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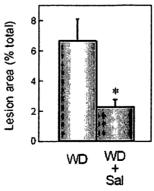
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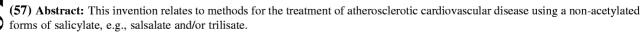
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(54) Title: TREATMENT OF CARDIOVASCULAR DISEASE WITH SALICYLATES

FIG. 3A Whole aorta en face (n=7)







## Treatment of Cardiovascular Disease With Salicylates

#### CROSS-REFERENCE TO RELATED APPLICATIONS

This application is related to U.S. Provisional Patent Applications Serial No. 60/958,185, filed on July 3, 2007, and 61/051,228, filed May 7, 2008, the entire contents of which are hereby incorporated by reference.

#### FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

This invention was made with Government support under Grant No. P50HL083813 awarded by the National Institutes of Health. The Government has certain rights in the invention.

#### **TECHNICAL FIELD**

This invention relates to methods for the treatment of atherosclerotic cardiovascular disease (ACVD) using an NF-kB inhibitor, e.g., non-acetylated forms of salicylate, e.g., salsalate and trilisate.

15 BACKGROUND

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Each year, cardiovascular disease (CVD) kills more Americans than cancer (United States Dept. of Health and Human Services, Center for Disease Control, Chronic Disease Overview. (1999)). Diseases of the heart alone caused 30% of all deaths (Id.). ACVD is the number one cause of death and disability in the United States and in most of Europe. In large part, the rise in ACVD is due to an increase in the risk factors for ACVD: abdominal obesity, Western diet and a sedentary life-style. Inflammation participates in the pathogenesis of cardiovascular disease (CVD) (Hansson, N. Engl. J. Med., 352(16):1685-1695 (2005); Hansson and Libby, Nat. Rev. Immunol. 6(7):508-519 (2006)).

25 SUMMARY

The present inventors have discovered that NF-κB mediated inflammation is a "common soil" involved in the pathogenesis of a number of inflammatory diseases associated with one or more of: abdominal obesity, Western diet and a sedentary lifestyle, including atherosclerotic cardiovascular disease (ACVD). Therefore, the present methods include the systemic inhibition of NF-κB by administration of high

doses of non-acetylated forms of salicylate, e.g., salsalate, trilisate, and other salicylate esters, amides and thioesters and salicylates having substituents on the ring (e.g., halogen, methyl, or halogenated methyl groups), for preventing or retarding the development or progression of cardiovascular diseases, e.g., atheroma formation.

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In one aspect, the invention provides methods for treating or preventing the development or progression of atherosclerotic cardiovascular disease (ACVD) in a subject. The methods include evaluating the subject for the presence of ACVD, or for the presence of one or more risk factors for ACVD; if the subject has or is at risk for ACVD, selecting the subject, and administering to the subject a therapeutically effective amount of an NF-kappaB inhibitor, e.g., a non-acetylated form of salicylate. In some embodiments, the methods of evaluating the subjects for the presence of ACVD include determining triglyceride levels, free fatty acid levels, cholesterol levels, and/or blood pressure, e.g., diastolic blood pressure, and selecting the subject based on the results of the determination. One or more other tests can also be used, e.g., angiogram (arteriogram); cholesterol test; chest x-ray; CT (computed tomography) scan; duplex scanning; echocardiogram; electrocardiogram (ECG or EKG); exercise stress test (cardiac stress test); intravascular ultrasound; MRI (magnetic resonance imaging) scan; PET (positron emission tomography) scan; and/or pharmacologic stress test.

In some embodiments, if the subject has ACVD, the methods include administering an NF-kappaB inhibitor. In some embodiments, the methods include administering about 2-5 g/day, e.g., about 3-4.5 g/day of the non-acetylated form of salicylate.

In some embodiments, the non-acetylated form of salicylate is salsalate or trilisate. In some embodiments, the ACVD is associated with elevated triglyceride levels, free fatty acid levels, total cholesterol levels, high LDL, and/or low HDL, and the methods include determining and optionally monitoring said levels. In some embodiments, the ACVD is associated with elevated diastolic blood pressure, and the methods include determining and optionally monitoring blood pressure, e.g., diastolic blood pressure, in the subject. Monitoring levels of an analyte or of blood pressure include determining the level at least twice, i.e., one to establish a base line level, and at least a second time to determine what effect the salsalate treatment is having on the analyte or blood pressure. In general, the determinations will be made at sequential

time points, e.g., days or weeks apart, e.g., at least two weeks apart. In some embodiments, the methods include administering an amount of salsalate sufficient to reduce levels of triglycerides, free fatty acids, and/or cholesterol, or to reduce blood pressure, e.g., diastolic blood pressure.

In some embodiments, the subject is not suffering from pain, e.g., does not have severe or substantial pain.

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Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Methods and materials are described herein for use in the present invention; other, suitable methods and materials known in the art can also be used. The materials, methods, and examples are illustrative only and not intended to be limiting. All publications, patent applications, patents, sequences, database entries, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control.

Other features and advantages of the invention will be apparent from the following detailed description and figures, and from the claims.

## **DESCRIPTION OF DRAWINGS**

FIGs. 1A-C are a trio of images of aortas from Ldlr-/- mice fed chow (1A), Western diet (1B), or Western diet (1C), plus salicylate for 6 months. Aortas were removed and are displayed en face.

FIGs. 2A-D are photomicrographs of cross sections of the aortic roots of Ldlr-/- mice fed Western diet (2A-B) or Western diet plus salicylate (2C-D). Lipid in the fatty plaques is stained with oil red O (red in the original).

