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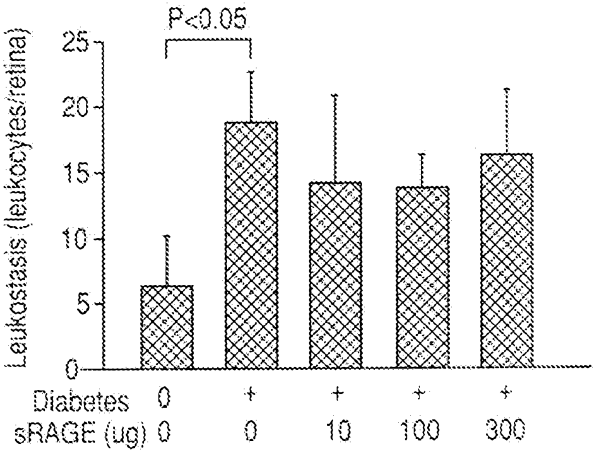

FIG. 1

(57) Abstract: The present invention provides novel therapeutics and methods of treatment for diseases associated with activation of the advanced glycation endproducts receptor (RAGE).

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RAGE FUSION PROTEINS

[0001] This application claims priority to United States provisional application for patent number 60/943, 994, filed June 14, 2007, the entire contents of which are specifically incorporated herein by reference.

FIELD OF THE INVENTION

[0002] The present invention relates generally to advanced glycation end products ("AGE") and more specifically to certain fusion proteins that comprise the receptor for advanced glycation end products ("RAGE"). Fusion proteins of the invention bind to AGE and other RAGE ligands (e.g., S100 and HMGB1) and compositions comprising fusion proteins of the invention may be used for the treatment of diseases.

BACKGROUND

[0003] Advanced glycation end products (AGE) are the result of nonenzymatic glycation and oxidation of proteins. They appear under stress related circumstances including in autoimmune connective tissue diseases, and may form in inflamed tissue due to the oxidation or the myeloperoxidase pathway. AGE have been implicated in a number of diabetes related complications. For example, the characteristic structural changes of diabetic nephropathy, thickened glomerular basement membrane and mesangial expansion, are accompanied by accumulation of AGE, leading to glomerulosclerosis and interstitial fibrosis. Prolonged infusion of nondiabetic rats with AGE has led to the development of similar morphological changes and significant proteinuria. AGE inhibitors such as aminoguanidine have been shown to prevent diabetic nephropathy in diabetic animal models and were recently shown to do the same in one clinical trial on diabetic patients. Also, AGE are a well validated therapeutic target for diabetic retinopathy. Extensive diabetic murine and rat studies have demonstrated the benefit of inhibiting AGE formation in treating this disease.

[0004] Atherosclerosis is significantly accelerated in diabetic patients and is associated with greater risk of cardiovascular and cerebrovascular mortality. Animal and human studies have suggested that AGE play a significant role in the formation and progression of atherosclerotic lesions. Increased AGE accumulation in the diabetic vascular tissues has been associated with changes in endothelial cell, macrophage, and smooth muscle cell function.

[0005] AGE interact with cell surface receptors on monocytes, macrophages, endothelial cells of the micro vasculature, smooth muscle cells, mesengial cells, and neurons. The receptor for advanced glycation end products (RAGE) is a member of the immunoglobulin superfamily of cell surface receptors. RAGE is made up of three extracellular immunoglobulin-like domains, a transmembrane domain, and a cytoplasmic domain that is involved in signaling. RAGE binds multiple ligands in addition to AGE including S100/calgranulins, amphoterin/HMGB 1 , and amyloid fibrils. RAGE acts through a signal cascade involving NF-κB. RAGE expression is up-regulated in the presence of RAGE ligands and is elevated in joints of subjects with rheumatoid arthritis (RA).

[0006] RAGE has a secreted isoform lacking a transmembrane domain called soluble RAGE (sRAGE). Administration of sRAGE has been shown to restore wound healing (Goova, et al. (2001) *Am. J. Pathol.* 159, 513-525) and suppress diabetic atherosclerosis (Park, et al. (1998) *Nat Med.* 4(9): 1025-31). Fusion proteins consisting of a RAGE ligand binding element and an immunoglobulin element are discussed in WO 2004/016229 A2 (Wyeth, Madison, NJ) and US Patent App. Publication 2006/0057679 Al (O'Keefe, T. et al.).

[0007] There exists a need for novel methods of treatment of AGE-mediated diseases, such as diseases that are associated with an elevated amount of AGE. This need and others are met by the present invention.

SUMMARY OF THE INVENTION

[0008] The present invention provides materials and methods for the treatment of diseases associated with an elevated amount of AGE. In one embodiment, the present invention provides a fusion protein comprising at least one polypeptide comprising: (a) a first amino acid sequence at least 95% identical to a mammalian receptor for advanced glycation end product (RAGE) ligand binding domain, the first amino acid sequence capable of binding a RAGE ligand; and (b) a second amino acid sequence at least 95% identical to a human heavy chain immunoglobulin IgG4 constant domain or a fragment thereof; wherein the first amino acid sequence comprises at least one mutation relative to a wild type RAGE ligand binding domain.

[0008a] In another aspect, the invention provides an isolated fusion protein comprising at least one polypeptide comprising:

(a) a first amino acid sequence comprising an amino acid sequence selected from the

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group consisting of amino acids 1-301, amino acids 24-301, amino acids 1-344, or amino acids 24-344 of SEQ ID NO:6; and

(b) a second amino acid sequence at least 95% identical to a human heavy chain immunoglobulin IgG4 constant domain or a fragment thereof.

[0008b] In a further aspect, the invention provides an isolated nucleic acid encoding a fusion protein comprising at least one polypeptide comprising:

(a) a first amino acid sequence comprising an amino acid sequence selected from the group consisting of amino acids 1-301, amino acids 24-301, amino acids 1-344, or amino acids 24-344 of SEQ ID NO:6; and

(b) a second amino acid sequence at least 95% identical to a human heavy chain immunoglobulin IgG4 constant domain or a fragment thereof.

[0008c] In one embodiment of the invention, a fusion protein of the invention may further comprise a linker sequence between the first amino acid sequence and the second amino acid sequence.

In some embodiments, the RAGE ligand binding domain may be from a mammalian RAGE, for example, a human RAGE. A suitable mammalian RAGE ligand binding domain may comprise amino acids 1-344 of SEQ ID NO: 6 or amino acids 24-344 of SEQ ID NO: 6. In one embodiment, a fusion protein of the invention may comprise an amino acid sequence selected from the group consisting of SEQ ID NO: 2, SEQ ID NO:4, SEQ ID NO:6, and SEQ ID NO:8. In one embodiment, an isolated fusion protein of the invention comprises SEQ ID NO:6 or SEQ ID NO:8. In another embodiment, an isolated fusion protein of the invention consists of SEQ ID NO:6 or SEQ ID NO:8. In some embodiments of the invention, fusion proteins of the invention may further comprise a linker between the RAGE amino acid sequence and the IgG4 amino acid sequence. The present invention also contemplates nucleic acid molecules (e.g., DNA or RNA molecules) encoding the fusion proteins of the invention as well as host cells expressing the nucleic acid molecules encoding the fusion proteins of the invention.

[0009] The present invention further provides for a pharmaceutical composition comprising a fusion protein of the invention and a pharmaceutically acceptable excipient or diluent.

[0010] The present invention provides methods of treating diseases mediated by AGE. Such diseases include any disease characterized by an increased amount of AGE in a subject, for example, a mammal such as a human. Methods of treating an AGE-mediated disease comprise administering to a subject having an AGE-mediated disease a therapeutically effective amount of a pharmaceutical composition comprising a fusion protein of the invention. Examples of diseases that can be treated by the methods of the invention include, but are not limited to, diabetic nephropathy, rheumatoid arthritis, and autoimmune diseases such as dermatitis, glomerulonephritis, multiple sclerosis, uveitis ophthalmia, autoimmune pulmonary inflammation, insulin dependent diabetes mellitus, autoimmune inflammatory eye, systemic lupus erythematosus, insulin resistance, rheumatoid arthritis, diabetic retinopathy, and scleroderma. Any fusion protein of the invention may be used in the practice of the methods of the invention. In one embodiment, methods of the invention may be practiced using a fusion protein comprising SEQ ID NO:6 or SEQ ID NO:8. In another embodiment, methods of the invention may be practiced using a fusion protein that consists of SEQ ID NO:6 or SEQ ID NO:8.

[0011] In another embodiment of the invention, the present invention provides methods of lowering the levels of ligand bound by RAGE in a mammal (e.g., a human) in need thereof. Such methods may comprise administering to the mammal a RAGE ligand-lowering amount of a fusion protein of the invention.

[0012] In other embodiments, the invention provides for a recombinant expression vector comprising the DNA sequences of the invention; a host cell transformed, transduced, or transfected with the vector; and a process for producing a fusion protein, which comprises culturing a host cell transformed, transduced or transfected with a nucleic acid encoding a fusion protein of the invention under conditions suitable to effect expression of the fusion protein.

[0013] The invention further provides compositions comprising the present fusion protein or fragments thereof. In some embodiments, the invention includes compositions comprising the present fusion protein or fragments thereof to which a radioisotope, chelator, toxin, fluorochrome, biotin, peptide epitopes such as his-tags, myc-tags, or sugars are directly or indirectly attached. Other embodiments of the invention include the present fusion protein fused with another protein for the purposes of altering the biological half-life or function and glycosylation variants of the fusion protein.

[0014] These and other aspects of the present invention will become evident upon reference to the following detailed description.

BRIEF DESCRIPTION OF THE DRAWINGS

[0015] Figure 1 is a bar graph showing the effects of an exemplary RAGE-Ig fusion protein on leukostasis in a streptozotocin-induced diabetic mouse model.

[0016] Figures 2A-2D are bar graphs showing the effects of an exemplary RAGE-Ig fusion protein on retinal vascular permeability in various retinal layers in a streptozotocin-induced diabetic mouse model.

[0017] Figure 3 is a bar graph showing the effects of an exemplary RAGE-Ig fusion protein on the nitration of retinal proteins in a streptozotocin-induced diabetic mouse model.

[0018] Figure 4 is a bar graph showing the effects of an exemplary RAGE-Ig fusion protein on the retinal expression of ICAM in a streptozotocin-induced diabetic mouse model.

[0019] Figure 5A is a bar graph showing the effects of an exemplary RAGE-Ig fusion protein on the number of acellular capillaries observed per square mm of retinal tissue in diabetic mice after 10 months of diabetes. Figure 5B is a bar graph showing the effects of an exemplary RAGE-Ig fusion protein on the number of pericyte ghosts per 1000 capillary cells observed in diabetic mice after 10 months of diabetes.

[0020] Figure 6 is a bar graph showing the effects of an exemplary RAGE-Ig fusion protein on the 50% response to touch threshold in diabetic mice after 10 months of diabetes.

[0021] Figure 7 provides a flow chart showing the experimental protocol of Example 3.

[0022] Figure 8 is a line graph showing the effects of an exemplary RAGE-Ig fusion protein on test animal weights in a type-II collagen induced arthritis mouse model.

[0023] Figure 9 is a bar graph showing the effects of an exemplary RAGE-Ig fusion protein on the incidence of arthritis in a type-II collagen induced arthritis mouse model.

[0024] Figure 10 is a bar graph showing the effects of an exemplary RAGE-Ig fusion protein on the onset of arthritis in a type-II collagen induced arthritis mouse model.

[0025] Figure 11 is a line graph showing the effects of an exemplary RAGE-Ig fusion protein on incidence of arthritis as a function of time in a type-II collagen induced arthritis mouse model.

[0026] Figure 12 is a line graph showing the effects of an exemplary RAGE-Ig fusion protein on the severity of arthritis as a function of time in a type-II collagen induced arthritis mouse model.

[0027] Figure 13 is a line graph showing the effects of an exemplary RAGE-Ig fusion protein on the number of arthritic paws observed as a function of time in a type-II collagen induced arthritis mouse model.

[0028] Figures 14A-14D are photomicrographs showing the effects of an exemplary RAGE-Ig fusion protein on joint morphology as a function of increasing amounts of the fusion protein in a type-II collagen induced arthritis mouse model.

[0029] Figure 15 is a bar graph showing the effects of an exemplary RAGE-Ig fusion protein on synovitis (black bars) and pannus (grey bars) in a type-II collagen induced arthritis mouse model.

[0030] Figure 16 is a bar graph showing the effects of an exemplary RAGE-Ig fusion protein on marginal erosion (black bars) and architectural changes (grey bars) in a type-II collagen induced arthritis mouse model.

[0031] Figure 17 is a bar graph showing the effects of an exemplary RAGE-Ig fusion protein on overall histological arthritis score in a type-II collagen induced arthritis mouse model.

[0032] Figure 18 is a bar graph showing the effects of an exemplary RAGE-Ig fusion protein on joint matrix protein loss in a type-II collagen induced arthritis mouse model.

[0033] Figures 19A-19D are photomicrographs of toluidine blue stained sections showing the effects of an exemplary RAGE-Ig fusion protein on joint matrix protein loss in a type-II collagen induced arthritis mouse model.

DETAILED DESCRITPION OF THE INVENTION

[0034] Definitions

[0035] As used herein the terms "receptor for advanced glycation end product" or RAGE refer to proteins having amino acid sequences that are substantially similar to the native mammalian RAGE amino acid sequences and function to bind one or more RAGE ligands in a ligand-receptor specific manner. The terms "advanced glycation end product" and "AGE" refer to a heterogeneous group of molecules formed from the nonenzymatic reaction of reducing sugars with free amino groups of proteins, lipids, and nucleic acids as described above.

[0036] As used herein, a "RAGE ligand binding domain" or "RAGE-LBD" refers to any mammalian RAGE protein or any portion of a mammalian RAGE protein that retains the ability to bind a RAGE ligand in a ligand-receptor specific manner. Specifically, without limitation, a RAGE ligand binding domain includes a

polypeptide having one or more extracellular domains of a transmembrane RAGE protein. With reference to Table 6, a suitable RAGE-LBD may comprise at least amino acids 1-99, or amino acids 24-99, or amino acids 1-208, or amino acids 24-208, or amino acids 1-301, or amino acids 24-301, or amino acids 1-344, or amino acids 24-344 of SEQ IDNO:6.

[0037] The term "isolated," as used in the context of this specification to define the purity of the fusion protein, means that the protein is substantially free of other proteins with which it is associated during production, including without limitation substantially free of other proteins present during expression of the fusion protein in a cell culture medium. For example, an isolated protein of the invention may comprise 1- 25%, 20-25%, 15-20%, 10-15%, 5-10%, 1-5% or less than about 2% by mass of protein contaminants residual of production processes. Compositions comprising isolated proteins of the invention, however, can contain other proteins added as stabilizers, carriers, excipients or co-therapeutics.

[0038] As used herein, "protein" and "polypeptide" are interchangeable.

[0039] As used herein "treating" a disease or disorder refers to improving at least one sign or symptom of the subject's disease or disorder.

[0040] The term "nucleic acid" refers to polynucleotides such as deoxyribonucleic acid (DNA), and, where appropriate, ribonucleic acid (RNA). The term should also be understood to include, as equivalents, analogs of either RNA or DNA made from nucleotide analogs, and, as applicable to the embodiment being described, single (sense or antisense) and double-stranded polynucleotides.

[0041] The term "or" is used herein to mean, and is used interchangeably with, the term "and/or," unless context clearly indicates otherwise.

[0042] The term "percent identical" refers to sequence identity between two amino acid sequences or between two nucleotide sequences. Percent identity can be determined by comparing a position in each sequence which may be aligned for purposes of comparison. Expression as a percentage of identity refers to a function of the number of identical amino acids or nucleic acids at positions shared by the compared sequences. Various alignment algorithms and/or programs may be used, including FASTA, BLAST, or ENTREZ. FASTA and BLAST are available as a part of the GCG sequence analysis package (University of Wisconsin, Madison, Wis.), and

can be used with, e.g., default settings. ENTREZ is available through the National Center for Biotechnology Information, National Library of Medicine, National Institutes of Health, Bethesda, Md. In one embodiment, the percent identity of two sequences can be determined by the GCG program with a gap weight of 1, e.g., each amino acid gap is weighted as if it were a single amino acid or nucleotide mismatch between the two sequences.

[0043] Sequence identity may be determined by comparing a reference sequence or a subsequence of the reference sequence to a test sequence (e.g., a nucleotide sequence, an amino acid sequence, etc.). The reference sequence and the test sequence are optimally aligned over an arbitrary number of residues termed a comparison window. In order to obtain optimal alignment, additions or deletions, such as gaps, may be introduced into the test sequence. The percent sequence identity is determined by determining the number of positions at which the same residue is present in both sequences and dividing the number of matching positions by the total length of the sequences in the comparison window and multiplying by 100 to give the percentage. In addition to the number of matching positions, the number and size of gaps is also considered in calculating the percentage sequence identity.

[0044] Sequence identity is typically determined using computer programs. A representative program is the BLAST (Basic Local Alignment Search Tool) program publicly accessible at the National Center for Biotechnology Information (NCBI, http://www.ncbi.nlm.nih.gov/). This program compares segments in a test sequence to sequences in a database to determine the statistical significance of the matches, then identifies and reports only those matches that are more significant than a threshold level. A suitable version of the BLAST program is one that allows gaps, for example, version 2.X (Altschul, et al., Nucleic Acids Res 25(17):3389-402, 1997). Additional suitable programs for identifying proteins with sequence identity to the proteins of the invention include, but are not limited to, PHI-BLAST (Pattern Hit Initiated BLAST, Zhang, et al., Nucleic Acids Res 26(17):3986-90, 1998) and PSI-BLAST (Position-Specific Iterated BLAST, Altschul, et al., Nucleic Acids Res 25(17):3389-402, 1997). The programs are publicly available at the NCBI web site listed above and may be used with the default settings in order to determine sequence identity according to the invention.

[0045] Fusion Proteins

[0046] The present invention provides an isolated fusion protein comprising at least one polypeptide comprising: (a) a first amino acid sequence at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical to a mammalian receptor for advanced glycation end product (RAGE) ligand binding domain, the first amino acid sequence capable of binding a RAGE ligand; and (b) a second amino acid sequence at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical to a human heavy chain immunoglobulin IgG4 constant domain or a fragment thereof, wherein the first amino acid sequence comprises at least one mutation, or at least two mutations, or at least three mutations, or 1-4 mutations, or 1-10 mutations relative to a wild type RAGE ligand binding domain. Examples of mutations that may be made in the first amino acid sequence are those that increase the stability of the fusion protein, for example, by making the RAGE ligand binding domain more resistant to proteolytic degradation, such as those that make the fusion protein more resistant to furin-like proteases. Suitable fragments of the second amino acid sequence include fragments that retain the ability to increase the serum half-life of the fusion proteins of which they are part relative to the serum half life of the same first amino acid sequence alone. Preferably the first amino acid sequence and the second amino acid sequence are derived from human RAGE ligand binding domain and human IgG4.

