

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization

International Bureau



(10) International Publication Number

WO 2014/043058 A1

(43) International Publication Date

20 March 2014 (20.03.2014)

(51) International Patent Classification:

A01N 63/02 (2006.01) A01P 5/00 (2006.01)

(21) International Application Number:

PCT/US2013/058866

(22) International Filing Date:

10 September 2013 (10.09.2013)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

61/700,054 12 September 2012 (12.09.2012) US

(71) Applicant: BAYER CROPSCIENCE LP [US/US]; 2 T.W. Alexander Drive, Research Triangle Park, NC 27709 (US).

(72) Inventors: RIGGS, Jennifer, Lynn; 2521-101 Gordon Glen Court, Raleigh, NC 27617 (US). CALA, Eder Leonardo, Sastoque; 3907 Eagle Ridge Lane, Opelika, AL 36801 (US). KLOEPPEP, Joseph, W.; 735 S. Gay Street, Auburn, AL 36830 (US). LAWRENCE, Kathryn, Kay Scott; 307 Willow Creek Road, Auburn, AL 36832 (US). RUSSI, Juan David, Castillo; 313 E. Thatch Avenue, Apt. 105, Auburn, AL 36830 (US).

(74) Agent: MCBEE, Susan, E. Shaw; Miles & Stockbridge PC, 1751 Pinnacle Drive, Suite 500, Tysons Corner, VA 22102-3833 (US).

(81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IS, JP, KE, KG, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LT, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, KM, ML, MR, NE, SN, TD, TG).

Published:

— with international search report (Art. 21(3))



WO 2014/043058 A1

(54) Title: COMPOSITIONS AND METHODS FOR CONTROLLING PLANT-PARASITE NEMATODE

(57) Abstract: An entity having anti-nematodal activity derived from *Bacillus firmus* bacteria selected from the group consisting of isolated biosurfactant, isolated protease enzyme, isolated amylase, isolated lipase, and isolated cellulase, is disclosed. The disclosure also provides for compositions comprising the same, methods of making the same, methods of controlling plant-parasite nematode, and methods of protecting plants.

COMPOSITIONS AND METHODS FOR CONTROLLING PLANT-PARASITE NEMATODE

CROSS REFERENCE TO RELATED APPLICATION

[0001] This application claims priority to U.S. Provisional Application Serial No. 61/700,054, filed September 12, 2012, the content of which is herein incorporated by reference in its entirety.

FIELD

[0002] An entity having anti-nematodal activity derived from *Bacillus firmus* bacteria selected from the group consisting of isolated biosurfactant, isolated protease enzyme, isolated amylase, isolated lipase, and isolated cellulase, is disclosed. The disclosure also provides for compositions comprising the same, methods of making the same, methods of controlling plant-parasite nematode, and methods of protecting plants.

BACKGROUND

[0003] Bacteria are an important group of natural antagonists of plant-parasitic nematodes. Bacteria are distributed broadly, have diverse modes of action, and have broad host ranges (Tian, B. *et al.*). They exhibit diverse modes of action against nematodes that include parasitism, production of toxins, antibiotics, or lytic enzymes; induce systemic resistance, and promote plant health (Aatlen, P.M. *et al.*; Kerry, B.R. 2000; Kerry, B.R. 1987; Siddiqui, Z.A. *et al.*; Stirling, G.R. 1991; Tian, B. *et al.*; Van Loon, L.C. *et al.*). Furthermore, bacteria can be in direct contact with the entrance sites of the nematodes and influence root exudates that can affect the nematode development (Sikora, R.A. 1992). The genera *Pasteuria*, *Pseudomonas*, and *Bacillus* have shown promising potential for nematode biocontrol (Meyer S.L.F; Siddiqui, Z.A. *et al.*; Stirling, G.R.; Tian, B. *et al.*).

[0004] *Bacillus firmus* strain GB-126 is a nematode biocontrol agent registered initially in a bionematicide in Israel under the trade name of BIONEM® WP (Blachinsky, D. *et al.*; Keren-Zur *et al.*). This formulation was shown to reduce galling index caused by *Meloidogyne* spp. on cucumber and tomato plots (Keren-Zur *et al.*). Also, under field conditions, suppression of *Meloidogyne* spp. was observed within 2 months of transplanting cucumbers and continued through the end of the experiments (Giannakou, O.I. *et al.*, 2004). Control provided by *B. firmus* GB-126 was less effective than the soil fumigant dazomet. However, its combination with soil solarization improves

nematode control giving results similar to dazomet use (Giannakou, O.I. *et al.* 2007). Furthermore, when *B. firmus* GB-126 was evaluated in tomato seedlings in the greenhouse, it reduced gall formation by 91 %, final nematode population by 76%, and the number of *M. incognita* eggs by 45% (Terefe, M. *et al.*).

[0005] In other study, a formulation of *B. firmus* that contains seaweed extract (BIONEM® L) was able to reduce *Helicotylenchus* spp. and *Tylenchorhynchus* spp. in golf greens (Wick, R.L. 2006). Furthermore, synergism of *B. firmus* with other nematode biocontrol agents has been reported to improve nematode reduction (Mendoza, A.R. *et al.* 2009). In banana, *B. firmus* was evaluated against *R. similis* and applied in combination with *F. oxysporum* and *P. lilacinus*, which reduced the infection of this migratory endoparasitic nematode (Mendoza, A.R. *et al.* 2009). Under *in vitro* conditions *B. firmus* was evaluated against the plant parasitic nematodes *Radopholus similis*, *Meloidogyne incognita*, and *Ditylenchus dipsaci*. *Bacillus firmus* produced bioactive secondary metabolites that were toxic to these nematode juveniles and reduced egg hatching (Mendoza, A.R. *et al.* 2008).

[0006] Thus, previous studies have demonstrated the antagonistic effect of bioactive secondary metabolites of *Bacillus firmus* GB-126 against plant parasitic nematodes. Nevertheless, the types of secondary metabolites and enzymatic properties involved and the role of possible induction of plant resistance have not been evaluated. Fewer and fewer traditional nematicide are available today and many that are available are very expensive or not environmentally and user friendly. Identification of a biological means to protection plants from pathogenic nematodes fits well into today's integrated pest management platform for agriculture. Understanding the single or multiple modes of actions of a biocontrol microbe can assist in the optimization of the growth of the microbe prior to formulating into a commercial product. Studies that evaluate the purpose of the various secondary metabolites can define which of the many metabolites should be optimized during the growth of the microbe. There are means available to optimize the growth media of the microbe to enhance production of one metabolite or another.

[0007] An object of the present invention is to provide a novel entity having anti-nematodal activity derived from *Bacillus firmus* bacteria selected from the group consisting of isolated biosurfactant, isolated protease, isolated amylase, isolated lipase, and isolated cellulase, is disclosed. Another object is to provide for compositions comprising the same. Another object is to provide for methods of making the same.

[0008] A further object is to provide for methods of controlling plant-parasite nematode, and methods of protecting plants.

SUMMARY

[0009] In an aspect, the disclosure provides for an a novel entity having anti-nematodal activity derived from *Bacillus firmus* bacteria selected from the group consisting of isolated biosurfactant, isolated protease, isolated amylase, isolated lipase, and isolated cellulose.

[0010] In an aspect, the disclosure provides for a novel composition having anti-nematodal activity comprising at least one entity derived from *Bacillus firmus* bacteria selected from the group consisting of an isolated biosurfactant, an isolated protease, an isolated amylase, an isolated lipase, and an isolated cellulase.

[0011] In an aspect, the disclosure provides for an extract of a *Bacillus firmus* culture having the following characteristics: anti-nematodal activity and presence of a biosurfactant.

[0012] In an aspect, the disclosure provides for an extract of a *Bacillus firmus* culture having the following characteristics: anti-nematodal activity and activity selected from the group consisting of protease activity, amylase activity, lipase activity, and cellulase activity.

[0013] In an aspect, the disclosure also provides for a method of making the same.

[0014] The disclosure also provides for a method of controlling plant-parasitic nematode by applying an entity or composition as described herein to a plant, the seed material, or the area on which the plant grows. In an aspect, the disclosure provides for a method of protecting a plant by applying an entity or composition as described herein to a plant, the seed material, or the area on which the plant grows.

[0015] In an aspect, the plant-parasitic nematode is *Rotylenchulus reniformis*.

BRIEF DESCRIPTION OF THE FIGURES

[0016] Fig. 1 sets forth the effects of *B. firmus* metabolites at 100% and 50% concentration on *R. reniformis* eggs as compared to water and media control.

[0017] Fig. 2 sets forth the effects of *B. firmus* metabolites at 100% and 50% concentration on *R. reniformis* second stage juveniles as compared to water and media control.

