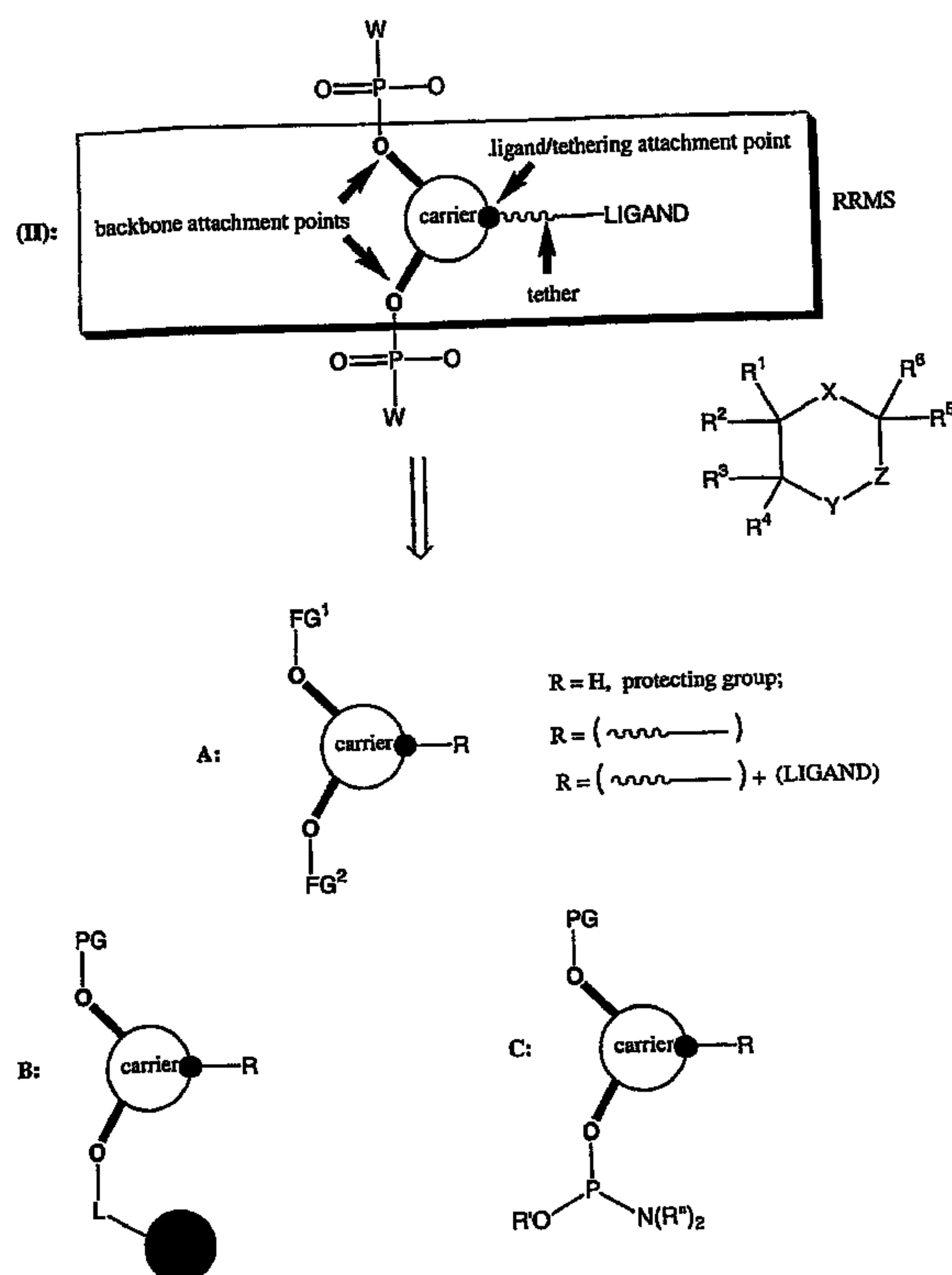




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(54) Title: CHEMICALLY MODIFIED OLIGONUCLEOTIDES



(57) **Abrégé/Abstract:**

This invention relates composition and methods for making and using chemically modified oligonucleotides agents for inhibiting gene expression.



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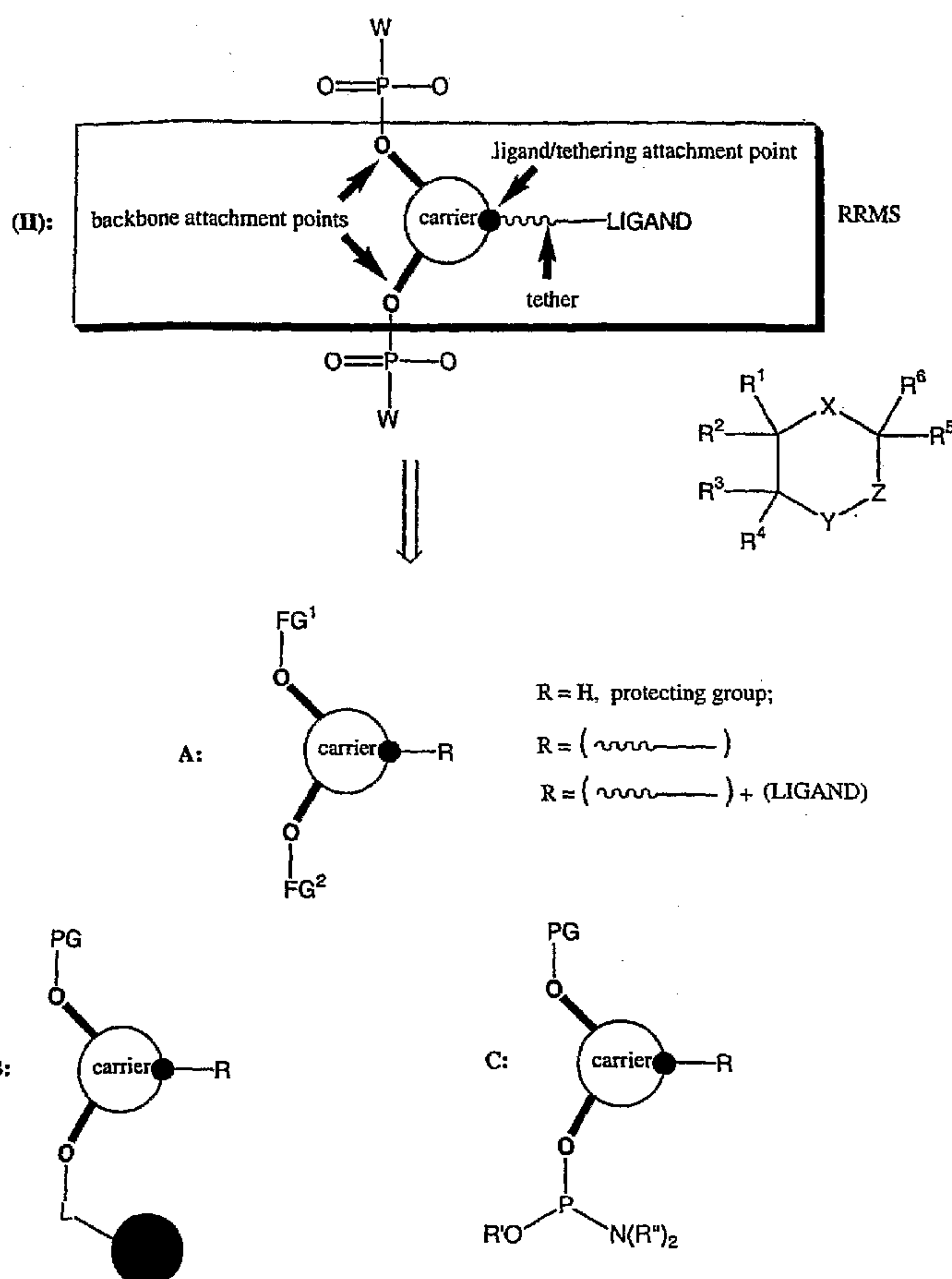
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[Continued on next page]

(54) Title: CHEMICALLY MODIFIED OLIGONUCLEOTIDES



(57) Abstract: This invention relates composition and methods for making and using chemically modified oligonucleotides agents for inhibiting gene expression.

WO 2006/020768 A2

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**Declarations under Rule 4.17:**

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CHEMICALLY MODIFIED OLIGONUCLEOTIDES

RELATED APPLICATIONS

This application claims the benefit of U.S. Provisional Application No. 60/600,703, filed August 10, 2004, which is incorporated herein by reference in its entirety.

5

TECHNICAL FIELD

This invention relates generally to chemically modified oligonucleotides useful for modulating gene expression. More particularly, the invention relates to single stranded chemically modified oligonucleotides for inhibiting gene expression and to methods of making
10 and using the modified oligonucleotides.

BACKGROUND

A variety of nucleic acid species are capable of modifying gene expression. These include antisense RNA, siRNA, microRNA, RNA and DNA aptamers, and decoy RNAs. Each of these nucleic acid species can inhibit gene expression by a different mechanism.

