



(86) Date de dépôt PCT/PCT Filing Date: 2000/03/09
(87) Date publication PCT/PCT Publication Date: 2000/09/21
(45) Date de délivrance/Issue Date: 2011/11/15
(85) Entrée phase nationale/National Entry: 2001/09/10
(86) N° demande PCT/PCT Application No.: CA 2000/000240
(87) N° publication PCT/PCT Publication No.: 2000/055326
(30) Priorité/Priority: 1999/03/12 (US60/123,966)

(51) Cl.Int./Int.Cl. *C12N 15/85* (2006.01),
A61K 39/118 (2006.01), *A61P 31/04* (2006.01),
C07K 14/295 (2006.01), *C07K 16/12* (2006.01),
C07K 19/00 (2006.01), *C12N 15/31* (2006.01),
C12N 15/62 (2006.01), *C12P 21/02* (2006.01),
C12Q 1/00 (2006.01), *G01N 33/569* (2006.01)

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(54) Titre : ANTIGENES (CHLAMYDIA), FRAGMENTS D'ADN CORRESPONDANTS, ET LEURS UTILISATIONS
(54) Title: CHLAMYDIA ANTIGENS AND CORRESPONDING DNA FRAGMENTS AND USES THEREOF

(57) **Abrégé/Abstract:**

The present invention provides a method of nucleic acid, including DNA, immunization of a host, including humans, against disease caused by infection by strain of Chlamydia, specifically *C. pneumoniae*, employing a vector containing a nucleotide sequence encoding a 60kDa cysteine-rich membrane protein of a strain of Chlamydia pneumoniae and a promoter to effect expression of the 60kDa cysteine-rich membrane protein gene in the host. Modifications are possible within the scope of this invention.



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**PCT**WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁷ : C12N 15/31, C07K 14/295, A61K 39/02, C07K 16/12, C12N 15/62, C12Q 1/04, 1/70	A1	(11) International Publication Number: WO 00/55326 (43) International Publication Date: 21 September 2000 (21.09.00)
(21) International Application Number: PCT/CA00/00240 (22) International Filing Date: 9 March 2000 (09.03.00) (30) Priority Data: 60/123,966 12 March 1999 (12.03.99) US (71) Applicant (for all designated States except US): AVENTIS PASTEUR LIMITED [CA/CA]; 1755 Steeles Avenue West, Toronto, Ontario M2R 3T4 (CA). (72) Inventors; and (75) Inventors/Applicants (for US only): MURDIN, Andrew, D. [CA/CA]; 146 Rhodes Circle, Newmarket, Ontario L3X 1V2 (CA). OOMEN, Raymond, P. [CA/CA]; 29 Kennedy Street W., Aurora, Ontario L4G 2L6 (CA). WANG, Joe [CA/CA]; 48 29th Street, Etobicoke, Ontario M8W 3A8 (CA). DUNN, Pamela [GB/CA]; Apartment 703, 370 Kaneff Crescent, Mississauga, Ontario L5A 4B8 (CA). (74) Agents: WHEELER, Michael, E. et al.; Smart & Biggar, 900-55 Metcalfe Street, P.O. Box 2999, Station D, Ottawa, Ontario K1P 5Y6 (CA).		(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i>
(54) Title: <i>CHLAMYDIA</i> ANTIGENS AND CORRESPONDING DNA FRAGMENTS AND USES THEREOF (57) Abstract The present invention provides a method of nucleic acid, including DNA, immunization of a host, including humans, against disease caused by infection by strain of <i>Chlamydia</i>, specifically <i>C. pneumoniae</i>, employing a vector containing a nucleotide sequence encoding a 60kDa cysteine-rich membrane protein of a strain of <i>Chlamydia pneumoniae</i> and a promoter to effect expression of the 60kDa cysteine-rich membrane protein gene in the host. Modifications are possible within the scope of this invention.		

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TITLE OF INVENTION

CHLAMYDIA ANTIGENS AND CORRESPONDING DNA FRAGMENTS
AND USES THEREOF

FIELD OF INVENTION

5 The present invention relates to the *Chlamydia*
60kDa cysteine-rich membrane protein and corresponding DNA
molecules, which can be used to prevent and treat *Chlamydia*
infection in mammals, such as humans.

BACKGROUND OF THE INVENTION

10 Chlamydiae are prokaryotes. They exhibit
morphologic and structural similarities to gram-negative
bacteria including a trilaminar outer membrane, which
contains lipopolysaccharide and several membrane proteins
that are structurally and functionally analogous to proteins
15 found in *E. coli*. They are obligate intra-cellular
parasites with a unique biphasic life cycle consisting of a
metabolically inactive but infectious extracellular stage
and a replicating but non-infectious intracellular stage.
The replicative stage of the life-cycle takes place within a
20 membrane-bound inclusion which sequesters the bacteria away
from the cytoplasm of the infected host cell.

C. pneumoniae is a common human pathogen,
originally described as the TWAR strain of *Chlamydia*
psittaci but subsequently recognised to be a new species.
25 *C. pneumoniae* is antigenically, genetically and
morphologically distinct from other chlamydia species
(*C. trachomatis*, *C. pecorum* and *C. psittaci*). It shows 10%
or less DNA sequence homology with either of *C. trachomatis*
or *C. psittaci*.

C. pneumoniae is a common cause of community acquired pneumonia, only less frequent than *Streptococcus pneumoniae* and *Mycoplasma pneumoniae* (Grayston et al. (1995) Journal of Infectious Diseases 168:1231; Campos et al. (1995) Investigation of Ophthalmology and Visual Science 36:1477). It can also cause upper respiratory tract symptoms and disease, including bronchitis and sinusitis (Grayston et al. (1995) Journal of Infectious Diseases 168:1231; Grayston et al (1990) Journal of Infectious Diseases 161:618; Marrie (1993) Clinical Infectious Diseases. 18:501; Wang et al (1986) Chlamydial infections Cambridge University Press, Cambridge. p. 329. The great majority of the adult population (over 60%) has antibodies to *C. pneumoniae* (Wang et al (1986) Chlamydial infections. Cambridge University Press, Cambridge. p. 329), indicating past infection which was unrecognized or asymptomatic.

C. pneumoniae infection usually presents as an acute respiratory disease (i.e., cough, sore throat, hoarseness, and fever; abnormal chest sounds on auscultation). For most patients, the cough persists for 2 to 6 weeks, and recovery is slow. In approximately 10% of these cases, upper respiratory tract infection is followed by bronchitis or pneumonia. Furthermore, during a *C. pneumoniae* epidemic, subsequent co-infection with pneumococcus has been noted in about half of these pneumonia patients, particularly in the infirm and the elderly. As noted above, there is more and more evidence that *C. pneumoniae* infection is also linked to diseases other than respiratory infections.

The reservoir for the organism is presumably people. In contrast to *C. psittaci* infections, there is no known bird or animal reservoir. Transmission has not been clearly defined. It may result from direct contact with secretions, from fomites, or from airborne spread. There is a long incubation period, which may last for many months. Based on analysis of epidemics, *C. pneumoniae* appears to spread slowly through a population

(case-to-case interval averaging 30 days) because infected persons are inefficient transmitters of the organism.

Susceptibility to *C. pneumoniae* is universal. Reinfections occur during adulthood, following the primary infection as a
5 child. *C. pneumoniae* appears to be an endemic disease throughout the world, noteworthy for superimposed intervals of increased incidence (epidemics) that persist for 2 to 3 years. *C. trachomatis* infection does not confer cross-immunity to *C. pneumoniae*. Infections are easily treated with oral
10 antibiotics, tetracycline or erythromycin (2 g/d, for at least 10 to 14 d). A recently developed drug, azithromycin, is highly effective as a single-dose therapy against chlamydial infections.

In most instances, *C. pneumoniae* infection is often
15 mild and without complications, and up to 90% of infections are subacute or unrecognized. Among children in industrialized countries, infections have been thought to be rare up to the age of 5 y, although a recent study (E Normann et al, Chlamydia pneumoniae in children with acute respiratory tract infections,
20 Acta Paediatrica, 1998, Vol 87, Iss 1, pp 23-27) has reported that many children in this age group show PCR evidence of infection despite being seronegative, and estimates a prevalence of 17-19% in 2-4 y olds. In developing countries, the seroprevalence of *C. pneumoniae* antibodies among young children
25 is elevated, and there are suspicions that *C. pneumoniae* may be an important cause of acute lower respiratory tract disease and mortality for infants and children in tropical regions of the world.

From seroprevalence studies and studies of local
30 epidemics, the initial *C. pneumoniae* infection usually happens between the ages of 5 and 20 y. In the USA, for example, there are estimated to be 30,000 cases of childhood pneumonia each year caused by *C. pneumoniae*. Infections may cluster among

groups of children or young adults (e.g., school pupils or military conscripts).

C. pneumoniae causes 10 to 25% of community-acquired lower respiratory tract infections (as reported from Sweden, Italy, Finland, and the USA). During an epidemic, *C. pneumonia* infection may account for 50 to 60% of the cases of pneumonia. During these periods, also, more episodes of mixed infections with *S. pneumoniae* have been reported.

Reinfection during adulthood is common; the clinical presentation tends to be milder. Based on population seroprevalence studies, there tends to be increased exposure with age, which is particularly evident among men. Some investigators have speculated that a persistent, asymptomatic *C. pneumoniae* infection state is common.

In adults of middle age or older, *C. pneumoniae* infection may progress to chronic bronchitis and sinusitis. A study in the USA revealed that the incidence of pneumonia caused by *C. pneumoniae* in persons younger than 60 years is 1 case per 1,000 persons per year; but in the elderly, the disease incidence rose three-fold. *C. pneumoniae* infection rarely leads to hospitalization, except in patients with an underlying illness.

Of considerable importance is the association of atherosclerosis and *C. pneumoniae* infection. There are several epidemiological studies showing a correlation of previous infections with *C. pneumoniae* and heart attacks, coronary artery and carotid artery disease (Saikku et al. (1988) Lancet;ii:983; Thom et al. (1992) JAMA 268:68; Linnanmaki et al. (1993), Circulation 87:1030; Saikku et al. (1992) Annals Internal Medicine 116:273; Melnick et al (1993) American Journal of Medicine 95:499). Moreover, the organisms has been detected in atheromas and fatty streaks of the coronary, carotid, peripheral arteries and aorta (Shor et al. (1992) South African. Medical Journal 82:158; Kuo et al. (1993) Journal of Infectious Diseases

167:841; Kuo et al. (1993) Arteriosclerosis and Thrombosis
13:1500; Campbell et al (1995) Journal of Infectious Diseases
172:585; Chiu et al. Circulation, 1997 (In Press)). Viable
C. pneumoniae has been recovered from the coronary and carotid
5 artery (Ramirez et al (1996) Annals of Internal Medicine
125:979; Jackson et al. Abst. K121, p272, 36th ICAAC, 15-18
Sept. 1996, New Orleans). Furthermore, it has been shown that
C. pneumoniae can induce changes of atherosclerosis in a rabbit
model (Fong et al (1997) Journal of Clinical Microbiology
10 35:48). Taken together, these results indicate that it is
highly probable that *C. pneumoniae* can cause atherosclerosis in
humans, though the epidemiological importance of chlamydial
atherosclerosis remains to be demonstrated.

A number of recent studies have also indicated an
15 association between *C. pneumoniae* infection and asthma.
Infection has been linked to wheezing, asthmatic bronchitis,
adult-onset asthma and acute exacerbations of asthma in adults,
and small-scale studies have shown that prolonged antibiotic
treatment was effective at greatly reducing the severity of the
20 disease in some individuals (Hahn DL, et al. Evidence for
Chlamydia pneumoniae infection in steroid-dependent asthma. Ann
Allergy Asthma Immunol. 1998 Jan; 80(1): 45-49.; Hahn DL, et al.
Association of *Chlamydia pneumoniae* IgA antibodies with recently
symptomatic asthma. Epidemiol Infect. 1996 Dec; 117(3): 513-
25 517; Bjornsson E, et al. Serology of chlamydia in relation to
asthma and bronchial hyperresponsiveness. Scand J Infect Dis.
1996; 28(1): 63-69.; Hahn DL. Treatment of *Chlamydia pneumoniae*
infection in adult asthma: a before-after trial. J Fam Pract.
1995 Oct; 41(4): 345-351.; Allegra L, et al. Acute exacerbations
30 of asthma in adults: role of *Chlamydia pneumoniae* infection.
Eur Respir J. 1994 Dec; 7(12): 2165-2168.; Hahn DL, et al.
Association of *Chlamydia pneumoniae* (strain TWAR) infection with
wheezing, asthmatic bronchitis, and adult-onset asthma. JAMA.
1991 Jul 10; 266(2): 225-230).

In light of these results a protective vaccine against *C. pneumoniae* infection would be of considerable importance. There is not yet an effective vaccine for any human chlamydial infection. It is conceivable that an effective vaccine can be developed using physically or chemically inactivated Chlamydiae. However, such a vaccine does not have a high margin of safety. In general, safer vaccines are made by genetically manipulating the organism by attenuation or by recombinant means. Accordingly, a major obstacle in creating an effective and safe vaccine against human chlamydial infection has been the paucity of genetic information regarding Chlamydia, specifically *C. pneumoniae*.

Studies with *C. trachomatis* and *C. psittaci* indicate that safe and effective vaccine against Chlamydia is an attainable goal. For example, mice which have recovered from a lung infection with *C. trachomatis* are protected from infertility induced by a subsequent vaginal challenge (Pal et al. (1996) Infection and Immunity.64:5341). Similarly, sheep immunized with inactivated *C. psittaci* were protected from subsequent chlamydial-induced abortions and stillbirths (Jones et al. (1995) Vaccine 13:715). Protection from chlamydial infections has been associated with Th1 immune responses, particularly the induction of INF γ - producing CD4+T-cells (Igietsemes et al. (1993) Immunology 5:317). The adoptive transfer of CD4+ cell lines or clones to nude or SCID mice conferred protection from challenge or cleared chronic disease (Igietseme et al (1993) Regional Immunology 5:317; Magee et al (1993) Regional Immunology 5: 305), and *in vivo* depletion of CD4+ T cells exacerbated disease post-challenge (Landers et al (1991) Infection & Immunity 59:3774; Magee et al (1995) Infection & Immunity 63:516). However, the presence of sufficiently high titres of neutralising antibody at mucosal surfaces can also exert a protective effect (Cotter et al. (1995) Infection and Immunity 63:4704).

Antigenic variation within the species *C. pneumoniae* is not well documented due to insufficient genetic information, though variation is expected to exist based on *C. trachomatis*. Serovars of *C. trachomatis* are defined on the basis of antigenic variation in the major outer membrane protein (MOMP), but published *C. pneumoniae* MOMP gene sequences show no variation between several diverse isolates of the organism (Campbell et al (1990) Infection and Immunity 58:93; McCafferty et al (1995) Infection and Immunity 63:2387-9; Knudsen et al (1996) Third Meeting of the European Society for Chlamydia Research, Vienna). The gene encoding a 76 kDa antigen has been cloned from a single strain of *C. pneumoniae* and the sequence published (Perez Melgosa et al., Infect. Immun. 1994. 62:880). An operon encoding the 9 kDa and 60 kDa cyteine-rich outer membrane protein genes has been described (Watson et al., Nucleic Acids Res (1990) 18:5299; Watson et al., Microbiology (1995) 141:2489). Many antigens recognized by immune sera to *C. pneumoniae* are conserved across all chlamydiae, but 98 kDa, 76 kDa and several other proteins may be *C. pneumoniae*-specific (Perez Melgosa et al., Infect. Immun. 1994. 62:880; Melgosa et al., FEMS Microbiol Lett (1993) 112 :199;, Campbell et al., J Clin Microbiol (1990) 28 :1261; Iijima et al., J Clin Microbiol (1994) 32:583). An assessment of the number and relative frequency of any *C. pneumoniae* serotypes, and the defining antigens, is not yet possible. The entire genome sequence of *C. pneumoniae* strain CWL-029 is now known (<http://chlamydia-www.berkeley.edu:4231/>) and as further sequences become available a better understanding of antigenic variation may be gained.

Many antigens recognised by immune sera to *C. pneumoniae* are conserved across all chlamydiae, but 98kDa, 76 kDa and 54 kDa proteins appear to be *C. pneumoniae*-specific (Campos et al. (1995) Investigation of Ophthalmology and Visual Science 36:1477; Marrie (1993) Clinical Infectious Diseases.

18:501; Wiedmann-Al-Ahmad M, et al. Reactions of polyclonal and neutralizing anti-p54 monoclonal antibodies with an isolated, species-specific 54-kilodalton protein of *Chlamydia pneumoniae*. Clin Diagn Lab Immunol. 1997 Nov; 4(6): 700-704).

5 Immunoblotting of isolates with sera from patients does show variation of blotting patterns between isolates, indicating that serotypes *C. pneumoniae* may exist (Grayston et al. (1995) Journal of Infectious Diseases 168:1231; Ramirez et al (1996) Annals of Internal Medicine 125:979). However, the
10 results are potentially confounded by the infection status of the patients, since immunoblot profiles of a patient's sera change with time post-infection. An assessment of the number and relative frequency of any serotypes, and the defining antigens, is not yet possible.

