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TUSCHL et al.(10) **Pub. No.: US 2012/0246747 A1**(43) **Pub. Date: Sep. 27, 2012**(54) **RNA-INTERFERENCE BY
SINGLE-STRANDED RNA MOLECULES**(75) Inventors: **Thomas TUSCHL**, New York, NY
(US); **Javier Martinez**, New York,
NY (US); **Agnieszka**
Patkaniowska, New York, NY
(US); **Henning Urlaub**, Goettingen
(DE); **Reinhard Lührmann**,
Marburg-Michelbach (DE)(73) Assignee: **Max-Planck-Gesellschaft zur**
Foerderung der Wissenschaften
e.V., Muenchen (DE)(21) Appl. No.: **13/329,710**(22) Filed: **Dec. 19, 2011****Related U.S. Application Data**(62) Division of application No. 10/520,470, filed on Jan. 7,
2005, now Pat. No. 8,101,348, filed as application No.
PCT/EP2003/007516 on Jul. 10, 2003.(30) **Foreign Application Priority Data**

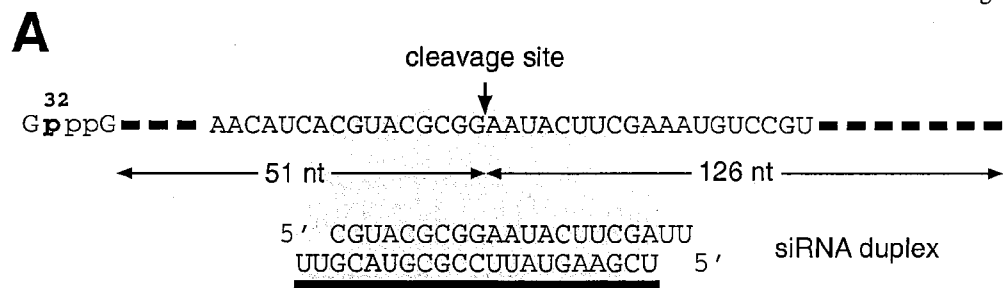
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A01K 67/027 (2006.01)
C12N 5/10 (2006.01)(52) **U.S. Cl.** **800/13; 435/325; 435/375; 536/24.5**(57) **ABSTRACT**

The present invention relates to sequence and structural features of single-stranded (ss)RNA molecules required to mediate target-specific nucleic acid modifications by RNA-interference (RNAi), such as target mRNA degradation and/or DNA methylation.

Figure 1



B

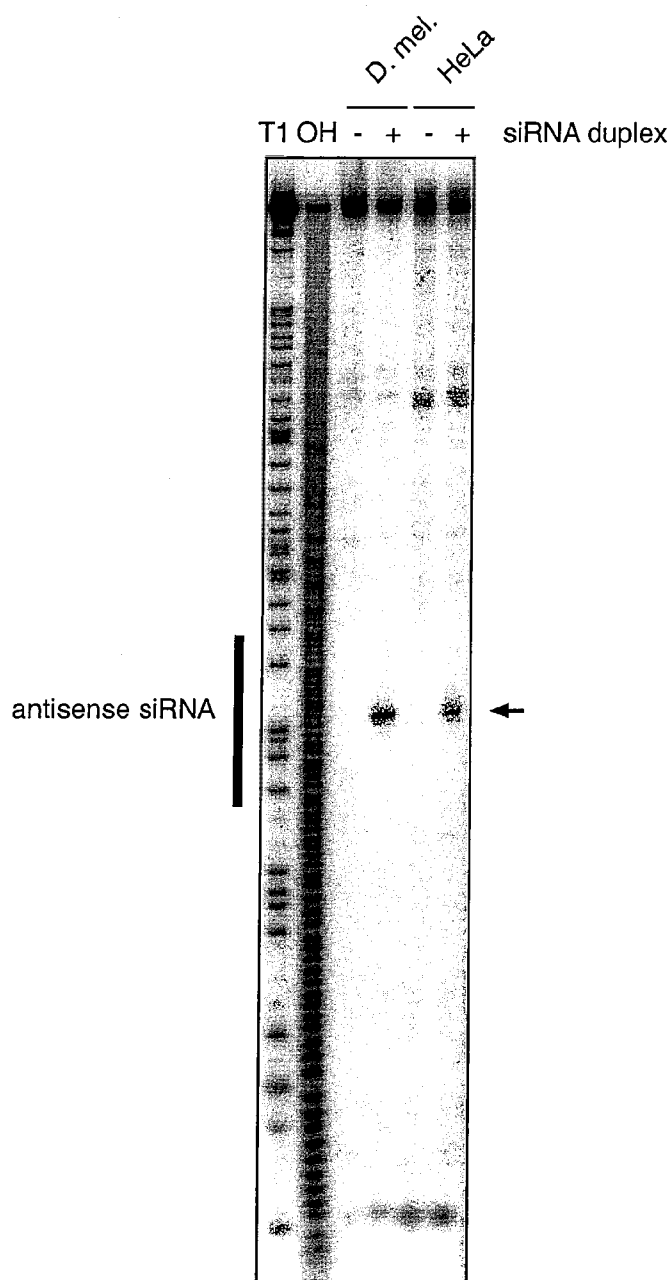


Figure 2

A



B

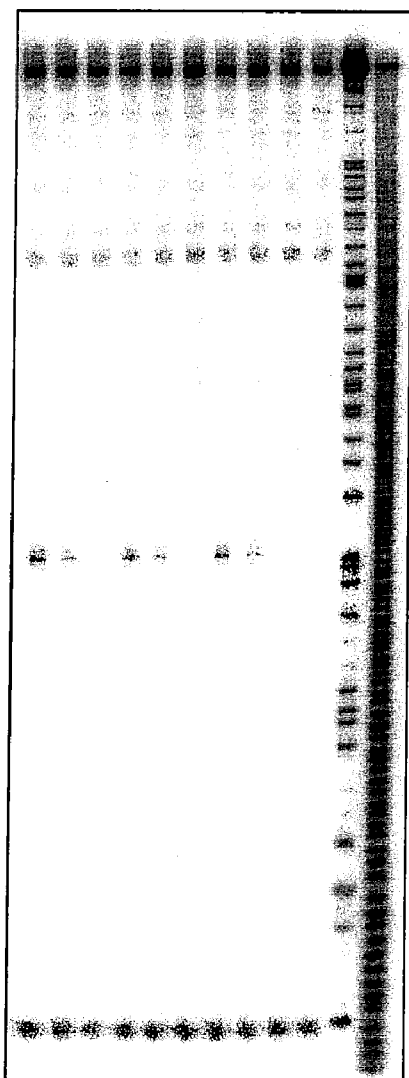
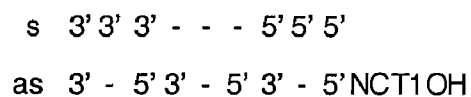


Figure 4

				0		15					
time point of competitor siRNA addition (min)				—		—					
siRNA duplexes	competitor (nM)	100	0	0	1000	10	1000				
	specific (nM)	0	100	← 10 →							
HeLa S100		T1	-	-	+	+	+	+	+	+	+

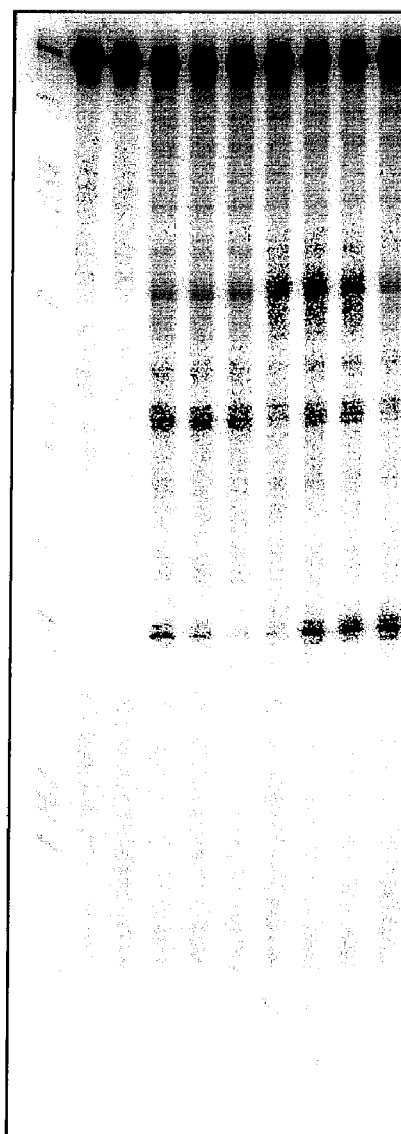


Figure 5

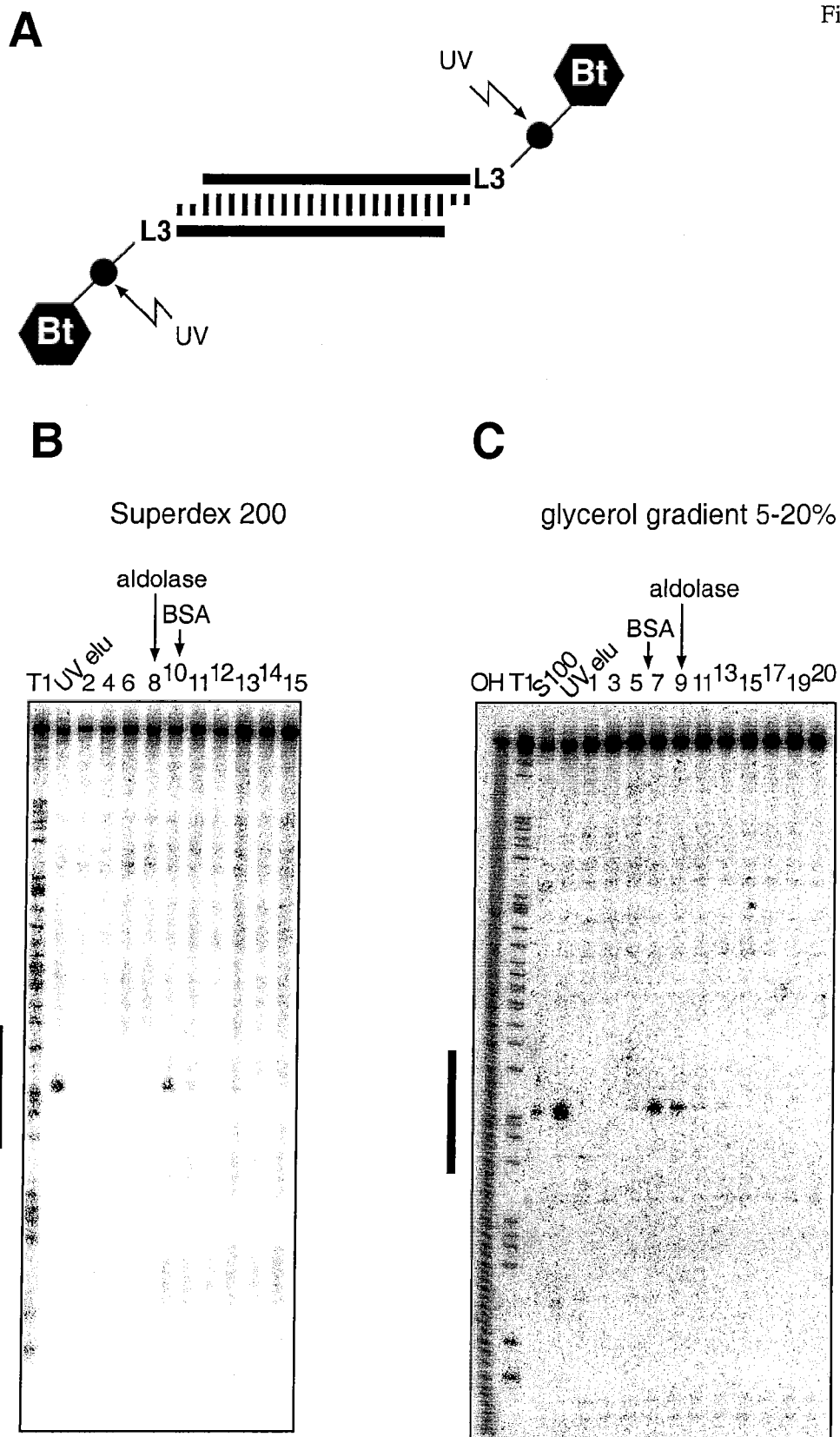


Figure 6

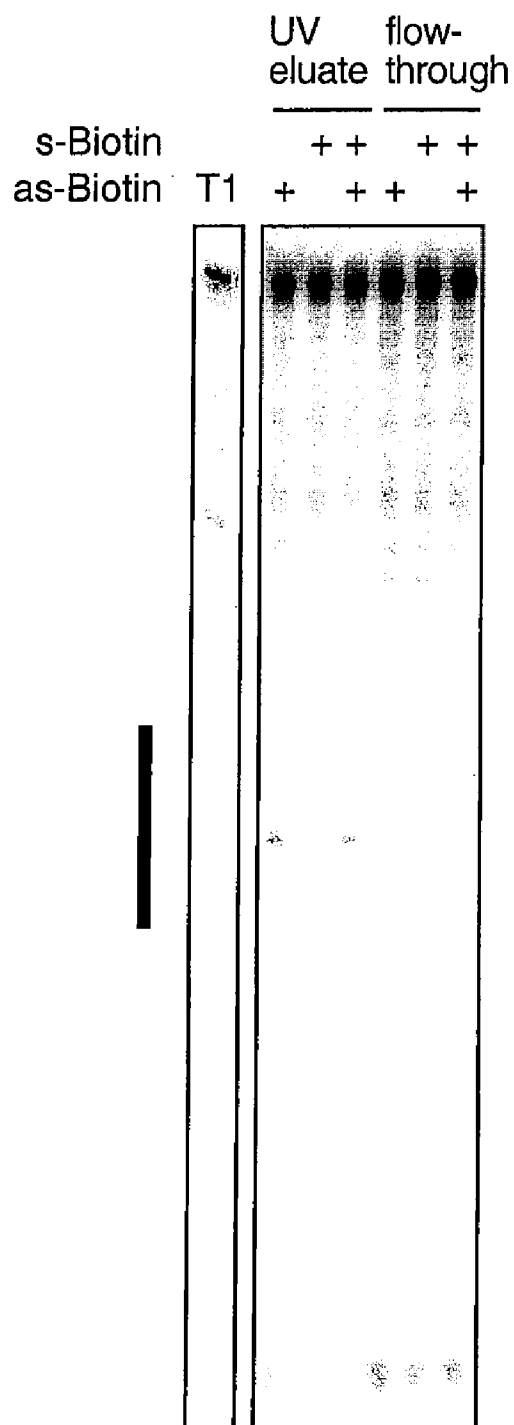
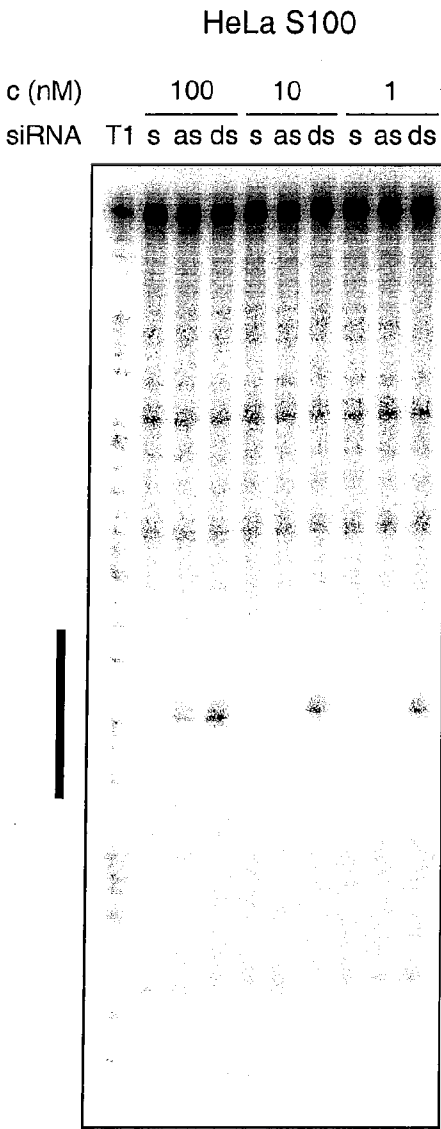


Figure 7

A



B

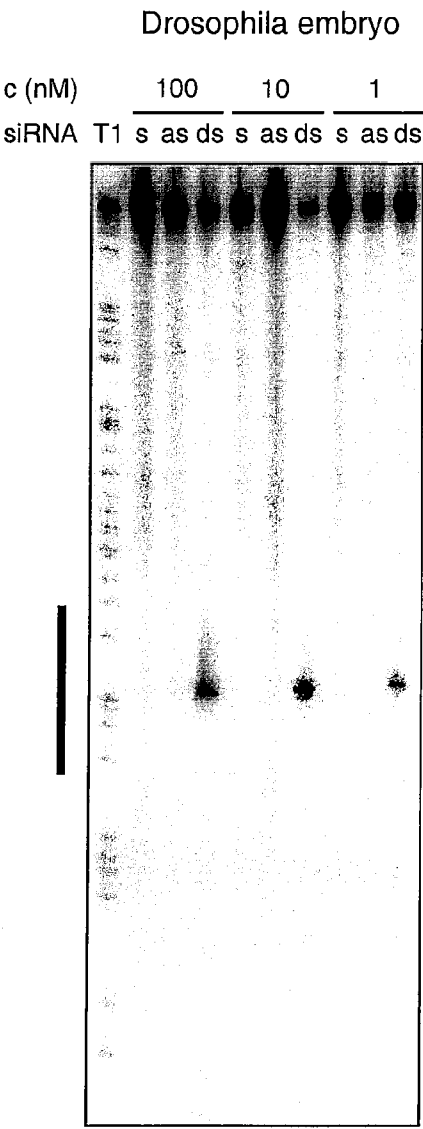


Figure 8

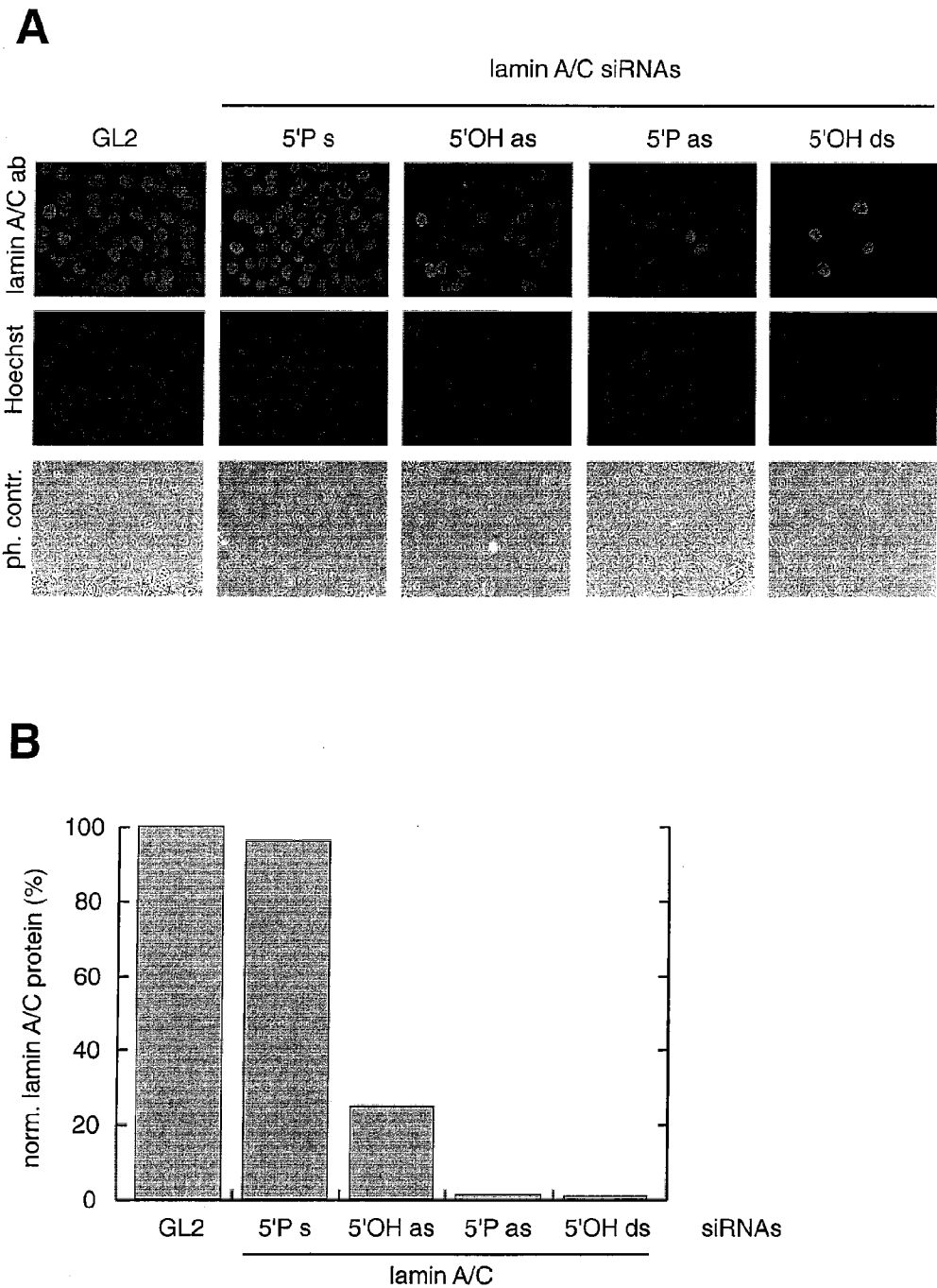
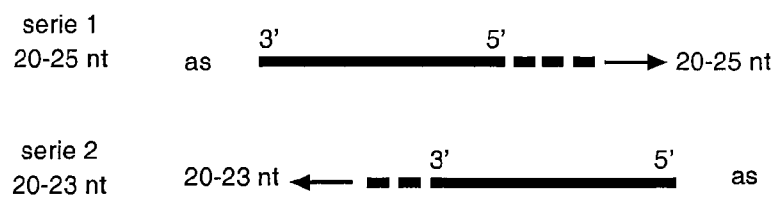


Figure 9

A



B

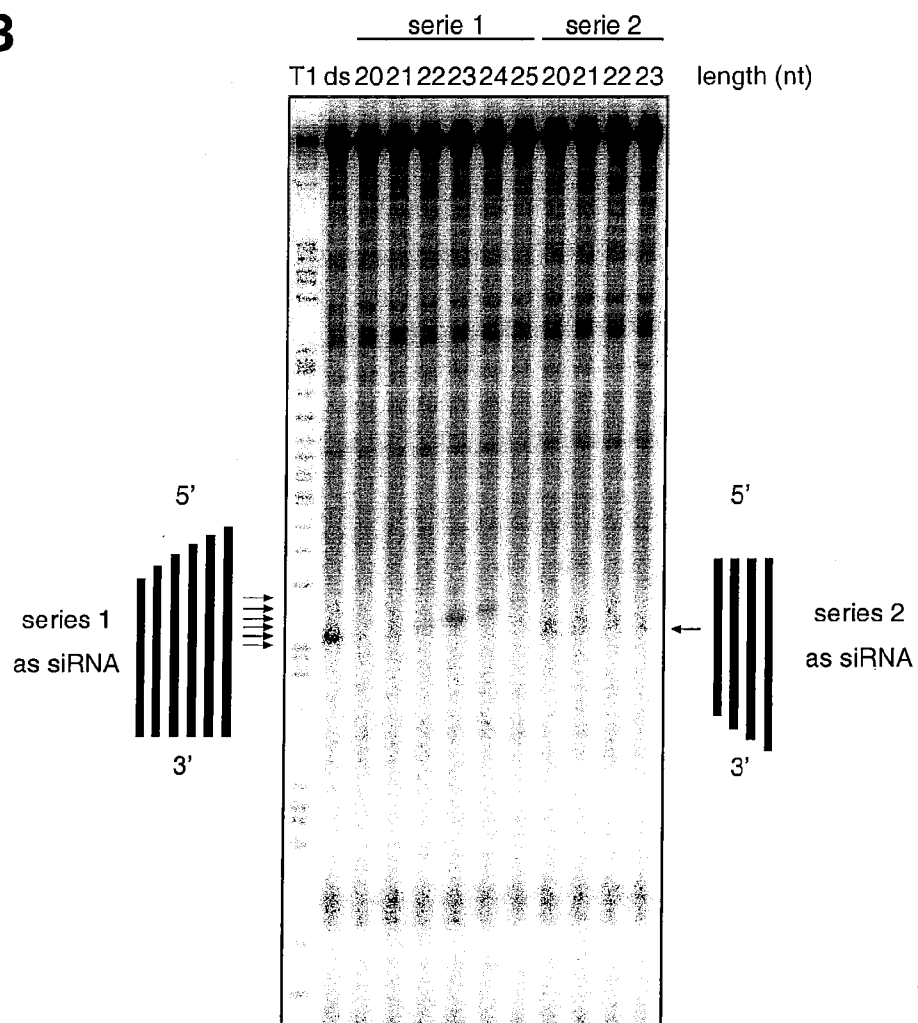


Figure 10

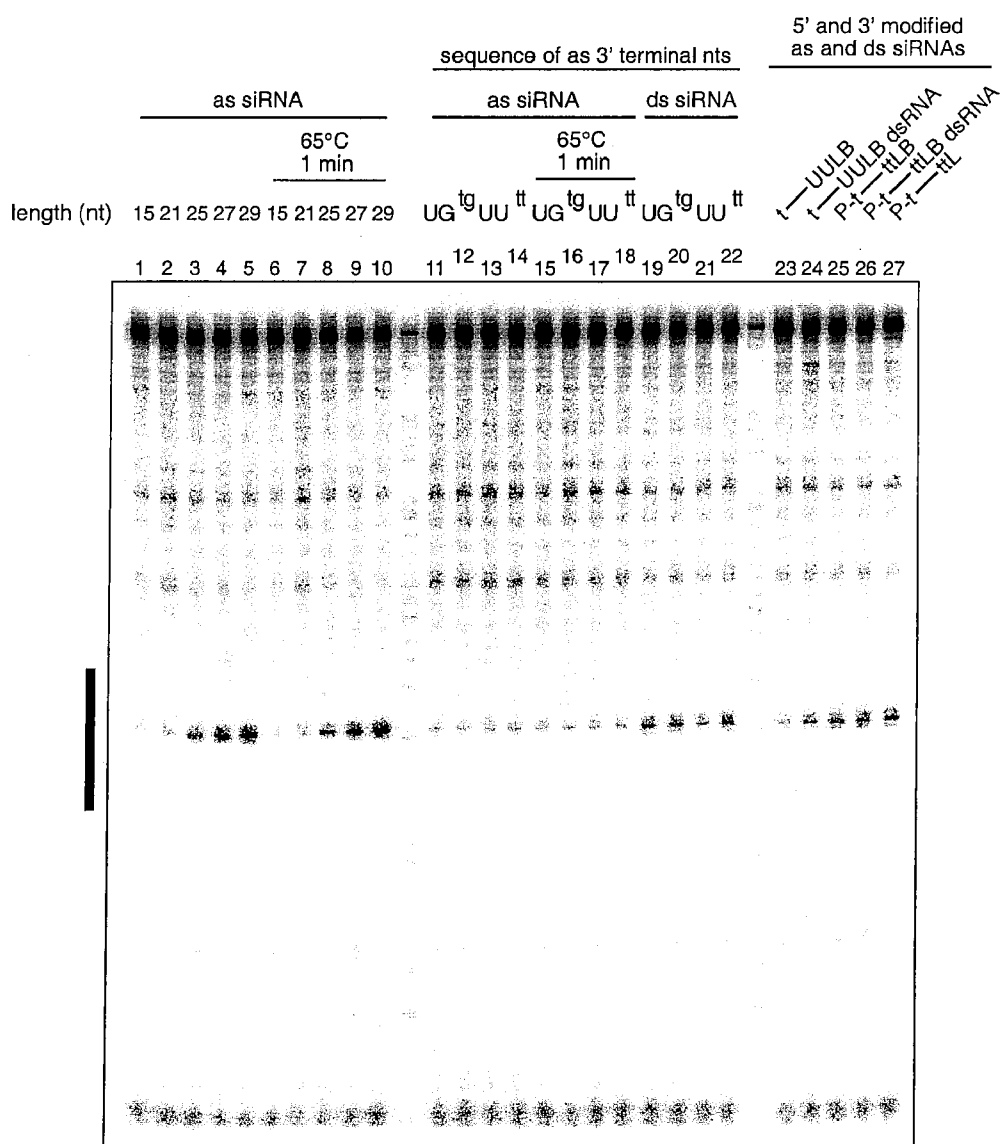


Figure 11

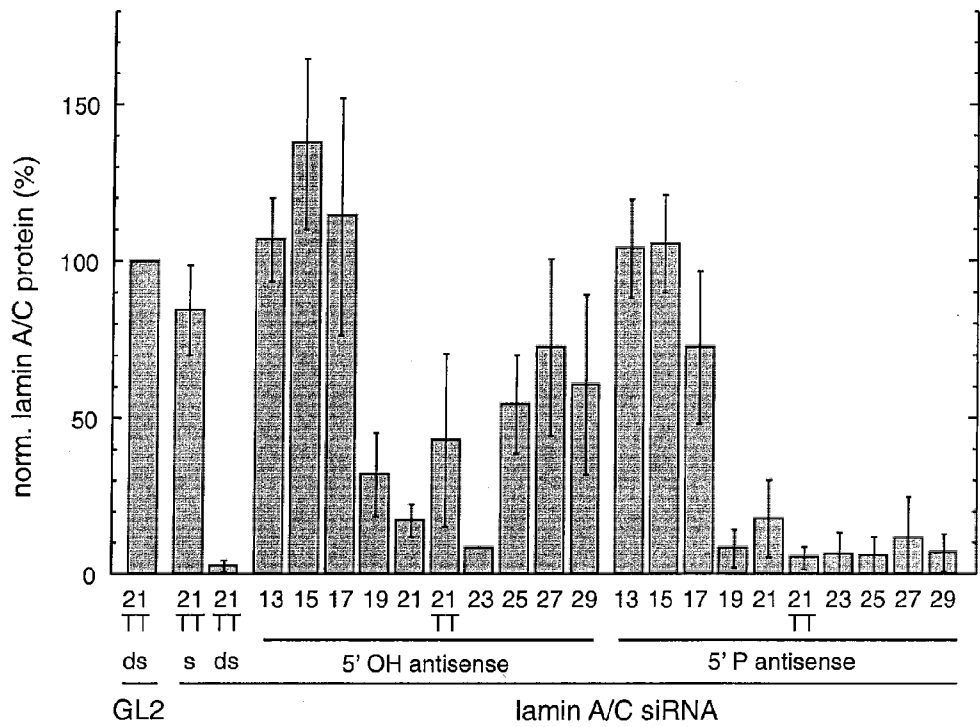


Figure 12

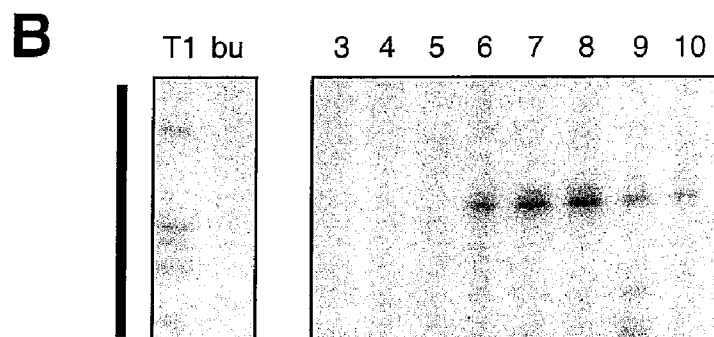
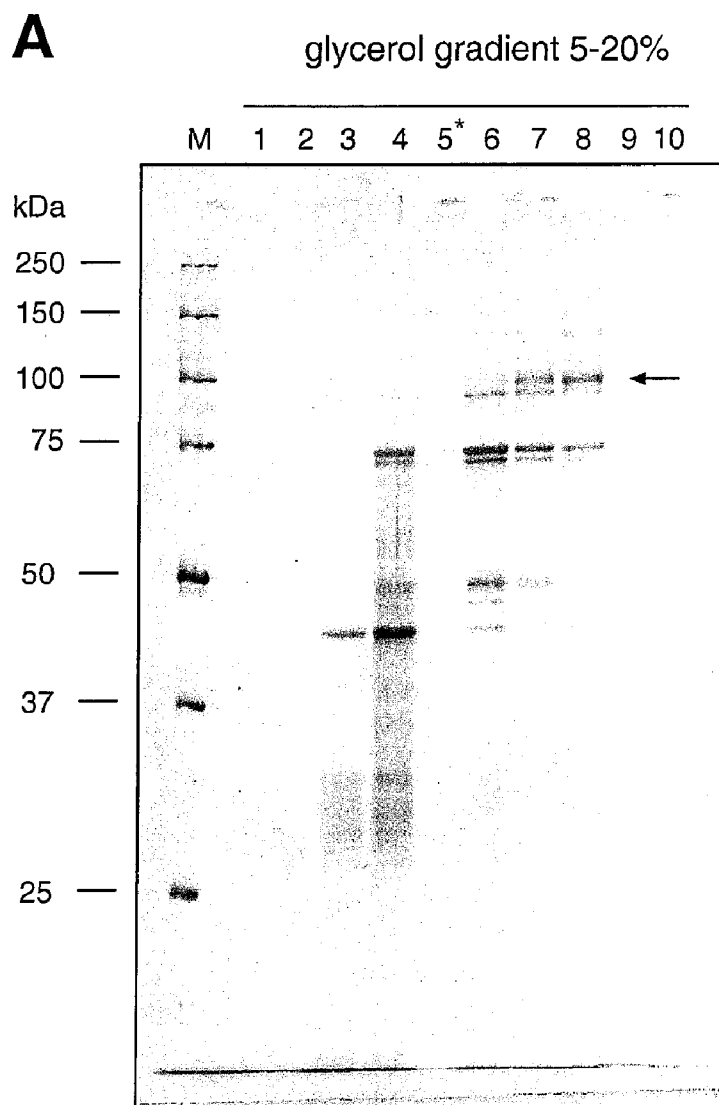
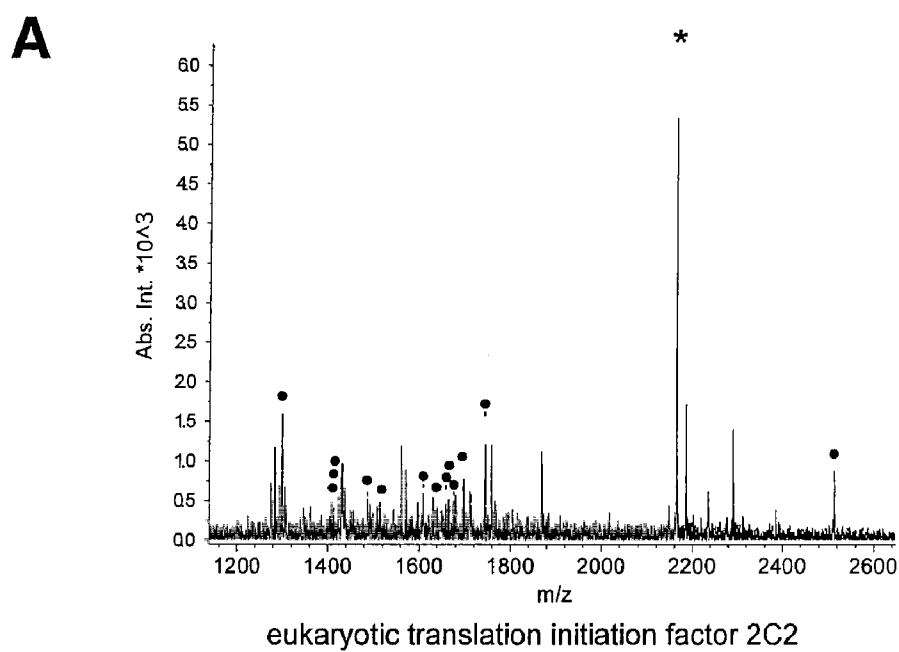
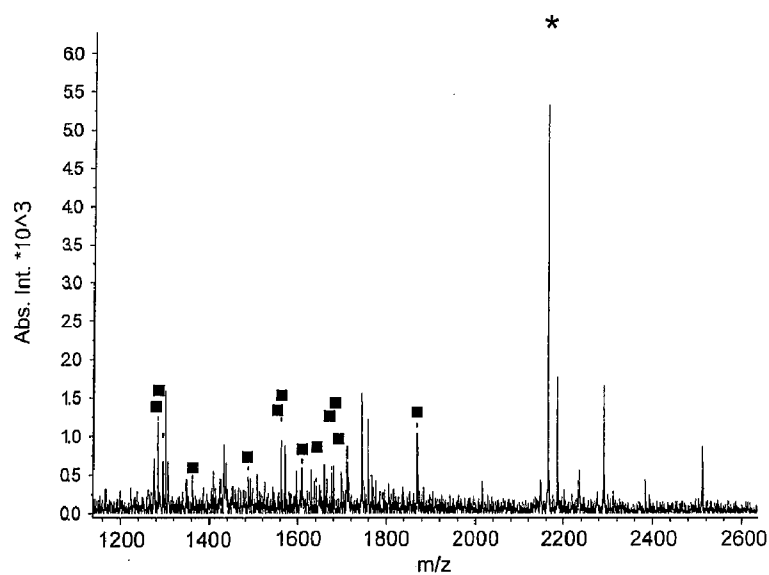