[0047] Fusion proteins of the invention may comprise one or more amino acid sequences in addition to a RAGE ligand binding domain and an IgG4 constant domain or fragment thereof. For example, a fusion protein of the invention may comprise a linker sequence which may be inserted between the RAGE ligand binding domain and the IgG sequence. Fusion proteins of the invention may comprise one or more tag sequences, for example, purification tag sequences such as 6-Histidines. Fusion proteins of the invention may comprise one or more epitopes recognized by commercially available antibodies, for example, c-myc (EQKLISEEDL, SEQ ID NO: 9) and hemagglutinin (YPYDVPDYA, SEQ ID NO:10) derived from an epitope tag of the influenza hemagglutinin protein.

[0048] Any mammalian RAGE protein known to those of skill in the art may be used in the practice of the present invention. Preferably the extracellular domain of the RAGE protein will be used to identify a ligand binding domain that can be mutated and used as the first amino acid sequence of the fusion protein. Suitable example of mammalian RAGE proteins include, but are not limited to, primate, human (e.g.,

GenBank accession no. NP_001127 and NP_751947), murine (e.g., GenBank accession no. NP_031451), canine (e.g., GenBank accession no. AAQ81297), rat (e.g., GenBank accession no. NP_445788), feline, bovine (e.g., GenBank accession no. AAI20128), ovine, equine and porcine (e.g., GenBank accession no. AAQ73283) RAGE domains.

[0049] RAGE amino acid sequences comprising one or more changes or modifications with respect to the wild type sequence may be used in the present invention. Such changes or modifications include, but are not limited to, point mutations, deletions from the N-terminal, deletions from the C-terminal, internal deletions, and combinations thereof. Any change or modification may be introduced into a RAGE sequence for use in the present invention so long as the resulting protein retains biological activity, e.g., the ability to bind one or more RAGE ligands. The fusion proteins of the invention also include those with or without endogenous glycosylation patterns, including without limitation, fusion proteins in which the first amino acid sequence is derived from a mammalian RAGE ligand binding domain with or without associated native-pattern glycosylation of the binding domain.

[0050] Any suitable IgG Fc region may be used in the practice of the invention, preferably, from an IgG4 molecule, for example, amino acid residues 149-473 of GenBank accession no. AAH25985. An IgG region for use in the present invention may be an IgG4 Fc region and may comprise one or more of the CH2 and CH3 regions of the IgG4 molecule.

[0051] Examples of suitable fusion proteins are provided in the following tables.

[0052] Table 1 provides the nucleotide sequence of a human RAGE-IgG4 Fc fusion protein gene sequence.

[0053] Table 1: Human RAGE-IgG4 Fc Fusion Gene Sequence (SEQ ID NO:1).

ATGGCAGCCGGAACAGCAGTTGGAGCCTGGGTGCTGGTCCTCAGTCTGTGGGGGGCAGTAGTAGGTGCTCA
AAACATCACAGCCCGGATTGGCGAGCCACTGGTGCTGAAGTGTAAGGGGGCCCCCAAGAAACCACCCCAGC
GGCTGGAATGGAAACTGAACACAGGCCGGACAGAAGCCTGGAAGGTCCTGTCTCCCCAGGGAGGAGGCCCC
TGGGACAGTGTGGCTCGTGTCCTTCCCAACGGCTCCCTCTTCCTTCCGGCTGTCGGGATCCAGGATGAGGG
GATTTTCCGGTGCCAGGCAATGAACAGGAATGGAAAGGAGACCAAGTCCAACTACCGAGTCCGTGTCTACC
AGATTCCTGGGAAGCCAGAAATTGTAGATTCTGCCTCTGAACTCACGGCTGGTGTTCCCAATAAGGTGGGG
ACATGTGTGTCAGAGGGAAGCTACCCTGCAGGGACTCTTAGCTGGCACTTGGATGGGAAGCCCCTGGTGCC
GAATGAGAAGGGAGTATCTGTGAAGGAACAGACCAGGAGACACCCTGAGACAGGGCTCTTCACACTGCAGT
CGGAGCTAATGGTGACCCCAGCCCGGGGAGGAGATCCCCGTCCCACCTTCTCCTGTAGCTTCAGCCCAGGC
CTTCCCCGACACCGGGCCTTGCGCACAGCCCCCATCCAGCCCCGTGTCTGGGAGCCTGTGCCTCTGGAGGA
GGTCCAATTGGTGGTGGAGCCAGAAGGTGGACGAGTAGCTCCTGGTGGAACCGTAACCCTGACCTGTGAAG
TCCCTGCCCAGCCCTCTCCTCAAATCCACTGGATGAAGGATGGTGTGCCCTTGCCCCTTCCCCCCAGCCCT
GTGCTGATCCTCCCTGAGATAGGGCCTCAGGACCAGGGAACCTACAGCTGTGTGGCCACCCATTCCAGCCA
CGGGCCCCAGGAAAGCCGTGCTGTCAGCATCAGCATCATCGAACCACGCCGAGGAGGGGCCAACTGCAGGCT
CTGTGGGAGGATCAGGGCTGGGAACTCTAGCCCTGGCCGCTTCCACCAAGGGCCCATCCGTCTTCCCCCTG
GCGCCCTGCTCCAGGAGCACCTCCGAGAGCACAGCCGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGA
ACCGGTGACGGTGTCGTGGAACTCAGGCGCCCTGACCAGCGGCGTGCACACCTTCCCGGCTGTCCTACAGT
CCTCAGGACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCCTCCAGCAGCTTGGGCACGAAGACCTACACC
TGCAACGTAGATCACAAGCCCAGCAACACCAAGGTGGACAAGAGAGTTGAGTCCAAATATGGTCCCCCATG
CCCATCATGCCCAGCACCTGAGTTCCTGGGGGGACCATCAGTCTTCCTGTTCCCCCCAAAACCCAAGGACA
CTCTCATGATCTCCCGGACCCCTGAGGTCACGTGCGTGGTGGTGGACGTGAGCCAGGAAGACCCCGAGGTC
CAGTTCAACTGGTACGTGGATGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCAGTTCAA
CAGCACGTACCGTGTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAACGGCAAGGAGTACAAGT
GCAAGGTCTCCAACAAAGGCCTCCCGTCCTCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCAGCCCCGA
GAGCCACAGGTGTACACCCTGCCCCCATCCCAGGAGGAGATGACCAAGAACCAGGTCAGCCTGACCTGCCT
GGTCAAAGGCTTCTACCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGAACAACTACA
AGACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCCTCTACAGCAGGCTAACCGTGGACAAGAGC
AGGTGGCAGGAGGGGAATGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACTACACACAGAA
GAGCCTCTCCCTGTCTCTCGGGAAATGA

[0054] Bold text is the coding sequence for the RAGE signal sequence, normal text is the coding sequence for human RAGE, and underlined text is the coding sequence for IgG4 Fc region.

[0055] Table 2: Amino acid sequence of a human RAGE-IgG4Fc fusion protein (SEQ ID NO:2).

MAAGTAVGAW	**VLVLSLWGAV**	**VGA**QNITARI	GEPLVLKCKG	APKKPPQRLE	50
WELNTGRTEA	WKVLSPQGGG	PWDSVARVLP	NGSLFLPAVG	IQDEGIFRCQ	100
AMNRNGKETK	SNYRVRVYQI	PGKPEIVDSA	SELTAGVPNK	VGTCVSEGSY	150
PAGTLSWHLD	GKPLVPNEKG	VSVKEQTRRH	PETGLFTLQS	ELMVTPARGG	200
DPRPTFSCSF	SPGLPRHRAL	RTAPIQPRVW	EPVPLEEVQL	VVEPEGGAVA	250
PGGTVTLTCE	VPAQPSPQIH	WMKDGVPLPL	PPSPVLILPE	IGPQDQGTYS	300
CVATHSSHGP	QESRAVSISI	IEPGEEGPTA	GSVGGSGLGT	LALAASTKGP	350
SVFPLAPCSR	STSESTAALG	CLVKDYFPEP	VTVSWNSGAL	TSGVHTFPAV	400
LQSSGLYSLS	SVVTVPSSSL	GTKTYTCNVD	HKPSNTKVDK	RVESKYGPPC	450
PSCPAPEFLG	GPSVFLFPPK	PKDTLMISRT	PEVTCVVVDV	SQEDPEVQFN	500
WYVDGVEVHN	AKTKPREEQF	NSTYRVVSVL	TVLHQDWLNG	KEYKCKVSNK	550
GLPSSIEKTI	SKAKGQPREP	QVYTLPPSQE	EMTKNQVSLT	CLVKGFYPSD	600
IAVEWESNGQ	PENNYKTTPP	VLDSDGSFFL	YSRLTVDKSR	WQEGNVFSCS	650
VMHEALHNHY	TQKSLSLSLG	K			671

[0056] Bold text is the amino acid sequence for the RAGE signal sequence, normal text is the amino acid sequence for human RAGE, and underlined text is the amino acid sequence for IgG4 Fc region.

[0057] Table 3: Human RAGE-Linker-IgG4 Fc Fusion Gene Sequence (SEQ ID NO:3).

ATGGCAGCCGGAACAGCAGTTGGAGCCTGGGTGCTGGTCCTCAGTCTGTGGGGGGCAGTAGTAGGTGCTCA
AAACATCACAGCCCCGGATTGGCGAGCCACTGGTGCTGAAGTGTAAGGGGGCCCCCAAGAAACCACCCCAGC
GGCTGGAATGGAAACTGAACACAGGCCGGACAGAAGCCTGGAAGGTCCTGTCTCCCCAGGGAGGAGGCCCC
TGGGACAGTGTGGCTCGTGTCCTTCCCAACGGCTCCCTCTTCCTTCCGGCTGTCGGGATCCAGGATGAGGG
GATTTTCCGGTGCCAGGCAATGAACAGGAATGGAAAGGAGACCAAGTCCAACTACCGAGTCCGTGTCTACC
AGATTCCTGGGAAGCCAGAAATTGTAGATTCTGCCTCTGAACTCACGGCTGGTGTTCCCAATAAGGTGGGG
ACATGTGTGTCAGAGGGAAGCTACCCTGCAGGGACTCTTAGCTGGCACTTGGATGGGAAGCCCCTGGTGCC
GAATGAGAAGGGAGTATCTGTGAAGGAACAGACCAGGAGACACCCTGAGACAGGGCTCTTCACACTGCAGT
CGGAGCTAATGGTGACCCCAGCCCCGGGGAGGAGATCCCCGTCCCACCTTCTCCTGTAGCTTCAGCCCAGGC
CTTCCCCGACACCGGGCCTTGCGCACAGCCCCCATCCAGCCCCGTGTCTGGGAGCCTGTGCCTCTGGAGGA
GGTCCAATTGGTGGTGGAGCCAGAAGGTGGAGCAGTAGCTCCTGGTGGAACCGTAACCCTGACCTGTGAAG
TCCCTGCCCAGCCCTCTCCTCAAATCCACTGGATGAAGGATGGTGTGCCCTTGCCCCTTCCCCCCAGCCCT
GTGCTGATCCTCCCTGAGATAGGGCCTCAGGACCAGGAACCTACAGCTGTGTGGCCACCCATTCCAGCCA
CGGGCCCCAGGAAAGCCGTGCTGTCAGCATCAGCATCATCGAACCAGGCGAGGAGGGGCCAACTGCAGGCT
CTGTGGGAGGATCAGGGCTGGGAACTCTAGCCCTGGCCCGGTACGGCTCCGGAAGTGGGGCTTCCACCAAG
GGCCCATCCGTCTTCCCCCTGGCGCCCTGCTCCAGGAGCACCTCCGAGAGCACAGCCGCCCTGGGCTGCCT
GGTCAAGGACTACTTCCCCGAACCGGTGACGGTGTCGTGGAACTCAGGCGCCCTGACCAGCGGCGTGCACA
CCTTCCCGGCTGTCCTACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCCTCCAGCAGC
TTGGGCACGAAGACCTACACCTGCAACGTAGATCACAAGCCCAGCAACACCAAGGTGGACAAGAGAGTTGA
GTCCAAATATGGTCCCCCATGCCCATCATGCCCAGCACCTGAGTTCCTGGGGGGGACCATCAGTCTTCCTGT
TCCCCCCAAAACCCAAGGACACTCTCATGATCTCCCGGACCCCTGAGGTCACGTGCGTGGTGGTGGACGTG
AGCCAGGAAGACCCCGAGGTCCAGTTCAACTGGTACGTGGATGGCGTGGAGGTGCATAATGCCAAGACAAA
GCCGCGGGAGGAGCAGTTCAACAGCACGTACCGTGTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGC
TGAACGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGGCCTCCCGTCCTCCATCGAGAAAACCATCTCC
AAAGCCAAAGGGCAGCCCCGAGAGCCACAGGTGTACACCCTGCCCCCATCCCAGGAGGAGATGACCAAGAA
CCAGGTCAGCCTGACCTGCCTGGTCAAAGGCTTCTACCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATG
GGCAGCCGGAGAACAACTACAAGACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCCTCTACAGC
AGGCTAACCGTGGACAAGAGCAGGTGGCAGGAGGGGAATGTCTTCTCATGCTCCGTGATGCATGAGGCTCT
GCACAACCACTACACACAGAAGAGCCTCTCCCTGTCTCTCGGGAAATGA

[0058] Bold text is the coding sequence for the RAGE signal sequence, normal text is the coding sequence for human RAGE, double underline text is the coding sequence

for the peptide linker, and single underlined text is the coding sequence for IgG4 Fc region.

[0059] Table 4: Amino acid sequence of a human RAGE-Linker-IgG4Fc fusion protein (SEQ ID NO:4).

MAAGTAVGAW VLVLSLWGAV VGAQNITARI GEPLVLKCKG APKKPPQRLE 50
WKLNTGRTEA WKVLSPQGGG PWDSVARVLP NGSLFLPAVG IQDEGIFRCQ 100
AMNRNGKETK SNYRVRVYQI PGKPEIVDSA SELTAGVPNK VGTCVSEGSY 150
PAGTLSWHLD GKPLVPNEKG VSVKEQTRRH PETGLFTLQS ELMVTPARGG 200
DPRPTFSCSF SPGLPRHRAL RTAPIQPRVW EPVPLEEVQL VVEPEGGAVA 250
PGGTVTLTCE VPAQPSPQIH WMKDGVPLPL PPSPVLILPE IGPQDQGTYS 300
CVATHSSHGP QESRAVSISI IEPGEEGPTA GSVGGSGLGT LALAGSGSGS 350
GASTKGPSVF PLAPCSRSTS ESTAALGCLV KDYFPEPVTV SWNSGALTSG 400
VHTFPAVLQS SGLYSLSSVV TVPSSSLGTK TYTCNVDHKP SNTKVDKRVE 450
SKYGPPCPSC PAPEFLGGPS VFLFPPKPKD TLMISRTPEV TCVVVDVSQE 500
DPEVQFNWYV DGVEVHNAKT KPREEQFNST YRVVSVLTVL HQDWLNGKEY 550
KCKVSNKGLP SSIEKTISKA KGQPREPQVY TLPPSQEEMT KNQVSLTCLV 600
KGFYPSDIAV EWESNGQPEN NYKTTPPVLD SDGSFFLYSR LTVDKSRWQE 650
GNVFSCSVMH EALHNHYTQK SLSLSLGK 678

[0060] Bold text is the amino acid sequence for the RAGE signal sequence, normal text is the amino acid sequence for human RAGE, double underline text is the amino acid sequence for the peptide linker, and single underlined text is the amino acid sequence for IgG4 Fc region.

[0061] Table 5: Human RAGE variant-IgG4 Fc Fusion Gene Sequence (SEQ ID NO:5).

ATGGCAGCCGGAACAGCAGTTGGAGCCTGGGTGCTGGTCCTCAGTCTGTGGGGGGCAGTAGTAGGTGCTCAAAAC
ATCACAGCCCGGATTGGCGAGCCACTGGTGCTGAAGTGTAAGGGGGCCCCCAAGAAACCACCCCAGCGGCTGGAA
TGGAAACTGAACACAGGCCGGACAGAAGCTTGGAAGGTCCTGTCTCCCCAGGGAGGAGGCCCCTGGGACAGTGTG
GCTCGTGTCCTTCCCAACGGCTCCCTCTTCCTTCCGGCTGTCGGGATCCAGGATGAGGGGATTTTCCGGTGCCAG
GCAATGAACAGGAATGGAAAGGAGACCAAGTCCAACTACCGAGTCCGTGTCTACCAGATTCCTGGGAAGCCAGAA
ATTGTAGATTCTGCCTCTGAACTCACGGCTGGTGTTCCCAATAAGGTGGGGACATGTGTGTCAGAGGGAAGCTAC
CCTGCAGGGACTCTTAGCTGGCACTTGGATGGGAAGCCCCTGGTGCCGAATGAGAAGGGAGTATCTGTGAAGGAA
CAGACCAGGAGACACCCTGAGACAGGGCTCTTCACACTGCAGTCGGAGCTAATGGTGACCCCAGCCCGGGGAGGA
GATCCCCGTCCCACCTTCTCCTGTAGCTTCAGCCCAGGCCTTCCCCGAC**G**CCGGGCCTTGC**A**CACAGCCCCCATC
CAGCCCCGTGTCTGGGAGCCTGTGCCTCTGGAGGAGGTCCAATTGGTGGTGGAGCCAGAAGGTGGAGCAGTAGCT
CCTGGTGGAACCGTAACCCTGACCTGTGAAGTCCCTGCCCAGCCCTCTCCTCAAATCCACTGGATGAAGGATGGT
GTGCCCTTGCCCCTTCCCCCCAGCCCTGTGCTGATCCTCCCTGAGATAGGGCCTCAGGACCAGGGAACCTACAGC
TGTGTGGCCACCCATTCCAGCCACGGGCCCCAGGAAAGCCGTGCTGTCAGCATCAGCATCATCGAACCAGGCGAG
GAGGGCCAACTGCAGGCTCTGTGGGAGGATCAGGGCTGGGAACTCTAGCCCTGGCCGCTTCCACCAAGGGCCCA
TCCGTCTTCCCCCTGGCGCCCTGCTCCAGGAGCACCTCCGAGAGCACAGCCGCCCTGGGCTGCCTGGTCAAGGAC
TACTTCCCCGAACCGGTGACGGTGTCGTGGAACTCAGGCGCCCTGACCAGCGGCGTGCACACCTTCCCGGCTGTC
CTACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCCTCCAGCAGCTTGGGCACGAAGACCTAC
ACCTGCAACGTAGATCACAAGCCCAGCAACACCAAGGTGGACAAGAGAGTTGAGTCCAAATATGGTCCCCCATGC
CCATCATGCCCAGCACCTGAGTTCCTGGGGGGACCATCAGTCTTCCTGTTCCCCCCAAAACCCAAGGACACTCTC
ATGATCTCCCGGACCCCTGAGGTCACGTGCGTGGTGGTGGACGTGAGCCAGGAAGACCCCGAGGTCCAGTTCAAC
TGGTACGTGGATGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCAGTTCAACAGCACGTACCGT
GTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAACGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAA
GGCCTCCCGTCCTCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCAGCCCCGAGAGCCACAGGTGTACACCCTG
CCCCCATCCCAGGAGGAGATGACCAAGAACCAGGTCAGCCTGACCTGCCTGGTCAAAGGCTTCTACCCCAGCGAC
ATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGAACAACTACAAGACCACGCCTCCCGTGCTGGACTCCGAC
GGCTCCTTCTTCCTCTACAGCAGGCTAACCGTGGACAAGAGCAGGTGGCAGGAGGGGAATGTCTTCTCATGCTCC
GTGATGCATGAGGCTCTGCACAACCACTACACACAGAAGAGCCTCTCCCTGTCTCTCGGGAAATGA

[0062] Bold text is the coding sequence for the RAGE signal sequence, normal text is the coding sequence for human RAGE variant, bold wavy underline letters are sites of the point mutations introduced into the variant hRAGE sequence, and underlined text is the coding sequence for IgG4 Fc region.

