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(57) Abstract: Novel human polynucleotide and polypeptide sequences are disclosed that can be used in therapeutic, diagnostic, and pharmacogenomic applications.

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NOVEL HUMAN LIPOCALIN HOMOLOGS AND
POLYNUCLEOTIDES ENCODING THE SAME

The present application claims the benefit of U.S. Provisional Application Number 60/203,874 which was filed on May 12, 2000 and is herein incorporated by reference in its entirety.

1. INTRODUCTION

The present invention relates to the discovery, identification, and characterization of novel human polynucleotides encoding proteins that share sequence similarity with mammalian lipocalin and prostaglandin D synthase proteins. The invention encompasses the described polynucleotides, host cell expression systems, the encoded proteins, fusion proteins, polypeptides and peptides, antibodies to the encoded proteins and peptides, and genetically engineered animals that either lack or over express the disclosed polynucleotides, antagonists and agonists of the proteins, and other compounds that modulate the expression or activity of the proteins encoded by the disclosed polynucleotides that can be used for diagnosis, drug screening, clinical trial monitoring, the treatment of diseases and disorders, or cosmetic or nutraceutical applications.

2. BACKGROUND OF THE INVENTION

Lipocalins are members of the calycin superfamily, a structurally related family of proteins that can be involved in the transport or binding of hydrophobic molecules. Prostaglandin synthase proteins have been implicated in central nervous system (CNS) function, smooth muscle

contraction and relaxation, and inhibition of platelet aggregation.

3. SUMMARY OF THE INVENTION

The present invention relates to the discovery,
5 identification, and characterization of nucleotides that
encode novel human proteins, and the corresponding amino acid
sequences of these proteins. The novel human proteins (NHPs)
described for the first time herein share structural
similarity with mammalian lipocalins and prostaglandin D
10 synthase.

The novel human nucleic acid sequences described herein,
encode alternative proteins/open reading frames (ORFs) of
184, 68, 112, 52, 192, 76, 120, 60, 198, 82, 126, 66, 143,
and 71 amino acids in length (see respectively SEQ ID NOS: 2,
15 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, and 28).

The invention also encompasses agonists and antagonists
of the described NHPs, including small molecules, large
molecules, mutant NHPs, or portions thereof, that compete
with native NHP, peptides, and antibodies, as well as
20 nucleotide sequences that can be used to inhibit the
expression of the described NHPs (e.g., antisense and
ribozyme molecules, and gene or regulatory sequence
replacement constructs) or to enhance the expression of the
described NHP polynucleotides (e.g., expression constructs
25 that place the described polynucleotide under the control of
a strong promoter system), and transgenic animals that
express a NHP transgene, or "knock-outs" (which can be
conditional) that do not express a functional NHP. Knock-out
mice can be produced in several ways, one of which involves
30 the use of mouse embryonic stem cells ("ES cells") lines that
contain gene trap mutations in a murine homolog of at least

one of the described NHPs. When the unique NHP sequences described in SEQ ID NOS:1-28 are "knocked-out" they provide a method of identifying phenotypic expression of the particular gene as well as a method of assigning function to previously
5 unknown genes. Additionally, the unique NHP sequences described in SEQ ID NOS:1-28 are useful for the identification of coding sequence and the mapping a unique gene to a particular chromosome.

Further, the present invention also relates to processes
10 for identifying compounds that modulate, *i.e.*, act as agonists or antagonists, of NHP expression and/or NHP activity that utilize purified preparations of the described NHPs and/or NHP product, or cells expressing the same. Such compounds can be used as therapeutic agents for the treatment
15 of any of a wide variety of symptoms associated with biological disorders or imbalances.

4. DESCRIPTION OF THE SEQUENCE LISTING AND FIGURES

The Sequence Listing provides the sequences of the
20 described NHP ORFs that encode the described NHP amino acid sequences.

5. DETAILED DESCRIPTION OF THE INVENTION

The NHPs, described for the first time herein, are novel
25 proteins that are expressed in, *inter alia*, human cell lines, human colon, brain, hypothalamus, and gene trapped human cells.

The present invention encompasses the nucleotides presented in the Sequence Listing, host cells expressing such
30 nucleotides, the expression products of such nucleotides, and: (a) nucleotides that encode mammalian homologs of the described genes, including the specifically described NHPs,

and the NHP products; (b) nucleotides that encode one or more portions of the NHPs that correspond to functional domains, and the polypeptide products specified by such nucleotide sequences, including but not limited to the novel regions of
5 any active domain(s); (c) isolated nucleotides that encode mutant versions, engineered or naturally occurring, of the described NHPs in which all or a part of at least one domain is deleted or altered, and the polypeptide products specified by such nucleotide sequences, including but not limited to
10 soluble proteins and peptides in which all or a portion of the signal sequence is deleted; (d) nucleotides that encode chimeric fusion proteins containing all or a portion of a coding region of a NHP, or one of its domains (e.g., a receptor or ligand binding domain, accessory protein/self-
15 association domain, etc.) fused to another peptide or polypeptide; or (e) therapeutic or diagnostic derivatives of the described polynucleotides such as oligonucleotides, antisense polynucleotides, ribozymes, dsRNA, or gene therapy constructs comprising a sequence first disclosed in the
20 Sequence Listing.

As discussed above, the present invention includes:
(a) the human DNA sequences presented in the Sequence Listing (and vectors comprising the same) and additionally contemplates any nucleotide sequence encoding a contiguous
25 NHP open reading frame (ORF) that hybridizes to a complement of a DNA sequence presented in the Sequence Listing under highly stringent conditions, e.g., hybridization to filter-bound DNA in 0.5 M NaHPO₄, 7% sodium dodecyl sulfate (SDS), 1 mM EDTA at 65°C, and washing in 0.1xSSC/0.1% SDS at 68°C
30 (Ausubel F.M. *et al.*, eds., 1989, Current Protocols in Molecular Biology, Vol. I, Green Publishing Associates, Inc., and John Wiley & sons, Inc., New York, at p. 2.10.3) and

encodes a functionally equivalent gene product. Additionally contemplated are any nucleotide sequences that hybridize to the complement of a DNA sequence that encodes and expresses an amino acid sequence presented in the Sequence Listing
5 under moderately stringent conditions, e.g., washing in 0.2xSSC/0.1% SDS at 42°C (Ausubel et al., 1989, *supra*), yet still encodes a functionally equivalent NHP product. Functional equivalents of a NHP include naturally occurring NHPs present in other species and mutant NHPs whether
10 naturally occurring or engineered (by site directed mutagenesis, gene shuffling, directed evolution as described in, for example, U.S. Patent No. 5,837,458). The invention also includes degenerate nucleic acid variants of the disclosed NHP polynucleotide sequences.

15 Additionally contemplated are polynucleotides encoding NHP ORFs, or their functional equivalents, encoded by polynucleotide sequences that are about 99, 95, 90, or about 85 percent similar or identical to corresponding regions of the nucleotide sequences of the Sequence Listing (as measured
20 by BLAST sequence comparison analysis using, for example, the GCG sequence analysis package (Madison, Wisconsin) using standard default settings).

The invention also includes nucleic acid molecules, preferably DNA molecules, that hybridize to, and are
25 therefore the complements of, the described NHP gene nucleotide sequences. Such hybridization conditions may be highly stringent or less highly stringent, as described above. In instances where the nucleic acid molecules are deoxyoligonucleotides ("DNA oligos"), such molecules are
30 generally about 16 to about 100 bases long, or about 20 to about 80, or about 34 to about 45 bases long, or any variation or combination of sizes represented therein that

incorporate a contiguous region of sequence first disclosed in the Sequence Listing. Such oligonucleotides can be used in conjunction with the polymerase chain reaction (PCR) to screen libraries, isolate clones, and prepare cloning and
5 sequencing templates, etc.

