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FGFR-Fc FUSION PROTEIN AND USE THEREOF**FIELD OF INVENTION**

The present invention belongs to the field of biotechnology and relates to the treatment of diseases, especially the treatment of FGF overexpression-related diseases. Particularly, the present invention relates to FGFR-Fc fusion proteins and the use thereof in the treatment of angiogenesis regulation-related diseases. More particularly, the present invention relates to isolated soluble FGFR-Fc fusion proteins and their applications in manufacture of the medicament for the treatment of angiogenesis regulation-related diseases.

BACKGROUND OF THE INVENTION

Angiogenesis is one of the primary factors resulting in the growth and metastasis of malignant tumor [1]. The process of angiogenesis is regulated by many factors, among which some factors promote angiogenesis, while some factors inhibit angiogenesis, and as a result, the regulation of angiogenesis is a very complicated dynamic equilibrium process [2]. The anti-angiogenesis treatment is intended to control the growth of tumor by blocking angiogenic stimulating factors or preventing angiogenesis in the tumor using angiogenesis inhibitors. At present, a large amount of angiogenic stimulating factors are known, such as, for example, vascular endothelial growth factor (VEGF), fibroblast growth factor (FGF), hepatocyte growth factor (HGF) etc., which may stimulate the division and differentiation of vascular endothelial cells and the morphogenesis of blood vessels. Among these factors mentioned above, it is now known that VEGF is the most angiogenesis-specific and the most effective growth factor [3, 4].

In a hypoxic environment inside the tumor tissue, a plenty of VEGFs are secreted by the tumor cells, which induce the division and migration of vascular

endotheliocytes, finally resulting in the establishment of tumor vascular network. It has been demonstrated by a lot of animal experiments that the inhibition of VEGF may prevent angiogenesis, and further inhibit the growth of tumor. For this reason, VEGF and its receptors become important targets for anti-angiogenesis medicaments. At present, anti-angiogenesis medicaments demonstrated in clinical trials to have remarkable efficacy include Bevacizumab (under the trade name of Avastin), which is able to block VEGF directly and inhibit the tumor angiogenesis. Bevacizumab was approved for marketing by FDA in 2004, and as a first-line drug for rectal cancer, it is the first marketing-approved drug that plays a role in anticarcinogenesis by inhibiting angiogenesis. Avastin is a humanized anti-VEGF monoclonal antibody, which is produced by a famous US biotechnology company Genentech. In a large-scale Phase III clinical trial, the combined therapy by Avastin and chemotherapy may significantly extend the survival time of the patients suffered from many kinds of cancers, including rectal cancer, lung cancer, breast cancer and renal cancer, etc. [5, 6] The clinical success of Avastin is a landmark, demonstrating that the anti-angiogenesis treatment using tumor vascular system as the target is a clinically effective measure and provide a new path for the tumor treatment. In western countries, Avastin has already been widely used for tumor therapy and is one of the drugs holding the largest global market share.

Besides Avastin, several drugs for anti-VEGF signaling are also under the late phase of human clinical trial in foreign countries, which are expected for clinical application in the next several years. Among others, Aflibercept (also called as VEGF-Trap), developed by the cooperation between US biotechnology company Regeneron and Sanofi-Aventis, is now under the large-scale Phase III clinical trial [7]. An anti-VEGF receptor II (VEGFR2) monoclonal antibody drug IMC-1121B (Imclone) is also under the Phase III clinical trial [8]. In China, the development of new medicaments now also focuses on the anti-tumor medicament using anti-angiogenesis as the target. Medicaments using VEGF and its receptor, or other angiogenic targets are under the development by several Chinese companies or

research institutions. These new drugs will definitely provide new choices for cancer therapy and new hope for the patients.

Great progress has been achieved in the clinical treatment of tumor using anti-VEGF medicament, however, it has also been shown by the clinical trial that the anti-VEGF treatment are also considerably limited. From the point of the effect of tumor treatment, Avastin may extend the half survival time of the colon cancer patient for about 3-4 months [9, 10], and extend the half survival time of the breast cancer patient for about 7-8 months [11], and thus, Avastin cannot effectively inhibit the growth of tumor blood vessel in a long term. Therefore, the problem how to further improve the effect of clinical treatment using anti-angiogenesis method need to be solved by tumor investigators and is also the main point of the research and development of the next generation anti-angiogenesis medicament.

The primary causes resulting in the failure of anti-VEGF treatment or the appearance of resistance may depend on the regulation of tumor angiogenesis by a plurality of factors. Although VEGF plays an important role in angiogenesis, it is not the only angiogenesis stimulating factor. Meanwhile, owing to the heterogeneity of tumor cells, the complexity of tumor microenvironment and the compensatory response mechanism of body, when the activity of VEGF is inhibited for a long period of time, other angiogenesis stimulating factors would be expressed [12], and thus the growth of tumor blood vessel is no longer dependent on VEGF signaling path. The variation of angiogenesis factors expressed by the tumor was studied during anti-VEGFR2 treatment for pancreatic tumor by Prof. Hanahan's group (University of California, San Francisco, US), indicating that the expression of several genes changed during anti-VEGF treatment, in which the expression of FGF-2 significantly increased. It has been shown that the expression of FGF, especially FGF-2, increased significantly in the tumor resistant to anti-VEGF treatment so that angiogenesis was activated again and the tumor repopulation was inhibited after blocking FGF signal pathway [13]. It may be seen that the over-expression of FGF-2 is closely related to the ability of tumor to escape from anti-VEGF treatment. Therefore, we believe the

angiogenesis of tumor may be efficiently prevented and the tumor growth may be inhibited by blocking FGF pathway, so that angiogenesis-related diseases can be treated alone by anti-FGF treatment or by a combination therapy of anti-FGF and anti-VEGF treatment.

Fibroblast growth factor (FGF) is a growth factor family for heparin-binding, and there are 22 family members (FGF 1-14, 16-23) in the mammals. FGF plays an important role in many biological functions, for example, cell proliferation, differentiation, migration, angiogenesis and tumorigenesis etc. Fibroblast growth factor receptor (FGFR) is the receptor which binds the family members of fibroblast growth factor. FGF may bind FGFR and activate the downstream signal pathway, which plays an important role in a physiological and pathological process, such as embryogenesis, development, vasculogenesis, vasodilatation, neuroregulation, ischemia protection, wound healing and tumorigenesis etc. [14, 15] It has already been demonstrated that overexpression of FGF/FGFR *in vivo* is closely related to many diseases including tumors (such as fibroma, neuroglioma, melanoma, prostate carcinoma, lymphomata, leukaemia, urinary system cancer etc.), skeletal system diseases (dwarfism, craniosynostosis, achondroplasia, acanthosis nigricans) and renal failure etc. It has already been reported that increased expression level of FGF and its receptor may directly promote the survival and proliferation of tumor cells, and the survival of hepatic carcinoma cells is significantly reduced by down-regulation of FGF by siRNA [22]. Therefore, it is expected to block FGF pathway by constructing an FGFR-Fc fusion protein that is able to antagonize FGF, which is of the potential for treating FGF overexpression-related diseases.

At present, few researches focus on the development of new anti-angiogenesis medicament using FGF and its receptor as the target in clinical trials. For example, FP-1039, a fusion protein composed of whole extracellular domain of human FGFR1 and human IgG1 Fc fragment, is developed by a US company Five Prime and now in volunteer recruitment stage of Phase I clinical trial. However, it has been suggested by researches of Wang and Olsen that the first Ig-like domain of the

extracellular domain of human FGFR1 and the linking fragment between the first and the second Ig-like domain of the extracellular domain of human FGFR1 may inhibit binding of FGFR1 and FGF [20, 21].

Therefore, it is expected to block FGF pathway by constructing an FGFR-Fc fusion protein that is able to antagonize FGF so that angiogenesis may be efficiently inhibited or it may act on tumor cells directly and inhibit their growth, and it is of the potential for treating FGF overexpression-related diseases to cure angiogenesis-related diseases such as tumors.

It is to be understood that, if any prior art publication is referred to herein, such reference does not constitute an admission that the publication forms a part of the common general knowledge in the art, in Australia or any other country.

SUMMARY

The space structure of protein is closely related to its biological function. The FGF binding capacity is directly influenced by difference among the conformations of each Ig-like domain of the extracellular domain of FGFR and the linking fragment. Different fusion proteins, composed of the FGFR extracellular domain fragments of various lengths and IgG Fc, are constructed by means of genetic engineering to obtain fusion proteins with different conformations, so that the fusion protein with high efficiency of FGF binding and biological activity can be screened.

There are four FGFR genes in the mammals: fgfr1-fgfr4. Fibroblast growth factor receptor is composed of the extracellular domain, transmembrane domain and intracellular domain. There are many members in FGFR family, which have similar extracellular domain but vary in the ligand binding property and kinase domain. Their extracellular domains include three immunoglobulin-like (Ig-like) domains: the first Ig-like domain, the second Ig-like domain and the third Ig-like domain, and there is a sequence between the first and the second Ig-like domain, which is referred as the intermediate functional sequence of the Ig-like domain of FGFR (IFS for short herein)

in this specification. The intermediate functional sequence may comprise one acidic amino acid segment, which is referred as acidic box (AB).

In a first aspect, the invention provides an isolated soluble fusion protein of fibroblast growth factor receptor (FGFR), comprising: a part derived from the FGFR extracellular domain and an immunoglobulin Fc region, wherein the part derived from the FGFR extracellular domain consists of an amino acid sequence corresponding to positions 1-241 of SEQ ID NO: 12, positions 1-230 of SEQ ID NO: 13, or positions 1-224 of SEQ ID NO: 14 or encoded by a nucleotide sequence corresponding to positions 1-723 of SEQ ID NO: 19, positions 1-690 of SEQ ID NO: 20, or positions 1-672 of SEQ ID NO: 21.

In a second aspect, the invention provides an isolated soluble fusion protein of fibroblast growth factor receptor (FGFR), wherein said protein consists of an amino acid sequence of any one of SEQ ID NOs: 12-14, or is encoded by a nucleic acid molecule consisting of a nucleotide sequence of any one of SEQ ID NOs: 19-21.

Disclosed herein is an isolated soluble fusion protein of fibroblast growth factor receptor (FGFR), which comprises: the part derived from the intermediate functional sequence (also referred as IFS herein) of the Ig-like domain of FGFR, the second Ig-like domain (also referred as D2 herein) of FGFR, the third Ig-like domain (also referred as D3 herein) of FGFR and immunoglobulin Fc region.

Also disclosed is a fusion protein, which comprises or consists of: the part derived from the intermediate functional sequence region of the Ig-like domain of FGFR, the second Ig-like domain of FGFR, the third Ig-like domain of FGFR and immunoglobulin Fc region. In some embodiments, the part derived from IFS contains no acidic box. In some other embodiments, the part of IFS has the amino acid sequence of position 134 to position 162, position 145 to position 162 or position 151 to position 162 of SEQ ID NO: 1, or has the amino acid sequence sharing at least 90% identity, preferably 93%, 95%, 97%, 98% or 99% identity, with the amino acid sequence of position 134 to position 162, position 145 to position 162 or position 151 to position 162 of SEQ ID NO: 1.

Also disclosed is a fusion protein, which comprises or consists of: the first Ig-like domain (also referred to as D1 herein) of FGFR or a moiety thereof, the part derived from the intermediate functional sequence region of the Ig-like domain of FGFR, the second Ig-like domain of FGFR, the third Ig-like domain of FGFR and immunoglobulin Fc region. Preferably, said D1 domain or a moiety thereof possesses:

the amino acid sequence corresponding to position 40 to position 118 of SEQ ID NO: 1, or the amino acid sequence sharing at least 70% identity, preferably at least 80%, 90%, 93%, 95%, 97%, 98% or 99% identity with the sequence of position 40 to position 118 of SEQ ID NO: 1; or

the amino acid sequence corresponding to position 77 to position 118 of SEQ ID NO: 1, or the amino acid sequence sharing at least 70% identity, preferably at least 80%, 90%, 93%, 95%, 97%, 98% or 99% identity with the amino acid sequence of position 77 to position 118 of SEQ ID NO: 1.

Also disclosed is a fusion protein, which comprises or consists of: the intermediate functional sequence region of the Ig-like domain of FGFR or a moiety thereof, the second Ig-like domain of FGFR, the third Ig-like domain of FGFR and immunoglobulin Fc region, wherein:

the second Ig-like domain of FGFR has the amino acid sequence corresponding to position 163 to position 247 of SEQ ID NO: 1, or the amino acid sequence sharing at least 70% identity, preferably at least 80%, 90%, 93%, 95%, 97%, 98% or 99% identity with the amino acid sequence of position 163 to position 247 of SEQ ID NO: 1; and/or

the third Ig-like domain of FGFR has the amino acid sequence corresponding to position 270 to position 359 of SEQ ID NO: 1, or the amino acid sequence sharing at least 70% identity, preferably at least 80%, 90%, 93%, 95%, 97%, 98% or 99% identity with the amino acid sequence of position 270 to position 359 of SEQ ID NO: 1.

Also disclosed is a fusion protein, which comprises a region derived from the extracellular domain of FGFR and immunoglobulin Fc region or composed thereof, wherein the region derived from the extracellular domain of FGFR:

(1) comprises an amino acid sequence corresponding to any one of positions 1-353 of SEQ ID NO: 9, positions 1-299 of SEQ ID NO: 10, positions 1-273 of SEQ ID NO: 11, positions 1-241 of SEQ ID NO: 12, positions 1-230 of SEQ ID NO: 13, positions 1-224 of SEQ ID NO: 14 and positions 1-219 of SEQ ID NO: 15, or encoded by a nucleotide sequence corresponding to any one of positions 1-1059 of SEQ ID NO: 16, positions 1-897 of SEQ ID NO: 17, positions 1-819 of SEQ ID NO: 18, positions 1-723 of SEQ ID NO: 19, positions 1-690 of SEQ ID NO: 20, positions 1-672 of SEQ ID NO: 21, and positions 1-657 of SEQ ID NO: 22;

(2) comprises or consists of an amino acid sequence having at least 70% identity, optionally at least 80%, 90%, 93%, 95%, 97%, 98% or 99% identity with an amino acid sequence corresponding to any one of positions 1-353 of SEQ ID NO: 9, positions 1-299 of SEQ ID NO: 10, positions 1-273 of SEQ ID NO: 11, positions 1-241 of SEQ ID NO: 12, positions 1-230 of SEQ ID NO: 13, positions 1-224 of SEQ ID NO: 14 and positions 1-219 of SEQ ID NO: 15; or

(3) comprises or consists of an amino acid sequence encoded by a nucleotide sequence having at least 70% identity, optionally at least 80%, 90%, 93%, 95%, 97%, 98% or 99% identity with a nucleotide sequence corresponding to any one of positions 1-1059 of SEQ ID NO: 16, positions 1-879 of SEQ ID NO: 17, positions 1-819 of SEQ ID NO: 18, positions 1-723 of SEQ ID NO: 19, positions 1-690 of SEQ ID NO: 20, positions 1-672 of SEQ ID NO: 21 and positions 1-657 of SEQ ID NO: 22.

Also disclosed is a fusion protein, said protein:

(1) comprises the amino acid sequence indicated by any one of SEQ ID NOs: 9-15, or the amino acid sequence encoded by the nucleotide sequence indicated by any one of SEQ ID NOs: 16-22;

(2) comprises or consists of the amino acid sequence sharing at least 70% identity, preferably at least 80%, 90%, 93%, 95%, 97%, 98% or 99% identity, with the amino acid sequence indicated by any one of SEQ ID NOs: 9-15; or

(3) comprises or consists of the amino acid sequence encoded by the nucleotide sequence sharing at least 70% identity, preferably at least 80%, 90%, 93%, 95%, 97%, 98% or 99% identity, with the nucleotide sequence indicated by any one of SEQ ID NOs: 16-22.

Preferably, in the fusion protein, the immunoglobulin Fc region is human IgG1 Fc region, and more preferably, it comprises:

the amino acid sequence corresponding to SEQ ID NO: 7, or the amino acid sequence sharing at least 70% identity, preferably at least 80%, 90%, 93%, 95%, 97%, 98% or 99% identity, with the amino acid sequence of SEQ ID NO: 7; or

the amino acid sequence encoded by the nucleotide sequence corresponding to SEQ ID NO: 8, or the amino acid sequence encoded by the nucleotide sequence sharing at least 70% identity, preferably at least 80%, 90%, 93%, 95%, 97%, 98% or 99% identity, with the nucleotide sequence of SEQ ID NO: 8.

In one embodiment of the present invention, the immunoglobulin Fc region is located at the C-terminus of the fusion protein.

Also disclosed is a fusion protein precursor comprising a secretory signal peptide region, for example, VEGFR1 signal peptide region, and preferably, the secretory signal peptide region has the amino acid sequence of position 1 to position 26 of SEQ ID NO: 2 or the amino acid sequence encoded by the nucleotide sequence of SEQ ID NO: 23. Preferably, the signal peptide region is located at the N-terminus of the precursor.

Also disclosed is a fusion protein which sequentially comprises from the N-terminus to the C-terminus: the part derived from IFR, D2, D3 and immunoglobulin Fc region.

In another embodiment, the domains and/or regions involved in the fusion protein of the present invention are linked directly and/or by a linker. In one embodiment, the region derived from the extracellular domain of FGFR and immunoglobulin Fc region are linked directly. In another embodiment, the region derived from the extracellular domain of FGFR and immunoglobulin Fc region are linked by a linker.

In one embodiment, the fusion protein of the present invention inhibits angiogenesis. In another embodiment, the fusion protein of the present invention binds FGF, preferably FGF2, *in vivo* and/or *in vitro*. In another embodiment, the fusion protein of the present invention inhibits tumor cells directly.

Also disclosed is an FGFR-Fc fusion protein, which comprises a part derived from the extracellular domain of FGFR and a part derived from immunoglobulin Fc region. Particularly, the part derived from the extracellular domain of FGFR is the part derived from the extracellular domain of FGFR1. Preferably, the immunoglobulin Fc region is human immunoglobulin Fc region, for example, human IgG1 Fc region. In one embodiment of the present invention, the FGFR-Fc fusion protein has the capacity of binding and/or antagonizing FGF, and thus, may inhibit angiogenesis.

In the FGFR-Fc fusion protein, the part derived from the extracellular domain of FGFR may comprise one or more selected from the group consisting of: D1 domain or a moiety thereof, the part derived from IFS, D2 domain or a moiety thereof and D3 domain or a moiety thereof. In one embodiment, the part derived from the extracellular domain of FGFR may comprise D1 or a moiety thereof, the part derived from IFS, D2 domain and D3 domain. In another embodiment, the part derived from the extracellular domain of FGFR may comprise the part derived from IFS, D2 domain and D3 domain, and preferably, the part derived from IFS has the amino acid sequence corresponding to position 134 to position 162, position 145 to position 162 or position 151 to position 162 of SEQ ID NO: 1. In some preferable embodiments of the present invention, the FGFR-Fc fusion protein contains no D1 or a moiety thereof. In some other preferable embodiments of the present invention, the FGFR-Fc fusion protein contains no part from IFS other than the amino acid sequence corresponding to position 134 to position 162, position 145 to position 162 or position 151 to position 162 of SEQ ID NO: 1.

In some embodiments of the present invention, the order from the N-terminus to the C-terminus of each region and/or each domain involved in the FGFR-Fc fusion protein may

be any order. In some other embodiments, said order can be as shown in Fig. 1. In some other embodiments, said order may be different from the order shown in Fig. 1.

In some embodiments, the FGFR-Fc fusion protein of the present invention further comprises one or more intrachain disulfide bonds, and preferably, comprises one or more intra-chain disulfide bonds in the Ig-like domain.