[0018] Fig. 3 sets forth tests of biosurfactant production of *B. firmus* GB-126. (A) represents the positive emulsification of kerosene. (B) represents halo formation due to cell lysis in blood agar. (C) represents positive oil drop collapse test.

- [0019]** Fig. 4 sets forth the effect of biosurfactant produced by *B. firmus* on *R. reniformis* second stage juveniles after 30 minutes of inoculation (P≤0.05) as compared to BP media and water controls and *B. firmus* bacteria.
- [0020]** Fig. 5 sets forth the effect of purified biosurfactant produced by *B. firmus* at different concentrations (ppm) on *R. reniformis* second stage juveniles after 30 minutes of inoculation under in vitro conditions (P≤0.05).
- [0021]** Fig. 6 sets forth the photos of second stage juveniles of *R. reniformis* form (control and after application of 2 ppm *B. firmus* biosurfactant) observed under SEM in *in vitro* trial.
- [0022]** Fig. 7 sets forth photos of growth of cotton plants after systemic resistance trial of *B. firmus* GM-126 and *S. maracescens* against *R. reniformis* in cotton plants.
- [0023]** Fig. 8 sets forth the enzyme reaction test of *B. firmus* GB-126: (A) represents production of proteases, (B) represents production of amylases, (C) represents production of cellulases, and (D) represents production of chitinases.
- [0024]** Fig. 9 sets forth *R. reniformis* life stages with cotton seeds treated with *B. firmus* GB-126 at 7 x 10⁶ cfu/seed under greenhouse conditions in autoclaved soil (P≤0.05).

DETAILED DESCRIPTION

- [0025]** In an aspect, the disclosure provides for an a novel entity having anti-nematodal activity derived from *Bacillus firmus* bacteria selected from the group consisting of isolated biosurfactant, isolated protease, isolated amylase, isolated lipase, and isolated cellulase.
- [0026]** Anti-nematodal activity is understood as meaning any activity useful in controlling or killing nematodes, such as reducing nematode egg hatching or paralyzing juveniles of nematodes. Testing for anti-nematodal activity is done both in-vitro and in-vivo through laboratory and greenhouse procedures. In-vitro testing can involve testing eggs and/or juvenile nematodes in selective media in small 96-well plates and measurements done by microscopic evaluations. Proper control treatments would be combined to monitor and compare activity. Under greenhouse studies soil was autoclaved using two 90 minute cycles at 130°C at 1.0 kg/cm³ pressure with a 24 hour cool down between cycles to remove any natural competition for the microflora. The vermiform stages were extracted from the soil by the modified gravity screening and sucrose centrifugation-flotation. Eggs stages were extracted from cotton roots by shaking the root system in a 1% NaOCl solution for four minutes at 120 rpm. The nematode suspension was collected and rinsed with water through a 25 µm sieve. Females in

roots were stained with acid fuchsin to facilitate enumeration of the females invading the root. Variables measured included plant height, shoot and root weights, females and eggs per gram of root, and the number of vermiform life stages in soil.

[0027] In an aspect, the strain of *Bacillus firmus* is GB-126. In an aspect, the entity described herein has anti-nematodal activity against Reniform nematode, *Rotylenchulus spp.*; Dagger nematode, *Xiphinema spp.*; Lance nematode, *Hoplolaimus spp.*; Pin nematode, *Paratylenchus spp.*; Ring nematode, *Criconemoides spp.*; Rootknot nematode, *Meloidogyne spp.*; Sheath nematode, *Hemicyclophora spp.*; Spiral nematode, *Helicotylenchus spp.*; Stubbyroot nematode, *Trichodorus spp.*; Cyst nematode, *Heterodera spp.*; Sting nematode, *Belonolaimus, spp.*; and/or Stunt nematode, *Tylenchorhynchus spp.*

[0028] In an aspect, the entity as described herein is a biosurfactant obtainable by a method comprising: obtaining a medium in which *Bacillus firmus* bacteria has been grown and precipitating the biosurfactant from the medium.

[0029] Biosurfactant is understood to mean microbially produced surface-active compounds. They are amphiphilic molecules with both hydrophilic and hydrophobic regions causing them to aggregate at the interfaces between fluids with different polarities such as water and hydrocarbons (Jennings E. R. *et al.*, 2000).

[0030] Multiple initial tests were conducted to determine the production of biosurfactant from *B. firmus*. The bacterium was grown under blood agar, where a positive production of biosurfactant is indicated by a transparent halo around the bacterial colony. Additional testing, *B. firmus* was cultured on nutrient broth at 30°C for 24 hours. Subsequently, the living cells were recovered by centrifugation at 5,181 xg for 15 minutes, and cells were washed twice with NaCl 0.85% (w/v) and later suspended in 5 ml of NaCl 0.85% (w/v). They were used to inoculate 45 ml of saline Davis minimal broth with an inoculum ratio of 1% (v/v). The composition was K₂HPO₄ 5.23 g/l, KH₂PO₄ 1.91 g/l, MgSO₄ 0.09 g/l, (NH₄)₂SO₄ 1 g/l, as well as 1 ml/l of trace elements solution (CoCl₃ 20 mg/l, H₃BO₃ 30 mg/l, ZnSO₄ 10 mg/l, Cu₂SO₄ 1 mg/l, Na₂MoO₄ 3 mg/l, FeSO₄ 10 mg/l and MgSO₄ 2.6 mg/l). Cultures were incubated at 30 °C ± 2 at 150 rpm for 3 days. Again *B. firmus* living cells were separated from the supernatant by centrifugation (20 minutes at 4000 xg). The supernatant was filtered through a Millipore filter 0.45-0.22 µm to obtain the final bacterial biosurfactant product. This final product was autoclaved twice for 30 minutes at 120°C at 1 kg/cm³ pressure to kill all the bacterium's living cells and inactivate its enzymes. emulsifying activity of the cell-free supernatant was evaluated by mixing 0.5 ml with 0.5 ml of kerosene and 4 ml of distilled water to a disposable culture tube (borosilicate glass 16x150 mm). The negative control consisted of distilled water and kerosene, and the positive

control consisted of distilled water, kerosene, and Triton X-100 (100 mg/ml). Each tube was agitated in a vortex for 1 min and was left to stand for 24 hours. The height of the emulsification ring was then measured in millimeters and compared to that of the chemical emulsifier. If there was positive production of surfactant, the kerosene emulsified it and produced foam. The third test consisted of an oil drop collapse in which one drop of the supernatant was placed on parafilm paper and a drop of oil was placed on top of it. If the drop of oil increased its diameter compared to the media control, the bacteria was considered to have produced a biosurfactant. Finally, the biosurfactant product was again tested under *in vitro* conditions in 96 well-plate against second stage juveniles of *R. reniformis*. Volumes of 100 μ L of the treatments were transferred to each well, which contained approximately 16 juveniles of *R. reniformis*. Treatments for this trial were *i*) water control, *ii*) BPM control, *iii*) biosurfactant 100%, and *iv*) *B. firmus* 15 \times 10⁷ cfu/ml. Each treatment was replicated 8 times, and the entire trial was repeated twice. The number of juveniles paralyzed or dead was recorded 30 minutes after inoculation. Data were analyzed on SAS 9.1 (SAS Institute Inc.).

[0031] In an aspect, the medium in which *Bacillus firmus* bacteria has been grown is obtained by growing *B. firmus* bacteria aerobically in medium. There are numerous mediums in which the bacteria can grow on, for example, a minimal salt medium supplemented with yeast extract and glucose. In an aspect, for biosurfactant isolation, bacterial cells are removed from the surfactant-containing medium by centrifugation, and the biosurfactant is precipitated from the supernatant by adding an acid. The acid precipitates are recovered by centrifugation and are extracted with a solvent. After precipitation with an acid, the crude fraction dissolved in the solvent is evaporated, and the final purified biosurfactant product is diluted in distilled water at specific concentrations.

[0032] In an aspect, the entity as described herein is an isolated protease. Protease is understood to mean any polypeptides or complex of polypeptides or fragments of polypeptides having protease activity. Production of protease enzymes were evaluated by use of milk agar assay. A clear halo following 24 hr period of incubation is recorded as a positive indication for the production of proteases.

[0033] In an aspect, the entity as described herein is an isolated amylase. Amylase is understood to mean any polypeptides or complex of polypeptides or fragments of polypeptides having amylase activity. Production of amylase enzymes were evaluated by use of starch agar assay. A clear halo following 24 hr period of incubation is recorded as a positive indication for the production of amylases.