Figure 13 A



Observed	Mr (expt)	Mr (calc)	Delta	Position	Miss	Peptide
1299.67	1298.67	1298.73	-0.07	413 - 424	0	VLQPPSILYGGR
1402.64	1401.64	1401.74	-0.10	637 - 648	0	QEIIQDLAAMVR Oxidation(M)
1413.62	1412.61	1412.73	-0.12	169 - 180	1	HLPSMRYTPVGR
1423.60	1422.59	1422.71	-0.12	356 - 367	1	KLTDNQTSTMIR Oxidation(M)
1486.56	1485.56	1485.66	-0.10	495 - 507	0	YAQGADSVPEPMFR Oxidation(M)
1513.71	1512.70	1512.80	-0.10	112 - 125	1	DKVELEVTLPGEGK
1608.67	1607.66	1607.69	-0.03	481 - 494	0	DAGMPIQGQPCFCK
1635.84	1634.83	1634.85	-0.02	85 - 98	1	TQIFGDRKPVFDGR
1658.85	1657.85	1657.84	0.01	368 - 382	2	ATARSAPDRQEEISK
1663.85	1662.85	1662.91	-0.06	698 - 711	1	DYQPGITFIVVQKR
1675.79	1674.78	1674.84	-0.06	372 - 385	2	SAPDRQEEISKLMR Oxidation(M)
1696.77	1695.76	1695.84	-0.08	323 - 336	0	YPHLPCLQVGQEQK
1743.75	1742.74	1742.77	-0.03	181 - 197	0	SFFTASEGCSNPLGGGR
2511.07	2510.06	2510.12	-0.05	816 - 838	1	YHLVDKEHDSAEGSHTSGQSNGR

Figure 13 B

B

eukaryotic translation initiation factor 2C1

Observed	Mr (expt)	Mr (calc)	Delta	Position	Miss	Peptide
1283.66	1282.65	1282.74	-0.09	410 - 421	0	VLPAPILQYGGK
1294.65	1293.64	1293.67	-0.03	794 - 805	0	SVSIPAPAYYAR
1361.61	1360.60	1360.70	-0.10	553 - 564	0	TSPQTLNLCLEK
1486.56	1485.56	1485.66	-0.10	492 - 504	0	YAQGADSVPEPMFR
1560.76	1559.75	1559.83	-0.08	97 - 110	0	NIYTVTALPIGNER
1561.76	1560.75	1560.78	-0.02	111 - 124	1	VDFEVTIPGEGKDR
1608.67	1607.66	1607.69	-0.03	478 - 491	0	DAGMPIQGQPCFCK
1640.74	1639.73	1639.82	-0.08	240 - 253	0	NIDEQPKPLTDSQR
1675.79	1674.78	1674.84	-0.06	369 - 382	2	SAPDRQEEISRLMK
1679.86	1678.85	1678.90	-0.05	695 - 708	1	DYQPGITYIVVQKR
1696.77	1695.76	1695.84	-0.08	320 - 333	0	YPHLPCLQVGQEQK
1867.85	1866.85	1866.87	-0.02	178 - 194	0	SFFSPPEGYYHPLGGGR

Fig.13C

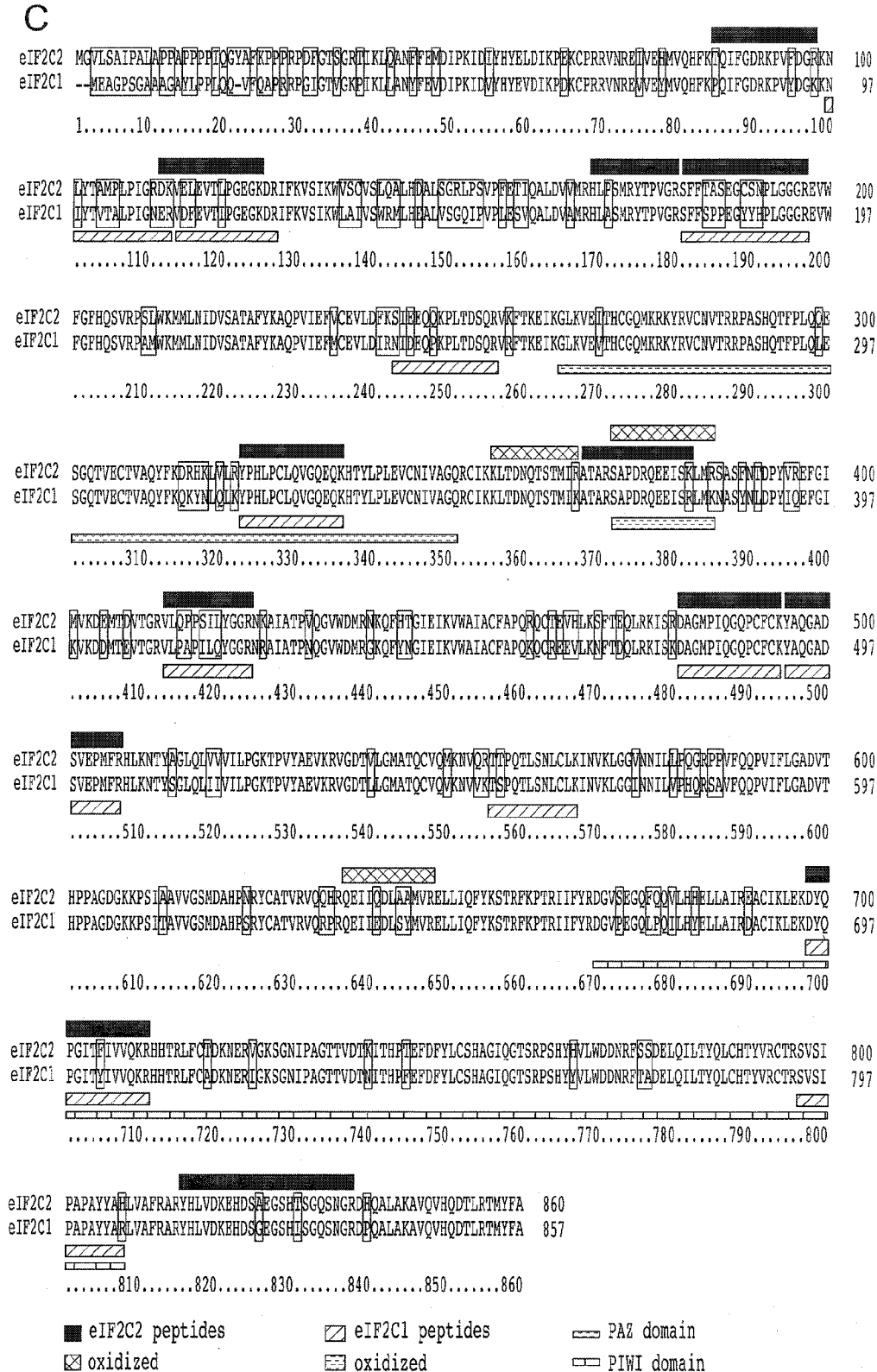


Figure 14

>eIF2C1, predicted protein sequence
 MEAGPSGAAAGAYLPPLQQVFQAPRRPGIGTVGKPIKLLANYFEVDIPKIDVYHYEVDIKPD
 KCPRRVNREVVEYMVQHFQKQIFGDRKPVYDGKKNIYTVTALPIGNERVDFEVTIPGEGKDR
 IFKVSISKWLAIIVSWRMLHEALVSGQIPVPLESVQALDVAMRHLASMRYPVGRSFFSPPEGY
 YHPLGGGREVWFGFHQSVRPAMWKMLNIDVSATAFYKAQPVIEFMCEVLDIRNIDEQPKPL
 TDSQRVRFTEIKGLKVEVTHCGQMKRKYRVCNVTRRPASHQTFPLQLESGQTVECTVAQYF
 KQKYNLQQLKYPHLPCLQVGQEQKHTYLPLEVCNIVAGQRCIKKLTNDQSTMIKATARSAPD
 RQEEISRLMKNASYNLDPYIQEFGIKVKDDMTTEVTGRVLPAPILQYGGRNRAIATPNQGVWD
 MRGKQFYNGIEIKVWAIACFAPQKQCREEVLKNFTDQLRKISKDAGMPIQGQPCFCKYAQGA
 DSVEPMFRHLKNTYSGQLIIVILPGKTPVYAEVKRVGDTLLGMATQCVQVKNVVKTSPTL
 SNLCLKINVKLGGINNILVPHQRSVAVFQQPVIFLGADVTHPPAGDGKKPSITAVVGSMDAHP
 SRYCATVRVQRPQEIIEDLSYMVRELLIQFYKSTRFKPTRIIFYRDGVPEGQLPQILHYEL
 LAIRDACIKLEKDYQPGITYIVVQKRHHTRLFCADKNERIGKSGNIPAGTTVDTNITHPFEF
 DFYLCSHAGIQGTSRPSHYVWLWDDNRFADDELQILTYQLCHTYVRCTRSVSIAPAYYARL
 VAFRARYHLVDKEHDSGEGSHISGQSNGRDPQALAKAVQVHQDTLRTMYFA

>eIF2C2, predicted protein sequence
 MGVLSAIPALAPPAPPPPIQGYAFKPPRPDFGTSGRTIKLQANFFEMDIPKIDIYHYELDI
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 KDRIKFVSIKWVSCVSLQALHDALSGRLPSVPFETIQALDVVMRHLPSMRYPVGRSFFTAS
 EGCSNPLGGGREVWFGFHQSVRPSLWKMLNIDVSATAFYKAQPVIEFVCEVLDFKSIIEEQQ
 KPLTDSQRVKFTKEIKGLKVEITHCGQMKRKYRVCNVTRRPASHQTFPLQLESGQTVECTVA
 QYFKDRHKLVLRYPHLPCLQVGQEQKHTYLPLEVCNIVAGQRCIKKLTNDQSTMIRATARS
 APDRQEEISKLMRSASFNTDPYVREFGIMVKDEMTDVTGRVLQPPSILYGGRNKAIATPVQG
 VWDMRNKQFHTGIEIKVWAIACFAPQRCQTEVHLKSFTEQLRKISRDAGMPIQGQPCFCKYA
 QGADSVEPMFRHLKNTYAGLQLVVVILPGKTPVYAEVKRVGDTVLGMATQCVQMKNVQRTTP
 QTLNCLKINVKLGGVNNILLPQGRPPVFQQPVIFLGADVTHPPAGDGKKPSIAAVVGSMD
 AHPNRYCATVRVQQHRQEI IQDLAAMVRELLIQFYKSTRFKPTRIIFYRDGVSEGQFQVVLH
 HELLAIREACIKLEKDYQPGITFIVVQKRHHTRLFCTDKNERVGKSGNIPAGTTVDTKITHP
 TEFDFYLCSHAGIQGTSRPSHYVWLWDDNRFSSDELQILTYQLCHTYVRCTRSVSIAPAYY
 AHLVAFRARYHLVDKEHDSAEGSHTSGQSNGRDHQALAKAVQVHQDTLRTMYFA

>eIF2C3, predicted protein sequence
 SRSRVVPVPGPAAAAAPCPAPASPRRHPSANIPEIKRYAAAAAAAAGPGAGGAGDRGEAAPAA
 AMEALGPGPPASLFPQPRRPGLTGTVGKPIRLLANHFQVQIPKIDVYHYDVIDKPEKRPRRVN
 REVVDTMVRHFQMIFGDRQPGYDGKRNMYTAHPLPIGRDRVDMEVTLPGEGKDQTFKVSQ
 WVSVSLQLLLLEALAGHLNEVPDDSVQALDVITRHLPSMRYPVGRSFFSPPEGYYHPLGGG
 REVWFGFHQSVRPAMWNMMLNIDVSATAFYRAQPIIEFMCEVLDIQNINEQTKPLTDSQRVK
 FTKEIRGLKVEVTHCGQMKRKYRVCNVTRRPASHQTFPLQLENGQAMECTVAQYFKQKYSQ
 LKYPHLPCLQVGQEQKHTYLPLEVCNIVAGQRCIKKLTNDQSTMIKATARSAPDRQEEISR
 LVKSNMVGPDYPYLKEFGIVVHNEMTELTGRVLPAPMLQYGGRNKTVATPNQGVWDMRGKQ
 FYAGIEIKVWAVACFAPQKQCREDLLKSFTDQLRKISKDAGMPIQGQPCFCKYAQGADSVEP
 MFKHLKMTYVGLQLIVVILPGKTPVYAEVKRVGDTLLGMATQCVQVKNVVKTSPTLNLCL
 KINAKLGGINNVLVPHQRPSVVFQQPVIFLGADVTHPPAGDGKKPSIAAVVGSMDGHPSRYCA
 TVRVQTSRQEI SQELLYSQEVIQDLTNMVRELLIQFYKSTRFKPTRIIFYRGGVSEGQMKQV
 AWPELIAIRKACISLEEDYRPGITYIVVQKRHHTRLFCADKTERVGKSGNVAGTTVDSTIT
 HPSEDFYLCSHAGIQGTSRPSHYQVLWDDNCFADDELQLLTYQLCHTYVRCTRSVSIAPAYY
 YARLVAFRARYHLVDKDHDSEAEGSHVSGQSNGRDPQALAKAVQIHHDTHQHTMYFA

Figure 14

>eIF2C4, predicted protein sequence

AGPAGAQPLLMVPRRPGYGTMGKPIKLLANCFQVEIPKIDVYLYEVDIKPDKCPRRVNREVV
DSMVQHFKVTIFGDRRPVYDGGKRSLYTANPLPVATGTGVDLDVTLPGEGGKDRPFKVSIFVVS
RVSWHLLHEVLTGRTLPEPELELDKPISTNPVHAVDVLRHLPSMKYTPVGRSFFSAPEGYDH
PLGGGREVWFGFHQSVRPAMWKMLNIDVSATAFYKAQPVIFQMCEVLDIHNIDEQPRPLTD
SHRVKFTKEIKGLKVEVTHCGTMRKRYVCNVTRRPASHQTFPLQLENGQTVERTVAQYFRE
KYTLQLKYPHLPCLQVQGEQKHTYLPLEV CNIVAGQRCIKKLTNDQTSTMIKATARSAPDRQ
EETSRLVRSANYETDPFVQEFQFKVRDEMAHVTGRVLPAPMLQYGGRNRTVATPSHGVWDMR
GKQFHTGVEIKMWAIACFATQRQCREEILKGFTDQLRKISKDAGMPIQGQPCFKYAQGADS
VEPMFRHLKNTYSGLQLIIVILPGKTPVYAEVKRVGDTLLGMATQCVQVKNVIKTSPTLSN
LCLKINVKLGGINNILVPHQRPSVFQQPVIFLGADVTHPPAGDGKKPSIAAVVGSMDAHP SR
YCATVRVQRPRQEIQLDLASMVRELLIQFYKSTRFKPTRIIFYRDGVSEGQFRQVLYYELLA
IREACISLEKDYQPGITYIVVQKRHHTRLFCAADRTERVGRSGNIPAGTTVDTDITHPYEFD
YLC SHAGIQGTSRPSHYHVLWDDNCFDADELQLLTYQLCHTYVRCRTRSVSIPAPAYYAH LVA
FRARYHLVDKEHDSAEGSHVSGQSNGRDPQALAKAVQIHQDTLRTMYFA

>HILI, predicted protein sequence

ISSGDAGSTFMERGVKNKQDFMDLSICTREKLAHVRNCKTGSSGIPVKLVTNLFNLDFPQDW
QLYQYHVITYIPDLASRRRLRIALLYSHSELSNKAKAFDGAIFLSQKLEEKVTELSSETQRGE
TIKMTITLKRRLPSSSPVCIQVFNIIFRKILKKLSMYQIGRNFYNPSEPMEIPQHKLSLWPG
FAISVSYFERKLLFSADVSYKVLARNETVLEFMTALCQRTGLSCFTQTCEKQLIGLIVLTRYN
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AQLAHLIPELCLTGLTDQATSDFQLMKAVAETRLSPSGRQORLARLVNDIQRNTNARFEL
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QLLSSVAESSNTSSRLSVIVVRKKCMPRFFTEMNRTVQNPPLGTVDSEATRNEWQYDFYL
ISQVACRGTVSPTYYNVIYDDNGLKPDHMQRLTFKLCHLYYNWPGIVSVPA PCQYAHKLTF L
VAQSIHKEPSLELANHLFY L

>HIWI, predicted protein sequence

MTGRARARARGRARGQETAQLVGSTASQQPGYIQPRPQPPPAEGELFGRGRQRGTAGGTAKS
QGLQISAGFQELSLAERGGRRRDFHDLGVNTRQNL DHVKESKTGSSGIIVRLSTNHFRLTSR
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LLSSNRKDKYDAIKKYLCTDCPTPSQCVVARTLGKQQTVMAIATKIALQMNCKMGGELWRVD
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LRAWNSCNEYMPSRIIVYRDGVGDGQLKTLVNYEVPQFLDCLKLSIGRGYNPRLTVIVVKKRV
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Fig.15

eIF2C3	SRSRVVPCPGAAAPCPAPASPRRHPSANTPEIKRYAANAAGAGGAGGAGCGTAAPANAAMEALCPPPASLFP	100
eIF2C4	AGPACAOPLLMPRRPGYGTACKPIKLLANCFQV	34
eIF2C1	MEAGPSGAAGAYLPLOQVFOAPRRPGIGTVGKPIKLLANFEM	45
eIF2C2	MGVSAIPALAPPAPPPPTQGYAKPPRRPFGTSGRTIKLOANFEM	48
HILI	ISSGAGSTFMERGKKNKQDFMDL	24
HIWI	MTFARARARGHARGOETAQLVSTASQOPGYTQPPQPPAGELFCRGROGTAGGTAKSGLQISAGFOELSLAERGR-RRDFIDL	89
ruler	1.....10.....20.....30.....40.....50.....60.....70.....80.....90.....100	
eIF2C3	QTPKIDVYHYVDIKPEKR-PRRNREVDVTVRHFQMOLFGDRQPGVDGKRN-MYTAAMPLPTGRDNDMNTLPCEG-KDQTFKVSVMVWSVM	191
eIF2C4	ETPKIDVYHYVDIKPEK-PRRNREVDVSNVQHFQMTIFGDRRPFVDGKRS-LYTAMPLPMTATGVOLDVTLPGEGGKDRPKVSTIKVFSRV	126
eIF2C1	DTPKIDVYHYVDIKPEK-PRRNREVDVYVQHFQMTIFGDRKPFVDGKKN-LYTAMPLPTGNRVDFEVTLPCEG-KDRIFKVSTIKVLAIV	136
eIF2C2	DTPKIDVYHYVDIKPEK-PRRNREVDVYVQHFQMTIFGDRKPFVDGKKN-LYTAMPLPTGRDVELEMTLPCEG-KDRIFKVSTIKVWSCV	139
HILI	SICTRKLAHVNRCKTSSSTIPKLVNLFNLDFPDQWLYQYHVITYIPDLASRRRLTALLYSHSELNKAFAFGAILFLSQLEEKVTLSSSTORGE	124
HIWI	GVNTRONLDHVKESKTSSTIVRLSTNHFRLTSRPDVALYQYHIDYVNLMEARNLSALLFOHEDLIGKCHAFDGTILFLPKMLQOKVTEVFSKTRNGE	189
ruler110.....120.....130.....140.....150.....160.....170.....180.....190.....200	
eIF2C3	SLQLLEALAGHLN-EPPDISVQALDVTVR-HLPSMRYTPVGRSFFSPPEGYVHPLGGGREVWFGFHQSVRPAMWMMMLNIDVSATAFYKAQF	282
eIF2C4	SWELLHEVLGRTLPLEPLEDKPTSTNPVHADVTLR-HLPSMRYTPVGRSFFSAPEGYDHPPLGGGREVWFGFHQSVRPAMWMMMLNIDVSATAFYKAQF	225
eIF2C1	SWRMILHEALVSGQTP-MPLESVOALDVAMR-HLASMRYTPVGRSFFSPPEGYVHPLGGGREVWFGFHQSVRPAMWMMMLNIDVSATAFYKAQF	227
eIF2C2	SLQALHDALSCRLPS-MPFETIQALDVVMR-HLPSMRYTPVGRSFFTASEGCSNPLGGGREVWFGFHQSVRPSLWMMMLNIDVSATAFYKAQF	230
HILI	TUKMILTAKRELPSS-SPVCIQVFNLIIFRKILKLSMYQIGRNFYNPSEPMETPOHK-LSLWPGFALSVMYFERKILLSADVSKVLR-NET	213
HIWI	DVNFILITLITNELPPT-SPITCLOFNLIIFRRLKIMNLOIGRMVYVNPDEPIDPSHR-LVWPGFTITSLQYENSIMLCIDVSHKVL-SET	278
ruler210.....220.....230.....240.....250.....260.....270.....280.....290.....300	
eIF2C3	VIEFMCBVLDIRNIDEQPKPLTDSQVWKTKETKGLKVEITHCGMKRKRYVQVWTRRPASHOTFPLENGCTVETVAQYFKEKYTLQKYPHLPCLQ	382
eIF2C4	VIEFMCBVLDIRNIDEQPKPLTDSQVWKTKETKGLKVEITHCGMKRKRYVQVWTRRPASHOTFPLENGCTVETVAQYFKEKYTLQKYPHLPCLQ	325
eIF2C1	VIEFMCBVLDIRNIDEQPKPLTDSQVWKTKETKGLKVEITHCGMKRKRYVQVWTRRPASHOTFPLENGCTVETVAQYFKEKYTLQKYPHLPCLQ	327
eIF2C2	VIEFVCEVLDIFKSTDEQPKPLTDSQVWKTKETKGLKVEITHCGMKRKRYVQVWTRRPASHOTFPLENGCTVETVAQYFKEKYTLQKYPHLPCLQ	330
HILI	VLEFMTALCORTGLS-CFTQTCQKQLGLIWLITRYN-NRTYSIDDIDWSVKPHTHTFOKR-DGTEITTYVDYKQOYDITVSDLNQPMNV	298
HIWI	VLDFTMNFYHOTEH-KFQEDVSKELTGLVWLTKYN-NKTYVDDIDWDONPKSTFKKA-DGSEVSFLEYRKOYNOETIDLKOPMLV	363
ruler310.....320.....330.....340.....350.....360.....370.....380.....390.....400	
eIF2C3	WQEQKHHT-VLPLEVCNIVAGORCIKKLTDNOT-STMIKATARSAPDROEELSKVKSNSMVGCPDPLKEFGIMVHNMETLTGRVLPAPMLQ	
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eIF2C1	WQEQKHHT-VLPLEVCNIVAGORCIKKLTDNOT-STMIKATARSAPDROEELSKVMSASYNL-DPVIQEFQVVKDDMTETGRVLPAPMLQ	
eIF2C2	WQEQKHHT-VLPLEVCNIVAGORCIKKLTDNOT-STMIKATARSAPDROEELSKVMSASNT-DPVIQEFQVVKDDMTETGRVLPAPMLQ	
HILI	SLKKKRN-DNSEAQLAHLPELCFLTGLTDQATSDFOLKVAEKITFLSPSGROORLRLVDNIQNTNARFEITMGLHFGS-QISLTGRVPSKIL	
HIWI	SQKRRRCGCGTLPQAPMLPELCVLTGLTDKMRDNFMVKDLAVHTRLTPEQROREVCRLDYTHNDVMORELDMCLSPDSNLSFGRTILOTEKH	
ruler410.....420.....430.....440.....450.....460.....470.....480.....490.....500	

Fig.15 (cont.)

eIF2C3 NGG-RNRATVATPAGVWDMRGKQFYAGIEIKVWAVACFAPQOCREDLLNSFTDQLRKISKDAGMHTCCQCFCKYACGADSVPEPFRHNTYVGLQLI 573
eIF2C4 YCG-RNRATVATPAGVWDMRGKQFYAGIEIKVWAVACFAPQOCREHLLKGTDTQLRKISKDAGMHTCCQCFCKYACGADSVPEPFRHNTYVGLQLI 514
eIF2C1 YCG-RNRATVATPAGVWDMRGKQFYAGIEIKVWAVACFAPQOCREHLLKGTDTQLRKISKDAGMHTCCQCFCKYACGADSVPEPFRHNTYVGLQLI 516
eIF2C2 YCG-RNRATVATPAGVWDMRGKQFYAGIEIKVWAVACFAPQOCREHLLKGTDTQLRKISKDAGMHTCCQCFCKYACGADSVPEPFRHNTYVGLQLI 519
HILI MDHICOP-VAADWSKDIRTCKTINAOSLNTWLILCSDR-----TETVAESFLNCLRRVAGSMCFN-----NM 459
HIWI CCGKTFDYNFQADWSKETRGAPLISVXPLEDMLLIVTR-----NYEANSLLIOMLFKVIIPAGMOMRKATMLEVDDRTTEAMLRVLOOKVTADTQ--INV 557
ruler510.....520.....530.....540.....550.....560.....570.....580.....590.....600

eIF2C3 VVILPKPTVYAEVKR/GDTLLGNATQCVQKMMK--TSPQTLNLCIKINAKLGGINNVLVPHQPSVFOQPVIFLGADVTHPAGDCKKPSIAAVVG 671
eIF2C4 VVILPKPTVYAEVKR/GDTLLGNATQCVQKMMK--TSPQTLNLCIKINAKLGGINNVLVPHQPSVFOQPVIFLGADVTHPAGDCKKPSIAAVVG 612
eIF2C1 VVILPKPTVYAEVKR/GDTLLGNATQCVQKMMK--TSPQTLNLCIKINAKLGGINNVLVPHQPSVFOQPVIFLGADVTHPAGDCKKPSIAAVVG 614
eIF2C2 VVILPKPTVYAEVKR/GDTLLGNATQCVQKMMK--TTPQTLNLCIKINAKLGGINNVLVPHQPSVFOQPVIFLGADVTHPAGDCKKPSIAAVVG 617
HILI CILPSNQTYYSIKKYLSSDCVPSQVLPARTLNKQGMMSIATKIAMQTXLGG-----ELNVEIPLKSLMVVGDVCKDALS--DMMVGCVA 551
HIWI CLSSNRKDYDAIKKYLCTDPTPSQVWARTLKQOTVMAIAKIALDMCKMG-----ELNVEIPLKSLMVVGDVCKDALS--RSTAGFVA 649
ruler610.....620.....630.....640.....650.....660.....670.....680.....690.....700

eIF2C3 SMDGHPSPRYCAIVRQ/SRQETSOE LNSQVIOITNMVRELLIOFYKSTRFKPTRLIYVRCGVSEGOVQVAMPGLIATKACTSLEKDYQPGITIV 771
eIF2C4 SMDGHPSPRYCAIVRQ/SRQETSOE LNSQVIOITNMVRELLIOFYKSTRFKPTRLIYVRCGVSEGOVQVAMPGLIATKACTSLEKDYQPGITIV 702
eIF2C1 SMDGHPSPRYCAIVRQ/SRQETSOE LNSQVIOITNMVRELLIOFYKSTRFKPTRLIYVRCGVSEGOVQVAMPGLIATKACTSLEKDYQPGITIV 704
eIF2C2 SMDGHPSPRYCAIVRQ/SRQETSOE LNSQVIOITNMVRELLIOFYKSTRFKPTRLIYVRCGVSEGOVQVAMPGLIATKACTSLEKDYQPGITIV 707
HILI SVNPRTIWFPSRCLQRTWT-----DVADCLKVFMTCALNKWKV/NHDLPARTIVRAGVGGGLKFLIEVEVPOLLSSVAESSNTSSRLSVIV 641
HIWI STNGNTIWFPSRCLQRTWT-----ELVDGLKVCLOALRAWSCNEMPSRIIYVRCGVGGGLKFLIEVEVPOLLCKSTGRGYNRLIVIV 739
ruler710.....720.....730.....740.....750.....760.....770.....780.....790.....800

eIF2C3 VQKRHHTRLFCDKMERVCKSGNPAGTIVDTIT-HPSEDFYLCSHAGIGQTSRPSHYV/LWDDNFTADELQLITVQLCHTVVCTRSVSTIPAPAYY 870
eIF2C4 VQKRHHTRLFCDKMERVCKSGNPAGTIVDTIT-HPSEDFYLCSHAGIGQTSRPSHYV/LWDDNFTADELQLITVQLCHTVVCTRSVSTIPAPAYY 801
eIF2C1 VQKRHHTRLFCDKMERVCKSGNPAGTIVDTIT-HPSEDFYLCSHAGIGQTSRPSHYV/LWDDNFTADELQLITVQLCHTVVCTRSVSTIPAPAYY 803
eIF2C2 VQKRHHTRLFCDKMERVCKSGNPAGTIVDTIT-HPSEDFYLCSHAGIGQTSRPSHYV/LWDDNFTADELQLITVQLCHTVVCTRSVSTIPAPAYY 806
HILI VRKCKMRFFFEMN-----RTVQNPPLGTWVDSATRNEQYDFYLSQVACRGTVSPTYNWIYDNGLPDHWQRLTFKLCHLYNWPGLVSPAPCOY 737
HIWI VQKRHHTRLFCDKMERVCKSGNPAGTIVDTIT-HPSEDFYLCSHAGIGQTSRPSHYV/LWDDNFTADELQLITVQLCHTVVCTRSVSTIPAPAYY 834
ruler810.....820.....830.....840.....850.....860.....870.....880.....890.....900

eIF2C3 ARLVAFRARYHLVDKEHDSAGSHVSGOSNGRDPQALAKAVQHDITLRTMYFA 924
eIF2C4 ARLVAFRARYHLVDKEHDSAGSHVSGOSNGRDPQALAKAVQHDITLRTMYFA 855
eIF2C1 ARLVAFRARYHLVDKEHDSAGSHVSGOSNGRDPQALAKAVQHDITLRTMYFA 857
eIF2C2 ARLVAFRARYHLVDKEHDSAGSHVSGOSNGRDPQALAKAVQHDITLRTMYFA 860
HILI AKKLT-----LVASIHKEP-----SLPLANLTYL 764
HIWI AKKLT-----LVASIHKEP-----NLSLSNRLYYL 861
ruler910.....920.....930.....940.....950.....

