CONJUGATES FOR BONE IMAGING AND BONE CANCER THERAPY.

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Description

Technical Field

The present invention relates generally to conjugates formed from calcified tissue targeting agents and radiolabeled compounds such as chelates, and methods of using these compositions. This invention is more particularly related to conjugates which are useful for bone matrix imaging and bone cancer therapy.

Background of the Invention

The invasive growth of cancer cells into bone tissues often results in severe pain syndromes and may ultimately result in death. Ionizing radiation has been used to achieve a reduction in pain. In particular, external radiation therapy has been applied in cases where bone pain is restricted to focal areas. For disseminated bone metastases, however, a treatment which focuses the radiation dose to bone tissue and particularly to bone metastases is desirable.

Prior to therapy, it is necessary to obtain reliable diagnostic information and to this end several approaches have been tried. An approach that has been the subject of a number of patents, for example, U.S. Patent No. 3,974,268, to Subramanian et al., involves the use of a technetium-99m (99mTc) diphosphonate complex as a skeletal-imaging agent. This approach utilizes a diphosphonate as both the bone-seeking agent and the complexing agent for the radionuclide. Such an approach has at least three major disadvantages. First, by involving the bone-seeking agent in the complexation of the metal radionuclide, the ability of the agent to direct, or "target," the radionuclide to the desired site may be diminished. Second, because diphosphonates are known to form polymers, a preparation of diphosphonates and radionuclides typically includes multiple forms of diphosphonate-radionuclide complexes which have differing charge and uptake properties. Third, the complex formed between a diphosphonate and a radionuclide is not of optimal stability which may necessitate purification of the diphosphonate complex prior to use. Further, even a purified diphosphonate complex may lose the radionuclide during its use.

Thus, there is a need in the art for compositions which are capable of both stably binding a radionuclide and preferentially delivering it to calcified tissue. The present invention fulfills this need and further provides other related advantages.

Summary of the Invention

Briefly stated, the present invention, in one aspect, provides conjugates formed from calcified tissue targeting agents and radiolabeled compounds. According to the present invention there is provided a conjugate comprising a radiolabeled chelate and a targeting agent that associates with calcified tissue, wherein said chelate contains at least four donor atoms selected from nitrogen atoms or sulphur atoms or combinations thereof, and wherein said chelate does not function as said targeting agent. The chelate may be formed from chelating compounds selected from the group consisting of N₂S₂, N₃S, N₂S₃, N₂S₄ and N₂S₅ as defined below.

There are disclosed kits for producing for radiopharmaceutical use the conjugates described above.

Another aspect of the invention provides the conjugates described above for use for diagnostic and therapeutic purposes.

Other aspects of the invention will become evident upon reference to the following detailed description and attached drawings.

Brief Description of the Drawings

- Figure 1 shows a comparison of ⁹⁹mTc-MDP versus ⁹⁹mTc-N₂S₂-ADP.
- Figure 2 shows a comparison of ⁹⁹mTc-ADP, ⁹⁹mTc-N₂S₂-ADP and ⁹⁹mTc-N₂S-ADP.

Detailed Description of the Invention

Prior to setting forth the invention, it may be helpful to an understanding thereof to set forth definitions of certain terms to be used hereinafter.

Calcified tissue - as used herein, includes bone as well as other tissue which may be undergoing pathological calcification.

Targeting agent - is any molecule that has the capacity to associate with calcified tissue.
As noted above, the present invention provides conjugates formed from calcified tissue targeting agents and radiolabeled compounds ("radiolabeled compound-calcified tissue targeting agent conjugates"). Thus, unlike a $^{99m}$Tc-diphosphonate complex for example, the targeting agent in the conjugates is not solely responsible for binding the radionuclide. One advantage of the conjugates is that the radionuclide is afforded greater stability and thus more likely to reach and remain on the target site. In the embodiment in which the radiolabeled compound is a chelating compound, the increased stability and ease of preparation of the conjugate is due, in part, to the superior ability of the chelating compound, relative to a diphosphonate for example, to chelate a metal radionuclide.

Another advantage of the conjugates of the present invention is that the number of different complexes that may be formed from a single conjugate are limited as opposed to the multiple complexes which are possible when a diphosphonate performs the complexation. Thus, conjugates for diagnosis and therapy may be made in a reproducible manner. Yet another advantage of the conjugates is that the targeting agent by not having to complex the metal radionuclide is unencumbered and therefore more capable of performing its targeting role.

The conjugates of the present invention have the formula:

\[ R-X-T \]

wherein \( R \) is a radiolabeled compound, \( X \) is a chemical bond or a linking group, and \( T \) is a targeting agent.

The symbols \( R \), \( X \) and \( T \) are described in more detail as follows.

\( T \) of the formula for the conjugates is a targeting agent. As noted above, a targeting agent has the capacity to associate with calcified tissue. Examples of calcified tissue include bone and other tissues which may be undergoing pathological calcification. The association of a targeting agent with calcified tissue may take a variety of forms, including sequestration by the tissue. Targeting agents will be taken up to a greater extent by those areas of the body which are undergoing more active calcification. Localized areas of abnormal accumulation of the conjugates of the present invention may be seen in primary malignancies of the bone, metastatic malignancies of the bone, acute or chronic osteomyelitis, arthritides, recent fractures, areas of ectopic calcification, Paget's disease, regional migratory osteoporosis, areas of aseptic necrosis, and, in general, any pathological situation involving bone in which there is increased osteogenic activity or localized increased osseous blood perfusion.

Examples of targeting agents include phosphorus-containing compounds such as those which contain C-O-P bonds (phosphates such as those described in U.S. Patent No. 4,582,700), P-O-P bonds (pyrophosphates or polyphosphates such as those described in U.S. Patent No. 4,016,249), C-C-P bonds (phosphonates such as those described in U.S. Patent Nos. 4,233,284 and 4,642,229), P-C-P bonds (diphosphonates such as those described in U.S. Patent No. 4,233,284), P-N-P bonds (imidodiphosphonates such as those described in U.S. Patent No. 3,974,268), or combinations or derivatives thereof. The targeting agents may be used in a variety of forms, e.g., mono-, di- and polyphosphonates. Of the phosphonates, particularly preferred are methylene diphosphonate (MDP) and 4,5-diamino-1-hydroxypentane-1,1-diphosphonate, and 3-amino-1-hydroxypropane-1,1-diphosphonate (ADP).

\( X \) of the formula for the conjugates is a chemical bond or a linking group. Illustrative examples of linking groups include: -(CH$_2$)$_n$-, -O-, -(CHOH)$_n$-, -S-, -NH-, -N substituted-, -C(O)-O-, -C(O)-NH-, -C(O)-S-, and combinations thereof.

\( R \) of the formula for the conjugates may be (a) a chelate comprising a chelating compound and a metal, or metal oxide, radionuclide, or (b) a radiolabeled compound in which the radionuclide is bonded covalently. One conjugate comprises a chelate and a targeting agent that is capable of associating with calcified tissue. The chelate may be formed from chelating compounds that include those containing various combinations of nitrogen and sulfur atoms as the donor atoms for binding a metal or metal oxide. Typically, the chelating compound comprises at least four donor atoms selected from sulfur donor atoms, nitrogen donor atoms, or combinations thereof. European Patent Application Publication No. 0188256, published July 23, 1986, discloses representative chelating compounds and their synthesis. One example of a chelating compound containing two nitrogens and two sulfurs is referred to herein as "N$_2$S$_2". Other chelating compounds included within the invention have different numbers of nitrogen and sulfur atoms. Examples of these chelating compounds are identified in like manner herein as "N$_n$S$_m". Other chelating compounds described in more detail below. In addition, the following U.S. patent applications are hereby incorporated in their entirety by reference: U.S.S.N. 065,017 (filed June 19, 1987), "Metal Radionuclide Labeled Proteins for Diagnosis and Therapy"; U.S.S.N. 172,004 (filed March 23, 1988), "Metal-Radionuclide-Labeled Proteins and Glycoproteins for Diagnosis and Therapy"; U.S.S.N. 201,134 (filed May 31, 1988), "Metal Radionuclide Chelating Compounds for Improved Chelation Kinetics";
and U.S.S.N. 157,284 (filed February 17, 1988), "Anchimeric Radiometal Chelating Compounds."

