subcutaneously. As a result, the tumor with a size of about 57 mm³ at the time of administration of the therapeutic agent for a cancer(s) completely regressed 14 days after the first administration.

[0079]

5 (2) Immune Inducibility Assay

Blood from the canine patient in which the anti-tumor effect was obtained in the administration test in the above-described (1) was collected before administration of the therapeutic agent for a cancer(s), and 10 days and 30 days after the first administration. Peripheral blood mononuclear cells were isolated according to a
10 conventional method, and by the ELISPOT assay for IFN γ using it, the immune inducibility of each administered recombinant protein was assayed.

[0080]

In a 96-well plate manufactured by Millipore (MultiScreen-IP, MAIPS 4510), 100 µL/well of 70% ethanol was placed and the plate was left to stand for 5 minutes,
15 followed by removal of the ethanol by aspiration. The plate was washed with sterile water and 300 µl/well of 200 mM Sodium Bicarbonate (pH8.2) was placed therein. After leaving it to stand for 5 minutes, Sodium Bicarbonate was removed by aspiration, and then the plate was washed. Subsequently, 0.5 µl/well of anti-canine interferon γ monoclonal antibody (manufactured by R&D, clone 142529, MAB781)
20 mixed with 200 mM Sodium Bicarbonate was placed in wells, and the plate was incubated at 37°C overnight to immobilize the primary antibody. After removal of the primary antibody by aspiration, 300 µL/well of a blocking solution (1% BSA-5% sucrose-200 mM Sodium Bicarbonate (pH8.2)) was added to the wells, and the plate was incubated at 4°C overnight to block the plate. After removal of the blocking
25 solution by aspiration, 300 µL/well of 10% fetal calf serum-containing RPMI medium (manufactured by Invitrogen) was placed in the wells, and the plate was left to stand for 5 minutes, followed by removal of the medium by aspiration.

Subsequently, 5×10^5 cells/well of the canine peripheral blood mononuclear cells suspended in 10% fetal calf serum-containing RPMI medium were placed in the plate, and 10 μ L/well of the canine-derived polypeptide or human-derived polypeptide used in each administration was added thereto, followed by culturing the cells under the conditions of 37°C and 5% CO₂ for 24 hours, to allow immunocytes that might exist in the peripheral blood mononuclear cells to produce interferon γ . After the culture, the medium was removed, and the wells were washed 6 times with a washing solution (0.1% Tween20-200mM Sodium Bicarbonate (pH8.2)). In each well, 100 μ L of rabbit anti-dog polyclonal antibody 1000-fold diluted with the above-described blocking solution was placed, and the plate was incubated at 4°C overnight. After washing the wells 3 times with the above-described washing solution, 100 μ L of HRP-labeled anti-rabbit antibody 1000-fold diluted with the above-described blocking solution was placed in each well, and the reaction was allowed to proceed at 37°C for 2 hours. After washing the wells 3 times with the above-described washing solution, the resultant was colored with Konica Immunostain (manufactured by Konica), and the wells were washed with water to stop the reaction. Thereafter, the membrane was dried, and the number of the appeared spots was counted using KS ELISPOT (manufactured by Carl Zeiss, Inc.).

[0081]

As a result, in either canine patient to which the canine polypeptide or the human polypeptide was administered, peripheral blood mononuclear cells sampled before the administration of the polypeptide showed no spots. On the other hand, in the canine patient to which the canine polypeptide was administered, peripheral blood mononuclear cells sampled 10 days and 30 days after the administration showed 20 and 36 spots, respectively. In the canine patient to which the human polypeptide was administered, peripheral blood mononuclear cells sampled 10 days and 30 days after the administration showed 24 and 36 spots, respectively.

[0082]

From the above results, it is confirmed that immunocytes which specifically react with the administered recombinant protein and produce interferon γ were induced in all of the canine patients to which the recombinant protein was administered, and it is thought that the anti-tumor effect described in (1) was exerted by immunoreactions in which these immunocytes are mainly involved.

[0083]

Example B-1: Acquisition of Novel Cancer Antigen Protein by SEREX Method

(1) Preparation of cDNA Library

Total RNA was prepared from testis tissue of a healthy dog by the Acid guanidium-Phenol-Chloroform method, and poly(A) RNA was purified using Oligotex-dT30 mRNA purification Kit (manufactured by Takara Shuzo Co., Ltd.) in accordance with the protocol attached to the kit.

[0084]

Using the obtained mRNA (5 μg), a dog testis cDNA phage library was synthesized. Preparation of the cDNA phage library was carried out using cDNA Synthesis Kit, ZAP-cDNA Synthesis Kit, and ZAP-cDNA Gigapack III Gold Cloning Kit (manufactured by STRATAGENE) in accordance with the protocols attached to the kits. The size of the prepared cDNA phage library was 1.3×10^6 pfu/ml.

[0085]

(2) Screening of cDNA Library with Serum

Using the dog testis-derived cDNA phage library prepared as described above, immunoscreening was carried out. More particularly, host *E. coli* cells (XL1-Blue MRF') were infected with the library such that 2,340 clones should appear on an NZY agarose plate having the size of $\Phi 90 \times 15$ mm, and cultured at 42°C for 3 to 4 hours to allow the phage to form plaques. The plate was covered with nitrocellulose membrane (Hybond C Extra: manufactured by GE Healthcare Bio-Science)

impregnated with IPTG (isopropyl- β -D-thiogalactoside) at 37°C for 4 hours to induce and express proteins, which were thus transferred to the membrane. Subsequently, the membrane was recovered and soaked in TBS (10 mM Tris-HCl, 150 mM NaCl; pH 7.5) containing 0.5% non-fat dry milk, followed by shaking at 4°C overnight to suppress non-specific reactions. This filter was allowed to react with 500-fold diluted canine patient serum at room temperature for 2 to 3 hours.

[0086]

As the above-described canine patient serum, serum collected from canine patients suffering from tumor proximal to the anus was used. The serum was stored at -80°C and pretreated immediately before use. The method of the pretreatment of the serum was as follows. That is, host *E. coli* cells (XL1-Blue MRF') were infected with λ ZAP Express phage to which no foreign gene was inserted, and then cultured on NZY plate medium at 37°C overnight. Subsequently, the buffer of 0.2 M NaHCO₃, pH 8.3 containing 0.5 M NaCl was added to the plate, and the plate was left to stand at 4°C for 15 hours, followed by collecting the supernatant as an *E. coli*/phage extract. Thereafter, the collected *E. coli*/phage extract was allowed to flow through an NHS column (manufactured by GE Healthcare Bio-Science) to immobilize proteins derived from the *E. coli*/phage thereon. The serum from the canine patients was allowed to flow through and react with this protein-immobilized column to remove antibodies adsorbed on *E. coli* and/or the phage. The serum fraction that passed through the column was 500-fold diluted with TBS containing 0.5% non-fat dry milk, and the resulting diluent was used as the material for the immunoscreening.

[0087]

The membrane on which the thus treated serum and the above-described fusion protein were blotted was washed 4 times with TBS-T (0.05% Tween 20/TBS), and allowed to react with goat anti-dog IgG (Goat anti Dog IgG-h+I HRP conjugated:

manufactured by BETHYL Laboratories) 5000-fold diluted with TBS containing 0.5% non-fat dry milk as a secondary antibody at room temperature for 1 hour, followed by detection by the enzyme coloring reaction using the NBT/BCIP reaction solution (manufactured by Roche). Colonies at positions where a positive coloring reaction was observed were recovered from the NZY agarose plate having the size of $\Phi 90 \times 15$ mm, and dissolved in 500 μ l of SM buffer (100 mM NaCl, 10 mM MgClSO₄, 50 mM Tris-HCl, 0.01% gelatin; pH 7.5). The screening was repeated as a second and third screening in the same manner as described above until a single coloring reaction-positive colony was obtained, thereby isolating one positive clone after screening of 30,940 phage clones reactive with IgG in the serum.

[0088]

(3) Homology Search of Isolated Antigen Gene

To subject the single positive clone isolated by the above-described method to a base sequence analysis, an operation of conversion of the phage vector to a plasmid vector was carried out. More particularly, 200 μ l of a solution prepared to contain a host *E. coli* (XL1-Blue MRF') such that the absorbance OD₆₀₀ should be 1.0 was mixed with 100 μ l of a purified phage solution and further with 1 μ l of ExAssist helper phage (manufactured by STRATAGENE), and the reaction was allowed to proceed at 37°C for 15 minutes. To the reaction mixture, 3 ml of LB medium was added, and the mixture was cultured at 37°C for 2.5 to 3 hours, followed by immediate incubation in a water bath at 70°C for 20 minutes. The mixture was then centrifuged at 4°C at 1000 \times g for 15 minutes, and the supernatant was recovered as a phagemid solution. Subsequently, 200 μ l of a solution prepared to contain a phagemid host *E. coli* (SOLR) such that the absorbance OD₆₀₀ should be 1.0 was mixed with 10 μ l of a purified phage solution, and the reaction was allowed to proceed at 37°C for 15 minutes. Thereafter, 50 μ l of the reaction mixture was plated on ampicillin (final concentration: 50 μ g/ml)-containing LB agar medium, and

cultured at 37°C overnight. A single colony of transformed SOLR was recovered and cultured in ampicillin (final concentration: 50 µg/ml)-containing LB medium at 37°C, followed by purification of plasmid DNA having an insert of interest using QIAGEN plasmid Miniprep Kit (manufactured by Qiagen).

5 [0089]

The purified plasmid was subjected to an analysis of the entire sequence of the insert by the primer walking method using the T3 primer described in SEQ ID NO:5 and the T7 primer described in SEQ ID NO:6. By this sequence analysis, the gene sequence described in SEQ ID NO:15 was obtained. Using the base sequence and the amino acid sequence of this gene, homology search against known genes was carried out using a homology search program BLAST (10 <http://www.ncbi.nlm.nih.gov/BLAST/>). As a result, it was revealed that the obtained gene is the calmegin gene. The human homologous factor of the canine calmegin gene was human calmegin (homology: base sequence, 90%; amino acid sequence, 89%). The base sequence of human calmegin is shown in SEQ ID NO:17, and the amino acid sequence thereof is shown in SEQ ID NO:18.

15 [0090]

(4) Analysis of Expression in Each Tissue

The expression of the gene, which was obtained by the above-described method, in normal tissues and various cell lines of dog and human were investigated by the RT-PCR (Reverse Transcription-PCR) method. The reverse transcription reaction was carried out as follows. That is, total RNA was extracted from 50 to 100 mg of each tissue or 5 to 10×10^6 cells of each cell line using TRIZOL reagent (manufactured by Invitrogen) in accordance with the protocol attached to the kit. Using this total RNA, cDNA was synthesized by Superscript First-Strand Synthesis System for RT-PCR (manufactured by Invitrogen) in accordance with the protocol attached to the kit. As the cDNAs from human normal tissues (brain, hippocampus,

testis, colon and placenta), Gene Pool cDNA (manufactured by Invitrogen), QUICK-Clone cDNA (manufactured by CLONTECH) and Large-Insert cDNA Library (manufactured by CLONTECH) were used. The PCR reactions were carried out as follows using primers (described in SEQ ID NOs:19 and 20) specific to the obtained gene. That is, respective reagents and the attached buffer were mixed such that the mixture should contain 0.25 μ l of the sample prepared by the reverse transcription reaction, 2 μ M each of the above primers, 0.2 mM each of dNTP and 0.65 U of ExTaq polymerase (manufactured by Takara Shuzo Co., Ltd.) in a total volume of 25 μ l, and the reaction was carried out with 30 cycles of 94°C for 30 seconds, 55°C for 30 seconds and 72°C for 1 minute using Thermal Cycler (manufactured by BIO RAD). The above-described gene-specific primers were those which amplify the regions of the 755th to 1318th bases of the base sequence of SEQ ID NO:15 (canine calmegin gene) and the 795th to 1358th bases of the base sequence of SEQ ID NO:17 (human calmegin gene), and can be used for investigation of the expression of both the canine calmegin gene and the human calmegin gene. As a control for comparison, primers (described in SEQ ID NOs:9 and 10) specific to GAPDH were used simultaneously. As a result, as shown in Fig. 3, strong expression of the canine calmegin gene was observed in testis among the normal dog tissues, and on the other hand, strong expression was observed in canine tumor cell lines. Expression of the human calmegin gene was confirmed, as is the case with the canine calmegin gene, only in testis among the human normal tissues, but the expression was detected in brain tumor, leukemia and esophagus cancer cells among human cancer cell lines. Thus, the human calmegin gene was also confirmed to be specifically expressed in testis and cancer cells.

[0091]

In Fig. 3, reference numeral 1 in the ordinate indicates the expression pattern of the calmegin gene, and reference numeral 2 indicates the expression pattern of the

GAPDH gene as a control for comparison.

[0092]

Example B-2: Preparation of Canine and Human Calmegin Proteins

(1) Preparation of Recombinant Protein

5 Based on the gene of SEQ ID NO:15 obtained in Example B-1, a recombinant protein was prepared by the following method. Respective reagents and the attached buffer were mixed such that the mixture should contain 1 μ l of the vector that was prepared from the phagemid solution obtained in Example B-1 and was subjected to the sequence analysis, 0.4 μ M each of two kinds of primers having
10 *Bam*HI and *Eco*RI restriction sites (described in SEQ ID NOs:21 and 22), 0.2 mM dNTP and 1.25 U of PrimeSTAR HS polymerase (manufactured by Takara Shuzo Co., Ltd.) in a total volume of 50 μ l, and PCR was carried out with 30 cycles of 98°C for 10 seconds, 55°C for 15 seconds and 72°C for 2 minutes using Thermal Cycler (manufactured by BIO RAD). The above-described two kinds of primers were
15 those which amplify the region encoding the entire amino acid sequence of SEQ ID NO:16. After the PCR, the amplified DNA was subjected to electrophoresis using 1% agarose gel, and a DNA fragment of about 1.9 kbp was purified using QIAquick Gel Extraction Kit (manufactured by QIAGEN).

[0093]

20 The purified DNA fragment was ligated into a cloning vector pCR-Blunt (manufactured by Invitrogen). *E. coli* was transformed with the resulting ligation product, and plasmids were recovered thereafter, followed by confirming, by sequencing, that the amplified gene fragment matches the sequence of interest. The plasmid that matched the sequence of interest was treated with restriction enzymes
25 *Bam*HI and *Eco*RI and purified using QIAquick Gel Extraction Kit, followed by inserting the gene sequence of interest into an expression vector for *E. coli*, pET30a (manufactured by Novagen) that had been treated with *Bam*HI and *Eco*RI. Usage

of this vector enables production of a His-tag fusion recombinant protein. *E. coli* for expression, BL21 (DE3), was transformed with this plasmid, and expression of the protein of interest was induced in *E. coli* with 1 mM IPTG.

[0094]

5 On the other hand, based on the gene of SEQ ID NO:17, a recombinant protein of the human homologous gene was prepared by the following method. Respective reagents and the attached buffer were mixed such that the mixture should contain 1 μ l of the cDNA prepared in Example B-1 whose expression could be confirmed by the RT-PCR method in various tissues/cells, 0.4 μ M each of two kinds
10 of primers having *Eco*RI and *Xho*I restriction sites (described in SEQ ID NOs:23 and 24), 0.2 mM dNTP and 1.25 U of PrimeSTAR HS polymerase (manufactured by Takara Shuzo Co., Ltd.) in a total volume of 50 μ l, and PCR was carried out with 30 cycles of 98°C for 10 seconds, 55°C for 15 seconds and 72°C for 2 minutes using Thermal Cycler (manufactured by BIO RAD). The above-described two kinds of
15 primers were those which amplify the region encoding the entire amino acid sequence of SEQ ID NO:18. After the PCR, the amplified DNA was subjected to electrophoresis using 1% agarose gel, and a DNA fragment of about 1.9 kbp was purified using QIAquick Gel Extraction Kit (manufactured by QIAGEN).

[0095]

20 The purified DNA fragment was ligated into a cloning vector pCR-Blunt (manufactured by Invitrogen). *E. coli* was transformed with the resulting ligation product, and plasmids were recovered thereafter, followed by confirming, by sequencing, that the amplified gene fragment matches the sequence of interest. The plasmid that matched the sequence of interest was treated with restriction enzymes
25 *Eco*RI and *Xho*I and purified using QIAquick Gel Extraction Kit, followed by inserting the gene sequence of interest into an expression vector for *E. coli*, pET30a (manufactured by Novagen) that had been treated with *Eco*RI and *Xho*I. Usage of

this vector enables production of a His-tag fusion recombinant protein. *E. coli* for expression, BL21 (DE3), was transformed with this plasmid, and expression of the protein of interest was induced in *E. coli* with 1 mM IPTG.

[0096]

5 (2) Purification of Recombinant Protein

The above-obtained recombinant *E. coli* cells that expressed SEQ ID NO:15 and SEQ ID NO:17, respectively, were cultured in 30 µg/ml kanamycin-containing LB medium at 37°C until the absorbance at 600 nm reached about 0.7, and then isopropyl-β-D-1-thiogalactopyranoside was added thereto such that its final
10 concentration should be 1 mM, followed by culturing them at 37°C for 4 hours. Subsequently, the cells were collected by centrifugation at 4,800 rpm for 10 minutes. The pellet of the cells was suspended in phosphate-buffered saline and further subjected to centrifugation at 4,800 rpm for 10 minutes to wash the cells.

[0097]

15 The obtained pellet of *E. coli* cells was suspended in 20 mM phosphate buffer (pH 7.0) and subjected to sonication on ice. The sonicated solution of *E. coli* was centrifuged at 6,000 rpm for 20 minutes to obtain the supernatant as the soluble fraction and the precipitate as the insoluble fraction.

[0098]

20 The soluble fraction was placed in an ion-exchange column (carrier: SP Sepharose (trademark) Fast Flow (GE Health Care); column volume: 5mL; equilibration buffer: 20 mM phosphate buffer (pH 7.0)). The column was washed with 10 column volumes of 20 mM phosphate buffer (pH 7.0), and elution was carried out with density gradient of salt by 0.3 M-1.0 M sodium chloride-containing
25 20 mM phosphate buffer (pH 7.0). Six column volumes of the eluted fraction was collected in each elution step.

[0099]

Among these eluted fractions, the 1st to 6th fractions eluted with 0.3 M sodium chloride-containing 20 mM phosphate buffer (pH 7.0) and the 1st fraction eluted with 1.0 M sodium chloride-containing 20 mM phosphate buffer (pH 7.0) were combined, and the resulting solution was subjected to additional purification by a secondary column.

[0100]

For the secondary column, a column carrier Bio gel HT Type II (BioRad) was used. The column volume was 5 mL. The column was equilibrated with 10 column volumes of 0.3 M sodium chloride-containing 20 mM phosphate buffer (pH 7.0), and the above-described eluted fractions were placed in the column. The fractions that were not adsorbed to the column was washed away with 10 column volumes of 0.3 M sodium chloride-containing 20 mM phosphate buffer (pH 7.0) and 0.1 M phosphate buffer (pH 7.0), and elution was carried out with 0.2 M phosphate buffer (pH 7.0) to obtain a purified fraction, which was used as the material for administration tests thereafter. The proteins of interest in the eluted fractions were confirmed by Coomassie staining carried out according to a conventional method. Among these, the canine calmegin protein is shown in Fig. 4.

[0101]

To 1 ml of a reaction buffer (20 mM Tris-HCl, 50 mM NaCl, 2 mM CaCl₂; pH 7.4), 200 µl of the purified preparation obtained by the above-described method was aliquoted, and 2 µl of enterokinase (manufactured by Novagen) was then added thereto, followed by leaving it to stand at room temperature overnight to cleave His tag. The resulting product was purified using Enterokinase Cleavage Capture Kit (manufactured by Novagen) in accordance with the protocol attached to the kit. Subsequently, the buffer contained in 1.2 ml of the purified preparation obtained by the above-described method was replaced with physiological phosphate buffer (manufactured by Nissui Pharmaceutical) by ultrafiltration using NANOSEP 10K

OMEGA (manufactured by PALL), and the resulting solution was filtered aseptically using HT Tuffryn Acrodisc 0.22 μm (manufactured by PALL) and used in the following experiments.

[0102]

5 Example B-3: Test of Administration of Recombinant Protein to Cancer-bearing Dogs

(1) Antitumor Assay

The anti-tumor effect of the two kinds of recombinant proteins which were purified as described above was assessed in two individuals of cancer-bearing dogs having epidermal tumor (2 individuals having mammary gland tumor).

[0103]

An equal amount of Freund's incomplete adjuvant (manufactured by Wako Pure Chemicals) was mixed with 100 μg (0.5 ml) of the recombinant canine calmegin and human calmegin proteins, respectively, to prepare therapeutic agents for a cancer(s). Each of these agents was administered to a regional lymph node in the vicinity of the tumor a total of 3 times, by carrying out the subsequent administrations 3 days and 7 days after the first administration. As a result, the tumors with a size of about 45 mm^3 and 78 mm^3 , respectively, at the time of administration of the therapeutice agents were reduced to 27 mm^3 and 46 mm^3 , respectively, 10 days after the first administration; 15 mm^3 and 26 mm^3 , respectively, 20 days after the first administration; and to 7 mm^3 and 15 mm^3 , respectively, 30 days after the first administration.

[0104]

Further, to a canine patient suffering from malignant melanoma, a mixture of 100 μg (0.5 ml) of the above-described canine calmegin protein and 0.5 ml of Freund's incomplete adjuvant was administered a total of 3 times in the same manner as described above. Further, concurrently with the respective administrations, 100

µg of canine interleukin 12 was administered subcutaneously. As a result, the tumor with a size of about 38 mm³ at the time of administration of the therapeutic agent completely regressed 21 days after the first administration of the therapeutic agent.

5 [0105]

(2) Immune Inducibility Assay

Blood from the canine patient in which the anti-tumor effect was obtained in the administration test in the above-described (1) was collected before administration of the therapeutic agent for a cancer(s) and 10 days and 30 days after the first
10 administration. Peripheral blood mononuclear cells were isolated according to a conventional method, and by the ELISPOT assay for IFN γ using it, the immune inducibility of each administered recombinant protein was assayed.

[0106]

In a 96-well plate manufactured by Millipore (MultiScreen-IP, MAIPS 4510),
15 100 µL/well of 70% ethanol was placed and the plate was left to stand for 5 minutes, followed by removal of the ethanol by aspiration. The plate was washed with sterile water and 300 µl/well of 200 mM Sodium Bicarbonate (pH8.2) was placed therein. After leaving it to stand for 5 minutes, Sodium Bicarbonate was removed by aspiration, and then the plate was washed. Subsequently, 0.5 µg/well of anti-canine
20 interferon γ monoclonal antibody (manufactured by R&D, clone 142529, MAB781) mixed with 200 mM Sodium Bicarbonate was placed in wells, and the plate was incubated at 37°C overnight to immobilize the primary antibody. After removal of the primary antibody by aspiration, 300 µL/well of a blocking solution (1% BSA-5% sucrose-200 mM Sodium Bicarbonate (pH8.2)) was added to the wells, and the plate
25 was incubated at 4°C overnight to block the plate. After removal of the blocking solution by aspiration, 300 µL/well of 10% fetal calf serum-containing RPMI medium (manufactured by Invitrogen) was placed in the wells, and the plate was left

to stand for 5 minutes, followed by removal of the medium by aspiration.

Subsequently, 5×10^5 cells/well of the canine peripheral blood mononuclear cells suspended in 10% fetal calf serum-containing RPMI medium were placed in the plate, and 10 μ L/well of the canine calmegin or human calmegin protein used in each administration was added thereto, followed by culturing the cells under the conditions of 37°C and 5% CO₂ for 24 hours, to allow immunocytes that might exist in the peripheral blood mononuclear cells to produce interferon γ . After the culture, the medium was removed, and the wells were washed 6 times with a washing solution (0.1% Tween20-200mM Sodium Bicarbonate (pH8.2)). In each well, 100 μ L of rabbit anti-dog polyclonal antibody 1000-fold diluted with the above-described blocking solution was placed, and the resulting mixture was incubated at 4°C overnight. After washing the wells 3 times with the above-described washing solution, 100 μ L of HRP-labeled anti-rabbit antibody 1000-fold diluted with the above-described blocking solution was placed in each well, and the reaction was allowed to proceed at 37°C for 2 hours. After washing the wells 3 times with the above-described washing solution, the resultant was colored with Konica Immunostain (manufactured by Konica), and the wells were washed with water to stop the reaction. Thereafter, the membrane was dried, and image processing of the wells was carried out, followed by counting the number of spot-forming cells (SFC) using KS ELISPOT compact system (Carl Zeiss, Inc., Germany).