Figure 16

>eIF2C1, cDNA sequence of predicted ORF
ATGGAAGCGGGACCCCTCGGGAGCAGCTGCGGGCGCTTACCTGCCCCCCTGCAGCAGGTGTT
CCAGGCACCTCGCCGGCCTGGCATTGGCACTGTGGGGAAACCAATCAAGCTCCTGGCCAATT
ACTTTGAGGTGGACATCCCTAAGATCGACGTGTACCACTACGAGGTGGACATCAAGCCGGAT
AAGTGTCCCCGTAGAGTCAACCGGGAAGTGGTGAATACATGGTCCAGCATTTCAAGCCTCA
GATCTTTGGTGATCGCAAGCCTGTGTATGATGGAAAGAAGAACATTTACACTGTCACAGCAC
TGCCCATTGGCAACGAACGGGTCGACTTTGAGGTGACAATCCCTGGGGAAGGGAAGGATCGA
ATCTTTAAGGTCTCCATCAAGTGGCTAGCCATTGTGAGCTGGCGAATGCTGCATGAGGCCCT
GGTCAGCGGCCAGATCCCTGTTCCCTTGGAGTCTGTGCAAGCCCTGGATGTGGCCATGAGGC
ACCTGGCATCCATGAGGTACACCCCTGTGGGCCGCTCCTTCTTCTCACC GCCTGAGGGCTAC
TACCACCCGCTGGGGGGTGGGCGCGAGGTCTGGTTGCGCTTTTACCAGTCTGTGCGCCCTGC
CATGTGGAAGATGATGCTCAACATTGATGTCTCAGCCACTGCCTTTTATAAGGCACAGCCAG
TGATTGAGTTCATGTGTGAGGTGCTGGACATCAGGAACATAGATGAGCAGCCCAAGCCCCCTC
ACGGACTCTCAGCGCGTTTCGCTTCACCAAGGAGATCAAGGGCCTGAAGGTGGAAGTCACCCA
CTGTGGACAGATGAAGAGGAAGTACCGCGTGTGTAATGTTACCCGTCGCCCTGCTAGCCATC
AGACATTCCCTTTACAGCTGGAGAGTGGACAGACTGTGGAGTGCACAGTGGCACAGTATTTT
AAGCAGAAATATAACCTTCAGCTCAAGTATCCCCATCTGCCCTGCCTACAAGTTGGCCAGGA
ACAAAAGCATACCTACCTTCCCCTAGAGGTCTGTAACATTGTGGCTGGGCAGCGCTGTATTA
AAAAGCTGACCGACAACCAGACCTCGACCATGATAAAGGCCACAGCTAGATCCGCTCCAGAC
AGACAGGAGGAGATCAGTCGCCTGATGAAGAATGCCAGCTACAACCTAGATCCCTACATCCA
GGAATTGGGATCAAAGTGAAGGATGACATGACGGAGGTGACAGGGCGAGTGTGCTGCCGCGC
CCATCTTGCAGTACGGCGGCCGGAACCGGGCCATTGCCACACCCAATCAGGGTGTCTGGGAC
ATGCGGGGGAACAGTTCTACAATGGGATTGAGATCAAAGTCTGGGCCATCGCCTGCTTCGC
ACCCCAAAAACAGTGTGCGAGAAGAGGTGCTCAAGAACTTCACAGACCAGCTGCGGAAGATTT
CCAAGGATGCGGGGATGCCTATCCAGGGTCAACCTTGTCTTCTGCAAATATGCACAGGGGGCA
GACAGCGTGGAGCCTATGTTCCGGCATCTCAAGAACACCTACTCAGGGCTGCAGCTCATTAT
TGTCATCCTGCCAGGGAAGACGCCGGTGTATGCTGAGGTGAAACGTGTGCGAGATACACTCT
TGGGAATGGCTACGCAGTGTGTGAGGTGAAGAACGTGGTCAAGACCTCACCTCAGACTCTG
TCCAACCTCTGCCCTCAAGATCAATGTCAAACCTTGGTGGCATTAACAACATCCTAGTCCCACA
CCAGCGCTCTGCCGTTTTTTCAACAGCCAGTGATATTCTTGGGAGCAGATGTTACACACCCCC
CAGCAGGGGATGGGAAAAAACCTTCTATCACAGCAGTGGTAGGCAGTATGGATGCCCACCCC
AGCCGATACTGTGCTACTGTGCGGGTACAGCGACCACGGCAAGAGATCATTGAAGACTTGTC
CTACATGGTGCGTGAGCTCCTCATCCAATTCTACAAGTCCACCCGTTTCAAGCCTACCCGCA
TCATCTTCTACCGAGATGGGGTGCCCTGAAGGCCAGCTACCCAGATACTCCACTATGAGCTA
CTGGCCATTTCGTGATGCCCTGCATCAAACCTGGAAAAGGACTACCAGCCTGGGATCACTTATAT
TGTGGTGCAGAAACGCCATCACACCCGCCCTTTTCTGTGCTGACAAGAATGAGCGAATTGGGA
AGAGTGGTAACATCCCAGCTGGGACCACAGTGGACACCAACATCACCCACCCATTTGAGTTT
GACTTCTATCTGTGCAGCCACGCAGGCATCCAGGGCACCAGCCGACCATCCCATTACTATGT
TCTTTGGGATGACAACCGTTTTACAGCAGATGAGCTCCAGATCCTGACGTACCAGCTGTGCC
ACACTTACGTACGATGCACACGCTCTGTCTCTATCCCAGCACCTGCCCTACTATGCCCCGCTG
GTGGCTTTCCGGGCACGATACCACCTGGTGGACAAGGAGCATGACAGTGGAGAGGGGAGCCA
CATATCGGGGCAGAGCAATGGGCGGGACCCCCAGGCCCTGGCCAAAGCCGTGCAGGTTTACC
AGGATACTCTGCGCACCATGTACTTCGCT

Figure 16

>eIF2C2, cDNA sequence of predicted ORF
ATGGGTGTTCTCTCTGCCATTCCCGCACTTGCACCTCCTGCGCCGCCGCCCCCATCCAAGG
ATATGCCTTCAAGCCTCCACCTAGACCCGACTTTGGGACCTCCGGGAGAACAATCAAATTAC
AGGCCAATTTCTTCGAAATGGACATCCCCAAAATTGACATCTATCATTATGAATTGGATATC
AAGCCAGAGAAGTGCCCCGAGGAGAGTTAACAGGGAAATCGTGGAACACATGGTCCAGCACTT
TAAAACACAGATCTTTGGGGATCGGAAGCCCGTGTTTGACGGCAGGAAGAATCTATACACAG
CCATGCCCCCTTCCGATTGGGAGGGACAAGGTGGAGCTGGAGGTCACGCTGCCAGGAGAAGGC
AAGGATCGCATCTTCAAGGTGTCCATCAAGTGGGTGTCTGCGTGAGCTTGCAGGCGTTACA
CGATGCACTTTCAAGGCGGCTGCCAGCGTCCCTTTTGAGACGATCCAGGCCCTGGACGTGG
TCATGAGGCACTTGCCATCCATGAGGTACACCCCCGTGGGCCGCTCCTTCTTCACCGCGTCC
GAAGGCTGCTCTAACCCCTCTTGGCGGGGGCCGAGAAGTGTGGTTTGGCTTCCATCAGTCCGT
CCGGCCTTCTCTCTGGAAAATGATGCTGAATATTGATGTGTCAGCAACAGCGTTTACAAGG
CACAGCCAGTAATCGAGTTTGTGTTGTGAAGTTTGGATTTTAAAAGTATTGAAGAACAACAA
AAACCTCTGACAGATTCCCAAAGGGTAAAGTTTACCAAAGAAATTAAAGGTCTAAAGGTGGA
GATAACGCACGTGTGGGCAGATGAAGAGGAAGTACCGTGTCTGCAATGTGACCCGGCGGCCCG
CCAGTCACCAAACATTCCCCTGCAGCAGGAGAGCGGGCAGACGGTGGAGTGCACGGTGGCC
CAGTATTTCAAGGACAGGCACAAGTTGGTTCTGCGCTACCCCCACCTCCCATGTTTACAAGT
CGGACAGGAGCAGAAACACACCTACCTTCCCCTGGAGGTCTGTAAACATTGTGGCAGGACAAA
GATGTATTAATAAATTAACGGACAATCAGACCTCAACCATGATCAGAGCAACTGCTAGGTCTG
GCGCCCGATCGGCAAGAAGAGATTAGCAAATTGATGCGAAGTGCAAGTTTCAACACAGATCC
ATACGTCCGTGAATTTGGAATCATGGTCAAAGATGAGATGACAGACGTGACTGGGCGGGTGC
TGCAGCCGCCCTCCATCCTCTACGGGGGCAGGAATAAAGCTATTGCGACCCCTGTCCAGGGC
GTCTGGGACATCGGGAACAAGCAGTTCCACACGGGCATCGAGATCAAGGTGTGGCCATTGC
GTGCTTCGCCCCCAGCGCCAGTGCACGGAAGTCCATCTGAAGTCTTTCACAGAGCAGCTCA
GAAAGATCTCGAGAGACGCTGGCATGCCCATCCAGGGCCAGCCGTGCTTCTGCAATACGCG
CAGGGGGCGGACAGCGTGGAGCCCATGTTCCGGCACCTGAAGAACACGTATGCGGGCCTGCA
GCTGGTGGTGGTTCATCCTGCCCCGCAAGACGCCCCTGTACGCCGAGGTCAAGCGCGTGGGAG
ACACGGTGCTGGGGATGGCCACGCAGTGCCTGCAGATGAAGAACGTGCAGAGGACCACGCCA
CAGACCCTGTCCAACCTTTGCCTGAAGATCAACGTCAAGCTGGGAGGCGTGAACAACATCCT
GCTGCCCCAGGGCAGGCCGCCGGTGTTCACGACGCCCGTCATCTTCTGGGAGCAGACGTCA
CTCACCCCCCGCCGGGGATGGGAAGAAGCCCTCCATTGCCGCCGTGGTGGGCAGCATGGAC
GCCCCACCCAATCGCTACTGCGCCACCGTGCAGTGCAGCAGCACCAGGAGATCATACA
AGACCTGGCCGCCATGGTCCGCGAGCTCCTCATCCAGTTCTACAAGTCCACGCGCTTCAAGC
CCACCCGCATCATCTTCTACCGCGACGGTGTCTCTGAAGGCCAGTTCCAGCAGGTTCTCCAC
CACGAGTTGCTGGCCATCCGTGAGGCCTGTATCAAGCTAGAAAAAGACTACCAGCCCGGGAT
CACCTTCATCGTGGTGCAGAAGAGGCACCACACCCGGCTCTTCTGCACTGACAAGAACGAGC
GGGTTGGGAAAAGTGGAAACATTCCAGCAGGCACGACTGTGGACACGAAAATCACCCACCCC
ACCGAGTTCGACTTCTACCTGTGTAGTCACGCTGGCATCCAGGGGACAAGCAGGCCTTCGCA
CTATCACGTCTCTGGGACGACAATCGTTTCTCTCTGATGAGCTGCAGATCCTAACCTACC
AGCTGTGTACACCTACGTGCGCTGCACACGCTCCGTGTCCATCCAGCGCCAGCATACTAC
GCTCACCTGGTGGCCTTCCGGGCCAGGTACCACCTGGTGGATAAGGAACATGACAGTGCTGA
AGGAAGCCATACCTCTGGGCAGAGTAACGGGCGAGACCACCAAGCACTGGCCAAGGCGGTCC
AGGTTACCAAGACACTCTGCGCACCATGTACTTTGCT

Figure 16

>eIF2C3, cDNA sequence of predicted ORF

```
AGCCGGAGCCGGGTCCCTGTCCCCGGGGCCGGGCGCCGCCGCCGCCCTGCCAGCGCCCGC
GTCTCCCGCGGCGCCACCCACGCGCAATATTCGGGAGATCAAGCGTTACGCGGCGGCGGCGG
CGGCGGCGGCGGCGGCGGCGGAGCGGGAGGCGCGGGGACCGGGCGAGGCGGCGGCGGCGGCGG
GCCATGGAGGCGCTGGGACCCGGACCTCCGGCTAGCCTGTTTCAGCCACCTCGTCGTCCTGG
CCTTGGAACCTGTTGGAAAACCAATTGCACTGTTAGCCAATCATTTTCAGGTTGAGATTCCCTA
AAATAGATGTGTATCACTATGATGTGGATATTAAGCCTGAAAAACGGCCTCGTAGAGTCAAC
AGGGAGGTAGTAGATACAATGGTGCGGCACTTCAAGATGCAAATATTTGGTGATCGGCAGCC
TGGGTATGATGGCAAAGAAACATGTACACAGCACATCCACTACCAATTGGACGGGATAGGG
TTGATATGGAGGTGACTCTTCCAGGCGAGGGTAAAGACCAAACATTTAAAGTGCTGTTCAG
TGGGTGTCAGTTGTGAGCCTTCAGTTGCTTTTAGAAGCTTTGGCTGGGCACTTGAATGAAGT
CCCAGATGACTCAGTACAAGCACTTGATGTTATCACAAGACACCTTCCCTCCATGAGGTACA
CCCCAGTGGGCGGTTCCCTTTTCTCACCCCCGGAAGGTTACTACCACCTCTGGGAGGGGGC
AGGGAGGTCTGGTTTGGTTTTCATCAGTCTGTGAGACCTGCCATGTGGAATATGATGCTCAA
CATTGATGTATCTGCAACTGCTTTCTACCGGGCTCAGCCTATCATTGAGTTCATGTGTGAGG
TTTGTAGACATTCAGAACATCAATGAACAGACCAAACCTCTAACAGACTCCCAGCGTGTCAAA
TTTACCAAAGAAATCAGAGGTCTCAAAGTTGAGGTGACCCACTGTGGACAGATGAAACGAAA
ATACCGAGTTTGTAAATGTGACTAGACGGCCAGCCAGTCATCAAACCTTTTCCCTTGCAGCTAG
AAAACGGTCAAGCTATGGAATGTACAGTAGCTCAATATTTTAAGCAAAAGTATAGTCTGCAA
CTGGAATACCCCATCTTCCCTGTCTCCAAGTGGGACAAGAACAAGCAATACATACTTGCC
ACTCGAGGTCTGTAATATAGTGGCAGGACAGCGATGTATCAAGAAGCTCACAGACAATCAGA
CTTCCACAATGATCAAAGCTACAGCAAGATCTGCTCCTGACAGACAGGAAGAGATCAGTAGA
CTGGTGAAGAGCAACAGTATGGTGGGTGGACCTGATCCATACCTTAAAGAAATTTGGTATTGT
TGTCCACAATGAAATGACAGAGCTCACAGGCAAGGTTACTTCCAGCACCAATGCTGCAATATG
GAGGCCGGAATAAAACAGTAGCCACACCCAACAGGGTGTCTGGGACATGCGAGGAAAGCAG
TTTTATGCTGGCATTGAAATTAAGTTTGGGCAGTTGCTTGTTTTGCACCTCAGAAACAATG
TAGGAAGATTTACTAAAGAGTTTCACTGACCAAGCTGCGTAAATCTCTAAGGATGCAGGAA
TGCCCATCCAGGGTCAGCCATGTTTCTGCAAGTATGCACAAGGTGCAGACAGTGTGGAGCCT
ATGTTTAAACATCTGAAAATGACTTATGTGGGCCTACAGCTAATAGTGGTTATCCTGCCTGG
AAAGACACCAGTATATGCGGAGGTGAAACGTGTTGGAGATACCCTTCTAGGTATGGCCACAC
AGTGTGTCCAGGTAAAAATGTAGTGAAGACCTCACCTCAAACCTTTCCAATCTTTGCCTG
AAGATAAATGCAAACTTGGAGGAATTAACAATGTGCTTGTGCCTCATCAAAGGCCCTCGGT
GTTCCAGCAGCCTGTCACTTCTCTGGGAGCGGATGTCACACACCCCCAGCAGGGGATGGGA
AGAAACCTTCCATTGCTGTGCTGTGGTGGCAGTATGGATGGCCACCCAGCCGGTACTGTGCC
ACCGTTCCGGTGCAGACTTCCCGGCAGGAGATCTCCAAGAGCTCCTCTACAGTCAAGAGGT
CATCCAGGACCTGACTAACATGGTTCGAGAGCTGCTGATTCAAGTCTACAAATCCACACGCT
TCAAACCCACTCGGATCATCTATTACCGTGGAGGGGTATCTGAGGGACAAATGAAACAGGTA
GCTTGGCCAGAACTAATAGCAATTGAAAGGCATGTATTAGCTTGGAAAGAGATTACCGGCC
AGGAATAACTTATATTGTGGTGCAAAAAGACATCACACACGACTCTTCTGTGCAGATAAAA
CAGAAAGGGTAGGGAAAAGTGGCAATGTACCAGCAGGCACTACAGTGGATAGTACCATCACA
CATCCATCTGAGTTTGACTTTTACCTCTGTAGTCATGCAGGAATTCAGGGAACAGCCGTCC
CTCATTACCAAGGTCTTGTGGGATGACAACCTGCTTCACTGCAGATGAACTCCAGCTACTGA
CTTACCAGCTGTGTACACCTATGTGAGGTGCACTCGCTCAGTCTCTATTCCAGCCCCCTGCA
TATTATGCCCGGCTTGTAGCATTTAGGGCAAGGTATCATCTGGTGGATAAAGATCATGACAG
TGCGGAAGGCAGTCATGTGTGAGGACAGAGCAACGGCCGGGATCCTCAGGCCTTGGCTAAGG
CTGTGCAATCCACCATGATACCCAGCACACGATGTATTTTGCC
```

Figure 16

>eIF2C4, cDNA sequence of predicted ORF
GCAGGACCCGCTGGGGCCCCAGCCCCCTACTCATGGTGCCAGAAAGACCTGGCTATGGCACCAT
GGGCAAACCCATTAAACTGCTGGCTAACTGTTTTCAAGTTGAAATCCCAAAGATTGATGTCT
ACCTCTATGAGGTAGATATTAAACCAGACAAGTGTCTTAGGAGAGTGAACAGGGAGGTGGTT
GACTCAATGGTTCAGCATTTTAAAGTAAGTATATTTGGAGACCGTAGACCAGTTTATGATGG
AAAAAGAAGTCTTTACACCGCCAATCCACTTCCTGTGGCAACTACAGGGGTAGATTTAGACG
TTACTTTACCTGGGGAAGGTGGAAAAGATCGACCTTTCAAGGTGTCAATCAAATTTGTCTCT
CGGGTGAGTTGGCACCTACTGCATGAAGTACTGACAGGACGGACCTTGCCCTGAGCCACTGGA
ATTAGACAAGCCAATCAGCACTAACCCTGTCCATGCCGTTGATGTGGTGCTACGACATCTGC
CCTCCATGAAATACACACCTGTGGGGCGTTCATTTTCTCCGCTCCAGAAGGATATGACCAC
CCTCTGGGAGGGGGCAGGGGAAGTGTGGTTTGGATTCCATCAGTCTGTTCCGGCTGCCATGTG
GAAAATGATGCTTAATATCGATGTTTCTGCCACTGCCTTCTACAAAGCACAACTGTAAATTC
AGTTCATGTGTGAAGTCTTGATATTCATAATATTGATGAGCAACCAAGACCTCTGACTGAT
TCTCATCGGGTAAAATTCACCAAAGAGATAAAAGGTTTGAAGGTTGAAGTGACTCATTGTGG
ACAATGAGACGGAAATACCGTGTGTGTAATGTAACAAGGAGGCCTGCCAGTCATCAAACCT
TTCCTTTACAGTTAGAAAACGGCCAAACTGTGGAGAGAACAGTAGCGCAGTATTTACAGAGAA
AAGTATACTCTTCAGCTGAAGTACCCGCACCTTCCCTGTCTGCAAGTCGGGCAGGAACAGAA
ACACACCTACCTGCCACTAGAAAGTCTGTAATATTGTGGCAGGGCAACGATGTATCAAGAAAGC
TAACAGACAATCAGACTTCCACTATGATCAAGGCAACAGCAAGATCTGCACCAGATAGACAA
GAGGAAATTAGCAGATTGGTAAGAAAGTGCAAATTATGAAACAGATCCATTTGTTTCAGGAGTT
TCAATTTAAAGTTCGGGATGAAATGGCTCATGTAAGTGGACGCGTACTTCCAGCACCTATGC
TCCAGTATGGAGGACGGAATCGGACAGTAGCAACACCGAGCCATGGAGTATGGGACATGCCGA
GGGAAACAATTCACACAGGAGTTGAAATCAAATGTGGGCTATCGCTTGTTTTGCACACA
GAGGCAGTGCAGAGAAGAAATATTGAAGGGTTTACAGACCAGCTGCGTAAGATTTCTAAGG
ATGCAGGGATGCCCATCCAGGGCCAGCCATGCTTCTGCAAATATGCACAGGGGGCAGACAGC
GTAGAGCCCATGTTCCGGCATCTCAAGAACACATATTCTGGCCTACAGCTTATTATCGTCAT
CCTGCCGGGGAAGACACCAGTGTATGCCGAAGTGAAACGTGTAGGAGACACACTTTTGGGTA
TGGCTACACAATGTGTTCAAGTCAAGAATGTAATAAAAACATCTCCTCAAACCTCTGTCAAAC
TTGTGCCATAAGATAAATGTTAAACTCGGAGGGGATCAATAATATTCTTGTACCTCATCAAAG
ACCTTCTGTGTTCCAGCAACCAGTGATCTTTTTGGGAGCCGATGTCACTCATCCACCTGCTG
GTGATGGAAAGAAGCCTTCTATTGCTGCTGTTGTAGGTAGTATGGATGCACACCCAAGCAGA
TACTGTGCCACAGTAAGAGTTCAGAGACCCCGACAGGAGATCATCCAGGACTTGGCCTCCAT
GGTCCGGGAACCTTCTTATTCAATTTTATAAGTCAACTCGGTTCAAGCCTACTCGTATCATCT
TTTATCGGGATGGTGTTCAGAGGGGCAGTTTAGGCAGGTATTATATTATGAACTACTAGCA
ATTCGAGAAGCCTGCATCAGTTTGGAGAAAGACTATCAACCTGGAATAACCTACATTGTAGT
TCAGAAGAGACATCACACTCGATTATTTTGTGCTGATAGGACAGAAAGGTTGGAAGAAGTG
GCAATATCCCAGCTGGAACAACAGTTGATACAGACATTACACACCCATATGAGTTTCGATTTT
TACCTCTGTAGCCATGCTGGAATACAGGGTACCAGTCGTCCTTCACACTATCATGTTTTATG
GGATGATAACTGCTTTACTGCAGATGAACTTCAGCTGCTAACTTACCAGCTCTGCCACACTT
ACGTACGCTGTACACGATCTGTTTCTATACCTGCACCAGCGTATTATGCTCACCTGGTAGCA
TTTAGAGCCAGATATCATCTTGTGGACAAAGAACATGACAGTGTGAAGGAAGTCACGTTTC
AGGACAAAGCAATGGGCGAGATCCACAAGCTCTTGCCAAGGCTGTACAGATTCACCAAGATA
CCTTACGCACAATGTACTTCGCTTAA

Figure 16

>HILI, cDNA sequence of predicted ORF
ATATCTTCTGGTGATGCTGGAAGTACCTTCATGGAAAAGAGGTGTGAAAAACAAACAGGACTT
TATGGATTTTGTAGTATCTGTACCAGAGAAAAATTGGCAGATGTGAGAAATTGTAAAAACAGGTT
CCAGTGGAATACCTGTGAACTGGTTACAAACCTCTTTAACTTAGATTTTCCCCAAGACTGG
CAGCTATACCAGTACCATGTGACATATATTCAGATTTAGCATCTAGAAGGCTGAGAATTGC
TTTACTTTTATAGTCATAGTGAACCTTTCCAACAAAGCAAAAGCATTTCGACGGTGCCATCCTTT
TTCCTGTACAAAAAGCTAGAAGAAAAGGTCACAGAGTTGTCAAGTGAAACTCAAAGAGGTGAG
ACTATAAAGATGACTATCACCTGAAGAGGGAGCTGCCATCAAGTTCTCCCGTGTGCATCCA
GGTCTTCAATATCATCTTCAGAAAGATCCTCAAAAAGTTGTCCATGTACCAAATTGGACGGA
ACTTCTATAATCCTTCAGAGCCAATGGAAATTCCCCAGCACAAATTATCCCTTTGGCCTGGG
TTTGCCATTTCTGTGTATATTTTGAAAGGAAGCTCCTGTTTAGTGTCTGATGTGAGTTACAA
AGTCCTCCGGAATGAGACGGTTCTGGAATTCATGACTGCTCTCTGTCAAAGAACTGGCTTGT
CCTGTTTTACCCAGACGTGTGAGAAGCAGCTAATAGGGCTCATTGTCCTTACAAGATACAAT
AACAGAACCTACTCCATTGATGACATTCAGTGGTCAGTGAAGCCACACACACCTTTCAGAA
GCGGGATGGCACCGAGATCACCTATGTGGATTACTACAAGCAGCAGTATGATATTACTGTAT
CGGACCTGAATCAGCCCATGCTTGTGTAGTCTGTTAAAGAAGAAGAGAAATGACAACAGTGAG
GCTCAGCTCGCCACCTGATACCTGAGCTCTGCTTTCTAACAGGGCTGACTGACCAGGCAAC
ATCTGATTTCCAGCTGATGAAGGCTGTGGCTGAAAAGACACGTCTCAGTCCTTCAGGCCGGC
AGCAGCGCCTGGCCAGGCTTGTGGACAACATCCAGAGGAATACCAATGCTCGCTTTGAACTA
GAGACCTGGGGAGTGCATTTTGGAAGCCAGATATCTCTGACTGGCCGGATTGTGCCTTCAGA
AAAAATATTAATGCAAGACCACATATGTCAACCTGTGTCTGCTGCTGACTGGTCCAAGGATA
TTCGAACCTTGCAAGATTTTAAATGCACAGTCTTTGAATACCTGGTTGATTTTATGTAGCGAC
AGAACTGAATATGTTGCCGAGAGCTTTCTGAACTGCTTGAGAAGAGTTGCAGGTTCATGGG
ATTTAATGTAATGTGCATTCCTGCCCTTCTAATCAGAAGACCTATTATGATTCCATTAAAAAAT
ATTTGAGCTCAGACTGCCCAGTCCCAAGCCAATGTGTGCTTGCTCGGACCTTGAATAAACAG
GGCATGATGATGAGTATCGCCACCAAGATCGCTATGCAGATGACTTGCAAGCTCGGAGGCCGA
GCTGTGGGCTGTGGAAATACCTTTAAAGTCCCTGATGGTGGTTCGGTATTGATGTCTGTAAAG
ATGCACCTCAGCAAGGACGTGATGGTTGTTGGATGCGTGGCCAGTGTTAACCCAGAAATCACC
AGGTGGTTTTTCCCGCTGTATCCTTCAGAGAACAATGACTGATGTTGCAGATTGCTTGAAAGT
TTTCATGACTGGAGCACTCAACAAATGGTACAAGTACAATCATGATTTGCCAGCACGGATAA
TTGTGTACCGTGTCTGGTGTAGGGGATGGTCAGCTGAAAACACTTATTGAATATGAAGTCCCA
CAGCTGCTGAGCAGTGTGGCAGAATCCAGCTCAAATACCAGCTCAAGACTGTCGGTGATTGT
GGTCAGGAAGAAGTGCATGCCACGATTCTTTACCGAAATGAACCGCACTGTACAGAACCCCC
CACTTGGCACTGTTGTGGATTGAGAAGCAACACGTAACGAATGGCAGTATGACTTTTATCTG
ATCAGCCAGGTGGCCTGCCGGGGAAGTGTAGTCTTACCTACTATAATGTCATCTATGATGA
CAACGGCTTGAAAGCCCGACCATATGCAGAGACTTACATTCAAATTGTGCCACCTGTACTACA
ACTGGCCGGGCATAGTCAGTGTCCAGCACCATGTGAGTATGCTCACAAGCTGACCTTTCTG
GTGGCACAAAGCATTCATAAAGAACCAGTCTGGAATTAGCCAACCATCTCTTCTACCTG

Figure 16

>HIWI, cDNA sequence of predicted ORF

ATGACTGGGAGAGCCCGAGCCAGAGCCAGAGGAAGGGCCCGGGTCAGGAGACAGCGCAGCT
GGTGGGCTCCACTGCCAGTCAGCAACCTGGTTATATTCAGCCTAGGCCTCAGCCGCCACCAG
CAGAGGGGAATTATTTGGCCGTGGACGGCAGAGAGGAACAGCAGGAGGAACAGCCAAGTCA
CAAGGACTCCAGATATCTGCTGGATTTCAGGAGTTATCGTTAGCAGAGAGAGGAGGTCGTCG
TAGAGATTTTCATGATCTTGGTGTGAATACAAGGCAGAACCTAGACCATGTTAAAGAATCAA
AAACAGGTTCTTCAGGCATTATAGTAAGGTTAAGCACTAACCATTTCCGGCTGACATCCCGT
CCCCAGTGGGCCCTTATATCAGTATCACATTGACTATAACCCACTGATGGAAGCCAGAAGACT
CCGTTACAGCTCTTCTTTTCAACACGAAGATCTAATTGGAAAGTGCCATGCTTTTGTATGGAA
CGATATTATTTTACCTAAAAGACTACAGCAAAAGGTTACTGAAGTTTTTAGTAAGACCCGG
AATGGAGAGGATGTGAGGATAACGATCACTTTAACAAATGAACTTCCACCTACATCACCAAC
TTGTTTGCAGTTCTATAATATTATTTTCAGGAGGCTTTTGAAAATCATGAATTTGCAACAAA
TTGGACGAAATTATTATAACCCAAATGACCAATTGATATTCCAAGTCACAGGTTGGTGATT
TGGCCTGGCTTCACTACTTCCATCCTTCAGTATGAAAACAGCATCATGCTCTGCCTGACGT
TAGCCATAAAGTCTTCTGAAGTGAGACTGTTTTGGATTTTCATGTTCAACTTTTATCATCAGA
CAGAAGAACATAAATTTCAAGAACAAGTTTCCAAAGAATAATAGGTTTAGTTGTTCTTACC
AAGTATAACAATAAGACATACAGAGTGGATGATATTGACTGGGACCAGAATCCCAAGAGCAC
CTTTAAGAAAAGCCGACGGCTCTGAAGTCAGCTTCTTAGAATACTACAGGAAGCAATACAACC
AAGAGATCACCGACTTGAAGCAGCCTGTCTTGGTCAGCCAGCCCAAGAGAAGCGGGGCCCT
GGGGGACACTGCCAGGGCCTGCCATGCTCATTCTGAGCTCTGCTATCTTACAGGTCCTAAC
TGATAAAATGCGTAATGATTTTAACGTGATGAAAGACTTAGCCGTTCATACAAGACTAACCTC
CAGAGCAAAGGCAGCGTGAAGTGAGGACGACTCATTGATTACATTCAAAAAACGATAATGTT
CAAAGGGAGCTTCGAGACTGGGGTTTGAGCTTTGATTCCAACCTACTGTCTTCTCAGGAAG
AATTTTGCAAACAGAAAAGATTCACCAAGGTGGAAAAACATTTGATTACAATCCACAATTTG
CAGATTGGTCCAAAGAAACAAGAGGTGCACCATTAAATTAGTGTTAAGCCACTAGATAACTGG
CTGTTGATCTATACGCGAAGAAATTATGAAGCAGCCAATTCATTGATACAAAATCTATTTAA
AGTTACACCAGCCATGGGCATGCAAATGAGAAAAGCAATAATGATTGAAGTGGATGACAGAA
CTGAAGCCTACTTAAGAGTCTTACAGCAAAAGGTCACAGCAGACACCCAGATAGTTGTCTGT
CTGTTGTCAAGTAATCGGAAGGACAAATACGATGCTATTAAAAAATACCTGTGTACAGATTG
CCCTACCCCAAGTCAGTGTGTGGTGGCCCGAACCTTAGGCAAACAGCAAACCTGTCATGGCCA
TTGCTACAAAGATTGCCCTACAGATGAACTGCAAGATGGGAGGAGAGCTCTGGAGGGTGGAC
ATCCCCCTGAAGCTCGTGATGATCGTTGGCATCGATTGTTACCATGACATGACAGCTGGGCG
GAGGTCAATCGCAGGATTTGTTGCCAGCATCAATGAAGGGATGACCCGCTGGTTCTCACGCT
GCATATTTCAGGATAGAGGACAGGAGCTGGTAGATGGGCTCAAAGTCTGCCTGCAAGCGGCT
CTGAGGGCTTGGAATAGCTGCAATGAGTACATGCCAGCCGGATCATCGTGTACCGCGATGG
CGTAGGAGACGGCCAGCTGAAAACACTGGTGAACCTACGAAGTGCCACAGTTTTTGGATTGTC
TAAAATCCATTGGTAGAGGTTACAACCTAGACTAACGGTAATTGTGGTGAAGAAAAGAGTG
AACACCAGATTTTTTGTCTCAGTCTGGAGGAAGACTTCAGAATCCACTTCCTGGAACAGTTAT
TGATGTAGAGGTTACCAGACCAGAATGGTATGACTTTTTTATCGTGAGCCAGGCTGTGAGAA
GTGGTAGTGTTTCTCCACACATTACAATGTCTATGACAACAGCGGCTGAAGCCAGAC
CACATACAGCGCTTGACCTACAAGCTGTGCCACATCTATTACAACCTGGCCAGGTGTCATTG
TGTTCTCTGCTCCTTGCCAGTACGCCACAAGCTGGCTTTTCTTGTGGCCAGAGTATTCACA
GAGAGCCAAATCTGTCACTGTCAAACCGCCTTTACTACCTC

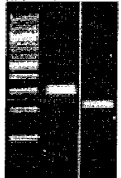
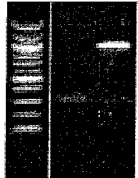
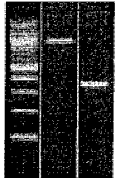

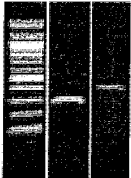
Figure 17

A

Gene name	1 st primer pair (5'-3')	2 nd primer pair (5'-3')	Expected length (bp)
eIF2C1	GAGGTCTGTAACATTGTGGC*	GAGGTCTGTAACATTGTGGC*	287
	CGGTAGAAGATGATGCGGGT	AAGTTCTTGAGCACCTCTTCTCGA	
	GAGGTCTGTAACATTGTGGC	CCACACCAGCGCTCTGCC	207
	CGGTAGAAGATGATGCGGGT	CTCACGCACCATGTAGGA	
eIF2C2	GAGGTCTGTAACATTGTGGC	ATCCTGCTGCCCCAAGGG	186
	CGGTAGAAGATGATGCGGGT	GATCTCCTGCCGGTGCTG	
	GAGGTCTGTAACATTGTGGC*	GAGGTCTGTAACATTGTGGC*	891
	CGGTAGAAGATGATGCGGGT	GATCTCCTGCCGGTGCTG	
eIF2C3	AGAGCAACAGTATGGTGGGTGGAC	CCTCTACAGTCAAGAGGT	334
	TGGATGTGTGATGGTACT*	TGGATGTGTGATGGTACT*	
	CACCTGAATGAAGTCCCA	AGAGCAACAGTATGGTGGGTGGAC	808
	TCCTGGATGACCTCTTGACTGTAG*	TCCTGGATGACCTCTTGACTGTAG*	
eIF2C4	TCCGGCATCTCAAGAACACATATTCT	ATCCAGGACTTGGCCTCC	324
	GAAGTCAATATGGGTGTGTAATGTCTG*	GAAGTCAATATGGGTGTGTAATGTCTG*	
HILI	CAGCACAAATTATCCCTT*	CAGCACAAATTATCCCTT*	264
	CGGCCTGAAGGACTGAGACGTGT	GTGTGTGGGCTTCACTGA	
	TCTCTGTCAAAGAACTGGCTTGTCT*	TCTCTGTCAAAGAACTGGCTTGTCT*	393
	CTGTACAGTGCGGTTCAT	CGGCCTGAAGGACTGAGACGTGT	

* primers used in both reactions (semi-nested PCR)

B

Gene name	eIF2C1		eIF2C2		eIF2C3		eIF2C4	HILI	
Expected length (bp)	287	207	186	891	808	334	324	264	393
PCR products									

RNA-INTERFERENCE BY SINGLE-STRANDED RNA MOLECULES

[0001] This application is a divisional of U.S. Ser. No. 10/520,470 filed Jan. 7, 2005, which is a 35 U.S.C. 371 National Phase Entry Application from PCT/EP2003/007516, filed Jul. 10, 2003, which claims the benefit of European Patent Application Nos. 02015532.1 filed Jul. 10, 2002 and 02018906.4 filed Aug. 23, 2002, the disclosures of which is incorporated herein in their entirety by reference.