The N2S2 metal chelating compounds may be dithio, diamino, diaminocarboxylic acids, amino/thio/amido combinations or derivatives thereof, e.g., an N,N'-bis-mercaptoacetyl, (w-x)-diamino carboxylic acid, (x is 1 or 2), esters capable of forming an amide bond in an aqueous medium, and intermediates to the chelate. An example of these compounds has the following formula:

\[
\begin{align*}
\text{wherein:} \\
\text{one of } Z_1, Z_2, Z_3 \text{ or } Z_4 \text{ is } RCW-(HNV)_nY, \text{ and the others are } =O \text{ or } H_2; \\
R \text{ is a divalent organic radical of at least 1 carbon atom and typically not more than 10, usually not more than 6 carbon atoms, usually from 1 to 3 carbon atoms having from 0 to 2 heteroatoms which are chalcogen (O, S) or nitrogen and is aliphatic, alicyclic, aromatic or heterocyclic, preferably aliphatic having from 0 to 2, usually 0 to 1 site of aliphatic unsaturation (e.g., ethylenic) and of from 1 to 2 carbon atoms;}
W \text{ is oxygen or imino (} =O \text{ or } =NH), \text{ with the proviso that when } Y \text{ is } -NH_2 \text{ or } NHNH_2, \text{ the } W \text{ bonded to the carbon atom bonded to } Y \text{ is } H_2; \\
V \text{ is } RCW, \text{ where the two } RCWs \text{ may be the same or different, usually being of from 1 to 8, more usually being of from 1 to 6 carbon atoms, preferably of from 2 to 3 carbon atoms;}
\end{align*}
\]

T is an acyl or acylthio radical of from 2 to 10, usually 2-8 carbon atoms, either a hydrocarbyl acyl or substituted acrylradical, usually aryl (e.g., phenyl) or alkyl (e.g., methyl), an organic sulfhydryl radical of from 1 to 10 carbon atoms, either substituted or unsubstituted hydrocarbyl; a heterocycle, particularly a chalcogen (O, S) heterocycle; an acylamidomethylene, where the acyl group is as defined above; hydrogen; sulfonato; an alkali metal ion; or the two T's may be taken together to define a polyvalent metal radionuclide, as the metal ion or metal ion oxide;

Substituents include nitro, cyano, inert halo (aryl or polyhalo), non-oxo-carbonyl (carboxylic acid, amide and ester), and the like;

Y is a chemically reactive moiety and is defined below;
A's are the same or different and are hydrogen, carboxylate or lower alkyl of from 1 to 6 carbon atoms, usually of from 1 to 3 carbon atoms, particularly methyl, usually hydrogen; and
X is a bond, methylene or CHZ+;
where T is other than M or H, Y will be other than a targeting agent.

Preferred sulfur protecting groups are those that result in the formation of a thioacetal, hemithioacetal, thioester or acetamidomethyl substituent. Particularly preferred groups include p-anisylidine, acetonyl, tetrahydrylfuranyl, ethoxyethyl, tetrahydrylpyranyl, acetamidomethyl and derivatives thereof. When conjugated to a targeting agent, the protecting groups may be removed and left as sulfydryls either during storage or just prior to radiolabeling.
A preferred group of $N_2S_2$ compounds will have one of the following formulas:

$$(\Lambda)_2S\overset{\text{M}}{\text{N}}(\Lambda)_2$$

wherein all of the symbols have been defined previously except for $M$ and $T'$, and wherein:

$M$ is a radionuclide capable of being chelated as the metal ion or metal ion oxide; and

$T'$ is a sulfur protective group, which includes acyl, acylthio, hydrocarbylthio or substituted-hydrocarbylthio or heterocyclicthio, where the acyl and hydrocarbly groups may be aliphatic, alicyclic, aromatic or combinations thereof and the acyl group further includes heterocyclic, wherein acyl is normally carboxyacyl; $T'$ will generally be of from 2 to 10 carbon atoms, usually 2 to 8 carbon atoms when acyl, where substituents will include non-oxo-carbonyl (carboxy), halo (aryl), particularly fluoro and chloro, cyano and nitro. Preferred sulfur-protecting groups are those that result in the formation of a thioacetal, hemithioacetal, thioester or acetamidomethyl substituent. Particularly preferred groups include $p$-anisylidine, acetonyl, tetrahydrylfuranyl, ethoxyethyl, tetrahydrylpyranyl, acetamidomethyl and derivatives thereof. When conjugated to a targeting agent, the protecting groups may be removed and left as sulphydryls either during storage or just prior to radiolabeling.

$N_2S_2$ type chelate compounds will for the most part have the following formula:

$$(A')_3S\overset{\text{M}}{\text{N}}(A')_2$$

wherein:

one of $Z^1$, $Z^2$, $Z^3$ or $Z^4$ is $R'CW'(HNV')n'Y'$, and the others are $=O$ or $H_2$;

$(A')$'s are the same or different and are hydrogen, carboxylate or lower alkyl or from 1 to 6, usually 1 to 3 carbon atoms, particularly methyl, usually hydrogen;

$n'$ is 0 or 1;

$V'$ is $R'CW'$, where the $(R'CW')$'s may be the same or different, usually being of from 1 to 8, more usually of from 1 to 6 carbon atoms, preferably of from 2 to 3 carbon atoms;

$W'$ is oxygen or imino ($=N$ or $=0$), with the proviso that when $Y'$ is $=NH_2$ or $NNH_2$, the $W'$ bonded to the carbon atom bonded to $Y$ is $H_2$;

$M$ is a radionuclide capable of being chelated as the metal ion or metal ion oxide;

$X'$ is a bond, methylene or CHZ$^{2+}$;
R' is an aliphatic divalent radical of from 1 to 6, usually from 1 to 3 carbon atoms, having from 0 to 1 site of aliphatic unsaturation and 0 to 2 heteroatoms, usually straight chain and preferably methylene or polymethylene of from 2 to 3 carbon atoms; and

Y' is a chemically reactive moiety as defined below.

The dashed lines in the formulae presented for the chelate compounds of the invention represent four coordinate covalent bonds between the metal radionuclide M and each of the two sulfur and the two nitrogen atoms shown in the formulae. Thus, the metal radionuclide is bound through relatively stable bonds in the chelate compounds of the invention.

An example of a N₂S₂ conjugate has the formula:

![Formula Image]

The N₂S metal chelating compounds will have, for the most part, the following formula:

![Formula Image]

wherein:

- T is H or a sulfur protecting group;
- each X independently represents H₂ or O;
- each R independently represents a substituent selected from the group consisting of hydrogen; alkyl; carboxylate; sulfonate; geminal dialkyl; a non-alkyl side chain of an amino acid other than cysteine (alkyl side chains being covered when R is an alkyl group); and -(CH₂)ₙ-Z;
- Z represents a chemically reactive moiety;
- n is an integer of from 1 to about 4; and
- R' is H₂; -(CH₂)ₙ-Z; or an alkyl group having one or more polar groups substituted thereon;

wherein the compound comprises at least one -(CH₂)ₙ-Z substituent.

When Z is -NH₂, n should be at least 2. When Z is other than -COOH, n preferably is 3.

The sulfur protecting group may be selected from alkyl, aryl, acyl (preferably alkanoyl or benzoyl), thioacyl groups having 1 to about 7 carbons, and organothio groups having 1 to about 10 carbons. Preferred sulfur protecting groups are those that result in the formation of a thioacetal, hemithioacetal, thioester or acetamidomethyl substituent. Particularly preferred groups include p-anisylidine, acetonyl, tetrahydrylfuranyl, ethoxyethyl, tetrahydrylpyranyl, acetamidomethyl and derivatives thereof. When conjugated to a targeting agent, the protecting groups may be removed and left as sulfydryls either during storage or just prior to radiolabeling.

For the R groups, the alkyl groups generally contain from 1 to 7 carbons, preferably from 1 to 4 carbons, and most preferably represent methyl.
An example of a N$_3$S conjugate has the formula:

\[
\begin{array}{c}
\text{N} \quad \text{Tc}^- \\
\text{S} \quad \text{N} \\
\text{O} \quad \text{O} \\
\text{N} \quad \text{H} \quad \text{CH}_2 \quad \text{R} \\
\text{S} \quad \text{PO}_3^- \\
\end{array}
\]

The N$_2$S$_3$ and N$_2$S$_4$ chelating compounds have the following formula:

\[
\begin{array}{c}
X_1 \quad Q \quad Z \\
\alpha \quad \text{(CH}_2\text{n)} \quad \beta \\
X_2 \quad A \quad X_3 \\
\text{A} \quad X_4 \\
Y \quad Y' \\
R_5 \quad R_6
\end{array}
\]

Examples of specific embodiments of the elements include the following.

X$_1$ and X$_2$ may be H or an oxy group (=O), but both are not =O. Likewise, X$_3$ and X$_4$ may be H or =O, but both are not =O. By selecting =O for X$_1$ or X$_2$, the N interposed between the carbons to which X$_1$ and X$_2$ are bonded will be an amide. Likewise, by selecting =O for X$_3$ or X$_4$, the N interposed between the carbons to which X$_3$ and X$_4$ are bonded will be an amide. Thus, a compound with zero, one or two amides may be formed by the appropriate selection of X$_1$, X$_2$, X$_3$ and X$_4$. Amide nitrogens, relative to amine nitrogens, afford greater stability to the complex formed with a metal, but at the expense of a diminished acceleration of complex formation. Thus, by selection of X$_1$, X$_2$, X$_3$ and X$_4$, compounds with a wide variety of chelating properties may be formed.