In one embodiment of the present invention, the FGFR-Fc fusion protein can be produced by expression of the nucleic acid comprising the nucleotide sequence indicated by any one of SEQ ID NOs: 16-22 in a mammalian cell line. Particularly, the mammalian cell line is CHO cell line.

Additionally, a FGFR-Fc fusion protein is also disclosed in which domains and/or regions involved in the fusion protein are operatively linked and/or by a linker.

In a third aspect, the invention provides an isolated nucleic acid molecule encoding the fusion protein of the first or second aspect.

Also disclosed is an isolated nucleic acid molecule which encodes the fusion protein or the precursor of the fusion protein of the present invention is provided. Preferably, the nucleic acid molecule comprises the nucleotide sequence indicated by any one of SEQ ID NOs: 16-22.

In a fourth aspect of the present invention, a vector comprising the nucleic acid molecule of the third aspect is provided.

In a fifth aspect of the present invention, an isolated and/or non-human cell, optionally a CHO cell, transfected by the vector of the fourth aspect provided.

Also disclosed is a composition comprising the fusion protein of the present invention, which is mixed with a pharmaceutically acceptable carrier, is provided.

In a sixth aspect of the present invention, a pharmaceutical composition comprising the fusion protein of the first or second aspect, the nucleic acid molecule of the third aspect, the vector of the fourth aspect, or the cell of the fifth aspect and a pharmaceutically acceptable carrier is provided.

Also disclosed is a method for producing the angiogenesis-inhibitory fusion protein, which is carried out by expressing the fusion protein in prokaryotic cells or eukaryotic cells, especially, in mammalian cell lines.

Also disclosed is a method for producing the angiogenesis-inhibitory fusion protein, which is carried out by expressing the nucleic acid molecule in mammalian cell lines. Preferably, the mammalian cell line is CHO cell line.

In a seventh aspect, the invention provides use of the fusion protein of the first or second aspect, the nucleic acid molecule of the third aspect, the vector of the fourth aspect, or the cell of the fifth aspect in the manufacture of a medicament for inhibiting angiogenesis in a mammal.

In an eighth aspect, the invention provides use of the fusion protein of the first or second aspect, the nucleic acid molecule of the third aspect, the vector of the fourth aspect, or the cell of the fifth aspect in the manufacture of a medicament for treating or preventing a tumor, optionally a solid tumor, in a subject.

In a ninth aspect, the invention provides a method for inhibiting angiogenesis in a mammal in need thereof, comprising administering an effective amount of the fusion protein of the first or second aspect, the nucleic acid molecule of the third aspect, the vector of the fourth aspect, the cell of the fifth aspect, or the pharmaceutical composition of the sixth aspect to the mammal.

In a tenth aspect, the invention provides a method for treating or preventing a tumor, optionally a solid tumor, in a subject, comprising administering an effective amount of the fusion protein of the first or second aspect, the nucleic acid molecule of the third aspect, the vector of the fourth aspect, the cell of the fifth aspect, or the pharmaceutical composition of the sixth aspect to the subject.

Also disclosed is a method for inhibition of angiogenesis is provided, which comprises administering angiogenesis-inhibiting effective amount of the FGFR-Fc fusion protein, the nucleic acid molecule encoding the protein, the vector comprising the nucleic acid molecule

and/or a pharmaceutical composition comprising any one mentioned above to the subject in need thereof. Preferably, the method is carried out in the mammals.

Also disclosed is a method for the treatment or prevention of tumors in the mammals is provided, which comprises administering therapeutically or preventively effective amount of the FGFR-Fc fusion protein, the nucleic acid molecule encoding the protein, the vector comprising the nucleic acid molecule and/or a pharmaceutical composition comprising any one mentioned above to the subject in need thereof, and preferably, the tumor is a solid tumor.

Also disclosed is a method for the treatment or prevention of ophthalmic angiogenesis-related diseases in the mammals is provided, which comprises administering therapeutically or preventively effective amount of the FGFR-Fc fusion protein, the nucleic acid molecule encoding the protein, the vector comprising the nucleic acid molecule and/or a pharmaceutical composition comprising any one mentioned above to the subject in need thereof, and preferably, the ophthalmic angiogenesis-related disease is age-related macular degeneration.

Also disclosed is use of the FGFR-Fc fusion protein, the nucleic acid molecule encoding the protein, the vector comprising the nucleic acid molecule and/or a pharmaceutical composition comprising any one mentioned above in manufacture of a medicament for inhibiting angiogenesis. Also disclosed is use of the FGFR-Fc fusion protein, the nucleic acid molecule encoding the protein, the vector comprising the nucleic acid molecule and/or a pharmaceutical composition comprising any one mentioned above in manufacture of a medicament for the treatment or prevention of angiogenesis-related diseases, and preferably, the angiogenesis-related disease is a tumor or ophthalmic angiogenesis-related disease.

In view of different provisions for the subject protected in the patent systems of different countries, the disclosure has further provided the pharmaceutical uses corresponding to the methods mentioned above and the medicines for the intended uses. These various pharmaceutical uses and medicines are also covered in the protection scope of the present invention, as if they were already specifically described in the present disclosure.

In the disclosure, only some specific embodiments claimed for protection are illustrated by way of example, in which the technical features described in one or more technical proposals can be combined with any one or more technical proposals, and these technical proposals obtained by combination are also covered in the protection scope of the application, as if these technical proposals obtained by combination were already specifically described in the disclosure.

With reference to the accompanying figures and the description in more detail below, the present invention will be illustrated by way of example only. It should be understood that the description below is only illustrated by way of example for the technical solutions claimed for protection by the present invention, and not regarded

as any limitation on these technical solutions. The protection scope of the present invention shall be defined by the claims as appended.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 is a structural representation of FGFR1-Fc fusion protein. FGFR1-Fc fusion protein is represented by a solid line, and the deleted amino acid is represented by a dash line; the antibody-like domain is represented by a circle; different antibody-like domains are represented by number 1-3; the disulfide bond is represented by s s; human IgG1 Fc is represented by a grey box; VEGFR1 signal peptide is represented by SP; the acidic box sequence is represented by a box with letter AB.

Fig. 2 shows the comparison of FGF-2 binding among various FGFR1-Fc fusion proteins. Binding of heparin (100 ng/mL) containing FGF-2 (50 ng/mL) or FGF-2 (50 ng/mL) alone to each FGFR1-Fc fusion protein (20 ng/mL) is detected by ELISA.

Fig. 3 shows SDS-PAGE of 26# FGFR1-Fc fusion protein.

Fig. 4 shows the binding of FGF-2 to a gradient concentration of 26# FGFR1-Fc fusion protein.

Fig. 5 shows the affinity between 26# FGFR1-Fc fusion protein and FGF-2.

Fig. 6 shows the effect of 26# FGFR1-Fc fusion protein on the HUVEC cell division induced by FGF-2.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

Definitions

Unless otherwise defined, all scientific terms used herein have the same meaning as commonly understood by those skilled in the art. With regard to the definitions and terms in the art, reference may be made to Current Protocols in Molecular Biology

(Ausubel) by the skilled one. Standard three- and/or one-letter code used for expressing one of 20 common L-amino acids in the art are adopted as the abbreviation of amino acid residues.

Although the number ranges and approximate parameter values are given in a broad range in the present invention, all numbers in the specific examples are described as precise as possible. However, certain errors exist in any numerical values essentially, which may be resulted from the standard deviation during the measurement for each of them. Additionally, it should be understood that all ranges disclosed herein encompass any and all possible subranges contained therein. For example, it should be understood that the range "from 1 to 10" as described herein encompasses any and all possible subranges between the minimum 1 and the maximum 10 (including the endpoints); i.e., all subranges started from the minimum 1 or more, for example 1 to 6.1, and all subranges ended at the maximum 10 or less, for example 5.5 to 10. Additionally, it should be understood that any reference referred as "incorporated herein" is incorporated in its entirety.

Additionally, it should be noted that unless otherwise clearly and explicitly stated, the singular form includes the plural referent, as used in the present invention. The term "or" and the term "and/or" are used interchangeably, unless otherwise clearly indicated in the context.

In the claims which follow and in the description of the invention, except where the context requires otherwise due to express language or necessary implication, the word "comprise" or variations such as "comprises" or "comprising" is used in an inclusive sense, i.e. to specify the presence of the stated features but not to preclude the presence or addition of further features in various embodiments of the invention.

As used herein, the term "Fc", "Fc region", "Fc fragment" or "immunoglobulin Fc region" refers to the crystallizable fragment of immunoglobulin, and in the present invention, said Fc region is preferably the human IgG1 Fc region.

The term "Fc fusion protein" refers to the antibody-like molecule which incorporates the binding specificity of a heterologous protein and the effector function of a constant region of an immunoglobulin. In the term of molecular structure, a Fc fusion protein comprises the amino acid sequence having the required binding specificity and the sequence of a constant region of an immunoglobulin. A Fc fusion protein molecule generally comprises a binding site of a receptor or a ligand. The sequence of immunoglobulin constant region may be derived from any

immunoglobulin, for example, IgG-1, IgG-2, IgG-3 or IgG-4 subtype, IgA (including IgA-1 and IgA-2), IgE, IgD or IgM.

The term "soluble" protein as used herein refers to the protein which may be dissolved in an aqueous solution at a biologically relevant temperature, pH level and osmotic pressure. The "soluble fusion protein" as used herein is intended to mean that the fusion protein does not contain a transmembrane region or an intracellular region.

As used herein, the term "isolated" refers to the following substance and/or entity: (1) which is isolated from at least some components which is present when initially produced (in natural environment and/or in a experiment device) and related thereto and/or (2) which is produced, prepared and/or manufactured artificially. The isolated substance and/or entity may be isolated from at least about 10%, about 20%, about 30%, about 40%, about 50%, about 60%, about 70%, about 80%, about 90%, about 95%, about 98%, about 99%, substantially 100% or 100% other components related to it initially.

The term "part" and "fragment" interchangeably refer to a part of polypeptide, nucleic acid or other molecular constructs.

The term "Ig-like domain" as used herein refers to immunoglobulin-like domain, which may be found in a plurality of protein families and involved in many biological functions, including cell-cell recognition, cell surface receptor, immune function and the like.

Fibroblast growth factor (FGF) is a heparin-binding growth factor family, which has 22 family members in the mammals (FGF 1-14, 16-23). FGF has many important biological functions, such as cell multiplication, differentiation, migration, angiogenesis and tumorigenesis. FGF exerts many biological functions by binding and activating the cell surface FGF receptor (FGFR). (See, for example, Eswarakumar *et al. Cytokine Growth Factor Rev.* 16: 139-149, 2005). Fibroblast growth factor receptor (FGFR) is the receptor that binds the family members of fibroblast growth factor. A part of fibroblast growth factor receptor is involved in the disease process. In the mammals, there are 4 FGFR genes: fgfr1-fgfr4. The fibroblast growth factor

receptor is composed of extracellular domain, transmembrane domain and intracellular domain. There are many members in FGFR family, which are different from each other in the term of ligand binding properties and kinase domains. However, the extracellular domains thereof are similar. There are three immunoglobulin-like (Ig-like) domains contained in their extracellular domains: the first Ig-like domain, the second Ig-like domain and the third Ig-like domain, and there is also a sequence contained between the first and the second Ig-like domain. Said sequence contained between the first and the second Ig-like domain is referred herein as the intermediate functional sequence region of the Ig-like domain of FGFR. Said intermediate regulation sequence comprises a region of acidic amino acids, referred as acidic box (AB).

As used herein, the term "the first Ig-like domain of FGFR" or "the first Ig-like domain" refers to the first Ig-like domain in the protein FGFR from the N-terminus, which has for example the amino acid sequence corresponding to position 40 to position 118 of SEQ ID NO: 1. Similarly, the term "the second Ig-like domain of FGFR" or "the second Ig-like domain" refers to the second Ig-like domain in the protein FGFR from the N-terminus, which has for example the amino acid sequence corresponding to position 163 to position 247 of SEQ ID NO: 1; the term "the third Ig-like domain of FGFR" or "the third Ig-like domain" refers to the first Ig-like domain in the protein FGFR from the N-terminus, which has for example the amino acid sequence corresponding to position 270 to position 359 of SEQ ID NO: 1. Preferably, the FGFR is FGFR1, and the first Ig-like domain of FGFR is the first Ig-like domain of FGFR1, and the second Ig-like domain of FGFR is the second Ig-like domain of FGFR1, and the third Ig-like domain of FGFR is the third Ig-like domain of FGFR1.

A part of sequence of hFGFR1 is given as follows, in which each Ig-like domain is shown in shaded area sequentially, see <http://www.ncbi.nlm.nih.gov/protein/AAH15035.1>

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MWSWKCLLFWAVLVTATLCTARPSPTLPEQAQPWGAPVEVESFLVHPGDL
 QLRCLRDDVQSINWLRDGVQLAESNRTRITGEEVEVQDSVPADSGLYACVT
 SSPSGSDTTYFSVNVSDALPSSSEDDDDDDSSSEEKETDNTKPNPVAPYWTSP
 EKMEKKLHAVPAAKTVKFKCPSSG180TPNPTLRWLKNGKEFKPDHRIGGYK
 VRYATWSIMDSVVP SDKGNYTCIVENEYGSINHTYQLDVVERSPHRPILQAG
 LPANKTVALGSNVEFMCKVYSDPQPHIQWLKHIEVNGSKIGP300DNLPPYVQI
 LKTAGVNTTDKEMEVLHLRNVSFEDAGEYTCLAGNSIGLSHHS AWLTVLEAL
 EER

The amino acid sequence of FGFR1 may be found in SEQ ID NO: 1, and its encoding nucleotide sequence may be found in SEQ ID NO: 4.

As used herein, the term "the intermediate functional sequence region of the Ig-like domain of FGFR" or "the intermediate functional sequence of the Ig-like domain of FGFR" or "IFS" refers to the sequence between the first Ig-like domain and the second Ig-like domain in the protein FGFR, and preferably, IFS sequence has the amino acid sequence corresponding to position 118 to position 162 of SEQ ID NO: 1. Unexpectedly, it has been found by the present inventor that there is a significant effect of the intermediate functional sequence region on the function of the Ig-like domain. In some embodiments of the present invention a FGFR fusion protein, which comprises a plurality of parts of various lengths derived from the intermediate functional sequence region, and particularly preferably, the part derived from the intermediate functional sequence region contains no acidic box. More preferably, the part derived from IFS has the amino acid sequence corresponding to position 134 to position 162, position 145 to position 162 or position 151 to position 162 of SEQ ID NO: 1. The protein FGFR is preferably FGFR1 (SEQ ID NO: 1), especially the protein human FGFR1. The amino acid sequence of the protein human FGFR1 may be found in SEQ ID NO: 1, and its cDNA sequence may be found in SEQ ID NO: 4.

The term "FGFR" as used herein refers to fibroblast growth factor receptor, which may be FGFR1, FGFR2, FGFR3 and/or FGFR4. Preferably, the FGFR in the present invention is FGFR1, more preferably, human FGFR1.

As used herein, the term "degenerate variant" is intended to mean that the degenerate variant comprises a degenerate change at the third position of the amino acid codon so that the degenerate variants encode the same amino acid, for example the wobble position of a triplet code comprising one or more changed variants (also referred as synonymous variant).

As used herein, the term "subject" refers to mammals, such as human. However, it may also be other animals, such as domesticated animals (such as dog and cat etc.), livestock (such as cattle, sheep, pig and horse etc.) or experimental animals (such as monkey, rat, mouse, rabbit and guinea pig etc.).

As used herein, the term "percentage identity", "homology" or "identity" refers to the sequence identity between two amino acid sequences or nucleic acid sequences. The percentage identity may be determined by alignment between two sequences, and the percentage identity refers to the amount of the same residue (i.e., amino acid or nucleotide) at the same position in the sequence aligned. Sequence alignment and comparison may be performed using standard algorithms in the art (for example Smith and Waterman, 1981, *Adv. Appl. Math.* 2: 482; Needleman and Wunsch, 1970, *J. Mol. Biol.* 48: 443; Pearson and Lipman, 1988, *Proc. Natl. Acad. Sci., USA*, 85: 2444) or by the computerized versions of these algorithms (Wisconsin Genetics Software Package Release 7.0, Genetics Computer Group, 575 Science Drive, Madison, WI). Said computerized versions publicly available are BLAST and FASTA. Additionally, ENTREZ available through National Institutes of Health (Bethesda MD) may be used for sequence alignment. When BLAST and GAP-BLAST are used, default parameters for each program (for example, BLASTN, available on the website of National Center for Biotechnology Information) may be used. In one embodiment, the percentage identity between two sequences may be determined using GCG with a gap-weight of 1 so that the giving weight of each amino acid gap seems as if it is a single amino acid mismatch between two sequences. Alternatively, ALIGN (version

2.0), which is a part of GCG (Accelrys, San Diego, CA) Sequence Alignment Software Package, may be used.

As used herein, the term "hybridization" refers to the process by which a stable double-stranded polynucleotide is formed by non-covalent bonding between two single stranded polynucleotides. The term "hybridization" also may refer to triple-stranded hybridization. The double stranded polynucleotide (generally) produced is the "hybrid" or "duplex". "The condition for hybridization" generally includes a salt concentration lower than about 1 M, and more generally, lower than about 500 mM, and lower than about 200 mM. The hybridization temperature may be as low as 5°C, but it usually higher than about 22°C, and more usually higher than about 30°C, and preferably higher than about 37°C. Hybridization is usually carried out under strict conditions (i.e., the conditions under which the probe will hybridize to its target sequence). Strict hybridization conditions are dependent on the sequence and will be varied under different conditions. Higher hybridization temperature will be probably required by longer segments for specific hybridization. Since the hybridization stringency may be influenced by other factors (including base composition and length of the complementary strand, the presence of organic solvent and the degree of base mismatch), the combination of parameters is more important than the absolute value of any single parameter. Generally, the strict condition is selected as 5°C lower than the T_m of the sequence under certain ionic strength and pH. Exemplary strict conditions include pH 7.0 to 8.3, sodium ion (or other salts) concentration of at least 0.01 M to no more than 1 M and temperature of at least 25°C. For strict conditions, see, for example Sambrook, Fritsche and Maniatis. "Molecular Cloning A laboratory Manual", 2nd edition, Cold Spring Harbor Press (1989) and Anderson "Nucleic Acid Hybridization", 1st edition, BIOS Scientific Publishers Limited (1999), which are incorporated herein by reference for all purposes mentioned above.

As used herein, the term "linker", "peptide linker", "linking sequence" or "linker sequence" refers to a short amino acid sequence by which individual domain and/or region involved in the present fusion protein are linked together, and the length of the short amino acid sequence is generally 0-20 amino acids, and preferably, 2-10 amino acids.

As used herein, the term of "the amino acid sequence corresponding to SEQ ID NO: N" in a fusion protein or part or domain is intended to mean said fusion protein or part or domain has the amino acid sequence substantially as indicated by SEQ ID NO: N, and preferably, containing no more than 1, 2, 3, 4, 5, 10 or 20 substitutions, additions and deletions of amino acids, and yet preferably, said fusion protein or part or domain shares at least 80%, 90%, 93%, 95%, 97%, 98% or 99% identity with the amino acid sequence of SEQ ID NO: N, and more preferably, said fusion protein or part or domain has the amino acid sequence as indicated by SEQ ID NO: N.

