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(54) **Extracelluláris Streptomyces koganeinsis hialuronidáz**

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(54) **EXTRACELLULAR YALURONIDASE FROM STREPTOMYCES KOGANEIENSIS**

EXTRAZELLULÄRE HYALURONIDASE AUS STREPTOMYCES KOGANEIENSIS

YALURONIDASE EXTRACELLULAIRE DE STREPTOMYCES KOGANEIENSIS

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Description**State of the technique**

5 [0001] Hyaluronidase is a hydrolytic enzyme that cleaves hyaluronic acid to D-glucuronic acid and N-acetylglucosamine; in varying manner, it is also able to degrade other acid mucopolysaccharides of the connective tissue. For example, high concentrations of hyaluronidase are found, in the buccal apparatus of leeches, in the venoms of snakes, bees, scorpions and in the culture supernatants of pathogenic bacteria such as pneumococci, β -hemolytic streptococci and *Staphylococcus aureus*. In the human body hyaluronidase is found in the cornea, ciliary body, spleen, skin and
10 testicles. High amounts of hyaluronidase are also found in spermatozoa, thus allowing them to cross the hyaluronic acid barrier that protects the egg cells.

[0002] Hyaluronidase is used in medicine in the treatment of edema, local inflammatory states, hemorrhoids and chilblains and to facilitate the subcutaneous administration of some active ingredients. Some hyaluronidases were also reported to be able to determine a significant reduction in the size of myocardial infarction [1]. In the veterinary field it is
15 used in antibiotic solutions for the treatment of animal diseases, such as bovine mastitis. Furthermore, hyaluronidase can be used as an analytical reagent in some biological assays, for example in the quali-quantitative determination of hyaluronic acid.

[0003] The industrial-scale production and purification of bacterial or animal hyaluronidases are difficult due to the fact that the enzyme becomes unstable in aqueous solution and loses activity upon purification.

20 [0004] US 4,258,134 and the corresponding European patent EP 0 005 751 [2] disclose a hyaluronidase obtained by dialysis and DEAE-and CM-cellulose ion exchange chromatography of the culture supernatant of *Streptomyces koganeiensis* ATCC 31399.

[0005] It has now been found that such protein fraction, obtained after the two chromatography steps, is indeed made of numerous protein components (about 68 in bidimensional electrophoresis), but only one of them has high hyaluronidase activity and marked stability.
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Detailed description of the invention

30 [0006] The invention relates to hyaluronidase from *Streptomyces koganeiensis* ATCC 31399 comprising the N-terminal amino acid sequence shown in SEQ ID No. 1, and having molecular weight of 21.6 kDa, isoelectric point (pI) ranging between 4.4-4.8 and enzyme activity equal to or higher than 40,000 I.U./mg.

[0007] The hyaluronidase according to the invention can be obtained by a process including the following steps:

- 35 a) subjecting the supernatant obtained from fermentation of *Streptomyces koganeiensis* ATCC 31394 to weak cation-exchange chromatography and isolating the protein fraction with hyaluronidase activity;
- b) subjecting the protein fraction with hyaluronidase activity obtained in step a) to diafiltration and strong anion-exchange chromatography and isolating the protein fraction with hyaluronidase activity;
- c) subjecting the protein fraction with hyaluronidases activity obtained in step b) to strong cation-exchange chromatography and isolating the protein fraction with hyaluronidase activity;
- 40 d) subjecting the protein fraction with hyaluronidase activity obtained in step c) to strong anion-exchange chromatography and isolating the protein fraction with hyaluronidase activity.

[0008] Fermentation of the microorganism can be carried out by known methods, particularly the method disclosed in US 4,258,134. The supernatant obtained upon fermentation is then collected, centrifuged and filtered. Furthermore,
45 before being subjected to the chromatography steps according to the invention, the supernatant can be subjected to further treatments aimed at removing residual particulate from the culture, by methods and techniques known to the skilled person. Usually, the centrifuged and filtered supernatant is subjected to concentration and dialysis. Typically, concentration is carried out by ultrafiltration on appropriate polyethersulfone filters with cut-off ranging between 5 and 15 kDa, preferably of 10 kDa; usually the supernatant is concentrated 8 to 12 fold, preferably 10 fold. Once the presence
50 of hyaluronidase activity is verified by a proper assay, for example the modified assay of Dorfman [3], the concentrated supernatant is dialysed with a buffer solution that is chosen depending on the weak cation-exchange resin used in step a), in such a way that hyaluronidase is at such pH conditions as to be able to bind to the resin; this resin comprises carboxyalkyl exchange groups, preferably carboxymethyl groups, such as the "CM-Sepharose® Fast Flow" resin. After properly equilibrating the resin with the same buffer solution in which dialysis was carried out, the sample is loaded and
55 elution is then performed with the same solution to remove unbound proteins, after which the pH is increased to elute bound proteins. When the "CM-Sepharose® Fast Flow" resin is used, dialysis, resin equilibration and unbound proteins elution are carried out with 50 mM sodium acetate solution at pH 4.0, while bound proteins elution is carried out with 50 mM sodium acetate solution at pH 4.5. The bound proteins that exhibit high hyaluronidase activity are pooled in a single

fraction and subjected to strong anion-exchange chromatography [step b)]. Before proceeding, the pooled fractions are clarified by diafiltration, which is carried out by means and methods known to the skilled person, using a buffer solution that allows hyaluronidase to bind to the strong anion-exchange resin used in step b). Such resin comprises trialkylammonium groups, typically trimethylammonium groups, such as the HiTrap[®] Q XL resin (5 ml column). After equilibrating the resin with the same solution used for diafiltration, the fraction obtained in step a) is loaded and elution is then performed with the same solution to remove unbound proteins; then the eluent ionic strength is progressively increased to elute bound proteins. According to a preferred embodiment, the HiTrap[®] Q XL resin is used and diafiltration, column equilibration and unbound proteins elution are carried out with 50 mM Tris-HCl buffer solution at pH 8, while bound proteins elution is carried out by adding NaCl at increasing concentrations to the eluent. By first eluting with 50 mM Tris-HCl, 35 mM NaCl solution at pH 8 and then 50 mM Tris-HCl, 200 mM NaCl solution at pH 8 two fractions are thus obtained, only the second of which, eluted with the solution containing 200 mM NaCl, exhibits hyaluronidase activity. Such fraction is diluted 8 - 12 fold, preferably 10 fold, with a buffer solution that allows hyaluronidase to bind to the strong cation-exchange resin used for step c). The resin comprises sulfonic groups, preferably sulfonyl alkyl groups, even more preferably sulfonyl propyl groups; according to a particularly preferred embodiment of the invention, the "HiTrap[®] SP FF" resin is used. Practically, after equilibrating the resin with the same buffer solution in which the hyaluronidase fraction obtained in step b) was diluted and loading the sample, washing with the same buffer solution (about 20 bed volumes) is carried out, after which bound proteins elution is then performed, by progressively increasing the eluent pH. Typically, for the HiTrap[®] SP FF resin, dilution, column equilibration and washing are carried out with 20 mM sodium phosphate buffer solution at pH 4, while elution is carried out with 50 mM sodium phosphate buffer at pH 4.8; the fractions with high hyaluronidase activity are pooled in a single fraction, which is subjected to strong anion-exchange chromatography [step d)]. Usually, before chromatography, such fraction is diluted 8 - 12 fold, preferably 10 fold, in a proper equilibration buffer, which allows hyaluronidase to bind to the chosen resin. The resin for step d) is a strong anion-exchange resin comprising quaternary ammonium groups; preferably, a Resource Q[®] column is used. After loading the sample, washing with the same equilibration buffer is performed and bound proteins elution is then performed by progressively decreasing the pH by 0.5 units, to pH 4. When a Resource Q[®] column is used, hyaluronidase fraction dilution, column equilibration and washing after sample loading are carried out with 20 mM sodium acetate at pH 5.5; by progressively decreasing the pH as defined above, a first fraction of proteins at pH 5 and two fractions of proteins at pH 4 are obtained; the second fraction eluted at pH 4 has absorbance at 280 nm and enzyme activity higher than the other two fractions, as Figure 2 shows. By subjecting such fraction to 12% SDS-PAGE chromatography and silver staining (Figure 6), a single protein band with apparent molecular weight of about 25 kDa is observed. Particularly, 99% of hyaluronidase purified by the above described process has apparent molecular weight of about 25 kDa. Such protein only comprises about 5% of the hyaluronidase present in the supernatant obtained upon fermentation; hence, the process allows obtaining about 20-fold enrichment with about 30% yield.

[0009] With respect to other hyaluronidases known to date, that of the invention is highly stable in aqueous solution, is not sensitive towards the action of proteolytic enzymes and has HPLC purity higher than 98% (Figs. 5a-5h), which is required for therapeutic use; hence, it may be used, alone or in combination with other active ingredients, in the preparation of pharmaceutical or veterinary compositions for the treatment of diseases in which it is necessary or advantageous to degrade the hyaluronic acid present in the organ or tissue affected by the disease.

[0010] Thanks to high stability in aqueous solution, the hyaluronidase of the invention can also be formulated in the form of aqueous based compositions, such as solutions, hydrophilic creams, hydrogels, in addition to the form of lipophilic products such as ointments or oily creams.

[0011] With regard to human use, the hyaluronidase of the invention can be used for preparation of pharmaceutical compositions for the treatment of edema, particularly traumatic edema, or inflammatory states, such as the hemorrhoidal syndrome; furthermore, it can be used for the preparation of compositions for the treatment of chilblains. The hyaluronidase of the invention can also be used in combination with other drugs whose bioavailability increase is necessary or advantageous.

[0012] For example, for the treatment of traumatic edema, combinations of the hyaluronidase according to the invention with anticoagulant and/or fibrinolytic agents are particularly advantageous. Such combinations may also optionally contain one or more steroidal or nonsteroidal anti-inflammatory agents. Furthermore, sulfated hyaluronic acid, which is also known to have antithrombotic and anticoagulant properties, in addition to anti-inflammatory properties, may advantageously be combined with these compositions. An example of sulfated hyaluronic acid that can be used to that end is disclosed, for example, in EP 0702699.

[0013] Combinations of the hyaluronidase according to the invention with other active ingredients are also advantageous in the case of injectable preparations containing particularly high molecular weight active ingredients, for example monoclonal antibodies, cytokines or enzymes, which are usually administered intravenously; hyaluronidase allows administering them subcutaneously, according to the so-called EASI (Enzymatically-Augmented Subcutaneous Infusion) procedure, which is mainly employed for fluid replacement in terminal patients, in such a way to limit or avoid nursing care. The hyaluronidase according to the invention can also be employed for the preparation of pharmaceutical com-

positions for the treatment of resistant solid tumors; in fact, by degrading hyaluronic acid, it lowers the interstitial fluid pressure in the tumor mass, retarding or inhibiting its growth. For the same reason, it also increases the effectiveness of antitumoral active ingredients optionally combined therewith. Hence, a further aspect of the invention relates to pharmaceutical compositions containing hyaluronidase in combination with one or more antitumoral active ingredients, such as vinca alkaloids (vinblastine, vincristine, vinorelbine) and taxanes (paclitaxel).

