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(57) **Abrégé/Abstract:**

Some aspects of this disclosure provide compositions, methods, and kits for improving the specificity of RNA-programmable endonucleases, such as Cas9. Also provided are variants of Cas9, e.g., Cas9 dimers and fusion proteins, engineered to have improved specificity for cleaving nucleic acid targets. Also provided are compositions, methods, and kits for site-specific nucleic acid modification using Cas9 fusion proteins (e.g., nuclease-inactivated Cas9 fused to a nuclease catalytic domain or a recombinase catalytic domain). Such Cas9 variants are useful in clinical and research settings involving site-specific modification of DNA, for example, genomic modifications.



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- (54) Title: CAS9 VARIANTS AND USES THEREOF
- (57) Abstract: Some aspects of this disclosure provide compositions, methods, and kits for improving the specificity of RNA-programmable endonucleases, such as Cas9. Also provided are variants of Cas9, e.g., Cas9 dimers and fusion proteins, engineered to have improved specificity for cleaving nucleic acid targets. Also provided are compositions, methods, and kits for site-specific nucleic acid modification using Cas9 fusion proteins (e.g., nuclease-inactivated Cas9 fused to a nuclease catalytic domain or a recombinase catalytic domain). Such Cas9 variants are useful in clinical and research settings involving site-specific modification of DNA, for example, genomic modifications.

CAS9 VARIANTS AND USES THEREOF

RELATED APPLICATION

[0001] This application claims priority under 35 U.S.C. § 365(c) to U.S. application, U.S.S.N 14/320,498, filed June 30, 2014, and to U.S. application, U.S.S.N 14/320,467, filed June 30, 2014, and also claims priority under 35 U.S.C. § 119(e) to U.S. provisional applications 61/874,609, filed September 6, 2013; 61/915,414, filed December 12, 2013; and 61/980,315, filed April 16, 2014; each of which is incorporated herein by reference.

BACKGROUND OF THE INVENTION

[0002] Site-specific endonucleases theoretically allow for the targeted manipulation of a single site within a genome and are useful in the context of gene targeting for therapeutic and research applications. In a variety of organisms, including mammals, site-specific endonucleases have been used for genome engineering by stimulating either non-homologous end joining or homologous recombination. In addition to providing powerful research tools, site-specific nucleases also have potential as gene therapy agents, and two site-specific endonucleases have recently entered clinical trials: (1) CCR5-2246, targeting a human CCR-5 allele as part of an anti-HIV therapeutic approach (NCT00842634, NCT01044654, NCT01252641); and (2) VF24684, targeting the human VEGF-A promoter as part of an anti-cancer therapeutic approach (NCT01082926).

[0003] Specific cleavage of the intended nuclease target site without or with only minimal off-target activity is a prerequisite for clinical applications of site-specific endonuclease, and also for high-efficiency genomic manipulations in basic research applications. For example, imperfect specificity of engineered site-specific binding domains has been linked to cellular toxicity and undesired alterations of genomic loci other than the intended target. Most nucleases available today, however, exhibit significant off-target activity, and thus may not be suitable for clinical applications. An emerging nuclease platform for use in clinical and research settings are the RNA-guided nucleases, such as Cas9. While these nucleases are able to bind guide RNAs (gRNAs) that direct cleavage of specific target sites, off-target activity is still observed for certain Cas9:gRNA complexes (Pattanayak *et al.*, “High-throughput profiling of off-target DNA cleavage reveals RNA-programmed Cas9 nuclease specificity.” *Nat Biotechnol.* 2013; doi: 10.1038/nbt.2673). Technology for engineering nucleases with improved specificity is therefore needed.

[0004] Another class of enzymes useful for targeted genetic manipulations are site-specific recombinases (SSRs). These enzymes perform rearrangements of DNA segments by recognizing and binding to short DNA sequences, at which they cleave the DNA backbone, exchange the two DNA helices involved and rejoin the DNA strands. Such rearrangements allow for the targeted insertion, inversion, excision, or translocation of DNA segments. However, like site-specific endonucleases, naturally-occurring SSRs typically recognize and bind specific consensus sequences, and are thus limited in this respect. Technology for engineering recombinases with altered and/or improved specificity is also needed.

SUMMARY OF THE INVENTION

[0005] Some aspects of this disclosure are based on the recognition that the reported toxicity of some engineered site-specific endonucleases is based on off-target DNA cleavage. Thus certain aspects described herein relate to the discovery that increasing the number of sequences (*e.g.*, having a nuclease bind at more than one site at a desired target), and/or splitting the activities (*e.g.*, target binding and target cleaving) of a nuclease between two or more proteins, will increase the specificity of a nuclease and thereby decrease the likelihood of off-target effects. Accordingly, some aspects of this disclosure provide strategies, compositions, systems, and methods to improve the specificity of site-specific nucleases, in particular, RNA-programmable endonucleases, such as Cas9 endonuclease. Certain aspects of this disclosure provide variants of Cas9 endonuclease engineered to have improved specificity.

[0006] Other aspects of this disclosure are based on the recognition that site-specific recombinases (SSRs) available today are typically limited to recognizing and binding distinct consensus sequences. Thus certain aspects described herein relate to the discovery that fusions between RNA-programmable (nuclease-inactivated) nucleases (or RNA-binding domains thereof), and a recombinase domain, provide novel recombinases theoretically capable of binding and recombining DNA at any site chosen, *e.g.*, by a practitioner (*e.g.*, sites specified by guide RNAs (gRNAs) that are engineered or selected according the sequence of the area to be recombined). Such novel recombinases are therefore useful, *inter alia*, for the targeted insertion, deletion, inversion, translocation or other genomic modifications. Thus, also provided are methods of using these inventive recombinase fusion proteins, *e.g.*, for such targeted genomic manipulations.

[0007] Accordingly, one embodiment of the disclosure provides fusion proteins and dimers thereof, for example, fusion proteins comprising two domains: (i) a nuclease-

inactivated Cas9 domain; and (ii) a nuclease domain (*e.g.*, a monomer of the FokI DNA cleavage domain). See *e.g.*, Figures 1A, 6D. The fusion protein may further comprise a nuclear localization signal (NLS) domain, which signals for the fusion proteins to be transported into the nucleus of a cell. In some embodiments, one or more domains of the fusion proteins are separated by a linker. In certain embodiments, the linker is a non-peptidic linker. In certain embodiments, the linker is a peptide linker. In the case of peptide linkers, the peptide linker may comprise an XTEN linker, an amino acid sequence comprising one or more repeats of the tri-peptide GGS, or any sequence as provided in Figure 12A. In some embodiments, the fusion protein is encoded by a nucleotide sequence set forth as SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, or SEQ ID NO:12, or a variant or fragment of any one of SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, or SEQ ID NO:12. The nuclease-inactivated Cas9 domain is capable of binding a guide RNA (gRNA). In certain embodiments, having dimers of such fusion protein each comprising a gRNA binding two distinct regions of a target nucleic acid provides for improved specificity, for example as compared to monomeric RNA-guided nucleases comprising a single gRNA to direct binding to the target nucleic acid.

[0008] According to another aspect of the invention, methods for site-specific DNA cleavage using the inventive Cas9 variants are provided. The methods typically comprise (a) contacting DNA with a fusion protein of the invention (*e.g.*, a fusion protein comprising a nuclease-inactivated Cas9 domain and a FokI DNA cleavage domain), wherein the inactive Cas9 domain binds a gRNA that hybridizes to a region of the DNA; (b) contacting the DNA with a second fusion protein (*e.g.*, a fusion protein comprising a nuclease-inactivated Cas9 and FokI DNA cleavage domain), wherein the inactive Cas9 domain of the second fusion protein binds a second gRNA that hybridizes to a second region of DNA; wherein the binding of the fusion proteins in steps (a) and (b) results in the dimerization of the nuclease domains of the fusion proteins, such that the DNA is cleaved in a region between the bound fusion proteins. In some embodiments, the gRNAs of steps (a) and (b) hybridize to the same strand of the DNA, or the gRNAs of steps (a) and (b) hybridize to opposite strands of the DNA. In some embodiments, the gRNAs of steps (a) and (b) hybridize to regions of the DNA that are no more than 10, no more than 15, no more than 20, no more than 25, no more than 30, no more than 40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart. In some embodiments, the DNA is in a cell, for example, a eukaryotic cell or a prokaryotic cell, which may be in an individual, such as a human.

[0009] According to another embodiment, a complex comprising a dimer of fusion proteins of the invention (*e.g.*, a dimer of a fusion protein comprising a nuclease-inactivated Cas9 and a FokI DNA cleavage domain) are provided. In some embodiments, the nuclease-inactivated Cas9 domain of each fusion protein of the dimer binds a single extended gRNA, such that one fusion protein of the dimer binds a portion of the gRNA, and the other fusion protein of the dimer binds another portion of the gRNA. See *e.g.*, Figure 1B. In some embodiments, the gRNA is at least 50, at least 75, at least 100, at least 150, at least 200, at least 250, or at least 300 nucleotides in length. In some embodiments, the regions of the extended gRNA that hybridize to a target nucleic acid comprise 15-25, 19-21, or 20 nucleotides.

[0010] In another embodiment, methods for site-specific DNA cleavage are provided comprising contacting a DNA with a complex of two inventive fusion proteins bound to a single extended gRNA. In some embodiments, the gRNA contains two portions that hybridize to two separate regions of the DNA to be cleaved; the complex binds the DNA as a result of the portions of the gRNA hybridizing to the two regions; and binding of the complex results in dimerization of the nuclease domains of the fusion proteins, such that the domains cleave the DNA in a region between the bound fusion proteins. In some embodiments, the two portions of the gRNA hybridize to the same strand of the DNA. In other embodiments, the two portions of the gRNA hybridize to opposing strands of the DNA. In some embodiments, the two portions of the gRNA hybridize to regions of the DNA that are no more 10, no more than 15, no more than 20, no more than 25, no more than 30, no more than 40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart. In some embodiments, the DNA is in a cell, for example, a eukaryotic cell or a prokaryotic cell, which may be in an individual, such as a human.

[0011] According to another embodiment of the invention, split Cas9 proteins (including fusion proteins comprising a split Cas9 protein) comprising fragments of a Cas9 protein are provided. In some embodiments, a protein is provided that includes a gRNA binding domain of Cas9 but does not include a DNA cleavage domain. In other embodiments, proteins comprising a DNA cleavage domain of Cas9, but not a gRNA binding domain, are provided. In some embodiments, a fusion protein comprising two domains: (i) a nuclease-inactivated Cas9 domain, and (ii) a gRNA binding domain of Cas9 are provided, for example, wherein domain (ii) does not include a DNA cleavage domain. See *e.g.*, Figure 2B. In some embodiments, fusion proteins comprising two domains: (i) a nuclease-inactivated Cas9 domain, and (ii) a DNA cleavage domain are provided, for example,

wherein domain (ii) does not include a gRNA binding domain. See *e.g.*, Figure 2C (fusion protein on right side, comprising a “B” domain). In some embodiments, protein dimers of any of the proteins described herein are provided. For example, in some embodiments, a dimer comprises two halves of a split Cas9 protein, for example, (i) a protein comprising a gRNA binding domain of Cas9, but not a DNA cleavage domain, and (ii) a protein comprising a DNA cleavage domain of Cas9, but not a gRNA binding domain. See *e.g.*, Figure 2A. In some embodiments, a dimer comprises one half of a split Cas9 protein, and a fusion protein comprising the other half of the split Cas9 protein. See *e.g.*, Figure 2B. For example, in certain embodiments such a dimer comprises (i) a protein comprising a gRNA binding domain of Cas9, but not a DNA cleavage domain, and (ii) a fusion protein comprising a nuclease-inactivated Cas9 and a DNA cleavage domain. In other embodiments, the dimer comprises (i) a protein comprising a DNA cleavage domain of Cas9, but not a gRNA binding domain, and (ii) a fusion protein comprising a nuclease-inactivated Cas9 and a gRNA binding domain of Cas9. In some embodiments, a dimer is provided that comprises two fusion proteins, each fusion protein comprising a nuclease-inactivated Cas9 and one half of a split Cas9. See *e.g.*, Figure 2C. For example, in certain embodiments, such a dimer comprises: (i) a fusion protein comprising a nuclease-inactivated Cas9 and a gRNA binding domain of Cas9, and (ii) a fusion protein comprising a nuclease-inactivated Cas9 and a DNA cleavage domain. In some embodiments, any of the provided protein dimers is associated with one or more gRNA(s).

[0012] In some embodiments, methods for site-specific DNA cleavage utilizing the inventive protein dimers are provided. For example, in some embodiments, such a method comprises contacting DNA with a protein dimer that comprises (i) a protein comprising a gRNA binding domain of Cas9, but not a DNA cleavage domain, and (ii) a protein comprising a DNA cleavage domain of Cas9, but not a gRNA binding domain, wherein the dimer binds a gRNA that hybridizes to a region of the DNA, and cleavage of the DNA occurs. See *e.g.*, Figure 2A. In some embodiments, the protein dimer used for site-specific DNA cleavage comprises (i) a protein comprising a gRNA binding domain of Cas9, but not a DNA cleavage domain, and (ii) a fusion protein comprising a nuclease-inactivated Cas9 and a DNA cleavage. See *e.g.*, Figure 2B. In some embodiments, the dimer used for site-specific DNA cleavage comprises (i) a protein comprising a DNA cleavage domain of Cas9, but not a gRNA binding domain, and (ii) a fusion protein comprising a nuclease-inactivated Cas9 and a gRNA binding domain of Cas9. In some embodiments, the protein dimer binds two gRNAs that hybridize to two regions of the DNA, and cleavage of the DNA occurs. See *e.g.*, Figure

2B. In some embodiments, the two gRNAs hybridize to regions of the DNA that are no more than 10, no more than 15, no more than 20, no more than 25, no more than 30, no more than 40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart. In some embodiments, the dimer used for site-specific DNA cleavage comprises two fusion proteins: (i) a fusion protein comprising a nuclease-inactivated Cas9 and a gRNA binding domain of Cas9, and (ii) a fusion protein comprising a nuclease-inactivated Cas9 and a DNA cleavage domain. In some embodiments, the protein dimer binds three gRNAs that hybridize to three regions of the DNA, and cleavage of the DNA occurs. Having such an arrangement, *e.g.*, targeting more than one region of a target nucleic acid, for example using dimers associated with more than one gRNA (or a gRNA comprising more than one region that hybridizes to the target) increases the specificity of cleavage as compared to a nuclease binding a single region of a target nucleic acid. In some embodiments, the three gRNAs hybridize to regions of the DNA that are no more than 10, no more than 15, no more than 20, no more than 25, no more than 30, no more than 40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart between the first and second, and the second and third regions. In some embodiments, the DNA is in a cell, for example, a eukaryotic cell or a prokaryotic cell, which may be in an individual, such as a human.

[0013] According to another embodiment, minimal Cas9 proteins are provided, for example, wherein the protein comprises N- and/or C-terminal truncations and retains RNA binding and DNA cleavage activity. In some embodiments, the N-terminal truncation removes at least 5, at least 10, at least 15, at least 20, at least 25, at least 40, at least 40, at least 50, at least 75, at least 100, or at least 150 amino acids. In some embodiments, the C-terminal truncation removes at least 5, at least 10, at least 15, at least 20, at least 25, at least 40, at least 40, at least 50, at least 75, at least 100, or at least 150 amino acids. In some embodiments, the minimized Cas9 protein further comprises a bound gRNA.

[0014] In some embodiments, methods for site-specific DNA cleavage are provided comprising contacting a DNA with minimized Cas9 protein:gRNA complex.

[0015] According to another embodiment, dimers of Cas9 (or fragments thereof) wherein the dimer is coordinated through a single gRNA are provided. In some embodiments, the single gRNA comprises at least two portions that (i) are each able to bind a Cas9 protein and (ii) each hybridize to a target nucleic acid sequence (*e.g.*, DNA sequence). In some embodiments, the portions of the gRNA that hybridize to the target nucleic acid each comprise no more than 5, no more than 10, or no more than 15 nucleotides complementary to

the target nucleic acid sequence. In some embodiments, the portions of the gRNA that hybridize to the target nucleic acid are separated by a linker sequence. In some embodiments, the linker sequence hybridizes to the target nucleic acid. See *e.g.*, Figure 4. In some embodiments, methods for site-specific DNA cleavage are provided comprising contacting DNA with a dimer of Cas9 proteins coordinated through a single gRNA.

[0016] According to another embodiment, the disclosure provides fusion proteins and dimers and tetramers thereof, for example, fusion proteins comprising two domains: (i) a nuclease-inactivated Cas9 domain; and (ii) a recombinase catalytic domain. See, *e.g.*, Figure 5. The recombinase catalytic domain, in some embodiments, is derived from the recombinase catalytic domain of Hin recombinase, Gin recombinase, or Tn3 resolvase. The nuclease-inactivated Cas9 domain is capable of binding a gRNA, *e.g.*, to target the fusion protein to a target nucleic acid sequence. The fusion proteins may further comprise a nuclear localization signal (NLS) domain, which signals for the fusion proteins to be transported into the nucleus of a cell. In some embodiments, one or more domains of the fusion proteins are separated by a linker. In certain embodiments, the linker is a non-peptidic linker. In certain embodiments, the linker is a peptide linker. In the case of peptide linkers, the peptide linker may comprise an XTEN linker, an amino acid sequence comprising one or more repeats of the tri-peptide GGS, or any sequence as provided in Figure 12A.

[0017] In another embodiment, methods for site-specific recombination are provided, which utilize the inventive RNA-guided recombinase fusion proteins described herein. In some embodiments, the method is useful for recombining two separate DNA molecules, and comprises (a) contacting a first DNA with a first RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain binds a first gRNA that hybridizes to a region of the first DNA; (b) contacting the first DNA with a second RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain of the second fusion protein binds a second gRNA that hybridizes to a second region of the first DNA; (c) contacting a second DNA with a third RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain of the third fusion protein binds a third gRNA that hybridizes to a region of the second DNA; and (d) contacting the second DNA with a fourth RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain of the fourth fusion protein binds a fourth gRNA that hybridizes to a second region of the second DNA, wherein the binding of the fusion proteins in steps (a) - (d) results in the tetramerization of the recombinase catalytic domains of the fusion proteins, under conditions such that the DNAs are recombined. In some embodiments, methods for site-specific recombination

between two regions of a single DNA molecule are provided. In some embodiments, the method comprises (a) contacting a DNA with a first RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain binds a first gRNA that hybridizes to a region of the DNA; (b) contacting the DNA with a second RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain of the second fusion protein binds a second gRNA that hybridizes to a second region of the DNA; (c) contacting the DNA with a third RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain of the third fusion protein binds a third gRNA that hybridizes to a third region of the DNA; (d) contacting the DNA with a fourth RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain of the fourth fusion protein binds a fourth gRNA that hybridizes to a fourth region of the DNA; wherein the binding of the fusion proteins in steps (a) - (d) results in the tetramerization of the recombinase catalytic domains of the fusion proteins, under conditions such that the DNA is recombined. In some embodiments involving methods for site-specific recombination, gRNAs hybridizing to the same DNA molecule hybridize to opposing strands of the DNA molecule. In some embodiments, *e.g.*, involving site-specific recombination of a single DNA molecule, two gRNAs hybridize to one strand of the DNA, and the other two gRNAs hybridize to the opposing strand. In some embodiments, the gRNAs hybridize to regions of their respective DNAs (*e.g.*, on the same strand) that are no more than 10, no more than 15, no more than 20, no more than 25, no more than 30, no more than 40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart. In some embodiments, the DNA is in a cell, for example, a eukaryotic cell or a prokaryotic cell, which may be in or obtained from an individual, such as a human.

[0018] According to another embodiment, polynucleotides are provided, for example, that encode any of the Cas9 proteins described herein (*e.g.*, Cas9 variants, Cas9 dimers, Cas9 fusion proteins, Cas9 fragments, minimized Cas9 proteins, Cas9 variants without a cleavage domain, Cas9 variants without a gRNA domain, Cas9-recombinase fusions, *etc.*). In some embodiments, polynucleotides encoding any of the gRNAs described herein are provided. In some embodiments, polynucleotides encoding any inventive Cas9 protein described herein and any combination of gRNA(s) as described herein are provided. In some embodiments, vectors that comprise a polynucleotide described herein are provided. In some embodiments, vectors for recombinant protein expression comprising a polynucleotide encoding any of the Cas9 proteins and/or gRNAs described herein are provided. In some

embodiments, cells comprising genetic constructs for expressing any of the Cas9 proteins and/or gRNAs described herein are provided.

[0019] In some embodiments, kits are provided. For example, kits comprising any of the Cas9 proteins and/or gRNAs described herein are provided. In some embodiments, kits comprising any of the polynucleotides described herein, *e.g.*, those encoding a Cas9 protein and/or gRNA, are provided. In some embodiments, kits comprising a vector for recombinant protein expression, wherein the vectors comprise a polynucleotide encoding any of the Cas9 proteins and/or gRNAs described herein, are provided. In some embodiments, kits comprising a cell comprising genetic constructs for expressing any of the Cas9 proteins and/or gRNAs described herein are provided.

[0020] Other advantages, features, and uses of the invention will be apparent from the Detailed Description of Certain Non-Limiting Embodiments of the Invention; the Drawings, which are schematic and not intended to be drawn to scale; and the Claims.

BRIEF DESCRIPTION OF THE DRAWINGS

[0021] **Figure 1** is a schematic detailing certain embodiments of the invention. (A) In this embodiment, nuclease-inactivated Cas9 protein is fused to a monomer of the FokI nuclease domain. Double-strand DNA-cleavage is achieved through dimerization of FokI monomers at the target site and is dependent on the simultaneous binding of two distinct Cas9:gRNA complexes. (B) In this embodiment, an alternate configuration is provided, wherein two Cas9-FokI fusions are coordinated through the action of a single extended gRNA containing two distinct gRNA motifs. The gRNA motifs comprise regions that hybridize the target in distinct regions, as well as regions that bind each fusion protein. The extended gRNA may enhance cooperative binding and alter the specificity profile of the fusions.

[0022] **Figure 2** is a schematic detailing certain embodiments of the invention. (A) In this embodiment, dimeric split Cas9 separates A) gRNA-binding ability from B) dsDNA cleavage. DNA cleavage occurs when both halves of the protein are co-localized to associate and refold into a nuclease-active state. (B) In this embodiment, nuclease-inactivated Cas9 mutant is fused to the A-half (or in some embodiments, the B-half) of the split Cas9 nuclease. Upon binding of both the Cas9-A-half (or Cas9-B-half) fusion and the inactive gRNA-binding Cas9 B-half (or A-half, respectively) at the target site, dsDNA is enabled following split protein reassembly. This split Cas9-pairing can use two distinct gRNA-binding Cas9 proteins to ensure the split nuclease-active Cas9 reassembles only on the

correct target sequence. (C) In this embodiment, nuclease-inactivated Cas9 mutant is fused to the A-half of the split Cas9 nuclease. A separate nuclease-inactivated Cas9 mutant is fused to the B-half of the split Cas9 nuclease. Upon binding of one nuclease-inactivated Cas9 mutant to a gRNA target site and binding of the other nuclease-inactivated Cas9 mutant to a second gRNA target site, the split Cas9 halves can dimerize and bind a third gRNA target to become a fully active Cas9 nuclease that can cleave dsDNA. This split Cas9-pairing uses three distinct gRNA-binding Cas9 proteins to ensure the split nuclease-active Cas9 reassembles only on the correct target sequence. Any other DNA-binding domain in place of the inactive Cas9 (zinc fingers, TALE proteins, *etc.*) can be used to complete the reassembly of the split Cas9 nuclease.

[0023] **Figure 3** shows schematically a minimal Cas9 protein that comprises the essential domains for Cas9 activity. Full-length Cas9 is a 4.1 kb gene which results in a protein of >150 kDa. Specific deletions and/or truncations decrease the size of Cas9 without affecting its activity (*e.g.*, gRNA binding and DNA cleavage activity). The minimized Cas9 protein increases the efficacy of, for example,, delivery to cells using viral vectors such as AAV (accommodating sequences ~ <4700 bp) or lentivirus (accommodating sequences ~ <9 kb), or when pursuing multiplexed gRNA/Cas9 approaches.

[0024] **Figure 4** shows how two Cas9 proteins can be coordinated through the action of a single extended RNA containing two distinct gRNA motifs. Each gRNA targeting region is shortened, such that a single Cas9:gRNA unit cannot bind efficiently by itself. The normal 20 nt targeting sequence has been altered so that some portion (*e.g.*, the 5' initial 10 nt) has been changed to some non-specific linker sequence, such as AAAAAAAAAA (SEQ ID NO:13), with only 10 nt of the gRNA remaining to direct target binding (alternatively this 5' 10 nt is truncated entirely). This "low-affinity" gRNA unit exists as part of a tandem gRNA construct with a second, distinct low-affinity gRNA unit downstream, separated by a linker sequence. In some embodiments, there are more than two low-affinity gRNA units (*e.g.*, at least 3, at least 4, at least 5, *etc.*). In some embodiments, the linker comprises a target nucleic acid complementary sequence (*e.g.*, as depicted by the linker region contacting the DNA target).

[0025] **Figure 5** shows schematically, how Cas9-recombinase fusions can be coordinated through gRNAs to bind and recombine target DNAs at desired sequences (sites). (A, B) Nuclease-inactivated Cas9 (dCas9) protein is fused to a monomer of a recombinase domain (Rec). Site-specific recombination is achieved through dimerization (A) of the recombinase catalytic domain monomers at the target site, and then tetramerization (B) of

two dimers assembled on separate Cas9-recombination sites. The fusion to dCas9:gRNA complexes determines the sequence identity of the flanking target sites while the recombinase catalytic domain determines the identity of the core sequence (the sequence between the two dCas9-binding sites). (B) Recombination proceeds through strand cleavage, exchange, and re-ligation within the dCas9-recombinase tetramer complex.

[0026] **Figure 6** shows architectures of Cas9 and *FokI*-dCas9 fusion variants. (A) Cas9 protein in complex with a guide RNA (gRNA) binds to target DNA. The *S. pyogenes* Cas9 protein recognizes the PAM sequence NGG, initiating unwinding of dsDNA and gRNA:DNA base pairing. (B) *FokI*-dCas9 fusion architectures tested. Four distinct configurations of NLS, *FokI* nuclease, and dCas9 were assembled. Seventeen (17) protein linker variants were also tested. (C) gRNA target sites tested within GFP. Seven gRNA target sites were chosen to test *FokI*-dCas9 activity in an orientation in which the PAM is distal from the cleaved spacer sequence (orientation A). Together, these seven gRNAs enabled testing of *FokI*-dCas9 fusion variants across spacer lengths ranging from 5 to 43 bp. See Figure 9 for guide RNAs used to test orientation B, in which the PAM is adjacent to the spacer sequence. (D) Monomers of *FokI* nuclease fused to dCas9 bind to separate sites within the target locus. Only adjacently bound *FokI*-dCas9 monomers can assemble a catalytically active *FokI* nuclease dimer, triggering dsDNA cleavage. The sequences shown in (C) are identified as follows: “EmGFP (bp 326-415)” corresponds to SEQ ID NO:204; “G1” corresponds to SEQ ID NO:205; “G2” corresponds to SEQ ID NO:206; “G3” corresponds to SEQ ID NO:207; “G4” corresponds to SEQ ID NO:208; “G5” corresponds to SEQ ID NO:209; “G6” corresponds to SEQ ID NO:210; and “G7” corresponds to SEQ ID NO:211.

[0027] **Figure 7** shows genomic DNA modification by fCas9, Cas9 nickase, and wild-type Cas9. (A) shows a graph depicting GFP disruption activity of fCas9, Cas9 nickase, or wild-type Cas9 with either no gRNA, or gRNA pairs of variable spacer length targeting the *GFP* gene in orientation A. (B) is an image of a gel showing Indel modification efficiency from PAGE analysis of a Surveyor cleavage assay of renatured target-site DNA amplified from cells treated with fCas9, Cas9 nickase, or wild-type Cas9 and two gRNAs spaced 14 bp apart targeting the *GFP* site (gRNAs G3 and G7; Figure 6C), each gRNA individually, or no gRNAs. The Indel modification percentage is shown below each lane for samples with modification above the detection limit (~2%). (C-G) show graphs depicting Indel modification efficiency for (C) two pairs of gRNAs spaced 14 or 25 bp apart targeting the *GFP* site, (D) one pair of gRNAs spaced 19 bp apart targeting the *CLTA* site, (E) one pair of

gRNAs spaced 23 bp apart targeting the *EMX* site, (F) one pair of gRNAs spaced 16 bp apart targeting the *HBB* site, and (G) two pairs of gRNAs spaced 14 or 16 bp apart targeting the *VEGF* site. Error bars reflect standard error of the mean from three biological replicates performed on different days.

[0028] **Figure 8** shows the DNA modification specificity of fCas9, Cas9 nickase, and wild-type Cas9. (A) shows a graph depicting GFP gene disruption by wild-type Cas9, Cas9 nickase, and fCas9 using gRNA pairs in orientation A. High activity of fCas9 requires spacer lengths of ~15 and 25 bp, roughly one DNA helical turn apart. (B) shows a graph depicting GFP gene disruption using gRNA pairs in orientation B. Cas9 nickase, but not fCas9, accepts either orientation of gRNA pairs. (C) shows a graph depicting GFP gene disruption by fCas9, but not Cas9 nickase or wild-type Cas9, which depends on the presence of two gRNAs. Four single gRNAs were tested along with three gRNA pairs of varying spacer length. In the presence of gRNA pairs in orientation A with spacer lengths of 14 or 25 bp (gRNAs 1+5, and gRNAs 3+7, respectively), fCas9 is active, but not when a gRNA pair with a 10-bp spacer (gRNAs 1+4) is used. In (A-C), “no treatment” refers to cells receiving no plasmid DNA. (D-F) show graphs depicting the indel mutation frequency from high-throughput DNA sequencing of amplified genomic on-target sites and off-target sites from human cells treated with fCas9, Cas9 nickase, or wild-type Cas9 and (D) two gRNAs spaced 19 bp apart targeting the *CLTA* site (gRNAs C1 and C2), (E) two gRNAs spaced 23 bp apart targeting the *EMX* site (gRNAs E1 and E2), or (F, G) two gRNAs spaced 14 bp apart targeting the *VEGF* site (gRNAs V1 and V2). (G) shows a graph depicting two in-depth trials to measure genome modification at *VEGF* off-target site 1. Trial 1 used 150 ng of genomic input DNA and $> 8 \times 10^5$ sequence reads for each sample; trial 2 used 600 ng of genomic input DNA and $> 23 \times 10^5$ sequence reads for each sample. In (D-G), all significant (P value < 0.005 Fisher’s Exact Test) indel frequencies are shown. P values are listed in Table 3. For (D-F) each on- and off-target sample was sequenced once with $> 10,000$ sequences analyzed per on-target sample and an average of 76,260 sequences analyzed per off-target sample (Table 3). The sequences shown in (C) are identified as follows, from top to bottom: the sequence found at the top of Figure 8C corresponds to SEQ ID NO:204; “G1” corresponds to SEQ ID NO:205; “G3” corresponds to SEQ ID NO:207; “G5” corresponds to SEQ ID NO:209; “G7” corresponds to SEQ ID NO:211; “G1+4” corresponds to SEQ ID NO:205 and SEQ ID NO:208; “G1+5” corresponds to SEQ ID NO:205 and SEQ ID NO:209; “G3+7” corresponds to SEQ ID NO:207 and SEQ ID NO:211.

[0029] **Figure 9** shows the target DNA sequences in a genomic GFP gene. Seven gRNA target sites were chosen to test FokI-dCas9 candidate activity in an orientation in which the PAM is adjacent to the cleaved spacer sequence (orientation B). Together, these seven gRNAs enabled testing of FokI-dCas9 fusion variants across six spacer lengths ranging from 4 to 42 bp. The sequences shown are identified as follows: “EmGFP (bp 297-388)” corresponds to SEQ ID NO:212; “G8” corresponds to SEQ ID NO:213; “G9” corresponds to SEQ ID NO:214; “G10” corresponds to SEQ ID NO:215; “G11” corresponds to SEQ ID NO:216; “G12” corresponds to SEQ ID NO:217; “G13” corresponds to SEQ ID NO:218; and “G14” corresponds to SEQ ID NO:219.

[0030] **Figure 10** shows a GFP disruption assay for measuring genomic DNA-modification activity. (A) depicts schematically a HEK293-derived cell line constitutively expressing a genomically integrated EmGFP gene used to test the activity of candidate FokI-dCas9 fusion constructs. Co-transfection of these cells with appropriate nuclease and gRNA expression plasmids leads to dsDNA cleavage within the EmGFP coding sequence, stimulating error-prone NHEJ and generating indels that can disrupt the expression of GFP, leading to loss of cellular fluorescence. The fraction of cells displaying a loss of GFP fluorescence is then quantitated by flow cytometry. (B) shows typical epifluorescence microscopy images at 200x magnification of EmGFP-HEK293 cells before and after co-transfection with wild-type Cas9 and gRNA expression plasmids.

[0031] **Figure 11** shows a graph depicting the activities of FokI-dCas9 fusion candidates combined with gRNA pairs of different orientations and varying spacer lengths. The fusion architectures described in Figure 6B were tested for functionality by flow cytometry using the GFP loss-of-function reporter across all (A) orientation A gRNA spacers and (B) orientation B gRNA spacers (Figure 6C and Figure 9). All FokI-dCas9 fusion data shown are the results of single trials. Wild-type Cas9 and Cas9 nickase data are the average of two replicates, while the ‘no treatment’ negative control data is the average of 6 replicates, with error bars representing one standard deviation. The grey dotted line across the Y-axis corresponds to the average of the ‘no treatment’ controls performed on the same day. The sequence shown as “(GGS)x3” corresponds to SEQ ID NO:14.

[0032] **Figure 12** shows the optimization of protein linkers in NLS-FokI-dCas9. (A) shows a table of all linker variants tested. Wild-type Cas9 and Cas9 nickase were included for comparison. The initial active construct NLS-FokI-dCas9 with a (GGS)₃ (SEQ ID NO:14) linker between FokI and dCas9 was tested across a range of alternate linkers. The final choice of linkers for fCas9 is highlighted. (B) shows a graph depicting the activity of

FokI-dCas9 fusions with linker variants. Each variant was tested across a range of spacer lengths from 5 to 43 bp using gRNA pair orientation A. A control lacking gRNA (“no gRNA”) was included for each separate fusion construct. NLS-FokI-dCas9 variant L8 showed the best activity, approaching the activity of Cas9 nickase. Variants L4 through L9 show peak activity with 14- and 25-bp spacer lengths, suggesting two optimal spacer lengths roughly one helical turn of dsDNA apart. The sequences shown in (A) are identified as follows: GGSGGSGGS corresponds to SEQ ID NO:14; GGSGGSGGSGGSGGSGGS corresponds to SEQ ID NO:15; MKIIEQLPSA corresponds to SEQ ID NO:22; VRHKLKRVGS corresponds to SEQ ID NO:23; VPFLLEPDNINGKTC corresponds to SEQ ID NO:19; GHGTGSTGSGSS corresponds to SEQ ID NO:24; MSRPDPA corresponds to SEQ ID NO:25; GSAGSAAGSGEF corresponds to SEQ ID NO:20; SGSETPGTSESA corresponds to SEQ ID NO:17; SGSETPGTSESATPES corresponds to SEQ ID NO:16; SGSETPGTSESATPEGGSGGS corresponds to SEQ ID NO:18; GGSM corresponds to SEQ ID NO:301; and SIVAQLSRPDPA corresponds to SEQ ID NO:21.

[0033] **Figure 13** shows target DNA sequences in endogenous human EMX, VEGF, CLTA, and HBB genes. The gRNA target sites tested within endogenous human EMX, VEGF, CLTA, and HBB genes are shown. Thirteen gRNA target sites were chosen to test the activity of the optimized fCas9 fusion in an orientation in which the PAM is distal from the cleaved spacer sequence (orientation A). Together, these 13 gRNAs enabled testing of fCas9 fusion variants across eight spacer lengths ranging from 5 to 47 bp. The sequences shown are identified as follows: “CLTA-1” corresponds to SEQ ID NO:220; “C1” corresponds to SEQ ID NO:221; “C2” corresponds to SEQ ID NO:222; “C3” corresponds to SEQ ID NO:224; “C4” corresponds to SEQ ID NO:225; “HBC” corresponds to SEQ ID NO:226; “H1” corresponds to SEQ ID NO:227; “H2” corresponds to SEQ ID NO:228; “H3” corresponds to SEQ ID NO:229; “H4” corresponds to SEQ ID NO:230; “H5” corresponds to SEQ ID NO:231; “H6” corresponds to SEQ ID NO:232; “H7” corresponds to SEQ ID NO:233; “EMX” corresponds to SEQ ID NO:234; “E1” corresponds to SEQ ID NO:235; “E2” corresponds to SEQ ID NO:236; “E3” corresponds to SEQ ID NO:237; “VEGF” corresponds to SEQ ID NO:238; “V1” corresponds to SEQ ID NO:239; “V2” corresponds to SEQ ID NO:240; “V3” corresponds to SEQ ID NO:241; and “V4” corresponds to SEQ ID NO:242.

[0034] **Figure 14** shows graphs depicting spacer length preference of genomic DNA modification by fCas9, Cas9 nickase, and wild-type Cas9. Indel modification efficiency for (A) pairs of gRNAs targeting the GFP site, (B) pairs of gRNAs targeting the CLTA site, (C)

pairs of gRNAs targeting the EMX site (D) pairs of gRNAs targeting the HBB site, and (E) pairs of gRNAs targeting the VEGF site. Error bars reflect standard error of the mean from three biological replicates performed on different days.

[0035] **Figure 15** shows graphs depicting the efficiency of genomic DNA modification by fCas9, Cas9 nickase, and wild-type Cas9 with varying amounts of Cas9 and gRNA expression plasmids. Indel modification efficiency from a Surveyor assay of renatured target-site DNA amplified from a population of cells treated with fCas9, Cas9 nickase, or wild-type Cas9 and two target site gRNAs. Either 700 ng of Cas9 expression plasmid with 250 ng of gRNA expression plasmid (950 ng total), 350 ng of Cas9 expression plasmid with 125 ng of gRNA expression plasmid (475 ng in total), 175 ng of Cas9 expression plasmid with 62.5 ng of gRNA expression plasmid (238 ng in total) or 88 ng of Cas9 expression plasmid with 31 ng of gRNA expression plasmid (119 ng in total) were transfected with an appropriate amount of inert, carrier plasmid to ensure uniform transfection of 950 ng of plasmid across all treatments. Indel modification efficiency for (A) gRNAs spaced 19-bp apart targeting the CLTA site, (B) gRNAs spaced 23 bp apart targeting the EMX site, and (C) gRNAs spaced 14 bp apart targeting the VEGF site. Error bars represent the standard error of the mean from three biological replicates performed on separate days.

[0036] **Figure 16** shows the ability of fCas9, Cas9 nickase, and wild-type Cas9 to modify genomic DNA in the presence of a single gRNA. (A) shows images of gels depicting Surveyor assay of a genomic GFP target from DNA of cells treated with the indicated combination of Cas9 protein and gRNA(s). Single gRNAs do not induce genome modification at a detectable level (< 2% modification) for both fCas9 and Cas9 nickase. Wild-type Cas9 effectively modifies the GFP target for all tested single and paired gRNAs. For both fCas9 and Cas9 nickase, appropriately paired gRNAs induce genome modification at levels comparable to those of wild-type Cas9. (B) shows a graph depicting the results from sequencing GFP on-target sites amplified from 150 ng genomic DNA isolated from human cells treated with a plasmid expressing either wild-type Cas9, Cas9 nickase, or fCas9 and either a single plasmid expressing a single gRNAs (G1, G3, G5 or G7), or two plasmids each expressing a different gRNA (G1+G5, or G3+G7). As a negative control, transfection and sequencing were performed in triplicate as above without any gRNA expression plasmids. Error bars represent s.d. Sequences with more than one insertion or deletion at the GFP target site (the start of the G1 binding site to the end of the G7 binding site) were considered indels. Indel percentages were calculated by dividing the number of indels by total number

of sequences. While wild-type Cas9 produced indels across all gRNA treatments, fCas9 and Cas9 nickase produced indels efficiently (> 1%) only when paired gRNAs were present. Indels induced by fCas9 and single gRNAs were not detected above the no-gRNA control, while Cas9 nickase and single gRNAs modified the target GFP sequence at an average rate of 0.12%.

[0037] **Figure 17** shows a graph depicting how fCas9 indel frequency of genomic targets reflects gRNA pair spacer length preference. The graph shows the relationship between spacer length (number of bp between two gRNAs) and the indel modification efficiency of fCas9 normalized to the indel modification efficiency of the same gRNAs co-expressed with wild-type Cas9 nuclease. Colored triangles below the X-axis denote spacer lengths that were tested but which yielded no detectable indels for the indicated target gene. These results suggest that fCas9 requires ~15 bp or ~25 bp between half-sites to efficiently cleave DNA.

[0038] **Figure 18** shows modifications induced by Cas9 nuclease, Cas9 nickases, or fCas9 nucleases at endogenous loci. (A) shows examples of modified sequences at the VEGF on-target site with wild-type Cas9 nuclease, Cas9 nickases, or fCas9 nucleases and a single plasmid expressing two gRNAs targeting the VEGF on-target site (gRNA V1 and gRNA V2). For each example shown, the unmodified genomic site is the first sequence, followed by the top eight sequences containing deletions. The numbers before each sequence indicate sequencing counts. The gRNA target sites are bold and capitalized. (B) is an identical analysis as in (A) for VEGF off-target site 1 VEG_Off1. (C) shows the potential binding mode of two gRNAs to VEGF off-target site 1. The top strand is bound in a canonical mode, while the bottom strand binds the second gRNA, gRNA V2, through gRNA:DNA base pairing that includes G:U base pairs. The sequences shown in (A) are identified, top to bottom, as follows: SEQ ID NO:243; SEQ ID NO:244; SEQ ID NO:245; SEQ ID NO:246; SEQ ID NO:247; SEQ ID NO:248; SEQ ID NO:249; SEQ ID NO:250; SEQ ID NO:251; SEQ ID NO:252; SEQ ID NO:253; SEQ ID NO:254; SEQ ID NO:255; SEQ ID NO:256; SEQ ID NO:257; SEQ ID NO:258; SEQ ID NO:259; SEQ ID NO:260; SEQ ID NO:261; SEQ ID NO:262; SEQ ID NO:263; SEQ ID NO:264; SEQ ID NO:265; SEQ ID NO:266; SEQ ID NO:267; SEQ ID NO:268; and SEQ ID NO:269. The sequences shown in (B) are identified, top to bottom, as follows: SEQ ID NO:270; SEQ ID NO:271; SEQ ID NO:272; SEQ ID NO:273; SEQ ID NO:274; SEQ ID NO:275; SEQ ID NO:276; SEQ ID NO:277; SEQ ID NO:278; SEQ ID NO:279; SEQ ID NO:280; SEQ ID NO:281; SEQ ID NO:282; SEQ ID NO:283; SEQ ID NO:284; SEQ ID NO:285; SEQ ID NO:286;

SEQ ID NO:287; SEQ ID NO:288; SEQ ID NO:289; SEQ ID NO:290; SEQ ID NO:291; SEQ ID NO:292; SEQ ID NO:293; SEQ ID NO:294; SEQ ID NO:295; and SEQ ID NO:296. The sequences shown in (C) are identified, top to bottom, as follows: SEQ ID NO:297; SEQ ID NO:298; SEQ ID NO:299; and SEQ ID NO:300.

[0039] **Figure 19** shows the target DNA sequences in a genomic CCR5 gene. (A) Eight gRNA target sites were identified for testing Cas9 variant (*e.g.*, FokI-dCas9) activity in an orientation in which the PAM is adjacent to the cleaved spacer sequence (orientation A). (B) Six gRNA target sites were identified for testing Cas9 variant (*e.g.*, FokI-dCas9) activity in an orientation in which the PAM is adjacent to the cleaved spacer sequence (orientation B). Together, these fourteen gRNAs enable testing of Cas9 fusion variants across spacer lengths ranging from 0 to 74 bp. The sequences shown in (A) are identified as follows: “CRA” corresponds to SEQ ID NO:302; “CRA-1” corresponds to SEQ ID NO:303; “CRA-2” corresponds to SEQ ID NO:304; “CRA-3” corresponds to SEQ ID NO:305; “CRA-4” corresponds to SEQ ID NO:306; “CRA-5” corresponds to SEQ ID NO:307; “CRA-6” corresponds to SEQ ID NO:308; “CRA-7” corresponds to SEQ ID NO:309; and “CRA-8” corresponds to SEQ ID NO:310. The sequences shown in (B) are identified as follows: “CRB” corresponds to SEQ ID NO:311; “CB-1” corresponds to SEQ ID NO:312; “CB-2” corresponds to SEQ ID NO:313; “CB-3” corresponds to SEQ ID NO:314; “CB-4” corresponds to SEQ ID NO:315; “CB-5” corresponds to SEQ ID NO:316; and “CB-6” corresponds to SEQ ID NO:317.

[0040] **Figure 20** depicts a vector map detailing an exemplary plasmid containing a FokI-dCas9 (fCas9) construct.

DEFINITIONS

[0041] As used herein and in the claims, the singular forms “a,” “an,” and “the” include the singular and the plural reference unless the context clearly indicates otherwise. Thus, for example, a reference to “an agent” includes a single agent and a plurality of such agents.

[0042] The term “Cas9” or “Cas9 nuclease” refers to an RNA-guided nuclease comprising a Cas9 protein, or a fragment thereof (*e.g.*, a protein comprising an active or inactive DNA cleavage domain of Cas9, and/or the gRNA binding domain of Cas9). A Cas9 nuclease is also referred to sometimes as a casn1 nuclease or a CRISPR (clustered regularly interspaced short palindromic repeat)-associated nuclease. CRISPR is an adaptive immune system that provides protection against mobile genetic elements (viruses, transposable

elements and conjugative plasmids). CRISPR clusters contain spacers, sequences complementary to antecedent mobile elements, and target invading nucleic acids. CRISPR clusters are transcribed and processed into CRISPR RNA (crRNA). In type II CRISPR systems correct processing of pre-crRNA requires a trans-encoded small RNA (tracrRNA), endogenous ribonuclease 3 (rnc) and a Cas9 protein. The tracrRNA serves as a guide for ribonuclease 3-aided processing of pre-crRNA. Subsequently, Cas9/crRNA/tracrRNA endonucleolytically cleaves linear or circular dsDNA target complementary to the spacer. The target strand not complementary to crRNA is first cut endonucleolytically, then trimmed 3'-5' exonucleolytically. In nature, DNA-binding and cleavage typically requires protein and both RNA. However, single guide RNAs ("sgRNA", or simply "gRNA") can be engineered so as to incorporate aspects of both the crRNA and tracrRNA into a single RNA species. See *e.g.*, Jinek M., Chylinski K., Fonfara I., Hauer M., Doudna J.A., Charpentier E. *Science* 337:816-821(2012), the entire contents of which is hereby incorporated by reference. Cas9 recognizes a short motif in the CRISPR repeat sequences (the PAM or protospacer adjacent motif) to help distinguish self versus non-self. Cas9 nuclease sequences and structures are well known to those of skill in the art (see, *e.g.*, "Complete genome sequence of an M1 strain of *Streptococcus pyogenes*." Ferretti et al., J.J., McShan W.M., Ajdic D.J., Savic D.J., Savic G., Lyon K., Primeaux C., Sezate S., Suvorov A.N., Kenton S., Lai H.S., Lin S.P., Qian Y., Jia H.G., Najar F.Z., Ren Q., Zhu H., Song L., White J., Yuan X., Clifton S.W., Roe B.A., McLaughlin R.E., *Proc. Natl. Acad. Sci. U.S.A.* 98:4658-4663(2001); "CRISPR RNA maturation by trans-encoded small RNA and host factor RNase III." Deltcheva E., Chylinski K., Sharma C.M., Gonzales K., Chao Y., Pirzada Z.A., Eckert M.R., Vogel J., Charpentier E., *Nature* 471:602-607(2011); and "A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity." Jinek M., Chylinski K., Fonfara I., Hauer M., Doudna J.A., Charpentier E. *Science* 337:816-821(2012), the entire contents of each of which are incorporated herein by reference). Cas9 orthologs have been described in various species, including, but not limited to, *S. pyogenes* and *S. thermophilus*. Additional suitable Cas9 nucleases and sequences will be apparent to those of skill in the art based on this disclosure, and such Cas9 nucleases and sequences include Cas9 sequences from the organisms and loci disclosed in Chylinski, Rhun, and Charpentier, "The tracrRNA and Cas9 families of type II CRISPR-Cas immunity systems" (2013) *RNA Biology* 10:5, 726-737; the entire contents of which are incorporated herein by reference. In some embodiments, a Cas9 nuclease has an inactive (*e.g.*, an inactivated) DNA cleavage domain. A nuclease-inactivated Cas9 protein may interchangeably be referred to as a "dCas9" protein (for nuclease "dead" Cas9). In some

embodiments, dCas9 corresponds to, or comprises in part or in whole, the amino acid set forth as SEQ ID NO:5, below. In some embodiments, variants of dCas9 (*e.g.*, variants of SEQ ID NO:5) are provided. For example, in some embodiments, variants having mutations other than D10A and H840A are provided, which *e.g.*, result in nuclease inactivated Cas9 (dCas9). Such mutations, by way of example, include other amino acid substitutions at D10 and H840, or other substitutions within the nuclease domains of Cas9 (*e.g.*, substitutions in the HNH nuclease subdomain and/or the RuvC1 subdomain). In some embodiments, variants or homologues of dCas9 (*e.g.*, variants of SEQ ID NO:5) are provided which are at least about 70% identical, at least about 80% identical, at least about 90% identical, at least about 95% identical, at least about 98% identical, at least about 99% identical, at least about 99.5% identical, or at least about 99.9% to SEQ ID NO:5. In some embodiments, variants of dCas9 (*e.g.*, variants of SEQ ID NO:5) are provided having amino acid sequences which are shorter, or longer than SEQ ID NO:5, by about 5 amino acids, by about 10 amino acids, by about 15 amino acids, by about 20 amino acids, by about 25 amino acids, by about 30 amino acids, by about 40 amino acids, by about 50 amino acids, by about 75 amino acids, by about 100 amino acids or more.

[0043] dCas9 (D10A and H840A):

MDKKYSIGLAIGTNSVGWAVITDEYKVPSSKKFKVLGNTDRHSIKKNLIGALLFDSGETAEATRLKRTARRRYTRR
KNRICYLQEIFSNEMAKVDDSFHRLSESLVEEDKKHERHPIFGNIVDEVAYHEKYPTIYHLRKKLV DSTDKAD
LRLIYLALAHMIKFRGHFLIEGDLNPDNSDVKLFIQLVQTYNQLFEEENPINASGVDAKAILSARLSKSRRLNL
IAQLPGEKKNGLFGNLIALSLGLTPNFKSNFDLAEDAKQLSKD TYDDDLNLLAQIGDQYADLFLAAKNLSDAI
LLSDILRVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQLPEKYKEIFFDQSKNGYAGYIDGGASQEEFYKF
IKPILEKMDGTEELLVKLNREDLLRKQRTFDNGSIPHQIHLGELHAILRRQEDFYFPLKDNREKIEKILTFRIPY
YVGPLARGNSRFAMTRKSEETITPWNFEVVDKGASQSFIERMTNFDKNLPNEKVLPHSLLYEYFTVYNELT
KVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKIECFDSVEISGVEDRFNASLGTYHDLLKI
IKDKDFLDNEENEDILEDIVLTTLTFEDREMIEERLKYAHLFDDKVMKQLKRRRYTGWGRLSRKLINGIRDKQS
GKTILDFLKS DGFANRNFQMQLIHDDSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPAIIKKGILQTVKVVDELVKV
MGRHKPENIVIEMARENQTTQKGQKNSRERMKRIEEGIKELGSQILKEHPVENTQLQNEKLYLYYLQNGRDMYVD
QELDINRLSDYDVDAIVPQSFLKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKKMKNYWRQLLNAKLITQRKFDNL
TKAERGGLSELKDAGFIKRQLVETRQITKHVAQILDSRMNTKYDENDKLIREVKVITLKS KLVSDFRKDFQFYKV
REINNYHHAHDAYLNAVVG TALIKKYPKLESEFVYGDYKVYDVRKMIKSEQEIGKATAKYFFYSNIMNFFKTEI
TLANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVL SMPQVNIVKKTEVQTGGFSKESILPKRNSDKLIARKKD
WDPKKYGGFDSPTVAYSVLVAKVEKGKSKKLKSVKELLGITIMERSSEFKNPIDFLEAKGYKEVKKDLIIKL PK
YSLFELENGRKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLGKSPEDNEQKQLFVEQHKHYLDEII EQISE
FSKRVLADANLDKVL SAYNKH RDKPIREQAENIIHLFTLTNLGAPAAFKYFDTTIDRKRYTSTKEVLDATLIHQ
SITGLYETRIDLSQLGGD (SEQ ID NO:5)

[0044] Methods for generating a Cas9 protein (or a fragment thereof) having an inactive DNA cleavage domain are known (See, *e.g.*, the Examples; and Jinek *et al.*, *Science*. 337:816-821(2012); Qi *et al.*, “Repurposing CRISPR as an RNA-Guided Platform for Sequence-Specific Control of Gene Expression” (2013) *Cell*. 28;152(5):1173-83, the entire

contents of each of which are incorporated herein by reference). For example, the DNA cleavage domain of Cas9 is known to include two subdomains, the HNH nuclease subdomain and the RuvC1 subdomain. The HNH subdomain cleaves the strand complementary to the gRNA, whereas the RuvC1 subdomain cleaves the non-complementary strand. Mutations within these subdomains can silence the nuclease activity of Cas9. For example, the mutations D10A and H840A completely inactivate the nuclease activity of *S. pyogenes* Cas9 (See *e.g.*, the Examples; and Jinek *et al.*, *Science*. 337:816-821(2012); Qi *et al.*, *Cell*. 28;152(5):1173-83 (2013)). In some embodiments, proteins comprising fragments of Cas9 are provided. For example, in some embodiments, a protein comprises one of two Cas9 domains: (1) the gRNA binding domain of Cas9; or (2) the DNA cleavage domain of Cas9. In some embodiments, proteins comprising Cas9 or fragments thereof are referred to as “Cas9 variants.” A Cas9 variant shares homology to Cas9, or a fragment thereof. For example a Cas9 variant is at least about 70% identical, at least about 80% identical, at least about 90% identical, at least about 95% identical, at least about 98% identical, at least about 99% identical, at least about 99.5% identical, or at least about 99.9% to wild type Cas9. In some embodiments, the Cas9 variant comprises a fragment of Cas9 (*e.g.*, a gRNA binding domain or a DNA-cleavage domain), such that the fragment is at least about 70% identical, at least about 80% identical, at least about 90% identical, at least about 95% identical, at least about 98% identical, at least about 99% identical, at least about 99.5% identical, or at least about 99.9% to the corresponding fragment of wild type Cas9. In some embodiments, wild type Cas9 corresponds to Cas9 from *Streptococcus pyogenes* (NCBI Reference Sequence: NC_017053.1, SEQ ID NO:1 (nucleotide); SEQ ID NO:2 (amino acid)).

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 QIGDQYADLFLAAKNLSDAILLSDILRVNSEITKAPLSASMIKRYDEHHQDLTLLKALVRQQLPEKYKEI
 FFDQSKNGYAGYIDGGASQEEFYKF IKPILEKMDGTEELLVKLNREDLLRKQRTFDNGSIPHQIHLGELH
 AILRRQEDFYFPLKDNREKIEKILTFRIPYVVGPLARGNSRFWMTRKSEETITPWNFEEVVDKGASAQS
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 SLTFKEDIQKAQVSGQGHSLSHEQIANLAGSPA IKKGILQTVKIVDELVKVMGHKPENIVIEMARENQTTQ
 KGQKNSRERMKRIEEGIKELGSQILKEHPVENTQLQNEKLYLYYLQNGRDMYVDQELDINRLSDYDVDHI
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 RKKDWD PKKYGGFDSPTVAYSVLVVAKEVGKSKKLKSVKELLGITIMERSSSF EKNPIDFLEAKGYKEVK
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 DRKRYTSTKEVLDATLIHQ SITGLYETRIDLSQLGGD (SEQ ID NO:2)

[0045] In some embodiments, wild type Cas9 corresponds to, or comprises, SEQ ID NO:3 (nucleotide) and/or SEQ ID NO:4 (amino acid).

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 LRLIYLALAHMIKFRGHFLIEGDLNPDNSDVKLFIQLVQTYNQLFEENPINASGVDAKAILSARLSKSRLENL
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 SITGLYETRIDLSQLGGD (SEQ ID NO:4)

[0046] In some embodiments, Cas9 refers to Cas9 from: *Corynebacterium ulcerans* (NCBI Refs: NC_015683.1, NC_017317.1); *Corynebacterium diphtheria* (NCBI Refs: NC_016782.1, NC_016786.1); *Spiroplasma syrphidicola* (NCBI Ref: NC_021284.1); *Prevotella intermedia* (NCBI Ref: NC_017861.1); *Spiroplasma taiwanense* (NCBI Ref: NC_021846.1); *Streptococcus iniae* (NCBI Ref: NC_021314.1); *Belliella baltica* (NCBI Ref: NC_018010.1); *Psychroflexus torquis* (NCBI Ref: NC_018721.1); *Streptococcus thermophilus* (NCBI Ref: YP_820832.1), *Listeria innocua* (NCBI Ref: NP_472073.1), *Campylobacter jejuni* (NCBI Ref: YP_002344900.1) or *Neisseria meningitidis* (NCBI Ref: YP_002342100.1).

[0047] The terms “conjugating,” “conjugated,” and “conjugation” refer to an association of two entities, for example, of two molecules such as two proteins, two domains (e.g., a binding domain and a cleavage domain), or a protein and an agent, e.g., a protein binding domain and a small molecule. In some aspects, the association is between a protein (e.g., RNA-programmable nuclease) and a nucleic acid (e.g., a guide RNA). The association can be, for example, via a direct or indirect (e.g., via a linker) covalent linkage. In some embodiments, the association is covalent. In some embodiments, two molecules are conjugated via a linker connecting both molecules. For example, in some embodiments where two proteins are conjugated to each other, e.g., a binding domain and a cleavage domain of an engineered nuclease, to form a protein fusion, the two proteins may be conjugated via a polypeptide linker, e.g., an amino acid sequence connecting the C-terminus of one protein to the N-terminus of the other protein.

[0048] The term “consensus sequence,” as used herein in the context of nucleic acid sequences, refers to a calculated sequence representing the most frequent nucleotide residues found at each position in a plurality of similar sequences. Typically, a consensus sequence is determined by sequence alignment in which similar sequences are compared to each other and similar sequence motifs are calculated. In the context of nuclease target site sequences, a consensus sequence of a nuclease target site may, in some embodiments, be the sequence most frequently bound, or bound with the highest affinity, by a given nuclease. In the context of recombinase target site sequences, a consensus sequence of a recombinase target site may, in some embodiments, be the sequence most frequently bound, or bound with the highest affinity, by a given recombinase.

[0001] The term “engineered,” as used herein refers to a protein molecule, a nucleic acid, complex, substance, or entity that has been designed, produced, prepared, synthesized, and/or manufactured by a human. Accordingly, an engineered product is a product that does not occur in nature.

[0049] The term “effective amount,” as used herein, refers to an amount of a biologically active agent that is sufficient to elicit a desired biological response. For example, in some embodiments, an effective amount of a nuclease may refer to the amount of the nuclease that is sufficient to induce cleavage of a target site specifically bound and cleaved by the nuclease. In some embodiments, an effective amount of a recombinase may refer to the amount of the recombinase that is sufficient to induce recombination at a target site specifically bound and recombined by the recombinase. As will be appreciated by the skilled artisan, the effective amount of an agent, *e.g.*, a nuclease, a recombinase, a hybrid protein, a fusion protein, a protein dimer, a complex of a protein (or protein dimer) and a polynucleotide, or a polynucleotide, may vary depending on various factors as, for example, on the desired biological response, the specific allele, genome, target site, cell, or tissue being targeted, and the agent being used.

[0050] The term “homologous,” as used herein is an art-understood term that refers to nucleic acids or polypeptides that are highly related at the level of nucleotide and/or amino acid sequence. Nucleic acids or polypeptides that are homologous to each other are termed “homologues.” Homology between two sequences can be determined by sequence alignment methods known to those of skill in the art. In accordance with the invention, two sequences are considered to be homologous if they are at least about 50-60% identical, *e.g.*, share identical residues (*e.g.*, amino acid residues) in at least about 50-60% of all residues comprised in one or the other sequence, at least about 70% identical, at least about 80%

identical, at least about 90% identical, at least about 95% identical, at least about 98% identical, at least about 99% identical, at least about 99.5% identical, or at least about 99.9% identical, for at least one stretch of at least 20, at least 30, at least 40, at least 50, at least 60, at least 70, at least 80, at least 90, at least 100, at least 120, at least 150, or at least 200 amino acids.

[0051] The term “linker,” as used herein, refers to a chemical group or a molecule linking two adjacent molecules or moieties, *e.g.*, a binding domain (*e.g.*, dCas9) and a cleavage domain of a nuclease (*e.g.*, FokI). In some embodiments, a linker joins a nuclear localization signal (NLS) domain to another protein (*e.g.*, a Cas9 protein or a nuclease or recombinase or a fusion thereof). In some embodiments, a linker joins a gRNA binding domain of an RNA-programmable nuclease and the catalytic domain of a recombinase. In some embodiments, a linker joins a dCas9 and a recombinase. Typically, the linker is positioned between, or flanked by, two groups, molecules, or other moieties and connected to each one via a covalent bond, thus connecting the two. In some embodiments, the linker is an amino acid or a plurality of amino acids (*e.g.*, a peptide or protein). In some embodiments, the linker is an organic molecule, group, polymer, or chemical moiety. In some embodiments, the linker is a peptide linker. In some embodiments, the peptide linker is any stretch of amino acids having at least 1, at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 15, at least 20, at least 25, at least 30, at least 40, at least 50, or more amino acids. In some embodiments, the peptide linker comprises repeats of the tri-peptide Gly-Gly-Ser, *e.g.*, comprising the sequence (GGG)_n, wherein n represents at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more repeats. In some embodiments, the linker comprises the sequence (GGG)₆ (SEQ ID NO:15). In some embodiments, the peptide linker is the 16 residue “XTEN” linker, or a variant thereof (See, *e.g.*, the Examples; and Schellenberger *et al.* A recombinant polypeptide extends the in vivo half-life of peptides and proteins in a tunable manner. *Nat. Biotechnol.* **27**, 1186–1190 (2009)). In some embodiments, the XTEN linker comprises the sequence SGSETPGTSESATPES (SEQ ID NO:16), SGSETPGTSESA (SEQ ID NO:17), or SGSETPGTSESATPEGGSGGS (SEQ ID NO:18). In some embodiments, the peptide linker is any linker as provided in Figure 12A, for example, one or more selected from VPFLLEPDNINGKTC (SEQ ID NO:19), GSAGSAAGSGEF (SEQ ID NO:20), SIVAQLSRPDPA (SEQ ID NO:21), MKIIEQLPSA (SEQ ID NO:22), VRHKLKRVGS (SEQ ID NO:23), GHGTGSTGSGSS (SEQ ID NO:24), MSRPDPA (SEQ ID NO:25); or GGSM (SEQ ID NO:301).

[0052] The term “mutation,” as used herein, refers to a substitution of a residue within a sequence, *e.g.*, a nucleic acid or amino acid sequence, with another residue, or a deletion or insertion of one or more residues within a sequence. Mutations are typically described herein by identifying the original residue followed by the position of the residue within the sequence and by the identity of the newly substituted residue. Various methods for making the amino acid substitutions (mutations) provided herein are well known in the art, and are provided by, for example, Green and Sambrook, *Molecular Cloning: A Laboratory Manual* (4th ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (2012)).