FIGs. 3A-B are bar graphs showing the results of quantification of lesion sizes in Ldlr-/-mice. 3A, data for 7 mice in each group treated as shown in FIGs. 1A-C. 3B, data for 6 mice/group as analyzed in FIGs. 2A-D. WD = Western diet; WD + Sal = Western diet plus salicylate.

FIGs. 4A-C are bar graphs showing Metabolic parameters in subjects prior to treatment (Pre, grey boxes) and after 2 wk treatment with 4.5 g/d (n = 7) or 3.0 g/d (n = 9) salsalate (Post, black boxes). Fasting data are displayed for (A) triglyceride (TG, mg/mlx0.0113 = mm0l/L), (B) total cholesterol (mg/dl x 0.0259 = mm0l/L), and (C) free fatty acids (FFA, mM). Data represent mean  $\pm$  SEM; \*P<0.05, \*\*P<0.005.

FIGS. 5A-B are line graphs showing levels of free fatty acids (FFA) in response to the liquid meal for subjects treated with the highest tolerated salsalate dose (5A) or placebo (5B) either before (Pre, dashed grey lines) or after (Post, solid black lines) the 4 wk treatment with salsalate. Data represent mean ± SEM.

#### **DETAILED DESCRIPTION**

The present inventors have discovered that NF-κB mediated inflammation is a "common soil" involved in the pathogenesis of inflammatory cardiovascular disease, including atherosclerosis. Therefore, the present methods include the administration of high doses of non-acetylated forms of salicylate, e.g., salsalate, for preventing or retarding these diseases, e.g., preventing or retarding atheroma formation, even in the presence of continued high-risk behavior such as continued adherence to a high-fat Western diet and/or sedentary life style.

## **General Methodology**

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The methods described herein, in general include the treatment or prevention of inflammation-mediated disorders that are associated with abdominal obesity, Western diet and/or a sedentary life-style by administering (e.g., orally administering) systemic doses of non-acetylated forms of salicylate, e.g., salsalate, diflunisal, or trilisate (e.g., choline-magnesium trilisate (CMT)). Salsalate is currently commercially available under the following brand names: Argesic®-SA; Disalcid®; Mono-Gesic®; Salflex®; and Salsitab®, and is presently prescribed to reduce the pain and inflammation (swelling) caused by arthritis or other inflammatory conditions. The conventional dosage is generally about 3000 mg daily, given orally in divided doses as follows: 1) two doses of two 750 mg tablets: 2) two doses of three 500 mg tablets/capsules; or 3) three doses of two 500 mg tablets/capsules. In some embodiments, the present methods generally include the administration of doses ranging from about 3000 to 4500 mg per day, but can also be lower, e.g., 2000 to 2500 mg per day, e.g., in subjects unable to tolerate the higher doses.

The methods described herein will generally include determining whether a subject has an inflammatory cardiovascular disease, or is at high risk for developing a inflammatory cardiovascular disease, and selecting the patient if they have or are at risk for such a disorder. In some embodiments of the methods described herein, the subjects are pain free, e.g., do not have joint pain, e.g., are not taking (or have not

been prescribed) a non-acetylated forms of salicylate for the treatment of pain. In some embodiments, the subjects do not have diabetes or insulin resistance. Subjects who are at risk of ACVD include those who have one or more of the following risk factors: adherence to a Western diet, sedentary lifestyle, abdominal obesity, hypertension, dyslipidemia, or have elevated circulating levels of C-reactive protein CRP or other markers of inflammation, or low circulating levels of adiponectin,

The methods described herein include treating subjects who have, or are at risk of developing, cardiovascular disease, e.g., atherosclerosis. In some embodiments, the methods described herein include the systemic administration of high doses of non-acetylated salicylates in overweight (Body Mass Index (BMI) of 25-29) or obese (BMI of  $\geq$  30) non-diabetic subjects who are pain free, to treat patients with ACVD or to prevent the development of atherosclerotic cardiovascular disease, e.g., subjects who are at risk for developing such disease.

## Non-Acetylated Salicylates

The methods described herein can include the administration of a non-acetylated form of salicylate. A number of such compounds are known in the art, including salsalate, trilisate, and other salicylate esters, amides and thioesters and salicylates having substituents on the ring (e.g., halogen, methyl, or hologenated methyl groups). All of the compounds useful in the methods described herein will have NF-kappaB inhibitory activity in a standard *in vitro* assay.

## <u>Salsalate</u>

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Salsalate (salicylsalicylic acid) is a dimer of salicylic acid; its structural formula is shown below.

#### Chemical Structure:

## <u>Trilisate</u>

Trilisate (choline magnesium trisalicylate) is freely soluble in water. The absolute structure of choline magnesium trisalicylate is not known at this time. Choline magnesium trisalicylate has a molecular formula of  $C_{26}H_{29}O_{10}NMg$ , and a

molecular weight of 539.8. When dissolved in water, trilisate appears to form 5 ions (1 choline ion, 1 magnesium ion, and 3 salicylate ions).

### Other Non-Acetylated Salicylates

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A number of other non-acetylated forms of salicylates are known in the art, including sodium salicylate, choline salicylate, magnesium salicylate, and diflunisal, as well as those that are ring-substituted forms, e.g., ortho, meta, or para-substituted forms; substituents include, e.g., halogen, methyl, or halogenated methyl groups. See, e.g., Soltani and De Brabander, Angewandte Chemie, 117(1), 1724-1727; Furst, Arth. & Rheum. 37(1),1-9 (1994); and Goodman and Gilman, The Pharmacological Basis of Therapeutics, 11th Edition, McGraw Hill (2005).