[0034] In an aspect, the entity as described herein is an isolated lipase. Lipase is understood to mean any polypeptides or complex of polypeptides or fragments of polypeptides having lipase activity. Production of lipase enzymes were evaluated by use of trybutirin agar assay. A clear halo following 24 hr period of incubation is recorded as a positive indication for the production of lipases.

[0035] In an aspect, the entity as described herein is an isolated cellulase. Cellulase is understood to mean any polypeptides or complex of polypeptides or fragments of polypeptides having cellulase activity. Production of cellulase enzymes were evaluated by use of carboximetil cellulose (CMC) agar agar assay. A clear halo following 24 hr period of incubation is recorded as a positive indication for the production of cellulases.

[0036] In an aspect, the disclosure provides for a novel composition having anti-nematodal activity comprising at least one entity derived from *Bacillus firmus* bacteria selected from the group consisting of an isolated biosurfactant, an isolated protease, an isolated amylase, an isolated lipase, and an isolated cellulase.

[0037] In an aspect, the strain of *Bacillus firmus* is GB-126. In an aspect, the composition described herein has anti-nematodal activity against *Rotlyenchulus reniformis*.

[0038] In an aspect, the composition described herein comprises an acid precipitate from a culture medium in which the *Bacillus firmus* bacteria was grown. In an aspect, the acid precipitate described herein is obtainable by a method comprising: adding an acid to a medium in which a *Bacillus firmus* bacteria has been grown to generate an acid precipitate, and (b) isolating the acid precipitate from the medium. In another aspect, the acid precipitate has anti-nemadodal activity.

[0039] In an aspect, the medium in which *Bacillus firmus* bacteria has been grown is obtained by growing *B. firmus* bacteria aerobically in medium. There are numerous mediums in which the bacteria can grow on, for example, a minimal salt medium supplemented with yeast extract and glucose. In an aspect, for biosurfactant isolation, bacterial cells are removed from the surfactant-containing medium by centrifugation, and the biosurfactant is precipitated from the supernatant by adding an acid. The acid precipitates are recovered by centrifugation and are extracted with a solvent. After precipitation with an acid, the crude fraction dissolved in the solvent is evaporated, and the final purified biosurfactant product is diluted in distilled water at specific concentrations.

[0040] In an aspect, the composition as described herein comprises of an isolated biosurfactant. In an aspect, the isolated biosurfactant concentration in the composition as described herein is greater than 0.5 ppm. In an aspect, the isolated biosurfactant concentration

in the composition described herein is at least 1 ppm. In an aspect, the isolated biosurfactant concentration in the composition described herein is from 1 to 2 ppm. In an aspect the isolated biosurfactant concentration in the composition described herein is at least 2 ppm. In an aspect, the isolated biosurfactant concentration in the composition described herein is from 1 to 5 ppm. In an aspect, the isolated biosurfactant concentration in the composition described herein is from 2 to 5 ppm.

- [0041] In an aspect, the composition as described herein is an isolated protease.
- [0042] In an aspect, the composition as described herein is an isolated amylase.
- [0043] In an aspect, the composition as described herein is an isolated lipase.
- [0044] In an aspect, the composition as described herein is an isolated cellulase.
- [0045] In an aspect, the disclosure provides for an extract of a *Bacillus firmus* bacteria having the following characteristics: anti-nematodal activity and presence of a biosurfactant.
- [0046] In an aspect, the disclosure provides for an extract of a *Bacillus firmus* bacteria having the following characteristics: anti-nematodal activity and activity selected from the group consisting of protease activity, amylase activity, lipase activity, and cellulase activity.
- [0047] Extract shall refer to any fraction extracted from reference material.
- [0048] The disclosure also provides for a method of controlling plant-parasitic nematode by applying an entity or composition as described herein to a plant, the seed material (e.g. grains, seeds or vegetative propagation organs such as tubers or shoot parts with buds) or the area on which the plants grow (e.g. the area under cultivation).
- [0049] In an aspect, the disclosure provides for a method of protecting a plant by applying an entity or composition as described herein to a plant, the seed material, or the area on which the plant grows.
- [0050] All plants and plant parts can be treated in accordance with the invention. Plants are to be understood as meaning in the present context all plants and plant populations such as desired and undesired wild plants or crop plants (inclusive of naturally occurring crop plants). Crop plants can be plants which can be obtained by conventional plant breeding and optimisation methods or by biotechnological and recombinant methods or by combinations of these methods, inclusive of the transgenic plants and inclusive of the plant varieties protectable or not protectable by plant breeders' rights. Plant parts are to be understood as meaning all aerial and subterranean plant parts and organs of the plants such as shoot, leaf, flower and root, examples which may be mentioned being leaves, needles, stalks, trunks,

flowers, fruiting bodies, fruits, seeds, roots, tubers and rhizomes. The plant parts also include vegetative and generative propagation material, for example cuttings, tubers, rhizomes, seedlings and seeds.

[0051] In an aspect, the plant selected from the group consisting of agronomic crops, corn (field, seed & popcorn), sorghum, wheat, barley, rye, oats, rice, forage grasses, soybeans, canola, peanuts, cotton, alfalfa, fruiting vegetables (peppers & tomatoes), melons, squash, pumpkins, cucumbers and potatoes.

[0052] In an aspect, the plant-parasitic nematode is Reniform nematode, *Rotylenchulus* spp.; Dagger nematode, *Xiphinema* spp.; Lance nematode, *Hoplolaimus* spp.; Pin nematode, *Paratylenchus* spp.; Ring nematode, *Criconemoides* spp.; Rootknot nematode, *Meloidogyne* spp.; Sheath nematode, *Hemicyclophora* spp.; Spiral nematode, *Helicotylenchus* spp.; Stubbyroot nematode, *Trichodorus* spp.; Cyst nematode, *Heterodera* spp.; Sting nematode, *Belonolaimus*, spp.; and/or Stunt nematode, *Tylenchorhynchus* spp.

[0053] In an aspect, the plant is cotton. In an aspect the plant is agronomic crops. In an aspect, the plant is corn (field, seed & popcorn). In an aspect, the plant is sorghum. In an aspect, the plant is wheat. In an aspect, the plant is barley. In an aspect, the plant is rye. In an aspect, the plant is oat. In an aspect, the plant is rice. In an aspect, the plant is forage grasses. In an aspect, the plant is soybeans. In an aspect, the plant is canola. In an aspect, the plant is peanuts. In an aspect, the plant is cotton. In an aspect, the plant is alfalfa. In an aspect, the plant is fruiting vegetables (peppers & tomatoes). In an aspect, the plant is melon. In an aspect, the plant is squash. In an aspect, the plant is pumpkin. In an aspect, the plant is cucumber. In an aspect, the plant is potato.

[0054] The following examples serve to illustrate certain aspects of the disclosure and are not intended to limit the disclosure.

[0055]

EXAMPLES

[0056] *In vitro* tests against *R. reniformis*.

An initial test was conducted to evaluate the effect of the metabolites produced by *B. firmus* against second stage juveniles and egg hatching of *R. reniformis*. Eggs of *R. reniformis* were extracted from cotton roots by shaking the root system in a 1% NaOCl solution for four minutes at 120 rpm. The suspension of eggs was collected and rinsed with water through a 25 µm sieve (8). Eggs were rinsed with streptomycin sulfate (300 mg/L) and

chlortetracycline (12.5 mg/L) for bacterial disinfection, then with metalaxyl (25mL/L) and iprodione (20mL/L) for fungal disinfection, then finally with distilled water. For the second stage juvenile trial, the eggs were placed in a modified Baerman dish on a slide warming tray at 27°C. Second stage juveniles were hatched after three days. To obtain the bacterial metabolite, *Bacillus firmus* GB-126 was grown in 50 mL of Tryptic Soy Broth (TSB) (BACTO™) for four days and then placed in 50 mL plastic tubes and centrifuged for 20 minutes at 4000x g. The supernatant was collected and filtered through a Millipore filter 0.45-0.22 µm to obtain the final bacterial metabolite product (CFE). *In vitro* trials were conducted on 96-well plates, where volumes of 100µL of the treatments were transferred to each well, which contained approximately 16 juveniles or 20 eggs of *R. reniformis*. Treatments were *i*) water control, *ii*) TSB media control, *iii*) metabolite 100%, and *iv*) metabolite 50%. Each treatment had 6 replications, and the entire trial was repeated twice. The number of eggs hatched releasing juveniles was recorded at 0, 24, 48, and 72 hours after inoculation. For the second stage juveniles, number of moving and paralyzed nematodes was recorded at 0, 1, 2, 4, 6, and 12 hours after inoculation. Data were analyzed on SAS 9.1 (SAS Institute Inc.) using the GLIMMIX procedure where the distributional assumption was evaluated with the student panel graphs. Dunnett's option was used to assess the differences with the water and TSB controls.