A is a hydrogen (H), alkyl group of C$_6$ or less, -CH$_2$-CH$_2$-S-R$_1$ or -CO-CH$_2$-S-R$_1$, except when either X$_1$ or X$_2$ is =O, A is H. Similarly, A' is H, alkyl group of C$_6$ or less, -CH$_2$-CH$_2$-S-R$_2$ or -CO-CH$_2$-S-R$_2$, except when either X$_3$ or X$_4$ is =O, A' is H. Y is -CH$_2$-S-R$_3$, or H, when A is H or an alkyl group of C$_6$ or less and A' is H or an alkyl group of C$_6$ or less. Alternatively, when either A or A' or both are not H or an alkyl group of C$_6$ or less, then Y is H. Similarly, Y' is -CH$_2$-S-R$_4$, or H, when A is H or an alkyl group of C$_6$ or less and A' is H or an alkyl group of C$_6$ or less. However, Y and Y' are both not H when A is H or an alkyl group of
C₆ or less and A' is H or an alkyl group of C₆ or less. Alternatively, when either A or A' or both are not H or an alkyl group of C₆ or less, then Y is H. Similarly, Y' is -CH₂-S-FU, or H, when A is H or an alkyl group of C₆ or less and A' is H or an alkyl group of C₆ or less. However, Y and Y' are both not H when A is H or an alkyl group of C₆ or less and A' is H or an alkyl group of C₆ or less. Alternatively, when either A or A' or both are not H or an alkyl group of C₆ or less, then Y' is H. Thus, compounds of the formula depicted above may be formed containing two nitrogens and three or four sulfurs ("N₂S₃" and "N₂S₄", respectively). For "N₂S₄" compounds, two of the sulfurs are the sulfurs bearing R₅ and R₆ and the remaining two sulfurs are from A and A' or Y and Y'. The following formulae depict examples of N₂S₄ compounds in which two sulfurs are from Y and Y' (A) or in which two sulfurs are from A and A' (B).

R₁, R₂, R₃, R₆, R₅, and R₆ are independently selected from sulfur protecting groups. Groups which may be used include any of the alkyl, acyl and aryl groups, disulfides and bunte salts known by those skilled in the art. Preferred sulfur protecting groups are those that result in the formation of a thioacetal, hemithioacetal, thioester or acetamidomethyl substituent. Particularly preferred groups include p-anisylidine, acetonil, tetrahydrylfuranyl, ethoxyethyl, tetrahydrylpyranyl, acetamidomethyl and derivatives thereof. When conjugated to a targeting agent, the protecting groups may be removed and left as sulfdryls either during storage or just prior to radiolabeling.

Q may be H or a polar group. One function of a polar group is to increase the hydrophilicity of the compound, e.g., in order to increase its aqueous solubility. Groups which may be used include carboxylates, sulfonates and secondary alcohols. A preferred group is -CH₂-COOH. Q may be attached to one of the positions designated as α, β, and γ. Because the number of methylene carbons at the γ position is defined by n which may be greater than one, the γ position includes additional points for attachment of Q.

The distance by which the nitrogen atoms are separated may be increased by interposing methylene (-CH₂-) groups between the carbons bonded to the nitrogens. When the number of -CH₂- groups, represented by n, is greater than zero, then the number of carbon atoms separating the nitrogen atoms in compound I is increased accordingly. Preferred integers for n are 0 to 2.

Z is -(W)m-R'. W is a group that functions as a "spacer arm" and may be useful to distance R' from the chelating portion of the compound. Groups which may be used include methylene (-CH₂-), methylenecarbonyl (-CH₂-CO-), or combinations thereof. The number, m, of groups such as these would be typically 0 to 30 and preferably 0 to 5.

Z, or R' when m is 0, may be attached to one of the positions designated as α, β, and γ. Because the number of methylene carbons at the γ position is defined by n which may be greater than one, the γ position includes additional points for attachment of a Z or an R'.

R' is a chemically reactive moiety. The moiety may be strongly electrophilic or nucleophilic and thereby be available for reacting directly with a targeting agent. Alternatively, the moiety may be a weaker electrophile or nucleophile and therefore require activation prior to the conjugation with a targeting agent. This alternative would be desirable where it is necessary to delay activation of R' until a compound has been formed. In either scenario R' is chemically reactive, the scenarios differ by whether following formation of a compound, R' is reacted directly with a targeting agent or is reacted first with one or more chemicals to
render $R'$ capable of reacting with a targeting agent. A discussion of reactions illustrative of a direct reaction with $R'$ and of activation of $R'$ is found below.

$N_3S_3$ compounds which contain three nitrogens and three sulfurs, have the following formula:

\[
\begin{align*}
\text{N}_1 & \text{N}_2 \text{N}_3 \\
\text{S}_1 & \text{S}_2 \text{S}_3
\end{align*}
\]

Examples of specific embodiments of the elements include the following.

$R_1$, $R_2$, and $R_3$ are independently selected from sulfur protecting groups. Groups which may be used include any of the alkyl, acyl, aryl groups, disulfides and bunte salts known by those skilled in the art. Preferred groups are those that result in on acyl, a thioacetol or a hemithioacetal. Particularly preferred groups include thioesters, p-anisylidine, acetonyl, tetrahydryfuranyl, ethoxyethyl, tetrahydropyranyl, acetamidomethyl, and derivatives thereof.

$X_1$ and $X_2$ are independently selected from hydrogen (H) and an alkyl group of C6 or less. $X_3$ is an H, an alkyl group of C6 or less, or Z. $X_4$, $X_5$, and $X_6$ are independently selected from H and $=0$. The selection of $=0$ results in the presence of an amide. Thus, a compound with zero, one, two or three amides may be formed by the appropriate selection of $X_4$, $X_5$, and $X_6$. Amide nitrogens, relative to amine nitrogens, afford greater stability to the complex formed with a metal, but at the expense of a diminished acceleration of complex formation. Thus, by selection of $X_4$, $X_5$, and $X_6$, compounds with a wide variety of chelating properties may be formed.

$Q$ may be H or a polar group. One function of a polar group is to increase the hydrophilicity of the compound, e.g., in order to increase its aqueous solubility. Groups which may be used include carboxylates, sulfonates and secondary alcohols. A preferred group is $-\text{CH}_2-\text{COOH}$. $Q$ may be attached to one of the positions designated as $\alpha$, $\beta$, $\gamma$ and $\delta$. Because the number of methylene carbons at the $\delta$ position is defined by $n$ which may be greater than one, the $\delta$ position includes additional points for attachment of $Q$.

The distance by which the nitrogen atoms are separated may be increased by interposing methylene ($-\text{CH}_2-$) groups between the carbons bonded to the nitrogens. When the number of $-\text{CH}_2-$ groups, represented by $n$, is greater than zero, then the number of carbon atoms separating the nitrogen atoms in compound II is increased accordingly. Preferred integers for $n$ are 0 to 4.

$Z$ is $-(W)_m-R'$. $W$ is a group that functions as a "spacer arm" and may be useful to distance $R'$ from the chelating portion of the compound. Groups which may be used include methylene ($-\text{CH}_2-$), methylenedioxy ($-\text{CH}_2-\text{O}-$), methylenecarbonyl ($-\text{CH}_2-\text{CO}-$), or combinations thereof. The number, $m$, of groups such as these would be typically 0 to 30 and preferably 0 to 5.

$Z$, or $R'$ when $m$ is 0, may be attached to $X_4$ or to one of the positions designated as $\alpha$, $\beta$, $\gamma$, and $\delta$. Because the number of methylene carbons at the $\delta$ position is defined by $n$ which may be greater than one, the $\delta$ position includes additional points for attachment of a Z or an R'.

$R'$ is a chemically reactive group and the discussion of it is found below.

The $N_3S_3$ compound formula, the carbon designated as $\beta$ may be bonded to any one of the carbons designated as $\alpha$, $\gamma$ and $\delta$. The following formulae depict compounds in which the $\beta$ carbon is bonded to the $\gamma$ carbon (A) and the $\beta$ carbon is bonded to the $\delta$ carbon (B).
Preferred metals and metal oxides for any of the chelating compounds described above are those that are radionuclides. Particularly preferred are $^{54}$Cu, $^{67}$Cu, $^{97}$Ru, $^{99m}$Tc, $^{105}$Rh, $^{109}$Pd, $^{186}$Re, $^{188}$Re, $^{198}$Au, $^{199}$Au, $^{203}$Pb, $^{212}$Pb, and $^{212}$Bi.

Three methods are provided for producing the conjugates described herein. The two methods for producing a conjugate from a fully formed compound suitable for radiolabelling have a different order for the addition of the targeting agent. In the first method the targeting agent is joined to a radiolabeled compound, e.g., after a metal or metal oxide has been added to a chelating compound. Conversely, in the second method the targeting agent is joined to a fully formed but unlabeled compound, e.g., before a metal or metal oxide has been added. In both methods, however, a conjugate has a targeting agent joined via a "chemically reactive moiety" (also herein called a "functional group suitable for conjugation"). As noted above, a targeting agent may be joined by a direct reaction with a chemically reactive moiety or following the activation of a chemically reactive moiety.