[0107]

As a result, in either canine patient to which canine calmegin or human calmegin was administered, peripheral blood mononuclear cells sampled before the administration showed no spots. On the other hand, in the canine patient to which canine calmegin was administered, peripheral blood mononuclear cells sampled 10 days and 30 days after the administration showed 15 and 45 spots, respectively. In the canine patient to which human calmegin was administered, peripheral blood

mononuclear cells sampled 10 days and 30 days after the administration showed 12 and 39 spots, respectively.

[0108]

From the above results, it is confirmed that immunocytes which specifically
5 react with the administered recombinant protein and produce interferon γ were induced in all of the canine patients to which the recombinant protein was administered, and it is thought that the anti-tumor effect described in (1) was exerted by immunoreactions in which these immunocytes are mainly involved.

[0109]

10 Example C-1: Acquisition of Novel Cancer Antigen Protein by SEREX Method

(1) Preparation of cDNA Library

Total RNA was prepared from testis tissue of a healthy dog by the Acid
guanidium-Phenol-Chloroform method, and poly(A) RNA was purified using
Oligotex-dT30 mRNA purification Kit (manufactured by Takara Shuzo Co., Ltd.) in
15 accordance with the protocol attached to the kit.

[0110]

Using the obtained mRNA (5 μ g), a dog testis cDNA phage library was
synthesized. Preparation of the cDNA phage library was carried out using cDNA
Synthesis Kit, ZAP-cDNA Synthesis Kit, and ZAP-cDNA Gigapack III Gold Cloning
20 Kit (manufactured by STRATAGENE) in accordance with the protocols attached to the kits. The size of the prepared cDNA phage library was 1.3×10^6 pfu/ml.

[0111]

(2) Screening of cDNA Library with Serum

Using the dog testis-derived cDNA phage library prepared as described above,
25 immunoscreening was carried out. More particularly, host *E. coli* cells (XL1-Blue MRF') were infected with the library such that 2,340 clones should appear on an NZY agarose plate having the size of $\Phi 90 \times 15$ mm, and cultured at 42°C for 3 to 4

hours to allow the phage to form plaques. The plate was covered with nitrocellulose membrane (Hybond C Extra: manufactured by GE Healthcare Bio-Science) impregnated with IPTG (isopropyl- β -D-thiogalactoside) at 37°C for 4 hours to induce and express proteins, which were thus transferred to the membrane. Subsequently, the membrane was recovered and soaked in TBS (10 mM Tris-HCl, 150 mM NaCl; pH 7.5) containing 0.5% non-fat dry milk, followed by shaking it at 4°C overnight to suppress non-specific reactions. This filter was allowed to react with 500-fold diluted canine patient serum at room temperature for 2 to 3 hours.

[0112]

As the above-described canine patient serum, serum collected from canine patients suffering from breast cancer was used. The serum was stored at -80°C and pretreated immediately before use. The method of the pretreatment of the serum was as follows. That is, host *E. coli* cells (XL1-Blue MRF') were infected with λ ZAP Express phage to which no foreign gene was inserted, and then cultured on NZY plate medium at 37°C overnight. Subsequently, the buffer of 0.2 M NaHCO₃, pH 8.3 containing 0.5 M NaCl was added to the plate, and the plate was left to stand at 4°C for 15 hours, followed by collecting the supernatant as an *E. coli*/phage extract. Thereafter, the collected *E. coli*/phage extract was allowed to flow through an NHS column (manufactured by GE Healthcare Bio-Science) to immobilize proteins derived from the *E. coli*/phage thereon. The serum from the canine patients was allowed to flow through and react with this protein-immobilized column to remove antibodies adsorbed on *E. coli* and/or the phage. The serum fraction that passed through the column was 500-fold diluted with TBS containing 0.5% non-fat dry milk, and the resulting diluent was used as the material for the immunoscreening.

[0113]

The membrane on which the thus treated serum and the above-described fusion protein were blotted was washed 4 times with TBS-T (0.05% Tween 20/TBS),

and allowed to react with goat anti-dog IgG (Goat anti Dog IgG-h+I HRP conjugated: manufactured by BETHYL Laboratories) 5,000-fold diluted with TBS containing 0.5% non-fat dry milk as a secondary antibody at room temperature for 1 hour, followed by detection by the enzyme coloring reaction using the NBT/BCIP reaction solution (manufactured by Roche). Colonies at positions where a positive coloring reaction was observed were recovered from the NZY agarose plate having the size of $\Phi 90 \times 15$ mm, and dissolved in 500 μ l of SM buffer (100 mM NaCl, 10 mM MgClSO₄, 50 mM Tris-HCl, 0.01% gelatin; pH 7.5). The screening was repeated as a second and third screening in the same manner as described above until a single coloring reaction-positive colony was obtained, thereby isolating one positive clone after screening of 30,940 phage clones reactive with IgG in the serum.

[0114]

(3) Homology Search of Isolated Antigen Gene

To subject the single positive clone isolated by the above-described method to a base sequence analysis, an operation of conversion of the phage vector to a plasmid vector was carried out. More particularly, 200 μ l of a solution prepared to contain a host *E. coli* (XL1-Blue MRF') such that the absorbance OD₆₀₀ should be 1.0 was mixed with 100 μ l of a purified phage solution and further with 1 μ l of ExAssist helper phage (manufactured by STRATAGENE), and the reaction was allowed to proceed at 37°C for 15 minutes. To the reaction mixture, 3 ml of LB medium was added, and the mixture was cultured at 37°C for 2.5 to 3 hours, followed by immediate incubation in a water bath at 70°C for 20 minutes. The mixture was then centrifuged at 4°C at 1000 \times g for 15 minutes, and the supernatant was recovered as a phagemid solution. Subsequently, 200 μ l of a solution prepared to contain a phagemid host *E. coli* (SOLR) such that the absorbance OD₆₀₀ should be 1.0 was mixed with 10 μ l of a purified phage solution, and the reaction was allowed to proceed at 37°C for 15 minutes. Thereafter, 50 μ l of the reaction mixture was

plated on ampicillin (final concentration: 50 µg/ml)-containing LB agar medium, and cultured at 37°C overnight. A single colony of transformed SOLR was recovered and cultured in ampicillin (final concentration: 50 µg/ml)-containing LB medium at 37°C, followed by purification of plasmid DNA having an insert of interest using
5 QIAGEN plasmid Miniprep Kit (manufactured by Qiagen).

[0115]

The purified plasmid was subjected to an analysis of the entire sequence of the insert by the primer walking method using the T3 primer described in SEQ ID NO:5 and the T7 primer described in SEQ ID NO:6. By this sequence analysis, the
10 gene sequence described in SEQ ID NO:25 was obtained. Using the base sequence and the amino acid sequence of this gene, homology search against known genes was carried out using a homology search program BLAST

(<http://www.ncbi.nlm.nih.gov/BLAST/>). As a result, it was revealed that the obtained gene has 99% homology (which was calculated only in the overlapping
15 region) to the *CEP* gene described in SEQ ID NO:27 in terms of base sequence and amino acid sequence, so that the gene was judged as the *CEP* gene. The human homologous factor of the canine CEP was human CEP (homology to the *CEP* gene described in SEQ ID NO:25: base sequence, 87%; amino acid sequence, 84%). The base sequence of human CEP is shown in SEQ ID NO:29, and the amino acid
20 sequence thereof is shown in SEQ ID NO:30.

[0116]

(4) Analysis of Expression in Each Tissue

The expression of the gene, which was obtained by the above-described method, in normal tissues and various cell lines of dog and human were investigated
25 by the RT-PCR (Reverse Transcription-PCR) method. The reverse transcription reaction was carried out as follows. That is, total RNA was extracted from 50 to 100 mg of each tissue or 5 to 10×10^6 cells of each cell line using TRIZOL reagent

(manufactured by Invitrogen) in accordance with the protocol attached to the kit. Using this total RNA, cDNA was synthesized by Superscript First-Strand Synthesis System for RT-PCR (manufactured by Invitrogen) in accordance with the protocol attached to the kit. As the cDNAs from human normal tissues (brain, hippocampus, testis, colon and placenta), Gene Pool cDNA (manufactured by Invitrogen), QUICK-Clone cDNA (manufactured by CLONTECH) and Large-Insert cDNA Library (manufactured by CLONTECH) were used. The PCR reactions were carried out as follows using primers (described in SEQ ID NOs:31 and 32) specific to the obtained gene. That is, respective reagents and the attached buffer were mixed such that the mixture should contain 0.25 µl of the sample prepared by the reverse transcription reaction, 2 µM each of the above primers, 0.2 mM each of dNTP and 0.65 U of ExTaq polymerase (manufactured by Takara Shuzo Co., Ltd.) in a total volume of 25 µl, and the reaction was carried out with 30 cycles of 94°C for 30 seconds, 55°C for 30 seconds and 72°C for 30 seconds using Thermal Cycler (manufactured by BIO RAD). The above-described gene-specific primers were those which amplify the regions of the 4582nd to 5124th bases of the base sequences of SEQ ID NOs:25 and 27 (canine *CEP* gene) and the 4610th to 5152nd bases of the base sequence of SEQ ID NO:29 (human *CEP* gene), and can be used for investigation of the expression of both the canine *CEP* gene and the human *CEP* gene. As a control for comparison, primers (described in SEQ ID NOs:9 and 10) specific to GAPDH were used simultaneously. As a result, as shown in Fig. 5, strong expression of the canine *CEP* gene was observed in testis among the normal dog tissues, and on the other hand, strong expression was observed in the canine breast cancer cell line. Expression of the human *CEP* gene was confirmed, as is the case with the canine *CEP* gene, only in testis among the human normal tissues, but the expression was detected in brain tumor, leukemia and esophagus cancer cells among human cancer cell lines, and especially, strong expression was observed in the leukemia cell line.

Thus, the human *CEP* gene was also confirmed to be specifically expressed in testis and cancer cells.

[0117]

In Fig. 5, reference numeral 1 in the ordinate indicates the expression pattern of the *CEP* gene, and reference numeral 2 indicates the expression pattern of the *GAPDH* gene as a control for comparison.

[0118]

Example C-2: Preparation of Canine and Human CEPs

(1) Preparation of Recombinant Protein

Based on the gene of SEQ ID NO:25 obtained in Example C-1, a recombinant protein was prepared by the following method. Respective reagents and the attached buffer were mixed such that the mixture should contain 1 μ l of the vector that was prepared from the phagemid solution obtained in Example C-1 and was subjected to the sequence analysis, 0.4 μ M each of two kinds of primers having *Bam*HI and *Sal*I restriction sites (described in SEQ ID NOs:33 and 34), 0.2 mM dNTP and 1.25 U of PrimeSTAR HS polymerase (manufactured by Takara Shuzo Co., Ltd.) in a total volume of 50 μ l, and PCR was carried out with 30 cycles of 98°C for 10 seconds, 55°C for 5 seconds and 72°C for 7 minutes using Thermal Cycler (manufactured by BIO RAD). The above-described two kinds of primers were those which amplify the region encoding the entire amino acid sequence of SEQ ID NO:26. After the PCR, the amplified DNA was subjected to electrophoresis using 1% agarose gel, and a DNA fragment of about 7.0 kbp was purified using QIAquick Gel Extraction Kit (manufactured by QIAGEN).

[0119]

The purified DNA fragment was ligated into a cloning vector pCR-Blunt (manufactured by Invitrogen). *E. coli* was transformed with the resulting ligation product, and plasmids were recovered thereafter, followed by confirming, by

sequencing, that the amplified gene fragment matches the sequence of interest. The plasmid that matched the sequence of interest was treated with restriction enzymes *Bam*HI and *Sal*I and purified using QIAquick Gel Extraction Kit, followed by inserting the gene sequence of interest into an expression vector for *E. coli*, pET30a (manufactured by Novagen) that had been treated with *Bam*HI and *Sal*I. Usage of this vector enables production of a His-tag fusion recombinant protein. *E. coli* for expression, BL21 (DE3), was transformed with this plasmid, and expression of the protein of interest was induced in *E. coli* with 1 mM IPTG. In the same manner, based on the gene of SEQ ID NO:27, using the canine testis cDNA as a template and two kinds of primers having *Bam*HI and *Sal*I restriction sites (SEQ ID NOs:33 and 35), a recombinant protein of the registered canine *CEP* gene was prepared. The above-described two kinds of primers were those which amplify the region of about 7.8 kbp encoding the entire amino acid sequence of SEQ ID NO:28.

[0120]

Further, based on the gene of SEQ ID NO:29, a recombinant protein of the human homologous gene was prepared by the following method. Respective reagents and the attached buffer were mixed such that the mixture should contain 1 μ l of the cDNA prepared in Example C-1 whose expression could be confirmed by the RT-PCR method in various tissues/cells, 0.4 μ M each of two kinds of primers having *Bam*HI and *Sal*I restriction sites (described in SEQ ID NOs:36 and 37), 0.2 mM dNTP and 1.25 U of PrimeSTAR HS polymerase (manufactured by Takara Shuzo Co., Ltd.) in a total volume of 50 μ l, and PCR was carried out with 30 cycles of 98°C for 10 seconds, 55°C for 5 seconds and 72°C for 7 minutes using Thermal Cycler (manufactured by BIO RAD). The above-described two kinds of primers were those which amplify the region encoding the entire amino acid sequence of SEQ ID NO:30. After the PCR, the amplified DNA was subjected to electrophoresis using 1% agarose gel, and a DNA fragment of about 7.0 kbp was purified using

QIAquick Gel Extraction Kit (manufactured by QIAGEN).

[0121]

The purified DNA fragment was ligated into a cloning vector pCR-Blunt (manufactured by Invitrogen). *E. coli* was transformed with the resulting ligation product, and plasmids were recovered thereafter, followed by confirming, by sequencing, that the amplified gene fragment matches the sequence of interest. The plasmid that matched the sequence of interest was treated with restriction enzymes *Bam*HI and *Sal*I and purified using QIAquick Gel Extraction Kit, followed by inserting the gene sequence of interest into an expression vector for *E. coli*, pET30a (manufactured by Novagen) that had been treated with *Bam*HI and *Sal*I. Usage of this vector enables production of a His-tag fusion recombinant protein. *E. coli* for expression, BL21 (DE3), was transformed with this plasmid, and expression of the protein of interest was induced in *E. coli* with 1 mM IPTG.

[0122]

15 (2) Purification of Recombinant Protein

The above-obtained recombinant *E. coli* cells that expressed SEQ ID NO:25, SEQ ID NO:27, and SEQ ID NO:29, respectively, were cultured in 30 µg/ml kanamycin-containing LB medium at 37°C until the absorbance at 600 nm reached about 0.7, and then isopropyl-β-D-1-thiogalactopyranoside was added thereto such that its final concentration should be 1 mM, followed by culturing them at 30°C for 20 hours. Subsequently, the cells were collected by centrifugation at 4,800 rpm for 10 minutes. The pellet of the cells was suspended in phosphate-buffered saline and further subjected to centrifugation at 4,800 rpm for 10 minutes to wash the cells.

[0123]

25 The cells were suspended in phosphate-buffered saline and subjected to sonication on ice. The sonicated solution of *E. coli* was centrifuged at 7000 rpm for 20 minutes to obtain the supernatant as the soluble fraction and the precipitate as the

insoluble fraction. The insoluble fraction was suspended in 4% Triton X-100 solution and the resulting suspension was centrifuged at 7000 rpm for 20 minutes. This operation was repeated twice and an operation of removal of proteases was carried out. The residue was suspended in 8 M urea-containing 10 mM Tris-HCl, 100 mM phosphate buffer (hereinafter referred to as 8 M urea solution) and a protease inhibitor cocktail solution, and the resulting suspension was left to stand at 4°C for 20 hours to denature proteins.

[0124]

Thereafter, the suspension was centrifuged at 7,000 rpm for 20 minutes, and the obtained soluble fraction was placed in a nickel chelate column prepared by a conventional method (carrier: Chelating Sepharose (trademark) Fast Flow (GE Health Care); column volume: 5mL; equilibration buffer: 8M urea solution), followed by leaving it to stand at 4°C overnight. The supernatant was recovered by centrifugation of this column carrier at 1,500 rpm for 5 minutes, and the column carrier was suspended in phosphate-buffered saline, followed by refilling the column with the resulting suspension. The fraction that was not adsorbed to the column was washed away with 5 column volumes of 8 M urea solution, 10 column volumes of 0.5 M sodium chloride-containing 0.1 M acetate buffer (pH 5.0) and 10 mM imidazole-containing 20 mM phosphate buffer (pH 8.0), and elution was immediately carried out with a five-step density gradient of 100 mM-500 mM imidazole to obtain a purified fraction, which was used as the material for administration tests thereafter. The proteins of interest in respective eluted fractions were confirmed by Coomassie staining carried out according to a conventional method. Among these, the recombinant canine CEP described in SEQ ID NO:26 is shown in Fig. 6.

[0125]

To 1 ml of a reaction buffer (20 mM Tris-HCl, 50 mM NaCl, 2 mM CaCl₂;

pH 7.4), 200 μ l of the purified preparation obtained by the above-described method was aliquoted, and 2 μ l of enterokinase (manufactured by Novagen) was then added thereto, followed by leaving it to stand at room temperature overnight to cleave His tag. The resulting product was purified using Enterokinase Cleavage Capture Kit (manufactured by Novagen) in accordance with the protocol attached to the kit. Subsequently, the buffer contained in 1.2 ml of the purified preparation obtained by the above-described method was replaced with physiological phosphate buffer (manufactured by Nissui Pharmaceutical) by ultrafiltration using NANOSEP 10K OMEGA (manufactured by PALL), and the resulting solution was filtered aseptically using HT Tuffryn Acrodisc 0.22 μ m (manufactured by PALL) and used in the following experiments.

[0126]

Example C-3: Test of Administration of Recombinant Protein to Cancer-bearing Dogs

(1) Antitumor Assay

The anti-tumor effect of the two kinds of recombinant proteins which were purified as described above was assessed in two individuals of cancer-bearing dogs having epidermal tumor (2 individuals having perianal adenoma).

[0127]

An equal amount of Freund's incomplete adjuvant (manufactured by Wako Pure Chemicals) was mixed with 100 μ g (0.5 ml) each of the recombinant canine CEP described in SEQ ID NO:26 and human CEP purified as described above to prepare therapeutic agents for a cancer(s). Each of these agents was administered to a regional lymph node in the vicinity of the tumor a total of 3 times, by carrying out the subsequent administrations 3 days and 7 days after the first administration. As a result, the tumors with a size of about 87 mm³ and 69 mm³ at the time of administration of the therapeutic agents, respectively, were reduced to 69 mm³ and

56 mm³, respectively, 10 days after the first administration; 24 mm³ and 31 mm³, respectively, 20 days after the first administration; and to 10 mm³ and 8 mm³, respectively, 30 days after the first administration of the therapeutic agent.

[0128]

5 Further, to a canine patient suffering from mammary adenocarcinoma, a mixture of 100 µg (0.5 ml) of the canine CEP protein described in SEQ ID NO:26 with 0.5 ml of Freund's incomplete adjuvant was administered a total of 3 times in the same manner as described above. Further, concurrently with the respective
10 administrations, 10 MU of "Intercat" which is a recombinant feline interferon was administered subcutaneously. As a result, the tumor with a size of about 126 mm³ at the time of administration of the therapeutic agent completely regressed 26 days after the first administration of the therapeutic agent. Similarly, in the case where the canine CEP described in SEQ ID NO:28 was used, an anti-tumor effect was also
15 observed in a cancer-bearing dog.

[0129]

Further, to a canine patient of mastocytoma, a mixture of 100 µg (0.5 ml) of the canine CEP protein described in SEQ ID NO:26 with 0.5 ml of Freund's incomplete adjuvant was administered a total of 3 times in the same manner as described above. Further, concurrently with the respective administrations, 100 µg
20 of canine interleukin-12 was subcutaneously administered. As a result, the tumor with a size of about 83 mm³ at the time of administration of the therapeutic agent completely regressed 18 days after the first administration of the therapeutic agent.

[0130]

(2) Immune Inducibility Assay

25 Blood from the canine patient suffering from perianal adenoma in which the anti-tumor effect was obtained in the administration test in the above-described (1) was collected before administration of the therapeutic agent for a cancer(s) and 10

days and 30 days after the first administration. Peripheral blood mononuclear cells were isolated according to a conventional method, and by the ELISPOT assay for IFN γ using it, the immune inducibility of each administered protein was assayed.

[0131]

5 In a 96-well plate manufactured by Millipore (MultiScreen-IP, MAIPS 4510), 100 μ L/well of 70% ethanol was placed and the plate was left to stand for 5 minutes, followed by removal of the ethanol by aspiration. The plate was washed with sterile water and 300 μ L/well of 200 mM Sodium Bicarbonate (pH8.2) was placed therein. After leaving it to stand for 5 minutes, Sodium Bicarbonate was removed by
10 aspiration, and then the plate was washed. Subsequently, 0.5 μ L/well of anti-canine interferon γ monoclonal antibody (manufactured by R&D, clone 142529, MAB781) mixed with 200 mM Sodium Bicarbonate was placed in wells, and the plate was incubated at 37°C overnight to immobilize the primary antibody. After removal of the primary antibody by aspiration, 300 μ L/well of a blocking solution (1% BSA-5% sucrose-200 mM Sodium Bicarbonate (pH8.2)) was added to the wells, and the plate
15 was incubated at 4°C overnight to block the plate. After removal of the blocking solution by aspiration, 300 μ L/well of 10% fetal calf serum-containing RPMI medium (manufactured by Invitrogen) was placed in the wells, and the plate was left to stand for 5 minutes, followed by removal of the medium by aspiration.
20 Subsequently, 5×10^5 cells/well of the canine peripheral blood mononuclear cells suspended in 10% fetal calf serum-containing RPMI medium were placed in the plate, and 10 μ L/well of the canine CEP described in SEQ ID NO:26 or the human CEP used in each administration was added thereto, followed by culturing the cells under the conditions of 37°C and 5% CO₂ for 24 hours, to allow immunocytes that might
25 exist in the peripheral blood mononuclear cells to produce interferon γ . After the culture, the medium was removed, and the wells were washed 6 times with a washing solution (0.1% Tween20-200 mM Sodium Bicarbonate (pH8.2)). In each well, 100

μL of rabbit anti-canine polyclonal antibody 1000-fold diluted with the above-described blocking solution was placed, and the plate was incubated at 4°C overnight. After washing the wells 3 times with the above-described washing solution, 100 μL of HRP-labeled anti-rabbit antibody 1000-fold diluted with the above-described
5 blocking solution was placed in each well, and the reaction was allowed to proceed at 37°C for 2 hours. After washing the wells 3 times with the above-described washing solution, the resultant was colored with Konica Immunostain (manufactured by Konica), and the wells were washed with water to stop the reaction. Thereafter, the membrane was dried, and image processing of the wells was carried out, followed
10 by counting the number of spot-forming cells (SFC) using KS ELISPOT compact system (Carl Zeiss, Inc., Germany).

[0132]

As a result, in either canine patient to which the canine CEP described in SEQ ID NO: 26 or the human CEP was administered, peripheral blood mononuclear cells
15 sampled before the administration showed no spots. On the other hand, in the canine patient to which the canine CEP was administered, peripheral blood mononuclear cells sampled 10 days and 30 days after the administration showed 23 and 52 spots, respectively. In the canine patient to which the human CEP was administered, peripheral blood mononuclear cells sampled 10 days and 30 days after
20 the administration showed 19 and 49 spots, respectively.

[0133]

From the above results, it is confirmed that immunocytes which specifically react with the administered recombinant protein and produce interferon γ were induced in all of the canine patients to which the recombinant protein was
25 administered, and it is thought that the anti-tumor effect described in (1) was exerted by immunoreactions in which these immunocytes are mainly involved.

[0134]

Example D-1: Acquisition of Novel Cancer Antigen Protein by SEREX Method

(1) Preparation of cDNA Library

Total RNA was prepared from testis tissue of a healthy dog by the Acid guanidium-Phenol-Chloroform method, and poly(A) RNA was purified using
5 Oligotex-dT30 mRNA purification Kit (manufactured by Takara Shuzo Co., Ltd.) in accordance with the protocol attached to the kit.

[0135]

Using the obtained mRNA (5 μ g), a dog testis cDNA phage library was synthesized. Preparation of the cDNA phage library was carried out using cDNA
10 Synthesis Kit, ZAP-cDNA Synthesis Kit, and ZAP-cDNA Gigapack III Gold Cloning Kit (manufactured by STRATAGENE) in accordance with the protocols attached to the kits. The size of the prepared cDNA phage library was 1.3×10^6 pfu/ml.