Alternatively, such NHP oligonucleotides can be used as hybridization probes for screening libraries, and assessing gene expression patterns (particularly using a micro array or high-throughput "chip" format). Additionally, a series of
10 the described NHP oligonucleotide sequences, or the complements thereof, can be used to represent all or a portion of the described NHP sequences. An oligonucleotide or polynucleotide sequence first disclosed in at least a portion of one or more of the sequences of SEQ ID NOS: 1-28
15 can be used as a hybridization probe in conjunction with a solid support matrix/substrate (resins, beads, membranes, plastics, polymers, metal or metallized substrates, crystalline or polycrystalline substrates, etc.). Of particular note are spatially addressable arrays (*i.e.*, gene
20 chips, microtiter plates, etc.) of oligonucleotides and polynucleotides, or corresponding oligopeptides and polypeptides, wherein at least one of the biopolymers present on the spatially addressable array comprises an oligonucleotide or polynucleotide sequence first disclosed in
25 at least one of the sequences of SEQ ID NOS: 1-28, or an amino acid sequence encoded thereby. Methods for attaching biopolymers to, or synthesizing biopolymers on, solid support matrices, and conducting binding studies thereon are disclosed in, *inter alia*, U.S. Patent Nos. 5,700,637,
30 5,556,752, 5,744,305, 4,631,211, 5,445,934, 5,252,743, 4,713,326, 5,424,186, and 4,689,405 the disclosures of which are herein incorporated by reference in their entirety.

Addressable arrays comprising sequences first disclosed in SEQ ID NOS:1-28 can be used to identify and characterize the temporal and tissue specific expression of a gene. These addressable arrays incorporate oligonucleotide sequences of
5 sufficient length to confer the required specificity, yet be within the limitations of the production technology. The length of these probes is within a range of between about 8 to about 2000 nucleotides. Preferably the probes consist of 60 nucleotides and more preferably 25 nucleotides from the
10 sequences first disclosed in SEQ ID NOS:1-28.

For example, a series of the described oligonucleotide sequences, or the complements thereof, can be used in chip format to represent all or a portion of the described sequences. The oligonucleotides, typically between about 16
15 to about 40 (or any whole number within the stated range) nucleotides in length can partially overlap each other and/or the sequence may be represented using oligonucleotides that do not overlap. Accordingly, the described polynucleotide sequences shall typically comprise at least about two or
20 three distinct oligonucleotide sequences of at least about 8 nucleotides in length that are each first disclosed in the described Sequence Listing. Such oligonucleotide sequences can begin at any nucleotide present within a sequence in the Sequence Listing and proceed in either a sense (5'-to-3')
25 orientation vis-a-vis the described sequence or in an antisense orientation.

Microarray-based analysis allows the discovery of broad patterns of genetic activity, providing new understanding of gene functions and generating novel and unexpected insight
30 into transcriptional processes and biological mechanisms. The use of addressable arrays comprising sequences first disclosed in SEQ ID NOS:1-28 provides detailed information

about transcriptional changes involved in a specific pathway, potentially leading to the identification of novel components or gene functions that manifest themselves as novel phenotypes.

5 Probes consisting of sequences first disclosed in SEQ ID NOS:1-28 can also be used in the identification, selection and validation of novel molecular targets for drug discovery. The use of these unique sequences permits the direct confirmation of drug targets and recognition of drug
10 dependent changes in gene expression that are modulated through pathways distinct from the drugs intended target. These unique sequences therefore also have utility in defining and monitoring both drug action and toxicity.

As an example of utility, the sequences first disclosed
15 in SEQ ID NOS:1-28 can be utilized in microarrays or other assay formats, to screen collections of genetic material from patients who have a particular medical condition. These investigations can also be carried out using the sequences first disclosed in SEQ ID NOS:1-28 *in silico* and by comparing
20 previously collected genetic databases and the disclosed sequences using computer software known to those in the art.

Thus the sequences first disclosed in SEQ ID NOS:1-28 can be used to identify mutations associated with a particular disease and also as a diagnostic or prognostic
25 assay.

Although the presently described sequences have been specifically described using nucleotide sequence, it should be appreciated that each of the sequences can uniquely be described using any of a wide variety of additional
30 structural attributes, or combinations thereof. For example, a given sequence can be described by the net composition of the nucleotides present within a given region of the sequence

in conjunction with the presence of one or more specific oligonucleotide sequence(s) first disclosed in the SEQ ID NOS: 1-28. Alternatively, a restriction map specifying the relative positions of restriction endonuclease digestion sites, or various palindromic or other specific oligonucleotide sequences can be used to structurally describe a given sequence. Such restriction maps, which are typically generated by widely available computer programs (e.g., the University of Wisconsin GCG sequence analysis package, SEQUENCHER 3.0, Gene Codes Corp., Ann Arbor, MI, etc.), can optionally be used in conjunction with one or more discrete nucleotide sequence(s) present in the sequence that can be described by the relative position of the sequence relative to one or more additional sequence(s) or one or more restriction sites present in the disclosed sequence.

For oligonucleotide probes, highly stringent conditions may refer, e.g., to washing in 6xSSC/0.05% sodium pyrophosphate at 37°C (for 14-base oligos), 48°C (for 17-base oligos), 55°C (for 20-base oligos), and 60°C (for 23-base oligos). These nucleic acid molecules may encode or act as NHP gene antisense molecules, useful, for example, in NHP gene regulation (for and/or as antisense primers in amplification reactions of NHP gene nucleic acid sequences). With respect to NHP gene regulation, such techniques can be used to regulate biological functions. Further, such sequences may be used as part of ribozyme and/or triple helix sequences that are also useful for NHP gene regulation.

Inhibitory antisense or double stranded oligonucleotides can additionally comprise at least one modified base moiety which is selected from the group including but not limited to 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xantine, 4-acetylcytosine,

5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-
2-thiouridine, 5-carboxymethylaminomethyluracil,
dihydrouracil, beta-D-galactosylqueosine, inosine,
N6-isopentenyladenine, 1-methylguanine, 1-methylinosine,
5 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine,
3-methylcytosine, 5-methylcytosine, N6-adenine,
7-methylguanine, 5-methylaminomethyluracil,
5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine,
5'-methoxycarboxymethyluracil, 5-methoxyuracil,
10 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid
(v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine,
5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil,
5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-
5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-
15 3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine.

The antisense oligonucleotide can also comprise at least one modified sugar moiety selected from the group including but not limited to arabinose, 2-fluoroarabinose, xylulose, and hexose.

20 In yet another embodiment, the antisense oligonucleotide will comprise at least one modified phosphate backbone selected from the group consisting of a phosphorothioate, a phosphorodithioate, a phosphoramidothioate, a phosphoramidate, a phosphordiamidate, a methylphosphonate, an
25 alkyl phosphotriester, and a formacetal or analog thereof.

In yet another embodiment, the antisense oligonucleotide is an α -anomeric oligonucleotide. An α -anomeric oligonucleotide forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual β -units,
30 the strands run parallel to each other (Gautier et al., 1987, Nucl. Acids Res. 15:6625-6641). The oligonucleotide is a 2'-O-methylribonucleotide (Inoue et al., 1987, Nucl. Acids Res.

15:6131-6148), or a chimeric RNA-DNA analogue (Inoue *et al.*, 1987, FEBS Lett. 215:327-330). Alternatively, double stranded RNA can be used to disrupt the expression and function of a targeted NHP.

5 Oligonucleotides of the invention can be synthesized by standard methods known in the art, e.g. by use of an automated DNA synthesizer (such as are commercially available from Biosearch, Applied Biosystems, etc.). As examples, phosphorothioate oligonucleotides can be synthesized by the
10 method of Stein *et al.* (1988, Nucl. Acids Res. 16:3209), and methylphosphonate oligonucleotides can be prepared by use of controlled pore glass polymer supports (Sarin *et al.*, 1988, Proc. Natl. Acad. Sci. U.S.A. 85:7448-7451), etc.

Low stringency conditions are well known to those of
15 skill in the art, and will vary predictably depending on the specific organisms from which the library and the labeled sequences are derived. For guidance regarding such conditions see, for example, Sambrook *et al.*, 1989, Molecular Cloning, A Laboratory Manual (and periodic updates thereof),
20 Cold Springs Harbor Press, N.Y.; and Ausubel *et al.*, 1989, Current Protocols in Molecular Biology, Green Publishing Associates and Wiley Interscience, N.Y.