[0053] The term “nuclease,” as used herein, refers to an agent, for example, a protein, capable of cleaving a phosphodiester bond connecting two nucleotide residues in a nucleic acid molecule. In some embodiments, “nuclease” refers to a protein having an inactive DNA cleavage domain, such that the nuclease is incapable of cleaving a phosphodiester bond. In some embodiments, a nuclease is a protein, *e.g.*, an enzyme that can bind a nucleic acid molecule and cleave a phosphodiester bond connecting nucleotide residues within the nucleic acid molecule. A nuclease may be an endonuclease, cleaving a phosphodiester bonds within a polynucleotide chain, or an exonuclease, cleaving a phosphodiester bond at the end of the polynucleotide chain. In some embodiments, a nuclease is a site-specific nuclease, binding and/or cleaving a specific phosphodiester bond within a specific nucleotide sequence, which is also referred to herein as the “recognition sequence,” the “nuclease target site,” or the “target site.” In some embodiments, a nuclease is a RNA-guided (*i.e.*, RNA-programmable) nuclease, which is associated with (*e.g.*, binds to) an RNA (*e.g.*, a guide RNA, “gRNA”) having a sequence that complements a target site, thereby providing the sequence specificity of the nuclease. In some embodiments, a nuclease recognizes a single stranded target site, while in other embodiments, a nuclease recognizes a double-stranded target site, for example, a double-stranded DNA target site. The target sites of many naturally occurring nucleases, for example, many naturally occurring DNA restriction nucleases, are well known to those of skill in the art. In many cases, a DNA nuclease, such as EcoRI, HindIII, or BamHI, recognize a palindromic, double-stranded DNA target site of 4 to 10 base pairs in length, and cut each of the two DNA strands at a specific position within the target site. Some endonucleases cut a double-stranded nucleic acid target site symmetrically, *i.e.*, cutting both strands at the same position so that the ends comprise base-paired nucleotides, also referred to herein as blunt ends. Other endonucleases cut a double-stranded nucleic acid target sites asymmetrically, *i.e.*, cutting each strand at a different position so that the ends comprise

unpaired nucleotides. Unpaired nucleotides at the end of a double-stranded DNA molecule are also referred to as “overhangs,” *e.g.*, as “5'-overhang” or as “3'-overhang,” depending on whether the unpaired nucleotide(s) form(s) the 5' or the 3' end of the respective DNA strand. Double-stranded DNA molecule ends ending with unpaired nucleotide(s) are also referred to as sticky ends, as they can “stick to” other double-stranded DNA molecule ends comprising complementary unpaired nucleotide(s). A nuclease protein typically comprises a “binding domain” that mediates the interaction of the protein with the nucleic acid substrate, and also, in some cases, specifically binds to a target site, and a “cleavage domain” that catalyzes the cleavage of the phosphodiester bond within the nucleic acid backbone. In some embodiments a nuclease protein can bind and cleave a nucleic acid molecule in a monomeric form, while, in other embodiments, a nuclease protein has to dimerize or multimerize in order to cleave a target nucleic acid molecule. Binding domains and cleavage domains of naturally occurring nucleases, as well as modular binding domains and cleavage domains that can be fused to create nucleases binding specific target sites, are well known to those of skill in the art. For example, the binding domain of RNA-programmable nucleases (*e.g.*, Cas9), or a Cas9 protein having an inactive DNA cleavage domain, can be used as a binding domain (*e.g.*, that binds a gRNA to direct binding to a target site) to specifically bind a desired target site, and fused or conjugated to a cleavage domain, for example, the cleavage domain of FokI, to create an engineered nuclease cleaving the target site. In some embodiments, Cas9 fusion proteins provided herein comprise the cleavage domain of FokI, and are therefore referred to as “fCas9” proteins. In some embodiments, the cleavage domain of FokI, *e.g.*, in a fCas9 protein corresponds to, or comprises in part or whole, the amino acid sequence (or variants thereof) set forth as SEQ ID NO:6, below. In some embodiments, variants or homologues of the FokI cleavage domain include any variant or homologue capable of dimerizing (*e.g.*, as part of fCas9 fusion protein) with another FokI cleavage domain at a target site in a target nucleic acid, thereby resulting in cleavage of the target nucleic acid. In some embodiments, variants of the FokI cleavage domain (*e.g.*, variants of SEQ ID NO:6) are provided which are at least about 70% identical, at least about 80% identical, at least about 90% identical, at least about 95% identical, at least about 98% identical, at least about 99% identical, at least about 99.5% identical, or at least about 99.9% to SEQ ID NO:6. In some embodiments, variants of the FokI cleavage domain (*e.g.*, variants of SEQ ID NO:6) are provided having an amino acid sequence which is shorter, or longer than SEQ ID NO:6, by about 5 amino acids, by about 10 amino acids, by about 15 amino acids, by about 20 amino acids, by about 25 amino acids, by

about 30 amino acids, by about 40 amino acids, by about 50 amino acids, by about 75 amino acids, by about 100 amino acids or more.

[0054] Cleavage domain of FokI:

GSQLVKSELEEKKSELRHKLKYVPHEYIELIEIARNSTQDRILEMKVMEFFMKVYGYRGKHLGGSRKPDGAIYTV
GSPIDYGVIVDTKAYSGGYNLPIGQADEMQRYVEENQTRNKHINPNEWVKVYPSSVTEFKFLFVSGHFKGNYKAQ
LTRLNHITNCNGAVLSVEELLIGGEMIKAGTLTLEEVRKFNNGEINF (SEQ ID NO:6)

[0055] The terms “nucleic acid” and “nucleic acid molecule,” as used herein, refer to a compound comprising a nucleobase and an acidic moiety, *e.g.*, a nucleoside, a nucleotide, or a polymer of nucleotides. Typically, polymeric nucleic acids, *e.g.*, nucleic acid molecules comprising three or more nucleotides are linear molecules, in which adjacent nucleotides are linked to each other via a phosphodiester linkage. In some embodiments, “nucleic acid” refers to individual nucleic acid residues (*e.g.* nucleotides and/or nucleosides). In some embodiments, “nucleic acid” refers to an oligonucleotide chain comprising three or more individual nucleotide residues. As used herein, the terms “oligonucleotide” and “polynucleotide” can be used interchangeably to refer to a polymer of nucleotides (*e.g.*, a string of at least three nucleotides). In some embodiments, “nucleic acid” encompasses RNA as well as single and/or double-stranded DNA. Nucleic acids may be naturally occurring, for example, in the context of a genome, a transcript, an mRNA, tRNA, rRNA, siRNA, snRNA, gRNA, a plasmid, cosmid, chromosome, chromatid, or other naturally occurring nucleic acid molecule. On the other hand, a nucleic acid molecule may be a non-naturally occurring molecule, *e.g.*, a recombinant DNA or RNA, an artificial chromosome, an engineered genome, or fragment thereof, or a synthetic DNA, RNA, DNA/RNA hybrid, or including non-naturally occurring nucleotides or nucleosides. Furthermore, the terms “nucleic acid,” “DNA,” “RNA,” and/or similar terms include nucleic acid analogs, *i.e.* analogs having other than a phosphodiester backbone. Nucleic acids can be purified from natural sources, produced using recombinant expression systems and optionally purified, chemically synthesized, *etc.* Where appropriate, *e.g.*, in the case of chemically synthesized molecules, nucleic acids can comprise nucleoside analogs such as analogs having chemically modified bases or sugars, and backbone modifications. A nucleic acid sequence is presented in the 5' to 3' direction unless otherwise indicated. In some embodiments, a nucleic acid is or comprises natural nucleosides (*e.g.* adenosine, thymidine, guanosine, cytidine, uridine, deoxyadenosine, deoxythymidine, deoxyguanosine, and deoxycytidine); nucleoside analogs (*e.g.*, 2-aminoadenosine, 2-thiothymidine, inosine, pyrrolo-pyrimidine, 3-methyl adenosine,

5-methylcytidine, 2-aminoadenosine, C5-bromouridine, C5-fluorouridine, C5-iodouridine, C5-propynyl-uridine, C5-propynyl-cytidine, C5-methylcytidine, 2-aminoadenosine, 7-deazaadenosine, 7-deazaguanosine, 8-oxoadenosine, 8-oxoguanosine, O(6)-methylguanine, and 2-thiocytidine); chemically modified bases; biologically modified bases (*e.g.*, methylated bases); intercalated bases; modified sugars (*e.g.*, 2'-fluororibose, ribose, 2'-deoxyribose, arabinose, and hexose); and/or modified phosphate groups (*e.g.*, phosphorothioates and 5'-*N*-phosphoramidite linkages).

[0056] The term “pharmaceutical composition,” as used herein, refers to a composition that can be administrated to a subject in the context of treatment and/or prevention of a disease or disorder. In some embodiments, a pharmaceutical composition comprises an active ingredient, *e.g.*, a nuclease or recombinase fused to a Cas9 protein, or fragment thereof (or a nucleic acid encoding a such a fusion), and optionally a pharmaceutically acceptable excipient. In some embodiments, a pharmaceutical composition comprises inventive Cas9 variant/fusion (*e.g.*, fCas9) protein(s) and gRNA(s) suitable for targeting the Cas9 variant/fusion protein(s) to a target nucleic acid. In some embodiments, the target nucleic acid is a gene. In some embodiments, the target nucleic acid is an allele associated with a disease, whereby the allele is cleaved by the action of the Cas9 variant/fusion protein(s). In some embodiments, the allele is an allele of the *CLTA* gene, the *EMX* gene, the *HBB* gene, the *VEGF* gene, or the *CCR5* gene. See *e.g.*, the Examples; Figures 7, 8, 13, 14, 15, 17 and 19.

[0057] The term “proliferative disease,” as used herein, refers to any disease in which cell or tissue homeostasis is disturbed in that a cell or cell population exhibits an abnormally elevated proliferation rate. Proliferative diseases include hyperproliferative diseases, such as pre-neoplastic hyperplastic conditions and neoplastic diseases. Neoplastic diseases are characterized by an abnormal proliferation of cells and include both benign and malignant neoplasias. Malignant neoplasia is also referred to as cancer. In some embodiments, the compositions and methods provided herein are useful for treating a proliferative disease. For example, in some embodiments, pharmaceutical compositions comprising Cas9 (*e.g.*, fCas9) protein(s) and gRNA(s) suitable for targeting the Cas9 protein(s) to an *VEGF* allele, whereby the allele is inactivated by the action of the Cas9 protein(s). See, *e.g.*, the Examples.

[0058] The terms “protein,” “peptide,” and “polypeptide” are used interchangeably herein, and refer to a polymer of amino acid residues linked together by peptide (amide) bonds. The terms refer to a protein, peptide, or polypeptide of any size, structure, or function.

Typically, a protein, peptide, or polypeptide will be at least three amino acids long. A protein, peptide, or polypeptide may refer to an individual protein or a collection of proteins. One or more of the amino acids in a protein, peptide, or polypeptide may be modified, for example, by the addition of a chemical entity such as a carbohydrate group, a hydroxyl group, a phosphate group, a farnesyl group, an isofarnesyl group, a fatty acid group, a linker for conjugation, functionalization, or other modification, *etc.* A protein, peptide, or polypeptide may also be a single molecule or may be a multi-molecular complex. A protein, peptide, or polypeptide may be just a fragment of a naturally occurring protein or peptide. A protein, peptide, or polypeptide may be naturally occurring, recombinant, or synthetic, or any combination thereof. The term “fusion protein” as used herein refers to a hybrid polypeptide which comprises protein domains from at least two different proteins. One protein may be located at the amino-terminal (N-terminal) portion of the fusion protein or at the carboxy-terminal (C-terminal) protein thus forming an “amino-terminal fusion protein” or a “carboxy-terminal fusion protein,” respectively. Any of the proteins provided herein may be produced by any method known in the art. For example, the proteins provided herein may be produced via recombinant protein expression and purification, which is especially suited for fusion proteins comprising a peptide linker. Methods for recombinant protein expression and purification are well known, and include those described by Green and Sambrook, *Molecular Cloning: A Laboratory Manual* (4th ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (2012)), the entire contents of which are incorporated herein by reference.

[0059] The term “RNA-programmable nuclease,” and “RNA-guided nuclease” are used interchangeably herein and refer to a nuclease that forms a complex with (*e.g.*, binds or associates with) one or more RNA that is not a target for cleavage. In some embodiments, an RNA-programmable nuclease, when in a complex with an RNA, may be referred to as a nuclease:RNA complex. Typically, the bound RNA(s) is referred to as a guide RNA (gRNA). gRNAs can exist as a complex of two or more RNAs, or as a single RNA molecule. gRNAs that exist as a single RNA molecule may be referred to as single-guide RNAs (sgRNAs), though “gRNA” is used interchangeably to refer to guide RNAs that exist as either single molecules or as a complex of two or more molecules. Typically, gRNAs that exist as single RNA species comprise two domains: (1) a domain that shares homology to a target nucleic acid (*e.g.*, and directs binding of a Cas9 complex to the target); and (2) a domain that binds a Cas9 protein. In some embodiments, domain (2) corresponds to a sequence known as a tracrRNA, and comprises a stem-loop structure. For example, in some embodiments, domain (2) is homologous to a tracrRNA as depicted in Figure 1E of Jinek *et*

al., *Science* 337:816-821(2012), the entire contents of which is incorporated herein by reference. Other examples of gRNAs (*e.g.*, those including domain 2) can be found in U.S. Provisional Patent Application, U.S.S.N. 61/874,682, filed September 6, 2013, entitled “Switchable Cas9 Nucleases And Uses Thereof;” U.S. Provisional Patent Application, U.S.S.N. 61/874,746, filed September 6, 2013, entitled “Delivery System For Functional Nucleases;” PCT Application WO 2013/176722, filed March 15, 2013, entitled “Methods and Compositions for RNA-Directed Target DNA Modification and for RNA-Directed Modulation of Transcription;” and PCT Application WO 2013/142578, filed March 20, 2013, entitled “RNA-Directed DNA Cleavage by the Cas9-crRNA Complex;” the entire contents of each are hereby incorporated by reference in their entirety. Still other examples of gRNAs and gRNA structure are provided herein. See *e.g.*, the Examples. In some embodiments, a gRNA comprises two or more of domains (1) and (2), and may be referred to as an “extended gRNA.” For example, an extended gRNA will *e.g.*, bind two or more Cas9 proteins and bind a target nucleic acid at two or more distinct regions, as described herein. The gRNA comprises a nucleotide sequence that complements a target site, which mediates binding of the nuclease/RNA complex to said target site, providing the sequence specificity of the nuclease:RNA complex. In some embodiments, the RNA-programmable nuclease is the (CRISPR-associated system) Cas9 endonuclease, for example Cas9 (Csn1) from *Streptococcus pyogenes* (see, *e.g.*, “Complete genome sequence of an M1 strain of *Streptococcus pyogenes*.” Ferretti J.J., McShan W.M., Ajdic D.J., Savic D.J., Savic G., Lyon K., Primeaux C., Sezate S., Suvorov A.N., Kenton S., Lai H.S., Lin S.P., Qian Y., Jia H.G., Najar F.Z., Ren Q., Zhu H., Song L., White J., Yuan X., Clifton S.W., Roe B.A., McLaughlin R.E., *Proc. Natl. Acad. Sci. U.S.A.* 98:4658-4663(2001); “CRISPR RNA maturation by trans-encoded small RNA and host factor RNase III.” Deltcheva E., Chylinski K., Sharma C.M., Gonzales K., Chao Y., Pirzada Z.A., Eckert M.R., Vogel J., Charpentier E., *Nature* 471:602-607(2011); and “A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity.” Jinek M., Chylinski K., Fonfara I., Hauer M., Doudna J.A., Charpentier E. *Science* 337:816-821(2012), the entire contents of each of which are incorporated herein by reference.

[0060] Because RNA-programmable nucleases (*e.g.*, Cas9) use RNA:DNA hybridization to determine target DNA cleavage sites, these proteins are able to cleave, in principle, any sequence specified by the guide RNA. Methods of using RNA-programmable nucleases, such as Cas9, for site-specific cleavage (*e.g.*, to modify a genome) are known in the art (see *e.g.*, Cong, L. *et al.* Multiplex genome engineering using CRISPR/Cas systems.

Science **339**, 819-823 (2013); Mali, P. *et al.* RNA-guided human genome engineering via Cas9. *Science* **339**, 823-826 (2013); Hwang, W.Y. *et al.* Efficient genome editing in zebrafish using a CRISPR-Cas system. *Nature biotechnology* **31**, 227-229 (2013); Jinek, M. *et al.* RNA-programmed genome editing in human cells. *eLife* **2**, e00471 (2013); Dicarlo, J.E. *et al.* Genome engineering in *Saccharomyces cerevisiae* using CRISPR-Cas systems. *Nucleic acids research* (2013); Jiang, W. *et al.* RNA-guided editing of bacterial genomes using CRISPR-Cas systems. *Nature biotechnology* **31**, 233-239 (2013); the entire contents of each of which are incorporated herein by reference).

[0061] The term “recombinase,” as used herein, refers to a site-specific enzyme that mediates the recombination of DNA between recombinase recognition sequences, which results in the excision, integration, inversion, or exchange (*e.g.*, translocation) of DNA fragments between the recombinase recognition sequences. Recombinases can be classified into two distinct families: serine recombinases (*e.g.*, resolvases and invertases) and tyrosine recombinases (*e.g.*, integrases). Examples of serine recombinases include, without limitation, Hin, Gin, Tn3, β -six, CinH, ParA, $\gamma\delta$, Bxb1, ϕ C31, TP901, TG1, ϕ BT1, R4, ϕ RV1, ϕ FC1, MR11, A118, U153, and gp29. Examples of tyrosine recombinases include, without limitation, Cre, FLP, R, Lambda, HK101, HK022, and pSAM2. The serine and tyrosine recombinase names stem from the conserved nucleophilic amino acid residue that the recombinase uses to attack the DNA and which becomes covalently linked to the DNA during strand exchange. Recombinases have numerous applications, including the creation of gene knockouts/knock-ins and gene therapy applications. See, *e.g.*, Brown *et al.*, “Serine recombinases as tools for genome engineering.” *Methods*. 2011;53(4):372-9; Hirano *et al.*, “Site-specific recombinases as tools for heterologous gene integration.” *Appl. Microbiol. Biotechnol.* 2011; 92(2):227-39; Chavez and Calos, “Therapeutic applications of the Φ C31 integrase system.” *Curr. Gene Ther.* 2011;11(5):375-81; Turan and Bode, “Site-specific recombinases: from tag-and-target- to tag-and-exchange-based genomic modifications.” *FASEB J.* 2011; 25(12):4088-107; Venken and Bellen, “Genome-wide manipulations of *Drosophila melanogaster* with transposons, Flp recombinase, and Φ C31 integrase.” *Methods Mol. Biol.* 2012; 859:203-28; Murphy, “Phage recombinases and their applications.” *Adv. Virus Res.* 2012; 83:367-414; Zhang *et al.*, “Conditional gene manipulation: Cre-ating a new biological era.” *J. Zhejiang Univ. Sci. B.* 2012; 13(7):511-24; Karpenshif and Bernstein, “From yeast to mammals: recent advances in genetic control of homologous recombination.” *DNA Repair (Amst)*. 2012; 1;11(10):781-8; the entire contents of each are hereby incorporated by reference in their entirety. The recombinases provided herein are not meant

to be exclusive examples of recombinases that can be used in embodiments of the invention. The methods and compositions of the invention can be expanded by mining databases for new orthogonal recombinases or designing synthetic recombinases with defined DNA specificities (See, *e.g.*, Groth et al., “Phage integrases: biology and applications.” *J. Mol. Biol.* 2004; 335, 667-678; Gordley et al., “Synthesis of programmable integrases.” *Proc. Natl. Acad. Sci. U S A.* 2009; 106, 5053-5058; the entire contents of each are hereby incorporated by reference in their entirety). Other examples of recombinases that are useful in the methods and compositions described herein are known to those of skill in the art, and any new recombinase that is discovered or generated is expected to be able to be used in the different embodiments of the invention. In some embodiments, the catalytic domains of a recombinase are fused to a nuclease-inactivated RNA-programmable nuclease (*e.g.*, dCas9, or a fragment thereof), such that the recombinase domain does not comprise a nucleic acid binding domain or is unable to bind to a target nucleic acid (*e.g.*, the recombinase domain is engineered such that it does not have specific DNA binding activity). Recombinases lacking DNA binding activity and methods for engineering such are known, and include those described by Klippel et al., “Isolation and characterisation of unusual gin mutants.” *EMBO J.* 1988; 7: 3983–3989; Burke et al., “Activating mutations of Tn3 resolvase marking interfaces important in recombination catalysis and its regulation. *Mol Microbiol.* 2004; 51: 937–948; Olorunniji et al., “Synapsis and catalysis by activated Tn3 resolvase mutants.” *Nucleic Acids Res.* 2008; 36: 7181–7191; Rowland et al., “Regulatory mutations in Sin recombinase support a structure-based model of the synaptosome.” *Mol Microbiol.* 2009; 74: 282–298; Akopian et al., “Chimeric recombinases with designed DNA sequence recognition.” *Proc Natl Acad Sci USA.* 2003;100: 8688–8691; Gordley et al., “Evolution of programmable zinc finger-recombinases with activity in human cells. *J Mol Biol.* 2007; 367: 802–813; Gordley et al., “Synthesis of programmable integrases.” *Proc Natl Acad Sci USA.* 2009;106: 5053–5058; Arnold et al., “Mutants of Tn3 resolvase which do not require accessory binding sites for recombination activity.” *EMBO J.* 1999;18: 1407–1414; Gaj et al., “Structure-guided reprogramming of serine recombinase DNA sequence specificity.” *Proc Natl Acad Sci USA.* 2011;108(2):498-503; and Proudfoot et al., “Zinc finger recombinases with adaptable DNA sequence specificity.” *PLoS One.* 2011;6(4):e19537; the entire contents of each are hereby incorporated by reference. For example, serine recombinases of the resolvase-invertase group, *e.g.*, Tn3 and $\gamma\delta$ resolvases and the Hin and Gin invertases, have modular structures with autonomous catalytic and DNA-binding domains (See, *e.g.*, Grindley et al., “Mechanism of site-specific recombination.” *Ann Rev Biochem.* 2006; 75: 567–605, the entire contents of

which are incorporated by reference). The catalytic domains of these recombinases are thus amenable to being recombined with nuclease-inactivated RNA-programmable nucleases (*e.g.*, dCas9, or a fragment thereof) as described herein, *e.g.*, following the isolation of ‘activated’ recombinase mutants which do not require any accessory factors (*e.g.*, DNA binding activities) (See, *e.g.*, Klippel *et al.*, “Isolation and characterisation of unusual gin mutants.” *EMBO J.* 1988; 7: 3983–3989; Burke *et al.*, “Activating mutations of Tn3 resolvase marking interfaces important in recombination catalysis and its regulation. *Mol Microbiol.* 2004; 51: 937–948; Olorunniji *et al.*, “Synapsis and catalysis by activated Tn3 resolvase mutants.” *Nucleic Acids Res.* 2008; 36: 7181–7191; Rowland *et al.*, “Regulatory mutations in Sin recombinase support a structure-based model of the synaptosome.” *Mol Microbiol.* 2009; 74: 282–298; Akopian *et al.*, “Chimeric recombinases with designed DNA sequence recognition.” *Proc Natl Acad Sci USA.* 2003;100: 8688–8691). Additionally, many other natural serine recombinases having an N-terminal catalytic domain and a C-terminal DNA binding domain are known (*e.g.*, phiC31 integrase, TnpX transposase, IS607 transposase), and their catalytic domains can be co-opted to engineer programmable site-specific recombinases as described herein (See, *e.g.*, Smith *et al.*, “Diversity in the serine recombinases.” *Mol Microbiol.* 2002;44: 299–307, the entire contents of which are incorporated by reference). Similarly, the core catalytic domains of tyrosine recombinases (*e.g.*, Cre, λ integrase) are known, and can be similarly co-opted to engineer programmable site-specific recombinases as described herein (See, *e.g.*, Guo *et al.*, “Structure of Cre recombinase complexed with DNA in a site-specific recombination synapse.” *Nature.* 1997; 389:40–46; Hartung *et al.*, “Cre mutants with altered DNA binding properties.” *J Biol Chem* 1998; 273:22884–22891; Shaikh *et al.*, “Chimeras of the Flp and Cre recombinases: Tests of the mode of cleavage by Flp and Cre. *J Mol Biol.* 2000; 302:27–48; Rongrong *et al.*, “Effect of deletion mutation on the recombination activity of Cre recombinase.” *Acta Biochim Pol.* 2005; 52:541–544; Kilbride *et al.*, “Determinants of product topology in a hybrid Cre-Tn3 resolvase site-specific recombination system.” *J Mol Biol.* 2006; 355:185–195; Warren *et al.*, “A chimeric cre recombinase with regulated directionality.” *Proc Natl Acad Sci USA.* 2008 105:18278–18283; Van Duyne, “Teaching Cre to follow directions.” *Proc Natl Acad Sci USA.* 2009 Jan 6;106(1):4-5; Numrych *et al.*, “A comparison of the effects of single-base and triple-base changes in the integrase arm-type binding sites on the site-specific recombination of bacteriophage λ .” *Nucleic Acids Res.* 1990; 18:3953–3959; Tirumalai *et al.*, “The recognition of core-type DNA sites by λ integrase.” *J Mol Biol.* 1998; 279:513–527; Aihara *et al.*, “A conformational switch controls the DNA cleavage activity of λ integrase.” *Mol*

Cell. 2003; 12:187–198; Biswas *et al.*, “A structural basis for allosteric control of DNA recombination by λ integrase.” *Nature*. 2005; 435:1059–1066; and Warren *et al.*, “Mutations in the amino-terminal domain of λ -integrase have differential effects on integrative and excisive recombination.” *Mol Microbiol*. 2005; 55:1104–1112; the entire contents of each are incorporated by reference).

[0062] The term “recombine,” or “recombination,” in the context of a nucleic acid modification (*e.g.*, a genomic modification), is used to refer to the process by which two or more nucleic acid molecules, or two or more regions of a single nucleic acid molecule, are modified by the action of a recombinase protein (*e.g.*, an inventive recombinase fusion protein provided herein). Recombination can result in, *inter alia*, the insertion, inversion, excision or translocation of nucleic acids, *e.g.*, in or between one or more nucleic acid molecules.

[0063] The term “subject,” as used herein, refers to an individual organism, for example, an individual mammal. In some embodiments, the subject is a human. In some embodiments, the subject is a non-human mammal. In some embodiments, the subject is a non-human primate. In some embodiments, the subject is a rodent. In some embodiments, the subject is a sheep, a goat, a cattle, a cat, or a dog. In some embodiments, the subject is a vertebrate, an amphibian, a reptile, a fish, an insect, a fly, or a nematode. In some embodiments, the subject is a research animal. In some embodiments, the subject is genetically engineered, *e.g.*, a genetically engineered non-human subject. The subject may be of either sex and at any stage of development.

[0064] The terms “target nucleic acid,” and “target genome,” as used herein in the context of nucleases, refer to a nucleic acid molecule or a genome, respectively, that comprises at least one target site of a given nuclease. In the context of fusions comprising a (nuclease-inactivated) RNA-programmable nuclease and a recombinase domain, a “target nucleic acid” and a “target genome” refers to one or more nucleic acid molecule(s), or a genome, respectively, that comprises at least one target site. In some embodiments, the target nucleic acid(s) comprises at least two, at least three, or at least four target sites. In some embodiments, the target nucleic acid(s) comprise four target sites.

[0065] The term “target site” refers to a sequence within a nucleic acid molecule that is either (1) bound and cleaved by a nuclease (*e.g.*, Cas9 fusion proteins provided herein), or (2) bound and recombined (*e.g.*, at or nearby the target site) by a recombinase (*e.g.*, a dCas9-recombinase fusion protein provided herein). A target site may be single-stranded or double-stranded. In the context of RNA-guided (*e.g.*, RNA-programmable) nucleases (*e.g.*, a protein

dimer comprising a Cas9 gRNA binding domain and an active Cas9 DNA cleavage domain or other nuclease domain such as FokI), a target site typically comprises a nucleotide sequence that is complementary to the gRNA(s) of the RNA-programmable nuclease, and a protospacer adjacent motif (PAM) at the 3' end adjacent to the gRNA-complementary sequence(s). In some embodiments, such as those involving fCas9, a target site can encompass the particular sequences to which fCas9 monomers bind, and/or the intervening sequence between the bound monomers that are cleaved by the dimerized FokI domains (See *e.g.*, the Examples; and Figures 1A, 6D). In the context of fusions between RNA-guided (*e.g.*, RNA-programmable, nuclease-inactivated) nucleases and a recombinase (*e.g.*, a catalytic domain of a recombinase), a target site typically comprises a nucleotide sequence that is complementary to the gRNA of the RNA-programmable nuclease domain, and a protospacer adjacent motif (PAM) at the 3' end adjacent to the gRNA-complementary sequence. For example, in some embodiments, four recombinase monomers are coordinated to recombine a target nucleic acid(s), each monomer being fused to a (nuclease-inactivated) Cas9 protein guided by a gRNA. In such an example, each Cas9 domain is guided by a distinct gRNA to bind a target nucleic acid(s), thus the target nucleic acid comprises four target sites, each site targeted by a separate dCas9-recombinase fusion (thereby coordinating four recombinase monomers which recombine the target nucleic acid(s)). For the RNA-guided nuclease Cas9 (or gRNA-binding domain thereof) and inventive fusions of Cas9, the target site may be, in some embodiments, 17-20 base pairs plus a 3 base pair PAM (*e.g.*, NNN, wherein N independently represents any nucleotide). Typically, the first nucleotide of a PAM can be any nucleotide, while the two downstream nucleotides are specified depending on the specific RNA-guided nuclease. Exemplary target sites (*e.g.*, comprising a PAM) for RNA-guided nucleases, such as Cas9, are known to those of skill in the art and include, without limitation, NNG, NGN, NAG, and NGG, wherein N independently represents any nucleotide. In addition, Cas9 nucleases from different species (*e.g.*, *S. thermophilus* instead of *S. pyogenes*) recognizes a PAM that comprises the sequence NGGNG. Additional PAM sequences are known, including, but not limited to, NNAGAAW and NAAR (see, *e.g.*, Esvelt and Wang, *Molecular Systems Biology*, 9:641 (2013), the entire contents of which are incorporated herein by reference). In some aspects, the target site of an RNA-guided nuclease, such as, *e.g.*, Cas9, may comprise the structure [N_z]-[PAM], where each N is, independently, any nucleotide, and z is an integer between 1 and 50, inclusive. In some embodiments, z is at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at

least 17, at least 18, at least 19, at least 20, at least 25, at least 30, at least 35, at least 40, at least 45, or at least 50. In some embodiments, *z* is 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50. In some embodiments, *z* is 20. In some embodiments, “target site” may also refer to a sequence within a nucleic acid molecule that is bound but not cleaved by a nuclease. For example, certain embodiments described herein provide proteins comprising an inactive (or inactivated) Cas9 DNA cleavage domain. Such proteins (*e.g.*, when also including a Cas9 RNA binding domain) are able to bind the target site specified by the gRNA; however, because the DNA cleavage site is inactivated, the target site is not cleaved by the particular protein. However, such proteins as described herein are typically conjugated, fused, or bound to another protein (*e.g.*, a nuclease) or molecule that mediates cleavage of the nucleic acid molecule. In other embodiments, such proteins are conjugated, fused, or bound to a recombinase (or a catalytic domain of a recombinase), which mediates recombination of the target nucleic acid. In some embodiments, the sequence actually cleaved or recombined will depend on the protein (*e.g.*, nuclease or recombinase) or molecule that mediates cleavage or recombination of the nucleic acid molecule, and in some cases, for example, will relate to the proximity or distance from which the inactivated Cas9 protein(s) is/are bound.

[0066] In the context of inventive proteins that dimerize (or multimerize), for example, dimers of a protein comprising a nuclease-inactivated Cas9 (or a Cas9 RNA binding domain) and a DNA cleavage domain (*e.g.*, FokI cleavage domain or an active Cas9 cleavage domain), or fusions between a nuclease-inactivated Cas9 (or a Cas9 gRNA binding domain) and a recombinase (or catalytic domain of a recombinase), a target site typically comprises a left-half site (bound by one protein), a right-half site (bound by the second protein), and a spacer sequence between the half sites in which the cut or recombination is made. In some embodiments, either the left-half site or the right half-site (and not the spacer sequence) is cut or recombined. In other embodiments, the spacer sequence is cut or recombined. This structure ([left-half site]-[spacer sequence]-[right-half site]) is referred to herein as an LSR structure. In some embodiments, the left-half site and/or the right-half site correspond to an RNA-guided target site (*e.g.*, a Cas9 target site). In some embodiments, either or both half-sites are shorter or longer than *e.g.*, a typical region targeted by Cas9, for example shorter or longer than 20 nucleotides. In some embodiments, the left and right half sites comprise different nucleic acid sequences. In some embodiments involving inventive nucleases, the target site is a sequence comprising three (3) RNA-guided nuclease target site sequences, for

example, three sequences corresponding to Cas9 target site sequences (See, *e.g.*, Figure 2C), in which the first and second, and second and third Cas9 target site sequences are separated by a spacer sequence. In some embodiments, the spacer sequence is at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at least 17, at least 18, at least 19, at least 20, at least 25, at least 30, at least 35, at least 40, at least 45, at least 50, at least 60, at least 70, at least 80, at least 90, at least 100, at least 125, at least 150, at least 175, at least 200, or at least 250 bp long. In some embodiments, the spacer sequence is between approximately 15 bp and approximately 25 bp long. In some embodiments, the spacer sequence is approximately 15 bp long. In some embodiments, the spacer sequence is approximately 25 bp long.

[0067] The term “Transcriptional Activator-Like Effector,” (TALE) as used herein, refers to bacterial proteins comprising a DNA binding domain, which contains a highly conserved 33-34 amino acid sequence comprising a highly variable two-amino acid motif (Repeat Variable Diresidue, RVD). The RVD motif determines binding specificity to a nucleic acid sequence and can be engineered according to methods known to those of skill in the art to specifically bind a desired DNA sequence (see, *e.g.*, Miller, Jeffrey; et.al. (February 2011). “A TALE nuclease architecture for efficient genome editing”. *Nature Biotechnology* **29** (2): 143–8; Zhang, Feng; *et.al.* (February 2011). “Efficient construction of sequence-specific TAL effectors for modulating mammalian transcription” *Nature Biotechnology* **29** (2): 149–53; Geißler, R.; Scholze, H.; Hahn, S.; Streubel, J.; Bonas, U.; Behrens, S. E.; Boch, J. (2011), Shiu, Shin-Han. ed. “Transcriptional Activators of Human Genes with Programmable DNA-Specificity”. *PLoS ONE* **6** (5): e19509; Boch, Jens (February 2011). “TALEs of genome targeting”. *Nature Biotechnology* **29** (2): 135–6; Boch, Jens; et.al. (December 2009). “Breaking the Code of DNA Binding Specificity of TAL-Type III Effectors”. *Science* **326** (5959): 1509–12; and Moscou, Matthew J.; Adam J. Bogdanove (December 2009). “A Simple Cipher Governs DNA Recognition by TAL Effectors” *Science* **326** (5959): 1501; the entire contents of each of which are incorporated herein by reference). The simple relationship between amino acid sequence and DNA recognition has allowed for the engineering of specific DNA binding domains by selecting a combination of repeat segments containing the appropriate RVDs.

[0068] The term “Transcriptional Activator-Like Element Nuclease,” (TALEN) as used herein, refers to an artificial nuclease comprising a transcriptional activator-like effector DNA binding domain to a DNA cleavage domain, for example, a FokI domain. A number of modular assembly schemes for generating engineered TALE constructs have been reported

(see *e.g.*, Zhang, Feng; *et.al.* (February 2011). “Efficient construction of sequence-specific TAL effectors for modulating mammalian transcription”. *Nature Biotechnology* **29** (2): 149–53; Geißler, R.; Scholze, H.; Hahn, S.; Streubel, J.; Bonas, U.; Behrens, S. E.; Boch, J. (2011), Shiu, Shin-Han. ed. “Transcriptional Activators of Human Genes with Programmable DNA-Specificity”. *PLoS ONE* **6** (5): e19509; Cermak, T.; Doyle, E. L.; Christian, M.; Wang, L.; Zhang, Y.; Schmidt, C.; Baller, J. A.; Somia, N. V. *et al.* (2011). “Efficient design and assembly of custom TALEN and other TAL effector-based constructs for DNA targeting”. *Nucleic Acids Research*; Morbitzer, R.; Elsaesser, J.; Hausner, J.; Lahaye, T. (2011). “Assembly of custom TALE-type DNA binding domains by modular cloning”. *Nucleic Acids Research*; Li, T.; Huang, S.; Zhao, X.; Wright, D. A.; Carpenter, S.; Spalding, M. H.; Weeks, D. P.; Yang, B. (2011). “Modularly assembled designer TAL effector nucleases for targeted gene knockout and gene replacement in eukaryotes”. *Nucleic Acids Research*; Weber, E.; Gruetzner, R.; Werner, S.; Engler, C.; Marillonnet, S. (2011). Bendahmane, Mohammed. ed. “Assembly of Designer TAL Effectors by Golden Gate Cloning”. *PLoS ONE* **6** (5): e19722; the entire contents of each of which are incorporated herein by reference).

[0069] The terms “treatment,” “treat,” and “treating,” refer to a clinical intervention aimed to reverse, alleviate, delay the onset of, or inhibit the progress of a disease or disorder, or one or more symptoms thereof, as described herein. As used herein, the terms “treatment,” “treat,” and “treating” refer to a clinical intervention aimed to reverse, alleviate, delay the onset of, or inhibit the progress of a disease or disorder, or one or more symptoms thereof, as described herein. In some embodiments, treatment may be administered after one or more symptoms have developed and/or after a disease has been diagnosed. In other embodiments, treatment may be administered in the absence of symptoms, *e.g.*, to prevent or delay onset of a symptom or inhibit onset or progression of a disease. For example, treatment may be administered to a susceptible individual prior to the onset of symptoms (*e.g.*, in light of a history of symptoms and/or in light of genetic or other susceptibility factors). Treatment may also be continued after symptoms have resolved, for example, to prevent or delay their recurrence.

[0070] The term “vector” refers to a polynucleotide comprising one or more recombinant polynucleotides of the present invention, *e.g.*, those encoding a Cas9 protein (or fusion thereof) and/or gRNA provided herein. Vectors include, but are not limited to, plasmids, viral vectors, cosmids, artificial chromosomes, and phagemids. The vector is able to replicate in a host cell and is further characterized by one or more endonuclease restriction

sites at which the vector may be cut and into which a desired nucleic acid sequence may be inserted. Vectors may contain one or more marker sequences suitable for use in the identification and/or selection of cells which have or have not been transformed or genomically modified with the vector. Markers include, for example, genes encoding proteins which increase or decrease either resistance or sensitivity to antibiotics (*e.g.*, kanamycin, ampicillin) or other compounds, genes which encode enzymes whose activities are detectable by standard assays known in the art (*e.g.*, β -galactosidase, alkaline phosphatase, or luciferase), and genes which visibly affect the phenotype of transformed or transfected cells, hosts, colonies, or plaques. Any vector suitable for the transformation of a host cell (*e.g.*, *E. coli*, mammalian cells such as CHO cell, insect cells, *etc.*) as embraced by the present invention, for example, vectors belonging to the pUC series, pGEM series, pET series, pBAD series, pTET series, or pGEX series. In some embodiments, the vector is suitable for transforming a host cell for recombinant protein production. Methods for selecting and engineering vectors and host cells for expressing proteins (*e.g.*, those provided herein), transforming cells, and expressing/purifying recombinant proteins are well known in the art, and are provided by, for example, Green and Sambrook, *Molecular Cloning: A Laboratory Manual* (4th ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (2012)).

[0071] The term “zinc finger,” as used herein, refers to a small nucleic acid-binding protein structural motif characterized by a fold and the coordination of one or more zinc ions that stabilize the fold. Zinc fingers encompass a wide variety of differing protein structures (see, *e.g.*, Klug A, Rhodes D (1987). “Zinc fingers: a novel protein fold for nucleic acid recognition”. *Cold Spring Harb. Symp. Quant. Biol.* 52: 473–82, the entire contents of which are incorporated herein by reference). Zinc fingers can be designed to bind a specific sequence of nucleotides, and zinc finger arrays comprising fusions of a series of zinc fingers, can be designed to bind virtually any desired target sequence. Such zinc finger arrays can form a binding domain of a protein, for example, of a nuclease, *e.g.*, if conjugated to a nucleic acid cleavage domain. Different types of zinc finger motifs are known to those of skill in the art, including, but not limited to, Cys₂His₂, Gag knuckle, Treble clef, Zinc ribbon, Zn₂/Cys₆, and TAZ2 domain-like motifs (see, *e.g.*, Krishna SS, Majumdar I, Grishin NV (January 2003). “Structural classification of zinc fingers: survey and summary”. *Nucleic Acids Res.* 31 (2): 532–50). Typically, a single zinc finger motif binds 3 or 4 nucleotides of a nucleic acid molecule. Accordingly, a zinc finger domain comprising 2 zinc finger motifs may bind 6-8 nucleotides, a zinc finger domain comprising 3 zinc finger motifs may bind 9-

12 nucleotides, a zinc finger domain comprising 4 zinc finger motifs may bind 12-16 nucleotides, and so forth. Any suitable protein engineering technique can be employed to alter the DNA-binding specificity of zinc fingers and/or design novel zinc finger fusions to bind virtually any desired target sequence from 3 – 30 nucleotides in length (see, e.g., Pabo CO, Peisach E, Grant RA (2001). “Design and selection of novel cys²His² Zinc finger proteins”. *Annual Review of Biochemistry* 70: 313–340; Jamieson AC, Miller JC, Pabo CO (2003). “Drug discovery with engineered zinc-finger proteins”. *Nature Reviews Drug Discovery* 2 (5): 361–368; and Liu Q, Segal DJ, Ghiara JB, Barbas CF (May 1997). “Design of polydactyl zinc-finger proteins for unique addressing within complex genomes”. *Proc. Natl. Acad. Sci. U.S.A.* 94 (11); the entire contents of each of which are incorporated herein by reference). Fusions between engineered zinc finger arrays and protein domains that cleave a nucleic acid can be used to generate a “zinc finger nuclease.” A zinc finger nuclease typically comprises a zinc finger domain that binds a specific target site within a nucleic acid molecule, and a nucleic acid cleavage domain that cuts the nucleic acid molecule within or in proximity to the target site bound by the binding domain. Typical engineered zinc finger nucleases comprise a binding domain having between 3 and 6 individual zinc finger motifs and binding target sites ranging from 9 base pairs to 18 base pairs in length. Longer target sites are particularly attractive in situations where it is desired to bind and cleave a target site that is unique in a given genome.

[0072] The term “zinc finger nuclease,” as used herein, refers to a nuclease comprising a nucleic acid cleavage domain conjugated to a binding domain that comprises a zinc finger array. In some embodiments, the cleavage domain is the cleavage domain of the type II restriction endonuclease FokI. Zinc finger nucleases can be designed to target virtually any desired sequence in a given nucleic acid molecule for cleavage, and the possibility to design zinc finger binding domains to bind unique sites in the context of complex genomes allows for targeted cleavage of a single genomic site in living cells, for example, to achieve a targeted genomic alteration of therapeutic value. Targeting a double-strand break to a desired genomic locus can be used to introduce frame-shift mutations into the coding sequence of a gene due to the error-prone nature of the non-homologous DNA repair pathway. Zinc finger nucleases can be generated to target a site of interest by methods well known to those of skill in the art. For example, zinc finger binding domains with a desired specificity can be designed by combining individual zinc finger motifs of known specificity. The structure of the zinc finger protein Zif268 bound to DNA has informed much of the work in this field and the concept of obtaining zinc fingers for each of the 64 possible

base pair triplets and then mixing and matching these modular zinc fingers to design proteins with any desired sequence specificity has been described (Pavletich NP, Pabo CO (May 1991). “Zinc finger-DNA recognition: crystal structure of a Zif268-DNA complex at 2.1 Å”. *Science* **252** (5007): 809–17, the entire contents of which are incorporated herein). In some embodiments, separate zinc fingers that each recognizes a 3 base pair DNA sequence are combined to generate 3-, 4-, 5-, or 6-finger arrays that recognize target sites ranging from 9 base pairs to 18 base pairs in length. In some embodiments, longer arrays are contemplated. In other embodiments, 2-finger modules recognizing 6-8 nucleotides are combined to generate 4-, 6-, or 8- zinc finger arrays. In some embodiments, bacterial or phage display is employed to develop a zinc finger domain that recognizes a desired nucleic acid sequence, for example, a desired nuclease target site of 3-30 bp in length. Zinc finger nucleases, in some embodiments, comprise a zinc finger binding domain and a cleavage domain fused or otherwise conjugated to each other via a linker, for example, a polypeptide linker. The length of the linker determines the distance of the cut from the nucleic acid sequence bound by the zinc finger domain. If a shorter linker is used, the cleavage domain will cut the nucleic acid closer to the bound nucleic acid sequence, while a longer linker will result in a greater distance between the cut and the bound nucleic acid sequence. In some embodiments, the cleavage domain of a zinc finger nuclease has to dimerize in order to cut a bound nucleic acid. In some such embodiments, the dimer is a heterodimer of two monomers, each of which comprise a different zinc finger binding domain. For example, in some embodiments, the dimer may comprise one monomer comprising zinc finger domain A conjugated to a FokI cleavage domain, and one monomer comprising zinc finger domain B conjugated to a FokI cleavage domain. In this non-limiting example, zinc finger domain A binds a nucleic acid sequence on one side of the target site, zinc finger domain B binds a nucleic acid sequence on the other side of the target site, and the dimerize FokI domain cuts the nucleic acid in between the zinc finger domain binding sites.

DETAILED DESCRIPTION OF CERTAIN EMBODIMENTS OF THE INVENTION

[0073] Site-specific nucleases and site-specific recombinases are powerful tools for targeted genome modification *in vitro* and *in vivo*. It has been reported that nuclease cleavage in living cells triggers a DNA repair mechanism that frequently results in a modification of the cleaved and repaired genomic sequence, for example, via homologous recombination. Accordingly, the targeted cleavage of a specific unique sequence within a genome opens up new avenues for gene targeting and gene modification in living cells,

including cells that are hard to manipulate with conventional gene targeting methods, such as many human somatic or embryonic stem cells. Another approach utilizes site-specific recombinases, which possess all the functionality required to bring about efficient, precise integration, deletion, inversion, or translocation of specified DNA segments.

[0074] Nuclease-mediated modification of disease-related sequences, *e.g.*, the CCR-5 allele in HIV/AIDS patients, or of genes necessary for tumor neovascularization, can be used in the clinical context, and two site specific nucleases are currently in clinical trials (Perez, E.E. *et al.*, “Establishment of HIV-1 resistance in CD4+ T cells by genome editing using zinc-finger nucleases.” *Nature biotechnology*. 26, 808-816 (2008); ClinicalTrials.gov identifiers: NCT00842634, NCT01044654, NCT01252641, NCT01082926). Accordingly, nearly any genetic disease can be treated using site-specific nucleases and/or recombinases and include, for example, diseases associated with triplet expansion (*e.g.*, Huntington’s disease, myotonic dystrophy, spinocerebellar ataxias, *etc.*), cystic fibrosis (by targeting the *CFTR* gene), hematological disease (*e.g.*, hemoglobinopathies), cancer, autoimmune diseases, and viral infections. Other diseases that can be treated using the inventive compositions and/or methods provided herein include, but are not limited to, achondroplasia, achromatopsia, acid maltase deficiency, adenosine deaminase deficiency, adrenoleukodystrophy, aicardi syndrome, alpha-1 antitrypsin deficiency, alpha-thalassemia, androgen insensitivity syndrome, apert syndrome, arrhythmogenic right ventricular, dysplasia, ataxia telangiectasia, Barth syndrome, beta-thalassemia, blue rubber bleb nevus syndrome, Canavan disease, chronic granulomatous diseases (CGD), cri du chat syndrome, Dercum’s disease, ectodermal dysplasia, Fanconi anemia, fibrodysplasia ossificans progressive, fragile X syndrome, galactosemia, Gaucher’s disease, generalized gangliosidoses (*e.g.*, GM1), hemochromatosis, the hemoglobin C mutation in the 6th codon of beta-globin (HbC), hemophilia, Hurler Syndrome, hypophosphatasia, Klinefelter syndrome, Krabbes Disease, Langer-Giedion Syndrome, leukocyte adhesion deficiency (LAD), leukodystrophy, long QT syndrome, Marfan syndrome, Moebius syndrome, mucopolysaccharidosis (MPS), nail patella syndrome, nephrogenic diabetes insipidus, neurofibromatosis, Neimann-Pick disease, osteogenesis imperfecta, porphyria, Prader-Willi syndrome, progeria, Proteus syndrome, retinoblastoma, Rett syndrome, Rubinstein-Taybi syndrome, Sanfilippo syndrome, severe combined immunodeficiency (SCID), Shwachman syndrome, sickle cell disease (sickle cell anemia), Smith-Magenis syndrome, Stickler syndrome, Tay-Sachs disease, Thrombocytopenia Absent Radius (TAR) syndrome, Treacher Collins syndrome, trisomy, tuberous sclerosis, Turner’s syndrome, urea cycle disorder, von Hippel-Landau disease,

Waardenburg syndrome, Williams syndrome, Wilson's disease, Wiskott-Aldrich syndrome, and X-linked lymphoproliferative syndrome (XLP).

[0075] One aspect of site-specific genomic modification is the possibility of off-target nuclease or recombinase effects, *e.g.*, the cleavage or recombination of genomic sequences that differ from the intended target sequence by one or more nucleotides. Undesired side effects of off-target cleavage/recombination range from insertion into unwanted loci during a gene targeting event to severe complications in a clinical scenario. Off-target cleavage or recombination of sequences encoding essential gene functions or tumor suppressor genes by an endonuclease or recombinase administered to a subject may result in disease or even death of the subject. Accordingly, it is desirable to employ new strategies in designing nucleases and recombinases having the greatest chance of minimizing off-target effects.

[0076] The methods and compositions of the present disclosure represent, in some aspects, an improvement over previous methods and compositions providing nucleases (and methods of their use) and recombinases (and methods of their use) engineered to have improved specificity for their intended targets. For example, nucleases and recombinases known in the art, both naturally occurring and those engineered, typically have a target (*e.g.*, DNA) binding domain that recognizes a particular sequence. Additionally, known nucleases and recombinases may comprise a DNA binding domain and a catalytic domain in a single protein capable of inducing cleavage or recombination, and as such the chance for off-target effects are increased as cleavage or recombination likely occurs upon off-target binding of the nuclease or recombinase, respectively. Aspects of the present invention relate to the recognition that increasing the number of sequences (*e.g.*, having a nuclease bind at more than one site at a desired target), and/or splitting the activities (*e.g.*, target binding and target cleaving) of a nuclease between two or more proteins, will increase the specificity of a nuclease and thereby decrease the likelihood of off-target effects. Other aspects of the present invention relate to the recognition that fusions between the catalytic domain of recombinases (or recombinases having inactive DNA binding domains) and nuclease-inactivated RNA-programmable nucleases allow for the targeted recombination of DNA at any location.

[0077] In the context of site-specific nucleases, the strategies, methods, compositions, and systems provided herein can be utilized to improve the specificity of any site-specific nuclease, for example, variants of the Cas9 endonuclease, Zinc Finger Nucleases (ZFNs) and Transcription Activator-Like Effector Nucleases (TALENs). Suitable nucleases for

modification as described herein will be apparent to those of skill in the art based on this disclosure.

[0078] In certain embodiments, the strategies, methods, compositions, and systems provided herein are utilized to improve the specificity of the RNA-guided (*e.g.*, RNA-programmable) endonuclease Cas9. Whereas typical endonucleases recognize and cleave a single target sequence, Cas9 endonuclease uses RNA:DNA hybridization to determine target DNA cleavage sites, enabling a single monomeric protein to cleave, in principle, any sequence specified by the guide RNA (gRNA). While Cas9:guide RNA complexes have been successfully used to modify both cells (Cong, L. *et al.* Multiplex genome engineering using CRISPR/Cas systems. *Science*. 339, 819-823 (2013); Mali, P. *et al.* RNA-guided human genome engineering via Cas9. *Science*. 339, 823-826 (2013); Jinek, M. *et al.* RNA-programmed genome editing in human cells. *eLife* 2, e00471 (2013)) and organisms (Hwang, W.Y. *et al.* Efficient genome editing in zebrafish using a CRISPR-Cas system. *Nature Biotechnology*. 31, 227-229 (2013)), a study using Cas9:guide RNA complexes to modify zebrafish embryos observed toxicity (*e.g.*, off-target effects) at a rate similar to that of ZFNs and TALENs (Hwang, W.Y. *et al.* *Nature Biotechnology*. 31, 227-229 (2013)). Further, while recently engineered variants of Cas9 that cleave only one DNA strand (“nickases”) enable double-stranded breaks to be specified by two distinct gRNA sequences (Cho, S. W. *et al.* Analysis of off-target effects of CRISPR/Cas-derived RNA-guided endonucleases and nickases. *Genome Res.* **24**, 132–141 (2013); Ran, F. A. *et al.* Double Nicking by RNA-Guided CRISPR Cas9 for Enhanced Genome Editing Specificity. *Cell* **154**, 1380–1389 (2013); Mali, P. *et al.* CAS9 transcriptional activators for target specificity screening and paired nickases for cooperative genome engineering. *Nat. Biotechnol.* **31**, 833–838 (2013)), these variants still suffer from off-target cleavage activity (Ran, F. A. *et al.* Double Nicking by RNA-Guided CRISPR Cas9 for Enhanced Genome Editing Specificity. *Cell* **154**, 1380–1389 (2013); Fu, Y., *et al.*, Improving CRISPR-Cas nuclease specificity using truncated guide RNAs. *Nat. Biotechnol.* (2014)) arising from the ability of each monomeric nickase to remain active when individually bound to DNA (Cong, L. *et al.* Multiplex Genome Engineering Using CRISPR/Cas Systems. *Science* **339**, 819–823 (2013); Jinek, M. *et al.* *Science* **337**, 816–821 (2012); Gasiunas, G., *et al.*, Cas9-crRNA ribonucleoprotein complex mediates specific DNA cleavage for adaptive immunity in bacteria. *Proc. Natl. Acad. Sci.* **109**, E2579–E2586 (2012). Accordingly, aspects of the present disclosure aim at reducing the chances for Cas9 off-target effects using novel engineered Cas9 variants. In one example, a Cas9 variant (*e.g.*, fCas9) is provided which has improved specificity as compared to the

Cas9 nickases or wild type Cas9, exhibiting, *e.g.*, >10-fold, >50-fold, >100-fold, >140-fold, > 200-fold, or more, higher specificity than wild type Cas9 (see *e.g.*, the Examples).

[0079] Other aspects of the present disclosure provide strategies, methods, compositions, and systems utilizing inventive RNA-guided (*e.g.*, RNA-programmable) Cas9-recombinase fusion proteins. Whereas typical recombinases recognize and recombine distinct target sequences, the Cas9-recombinase fusions provided herein use RNA:DNA hybridization to determine target DNA recombination sites, enabling the fusion proteins to recombine, in principle, any region specified by the gRNA(s).

[0080] While of particular relevance to DNA and DNA-cleaving nucleases and/or recombinases, the inventive concepts, methods, strategies and systems provided herein are not limited in this respect, but can be applied to any nucleic acid:nuclease or nucleic acid:recombinase system.

Nucleases

[0081] Some aspects of this disclosure provide site-specific nucleases with enhanced specificity that are designed using the methods and strategies described herein. Some embodiments of this disclosure provide nucleic acids encoding such nucleases. Some embodiments of this disclosure provide expression constructs comprising such encoding nucleic acids (See, *e.g.*, Figure 20). For example, in some embodiments an isolated nuclease is provided that has been engineered to cleave a desired target site within a genome. In some embodiments, the isolated nuclease is a variant of an RNA-programmable nuclease, such as a Cas9 nuclease.

[0082] In one embodiment, fusion proteins are provided comprising two domains: (i) an RNA-programmable nuclease (*e.g.*, Cas9 protein, or fragment thereof) domain fused or linked to (ii) a nuclease domain. For example, in some aspects, the Cas9 protein (*e.g.*, the Cas9 domain of the fusion protein) comprises a nuclease-inactivated Cas9 (*e.g.*, a Cas9 lacking DNA cleavage activity; “dCas9”) that retains RNA (gRNA) binding activity and is thus able to bind a target site complementary to a gRNA. In some aspects, the nuclease fused to the nuclease-inactivated Cas9 domain is any nuclease requiring dimerization (*e.g.*, the coming together of two monomers of the nuclease) in order to cleave a target nucleic acid (*e.g.*, DNA). In some embodiments, the nuclease fused to the nuclease-inactivated Cas9 is a monomer of the FokI DNA cleavage domain, *e.g.*, thereby producing the Cas9 variant referred to as fCas9. The FokI DNA cleavage domain is known, and in some aspects corresponds to amino acids 388–583 of FokI (NCBI accession number J04623). In some

embodiments, the FokI DNA cleavage domain corresponds to amino acids 300-583, 320-583, 340-583, or 360-583 of FokI. See also Wah *et al.*, “Structure of FokI has implications for DNA cleavage” *Proc. Natl. Acad. Sci. USA*. 1998; 1;95(18):10564-9; Li *et al.*, “TAL nucleases (TALNs): hybrid proteins composed of TAL effectors and FokI DNA-cleavage domain” *Nucleic Acids Res.* 2011; 39(1):359-72; Kim *et al.*, “Hybrid restriction enzymes: zinc finger fusions to Fok I cleavage domain” *Proc. Natl Acad. Sci. USA*. 1996; 93:1156–1160; the entire contents of each are herein incorporated by reference). In some embodiments, the FokI DNA cleavage domain corresponds to, or comprises in part or whole, the amino acid sequence set forth as SEQ ID NO:6. In some embodiments, the FokI DNA cleavage domain is a variant of FokI (*e.g.*, a variant of SEQ ID NO:6), as described herein.

[0083] In some embodiments, a dimer of the fusion protein is provided, *e.g.*, dimers of fCas9. For example, in some embodiments, the fusion protein forms a dimer with itself to mediate cleavage of the target nucleic acid. In some embodiments, the fusion proteins, or dimers thereof, are associated with one or more gRNAs. In some aspects, because the dimer contains two fusion proteins, each having a Cas9 domain having gRNA binding activity, a target nucleic acid is targeted using two distinct gRNA sequences that complement two distinct regions of the nucleic acid target. See, *e.g.*, Figures 1A, 6D. Thus, in this example, cleavage of the target nucleic acid does not occur until both fusion proteins bind the target nucleic acid (*e.g.*, as specified by the gRNA:target nucleic acid base pairing), and the nuclease domains dimerize (*e.g.*, the FokI DNA cleavage domains; as a result of their proximity based on the binding of the Cas9:gRNA domains of the fusion proteins) and cleave the target nucleic acid, *e.g.*, in the region between the bound Cas9 fusion proteins (the “spacer sequence”). This is exemplified by the schematics shown in Figures 1A and 6D. This approach represents a notable improvement over wild type Cas9 and other Cas9 variants, such as the nickases (Ran *et al.* Double Nicking by RNA-Guided CRISPR Cas9 for Enhanced Genome Editing Specificity. *Cell* **154**, 1380–1389 (2013); Mali *et al.* CAS9 transcriptional activators for target specificity screening and paired nickases for cooperative genome engineering. *Nat. Biotechnol.* **31**, 833–838 (2013)), which do not require the dimerization of nuclease domains to cleave a nucleic acid. These nickase variants, as described in the Examples, can induce cleaving, or nicking upon binding of a single nickase to a nucleic acid, which can occur at on- and off-target sites, and nicking is known to induce mutagenesis. An exemplary nucleotide encoding a Cas9 nickase (SEQ ID NO:7) and an exemplary amino acid sequence of Cas9 nickase (SEQ ID NO:8) are provided below. As the variants provided herein require the binding of two Cas9 variants in proximity to one another

to induce target nucleic acid cleavage, the chances of inducing off-target cleavage is reduced. See, *e.g.*, the Examples. For example, in some embodiments, a Cas9 variant fused to a nuclease domain (*e.g.*, fCas9) has an on-target:off-target modification ratio that is at least 2-fold, at least 5-fold, at least 10-fold, at least 20-fold, at least 30-fold, at least 40-fold, at least 50-fold, at least 60-fold, at least 70-fold, at least 80-fold, at least 90-fold, at least 100-fold, at least 110-fold, at least 120-fold, at least 130-fold, at least 140-fold, at least 150-fold, at least 175-fold, at least 200-fold, at least 250-fold, or more higher than the on-target:off-target modification ratio of a wild type Cas9 or other Cas9 variant (*e.g.*, nickase). In some embodiments, a Cas9 variant fused to a nuclease domain (*e.g.*, fCas9) has an on-target:off-target modification ratio that is between about 60- to 180-fold, between about 80- to 160-fold, between about 100- to 150-fold, or between about 120- to 140-fold higher than the on-target:off-target modification ratio of a wild type Cas9 or other Cas9 variant. Methods for determining on-target:off-target modification ratios are known, and include those described in the Examples. In certain embodiments, the on-target:off-target modification ratios are determined by measuring the number or amount of modifications of known Cas9 off-target sites in certain genes. For example, the Cas9 off-target sites of the *CLTA*, *EMX*, and *VEGF* genes are known, and modifications at these sites can be measured and compared between test proteins and controls. The target site and its corresponding known off-target sites (see, *e.g.*, Table 5 for *CLTA*, *EMX*, and *VEGF* off-target sites) are amplified from genomic DNA isolated from cells (*e.g.*, HEK293) treated with a particular Cas9 protein or variant. The modifications are then analyzed by high-throughput sequencing. Sequences containing insertions or deletions of two or more base pairs in potential genomic off-target sites and present in significantly greater numbers (P value < 0.005, Fisher's exact test) in the target gRNA-treated samples versus the control gRNA-treated samples are considered Cas9 nuclease-induced genome modifications.