[0136]

(2) Screening of cDNA Library with Serum

15 Using the dog testis-derived cDNA phage library prepared as described above, immunoscreening was carried out. More particularly, host *E. coli* cells (XL1-Blue MRF') were infected with the library such that 2,340 clones should appear on an NZY agarose plate having the size of $\Phi 90 \times 15$ mm, and cultured at 42°C for 3 to 4 hours to allow the phage to form plaques. The plate was covered with nitrocellulose
20 membrane (Hybond C Extra: manufactured by GE Healthcare Bio-Science) impregnated with IPTG (isopropyl- β -D-thiogalactoside) at 37°C for 4 hours to induce and express proteins, which were thus transferred to the membrane. Subsequently, the membrane was recovered and soaked in TBS (10 mM Tris-HCl, 150 mM NaCl; pH 7.5) containing 0.5% non-fat dry milk, followed by shaking at 4°C overnight to
25 suppress non-specific reactions. This filter was allowed to react with 500-fold diluted canine patient serum at room temperature for 2 to 3 hours.

[0137]

As the above-described canine patient serum, serum collected from canine patients suffering from breast cancer was used. The serum was stored at -80°C and pretreated immediately before use. The method of the pretreatment of the serum was as follows. That is, host *E. coli* cells (XL1-Blue MRF') were infected with λ ZAP Express phage to which no foreign gene was inserted, and then cultured on NZY plate medium at 37°C overnight. Subsequently, the buffer of 0.2 M NaHCO₃, pH 8.3 containing 0.5 M NaCl was added to the plate, and the plate was left to stand at 4°C for 15 hours, followed by collecting the supernatant as an *E. coli*/phage extract. Thereafter, the collected *E. coli*/phage extract was allowed to flow through an NHS column (manufactured by GE Healthcare Bio-Science) to immobilize proteins derived from the *E. coli*/phage thereon. The serum from the canine patients was allowed to flow through and react with this protein-immobilized column to remove antibodies adsorbed on *E. coli* and/or the phage. The serum fraction that passed through the column was 500-fold diluted with TBS containing 0.5% non-fat dry milk, and the resulting diluent was used as the material for the immunoscreening.

[0138]

The membrane on which the thus treated serum and the above-described fusion protein were blotted was washed 4 times with TBS-T (0.05% Tween 20/TBS), and allowed to react with goat anti-dog IgG (Goat anti Dog IgG-h+I HRP conjugated: manufactured by BETHYL Laboratories) 5,000-fold diluted with TBS containing 0.5% non-fat dry milk as a secondary antibody at room temperature for 1 hour, followed by detection by the enzyme coloring reaction using the NBT/BCIP reaction solution (manufactured by Roche). Colonies at positions where a positive coloring reaction was observed were recovered from the NZY agarose plate having the size of $\Phi 90 \times 15$ mm, and dissolved in 500 μ l of SM buffer (100 mM NaCl, 10 mM MgClSO₄, 50 mM Tris-HCl, 0.01% gelatin; pH 7.5). The screening was repeated as a second and third screening in the same manner as described above until a single

coloring reaction-positive colony was obtained, thereby isolating one positive clone after screening of 30,940 phage clones reactive with IgG in the serum.

[0139]

(3) Homology Search of Isolated Antigen Gene

5 To subject the single positive clone isolated by the above-described method to a base sequence analysis, an operation of conversion of the phage vector to a plasmid vector was carried out. More particularly, 200 μ l of a solution prepared to contain a host *E. coli* (XL1-Blue MRF') such that the absorbance OD₆₀₀ should be 1.0 was mixed with 100 μ l of a purified phage solution and further with 1 μ l of ExAssist
10 helper phage (manufactured by STRATAGENE), and the reaction was allowed to proceed at 37°C for 15 minutes. To the reaction mixture, 3 ml of LB medium was added, and the mixture was cultured at 37°C for 2.5 to 3 hours, followed by immediate incubation in a water bath at 70°C for 20 minutes. The mixture was then centrifuged at 4°C at 1000 \times g for 15 minutes, and the supernatant was recovered as a
15 phagemid solution. Subsequently, 200 μ l of a solution prepared to contain a phagemid host *E. coli* (SOLR) such that the absorbance OD₆₀₀ should be 1.0 was mixed with 10 μ l of a purified phage solution, and the reaction was allowed to proceed at 37°C for 15 minutes. Thereafter, 50 μ l of the reaction mixture was plated on ampicillin (final concentration: 50 μ g/ml)-containing LB agar medium, and
20 cultured at 37°C overnight. A single colony of transformed SOLR was recovered and cultured in ampicillin (final concentration: 50 μ g/ml)-containing LB medium at 37°C, followed by purification of plasmid DNA having an insert of interest using QIAGEN plasmid Miniprep Kit (manufactured by Qiagen).

[0140]

25 The purified plasmid was subjected to an analysis of the entire sequence of the insert by the primer walking method using the T3 primer described in SEQ ID NO:5 and the T7 primer described in SEQ ID NO:6. By this sequence analysis, the

gene sequence described in SEQ ID NO:38 was obtained. Using the base sequence and the amino acid sequence of this gene, homology search against known genes was carried out using a homology search program BLAST

(<http://www.ncbi.nlm.nih.gov/BLAST/>). As a result, it was revealed that the

5 obtained gene is the *TRIP11* gene. The human homologous factor of canine TRIP11 was human TRIP11 (homology: base sequence, 88%; amino acid sequence, 86%). The base sequence of human TRIP11 is shown in SEQ ID NO:40, and the amino acid sequence thereof is shown in SEQ ID NO:41.

[0141]

10 (4) Analysis of Expression in Each Tissue

The expression of the gene, which was obtained by the above-described method, in normal tissues and various cell lines of dog and human were investigated by the RT-PCR (Reverse Transcription-PCR) method. The reverse transcription reaction was carried out as follows. That is, total RNA was extracted from 50 to
15 100 mg of each tissue or 5 to 10×10^6 cells of each cell line using TRIZOL reagent (manufactured by Invitrogen) in accordance with the protocol attached to the kit. Using this total RNA, cDNA was synthesized by Superscript First-Strand Synthesis System for RT-PCR (manufactured by Invitrogen) in accordance with the protocol attached to the kit. As the cDNAs from human normal tissues (brain, hippocampus,
20 testis, colon and placenta), Gene Pool cDNA (manufactured by Invitrogen), QUICK-Clone cDNA (manufactured by CLONTECH) and Large-Insert cDNA Library (manufactured by CLONTECH) were used. The PCR reactions were carried out as follows using primers (described in SEQ ID NOs:42 and 43) specific to the obtained gene. That is, respective reagents and the attached buffer were mixed such that the
25 mixture should contain 0.25 μ l of the sample prepared by the reverse transcription reaction, 2 μ M each of the above primers, 0.2 mM each of dNTP and 0.65 U of ExTaq polymerase (manufactured by Takara Shuzo Co., Ltd.) in a total volume of 25

μl, and the reaction was carried out with 30 cycles of 94°C for 30 seconds, 55°C for 30 seconds and 72°C for 1.5 minutes using Thermal Cycler (manufactured by BIO RAD). The above-described gene-specific primers were those which amplify the regions of the 1519th to 2957th bases of the base sequence of SEQ ID NO:38 (canine *TRIP11* gene) and the 1872nd to 3310th bases of the base sequence of SEQ ID NO:40 (human *TRIP11* gene), and can be used for investigation of the expression of both the canine *TRIP11* gene and the human *TRIP11* gene. As a control for comparison, primers (described in SEQ ID NOs:9 and 10) specific to GAPDH were used simultaneously. As a result, as shown in Fig. 7, strong expression of the canine *TRIP11* gene was observed in testis among the normal dog tissues, and on the other hand, strong expression was observed in the canine breast cancer cell line. Expression of the human gene was confirmed, as is the case with the canine *TRIP11* gene, only in testis among the human normal tissues, but the expression was detected in many types of cancer cell lines such as brain tumor, leukemia, breast cancer, lung cancer and esophagus cancer cell lines among human cancer cell lines. Thus, the human *TRIP11* gene was also confirmed to be specifically expressed in testis and cancer cells.

[0142]

In Fig. 7, reference numeral 1 in the ordinate indicates the expression pattern of the *TRIP11* gene, and reference numeral 2 indicates the expression pattern of the *GAPDH* gene as a control for comparison.

[0143]

Example D-2: Preparation of Canine and Human *TRIP11* Proteins

(1) Preparation of Recombinant Protein

Based on the gene of SEQ ID NO:38 obtained in Example D-1, a recombinant protein was prepared by the following method. Respective reagents and the attached buffer were mixed such that the mixture should contain 1 μl of the vector

which was prepared from the phagemid solution obtained in Example D-1 and was subjected to the sequence analysis, 0.4 μ M each of two kinds of primers having *SalI* and *XhoI* restriction sites (described in SEQ ID NOs:44 and 45), 0.2 mM dNTP and 1.25 U of PrimeSTAR HS polymerase (manufactured by Takara Shuzo Co., Ltd.) in a total volume of 50 μ l, and PCR was carried out with 30 cycles of 98°C for 10 seconds, 55°C for 5 seconds and 72°C for 6 minutes using Thermal Cycler (manufactured by BIO RAD). The above-described two kinds of primers were those which amplify the region encoding the entire amino acid sequence of SEQ ID NO:39. After the PCR, the amplified DNA was subjected to electrophoresis using 1% agarose gel, and a DNA fragment of about 6.0 kbp was purified using QIAquick Gel Extraction Kit (manufactured by QIAGEN).

[0144]

The purified DNA fragment was ligated into a cloning vector pCR-Blunt (manufactured by Invitrogen). *E. coli* was transformed with the resulting ligation product, and plasmids were recovered thereafter, followed by confirming, by sequencing, that the amplified gene fragment matches the sequence of interest. The plasmid that matched the sequence of interest was treated with restriction enzymes *SalI* and *XhoI* and purified using QIAquick Gel Extraction Kit, followed by inserting the gene sequence of interest into an expression vector for *E. coli*, pET30b (manufactured by Novagen) that had been treated with *SalI* and *XhoI*. Usage of this vector enables production of a His-tag fusion recombinant protein. *E. coli* for expression, BL21 (DE3), was transformed with this plasmid, and expression of the protein of interest was induced in *E. coli* with 1 mM IPTG.

[0145]

Further, based on the gene of SEQ ID NO:40, a recombinant protein of the human homologous gene was prepared by the following method. Respective reagents and the attached buffer were mixed such that the mixture should contain 1

5 μ l of the cDNA prepared in Example D-1 whose expression could be confirmed by the RT-PCR method in various tissues/cells, 0.4 μ M each of two kinds of primers having *Nde*I and *Kpn*I restriction sites (described in SEQ ID NOs:46 and 47), 0.2 mM dNTP and 1.25 U of PrimeSTAR HS polymerase (manufactured by Takara Shuzo Co., Ltd.) in a total volume of 50 μ l, and PCR was carried out with 30 cycles of 98°C for 10 seconds, 55°C for 5 seconds and 72°C for 6 minutes using Thermal Cycler (manufactured by BIO RAD). The above-described two kinds of primers were those which amplify the region encoding the entire amino acid sequence of SEQ ID NO:41. After the PCR, the amplified DNA was subjected to electrophoresis using 1% agarose gel, and a DNA fragment of about 6.0 kbp was purified using QIAquick Gel Extraction Kit (manufactured by QIAGEN).

[0146]

15 The purified DNA fragment was ligated into a cloning vector pCR-Blunt (manufactured by Invitrogen). *E. coli* was transformed with the resulting ligation product, and plasmids were recovered thereafter, followed by confirming, by sequencing, that the amplified gene fragment matches the sequence of interest. The plasmid that matched the sequence of interest was treated with restriction enzymes *Nde*I and *Kpn*I and purified using QIAquick Gel Extraction Kit, followed by inserting the gene sequence of interest into an expression vector for *E. coli*, pET30b (manufactured by Novagen) that had been treated with *Nde*I and *Kpn*I. Usage of this vector enables production of a His-tag fusion recombinant protein. *E. coli* for expression, BL21 (DE3), was transformed with this plasmid, and expression of the protein of interest was induced in *E. coli* with 1 mM IPTG.

[0147]

25 (2) Purification of Recombinant Proteins

The above-obtained recombinant *E. coli* cells that expressed SEQ ID NO:38 and SEQ ID NO:40, respectively, were cultured in 30 μ g/ml kanamycin-containing

LB medium at 37°C until the absorbance at 600 nm reached about 0.7, and then isopropyl- β -D-1-thiogalactopyranoside was added thereto such that its final concentration should be 1 mM, followed by culturing them at 30°C for 20 hours. Subsequently, the cells were collected by centrifugation at 4,800 rpm for 10 minutes. The pellet of the cells was suspended in phosphate-buffered saline and further subjected to centrifugation at 4,800 rpm for 10 minutes to wash the cells.

[0148]

The obtained pellet of *E. coli* cells was suspended in phosphate-buffered saline and subjected to sonication on ice. The sonicated solution of *E. coli* was centrifuged at 7,000 rpm for 15 minutes to obtain the supernatant as the soluble fraction and the precipitate as the insoluble fraction.

[0149]

The insoluble fraction was suspended in 4% Triton X-100 solution and the resulting suspension was centrifuged at 7,000 rpm for 10 minutes. This operation was repeated twice and an operation of removal of proteases was carried out. Thereafter, the residue was suspended in phosphate-buffered saline and an operation of removal of the surfactant was carried out.

[0150]

The residue was suspended in 6M guanidine hydrochloride-containing 20 mM phosphate buffer (pH 8.0), and the resulting suspension was left to stand at 4°C for 20 hours to denature proteins. Thereafter, the suspension was centrifuged at 7,000 rpm for 20 minutes, and the obtained soluble fraction was placed in a nickel chelate column prepared by a conventional method (carrier: Chelating Sepharose (trademark) Fast Flow (GE Health Care); column volume: 5mL; equilibration buffer: 6M guanidine hydrochloride-containing 20 mM phosphate buffer (pH 8.0)). The fraction that was not adsorbed to the column was washed away with 10 column volumes of 6 M sodium chloride-containing 20 mM phosphate buffer (pH 8.0) and

10 mM imidazole-containing 20 mM phosphate buffer (pH 8.0), and elution was immediately carried out with a four-step density gradient of 50 mM-500 mM imidazole to obtain a purified fraction, which was used as the material for administration tests thereafter. The proteins of interest in the eluted fractions were confirmed by Coomassie staining carried out according to a conventional method. Among these, the canine TRIP11 protein is shown in Fig. 8.

[0151]

To 1 ml of a reaction buffer (20 mM Tris-HCl, 50 mM NaCl, 2 mM CaCl₂; pH 7.4), 200 µl of the purified preparation obtained by the above-described method was aliquoted, and 2 µl of enterokinase (manufactured by Novagen) was then added thereto, followed by leaving it to stand at room temperature overnight to cleave His tag. The resulting product was purified using Enterokinase Cleavage Capture Kit (manufactured by Novagen) in accordance with the protocol attached to the kit. Subsequently, the buffer contained in 1.2 ml of the purified preparation obtained by the above-described method was replaced with physiological phosphate buffer (manufactured by Nissui Pharmaceutical) by ultrafiltration using NANOSEP 10K OMEGA (manufactured by PALL), and the resulting solution was filtered aseptically using HT Tuffryn Acrodisc 0.22 µm (manufactured by PALL) and used in the following experiments.

[0152]

Example D-3: Test of Administration of Recombinant Protein to Cancer-bearing Dogs

(1) Antitumor Assay

The anti-tumor effect of the two kinds of recombinant proteins which were purified as described above was assessed in two individuals of cancer-bearing dogs having epidermal tumor (2 individuals having mammary gland tumor).

[0153]

Therapeutic agents for a cancer(s) were prepared by mixing 0.5 ml of Freund's incomplete adjuvant (manufactured by Wako Pure Chemicals) with 100 μg (0.5 ml) of the recombinant canine TRIP11 and human TRIP11 proteins, respectively, purified as described above. Each of these agents was administered to a regional lymph node in the vicinity of the tumor a total of 3 times, by carrying out the subsequent administrations 3 days and 7 days after the first administration. As a result, the tumors with a size of about 75 mm^3 and 102 mm^3 , respectively, at the time of administration of the therapeutic agents were reduced to 63 mm^3 and 85 mm^3 , respectively, 10 days after the first administration; 35 mm^3 and 42 mm^3 , respectively, 20 days after the first administration; and to 15 mm^3 and 19 mm^3 , respectively, 30 days after the first administration of the therapeutic agent for a cancer(s).

[0154]

Further, to a canine patient suffering from mastocytoma, a mixture of 100 μg (0.5 ml) of the canine TRP11 protein with 0.5 ml of Freund's incomplete adjuvant was administered a total of 3 times in the same manner as described above. Concurrently with the respective administrations, 100 μg of canine interleukin-12 was subcutaneously administered. As a result, the tumor with a size of about 165 mm^3 at the time of administration of the therapeutic agent completely regressed 23 days after the first administration of the therapeutic agent.

[0155]

(2) Immune Inducibility Assay

Blood from the canine patient suffering from mammary gland tumor in which the anti-tumor effect was obtained in the administration test in the above-described (1) was collected. Peripheral blood mononuclear cells were isolated according to a conventional method, and by the ELISPOT assay for $\text{IFN}\gamma$ using it, the immune inducibility of each administered protein was assayed.

[0156]

In a 96-well plate manufactured by Millipore (MultiScreen-IP, MAIPS 4510), 100 μL /well of 70% ethanol was placed and the plate was left to stand for 5 minutes, followed by removal of the ethanol by aspiration. The plate was washed with sterile water and 300 μL /well of 200 mM Sodium Bicarbonate (pH8.2) was placed therein.

5 After leaving it to stand for 5 minutes, Sodium Bicarbonate was removed by aspiration, and then the plate was washed. Subsequently, 0.5 μg /well of anti-canine interferon γ monoclonal antibody (manufactured by R&D, clone 142529, MAB781) mixed with 200 mM Sodium Bicarbonate was placed in wells, and the plate was incubated at 37°C overnight to immobilize the primary antibody. After removal of

10 the primary antibody by aspiration, 300 μL /well of a blocking solution (1% BSA-5% sucrose-200 mM Sodium Bicarbonate (pH8.2)) was added to the wells, and the plate was incubated at 4°C overnight to block the plate. After removal of the blocking solution by aspiration, 300 μL /well of 10% fetal calf serum-containing RPMI medium (manufactured by Invitrogen) was placed in the wells and the plate was left

15 to stand for 5 minutes, followed by removal of the medium by aspiration. Subsequently, 5×10^5 cells/well of the canine peripheral blood mononuclear cells suspended in 10% fetal calf serum-containing RPMI medium were placed in the plate, and 10 μL /well of the canine TRIP11 or the human TRIP11 protein used in each administration was added thereto, followed by culturing the cells under the

20 conditions of 37°C and 5% CO_2 for 24 hours, to allow immunocytes that might exist in the peripheral blood mononuclear cells to produce interferon γ . After the culture, the medium was removed, and the wells were washed 6 times with a washing solution (0.1% Tween20-200mM Sodium Bicarbonate (pH8.2)). In each well, 100 μL of rabbit anti-dog polyclonal antibody 1000-fold diluted with the above-described

25 blocking solution was placed, and the plate was incubated at 4°C overnight. After washing the wells 3 times with the above-described washing solution, 100 μL of HRP-labeled anti-rabbit antibody 1,000-fold diluted with the above-described

blocking solution was placed in each well, and the reaction was allowed to proceed at 37°C for 2 hours. After washing the wells 3 times with the above-described washing solution, the resultant was colored with Konica Immunostain (manufactured by Konica), and the wells were washed with water to stop the reaction. Thereafter, 5 the membrane was dried, and image processing of the wells was carried out, followed by counting the number of spot-forming cells (SFC) using KS ELISPOT compact system (Carl Zeiss, Inc., Germany).

[0157]

As a result, in either canine patient to which the canine TRIP11 protein or the 10 human TRIP11 protein was administered, peripheral blood mononuclear cells sampled before the administration showed no spots. On the other hand, in the canine patient to which the canine TRIP11 was administered, peripheral blood mononuclear cells sampled 10 days and 30 days after the administration showed 26 and 65 spots, respectively. In the canine patient to which the human TRIP11 was 15 administered, peripheral blood mononuclear cells sampled 10 days and 30 days after the administration showed 31 and 72 spots, respectively.

[0158]

From the above results, it is confirmed that immunocytes which specifically react with the administered recombinant protein and produce interferon γ were 20 induced in all of the canine patients to which the recombinant protein was administered, and it is thought that the anti-tumor effect described in the above-described (1) was exerted by immunoreactions in which these immunocytes are mainly involved.

SEQUENCE LISTING IN ELECTRONIC FORM

In accordance with Section 111(1) of the Patent Rules, this description contains a sequence listing in electronic form in ASCII text format (file: 72643-116 Seq 16-05-10 v1.txt).

A copy of the sequence listing in electronic form is available from the Canadian Intellectual Property Office.

The sequences in the sequence listing in electronic form are reproduced in the following table.

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 Lys Lys Val Asp Gly Lys Leu Leu Cys Trp Leu Cys Thr Leu Ser Tyr
 120 125 130

aaa cgg gtc ctt cag aag acc aaa gag cag agg aaa cac ctg agt agc 546
 Lys Arg Val Leu Gln Lys Thr Lys Glu Gln Arg Lys His Leu Ser Ser
 135 140 145

tct tct cgt gct ggc cac cag gag aag gag cag tat agt cgc ctg agt 594
 Ser Ser Arg Ala Gly His Gln Glu Lys Glu Gln Tyr Ser Arg Leu Ser
 150 155 160

ggt ggt ggc cat tat aac agc cag aaa aca ctt tct aca tct tca att 642
 Gly Gly Gly His Tyr Asn Ser Gln Lys Thr Leu Ser Thr Ser Ser Ile
 165 170 175 180

caa aat gaa atc cca aag aaa aag tcc aag ttt gag tca atc aca act	690
Gln Asn Glu Ile Pro Lys Lys Lys Ser Lys Phe Glu Ser Ile Thr Thr	
185 190 195	
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Asn Gly Asp Ser Phe Ser Pro Asp Leu Ala Leu Asp Ser Pro Gly Thr	
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Asp His Phe Val Ile Ile Ala Gln Leu Lys Glu Glu Val Ala Thr Leu	
215 220 225	
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Lys Lys Met Leu His Gln Lys Asp Gln Met Ile Leu Glu Lys Glu Lys	
230 235 240	
aag att aca gag ttg aag gct gat ttt cag tac cag gaa tcg cag atg	882
Lys Ile Thr Glu Leu Lys Ala Asp Phe Gln Tyr Gln Glu Ser Gln Met	
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aga gcc aaa atg aac cag atg gag aaa acc cac aaa gaa gtc aca gaa	930
Arg Ala Lys Met Asn Gln Met Glu Lys Thr His Lys Glu Val Thr Glu	
265 270 275	
caa ctg cag gcc aaa aac cga gag ctc ctg aag cag gca gct gct ttg	978
Gln Leu Gln Ala Lys Asn Arg Glu Leu Leu Lys Gln Ala Ala Ala Leu	
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tcc aag agc aag aag tca gag aag tca gga gct ata acc tct cca tga	1026
Ser Lys Ser Lys Lys Ser Glu Lys Ser Gly Ala Ile Thr Ser Pro	
295 300 305	
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gagatactta agggaggtag caaagatttg aaccgtctgt ctttttaagt aagggcagaa	1926
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 <213> Homo sapiens