As used herein, the term "FGFR-Fc fusion protein" refers to a fusion protein which comprises the part derived from the extracellular domain of FGFR and the part derived from the immunoglobulin Fc region, wherein the part derived from the extracellular domain of FGFR may: (1) comprise the amino acid sequence sharing at least 70% identity, preferably at least 80%, 90%, 93%, 95%, 97%, 98% or 99% identity, with the amino acid sequence indicated by any one of SEQ ID NOs: 9-15 or composed thereof; (2) comprise the amino acid sequence encoded by the nucleotide sequence sharing at least 70% identity, preferably at least 80%, 90%, 93%, 95%, 97%, 98% or 99% identity, with the nucleotide sequence indicated by any one of SEQ ID NOs: 16-22 or composed thereof; or (3) possess the amino acid sequence indicated by any one of SEQ ID NOs: 9-15, or the amino acid sequence encoded by the nucleotide sequence indicated by any one of SEQ ID NOs: 16-22.

In some preferable embodiments, the FGFR-Fc fusion protein may be encoded by the nucleic acid, in which the nucleotide sequence encoding the part derived from the extracellular domain of FGFR comprises the sequence of which the complementary

sequence is hybridized with the nucleotide sequence as indicated by any one of SEQ ID NOs: 16-22 under stringent conditions, or comprises the degenerative variant of the nucleotide sequence as indicated by any one of SEQ ID NOs: 16-22. In some preferable embodiments, the nucleotide sequence encoding the immunoglobulin Fc region comprises the sequence of which the complementary sequence is hybridized with the nucleotide sequence indicated by SEQ ID NO: 8 under stringent conditions, or comprises the degenerative variant of the nucleotide sequence indicated by SEQ ID NO: 8.

In other preferable embodiments, the FGFR-Fc fusion protein includes the FGFR-Fc fusion protein variant. In one embodiment, the variant includes the variant which contains no more than 2, 3, 4, 5 or 10 substitutions, additions or deletions of amino acid in the part derived from IFS corresponding to the amino acid sequence indicated by position 134 to position 162, position 145 to position 162 or position 151 to position 162 of SEQ ID NO: 1, and preferably, the variant retains the angiogenesis-inhibitory capacity. In another embodiment, the variant includes the variant which contains no more than 2, 3, 4, 5, 10 or 20 substitutions, additions or deletions of amino acid in D2 domain corresponding to the amino acid sequence indicated by position 163 to position 247 of SEQ ID NO: 1, and preferably, the variant retains the angiogenesis-inhibitory capacity. In another embodiment, the variant includes the variant which contains no more than 2, 3, 4, 5, 10 or 20 substitutions, additions or deletions of amino acid in D3 domain corresponding to the amino acid sequence indicated by position 270 to position 359 of SEQ ID NO: 1, and preferably, the variant retains the angiogenesis-inhibitory capacity. In another embodiment, the substitution, addition or deletion is located at the linker or the linking part.

In addition to the naturally occurring modifications in the part derived from the extracellular domain of FGFR and the part derived from immunoglobulin Fc region, other post-translational modifications may also be comprised in the FGFR-Fc fusion

protein. Such modifications include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, esterification and acylation. As a result, non-amino acid component may be comprised in the modified FGFR-Fc fusion protein, for example polyethylene glycol, lipid, polysaccharide or monosaccharide, and phosphoric acid. Effect of such non-amino acid components on the function of the FGFR-Fc fusion protein may be tested as described for other FGFR-Fc fusion protein variants herein. When FGFR-Fc fusion protein is produced in a cell, post-translational processing is also possibly important for correct folding and/or protein function. Special cell machines and unique mechanisms exist in different cells (for example CHO, HeLa, MDCK, 293, WI38, NIH-3T3 or HEK293) for these post-translational activities, and different cells may be selected to make sure correct modification and processing of FGFR-Fc fusion protein.

The fusion protein as described herein may be produced by any method known in the art. For example, it may be produced by chemical synthesis or from nucleic acid expression. The peptides used in the present invention may be easily prepared according to the established standard liquid, or preferably, solid phase peptide synthesis method known in the art (see, for example J. M. Stewart and J. D. Young, *Solid Phase Peptide Synthesis*, 2nd edition, Pierce Chemical Company, Rockford, Illinois (1984), in M. Bodanzsky, and A. Bodanzsky, *The Practice of Peptide Synthesis*, Springer Verlag, New York (1984)). The fusion protein may be produced by the techniques known in the art so that one or more intramolecular crosslinkings may be formed between the cysteine residues located in the polypeptide sequence expected to be comprised in the protein (see, for example US patent No. 5478925). In addition, general modifications may be performed to the protein described herein by adding cysteine or biotin to the C-terminus or N-terminus of the protein.

As used herein, "therapeutically effective amount" or "effective amount" refers to the dosage which is sufficient to show the benefit to the subject administrated. The actually administrated dosage, the rate and the time course of administration are

dependent on the condition of the patient and the severity of the disease. Finally, the physician is responsible for the prescription (for example decision on the dosage etc.) and will make a decision for the treatment, usually by considering the disease treated, individual condition of the patient, position of delivery, the method for administration and other factors known to the physician.

A series of isolated soluble FGFR-Fc fusion proteins are constructed by the present inventor, which may bind FGF and effectively inhibit the cell division induced by FGF. The fusion protein preferably comprises: the part derived from IFS, D2, D3 and immunoglobulin Fc region.

Unexpectedly, it has also been found by the present inventor that the binding of FGF by the fusion protein is significantly influenced by the length of the part derived from IFS. Therefore, in some embodiments of the present invention fusion proteins comprising the parts derived from IFS with various lengths. Preferably, the part derived from IFS comprises no acidic box, and more preferably, it has the amino acid sequence corresponding to position 134 to position 162, position 145 to position 162 or position 151 to position 162 of SEQ ID NO: 1. In some preferable embodiments, the part derived from IFS comprises the fusion protein corresponding to the amino acid sequence indicated by position 145 to position 162 of SEQ ID NO: 1, which has extremely high FGF affinity and may particularly effectively inhibit the cell division induced by FGF.

In some embodiments of the present invention, a soluble FGFR-Fc fusion protein is provided, which comprises: D1, a part derived from IFS, D2, D3 and immunoglobulin Fc region. Preferably, the part derived from IFS comprises no acidic box, and more preferably, it has the amino acid sequence corresponding to position 134 to position 162, position 145 to position 162 or position 151 to position 162 of SEQ ID NO: 1.

In some other embodiments of the present invention, a soluble FGFR-Fc fusion protein is provided, which comprises: a part of D1, a part derived from IFS, D2, D3

and immunoglobulin Fc region. Preferably, the part derived from IFS comprises no acidic box, and more preferably, it has the amino acid sequence corresponding to position 134 to position 162, position 145 to position 162 or position 151 to position 162 of SEQ ID NO: 1.

In some other embodiments of the present invention, a soluble FGFR-Fc fusion protein is provided, which is composed of: a part derived from IFS, D2, D3 and immunoglobulin Fc region. Preferably, the part derived from IFS comprises no acidic box, and more preferably, it has the amino acid sequence corresponding to position 134 to position 162, position 145 to position 162 or position 151 to position 162 of SEQ ID NO: 1.

In some other embodiments of the present invention, a soluble FGFR-Fc fusion protein is provided, which is sequentially composed of, from the N-terminus to the C-terminus, a part derived from IFS, D2, D3 and immunoglobulin Fc region. Preferably, the part derived from IFS comprises no acidic box, and more preferably, it has the amino acid sequence corresponding to position 134 to position 162, position 145 to position 162 or position 151 to position 162 of SEQ ID NO: 1.

In some other embodiments of the present invention, an FGFR-Fc fusion protein is provided, which may inhibit tumor cells directly or indirectly. Preferably, the FGFR-Fc fusion protein of the present invention inhibits tumor cells directly. More preferably, the growth of tumor cells is inhibited by the FGFR-Fc fusion protein of the present invention by at least 10%, 20%, 30%, 40%, 50%, 80%, 90% and 95% etc. The tumor cells may be any tumor cells, for example, leukaemia, lung cancer, liver cancer, head and neck cancer, stomach cancer, bladder cancer, carcinoma of uterine cervix etc. Particularly, the inhibition is achieved by direct binding to tumor cells.

In some embodiments, the present invention includes use of (i) FGFR-Fc fusion protein, or (ii) the polynucleotide encoding such fusion protein, in the preparation of the compositions or medicaments for the treatment of diseases mediated by or related to angiogenesis. For example, in one embodiment, the present invention includes use

of (i) FGFR-Fc fusion protein, or (ii) the polynucleotide encoding such fusion protein in the preparation of the medicaments as an angiogenesis inhibitor.

In some embodiments, the FGFR-Fc fusion protein according to the present invention may be produced by the expression of the nucleotide sequence as indicated by any one of SEQ ID NOs: 16-22 in a mammalian cell line. In particular, the mammalian cell line is CHO cell line.

Additionally, in the present invention, the FGFR-Fc fusion protein as described below is provided, in which a part derived from the extracellular domain of FGFR may be fused with the immunoglobulin Fc region with or without a linker.

In some other embodiments, the present invention includes the isolated nucleic acid molecules encoding the FGFR-Fc fusion protein, and the present invention also includes use of these molecules in manufacture of a medicament. The nucleic acid may be recombinant, synthetic or produced by any available methods in the art, and the method includes cloning by means of using a standard technique.

In some other embodiments, the present invention includes a vector comprising the isolated nucleic acid molecule of the present invention. The vector may be an expression vector, in which the nucleic acid is operatively linked to a control sequence which is able to provide the expression of the nucleic acid in a host cell. A plurality of vectors may be used. For example, suitable vectors may include virus (for example poxvirus, adenovirus, baculovirus etc.); yeast vector, bacteriophage, chromosome, artificial chromosome, plasmid, cosmid.

In some embodiments, the present invention further includes the cells transfected by these vectors so that the FGFR-Fc fusion protein is expressed. The host cell suitable for the present invention may be prokaryotic cell or eukaryotic cell. They include bacteria, for example *E. coli*, yeast, insect cell and mammalian cell. The mammalian cell lines that may be used include, but are not limited to, Chinese Hamster Ovary (CHO) cell, baby hamster kidney cell, NS0 mouse myeloma cell, monkey and human cell lines, and derivate cell lines thereof, etc.

In another aspect of the present invention, a method for angiogenesis inhibition is provided, comprising administrating the FGFR-Fc fusion protein of the present invention to the subject in need thereof. Preferably, the method is carried out in the mammals.

In another aspect of the present invention, a method for binding FGF *in vitro* or *in vivo* is provided, which comprises contacting FGF to the fusion protein according to the present invention.

In another aspect of the present invention, a method for the treatment or prevention of tumors in the mammals is provided, which comprises administrating the FGFR-Fc fusion protein of the present invention to the subject in need thereof, and preferably, the tumor is a solid tumor.

In another aspect of the present invention, a method for the treatment or prevention of ophthalmic angiogenesis-related diseases in the mammals is provided, which comprises administrating the FGFR-Fc fusion protein of the present invention to the subject in need thereof, and preferably, the ophthalmic angiogenesis-related disease is age-related macular degeneration.

The present invention also relates to use of the FGFR-Fc fusion protein in the preparation of medicaments for angiogenesis inhibition. Additionally, the present invention also relates to use of the FGFR-Fc fusion protein in the preparation of medicaments for the treatment or prevention of angiogenesis-related diseases, and preferably, angiogenesis-related diseases are tumors or ophthalmic angiogenesis-related disease.

The angiogenesis-related diseases as described in the present invention include, but are not limited to, angiogenesis-dependent cancers, comprising, for example, solid tumor, hematogenic tumor (for example leukaemia) and tumor metastasis; benign tumor, for example, angioma, acoustic neuroma, neurofibroma, trachoma and pyogenic granuloma; rheumatoid arthritis; psoriasis; rubeosis; Osler-Webber

Syndrome; myocardial angiogenesis; plaque neovascularization; telangiectasia; hemophiliac joint and angiofibroma.

In some embodiments of the methods described, one or more FGFR-Fc fusion proteins may be administered together (simultaneously) or at a different time (sequentially). Additionally, the fusion protein may be administered together with additional medicament used for cancer treatment or angiogenesis inhibition.

In some embodiments, the method disclosed in the present invention may be used alone. Alternatively, the subject method may be combined with other conventional anticancer therapies for the treatment or prevention of proliferative diseases (for example tumor). For example, these methods may be used for the prevention of cancers, the prevention of cancer relapse and postoperative metastasis, and may be used as a supplement for other cancer therapies. As disclosed in the present invention, the effectiveness of conventional cancer therapies (for example, chemotherapy, radiotherapy, phototherapy, immunotherapy and operation) may be enhanced by using target polypeptide therapeutic agents.

In ophthalmology, angiogenesis is related to, for example, diabetic retinopathy, retinopathy of prematurity, age-related macular degeneration, corneal transplantation rejection, neovascular glaucoma and RLF (retrolental fibroplasia). The FGFR-Fc fusion protein disclosed herein may be administered inside the eye or by other routes. Other diseases related to angiogenesis in ophthalmology include, but not limited to, epidemic keratoconjunctivitis, Vitamin A deficiency, contact lens overwear, atopic keratitis, superior limbic keratitis, pterygium keratitis sicca, sjogren, acne rosacea, phlyctenosis, syphilis, Mycobacteria infection, lipid degeneration, chemical burn, bacterial ulcer, fungal ulcer, Herpes simplex infection, Herpes zoster infection, protozoan infection, Kaposi sarcoma, Mooren ulcer, Terrien's marginal degeneration, marginal keratolysis, rheumatoid arthritis, systemic lupus, polyarteritis, trauma, Wegeners sarcoidosis, Scleritis, Steven's Johnson disease, periphigoid radial keratotomy and corneal graft rejection, sickle cell anemia, sarcoid, pseudoxanthoma

elasticum, Pagets disease, vein occlusion, artery occlusion, carotid obstructive disease, chronic uveitis/vitritis, mycobacterial infections, Lyme's disease, systemic lupus erythematosus, retinopathy of prematurity, Eales disease, Bechets disease, infection resulting in retinitis or choroiditis, presumed ocular histoplasmosis, Bests disease, myopia, optic pit, Stargarts disease, pars planitis, chronic retinal detachment, hyperviscosity syndromes, toxoplasmosis, trauma and post-laser complication. Other diseases include, but not limited to, rubeosis (neovasculariation of the angle) related diseases and diseases induced by abnormal hyperplasia of the fibrous blood vessel or fibrous tissue, including all kinds of proliferative vitreoretinopathy.

Administration

The fusion protein of the present invention may be administrated alone, but preferably, as a pharmaceutical composition which usually comprises a suitable pharmaceutical excipient, diluent or carrier selected according to the intended administration route. The fusion protein may be administrated to the patient in need thereof by any suitable route. A precise dosage will be dependent on many factors, including exact properties of the fusion protein.

Some suitable administration routes include (but are not limited to) oral, rectal, nasal, topical (including buccal and sublingual), subcutaneous, vaginal or parenteral (including subcutaneous, intramuscular, intravenous, intracutaneous, intrathecal and extradural) administration.

For intravenous injection and injection at the focal site, active ingredients are present in the form of a parenterally-acceptable aqueous solution, which is free of pyrogen and has appropriate pH value, isotonicity and stability.

A suitable solution may be well formulated by the skilled one in the art using, for example, isotonic excipients such as sodium chloride injection, Ringer's injection, Ringer's lactate injection. As required, preservative, stabilizer, buffering agent,

antioxidant and/or some other additives may be added. The pharmaceutical composition orally administered may be in a form of tablet, capsule, powder or oral liquid etc. Solid carrier, such as gelatin or adjuvant, may be comprised in a tablet. Liquid pharmaceutical composition usually comprises liquid carrier, such as water, petroleum, animal or vegetable oil, mineral oil or synthetic oil. Also included may be normal saline solution, glucose or other sugar solutions or glycols such as ethylene glycol, propylene glycol or polyethylene glycol.

Examples of the techniques and schemes as mentioned above and other techniques and schemes as used according to the present invention may be found in Remington's Pharmaceutical Sciences, 16th edition, Oslo, A. (ed), 1980.

Cloning of the fusion protein and construction of the expression plasmid

The FGF receptor fragment are obtained from the amplification of the cDNA template of corresponding receptor through PCR, and IgG1 Fc fragment is obtained from the cDNA amplification of the human-derived IgG1 through PCR. When PCR primers are designed, linking sequences are introduced between different fragments so that these different fragments may be finally linked by overlap PCR to form reading frames for different fusion proteins, and endonuclease BspE I and Pst I site are added to both ends of the cDNA. The cDNAs for different fusion proteins may be cloned to the expression plasmid after digestion by BspE I and Pst I. The plasmid after cloning may be determined by endonuclease digestion, electrophoresis and finally DNA sequencing.

Expression and purification of the fusion protein

The present fusion protein may be expressed and purified by techniques commonly used in the art. DNA from corresponding fusion protein plasmid was purified using plasmid purification kit (MAX) available from Qiagen, and the

concentration of plasmid DNA was determined using UV spectrophotometry, and the plasmid was transfected to CHO cell using FUGENE 6 liposome (Roche). Specific method for transfection was performed according to the specification of the product. Based on the expression amount required for the proteins, two methods were employed in the present invention for protein expression: (1) transient expression, in which the fusion protein contained culture supernatant was usually harvested 48-72 h after transfection, and the relative content of the fusion protein was then determined using human IgG ELISA so that the fusion protein may be rapidly and efficiently obtained; (2) establishing a stable cell line and producing the common DHFR-defective CHO cell expression system using the recombinant protein medicament expression, the basic process of which includes cell transfection, selection of stably transfected cell, clone screening, stress amplification, culture medium and process optimization etc., and finally realizing a large-scale suspension culture of CHO engineering cell strain in a serum free culture medium. The culture product was collected and the fusion protein was purified using Protein A affinity column. The purified protein was analyzed by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE), and subsequently all eluates in which the required expression product was contained were combined and filtered using a 0.22 μm filter, and then protein quantification was carried out according to a plurality of methods such as Lowry protein assay. The volume of CHO cell culture in the present invention was at a level of 10 L bioreactor, through which the fusion protein obtained after purification could satisfy the protein amount required in the animal experiments, and also a basis was established for future scaling-up.

Neutralization of FGF by the fusion protein was validated at a level of protein

After the fusion protein expressed by CHO was obtained, the binding capacity of the fusion protein to FGF is evaluated in the present invention at a level of protein. Binding experiment and affinity experiment were performed for validation in the

present invention, in which steps of the binding experiment included: after initially coated by FGF-2 on a 96-well ELISA plate, the coated well was blocked by BSA followed by adding each fusion protein at the same concentration, and then a secondary antibody to human IgG Fc-HRP was added after washing, and the samples were developed, stopped and read at 450nm on a ELISA plate, and finally the fusion protein which had binding capacity to FGF-2 was screened based on the signal strength. The affinity experiment was performed in order to determine the affinity of the fusion protein to FGF-2 in the solution system, which comprised the following steps: FGF-2 was initially coated on a 96-well ELISA plate to capture the antibody, and then the coated well was blocked by BSA, and subsequently a mixture of the fusion protein and FGF-2 which was previously prepared and incubated were added with a gradient of diluted standards, and after incubation, an HRP-labeled detection antibody was added (using antibody 2 which specifically detected free VEGF or FGF-2), and subsequently the samples were developed, stopped and read at 450nm on a ELISA plate, and finally the relative concentration of free FGF-2 was detected in the mixture of the fusion protein and FGF-2. Through the experiments above, the fusion protein having a blocking effect on FGF-2 was screened.