Table 1 demonstrates the statistical significance of Fig. 1.

Table 1

Treatments			Statistical Comparison; P<0.05 (Hr after Combining)		
			24	48	72
Water Control	vs	CFE100%	0.11	0.03	0.03
Water Control	vs	CFE 50%	0.11	0.04	0.03
Media Ctl	vs	CFE 100%	0.24	≤0.001	≤0.001
Media Ctl	vs	CFE 50%	0.28	≤0.001	≤0.001

Table 2 demonstrates the statistical significant of Fig. 2.

Table 2

Statistical Comparison; P<0.05 (Hr after Combining)					
Treatments		1	2	4	6
					12

Control	vs	CFE 100%	≤0.001	≤0.001	≤0.001	≤0.001	≤0.001
Control	vs	CFE 50%	≤0.005	≤0.009	≤0.001	≤0.001	≤0.001
Media Ctl	vs	CFE 100%	0.04	0.003	≤0.001	≤0.001	0.02
Media Ctl	vs	CFE 50%	0.12	0.024	≤0.001	≤0.001	0.03

[0057] Enzyme characterization of *B. firmus*.

Different enzymatic properties of *B. firmus* GB-126 were evaluated to test their capacity to degrade different media. Production of enzymes was evaluated as positive when a transparent halo was formed around the bacterium culture. *Bacillus firmus* GB-126 was grown for 24 hours on milk agar to test the production of proteases, starch agar for the production of amylases, carboximethyl cellulose (CMC) agar for the production of cellulases, chitinase agar for the production of chitinases, and trybutirin agar for the production of lipases. The CMC agar and chitinase agar required the application of 5 ml of congo red to stain the media and a transparent halo after 24 hours of culturing the bacteria. In the case of the starch media, culture was stained with lugol.

[0058] Determination of production of biosurfactant by *B. firmus*.

Three initial tests were conducted to determine the production of biosurfactant from *B. firmus*. In the first test, the bacterium was grown under blood agar, where a positive production of biosurfactant is indicated by a transparent halo around the bacterial colony. For the second and third tests, *B. firmus* was cultured on nutrient broth at 30°C for 24 hours. Subsequently, the living cells were recovered by centrifugation at 5,181 xg for 15 minutes, and cells were washed twice with NaCl 0.85% (w/v) and later suspended in 5 ml of NaCl 0.85% (w/v). They were used to inoculate 45 ml of saline Davis minimal broth with an inoculum ratio of 1% (v/v). The composition was K₂HPO₄ 5.23 g/l, KH₂PO₄ 1.91 g/l, MgSO₄ 0.09 g/l, (NH₄)₂SO₄ 1 g/l, as well as 1 ml/l of trace elements solution (CoCl₃ 20 mg/l, H₃BO₃ 30 mg/l, ZnSO₄ 10 mg/l, Cu₂SO₄ 1 mg/l, Na₂MoO₄ 3 mg/l, FeSO₄ 10 mg/l and MgSO₄ 2.6 mg/l). Cultures were incubated at 30 °C ± 2 at 150 rpm for 3 days. Again *B. firmus* living cells were separated from the supernatant by centrifugation (20 minutes at 4000x g). The supernatant was filtered through a Millipore filter 0.45-0.22 µm to obtain the final bacterial biosurfactant product. This final product was autoclaved twice for 30 minutes at 120°C at 1 kg/cm³ pressure to kill all the bacterium's living cells and inactivate its enzymes.

[0059] In the second test, emulsifying activity of the cell-free supernatant was evaluated by mixing 0.5 ml with 0.5 ml of kerosene and 4 ml of distilled water to a disposable culture tube (borosilicate glass 16x150 mm). The negative control consisted of distilled water and kerosene, and the positive control consisted of distilled water, kerosene, and Triton X-100 (100 mg/ml). Each tube was agitated in a vortex for 1 min and was left to stand for 24 hours. The height of the emulsification ring was then measured in millimeters and compared to that of the chemical emulsifier. If there was positive production of surfactant, the kerosene emulsified it and produced foam. The third test consisted of an oil drop collapse in which one drop of the supernatant was placed on parafilm paper and a drop of oil was placed on top of it. If the drop of oil increased its diameter compared to the media control, the bacteria was considered to have produced a biosurfactant.

Finally, the biosurfactant product was again tested under *in vitro* conditions in 96 well-plate against second stage juveniles of *R. reniformis*. Volumes of 100 μ L of the treatments were transferred to each well, which contained approximately 16 juveniles of *R. reniformis*. Treatments for this trial were *i*) water control, *ii*) BPM control, *iii*) biosurfactant 100%, and *iv*) *B. firmus* 15 \times 10⁷ cfu/ml. Each treatment was replicated 8 times, and the entire trial was repeated twice. The number of juveniles paralyzed or dead was recorded 30 minutes after inoculation. Data were analyzed on SAS 9.1 (SAS Institute Inc.) as described previously.

[0060] Purification and production of biosurfactant and in vitro evaluations at different concentrations against *R. reniformis*.

For biosurfactant production, *B. firmus* GB-126 was grown aerobically on minimal salt medium containing (per liter) KH₂PO₄ (2.0 g), K₂HPO₄ (5.0 g), (NH₄)₂SO₄ (3.0 g), NaNO₃ (2.0 g), NaCl (0.1 g), MgSO₄ H₂O (0.2 g), FeSO₄ 7H₂O (0.01 g), CaCl₂ (0.01 g), and 1 ml of a trace element solution. The stock solution of trace elements contained (per liter) ZnSO₄ 7H₂O (2.32 g), MnSO₄ 4H₂O (1.78 g), H₃BO₃ (0.56 g), CuSO₄ 5H₂O (1 g), Na₂MoO₄ 7H₂O (0.39 g), CoCl₂ 6H₂O (0.42 g), EDTA (1 g), NiCl₂ 6H₂O (0.004g), and KI (0.66 g). The medium was supplemented with 0.05% yeast extract (Vater, J. *et al.*) Glucose was added as a carbon source at a concentration of 2% (wt/vol). The medium pH was 7.1 to 7.2. The organism was grown at 37°C for 48 h in 2-liter Erlenmeyer flasks containing 800 ml of medium and shaken at 200 rpm in a shaker incubator.

[0061] For biosurfactant isolation, bacterial cells were removed from the surfactant-containing medium by centrifugation (13000x g for 15 min at 4°C). The biosurfactant was

precipitated from the supernatant by adding 6 N HCl to obtain a final pH of 2.0. The acid precipitates were recovered by centrifugation (13000x g for 15 min at 4°C) and were extracted with dichloromethane or methanol (lipopeptide fraction). When methanol was used as the solvent, the extract was neutralized immediately to avoid formation of methyl esters. After precipitation with HCl, the crude fraction dissolved in methanol or dichloromethane was evaporated in a rotary evaporator (Model Buchi R) under a vacuum pump (Model Gem 8890) (Vater, J. *et al.*). The final purified biosurfactant product was diluted in distilled water at concentrations of 2 ppm, 1 ppm, 0.5 ppm, 0.2 ppm, 0.1 ppm, and 0.02 ppm. These concentrations were evaluated, compared to distilled water control under *in vitro* conditions on second stage juveniles as described above, with 8 replications per treatment, and repeated twice.

[0062] Greenhouse trials.

To evaluate if *B. firmus* GB-126 induces systemic resistance the following treatments were tested in a split root system. Treatments consisted of *i*) water control without nematodes, *ii*) water control with nematodes, *iii*) *B. firmus* 1x10⁷ cfu/mL, *iv*) *B. firmus* 1x10⁶ cfu/mL, or *v*) *Serratia marcescens* 1x10⁷ cfu/mL. Stoneville 5458 B2RF cotton seeds were germinated in potting mixing soil under greenhouse conditions. Emerging root radicals approximately 2.5 cm in length were split with a razer blade. At 5 days after planting (DAP), soil was removed from the roots and divided into two equal halves. Plants were planted in 960 cm³ pots with each root half in a different cup. At 7 days after splitting the roots, a suspension of 50 mL of treatments was applied on the left side of the root. Five days later, the right side of the root was inoculated with 500 second stage juveniles of *R. reniformis*. The trial was harvested 45 DAP, and plant height, root fresh weight, and number of females and eggs per gram of root were measured. Each treatment had 6 replications and the entire trial was repeated twice.