86

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 35 40 45
 Ser Lys Thr Asn Thr Ile Cys Lys Lys Cys Ala Gln Asn Val Gln Leu
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 Tyr Gly Thr Pro Lys Pro Cys Gln Tyr Cys Asn Ile Ile Ala Ala Phe
 65 70 75 80
 Ile Gly Asn Lys Cys Gln Arg Cys Thr Asn Ser Glu Lys Lys Tyr Gly
 85 90 95
 Pro Pro Tyr Ser Cys Glu Gln Cys Lys Gln Gln Cys Ala Phe Asp Arg
 100 105 110
 Lys Asp Asp Arg Lys Lys Val Asp Gly Lys Leu Leu Cys Trp Leu Cys
 115 120 125
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 130 135 140
 His Leu Ser Ser Ser Ser Arg Ala Gly His Gln Glu Lys Glu Gln Tyr
 145 150 155 160
 Ser Arg Leu Ser Gly Gly Gly His Tyr Asn Ser Gln Lys Thr Leu Ser
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 Thr Ser Ser Ile Gln Asn Glu Ile Pro Lys Lys Lys Ser Lys Phe Glu
 180 185 190
 Ser Ile Thr Thr Asn Gly Asp Ser Phe Ser Pro Asp Leu Ala Leu Asp
 195 200 205
 Ser Pro Gly Thr Asp His Phe Val Ile Ile Ala Gln Leu Lys Glu Glu
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 Val Ala Thr Leu Lys Lys Met Leu His Gln Lys Asp Gln Met Ile Leu
 225 230 235 240
 Glu Lys Glu Lys Lys Ile Thr Glu Leu Lys Ala Asp Phe Gln Tyr Gln
 245 250 255
 Glu Ser Gln Met Arg Ala Lys Met Asn Gln Met Glu Lys Thr His Lys
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Glu Asn Ser Glu Glu Ile Asp Val Asn Glu Gly Glu Leu Pro Ser Glu	
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Ile Asn Tyr Lys Thr Pro Gln Pro Met Gly Glu Val Tyr Phe Thr Glu	
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Thr Phe Asp Ser Gly Arg Leu Ala Gly Trp Val Leu Ser Lys Ala Lys	
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Lys Asp Asp Thr Asp Ala Glu Ile Ser Ile Tyr Asp Gly Arg Trp Glu	
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ata gaa gaa ttg aaa gaa aac cga gtg cct ggt gac aga ggg ctg gta	397
Ile Glu Glu Leu Lys Glu Asn Arg Val Pro Gly Asp Arg Gly Leu Val	
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ctg aaa tct aga gca aag cat cat gca ata gct gct gta tta gca aaa	445
Leu Lys Ser Arg Ala Lys His His Ala Ile Ala Ala Val Leu Ala Lys	
115 120 125	
ccc ttc att ttt gct gac aaa ccc ttg atc gtt caa tat gaa gta aat	493
Pro Phe Ile Phe Ala Asp Lys Pro Leu Ile Val Gln Tyr Glu Val Asn	
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Phe Gln Asp Gly Ile Asp Cys Gly Gly Ala Tyr Ile Lys Leu Leu Ala	
145 150 155 160	
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Asp Thr Asp Gly Leu Asn Leu Glu Asn Phe Tyr Asp Lys Thr Ser Tyr	
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Thr Ile Met Phe Gly Pro Asp Lys Cys Gly Glu Asp Tyr Lys Leu His	
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Phe Ile Phe Arg His Lys His Pro Lys Thr Gly Val Phe Glu Glu Lys	
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His Ala Lys Pro Pro Asp Val Asp Leu Lys Lys Phe Phe Thr Asp Arg	
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Lys Thr His Leu Tyr Thr Leu Val Met Asn Pro Asp Asp Thr Phe Glu	
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Val Val Pro Pro Ile Asn Pro Pro Lys Glu Ile Glu Asp Pro Ser Asp	
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Lys Lys Pro Asp Glu Trp Asp Glu Arg Ala Lys Ile Pro Asp Pro Ser	
275 280 285	
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Ala Val Lys Pro Glu Asp Trp Asp Glu Ser Glu Pro Ala Gln Ile Glu	
290 295 300	
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Asp Leu Ser Val Val Lys Pro Asp Gly Trp Leu Asp Asp Glu Pro Lys	
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Phe Ile Pro Asp Pro Asn Ala Glu Lys Pro Asp Asp Trp Asn Glu Asp	
325 330 335	
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Met Asp Gly Glu Trp Glu Ala Pro Arg Ile Ser Asn Pro Ala Cys Arg	
340 345 350	
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Ile Gly Cys Gly Glu Trp Ser Pro Pro Met Ile Asp Asn Pro Lys Tyr	
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aaa gga gta tgg aga cct cca atg ata gat aat cct aac tac cag gga	1213
Lys Gly Val Trp Arg Pro Pro Met Ile Asp Asn Pro Asn Tyr Gln Gly	
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Ile Trp Ser Pro Arg Lys Ile Pro Asn Pro Asp Tyr Phe Glu Asp Asp	
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His Pro Phe Leu Leu Thr Ser Phe Arg Ala Leu Gly Leu Glu Leu Trp	
405 410 415	
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Ser Met Thr Ser Asn Ile Tyr Phe Asp Asn Phe Ile Ile Cys Ser Glu	
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Leu Val Ala Asn Ala Asn Glu Pro Gly Ile Phe Lys Gln Leu Met Ala	
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Ala Ala Glu Glu Arg Pro Trp Leu Trp Leu Ile Tyr Phe Val Thr Ala	
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485 490 495	

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cca caa aca aag gga gca cta gag caa gaa gtg aag gaa aag aaa gct 1645
Pro Gln Thr Lys Gly Ala Leu Glu Gln Glu Val Lys Glu Lys Lys Ala
515 520 525

gcc ctg gag aaa cca gta gac ttg gaa gaa gaa aaa aag caa agt gat 1693
Ala Leu Glu Lys Pro Val Asp Leu Glu Glu Glu Lys Lys Gln Ser Asp
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ggt gaa act gtt gaa aaa gaa gag gaa gct gaa cct gag gaa aag agt 1741
Gly Glu Thr Val Glu Lys Glu Glu Glu Ala Glu Pro Glu Glu Lys Ser
545 550 555 560

gaa gaa gaa att gaa atc ata gaa gga caa gaa gaa ggt aat aaa tca 1789
Glu Glu Glu Ile Glu Ile Ile Glu Gly Gln Glu Glu Gly Asn Lys Ser
565 570 575

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Asn Lys Ser Gly Ser Glu Asp Glu Met Lys Glu Ala Asp Glu Ser Thr
580 585 590

gga tct gga gat ggg cca gtg aag tca gtg cgc aaa aga aga gta cga 1885
Gly Ser Gly Asp Gly Pro Val Lys Ser Val Arg Lys Arg Arg Val Arg
595 600 605

aag gaa taa actatattca agtattttta attcctgagc gagatatttg 1934
Lys Glu
610

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<211> 610
<212> PRT
<213> Canis familiaris

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Glu Asn Ser Glu Glu Ile Asp Val Asn Glu Gly Glu Leu Pro Ser Glu
35 40 45
Ile Asn Tyr Lys Thr Pro Gln Pro Met Gly Glu Val Tyr Phe Thr Glu
50 55 60
Thr Phe Asp Ser Gly Arg Leu Ala Gly Trp Val Leu Ser Lys Ala Lys
65 70 75 80
Lys Asp Asp Thr Asp Ala Glu Ile Ser Ile Tyr Asp Gly Arg Trp Glu
85 90 95

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Leu	Lys	Ser	Arg	Ala	Lys	His	His	Ala	Ile	Ala	Ala	Val	Leu	Ala	Lys
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Pro	Phe	Ile	Phe	Ala	Asp	Lys	Pro	Leu	Ile	Val	Gln	Tyr	Glu	Val	Asn
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Phe	Gln	Asp	Gly	Ile	Asp	Cys	Gly	Gly	Ala	Tyr	Ile	Lys	Leu	Leu	Ala
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Asp	Thr	Asp	Gly	Leu	Asn	Leu	Glu	Asn	Phe	Tyr	Asp	Lys	Thr	Ser	Tyr
				165					170					175	
Thr	Ile	Met	Phe	Gly	Pro	Asp	Lys	Cys	Gly	Glu	Asp	Tyr	Lys	Leu	His
			180					185					190		
Phe	Ile	Phe	Arg	His	Lys	His	Pro	Lys	Thr	Gly	Val	Phe	Glu	Glu	Lys
		195					200					205			
His	Ala	Lys	Pro	Pro	Asp	Val	Asp	Leu	Lys	Lys	Phe	Phe	Thr	Asp	Arg
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Lys	Thr	His	Leu	Tyr	Thr	Leu	Val	Met	Asn	Pro	Asp	Asp	Thr	Phe	Glu
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Val	Leu	Ile	Asp	Gln	Val	Val	Val	Asn	Gln	Gly	Ser	Leu	Leu	Glu	Asp
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Phe	Ile	Pro	Asp	Pro	Asn	Ala	Glu	Lys	Pro	Asp	Asp	Trp	Asn	Glu	Asp
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385					390					395					400
His	Pro	Phe	Leu	Leu	Thr	Ser	Phe	Arg	Ala	Leu	Gly	Leu	Glu	Leu	Trp
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Ser	Met	Thr	Ser	Asn	Ile	Tyr	Phe	Asp	Asn	Phe	Ile	Ile	Cys	Ser	Glu
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Lys	Glu	Thr	Ala	Asp	Arg	Trp	Ala	Ala	Asp	Gly	Trp	Gly	Val	Lys	Ile
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		515					520					525			
Ala	Leu	Glu	Lys	Pro	Val	Asp	Leu	Glu	Glu	Glu	Lys	Lys	Gln	Ser	Asp
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Gly	Glu	Thr	Val	Glu	Lys	Glu	Glu	Glu	Ala	Glu	Pro	Glu	Glu	Lys	Ser
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 Lys Glu
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 <222> (102)..(1934)

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 Phe Trp Leu Cys Leu Gly Leu Leu Phe Ile Ser Ile Asn Ala Glu Phe
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atg gat gat gat gtt gag acg gaa gac ttt gaa gaa aat tca gaa gaa 212
 Met Asp Asp Asp Val Glu Thr Glu Asp Phe Glu Glu Asn Ser Glu Glu
 25 30 35

att gat gtt aat gaa agt gaa ctt tcc tca gag att aaa tat aag aca 260
 Ile Asp Val Asn Glu Ser Glu Leu Ser Ser Glu Ile Lys Tyr Lys Thr
 40 45 50

cct caa cct ata gga gaa gta tat ttt gca gaa act ttt gat agt gga 308
 Pro Gln Pro Ile Gly Glu Val Tyr Phe Ala Glu Thr Phe Asp Ser Gly
 55 60 65

agg ttg gct gga tgg gtc tta tca aaa gca aag aaa gat gac atg gat 356
 Arg Leu Ala Gly Trp Val Leu Ser Lys Ala Lys Lys Asp Asp Met Asp
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gag gaa att tca ata tac gat gga aga tgg gaa att gaa gag ttg aaa 404
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 90 95 100

gaa aac cag gta cct ggt gac aga gga ctg gta tta aaa tct aga gca 452
 Glu Asn Gln Val Pro Gly Asp Arg Gly Leu Val Leu Lys Ser Arg Ala
 105 110 115

aag cat cat gca ata tct gct gta tta gca aaa cca ttc att ttt gct 500
 Lys His His Ala Ile Ser Ala Val Leu Ala Lys Pro Phe Ile Phe Ala
 120 125 130

gat aaa ccc ttg ata gtt caa tat gaa gta aat ttt caa gat ggt att 548
 Asp Lys Pro Leu Ile Val Gln Tyr Glu Val Asn Phe Gln Asp Gly Ile
 135 140 145

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Asp	Cys	Gly	Gly	Ala	Tyr	Ile	Lys	Leu	Leu	Ala	Asp	Thr	Asp	Asp	Leu	
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Ile	Leu	Glu	Asn	Phe	Tyr	Asp	Lys	Thr	Ser	Tyr	Ile	Ile	Met	Phe	Gly	
			170						175					180		
cca	gat	aaa	tgt	gga	gaa	gat	tat	aaa	ctt	cat	ttt	atc	ttc	aga	cat	692
Pro	Asp	Lys	Cys	Gly	Glu	Asp	Tyr	Lys	Leu	His	Phe	Ile	Phe	Arg	His	
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Asp	Val	Asp	Leu	Lys	Lys	Phe	Phe	Thr	Asp	Arg	Lys	Thr	His	Leu	Tyr	
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Thr	Leu	Val	Met	Asn	Pro	Asp	Asp	Thr	Phe	Glu	Val	Leu	Val	Asp	Gln	
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Lys	Pro	Ala	Gly	Trp	Leu	Asp	Asp	Glu	Pro	Lys	Phe	Ile	Pro	Asp	Pro	
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Glu	Ala	Pro	Gln	Ile	Leu	Asn	Pro	Ala	Cys	Arg	Ile	Gly	Cys	Gly	Glu	
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Trp	Lys	Pro	Pro	Met	Ile	Asp	Asn	Pro	Lys	Tyr	Lys	Gly	Val	Trp	Arg	
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Pro	Pro	Leu	Val	Asp	Asn	Pro	Asn	Tyr	Gln	Gly	Ile	Trp	Ser	Pro	Arg	
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Thr Ser Phe Ser Ala Leu Gly Leu Glu Leu Trp Ser Met Thr Ser Asp	
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Ile Tyr Phe Asp Asn Phe Ile Ile Cys Ser Glu Lys Glu Val Ala Asp	
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Pro Trp Leu Trp Leu Ile Tyr Leu Val Thr Ala Gly Val Pro Ile Ala	
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Glu Asn Ser Glu Glu Ile Asp Val Asn Glu Ser Glu Leu Ser Ser Glu
35          40          45
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Thr Phe Asp Ser Gly Arg Leu Ala Gly Trp Val Leu Ser Lys Ala Lys
65          70          75          80
Lys Asp Asp Met Asp Glu Glu Ile Ser Ile Tyr Asp Gly Arg Trp Glu
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Ile Glu Glu Leu Lys Glu Asn Gln Val Pro Gly Asp Arg Gly Leu Val
100         105         110
Leu Lys Ser Arg Ala Lys His His Ala Ile Ser Ala Val Leu Ala Lys
115         120         125
Pro Phe Ile Phe Ala Asp Lys Pro Leu Ile Val Gln Tyr Glu Val Asn
130         135         140
Phe Gln Asp Gly Ile Asp Cys Gly Gly Ala Tyr Ile Lys Leu Leu Ala
145         150         155         160
Asp Thr Asp Asp Leu Ile Leu Glu Asn Phe Tyr Asp Lys Thr Ser Tyr
165         170         175
Ile Ile Met Phe Gly Pro Asp Lys Cys Gly Glu Asp Tyr Lys Leu His
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Phe Ile Phe Arg His Lys His Pro Lys Thr Gly Val Phe Glu Glu Lys
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97

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 His Pro Phe Leu Leu Thr Ser Phe Ser Ala Leu Gly Leu Glu Leu Trp
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 Ser Met Thr Ser Asp Ile Tyr Phe Asp Asn Phe Ile Ile Cys Ser Glu
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 Lys Glu Val Ala Asp His Trp Ala Ala Asp Gly Trp Arg Trp Lys Ile
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 Ala Ala Glu Gly His Pro Trp Leu Trp Leu Ile Tyr Leu Val Thr Ala
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 Gly Val Pro Ile Ala Leu Ile Thr Ser Phe Cys Trp Pro Arg Lys Val
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 Ser Ser His Ser Pro Ser Pro Pro Ser Leu Thr Ser Asn Met Arg Ser
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agg tca ctt tcg cct cta agt gga tct gag act ctg cct ttt cat ttt 144
 Arg Ser Leu Ser Pro Leu Ser Gly Ser Glu Thr Leu Pro Phe His Phe
 35 40 45

gga gga ccg tgg cat gag caa gtt gag att aca gat gaa agc aca gtg 192
 Gly Gly Pro Trp His Glu Gln Val Glu Ile Thr Asp Glu Ser Thr Val
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gtt tta gac tac caa gac cat aaa gaa gct gat tca cat gca gga gtc 240
 Val Leu Asp Tyr Gln Asp His Lys Glu Ala Asp Ser His Ala Gly Val
 65 70 75 80

cga tat att aca gag gcc ctt gtt aga aaa ctt act aaa cag gac aat 288
 Arg Tyr Ile Thr Glu Ala Leu Val Arg Lys Leu Thr Lys Gln Asp Asn
 85 90 95

ttg gcc ttg gta aaa tct ctg aac ctt tca ctt gct aaa ggt ggt ggc 336
 Leu Ala Leu Val Lys Ser Leu Asn Leu Ser Leu Ala Lys Gly Gly Gly
 100 105 110

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 Lys Lys Phe Arg Cys Ile Glu Asn Leu Glu Lys Cys Val Lys Leu Glu
 115 120 125

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 130 135 140

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 Gln Asp Val Ser Lys Leu Lys Pro Leu Gln Asp Leu Thr Ser Leu Ile
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 225 230 235 240

100

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Ser Gln Asp Arg Gln Glu Ala Phe Ala Arg Phe Ser Leu Asp Glu Val	
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Glu Arg Leu Glu Arg Asp Leu Glu Lys Lys Thr Met Glu Thr Glu Glu	
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Val	Asp	Gly	Leu	Ile	Arg	Pro	Glu	Glu	Val	Ala	Ala	Cys	Val	Asp	Glu	
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cta	agg	aaa	aaa	ctg	aag	tca	gga	gct	ggg	gaa	atg	aga	atc	cat	act	2496
Leu	Arg	Lys	Lys	Leu	Lys	Ser	Gly	Ala	Gly	Glu	Met	Arg	Ile	His	Thr	
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cct	tca	gat	gtc	tta	ggg	aaa	agt	ctt	gct	gac	ttg	cag	aag	caa	ttc	2544
Pro	Ser	Asp	Val	Leu	Gly	Lys	Ser	Leu	Ala	Asp	Leu	Gln	Lys	Gln	Phe	
		835					840					845				
agt	gag	atc	ctg	gca	cg	tcc	cag	tgg	gaa	aga	cag	gaa	gca	caa	gtg	2592
Ser	Glu	Ile	Leu	Ala	Arg	Ser	Gln	Trp	Glu	Arg	Gln	Glu	Ala	Gln	Val	
	850					855					860					
aga	gag	aga	aaa	ctc	cag	gag	gaa	atg	gct	ctg	caa	caa	gag	aaa	ctg	2640
Arg	Glu	Arg	Lys	Leu	Gln	Glu	Glu	Met	Ala	Leu	Gln	Gln	Glu	Lys	Leu	
865					870					875					880	
gcg	agc	gga	caa	gag	gag	ttc	agg	cac	gcc	tgc	gag	agg	gcc	ctg	gaa	2688
Ala	Ser	Gly	Gln	Glu	Glu	Phe	Arg	His	Ala	Cys	Glu	Arg	Ala	Leu	Glu	
				885					890					895		
gcc	cga	att	agt	ttt	gat	aag	agg	cag	cac	gaa	gca	aga	atc	cag	cag	2736
Ala	Arg	Ile	Ser	Phe	Asp	Lys	Arg	Gln	His	Glu	Ala	Arg	Ile	Gln	Gln	
			900					905					910			
ttg	gag	aat	gaa	att	cac	tat	ttg	caa	gaa	aat	cta	aaa	agt	atg	gag	2784
Leu	Glu	Asn	Glu	Ile	His	Tyr	Leu	Gln	Glu	Asn	Leu	Lys	Ser	Met	Glu	
		915					920					925				
gaa	atc	caa	ggt	ctc	aca	gac	ctc	caa	ctt	cag	gaa	gct	gat	gaa	gag	2832
Glu	Ile	Gln	Gly	Leu	Thr	Asp	Leu	Gln	Leu	Gln	Glu	Ala	Asp	Glu	Glu	
	930					935					940					
aag	gag	aga	att	ctg	gcc	caa	ctc	cg	gag	tta	gag	aaa	aag	aag	aaa	2880
Lys	Glu	Arg	Ile	Leu	Ala	Gln	Leu	Arg	Glu	Leu	Glu	Lys	Lys	Lys	Lys	
945					950					955					960	

ctt gag gat gcc aag tct cag gag cag ttt ctt gga tta gat aga gaa Leu Glu Asp Ala Lys Ser Gln Glu Gln Phe Leu Gly Leu Asp Arg Glu	2928
965 970 975	
ttg aag aag cta aag aaa gct gtg gct gcc tct gat aag ctg gcc aca Leu Lys Lys Leu Lys Lys Ala Val Ala Ala Ser Asp Lys Leu Ala Thr	2976
980 985 990	
gct gag ctc acc att gcc aaa gac cag ctc aag tcc ctt cat gga act Ala Glu Leu Thr Ile Ala Lys Asp Gln Leu Lys Ser Leu His Gly Thr	3024
995 1000 1005	
gtg atg aaa att aac cag gag cga gca gag gag ctg cag gag acg Val Met Lys Ile Asn Gln Glu Arg Ala Glu Glu Leu Gln Glu Thr	3069
1010 1015 1020	
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1055 1060 1065	
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1070 1075 1080	
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1085 1090 1095	
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1115 1120 1125	
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1145 1150 1155	
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1160 1165 1170	
aca gcc tcc act ccg gtt aga aaa cca cat cgt gga cgg cag gat Thr Ala Ser Thr Pro Val Arg Lys Pro His Arg Gly Arg Gln Asp	3564
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tat tct cct atc agg agt ggg tta cat aaa tcg ttc tca aat aga Tyr Ser Pro Ile Arg Ser Gly Leu His Lys Ser Phe Ser Asn Arg 1205 1210 1215	3654
gac gca gac agt gga gga gat agc cag gaa gag agc gag cta gat Asp Ala Asp Ser Gly Gly Asp Ser Gln Glu Glu Ser Glu Leu Asp 1220 1225 1230	3699
gac caa gaa gac cac cca ttt gta cct cct cct gga tac atg atg Asp Gln Glu Asp His Pro Phe Val Pro Pro Pro Gly Tyr Met Met 1235 1240 1245	3744
tac act gtg ttt cct gat ggt tct cct gta ccc cag ggc atg gcc Tyr Thr Val Phe Pro Asp Gly Ser Pro Val Pro Gln Gly Met Ala 1250 1255 1260	3789
ctg tat gca ccc cct cct ccc ttg ccc aac aat agc cag cct ctt Leu Tyr Ala Pro Pro Pro Pro Leu Pro Asn Asn Ser Gln Pro Leu 1265 1270 1275	3834
gac ctt ggc act gtt gtt tat ggc cca cct cct gtt ggg gct ccc Asp Leu Gly Thr Val Val Tyr Gly Pro Pro Pro Val Gly Ala Pro 1280 1285 1290	3879
atc gtg tat ggg cct cca cct ccc aac ttc tcc gta ccc ctc atc Ile Val Tyr Gly Pro Pro Pro Pro Asn Phe Ser Val Pro Leu Ile 1295 1300 1305	3924
ccc gtg ggt gtg ctg cac tgc aat gtc cca gaa cac cat aac ttg Pro Val Gly Val Leu His Cys Asn Val Pro Glu His His Asn Leu 1310 1315 1320	3969
gag aat gaa gtt tct aga tta gaa gac ata atg cag cat tta aaa Glu Asn Glu Val Ser Arg Leu Glu Asp Ile Met Gln His Leu Lys 1325 1330 1335	4014
tct ggg aaa cgg gaa cag tgc atg aaa aca ccc aag ctg cag tcg Ser Gly Lys Arg Glu Gln Cys Met Lys Thr Pro Lys Leu Gln Ser 1340 1345 1350	4059
gag aaa gaa ctc gca gag ctg cag cat aac att gat ggt ctt ttg Glu Lys Glu Leu Ala Glu Leu Gln His Asn Ile Asp Gly Leu Leu 1355 1360 1365	4104
caa gag aag aaa gac tta gag cat gaa gta gaa gaa tta cat aga Gln Glu Lys Lys Asp Leu Glu His Glu Val Glu Glu Leu His Arg 1370 1375 1380	4149
acc atc caa aaa cat caa cag cga aaa gat ttc att gat gga aac Thr Ile Gln Lys His Gln Gln Arg Lys Asp Phe Ile Asp Gly Asn 1385 1390 1395	4194
gtt gag agt ctt gtg aat gat cta gaa ata gag aag tca ctc aaa Val Glu Ser Leu Val Asn Asp Leu Glu Ile Glu Lys Ser Leu Lys 1400 1405 1410	4239