## Other NF-kappaB Inhibitors

In some embodiments, other NF-kappaB inhibitors can be used in place of or in addition to a non-acetylated form of salicylate, e.g., PS341, dehydroxymethyl-epoxyquinomicin (DHMEQ), include 2-chloro-N- [3, 5di (trifluoromethyl) phenyl]-4-(trifluoromethyl) pyrimidine-5-carboxamide (also known as SP-100030); 3,4-dihydro-4,4-dimethyl-2H-1,2-benzoselenazine (also known as BXT-51072); declopramide (also known as Oxi-104); dexlipotam; aryl, substituted or unsubstituted aralkyl, allyl, and substituted or unsubstituted, linear, branched, or cyclic alkyl esters of salicylic acid; nabumetone; sulindac sulfide; sulindac sulfone; sulfasalazine; and pharmaceutically acceptable salts thereof.

## Inflammation and atherogenesis.

An increasingly accepted sequence of events appears to link inflammation to the development and progression of atheroma formation (Blake and Ridker, J. Intern. Med. 252, 283-294 (2002); Libby, Circulation 104, 365-372 (2001)). For example, signs of inflammation accompany even the earliest accumulation of lipid within the arterial wall. Circulating leukocytes don't adhere to normal endothelium, but during an atherogenic diet the endothelium begins to express adhesion molecules that bind circulating leukocytes. These cell adhesion molecules include P- and E-selectins, ICAM-1, VCAM-1 and members of the immunoglobulin family. The selectins mediate transient rolling of the leucocytes along the endothelium while stronger attachments are mediated by ICAM-1 and VCAM-1 (Adams et al., Lancet 343, 831-836 (1994); Tedder et al., FASEB J. 9, 866-873 (1995)). Mice harboring genetic

deletions of E- or P-selectin or ICAM-1 develop less atherosclerosis than normal mice (Collins et al., J. Exp. Med. 191, 189-194 (2000); Dong et al., J. Clin. Invest 102, 145-152 (1998); Mayadas et al., Cell 74, 541-554 (1993); Nageh et al., Arterioscler. Thromb. Vasc. Biol. 17, 1517-1520 (1997)).

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Adherent inflammatory cells can migrate into the subendothelial space, where they contribute to the local inflammatory response and express a variety of chemoattractants that recruit additional cells as well as augmenting the differentiation of blood derived monocytes into macrophage foam cells and promoting the expression of modified lipoproteins on the macrophage surface (Gu et al., Mol Cell 2, 275-281 (1998); Qiao et al., Am. J. Pathol. 150, 1687-1699 (1997)). Cytokines and chemokines produced locally that participate in these processes include macrophage chemotactic protein (MCP-1) and the macrophage inflammatory proteins MIP-1α, MIP-1 $\beta$ , MIP-2, and MIP-3 $\alpha$ . T-cell activation leads to expression of IFN- $\gamma$  and lymphotoxin, which further amplify the proinflammatory state. Macrophages, endothelial cells and smooth muscles cells produce TNF-α (Barath et al., Am. J. Cardiol. 65, 297-302 (1990); Warner and Libby, J. Immunol. 142, 100-109 (1989)), which along with IFN-γ and IL-1β stimulates the local production of IL-6 in human atheroma (Rus et al., Atherosclerosis 127, 263-271 (1996); Seino et al., Cytokine 6, 87-91 (1994)). Paul Ridker has suggested that IL-6, possibly produced by atheromas, is the main hepatic stimulus for the acute phase reactant, C-reactive protein (CRP) (Blake and Ridker, J. Intern. Med. 252, 283-294 (2002)).

The CD40 ligand and its receptor CD40 are expressed by a wide variety of inflammatory cells. Ligation of CD40 triggers the expression of a host of proinflammatory mediators such as cytokines, chemokines, ICAM-1 and VCAM-1 and matrix metalloproteinases (MMPs) (Schonbeck and Libby, Circ. Res. 89, 1092-1103 (2001)). The latter can breakdown the collagen as well as tissue factor, an important mediator of thrombosis (Mach et al., Circulation 96, 396-399 (1997)).

As atherosclerotic lesions mature, the accumulation of foam cells leads to the formation of a lipid pool, rich in prothrombotic tissue factor. Smooth muscle cells produce collagen, which contributes to the strength of the fibrous cap and shields the circulating blood from the prothrombotic lipid pool. The synthesis and breakdown of collagen in the fibrous cap is dynamically mediated by inflammatory signals (Libby, Circulation 91, 2844-2850 (1995)). PDGF and TGF-β increase the rate of collagen

production. Conversely, IFN-γ halts collagen synthesis by smooth muscle cells. Activated macrophages secrete matrix metalloproteinases, which degrade collagen and render the fibrous cap weak and prone to rupture (Libby, Circulation 104, 365-372 (2001)). Thus a dynamic balance is maintained between collagen synthesis and breakdown. If pro-inflammatory forces predominate, the fibrous cap may thin and eventually rupture, with release of the prothrombotic lipid pool into the lumen. This may herald the onset of an acute ischemic event.

## Selection of Subjects

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The methods described herein include selecting subjects for treatment on the basis that they have, or are at risk of developing, ACVD. Methods for diagnosing ACVD are well known in the art, and include evaluating the subjects for the presence of ACVD include performing a diagnostic test, e.g., determining triglyceride levels, free fatty acid levels, cholesterol levels, and/or blood pressure, e.g., diastolic blood pressure, and selecting the subject based on the results of the diagnostic test. One or more other diagnostic tests can also be used, e.g., angiogram (arteriogram); cholesterol test; chest x-ray; CT (computed tomography) scan; duplex scanning; echocardiogram; electrocardiogram (ECG or EKG); exercise stress test (cardiac stress test); intravascular ultrasound; MRI (magnetic resonance imaging) scan; PET (positron emission tomography) scan; and/or pharmacologic stress test. The presence of detectable coronary calcium is one suitable indicator of the presence of ACVD.