15

SUMMARY

The invention features methods and compositions for modulating, either inhibiting or up-regulating, gene expression by using a conjugate of an oligonucleotide agent (a term which is defined below) and a ligand. The compositions include conjugated oligonucleotide agents as well as conjugated monomers that are the components of or can be used to make the conjugated
20 oligonucleotide agents. The conjugated oligonucleotide agents can modify gene expression by targeting and binding to a nucleic acid, *e.g.*, a pre-mRNA, an mRNA, a microRNA (miRNA), a mi-RNA precursor (pre-miRNA), or DNA, or to a protein. An oligonucleotide agent featured in the invention can be, *e.g.*, an miRNA, antisense RNA, decoy RNA, DNA, or aptamer.

Thus, the invention features an oligonucleotide agent coupled to a ligand. The ligand can
25 be attached to the oligonucleotide agent through a monomer, *e.g.*, a chemically modified monomer that is integrated into the oligonucleotide agent. In a preferred embodiment, the coupling is by a tether or a linker (or both) as described herein, and the complex has the formula represented by:

Ligand- [linker]_{optional} -[tether]_{optional} – oligonucleotide agent

While, in most cases, embodiments are described with respect to an oligonucleotide agent
5 including a number of nucleotides, the invention includes monomeric subunits having the structure:

Ligand- [linker]_{optional} -[tether]_{optional} – monomer

10 Methods of making and incorporating the monomers into the oligonucleotide agents and methods of using of those agents are included in the invention.

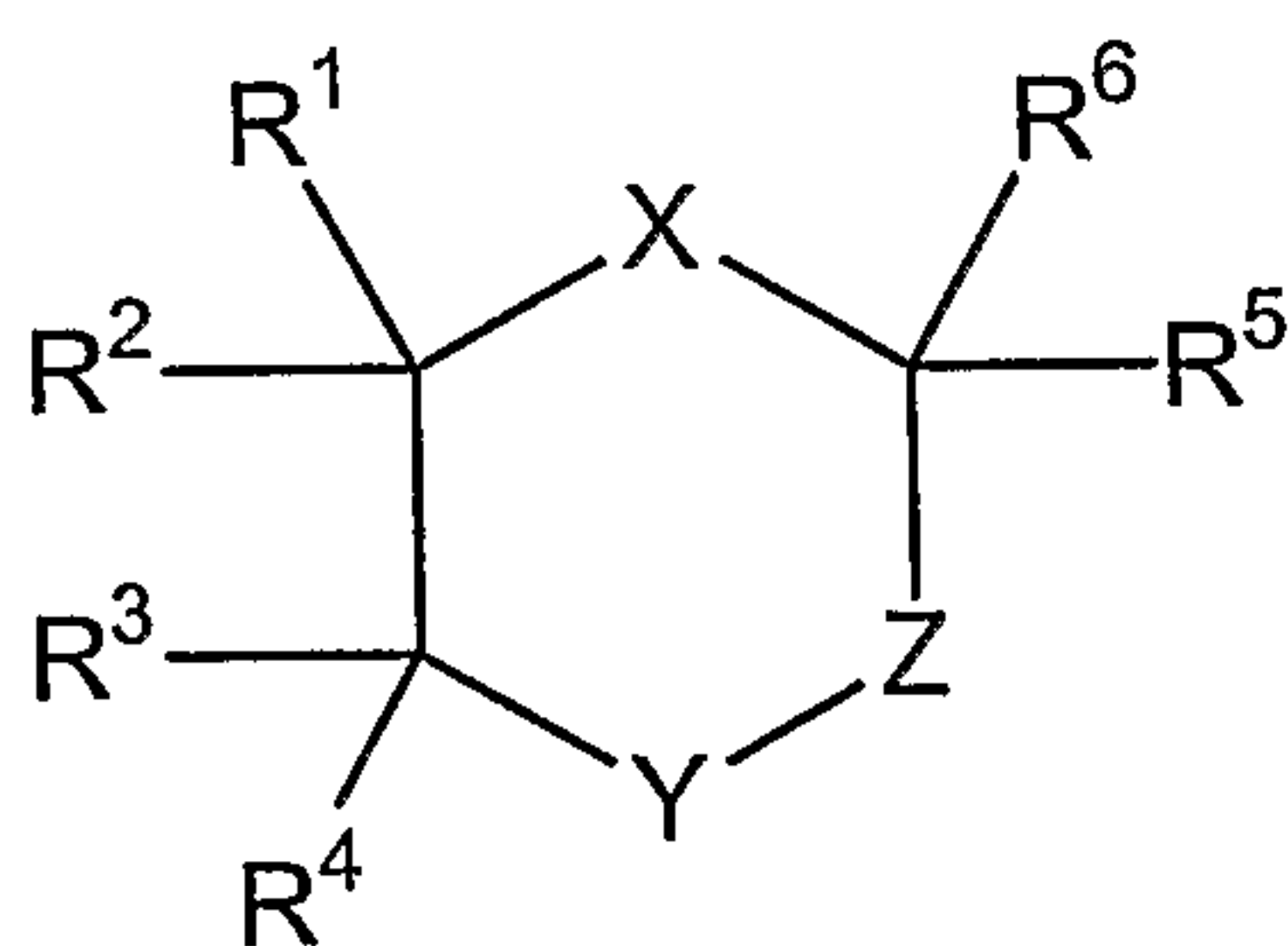
In preferred embodiments, the sugar, *e.g.*, the ribose sugar of one or more of the nucleotides, (*e.g.*, ribonucleotide, deoxynucleotide, or modified nucleotide) subunits of an oligonucleotide agent can be replaced with another moiety, *e.g.*, a non-carbohydrate (preferably
15 cyclic) carrier. A nucleotide subunit in which the sugar of the subunit has been so replaced is referred to herein as a sugar replacement modification subunit (SRMS). This is often referred to herein as a “tether.” A cyclic carrier may be a carbocyclic ring system, *i.e.*, all ring atoms are carbon atoms or a heterocyclic ring system, *i.e.*, one or more ring atoms may be a heteroatom, *e.g.*, nitrogen, oxygen, or sulfur. The cyclic carrier may be a monocyclic ring system, or may
20 contain two or more rings, *e.g.* fused rings. The cyclic carrier may be a fully saturated ring system, or it may contain one or more double bonds.

The carriers further include (i) at least two “backbone attachment points” and (ii) at least one “tethering attachment point.” A “backbone attachment point” as used herein refers to a functional group, *e.g.* a hydroxyl group, or generally, a bond available for, and that is suitable for
25 incorporation of the carrier into the backbone, *e.g.*, the phosphate, or modified phosphate, *e.g.*, sulfur containing, backbone, of a ribonucleic acid. A “tethering attachment point” as used herein refers to a constituent ring atom of the cyclic carrier, *e.g.*, a carbon atom or a heteroatom (distinct from an atom which provides a backbone attachment point), that connects a selected moiety. The moiety can be, *e.g.*, a ligand, *e.g.*, a targeting or delivery moiety, or a moiety which alters a
30 physical property. One of the most preferred moieties is a moiety which promotes entry into a cell, *e.g.*, a lipophilic moiety, *e.g.*, cholesterol. While not wishing to be bound by theory it is

believed the attachment of a lipophilic agent increases the lipophilicity of an oligonucleotide agent. Optionally, the selected moiety is connected by an intervening tether to the cyclic carrier. Thus, it will often include a functional group, *e.g.*, an amino group, or generally, provide a bond, that is suitable for incorporation or tethering of another chemical entity, *e.g.*, a ligand to the
 5 constituent ring.

Incorporation of one or more SRMSs described herein into an oligonucleotide agent, particularly when tethered to an appropriate entity, can confer one or more new properties to the oligonucleotide agent and/or alter, enhance or modulate one or more existing properties in the oligonucleotide agent. *E.g.*, it can alter one or more of lipophilicity or nuclease resistance.
 10 Incorporation of one or more SRMSs described herein into an oligonucleotide agent can, particularly when the SRMS is tethered to an appropriate entity, modulate, *e.g.*, increase, binding affinity of an oligonucleotide agent to a target RNA, *e.g.*, a pre-mRNA, mRNA, or miRNA of the subject or a pathogen of the subject. Incorporation of one or more SRMSs can alter distribution, target the oligonucleotide agent to a particular part of the body, modify the
 15 interaction with nucleic acid binding proteins (*e.g.*, during RISC formation and strand separation), or increase sequence specificity, *e.g.*, to inhibit off-site targeting.