15 Accordingly, a need exists for identifying and isolating polynucleotide sequences of *C. pneumoniae* for use in preventing and treating Chlamydia infection.

SUMMARY OF THE INVENTION

20 The present invention provides purified and isolated polynucleotide molecules that encode the *Chlamydia* polypeptides designated 60kDa cysteine-rich membrane proteine (SEQ ID No: 1) which can be used in methods to prevent, treat, and diagnose *Chlamydia* infection. In one form of the invention, the
25 polynucleotide molecules are DNA that encode the polypeptide of SEQ ID No: 2.

Another form of the invention provides polypeptides corresponding to the isolated DNA molecules. The amino acid sequence of the corresponding encoded polypeptide is shown as
30 SEQ ID No: 2.

Those skilled in the art will readily understand that the invention, having provided the polynucleotide sequences encoding the *Chlamydia* 60kDa cysteine-rich membrane protein, also provides polynucleotides encoding fragments derived from

such a polypeptide. Moreover, the invention is understood to provide mutants and derivatives of such polypeptides and fragments derived therefrom, which result from the addition, deletion, or substitution of non-essential amino acids as
5 described herein. Those skilled in the art would also readily understand that the invention, having provided the polynucleotide sequences encoding *Chlamydia* polypeptides, further provides monospecific antibodies that specifically bind to such polypeptides.

10 The present invention has wide application and includes expression cassettes, vectors, and cells transformed or transfected with the polynucleotides of the invention. Accordingly, the present invention further provides (i) a method for producing a polypeptide of the invention in a recombinant
15 host system and related expression cassettes, vectors, and transformed or transfected cells; (ii) a vaccine, or a live vaccine vector such as a pox virus, *Salmonella typhimurium*, or *Vibrio cholerae* vector, containing a polynucleotide of the invention, such vaccines and vaccine vectors being useful for,
20 e.g., preventing and treating *Chlamydia* infection, in combination with a diluent or carrier, and related pharmaceutical compositions and associated therapeutic and/or prophylactic methods; (iii) a therapeutic and/or prophylactic use of an RNA or DNA molecule of the invention, either in a
25 naked form or formulated with a delivery vehicle, a polypeptide or combination of polypeptides, or a monospecific antibody of the invention, and related pharmaceutical compositions; (iv) a method for diagnosing the presence of *Chlamydia* in a biological sample, which can involve the use of a DNA or RNA molecule, a
30 monospecific antibody, or a polypeptide of the invention; and (v) a method for purifying a polypeptide of the invention by antibody-based affinity chromatography.

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One aspect of the invention relates a vaccine vector comprising an isolated nucleic acid molecule operably linked to a promoter functional in a mammalian cell, wherein the nucleic acid molecule encodes a polypeptide is selected
5 from any of: (a) SEQ ID No:2; (b) an immunogenic fragment consisting of at least 50 consecutive amino acids from SEQ ID No:2; and (c) a polypeptide which has been modified without loss of immunogenicity, wherein said modified polypeptide is at least 90% identical in amino acid sequence
10 over the full length of SEQ ID No:2; wherein the polypeptide, when used to immunize an animal, confers immunoprotection to the animal against Chlamydia.

Another aspect of the invention relates to a vaccine composition comprising a protein and a compound
15 pharmaceutically acceptable for use in a vaccine, wherein the protein comprises the amino acid sequence selected from any of: (a) SEQ ID No:2; (b) an immunogenic fragment consisting of at least 50 consecutive amino acids from SEQ ID No:2; and (c) a polypeptide which has been modified
20 without loss of immunogenicity, wherein said modified polypeptide is at least 90% identical in amino acid sequence over the full length of SEQ ID No:2; wherein the amino acid sequence, when used to immunize an animal, confers immunoprotection to the animal against Chlamydia, and
25 wherein the compound facilitates delivery and/or enhance an immune response to the part of the protein having the amino acid sequence.

Another aspect of the invention relates to use of a nucleic acid molecule encoding an amino acid sequence, or
30 of a protein comprising the amino acid sequence, in the manufacture of a vaccine against Chlamydia, wherein the

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amino acid sequence is selected from any of: (a) SEQ ID No:2; (b) an immunogenic fragment consisting of at least 50 consecutive amino acids from SEQ ID No:2; and (c) a polypeptide which has been modified without loss of immunogenicity, wherein said modified polypeptide is at least 90% identical in amino acid sequence over the full length of SEQ ID No:2; wherein the amino acid sequence, when used to immunize an animal, confers immunoprotection to the animal against Chlamydia.

10 Another aspect of the invention relates to a vaccination kit comprising the vaccine vector as disclosed herein in unit dosage form and instructions for its use in vaccinating a subject against Chlamydia infection.

15 Another aspect of the invention relates to a vaccination kit comprising the vaccine composition as disclosed herein in unit dosage form and instructions for its use in vaccinating a subject against Chlamydia infection.

BRIEF DESCRIPTION OF THE DRAWINGS

The present invention will be further understood from the following description with reference to the drawings, in which:

5 Figure 1 shows the nucleotide sequence of the 60kDa cysteine-rich membrane protein gene (SEQ ID No: 1) and the deduced amino acid sequence of the 60kDa cysteine-rich membrane protein from *Chlamydia pneumoniae* (SEQ ID No: 2).

10 Figure 2 shows the restriction enzyme analysis of the *C. pneumoniae* 60kDa cysteine-rich membrane protein gene.

 Figure 3 shows the construction and elements of plasmid pCACRMP60.

 Figure 4 illustrates protection against *C. pneumoniae* infection by pCACRMP60 following DNA immunization.

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DETAILED DESCRIPTION OF INVENTION

 An open reading frame (ORF) encoding the chlamydial 60kDa cysteine-rich membrane protein has been identified from the *C. pneumoniae* genome. The gene encoding this protein has
20 been inserted into an expression plasmid and shown to confer immune protection against chlamydial infection. Accordingly, this 60kDa cysteine-rich membrane protein and related polypeptides can be used to prevent and treat *Chlamydia* infection.

25 According to a first aspect of the invention, isolated polynucleotides are provided which encode *Chlamydia* polypeptides, whose amino acid sequences are shown in SEQ ID No: 2.

 The term "isolated polynucleotide" is defined as a
30 polynucleotide removed from the environment in which it naturally occurs. For example, a naturally-occurring DNA molecule present in the genome of a living bacteria or as part of a gene bank is not isolated, but the same molecule separated from the remaining part of the bacterial genome, as a result of,

e.g., a cloning event (amplification), is isolated. Typically, an isolated DNA molecule is free from DNA regions (e.g., coding regions) with which it is immediately contiguous at the 5' or 3' end, in the naturally occurring genome. Such isolated
5 polynucleotides may be part of a vector or a composition and still be defined as isolated in that such a vector or composition is not part of the natural environment of such polynucleotide.

The polynucleotide of the invention is either RNA or
10 DNA (cDNA, genomic DNA, or synthetic DNA), or modifications, variants, homologs or fragments thereof. The DNA is either double-stranded or single-stranded, and, if single-stranded, is either the coding strand or the non-coding (anti-sense) strand. Any one of the sequences that encode the polypeptides of the
15 invention as shown in SEQ ID No: 1 is (a) a coding sequence, (b) a ribonucleotide sequence derived from transcription of (a), or (c) a coding sequence which uses the redundancy or degeneracy of the genetic code to encode the same polypeptides. By "polypeptide" or "protein" is meant any chain of amino acids,
20 regardless of length or post-translational modification (e.g., glycosylation or phosphorylation). Both terms are used interchangeably in the present application.

Consistent with the first aspect of the invention, amino acid sequences are provided which are homologous to SEQ ID
25 No: 2. As used herein, "homologous amino acid sequence" is any polypeptide which is encoded, in whole or in part, by a nucleic acid sequence which hybridizes at 25-35°C below critical melting temperature (T_m), to any portion of the nucleic acid sequence of SEQ ID No: 1. A homologous amino acid sequence is one that
30 differs from an amino acid sequence shown in SEQ ID No: 2 by one or more conservative amino acid substitutions. Such a sequence also encompass serotypic variants (defined below) as well as sequences containing deletions or insertions which retain inherent characteristics of the polypeptide such as

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immunogenicity. Preferably, such a sequence is at least 75%, more preferably 80%, and most preferably 90% identical to SEQ ID No: 2.

Homologous amino acid sequences include sequences that are identical or substantially identical to SEQ ID No: 2. By "amino acid sequence substantially identical" is meant a sequence that is at least 90%, preferably 95%, more preferably 97%, and most preferably 99% identical to an amino acid sequence of reference and that preferably differs from the sequence of reference by a majority of conservative amino acid substitutions.

Conservative amino acid substitutions are substitutions among amino acids of the same class. These classes include, for example, amino acids having uncharged polar side chains, such as asparagine, glutamine, serine, threonine, and tyrosine; amino acids having basic side chains, such as lysine, arginine, and histidine; amino acids having acidic side chains, such as aspartic acid and glutamic acid; and amino acids having nonpolar side chains, such as glycine, alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan, and cysteine.

Homology is measured using sequence analysis software such as Sequence Analysis Software Package of the Genetics Computer Group, University of Wisconsin Biotechnology Center, 1710 University Avenue, Madison, WI 53705. Amino acid sequences are aligned to maximize identity. Gaps may be artificially introduced into the sequence to attain proper alignment. Once the optimal alignment has been set up, the degree of homology is established by recording all of the positions in which the amino acids of both sequences are identical, relative to the total number of positions.

Homologous polynucleotide sequences are defined in a similar way. Preferably, a homologous sequence is one that is

at least 45%, more preferably 60%, and most preferably 85% identical to the coding sequence of SEQ ID No: 1.

Consistent with the first aspect of the invention, polypeptides having a sequence homologous to SEQ ID No: 2 include naturally-occurring allelic variants, as well as mutants or any other non-naturally occurring variants that retain the inherent characteristics of the polypeptide of SEQ ID No: 2.

As is known in the art, an allelic variant is an alternate form of a polypeptide that is characterized as having a substitution, deletion, or addition of one or more amino acids that does not alter the biological function of the polypeptide. By "biological function" is meant the function of the polypeptide in the cells in which it naturally occurs, even if the function is not necessary for the growth or survival of the cells. For example, the biological function of a porin is to allow the entry into cells of compounds present in the extracellular medium. Biological function is distinct from antigenic property. A polypeptide can have more than one biological function.

Allelic variants are very common in nature. For example, a bacterial species such as *C. pneumoniae*, is usually represented by a variety of strains that differ from each other by minor allelic variations. Indeed, a polypeptide that fulfills the same biological function in different strains can have an amino acid sequence (and polynucleotide sequence) that is not identical in each of the strains. Despite this variation, an immune response directed generally against many allelic variants has been demonstrated. In studies of the *Chlamydial* MOMP antigen, cross-strain antibody binding plus neutralization of infectivity occurs despite amino acid sequence variation of MOMP from strain to strain, indicating that the MOMP, when used as an immunogen, is tolerant of amino acid variations.

Polynucleotides encoding homologous polypeptides or allelic variants are retrieved by polymerase chain reaction (PCR) amplification of genomic bacterial DNA extracted by conventional methods. This involves the use of synthetic oligonucleotide primers matching upstream and downstream of the 5' and 3' ends of the encoding domain. Suitable primers are designed according to the nucleotide sequence information provided in SEQ ID No:1. The procedure is as follows: a primer is selected which consists of 10 to 40, preferably 15 to 25 nucleotides. It is advantageous to select primers containing C and G nucleotides in a proportion sufficient to ensure efficient hybridization; *i.e.*, an amount of C and G nucleotides of at least 40%, preferably 50% of the total nucleotide content. A standard PCR reaction contains typically 0.5 to 5 Units of Taq DNA polymerase per 100 μ L, 20 to 200 μ M deoxynucleotide each, preferably at equivalent concentrations, 0.5 to 2.5 mM magnesium over the total deoxynucleotide concentration, 10^5 to 10^6 target molecules, and about 20 pmol of each primer. About 25 to 50 PCR cycles are performed, with an annealing temperature 15°C to 5°C below the true T_m of the primers. A more stringent annealing temperature improves discrimination against incorrectly annealed primers and reduces incorporation of incorrect nucleotides at the 3' end of primers. A denaturation temperature of 95°C to 97°C is typical, although higher temperatures may be appropriate for dematuration of G+C-rich targets. The number of cycles performed depends on the starting concentration of target molecules, though typically more than 40 cycles is not recommended as non-specific background products tend to accumulate.

30 An alternative method for retrieving polynucleotides encoding homologous polypeptides or allelic variants is by hybridization screening of a DNA or RNA library. Hybridization procedures are well-known in the art and are described in

Ausubel et al., (Ausubel et al., Current Protocols in Molecular Biology, John Wiley & Sons Inc., 1994), Silhavy et al. (Silhavy et al. Experiments with Gene Fusions, Cold Spring Harbor Laboratory Press, 1984), and Davis et al. (Davis et al. A Manual
5 for Genetic Engineering: Advanced Bacterial Genetics, Cold Spring Harbor Laboratory Press, 1980)). Important parameters for optimizing hybridization conditions are reflected in a formula used to obtain the critical melting temperature above which two complementary DNA strands separate from each other
10 (Casey & Davidson, Nucl. Acid Res. (1977) 4:1539). For polynucleotides of about 600 nucleotides or larger, this formula is as follows: $T_m = 81.5 + 0.41 \times (\% \text{ G+C}) + 16.6 \log (\text{cation ion concentration}) - 0.63 \times (\% \text{ formamide}) - 600/\text{base number}$. Under appropriate stringency conditions, hybridization temperature
15 (T_h) is approximately 20 to 40°C, 20 to 25°C, or, preferably 30 to 40°C below the calculated T_m . Those skilled in the art will understand that optimal temperature and salt conditions can be readily determined.

For the polynucleotides of the invention, stringent
20 conditions are achieved for both pre-hybridizing and hybridizing incubations (i) within 4-16 hours at 42°C, in 6 x SSC containing 50% formamide, or (ii) within 4-16 hours at 65°C in an aqueous 6 x SSC solution (1 M NaCl, 0.1 M sodium citrate (pH 7.0)). Typically, hybridization experiments are performed at a
25 temperature from 60 to 68°C, e.g. 65°C. At such a temperature, stringent hybridization conditions can be achieved in 6xSSC, preferably in 2xSSC or 1xSSC, more preferably in 0.5xSSC, 0.3xSSC or 0.1xSSC (in the absence of formamide). 1xSSC contains 0.15 M NaCl and 0.015 M sodium citrate.

30 Useful homologs and fragments thereof that do not occur naturally are designed using known methods for identifying regions of an antigen that are likely to tolerate amino acid sequence changes and/or deletions. As an example, homologous polypeptides from different species are compared; conserved

sequences are identified. The more divergent sequences are the most likely to tolerate sequence changes. Homology among sequences may be analyzed using, as an example, the BLAST homology searching algorithm of Altschul et al., Nucleic Acids Res.; 25:3389-3402 (1997). Alternatively, sequences are modified such that they become more reactive to T- and/or B-cells, based on computer-assisted analysis of probable T- or B-cell epitopes. Yet another alternative is to mutate a particular amino acid residue or sequence within the polypeptide *in vitro*, then screen the mutant polypeptides for their ability to prevent or treat Chlamydia infection according to the method outlined below.

A person skilled in the art will readily understand that by following the screening process of this invention, it will be determined without undue experimentation whether a particular homolog of SEQ ID No. 2 may be useful in the prevention or treatment of Chlamydia infection. The screening procedure comprises the steps:

- (i) immunizing an animal, preferably mouse, with the test homolog or fragment;
- (ii) inoculating the immunized animal with Chlamydia; and
- (iii) selecting those homologs or fragments which confer protection against Chlamydia.

By "conferring protection" is meant that there is a reduction in severity of any of the effects of Chlamydia infection, in comparison with a control animal which was not immunized with the test homolog or fragment.

Consistent with the first aspect of the invention, polypeptide derivatives are provided that are partial sequences of SEQ ID No. 2, partial sequences of polypeptide sequences homologous to SEQ ID No. 2, polypeptides derived from full-length polypeptides by internal deletion, and fusion proteins.

It is an accepted practice in the field of immunology

to use fragments and variants of protein immunogens as vaccines, as all that is required to induce an immune response to a protein is a small (e.g., 8 to 10 amino acid) immunogenic region of the protein. Various short synthetic peptides corresponding
5 to surface-exposed antigens of pathogens other than *Chlamydia* have been shown to be effective vaccine antigens against their respective pathogens, e.g. an 11 residue peptide of murine mammary tumor virus (Casey & Davidson, Nucl. Acid Res. (1977) 4:1539), a 16-residue peptide of Semliki Forest virus (Snijders
10 et al., 1991. J. Gen. Virol. 72:557-565), and two overlapping peptides of 15 residues each from canine parvovirus (Langeveld et al., Vaccine 12(15):1473-1480, 1994).