DESCRIPTION

[0002] The present invention relates to sequence and structural features of single-stranded (ss)RNA molecules required to mediate target-specific nucleic acid modifications by RNA-interference (RNAi), such as target mRNA degradation and/or DNA methylation.

[0003] Most eukaryotes possess a cellular defense system protecting their genomes against invading foreign genetic elements. Insertion of foreign elements is believed to be generally accompanied by formation of dsRNA that is interpreted by the cell as a signal for unwanted gene activity (e.g. Ahlquist, Science 296 (2002), 1270-1273; Fire et al., Nature 391 (1998), 806-811). Dicer RNase III rapidly processes dsRNA to small dsRNA fragments of distinct size and structure (e.g. Bernstein et al., Nature 409 (2001), 363-366), the small interfering RNAs (siRNAs) (Elbashir et al., Genes & Dev. 15 (2001 b), 188-200), which direct the sequence-specific degradation of the single-stranded mRNAs of the invading genes. siRNA duplexes have 2- to 3-nt 3' overhanging ends and contain 5' phosphate and free 3' hydroxyl termini (WO 02/44321). The process of posttranscriptional dsRNA-dependent gene silencing is commonly referred to as RNA interference (RNAi), and in some instances is also linked to transcriptional silencing.

[0004] Experimental introduction of siRNA duplexes into mammalian cells is now widely used to disrupt the activity of cellular genes homologous in sequence to the introduced dsRNA. Used as a reverse genetic approach, siRNA-induced gene silencing accelerates linking of gene sequence to biological function. siRNA duplexes are short enough to bypass general dsRNA-induced unspecific effects in vertebrate animal and mammalian cells. siRNAs may also be expressed intracellularly from introduced expression plasmids or viral vectors providing an alternative to chemical RNA synthesis. Therefore, an understanding of how siRNAs act in mammalian systems is important for refining this gene silencing technology and for producing gene-specific therapeutic agents.

[0005] Biochemical studies have begun to unravel the mechanistic details of RNAi. The first cell-free systems were developed using *D. melanogaster* cell or embryo extracts, and were followed by the development of in vitro systems from *C. elegans* embryo and mouse embryonal carcinoma cells. While the *D. melanogaster* lysates support the steps of dsRNA processing and sequence-specific mRNA targeting, the latter two systems only recapitulate the first step.

[0006] RNAi in *D. melanogaster* extracts is initiated by ATP-dependent processing of long dsRNA to siRNAs by Dicer RNase III (e.g. Bernstein et al., (2001), supra). Thereafter, siRNA duplexes are assembled into a multi-component complex, which guides the sequence-specific recognition of the target mRNA and catalyzes its cleavage (e.g. Elbashir

(2001 b), supra). This complex is referred to as RNA-induced silencing complex (RISC) (Hammond et al., Nature 404 (2000), 293-296). siRNAs in *D. melanogaster* are predominantly 21- and 22-nt, and when paired in a manner to contain a 2-nt 3' overhanging structure effectively enter RISC (Elbashir et al., EMBO J. 20 (2001 c), 6877-6888). Mammalian systems have siRNAs of similar size, and siRNAs of 21- and 22-nt also represent the most effective sizes for silencing genes expressed in mammalian cells (e.g. Elbashir et al., Nature 411 (2001 a), 494-498, Elbashir et al., Methods 26 (2002), 199-213).

[0007] RISC assembled on siRNA duplexes in *D. melanogaster* embryo lysate targets homologous sense as well as antisense single-stranded RNAs for degradation. The cleavage sites for sense and antisense target RNAs are located in the middle of the region spanned by the siRNA duplex. Importantly, the 5'-end, and not the 3' end, of the guide siRNA sets the ruler for the position of the target RNA cleavage. Furthermore, a 5' phosphate is required at the target-complementary strand of a siRNA duplex for RISC activity, and ATP is used to maintain the 5' phosphates of the siRNAs (Nykänen et al., Cell 107 (2001), 309-321). Synthetic siRNA duplexes with free 5' hydroxyls and 2-nt 3' overhangs are so readily phosphorylated in *D. melanogaster* embryo lysate that the RNAi efficiencies of 5'-phosphorylated and non-phosphorylated siRNAs are not significantly different (Elbashir et al. (2001 c), supra).

[0008] Unwinding of the siRNA duplex must occur prior to target RNA recognition. Analysis of ATP requirements revealed that the formation of RISC on siRNA duplexes required ATP in lysates of *D. melanogaster*. Once formed, RISC cleaves the target RNA in the absence of ATP. The need for ATP probably reflects the unwinding step and/or other conformational rearrangements. However, it is currently unknown if the unwound strands of an siRNA duplex remain associated with RISC or whether RISC only contains a single-stranded siRNA.

[0009] A component associated with RISC was identified as Argonaute2 from *D. melanogaster* Schneider 2 (S2) cells (Hammond et al., Science 293 (2001 a), 1146-1150), and is a member of a large family of proteins. The family is referred to as Argonaute or PPD family and is characterized by the presence of a PAZ domain and a C-terminal Piwi domain, both of unknown function (Cerutti et al., Trends Biochem. Sci. (2000), 481-482); Schwarz and Zamore, Genes & Dev. 16 (2002), 1025-1031). The PAZ domain is also found in Dicer. Because Dicer and Argonaute2 interact in S2 cells, PAZ may function as a protein-protein interaction motif. Possibly, the interaction between Dicer and Argonaute2 facilitates siRNA incorporation into RISC. In *D. melanogaster*, the Argonaute family has five members, most of which were shown to be involved in gene silencing and development. The mammalian members of the Argonaute family are poorly characterized, and some of them have been implicated in translational control, microRNA processing and development. The biochemical function of Argonaute proteins remains to be established and the development of more biochemical systems is crucial.

[0010] Here we report on the analysis of human RISC in extracts prepared from HeLa cells. The reconstitution of RISC and the mRNA targeting step revealed that RISC is a ribonucleoprotein complex that is composed of a single-stranded siRNA. Once RISC is formed the incorporated siRNA can no longer exchange with free siRNAs. Surprisingly, RISC can be reconstituted in HeLa S100 extracts pro-

viding single-stranded siRNAs. Introducing 5' phosphorylated single-stranded antisense siRNAs into HeLa cells potently silences an endogenous gene with similar efficiency than duplex siRNA.

[0011] The object underlying the present invention is to provide novel agents capable of mediating target-specific RNAi.

[0012] The solution of this problems is provided by the use of a single-stranded RNA molecule for the manufacture of an agent for inhibiting the expression of said target transcript. Surprisingly, it was found that single-stranded RNA molecules are capable of inhibiting the expression of target transcripts by RNA-interference (RNAi).

[0013] The length of the single-stranded RNA molecules is preferably from 14-50 nt, wherein at least the 14 to 20 5'-most nucleotides are substantially complementary to the target RNA transcript. The RNA oligonucleotides may have a free 5' hydroxyl moiety, or a moiety which is 5' phosphorylated (by means of chemical synthesis or enzymatic reactions) or which is modified by 5'-monophosphate analogues.

[0014] The inhibition of target transcript expression may occur in vitro, e.g. in eucaryotic, particularly mammalian cell cultures or cell extracts. On the other hand, the inhibition may also occur in vivo i.e. in eucaryotic, particularly mammalian organisms including human beings.

[0015] Preferably, the single-stranded RNA molecule has a length from 15-29 nucleotides. The RNA-strand may have a 3' hydroxyl group. In some cases, however, it may be preferable to modify the 3' end to make it resistant against 3' to 5' exonucleases. Tolerated 3'-modifications are for example terminal 2'-deoxy nucleotides, 3' phosphate, 2',3'-cyclic phosphate, C3 (or C6, C7, C12) aminolinker, thiol linkers, carboxyl linkers, non-nucleotidic spacers (C3, C6, C9, C12, abasic, triethylene glycol, hexaethylene glycol), biotin, fluoresceine, etc.

[0016] The 5'-terminus comprises an OH group, a phosphate group or an analogue thereof. Preferred 5' phosphate modifications are 5'-monophosphate $((\text{HO})_2(\text{O})\text{P}-\text{O}-5')$, 5'-diphosphate $((\text{HO})_2(\text{O})\text{P}-\text{O}-\text{P}(\text{HO})(\text{O})-\text{O}-5')$, 5'-triphosphate $((\text{HO})_2(\text{O})\text{P}-\text{O}-\text{P}(\text{HO})(\text{O})\text{P}-\text{O}-\text{P}(\text{HO})(\text{O})-\text{O}-5')$, 5'-guanosine cap (7-methylated or non-methylated) $(7\text{m-G-O}-5'-(\text{HO})(\text{O})\text{P}-\text{O}-\text{P}(\text{HO})(\text{O})\text{P}-\text{O}-\text{P}(\text{HO})(\text{O})-\text{O}-5')$, 5'-adenosine cap. (Aapp), and any modified or unmodified nucleotide cap structure $(\text{N}-\text{O}-5'-(\text{HO})(\text{O})\text{P}-\text{O}-\text{P}(\text{HO})(\text{O})\text{P}-\text{O}-\text{P}(\text{HO})(\text{O})-\text{O}-5')$, 5'-monothiophosphate (phosphorothioate; $(\text{HO})_2(\text{S})\text{P}-\text{O}-5'$), 5'-monodithiophosphate (phosphorodithioate; $(\text{HO})(\text{HS})(\text{S})\text{P}-\text{O}-5'$), 5'-phosphorothiolate $((\text{HO})_2(\text{O})\text{P}-\text{S}-5')$; any additional combination of oxygen/sulfur replaced monophosphate, diphosphate and triphosphates (e.g. 5'-alpha-thiotriphosphate, 5'-gamma-thiotriphosphate, etc.), 5'-phosphoramidates $((\text{HO})_2(\text{O})\text{P}-\text{NH}-5')$, $(\text{HO})(\text{NH}_2)(\text{O})\text{P}-\text{O}-5'$), 5'-alkylphosphonates $(\text{R}=\text{alkyl}=\text{methyl, ethyl, isopropyl, propyl, etc., e.g. RP}(\text{OH})(\text{O})-\text{O}-5', (\text{OH})_2(\text{O})\text{P}-5'-\text{CH}_2-)$, 5'-alkyletherphosphonates $(\text{R}=\text{alkylether}=\text{methoxymethyl (MeOCH}_2-), \text{ethoxymethyl, etc., e.g. RP}(\text{OH})(\text{O})-\text{O}-5'-)$.

[0017] The sequence of the RNA molecule of the present invention has to have a sufficient identity to a nucleic acid target molecule in order to mediate target-specific RNAi. Thus the single-stranded RNA molecule of the present invention is substantially complementary to the target transcript.

[0018] The target RNA cleavage reaction guided by the single-stranded RNA molecules of the present invention is highly sequence-specific. However, no all positions of the

RNA molecule contribute equally to target recognition. Mismatches, particularly at the 3'-terminus of the single-stranded RNA molecule, more particularly the residues 3' to the first 20 nt of the single-stranded RNA molecule are tolerated. Especially preferred are single-stranded RNA molecules having at the 5'-terminus at least 15 and preferably at least 20 nucleotides which are completely complementary to a predetermined target transcript or have at only mismatch and optionally up to 35 nucleotides at the 3'-terminus which may contain 1 or several, e.g. 2, 3 or more mismatches.

[0019] In order to enhance the stability of the single-stranded RNA molecules, the 3'-ends may be stabilized against degradation, e.g. they may be selected such that they consist of purine nucleotides, particularly adenosine or guanosine nucleotides. Alternatively or additionally, 3' nucleotides may be substituted by modified nucleotide analogues, including backbone modifications of ribose and/or phosphate residues.

[0020] In an especially preferred embodiment of the present invention the RNA molecule may contain at least one modified nucleotide analogue. The nucleotide analogues may be located at positions where the target-specific activity, e.g. the RNAi mediating activity is not substantially affected, e.g. in a region at the 5'-end and/or the 3'-end of the RNA molecule. Particularly, the 3'-terminus may be stabilized by incorporating modified nucleotide analogues, such as non-nucleotidic chemical derivatives such as C3 (or C6, C7, C12) aminolinker, thiol linkers, carboxyl linkers, non-nucleotidic spacers (C3, C6, C9, C12, abasic, triethylene glycol, hexaethylene glycol), biotin, fluoresceine, etc. A further modification, by which the nuclease resistance of the RNA molecule may be increased, is by covalent coupling of inverted nucleotides, e.g. 2'-deoxyribonucleotides or ribonucleotides to the 3'-end of the RNA molecule. A preferred RNA molecule structure comprises: 5'-single-stranded siRNA-3'-O-P(O)(OH)-O-3'-N, wherein N is a nucleotide, e.g. a 2'-deoxyribonucleotide or ribonucleotide, typically an inverted thymidine residue, or an inverted oligonucleotide structure, e.g. containing up to 5 nucleotides.

[0021] Preferred nucleotide analogues are selected from sugar- or backbone-modified ribonucleotides. It should be noted, however, that also nucleobase-modified ribonucleotides, i.e. ribonucleotides, containing a non-naturally occurring nucleobase instead of a naturally occurring nucleobase such as uridines or cytidines modified at the 5-position, e.g. 5-(2-amino)propyl uridine, 5-bromo uridine; 5-methyl-cytidine; adenosines and guanosines modified at the 8-position, e.g. 8-bromo guanosine; deaza nucleotides, e.g. 7-deaza-adenosine; O- and N-alkylated nucleotides, e.g. N6-methyl adenosine are suitable. In preferred sugar-modified ribonucleotides the 2' OH-group is replaced by a group selected from H, OR, R, halo, SH, SR, NH_2 , NHR, NR_2 or CN, wherein R is $\text{C}_1\text{-C}_6$ alkyl, alkenyl, alkynyl or methoxyethoxy, and halo is F, Cl, Br or I. In preferred backbone-modified ribonucleotides the phosphoester group connecting to adjacent ribonucleotides is replaced by a modified group, e.g. a phosphorothioate, phosphorodithioate, N3'-O5'- and/or N5'-O3' phosphoramidate group. It should be noted that the above modifications may be combined. For example, complementary or non-complementary nucleotides at the 3'-terminus, particularly after at least 15, more particularly after at least 20 5'-terminal nucleotides may be modified without significant loss of activity.

[0022] The single-stranded RNA molecule of the invention may be prepared by chemical synthesis. Methods of synthesizing RNA molecules are known in the art.

[0023] The single-stranded RNAs can also be prepared by enzymatic transcription from synthetic DNA templates or from DNA plasmids isolated from recombinant bacteria and subsequent 5'-terminal modification. Typically, phage RNA polymerases are used such as T7, T3 or SP6 RNA polymerase.

[0024] A further aspect of the present invention relates to a method of mediating RNA interference in a cell or an organism comprising the steps:

[0025] (a) contacting the cell or organism with the single-stranded RNA molecule of the invention under conditions wherein target-specific nucleic acid modifications may occur and

[0026] (b) mediating a target-specific nucleic acid modification effected by the single-stranded RNA towards a target nucleic acid having a sequence portion substantially complementary to the single-stranded RNA.

[0027] Preferably the contacting step (a) comprises introducing the single-stranded RNA molecule into a target cell, e.g. an isolated target cell, e.g. in cell culture, a unicellular microorganism or a target cell or a plurality of target cells within a multicellular organism. More preferably, the introducing step comprises a carrier-mediated delivery, e.g. by liposomal carriers and/or by injection. Further suitable delivery systems include Oligofectamine (Invitrogen) and Transit-TKO siRNA Transfection reagent (Mirus)

[0028] The method of the invention may be used for determining the function of a gene in a cell or an organism or even for modulating the function of a gene in a cell or an organism, being capable of mediating RNA interference.

[0029] The cell is preferably a eukaryotic cell or a cell line, e.g. a plant cell or an animal cell, such as a mammalian cell, e.g. an embryonic cell, a pluripotent stem cell, a tumor cell, e.g. a teratocarcinoma cell or a virus-infected cell. The organism is preferably a eukaryotic organism, e.g. a plant or an animal, such as a mammal, particularly a human.

[0030] The target gene to which the RNA molecule of the invention is directed may be associated with a pathological condition. For example, the gene may be a pathogen-associated gene, e.g. a viral gene, a tumor-associated gene or an autoimmune disease-associated gene. The target gene may also be a heterologous gene expressed in a recombinant cell or a genetically altered organism. By determining or modulating, particularly, inhibiting the function of such a gene valuable information and therapeutic benefits in the agricultural field or in the medicine or veterinary medicine field may be obtained.

[0031] The ssRNA is usually administered as a pharmaceutical composition. The administration may be carried out by known methods, wherein a nucleic acid is introduced into a desired target cell in vitro or in vivo. Commonly used gene transfer techniques include calcium phosphate, DEAE-dextran, electroporation and microinjection and viral methods (Graham, F. L. and van der Eb, A. J. (1973) *Virology* 52, 456; McCutchan, J. H. and Pagano, J. S. (1968), *J. Natl. Cancer Inst.* 41, 351; Chu, G. et al (1987), *Nucl. Acids Res.* 15, 1311; Fraley, R. et al. (1980), *J. Biol. Chem.* 255, 10431; Capecchi, M. R. (1980), *Cell* 22, 479). A recent addition to this arsenal of techniques for the introduction of nucleic acids into cells is the use of cationic liposomes (Feigner, P. L. et al. (1987), *Proc. Natl. Acad. Sci USA* 84, 7413). Commercially available

cationic lipid formulations are e.g. Tfx 50 (Promega) or Lipofectamin2000 (Life Technologies). A further preferred method for the introduction of RNA into a target organism, particularly into a mouse, is the high-pressure tail vein injection (Lewis, D. L. et al. (2002), *Nat. Genet.* 29, 29; McCaffrey, A. P. et al. (2002), *Nature* 418, 38-39).

[0032] Herein, a buffered solution comprising the single-stranded RNA (e.g. about 2 ml) is injected into the tail vein of the mouse within 10 s.

[0033] Thus, the invention also relates to a pharmaceutical composition containing as an active agent at least one single-stranded RNA molecule as described above and a pharmaceutical carrier. The composition may be used for diagnostic and for therapeutic applications in human medicine or in veterinary medicine.

[0034] For diagnostic or therapeutic applications, the composition may be in form of a solution, e.g. an injectable solution, a cream, ointment, tablet, suspension or the like. The composition may be administered in any suitable way, e.g. by injection, by oral, topical, nasal, rectal application etc. The carrier may be any suitable pharmaceutical carrier. Preferably, a carrier is used, which is capable of increasing the efficacy of the RNA molecules to enter the target-cells. Suitable examples of such carriers are liposomes, particularly cationic liposomes. A further preferred administration method is injection.

[0035] A further preferred application of the RNAi method is a functional analysis of eukaryotic cells, or eukaryotic non-human organisms, preferably mammalian cells or organisms and most preferably human cells, e.g. cell lines such as HeLa or 293 or rodents, e.g. rats and mice. By transfection with suitable single-stranded RNA molecules which are homologous to a predetermined target gene or DNA molecules encoding a suitable single-stranded RNA molecule a specific knockout phenotype can be obtained in a target cell, e.g. in cell culture or in a target organism. The presence of short single-stranded RNA molecules does not result in an interferon response from the host cell or host organism.

[0036] In an especially preferred embodiment, the RNA molecule is administered associated with biodegradable polymers, e.g. polypeptides, poly(D,L-lactic-co-glycolic acid) (PLGA), polylysine or polylysine conjugates, e.g. polylysine-graft-imidazole acetic acid, or poly(beta-amino ester) or microparticles, such as microspheres, nanoparticles or nanospheres. More preferably the RNA molecule is covalently coupled to the polymer or microparticle, wherein the covalent coupling particularly is effected via the 3'-terminus of the RNA molecule.

[0037] Further, the invention relates to a pharmaceutical composition for inhibiting the expression of a target transcript by RNAi comprising as an active agent a single-stranded RNA molecule having a length from 14-50, preferably 15-29 nucleotides wherein at least the 14-20 5'-most nucleotides are substantially complementary to said target transcript.

[0038] Furthermore, the invention relates to a method for the prevention or treatment of a disease associated with over-expression of at least one target gene comprising administering a subject in need thereof a single-stranded RNA molecule having a length from 14-50, preferably 15-29 nucleotides wherein at least the 14-20 5'-most nucleotides are substantially complementary to a target transcript in an amount which is therapeutically effective for RNAi.

[0039] Still, a further subject matter of the invention is a eukaryotic cell or a eukaryotic non-human organism exhibit-

ing a target gene-specific knockout phenotype comprising an at least partially deficient expression of at least one endogenous target gene wherein said cell or organism is transfected with at least one single-stranded RNA molecule capable of inhibiting the expression of at least one endogenous target gene. It should be noted that the present invention allows the simultaneous delivery of several antisense RNAs of different sequences, which are either cognate to a different or the same target gene.

[0040] Gene-specific knockout phenotypes of cells or non-human organisms, particularly of human cells or non-human mammals may be used in analytic procedures, e.g. in the functional and/or phenotypical analysis of complex physiological processes such as analysis of gene expression profiles and/or proteomes. For example, one may prepare the knock-out phenotypes of human genes in cultured cells which are assumed to be regulators of alternative splicing processes. Among these genes are particularly the members of the SR splicing factor family, e.g. ASF/SF2, SC35, SRp20, SRp40 or SRp55. Further, the effect of SR proteins on the mRNA profiles of predetermined alternatively spliced genes such as CD44 may be analysed. Preferably the analysis is carried out by high-throughput methods using oligonucleotide based chips.

[0041] Using RNAi based knockout technologies, the expression of an endogenous target gene may be inhibited in a target cell or a target organism. The endogenous gene may be complemented by an exogenous target nucleic acid coding for the target protein or a variant or mutated form of the target protein, e.g. a gene or a cDNA, which may optionally be fused to a further nucleic acid sequence encoding a detectable peptide or polypeptide, e.g. an affinity tag, particularly a multiple affinity tag. Variants or mutated forms of the target gene differ from the endogenous target gene in that they encode a gene product which differs from the endogenous gene product on the amino acid level by substitutions, insertions and/or deletions of single or multiple amino acids. The variants or mutated forms may have the same biological activity as the endogenous target gene. On the other hand, the variant or mutated target gene may also have a biological activity, which differs from the biological activity of the endogenous target gene, e.g. a partially deleted activity, a completely deleted activity, an enhanced activity etc.

[0042] The complementation may be accomplished by coexpressing the polypeptide encoded by the exogenous nucleic acid, e.g. a fusion protein comprising the target protein and the affinity tag and the double stranded RNA molecule for knocking out the endogenous gene in the target cell. This coexpression may be accomplished by using a suitable expression vector expressing both the polypeptide encoded by the exogenous nucleic acid, e.g. the tag-modified target protein and the single-stranded RNA molecule or alternatively by using a combination of expression vectors. Proteins and protein complexes which are synthesized de novo in the target cell will contain the exogenous gene product, e.g. the modified fusion protein. In order to avoid suppression of the exogenous gene product expression by the RNAi molecule, the nucleotide sequence encoding the exogenous nucleic acid may be altered on the DNA level (with or without causing mutations on the amino acid level) in the part of the sequence which is homologous to the single-stranded RNA molecule. Alternatively, the endogenous target gene may be complemented by corresponding nucleotide sequences from other species, e.g. from mouse.

[0043] Preferred applications for the cell or organism of the invention is the analysis of gene expression profiles and/or proteomes. In an especially preferred embodiment an analysis of a variant or mutant form of one or several target proteins is carried out, wherein said variant or mutant forms are reintroduced into the cell or organism by an exogenous target nucleic acid as described above. The combination of knockout of an endogenous gene and rescue, by using mutated, e.g. partially deleted exogenous target has advantages compared to the use of a knockout cell. Further, this method is particularly suitable for identifying functional domains of the target protein. In a further preferred embodiment a comparison, e.g. of gene expression profiles and/or proteomes and/or phenotypic characteristics of at least two cells or organisms is carried out. These organisms are selected from:

[0044] (i) a control cell or control organism without target gene inhibition,

[0045] (ii) a cell or organism with target gene inhibition and

[0046] (iii) a cell or organism with target gene inhibition plus target gene complementation by an exogenous target nucleic acid.

[0047] The method and cell of the invention may also be used in a procedure for identifying and/or characterizing pharmacological agents, e.g. identifying new pharmacological agents from a collection of test substances and/or characterizing mechanisms of action and/or side effects of known pharmacological agents.

[0048] Thus, the present invention also relates to a system for identifying and/or characterizing pharmacological agents acting on at least one target protein comprising:

[0049] (a) a eukaryotic cell or a eukaryotic non-human organism capable of expressing at least one endogenous target gene coding for said target protein,

[0050] (b) at least one single-stranded RNA molecule capable of inhibiting the expression of said at least one endogenous target gene by RNAi and

[0051] (c) a test substance or a collection of test substances wherein pharmacological properties of said test substance or said collection are to be identified and/or characterized.

[0052] Further, the system as described above preferably comprises:

[0053] (d) at least one exogenous target nucleic acid coding for the target protein or a variant or mutated form of the target protein wherein said exogenous target nucleic acid differs from the endogenous target gene on the nucleic acid level such that the expression of the exogenous target nucleic acid is substantially less inhibited by the single-stranded RNA molecule than the expression of the endogenous target gene.

[0054] Furthermore, the RNA knockout complementation method may be used for preparative purposes, e.g. for the affinity purification of proteins or protein complexes from eukaryotic cells, particularly mammalian cells and more particularly human cells. In this embodiment of the invention, the exogenous target nucleic acid preferably codes for a target protein which is fused to an affinity tag.

[0055] The preparative method may be employed for the purification of high molecular weight protein complexes which preferably have a mass of ≥ 150 kD and more preferably of >500 kD and which optionally may contain nucleic acids such as RNA. Specific examples are the heterotrimeric protein complex consisting of the 20 kD, 60 kD and 90 kD

proteins of the U4/U6 snRNP particle, the splicing factor SF3b from the 17S U2 snRNP consisting of 5 proteins having molecular weights of 14, 49, 120, 145 and 155 kD and the 25S U4/U6/U5 tri-snRNP particle containing the U4, U5 and U6 snRNA molecules and about 30 proteins, which has a molecular weight of about 1.7 MD.

[0056] This method is suitable for functional proteome analysis in mammalian cells, particularly human cells.

[0057] Finally, the invention relates to a purified and isolated mammalian, particularly human RNA-induced silencing complex (RISC) having an apparent molecular weight of less than about 150-160 kDa, e.g. about 120 to 150-160 kDa. The RISC comprises polypeptide and optionally nucleic acid components, particularly single-stranded RNA molecules as described above. The RISC may be used as a target for diagnosis and/or therapy, as a diagnostic and/or therapeutic agent itself, as a molecular-biological reagent or as component in a screening procedure for the identification and/or characterization of pharmaceutical agents.

[0058] Polypeptide components of RISC preferably comprise members of the Argonaute family of proteins, and contain eIF2C1 and/or eIF2C2, and possibly at least one other expressed eIF2C family member, particularly selected from eIF2C3, eIF2C4, HILI and HIWI.

[0059] Expression or overexpression of one or several proteins present in RISC in suitable host cells, e.g. eukaryotic cells, particularly mammalian cells, is useful to assist an RNAi response. These proteins may also be expressed or overexpressed in transgenic animals, e.g. vertebrates, particularly mammals, to produce animals particularly sensitive to injected single-stranded or double-stranded siRNAs. Further, the genes encoding the proteins may be administered for therapeutic purposes, e.g. by viral or non-viral gene delivery vectors.

[0060] It is also conceivable to administer a siRNA/eIF2C1 or 2 complex directly by the assistance of protein transfection reagents (e.g. Amphoteric Protein Transfection Reagents, ProVectin protein (Imgenex), or similar products) rather than RNA/DNA transfection. This may have technical advantages over siRNA transfection that are limited to nucleic acid transfection.

[0061] Alternatively to the application of siRNAs as synthetic double-stranded or single-stranded siRNAs, it is conceivable to also administer an antisense siRNA precursor molecule in the form of a hairpin stem-loop structure comprising 19 to 29 base pairs in the stem with or without 5' or 3' overhanging ends on one side of the duplex and a nucleotide or non-nucleotide loop on the other end. Preferably, the hairpin structure has a 3' overhang of from 1-5 nucleotides. Further, the precursor may contain modified nucleotides as described above, particularly in the loop and/or in the 3' portion, particularly in the overhang. The siRNA or precursors of siRNAs may also be introduced by viral vectors or RNA expression systems into a RISC compound, e.g. eIF2C1 and/or 2 overexpressing organism or cell line. The siRNA precursors may also be generated by direct expression within an organism or cell line. This may be achieved by transformation with a suitable expression vector carrying a nucleic acid template operatively linked to an expression control sequence to express the siRNA precursor.

[0062] Further, the present invention is explained in more detail in the following figures and examples.

FIGURE LEGENDS

[0063] FIG. 1. HeLa cytoplasmic S100 extracts show siRNA-dependent target RNA cleavage.