The step of combining a targeting agent with a labeled or unlabeled compound may be performed by direct reaction of the targeting agent with a chemically reactive moiety. Alternatively, it may be desirable to include a preparatory step before the step of combining. For example, a targeting agent may be modified in preparation for a direct reaction with a chemically reactive moiety. The modification of a targeting agent includes reaction with any of the numerous bifunctional reagents reported in the literature.

A direct reaction with a chemically reactive moiety by modified or unmodified targeting agent is intended to mean that a chemically reactive moiety is capable of reacting with the modified or unmodified targeting agent. For example, a chemically reactive moiety may be an alkyl group containing a good leaving group, e.g., a halide, or a carbonyl-containing group, such as an anhydride, acid halide, or "active ester." The term "active ester" is known to refer to esters which are highly reactive in nucleophilic substitution reactions. In the present invention, the modified or unmodified targeting agent would be the nucleophile. Typically the esters will be activated phenols and cyclic compounds based on hydroxylamine. Examples of commonly used ester groups are tetrafluorophenyl, N-hydroxysuccinimimidyl, nitrophenyl, isothiocyanate and substituted isothiocyanates. Alternatively, a chemically reactive moiety may be a nucleophilic group, such as an amino or sulphydryl group, which is capable of reacting with a modified targeting agent, e.g., containing a maleimide group.

Another way to perform a step in preparation for the step of combining a targeting agent and a labeled or unlabeled compound is to convert a chemically reactive moiety to a form capable of reacting with the targeting agent. Examples of conversions of a chemically reactive moiety include where a chemically reactive moiety is a carboxyl group and is then activated. Activation of a carboxyl group includes formation of an "active ester" as defined above. Another example of a conversion is where a chemically reactive moiety is a succinimide derivative containing a protective group, such as phenylsulfonyl. Upon removal of the group, the succinimide is converted to a maleimide which is highly reactive in nucleophilic addition reactions. Alternatively, a chemically reactive moiety may be a nucleophilic group, such as an amino or
sulfhydryl group, and the conversion comprises reaction with a bifunctional reagent. It will be evident to one skillful in the art that a variety of bifunctional reagents, both homobifunctional and heterobifunctional, may be employed within the present invention.

A third method for providing a conjugate is to incorporate a targeting agent into a compound, which is suitable for radiolabeling, as the compound is being synthesized. In other words, a targeting agent is covalently attached to a precursor of a compound suitable for radiolabeling. An example of such a compound is a precursor of one of the chelating compounds described above. Following attachment of a targeting agent to a precursor of a compound, the synthesis of the compound is completed so that it will be suitable for radiolabeling.

The above-described chelating compounds are radiolabeled with an appropriate radionuclide to produce the corresponding chelate. The radiolabeling step may be done either before or after the compound is attached to the targeting agent. Conventional procedures are used for radiolabeling the chelating compounds. For example, pertechnetate (99mTcO₄⁻) or perrhenate (186 or 188 ReO₄⁻) are generally contacted with a chelating compound in the presence of a reducing agent to effect reduction of the radionuclide to an oxidation state at which chelation can occur. Many suitable reducing agents are known, including but not limited to stannous ion, (e.g., in the form of stannous salts such as stannous chloride or stannous fluoride), metallic tin, dithionite, formamidine sulfonic acid, ferrous ion (e.g., in the form of ferrous salts such as ferrous chloride, ferrous sulfate, or ferrous ascorbate), and alkali salts of borohydride. Preferred reducing agents are stannous salts.

Alternatively, the pertechnetate or perrhenate may be reduced in the presence of a relatively labile complexing agent such as gluconic acid or citric acid to form intermediate exchange complexes (99mTc-glucurate or 186Re-citrate). When the exchange complexes are contacted with the chelating compound under appropriate reaction conditions, the radionuclide metal is transferred to the chelating compound, thereby producing a radionuclide metal chelate.

The formation of intermediate exchange complexes is desirable for the radiolabeling of a number of chelating compounds with certain radionuclides. Improved yields of the radiolabeled chelate may be achieved through the use of complexing agents, compared to the yields resulting from reaction of chelating compounds alone with a radionuclide.

The exchange complex may serve to stabilize the radionuclide. For example, certain radionuclides are available in oxidation states at which chelation by the above-described chelating compounds does not occur. Reducing agents are used to effect reduction of the radionuclide to an oxidation state at which chelation can occur, as described for 99mTc and 188Re as above. Contacting the radionuclide with a reducing agent in the presence of a complexing agent to form an exchange complex serves to stabilize the reduced radionuclide thereby minimizing reoxidation thereof. Exchange complexes may be used to minimize reoxidation of such "redox active" radionuclides as radioisotopes of Tc, Re, Ru, Rh, Fe, Co, Cr, V, Sn, and Os, among others.

Exchange complexes also may be used to stabilize certain radionuclides so that polymerization thereof (e.g., formation of hydroxides) is minimized. Radionuclides that are susceptible to polymerization include, but are not limited to, radioisotopes of W, Mo, Ni, and Nb.

The exchange complexes often have a faster rate of formation (i.e., faster kinetics of complex formation), but a lower stability with respect to radionuclide binding than the radionuclide metal chelate compounds. Thus, the complexing agent binds the radionuclide much more quickly than does the chelating compound. The rapid formation of the exchange complex stabilizes the radionuclide, whereas the radionuclide would remain unbound longer (providing more opportunity for reoxidation or polymerization) if contacted with the chelating compound alone.

Radionuclides may be reacted with complexing agents at a wide range of temperatures to form exchange complexes (e.g., from 0°C to 100°C, preferably from room temperature to about 37°C). For convenience, the reaction is generally conducted at room temperature (about 18°C - 25°C) for many complexing agents. Due to the fast kinetics of complex formation, reaction for less than one hour, preferably about 10 minutes, is sufficient for exchange complex formation with most complexing agents.

Since exchange complexes are less stable (e.g., less thermodynamically stable) than the final chelate compound with respect to radionuclide binding, contacting the exchange complex with the chelating compound may result in transfer of the radionuclide to the chelating compound. Heating is often required to promote transfer of the radionuclide. A radionuclide may be reacted with a reducing agent in the presence of a complexing agent to form an exchange complex. The chelating compound then is added to the reaction mixture, followed by heating to promote transfer of the radionuclide. Alternatively, the radionuclide, reducing agent, complexing agent, and chelating compound all may be added to a single reaction mixture and heated together to form the final chelate. The improved yields of the radionuclide chelate that result when a
complexing agent is used demonstrate that the complexing agents facilitate formation of the chelate.

Lyophilized preparations comprising two or more reagents may be used in the radiolabelling reactions. For example, a reducing agent and complexing agent may be combined in an aqueous solution and then lyophilized. Optionally, one or more stabilizer compounds may be added prior to lyophilization. Many such stabilizer compounds are known. See, for example, U.S. Patent Nos. 4,440,738 and 4,510,125. Advantageously, gentisic acid is added to stabilize (e.g., to prevent oxidation of) a stannous ion reducing agent. A "filler compound" also may be added to the solution to provide bulk or mass and to aid in the lyophilization process. Good results have been achieved using lactose as the filler compound.

An appropriate solvent is added to reconstitute the lyophilized preparation and the radionuclide is then added to initiate the radiolabelling reaction. Alternatively, an aqueous solution eluted off a radionuclide generator may be added directly to the lyophilized preparation.

A number of compounds are suitable for use as complexing agents. Examples of complexing agents that have been used to facilitate transfer of metals to proteins or strong chelating agents attached thereto include the polyhydroxycarboxylates, glucoheptonate, Burchiel et al., J. Nucl. Med. 27:896 (1986), and tartrate, Kasina et al., Proc. Intl. Radio. Chem. Symp. 269-71 (1986). Additional examples of complexing agents include, but are not limited to, gluconic acid, glucoheptonic acid, methylene diphosphonate, glyceric acid, glycolic acid, mannitol, oxalic acid, malonic acid, succinic acid, N,N'-bis(2-hydroxyethyl) ethylene diamine, citric acid, ascorbic acid and gentisic acid. Good results are obtained using gluconic acid or glucoheptonic acid as the complexing agent for 99mTc, and citric acid as the complexing agent for 186 or 188Re.