[0014] A further therapeutic use of the hyaluronidase according to the invention relates to the treatment of IgE-mediated allergic forms via enzyme potentiated desensitization (EPD = *Enzyme Potentiated Desensitization*), which consists of administering extremely low doses of allergens to desensitize subjects susceptible thereto. By associating hyaluronidase with an allergen it is possible to increase the treatment effectiveness, since the allergen more readily reaches the site of action. Hence, a further object of the invention consists of pharmaceutical compositions containing hyaluronidase in association with one or more allergens that induce IgE-mediated allergic reactions. Hyaluronidase also finds use as a diffusion factor of drugs for odontological use in the treatment of oral cavity diseases, for example local anaesthetics and antibiotics; hence, according to a further aspect, the invention relates to pharmaceutical compositions containing the hyaluronidase according to the invention in association with one or more local anaesthetics or antibiotics.

[0015] In ophthalmology, hyaluronidase allows to markedly expedite the treatment of spontaneous vitreous haemorrhages and can be used, alone or in combination with other active ingredients, in the preparation of pharmaceutical forms for ophthalmic use, such as solutions, suspensions, gels, creams and ointments, for the treatment of said haemorrhages.

[0016] With regard to veterinary use instead, a disease that can effectively be treated with the hyaluronidase of the invention is bovine mastitis; in that case, hyaluronidase can be administered in combination with antibiotics, such as penicillin G, I-IV generation cephalosporins and potentiated aminopenicillins.

[0017] Pharmaceutical compositions can be prepared by techniques and excipients known to the skilled person, for example according to what is described in Remington, "The Science and the Practice of Pharmacy", 21st ed. (Lippincott, Williams & Wilkins); such compositions include, particularly, injectable preparations and topical preparations for dermal, transdermal and ophthalmic application. Particularly, topical preparations for epidermal application can be selected from creams, gels, ointments and spray solutions, while topical preparations for ophthalmic application can be selected from creams, gels, ointments solutions and suspensions. As previously noted, thanks to its stability in aqueous solution, the hyaluronidase according to the invention can be formulated in aqueous based products; the choice between an aqueous and oily based formulation can be made by a skilled technician based on common knowledge in the field of pharmaceutical technology, depending on the other components present in the composition.

[0018] Finally, the hyaluronidase according to the invention can be used as a reagent in biochemical assays for the quali-quantitative determination of hyaluronic acid.

Description of the figures

[0019]

FIG. 1: 2D electrophoresis of the CM-cellulose fraction positive for hyaluronidase activity deriving from culture supernatant of *Streptomyces koganeiensis* ATCC 31394.

FIG. 2: chromatogram of *Streptomyces koganeiensis* hyaluronidase obtained upon Resource Q[®] column chromatography (step d).

FIG. 3: 12% SDS-PAGE protein pattern of the fractions obtained at the end of each purification step according to the invention compared to the supernatant protein pattern.

FIG. 4: analysis of hyaluronidase purity by HPLC on Bio-Sil SEC gel filtration column [step d)].

FIGs. 5a-4h: SDS-PAGE analysis and absorption spectra of the N-terminal sequencing of hyaluronidase obtained in step d).

FIG. 6: non-denaturing SDS-PAGE analysis for determination of hyaluronidase activity; comparison between hyaluronidase obtained in step d) and hyaluronidase obtained according to US 4,258,134.

FIG. 7: mass spectrometry determination of the molecular weight of hyaluronidase obtained in step d).

FIG. 8: determination of the isoelectric point of hyaluronidase obtained in step d).

FIG. 9: comparison between the enzyme activities of some commercially available hyaluronidases and the hyaluronidase according to the invention.

EXPERIMENTAL PART

[0020] The invention will now be disclosed in greater detail in the following experimental part, which illustrates the best mode of carrying out the invention, which is not to be intended in a limiting sense.

MATERIALS AND METHODS

Culture of the microorganism

5 [0021] *S. koganeiensis* was obtained from American Type Culture Collection (ATCC 31394) and cultured as described in [2]. Briefly, the microorganism was grown in 1 litre of culture medium [(20 g/l yeast extract (Organotechnie) and 5 g/l soy peptone (Solabia), pH 6.9)] at 30 °C, shaking at 150 rpm and for about 16 h. Upon growth, the culture was used to inoculate a 50 litre fermentor (Biostat U, B.BRAUN) containing 30 litres of appropriate medium [(10 g/l yeast extract (Organotechnie), 5 g/l soy peptone (Solabia), 3 g/l malt extract (Costantino), 3 g/l dextrin type I (Sigma), 0.2 g/l antifoam (Sigma)]. Before inoculation, the pH was brought to 7.0 with NaOH; during fermentation the pH was monitored, but not controlled, and the temperature was kept at 30 °C throughout fermentation, while shaking was kept at 300 rpm, with aeration of 1.6 VVM (volume of air per volume of culture medium per minute). The fermentation lasted 48 h, a time that corresponded to the highest production of hyaluronidase enzyme activity (1×10^5 - 1.3×10^5 I.U./l) in the culture supernatant.

10 [0022] At the end of the fermentation the culture was centrifuged at 5000 rpm for 30 min at 4 °C (SORVALL Evolution RC) and filtered with 0.2 µm polyethersulfone tangential flow filters, in such a way to eliminate the *Streptomyces koganeiensis* biomass (which occurred in the form of 1-4 mm diameter roundish hyphal aggregates) and obtain a clarified supernatant containing hyaluronidase.

Determination of hyaluronidase activity

20 [0023] Hyaluronidase activity was measured by the modified method of Dorfman [3]. Briefly, the product obtained from DEAE- and CM-cellulose chromatographies was diluted in 0.03 M phosphate buffer, 0.82% NaCl, pH 6.3 and 1 ml of the solution thus obtained was mixed with 1 ml of substrate buffer (0.03 M phosphate buffer, 0.82% NaCl, pH 6.3) containing 0.5 mg hyaluronic acid. Enzymatic digestion was carried out at 37 °C for 30 min and at the end of the incubation process turbidity was generated by adding 4 ml of horse serum based acid solution (SIGMA). The optical density at 640 nm was measured exactly 30 min after adding the horse serum based acid solution. A standard of mammalian testicle hyaluronidase (EDQM, FIP Hyaluronidase, H1115000) containing 328 I.U./mg was used to construct a standard curve and the sample activity (in units) was calculated using this curve.

Chromatographies

30 [0024] The chromatography resins and columns were purchased from GE Healthcare Life Sciences and kept according to the specifications provided by the supplier. The equilibration and elution stages were carried out with a Fast Performance Liquid Chromatography system (FPLC; AKTA explorer 100, GE Healthcare) at a flow of 40-50 ml/min for the first chromatography and 5 ml/min for the following chromatographies. At the end of each chromatography step the hyaluronidase activity was verified with the modified assay of Dorfman described in the previous step.

35 [0025] For purity analyses by gel filtration the LC-10AD HPLC instrument (SHIMADZU) with a Bio-Sil SEC column (BIO-RAD) was used, eluting with 0.05 M NaH_2PO_4 , 0.05 M Na_2HPO_4 , 0.15 M NaCl, pH 6.6, at 1 ml/min. The absorption wavelength used was 214 nm (SPD-10A, SHIMADZU). The protein purity was determined using the LC solution 1.21 SP1 software.

SDS-PAGE electrophoresis

45 [0026] Polyacrylamide gel electrophoresis analyses in the presence of sodium dodecyl sulfate (SDS-PAGE) were carried out using the method of Laemmli [4] on 12% polyacrylamide gel, using a Mini-PROTEAN 3 (BIO-RAD) according to the supplier's instructions. The molecular weight of the purified protein was estimated by comparison with low molecular weight standard proteins (BIO-RAD).

Bidimensional electrophoresis and isoelectric focusing

50 [0027] The protein fraction to be analyzed was mixed in proper loading buffer and loaded on pH 3-10 IPG strips (ReadyStrips 7 cm, BIO-RAD); the strip was incubated at 25 °C until sample absorption and loaded on the PROTEAN IEF Cell (BIO-RAD) for isoelectric focusing (IEF).

55 [0028] At the end of the isoelectric focusing run (first dimension) the strip was equilibrated in proper loading and running buffer, then it was loaded in the second dimension on 12% SDS-PAGE, using Mini-PROTEAN 3 cells (BIO-RAD).

Densitometric analyses

[0029] Polyacrylamide gels properly stained with Silver Stain Plus (BIO-RAD) or Coomassie (BIO-RAD), were acquired with a laboratory imager ImageQuant 300 TL (GE Healthcare), while (quantitative and qualitative) analyses were carried out employing the ImageQuant TL image analysis software (GE Healthcare). Image analyses on 2D SDS-PAGE polyacrylamide gels were instead carried out employing the ImageMaster 2D Platinum 6.0 software (GE Healthcare).

Mass spectrometry

[0030] Mass spectrometry analyses for molecular weight determination were carried out using the Ultraflex III TOF/TOF mass spectrometer (BRUKER) and Bruker Protein Mix 1 markers, while protein identification was carried out using the peptide accurate mass values determined by MALDI-MS Voyager DE-PRO system (Applied Biosystems).

Sequencing of the N-terminal end

[0031] N-Terminal amino acid sequencing was carried out according to the Edman degradation method using a pulsed liquid-phase automated protein sequencer (ABI-Perkin Elmer Mod. 477A). The BLAST software [5] was used to carry out homology searches on the GenBank data bank and that of the genome project of web-available *Streptomyces* species.

Comparison between the enzyme activities of different types of hyaluronidases

[0032] The enzymatic potential of the hyaluronidase according to the invention was evaluated by comparison with the enzyme activities of some of the most used commercially available hyaluronidase (FIP hyaluronidase standard, bovine testicle hyaluronidase type I-S (SIGMA), bovine testicle hyaluronidase type VI-S (SIGMA), sheep testicle hyaluronidase of type V (SIGMA), *Streptomyces hyalurolyticus* hyaluronidase (SIGMA)), using the above described enzyme assay and the activity value was plotted in Figure 9 as I.U./mg (protein concentration was determined by BCA Protein Assay Reagent Kit, PIERCE).

EXAMPLE 1 (REFERENCE EXAMPLE) - OBTAINMENT, PURIFICATION AND CHARACTERIZATION OF STREPTOMYCES KOGANEIENSIS HYALURONIDASE ACCORDING TO US 4,258,134

[0033] *S. koganeiensis* 31394 ATCC was cultured as described in Materials and Methods; the supernatant obtained from centrifugation, properly filtered with tangential flow filters, was subjected to weak anion-exchange chromatography on DEAE-cellulose. Briefly, 1.2 kg of DEAE-cellulose was equilibrated with 25 mM sodium phosphate buffer at pH 7.0 and packed, then the supernatant, clarified in the same buffer, was loaded on the column and eluted with 25 mM sodium phosphate buffer at pH 7.0 containing 250 mM NaCl; after chromatography, the fraction having hyaluronidase activity was collected and concentrated by ultrafiltration, dialysed with 10 volumes of acetate buffer (pH 5.0) and run through weak cation-exchange chromatography using a CM-cellulose column. Elution was performed by 0.005-0.1 M acetate buffer elution gradient. The positive fraction for hyaluronidase activity was collected upon chromatography, concentrated by ultrafiltration and dialysed with 10 volumes of distilled water (MilliQ, Millipore). The product thus obtained was filtered on 0.2 μ m polyethersulfone filters and subjected to assay for determination of hyaluronidase activity, SDS-PAGE, bidimensional electrophoresis and densitometric analysis.