cac	cat	gaa	gat	att	gtt	gat	gaa	att	gaa	tgt	att	gag	agg	acc	4284
His	His	Glu	Asp	Ile	Val	Asp	Glu	Ile	Glu	Cys	Ile	Glu	Arg	Thr	
	1415					1420					1425				
ctt	ctg	aag	cgc	cgt	gca	gag	ctc	agg	gaa	gcc	gac	cgg	ctg	ctg	4329
Leu	Leu	Lys	Arg	Arg	Ala	Glu	Leu	Arg	Glu	Ala	Asp	Arg	Leu	Leu	
	1430					1435					1440				
acg	gag	gct	gaa	agt	gaa	ctt	tca	tgc	acg	aaa	gag	aaa	aca	aaa	4374
Thr	Glu	Ala	Glu	Ser	Glu	Leu	Ser	Cys	Thr	Lys	Glu	Lys	Thr	Lys	
	1445					1450					1455				
cat	gct	gtt	gag	aag	ttc	act	gat	gcc	aag	aga	aat	tta	ttg	caa	4419
His	Ala	Val	Glu	Lys	Phe	Thr	Asp	Ala	Lys	Arg	Asn	Leu	Leu	Gln	
	1460					1465					1470				
act	gag	aaa	gat	gct	gag	gag	tta	gaa	agg	aga	gcc	cag	gaa	act	4464
Thr	Glu	Lys	Asp	Ala	Glu	Glu	Leu	Glu	Arg	Arg	Ala	Gln	Glu	Thr	
	1475					1480					1485				
gcc	att	aac	ctc	gtc	aaa	gcc	gac	cag	cag	ctg	aga	ttg	ctc	cag	4509
Ala	Ile	Asn	Leu	Val	Lys	Ala	Asp	Gln	Gln	Leu	Arg	Leu	Leu	Gln	
	1490					1495					1500				
gct	gac	acg	aag	gat	ttg	gag	cag	cac	aaa	atg	gag	caa	gag	gaa	4554
Ala	Asp	Thr	Lys	Asp	Leu	Glu	Gln	His	Lys	Met	Glu	Gln	Glu	Glu	
	1505					1510					1515				
atc	ttg	aaa	gaa	ata	aac	aaa	gtt	gtt	gca	gca	aaa	gac	tca	gac	4599
Ile	Leu	Lys	Glu	Ile	Asn	Lys	Val	Val	Ala	Ala	Lys	Asp	Ser	Asp	
	1520					1525					1530				
ttc	cag	agc	cta	aac	aag	aag	aag	gaa	gta	ctg	aca	gga	gag	ctg	4644
Phe	Gln	Ser	Leu	Asn	Lys	Lys	Lys	Glu	Val	Leu	Thr	Gly	Glu	Leu	
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cag	aaa	ctc	cag	aag	gac	att	gag	act	gca	cgg	cac	aat	gag	gat	4689
Gln	Lys	Leu	Gln	Lys	Asp	Ile	Glu	Thr	Ala	Arg	His	Asn	Glu	Asp	
	1550					1555					1560				
cag	cac	ctg	cag	gtc	ctt	aaa	gag	tcg	gag	acc	ctc	ctg	cag	gcc	4734
Gln	His	Leu	Gln	Val	Leu	Lys	Glu	Ser	Glu	Thr	Leu	Leu	Gln	Ala	
	1565					1570					1575				
aag	aaa	gct	gag	ctg	gaa	aat	ctg	aaa	agc	cag	gtg	tca	gga	cag	4779
Lys	Lys	Ala	Glu	Leu	Glu	Asn	Leu	Lys	Ser	Gln	Val	Ser	Gly	Gln	
	1580					1585					1590				
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Gln	Gln	Glu	Met	Ala	Val	Leu	Asp	Arg	Glu	Leu	Gly	His	Lys	Lys	
	1595					1600					1605				
gaa	gag	ctg	cat	ctc	ctc	cag	gaa	agc	atg	gtc	cag	gcc	aaa	gct	4869
Glu	Glu	Leu	His	Leu	Leu	Gln	Glu	Ser	Met	Val	Gln	Ala	Lys	Ala	
	1610					1615					1620				
gac	ctc	cag	gaa	gca	ctg	aga	cta	gga	gaa	agc	gaa	gta	act	gag	4914
Asp	Leu	Gln	Glu	Ala	Leu	Arg	Leu	Gly	Glu	Ser	Glu	Val	Thr	Glu	
	1625					1630					1635				

aag tgc aat cac att agg gaa gta aaa tct ctt ctg gaa gaa ctc	4959
Lys Cys Asn His Ile Arg Glu Val Lys Ser Leu Leu Glu Glu Leu	
1640 1645 1650	
agt ttt cag aaa gga gaa ctg aat gtc cag atc agt gaa aaa aaa	5004
Ser Phe Gln Lys Gly Glu Leu Asn Val Gln Ile Ser Glu Lys Lys	
1655 1660 1665	
act caa ctt gca ctc ata aag cag gaa att gaa aaa gag gaa gac	5049
Thr Gln Leu Ala Leu Ile Lys Gln Glu Ile Glu Lys Glu Glu Asp	
1670 1675 1680	
aat ctt cag gta gtt tta ggg caa atg tct aaa cat aaa act gaa	5094
Asn Leu Gln Val Val Leu Gly Gln Met Ser Lys His Lys Thr Glu	
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cta aag aat att ctg gac atg ttg caa ctt gaa aat aat gag ctg	5139
Leu Lys Asn Ile Leu Asp Met Leu Gln Leu Glu Asn Asn Glu Leu	
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caa ggt ttg aag ctc caa cat gac caa aag atg tct gaa tta gag	5184
Gln Gly Leu Lys Leu Gln His Asp Gln Lys Met Ser Glu Leu Glu	
1715 1720 1725	
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Lys Thr Arg Val Glu Val Leu Glu Glu Lys Leu Glu Leu Glu Ser	
1730 1735 1740	
ctg cag cag gca gcc ctg cga cag aga ggg gag ata gag tgg cag	5274
Leu Gln Gln Ala Ala Leu Arg Gln Arg Gly Glu Ile Glu Trp Gln	
1745 1750 1755	
aag cag ctc ctc cag agg aac aca cag gaa gta gag cgg atg act	5319
Lys Gln Leu Leu Gln Arg Asn Thr Gln Glu Val Glu Arg Met Thr	
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gct gag acc cga gca tta cag tcg tgt gtt gag tct ttg tgc aaa	5364
Ala Glu Thr Arg Ala Leu Gln Ser Cys Val Glu Ser Leu Cys Lys	
1775 1780 1785	
gaa aag caa gat ctc gaa gaa aaa cag gac agc tgg gaa aag aag	5409
Glu Lys Gln Asp Leu Glu Glu Lys Gln Asp Ser Trp Glu Lys Lys	
1790 1795 1800	
ttg gca cag acc aaa cgg gtt cta gca gct gca gaa gag gac agc	5454
Leu Ala Gln Thr Lys Arg Val Leu Ala Ala Ala Glu Glu Asp Ser	
1805 1810 1815	
gag atg gag cgg gca cgc tta gaa aag ttg gaa ctg gac gcc agg	5499
Glu Met Glu Arg Ala Arg Leu Glu Lys Leu Glu Leu Asp Ala Arg	
1820 1825 1830	
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Lys Leu Gln Gln Glu Leu Asp Gln Arg Asn Arg Glu Lys Leu Ser	
1835 1840 1845	
ctg cat caa gac ctg gca gtg gtg cag cag cag cta caa gaa aaa	5589
Leu His Gln Asp Leu Ala Val Val Gln Gln Gln Leu Gln Glu Lys	
1850 1855 1860	

cag gaa gca gta aac tca tta	cag aag gaa cta act	gat gtc cag	5634
Gln Glu Ala Val Asn Ser Leu	Gln Lys Glu Leu Thr	Asp Val Gln	
1865	1870	1875	
gag cat ttg gac cta gca gaa	cag gag gtg ctc tgc	acc acc aag	5679
Glu His Leu Asp Leu Ala Glu	Gln Glu Val Leu Cys	Thr Thr Lys	
1880	1885	1890	
cgc aag gac gca ctg ctc agc	gaa cag acc agg ctc	gag aag gac	5724
Arg Lys Asp Ala Leu Leu Ser	Glu Gln Thr Arg Leu	Glu Lys Asp	
1895	1900	1905	
gtg ggt gaa tgg acg aag aag	ttt gaa gac tgc cag	aaa gaa ggg	5769
Val Gly Glu Trp Thr Lys Lys	Phe Glu Asp Cys Gln	Lys Glu Gly	
1910	1915	1920	
gag aca aag cag caa cag ctt	caa ggg ctt cag aag	gag att gaa	5814
Glu Thr Lys Gln Gln Gln Leu	Gln Gly Leu Gln Lys	Glu Ile Glu	
1925	1930	1935	
gga aac gag gcg aag cta gcc	caa caa gaa atg atg	ttt cag aga	5859
Gly Asn Glu Ala Lys Leu Ala	Gln Gln Glu Met Met	Phe Gln Arg	
1940	1945	1950	
ctc cag aaa gag cga gaa tgt	gaa gaa aaa aag tta	gaa gct agt	5904
Leu Gln Lys Glu Arg Glu Cys	Glu Glu Lys Lys Leu	Glu Ala Ser	
1955	1960	1965	
aaa gtg act ctg aag gag cag	cag caa cag ctg gaa	aag gaa ttg	5949
Lys Val Thr Leu Lys Glu Gln	Gln Gln Gln Leu Glu	Lys Glu Leu	
1970	1975	1980	
atg gag cag aaa ggc aag ctg	gac cag gtg ctc gct	aag ctc ttg	5994
Met Glu Gln Lys Gly Lys Leu	Asp Gln Val Leu Ala	Lys Leu Leu	
1985	1990	1995	
gtg gct gag gag cgt gtc agg	acc ttg cag gag gag	gga agg tgg	6039
Val Ala Glu Glu Arg Val Arg	Thr Leu Gln Glu Glu	Gly Arg Trp	
2000	2005	2010	
agc gag acc ctg gag aag acg	ctc tcc cag acc aag	cga cag ctt	6084
Ser Glu Thr Leu Glu Lys Thr	Leu Ser Gln Thr Lys	Arg Gln Leu	
2015	2020	2025	
tca gaa cgg gag cag cag tta	ctg gcc aag tca gac	gag ctg ctg	6129
Ser Glu Arg Glu Gln Gln Leu	Leu Ala Lys Ser Asp	Glu Leu Leu	
2030	2035	2040	
gcc ctg cag aag gag acg gac	tcc atg agg gcg gac	ttc agc ctc	6174
Ala Leu Gln Lys Glu Thr Asp	Ser Met Arg Ala Asp	Phe Ser Leu	
2045	2050	2055	
ttg cgc aac cag ttc ctg aca	gaa aga aag aaa gcc	gag aag cag	6219
Leu Arg Asn Gln Phe Leu Thr	Glu Arg Lys Lys Ala	Glu Lys Gln	
2060	2065	2070	
gtg gcc agc ctg aag gaa gcc	ctt aag atc cag cgg	agc caa ctg	6264
Val Ala Ser Leu Lys Glu Ala	Leu Lys Ile Gln Arg	Ser Gln Leu	
2075	2080	2085	

gag aag aac ctt ctg gag caa aag cag gag aac agc tgc atg cag	6309
Glu Lys Asn Leu Leu Glu Gln Lys Gln Glu Asn Ser Cys Met Gln	
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agg gag atg gca acc atc gaa cag gtg gcc cag gac aac cac gag	6354
Arg Glu Met Ala Thr Ile Glu Gln Val Ala Gln Asp Asn His Glu	
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cgg gcc cgg cgc cta atg agg gag ctc aac cag atg cag cgc gag	6399
Arg Ala Arg Arg Leu Met Arg Glu Leu Asn Gln Met Gln Arg Glu	
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tac gtg gag ctc agg aaa cag atg aca aac caa aag gat ttg gaa	6444
Tyr Val Glu Leu Arg Lys Gln Met Thr Asn Gln Lys Asp Leu Glu	
2135 2140 2145	
aga aga cag atg gaa atc agt gat gcg atg caa gca ctt aaa tgt	6489
Arg Arg Gln Met Glu Ile Ser Asp Ala Met Gln Ala Leu Lys Cys	
2150 2155 2160	
gag gtg aaa gat gaa atc cga acc agc ctg aag aat ctc aac cag	6534
Glu Val Lys Asp Glu Ile Arg Thr Ser Leu Lys Asn Leu Asn Gln	
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Phe Leu Pro Glu Leu Pro Ala Asp Leu Glu Ala Leu Leu Glu Arg	
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Asn Glu Asn Leu Gly Gly Gly Leu Glu Ser Leu Lys Glu Asn Phe	
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ccg ttt acc gtg agc gac aga cca tca tct tgc gaa gag aaa ctg	6669
Pro Phe Thr Val Ser Asp Arg Pro Ser Ser Cys Glu Glu Lys Leu	
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Asn Phe Gly Gln Ala His Val Ala Asp Glu Gln Trp Arg Gly Glu	
2225 2230 2235	
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Ala Leu Arg Glu Lys Leu Arg His Arg Glu Asp Arg Leu Lys Ala	
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Gln Leu Arg Arg Cys Met Ser Lys Gln Ala Glu Val Leu Ser Glu	
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Gly Arg Arg Arg Thr Glu Gly Thr Leu His Ser Leu Arg Arg Gln	
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Val Asp Ala Leu Gly Glu Leu Val Thr Ser Thr Ser Gly Asp Ser	
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Ala Ser Thr Arg Ser Leu Ser Arg Thr Glu Gly Ser Leu Ala Glu	
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 Asp Glu Pro Pro Gly Pro Ser Gln Ser Ser Arg Arg Leu Pro Arg
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 Gly Pro Ser Pro Arg Leu Asp Ala His Arg Pro
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 Val Leu Asp Tyr Gln Asp His Lys Glu Ala Asp Ser His Ala Gly Val
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 Arg Tyr Ile Thr Glu Ala Leu Val Arg Lys Leu Thr Lys Gln Asp Asn
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 Lys Glu Gln Gln Leu Asp Ile Met Asn Lys Gln Tyr Lys Gln Leu Glu
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 Ser Arg Leu Asp Glu Ile Leu Ser Arg Ile Ala Lys Glu Thr Glu Glu
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 Glu Ala Leu Lys Lys Asp Leu Glu Ser Val Ile Ser Gly Leu Gln Glu
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 Tyr Leu Glu Thr Val Lys Gly Gln Ala Arg Gln Ala Gln Asn Glu Cys
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 Tyr Glu Ala Glu Leu Glu Ala Gln Leu Lys Ile Arg Asp Ala Glu Ala
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Pro	Val	Gly	Val	Leu	His	Cys	Asn	Val	Pro	Glu	His	His	Asn	Leu
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Glu	Asn	Glu	Val	Ser	Arg	Leu	Glu	Asp	Ile	Met	Gln	His	Leu	Lys
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Gln	Glu	Lys	Lys	Asp	Leu	Glu	His	Glu	Val	Glu	Glu	Leu	His	Arg
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Thr	Ile	Gln	Lys	His	Gln	Gln	Arg	Lys	Asp	Phe	Ile	Asp	Gly	Asn
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Val	Glu	Ser	Leu	Val	Asn	Asp	Leu	Glu	Ile	Glu	Lys	Ser	Leu	Lys
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His	His	Glu	Asp	Ile	Val	Asp	Glu	Ile	Glu	Cys	Ile	Glu	Arg	Thr
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 Arg Ser Leu Ser Pro Leu Ser Gly Ser Glu Thr Leu Pro Phe His Phe
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 gga gga ccg tgg cat gag caa gtt gag att aca gat gaa agc aca gtg 192
 Gly Gly Pro Trp His Glu Gln Val Glu Ile Thr Asp Glu Ser Thr Val
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 gtt tta gac tac caa gac cat aaa gaa gct gat tca cat gca gga gtc 240
 Val Leu Asp Tyr Gln Asp His Lys Glu Ala Asp Ser His Ala Gly Val
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115

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Gln	Lys	Thr	Met	Glu	Leu	Met	Arg	Ala	Cys	Gln	Lys	Gln	Tyr	Glu	Met	
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Ala	Ser	Pro	Thr	Asp	Ile	Gln	Leu	Glu	Asp	Lys	Glu	Lys	Lys	Ile	Ser	
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Ala	Ala	Gln	Thr	Arg	Leu	Ser	Glu	Leu	His	Asp	Glu	Ile	Glu	Lys	Ala	
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465					470					475					480	
gct	ata	caa	ctt	aaa	aaa	att	tca	gaa	gcg	gag	aaa	gac	ctt	ctt	ttc	1488
Ala	Ile	Gln	Leu	Lys	Lys	Ile	Ser	Glu	Ala	Glu	Lys	Asp	Leu	Leu	Phe	
				485					490					495		
aag	cag	ttg	agt	ggt	agg	ata	cag	ctt	ctc	aat	aaa	tta	cgc	caa	gaa	1536
Lys	Gln	Leu	Ser	Gly	Arg	Ile	Gln	Leu	Leu	Asn	Lys	Leu	Arg	Gln	Glu	
			500					505					510			
gct	gtg	gat	cta	gaa	aca	cag	atg	gaa	aag	caa	agg	caa	gaa	att	ggt	1584
Ala	Val	Asp	Leu	Glu	Thr	Gln	Met	Glu	Lys	Gln	Arg	Gln	Glu	Ile	Gly	
		515					520					525				
gaa	aag	cag	aat	gag	atc	aag	gac	ctg	gaa	ata	gtc	aca	gat	agc	ctg	1632
Glu	Lys	Gln	Asn	Glu	Ile	Lys	Asp	Leu	Glu	Ile	Val	Thr	Asp	Ser	Leu	
	530					535					540					
gat	tcc	aga	gac	cca	aaa	cat	tgc	cat	atg	aag	gct	cag	aaa	aga	ggt	1680
Asp	Ser	Arg	Asp	Pro	Lys	His	Cys	His	Met	Lys	Ala	Gln	Lys	Arg	Gly	
545					550					555					560	

aaa gaa caa caa ctt gac att atg aac aag cag tac aaa cag ctt gaa	1728
Lys Glu Gln Gln Leu Asp Ile Met Asn Lys Gln Tyr Lys Gln Leu Glu	
565 570 575	
agc cgt ttg gat gag ata ctt tct aga att gcc aaa gaa act gaa gag	1776
Ser Arg Leu Asp Glu Ile Leu Ser Arg Ile Ala Lys Glu Thr Glu Glu	
580 585 590	
att aag gac ctt gaa gaa cag ctt act gaa gga caa ata gcc gca aac	1824
Ile Lys Asp Leu Glu Glu Gln Leu Thr Glu Gly Gln Ile Ala Ala Asn	
595 600 605	
gaa gcc ctg aag aag gac tta gaa agt gtc atc agt ggg ttg caa gaa	1872
Glu Ala Leu Lys Lys Asp Leu Glu Ser Val Ile Ser Gly Leu Gln Glu	
610 615 620	
tac ctg gag act gtc aaa ggt cag gcc cgt cag gcc cag aat gag tgc	1920
Tyr Leu Glu Thr Val Lys Gly Gln Ala Arg Gln Ala Gln Asn Glu Cys	
625 630 635 640	
aga aag cta cag gat gag aag gag aca ttg ctg cag aga ttg agt gag	1968
Arg Lys Leu Gln Asp Glu Lys Glu Thr Leu Leu Gln Arg Leu Ser Glu	
645 650 655	
gtc gag cag gag agg gac caa ctg gaa ata gtg gcc ata gat gca gaa	2016
Val Glu Gln Glu Arg Asp Gln Leu Glu Ile Val Ala Ile Asp Ala Glu	
660 665 670	
aat atg agg aag gag ctc gca gaa ctg gag aat gcc ctc cag gag cag	2064
Asn Met Arg Lys Glu Leu Ala Glu Leu Glu Asn Ala Leu Gln Glu Gln	
675 680 685	
cat gag gtg aat ata tct ctg cag cag acc cag gga gat ctc agt gcc	2112
His Glu Val Asn Ile Ser Leu Gln Gln Thr Gln Gly Asp Leu Ser Ala	
690 695 700	
tat gag gct gag cta gag gct cag ctg aaa ata cgg gat gct gaa gcc	2160
Tyr Glu Ala Glu Leu Glu Ala Gln Leu Lys Ile Arg Asp Ala Glu Ala	
705 710 715 720	
aac cag ctc aag gag gag ttg gaa aaa ctt aga agg ttg agc cag tta	2208
Asn Gln Leu Lys Glu Glu Leu Glu Lys Leu Arg Arg Leu Ser Gln Leu	
725 730 735	
gaa caa tcg gcc ctt caa gca gag ctt gag aag gaa aag caa gcc ttc	2256
Glu Gln Ser Ala Leu Gln Ala Glu Leu Glu Lys Glu Lys Gln Ala Phe	
740 745 750	
aag act gct gtc aaa aaa gcc cag ctc tca gaa gga aag gac caa gaa	2304
Lys Thr Ala Val Lys Lys Ala Gln Leu Ser Glu Gly Lys Asp Gln Glu	
755 760 765	
aat agt gag ctc cgc aca caa ctc caa cag ctg cag gat gac aat gac	2352
Asn Ser Glu Leu Arg Thr Gln Leu Gln Gln Leu Gln Asp Asp Asn Asp	
770 775 780	
cta ttg aaa cag caa ctt aaa gat ttc cag agt cac ctt aac cat gtg	2400
Leu Leu Lys Gln Gln Leu Lys Asp Phe Gln Ser His Leu Asn His Val	
785 790 795 800	

ggt gat ggt ttg att cgt cca gaa gaa gtg gca gct tgt gtg gat gag Val Asp Gly Leu Ile Arg Pro Glu Glu Val Ala Ala Cys Val Asp Glu	2448
805 810 815	
cta agg aaa aaa ctg aag tca gga gct ggg gaa atg aga atc cat act Leu Arg Lys Lys Leu Lys Ser Gly Ala Gly Glu Met Arg Ile His Thr	2496
820 825 830	
cct tca gat gtc tta ggg aaa agt ctt gct gac ttg cag aag caa ttc Pro Ser Asp Val Leu Gly Lys Ser Leu Ala Asp Leu Gln Lys Gln Phe	2544
835 840 845	
agt gag atc ctg gca cgc tcc cag tgg gaa aga cag gaa gca caa gtg Ser Glu Ile Leu Ala Arg Ser Gln Trp Glu Arg Gln Glu Ala Gln Val	2592
850 855 860	
aga gag aga aaa ctc cag gag gaa atg gct ctg caa caa gag aaa ctg Arg Glu Arg Lys Leu Gln Glu Glu Met Ala Leu Gln Gln Glu Lys Leu	2640
865 870 875 880	
gcg agc gga caa gag gag ttc agg cac gcc tgc gag agg gcc ctg gaa Ala Ser Gly Gln Glu Glu Phe Arg His Ala Cys Glu Arg Ala Leu Glu	2688
885 890 895	
gcc cga att agt ttt gat aag agg cag cac gaa gca aga atc cag cag Ala Arg Ile Ser Phe Asp Lys Arg Gln His Glu Ala Arg Ile Gln Gln	2736
900 905 910	
ttg gag aat gaa att cac tat ttg caa gaa aat cta aaa agt atg gag Leu Glu Asn Glu Ile His Tyr Leu Gln Glu Asn Leu Lys Ser Met Glu	2784
915 920 925	
gaa atc caa ggt ctc aca gac ctc caa ctt cag gaa gct gat gaa gag Glu Ile Gln Gly Leu Thr Asp Leu Gln Leu Gln Glu Ala Asp Glu Glu	2832
930 935 940	
aag gag aga att ctg gcc caa ctc cgg gag tta gag aaa aag aag aaa Lys Glu Arg Ile Leu Ala Gln Leu Arg Glu Leu Glu Lys Lys Lys Lys	2880
945 950 955 960	
ctt gag gat gcc aag tct cag gag cag ttt ctt gga tta gat aga gaa Leu Glu Asp Ala Lys Ser Gln Glu Gln Phe Leu Gly Leu Asp Arg Glu	2928
965 970 975	
ttg aag aag cta aag aaa gct gtg gct gcc tct gat aag ctg gcc aca Leu Lys Lys Leu Lys Lys Ala Val Ala Ala Ser Asp Lys Leu Ala Thr	2976
980 985 990	
gct gag ctc acc att gcc aaa gac cag ctc aag tcc ctt cat gga act Ala Glu Leu Thr Ile Ala Lys Asp Gln Leu Lys Ser Leu His Gly Thr	3024
995 1000 1005	
gtg atg aaa att aac cag gag cga gca gag gag ctg cag gag acg Val Met Lys Ile Asn Gln Glu Arg Ala Glu Glu Leu Gln Glu Thr	3069
1010 1015 1020	
gag agg ttc agc aga aag gca gca caa gca gct agg gat ctg atc Glu Arg Phe Ser Arg Lys Ala Ala Gln Ala Ala Arg Asp Leu Ile	3114
1025 1030 1035	

cga gca gaa gcg gag att gaa ctc ctg cag aag ctt ctc aga gat	3159
Arg Ala Glu Ala Glu Ile Glu Leu Leu Gln Lys Leu Leu Arg Asp	
1040 1045 1050	
aaa gag gag cag ttt cga aat gag att gag aaa gta gat gtc ggc	3204
Lys Glu Glu Gln Phe Arg Asn Glu Ile Glu Lys Val Asp Val Gly	
1055 1060 1065	
tct gga gga gca aag tca cag atg ctg gag atg gag aaa cta aat	3249
Ser Gly Gly Ala Lys Ser Gln Met Leu Glu Met Glu Lys Leu Asn	
1070 1075 1080	
gag aca atg gag agg caa aga aca gag att gct agg ctg agg aat	3294
Glu Thr Met Glu Arg Gln Arg Thr Glu Ile Ala Arg Leu Arg Asn	
1085 1090 1095	
tta cta gac ctc acc ggg gct gat aac aaa gga aac ttt gaa aat	3339
Leu Leu Asp Leu Thr Gly Ala Asp Asn Lys Gly Asn Phe Glu Asn	
1100 1105 1110	
gtt ttg gaa gaa att gct gaa ctt cga cgt gaa gtt tct cat cag	3384
Val Leu Glu Glu Ile Ala Glu Leu Arg Arg Glu Val Ser His Gln	
1115 1120 1125	
aat gat tac atc agc agc atg aca gat cct ttc aaa aga cga ggc	3429
Asn Asp Tyr Ile Ser Ser Met Thr Asp Pro Phe Lys Arg Arg Gly	
1130 1135 1140	
tat tgg tac ttt atg cca cca cca tca tca tca aaa gtt tcc agc	3474
Tyr Trp Tyr Phe Met Pro Pro Pro Ser Ser Ser Lys Val Ser Ser	
1145 1150 1155	
cac agt tcc cag gcc acc aag gac tct ggt gtt ggc cta aag tac	3519
His Ser Ser Gln Ala Thr Lys Asp Ser Gly Val Gly Leu Lys Tyr	
1160 1165 1170	
aca gcc tcc act ccg gtt aga aaa cca cat cgt gga cgg cag gat	3564
Thr Ala Ser Thr Pro Val Arg Lys Pro His Arg Gly Arg Gln Asp	
1175 1180 1185	
gga aag gag aac agt ggg cct cca cct gcc tca gga tac tgg gtg	3609
Gly Lys Glu Asn Ser Gly Pro Pro Pro Ala Ser Gly Tyr Trp Val	
1190 1195 1200	
tat tct cct atc agg agt ggg tta cat aaa tcg ttc tca aat aga	3654
Tyr Ser Pro Ile Arg Ser Gly Leu His Lys Ser Phe Ser Asn Arg	
1205 1210 1215	
gac gca gac agt gga gga gat agc cag gaa gag agc gag cta gat	3699
Asp Ala Asp Ser Gly Gly Asp Ser Gln Glu Glu Ser Glu Leu Asp	
1220 1225 1230	
gac caa gaa gac cac cca ttt gta cct cct cct gga tac atg atg	3744
Asp Gln Glu Asp His Pro Phe Val Pro Pro Pro Gly Tyr Met Met	
1235 1240 1245	
tac act gtg ttt cct gat ggt tct cct gta ccc cag ggc atg gcc	3789
Tyr Thr Val Phe Pro Asp Gly Ser Pro Val Pro Gln Gly Met Ala	
1250 1255 1260	