Alternatively, suitably labeled NHP nucleotide probes can be used to screen a human genomic library using
25 appropriately stringent conditions or by PCR. The identification and characterization of human genomic clones is helpful for identifying polymorphisms (including, but not limited to, nucleotide repeats, microsatellite alleles, single nucleotide polymorphisms, or coding single nucleotide
30 polymorphisms), determining the genomic structure of a given locus/allele, and designing diagnostic tests. For example,

sequences derived from regions adjacent to the intron/exon boundaries of the human gene can be used to design primers for use in amplification assays to detect mutations within the exons, introns, splice sites (e.g., splice acceptor
5 and/or donor sites), etc., that can be used in diagnostics and pharmacogenomics.

Further, a NHP gene homolog can be isolated from nucleic acid from an organism of interest by performing PCR using two degenerate or "wobble" oligonucleotide primer pools designed
10 on the basis of amino acid sequences within the NHP products disclosed herein. The template for the reaction may be total RNA, mRNA, and/or cDNA obtained by reverse transcription of mRNA prepared from human or non-human cell lines or tissue known or suspected to express an allele of a NHP gene.

15 The PCR product can be subcloned and sequenced to ensure that the amplified sequences represent the sequence of the desired NHP gene. The PCR fragment can then be used to isolate a full length cDNA clone by a variety of methods. For example, the amplified fragment can be labeled and used
20 to screen a cDNA library, such as a bacteriophage cDNA library. Alternatively, the labeled fragment can be used to isolate genomic clones via the screening of a genomic library.

PCR technology can also be used to isolate full length
25 cDNA sequences. For example, RNA can be isolated, following standard procedures, from an appropriate cellular or tissue source (i.e., one known, or suspected, to express a NHP gene). A reverse transcription (RT) reaction can be performed on the RNA using an oligonucleotide primer specific
30 for the most 5' end of the amplified fragment for the priming of first strand synthesis. The resulting RNA/DNA hybrid may then be "tailed" using a standard terminal transferase

reaction, the hybrid may be digested with RNase H, and second strand synthesis may then be primed with a complementary primer. Thus, cDNA sequences upstream of the amplified fragment can be isolated. For a review of cloning strategies
5 that can be used, see e.g., Sambrook et al., 1989, *supra*.

A cDNA encoding a mutant NHP gene can be isolated, for example, by using PCR. In this case, the first cDNA strand may be synthesized by hybridizing an oligo-dT oligonucleotide to mRNA isolated from tissue known or suspected to be
10 expressed in an individual putatively carrying a mutant NHP allele, and by extending the new strand with reverse transcriptase. The second strand of the cDNA is then synthesized using an oligonucleotide that hybridizes specifically to the 5' end of the normal gene. Using these
15 two primers, the product is then amplified via PCR, optionally cloned into a suitable vector, and subjected to DNA sequence analysis through methods well known to those of skill in the art. By comparing the DNA sequence of the mutant NHP allele to that of a corresponding normal NHP
20 allele, the mutation(s) responsible for the loss or alteration of function of the mutant NHP gene product can be ascertained.

Alternatively, a genomic library can be constructed using DNA obtained from an individual suspected of or known
25 to carry a mutant NHP allele (e.g., a person manifesting a NHP-associated phenotype such as, for example, obesity, behavioral disorders, colitis or spastic colon, high blood pressure, depression, infertility, etc.), or a cDNA library can be constructed using RNA from a tissue known, or
30 suspected, to express a mutant NHP allele. A normal NHP gene, or any suitable fragment thereof, can then be labeled and used as a probe to identify the corresponding mutant NHP

allele in such libraries. Clones containing mutant NHP gene sequences can then be purified and subjected to sequence analysis according to methods well known to those skilled in the art.

5 Additionally, an expression library can be constructed utilizing cDNA synthesized from, for example, RNA isolated from a tissue known, or suspected, to express a mutant NHP allele in an individual suspected of or known to carry such a mutant allele. In this manner, gene products made by the
10 putatively mutant tissue can be expressed and screened using standard antibody screening techniques in conjunction with antibodies raised against a normal NHP product, as described below. (For screening techniques, see, for example, Harlow, E. and Lane, eds., 1988, "Antibodies: A Laboratory Manual",
15 Cold Spring Harbor Press, Cold Spring Harbor, NY.)

 Additionally, screening can be accomplished by screening with labeled NHP fusion proteins, such as, for example, alkaline phosphatase-NHP or NHP-alkaline phosphatase fusion proteins. In cases where a NHP mutation results in an
20 expressed gene product with altered function (e.g., as a result of a missense or a frameshift mutation), polyclonal antibodies to a NHP are likely to cross-react with a corresponding mutant NHP gene product. Library clones detected via their reaction with such labeled antibodies can
25 be purified and subjected to sequence analysis according to methods well known in the art.

 The invention also encompasses (a) DNA vectors that contain any of the foregoing NHP coding sequences and/or their complements (i.e., antisense); (b) DNA expression
30 vectors that contain any of the foregoing NHP coding sequences operatively associated with a regulatory element that directs the expression of the coding sequences (for

example, baculo virus as described in U.S. Patent No. 5,869,336 herein incorporated by reference); (c) genetically engineered host cells that contain any of the foregoing NHP coding sequences operatively associated with a regulatory
5 element that directs the expression of the coding sequences in the host cell; and (d) genetically engineered host cells that express an endogenous NHP gene under the control of an exogenously introduced regulatory element (*i.e.*, gene activation). As used herein, regulatory elements include,
10 but are not limited to, inducible and non-inducible promoters, enhancers, operators and other elements known to those skilled in the art that drive and regulate expression. Such regulatory elements include but are not limited to the cytomegalovirus (hCMV) immediate early gene, regulatable,
15 viral elements (particularly retroviral LTR promoters), the early or late promoters of SV40 adenovirus, the *lac* system, the *trp* system, the *TAC* system, the *TRC* system, the major operator and promoter regions of phage lambda, the control regions of fd coat protein, the promoter for
20 3-phosphoglycerate kinase (PGK), the promoters of acid phosphatase, and the promoters of the yeast α -mating factors.

The present invention also encompasses antibodies and anti-idiotypic antibodies (including Fab fragments), antagonists and agonists of the NHP, as well as compounds or
25 nucleotide constructs that inhibit expression of a NHP gene (transcription factor inhibitors, antisense and ribozyme molecules, or gene or regulatory sequence replacement constructs), or promote the expression of a NHP (*e.g.*, expression constructs in which NHP coding sequences are
30 operatively associated with expression control elements such as promoters, promoter/enhancers, etc.).

The NHPs or NHP peptides, NHP fusion proteins, NHP nucleotide sequences, antibodies, antagonists and agonists can be useful for the detection of mutant NHPs or inappropriately expressed NHPs for the diagnosis of disease.

5 The NHP proteins or peptides, NHP fusion proteins, NHP nucleotide sequences, host cell expression systems, antibodies, antagonists, agonists and genetically engineered cells and animals can be used for screening for drugs (or high throughput screening of combinatorial libraries)

10 effective in the treatment of the symptomatic or phenotypic manifestations of perturbing the normal function of NHP in the body. The use of engineered host cells and/or animals may offer an advantage in that such systems allow not only for the identification of compounds that bind to the

15 endogenous receptor for an NHP, but can also identify compounds that trigger NHP-mediated activities or pathways.