[0034] With regard to bidimensional electrophoresis, whose result is reported in Figure 1, a 600 μ l aliquot of the fraction with hyaluronidase activity obtained from chromatography was concentrated to a volume of about 20 μ l (about 864 U of hyaluronidase) using BIOMAX 5k columns (Millipore). The concentrated aliquot was mixed with 125 μ l of loading buffer (8 M urea, 2% CHAPS, 50 mM dithiothreitol (DTT), 0.2% (w/v) Bio-Lyte 2/10 ampholyte and bromophenol blue) and loaded on pH 3-10 IPG strips (ReadyStrips 7 cm, BIO-RAD), incubating the sample on the strip at 25 °C for 11 h. After 11 h of sample absorption, the strip was loaded on the PROTEAN IEF Cell (BIO-RAD) for isoelectric focusing (IEF).

[0035] At the end of the isoelectric focusing run (first dimension) the strip was first equilibrated for 15 min with a first buffer [(6 M urea, 2% SDS, 0.375 M Tris-HCl (pH 8.8), 20% glycerol, and 2% (w/v) DTT], then with a second buffer [6 M urea, 2% SDS, 0.375 M Tris-HCl (pH 8.8), 20% glycerol]. After equilibration, the strip was loaded in the second dimension on 12% SDS-PAGE, using Mini-PROTEAN 3 cells (BIO-RAD). After the electrophoresis run, the gel was stained with Coomassie PhastGel Blue R and analyzed, after scanning, by imaging software as described in materials and methods (Figure 1).

EXAMPLE 2 - OBTAINMENT, PURIFICATION AND CHARACTERIZATION OF THE *STREPTOMYCES KOGANEIENSIS* HYALURONIDASE ACCORDING TO THE INVENTION**2a) Obtainment and purification****Sample preparation**

[0036] The clarified supernatant (about 30 litres), obtained from fermentation of *Streptomyces koganeiensis* ATCC 31394, as described in Materials and Methods, was concentrated 10 fold by ultrafiltration through 10-kDa cut-off polyethersulfone filters and its hyaluronidase activity was measured. The concentrated supernatant was then dialysed (10 volumes) with 50 mM sodium acetate solution at pH 4.0 and subjected to step a).

Step a) Weak cation-exchange chromatography

[0037] The concentrated and dialysed supernatant was loaded on 200 ml of CM-Sepharose® Fast Flow resin (GE Healthcare), packed in a XK-50 column (GE Healthcare) and equilibrated with 10 bed volumes (bed volumes, BV) of 50 mM sodium acetate buffer at pH 4.0.

[0038] After loading, the column was washed with 3 BV of the same buffer, then bound proteins were eluted with 3 BV of 50 mM sodium acetate buffer solution at pH 4.5. Eluted proteins were collected in a single fraction having volume of about 200 ml and subjected to hyaluronidase activity assay.

Step b) Diafiltration and strong anion-exchange chromatography

[0039] The enzymatically active fraction obtained in step a) was subjected to diafiltration for 10 times with 50 mM Tris-HCl, pH 8, equilibration buffer, after which it was loaded on a HiTrap® Q XL column (5 ml), previously equilibrated with 20 BV of the same buffer. After loading the sample, washing with 20 BV of buffer was carried out, then bound proteins were first eluted with 12 BV of 50 mM Tris-HCl buffer, 35 mM NaCl at pH 8 to remove impurities comprised of inactive proteins, after which the enzymatically active fraction was eluted with 14 BV of 50 mM Tris-HCl buffer, 200 mM NaCl at pH 8 and collected in a final volume of about 50 ml. Equilibration, washing and elution were carried out at 5 ml/min flow.

Step c) Strong cation-exchange chromatography

[0040] The fraction deriving from step b) was diluted 10 fold with 20 mM sodium acetate buffer at pH 4 and loaded on HiTrap® SP FF column, previously equilibrated with 20 BV of the same solution. After a first washing with 20 BV of the same solution, bound proteins were eluted with 10 BV of 50 mM sodium phosphate buffer at pH 4. Eluted proteins were collected in a fraction having a volume of about 45 ml and subjected to hyaluronidase enzyme activity assay. Equilibration, washing and elution were carried out at 5 ml/min flow.

Step d) Strong anion-exchange chromatography

[0041] The enzymatically active fraction obtained in step c) was diluted 10 fold in 20 mM sodium acetate buffer at pH 5.5 and loaded on a Resource Q® column, previously equilibrated with 20 BV of the same buffer. After loading the sample, washing with 20 BV of the same buffer was carried out, then pH was lowered to 5 and elution with 10 BV of solution was carried out in such a way to remove the impurities comprised of inactive proteins. The pH was then further lowered to 4 and elution with 15 BV of buffer was carried out. The second peak eluted at this pH value, having higher absorbance, was collected in a final volume of about 10-15 ml and subjected to ultrafiltration and dialysis with 10 volumes of MilliQ water (Millipore). Equilibration, washing and elution were carried out at 5 ml/min flow.

[0042] All of the eluted protein fractions, either enzymatically active or inactive or little active, were then analyzed by 12% SDS-PAGE as described in Materials and Methods and then stained with silver stain according to the instructions provided by the supplier; in all of the fractions with the highest hyaluronidase activity a more marked protein band at about 25 kDa was present (Figure 3).

2b) Analysis and characterization**HPLC analysis by gel filtration**

[0043] The fraction obtained in step d) was subjected to gel filtration column HPLC as described in Materials and Methods. The result of the analysis is reported in Figure 4.

Mass spectrometry

5 [0044] The fraction obtained in step d) was subjected to 12% SDS-PAGE electrophoresis. At the end of the run the gel was stained with Coomassie Brilliant Blue G-250 (BIO-RAD) and the protein excised from the gel was digested with trypsin (BIO-RAD). A peptide mass pattern was obtained using the MALDI-MS Voyager DE-PRO system (Applied Biosystems). The obtained peptide masses were used for the data bank searches for the protein identification.

N-Terminal sequencing

10 [0045] The fraction obtained in step d) was subjected to SDS-PAGE electrophoresis on 12% gel, as described above, then blotting to polyvinylidene difluoride membrane (BIO-RAD) and stained according to the instructions provided by the supplier. The band was excised with a scalpel, trying to obtain a piece of the smallest possible size (3 mm x 10 mm) and was loaded in the sequencer reaction chamber.

Determination of hyaluronidase activity by non-denaturing SDS-PAGE

15 [0046] The protein samples to be analyzed for hyaluronidase enzyme activity [hyaluronidase and unbound proteins obtained in step d) and hyaluronidase obtained according to US 4,258,134 chromatography] were separated in duplicate on native 8% polyacrylamide gel impregnated with 0.17 mg/ml of hyaluronic acid. After the electrophoresis run, the gel was washed three times with 0.1 M sodium formate, 0.05 M NaCl, pH 4.0, and incubated overnight in the same solution at 37 °C. The gel was washed three times in 3% acetic acid and stained for 2 h at room temperature in 0.5% (w/v) Alcian blue (SIGMA) and 3% acetic acid solution. The gel was then destained in 7% sodium acetate solution for at least 1 h.

20 [0047] Proteins that exhibited hyaluronidase activity were detected as pale bands on the gel blue background [6]. Native-PAGE prepared in the same way, but stained with Coomassie Brilliant Blue G-250 (BIO-RAD) was used as a control (Figure 6).

Passive elution of the protein from native polyacrylamide gel

30 [0048] The protein band with hyaluronidase activity was excised from the native polyacrylamide gel stained with Coomassie Brilliant Blue G-250 and placed within a 1.5 ml sterile tube. 0.5 ml of elution buffer (50 mM Tris-HCl, 150 mM NaCl, and 0.1 mM EDTA; pH 7.5) was added to the excised piece of gel, in such a way that it was completely immersed. The piece of gel was homogenized with a sterile pestle and incubated in an orbital shaker at 30 °C overnight. After incubation, the homogenized gel was centrifuged at 5,000-10,000 x g (5402 centrifuge, eppendorf) for 10 min and the supernatant was very carefully taken out and transferred to a new 1.5 ml tube. After being concentrated 10 fold with BIOMAX 5k column (Millipore), the supernatant was verified for the presence of the eluted protein by SDS-PAGE, and afterwards it was subjected to hyaluronidase activity assay and, as a confirmation, N-terminal sequencing (Figure 5).

Determination of the protein molecular weight by mass spectrometry

40 [0049] 1 μ l hyaluronidase (about 0.5 μ g) was mixed with 1 μ l of a solution comprised of 20 μ g/ μ l sinapinic acid (SA) in 50% acetonitrile with 0.1% trifluoroacetic acid (TFA). The obtained mixture was transferred onto MALDI plate and subjected to analysis as noted in Materials and Methods. The result of the analysis is reported in Figure 7.

Isoelectric focusing

45 [0050] A 20 μ l aliquot (20 μ g) of hyaluronidase obtained in step d), was mixed with 125 μ l of loading buffer [8 M urea, 2% CHAPS, 50 mM dithiothreitol (DTT), 0.2% (w/v) Bio-Lyte 2/10 ampholyte and bromophenol blue] and loaded on pH 3-10 IPG strip (ReadyStrips 7 cm, BIO-RAD), incubating the sample on the strip at 25 °C for 11 h. After 11 h of sample absorption, the strip was loaded for isoelectric focusing (IEF) on the PROTEAN IEF Cell (BIO-RAD). At the end of the isoelectric focusing run, the strip was dried with filter paper (Whatman), moistened with MilliQ water, and stained for 45 min using IEF Gel Staining solution (BIO-RAD). The strip was destained for 1 h or longer with destaining solution (Destain solution, Coomassie R-250, BIO-RAD). The sample isoelectric point was determined by comparison with the reference standards isoelectric points (IEF Marker pH 3-10, SERVA). The result of the analysis is reported in Figure 8.

Comparison between the enzyme activities of different types of hyaluronidases

55 [0051] The result of this assay demonstrates that the hyaluronidase according to the invention has an activity about three times higher than the most active among those used for the comparison (Figure 9).

Sequencing

[0052] Sequencing of the N-terminal end, which was carried out as described in materials and methods, allowed to establish that it contains the following amino acid sequence:

Ala-Gly-Glu-Asn-Gly-Ala-Thr-Thr-Thr-Phe-Asp-Gly-Pro-Val-Ala (SEQ ID No. 1)

EXAMPLE 3 - PHARMACEUTICAL PREPARATIONS**Preparation 1 - Hydrophilic gel****[0053]**

Components	Amounts (I.U. or mg/1 g of hydrogel)
Hyaluronidase	150 I.U.
Carbomer 974P	15 mg
Glycerol	100 mg
Propylene glycol	66.75 mg
Triethanolamine (TEA)	13.25 mg
Polyethylene glycol 400	66.75 mg
Methyl p-hydroxybenzoate	2 mg
Propyl p-hydroxybenzoate	0.2 mg
Purified water	q.s. to 1 g

[0054] Methyl- and propyl-paraben were dissolved in purified water at 80 °C. After the solution cooled down to room temperature, hyaluronidase was added, shaking until completely dissolved, after which PEG 400 was added, continuing to shake until dissolved. To this solution Carbomer® 974P was added, continuing to shake until homogeneous dispersion and complete hydration thereof, then TEA was added to obtain the aqueous phase gelation. Finally, always under shaking, glycerol and propylene glycol were added.