Accordingly, in one aspect, the invention features, an oligonucleotide agent preferably comprising at least one subunit having the structure of formula (I) :



(I)

wherein:

X is N(CO)R⁷, NR⁷ or CH₂;

Y is NR⁸, O, S, CR⁹R¹⁰, or absent;

Z is CR¹¹R¹² or absent;

Each of R^1, R^2, R^3, R^4, R^9 , and R^{10} is, independently, H, OR^a , OR^b , $(CH_2)_nOR^a$, or $(CH_2)_nOR^b$, provided that at least one of R^1, R^2, R^3, R^4, R^9 , and R^{10} is OR^a or OR^b and that at least one of R^1, R^2, R^3, R^4, R^9 , and R^{10} is $(CH_2)_nOR^a$, or $(CH_2)_nOR^b$ (when the SRMS is terminal, one of R^1, R^2, R^3, R^4, R^9 , and R^{10} will include R^a and one will include R^b ; when the SRMSS is internal, two of R^1, R^2, R^3, R^4, R^9 , and R^{10} will each include an R^b); further provided that preferably OR^a may only be present with $(CH_2)_nOR^b$ and $(CH_2)_nOR^a$ may only be present with OR^b ;

Each of R^5, R^6, R^{11} , and R^{12} is, independently, H, C_1 - C_6 alkyl optionally substituted with 1-3 R^{13} , or $C(O)NHR^7$; or R^5 and R^{11} together are C_3 - C_8 cycloalkyl optionally substituted with R^{14} ;

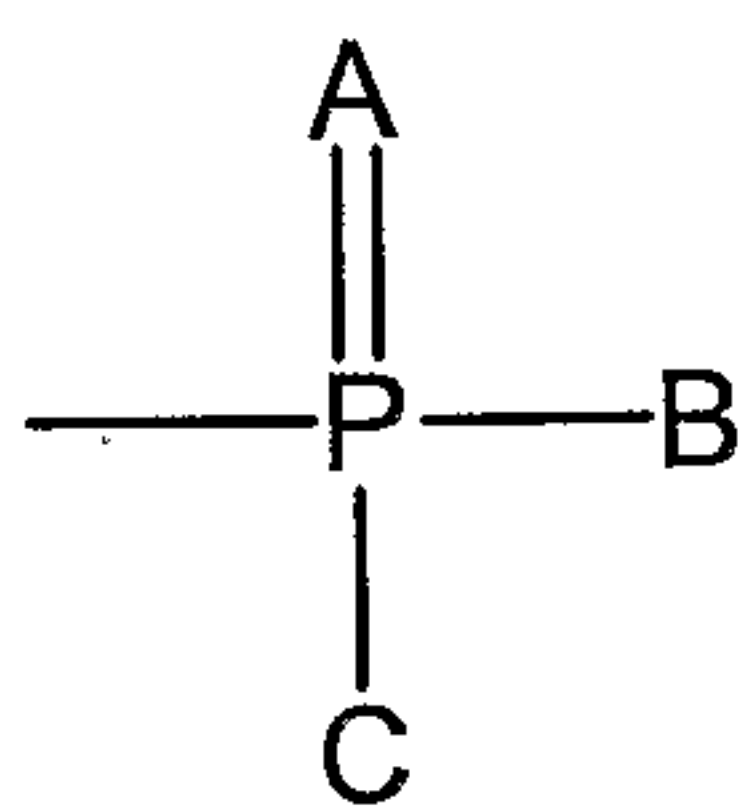
R^7 can be a ligand, *e.g.*, R^7 can be R^d , or R^7 can be a ligand tethered indirectly to the carrier, *e.g.*, through a tethering moiety, *e.g.*, C_1 - C_{20} alkyl substituted with NR^cR^d ; or C_1 - C_{20} alkyl substituted with $NHC(O)R^d$;

R^8 is C_1 - C_6 alkyl;

R^{13} is hydroxy, C_1 - C_4 alkoxy, or halo;

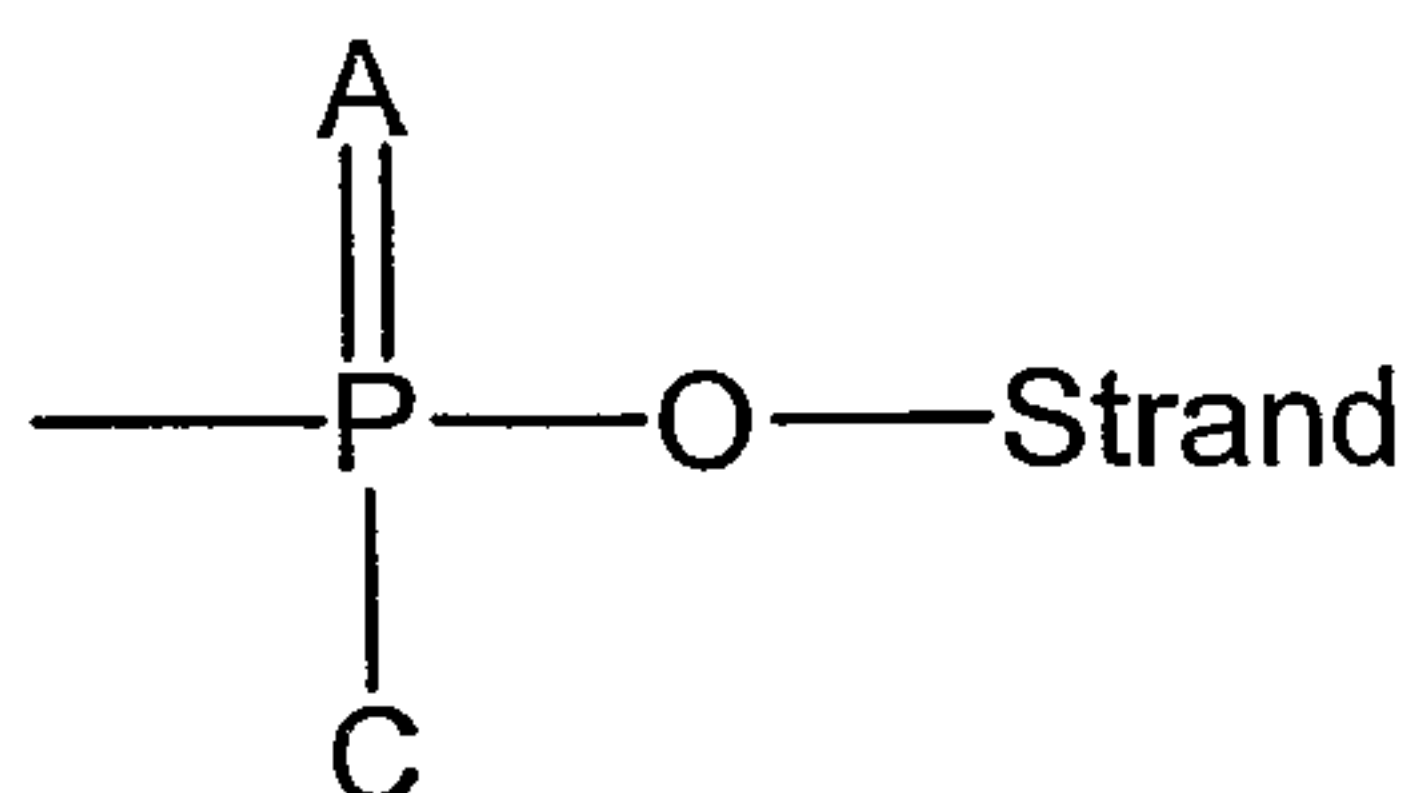
R^{14} is NR^cR^7 ;

R^a is:



;