Risk factors for ACVD are also well known and include high cholesterol in the blood (i.e., total blood cholesterol levels of 200 mg/dL); high low-density lipoprotein (LDL) in the blood (i.e., in subjects with heart disease or diabetes, LDL levels of 100 mg/dL or higher, or in the absence of other risk factors, LDL levels of 160 mg/dL or higher); a low level of high-density lipoprotein (HDL) in the blood (i.e., HDL levels of less than 40 mg/dL); high blood pressure (i.e., 140/90 mm Hg or higher); triglyceride levels (i.e., triglyceride levels above 150 mm/dL); tobacco smoke; diabetes mellitus; obesity (i.e., BMI of 25 or above); waist circumference (e.g., above 40 inches for men and 35 inches for women); waist-to-hip ratio (e.g., above about 1.0 for men or 0.90 or women, depending on subject, e.g., health status, age and race of the subject); inactive lifestyle (i.e., less than 30-60 minutes of physical activity on most days); age (i.e., age 45 for men and age 55 for women; and family history of heart disease (e.g., parents who developed coronary heart disease before

age 55). In some embodiments, the methods include selecting a subject based on the presence of one or more of these risk factors. For example, the methods can include determining whether a subject has heart disease or diabetes, and evaluating LDL cholesterol levels, and selecting subjects who have one or both of heart disease or diabetes in addition to LDL levels of 100 mg/dL or higher. In some embodiments, the methods further include determining levels of C-reactive protein (CRP) in the subject, and if the levels are above a preselected threshold (see, e.g., Blake and Ridker, Arter. Thromb. Vasc. Biol. 22:1512 (2002)), selecting the subject.

The methods described herein can be administered in conjunction with one or more other treatments for ACVD, e.g., pharmacological treatment (e.g., anticoagulants, beta blockers; bile acid sequestrants; calcium channel blockers; ezetimibe; fibrates; glycoprotein IIb/IIIa receptor inhibitors; niacin (nicotinic acid); nitrates; platelet inhibitors; statins (HMG-CoA reductase inhibitors); and/or thrombolytics); surgeries and procedures (e.g., angioplasty; stent placement; coronary artery bypass surgery; carotid artery surgery; atherectomy); medical devices (e.g., stents; drug-eluting stents; cardiac angioplasty device; atherectomy device); lifestyle modification (e.g., control high blood pressure; control high blood cholesterol; prevent and manage diabetes mellitus; exercise; maintain a healthy weight; eat a healthy diet; quit or do not start smoking).

20 Examples

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The invention is further described in the following examples, which do not limit the scope of the invention described in the claims.

Example 1: Salicylate-Mediated Inhibition of NF-κB Decreases Risk for Atherosclerosis

To evaluate the hypothesis that NF- $\kappa B$  activation by Western diet or obesity promotes atherogenesis as well as insulin resistance, experiments were conducted to determine whether anti-inflammatory salicylate therapy is a useful route for treating the metabolic syndrome.

Established models for studying atherosclerosis were used in these experiments, specifically the LDL receptor-deficient mice (Ldlr-/-) transgenic mice, which are on a C57Bl/6 background. These mice were selected based on the magnitude of hypercholesterolemia (~500 mg/dl in Ldlr-/-) and the impression that Ldlr-/- was an inflammation-driven model.

The mice were treated orally with salicylate, which provided therapeutic levels (>4 mg/dl) in the circulation.

The mice were fed normal rodent chow (4.5% fat g/g; Ralston Purina Co.) until the start of the experiment. Mice were housed in a VAF, pathogen-free transgenic facility on a 12 hour light-dark cycle with free access to food and water. Experiments were initiated when the mice were weaned at 4 weeks of age. Study groups included standard chow, plus or minus salicylate, and atherogenic, Western diet, plus or minus salicylate. The high fat (21% g/g), high cholesterol (0.15%), cholate-free diet was commercially available (Harlan Teklad 88137). The salicylate/diet trials proceeded for eight and 24 weeks. These time points were chosen to assess the impact of salicylate-mediated NF-κB inhibition on both early foam cell lesions and later, more complex atherosclerotic lesions.

Quantitation of atherosclerosis in the mouse.

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Several methods have been described to quantitate atherosclerosis lesion development in the mouse. Classically, lesions are visualized on cross sections of the aorta beginning at the level of the aortic sinus. A second method does not section the artery, but rather opens the vessel with a ventral incision and presents the entire surface of the thoracic and abdominal aorta en face. Both methods employ image analysis software to quantitate lesion size using digitized images. Both methods have been successfully employed, see, e.g., Bjorkbacka et al., Nat. Med. 10, 416-421 (2004). The system permits lesion area to be quantified, as well as for high-resolution morphologic analysis to be investigated. Quantification of atherosclerotic lesions was done by evaluation of lesion size in the aortic arch or descending thoracic and abdominal aorta.

At the trial termination, mice were anesthetized by i.p. injection with Avertin (0.4 mg/g body wt), and the arterial system was perfusion fixed with 10% formalin. Using an Olympus SZX12 dissecting microscope, the heart and arterial tree were removed and cleaned of peripheral fat. The heart was separated from the aortic root and embedded in optimal cutting temperature compound (OCT, Tissue-Tek). The heart was sectioned to the beginning of the aortic sinus, following which every third section (5  $\mu$ m) was collected for analysis (400  $\mu$ m total), stained with oil red O, and counterstained with hematoxylin.