120

ctg	tat	gca	ccc	cct	cct	ccc	ttg	ccc	aac	aat	agc	cag	cct	ctt	3834
Leu	Tyr	Ala	Pro	Pro	Pro	Pro	Leu	Pro	Asn	Asn	Ser	Gln	Pro	Leu	
	1265					1270					1275				
gac	ctt	ggc	act	gtt	gtt	tat	ggc	cca	cct	cct	gtt	ggg	gct	ccc	3879
Asp	Leu	Gly	Thr	Val	Val	Tyr	Gly	Pro	Pro	Pro	Val	Gly	Ala	Pro	
	1280					1285					1290				
atc	gtg	tat	ggg	cct	cca	cct	ccc	aac	ttc	tcc	gta	ccc	ctc	atc	3924
Ile	Val	Tyr	Gly	Pro	Pro	Pro	Pro	Asn	Phe	Ser	Val	Pro	Leu	Ile	
	1295					1300					1305				
ccc	gtg	ggt	gtg	ctg	cac	tgc	aat	gtc	cca	gaa	cac	cat	aac	ttg	3969
Pro	Val	Gly	Val	Leu	His	Cys	Asn	Val	Pro	Glu	His	His	Asn	Leu	
	1310					1315					1320				
gag	aat	gaa	gtt	tct	aga	tta	gaa	gac	ata	atg	cag	cat	tta	aaa	4014
Glu	Asn	Glu	Val	Ser	Arg	Leu	Glu	Asp	Ile	Met	Gln	His	Leu	Lys	
	1325					1330					1335				
tct	ggg	aaa	cgg	gaa	cag	tgc	atg	aaa	aca	ccc	aag	ctg	cag	tcg	4059
Ser	Gly	Lys	Arg	Glu	Gln	Cys	Met	Lys	Thr	Pro	Lys	Leu	Gln	Ser	
	1340					1345					1350				
gag	aaa	gaa	ctc	gca	gag	ctg	cag	cat	aac	att	gat	ggt	ctt	ttg	4104
Glu	Lys	Glu	Leu	Ala	Glu	Leu	Gln	His	Asn	Ile	Asp	Gly	Leu	Leu	
	1355					1360					1365				
caa	gag	aag	aaa	gac	tta	gag	cat	gaa	gta	gaa	gaa	tta	cat	aga	4149
Gln	Glu	Lys	Lys	Asp	Leu	Glu	His	Glu	Val	Glu	Glu	Leu	His	Arg	
	1370					1375					1380				
acc	atc	caa	aaa	cat	caa	cag	cga	aaa	gat	ttc	att	gat	gga	aac	4194
Thr	Ile	Gln	Lys	His	Gln	Gln	Arg	Lys	Asp	Phe	Ile	Asp	Gly	Asn	
	1385					1390					1395				
gtt	gag	agt	ctt	gtg	aat	gat	cta	gaa	ata	gag	aag	tca	ctc	aaa	4239
Val	Glu	Ser	Leu	Val	Asn	Asp	Leu	Glu	Ile	Glu	Lys	Ser	Leu	Lys	
	1400					1405					1410				
cac	cat	gaa	gat	att	gtt	gat	gaa	att	gaa	tgt	att	gag	agg	acc	4284
His	His	Glu	Asp	Ile	Val	Asp	Glu	Ile	Glu	Cys	Ile	Glu	Arg	Thr	
	1415					1420					1425				
ctt	ctg	aag	cgc	cgt	gca	gag	ctc	agg	gaa	gcc	gac	cgg	ctg	ctg	4329
Leu	Leu	Lys	Arg	Arg	Ala	Glu	Leu	Arg	Glu	Ala	Asp	Arg	Leu	Leu	
	1430					1435					1440				
acg	gag	gct	gaa	agt	gaa	ctt	tca	tgc	acg	aaa	gag	aaa	aca	aaa	4374
Thr	Glu	Ala	Glu	Ser	Glu	Leu	Ser	Cys	Thr	Lys	Glu	Lys	Thr	Lys	
	1445					1450					1455				
cat	gct	gtt	gag	aag	ttc	act	gat	gcc	aag	aga	aat	tta	ttg	caa	4419
His	Ala	Val	Glu	Lys	Phe	Thr	Asp	Ala	Lys	Arg	Asn	Leu	Leu	Gln	
	1460					1465					1470				
act	gag	aaa	gat	gct	gag	gag	tta	gaa	agg	aga	gcc	cag	gaa	act	4464
Thr	Glu	Lys	Asp	Ala	Glu	Glu	Leu	Glu	Arg	Arg	Ala	Gln	Glu	Thr	
	1475					1480					1485				

gcc att aac ctc gtc aaa gcc gac cag cag ctg aga ttg ctc cag	4509
Ala Ile Asn Leu Val Lys Ala Asp Gln Gln Leu Arg Leu Leu Gln	
1490 1495 1500	
gct gac acg aag gat ttg gag cag cac aaa atg gag caa gag gaa	4554
Ala Asp Thr Lys Asp Leu Glu Gln His Lys Met Glu Gln Glu Glu	
1505 1510 1515	
atc ttg aaa gaa ata aac aaa gtt gtt gca gca aaa gac tca gac	4599
Ile Leu Lys Glu Ile Asn Lys Val Val Ala Ala Lys Asp Ser Asp	
1520 1525 1530	
ttc cag agc cta aac aag aag aag gaa gta ctg aca gga gag ctg	4644
Phe Gln Ser Leu Asn Lys Lys Lys Glu Val Leu Thr Gly Glu Leu	
1535 1540 1545	
cag aaa ctc cag aag gac att gag act gca cgg cac aat gag gat	4689
Gln Lys Leu Gln Lys Asp Ile Glu Thr Ala Arg His Asn Glu Asp	
1550 1555 1560	
cag cac ctg cag gtc ctt aaa gag tcg gag acc ctc ctg cag gcc	4734
Gln His Leu Gln Val Leu Lys Glu Ser Glu Thr Leu Leu Gln Ala	
1565 1570 1575	
aag aaa gct gag ctg gaa aat ctg aaa agc cag gtg tca gga cag	4779
Lys Lys Ala Glu Leu Glu Asn Leu Lys Ser Gln Val Ser Gly Gln	
1580 1585 1590	
cag cag gag atg gcc gtc ttg gac agg gag tta gga cac aag aag	4824
Gln Gln Glu Met Ala Val Leu Asp Arg Glu Leu Gly His Lys Lys	
1595 1600 1605	
gaa gag ctg cat ctc ctc cag gaa agc atg gtc cag gcc aaa gct	4869
Glu Glu Leu His Leu Leu Gln Glu Ser Met Val Gln Ala Lys Ala	
1610 1615 1620	
gac ctc cag gaa gca ctg aga cta gga gaa agt gaa gta act gag	4914
Asp Leu Gln Glu Ala Leu Arg Leu Gly Glu Ser Glu Val Thr Glu	
1625 1630 1635	
aag tgc aat cac att agg gaa gta aaa tct ctt ctg gaa gaa ctc	4959
Lys Cys Asn His Ile Arg Glu Val Lys Ser Leu Leu Glu Glu Leu	
1640 1645 1650	
agt ttt cag aaa gga gaa ctg aat gtc cag atc agt gaa aaa aaa	5004
Ser Phe Gln Lys Gly Glu Leu Asn Val Gln Ile Ser Glu Lys Lys	
1655 1660 1665	
act caa ctt gca ctc ata aag cag gaa att gaa aaa gag gaa gac	5049
Thr Gln Leu Ala Leu Ile Lys Gln Glu Ile Glu Lys Glu Glu Asp	
1670 1675 1680	
aat ctt cag gta gtt tta ggg caa atg tct aaa cat aaa act gaa	5094
Asn Leu Gln Val Val Leu Gly Gln Met Ser Lys His Lys Thr Glu	
1685 1690 1695	
cta aag aat att ctg gac atg ttg caa ctt gaa aat aat gag ctg	5139
Leu Lys Asn Ile Leu Asp Met Leu Gln Leu Glu Asn Asn Glu Leu	
1700 1705 1710	

caa ggt	ttg aag	ctc caa	cat gac	caa aag	atg tct	gaa tta	gag	5184
Gln Gly	Leu Lys	Leu Gln	His Asp	Gln Lys	Met Ser	Glu Leu	Glu	
1715			1720		1725			
aag act	cgg gtt	gaa gtg	ctg gag	gag gag	aaa ctg	gag tta	gag agt	5229
Lys Thr	Arg Val	Glu Val	Leu Glu	Glu Glu	Lys Leu	Glu Leu	Glu Ser	
1730			1735		1740			
ctg cag	cag gca	gcc ctg	cga cag	aga ggg	gag ata	gag tgg	cag	5274
Leu Gln	Gln Ala	Ala Leu	Arg Gln	Arg Gly	Glu Ile	Glu Trp	Gln	
1745			1750		1755			
aag cag	ctc ctc	cag agg	aac aca	cag gaa	gta gag	cgg atg	act	5319
Lys Gln	Leu Leu	Gln Arg	Asn Thr	Gln Glu	Val Glu	Arg Met	Thr	
1760			1765		1770			
gct gag	acc cga	gca tta	cag tca	tgt gtt	gag tct	ttg tgc	aaa	5364
Ala Glu	Thr Arg	Ala Leu	Gln Ser	Cys Val	Glu Ser	Leu Cys	Lys	
1775			1780		1785			
gaa aag	caa gat	ctc gaa	gaa gaa	aaa cag	gac agc	tgg gaa	aag aag	5409
Glu Lys	Gln Asp	Leu Glu	Glu Glu	Lys Gln	Asp Ser	Trp Glu	Lys Lys	
1790			1795		1800			
ttg gca	cag acc	aaa cgg	gtt cta	gca gct	gca gaa	gag gac	agc	5454
Leu Ala	Gln Thr	Lys Arg	Val Leu	Ala Ala	Ala Glu	Glu Asp	Ser	
1805			1810		1815			
gag atg	gag cgg	gca cgc	tta gaa	aag ttg	gaa ctg	gac gcc	agg	5499
Glu Met	Glu Arg	Ala Arg	Leu Glu	Lys Leu	Glu Leu	Asp Ala	Arg	
1820			1825		1830			
aag ctg	cag cag	gag ttg	gac caa	cga aac	agg gag	aag ctc	tcc	5544
Lys Leu	Gln Gln	Glu Leu	Asp Gln	Arg Asn	Arg Glu	Lys Leu	Ser	
1835			1840		1845			
ctg cat	caa gac	ctg gca	gtg gtg	cag cag	cag cta	caa gaa	aaa	5589
Leu His	Gln Asp	Leu Ala	Val Val	Gln Gln	Gln Leu	Gln Glu	Lys	
1850			1855		1860			
cag gaa	gca gta	aac tca	tta cag	aag gaa	cta gct	gat gtc	cag	5634
Gln Glu	Ala Val	Asn Ser	Leu Gln	Lys Glu	Leu Ala	Asp Val	Gln	
1865			1870		1875			
gag cat	ttg gac	cta gca	gaa gag	gag gtg	ctc tgc	acc acc	aag	5679
Glu His	Leu Asp	Leu Ala	Glu Gln	Glu Val	Leu Cys	Thr Thr	Lys	
1880			1885		1890			
cgc aag	gac gca	ctg ctc	agc gaa	cag acc	agg ctc	gag aag	gac	5724
Arg Lys	Asp Ala	Leu Leu	Ser Glu	Gln Thr	Arg Leu	Glu Lys	Asp	
1895			1900		1905			
gtg ggt	gaa tgg	acg aag	aag ttt	gaa gac	tgc cag	aaa gaa	ggg	5769
Val Gly	Glu Trp	Thr Lys	Lys Phe	Glu Asp	Cys Gln	Lys Glu	Gly	
1910			1915		1920			
gag aca	aag cag	caa cag	ctt caa	ggg ctt	cag aag	gag att	gaa	5814
Glu Thr	Lys Gln	Gln Gln	Leu Gln	Gly Leu	Gln Lys	Glu Ile	Glu	
1925			1930		1935			

gga aac gag gcg aag cta gcc caa caa gaa atg atg ttt cag aga	5859
Gly Asn Glu Ala Lys Leu Ala Gln Gln Glu Met Met Phe Gln Arg	
1940 1945 1950	
ctc cag aaa gag cga gaa tgt gaa gaa aaa aag tta gaa gct agt	5904
Leu Gln Lys Glu Arg Glu Cys Glu Glu Lys Lys Leu Glu Ala Ser	
1955 1960 1965	
aaa gtg act ctg aag gag cag cag caa cag ctg gaa aag gaa ttg	5949
Lys Val Thr Leu Lys Glu Gln Gln Gln Glu Leu Glu Lys Glu Leu	
1970 1975 1980	
atg gag cag aaa ggc aag ctg gac cag gtg ctc gct aag ctc ttg	5994
Met Glu Gln Lys Gly Lys Leu Asp Gln Val Leu Ala Lys Leu Leu	
1985 1990 1995	
gtg gct gag gag cgt gtc agg acc ttg cag gag gag gga agg tgg	6039
Val Ala Glu Glu Arg Val Arg Thr Leu Gln Glu Glu Gly Arg Trp	
2000 2005 2010	
agc gag acc ctg gag aag acg ctc tcc cag acc aag cga cag ctt	6084
Ser Glu Thr Leu Glu Lys Thr Leu Ser Gln Thr Lys Arg Gln Leu	
2015 2020 2025	
tca gaa cgg gag cag cag tta ctg gcc aag tca gac gag ctg ctg	6129
Ser Glu Arg Glu Gln Gln Leu Leu Ala Lys Ser Asp Glu Leu Leu	
2030 2035 2040	
gcc ctg cag aag gag acg gac tcc atg agg gcg gac ttc agc ctc	6174
Ala Leu Gln Lys Glu Thr Asp Ser Met Arg Ala Asp Phe Ser Leu	
2045 2050 2055	
ttg cgc aac cag ttc ctg aca gaa aga aag aaa gcc gag aag cag	6219
Leu Arg Asn Gln Phe Leu Thr Glu Arg Lys Lys Ala Glu Lys Gln	
2060 2065 2070	
gtg gcc agc ctg aag gaa gcc ctt aag atc cag cgg agc caa ctg	6264
Val Ala Ser Leu Lys Glu Ala Leu Lys Ile Gln Arg Ser Gln Leu	
2075 2080 2085	
gag aag aac ctt ctg gag caa aag cag gag aac agc tgc atg cag	6309
Glu Lys Asn Leu Leu Glu Gln Lys Gln Glu Asn Ser Cys Met Gln	
2090 2095 2100	
agg gag atg gca acc atc gaa cag gtg gcc cag gac aac cac gag	6354
Arg Glu Met Ala Thr Ile Glu Gln Val Ala Gln Asp Asn His Glu	
2105 2110 2115	
cgg gcc cgg cgc ctg atg agg gag ctc aac cag atg cag cgc gag	6399
Arg Ala Arg Arg Leu Met Arg Glu Leu Asn Gln Met Gln Arg Glu	
2120 2125 2130	
tac gtg gag ctc agg aaa cag atg aca aac caa aag gat ttg gaa	6444
Tyr Val Glu Leu Arg Lys Gln Met Thr Asn Gln Lys Asp Leu Glu	
2135 2140 2145	
aga aga cag atg gaa atc agt gat gcg atg caa gca ctt aaa tgt	6489
Arg Arg Gln Met Glu Ile Ser Asp Ala Met Gln Ala Leu Lys Cys	
2150 2155 2160	

gag	gtg	aaa	gat	gaa	atc	cga	acc	agc	ctg	aag	aat	ctc	aac	cag	6534
Glu	Val	Lys	Asp	Glu	Ile	Arg	Thr	Ser	Leu	Lys	Asn	Leu	Asn	Gln	
	2165					2170					2175				
ttt	ctt	cca	gag	ctg	cca	gcg	gac	ctg	gag	gcc	ctt	ctg	gaa	agg	6579
Phe	Leu	Pro	Glu	Leu	Pro	Ala	Asp	Leu	Glu	Ala	Leu	Leu	Glu	Arg	
	2180					2185					2190				
aat	gag	aac	ctt	gga	gga	ggc	ttg	gag	agc	ttg	aaa	gag	aat	ttc	6624
Asn	Glu	Asn	Leu	Gly	Gly	Gly	Leu	Glu	Ser	Leu	Lys	Glu	Asn	Phe	
	2195					2200					2205				
ccg	ttt	acc	gtg	agc	gac	aga	cca	tca	tct	tgc	gaa	gag	aaa	ctg	6669
Pro	Phe	Thr	Val	Ser	Asp	Arg	Pro	Ser	Ser	Cys	Glu	Glu	Lys	Leu	
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aat	ttt	ggc	cag	gct	cac	gtg	gcg	gat	gaa	cag	tgg	cgg	gga	gag	6714
Asn	Phe	Gly	Gln	Ala	His	Val	Ala	Asp	Glu	Gln	Trp	Arg	Gly	Glu	
	2225					2230					2235				
gca	ctc	cgg	gag	aag	ctg	cgc	cac	cgc	gag	gac	cgg	ctc	aag	gcc	6759
Ala	Leu	Arg	Glu	Lys	Leu	Arg	His	Arg	Glu	Asp	Arg	Leu	Lys	Ala	
	2240					2245					2250				
cag	ctg	cgc	cgc	tgc	atg	tcc	aag	cag	gcc	gag	gtg	ctg	agc	gag	6804
Gln	Leu	Arg	Arg	Cys	Met	Ser	Lys	Gln	Ala	Glu	Val	Leu	Ser	Glu	
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ggc	cgg	cgg	cgc	acg	gag	ggg	acc	ctg	cac	agc	ctg	cgg	cgg	cag	6849
Gly	Arg	Arg	Arg	Thr	Glu	Gly	Thr	Leu	His	Ser	Leu	Arg	Arg	Gln	
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gtg	gac	gcc	ctg	ggc	gag	ctg	gtc	acc	agc	act	tcc	ggg	gac	tcc	6894
Val	Asp	Ala	Leu	Gly	Glu	Leu	Val	Thr	Ser	Thr	Ser	Gly	Asp	Ser	
	2285					2290					2295				
gcg	tcc	acc	cgc	agt	ctg	tcg	cgc	acc	gag	ggc	tcg	ctc	gcc	gag	6939
Ala	Ser	Thr	Arg	Ser	Leu	Ser	Arg	Thr	Glu	Gly	Ser	Leu	Ala	Glu	
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gac	gaa	ccg	ccg	ggg	ccc	agc	cag	gag	ctg	cac	gtg	ctg	ggg	tcg	6984
Asp	Glu	Pro	Pro	Gly	Pro	Ser	Gln	Glu	Leu	His	Val	Leu	Gly	Ser	
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Gly	Gly	Ser	Asp	Arg	Gly	Gly	Gly	Arg	Gly	Gly	Gly	Arg	Lys	Gly	
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ctt	tcc	cga	cgc	cgc	cgc	tgg	aac	cac	gga	gaa	gcg	cgc	ctc	ggc	7074
Leu	Ser	Arg	Arg	Arg	Arg	Trp	Asn	His	Gly	Glu	Ala	Arg	Leu	Gly	
	2345					2350					2355				
ccg	cgg	agg	ccc	cca	cgg	gag	ggg	gca	ggg	cgg	ggc	gcg	gcc	ttc	7119
Pro	Arg	Arg	Pro	Pro	Arg	Glu	Gly	Ala	Gly	Arg	Gly	Ala	Ala	Phe	
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cga	gcc	ttg	gtc	tcc	tgc	tcc	cgc	cct	gca	gag	ctc	ccg	gcg	gct	7164
Arg	Ala	Leu	Val	Ser	Cys	Ser	Arg	Pro	Ala	Glu	Leu	Pro	Ala	Ala	
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ccc ccg agg ccc gtc gcc gcg gct gga cgc gca ccg acc ctg agg	7209
Pro Pro Arg Pro Val Ala Ala Ala Gly Arg Ala Pro Thr Leu Arg	
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Thr Arg Arg Thr Arg Arg Pro Gly Val Pro Ser Glu Arg Phe Leu	
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Arg Val Arg Gly His Gln Ala His Gly Lys Ala Arg Pro Cys Gly	
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Lys Ser Arg Glu Arg Asn Pro Asp Ala Arg Ala Gly Leu Trp Ala	
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ttg gaa acg tgt tgc cgt aaa agc agc gcc cgc ggc tgc gga ctt	7389
Leu Glu Thr Cys Cys Arg Lys Ser Ser Ala Arg Gly Cys Gly Leu	
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gaa gcc ccg aac tgc cgc cgt gcc cgg tgc gga gcg agc gtg cgg	7434
Glu Ala Pro Asn Cys Arg Arg Ala Arg Cys Gly Ala Ser Val Arg	
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Tyr Pro Leu Val Pro Arg Gly Arg Thr Gly Arg Gly Ala Val Thr	
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Pro Trp Gly Arg Leu Gln Ser Arg Gly Thr Arg Thr Thr Pro Arg	
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Pro Val Arg Arg Glu His Pro Gln His Gln Glu Arg Pro Pro Gly	
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Arg Val Thr Ala Ala His Thr Glu Thr Ala Pro Pro Arg Arg Val	
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Phe His Ala Arg Val Ala Val Gly Glu Val Ser Leu Gly Pro Gly	
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Arg Gly Leu Glu Arg Thr Arg Gly Gly Gly Gly Gly Ala Gly Ala	
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Gly Leu Leu Ala Glu Ala Ala Ala Thr Ala Arg Cys Ala Asp Pro	
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Ser Thr Asp Pro Ser Ala	
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<210> 28