Accordingly, it will be readily apparent to one skilled in the art, having read the present description, that
15 partial sequences of SEQ ID No: 2 or their homologous amino acid sequences are inherent to the full-length sequences and are taught by the present invention. Such polypeptide fragments preferably are at least 12 amino acids in length. Advantageously, they are at least 20 amino acids, preferably at
20 least 50 amino acids, more preferably at least 75 amino acids, and most preferably at least 100 amino acids in length.

Polynucleotides of 30 to 600 nucleotides encoding partial sequences of sequences homologous to SEQ ID No: 2 are retrieved by PCR amplification using the parameters outlined
25 above and using primers matching the sequences upstream and downstream of the 5' and 3' ends of the fragment to be amplified. The template polynucleotide for such amplification is either the full length polynucleotide homologous to SEQ ID No: 1, or a polynucleotide contained in a mixture of
30 polynucleotides such as a DNA or RNA library. As an alternative method for retrieving the partial sequences, screening hybridization is carried out under conditions described above and using the formula for calculating T_m . Where fragments of 30 to 600 nucleotides are to be retrieved, the calculated T_m is

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corrected by subtracting (600/polynucleotide size in base pairs) and the stringency conditions are defined by a hybridization temperature that is 5 to 10°C below T_m . Where oligonucleotides shorter than 20-30 bases are to be obtained, the formula for calculating the T_m is as follows: $T_m = 4 \times (G+C) + 2 \times (A+T)$. For example, an 18 nucleotide fragment of 50% G+C would have an approximate T_m of 54°C. Short peptides that are fragments of SEQ ID No: 2 or its homologous sequences, are obtained directly by chemical synthesis (E. Gross and H. J. Meinhofer, 4 The Peptides: Analysis, Synthesis, Biology; Modern Techniques of Peptide Synthesis, John Wiley & Sons (1981), and M. Bodanzki, Principles of Peptide Synthesis, Springer -Verlag (1984)).

Useful polypeptide derivatives, e.g., polypeptide fragments, are designed using computer-assisted analysis of amino acid sequences. This would identify probable surface-exposed, antigenic regions (Hughes et al., 1992. Infect. Immun. 60(9):3497). Analysis of 6 amino acid sequences contained in SEQ ID No: 2, based on the product of flexibility and hydrophobicity propensities using the program SEQSEE (Wishart DS, et al. "SEQSEE: a comprehensive program suite for protein sequence analysis." *Comput Appl Biosci*. 1994 Apr;10(2):121-32), can reveal potential B- and T-cell epitopes which may be used as a basis for selecting useful immunogenic fragments and variants. This analysis uses a reasonable combination of external surface features that is likely to be recognized by antibodies. Probable T-cell epitopes for HLA-A0201 MHC subclass may be revealed by an algorithm that emulate an approach developed at the NIH (Parker KC, et al. "Peptide binding to MHC class I molecules: implications for antigenic peptide prediction." *Immunol Res* 1995;14(1):34-57).

Epitopes which induce a protective T cell-dependent immune response are present throughout the length of the polypeptide. However, some epitopes may be masked by secondary and tertiary structures of the polypeptide. To reveal such

masked epitopes large internal deletions are created which remove much of the original protein structure and exposes the masked epitopes. Such internal deletions sometimes effect the additional advantage of removing immunodominant regions of high
5 variability among strains.

Polynucleotides encoding polypeptide fragments and polypeptides having large internal deletions are constructed using standard methods (Ausubel et al., Current Protocols in Molecular Biology, John Wiley & Sons Inc., 1994). Such methods
10 include standard PCR, inverse PCR, restriction enzyme treatment of cloned DNA molecules, or the method of Kunkel et al. (Kunkel et al. Proc. Natl. Acad. Sci. USA (1985) 82:448). Components for these methods and instructions for their use are readily available from various commercial sources such as
15 Stratagene. Once the deletion mutants have been constructed, they are tested for their ability to prevent or treat Chlamydia infection as described above.

As used herein, a fusion polypeptide is one that contains a polypeptide or a polypeptide derivative of the
20 invention fused at the N- or C-terminal end to any other polypeptide (hereinafter referred to as a peptide tail). A simple way to obtain such a fusion polypeptide is by translation of an in-frame fusion of the polynucleotide sequences, i.e., a hybrid gene. The hybrid gene encoding the fusion polypeptide is
25 inserted into an expression vector which is used to transform or transfect a host cell. Alternatively, the polynucleotide sequence encoding the polypeptide or polypeptide derivative is inserted into an expression vector in which the polynucleotide encoding the peptide tail is already present. Such vectors and
30 instructions for their use are commercially available, e.g. the pMal-c2 or pMal-p2 system from New England Biolabs, in which the peptide tail is a maltose binding protein, the glutathione-S-transferase system of Pharmacia, or the His-Tag system available from Novagen. These and other expression systems provide

convenient means for further purification of polypeptides and derivatives of the invention.

An advantageous example of a fusion polypeptide is one where the polypeptide or homolog or fragment of the invention is fused to a polypeptide having adjuvant activity, such as subunit B of either cholera toxin or *E. coli* heat-labile toxin. Another advantageous fusion is one where the polypeptide, homolog or fragment is fused to a strong T-cell epitope or B-cell epitope. Such an epitope may be one known in the art (e.g. the Hepatitis B virus core antigen, D.R. Millich et al., "Antibody production to the nucleocapsid and envelope of the Hepatitis B virus primed by a single synthetic T cell site", Nature. 1987. 329:547-549), or one which has been identified in another polypeptide of the invention based on computer-assisted analysis of probable T- or B-cell epitopes. Consistent with this aspect of the invention is a fusion polypeptide comprising T- or B-cell epitopes from SEQ ID No: 2 or its homolog or fragment, wherein the epitopes are derived from multiple variants of said polypeptide or homolog or fragment, each variant differing from another in the location and sequence of its epitope within the polypeptide. Such a fusion is effective in the prevention and treatment of Chlamydia infection since it optimizes the T- and B-cell response to the overall polypeptide, homolog or fragment.

To effect fusion, the polypeptide of the invention is fused to the N-, or preferably, to the C-terminal end of the polypeptide having adjuvant activity or T- or B-cell epitope. Alternatively, a polypeptide fragment of the invention is inserted internally within the amino acid sequence of the polypeptide having adjuvant activity. The T- or B-cell epitope may also be inserted internally within the amino acid sequence of the polypeptide of the invention.

Consistent with the first aspect, the polynucleotides of the invention also encode hybrid precursor polypeptides containing heterologous signal peptides, which mature into

polypeptides of the invention. By "heterologous signal peptide" is meant a signal peptide that is not found in naturally-occurring precursors of polypeptides of the invention.

Polynucleotide molecules according to the invention, including RNA, DNA, or modifications or combinations thereof, have various applications. A DNA molecule is used, for example, (i) in a process for producing the encoded polypeptide in a recombinant host system, (ii) in the construction of vaccine vectors such as poxviruses, which are further used in methods and compositions for preventing and/or treating *Chlamydia* infection, (iii) as a vaccine agent (as well as an RNA molecule), in a naked form or formulated with a delivery vehicle and, (iv) in the construction of attenuated *Chlamydia* strains that can over-express a polynucleotide of the invention or express it in a non-toxic, mutated form.

Accordingly, a second aspect of the invention encompasses (i) an expression cassette containing a DNA molecule of the invention placed under the control of the elements required for expression, in particular under the control of an appropriate promoter; (ii) an expression vector containing an expression cassette of the invention; (iii) a procaryotic or eucaryotic cell transformed or transfected with an expression cassette and/or vector of the invention, as well as (iv) a process for producing a polypeptide or polypeptide derivative encoded by a polynucleotide of the invention, which involves culturing a procaryotic or eucaryotic cell transformed or transfected with an expression cassette and/or vector of the invention, under conditions that allow expression of the DNA molecule of the invention and, recovering the encoded polypeptide or polypeptide derivative from the cell culture.

A recombinant expression system is selected from procaryotic and eucaryotic hosts. Eucaryotic hosts include yeast cells (e.g., *Saccharomyces cerevisiae* or *Pichia pastoris*), mammalian cells (e.g., COS1, NIH3T3, or JEG3 cells), arthropods

cells (e.g., *Spodoptera frugiperda* (SF9) cells), and plant cells. A preferred expression system is a procaryotic host such as *E. coli*. Bacterial and eucaryotic cells are available from a number of different sources including commercial sources to
5 those skilled in the art, e.g., the American Type Culture Collection (ATCC; Rockville, Maryland). Commercial sources of cells used for recombinant protein expression also provide instructions for usage of the cells.

The choice of the expression system depends on the
10 features desired for the expressed polypeptide. For example, it may be useful to produce a polypeptide of the invention in a particular lipidated form or any other form.

One skilled in the art would readily understand that not all vectors and expression control sequences and hosts would
15 be expected to express equally well the polynucleotides of this invention. With the guidelines described below, however, a selection of vectors, expression control sequences and hosts may be made without undue experimentation and without departing from the scope of this invention.

20 In selecting a vector, the host must be chosen that is compatible with the vector which is to exist and possibly replicate in it. Considerations are made with respect to the vector copy number, the ability to control the copy number, expression of other proteins such as antibiotic resistance. In
25 selecting an expression control sequence, a number of variables are considered. Among the important variable are the relative strength of the sequence (e.g. the ability to drive expression under various conditions), the ability to control the sequence's function, compatibility between the polynucleotide to be
30 expressed and the control sequence (e.g. secondary structures are considered to avoid hairpin structures which prevent efficient transcription). In selecting the host, unicellular hosts are selected which are compatible with the selected vector, tolerant of any possible toxic effects of the expressed

product, able to secrete the expressed product efficiently if such is desired, to be able to express the product in the desired conformation, to be easily scaled up, and to which ease of purification of the final product.

5 The choice of the expression cassette depends on the host system selected as well as the features desired for the expressed polypeptide. Typically, an expression cassette includes a promoter that is functional in the selected host system and can be constitutive or inducible; a ribosome binding
10 site; a start codon (ATG) if necessary; a region encoding a signal peptide, e.g., a lipidation signal peptide; a DNA molecule of the invention; a stop codon; and optionally a 3' terminal region (translation and/or transcription terminator). The signal peptide encoding region is adjacent to the
15 polynucleotide of the invention and placed in proper reading frame. The signal peptide-encoding region is homologous or heterologous to the DNA molecule encoding the mature polypeptide and is compatible with the secretion apparatus of the host used for expression. The open reading frame constituted by the DNA
20 molecule of the invention, solely or together with the signal peptide, is placed under the control of the promoter so that transcription and translation occur in the host system. Promoters and signal peptide encoding regions are widely known and available to those skilled in the art and include, for
25 example, the promoter of *Salmonella typhimurium* (and derivatives) that is inducible by arabinose (promoter araB) and is functional in Gram-negative bacteria such as *E. coli* (as described in U.S. Patent No. 5,028,530 and in Cagnon et al., (Cagnon et al., Protein Engineering (1991) 4(7):843)); the
30 promoter of the gene of bacteriophage T7 encoding RNA polymerase, that is functional in a number of *E. coli* strains expressing T7 polymerase (described in U.S. Patent No. 4,952,496); OspA lipidation signal peptide ; and RlpB

lipidation signal peptide (Takase et al., J. Bact. (1987) 169:5692).

The expression cassette is typically part of an expression vector, which is selected for its ability to replicate in the chosen expression system. Expression vectors (e.g., plasmids or viral vectors) can be chosen, for example, from those described in Pouwels et al. (Cloning Vectors: A Laboratory Manual 1985, Supp. 1987). Suitable expression vectors can be purchased from various commercial sources.

10 Methods for transforming/transfecting host cells with expression vectors are well-known in the art and depend on the host system selected as described in Ausubel et al., (Ausubel et al., Current Protocols in Molecular Biology, John Wiley & Sons Inc., 1994).

15 Upon expression, a recombinant polypeptide of the invention (or a polypeptide derivative) is produced and remains in the intracellular compartment, is secreted/excreted in the extracellular medium or in the periplasmic space, or is embedded in the cellular membrane. The polypeptide is recovered in a
20 substantially purified form from the cell extract or from the supernatant after centrifugation of the recombinant cell culture. Typically, the recombinant polypeptide is purified by antibody-based affinity purification or by other well-known methods that can be readily adapted by a person skilled in the
25 art, such as fusion of the polynucleotide encoding the polypeptide or its derivative to a small affinity binding domain. Antibodies useful for purifying by immunoaffinity the polypeptides of the invention are obtained as described below.

A polynucleotide of the invention can also be useful
30 as a vaccine. There are two major routes, either using a viral or bacterial host as gene delivery vehicle (live vaccine vector) or administering the gene in a free form, e.g., inserted into a plasmid. Therapeutic or prophylactic efficacy of a polynucleotide of the invention is evaluated as described below.

Accordingly, a third aspect of the invention provides (i) a vaccine vector such as a poxvirus, containing a DNA molecule of the invention, placed under the control of elements required for expression; (ii) a composition of matter comprising
5 a vaccine vector of the invention, together with a diluent or carrier; specifically (iii) a pharmaceutical composition containing a therapeutically or prophylactically effective amount of a vaccine vector of the invention; (iv) a method for inducing an immune response against *Chlamydia* in a mammal (e.g.,
10 a human; alternatively, the method can be used in veterinary applications for treating or preventing *Chlamydia* infection of animals, e.g., cats or birds), which involves administering to the mammal an immunogenically effective amount of a vaccine vector of the invention to elicit a protective or therapeutic
15 immune response to *Chlamydia* ; and particularly, (v) a method for preventing and/or treating a *Chlamydia* (e.g., *C. trachomatis*, *C. psittaci*, *C. pneumonia*, *C. pecorum*) infection, which involves administering a prophylactic or therapeutic amount of a vaccine vector of the invention to an
20 infected individual. Additionally, the third aspect of the invention encompasses the use of a vaccine vector of the invention in the preparation of a medicament for preventing and/or treating *Chlamydia* infection.

As used herein, a vaccine vector expresses one or
25 several polypeptides or derivatives of the invention. The vaccine vector may express additionally a cytokine, such as interleukin-2 (IL-2) or interleukin-12 (IL-12), that enhances the immune response (adjuvant effect). It is understood that each of the components to be expressed is placed under the
30 control of elements required for expression in a mammalian cell.

Consistent with the third aspect of the invention is a composition comprising several vaccine vectors, each of them capable of expressing a polypeptide or derivative of the invention. A composition may also comprise a vaccine vector

capable of expressing an additional *Chlamydia* antigen, or a subunit, fragment, homolog, mutant, or derivative thereof; optionally together with or a cytokine such as IL-2 or IL-12.

Vaccination methods for treating or preventing
5 infection in a mammal comprises use of a vaccine vector of the invention to be administered by any conventional route, particularly to a mucosal (e.g., ocular, intranasal, oral, gastric, pulmonary, intestinal, rectal, vaginal, or urinary tract) surface or via the parenteral (e.g., subcutaneous,
10 intradermal, intramuscular, intravenous, or intraperitoneal) route. Preferred routes depend upon the choice of the vaccine vector. Treatment may be effected in a single dose or repeated at intervals. The appropriate dosage depends on various parameters understood by skilled artisans such as the vaccine
15 vector itself, the route of administration or the condition of the mammal to be vaccinated (weight, age and the like).

Live vaccine vectors available in the art include viral vectors such as adenoviruses and poxviruses as well as bacterial vectors, e.g., *Shigella*, *Salmonella*, *Vibrio cholerae*,
20 *Lactobacillus*, Bacille bilié de Calmette-Guérin (BCG), and *Streptococcus*.

An example of an adenovirus vector, as well as a method for constructing an adenovirus vector capable of expressing a DNA molecule of the invention, are described in
25 U.S. Patent No. 4,920,209. Poxvirus vectors include vaccinia and canary pox virus, described in U.S. Patent No. 4,722,848 and U.S. Patent No. 5,364,773, respectively. (Also see, e.g., Tartaglia et al., Virology (1992) 188:217) for a description of a vaccinia virus vector and Taylor et al, Vaccine (1995) 13:539
30 for a reference of a canary pox.) Poxvirus vectors capable of expressing a polynucleotide of the invention are obtained by homologous recombination as described in Kieny et al., Nature (1984) 312:163 so that the polynucleotide of the invention is inserted in the viral genome under appropriate conditions for

expression in mammalian cells. Generally, the dose of vaccine viral vector, for therapeutic or prophylactic use, can be of from about 1×10^4 to about 1×10^{11} , advantageously from about 1×10^7 to about 1×10^{10} , preferably of from about 1×10^7 to about 1×10^9 5 plaque-forming units per kilogram. Preferably, viral vectors are administered parenterally; for example, in 3 doses, 4 weeks apart. It is preferable to avoid adding a chemical adjuvant to a composition containing a viral vector of the invention and thereby minimizing the immune response to the viral vector 10 itself.