In one embodiment of the invention, the complexing agent is a compound of the formula:

\[
\begin{align*}
\text{R}_1 & \quad \text{HO—} \quad \text{C—} \quad \text{CO}_2\text{H} \\
\text{R}_2 &
\end{align*}
\]

wherein R1 and R2 are independently selected from hydrogen or lower alkyl (C1-C4) groups. Alternatively, R1 is hydrogen or a lower alkyl group (C1-C4) and R2 is selected from -CH2 COOH, -CH2OH, or -COOH. In another alternative, R1 is OH and R2 is -C(OH)2 COOH. Examples of some of the many complexing agents of this formula are malic acid, citramalic acid, 2-hydroxy-isobutyric acid, 2-ethyl-2-hydroxybutyric acid, glyceric acid, hydroxymalonic acid, 2,3-dihydroxy tartaric acid, and lactic acid. The structures of two complexing agents that have been used with good results are as follows:

- 2-Hydroxyisobutyric Acid

\[
\begin{align*}
\text{CH}_3 & \quad \text{HO—} \quad \text{C—} \quad \text{CO}_2\text{H} \\
\text{CH}_3 &
\end{align*}
\]

- 2-Ethyl-2-hydroxybutyric Acid

\[
\begin{align*}
\text{CH}_2\text{CH}_3 & \quad \text{HO—} \quad \text{C—} \quad \text{CO}_2\text{H} \\
\text{CH}_2\text{CH}_3 &
\end{align*}
\]

The radionuclide chelate compounds produced by using the above-described exchange complexes are used in the preparation of the conjugates of the present invention that comprise a radiolabeled chelate compound attached to a targeting agent that is capable of associating with calcified tissue. The chelates produced using these exchange complexes also may be attached to other molecules to radiolabel the molecules.
For example, a chemically reactive moiety (a conjugation group) on a chelate may be reacted with a functional group on a targeting protein to bind the chelate thereto. See, for example, European Patent Application Publication No. 188,256. The targeting protein is a protein such as an antibody (preferably a monoclonal antibody) that binds to a desired target site within a human or mammalian host. The radiolabeled targeting proteins have diagnostic or therapeutic use, depending on the radionuclide in the chelate. An example of a target site is a tumor (cancer cells).

The present invention also provides a method for radiolabelling a chelating compound to form a radionuclide metal chelate. The method comprises reacting a radionuclide metal with a complexing agent to form an exchange complex, and contacting the exchange complex with a chelating compound under conditions such that the radionuclide metal is transferred from the exchange complex to the chelating compound, thereby forming said radionuclide metal chelate. The complexing agent is a compound of the formula:

\[
\text{HO} - \text{C} - \text{CO}_2\text{H}
\]

as described above. Preferably, the complexing agent is 2-hydroxyisobutyric acid or 2-ethyl-2-hydroxybutyric acid.

The chelating compound may be selected from any of the above-described chelating compounds comprising nitrogen and sulfur donor atoms. The radionuclide may be a radioisotope of technetium or rhenium, among others. The radionuclide metal may be reacted with the complexing agent in the presence of a reducing agent, and stabilizers and fillers also may be present. The chelating compound may be present in the reaction mixture during reaction of the radionuclide with the complexing agent. Alternatively, the chelating compound may be added after formation of the exchange complex, as described above.

The conditions under which the exchange complex is contacted with the chelating compound will vary according to such factors as the chemical structure of the particular complexing agent and chelating compound used. The reaction mixture containing the exchange complex and the chelating compound commonly is heated to promote transfer of the radionuclide, often to a temperature between 50 °C and 100 °C. Other reaction conditions (e.g. pH) are chosen based on such factors as the conditions required for displacement of protecting groups from sulfur donor atoms.

Kits comprising a chelating compound and an exchange agent selected from 2-hydroxyisobutyric acid and 2-ethyl-2-hydroxybutyric acid also are disclosed. The kits are useful for preparing radionuclide metal chelates in accordance with the above-described method.

Kits are disclosed for producing radiolabeled compound-targeting agent conjugates for radiopharmaceutical use. Two types of diagnostic and therapeutic kits are prepared for use in the administration of chelate-targeting agent conjugates. The first type of kit ("pre-formed") comprises a compound to be radiolabeled and a targeting agent, each in separate containers. The compound is labeled with a radionuclide and then the resulting radiolabeled compound is joined to the targeting agent. Thus, the radionuclide is added to the compound prior to the addition of the targeting agent.

The second type of kit ("post-formed") comprises in one container a compound to be radiolabeled attached to a targeting agent and in another container one or more reagents for facilitating the radiolabeling, e.g., a reducing agent and a complexing agent which in combination are capable of allowing a metal, or metal oxide, radionuclide to form an exchange complex. Thus, the radionuclide is added to the compound after the addition of the targeting agent.

Preferred radionuclides for use in conjunction with a diagnostic kit include \( ^{99m}\text{Tc} \), \( ^{57}\text{Ru} \) and \( ^{203}\text{Pb} \) and with a therapeutic kit include \( ^{186}\text{Re} \), \( ^{188}\text{Re} \), \( ^{67}\text{Cu} \), \( ^{105}\text{Rh} \), \( ^{198}\text{Au} \), \( ^{199}\text{Au} \) and \( ^{212}\text{Bi} \). The discussion provided above regarding targeting agent is applicable here as well.

Yet another aspect of the invention provides the radiolabeled compound-targeting agent conjugates described above for use for diagnostic and therapeutic purposes. Targeting agents, such as those described above, direct the conjugates to calcified tissue. Cancers and other disease states lead to increased pathological calcification. The targeting agents associate to a greater extent with those tissues undergoing increased calcification. Thus, abnormal calcified tissue, e.g., bone cancer, may be distinguishable from normal calcified tissue by increased levels of radioactivity delivered by a conjugate of the
invention. Uses of the conjugates as skeletal imaging or scanning agents include: to visualize areas of altered blood flow to the bone and altered osteogenic activity, including suspected bone lesions not shown on an X-ray bone survey performed as part of a work-up in patients with known or suspected malignancy, to follow the response of metastatic or primary bone lesions to radiation therapy, to diagnose arthritis, metabolic bone disease and osteomyelitis, and to diagnose and determine the healing rate of bone fractures.

The diagnostic conjugates of the invention may be for use to detect the presence or absence of a calcified tissue target site, e.g., bone cancer, within a mammalian host. The use comprises the steps of administering to a mammal a diagnostically effective dose of one of the conjugates described above, where the conjugate is capable of associating with the target site. This step is followed by a step of detecting the biodistribution of the radionuclide in the mammal to determine the presence or absence of the target site in the host.

A diagnostically effective dose of a conjugate is generally from about 5 to about 35 and typically from about 10 to about 30 mCi per 70 kg body weight. The precise dose may depend upon the particular route of administration, e.g., intravenous, intracompartmental, intraarticular or intratumoral. It will be evident to one skilled in the art how to determine the optimal effective dose for a particular chelate-targeting agent conjugate and a particular route of administration. The discussion provided above regarding targeting agent is applicable here as well. Preferred radionuclides include $^{99m}$Tc and $^{203}$Pb. A preferred mammal is man.

To summarize the examples which follow, Example I describes the conjugation of a diphosphonate to a chelating compound. Example II describes the incorporation of a diphosphonate during synthesis of a chelating compound. Example III provides the preparation of a radiolabeled chelate and its conjugation with a bone-seeking agent. Example IV provides the preparation of a bone-seeking agent-chelating compound conjugate, followed by labeling with $^{99m}$Tc or $^{188}$Re. Example V describes biodistribution studies in mice.

The following examples are offered by way of illustration and not by way of limitation.
EXAMPLES

EXAMPLE I

Conjugation of Diphosphonate to Chelating Compound

A solution of 3-amino-1-hydroxypropane-1,1-diphosphonic acid (2) and 4.0 equivalents of sodium hydrogen carbonate in water is stirred while a solution of 1 in 1,2-dimethoxyethane is added dropwise over a period of 0.5 hours. The mixture is then stirred at room temperature for 2.0. The solvent is removed in vacuo and the resulting residue is eluted through a column of DEAE sephadex using aqueous triethylammonium bicarbonate. The product containing fractions are combined and concentrated to a residue. The final product is isolated as a sodium salt by precipitation from acetone with sodium perchlorate.

EXAMPLE II

Incorporation of Diphosphonate During Synthesis of Chelating Compound

A. Synthesis of 4,5-Diamino-1-Hydroxypentane-1,1-Diphosphonic Acid

A mixture of 4,5-diaminopentanoic acid dihydrochloride and phosphorous acid (2.7 equivalents) in n-propyl sulfone is heated at 100°C while 2.1 equivalents of phosphorous trichloride is added dropwise. The reaction mixture is heated for three hours, then 25 ml of water is added and the heating continued for 1.5 hours. The solution is filtered and transferred to a separating funnel. The aqueous layer is transferred to a flask and 20 mls of methanol is added dropwise and the mixture is refluxed overnight. The mixture is then cooled to give a precipitate. The solution is decanted off and 10 ml of H₂O is added to the gummy
precipitate and the mixture is then heated to reflux for 1.5 hours. The mixture is cooled to give a precipitate which is collected by filtration and washed with cold methanol/water then washed with ether. The final product is then air dried.