Preparation 2 - Hydrophilic cream (o/w emulsion)**[0055]**

Components	Amounts (I.U. or mg/1 g of cream)
Hyaluronidase	150 I.U.
Tefose 1500	110 mg
Glycerol	80 mg
Stearic acid	33 mg
Liquid paraffin	40 mg
Methyl p-hydroxybenzoate	1 mg
Purified water	q.s. to 1 g

[0056] For the preparation of the oily phase, liquid paraffin, stearic acid and Tefose® 1500 were melted under shaking at 50 °C. Separately, the aqueous phase was prepared by initial solution of methyl-paraben at 80 °C, subsequent cooling to room temperature and incorporation of glycerol and hyaluronidase under shaking until completely dissolved.

[0057] The aqueous phase was added to the oily phase, proceeding with emulsification, after which the obtained o/w emulsion was cooled under shaking to room temperature.

Preparation 3 - Ointment**[0058]**

5

Components	Amounts (I.U. or mg/1 g of ointment)
Hyaluronidase	150 I.U.
Light liquid paraffin	200 mg
White vaseline	q.s. to 1 g

10

[0059] The ointment base was prepared by melting light liquid paraffin and white vaseline under shaking at 70 °C. After cooling to room temperature, hyaluronidase was incorporated, mixing until obtaining a homogeneous suspension.

15

Preparation 4 - Lipogel**[0060]**

20

Components	Amounts (I.U. or mg/1 g of lipogel)
Hyaluronidase	150 I.U.
Hydrogenated castor oil	10 mg
Cetostearyl alcohol	50 mg
white vaseline	365 mg
Light liquid paraffin	q.s. to 1 g

25

30

[0061] Light liquid paraffin, white vaseline and the cetylstearyl alcohol were melted under shaking at 90 °C, after which, under shaking, hydrogenated castor oil (lipogelation agent) was added until homogeneous solution. After slowly cooling to room temperature, hyaluronidase was incorporated, mixing until obtaining a homogeneous suspension.

Preparation 5 - Injectable solutions for intramuscular or subcutaneous use

35

[0062]

Components	Amounts (I.U. or mg/ml of sol.)
Hyaluronidase	200 I.U.
Lactose	0.93
Potassium Phosphate Dibasic	0.36
Potassium Phosphate Monobasic	0.23
Sodium chloride	9

40

45

[0063] Lactose and hyaluronidase were dissolved, under shaking, in buffered saline at pH 6.4 - 7.2, prepared at room temperature and the solution thus obtained was filtered on 0.22-micron filters.

50

References**[0064]**

55

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SEQUENCE LISTING

10 **[0065]**

<110> Fidia Farmaceutici SpA

<120> IALURONIDASI EXTRACELLULARE DA STREPTOMYCES KOGANEIENSIS

15

<130> P03279IT

<160> 1

20

<170> PatentIn version 3.3

<210> 1

<211> 15

<212> PRT

25

<213> Streptomyces koganeiensis ATCC 31394

<400> 1

30

Ala Gly Glu Asn Gly Ala Thr Thr Thr Phe Asp Gly Pro Val Ala
1 5 10 15

Claims

35

1. Hyaluronidase from *Streptomyces koganeiensis* ATCC 31394 comprising the N-terminal amino acid sequence shown in SEQ ID No. 1 and having a molecular weight of 21,6 kDa, an isoelectric point (pI) ranging from 4.4 to 4.8 and an enzymatic activity equal to or higher than 40,000 I.U./mg.

40

2. A process for the preparation of the hyaluronidase according to claim 1 comprising the following steps:

- a) submitting the supernatant obtained from the fermentation of *Streptomyces koganeiensis* ATCC 31394 to weak cation-exchange chromatography and isolating the protein fraction with hyaluronidase activity;
- b) submitting the protein fraction with hyaluronidase activity from step a) to diafiltration and strong anion-exchange chromatography and isolating the protein fraction with hyaluronidase activity;
- c) submitting the protein fraction with hyaluronidase activity from step b) to strong cation-exchange chromatography and isolating the protein fraction with hyaluronidase activity;
- d) submitting the protein fraction with hyaluronidase activity from step c) to strong anion-exchange chromatography and isolating the protein fraction with hyaluronidase activity.

50

3. The hyaluronidase according to claim 1 for use as a medicament.

4. The hyaluronidase according to claim 3 for use in the treatment of diseases in which degradation of hyaluronic acid in the affected tissues or organs is advantageous or desirable.

55

5. The hyaluronidase for the use according to claim 4 wherein the disease is selected from edema, inflammation, chilblain, solid tumors, IgE-mediated allergies, oral diseases and spontaneous vitreous haemorrhages.

6. The hyaluronidase for the use according to claim 4 wherein the disease is bovine mastitis.
7. Pharmaceutical or veterinary compositions containing the hyaluronidase of claim 1 in admixture with suitable excipients and/or carriers.
8. The compositions according to claim 7 in the form of injectable preparations or of topical preparations for epidermal, transdermal or ophthalmic application.
9. The compositions according to claim 7 or 8 further containing one or more active principles selected from: steroidal and non-steroidal antiinflammatories, antitumor agents, allergens, local anesthetics, antibiotics, monoclonal antibodies, cytokines, enzymes and sulphated hyaluronic acid.
10. The compositions according to claim 9) in which the active principle is sulphated hyaluronic acid and the topical composition for epidermal application is in the form of cream, gel, ointment or spray.
11. Use of the hyaluronidase according to claim 1 as a reagent in biochemical assays for the quali/quantitative determination of hyaluronic acid.

Patentansprüche

1. Hyaluronidase von *Streptomyces koganei* ATCC 31394, umfassend die N-terminale Aminosäuresequenz, die in SEQ ID NR. 1 gezeigt ist, und mit einem Molekulargewicht von 21,6 kDa, einem isoelektrischen Punkt (pI) im Bereich von 4,4 bis 4,8 und einer enzymatischen Aktivität, die mindestens 40.000 IE/mg entspricht.
2. Verfahren zur Herstellung der Hyaluronidase nach Anspruch 1, umfassend die folgenden Schritte:
- a) Unterziehen des aus der Fermentation von *Streptomyces koganei* ATCC 31394 erhaltenen Überstands einer schwachen Kationenaustauschchromatographie und Isolieren der Proteinfraktion mit Hyaluronidaseaktivität;
 - b) Unterziehen der Proteinfraktion mit Hyaluronidaseaktivität aus Schritt a) einer Diafiltration und einer starken Anionenaustauschchromatographie und Isolieren der Proteinfraktion mit Hyaluronidaseaktivität;
 - c) Unterziehen der Proteinfraktion mit Hyaluronidaseaktivität aus Schritt b) einer starken Kationenaustauschchromatographie und Isolieren der Proteinfraktion mit Hyaluronidaseaktivität;
 - d) Unterziehen der Proteinfraktion mit Hyaluronidaseaktivität aus Schritt c) einer starken Anionenaustauschchromatographie und Isolieren der Proteinfraktion mit Hyaluronidaseaktivität.
3. Hyaluronidase nach Anspruch 1 zur Verwendung als Medikament.
4. Hyaluronidase nach Anspruch 3 zur Verwendung bei der Behandlung von Krankheiten, bei denen der Abbau von Hyaluronsäure in den betroffenen Geweben oder Organen vorteilhaft oder wünschenswert ist.
5. Hyaluronidase zur Verwendung nach Anspruch 4, wobei die Krankheit aus Ödem, Entzündung, Frostbeulen, festen Tumoren, IgE-vermittelten Allergien, oralen Krankheiten und spontanen Glaskörperblutungen ausgewählt ist.
6. Hyaluronidase zur Verwendung nach Anspruch 4, wobei es sich bei der Krankheit um Mastitis beim Rind handelt.
7. Pharmazeutische oder tiermedizinische Zusammensetzungen, welche die Hyaluronidase nach Anspruch 1 in Mischung mit geeigneten sonstigen Bestandteilen und/oder Trägern enthalten.
8. Zusammensetzungen nach Anspruch 7 in der Form von injizierbaren Zubereitungen oder topischer Zubereitungen für die epidermale, transdermale oder ophthalmische Verabreichung.
9. Zusammensetzungen nach Anspruch 7 oder 8, ferner enthaltend mindestens ein Wirkprinzip, ausgewählt aus: steroidal und nicht-steroidal entzündungshemmenden Mitteln, Antitumormitteln, Allergenen, Lokalanästhetika, Antikörpern, monoklonalen Antikörpern, Zytokinen, Enzymen und sulfierter Hyaluronsäure.
10. Zusammensetzungen nach Anspruch 9, bei denen es sich bei dem Wirkprinzip um sulfierte Hyaluronsäure handelt

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und die topische Zusammensetzung für die epidermale Verabreichung in der Form einer Creme, eines Gels, einer Salbe oder eines Sprays vorliegt.

- 5 11. Verwendung der Hyaluronidase nach Anspruch 1 als Reagenz in biochemischen Assays zur quali-/quantitativen Bestimmung von Hyaluronsäure.

Revendications

- 10 1. Hyaluronidase de *Streptomyces koganeiensis* ATCC 31394 comprenant la séquence d'acides aminés N-terminale présentée dans SEQ ID NO : 1 et ayant une masse moléculaire de 21,6 kDa, un point isoélectrique (pI) dans la plage de 4,4 à 4,8 et une activité enzymatique supérieure ou égale à 40 000 U.I./mg.

- 15 2. Procédé de préparation de la hyaluronidase selon la revendication 1, ledit procédé comprenant les étapes suivantes :

a) soumission du surnageant obtenu de la fermentation de *Streptomyces koganeiensis* ATCC 31394 à une chromatographie d'échange de cations faible puis isolement de la fraction protéique ayant une activité hyaluronidase ;

20 b) soumission de la fraction protéique ayant une activité hyaluronidase de l'étape a) à une diafiltration et à une chromatographie d'échange d'anions forte puis isolement de la fraction protéique ayant une activité hyaluronidase ;

c) soumission de la fraction protéique ayant une activité hyaluronidase de l'étape b) à une chromatographie d'échange de cations forte puis isolement de la fraction protéique ayant une activité hyaluronidase ;

25 d) soumission de la fraction protéique ayant une activité hyaluronidase de l'étape c) à une chromatographie d'échange d'anions forte puis isolement de la fraction protéique ayant une activité hyaluronidase.

3. Hyaluronidase la revendication 1, pour son utilisation en tant que médicament.

- 30 4. Hyaluronidase la revendication 3, pour son utilisation dans le traitement de maladies dans lesquelles la dégradation de l'acide hyaluronique dans les tissus ou les organes affectés est avantageuse ou souhaitable.