[0063] Table 6: Amino acid sequence of a human RAGE variant-IgG4Fc fusion protein (SEQ ID NO:6).

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   1 MAAGTAVGAW VLVLSLWGAV VGAQNITARI GEPLVLKCKG APKKPPQRLE
  51 WKLNTGRTEA WKVLSPQGGG PWDSVARVLP NGSLFLPAVG IQDEGIFRCQ
 101 AMNRNGKETK SNYRVRVYQI PGKPEIVDSA SELTAGVPNK VGTCVSEGSY
 151 PAGTLSWHLD GKPLVPNEKG VSVKEQTRRH PETGLFTLQS ELMVTPARGG
 201 DPRPTFSCSF SPGLPRRRAL HTAPIQPRVW EPVPLEEVQL VVEPEGGAVA
 251 PGGTVTLTCE VPAQPSPQIH WMKDGVPLPL PPSPVLILPE IGPQDQGTYS
 301 CVATHSSHGP QESRAVSISI IEPGEEGPTA GSVGGSGLGT LALAASTKGP
 351 SVFPLAPCSR STSESTAALG CLVKDYFPEP VTVSWNSGAL TSGVHTFPAV
 401 LQSSGLYSLS SVVTVPSSSL GTKTYTCNVD HKPSNTKVDK RVESKYGPPC
 451 PSCPAPEFLG GPSVFLFPPK PKDTLMISRT PEVTCVVVDV SQEDPEVQFN
 501 WYVDGVEVHN AKTKPREEQF NSTYRVVSVL TVLHQDWLNG KEYKCKVSNK
 551 GLPSSIEKTI SKAKGQPREP QVYTLPPSQE EMTKNQVSLT CLVKGFYPSD
 601 IAVEWESNGQ PENNYKTTPP VLDSDGSFFL YSRLTVDKSR WQEGNVFSCS
 651 VMHEALHNHY TQKSLSLSLG K
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[0064] Bold text is the amino acid sequence for the RAGE signal sequence, normal text is the amino acid sequence for human RAGE variant, bold wavy underline letters are sites of the point mutations introduced into the variant hRAGE, and underlined text is the amino acid sequence for IgG4 Fc region.

[0065] Table 7: Human RAGE variant-Linker-IgG4 Fc Fusion Gene Sequence (SEQ ID NO:7).

ATGGCAGCCGGAACAGCAGTTGGAGCCTGGGTGCTGGTCCTCAGTCTGTGGGGGGCAGTAGTAGGTGCTCAAAAC
ATCACAGCCCCGGATTGGCGAGCCACTGGTGCTGAAGTGTAAGGGGGCCCCCAAGAAACCACCCCAGCGGCTGGAA
TGGAAACTGAACACAGGCCGGACAGAAGCTTGGAAGGTCCTGTCTCCCCAGGGAGGAGGCCCCTGGGACAGTGTG
GCTCCGTGTCCTTCCCAACGGCTCCCTCTTCCTTCCGGCTGTCGGGATCCAGGATGAGGGGATTTTCCGGTGCCAG
GCAATGAACAGGAATGGAAAGGAGACCAAGTCCAACTACCGAGTCCGTGTCTACCAGATTCCTGGGAAGCCAGAA
ATTGTAGATTCTGCCTCTGAACTCACGGCTGGTGTTCCCAATAAGGTGGGGACATGTGTGTCAGAGGGAAGCTAC
CCTGCAGGGACTCTTAGCTGGCACTTGGATGGGAAGCCCCTGGTGCCGAATGAGAAGGGAGTATCTGTGAAGGAA
CAGACCAGGAGACACCCTGAGACAGGGCTCTTCACACTGCAGTCGGAGCTAATGGTGACCCCAGCCCGGGGAGGA
GATCCCCGTCCCACCTTCTCCTGTAGCTTCAGCCCAGGCCTTCCCCGAC**G**CCGGGCCTTGC**A**CACAGCCCCCATC
CAGCCCCGTGTCTGGGAGCCTGTGCCTCTGGAGGAGGTCCAATTGGTGGTGGAGCCAGAAGGTGGAGCAGTAGCT
CCTGGTGGAACCGTAACCCTGACCTGTGAAGTCCCTGCCCAGCCCCTCTCCTCAAATCCACTGGATGAAGGATGGT
GTGCCCTTGCCCCTTCCCCCCAGCCCTGTGCTGATCCTCCCTGAGATAGGGCCTCAGGACCAGGGAACCTACAGC
TGTGTGGCCACCCATTCCAGCCACGGGCCCCAGGAAAGCCGTGCTGTCAGCATCAGCATCATCGAACCAGGCGAG
GAGGGCCAACTGCAGGCTCTGTGGGAGGATCAGGGCTGGGAACTCTAGCCCTGGCCCGGTAGCGGCTCCGGAAGT
GGGGCTTCCACCAAGGGCCCATCCGTCTTCCCCCTGGCGCCCTGCTCCAGGAGCACCTCCGAGAGCACAGCCGCC
CTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGGTGACGGTGTCGTGGAACTCAGGCGCCCTGACCAGCGGC
GTGCACACCTTCCCGGCTGTCCTACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCCTCCAGC
AGCTTGGGCACGAAGACCTACACCTGCAACGTAGATCACAAGCCCAGCAACACCAAGGTGGACAAGAGAGTTGAG
TCCAAATATGGTCCCCCATGCCCATCATGCCCAGCACCTGAGTTCCTGGGGGGACCATCAGTCTTCCTGTTCCCC
CCAAAACCCAAGGACACTCTCATGATCTCCCGGACCCCTGAGGTCACGTGCGTGGTGGTGGACGTGAGCCAGGAA
GACCCCGAGGTCCAGTTCAACTGGTACGTGGATGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAG
CAGTTCAACAGCACGTACCGTGTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAACGGCAAGGAGTAC
AAGTGCAAGGTCTCCAACAAAGGCCTCCCGTCCTCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCAGCCCCGA
GAGCCACAGGTGTACACCCTGCCCCCATCCCAGGAGGAGATGACCAAGAACCAGGTCAGCCTGACCTGCCTGGTC
AAAGGCTTCTACCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGAACAACTACAAGACCACG
CCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCCTCTACAGCAGGCTAACCGTGGACAAGAGCAGGTGGCAGGAG
GGGAATGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACTACACACAGAAGAGCCTCTCCCTGTCT
CTCGGGAAATGA

[0066] Bold text is the coding sequence for the RAGE signal sequence, normal text is the coding sequence for human RAGE variant, bold wavy underline letters are sites of the point mutations introduced into the variant hRAGE sequence, double underline text is the sequence encoding a peptide linker, and underlined text is the coding sequence for IgG4 Fc region.

[0067] Table 8: Amino acid sequence of human RAGE variant-Linker-IgG4 Fc (SEQ ID NO:8).

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  1 MAAGTAVGAW VLVLSLWGAV VGAQNITART GEPLVLKCKG APKKPPQRLE
 51 WKLNTGRTEA WKVLSPQGGG PWDSVARVLP NGSLFLPAVG IQDEGIFRCQ
101 AMNRNGKETK SNYRVRVYQI PGKPEIVDSA SELTAGVPNK VGTCVSEGSY
151 PAGTLSWHLD GKPLVPNEKG VSVKEQTRRH PETGLFTLQS ELMVTPARGG
201 DPRPTFSCSF SPGLPRRRAL HTAFIQPRVW EPVPLEEVQL VVEPEGGAVA
251 PGGTVTLTCE VPAQPSPQIH WMKDGVPLPL PPSPVLILPE IGPQDQGTYS
301 CVATHSSHGP QESRAVSISI IEPGEEGPTA GSVGGSGLGT LALAGSGSGS
351 GASTKGPSVF PLAPCSRSTS ESTAALGCLV KDYFPEPVTV SWNSGALTSG
401 VHTFPAVLQS SGLYSLSSVV TVPSSSLGTK TYTCNVDHKP SNTKVDERVE
451 SKYGPPCPSC PAPEFLGGPS VFLFPPKPKD TLMISRTPEV TCVVVDVSQE
501 DPEVQFNWYV DGVEVHNAKT KPREEQFNST YRVVSVLTVL HQDWLNGKEY
551 KCKVSNKGLP SSIEKTISKA KGQPREPQVY TLPPSQEEMT KNQVSLTCLV
601 KGFYPSDIAV EWESNGQPEN NYKTTPPVLD SDGSFFLYSR LTVDKSRWQE
651 GNVFSCSVMH EALHNHYTQK SLSLSLGK
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[0068] Bold text is the amino acid sequence for the RAGE signal sequence, normal text is the amino acid sequence for human RAGE variant, bold wavy underline letters are sites of the point mutations introduced into the variant hRAGE, double underline text is the amino acid sequence for the peptide linker, and underlined text is the amino acid sequence for IgG4 Fc region.

EXPRESSION OF RAGE FUSION PROTEINS

[0069] Fusion proteins of the invention may be produced in any protein expression system known to those skilled in the art, for example, eukaryotic expression systems, bacterial expression systems, and viral expression systems. A variety of host-expression vector systems may be utilized to express the fusion protein of the invention. Such host systems represent vehicles in which the fusion proteins of the invention may be produced and from which they may be subsequently purified. Such systems include, but are not limited to microorganisms such as bacteria, yeast, insect cells, or plant cells. RAGE expressed in yeast or mammalian expression systems, e.g., COS-7 cells, may be similar or slightly different in molecular weight and glycosylation pattern than the native molecules, depending upon the expression system. Expression of RAGE DNAs in bacteria such as E. coli provides non-glycosylated molecules. Different glycosylation patterns may be obtained using baculoviral expression systems in insect cells. Functional mutant analogs of mammalian RAGE having inactivated N-glycosylation sites can be produced by oligonucleotide synthesis and ligation or by site-specific mutagenesis techniques.

These analog proteins can be produced in a homogeneous, reduced-carbohydrate form in good yield using yeast expression systems.

[0070] Nucleic acid molecules encoding fusion proteins of the invention may be obtained, and the nucleotide sequence of the polynucleotides determined, by any method known in the art. In view of the teachings herein and the known RAGE polypeptide sequences and their identified or identifiable ligand binding elements, and the known sequences for heavy chain IgG constant domains, nucleotide sequences encoding these polypeptides can be determined using methods well known in the art, i.e. the nucleotide codons known to encode the particular amino acids may be assembled in such a way to generate a nucleic acid that encodes the fusion protein of the invention. Nucleotide codons may be selected based upon the expression system used, for example, by selecting codons that correspond to more abundant tRNA molecules present in the expression system, a higher level of fusion protein may be expressed. Such a polynucleotide encoding the fusion protein may be assembled from chemically synthesized oligonucleotides (e.g. as described in Kutmeier et. Al., Biotechniques 17:242(1994), which, briefly, involves the synthesis of overlapping oligonucleotides containing portions of the sequence encoding the fusion protein, annealing and ligating of those ologonucloetides, and then amplification of the ligated oligonucleotides by polymerase chain reaction(s) (PCR).

[0071] Recombinant expression of a fusion protein of the invention (including other molecules comprising or alternatively consisting of fusion protein fragments or variants thereof) may require construction of an expression vector(s) containing a polynucleotide that encodes the fusion protein. Once a polynucleotide encoding the fusion protein of the invention has been obtained, the vector(s) for the production of the fusion protein may be produced by recombinant DNA technology using techniques well known in the art. Such expression vectors containing RAGE-Fc coding sequences may also contain appropriate transcriptional and translational control signals/sequences, for example, ribosome binding sites (i.e., Kozak sequences), internal ribosome entry sites (IRES), and polyadenylation sites etc.

[0072] Nucleic acid molecules encoding fusion proteins of the invention may be transferred to mammalian cells utilizing replication-defective retroviral vectors (e.g., vectors derived from Moloney murine leukemia virus (MLV) or HIV) and pseudotyped with vesicular stomatitis virus G protein (VSV-G) to stably insert single

copies of nucleic acid molecules encoding the fusion protein of the invention into dividing cells. Retroviral vectors deliver genes coded as RNA that, after entering the cell, are reverse transcribed to DNA and integrated stably into the genome of the host cell. Multiple gene insertions in a single cell may increase the expression and secretion of the fusion protein. Multiple rounds of infection may also increase the number of gene copies integrated and thus the amount of expressed fusion protein. The integrated gene(s) encoding the fusion protein are maintained in the cells through cell division by virtue of their presence in the genome.

[0073] In some embodiments, the present invention provides a stable cell line that expresses fusion proteins of the invention. One suitable method for the rapid generation of stable, high protein expressing mammalian cell lines is using the GPEx™ expression system (Gala Biotech, a business unit of Catalent Pharma Solutions, Middleton, WI., Bleck, Gregory T., Bioprocessingjournal.com September/October 2005 p1-7). Such a method may entail producing a replication defective, pseudotyped retroviral vector based on MMLV and transducing mammalian cells (for example, CHO cells) with the vector. The vector may integrate into the genome of the cells thereby producing a stable cell line.

PURIFICATION OF ISOLATED FUSION PROTEIN

[0074] Isolated fusion proteins of the invention may be prepared by culturing suitable host/vector systems to express the recombinant translation products of the present DNA sequences, which are then purified from culture media or cell extracts using techniques well known in the art.

[0075] For example, supernatants from systems which secrete recombinant protein into culture media can be first concentrated using a commercially available protein concentration filter, for example, an Amicon or Millipore Pellicon ultra filtration unit. Following the concentration step, the concentrate can be applied to a suitable purification matrix. For example, a suitable affinity matrix can comprise, for example, an AGE or lectin or Protein A or Protein G or antibody molecule bound to a suitable support. Alternatively, an anion exchange resin can be employed, for example, a matrix or substrate having pendant diethylaminoethyl (DEAE) groups. The matrices can be acrylamide, agarose, dextran, cellulose or other types commonly employed in protein purification. Alternatively, a cation exchange step can be

employed. Suitable cation exchangers include various insoluble matrices comprising sulfopropyl or carboxymethyl groups. Sulfopropyl groups are preferred.

[0076] Recombinant protein produced in bacterial culture is usually isolated by initial extraction from cell pellets, followed by one or more concentration, salting-out, aqueous ion exchange or size exclusion chromatography steps. Finally, high performance liquid chromatography (HPLC) can be employed for final purification steps. Microbial cells employed in expression of recombinant mammalian RAGE can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents.

[0077] Fermentation of yeast which expresses the fusion protein of the invention as a secreted protein greatly simplifies purification. Secreted recombinant protein resulting from a large-scale fermentation can be purified by methods analogous to those disclosed by Urdal et al. (J. Chromatog. 296:171, 1984). This reference describes two sequential, reversed-phase HPLC steps for purification of recombinant human GMCSF on a preparative HPLC column.

PHARMACEUTICAL COMPOSITIONS

[0078] Fusion proteins of the invention may be formulated in a manner suitable for administration to a subject in need thereof, e.g., may be formulated as pharmaceutical compositions. Compositions of the invention may comprise one or more pharmaceutically-acceptable carrier, excipient or diluent. As used herein "pharmaceutically-acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like that are physiologically compatible. In one embodiment, the carrier is suitable for parenteral administration. A carrier may be suitable for administration into the central nervous system (e.g., intraspinally or intracerebrally). Alternatively, the carrier can be suitable for intravenous, subcutaneous, intraperitoneal or intramuscular administration. In another embodiment, the carrier is suitable for oral administration. Pharmaceutically-acceptable carriers include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the fusion proteins of the invention,

use thereof in the pharmaceutical compositions of the invention is contemplated. Supplementary active compounds can also be incorporated into the compositions.

[0079] Suitable carriers are typically nontoxic to recipients at the dosages and concentrations employed. Ordinarily, the preparation of pharmaceutical compositions of the invention entails combining the fusion protein of the invention with one or more of buffers, antioxidants such as ascorbic acid, low molecular weight (less than about 10 residues) polypeptides, proteins, amino acids, carbohydrates including glucose, trehalose, sucrose or dextrins, chelating agents such as EDTA, glutathione and other stabilizers and excipients. Neutral buffered saline or saline mixed with conspecific serum albumin are exemplary appropriate diluents.

THERAPEUTIC ADMINISTRATION OF FUSION PROTEINS OF THE INVENTION

[0080] The present invention contemplates the administration of the fusion proteins of the invention in the form of a pharmaceutical composition comprising the fusion protein of the invention and a pharmaceutically acceptable diluent or carrier to a subject (e.g., a mammal particularly a human) in need thereof. The present invention also provides a method for treating human disease with such compositions.