[0063] To evaluate the response of *R. reniformis* to cotton seeds treated with *B. firmus* GB-126, a trial in autoclaved soil was conducted in the Plant Science Research Center (PSRC) of Auburn University. Cotton seeds from cultivar Stoneville 5458 B2RF were treated with *B. firmus* GB126 by the manufacturer in a liquid seed dresser Hege1 1 (Hege Maschinen GmbH, Germany). Presence of the bacterium in the seed was confirmed by culturing the treated seed on Tryptic Soy Agar (TSA) adjusted to pH 8.0 and recording the growth after 16 hours. The soil was a Decatur silty clay loam (sand-silt-clay: 17.5-51.3-31.2%; nitrogen: 0.16%; organic matter: 2.2; pH 7.24) from the Tennessee Valley Research and Extension Center (TVREC) near Belle Mina, AL. The soil was autoclaved using two 90-

minute cycles at 130°C at 15 psi with a 24 hour cool down between cycles. Seed treatments were as follows: *i*) untreated seed with nematodes; *ii*) imidacloprid (500 g ai/100kg) a standard insecticide; *iii*) *B. firmus* (7×10^4 cfu/seed) plus imidacloprid (500 g ai/100kg); *iv*) *B. firmus* (7×10^5 cfu/seed) plus imidacloprid (500 g ai/100kg); and *v*) *B. firmus* (7×10^6 cfu/seed) plus imidacloprid (500 g ai/100kg). The standard insecticide seed treatment imidacloprid was included because this insecticide is commonly applied as a seed treatment and was tested to determine if it has any effect on *B. firmus* or *R. reniformis* life stages.

[0064] *Rotylenchulus reniformis* vermiform life stages were extracted from the soil by modified gravity screening and sucrose centrifugation-flotation. Eggs were extracted from cotton roots by shaking the root system in a 1% NaOCl solution for four minutes at 120 rpm. The nematode suspension was rinsed with water and collected on a 25 µm sieve. Females in roots were stained with acid fuchsin to facilitate enumeration of the females invading the root. Vermiform life stages and eggs were counted under an inverted TS 100 Nikon microscope at 40x magnification. Females embedded in the root systems were quantified at 5x magnification utilizing the Nikon SMZ800 compound microscope. Variables measured were plant height, shoot and root weight, females and eggs per gram of root, and the number of vermiform life stages in 500 cm³ of soil. Greenhouse average temperature where plants were grown was 29°C. Soil moisture was maintained between 40-60% of the maximum water holding capacity. Data were analyzed in SAS 9.1 (SAS Institute Inc.). The distributional assumption was evaluated with the student panel graphs of the GLIMMIX procedure. Dunnett's option was used to assess the differences with the untreated control.

[0065] *In Vitro and Greenhouse Results*

In the first *in vitro* trial, *R. reniformis* egg hatch was reduced at 48 and 72 hours, when eggs were exposed to *B. firmus* metabolites at 100% and 50%, when compared to the water and media control ($P \leq 0.01$) (Fig. 1). Furthermore, paralysis of second stage juveniles of *R. reniformis* observed within one hour of inoculation in 100% and 50% metabolite through 12 hours when all the second stage juveniles were paralyzed ($P \leq 0.01$) (Fig. 2). No differences were observed between the water and the media controls in these trials ($P \leq 0.99$).

[0066] Biosurfactant production was confirmed by the emulsification of kerosene, oil drop collapse, and halo formation in blood agar (Fig. 3). Biosurfactant and living cells of *B. firmus* at a concentration of 15×10^7 cfu/mL paralyzed all the second stage juveniles within 30 minutes after inoculation when compared to the BP media and water controls

($P \leq 0.01$) (Fig. 3). There were no differences between the two controls ($P \leq 0.99$). Finally, in the last *in vitro* trial where the pure *B. firmus* biosurfactant was evaluated at different concentrations, the biosurfactant at 2 ppm and 1 ppm paralyzed 100% and 45.9%, of the second stage juveniles of *R. reniformis*, respectively, within 30 minutes (Fig. 4).

[0067] These two concentrations produced an increase in paralysis of the second stage juveniles compared to the water control ($P \leq 0.001$). Biosurfactant concentrations of 0.5 ppm, 0.2 ppm, 0.1 ppm, and 0.02 ppm did not paralyze second stage juveniles and were not different from the water control ($P \leq 0.932$). When second stage juveniles from the water control and 2 ppm treatments were observed under SEM, no mechanical damage to the cuticle was observed (Fig. 5A, B). The enzymatic profile of *Bacillus firmus* GB-126 indicated a high enzymatic activity for proteases, amylases, and cellulases forming a transparent halo in milk agar, starch agar, and CMC agar, respectively, within 24 hours. In contrast, no production of chitinases was observed under chitinase agar (Fig. 7).

[0068] *Bacillus firmus* GB-126 inhibits the hatch of *R. reniformis* eggs and paralyzes second stage juveniles under *in vitro* conditions using secondary metabolites from this bacterium and also living cells at a concentration of 15×10^7 cfu/ml. In greenhouse trials, *B. firmus* GB-126 applied as a seed treatment at a rate of 7×10^6 cfu/seed reduced number of *R. reniformis* females in the root and juveniles in soil within the first 30 days of planting. The effect of the insecticide imidacloprid, which is used as a seed treatment and formulated with this bacterium, did not show any nematicidal activity.

[0069] The results demonstrate that the mode of action of *B. firmus* GB-126 against nematodes is a secondary metabolite toxic to nematodes. This secondary metabolite is a biosurfactant which is responsible for the paralysis of *R. reniformis* juveniles and inhibition of egg hatch under *in vitro* conditions. *Bacillus firmus* GB-126 biosurfactant needs a minimum concentration of 1 ppm to paralyze second stage juveniles within 30 minutes.

[0070] The presence of enzymes (amylases, cellulases, and proteases) suggests the possibility that *B. firmus* GB-126 can have other modes of action against *R. reniformis* and other nematode species during different stages of the life cycle. The production proteases by this bacterium can affect on egg hatching and cause nematode paralysis.

[0071] The induction of systemic resistance trial indicated cotton plants treated with *B. firmus* GB-126 (1×10^6 cfu/ml) was taller than control with nematodes ($P \leq 0.05$) and *S. marcevens* (1×10^9 cfu/ml) treatment ($P \leq 0.01$). There were no differences in left or right root

fresh weights or the number of *R. reniformis* females and eggs among the treatments ($P \leq 0.99$) (Fig. 6). In contrast, *B. firmus* GB-126 at a rate of 7×10^6 cfu/seed reduced the number of females per gram of root ($P \leq 0.001$) and juveniles per 500 cm^3 of soil 30 days after planting ($P \leq 0.01$) (Fig. 8). The insecticide imidacloprid did not have any effect on cotton plant growth or *R. reniformis* life stages. Induction of systemic resistance of *B. firmus* GB-126 in cotton plants was not observed at the concentrations evaluated.

[0072] In summary, the biocontrol activity of *B. firmus* GB-126 observed in previous trials under greenhouse and field conditions where eggs and juvenile stages were reduced can be explained because the bacterium is producing a biosurfactant that is toxic to the plant-parasitic nematode. No ISR was observed at rates tested. However, *B. firmus* GB-126 possibly has other mechanisms of action against *R. reniformis* due to the presence of proteases that can be deleterious to the nematode.

1. Tian, B., Yang, J., Zhang, K. 2007. *Bacteria used in the biological control of plant-parasitic nematodes: populations, mechanisms of action, and future prospects.* FEMS Microbiol Ecol 61: 197-213.
2. Aatlen, P.M., Vitour, D., Blanvillain, D., Gowen, S.R., Sutra, L. 1998. *Effect of the rhizosphere fluorescent Pseudomonas strains on plant-parasitic nematodes Radopholus similis and Meloidogyne spp.* Letter in Applied Microbiology 27:357-361.
3. Kerry, B.R. 2000. *Rhizosphere interactions and the exploitation of microbial agents for the biological control of plant parasitic nematodes.* Annual Review of Phytopathology 38:423-441.
4. Kerry, B.R. 1987. *Principles and practice of nematode control in crops*, p 233-263. In Brown RH, Kerry BR (eds.), Biological control. Academic Press. London.
5. Siddiqui, Z.A., Mahmood, I. 1999. *Role of bacteria in the management of plant parasitic nematodes: a review.* Bioresource Technol 69:167-179;
6. Stirling, G.R. 1991. *Biological control of plant-parasitic nematodes.* CAB International. Wallingford.
7. Van Loon, L.C., Baker, P.A., Pieterse, C.M. 1998. *Systemic Resistance induced by rhizosphere bacteria.* Annual Review of Phytopathology 36:453-483.
8. Sikora, R.A. 1992. *Management of the antagonistic potential in agriculture ecosystems for the biological control of plant parasitic nematodes.* Annual Review of Phytopathology 30: 245-270.
9. Meyer, S.L.F. 2003. United States Department of Agriculture – Agricultural Research Service research programs on microbes for management of plant-parasitic nematodes. Pest Management Science 59:665-670.
10. Blachinsky, D., Antonov, J., Bercovitz, A., El-ad, B., Feldman, K., Husid, A., Lazare, M., Shamai, I., Keren-Zur, M. 2007. *BioNemWP: a unique tool for nematode control.* <http://agrogreen.co.il/PDF/Bionem%20IOBC%202006.pdf>.
11. Keren-Zur, M., Antonov, J., Bercovitz, A., Feldman, K., Husid, A., Kenan, G., Markov, N. Rebhun, M. 2000. *Bacillus firmus formulations for the safe control of root-knot nematodes.* Proceedings of Brighton Crop Protection Conference: Pest and Diseases 1:47-52
12. Giannakou, O.I., Karpouzas, D.G., Prophetou-Athanasiadou, D.A. 2004. *A novel nonchemical nematicide for the control of root-knot nematodes.* Applied Soil Ecology 26:69- 79