Neutralization of FGF by the fusion protein was validated at a cellular level

After the binding capacity of the fusion protein to FGF-2 was determined at a level of protein, its angiogenesis-inhibiting effect will be further validated at a cellular level in the present invention. The inhibition capacity of the fusion protein on the division and migration of the vascular endotheliocyte is examined by the division test using human umbilical vein endothelial cell (HUVEC) and the HUVEC cell migration test. The inhibition capacity of the fusion protein on the division of HUVEC cell can be examined by the HUVEC cell division test, which comprises the following steps during the experiment: 3000 HUVEC cells/well were inoculated to a 96-well plate and cultured at 37°C in an incubator supplemented with 5% CO₂, and then FGF-2 as

well as a mixture of the fusion protein at different concentrations with FGF-2 are added respectively, and after culturing for another 3-4 days, 10% CCK-8 is added and cultured for 2 h before the sample is read at 450 nm on a ELISA plate. The inhibition capacity of the fusion protein on the division of vascular endotheliocyte induced by FGF-2 was evaluated based on the difference of absorbance, and the median effective concentration of the fusion protein was obtained for FGF-2 inhibition. The inhibition capacity of the fusion protein on HUVEC cell migration was examined by the HUVEC cell migration test, which comprises the following steps during the experiment: 50000 HUVEC cells as well as the fusion protein at various concentrations were initially inoculated in the upper chamber, while 600 μ L FGF-2 containing culture liquid was added into the lower chamber, and subsequently, the sample was cultured at 37°C in an incubator supplemented with 5% CO₂ for 20-24 h before cells on the face side of the membrane of the upper chamber were removed, and then cells on the back side of the membrane were fixed, stained and washed with PBS before observed and counted under an inverted microscope. The migration of HUVEC cells induced by the stimulation of FGF-2 was demonstrated by counting the HUVEC cells on the back side of the membrane, and the inhibition capacity of the fusion protein on the migration of the vascular endotheliocyte was tested by adding the fusion protein at various concentrations into the culture liquid. Through the experiments mentioned above, the inhibition capacity of the new fusion protein constructed in the present invention was validated on the division and migration of the vascular endotheliocyte induced by FGF-2, which also provided a basis for future animal experiments.

Tumor growth-inhibiting capacity of the fusion protein was validated by the tumor model

After the blocking effect of the new fusion protein in the present invention on FGF-2 signal was demonstrated by experiments at a protein level and a cellular level,

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its anti-tumor capacity would be tested in animal tumor models in the present invention. In the present invention, the anti-angiogenesis and anti-tumor effect of the fusion protein would be validated by models commonly used in searching medicaments for angiogenesis and tumor, for example, LLC mouse lung cancer, U87 gliocytoma, B16 melanoma and so on. In animal experiments, in addition to conventional control groups, control medicaments, such as VEGF-Trap, FP-1039, would also be included so as to obtain comparative data for anti-tumor capacity. During experiments, 100 μL tumor cell liquid with appropriate amount was subcutaneously injected into C57 mouse on one side of the back, and the tumor volume was measured with a vernier caliper twice a week. Upon the tumor grew to about 200 mm^3 , the fusion protein at various concentrations was subcutaneously injected and the mice were sacrificed after 2-3 weeks. Subsequently, the tumor volume was measured with a vernier caliper, and the anti-tumor effect of the fusion protein was validated by the size of the tumor. Furthermore, individual tumor tissue was analyzed using methods such as immunohistochemistry to investigate the regulation mechanism of angiogenesis.

Examples

Example 1: Construction of recombinant expression plasmid for FGFR1-Fc fusion protein

The FGF receptor fragment is obtained from the amplification of the cDNA templet of FGF receptor through PCR, and IgG1 Fc fragment is obtained from the cDNA amplification of the human-derived IgG1 through PCR. A commercially available cDNA (PCR Ready First Strand cDNA, derived from human adult colon cancer tissue, BioChain) was used as the template for FGFR1 fragment. Total RNA was extracted from the blood of healthy human subjects using human blood RNA extraction kit (QIAGEN). According to the manufacturer's instruction of reverse transcription kit (Promega), RT-PCR was performed using M-MLV reverse

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transcriptase (Promega) so that RNA was reversely transcribed to cDNA which was used as the template for IgG1 Fc fragment. RT-PCR was performed according to the manufacturer's instruction of reverse transcription kit, which has the following steps: Oligo dT, dNTP, total RNA and DEPC H₂O were mixed homogeneously and reacted at 70°C for 10 min before placed on ice for 5 min, and subsequently RNase inhibitor, M-MLV reverse transcriptase and reaction buffer were added. The mixture was reacted at 42°C for 1 h and subsequently at 70°C for 15 min, and the cDNA obtained may be used as the template.

Various FGFR1 fragments were individually amplified by PCR using the cDNA from human adult colon cancer tissue as the template (the primers were listed in table 1), and IgG1 Fc fragment was amplified by PCR using human blood cDNA as the template (the primers were listed in table 1 and 2). The reaction conditions for the PCR were as follows: 5 min of pre-denaturalization at 98°C, total 30 cycles of 30 s of denaturalization at 98°C, 45 s of annealing at 56°C and 2 min of extension at 72°C, and finally another 10 min of extension. When PCR primers were designed, 20 or more complementary base sequences were introduced as the linking sequence between FGFR1 fragment and IgG1 Fc fragment so that the FGFR1 fragment and IgG1 Fc fragment may be subsequently linked by overlap PCR to form reading frames for different fusion proteins, and at the same time, restriction endonuclease BspE I and Pst I site were added at both ends of the PCR product.

Subsequently, overlap PCR was carried out to obtain each FGFR1-Fc fusion protein fragment by amplification. The process of the overlap PCR reaction may be divided into two rounds, in which the fragment required for linking and containing no primer was included in the first round with reaction conditions as follows: 5 min of pre-denaturalization at 98°C, 6 cycles of 30 s of denaturalization at 98°C, 45 s of annealing at 56°C and 5 min of extension at 72°C, and finally another 10 min of extension at 72°C; after the first round, the second round of PCR was carried out by adding the primers for both ends with reaction conditions as follows: 5 min of

pre-denaturalization at 98°C, 30 cycles of 30 s of denaturalization at 98°C, 45 s of annealing at 56°C and 2 min of extension at 72°C, and finally another 10 min of extension at 72°C; through the process above, reading frames for different fusion proteins were spliced, and at the same time, restriction endonuclease BspE I and Pst I site were added at both ends of the cDNA.

After amplification, the fragments amplified by PCR were purified using QIAquick PCR purification kit (QIAGEN). cDNAs of various fusion proteins and the eucaryotic expression plasmid pSV2-dhfr (ATCC) were digested by BspE I and Pst I, respectively. Subsequently, 1% agarose gel electrophoresis was performed on the digested samples under a voltage of 90 V. Target fragments were recovered using QIAquick gel extraction kit (QIAGEN) before ligating at 16°C for 1 h using a ligase (NEB). The mixture for ligation reaction was transformed to the competent Top10 *E. coli* under the conditions of 90 s of reaction at 42°C followed by 3 min of standing on ice. After the sterile LB culture broth (free of antibody) added, the mixture was shaken at 250 rpm in a shaker at 37°C for 1 h before coating on a LB plate supplemented with ampicillin. The plate was cultured overnight in a thermostated incubator at 37°C, and then single colonies were picked out and transferred to an ampicillin-containing LB culture broth. The inoculated culture broth was shaken at 250 rpm in a shaker at 37°C overnight before the plasmid was extracted using alkaline lysis. Subsequently, the sample was digested by restriction endonuclease before evaluated by 1% agarose gel electrophoresis under a voltage of 90 V. The recombinant plasmid with correct endonuclease digestion was confirmed by DNA sequencing. Based on the steps above, 19#, 13#, 22#, 23#, 26#, 29# and 8# expression plasmid for FGFR1-Fc fusion protein were constructed. The protein sequence of FGFR1-Fc in each fusion protein and its encoding nucleotide sequence were listed in Table 3. The schematic diagram of the fusion protein structure was shown in Fig. 1.

Table 1: Primers used for amplification of FGFR1 fragment

Fusion	Upstream primer	Downstream primer
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protein		
19#	19#-FGFR1For (SEQ ID NO: 24) TAGTTCCGGAAGGCCGTCCCCGACCTTGCCTG	FGFR1Rev (SEQ ID NO: 31) GTTTTGTCTCCAGGTAC AGGGGCGAGGTC
13#	13#-FGFR1For (SEQ ID NO: 25) TAGTTCCGGAATAATCGCACCCGCATCACAG	FGFR1Rev
22#	22#-FGFR1For (SEQ ID NO: 26) TAGTTCCGGAGTAACCAGCAGCCCCTCGGGC	FGFR1Rev
23#	23#-FGFR1For (SEQ ID NO: 27) TAGTTCCGGATCCTCTTCAGAGGAGAAAGAAAC	FGFR1Rev
26#	26#-FGFR1For (SEQ ID NO: 28) TAGTTCCGGAACCTAACCCCGTAGCTCCAT	FGFR1Rev
29#	29#-FGFR1For (SEQ ID NO: 29) TAGTTCCGGACCATATTGGACATCCCCAGAAAAG	FGFR1Rev
8#	8#-FGFR1For (SEQ ID NO: 30) CTAGCTCCGGACCAGAAAAGATGGAAAAGAAATTGC	FGFR1Rev

Table 2: Primers used for amplification of IgG1 Fc fragment

	Upstream primer	Downstream primer
IgG1 Fc fragment	FcFor (SEQ ID NO: 32) CTGTACCTGGAGGACAAACTCACACATGC	FcRev (SEQ ID NO: 33) GATATCTGCAGTCATTT ACCCGGAGACAGG

Table 3: Protein sequences and nucleotide sequences for FGFR1-Fc fusion proteins

Fusion protein	Upstream primer	Downstream primer
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19#	SEQ ID NO: 9	SEQ ID NO: 16
13#	SEQ ID NO: 10	SEQ ID NO: 17
22#	SEQ ID NO: 11	SEQ ID NO: 18
23#	SEQ ID NO: 12	SEQ ID NO: 19
26#	SEQ ID NO: 13	SEQ ID NO: 20
29#	SEQ ID NO: 14	SEQ ID NO: 21
8#	SEQ ID NO: 15	SEQ ID NO: 22

Example 2: Transient expression and quantification of the fusion proteins

The DNA of individual fusion protein plasmid was purified using MAX Plasmid Purification Kit (Qiagen). The concentration of the plasmid DNA was determined by UV spectrophotometry. 1 µg recombinant plasmid and 6 µL liposome (FuGENE 6 Transfection Reagent, Roche) were homogeneously mixed into 100 µL fresh IMDM culture broth (GIBCO); after standing for 15 min, the mixture was added to the CHO cells (ATCC) cultured overnight after inoculation at a cell density of $3 \times 10^5/\text{mL}$ into a 6-well plate; the mixture was cultured at 37°C in an incubator supplemented with 5% CO₂ for 48 h with a cell complete culture broth (IMDM medium containing 10% FBS, 1% HT and 1% glutamine, all supplied by GIBCO); subsequently, the supernatant was collected and determined for the relative content of the fusion protein using human IgG ELISA kit for protein quantification (BETHYL). The relative content of the fusion protein expressed and secreted by CHO was determined with the following steps: 100 µL anti-human IgG-Fc protein (10 µg/mL) purified by affinity was coated to a 96-well ELISA plate (IMMULON) and subsequently washed for 5 times using 300 µL PBST washing solution; each coated well was blocked with 200 µL freshly prepared blocking working solution (blocking stock solution : PBS=1: 19) and incubated at 37°C for 1 h; after washed in 300 µL PBST washing solution for 5 times, 100 µL IgG solution diluted in a gradient (200 ng/mL original concentration and

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diluted by PBS in the multiple proportion of 1: 2) as a standard and 100 μ L culture supernatant of each fusion protein diluted in a gradient (starting with the concentration of each culture supernatant, and diluted by PBS in the multiple proportion of 1: 5) were added to each well and incubated at 37°C for 2h; after washed in 300 μ L PBST washing solution for 5 times, 100 μ L anti-human IgG Fc-HRP secondary antibodies diluted with PBS in a ratio of 1: 10000 was added and incubated at 37°C for 1h; after washed, the well was developed by adding 100 μ L developing solution (KPL); finally, after the development was stopped by adding 100 μ L stopping solution (KPL), the absorbance of the ELISA plate was read at a wavelength of 450 nm on a ELISA reader. The concentrations of various fusion proteins may thereby be determined according to the standard curve.

Example 3: Binding of the fusion proteins

The binding capacity of 19#, 13#, 22#, 23#, 26#, 29# and 8# fusion protein constructed above to FGF-2 was detected by ELISA. Initially, a 96-well ELISA plate (IMMULON Company) was coated by 100 μ L solution containing 50 ng/mL FGF-2 (R&D Systems) as well as containing 100 ng/mL heparin (Sigma Company) and 50 ng/mL FGF-2. Subsequently, the plate was washed by 300 μ L PBST washing solution for 5 times before each coated well was blocked by 200 μ L freshly prepared blocking working solution (KPL Company) (blocking stock solution : PBS = 1:19) and incubated at 37°C for 1 h. After washed in 300 μ L PBST washing solution for 5 times, 100 μ L solutions of various fusion proteins (dissolve in PBS, pH=7.2, concentration of 20 ng/ml) were added and incubated at 37°C for 2 h. After washed in 300 μ L PBST washing solution for 5 times, 100 μ L secondary antibody to human IgG Fc-HRP (BETHYL Company) diluted with PBS in a ratio of 1:10000 was added and incubated at 37°C for 1h. After washed in 300 μ L PBST washing solution for 5 times, the well was developed to the presence of color at room temperature in a dark place by adding 100 μ L developing solution (KPL Company), and finally the development was

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stopped by adding 100 μ L stopping solution (KPL Company) before the absorbance of the ELISA plate was read at a wavelength of 450 nm on a ELISA reader. The higher the binding capacity of the fusion protein to FGF2 was, the larger the absorbance was and the stronger the signal was. Based on the strength of the signal, 26# fusion protein was determined to have the highest binding capacity to FGF-2. Comparison of FGF-2 binding among various fusion proteins was shown in Fig. 2. It can be seen from Fig. 2 that 19#, 13#, 22#, 23#, 26# and 29# fusion protein constructed in the present invention bound to FGF at different extents in the presence of heparin, and particularly, the binding extent of 23#, 26# and 29# was extremely higher than control, and higher than that of 19#, 13# and 22#, indicating that the fusion proteins containing no acidic box according to the present invention had excellent effect. Among others, especially high binding extent was demonstrated by 26#, which presented us a clue that the fusion protein of the present invention had significantly better binding effect when comprises a part of certain length derived from the intermediate functional sequence of the Ig-like domain of FGFR.

Example 4: Stable expression and purification of the fusion proteins

DHFR-defective CHO cells (ATCC) were transfected by the recombinant expression plasmid of 26# fusion protein (possessing a high FGF-2 binding capacity) through a liposome (Roche). Particularly, 5 μ g recombinant plasmid and 30 μ L liposome (FuGENE 6 Transfection Reagent, Roche) were homogeneously mixed into 100 μ L fresh IMDM culture broth (GIBCO); after standing for 15 min, the mixture was added to the DHFR-defective CHO cells (ATCC) cultured overnight after inoculation at a cell density of 3×10^5 /mL in a 10 cm culture dish (Corning); the mixture was cultured at 37°C in an incubator supplemented with 5% CO₂ for 2-3 days with a cell complete culture broth containing 10% FBS, 1% HT and 1% glutamine in a IMDM culture medium (all supplied by GIBCO); subsequently, the cells were digested by trypsin (GIBCO), inoculated at a cell density of 3×10^5 /mL in 30 mL

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serum-free 302 culture medium (SAFC) in a flask, and selectively cultured at 37°C in an incubator supplemented with 5% CO₂ at 100 rpm to a cell density of 10⁶/mL. Subsequently, 3000 cells were inoculated into a 10 cm culture dish (Corning) (the culture broth containing 10% FBS and 1% glutamine in an IMDM culture medium) and cultured at 37°C in an incubator supplemented with 5% CO₂ to form single clones. These single clones were picked out and cultured in a 96-well plate (Corning). The relative content of the fusion protein expressed and secreted by each individual single clone was determined using a human IgG ELISA kit for protein quantification (BETHYL) under the same conditions and steps as described in Example 2 for the determination of the relative content of the fusion protein. The clone with the highest expression amount was screened out and transferred to a 6-well plate for culturing to a confluence rate of about 70%. The cells were digested by trypsin and transferred to a 10 cm culture dish. Subsequently, gradual stress amplification was carried out by adding methotrexate (MTX, Sigma) with various concentrations (10 nM, 20 nM, 50 nM, 100 nM, 200 nM and 500 nM). After stress amplification, the cells were digested by trypsin and inoculated at a cell density of 3×10⁵/mL in a flask. The expression amount of a single cell was determined so that genetically engineered stains of CHO were obtained for expressing a particular fusion protein. Finally, large-scale suspension culture (volume of 10L) of the genetically engineered stain of CHO was carried out at 37°C, 5% CO₂, 40% dissolved oxygen and 80 rpm in a serum-free 302 culture medium (pH 7.0, SAFC). The culture product was collected by centrifugation. After the supernatant was filtered using 0.45 μm filter membrane (Millipore), affinity chromatography was performed according to the instruction manual of Protein A affinity column (GE) with the specific steps as follows: initially, a protein A affinity column was equilibrated by a PBS buffer (pH 7.0); subsequently, the supernatant was loaded on the column and washed again with the PBS buffer; finally, the column was eluted with a citric acid buffer (pH 3.0), and the eluent was collected and filtered by a 0.45 μm filter membrane. After virus inactivation by adding S/D (0.3% tributyl phosphate/1% Tween 80) at 24°C for 6 h, the target protein was further purified by a

molecular sieve chromatography with the following steps: first, the eluent obtained from the Protein A affinity chromatography was dialyzed in a dialysis bag against a PBS buffer; subsequently, the sample was concentrated in a 10 KD ultrafiltration cup (Millipore); the sample concentrated using the ultrafiltration cup was then loaded on a molecular sieve chromatography column Superdex 200 (GE) equilibrated by a PBS buffer, and subsequently the column was eluted with a PBS buffer and the eluting peak was collected. The purified protein was analyzed by SDS-PAGE (Fig. 3); and subsequently, the eluates containing the required expression product was combined and filtered with a 0.22 μm filter membrane (Millipore) before the protein content was determined using many methods such Lowry protein assay.

Example 5: Gradient-binding experiment of the fusion proteins

The binding capacities of the fusion proteins as constructed above to FGF-2 were detected by ELISA, similarly as in Example 3. Initially, a 96-well ELISA plate was coated by 100 μL solution containing 50 ng/mL FGF-2 (R&D Systems). Subsequently, the plate was washed in 300 μL PBST washing solution for 5 times before each coated well was blocked by 200 μL freshly prepared blocking working solution (KPL) (blocking stock solution : PBS = 1: 19) and incubated at 37°C for 1 h. After washed in 300 μL PBST washing solution for 5 times, 100 μL solutions containing various fusion proteins at different concentrations (the starting content of protein was 16000 pM, and was diluted in a ratio of 1: 3) were added and incubated at 37°C for 2 h. After washed in 300 μL PBST washing solution for 5 times, 100 μL anti-human IgG Fc-HRP secondary antibody (BETHYL) diluted with PBS in a ratio of 1: 10000 was added and incubated at 37°C for 1 h. After washed in 300 μL PBST washing solution for 5 times, the well was developed by adding 100 μL developing solution (KPL), and finally the development was stopped by adding 100 μL stopping solution (KPL) before the absorbance of the ELISA plate was read at a wavelength of 450 nm on a ELISA reader. Based on the intensity of the signal, the gradient binding capacities of

the fusion proteins to FGF-2 were determined. In the experiment procedure mentioned above, specific conditions and steps may be found in Example 3. Gradient binding of 26# fusion protein to FGF-2 was compared in Fig. 4. It can be seen that the binding capacity of 26# fusion protein to FGF-2 was dose-dependent. It has been suggested by this example that the binding capacity to FGF-2 increased with an enhanced molar concentration of 26# fusion protein, manifested by a stronger signal at a wavelength of 450 nm; while the binding capacity to FGF-2 decreased correspondingly with a gradient dilution of the molar concentration of 26# fusion protein.