Finally, the NHP products, or modified/processed forms thereof, can be used as therapeutics. For example, soluble derivatives such as NHP peptides/domains corresponding to the

20 NHPs, NHP fusion protein products (especially NHP-Ig fusion proteins, *i.e.*, fusions of a NHP, or a domain of a NHP, to an IgFc), NHP antibodies and anti-idiotypic antibodies (including Fab fragments), antagonists or agonists (including compounds that modulate or act on downstream targets in a

25 NHP-mediated pathway) can be used to directly treat diseases or disorders. For instance, the administration of an effective amount of soluble NHP, or a NHP-IgFc fusion protein or an anti-idiotypic antibody (or its Fab) that mimics the NHP could activate or effectively antagonize the endogenous

30 NHP receptor. Nucleotide constructs encoding such NHP products can be used to genetically engineer host cells to express such products *in vivo*; these genetically engineered

cells function as "bioreactors" in the body delivering a continuous supply of a NHP, a NHP peptide, or a NHP fusion protein to the body. Nucleotide constructs encoding functional NHPs, mutant NHPs, as well as antisense and
5 ribozyme molecules can also be used in "gene therapy" approaches for the modulation of NHP expression. Thus, the invention also encompasses pharmaceutical formulations and methods for treating biological disorders such as, for example, hyperthyroidism and/or hypothyroidism.

10 Various aspects of the invention are described in greater detail in the subsections below.

5.1 THE NHP SEQUENCES

The cDNA sequences and the corresponding deduced amino
15 acid sequences of the described NHPs are presented in the Sequence Listing. The NHP nucleotides were obtained from clustered human gene trapped sequences, and cDNA products isolated from human hypothalamus, testis, and colon mRNA. The described sequences share substantial structural
20 similarity with a variety of proteins, including, but not limited to, lipocalin proteins, prostaglandin D synthases and D isomerases, lactoglobulin, and microglobulin. Because of their medical significance, lipocalin protein homologs have been subject to considerable scientific scrutiny as evidenced
25 in U.S. Patent No. 6,020,163, herein incorporated by reference, which describes various applications, uses, and compositions in which lipocalin homologs such as the presently described NHPs can be advantageously applied.

An additional application of the described novel human
30 polynucleotide sequences is their use in the molecular mutagenesis/evolution of proteins that are at least partially encoded by the described novel sequences using, for example,

polynucleotide shuffling or related methodologies. Such approaches are described in U.S. Patents Nos. 5,830,721 and 5,837,458 which are herein incorporated by reference in their entirety.

5 NHP gene products can also be expressed in transgenic animals. Animals of any species, including, but not limited to, worms, mice, rats, rabbits, guinea pigs, pigs, micro-pigs, birds, goats, and non-human primates, e.g., baboons, monkeys, and chimpanzees may be used to generate NHP
10 transgenic animals.

Any technique known in the art may be used to introduce a NHP transgene into animals to produce the founder lines of transgenic animals. Such techniques include, but are not limited to pronuclear microinjection (Hoppe, P.C. and Wagner,
15 T.E., 1989, U.S. Pat. No. 4,873,191); retrovirus mediated gene transfer into germ lines (Van der Putten *et al.*, 1985, Proc. Natl. Acad. Sci., USA 82:6148-6152); gene targeting in embryonic stem cells (Thompson *et al.*, 1989, Cell 56:313-321); electroporation of embryos (Lo, 1983, Mol Cell. Biol.
20 3:1803-1814); and sperm-mediated gene transfer (Lavitrano *et al.*, 1989, Cell 57:717-723); etc. For a review of such techniques, see Gordon, 1989, Transgenic Animals, Intl. Rev. Cytol. 115:171-229, which is incorporated by reference herein in its entirety.

25 The present invention provides for transgenic animals that carry the NHP transgene in all their cells, as well as animals which carry the transgene in some, but not all their cells, *i.e.*, mosaic animals or somatic cell transgenic animals. The transgene may be integrated as a single
30 transgene or in concatamers, e.g., head-to-head tandems or head-to-tail tandems. The transgene may also be selectively

introduced into and activated in a particular cell type by following, for example, the teaching of Lasko et al., 1992, Proc. Natl. Acad. Sci. USA 89:6232-6236. The regulatory sequences required for such a cell-type specific activation
5 will depend upon the particular cell type of interest, and will be apparent to those of skill in the art.

When it is desired that a NHP transgene be integrated into the chromosomal site of the endogenous NHP gene, gene targeting is preferred. Briefly, when such a technique is to
10 be utilized, vectors containing some nucleotide sequences homologous to the endogenous NHP gene are designed for the purpose of integrating, via homologous recombination with chromosomal sequences, into and disrupting the function of the nucleotide sequence of the endogenous NHP gene (*i.e.*,
15 "knockout" animals).

The transgene can also be selectively introduced into a particular cell type, thus inactivating the endogenous NHP gene in only that cell type, by following, for example, the teaching of Gu et al., 1994, Science, 265:103-106. The
20 regulatory sequences required for such a cell-type specific inactivation will depend upon the particular cell type of interest, and will be apparent to those of skill in the art.

Once transgenic animals have been generated, the expression of the recombinant NHP gene may be assayed
25 utilizing standard techniques. Initial screening may be accomplished by Southern blot analysis or PCR techniques to analyze animal tissues to assay whether integration of the transgene has taken place. The level of mRNA expression of the transgene in the tissues of the transgenic animals may
30 also be assessed using techniques which include but are not limited to Northern blot analysis of tissue samples obtained from the animal, *in situ* hybridization analysis, and RT-PCR.

Samples of NHP gene-expressing tissue, may also be evaluated immunocytochemically using antibodies specific for the NHP transgene product.

5

5.2 NHPS AND NHP POLYPEPTIDES

NHPs, polypeptides, peptide fragments, mutated, truncated, or deleted forms of the NHPs, and/or NHP fusion proteins can be prepared for a variety of uses. These uses include but are not limited to, use as protein therapeutics, the generation of antibodies, as reagents in diagnostic assays, the identification of other cellular gene products related to a NHP, as reagents in assays for screening for compounds that can be used as pharmaceutical reagents useful in the therapeutic treatment of mental, biological, or medical disorders and diseases. Given the similarity information and expression data, the described NHPs can be targeted (by drugs, oligos, antibodies, etc,) in order to treat disease, or to therapeutically augment the efficacy of therapeutic agents.

The Sequence Listing discloses the amino acid sequences encoded by the described NHP sequences. The NHPs display initiator methionines in DNA sequence contexts consistent with a translation initiation site, and a signal sequence characteristic of secreted proteins. Those NHP ORFs that initiate with tandem methionines can use either methionine as the N-terminal amino acid sequence of the NHP precursor protein (that is subsequently cleaved during cleavage/removal of the signal sequence).

The NHP amino acid sequences of the invention include the amino acid sequences presented in the Sequence Listing as well as analogues and derivatives thereof. Further, corresponding NHP homologues from other species are

encompassed by the invention. In fact, any NHP protein encoded by the NHP nucleotide sequences described above are within the scope of the invention, as are any novel polynucleotide sequences encoding all or any novel portion of
5 an amino acid sequence presented in the Sequence Listing. The degenerate nature of the genetic code is well known, and, accordingly, each amino acid presented in the Sequence Listing, is generically representative of the well known nucleic acid "triplet" codon, or in many cases codons, that
10 can encode the amino acid. As such, as contemplated herein, the amino acid sequences presented in the Sequence Listing, when taken together with the genetic code (see, for example, Table 4-1 at page 109 of "Molecular Cell Biology", 1986, J. Darnell *et al.* eds., Scientific American Books, New York, NY,
15 herein incorporated by reference) are generically representative of all the various permutations and combinations of nucleic acid sequences that can encode such amino acid sequences.

The invention also encompasses proteins that are
20 functionally equivalent to the NHPs encoded by the presently described nucleotide sequences as judged by any of a number of criteria, including, but not limited to, the ability to bind and cleave a substrate of a NHP, or the ability to effect an identical or complementary downstream pathway, or a
25 change in cellular metabolism (e.g., proteolytic activity, ion flux, tyrosine phosphorylation, transport, etc.). Such functionally equivalent NHP proteins include, but are not limited to, additions or substitutions of amino acid residues within the amino acid sequence encoded by the NHP nucleotide
30 sequences described above, but which result in a silent change, thus producing a functionally equivalent gene product. Amino acid substitutions may be made on the basis

of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved. For example, nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine; polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine; positively charged (basic) amino acids include arginine, lysine, and histidine; and negatively charged (acidic) amino acids include aspartic acid and glutamic acid.