[0081] Typically, methods of the invention will comprise administering a pharmaceutical composition comprising a pharmaceutically effective amount of a fusion protein of the invention. The pharmaceutically effective amount employed may vary according to factors such as the disease state, age, sex, and weight of the individual.

[0082] A pharmaceutically effective amount of a fusion protein of the invention may be from about 1 μg fusion protein/1 kg body weight of subject to about 500 mg fusion protein/ 1 kg body weight of subject, or from about 10 μg fusion protein/1 kg body weight of subject to about 500 mg fusion protein/ 1 kg body weight of subject, or from about 100 μg fusion protein/1 kg body weight of subject to about 500 mg fusion protein/ 1 kg body weight of subject, or from about 1 mg fusion protein/1 kg body weight of subject to about 500 mg fusion protein/ 1 kg body weight of subject, or from about 10 mg fusion protein/1 kg body weight of subject to about 500 mg fusion protein/ 1 kg body weight of subject, or from about 100 mg fusion protein/1 kg body weight of subject to about 500 mg fusion protein/ 1 kg body weight of subject, or from about 100 μg fusion protein/1 kg body weight of subject to about 25 mg fusion

protein/ 1 kg body weight of subject, or from about 1 mg fusion protein/1 kg body weight of subject to about 25 mg fusion protein/ 1 kg body weight of subject, or from about 5 mg fusion protein/1 kg body weight of subject to about 25 mg fusion protein/ 1 kg body weight of subject, or from about 10 mg fusion protein/1 kg body weight of subject to about 25 mg fusion protein/ 1 kg body weight of subject, or from about 15 mg fusion protein/1 kg body weight of subject to about 25 mg fusion protein/ 1 kg body weight of subject, or from about 100 µg fusion protein/1 kg body weight of subject to about 10 mg fusion protein/ 1 kg body weight of subject, or from about 1 mg fusion protein/1 kg body weight of subject to about 10 mg fusion protein/ 1 kg body weight of subject, or from about 2.5 mg fusion protein/1 kg body weight of subject to about 10 mg fusion protein/ 1 kg body weight of subject, or from about 5 mg fusion protein/1 kg body weight of subject to about 10 mg fusion protein/ 1 kg body weight of subject, or from about 7.5 mg fusion protein/1 kg body weight of subject to about 10 mg fusion protein/ 1 kg body weight of subject.

[0083] In some embodiments, a pharmaceutically effective amount of a fusion protein of the invention may be 0.5 mg fusion protein/1 kg body weight of subject, 1 mg fusion protein/1 kg body weight of subject, 2 mg fusion protein/1 kg body weight of subject, 3 mg fusion protein/1 kg body weight of subject, 4 mg fusion protein/1 kg body weight of subject, 5 mg fusion protein/1 kg body weight of subject, 6 mg fusion protein/1 kg body weight of subject, 7 mg fusion protein/1 kg body weight of subject, 8 mg fusion protein/1 kg body weight of subject, 9 mg fusion protein/1 kg body weight of subject, or10 mg fusion protein/1 kg body weight of subject.

[0084] A unit dosage form refers to physically discrete units suited as unitary dosages for the mammalian subjects to be treated; each unit containing a predetermined quantity of the fusion protein of the invention calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. A unit dosage form of a fusion protein of the invention may be from about 1mg to about 1000 mg, from about 25 mg to about 1000 mg, from about 50 mg to about 1000 mg, from about 100 mg to about 1000 mg, from about 250 mg to about 1000 mg, from about 500 mg to about 1000 mg, from about 100 mg to about 500 mg, from about 200 mg to about 500 mg, from about 300 to about 500 mg, or from about 400 mg to about 500 mg. A unit dose of a fusion protein of the invention may be about 100 mg, 200 mg, 300 mg, 400 mg, 500 mg, 600 mg, or 700 mg.

[0085] Compositions of the invention may comprise fusion proteins of the invention at a level of from about 0.1 wt% to about 20 wt%, from about 0.1 wt% to about 18 wt%, from about 0.1 wt% to about 16 wt%, from about 0.1 wt% to about 14 wt%, from about 0.1 wt% to about 12 wt%, from about 0.1 wt% to about 10 wt%, from about 0.1 wt% to about 8 wt%, from about 0.1 wt% to about 6 wt%, from about 0.1 wt% to about 4 wt%, from about 0.1 wt% to about 2 wt%, from about 0.1 wt% to about 1 wt%, from about 0.1 wt% to about 0.9 wt%, from about 0.1 wt% to about 0.8 wt%, from about 0.1 wt% to about 0.7 wt%, from about 0.1 wt% to about 0.6 wt%, from about 0.1 wt% to about 0.5 wt%, from about 0.1 wt% to about 0.4 wt%, from about 0.1 wt% to about 0.3 wt%, or from about 0.1 wt% to about 0.2 wt% of the total weight of the composition.

[0086] Pharmaceutical compositions of the invention may comprise one or more fusion proteins of the invention at a level of from about 1 wt% to about 20 wt%, from about 1 wt% to about 18 wt%, from about 1 wt% to about 16 wt%, from about 1 wt% to about 14 wt%, from about 1 wt% to about 12 wt%, from about 1 wt% to about 10 wt%, from about 1 wt% to about 9 wt%, from about 1 wt% to about 8 wt%, from about 1 wt% to about 7 wt%, from about 1 wt% to about 6 wt%, from about 1 wt% to about 5 wt%, from about 1 wt% to about 4 wt%, from about 1 wt% to about 3 wt%, or from about 1 wt% to about 2 wt% of the total weight of the composition. Pharmaceutical compositions of the invention may comprise one or more fusion proteins of the invention at a level of about 0.1 wt%, about 0.2 wt%, about 0.3 wt%, about 0.4 wt%, about 0.5 wt%, about 0.6 wt%, about 0.7 wt%, about 0.8 wt%, about 0.9 wt%, about 1 wt%, about 2 wt%, about 3 wt%, about 4 wt%, about 5 wt%, about 6 wt%, about 7 wt%, about 8 wt%, or about 9 wt% based on the total weight of the composition.

[0087] Dosage regimens may be adjusted to provide the optimum therapeutic response. For example, a single bolus may be administered, several divided doses may be administered over time or the dose may be proportionally reduced or increased as indicated by the exigencies of the therapeutic situation. It is especially advantageous to formulate parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Compositions of the invention may be formulated and administered by intravenous, intramuscular, or subcutaneous

injection. In some embodiments, compositions of the invention may be administered subcutaneously or intramuscularly.

[0088] In some embodiments a dosage regimen may entail administering repeat doses, for example, administering a weekly dose. Treatment regimens may entail a weekly dose for one period of time (for example, for four weeks) followed by a less frequent "maintenance" dosage regimen (for example, one monthly or once bimonthly). Dosage regimens may be adjusted to achieve the desired therapeutic outcomes.

[0089] Methods of the invention include methods for suppressing AGE-dependent inflammatory responses in humans comprising administering an effective amount of a pharmaceutical composition comprising one or more fusion protein of the invention.

[0090] Methods of the invention include methods of inhibiting AGE-mediated biological activity comprising administering a pharmaceutical composition comprising one or more fusion proteins of the invention. As discussed above, AGE has been implicated in a variety of diseases or conditions such as autoimmune diseases. Autoimmune disorders diseases or conditions that may be treated, ameliorated, detected, diagnosed, prognosed or monitored using the fusion protein of the invention include but are not limited to dermatitis, glomerulonephritis, multiple sclerosis, uveitis ophthalmia, autoimmune pulmonary inflammation, insulin dependent diabetes mellitus, autoimmune inflammatory eye, systemic lupus erythematosus, insulin resistance, rheumatoid arthritis, diabetic retinopathy, and scleroderma.

[0091] Other disorders that may be treated or prevented with the methods of the invention may be characterized generally as including any disorder in which an affected cell exhibits elevated expression of RAGE or of one or more RAGE ligands, or any disorder that is treatable (i.e., one or more symptoms may be eliminated or ameliorated) by a decrease in RAGE function. For example, RAGE function can be decreased by administration of an agent that disrupts the interaction between RAGE and a RAGE ligand.

[0092] The increased expression of RAGE is associated with several pathological states, such as diabetic vasculopathy, nephropathy, retinopathy, neuropathy, and other disorders, including Alzheimer's disease and immune/inflammatory reactions of blood

vessel walls. RAGE ligands are produced in tissue affected with many inflammatory disorders, including arthritis (such as rheumatoid arthritis). Depositions of amyloid in tissues cause a variety of toxic effects on cells and are characteristic of diseases termed amyloidoses. RAGE binds to beta-sheet fibrillar material, such as that found in amyloid-beta peptide, Abeta, amylin, serum amyloid A and prion-derived peptides. RAGE is also expressed at increased levels in tissues having amyloid structures. Accordingly, RAGE is involved in amyloid disorders. The RAGE-amyloid interaction is thought to result in oxidative stress leading to neuronal degeneration.

[0093] A variety of RAGE ligands, and particularly those of the S100/calgranulin and Amphoterin (HMGB) families are produced in inflamed tissues. This observation is true both for acute inflammation, such as that seen in response to a lipopolysaccharide challenge (as in sepsis) and for chronic inflammation, such as that seen in various forms of arthritis, ulcerative colitis, inflammatory bowel disease, etc. Cardiovascular diseases, and particularly those arising from atherosclerotic plaques, are also thought to have a substantial inflammatory component. Such diseases include occlusive, thrombotic and embolic diseases, such as angina, fragile plaque disorder and embolic stroke, respectively. Tumor cells also evince an increased expression of a RAGE ligand, particularly amphoterin, indicating that cancers are also a RAGE-related disorder. Furthermore, the oxidative effects and other aspects of chronic inflammation may have a contributory effect to the genesis of certain tumors.

[0094] AGE are a therapeutic target for rheumatoid arthritis and other inflammatory diseases.

[0095] Accordingly, the RAGE-related disorders that may be treated with an inventive compositions include, in addition to the autoimmune disorders discussed above: amyloidoses (such as Alzheimer's disease), Crohn's disease, acute inflammatory diseases (such as sepsis), shock (e.g., septic shock, hemorrhagic shock), cardiovascular diseases (e.g., atherosclerosis, stroke, fragile plaque disorder, angina and restenosis), diabetes (and particularly cardiovascular diseases in diabetics), complications of diabetes, prion-related disorders, cancers, vasculitis and other vasculitis syndromes such as necrotizing vasculitides, nephropathies, retinopathies, and neuropathies.

[0096] The following examples are provided for illustrative purposes only, and are in no way intended to limit the scope of the present invention.

EXAMPLES

[0097] In the following examples, experiments in mice were performed with a fusion protein comprising extracellular domains of mouse RAGE (amino acid residues 1-342) fused to the hinge, CH2 and CH3 domains of the mouse IgG2a heavy chain FC region. The construct was expressed in CHO cells using the GPEx™ expression system. The sequence of the mouse RAGE sequence used is provided in the following table.

[0098] Table 9 (SEQ ID NO:11) Sequence of mouse RAGE

MPAGTAARAW VLVLALWGAV AGGQNITART GEPLVLSCKG APKKPPQQLE WKLNTGRTEA
WKVLSPQGGP WDSVARILPN GSLLLPATGI VDEGTFRCRA TNRRGKEVKS NYRVRVYQIP
GKPEIVDPAS ELTASVPNKV GTCVSEGSYP AGTLSWHLDG KLLIPDGKET LVKEETRRHP
ETGLFTLRSE LTVIPTQGGT HPTFSCSFSL GLPRRRPLNT APIQLRVREP GPPEGTQLLV
EPEGGIVAPG GTVTLTCAIS AQPPPQVHWI KDGAPLPLAP SPVLLLPEVG HEDEGTYSCV
ATHPSHGPQE SPPVSIRVTE TGDEGPAEGS VGESGLGTLA LAEPRGPTIK PCPPCKCPAP
NLLGGPSVFI FPPKIKDVLM ISLSPIVTCV VVDVSEDDPD VQISWFVNNV EVHTAQTQTH
REDYNSTLRV VSALPIQHQD WMSGKEFKCK VNNKDLPAPI ERTISKPKGS VRAPQVYVLP
PPEEEMTKKQ VTLTCMVTDF MPEDIYVEWT NNGKTELNYK NTEPVLDSDG SYFMYSKLRV
EKKNWVERNS YSCSVVHEGL HNHHTTKSFS RTPGK

where RAGE signal peptide = plain underline, RAGE extracellular domain = no underline, Mouse IgG2a hinge region = double underline, Mouse IgG2a CH2 region = dash underline, and Mouse IgG2a CH3 region = wavy underline.

EXAMPLE 1

[0099] Effect of RAGE fusion proteins of the invention on streptozotocin induced diabetes in mice.

[00100] Streptozotocin induced diabetes in mice is an art recognized model for diabetes induced retinal changes (see Obrosova IG, Drel VR, Kumagai AK, Szabo C, Pacher P, Stevens MJ. Early diabetes-induced biochemical changes in the retina: comparison of rat and mouse models. *Diabetologia.* 2006 Oct; 49(10):2525-33.)

[0100] The present experiment involved 5 treatment groups containing 15 C57BL/6 mice per group: 1) non-diabetic control; 2) diabetic control containing mice treated with streptozotocin at 45mg/kg on 5 consecutive days before the study starts to induce diabetes; 3) streptozotocin treated mice that also received 10 μg/day mRAGE-

IgG2aFc injected IP, 3 injections/week; 4) streptozotocin treated mice that also received 100 μg/day mRAGE-IgG2aFc injected IP, 3 injections/week; and 5) streptozotocin treated mice that also received 300 μg/day mRAGE-IgG2aFc injected IP, 3 injections/week.

[0101] During the study, the mice were assessed for body weight, blood glucose, glycohemoglobin (GHb), albuminuria, and tactile sensitivity as measure of sensory nerve function. The mice were sacrificed at the end of the study and assessed for retinal vascular permeability using a fluorescent probe, leukocyte adherence to retinal capillaries, and NF-κB-regulated protein expression (COX-2, ICAM, iNOS).

[0102] Results From Two Month Long Study

[0103] The effects of RAGE-Ig fusion protein on the development of diabetes-induced alterations in retinal physiology and metabolism in C57Bl/6J mice were studied. The fusion protein was administered intraperitoneally at 3 different concentrations (10 μg, 100μg, and 300 μg) three times per week. No adverse effects of any dose of drug on body weight gain or overall health of the diabetic mice was seen. Nonfasted blood glucose levels were 155 ±24 mg/dl (mean ±SD), 358 ±38, 417 ±36, 376 ±36, and 370 ±55 in the Non-diabetic control, Diabetic control, Diabetic + 10 μg RAGE-Ig fusion protein, Diabetic + 100 μg RAGE-Ig fusion protein, and Diabetic + 300 μg RAGE-Ig fusion protein groups, respectively.

[0104] Parameters related to retinopathy measured in the short-term studies were (1) leukostasis, (2) permeability of endogenous albumin from retinal vessels, (3) nitration of retinal proteins, and (4) expression of retinal ICAM and COX-2.

[0105] 1. Leukostasis.

[0106] Methods: At 2 months of diabetes, blood was removed from the vasculature of anesthetized animals by complete perfusion with PBS via a heart catheter. Animals then were perfused with fluorescein-coupled Concanavalin A lectin (20 μg/ml in PBS; Vector Laboratories, Burlingame, CA) as described previously (see Joussen et al., *FASEB J.* 2004 Sep;18(12):1450-2). Flat-mounted retinas were imaged via fluorescence microscopy, and the number of leukocytes adherent to the vascular wall was counted.

[0107] Results: A significant increase in leukostasis was demonstrated in mice that had been diabetic for 2 months compared to the nondiabetics ($P < 0.05$). Leukostasis

was not inhibited in any of the groups treated with the RAGE-Ig fusion protein (see Figure 1).

[0108] 2. Vascular permeability

[0109] Methods: At 2 months of diabetes, eyes were cryosectioned (10 μm), fixed in methanol for 10 min, and washed 4x in PBS. Each section was incubated in sheep anti-mouse serum albumin (Abcam, Cambridge MA; AB8940; 1:2000 dilution) for 2 hrs. After washing, sections were incubated in FTIC-labeled secondary antibody (AB 6743; 1:1000 dilution) for 90 min. Under fluorescence microscopy, the average amount of fluorescence was determined in 3 different sites for each of 4 retinal layers (inner plexiform layer, inner nuclear layer, outer plexiform layer, outer nuclear layer). The amount of fluorescence in each site was the average of 10 random measurements, and the amount of fluorescence in each retinal layer was the average of fluorescence in each of the 3 different sites within that layer.

[0110] Results:

[0111] Diabetes resulted in a significant increase in the fluorescence in the nonvascular retina (ie, due to albumin leaking out of the vessels) in each of the 4 retinal layers studied. The results are shown in Figure 2 (2A inner plexiform layer, 2B inner nuclear layer, 2C outer plexiform layer, 2D outer nuclear layer). To assess albumin in the inner and outer nuclear layers, we intentionally measured in the thin space between nuclei, so these numbers might not be as strong as those from the plexiform layers, where there were no nuclei to impair our measurements.

[0112] 3. Nitration of retinal proteins

[0113] Methods: At 2 months of diabetes, retinas were isolated and homogenized. Dot-blots were made, blotting 50 μg protein homogenate from each animal onto nitrocellulose membrane. Membranes were blocked with milk (5%), washed, and immunostained using anti-nitrotyrosine (Upstate Biotechnology, Inc. #05-233; 1:500 dilution) for 2 hrs, and then stained with secondary antibody (Bio-Rad goat anti-mouse IgG-HRP conjugate; 1:1000 dilution) for 1 hour. After extensive washing, immunostaining detected by the antibody was visualized by enhanced chemiluminescence (ECL, Santa Cruz Biotechnology, Santa Cruz, CA). Immunostain-dependent chemiluminescence was recording on film, and the density of

the immunostained dots quantitated. Results are expressed as a percent of values detected in the nondiabetic controls.

[0114] Results:

[0115] Results are shown in Figure 3. Retinal homogenates from diabetic mice showed the expected increase in nitration of proteins. The therapy inhibited this post-translational modification in a dose-dependent manner. Nitration of proteins is regarded to be a parameter of both oxidative and nitrative stress.