[0084] Cas9 nickase (nucleotide sequence):

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ATGGACTATAAGGACCACGACGGAGACTACAAGGATCATGATATTGATTACAAAGACGATGACGATAAGATGGCC
CCAAAGAAGAAGCGGAAGGTCGGTATCCACGGAGTCCCAGCAGCCGATAAAAAGTATTCTATTGGTTTAGCTATC
GGCACTAATTCCGTTGGATGGGCTGTCATAACCGATGAATACAAAGTACCTTCAAAGAAATTTAAGGTGTTGGGG
AACACAGACCGTCATTTCGATTAAAAAGAATCTTATCGGTGCCCTCCTATTTCGATAGTGGCGAAACGGCAGAGGCG
ACTCGCCTGAAACGAACCGCTCGGAGAAGGTATACACGTCGCAAGAACCGAATATGTTACTTACAAGAAATTTTT
AGCAATGAGATGGCCAAAGTTGACGATTCTTTCTTTTACCGTTTGGGAAGAGTCCTTCCTTGTCGAAGAGGACAAG
AAACATGAACGGCACCCCATCTTTGGAAACATAGTAGATGAGGTGGCATATCATGAAAAGTACCCAACGATTTAT
CACCTCAGAAAAAGCTAGTTGACTCAACTGATAAAGCGGACCTGAGGTTAATCTACTTGGCTCTTGCCCATATG
ATAAAGTTCCGTGGGCACTTTCTCATTGAGGGTGATCTAAATCCGGACAACCTCGGATGTGACAAACTGTTTCATC
CAGTTAGTACAAACCTATAATCAGTTGTTTGAAGAGAACCCTATAAATGCAAGTGGCGTGGATGCGAAGGCTATT
CTTAGCGCCCGCCTCTCTAAATCCCGACGGCTAGAAAACCTGATCGCACAATTACCCGGAGAGAAGAAAAATGGG
TTGTTTCGGTAACCTTATAGCGCTCTCACTAGGCCTGACACCAAATTTTAAGTCGAACTTCGACTTAGCTGAAGAT
```

GCCAAATTGCAGCTTAGTAAGGACACGTACGATGACGATCTCGACAATCTACTGGCACAAATTGGAGATCAGTAT
 GCGGACTTATTTTTGGCTGCCAAAACCTTAGCGATGCAATCCTCCTATCTGACATACTGAGAGTTAATACTGAG
 ATTACCAAGGCGCCGTTATCCGCTTCAATGATCAAAAGGTACGATGAACATCACCAAGACTTGACACTTCTCAAG
 GCCCTAGTCCGTCAGCAACTGCCTGAGAAATATAAGGAAATATTCTTTGATCAGTCGAAAAACGGGTACGCAGGT
 TATATTGACGGCGGAGCGAGTCAAGAGGAATTCTACAAGTTTATCAAACCCATATTAGAGAAGATGGATGGGACG
 GAAGAGTTGCTTGTAAACTCAATCGCGAAGATCTACTGCGAAAGCAGCGGACTTTTCGACAACGGTAGCATTCCA
 CATCAAATCCACTTAGGGCAATTGCATGCTATACTTAGAAGGCAGGAGGATTTTTATCCGTTCTCAAAGACAAT
 CGTGAAAAGATTGAGAAAATCCTAACCTTTTCGCATACCTTACTATGTGGGACCCCTGGCCCGAGGGAACTCTCGG
 TTCGCATGGATGACAAGAAAGTCCGAAGAAACGATTACTCCATGGAATTTTGAGGAAGTTGTCGATAAAGGTGCG
 TCAGCTCAATCGTTCATCGAGAGGATGACCAACTTTGACAAGAATTTACCGAACGAAAAAGTATTGCCTAAGCAC
 AGTTTACTTTACGAGTATTTACAGTGTACAATGAACTCACGAAAGTTAAGTATGTCACTGAGGGCATGCGTAAA
 CCCGCCTTTCTAAGCGGAGAACAGAAGAAAGCAATAGTAGATCTGTTATTCAAGACCAACCGCAAAGTGACAGTT
 AAGCAATTGAAAGAGGACTACTTTAAGAAAATTGAATGCTTCGATTCTGTGCGAGATCTCCGGGGTAGAAGATCGA
 TTTAATGCGTCACTTGGTACGTATCATGACCTCCTAAAGATAATTAAAGATAAGGACTTCCTGGATAACGAAGAG
 AATGAAGATATCTTAGAAGATATAGTGTTGACTCTTACCCTCTTTGAAGATCGGGAAATGATTGAGGAAAGACTA
 AAAACATACGCTCACCTGTTTCGACGATAAGGTTATGAAACAGTTAAAGAGGCGTCGCTATACGGGCTGGGGACGA
 TTGTGCGGGAAACTTATCAACGGGATAAGAGACAAGCAAAGTGGTAAACTATTCTCGATTTTCTAAAGAGCGAC
 GGCTTCGCCAATAGGAACTTTATGCAGCTGATCCATGATGACTCTTTAACCTTCAAAGAGGATATACAAAAGGCA
 CAGGTTTCCGGACAAGGGGACTCATTGCACGAACATATTGCGAATCTTGCTGGTTTCGCCAGCCATCAAAAAGGGC
 ATACTCCAGACAGTCAAAGTAGTGATGAGCTAGTTAAGGTCATGGGACGTCACAAACCGGAAAACATTGTAATC
 GAGATGGCACGCGAAAATCAAACGACTCAGAAGGGGCAAAAAACAGTCGAGAGCGGATGAAGAGAATAGAAGAG
 GGTATTAAAGAACTGGGCAGCCAGATCTTAAAGGAGCATCCTGTGGAAAATACCCAATTGCAGAACGAGAACTT
 TACCTCTATTACCTACAAAATGGAAGGGACATGTATGTTGATCAGGAACCTGGACATAAACCGTTTTATCTGATTAC
 GACGTCGATCACATTGTACCCCAATCCTTTTTTGAAGGACGATTCAATCGACAATAAAGTGCTTACACGCTCGGAT
 AAGAACCGAGGGAAAAGTGACAATGTTCCAAGCGAGGAAGTCGTAAAGAAAATGAAGAACTATTGGCGGCAGCTC
 CTAAATGCGAACTGATAACGCAAAGAAAGTTTCGATAACTTAACTAAAGCTGAGAGGGGTGGCTTGTCTGAACCTT
 GACAAGGCCGGATTTATTAAACGTCAGCTCGTGGAACCCGCCAAATCACAAAGCATGTTGCACAGATACTAGAT
 TCCCGAATGAATACGAAATACGACGAGAACGATAAGCTGATTCGGGAAGTCAAAGTAATCACTTTAAAGTCAAAA
 TTGGTGTGCGGACTTCAGAAAGGATTTTCAATTCTATAAAGTTAGGGAGATAAATAACTACCACCATGCGCACGAC
 GCTTATCTTAATGCCGTCGTAGGGACCGCACTCATTAAAGAAATACCCGAAGCTAGAAAGTGAGTTTGTGTATGGT
 GATTACAAAGTTTATGACGTCCGTAAGATGATCGCGAAAAGCGAACAGGAGATAGGCAAGGCTACAGCCAAATAC
 TTCTTTTATTCTAACATTATGAATTTCTTTAAGACGGAAATCACTCTGGCAAACGGAGAGATACGCAAACGACCT
 TTAATTGAAACCAATGGGGAGACAGGTGAAATCGTATGGGATAAGGGCCGGGACTTCGCGACGGTGAGAAAAGTT
 TTGTCCATGCCCCAAGTCAACATAGTAAAGAAAAGTTGAGGTGCAGACCGGAGGGTTTTTCAAAGGAATCGATTCTT
 CAAAAAGGAATAGTGATAAGCTCATCGCTCGTAAAAAGGACTGGGACCCGAAAAAGTACGGTGGCTTCGATAGC
 CCTACAGTTGCCTATTCTGTCTTAGTAGTGCCAAAAGTTGAGAAGGGGAAAATCCAAGAACTGAAGTCAGTCAAA
 GAATTATTGGGGATAACGATTATGGAGCGCTCGTCTTTTGAAAAGAACCCCATCGACTTCCTTGAGGCGAAAGGT
 TACAAGGAAGTAAAAAAGGATCTCATAATTAACTACCAAAGTATAGTCTGTTTGAGTTAGAAAATGGCCGAAAA
 CGGATGTTGGCTAGCGCCGGAGAGCTTCAAAGGGGAACGAACCTCGCACTACCGTCTAAATACGTGAATTTCTTG
 TATTTAGCGTCCCATTACGAGAAGTTGAAAGGTTACCTGAAGATAACGAACAGAAGCAACTTTTTTGTGAGCAG
 CACAAACATTATCTCGACGAAATCATAGAGCAAATTTTCGGAATTCAGTAAGAGAGTCATCCTAGCTGATGCCAAT
 CTGGACAAAGTATTAAGCGCATAACAAGCACAGGGATAAACCCATACGTGAGCAGGCGGAAAAATATTATCCAT
 TTGTTTACTCTTACCAACCTCGGCGCTCCAGCCGCATTCAAGTATTTTGACACAACGATAGATCGCAAACGATAC
 ACTTCTACCAAGGAGGTGCTAGACGCGACACTGATTCACCAATCCATCACGGGATTATATGAAACTCGGATAGAT
 TTGTCACAGCTTGGGGGTGAC (SEQ ID NO:7)

[0085] Cas9 nickase (D10A)(amino acid sequence):

MDKKYSIGLAIGTNSVGWAVITDEYKVPSKKFKVLGNTDRHSIKKNLIGALLFDSGETAEATRLKRTARRRYTRR
 KNRICYLQEIFSNEMAKVDDSFHRLSEESFLVEEDKKHERHPIFGNIVDEVAYHEKYPTIYHLRKKLVDSTDKAD
 LRLIYLALAHMIKFRGHFLIEGDLNPDNSDVKLFIQLVQTYNQLFEEENPINASGVDAKAILSARLSKSRLENL
 IAQLPGEKKNGLFGNLIALSLGLTPNFKSNFDLAEDAKLQLSKDITYDDDLNLLAQIGDQYADLFLAAKNLSDAI
 LLSDILRVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQLPEKYKEIFFDQSKNGYAGYIDGGASQEEFYKF
 IKPILEKMDGTEELLVKLNREDLLRKQRTFDNGSIPHQIHLGELHAILRRQEDFYFPLKDNREKIEKILTFRIPY
 YVGPLARGNSRFAWMTRKSEETITPWNFEVVDKGASAQSFIERMTNFDKNLPNEKVLPHKSLLYEYFTVYNELT
 KVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTQKQLKEDYFKKIECFDSVEISGVEDRFNASLGTYHDLLKI
 IKDKDFLDNEENEDILEDIVLTTLTFEDREMIEERLKYAHLFDDKVMKQLKRRRYTGWGRLSRKLINGIRDKQS
 GKTILDFLKSDGFANRNFMLIHDDSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPAIIKKGILQTVKVVDDELVKV
 MGRHKPENIVIEARENQTTQKGQKNSRERMKRIEEGIKELGSQILKEHPVENTQLQNEKLYLYYLQNGRDMYVD
 QELDINRLSDYDVDHIVPQSFLKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKKMKNYWRQLLNAKLITQRKFDNL

TKAERGGLSELDKAGFIKRQLVETRQITKHVAQILD SRMNTKYDENDKLIREVKVITLKS KLVSDFRKDFQFYKV
 REINNYHHAHDAYLNAVVG TALIKKYPKLESEFVYGDYKVYDVRKMI AKSEQEIGKATAKYFFYSNIMNFFKTEI
 T LANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVL SMPQVNIVKKTEVQTGGFSKESILPKRNSDKLIARKKD
 WDPKKYGGFDSPTVAYSVLV VAKVEKGKSKKLKSVKELLGITIMERS SFEKNPIDFLEAKGYKEVKKDLIIKLPK
 YSLFELENGRKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLKGSPEDNEQKQLFVEQHKHYLDEIIIEQISE
 FSKRVILADANLDKVL SAYNKH RDKPIREQAENI IHLFTLTNLGAPAAFKYFDTTIDRKRYTSTKEVLDATLIHQ
 SITGLYETRIDLSQLGGD (SEQ ID NO:8)

[0086] In some embodiments, the gRNAs which bind the Cas9 variants (*e.g.*, fCas9) can be oriented in one of two ways, with respect to the spacer sequence, deemed the “A” and “B” orientations. In orientation A, the region of the gRNAs that bind the PAM motifs is distal to the spacer sequence with the 5' ends of the gRNAs adjacent to the spacer sequence (Figure 6C); whereas in orientation B, the region of the gRNAs that bind the PAM motifs is adjacent to the spacer sequence (Figure 9). In some embodiments, the gRNAs are engineered or selected to bind (*e.g.*, as part of a complex with a Cas9 variant, such as fCas9) to a target nucleic acid in the A or B orientation. In some embodiments, the gRNAs are engineered or selected to bind (*e.g.*, as part of a complex with a Cas9 variant such as fCas9) to a target nucleic acid in the A orientation. In some embodiments, the gRNAs are engineered or selected to bind (*e.g.*, as part of a complex with a Cas9 variant, such as fCas9) to a target nucleic acid in the B orientation.

[0087] In some embodiments, the domains of the fusion protein are linked via a linker *e.g.*, as described herein. In certain embodiments, the linker is a peptide linker. In other embodiments, the linker is a non-peptidic linker. In some embodiments, a functional domain is linked via a peptide linker (*e.g.*, fused) or a non-peptidic linker to an inventive fusion protein. In some embodiments, the functional domain is a nuclear localization signal (NLS) domain. An NLS domain comprises an amino acid sequence that “tags” or signals a protein for import into the cell nucleus by nuclear transport. Typically, this signal consists of one or more short sequences of positively charged lysines or arginines exposed on the protein surface. NLS sequences are well known in the art (See *e.g.*, Lange et al., “Classical nuclear localization signals: definition, function, and interaction with importin alpha.” *J Biol Chem.* 2007 Feb 23;282(8):5101-5; the entire contents of which is hereby incorporated by reference), and include, for example those described in the Examples section. In some embodiments, the NLS sequence comprises, in part or in whole, the amino acid sequence MAPKKKRKVG IHRGVP (SEQ ID NO:318). The domains (*e.g.*, two or more of a gRNA binding domain (dCas9 domain), a catalytic nuclease domain, and a NLS domain) associated via a linker can be linked in any orientation or order. For example, in some embodiments, any domain can be at the N-terminus, the C-terminus, or in between the domains at the N-

and C-termini of the fusion protein. In some embodiments, the orientation or order of the domains in an inventive fusion protein are as provided in Figure 6B. In some embodiments, wherein the fusion protein comprises three domains (*e.g.*, a gRNA binding domain (*e.g.*, dCas9 domain), a nuclease domain (*e.g.*, FokI), and an NLS domain), each domain is connected via a linker, as provided herein. In some embodiments, the domains are not connected via a linker. In some embodiments, one or more of the domains is/are connected via a linker.

[0088] In some embodiments, an inventive fusion protein (*e.g.*, fCas9) comprising three domains (*e.g.*, a gRNA binding domain (*e.g.*, dCas9 domain), a nuclease domain (*e.g.*, FokI), and an NLS domain) is encoded by a nucleotide sequence (or fragment or variant thereof) set forth as SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, SEQ ID NO:12, or SEQ ID NO:319, as shown below.

[0089] fCas9 (*e.g.*, dCas9-NLS-GGS3linker-FokI):

ATGGATAAAAAGTATTCTATTGGTTTAGCTATCGGCACCTAATTCCGTTGGATGGGCTGTCATAACCGATGAATAC
AAAGTACCTTCAAAGAAATTTAAGGTGTTGGGGAACACAGACCGTCATTTCGATTAAAAAGAATCTTATCGGTGCC
CTCCTATTTCGATAGTGGCGAAACGGCAGAGGCGACTCGCCTGAAACGAACCGCTCGGAGAAGGTATACACGTCGC
AAGAACCGAATATGTTACTTACAAGAAATTTTGTAGCAATGAGATGGCCAAAGTTGACGATTCTTTCTTTTACCGT
TTGGAAGAGTCCTTCCTTGTCGAAGAGGACAAGAAACATGAACGGCACCCCATCTTTGGAAACATAGTAGATGAG
GTGGCATATCATGAAAAGTACCCAACGATTTATCACCTCAGAAAAAGCTAGTTGACTCAACTGATAAAGCGGAC
CTGAGGTTAATCTACTTGGCTCTTGCCCATATGATAAAGTTCCGTGGGCACTTTCTCATTGAGGGTGATCTAAAT
CCGGACAACCTCGGATGTCGACAACTGTTTCATCCAGTTAGTACAAACCTATAATCAGTTGTTTGAAGAGAACCCT
ATAAATGCAAGTGGCGTGGATGCGAAGGCTATTCTTAGCGCCCGCCTCTCTAAATCCCGACGGCTAGAAAACCTG
ATCGCACAATTACCCGGAGAGAAGAAAAATGGGTTGTTTCGGTAACCTTATAGCGCTCTCACTAGGCCTGACACCA
AATTTTAAGTCGAACTTCGACTTAGCTGAAGATGCCAAATTGCAGCTTAGTAAGGACACGTACGATGACGATCTC
GACAATCTACTGGCACAATTTGGAGATCAGTATGCGGACTTATTTTTGGCTGCCAAAACCTTAGCGATGCAATC
CTCCTATCTGACATACTGAGAGTTAATACTGAGATTACCAAGGCGCCGTTATCCGCTTCAATGATCAAAAGGTAC
GATGAACATCACCAAGACTTGACACTTCTCAAGGCCCTAGTCCGTCAGCAACTGCCTGAGAAATATAAGGAAATA
TTCTTTGATCAGTCGAAAAACGGGTACGCAGGTTATATTGACGGCGGAGCGAGTCAAGAGGAATTCTACAAGTTT
ATCAAACCCATATTAGAGAAGATGGATGGGACGGAAGAGTTGCTTGTAAACTCAATCGCGAAGATCTACTGCGA
AAGCAGCGGACTTTTCGACAACGGTAGCATTCACATCAAATCCACTTAGGCGAATTGCATGCTATACTTAGAAGG
CAGGAGGATTTTTATCCGTTCTCAAAGACAATCGTGAAAAGATTGAGAAAATCCTAACCTTTTCGCATACCTTAC
TATGTGGGACCCCTGGCCCGAGGGAACCTCTCGGTTTCGCATGGATGACAAGAAAGTCCGAAGAAACGATTACTCCA
TGGAATTTTGAGGAAGTTGTCGATAAAGGTGCGTCAGCTCAATCGTTTCATCGAGAGGATGACCAACTTTGACAAG
AATTTACCGAACGAAAAAGTATTGCCCTAAGCACAGTTTACTTTACGAGTATTTACAGTGTACAATGAACTCACG
AAAGTTAAGTATGTCAGTGGGTCATGCGTAAACCCGCCTTTCTAAGCGGAGAACAGAAGAAAGCAATAGTAGAT
CTGTTATTCAAGACCAACCGCAAAGTGACAGTTAAGCAATTGAAAGAGGACTACTTTAAGAAAATTGAATGCTTC
GATTCTGTGTCGAGATCTCCGGGGTAGAAGATCGATTTAATGCGTCACCTTGGTACGTATCATGACCTCCTAAAGATA
ATTAAAGATAAGGACTTCCTGGATAACGAAGAGAATGAAGATATCTTAGAAGATATAGTGTTGACTCTTACCCTC
TTTGAAGATCGGGAAATGATTGAGGAAAGACTAAAAACATACGCTCACCTGTTTCGACGATAAGGTTATGAAACAG
TTAAAGAGGCGTCGCTATACGGGCTGGGGACGATTGTGCGCGGAAACTTATCAACGGGATAAGAGACAAGCAAAGT
GGTAAACTATTCTCGATTTTCTAAAGAGCGACGGCTTCGCCAATAGGAACTTTATGCAGCTGATCCATGATGAC
TCTTTAACCTTCAAAGAGGATATACAAAAGGCACAGGTTTCCGGACAAGGGGACTCATTGCACGAACATATTGCG
AATCTTGCTGGTTTCGCCAGCCATCAAAAAGGGCATACTCCAGACAGTCAAAGTAGTGGATGAGCTAGTTAAGGTC
ATGGGACGTCACAAACCGGAAAACATTGTAATCGAGATGGCACGCGAAAATCAAACGACTCAGAAGGGGCAAAAA
AACAGTCGAGAGCGGATGAAGAGAATAGAAGAGGGTATTAAAGAACTGGGCAGCCAGATCTTAAAGGAGCATCCT
GTGGAATAATACCAATTGCAGAACGAGAACTTTACCTCTATTACCTACAAAATGGAAGGGACATGTATGTTGAT
CAGGAACTGGACATAAACCGTTTATCTGATTACGACGTCGATGCCATTGTACCCCAATCCTTTTTTGAAGGACGAT
TCAATCGACAATAAAGTGCTTACACGCTCGGATAAGAACCGAGGGAAAAGTGACAATGTTCCAAGCGAGGAAGTC
GTAAAGAAAATGAAGAACTATTGGCGGCAGCTCCTAAATGCGAACTGATAACGCAAAGAAAGTTTCGATAACTTA

ACTAAAGCTGAGAGGGGTGGCTTGTCTGAACTTGACAAGGCCGGATTTATTAAACGTCAGCTCGTGGAACCCGC
CAAATCACAAAGCATGTTGCACAGATACTAGATTCCCGAATGAATACGAAATACGACGAGAACGATAAGCTGATT
CGGGAAGTCAAAGTAATCACTTTAAAGTCAAATTTGGTGTGCGGACTTCAGAAAGGATTTTCAATTCTATAAAGTT
AGGGAGATAAATAACTACCACCATGCGCACGACGCTTATCTTAATGCCGTCGTAGGGACCGCACTCATTAAAGAA
TACCCGAAGCTAGAAAGTGAGTTTGTGTATGGTGATTACAAAGTTTATGACGTCCGTAAGATGATCGCGAAAAGC
GAACAGGAGATAGGCAAGGCTACAGCCAAATACTTCTTTTTATTCTAACATTATGAATTTCTTTAAGACGGAAATC
ACTCTGGCAAACGGAGAGATACGCAAACGACCTTTAATTGAAACCAATGGGGAGACAGGTGAAATCGTATGGGAT
AAGGGCCGGGACTTCGCGACGGTGAGAAAAGTTTTGTCCATGCCCCAAGTCAACATAGTAAAGAAAACCTGAGGTG
CAGACCGGAGGGTTTTCAAAGGAATCGATTCTTCCAAAAAGGAATAGTGATAAGCTCATCGCTCGTAAAAAGGAC
TGGGACCCGAAAAAGTACGGTGGCTTCGATAGCCCTACAGTTGCCTATTCTGTCTTAGTAGTGCCAAAAGTTGAG
AAGGGAAAATCCAAGAACTGAAGTCAGTCAAAGAATTATTGGGGATAACGATTATGGAGCGCTCGTCTTTTGAA
AAGAACCCCATCGACTTCCTTGAGGCGAAAGGTTACAAGGAAGTAAAAAAGGATCTCATAATTAACTACCAAAG
TATAGTCTGTTTGAGTTAGAAAATGGCCGAAAACGGATGTTGGCTAGCGCCGGAGAGCTTCAAAGGGGAAACGAA
CTCGCACTACCGTCTAAATACGTGAATTTCTGTATTTAGCGTCCCATTACGAGAAGTTGAAAGGTTTACCTGAA
GATAACGAACAGAAGCAACTTTTTGTTGAGCAGCACAAACATTATCTCGACGAAATCATAGAGCAAATTTTCGGAA
TTCAGTAAGAGAGTCATCCTAGCTGATGCCAATCTGGACAAAGTATTAAGCGCATAACAACAGCACAGGGATAAA
CCCATACGTGAGCAGGCGGAAAATATTATCCATTTGTTTACTCTTACCAACCTCGGCGCTCCAGCCGCATTCAAG
TATTTTGACACAACGATAGATCGCAAACGATACACTTCTACCAAGGAGGTGCTAGACGCGACACTGATTCACCAA
TCCATCACGGGATTATATGAAACTCGGATAGATTTGTACAGCTTGGGGGTGACGGATCCCCCAAGAAGAAGAGG
AAAGTCTCGAGCGACTACAAAGACCATGACGGTGATTATAAAGATCATGACATCGATTACAAGGATGACGATGAC
AAGGCTGCAGGATCAGGTGGAAGTGGCGGCAGCGGAGGTTCGGATCCCAACTAGTCAAAGTGAACTGGAGGAG
AAGAAATCTGAACTTCGTCATAAATTGAAATATGTGCCTCATGAATATATTGAATTAATTGAAATTGCCAGAAAT
TCCACTCAGGATAGAATTCTTGAAATGAAGGTAATGGAATTTTTTATGAAAGTTTATGGATATAGAGGTAAACAT
TTGGGTGGATCAAGGAAACCGGACGGAGCAATTTATACTGTGCGATCTCCTATTGATTACGGTGTGATCGTGGAT
ACTAAAGCTTATAGCGGAGGTTATAATCTGCCAATTGGCCAAGCAGATGAAATGCAACGATATGTCGAAGAAAAT
CAAACACGAAACAAACATATCAACCCTAATGAATGGTGGAAAGTCTATCCATCTTCTGTAACGGAATTTAAGTTT
TTATTTGTGAGTGGTCACTTTAAAGGAACTACAAAGCTCAGCTTACACGATTAAATCATATCACTAATTGTAAT
GGAGCTGTTCTTAGTGTAGAAGAGCTTTTAATTGGTGGAGAAATGATTAAAGCCGGGCACATTAACCTTAGAGGAA
GTCAGACGGAAATTTAATAACGGCGAGATAAACTTT (SEQ ID NO:9)

[0090] fCas9 (*e.g.*, NLS- dCas9-GGS3linker –FokI):

ATGGACTACAAAGACCATGACGGTGATTATAAAGATCATGACATCGATTACAAGGATGACGATGACAAGATGGCC
CCCAAGAAGAAGAGGAAGGTGGGCATTCACCGCGGGGTACCTATGGATAAAAAGTATTCTATTGGTTTAGCTATC
GGCACTAATTCCGTTGGATGGGCTGTCATAACCGATGAATACAAAGTACCTTCAAAGAAATTTAAGGTGTTGGGG
AACACAGACCGTCATTTCGATTAAAAAGAATCTTATCGGTGCCCTCCTATTTCGATAGTGGCGAAACGGCAGAGGCG
ACTCGCCTGAAACGAACCGCTCGGAGAAGGTATACACGTCGCAAGAACCGAATATGTTACTTACAAGAAATTTTT
AGCAATGAGATGGCCAAAGTTGACGATTCTTTCTTTACCGTTTGAAGAGTCCCTTCCTTGTCGAAGAGGACAAG
AAACATGAACGGCACCCCATCTTTGGAAACATAGTAGATGAGGTGGCATATCATGAAAAGTACCCAACGATTTAT
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[0091] fCas9 (*e.g.*, FokI-GGS3linker-dCas9-NLS):

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[0092] fCas9 (*e.g.*, NLS -FokI-GGS3linker-dCas9):

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[0093] fCas9:

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AAAGCAATAGTAGATCTGTTATTCAAGACCAACCGCAAAGTGACAGTTAAGCAATTGAAAGAGGACTACTTTAAG
AAAATTGAATGCTTCGATTCTGTGCGAGATCTCCGGGGTAGAAGATCGATTTAATGCGTCACTTGGTACGTATCAT
GACCTCCTAAAGATAATTAAAGATAAGGACTTCCTGGATAACGAAGAGAATGAAGATATCTTAGAAGATATAGTG
TTGACTCTTACCCTCTTTGAAGATCGGGAAATGATTGAGGAAAGACTAAAAACATACGCTCACCTGTTTCGACGAT
AAGGTTATGAAACAGTTAAAGAGGCGTCGCTATACGGGCTGGGGACGATTGTTCGCGGAAACTTATCAACGGGATA
AGAGACAAGCAAAGTGGTAAACTATTCTCGATTTTCTAAAGAGCGACGGCTTCGCCAATAGGAACTTTATGCAG
CTGATCCATGATGACTCTTTAACCTTCAAAGAGGATATACAAAAGGCACAGGTTTCCGGACAAGGGGACTCATTG
CACGAACATATTGCGAATCTTGCTGGTTTCGCCAGCCATCAAAAAGGGCATACTCCAGACAGTCAAAGTAGTGGAT
GAGCTAGTTAAGGTCATGGGACGTCACAAACCGGAAAACATTGTAATCGAGATGGCACGCGAAAATCAAACGACT
CAGAAGGGGCAAAAAACAGTCGAGAGCGGATGAAGAGAATAGAAGAGGGTATTAAAGAACTGGGCAGCCAGATC
TTAAAGGAGCATCCTGTGGAAAATACCCAATTGCAGAACGAGAACTTTACCTCTATTACCTACAAAATGGAAGG
GACATGTATGTTGATCAGGAACCTGGACATAAACCGTTTATCTGATTACGACGTCGATGCCATTGTACCCCAATCC
TTTTTGAAGGACGATTCAATCGACAATAAAGTGCTTACACGCTCGGATAAGAACCGAGGGGAAAAGTGACAATGTT
CCAAGCGAGGAAGTCGTAAAGAAAATGAAGAACTATTGGCGGCAGCTCCTAAATGCGAACTGATAACGCAAAGA
AAGTTCGATAACTTAACTAAAGCTGAGAGGGGTGGCTTGCTGAACTTGACAAGGCCGGATTTATTAAACGTCAG
CTCGTGGAACCCGCCAAATCACAAAGCATGTTGCACAGATACTAGATTCCCGAATGAATACGAAATACGACGAG
AACGATAAGCTGATTCGGGAAGTCAAAGTAATCACTTTAAAGTCAAATTTGGTGTCTGGACTTCAGAAAGGATTTT
CAATTCTATAAAGTTAGGGAGATAAATAACTACCACCATGCGCACGACGCTTATCTTAATGCCGTCGTAGGGACC
GCACTCATTAAGAAATACCCGAAGCTAGAAAGTGAGTTTGTGTATGGTGATTACAAAGTTTATGACGTCCGTAAG
ATGATCGCGAAAAGCGAACAGGAGATAGGCAAGGCTACAGCCAAATACTTCTTTTATTCTAACATTATGAATTTCT
TTTAAGACGGAAATCACTCTGGCAAACGGAGAGATACGCAAACGACCTTTAATTGAAACCAATGGGGAGACAGGT
GAAATCGTATGGGATAAGGGCCGGGACTTCGCGACGGTGAGAAAAGTTTTGTCCATGCCCCAAGTCAACATAGTA
AAGAAAACCTGAGGTGCAGACCGGAGGGTTTTCAAAGGAATCGATTCTTCCAAAAGGAATAGTGATAAGCTCATC
GCTCGTAAAAAGGACTGGGACCCGAAAAAGTACGGTGGCTTCGATAGCCCTACAGTTGCCTATTCTGTCTCTAGTA
GTGGCAAAGTTGAGAAGGGAAAATCCAAGAACTGAAGTCAGTCAAAGAATTATTGGGGATAACGATTATGGAG
CGCTCGTCTTTTGAAAAGAACCCCATCGACTTCCTTGAGGCGAAAGGTTACAAGGAAGTAAAAAAGGATCTCATA
ATTAAACTACCAAAGTATAGTCTGTTTGAGTTAGAAAATGGCCGAAAACGGATGTTGGCTAGCGCCGGAGAGCTT
CAAAGGGGAACGAACCTCGCACTACCGTCTAAATACGTGAATTTCTGTATTTAGCGTCCCATTACGAGAAGTTG
AAAGGTTACCTGAAGATAACGAACAGAAGCAACTTTTTGTTGAGCAGCACAAACATTATCTCGACGAAATCATA
GAGCAAATTTTCGAATTCAGTAAGAGAGTCATCCTAGCTGATGCCAATCTGGACAAAGTATTAAGCGCATACAAC
AAGCACAGGGATAAACCCATACGTGAGCAGGCGGAAAATATTATCCATTTGTTTACTCTTACCAACCTCGGCGCT
CCAGCCGCATTCAAGTATTTTGACACAACGATAGATCGCAAACGATACACTTCTACCAAGGAGGTGCTAGACGCG

ACACTGATTCACCAATCCATCACGGGATTATATGAAACTCGGATAGATTTGTCACAGCTTGGGGGTGAC (SEQ ID NO:319)

[0094] In some embodiments, an inventive fusion protein (*e.g.*, fCas9) corresponds to or is encoded by a homologue of any one of SEQ ID NO:9-12 or SEQ ID NO:319.

[0095] In some embodiments, an inventive fusion protein (*e.g.*, fCas9) comprises, in part or in whole, one or more of the amino acid sequences set forth as SEQ ID NO:5, SEQ ID NO:320, SEQ ID NO:6, SEQ ID NO:16, SEQ ID NO:318, and SEQ ID NO:321, as provided herein and shown below. The various domains corresponding to SEQ ID NO:5, SEQ ID NO:320, SEQ ID NO:6, SEQ ID NO:16, SEQ ID NO:318, and SEQ ID NO:321 may be arranged in any order with respect to each other. For example, in some embodiments, a dCas9 domain (*e.g.*, SEQ ID NO:5 or SEQ ID NO:320) is at the amino or carboxy terminus, or is somewhere in between the amino and carboxy termini. Similarly, each of the other domains corresponding to SEQ ID NO:6, SEQ ID NO:16, SEQ ID NO:318, and SEQ ID NO:321 may be at the amino or carboxy terminus, or somewhere in between the amino and carboxy termini of an inventive fusion protein (*e.g.*, fCas9). Examples of inventive fusion proteins having various domain arrangements include the inventive fusion proteins corresponding to SEQ ID NOs:9-12 and SEQ ID NO:319. In some embodiments, an inventive fusion protein comprises additional or other domains, such as other linkers, other NLS domains, other nuclease domains, or other Cas9 domains, which may be in addition to or substituted for any of the domains as provided herein.

[0096] FokI cleavage domain:

GSQLVKSELEEKSELHRKLKYPHEYIELIEIARNSTQDRILEMKVMEFFMKVYGYRGKHLGGSRKPDGAIYTV
GSPIDYGVIVDTKAYSGGYNLPIGQADEMQRYVEENQTRNKHINPNEWKVPSSVTEFKFLFVSGHFKNYKAQ
LTRLNHITNCNGAVLSVEELLIGGEMIKAGTLTLEEVRRKFNNGEINF (SEQ ID NO:6)

[0097] dCas9:

DKKYSIGLAIGTNSVGWAVITDEYKVPSKKFKVLGNTDRHSIKKNLIGALLFDSGETAEATRLKRTARRRYTRRK
NRICYLQEIFSNEMAKVDDSFHRLEESFLVEEDKKHERHPIFGNIVDEVAYHEKYPTIYHLRKKLVDSTDKADL
RLIYLALAHMIKFRGHFLIEGDLNPDNSDVKLFIQLVQTYNQLFEENPINASGVDAKAILSARLSKSRLENLI
AQLPGEKKNGLFGNLIASLGLTPNFKSNFDLAEDAKLQLSKDQYADLFLAAKNLSDAIL
LSDILRVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQLPEKYKEIFFDQSKNGYAGYIDGGASQEEFYKFI
KPILEKMDGTEELLVKLNREDLLRKQRTFDNGSIPHQIHLGELHAILRRQEDFYFPLKDNREKIEKILTFRIPYY
VGPLARGNSRFAWMTRKSEETITPWNFEEVVDKGASAQSFIERMTNFDKNLPNEKVLPHKSLLEYFTVYNELTK
VKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKIECFDSVEISGVEDRFNASLGTYHDLKII
KDKDFLDNEENEDILEDIVLTTLTFEDREMIEERLKYAHLFDDKVMKQLKRRRYTGWGRLSRKLINGIRDKQSG
KTILDFLKSDGFANRNFQMQLIHDDSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPAIAKKGILQTVKVVDELVKVM
GRHKPENIVIMARENQTTQKGQKNSRERMKRIEELGSGILKEHPVENTQLQNEKLYLYYLQNGRDMYVDQ
ELDINRLSDYDVAIVPQSFLKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKKMKNYWRQLLNAKLITQRKFDNLT
KAERGGLSELDKAGFIKRQLVETRQITKHVAQILDSRMNTKYDENDKLIREVKVITLKSCLVSDFRKDFQFYKVR

EINNYHHAHDAYLNAVVGITALIKKYPKLESEFVYGDYKVYDVRKMIKSEQEIGKATAYFFYSNIMNFFKTEIT
 LANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVLSPQVNIIVKKTEVQTGGFSKESILPKRNSDKLIARKKDW
 DPKKYGGFDSPTVAYSVLVVAKEKGKSKKLKSVKELLGITIMERSSEFKNPIDFLEAKGYKEVKKDLIIKLPKY
 SLFELENGRKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLKGSPEDEQKQLFVEQHKHYLDEIIEQISEF
 SKRVILADANLDKVL SAYNKH RD KPIREQAENIIHLFTLTNLGAPAAFKYFDTTIDRKRYTSTKEVLDATLIHQ
 ITGLYETRIDLSQLGGD (SEQ ID NO:320)

[0098] 3xFLAG TAG:

MDYKDHDGDYKDHDIDYKDDDDK (SEQ ID NO:321)

[0099] NLS domain:

MAPKKRKVGIIHRGVP (SEQ ID NO:318)

[00100] XTEN linker:

SGSETPGTSESATPES (SEQ ID NO:16)

[00101] In some embodiments, the fusion proteins forming the dimer are coordinated through the action of a single extended gRNA (*e.g.*, as opposed to two separate gRNAs, each binding a monomer of the fusion protein dimer). Thus, in some aspects, the single extended gRNA contains at least two portions, separated by a linker sequence, that complement the target nucleic acid (*e.g.*, bind the target nucleic acid at two distinct sites), and the gRNA is able to bind at least two fusion proteins, as described herein. This is exemplified by the schematic shown in Figure 1B. In some embodiments, the linker sequence separating the two portions in the extended gRNA has complementarity with the target sequence. In some embodiments, the extended gRNA is at least 50, at least 60, at least 70, at least 80, at least 90, at least 100, at least 125, at least 150, at least 175, at least 200, at least 250, at least 300, at least 350, at least 400, at least 500, at least 600, at least 700, at least 800, at least 900, or at least 1000 or more nucleotides in length. Whether the fusion proteins are coordinated through separate or a single gRNA, to form dimers that can cleave a target nucleic acid, it is expected that the specificity of such cleavage is enhanced (*e.g.*, reduced or no off-target cleavage) as compared to nucleases having a single target nucleic acid binding site. Methods for determining the specificity of a nuclease are known (see *e.g.*, published PCT Application, WO 2013/066438; pending provisional application US 61/864,289; and Pattanayak, V., Ramirez, C.L., Joung, J.K. & Liu, D.R. Revealing off-target cleavage specificities of zinc-finger nucleases by in vitro selection. *Nature Methods* **8**, 765-770 (2011), the entire contents of each of which are incorporated herein by reference).

[00102] According to another embodiment, dimers of Cas9 protein are provided. In some embodiments, the dimers are coordinated through the action of a single extended gRNA that comprises at least two portions that complement the target nucleic acid. In some embodiments, the portions complementary to the target nucleic acid comprise no more than 25, no more than 24, no more than 23, no more than 22, no more than 21, no more than 20, no more than 19, no more than 18, no more than 17, no more than 16, no more than 15, no more than 14, no more than 13, no more than 12, no more than 11, no more than 10, no more than 9, no more than 8, no more than 7, no more than 6, or no more than 5 nucleotides that complement the target nucleic acid. In some embodiments, the portions complementary to the target nucleic acid comprise 5-30, 5-25, or 5-20 nucleotides. In some embodiments, the portions complementary to the target nucleic acid comprise 15-25, 19-21, or 20 nucleotides. In some embodiments, the portions comprise the same number of nucleotides that complement the target nucleic acid. In some embodiments, the portions comprise different numbers of nucleotides that complement the target nucleic acid. For example, in some embodiments, the extended gRNA comprises two portions that complement (*e.g.*, and hybridize to) the target nucleic acid, each portion comprising 5-19, 10-15, or 10 nucleotides that complement the target nucleic acid. Without wishing to be bound by any particular theory, having the portions comprise fewer than approximately 20 nucleotides typical of gRNAs (*e.g.*, having the portions comprise approximately 5-19, 10-15, or 10 complementary nucleotides), ensures that a single Cas9:gRNA unit cannot bind efficiently by itself. Thus the cooperative binding between Cas9 proteins coordinated by such an extended gRNA improves the specificity and cleavage of intended target nucleic acids. In some embodiments, the linker sequence separating the two portions of the extended gRNA has complementarity with the target sequence. For example, in some embodiments, the linker sequence has at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 20, at least 25, at least 30, at least 40, or at least 50 nucleotides that complement the target nucleic acid. Without wishing to be bound by any particular theory, it is believed that having an extended gRNA that comprises multiple binding sites (*e.g.*, multiple low-affinity binding sites), including those that are bound by a Cas9 protein as well as those in the linker sequence, provides for increased specificity by promoting cooperative binding. Certain aspects of this embodiment are shown in Figure 4. In some embodiments, any of the Cas9 proteins described herein may be coordinated through a single extended gRNA.

[00103] In another embodiment, proteins comprising a fragment of an RNA-programmable nuclease (*e.g.*, Cas9) are provided. For example, in some embodiments, a protein comprising the gRNA binding domain of Cas9 is provided. In some embodiments, the protein comprising the gRNA binding domain of Cas9 does not comprise a DNA cleavage domain (referred to herein as the “A-half” of Cas9). In other embodiments, proteins comprising the DNA cleavage domain(s) (*e.g.*, the HNH, RuvC1 subdomains) of Cas9 are provided. In some embodiments, the “DNA cleavage domain” refers collectively to the portions of Cas9 required for double-stranded DNA cleavage (*e.g.*, the HNH, RuvC1 subdomains). In some embodiments, the protein comprising the DNA cleavage domain of Cas9 does not comprise a gRNA binding domain (referred to herein as the “B-half” of Cas9). In some embodiments, dimers are provided that comprise (i) a protein comprising the gRNA binding domain of Cas9 (*e.g.*, the A-half), and (ii) a protein comprising the DNA cleavage domain of Cas9 (*e.g.*, the B-half). In some embodiments, the dimer is bound by a gRNA. For example, such dimers are expected to recapitulate the binding and cleaving activities of a full length Cas9 protein. In some embodiments, such dimers are referred to herein as “dimeric split Cas9.” Using a dimeric split Cas9 to cleave a target nucleic acid is expected to provide for increased specificity as compared to a single full length Cas9 protein because both halves of the protein must be co-localized to associate and re-fold into a nuclease-active state. This strategy is shown in the schematic of Figure 2A.

[00104] In some embodiments, fusion proteins comprising two domains are provided: (i) a protein capable of specifically binding a target nucleic acid (*e.g.*, a nuclease-inactivated RNA programmable nuclease, such as a nuclease-inactivated Cas9, as described herein) fused or linked to (ii) a fragment of an RNA-programmable nuclease (*e.g.*, the A- or B-half of Cas9, as described herein). In some embodiments, domain (i) of the aforementioned fusion protein comprises a DNA binding domain, for example, a DNA binding domain of a zinc finger or TALE protein. In some embodiments, the fusion protein comprises (i) a nuclease-inactivated Cas9, and (ii) a gRNA binding domain of Cas9 (*e.g.*, Cas9 A-half). In some embodiments, domain (ii) of the fusion protein does not include a DNA cleavage domain. In other embodiments, the fusion protein comprises (i) a nuclease-inactivated Cas9, and (ii) a DNA cleavage domain (*e.g.*, Cas9 B-half). In some embodiments, domain (ii) of the fusion protein does not include a gRNA binding domain.

[00105] In some embodiments, dimers are provided that comprise two proteins: (i) a fusion protein comprising a nuclease-inactivated Cas9 and a gRNA binding domain of Cas9 (*e.g.*, nuclease-inactivated Cas9 fused to Cas9 A-half), and (ii) a protein comprising the DNA

cleavage domain of Cas9 (*e.g.*, Cas9 B-half). In other embodiments, the dimer comprises (i) a fusion protein comprising a nuclease-inactivated Cas9 and a DNA cleavage domain of Cas9 (*e.g.*, nuclease-inactivated Cas9 fused to Cas9 B-half), and (ii) a protein comprising the gRNA binding domain of Cas9 (*e.g.*, Cas9 A-half). In some embodiments, the protein dimers include one or more gRNAs. For example, in some embodiments, the dimers include two gRNAs: one bound by the nuclease-inactivated Cas9 domain of the fusion protein; the other bound by the A-half domain (*e.g.*, either the A-half of the fusion protein, or the A-half of the dimer not part of the fusion protein). Such a dimer (*e.g.*, associated with two gRNAs having sequences binding separate regions of a target nucleic acid) is expected to have improved specificity compared to *e.g.*, a Cas9 protein having a single gRNA. This strategy is shown in Figure 2B.

[00106] In some embodiments, a protein dimer is provided that comprises two fusion proteins: (i) a fusion protein comprising a nuclease-inactivated Cas9 and a gRNA binding domain of Cas9 (*e.g.*, a nuclease-inactivated Cas9 fused to a Cas9 A-half), and (ii) a fusion protein comprising a nuclease-inactivated Cas9 and a DNA cleavage domain (*e.g.*, a nuclease-inactivated Cas9 fused to a Cas9 B-half). In some embodiments, the dimer is associated with (*e.g.*, binds) one or more distinct gRNAs. For example, in some embodiments, the dimer is associated with two or three gRNAs. In some embodiments, the dimer is associated with three gRNAs. For example, upon binding of one nuclease-inactivated Cas9:gRNA to a region of a nucleic acid target, and binding of the other nuclease-inactivated Cas9:gRNA to a second region of the nucleic acid target, the split Cas9 halves (*e.g.*, A-half and B-half of the fusion proteins) can dimerize and bind a third gRNA complementary to a third region of the nucleic acid target, to become a fully active Cas9 nuclease, which can cleave dsDNA. This strategy is illustrated in Figure 2C.