- 35 5. Hyaluronidase pour son utilisation selon la revendication 4, dans laquelle la maladie est sélectionnée parmi un oedème, une inflammation, une engelure, des tumeurs solides, des allergies médiées par les IgE, des maladies orales et des hémorragies spontanées du vitré.

6. Hyaluronidase pour son utilisation selon la revendication 4, dans laquelle la maladie est la mastite bovine.

- 40 7. Compositions pharmaceutiques ou vétérinaires contenant la hyaluronidase selon la revendication 1 en mélange avec des excipients et/ou des véhicules adaptés.

8. Compositions selon la revendication 7, sous la forme de préparations injectables ou de préparations topiques pour une application épidermique, transdermique ou ophtalmique.

- 45 9. Compositions selon la revendication 7 ou la revendication 8, contenant en outre un ou plusieurs principes actifs sélectionnés parmi : les anti-inflammatoires stéroïdiens et non stéroïdiens, les agents antitumoraux, les allergènes, les anesthésiques locaux, les antibiotiques, les anticorps monoclonaux, les cytokines, les enzymes et l'acide hyaluronique sulfaté.

- 50 10. Compositions selon la revendication 9, dans lesquelles le principe actif est l'acide hyaluronique sulfaté et la composition topique pour application épidermique se trouve sous la forme d'une crème, d'un gel, d'un onguent ou d'une pulvérisation.

- 55 11. Utilisation de la hyaluronidase la revendication 1, en tant que réactif dans des analyses biochimiques pour la détermination quali/quantitative de l'acide hyaluronique.