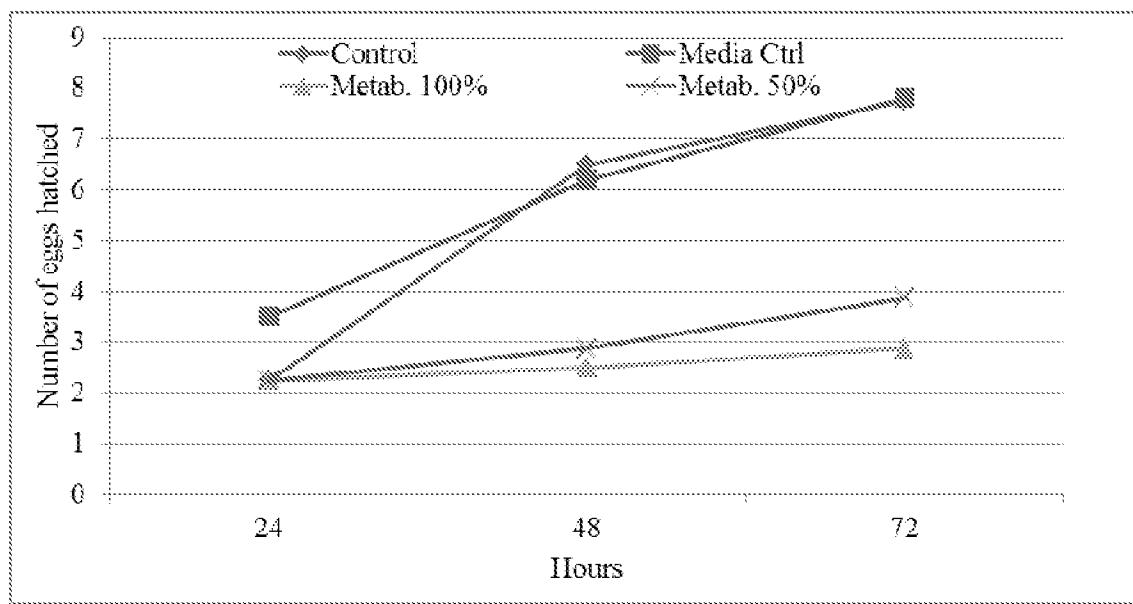
13. Giannakou, O.I., Anastasiadis, A.I., Gowen, S.R., Prophetou-Athanasiadou, D.A. 2007. *Effects of a non-chemical nematicide combined with soil solarization for the control of root-knot nematodes*. Crop Protection 26:1644-1654
14. Terefe, M., Tefera, T., Sahuja, P.K. 2009. *Effect of a formulation of *Bacillus firmus* on root-knot nematode *Meloidogyne incognita* infestation and the growth of tomato plants in the greenhouse and nursery*. Journal of Invertebrate Pathology 100:94-99
15. Wick, R.L. 2006. *Seeking alternatives to Nemacur: Evaluation of natural suppression and testing new products*. Summary report. Available: <http://www.nerf.org/15.pdf>
16. Mendoza, A.R., Sikora, R. 2009. *Biological control of *Radopholus similis* in banana by combined application of the mutualistic endophyte *Fusarium oxysporum* strain 162, the egg pathogen *Paecilomyces lilacinus* strain 251 and the antagonistic bacteria *Bacillus firmus**. BioControl 54:263-273
17. Mendoza, A.R., Kiewnick, S., Sikora, R. 2008. *In vitro activity of *Bacillus firmus* against the burrowing nematode *Radopholus similis*, the root-knot nematode *Meloidogyne incognita*, and the stem nematode *Ditylenchus dipsaci**. Biocontrol Science Technology 18:377-389
18. Vater, J., Kablitz, B., Wilde, C., Franke, P., Mehta, N., Cameotra, S.S. 2002. Matrixassisted laser desorption ionization-time of flight mass spectrometry of lipopeptide biosurfactants in whole cells and culture filtrates of *Bacillus subtilis* C-1 isolated from petroleum sludge. Applied and Environmental Microbiology 68:6210-6219
19. Jennings, E. R. and Tanner, R. S. 2000. Biosurfactant-producing Bacteria Found in Contaminated and Uncontaminated Soils, *Proceedings of the 2000 Conference on Hazardous Waste Research*. 299-306

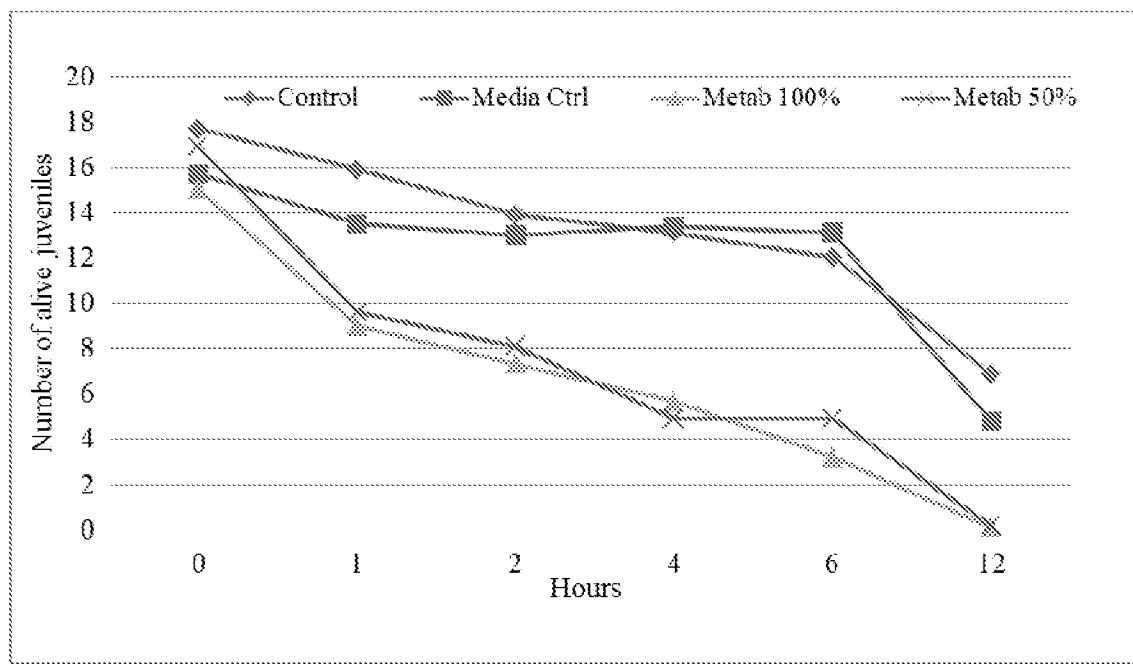
The Claims:

1. An entity having anti-nematodal activity derived from a bacterium having anti-nematodal activity, wherein the entity is at least one selected from the group consisting of an isolated biosurfactant, an isolated protease, an isolated amylase, an isolated lipase, and an isolated cellulase.
2. The entity according to claim 1, wherein the bacterium is of the species *Bacillus firmus*.
3. The entity according to claim 2, wherein the strain of *Bacillus firmus* is strain GB-126.
4. The entity according to any of claims 1-3, wherein the entity is at least one biosurfactant obtainable by a method comprising:
 - A. obtaining a medium in which the bacterium has been grown,
 - B. precipitating biosurfactant from the medium.
5. The entity according to any of claims 1-4, wherein the entity has anti-nematodal activity against *Rotlyenchulus reniformis*.
6. A composition having anti-nematodal activity comprising at least one entity according to claims 1-5.
7. The composition according to claim 6, wherein the bacterium is of the species *Bacillus firmus*.
8. The composition according to claim 7, wherein the strain of *Bacillus firmus* is strain GB-126.
9. The composition according to any of claims 6-8, wherein the composition has anti-nematodal activity against *Rotlyenchulus reniformis*.