A variety of host-expression vector systems can be used to express the NHP nucleotide sequences of the invention. Where, as in the present instance, the NHP peptide or polypeptide is thought to be membrane protein, the hydrophobic regions of the protein can be excised and the resulting soluble peptide or polypeptide can be recovered from the culture media. Such expression systems also encompass engineered host cells that express a NHP, or functional equivalent, *in situ*. Purification or enrichment of a NHP from such expression systems can be accomplished using appropriate detergents and lipid micelles and methods well known to those skilled in the art. However, such engineered host cells themselves may be used in situations where it is important not only to retain the structural and functional characteristics of the NHP, but to assess biological activity, e.g., in drug screening assays.

The expression systems that can be used for purposes of the invention include but are not limited to microorganisms such as bacteria (e.g., *E. coli*, *B. subtilis*) transformed with recombinant bacteriophage DNA, plasmid DNA or cosmid DNA expression vectors containing NHP nucleotide sequences; yeast (e.g., *Saccharomyces*, *Pichia*) transformed with recombinant

yeast expression vectors containing NHP nucleotide sequences;
insect cell systems infected with recombinant virus expres-
sion vectors (e.g., baculovirus) containing NHP sequences;
plant cell systems infected with recombinant virus expression
5 vectors (e.g., cauliflower mosaic virus, CaMV; tobacco mosaic
virus, TMV) or transformed with recombinant plasmid expres-
sion vectors (e.g., Ti plasmid) containing NHP nucleotide
sequences; or mammalian cell systems (e.g., COS, CHO, BHK,
293, 3T3) harboring recombinant expression constructs
10 containing promoters derived from the genome of mammalian
cells (e.g., metallothionein promoter) or from mammalian
viruses (e.g., the adenovirus late promoter; the vaccinia
virus 7.5K promoter).

In bacterial systems, a number of expression vectors may
15 be advantageously selected depending upon the use intended
for the NHP product being expressed. For example, when a
large quantity of such a protein is to be produced for the
generation of pharmaceutical compositions of or containing
NHP, or for raising antibodies to a NHP, vectors that direct
20 the expression of high levels of fusion protein products that
are readily purified may be desirable. Such vectors include,
but are not limited, to the *E. coli* expression vector pUR278
(Ruther et al., 1983, EMBO J. 2:1791), in which a NHP coding
sequence may be ligated individually into the vector in frame
25 with the *lacZ* coding region so that a fusion protein is
produced; pIN vectors (Inouye & Inouye, 1985, Nucleic Acids
Res. 13:3101-3109; Van Heeke & Schuster, 1989, J. Biol. Chem.
264:5503-5509); and the like. pGEX vectors (Pharmacia or
American Type Culture Collection) can also be used to express
30 foreign polypeptides as fusion proteins with glutathione
S-transferase (GST). In general, such fusion proteins are

soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. The PGEX vectors are designed to include thrombin or factor Xa protease cleavage
5 sites so that the cloned target gene product can be released from the GST moiety.

In an insect system, *Autographa californica* nuclear polyhydrosis virus (AcNPV) is used as a vector to express foreign genes. The virus grows in *Spodoptera frugiperda*
10 cells. A NHP gene coding sequence may be cloned individually into non-essential regions (for example the polyhedrin gene) of the virus and placed under control of an AcNPV promoter (for example the polyhedrin promoter). Successful insertion of NHP gene coding sequence will result in inactivation of
15 the polyhedrin gene and production of non-occluded recombinant virus (*i.e.*, virus lacking the proteinaceous coat coded for by the polyhedrin gene). These recombinant viruses are then used to infect *Spodoptera frugiperda* cells in which the inserted gene is expressed (*e.g.*, see Smith *et al.*,
20 1983, J. Virol. 46:584; Smith, U.S. Patent No. 4,215,051).

In mammalian host cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, the NHP nucleotide sequence of interest may be ligated to an
25 adenovirus transcription/translation control complex, *e.g.*, the late promoter and tripartite leader sequence. This chimeric gene may then be inserted in the adenovirus genome by *in vitro* or *in vivo* recombination. Insertion in a non-essential region of the viral genome (*e.g.*, region E1 or E3)
30 will result in a recombinant virus that is viable and capable of expressing a NHP product in infected hosts (*e.g.*, See

Logan & Shenk, 1984, Proc. Natl. Acad. Sci. USA 81:3655-3659). Specific initiation signals may also be required for efficient translation of inserted NHP nucleotide sequences. These signals include the ATG initiation codon and adjacent
5 sequences. In cases where an entire NHP gene or cDNA, including its own initiation codon and adjacent sequences, is inserted into the appropriate expression vector, no additional translational control signals may be needed. However, in cases where only a portion of a NHP coding
10 sequence is inserted, exogenous translational control signals, including, perhaps, the ATG initiation codon, must be provided. Furthermore, the initiation codon must be in phase with the reading frame of the desired coding sequence to ensure translation of the entire insert. These exogenous
15 translational control signals and initiation codons can be of a variety of origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of appropriate transcription enhancer elements, transcription terminators, etc. (See Bitter et al., 1987, Methods in
20 Enzymol. 153:516-544).

In addition, a host cell strain may be chosen that modulates the expression of the inserted sequences, or modifies and processes the gene product in the specific fashion desired. Such modifications (e.g., glycosylation)
25 and processing (e.g., cleavage) of protein products may be important for the function of the protein. Different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins and gene products. Appropriate cell lines or host systems
30 can be chosen to ensure the correct modification and processing of the foreign protein expressed. To this end, eukaryotic host cells which possess the cellular machinery

for proper processing of the primary transcript, glycosylation, and phosphorylation of the gene product may be used. Such mammalian host cells include, but are not limited to, CHO, VERO, BHK, HeLa, COS, MDCK, 293, 3T3, WI38, and in particular, human cell lines.

For long-term, high-yield production of recombinant proteins, stable expression is preferred. For example, cell lines which stably express the NHP sequences described above can be engineered. Rather than using expression vectors which contain viral origins of replication, host cells can be transformed with DNA controlled by appropriate expression control elements (e.g., promoter, enhancer sequences, transcription terminators, polyadenylation sites, etc.), and a selectable marker. Following the introduction of the foreign DNA, engineered cells may be allowed to grow for 1-2 days in an enriched media, and then are switched to a selective media. The selectable marker in the recombinant plasmid confers resistance to the selection and allows cells to stably integrate the plasmid into their chromosomes and grow to form foci which in turn can be cloned and expanded into cell lines. This method may advantageously be used to engineer cell lines which express the NHP product. Such engineered cell lines may be particularly useful in screening and evaluation of compounds that affect the endogenous activity of the NHP product.

A number of selection systems may be used, including but not limited to the herpes simplex virus thymidine kinase (Wigler, et al., 1977, Cell 11:223), hypoxanthine-guanine phosphoribosyltransferase (Szybalska & Szybalski, 1962, Proc. Natl. Acad. Sci. USA 48:2026), and adenine phosphoribosyltransferase (Lowy, et al., 1980, Cell 22:817) genes can be employed in tk⁻, hgp⁻ or aprt⁻ cells,

respectively. Also, antimetabolite resistance can be used as the basis of selection for the following genes: dhfr, which confers resistance to methotrexate (Wigler, *et al.*, 1980, Natl. Acad. Sci. USA 77:3567; O'Hare, *et al.*, 1981, Proc. Natl. Acad. Sci. USA 78:1527); gpt, which confers resistance to mycophenolic acid (Mulligan & Berg, 1981, Proc. Natl. Acad. Sci. USA 78:2072); neo, which confers resistance to the aminoglycoside G-418 (Colberre-Garapin, *et al.*, 1981, J. Mol. Biol. 150:1); and hygromycin (Santerre, *et al.*, 1984, Gene 30:147).