FIG. 1

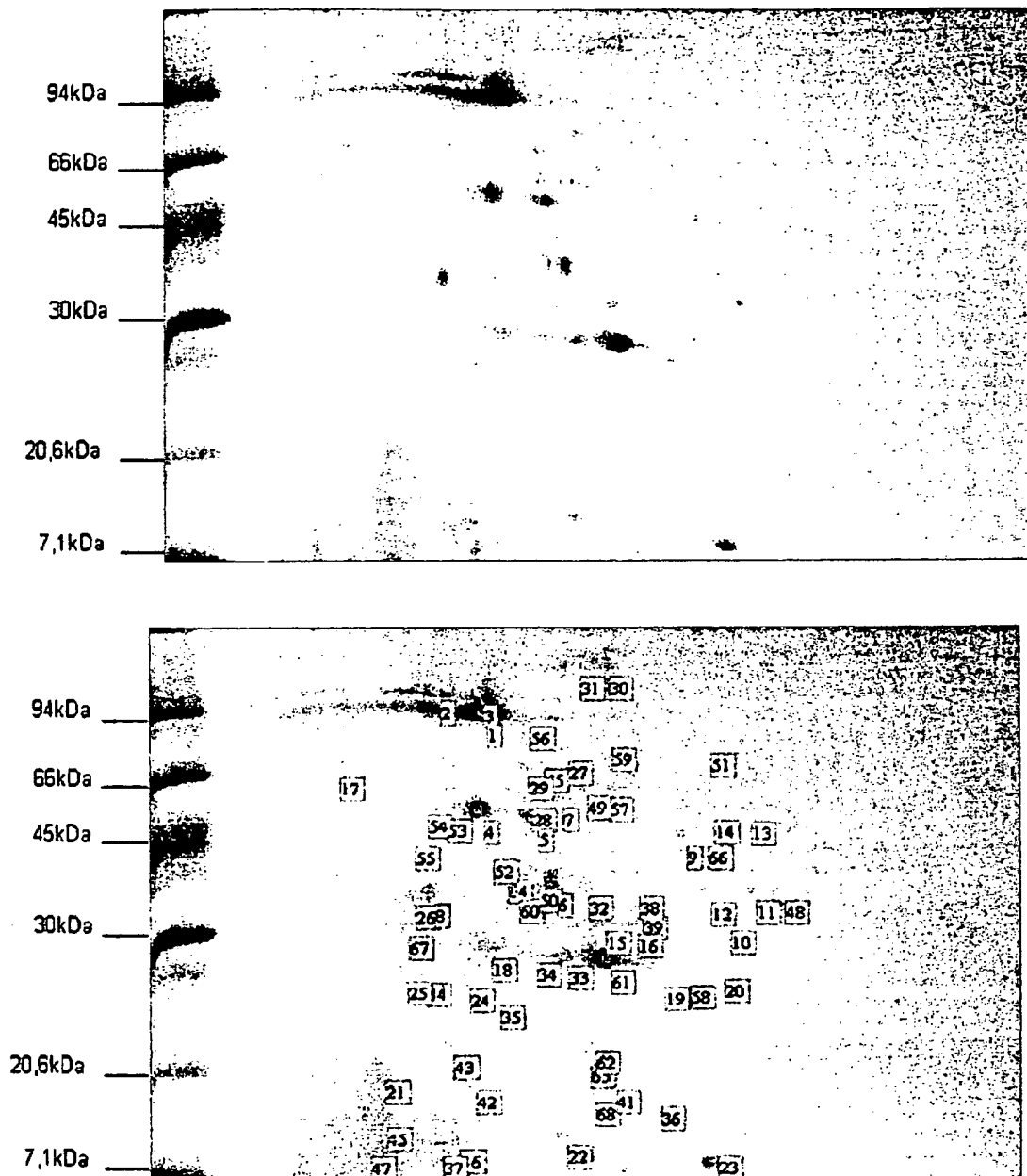


FIG. 2

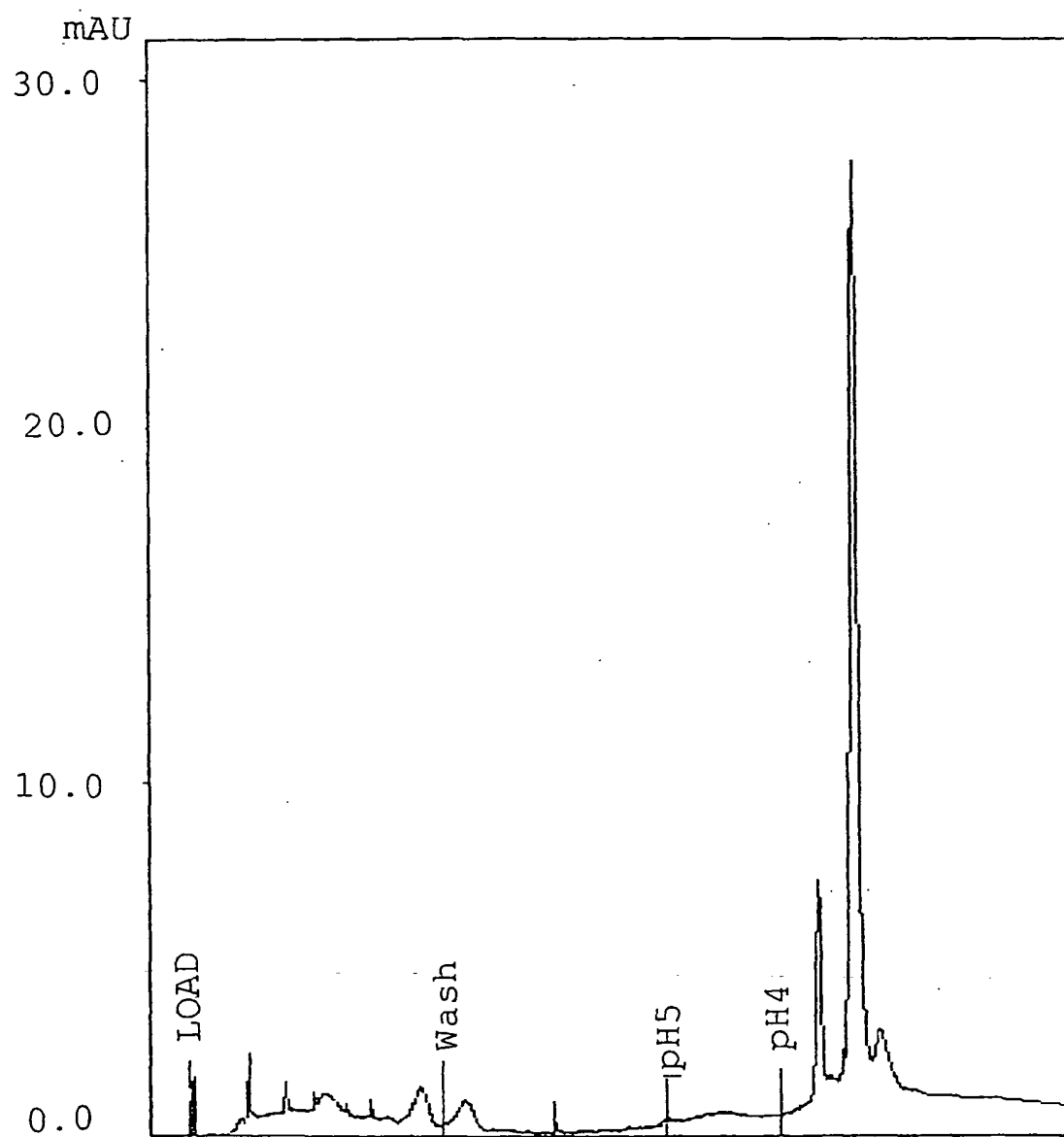


FIG. 3

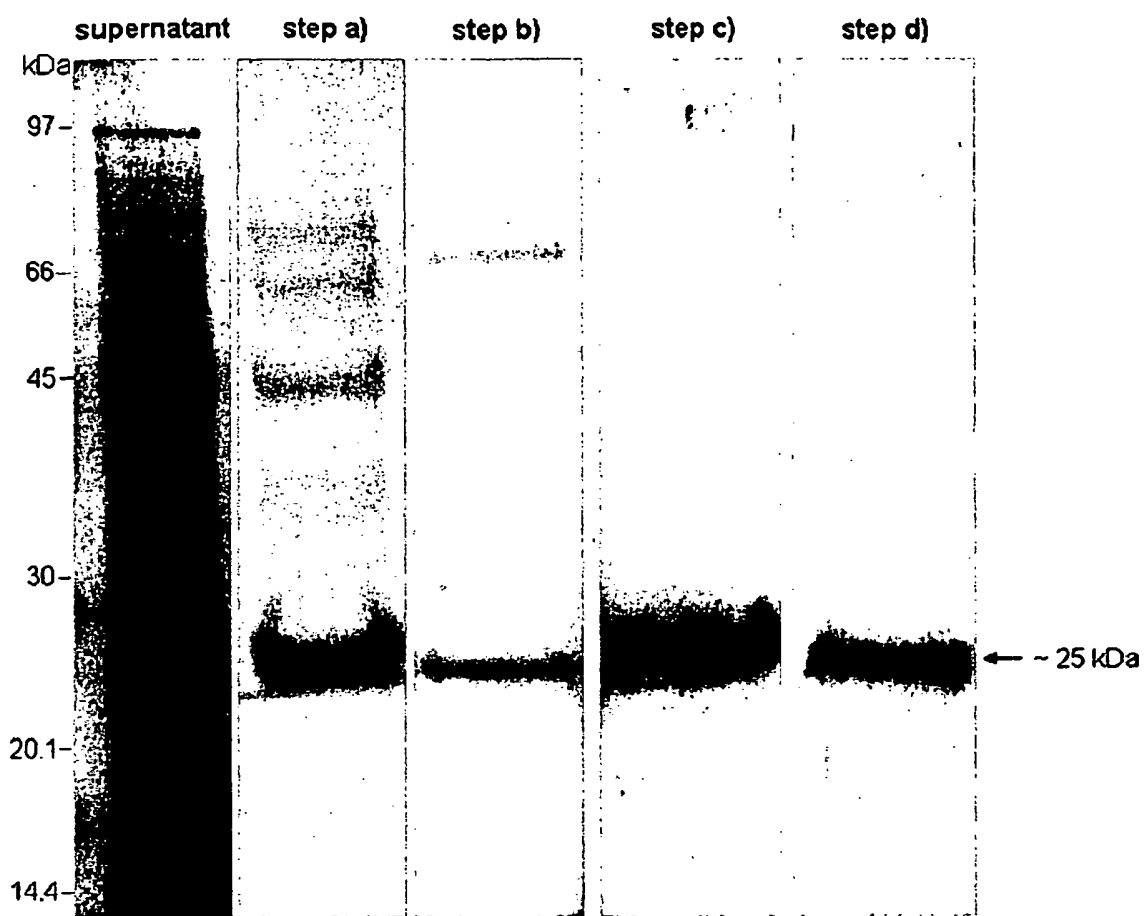


FIG. 4

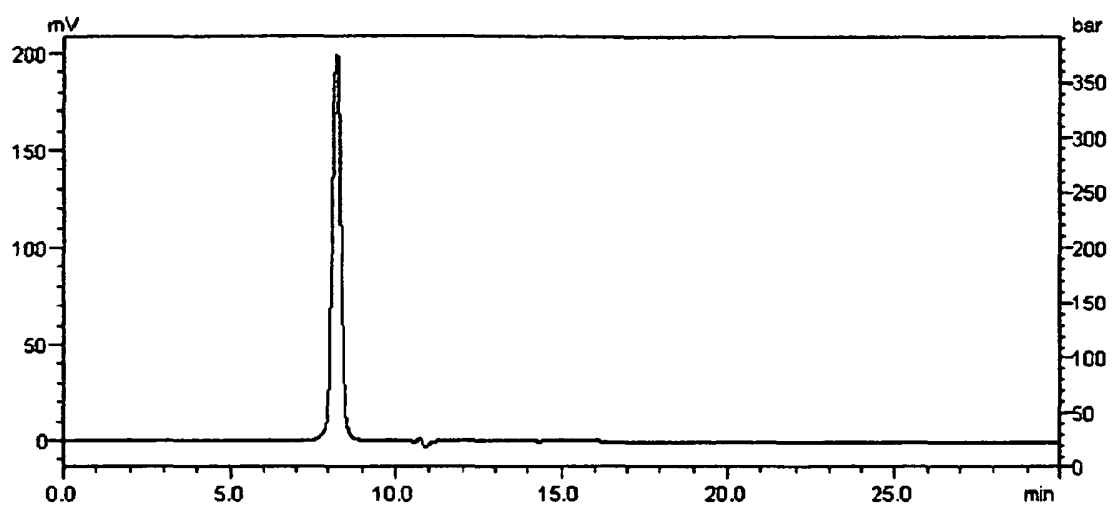
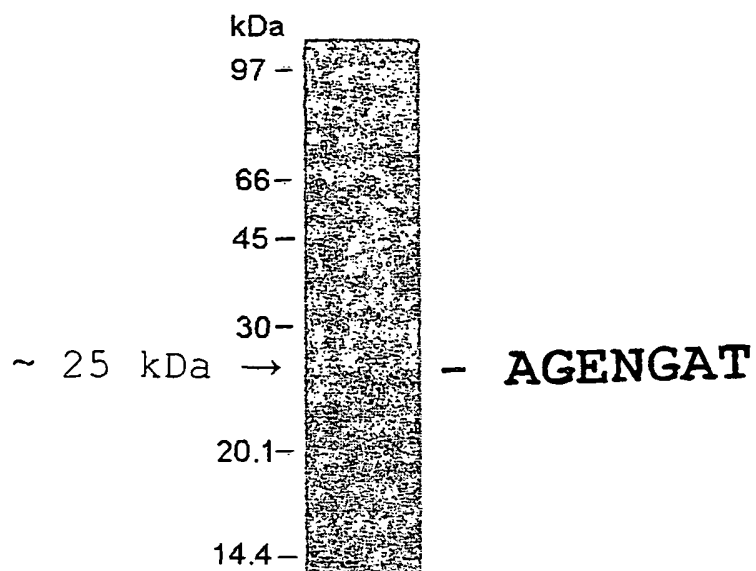


FIG. 5a



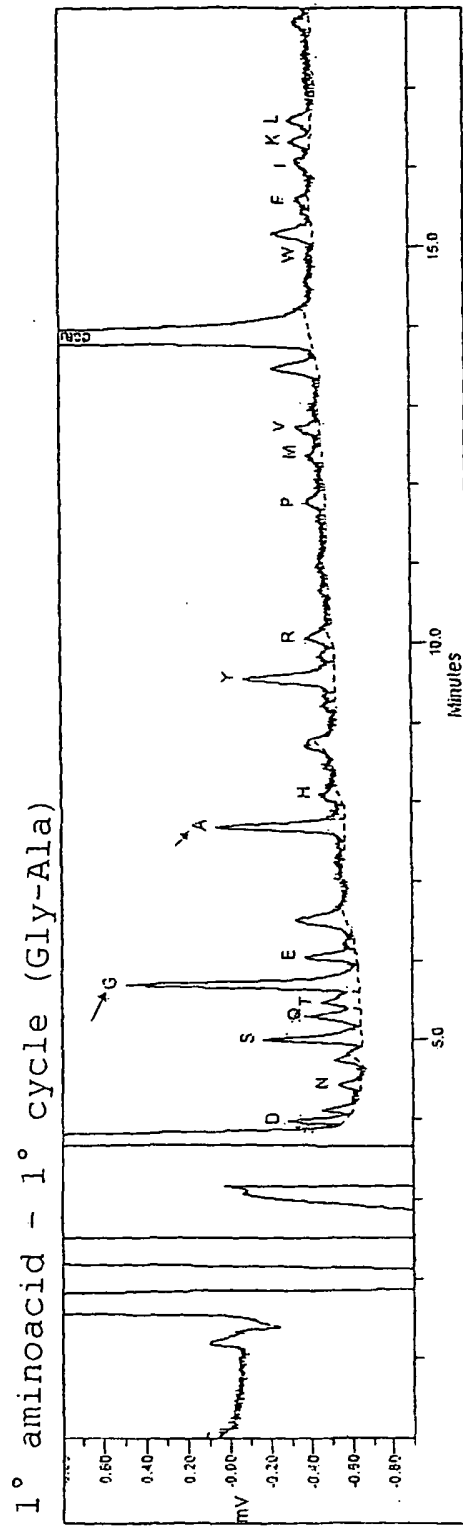


Fig. 5b

Fig. 5c

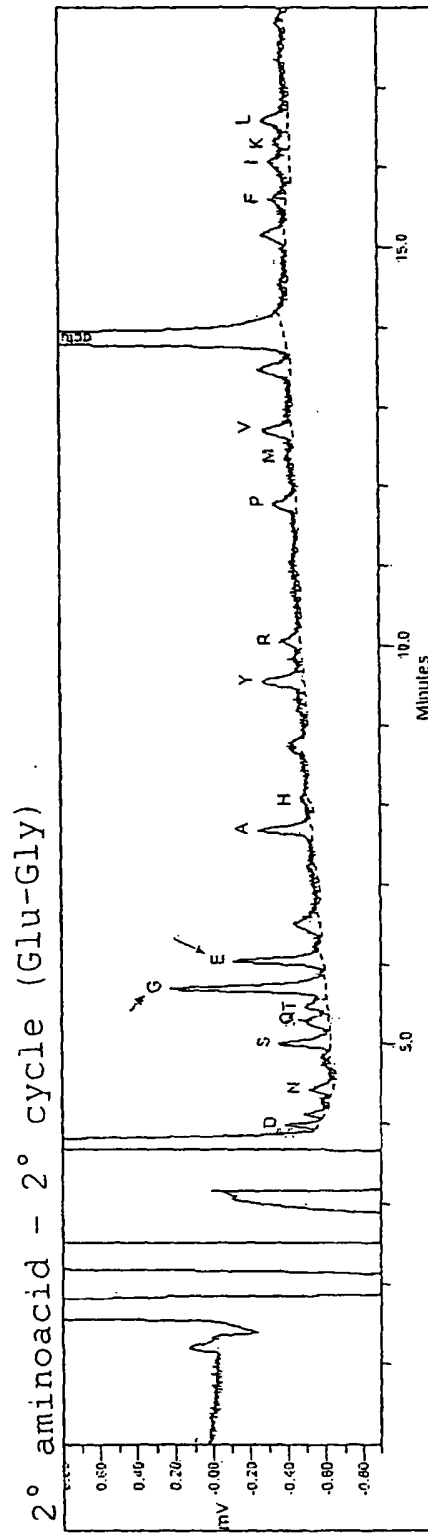
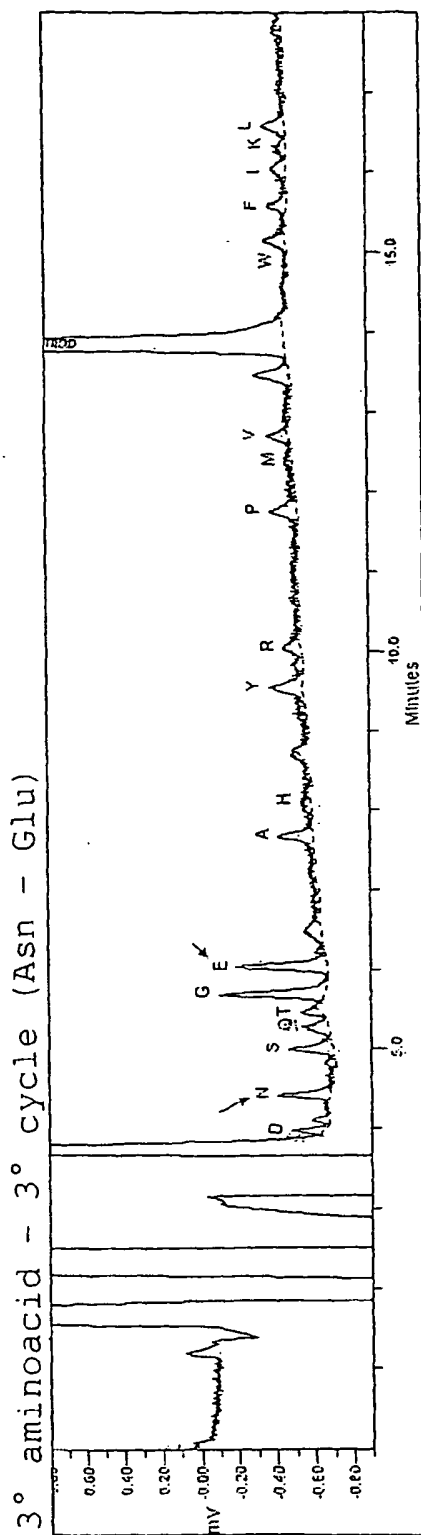