[00107] According to another aspect of the invention, minimized Cas9 proteins are provided. By “minimized,” it is meant that the Cas9 protein comprises amino acid deletions and/or truncations, as compared to the wild type protein, but retains gRNA binding activity, DNA cleavage activity, or both. Any of the embodiments herein describing Cas9 proteins (*e.g.*, split Cas9 proteins, Cas9 A-half, Cas9 B-half, nuclease-inactivated Cas9 fusion proteins, *etc.*) can utilize a minimized Cas9 protein. In some embodiments, minimized Cas9 proteins comprising N-terminal deletions and/or truncations are provided. In some embodiments, minimized Cas9 proteins comprising C-terminal deletions and/or truncations are provided. In some embodiments, minimized Cas9 proteins are provided that comprise N- and/or C-terminal deletions and/or truncations. In some embodiments, the minimized Cas9

protein retains both gRNA binding and DNA cleavage activities. In some embodiments, the minimized Cas9 protein comprises an N-terminal truncation that removes at least 5, at least 10, at least 15, at least 20, at least 25, at least 40, at least 40, at least 50, at least 75, at least 100, at least 150, at least 200, at least 250, at least 300, at least 350, at least 400, at least 450, or at least 500 amino acids. In some embodiments, the minimized Cas9 protein comprises a C-terminal truncation that removes at least 5, at least 10, at least 15, at least 20, at least 25, at least 40, at least 40, at least 50, at least 75, at least 100, at least 150, at least 200, at least 250, at least 300, at least 350, at least 400, at least 450, or at least 500 amino acids. In some embodiments, deletions are made within Cas9, for example in regions not affecting gRNA binding and/or DNA cleavage. In some embodiments, the minimized Cas9 protein is associated with one or more gRNAs. In certain embodiments, the minimized Cas9 protein is associated with one gRNA.

Recombinases

[00108] Some aspects of this disclosure provide RNA-guided recombinase fusion proteins that are designed using the methods and strategies described herein. Some embodiments of this disclosure provide nucleic acids encoding such recombinases. Some embodiments of this disclosure provide expression constructs comprising such encoding nucleic acids. For example, in some embodiments an isolated recombinase is provided that has been engineered to recombine a desired target site (*e.g.*, a site targeted by one or more gRNAs bound to one or more of the engineered recombinases) within a genome, *e.g.*, with another site in the genome or with an exogenous nucleic acid. In some embodiments, the isolated recombinase comprises a variant of an RNA-programmable nuclease, such as a Cas9 nuclease. In some embodiments, the Cas9 variant is a nuclease-inactivated Cas9 (*e.g.*, dCas9). In some embodiments, dCas9 is encoded by a nucleotide sequence comprising in part or in whole, SEQ ID NO:5 or SEQ ID NO:320. In some embodiments, dCas9 is encoded by a nucleotide sequence comprising a variant of SEQ ID NO:5 or SEQ ID NO:320.

[00109] In one embodiment, an RNA-guided recombinase fusion protein is provided. Typically, the fusion protein comprises two or more domains. In some embodiments, the fusion protein comprises two domains. In some embodiments, one of the two or more domains is a nuclease-inactivated Cas9 (or fragment thereof, *e.g.*, Cas9 A-half), for example, those described herein (*e.g.*, dCas9). The Cas9 domain of the recombinase fusion protein is capable of binding one or more gRNAs, and thereby directs or targets the recombinase fusion protein(s) to a target nucleic acid, *e.g.*, as described herein. Another domain of the two or

more domains is a recombinase, or a fragment thereof, *e.g.*, a catalytic domain of a recombinase. By “catalytic domain of a recombinase,” it is meant that a fusion protein includes a domain comprising an amino acid sequence of (*e.g.*, derived from) a recombinase, such that the domain is sufficient to induce recombination when contacted with a target nucleic acid (either alone or with additional factors including other recombinase catalytic domains which may or may not form part of the fusion protein). In some embodiments, a catalytic domain of a recombinase excludes a DNA binding domain of the recombinase. In some embodiments, the catalytic domain of a recombinase includes part or all of a recombinase, *e.g.*, the catalytic domain may include a recombinase domain and a DNA binding domain, or parts thereof, or the catalytic domain may include a recombinase domain and a DNA binding domain that is mutated or truncated to abolish DNA binding activity. Recombinases and catalytic domains of recombinases are known to those of skill in the art, and include, for example, those described herein. In some embodiments, the catalytic domain is derived from any recombinase. In some embodiments, the recombinase catalytic domain is a catalytic domain of a Tn3 resolvase, a Hin recombinase, or a Gin recombinase. In some embodiments, the catalytic domain comprises a Tn3 resolvase (*e.g.*, Stark Tn3 recombinase) that is encoded by a nucleotide sequence comprising, in part or in whole, SEQ ID NO:322, as provided below. In some embodiments, a Tn3 catalytic domain is encoded by a variant of SEQ ID NO:322. In some embodiments, a Tn3 catalytic domain is encoded by a polynucleotide (or a variant thereof) that encodes the polypeptide corresponding to SEQ ID NO:325. In some embodiments, the catalytic domain comprises a Hin recombinase that is encoded by a nucleotide sequence comprising, in part or in whole, SEQ ID NO:323, as provided below. In some embodiments, a Hin catalytic domain is encoded by a variant of SEQ ID NO:323. In some embodiments, a Hin catalytic domain is encoded by a polynucleotide (or a variant thereof) that encodes the polypeptide corresponding to SEQ ID NO:326. In some embodiments, the catalytic domain comprises a Gin recombinase (*e.g.*, Gin beta recombinase) that is encoded by a nucleotide sequence comprising, in part or in whole, SEQ ID NO:324, as provided below. In some embodiments, a Gin catalytic domain is encoded by a variant of SEQ ID NO:324. In some embodiments, a Gin catalytic domain is encoded by a polynucleotide (or a variant thereof) that encodes the polypeptide corresponding to SEQ ID NO:327.

[00110] Stark Tn3 recombinase (nucleotide: SEQ ID NO:322; amino acid: SEQ ID NO:325):

ATGGCCCTGTTTGGCTACGCACGCGTGTCTACCAGTCAACAGTCACTCGATTTGCAAGTGAGGGCTCTTAAAGAT
GCCGGAGTGAAGGCAAACAGAATTTTTTACTGATAAGGCCAGCGGAAGCAGCACAGACAGAGAGGGGCTGGATCTC
CTGAGAATGAAGGTAAAGGAGGGTGATGTGATCTTGGTCAAAAAATTGGATCGACTGGGGAGAGACACAGCTGAT
ATGCTTCAGCTTATTAAAGAGTTTGACGCTCAGGGTGTTGCCGTGAGGTTTATCGATGACGGCATCTCAACCGAC
TCCTACATTGGTCTTATGTTTGTGACAATTTTGTCCGCTGTGGCTCAGGCTGAGCGGAGAAGGATTCTCGAAAGG
ACGAATGAGGGACGGCAAGCAGCTAAGTTGAAAGGTATCAAATTTGGCAGACGAAGG (SEQ ID NO:322)

MALFGYARVSTSQQSLDLQVRALKDAGVKANRIFTDKASGSSTDREGLDLLRMKVKEGDVILVKKLDRLGRDTAD
MLQLIKEFDAQGVAVRFIDDGISTDSYIGLMFVTILSAVAQAERRRILERTNEGRQAARKLKGIFGRRR (SEQ
ID NO:325)

[00111] Hin Recombinase (nucleotide: SEQ ID NO:323; amino acid: SEQ ID NO:326):

ATGGCAACCATTGGCTACATAAGGGTGTCTACCATCGACCAAATATCGACCTGCAGCGCAACGCTCTGACATCC
GCCAACTGCGATCGGATCTTCGAGGATAGGATCAGTGGCAAGATCGCCAACCGGCCCGGTCTGAAGCGGGCTCTG
AAGTACGTGAATAAGGGCGATACTCTGGTTGTGTGGAAGTTGGATCGCTTGGGTAGATCAGTGAAGAATCTCGTA
GCCCTGATAAGCGAGCTGCACGAGAGGGGTGCACATTTCCATTCTCTGACCGATTCCATCGATACGTCTAGCGCC
ATGGGCCGATTCTTCTTTTACGTCATGTCCGCCCTCGCTGAAATGGAGCGCGAAGTTATTGTTGAACGGACTTTG
GCTGGACTGGCAGCGGCTAGAGCACAGGGCCGACTTGA (SEQ ID NO:323)

MATIGYIRVSTIDQNIDLQRNALTSANCDRIFEDRISGKIANRPGLKRALKYVNKGDTLVVWKLDRLGRSVKNLV
ALISELHERGAHFHSLTDSIDTSSAMGRFFFYVMSALAEMERELIVERTLAGLAAARAQGR LG (SEQ ID
NO:326)

[00112] Gin beta recombinase (nucleotide: SEQ ID NO:324; amino acid: SEQ ID NO:327):

ATGCTCATTGGCTATGTAAGGGTCAGCACCAATGACCAAACACAGACTTGCAACGCAATGCTTTGGTTTGCGCC
GGATGTGAACAGATATTTGAAGATAAACTGAGCGGCACTCGGACAGACAGACCTGGGCTTAAGAGAGCACTGAAA
AGACTGCAGAAGGGGGACACCCTGGTCGTCTGGAACTGGATCGCCTCGGACGCAGCATGAAACATCTGATTAGC
CTGGTTGGTGAGCTTAGGGAGAGAGGAATCAACTTCAGAAGCCTGACCGACTCCATCGACACCAGTAGCCCCATG
GGACGATTCTTCTTCTATGTGATGGGAGCACTTGCTGAGATGGAAAGAGAGCTTATTATCGAAAGAACTATGGCT
GGTATCGCTGCTGCCCCGGAACAAAGGCAGACGGTTCGGCAGACCGCCGAAGAGCGGC (SEQ ID NO:324)

MLIGYVRVSTNDQNTDLQRNALVCAGCEQIFEDKLSGTRTDRPGLKRALKRLQKGDTLVVWKLDRLGRSMKHLIS
LVGELRERGINFRSLTDSIDTSSPMGRFFFYVMGALAEMERELI IERTMAGIAAARNKGRRFGRPPKSG (SEQ
ID NO:327)

[00113] In some embodiments, the recombinase catalytic domain is fused to the N-terminus, the C-terminus, or somewhere in between the N- and C-termini of a Cas9 protein (*e.g.*, sCas9). In some embodiments, the fusion protein further comprises a nuclear localization signal (NLS; *e.g.*, any of those provided herein). For example, in some embodiments, the general architecture of exemplary RNA-guided recombinase fusion proteins (*e.g.*, Cas9-recombinase fusions) comprise one of the following structures:

[NH₂]-[Cas9]-[recombinase]-[COOH],
 [NH₂]-[recombinase]-[Cas9],
 [NH₂]-[NLS]-[Cas9]-[recombinase]-[COOH],
 [NH₂]-[NLS]-[recombinase]-[Cas9]-[COOH],
 [NH₂]-[Cas9]-[NLS]-[recombinase]-[COOH],
 [NH₂]-[recombinase]-[NLS]-[Cas9]-[COOH],
 [NH₂]-[Cas9]-[recombinase]-[NLS]-[COOH], or
 [NH₂]-[recombinase]-[Cas9]-[NLS]-[COOH]

wherein NLS is a nuclear localization signal, NH₂ is the N-terminus of the fusion protein, and COOH is the C-terminus of the fusion protein. In some embodiments, a linker is inserted between the Cas9 domain and the recombinase domain, *e.g.*, any linker provided herein. Additional features, such as sequence tags (*e.g.*, any of those provided herein), may also be present.

Pharmaceutical compositions

[00114] In some embodiments, any of the nucleases (*e.g.*, fusion proteins comprising nucleases or nuclease domains) and recombinases (*e.g.*, fusion proteins comprising recombinases or recombinase catalytic domains) described herein are provided as part of a pharmaceutical composition. For example, some embodiments provide pharmaceutical compositions comprising a nuclease and/or recombinase as provided herein, or a nucleic acid encoding such a nuclease and/or recombinase, and a pharmaceutically acceptable excipient. Pharmaceutical compositions may optionally comprise one or more additional therapeutically active substances.

[00115] In some embodiments, compositions provided herein are administered to a subject, for example, to a human subject, in order to effect a targeted genomic modification within the subject. In some embodiments, cells are obtained from the subject and are contacted with a nuclease and/or recombinase *ex vivo*. In some embodiments, cells removed from a subject and contacted *ex vivo* with an inventive nuclease and/or recombinase are re-introduced into the subject, optionally after the desired genomic modification has been effected or detected in the cells. Although the descriptions of pharmaceutical compositions provided herein are principally directed to pharmaceutical compositions which are suitable for administration to humans, it will be understood by the skilled artisan that such compositions are generally suitable for administration to animals of all sorts. Modification of pharmaceutical compositions suitable for administration to humans in order to render the

compositions suitable for administration to various animals is well understood, and the ordinarily skilled veterinary pharmacologist can design and/or perform such modification with merely ordinary, if any, experimentation. Subjects to which administration of the pharmaceutical compositions is contemplated include, but are not limited to, humans and/or other primates; mammals, domesticated animals, pets, and commercially relevant mammals such as cattle, pigs, horses, sheep, cats, dogs, mice, and/or rats; and/or birds, including commercially relevant birds such as chickens, ducks, geese, and/or turkeys.

[00116] Formulations of the pharmaceutical compositions described herein may be prepared by any method known or hereafter developed in the art of pharmacology. In general, such preparatory methods include the step of bringing the active ingredient into association with an excipient, and then, if necessary and/or desirable, shaping and/or packaging the product into a desired single- or multi-dose unit.

[00117] Pharmaceutical formulations may additionally comprise a pharmaceutically acceptable excipient, which, as used herein, includes any and all solvents, dispersion media, diluents, or other liquid vehicles, dispersion or suspension aids, surface active agents, isotonic agents, thickening or emulsifying agents, preservatives, solid binders, lubricants and the like, as suited to the particular dosage form desired. Remington's *The Science and Practice of Pharmacy*, 21st Edition, A. R. Gennaro (Lippincott, Williams & Wilkins, Baltimore, MD, 2006; incorporated in its entirety herein by reference) discloses various excipients used in formulating pharmaceutical compositions and known techniques for the preparation thereof. See also PCT application PCT/US2010/055131, incorporated in its entirety herein by reference, for additional suitable methods, reagents, excipients and solvents for producing pharmaceutical compositions comprising a nuclease. Except insofar as any conventional excipient medium is incompatible with a substance or its derivatives, such as by producing any undesirable biological effect or otherwise interacting in a deleterious manner with any other component(s) of the pharmaceutical composition, its use is contemplated to be within the scope of this disclosure.

[00118] In some embodiments, compositions in accordance with the present invention may be used for treatment of any of a variety of diseases, disorders, and/or conditions, including but not limited to one or more of the following: autoimmune disorders (*e.g.* diabetes, lupus, multiple sclerosis, psoriasis, rheumatoid arthritis); inflammatory disorders (*e.g.* arthritis, pelvic inflammatory disease); infectious diseases (*e.g.* viral infections (*e.g.*, HIV, HCV, RSV), bacterial infections, fungal infections, sepsis); neurological disorders (*e.g.* Alzheimer's disease, Huntington's disease; autism; Duchenne muscular dystrophy);

cardiovascular disorders (*e.g.* atherosclerosis, hypercholesterolemia, thrombosis, clotting disorders, angiogenic disorders such as macular degeneration); proliferative disorders (*e.g.* cancer, benign neoplasms); respiratory disorders (*e.g.* chronic obstructive pulmonary disease); digestive disorders (*e.g.* inflammatory bowel disease, ulcers); musculoskeletal disorders (*e.g.* fibromyalgia, arthritis); endocrine, metabolic, and nutritional disorders (*e.g.* diabetes, osteoporosis); urological disorders (*e.g.* renal disease); psychological disorders (*e.g.* depression, schizophrenia); skin disorders (*e.g.* wounds, eczema); blood and lymphatic disorders (*e.g.* anemia, hemophilia); *etc.*

Methods for site-specific nucleic acid cleavage

[00119] In another embodiment of this disclosure, methods for site-specific nucleic acid (*e.g.*, DNA) cleavage are provided. In some embodiments, the methods comprise contacting a DNA with any of the Cas9:gRNA complexes described herein. For example, in some embodiments, the method comprises contacting a DNA with a fusion protein (*e.g.*, fCas9) that comprises two domains: (i) a nuclease-inactivated Cas9 (dCas9); and (ii) a nuclease (*e.g.*, a FokI DNA cleavage domain), wherein the wherein the inactive Cas9 domain binds a gRNA that hybridizes to a region of the DNA. In some embodiments, the method further comprises contacting the DNA with a second fusion protein described herein (*e.g.*, fCas9), wherein the nuclease-inactivated Cas9 (dCas9) domain of the second fusion protein binds a second gRNA that hybridizes to a second region of DNA, wherein the binding of the fusion proteins results in the dimerization of the nuclease domains of the fusion proteins, such that the DNA is cleaved in a region between the bound fusion proteins. See *e.g.*, Figures 1A, 6D. In some embodiments, the gRNAs bound to each fusion protein hybridize to the same strand of the DNA, or they hybridize to opposing strands of the DNA. In some embodiments, the gRNAs hybridize to regions of the DNA that are no more than 10, no more than 15, no more than 20, no more than 25, no more than 30, no more than 40, 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart. The region between the bound Cas9:gRNA complexes may be referred to as the “spacer sequence,” which is typically where the target nucleic acid is cleaved. See, *e.g.*, Figures 6C-D. In some embodiments, the spacer sequence is at least 5, at least 10, at least 15, at least 20, at least 25, at least 30 at least 35, at least 40, at least 45, at least 50, at least 60, at least 70, at least 80, at least 90, or at least 100 base pairs in length. In some embodiments, the spacer sequence is between about 5 and about 50 base pairs, about 10 and about 40, or about 15 and about 30 base pairs in length. In some embodiments, the spacer sequence is

about 15 to about 25 base pairs in length. In some embodiments, the spacer sequence is about 15, about 20, or about 25 base pairs in length. In some embodiments, the Cas9:gRNA complexes are bound in the A orientation, as described herein. In some embodiments, the Cas9:gRNA complexes are bound in the B orientation, as described herein. In some embodiments, the method has an on-target:off-target modification ratio that is at least 2-fold, at least 5-fold, at least 10-fold, at least 20-fold, at least 30-fold, at least 40-fold, at least 50-fold, at least 60-fold, at least 70-fold, at least 80-fold, at least 90-fold, at least 100-fold, at least 110-fold, at least 120-fold, at least 130-fold, at least 140-fold, at least 150-fold, at least 175-fold, at least 200-fold, or at least 250-fold or more higher than the on-target:off-target modification ratio of methods utilizing a wild type Cas9 or other Cas9 variant. In some embodiments, the method has an on-target:off-target modification ratio that is between about 60- to about 180-fold, between about 80- to about 160-fold, between about 100- to about 150-fold, or between about 120- to about 140-fold higher than the on-target:off-target modification ratio of methods utilizing a wild type Cas9 or other Cas9 variant. Methods for determining on-target:off-target modification ratios are known, and include those described in the Examples. In some embodiments, the fusion proteins are coordinated or associated through a single gRNA, *e.g.*, as described herein.

[00120] In some embodiments, the method comprises contacting a nucleic acid with a dimer of Cas9 proteins (or fragments thereof) coordinated with (*e.g.*, bound by) a single gRNA as described herein. In some embodiments, the single gRNA comprises at least two portions that hybridize to the nucleic acid. In some embodiments, the portions comprise at least 5, at least 10, at least 15, or at least 19 complementary nucleotides. In some embodiments, the portions comprise fewer than 20 complementary nucleotides. In some embodiments, a linker sequence separates the portions, wherein the linker sequence also comprises nucleotides complementary to the target nucleic acid (*e.g.*, but are not bound by a Cas9 protein). In some embodiments, the linker sequence does not hybridize to the target nucleic acid.

[00121] In some embodiments, the methods comprise contacting a DNA with a protein dimer of fusion proteins described herein, wherein the fusion proteins are bound by one or more gRNAs. For example, in some embodiments, one fusion protein of the dimer comprises a gRNA binding domain of Cas9 (*e.g.*, Cas9 A-half), wherein the protein does not comprise a DNA cleavage domain (*e.g.*, Cas9 B-half); and the other fusion protein of the dimer comprises a DNA cleavage domain of Cas9 (*e.g.*, Cas9 B-half), wherein the protein does not comprise a gRNA binding domain (*e.g.*, Cas9 A-half). Thus, in some embodiments, the

binding of a gRNA (*e.g.*, that hybridizes to a target nucleic acid) to one or both of the monomers of the dimer co-localizes the dimer to the target nucleic acid, allowing the dimer to re-fold into a nuclease-active state and cleave the target nucleic acid.

[00122] In some embodiments, the method comprises contacting a nucleic acid with protein dimers comprising two proteins: (i) a fusion protein comprising a nuclease-inactivated Cas9 and a gRNA binding domain of Cas9 (*e.g.*, nuclease-inactivated Cas9 fused to Cas9 A-half), and (ii) a protein comprising the DNA cleavage domain of Cas9 (*e.g.*, Cas9 B-half). In other embodiments, the dimer comprises (i) a fusion protein comprising a nuclease-inactivated Cas9 and a DNA cleavage domain of Cas9 (*e.g.*, nuclease-inactivated Cas9 fused to Cas9 B-half), and (ii) a protein comprising the gRNA binding domain of Cas9 (*e.g.*, Cas9 A-half). In some embodiments, the protein dimers are associated with one or more gRNAs. For example, in some embodiments, the dimers are associated with two gRNAs: one bound by the nuclease-inactivated Cas9 domain of the fusion protein; the other bound by the A-half domain (*e.g.*, either the A-half of the fusion protein, or the A-half of the dimer not part of the fusion protein). In some embodiments, the protein dimer comprises (i) a fusion protein comprising a nuclease-inactivated Cas9 and a gRNA binding domain of Cas9 (*e.g.*, a nuclease-inactivated Cas9 fused to a Cas9 A-half), and (ii) a fusion protein comprising a nuclease-inactivated Cas9 and a DNA cleavage domain (*e.g.*, a nuclease-inactivated Cas9 fused to a Cas9 B-half). In some embodiments, the dimer is associated with one or more distinct gRNAs. For example, in some embodiments, the dimer is associated with two or three gRNAs. In some embodiments, the dimer is associated with three gRNAs. For example, upon binding of one nuclease-inactivated Cas9:gRNA to a region of a nucleic acid target, and binding of the other nuclease-inactivated Cas9:gRNA to a second region of the nucleic acid target, the split Cas9 halves (*e.g.*, A-half and B-half of the fusion proteins) dimerize and bind a third gRNA complementary to a third region of the nucleic acid target, to become a fully active Cas9 nuclease leading to cleavage of the target DNA.

[00123] In some embodiments, a method for site-specific cleavage of a nucleic acid comprises contacting a nucleic acid (*e.g.*, DNA) with a minimized Cas9 protein (*e.g.*, as described herein) associated with a gRNA.

[00124] In some embodiments, any of the methods provided herein can be performed on DNA in a cell, for example a bacterium, a yeast cell, or a mammalian cell. In some embodiments, the DNA contacted by any Cas9 protein provided herein is in a eukaryotic cell. In some embodiments, the methods can be performed on a cell or tissue *in vitro* or *ex vivo*. In

some embodiments, the eukaryotic cell is in an individual, such as a patient or research animal. In some embodiments, the individual is a human.

Methods for site-specific recombination

[00125] In another embodiment of this disclosure, methods for site-specific nucleic acid (*e.g.*, DNA) recombination are provided. In some embodiments, the methods are useful for inducing recombination of or between two or more regions of two or more nucleic acid (*e.g.*, DNA) molecules. In other embodiments, the methods are useful for inducing recombination of or between two or more regions in a single nucleic acid molecule (*e.g.*, DNA). In some embodiments, the recombination of one or more target nucleic acid molecules requires the formation of a tetrameric complex at the target site. Typically, the tetramer comprises four (4) inventive RNA-guided recombinase fusion proteins (*e.g.*, a complex of any four inventive recombinase fusion protein provided herein). In some embodiments, each recombinase fusion protein of the tetramer targets a particular DNA sequence via a distinct gRNA bound to each recombinase fusion protein (See, *e.g.*, Figure 5).

[00126] In some embodiments, the method for site-specific recombination between two DNA molecules comprises (a) contacting a first DNA with a first RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain binds a first gRNA that hybridizes to a region of the first DNA; (b) contacting the first DNA with a second RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain of the second fusion protein binds a second gRNA that hybridizes to a second region of the first DNA; (c) contacting a second DNA with a third RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain of the third fusion protein binds a third gRNA that hybridizes to a region of the second DNA; and (d) contacting the second DNA with a fourth RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain of the fourth fusion protein binds a fourth gRNA that hybridizes to a second region of the second DNA. The binding of the fusion proteins in steps (a) - (d) results in the tetramerization of the recombinase catalytic domains of the fusion proteins, such that the DNAs are recombined. In some embodiments, the gRNAs of steps (a) and (b) hybridize to opposing strands of the first DNA, and the gRNAs of steps (c) and (d) hybridize to opposing strands of the second DNA. In some embodiments, the target sites of the gRNAs of steps (a) - (d) are spaced to allow for tetramerization of the recombinase catalytic domains. For example, in some embodiments, the target sites of the gRNAs of steps (a) - (d) are no more than 10, no more 15, no more than 20, no more than 25, no more than 30, no more than

40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart. In some embodiments, the two regions of the two DNA molecules being recombined share homology, such that the regions being recombined are at least 80%, at least 90%, at least 95%, at least 98%, or are 100% homologous.

[00127] In another embodiment, methods for site-specific recombination between two regions of a single DNA molecule are provided. In some embodiments, the methods comprise (a) contacting a DNA with a first RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain binds a first gRNA that hybridizes to a region of the DNA; (b) contacting the DNA with a second RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain of the second fusion protein binds a second gRNA that hybridizes to a second region of the DNA; (c) contacting the DNA with a third RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain of the third fusion protein binds a third gRNA that hybridizes to a third region of the DNA; and (d) contacting the DNA with a fourth RNA-guided recombinase fusion protein, wherein the nuclease-inactivated Cas9 domain of the fourth fusion protein binds a fourth gRNA that hybridizes to a fourth region of the DNA. The binding of the fusion proteins in steps (a) - (d) results in the tetramerization of the recombinase catalytic domains of the fusion proteins, such that the DNA is recombined. In some embodiments, two of the gRNAs of steps (a) - (d) hybridize to the same strand of the DNA, and the other two gRNAs of steps (a)- (d) hybridize to the opposing strand of the DNA. In some embodiments, the gRNAs of steps (a) and (b) hybridize to regions of the DNA that are no more 10, no more than 15, no more than 20, no more than 25, no more than 30, no more than 40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart, and the gRNAs of steps (c) and (d) hybridize to regions of the DNA that are no more than 10, no more than 15, no more than 20, no more than 25, no more than 30, no more than 40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart. In some embodiments, the two regions of the DNA molecule being recombined share homology, such that the regions being recombined are at least 80%, at least 90%, at least 95%, at least 98%, or are 100% homologous.

[00128] In some embodiments, any of the inventive methods for site-specific recombination are amenable for inducing recombination, such that the recombination results in excision (*e.g.*, a segment of DNA is excised from a target DNA molecule), insertion (*e.g.*, a segment of DNA is inserted into a target DNA molecule), inversion (*e.g.*, a segment of DNA is inverted in a target DNA molecule), or translocation (*e.g.*, the exchange of DNA

segments between one or more target DNA molecule(s)). In some embodiments, the particular recombination event (*e.g.*, excision, insertion, inversion, translocation, *etc.*) depends, *inter alia*, on the orientation (*e.g.*, with respect to the target DNA molecule(s)) of the bound RNA-guided recombinase fusion protein(s). In some embodiments, the orientation, or direction, in which a RNA-guided recombinase fusion protein binds a target nucleic acid can be controlled, *e.g.*, by the particular sequence of the gRNA bound to the RNA-guided recombinase fusion protein(s). Methods for controlling or directing a particular recombination event are known in the art, and include, for example, those described by Turan and Bode, “Site-specific recombinases: from tag-and-target- to tag-and-exchange-based genomic modifications.” *FASEB J.* 2011; Dec;25(12):4088-107, the entire contents of which are hereby incorporated by reference.

[00129] In some embodiments, any of the methods for site-specific recombination can be performed *in vivo* or *in vitro*. In some embodiments, any of the methods for site-specific recombination are performed in a cell (*e.g.*, recombine genomic DNA in a cell). The cell can be prokaryotic or eukaryotic. The cell, such as a eukaryotic cell, can be in an individual, such as a subject, as described herein (*e.g.*, a human subject). The methods described herein are useful for the genetic modification of cells *in vitro* and *in vivo*, for example, in the context of the generation of transgenic cells, cell lines, or animals, or in the alteration of genomic sequence, *e.g.*, the correction of a genetic defect, in a cell in or obtained from a subject. In some embodiments, a cell obtained from a subject and modified according to the methods provided herein, is re-introduced into a subject (*e.g.*, the same subject), *e.g.*, to treat a disease, or for the production of genetically modified organisms in agriculture or biological research.