[0116] 4. Expression of retinal ICAM and COX-2

[0117] Methods: Retinas were isolated and sonicated, and the supernatant used as whole retinal extract. Samples (50 μg) were fractionated by SDS-PAGE, electroblotted to nitrocellucose membrane, and membranes blocked in Tris-buffered saline containing 0.02% Tween 20 and 5% nonfat milk. Antibodies for ICAM-1 (1:200 dilution; Santa Cruz Biotechnology) and COX-2 were applied, followed by secondary antibody for 1 hour. After washing, results were visualized by enhanced chemiluminescence.

[0118] Results:

[0119] Results are shown in Figure 4. Since ICAM-1 expression on endothelial cells plays a critical role in adhesion of white blood cells to the vessel wall (leukostasis), we measured the effect of diabetes and the therapy on expression of ICAM-1 in retina. Two months of diabetes did result in a significant increase in expression of retinal ICAM-1. Administration of the RAGE-Ig fusion protein resulted in a dose-dependent decrease in expression of the ICAM, and the highest dose significantly inhibited this expression.

[0120] Expression of an immunostained band consistent with the molecular weight for COX-2 did not increase in diabetes and did not change in animals getting the therapy (not shown).

[0121] The endpoints use in this short term study of the effects of the RAGE-Ig fusion protein were selected because all have been found to be associated with the development of the early (degenerative) stages of diabetic retinopathy, ie, various therapies that have been found to inhibit diabetes-induced degeneration of retinal capillaries also have inhibited these defects.

[0122] Inhibition of RAGE did inhibit abnormalities related to vascular permeability and nitrative stress in the retina. Nitrative stress also is regarded as a marker of oxidative stress. The RAGE inhibitor, however, did not inhibit abnormalities related to leukostasis.

EXAMPLE 2

[0123] Effect of RAGE fusion proteins of the invention on long-term streptozotocin induced diabetes in mice.

[0124] Streptozotocin induced diabetes in mice is an art recognized model for diabetes induced retinal changes (see Obrosova IG, Drel VR, Kumagai AK, Szabo C, Pacher P, Stevens MJ. Early diabetes-induced biochemical changes in the retina: comparison of rat and mouse models. *Diabetologia.* 2006 Oct: 49(10):2525-33.)

[0125] The long term studies involved 5 treatment groups containing 25 C57BL/6 mice per group: 1) non-diabetic control; 2) diabetic control containing mice treated with streptozotocin at 45mg/kg on 5 consecutive days before the study starts to induce diabetes; 3) streptozotocin treated mice that also received 10 μg/day mRAGE-IgG2aFc injected IP, 3 injections/week; 4) streptozotocin treated mice that also received 100 μg/day mRAGE-IgG2aFc injected IP, 3 injections/week; and 5) streptozotocin treated mice that also received 300 μg/day mRAGE-IgG2aFc injected IP, 3 injections/week.

[0126] During the study, the mice were assessed for body weight, blood glucose, glycohemoglobin (GHb), albuminuria, and tactile sensitivity as measure of sensory nerve function. The mice were sacrificed at the end of the study and assessed for quantitative histopathology and neurodegeneration in the retina.

[0127] Parameters related to retinopathy measured in the long-term study were (1) acellular capillaries, (2) pericyte ghosts, and (3) ganglion cells. As a marker of peripheral neuropathy, sensitivity of the paw to light touch was also measured in the long-term study.

[0128] Diabetes-induced retinal histopathology

[0129] After 10 mos of diabetes, eyes were fixed in formalin, and one retina from each animal was isolated, washed in running water overnight, and digested for 2 hrs in crude trypsin solution as we have reported previously. The retinal vasculature was

isolated by gently dislodging neural cells with a "brush" made from a single hair. When totally cleaned of neural cells, the isolated vasculature was laid out on a glass microscope slide, dried overnight, stained with hematoxylin and periodic acid—Schiff, dehydrated and coverslipped. Degenerate (acellular) capillaries were quantitated in 6-7 field areas corresponding to the mid-retina (200X magnification) in a masked manner. Acellular capillaries were identified as capillary-sized vessel tubes having no nuclei anywhere along their length, and were reported per square millimeter of retinal area. Pericyte ghosts were estimated from the prevalence of protruding "bumps" in the capillary basement membranes from which pericytes had disappeared. At least 1,000 capillary cells (endothelial cells and pericytes) in 5 field areas in the mid-retina (400X magnification) in a masked manner were examined. Ghosts on any already acellular vessel were excluded.

[0130] To study the effects of diabetes on retinal neurodegeneration, cells in the ganglion cell layer were counted. Formalin-fixed eyes were embedded in paraffin, sectioned sagittally through the retina, going through the optic nerve, and stained with hematoxylin-eosin. The number of cells in the ganglion cell layer were counted in two areas (mid-retina and posterior retina adjacent to optic nerve) on both sides of the optic nerve. Comparable areas from both sides of the optic nerve were averaged, and expressed per unit length.

[0131] Results. As expected base on previous work, long-term diabetes resulted in a significant increase in the number of degenerate, acellular capillaries in the retina (Figure 5A). All doses of the RAGE-Ig fusion protein significantly inhibited this capillary degeneration, without having any effect on the severity of hyperglycemia. Diabetes also tended to increase pericyte degeneration (pericyte ghosts), but the results did not achieve statistical significance (Figure 5B). We previously have found pericyte loss to be much more difficult to detect in diabetic C57Bl/6 mice compared to diabetic rats or larger species, and we now regard it as an unreliable parameter of vascular disease in this model. Perhaps s a result of the failure to detect significant pericyte loss in control diabetics, we did not detect any effect of the RAGE-Ig fusion protein on pericyte loss in these mice.

[0132] Diabetes did not induce a decrease in the number of cells in the retinal ganglion cell layer (ie, neurodegeneration) in these C57Bl/6 mice. This finding was consistent with a prior study of this mouse model. In the absence of an effect of

diabetes on the retinal neurodegeneration, we are unable to assess whether or not the inhibitor would have had an effect on the neurodegeneration.

[0133] Sensitivity to light touch (a marker of peripheral neuropathy).

[0134] Patients with diabetic neuropathy may exhibit a variety of aberrant sensations including spontaneous pain, pain evoked by light touch and hyperalgesia. There is accumulating data that diabetic rodents reproduce this hyperalgesia, and develop a tactile allodynia. In rodents, this is measured as the paw tactile response threshold.

[0135] Methods: Mice (8 mos diabetes) were transferred to a testing cage with a wire mesh bottom and allowed to acclimatize for 10 to 15 min. Von Frey filaments were used to determine the 50% mechanical withdrawal threshold for foot withdrawal. A series of filaments with logarithmically increasing stiffness, starting with one that had a buckling weight of 0.6 g, were applied in sequence to the plantar surface of the right hind paw with a pressure that caused the filament to buckle. Lifting of the paw was recorded as a positive response and a lighter filament was chosen for the next measurement. If there was no response after 5 seconds, the next heaviest filament was used afterwards. This method was continued until four measurements had been made after an initial change in the behavior or until five consecutive negative (6 g) or four consecutive positive (0.4 g) responses had occurred. The resulting sequence of positive and negative scores was used to calculate the 50% withdrawal response threshold.

[0136] Results: Diabetes significantly increased the sensitivity of the paw to light touch, meaning that it required a lower amount of pressure for diabetic animals to withdraw their paw than did nondiabetic animals (Figure 6). This diabetes-induced defect was significantly inhibited at each dose of the sRAGE-Ig fusion protein.

[0137] Retinopathy: The studies conducted using the RAGE-Ig fusion protein were conducted for 2 durations of diabetes: (1) long-term (10 mo) studies to assess the effect of the therapy on long-term histopathology of diabetic retinopathy that develops in mice, and (2) 2-3 mo studies to assess physiologic and molecular effects of the therapy that presumably underlie the effects on long-term histopathology. The physiologic and molecular endpoints studied with respect to effects of the RAGE-Ig fusion protein were selected because all have been found in other studies to be associated with (and likely causally related to) the development of the early

(degenerative) stages of diabetic retinopathy. All three doses of the therapy clearly and significantly inhibited the diabetes-induced degeneration of the retinal vasculature. Likewise, all three doses of the drug seemed to inhibit also diabetes-induced increase in retinal permeability in these mice. These findings are of major clinical significance, because the early (nonproliferative) stages of diabetic retinopathy still are defined based on vascular pathology (vascular nonperfusion and degeneration, and increased permeability).

[0138] The effect of the therapy on the measured molecular and physiologic endpoints in retinas from diabetic mice was mixed. Inhibition of RAGE did inhibit abnormalities related to nitrative stress, a marker of oxidative stress in the retina. The RAGE inhibitor, however, did not inhibit abnormalities related to leukostasis. The lack of effect of the therapy on leukostasis is surprising in that another group recently reported that their sRAGE did inhibit the increase in leukostasis in diabetes. Evidence that we have generated since the start of our studies using the RAGE-Ig fusion protein (Diabetes 57:1387-93, 2008), however, indicates that effects of a drug therapy on retinal leukostasis in diabetes does not predict the effect of the therapy on the degeneration of retinal capillaries in diabetes. Thus, the lack of the therapy on retinal leukostasis in no way diminishes the significance of the observed effects of the drug.

[0139] Surprisingly, there appeared to be a dose effect of the drug with respect to expression of ICAM-1 and nitration of proteins in retinas from the diabetic animals, whereas this dose effect was not apparent on retinal capillary permeability and degeneration. This would seem to suggest that neither ICAM nor nitration are involved in the retinal vascular defects in diabetes, although we have data using ICAM-1 knockout animals that argues against this conclusion.

[0140] It is clear that the drug did get to the retina, did exert biologic effects, and did demonstrate a significant ability of the drug to inhibit at least the early vascular lesions of diabetic retinopathy.

[0141] Sensory neuropathy: It has been postulated by others that advanced glycation endproducts (AGEs) and interaction of these AGEs with RAGE induce oxidative stress, upregulate NF-kB and various NF-kB-mediated proinflammatory genes in the nerves, and exaggerate neurological dysfunction, including altered pain sensation. The present data is consistent with evidence that RAGE-mediated signaling

contributes to the development of at least some aspects of diabetic neuropathy, and provides evidence that the sRAGE-Ig fusion protein inhibits this process in long-term studies.

EXAMPLE 3

[0142] Evaluation of a RAGE-Ig fusion protein using the Type II Collagen-Induced Arthritis Mouse Model.

[0143] Immunization of susceptible strains of mice with type II collagen, the major component of joint cartilage, induces a progressive, inflammatory arthritis (Wooley et al. *Journal of Experimental Medicine* 1981; 154:688-700). Collagen induced arthritis (CIA) is characterized clinically by erythema and edema, with affected paw width increases of typically 100%. A clinical scoring index has been developed to assess disease progression to joint distortion and spondylitis (Wooley, *Methods In Enzymology* 1988; 162:361-373). Histopathology of affected joints reveals synovitis, pannus formation, and cartilage and bone erosion, which may also be represented by an index. Immunological laboratory findings include high antibody levels to type II collagen, and hypergammaglobulinemia. This model is now well established for testing of immunotherapeutic approaches to joint disease (Staines et al. *British Journal of Rheumatology* 1994; 33(9):798-807), and has been successfully employed for the study of both biological and pharmacological agents for the treatment of rheumatoid arthritis (RA) (Wooley et al. *Arthritis Rheum* 1993; 36:1305-1314, and Wooley et al. *Journal of Immunology* 1993; 151:6602-6607).

[0144] Antagonism of the RAGE receptor is recognized as a potential therapeutic target in RA. Blockade of RAGE in mice with collagen-induced-arthritis resulted in the suppression of the clinical and histologic evidence of arthritis, and disease amelioration was associated with a reduction in the levels of TNFα, IL-6, and matrix metalloproteinases MMP-3, MMP-9 and MMP-13 in arthritic paw tissue (Hofmann et al. *Genes Immun* 2002; 3(3):123-135). This indicates that the collagen induced arthritis is sensitive to RAGE targeted therapy.

[0145] This experiment will evaluate the influence of RAGE-Ig fusion protein on CIA at three doses administered from the time of immunization with type II collagen. The study design is shown in Figure 7.

[0146] Forty DBA/1 LacJ mice 8-10 weeks of age were obtained from Jackson Labs, and acclimatized in the facility for a minimum of 10 days prior to experimentation. All animals weighed >16 grams at the start of the experiment. Mice were divided into one of four treatment groups: 1) 100 µl sterile PBS by i.p. injection daily; 2) 100 µl sterile PBS containing RAGE-Ig fusion protein at 10µg by i.p. injection daily; 3) 100 µl sterile PBS containing RAGE-Ig fusion protein at 100 µg by i.p. injection daily; and 4) 100 µl sterile PBS containing RAGE-Ig fusion protein at 300 µg by i.p. injection daily.

[0147] Three days after the initial dosing, all mice were injected with 100 µg bovine type II collagen in Freund's complete adjuvant (FCA) intradermally at the base of the tail. Mice were monitored by daily examination for the onset of disease, which was recorded. Mice were weighed weekly, and overall health status noted. Arthritis affected animals were clinically assessed five times per week until ten weeks after immunization, and paw measurements were made three times per week. Mice without signs of arthritis ten weeks after immunization were considered disease negative.

[0148] RESULTS

[0149] Overall Health and Toxicity. No acute toxic episodes occurred during the trial, and all animals survived the duration of the experiment. The treatment was well tolerated, and no adverse signs such as fur matting or irritation were observed. The mouse weights (Figure 8) indicate minor changes in weight over the course of the trial, which is typical due to transient weight loss in individual animals corresponding to the onset of disease. None of these variations between the groups reached statistical significance.

[0150] Incidence and Onset of Arthritis. The terminal incidence of collagen arthritis in the trial is shown in Figure 9. The control mice reached 100% onset, which is not unusual in classic collagen arthritis model, where the typical incidence ranges from 80%-100%. Mice treated with 10 µg day RAGE reached an incidence of 80%, which was not a significant reduction in incidence. Mice treated with 100 µg day RAGE exhibited a 60% incidence of arthritis, which was significantly lower than the control group ($p<0.05$). Surprising the incidence of arthritis in mice treated with 300 µg RAGE was 100%, and thus similar to the control incidence.

[0151] The mean (and SEM) of the day of disease onset is shown in Figure 10. Disease onset was typical in the control group, with a mean day with the first appearance of arthritis of 38.6. Disease onset in mice treated with either 10 μg or 100 μg RAGE was nominally delayed to 42.5, which did not achieved statistical significance. However, disease onset was significantly delayed ($p<0.05$) in mice treated with RAGE at 300 μg. Therefore, although mice at the high dose did not exhibit a reduction in disease incidence, the time to the development of clinically overt arthritis was markedly increased.

[0152] The modulation of the onset of disease by RAGE treatment may be readily assessed by the plot of disease incidence over time (Figure 11). The typical rapid disease onset characteristic of CIA is observed in the control group, while mice treated with RAGE at either 10 μg or 100 μg resulted in a delay of disease onset and a lower terminal incidence of arthritis. For approximately eight weeks, mice treated with 300 μg RAGE developed disease in a similar pattern, but a series of late arthritic animals resulted in a high disease incidence but delayed disease onset.

[0153] Disease Severity and Progression. Analysis of the cumulative joint score in treated and control animals revealed significant effects of RAGE therapy on the severity of collagen-induced arthritis (Figure 12). Control mice developed the typical chronic progressing disease, with a marked increase in the cumulative arthritis index. In contrast, mice treated with RAGE at any dose exhibited a marked decrease in the arthritis score. The difference between the control and treated groups achieved a high level of statistical significance ($p<0.001$) from Day 43 post immunization, and this difference was maintained throughout the trial. Although RAGE therapy of 100 μg/day achieved the lowest arthritis cumulative score, there were no significant differences between the RAGE groups with respect to the arthritis score, suggesting that a 'threshold' effect was achieved, rather than a classic dose dependant effect.

[0154] The analysis on the influence of RAGE therapy on the number of arthritic paws (Figure 13) does show a significant effect on the progression of the disease. Again, a significant influence was observed on the number of involved paws from Day 43 on. The level of significance varied from $p<0.001$ to $p<0.025$, which may reflect that the influence of RAGE was more pronounced upon disease severity than arthritis progression; however, the maximum number of involved paws (40) is more restricted than the maximum cumulative disease score (120). Again, there were no

significant variations between the RAGE treated groups, although the 100 μg RAGE group did exhibit the highest level of retardation of arthritis.

[0155] The results suggest that administration of RAGE protein exerted a marked effect upon collagen-induced arthritis when administered using a prophylactic protocol. There were no overt toxic effects of RAGE injection at any dose, and the treatment appeared to be very well tolerated. The overall disease incidence was significantly reduced in mice receiving 100 μg daily, and a delay of disease onset was observed in mice treated with 300 μg/day. However, the most obvious indication of clinical activity was observed in the reduction of disease score and arthritic paw count, where a wide separation between RAGE treated mice and control animals was detected from Day 43 post immunization. At this point, control animals underwent the typical progression of severe arthritis, while RAGE treatment at all doses retarded the disease progression.

[0156] Histopathological assessment: Limbs from all mice were removed at the completion of the clinical assessment study, and stored in neutral buffered formalin solution. Joints were decalcified for 18 days in 10% formic acid, dehydrated, and embedded in paraffin blocks. Sections were cut along a longitudinal axis, mounted and stained with either hematoxylin and eosin or Toluidine Blue. Specimens were cut to approximately the mid line, and then sagital central samples mounted for evaluation. This allowed a consistent geographic evaluation. Five to ten samples were mounted (usually 4 - 6 samples per slide). After staining, the slides were permanently bonded with coverslips. A minimum of 3 separate sections per specimen were evaluated in a blinded fashion, with the evaluator unaware of the group assignment. On front limbs, all elbow, wrist, and metacarpal joints were scored, while all knee, ankle, and metatarsal joints were scored on the rear paws. Digits were not evaluated, since the sectioning procedure eliminates most PIP joints. Slides were evaluated for the presence of synovitis, pannus formation, marginal erosions, architectural changes (mostly subluxation), and destruction. An overall score, based on these collective points, was then assigned to each section. The scoring system was based as follows:

[0157] Synovitis was judged by the thickness of the synovial membrane, and scored as follows: 0 for less than 3 cells thick; 1 for 3 - 5 cells thick; 2 for 6 - 10 cells thick; 3 for 10 - 20 cells thick; and 4 for 20 - 30 cells thick.

[0158] Pannus formation was scored as follows: 0 for no pannus formation; 1 for microvillus present; 2 for clear pannus attachment; 3 for marked pannus attachment; and 4 for joint space filled by pannus.