Non-toxicogenic *Vibrio cholerae* mutant strains that are useful as a live oral vaccine are known. Mekalanos et al., Nature (1983) 306:551 and U.S. Patent No. 4,882,278 describe strains which have a substantial amount of the coding sequence 15 of each of the two *ctxA* alleles deleted so that no functional *cholerae* toxin is produced. WO 92/11354 describes a strain in which the *irgA* locus is inactivated by mutation; this mutation can be combined in a single strain with *ctxA* mutations. WO 94/01533 describes a deletion mutant lacking functional *ctxA* and 20 *attRS1* DNA sequences. These mutant strains are genetically engineered to express heterologous antigens, as described in WO 94/19482. An effective vaccine dose of a *Vibrio cholerae* strain capable of expressing a polypeptide or polypeptide derivative encoded by a DNA molecule of the invention contains 25 about 1×10^5 to about 1×10^9 , preferably about 1×10^6 to about 1×10^8 , viable bacteria in a volume appropriate for the selected route of administration. Preferred routes of administration include all mucosal routes; most preferably, these vectors are administered intranasally or orally.

30 Attenuated *Salmonella typhimurium* strains, genetically engineered for recombinant expression of heterologous antigens or not, and their use as oral vaccines are described in Nakayama et al. (Bio/Technology (1988) 6:693) and WO 92/11361. Preferred routes of administration include all mucosal routes;

most preferably, these vectors are administered intranasally or orally.

Other bacterial strains used as vaccine vectors in the context of the present invention are described for *Shigella*
5 *flexneri* in High et al., EMBO (1992) 11:1991 and Sizemore et al., Science (1995) 270:299; for *Streptococcus gordonii* in Medaglini et al., Proc. Natl. Acad. Sci. USA (1995) 92:6868; and for Bacille Calmette Guerin in Flynn J.L., Cell. Mol. Biol. (1994) 40 (suppl. I):31, WO 88/06626, WO 90/00594, WO 91/13157,
10 WO 92/01796, and WO 92/21376.

In bacterial vectors, the polynucleotide of the invention is inserted into the bacterial genome or remains in a free state as part of a plasmid.

The composition comprising a vaccine bacterial vector
15 of the present invention may further contain an adjuvant. A number of adjuvants are known to those skilled in the art. Preferred adjuvants are selected as provided below.

Accordingly, a fourth aspect of the invention provides (i) a composition of matter comprising a polynucleotide of the
20 invention, together with a diluent or carrier; (ii) a pharmaceutical composition comprising a therapeutically or prophylactically effective amount of a polynucleotide of the invention; (iii) a method for inducing an immune response against *Chlamydia* in a mammal by administration of an
25 immunogenically effective amount of a polynucleotide of the invention to elicit a protective immune response to *Chlamydia*; and particularly, (iv) a method for preventing and/or treating a *Chlamydia* (e.g., *C. trachomatis*, *C. psittaci*, *C. pneumoniae*, or *C. pecorum*) infection, by administering a prophylactic or
30 therapeutic amount of a polynucleotide of the invention to an infected individual. Additionally, the fourth aspect of the invention encompasses the use of a polynucleotide of the invention in the preparation of a medicament for preventing and/or treating *Chlamydia* infection. A preferred use includes

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the use of a DNA molecule placed under conditions for expression in a mammalian cell, especially in a plasmid that is unable to replicate in mammalian cells and to substantially integrate in a mammalian genome.

5 Use of the polynucleotides of the invention include their administration to a mammal as a vaccine, for therapeutic or prophylactic purposes. Such polynucleotides are used in the form of DNA as part of a plasmid that is unable to replicate in a mammalian cell and unable to
10 integrate into the mammalian genome. Typically, such a DNA molecule is placed under the control of a promoter suitable for expression in a mammalian cell. The promoter functions either ubiquitously or tissue-specifically. Examples of non-tissue specific promoters include the early
15 Cytomegalovirus (CMV) promoter (described in U.S. Patent No. 5,168,062) and the Rous Sarcoma Virus promoter (described in Norton & Coffin, *Molec. Cell Biol.* (1985) 5:281). An example of a tissue-specific promoter is the desmin promoter which drives expression in muscle cells
20 (Li *et al.*, *Gene* (1989) 78:243, Li & Paulin, *J. Biol. Chem.* (1991) 266:6562 and Li & Paulin, *J. Biol. Chem.* (1993) 268:10403). Use of promoters is well-known to those skilled in the art. Useful vectors are described in numerous publications, specifically WO 94/21797 and Hartikka *et al.*,
25 Human Gene Therapy (1996) 7:1205.

Polynucleotides of the invention which are used as vaccines encode either a precursor or a mature form of the corresponding polypeptide. In the precursor form, the signal peptide is either homologous or heterologous. In the
30 latter case, a eucaryotic leader sequence such as the leader sequence of the tissue-type plasminogen factor (tPA) is preferred.

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As used herein, a composition of the invention contains one or several polynucleotides with optionally at least one additional polynucleotide encoding another *Chlamydia* antigen such as urease subunit A, B, or both, or a
5 fragment, derivative, mutant, or analog thereof. The composition may also contain an

additional polynucleotide encoding a cytokine, such as interleukin-2 (IL-2) or interleukin-12 (IL-12) so that the immune response is enhanced. These additional polynucleotides are placed under appropriate control for expression.

5 Advantageously, DNA molecules of the invention and/or additional DNA molecules to be included in the same composition, are present in the same plasmid.

Standard techniques of molecular biology for preparing and purifying polynucleotides are used in the preparation of
10 polynucleotide therapeutics of the invention. For use as a vaccine, a polynucleotide of the invention is formulated according to various methods outlined below.

One method utilizes the polynucleotide in a naked form, free of any delivery vehicles. Such a polynucleotide is
15 simply diluted in a physiologically acceptable solution such as sterile saline or sterile buffered saline, with or without a carrier. When present, the carrier preferably is isotonic, hypotonic, or weakly hypertonic, and has a relatively low ionic strength, such as provided by a sucrose solution, e.g., a
20 solution containing 20% sucrose.

An alternative method utilizes the polynucleotide in association with agents that assist in cellular uptake. Examples of such agents are (i) chemicals that modify cellular permeability, such as bupivacaine (see, e.g., WO 94/16737), (ii)
25 liposomes for encapsulation of the polynucleotide, or (iii) cationic lipids or silica, gold, or tungsten microparticles which associate themselves with the polynucleotides.

Anionic and neutral liposomes are well-known in the
30 art (see, e.g., Liposomes: A Practical Approach, RPC New Ed, IRL press (1990), for a detailed description of methods for making liposomes) and are useful for delivering a large range of products, including polynucleotides.

Cationic lipids are also known in the art and are commonly used for gene delivery. Such lipids include LipofectinTM also known as DOTMA (N-[1-(2,3-dioleyloxy)propyl]-N,N,N-trimethylammonium chloride), DOTAP (1,2-bis(oleyloxy)-3-(trimethylammonio)propane), DDAB (dimethyldioctadecylammonium bromide), DOGS (dioctadecylamidoglycyl spermine) and cholesterol derivatives such as DC-Chol (3 beta-(N-(N',N'-dimethyl aminomethane)-carbamoyl) cholesterol). A description of these cationic lipids can be found in EP 187,702, 10 WO 90/11092, U.S. Patent No. 5,283,185, WO 91/15501, WO 95/26356, and U.S. Patent No. 5,527,928. Cationic lipids for gene delivery are preferably used in association with a neutral lipid such as DOPE (dioleoyl phosphatidylethanolamine), as described in WO 90/11092 as an example.

15 Formulation containing cationic liposomes may optionally contain other transfection-facilitating compounds. A number of them are described in WO 93/18759, WO 93/19768, WO 94/25608, and WO 95/02397. They include spermine derivatives useful for facilitating the transport of DNA through the nuclear 20 membrane (see, for example, WO 93/18759) and membrane-permeabilizing compounds such as GALA, Gramicidine S, and cationic bile salts (see, for example, WO 93/19768).

Gold or tungsten microparticles are used for gene delivery, as described in WO 91/00359, WO 93/17706, and Tang et 25 al. Nature (1992) 356:152. The microparticle-coated polynucleotide is injected via intradermal or intraepidermal routes using a needleless injection device ("gene gun"), such as those described in U.S. Patent No. 4,945,050, U.S. Patent No. 5,015,580, and WO 94/24263.

30 The amount of DNA to be used in a vaccine recipient depends, e.g., on the strength of the promoter used in the DNA construct, the immunogenicity of the expressed gene product, the condition of the mammal intended for administration (e.g., the weight, age, and general health of the mammal), the mode of

administration, and the type of formulation. In general, a therapeutically or prophylactically effective dose from about 1 μ g to about 1 mg, preferably, from about 10 μ g to about 800 μ g and, more preferably, from about 25 μ g to about 250 μ g, can be administered to human adults. The administration can be achieved in a single dose or repeated at intervals.

The route of administration is any conventional route used in the vaccine field. As general guidance, a polynucleotide of the invention is administered via a mucosal surface, e.g., an ocular, intranasal, pulmonary, oral, intestinal, rectal, vaginal, and urinary tract surface; or via a parenteral route, e.g., by an intravenous, subcutaneous, intraperitoneal, intradermal, intraepidermal, or intramuscular route. The choice of administration route depends on the formulation that is selected. A polynucleotide formulated in association with bupivacaine is advantageously administered into muscles. When a neutral or anionic liposome or a cationic lipid, such as DOTMA or DC-Chol, is used, the formulation can be advantageously injected via intravenous, intranasal (aerosolization), intramuscular, intradermal, and subcutaneous routes. A polynucleotide in a naked form can advantageously be administered via the intramuscular, intradermal, or subcutaneous routes.

Although not absolutely required, such a composition can also contain an adjuvant. If so, a systemic adjuvant that does not require concomitant administration in order to exhibit an adjuvant effect is preferable such as, e.g., QS21, which is described in U.S. Patent No. 5,057,546.

The sequence information provided in the present application enables the design of specific nucleotide probes and primers that are used for diagnostic purposes. Accordingly, a fifth aspect of the invention provides a nucleotide probe or primer having a sequence found in or derived by degeneracy of the genetic code from a sequence shown in SEQ ID No:1.

The term "probe" as used in the present application refers to DNA (preferably single stranded) or RNA molecules (or modifications or combinations thereof) that hybridize under the stringent conditions, as defined above, to nucleic acid molecules having SEQ ID No:1 or to sequences homologous to SEQ ID No:1, or to its complementary or anti-sense sequence. Generally, probes are significantly shorter than full-length sequences. Such probes contain from about 5 to about 100, preferably from about 10 to about 80, nucleotides. In particular, probes have sequences that are at least 75%, preferably at least 85%, more preferably 95% homologous to a portion of SEQ ID No:1 or that are complementary to such sequences. Probes may contain modified bases such as inosine, methyl-5-deoxycytidine, deoxyuridine, dimethylamino-5-deoxyuridine, or diamino-2, 6-purine. Sugar or phosphate residues may also be modified or substituted. For example, a deoxyribose residue may be replaced by a polyamide (Nielsen *et al.*, Science (1991) 254:1497) and phosphate residues may be replaced by ester groups such as diphosphate, alkyl, arylphosphonate and phosphorothioate esters. In addition, the 2'-hydroxyl group on ribonucleotides may be modified by including such groups as alkyl groups.

Probes of the invention are used in diagnostic tests, as capture or detection probes. Such capture probes are conventionally immobilized on a solid support, directly or indirectly, by covalent means or by passive adsorption. A detection probe is labelled by a detection marker selected from: radioactive isotopes, enzymes such as peroxidase, alkaline phosphatase, and enzymes able to hydrolyze a chromogenic, fluorogenic, or luminescent substrate, compounds that are chromogenic, fluorogenic, or luminescent, nucleotide base analogs, and biotin.

Probes of the invention are used in any conventional hybridization technique, such as dot blot (Maniatis *et al.*,

Molecular Cloning: A Laboratory Manual (1982) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York), Southern blot (Southern, J. Mol. Biol. (1975) 98:503), northern blot (identical to Southern blot with the exception that RNA is used as a target), or the sandwich technique (Dunn et al., Cell (1977) 12:23). The latter technique involves the use of a specific capture probe and/or a specific detection probe with nucleotide sequences that at least partially differ from each other.

10 A primer is a probe of usually about 10 to about 40 nucleotides that is used to initiate enzymatic polymerization of DNA in an amplification process (e.g., PCR), in an elongation process, or in a reverse transcription method. Primers used in diagnostic methods involving PCR are labeled by methods known in
15 the art.

As described herein, the invention also encompasses (i) a reagent comprising a probe of the invention for detecting and/or identifying the presence of *Chlamydia* in a biological material; (ii) a method for detecting and/or identifying the
20 presence of *Chlamydia* in a biological material, in which (a) a sample is recovered or derived from the biological material, (b) DNA or RNA is extracted from the material and denatured, and (c) exposed to a probe of the invention, for example, a capture, detection probe or both, under stringent hybridization
25 conditions, such that hybridization is detected; and (iii) a method for detecting and/or identifying the presence of *Chlamydia* in a biological material, in which (a) a sample is recovered or derived from the biological material, (b) DNA is extracted therefrom, (c) the extracted DNA is primed with at
30 least one, and preferably two, primers of the invention and amplified by polymerase chain reaction, and (d) the amplified DNA fragment is produced.

It is apparent that disclosure of polynucleotide sequences of SEQ ID No:1, its homologs and partial sequences

enable their corresponding amino acid sequences. Accordingly, a sixth aspect of the invention features a substantially purified polypeptide or polypeptide derivative having an amino acid sequence encoded by a polynucleotide of the invention.

5 A "substantially purified polypeptide" as used herein is defined as a polypeptide that is separated from the environment in which it naturally occurs and/or that is free of the majority of the polypeptides that are present in the environment in which it was synthesized. For example, a
10 substantially purified polypeptide is free from cytoplasmic polypeptides. Those skilled in the art would readily understand that the polypeptides of the invention may be purified from a natural source, *i.e.*, a *Chlamydia* strain, or produced by recombinant means.

15 Consistent with the sixth aspect of the invention are polypeptides, homologs or fragments which are modified or treated to enhance their immunogenicity in the target animal, in whom the polypeptide, homolog or fragments are intended to confer protection against *Chlamydia*. Such modifications or
20 treatments include: amino acid substitutions with an amino acid derivative such as 3-methylhistidine, 4-hydroxyproline, 5-hydroxylysine etc., modifications or deletions which are carried out after preparation of the polypeptide, homolog or fragment, such as the modification of free amino, carboxyl or hydroxyl
25 side groups of the amino acids.

 Identification of homologous polypeptides or polypeptide derivatives encoded by polynucleotides of the invention which have specific antigenicity is achieved by screening for cross-reactivity with an antiserum raised against
30 the polypeptide of reference having an amino acid sequence of SEQ ID No:1. The procedure is as follows: a monospecific hyperimmune antiserum is raised against a purified reference polypeptide, a fusion polypeptide (for example, an expression product of MBP, GST, or His-tag systems, the description and

instructions for use of which are contained in Invitrogen product manuals for pcDNA3.1/Myc-His(+) A, B, and C and for the Xpress[™] System Protein Purification), or a synthetic peptide predicted to be antigenic. Where an antiserum is raised against
5 a fusion polypeptide, two different fusion systems are employed. Specific antigenicity can be determined according to a number of methods, including Western blot (Towbin et al., Proc. Natl. Acad. Sci. USA (1979) 76:4350), dot blot, and ELISA, as described below.

10 In a Western blot assay, the product to be screened, either as a purified preparation or a total *E. coli* extract, is submitted to SDS-Page electrophoresis as described by Laemmli (Nature (1970) 227:680). After transfer to a nitrocellulose membrane, the material is further incubated with the
15 monospecific hyperimmune antiserum diluted in the range of dilutions from about 1:5 to about 1:5000, preferably from about 1:100 to about 1:500. Specific antigenicity is shown once a band corresponding to the product exhibits reactivity at any of the dilutions in the above range.

20 In an ELISA assay, the product to be screened is preferably used as the coating antigen. A purified preparation is preferred, although a whole cell extract can also be used. Briefly, about 100 μ l of a preparation at about 10 μ g protein/ml are distributed into wells of a 96-well polycarbonate ELISA
25 plate. The plate is incubated for 2 hours at 37°C then overnight at 4°C. The plate is washed with phosphate buffer saline (PBS) containing 0.05% Tween 20 (PBS/Tween buffer). The wells are saturated with 250 μ l PBS containing 1% bovine serum albumin (BSA) to prevent non-specific antibody binding. After 1
30 hour incubation at 37°C, the plate is washed with PBS/Tween buffer. The antiserum is serially diluted in PBS/Tween buffer containing 0.5% BSA. 100 μ l of dilutions are added per well. The plate is incubated for 90 minutes at 37°C, washed and evaluated according to standard procedures. For example, a goat

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anti-rabbit peroxidase conjugate is added to the wells when specific antibodies were raised in rabbits. Incubation is carried out for 90 minutes at 37°C and the plate is washed. The reaction is developed with the appropriate substrate and the
5 reaction is measured by colorimetry (absorbance measured spectrophotometrically). Under the above experimental conditions, a positive reaction is shown by O.D. values greater than a non immune control serum.