For quantification of lesions in the descending aorta, the aorta was dissected free of all branching vessels, and stained, en face, with Sudan IV for 15 minutes, destained with 70% ethanol for 30 seconds, and stored in water. Stained aortas were incised ventrally from the aortic root to the bifurcation of the iliac arteries, laid flat on glass slides and covered by a coverslip. Images of stained aortas were acquired with a SPOT INSIGHT QE color digital camera with a Nikon MICRO-NIKKOR 60 mm f2.8 D macro lens using SPOT V3.5.7 software. Lesion areas were selected using IP LAB SPECTRUM software that utilizes pre-defined Sudan IV staining parameters to distinguish atherosclerotic regions within the aorta. Lesion areas were calculated as a percent of the total area of the whole aorta (or defined regions of the aorta such as the aortic arch, thoracic, abdominal, ileal areas). The tissue preparation, sectioning, and staining were performed as described in (Bjorkbacka et al., Nat. Med. 10, 416-421 (2004)).

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Shown in Figs. 1A-C is an example of the effect of salicylate in inhibiting plaque (fatty streak) formation in the descending aortas of Ldlr-/- mice. There was no detectable "plaque" in animals fed normal chow(Fig. 1A), yet a significant amount of plaque formation was seen in aortas of mice fed Western diet, particularly in the arch after the aorta exits the heart (Fig. 1B, arrow). Smaller lesions were seen throughout the thoracic and abdominal aorta (Fig. 1B). Salicylate treatment markedly reduced the plaque volume in mice on Western diet (Fig. 1C).

Frozen sections of the aortic sinus were also immunostained with antibodies against macrophage (F4/80 or MAC3), smooth muscle (alpha-actin) and endothelial (CD31) markers. Images of stained sections are acquired as described above and positively stained cells will be selected by IP Lab Spectrum software; immunoreactivity will be quantified as a percentage of total lesion area.

Images were acquired using computerized image analysis software linked to a Hitachi 3 CCD color video camera (Model HV-C20) mounted on a Nikon E600 microscope. The 4X PLANFLUOR objective was used to encompass the entire aortic sinus in the video image, while the 10X, 20X and 40X, objectives can be used for higher resolution detail. Magnification was calibrated using an in-view micrometer. Mean lesion area was determined from 12 digitally captured sections per mouse using a Wacom CINTIQ tablet to outline the lesion areas and IP Lab Spectrum software v.3.6 to calculate lesion area.

Statistical analyses were performed as follows. Power calculations with a threshold of detecting at least a 20% difference in lesion size predicted that 9 mice per experimental group would be required to show a statistically significant difference (using a two-tailed test with alpha = 0.05) with a 90% probability. Twelve mice of each sex were therefore used for each group, providing a small cushion for any unexpected animal losses. Should higher losses occur, only 7 mice per group are required to give an 80% probability of detecting a 20% difference.

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Comparisons of atherosclerosis outcomes were analyzed by one-way analysis of variance (ANOVA) and Student's t-test. Data were plotted as mean lesion area  $\pm$  SEM.

Data from the whole aorta en face preparations (Figs. 1A-C) and cross sectional analyses of the aortic root (Figs. 2A-D), from larger numbers of similarly treated animals, were quantified using these methods; the results are shown in Figs. 3A-B. The overwhelming conclusion was that the anti-inflammatory drug salicylate has a significant and beneficial effect in reducing plaque volume in this animal model of atherosclerosis.

These preclinical data support the use of salicylates as new therapeutic modalities in the treatment and/or prevention of the development or progression of cardiovascular disease, e.g., atheroma formation.

Lipoprotein and lipid profiles of mouse serum.

Blood from overnight fasted mice is collected from the retro-orbital plexus, allowed to clot on ice, and centrifuged at 12000 rpm for 15 min at 4 oC. Serum is collected and concentrations of total cholesterol and triglycerides will be determined enzymatically (Sigma) as described (Eberhart et al., J. Clin. Endocrinol. Metab. 83, 836-846 (1998)). Plasma from animals in identical treatment groups is pooled and fractionated by fast pressure liquid chromatography (FPLC) on two Superose 6B columns linked in series (Id.). Cholesterol concentrations are measured on each fraction.

Measurement of pro-inflammatory molecules by microarray and QRT-PCR in arterial lesions.

In addition to the studies described above, 6 mice of each genotype (LIKK/Ldlr-/-, FIKK/Ldlr-/-, LISR/Ldlr-/-, FISR/Ldlr-/-, and Ldlr-/-) are fed a Western diet for 8 weeks and the aortic sinus will be isolated for analysis of

inflammatory gene expression (Bjorkbacka et al (Nat. Med. 10, 416-421 (2004)). Mice are anesthetized by i.p. injection with Avertin (0.4 mg/g body wt), and the arterial system is perfused with RNA Later (Ambion). Using an Olympus SZX12 dissecting microscope, the aortic arch is separated from the heart and the mid thoracic aorta and placed in Trizol reagent (Invitrogen). Total RNA is isolated and 50 ng amplified using the MessageAmp aRNA kit (Ambion). RNA is differentially labeled with Cy3 and Cy5 dyes using the Micromax ASAP RNA labeling kit (Perkin Elmer) and hybridized to spotted oligonucleotide arrays. RNA from LIKK/Ldlr-/-, FIKK/Ldlr-/-, FISR/Ldlr-/-, and Ldlr-/- arteries is compared to assess changes in aortic gene expression due to the activation or inhibition of NF-κB in fat or liver. Genes that are found to be differentially expressed in the aortic sinus by microarray analysis will be confirmed by QRT-PCR as we previously described (Cai et al., Cell 119, 285-298 (2004); Ceriello et al., Arterioscler. Thromb. Vasc. Biol. 24, 816-823 (2004)).