B. Synthesis of \( \text{N}_2\text{S}_2 \) Ligand Diphosphonic Acid 1:

\[
\begin{align*}
\text{OH} & \quad \text{PO} \\
\text{NH}_2 & \quad \text{NH}_2 \\
\begin{array}{c}
\text{O} \\
\text{PO(OH)}_2
\end{array} & \quad \begin{array}{c}
\text{RS} \text{CO}_2\text{R} \\
\text{DME, NaHCO}_3, \text{H}_2\text{O}
\end{array}
\end{align*}
\]

\( R = \) active ester, e.g.

\( R' = \) sulfur protective group, e.g., etc. The \( \text{N}_2\text{S}_2 \) ligand diphosphonic acid is prepared in a manner similar to that described in M. Bodansky, Reactivity and Structure Concepts in Organic Chemistry, Vol. 21, The Practice of Peptide Synthesis, Springer-Verlag, 1984, p. 138:

\[
\begin{align*}
\text{O} & \quad \text{N} \\
\text{O} & \quad \text{CH}_3 \\
\text{N} & \quad \text{H}
\end{align*}
\]

A solution of 4,5-diamino-1-hydroxypentane-1,1-diphosphonic acid and 4.0 equivalents of sodium hydrogen carbonate in water is stirred while a solution of \( \text{S-(1-ethoxy)ethylmercaptoacetic acid succinimidyl ester} \) (2.5 equivalents) in 1,2-dimethoxyethane is added dropwise over a period of 0.5 hour. The mixture is then stirred at room temperature for an additional 3.0 hours. The solvents are removed in vacuo and the resulting residue is eluted through a column of DEAE sephadex using aqueous triethylammonium bicarbonate. The product containing fractions are combined and concentrated to a residue. The product is isolated by dissolving the residue in acetone and precipitating the material as the sodium salt by addition of a solution of sodium perchlorate in acetone.

EXAMPLE III

Preparation of Radiolabeled Chelate and Conjugation With Bone-Seeking Agent

A. Radiolabeling of Chelating Compounds

1. Technetium-99m labeling

Method a: To 100 \( \mu\)L of solution containing 5 mg of sodium gluconate and 0.1 mg of \( \text{SnCl}_2 \) in water, at \( \text{pH} = 6.1 \), 1 mL of \( 99m\text{TcO}_4^- \) (pertechnetate; 75-100 mCi) is added. After incubation at room temperature for 10 min. to form a \( 99m\text{Tc}-\text{gluconate complex} \), 100 \( \mu\)g of a chelating compound dissolved in i-
propanol:acetic acid, 90:10, at 1 mg/mL, 80 µl of 0.2 N HCl and 200 µL of i-propanol are added in that order. The reaction mixture is heated to 75 °C for 15 min., then cooled in ice for 5 min.

Method b: Alternatively, 1.0 mL of 99mTc generator eluted pertechnetate (75-100 mCi) is added to a vial containing a lyophilized mixture comprising sodium gluconate, stannous chloride, 0.1 mg; gentisic acid, 0.1 mg; lactose, 25 mg at pH 6.1. The vial is agitated gently to mix the contents by inversion and then incubated at room temperature for 10 min. to form a 99mTc–gluconate complex. Then 100 µg of a chelating compound dissolved in i-propanol:acetic acid, 90:10, at 1 mg/mL, 80 µl of 0.2 N HCl and 200 µL of i-propanol are added in that order. The reaction mixture is heated to 75 °C for 15 min., then cooled in ice for 5 min.

2. Rhenium-188 labeling

The 188 Re chelate of the chelating compound is prepared by a similar procedure. Sodium perrhenate (3 mL, 30 mCi/mL produced from a 188 W/188 Re generator) is added to a vial containing a lyophilized mixture comprising citric acid, 75 mg; stannous chloride, 0.75 mg; gentisic acid, 0.25 mg and lactose, 100 mg. The vial is agitated gently to mix the contents, then incubated at room temperature for 10 min. To a separate vial containing 0.50 mg of the chelating compound, 0.50 mL of 2-propanol is added and the vial is agitated for 2 min. to completely dissolve the compound. Then 0.3 mL of this solution is transferred to the vial containing the 188 Re-citrate complex. The reaction mixture is heated to 75 °C for 15 min., then cooled in ice for 5 min.

3. Rhenium-186 labeling

The 186 Re chelate of the chelating compound is prepared by a procedure similar to the 188 Re procedure. Perrhenate 186ReO4 as sodium, lithium, ammonium or any other suitable counter ion is obtained from irradiation of 185 Re (0.1 to few mg) in a nuclear reactor dissolution and purification of the original solution by previously described procedure (Vanderheyden et al., Inorg. Chem. 24:1666, 1985) or any other suitable procedure. To the 186ReO4 vial is added 0.75 mL reconstituted solution made of 1.0 mL of sterile water added to a vial containing a lyophilized mixture comprising citric acid 25 mg, stannous chloride 1 mg, gentisic acid 1 mg and lactose 75 mg.

The vial is agitated gently to mix the contents, then incubated at room temperature for 10 min. To the separate vial containing the chelating compound, 2-propanol is added to obtain a completely dissolved solution. An appropriate amount of this solution to give a ligand to rhenium mole ratio of 1:5 (preferentially between 1 and 1.5 ligand to rhenium mole ratio) is then transferred to the vial containing the 186 Re-citrate complex. The reaction mixture is heated to 75 °C for 20 min. then cooled on ice for 5 min.

B. Conjugation of Radiolabeled Chelate With Bone-Seeking Agent

To the above 99mTc chelate, 100 µl of bicarbonate buffer is added so that the pH of the solution is about 6.0. Next, 100-500 µl of a solution containing a bone-seeking agent at 0.1 to 10 mg/mL is added in the same buffer to reach a final pH of 8 to 9.5. The reaction mixtures are incubated at room temperature for 5-30 min. as necessary. The ITLC procedure (D. Bernier et al. (eds.), Nuclear Medicine Technology and Techniques, The C.V. Mosby Co., St. Louis, 1981, pp. 172-174) using 100% CH3CN is utilized to determine the percentage of the conjugate.

If necessary, an extra purification step is added to increase the purity of the conjugate. This involves passing the conjugate through on ion exchange (DEAE, CAC, Sephadex, ACCEL® or the like) or reverse phase column. The conjugate is either collected in the flow through or eluted with solution of appropriate ionic strength.

EXAMPLE IV

Preparation of Bone-Seeking Agent-Chelating Compound Conjugate Followed by Labeling With 99mTc or 188 Re

A. Preparation of the Conjugate

The antibody conjugation reaction is performed in a final volume of 4.0 mL: 1 mg of a chelating compound (61.67 µmol), 1.1 mg of a bone-seeking agent, 1-2 mL of distilled dimethyl formamide (if
B. Radiolabeling of the Conjugate

1. Technetium-99m labeling

Method a: Stannous gluconate kits are prepared containing 5 mg sodium gluconate, 100 µg stannous chloride, 100 µg of chelating bone-seeking ligand and 25 mg of lactose. The pH is contained between 2 and 7 using HCl, Acetic acid or NaOH. To the stannous gluconate kit is added 1.0 mL sodium pertechnetate (Tc-99m) about 50 mCi. The vial is then adjusted to 7-7.4. The percent formation of the conjugate, remaining TcO⁺ and hydrolyzed reduced technetium is determined by ITLC SG using 100% CH₃CN and HPLC reverse phase using a C₁₈ column and a mobile phase ranging between 0% and 40% acetonitrile/phosphate buffer. The technetium incorporation typically ranges from 75% to 99% on a chelating bone-seeking conjugate.

Method b: Sodium perrhenate (3 mL, 15 mCi, produced from a ¹⁸⁸W/¹⁸⁸Re generator) is added to a vial containing a lyophilized mixture comprising: citric acid, 75 mg; stannous chloride 0.75 mg; gentisic acid 3 mg; lactose 75 mg and 0.1-1 mg of the chelating bone-seeking ligand. The pH is contained between 1.5 and 7 using HCl, acetic acid or NaOH. The vial is heated at 75°-100°C for 5-15 minutes. The percent formation of the conjugate, remaining TcO⁺ and hydrolyzed reduced technetium is determined by ITLC SG using 100% CH₃CN and HPLC reverse phase using a C₁₈ column and a mobile phase ranging between 0% and 40% acetonitrile/phosphate buffer. The technetium incorporation typically ranges from 75% to 99% on a chelating bone-seeking conjugate.

Method c: Stannous tartrate kits are prepared in an evacuated vial under nitrogen to contain 0.5 mL of disodium tartrate (10 mg/ml) and 0.1 mL stannous chloride (1.0 mg/ml in ethanol). The pH of the solution is kept between 4 and 7, preferably 6.0. To this stannous solution is added 0.5 mL of sodium dithionite: a 100 µL aliquot is removed and quickly added to the pertechnetate liquid vial. The reaction vial is heated at 60°-75°C for 5-15 min. After a few minutes cooling at room temperature, 100 µL of 1 N HCl and 0.3 µL of 0.2 M sodium phosphate pH 7.4 are added. The percent formation of the conjugate, remaining TcO⁺ and hydrolyzed reduced technetium is determined by ITLC SG using 100% CH₃CN and HPLC reverse phase using a C₁₈ column and a mobile phase ranging between 0% and 40% acetonitrile/phosphate buffer. The technetium incorporation typically ranges from 75% to 99% on a chelating bone-seeking conjugate.