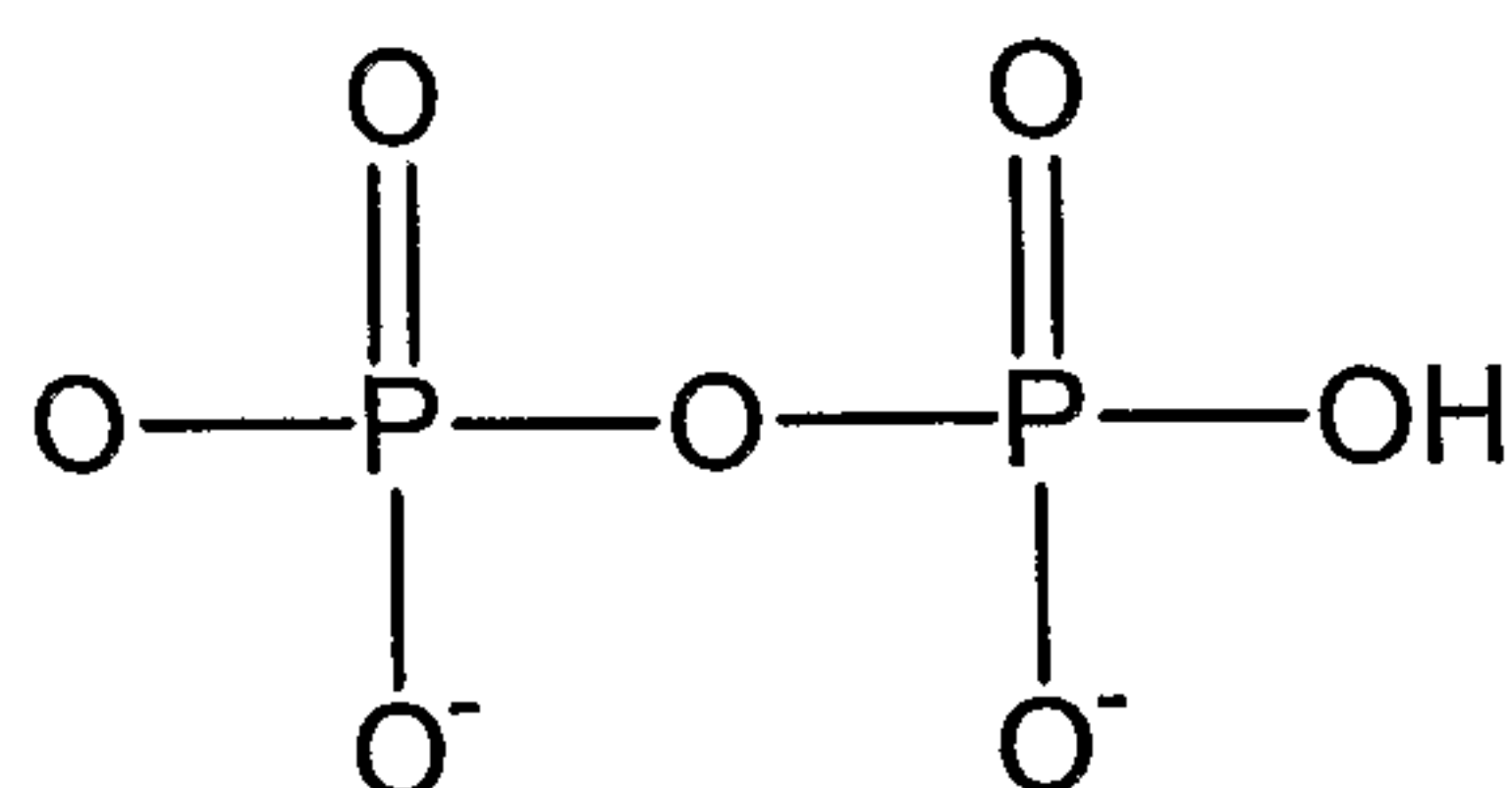
R^b is:



;

Each of A and C is, independently, O or S;

B is OH, O⁻, or



;

5 R^c is H or C₁-C₆ alkyl;

R^d is H or a ligand, *e.g.*, a lipophilic ligand, *e.g.*, cholesterol; and

n is 1-4.

Embodiments can include one or more of the following features:

10

R¹ can be CH₂OR^a and R³ can be OR^b; or R¹ can be CH₂OR^a and R⁹ can be OR^b; or R¹ can be CH₂OR^a and R² can be OR^b.

15 R¹ can be CH₂OR^b and R³ can be OR^b; or R¹ can be CH₂OR^b and R⁹ can be OR^b; or R¹ can be CH₂OR^b and R² can be OR^b; or R¹ can be CH₂OR^b and R³ can be OR^a; or R¹ can be CH₂OR^b and R⁹ can be OR^a; or R¹ can be CH₂OR^b and R² can be OR^a.

R¹ can be OR^a and R³ can be CH₂OR^b; or R¹ can be OR^a and R⁹ can be CH₂OR^b; or R¹ can be OR^a and R² can be CH₂OR^b.

20 R¹ can be OR^b and R³ can be CH₂OR^b; or R¹ can be OR^b and R⁹ can be CH₂OR^b; or R¹ can be OR^b and R² can be CH₂OR^b; or R¹ can be OR^b and R³ can be CH₂OR^a; or R¹ can be OR^b and R⁹ can be CH₂OR^a; or R¹ can be OR^b and R² can be CH₂OR^a.

R³ can be CH₂OR^a and R⁹ can be OR^b; or R³ can be CH₂OR^a and R⁴ can be OR^b.

R³ can be CH₂OR^b and R⁹ can be OR^b; or R³ can be CH₂OR^b and R⁴ can be OR^b; or R³ can be CH₂OR^b and R⁹ can be OR^a; or R³ can be CH₂OR^b and R⁴ can be OR^a.

25 R³ can be OR^b and R⁹ can be CH₂OR^a; or R³ can be OR^b and R⁴ can be CH₂OR^a; or R³ can be OR^b and R⁹ can be CH₂OR^b; or R³ can be OR^b and R⁴ can be CH₂OR^b.

R³ can be OR^a and R⁹ can be CH₂OR^b; or R³ can be OR^a and R⁴ can be CH₂OR^b.

R⁹ can be CH₂OR^a and R¹⁰ can be OR^b.

R^9 can be CH_2OR^b and R^{10} can be OR^b ; or R^9 can be CH_2OR^b and R^{10} can be OR^a .

In a preferred embodiment the ribose is replaced with a pyrroline scaffold or with a 4-hydroxyproline-derived scaffold, and X is $N(CO)R^7$ or NR^7 , Y is CR^9R^{10} , and Z is absent.

R^1 and R^3 can be *cis* or R^1 and R^3 can be *trans*.

5 n can be 1.

A can be O or S.

R^1 can be $(CH_2)_nOR^b$ and R^3 can be OR^b ; or R^1 can be $(CH_2)_nOR^a$ and R^3 can be OR^b .

R^7 can be $(CH_2)_5NHR^d$ or $(CH_2)_5NHR^d$. R^d can be chosen from a folic acid radical; a cholesterol radical; a carbohydrate radical; a vitamin A radical; a vitamin E radical; a vitamin K radical. Preferably, R^d is a cholesterol radical.

R^1 can be OR^b and R^3 can be $(CH_2)_nOR^b$; or R^1 can be OR^b and R^3 can be $(CH_2)_nOR^a$; or R^1 can be OR^a and R^3 can be $(CH_2)_nOR^b$; or R^1 can be $(CH_2)_nOR^b$ and R^9 can be OR^a .

R^1 and R^9 can be *cis* or R^1 and R^9 can be *trans*.

R^1 can be OR^a and R^9 can be $(CH_2)_nOR^b$; or R^1 can be $(CH_2)_nOR^b$ and R^9 can be OR^b ; or R^1 can be $(CH_2)_nOR^a$ and R^9 can be OR^b ; or R^1 can be OR^b and R^9 can be $(CH_2)_nOR^b$; or R^1 can be OR^b and R^9 can be $(CH_2)_nOR^a$.

R^3 can be $(CH_2)_nOR^b$ and R^9 can be OR^a ; or R^3 can be $(CH_2)_nOR^b$ and R^9 can be OR^b ; or R^3 can be $(CH_2)_nOR^a$ and R^9 can be OR^b ; or R^3 can be OR^a and R^9 can be $(CH_2)_nOR^b$; R^3 can be OR^b and R^9 can be $(CH_2)_nOR^b$; or R^3 can be OR^b and R^9 can be $(CH_2)_nOR^a$.

20 R^3 and R^9 can be *cis* or R^3 and R^9 can be *trans*.

In other preferred embodiments the ribose is replaced with a piperidine scaffold, and X is $N(CO)R^7$ or NR^7 , Y is CR^9R^{10} , and Z is $CR^{11}R^{12}$.

R^9 can be $(CH_2)_nOR^b$ and R^{10} can be OR^a .

n can be 1 or 2.

25 R^9 can be $(CH_2)_nOR^b$ and R^{10} can be OR^b ; or R^9 can be $(CH_2)_nOR^a$ and R^{10} can be OR^b .

A can be O or S.

R^7 can be $(CH_2)_5NHR^d$ or $(CH_2)_5NHR^d$. R^d can be selected from a folic acid radical; a cholesterol radical; a carbohydrate radical; a vitamin A radical; a vitamin E radical; a vitamin K radical. Preferably, R^d is a cholesterol radical.

30 R^3 can be $(CH_2)_nOR^b$ and R^4 can be OR^a ; or R^3 can be $(CH_2)_nOR^b$ and R^4 can be OR^b ; or R^3 can be $(CH_2)_nOR^a$ and R^4 can be OR^b .

R^1 can be $(CH_2)_nOR^b$ and R^2 can be OR^a ; or R^1 can be $(CH_2)_nOR^b$ and R^2 can be OR^b ; or R^1 can be $(CH_2)_nOR^a$ and R^2 can be OR^b .

R^3 can be $(CH_2)_nOR^b$ and R^9 can be OR^a .

R^3 and R^9 can be *cis*, or R^3 and R^9 can be *trans*.

5 R^3 can be $(CH_2)_nOR^b$ and R^9 can be OR^b ; or R^3 can be $(CH_2)_nOR^b$ and R^9 can be OR^a ; or R^3 can be $(CH_2)_nOR^a$ and R^9 can be OR^b .

R^1 can be $(CH_2)_nOR^b$ and R^3 can be OR^a .

R^1 and R^3 can be *cis*, or R^1 and R^3 can be *trans*.

R^3 can be OR^a and R^9 can be $(CH_2)_nOR^b$.