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# <u>Example 2: Salsalate-Mediated Reduction of Triglyceride, Free Fatty Acid and</u> Cholesterol Levels in Human Plasma

To evaluate the hypothesis that Salsalate treatment has beneficiary effects on risk factors involved in the development of ACVD, experiments were conducted to determine the effect of Salsalate administration on triglyceride and free fatty acid levels in human plasma. Baseline characteristics of the population of subjects are shown in Table 1.

TABLE 1. Subject characteristics for the open label 4.5 g/d and 3.0 g/d trials (mean  $\pm$  SD). F, female; M, male; BMI, body mass index; FBG, fasting blood glucose (mg/dlx0.0555 = mmol/l).

Salsalate dose	4.5 (g/d)	3.0 (g/d)
Glucose Tolerance	2 IGT, 5 T2D	9 T2D
Gender	5F, 2M	6F, 3M
Age (yr)	$49~\pm~9$	$51 \pm 3$
BMI (kg/m <sup>2</sup> )	$32 \pm 6$	$34 \pm 3$
FBS (mmol/l)	$6.2 \pm 2.4$	$11.1 \pm 1.3$
HbA1c (%)	$6.3 \ \pm \ 1.7$	$8.1~\pm~0.5$

The study used an open-label trial design of two weeks duration, with one dose at 4.5 g/d salsalate (1.5 g three-times daily) to match the dosage and duration used historically to improve glycosuria (Ebstein, Berliner Klinische Wochenschrift; 13:337-340 (1876); Gilgore, Diabetes, 9(5), 392-393 (1960); Hundal et al., J. Clin. Invest., 109(10), 1321-1326 (2002)) and the second at 3 g/d (1.5 g twice daily) as recommended to minimize side effects (Caraco Pharmaceuticals). Antihyperglycemic medications were discontinued one month prior to baseline. Subjects were instructed to monitor fasting blood glucose levels and with symptoms of hyperglycemia or hypoglycemia, and to avoid changing dietary or exercise habits. Subjects received misoprostol (200 µg orally four times daily), beginning three days prior and throughout the 2 wk treatment period, to compare results to those from the previously conducted high-dose aspirin study (Hundal et al., J. Clin. Invest. 109(10), 1321-1326 (2002)).

Plasma glucose was measured with a Beckman Instrument Glucose Analyzer. Immunoassays were performed in duplicate by commercial assay including RIA for insulin and C-peptide (Diagnostic Systems), adiponectin (LINCO), and ELISA for free fatty acids (WAKO), IL-6 (R&D) and sCD40L (Bender Medsystems) and nitrite (Cayman). hsCRP was analyzed by immunoturbidometry (WAKO). Glycated albumin was evaluated by Hitachi 911 lipid and protein analyzer and kits from AsahiKasei (Tokyo, JP). Deuterated glucose (6,6-[2H2]-glucose) enrichment was measured by gas chromatography-mass spectrometry analysis (Ferrannini, Metabolism, 37(3):287-301 (1988)). NF-κB DNA binding activity was assessed by ELISA using neutra-avidin plates (Pierce) pre-immobilized with 5'-biotin-labeled, double-stranded NF-κB binding oligonucleotides. Isolated peripheral blood mononuclear cells (PBMC) were lysed with Passive Lysis Buffer (Promega). DNA-bound NF-κB was detected using anti-p65 antibody (Santa Cruz sc-372), and quantified using HRP-conjugated anti-rabbit IgG.

Salsalate therapy was accompanied by decreases in fasting triglycerides (Fig. 4A), 40% ( $2.0 \pm 0.4$  vs  $1.2 \pm 0.2$  mmol/L [ $174 \pm 37$  vs  $105 \pm 20$  mg/dl], P = 0.007) at 4.5 g/d and 11% ( $1.7 \pm 0.5$  vs  $1.5 \pm 0.5$  mmol/L [ $150 \pm 47$  vs  $133 \pm 46$  mg/dl], P = 0.007) at 3.0 g/d. Other changes in fasting lipids were seen only at the higher dose, including reductions in total cholesterol by 12% ( $5.2 \pm 0.5$  vs  $4.6 \pm 0.4$  mmol/L [ $201 \pm 19$  vs  $177 \pm 14$  mg/dl], P = 0.04) (Fig. 4B) and non-esterified free

fatty acids (FFA, Fig. 4C) by 28% (0.71  $\pm$  0.05 vs 0.51  $\pm$  0.05 mM, P = 0.02) with major fatty acid subtypes reduced equivalently.

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In another study, a group of diabetic subjects participated in a double-masked, placebo-controlled parallel study design of one-month duration. At entry, participants were treated by lifestyle modification alone or in conjunction with either sulfonylurea or biguanide therapy (stably dosed for >4 wk and continued for the trial duration). After an overnight fast, subjects received a mixed meal tolerance test with blood samples collected at baseline and every 30 min for 2 hours (8 oz. BOOST<sup>TM</sup> High Protein Drink, Novartis; 240 calories; 41 g carbohydrate, 10 g protein, 4 g fat). Subjects were then randomized to receive placebo or salsalate (4.0 g/d), orally, divided twice daily. Misoprostol was not administered to subjects in this trial. Baseline characteristics of the population of subjects are shown in Table 2.