Fig. 5d



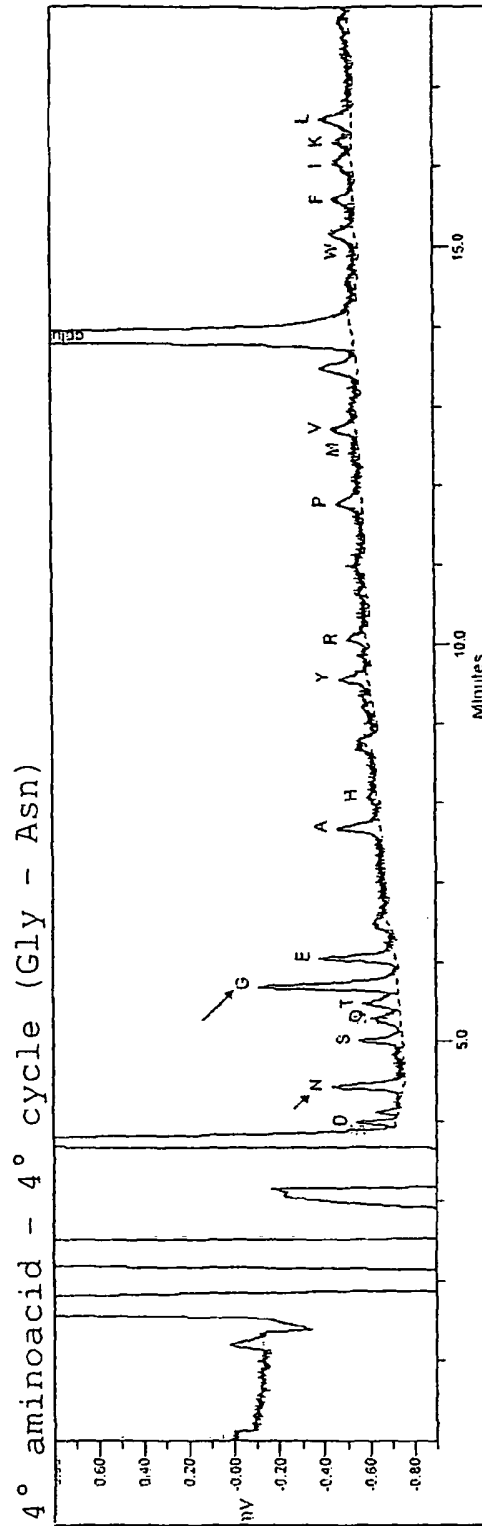
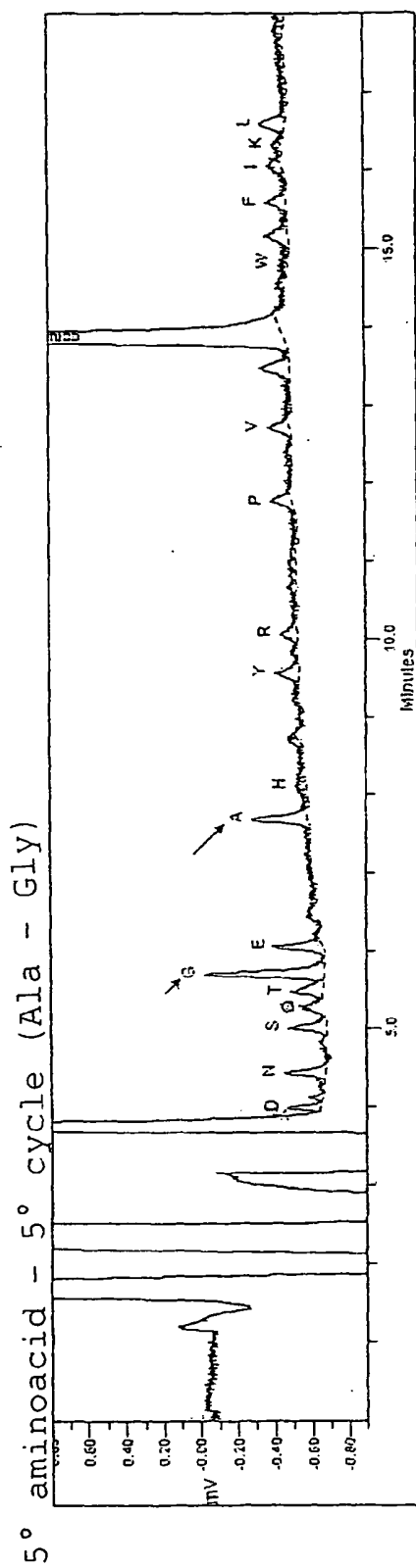