10 R^1 can be OR^a and R^3 can be $(CH_2)_nOR^b$.

In other preferred embodiments the ribose is replaced with a piperazine scaffold, and X is $N(CO)R^7$ or NR^7 , Y is NR^8 , and Z is $CR^{11}R^{12}$.

R^1 can be $(CH_2)_nOR^b$ and R^3 can be OR^a .

R^1 and R^3 can be *cis* or R^1 and R^3 can be *trans*.

15 n can be 1.

R^1 can be $(CH_2)_nOR^b$ and R^3 can be OR^b ; or R^1 can be $(CH_2)_nOR^a$ and R^3 can be OR^b .

A can be O or S, preferably S.

R^7 can be $(CH_2)_5NHR^d$ or $(CH_2)_5NHR^d$. R^d can be chosen from the group of a folic acid radical; a cholesterol radical; a carbohydrate radical; a vitamin A radical; a vitamin E radical; a
20 vitamin K radical. Preferably, R^d is a cholesterol radical.

R^8 can be CH_3 .

R^1 can be OR^a and R^3 can be $(CH_2)_nOR^b$.

In other preferred embodiments the ribose is replaced with a morpholino scaffold, and X is $N(CO)R^7$ or NR^7 , Y is O, and Z is $CR^{11}R^{12}$.

25 R^1 can be $(CH_2)_nOR^b$ and R^3 can be OR^a .

R^1 and R^3 can be *cis*, or R^1 and R^3 can be *trans*.

n can be 1.

R^1 can be $(CH_2)_nOR^b$ and R^3 can be OR^b ; or R^1 can be $(CH_2)_nOR^a$ and R^3 can be OR^b .

A can be O or S.

R^7 can be $(CH_2)_5NHR^d$ or $(CH_2)_5NHR^d$. R^d can be chosen from the group of a folic acid radical; a cholesterol radical; a carbohydrate radical; a vitamin A radical; a vitamin E radical; a vitamin K radical. Preferably, R^d is a cholesterol radical.

R^8 can be CH_3 .

5 R^1 can be OR^a and R^3 can be $(CH_2)_nOR^b$.

In other preferred embodiments the ribose is replaced with a decalin scaffold, and X is CH_2 ; Y is CR^9R^{10} ; and Z is $CR^{11}R^{12}$; and R^5 and R^{11} together are C^6 cycloalkyl.

R^6 can be $C(O)NHR^7$.

R^{12} can be hydrogen.

10 R^6 and R^{12} can be *trans*.

R^3 can be OR^a and R^9 can be $(CH_2)_nOR^b$.

R^3 and R^9 can be *cis*, or R^3 and R^9 can be *trans*.

n can be 1 or 2.

R^3 can be OR^b and R^9 can be $(CH_2)_nOR^b$; or R^3 can be OR^b and R^9 can be $(CH_2)_nOR^a$.

15 A can be O or S.

R^7 can be $(CH_2)_5NHR^d$ or $(CH_2)_5NHR^d$. R^d can be chosen from the group of a folic acid radical; a cholesterol radical; a carbohydrate radical; a vitamin A radical; a vitamin E radical; a vitamin K radical. Preferably, R^d is a cholesterol radical.

20 In other preferred embodiments the ribose is replaced with a decalin/indane scaffold, e.g., X is CH_2 ; Y is CR^9R^{10} ; and Z is $CR^{11}R^{12}$; and R^5 and R^{11} together are C^5 cycloalkyl.

R^6 can be CH_3 .

R^{12} can be hydrogen.

R^6 and R^{12} can be *trans*.

R^3 can be OR^a and R^9 can be $(CH_2)_nOR^b$.

25 R^3 and R^9 can be *cis*, or R^3 and R^9 can be *trans*.

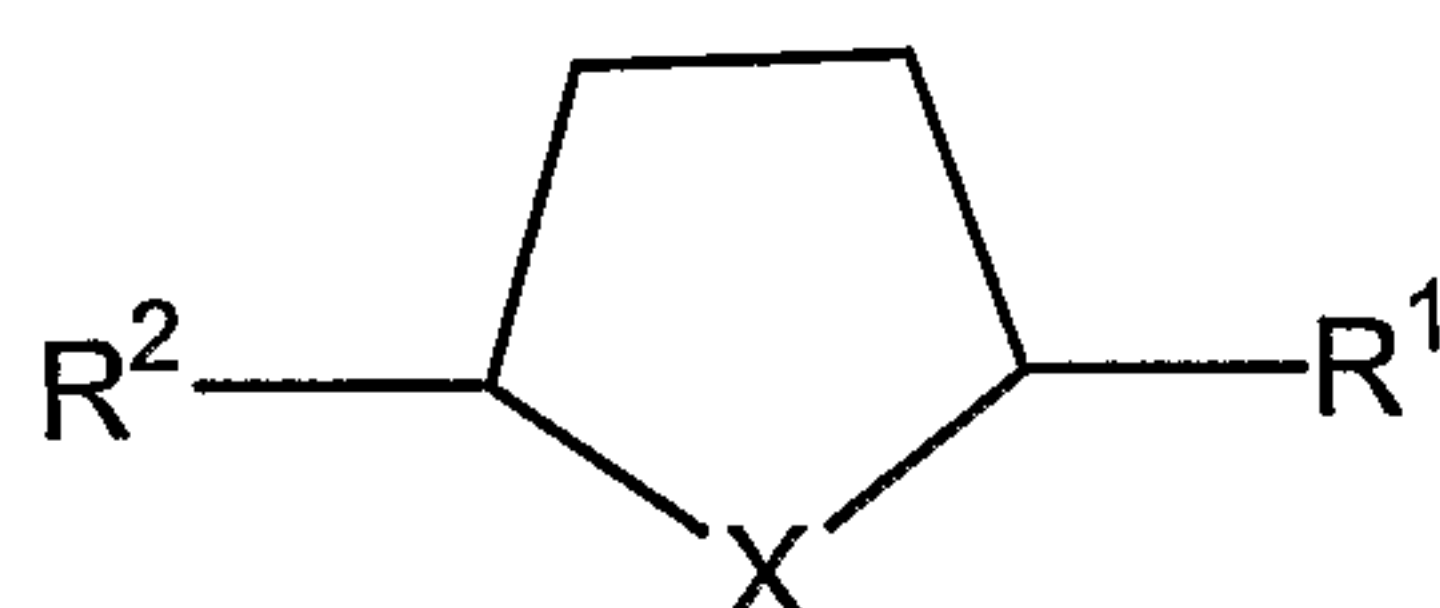
n can be 1 or 2.

R^3 can be OR^b and R^9 can be $(CH_2)_nOR^a$; or R^3 can be OR^b and R^9 can be $(CH_2)_nOR^a$.

A can be O or S.

30 R^{14} can be $N(CH_3)R^7$. R^7 can be $(CH_2)_5NHR^d$ or $(CH_2)_5NHR^d$. R^d can be chosen from the group of a folic acid radical; a cholesterol radical; a carbohydrate radical; a vitamin A radical; a vitamin E radical; a vitamin K radical. Preferably, R^d is a cholesterol radical.

In another aspect, this invention features an oligonucleotide agent comprising at least one subunit having a structure of formula (II):



(II)

X is $\text{N}(\text{CO})\text{R}^7$ or NR^7 ;

Each of R^1 and R^2 is, independently, OR^a , OR^b , $(\text{CH}_2)_n\text{OR}^a$, or $(\text{CH}_2)_n\text{OR}^b$, provided that one of R^1 and R^2 is OR^a or OR^b and the other is $(\text{CH}_2)_n\text{OR}^a$ or $(\text{CH}_2)_n\text{OR}^b$ (when the SRMS is terminal, one of R^1 or R^2 will include R^a and one will include R^b ; when the SRMSS is internal, both R^1 and R^2 will each include an R^b); further provided that preferably OR^a may only be present with $(\text{CH}_2)_n\text{OR}^b$ and $(\text{CH}_2)_n\text{OR}^a$ may only be present with OR^b ;