Example 6: Affinity experiment of the fusion proteins

The affinity of the fusion protein to FGF-2 in a solution system was determined by an affinity experiment. Initially, a 96-well ELISA plate was coated by 100 μ L solution containing 2.0 μ g/mL FGF-2 capture antibody (R&D Systems). Subsequently, the plate was washed in 300 μ L PBST washing solution for 5 times before each coated well was blocked by a blocking working solution (KPL) (as seen in Example 3) and incubated at 37°C for 1 h. After washed in 300 μ L PBST washing solution for 5 times, previously prepared and incubated (4 °C overnight) mixture of the fusion proteins and FGF-2 as well as the standard (R&D Systems) diluted in a gradient were added, in which the specific preparation procedure was as follows: the starting concentration of 26# fusion protein was 400 pM (dissolved in PBS) and diluted in a gradient ratio of 2-fold, and the solutions of the fusion protein were 1: 1 mixed with 20 pM FGF-2 solution (dissolved in PBS), and that is, the starting final concentration of each fusion protein was 200pM, and the final concentration of FGF-2 was 10 pM in the mixture solution prepared. The plate was incubated at 37°C for 2 h and washed in 300 μ L PBST washing solution for 5 times before 100 μ L FGF-2 detection antibody solution (250 ng/mL) was added (R&D systems, which may specifically detect free antibodies against FGF-2). The plate was incubated at 37°C for 2 h and washed in 300 μ L PBST washing solution for 5 times, and subsequently, HRP labeled streptavidin

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(R&D systems) was added (diluted by PBS in 1: 200). The plate was incubated at 37°C for 2 h and washed in 300 µL PBST washing solution for 5 times before the well was developed at room temperature in a dark place for an appropriate duration (about 15-30 min) by adding 100 µL developing solution (KPL). Finally, after the development was stopped by adding 100 µL stopping solution (KPL), the absorbance of the ELISA plate was read at a wavelength of 450 nm on a ELISA reader. The relative concentration of free FGF-2 in the mixture of the fusion protein and FGF-2 was determined. The affinity between 26# fusion protein and FGF-2 in a solution system can be seen in Fig. 5. As demonstrated in this Example, 26# fusion protein had high affinity to FGF-2 in a solution system. The affinity increased with an enhanced concentration, which is manifested as a decreased amount of free FGF-2 with an enhanced concentration of the fusion protein. The affinity between 26# fusion protein and FGF-2 in a solution system can be seen in Fig. 5. As demonstrated in this Example, 26# fusion protein had affinity to FGF-2 in a solution system. The affinity increased with an enhanced concentration, which is manifested as a decreased amount of free FGF-2.

Example 7: Inhibitory test for division on human umbilical vein endothelial cell

The inhibitory ability of the fusion proteins on the division of vascular endothelial cells was examined in a division test for human umbilical vein endothelial cell (HUVEC). HUVEC cells (AllCells) were cultured to the exponential growth phase in an HUVEC complete medium (AllCells) at 37°C in an incubator supplemented with 5% CO₂. HUVEC cells were counted after digested by trypsin. 3000 HUVEC cells were inoculated per well in an HUVEC basal medium containing 1% FBS (AllCells) in a 96-well plate. The plate was cultured overnight at 37°C in an incubator supplemented with 5% CO₂.

100 µL FGF-2 (R&D Systems) solution (final concentration of 5 ng/mL) diluted by an HUVEC basal medium containing 1% FBS, as well as 100 µL mixture of

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various amount of 26# fusion protein and FGF-2 (in which the final concentration of the fusion protein was 40 pM, diluted in an HUVEC basal medium containing 1% FBS with a ratio of 1:10, and the final concentration of FGF-2 was 5 ng/mL) were added and cultured for another 3-4 days. Subsequently, the culture medium was taken out and a culture medium containing 10% CCK-8 (DOJINDO) was added for another 2 h of culture before the absorbance of the 96-well plate was read directly at a wavelength of 450 nm on an ELISA reader. Based on the difference of the absorbance, the inhibitory ability of the fusion protein on the division of vascular endothelial cells induced by FGF-2 was determined. The effect of the fusion protein on HUVEC cell division induced by FGF-2 was shown in Fig. 6. As demonstrated in this Example, 26# fusion protein has biological activity and function at the cellular level, which can inhibit HUVEC cell division induced by FGF-2, and has the binding capacity to FGF-2. Such binding capacity increases as the molar concentration of 26# fusion protein increases, which is indicated by the inhibition of HUVEC cell division induced by FGF-2.

The present invention has already been illustrated by specific examples. However, it will be appreciated by a person of ordinary skill in the art that the present invention is not limited to each specific embodiments. Various changes and modifications may be made by a person of ordinary skill under the scope of the present invention, and each technical feature mentioned in the specification may be combined without departing from the spirit and scope of the invention. Such changes and modifications fall into the scope of the present invention.

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CLAIMS

1. An isolated soluble fusion protein of fibroblast growth factor receptor (FGFR), comprising: a part derived from the FGFR extracellular domain and an immunoglobulin Fc region, wherein the part derived from the FGFR extracellular domain consists of an amino acid sequence corresponding to positions 1-241 of SEQ ID NO: 12, positions 1-230 of SEQ ID NO: 13, or positions 1-224 of SEQ ID NO: 14 or encoded by a nucleotide sequence corresponding to positions 1-723 of SEQ ID NO: 19, positions 1-690 of SEQ ID NO: 20, or positions 1-672 of SEQ ID NO: 21.
2. An isolated soluble fusion protein of fibroblast growth factor receptor (FGFR), wherein said protein consists of an amino acid sequence of any one of SEQ ID NOs: 12-14, or is encoded by a nucleic acid molecule consisting of a nucleotide sequence of any one of SEQ ID NOs: 19-21.
3. An isolated nucleic acid molecule encoding the fusion protein of claim 1 or claim 2.
4. A vector comprising the nucleic acid molecule of claim 3.
5. A cell, optionally a CHO cell, transfected by the vector of claim 4.
6. A pharmaceutical composition, comprising the fusion protein of claim 1 or claim 2, the nucleic acid molecule of claim 3, the vector of claim 4, or the cell of claim 5, and a pharmaceutically acceptable carrier.
7. Use of the fusion protein of claim 1 or claim 2, the nucleic acid molecule of claim 3, the vector of claim 4, or the cell of claim 5 in the manufacture of a medicament for inhibiting angiogenesis in a mammal.

8. Use of the fusion protein of claim 1 or claim 2, the nucleic acid molecule of claim 3, the vector of claim 4, or the cell of claim 5 in the manufacture of a medicament for treating or preventing a tumor, optionally a solid tumor, in a subject.

9. A method for inhibiting angiogenesis in a mammal in need thereof, comprising administering an effective amount of the fusion protein of claim 1 or claim 2, the nucleic acid molecule of claim 3, the vector of claim 4, the cell of claim 5, or the pharmaceutical composition of claim 6 to the mammal.

10. A method for treating or preventing a tumor, optionally a solid tumor, in a subject, comprising administering an effective amount of the fusion protein of claim 1 or claim 2, the nucleic acid molecule of claim 3, the vector of claim 4, the cell of claim 5, or the pharmaceutical composition of claim 6 to the subject.

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Figure 1

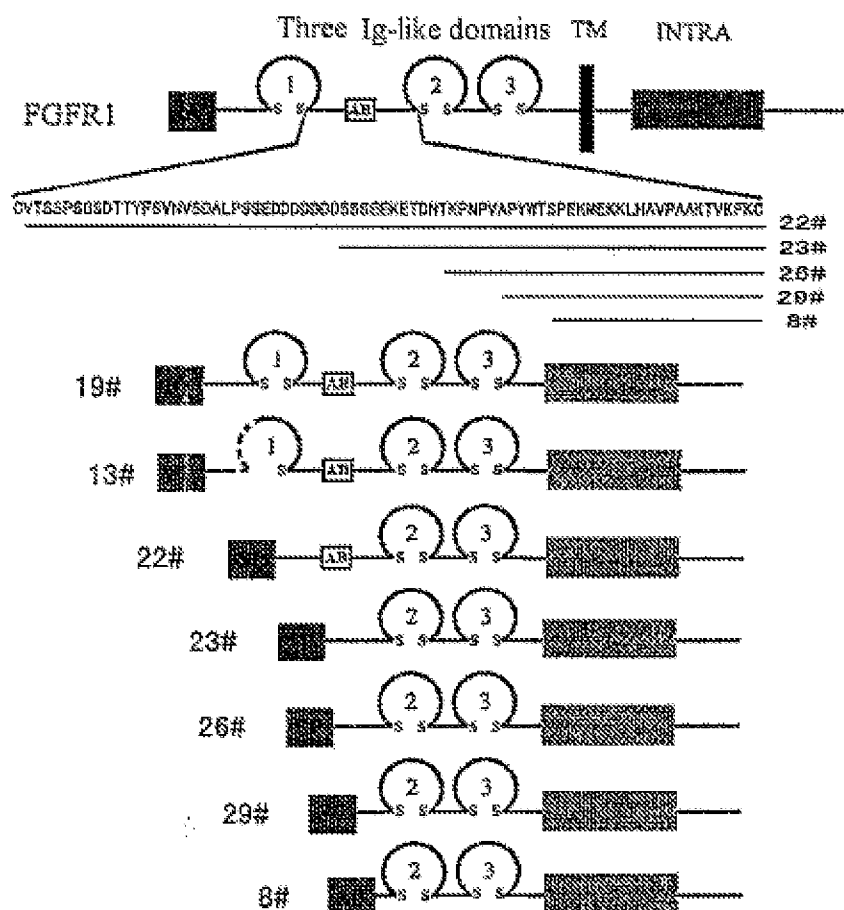


Figure 2

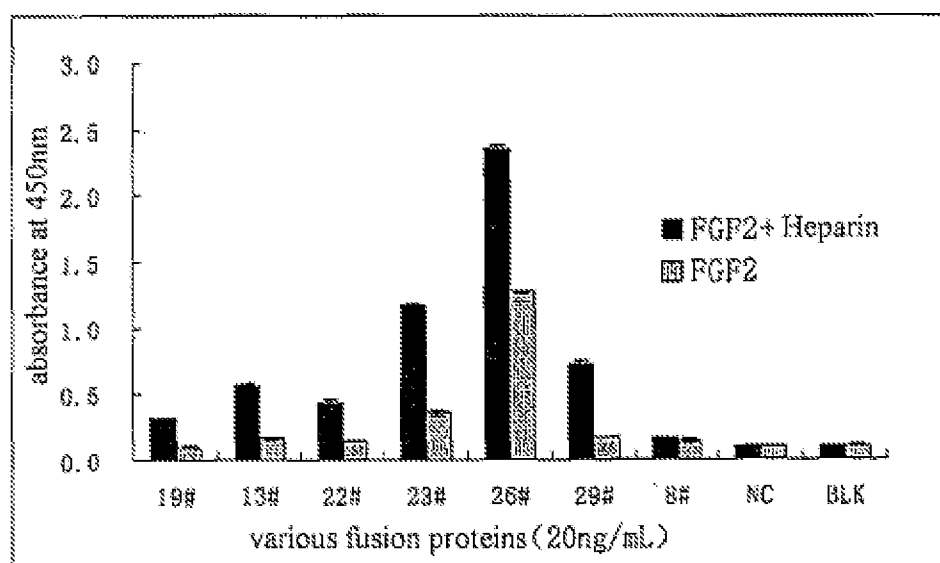


Figure 3

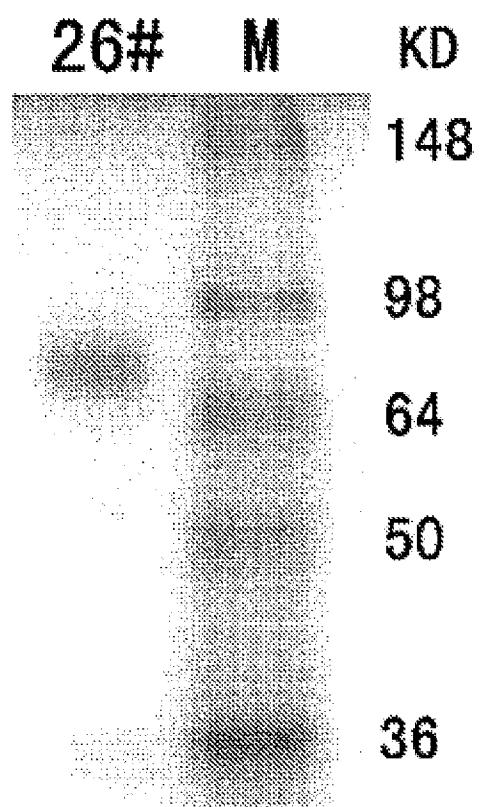


Figure 4

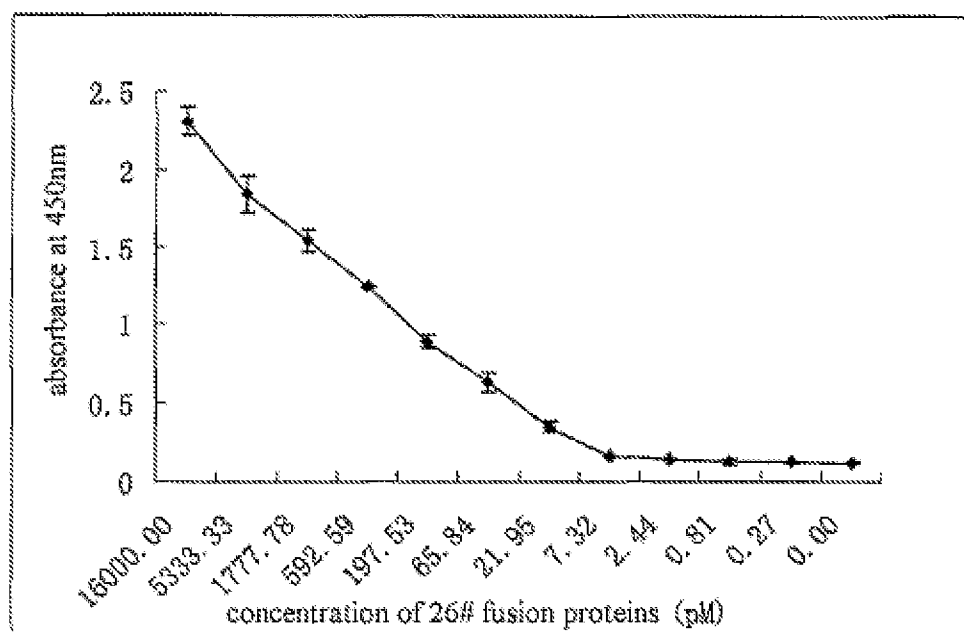


Figure 5

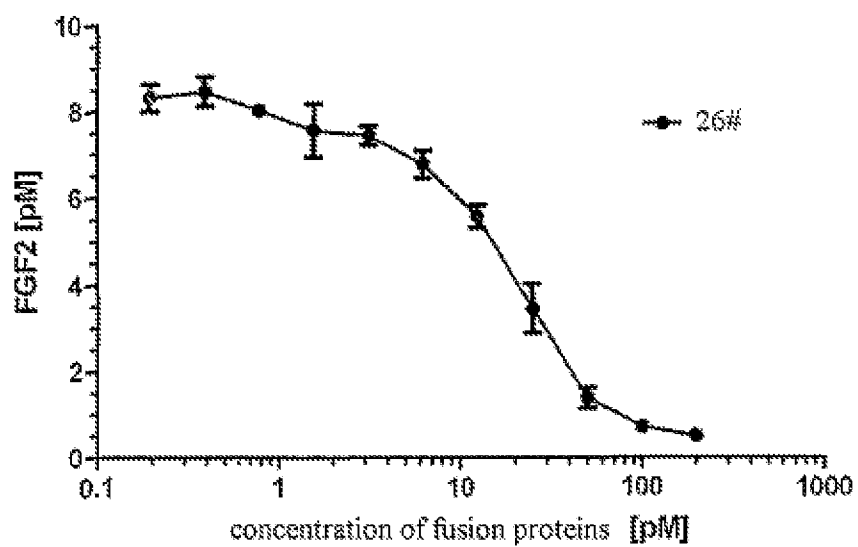
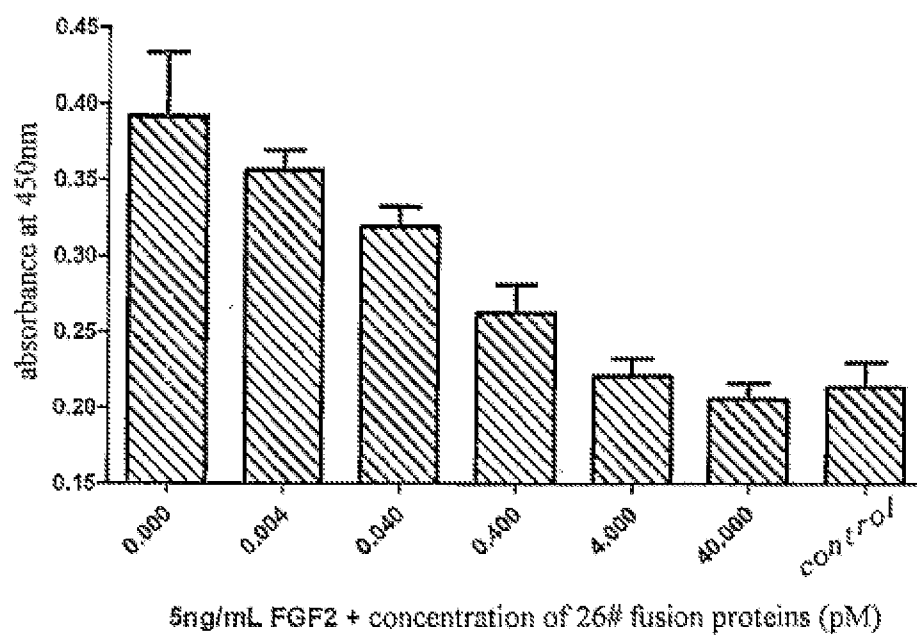


Figure 6

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 210 215 220
 Asn Tyr Thr Cys Ile Val Glu Asn Glu Tyr Gly Ser Ile Asn His Thr
 225 230 235 240
 Tyr Gln Leu Asp Val Val Glu Arg Ser Pro His Arg Pro Ile Leu Gln
 245 250 255
 Ala Gly Leu Pro Ala Asn Lys Thr Val Ala Leu Gly Ser Asn Val Glu
 260 265 270
 Phe Met Cys Lys Val Tyr Ser Asp Pro Gln Pro His Ile Gln Trp Leu
 275 280 285
 Lys His Ile Glu Val Asn Gly Ser Lys Ile Gly Pro Asp Asn Leu Pro
 290 295 300
 Tyr Val Gln Ile Leu Lys Thr Ala Gly Val Asn Thr Thr Asp Lys Glu
 305 310 315 320
 Met Glu Val Leu His Leu Arg Asn Val Ser Phe Glu Asp Ala Gly Glu
 325 330 335
 Tyr Thr Cys Leu Ala Gly Asn Ser Ile Gly Leu Ser His His Ser Ala
 340 345 350
 Trp Leu Thr Val Leu Glu Ala Leu Glu Glu Arg Pro Ala Val Met Thr
 355 360 365
 Ser Pro Leu Tyr Leu Glu Ile Ile Ile Tyr Cys Thr Gly Ala Phe Leu
 370 375 380
 Ile Ser Cys Met Val Gly Ser Val Ile Val Tyr Lys Met Lys Ser Gly
 385 390 395 400
 Thr Lys Lys Ser Asp Phe His Ser Gln Met Ala Val His Lys Leu Ala
 405 410 415