[0159] Marginal erosions were scored as follows: 0 for no erosions visible; 1 for minor indentation in area of capsular attachment; 2 for clear erosions of cartilage; 3 for erosions extend into subchondral bone; and 4 for major erosion of bone and cartilage.

[0160] Architectural changes were scored as follows: 0 for normal joint architecture; 1 for edematous changes; 2 for minor subluxation of articulating surfaces; 3 for major subluxation of articulating surfaces; 4 for complete fibrosis and collagen bridging.

[0161] The overall score reflects: 0 for classical normal joint appearance; 1 for minor changes; consistent with remission; may be clinically normal; 2 for definite inflammatory arthritis; 3 major inflammatory, erosive disease; and 4 for destructive, erosive arthritis.

[0162] Cartilage and Bone Matrix degradation. Serial sections were stained for cartilage matrix components using the histochemical stain Toluidine Blue. The toluidine blue sections were evaluated for proteoglycan loss. The staining at the articular surface was compared to staining at the growth plate, and was scored as follows: 0 for No proteoglycan loss; Normal Toluidine Blue staining; 1 for Minor proteoglycan loss; Some loss of staining from the superficial cartilage; 2 for Moderate proteoglycan loss; Weak staining of superficial cartilage; 3 for Significant proteoglycan loss; No Toluidine Blue staining of superficial cartilage;and 4 for Major proteoglycan loss; No Toluidine Blue staining of deep cartilage.

[0163] RESULTS

[0164] Histological Findings of Collagen-induced arthritis. Sections were assessed for the inflammatory and erosive parameters of disease. The appearance of the arthritis (Figure 14) reveals typical inflammatory erosive disease pathology for this time point in the control (PBS treated) group, with the typical arthritic features of synovial hypertrophy and hyperplasia, with marked pannus attachment and marginal erosions.

[0165] Treatment with the RAGE-Ig fusion protein at 10 μg/ml (Figure 14B) resulted in moderate changes in the inflammatory and erosive parameters, with an overall

improvement in the appearance of erosions and disrupted cartilage surfaces. Treatment with the RAGE-Ig fusion protein at 100 μg/ml (Figure 14C) resulted in a reduction in pannus formation and erosions compared with the control, and the overall difference was quite marked. However, administration of the RAGE-Ig fusion protein at 300 μg/ml (Figure 14D) resulted in arthritis that appeared somewhat less severe than the pathology seen in the saline control, but was nevertheless quite severe, with synovial hypertrophy and hyperplasia, with marked pannus attachment and marginal erosions.

[0166] Analysis of the inflammatory scores (Figure 15) revealed a reduction in the inflammation in mice treated with the RAGE-Ig fusion protein at all doses when compared with control (saline-treated) animals. However the synovitis was significantly reduced ($p<0.05$) only in the 100 μg/ml group, and the pannus formation showed similar reductions in score ($p<0.03$). The reductions in the inflammatory disease parameters observed using the RAGE-Ig fusion protein at either 10 μg/ml or 300μg/ml failed to reach statistical significance.

[0167] Assessment of changes in the erosive features (erosions and changes in joint architecture) of collagen-induced arthritis showed a similar pattern of effects. A significant reduction ($p<0.01$) in joint erosions was observed between the group treated with the RAGE-Ig fusion protein at 100 μg/ml when compared with control (saline-treated) animals (Figure 16), while the reductions observed in mice treated with the RAGE-Ig fusion protein at 10 μg/ml and 300 μg/ml did not reach significance.

[0168] The combination of the histopathological parameters into an overall histological arthritis score (Figure 17) reflected the findings of the individual pathology parameters. Significant differences between the control (saline) treated animals and mice treated with the RAGE-Ig fusion protein at 100 μg/ml ($p<0.02$) were observed, and the overall score in mice treated at 10 μg/ml just achieved significance ($p=0.05$), while no significant reductions in the overall disease scores were observed using the RAGE-Ig fusion protein at 300 μg/ml.

[0169] The Toluidine Blue stained sections were examined to determine whether the RAGE-Ig fusion protein influenced the loss of matrix proteins from the arthritic joint. The data (shown in Figures 18 and 19) suggest that the RAGE-Ig fusion protein did

protect against proteoglycan loss, but this effect was only statistically significant ($p<0.05$) at the 100 μg/ml dose. The PBS control group exhibits major loss of cartilage matrix (proteoglycans and collagens), and a marked loss of staining at the proximal cartilage surface is seen in mice treated with the RAGE-Ig fusion protein at 300 μg/ml. In contrast, there is good preservation of the matrix protein with administration of the RAGE-Ig fusion protein at 10 μg/ml or 100 μg/ml.

[0170] The histological findings confirm the clinical data that indicate that treatment of collagen-induced arthritis with the RAGE-Ig fusion protein resulted in an effect on the incidence and severity of the disease. The histological parameters reached high of levels statistical significance in mice treated with 100 μg/ml, and achieved statistical significance on the overall pathology in mice treated with 10 μg/ml. RAGE-Ig fusion protein at 100 μg/ml achieved good preservation of the joint structure, and a significant reduction of all the parameters of arthritis under evaluation. The overall impression is that the RAGE-Ig fusion protein blocked the erosive phase of arthritis, since the degree of inflammatory changes was less influenced than the secondary disease parameters. Mice treated with the RAGE-Ig fusion protein at 300 μg/ml were not protected to the same degree as the lower doses, again raising the possibility of a suppressive response to this level of protein administration. Overall, these findings are in agreement with the clinical observations made in the study, and demonstrate that the RAGE-Ig fusion protein can exert an anti- arthritic effect.

[0171] While the invention has been described in detail, and with reference to specific embodiments thereof, it will be apparent to one of ordinary skill in the art that various changes and modifications can be made therein without departing from the spirit and scope thereof and such changes and modifications may be practiced within the scope of the appended claims. All patents and publications herein are incorporated by reference to the same extent as if each individual publication was specifically and individually indicated to be incorporated by reference in their entirety.

[0172] The reference in this specification to any prior publication (or information derived from it), or to any matter which is known, is not, and should not be taken as an acknowledgment or admission or any form of suggestion that that prior publication (or information derived from it) or known matter forms part of the common general knowledge in the field of endeavour to which this specification

relates.

[0173] Throughout this specification and the claims which follow, unless the context requires otherwise, the word "comprise", and variations such as "comprises" and "comprising", will be understood to imply the inclusion of a stated integer or step or group of integers or steps but not the exclusion of any other integer or step or group of integers or steps.

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THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:

1. An isolated fusion protein comprising at least one polypeptide comprising:

(a) a first amino acid sequence comprising an amino acid sequence selected from the group consisting of amino acids 1-301, amino acids 24-301, amino acids 1-344, or amino acids 24-344 of SEQ ID NO:6; and

(b) a second amino acid sequence at least 95% identical to a human heavy chain immunoglobulin IgG4 constant domain or a fragment thereof.

2. The isolated fusion protein according to claim 1, wherein the first amino acid sequence comprises amino acids 1-344 of SEQ ID NO:6.

3. The isolated fusion protein according to claim 1 or claim 2, wherein the fusion protein comprises an amino acid sequence selected from the group consisting of SEQ ID NO:6, and SEQ ID NO:8.

4. The isolated fusion protein according to any one of claims 1 to 3, wherein the fusion protein comprises the amino acid sequence of SEQ ID NO:6.

5. The isolated fusion protein according to any one of claims 1 to 4, wherein the amino acid sequence of the fusion protein consists of SEQ ID NO:6.

6. The isolated fusion protein according to any one of claims 1 to 5, further comprising a linker between the first amino acid sequence and the second amino acid sequence.

7. An isolated nucleic acid encoding a fusion protein comprising at least one polypeptide comprising:

(a) a first amino acid sequence comprising an amino acid sequence selected from the group consisting of amino acids 1-301, amino acids 24-301, amino acids 1-344, or amino acids 24-344 of SEQ ID NO:6; and

(b) a second amino acid sequence at least 95% identical to a human heavy chain immunoglobulin IgG4 constant domain or a fragment thereof.

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8. An isolated nucleic acid according to claim 7, wherein the first amino acid sequence comprises amino acids 1-344 of SEQ ID NO:6.

9. The isolated nucleic acid according to claim 7 or claim 8, wherein the fusion protein comprises an amino acid sequence selected from the group consisting of SEQ ID NO:6, and SEQ ID NO:8.

10. The isolated nucleic acid according to any one of claims 7 to 9, wherein the fusion protein comprises SEQ ID NO:6.

11. The isolated nucleic acid according to any one of claims 7 to 10, wherein the fusion protein has an amino acid sequence that consists of SEQ ID NO:6.

12. An isolated nucleic acid according to any one of claims 7 to 11, further comprising a sequence encoding a linker between the first amino acid sequence and the second amino acid sequence.

13. A recombinant host cell comprising the nucleic acid of any one of claims 7 to 12.

14. A pharmaceutical composition comprising the isolated fusion protein of any one of claims 1 to 6 and a pharmaceutically acceptable carrier.

15. A method of treating an AGE-mediated disease, comprising:
administering to a mammal having an AGE-mediated disease a therapeutically effective amount of the pharmaceutical composition of claim 14.

16. The method according to claim 15, wherein the disease is diabetic nephropathy.

17. The method according to claim 15, wherein the disease is rheumatoid arthritis.

18. The method according to claim 15, wherein the disease is an autoimmune disease.

19. The method according to claim 15, wherein the disease is selected from the group

consisting of dermatitis, glomerulonephritis, multiple sclerosis, uveitis ophthalmia, autoimmune pulmonary inflammation, insulin dependent diabetes mellitus, autoimmune inflammatory eye, systemic lupus erythematosus, insulin resistance, rheumatoid arthritis, diabetic retinopathy, scleroderma, Alzheimer's disease and diabetes.

20. The method according to any one of claims 15 to 19, wherein the fusion protein comprises the amino acid sequence of SEQ ID NO:6.

21. The method according to any one of claims 15 to 20, wherein the amino acid sequence of the fusion protein consists of SEQ ID NO:6.

22. A method of lowering the levels of ligand bound by RAGE in a mammal in need thereof which comprises administering to the mammal a RAGE ligand-lowering amount of a fusion protein according to any one of claims 1 to 6.

23. The fusion protein according to any one of claims 1 to 6, the nucleic acid according to any one of claims 7 to 12, the cell according to claim 13, the composition according to claim 14, or the method according to any one of claims 15 to 22, substantially as hereinbefore described with reference to the Examples and/or Figures.

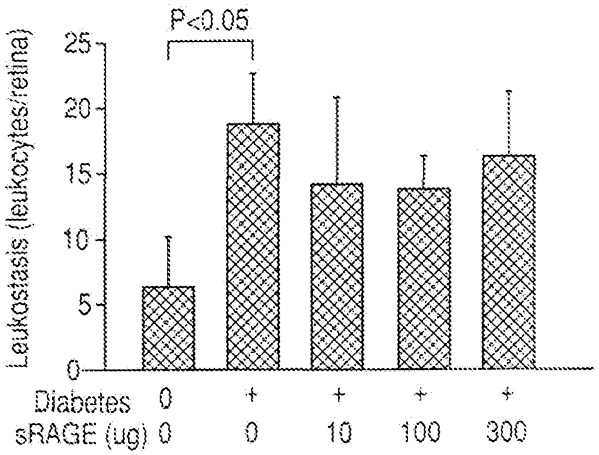
P<0.05
Leukostasis (leukocytes/retina)
25
20
15
10
5
0
Diabetes 0 + + + +
sRAGE (ug) 0 0 10 100 300

FIG. 1

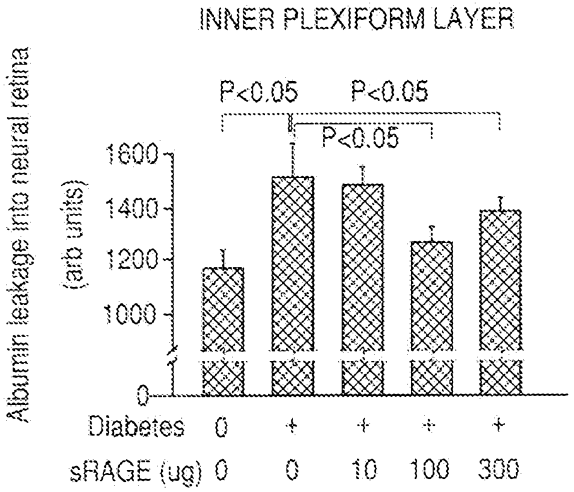
INNER PLEXIFORM LAYER
Albumin leakage into neural retina
(arb units)
P<0.05
P<0.05
P<0.05
1600
1400
1200
1000
0
Diabetes 0 + + + +
sRAGE (ug) 0 0 10 100 300

FIG. 2A

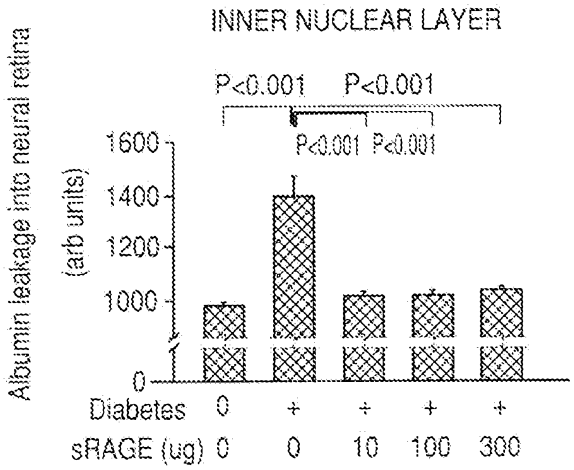
INNER NUCLEAR LAYER
Albumin leakage into neural retina
(arb units)
P<0.001
P<0.001
P<0.001 P<0.001
1600
1400
1200
1000
0
Diabetes 0 + + + +
sRAGE (ug) 0 0 10 100 300

FIG. 2B

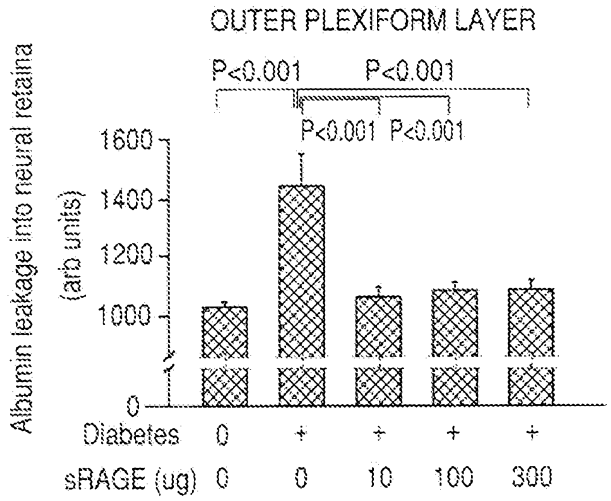
OUTER PLEXIFORM LAYER
P<0.001
P<0.001
P<0.001
P<0.001
Albumin leakage into neural retina
(arb units)
1600
1400
1200
1000
0
Diabetes 0 + + + +
sRAGE (ug) 0 0 10 100 300

FIG. 2C

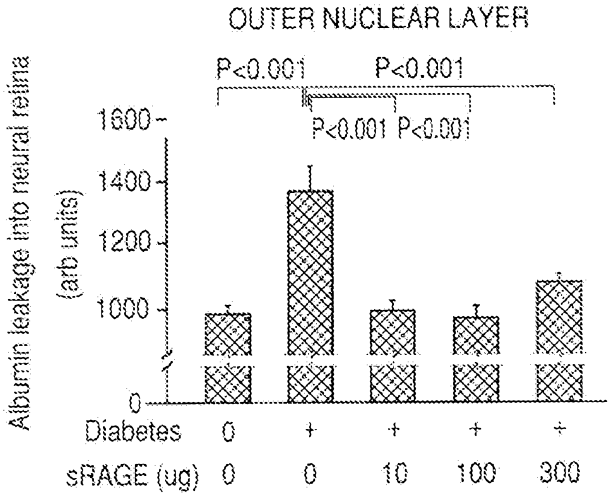
OUTER NUCLEAR LAYER
P<0.001
P<0.001
P<0.001
P<0.001
Albumin leakage into neural retina
(arb units)
1600
1400
1200
1000
0
Diabetes 0 + + + +
sRAGE (ug) 0 0 10 100 300

FIG. 2D

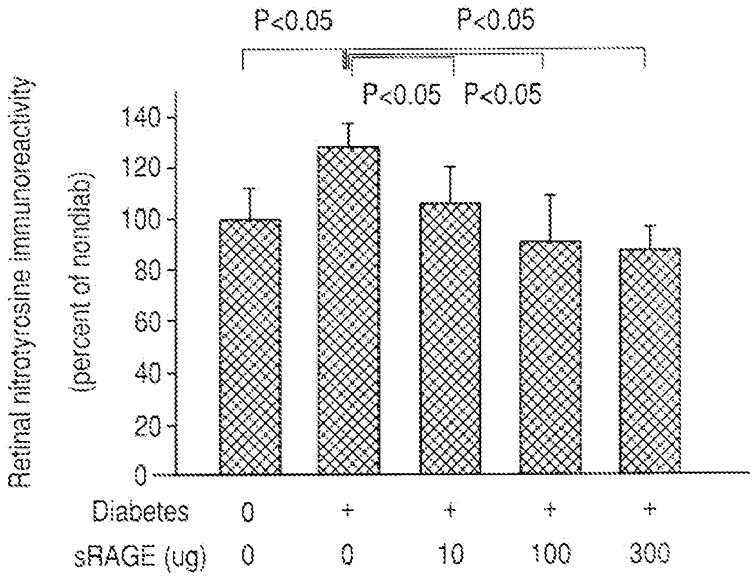
P<0.05
P<0.05
P<0.05
P<0.05
Retinal nitrotyrosine immunoreactivity
(percent of nondiab)
140
120
100
80
60
40
20
0
Diabetes 0 + + + +
sRAGE (ug) 0 0 10 100 300