[0064] (A) Representation of the 177-nt ³²P-cap-labeled target RNA with the targeting siRNA duplex. Target RNA cleavage site and the length of the expected cleavage products is also shown. The fat black line positioned under the antisense siRNA is used in the following figures as symbol to indicate the region of the target RNA, which is complementary to the antisense siRNA sequence. (B) Comparison of the siRNA mediated target RNA cleavage using the previously established *D. melanogaster* embryo in vitro system and HeLa cell S100 cytoplasmic extract. 10 nM cap-labeled target RNA was incubated with 100 nM siRNA as described in materials. Reaction products were resolved on a 6% sequencing gel. Position markers were generated by partial RNase T1 digestion (T1) and partial alkaline hydrolysis (OH) of the cap-labeled target RNA. The arrow indicates the 5' cleavage product, the fragment is unlabeled and therefore invisible.

[0065] FIG. 2. Chemical modification of the 5' end of the antisense but not the sense siRNAs prevents sense target RNA cleavage in HeLa S100 extracts. (A) Illustration of the possible 5' and 3' aminolinker modifications of the sense and antisense strands of a siRNA duplex. L5 represents a 6-carbon chain aminolinker connected via a 5'-phosphodiester linkage, L3 represents a 7-carbon aminolinker connected via a phosphodiester bond to the terminal 3' phosphate. s, sense; as, antisense. (B) Target RNA cleavage testing various combinations of 5' and 3' aminolinker-modified siRNA duplexes. NC (negative control) shows an incubation reaction of the target RNA in the absence of siRNA duplex. T1, RNase T1 ladder; OH, partial alkaline hydrolysis ladder.

[0066] FIG. 3. siRNA containing 3'-terminal phosphates are subjected to ligation as well as dephosphorylation reactions.

[0067] (A) Sequence of the radiolabeled siRNA duplex. The labeled nucleotide was joined to synthetic 20-nt antisense siRNA by T4 RNA ligation of ³²pCp. The various combinations of 5' and 3' hydroxyl/phosphate were prepared as described in materials. X and Y indicate 5' and 3' modifications of the antisense siRNA. (B) Fate of the antisense siRNA during incubation of the modified siRNA duplexes in HeLa S100 extract in the presence of non-radiolabeled target RNA. The different phosphorylated forms of the antisense siRNA were distinguished based on their gel mobility. Identical results were obtained when using 5' phosphorylated sense siRNA or when leaving out the target RNA during incubation. Ligation products are only observed when 3' phosphates were present on the labeled antisense siRNA.

[0068] FIG. 4: RISC is a stable complex that does not rapidly exchange bound siRNA.

[0069] Increasing concentrations of non-specific siRNA compete with target-specific RISC formation when added simultaneously to HeLa S100 extracts (lanes 4 to 7). However, when the unspecific siRNA duplex is added 15 min after pre-incubation with the specific siRNA duplex, no more competition was observed (3 lanes to the right). T1, RNase T1 ladder.

[0070] FIG. 5. Partial purification of human RISC.

[0071] (A) Graphical representation of the structure of the biotinylated siRNA duplex used for affinity purification of siRNA-associated factors. L3 indicates a C7-aminolinker that was conjugated to a photo-cleavable biotin N-hydroxysuccinimide ester; UV indicates photocleavage of the UV-sensitive linkage to release affinity selected complexes under native conditions. (B) Superdex-200 gel filtration analysis of siRNA-protein complexes (siRNPs) recovered

by UV treatment/elution (UV elu) from the streptavidin affinity column. Fractions were assayed for their ability to sequence-specifically cleave the cap-labeled target RNA. The number of the 10 collected fractions and the relative positions of the aldolase (158 kDa) and BSA (66 kDa) size markers are indicated. (C) Glycerol gradient (5%-20%) sedimentation of siRNPs recovered by UV treatment/elution from the streptavidin affinity column. For legend, see (B). When monitoring the precise size of target RNA cleavage fragments using internally ^{32}P -UTP-labeled, capped mRNA, the sum is equal to the full-length transcript, thus indicating that target RNA is indeed only cleaved once in the middle of the region spanned by the siRNA.

[0072] FIG. 6. RISC contains a single-stranded siRNA.

[0073] siRNPs were subjected to affinity selection after incubation using siRNA duplexes with one or both strands biotinylated. The eluate recovered after UV treatment or the unbound fraction after streptavidin affinity selection (flow-through) was assayed for target RNA degradation. If the antisense strand was biotinylated, all sense target RNA-cleaving RISC was bound to the streptavidin beads, while sense siRNA biotinylation resulted in RISC activity of the flow-through. The cleavage reaction in the flow-through fraction was less efficient than in the UV eluate, because affinity-selected RISC was more concentrated.

[0074] FIG. 7. Single-stranded antisense siRNAs reconstitute RISC in HeLa S100 extracts.

[0075] Analysis of RISC reconstitution using single-stranded or duplex siRNAs comparing HeLa S100 extracts (A) and the previously described *D. melanogaster* embryo lysate (B). Different concentrations of single-stranded siRNAs (s, sense; as, antisense) and duplex siRNA (ds) were tested for specific targeting of cap-labeled substrate RNA. 100 nM concentrations of the antisense siRNA reconstituted RISC in HeLa S100 extract, although at reduced levels in comparison to the duplex siRNA. Reconstitution with single-stranded siRNAs was almost undetectable in *D. melanogaster* lysate, presumably because of the higher nuclease activity in this lysate causing rapid degradation of uncapped single-stranded RNAs.

[0076] FIG. 8. Single-stranded antisense siRNAs mediate gene silencing in HeLa cells.

[0077] (A) Silencing of nuclear envelope protein lamin A/C. Fluorescence staining of cells transfected with lamin A/C-specific siRNAs and GL2 luciferase (control) siRNAs. Top row, staining with lamin A/C specific antibody; middle row, Hoechst staining of nuclear chromatin; bottom row, phase contrast images of fixed cells. (B) Quantification of lamin A/C knockdown after Western blot analysis. The blot was stripped after lamin A/C probing and reprobed with vimentin antibody. Quantification was performed using a Lumi-Imager (Roche) and LumiAnalyst software to quantitate the ECL signals (Amersham Biosciences), differences in gel loading were corrected relative to non-targeted vimentin protein levels. The levels of lamin A/C protein were normalized to the non-specific GL2 siRNA duplex.

[0078] FIG. 9. Antisense siRNAs of different length direct target RNA cleavage in HeLa S100 extracts.

[0079] (A) Graphical representation of the experiment. Antisense siRNAs were extended towards the 5' side (series 1, 20 to 25-nt) or the 3' side (series 2, 20 to 23-nt).

[0080] (B) Target RNA cleavage using the antisense siRNAs described in (A). HeLa S100 extract was incubated

with 10 nM cap-labeled target RNA and 100 nM antisense siRNAs at 30° C. for 2.5 h. Reaction products were resolved on a 6% sequencing gel. Position markers were generated by partial RNase T1 digestion (T1) and partial alkaline hydrolysis (OH) of the cap-labeled target RNA. Arrows indicate the position of the 5' cleavage products generated by the different antisense siRNAs. The fat black lines on the left (series 1) and the right (series 2) indicate the region of the target RNA, which is complementary to the antisense siRNA sequences.

[0081] FIG. 10. Length dependence of antisense siRNAs and effect of terminal modifications for targeting RNA cleavage in HeLa S100 extracts.

[0082] HeLa S100 extract was incubated with 10 nM cap-labeled target RNA and 100 nM antisense siRNAs at 30° C. for 2.5h. Reaction products were resolved on a 6% sequencing gel. Position markers were generated by partial RNase T1 digestion (T1) of the cap-labeled target RNA. The fat black line on the left indicates the region of the target RNA, which is complementary to the 21-nt antisense siRNA sequence. The siRNA sequences used in each experiment are listed below (sense and antisense siRNAs are listed together, they were pre-annealed to form duplex siRNAs). p, phosphate; t, 2'-deoxythymidine, c, 2'-deoxycytidine, g, 2'-deoxyguanosine; L, aminolinker, B, photocleavable biotin; A,C,G,U, ribonucleotides.

Lane	Sense siRNA (5'-3')	Antisense siRNA (5'-3')
1		pUCGAAGUAUCCG CG
2		pUCGAAGUAUCCG CGUACGUG
3		pUCGAAGUAUCCG CGUACGUGAUGU
4		pUCGAAGUAUCCG CGUACGUGAUGUUC
5		pUCGAAGUAUCCG CGUACGUGAUGUUC AC
6		pUCGAAGUAUCCG CG
7		pUCGAAGUAUCCG CGUACGUG
8		pUCGAAGUAUCCG CGUACGUGAUGU
9		pUCGAAGUAUCCG CGUACGUGAUGUUC
10		pUCGAAGUAUCCG CGUACGUGAUGUUC AC
11		pUCGAAGUAUCCG CGUACGUG
12		pUCGAAGUAUCCG CGUACGtg
13		pUCGAAGUAUCCG CGUACGUU

-continued

Lane	Sense siRNA (5'-3')	Antisense siRNA (5'-3')
14		pUCGAAGUAUCCG CGUACGtt
15		pUCGAAGUAUCCG CGUACGUG
16		pUCGAAGUAUCCG CGUACGtg
17		pUCGAAGUAUCCG CGUACGUU
18		pUCGAAGUAUCCG CGUACGtt
19	CGUACGCGGAUACUUCG AAA	pUCGAAGUAUCCG CGUACGUG
20	CGUACGCGGAUACUUCG AAA	pUCGAAGUAUCCG CGUACGtg
21	CGUACGCGGAUACUUCG AAA	pUCGAAGUAUCCG CGUACGUU
22	CGUACGCGGAUACUUCG AAA	pUCGAAGUAUCCG CGUACGtt
23		tCGAAGUAUCCGC GUACGUULB
24	cGUACGCGGAUACUUCG AUULB	tCGAAGUAUCCGC GUACGUULB
25		ptCGAAGUAUCCGC GUACGttLB
26	cGUACGCGGAUACUUCG AttLB	ptCGAAGUAUCCGC GUACGttLB
27		ptCGAAGUAUCCGC GUACGttL

[0083] FIG. 11: Single-stranded antisense siRNAs mediate gene silencing in HeLa cells.

[0084] Quantification of lamin A/C knockdown after Western blot analysis. The blot was stripped after lamin A/C probing and reprobed with vimentin antibody. Quantification was performed using a Lumi-Imager (Roche) and LumiAnalyst software to quantitate the ECL signals (Amersham Biosciences), differences in gel loading were corrected relative to non-targeted vimentin protein levels. The levels of lamin A/C protein were normalized to the non-specific GL2 siRNA duplex.

[0085] FIG. 12. Protein composition of affinity purified RISC.

[0086] (A) Silver-stained SDS-PAGE gel of affinity-selected ribonucleoprotein complexes after glycerol gradient (5%-20%) sedimentation. The arrow indicates the band containing eIF2C1 and eIF2C2. Molecular size markers are indicated on the left. The asterisk indicates a fraction for which the protein pellet was lost after precipitation. (B) Target RNA cleavage assay of the collected fractions. RISC activity peaked in fraction 7 and 8; bu, buffer.

[0087] FIG. 13. Mass spectrometric characterization of eIF2C1 and eIF2C2.

[0088] The 100 kDa band was analysed by mass spectrometry. Mass spectrum indicating the peptide peaks corresponding to eIF2C2 (A) and eIF2C1 (B).

[0089] (C) Alignment of eIF2C2 and eIF2C1 amino-acid sequences indicating the position of the identified peptides. Sequence differences are indicated by yellow boxes.

[0090] FIG. 14. Predicted amino-acid sequences of the six human Argonaute protein family members.

[0091] FIG. 15. Alignment of the sequences of the six human Argonaute protein family members.

[0092] Predicted sequences of human eIF2C1-4, HILI and HIWI have been aligned using ClustalX program.

[0093] FIG. 16. Predicted cDNA sequences of the six human Argonaute protein family members.

[0094] FIG. 17. AU members of the Argonaute family but HIWI are expressed in HeLa cells.

[0095] RT-PCR analysis on polyA RNA from HeLa cells. (A) Primers (forward and reverse) used for nested and semi-nested PCR amplification of the different Argonautes and expected length of the PCR products. (B) Agarose gel electrophoresis of the obtained PCR products, confirming the expected length. Left lanes, 100 by DNA ladder.

EXAMPLE

1. Material and Methods

[0096] 1.1 siRNA Synthesis and Biotin Conjugation

[0097] siRNAs were chemically synthesized using RNA phosphoramidites (Prologo, Hamburg, Germany) and deprotected and gel-purified as described previously. 5' aminolinkers were introduced by coupling MMT-C6-aminolinker phosphoramidite (Prologo, Hamburg), 3' C7-aminolinkers were introduced by assembling the oligoribonucleotide chain on 3'-aminomodifier (TFA) C7 Icaa control pore glass support (Chemgenes, Mass., USA). The sequences for GL2 luciferase siRNAs were as described (Elbashir et al., 2001a, supra). If 5'-phosphates were to be introduced, 50 to 100 nmoles of synthetic siRNAs were treated with T4 polynucleotide kinase (300 p1 reaction, 2.5 mM ATP, 70 mM Tris-HCl, pH 7.6, 10 mM MgCl₂, 5 mM DTT, 30 U T4 PNK, New England Biolabs, 45 min, 37° C.) followed by ethanol precipitation.

[0098] 3' Terminal ³²pCp labeling (FIG. 3) was performed in a 30 µl reaction (17 µM siRNA, 0.5 µM ³²pCp (110 TBq/mmol), 15% DMSO, 20 U T4 RNA ligase, NEB, and 1x NEB-supplied reaction buffer) for 1.5 h at 37° C., and gel-purified. One half of the pCp-labeled RNA was dephosphorylated (25 µl reaction, 500 U alkaline phosphatase, Roche, and Roche-supplied buffer, 30 min, 50° C.), followed by phenol/chloroform extraction and ethanol precipitation. Half of this reaction was 5' phosphorylated (20 µl reaction, 2 units T4 polynucleotide kinase, NEB, 10 mM ATP, NEB-supplied buffer, 60 min, 37° C.). A quarter of the initial pCp-labeled siRNA was also 5' phosphorylated (10 µl reaction, 10 units 3' phosphatase-free T4 polynucleotide kinase, Roche, 10 mM ATP, Roche-supplied buffer, 3 min, 37° C.).

[0099] For conjugation to biotin, 20 to 65 nmoles of fully deprotected aminolinker-modified siRNA were dissolved in 100 µl of 100 mM sodium borate buffer (pH 8.5) and mixed with a solution of 1 mg of EZ-Link NHS-PC-LC-Biotin (Pierce, Ill., USA) in 100 µl of anhydrous dimethylformamide. The solution was incubated for 17 h at 25° C. in the dark. Subsequently, siRNAs were precipitated by the addition of 60 µl 2 M sodium acetate (pH 6.0) and 1 ml ethanol. The RNA pellet was collected by centrifugation and biotin-conjugated siRNA was separated from non-reacted siRNA on a preparative denaturing 18% acrylamide gel (40 cm length) in

the dark. The RNA bands were visualized by 254 nm UV shadowing and minimized exposure time. The bands were excised, and the RNA was eluted overnight in 0.3 M NaCl at 4° C. and recovered by ethanol precipitation. siRNA duplexes were formed as previously described (Elbashir et al., Methods 26 (2002), 199-213).

1.2 Preparation of S100 Extracts from HeLa Cells

[0100] Cytoplasm from HeLa cells adapted to grow at high density was prepared following the Dignam protocol for isolation of HeLa cell nuclei (Dignam et al., Nucleic Acids Res. 11 (1983), 1475-1489). The cytoplasmic fraction was supplemented with KCl, MgCl₂ and glycerol to final concentrations of 100 mM, 2 mM and 10%, respectively. At this stage, the extracts can be stored frozen at -70° C. after quick-freezing in liquid nitrogen without loss of activity. S100 extracts were prepared by ultracentrifugation at 31,500 rpm for 60 minutes at 4° C. using a Sorvall T-865 rotor. The protein concentration of HeLa S100 extract varied between 4 to 5 mg/ml as determined by Bradford assay.

1.3 Affinity Purification of RISC with 3' Biotinylated siRNA Duplexes

[0101] For affinity purification of siRNA-associated protein complexes from HeLa S100 extracts, 10 nM of a 3' double-biotinylated siRNA duplex were incubated in 0.2 mM ATP, 0.04 mM GTP, 10 U/ml RNasin, 6 µg/ml creatine kinase, and 5 mM creatine phosphate in 60% S100 extract at 30° C. for 30 to 60 min and gentle rotation. Thereafter, 1 ml slurry of Immobilized Neutravidin Biotin Binding Protein (Pierce, IL, USA) was added per 50 ml of reaction solution and the incubation was continued for another 60 to 120 min at 30° C. with gentle rotation. The Neutravidin beads were then collected at 2000 rpm for 2 minutes at 4° C. in a Heraeus Megafuge 1.0 R centrifuge using a swinging bucket rotor type 2704. Effective capturing of RISC components after affinity selection was confirmed by assaying the supernatant for residual RISC activity with and without supplementing fresh siRNA duplexes. The collected Neutravidin beads were washed with 10 volumes of buffer A relative to the bead volume (30 mM HEPES, pH 7.4, 100 mM KCl, 2 mM MgCl₂, 0.5 mM DTT, 10% glycerol) followed by washing with 5 volumes of buffer B (same as buffer A with only 3% glycerol content). The beads were transferred to a 0.8×4 cm Poly-Prep chromatography column (BioRad; CA, USA) by resuspending in 3 volumes of buffer B at 4° C., followed by 10 volumes of washing with buffer B. Washing of the beads was continued by 10 volumes of buffer B increased to 300 mM KCl. The column was then reequilibrated with regular buffer B. To recover native siRNA-associated complexes, the column was irradiated in the cold room by placing it at a 2 cm distance surrounded by four 312 nm UV lamps (UV-B tube, 8 W, Herotab, Germany) for 30 minutes. To recover the photo-cleaved siRNP solution, the column was placed into a 50 ml Falcon tube and centrifuged at 2000 rpm for 1 minute at 4° C. using again the 2704 rotor. For full recovery of siRNPs, the beads were once again resuspended in buffer B followed by a second round of UV treatment for 15 minutes. Both eluates were pooled and assayed for target RNA degradation.

1.4 Target RNA Cleavage Assays

[0102] Cap-labeled target RNA of 177 nt was generated as described (Elbashir et al., EMBO J. 20 (2001 c), 6877-6888) except that his-tagged guanylyl transferase was expressed in *E. coli* from a plasmid generously provided by J. Wilusz and purified to homogeneity. If not otherwise indicated, 5' phos-

phorylated siRNA or siRNA duplex was pre-incubated in supplemented HeLa S100 extract at 30° C. for 15 min prior to addition of cap-labeled target RNA. After addition of all components, final concentrations were 100 nM siRNA, 10 nM target RNA, 1 mM ATP, 0.2 mM GTP, 10 U/ml RNasin, 30 µg/ml creatine kinase, 25 mM creatine phosphate, 50% S100 extract. Incubation was continued for 2.5 h. siRNA-mediated target RNA cleavage in *D. melanogaster* embryo lysate was performed as described (Zamore et al., Cell 101 (2000), 25-33). Affinity-purified RISC in buffer B was assayed for target RNA cleavage without preincubation nor addition of extra siRNA (10 nM target RNA, 1 mM ATP, 0.2 mM GTP, 10 U/ml RNasin, 30 µg/ml creatine kinase, 25 mM creatine phosphate, 50% RISC in buffer B). Cleavage reactions were stopped by the addition of 8 vols of proteinase K buffer (200 mM Tris-HCl pH 7.5, 25 mM EDTA, 300 mM NaCl, 2% w/v SDS). Proteinase K, dissolved in 50 mM Tris-HCl pH 8.0, 5 mM CaCl₂, 50% glycerol, was added to a final concentration of 0.6 mg/ml and processed as described (Zamore et al. (2000), supra). Samples were separated on 6% sequencing gels.

1.5 Analytical Gel Filtration

[0103] UV-eluates in buffer B were fractionated by gel filtration using a Superdex 200 PC 3.2/30 column (Amersham Biosciences) equilibrated with buffer A on a SMART system (Amersham Biosciences). Fractionation was performed by using a flow rate of 40 µl/minute and collecting 100 µl fractions. Fractions were assayed for specific target RNA cleavage. Size calibration was performed using molecular size markers thyroglobulin (669 kDa), ferritin (440 kDa), catalase (232 kDa), aldolase (158 kDa) and BSA (66 kDa) (Amersham Biosciences).

1.6 Glycerol Gradient Sedimentation

[0104] UV-eluates were layered on top of 4 ml linear 5% to 20% (w/w) glycerol gradient adjusted to 30 mM HEPES, pH 7.4, 100 mM KCl, 2 mM MgCl₂, 0.5 mM DTT. Centrifugation was performed at 35000 rpm for 14.5 h at 4° C. using a Sorvall SW 60 rotor. Twenty fractions of 0.2 ml volume were removed sequentially from the top and 15 µl aliquots were used to assay for target RNA cleavage.

2. Results

[0105] 2.1 A Human Biochemical System for siRNA Functional Analysis

[0106] We were interested in assaying siRNA-mediated target RNA degradation in human cell extracts, because siRNAs are powerful reagents to knockdown gene expression in human cells but the action of siRNAs in human cells was uncertain. To investigate whether siRNAs guide target RNA degradation in human cells with a similar mechanism to the one observed in *D. melanogaster* (e.g. Elbashir et al. (2001 b), supra), we prepared substrates for targeted mRNA degradation as described previously (Elbashir et al. (2001 c), supra). A 5'—³²P-cap-labeled, 177-nt RNA transcript, derived from a segment of the firefly luciferase gene, was incubated in HeLa cell S100 or *D. melanogaster* embryo extracts with a 21-nt siRNA duplex in the presence of an ATP regeneration system (FIG. 1A, B). siRNA cleavage assays were performed at 25° C. in *D. melanogaster* lysate and at 30° C. in HeLa

S100 extracts for 2.5 h. After deproteinization using proteinase K, the reaction products were separated on a 6% sequencing gel.

[0107] Similar to the previous observation in *D. melanogaster* lysate, we observed the appearance of a cleavage product in HeLa S100 extract at exactly the same position, thus indicating that the siRNA duplex guides target RNA cleavage in the human system with the same specificity and mechanism. The cleavage reaction appeared less efficient when compared to the *D. melanogaster* system, but this could be explained by the 5-fold lower total protein concentration of HeLa S100 extracts (25 mg/ml vs. 5 mg/ml). Similar to *D. melanogaster* lysates, siRNA duplexes without 5' phosphate were rapidly 5' phosphorylated in HeLa S100 extracts (see below) and the ability to cleave the target RNA was independent of the presence of a 5' phosphate on the synthetic siRNA duplexes.

[0108] Comparative analysis of the efficiency of siRNA duplexes of different length in *D. melanogaster* lysate and in transfected mammalian cells indicated that the differences in silencing efficiencies between 20- to 25-nt siRNA duplexes were less pronounced in mammalian cells than in *D. melanogaster* (Elbashir et al. (2002), supra). Duplexes of 24- and 25-nt siRNAs were inactive in *D. melanogaster* lysate, whereas the same duplexes were quite effective for silencing when introduced by transfection into HeLa cells. We therefore asked whether siRNA duplexes of 20- to 25-nt are able to reconstitute RISC also with approximately equal efficiency. Indeed, we observed no large differences in our biochemical assay, and the position of target RNA cleavage was as predicted according to the cleavage guiding rules established in *D. melanogaster* lysate (data not shown). Our biochemical results therefore support the in vivo observations.

2.2 5' Modification of the Guide siRNA Inhibits RISC Activity

[0109] Modification of siRNAs at their termini is important for developing siRNA-based affinity purification schemes or for conjugating reporter tags for biophysical measurements. The most common method for introducing reactive side chains into nucleic acids is by chemical synthesis using aminolinker derivatives (Eckstein (1991), Oligonucleotides and analogues, 2nd Ed., Oxford UK, Oxford University Press). After complete deprotection of the oligonucleotide, the primary amine is typically reacted with the N-hydroxysuccinimidyl ester of the desired compound. We have introduced 5' and 3' aminolinkers with six and seven methylene groups as spacers, respectively. The linker-modified siRNA duplexes were tested for mediating target RNA degradation in HeLa S100 extract (FIG. 2A, B). Modification of the 5'-end of the antisense guide siRNA abolished target RNA cleavage, while modification of neither the sense 5'-end nor of both 3'-ends showed any inhibitory effect. In an identical experiment using *D. melanogaster* embryo lysate, we observed a similar pattern of RISC activity although the duplex carrying the 5' aminolinker-modified antisense siRNA showed some residual activity (data not shown). Presumably, introduction of additional atoms or the change in terminal phosphate electric charge at the 5'-end of the antisense siRNA interfered with its ability to function as guide RNA. The critical function of the guide siRNAs 5' end was previously documented (Elbashir et al. (2001 c), supra).

[0110] The ability to modify siRNAs at their 3'-end suggests that siRNAs do not play a major role for priming dsRNA synthesis and do not act as primers for degenerative PCR. The

fate of a siRNA in HeLa S100 extracts was followed directly by incubation of an internally ^{32}P -labeled siRNA duplexes. The radiolabeled antisense siRNA strand was also prepared with different 5' and 3' phosphate modifications (FIG. 3A). All described combinations of siRNA duplexes were fully competent for RISC-dependent target RNA degradation (data not shown). As previously observed for *D. melanogaster* lysates (Nykänen et al. (2001), supra), rapid 5' phosphorylation of siRNA duplexes with free 5' hydroxyl termini was apparent. To our surprise, we noted that a small fraction of the 3' phosphorylated antisense siRNA could be ligated to the opposing 5' hydroxyl of the sense siRNA producing a lower mobility band. The inter-strand ligation was confirmed by changing the length of the unlabeled sense siRNA, which resulted in the expected mobility changes of the ligation product (data not shown). RNA ligase activity was previously observed in HeLa S100 extracts and it is mediated by two enzymatic activities (e.g. Vicente and Filipowicz, Eur. J. Biochem., 176 (1988), 431-439). The 3' terminal phosphate is first converted to a 2',3'-cyclic phosphate requiring ATP and 3' terminal phosphate cyclase. Thereafter, the opposing 5' hydroxyl is ligated to the cyclic phosphate end by an as yet uncharacterized RNA ligase. We chemically synthesized the predicted 5' phosphorylated, 42-nt ligation product and found that it is unable to mediate target RNA cleavage, presumably because it can not form activated RISC. The majority of the 3' phosphorylated duplexes siRNA was gradually dephosphorylated at its 3' end and emerged chemically similar to naturally generated siRNA. Together, these observations indicate that the cell has a mechanism to preserve the integrity of siRNAs. We were unable to detect a proposed siRNA-primed polymerization product (FIG. 3B), suggesting that siRNAs do not function as primers for template-dependent dsRNA synthesis in our system. However, we acknowledge that a proposed RNA-dependent polymerase activity may have been inactivated during preparation of our extracts.

2.3 siRNAs Incorporated into RISC do not Compete with a Pool of Free siRNAs

[0111] In order to analyze RISC assembly and stability, we tested whether target-unspecific siRNA duplexes were able to compete with target-specific siRNA duplexes. When specific and non-specific siRNA duplexes were co-incubated in HeLa S100 extracts, increasing concentrations of unspecific siRNA duplex competed with the formation of target-specific RISC (FIG. 4, left lanes). However, when target-specific siRNAs were pre-incubated in HeLa S100 extract for 15 min in the absence of competitor siRNA duplex, the assembled siRNA in the target-specific RISC could no longer be competed with the target-unspecific siRNA duplex (FIG. 4, right lanes). This result suggests that RISC is formed during the first 15 minutes of incubation and that siRNAs were irreversibly associated with the protein components of RISC during the 2.5 h time window of the experiment.

2.4 Purification of Human RISC

[0112] After having the 3' termini of siRNAs defined as the most suitable position for chemical modification, a photocleavable biotin derivative was conjugated to the 3' aminolinker-modified siRNAs. A photo-cleavable biotin derivative was selected because of the advantage of recovering RISC under non-denaturing conditions after capturing complexes on streptavidin-coated affinity supports. 3' Conjugation of biotin to the sense, antisense or to both of the strands did not

affect target RNA cleavage when compared to non-biotinylated siRNAs (data not shown). siRNA duplexes with biotin residues on both 3' ends were therefore used for affinity purification (FIG. 5A). The double biotinylated siRNA duplex was incubated in HeLa S100 extracts in the presence of ATP, GTP, creatine phosphate, and creatine kinase for ATP regeneration. Thereafter, streptavidin-conjugated agarose beads were added to capture the biotinylated siRNA ribonucleoprotein complexes (siRNPs) including RISC. After extensive washing of the collected beads, the siRNPs were released by UV irradiation at 312 nm. The eluate cleaved target RNA sequence-specifically, thus indicating that RISC was recovered in its native state from the resin. (FIG. 5B, C, lane UV elu). The flow-through from the affinity selection showed no detectable RISC activity indicating complete binding of RISC by the beads (FIG. 6). The affinity eluate was further analyzed by applying it onto a Superdex 200 gel filtration column (FIG. 5B) as well as a 5%-20% glycerol gradient ultra-centrifugation (FIG. 5C). Individual fractions were collected and assayed for target RNA cleavage without the addition of any further siRNA. RISC activity appeared between the molecular size markers aldolase (158 kDa) and BSA (66 kDa) after gel filtration or glycerol gradient centrifugation (FIG. 5B, C). The molecular size of human RISC is therefore estimated to be between 90 and 160 kDa, significantly smaller than the complex previously analyzed in *D. melanogaster* lysates (Hammond et al. (2000), supra; Nykänen et al. (2001), supra). The small size of RISC suggests that Dicer (210 kDa) is not contained in RISC and that the formation of RISC from synthetic siRNAs may occur independently of Dicer. While these results do not rule out a role for Dicer during assembly of RISC, they emphasize the absence of Dicer in RISC.

2.5 RISC Contains a Single siRNA Strand and can be Reconstituted Using Single-Stranded siRNAs

[0113] Two models are currently discussed concerning the siRNA strand composition of RISC. The first model suggests that both strands of the initially added siRNA duplex are physically present in RISC, but in an unwound conformation. The second model proposes that RISC carries only a single siRNA strand, implying loss of one of the siRNA strands during assembly. The latter model has been favored based on the analogy to miRNA precursor processing, where only one 21-nt strand accumulated from a dsRNA hairpin precursor. The molecular basis for the asymmetry of the miRNA precursor processing reaction is not yet understood. Because siRNAs have symmetric 2-nt 3'-overhangs it is assumed that siRNA duplexes enter RISC with equal probability for both orientations, thus giving rise to distinct sense and antisense targeting RISCs.

[0114] To address the constitution of siRNAs in RISC, we affinity selected the assembled complexes with siRNA duplexes that were biotinylated at only one of the two constituting strands or both (FIG. 6). If both strands were present together in RISC, the cleavage activity should be affinity selected on Neutravidin independently of the position of the biotin residue. In contrast, we observed target RNA cleavage from UV eluates after streptavidin selection only for siRNA duplexes with biotin conjugated to the antisense strand, but not the sense strand (FIG. 6). RISC activity, assembled on siRNA duplexes with only the sense siRNA biotinylated, remained in the flow-through. These data suggest that RISC contains only a single-stranded RNA molecule.

[0115] To assess whether single-stranded siRNAs may be able to reconstitute RISC, single-stranded 5' phosphorylated siRNAs as well as the siRNA duplex were incubated at concentrations between 1 to 100 nM with cap-labeled target RNA in HeLa S100 extract (FIG. 7A). At 100 nM single-stranded antisense siRNA, we detected RISC-specific target RNA cleavage, thus confirming that single-stranded siRNAs are present in RISC. At lower concentrations of single-stranded siRNAs, RISC formation remained undetectable while duplex siRNAs were effectively forming RISC even at 1 nM concentration. Therefore, a specific pathway exists which converts double-stranded siRNA into single-stranded siRNA containing RISC. Using *D. melanogaster* embryo lysate, we were unable to detect RISC activity from antisense siRNA (FIG. 7B), presumably because of the high load of single-strand specific ribonucleases (Elbashir et al. (2001 b), supra). Furthermore, 5' phosphorylated 20- to 25-nt antisense siRNAs were able to mediate RISC-specific target RNA degradation in HeLa S100 extract producing the same target RNA cleavage sites as duplex siRNAs of this length (data not shown).

[0116] Finally, we tested single-stranded and duplex siRNAs for targeting of an endogenous gene in HeLa cells following our standard protocol previously established for silencing of lamin A/C. 200 nM concentrations of single-stranded siRNAs with and without 5' phosphate and 100 nM concentrations of duplex siRNAs were transfected into HeLa cells. Lamin A/C levels were monitored 48 h later using immunofluorescence (FIG. 8A) and quantitative luminescence-based Western blot analysis (FIG. 8B). non-phosphorylated antisense siRNA caused a substantial knockdown of lamin A/C to about 25% of its normal level while 5' phosphorylated siRNAs reduced the lamin A/C content to less than 5%, similar to the reduction observed with the lamin A/C 5' phosphorylated (data not shown) or non-phosphorylated duplex siRNA (FIG. 8). Sense siRNA and GL2 unspecific siRNA did not affect lamin A/C levels. The levels of non-targeted vimentin protein were monitored and used for normalizing of the loading of the lanes of the lamin A/C Western blots.

[0117] Gene silencing was also observed with phosphorylated as well as non-phosphorylated antisense siRNAs ranging in size between 19 to 29 nt. The phosphorylated antisense siRNAs were consistently better performing than the non-phosphorylated antisense, and their silencing efficiencies were comparable to that of the conventional duplex siRNA (FIG. 11).