[00130] In applications in which it is desirable to recombine two or more nucleic acids so as to insert a nucleic acid sequence into a target nucleic acid, a nucleic acid comprising a donor sequence to be inserted is also provided, *e.g.*, to a cell. By a “donor sequence” it is meant a nucleic acid sequence to be inserted at the target site induced by one or more RNA-guided recombinase fusion protein(s). In some embodiments, *e.g.*, in the context of genomic modifications, the donor sequence will share homology to a genomic sequence at the target site, *e.g.*, 1%, 2%, 3%, 4%, 5%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 85%, 90%, 95%, or 100% homology with the nucleotide sequences flanking the target site, *e.g.*, within about 100 bases or less of the target site, *e.g.* within about 90 bases, within about 80 bases, within about 70 bases, within about 60 bases, within about 50 bases, within about 40 bases, within about 30 bases, within about 15 bases, within about 10 bases, within about 5 bases, or

immediately flanking the target site. In some embodiments, the donor sequence does not share any homology with the target nucleic acid, *e.g.*, does not share homology to a genomic sequence at the target site. Donor sequences can be of any length, *e.g.*, 10 nucleotides or more, 50 nucleotides or more, 100 nucleotides or more, 250 nucleotides or more, 500 nucleotides or more, 1000 nucleotides or more, 5000 nucleotides or more, 10000 nucleotides or more, 100000 nucleotides or more, *etc.*

[00131] Typically, the donor sequence is not identical to the target sequence that it replaces or is inserted into. In some embodiments, the donor sequence contains at least one or more single base changes, insertions, deletions, inversions or rearrangements with respect to the target sequence (*e.g.*, target genomic sequence). In some embodiments, donor sequences also comprise a vector backbone containing sequences that are not homologous to the DNA region of interest and that are not intended for insertion into the DNA region of interest.

[00132] The donor sequence may comprise certain sequence differences as compared to the target (*e.g.*, genomic) sequence, for example restriction sites, nucleotide polymorphisms, selectable markers (*e.g.*, drug resistance genes, fluorescent proteins, enzymes *etc.*), which can be used to assess for successful insertion of the donor sequence at the target site or in some cases may be used for other purposes (*e.g.*, to signify expression at the targeted genomic locus). In some embodiments, if located in a coding region, such nucleotide sequence differences will not change the amino acid sequence, or will make silent amino acid changes (*e.g.*, changes which do not affect the structure or function of the protein). In some embodiments, these sequences differences may include flanking recombination sequences such as FLPs, loxP sequences, or the like, that can be activated at a later time for removal of *e.g.*, a marker sequence.

[00133] The donor sequence may be provided to the cell as single-stranded DNA, single-stranded RNA, double-stranded DNA, or double-stranded RNA. It may be introduced into a cell in linear or circular form. If introduced in linear form, the ends of the donor sequence may be protected (*e.g.*, from exonucleolytic degradation) by methods known to those of skill in the art. For example, one or more dideoxynucleotide residues are added to the 3' terminus of a linear molecule and/or self-complementary oligonucleotides are ligated to one or both ends. See, *e.g.*, Chang *et al.*, *Proc. Natl. Acad. Sci. USA*. 1987; 84:4959-4963; Nehls *et al.*, *Science*. 1996; 272:886-889. In some embodiments, a donor sequence can be introduced into a cell as part of a vector molecule having additional sequences such as, for example, replication origins, promoters and genes encoding antibiotic resistance. In some

embodiments, donor sequences can be introduced as naked nucleic acid, as nucleic acid complexed with an agent such as a liposome or poloxamer, or can be delivered by viruses (*e.g.*, adenovirus, AAV, *etc.*).

Polynucleotides, Vectors, Cells, Kits

[00134] In another embodiment of this disclosure, polynucleotides encoding one or more of the inventive proteins and/or gRNAs are provided. For example, polynucleotides encoding any of the proteins described herein are provided, *e.g.*, for recombinant expression and purification of isolated nucleases and recombinases, *e.g.*, comprising Cas9 variants. In some embodiments, an isolated polynucleotide comprises one or more sequences encoding a Cas9 half site (*e.g.*, A-half and/or B-half). In some embodiments, an isolated polynucleotide comprises one or more sequences encoding a Cas9 fusion protein, for example, any of the Cas9 fusion proteins described herein (*e.g.*, those comprising a nuclease-inactivated Cas9). In some embodiments, an isolated polynucleotides comprises one or more sequences encoding a gRNA, alone or in combination with a sequence encoding any of the proteins described herein.

[00135] In some embodiments, vectors encoding any of the proteins described herein are provided, *e.g.*, for recombinant expression and purification of Cas9 proteins, and/or fusions comprising Cas9 proteins (*e.g.*, variants). In some embodiments, the vector comprises or is engineered to include an isolated polynucleotide, *e.g.*, those described herein. In some embodiments, the vector comprises one or more sequences encoding a Cas9 protein (as described herein), a gRNA, or combinations thereof, as described herein. Typically, the vector comprises a sequence encoding an inventive protein operably linked to a promoter, such that the fusion protein is expressed in a host cell.

[00136] In some embodiments, cells are provided, *e.g.*, for recombinant expression and purification of any of the Cas9 proteins provided herein. The cells include any cell suitable for recombinant protein expression, for example, cells comprising a genetic construct expressing or capable of expressing an inventive protein (*e.g.*, cells that have been transformed with one or more vectors described herein, or cells having genomic modifications, for example, those that express a protein provided herein from an allele that has been incorporated in the cell's genome). Methods for transforming cells, genetically modifying cells, and expressing genes and proteins in such cells are well known in the art, and include those provided by, for example, Green and Sambrook, *Molecular Cloning: A Laboratory Manual* (4th ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.

(2012)) and Friedman and Rossi, *Gene Transfer: Delivery and Expression of DNA and RNA, A Laboratory Manual* (1st ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (2006)).

[00137] Some aspects of this disclosure provide kits comprising a Cas9 variant and/or nuclease and/or recombinase, as provided herein. In some embodiments, the kit comprises a polynucleotide encoding an inventive Cas9 variant, nuclease, and/or recombinase, *e.g.*, as provided herein. In some embodiments, the kit comprises a vector for recombinant protein expression, wherein the vector comprises a polynucleotide encoding any of the proteins provided herein. In some embodiments, the kit comprises a cell (*e.g.*, any cell suitable for expressing Cas9 proteins or fusions comprising Cas9 proteins, such as bacterial, yeast, or mammalian cells) that comprises a genetic construct for expressing any of the proteins provided herein. In some embodiments, any of the kits provided herein further comprise one or more gRNAs and/or vectors for expressing one or more gRNAs. In some embodiments, the kit comprises an excipient and instructions for contacting the nuclease and/or recombinase with the excipient to generate a composition suitable for contacting a nucleic acid with the nuclease and/or recombinase such that hybridization to and cleavage and/or recombination of a target nucleic acid occurs. In some embodiments, the composition is suitable for delivering a Cas9 protein to a cell. In some embodiments, the composition is suitable for delivering a Cas9 protein to a subject. In some embodiments, the excipient is a pharmaceutically acceptable excipient.

[00138] The function and advantage of these and other embodiments of the present invention will be more fully understood from the Examples below. The following Examples are intended to illustrate the benefits of the present invention and to describe particular embodiments, but are not intended to exemplify the full scope of the invention. Accordingly, it will be understood that the Examples are not meant to limit the scope of the invention.

EXAMPLES

Example 1: Fusion of Inactivated Cas9 to FokI Nuclease Improves Genome Modification Specificity

Methods:

Oligonucleotides and PCR

[00139] All oligonucleotides were purchased from Integrated DNA Technologies (IDT). Oligonucleotide sequences are listed in Table 1. PCR was performed with 0.4 μ L of

2 U/ μ L Phusion Hot Start Flex DNA polymerase (NEB) in 50 μ L with 1x HF Buffer, 0.2 mM dNTP mix (0.2 mM dATP, 0.2 mM dCTP, 0.2 mM dGTP, 0.2 mM dTTP) (NEB), 0.5 μ M of each primer and a program of: 98 °C, 1 min; 35 cycles of [98 °C, 15 s; 65 °C, 15 s; 72 °C, 30 s] unless otherwise noted.

Construction of FokI-dCas9, Cas9 Nickase and gRNA Expression Plasmids

[00140] The human codon-optimized *streptococcus pyogenes* Cas9 nuclease with NLS and 3xFLAG tag (Addgene plasmid 43861)² was used as the wild-type Cas9 expression plasmid. PCR (72 °C, 3 min) products of wild-type Cas9 expression plasmid as template with Cas9_Exp primers listed in Table 1 below were assembled with Gibson Assembly Cloning Kit (New England Biolabs) to construct Cas9 and FokI-dCas9 variants. Expression plasmids encoding a single gRNA construct (gRNA G1 through G13) were cloned as previously described. Briefly, gRNA oligonucleotides listed in **Table 1** containing the 20-bp protospacer target sequence were annealed and the resulting 4-bp overhangs were ligated into BsmBI-digested gRNA expression plasmid. gRNA expression plasmids encoding expression of two separate gRNA constructs from separate promoters on a single plasmid were cloned in a two-step process. First, one gRNA (gRNA E1, V1, C1, C3, H1, G1, G2 or G3) was cloned as above and used as template for PCR (72 °C, 3 min) with PCR_Pla-fwd and PCR_Pla-rev primers, 1 μ l DpnI (NEB) was added, and the reaction was incubated at 37 °C for 30 min and then subjected to QIAquick PCR Purification Kit (Qiagen) for the “1st gRNA + vector DNA”. PCR (72 °C, 3 min) of 100 pg of BsmBI-digested gRNA expression plasmid as template with PCR_gRNA-fwd1, PCR_gRNA-rev1, PCR_gRNA-rev2 and appropriate PCR_gRNA primer listed in **Table 1** was DpnI treated and purified as above for the “2nd gRNA instert DNA”. ~200 ng of “1st gRNA + vector DNA” and ~200 ng of “2nd gRNA instert DNA” were blunt-end ligated in 1x T4 DNA Ligase Buffer, 1 μ l of T4 DNA Ligase (400 U/ μ l, NEB) in a total volume of 20 μ l at room temperature (~21 °C) for 15 min. For all cloning, 1 μ l of ligation or assembly reaction was transformed into Mach1 chemically competent cells (Life Technologies).

Table 1. Oligonucleotides. ‘/5Phos/’ indicates 5’ phosphorylated oligonucleotides.

dCas9-NLS-FokI primers:	
Cas9_Exp_CNF_Fok1+Plas-Fwd	CGGCGAGATAAACTTTTAA TGACCGGTCATCATCACCA (SEQ ID NO:26)
Cas9_Exp_CNF_Cas9coD10-Rev	CCAACGGAATTAGTGCCGATAGCTAAACCAATAGAATACTTTTATC (SEQ ID NO:27)

Cas9_Exp_CNF_Cas9coD10-Fwd	GATAAAAAGTATTCTATTGGTTTAGCTATCGGCACTAATTCCGTTGG (SEQ ID NO:28)
Cas9_Exp_CNF_Cas9coH850-Rev	TTCAAAAAGGATTGGGGTACAATGGCATCGACGTCGTAATCAGATAAAC (SEQ ID NO:29)
Cas9_Exp_CNF_Cas9coH850-Fwd	GTTTATCTGATTACGACGTCGATGCCATTGTACCCCAATCCTTTTTGA A (SEQ ID NO:30)
Cas9_Exp_CNF_(Cas9)NLS+GGs-Fok-Rev	TTGGGATCCAGAACCTCCTCCTGCAGCCTTGTCATCG (SEQ ID NO:31)
Cas9_Exp_CNF_(Cas9)NLS+GGs3-Fok-Rev	TTGGGATCCAGAACCTCC GCTGCCGCCACTTCCACCTGA TCCTGCAGCCTTGTCATCG (SEQ ID NO:32)
Cas9_Exp_CNF_(Cas9)NLS+GGs-Fok-Fwd	CGATGACAAGGCTGCAGGAGGAGGTTCTGGATCCCAA (SEQ ID NO:33)
Cas9_Exp_CNF_(Cas9)NLS+GGs3-Fok-Fwd	CGATGACAAGGCTGCAGGA TCAGGTGGAAGTGGCGGCAGC GGAGGTTCTGGATCCCAA (SEQ ID NO:34)
Cas9_Exp_CNF_Fok1+Plas-Rev	TGGTGATGATGACCGGTCA TTAAAAGTTTATCTCGCCG (SEQ ID NO:35)
NLS-dCas9-FokI primers:	
Cas9_Exp_NCF_Fok1+Plas-Fwd	CGGCGAGATAAACTTTTAA TGACCGGTCATCATCACCA (SEQ ID NO:36)
Cas9_Exp_NCF_PlasS+FLAG(NLS-Fok1-Rev	TAGGGAGAGCCGCCACCATGGACTACAAAGACCATGACGG (SEQ ID NO:37)
Cas9_Exp_NCF_NLS+Cas9coD10-Rev	TAAACCAATAGAATACTTTTTATC CATAGGTACCCCGCGGTGAATG (SEQ ID NO:38)
Cas9_Exp_NCF_Cas9coD10-Fwd	GATAAAAAGTATTCTATTGGTTTAGCTATCGGCACTAATTCCGTTGG (SEQ ID NO:39)
Cas9_Exp_NCF_Cas9coH850-Rev	TTCAAAAAGGATTGGGGTACAATGGCATCGACGTCGTAATCAGATAAAC (SEQ ID NO:40)
Cas9_Exp_NCF_Cas9coH850-Fwd	GTTTATCTGATTACGACGTCGATGCCATTGTACCCCAATCCTTTTTGA A (SEQ ID NO:41)
Cas9_Exp_NCF_Cas9End+GGs-Fok-Rev	TTGGGATCCAGAACCTCCGTCACCCCCAAGCTGTG (SEQ ID NO:42)
Cas9_Exp_NCF_Cas9End+GGs3-Fok-Rev	TTGGGATCCAGAACCTCC GCTGCCGCCACTTCCACCTGA GTCACCCCCAAGCTGTG (SEQ ID NO:43)
Cas9_Exp_NCF_Cas9End+GGs-Fok-Fwd	CACAGCTTGGGGGTGACGGAGGTTCTGGATCCCAA (SEQ ID NO:44)
Cas9_Exp_NCF_Cas9End+GGs3-Fok-Fwd	CACAGCTTGGGGGTGAC TCAGGTGGAAGTGGCGGCAGC GGAGGTTCTGGATCCCAA (SEQ ID NO:45)
Cas9_Exp_NCF_Fok1+Plas-Rev	TGGTGATGATGACCGGTCA TTAAAAGTTTATCTCGCCG (SEQ ID NO:46)
FokI-dCas9-NLS primers:	
Cas9_Exp_FCN_PlasS+Fok-Fwd	TAGGGAGAGCCGCCACCATGGGATCCCAACTAGTCAAAAAG (SEQ ID NO:47)
Cas9_Exp_FCN_Fok1GGS+Cas-Rev	ACCAATAGAATACTTTTTATCCATGCTGCCACCAAAGTTTATCTC (SEQ ID NO:48)
Cas9_Exp_FCN_Fok1GGS3+Cas-Rev	ACCAATAGAATACTTTTTATCCATGCTGCCGCCACTTCCACCTG (SEQ ID NO:49)
Cas9_Exp_FCN_Cas9coD10-Fwd	GATAAAAAGTATTCTATTGGTTTAGCTATCGGCACTAATTCCGTTGG (SEQ ID NO:50)
Cas9_Exp_FCN_Cas9coH850-Rev	CCAACGGAATTAGTGCCGATAGCTAAACCAATAGAATACTTTTTATC (SEQ ID NO:51)
Cas9_Exp_FCN_Cas9coH850-Fwd	GTTTATCTGATTACGACGTCGATGCCATTGTACCCCAATCCTTTTTGA A (SEQ ID NO:52)
Cas9_Exp_FCN_Cas9End+PlasmidEn-Rev	TGGTGATGATGACCGGTCA GTCACCCCCAAGCTGTG (SEQ ID NO:53)
Cas9_Exp_FCN_Cas9End+PlasmidEn-Fwd	CACAGCTTGGGGGTGAC TGACCGGTCATCATCACCA (SEQ ID NO:54)
Cas9_Exp_FCN_PlasS+Fok-Rev	CTTTTGACTAGTTGGGATCCCATGGTGGCGGCTCTCCCTA (SEQ ID NO:55)

gRNA_G1-top	ACACCCCTCGAACTTCACCTCGGCGG (SEQ ID NO:56)
gRNA_G2-top	ACACCGTCGCCCTCGAACTTCACCTG (SEQ ID NO:57)
gRNA_G3-top	ACACCCAGCTCGATGCGGTTACACCAG (SEQ ID NO:58)
gRNA_G4-top	ACACCGGTGAACCGCATCGAGCTGAG (SEQ ID NO:59)
gRNA_G5-top	ACACCGCTGAAGGGCATCGACTTCAG (SEQ ID NO:60)
gRNA_G6-top	ACACCGGCATCGACTTCAAGGAGGAG (SEQ ID NO:61)
gRNA_G7-top	ACACCCAAGGAGGACGGCAACATCCG (SEQ ID NO:62)
gRNA_G8-top	ACACCACCATCTTCTTCAAGGACGAG (SEQ ID NO:63)
gRNA_G9-top	ACACCCAATAACAAGACCCGCGCCGG (SEQ ID NO:64)
gRNA_G10-top	ACACCCCGCGCCGAGGTGAAGTTCGG (SEQ ID NO:65)
gRNA_G11-top	ACACCGAAGTTCGAGGGCGACACCCG (SEQ ID NO:66)
gRNA_G12-top	ACACCTTCGAACTTCACCTCGGCGCG (SEQ ID NO:67)
gRNA_G13-top	ACACCTCAGCTCGATGCGGTTACCCG (SEQ ID NO:68)
gRNA_G14-top	ACACCCGATGCCCTTCAGCTCGATGG (SEQ ID NO:69)
gRNA_G1-bottom	AAAACCGCCGAGGTGAAGTTCGAGGG (SEQ ID NO:70)
gRNA_G2-bottom	AAAACAGGTGAAGTTCGAGGGCGACG (SEQ ID NO:71)
gRNA_G3-bottom	AAAACCTGGTGAACCGCATCGAGCTGG (SEQ ID NO:72)
gRNA_G4-bottom	AAAACCTCAGCTCGATGCGGTTACCCG (SEQ ID NO:73)
gRNA_G5-bottom	AAAACCTGAAGTCGATGCCCTTCAGCG (SEQ ID NO:74)
gRNA_G6-bottom	AAAACCTCCTCCTTGAAGTCGATGCCG (SEQ ID NO:75)
gRNA_G7-bottom	AAAACGGATGTTGCCGTCCTCCTTGG (SEQ ID NO:76)
gRNA_G8-bottom	AAAACCTCGTCCTTGAAGAAGATGGTG (SEQ ID NO:77)
gRNA_G9-bottom	AAAACCGGCGCGGGTCTTGTAGTTGG (SEQ ID NO:78)
gRNA_G10-bottom	AAAACCGAACTTCACCTCGGCGCGGG (SEQ ID NO:79)
gRNA_G11-bottom	AAAACGGGTGTCGCCCTCGAACTTCG (SEQ ID NO:80)
gRNA_G12-bottom	AAAACGCGCCGAGGTGAAGTTCGAAG (SEQ ID NO:81)
gRNA_G13-bottom	AAAACGGTGAACCGCATCGAGCTGAG (SEQ ID NO:82)
gRNA_G14-bottom	AAAACCATCGAGCTGAAGGGCATCGG (SEQ ID NO:83)
gRNA_C1-top	ACACCTGGCCTGCTTGCTAGACTTGG (SEQ ID NO:84)
gRNA_C3-top	ACACCGCAGATGTAGTGTTTCCACAG (SEQ ID NO:85)
gRNA_H1-top	ACACCCTTGCCCCACAGGGCAGTAAG (SEQ ID NO:86)
gRNA_E1-top	ACACCGAGTCCGAGCAGAAGAAGAAG (SEQ ID NO:87)
gRNA_V1-top	ACACCGGGTGGGGGGAGTTTGCTCCG (SEQ ID NO:88)
gRNA_C1-bottom	AAAACCAAGTCTAGCAAGCAGGCCAG (SEQ ID NO:89)
gRNA_C3-bottom	AAAACCTGTGGAAACACTACATCTGCG (SEQ ID NO:90)
gRNA_H1-bottom	AAAACCTTCTTCTTCTGCTCGGACTCG (SEQ ID NO:91)
gRNA_E1-bottom	AAAACCTTACTGCCCTGTGGGGCAAGG (SEQ ID NO:92)
gRNA_V1-bottom	AAAACGGAGCAAACCTCCCCCACCAG (SEQ ID NO:93)
PCR_Pla-fwd	AGG AAA GAA CAT GTG AGC AAA AG (SEQ ID NO:94)
PCR_Pla-rev	CAGCGAGTCAGTGAGCGA (SEQ ID NO:95)
PCR_gRNA-fwd1	CTGTACAAAAAAGCAGGCTTTA (SEQ ID NO:96)
PCR_gRNA-rev1	AACGTAGGTCTCTACCGCTGTACAAAAAAGCAGGCTTTA (SEQ ID NO:97)
PCR_gRNA-rev2	AAAAAAAGCACCGACTCGGTGCCACTTTTTCAAGTTGATAACGGACT AGCCTTATTTAACTTGCTATTCTAGCTCTAAAAC (SEQ ID NO:98)

PCR_gRNA_G1	TTGCTATTTCTAGCTCTAAAACCGCCGAGGTGAAGTTCGAGGGGTGT TTCGTCCTTTCCA (SEQ ID NO:99)
PCR_gRNA_G2	TTGCTATTTCTAGCTCTAAAACAGGTGAAGTTCGAGGGCGACGGTGT TTCGTCCTTTCCA (SEQ ID NO:100)
PCR_gRNA_G3	TTGCTATTTCTAGCTCTAAAACCTGGTGAACCGCATCGAGCTGGGTGT TTCGTCCTTTCCA (SEQ ID NO:101)
PCR_gRNA_G4	TTGCTATTTCTAGCTCTAAAACCTCAGCTCGATGCGGTTCACCGGTGTT TCGTCCTTTCCA (SEQ ID NO:102)
PCR_gRNA_G5	TTGCTATTTCTAGCTCTAAAACCTGAAGTCGATGCCCTTCAGCGGTGTT TCGTCCTTTCCA (SEQ ID NO:103)
PCR_gRNA_G6	TTGCTATTTCTAGCTCTAAAACCTCCTCCTTGAAGTCGATGCCGGTGT TCGTCCTTTCCA (SEQ ID NO:104)
PCR_gRNA_G7	TTGCTATTTCTAGCTCTAAAACGGATGTTGCCGTCCTCCTTGGGTGTT TCGTCCTTTCCA (SEQ ID NO:105)
PCR_gRNA_C2	TTGCTATTTCTAGCTCTAAAACGCTTGAGGGAGATGAGGACTGGTGT TTCGTCCTTTCCA (SEQ ID NO:106)
PCR_gRNA_C4	TTGCTATTTCTAGCTCTAAAACATGACTGTGAAGAGCTTCACGGTGT TTCGTCCTTTCCA (SEQ ID NO:107)
PCR_gRNA_E2	TTGCTATTTCTAGCTCTAAAACGAGGACAAAGTACAAACGGCGGTGT TTCGTCCTTTCCA (SEQ ID NO:108)
PCR_gRNA_E3	TTGCTATTTCTAGCTCTAAAACGAACCGGAGGACAAAGTACAGGTGT TTCGTCCTTTCCA (SEQ ID NO:109)
PCR_gRNA_H2	TTGCTATTTCTAGCTCTAAAACCACTTCATCCACGGGTGTT TCGTCCTTTCCA (SEQ ID NO:110)
PCR_gRNA_H3	TTGCTATTTCTAGCTCTAAAACGGGCCTCACCACCAACTTCAGGTGT TTCGTCCTTTCCA (SEQ ID NO:111)
PCR_gRNA_H4	TTGCTATTTCTAGCTCTAAAACGCCAGGGCCTCACCACCAAGGTGT TTCGTCCTTTCCA (SEQ ID NO:112)
PCR_gRNA_H5	TTGCTATTTCTAGCTCTAAAACACCTGCCAGGGCCTCACCAGGTGT TTCGTCCTTTCCA (SEQ ID NO:113)
PCR_gRNA_H6	TTGCTATTTCTAGCTCTAAAACCTGATACCAACCTGCCAGGGGGTGT TTCGTCCTTTCCA (SEQ ID NO:114)
PCR_gRNA_H7	TTGCTATTTCTAGCTCTAAAACCTGTCTTGTAACCTTGGTGT TCGTCCTTTCCA (SEQ ID NO:115)
PCR_gRNA_V2	TTGCTATTTCTAGCTCTAAAACGCTCTGGCTAAAGAGGGAATGGTGT TTCGTCCTTTCCA (SEQ ID NO:116)
PCR_gRNA_V3	TTGCTATTTCTAGCTCTAAAACCGGCTCTGGCTAAAGAGGGAGGTGT TTCGTCCTTTCCA (SEQ ID NO:117)
PCR_gRNA_V4	TTGCTATTTCTAGCTCTAAAACCTCTGCACACCCCGGCTCTGGGGTGT TCGTCCTTTCCA (SEQ ID NO:118)
Survey_GFP-fwd	TACGGCAAGCTGACCCTGAA (SEQ ID NO:119)
Survey_GFP-rev	GTCCATGCCGAGAGTGATCC (SEQ ID NO:120)
Survey_CLTA-fwd	GCCAGGGGCTGTTATCTTGG (SEQ ID NO:121)
Survey_CLTA-rev	ATGCACAGAAGCACAGGTTGA (SEQ ID NO:122)
Survey_EMX-fwd	CTGTGTCCTCTTCCTGCCCT (SEQ ID NO:123)
Survey_EMX-rev	CTCTCCGAGGAGAAGGCCAA (SEQ ID NO:124)
Survey_HBB-fwd	GGTAGACCACCAGCAGCCTA (SEQ ID NO:125)
Survey_HBB-rev	CAGTGCCAGAAGAGCCAAGG (SEQ ID NO:126)
Survey_VEGF-fwd	CCACACAGCTTCCCGTTCTC (SEQ ID NO:127)
Survey_VEGF-rev	GAGAGCCGTTCCCTCTTTGC (SEQ ID NO:128)
HTS_EXM_ON-fwd	CCTCCCCATTGGCCTGCTTC (SEQ ID NO:129)
HTS_EXM_Off1-fwd	TCGTCCTGCTCTCACTTAGAC (SEQ ID NO:130)
HTS_EXM_Off2-fwd	TTTTGTGGCTTGGCCCCAGT (SEQ ID NO:131)
HTS_EXM_Off3-fwd	TGCAGTCTCATGACTTGGCCT (SEQ ID NO:132)

HTS_EXM_Off4-fwd	TTCTGAGGGCTGCTACCTGT (SEQ ID NO:133)
HTS_VEFG_ON-fwd	ACATGAAGCAACTCCAGTCCCA (SEQ ID NO:134)
HTS_EXM_Off1-fwd	AGCAGACCCACTGAGTCAACTG (SEQ ID NO:135)
HTS_EXM_Off2-fwd	CCCGCCACAGTCGTGTCAT (SEQ ID NO:136)
HTS_EXM_Off3-fwd	CGCCCCGGTACAAGGTGA (SEQ ID NO:137)
HTS_EXM_Off4-fwd	GTACCGTACATTGTAGGATGTTT (SEQ ID NO:138)
HTS_CLTA2_ON-fwd	CCTCATCTCCCTCAAGCAGGC (SEQ ID NO:139)
HTS_CLTA2_Off1-fwd	ATTCTGCTCTTGAGGTTATTTGT (SEQ ID NO:140)
HTS_CLTA2_Off2-fwd	CACCTCTGCCTCAAGAGCAGAAAA (SEQ ID NO:141)
HTS_CLTA2_Off3-fwd	TGTGTGTGTGTGTGTGTAGGACT (SEQ ID NO:142)
HTS_EXM_ON-rev	TCATCTGTGCCCCCTCCCTCC (SEQ ID NO:143)
HTS_EXM_Off-rev	CGAGAAGGAGGTGCAGGAG (SEQ ID NO:144)
HTS_EXM_Off-rev	CGGGAGCTGTTTCAGAGGCTG (SEQ ID NO:145)
HTS_EXM_Off-rev	CTCACCTGGGCGAGAAAGGT (SEQ ID NO:146)
HTS_EXM_Off-rev	AAAACCTCAAAGAAATGCCCAATCA (SEQ ID NO:147)
HTS_VEFG_ON-rev	AGACGCTGCTCGCTCCATTC (SEQ ID NO:148)
HTS_EXM_Off1-rev	ACAGGCATGAATCACTGCACCT (SEQ ID NO:149)
HTS_EXM_Off2-rev	GCGGCAACTTCAGACAACCGA (SEQ ID NO:150)
HTS_EXM_Off3-rev	GACCCAGGGGCACCAAGTT (SEQ ID NO:151)
HTS_EXM_Off4-rev	CTGCCTTCATTGCTTAAAAGTGGAT (SEQ ID NO:152)
HTS_CLTA2_ON-rev	ACAGTTGAAGGAAGGAAACATGC (SEQ ID NO:153)
HTS_CLTA2_Off1-rev	GCTGCATTTGCCCATTTCCA (SEQ ID NO:154)
HTS_CLTA2_Off2-rev	GTTGGGGGAGGAGGAGCTTAT (SEQ ID NO:155)
HTS_CLTA2_Off3-rev	CTAAGAGCTATAAGGGCAAATGACT (SEQ ID NO:156)

Modification of Genomic GFP

[00141] HEK293-GFP stable cells (GenTarget) were used as a cell line constitutively expressing an Emerald GFP gene (GFP) integrated on the genome. Cells were maintained in Dulbecco's modified Eagle medium (DMEM, Life Technologies) supplemented with 10% (vol/vol) fetal bovine serum (FBS, Life Technologies) and penicillin/streptomycin (1x, Amresco). 5×10^4 HEK293-GFP cells were plated on 48-well collagen coated Biocoat plates (Becton Dickinson). One day following plating, cells at ~75% confluence were transfected with Lipofectamine 2000 (Life Technologies) according to the manufacturer's protocol. Briefly, 1.5 μ L of Lipofectamine 2000 was used to transfect 950 ng of total plasmid (Cas9 expression plasmid plus gRNA expression plasmids). 700 ng of Cas9 expression plasmid, 125 ng of one gRNA expression plasmid and 125 ng of the paired gRNA expression plasmid with the pairs of targeted gRNAs listed in Figure 6D and Figure 9A. Separate wells were transfected with 1 μ g of a near-infrared iRFP670 (Addgene plasmid 45457)³² as a transfection control. 3.5 days following transfection, cells were trypsinized and resuspended in DMEM supplemented with 10% FBS and analyzed on a C6 flow cytometer (Accuri) with

a 488 nm laser excitation and 520 nm filter with a 20 nm band pass. For each sample, transfections and flow cytometry measurements were performed once.

T7 Endonuclease I Surveyor Assays of Genomic Modifications

[00142] HEK293-GFP stable cells were transfected with Cas9 expression and gRNA expression plasmids as described above. A single plasmid encoding two separate gRNAs was transfected. For experiments titrating the total amount of expression plasmids (Cas9 expression + gRNA expression plasmid), 700/250, 350/125, 175/62.5, 88/31 ng of Cas9 expression plasmid/ng of gRNA expression plasmid were combined with inert carrier plasmid, pUC19 (NEB), as necessary to reach a total of 950 ng transfected plasmid DNA.

[00143] Genomic DNA was isolated from cells 2 days after transfection using a genomic DNA isolation kit, DNAdvance Kit (Agencourt). Briefly, cells in a 48-well plate were incubated with 40 μ L of trypsin for 5 min at 37 °C. 160 μ L of DNAdvance lysis solution was added and the solution incubated for 2 hr at 55 °C and the subsequent steps in the Agencourt DNAdvance kit protocol were followed. 40 ng of isolated genomic DNA was used as template to PCR amplify the targeted genomic loci with flanking Survey primer pairs specified in Table 1. PCR products were purified with a QIAquick PCR Purification Kit (Qiagen) and quantified with Quant-iT™ PicoGreen® dsDNA Kit (Life Technologies). 250ng of purified PCR DNA was combined with 2 μ L of NEBuffer 2 (NEB) in a total volume of 19 μ L and denatured then re-annealed with thermocycling at 95 °C for 5 min, 95 to 85 °C at 2 °C/s; 85 to 20 °C at 0.2 °C/s. The re-annealed DNA was incubated with 1 μ L of T7 Endonuclease I (10 U/ μ L, NEB) at 37 °C for 15 min. 10 μ L of 50% glycerol was added to the T7 Endonuclease reaction and 12 μ L was analyzed on a 5% TBE 18-well Criterion PAGE gel (Bio-Rad) electrophoresed for 30 min at 150 V, then stained with 1x SYBR Gold (Life Technologies) for 30 min. Cas9-induced cleavage bands and the uncleaved band were visualized on an AlphaImager HP (Alpha Innotech) and quantified using ImageJ software.³³ The peak intensities of the cleaved bands were divided by the total intensity of all bands (uncleaved + cleaved bands) to determine the fraction cleaved which was used to estimate gene modification levels as previously described.²⁸ For each sample, transfections and subsequent modification measurements were performed in triplicate on different days.

High-throughput Sequencing of Genomic Modifications

[00144] HEK293-GFP stable cells were transfected with Cas9 expression and gRNA expression plasmids, 700 ng of Cas9 expression plasmid plus 250 ng of a single plasmid

expression a pair of gRNAs were transfected (high levels) and for just Cas9 nuclease, 88 ng of Cas9 expression plasmid plus 31 ng of a single plasmid expression a pair of gRNAs were transfected (low levels). Genomic DNA was isolated as above and pooled from three biological replicates. 150 ng or 600 ng of pooled genomic DNA was used as template to amplify by PCR the on-target and off-target genomic sites with flanking HTS primer pairs specified in Table 1. Relative amounts of crude PCR products were quantified by gel electrophoresis and samples treated with different gRNA pairs or Cas9 nuclease types were separately pooled in equimolar concentrations before purification with the QIAquick PCR Purification Kit (Qiagen). ~500 ng of pooled DNA was run a 5% TBE 18-well Criterion PAGE gel (BioRad) for 30 min at 200 V and DNAs of length ~125 bp to ~300 bp were isolated and purified by QIAquick PCR Purification Kit (Qiagen). Purified DNA was PCR amplified with primers containing sequencing adaptors, purified and sequenced on a MiSeq high-throughput DNA sequencer (Illumina) as described previously.¹

Data Analysis

[00145] Illumina sequencing reads were filtered and parsed with scripts written in Unix Bash. All scripts were written in bash.

[00146] The Patmatch program³⁸ was used to search the human genome (GRCh37/hg19 build) for pattern sequences corresponding to Cas9 binding sites (CCN N²⁰ spacer N²⁰NGG for Orientation A and N²⁰NGG spacer CCN N²⁰ for Orientation B). The steps for the identification of ingels in sequences of genomic sites can be found below:

[00147] 1) Sequence reads were initially filtered removing reads of less than 50 bases and removing reads with greater than 10% of the Illumina base scores not being B-J:

[00148] Example SeqA-1st read:

[00149] TTCTGAGGGCTGCTACCTGTACATCTGCACAAGATTGCCTTTACTCC
ATGCCTTTCTTCTTCTGCTCTAACTCTGACAATCTGTCTTGCCATGCCATAAGCCC
CTATTCTTTCTGTAAACCCCAAGATGGTATAAAAGCATCAATGATTGGGC (SEQ ID
NO:157)

[00150] Example SeqA-2st read:

[00151] AAAACTCAAAGAAATGCCCAATCATTGATGCTTTTATACCATCTTG
GGGTTACAGAAAGAATAGGGGCTTATGGCATGGCAAGACAGATTGTCAGAGTTA
GAGCAGAAGAAGAAAGGCATGGAGTAAAGGCAATCTTGTGCAGATGTACAGGT
AA (SEQ ID NO:158)

[00152] 2) Find **the first 20 bases four bases from the start** of the reverse complement of SeqA-2nd read in SeqA-1st read allowing for 1 mismatch:

[00153] Reverse complement of SeqA-2nd read:

[00154] TTACCTGTACATCTGCACAAGATTGCCTTTACTCCATGCCTTTCTT
CTTCTGCTCTAACTCTGACAATCTGTCTTGCCATGCCATAAGCCCCTATTCTTTCT
GTAACCCCAAGATGGTATAAAAGCATCAATGATTGGGCATTTCTTTGAGTTTT
(SEQ ID NO:159)

[00155] Position in SeqA-1st read

TTCTGAGGGCTGCTACCTGTACATCTGCACAAGATTGCCTTTACTCCATGCCTTT
CTTCTTCTGCTCTAACTCTGACAATCTGTCTTGCCATGCCATAAGCCCCTATTCTT
TCTGTAACCCCAAGATGGTATAAAAGCATCAATGATTGGGC (SEQ ID NO:160)

[00156] 3) Align and then combine sequences, removing any sequence with greater than 5% mismatches in the simple base pair alignment:

[00157] Combination of SeqA-1st read and SeqA-2nd read:

[00158] TTCTGAGGGCTGCTACCTGTACATCTGCACAAGATTGCCTTTACTC
CATGCCTTTCTTCTTCTGCTCTAACTCTGACAATCTGTCTTGCCATGCCATAAGCC
CCTATTCTTTCTGTAACCCCAAGATGGTATAAAAGCATCAATGATTGGGCATTTC
TTTGAGTTTT (SEQ ID NO:161)

[00159] 4) To identify the target site the flanking genomic sequences were searched for with the Patmatch program³⁸ allowing for varying amounts of bases from 1 to 300 between the flanking genomic sequences (Table 2):

Table 2. Patmatch Sequences

Target Site	Downstream genomic sequence	Upstream genomic sequence
EMX_On	GGCCTGCTTCGTGGCAATGC (SEQ ID NO:162)	ACCTGGGCCAGGGAGGGAGG (SEQ ID NO:163)
EMX_Off1	CTCACTTAGACTTTCTCTCC (SEQ ID NO:164)	CTCGGAGTCTAGCTCCTGCA (SEQ ID NO:165)

EMX_Off2	TGGCCCCAGTCTCTCTTCTA (SEQ ID NO:166)	CAGCCTCTGAACAGCTCCCG (SEQ ID NO:167)
EMX_Off3	TGACTTGGCCTTTGTAGGAA (SEQ ID NO:168)	GAGGCTACTGAAACATAAGT (SEQ ID NO:169)
EMX_Off4	TGCTACCTGTACATCTGCAC (SEQ ID NO:170)	CATCAATGATTGGGCATTTC (SEQ ID NO:171)
VEG_On	ACTCCAGTCCCAAATATGTA (SEQ ID NO:172)	ACTAGGGGGGCGCTCGGCCAC (SEQ ID NO:173)
VEG_Off1	CTGAGTCAACTGTAAGCATT (SEQ ID NO:174)	GGCCAGGTGCAGTGATTCAT (SEQ ID NO:175)
VEG_Off2	TCGTGTCATCTTGTTTGTGC (SEQ ID NO:176)	GGCAGAGCCCAGCGGACACT (SEQ ID NO:177)
VEG_Off3	CAAGGTGAGCCTGGGTCTGT (SEQ ID NO:178)	ATCACTGCCCAAGAAGTGCA (SEQ ID NO:179)
VEG_Off4	TTGTAGGATGTTTAGCAGCA (SEQ ID NO:180)	ACTTGCTCTCTTTAGAGAAC (SEQ ID NO:181)
CLT2_On	CTCAAGCAGGCCCGCTGGT (SEQ ID NO:182)	TTTTGGACCAAACCTTTTGT (SEQ ID NO:183)
CLT2_Off1	TGAGGTTATTTGTCCATTGT (SEQ ID NO:184)	TAAGGGGAGTATTACACCA (SEQ ID NO:185)
CLT2_Off2	TCAAGAGCAGAAAATGTGAC (SEQ ID NO:186)	CTTGCAGGGACCTTCTGATT (SEQ ID NO:187)
CLT2_Off3	TGTGTGTAGGACTAACTCT (SEQ ID NO:188)	GATAGCAGTATGACCTTGGG (SEQ ID NO:189)

[00160] Any target site sequences corresponding to the same size as the reference genomic site in the human genome (GRCh37/hg19 build) were considered unmodified and any sequences not the reference size were considered potential insertions or deletions. Sequences not the reference size were aligned with ClustalW³⁹ to the reference genomic site. Aligned sequences with more than one insertion or one deletion in the DNA spacer sequence in or between the two half-site sequences were considered indels. Since high-throughput sequencing can result in insertions or deletions of one base pairs (mis-phasing) at a low but relevant rates - indels of two bp are more likely to arise from Cas9 induced modifications.

[00161] Sample sizes for sequencing experiments were maximized (within practical experimental considerations) to ensure greatest power to detect effects. Statistical analyses for Cas9-modified genomic sites in Table 3 were performed as previously described³⁴ with multiple comparison correction using the Bonferroni method.