Alternatively, any fusion protein can be readily purified by utilizing an antibody specific for the fusion protein being expressed. For example, a system described by Janknecht *et al.* allows for the ready purification of non-denatured fusion proteins expressed in human cell lines (Janknecht, *et al.*, 1991, Proc. Natl. Acad. Sci. USA 88:8972-8976). In this system, the gene of interest is subcloned into a vaccinia recombination plasmid such that the gene's open reading frame is translationally fused to an amino-terminal tag consisting of six histidine residues. Extracts from cells infected with recombinant vaccinia virus are loaded onto Ni²⁺-nitriloacetic acid-agarose columns and histidine-tagged proteins are selectively eluted with imidazole-containing buffers.

Also encompassed by the present invention are fusion proteins that direct the NHP to a target organ and/or facilitate transport across the membrane into the cytosol. Conjugation of NHPs to antibody molecules or their Fab fragments could be used to target cells bearing a particular epitope. Attaching the appropriate signal sequence to the NHP would also transport the NHP to the desired location

within the cell. Alternatively targeting of NHP or its nucleic acid sequence might be achieved using liposome or lipid complex based delivery systems. Such technologies are described in Liposomes: A Practical Approach, New, RRC ed.,
5 Oxford University Press, New York and in U.S. Patents Nos. 4,594,595, 5,459,127, 5,948,767 and 6,110,490 and their respective disclosures which are herein incorporated by reference in their entirety. Additionally embodied are novel protein constructs engineered in such a way that they
10 facilitate transport of the NHP to the target site or desired organ, where they cross the cell membrane and/or the nucleus where the NHP can exert its functional activity. This goal may be achieved by coupling of the NHP to a cytokine or other ligand that provides targeting specificity, and/or to a
15 protein transducing domain (see generally U.S. applications Ser. No. 60/111,701 and 60/056,713, both of which are herein incorporated by reference, for examples of such transducing sequences) to facilitate passage across cellular membranes and can optionally be engineered to include nuclear
20 localization sequences.

5.3 ANTIBODIES TO NHP PRODUCTS

Antibodies that specifically recognize one or more epitopes of a NHP, or epitopes of conserved variants of a
25 NHP, or peptide fragments of a NHP are also encompassed by the invention. Such antibodies include but are not limited to polyclonal antibodies, monoclonal antibodies (mAbs), humanized or chimeric antibodies, single chain antibodies, Fab fragments, F(ab')₂ fragments, fragments produced by a Fab
30 expression library, anti-idiotypic (anti-Id) antibodies, and epitope-binding fragments of any of the above.

The antibodies of the invention may be used, for example, in the detection of NHP in a biological sample and may, therefore, be utilized as part of a diagnostic or prognostic technique whereby patients may be tested for abnormal amounts of NHP. Such antibodies may also be utilized in conjunction with, for example, compound screening schemes for the evaluation of the effect of test compounds on expression and/or activity of a NHP gene product. Additionally, such antibodies can be used in conjunction gene therapy to, for example, evaluate the normal and/or engineered NHP-expressing cells prior to their introduction into the patient. Such antibodies may additionally be used as a method for the inhibition of abnormal NHP activity. Thus, such antibodies may, therefore, be utilized as part of treatment methods.

For the production of antibodies, various host animals may be immunized by injection with the NHP, an NHP peptide (e.g., one corresponding to a functional domain of an NHP), truncated NHP polypeptides (NHP in which one or more domains have been deleted), functional equivalents of the NHP or mutated variant of the NHP. Such host animals may include but are not limited to pigs, rabbits, mice, goats, and rats, to name but a few. Various adjuvants may be used to increase the immunological response, depending on the host species, including but not limited to Freund's adjuvant (complete and incomplete), mineral salts such as aluminum hydroxide or aluminum phosphate, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, and potentially useful human adjuvants such as BCG (bacille Calmette-Guerin) and *Corynebacterium parvum*. Alternatively, the immune response could be enhanced by combination and or coupling with molecules such as keyhole

limpet hemocyanin, tetanus toxoid, diphtheria toxoid, ovalbumin, cholera toxin or fragments thereof. Polyclonal antibodies are heterogeneous populations of antibody molecules derived from the sera of the immunized animals.

5 Monoclonal antibodies, which are homogeneous populations of antibodies to a particular antigen, can be obtained by any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique of
10 Kohler and Milstein, (1975, Nature 256:495-497; and U.S. Patent No. 4,376,110), the human B-cell hybridoma technique (Kosbor *et al.*, 1983, Immunology Today 4:72; Cole *et al.*, 1983, Proc. Natl. Acad. Sci. USA 80:2026-2030), and the EBV-hybridoma technique (Cole *et al.*, 1985, Monoclonal Antibodies
15 And Cancer Therapy, Alan R. Liss, Inc., pp. 77-96). Such antibodies may be of any immunoglobulin class including IgG, IgM, IgE, IgA, IgD and any subclass thereof. The hybridoma producing the mAb of this invention may be cultivated *in vitro* or *in vivo*. Production of high titers of mAbs *in vivo*
20 makes this the presently preferred method of production.

 In addition, techniques developed for the production of "chimeric antibodies" (Morrison *et al.*, 1984, Proc. Natl. Acad. Sci., 81:6851-6855; Neuberger *et al.*, 1984, Nature, 312:604-608; Takeda *et al.*, 1985, Nature, 314:452-454) by
25 splicing the genes from a mouse antibody molecule of appropriate antigen specificity together with genes from a human antibody molecule of appropriate biological activity can be used. A chimeric antibody is a molecule in which different portions are derived from different animal species,
30 such as those having a variable region derived from a murine mAb and a human immunoglobulin constant region. Such

technologies are described in U.S. Patents Nos. 6,075,181 and 5,877,397 and their respective disclosures which are herein incorporated by reference in their entirety. Also encompassed by the present invention is the use of fully
5 humanized monoclonal antibodies as described in US Patent No. 6,150,584 and respective disclosures which are herein incorporated by reference in their entirety.

Alternatively, techniques described for the production of single chain antibodies (U.S. Patent 4,946,778; Bird,
10 1988, *Science* 242:423-426; Huston *et al.*, 1988, *Proc. Natl. Acad. Sci. USA* 85:5879-5883; and Ward *et al.*, 1989, *Nature* 341:544-546) can be adapted to produce single chain antibodies against NHP gene products. Single chain antibodies are formed by linking the heavy and light chain
15 fragments of the Fv region via an amino acid bridge, resulting in a single chain polypeptide.

Antibody fragments which recognize specific epitopes may be generated by known techniques. For example, such fragments include, but are not limited to: the F(ab')₂
20 fragments which can be produced by pepsin digestion of the antibody molecule and the Fab fragments which can be generated by reducing the disulfide bridges of the F(ab')₂ fragments. Alternatively, Fab expression libraries may be constructed (Huse *et al.*, 1989, *Science*, 246:1275-1281) to
25 allow rapid and easy identification of monoclonal Fab fragments with the desired specificity.

Antibodies to a NHP can, in turn, be utilized to generate anti-idiotypic antibodies that "mimic" a given NHP, using techniques well known to those skilled in the art.
30 (See, e.g., Greenspan & Bona, 1993, *FASEB J* 7(5):437-444; and Nissinoff, 1991, *J. Immunol.* 147(8):2429-2438). For example

antibodies which bind to a NHP domain and competitively inhibit the binding of NHP to its cognate receptor can be used to generate anti-idiotypes that "mimic" the NHP and, therefore, bind and activate or neutralize a receptor. Such
5 anti-idiotypic antibodies or Fab fragments of such anti-idiotypes can be used in therapeutic regimens involving a NHP mediated pathway.

The present invention is not to be limited in scope by the specific embodiments described herein, which are intended
10 as single illustrations of individual aspects of the invention, and functionally equivalent methods and components are within the scope of the invention.. Indeed, various modifications of the invention, in addition to those shown and described herein will become apparent to those skilled in
15 the art from the foregoing description. Such modifications are intended to fall within the scope of the appended claims. All cited publications, patents, and patent applications are herein incorporated by reference in their entirety.