In a dot blot assay, a purified product is preferred,
10 although a whole cell extract can also be used. Briefly, a solution of the product at about 100 µg/ml is serially two-fold diluted in 50 mM Tris-HCl (pH 7.5). 100 µl of each dilution are applied to a nitrocellulose membrane 0.45 µm set in a 96-well dot blot apparatus (Biorad). The buffer is removed by applying
15 vacuum to the system. Wells are washed by addition of 50 mM Tris-HCl (pH 7.5) and the membrane is air-dried. The membrane is saturated in blocking buffer (50 mM Tris-HCl (pH 7.5) 0.15 M NaCl, 10 g/L skim milk) and incubated with an antiserum dilution from about 1:50 to about 1:5000, preferably about 1:500. The
20 reaction is revealed according to standard procedures. For example, a goat anti-rabbit peroxidase conjugate is added to the wells when rabbit antibodies are used. Incubation is carried out 90 minutes at 37°C and the blot is washed. The reaction is developed with the appropriate substrate and stopped. The
25 reaction is measured visually by the appearance of a colored spot, e.g., by colorimetry. Under the above experimental conditions, a positive reaction is shown once a colored spot is associated with a dilution of at least about 1:5, preferably of at least about 1:500.

30 Therapeutic or prophylactic efficacy of a polypeptide or derivative of the invention can be evaluated as described below. A seventh aspect of the invention provides (i) a composition of matter comprising a polypeptide of the invention together with a diluent or carrier; specifically (ii) a

pharmaceutical composition containing a therapeutically or prophylactically effective amount of a polypeptide of the invention; (iii) a method for inducing an immune response against *Chlamydia* in a mammal, by administering to the mammal an
5 immunogenically effective amount of a polypeptide of the invention to elicit a protective immune response to *Chlamydia*; and particularly, (iv) a method for preventing and/or treating a *Chlamydia* (e.g., *C. trachomatis*, *C. psittaci*, *C. pneumoniae*, or *C. pecorum*) infection, by administering a prophylactic or
10 therapeutic amount of a polypeptide of the invention to an infected individual. Additionally, the seventh aspect of the invention encompasses the use of a polypeptide of the invention in the preparation of a medicament for preventing and/or treating *Chlamydia* infection.

15 As used herein, the immunogenic compositions of the invention are administered by conventional routes known the vaccine field, in particular to a mucosal (e.g., ocular, intranasal, pulmonary, oral, gastric, intestinal, rectal, vaginal, or urinary tract) surface or via the parenteral (e.g.,
20 subcutaneous, intradermal, intramuscular, intravenous, or intraperitoneal) route. The choice of administration route depends upon a number of parameters, such as the adjuvant associated with the polypeptide. If a mucosal adjuvant is used, the intranasal or oral route is preferred. If a lipid
25 formulation or an aluminum compound is used, the parenteral route is preferred with the sub-cutaneous or intramuscular route being most preferred. The choice also depends upon the nature of the vaccine agent. For example, a polypeptide of the invention fused to CTB or LTB is best administered to a mucosal
30 surface.

As used herein, the composition of the invention contains one or several polypeptides or derivatives of the invention. The composition optionally contains at least one

additional *Chlamydia* antigen, or a subunit, fragment, homolog, mutant, or derivative thereof.

For use in a composition of the invention, a polypeptide or derivative thereof is formulated into or with
5 liposomes, preferably neutral or anionic liposomes, microspheres, ISCOMS, or virus-like-particles (VLPs) to facilitate delivery and/or enhance the immune response. These compounds are readily available to one skilled in the art; for example, see *Liposomes: A Practical Approach*, RCP New Ed, IRL
10 press (1990).

Adjuvants other than liposomes and the like are also used and are known in the art. Adjuvants may protect the antigen from rapid dispersal by sequestering it in a local deposit, or they may contain substances that stimulate the host
15 to secrete factors that are chemotactic for macrophages and other components of the immune system. An appropriate selection can conventionally be made by those skilled in the art, for example, from those described below (under the eleventh aspect of the invention).

20 Treatment is achieved in a single dose or repeated as necessary at intervals, as can be determined readily by one skilled in the art. For example, a priming dose is followed by three booster doses at weekly or monthly intervals. An appropriate dose depends on various parameters including the
25 recipient (e.g., adult or infant), the particular vaccine antigen, the route and frequency of administration, the presence/absence or type of adjuvant, and the desired effect (e.g., protection and/or treatment), as can be determined by one skilled in the art. In general, a vaccine antigen of the
30 invention is administered by a mucosal route in an amount from about 10 μ g to about 500 mg, preferably from about 1 mg to about 200 mg. For the parenteral route of administration, the dose usually does not exceed about 1 mg, preferably about 100 μ g.

When used as vaccine agents, polynucleotides and polypeptides of the invention may be used sequentially as part of a multistep immunization process. For example, a mammal is initially primed with a vaccine vector of the invention such as
5 a pox virus, e.g., via the parenteral route, and then boosted twice with the polypeptide encoded by the vaccine vector, e.g., via the mucosal route. In another example, liposomes associated with a polypeptide or derivative of the invention is also used for priming, with boosting being carried out mucosally using a
10 soluble polypeptide or derivative of the invention in combination with a mucosal adjuvant (e.g., LT).

A polypeptide derivative of the invention is also used in accordance with the seventh aspect as a diagnostic reagent for detecting the presence of anti-*Chlamydia* antibodies, e.g.,
15 in a blood sample. Such polypeptides are about 5 to about 80, preferably about 10 to about 50 amino acids in length. They are either labeled or unlabeled, depending upon the diagnostic method. Diagnostic methods involving such a reagent are described below.

20 Upon expression of a DNA molecule of the invention, a polypeptide or polypeptide derivative is produced and purified using known laboratory techniques. As described above, the polypeptide or polypeptide derivative may be produced as a fusion protein containing a fused tail that facilitates
25 purification. The fusion product is used to immunize a small mammal, e.g., a mouse or a rabbit, in order to raise antibodies against the polypeptide or polypeptide derivative (monospecific antibodies). Accordingly, an eighth aspect of the invention provides a monospecific antibody that binds to a polypeptide or
30 polypeptide derivative of the invention.

By "monospecific antibody" is meant an antibody that is capable of reacting with a unique naturally-occurring *Chlamydia* polypeptide. An antibody of the invention is either polyclonal or monoclonal. Monospecific antibodies may be

recombinant, e.g., chimeric (e.g., constituted by a variable region of murine origin associated with a human constant region), humanized (a human immunoglobulin constant backbone together with hypervariable region of animal, e.g., murine, 5 origin), and/or single chain. Both polyclonal and monospecific antibodies may also be in the form of immunoglobulin fragments, e.g., F(ab)'₂ or Fab fragments. The antibodies of the invention are of any isotype, e.g., IgG or IgA, and polyclonal antibodies are of a single isotype or a mixture of isotypes.

10 Antibodies against the polypeptides, homologs or fragments of the present invention are generated by immunization of a mammal with a composition comprising said polypeptide, homolog or fragment. Such antibodies may be polyclonal or monoclonal. Methods to produce polyclonal or monoclonal 15 antibodies are well known in the art. For a review, see "Antibodies, A Laboratory Manual, Cold Spring Harbor Laboratory, Eds. E. Harlow and D. Lane (1988), and D.E. Yelton et al., 1981. Ann. Rev. Biochem. 50:657-680. For monoclonal antibodies, see Kohler & Milstein (1975) Nature 256:495-497.

20 The antibodies of the invention, which are raised to a polypeptide or polypeptide derivative of the invention, are produced and identified using standard immunological assays, e.g., Western blot analysis, dot blot assay, or ELISA (see, e.g., Coligan et al., Current Protocols in Immunology (1994) 25 John Wiley & Sons, Inc., New York, NY). The antibodies are used in diagnostic methods to detect the presence of a *Chlamydia* antigen in a sample, such as a biological sample. The antibodies are also used in affinity chromatography for purifying a polypeptide or polypeptide derivative of the 30 invention. As is discussed further below, such antibodies may be used in prophylactic and therapeutic passive immunization methods.

Accordingly, a ninth aspect of the invention provides (i) a reagent for detecting the presence of *Chlamydia* in a

biological sample that contains an antibody, polypeptide, or polypeptide derivative of the invention; and (ii) a diagnostic method for detecting the presence of *Chlamydia* in a biological sample, by contacting the biological sample with an antibody, a
5 polypeptide, or a polypeptide derivative of the invention, such that an immune complex is formed, and by detecting such complex to indicate the presence of *Chlamydia* in the sample or the organism from which the sample is derived.

Those skilled in the art will readily understand that
10 the immune complex is formed between a component of the sample and the antibody, polypeptide, or polypeptide derivative, whichever is used, and that any unbound material is removed prior to detecting the complex. It is understood that a polypeptide reagent is useful for detecting the presence of
15 anti-*Chlamydia* antibodies in a sample, e.g., a blood sample, while an antibody of the invention is used for screening a sample, such as a gastric extract or biopsy, for the presence of *Chlamydia* polypeptides.

For diagnostic applications, the reagent (i.e., the
20 antibody, polypeptide, or polypeptide derivative of the invention) is either in a free state or immobilized on a solid support, such as a tube, a bead, or any other conventional support used in the field. Immobilization is achieved using direct or indirect means. Direct means include passive
25 adsorption (non-covalent binding) or covalent binding between the support and the reagent. By "indirect means" is meant that an anti-reagent compound that interacts with a reagent is first attached to the solid support. For example, if a polypeptide reagent is used, an antibody that binds to it can serve as an
30 anti-reagent, provided that it binds to an epitope that is not involved in the recognition of antibodies in biological samples. Indirect means may also employ a ligand-receptor system, for example, where a molecule such as a vitamin is grafted onto the polypeptide reagent and the corresponding receptor immobilized

on the solid phase. This is illustrated by the biotin-streptavidin system. Alternatively, a peptide tail is added chemically or by genetic engineering to the reagent and the grafted or fused product immobilized by passive adsorption or
5 covalent linkage of the peptide tail.

Such diagnostic agents may be included in a kit which also comprises instructions for use. The reagent is labeled with a detection means which allows for the detection of the reagent when it is bound to its target. The detection means may
10 be a fluorescent agent such as fluorescein isocyanate or fluorescein isothiocyanate, or an enzyme such as horse radish peroxidase or luciferase or alkaline phosphatase, or a radioactive element such as ^{125}I or ^{51}Cr .

Accordingly, a tenth aspect of the invention provides
15 a process for purifying, from a biological sample, a polypeptide or polypeptide derivative of the invention, which involves carrying out antibody-based affinity chromatography with the biological sample, wherein the antibody is a monospecific antibody of the invention.

20 For use in a purification process of the invention, the antibody is either polyclonal or monospecific, and preferably is of the IgG type. Purified IgGs is prepared from an antiserum using standard methods (see, e.g., Coligan et al., Current Protocols in Immunology (1994) John Wiley & Sons, Inc.,
25 New York, NY.). Conventional chromatography supports, as well as standard methods for grafting antibodies, are described in, e.g., Antibodies: A Laboratory Manual, D. Lane, E. Harlow, Eds. (1988) and outlined below.

Briefly, a biological sample, such as an *C. pneumoniae*
30 extract preferably in a buffer solution, is applied to a chromatography material, preferably equilibrated with the buffer used to dilute the biological sample so that the polypeptide or polypeptide derivative of the invention (i.e., the antigen) is allowed to adsorb onto the material. The chromatography

material, such as a gel or a resin coupled to an antibody of the invention, is in either a batch form or a column. The unbound components are washed off and the antigen is then eluted with an appropriate elution buffer, such as a glycine buffer or a buffer
5 containing a chaotropic agent, e.g., guanidine HCl, or high salt concentration (e.g., 3 M MgCl₂). Eluted fractions are recovered and the presence of the antigen is detected, e.g., by measuring the absorbance at 280 nm.

An eleventh aspect of the invention provides (i) a
10 composition of matter comprising a monospecific antibody of the invention, together with a diluent or carrier; (ii) a pharmaceutical composition comprising a therapeutically or prophylactically effective amount of a monospecific antibody of the invention, and (iii) a method for treating or preventing a
15 *Chlamydia* (e.g., *C. trachomatis*, *C. psittaci*, *C. pneumoniae* or *C. pecorum*) infection, by administering a therapeutic or prophylactic amount of a monospecific antibody of the invention to an infected individual. Additionally, the eleventh aspect of the invention encompasses the use of a monospecific antibody of
20 the invention in the preparation of a medicament for treating or preventing *Chlamydia* infection.

The monospecific antibody is either polyclonal or monoclonal, preferably of the IgA isotype (predominantly). In passive immunization, the antibody is administered to a mucosal
25 surface of a mammal, e.g., the gastric mucosa, e.g., orally or intragastrically, advantageously, in the presence of a bicarbonate buffer. Alternatively, systemic administration, not requiring a bicarbonate buffer, is carried out. A monospecific antibody of the invention is administered as a single active
30 component or as a mixture with at least one monospecific antibody specific for a different *Chlamydia* polypeptide. The amount of antibody and the particular regimen used are readily determined by one skilled in the art. For example, daily administration of about 100 to 1,000 mg of antibodies over one

week, or three doses per day of about 100 to 1,000 mg of antibodies over two or three days, are effective regimens for most purposes.

Therapeutic or prophylactic efficacy are evaluated using standard methods in the art, e.g., by measuring induction of a mucosal immune response or induction of protective and/or therapeutic immunity, using, e.g., the *C. pneumoniae* mouse model. Those skilled in the art will readily recognize that the *C. pneumoniae* strain of the model may be replaced with another *Chlamydia* strain. For example, the efficacy of DNA molecules and polypeptides from *C. pneumoniae* is preferably evaluated in a mouse model using *C. pneumoniae* strain. Protection is determined by comparing the degree of *Chlamydia* infection to that of a control group. Protection is shown when infection is reduced by comparison to the control group. Such an evaluation is made for polynucleotides, vaccine vectors, polypeptides and derivatives thereof, as well as antibodies of the invention.

Adjuvants useful in any of the vaccine compositions described above are as follows.

Adjuvants for parenteral administration include aluminum compounds, such as aluminum hydroxide, aluminum phosphate, and aluminum hydroxy phosphate. The antigen is precipitated with, or adsorbed onto, the aluminum compound according to standard protocols. Other adjuvants, such as RIBI (ImmunoChem, Hamilton, MT), are used in parenteral administration.

Adjuvants for mucosal administration include bacterial toxins, e.g., the cholera toxin (CT), the *E. coli* heat-labile toxin (LT), the *Clostridium difficile* toxin A and the pertussis toxin (PT), or combinations, subunits, toxoids, or mutants thereof such as a purified preparation of native cholera toxin subunit B (CTB). Fragments, homologs, derivatives, and fusions to any of these toxins are also suitable, provided that they retain adjuvant activity. Preferably, a mutant having reduced

toxicity is used. Suitable mutants are described, e.g., in WO 95/17211 (Arg-7-Lys CT mutant), WO 96/06627 (Arg-192-Gly LT mutant), and WO 95/34323 (Arg-9-Lys and Glu-129-Gly PT mutant). Additional LT mutants that are used in the methods and
5 compositions of the invention include, e.g., Ser-63-Lys, Ala-69-Gly, Glu-110-Asp, and Glu-112-Asp mutants. Other adjuvants, such as a bacterial monophosphoryl lipid A (MPLA) of, e.g., *E. coli*, *Salmonella minnesota*, *Salmonella typhimurium*, or *Shigella flexneri*; saponins, or polylactide glycolide (PLGA)
10 microspheres, is also be used in mucosal administration.

Adjuvants useful for both mucosal and parenteral administrations include polyphosphazene (WO 95/02415), DC-chol (3 b-(N-(N',N'-dimethyl aminomethane)-carbamoyl) cholesterol; U.S. Patent No. 5,283,185 and WO 96/14831) and QS-21
15 (WO 88/09336).

Any pharmaceutical composition of the invention containing a polynucleotide, a polypeptide, a polypeptide derivative, or an antibody of the invention, is manufactured in a conventional manner. In particular, it is formulated with a
20 pharmaceutically acceptable diluent or carrier, e.g., water or a saline solution such as phosphate buffer saline. In general, a diluent or carrier is selected on the basis of the mode and route of administration, and standard pharmaceutical practice. Suitable pharmaceutical carriers or diluents, as well as
25 pharmaceutical necessities for their use in pharmaceutical formulations, are described in *Remington's Pharmaceutical Sciences*, a standard reference text in this field and in the USP/NF.

The invention also includes methods in which *Chlamydia*
30 infection are treated by oral administration of a *Chlamydia* polypeptide of the invention and a mucosal adjuvant, in combination with an antibiotic, an antacid, sucralfate, or a combination thereof. Examples of such compounds that can be administered with the vaccine antigen and the adjuvant are

antibiotics, including, e.g., macrolides, tetracyclines, and derivatives thereof (specific examples of antibiotics that can be used include azithromycin or doxycyclin or immunomodulators such as cytokines or steroids). In addition, compounds
5 containing more than one of the above-listed components coupled together, are used. The invention also includes compositions for carrying out these methods, i.e., compositions containing a *Chlamydia* antigen (or antigens) of the invention, an adjuvant, and one or more of the above-listed compounds, in a
10 pharmaceutically acceptable carrier or diluent.