[00162] Table 3, referred to in the Results below, shows (A) results from sequencing *CLTA* on-target and previously reported genomic off-target sites amplified from 150 ng genomic DNA isolated from human cells treated with a plasmid expressing either wild-type Cas9, Cas9 nickase, or fCas9 and a single plasmid expressing two gRNAs targeting the *CLTA* on-target site (gRNA C3 and gRNA C4). As a negative control, transfection and sequencing were performed as above, but using two gRNAs targeting the *GFP* gene on-target site (gRNA G1, G2 or G3 and gRNA G4, G5, G6 or G7). Indels: the number of observed sequences containing insertions or deletions consistent with any of the three Cas9 nuclease-induced cleavage. Total: total number of sequence counts while only the first 10,000 sequences were analyzed for the on-target site sequences. Modified: number of indels divided by total number of sequences as percentages. Upper limits of potential modification were calculated for sites with no observed indels by assuming there is less than one indel then dividing by the total sequence count to arrive at an upper limit modification percentage, or taking the theoretical limit of detection (1/49,500), whichever value was larger. P-values: For wild-type Cas9 nuclease, Cas9 nickase or fCas9 nuclease, P-values were calculated as previously reported¹⁸ using a two-sided Fisher's exact test between each sample treated with two gRNAs targeting the *CLTA* on-target site and the control sample treated with two gRNAs targeting the *GFP* on-target site. P-values of < 0.0045 were considered significant and shown based on conservative multiple comparison correction using the Bonferroni method. On:off specificity is the ratio of on-target to off-target genomic modification frequency for each site. (B) Shows experimental and analytic methods as in (A) applied to *EMX* target sites using a single plasmid expressing two gRNAs targeting the *EMX* on-target site (gRNA E1 and gRNA E2).

(C) shows experimental and analytic methods as in (A) applied to *VEGF* target sites using a single plasmid expressing two gRNAs targeting the *VEGF* on-target site (gRNA V1 and gRNA v2). (D) shows experimental and analytic methods as in (A) applied to *VEGF* on-target and *VEGF* off-target site 1 amplified from 600 ng genomic DNA to increase detection sensitivity to 1/198,000.

Table 3. Cellular modification induced by wild-type Cas9, Cas9 nickase, and fCas9 at on-target and off-target genomic sites.

(A)

Nuclease type:	wt Cas9	wt Cas9	Cas9 nickase	fCas9	wt Cas9	Cas9 nickase	fCas9
gRNA pair target:	<i>CLTA</i>	<i>CLTA</i>	<i>CLTA</i>	<i>CLTA</i>	<i>GFP</i>	<i>GFP</i>	<i>GFP</i>
Total expression plasmids (ng):	1000	125	1000	1000	1000	1000	1000
<u><i>CLTA</i> Sites</u>							
<u>CLT2_On</u>							
Indels	3528	1423	3400	575	3	13	5
Total	10000	10000	10000	10000	10000	10000	10000
Modified (%)	35.280	14.230	34.000	5.750	0.030	0.130	0.050
P-value	<1.0E-300	<1.0E-300	<1.0E-300	1.4E-163			
On:off specificity	1	1		1			
<u>CLT2_Off1</u>							
Indels	316	44	2	2	1	3	3
Total	60620	64755	71537	63079	93883	91306	82055
Modified (%)	0.521	0.068	0.003	0.003	<0.002	0.003	0.004
P-value	1.3E-126	2.1E-16					
On:off specificity	68	209		>2850			
<u>CLT2_Off2</u>							
Indels	11	5	3	1	1	1	2
Total	72596	51093	59632	35541	69114	64412	39978
Modified (%)	0.015	0.010	0.005	0.003	<0.002	<0.002	0.005
P-value	6.5E-03						
On:off specificity	2328	1454		>2850			
<u>CLT2_Off3</u>							
Indels	11	10	0	0	1	1	1
Total	52382	44212	54072	48668	55670	58707	54341
Modified (%)	0.021	0.023	<0.002	<0.002	<0.002	<0.002	<0.002
P-value	2.7E-03	3.5E-03					
On:off specificity	1680	629		>2850			

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(B)

Nuclease type:	wt Cas9	wt Cas9	Cas9 nickase	fCas9	wt Cas9	Cas9 nickase	fCas9
gRNA pair:	<i>EMX</i>	<i>EMX</i>	<i>EMX</i>	<i>EMX</i>	<i>GFP</i>	<i>GFP</i>	<i>GFP</i>
Total expression plasmids (ng):	1000	125	1000	1000	1000	1000	1000
<u>EMX Site</u>							
<u>EMX_On</u>							
Indels	5111	2683	2267	522	0	0	2
Total	10000	10000	10000	10000	10000	10000	10000
Modified (%)	51.110	26.830	22.670	5.220	<0.002	<0.002	0.020
P-value	<1.0E-300	<1.0E-300	<1.0E-300	1.0E-154			
On:off specificity	1	1	1	1			
<u>EMX_Off1</u>							
Indels	386	122	7	1	4	9	7
Total	109787	83420	124564	88424	102817	90020	96526
Modified (%)	0.352	0.146	0.006	<0.002	0.004	0.010	0.007
P-value	1.3E-103	2.8E-37					
On:off specificity	145	183	>11222	>2584			
<u>EMX_Off2</u>							
Indels	74	58	3	6	3	0	4
Total	98568	94108	105747	78871	81717	79469	79193
Modified (%)	0.075	0.062	0.003	0.008	0.004	<0.002	0.005
P-value	3.2E-16	1.4E-12					
On:off specificity	681	435	>11222	>2584			
<u>EMX_Off3</u>							
Indels	736	178	20	14	12	11	17
Total	72888	65139	82348	59593	74341	73408	75080
Modified (%)	1.010	0.273	0.024	0.023	0.016	0.015	0.023
P-value	2.5E-202	3.1E-44					
On:off specificity	51	98	>11222	>2584			
<u>EMX_Off4</u>							
Indels	4149	620	3	3	6	7	5
Total	107537	91695	91368	91605	111736	119643	128088
Modified (%)	3.858	0.676	0.003	0.003	0.005	0.006	0.004
P-value	<1.0E-300	1.9E-202					
On:off specificity	13	40	>11222	>2584			

(C)

Nuclease type:	wt Cas9	wt Cas9	Cas9 nickase	fCas9	wt Cas9	Cas9 nickase	fCas9
gRNA pair:	<i>VEGF</i>	<i>VEGF</i>	<i>VEGF</i>	<i>VEGF</i>	<i>GFP</i>	<i>GFP</i>	<i>GFP</i>

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Total expression plasmids (ng):	1000	125	1000	1000	1000	1000	1000
<u>VEGF Sites</u>							
<u>VEG_On</u>							
Indels	5253	2454	1230	1041	8	0	1
Total	10000	10000	10000	10000	10000	10000	10000
Modified (%)	52.530	24.540	12.300	10.410	0.080	<0.002	0.010
P-value	<1.0E-300	<1.0E-300	<1.0E-300	6.6E-286			
On:off specificity	1	1	1	1			
<u>VEG_Off1</u>							
Indels	2950	603	22	0	0	4	1
Total	82198	71163	90434	77557	74765	79738	74109
Modified (%)	3.589	0.847	0.024	<0.002	<0.002	0.005	<0.002
P-value	<1.0E-300	3.2E-188	2.5E-06				
On:off specificity	15	29	506	>5150			
<u>VEG_Off2</u>							
Indels	863	72	3	3	0	2	1
Total	102501	49836	119702	65107	54247	65753	61556
Modified (%)	0.842	0.144	0.003	0.005	<0.002	0.003	<0.002
P-value	3.5E-159	9.6E-24					
On:off specificity	62	170	>6090	>5150			
<u>VEG_Off3</u>							
Indels	260	33	3	2	3	1	0
Total	91277	83124	90063	84385	62126	68165	69811
Modified (%)	0.285	0.040	0.003	0.002	0.005	<0.002	<0.002
P-value	6.8E-54	1.0E-05					
On:off specificity	184	618	>6090	>5150			
<u>VEG_Off4</u>							
Indels	1305	149	3	2	3	2	4
Total	59827	41203	65964	57828	60906	61219	62162
Modified (%)	2.181	0.362	0.005	0.003	0.005	0.003	0.006
P-value	<1.0E-300	2.7E-54					
On:off specificity	24	68	>6090	>5150			

(D)

Nuclease type:	Cas9		Cas9	
	nickase	fCas9	nickase	fCas9
gRNA pair:	<i>VEGF</i>	<i>VEGF</i>	<i>GFP</i>	<i>GFP</i>
Total expression plasmids (ng):	1000	1000	1000	1000
<u>VEGF Sites</u>				
<u>VEG_On</u>				

Indels	2717	2122	10	13
Total	10000	10000	10000	10000
Modified (%)	27.170	21.220	0.100	0.130
P-value	<1.0E-300	<1.0E-300		
On:off specificity	1	1		
VEG_Off1				
Indels	67	30	3	2
Total	302573	233567	204454	190240
Modified (%)	0.022	0.013		
P-value	5.9E-12	2.5E-06		
On:off specificity	1227	1652		

Results

[00163] Recently engineered variants of Cas9 that cleave only one DNA strand (“nickases”) enable double-stranded breaks to be specified by two distinct gRNA sequences,⁵⁻⁷ but still suffer from off-target cleavage activity^{6,8} arising from the ability of each monomeric nickase to remain active when individually bound to DNA.⁹⁻¹¹ In contrast, the development of a *FokI* nuclease fusion to a catalytically dead Cas9 that requires simultaneous DNA binding and association of two *FokI*-dCas9 monomers to cleave DNA is described here. Off-target DNA cleavage of the engineered *FokI*-dCas9 (fCas9) is further reduced by the requirement that only sites flanked by two gRNAs ~15 or 25 base pairs apart are cleaved, a much more stringent spacing requirement than nickases. In human cells, fCas9 modified target DNA sites with efficiency comparable to that of nickases, and with > 140-fold higher specificity than wild-type Cas9. Target sites that conform to the substrate requirements of fCas9 are abundant in the human genome, occurring on average once every 34 bp.

[00164] In cells, Cas9:gRNA-induced double strand breaks can result in functional gene knockout through non-homologous end joining (NHEJ) or alteration of a target locus to virtually any sequence through homology-directed repair (HDR) with an exogenous DNA template.^{9,15,16} Cas9 is an especially convenient genome editing platform,¹⁷ as a genome editing agent for each new target site of interest can be accessed by simply generating the corresponding gRNA. This approach has been widely used to create targeted knockouts and gene insertions in cells and model organisms, and has also been recognized for its potential therapeutic relevance.

[00165] While Cas9:gRNA systems provide an unprecedented level of programmability and ease of use, studies¹⁻⁵ have reported the ability of Cas9 to cleave off-

target genomic sites, resulting in modification of unintended loci that can limit the usefulness and safety of Cas9 as a research tool and as a potential therapeutic. It was hypothesized that engineering Cas9 variants to cleave DNA only when two simultaneous, adjacent Cas9:DNA binding events take place could substantially improve genome editing specificity since the likelihood of two adjacent off-target binding events is much smaller than the likelihood of a single off-target binding event (approximately $1/n^2$ vs. $1/n$). Such an approach is distinct from the recent development of mutant Cas9 proteins that cleave only a single strand of dsDNA, such as nickases. Nickases can be used to nick opposite strands of two nearby target sites, generating what is effectively a double strand break, and paired Cas9 nickases can effect substantial on-target DNA modification with reduced off-target modification.^{5,6,8} Because each of the component Cas9 nickases remains catalytically active⁹⁻¹¹ and single-stranded DNA cleavage events are weakly mutagenic,^{18,19} nickases can induce genomic modification even when acting as monomers.^{5,7,16} Indeed, Cas9 nickases have been previously reported to induce off-target modifications in cells.^{6,8} Moreover, since paired Cas9 nickases can efficiently induce dsDNA cleavage-derived modification events when bound up to ~100 bp apart,⁶ the statistical number of potential off-target sites for paired nickases is larger than that of a more spatially constrained dimeric Cas9 cleavage system.

[00166] To further improve the specificity of the Cas9:gRNA system, an obligate dimeric Cas9 system is provided herein. In this example, fusing the *FokI* restriction endonuclease cleavage domain to a catalytically dead Cas9 (dCas9) created an obligate dimeric Cas9 that would cleave DNA only when two distinct *FokI*-dCas9:gRNA complexes bind to adjacent sites (“half-sites”) with particular spacing constraints (Figure 6D). In contrast with Cas9 nickases, in which single-stranded DNA cleavage by monomers takes place independently, the DNA cleavage of *FokI*-dCas9 requires simultaneous binding of two distinct *FokI*-dCas9 monomers because monomeric *FokI* nuclease domains are not catalytically competent.²¹ This approach increased the specificity of DNA cleavage relative to wild-type Cas9 by doubling the number of specified target bases contributed by both monomers of the *FokI*-dCas9 dimer, and offered improved specificity compared to nickases due to inactivity of monomeric *FokI*-dCas9:gRNA complexes, and the more stringent spatial requirements for assembly of a *FokI*-dCas9 dimer.

[00167] While fusions of Cas9 to short functional peptide tags have been described to enable gRNA-programmed transcriptional regulation,²² it is believed that no fusions of Cas9 with active enzyme domains have been previously reported. Therefore a wide variety of *FokI*-dCas9 fusion proteins were constructed and characterized with distinct configurations of

a *FokI* nuclease domain, dCas9 containing inactivating mutations D10A and H840A, and a nuclear localization sequence (NLS). *FokI* was fused to either the N- or C-terminus of dCas9, and varied the location of the NLS to be at either terminus or between the two domains (Figure 6B). The length of the linker sequence was varied as either one or three repeats of Gly-Gly-Ser (GGS) between the *FokI* and dCas9 domains. Since previously developed dimeric nuclease systems are sensitive to the length of the spacer sequence between half-sites,^{23,24} a wide range of spacer sequence lengths was tested between two gRNA binding sites within a test target gene, Emerald GFP (referred to hereafter as GFP) (Figure 6C and Figure 9). Two sets of gRNA binding-site pairs with different orientations were chosen within GFP. One set placed the pair of NGG PAM sequences distal from the spacer sequence, with the 5' end of the gRNA adjacent to the spacer (orientation A) (Figure 6C), while the other placed the PAM sequences immediately adjacent to the spacer (orientation B) (Figure 9). In total, seven pairs of gRNAs were suitable for orientation A, and nine were suitable for orientation B. By pairwise combination of the gRNA targets, eight spacer lengths were tested in both dimer orientations, ranging from 5 to 43 bp in orientation A, and 4 to 42 bp in orientation B. In total, DNA constructs corresponding to 104 pairs of *FokI*-dCas9:gRNA complexes were generated and tested, exploring four fusion architectures, 17 protein linker variants (described below), both gRNA orientations and 13 spacer lengths between half-sites.

[00168] To assay the activities of these candidate *FokI*-dCas9:gRNA pairs, a previously described flow cytometry-based fluorescence assay^{2,8} in which DNA cleavage and NHEJ of a stably integrated constitutively expressed GFP gene in HEK293 cells leads to loss of cellular fluorescence was used (Figure 10). For comparison, the initial set of *FokI*-dCas9 variants were assayed side-by-side with the corresponding Cas9 nickases and wild-type Cas9 in the same expression plasmid across both gRNA spacer orientation sets A and B. Cas9 protein variants and gRNA were generated in cells by transient co-transfection of the corresponding Cas9 protein expression plasmids together with the appropriate pair of gRNA expression plasmids. The *FokI*-dCas9 variants, nickases, and wild-type Cas9 all targeted identical DNA sites using identical gRNAs.

[00169] Most of the initial *FokI*-dCas9 fusion variants were inactive or very weakly active (Figure 11). The NLS-*FokI*-dCas9 architecture (listed from N to C terminus), however, resulted in a 10% increase of GFP-negative cells above corresponding the no-gRNA control when used in orientation A, with PAMs distal from the spacer (Figure 11A). In contrast, NLS-*FokI*-dCas9 activity was undetectable when used on gRNA pairs with

PAMs adjacent to the spacer (Figure 11B). Examination of the recently reported Cas9 structures^{25,26} reveals that the Cas9 N-terminus protrudes from the RuvCI domain, which contacts the 5' end of the gRNA:DNA duplex. Without wishing to be bound by any particular theory, it is speculated that this arrangement places an N-terminally fused *FokI* distal from the PAM, resulting in a preference for gRNA pairs with PAMs distal from the cleaved spacer (Figure 6D). While other *FokI*-dCas9 fusion pairings and the other gRNA orientation in some cases showed modest activity (Figure 11), NLS-*FokI*-dCas9 with gRNAs in orientation A were chosen for further development.

[00170] Next the protein linkers between the NLS and *FokI* domain, and between the *FokI* domain and dCas9 in the NLS-*FokI*-dCas9 architecture were optimized. 17 linkers with a wide range of amino acid compositions, predicted flexibilities, and lengths varying from 9 to 21 residues were tested (Figure 12A). Between the *FokI* domain and dCas9 a flexible 18-residue linker, (GGS)₆ (SEQ ID NO:15), and a 16-residue “XTEN” linker (*FokI*-L8 in Figure 12A) were identified based on a previously reported engineered protein with an open, extended conformation,²⁷ as supporting the highest levels of genomic GFP modification (Figure 12B).

[00171] The XTEN protein was originally designed to extend the serum half-life of translationally fused biologic drugs by increasing their hydrodynamic radius, acting as protein-based functional analog to chemical PEGylation.³⁵ Possessing a chemically stable, non-cationic, and non-hydrophobic primary sequence, and an extended conformation, it is hypothesized that a portion of XTEN could function as a stable, inert linker sequence for fusion proteins. The sequence of the XTEN protein tag from E-XTEN was analyzed, and repeating motifs within the amino acid sequence were aligned. The sequence used in the *FokI*-dCas9 fusion construct *FokI*-L8 (Figure 12A) was derived from the consensus sequence of a common E-XTEN motif, and a 16 amino acid sequence was chosen from within this motif to test as a *FokI*-dCas9 linker.

[00172] Many of the *FokI*-dCas9 linkers tested including the optimal XTEN linker resulted in nucleases with a marked preference for spacer lengths of ~15 and ~25 bp between half-sites, with all other spacer lengths, including 20 bp, showing substantially lower activity (Figure 12B). This pattern of linker preference is consistent with a model in which the *FokI*-dCas9 fusions must bind to opposite faces of the DNA double helix to cleave DNA, with optimal binding taking place ~1.5 or 2.5 helical turns apart. The variation of NLS-*FokI* linkers did not strongly affect nuclease performance, especially when combined with the XTEN *FokI*-dCas9 linker (Figure 12B).

[00173] In addition to assaying linkers between the FokI domain and dCas9 in the NLS-FokI-dCas9 architecture, four linker variants between the N-terminal NLS and the FokI domain were also tested (Figure 12A). Although a NLS-GSAGSAAGSGEF (SEQ ID NO:20)-FokI-dCas9 linker exhibited nearly 2-fold better GFP gene modification than the other NLS-FokI linkers tested when a simple GGS linker was used between the FokI and dCas9 domains (Figure 12B), the GSAGSAAGSGEF (SEQ ID NO:20) linker did not perform substantially better when combined with the XTEN linker between the FokI and dCas9 domains.

[00174] The NLS-GGS-*FokI*-XTEN-dCas9 construct consistently exhibited the highest activity among the tested candidates, inducing loss of GFP in ~15% of cells, compared to ~20% and ~30% for Cas9 nickases and wild-type Cas9 nuclease, respectively (Figure 7A). All subsequent experiments were performed using this construct, hereafter referred to as fCas9. To confirm the ability of fCas9 to efficiently modify genomic target sites, the T7 endonuclease I Surveyor assay²⁸ was used to measure the amount of mutation at each of seven target sites within the integrated GFP gene in HEK293 cells treated with fCas9, Cas9 nickase, or wild-type Cas9 and either two distinct gRNAs in orientation A or no gRNAs as a negative control. Consistent with the flow cytometry-based studies, fCas9 was able to modify the GFP target sites with optimal spacer lengths of ~15 or ~25 bp at a rate of ~20%, comparable to the efficiency of nickase-induced modification and approximately two-thirds that of wild-type Cas9 (Figure 7A-C).

[00175] Next the ability of the optimized fCas9 to modify four distinct endogenous genomic loci by Surveyor assay was evaluated. *CLTA* (two sites), *EMX* (two sites), *HBB* (six sites) *VEGF* (three sites), and were targeted with two gRNAs per site in orientation A spaced at various lengths (Figure 13). Consistent with the results of the experiments targeting GFP, at appropriately spaced target half-sites fCas9 induced efficient modification of all four genes, ranging from 8% to 22% target chromosomal site modification (Figure 7D-G and Figure 14). Among the gRNA spacer lengths resulting in the highest modification at each of the five genes targeted (including GFP), fCas9 induced on average 15.6% ($\pm 6.3\%$ s.d.) modification, while Cas9 nickase and wild-type Cas9 induced on average 22.1% ($\pm 4.9\%$ s.d.) and 30.4% ($\pm 3.1\%$ s.d.) modification, respectively, from their optimal gRNA pairs for each gene. Because decreasing the amount of Cas9 expression plasmid and gRNA expression plasmid during transfection generally did not proportionally decrease genomic modification activity for Cas9 nickase and fCas9 (Figure 15A-C), expression was likely not limiting under the conditions tested.

[00176] As the gRNA requirements of fCas9 potentially restricts the number of potential off-target substrates of fCas9, the effect of guide RNA orientation on the ability of fCas9, Cas9 nickase, and wild-type Cas9 to cleave target GFP sequences were compared. Consistent with previous reports,^{5,6,17} Cas9 nickase efficiently cleaved targets when guide RNAs were bound either in orientation A or orientation B, similar to wild-type Cas9 (Figure 8A, B). In contrast, fCas9 only cleaved the GFP target when guide RNAs were aligned in orientation A (Figure 8A). This orientation requirement further limits opportunities for undesired off-target DNA cleavage.

[00177] Importantly, no modification was observed by GFP disruption or Surveyor assay when any of four single gRNAs were expressed individually with fCas9, as expected since two simultaneous binding events are required for *FokI* activity (Figure 7B and Figure 8C). In contrast, GFP disruption resulted from expression of any single gRNA with wild-type Cas9 (as expected) and, for two single gRNAs, with Cas9 nickase (Figure 8C). Surprisingly, Surveyor assay revealed that although GFP was heavily modified by wild-type Cas9 with single gRNAs, neither fCas9 nor Cas9 nickase showed detectable modification (< ~2%) in cells treated with single gRNAs (Figure 16A). High-throughput sequencing to detect indels at the GFP target site in cells treated with a single gRNA and fCas9, Cas9 nickase, or wild-type Cas9 revealed the expected substantial level of modification by wild-type Cas9 (3-7% of sequence reads). Modification by fCas9 in the presence of any of the four single gRNAs was not detected above background (< ~0.03% modification), consistent with the requirement of fCas9 to engage two gRNAs in order to cleave DNA. In contrast, Cas9 nickases in the presence of single gRNAs resulted in modification levels ranging from 0.05% to 0.16% at the target site (Figure 16B). The detection of bona fide indels at target sites following Cas9 nickase treatment with single gRNAs confirms the mutagenic potential of genomic DNA nicking, consistent with previous reports.^{5,7,18,19}

[00178] The observed rate of nickase-induced DNA modification, however, did not account for the much higher GFP disruption signal in the flow cytometry assay (Figure 8C). Since the gRNAs that induced GFP signal loss with Cas9 nickase (gRNAs G1 and G3) both target the non-template strand of the GFP gene, and since targeting the non-template strand with dCas9 in the coding region of a gene is known to mediate efficient transcriptional repression,²⁹ it is speculated that Cas9 nickase combined with the G1 or G3 single guide RNAs induced substantial transcriptional repression, in addition to a low level of genome modification. The same effect was not seen for fCas9, suggesting that fCas9 may be more easily displaced from DNA by transcriptional machinery. Taken together, these results

indicate that fCas9 can modify genomic DNA efficiently and in a manner that requires simultaneous engagement of two guide RNAs targeting adjacent sites, unlike the ability of wild-type Cas9 and Cas9 nickase to cleave DNA when bound to a single guide RNA.

[00179] The above results collectively reveal much more stringent spacer, gRNA orientation, and guide RNA pairing requirements for fCas9 compared with Cas9 nickase. In contrast with fCas9 (Figure 17), Cas9 nickase cleaved sites across all spacers assayed (5- to 47- bp in orientation A and 4 to 42 bp in orientation B in this work) (Figure 8A, B). These observations are consistent with previous reports of Cas9 nickases modifying sites targeted by gRNAs with spacer lengths up to 100 bp apart.⁶ The more stringent spacer and gRNA orientation requirements of fCas9 compared with Cas9 nickase reduces the number of potential genomic off-target sites of the former by approximately 10-fold (Table 4). Although the more stringent spacer requirements of fCas9 also reduce the number of potential targetable sites, sequences that conform to the fCas9 spacer and dual PAM requirements exist in the human genome on average once every 34 bp (9.2×10^7 sites in 3.1×10^9 bp) (Table 4). It is also anticipated that the growing number of Cas9 homologs with different PAM specificities³⁰ are amenable for use as described herein, and will further increase the number of targetable sites using the fCas9 approach.

[00180] In Table 4 (A) column 2 shows the number of sites in the human genome with paired gRNA binding sites in orientation A allowing for a spacer length from -8 bp to 25 bp (column 1) between the two gRNA binding sites. gRNA binding sites in orientation A have the NGG PAM sequences distal from the spacer sequence (CCNN₂₀-spacer-N₂₀NGG). Column 3 shows the number of sites in the human genome with paired gRNA binding sites in orientation B allowing for a spacer length from 4 to 25 bp (column 1) between the two gRNA binding sites. gRNA binding sites in orientation B have the NGG PAM sequences adjacent to the spacer sequence (N₂₀NGG spacer CCNN₂₀). NC indicates the number of sites in the human genome was not calculated. Negative spacer lengths refer to target gRNA binding sites that overlap by the indicated number of base pairs. Table 4 (B) shows the sum of the number of paired gRNA binding sites in orientation A with spacer lengths of 13 to 19 bp, or 22 to 29 bp, the spacer preference of fCas9 (Figure 16). Sum of the number of paired gRNA binding sites with spacer lengths of -8 bp to 100 bp in orientation A, or 4 to 42 bp in orientation B, the spacer preference of Cas9 nickases (4 to 42 bp in orientation B is based on Figure 8B, C, and -8 bp to 100 bp in orientation A is based on previous reports^{36, 37}).

Table 4. Paired gRNA target site abundances for fCas9 and Cas9 nickase in the human genome.

(A)

Spacer length (b)	Number of paired gRNA sites in orientation A	Number of paired gRNA sites in orientation B
-8	6874293	NC
-7	6785996	NC
-6	6984064	NC
-5	7023260	NC
-4	6487302	NC
-3	6401348	NC
-2	6981383	NC
-1	7230098	NC
0	7055143	NC
1	6598582	NC
2	6877046	NC
3	6971447	NC
4	6505614	5542549
5	6098107	5663458
6	6254974	6819289
7	6680118	6061225
8	7687598	5702252
9	6755736	7306646
10	6544849	6387485
11	6918186	6172852
12	6241723	5799496
13	6233385	7092283
14	6298717	7882433
15	6181422	7472725
16	6266909	6294684
17	6647352	6825904
18	6103603	6973590
19	5896092	6349456
20	6000683	5835825
21	5858015	6056352
22	6116108	6531913
23	5991254	6941816
24	6114969	6572849
25	6135119	5671641

(B)

Cas9 variant	Preferred spacer lengths (bp)	Total sites
fCas9	13 to 19, or 22 to 29, in orientation A	92354891
Cas9 nickase	-8 to 100 in orientation A 4 to 42 in orientation B	953048977

[00181] To evaluate the DNA cleavage specificity of fCas9, the modification of known Cas9 off-target sites of *CLTA*, *EMX*, and *VEGF* genomic target sites were measured.^{1,2,6,8} The target site and its corresponding known off-target sites (Table 5) were amplified from genomic DNA isolated from HEK293 cells treated with fCas9, Cas9 nickase, or wild-type Cas9 and two gRNAs spaced 19 bp apart targeting the *CLTA* site, two gRNAs spaced 23 bp apart targeting the *EMX* site, two gRNAs spaced 14 bp apart targeting the *VEGF* site, or two gRNAs targeting an unrelated site (GFP) as a negative control. In total 11 off-target sites were analyzed by high-throughput sequencing.

[00182] The sensitivity of the high-throughput sequencing method for detecting genomic off-target cleavage is limited by the amount genomic DNA (gDNA) input into the PCR amplification of each genomic target site. A 1 ng sample of human gDNA represents only ~330 unique genomes, and thus only ~330 unique copies of each genomic site are present. PCR amplification for each genomic target was performed on a total of 150 ng of input gDNA, which provides amplicons derived from at most 50,000 unique gDNA copies. Therefore, the high-throughput sequencing assay cannot detect rare genome modification events that occur at a frequency of less than 1 in 50,000, or 0.002%.

Table 5. Known off-target substrates of Cas9 target sites in *EMX*, *VEGF*, and *CLTA*. List of genomic on-target and off-targets sites of the *EMX*, *VEGF*, and *CLTA* are shown with mutations from on-target in lower case and bold. PAMs are shown in upper case bold.

	Genomic target site
EMX_On	GAGTCCGAGCAGAAGAAGAA GGG (SEQ ID NO:190)
EMX_Off1	GAG g CCGAGCAGAAGAA ag A CGG (SEQ ID NO:191)
EMX_Off2	GAGTCC t AGCAG g AGAAGAA GaG (SEQ ID NO:192)
EMX_Off3	GAGTC ta AGCAGAAGAAGAA GaG (SEQ ID NO:193)
EMX_Off4	GAGT ta GAGCAGAAGAAGAA AGG (SEQ ID NO:194)
VEG_On	GGGTGGGGGGAGTTTGCTCCT TGG (SEQ ID NO:195)
VEG_Off1	GG a TGG a GGGAGTTTGCTCCT TGG (SEQ ID NO:196)
VEG_Off2	GGG a GGG t GGAGTTTGCTCCT TGG (SEQ ID NO:197)
VEG_Off3	c GG g GG a GGGAGTTTGCTCCT TGG (SEQ ID NO:198)
VEG_Off4	GGG ga GGGG a AGTTTGCTCCT TGG (SEQ ID NO:199)

CLT2_On GCAGATGTAGTGTTCACAGGG
(SEQ ID NO:200)
CLT2_Off1 aCAaATGTAGTaTTTCACAGGG
(SEQ ID NO:201)
CLT2_Off2 cCAGATGTAGTaTTcCCACAGGG
(SEQ ID NO:202)
CLT2_Off3 ctAGATGaAGTGcTTCCACATGG
(SEQ ID NO:203)

[00183] Sequences containing insertions or deletions of two or more base pairs in potential genomic off-target sites and present in significantly greater numbers (P value < 0.005, Fisher's exact test) in the target gRNA-treated samples versus the control gRNA-treated samples were considered Cas9 nuclease-induced genome modifications. For 10 of the 11 off-target sites assayed, fCas9 did not result in any detectable genomic off-target modification within the sensitivity limit of the assay (< 0.002%), while demonstrating substantial on-target modification efficiencies of 5% to 10% (Figure 8D-F and Table 3). The detailed inspection of fCas9-modified *VEGF* on-target sequences (Figure 18A) revealed a prevalence of deletions ranging from two to dozens of base pairs consistent with cleavage occurring in the DNA spacer between the two target binding sites, similar to the effects of *FokI* nuclease domains fused to zinc finger or TALE DNA-binding domains.³¹

[00184] In contrast, genomic off-target DNA cleavage was observed for wild-type Cas9 at all 11 sites assayed. Using the detection limit of the assay as an upper bound for off-target fCas9 activity, it was calculated that fCas9 has a much lower off-target modification rate than wild-type Cas9 nuclease. At the 11 off-target sites modified by wild-type Cas9 nuclease, fCas9 resulted in on-target:off-target modification ratios at least 140-fold higher than that of wild-type Cas9 (Figure 8D-F).