10. The composition according to any of claims 6-9 comprising at least one isolated biosurfactant.
11. The composition according to claim 10, wherein the biosurfactant concentration is greater than 0.5 ppm.
12. The composition according to any of claims 10-11, wherein the biosurfactant concentration is at least 1 ppm.
13. The composition according to any of claims 10-12 wherein the biosurfactant concentration is from 1 to 2 ppm.
14. The composition according to any of claims 10-12, wherein the biosurfactant concentration is at least 2 ppm.
15. The composition according to any of claims 6-14 comprising at least one acid precipitate from a culture medium in which the bacterium was grown.
16. The composition according to claim 15, wherein the at least one acid precipitate is obtainable by a method comprising:
 - (a) adding an acid to a medium in which a bacterium has been grown to generate an acid precipitate, and
 - (b) isolating the acid precipitate from the medium.
17. The composition according to claim 16, wherein the medium is obtained by growing bacterium at 37°C for 48 h in a flask containing a minimal salt medium supplemented with yeast extract and glucose.
18. The composition according to any of claims 16-17, wherein the at least one acid precipitate is generated by adding a mineral acid to the medium to obtain a final pH of 2.0.
19. The composition according to claim 18, wherein the mineral acid is HCl.

20. The composition according to any of claims 16-19, wherein the at least one acid precipitate is isolated from the medium by centrifugation and extracting the acid precipitate with dichloromethane or methanol.
21. A method of controlling plant-parasitic nematode by applying an entity according to any of claims 1-5 and/or a composition according to any of claims 6-20 to a plant, seed material, and/or the area on which the plant grows.
22. The method according to claim 21, wherein the plant-parasitic nematode is *Rotylenchulus reniformis*.
23. The method according to any of claims 21-22, wherein the plant is cotton.
24. A method of protecting a plant by applying a entity according to any of claims 1-5 and/or a composition according to any of claims 6-20 to a plant, seed material, and/or the area on which the plant grows.
25. An extract of a *Bacillus firmus* culture having the following characteristics: anti-nematodal activity and/or presence of a biosurfactant.
26. An extract of a *Bacillus firmus* culture having the following characteristics: anti-nematodal activity and/or activity selected from the group consisting of protease activity, amylase activity, lipase activity, and cellulase activity.
27. The use of an entity according to any of claims 1-5 and/or a composition according to any of claims 6-20 to protect a plant.
28. The use of an entity according to any of claims 1-5 and/or a composition according to any of claims 6-20 to control plant-parasitic nematode.