2.6 Protein Composition of RISC

[0118] In order to identify the protein components of the RNA-induced silencing complex (RISC) in HeLa S100 extract, the specific affinity selection previously outlined was used. UV eluates were fractionated on a 5-20% glycerol gradient, fractions were recovered from the gradient and analysed for protein composition and target RNA endonucleolytic activity. Two proteins of approximately 100 kDa were identified by mass spectrometry in the peak fraction of the endonucleolytic activity (FIG. 12, fractions 7 and 8), corresponding to eIF2C1 and eIF2C2/GERp95 (FIGS. 13A and B). These proteins are 82% similar and are both members of the Argonaute family (FIG. 13C). The first evidence that Argonaute proteins are part of RISC was provided by classical biochemical fractionation studies using dsRNA-transfected *D. melanogaster* S2 cells (Hammond et al., 2001, supra). The closest relative to *D. melanogaster* Argonaute2,

D. melanogaster Argonaute1, was recently shown to be required for RNAi (Williams and Rubin, PNAS USA 99 (2002), 6889-6894).

[0119] Mass spectrometry analysis also revealed the presence of three peptides belonging exclusively to the HILI member of the Argonaute family of proteins. The sequences of those peptides are: NKQDFMDLSICTR, is corresponding to positions 17-29 of the protein; TEYVAESFLNCLRR, corresponding to positions 436-449 of the protein, and; YNHDL-PARIIVYR, corresponding to positions 591-603 of the protein. This finding suggests that the protein HILI may also be part of RISC.

[0120] In human, the Argonaute family is composed of 6 members, eIF2C1, eIF2C2, eIF2C3, eIF2C4, HILI and HMI (FIG. 14). The alignment of the six predicted amino-acid sequences show a high conservation, in particular between the eIF2C members, and HILI and HIWI (FIG. 15). Predicted cDNA sequences encoding the Argonaute proteins are also shown (FIG. 16).

[0121] The expression of the human Argonaute proteins was also investigated in HeLa cells by RT-PCR analysis using total and poly (A) selected RNA. All members of the family but HIWI were detected (FIG. 17).

3. Discussion

[0122] The development of a human biochemical system for analysis of the mechanism of RNAi is important given the recent success of siRNA duplexes for silencing genes expressed in human cultured cells and the potential for becoming a sequence-specific therapeutic agent. Biochemical systems are useful for defining the individual steps of the RNAi process and for evaluating the constitution and molecular requirements of the participating macromolecular complexes. For the analysis of RNAi, several systems were developed, with the *D. melanogaster* systems being the most comprehensive as they enable to reconstitute dsRNA processing as well as the mRNA targeting. For mammalian systems, reconstitution of the mRNA targeting reaction has not yet been accomplished. Here, we describe the development and application of a biochemical system prepared from the cytoplasmic fraction of human HeLa cells, which is able to reconstitute the human mRNA-targeting RNA-induced silencing complex (RISC). Formation of RISC was accomplished using either 5' phosphorylated or non-phosphorylated siRNA duplexes; as well as single-stranded antisense siRNAs; non-phosphorylated siRNA duplexes and presumably also single-stranded antisense siRNAs are rapidly 5' phosphorylated in HeLa cell extracts (FIG. 3).

Biochemical Characterization of siRNA Function

[0123] Reconstitution of RISC activity was only observed using cytoplasmic HeLa extracts. HeLa nuclear extracts assayed under the same conditions did not support siRNA-specific target RNA cleavage, thus suggesting that RISC components are located predominantly in the cytoplasm (data not shown).

[0124] Modifications of the 5' and 3' termini of siRNAs were tested in order to assess the importance of the siRNA termini for the targeting step. It was found that the 5' end modification of the guide siRNA was more inhibitory for target RNA cleavage than 3' end modification. Introduction of the 3' biotin affinity tag into the target-complementary guide siRNA enabled us to affinity select sense-RNA-targeting RISC, whereas 3' biotinylation of the sense siRNA strand resulted in RISC activity in the flowthrough. Furthermore, the

single RNA strand composition of RISC was confirmed by reconstituting the sequence-specific endonuclease complex using 5'-phosphorylated single-stranded guide siRNA. The reconstitution of RISC from single-stranded siRNA was however less effective and required 10- to 100-fold higher concentrations compared to duplex siRNA. Reconstitution of RISC from single-stranded siRNA was undetectable using *D. melanogaster* embryo lysate, which is most likely explained by the high content of 5' to 3' exonucleases in embryo lysate.

[0125] The size of RISC in HeLa lysate was determined by gel filtration as well as glycerol gradient ultracentrifugation after streptavidin affinity purification with 3' biotinylated siRNA duplexes. Sizes for RISC in *D. melanogaster* systems have been reported within a range of less than 230 to 500 kDa, however size determinations were conducted without having affinity purified RISC. Our affinity-purified RISC sediments in a narrow range between the size markers of 66 and 158 kDa. The differences to the reported sizes for RISC are not species-specific as we observed a similar size for RISC in *D. melanogaster* S2 cell cytoplasmic extracts after affinity purification (data not shown).

[0126] It has previously been proposed that siRNAs act as primers for target RNA-templated dsRNA synthesis (Lipardi et al., Cell 107 (2001), 297-307) although homologs for such RNA-dependent RNA polymerases known to participate in gene silencing in other systems are not identified in *D. melanogaster* or mammalian genomes. Analysis of the fate of siRNA duplexes in the HeLa cell system did not provide evidence for such a siRNA-primed activity (FIG. 3), but indicates that the predominant pathway for siRNA-mediated gene silencing is sequence-specific endonucleolytic target RNA degradation.

Single-Stranded 5' Phosphorylated Antisense siRNAs as Triggers of Mammalian Gene Silencing

[0127] It was previously noted that introduction of sense and antisense RNAs of several hundred nucleotides in length into *C. elegans* was able to sequence-specifically silence homologous genes (Guo and Kemphues, Cell 81 (1995), 611-620). Later, it was suggested that the sense and antisense RNA preparation were contaminated with a small amount of dsRNA, which was responsible for the silencing effect and is a much more potent inducer of gene silencing (Fire et al. (1998), supra). It is however conceivable that antisense RNA directly contributed to initiation of silencing. Indeed, most recently it was shown that antisense RNAs between 22 and 40 nt, but not sense RNAs were able to activate gene silencing in *C. elegans* (Tijsterman et al., Science 295 (2002), 694-697). The authors, however, favored the hypothesis of siRNA-primed dsRNA synthesis.

[0128] We have shown that modification of the 3' ends of antisense siRNA did not interfere with reconstitution of RISC in the human system. Together, these observations suggest that the driving forces for gene silencing in *C. elegans* may be predominantly dsRNA synthesis followed by Dicer cleavage, while in human and possibly also in *D. melanogaster* RISC-specific target mRNA degradation predominates.

[0129] Targeting of endogenously expressed lamin A/C by transfection of duplex siRNA into HeLa cells was the first reported example of siRNA-induced gene silencing. Lamin A/C protein was drastically reduced by a lamin A/C-specific siRNA duplex within two days post transfection, while unspecific siRNA duplexes showed no effect. At the time, transfection of non-phosphorylated sense or antisense siRNA did not reveal a substantial effect on lamin A/C levels,

although more recently a minor reduction upon antisense siRNA transfection was noticed when similar concentrations of antisense siRNA were delivered as described in this study. However, the effect was not interpreted as RISC-specific effect. Assaying 5'-phosphorylated antisense siRNAs revealed a substantial increase in lamin A/C silencing. Probably, 5' phosphorylated siRNAs are more stable or enter RISC more rapidly. Alternatively, the 5' end of transfected single-stranded siRNA may be less rapidly phosphorylated in the cell in comparison to duplex siRNAs.

[0130] Finally, it should be noted that HeLa cells are generally poor in nucleases and represent one of the preferred mammalian systems for studying RNA-processing or transcription reactions in vivo and in vitro. However, it can be expected that 5' phosphorylated single-stranded antisense siRNAs are suitable to knockdown gene expression in other cell types or tissues with a different content of nucleases, since chemical strategies to improve nuclease resistance of single stranded RNA are available. The general silencing ability of various cell types may also depend on the relative levels of siRNA/miRNA-free eIF2C1 and eIF2C2 proteins capable of associating with exogenously delivered siRNAs.

[0131] In summary, single-stranded 5'-phosphorylated antisense siRNAs of 19- to 29-nt in size broaden the use of RNA molecules for gene silencing because they can enter the mammalian RNAi pathway in vitro as well as in vivo through reconstitution of RISC. Human eIF2C1 and/or eIF2C2 seem to play a critical role in this process. Considering the feasibility of modulating the stability and uptake properties of single-stranded RNAs, 5'-phosphorylated single-stranded antisense siRNAs may further expand the utility of RNAi-based gene silencing technology as tool for functional genomics as well as therapeutic applications.

[0132] Argonaute proteins are a distinct class of proteins, containing a PAZ and Piwi domain (Cerutti et al., 2000, supra) and have been implicated in many processes previously linked to post-transcriptional silencing, however only limited biochemical information is available.

[0133] Human eIF2C2 is the ortholog of rat GERp95, which was identified as a component of the Golgi complex or the endoplasmic reticulum and copurified with intracellular membranes (Cikaluk et al., Mol. Biol. Cell 10 (1999), 3357-3722). More recently, HeLa cell eIF2C2 was shown to be associated with microRNAs and components of the SMN

complex, a regulator of ribonucleoprotein assembly, suggesting that eIF2C2 plays a role in miRNA precursor processing or miRNA function (Mourelatos et al., Genes & Dev. 16 (2002), 720-728). A more provocative hypothesis is that miRNAs are also in a RISC-like complex, which could potentially mediate target RNA degradation, if only perfectly matched miRNA target mRNAs existed. Sequence analysis using cloned human and mouse, however, did not reveal the presence of such perfectly complementary sequences in the genomes (Lagos-Quintana et al., Science 294 (2001), 853-858). Therefore, miRNPs may only function as translational regulators of partially mismatched target mRNAs, probably by recruiting additional factors that prevent dissociation from mismatched target mRNAs.

[0134] Human eIF2C1 has not been linked to gene silencing previously, but it is more than 80% similar in sequence to eIF2C2 (Koesters et al., Genomics 61 (1999), 210-218). This similarity may indicate functional redundancy, but it is also conceivable that functional RISC may contain eIF2C1 and eIF2C2 heterodimers. The predicted molecular weight of this heterodimeric complex would be slightly larger than the observed size of 90-160 kDa, but because size fractionation is based on globular shape, we can not disregard this possibility at this time.

[0135] Due to the high conservation between the members of the Argonaute family, it is possible that peptides that derive from regions 100% conserved in the 6 predicted proteins, may belong to members others than eIF2C1 and eIF2C2. In this respect, three peptides were identified with masses corresponding to HILI, meaning that this protein might be also a component of RISC.

[0136] To precisely assess the protein composition of RISC, reconstitution of the siRNA-mediated target RNA cleavage must be achieved by using recombinant proteins which may be obtained by cloning and expression in suitable bacterial or eukaryotic systems.

[0137] We expect that the biochemical characterization or the siRNA-mediated target RNA degradation process will have immediate applications, such as the development of cell lines or transgenic animals overexpressing RISC components. The efficiency in targeting endogenous genes in those lines or organisms will be enhanced. Furthermore, a reconstituted in vitro system for RNAi will allow the design of more potent and specific siRNA to achieve gene silencing.

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<400> SEQUENCE: 20

ucgaaguauu ccgcguacgu g 21
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<210> SEQ ID NO 21
<211> LENGTH: 21
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
sense siRNA (5'-3')

<400> SEQUENCE: 21

cguacgcgga auacuucgaa a

21

<210> SEQ ID NO 22
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: RNA/DNA hybrid
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
antisense siRNA (5'-3')
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (20)..(20)
<223> OTHER INFORMATION: n = 2'-deoxythymidine
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (21)..(21)
<223> OTHER INFORMATION: n = 2'-deoxyguanosine

<400> SEQUENCE: 22

ucgaaguauu ccgcguacgn n

21

<210> SEQ ID NO 23
<211> LENGTH: 21
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
sense siRNA (5'-3')

<400> SEQUENCE: 23

cguacgcgga auacuucgaa a

21

<210> SEQ ID NO 24
<211> LENGTH: 21
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
antisense siRNA (5'-3')

<400> SEQUENCE: 24

ucgaaguauu ccgcguacgu u

21

<210> SEQ ID NO 25
<211> LENGTH: 21
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100

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sense sirna (5'-3')

<400> SEQUENCE: 25

cguacgcgga auacuucgaa a 21

<210> SEQ ID NO 26

<211> LENGTH: 21

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (1)..(21)

<223> OTHER INFORMATION: RNA/DNA hybrid

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: HeLa S100 antisense sirna (5'-3')

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (20)..(21)

<223> OTHER INFORMATION: n = 2'-deoxythymidine

<400> SEQUENCE: 26

ucgaaguauu ccgcguacgn n 21

<210> SEQ ID NO 27

<211> LENGTH: 21

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (1)..(21)

<223> OTHER INFORMATION: RNA/DNA hybrid

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: HeLa S100 antisense sirna (5'-3')

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (1)..(1)

<223> OTHER INFORMATION: n = 2'-deoxythymidine

<400> SEQUENCE: 27

ncgaaguauu ccgcguacgu u 21

<210> SEQ ID NO 28

<211> LENGTH: 21

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (1)..(21)

<223> OTHER INFORMATION: RNA/DNA hybrid

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: HeLa S100 sense sirna (5'-3')

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (1)..(1)

<223> OTHER INFORMATION: n = 2'-deoxycytidine

<400> SEQUENCE: 28

nguacgcgga auacuucgau u 21

<210> SEQ ID NO 29

<211> LENGTH: 21

<212> TYPE: DNA

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<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: RNA/DNA hybrid
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
 antisense siRNA (5'-3')
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(1)
<223> OTHER INFORMATION: n = 2'-deoxythymidine

<400> SEQUENCE: 29

ncgaaguauu ccgcguacgu u

21

<210> SEQ ID NO 30
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: RNA/DNA hybrid
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100 cells
 antisense siRNA (5'-3')
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: n = 2'-deoxythymidine

<400> SEQUENCE: 30

ncgaaguauu ccgcguacgn n

21

<210> SEQ ID NO 31
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: RNA/DNA hybrid
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
 sense siRNA (5'-3')
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(1)
<223> OTHER INFORMATION: n = 2'-deoxycytidine
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (20)..(21)
<223> OTHER INFORMATION: n = 2'-deoxythymidine

<400> SEQUENCE: 31

nguacgcgga auacuucgan n

21

<210> SEQ ID NO 32
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: RNA/DNA hybrid
<220> FEATURE:

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<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
        antisense siRNA (5'-3')
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: n = 2'-deoxythymidine

<400> SEQUENCE: 32

ncgaaguauu ccgcguacgn n                                     21

```

```

<210> SEQ ID NO 33
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: RNA/DNA hybrid
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
        antisense siRNA (5'-3')
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: n = 2'-deoxythymidine

<400> SEQUENCE: 33

ncgaaguauu ccgcguacgn n                                     21

```

```

<210> SEQ ID NO 34
<211> LENGTH: 13
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of HILI, corresponding to
        position 17-29 of the protein

<400> SEQUENCE: 34

Asn Lys Gln Asp Phe Met Asp Leu Ser Ile Cys Thr Arg
1           5           10

```

```

<210> SEQ ID NO 35
<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of HILI, corresponding to
        position 436-449 of the protein

<400> SEQUENCE: 35

Thr Glu Tyr Val Ala Glu Ser Phe Leu Asn Cys Leu Arg Arg
1           5           10

```

```

<210> SEQ ID NO 36
<211> LENGTH: 13
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of HILI, corresponding to
        position 591-603 of the protein

<400> SEQUENCE: 36

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-continued

Tyr Asn His Asp Leu Pro Ala Arg Ile Ile Val Tyr Arg
1 5 10

<210> SEQ ID NO 37
<211> LENGTH: 35
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
target RNA

<400> SEQUENCE: 37

aacaucacgu acgcggaaua cuucgaaaug uccgu 35

<210> SEQ ID NO 38
<211> LENGTH: 21
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
strand of siRNA duplex

<400> SEQUENCE: 38

cguacgcgga auacuucgau u 21

<210> SEQ ID NO 39
<211> LENGTH: 21
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
strand of siRNA duplex

<400> SEQUENCE: 39

ucgaaguauu ccgcguacgu u 21

<210> SEQ ID NO 40
<211> LENGTH: 21
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
strand of siRNA duplex

<400> SEQUENCE: 40

cguacgcgga auacuucgaa a 21

<210> SEQ ID NO 41
<211> LENGTH: 20
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
strand of siRNA duplex

<400> SEQUENCE: 41

ucgaaguauu ccgcguacgu 20

<210> SEQ ID NO 42
<211> LENGTH: 12

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<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 42

Val Leu Gln Pro Pro Ser Ile Leu Tyr Gly Gly Arg
1 5 10

<210> SEQ ID NO 43
<211> LENGTH: 12
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 43

Gln Glu Ile Ile Gln Asp Leu Ala Ala Met Val Arg
1 5 10

<210> SEQ ID NO 44
<211> LENGTH: 12
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 44

His Leu Pro Ser Met Arg Tyr Thr Pro Val Gly Arg
1 5 10

<210> SEQ ID NO 45
<211> LENGTH: 12
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 45

Lys Leu Thr Asp Asn Gln Thr Ser Thr Met Ile Arg
1 5 10

<210> SEQ ID NO 46
<211> LENGTH: 13
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 46

Tyr Ala Gln Gly Ala Asp Ser Val Glu Pro Met Phe Arg
1 5 10

<210> SEQ ID NO 47
<211> LENGTH: 14
<212> TYPE: PRT

-continued

<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 47

Asp Lys Val Glu Leu Glu Val Thr Leu Pro Gly Glu Gly Lys
1 5 10

<210> SEQ ID NO 48
<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 48

Asp Ala Gly Met Pro Ile Gln Gly Gln Pro Cys Phe Cys Lys
1 5 10

<210> SEQ ID NO 49
<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 49

Thr Gln Ile Phe Gly Asp Arg Lys Pro Val Phe Asp Gly Arg
1 5 10

<210> SEQ ID NO 50
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 50

Ala Thr Ala Arg Ser Ala Pro Asp Arg Gln Glu Glu Ile Ser Lys
1 5 10 15

<210> SEQ ID NO 51
<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 51

Asp Tyr Gln Pro Gly Ile Thr Phe Ile Val Val Gln Lys Arg
1 5 10

<210> SEQ ID NO 52
<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

-continued

<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 52

Ser Ala Pro Asp Arg Gln Glu Glu Ile Ser Lys Leu Met Arg
1 5 10

<210> SEQ ID NO 53
<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 53

Tyr Pro His Leu Pro Cys Leu Gln Val Gly Gln Glu Gln Lys
1 5 10

<210> SEQ ID NO 54
<211> LENGTH: 17
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 54

Ser Phe Phe Thr Ala Ser Glu Gly Cys Ser Asn Pro Leu Gly Gly Gly
1 5 10 15

Arg

<210> SEQ ID NO 55
<211> LENGTH: 23
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 55

Tyr His Leu Val Asp Lys Glu His Asp Ser Ala Glu Gly Ser His Thr
1 5 10 15

Ser Gly Gln Ser Asn Gly Arg
20

<210> SEQ ID NO 56
<211> LENGTH: 12
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 56

Val Leu Pro Ala Pro Ile Leu Gln Tyr Gly Gly Arg
1 5 10

-continued

<210> SEQ ID NO 57
<211> LENGTH: 12
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 57

Ser Val Ser Ile Pro Ala Pro Ala Tyr Tyr Ala Arg
1 5 10

<210> SEQ ID NO 58
<211> LENGTH: 12
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 58

Thr Ser Pro Gln Thr Leu Ser Asn Leu Cys Leu Lys
1 5 10

<210> SEQ ID NO 59
<211> LENGTH: 13
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 59

Tyr Ala Gln Gly Ala Asp Ser Val Glu Pro Met Phe Arg
1 5 10

<210> SEQ ID NO 60
<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 60

Asn Ile Tyr Thr Val Thr Ala Leu Pro Ile Gly Asn Glu Arg
1 5 10

<210> SEQ ID NO 61
<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 61

Val Asp Phe Glu Val Thr Ile Pro Gly Glu Gly Lys Asp Arg
1 5 10

<210> SEQ ID NO 62

-continued

<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: HeLa S100 cells
peptide fragment of eIF2C1 obtained by mass spectrometry

<400> SEQUENCE: 62

Asp Ala Gly Met Pro Ile Gln Gly Gln Pro Cys Phe Cys Lys
1 5 10

<210> SEQ ID NO 63
<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass
spectrometry

<400> SEQUENCE: 63

Asn Ile Asp Glu Gln Pro Lys Pro Leu Thr Asp Ser Gln Arg
1 5 10

<210> SEQ ID NO 64
<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass
spectrometry

<400> SEQUENCE: 64

Ser Ala Pro Asp Arg Gln Glu Glu Ile Ser Arg Leu Met Lys
1 5 10

<210> SEQ ID NO 65
<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass
spectrometry

<400> SEQUENCE: 65

Asp Tyr Gln Pro Gly Ile Thr Tyr Ile Val Val Gln Lys Arg
1 5 10

<210> SEQ ID NO 66
<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass
spectrometry

<400> SEQUENCE: 66

Tyr Pro His Leu Pro Cys Leu Gln Val Gly Gln Glu Gln Lys
1 5 10

<210> SEQ ID NO 67
<211> LENGTH: 17

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<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass
spectrometry

```

```

<400> SEQUENCE: 67

```

```

Ser Phe Phe Ser Pro Pro Glu Gly Tyr Tyr His Pro Leu Gly Gly Gly
1           5           10           15

```

```

Arg

```

```

<210> SEQ ID NO 68
<211> LENGTH: 857
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: eIF2C1, predicted protein sequence

```

```

<400> SEQUENCE: 68

```

```

Met Glu Ala Gly Pro Ser Gly Ala Ala Ala Gly Ala Tyr Leu Pro Pro
1           5           10           15

```

```

Leu Gln Gln Val Phe Gln Ala Pro Arg Arg Pro Gly Ile Gly Thr Val
20           25           30

```

```

Gly Lys Pro Ile Lys Leu Leu Ala Asn Tyr Phe Glu Val Asp Ile Pro
35           40           45

```

```

Lys Ile Asp Val Tyr His Tyr Glu Val Asp Ile Lys Pro Asp Lys Cys
50           55           60

```

```

Pro Arg Arg Val Asn Arg Glu Val Val Glu Tyr Met Val Gln His Phe
65           70           75           80

```

```

Lys Pro Gln Ile Phe Gly Asp Arg Lys Pro Val Tyr Asp Gly Lys Lys
85           90           95

```

```

Asn Ile Tyr Thr Val Thr Ala Leu Pro Ile Gly Asn Glu Arg Val Asp
100          105          110

```

```

Phe Glu Val Thr Ile Pro Gly Glu Gly Lys Asp Arg Ile Phe Lys Val
115          120          125

```

```

Ser Ile Lys Trp Leu Ala Ile Val Ser Trp Arg Met Leu His Glu Ala
130          135          140

```

```

Leu Val Ser Gly Gln Ile Pro Val Pro Leu Glu Ser Val Gln Ala Leu
145          150          155          160

```

```

Asp Val Ala Met Arg His Leu Ala Ser Met Arg Tyr Thr Pro Val Gly
165          170          175

```

```

Arg Ser Phe Phe Ser Pro Pro Glu Gly Tyr Tyr His Pro Leu Gly Gly
180          185          190

```

```

Gly Arg Glu Val Trp Phe Gly Phe His Gln Ser Val Arg Pro Ala Met
195          200          205

```

```

Trp Lys Met Met Leu Asn Ile Asp Val Ser Ala Thr Ala Phe Tyr Lys
210          215          220

```

```

Ala Gln Pro Val Ile Glu Phe Met Cys Glu Val Leu Asp Ile Arg Asn
225          230          235          240

```

```

Ile Asp Glu Gln Pro Lys Pro Leu Thr Asp Ser Gln Arg Val Arg Phe
245          250          255

```