[00185] Consistent with previous reports,^{5,6,8} Cas9 nickase also induced substantially fewer off-target modification events (1/11 off-target sites modified at a detectable rate) compared to wild-type Cas9. An initial high-throughput sequencing assay revealed significant (P value < 10⁻³, Fisher's Exact Test) modification induced by Cas9 nickases in 0.024% of sequences at *VEGF* off-target site 1. This genomic off-target site was not modified by fCas9 despite similar *VEGF* on-target modification efficiencies of 12.3% for Cas9 nickase and 10.4% for fCas9 (Figure 8F and Table 3C). Because Cas9 nickase-induced modification levels were within an order of magnitude of the limit of detection and fCas9 modification levels were undetected, the experiment was repeated with a larger input DNA samples and a greater number of sequence reads (150 versus 600 ng genomic DNA and > 8 x 10⁵ versus > 23 x 10⁵ reads for the initial and repeated experiments, respectively) to detect

off-target cleavage at this site by Cas9 nickase or fCas9. From this deeper interrogation, it was observed that Cas9 nickase and fCas9 both significantly modify (P value $< 10^{-5}$, Fisher's Exact Test) *VEGF* off-target site 1 (Figure 8G, Table 3D, Figure 18B). For both experiments interrogating the modification rates at *VEGF* off-target site 1, fCas9 exhibited a greater on-target:off-target DNA modification ratio than that of Cas9 nickase ($> 5,150$ and $1,650$ for fCas9, versus 510 and $1,230$ for Cas9 nickase, Figure 8G).

[00186] On either side of *VEGF* off-target site 1 there exist no other sites with six or fewer mutations from either of the two half-sites of the *VEGF* on-target sequence. The first 11 bases of one gRNA (V2) might hybridize to the single-stranded DNA freed by canonical Cas9:gRNA binding within *VEGF* off-target site 1 (Figure 18C). Through this gRNA:DNA hybridization it is possible that a second Cas9 nickase or fCas9 could be recruited to modify this off-target site at a very low, but detectable level. Judicious gRNA pair design could eliminate this potential mode of off-target DNA cleavage, as *VEGF* off-target site 1 is highly unusual in its ability to form 11 consecutive potential base pairs with the second gRNA of a pair. In general, fCas9 was unable to modify the genomic off-target sites tested because of the absence of any adjacent second binding site required to dimerize and activate the *FokI* nuclease domain.

[00187] The optimized *FokI*-dCas9 fusion architecture developed in this work modified all five genomic loci targeted, demonstrating the generality of using fCas9 to induce genomic modification in human cells, although modification with fCas9 was somewhat less efficient than with wild-type Cas9. The use of fCas9 is straightforward, requiring only that PAM sequences be present with an appropriate spacing and orientation, and using the same gRNAs as wild-type Cas9 or Cas9 nickases. The observed low off-target:on-target modification ratios of fCas9, > 140 -fold lower than that of wild-type Cas9, likely arises from the distinct mode of action of dimeric *FokI*, in which DNA cleavage proceeds only if two DNA sites are occupied simultaneously by two *FokI* domains at a specified distance (here, ~ 15 bp or ~ 25 bp apart) and in a specific half-site orientation. The resulting unusually low off-target activity of fCas9 enable applications of Cas9:gRNA-based technologies that require a very high degree of target specificity, such as *ex vivo* or *in vivo* therapeutic modification of human cells.

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Example 2: Targetting CCR5 for Cas9 variant-mediated inactivation

[00188] In addition to providing powerful research tools, site-specific nucleases also have potential as gene therapy agents, and site-specific zinc finger endonucleases have recently entered clinical trials: CCR5-2246, targeting a human CCR-5 allele as part of an anti-HIV therapeutic approach.

[00189] In a similar approach, the inventive Cas9 variants of the present disclosure may be used to inactivate CCR5, for example in autologous T cells obtained from a subject which, once modified by a Cas9 variant, are re-introduced into the subject for the treatment or prevention of HIV infection.

[00190] In this example, the CCR5 gene is targeted in T cells obtained from a subject. CCR5 protein is required for certain common types of HIV to bind to and enter T cells, thereby infecting them. T cells are one of the white blood cells used by the body to fight HIV.

[00191] Some people are born lacking CCR5 expression on their T cells and remain healthy and are resistant to infection with HIV. Others have low expression of CCR5 on their T cells, and their HIV disease is less severe and is slower to cause disease (AIDS).

[00192] In order to delete the CCR5 protein on the T cells, large numbers of T-cells are isolated from a subject. Cas9 variants (*e.g.*, fCas9) and gRNA capable of inactivating CCR5 are then delivered to the isolated T cells using a viral vector, *e.g.*, an adenoviral vector.

Examples of suitable Cas9 variants include those inventive fusion proteins provided herein. Examples of suitable target sequences for gRNAs targetting the CCR5 allele include those described in Figure 19, *e.g.*, SEQ ID NOs:303-310 and 312-317. The viral vector(s) capable of expressing the Cas9 variant and gRNA is/are added to the isolated T cells to knock out the CCR5 protein. When the T cells are returned to subject, there is minimal adenovirus or Cas9 variant protein present. The removal of the CCR5 protein on the T cells subjects receive, however, is permanent. The cells are then reintroduced to the subject for the treatment or prevention of HIV/AIDS.

Example 3: Cas9-recombinase fusion proteins

[00193] Exemplary Cas9-recombinase fusion proteins are provided below:

[00194] *dCas9-NLS-GGS3linker-Tn3*

MDKKYSIGLAIGTNSVGWAVITDEYKVPSKKFKVLGNTDRHSIKKNLIGALLFDSGE
TAEATRLKRTARRRYTRRKNRICYLQEIFSNEMAKVDDSFHRLEESFLVEEDKKHE
RHPIFGNIVDEVAYHEKYPTIYHLRKKLVSTDKADLRLIYLALAHMIKFRGHFLIEG
DLNPDNSDVKLFIQLVQTYNQLFEENPINASGVDAKAILSARLSKSRLENLIAQLP
GEKKNGLFGNLIALSLGLTPNFKSNFDLAEDAKLQLSKDITYDDDLNLLAQIGDQYA
DLFLAAKNLSDAILLSDILRVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQLP
KYKEIFFDQSKNGYAGYIDGGASQEEFYKFIKPILEKMDGTEELLVKLNREDLLRKQR
TFDNGSIPHQIHLGELHAILRRQEDFYFPLKDNREKIEKILTFRIPIYYVGPLARGNSRFA
WMTRKSEETITPWNFEEVVDKGASASQFIERMTNFDKNLPNEKVLPKHSLLYEYFTV
YNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKIECFD
SVEISGVEDRFNASLGTYHDLLKIIKDKDFLDNEENEDILEDIVLTTLTFEDREMIEERL
KTYAHLFDDKVMKQLKRRRYTGWGRLSRKLINGIRDKQSGKTILDFLKSDGFANRN
FMQLIHDDSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPAIIKKGILQTVKVVDDELVK
VMGRHKPENIVIEMARENQTTQKGQKNSRERMKRIEEGIKELGSQILKEHPVENTQL
QNEKLYLYYLQNGRDMYVDQELDINRLSDYDVDAIVPQSFLKDDSIDNKVLTRSDK
NRGKSDNVPSEEVVKKMKNYWRQLLNAKLITQRKFDNLTKAERGGLSELDKAGFIK
RQLVETRQITKHVAQILDSRMNTKYDENDKLIREVKVITLKSCLVSDFRKDFQFYKV
REINNYHHAHDAYLNAVVGTAIIKKYPKLESEFVYGDYKVYDVRKMIKSEQEIGK
ATAKYFFYSNIMNFFKTEITLANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVLSM
PQVNIVKKTEVQTGGFSKESILPKRNSDKLIARKKDWDPKKYGGFDSPTVAYSVLVV
AKVEKGKSKKLKSVKELLGITIMERSSSFENPIDFLEAKGYKEVKKDLIIKLPKYSLFE
LENGRKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLKGSPEDNEQKQLFVEQ
HKHYLDEIIEQISEFSKRVLADANLDKVLSAYNKHARDKPIREQAENIIHLFTLTNLGA
PAAFKYFDTTIDRKRYTSTKEVLDTLIHQSTGLYETRIDLSQLGGDMAPKKKRKVG
IHRGVPGGSGGSGGSMALFGYARVSTSQQSLDLQVRALKDAGVKANRIFTDKASGS
STDREGLDLLRMKVKEGDVILVKKLDRLGRDTADMLQLIKEFDAQGVAVRFIDDGIS
TDSYIGLMFVTILSAVAQAERRRILERTNEGRQAALKKGIFGRRR (SEQ ID NO:328)
(underline: nuclear localization signal; bold: linker sequence)

[00195] *NLS-dCas9-GGS3linker-Tn3*

MAPKKKRKVGIHRGVPMDDKKYSIGLAIGTNSVGWAVITDEYKVPSKKFKVLGNTDR
HSIKKNLIGALLFDSGETAEATRLKRTARRRYTRRKNRICYLQEIFSNEMAKVDDSF
HRLEESFLVEEDKKHERHPIFGNIVDEVAYHEKYPTIYHLRKKLVDSTDKADLRLIYL
ALAHMIKFRGHFLIEGDLNPDNSDVKLFIQLVQTYNQLFEENPINASGVDAKAILS
ARLSKSRLENLIAQLPGEKKNGLFGNLIALSLGLTPNFKSNFDLAEDAKLQLSKDTYD
DDLDNLLAQIGDQYADLFLAAKNLSDAILLSDILRVNTEITKAPLSASMIKRYDEHHQ
DLTLLKALVRQQQLPEKYKEIFFDQSKNGYAGYIDGGASQEEFYKFIKPILEKMDGTEE
LLVCLNREDLLRKQRTFDNGSIPHQIHLGELHAILRRQEDFYPFLKDNREKIEKILTFRI
PYYVGPLARGNSRFAWMTRKSEETITPWNFEEVVDKGASAQSFIERMTNFDKNLPN
EKVLPKHSLLYEYFTVYNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVT
VKQLKEDYFKKIECFDSVEISGVEDRFNASLGTYHDLLKIIKDKDFLDNEENEDILEDI
VLTTLTLFEDREMIEERLKTYAHLFDDKVMKQLKRRRYTGWGRLSRKLINGIRDKQS
GKTILDFLKSDGFANRNFMQLIHDDSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPA
IKKILQTVKVVDDELVKVMGRHKPENIVIAMARENQTTQKGQKNSRERMKRIEEGIK
ELGSQILKEHPVENTQLQNEKLYLYYLQNGRDMYVDQELDINRLSDYDVDAIVPQSF
LKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKKMKNYWRQLLNAKLITQRKFDNLTK
AERGGLSELDKAGFIKRQLVETRQITKHVAQILDSRMNTKYDENDKLIREVKVITLKS
KLVSDFRKDFQFYKVREINNYHHAHDAYLNAVVGTALIKYPKLESEFVYGDYKVY
DVRKMIKSEQEIGKATAKYFFYSNIMNFFKTEITLANGEIRKRPLIETNGETGEIVWD
KGRDFATVRKVLSPQVNIVKKTEVQTGGFSKESILPKRNSDKLIARKKDWDPKKY
GGFDSPTVAYSVLVAKVEKGKSKKLKSVKELLGITIMERSSFEKNPIDFLEAKGYKE
VKKDLIIKLPKYSLFELENGRKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLK
GSPEDNEQKQLFVEQHKHYLDEIIEQISEFSKRVILADANLDKVLSAYNKHRDKPIRE
QAENIIHLFTLTNLGAPAAFKYFDTTIDRKRYTSTKEVLDATLIHQSITGLYETRIDLSQ
LGGDGGSGGSGGSMALFGYARVSTSQQSLDLQVRALKDAGVKANRIFTDKASGSS
TDREGLDLLRMKVKEGDVILVKKLDRLGRDTADMLQLIKEFDAQGVAVRFIDDGIST
DSYIGLMFVTILSAVAQAERRRILERTNEGRQAAKLKGIFGRRR (SEQ ID NO:329)
 (underline: nuclear localization signal; bold: linker sequence)

[00196] *Tn3-GGS3linker-dCas9-NLS*

MALFGYARVSTSQQSLDLQVRALKDAGVKANRIFTDKASGSSSTDREGLDLLRMKV
 EGDVILVKKLDRLGRDTADMLQLIKEFDAQGVAVRFIDDGISTDSYIGLMFVTILSA
 VAQAERRRILERTNEGRQA AKLKGIFGRRR**GGSGGSGGSM**DDKKYSIGLAIGTNSVG
 WAVITDEYKVPSKKFKVLGNTDRHSIKKNLIGALLFDSGETAEATRLKRTARRRYTR
 RKNRICYLQEIFSNEMAKVDDSFHRLEESFLVEEDKKHERHPIFGNIVDEVAYHEKY
 PTIYHLRKKLVDSTDKADLR LIYLALAHMIKFRGHFLIEGDLNPDNSDVKLFIQLVQ
 TYNQLFEE NPINASGVDAKAILSARLSKSRLENLIAQLPGEKKNGLFGNLIALSLGLT
 PNFKS NFDLAEDAKLQLSKD TYDDDLNLLAQIGDQYADLFLAAKNLSDAILLSDIL
 RVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQQLPEKYKEIFFDQSKNGYAGYI
 DGGASQEEFYKFIKPILEKMDGTEELLVCLNREDLLRKQRTFDNGSIPHQIHLGELHAI
 LRRQEDFY PFLKDNREKIEKILTFRI PYYVGPLARGNSRFAWMTRKSEETITPWNFEE
 VVDKGASAQSFIERMTNFDKNLPNEKVLPKHSLLYEYFTVYNELTKVKYVTEGMRK
 PAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKIECFDSVEISGVEDRFNASLGTY
 HDLLKIIKDKDFLDNEENEDILEDIVLTTLTLFEDREMIEERLKTYAHLFDDKVMKQLK
 RRRYTGWGRLSRKLINGIRDKQSGKTILDFLKSDGFANRNFMQLIHDDSLTFKEDIQK
 AQVSGQGDSLHEHIANLAGSPA IKKILQTVKVVDDELVKVMGRHKPENIVIAMAREN
 QTTQKGQKNSRERMKRIEEGIKELGSQILKEHPVENTQLQNEKLYLYYLQNGRDMY
 VDQELDINRLSDYDVDAIVPQSFLKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKKM
 KNYWRQLLNAKLITQRKFDNLTKAERGGLSELDKAGFIKRQLVETRQITKHVAQILD

SRMNTKYDENDKLIREVKVITLKS KLVSDFRKDFQFYKVREINNYHHAHDAYLNAV
 VGTALIKKYPKLESEFVYGDYKVYDVRKMIAKSEQEIGKATAKYFFYSNIMNFFKTEI
 TLANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVL SMPQVNIVKKTEVQTGGFSK
 ESILPKRNSDKLIARKKDWD PKKYGGFDSPTVAYSVLVVAKVEKGKSKKLKSVKEL
 LGITIMERS SFEKNPIDFLEAKGYKEVKKDLIIKLPKYSLFELENGRKRMLASAGELQK
 GNELALPSKYVNFLYLASHYEKLKGSPEDNEQKQLFVEQHKHYLDEIIEQISEFSKRV
 ILADANLDKVLSAYNKH RDKPIREQAENIIHLFTLTNLGAPAAFKYFDTTIDRKRYTS
 TKEVL DATLIHQ SITGLYETRIDLSQLGGD MAPKKKRKVGIHRGVP (SEQ ID NO:330)
 (underline: nuclear localization signal; bold: linker sequence)

[00197] *NLS-Tn3-GGS3linker-dCas9*

MAPKKKRKVGIHRGVP MALFGYARVSTSQQSLDLQVRALKDAGVKANRIFTDKAS
 GSSTDREGLDLLRMKVKEGDVILVKKLDRLGRDTADMLQLIKEFDAQGVAVRFIDD
 GISTDSYIGLMFVTILSAVAQAERRRILERTNEGRQAAKLKGIFGRRRGGS GGS
 MDKKYSIGLAIGTNSVGWAVITDEYKVPSKKFKVLGNTDRHSIKKNLIGALLFDSGE
 TAEATRLKRTARRRYTRRKNRICYLQEIFS NEMAKVDDSFHRLEESFLVEEDKKHE
 RHPIFGNIVDEVAYHEKYPTIYHLRKKLVDSTDKADLRLIYLALAHMIKFRGHFLIEG
 DLNPDNSDVKLFIQLVQTYNQLFEENPINASGVDAKAILSARLSKSRRLLENLIAQLP
 GEKKNGLFGNLIALSLGLTPNFKSNFDLAEDAKLQLSKD TYDDDLDNLLAQIGDQYA
 DLFLAAKNLSDAILLSDILRVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQLP
 KYKEIFFDQSKNGYAGYIDGGASQEEFYKFIKPILEKMDGTEELLVKLNREDLLRKQR
 TFDNGSIPHQIHLGELHAILRRQEDFY PFLKDNREKIEKILTFRIPYYVGPLARGNSRFA
 WMTRKSEETITPWNFEEVVDKGASAQSFIERM TNFDKNLPNEKVLPHSLLYEYFTV
 YNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKIECFD
 SVEISGVEDRFNASLGTYHDLLKIIKDKDFLDNEENEDILEDIVLTLT LFEDREMIEERL
 KTYAHLFDDKVMKQLKRRRYTGWGRLSRKLINGIRDKQSGKTILDFLKSDGFANRN
 FMQLIHDDSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPA IKKGILQTVKVVDLVK
 VMGRHKPENIVIEMARENQTTQKGQKNSRERMKRIEEGIKELGSQILKEHPVENTQL
 QNEKLYLYYLQNGRDMYVDQELDINRLSDYDVDAIVPQSFLKDDSIDNKVLTRSDK
 NRGKSDNVPSEEVVKMKMKNYWRQLLNAKLITQRKFDNLTKAERGGLSELDKAGFIK
 RQLVETRQITKHVAQILDSRMNTKYDENDKLIREVKVITLKS KLVSDFRKDFQFYKV
 REINNYHHAHDAYLNAVVG TALI KYPKLESEFVYGDYKVYDVRKMIAKSEQEIGK
 ATAKYFFYSNIMNFFKTEITLANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVL SM
 PQVNIVKKTEVQTGGFSKESILPKRNSDKLIARKKDWD PKKYGGFDSPTVAYSVLV
 AKVEKGKSKKLKSVKELLGITIMERS SFEKNPIDFLEAKGYKEVKKDLIIKLPKYSLFE
 LENG RKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLKGSPEDNEQKQLFVEQ
 HKHYLDEIIEQISEFSKRVILADANLDKVLSAYNKH RDKPIREQAENIIHLFTLTNLGA
 PAAFKYFDTTIDRKRYTSTKEVL DATLIHQ SITGLYETRIDLSQLGGD (SEQ ID
 NO:331)

(underline: nuclear localization signal; bold: linker sequence)

[00198] *dCas9-NLS-GGS3linker-Hin*

MDKKYSIGLAIGTNSVGWAVITDEYKVPSKKFKVLGNTDRHSIKKNLIGALLFDSGE
 TAEATRLKRTARRRYTRRKNRICYLQEIFS NEMAKVDDSFHRLEESFLVEEDKKHE
 RHPIFGNIVDEVAYHEKYPTIYHLRKKLVDSTDKADLRLIYLALAHMIKFRGHFLIEG
 DLNPDNSDVKLFIQLVQTYNQLFEENPINASGVDAKAILSARLSKSRRLLENLIAQLP
 GEKKNGLFGNLIALSLGLTPNFKSNFDLAEDAKLQLSKD TYDDDLDNLLAQIGDQYA

DLFLAAKNLSDAILLSDILRVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQLP
 KYKEIFFDQSKNGYAGYIDGGASQEEFYKFIKPILEKMDGTEELLVKLNREDLLRKQR
 TFDNGSIPHQIHLGELHAILRRQEDFYFPLKDNREKIEKILTFRIPYYVGPLARGNSRFA
 WMTRKSEETITPWNFEEVVDKGASAQSFIERMTNFDKNLPNEKVLPKHSLLYEYFTV
 YNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKIECFD
 SVEISGVEDRFNASLGTYHDLLKIIKDKDFLDNEENEDILEDIVLTTLTFEDREMIEERL
 KTYAHLFDDKVMKQLKRRRYTGWGRLSRKLINGIRDKQSGKTILDFLKSDGFANRN
 FMQLIHDDSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPAIIKKGILQTVKVVDDELVK
 VMGRHKPENIVIAMARENQTTQKGQKNSRERMKRIEEGIKELGSQILKEHPVENTQL
 QNEKLYLYYLQNGRDMYVDQELDINRLSDYDVDAIVPQSFLKDDSIDNKVLTRSDK
 NRGKSDNVPSEEVVKKMKNYWRQLLNAKLITQRKFDNLTKAERGGLSELDKAGFIK
 RQLVETRQITKHVAQILDSRMNTKYDENDKLIREVKVITLKS KLVSDFRKDFQFYKV
 REINNYHHAHDAYLNAVVG TALIKKYPKLESEFVYGDYKVYDVRKMIAKSEQEIGK
 ATAKYFFYSNIMNFFKTEITLANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVLSM
 PQVNIVKKTEVQTGGFSKESILPKRNSDKLIARKKDWDPKKYGGFDSPTVAYSVLVV
 AKVEKGKSKKLKSVKELLGITIMERSSFEKNPIDFLEAKGYKEVKKDLIIKLPKYSLFE
 LENGKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLKGSPEDNEQKQLFVEQ
 HKHYLDEIIIEQISEFSKRVLADANLDKVLSAYNKHARDKPIREQAENIIHLFTLTNLGA
 PAAFKYFDTTIDRKRYTSTKEVL DATLIHQ SITGLYETRIDLSQLGGD MAPKKRKVG
IHRGVP **GGSGGSGGSM**ATIGYIRVSTIDQNIDLQRNALTSANCDRIFEDRISGKIANRP
 GLKRALKYVNKGDTLVVWKLDRGRSVKNLVALISELHERGAHFHSLTDSIDTSSA
 MGRFFFYVMSALAEMERELIVERTLAGLAAARAQGR LG (SEQ ID NO:332)
 (underline: nuclear localization signal; bold: linker sequence)

[00199] *NLS-dCas9-GGS3linker-Hin*

MAPKKRKRVGIHRGVPMDKKYSIGLAIGTNSVGWAVITDEYKVPSKKFKVLGNTDR
 HSIKKNLIGALLFDSGETAEATRLKRTARRRYTRRKNRICYLQEIFSNEMAKVDDSF
 HRLEESFLVEEDKKHERHPIFGNIVDEVAYHEKYPTIYHLRKKLVDSTDKADLRILIY
 ALAHMIKFRGHFLIEGDLNPDNSDVKLFIQLVQTYNQLFEENPINASGVDAKAILS
 RLSKSRRLENLIAQLPGEKKNGLFGNLIALSLGLTPNFKSNFDLAEDAKLQLSKDTYD
 DDLDNLLAQIGDQYADLFLAAKNLSDAILLSDILRVNTEITKAPLSASMIKRYDEHHQ
 DLTLLKALVRQQLP EKYKEIFFDQSKNGYAGYIDGGASQEEFYKFIKPILEKMDGTEE
 LLVKLNREDLLRKQR TFDNGSIPHQIHLGELHAILRRQEDFYFPLKDNREKIEKILTFRI
 PYYVGPLARGNSRFAWMTRKSEETITPWNFEEVVDKGASAQSFIERMTNFDKNLPN
 EKVLPKHSLLYEYFTVYNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVT
 VKQLKEDYFKKIECFDSVEISGVEDRFNASLGTYHDLLKIIKDKDFLDNEENEDILEDI
 VLTTLTFEDREMIEERL KTYAHLFDDKVMKQLKRRRYTGWGRLSRKLINGIRDKQS
 GKTILDFLKSDGFANRNFMQLIHDDSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPA
 I KKGILQTVKVVDDELVKVMGRHKPENIVIAMARENQTTQKGQKNSRERMKRIEEGIK
 ELGSQILKEHPVENTQLQNEKLYLYYLQNGRDMYVDQELDINRLSDYDVDAIVPQSF
 LKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKKMKNYWRQLLNAKLITQRKFDNLTK
 AERGGLSELDKAGFIKRQLVETRQITKHVAQILDSRMNTKYDENDKLIREVKVITLKS
 KLVSDFRKDFQFYKVREINNYHHAHDAYLNAVVG TALIKKYPKLESEFVYGDYKVY
 DVRKMIAKSEQEIGKATAKYFFYSNIMNFFKTEITLANGEIRKRPLIETNGETGEIVWD
 KGRDFATVRKVLSMPQVNIVKKTEVQTGGFSKESILPKRNSDKLIARKKDWDPKKY
 GGFDSPTVAYSVLVVAKVEKGKSKKLKSVKELLGITIMERSSFEKNPIDFLEAKGYKE
 VKKDLIIKLPKYSLFELENGKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLK
 GSPEDNEQKQLFVEQHKHYLDEIIIEQISEFSKRVLADANLDKVLSAYNKHARDKPIRE
 QAENIIHLFTLTNLGAPAAFKYFDTTIDRKRYTSTKEVL DATLIHQ SITGLYETRIDLSQL

LGGDGGSGGSGGSMATIGYIRVSTIDQNIDLQRNALTSANCDRIFEDRISGKIANRPG
LKRALKYVNKGDTLVVWKLDRGRSVKNLVALISELHERGAHFHSLTDSIDTSSAM
GRFFFYVMSALAEMERELIVERTLAGLAAARAQGRLG (SEQ ID NO:333)
(underline: nuclear localization signal; bold: linker sequence)

[00200] *Hin-GGS3linker-dCas9-NLS*

MATIGYIRVSTIDQNIDLQRNALTSANCDRIFEDRISGKIANRPG LKRALKYVNKGDT
LVVWKLDRGRSVKNLVALISELHERGAHFHSLTDSIDTSSAMGRFFFYVMSALAE
MERELIVERTLAGLAAARAQGRLG **GGGSGGSGGSM**DKKYSIGLAIGTNSVGWAVITD
EYKVPSKKFKVLGNTDRHSIKKNLIGALLFDSGETAEATRLKRTARRRYTRRKNRIC
YLQEIFSNEMAKVDDSFHRLEESFLVEEDKKHERHPIFGNIVDEVAYHEKYPTIYHL
RKKLVDSTDKADLRLIYLALAHMIKFRGHFLIEGDLNPDNSDVDKLFQQLVQTYNQL
FEENPINASGVDAKAILSARLSKSRRLENLIAQLPGEKKNGLFGNLIALSLGLTPNFKS
NFDLAEDAKLQLSKDTYDDDLNLLAQIGDQYADLFLAAKNLSDAILLSDILRVNTE
ITKAPLSASMIKRYDEHHQDLTLLKALVRQQLEPKYKEIFFDQSKNGYAGYIDGGAS
QEEFYKFIKPILEKMDGTEELLVKLNREDLLRKQRTFDNGSIPHQIHLGELHAILRRQE
DFYPFLKDNREKIEKILTRIPYYVGPLARGNSRFAWMTRKSEETITPWNFEEVVDKG
ASAQSFIERMTNFDKNLPNEKVLPKHSLLYEYFTVYNELTKVKYVTEGMRKPAFLSG
EQKKAIVDLLFKTNRKVTVKQLKEDYFKKIECFDSVEISGVEDRFNASLGTYHDLLKI
IKDKDFLDNEENEDILEDIVLTTLTFEDREMIEERLKTYAHLFDDKVMKQLKRRRYT
GWGRLSRKLINGIRDKQSGKTILDFLKSDGFANRNFQMQLIHDDSLTFKEDIQKAQVSG
QGDSLHEHIANLAGSPAIKKGILQTVKVVDDELVKVMGRHKPENIVIEMARENQTTQK
GQKNSRERMKRIEIEGKELGSQILKEHPVENTQLQNEKLYLYLQNGRDMYVDQEL
DINRLSDYDVDAIVPQSFLKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKKMKKNYWR
QLLNAKLITQRKFDNLTKAERGGLSELDKAGFIKRQLVETRQITKHVAQILDSRMNT
KYDENDKLIREVKVITLKSCLVSDFRKDFQFYKVREINNYHHAHDAYLNAVVGITALI
KKYPKLESEFVYGDYKVYDVRKMIKSEQEIGKATAKYFFYSNIMNFFKTEITLANG
EIRKRPLIETNGETGEIVWDKGRDFATVRKVLSMPQVNIVKKTEVQTGGFSKESILPK
RNSDKLIARKKDWDPKKYGGFDSPTVAYSVLVVAKEKGKSKKLKSVKELLGITIM
ERSSFENPIDFLEAKGYKEVKKDLIILPKYSLFELENGRKRMLASAGELQKGNELA
LPSKYVNFLYLASHYEKLKGSPEDEQKQLFVEQHKHYLDEIIEQISEFSKRVLADA
NLDKVLSAYNKHRDKPIREQAENIIHLFTLTNLGAPAAFKYFDTTIDRKRYTSTKEVL
DATLIHQSTGLYETRIDLSQLGGD MAPKKKRKVGIHRGVP (SEQ ID NO:334)
(underline: nuclear localization signal; bold: linker sequence)

[00201] *NLS-Hin-GGS3linker-dCas9*

MAPKKKRKVGIHRGVP MATIGYIRVSTIDQNIDLQRNALTSANCDRIFEDRISGKIAN
RPGLKRALKYVNKGDTLVVWKLDRGRSVKNLVALISELHERGAHFHSLTDSIDTSS
AMGRFFFYVMSALAEMERELIVERTLAGLAAARAQGRLG **GGGSGGSGGSM**DKKYSI
GLAIGTNSVGWAVITDEYKVPSKKFKVLGNTDRHSIKKNLIGALLFDSGETAEATRL
KRTARRRYTRRKNRICYLQEIFSNEMAKVDDSFHRLEESFLVEEDKKHERHPIFGNI
VDEVAYHEKYPTIYHLRKKLVDSTDKADLRLIYLALAHMIKFRGHFLIEGDLNPDNS
DVDKLFQQLVQTYNQLFEENPINASGVDAKAILSARLSKSRRLENLIAQLPGEKKNGL
FGNLIALSLGLTPNFKS NFDLAEDAKLQLSKDTYDDDLNLLAQIGDQYADLFLAAK
NLSDAILLSDILRVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQLEPKYKEIFFD
QSKNGYAGYIDGGASQEEFYKFIKPILEKMDGTEELLVKLNREDLLRKQRTFDNGSIP
HQIHLGELHAILRRQEDFYPFLKDNREKIEKILTRIPYYVGPLARGNSRFAWMTRKS

EETITPWNFEEVVDKGASAQSFIERMTNFDKNLPNEKVLPKHSLLYEYFTVYNELTK
VKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKIECFDSVEISGV
EDRFNASLGTYHDLLKIIKDKDFLDNEENEDILEDIVLTTLTFEDREMIEERLKTYAHL
FDDKVMKQLKRRRYTGWGRLSRKLINGIRDKQSGKTILDFLKSDGFANRNFQMQLIH
DDSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPAIIKKGILQTVKVVDDELVKVMGRHK
PENIVIAMARENQTTQKGQKNSRERMKRIEEGIKELGSQILKEHPVENTQLQNEKLYL
YYLQNGRDMYVDQELDINRLSDYDVDAIVPQSFLKDDSIDNKVLTRSDKNRGKSDN
VPSEEVVKMKMKNYWRQLLNAKLITQRKFDNLTKAERGGLSELDKAGFIKRQLVETR
QITKHVAQILDSRMNTKYDENDKLIREVKVITLKSCLVSDFRKDFQFYKVREINNYH
HAHDAYLNAVVGTAIIKKYPKLESEFVYGDYKVYDVRKMIKSEQEIGKATAKYFF
YSNIMNFFKTEITLANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVLSPQVNIKK
KTEVQTGGFSKESILPKRNSDKLIARKKDWDPKKYGGFDSPTVAYSVLVVAKEVKG
KSKKLKSVKELLGITIMERSSEFEKNPIDFLEAKGYKEVKKDLIIKLPKYSLELENGRK
RMLASAGELQKGNELALPSKYVNFLYLASHYEKLGKSPEDNEQKQLFVEQHKHYLD
EIIQISEFSKRVLADANLDKVL SAYNKHRDKPIREQAENIIHLFTLTNLGAPAAFKYF
DTTIDRKRYTSTKEVLDATLIHQSTGLYETRIDLSQLGGD (SEQ ID NO:335)

(underline: nuclear localization signal; bold: linker sequence)

[00202] *dCas9-NLS-GGS3linker-Gin*

MDKKYSIGLAIGTNSVGWAVITDEYKVPSKKFKVLGNTDRHSIKKNLIGALLFDSGE
TAEATRLKRTARRRYTRRKNRICYLQEIFSNEMAKVDDSFHRLEESFLVEEDKKHE
RHPIFGNIVDEVAYHEKYPTIYHLRKKLVDSTDKADLRLIYLALAHMIKFRGHFLIEG
DLNPDNSDVKLFIQLVQTYNQLFEENPINASGVDAKAILSARLSKSRLENLIAQLP
GEKKNGLFGNLIALSLGLTPNFKSNFDLAEDAKLQLSKDTYDDDLDNLLAQIGDQYA
DLFLAAKNLSDAILLSDILRVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQLPE
KYKEIFFDQSKNGYAGYIDGGASQEEFYKFIKPILEKMDGTEELLVKLNREDLLRKQR
TFDNGSIPHQIHLGELHAILRRQEDFYFPLKDNREKIEKILTFRIPYYVGPLARGNSRFA
WMTRKSEETITPWNFEEVVDKGASAQSFIERMTNFDKNLPNEKVLPKHSLLYEYFTV
YNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKIECFD
SVEISGVEDRFNASLGTYHDLLKIIKDKDFLDNEENEDILEDIVLTTLTFEDREMIEERL
KTYAHLFDDKVMKQLKRRRYTGWGRLSRKLINGIRDKQSGKTILDFLKSDGFANRN
FMQLIHDDSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPAIIKKGILQTVKVVDDELVK
VMGRHKPENIVIAMARENQTTQKGQKNSRERMKRIEEGIKELGSQILKEHPVENTQL
QNEKLYLYYLQNGRDMYVDQELDINRLSDYDVDAIVPQSFLKDDSIDNKVLTRSDK
NRGKSDNVPSEEVVKMKMKNYWRQLLNAKLITQRKFDNLTKAERGGLSELDKAGFIK
RQLVETRQITKHVAQILDSRMNTKYDENDKLIREVKVITLKSCLVSDFRKDFQFYKV
REINNYHHAHDAYLNAVVGTAIIKKYPKLESEFVYGDYKVYDVRKMIKSEQEIGK
ATAKYFFYSNIMNFFKTEITLANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVLSP
PQVNIKKTEVQTGGFSKESILPKRNSDKLIARKKDWDPKKYGGFDSPTVAYSVLVVA
KEVKGKSKKLKSVKELLGITIMERSSEFEKNPIDFLEAKGYKEVKKDLIIKLPKYSLE
LENGRKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLGKSPEDNEQKQLFVEQ
HKHYLDEIIQISEFSKRVLADANLDKVL SAYNKHRDKPIREQAENIIHLFTLTNLGA
PAAFKYFDTTIDRKRYTSTKEVLDATLIHQSTGLYETRIDLSQLGGDMAPKKKRKVG
IHRGVPGGSGGSGGSMLIGYVRVSTNDQNTDLQRNALVCAGCEQIFEDKLSGTRTD
RPGLKRALKRLQKGDTLVWVKLDRLGRSMKHLISLVGELRERGINFRSLTDSIDTSSP
MGRFFFYVMGALAEMERELIERTMAGIAAARNKGRRFRPPKSG (SEQ ID NO:336)

(underline: nuclear localization signal; bold: linker sequence)

[00203] *NLS-dCas9-GGS3linker-Gin*

MAPKKKRKVGIHRGVPMDDKKYSIGLAIGTNSVGWAVITDEYKVPSKKFKVLGNTDR
HSIKKNLIGALLFDSGETAEATRLKRTARRRYTRRKNRICYLQEIFSNE
MAKVDDDSFFHRLEESFLVEEDKKHERHPIFGNIVDEVAYHEKYPTIYHLRKKLVDSTDKADLR
LIYLALAHMIKFRGHFLIEGDLNPDNSDVKLFIQLVQTYNQLFEENPINASGVDAKAILS
ARLSKSRLENLIAQLPGEKKNGLFGNLIALSLGLTPNFKSNFDLAEDAKLQLSKDTYD
DDLDNLLAQIGDQYADLFLAAKNLSDAILLSDILRVNTEITKAPLSASMIKRYDEHHQ
DLTLLKALVRQQQLPEKYKEIFFDQSKNGYAGYIDGGASQEEFYKFIKPILEKMDGTEE
LLVKLNREDLLRKQRTFDNGSIPHQIHLGELHAILRRQEDFYPFLKDNREKIEKILTFRI
PYYVGPLARGNSRFAWMTRKSEETITPWNFEEVVDKGASAQSFIERMTNFDKNLPN
EKVLPHSLLYEYFTVYNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVT
VKQLKEDYFKKIECFDSVEISGVEDRFNASLGTYHDLLKIIKDKDFLDNEENEDILEDI
VLTLTLFEDREMIEERLKTYAHLFDDKVMKQLKRRRYTGWGRLSRKLINGIRDKQS
GKTILDFLKSDGFANRNFMQLIHDDSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPA
IKKILQTVKVVDELVKVMGRHKPENIVIEMARENQTTQKGQKNSRERMKRIEEGIK
ELGSQILKEHPVENTQLQNEKLYLYYLQNGRDMYVDQELDINRLSDYDVDAIVPQSF
LKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKKMKNYWRQLLNAKLITQRKFDNLTK
AERGGLSELDKAGFIKRQLVETRQITKHVAQILDSRMNTKYDENDKLIREVKVITLKS
KLVSDFRKDFQFYKVREINNYHHAHDAYLNAVVGTALIKKYPKLESEFVYGDYKVY
DVRKMIKSEQEIGKATAKYFFYSNIMNFFKTEITLANGEIRKRPLIETNGETGEIVWD
KGRDFATVRKVLSMPQVNIVKKTEVQTGGFSKESILPKRNSDKLIARKKDWDPKKY
GGFDSPTVAYSVLVAKVEKGKSKKLKSVKELLGITIMERSSFEKNPIDFLEAKGYKE
VKKDLIIKLPKYSLFELNGRKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLK
GSPEDNEQKQLFVEQHKHYLDEIIEQISEFSKRVILADANLDKVLSAYNKHRDKPIRE
QAENIIHLFTLTNLGAPAAFKYFDTTIDRKRYTSTKEVLDATLIHQSITGLYETRIDLSQ
LGGDGGSGGSGGSMLIGYVRVSTNDQNTDLQRNALVCAGCEQIFEDKLSGTRTDRP
GLKRALKRLQKGDTLVVWKLDRLGRSMKHLISLVGELRERGINFRSLTDSIDTSSPM
GRFFFYVMGALAEMERELIIERTMAGIAAARNKGRRFGRPPKSG (SEQ ID NO:337)
 (underline: nuclear localization signal; bold: linker sequence)

[00204] *Gin-GGS3linker-dCas9-NLS*

MLIGYVRVSTNDQNTDLQRNALVCAGCEQIFEDKLSGTRTDRPGLKRALKRLQKGD
TLVVWKLDRLGRSMKHLISLVGELRERGINFRSLTDSIDTSSPMGRFFFYVMGALAE
MERELIIERTMAGIAAARNKGRRFGRPPKSGGGSGGSGGSMDDKKYSIGLAIGTNSVG
WAVITDEYKVPSKKFKVLGNTDRHSIKKNLIGALLFDSGETAEATRLKRTARRRYTR
RKNRICYLQEIFSNEMAKVDDDSFFHRLEESFLVEEDKKHERHPIFGNIVDEVAYHEKY
PTIYHLRKKLVDSTDKADLRLIYLALAHMIKFRGHFLIEGDLNPDNSDVKLFIQLVQ
TYNQLFEENPINASGVDAKAILSARLSKSRLENLIAQLPGEKKNGLFGNLIALSLGLT
PNFKSNFDLAEDAKLQLSKDTYDDDLNLLAQIGDQYADLFLAAKNLSDAILLSDIL
RVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQQLPEKYKEIFFDQSKNGYAGYI
DGGASQEEFYKFIKPILEKMDGTEELLVKLNREDLLRKQRTFDNGSIPHQIHLGELHAI
LRRQEDFYPFLKDNREKIEKILTFRIPYYVGPLARGNSRFAWMTRKSEETITPWNFEE
VVDKGASAQSFIERMTNFDKNLPNEKVLPHSLLYEYFTVYNELTKVKYVTEGMRK
PAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKIECFDSVEISGVEDRFNASLGTY
HDLLKIIKDKDFLDNEENEDILEDIVLTLTLFEDREMIEERLKTYAHLFDDKVMKQLK
RRRYTGWGRLSRKLINGIRDKQSGKTILDFLKSDGFANRNFMQLIHDDSLTFKEDIQK
AQVSGQGDSLHEHIANLAGSPAIKKILQTVKVVDELVKVMGRHKPENIVIEMAREN
QTTQKGQKNSRERMKRIEEGIKELGSQILKEHPVENTQLQNEKLYLYYLQNGRDMY

VDQELDINRLSDYDVDAIVPQSFLKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKKM
 KNYWRQLLNAKLITQRKFDNLTKAERGGLSELDKAGFIKRQLVETRQITKHVAQILD
 SRMNTKYDENDKLIREVKVITLKSCLVSDFRKDFQFYKVREINNYHHAHDAYLNAV
 VGTALIKKYPKLESEFVYGDYKVYDVRKMIKSEQEIGKATAKYFFYSNIMNFFKTEI
 TLANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVLSPQVNIKKTEVQTGGFSK
 ESILPKRNSDKLIARKKDWDPKKYGGFDSPTVAYSVLVVAKEKGKSKKLKSVKEL
 LGITIMERSSEFEKNPIDFLEAKGYKEVKKDLIILPKYSLFELENGRKRMLASAGELQK
 GNELALPSKYVNFLYLASHYEKLKGSPEDNEQKQLFVEQHKHYLDEIIEQISEFSKR
 VILADANLDKVL SAYNKH RD KPIREQAENIIHLFTLTNLGAPAAFKYFDTTIDRKRYTS
 TKEVL DATLIHQ SITGLYETRIDLSQLGGD MAPKKKRKVGIHRGVP (SEQ ID NO:338)
 (underline: nuclear localization signal; bold: linker sequence)

[00205] *NLS-Gin-GGS3linker-dCas9*

MAPKKKRKVGIHRGVPMLIGYVRVSTNDQNTDLQRNALVCAGCEQIFEDKLSGTRT
 DRPGLKRALKRLQKGD TLV VWKLDRLGRSMKHLISLVGELRERGINFRSLTDSIDTS
 SPMGRFFFYVMGALAEMERELIERTMAGIAAARNKGRRFGRPPKSGGGSGGGSGGS
 MDKKYSIGLAIGTNSVGWAVITDEYKVPSKKFKVLGNTDRHSIKKNLIGALLFDSGE
 TAEATRLKRTARRRYTRRKNRICYLQEIFSNEMAKVDDSFHRLEESFLVEEDKKHE
 RHPIFGNIVDEVAYHEKYPTIYHLRKKLV DSTDKADRLIYLALAHMIKFRGHFLIEG
 DLNPDNSDVKLFIQLVQTYNQLFEENPINASGVDAKAILSARLSKSRLENLIAQLP
 GEKKNGLFGNLIALSLGLTPNFKSNFDLAEDAKLQLSKD TYDDDLDNLLAQIGDQYA
 DLFLAAKNLSDAILLSDILRVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQLP
 KYKEIFFDQSKNGYAGYIDGGASQEEFYKFIKPILEKMDGTEELLVKLNREDLLRKQR
 TFDNGSIPHQIHLGELHAILRRQEDFYFPLKDNREKIEKILTFRIPYYVGPLARGNSRFA
 WMTRKSEETITPWNFEEVVDKGASAQSFIERMTNFDKNLPNEKVLPKHSLLYEYFTV
 YNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTVKQLKEDYFKKIECFD
 SVEISGVEDRFNASLGT YHDLLKIIKDKDFLDNEENEDILEDIVLTTLTFEDREMIEERL
 KTYAHLFDDKVMKQLKRRRYTGWGRLSRKLINGIRDKQSGKTILDFLKSDGFANRN
 FMQLIHDDSLTFKEDIQKAQVSGQGDSLHEHIANLAGSPAIIKKGILQTVKVVDLVK
 VMGRHKPENIVIAMARENQTTQKGQKNSRERMKRIEEGIKELGSQILKEHPVENTQL
 QNEKLYLYYLQNGRDMYVDQELDINRLSDYDVDAIVPQSFLKDDSIDNKVLTRSDK
 NRGKSDNVPSEEVVKKMKNYWRQLLNAKLITQRKFDNLTKAERGGLSELDKAGFIK
 RQLVETRQITKHVAQILDSRMNTKYDENDKLIREVKVITLKSCLVSDFRKDFQFYKV
 REINNYHHAHDAYLNAVVG TALI K KYPKLESEFVYGDYKVYDVRKMIKSEQEIGK
 ATAKYFFYSNIMNFFKTEITLANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVLSP
 QVNIKKTEVQTGGFSKESILPKRNSDKLIARKKDWDPKKYGGFDSPTVAYSVLV
 VAKVEKGKSKKLKSVKEL LGITIMERSSEFEKNPIDFLEAKGYKEVKKDLIILPKYSLFE
 LENG RKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLKGSPEDNEQKQLFVEQ
 HKHYLDEIIEQISEFSKR VILADANLDKVL SAYNKH RD KPIREQAENIIHLFTLTNLGA
 PAAFKYFDTTIDRKRYTSTKEVL DATLIHQ SITGLYETRIDLSQLGGD (SEQ ID
 NO:339)
 (underline: nuclear localization signal; bold: linker sequence)

**Example 4: Introduction of a marker gene by homologous recombination using Cas9-
 recombinase fusion proteins**

[00206] A vector carrying a green fluorescent protein (GFP) marker gene flanked by genomic sequence of a host cell gene is introduced into a cell, along with an expression construct encoding a dCas9-recombinase fusion protein (any one of SEQ ID NO:328-339) and four appropriately designed gRNAs targetting the GFP marker gene and the genomic locus into which the GFP marker is recombined. Four dCas9-recombinase fusion proteins are coordinated at the genomic locus along with the GFP marker gene through the binding of the gRNAs (Figure 5B). The four recombinase domains of the fusion proteins tetramerize, and the recombinase activity of the recombinase domains of the fusion protein results in the recombination between the genomic locus and the marker gene, thereby introducing the marker gene into the genomic locus. Introduction of the marker gene is confirmed by GFP expression and/or by PCR.

EQUIVALENTS AND SCOPE

[00207] Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. The scope of the present invention is not intended to be limited to the above description, but rather is as set forth in the appended claims.

[00208] In the claims articles such as “a,” “an,” and “the” may mean one or more than one unless indicated to the contrary or otherwise evident from the context. Claims or descriptions that include “or” between one or more members of a group are considered satisfied if one, more than one, or all of the group members are present in, employed in, or otherwise relevant to a given product or process unless indicated to the contrary or otherwise evident from the context. The invention includes embodiments in which exactly one member of the group is present in, employed in, or otherwise relevant to a given product or process. The invention also includes embodiments in which more than one, or all of the group members are present in, employed in, or otherwise relevant to a given product or process.

[00209] Furthermore, it is to be understood that the invention encompasses all variations, combinations, and permutations in which one or more limitations, elements, clauses, descriptive terms, *etc.*, from one or more of the claims or from relevant portions of the description is introduced into another claim. For example, any claim that is dependent on another claim can be modified to include one or more limitations found in any other claim that is dependent on the same base claim. Furthermore, where the claims recite a composition, it is to be understood that methods of using the composition for any of the purposes disclosed herein are included, and methods of making the composition according to

any of the methods of making disclosed herein or other methods known in the art are included, unless otherwise indicated or unless it would be evident to one of ordinary skill in the art that a contradiction or inconsistency would arise.

[00210] Where elements are presented as lists, *e.g.*, in Markush group format, it is to be understood that each subgroup of the elements is also disclosed, and any element(s) can be removed from the group. It is also noted that the term “comprising” is intended to be open and permits the inclusion of additional elements or steps. It should be understood that, in general, where the invention, or aspects of the invention, is/are referred to as comprising particular elements, features, steps, *etc.*, certain embodiments of the invention or aspects of the invention consist, or consist essentially of, such elements, features, steps, *etc.* For purposes of simplicity those embodiments have not been specifically set forth *in haec verba* herein. Thus for each embodiment of the invention that comprises one or more elements, features, steps, *etc.*, the invention also provides embodiments that consist or consist essentially of those elements, features, steps, *etc.*