WHAT IS CLAIMED IS:

1. An isolated nucleic acid molecule comprising
at a nucleotide sequence encoding an amino acid sequence
5 drawn from the group consisting of SEQ ID NOS: 2, 4, 6, 8,
10, 12, 14, 16, 18, 20, 22, 24, 26, and 28.

2. An isolated nucleic acid molecule comprising
a nucleotide sequence that:
10 (a) encodes the amino acid sequence shown in SEQ
ID NO: 2; and
(b) hybridizes under stringent conditions to the
nucleotide sequence of SEQ ID NO: 1 or the
complement thereof.

15 3. An isolated nucleic acid molecule comprising
a nucleotide sequence that encodes the amino acid sequence
shown in SEQ ID NO: 2.

20 4. An isolated nucleic acid molecule comprising
a nucleotide sequence that encodes the amino acid sequence
shown in SEQ ID NO:18.

SEQUENCE LISTING

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 Val His Met Glu Phe Pro Gly Ala Asp Gly Cys Asn Gln Val Asp Ala
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 Glu Tyr Leu Lys Val Gly Ser Glu Gly His Phe Arg Val Pro Ala Leu
 100 105 110
 Gly Tyr Leu Asp Val Arg Ile Val Asp Thr Asp Tyr Ser Ser Phe Ala
 115 120 125
 Val Leu Tyr Ile Tyr Lys Glu Leu Glu Gly Ala Leu Ser Thr Met Val
 130 135 140
 Gln Leu Tyr Ser Arg Thr Gln Asp Val Ser Pro Gln Ala Leu Lys Ala
 145 150 155 160
 Phe Gln Asp Phe Tyr Pro Thr Leu Gly Leu Pro Glu Asp Met Met Val
 165 170 175
 Met Leu Pro Gln Ser Asp Ala Cys Asn Pro Glu Ser Lys Glu Ala Pro
 180 185 190

<210> 11
 <211> 231
 <212> DNA
 <213> homo sapiens

<400> 11
 atgggctgga gggcagggga ggggatgatg tcattcctgc tcggcgcaat cctgaccctg 60
 ctctgggcgc ccacggctca ggctgaggtt ctgctgcagc ctgacttcaa tgctgaaaag 120
 ggtcttcctg ggcaagaagg accacctgtc catgtccacc agggccatca ggcccacaga 180
 ggagggcggc ctccacgtcc acatggagtt cccgggggcg gacggctgta a 231

<210> 12
 <211> 76
 <212> PRT
 <213> homo sapiens

<400> 12
 Met Gly Trp Arg Ala Gly Glu Gly Met Met Ser Phe Leu Leu Gly Ala
 1 5 10 15
 Ile Leu Thr Leu Leu Trp Ala Pro Thr Ala Gln Ala Glu Val Leu Leu
 20 25 30
 Gln Pro Asp Phe Asn Ala Glu Lys Gly Leu Pro Gly Gln Glu Gly Pro
 35 40 45
 Pro Val His Val His Gln Gly His Gln Ala His Arg Gly Gly Arg Pro
 50 55 60

Pro Arg Pro His Gly Val Pro Gly Gly Gly Arg Leu
65 70 75

<210> 13
<211> 363
<212> DNA
<213> homo sapiens

<400> 13
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ctctgggagc ccacggctca ggctgaggtt ctgctgcagc ctgacttcaa tgctgaaaag 120
ttctcaggcc tctggtagct ggtctccatg gcctctgact gcagggctct cctgggcaag 180
aaggaccacc tgtccatgtc caccagggcc atcaggccca cagaggaggg cggcctccac 240
gtccacatgg agttcccggg ggcggacggc tgtaaccagg tggatgccga gtacctggag 300
tctctccatc ctccaccccc cgcctgtggg atgccttgtg ggacgtctct ttctattcaa 360
taa 363

<210> 14
<211> 120
<212> PRT
<213> homo sapiens

<400> 14
Met Gly Trp Arg Ala Gly Glu Gly Met Met Ser Phe Leu Leu Gly Ala
1 5 10 15
Ile Leu Thr Leu Leu Trp Ala Pro Thr Ala Gln Ala Glu Val Leu Leu
20 25 30
Gln Pro Asp Phe Asn Ala Glu Lys Phe Ser Gly Leu Trp Tyr Val Val
35 40 45
Ser Met Ala Ser Asp Cys Arg Val Phe Leu Gly Lys Lys Asp His Leu
50 55 60
Ser Met Ser Thr Arg Ala Ile Arg Pro Thr Glu Glu Gly Gly Leu His
65 70 75 80
Val His Met Glu Phe Pro Gly Ala Asp Gly Cys Asn Gln Val Asp Ala
85 90 95
Glu Tyr Leu Glu Ser Leu His Pro Pro Pro Pro Ala Cys Gly Met Pro
100 105 110
Cys Gly Thr Ser Leu Ser Ile Gln
115 120

<210> 15
<211> 183
<212> DNA
<213> homo sapiens

<400> 15
atgggctgga gggcagggga ggggatgatg tcattcctgc tcggcgcaat cctgaccctg 60
ctctgggagc ccacggctca ggctgaggtt ctgctgcagc ctgacttcaa tgctgaaaag 120
gtaccagggg cctctgctgt cctgtggttg gtgggagctg ggccccctgcc agagacaacg 180
tga 183

<210> 16
<211> 60
<212> PRT
<213> homo sapiens

<400> 16

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Met Gly Trp Arg Ala Gly Glu Gly Met Met Ser Phe Leu Leu Gly Ala
 1           5           10           15
Ile Leu Thr Leu Leu Trp Ala Pro Thr Ala Gln Ala Glu Val Leu Leu
          20           25           30
Gln Pro Asp Phe Asn Ala Glu Lys Val Pro Gly Ala Ser Ala Val Leu
          35           40           45
Trp Trp Val Gly Ala Gly Pro Leu Pro Glu Thr Thr
          50           55           60

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<210> 17

<211> 597

<212> DNA

<213> homo sapiens

<400> 17

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atgggctcag ctcacaccca agagaggagg gcaggggagg ggatgatgtc attcctgctc      60
ggcgcaatcc tgaccctgct ctgggcgccc acggctcagg ctgaggttct gctgcagcct      120
gacttcaatg ctgaaaagt ctcaggcctc tggtagctgg tctccatggc atctgactgc      180
agggctcttc tgggcaagaa ggaccacctg tccatgtcca ccagggccat caggcccaca      240
gaggagggcg gcctccacgt ccacatggag ttcccggggg cggacggctg taaccaggtg      300
gatgccgagt acctgaaggt gggctccgag ggacacttca gagtcccggc cttgggctac      360
ctggacgtgc gcatcggtga cacagactac agctccttcg ccgtccttta catctacaag      420
gagctggagg gggcgctcag caccatggtg cagctctaca gccggacca ggatgtgagt      480
ccccaggctc tgaaggcctt ccaggacttc taccgcaccc tggggctccc cgaggacatg      540
atggctcatgc tgcccagtc agatgcatgc aaccctgaga gcaaggaggc gccctga      597

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<210> 18

<211> 198

<212> PRT

<213> homo sapiens

<400> 18

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Met Gly Ser Ala His Thr Gln Glu Arg Arg Ala Gly Glu Gly Met Met
 1           5           10           15
Ser Phe Leu Leu Gly Ala Ile Leu Thr Leu Leu Trp Ala Pro Thr Ala
          20           25           30
Gln Ala Glu Val Leu Leu Gln Pro Asp Phe Asn Ala Glu Lys Phe Ser
          35           40           45
Gly Leu Trp Tyr Val Val Ser Met Ala Ser Asp Cys Arg Val Phe Leu
          50           55           60
Gly Lys Lys Asp His Leu Ser Met Ser Thr Arg Ala Ile Arg Pro Thr
        65           70           75           80
Glu Glu Gly Gly Leu His Val His Met Glu Phe Pro Gly Ala Asp Gly
          85           90           95
Cys Asn Gln Val Asp Ala Glu Tyr Leu Lys Val Gly Ser Glu Gly His
          100          105          110
Phe Arg Val Pro Ala Leu Gly Tyr Leu Asp Val Arg Ile Val Asp Thr
          115          120          125
Asp Tyr Ser Ser Phe Ala Val Leu Tyr Ile Tyr Lys Glu Leu Glu Gly
          130          135          140
Ala Leu Ser Thr Met Val Gln Leu Tyr Ser Arg Thr Gln Asp Val Ser
        145          150          155          160
Pro Gln Ala Leu Lys Ala Phe Gln Asp Phe Tyr Pro Thr Leu Gly Leu
          165          170          175
Pro Glu Asp Met Met Val Met Leu Pro Gln Ser Asp Ala Cys Asn Pro