TABLE 2. Subject characteristics for the placebo-controlled trial of salsalate at maximum tolerated dose (mean  $\pm$  SD). F, female; M, male; BMI, body mass index; FBG, fasting blood glucose; Met, metformin; SFU, sulfonylurea; LS, lifestyle. Chol, cholesterol (mg/dlx0.0259 = mmol/L)

	Es, mestyle: end, en	· · ·	
	Salsalate	Placebo	P value
Age (yr)	51 ± 12	54 ± 8	0.6
Gender	3 M, 5 F	5 M, 4 F	0.5
BMI (kg/m²)	$32.5 \pm 6.4$	31.5 ± 4.9	0.7
FBS (mmol/L)	$7.5 \pm 1.0$	$7.0 \pm 1.4$	0.4
HbA1c (%)	7.1 ± 1.2	$6.7 \pm 0.5$	0.3
SBP (mm Hg)	125 ± 12	132 ± 15	0.3
DBP (mm Hg)	71 ± 4	78 ± 10	0.1
Chol (mmol/L)	$3.9 \pm 0.8$	$4.7 \pm 0.7$	0.04
Treatment	5 Met, 1 SFU, 2 LS	6 Met, 1 SFU, 2 LS	ns

In this trial salsalate was associated with a 33% reduction in fasting free fatty acid concentrations  $(0.57 \pm 0.07 \text{ vs } 0.38 \pm 0.06 \text{ mM}, P = 0.0009)$ , which was sustained following a mixed meal challenge (Fig. 5A). This is consistent with previous findings in rodents treated with high-dose sodium salicylate or aspirin (Yuan et al., Science, 293(5535):1673-1677 (2001); Cai et al., Nat. Med., 11(2):183-190 (2005)) and humans treated with high-dose aspirin (Hundal et al., J. Clin. Invest., 109(10):1321-1326 (2002). This is an important result as patients with diabetes often

have elevated circulating lipids that potentially contribute to both the pathophysiology of type 2 diabetes and its associated complications.

# Example 3: Salsalate Mediated Reduction of Blood Pressure in Human Test Cohort

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To evaluate the hypothesis that Salsalate treatment has beneficiary effects on risk factors involved in the development of ACVD, experiments were conducted to determine the Salsalate mediated changes in blood pressure in the patients participating in the trial. Enrolled patients and study design were identical to the study described in Example 2.

Although blood pressure was already well controlled in these patients, there was an 8% drop in diastolic pressure in the group receiving 3.0 g/d ( $77 \pm 4$  vs  $71 \pm 5$  mmHg, P = 0.02); the downward trends in systolic blood pressure in both groups and diastolic blood pressure at 4.5 g/d did not reach statistical significance (Table 3).

TABLE 3: Indices for fasting subjects studied in the open label trials before salsalate therapy (Pre) and after 2 wk treatment (Post) (mean  $\pm$  SEM) (HDL and LDL mg/dlx0.0259 = mmol/L), Cr, creatinine mg/dlx88.4 =  $\mu$ mol/L)

	Salsalate 4.5 (g/d)		Salsalate 3.0 (g/d)			
	Pre	Post	P value	Pre	Post	P value
Weight	88.6 ±	88.4 ±	0.8	93.7 ±	94.2 ±	0.2
(kg)	8.3	8.4		7.2	7.1	
HDL	1.2 ±	$1.1 \pm$	0.01	1.1 ±	$1.1 \pm$	0.6
(mmol/L)	0.1	0.1		0.1	0.1	
LDL	$3.2 \pm$	$3.0 \pm$	0.2	$2.7 \pm$	$2.9~\pm$	0.2
(mmol/L)	0.5	0.4		0.3	0.3	
SBP	$135 \pm 4$	$128 \pm 5$	0.2	$135 \pm 6$	$125~\pm~7$	0.1
(mm Hg)						
DBP	$75 \pm 2$	$71 \pm 4$	0.3	$77 \pm 4$	$71 \pm 5$	0.02
(mm Hg)						
AST	$25 \pm 3$	$24 \pm 4$	0.6	$20 \pm 2$	$20 \pm 2$	0.7
(U/L)						
ALT	$21 \pm 1$	$24 \pm 2$	0.2	$26 \pm 4$	$22 \pm 3$	0.01
(U/L)						
Cr	$67 \pm 5$	$77 \pm 5$	0.005	$68 \pm 6$	$72 \pm 6$	0.04
(µmol/L)						

## OTHER EMBODIMENTS

It is to be understood that while the invention has been described in conjunction with the detailed description thereof, the foregoing description is intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims. Other aspects, advantages, and modifications are within the scope of the following claims.

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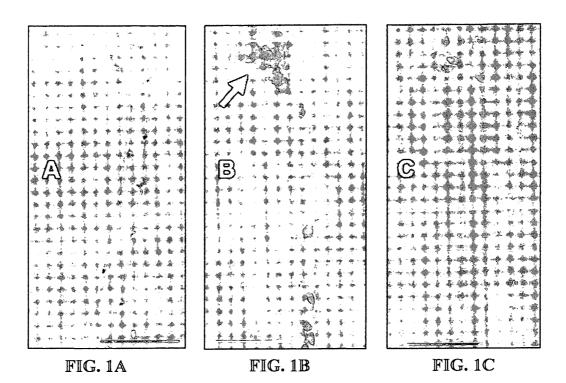
#### WHAT IS CLAIMED IS:

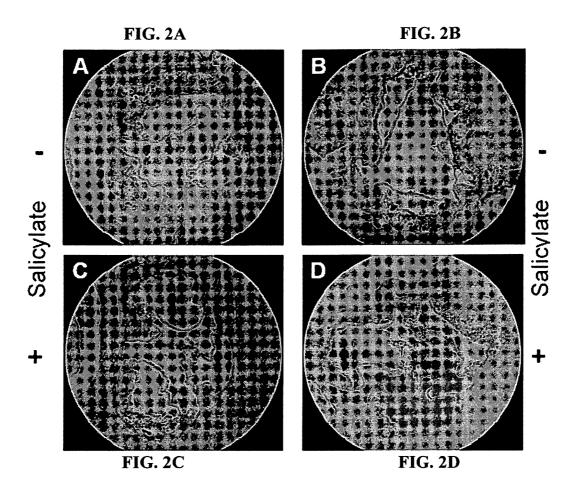
A method of treating or preventing the development or progression of
atherosclerotic cardiovascular disease (CVD) in a subject, the method comprising:
evaluating the subject for the presence of ACVD, or for the presence of one or
more risk factors for ACVD;
if the subject has or is at risk for ACVD, selecting the subject, and
administering to the subject a therapeutically effective amount of a non-acetylated
form of salicylate.