It has recently been shown that the 60kDa cysteine rich membrane protein contains a sequence cross-reactive with the murine alpha-myosin heavy chain epitope M7A-alpha, an epitope conserved in humans (Bachmaier et al., Science (1999)
15 283:1335). This cross-reactivity is proposed to contribute to the development of cardiovascular disease, so it may be beneficial to remove this epitope, and any other epitopes cross-reactive with human antigens, from the protein if it is to be used as a vaccine. Accordingly, a further embodiment of the
20 present invention includes the modification of the coding sequence, for example, by deletion or substitution of the nucleotides encoding the epitope from polynucleotides encoding the protein, as to improve the efficacy and safety of the protein as a vaccine. A similar approach may be appropriate for
25 any protective antigen found to have unwanted homologies or cross-reactivities with human antigens.

Amounts of the above-listed compounds used in the methods and compositions of the invention are readily determined by one skilled in the art. Treatment/immunization schedules are
30 also known and readily designed by one skilled in the art. For example, the non-vaccine components can be administered on days 1-14, and the vaccine antigen + adjuvant can be administered on days 7, 14, 21, and 28.

EXAMPLES

The above disclosure generally describes the present invention. A more complete understanding can be obtained by reference to the following specific examples. These examples
5 are described solely for purposes of illustration and are not intended to limit the scope of the invention. Changes in form and substitution of equivalents are contemplated as circumstances may suggest or render expedient. Although specific terms have been employed herein, such terms are
10 intended in a descriptive sense and not for purposes of limitation.

Example 1:

15 This example illustrates the preparation of a plasmid vector pCACRMP60 containing the 60kDa cysteine rich membrane protein gene.

The 60kDa cysteine rich membrane protein gene was amplified from *Chlamydia pneumoniae* genomic DNA by polymerase
20 chain reaction (PCR) using a 5' primer
(5' ATAAGAATGCGGCCGCCACCATGTCCAAACTCATCAGACGAGTAG 3') (SEQ ID No:3) and a 3' primer (5' GCGCCGGATCCGATACACGTGGGTATTTTCTGTG 3')
(SEQ ID No:4). The 5' primer contains a Not I restriction site, a ribosome binding site, an initiation codon and a sequence at
25 the 5' end of the 60kDa cysteine rich membrane protein coding sequence. The 3' primer includes the sequence encoding the C-terminal sequence of the 60kDa cysteine rich membrane protein and a Bam HI restriction site. The stop codon was excluded and an additional nucleotide was inserted to obtain an in-frame
30 fusion with the Histidine tag.

After amplification, the PCR fragment was purified using QIAquick™ PCR purification kit (Qiagen) and then digested with Not I and Bam HI and cloned into the pCA-Myc-His eukaryotic

expression vector describe in example 2 (Fig. 3) with transcription under control of the human CMV promoter.

Example 2:

5 This example illustrates the preparation of the eukaryotic expression vector pCA/Myc-His.

Plasmid pcDNA3.1(-)Myc-His C (Invitrogen) was restricted with Spe I and Bam HI to remove the CMV promoter and the remaining vector fragment was isolated. The CMV promoter and
10 intron A from plasmid VR-1012 (Vical) was isolated on a Spe I / Bam HI fragment. The fragments were ligated together to produce plasmid pCA/Myc-His. The Not I/Bam HI restricted PCR fragment containing the 60kDa cysteine rich membrane protein gene was ligated into the Not I and Bam HI restricted plasmid pCA/Myc-His
15 to produce plasmid pCACRMP60 (Fig 3).

The resulting plasmid, pCACRMP60, was transferred by electroporation into *E. coli* XL-1 blue (Stratagene) which was grown in LB broth containing 50 µg/ml of carbenicillin. The plasmid was isolated by Endo Free Plasmid Giga Kit™ (Qiagen)
20 large scale DNA purification system. DNA concentration was determined by absorbance at 260 nm and the plasmid was verified after gel electrophoresis and Ethidium bromide staining and comparison to molecular weight standards. The 5' and 3' ends of the gene were verified by sequencing using a LiCor model 4000 L
25 DNA sequencer and IRD-800 labelled primers.

Example 3:

This example illustrates the immunization of mice to achieve protection against an intranasal challenge of *C. pneumoniae*.
30

It has been previously demonstrated (Yang et. al., 1993) that mice are susceptible to intranasal infection with different isolates of *C. pneumoniae*. Strain AR-39 (Grayston, 1989) was used in Balb/c mice as a challenge infection model to

examine the capacity of chlamydia gene products delivered as naked DNA to elicit a protective response against a sublethal *C. pneumoniae* lung infection. Protective immunity is defined as an accelerated clearance of pulmonary infection.

5 Groups of 7 to 9 week old male Balb/c mice (8 to 10 per group) were immunized intramuscularly (i.m.) plus intranasally (i.n.) with plasmid DNA containing the coding sequence of *C.pneumoniae* 60kDa cysteine rich membrane protein as described in Example 1 and 2. Saline or the plasmid vector
10 lacking an inserted chlamydial gene was given to groups of control animals.

For i.m. immunization alternate left and right quadriceps were injected with 100µg of DNA in 50µl of PBS on three occasions at 0, 3 and 6 weeks. For i.n. immunization,
15 anaesthetized mice aspirated 50µl of PBS containing 50 µg DNA on three occasions at 0, 3 and 6 weeks. At week 8, immunized mice were inoculated i.n. with 5×10^5 IFU of *C. pneumoniae*, strain AR39 in 100µl of SPG buffer to test their ability to limit the growth of a sublethal *C. pneumoniae* challenge.

20 Lungs were taken from mice at days 5 and 9 post-challenge and immediately homogenised in SPG buffer (7.5% sucrose, 5mM glutamate, 12.5mM phosphate pH7.5). The homogenate was stored frozen at -70°C until assay. Dilutions of the homogenate were assayed for the presence of infectious chlamydia
25 by inoculation onto monolayers of susceptible cells. The inoculum was centrifuged onto the cells at 3000rpm for 1 hour, then the cells were incubated for three days at 35°C in the presence of 1µg/ml cycloheximide. After incubation the monolayers were fixed with formalin and methanol then
30 immunoperoxidase stained for the presence of chlamydial inclusions using convalescent sera from rabbits infected with *C.pneumoniae* and metal-enhanced DAB as a peroxidase substrate.

Figure 4 and Table 1 show that mice immunized i.n. and i.m. with pCACRMP60 had chlamydial lung titers less than 1400 in 4 of 4 cases at day 5 and less than 400 in 4 of 4 cases at day 9 whereas the range of values for control mice sham immunized with 5 saline was 1200-2100 IFU/lung (mean 1680) at day 5 and 1100-12400 IFU/lung (mean 4320) at day 9. DNA immunisation *per se* was not responsible for the observed protective effect since another plasmid DNA construct, pCAI422, failed to protect, with lung titers in immunised mice similar to those obtained for 10 saline-immunized control mice. The construct pCAI422 is identical to pCACRMP60 except that the nucleotide sequence encoding the 60kDa cysteine rich membrane protein is replaced with a *C.pneumoniae* nucleotide sequence encoding a non-protective protein.

Table 1

MOUSE	BACTERIAL LOAD (INCLUSION FORMING UNITS PER LUNG) IN THE LUNGS OF BALB/C MICE IMMUNIZED WITH VARIOUS DNA IMMUNIZATION CONSTRUCTS					
	IMMUNIZING CONSTRUCT					
	Saline	Saline	pCACRMP60	pCACRMP60	pCAI42 2	pCAI422
	Day 5	Day 9	Day 5	Day 9	Day 5	Day 9
1	1800	1100	200	300	2200	3100
2	1200	2100	100	0	1300	8100
3	2100	12400	500	200	700	500
4	1900	3900	1300	0	1000	1400
5	1400	2100				
MEAN	1680	4320	525	125	1300	3275
SD	370.14	4628.39	543.905 6	150.00	648.07	3392.52

SEQUENCE LISTING

gat aaa gaa ttt tat ccc tgt gaa gag gga cga tgt caa cct gta gag 411
Asp Lys Glu Phe Tyr Pro Cys Glu Glu Gly Arg Cys Gln Pro Val Glu
80 85 90

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gct	cag	caa	gag	tct	tgc	tac	gga	aga	ttg	tat	tct	gta	aaa	gta	aac	459
Ala	Gln	Gln	Glu	Ser	Cys	Tyr	Gly	Arg	Leu	Tyr	Ser	Val	Lys	Val	Asn	
			95					100					105			
gat	gat	tgc	aac	gta	gaa	att	tgc	cag	tcc	gtt	cca	gaa	tac	gct	act	507
Asp	Asp	Cys	Asn	Val	Glu	Ile	Cys	Gln	Ser	Val	Pro	Glu	Tyr	Ala	Thr	
		110					115					120				
gta	gga	tct	cct	tac	cct	att	gaa	atc	ctt	gct	ata	ggc	aaa	aaa	gat	555
Val	Gly	Ser	Pro	Tyr	Pro	Ile	Glu	Ile	Leu	Ala	Ile	Gly	Lys	Lys	Asp	
	125					130					135					
tgt	gtt	gat	gtt	gtg	att	aca	caa	cag	cta	cct	tgc	gaa	gct	gaa	ttc	603
Cys	Val	Asp	Val	Val	Ile	Thr	Gln	Gln	Leu	Pro	Cys	Glu	Ala	Glu	Phe	
140					145					150					155	
gta	agc	agt	gat	cca	gaa	aca	act	cct	aca	agt	gat	ggg	aaa	tta	gtc	651
Val	Ser	Ser	Asp	Pro	Glu	Thr	Thr	Pro	Thr	Ser	Asp	Gly	Lys	Leu	Val	
				160					165					170		
tgg	aaa	atc	gat	cgc	ctg	ggc	gca	gga	gat	aaa	tgc	aaa	att	act	gta	699
Trp	Lys	Ile	Asp	Arg	Leu	Gly	Ala	Gly	Asp	Lys	Cys	Lys	Ile	Thr	Val	
			175					180					185			
tgg	gta	aaa	cct	ctt	aaa	gaa	ggc	tgc	tgc	ttc	aca	gct	gct	act	gta	747
Trp	Val	Lys	Pro	Leu	Lys	Glu	Gly	Cys	Cys	Phe	Thr	Ala	Ala	Thr	Val	
		190					195					200				
tgt	gct	tgc	cca	gag	ctc	cgt	tct	tat	act	aaa	tgc	ggc	caa	cca	gcc	795
Cys	Ala	Cys	Pro	Glu	Leu	Arg	Ser	Tyr	Thr	Lys	Cys	Gly	Gln	Pro	Ala	
	205					210					215					
att	tgt	att	aag	caa	gaa	gga	cct	gac	tgt	gct	tgc	cta	aga	tgc	cct	843
Ile	Cys	Ile	Lys	Gln	Glu	Gly	Pro	Asp	Cys	Ala	Cys	Leu	Arg	Cys	Pro	
220					225					230					235	
gta	tgc	tac	aaa	atc	gaa	gta	gtg	aac	aca	gga	tct	gct	att	gcc	cgt	891
Val	Cys	Tyr	Lys	Ile	Glu	Val	Val	Asn	Thr	Gly	Ser	Ala	Ile	Ala	Arg	
				240				245						250		
aac	gta	act	gta	gat	aat	cct	gtt	ccc	gat	ggc	tat	tct	cat	gca	tct	939
Asn	Val	Thr	Val	Asp	Asn	Pro	Val	Pro	Asp	Gly	Tyr	Ser	His	Ala	Ser	
			255					260					265			
ggc	caa	aga	gtt	ctc	tct	ttt	aac	tta	gga	gac	atg	aga	cct	ggc	gat	987
Gly	Gln	Arg	Val	Leu	Ser	Phe	Asn	Leu	Gly	Asp	Met	Arg	Pro	Gly	Asp	
		270					275					280				
aaa	aag	gta	ttt	aca	gtt	gag	ttc	tgc	cct	caa	aga	aga	ggc	caa	atc	1035
Lys	Lys	Val	Phe	Thr	Val	Glu	Phe	Cys	Pro	Gln	Arg	Arg	Gly	Gln	Ile	
	285					290					295					
act	aac	gtt	gct	act	gta	act	tac	tgc	ggc	gga	cac	aaa	tgt	tct	gca	1083
Thr	Asn	Val	Ala	Thr	Val	Thr	Tyr	Cys	Gly	Gly	His	Lys	Cys	Ser	Ala	
300					305					310					315	

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aat gta act aca gtt gtt aat gag cct tgt gta caa gta aat atc tct	1131
Asn Val Thr Thr Val Val Asn Glu Pro Cys Val Gln Val Asn Ile Ser	
320 325 330	
ggg gct gat tgg tct tac gta tgt aaa cct gtg gag tac tct atc tca	1179
Gly Ala Asp Trp Ser Tyr Val Cys Lys Pro Val Glu Tyr Ser Ile Ser	
335 340 345	
gta tcg aat cct gga gac ttg gtt ctt cat gat gtc gtg atc caa gat	1227
Val Ser Asn Pro Gly Asp Leu Val Leu His Asp Val Val Ile Gln Asp	
350 355 360	
aca ctc cct tct ggt gtt aca gta ctc gaa gct cct ggt gga gag atc	1275
Thr Leu Pro Ser Gly Val Thr Val Leu Glu Ala Pro Gly Gly Glu Ile	
365 370 375	
tgc tgt aat aaa gtt gtt tgg cgt att aaa gaa atg tgc cca gga gaa	1323
Cys Cys Asn Lys Val Val Trp Arg Ile Lys Glu Met Cys Pro Gly Glu	
380 385 390 395	
acc ctc cag ttt aaa ctt gta gtg aaa gct caa gtt cct gga aga ttc	1371
Thr Leu Gln Phe Lys Leu Val Val Lys Ala Gln Val Pro Gly Arg Phe	
400 405 410	
aca aat caa gtt gca gta act agt gag tct aac tgc gga aca tgt aca	1419
Thr Asn Gln Val Ala Val Thr Ser Glu Ser Asn Cys Gly Thr Cys Thr	
415 420 425	
tct tgc gca gaa aca aca aca cat tgg aaa ggt ctt gca gct acc cat	1467
Ser Cys Ala Glu Thr Thr Thr His Trp Lys Gly Leu Ala Ala Thr His	
430 435 440	
atg tgc gta tta gac aca aat gat cct atc tgt gta gga gaa aat act	1515
Met Cys Val Leu Asp Thr Asn Asp Pro Ile Cys Val Gly Glu Asn Thr	
445 450 455	
gtc tat cgt atc tgt gta act aac cgt ggt tct gct gaa gat act aac	1563
Val Tyr Arg Ile Cys Val Thr Asn Arg Gly Ser Ala Glu Asp Thr Asn	
460 465 470 475	
gta tct tta atc ttg aag ttc tca aaa gaa ctt cag cca ata gct tct	1611
Val Ser Leu Ile Leu Lys Phe Ser Lys Glu Leu Gln Pro Ile Ala Ser	
480 485 490	
tca ggt cca act aaa gga acg att tca ggt aat acc gtt gtt ttc gac	1659
Ser Gly Pro Thr Lys Gly Thr Ile Ser Gly Asn Thr Val Val Phe Asp	
495 500 505	
gct tta cct aaa ctc ggt tct aag gaa tct gta gag ttt tct gtt acc	1707
Ala Leu Pro Lys Leu Gly Ser Lys Glu Ser Val Glu Phe Ser Val Thr	
510 515 520	
ttg aaa ggt att gct ccc gga gat gct cgc ggc gaa gct att ctt tct	1755
Leu Lys Gly Ile Ala Pro Gly Asp Ala Arg Gly Glu Ala Ile Leu Ser	
525 530 535	