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Lys Ser Ile Pro Leu Arg Arg Gln Val Thr Val Ser Ala Asp Ser Ser
420 425 430

Ala Ser Met Asn Ser Gly Val Leu Leu Val Arg Pro Ser Arg Leu Ser
435 440 445

Ser Ser Gly Thr Pro Met Leu Ala Gly Val Ser Glu Tyr Glu Leu Pro
450 455 460

Glu Asp Pro Arg Trp Glu Leu Pro Arg Asp Arg Leu Val Leu Gly Lys
465 470 475 480

Pro Leu Gly Glu Gly Cys Phe Gly Gln Val Val Leu Ala Glu Ala Ile
485 490 495

Gly Leu Asp Lys Asp Lys Pro Asn Arg Val Thr Lys Val Ala Val Lys
500 505 510

Met Leu Lys Ser Asp Ala Thr Glu Lys Asp Leu Ser Asp Leu Ile Ser
515 520 525

Glu Met Glu Met Met Lys Met Ile Gly Lys His Lys Asn Ile Ile Asn
530 535 540

Leu Leu Gly Ala Cys Thr Gln Asp Gly Pro Leu Tyr Val Ile Val Glu
545 550 555 560

Tyr Ala Ser Lys Gly Asn Leu Arg Glu Tyr Leu Gln Ala Arg Arg Pro
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Pro Gly Leu Glu Tyr Cys Tyr Asn Pro Ser His Asn Pro Glu Glu Gln
580 585 590

Leu Ser Ser Lys Asp Leu Val Ser Cys Ala Tyr Gln Val Ala Arg Gly
595 600 605

Met Glu Tyr Leu Ala Ser Lys Lys Cys Ile His Arg Asp Leu Ala Ala
610 615 620

Arg Asn Val Leu Val Thr Glu Asp Asn Val Met Lys Ile Ala Asp Phe
625 630 635 640

Gly Leu Ala Arg Asp Ile His His Ile Asp Tyr Tyr Lys Lys Thr Thr
645 650 655

Asn Gly Arg Leu Pro Val Lys Trp Met Ala Pro Glu Ala Leu Phe Asp
660 665 670

Arg Ile Tyr Thr His Gln Ser Asp Val Trp Ser Phe Gly Val Leu Leu
675 680 685

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Trp Glu Ile Phe Thr Leu Gly Gly Ser Pro Tyr Pro Gly Val Pro Val
690 695 700

Glu Glu Leu Phe Lys Leu Leu Lys Glu Gly His Arg Met Asp Lys Pro
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Ser Asn Cys Thr Asn Glu Leu Tyr Met Met Met Arg Asp Cys Trp His
725 730 735

Ala Val Pro Ser Gln Arg Pro Thr Phe Lys Gln Leu Val Glu Asp Leu
740 745 750

Asp Arg Ile Val Ala Leu Thr Ser Asn Gln Glu Tyr Leu Asp Leu Ser
755 760 765

Met Pro Leu Asp Gln Tyr Ser Pro Ser Phe Pro Asp Thr Arg Ser Ser
770 775 780

Thr Cys Ser Ser Gly Glu Asp Ser Val Phe Ser His Glu Pro Leu Pro
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Leu Lys Arg Arg
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Glu Leu Ser Leu Lys Gly Thr Gln His Ile Met Gln Ala Gly Gln Thr
35 40 45

Leu His Leu Gln Cys Arg Gly Glu Ala Ala His Lys Trp Ser Leu Pro
50 55 60

Glu Met Val Ser Lys Glu Ser Glu Arg Leu Ser Ile Thr Lys Ser Ala
65 70 75 80

Cys Gly Arg Asn Gly Lys Gln Phe Cys Ser Thr Leu Thr Leu Asn Thr
85 90 95

Ala Gln Ala Asn His Thr Gly Phe Tyr Ser Cys Lys Tyr Leu Ala Val

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 Pro Thr Ser Lys Lys Lys Glu Thr Glu Ser Ala Ile Tyr Ile Phe Ile
 115 120 125
 Ser Asp Thr Gly Arg Pro Phe Val Glu Met Tyr Ser Glu Ile Pro Glu
 130 135 140
 Ile Ile His Met Thr Glu Gly Arg Glu Leu Val Ile Pro Cys Arg Val
 145 150 155 160
 Thr Ser Pro Asn Ile Thr Val Thr Leu Lys Lys Phe Pro Leu Asp Thr
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 Leu Ile Pro Asp Gly Lys Arg Ile Ile Trp Asp Ser Arg Lys Gly Phe
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 Ile Ile Ser Asn Ala Thr Tyr Lys Glu Ile Gly Leu Leu Thr Cys Glu
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 Ala Thr Val Asn Gly His Leu Tyr Lys Thr Asn Tyr Leu Thr His Arg
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 Gln Thr Asn Thr Ile Ile Asp Val Gln Ile Ser Thr Pro Arg Pro Val
 225 230 235 240
 Lys Leu Leu Arg Gly His Thr Leu Val Leu Asn Cys Thr Ala Thr Thr
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 Pro Leu Asn Thr Arg Val Gln Met Thr Trp Ser Tyr Pro Asp Glu Lys
 260 265 270
 Asn Lys Arg Ala Ser Val Arg Arg Arg Ile Asp Gln Ser Asn Ser His
 275 280 285
 Ala Asn Ile Phe Tyr Ser Val Leu Thr Ile Asp Lys Met Gln Asn Lys
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 Asp Lys Gly Leu Tyr Thr Cys Arg Val Arg Ser Gly Pro Ser Phe Lys
 305 310 315 320
 Ser Val Asn Thr Ser Val His Ile Tyr Asp Lys Ala Phe Ile Thr Val
 325 330 335
 Lys His Arg Lys Gln Gln Val Leu Glu Thr Val Ala Gly Lys Arg Ser
 340 345 350
 Tyr Arg Leu Ser Met Lys Val Lys Ala Phe Pro Ser Pro Glu Val Val
 355 360 365
 Trp Leu Lys Asp Gly Leu Pro Ala Thr Glu Lys Ser Ala Arg Tyr Leu

370

375

Thr Arg Gly Tyr Ser Leu Ile Ile Lys Asp Val Thr Glu Glu Asp Ala
385 390 395 400

Gly Asn Tyr Thr Ile Leu Leu Ser Ile Lys Gln Ser Asn Val Phe Lys
405 410 415

Asn Leu Thr Ala Thr Leu Ile Val Asn Val Lys Pro Gln Ile Tyr Glu
420 425 430

Lys Ala Val Ser Ser Phe Pro Asp Pro Ala Leu Tyr Pro Leu Gly Ser
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Arg Gln Ile Leu Thr Cys Thr Ala Tyr Gly Ile Pro Gln Pro Thr Ile
450 455 460

Lys Trp Phe Trp His Pro Cys Asn His Asn His Ser Glu Ala Arg Cys
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Asp Phe Cys Ser Asn Asn Glu Glu Ser Phe Ile Leu Asp Ala Asp Ser
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Asn Met Gly Asn Arg Ile Glu Ser Ile Thr Gln Arg Met Ala Ile Ile
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Glu Gly Lys Asn Lys Met Ala Ser Thr Leu Val Val Ala Asp Ser Arg
515 520 525

Ile Ser Gly Ile Tyr Ile Cys Ile Ala Ser Asn Lys Val Gly Thr Val
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Gly Arg Asn Ile Ser Phe Tyr Ile Thr Asp Val Pro Asn Gly Phe His
545 550 555 560

Val Asn Leu Glu Lys Met Pro Thr Glu Gly Glu Asp Leu Lys Leu Ser
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Cys Thr Val Asn Lys Phe Leu Tyr Arg Asp Val Thr Trp Ile Leu Leu
580 585 590

Arg Thr Val Asn Asn Arg Thr Met His Tyr Ser Ile Ser Lys Gln Lys
595 600 605

Met Ala Ile Thr Lys Glu His Ser Ile Thr Leu Asn Leu Thr Ile Met
610 615 620

Asn Val Ser Leu Gln Asp Ser Gly Thr Tyr Ala Cys Arg Ala Arg Asn
625 630 635 640

Val Tyr Thr Gly Glu Glu Ile Leu Gln Lys Lys Glu Ile Thr Ile Arg

645

650

655

Asp Gln Glu Ala Pro Tyr Leu Leu Arg Asn Leu Ser Asp His Thr Val
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 Ala Ile Ser Ser Ser Thr Thr Leu Asp Cys His Ala Asn Gly Val Pro
 675 680 685
 Glu Pro Gln Ile Thr Trp Phe Lys Asn Asn His Lys Ile Gln Gln Glu
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 Pro Gly Ile Ile Leu Gly Pro Gly Ser Ser Thr Leu Phe Ile Glu Arg
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 Val Thr Glu Glu Asp Glu Gly Val Tyr His Cys Lys Ala Thr Asn Gln
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 Lys Gly Ser Val Glu Ser Ser Ala Tyr Leu Thr Val Gln Gly Thr Ser
 740 745 750
 Asp Lys Ser Asn Leu Glu Leu Ile Thr Leu Thr Cys Thr Cys Val Ala
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 Ala Thr Leu Phe Trp Leu Leu Leu Thr Leu Phe Ile Arg Lys Met Lys
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 Arg Ser Ser Ser Glu Ile Lys Thr Asp Tyr Leu Ser Ile Ile Met Asp
 785 790 795 800
 Pro Asp Glu Val Pro Leu Asp Glu Gln Cys Glu Arg Leu Pro Tyr Asp
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 Ile Lys Lys Ser Pro Thr Cys Arg Thr Val Ala Val Lys Met Leu Lys
 850 855 860
 Glu Gly Ala Thr Ala Ser Glu Tyr Lys Ala Leu Met Thr Glu Leu Lys
 865 870 875 880
 Ile Leu Thr His Ile Gly His His Leu Asn Val Val Asn Leu Leu Gly
 885 890 895
 Ala Cys Thr Lys Gln Gly Gly Pro Leu Met Val Ile Val Glu Tyr Cys
 900 905 910
 Lys Tyr Gly Asn Leu Ser Asn Tyr Leu Lys Ser Lys Arg Asp Leu Phe

915

920

925

Phe Leu Asn Lys Asp Ala Ala Leu His Met Glu Pro Lys Lys Glu Lys
 930 935 940

Met Glu Pro Gly Leu Glu Gln Gly Lys Lys Pro Arg Leu Asp Ser Val
 945 950 955 960

Thr Ser Ser Glu Ser Phe Ala Ser Ser Gly Phe Gln Glu Asp Lys Ser
 965 970 975

Leu Ser Asp Val Glu Glu Glu Glu Asp Ser Asp Gly Phe Tyr Lys Glu
 980 985 990

Pro Ile Thr Met Glu Asp Leu Ile Ser Tyr Ser Phe Gln Val Ala Arg
 995 1000 1005

Gly Met Glu Phe Leu Ser Ser Arg Lys Cys Ile His Arg Asp Leu
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Ala Ala Arg Asn Ile Leu Leu Ser Glu Asn Asn Val Val Lys Ile
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Cys Asp Phe Gly Leu Ala Arg Asp Ile Tyr Lys Asn Pro Asp Tyr
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Val Arg Lys Gly Asp Thr Arg Leu Pro Leu Lys Trp Met Ala Pro
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Glu Ser Ile Phe Asp Lys Ile Tyr Ser Thr Lys Ser Asp Val Trp
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Ser Tyr Gly Val Leu Leu Trp Glu Ile Phe Ser Leu Gly Gly Ser
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Pro Tyr Pro Gly Val Gln Met Asp Glu Asp Phe Cys Ser Arg Leu
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Arg Glu Gly Met Arg Met Arg Ala Pro Glu Tyr Ser Thr Pro Glu
 1115 1120 1125

Ile Tyr Gln Ile Met Leu Asp Cys Trp His Arg Asp Pro Lys Glu
 1130 1135 1140

Arg Pro Arg Phe Ala Glu Leu Val Glu Lys Leu Gly Asp Leu Leu
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Gln Ala Asn Val Gln Gln Asp Gly Lys Asp Tyr Ile Pro Ile Asn
 1160 1165 1170

Ala Ile Leu Thr Gly Asn Ser Gly Phe Thr Tyr Ser Thr Pro Ala

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1175

1180

1185

Phe Ser Glu Asp Phe Phe Lys Glu Ser Ile Ser Ala Pro Lys Phe
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Asn Ser Gly Ser Ser Asp Asp Val Arg Tyr Val Asn Ala Phe Lys
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Phe Met Ser Leu Glu Arg Ile Lys Thr Phe Glu Glu Leu Leu Pro
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Asn Ala Thr Ser Met Phe Asp Asp Tyr Gln Gly Asp Ser Ser Thr
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Leu Leu Ala Ser Pro Met Leu Lys Arg Phe Thr Trp Thr Asp Ser
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Lys Pro Lys Ala Ser Leu Lys Ile Asp Leu Arg Val Thr Ser Lys
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Ser Lys Glu Ser Gly Leu Ser Asp Val Ser Arg Pro Ser Phe Cys
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His Ser Ser Cys Gly His Val Ser Glu Gly Lys Arg Arg Phe Thr
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Pro Pro Asp Tyr Asn Ser Val Val Leu Tyr Ser Thr Pro Pro Ile
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Leu Gln Ile Thr Cys Arg Gly Gln Arg Asp Leu Asp Trp Leu Trp Pro
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Asn Asn Gln Ser Gly Ser Glu Gln Arg Val Glu Val Thr Glu Cys Ser
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Asp Gly Leu Phe Cys Lys Thr Leu Thr Ile Pro Lys Val Ile Gly Asn
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 Asp Thr Gly Ala Tyr Lys Cys Phe Tyr Arg Glu Thr Asp Leu Ala Ser
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 Val Ile Tyr Val Tyr Val Gln Asp Tyr Arg Ser Pro Phe Ile Ala Ser
 115 120 125
 Val Ser Asp Gln His Gly Val Val Tyr Ile Thr Glu Asn Lys Asn Lys
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 Thr Val Val Ile Pro Cys Leu Gly Ser Ile Ser Asn Leu Asn Val Ser
 145 150 155 160
 Leu Cys Ala Arg Tyr Pro Glu Lys Arg Phe Val Pro Asp Gly Asn Arg
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 Ile Ser Trp Asp Ser Lys Lys Gly Phe Thr Ile Pro Ser Tyr Met Ile
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 Ser Tyr Ala Gly Met Val Phe Cys Glu Ala Lys Ile Asn Asp Glu Ser
 195 200 205
 Tyr Gln Ser Ile Met Tyr Ile Val Val Val Val Gly Tyr Arg Ile Tyr
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 Asp Val Val Leu Ser Pro Ser His Gly Ile Glu Leu Ser Val Gly Glu
 225 230 235 240
 Lys Leu Val Leu Asn Cys Thr Ala Arg Thr Glu Leu Asn Val Gly Ile
 245 250 255
 Asp Phe Asn Trp Glu Tyr Pro Ser Ser Lys His Gln His Lys Lys Leu
 260 265 270
 Val Asn Arg Asp Leu Lys Thr Gln Ser Gly Ser Glu Met Lys Lys Phe
 275 280 285
 Leu Ser Thr Leu Thr Ile Asp Gly Val Thr Arg Ser Asp Gln Gly Leu
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 Tyr Thr Cys Ala Ala Ser Ser Gly Leu Met Thr Lys Lys Asn Ser Thr
 305 310 315 320
 Phe Val Arg Val His Glu Lys Pro Phe Val Ala Phe Gly Ser Gly Met
 325 330 335
 Glu Ser Leu Val Glu Ala Thr Val Gly Glu Arg Val Arg Ile Pro Ala
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Lys Tyr Leu Gly Tyr Pro Pro Pro Glu Ile Lys Trp Tyr Lys Asn Gly
 355 360 365
 Ile Pro Leu Glu Ser Asn His Thr Ile Lys Ala Gly His Val Leu Thr
 370 375 380
 Ile Met Glu Val Ser Glu Arg Asp Thr Gly Asn Tyr Thr Val Ile Leu
 385 390 395 400
 Thr Asn Pro Ile Ser Lys Glu Lys Gln Ser His Val Val Ser Leu Val
 405 410 415
 Val Tyr Val Pro Pro Gln Ile Gly Glu Lys Ser Leu Ile Ser Pro Val
 420 425 430
 Asp Ser Tyr Gln Tyr Gly Thr Thr Gln Thr Leu Thr Cys Thr Val Tyr
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 Ala Ile Pro Pro Pro His His Ile His Trp Tyr Trp Gln Leu Glu Glu
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 Glu Cys Ala Asn Glu Pro Ser Gln Ala Val Ser Val Thr Asn Pro Tyr
 465 470 475 480
 Pro Cys Glu Glu Trp Arg Ser Val Glu Asp Phe Gln Gly Gly Asn Lys
 485 490 495
 Ile Glu Val Asn Lys Asn Gln Phe Ala Leu Ile Glu Gly Lys Asn Lys
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 Thr Val Ser Thr Leu Val Ile Gln Ala Ala Asn Val Ser Ala Leu Tyr
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 Phe His Val Thr Arg Gly Pro Glu Ile Thr Leu Gln Pro Asp Met Gln
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 Pro Thr Glu Gln Glu Ser Val Ser Leu Trp Cys Thr Ala Asp Arg Ser
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 Thr Phe Glu Asn Leu Thr Trp Tyr Lys Leu Gly Pro Gln Pro Leu Pro
 580 585 590
 Ile His Val Gly Glu Leu Pro Thr Pro Val Cys Lys Asn Leu Asp Thr
 595 600 605
 Leu Trp Lys Leu Asn Ala Thr Met Phe Ser Asn Ser Thr Asn Asp Ile
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Leu Ile Met Glu Leu Lys Asn Ala Ser Leu Gln Asp Gln Gly Asp Tyr
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 Val Cys Leu Ala Gln Asp Arg Lys Thr Lys Lys Arg His Cys Val Val
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 Arg Gln Leu Thr Val Leu Glu Arg Val Ala Pro Thr Ile Thr Gly Asn
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 Thr Ala Ser Gly Asn Pro Pro Pro Gln Ile Met Trp Phe Lys Asp Asn
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 Cys Gln Ala Cys Ser Val Leu Gly Cys Ala Lys Val Glu Ala Phe Phe
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 Val Gly Thr Ala Val Ile Ala Met Phe Phe Trp Leu Leu Leu Val Ile
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 Ile Leu Arg Thr Val Lys Arg Ala Asn Gly Gly Glu Leu Lys Thr Gly
 785 790 795 800
 Tyr Leu Ser Ile Val Met Asp Pro Asp Glu Leu Pro Leu Asp Glu His
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 Cys Glu Arg Leu Pro Tyr Asp Ala Ser Lys Trp Glu Phe Pro Arg Asp
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 Ile Glu Ala Asp Ala Phe Gly Ile Asp Lys Thr Ala Thr Cys Arg Thr
 850 855 860
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 Ala Leu Met Ser Glu Leu Lys Ile Leu Ile His Ile Gly His His Leu
 885 890 895