<211> 2589

<212> PRT

<213> Canis familiaris

<400> 28

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Arg	Ser	Leu	Ser	Pro	Leu	Ser	Gly	Ser	Glu	Thr	Leu	Pro	Phe	His	Phe
		35					40					45			
Gly	Gly	Pro	Trp	His	Glu	Gln	Val	Glu	Ile	Thr	Asp	Glu	Ser	Thr	Val
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Val	Leu	Asp	Tyr	Gln	Asp	His	Lys	Glu	Ala	Asp	Ser	His	Ala	Gly	Val
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Arg	Tyr	Ile	Thr	Glu	Ala	Leu	Val	Arg	Lys	Leu	Thr	Lys	Gln	Asp	Asn
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Leu	Ala	Leu	Val	Lys	Ser	Leu	Asn	Leu	Ser	Leu	Ala	Lys	Gly	Gly	Gly
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Lys	Lys	Phe	Arg	Cys	Ile	Glu	Asn	Leu	Glu	Lys	Cys	Val	Lys	Leu	Glu
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Val	Leu	Asn	Leu	Ser	Tyr	Asn	Leu	Ile	Gly	Lys	Ile	Glu	Lys	Val	Asp
	130					135						140			
Lys	Leu	Leu	Lys	Leu	Arg	Glu	Leu	Asn	Leu	Ser	Tyr	Asn	Lys	Ile	Arg
145					150						155				160
Lys	Ile	Glu	Gly	Ile	Glu	Asn	Leu	Tyr	Asn	Leu	Gln	Lys	Leu	Asn	Leu
				165					170					175	
Ala	Gly	Asn	Glu	Ile	Glu	His	Ile	Pro	Val	Trp	Leu	Gly	Lys	Lys	Leu
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Lys	Ser	Leu	Arg	Ile	Leu	Asn	Leu	Lys	Gly	Asn	Lys	Ile	Ser	Ser	Leu
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Gln	Asp	Val	Ser	Lys	Leu	Lys	Pro	Leu	Gln	Asp	Leu	Thr	Ser	Leu	Ile
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Ile	Phe	His	Leu	Arg	Ser	Leu	Glu	Ser	Leu	Glu	Gly	Gln	Pro	Val	Thr
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Ser	Gln	Asp	Arg	Gln	Glu	Ala	Phe	Ala	Arg	Phe	Ser	Leu	Asp	Glu	Val
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		275					280					285			
Leu	Arg	Ser	Glu	Gln	Thr	Arg	Phe	Leu	Glu	Glu	Ile	Lys	Ser	Gln	Asp
	290					295						300			
Lys	Leu	Asn	Lys	Ser	Leu	Lys	Glu	Glu	Ala	Arg	Leu	Gln	Lys	Gln	Ser
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Tyr	Glu	Glu	Leu	Glu	Ser	Asn	Leu	Asn	Thr	Lys	Asn	Glu	Leu	Leu	Lys
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Gln	Lys	Thr	Met	Glu	Leu	Met	Arg	Ala	Cys	Gln	Lys	Gln	Tyr	Glu	Met
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Glu	Gln	Glu	Leu	Ala	Phe	Tyr	Lys	Ile	Asp	Ala	Lys	Phe	Glu	Pro	Leu
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Asn	Tyr	Tyr	Pro	Ser	Glu	Tyr	Val	Glu	Ile	Asp	Lys	Thr	Pro	Asp	Glu
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Ser	Pro	Tyr	Ile	Gly	Lys	Ser	Arg	Tyr	Lys	Arg	Asn	Met	Phe	Thr	Thr
385					390					395					400
Glu	Ser	Tyr	Ile	Ile	Ala	Asn	Ala	Gln	Thr	Val	Lys	Ile	Lys	Lys	Met
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Glu	Leu	Asp	Glu	Gly	Glu	Gln	Leu	Arg	Asn	Glu	His	Val	Asn	Leu	Gly
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Ala	Ser	Pro	Thr	Asp	Ile	Gln	Leu	Glu	Asp	Lys	Glu	Lys	Lys	Ile	Ser
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Ala	Ala	Gln	Thr	Arg	Leu	Ser	Glu	Leu	His	Asp	Glu	Ile	Glu	Lys	Ala
	450					455					460				
Glu	Gln	Gln	Ile	Leu	Arg	Ala	Thr	Glu	Glu	Phe	Lys	Gln	Leu	Glu	Glu
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Ala	Ile	Gln	Leu	Lys	Lys	Ile	Ser	Glu	Ala	Glu	Lys	Asp	Leu	Leu	Phe
				485					490					495	
Lys	Gln	Leu	Ser	Gly	Arg	Ile	Gln	Leu	Leu	Asn	Lys	Leu	Arg	Gln	Glu
			500					505					510		
Ala	Val	Asp	Leu	Glu	Thr	Gln	Met	Glu	Lys	Gln	Arg	Gln	Glu	Ile	Gly
		515					520					525			
Glu	Lys	Gln	Asn	Glu	Ile	Lys	Asp	Leu	Glu	Ile	Val	Thr	Asp	Ser	Leu
	530					535						540			
Asp	Ser	Arg	Asp	Pro	Lys	His	Cys	His	Met	Lys	Ala	Gln	Lys	Arg	Gly
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Lys	Glu	Gln	Gln	Leu	Asp	Ile	Met	Asn	Lys	Gln	Tyr	Lys	Gln	Leu	Glu
				565					570					575	
Ser	Arg	Leu	Asp	Glu	Ile	Leu	Ser	Arg	Ile	Ala	Lys	Glu	Thr	Glu	Glu
			580					585						590	
Ile	Lys	Asp	Leu	Glu	Glu	Gln	Leu	Thr	Glu	Gly	Gln	Ile	Ala	Ala	Asn
	595					600						605			
Glu	Ala	Leu	Lys	Lys	Asp	Leu	Glu	Ser	Val	Ile	Ser	Gly	Leu	Gln	Glu
	610					615						620			
Tyr	Leu	Glu	Thr	Val	Lys	Gly	Gln	Ala	Arg	Gln	Ala	Gln	Asn	Glu	Cys
625					630					635					640
Arg	Lys	Leu	Gln	Asp	Glu	Lys	Glu	Thr	Leu	Leu	Gln	Arg	Leu	Ser	Glu
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Val	Glu	Gln	Glu	Arg	Asp	Gln	Leu	Glu	Ile	Val	Ala	Ile	Asp	Ala	Glu
			660					665					670		
Asn	Met	Arg	Lys	Glu	Leu	Ala	Glu	Leu	Glu	Asn	Ala	Leu	Gln	Glu	Gln
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	690					695						700			
Tyr	Glu	Ala	Glu	Leu	Glu	Ala	Gln	Leu	Lys	Ile	Arg	Asp	Ala	Glu	Ala
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Asn	Gln	Leu	Lys	Glu	Glu	Leu	Glu	Lys	Leu	Arg	Arg	Leu	Ser	Gln	Leu
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Glu	Gln	Ser	Ala	Leu	Gln	Ala	Glu	Leu	Glu	Lys	Glu	Lys	Gln	Ala	Phe
			740					745					750		
Lys	Thr	Ala	Val	Lys	Lys	Ala	Gln	Leu	Ser	Glu	Gly	Lys	Asp	Gln	Glu
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	770					775						780			
Leu	Leu	Lys	Gln	Gln	Leu	Lys	Asp	Phe	Gln	Ser	His	Leu	Asn	His	Val
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Val	Asp	Gly	Leu	Ile	Arg	Pro	Glu	Glu	Val	Ala	Ala	Cys	Val	Asp	Glu
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Leu	Arg	Lys	Lys	Leu	Lys	Ser	Gly	Ala	Gly	Glu	Met	Arg	Ile	His	Thr
			820					825					830		
Pro	Ser	Asp	Val	Leu	Gly	Lys	Ser	Leu	Ala	Asp	Leu	Gln	Lys	Gln	Phe
		835					840						845		
Ser	Glu	Ile	Leu	Ala	Arg	Ser	Gln	Trp	Glu	Arg	Gln	Glu	Ala	Gln	Val
	850					855						860			
Arg	Glu	Arg	Lys	Leu	Gln	Glu	Glu	Met	Ala	Leu	Gln	Gln	Glu	Lys	Leu
865					870					875					880
Ala	Ser	Gly	Gln	Glu	Glu	Phe	Arg	His	Ala	Cys	Glu	Arg	Ala	Leu	Glu
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Ala	Arg	Ile	Ser	Phe	Asp	Lys	Arg	Gln	His	Glu	Ala	Arg	Ile	Gln	Gln
			900						905					910	
Leu	Glu	Asn	Glu	Ile	His	Tyr	Leu	Gln	Glu	Asn	Leu	Lys	Ser	Met	Glu
		915					920						925		

Glu Ile Gln Gly Leu Thr Asp Leu Gln Leu Gln Glu Ala Asp Glu Glu
 930 935 940
 Lys Glu Arg Ile Leu Ala Gln Leu Arg Glu Leu Glu Lys Lys Lys Lys
 945 950 955 960
 Leu Glu Asp Ala Lys Ser Gln Glu Gln Phe Leu Gly Leu Asp Arg Glu
 965 970 975
 Leu Lys Lys Leu Lys Lys Ala Val Ala Ala Ser Asp Lys Leu Ala Thr
 980 985 990
 Ala Glu Leu Thr Ile Ala Lys Asp Gln Leu Lys Ser Leu His Gly Thr
 995 1000 1005
 Val Met Lys Ile Asn Gln Glu Arg Ala Glu Glu Leu Gln Glu Thr
 1010 1015 1020
 Glu Arg Phe Ser Arg Lys Ala Ala Gln Ala Ala Arg Asp Leu Ile
 1025 1030 1035
 Arg Ala Glu Ala Glu Ile Glu Leu Leu Gln Lys Leu Leu Arg Asp
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 Lys Glu Glu Gln Phe Arg Asn Glu Ile Glu Lys Val Asp Val Gly
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 Ser Gly Gly Ala Lys Ser Gln Met Leu Glu Met Glu Lys Leu Asn
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 Glu Thr Met Glu Arg Gln Arg Thr Glu Ile Ala Arg Leu Arg Asn
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 Leu Leu Asp Leu Thr Gly Ala Asp Asn Lys Gly Asn Phe Glu Asn
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 Val Leu Glu Glu Ile Ala Glu Leu Arg Arg Glu Val Ser His Gln
 1115 1120 1125
 Asn Asp Tyr Ile Ser Ser Met Thr Asp Pro Phe Lys Arg Arg Gly
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 Tyr Trp Tyr Phe Met Pro Pro Pro Ser Ser Ser Lys Val Ser Ser
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 His Ser Ser Gln Ala Thr Lys Asp Ser Gly Val Gly Leu Lys Tyr
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 Thr Ala Ser Thr Pro Val Arg Lys Pro His Arg Gly Arg Gln Asp
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 Gly Lys Glu Asn Ser Gly Pro Pro Pro Ala Ser Gly Tyr Trp Val
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 Tyr Ser Pro Ile Arg Ser Gly Leu His Lys Ser Phe Ser Asn Arg
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 Asp Ala Asp Ser Gly Gly Asp Ser Gln Glu Glu Ser Glu Leu Asp
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 Tyr Thr Val Phe Pro Asp Gly Ser Pro Val Pro Gln Gly Met Ala
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 Leu Tyr Ala Pro Pro Pro Pro Leu Pro Asn Asn Ser Gln Pro Leu
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 Pro Val Gly Val Leu His Cys Asn Val Pro Glu His His Asn Leu
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 Glu Asn Glu Val Ser Arg Leu Glu Asp Ile Met Gln His Leu Lys
 1325 1330 1335
 Ser Gly Lys Arg Glu Gln Cys Met Lys Thr Pro Lys Leu Gln Ser
 1340 1345 1350
 Glu Lys Glu Leu Ala Glu Leu Gln His Asn Ile Asp Gly Leu Leu
 1355 1360 1365
 Gln Glu Lys Lys Asp Leu Glu His Glu Val Glu Glu Leu His Arg
 1370 1375 1380

Thr	Ile	Gln	Lys	His	Gln	Gln	Arg	Lys	Asp	Phe	Ile	Asp	Gly	Asn
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	1400					1405					1410			
His	His	Glu	Asp	Ile	Val	Asp	Glu	Ile	Glu	Cys	Ile	Glu	Arg	Thr
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Leu	Leu	Lys	Arg	Arg	Ala	Glu	Leu	Arg	Glu	Ala	Asp	Arg	Leu	Leu
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	1700					1705					1710			
Gln	Gly	Leu	Lys	Leu	Gln	His	Asp	Gln	Lys	Met	Ser	Glu	Leu	Glu
	1715					1720					1725			
Lys	Thr	Arg	Val	Glu	Val	Leu	Glu	Glu	Lys	Leu	Glu	Leu	Glu	Ser
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Glu	Lys	Gln	Asp	Leu	Glu	Glu	Lys	Gln	Asp	Ser	Trp	Glu	Lys	Lys
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Glu	Met	Glu	Arg	Ala	Arg	Leu	Glu	Lys	Leu	Glu	Leu	Asp	Ala	Arg
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Lys	Leu	Gln	Gln	Glu	Leu	Asp	Gln	Arg	Asn	Arg	Glu	Lys	Leu	Ser
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52

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gtc Val 1315	cct Pro	gaa Glu	cac His	cat His	aac Asn 1320	tta Leu	gag Glu	aat Asn	gaa Glu	gtt Val 1325	tct Ser	aga Arg	tta Leu	gaa Glu	4018
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cat His 1360	cat His	aat Asn	att Ile	gat Asp	gat Asp 1365	ctt Leu	ttg Leu	caa Gln	gag Glu	aag Lys 1370	aaa Lys	agc Ser	tta Leu	gag Glu	4153
tgt Cys 1375	gaa Glu	gta Val	gaa Glu	gaa Glu	tta Leu 1380	cat His	aga Arg	act Thr	gtc Val	cag Gln 1385	aaa Lys	cgt Arg	caa Gln	cag Gln	4198
caa Gln 1390	aag Lys	gac Asp	ttc Phe	att Ile	gat Asp 1395	gga Gly	aat Asn	gtt Val	gag Glu	agt Ser 1400	ctt Leu	atg Met	act Thr	gaa Glu	4243
cta Leu 1405	gaa Glu	ata Ile	gaa Glu	aaa Lys	tca Ser 1410	ctc Leu	aaa Lys	cat His	cat His	gaa Glu 1415	gat Asp	att Ile	gta Val	gat Asp	4288

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 2320 2325

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 Gly Gly Gln Trp Cys Glu Gln Val Glu Ile Ala Asp Glu Asn Asn Met
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 Leu Leu Asp Tyr Gln Asp His Lys Gly Ala Asp Ser His Ala Gly Val
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 Arg Tyr Ile Thr Glu Ala Leu Ile Lys Lys Leu Thr Lys Gln Asp Asn
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 Leu Ala Leu Ile Lys Ser Leu Asn Leu Ser Leu Ser Lys Asp Gly Gly
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 Lys Lys Phe Lys Tyr Ile Glu Asn Leu Glu Lys Cys Val Lys Leu Glu
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 Val Leu Asn Leu Ser Tyr Asn Leu Ile Gly Lys Ile Glu Lys Leu Asp
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147

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 Leu Pro Glu Leu Pro Ala Asp Leu Glu Ala Ile Leu Glu Arg Asn
 2180 2185 2190
 Glu Asn Leu Glu Gly Glu Leu Glu Ser Leu Lys Glu Asn Leu Pro
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 Phe Thr Met Asn Glu Gly Pro Phe Glu Glu Lys Leu Asn Phe Ser
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 Glu Lys Leu Arg His Arg Glu Asp Arg Leu Lys Ala Gln Leu Arg
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 His Cys Met Ser Lys Gln Ala Glu Val Leu Ile Lys Gly Lys Arg
 2255 2260 2265
 Gln Thr Glu Gly Thr Leu His Ser Leu Arg Arg Gln Val Asp Ala
 2270 2275 2280
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 2285 2290 2295
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149

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Gly Gln Val Gly Gly Ser Leu Ala Ser Leu Thr Gly Gln Ile Ser Asn
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Phe Thr Lys Asp Met Leu Met Glu Gly Thr Glu Glu Val Glu Ala Glu
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Leu Pro Asn Ser Arg Arg Lys Glu Val Glu Ala Ile His Ala Ile Leu
50 55 60

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Arg Ser Glu Asn Glu Arg Leu Lys Glu Leu Cys Thr Asp Leu Glu Glu
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Lys His Glu Ala Ser Glu Leu Gln Ile Lys Gln Gln Ser Thr Asn Tyr
85 90 95

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Arg Asn Gln Leu Gln Gln Lys Glu Val Glu Ile Ser His Leu Lys Ala
100 105 110

aga cag att gca ctg cag gat cag ttg ctg aag ctg cag tca gct gct 384
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115 120 125

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Gln Ser Ala His Ser Gly Ala Ser Ser Val Pro Ala Ala Leu Ala Ser
130 135 140

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Ser Pro Phe Ser Tyr Ser Val Ser His His Ala Ser Ala Phe His Asp
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Arg Leu Ser Asn Glu Val Ser Arg Leu Glu Ser Glu Val Gly His Trp
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Arg His Ile Ala Gln Thr Ser Lys Ala Gln Gly Ser Asn Ser Ser Asp
195 200 205

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Gln Ser Glu Ile Cys Lys Leu Gln Ser Ile Ile Lys Glu Leu Lys Gln
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Ile Arg Ser Gln Glu Ile Asp Asp His Gln His Glu Met Ser Val Leu
225 230 235 240

150

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gaa Glu	gaa Glu	tta Leu	cgt Arg 260	gac Asp	tat Tyr	gaa Glu	gaa Glu	cga Arg 265	att Ile	gaa Glu	gaa Glu	ctg Leu	gaa Glu 270	aat Asn	ctg Leu	816
tta Leu	gaa Glu	caa Gln 275	ggt Gly	ggc Gly	tca Ser	gga Gly	att Ile 280	gta Val	ata Ile	cct Pro	gat Asp	cac His 285	tca Ser	aaa Lys	atc Ile	864
cat His 290	gag Glu	atg Met	caa Gln	aaa Lys	act Thr	att Ile 295	cag Gln	aat Asn	cta Leu	caa Gln	act Thr 300	gaa Glu	aaa Lys	gta Val	gca Ala	912
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gaa Glu	att Ile	atg Met	aga Arg 405	ctg Leu	agt Ser	aat Asn	tta Leu	tac Tyr	cag Gln 410	gat Asp	aac Asn	agt Ser	ctc Leu	act Thr 415	gaa Glu	1248
gat Asp	aat Asn	ttg Leu	aaa Lys 420	ctt Leu	aaa Lys	atg Met	cat His	gtc Val	gaa Glu 425	ttt Phe	tta Leu	gaa Glu	aaa Lys 430	cag Gln	aag Lys	1296
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151

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gag Glu	tta Leu	gag Glu	gaa Glu 500	ttg Leu	gac Asp	aga Arg	caa Gln	aac Asn 505	caa Gln	gaa Glu	gct Ala	aca Thr	aag Lys 510	cac His	atg Met	1536
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atc Ile	att Ile 530	agt Ser	aaa Lys	ctg Leu	aga Arg	aaa Lys 535	gat Asp	cta Leu	aat Asn	gat Asp	gaa Glu 540	aac Asn	aag Lys	aga Arg	gtc Val	1632
cat His 545	caa Gln	ctt Leu	gaa Glu	gat Asp 550	gat Asp	aaa Lys	aag Lys	aat Asn	atg Met	act Thr 555	aaa Lys	gaa Glu	cta Leu	aat Asn	gtg Val 560	1680
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caa Gln	aat Asn	ctt Leu	tca Ser 660	gaa Glu	ata Ile	gaa Glu	cag Gln	ctc Leu 665	aat Asn	gac Asp	agt Ser	tta Leu	aac Asn 670	aaa Lys	gtt Val	2016
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tct Ser 705	ctg Leu	gaa Glu	aga Arg	aac Asn	act Thr 710	att Ile	gtg Val	gag Glu	gct Ala	cta Leu	aaa Lys 715	atg Met	gaa Glu	aaa Lys	gga Gly 720	2160

152

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ttt att gag aaa ctt aaa gaa aga agt tca gag ctt cag gag gaa tta Phe Ile Glu Lys Leu Lys Glu Arg Ser Ser Glu Leu Gln Glu Glu Leu 820 825 830	2496
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Glu Phe Phe Gln Glu Thr Lys Val Gln Ser Leu Asn Leu Glu Asn Gly	
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Ser Glu Lys His Asp Leu Ser Lys Ala Glu Thr Glu Arg Leu Val	
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163

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164

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165

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171

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<213> Homo sapiens

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20      25      30
Phe Thr Lys Asp Met Leu Met Glu Gly Thr Glu Glu Val Glu Ala Glu
35      40      45
Leu Pro Asp Ser Arg Thr Lys Glu Ile Glu Ala Ile His Ala Ile Leu
50      55      60
Arg Ser Glu Asn Glu Arg Leu Lys Lys Leu Cys Thr Asp Leu Glu Glu
65      70      75      80
Lys His Glu Ala Ser Glu Ile Gln Ile Lys Gln Gln Ser Thr Ser Tyr
85      90      95
Arg Asn Gln Leu Gln Gln Lys Glu Val Glu Ile Ser His Leu Lys Ala
100     105     110
Arg Gln Ile Ala Leu Gln Asp Gln Leu Leu Lys Leu Gln Ser Ala Ala
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Gln Ser Val Pro Ser Gly Ala Gly Val Pro Ala Thr Thr Ala Ser Ser
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Ser Phe Ala Tyr Gly Ile Ser His His Pro Ser Ala Phe His Asp Asp
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Asp Met Asp Phe Gly Asp Ile Ile Ser Ser Gln Gln Glu Ile Asn Arg
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Leu Ser Asn Glu Val Ser Arg Leu Glu Ser Glu Val Gly His Trp Arg
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His Ile Ala Gln Thr Ser Lys Ala Gln Gly Thr Asp Asn Ser Asp Gln
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Ser Glu Ile Cys Lys Leu Gln Asn Ile Ile Lys Glu Leu Lys Gln Asn
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Gln Gln Gly Gly Ser Gly Val Ile Glu Thr Asp Leu Ser Lys Ile Tyr
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Glu Met Gln Lys Thr Ile Gln Val Leu Gln Ile Glu Lys Val Glu Ser
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Thr Lys Lys Met Glu Gln Leu Glu Asp Lys Ile Lys Asp Ile Asn Lys
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Glu Glu Val Phe Arg Leu Gln Gln Ala Leu Ser Asp Ala Glu Asn Glu
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Glu	Thr	Ala	Leu	Gln	Leu	Ser	Val	Ser	Gln	Glu	Gln	Val	Lys	Gln
1655						1660					1665			
Tyr	Ala	Leu	Ser	Leu	Ala	Asn	Leu	Gln	Met	Val	Leu	Glu	His	Phe
1670						1675					1680			
Gln	Gln	Glu	Glu	Lys	Ala	Met	Tyr	Ser	Ala	Glu	Leu	Glu	Lys	Gln
1685						1690					1695			
Lys	Gln	Leu	Ile	Ala	Glu	Trp	Lys	Lys	Asn	Ala	Glu	Asn	Leu	Glu
1700						1705					1710			
Gly	Lys	Val	Ile	Ser	Leu	Gln	Glu	Cys	Leu	Asp	Glu	Ala	Asn	Ala
1715						1720					1725			
Ala	Leu	Asp	Ser	Ala	Ser	Arg	Leu	Thr	Glu	Gln	Leu	Asp	Val	Lys
1730						1735					1740			
Glu	Glu	Gln	Ile	Glu	Glu	Leu	Lys	Arg	Gln	Asn	Glu	Leu	Arg	Gln
1745						1750					1755			
Glu	Met	Leu	Asp	Asp	Val	Gln	Lys	Lys	Leu	Met	Ser	Leu	Ala	Asn
1760						1765					1770			
Ser	Ser	Glu	Gly	Lys	Val	Asp	Lys	Val	Leu	Met	Arg	Asn	Leu	Phe
1775						1780					1785			
Ile	Gly	His	Phe	His	Thr	Pro	Lys	Asn	Gln	Arg	His	Glu	Val	Leu
1790						1795					1800			
Arg	Leu	Met	Gly	Ser	Ile	Leu	Gly	Val	Arg	Arg	Glu	Glu	Met	Glu
1805						1810					1815			
Gln	Leu	Phe	His	Asp	Asp	Gln	Gly	Ser	Val	Thr	Arg	Trp	Met	Thr
1820						1825					1830			

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Gly Trp Leu Gly Gly Gly Ser Lys Ser Val Pro Asn Thr Pro Leu
 1835 1840 1845
 Arg Pro Asn Gln Gln Ser Val Val Asn Ser Ser Phe Ser Glu Leu
 1850 1855 1860
 Phe Val Lys Phe Leu Glu Thr Glu Ser His Pro Ser Ile Pro Pro
 1865 1870 1875
 Pro Lys Leu Ser Val His Asp Met Lys Pro Leu Asp Ser Pro Gly
 1880 1885 1890
 Arg Arg Lys Arg Asp Thr Asn Ala Pro Glu Ser Phe Lys Asp Thr
 1895 1900 1905
 Ala Glu Ser Arg Ser Gly Arg Arg Thr Asp Val Asn Pro Phe Leu
 1910 1915 1920
 Ala Pro Arg Ser Ala Ala Val Pro Leu Ile Asn Pro Ala Gly Leu
 1925 1930 1935
 Gly Pro Gly Gly Pro Gly His Leu Leu Leu Lys Pro Ile Ser Asp
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 1955 1960 1965
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CLAIMS:

1. An immunity-inducing agent to treat and/or prevent a calmegin-expressing cancer(s), said agent comprising as an effective ingredient any one of the polypeptides (a) to (c) below, said polypeptide having an immunity-inducing activity:
 - 5 (a) a polypeptide having the amino acid sequence shown in SEQ ID NO: 16 or 18 in SEQUENCE LISTING;
 - (b) a polypeptide having a sequence identity of not less than 89% to the full length of the polypeptide (a); and
 - (c) a polypeptide comprising the polypeptide (a) or (b) as a partial sequence.
- 10 2. The immunity-inducing agent according to claim 1, wherein said polypeptide (b) has a sequence identity of not less than 95% to the full length of said polypeptide (a).
3. The immunity-inducing agent according to claim 1, wherein said polypeptide having an immunity-inducing activity is a polypeptide having the amino acid sequence shown
15 in SEQ ID NO: 16 or 18 in SEQUENCE LISTING or a polypeptide comprising said polypeptide as a partial sequence.
4. The immunity-inducing agent according to claim 3, wherein said polypeptide having an immunity-inducing activity has the amino acid sequence shown in SEQ ID NO: 16 or 18 in SEQUENCE LISTING.
- 20 5. The immunity-inducing agent according to any one of claims 1 to 4, which is for use in a human, dog or cat.
6. The immunity-inducing agent according to claim 5, further comprising an immunoenhancer.