FIG. 3

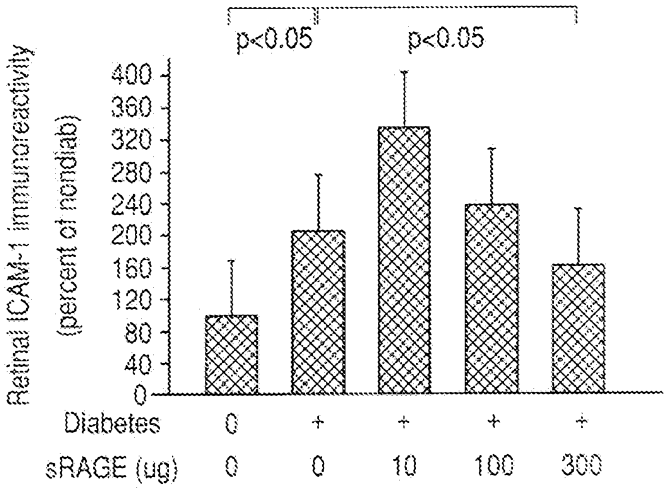
p<0.05
p<0.05
Retinal ICAM-1 immunoreactivity
(percent of nondiab)
400
360
320
280
240
200
160
120
80
40
0
Diabetes 0 + + + +
sRAGE (ug) 0 0 10 100 300

FIG. 4

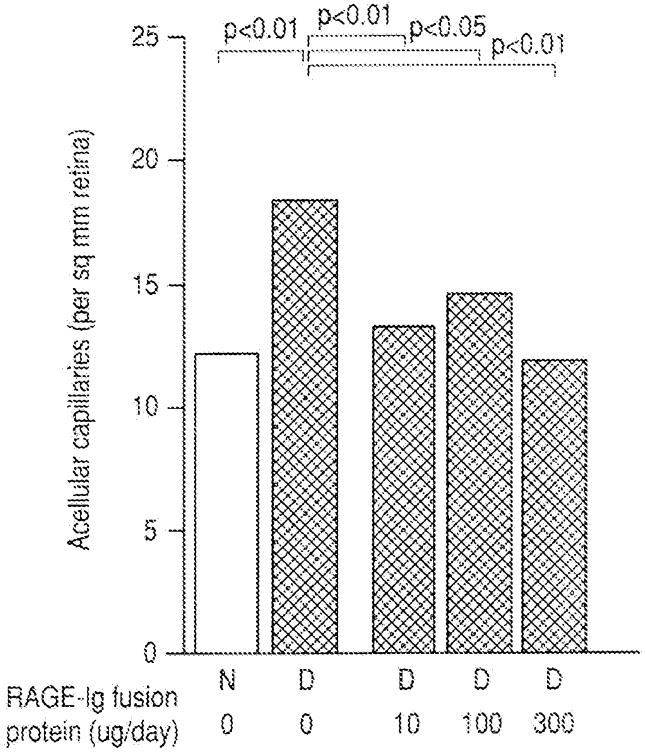
p<0.01
p<0.01
p<0.05
p<0.01
25
20
15
10
5
0
Acellular capillaries (per sq mm retina)
N
D
D
D
D
RAGE-Ig fusion protein (ug/day)
0
0
10
100
300

FIG. 5A

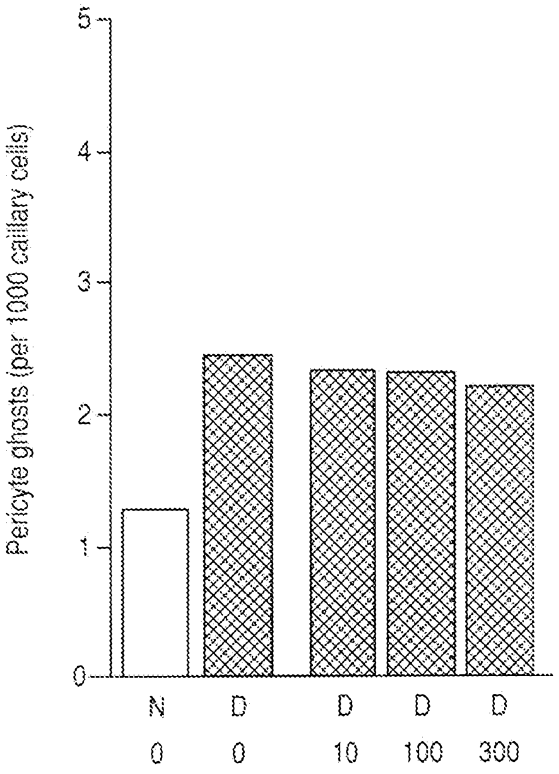
Pericyte ghosts (per 1000 caillary cells)
5
4
3
2
1
0
N
0
D
0
D
10
D
100
D
300

FIG. 5B

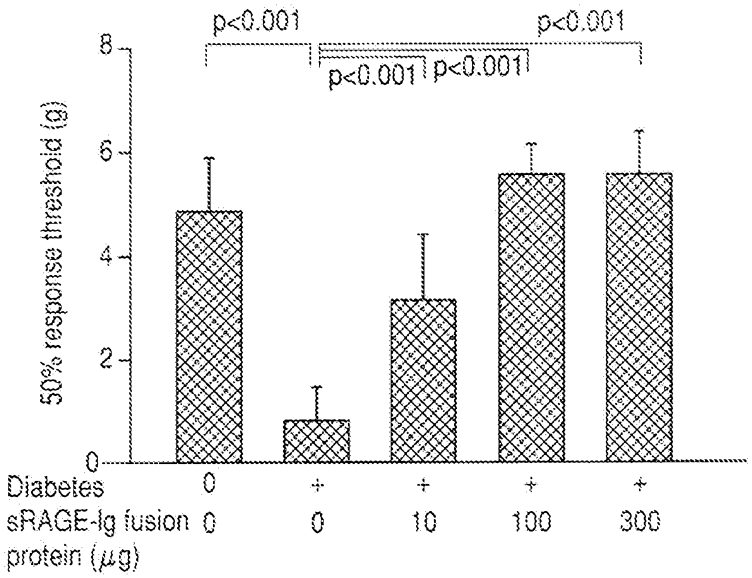
p<0.001
p<0.001
p<0.001
p<0.001
50% response threshold (g)
8
6
4
2
0
Diabetes 0 + + + +
sRAGE-Ig fusion protein (μg) 0 0 10 100 300

FIG. 6

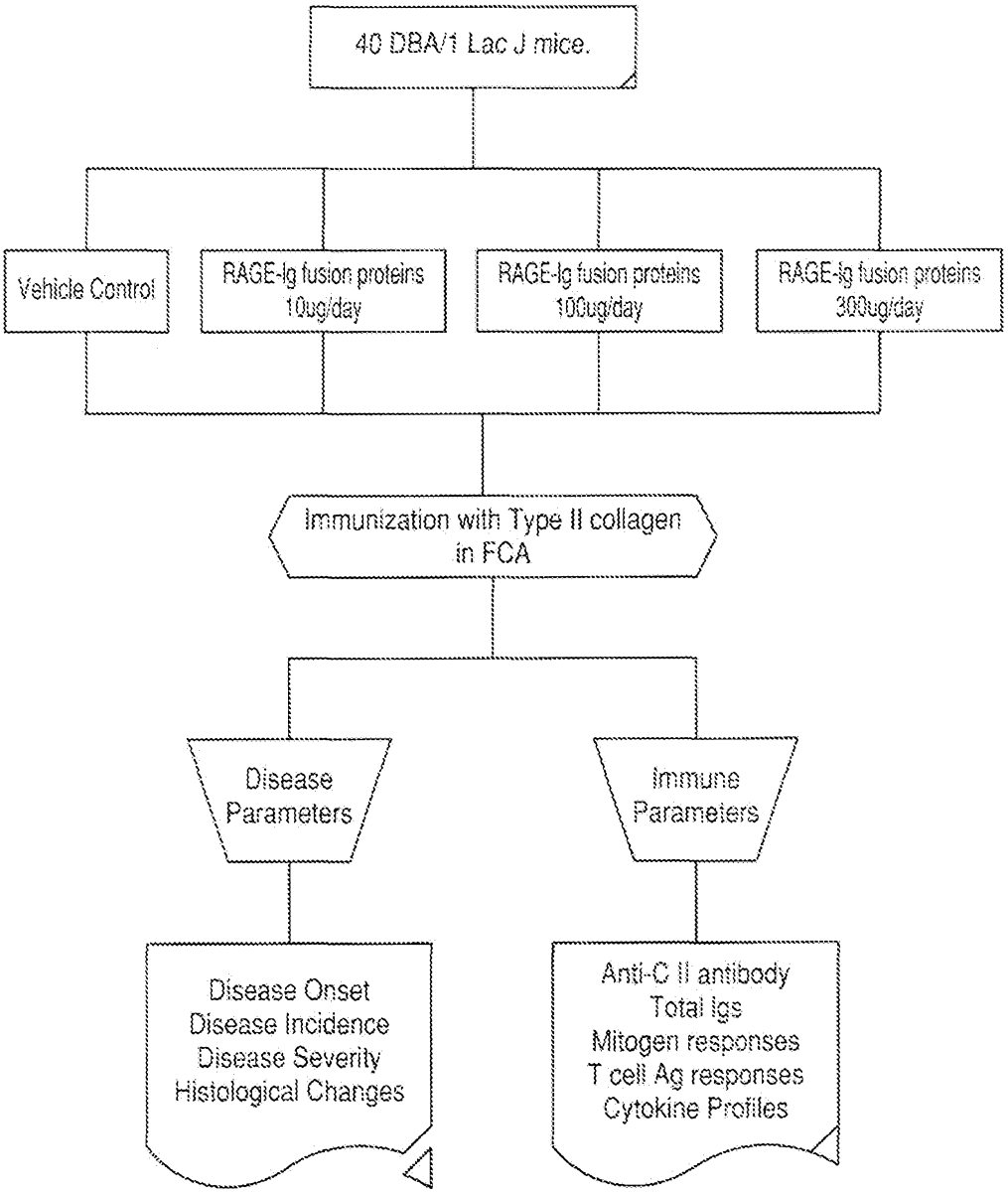
40 DBA/1 Lac J mice.
Vehicle Control
RAGE-Ig fusion proteins 10ug/day
RAGE-Ig fusion proteins 100ug/day
RAGE-Ig fusion proteins 300ug/day
Immunization with Type II collagen in FCA
Disease Parameters
Immune Parameters
Disease Onset
Disease Incidence
Disease Severity
Histological Changes
Anti-C II antibody
Total Igs
Mitogen responses
T cell Ag responses
Cytokine Profiles

FIG. 7

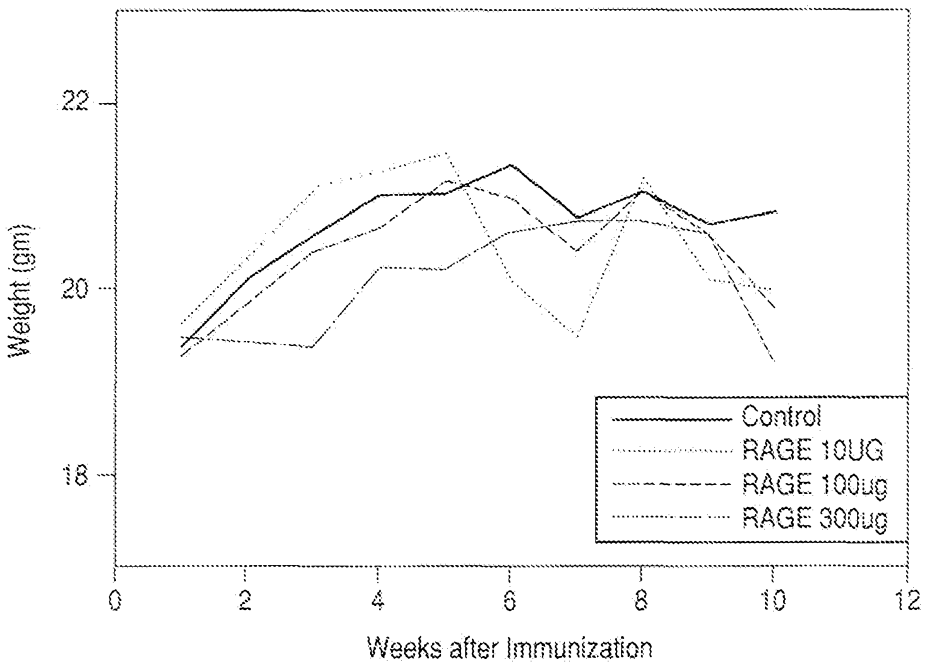
22
20
18
Weight (gm)
0
2
4
6
8
10
12
Weeks after Immunization
Control
RAGE 10UG
RAGE 100ug
RAGE 300ug

FIG. 8

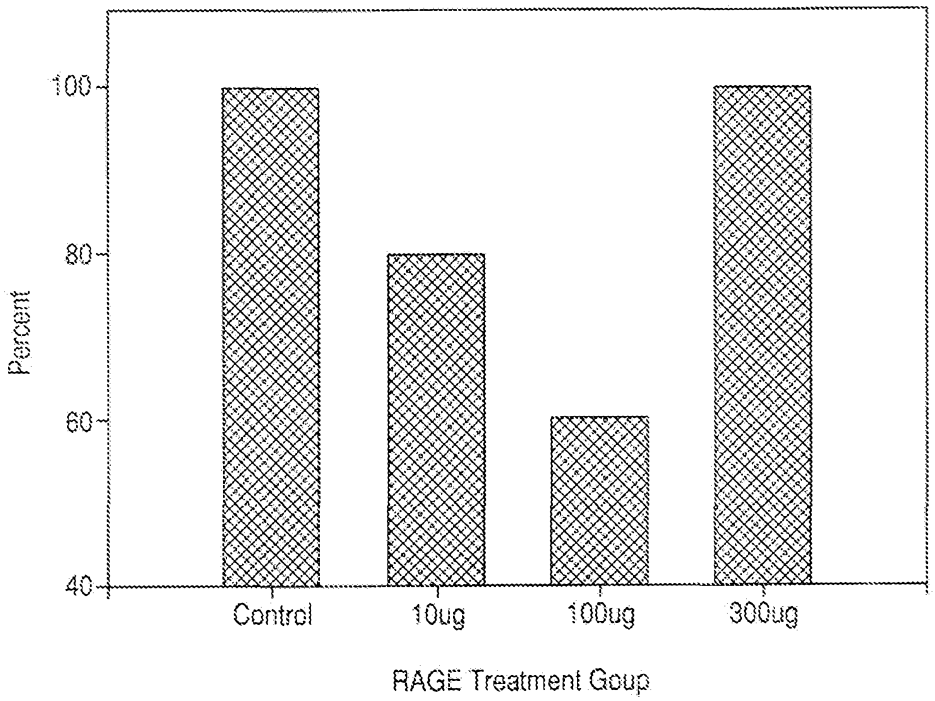
100
80
60
40
Percent
Control
10ug
100ug
300ug
RAGE Treatment Goup

FIG. 9

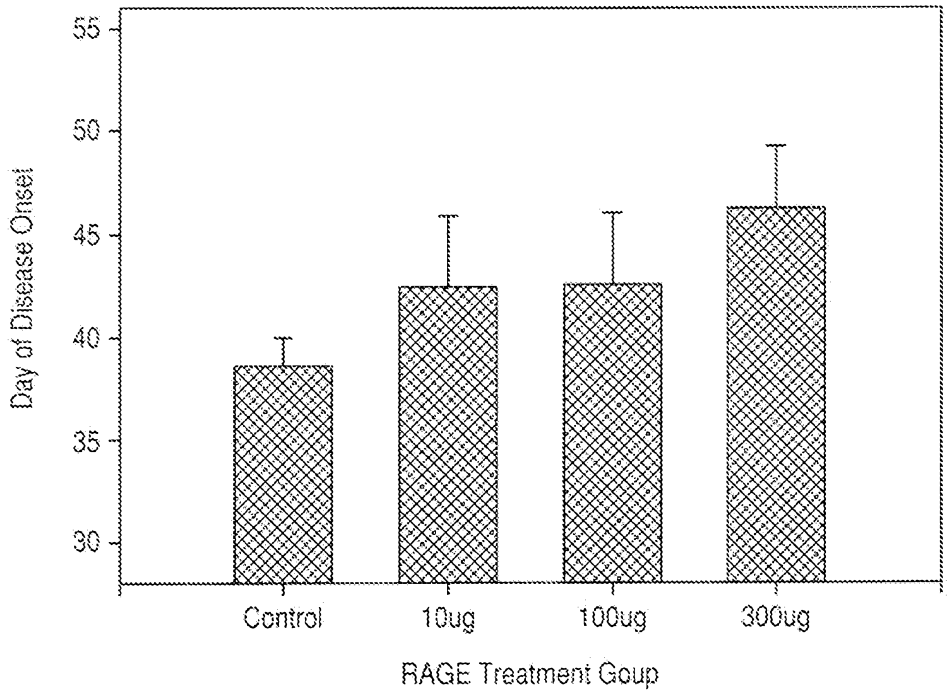
55
50
45
40
35
30
Day of Disease Onset
Control
10ug
100ug
300ug
RAGE Treatment Goup

FIG. 10

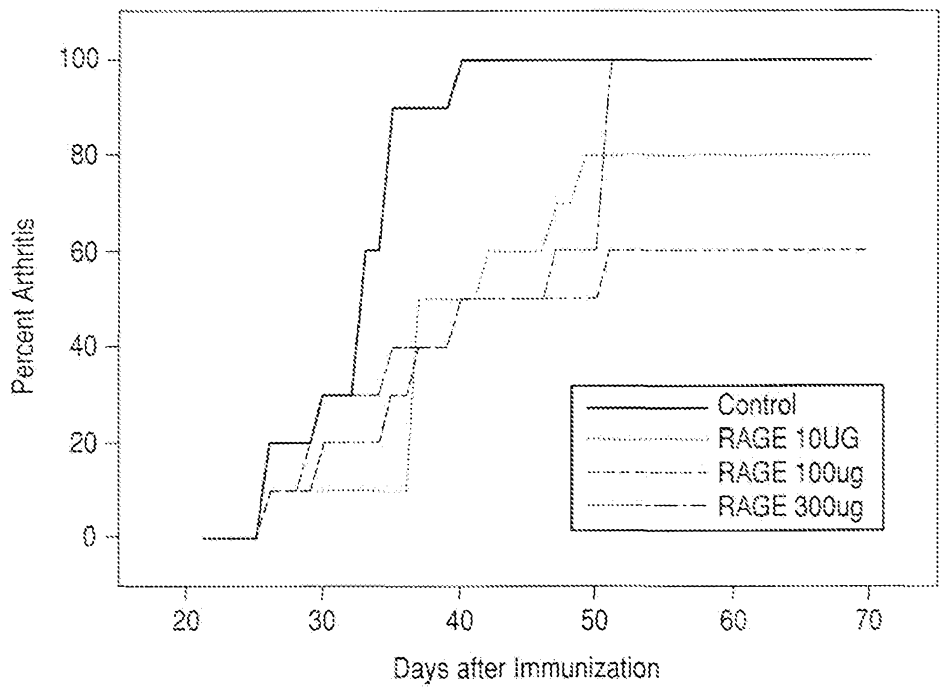
Percent Arthritis
100
80
60
40
20
0
20
30
40
50
60
70
Days after Immunization
Control
RAGE 10UG
RAGE 100ug
RAGE 300ug