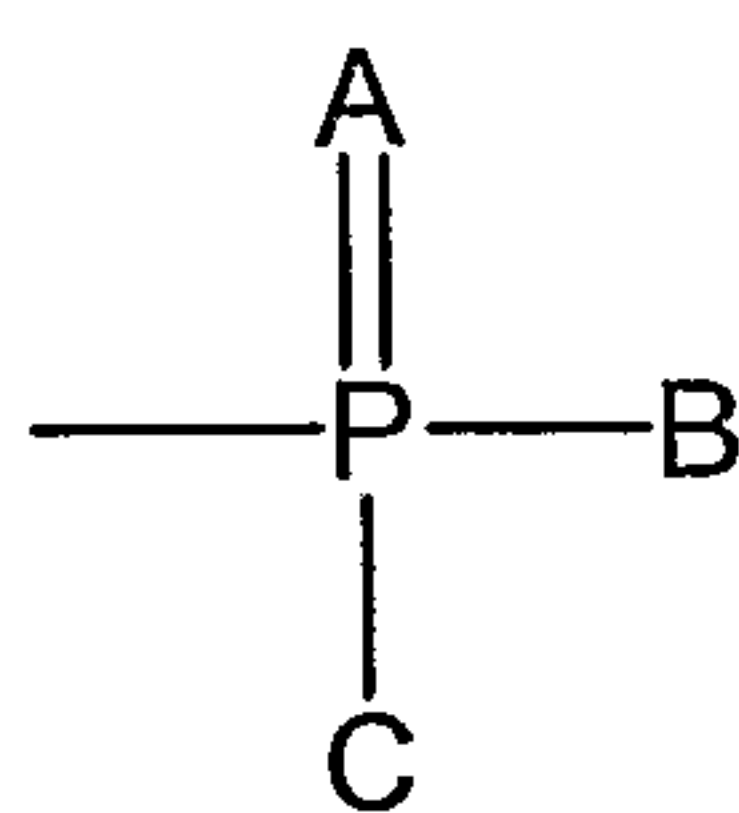
R^7 is C_1 - C_{20} alkyl substituted with NR^cR^d ;

R^8 is C_1 - C_6 alkyl;

R^{13} is hydroxy, C_1 - C_4 alkoxy, or halo;

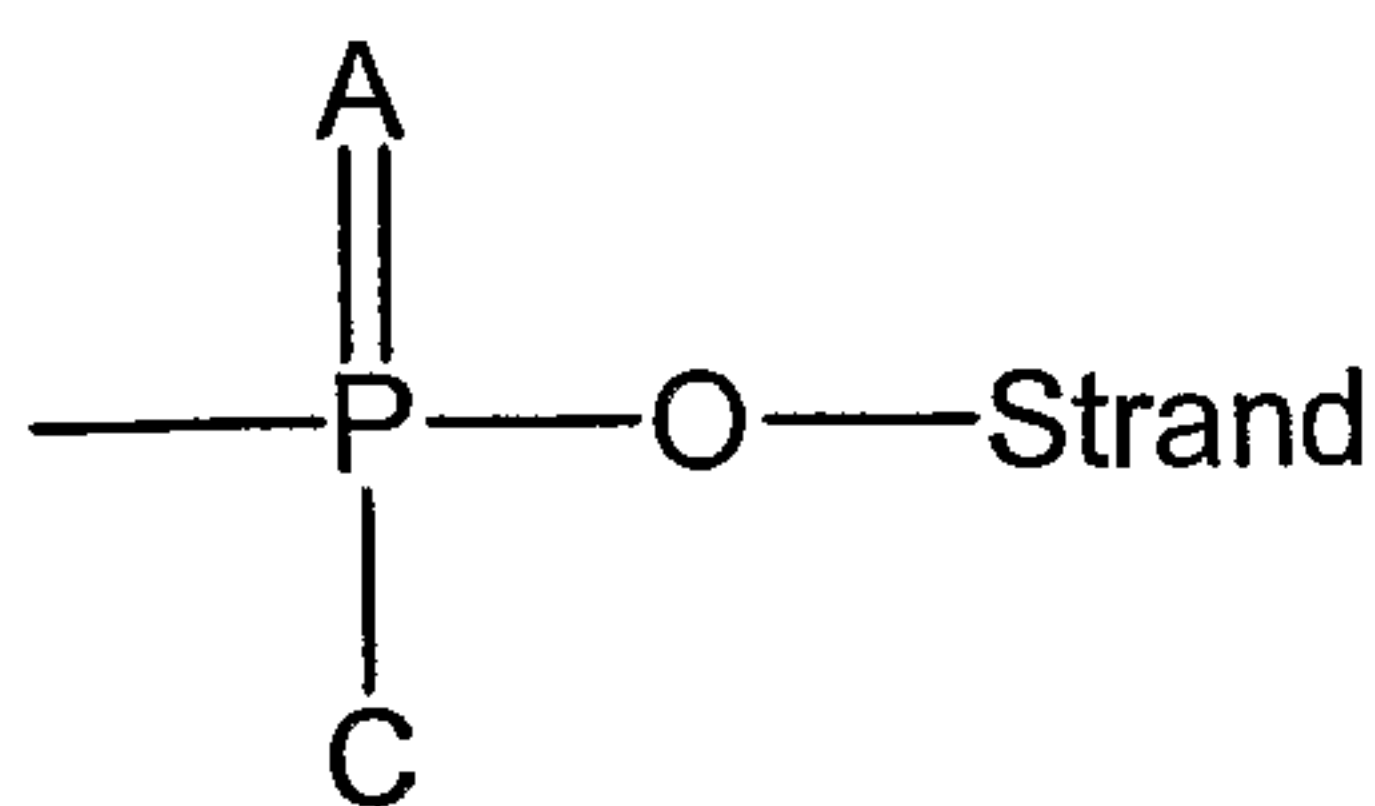
R^{14} is NR^cR^7 ;

R^a is:



;

R^b is

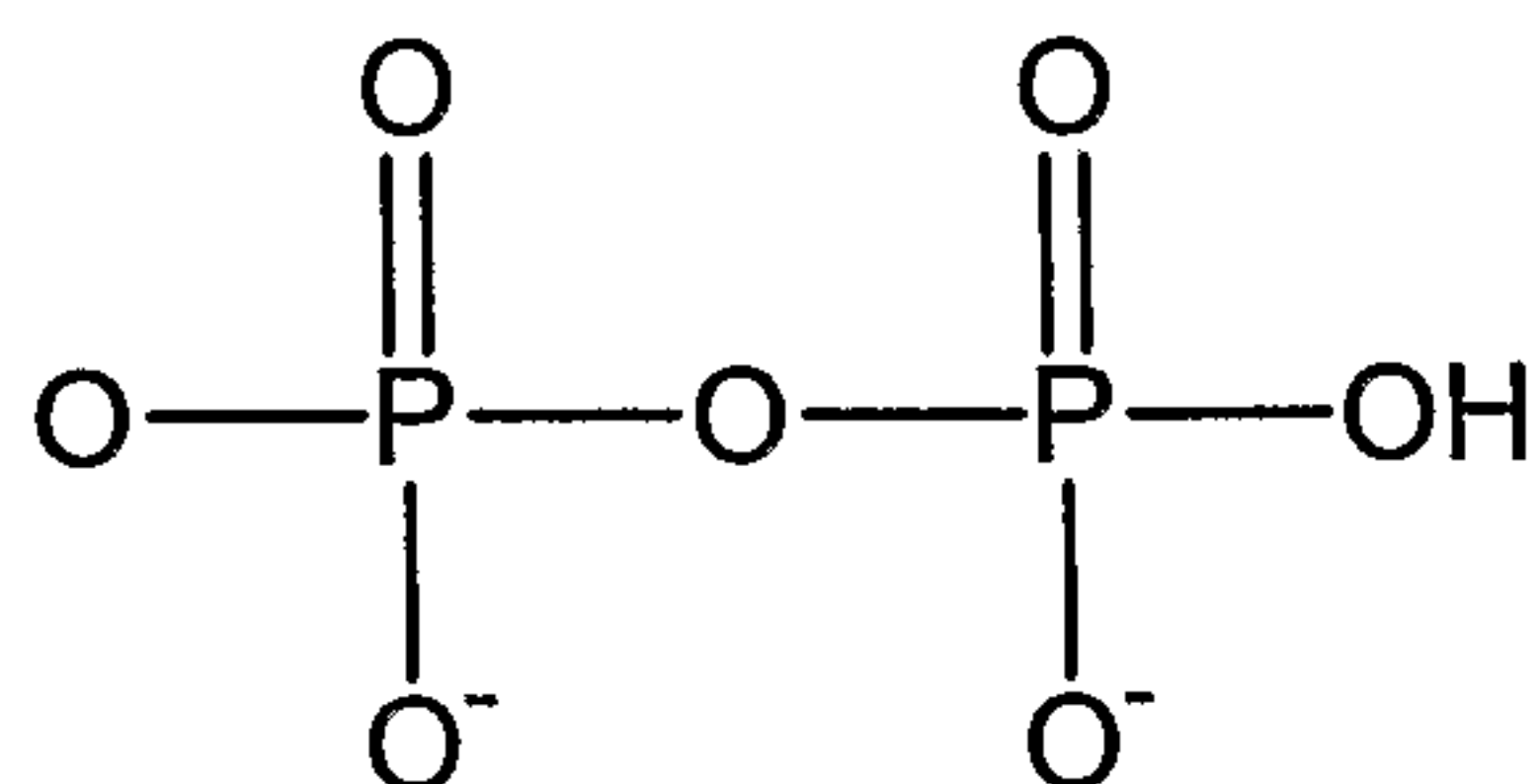


;

Each of A and C is, independently, O or S;

B is OH, O⁻, or

5

R^c is H or C₁-C₆ alkyl;R^d is H or a ligand; and

n is 1-4.