2. Rhenium-188 labeling

Method a: Stannous gluconate kits are prepared containing 5 mg sodium gluconate, 100 µg stannous chloride, 100 µg of chelating bone-seeking ligand and 25 mg of lactose. The pH is contained between 2 and 7 using HCl, Acetic acid or NaOH. To the stannous gluconate kit is added 1.0 mL sodium pertechnetate (Tc-99m) about 50 mCi. The vial is heated at 75°-100°C for 5-15 min. After a few minutes cooling at room temperature, 100 µL of 1 N NaOH and 1.0 ml of sodium pertechnetate (50 mCi). Extemporaneously, a stock solution containing a lyophilized mixture comprising: citric acid, 75 mg; stannous chloride 0.75 mg; gentisic acid 0.25-5 mg; lactose 75 mg and 0.1-1 mg of the chelating bone-seeking ligand. The pH is contained between 1.5 and 7 using HCl, acetic acid or NaOH. The vial is heated at 75°-100°C for 5-15 minutes. The pH is then adjusted to 7-7.4. The percent formation of the conjugate, remaining TcO⁺ and hydrolyzed reduced technetium is determined by ITLC SG using 100% CH₃CN and HPLC reverse phase using a C₁₈ column and a mobile phase ranging between 0% and 40% acetonitrile/phosphate buffer. The technetium incorporation typically ranges from 75% to 99% on a chelating bone-seeking conjugate.

Method b: Sodium perrhenate (3 mL, 15 mCi, produced from a ¹⁸⁸W/¹⁸⁸Re generator) is added to a vial containing a lyophilized mixture comprising: hydroxy isobutyric acid 50-300 mg; stannous chloride 0.5-5 mg; gentisic acid 0.25-5 mg; lactose 75 mg and 0.1-1 mg of the chelating bone-seeking ligand. The pH is contained between 1.5 and 7 using HCl, acetic acid or NaOH. The vial is incubated at room temperature for 5-60 min. After a few minutes cooling at necessary, the pH is raised to 8-9.5 using a sufficient amount of a carbonate buffer or NaOH. The vial is incubated again at room temperature for 5-60 min. The pH is then adjusted to 7-7.4. The percent formation of the conjugate, remaining rhenium oxide and hydrolyzed reduced rhenium is determined by ITLC SG using 100% CH₃CN and HPLC reverse phase using a C₁₈ column and a mobile phase ranging between 0%
and 40% acetonitrile/phosphate buffer. The rhenium incorporation typically ranges from 75% to 99% on a chelating bone-seeking conjugate.

3. Rhenium-186 labeling

Method a: The $^{186}$Re chelate form of the conjugate is obtained by a procedure similar to the $^{186}$Re labeling of the chelating compound described above. Perrhenate $^{185}$Re as sodium, lithium, ammonium or any other suitable counter ion is obtained from irradiation of $^{185}$Re (0.1 to few mg) in a nuclear reactor dissolution and purification of the original solution by previously described procedure (Vanderheyden et al., Inorg. Chem. 24:1666, 1985) or any other suitable procedure. To the purified perrhenate vial is added 0.75 mL of the reconstituted solution made of 1.0 mL of sterile water added to a vial containing a lyophilized mixture comprising citric acid 25 mg, stannous chloride 1 mg, gentisic acid 1 mg, lactose 75 mg and 0.1-10 mg of the chelating bone-seeking ligand. The vial is incubated at 75 °C for 15-30 min., then cooled at room temperature. The pH is then adjusted to 7.0-7.4. The percent formation of the conjugate, remaining rhenium oxide and hydrolyzed reduced rhenium is determined by ITLC SG using 100% CH3CN and HPLC reverse phase using a C18 column and a mobile phase ranging between 0% and 40% acetonitrile/phosphate buffer. The rhenium incorporation typically ranges from 75% to 99% on a chelating bone-seeking conjugate.

Method b: Perrhenate $^{186}$ReO4 (prepared as described above) is added to a vial containing a lyophilized mixture comprising a hydroxy isobutyric acid 50-300 mg; stannous chloride 0.5-5 mg; gentisic acid 0.25-5 mg; lactose 75 mg and 0.1-10 mg of the chelating bone-seeking ligand. The pH is contained between 1.5 and 7 using HCl, acetic acid or NaOH. The vial is incubated at room temperature for 5-60 min. or heated at 75°-100° for 5-15 min. After a few minutes cooling if necessary, the pH is raised to 8-9.5 using a sufficient amount of a carbonate buffer or NaOH. The vial is incubated again at room temperature for 5-60 min. The pH is then adjusted to 7-7.4. The percent formation of the conjugate, remaining rhenium oxide and hydrolyzed reduced rhenium is determined by ITLC SG using 100% CH3CN and HPLC reverse phase using a C18 column and a mobile phase ranging between 0% and 40% acetonitrile/phosphate buffer. The rhenium incorporation typically ranges from 75% to 99% on a chelating bone-seeking conjugate.

EXAMPLE V

Biodistribution Studies in Mice

Conjugates are injected into Balb/c mice. The biodistribution of the conjugate is analyzed for 2 hours after injection according to the method of Hwang et al., Cancer Res., 45: 4150-4155 (1985). The data is collected in terms of the percentage of the injected radioactivity per gram (% inj. dose/gram) of each specified tissue type and bone/tissue ratio of injected radioactivity. The tissue types evaluated are as follows: blood (BL); tail (TA); heart (TU); skin (SK); muscle (MU); bone (BO); lung (LU); liver (LI); spleen (SP); stomach (ST); thyroid (TH); kidney (KI); and intestine (IN). Figures 1 and 2 compare $^{99m}$Tc-labeled phosphonates, e.g., "MDP" and "ADP-KIT," with representative conjugates.

EXAMPLE VI

Preparation of Radiolabeled Chelate Compounds and Attachment Thereof to Targeting Agents

The compounds 2-hydroxy-isobutyric acid and 2-ethyl-2-hydroxy butyric acid were used as complexing agents in the radiolabeling of an N2S chelating compound having the following structure:
wherein \( T \) represents a sulfur protecting group and \( Q \) represents a conjugation group. One such compound comprises a hemithioacetal sulfur protecting group such as an ethoxyethyl group of the formula:

wherein the sulfur atom is the sulfur donor atom of the chelating compound. One example of a suitable conjugation group \( Q \) is an ester group such as a tetrafluorophenyl ester of the formula:

The chelating compound was radiolabeled with \(^{99m}\)Tc by combining the following reagents (solvents were purged with nitrogen):

- lactose (20 mg);
- 2-ethyl-2-hydroxybutyric acid (5 mg, 1 mg/mL in saline);
- gentisic acid (100 \( \mu \)g in 0.1 mL saline);
- sodium pertechnetate (100 \( \mu \)l eluent, about 100 mCi, from a \(^{99}\)Mo/\(^{99m}\)Tc generator, generators are commercially available from DuPont, Mediphysics, E.R. Squibb, or Mallinckrodt);
- \( \text{SnCl}_2 \) (100 \( \mu \)g in 0.1 mL ethanol, purged with \( \text{N}_2 \) prior to the addition of the reagents); and
- chelating compound (100 \( \mu \)g, 1 mg/mL in isopropyl alcohol).

The reaction vial was shaken and heated for ten minutes at 75\( ^\circ \)C. Since the ethoxyethyl sulfur protecting group is displaced by metal assisted acid cleavage during radiolabeling, the radiolabeling reaction was conducted at acidic pH, preferably about 2.2-2.5 for this chelating compound. Analysis by HPLC showed 81% ester and ITLC showed 1.1% TcO\(_2\). The radiolabeled chelate compound may be purified using any suitable chromatography column. Good results were achieved using a pD10 column.

In another radiolabeling reaction, 1 mL of diluted \(^{99m}\)Mo/\(^{99m}\)Tc generator eluent (about 75-100 mCi) and 100-300 \( \mu \)L isopropyl alcohol were added to a lyophilized preparation comprising:

- lactose (20 mg);
- 2-ethyl-2-hydroxybutyric acid (5 mg);
- chelating compound (50 \( \mu \)g);
- gentisic acid (100 \( \mu \)g); and
- \( \text{SnCl}_2 \) (100 \( \mu \)g).
The reaction vial was immediately placed in a boiling water bath at 95-100 °C. Aliquots were taken at 1, 3, 5, 10 and 15 minutes. The pH of the reaction mixture was 2.77.

The ester yields as a function of the time points listed above are 76%, 88%, 68%, 60% and 45%, respectively.

The corresponding TcO₂ values are 5.2%, 3.4%, 3% and 4% (ITLC as thioglycolates).