FIG. 11

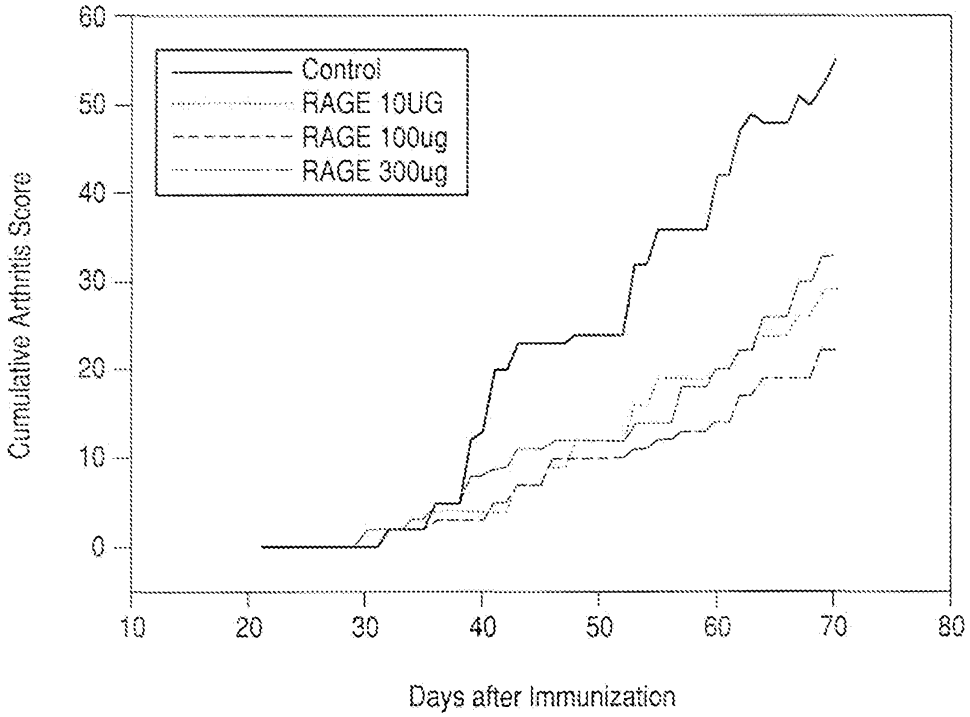
Cumulative Arthritis Score
60
50
40
30
20
10
0
Control
RAGE 10UG
RAGE 100ug
RAGE 300ug
10
20
30
40
50
60
70
80
Days after Immunization

FIG. 12

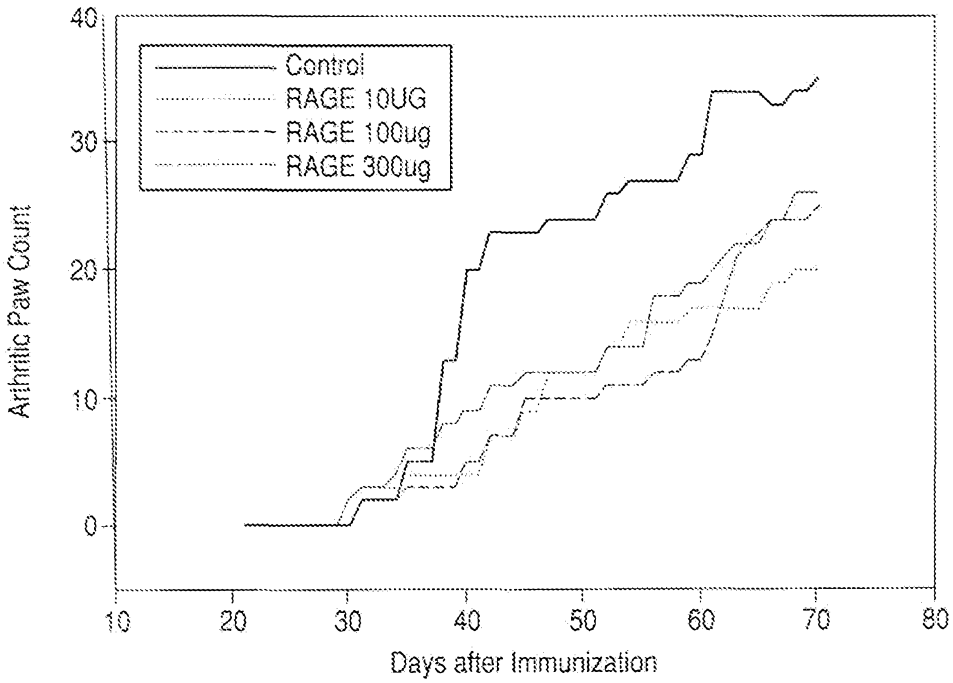
Control
RAGE 10UG
RAGE 100ug
RAGE 300ug
Arthritic Paw Count
0
10
20
30
40
10
20
30
40
50
60
70
80
Days after Immunization

FIG. 13

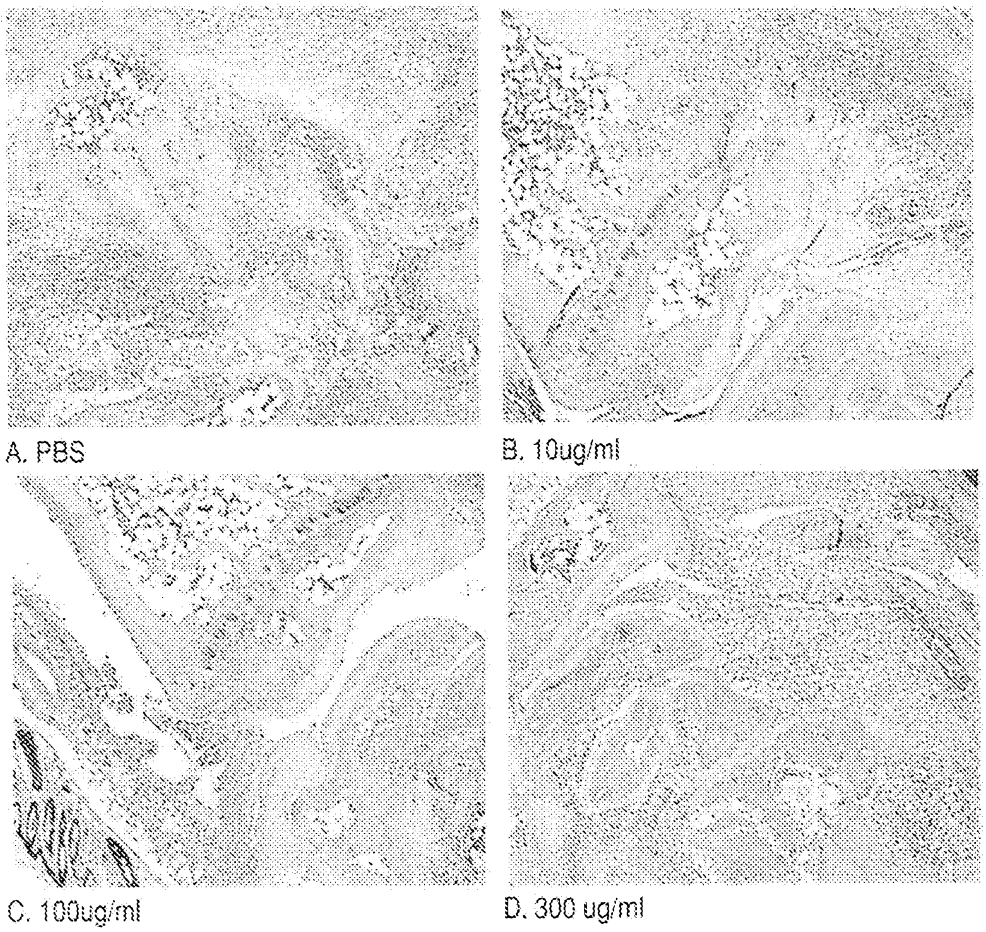

FIG. 14

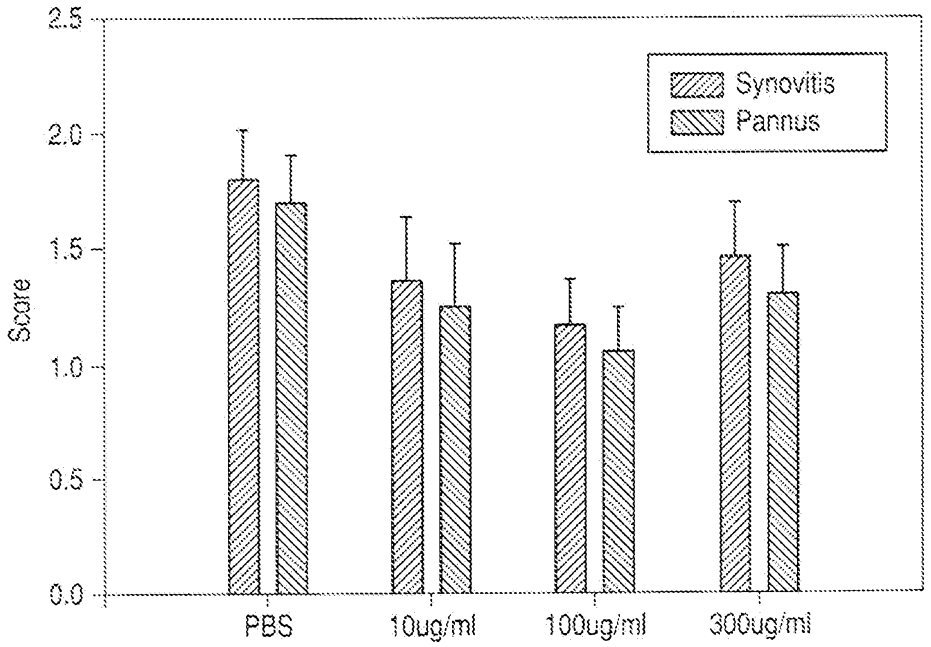
2.5
2.0
1.5
1.0
0.5
0.0
Score
Synovitis
Pannus
PBS
10ug/ml
100ug/ml
300ug/ml

FIG. 15

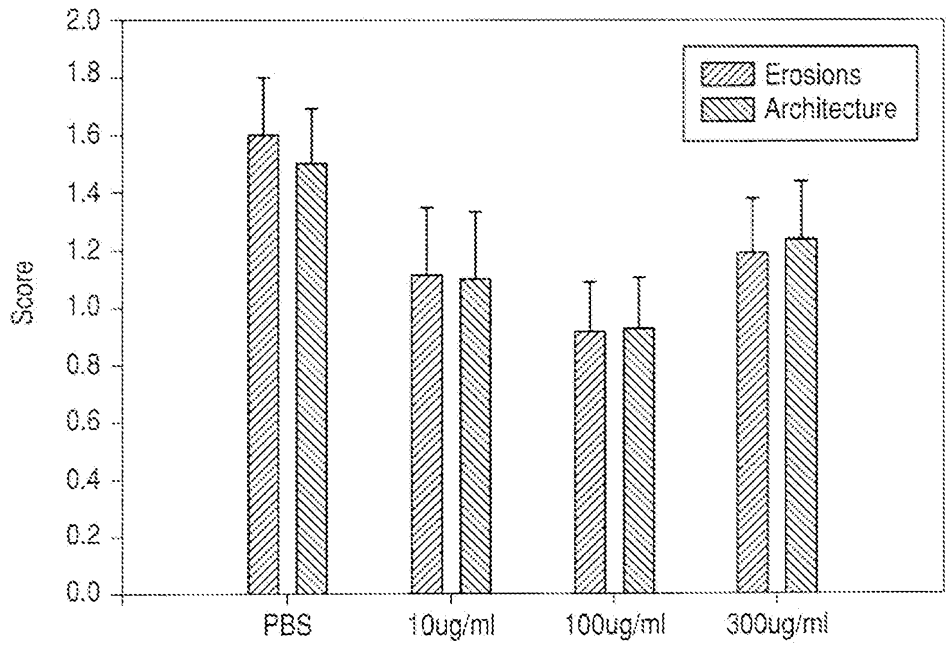
2.0
1.8
1.6
1.4
1.2
1.0
0.8
0.6
0.4
0.2
0.0
Score
Erosions
Architecture
PBS
10ug/ml
100ug/ml
300ug/ml

FIG. 16

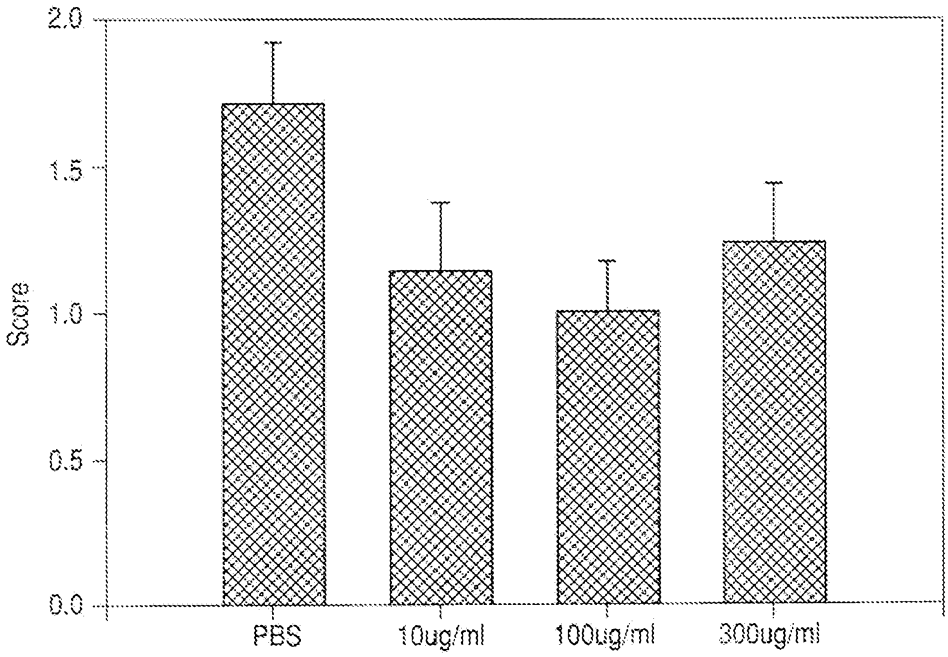

FIG. 17

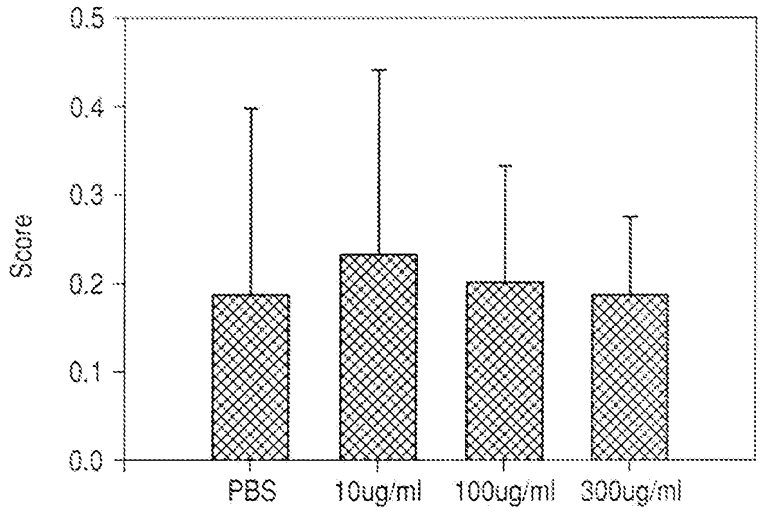

FIG. 18

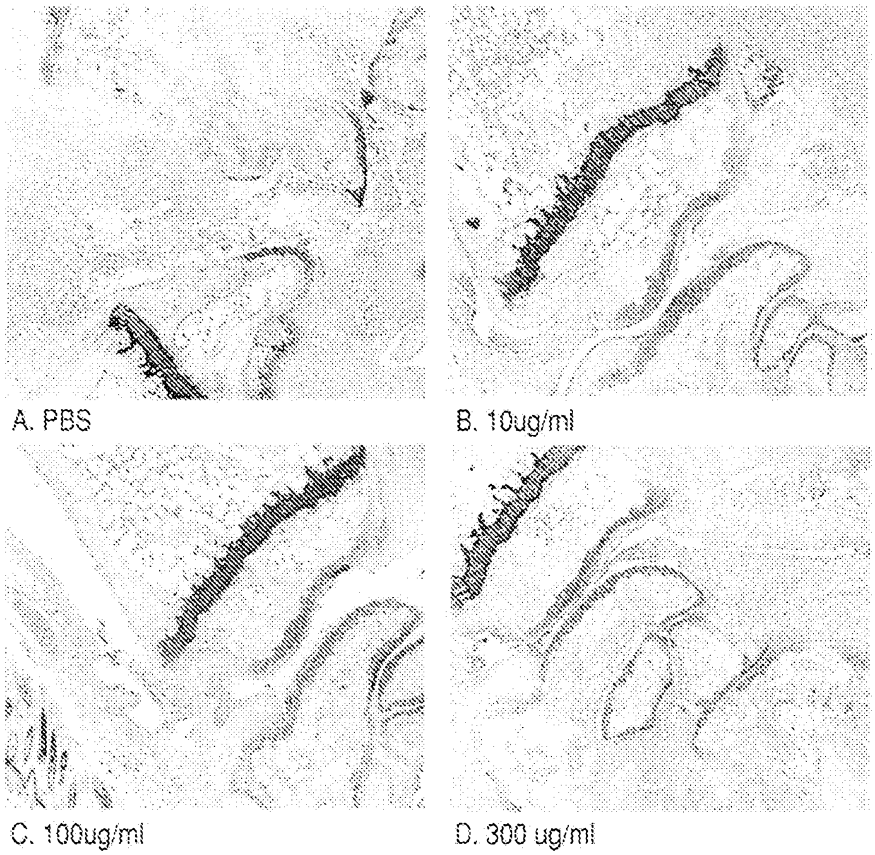

FIG. 19