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 Ser Asp Thr Leu Thr Ser Pro Val Ser Asp Thr Glu Asn Thr His Val
 540 545 550 555

tat taa attctaagga attatcctaa agcagagcga tattccgctc tgcttttagga 1859
 Tyr

tagctttcaa agaagtaccg ctttagtacc ttacgtacta aagcggtttt tttgttttat 1919

aagctcttca atccaatcgt agagtttctt aatcaaagat attatttaag tttctgaaat 1979

cctaagattt attttaaaag cccatctttt taggtatgta attaaaattt ttaattaagc 2039

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<213> Chlamydia pneumoniae

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 35 40 45

Val Pro Met Thr Ala Lys Lys Val Arg Leu Val Arg Arg Asn Lys Gln
 50 55 60

Pro Val Glu Gln Lys Ser Arg Gly Ala Phe Cys Asp Lys Glu Phe Tyr
 65 70 75 80

Pro Cys Glu Glu Gly Arg Cys Gln Pro Val Glu Ala Gln Gln Glu Ser
 85 90 95

Cys Tyr Gly Arg Leu Tyr Ser Val Lys Val Asn Asp Asp Cys Asn Val
 100 105 110

Glu Ile Cys Gln Ser Val Pro Glu Tyr Ala Thr Val Gly Ser Pro Tyr
 115 120 125

Pro Ile Glu Ile Leu Ala Ile Gly Lys Lys Asp Cys Val Asp Val Val
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Glu Thr Thr Pro Thr Ser Asp Gly Lys Leu Val Trp Lys Ile Asp Arg
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Leu	Gly	Ala	Gly	Asp	Lys	Cys	Lys	Ile	Thr	Val	Trp	Val	Lys	Pro	Leu
			180					185					190		
Lys	Glu	Gly	Cys	Cys	Phe	Thr	Ala	Ala	Thr	Val	Cys	Ala	Cys	Pro	Glu
		195					200					205			
Leu	Arg	Ser	Tyr	Thr	Lys	Cys	Gly	Gln	Pro	Ala	Ile	Cys	Ile	Lys	Gln
	210					215					220				
Glu	Gly	Pro	Asp	Cys	Ala	Cys	Leu	Arg	Cys	Pro	Val	Cys	Tyr	Lys	Ile
225					230					235					240
Glu	Val	Val	Asn	Thr	Gly	Ser	Ala	Ile	Ala	Arg	Asn	Val	Thr	Val	Asp
				245					250					255	
Asn	Pro	Val	Pro	Asp	Gly	Tyr	Ser	His	Ala	Ser	Gly	Gln	Arg	Val	Leu
			260					265					270		
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		275					280					285			
Val	Glu	Phe	Cys	Pro	Gln	Arg	Arg	Gly	Gln	Ile	Thr	Asn	Val	Ala	Thr
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Val	Thr	Tyr	Cys	Gly	Gly	His	Lys	Cys	Ser	Ala	Asn	Val	Thr	Thr	Val
305					310					315					320
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				325					330					335	
Tyr	Val	Cys	Lys	Pro	Val	Glu	Tyr	Ser	Ile	Ser	Val	Ser	Asn	Pro	Gly
			340					345					350		
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				405					410					415	
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			420					425					430		
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		435					440					445			
Thr	Asn	Asp	Pro	Ile	Cys	Val	Gly	Glu	Asn	Thr	Val	Tyr	Arg	Ile	Cys
	450					455					460				
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465					470					475					480

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Lys Phe Ser Lys Glu Leu Gln Pro Ile Ala Ser Ser Gly Pro Thr Lys
485 490 495

Gly Thr Ile Ser Gly Asn Thr Val Val Phe Asp Ala Leu Pro Lys Leu
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<213> primer

<400> 3

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<210> 4
<211> 34
<212> DNA
<213> primer

<400> 4

gcgccggatc cgatacacgt gggtatttttc tgtg 34

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CLAIMS:

1. A vaccine vector comprising an isolated nucleic acid molecule operably linked to a promoter functional in a mammalian cell, wherein the nucleic acid molecule encodes a polypeptide is selected from any of:

(a) SEQ ID No:2;

(b) an immunogenic fragment consisting of at least 50 consecutive amino acids from SEQ ID No:2; and

(c) a polypeptide which has been modified without loss of immunogenicity, wherein said modified polypeptide is at least 90% identical in amino acid sequence over the full length of SEQ ID No:2;

wherein the polypeptide, when used to immunize an animal, confers immunoprotection to the animal against Chlamydia.

2. The vaccine vector of claim 1 wherein the nucleic acid molecule encodes an immunogenic fragment consisting of at least 50 consecutive amino acids from SEQ ID No:2.

3. The vaccine vector of claim 1 wherein the nucleic acid molecule encodes a polypeptide which has been modified without loss of immunogenicity, wherein said modified polypeptide is at least 90% identical in amino acid sequence to SEQ ID No:2.

4. The vaccine vector of claim 1 comprising the nucleotide sequence set forth in SEQ ID No:1 operably linked to a promoter functional in a mammalian cell.

5. The vaccine vector of any one of claims 1 to 4 further comprising another nucleic acid encoding an additional polypeptide which enhances the immune response to

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the polypeptide having SEQ ID No:2, wherein the additional polypeptide is a Chlamydia polypeptide.

6. The vaccine vector of any one of claims 1 to 5 wherein the nucleic acid molecule encodes SEQ ID No:2.

5 7. The vaccine vector of any one of claims 1 to 6 wherein the promoter is a viral promoter.

8. The vaccine vector of claim 7 wherein the promoter is cytomegalovirus (CMV) promoter.

9. A vaccine comprising the vaccine vector according
10 to any one of claims 1 to 8 and a pharmaceutically acceptable carrier.

10. A vaccine comprising the vaccine vector according to any one of claims 1 to 8 and an adjuvant.

11. The vaccine vector according to any one of
15 claims 1 to 8 in unit dosage form.

12. The vaccine vector according to any one of claims 1 to 8 which is unable to replicate in mammalian cells and unable to integrate in a mammalian genome.

13. The vaccine vector according to any one of
20 claims 1 to 8 which, when used to immunize mice, reduces Chlamydial lung titer by at least 50% at 5 days post-challenge in mice intranasally infected with *C. pneumoniae*, compared to mice not immunized with the vaccine vector.

14. A vaccine composition comprising a protein and a
25 compound pharmaceutically acceptable for use in a vaccine, wherein the protein comprises the amino acid sequence selected from any of:

(a) SEQ ID No:2;

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(b) an immunogenic fragment consisting of at least 50 consecutive amino acids from SEQ ID No:2; and

(c) a polypeptide which has been modified without loss of immunogenicity, wherein said modified polypeptide is
5 at least 90% identical in amino acid sequence over the full length of SEQ ID No:2;

wherein the amino acid sequence, when used to immunize an animal, confers immunoprotection to the animal against Chlamydia, and

10 wherein the compound facilitates delivery and/or enhance an immune response to the part of the protein having the amino acid sequence.

15. The composition according to claim 14, wherein the protein comprises the amino acid sequence set forth in SEQ
15 ID No:2.

16. The composition according to claim 14, wherein the protein comprises an immunogenic fragment consisting of at least 50 consecutive amino acids from SEQ ID No:2 and wherein the immunogenic fragment, when used to immunize an
20 animal, confers immunoprotection to the animal against Chlamydia.

17. The composition according to claim 14, wherein the protein comprises a polypeptide which has been modified without loss of immunogenicity, wherein said modified
25 polypeptide is at least 90% identical in amino acid sequence over the full length of SEQ ID No:2.

18. The composition according to claim 14 wherein the protein is a fusion protein comprising the amino acid sequence fused with a heterologous polypeptide.

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19. The composition according to claim 15 wherein the protein is a fusion protein comprising SEQ ID No:2 fused with a heterologous polypeptide.

20. The composition according to claim 16 wherein the
5 protein is a fusion protein comprising the immunogenic fragment fused with a heterologous polypeptide.

21. The composition according to claim 17 wherein the protein is a fusion protein comprising the modified polypeptide fused with a heterologous polypeptide.

10 22. The composition according to any one of claims 18 to 21 wherein the heterologous polypeptide is a peptide tail for purifying the protein.

23. The composition according to any one of claims 14 to 22 wherein the compound is a liposome.

15 24. The composition according to claim 23 wherein the liposome is at least one liposome selected from the group consisting of neutral liposomes, anionic liposomes, microspheres, ISCOMS, and virus-like-particles (VLPs).

25. The composition according to any one of claims 14
20 to 22 wherein the compound is an adjuvant.

26. The composition according to claim 25 which is suitable for parenteral administration.

27. The composition according to claim 26 wherein the adjuvant is at least one adjuvant selected from the group
25 consisting of an aluminum compound, RIBI, polyphosphazene, DC-chol (3 β -(N-(N',N'-dimethyl aminomethane)-carbamoyl) cholesterol) and QS-21.

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28. The composition according to claim 27 wherein the adjuvant is aluminum hydroxide, aluminum phosphate, or aluminum hydroxy phosphate.

29. The composition according to claim 25 which is
5 suitable for mucosal administration.

30. The composition according to claim 29 wherein the adjuvant is at least one adjuvant selected from the group consisting of bacterial toxin, bacterial monophosphoryl lipid A (MPLA), saponin, polylactide glycolide (PLGA)
10 microsphere, polyphosphazene, DC-chol (3 β -(N-(N',N'-dimethyl aminomethane)-carbamoyl) cholesterol), and QS-21.

31. The composition according to claim 30 wherein the adjuvant is at least one bacterial toxin selected from the group consisting of cholera toxin (CT), E. coli heat-labile
15 toxin (LT), Clostridium difficile toxin A, pertussis toxin (PT), and combinations, subunits, toxoids, or mutants thereof that retain adjuvant activity and/or have reduced toxicity.

32. The composition according to claim 31 wherein the
20 adjuvant is at least one bacterial toxin selected from the group consisting of native cholera toxin subunit B (CTB), Arg-7-Lys CT mutant, Arg-192-Gly LT mutant, Arg-9-Lys PT mutant, Glu-129-Gly PT mutant, Ser-63-Lys LT mutant, Ala-69-Gly LT mutant, Glu-110-Asp LT mutant, and Glu-112-Asp LT
25 mutant.

33. The composition according to claim 30 wherein the adjuvant is bacterial monophosphoryl lipid A (MPLA) of E. coli, Salmonella minnesota, Salmonella typhimurium, or Shigella flexneri.

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34. The composition according to any one of claims 14 to 33 in unit dosage form.

35. Use of the vaccine vector as defined in any one of claims 1 to 13 for treating or preventing a Chlamydia
5 infection.

36. Use of the vaccine composition as defined in any one of claims 14 to 34 for treating or preventing a Chlamydia infection.

37. Use of the vaccine vector as defined in any one of
10 claims 1 to 13 or of the vaccine composition as defined in any one of claims 14 to 34, in combination with an antibiotic, an antacid, sucralfate, a cytokine immunomodulator or a combination thereof, for treating or preventing a Chlamydia infection.

15 38. The use according to claim 37 wherein the antibiotic is a macrolide, a tetracycline, or a derivative thereof.

39. The use according to claim 37 wherein the antibiotic is azithromycin or doxycyclin, and wherein
20 the immunomodulator is interleukin-2 (IL-2), interleukin-12 (IL-12), or a steroid.

40. Use of a nucleic acid molecule encoding an amino acid sequence, or of a protein comprising the amino acid sequence, in the manufacture of a vaccine against Chlamydia,
25 wherein the amino acid sequence is selected from any of:

(a) SEQ ID No:2;

(b) an immunogenic fragment consisting of at least 50 consecutive amino acids from SEQ ID No:2; and

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(c) a polypeptide which has been modified without loss of immunogenicity, wherein said modified polypeptide is at least 90% identical in amino acid sequence over the full length of SEQ ID No:2;

5 wherein the amino acid sequence, when used to immunize an animal, confers immunoprotection to the animal against Chlamydia.

41. A vaccination kit comprising the vaccine vector as defined in any one of claims 1 to 13 in unit dosage form and
10 instructions for its use in vaccinating a subject against Chlamydia infection.

42. A vaccination kit comprising the vaccine composition as defined in any one of claims 14 to 34 in unit dosage form and instructions for its use in vaccinating a
15 subject against Chlamydia infection.

43. A vaccination kit comprising the vaccine composition as defined in any one of claims 29 to 33, at least one additional compound selected from the group consisting of an antibiotic, an antacid, sucralfate, and a
20 cytokine immunomodulator, and instructions for using the composition and the additional compound in vaccinating a subject against Chlamydia infection.

44. The kit according to claim 43 wherein the antibiotic is a macrolide, a tetracycline, or a derivative
25 thereof.

45. The kit according to claim 43 wherein the antibiotic is azithromycin or doxycyclin, and wherein the immunomodulator is interleukin-2 (IL-2), interleukin-12 (IL-12), or a steroid.

Figure 1. Nucleotide and amino acid sequences (SEQ ID Nos: 1 and 2)
of the *C. pneumoniae* 60 kDa cysteine rich membrane protein

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atgcataggt agatccct atg tcc aaa ctc atc aga cga gta gtt acg gtc 171
      Met Ser Lys Leu Ile Arg Arg Val Val Thr Val
              1              5              10

ctt gcg cta acg agt atg gcg agt tgc ttt gcc agc ggg ggt ata gag 219
Leu Ala Leu Thr Ser Met Ala Ser Cys Phe Ala Ser Gly Gly Ile Glu
              15              20              25

gcc gct gta gca gag tct ctg att act aag atc gtc gct agt gcg gaa 267
Ala Ala Val Ala Glu Ser Leu Ile Thr Lys Ile Val Ala Ser Ala Glu
              30              35              40

aca aag cca gca cct gtt cct atg aca gcg aag aag gtt aga ctt gtc 315
Thr Lys Pro Ala Pro Val Pro Met Thr Ala Lys Lys Val Arg Leu Val
              45              50              55

cgt aga aat aaa caa cca gtt gaa caa aaa agc cgt ggt gct ttt tgt 363
Arg Arg Asn Lys Gln Pro Val Glu Gln Lys Ser Arg Gly Ala Phe Cys
              60              65              70              75

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Asp Lys Glu Phe Tyr Pro Cys Glu Glu Gly Arg Cys Gln Pro Val Glu
              80              85              90

gct cag caa gag tct tgc tac gga aga ttg tat tct gta aaa gta aac 459
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              95              100              105

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              110              115              120

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Val Gly Ser Pro Tyr Pro Ile Glu Ile Leu Ala Ile Gly Lys Lys Asp
              125              130              135

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Cys Val Asp Val Val Ile Thr Gln Gln Leu Pro Cys Glu Ala Glu Phe
              140              145              150              155

gta agc agt gat cca gaa aca act cct aca agt gat ggg aaa tta gtc 651
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              160              165              170

tgg aaa atc gat cgc ctg ggt gca gga gat aaa tgc aaa att act gta 699
Trp Lys Ile Asp Arg Leu Gly Ala Gly Asp Lys Cys Lys Ile Thr Val
              175              180              185

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Figure 1 (cont'd)

tgg	gta	aaa	cct	ctt	aaa	gaa	ggg	tgc	tgc	ttc	aca	gct	gct	act	gta	747
Trp	Val	Lys	Pro	Leu	Lys	Glu	Gly	Cys	Cys	Phe	Thr	Ala	Ala	Thr	Val	
		190					195					200				
tgt	gct	tgc	cca	gag	ctc	cgt	tct	tat	act	aaa	tgc	ggg	caa	cca	gcc	795
Cys	Ala	Cys	Pro	Glu	Leu	Arg	Ser	Tyr	Thr	Lys	Cys	Gly	Gln	Pro	Ala	
	205					210					215					
att	tgt	att	aag	caa	gaa	gga	cct	gac	tgt	gct	tgc	cta	aga	tgc	cct	843
Ile	Cys	Ile	Lys	Gln	Glu	Gly	Pro	Asp	Cys	Ala	Cys	Leu	Arg	Cys	Pro	
220					225					230					235	
gta	tgc	tac	aaa	atc	gaa	gta	gtg	aac	aca	gga	tct	gct	att	gcc	cgt	891
Val	Cys	Tyr	Lys	Ile	Glu	Val	Val	Asn	Thr	Gly	Ser	Ala	Ile	Ala	Arg	
				240					245					250		
aac	gta	act	gta	gat	aat	cct	gtt	ccc	gat	ggc	tat	tct	cat	gca	tct	939
Asn	Val	Thr	Val	Asp	Asn	Pro	Val	Pro	Asp	Gly	Tyr	Ser	His	Ala	Ser	
			255					260					265			
ggg	caa	aga	gtt	ctc	tct	ttt	aac	tta	gga	gac	atg	aga	cct	ggc	gat	987
Gly	Gln	Arg	Val	Leu	Ser	Phe	Asn	Leu	Gly	Asp	Met	Arg	Pro	Gly	Asp	
		270					275					280				
aaa	aag	gta	ttt	aca	gtt	gag	ttc	tgc	cct	caa	aga	aga	ggg	caa	atc	1035
Lys	Lys	Val	Phe	Thr	Val	Glu	Phe	Cys	Pro	Gln	Arg	Arg	Gly	Gln	Ile	
	285					290					295					
act	aac	gtt	gct	act	gta	act	tac	tgc	ggg	gga	cac	aaa	tgt	tct	gca	1083
Thr	Asn	Val	Ala	Thr	Val	Thr	Tyr	Cys	Gly	Gly	His	Lys	Cys	Ser	Ala	
300					305					310					315	
aat	gta	act	aca	gtt	gtt	aat	gag	cct	tgt	gta	caa	gta	aat	atc	tct	1131
Asn	Val	Thr	Thr	Val	Val	Asn	Glu	Pro	Cys	Val	Gln	Val	Asn	Ile	Ser	
				320					325					330		
ggg	gct	gat	tgg	tct	tac	gta	tgt	aaa	cct	gtg	gag	tac	tct	atc	tca	1179
Gly	Ala	Asp	Trp	Ser	Tyr	Val	Cys	Lys	Pro	Val	Glu	Tyr	Ser	Ile	Ser	
			335					340					345			
gta	tcg	aat	cct	gga	gac	ttg	gtt	ctt	cat	gat	gtc	gtg	atc	caa	gat	1227
Val	Ser	Asn	Pro	Gly	Asp	Leu	Val	Leu	His	Asp	Val	Val	Ile	Gln	Asp	
		350					355					360				
aca	ctc	cct	tct	ggg	gtt	aca	gta	ctc	gaa	gct	cct	ggg	gga	gag	atc	1275
Thr	Leu	Pro	Ser	Gly	Val	Thr	Val	Leu	Glu	Ala	Pro	Gly	Gly	Glu	Ile	
	365					370					375					
tgc	tgt	aat	aaa	gtt	gtt	tgg	cgt	att	aaa	gaa	atg	tgc	cca	gga	gaa	1323
Cys	Cys	Asn	Lys	Val	Val	Trp	Arg	Ile	Lys	Glu	Met	Cys	Pro	Gly	Glu	
380					385					390					395	
acc	ctc	cag	ttt	aaa	ctt	gta	gtg	aaa	gct	caa	gtt	cct	gga	aga	ttc	1371
Thr	Leu	Gln	Phe	Lys	Leu	Val	Val	Lys	Ala	Gln	Val	Pro	Gly	Arg	Phe	
				400					405					410		
aca	aat	caa	gtt	gca	gta	act	agt	gag	tct	aac	tgc	gga	aca	tgt	aca	1419
Thr	Asn	Gln	Val	Ala	Val	Thr	Ser	Glu	Ser	Asn	Cys	Gly	Thr	Cys	Thr	
			415					420					425			