Asn Val Val Asn Leu Leu Gly Ala Cys Thr Lys Pro Gly Gly Pro Leu
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 Met Val Ile Val Glu Phe Cys Lys Phe Gly Asn Leu Ser Thr Tyr Leu
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 Arg Ser Lys Arg Asn Glu Phe Val Pro Tyr Lys Thr Lys Gly Ala Arg
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 Phe Arg Gln Gly Lys Asp Tyr Val Gly Ala Ile Pro Val Asp Leu Lys
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 Arg Arg Leu Asp Ser Ile Thr Ser Ser Gln Ser Ser Ala Ser Ser Gly
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 Phe Val Glu Glu Lys Ser Leu Ser Asp Val Glu Glu Glu Glu Ala Pro
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 Glu Asp Leu Tyr Lys Asp Phe Leu Thr Leu Glu His Leu Ile Cys Tyr
 995 1000 1005
 Ser Phe Gln Val Ala Lys Gly Met Glu Phe Leu Ala Ser Arg Lys
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 Cys Ile His Arg Asp Leu Ala Ala Arg Asn Ile Leu Leu Ser Glu
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 Lys Asn Val Val Lys Ile Cys Asp Phe Gly Leu Ala Arg Asp Ile
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 1070 1075 1080
 Ile Gln Ser Asp Val Trp Ser Phe Gly Val Leu Leu Trp Glu Ile
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 Phe Ser Leu Gly Ala Ser Pro Tyr Pro Gly Val Lys Ile Asp Glu
 1100 1105 1110
 Glu Phe Cys Arg Arg Leu Lys Glu Gly Thr Arg Met Arg Ala Pro
 1115 1120 1125
 Asp Tyr Thr Thr Pro Glu Met Tyr Gln Thr Met Leu Asp Cys Trp
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 His Gly Glu Pro Ser Gln Arg Pro Thr Phe Ser Glu Leu Val Glu
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His Leu Gly Asn Leu Leu Gln Ala Asn Ala Gln Gln Asp Gly Lys
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Asp Tyr Ile Val Leu Pro Ile Ser Glu Thr Leu Ser Met Glu Glu
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Asp Ser Gly Leu Ser Leu Pro Thr Ser Pro Val Ser Cys Met Glu
1190 1195 1200

Glu Glu Glu Val Cys Asp Pro Lys Phe His Tyr Asp Asn Thr Ala
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Gly Ile Ser Gln Tyr Leu Gln Asn Ser Lys Arg Lys Ser Arg Pro
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Val Ser Val Lys Thr Phe Glu Asp Ile Pro Leu Glu Glu Pro Glu
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Val Lys Val Ile Pro Asp Asp Asn Gln Thr Asp Ser Gly Met Val
1250 1255 1260

Leu Ala Ser Glu Glu Leu Lys Thr Leu Glu Asp Arg Thr Lys Leu
1265 1270 1275

Ser Pro Ser Phe Gly Gly Met Val Pro Ser Lys Ser Arg Glu Ser
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Val Ala Ser Glu Gly Ser Asn Gln Thr Ser Gly Tyr Gln Ser Gly
1295 1300 1305

Tyr His Ser Asp Asp Thr Asp Thr Thr Val Tyr Ser Ser Glu Glu
1310 1315 1320

Ala Glu Leu Leu Lys Leu Ile Glu Ile Gly Val Gln Thr Gly Ser
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Pro Pro Val
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<210> 7
 <211> 223
 <212> PRT
 <213> Homo sapiens
 <400> 7

Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val
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Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr
 20 25 30

Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu
 35 40 45

Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys
 50 55 60

Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser
 65 70 75 80

Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys
 85 90 95

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Cys	Lys	Val	Ser	Asn	Lys	Ala	Leu	Pro	Ala	Pro	Ile	Glu	Lys	Thr	Ile
			100					105					110		
Ser	Lys	Ala	Lys	Gly	Gln	Pro	Arg	Glu	Pro	Gln	Val	Tyr	Thr	Leu	Pro
		115					120					125			
Pro	Ser	Arg	Asp	Glu	Leu	Thr	Lys	Asn	Gln	Val	Ser	Leu	Thr	Cys	Leu
		130				135					140				
Val	Lys	Gly	Phe	Tyr	Pro	Ser	Asp	Ile	Ala	Val	Glu	Trp	Glu	Ser	Asn
145					150					155					160
Gly	Gln	Pro	Glu	Asn	Asn	Tyr	Lys	Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser
			165						170					175	
Asp	Gly	Pro	Phe	Phe	Leu	Tyr	Ser	Lys	Leu	Thr	Val	Asp	Lys	Ser	Arg
			180					185					190		
Trp	Gln	Gln	Gly	Asn	Val	Phe	Ser	Cys	Ser	Val	Met	His	Glu	Ala	Leu
	195						200					205			
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<210> 8
 <211> 669
 <212> DNA
 <213> Homo sapiens

<400> 8	
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gacgtgagcc acgaagaccc tgagggtcaag ttcaactggt acgtggacgg cgtggaggtg	180
cataatgcca agacaaagcc gcgggaggag cagtacaaca gcacgtaccg tgtgggtcagc	240
gtcctcaccg tcctgcacca ggactggctg aatggcaagg agtacaagtg caaggctctcc	300
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gaaccacagg tgtacaccct gcccccatcc cgggatgagc tgaccaagaa ccaggtcagc	420
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gggcagccgg agaacaacta caagaccacg cctcccgctg tggactccga cggccccctc	540
ttcctctaca gcaagctcac cgtggacaag agcagggtggc agcaggggaa cgtcttctca	600
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ccgggtaaa	669

<210> 9
 <211> 580
 <212> PRT
 <213> Artificial sequence

<220>

<223> 19# fused protein

<400> 9

Arg Pro Ser Pro Thr Leu Pro Glu Gln Ala Gln Pro Trp Gly Ala Pro
1 5 10 15

Val Glu Val Glu Ser Phe Leu Val His Pro Gly Asp Leu Leu Gln Leu
20 25 30

Arg Cys Arg Leu Arg Asp Asp Val Gln Ser Ile Asn Trp Leu Arg Asp
35 40 45

Gly Val Gln Leu Ala Glu Ser Asn Arg Thr Arg Ile Thr Gly Glu Glu
50 55 60

Val Glu Val Gln Asp Ser Val Pro Ala Asp Ser Gly Leu Tyr Ala Cys
65 70 75 80

Val Thr Ser Ser Pro Ser Gly Ser Asp Thr Thr Tyr Phe Ser Val Asn
85 90 95

Val Ser Asp Ala Leu Pro Ser Ser Glu Asp Asp Asp Asp Asp Asp Asp
100 105 110

Ser Ser Ser Glu Glu Lys Glu Thr Asp Asn Thr Lys Pro Asn Pro Val
115 120 125

Ala Pro Tyr Trp Thr Ser Pro Glu Lys Met Glu Lys Lys Leu His Ala
130 135 140

Val Pro Ala Ala Lys Thr Val Lys Phe Lys Cys Pro Ser Ser Gly Thr
145 150 155 160

Pro Asn Pro Thr Leu Arg Trp Leu Lys Asn Gly Lys Glu Phe Lys Pro
165 170 175

Asp His Arg Ile Gly Gly Tyr Lys Val Arg Tyr Ala Thr Trp Ser Ile
180 185 190

Ile Met Asp Ser Val Val Pro Ser Asp Lys Gly Asn Tyr Thr Cys Ile
195 200 205

Val Glu Asn Glu Tyr Gly Ser Ile Asn His Thr Tyr Gln Leu Asp Val
210 215 220

Val Glu Arg Ser Pro His Arg Pro Ile Leu Gln Ala Gly Leu Pro Ala
225 230 235 240

Asn Lys Thr Val Ala Leu Gly Ser Asn Val Glu Phe Met Cys Lys Val
245 250 255

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Tyr Ser Asp Pro Gln Pro His Ile Gln Trp Leu Lys His Ile Glu Val
 260 265 270
 Asn Gly Ser Lys Ile Gly Pro Asp Asn Leu Pro Tyr Val Gln Ile Leu
 275 280 285
 Lys Thr Ala Gly Val Asn Thr Thr Asp Lys Glu Met Glu Val Leu His
 290 295 300
 Leu Arg Asn Val Ser Phe Glu Asp Ala Gly Glu Tyr Thr Cys Leu Ala
 305 310 315 320
 Gly Asn Ser Ile Gly Leu Ser His His Ser Ala Trp Leu Thr Val Leu
 325 330 335
 Glu Ala Leu Glu Glu Arg Pro Ala Val Met Thr Ser Pro Leu Tyr Leu
 340 345 350
 Glu Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu
 355 360 365
 Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu
 370 375 380
 Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser
 385 390 395 400
 His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu
 405 410 415
 Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr
 420 425 430
 Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn
 435 440 445
 Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro
 450 455 460
 Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln
 465 470 475 480
 Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val
 485 490 495
 Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val
 500 505 510
 Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro
 515 520 525

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Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr
 530 535 540

Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val
 545 550 555 560

Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu
 565 570 575

Ser Pro Gly Lys
 580

<210> 10
 <211> 526
 <212> PRT
 <213> artificial sequence

<220>
 <223> 13# fused protein

<400> 10

Lys Asn Arg Thr Arg Ile Thr Gly Glu Glu Val Glu Val Gln Asp Ser
 1 5 10 15

Val Pro Ala Asp Ser Gly Leu Tyr Ala Cys Val Thr Ser Ser Pro Ser
 20 25 30

Gly Ser Asp Thr Thr Tyr Phe Ser Val Asn Val Ser Asp Ala Leu Pro
 35 40 45

Ser Ser Glu Asp Asp Asp Asp Asp Asp Ser Ser Ser Glu Glu Lys
 50 55 60

Glu Thr Asp Asn Thr Lys Pro Asn Pro Val Ala Pro Tyr Trp Thr Ser
 65 70 75 80

Pro Glu Lys Met Glu Lys Lys Leu His Ala Val Pro Ala Ala Lys Thr
 85 90 95

Val Lys Phe Lys Cys Pro Ser Ser Gly Thr Pro Asn Pro Thr Leu Arg
 100 105 110

Trp Leu Lys Asn Gly Lys Glu Phe Lys Pro Asp His Arg Ile Gly Gly
 115 120 125

Tyr Lys Val Arg Tyr Ala Thr Trp Ser Ile Ile Met Asp Ser Val Val
 130 135 140

Pro Ser Asp Lys Gly Asn Tyr Thr Cys Ile Val Glu Asn Glu Tyr Gly
 145 150 155 160

Ser Ile Asn His Thr Tyr Gln Leu Asp Val Val Glu Arg Ser Pro His

Arg Pro Ile Leu Gln Ala Gly Leu Pro Ala Asn Lys Thr Val Ala Leu
 180 185 190
 Gly Ser Asn Val Glu Phe Met Cys Lys Val Tyr Ser Asp Pro Gln Pro
 195 200 205
 His Ile Gln Trp Leu Lys His Ile Glu Val Asn Gly Ser Lys Ile Gly
 210 215 220
 Pro Asp Asn Leu Pro Tyr Val Gln Ile Leu Lys Thr Ala Gly Val Asn
 225 230 235 240
 Thr Thr Asp Lys Glu Met Glu Val Leu His Leu Arg Asn Val Ser Phe
 245 250 255
 Glu Asp Ala Gly Glu Tyr Thr Cys Leu Ala Gly Asn Ser Ile Gly Leu
 260 265 270
 Ser His His Ser Ala Trp Leu Thr Val Leu Glu Ala Leu Glu Glu Arg
 275 280 285
 Pro Ala Val Met Thr Ser Pro Leu Tyr Leu Glu Asp Lys Thr His Thr
 290 295 300
 Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe
 305 310 315 320
 Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro
 325 330 335
 Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val
 340 345 350
 Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr
 355 360 365
 Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val
 370 375 380
 Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys
 385 390 395 400
 Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser
 405 410 415
 Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro
 420 425 430
 Ser Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val

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435

440

445

Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly
 450 455 460

Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp
 465 470 475 480

Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp
 485 490 495

Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His
 500 505 510

Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys
 515 520 525

<210> 11
 <211> 500
 <212> PRT
 <213> artificial sequence

<220>
 <223> 22# fused protein

<400> 11

Val Thr Ser Ser Pro Ser Gly Ser Asp Thr Thr Tyr Phe Ser Val Asn
 1 5 10 15

Val Ser Asp Ala Leu Pro Ser Ser Glu Asp Asp Asp Asp Asp Asp Asp
 20 25 30

Ser Ser Ser Glu Glu Lys Glu Thr Asp Asn Thr Lys Pro Asn Pro Val
 35 40 45

Ala Pro Tyr Trp Thr Ser Pro Glu Lys Met Glu Lys Lys Leu His Ala
 50 55 60

Val Pro Ala Ala Lys Thr Val Lys Phe Lys Cys Pro Ser Ser Gly Thr
 65 70 75 80

Pro Asn Pro Thr Leu Arg Trp Leu Lys Asn Gly Lys Glu Phe Lys Pro
 85 90 95

Asp His Arg Ile Gly Gly Tyr Lys Val Arg Tyr Ala Thr Trp Ser Ile
 100 105 110

Ile Met Asp Ser Val Val Pro Ser Asp Lys Gly Asn Tyr Thr Cys Ile
 115 120 125

Val Glu Asn Glu Tyr Gly Ser Ile Asn His Thr Tyr Gln Leu Asp Val
 130 135 140

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Val Glu Arg Ser Pro His Arg Pro Ile Leu Gln Ala Gly Leu Pro Ala
 145 150 155 160
 Asn Lys Thr Val Ala Leu Gly Ser Asn Val Glu Phe Met Cys Lys Val
 165 170 175
 Tyr Ser Asp Pro Gln Pro His Ile Gln Trp Leu Lys His Ile Glu Val
 180 185 190
 Asn Gly Ser Lys Ile Gly Pro Asp Asn Leu Pro Tyr Val Gln Ile Leu
 195 200 205
 Lys Thr Ala Gly Val Asn Thr Thr Asp Lys Glu Met Glu Val Leu His
 210 215 220
 Leu Arg Asn Val Ser Phe Glu Asp Ala Gly Glu Tyr Thr Cys Leu Ala
 225 230 235 240
 Gly Asn Ser Ile Gly Leu Ser His His Ser Ala Trp Leu Thr Val Leu
 245 250 255
 Glu Ala Leu Glu Glu Arg Pro Ala Val Met Thr Ser Pro Leu Tyr Leu
 260 265 270
 Glu Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu
 275 280 285
 Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu
 290 295 300
 Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser
 305 310 315 320
 His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu
 325 330 335
 Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr
 340 345 350
 Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn
 355 360 365
 Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro
 370 375 380
 Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln
 385 390 395 400
 Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val
 405 410 415

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Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val
 420 425 430

Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro
 435 440 445

Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr
 450 455 460

Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val
 465 470 475 480

Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu
 485 490 495

Ser Pro Gly Lys
 500

<210> 12
 <211> 468
 <212> PRT
 <213> artificial sequence

<220>
 <223> 23# fused protein

<400> 12

Ser Ser Ser Glu Glu Lys Glu Thr Asp Asn Thr Lys Pro Asn Pro Val
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Ala Pro Tyr Trp Thr Ser Pro Glu Lys Met Glu Lys Lys Leu His Ala
 20 25 30

Val Pro Ala Ala Lys Thr Val Lys Phe Lys Cys Pro Ser Ser Gly Thr
 35 40 45

Pro Asn Pro Thr Leu Arg Trp Leu Lys Asn Gly Lys Glu Phe Lys Pro
 50 55 60

Asp His Arg Ile Gly Gly Tyr Lys Val Arg Tyr Ala Thr Trp Ser Ile
 65 70 75 80

Ile Met Asp Ser Val Val Pro Ser Asp Lys Gly Asn Tyr Thr Cys Ile
 85 90 95

Val Glu Asn Glu Tyr Gly Ser Ile Asn His Thr Tyr Gln Leu Asp Val
 100 105 110

Val Glu Arg Ser Pro His Arg Pro Ile Leu Gln Ala Gly Leu Pro Ala
 115 120 125

Asn Lys Thr Val Ala Leu Gly Ser Asn Val Glu Phe Met Cys Lys Val

130

135

Tyr Ser Asp Pro Gln Pro His Ile Gln Trp Leu Lys His Ile Glu Val
145 150 155 160

Asn Gly Ser Lys Ile Gly Pro Asp Asn Leu Pro Tyr Val Gln Ile Leu
165 170 175

Lys Thr Ala Gly Val Asn Thr Thr Asp Lys Glu Met Glu Val Leu His
180 185 190

Leu Arg Asn Val Ser Phe Glu Asp Ala Gly Glu Tyr Thr Cys Leu Ala
195 200 205

Gly Asn Ser Ile Gly Leu Ser His His Ser Ala Trp Leu Thr Val Leu
210 215 220

Glu Ala Leu Glu Glu Arg Pro Ala Val Met Thr Ser Pro Leu Tyr Leu
225 230 235 240

Glu Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu
245 250 255

Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu
260 265 270

Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser
275 280 285

His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu
290 295 300

Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr
305 310 315 320

Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn
325 330 335

Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro
340 345 350

Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln
355 360 365

Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val
370 375 380

Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val
385 390 395 400

Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro

Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr
 420 425 430

Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val
 435 440 445

Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu
 450 455 460

Ser Pro Gly Lys
 465

<210> 13
 <211> 457
 <212> PRT
 <213> artificial sequence

<220>
 <223> 26# fused protein

<400> 13

Lys Pro Asn Pro Val Ala Pro Tyr Trp Thr Ser Pro Glu Lys Met Glu
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Lys Lys Leu His Ala Val Pro Ala Ala Lys Thr Val Lys Phe Lys Cys
 20 25 30

Pro Ser Ser Gly Thr Pro Asn Pro Thr Leu Arg Trp Leu Lys Asn Gly
 35 40 45

Lys Glu Phe Lys Pro Asp His Arg Ile Gly Gly Tyr Lys Val Arg Tyr
 50 55 60

Ala Thr Trp Ser Ile Ile Met Asp Ser Val Val Pro Ser Asp Lys Gly
 65 70 75 80

Asn Tyr Thr Cys Ile Val Glu Asn Glu Tyr Gly Ser Ile Asn His Thr
 85 90 95

Tyr Gln Leu Asp Val Val Glu Arg Ser Pro His Arg Pro Ile Leu Gln
 100 105 110

Ala Gly Leu Pro Ala Asn Lys Thr Val Ala Leu Gly Ser Asn Val Glu
 115 120 125

Phe Met Cys Lys Val Tyr Ser Asp Pro Gln Pro His Ile Gln Trp Leu
 130 135 140

Lys His Ile Glu Val Asn Gly Ser Lys Ile Gly Pro Asp Asn Leu Pro
 145 150 155 160

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Tyr Val Gln Ile Leu Lys Thr Ala Gly Val Asn Thr Thr Asp Lys Glu
 165 170 175
 Met Glu Val Leu His Leu Arg Asn Val Ser Phe Glu Asp Ala Gly Glu
 180 185 190
 Tyr Thr Cys Leu Ala Gly Asn Ser Ile Gly Leu Ser His His Ser Ala
 195 200 205
 Trp Leu Thr Val Leu Glu Ala Leu Glu Glu Arg Pro Ala Val Met Thr
 210 215 220
 Ser Pro Leu Tyr Leu Glu Asp Lys Thr His Thr Cys Pro Pro Cys Pro
 225 230 235 240
 Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys
 245 250 255
 Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val
 260 265 270
 Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr
 275 280 285
 Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu
 290 295 300
 Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His
 305 310 315 320
 Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys
 325 330 335
 Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln
 340 345 350
 Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu
 355 360 365
 Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro
 370 375 380
 Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn
 385 390 395 400
 Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu
 405 410 415
 Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val
 420 425 430

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Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln
435 440 445