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7. The immunity-inducing agent according to claim 6, wherein said immunoenhancer is at least one selected from the group consisting of Freund's incomplete adjuvant; Montanide; poly I:C and derivatives thereof; CpG oligonucleotides; interleukin-12; interleukin-18; interferon- α ; interferon- β ; interferon- ω ; interferon- γ ; and Flt3 ligand.
- 5 8. A method for preparing antigen-presenting cells *in vitro*, said method comprising bringing a polypeptide having the amino acid sequence shown in SEQ ID NO: 16 or 18 in SEQUENCE LISTING into contact with dendritic cells or B cells which have MHC class I molecules.
9. Use of any one of the polypeptides (a) to (c) below, said polypeptide having an
10 immunity-inducing activity for production of an immunity-inducing agent to treat and/or prevent a calmegin-expressing cancer(s):
- (a) a polypeptide having the amino acid sequence shown in SEQ ID NO: 16 or 18 in SEQUENCE LISTING;
- (b) a polypeptide having a sequence identity of not less than 89% to the full
15 length of the polypeptide (a); and
- (c) a polypeptide comprising the polypeptide (a) or (b) as a partial sequence.

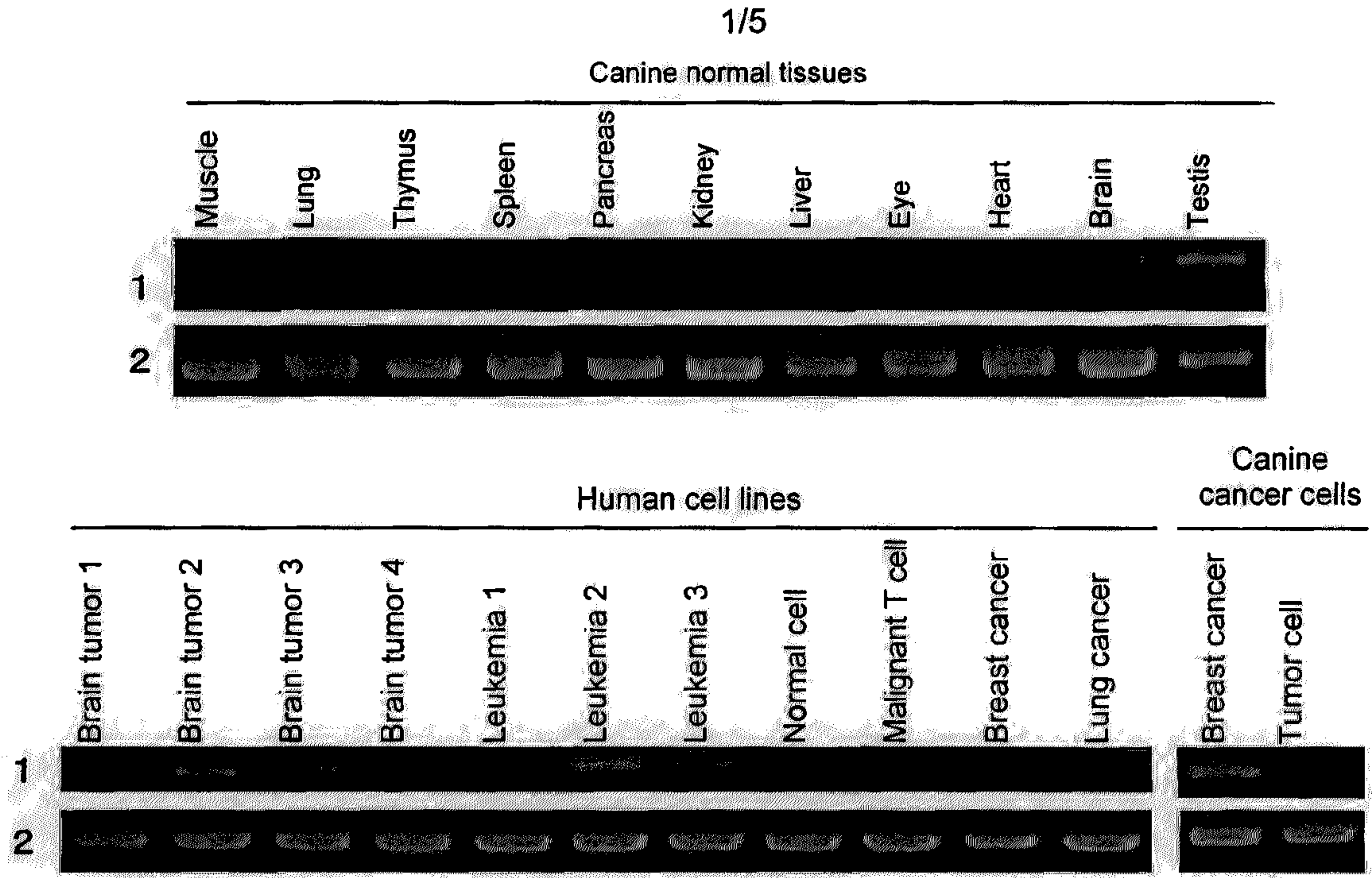


Fig.1

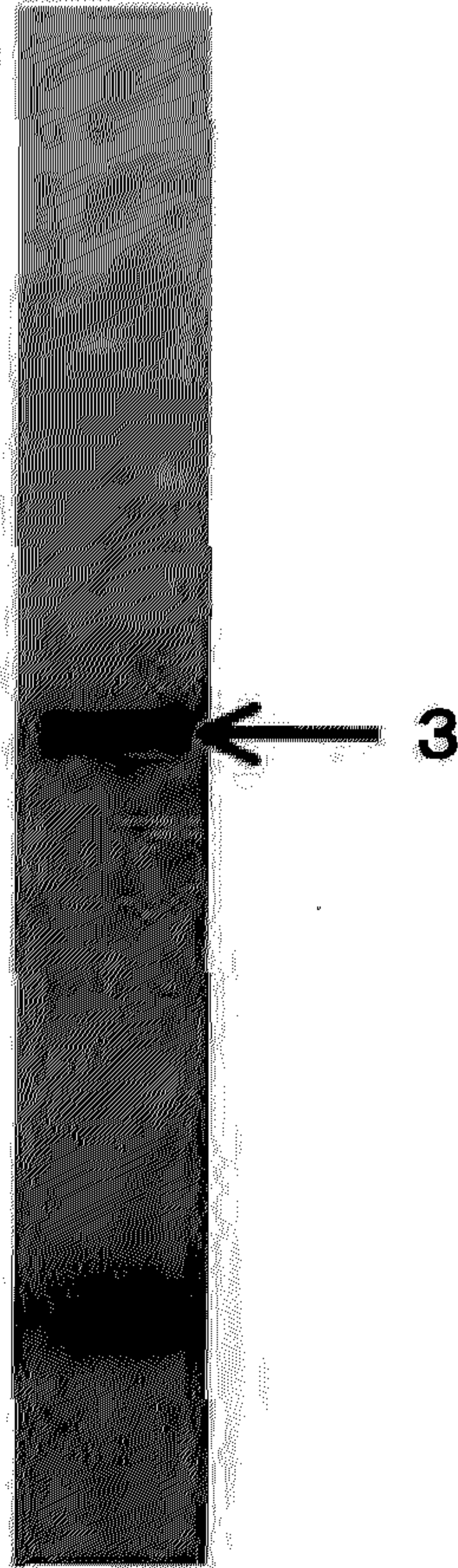


Fig.2

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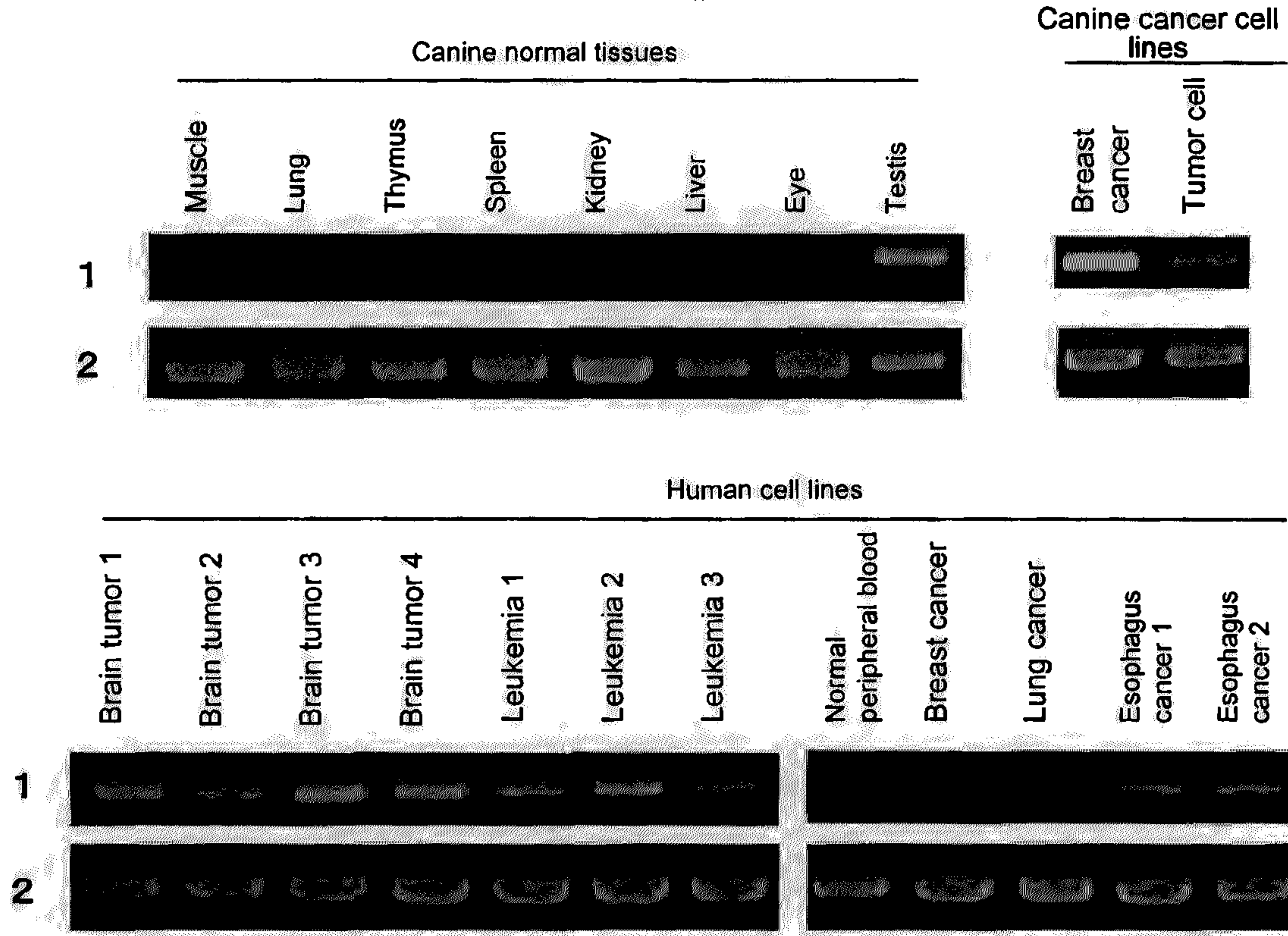


Fig.3

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Fig.4

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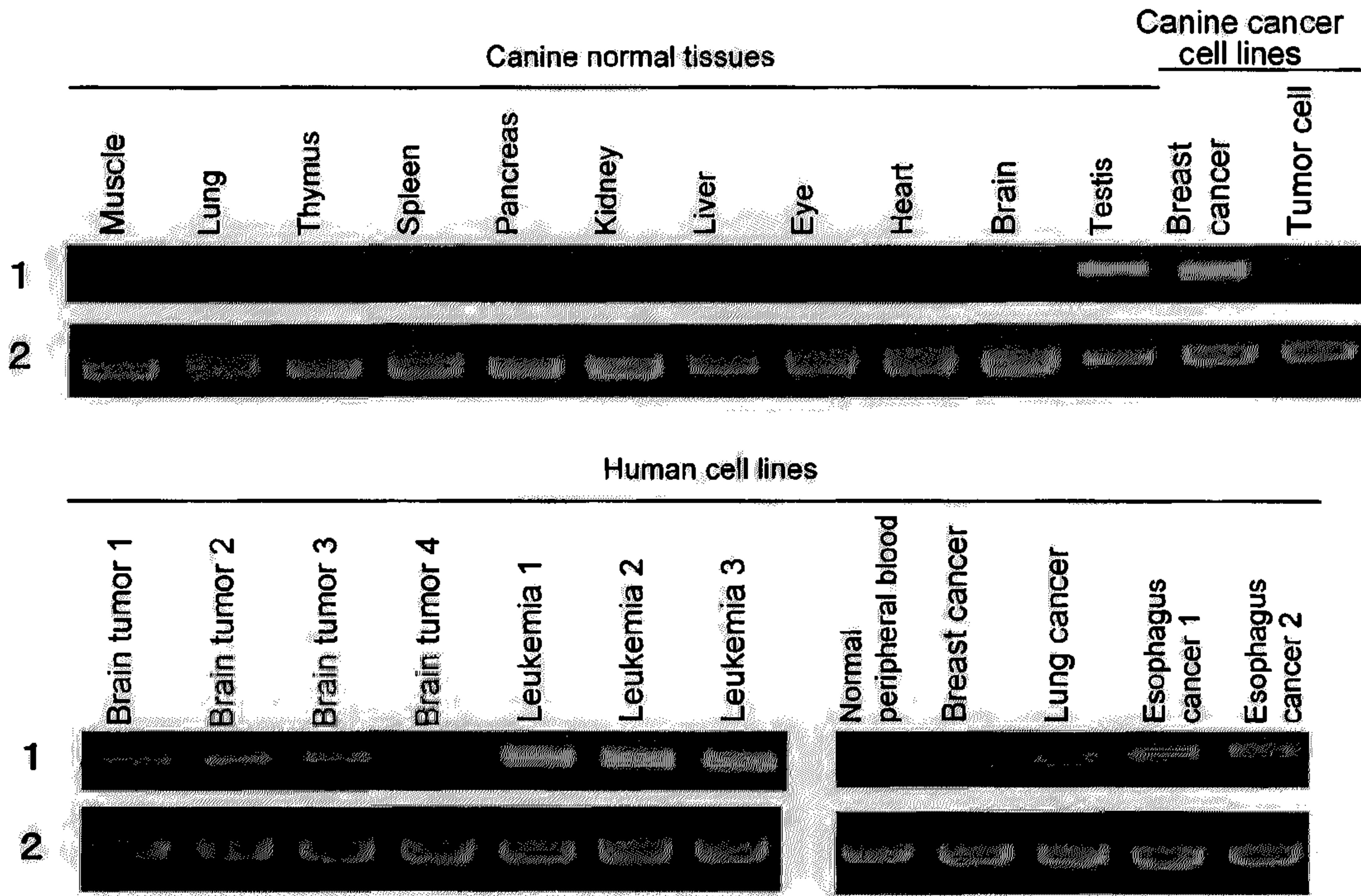


Fig.5

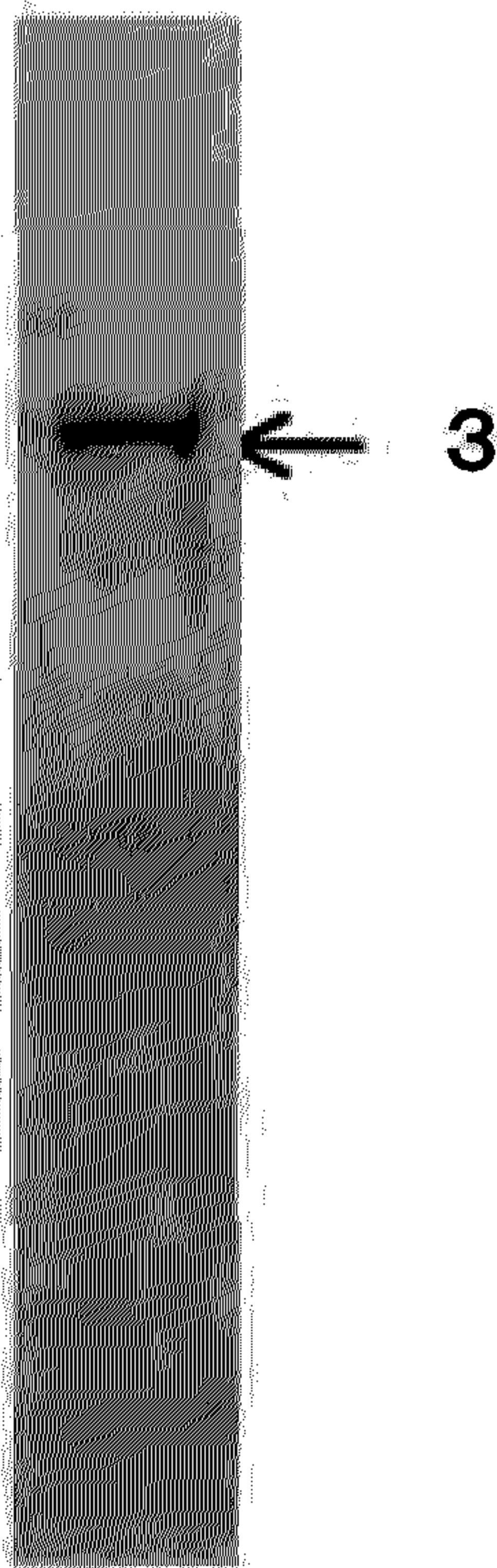


Fig.6

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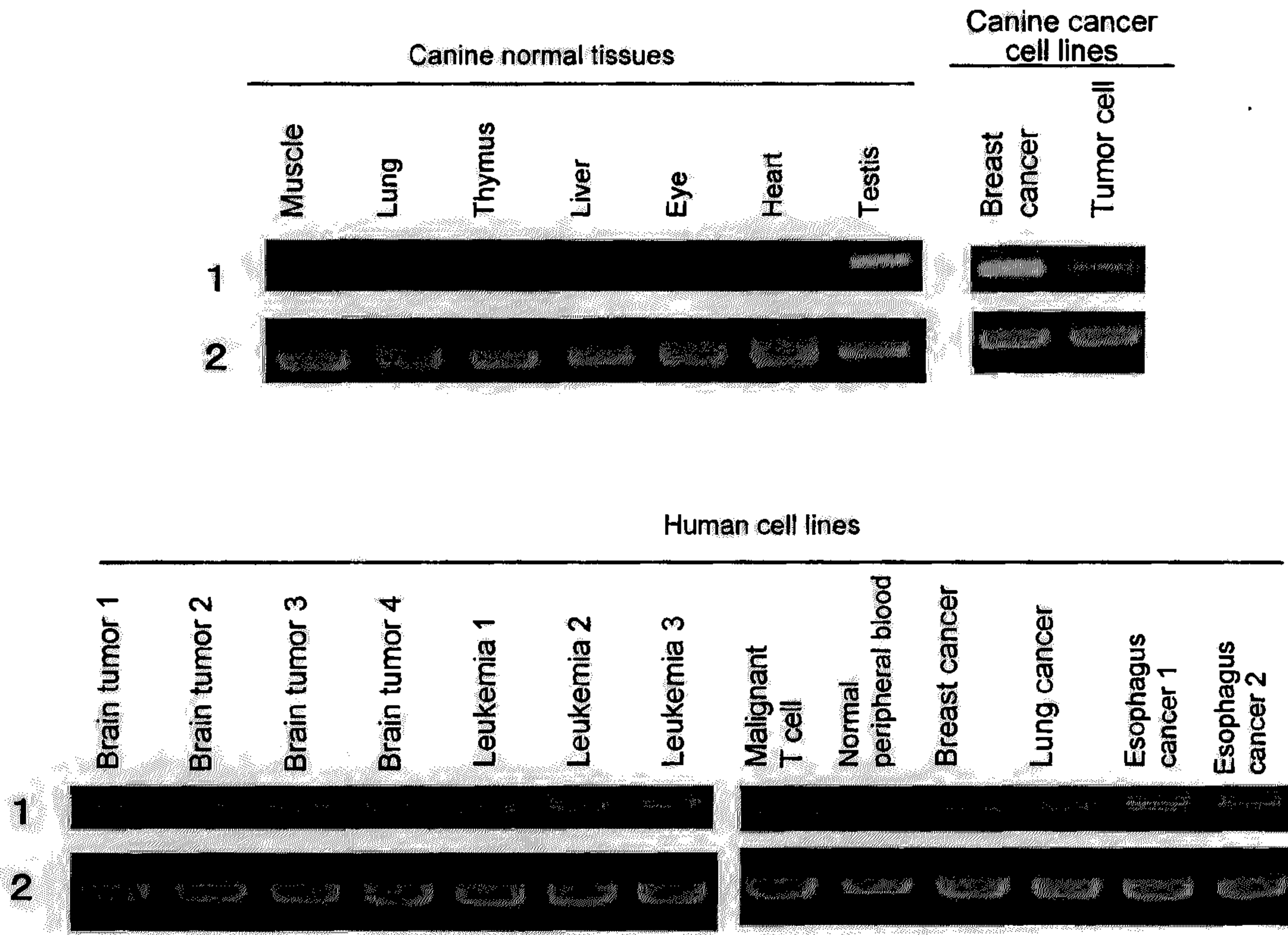


Fig.7

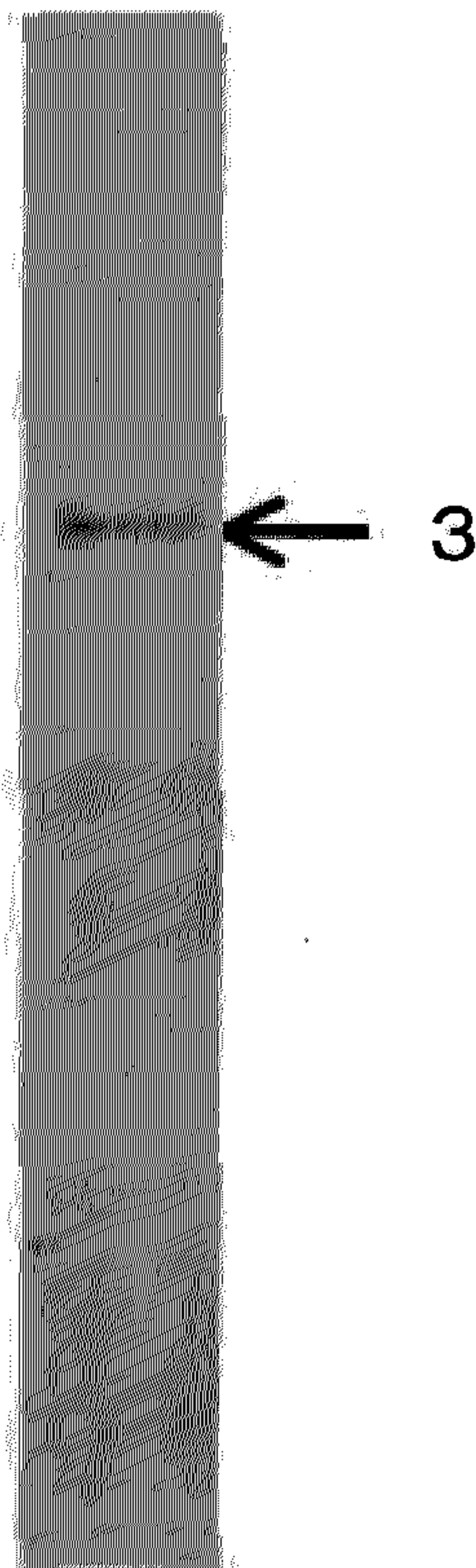


Fig.8