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Figure 1 (cont'd)

tct tgc gca gaa aca aca aca cat tgg aaa ggt ctt gca gct acc cat	1467
Ser Cys Ala Glu Thr Thr Thr His Trp Lys Gly Leu Ala Ala Thr His	
430 435 440	
atg tgc gta tta gac aca aat gat cct atc tgt gta gga gaa aat act	1515
Met Cys Val Leu Asp Thr Asn Asp Pro Ile Cys Val Gly Glu Asn Thr	
445 450 455	
gtc tat cgt atc tgt gta act aac cgt ggt tct gct gaa gat act aac	1563
Val Tyr Arg Ile Cys Val Thr Asn Arg Gly Ser Ala Glu Asp Thr Asn	
460 465 470 475	
gta tct tta atc ttg aag ttc tca aaa gaa ctt cag cca ata gct tct	1611
Val Ser Leu Ile Leu Lys Phe Ser Lys Glu Leu Gln Pro Ile Ala Ser	
480 485 490	
tca ggt cca act aaa gga acg att tca ggt aat acc gtt gtt ttc gac	1659
Ser Gly Pro Thr Lys Gly Thr Ile Ser Gly Asn Thr Val Val Phe Asp	
495 500 505	
gct tta cct aaa ctc ggt tct aag gaa tct gta gag ttt tct gtt acc	1707
Ala Leu Pro Lys Leu Gly Ser Lys Glu Ser Val Glu Phe Ser Val Thr	
510 515 520	
ttg aaa ggt att gct ccc gga gat gct cgc ggc gaa gct att ctt tct	1755
Leu Lys Gly Ile Ala Pro Gly Asp Ala Arg Gly Glu Ala Ile Leu Ser	
525 530 535	
tct gat aca ctg act tca cca gta tca gac aca gaa aat acc cac gtg	1803
Ser Asp Thr Leu Thr Ser Pro Val Ser Asp Thr Glu Asn Thr His Val	
540 545 550 555	
tat taa attctaagga attatcctaa agcagagcga tattccgctc tgcttttagga	1859
Tyr	
tagctttcaa agaagtaccg ctttagtacc ttacgtacta aagcgggtttt tttgttttat	1919
aagctcttca atccaatcgt agagtttctt aatcaaagat attattttaag tttctgaaat	1979
cctaagattt attttaaaaag cccatctttt taggtatgta attaaaattt ttaattaagc	2039
ttttcctagt gtaacctgct tctttaggaa ctacactagg agaacggtat gtcacaaat	2099
ctacatcccg ta	2111

Figure 2: Restriction enzyme analysis of the the *C. pneumoniae* 60 kDa cysteine rich membrane protein.

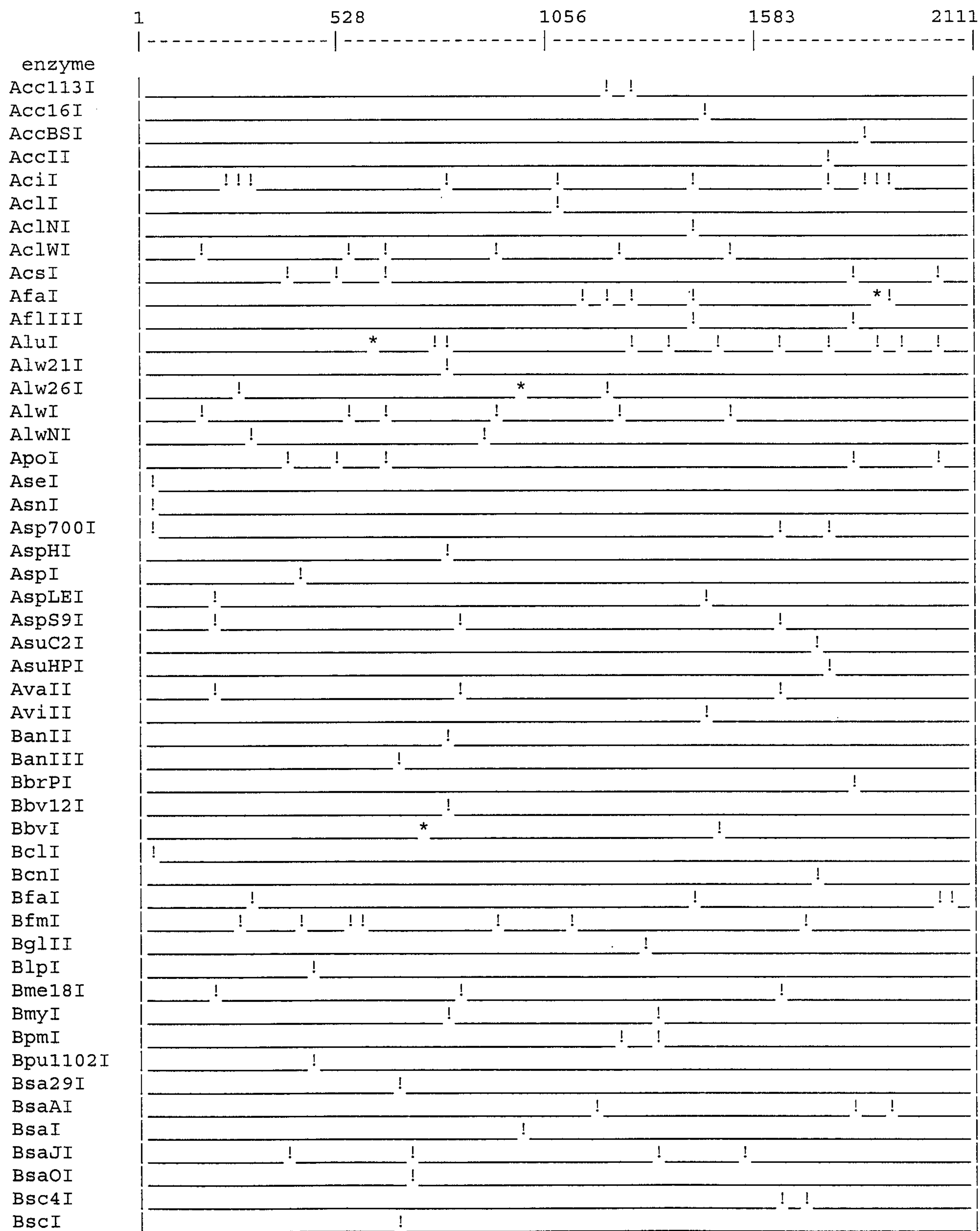


Figure 2 (cont'd)

Bse1I	! ! ! ! !
BseCI	! ! ! ! !
BseDI	! ! ! ! !
BseNI	! ! ! ! !
BsgI	! ! ! ! !
Bsh1236I	! ! ! ! !
Bsh1285I	! ! ! ! !
BsiEI	! ! ! ! !
BsiHKAI	! ! ! ! !
BsiLI	! ! ! ! !
BsiQI	! ! ! ! !
BsiSI	! ! ! ! !
BsiYI	! ! ! ! !
BsiZI	! ! ! ! !
BslI	! ! ! ! !
BsmAI	! ! ! ! !
BsmFI	! ! ! ! !
Bsp106I	! ! ! ! !
Bsp1286I	! ! ! ! !
Bsp1407I	! ! ! ! !
Bsp1720I	! ! ! ! !
BspHI	! ! ! ! !
BspLU11I	! ! ! ! !
BspMI	! ! ! ! !
BspXI	! ! ! ! !
BsrBI	! ! ! ! !
BsrGI	! ! ! ! !
BsrI	! ! ! ! !
BsrSI	! ! ! ! !
Bst2UI	! ! ! ! !
Bst71I	! ! ! ! !
BstBAI	! ! ! ! !
BstDEI	! ! ! ! !
BstDSI	! ! ! ! !
BstMCI	! ! ! ! !
BstNSI	! ! ! ! !
BstOI	! ! ! ! !
BstSFI	! ! ! ! !
BstSNI	! ! ! ! !
BstUI	! ! ! ! !
BstX2I	! ! ! ! !
BstYI	! ! ! ! !
Bsu15I	! ! ! ! !
Bsu6I	! ! ! ! !
Cac8I	! ! ! ! !
CelII	! ! ! ! !
Cfr13I	! ! ! ! !
Clal	! ! ! ! !
CviJI	! ! ! ! !
DdeI	! ! ! ! !
DpnI	! ! ! ! !
DraI	! ! ! ! !
DraII	! ! ! ! !
DsaI	! ! ! ! !
Eam1104I	! ! ! ! !
EarI	! ! ! ! !
Eco105I	! ! ! ! !
Eco24I	! ! ! ! !

Figure 2 (cont'd)

Eco31I	_____!_____
Eco47I	_____!_____!_____!
Eco57I	_____!_____!*
Eco72I	_____!_____!
EcoRI	_____!_____
EcoRII	_____!_____!_____!_____!_____!
EcoT22I	_____!_____
EcoT38I	_____!_____
FauI	_____!_____
FbaI	!_____
Fnu4HI	_____!_____!_____*
FokI	_____!_____!
FriOI	_____!_____
Fsp4HI	_____!_____!_____*
FspI	_____!_____!
GsuI	_____!_____!_____
HaeIII	_____!_____
HapII	_____!_____!
HgaI	_____!_____!
HgiEI	_____!_____!_____!
HhaI	_____!_____!_____!
Hin6I	_____!_____!_____!
HindII	_____!_____!_____
HindIII	_____!_____!
HinfI	_____!_____!_____!_____!_____!
HpaII	_____!_____!
HphI	_____!_____!
Hsp92II	_____!_____!_____!_____!
HspAI	_____!_____!_____!
ItaI	_____!_____!_____*
Ksp22I	!_____
Ksp632I	_____!_____!_____!
Kzo9I	!_____!_____!_____!_____!_____!_____!
MaeI	_____!_____!_____!
MaeII	_____!_____!_____!_____!_____!
MaeIII	_____!_____!_____!_____!_____!_____!
MboI	!_____!_____!_____!_____!_____!_____!
MboII	_____!_____!_____!_____!_____!_____!
MflI	_____!_____!_____!
MnlI	_____!_____!_____!_____!_____!
Mph1103I	_____!_____!
MroXI	!_____!_____!
MseI	!!_____!_____!_____!_____!_____!_____!_____!_____!_____!_____!
MspA1I	_____!!_____!_____!
MspR9I	_____!_____!_____!_____!_____!
MvnI	_____!_____!
MwoI	_____!!_____!_____*
NciI	_____!_____!
NdeI	_____!_____!
NlaIII	_____!_____!_____!_____!
NsiI	_____!_____!
NspBII	_____!!_____!_____!
NspI	_____!_____!_____!
PacI	_____!_____!
PaiI	_____!_____!
Ple19I	_____!_____!
PleI	_____!_____!_____!
PmaCI	_____!_____!

Figure 2 (cont'd)

PmeI	_____!
PmlI	_____!
Ppu10I	_____!
PpuMI	_____!
PshBI	!_____
Psp124BI	_____!
Psp1406I	_____!
Psp5II	_____!
PspPPI	_____!
PvuI	_____!
PvuII	_____!
RcaI	_____!
RsaI	_____!_!_!_!_!_!_*
SacI	_____!
SapI	_____!
Sau96I	_____!_____!
ScaI	_____!_!_!
ScrFI	_____!_____!_____!_!_!_!
SduI	_____!_____!
SfaNI	_____!_!_!
SfcI	_____!_!_!_!_!_!_!
SnaBI	_____!_____!
SpeI	_____!
Sse9I	*!_____!_!_!_!_!_!_*
SspBI	_____!_____!
TaiI	_____!_____!_____!_!_!
TaqI	_____!_____!_!_!
TfiI	_____!_____!
ThaI	_____!
Tru1I	!!_____!_!_!_!_!_!_!!_!_!_!*
Tru9I	!!_____!_!_!_!_!_!_!!_!_!_!*
TscI	_____!_____!_____!
TseI	_____*
Tsp509I	*!_____!_!_!_!_!_!_*
TspEI	*!_____!_!_!_!_!_!_*
TspRI	_____!
Tth111I	_____!
VspI	!_____
XhoII	_____!_____!_____!
XmnI	!_____!_____!
Zsp2I	_____!

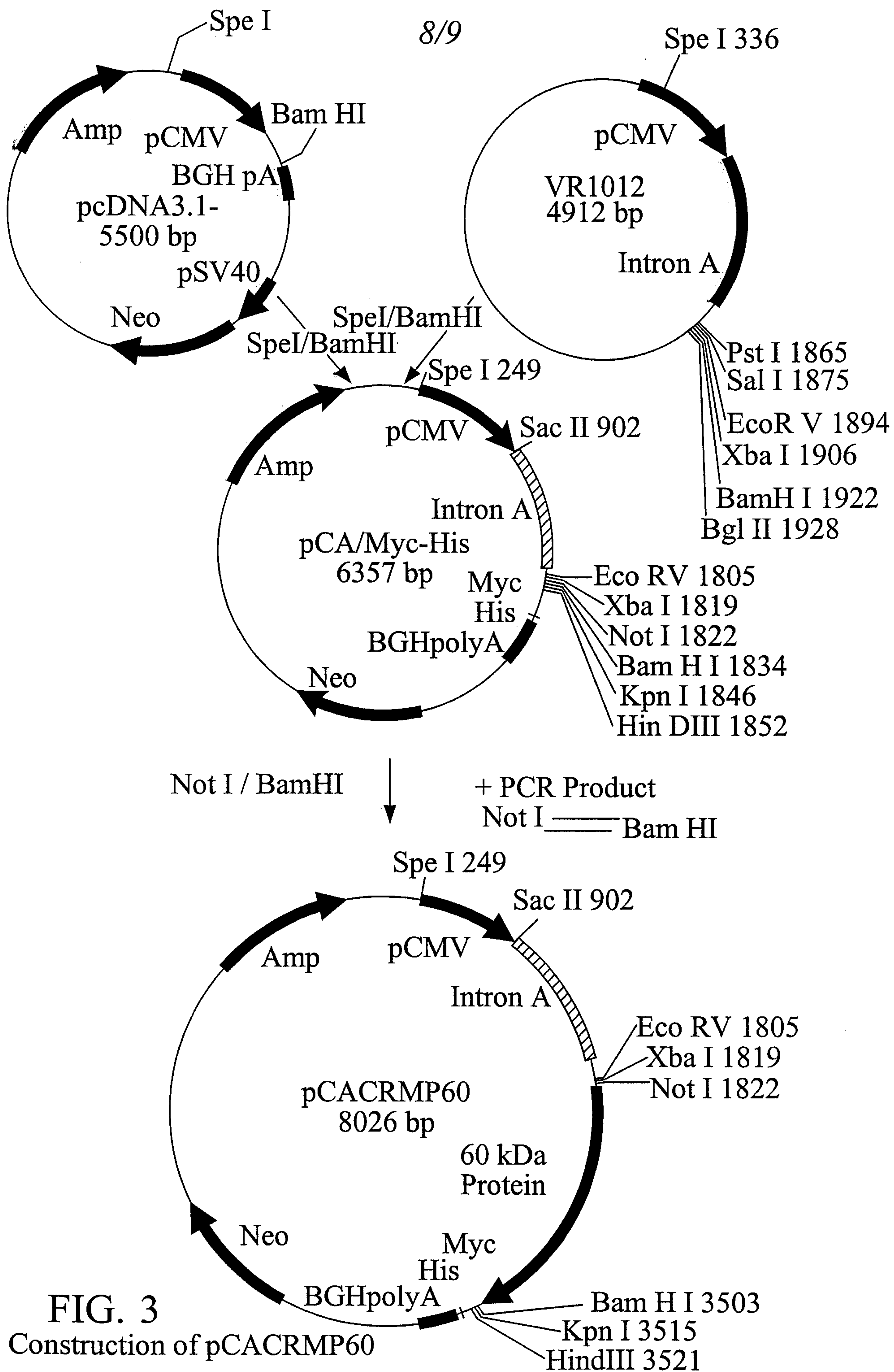


FIG. 3
Construction of pCACRMP60

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Figure 4: Protective efficacy of DNA immunization with pCACRMP60 against intranasal challenge of *C. pneumoniae*