**Fig 1**

**Fig 2**

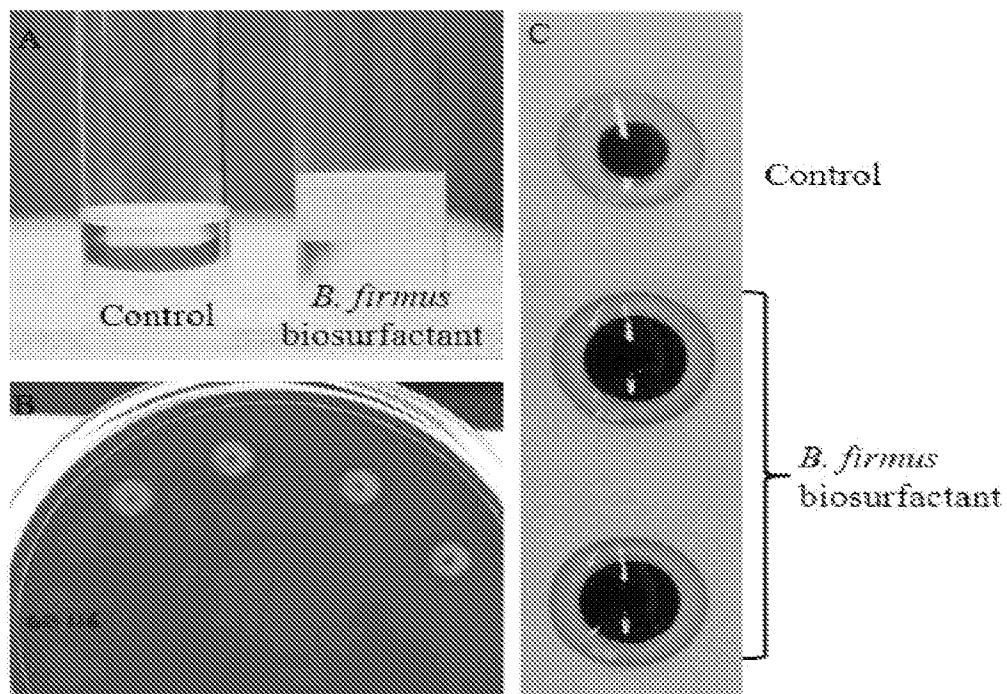


Fig. 3

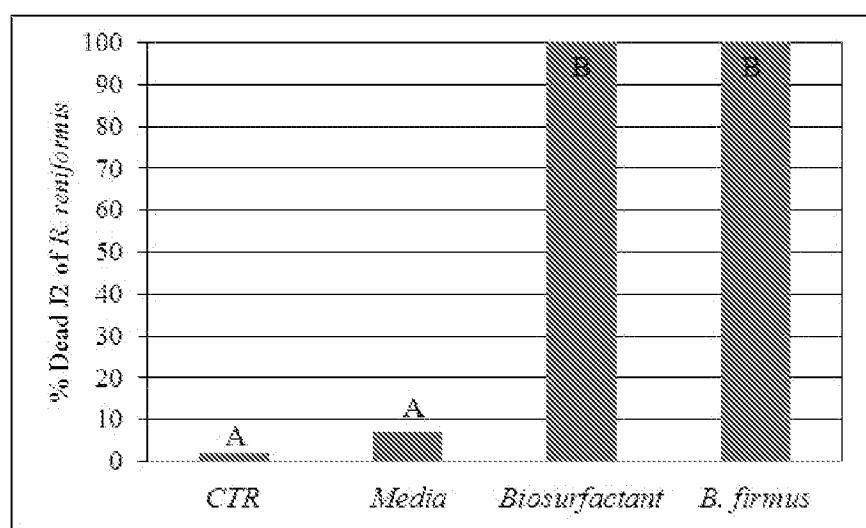


Fig. 4

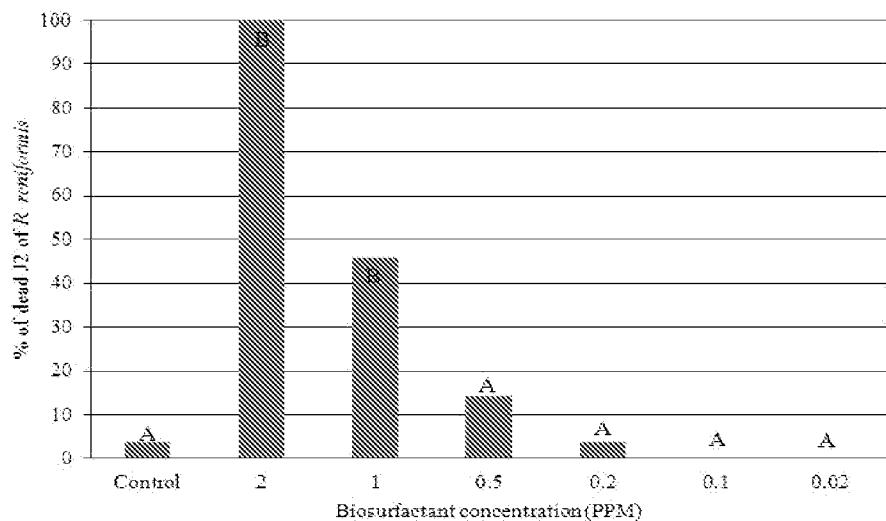


Fig. 5

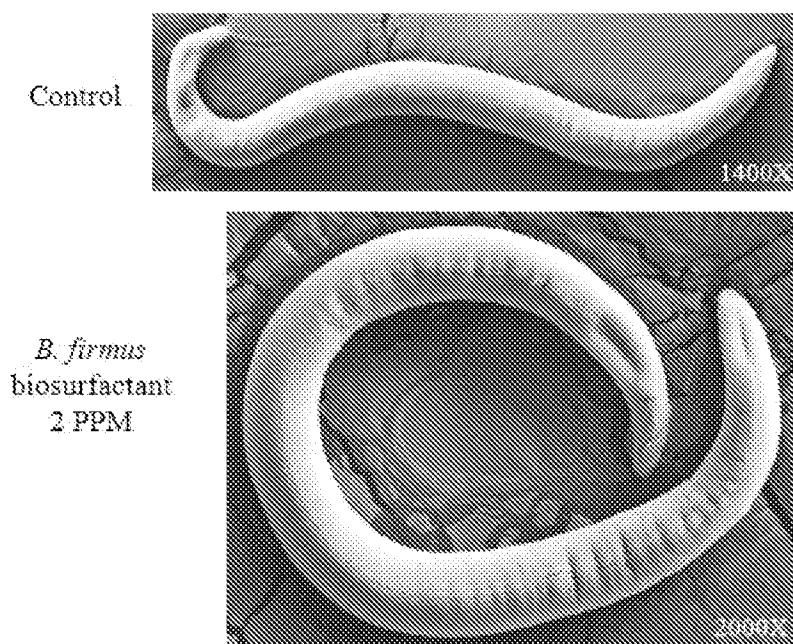


Fig. 6

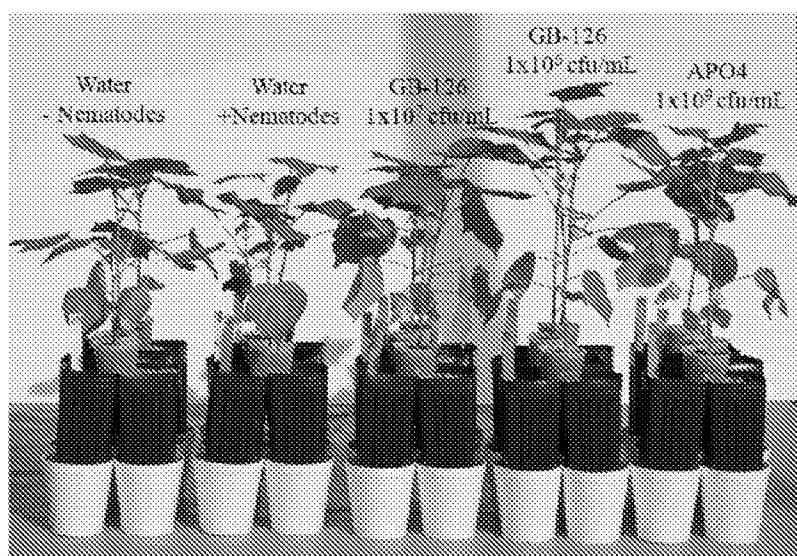


Fig. 7

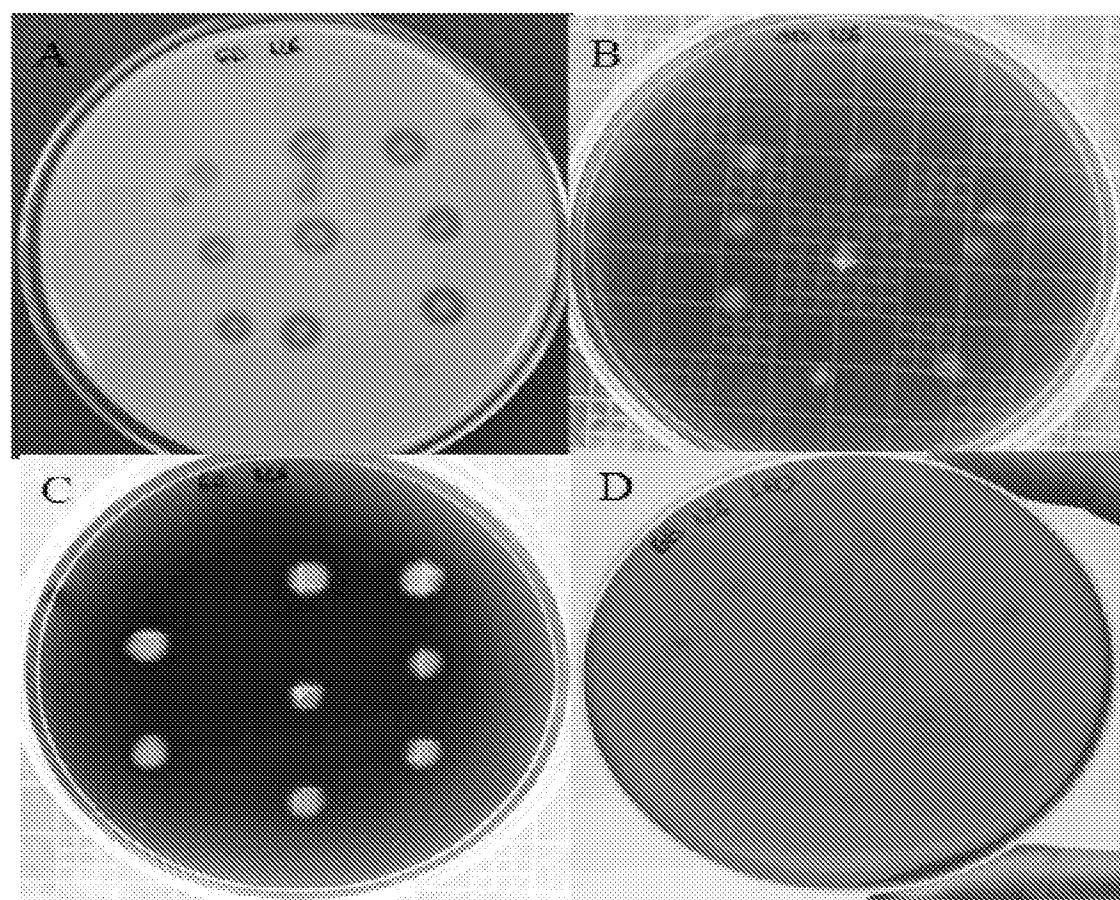


Fig. 8

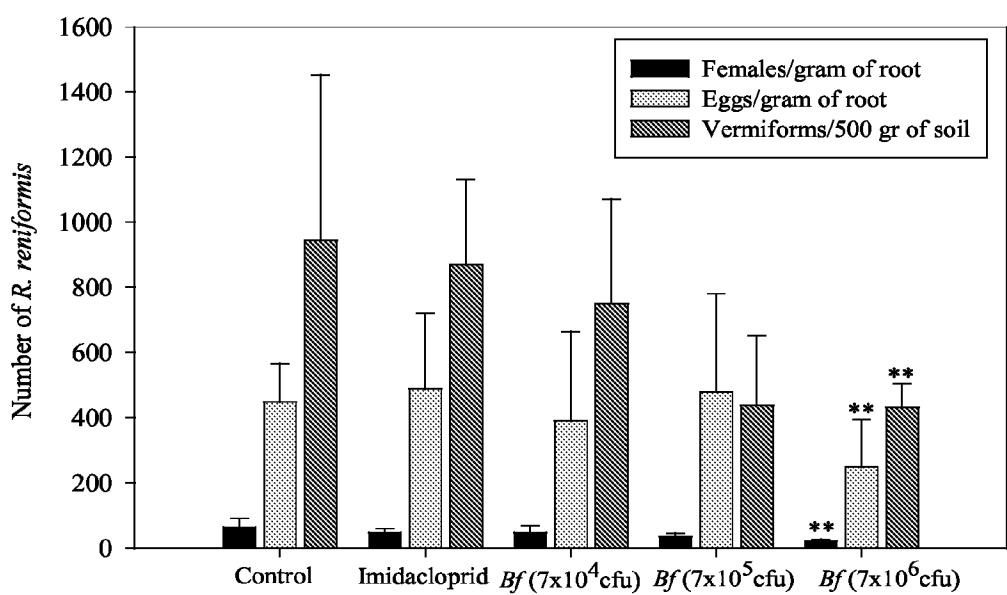


Fig. 9

INTERNATIONAL SEARCH REPORT

International application No
PCT/US2013/058866

A. CLASSIFICATION OF SUBJECT MATTER
INV. A01N63/02 A01P5/00
ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
A01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPO-Internal

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>Juan David Russi Castillo: "Biocontrol Studies of <i>Rotylenchulus reniformis</i> in Cotton Crops in Alabama", , 4 August 2012 (2012-08-04), pages 1-154, XP055084212, Retrieved from the Internet: URL:http://etd.auburn.edu/etd/bitstream/handle/10415/3318/PhD thesis w corrections July 20.pdf?sequence=2 [retrieved on 2013-10-16] Chapter III; page 76 - page 98 -----</p>	1-28



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance
"E" earlier application or patent but published on or after the international filing date
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
"O" document referring to an oral disclosure, use, exhibition or other means
"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
16 October 2013	24/10/2013
Name and mailing address of the ISA/ European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Fax: (+31-70) 340-3016	Authorized officer Molina de Alba, José