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180
Glu Ser Lys Glu Ala Pro
195

185

190

<210> 19
<211> 249
<212> DNA
<213> homo sapiens

<400> 19
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ggcgcaatcc tgaccctgct ctgggcgccc acggctcagg ctgaggttct gctgcagcct 120
gacttcaatg ctgaaaaggg tcttcctggg caagaaggac cacctgtcca tgtccaccag 180
ggccatcagg cccacagagg agggcggcct ccacgtccac atggagttcc cgggggcgga 240
cggtgtaa 249

<210> 20
<211> 82
<212> PRT
<213> homo sapiens

<400> 20
Met Gly Ser Ala His Thr Gln Glu Arg Arg Ala Gly Glu Gly Met Met
1 5 10 15
Ser Phe Leu Leu Gly Ala Ile Leu Thr Leu Leu Trp Ala Pro Thr Ala
20 25 30
Gln Ala Glu Val Leu Leu Gln Pro Asp Phe Asn Ala Glu Lys Gly Leu
35 40 45
Pro Gly Gln Glu Gly Pro Pro Val His Val His Gln Gly His Gln Ala
50 55 60
His Arg Gly Gly Arg Pro Pro Arg Pro His Gly Val Pro Gly Gly Gly
65 70 75 80
Arg Leu

<210> 21
<211> 381
<212> DNA
<213> homo sapiens

<400> 21
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ggcgcaatcc tgaccctgct ctgggcgccc acggctcagg ctgaggttct gctgcagcct 120
gacttcaatg ctgaaaagtt ctcaggcctc tggtagctgg tctccatggc atctgactgc 180
agggtcttcc tgggcaagaa ggaccacctg tccatgtcca ccagggccat caggccaca 240
gaggagggcg gcctccacgt ccacatggag ttcccggggg cggacggctg taaccagggtg 300
gatgccgagt acctggagtc tctccatcct ccaccccccg cctgtgggat gccttggtggg 360
acgtctcttt ctattcaata a 381

<210> 22
<211> 126
<212> PRT
<213> homo sapiens

<400> 22
Met Gly Ser Ala His Thr Gln Glu Arg Arg Ala Gly Glu Gly Met Met

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1           5           10           15
Ser Phe Leu Leu Gly Ala Ile Leu Thr Leu Leu Trp Ala Pro Thr Ala
                20           25           30
Gln Ala Glu Val Leu Leu Gln Pro Asp Phe Asn Ala Glu Lys Phe Ser
                35           40           45
Gly Leu Trp Tyr Val Val Ser Met Ala Ser Asp Cys Arg Val Phe Leu
                50           55           60
Gly Lys Lys Asp His Leu Ser Met Ser Thr Arg Ala Ile Arg Pro Thr
65           70           75           80
Glu Glu Gly Gly Leu His Val His Met Glu Phe Pro Gly Ala Asp Gly
                85           90           95
Cys Asn Gln Val Asp Ala Glu Tyr Leu Glu Ser Leu His Pro Pro Pro
                100           105           110
Pro Ala Cys Gly Met Pro Cys Gly Thr Ser Leu Ser Ile Gln
                115           120           125

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<210> 23
 <211> 201
 <212> DNA
 <213> homo sapiens

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<400> 23
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ggcgcaatcc tgaccctgct ctgggcgccc acggctcagg ctgaggttct gctgcagcct      120
gacttcaatg ctgaaaaggt accaggggcc tctgctgtcc tgtggtgggt gggagctggg      180
ccctgcccag agacaacgtg a                                     201

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<210> 24
 <211> 66
 <212> PRT
 <213> homo sapiens

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<400> 24
Met Gly Ser Ala His Thr Gln Glu Arg Arg Ala Gly Glu Gly Met Met
1           5           10           15
Ser Phe Leu Leu Gly Ala Ile Leu Thr Leu Leu Trp Ala Pro Thr Ala
                20           25           30
Gln Ala Glu Val Leu Leu Gln Pro Asp Phe Asn Ala Glu Lys Val Pro
                35           40           45
Gly Ala Ser Ala Val Leu Trp Trp Val Gly Ala Gly Pro Leu Pro Glu
                50           55           60
Thr Thr
65

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<210> 25
 <211> 432
 <212> DNA
 <213> homo sapiens

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<400> 25
atggcatctg actgcagggt cttcctgggc aagaaggacc acctgtccat gtccaccagg      60
gccatcaggc ccacagagga gggcggcctc cacgtccaca tggagttccc gggggcggac      120
ggctgtaacc aggtggatgc cgagtacctg aaggtgggct ccgagggaca cttcagagtc      180
ccggccttgg gctacctgga cgtgcgcata gtggacacag actacagctc cttcgccgct      240
ctttacatct acaaggagct ggagggggcg ctcagcacca tgggtgcagct ctacagccgg      300
accaggatg tgagtcccca ggctctgaag gccttcagg atttctaccc gaccctgggg      360

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ctccccgagg acatgatggg catgctgccc cagtcagatg catgcaaccc tgagagcaag 420
gaggcgccct ga 432

<210> 26
<211> 143
<212> PRT
<213> homo sapiens

<400> 26
Met Ala Ser Asp Cys Arg Val Phe Leu Gly Lys Lys Asp His Leu Ser
1 5 10 15
Met Ser Thr Arg Ala Ile Arg Pro Thr Glu Glu Gly Gly Leu His Val
20 25 30
His Met Glu Phe Pro Gly Ala Asp Gly Cys Asn Gln Val Asp Ala Glu
35 40 45
Tyr Leu Lys Val Gly Ser Glu Gly His Phe Arg Val Pro Ala Leu Gly
50 55 60
Tyr Leu Asp Val Arg Ile Val Asp Thr Asp Tyr Ser Ser Phe Ala Val
65 70 75 80
Leu Tyr Ile Tyr Lys Glu Leu Glu Gly Ala Leu Ser Thr Met Val Gln
85 90 95
Leu Tyr Ser Arg Thr Gln Asp Val Ser Pro Gln Ala Leu Lys Ala Phe
100 105 110
Gln Asp Phe Tyr Pro Thr Leu Gly Leu Pro Glu Asp Met Met Val Met
115 120 125
Leu Pro Gln Ser Asp Ala Cys Asn Pro Glu Ser Lys Glu Ala Pro
130 135 140

<210> 27
<211> 216
<212> DNA
<213> homo sapiens

<400> 27
atggcatctg actgcagggt cttcctgggc aagaaggacc acctgtccat gtccaccagg 60
gccatcaggc ccacagagga gggcggcctc caggtccaca tggagtccc gggggcggac 120
ggctgtaacc aggtggatgc cgagtacctg gagtctctcc atcctccacc cccgcctgt 180
gggatgcctt gtgggacgtc tctttctatt caataa 216

<210> 28
<211> 71
<212> PRT
<213> homo sapiens

<400> 28
Met Ala Ser Asp Cys Arg Val Phe Leu Gly Lys Lys Asp His Leu Ser
1 5 10 15
Met Ser Thr Arg Ala Ile Arg Pro Thr Glu Glu Gly Gly Leu His Val
20 25 30
His Met Glu Phe Pro Gly Ala Asp Gly Cys Asn Gln Val Asp Ala Glu
35 40 45
Tyr Leu Glu Ser Leu His Pro Pro Pro Pro Ala Cys Gly Met Pro Cys
50 55 60
Gly Thr Ser Leu Ser Ile Gln
65 70