```

Thr Lys Glu Ile Lys Gly Leu Lys Val Glu Val Thr His Cys Gly Gln
260          265          270

```

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Met	Lys	Arg	Lys	Tyr	Arg	Val	Cys	Asn	Val	Thr	Arg	Arg	Pro	Ala	Ser	275	280	285
His	Gln	Thr	Phe	Pro	Leu	Gln	Leu	Glu	Ser	Gly	Gln	Thr	Val	Glu	Cys	290	295	300
Thr	Val	Ala	Gln	Tyr	Phe	Lys	Gln	Lys	Tyr	Asn	Leu	Gln	Leu	Lys	Tyr	305	310	315
Pro	His	Leu	Pro	Cys	Leu	Gln	Val	Gly	Gln	Glu	Gln	Lys	His	Thr	Tyr	325	330	335
Leu	Pro	Leu	Glu	Val	Cys	Asn	Ile	Val	Ala	Gly	Gln	Arg	Cys	Ile	Lys	340	345	350
Lys	Leu	Thr	Asp	Asn	Gln	Thr	Ser	Thr	Met	Ile	Lys	Ala	Thr	Ala	Arg	355	360	365
Ser	Ala	Pro	Asp	Arg	Gln	Glu	Glu	Ile	Ser	Arg	Leu	Met	Lys	Asn	Ala	370	375	380
Ser	Tyr	Asn	Leu	Asp	Pro	Tyr	Ile	Gln	Glu	Phe	Gly	Ile	Lys	Val	Lys	385	390	395
Asp	Asp	Met	Thr	Glu	Val	Thr	Gly	Arg	Val	Leu	Pro	Ala	Pro	Ile	Leu	405	410	415
Gln	Tyr	Gly	Gly	Arg	Asn	Arg	Ala	Ile	Ala	Thr	Pro	Asn	Gln	Gly	Val	420	425	430
Trp	Asp	Met	Arg	Gly	Lys	Gln	Phe	Tyr	Asn	Gly	Ile	Glu	Ile	Lys	Val	435	440	445
Trp	Ala	Ile	Ala	Cys	Phe	Ala	Pro	Gln	Lys	Gln	Cys	Arg	Glu	Glu	Val	450	455	460
Leu	Lys	Asn	Phe	Thr	Asp	Gln	Leu	Arg	Lys	Ile	Ser	Lys	Asp	Ala	Gly	465	470	475
Met	Pro	Ile	Gln	Gly	Gln	Pro	Cys	Phe	Cys	Lys	Tyr	Ala	Gln	Gly	Ala	485	490	495
Asp	Ser	Val	Glu	Pro	Met	Phe	Arg	His	Leu	Lys	Asn	Thr	Tyr	Ser	Gly	500	505	510
Leu	Gln	Leu	Ile	Ile	Val	Ile	Leu	Pro	Gly	Lys	Thr	Pro	Val	Tyr	Ala	515	520	525
Glu	Val	Lys	Arg	Val	Gly	Asp	Thr	Leu	Leu	Gly	Met	Ala	Thr	Gln	Cys	530	535	540
Val	Gln	Val	Lys	Asn	Val	Val	Lys	Thr	Ser	Pro	Gln	Thr	Leu	Ser	Asn	545	550	555
Leu	Cys	Leu	Lys	Ile	Asn	Val	Lys	Leu	Gly	Gly	Ile	Asn	Asn	Ile	Leu	565	570	575
Val	Pro	His	Gln	Arg	Ser	Ala	Val	Phe	Gln	Gln	Pro	Val	Ile	Phe	Leu	580	585	590
Gly	Ala	Asp	Val	Thr	His	Pro	Pro	Ala	Gly	Asp	Gly	Lys	Lys	Pro	Ser	595	600	605
Ile	Thr	Ala	Val	Val	Gly	Ser	Met	Asp	Ala	His	Pro	Ser	Arg	Tyr	Cys	610	615	620
Ala	Thr	Val	Arg	Val	Gln	Arg	Pro	Arg	Gln	Glu	Ile	Ile	Glu	Asp	Leu	625	630	635
Ser	Tyr	Met	Val	Arg	Glu	Leu	Leu	Ile	Gln	Phe	Tyr	Lys	Ser	Thr	Arg	645	650	655
Phe	Lys	Pro	Thr	Arg	Ile	Ile	Phe	Tyr	Arg	Asp	Gly	Val	Pro	Glu	Gly	660	665	670
Gln	Leu	Pro	Gln	Ile	Leu	His	Tyr	Glu	Leu	Leu	Ala	Ile	Arg	Asp	Ala			

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675					680					685					
Cys	Ile	Lys	Leu	Glu	Lys	Asp	Tyr	Gln	Pro	Gly	Ile	Thr	Tyr	Ile	Val
690					695					700					
Val	Gln	Lys	Arg	His	His	Thr	Arg	Leu	Phe	Cys	Ala	Asp	Lys	Asn	Glu
705					710					715					720
Arg	Ile	Gly	Lys	Ser	Gly	Asn	Ile	Pro	Ala	Gly	Thr	Thr	Val	Asp	Thr
			725						730					735	
Asn	Ile	Thr	His	Pro	Phe	Glu	Phe	Asp	Phe	Tyr	Leu	Cys	Ser	His	Ala
			740					745					750		
Gly	Ile	Gln	Gly	Thr	Ser	Arg	Pro	Ser	His	Tyr	Tyr	Val	Leu	Trp	Asp
		755					760					765			
Asp	Asn	Arg	Phe	Thr	Ala	Asp	Glu	Leu	Gln	Ile	Leu	Thr	Tyr	Gln	Leu
	770					775					780				
Cys	His	Thr	Tyr	Val	Arg	Cys	Thr	Arg	Ser	Val	Ser	Ile	Pro	Ala	Pro
785					790					795					800
Ala	Tyr	Tyr	Ala	Arg	Leu	Val	Ala	Phe	Arg	Ala	Arg	Tyr	His	Leu	Val
			805						810					815	
Asp	Lys	Glu	His	Asp	Ser	Gly	Glu	Gly	Ser	His	Ile	Ser	Gly	Gln	Ser
			820					825					830		
Asn	Gly	Arg	Asp	Pro	Gln	Ala	Leu	Ala	Lys	Ala	Val	Gln	Val	His	Gln
		835					840					845			
Asp	Thr	Leu	Arg	Thr	Met	Tyr	Phe	Ala							
	850					855									

<210> SEQ ID NO 69

<211> LENGTH: 860

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: eIF2C2, predicted protein sequence

<400> SEQUENCE: 69

Met	Gly	Val	Leu	Ser	Ala	Ile	Pro	Ala	Leu	Ala	Pro	Pro	Ala	Pro	Pro
1			5						10				15		
Pro	Pro	Ile	Gln	Gly	Tyr	Ala	Phe	Lys	Pro	Pro	Pro	Arg	Pro	Asp	Phe
		20					25					30			
Gly	Thr	Ser	Gly	Arg	Thr	Ile	Lys	Leu	Gln	Ala	Asn	Phe	Phe	Glu	Met
	35					40					45				
Asp	Ile	Pro	Lys	Ile	Asp	Ile	Tyr	His	Tyr	Glu	Leu	Asp	Ile	Lys	Pro
	50				55					60					
Glu	Lys	Cys	Pro	Arg	Arg	Val	Asn	Arg	Glu	Ile	Val	Glu	His	Met	Val
65				70					75					80	
Gln	His	Phe	Lys	Thr	Gln	Ile	Phe	Gly	Asp	Arg	Lys	Pro	Val	Phe	Asp
			85					90					95		
Gly	Arg	Lys	Asn	Leu	Tyr	Thr	Ala	Met	Pro	Leu	Pro	Ile	Gly	Arg	Asp
	100						105					110			
Lys	Val	Glu	Leu	Glu	Val	Thr	Leu	Pro	Gly	Glu	Gly	Lys	Asp	Arg	Ile
	115					120					125				
Phe	Lys	Val	Ser	Ile	Lys	Trp	Val	Ser	Cys	Val	Ser	Leu	Gln	Ala	Leu
	130				135					140					
His	Asp	Ala	Leu	Ser	Gly	Arg	Leu	Pro	Ser	Val	Pro	Phe	Glu	Thr	Ile
145				150					155					160	

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Gln	Ala	Leu	Asp	Val	Val	Met	Arg	His	Leu	Pro	Ser	Met	Arg	Tyr	Thr	165	170	175	
Pro	Val	Gly	Arg	Ser	Phe	Phe	Thr	Ala	Ser	Glu	Gly	Cys	Ser	Asn	Pro	180	185	190	
Leu	Gly	Gly	Gly	Arg	Glu	Val	Trp	Phe	Gly	Phe	His	Gln	Ser	Val	Arg	195	200	205	
Pro	Ser	Leu	Trp	Lys	Met	Met	Leu	Asn	Ile	Asp	Val	Ser	Ala	Thr	Ala	210	215	220	
Phe	Tyr	Lys	Ala	Gln	Pro	Val	Ile	Glu	Phe	Val	Cys	Glu	Val	Leu	Asp	225	230	235	240
Phe	Lys	Ser	Ile	Glu	Glu	Gln	Gln	Lys	Pro	Leu	Thr	Asp	Ser	Gln	Arg	245	250	255	
Val	Lys	Phe	Thr	Lys	Glu	Ile	Lys	Gly	Leu	Lys	Val	Glu	Ile	Thr	His	260	265	270	
Cys	Gly	Gln	Met	Lys	Arg	Lys	Tyr	Arg	Val	Cys	Asn	Val	Thr	Arg	Arg	275	280	285	
Pro	Ala	Ser	His	Gln	Thr	Phe	Pro	Leu	Gln	Gln	Glu	Ser	Gly	Gln	Thr	290	295	300	
Val	Glu	Cys	Thr	Val	Ala	Gln	Tyr	Phe	Lys	Asp	Arg	His	Lys	Leu	Val	305	310	315	320
Leu	Arg	Tyr	Pro	His	Leu	Pro	Cys	Leu	Gln	Val	Gly	Gln	Glu	Gln	Lys	325	330	335	
His	Thr	Tyr	Leu	Pro	Leu	Glu	Val	Cys	Asn	Ile	Val	Ala	Gly	Gln	Arg	340	345	350	
Cys	Ile	Lys	Lys	Leu	Thr	Asp	Asn	Gln	Thr	Ser	Thr	Met	Ile	Arg	Ala	355	360	365	
Thr	Ala	Arg	Ser	Ala	Pro	Asp	Arg	Gln	Glu	Glu	Ile	Ser	Lys	Leu	Met	370	375	380	
Arg	Ser	Ala	Ser	Phe	Asn	Thr	Asp	Pro	Tyr	Val	Arg	Glu	Phe	Gly	Ile	385	390	395	400
Met	Val	Lys	Asp	Glu	Met	Thr	Asp	Val	Thr	Gly	Arg	Val	Leu	Gln	Pro	405	410	415	
Pro	Ser	Ile	Leu	Tyr	Gly	Gly	Arg	Asn	Lys	Ala	Ile	Ala	Thr	Pro	Val	420	425	430	
Gln	Gly	Val	Trp	Asp	Met	Arg	Asn	Lys	Gln	Phe	His	Thr	Gly	Ile	Glu	435	440	445	
Ile	Lys	Val	Trp	Ala	Ile	Ala	Cys	Phe	Ala	Pro	Gln	Arg	Gln	Cys	Thr	450	455	460	
Glu	Val	His	Leu	Lys	Ser	Phe	Thr	Glu	Gln	Leu	Arg	Lys	Ile	Ser	Arg	465	470	475	480
Asp	Ala	Gly	Met	Pro	Ile	Gln	Gly	Gln	Pro	Cys	Phe	Cys	Lys	Tyr	Ala	485	490	495	
Gln	Gly	Ala	Asp	Ser	Val	Glu	Pro	Met	Phe	Arg	His	Leu	Lys	Asn	Thr	500	505	510	
Tyr	Ala	Gly	Leu	Gln	Leu	Val	Val	Val	Ile	Leu	Pro	Gly	Lys	Thr	Pro	515	520	525	
Val	Tyr	Ala	Glu	Val	Lys	Arg	Val	Gly	Asp	Thr	Val	Leu	Gly	Met	Ala	530	535	540	
Thr	Gln	Cys	Val	Gln	Met	Lys	Asn	Val	Gln	Arg	Thr	Thr	Pro	Gln	Thr	545	550	555	560
Leu	Ser	Asn	Leu	Cys	Leu	Lys	Ile	Asn	Val	Lys	Leu	Gly	Gly	Val	Asn				

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565					570					575					
Asn	Ile	Leu	Leu	Pro	Gln	Gly	Arg	Pro	Pro	Val	Phe	Gln	Gln	Pro	Val
			580					585					590		
Ile	Phe	Leu	Gly	Ala	Asp	Val	Thr	His	Pro	Pro	Ala	Gly	Asp	Gly	Lys
		595					600					605			
Lys	Pro	Ser	Ile	Ala	Ala	Val	Val	Gly	Ser	Met	Asp	Ala	His	Pro	Asn
	610					615					620				
Arg	Tyr	Cys	Ala	Thr	Val	Arg	Val	Gln	Gln	His	Arg	Gln	Glu	Ile	Ile
	625					630					635				640
Gln	Asp	Leu	Ala	Ala	Met	Val	Arg	Glu	Leu	Leu	Ile	Gln	Phe	Tyr	Lys
			645						650					655	
Ser	Thr	Arg	Phe	Lys	Pro	Thr	Arg	Ile	Ile	Phe	Tyr	Arg	Asp	Gly	Val
			660					665					670		
Ser	Glu	Gly	Gln	Phe	Gln	Gln	Val	Leu	His	His	Glu	Leu	Leu	Ala	Ile
		675					680					685			
Arg	Glu	Ala	Cys	Ile	Lys	Leu	Glu	Lys	Asp	Tyr	Gln	Pro	Gly	Ile	Thr
	690					695					700				
Phe	Ile	Val	Val	Gln	Lys	Arg	His	His	Thr	Arg	Leu	Phe	Cys	Thr	Asp
	705					710					715				720
Lys	Asn	Glu	Arg	Val	Gly	Lys	Ser	Gly	Asn	Ile	Pro	Ala	Gly	Thr	Thr
			725						730					735	
Val	Asp	Thr	Lys	Ile	Thr	His	Pro	Thr	Glu	Phe	Asp	Phe	Tyr	Leu	Cys
			740					745					750		
Ser	His	Ala	Gly	Ile	Gln	Gly	Thr	Ser	Arg	Pro	Ser	His	Tyr	His	Val
		755					760					765			
Leu	Trp	Asp	Asp	Asn	Arg	Phe	Ser	Ser	Asp	Glu	Leu	Gln	Ile	Leu	Thr
	770					775					780				
Tyr	Gln	Leu	Cys	His	Thr	Tyr	Val	Arg	Cys	Thr	Arg	Ser	Val	Ser	Ile
	785					790					795				800
Pro	Ala	Pro	Ala	Tyr	Tyr	Ala	His	Leu	Val	Ala	Phe	Arg	Ala	Arg	Tyr
			805						810					815	
His	Leu	Val	Asp	Lys	Glu	His	Asp	Ser	Ala	Glu	Gly	Ser	His	Thr	Ser
			820					825					830		
Gly	Gln	Ser	Asn	Gly	Arg	Asp	His	Gln	Ala	Leu	Ala	Lys	Ala	Val	Gln
		835					840					845			
Val	His	Gln	Asp	Thr	Leu	Arg	Thr	Met	Tyr	Phe	Ala				
	850					855					860				

<210> SEQ ID NO 70

<211> LENGTH: 924

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: eIF2C3, predicted protein sequence

<400> SEQUENCE: 70

Ser	Arg	Ser	Arg	Val	Pro	Val	Pro	Gly	Pro	Gly	Ala	Ala	Ala	Ala	Pro
1				5				10						15	
Cys	Pro	Ala	Pro	Ala	Ser	Pro	Arg	Arg	His	Pro	Ser	Ala	Asn	Ile	Pro
			20				25						30		
Glu	Ile	Lys	Arg	Tyr	Ala	Ala	Ala	Ala	Ala	Ala	Ala	Ala	Gly	Pro	Gly
	35						40						45		

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Ala	Gly	Gly	Ala	Gly	Asp	Arg	Gly	Glu	Ala	Ala	Pro	Ala	Ala	Ala	Met
50					55						60				
Glu	Ala	Leu	Gly	Pro	Gly	Pro	Pro	Ala	Ser	Leu	Phe	Gln	Pro	Pro	Arg
65				70					75						80
Arg	Pro	Gly	Leu	Gly	Thr	Val	Gly	Lys	Pro	Ile	Arg	Leu	Leu	Ala	Asn
			85					90						95	
His	Phe	Gln	Val	Gln	Ile	Pro	Lys	Ile	Asp	Val	Tyr	His	Tyr	Asp	Val
			100				105						110		
Asp	Ile	Lys	Pro	Glu	Lys	Arg	Pro	Arg	Arg	Val	Asn	Arg	Glu	Val	Val
		115					120					125			
Asp	Thr	Met	Val	Arg	His	Phe	Lys	Met	Gln	Ile	Phe	Gly	Asp	Arg	Gln
	130					135						140			
Pro	Gly	Tyr	Asp	Gly	Lys	Arg	Asn	Met	Tyr	Thr	Ala	His	Pro	Leu	Pro
145					150					155					160
Ile	Gly	Arg	Asp	Arg	Val	Asp	Met	Glu	Val	Thr	Leu	Pro	Gly	Glu	Gly
			165					170						175	
Lys	Asp	Gln	Thr	Phe	Lys	Val	Ser	Val	Gln	Trp	Val	Ser	Val	Val	Ser
			180					185					190		
Leu	Gln	Leu	Leu	Leu	Glu	Ala	Leu	Ala	Gly	His	Leu	Asn	Glu	Val	Pro
		195					200					205			
Asp	Asp	Ser	Val	Gln	Ala	Leu	Asp	Val	Ile	Thr	Arg	His	Leu	Pro	Ser
	210					215					220				
Met	Arg	Tyr	Thr	Pro	Val	Gly	Arg	Ser	Phe	Phe	Ser	Pro	Pro	Glu	Gly
225					230					235					240
Tyr	Tyr	His	Pro	Leu	Gly	Gly	Gly	Arg	Glu	Val	Trp	Phe	Gly	Phe	His
			245					250						255	
Gln	Ser	Val	Arg	Pro	Ala	Met	Trp	Asn	Met	Met	Leu	Asn	Ile	Asp	Val
			260					265					270		
Ser	Ala	Thr	Ala	Phe	Tyr	Arg	Ala	Gln	Pro	Ile	Ile	Glu	Phe	Met	Cys
		275					280					285			
Glu	Val	Leu	Asp	Ile	Gln	Asn	Ile	Asn	Glu	Gln	Thr	Lys	Pro	Leu	Thr
	290					295					300				
Asp	Ser	Gln	Arg	Val	Lys	Phe	Thr	Lys	Glu	Ile	Arg	Gly	Leu	Lys	Val
305					310					315					320
Glu	Val	Thr	His	Cys	Gly	Gln	Met	Lys	Arg	Lys	Tyr	Arg	Val	Cys	Asn
			325					330						335	
Val	Thr	Arg	Arg	Pro	Ala	Ser	His	Gln	Thr	Phe	Pro	Leu	Gln	Leu	Glu
			340					345					350		
Asn	Gly	Gln	Ala	Met	Glu	Cys	Thr	Val	Ala	Gln	Tyr	Phe	Lys	Gln	Lys
		355					360					365			
Tyr	Ser	Leu	Gln	Leu	Lys	Tyr	Pro	His	Leu	Pro	Cys	Leu	Gln	Val	Gly
	370				375						380				
Gln	Glu	Gln	Lys	His	Thr	Tyr	Leu	Pro	Leu	Glu	Val	Cys	Asn	Ile	Val
385					390					395					400
Ala	Gly	Gln	Arg	Cys	Ile	Lys	Lys	Leu	Thr	Asp	Asn	Gln	Thr	Ser	Thr
			405					410						415	
Met	Ile	Lys	Ala	Thr	Ala	Arg	Ser	Ala	Pro	Asp	Arg	Gln	Glu	Glu	Ile
			420					425					430		
Ser	Arg	Leu	Val	Lys	Ser	Asn	Ser	Met	Val	Gly	Gly	Pro	Asp	Pro	Tyr
		435					440					445			
Leu	Lys	Glu	Phe	Gly	Ile	Val	Val	His	Asn	Glu	Met	Thr	Glu	Leu	Thr

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450					455					460				
Gly	Arg	Val	Leu	Pro	Ala	Pro	Met	Leu	Gln	Tyr	Gly	Gly	Arg	Lys
465					470					475				480
Thr	Val	Ala	Thr	Pro	Asn	Gln	Gly	Val	Trp	Asp	Met	Arg	Gly	Lys
				485					490					495
Phe	Tyr	Ala	Gly	Ile	Glu	Ile	Lys	Val	Trp	Ala	Val	Ala	Cys	Phe
			500					505					510	Ala
Pro	Gln	Lys	Gln	Cys	Arg	Glu	Asp	Leu	Leu	Lys	Ser	Phe	Thr	Asp
		515					520					525		Gln
Leu	Arg	Lys	Ile	Ser	Lys	Asp	Ala	Gly	Met	Pro	Ile	Gln	Gly	Gln
	530					535					540			Pro
Cys	Phe	Cys	Lys	Tyr	Ala	Gln	Gly	Ala	Asp	Ser	Val	Glu	Pro	Met
545					550					555				560
Lys	His	Leu	Lys	Met	Thr	Tyr	Val	Gly	Leu	Gln	Leu	Ile	Val	Ile
				565					570					575
Leu	Pro	Gly	Lys	Thr	Pro	Val	Tyr	Ala	Glu	Val	Lys	Arg	Val	Gly
			580					585					590	Asp
Thr	Leu	Leu	Gly	Met	Ala	Thr	Gln	Cys	Val	Gln	Val	Lys	Asn	Val
		595					600					605		Val
Lys	Thr	Ser	Pro	Gln	Thr	Leu	Ser	Asn	Leu	Cys	Leu	Lys	Ile	Asn
	610					615					620			Ala
Lys	Leu	Gly	Gly	Ile	Asn	Asn	Val	Leu	Val	Pro	His	Gln	Arg	Pro
625					630					635				640
Val	Phe	Gln	Gln	Pro	Val	Ile	Phe	Leu	Gly	Ala	Asp	Val	Thr	His
				645					650					655
Pro	Ala	Gly	Asp	Gly	Lys	Lys	Pro	Ser	Ile	Ala	Ala	Val	Val	Gly
			660					665					670	Ser
Met	Asp	Gly	His	Pro	Ser	Arg	Tyr	Cys	Ala	Thr	Val	Arg	Val	Gln
		675					680					685		Thr
Ser	Arg	Gln	Glu	Ile	Ser	Gln	Glu	Leu	Leu	Tyr	Ser	Gln	Glu	Val
		690				695					700			Ile
Gln	Asp	Leu	Thr	Asn	Met	Val	Arg	Glu	Leu	Leu	Ile	Gln	Phe	Tyr
705				710						715				720
Ser	Thr	Arg	Phe	Lys	Pro	Thr	Arg	Ile	Ile	Tyr	Tyr	Arg	Gly	Gly
				725					730					735
Ser	Glu	Gly	Gln	Met	Lys	Gln	Val	Ala	Trp	Pro	Glu	Leu	Ile	Ala
			740					745					750	Ile
Arg	Lys	Ala	Cys	Ile	Ser	Leu	Glu	Glu	Asp	Tyr	Arg	Pro	Gly	Ile
		755					760					765		Thr
Tyr	Ile	Val	Val	Gln	Lys	Arg	His	His	Thr	Arg	Leu	Phe	Cys	Ala
	770					775					780			Asp
Lys	Thr	Glu	Arg	Val	Gly	Lys	Ser	Gly	Asn	Val	Pro	Ala	Gly	Thr
785					790					795				800
Val	Asp	Ser	Thr	Ile	Thr	His	Pro	Ser	Glu	Phe	Asp	Phe	Tyr	Leu
				805					810					815
Ser	His	Ala	Gly	Ile	Gln	Gly	Thr	Ser	Arg	Pro	Ser	His	Tyr	Gln
			820					825					830	Val
Leu	Trp	Asp	Asp	Asn	Cys	Phe	Thr	Ala	Asp	Glu	Leu	Gln	Leu	Leu
		835					840					845		Thr
Tyr	Gln	Leu	Cys	His	Thr	Tyr	Val	Arg	Cys	Thr	Arg	Ser	Val	Ser
	850					855					860			Ile

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Pro Ala Pro Ala Tyr Tyr Ala Arg Leu Val Ala Phe Arg Ala Arg Tyr
865                870                875                880

His Leu Val Asp Lys Asp His Asp Ser Ala Glu Gly Ser His Val Ser
      885                890                895

Gly Gln Ser Asn Gly Arg Asp Pro Gln Ala Leu Ala Lys Ala Val Gln
      900                905                910

Ile His His Asp Thr Gln His Thr Met Tyr Phe Ala
      915                920

<210> SEQ ID NO 71
<211> LENGTH: 855
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: eIF2C4, predicted protein sequence

<400> SEQUENCE: 71

Ala Gly Pro Ala Gly Ala Gln Pro Leu Leu Met Val Pro Arg Arg Pro
1      5      10      15

Gly Tyr Gly Thr Met Gly Lys Pro Ile Lys Leu Leu Ala Asn Cys Phe
      20      25      30

Gln Val Glu Ile Pro Lys Ile Asp Val Tyr Leu Tyr Glu Val Asp Ile
      35      40      45

Lys Pro Asp Lys Cys Pro Arg Arg Val Asn Arg Glu Val Val Asp Ser
      50      55      60

Met Val Gln His Phe Lys Val Thr Ile Phe Gly Asp Arg Arg Pro Val
      65      70      75      80

Tyr Asp Gly Lys Arg Ser Leu Tyr Thr Ala Asn Pro Leu Pro Val Ala
      85      90      95

Thr Thr Gly Val Asp Leu Asp Val Thr Leu Pro Gly Glu Gly Gly Lys
      100     105     110

Asp Arg Pro Phe Lys Val Ser Ile Lys Phe Val Ser Arg Val Ser Trp
      115     120     125

His Leu Leu His Glu Val Leu Thr Gly Arg Thr Leu Pro Glu Pro Leu
      130     135     140

Glu Leu Asp Lys Pro Ile Ser Thr Asn Pro Val His Ala Val Asp Val
      145     150     155     160

Val Leu Arg His Leu Pro Ser Met Lys Tyr Thr Pro Val Gly Arg Ser
      165     170     175

Phe Phe Ser Ala Pro Glu Gly Tyr Asp His Pro Leu Gly Gly Gly Arg
      180     185     190

Glu Val Trp Phe Gly Phe His Gln Ser Val Arg Pro Ala Met Trp Lys
      195     200     205

Met Met Leu Asn Ile Asp Val Ser Ala Thr Ala Phe Tyr Lys Ala Gln
      210     215     220

Pro Val Ile Gln Phe Met Cys Glu Val Leu Asp Ile His Asn Ile Asp
      225     230     235     240

Glu Gln Pro Arg Pro Leu Thr Asp Ser His Arg Val Lys Phe Thr Lys
      245     250     255

Glu Ile Lys Gly Leu Lys Val Glu Val Thr His Cys Gly Thr Met Arg
      260     265     270

Arg Lys Tyr Arg Val Cys Asn Val Thr Arg Arg Pro Ala Ser His Gln

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275					280					285					
Thr	Phe	Pro	Leu	Gln	Leu	Glu	Asn	Gly	Gln	Thr	Val	Glu	Arg	Thr	Val
	290					295					300				
Ala	Gln	Tyr	Phe	Arg	Glu	Lys	Tyr	Thr	Leu	Gln	Leu	Lys	Tyr	Pro	His
	305				310					315					320
Leu	Pro	Cys	Leu	Gln	Val	Gly	Gln	Glu	Gln	Lys	His	Thr	Tyr	Leu	Pro
				325					330					335	
Leu	Glu	Val	Cys	Asn	Ile	Val	Ala	Gly	Gln	Arg	Cys	Ile	Lys	Lys	Leu
				340				345					350		
Thr	Asp	Asn	Gln	Thr	Ser	Thr	Met	Ile	Lys	Ala	Thr	Ala	Arg	Ser	Ala
		355					360					365			
Pro	Asp	Arg	Gln	Glu	Glu	Ile	Ser	Arg	Leu	Val	Arg	Ser	Ala	Asn	Tyr
		370				375					380				
Glu	Thr	Asp	Pro	Phe	Val	Gln	Glu	Phe	Gln	Phe	Lys	Val	Arg	Asp	Glu
	385				390					395					400
Met	Ala	His	Val	Thr	Gly	Arg	Val	Leu	Pro	Ala	Pro	Met	Leu	Gln	Tyr
				405					410					415	
Gly	Gly	Arg	Asn	Arg	Thr	Val	Ala	Thr	Pro	Ser	His	Gly	Val	Trp	Asp
			420					425					430		
Met	Arg	Gly	Lys	Gln	Phe	His	Thr	Gly	Val	Glu	Ile	Lys	Met	Trp	Ala
			435				440					445			
Ile	Ala	Cys	Phe	Ala	Thr	Gln	Arg	Gln	Cys	Arg	Glu	Glu	Ile	Leu	Lys
		450				455					460				
Gly	Phe	Thr	Asp	Gln	Leu	Arg	Lys	Ile	Ser	Lys	Asp	Ala	Gly	Met	Pro
	465				470					475					480
Ile	Gln	Gly	Gln	Pro	Cys	Phe	Cys	Lys	Tyr	Ala	Gln	Gly	Ala	Asp	Ser
				485					490					495	
Val	Glu	Pro	Met	Phe	Arg	His	Leu	Lys	Asn	Thr	Tyr	Ser	Gly	Leu	Gln
			500					505					510		
Leu	Ile	Ile	Val	Ile	Leu	Pro	Gly	Lys	Thr	Pro	Val	Tyr	Ala	Glu	Val
			515				520					525			
Lys	Arg	Val	Gly	Asp	Thr	Leu	Leu	Gly	Met	Ala	Thr	Gln	Cys	Val	Gln
		530				535					540				
Val	Lys	Asn	Val	Ile	Lys	Thr	Ser	Pro	Gln	Thr	Leu	Ser	Asn	Leu	Cys
				550						555					560
Leu	Lys	Ile	Asn	Val	Lys	Leu	Gly	Gly	Ile	Asn	Asn	Ile	Leu	Val	Pro
				565					570					575	
His	Gln	Arg	Pro	Ser	Val	Phe	Gln	Gln	Pro	Val	Ile	Phe	Leu	Gly	Ala
			580					585					590		
Asp	Val	Thr	His	Pro	Pro	Ala	Gly	Asp	Gly	Lys	Lys	Pro	Ser	Ile	Ala
			595				600					605			
Ala	Val	Val	Gly	Ser	Met	Asp	Ala	His	Pro	Ser	Arg	Tyr	Cys	Ala	Thr
		610				615					620				
Val	Arg	Val	Gln	Arg	Pro	Arg	Gln	Glu	Ile	Ile	Gln	Asp	Leu	Ala	Ser
	625				630					635					640
Met	Val	Arg	Glu	Leu	Leu	Ile	Gln	Phe	Tyr	Lys	Ser	Thr	Arg	Phe	Lys
				645					650					655	
Pro	Thr	Arg	Ile	Ile	Phe	Tyr	Arg	Asp	Gly	Val	Ser	Glu	Gly	Gln	Phe
			660					665					670		
Arg	Gln	Val	Leu	Tyr	Tyr	Glu	Leu	Leu	Ala	Ile	Arg	Glu	Ala	Cys	Ile
		675					680					685			

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Ser Leu Glu Lys Asp Tyr Gln Pro Gly Ile Thr Tyr Ile Val Val Gln
 690                695                700

Lys Arg His His Thr Arg Leu Phe Cys Ala Asp Arg Thr Glu Arg Val
 705                710                715                720

Gly Arg Ser Gly Asn Ile Pro Ala Gly Thr Thr Val Asp Thr Asp Ile
 725                730                735

Thr His Pro Tyr Glu Phe Asp Phe Tyr Leu Cys Ser His Ala Gly Ile
 740                745                750

Gln Gly Thr Ser Arg Pro Ser His Tyr His Val Leu Trp Asp Asp Asn
 755                760                765

Cys Phe Thr Ala Asp Glu Leu Gln Leu Leu Thr Tyr Gln Leu Cys His
 770                775                780

Thr Tyr Val Arg Cys Thr Arg Ser Val Ser Ile Pro Ala Pro Ala Tyr
 785                790                795                800

Tyr Ala His Leu Val Ala Phe Arg Ala Arg Tyr His Leu Val Asp Lys
 805                810                815

Glu His Asp Ser Ala Glu Gly Ser His Val Ser Gly Gln Ser Asn Gly
 820                825                830

Arg Asp Pro Gln Ala Leu Ala Lys Ala Val Gln Ile His Gln Asp Thr
 835                840                845

Leu Arg Thr Met Tyr Phe Ala
 850                855

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<210> SEQ ID NO 72
<211> LENGTH: 764
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: HILI, predicted protein sequence

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<400> SEQUENCE: 72

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Ile Ser Ser Gly Asp Ala Gly Ser Thr Phe Met Glu Arg Gly Val Lys
 1          5          10          15

Asn Lys Gln Asp Phe Met Asp Leu Ser Ile Cys Thr Arg Glu Lys Leu
 20          25          30

Ala His Val Arg Asn Cys Lys Thr Gly Ser Ser Gly Ile Pro Val Lys
 35          40          45

Leu Val Thr Asn Leu Phe Asn Leu Asp Phe Pro Gln Asp Trp Gln Leu
 50          55          60

Tyr Gln Tyr His Val Thr Tyr Ile Pro Asp Leu Ala Ser Arg Arg Leu
 65          70          75          80

Arg Ile Ala Leu Leu Tyr Ser His Ser Glu Leu Ser Asn Lys Ala Lys
 85          90          95

Ala Phe Asp Gly Ala Ile Leu Phe Leu Ser Gln Lys Leu Glu Glu Lys
 100         105         110

Val Thr Glu Leu Ser Ser Glu Thr Gln Arg Gly Glu Thr Ile Lys Met
 115         120         125

Thr Ile Thr Leu Lys Arg Glu Leu Pro Ser Ser Ser Pro Val Cys Ile
 130         135         140

Gln Val Phe Asn Ile Ile Phe Arg Lys Ile Leu Lys Lys Leu Ser Met
 145         150         155         160

Tyr Gln Ile Gly Arg Asn Phe Tyr Asn Pro Ser Glu Pro Met Glu Ile

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165								170				175			
Pro	Gln	His	Lys	Leu	Ser	Leu	Trp	Pro	Gly	Phe	Ala	Ile	Ser	Val	Ser
			180					185					190		
Tyr	Phe	Glu	Arg	Lys	Leu	Leu	Phe	Ser	Ala	Asp	Val	Ser	Tyr	Lys	Val
		195					200					205			
Leu	Arg	Asn	Glu	Thr	Val	Leu	Glu	Phe	Met	Thr	Ala	Leu	Cys	Gln	Arg
	210					215					220				
Thr	Gly	Leu	Ser	Cys	Phe	Thr	Gln	Thr	Cys	Glu	Lys	Gln	Leu	Ile	Gly
225					230					235					240
Leu	Ile	Val	Leu	Thr	Arg	Tyr	Asn	Asn	Arg	Thr	Tyr	Ser	Ile	Asp	Asp
				245					250					255	
Ile	Asp	Trp	Ser	Val	Lys	Pro	Thr	His	Thr	Phe	Gln	Lys	Arg	Asp	Gly
			260					265					270		
Thr	Glu	Ile	Thr	Tyr	Val	Asp	Tyr	Tyr	Lys	Gln	Gln	Tyr	Asp	Ile	Thr
		275					280					285			
Val	Ser	Asp	Leu	Asn	Gln	Pro	Met	Leu	Val	Ser	Leu	Leu	Lys	Lys	Lys
	290					295					300				
Arg	Asn	Asp	Asn	Ser	Glu	Ala	Gln	Leu	Ala	His	Leu	Ile	Pro	Glu	Leu
305					310					315					320
Cys	Phe	Leu	Thr	Gly	Leu	Thr	Asp	Gln	Ala	Thr	Ser	Asp	Phe	Gln	Leu
				325					330					335	
Met	Lys	Ala	Val	Ala	Glu	Lys	Thr	Arg	Leu	Ser	Pro	Ser	Gly	Arg	Gln
			340					345					350		
Gln	Arg	Leu	Ala	Arg	Leu	Val	Asp	Asn	Ile	Gln	Arg	Asn	Thr	Asn	Ala
		355					360					365			
Arg	Phe	Glu	Leu	Glu	Thr	Trp	Gly	Leu	His	Phe	Gly	Ser	Gln	Ile	Ser
						375					380				
Leu	Thr	Gly	Arg	Ile	Val	Pro	Ser	Glu	Lys	Ile	Leu	Met	Gln	Asp	His
385					390					395					400
Ile	Cys	Gln	Pro	Val	Ser	Ala	Ala	Asp	Trp	Ser	Lys	Asp	Ile	Arg	Thr
				405					410					415	
Cys	Lys	Ile	Leu	Asn	Ala	Gln	Ser	Leu	Asn	Thr	Trp	Leu	Ile	Leu	Cys
			420					425				430			
Ser	Asp	Arg	Thr	Glu	Tyr	Val	Ala	Glu	Ser	Phe	Leu	Asn	Cys	Leu	Arg
		435					440					445			
Arg	Val	Ala	Gly	Ser	Met	Gly	Phe	Asn	Val	Met	Cys	Ile	Leu	Pro	Ser
						455					460				
Asn	Gln	Lys	Thr	Tyr	Tyr	Asp	Ser	Ile	Lys	Lys	Tyr	Leu	Ser	Ser	Asp
465					470					475					480
Cys	Pro	Val	Pro	Ser	Gln	Cys	Val	Leu	Ala	Arg	Thr	Leu	Asn	Lys	Gln
				485					490					495	
Gly	Met	Met	Met	Ser	Ile	Ala	Thr	Lys	Ile	Ala	Met	Gln	Met	Thr	Cys
			500					505				510			
Lys	Leu	Gly	Gly	Glu	Leu	Trp	Ala	Val	Glu	Ile	Pro	Leu	Lys	Ser	Leu
		515					520					525			
Met	Val	Val	Gly	Ile	Asp	Val	Cys	Lys	Asp	Ala	Leu	Ser	Lys	Asp	Val
						535					540				
Met	Val	Val	Gly	Cys	Val	Ala	Ser	Val	Asn	Pro	Arg	Ile	Thr	Arg	Trp
545					550					555					560
Phe	Ser	Arg	Cys	Ile	Leu	Gln	Arg	Thr	Met	Thr	Asp	Val	Ala	Asp	Cys
				565					570					575	

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Leu Lys Val Phe Met Thr Gly Ala Leu Asn Lys Trp Tyr Lys Tyr Asn
 580 585 590
 His Asp Leu Pro Ala Arg Ile Ile Val Tyr Arg Ala Gly Val Gly Asp
 595 600 605
 Gly Gln Leu Lys Thr Leu Ile Glu Tyr Glu Val Pro Gln Leu Leu Ser
 610 615 620
 Ser Val Ala Glu Ser Ser Ser Asn Thr Ser Ser Arg Leu Ser Val Ile
 625 630 635 640
 Val Val Arg Lys Lys Cys Met Pro Arg Phe Phe Thr Glu Met Asn Arg
 645 650 655
 Thr Val Gln Asn Pro Pro Leu Gly Thr Val Val Asp Ser Glu Ala Thr
 660 665 670
 Arg Asn Glu Trp Gln Tyr Asp Phe Tyr Leu Ile Ser Gln Val Ala Cys
 675 680 685
 Arg Gly Thr Val Ser Pro Thr Tyr Tyr Asn Val Ile Tyr Asp Asp Asn
 690 695 700
 Gly Leu Lys Pro Asp His Met Gln Arg Leu Thr Phe Lys Leu Cys His
 705 710 715 720
 Leu Tyr Tyr Asn Trp Pro Gly Ile Val Ser Val Pro Ala Pro Cys Gln
 725 730 735
 Tyr Ala His Lys Leu Thr Phe Leu Val Ala Gln Ser Ile His Lys Glu
 740 745 750
 Pro Ser Leu Glu Leu Ala Asn His Leu Phe Tyr Leu
 755 760

<210> SEQ ID NO 73
 <211> LENGTH: 861
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: HIWI, predicted protein sequence

<400> SEQUENCE: 73

Met Thr Gly Arg Ala Arg Ala Arg Ala Arg Gly Arg Ala Arg Gly Gln
 1 5 10 15
 Glu Thr Ala Gln Leu Val Gly Ser Thr Ala Ser Gln Gln Pro Gly Tyr
 20 25 30
 Ile Gln Pro Arg Pro Gln Pro Pro Pro Ala Glu Gly Glu Leu Phe Gly
 35 40 45
 Arg Gly Arg Gln Arg Gly Thr Ala Gly Gly Thr Ala Lys Ser Gln Gly
 50 55 60
 Leu Gln Ile Ser Ala Gly Phe Gln Glu Leu Ser Leu Ala Glu Arg Gly
 65 70 75 80
 Gly Arg Arg Arg Asp Phe His Asp Leu Gly Val Asn Thr Arg Gln Asn
 85 90 95
 Leu Asp His Val Lys Glu Ser Lys Thr Gly Ser Ser Gly Ile Ile Val
 100 105 110
 Arg Leu Ser Thr Asn His Phe Arg Leu Thr Ser Arg Pro Gln Trp Ala
 115 120 125
 Leu Tyr Gln Tyr His Ile Asp Tyr Asn Pro Leu Met Glu Ala Arg Arg
 130 135 140
 Leu Arg Ser Ala Leu Leu Phe Gln His Glu Asp Leu Ile Gly Lys Cys

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145	150	155	160
His Ala Phe Asp Gly Thr Ile Leu Phe Leu Pro Lys Arg Leu Gln Gln	165	170	175
Lys Val Thr Glu Val Phe Ser Lys Thr Arg Asn Gly Glu Asp Val Arg	180	185	190
Ile Thr Ile Thr Leu Thr Asn Glu Leu Pro Pro Thr Ser Pro Thr Cys	195	200	205
Leu Gln Phe Tyr Asn Ile Ile Phe Arg Arg Leu Leu Lys Ile Met Asn	210	215	220
Leu Gln Gln Ile Gly Arg Asn Tyr Tyr Asn Pro Asn Asp Pro Ile Asp	225	230	235
Ile Pro Ser His Arg Leu Val Ile Trp Pro Gly Phe Thr Thr Ser Ile	245	250	255
Leu Gln Tyr Glu Asn Ser Ile Met Leu Cys Thr Asp Val Ser His Lys	260	265	270
Val Leu Arg Ser Glu Thr Val Leu Asp Phe Met Phe Asn Phe Tyr His	275	280	285
Gln Thr Glu Glu His Lys Phe Gln Glu Gln Val Ser Lys Glu Leu Ile	290	295	300
Gly Leu Val Val Leu Thr Lys Tyr Asn Asn Lys Thr Tyr Arg Val Asp	305	310	315
Asp Ile Asp Trp Asp Gln Asn Pro Lys Ser Thr Phe Lys Lys Ala Asp	325	330	335
Gly Ser Glu Val Ser Phe Leu Glu Tyr Tyr Arg Lys Gln Tyr Asn Gln	340	345	350
Glu Ile Thr Asp Leu Lys Gln Pro Val Leu Val Ser Gln Pro Lys Arg	355	360	365
Arg Arg Gly Pro Gly Gly Thr Leu Pro Gly Pro Ala Met Leu Ile Pro	370	375	380
Glu Leu Cys Tyr Leu Thr Gly Leu Thr Asp Lys Met Arg Asn Asp Phe	385	390	395
Asn Val Met Lys Asp Leu Ala Val His Thr Arg Leu Thr Pro Glu Gln	405	410	415
Arg Gln Arg Glu Val Gly Arg Leu Ile Asp Tyr Ile His Lys Asn Asp	420	425	430
Asn Val Gln Arg Glu Leu Arg Asp Trp Gly Leu Ser Phe Asp Ser Asn	435	440	445
Leu Leu Ser Phe Ser Gly Arg Ile Leu Gln Thr Glu Lys Ile His Gln	450	455	460
Gly Gly Lys Thr Phe Asp Tyr Asn Pro Gln Phe Ala Asp Trp Ser Lys	465	470	475
Glu Thr Arg Gly Ala Pro Leu Ile Ser Val Lys Pro Leu Asp Asn Trp	485	490	495
Leu Leu Ile Tyr Thr Arg Arg Asn Tyr Glu Ala Ala Asn Ser Leu Ile	500	505	510
Gln Asn Leu Phe Lys Val Thr Pro Ala Met Gly Met Gln Met Arg Lys	515	520	525
Ala Ile Met Ile Glu Val Asp Asp Arg Thr Glu Ala Tyr Leu Arg Val	530	535	540
Leu Gln Gln Lys Val Thr Ala Asp Thr Gln Ile Val Val Cys Leu Leu	545	550	555
			560