From the above data, the optimal time at 100 °C is three minutes. The ester yield was 88%. The heating for longer periods decreases the yield. The TcO₂ values are very good at all time points.

2-Hydroxyisobutyric acid may be substituted for 2-ethyl-2-hydroxybutyric acid as the complexing agent in the above radiolabeling reactions. Other chelating compounds may be used as well. In alternative radiolabeling reactions, the components other than the chelating compound are incubated for 10 minutes at room temperature to form the exchange complex. The chelating compound then is added, and the reaction mixture is heated to 75-100 °C to form the chelates.

The ⁹⁹ᵐTc-labeled chelate produced by the above radiolabeling procedures is attached to a targeting agent. A conjugation moiety on the chelate (e.g., the tetrafluorophenyl ester group, which survives the radiolabeling procedures conducted at acidic pH) is reacted with a functional group on the desired targeting agent to bind the chelate thereto.

Reactions for attaching chelates to bone-targeting agents are described above. Alternatively, the chelate may be reacted with a targeting protein such as an antibody in a buffered solution at a pH of about 9-11. The ester conjugation group will react with free amine groups on lysine residues of the protein.

Claims

1. A conjugate of a radiolabeled chelate and a targeting agent that associates with calcified tissue, wherein said chelate contains at least four donor atoms selected from nitrogen atoms or sulphur atoms or combinations thereof, and wherein said chelate does not function as said targeting agent.

2. The conjugate of Claim 1 wherein the chelate is formed from a chelating compound containing two nitrogen and two sulphur atoms, or three nitrogen and one sulphur atom or two nitrogen and three sulphur atoms, or two nitrogen and four sulphur atoms, or three nitrogen and three sulphur atoms, said nitrogen and sulphur atoms as donor atoms to which a radionuclide atom is bound.

3. The conjugate of claims 1 or 2 wherein the metal, or metal oxide, radionuclide of the chelate is selected from the group consisting of ⁶⁴Cu, ⁶⁷Cu, ⁵⁷Ru, ⁹⁹ᵐTc, ¹⁰⁵Rh, ¹⁰³Pd, ¹⁸⁵Re, ¹⁸⁸Re, ¹⁹⁸Au, ¹⁹⁹Au, ²⁰³Pb, ²¹²Pb, and ²¹²Bi.

4. The conjugate of Claim 1 wherein the targeting agent is a phosphorus-containing compound which contains a c-O-P bond, a P-O-P bond, a C-C-P bond, a P-C-P bond, a P-N-P bond or combinations thereof.

5. The conjugate of claim 1 wherein the calcified tissue is bone.

6. A conjugate of the formula:
7. A conjugate of the formula:

\[
\begin{align*}
&\text{NH-(CH}_2\text{)}^n\text{-C-OH} \\
&\text{CH}_2 \\
&\text{C=O} \\
&\text{PO}_3^3 \\
&\text{PO}_3^3 
\end{align*}
\]

wherein \( n \) is 1 to 4.

8. The conjugate of Claims 1 to 7 for use within a method of detecting the presence or absence of a calcified tissue target site.

9. The conjugate of Claims 1 to 7 for use as an active therapeutic substance.

10. The conjugate of Claim 8 or 9 wherein the calcified tissue target site is bone cancer.

Patentansprüche

1. Ein Konjugat eines radioaktiv markierten Chelats und eines targetierenden Agens, das mit kalzifiziertem Gewebe assoziiert, wobei besagtes Chelat mindestens vier Donoratome enthält, die aus Stickstoffatomen oder Schwefelatomen oder Kombinationen davon ausgewählt sind, und wobei besagtes Chelat nicht als besagtes targetierendes Agens fungiert.

2. Das Konjugat nach Anspruch 1, wobei das Chelat aus einer chelatbildenden Verbindung gebildet ist, die zwei Stickstoff- und zwei Schwefelatome, oder drei Stickstoff- und ein Schwefelatom oder zwei Stickstoff- und drei Schwefelatome, oder zwei Stickstoff- und vier Schwefelatome, oder drei Stickstoff- und drei Schwefelatome enthält, wobei besagte Stickstoff- und Schwefelatome als Donoratome fungieren, an die ein Radionuklidatom gebunden ist.

3. Das Konjugat nach Anspruch 1 oder 2, wobei das Metall-, oder Metalloxid-, Radionuklid des Chelates ausgewählt ist aus der Gruppe bestehend aus \( ^{64}\text{Cu}, ^{67}\text{Cu}, ^{97}\text{Ru}, ^{99m}\text{Tc}, ^{105}\text{Rh}, ^{109}\text{Pd}, ^{186}\text{Re}, ^{188}\text{Re}, ^{198}\text{Au}, ^{199}\text{Au}, ^{203}\text{Pb}, ^{212}\text{Pb}, ^{212}\text{Bi} \).

4. Das Konjugat nach Anspruch 1, wobei das targetierende Agens eine Phosphor-haltige Verbindung ist, die eine C-O-P-Bindung, eine P-O-P-Bindung, eine C-C-P-Bindung, eine P-C-P-Bindung, eine P-N-P-Bindung oder Kombinationen davon enthält.

5. Das Konjugat nach Anspruch 1, wobei das kalzifizierte Gewebe Knochen ist.
6. Ein Konjugat der Formel:

![Chemical Structure](image)

7. Ein Konjugat der Formel:

![Chemical Structure](image)

wobei n 1 bis 4 ist.

8. Das Konjugat nach den Ansprüchen 1 bis 7 zur Verwendung in einem Verfahren zum Nachweis der Anwesenheit oder Abwesenheit einer Target-Stelle in kalzifiziertem Gewebe.

9. Das Konjugat nach den Ansprüchen 1 bis 7 zur Verwendung als eine aktive therapeutische Substanz.

10. Das Konjugat nach Anspruch 8 oder 9, wobei die Target-Stelle in kalzifiziertem Gewebe Knochenkrebs ist.

Reventications

1. Conjugué d’un chélate radiomarqué et d’un agent cible qui s’associe avec un tissus calcifié, dans lequel le chélate contient au moins quatre atomes donneurs choisis parmi des atomes d’azote, des atomes de soufre ou des combinaisons de ces atomes et dans lequel le chélate n’agit pas comme agent cible.

2. Conjugué selon la revendication 1, dans lequel le chélate est formé d’un composé de chélatation contenant deux atomes d’azote et deux atomes de soufre, ou trois atomes d’azote et un atome de soufre, ou deux atomes d’azote et trois atomes de soufre, ou deux atomes d’azote et quatre atomes de soufre, ou trois atomes d’azote et trois atomes de soufre, les atomes d’azote et de soufre en tant qu’atomes donneurs auxquels un atome radionucléide est lié.
3. Conjugué selon la revendication 1 ou 2, dans lequel le métal, ou l'oxyde de métal, radionucléide du chélate est choisi parmi le groupe comprenant $^{64}\text{Cu}$, $^{67}\text{Cu}$, $^{97}\text{Ru}$, $^{99m}\text{Tc}$, $^{103}\text{Rh}$, $^{106}\text{Pd}$, $^{186}\text{Re}$, $^{188}\text{Re}$, $^{198}\text{Au}$, $^{199}\text{Au}$, $^{203}\text{Pb}$, $^{212}\text{Pb}$ et $^{212}\text{Bi}$.

4. Conjugué selon la revendication 1, dans lequel l'agent cible est un composé contenant du phosphore qui contient une liaison C-O-P, une liaison P-O-P, une liaison C-C-P, une liaison P-C-P, une liaison P-N-P, ou des combinaisons de ces liaisons.

5. Conjugué selon la revendication 1, dans lequel le tissu calcifié est un os.

6. Conjugué ayant la formule :

7. Conjugué ayant la formule :

où $n$ est compris entre 1 et 4.

8. Conjugué selon les revendications 1 à 7, destiné à être utilisé dans un procédé de détection de la présence ou de l'absence d'un site cible sur un tissu calcifié.

9. Conjugué selon les revendications 1 à 7, destiné à être utilisé comme substance thérapeutique active.

10. Conjugué selon la revendication 8 ou 9, dans lequel le site cible de tissu calcifié est un cancer osseux.
FIG. 1
COMPARISON OF MDP (SQUIBB) AND Tc-N2S2-ADP

% INJ. DOSE/GRAM.

Tissue: BL TA TU SK MU BO LU LI SP ST NE KI IN

FIG. 2A
-ADP-KIT vs ADP-N2S2 & ADP N3S-
-IN B/C AT 2HR

% INJ. DOSE/GRAM.

Tissue: BL TA TU SK MU BO LU LI SP ST TH KI IN

ADP-KIT ADP-N2S2 ADP-N3S
FIG. 2B

- Bone/Tissue Ratio -

ADP-KIT    ADP-N2S2    ADP-N3S

Tissue: BL TA TU SK MU BO LU LI SP ST TH KI IN

BL     TA     TU     SK     MU     BO     LU     LI     SP     ST     TH     KI     IN