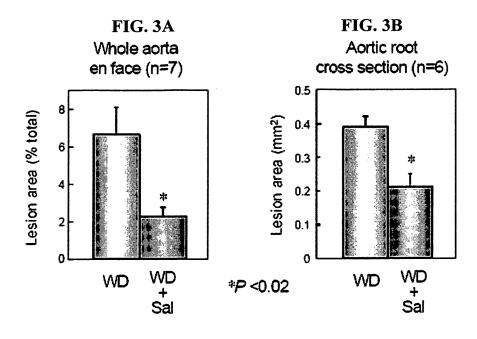
- 2. The method of claim 1, wherein if the subject has ACVD, the methods include administering about 3-4.5 g/day of the non-acetylated form of salicylate.
- 3. The method of claim 1, wherein the non-acetylated form of salicylate is salsalate or trilisate.
- 4. The method of claim 1, wherein the subject has ACVD.
- 5. The method of claim 1, wherein the subject is not suffering from pain.
- 6. The method of claim 1, wherein the subject does not have severe or substantial pain.
- 7. The method of claim 1, further comprising determining a level of triglycerides in the subject.
- 8. The method of claim 1, comprising monitoring levels of triglycerides in the subject, and administering an amount of salsalate sufficient to produce a decrease in triglyceride levels in the subject.
- 9. The method of claim 1, further comprising determining a level of free fatty acids in the subject.
- 10. The method of claim 1, further comprising monitoring levels of free fatty acids in the subject, and administering an amount of salsalate sufficient to produce a decrease in free fatty acid levels in the subject.

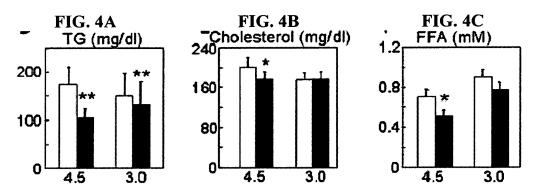
11. The method of claim 1, further comprising determining a level of cholesterol in the subject.

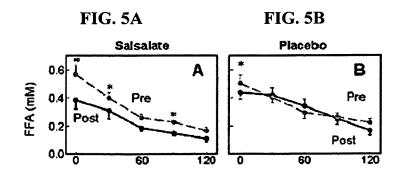
- 12. The method of claim 1, further comprising monitoring cholesterol levels in the subject, and administering an amount of salsalate sufficient to produce a decrease in cholesterol levels in the subject.
- 13. The method of claim 1, further comprising determining blood pressure in the subject.
- 14. The method of claim 1, further comprising monitoring diastolic blood pressure in the subject, and administering an amount of salsalate sufficient to produce a decrease in blood pressure in the subject.
- 15. The method of claim 14, wherein diastolic blood pressure is reduced.
- 16. The method of claim 1, wherein evaluating the subject for the presence of ACVD comprises performing one or more diagnostic tests for ACVD, and wherein the subject is selected if diagnostic test results indicate that the subject has ACVD.
- 17. The method of claim 1, wherein evaluating the subject for the presence of one or more risk factors for ACVD comprises determining one or more of total blood cholesterol levels; low-density lipoprotein (LDL) levels in the blood; high-density lipoprotein (HDL) levels in the blood; blood pressure; or triglyceride levels, and wherein the subject is selected if the results of the determination indicate that the subject has an increased risk of developing ACVD.











## INTERNATIONAL SEARCH REPORT

International application No. PCT/US08/69177

A. CLASSIFICATION OF SUBJECT MATTER IPC(8) - C12Q 1/68 (2008.04) USPC - 435/6				
	o International Patent Classification (IPC) or to both r DS SEARCHED	national classification and IPC		
	DS SEARCHED  ocumentation searched (classification system followed by	classification symbols)		
	2Q 1/68 (2008.04)	Classification by mooney		
Documentat	ion searched other than minimum documentation to the ex	xtent that such documents are included in the	fields searched	
Electronic da PATBASE	ata base consulted during the international search (name of	of data base and, where practicable, search te	rms used)	
C. DOCU	MENTS CONSIDERED TO BE RELEVANT			
Category*	Citation of document, with indication, where a	ppropriate, of the relevant passages	Relevant to claim No.	
Y	US 2007/0043120 A1 (BEYREUTHER et al) 22 Febru	ary 2007 (22.02.2007) entire document	1-17	
Y	US 5,380,747 A (MEDFORD et al) 10 January 1995 (	10.01.1995) entire document	1-17	
Y	US 6,844,149 B2 (GOLDMAN) 18 January 2005 (18.0	11.2005) entire document	7-17	
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Furthe	l er documents are listed in the continuation of Box C.		L	
	categories of cited documents:	"T" later document published after the inter		
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filing d	application or patent but published on or after the international ate ent which may throw doubts on priority claim(s) or which is	considered novel or cannot be considered	ered to involve an inventive	
cited to special	establish the publication date of another citation or other reason (as specified)	"Y" document of particular relevance; the considered to involve an inventive s	claimed invention cannot be step when the document is	
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the prior	ority date claimed actual completion of the international search	"&" document member of the same patent		
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