10 The oligonucleotide agent of the conjugate is substantially single-stranded and comprises from about 12 to about 29 subunits, preferably about 15 to about 25 subunits. An oligonucleotide agent that is substantially single-stranded includes at least 60%, 70%, 80%, or 90% or more nucleotides that are not duplexed.

Embodiments can include one or more of the features described above.

15 In a further aspect, this invention features an oligonucleotide agent having at least one subunit comprising formula (I) or formula (II).

In one aspect, this invention features an oligonucleotide agent having at least two subunits comprising formula (I) and/or formula (II).

20 In another aspect, this invention provides a method of making an oligonucleotide agent described herein having at least one subunit comprising formula (I) and/or (II). In a further aspect, this invention provides a method of modulating expression of a target gene. The method includes administering an oligonucleotide agent described herein having at least one subunit comprising formula (I) and/or (II) to a subject.

In one aspect, this invention features a pharmaceutical composition having an oligonucleotide agent described herein having at least one subunit comprising formula (I) and/or (II) and a pharmaceutically acceptable carrier.

SRMSs or tethers described herein may be incorporated into any oligonucleotide agent described herein. An oligonucleotide agent may include one or more of the SRMSs described herein. An SRMS can be introduced at one or more points in an oligonucleotide agent. An SRMS can be placed at or near (within 1, 2, or 3 positions) the 3' or 5' end of the oligonucleotide. In some embodiments, it is preferred to not have an SRMS at or near (within 1, 2, or 3 positions of) the 5' end of the oligonucleotide. An SRMS can be internal, and will preferably be positioned in regions not critical for binding to the target.

In an embodiment, an oligonucleotide agent may have an SRMS at (or within 1, 2, or 3 positions of) the 3' end.

In another embodiment, an oligonucleotide agent may have an SRMS at an internal position. In other embodiments, an oligonucleotide agent may have an SRMS at the 3' end and an SRMS at an internal position.

Other modifications to sugars, bases, or backbones described herein can be incorporated into the oligonucleotide agents.

The oligonucleotide agents can take an architecture or structure described herein.

The oligonucleotide agent can be selected to target any of a broad spectrum of genes, including any of the genes described herein.

In a preferred embodiment the oligonucleotide agent has an architecture (architecture refers to one or more of the overall length) described herein. In addition to the SRMS-containing bases of the oligonucleotide agents described herein can include nuclease resistant monomers (NRMs).

In another aspect, the invention features an oligonucleotide agent to which is conjugated a lipophilic moiety, *e.g.*, cholesterol, *e.g.*, by conjugation to an SRMS of an oligonucleotide agent. In a preferred embodiment, the lipophilic moiety enhances entry of the oligonucleotide agent into a cell. In a preferred embodiment, the cell is part of an organism, tissue, or cell line, *e.g.*, a primary cell line, immortalized cell line, or any type of cell line disclosed herein. Thus, the conjugated oligonucleotide agent can be used to inhibit expression of a target gene in an

organism, *e.g.*, a mammal, *e.g.*, a human, or to inhibit expression of a target gene in a cell line or in cells which are outside an organism.

The lipophilic moiety can be chosen, for example, from the group consisting of a lipid, cholesterol, oleyl, retinyl, cholesteryl residues, cholic acid, adamantane acetic acid, 1-pyrene
 5 butyric acid, dihydrotestosterone, 1,3-Bis-O(hexadecyl)glycerol, geranyloxyhexyl group, hexadecylglycerol, borneol, menthol, 1,3-propanediol, heptadecyl group, palmitic acid, myristic acid, O3-(oleoyl)lithocholic acid, O3-(oleoyl)cholenic acid, dimethoxytrityl, or phenoxazine. A preferred lipophilic moiety is cholesterol.

The oligonucleotide agent can have at least one subunit having formula (I) or formula (II)
 10 incorporated into it. The oligonucleotide agent can have one or more of any of the features described herein. For example, when the subunit is of formula (I), R^d can be cholesterol; X can be $N(CO)R^7$ or NR^7 , Y can be CR^9R^{10} , and Z can be absent, and R^1 can be $(CH_2)_nOR^b$ and R^3 can be OR^a ; X can be $N(CO)R^7$ or NR^7 , Y can be CR^9R^{10} , and Z can be $CR^{11}R^{12}$, and R^9 can be $(CH_2)_nOR^b$ and R^{10} can be OR^a ; X can be $N(CO)R^7$ or NR^7 , Y can be NR^8 , and Z can be
 15 $CR^{11}R^{12}$, and R^1 can be $(CH_2)_nOR^b$ and R^3 can be OR^a ; X can be CH_2 ; Y can be CR^9R^{10} ; and Z can be $CR^{11}R^{12}$, in which R^6 can be $C(O)NHR^7$; or X can be CH_2 ; Y can be CR^9R^{10} ; and Z can be $CR^{11}R^{12}$, in which R^{11} or R^{12} can be $C(O)NHR^7$ or R^5 and R^{11} together can be C_5 or C_6 cycloalkyl substituted with $N(CH_3)R^7$.

In a preferred embodiment, the lipophilic moiety, *e.g.*, a cholesterol, enhances entry of
 20 the oligonucleotide agent into a synoviocyte, myocyte, keratinocyte, hepatocyte, leukocyte, endothelial cell (*e.g.*, a kidney cell), B-cell, T-cell, epithelial cell, mesodermal cell, myeloid cell, neural cell, neoplastic cell, mast cell, or fibroblast cell. In certain aspects, a myocyte can be a smooth muscle cell or a cardiac myocyte, a fibroblast cell can be a dermal fibroblast, and a leukocyte can be a monocyte. In another preferred embodiment, the cell can be from an adherent
 25 tumor cell line derived from a tissue, such as bladder, lung, breast, cervix, colon, pancreas, prostate, kidney, liver, skin, or nervous system (*e.g.*, central nervous system).

In another aspect, the invention provides, methods of inhibiting expression of a target gene by providing an oligonucleotide agent to which a lipophilic moiety is conjugated, *e.g.*, a lipophilic conjugated oligonucleotide agent described herein, to a cell. In a preferred
 30 embodiment the conjugated oligonucleotide agent can be used to inhibit expression of a target gene in an organism, *e.g.*, a mammal, *e.g.*, a human, or to inhibit expression of a target gene in a

cell line or in cells which are outside an organism. In the case of a whole organism, the method can be used to inhibit expression of a gene, *e.g.*, a gene described herein, and treat a condition mediated by the gene. In the case of use on a cell which is not part of an organism, *e.g.*, a primary cell line, secondary cell line, tumor cell line, or transformed or immortalized cell line, the oligonucleotide agent to which a lipophilic moiety is conjugated can be used to inhibit expression of a gene, *e.g.*, one described herein. Cells which are not part of a whole organism can be used in an initial screen to determine if an oligonucleotide agent is effective in inhibiting expression of a gene. A test in cells which are not part of a whole organism can be followed by testing the oligonucleotide agent in a whole animal. In preferred embodiments, the oligonucleotide agent which is conjugated to a lipophilic moiety is administered to an organism, or contacted with a cell which is not part of an organism, in the absence of (or in a reduced amount of) other reagents that facilitate or enhance delivery, *e.g.*, a compound which enhances transit through the cell membrane. (A reduced amount can be an amount of such reagent which is reduced in comparison to what would be needed to get an equal amount of nonconjugated oligonucleotide agent into the target cell). *E.g.*, the oligonucleotide agent which is conjugated to a lipophilic moiety is administered to an organism, or contacted with a cell which is not part of an organism, in the absence (or reduced amount) of: an additional lipophilic moiety; a transfection agent, *e.g.*, concentrations of an ion or other substance which substantially alters cell permeability to an oligonucleotide agent; a transfecting agent such as LipofectamineTM (Invitrogen, Carlsbad, CA), Lipofectamine 2000TM, TransIT-TKOTM (Mirus, Madison, WI), FuGENE 6 (Roche, Indianapolis, IN), polyethylenimine, X-tremeGENE Q2 (Roche, Indianapolis, IN), DOTAP, DOSPER, MetafecteneTM (Biontex, Munich, Germany), and the like.

In a preferred embodiment the oligonucleotide agent is suitable for delivery to a cell *in vivo*, *e.g.*, to a cell in an organism. In another aspect, the oligonucleotide agent is suitable for delivery to a cell *in vitro*, *e.g.*, to a cell in a cell line.

An oligonucleotide agent to which a lipophilic moiety is attached can target any gene described herein and can be delivered to any cell type described herein, *e.g.*, a cell type in an organism, tissue, or cell line. Delivery of the oligonucleotide agent can be *in vivo*, *e.g.*, to a cell in an organism, or *in vitro*, *e.g.*, to a cell in a cell line.

In another aspect, the invention provides compositions of oligonucleotide agents described herein, and in particular compositions of an oligonucleotide agent to which a lipophilic

moiety is conjugated, *e.g.*, a lipophilic conjugated oligonucleotide agent described herein. In a preferred embodiment the composition is a pharmaceutically acceptable composition.