Lys Ser Leu Ser Leu Ser Pro Gly Lys
450 455

<210> 14
<211> 451
<212> PRT
<213> artificial sequence

<220>
<223> 29# fused protein

<400> 14

Pro Tyr Trp Thr Ser Pro Glu Lys Met Glu Lys Lys Leu His Ala Val
1 5 10 15

Pro Ala Ala Lys Thr Val Lys Phe Lys Cys Pro Ser Ser Gly Thr Pro
20 25 30

Asn Pro Thr Leu Arg Trp Leu Lys Asn Gly Lys Glu Phe Lys Pro Asp
35 40 45

His Arg Ile Gly Gly Tyr Lys Val Arg Tyr Ala Thr Trp Ser Ile Ile
50 55 60

Met Asp Ser Val Val Pro Ser Asp Lys Gly Asn Tyr Thr Cys Ile Val
65 70 75 80

Glu Asn Glu Tyr Gly Ser Ile Asn His Thr Tyr Gln Leu Asp Val Val
85 90 95

Glu Arg Ser Pro His Arg Pro Ile Leu Gln Ala Gly Leu Pro Ala Asn
100 105 110

Lys Thr Val Ala Leu Gly Ser Asn Val Glu Phe Met Cys Lys Val Tyr
115 120 125

Ser Asp Pro Gln Pro His Ile Gln Trp Leu Lys His Ile Glu Val Asn
130 135 140

Gly Ser Lys Ile Gly Pro Asp Asn Leu Pro Tyr Val Gln Ile Leu Lys
145 150 155 160

Thr Ala Gly Val Asn Thr Thr Asp Lys Glu Met Glu Val Leu His Leu
165 170 175

Arg Asn Val Ser Phe Glu Asp Ala Gly Glu Tyr Thr Cys Leu Ala Gly
180 185 190

Asn Ser Ile Gly Leu Ser His His Ser Ala Trp Leu Thr Val Leu Glu

195

200

205

Ala Leu Glu Glu Arg Pro Ala Val Met Thr Ser Pro Leu Tyr Leu Glu
210 215 220

Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly
225 230 235 240

Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met
245 250 255

Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His
260 265 270

Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val
275 280 285

His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr
290 295 300

Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly
305 310 315 320

Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile
325 330 335

Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val
340 345 350

Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val Ser
355 360 365

Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu
370 375 380

Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro
385 390 395 400

Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val
405 410 415

Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met
420 425 430

His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser
435 440 445

Pro Gly Lys
450

<210> 15

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<211> 446
 <212> PRT
 <213> artificial sequence

<220>
 <223> 8# fused protein

<400> 15

Pro Glu Lys Met Glu Lys Lys Leu His Ala Val Pro Ala Ala Lys Thr
 1 5 10 15

Val Lys Phe Lys Cys Pro Ser Ser Gly Thr Pro Asn Pro Thr Leu Arg
 20 25 30

Trp Leu Lys Asn Gly Lys Glu Phe Lys Pro Asp His Arg Ile Gly Gly
 35 40 45

Tyr Lys Val Arg Tyr Ala Thr Trp Ser Ile Ile Met Asp Ser Val Val
 50 55 60

Pro Ser Asp Lys Gly Asn Tyr Thr Cys Ile Val Glu Asn Glu Tyr Gly
 65 70 75 80

Ser Ile Asn His Thr Tyr Gln Leu Asp Val Val Glu Arg Ser Pro His
 85 90 95

Arg Pro Ile Leu Gln Ala Gly Leu Pro Ala Asn Lys Thr Val Ala Leu
 100 105 110

Gly Ser Asn Val Glu Phe Met Cys Lys Val Tyr Ser Asp Pro Gln Pro
 115 120 125

His Ile Gln Trp Leu Lys His Ile Glu Val Asn Gly Ser Lys Ile Gly
 130 135 140

Pro Asp Asn Leu Pro Tyr Val Gln Ile Leu Lys Thr Ala Gly Val Asn
 145 150 155 160

Thr Thr Asp Lys Glu Met Glu Val Leu His Leu Arg Asn Val Ser Phe
 165 170 175

Glu Asp Ala Gly Glu Tyr Thr Cys Leu Ala Gly Asn Ser Ile Gly Leu
 180 185 190

Ser His His Ser Ala Trp Leu Thr Val Leu Glu Ala Leu Glu Glu Arg
 195 200 205

Pro Ala Val Met Thr Ser Pro Leu Tyr Leu Glu Asp Lys Thr His Thr
 210 215 220

Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe
 225 230 235 240

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Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro
 245 250 255

Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val
 260 265 270

Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr
 275 280 285

Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val
 290 295 300

Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys
 305 310 315 320

Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser
 325 330 335

Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro
 340 345 350

Ser Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val
 355 360 365

Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly
 370 375 380

Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp
 385 390 395 400

Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp
 405 410 415

Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His
 420 425 430

Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys
 435 440 445

<210> 16
 <211> 1743
 <212> DNA
 <213> Artificial sequence

<220>
 <223> Nucleotide sequence of 19# fused protein

<400> 16
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 tcttcctggg tccaccccggtg tgacctgcta cagcttcgct gtcggctgctg ggacgatgtg 120
 cagagcatca actggctgctg ggacgggggtg cagctggctg aaagcaatcg caccgcgcatc 180

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acaggggagg aggtggaggt gcaggactcc gtgcccgcag actccggcct ctatgcttgc 240
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ctccccctct cggaggatga tgatgatgat gatgactcct cttcagagga gaaagaaaaca 360
gataacacca aaccaaacc cgtagctcca tattggacat cccagaaaa gatggaaaag 420
aaattgcatg cagtgcggc tgccaagaca gtgaagttca aatgcccctc cagtgggacc 480
ccaaacccca cactgcgctg gttgaaaaat ggcaaagaat tcaaacctga ccacagaatt 540
ggaggctaca aggtccgtta tgccacctgg agcatcataa tggactctgt ggtgccctct 600
gacaagggca actacacctg cattgtggag aatgagtacg gcagcatcaa ccacacatac 660
cagctggatg tcgtggagcg gtccccctac cggcccatcc tgcaagcagg gttgcccgcc 720
aacaaaacag tggccctggg tagcaacgtg gagttcatgt gtaagggtga cagtgacccg 780
cagccgcaca tccagtggct aaagcacatc gaggtgaatg ggagcaagat tggcccagac 840
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gagggtgctt acttaagaaa tgtctccttt gaggacgcag gggagtatac gtgcttggcg 960
ggtaactcta tcggactctc ccatcactct gcatggttga ccgttctgga agccctggaa 1020
gagaggccgg cagtgatgac ctgccccctg tacctggagg acaaaactca cacatgcccc 1080
ccgtgcccag cactgaact cctgggggga ccgtcagtct tcctcttccc cccaaaaccc 1140
aaggacaccc tcatgatctc ccggacccct gaggtcacat gcgtggtggt ggacgtgagc 1200
cacgaagacc ctgaggtcaa gttcaactgg tacgtggacg gcgtggaggt gcataatgcc 1260
aagacaaagc cgcgggagga gcagtacaac agcacgtacc gtgtggtcag cgtcctcacc 1320
gtcctgcacc aggactggct gaatggcaag gagtacaagt gcaaggctct caacaaagcc 1380
ctcccagccc ccatcgagaa aaccatctcc aaagccaaag ggcagccccg agaaccacag 1440
gtgtacaccc tgcccccatc ccgggatgag ctgaccaaga accaggtcag cctgacctgc 1500
ctggtcaaag gtttctatcc cagcgacatc gccgtggagt gggagagcaa tgggcagccg 1560
gagaacaact acaagaccac gcctcccgtg ctggactccg acggtcctt cttcctctac 1620
agcaagctca ccgtggacaa gagcaggtgg cagcagggga acgtcttctc atgctccgtg 1680
atgcatgagg ctctgcacaa ccactacacg cagaagagcc tctccctgtc tccgggtaaa 1740
tga 1743

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<210> 17
 <211> 1581
 <212> DNA
 <213> Artificial sequence

<220>
 <223> Nucleotide sequence of 13# fused protein

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<400> 17
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tccggcctct atgcttgcgt aaccagcagc cctcggggca gtgacaccac ctacttctcc 120

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gtcaatgttt cagatgctct cccctcctcg gaggatgatg atgatgatga tgactcctct	180
tcagaggaga aagaaacaga taacacccaaa ccaaaccctcg tagctccata ttggacatcc	240
ccagaaaaga tggaaaagaa attgcatgca gtgccggctg ccaagacagt gaagttcaaa	300
tgcccttcca gtgggacccc aaaccccaca ctgcgctggg tgaaaaatgg caaagaattc	360
aaacctgacc acagaattgg aggcatacaag gtccggtatg ccacctggag catcataatg	420
gactctgtgg tgccctctga caagggaac tacacctgca ttgtggagaa tgagtacggc	480
agcatcaacc acacatacca gctggatgtc gtggagcggg cccctcaccg gcccatcctg	540
caagcagggg tgcccgccaa caaacacagt gccctgggta gcaacgtgga gtcatgtgt	600
aagggtgtaca gtgacccgca gccgcacatc cagtggctaa agcacatcga ggtgaatggg	660
agcaagattg gccagacaa cctgcccttat gtccagatct tgaagactgc tggagttaat	720
accaccgaca aagagatgga ggtgcttcac ttaagaaatg tctcccttga ggacgcaggg	780
gagtatacgt gcttggcggg taactctatc ggactctccc atcactctgc atggttgacc	840
gttctggaag ccctggaaga gaggccggca gtgatgacct cggccctgta cctggaggac	900
aaaactcaca catgccacc gtgcccagca cctgaactcc tggggggacc gtcagtcttc	960
ctcttcccc caaaacccaa ggacaccctc atgatctccc ggacccctga ggtcacatgc	1020
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aagggtctcca acaaagccct ccagccccc atcgagaaaa ccatctccaa agccaaaggg	1260
cagccccgag aaccacaggt gtacaccctg ccccatccc gggatgagct gaccaagaac	1320
caggctcagc tgacctgcct ggtcaaaggc tcttatccca gcgacatcgc cgtggagtgg	1380
gagagcaatg ggcagccgga gaacaactac aagaccacgc ctcccgtgct ggactccgac	1440
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<210> 18
 <211> 1503
 <212> DNA
 <213> Artificial sequence

<220>
 <223> Nucleotide sequence of 22# fused protein

<400> 18	
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ctccccctct cggaggatga tgatgatgat gatgactcct cttcagagga gaaagaaaca	120
gataacacca aaccaaacc cgtagctcca tattggacat cccagaaaa gatggaaaag	180
aaattgcatg cagtgccggc tgccaagaca gtgaagttca aatgcccttc cagtgggacc	240

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ccaaacccca cactgcgctg gttgaaaaat ggcaaagaat tcaaacctga ccacagaatt 300
ggagggtaca aggtccgtta tgccacctgg agcatcataa tggactctgt ggtgccctct 360
gacaagggca actacacctg cattgtggag aatgagtagc gcagcatcaa ccacacatac 420
cagctggatg tcgtggagcg gtccccctac cggcccatcc tgcaagcagg gttgccccgc 480
aataaaacag tggccctggg tagcaacgtg gagttcatgt gtaagggtga cagtgaacccg 540
cagccgcaca tccagtggct aaagcacatc gaggtgaatg ggagcaagat tggcccagac 600
aacctgcctt atgtccagat ctgaagact gctggagtta ataccaccga caaagagatg 660
gaggtgtctc acttaagaaa tgtctccttt gaggacgcag gggagtatac gtgcttggcg 720
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gagaggccgg cagtgatgac ctgcacctg tacctggagg aaaaaactca cacatgcccc 840
ccgtgcccag cacctgaact cctgggggga ccgtcagtct tctcttccc cccaaaaccc 900
aaggacaccc tcattgatctc ccggacccct gaggtcacat gcgtgggtgt ggacgtgagc 960
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agcaagctca ccgtggacaa gagcaggtgg cagcagggga acgtcttctc atgctccgtg 1440
atgcatgagg ctctgcacaa ccactacacg cagaagagcc tctcctgtc tccgggtaaa 1500
tga 1503

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<210> 19
 <211> 1407
 <212> DNA
 <213> Artificial sequence

<220>
 <223> Nucleotide sequence of 23# fused protein

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<400> 19
tctctttcag aggagaaaga aacagataac accaaaccaa accccgtagc tccatattgg 60
acatccccag aaaagatgga aaagaaattg catgcagtgc cggctgcca gacagtgaag 120
ttcaaattgcc ctccagtgg gaccccaaac cccacactgc gctggttgaa aaatggcaaa 180
gaattcaaac ctgaccacag aattggaggc tacaaggctc gttatgccac ctggagcatc 240
ataatggact ctgtggtgcc ctctgacaag ggcaactaca cctgcattgt ggagaatgag 300
tacggcagca tcaaccacac ataccagctg gatgtcgtgg agcgggtccc tcaccggccc 360
atcctgcaag cagggttgcc cgccaacaaa acagtggccc tgggtagcaa cgtggagttc 420

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atgtgtaagg	tgtacagtga	cccgagccg	cacatccagt	ggctaaagca	catcgagggtg	480
aatgggagca	agattggccc	agacaacctg	ccttatgtcc	agatcttgaa	gactgctgga	540
gttaatacca	ccgacaaaga	gatggagggtg	cttcacttaa	gaaatgtctc	ctttgaggac	600
gcaggggagt	atacgtgctt	ggcgggtaac	tctatcggac	tctcccatca	ctctgcatgg	660
ttgaccgttc	tggaagccct	ggaagagagg	ccggcagtga	tgacctcgcc	cctgtacctg	720
gaggacaaaa	ctcacacatg	cccaccgtgc	ccagcacctg	aactcctggg	gggaccgtca	780
gtcttctctt	ccccccaaa	acccaaggac	acctcatga	tctccggac	ccctgagggtc	840
acatgcgtgg	tggtggacgt	gagccacgaa	gacctgagg	tcaagttcaa	ctggtacctg	900
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aagtgcagg	tctccaacaa	agccctccca	gcccccatcg	agaaaaccat	ctccaaagcc	1080
aaagggcagc	cccgagaacc	acaggtgtac	acctgcccc	catccccgga	tgagctgacc	1140
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gagtgggaga	gcaatgggca	gccggagAAC	aactacaaga	ccacgcctcc	cgtgctggac	1260
tccgaagggt	ctttcttctt	ctacagcaag	ctcaccgtgg	acaagagcag	gtggcagcag	1320
gggaacgtct	tctcatgctc	cgtgatgcat	gaggctctgc	acaaccacta	cacgcagaag	1380
agcctctccc	tgtctccggg	taaatga				1407

<210> 20
 <211> 1374
 <212> DNA
 <213> artificial sequence

<220>
 <223> Nucleotide sequence of 26# fused protein

<400> 20	
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gcagtgccgg	ctgccaagac agtgaagttc aaatgccctt ccagtgggac cccaaacccc 120
acactgcgct	ggttgaaaaa tggcaaagaa ttcaaacctg accacagaat tggaggctac 180
aagggtccgtt	atgccacctg gagcatcata atggactctg tggtgccctc tgacaagggc 240
aactacacct	gcattgtgga gaatgagtac ggcagcatca accacacata ccagctggat 300
gtcgtggagc	ggfccccctc ccggcccatc ctgcaagcag ggttgcccg ccaaaaaaca 360
gtggccctgg	gtagcaacgt ggagttcatg tgtaagggtg acagtgacct gcagccgcac 420
atccagtggc	taaagcacat cgagggtgaat gggagcaaga ttggcccaga caacctgcct 480
tatgtccaga	tcttgaagac tgctggagtt aataccaccg acaaagagat ggaggtgctt 540
cacttaagaa	atgtctctct tgaggacgca ggggagtata cgtgcttggc gggttaactc 600
atcggactct	cccatcactc tgcattggtg accgttctgg aagccctgga agagaggccg 660
gcagtgatga	cctcgccccc gtacctggag gacaaaactc acacatgccc accgtgcccc 720

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gcacctgaac	tcctgggggg	accgtcagtc	ttctctttcc	ccccaaaacc	caaggacacc	780
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cctgagggtca	agttcaactg	gtacgtggac	ggcgtggagg	tgcataatgc	caagacaaag	900
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caggactggr	tgaatggcaa	ggagtacaag	tgcaagggtct	ccaacaaagc	cctcccagcc	1020
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tacaagacca	cgctcccggt	gctggactcc	gacggctcct	tcttcctcta	cagcaagetc	1260
accgtggaca	agagcaggtg	gcagcagggg	aacgtcttct	catgctccgt	gatgcatgag	1320
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<210> 21
 <211> 1356
 <212> DNA
 <213> Artificial sequence

<220>
 <223> nucleotide sequence of 29# fused protein

<400> 21	
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aatggcaaaag	aattcaaaacc tgaccacaga attggagggt acaagggtccg ttatgccacc 180
tggagcatca	taatggactc tgtggtgccc tctgacaagg gcaactacac ctgcattgtg 240
gagaatgagt	acggcagcat caaccacaca taccagctgg atgtcgtgga gcgggtccccct 300
caccggccca	tcctgcaagc aggggttgcgc gccaaacaaa cagtggccctt gggtagcaac 360
gtggagttca	tgtgtaagggt gtacagtgac ccgcagccgc acatccagtg gctaaagcac 420
atcgagggtga	atgggagcaa gattggccca gacaacctgc cttatgtcca gatcttgaag 480
actgctggag	ttaataccac cgacaaagag atggagggtgc ttactttaag aaatgtctcc 540
tttgaggacg	caggggagta tacgtgcttg gcgggtaact ctatcggact ctcccatcac 600
tctgcatggt	tgaccgttct ggaagccctg gaagagagggc cggcagtgat gacctgcgcc 660
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cctgagggtca	catgcgtgggt ggtggacgtg agccacgaag accctgagggt caagttcaac 840
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aacagcacgt	accgtgtggt cagcgtcctc accgtcctgc accaggactg gctgaatggc 960
aaggagtaca	agtgcaagggt ctccaacaaa gccctcccag ccccatcga gaaaaccatc 1020
tccaaagcca	aagggcagcc ccgagaacca cagggtgtaca ccctgcccc atcccgggat 1080

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gtgctggact	ccgacggctc	cttcttcttc	tacagcaagc	tcaccgtgga	caagagcagg	1260
tggcagcagg	ggaacgtctt	ctcatgctcc	gtgatgcatg	aggctctgca	caaccactac	1320
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<210> 22
 <211> 1341
 <212> DNA
 <213> artificial sequence

<220>
 <223> Nucleotide sequence of 8# fused protein

<400> 22	
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tggaaaagaa	
attgcatgca	
gtgccggctg	
ccaagacagt	
gaagttcaaa	
tgcccttcca	120
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aaaccccaca	
ctgcgctggg	
tgaaaaatgg	
caaagaattc	
aaacctgacc	180
acagaattgg	
aggctacaag	
gtccgttatg	
ccacctggag	
catcataatg	
gactctgtgg	240
tgcctcttga	
caagggcaac	
tacacctgca	
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agcatcaacc	300
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gctggatgtc	
gtggagcggg	
ccccctaccg	
gcccattcctg	
caagcagggg	360
tgcctgcca	
caaaacagtg	
gccctgggta	
gcaacgtgga	
gttcatgtgt	
aaggtgtaca	420
gtgacccgca	
gccgcacatc	
cagtggctaa	
agcacatcga	
ggtgaatggg	
agcaagattg	480
gcccagacaa	
cctgccttat	
gtccagatct	
tgaagactgc	
tggagttaat	
accaccgaca	540
aagagatgga	
ggtgcttcac	
ttaagaaatg	
tctcctttga	
ggacgcaggg	
gagtatacgt	600
gcttggcggg	
taactctatc	
ggactctccc	
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