[00211] Where ranges are given, endpoints are included. Furthermore, it is to be understood that unless otherwise indicated or otherwise evident from the context and/or the understanding of one of ordinary skill in the art, values that are expressed as ranges can assume any specific value within the stated ranges in different embodiments of the invention, to the tenth of the unit of the lower limit of the range, unless the context clearly dictates otherwise. It is also to be understood that unless otherwise indicated or otherwise evident from the context and/or the understanding of one of ordinary skill in the art, values expressed as ranges can assume any subrange within the given range, wherein the endpoints of the subrange are expressed to the same degree of accuracy as the tenth of the unit of the lower limit of the range.

[00212] In addition, it is to be understood that any particular embodiment of the present invention may be explicitly excluded from any one or more of the claims. Where ranges are given, any value within the range may explicitly be excluded from any one or more of the claims. Any embodiment, element, feature, application, or aspect of the compositions and/or methods of the invention, can be excluded from any one or more claims. For purposes of brevity, all of the embodiments in which one or more elements, features, purposes, or aspects is excluded are not set forth explicitly herein.

[00213] All publications, patents and sequence database entries mentioned herein, including those items listed above, are hereby incorporated by reference in their entirety as if each individual publication or patent was specifically and individually indicated to be

incorporated by reference. In case of conflict, the present application, including any definitions herein, will control.

CLAIMS

What is claimed is:

1. A fusion protein comprising two domains: (i) a nuclease-inactivated Cas9; and (ii) a nuclease.
2. The fusion protein of claim 1, wherein the nuclease is a monomer of the FokI DNA cleavage domain.
3. A dimer of the fusion protein of claim 1 or 2.
4. The fusion protein of claim 1 or 2, wherein the nuclease-inactivated Cas9 domain binds a gRNA.
5. A method for site-specific DNA cleavage, comprising:
 - (a) contacting a DNA with a fusion protein of claim 4, wherein the inactive Cas9 domain binds a gRNA that hybridizes to a region of the DNA;
 - (b) contacting the DNA with a second fusion protein of claim 4, wherein the inactive Cas9 domain of the second fusion protein binds a second gRNA that hybridizes to a second region of DNA;wherein the binding of the fusion proteins in steps (a) and (b) results in the dimerization of the nuclease domains of the fusion proteins, such that the DNA is cleaved in a region between the bound fusion proteins.
6. The method of claim 5, wherein the gRNAs of steps (a) and (b) hybridize to the same strand of the DNA, or wherein the gRNAs of steps (a) and (b) hybridize to opposing strands of the DNA.
7. The method of claim 5 or 6, wherein the gRNAs of steps (a) and (b) hybridize to regions of the DNA that are no more than 10, no more than 15, no more than 20, no more than 25, no more than 30, no more than 40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart.

8. The method of any one of claims 5-7, wherein the DNA is in a cell.
9. The method of claim 8, wherein the cell is a eukaryotic cell.
10. The method of claim 9, wherein the cell is in an individual.
11. The method of claim 10, wherein the individual is a human.
12. A complex comprising a dimer of claim 3 and a single extended gRNA wherein the inactive Cas9 domain of each fusion protein of the dimer binds the gRNA, such that one fusion protein binds a portion of the gRNA, and the other fusion protein binds another portion of the gRNA.
13. The complex of claim 12, wherein the gRNA is at least 50, at least 75, at least 100, at least 150, at least 200, at least 250, or at least 300 nucleotides in length.
14. The complex of claim 12 or 13, wherein the regions of the extended gRNA that hybridize to a target nucleic acid comprise 15-25, 19-21, or 20 nucleotides.
15. A method for site-specific DNA cleavage comprising contacting a DNA with a complex of any one of claims 12-14, wherein:
 - (a) the gRNA contains two portions that hybridize to two separate regions of the DNA;
 - (b) the complex binds the DNA as a result of the portions of the gRNA hybridizing to the two regions; and
 - (c) binding of the complex results in the dimerization of the nuclease domains of each fusion protein, such that the domains cleave the DNA in a region between the bound fusion proteins.
16. The method of claim 15, wherein the two portions of the gRNA hybridize to the same strand of the DNA, or wherein the two portions of the gRNA hybridize to opposing strands of the DNA.

17. The method of claim 15 or 16, wherein the two portions of the gRNA hybridize to regions of the DNA that are no more than 10, no more than 15, no more than 20, no more than 25, no more than 30, no more than 40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart.
18. The method of any one of claims 15-17, wherein the DNA is in a cell.
19. The method of claim 18, wherein the cell is a eukaryotic cell.
20. The method of claim 19, wherein the cell is in an individual.
21. The method of claim 20, wherein the individual is a human.
22. A polynucleotide encoding a fusion protein of claims 1 or 2.
23. A vector comprising a polynucleotide of claim 22.
24. A vector for recombinant protein expression comprising a polynucleotide encoding a fusion protein of claim 1 or 2.
25. A cell comprising a genetic construct for expressing a fusion protein of claim 1 or 2.
26. A kit comprising a fusion protein of any of claims 1 or 2.
27. A kit comprising a polynucleotide encoding a fusion protein of claim 1 or 2.
28. A kit comprising a vector for recombinant protein expression, wherein the vector comprises a polynucleotide encoding a fusion protein of claim 1 or 2.
29. A kit comprising a cell that comprises a genetic construct for expressing a fusion protein of any of claims 1 or 2.
30. The kit of any one of claims 26-29, further comprising one or more gRNAs and/or vectors for expressing one or more gRNAs.

31. A protein comprising a gRNA binding domain of Cas9, wherein the protein does not comprise a DNA cleavage domain.
32. A protein comprising a DNA cleavage domain of Cas9, wherein the protein does not comprise a gRNA binding domain.
33. A fusion protein comprising two domains: (i) a nuclease-inactivated Cas9, and (ii) a gRNA binding domain of Cas9, wherein domain (ii) does not include a DNA cleavage domain.
34. A fusion protein comprising two domains: (i) a nuclease-inactivated Cas9, and (ii) a DNA cleavage domain, wherein domain (ii) does not include a gRNA binding domain.
35. A protein dimer comprising the proteins of claim 31 and 32.
36. A protein dimer comprising the proteins of claim 31 and 34.
37. A protein dimer comprising the proteins of claim 32 and 33.
38. A protein dimer comprising the proteins of claim 33 and 34.
39. The protein dimer of any one of claims 35-38, further comprising one or more gRNA.
40. A method for site-specific DNA cleavage comprising contacting a DNA with a protein dimer of claim 35, wherein the dimer binds a gRNA that hybridizes to a region of the DNA, and cleavage of the DNA occurs.
41. A method for site-specific DNA cleavage comprising contacting a DNA with a protein dimer of claim 36 or 37, wherein the protein dimer binds two gRNAs that hybridize to two regions of the DNA, and cleavage of the DNA occurs.
42. The method of claim 41, wherein the two gRNAs hybridize to regions of the DNA that are no more than 10, no more than 15, no more than 20, no more than 25, no more than

30, no more than 40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart.

43. A method for site-specific DNA cleavage comprising contacting a DNA with a protein dimer of claim 38, wherein the protein dimer binds three gRNAs that hybridize to three regions of the DNA, and cleavage of the DNA occurs.

44. The method of claim 43, wherein the three RNA hybridize to regions of the DNA that are no more than 10, no more than 15, no more than 20, no more than 25, no more than 30, no more than 40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart, as between the first and second, and second and third regions.

45. The method of any one of claims 40-44, wherein the DNA is in a cell.

46. The method of claim 45, wherein the cell is a eukaryotic cell.

47. The method of claim 46, wherein the cell is in an individual.

48. The method of claim 47, wherein the individual is a human.

49. A polynucleotide encoding a protein of claim 31, a protein of claim 32, a fusion protein of claim 33, a fusion protein of claim 34, or combinations thereof.

50. A vector comprising a polynucleotide of claim 49.

51. A vector for recombinant protein expression comprising a polynucleotide encoding a protein of claim 31, a protein of claim 32, a fusion protein of claim 33, a fusion protein of claim 34, or combinations thereof.

52. A cell comprising a genetic construct for expressing a protein of claim 31, a protein of claim 32, a fusion protein of claim 33, a fusion protein of claim 34, or combinations thereof.

53. A kit comprising a protein of claim 31, a protein of claim 32, a fusion protein of claim 33, a fusion protein of claim 34, or combinations thereof.
54. A kit comprising a polynucleotide encoding a protein of claim 31, a protein of claim 32, a fusion protein of claim 33, a fusion protein of claim 34, or combinations thereof.
55. A kit comprising a vector for recombinant protein expression, the vector comprising a polynucleotide encoding a protein of claim 31, a protein of claim 32, a fusion protein of claim 33, a fusion protein of claim 34, or combinations thereof.
56. A kit comprising a cell that comprises a genetic construct for expressing a protein of claim 31, a protein of claim 32, a fusion protein of claim 33, a fusion protein of claim 34, or combinations thereof.
57. The kit of any one of claims 53-56, further comprising one or more gRNAs and/or vectors for expressing one or more gRNAs.
58. A minimized Cas9 protein, wherein the protein comprises N- and/or C-terminal truncations and retains RNA binding and DNA cleavage activity.
59. The protein of claim 58, wherein the N-terminal truncation removes at least 5, at least 10, at least 15, at least 20, at least 25, at least 40, at least 40, at least 50, at least 75, at least 100, or at least 150 amino acids.
60. The protein of claim 58, wherein the C-terminal truncation removes at least 5, at least 10, at least 15, at least 20, at least 25, at least 40, at least 40, at least 50, at least 75, at least 100, or at least 150 amino acids.
61. The protein of any one of claims 58-60, further comprising a gRNA.
62. A method for site-specific DNA cleavage comprising contacting a DNA with the protein:gRNA complex of claim 61.
63. A polynucleotide encoding a protein of any one of claims 58-60.

64. A vector comprising a polynucleotide of claim 63.
65. A vector for recombinant protein expression comprising a polynucleotide encoding a protein of any one of claims 58-60.
66. A cell comprising a genetic construct for expressing a protein of any one of claims 58-60.
67. A kit comprising a protein of any one of claims 58-60.
68. A kit comprising a polynucleotide encoding a protein of any one of claims 58-60.
69. A kit comprising a vector for recombinant protein expression, the vector comprising a polynucleotide encoding a protein of any one of claims 58-60.
70. A kit comprising a cell that comprises a genetic construct for expressing a protein of any one of claims 58-60.
71. The kit of any one of claims 67-70, further comprising one or more gRNAs and/or vectors for expressing one or more gRNAs.
72. A dimer of Cas9 or fragments thereof, wherein the dimer is coordinated through a single gRNA.
73. The dimer of claim 72, wherein the single gRNA comprises at least two portions that (i) are each able to bind a Cas9 protein and (ii) each hybridize to a target nucleic acid.
74. The dimer of claim 73, wherein the portions of the gRNA that hybridize to the target nucleic acid each comprise no more than 5, no more than 10, or no more than 15 nucleotides complementary to the target nucleic acid.
75. The dimer of claim 74 or 74, wherein the portions of the gRNA are separated by a linker sequence.

76. The dimer of claim 75, wherein the linker sequence hybridizes to the target nucleic acid.
77. A method for site-specific DNA cleavage comprising contacting a DNA with a dimer of any one of claims 72-76.
78. A polynucleotide encoding a monomer of the dimer and/or the gRNA of any one of claims 72-76.
79. A vector comprising a polynucleotide of claim 78.
80. A vector for recombinant protein expression comprising a polynucleotide encoding a monomer of the dimer and/or the gRNA of any one of claims 72-76.
81. A cell comprising a genetic construct for expressing a monomer of the dimer and/or the gRNA of any one of claims 72-76.
82. A kit comprising the dimer of claim and/or gRNA of any one of claims 72-76.
83. A kit comprising a polynucleotide encoding a monomer of the dimer and/or the gRNA of any one of claims 72-76.
84. A kit comprising a vector for recombinant protein expression, the vector comprising a polynucleotide encoding a monomer of the dimer and/or the gRNA of any one of claims 72-76.
85. A kit comprising a cell that comprises a genetic construct for expressing a monomer of the dimer and/or the gRNA of any one of claims 72-76.
86. A fusion protein comprising two domains: (i) a nuclease-inactivated Cas9; and (ii) a recombinase catalytic domain.

87. The fusion protein of claim 86, wherein the recombinase catalytic domain is a monomer of the recombinase catalytic domain of Hin recombinase, Gin recombinase, or Tn3 resolvase.

88. The fusion protein of claim 86 or claim 87, wherein the nuclease-inactivated Cas9 domain binds a gRNA.

89. A dimer of the fusion protein of claim 88, wherein the dimer is bound to a nucleic acid, optionally wherein each fusion protein of the dimer is bound to opposite strands of the nucleic acid.

90. A tetramer of the fusion protein of claim 88, wherein the tetramer is bound to one or more nucleic acid(s).

91. A method for site-specific recombination between two DNA molecules, comprising:

(a) contacting a first DNA with a first fusion protein of claim 86, wherein the nuclease-inactivated Cas9 domain binds a first gRNA that hybridizes to a region of the first DNA;

(b) contacting the first DNA with a second fusion protein of claim 86, wherein the nuclease-inactivated Cas9 domain of the second fusion protein binds a second gRNA that hybridizes to a second region of the first DNA;

(c) contacting a second DNA with a third fusion protein of claim 86, wherein the nuclease-inactivated Cas9 domain of the third fusion protein binds a third gRNA that hybridizes to a region of the second DNA; and

(d) contacting the second DNA with a fourth fusion protein of claim 86, wherein the nuclease-inactivated Cas9 domain of the fourth fusion protein binds a fourth gRNA that hybridizes to a second region of the second DNA;

whereby the binding of the fusion proteins in steps (a) - (d) results in the tetramerization of the recombinase catalytic domains of the fusion proteins, under conditions such that the DNAs are recombined.

92. The method of claim 91, wherein the gRNAs of steps (a) and (b) hybridize to opposing strands of the first DNA, and the gRNAs of steps (c) and (d) hybridize to opposing strands of the second DNA.

93. The method of claim 91 or 92, wherein the gRNAs of steps (a) and (b); and/or the gRNAs of steps (c) and (d) hybridize to regions of their respective DNAs that are no more than 10, no more than 15, no more than 20, no more than 25, no more than 30, no more than 40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart.

94. A method for site-specific recombination between two regions of a single DNA molecule, comprising:

(a) contacting the DNA with a first fusion protein of claim 86, wherein the nuclease-inactivated Cas9 domain binds a first gRNA that hybridizes to a region of the DNA;

(b) contacting the DNA with a second fusion protein of claim 86, wherein the nuclease-inactivated Cas9 domain of the second fusion protein binds a second gRNA that hybridizes to a second region of the DNA;

(c) contacting the DNA with a third fusion protein of claim 86, wherein the nuclease-inactivated Cas9 domain of the third fusion protein binds a third gRNA that hybridizes to a third region of the DNA;

(d) contacting the DNA with a fourth fusion protein of claim 86, wherein the nuclease-inactivated Cas9 domain of the fourth fusion protein binds a fourth gRNA that hybridizes to a fourth region of the DNA;

whereby the binding of the fusion proteins in steps (a) - (d) results in the tetramerization of the recombinase catalytic domains of the fusion proteins, under conditions such that the DNA is recombined.

95. The method of claim 94, wherein two of the gRNAs of steps (a) - (d) hybridize to the same strand of the DNA, and the other two gRNAs of steps (a)- (d) hybridize to the opposing strand of the DNA.

96. The method of claim 94 or 95, wherein the gRNAs of steps (a) and (b) hybridize to regions of the DNA that are no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart, and the gRNAs of steps (c) and (d) hybridize to regions of the DNA that are no more than 10, no more than 15, no more than 20, no more than 25, no more than 30, no more than 40, no more than 50, no more than 60, no more than 70, no more than 80, no more than 90, or no more than 100 base pairs apart.

97. The method of any one of claims 91-96, wherein the DNA is in a cell.
98. The method of claim 97, wherein the cell is a eukaryotic cell or a prokaryotic cell.
99. The method of claim 98, wherein the cell is a eukaryotic cell in an individual.
100. The method of claim 99, wherein the individual is a human.
101. A polynucleotide encoding a fusion protein of claim 86 or 87.
102. A vector comprising a polynucleotide of claim 101.
103. A vector for recombinant protein expression comprising a polynucleotide encoding a fusion protein of claim 86 or 87.
104. A cell comprising a genetic construct for expressing a fusion protein of claims 86 or 87.
105. A kit comprising a fusion protein of any of claim 86 or 87.
106. A kit comprising a polynucleotide encoding a fusion protein of claim 86 or 87.
107. A kit comprising a vector for recombinant protein expression, wherein the vector comprises a polynucleotide encoding a fusion protein of claim 86 or 87.
108. A kit comprising a cell that comprises a genetic construct for expressing a fusion protein of any of claims 86 or 87.
109. The kit of any one of claims 105-108, further comprising one or more gRNAs and/or vectors for expressing one or more gRNAs.
110. The fusion protein of claim 1, further comprising a nuclear localization signal (NLS) domain.

111. The fusion protein of claim 110, wherein one or more domains are separated by a peptide linker or a non-peptide linker.

112. The fusion protein of claim 111, wherein the peptide linker comprises an XTEN linker, an amino acid sequence comprising one or more repeats of the tri-peptide GGS, or any sequence as provided in Figure 12A.

113. The fusion protein of claim 1, wherein the fusion protein is encoded by a nucleotide sequence set forth as SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, or SEQ ID NO:12, or a variant or fragment of any one of SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, or SEQ ID NO:12.

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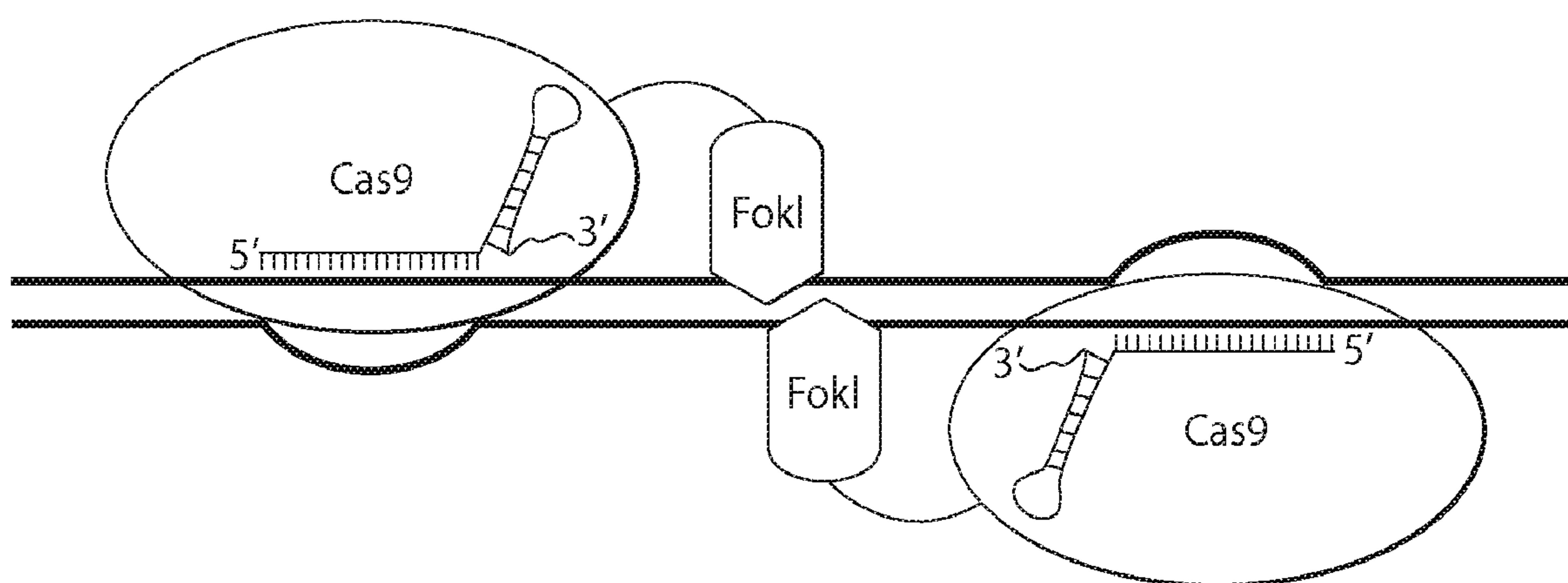


Fig. 1A

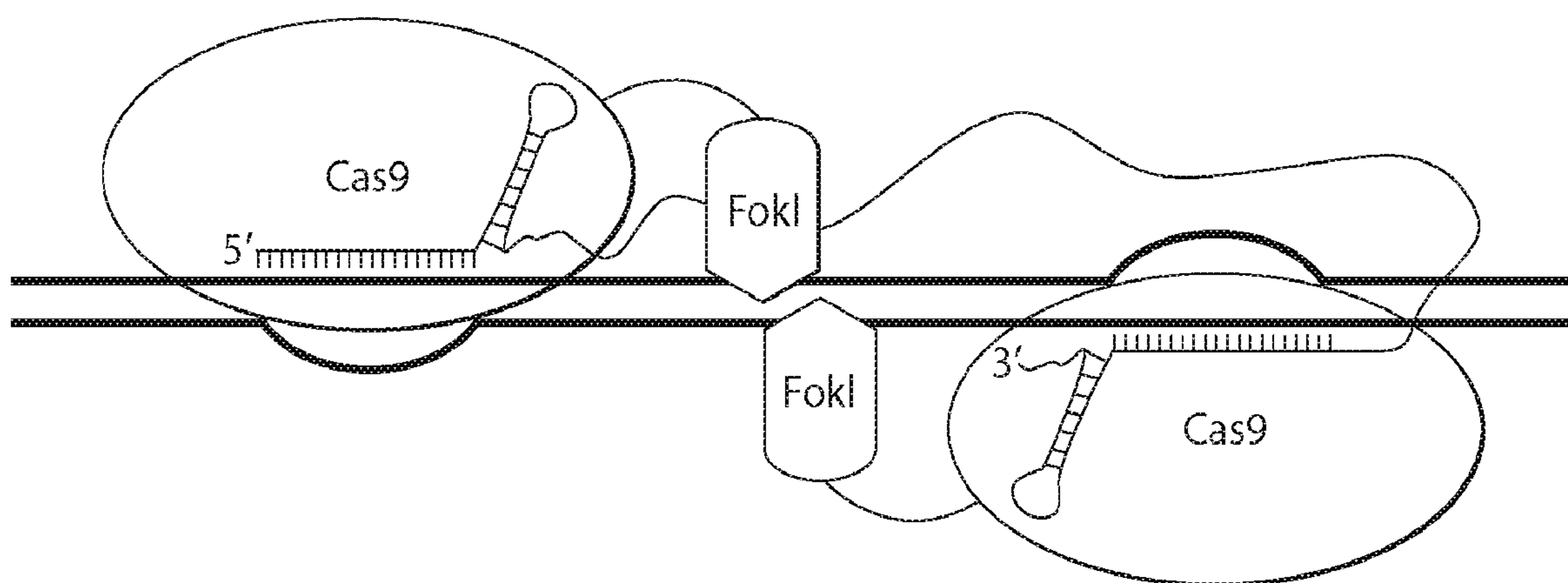


Fig. 1B

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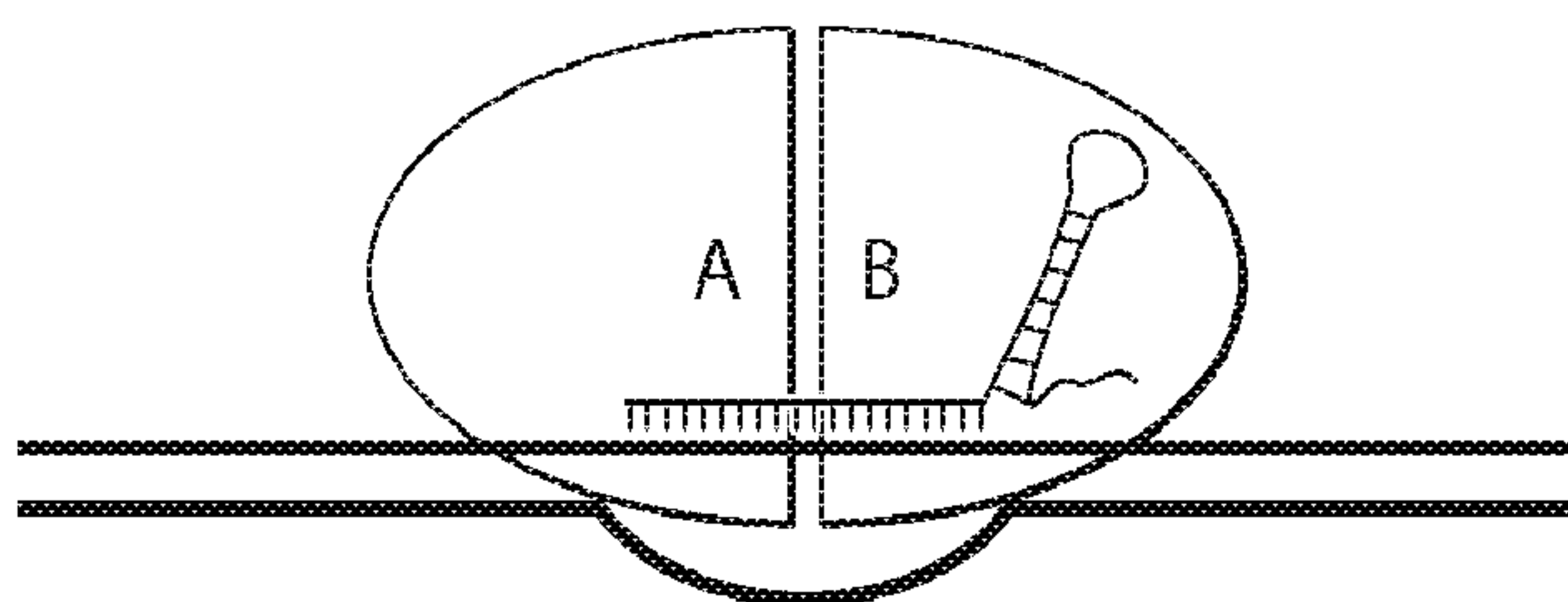


Fig. 2A

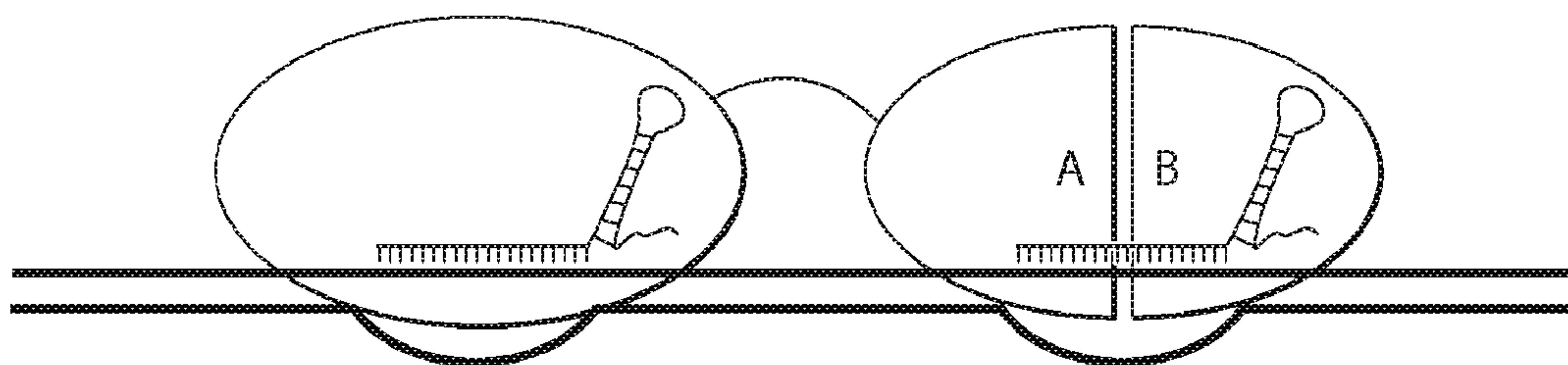


Fig. 2B

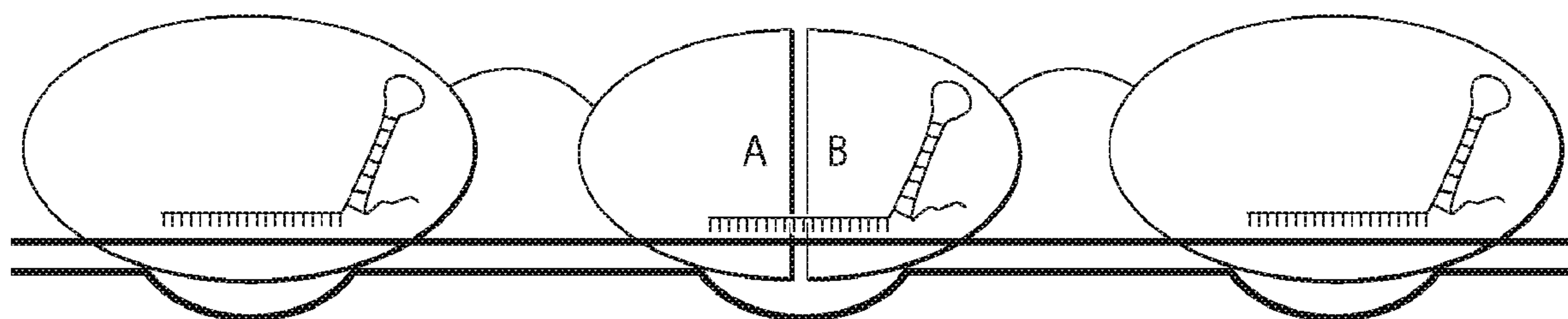


Fig. 2C

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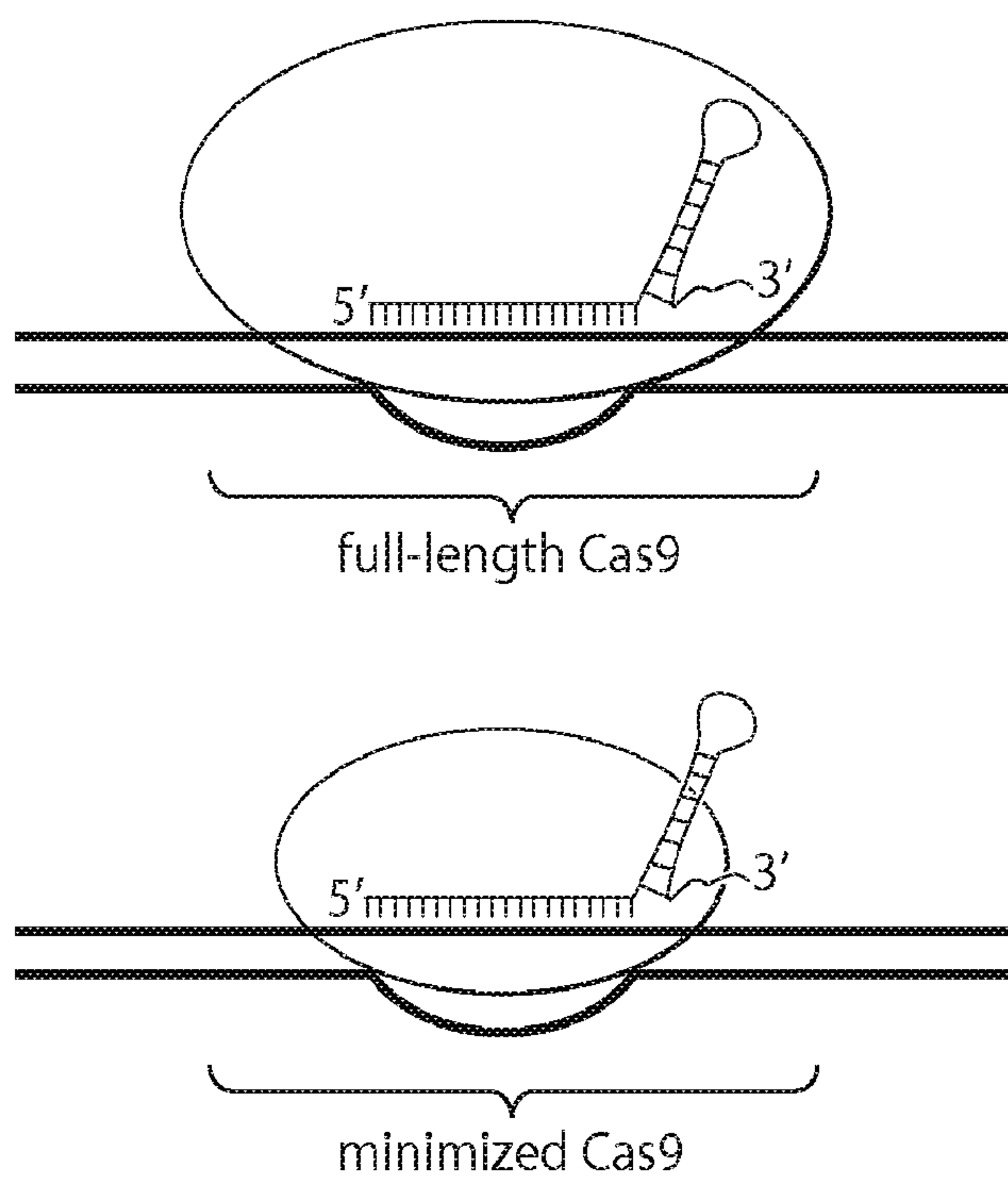


Fig. 3

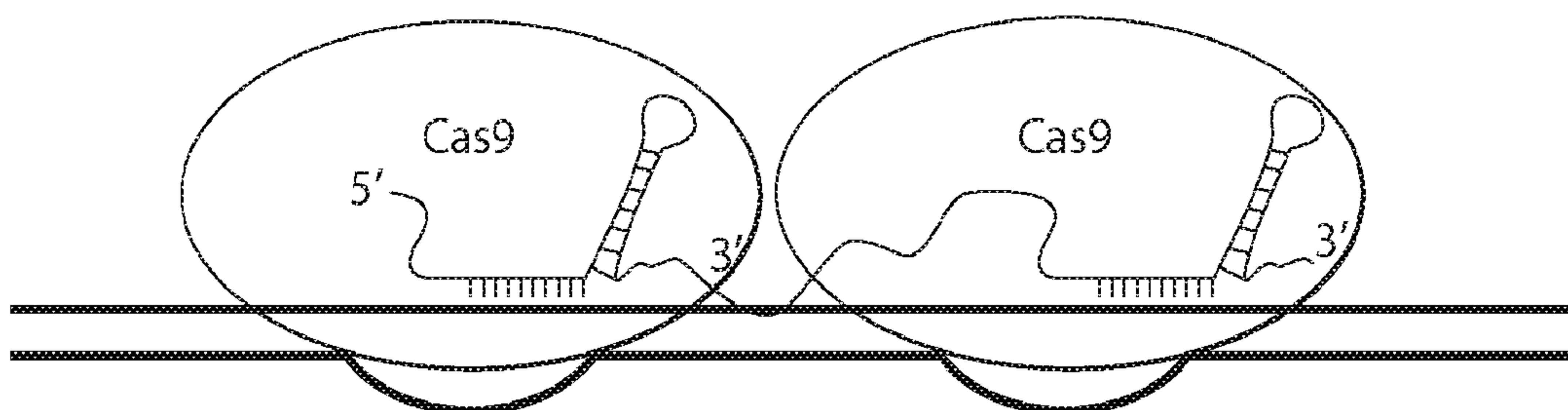


Fig. 4

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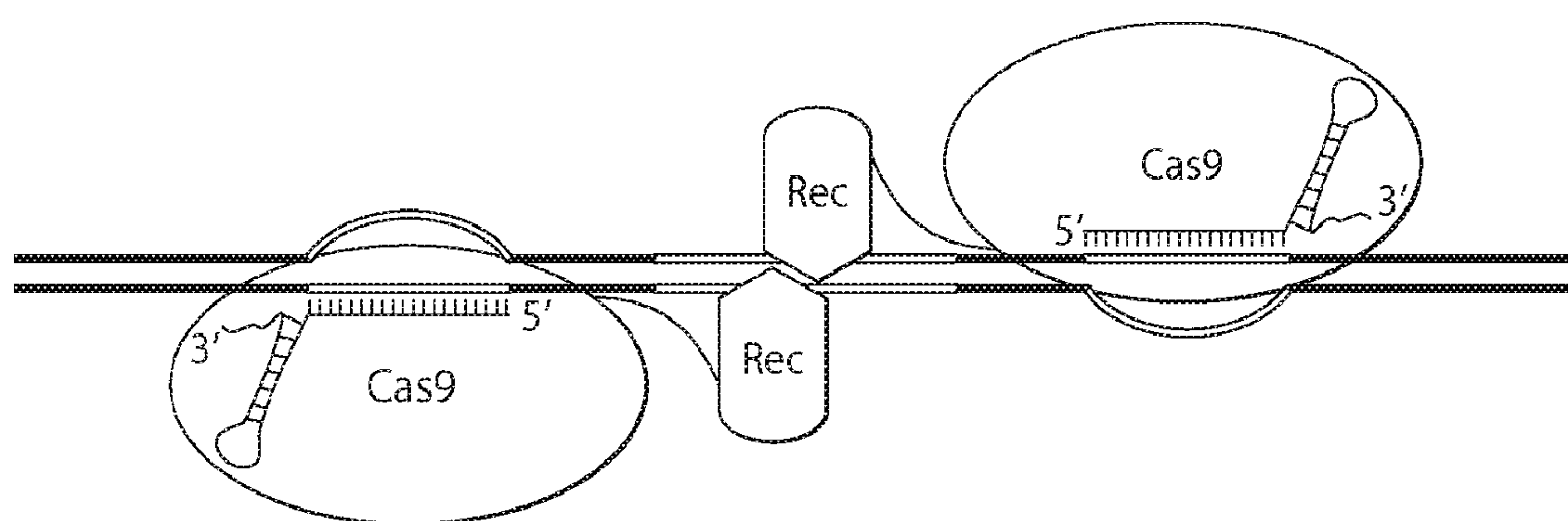


Fig. 5A

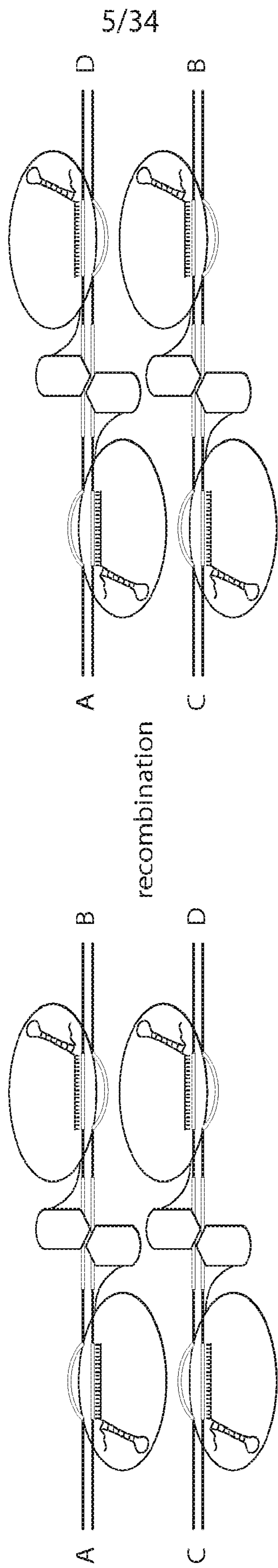


Fig. 5B

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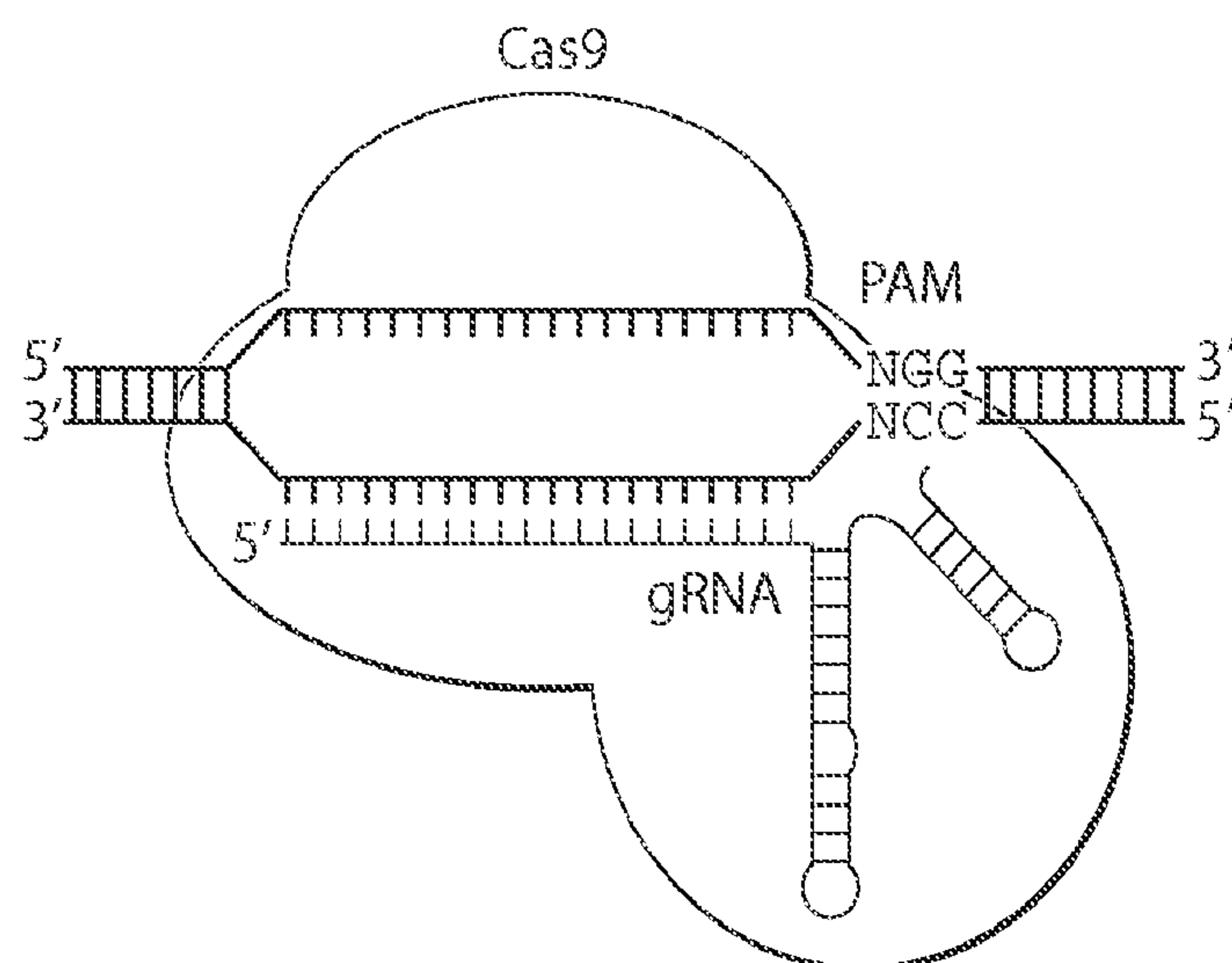


Fig. 6A

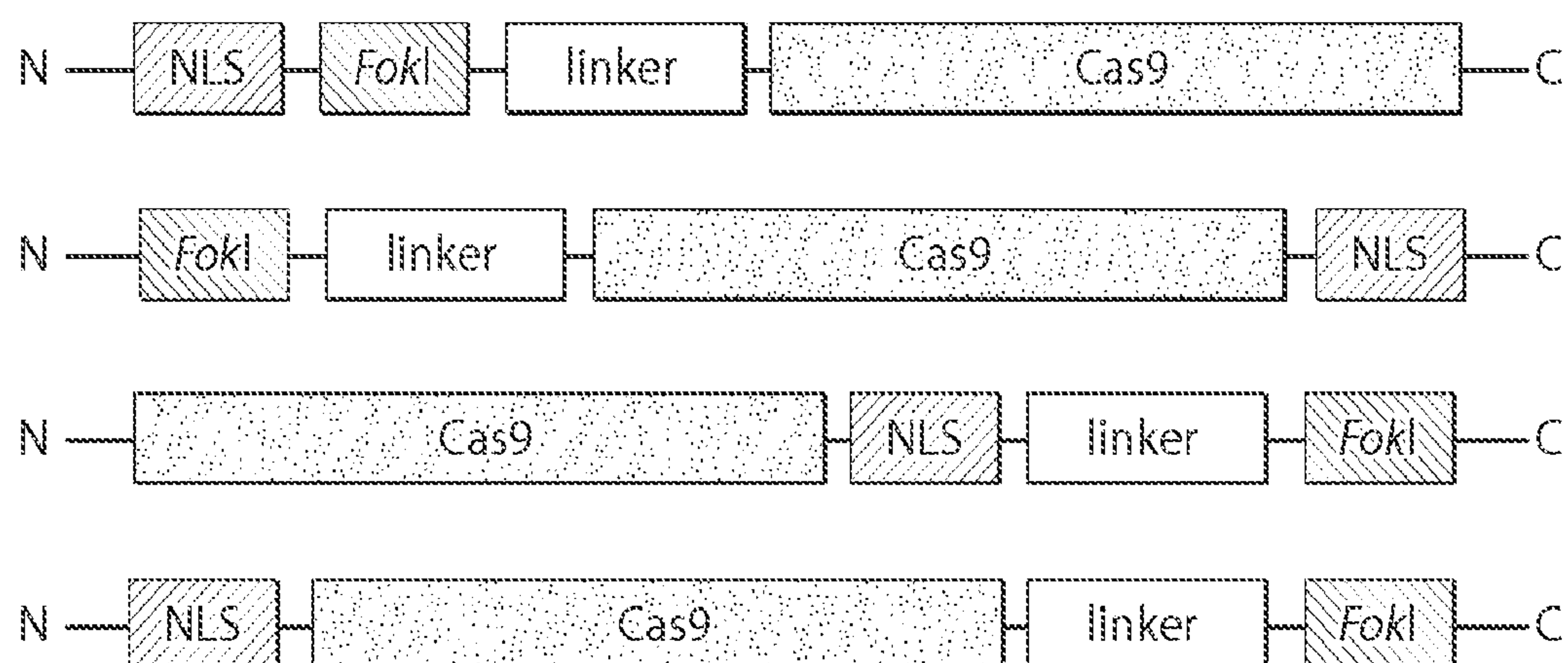


Fig. 6B

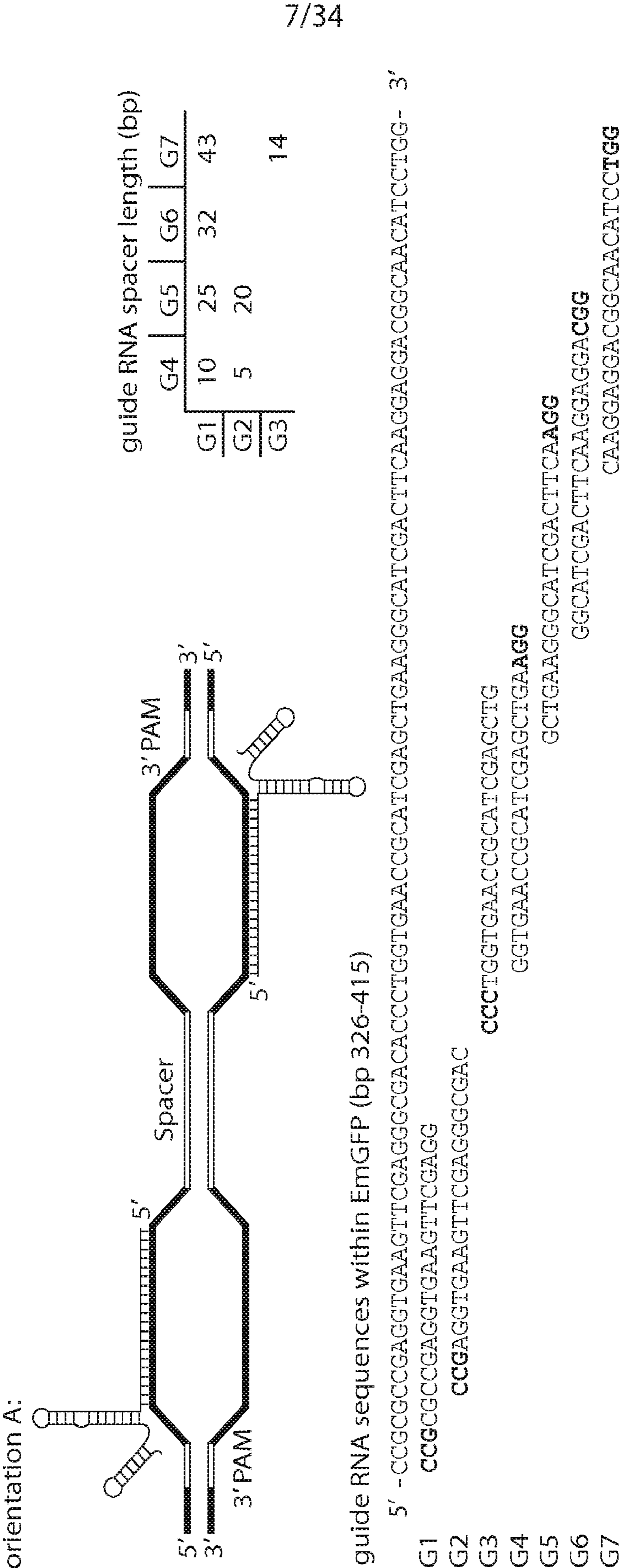


Fig. 6C

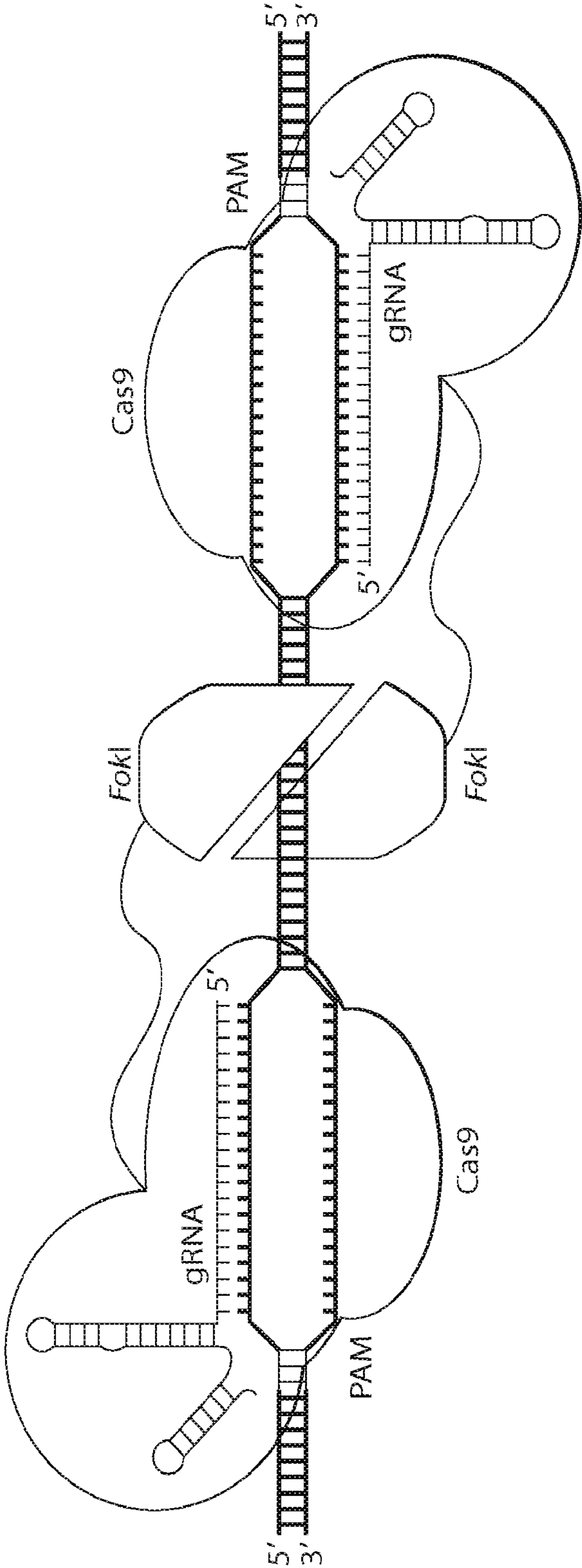


Fig. 6D

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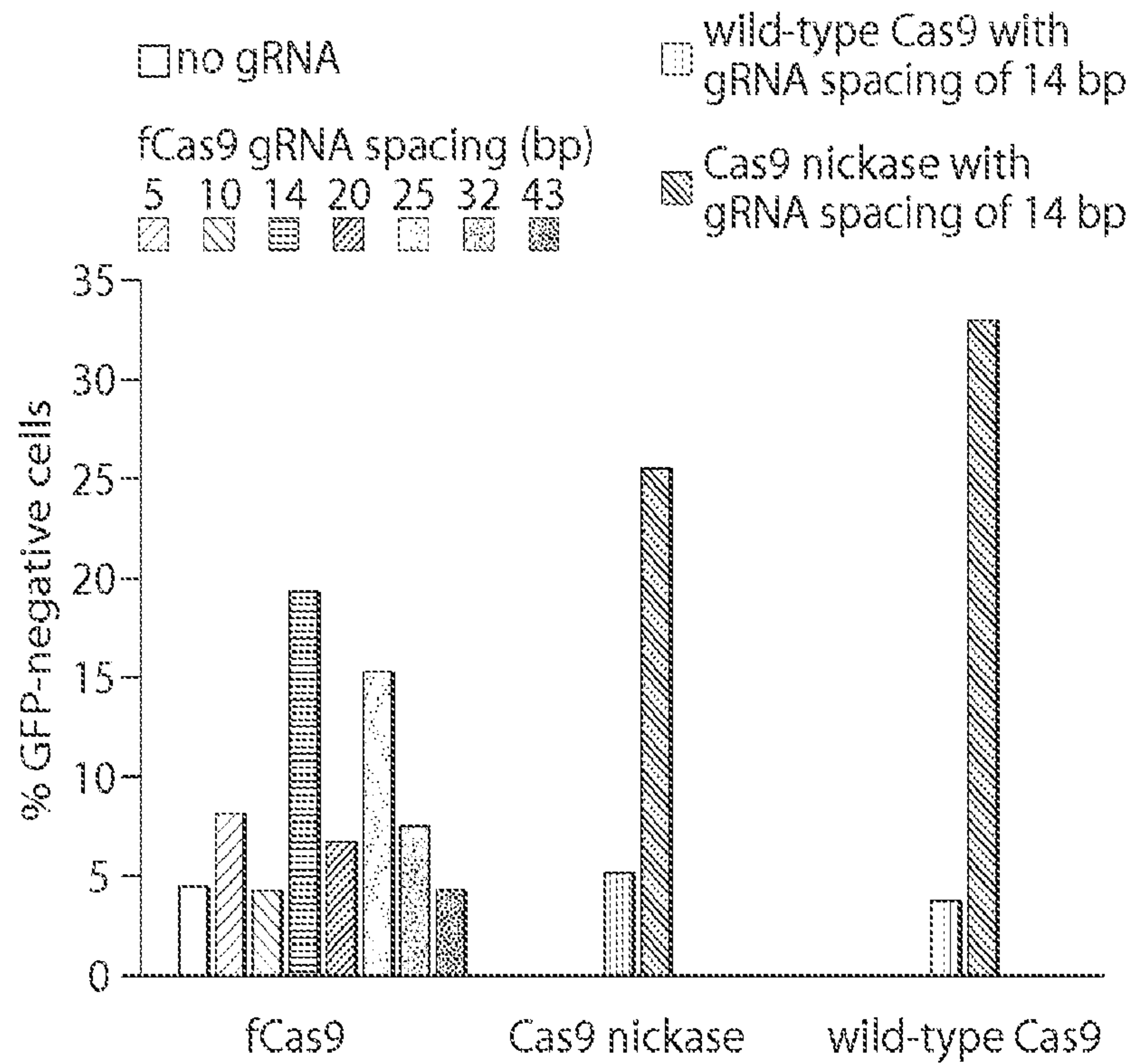


Fig. 7A

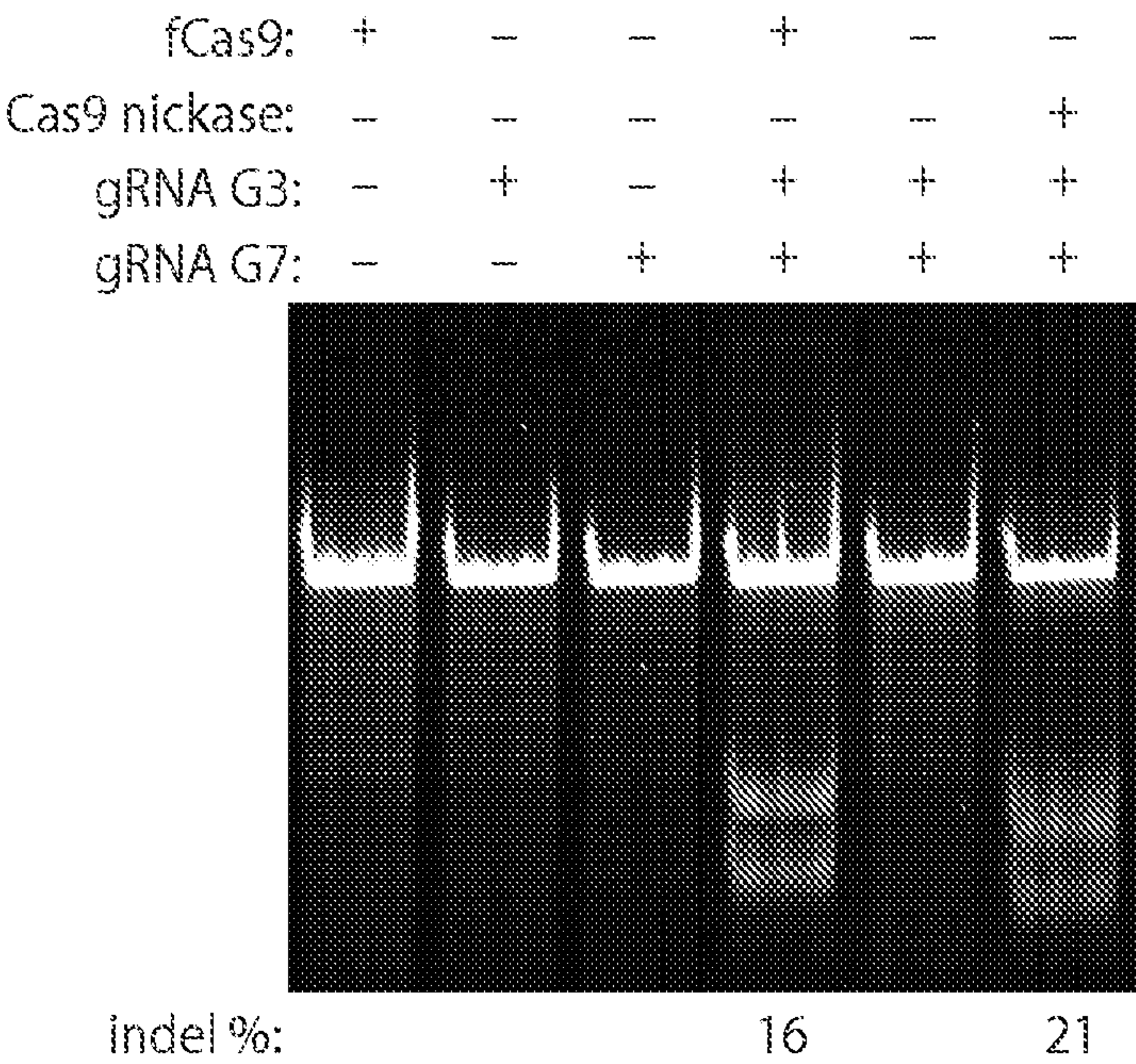


Fig. 7B

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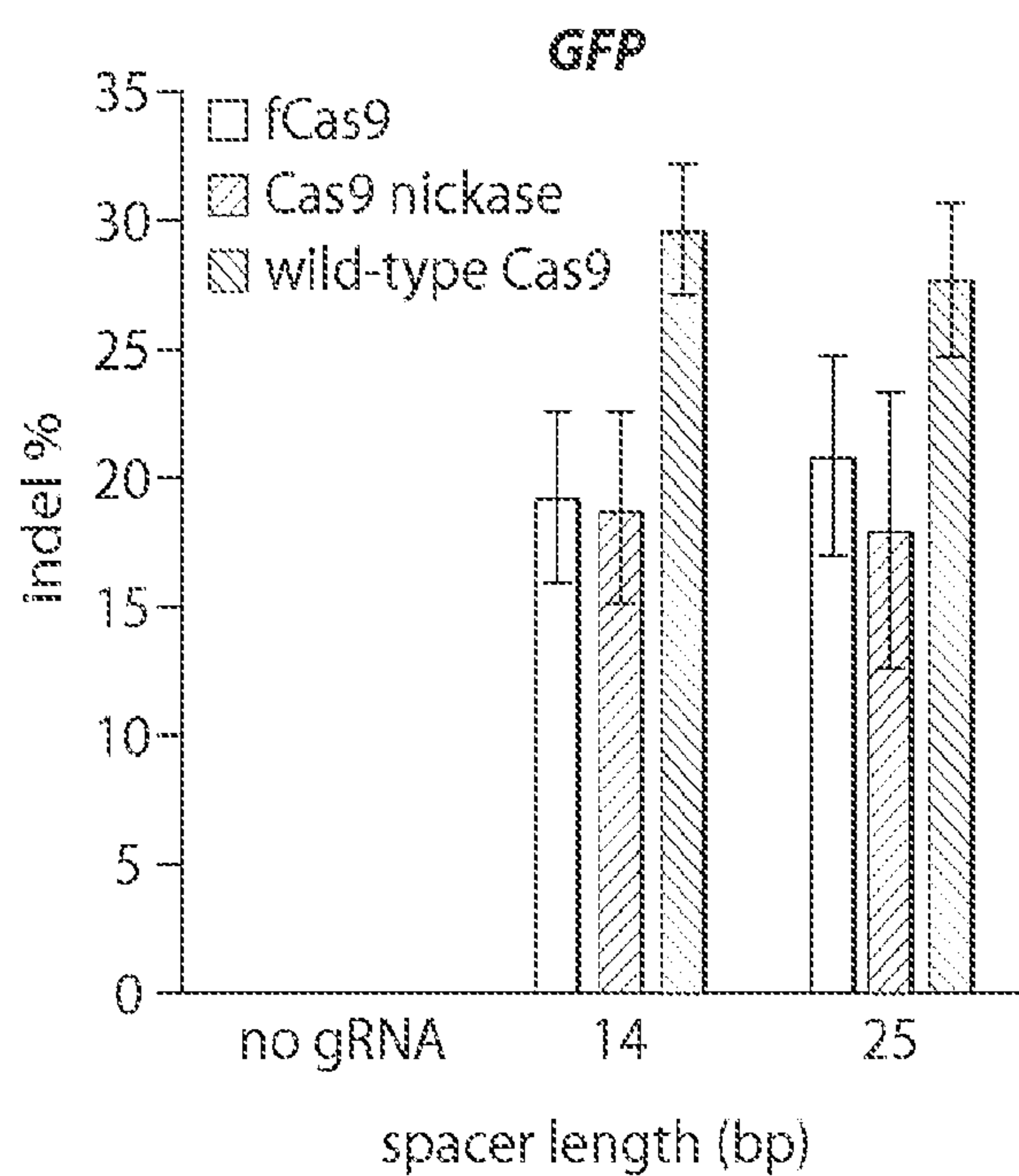


Fig. 7C

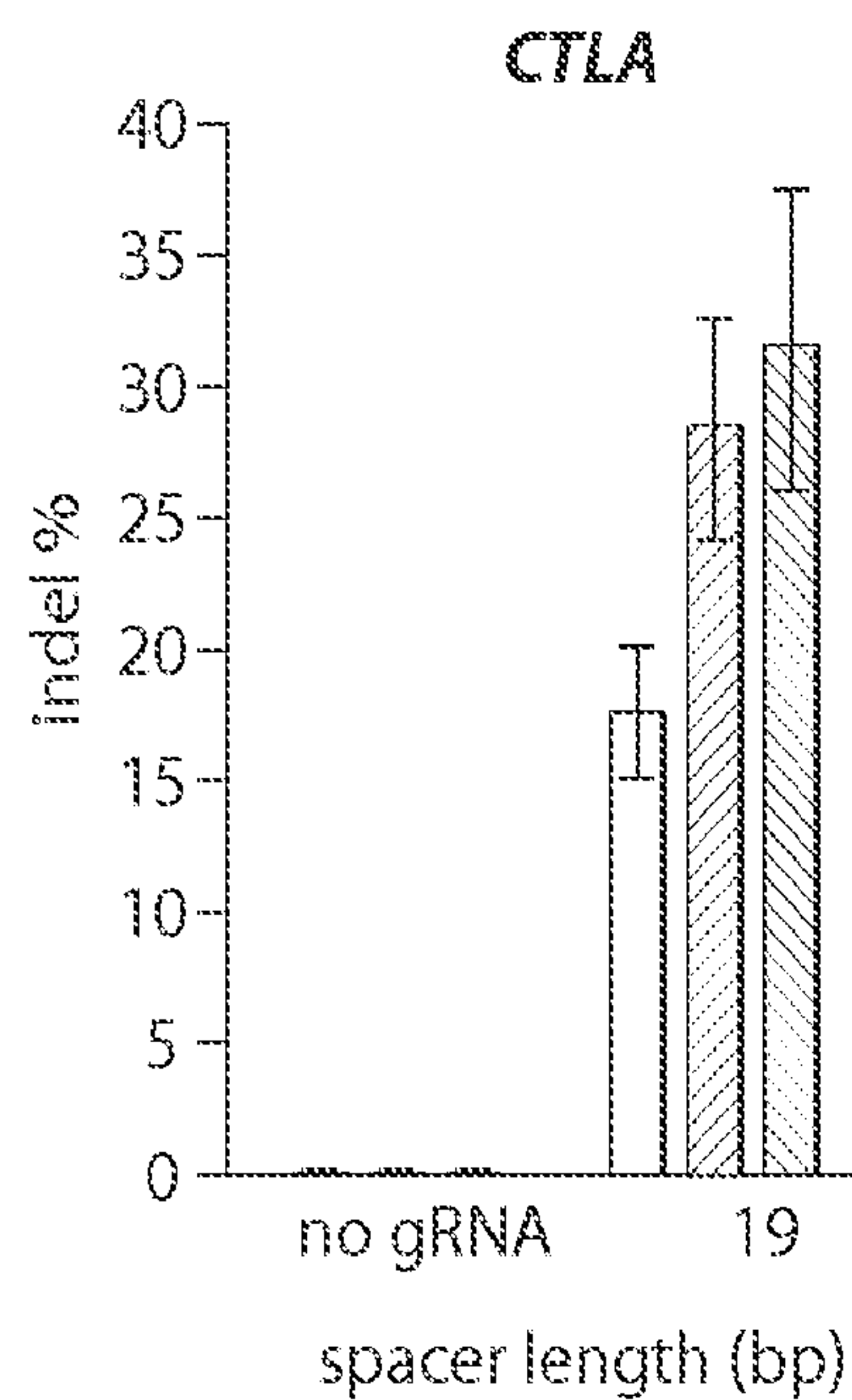


Fig. 7D

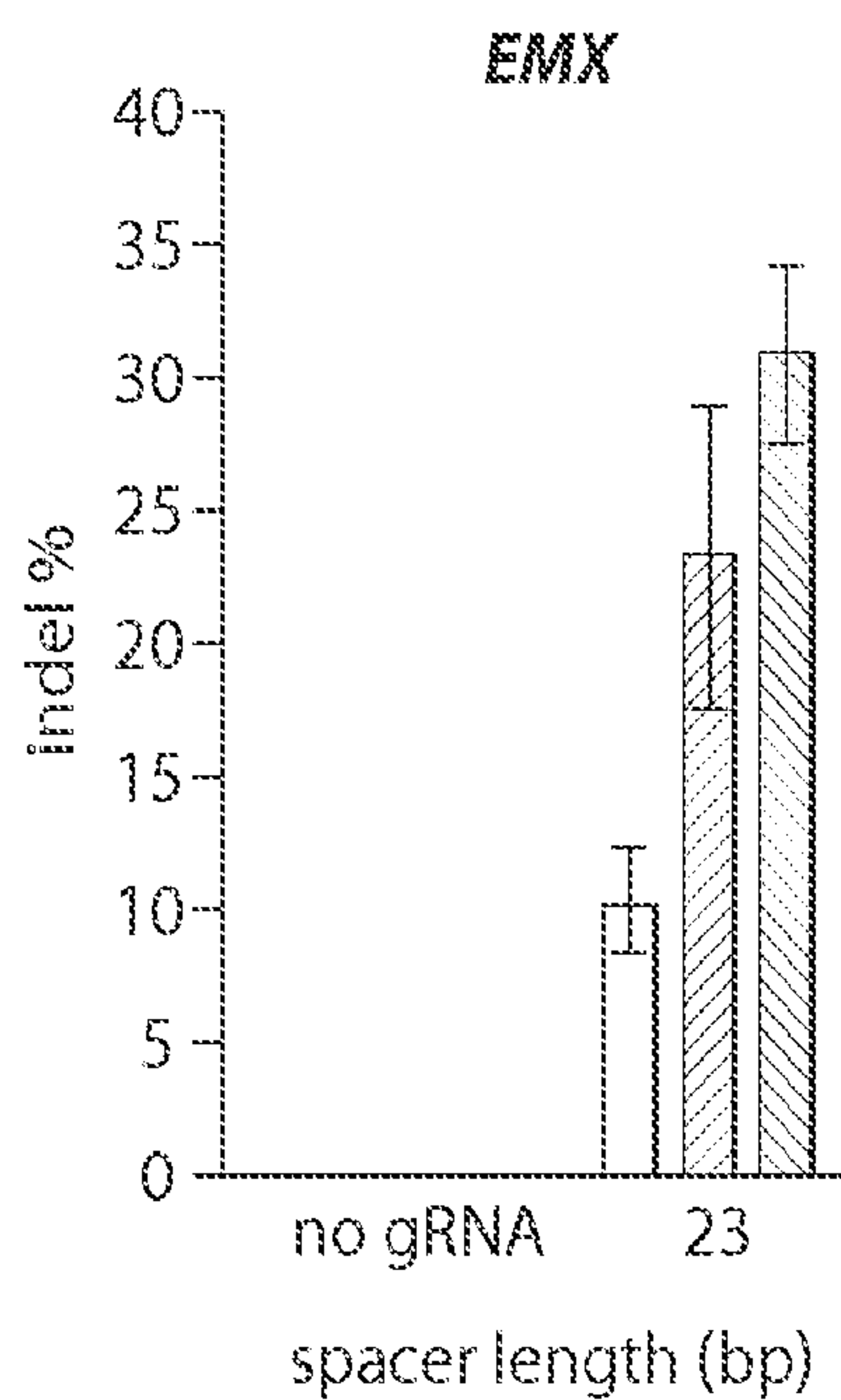


Fig. 7E

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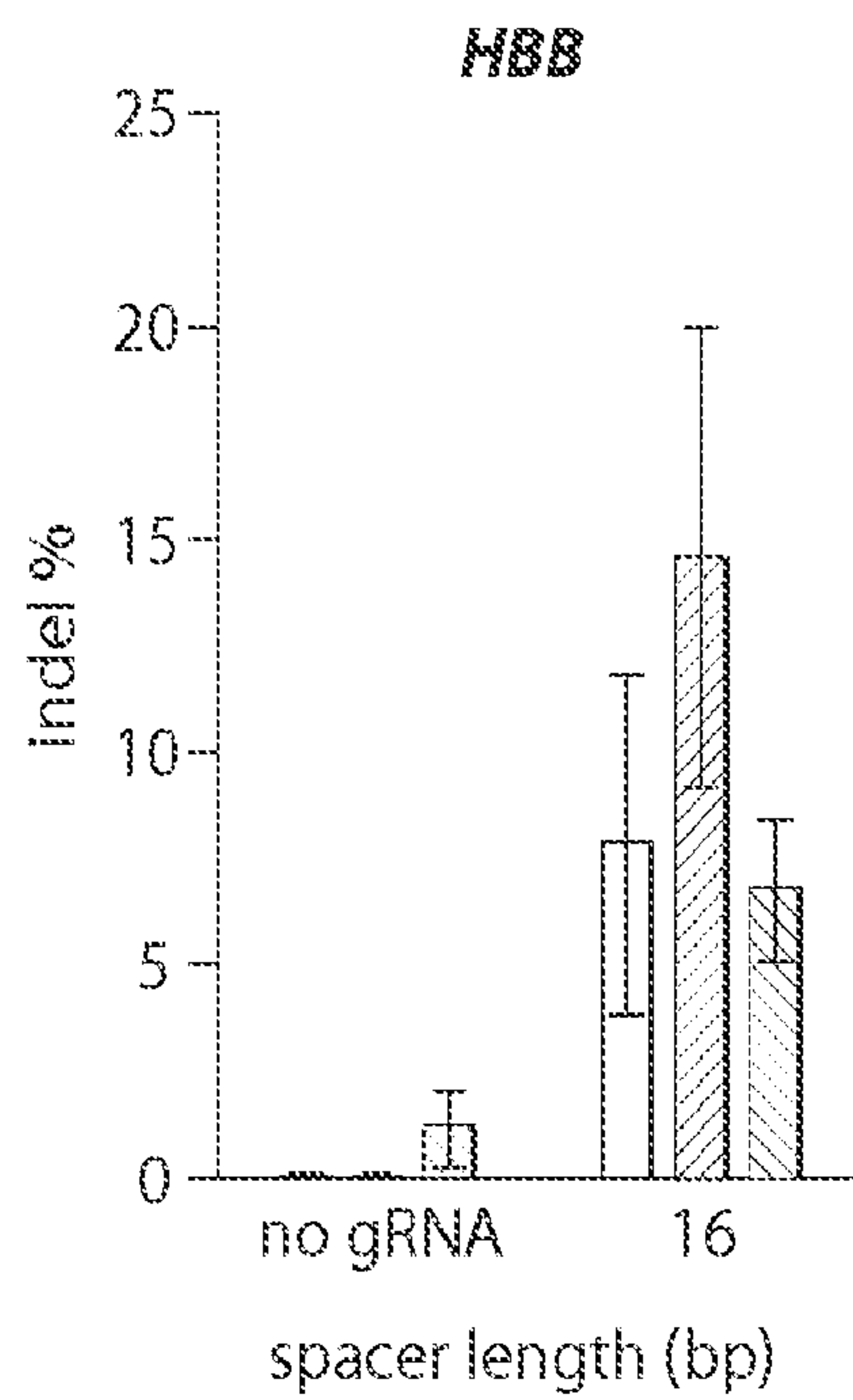


Fig. 7F

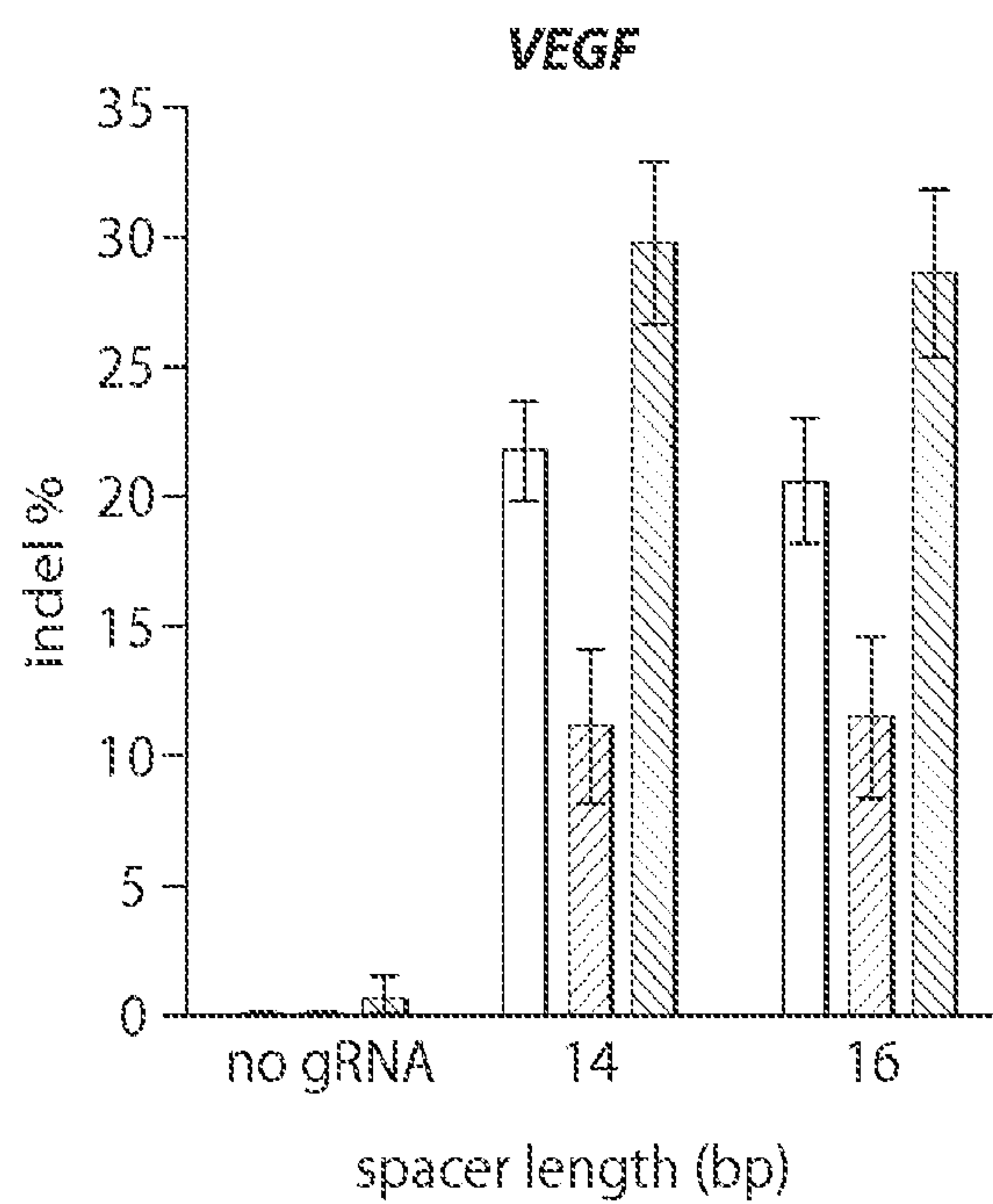


Fig. 7G

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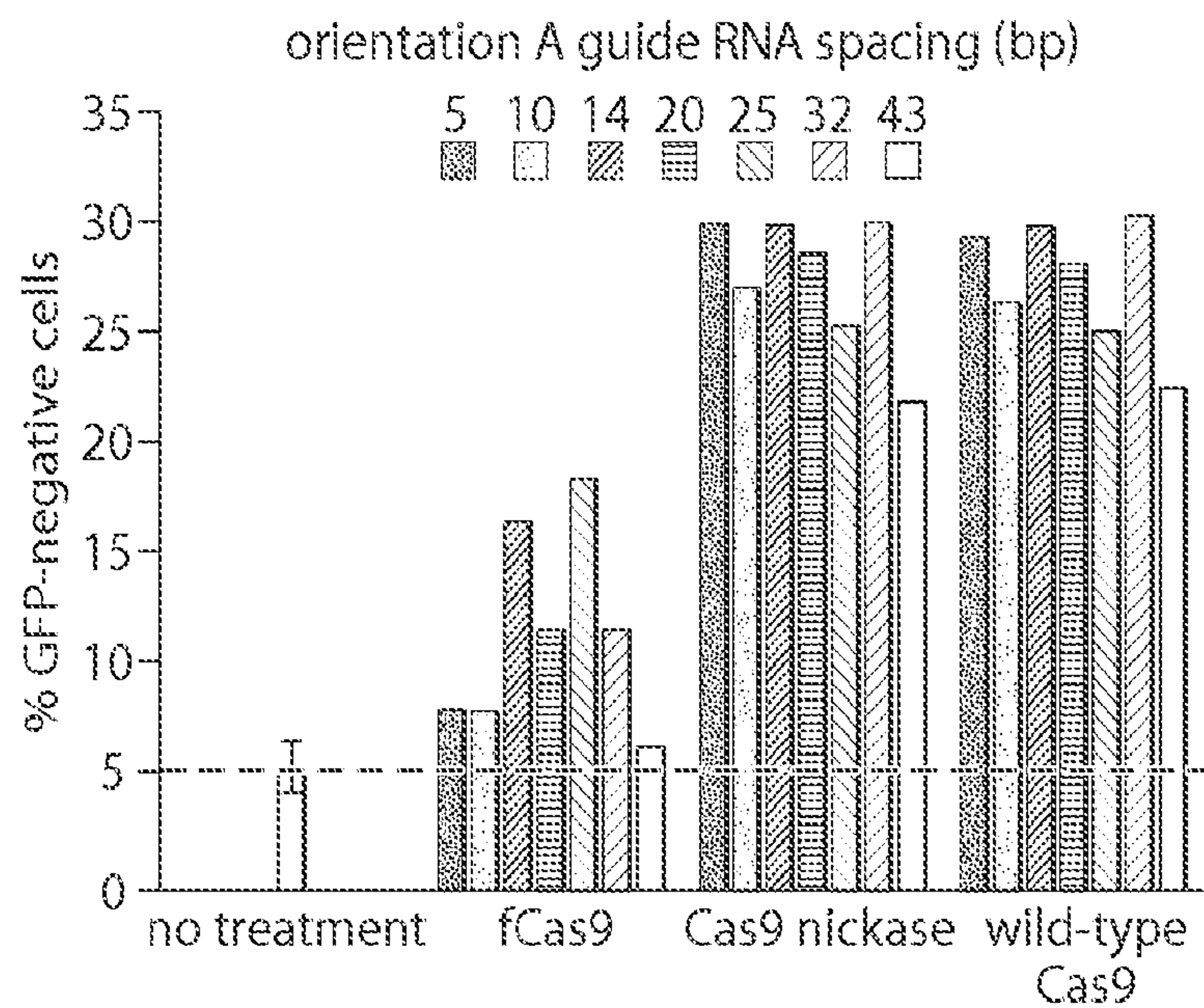


Fig. 8A

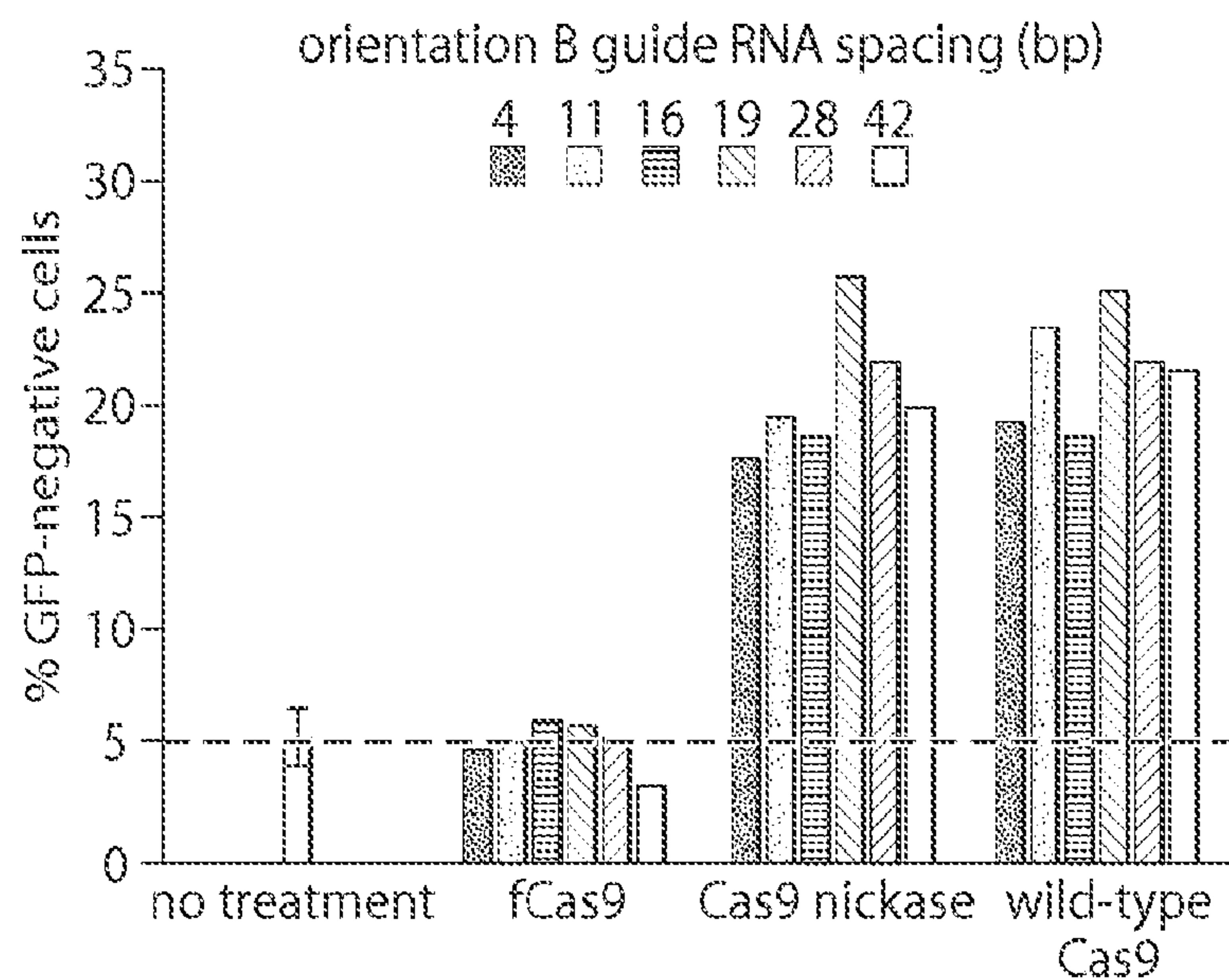


Fig. 8B

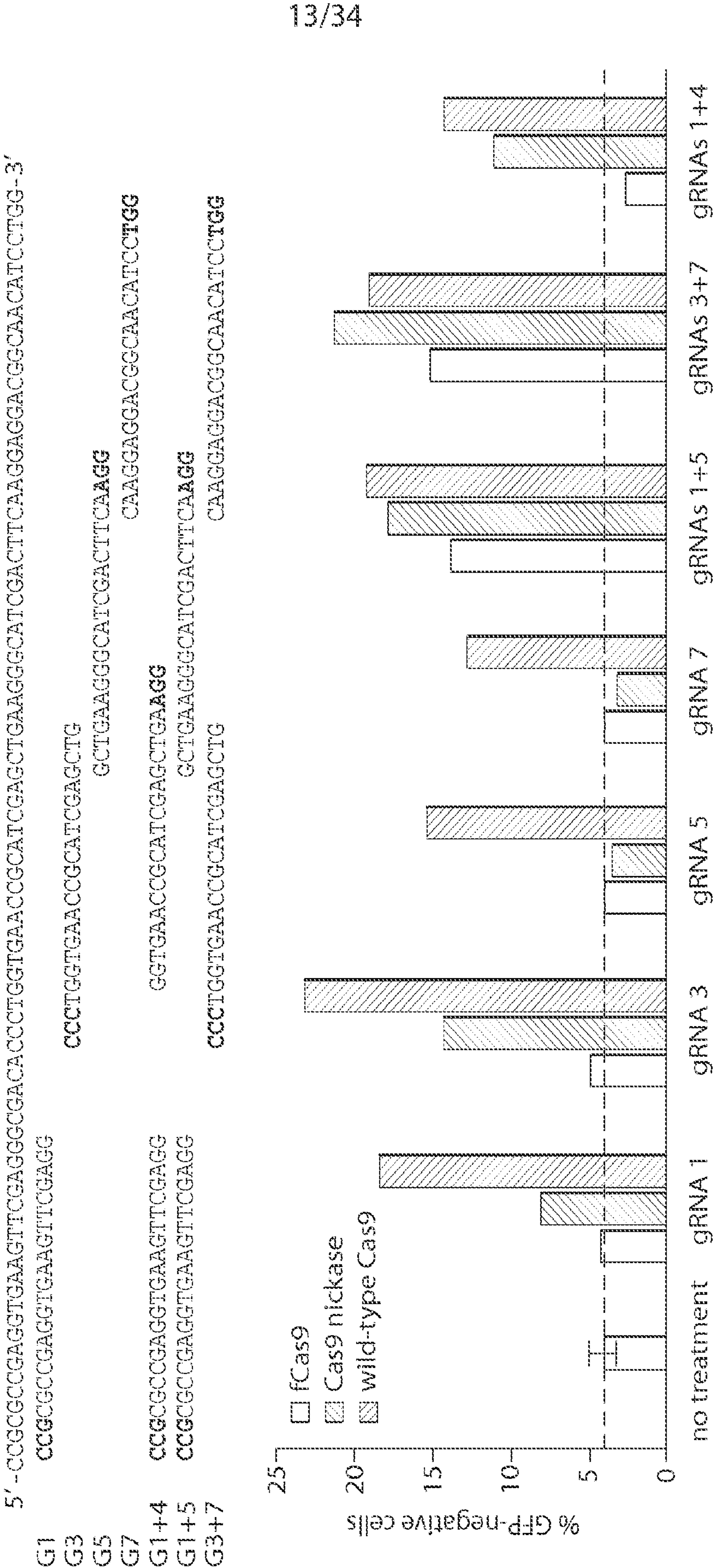


Fig. 8C

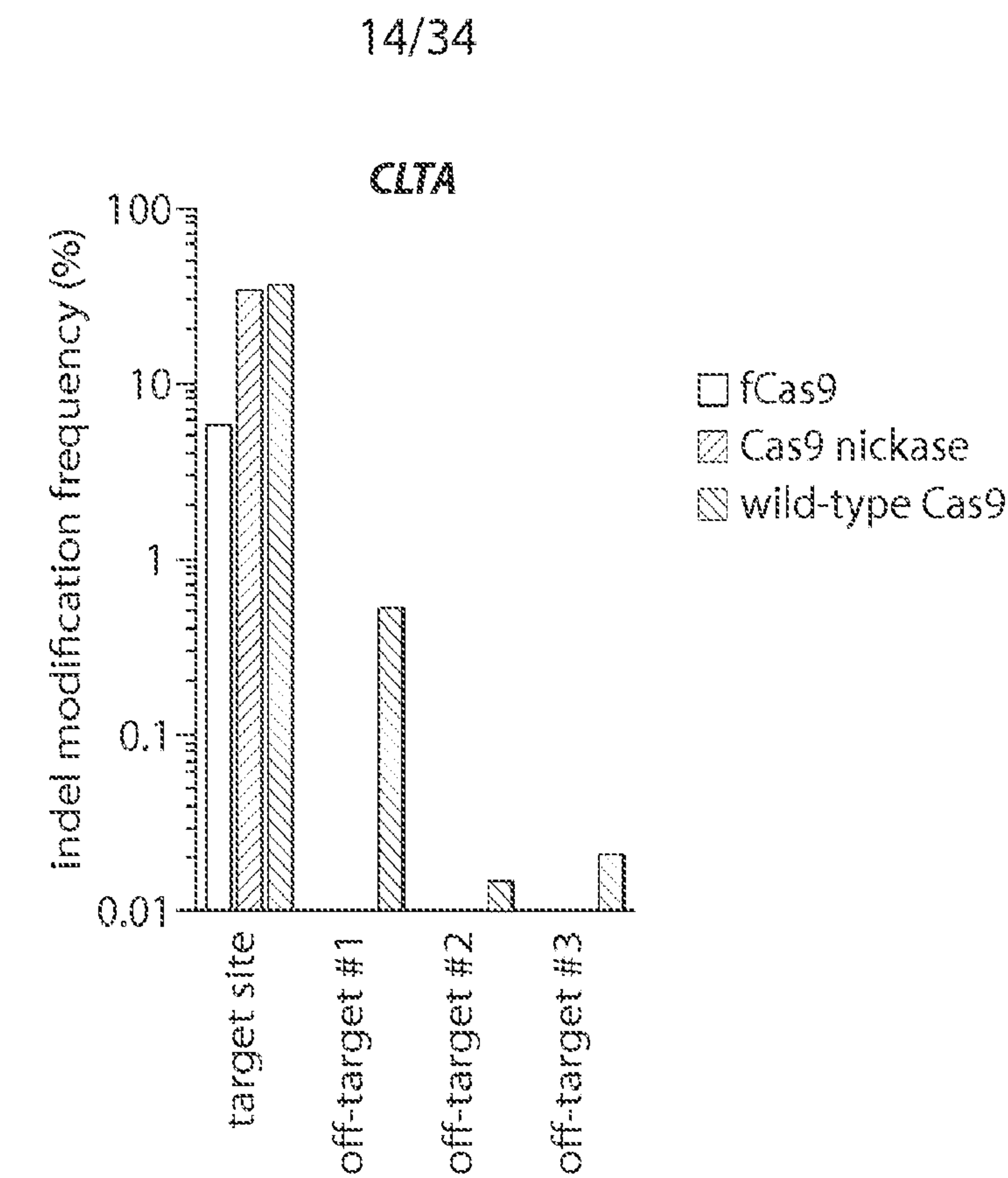


Fig. 8D

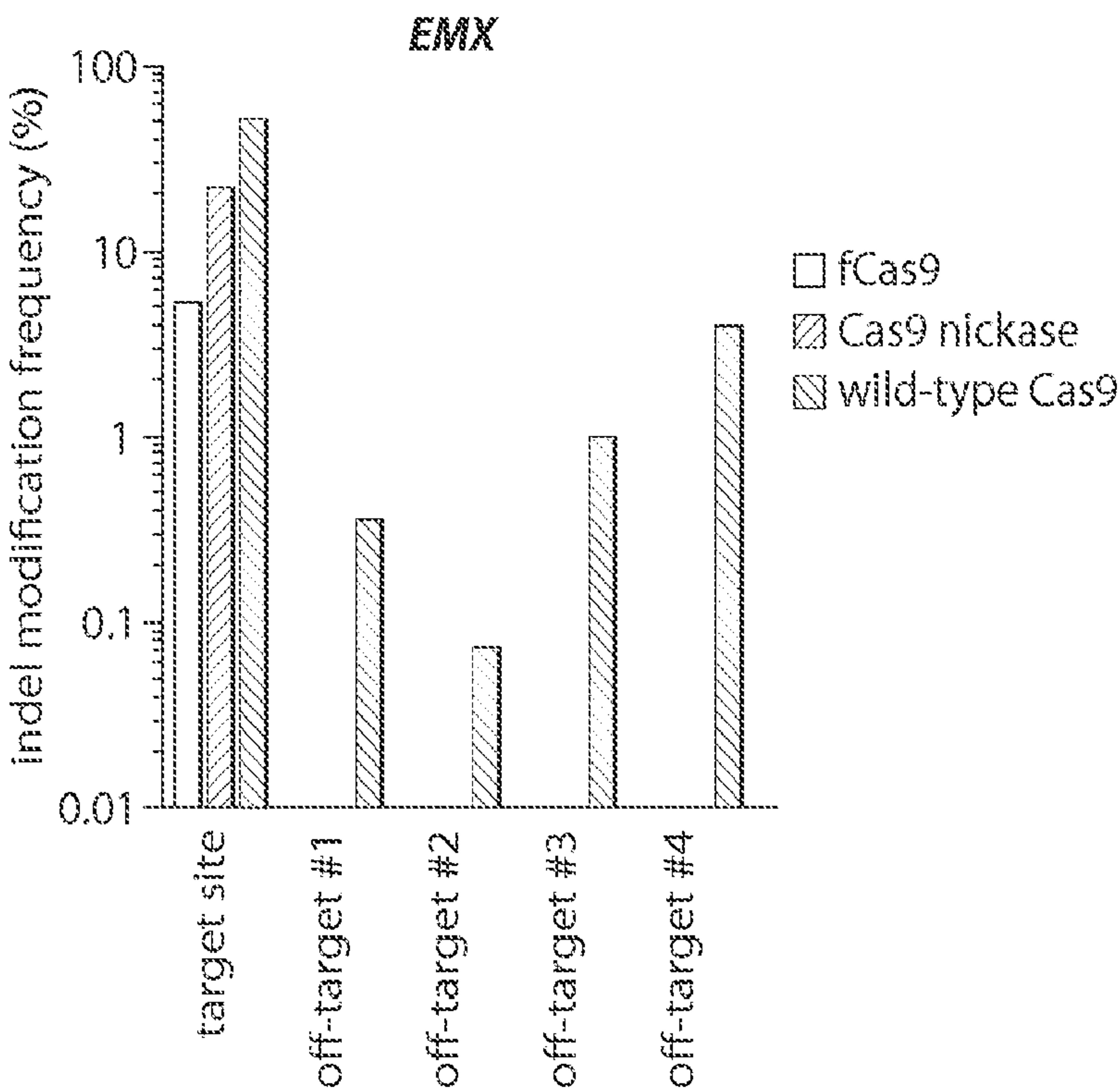


Fig. 8E

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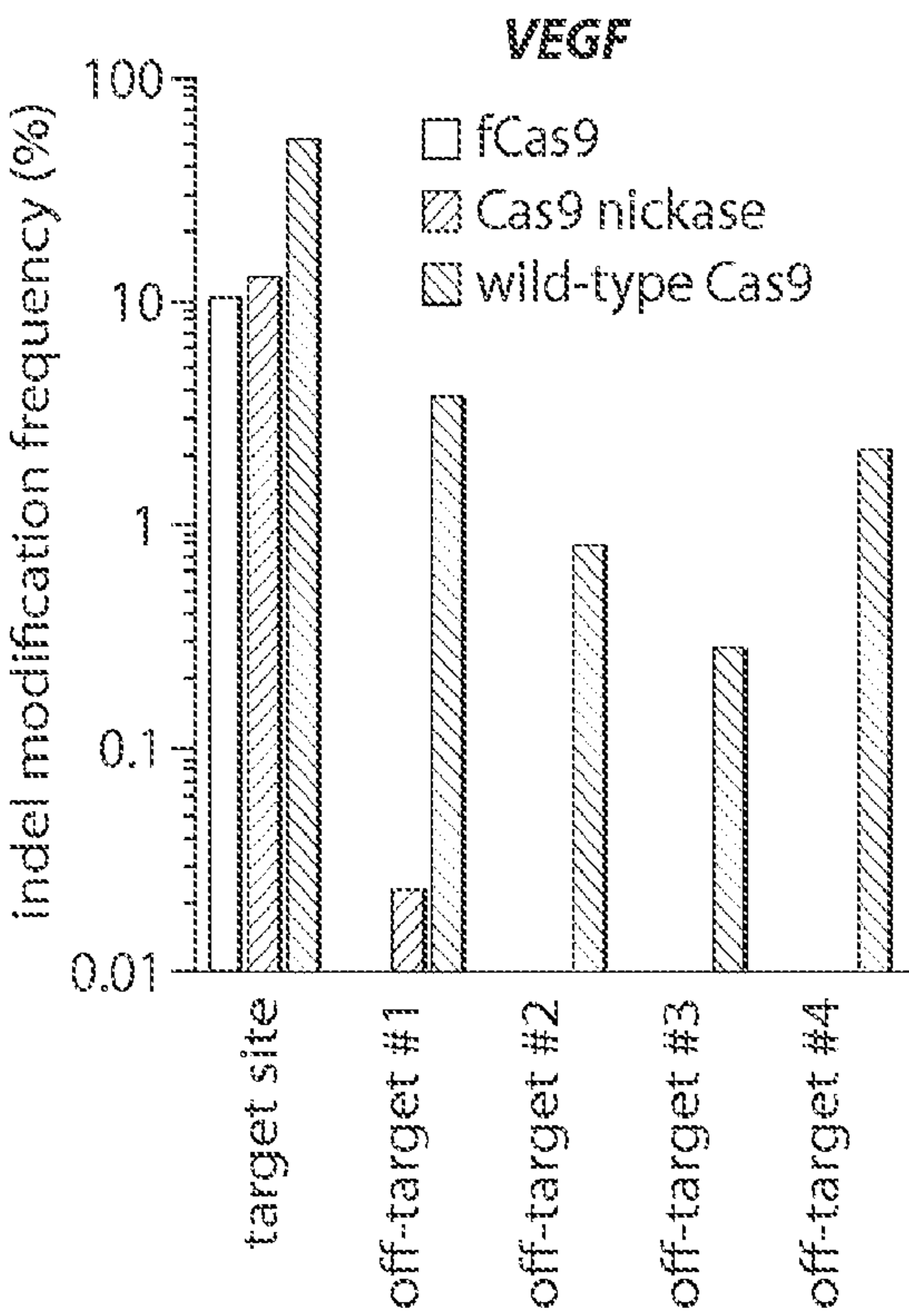


Fig. 8F

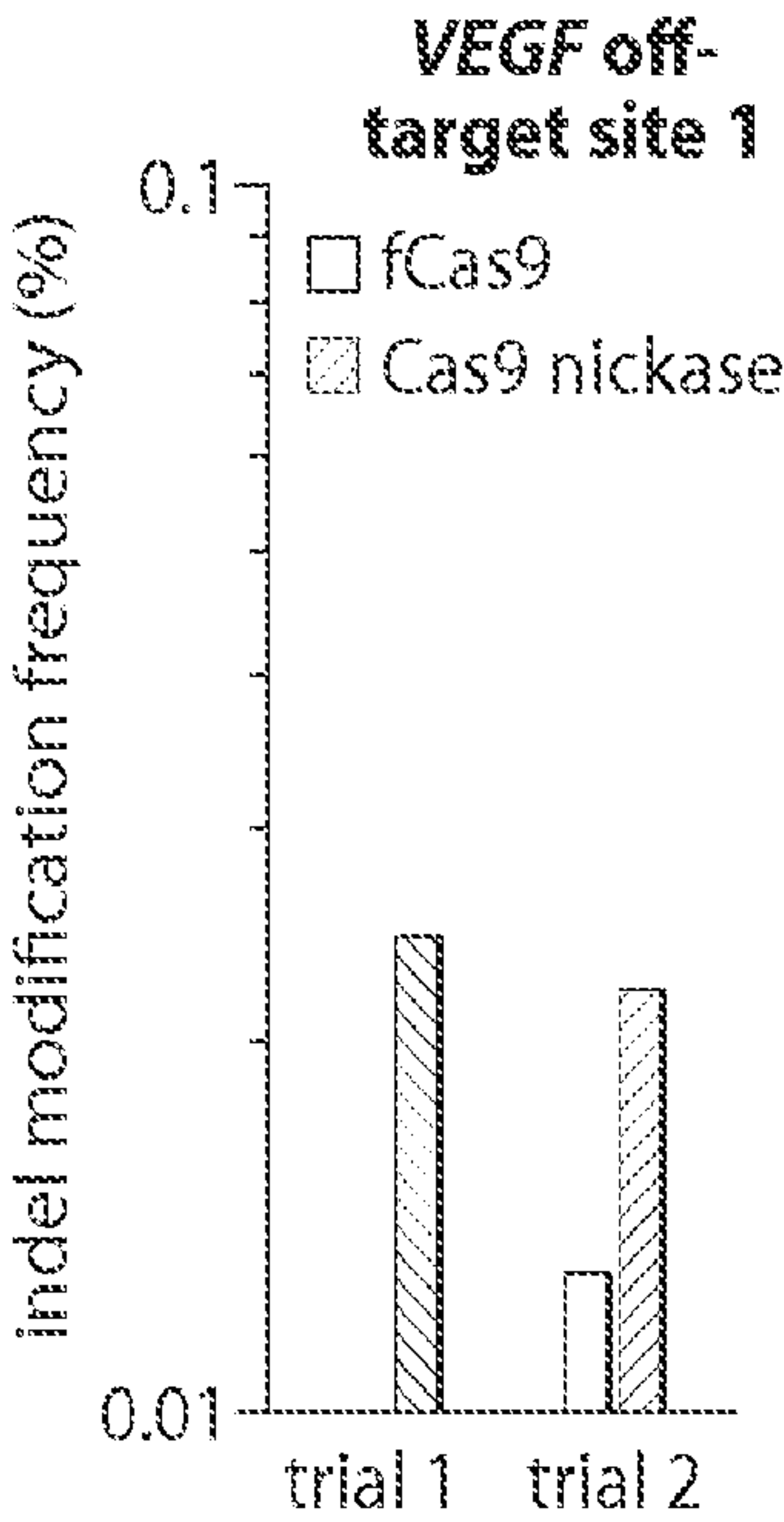


Fig. 8G

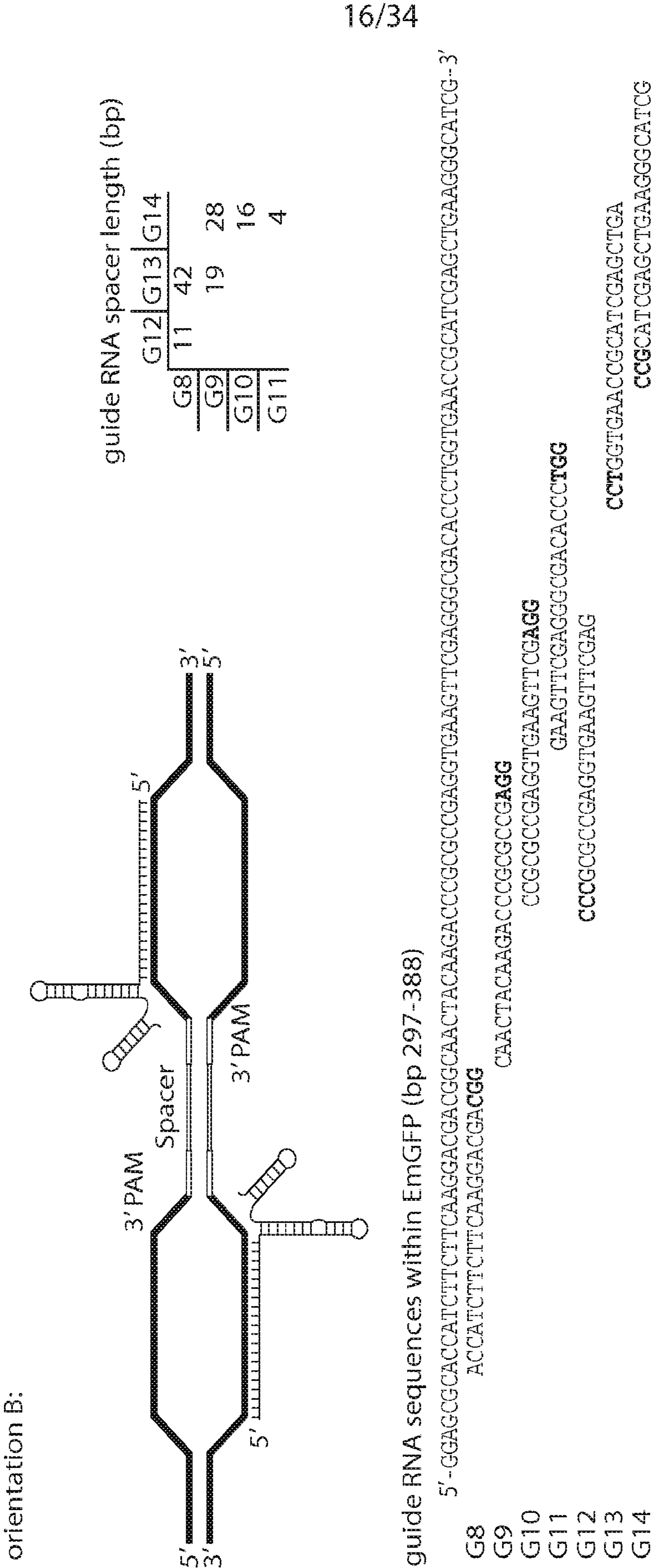


Fig. 9

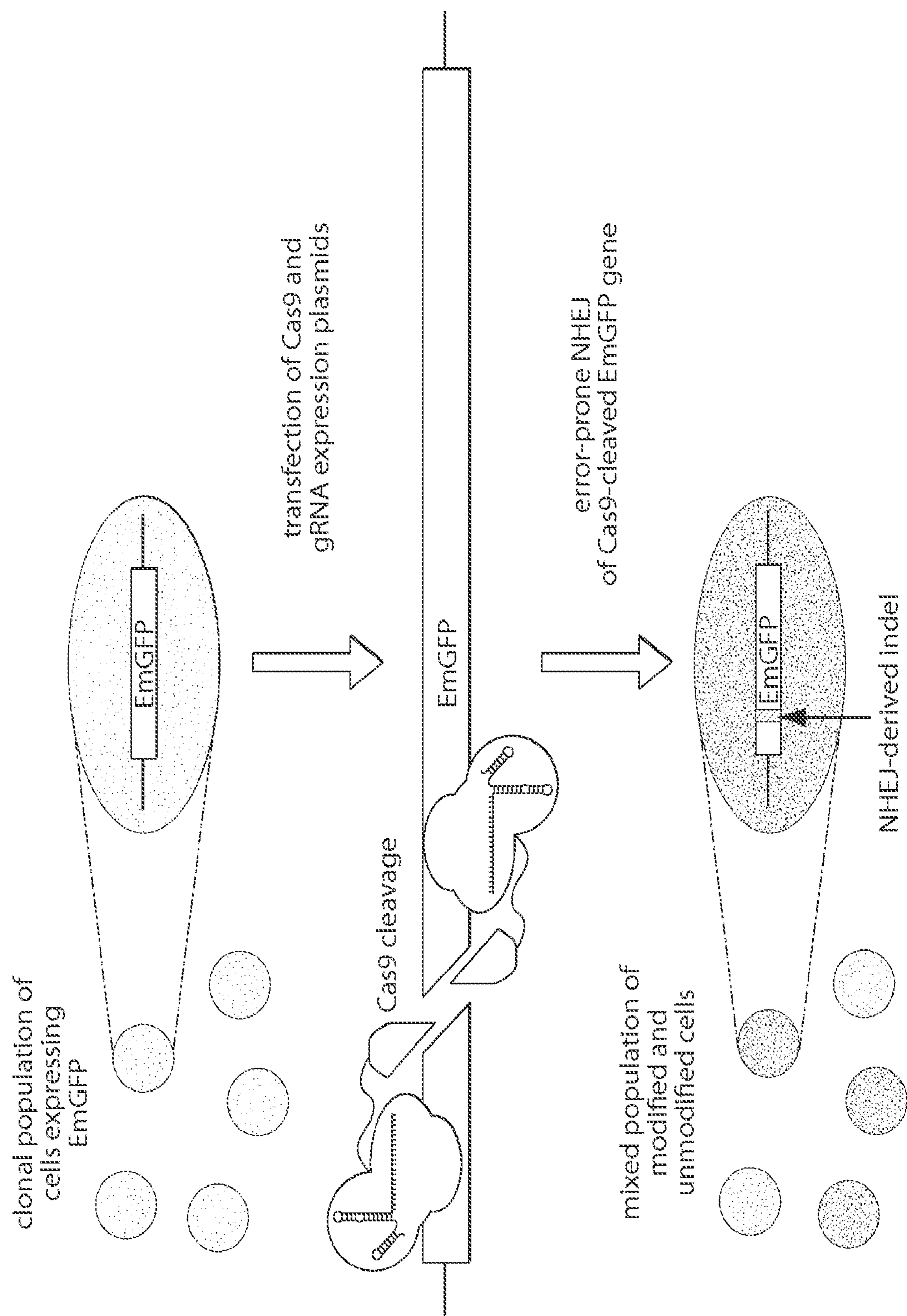
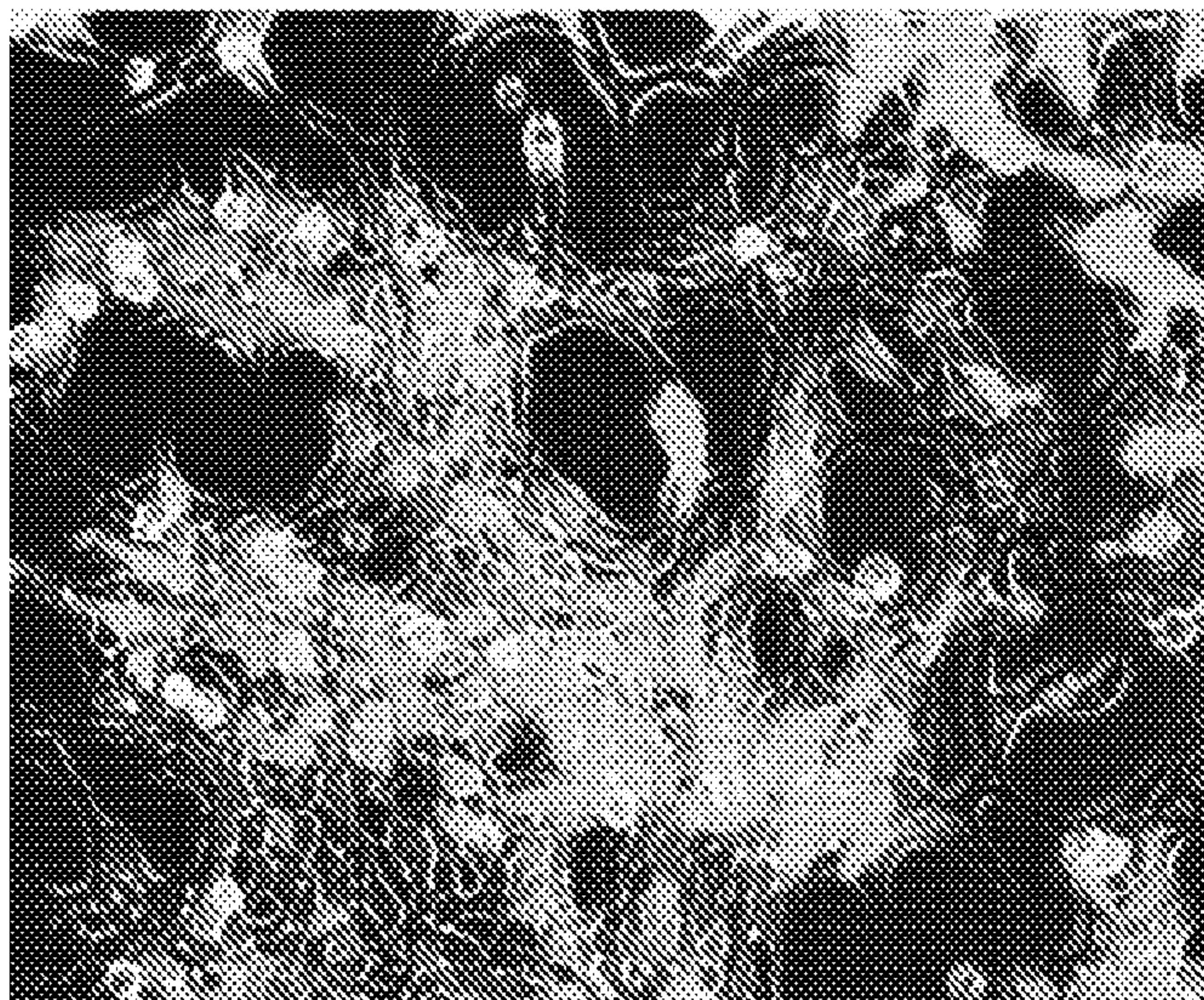


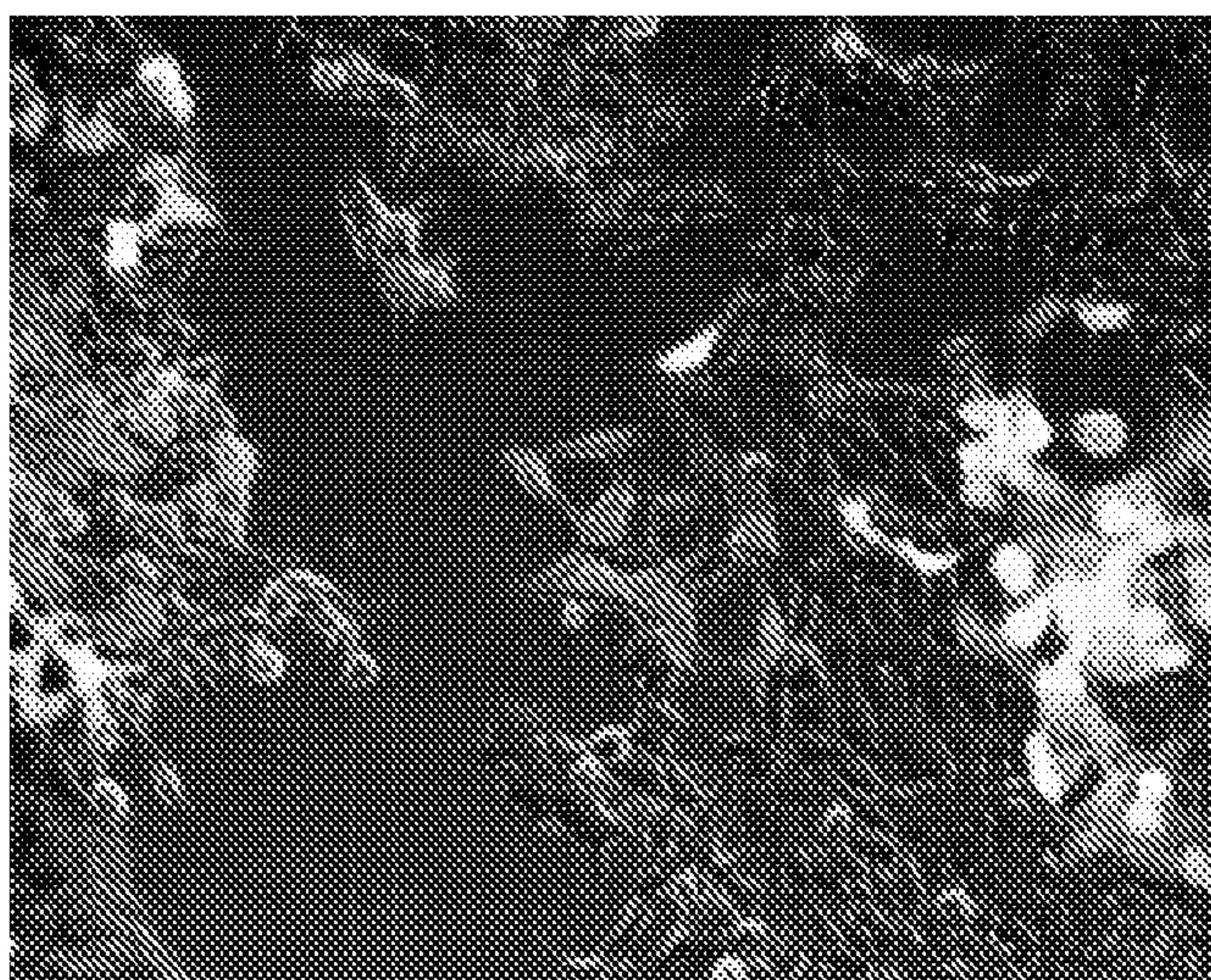
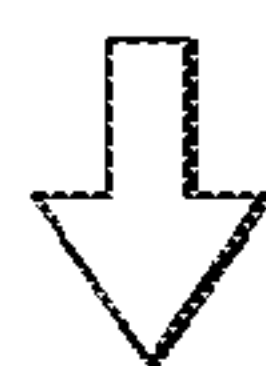
Fig. 10A

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untreated EmGFP-HEK293 cells



Cas9 and gRNA
expression plasmid
transfection



EmGFP-HEK293 cells transfected
with wild-type Cas9 and gRNAs

Fig. 10B

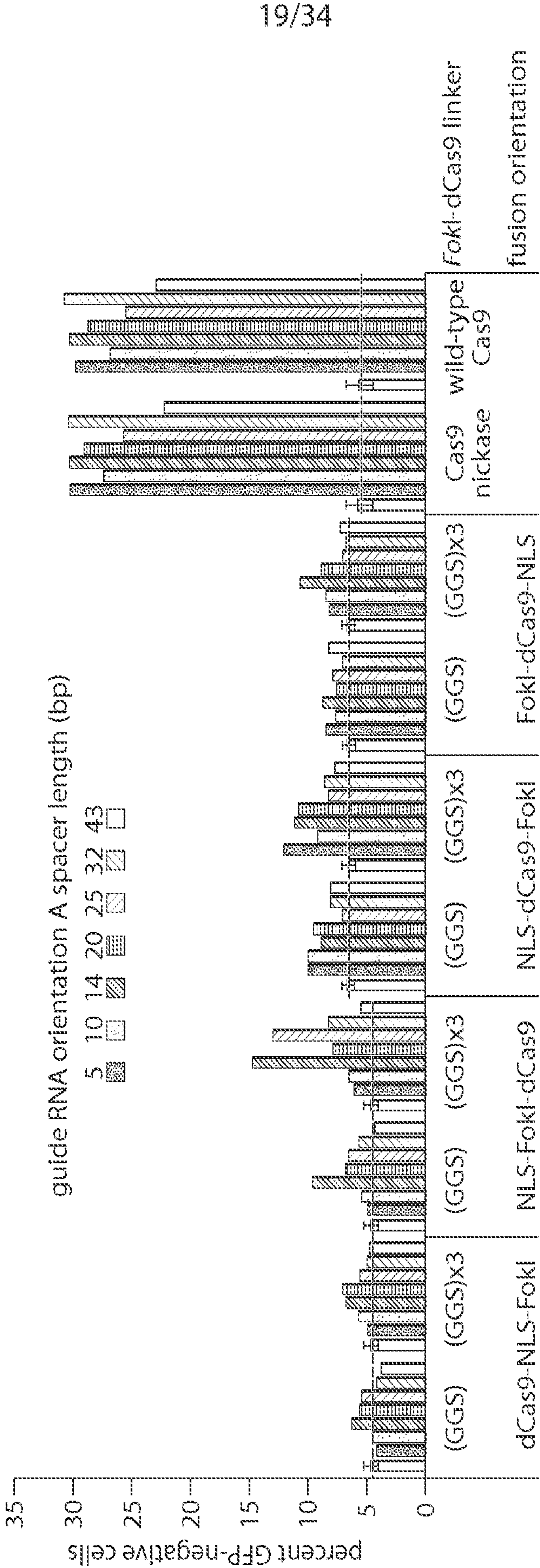


Fig. 11A

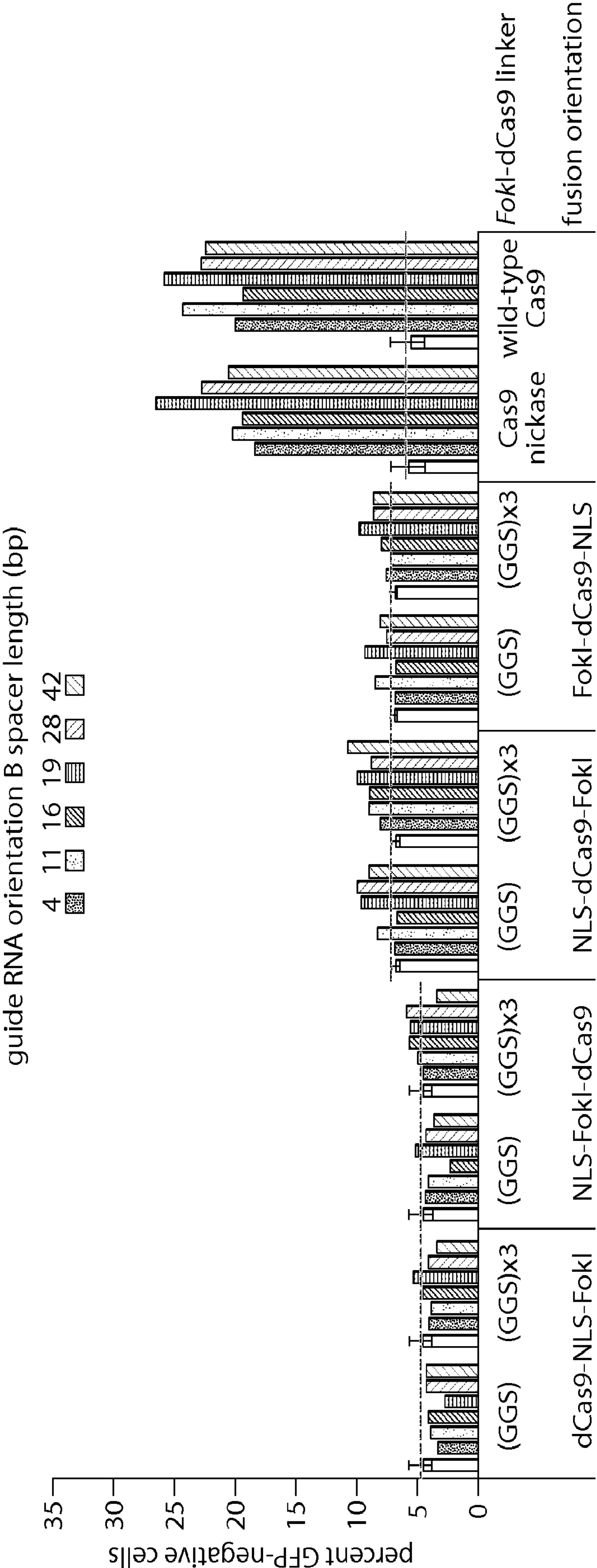


Fig. 11B

name	NLS-linker-Fok1	Fok1-linker-dCas9
<i>FokI</i> -(GGG)x3	GGG	GGSGGGSGGS
<i>FokI</i> -(GGG)x6	GGG	GGSGGGSGGS
<i>FokI</i> -L0	GGG	–
<i>FokI</i> -L1	GGG	MKIIEQLPSA
<i>FokI</i> -L2	GGG	VRHKLKRVGS
<i>FokI</i> -L3	GGG	VPFLLEPDNINGKTC
<i>FokI</i> -L4	GGG	GHGTGSTGSGSS
<i>FokI</i> -L5	GGG	MSRPDPA
<i>FokI</i> -L6	GGG	GSAGSAAGSGEF
<i>FokI</i> -L7	GGG	SGSETPGTSESA
<i>FokI</i> -L8	GGG	SGSETPGTSESATPES
<i>FokI</i> -L9	GGG	SGSETPGTSESATPEGGSGGS
NLS-(GGG)	GGG	GGSM
NLS-(GGG)x3	GGSGGGSGGS	GGSM
NLS-L1	VPFLLEPDNINGKTC	GGSM
NLS-L2	GSAGSAAGSGEF	GGSM
NLS-L3	SIVAQLSRPDPA	GGSM
wild-type Cas9	N/A	N/A
Cas9 nickase	N/A	N/A

Fig. 12A

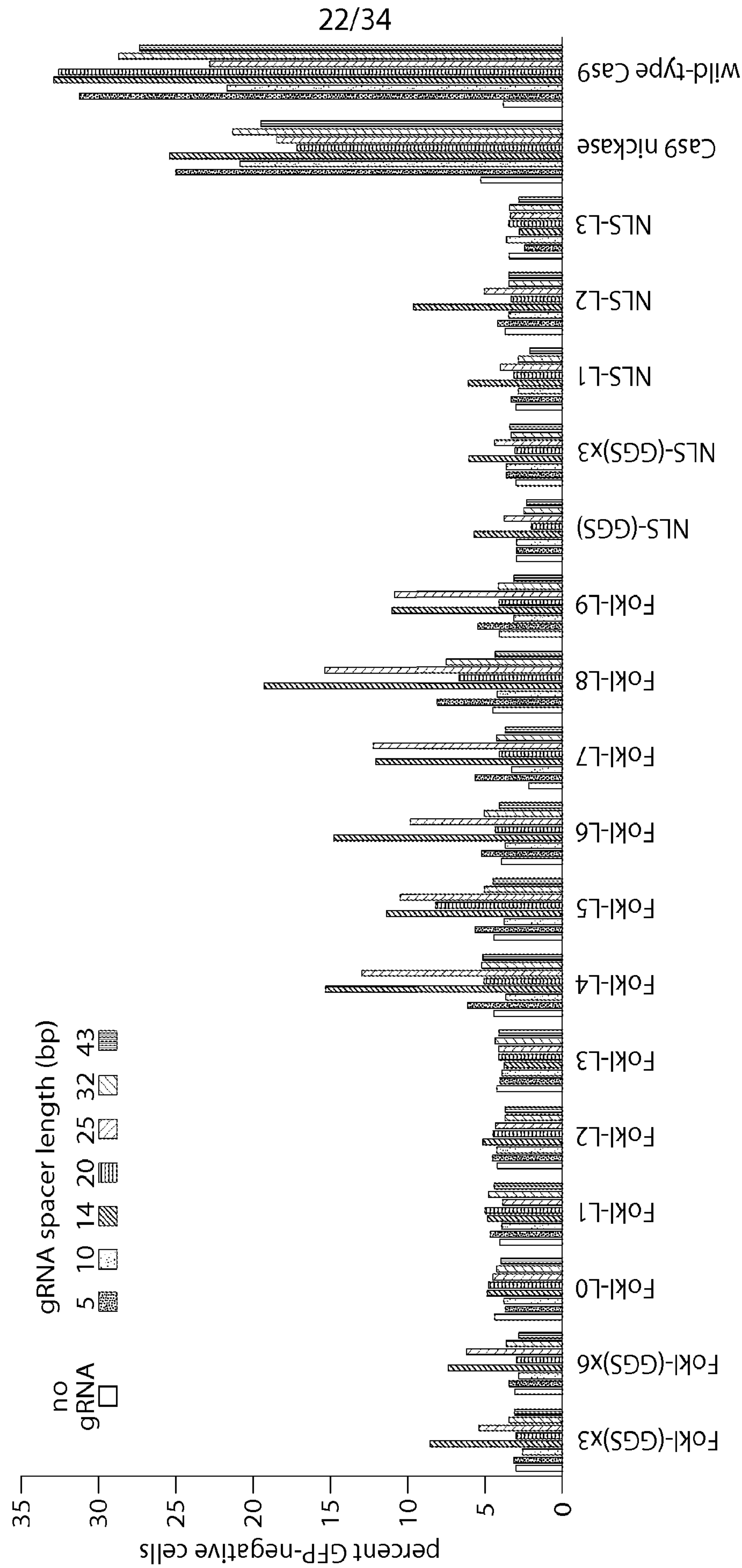


Fig. 12B

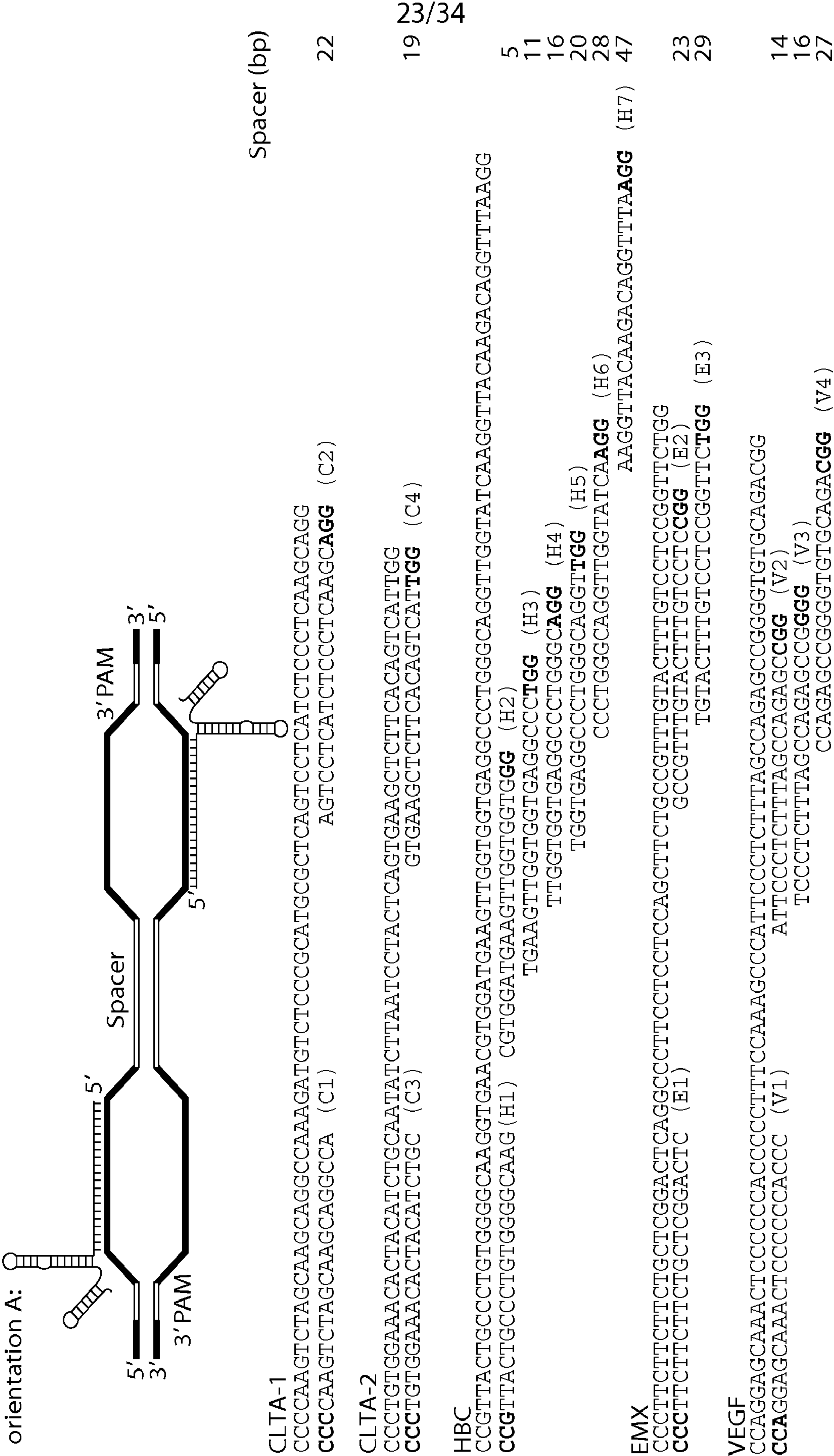


Fig. 13

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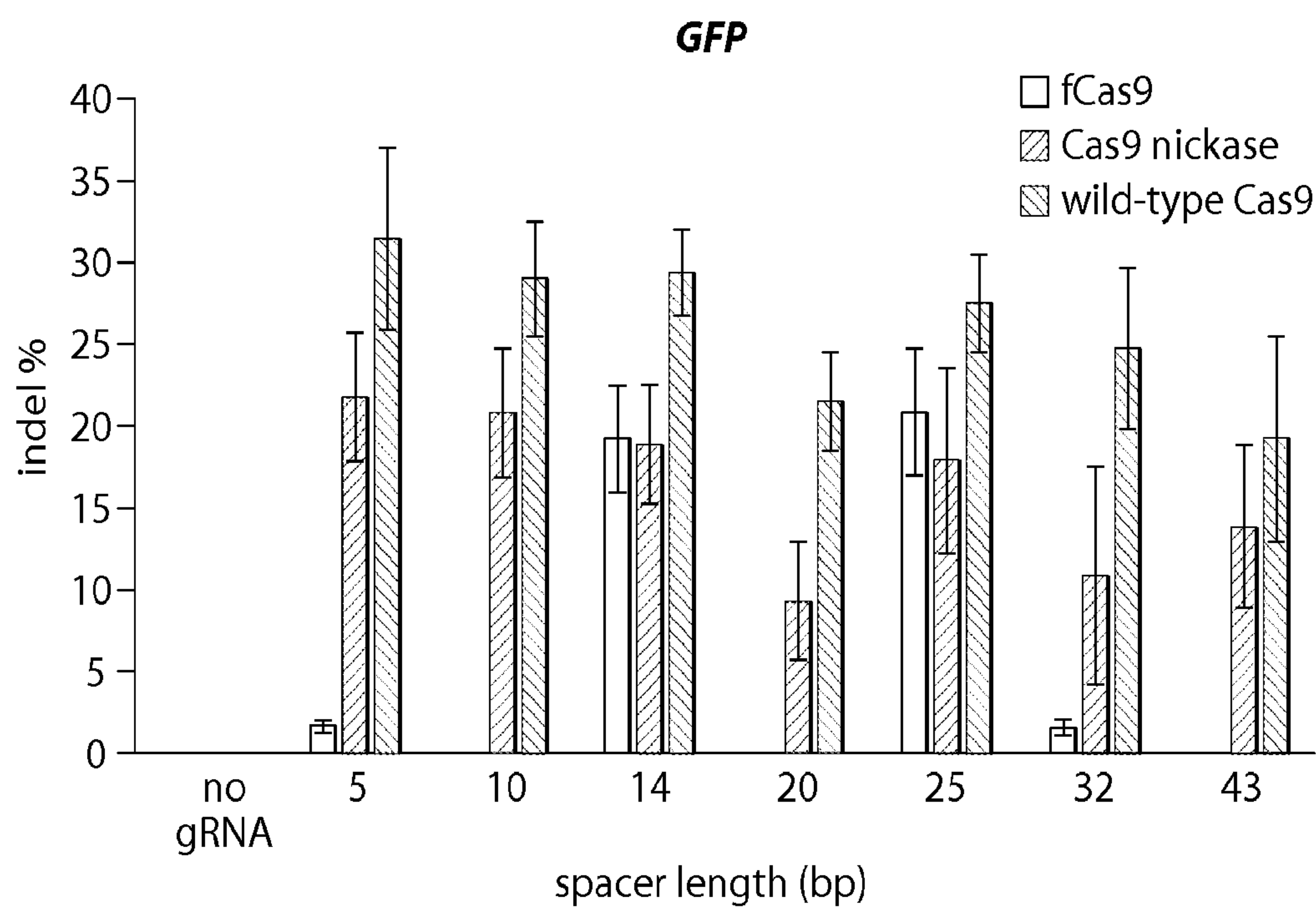


Fig. 14A

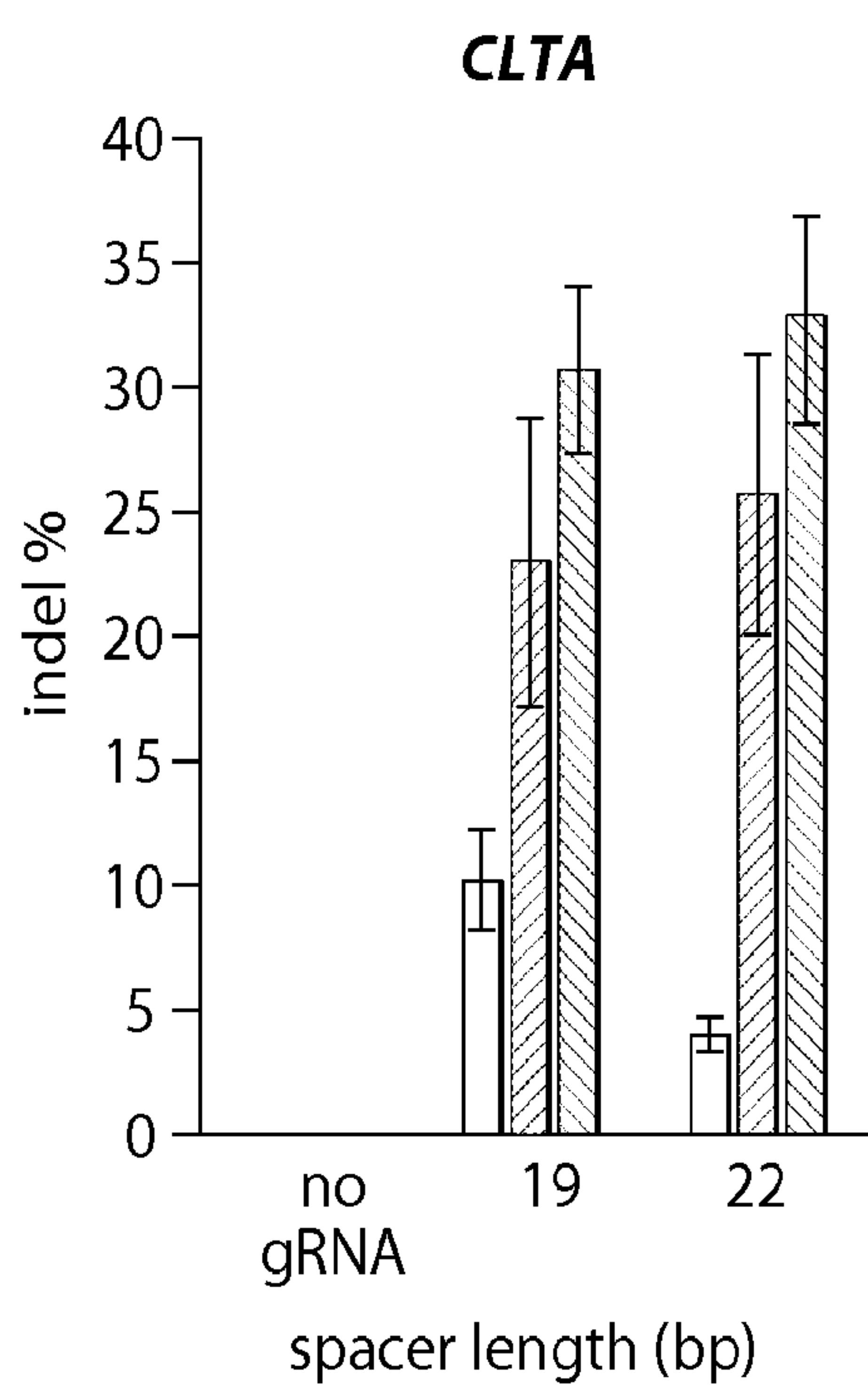


Fig. 14B

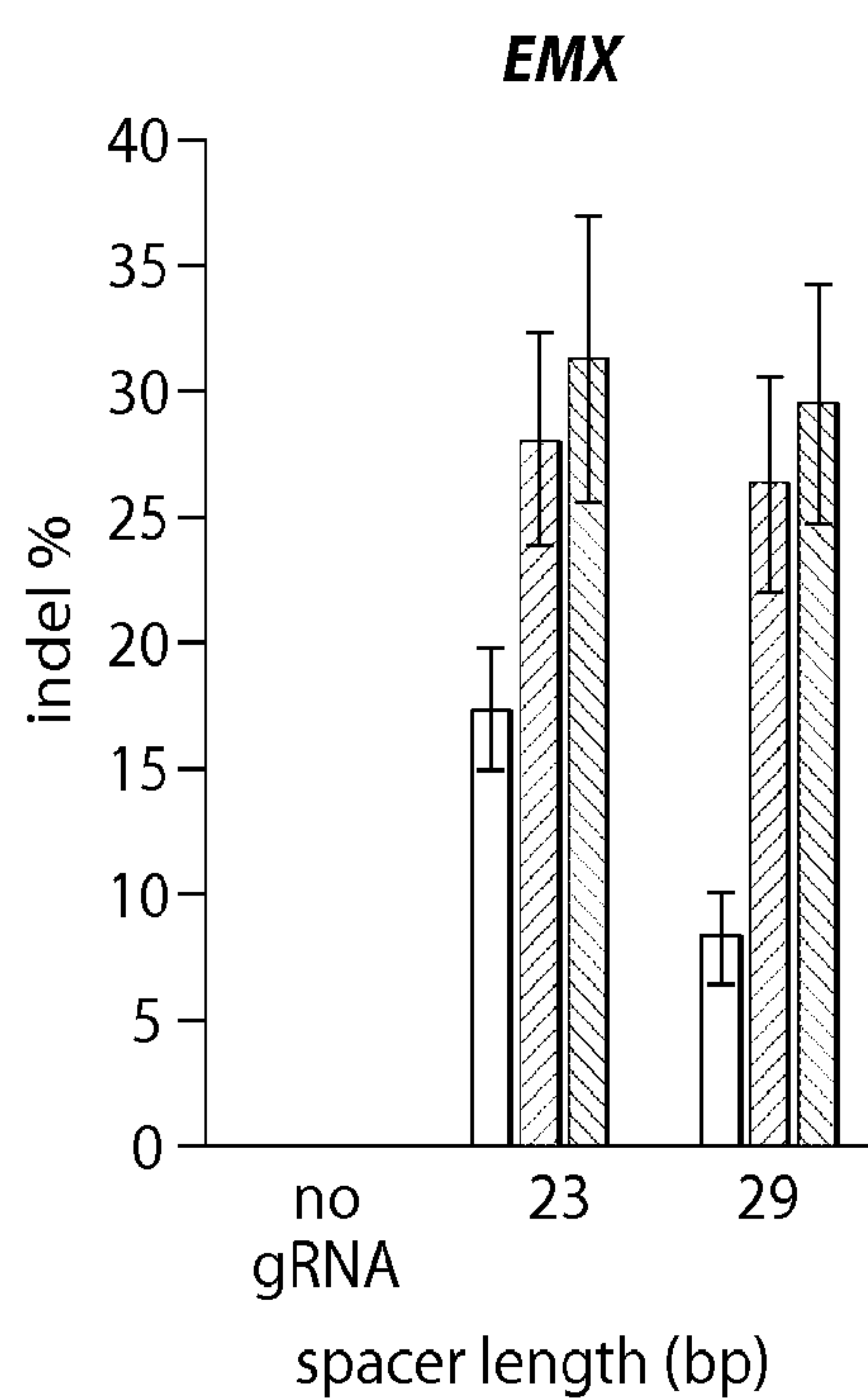


Fig. 14C

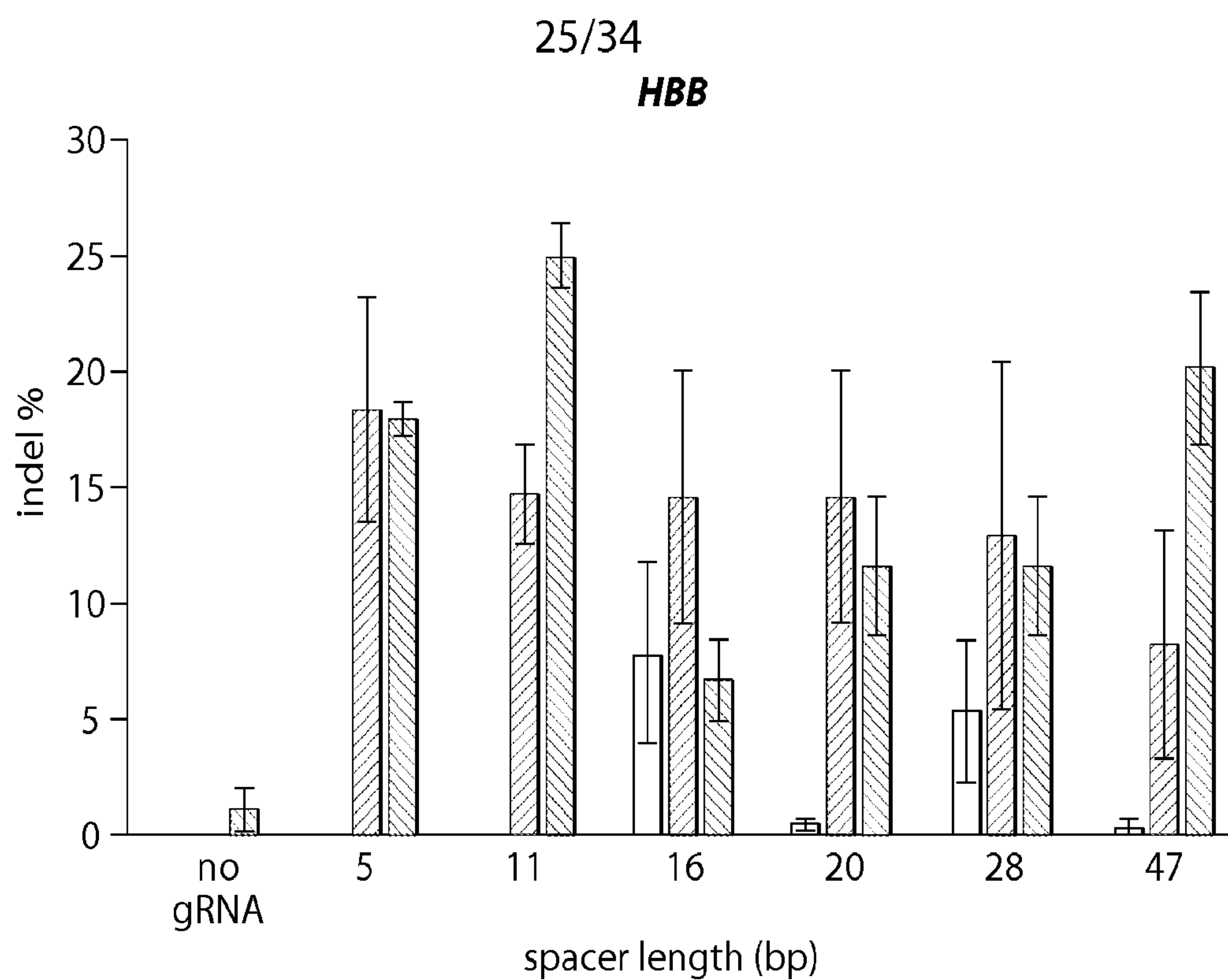


Fig. 14D

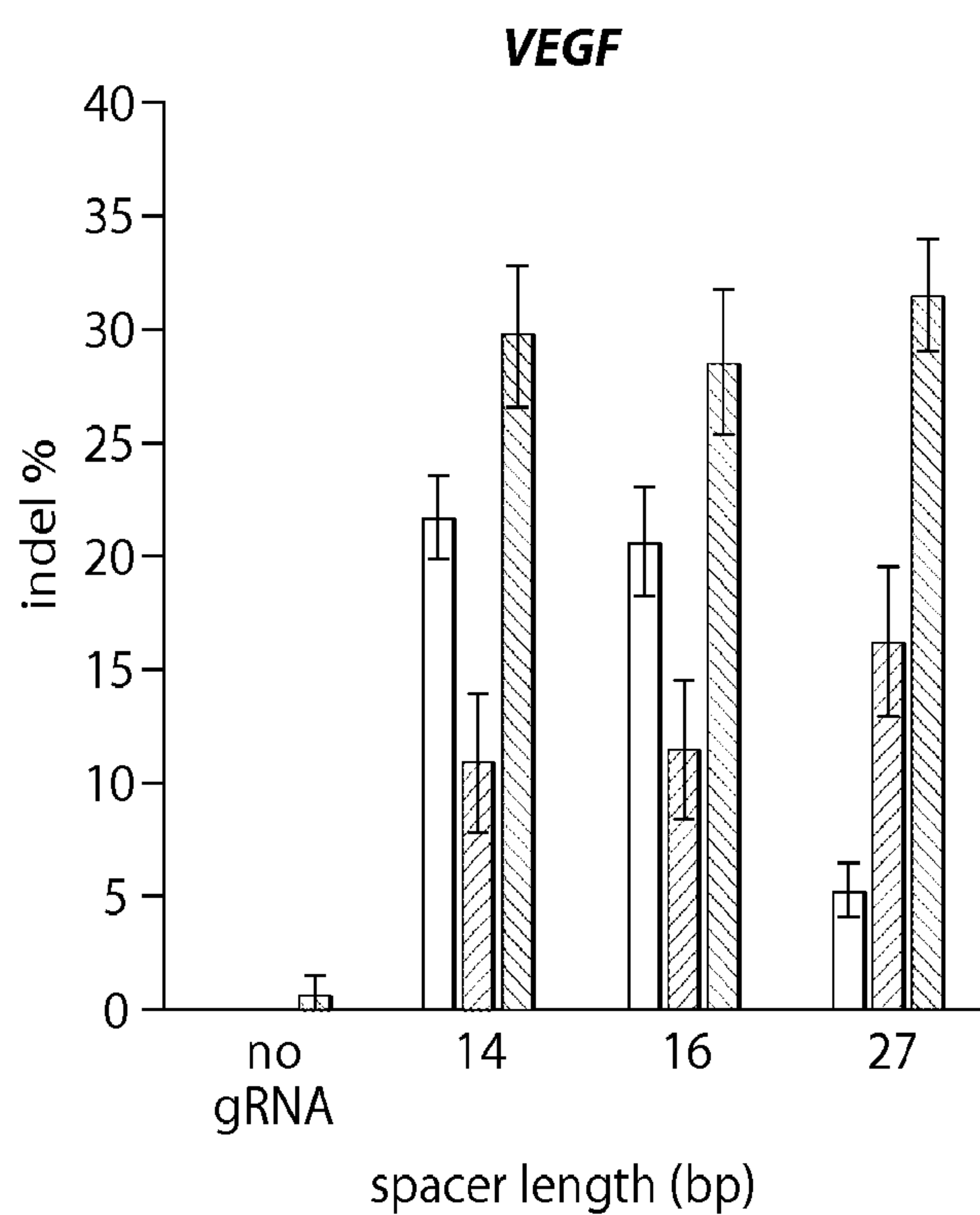


Fig. 14E

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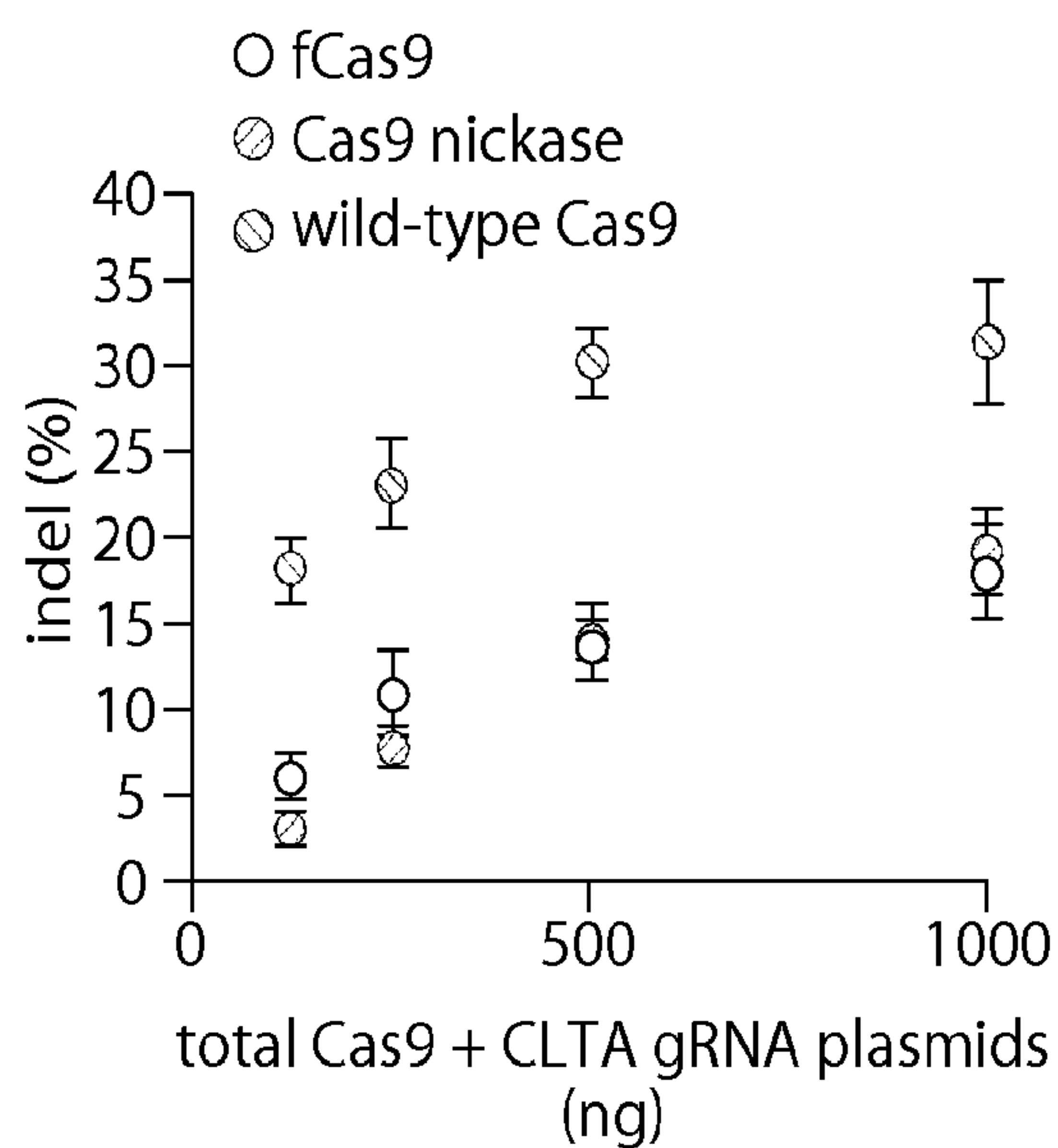


Fig. 15A

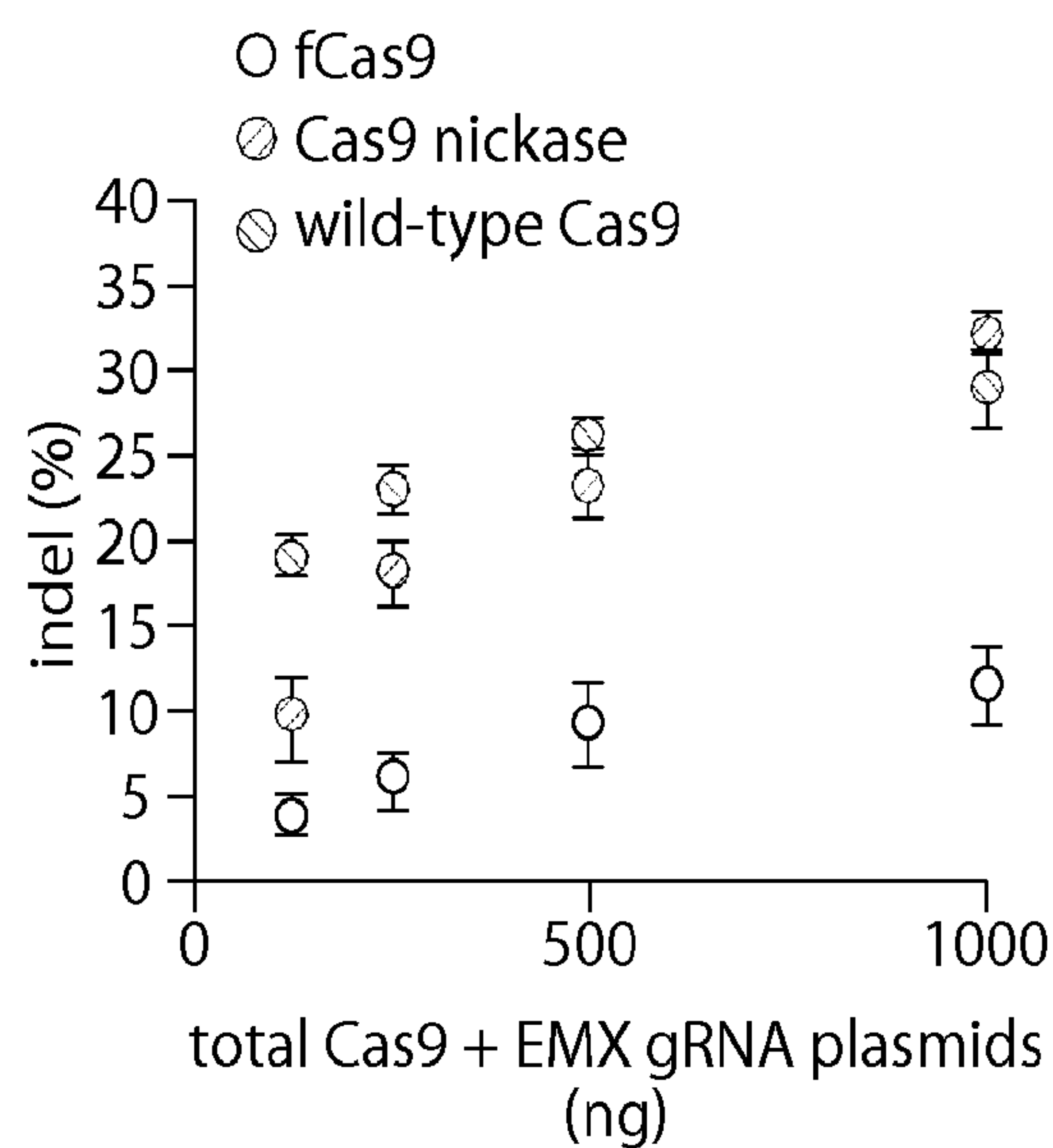


Fig. 15B

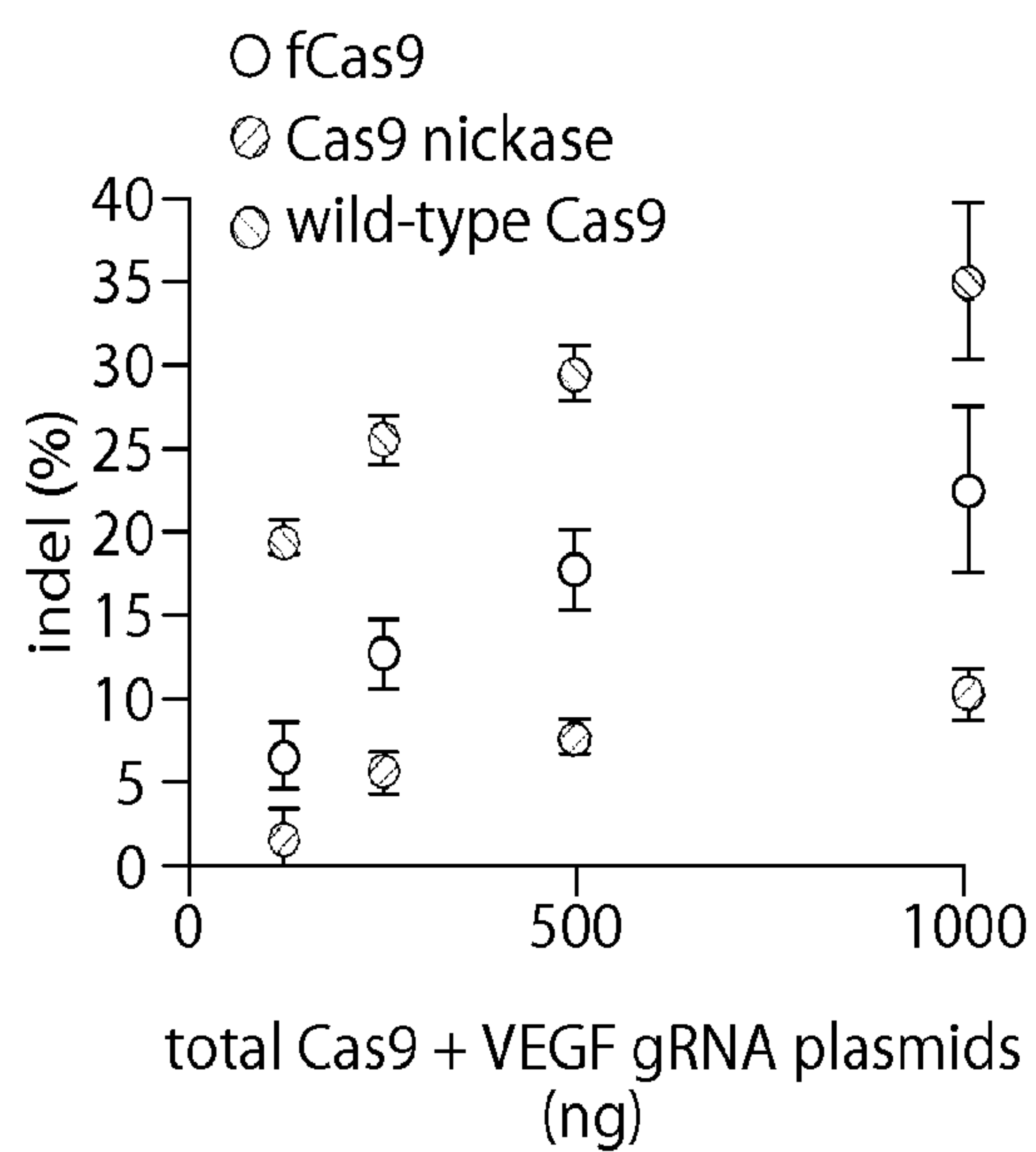


Fig. 15C

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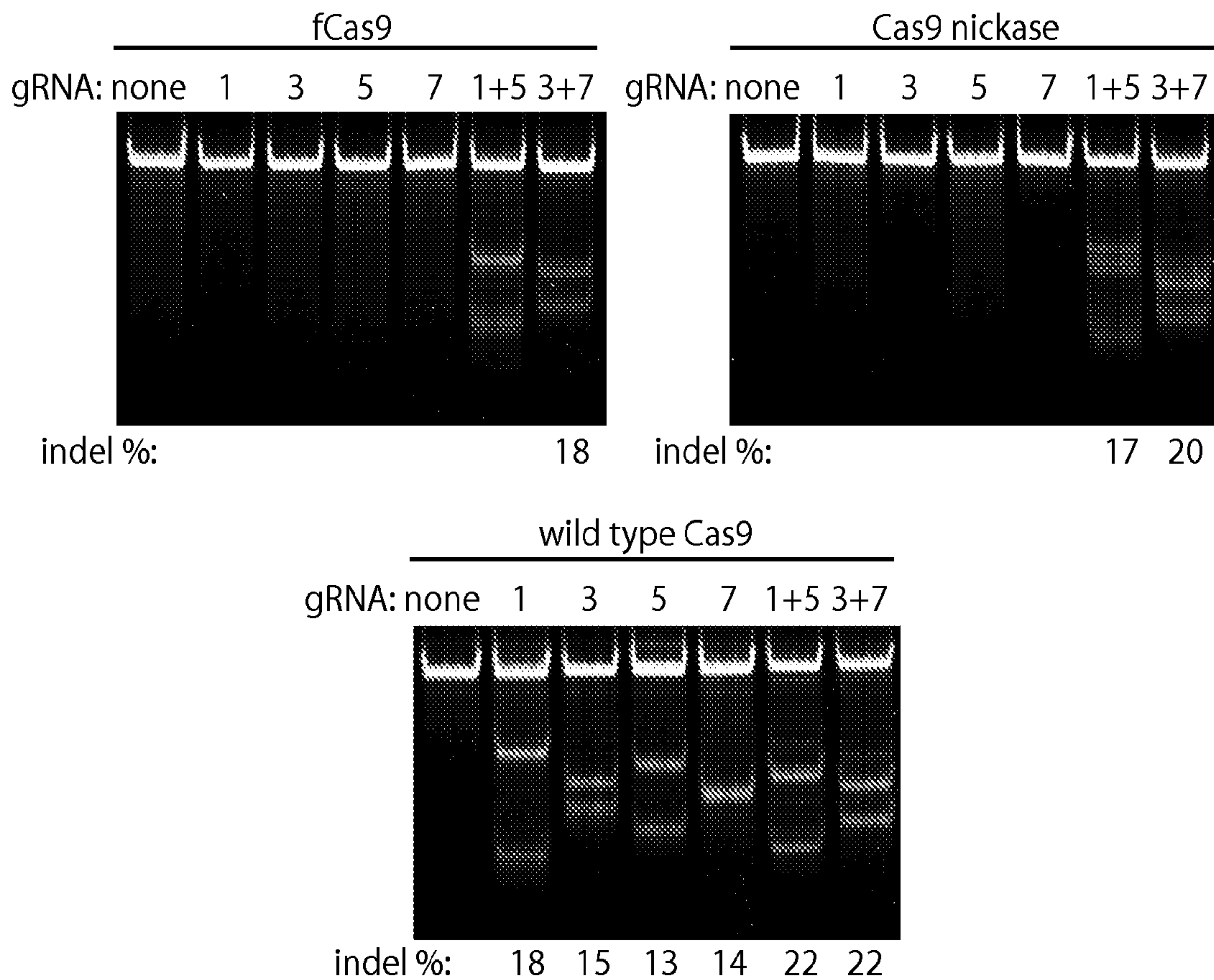


Fig. 16A

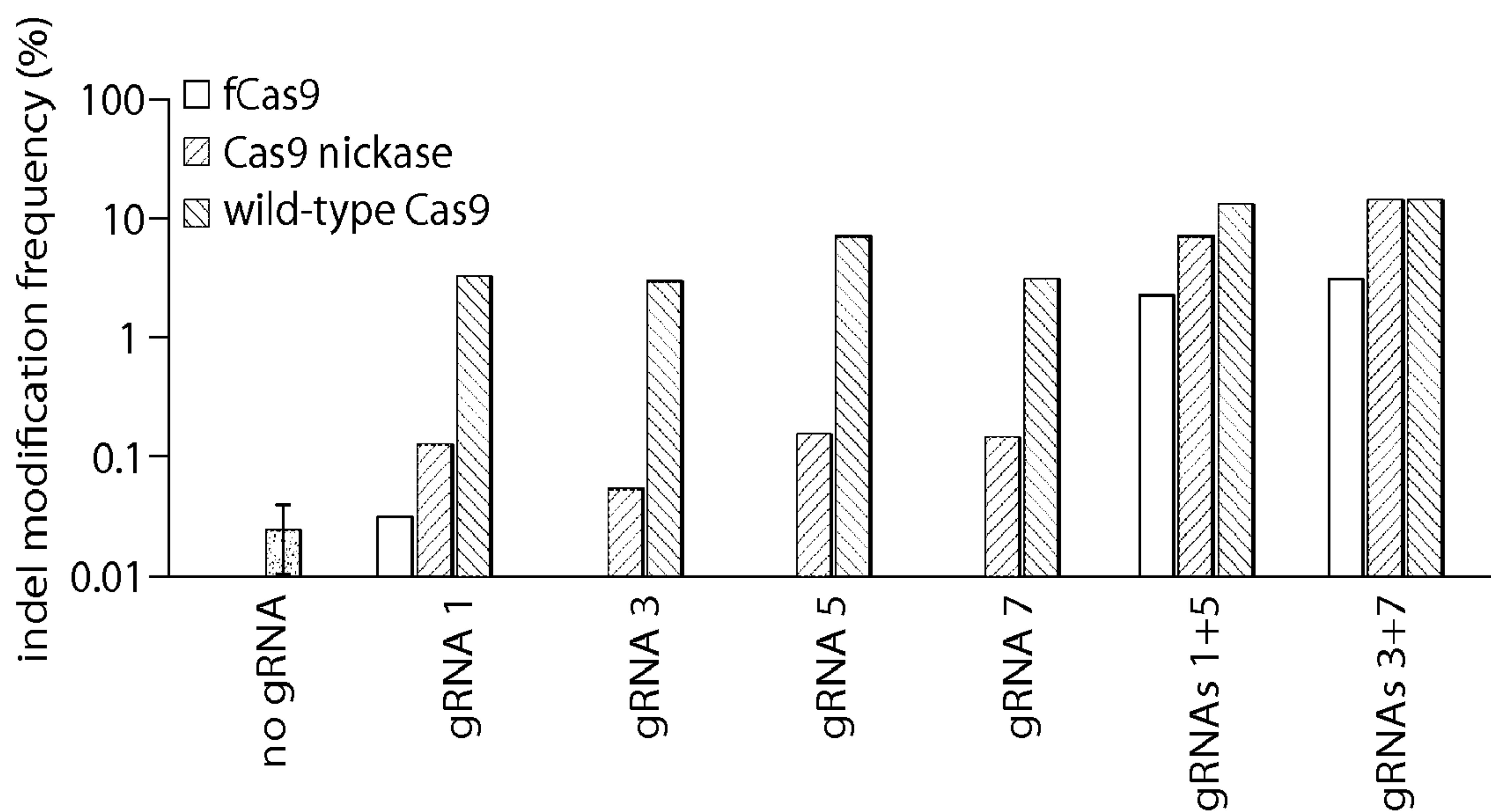


Fig. 16B

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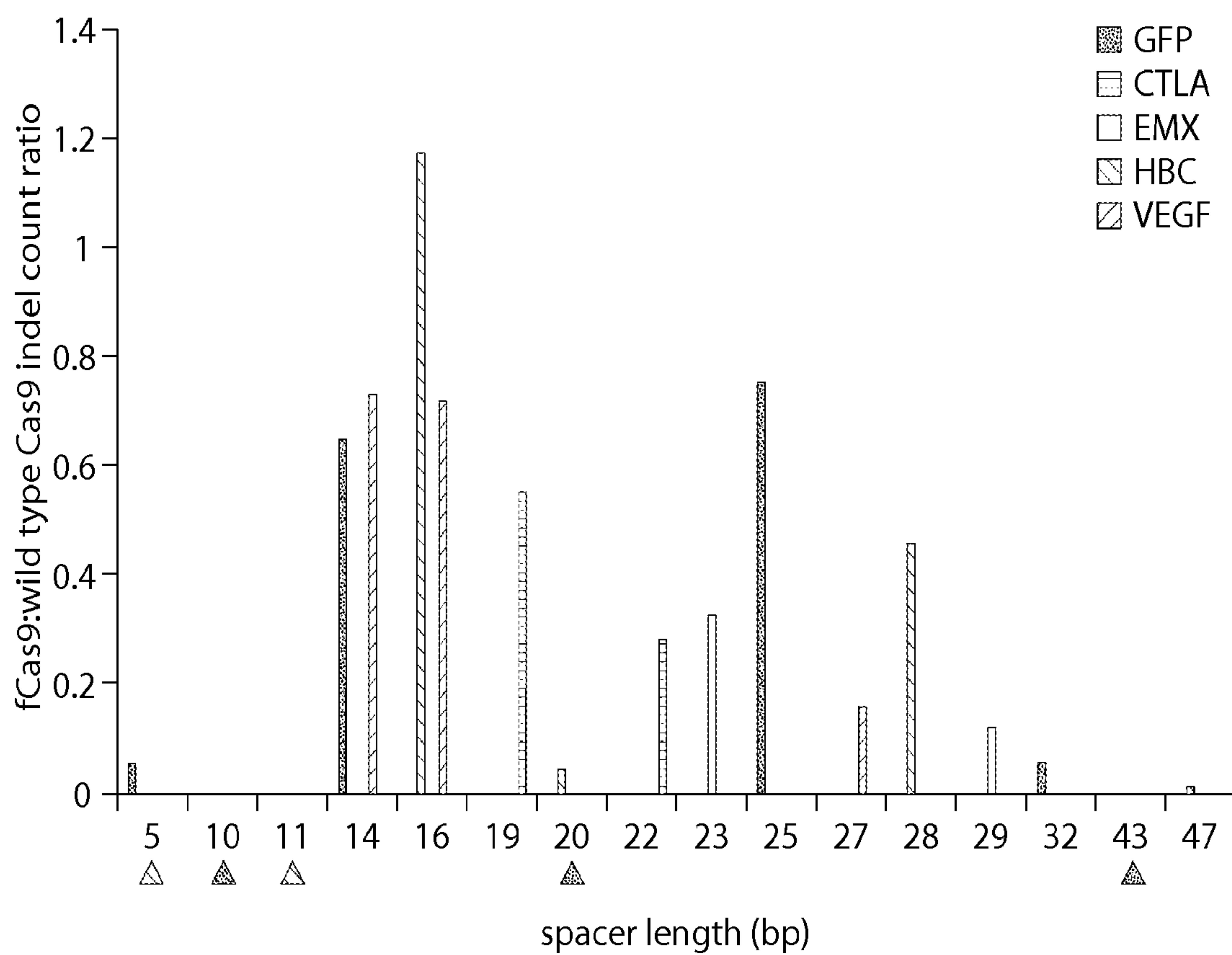


Fig. 17

Wild-type Cas9 nuclease modifications of *VEGF* on-target site:
4747 gctgtttgggaggtcagaaataggggtCCAGGAGCAAACTCCCCCACCCcctttccaaagcccATTCCCTCTTTAGCCAGAGCCGGggtgtgcagacggcagtc (ref)
4577 gctgtttgggaggtcagaaataggggtccaggA-----agccggggtgtgcagacggcagtc
58 gctgtttgggaggtcagaaataggggtccaggagcaaaactccccacccctttccaaagcccatcctctttagc-----cggggtgtgcagacggcagtc
54 gctgtttgggaggtcagaaataggggtccaggag-----agccggggtgtgcagacggcagtc
43 gctgtttgggaggtcagaaatag-----ccggggtgtgcagacggcagtc
33 gctgtttgggaggtcagaaataggggtccaggagc-----cggggtgtgcagacggcagtc
23 gctgtttgggaggtcagaaataggggtccag-----ccggggtgtgcagacggcagtc
22 gctgtttgggaggtcagaaataggggtccaggagcaaaactccccacccctttccaaagcccatcctctttagccag-----ggtgtgcagacggcagtc
18 gctgtttgggaggtcagaaataggggtccaggA-----t-----agccggggtgtgcagacggcagtc

Cas9 nickase modifications of *VEGF* on-target site:
8770 gctgtttgggaggtcagaaataggggtCCAGGAGCAAACTCCCCCACCCcctttccaaagcccATTCCCTCTTTAGCCAGAGCCGGggtgtgcagacggcagtc (ref)
78 gctgtttgggaggtcagaaataggggtccag-----acggcagtc
60 gctgtttgggaggtcagaaataggggtccaggagcaaaactccccacccctttccaaagcc-----ggggtgtgcagacggcagtc
58 gctgtttgggaggtcagaaataggggtcca-----aagcccatcctctttagccagagccggggtgtgcagacggcagtc
56 gctgtttgggaggtcagaaataggggt-----gtgcagacggcagtc
49 gctgtttgggaggtcagaaataggggtccag-----ccggggtgtgcagacggcagtc
37 gctgtttgggaggtcagaaataggggtccagg-----gtgtgcagacggcagtc
36 gctgtttgggaggtcagaaataggggtccaggagc-----cggggtgtgcagacggcagtc
27 gctgtttgggaggtcagaaatag-----ccggggtgtgcagacggcagtc

fCas9 nuclease modifications of *VEGF* on-target site:
8959 gctgtttgggaggtcagaaataggggtCCAGGAGCAAACTCCCCCACCCcctttccaaagcccATTCCCTCTTTAGCCAGAGCCGGggtgtgcagacggcagtc (ref)
125 gctgtttgggaggtcagaaataggggtccaggagcaaaactccccca-----agcccatcctctttagccagagccggggtgtgcagacggcagtc
121 gctgtttgggaggtcagaaataggggtccaggagcaaaactccccacccctt-----ttccctctttagccagagccggggtgtgcagacggcagtc
77 gctgtttgggaggtcagaaataggggtccaggagcaaaactccccacccct-----ttccctctttagccagagccggggtgtgcagacggcagtc
73 gctgtttgggaggtcagaaataggggtccaggagcaaaactccccca-----gcccatcctctttagccagagccggggtgtgcagacggcagtc
48 gctgtttgggaggtcagaaataggggtccaggagcaaaactccccacccct-----attccctctttagccagagccggggtgtgcagacggcagtc
44 gctgtttgggaggtcagaaataggggtccaggagcaaaactccccacccctt-----agccagagccggggtgtgcagacggcagtc
24 gctgtttgggaggtcagaaataggggtccaggagcaaaactccccacccctt-----aaagcccatcctctttagccagagccggggtgtgcagacggcagtc
22 gctgtttgggaggtcagaaataggggtccaggagcaaaactcccc-----aagcccatcctctttagccagagccggggtgtgcagacggcagtc

Fig. 18A

Wild-type Cas9 nuclease modifications of VEG_Off1:

79248 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccCCAGGAGCAA**ACTCCCTCCAT**CCCacaaaatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat (ref)
800 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccCCAGG--CAAAC**TCCCTCCAT**CCCacaaaatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
239 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacacccca-----tccacaaaatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
155 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacacccca---caaactccctccatccacaaaatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
90 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacacccca-----caaatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
71 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacac-----tccctccatccacaaaatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
54 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacacccc-----tccatccacaaaatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
53 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaaggga-----tgat
47 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgac-----tccctccatccacaaaatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat

Cas9 nickase modifications of VEG_Off1:

302573 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccCCAGGAGCAA**ACTCCCTCCAT**CCCacaaaatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat (ref)
28 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaaggga-----c-----atccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
13 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgac-----tccctccatccacaaaatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
11 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctga-----tcgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
4 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaagg--ca---tc--tg-atg--acaaaatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
2 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaaggacaaaactccctccatccacaaaatccgctccttagatgtgca-ccccacccctcctaagaataagaaggatgat
1 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaaggacaaaactccctccatccacaaaatccgctccttagatgtgca-ccccaccccaacctcctaagaataagaaggatgat
1 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaaggga-----tgatgcacaccccaacctcctaagaataagaaggatgat
1 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctga-----aatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat

fCas9 nuclease modifications of VEG_Off1:

233567 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccCCAGGAGCAA**ACTCCCTCCAT**CCCacaaaatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat (ref)
6 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaggagcaaaactccctc-----gtccttagatgtgcacaccccaacctcctaagaataagaaggatgat
5 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaaggacaaaactccctc-----tccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
4 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaaggacaaaactccctccat-----tccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
3 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaaggacaaaactccctccatccatccacaaaatccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
3 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaaggacaaaactccctcc-----tccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
2 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaaggacaaaactccctcc-----atccgctccttagatgtgcacaccccaacctcctaagaataagaaggatgat
1 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaaggacaaaactccctccatccacaaaatccgctccttagatgtgcacaccc-----tccataagaataagaaggatgat
1 cattcaacagatacttactgaatgctaataatgtctcagacaggacattctgacaccccaaggacaaaactccctccatccacaaaatccgctcc--agatgtgcacaccccaacctcctaagaataagaaggatgat

Fig. 18B

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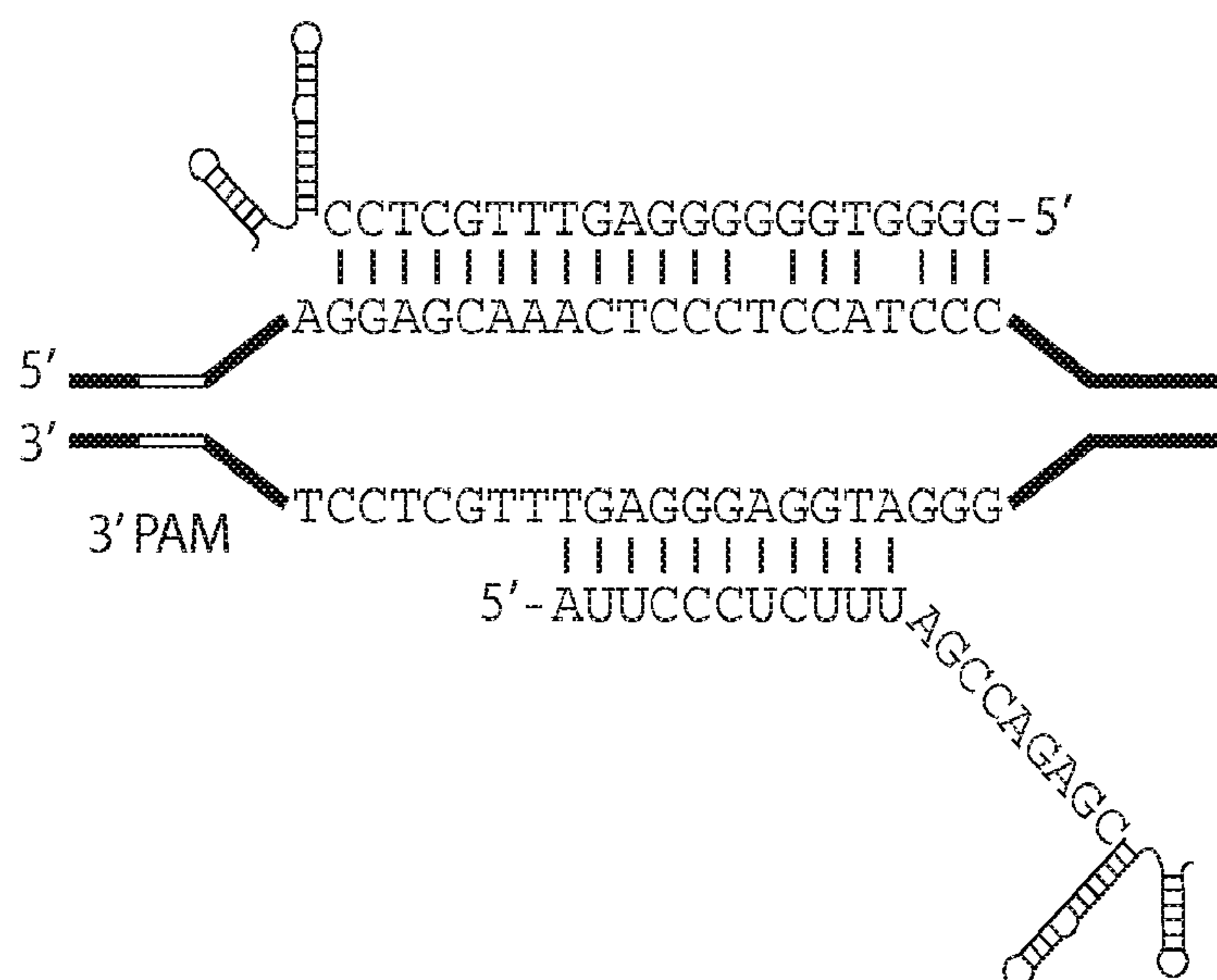


Fig. 18C

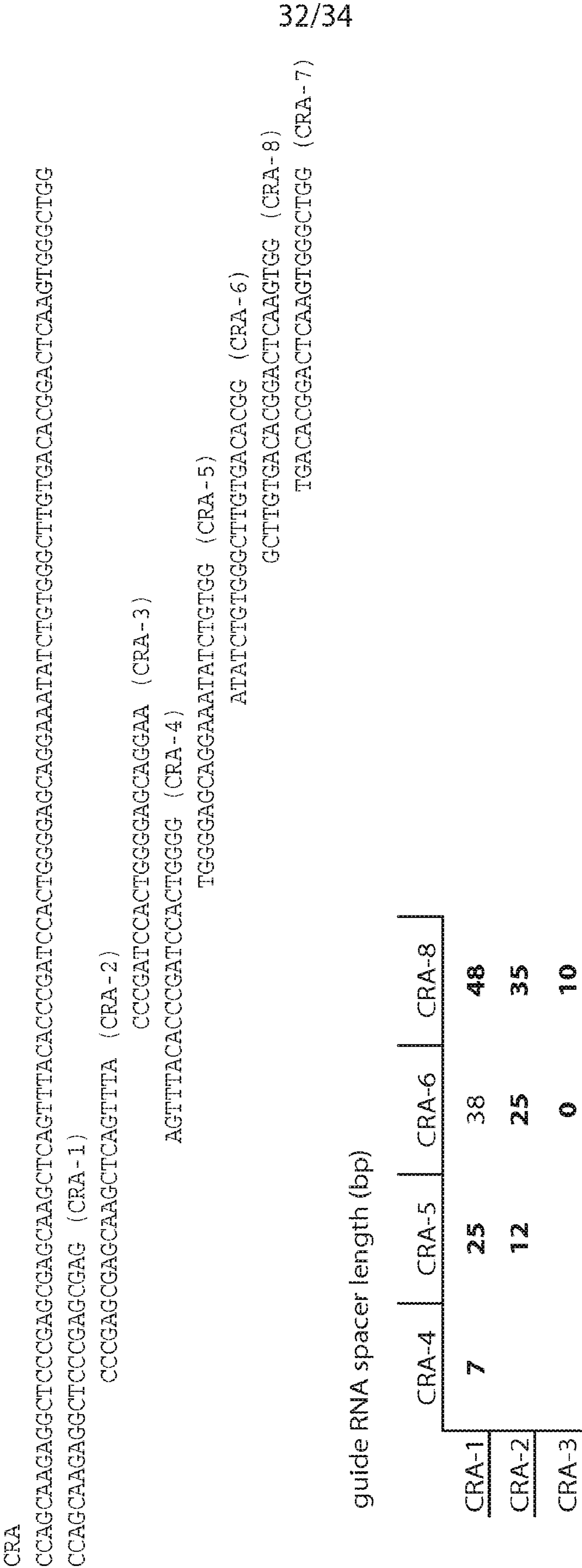


Fig. 19A

CRB

CCTGGCCATCTCTGACCTGTTTTTCCCTTCTTACTGTCCCTTCTGGGCTCACTATGCTGCCGCCAGTGGACTTTGGAATACAATGTGTCAACTCTTGACAGGGC
TCTATTTTATAGG

CCTGGCCATCTCTGACCTGTTTT (CB-1)

CCCCCTTCTGGGCTCACTATGCTG (CB-2)

CCGCCAGTGGGACTTTGGAAAT (CB-3)

TCACTATGCTGCCGCCAGTGGG (CB-4)

CAATGTGTCAACTCTTGACAGGG (CB-5)

TTGACAGGGCTCTATTTTATAGG (CB-6)

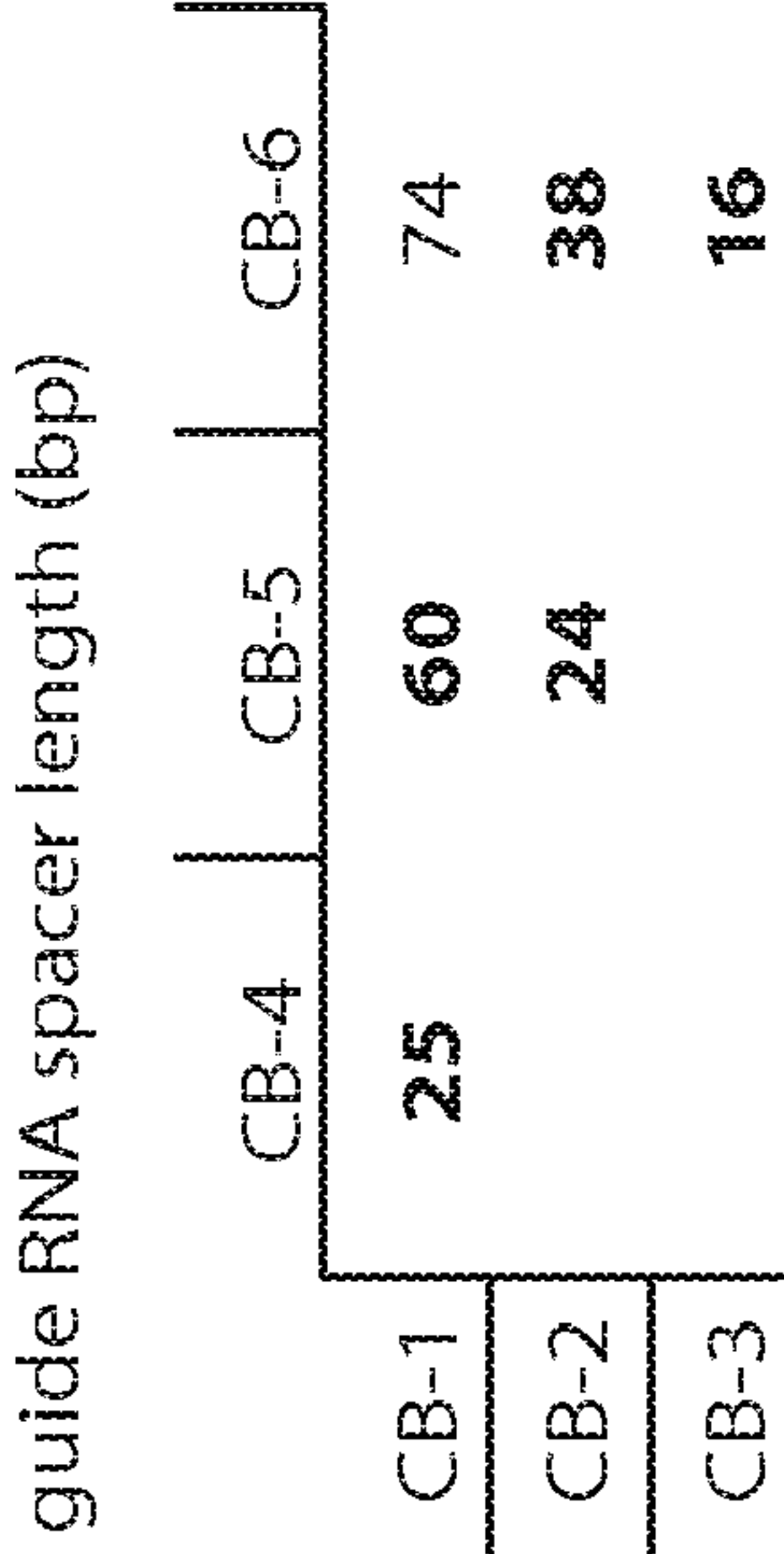


Fig. 19B

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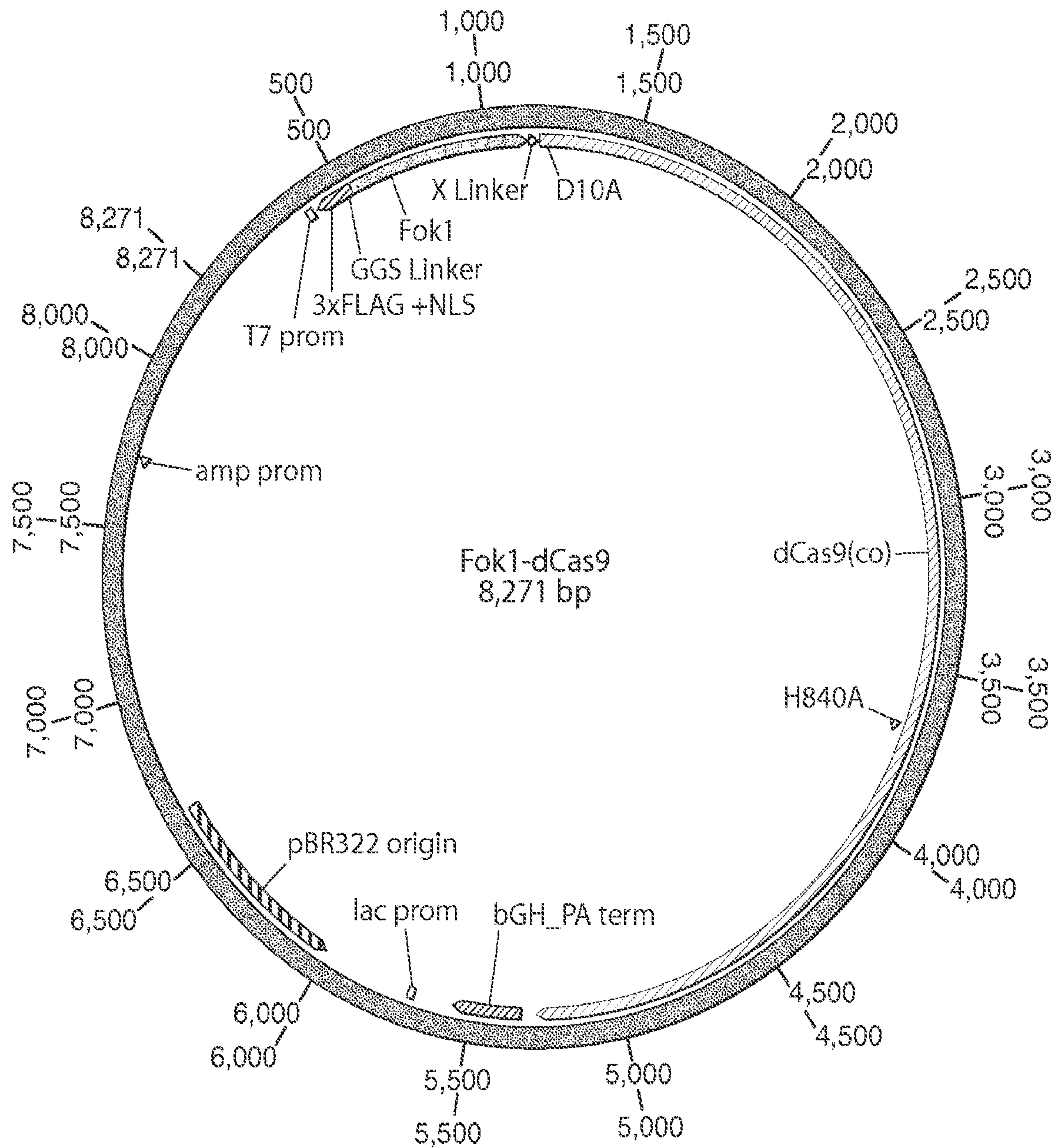


Fig. 20