Fig. 5e

Fig. 5f



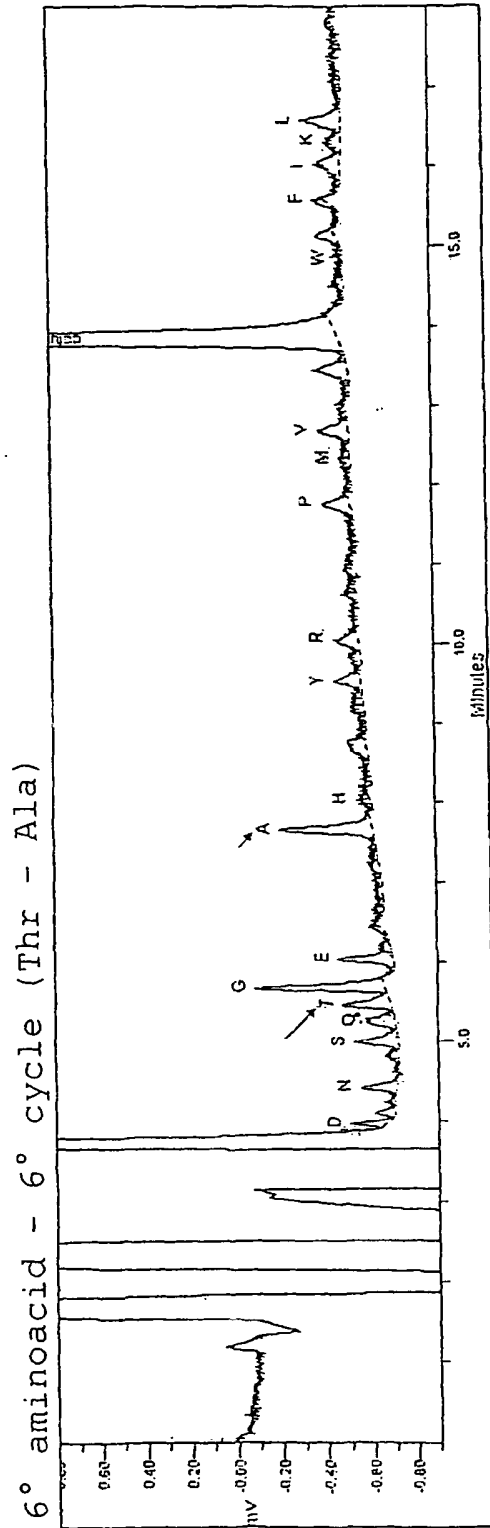


Fig. 59

Fig. 5h

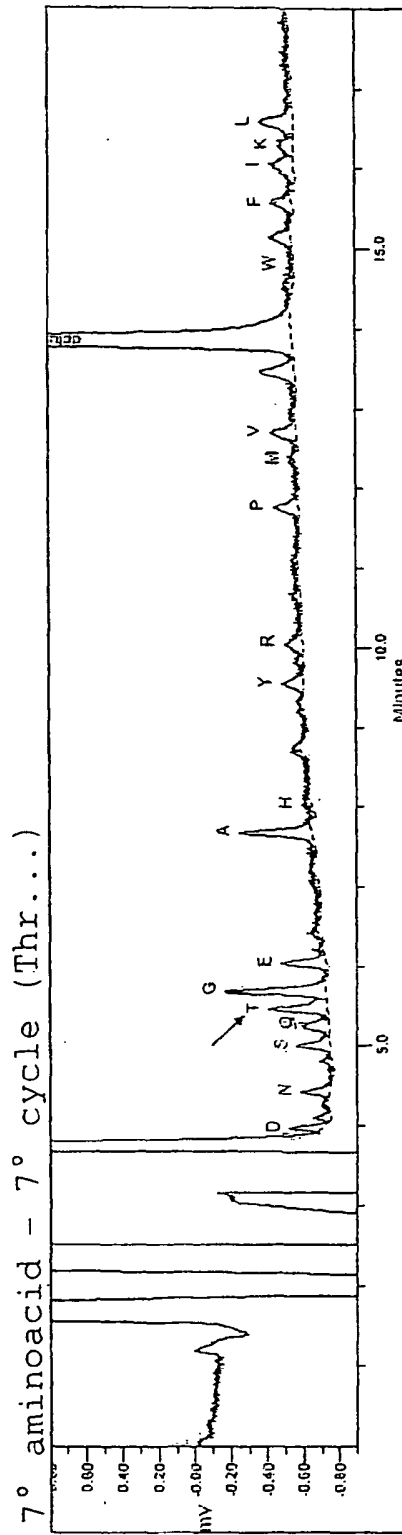


FIG. 6

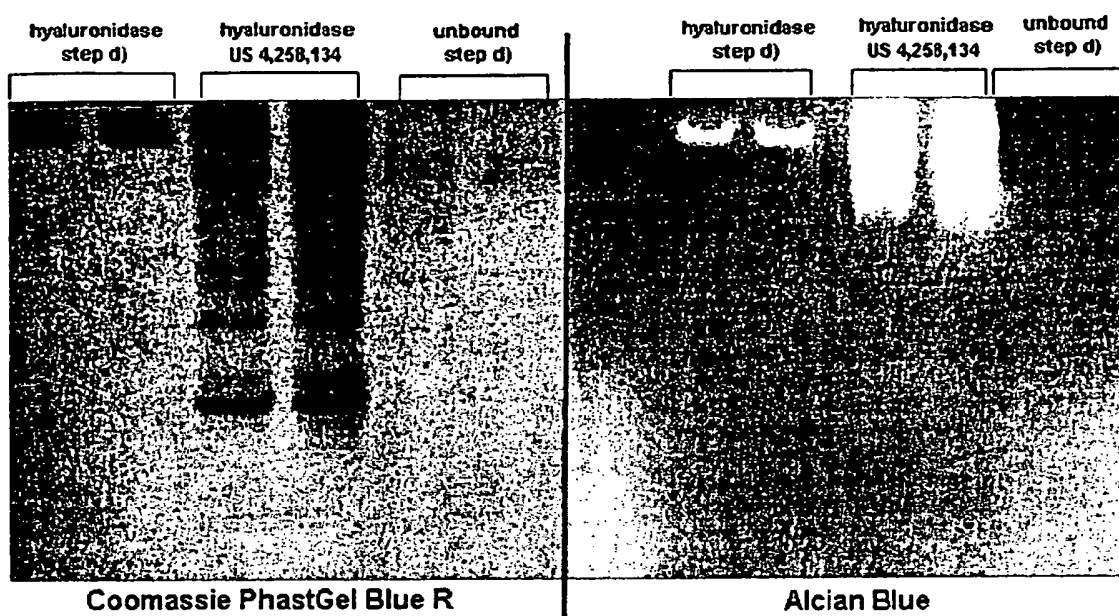


FIG. 7

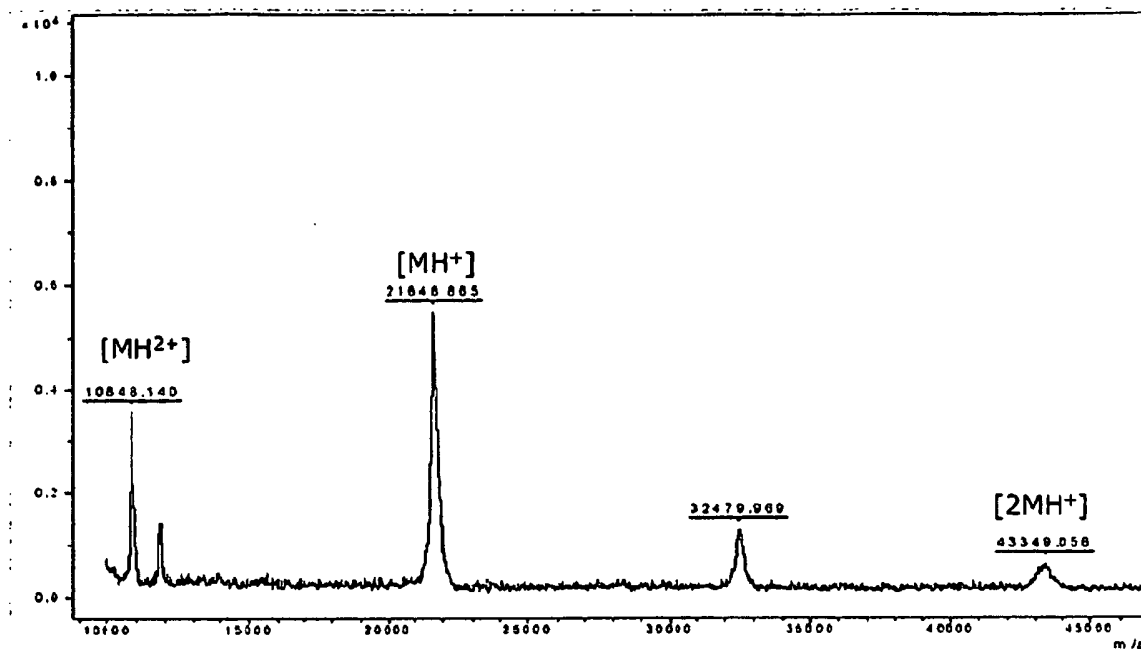


FIG. 8

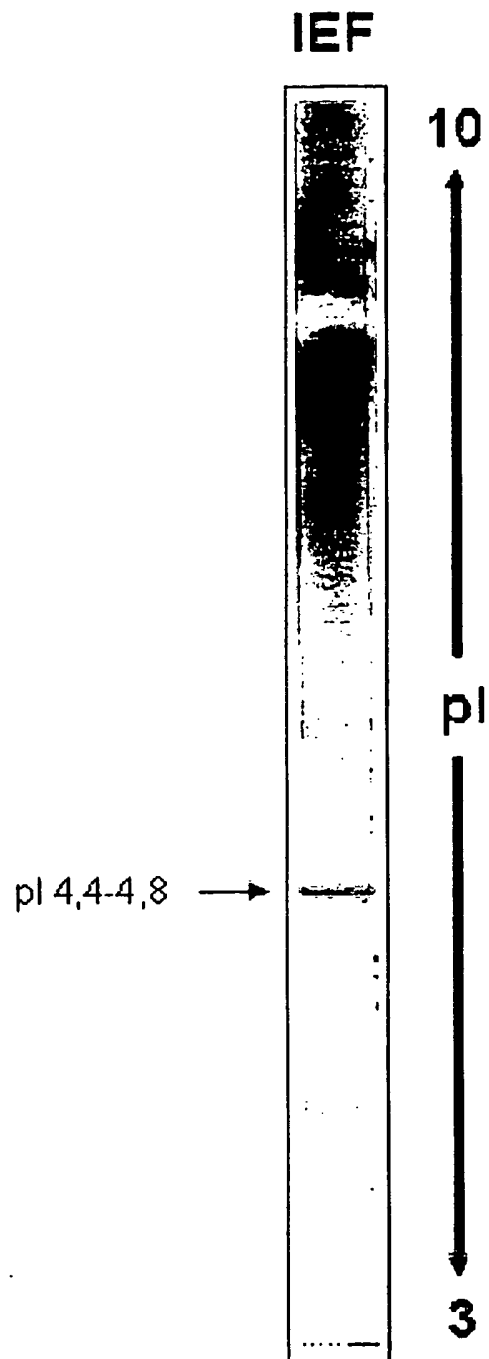
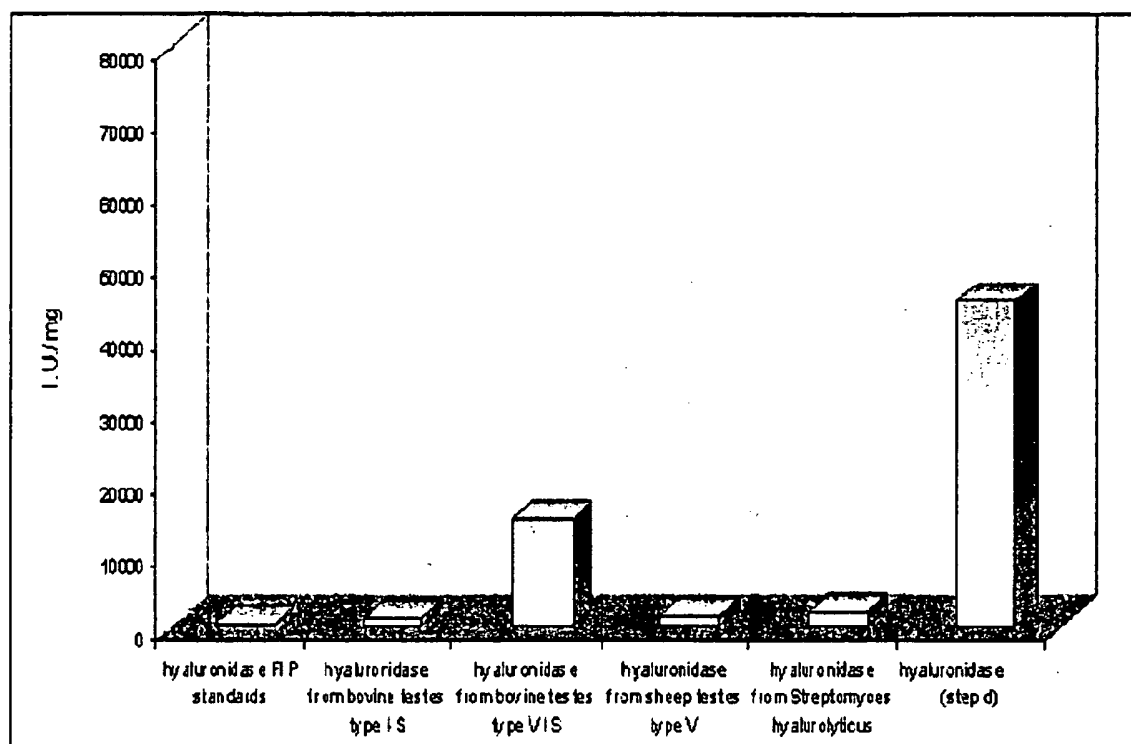


FIG. 9



REFERENCES CITED IN THE DESCRIPTION

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Extracelluláris *Streptomyces koganeiensis* hialuronidáz

Szabadalmi igénypontok

1. Extracelluláris *Streptomyces koganeiensis* ATCC 31394 hialuronidáz, mely az 1. számú N-terminális aminosavszekvenciát tartalmazza, és molekulatömege 21,6 kDa, izoelektromos pontja (pI) 4.4 - 4.8 és enzimaktivitása 40,000 I.U./mg vagy annál nagyobb.
2. Eljárás az 1. igénypont szerinti hialuronidáz előállítására, melynek során
 - a) *Streptomyces koganeiensis* ATCC 31394 fermentációja során nyert felülúszót enyhe kationcserélő kromatográfiának vetjük alá, és izoláljuk a hialuronidáz-aktivitású protein-frakciókat;
 - b) az a) lépésben nyert hialuronidáz-aktivitású protein-frakciókat erős anioncserélő kromatográfiának vetjük alá, és izoláljuk a hialuronidáz-aktivitású protein-frakciókat;
 - c) a b) lépésben nyert hialuronidáz-aktivitású protein-frakciókat erős kationcserélő kromatográfiának vetjük alá, és izoláljuk a hialuronidáz-aktivitású protein-frakciókat;
 - d) a c) lépésben nyert hialuronidáz-aktivitású protein-frakciókat erős anioncserélő kromatográfiának vetjük alá, és izoláljuk a hialuronidáz-aktivitású protein-frakciókat.
3. Az 1. igénypont szerinti hialuronidáz gyógyszerként történő alkalmazásra.
4. A 3. igénypont szerinti hialuronidáz olyan betegségek kezelésében történő alkalmazásra, ahol a hialuronsavnak az érintett szövetekben vagy szervekben történő lebontása előnyös vagy kívánatos.
5. Hialuronidáz a 4. igénypont szerinti alkalmazásra, ahol a betegség ödéma, gyulladás, fagyás, szilárd tumor, IgE által közvetített allergia, orális betegség és spontán üvegtesti vérzés.
6. Hialuronidáz a 4. igénypont szerinti alkalmazásra, ahol a betegség bovin mastitis.
7. Gyógyászati vagy állatgyógyászati készítmények, melyek az 1. igénypont szerinti hialuronidázt tartalmazzák, megfelelő adalékanyagokkal és/vagy hordozóanyagokkal keverve.

8. A 7. igénypont szerinti készítmények injektálható készítmény vagy epidermális, transzdermális vagy szemem keresztül adagolható topikális készítmény formában.

9. A 7. vagy 8. igénypont szerinti készítmények, melyek tartalmaznak továbbá szteroid vagy nem-szteroid típusú gyulladásgátlók, tumorelleses szerek, allergének, helyi érzéstelenítők, antibiotikumok, monoklonális antitestek, citokinek, enzimek és szulfatált hialuronsav közül választott egy vagy több hatóanyagot is.

10. A 9. igénypont szerinti készítmények, ahol a hatóanyag szulfatált hialuronsav és az epidermális adagolásra alkalmas topikális készítmény krém, gél, kenőcs vagy spray formájú.

11. Az 1. igénypont szerinti hialuronidáz alkalmazása biokémiai esszék reagenseként hialuronsav kvalitatív/kvantitatív meghatározására.