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Ser Ser Asn Arg Lys Asp Lys Tyr Asp Ala Ile Lys Lys Tyr Leu Cys
 565 570 575
 Thr Asp Cys Pro Thr Pro Ser Gln Cys Val Val Ala Arg Thr Leu Gly
 580 585 590
 Lys Gln Gln Thr Val Met Ala Ile Ala Thr Lys Ile Ala Leu Gln Met
 595 600 605
 Asn Cys Lys Met Gly Gly Glu Leu Trp Arg Val Asp Ile Pro Leu Lys
 610 615 620
 Leu Val Met Ile Val Gly Ile Asp Cys Tyr His Asp Met Thr Ala Gly
 625 630 635 640
 Arg Arg Ser Ile Ala Gly Phe Val Ala Ser Ile Asn Glu Gly Met Thr
 645 650 655
 Arg Trp Phe Ser Arg Cys Ile Phe Gln Asp Arg Gly Gln Glu Leu Val
 660 665 670
 Asp Gly Leu Lys Val Cys Leu Gln Ala Ala Leu Arg Ala Trp Asn Ser
 675 680 685
 Cys Asn Glu Tyr Met Pro Ser Arg Ile Ile Val Tyr Arg Asp Gly Val
 690 695 700
 Gly Asp Gly Gln Leu Lys Thr Leu Val Asn Tyr Glu Val Pro Gln Phe
 705 710 715 720
 Leu Asp Cys Leu Lys Ser Ile Gly Arg Gly Tyr Asn Pro Arg Leu Thr
 725 730 735
 Val Ile Val Val Lys Lys Arg Val Asn Thr Arg Phe Phe Ala Gln Ser
 740 745 750
 Gly Gly Arg Leu Gln Asn Pro Leu Pro Gly Thr Val Ile Asp Val Glu
 755 760 765
 Val Thr Arg Pro Glu Trp Tyr Asp Phe Phe Ile Val Ser Gln Ala Val
 770 775 780
 Arg Ser Gly Ser Val Ser Pro Thr His Tyr Asn Val Ile Tyr Asp Asn
 785 790 795 800
 Ser Gly Leu Lys Pro Asp His Ile Gln Arg Leu Thr Tyr Lys Leu Cys
 805 810 815
 His Ile Tyr Tyr Asn Trp Pro Gly Val Ile Arg Val Pro Ala Pro Cys
 820 825 830
 Gln Tyr Ala His Lys Leu Ala Phe Leu Val Gly Gln Ser Ile His Arg
 835 840 845
 Glu Pro Asn Leu Ser Leu Ser Asn Arg Leu Tyr Tyr Leu
 850 855 860

<210> SEQ ID NO 74
 <211> LENGTH: 2571
 <212> TYPE: DNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: eIF2C1, cDNA sequence of predicted ORF
 <400> SEQUENCE: 74

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 ttccaggcac ctgcgccggc tggcattggc actgtgggga aaccaatcaa gctcctggcc 120
 aattactttg aggtggacat ccctaagatc gacgtgtacc actacgaggt ggacatcaag 180
 ccggataagt gtccccgtag agtcaaccgg gaagtgggtg aatacatggt ccagcatttc 240

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aagcctcaga	tctttggtga	tcgcaagcct	gtgtatgatg	gaaagaagaa	catttacact	300
gtcacagcac	tgccatttgg	caacgaacgg	gtcgactttg	aggtgacaat	ccttggggaa	360
gggaaggatc	gaatctttaa	ggtctccatc	aagtggctag	ccattgtgag	ctggcgaatg	420
ctgcatgagg	ccttggctcag	cggccagatc	cctgttcctt	tggagtctgt	gcaagccctg	480
gatgtggcca	tgaggcacct	ggcatccatg	aggtaacccc	ctgtgggccc	ctccttcttc	540
tcaccgcctg	agggtacta	ccaccgctg	gggggtgggc	gcgaggtctg	gttcggcttt	600
caccagtctg	tcgcgcctgc	catgtggaag	atgatgctca	acattgatgt	ctcagccact	660
gccttttata	aggcacagcc	agtgattgag	ttcatgtgtg	agggtctgga	catcaggaac	720
atagatgagc	agcccaagcc	cctcacggac	tctcagcgcg	tctgcttcac	caaggagatc	780
aagggcctga	agggtgaagt	cacccactgt	ggacagatga	agaggaagta	ccgcgtgtgt	840
aatgttacc	gtcgcctgc	tagccatcag	acattcccct	tacagctgga	gagtggacag	900
actgtggagt	gcacagtggc	acagtatttc	aagcagaaat	ataacctca	gctcaagtat	960
ccccatctgc	cctgcctaca	agttggccag	gaacaaaagc	atactacct	tcccctagag	1020
gtctgtaaca	ttgtggctgg	gcagcgctgt	attaaaaagc	tgaccgacaa	ccagacctcg	1080
acatgataa	aggccacagc	tagatccgct	ccagacagac	aggaggagat	cagtcgcctg	1140
atgaagaatg	ccagctacaa	cttagatccc	tacatccagg	aatttgggat	caaagtgaag	1200
gatgacatga	cggaggtgac	agggcgagtg	ctgccggcgc	ccatcttgca	gtaccggcggc	1260
cggaaaccggg	ccattgccac	acccaatcag	ggtgtctggg	acatgcgggg	gaaacagttc	1320
tacaatggga	ttgagatcaa	agtctgggcc	atcgctctgt	tcgcacccca	aaaacagtgt	1380
cgagaagagg	tgctcaagaa	cttcacagac	cagctgcgga	agatttccaa	ggatgcgggg	1440
atgcctatcc	agggtcaacc	ttgtttctgc	aaatatgcac	agggggcaga	cagcgtggag	1500
cctatgttcc	ggcatctcaa	gaacacctac	tcagggtctg	agctcattat	tgtcatcctg	1560
ccaggaaga	cgcgggtgta	tgctgaggtg	aaacgtgtcg	gagatacact	cttgggaatg	1620
gctacgcagt	gtgtgcaggt	gaagaacgtg	gtcaagacct	cacctcagac	tctgtccaac	1680
ctctgcctca	agatcaatgt	caaacttggt	ggcattaaca	acatcctagt	cccacaccag	1740
cgctctgccg	tttttcaaca	gccagtata	ttctggggag	cagatgttac	acacccccca	1800
gcaggggatg	ggaaaaaacc	ttctatcaca	gcagtggtag	gcagtatgga	tgccaccccc	1860
agccgatact	gtgctactgt	gcgggtacag	cgaccacggc	aagagatcat	tgaagacttg	1920
tctacatgg	tgctgagct	cctcatccaa	ttctacaagt	ccaccgctt	caagcctacc	1980
cgcatcatct	tctaccgaga	tgggtgcct	gaaggccagc	tacccagat	actccactat	2040
gagctactgg	ccattcgtga	tgccctgcac	aaactggaaa	aggactacca	gcctgggatc	2100
acttatattg	tggtgcagaa	acgccatcac	acccgccttt	tctgtgctga	caagaatgag	2160
cgaattggga	agagtggtaa	catcccagct	gggaccacag	tggacaccaa	catcaccac	2220
ccatttgagt	ttgacttcta	tctgtgcagc	cacgcaggca	tccagggcac	cagccgacca	2280
tcccattact	atgttctttg	ggatgacaa	cgtttcacag	cagatgagct	ccagatcctg	2340
acgtaccagc	tgtgccacac	ttacgtacga	tgcacacgct	ctgtctctat	cccagcacct	2400
gcctactatg	ccgccttggt	ggctttccgg	gcacgatacc	acctgggtgga	caaggagcat	2460
gacagtggag	aggggagcca	catatcgggg	cagagcaatg	ggcgggaccc	ccaggccctg	2520

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gccccagccg tgcaggttca ccaggatact ctgcgcacca tgtacttcgc t 2571

<210> SEQ ID NO 75
 <211> LENGTH: 2580
 <212> TYPE: DNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: eIF2C2, cDNA sequence of predicted ORF

<400> SEQUENCE: 75

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ggatatgcct tcaagcctcc acctagacct gactttggga cctccgggag aacaatcaaa	120
ttacaggcca atttcttcga aatggacatc cccaaaattg acatctatca ttatgaattg	180
gatatcaagc cagagaagtg cccgaggaga gttaacaggg aaatcgtgga acacatggtc	240
cagcacttta aaacacagat ctttggggat cggaagcccg tgtttgacgg caggaagaat	300
ctatacacag ccattgcccct tccgattggg agggacaagg tggagctgga ggtcacgctg	360
ccaggagaag gcaaggatcg catcttcaag gtgtccatca agtgggtgtc ctgcgtgagc	420
ttgcaggcgt tacacgatgc actttcaggg cggctgcccc gcgtcccttt tgagacgatc	480
caggcccttg acgtggctcat gaggcacttg ccatccatga ggtacacccc cgtgggcccgc	540
tcctttttca ccgcgtccga aggctgctct aacctcttg gcgggggccc agaagtgttg	600
tttggtctcc atcagtcctg ccggccttct ctctggaaaa tgatgctgaa tattgatgtg	660
tcagcaacag cgtttttcaa ggcacagcca gtaatcgagt ttgtttgtga agttttggat	720
tttaaaagta ttgaagaaca acaaaaacct ctgacagatt cccaaagggt aaagtattacc	780
aaagaaatta aaggtctaaa ggtggagata acgcactgtg ggcagatgaa gaggaagtac	840
cgtgtctgca atgtgacctg gcggccccc agtcacccaa cattcccgtg gcagcaggag	900
agcgggcaga cgggtggagtg cacggtggcc cagtatttca aggacaggca caagttgggt	960
ctgcgtacc cccacctccc atgttttcaa gtcggacagg agcagaaaca cacctacctt	1020
cccttgagg tctgtaacat tgtggcagga caaagatgta ttaaaaaatt aacggacaat	1080
cagacctcaa ccatgatcag agcaactgct aggtcggcgc ccgacggca agaagagatt	1140
agcaaatgta tgcaagtgc aagtttcaac acagatccat acgtccgtga atttggaatc	1200
atggtcaaa atgagatgac agacgtgact gggcgggtgc tgcagccgc ctccatctc	1260
tacgggggca ggaataaagc tattgcgacc cctgtccagg gcgtctggga catgcggaac	1320
aagcagttcc acacgggcat cgagatcaag gtgtgggcca ttgcgtgctt cgcctccag	1380
cgccagtgca cggaagtcca tctgaagtcc ttcacagagc agtcagaaa gatctcgaga	1440
gacgtggca tgcccatcca ggccagccg tgcttctgca aatacgcgc gggggcggac	1500
agcgtggagc ccattgtccg gcacctgaag aacacgtatg cgggcctgca gctggtggtg	1560
gtcatctgc ccggcaagac gccctgtac gccgaggtca agcgcgtggg agacacggtg	1620
ctggggatgg ccacgcagtg cgtgcagatg aagaactgc agaggaccac gccacagacc	1680
ctgtccaacc ttgctgaa gatcaacgtc aagctgggag gcgtgaacaa catcctgctg	1740
ccccaggga gccgcgggt gttccagcag ccgctcatct ttctgggagc agacgtcact	1800
caccccccg ccggggatgg gaagaagccc tccattgccg ccgtgggtggg cagcatggac	1860

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gcccacccca atcgctactg cgccaccgtg cgcgtgcagc agcaccggca ggagatcata 1920
caagacctgg ccgccatggt ccgcgagctc ctcatccagt tctacaagtc cagcgcttc 1980
aagccccccc gcatcatctt ctaccgcgac ggtgtctctg aaggccagtt ccagcagggt 2040
ctccaccacg agttgtctgg catccgtgag gctgtatca agctagaaaa agactaccag 2100
ccccggatca ccttcacgtt ggtgcagaag aggcaccaca cccggctctt ctgcactgac 2160
aagaacgagc gggttgggaa aagtggaaac attccagcag gcacgactgt ggacacgaaa 2220
atcacccacc ccaccgagtt cgacttctac ctgtgtagtc acgtggcat ccaggggaca 2280
agcaggcctt cgcactatca cgtcctctgg gacgacaatc gtttctctc tgatgagctg 2340
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ccagcgccag catactacgc tcacctgggt gccttcgggg ccaggtagca cctggtggat 2460
aaggaacatg acagtgtgta aggaagccat acctctgggc agagtaacgg gcgagaccac 2520
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<210> SEQ ID NO 76

<211> LENGTH: 2772

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: eIF2C3, cDNA sequence of predicted ORF

<400> SEQUENCE: 76

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gcggcgccgc cggcgccccc cggagcggga ggcgcgcggg accggggcga ggcggccccc 180
gccgcgcgca tggaggcgct gggacccgga cctccggcta gcctgtttca gccacctcgt 240
cgtcctggcc ttggaactgt tggaaaacca attcgactgt tagccaatca ttttcagggt 300
cagattccta aaatagatgt gtatcactat gatgtggata ttaagcctga aaaacggcct 360
cgtagagtca acaggggaggt agtagataca atggtgcggc acttcaagat gcaaatatct 420
ggtgatcgcc agcctgggta tgatggcaaa agaaacatgt acacagcaca tccactacca 480
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gctgggcact tgaatgaagt ccagatgac tcagtacaag cacttgatgt taccacaaga 660
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ccagccagtc atcaaaactt tcctttgcag ctagaaaacg gtcaagctat ggaatgtaca 1080
gtagctcaat attttaagca aaagtatagt ctgcaactga aatcccccca tcttcctgt 1140
ctccaagtgg gacaagaaca aaagcataca tacttgccac tcgaggtctg taatatagtg 1200
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acagcaagat ctgctcctga cagacaggaa gagatcagta gactggtgaa gagcaacagt 1320
atgggtgggtg gacctgatcc ataccttaaa gaatttggtg ttgtgtgcca caatgaaatg 1380
acagagctca caggcagggt acttccagca ccaatgctgc aatatggagg ccggaataaa 1440
acagtagcca caccacaacca ggggtgtctgg gacatgcgag gaaagcagtt ttatgctggc 1500
attgaaatta aagtttgggc agttgcttgt ttgacacctc agaacaatg tagggaagat 1560
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<210> SEQ ID NO 77
<211> LENGTH: 2568
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C4, cDNA sequence of predicted ORF

<400> SEQUENCE: 77

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gcaggaccgc ctggggccca gcccctactc atgggtgcca gaagacctgg ctatggcacc 60
atgggcaaac ccattaaact gctggctaac tgttttcaag ttgaaatccc aaagattgat 120
gtctacctct atgaggtaga tattaacca gacaagtgtc ctaggagagt gaacaggag 180
gtggttgact caatggttca gcattttaaa gtaactatat ttggagaccg tagaccagtt 240
tatgatggaa aaagaagtct ttacaccgcc aatccacttc ctgtggcaac tacagggtga 300
gatttagacg ttactttacc tggggaagggt ggaaaagatc gaccttcaa ggtgtcaatc 360

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aaatttgtct	ctcgggtgag	ttggcaccta	ctgcatgaag	tactgacagg	acggaccttg	420
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gtgtctacgac	atctgccctc	catgaaatc	acacctgtgg	ggcgttcatt	ttctccgct	540
ccagaaggat	atgaccaccc	tctgggaggg	ggcaggaag	tgtggtttg	attccatcag	600
tctgttcggc	ctgccatgtg	gaaaatgatg	cttaatatcg	atgtttctgc	actgccttc	660
tacaaagcac	aacctgtaat	tcagtccatg	tgtgaagtcc	ttgatattca	taatattgat	720
gagcaaccaa	gacctctgac	tgattctcat	cgggtaaaat	tcaccaaaga	gataaaaggt	780
ttgaagggtg	aagtgactca	ttgtggaaca	atgagacgga	aataccgtgt	ttgtaattga	840
acaaggaggc	ctgccagtca	tcaaaccttt	cctttacagt	tagaaaacgg	ccaaactgtg	900
gagagaacag	tagcgcagta	tttcagagaa	aagtatactc	ttcagctgaa	gtacccgcac	960
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aatattgtgg	cagggcaacg	atgtatcaag	aagctaacag	acaatcagac	ttccactatg	1080
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gatgaaaga	agccttctat	tgctgctgtt	gtaggtagta	tggatgcaca	ccaagcaga	1860
tactgtgcca	cagtaagagt	tcagagaccc	cgacaggaga	tcattccagga	cttggcctcc	1920
atggtccggg	aacttcttat	tcaattttat	aagtcaactc	ggttcaagcc	tactcgtatc	1980
atcttttata	gggatggtgt	ttcagagggg	cagtttaggc	aggtattata	ttatgaacta	2040
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ggaagaagtg	gcaatatccc	agctggaaca	acagttgata	cagacattac	acacccatat	2220
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gctgaaggaa	gtcacgtttc	aggacaaagc	aatgggcgag	atccacaagc	tcttgccaag	2520
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<211> LENGTH: 2292
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HILI1, cDNA sequence of predicted ORF

<400> SEQUENCE: 78

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ggttccagtg gaatacctgt gaaactgggt acaaacctct ttaacttaga ttttcccca 180
gactggcagc tataccagta ccattgtgaca tatattccag atttagcatc tagaaggctg 240
agaattgctt tactttatag tcatagttaa ctttccaaca aagcaaaagc attcgacggc 300
gccatccttt ttctgtcaca aaagctagaa gaaaagggtc cagagttgtc aagtgaact 360
caaagagggt agactataaa gatgactatc accctgaaga gggagctgcc atcaagttct 420
cccggtgtgc tccaggtctt caatatcatc ttcagaaaga tcctcaaaa gttgtccatg 480
taccaaaattg gacggaactt ctataatcct tcagagccaa tggaaattcc ccagcacaaa 540
ttatcccttt ggctggggtt tgccatttct gtgtcatatt ttgaaaggaa gctcctgttt 600
agtgtgatg tgagttacaa agtcctccgg aatgagacgg ttctggaatt catgactgct 660
ctctgtcaaa gaactggctt gtcctgttcc acccagacgt gtgagaagca gctaataagg 720
ctcattgttc ttacaagata caataacaga acctactcca ttgatgacat tgactgggtc 780
gtgaagccca cacacacctt tcagaagcgg gatggcaccg agatcaccta tgtggattac 840
tacaagcagc agtatgatat tactgtatcg gacctgaatc agcccatgct tgttagtctg 900
ttaaagaaga agagaaatga caacagttag gctcagctcg cccacctgat acctgagctc 960
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gtggtcagga agaagtgcac gccacgatc tttaccgaaa tgaaccgcac tgtacagaac 1980
ccccacttg gcactgttgt ggattcagaa gcaacacgta acgaatggca gtatgacttt 2040

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tatctgatca gccaggtggc ctgccgggga actgttagtc ctacctacta taatgtcatc	2100
tatgatgaca acggcttgaa gcccgaccat atgcagagac ttacattcaa attgtgccac	2160
ctgtactaca actggccggg catagtcagt gtcccagcac catgtcagta tgctcacaag	2220
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ctcttctacc tg	2292

<210> SEQ ID NO 79
 <211> LENGTH: 2583
 <212> TYPE: DNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HIWI, cDNA sequence of predicted ORF

<400> SEQUENCE: 79

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ccagcagagg gggaattatt tggccgtgga cggcagagag gaacagcagg aggaacagcc	180
aagtcacaag gactccagat atctgctgga tttcaggagt tatcgttagc agagagagga	240
ggtcgtcgta gagattttca tgatcttggt gtgaatacaa ggcagaacct agaccatgtt	300
aaagaatcaa aaacaggttc ttcaggcatt atagtaaggt taagcactaa ccatttccgg	360
ctgacatccc gtcccagtg ggccttatat cagtatcaca ttgactataa cccactgatg	420
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catgcttttg atggaacgat attattttta cctaaaagac tacagcaaaa ggttactgaa	540
gttttttagta agaccggaa tggagaggat gtgaggataa cgatcacttt aacaaatgaa	600
cttcaccta catcaccaac ttgtttgcag ttctataata ttattttcag gaggttttg	660
aaaatcatga atttgaaca aattggacga aattattata acccaaata cccaattgat	720
attccaagtc acaggttggt gatttggcct ggcttcacta cttccatcct tcagtatgaa	780
aacagcatca tgctctgcac tgacgttagc cataaagtec ttcgaagtga gactgttttg	840
gattttcatgt tcaactttta tcatcagaca gaagaacata aatttcaaga acaagtttcc	900
aaagaactaa taggtttagt tgttcttacc aagtataaca ataagacata cagagtggat	960
gatattgact gggaccagaa tcccagagc acctttaaga aagccgacgg ctctgaagtc	1020
agcttcttag aatactacag gaagcaatac aaccaagaga tcaccgactt gaagcagcct	1080
gtcttggtca gccagcccaa gagaaggcgg ggccctgggg ggacactgcc agggcctgcc	1140
atgetcattc ctgagctctg ctatcttaca ggtctaactg ataaaatgcg taatgatattt	1200
aacgtgatga aagacttagc cgttcataca agactaactc cagagcaaag gcagcgtgaa	1260
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aagattcacc aagggtgaaa aacatttgat tacaatccac aatttgcaga ttggtccaaa	1440
gaaacaagag gtgcaccatt aattagtgtt aagccactag ataactggct gttgatctat	1500
acgcgaagaa attatgaagc agccaattca ttgatacaaa atctatttaa agttacacca	1560
gccatgggca tgcaaatgag aaaagcaata atgattgaag tggatgacag aactgaagcc	1620

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agcggcctga agccagacca catacagcgc ttgacctaca agctgtgcca catctattac	2460
aactggccag gtgtcattcg tgttcctgct ccttgccagt acgcccacaa gctggctttt	2520
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ctc	2583

<210> SEQ ID NO 80
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human
 gene
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: eIF2C1, primer (5'-3')

<400> SEQUENCE: 80

gaggtctgta acattgtggc	20
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<210> SEQ ID NO 81
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human
 gene
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: eIF2C1, primer (5'-3')

<400> SEQUENCE: 81

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<210> SEQ ID NO 82
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human
 gene
 <220> FEATURE:
 <221> NAME/KEY: misc_feature

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<223> OTHER INFORMATION: eIF2C1, primer (5'-3')

<400> SEQUENCE: 82

gaggtctgta acattgtggc 20

<210> SEQ ID NO 83

<211> LENGTH: 24

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: eIF2C1, primer (5'-3')

<400> SEQUENCE: 83

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<210> SEQ ID NO 84

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: eIF2C1, primer (5'-3')

<400> SEQUENCE: 84

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<210> SEQ ID NO 85

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: eIF2C1, primer (5'-3')

<400> SEQUENCE: 85

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<210> SEQ ID NO 86

<211> LENGTH: 18

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: eIF2C1, primer (5'-3')

<400> SEQUENCE: 86

ccacaccagc gctctgcc 18

<210> SEQ ID NO 87

<211> LENGTH: 18

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

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<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C1, primer (5'-3')

<400> SEQUENCE: 87

ctcacgcacc atgtagga 18

<210> SEQ ID NO 88
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C2, primer (5'-3')

<400> SEQUENCE: 88

gaggtctgta acattgtggc 20

<210> SEQ ID NO 89
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C2, primer (5'-3')

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cggtagaaga tgatgcgggt 20

<210> SEQ ID NO 90
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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C2, primer (5'-3')

<400> SEQUENCE: 90

atcctgctgc cccaaggg 18

<210> SEQ ID NO 91
<211> LENGTH: 18
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
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<223> OTHER INFORMATION: eIF2C2, primer (5'-3')

<400> SEQUENCE: 91

gatctcctgc cggtgctg 18

<210> SEQ ID NO 92

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<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C2, primer (5'-3')

<400> SEQUENCE: 92

gaggtctgta acattgtggc 20

<210> SEQ ID NO 93
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
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<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C2, primer (5'-3')

<400> SEQUENCE: 93

cggtagaaga tgatgcgggt 20

<210> SEQ ID NO 94
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C2, primer (5'-3')

<400> SEQUENCE: 94

gaggtctgta acattgtggc 20

<210> SEQ ID NO 95
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C2, primer (5'-3')

<400> SEQUENCE: 95

gatctcctgc cggtgctg 18

<210> SEQ ID NO 96
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C3, primer (5'-3')

<400> SEQUENCE: 96

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agagcaacag tatggtgggt ggac 24

<210> SEQ ID NO 97
<211> LENGTH: 18
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human
gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C3, primer (5'-3')

<400> SEQUENCE: 97

tggatgtgtg atggtact 18

<210> SEQ ID NO 98
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human
gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C3, primer (5'-3')

<400> SEQUENCE: 98

cctctacagt caagaggt 18

<210> SEQ ID NO 99
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<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human
gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C3, primer (5'-3')

<400> SEQUENCE: 99

tggatgtgtg atggtact 18

<210> SEQ ID NO 100
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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human
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<223> OTHER INFORMATION: eIF2C3, primer (5'-3')

<400> SEQUENCE: 100

cacttgaatg aagtccca 18

<210> SEQ ID NO 101
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<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human
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<220> FEATURE:
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<223> OTHER INFORMATION: eIF2C3, primer (5'-3')

<400> SEQUENCE: 101

tcctggatga cctcttgact gtag 24

<210> SEQ ID NO 102

<211> LENGTH: 24

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: eIF2C3, primer (5'-3')

<400> SEQUENCE: 102

agagcaacag tatggtgggt ggac 24

<210> SEQ ID NO 103

<211> LENGTH: 24

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: eIF2C3, primer (5'-3')

<400> SEQUENCE: 103

tcctggatga cctcttgact gtag 24

<210> SEQ ID NO 104

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: eIF2C4, primer (5'-3')

<400> SEQUENCE: 104

tcggcatct caagaacaca tattct 26

<210> SEQ ID NO 105

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: eIF2C4, primer (5'-3')

<400> SEQUENCE: 105

gaactcatat ggggtgtgtgtg 26

<210> SEQ ID NO 106

<211> LENGTH: 18

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

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<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C4, primer (5'-3')

<400> SEQUENCE: 106

atccaggact tggcctcc 18

<210> SEQ ID NO 107
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
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<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
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<223> OTHER INFORMATION: eIF2C4, primer (5'-3')

<400> SEQUENCE: 107

gaactcatat ggggtgtgtaa tgtctg 26

<210> SEQ ID NO 108
<211> LENGTH: 18
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HILI, primer (5'-3')

<400> SEQUENCE: 108

cagcacaaat tatccctt 18

<210> SEQ ID NO 109
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<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HILI, primer (5'-3')

<400> SEQUENCE: 109

cggcctgaag gactgagacg tgt 23

<210> SEQ ID NO 110
<211> LENGTH: 18
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
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<220> FEATURE:
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<223> OTHER INFORMATION: HILI, primer (5'-3')

<400> SEQUENCE: 110

cagcacaaat tatccctt 18

<210> SEQ ID NO 111

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<211> LENGTH: 18
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<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HILI, primer (5'-3')

<400> SEQUENCE: 111

gtgtgtgggc ttactga 18

<210> SEQ ID NO 112
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
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<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HILI, primer (5'-3')

<400> SEQUENCE: 112

tctctgtcaa agaactggct tgcct 26

<210> SEQ ID NO 113
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HILI, primer (5'-3')

<400> SEQUENCE: 113

ctgtacagtg cggttcat 18

<210> SEQ ID NO 114
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HILI, primer (5'-3')

<400> SEQUENCE: 114

tctctgtcaa agaactggct tgcct 26

<210> SEQ ID NO 115
<211> LENGTH: 23
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
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<223> OTHER INFORMATION: HILI, primer (5'-3')

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<400> SEQUENCE: 115

cggcctgaag gactgagacg tgt

23

1. Purified human RISC having a molecular weight of from up to about 150-160 kDa.

2. The RISC of claim 1 comprising at least one member of the Argonaute family of proteins.

3. The RISC of claim 1 containing eIF2C1 and/or eIFC2 and optionally at least one of eIFC3, eIFC4, HILI and HIWI.

4. The RISC of claim 1, further containing an RNA component, particularly a single-stranded RNA molecule.

5. The RISC of claim 4, wherein the single-stranded RNA molecule has a length from 14-50 nucleotides wherein at least the 14-20 5' most nucleotides are substantially complementary to a target transcript.

6. The RISC of claim 4, wherein said RNA molecule has a length from 15-29 nucleotides.

7. The RISC of claim 4, wherein said RNA molecule has a free 5' hydroxyl moiety or a moiety selected from phosphate groups or analogues thereof.

8. The RISC of claim 7, wherein said RNA molecule has a 5'-moiety selected from 5'-monophosphate ((HO)2(O)P—O-5'), 5'-diphosphate ((HO)2(O)P—O—P(HO)(O)—O-5'), 5'-triphosphate ((HO)2(O)P—O—(HO)(O)P—O—P(HO)(O)—O-5'), 5'-guanosine cap (7-methylated or non-methylated) (7m-G-0-5'-(HO)(O)P—O—(HO)(O)P—O—P(HO)(O)—O-5'), 5'-adenosine cap (Appp), and any modified or unmodified nucleotide cap structure (N—O-5'-(HO)(O)P—O—(HO)(O)P—O—P(HO)(O)—O-5'), 5'-monothiophosphate (phosphorothioate; (HO)2(S)P—O-5'), 5'-monodithiophosphate (phosphorodithioate; (HO)(HS)(S)P—O-5'), 5'-phosphorothiolate ((HO)2(O)P—S-5'); any additional combination of oxygen/sulfur replaced monophosphate, diphosphate and triphosphates (e.g. 5'-alpha-thiotriphosphate, 5'-gamma-thiotriphosphate, etc.), 5'-phosphoramidates ((HO)2(O)P—NH-5', (HO)(NH2)(O)P—O-5'), 5'-alkylphosphonates (R=alkyl=methyl, ethyl, isopropyl, propyl, etc., e.g. RP(OH)(O)—O-5', (OH)2(O)P-5'-CH2-), 5'-alkyletherphosphonates (R=alkylether=methoxymethyl (MeOCH2-), ethoxymethyl, etc., e.g. RP(OH)(O)—O-5'-).

9. The RISC of claim 1, wherein said RNA molecule is completely complementary to said target transcript, optionally with the exception of nucleotides that extend beyond position 20 (counted from the 5' terminus).

10. The RISC of claim 1, wherein said RNA molecule comprises at least one modified nucleotide analogue, which is preferably selected from sugar-backbone- and nucleobase-modified ribonucleotides and combinations thereof.

11. The RISC of claim 1, wherein said RNA molecule is associated with biodegradable polymers or microparticles, preferably wherein said association comprises a covalent coupling, in particular a covalent coupling via the 3'-terminus of the RNA molecule.

12. A host cell or non-human host organism capable of overexpressing RISC according to claim 1.

13. A method of enhancing RNAi in a cell or an organism comprising causing said cell or organism to overexpress at least one component of RISC according to claim 1.

14. The RISC molecule according to claim 1 for use as a target for diagnosis and/or therapy.

15. The RISC according to claim 1 for use as a diagnostic and/or therapeutic agent itself, as a molecular-biological reagent or as component in a screening procedure for identification and/or characterization of pharmaceutical agents.

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