In preferred embodiments, the composition, *e.g.*, pharmaceutically acceptable composition, is free of, has a reduced amount of, or is essentially free of other reagents that facilitate or enhance delivery, *e.g.*, compounds which enhance transit through the cell membrane. (A reduced amount can be an amount of such reagent which is reduced in comparison to what would be needed to get an equal amount of nonconjugated oligonucleotide agent into the target cell). *E.g.*, the composition is free of, has a reduced amount of, or is essentially free of: an additional lipophilic moiety; a transfection agent, *e.g.*, concentrations of an ion or other substance which substantially alters cell permeability to an oligonucleotide agent; a transfecting agent such as LipofectamineTM (Invitrogen, Carlsbad, CA), Lipofectamine 2000TM, TransIT-TKOTM (Mirus, Madison, WI), FuGENE 6 (Roche, Indianapolis, IN), polyethylenimine, XtremeGENE Q2 (Roche, Indianapolis, IN), DOTAP, DOSPER, MetafecteneTM (Biontex, Munich, Germany), and the like.

In a preferred embodiment the composition is suitable for delivery to a cell *in vivo*, *e.g.*, to a cell in an organism. In another aspect, the oligonucleotide agent is suitable for delivery to a cell *in vitro*, *e.g.*, to a cell in a cell line.

The SRMS-containing oligonucleotide agents can be used in any of the methods described herein, *e.g.*, to target any of the genes described herein or to treat any of the disorders described herein. They can be incorporated into any of the formulations, modes of delivery, delivery modalities, kits or preparations, *e.g.*, pharmaceutical preparations, described herein. *E.g.*, a kit which includes one or more of the oligonucleotide agents described herein, a sterile container in which the oligonucleotide agent is disclosed, and instructions for use.

The methods and compositions of the invention, *e.g.*, the SRMS-containing oligonucleotide agents described herein, can be used with any of the oligonucleotide agents described herein. In addition, the methods and compositions of the invention can be used for the treatment of any disease or disorder described herein, and for the treatment of any subject, *e.g.*, any animal, any mammal, such as any human.

The methods and compositions of the invention, *e.g.*, the SRMS-containing oligonucleotide agents described herein, can be used with any dosage and/or formulation described herein, as well as with any route of administration described herein.

The non-ribose scaffolds, as well as monomers and dimers of the SRMSs described herein are within the invention.

An "oligonucleotide agent" refers to a single stranded oligomer or polymer of ribonucleic acid (RNA) or deoxyribonucleic acid (DNA) or both or modifications thereof, which is antisense
5 with respect to its target. This term includes oligonucleotides composed of naturally-occurring nucleobases, sugars and covalent internucleoside (backbone) linkages as well as oligonucleotides having non-naturally-occurring portions which function similarly. Such modified or substituted oligonucleotides are often preferred over native forms because of desirable properties such as, for example, enhanced cellular uptake, enhanced affinity for nucleic acid target and increased
10 stability in the presence of nucleases.

Oligonucleotide agents include both nucleic acid targeting (NAT) oligonucleotide agents and protein-targeting (PT) oligonucleotide agents. NAT and PT oligonucleotide agents refer to single stranded oligomers or polymers of ribonucleic acid (RNA) or deoxyribonucleic acid (DNA) or both or modifications thereof. This term includes oligonucleotides composed of
15 naturally occurring nucleobases, sugars, and covalent internucleoside (backbone) linkages as well as oligonucleotides having non-naturally-occurring portions that function similarly. Such modified or substituted oligonucleotides are often preferred over native forms because of desirable properties such as, for example, enhanced cellular uptake, enhanced affinity for nucleic acid target, and/or increased stability in the presence of nucleases. NATs designed to bind to
20 specific RNA or DNA targets have substantial complementarity, *e.g.*, at least 70, 80, 90, or 100% complementary, with at least 10, 20, or 30 or more bases of a target nucleic acid, and include antisense RNAs, miRNAs, and other non-duplex structures which can modulate expression. Other NAT oligonucleotide agents include external guide sequence (EGS) oligonucleotides (oligozymes), DNazymes, and ribozymes. The NAT oligonucleotide agents can target any
25 nucleic acid, *e.g.*, a miRNA, a pre-miRNA, a pre-mRNA, an mRNA, or a DNA. These NAT oligonucleotide agents may or may not bind via Watson-Crick complementarity to their targets. PT oligonucleotide agents bind to protein targets, preferably by virtue of three-dimensional interactions, and modulate protein activity. They include decoy RNAs, aptamers, and the like.

The compounds in accordance with this invention preferably comprise from about 5 to
30 about 100 nucleobases, *e.g.*, from about 8 to about 75 nucleobases, *e.g.*, from about 8 to about 50 nucleobases. NAT oligonucleotide agents are preferably about 12 or about 15 nucleotides long,

more preferably about 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, or 30 nucleotides long. PT oligonucleotide agents are preferably about 18 nucleotides long, more preferably 23.

Particularly preferred compounds are miRNAs and antisense oligonucleotides, even more preferably those comprising from about 12 to about 30 nucleobases.

5 While not wishing to be bound by theory, an oligonucleotide agent may act by one or more of a number of mechanisms, including a cleavage-dependent or cleavage-independent mechanism. A cleavage-based mechanism can be RNase H dependent and/or can include RISC complex function. Cleavage-independent mechanisms include occupancy-based translational arrest, such as can be mediated by miRNAs, or binding of the oligonucleotide agent to a protein,
10 as do aptamers. Oligonucleotide agents may also be used to alter the expression of genes by changing the choice of splice site in a pre-mRNA. Inhibition of splicing can also result in degradation of the improperly processed message, thus down-regulating gene expression.

An oligonucleotide agent can be administered, *e.g.*, to a cell or to a human, in a single-stranded or double-stranded configuration. An oligonucleotide agent that is in a double-stranded
15 configuration is bound to a substantially complementary oligonucleotide strand. Delivery of an oligonucleotide agent in a double stranded configuration may confer certain advantages on the oligonucleotide agent, such as an increased resistance to nucleases. When an oligonucleotide agent is presented in a double stranded configuration, one or both of the oligonucleotide agent and the substantially complementary strand can include modifications, *e.g.*, any of the
20 modifications described herein, including base modifications, sugar modifications, tethered ligands, and the like.

An oligonucleotide agent featured in the invention can target a substantially complementary miRNA. For example, an oligonucleotide agent can target an endogenous miRNA, such as an miRNA associated with a particular disease or disorder. In one embodiment,
25 an oligonucleotide agent targets an miRNA that is upregulated in tumor cells, *e.g.*, lung tumor cells. For example, the oligonucleotide agent can target miR-21 (Shingara *et al.*, *Ambion TechNotes* 11(6), 2005). In another example, an oligonucleotide agent targets an miRNA that has been found to be upregulated in diffuse large B cell lymphoma (DLBCL) or in Burkitt's Lymphoma. For example, the oligonucleotide agent can target miR-155 (Eis *et al.*, *Proc. Natl.*
30 *Acad. Sci. U.S.A.* 102:3627-3632, 2005; Metzler *et al.*, *Genes Chromosom. Cancer* 39:167-169, 2004). In another embodiment, an oligonucleotide agent targets an miRNA that is enriched in

particular tissues or in particular cell types, *e.g.*, in pancreatic islet cells. For example, the oligonucleotide agent can target miR-375, a pancreatic islet-specific miRNA that has been shown to suppress glucose-induced insulin secretion (Poy *et al.*, *Nature* 432:226-230, 2004). In another embodiment, an miRNA oligonucleotide agent featured in the invention targets an endogenous miRNA that binds a gene for which underexpression is associated with disease. Targeting of such an miRNA with an miRNA oligonucleotide agent will result in upregulation of the disease-associated gene, thereby relieving symptoms of the disease or disorder.

Exemplary single stranded oligonucleotide agents can target RNAs encoding the following polypeptides: vascular endothelial growth factor (VEGF); Apolipoprotein B (ApoB); luciferase (luc); Androgen Receptor (AR); coagulation factor VII (FVII); hypoxia-inducible factor 1, alpha subunit (Hif-1 α); placenta growth factor (PLGF); Lamin A/C; and green fluorescent protein (GFP). Exemplary single stranded oligonucleotide agents are shown in Table 1 below. Additional suitable miRNA targets are described, *e.g.*, in John *et al.*, *PLoS Biology* 2:1862-1879, 2004 (correction in *PLoS* 3:1328, 2005), and The microRNA Registry (Griffiths-Jones S., *NAR* 32:D109-D111, 2004).

Table 1. Exemplary oligonucleotide agents

AL-SQ-NO:	Sequence (5' – 3' unless otherwise indicated)	Target
3186	GCACAUAGGAGAGAUGAGCUUs - Chol	VEGF
3191	Naproxen - sGUCAUCACACUGAAUACCAAUUs - Chol	ApoB
3209	CAUCACACUGAAUACCAAUdTdTts - Chol	Luc
3230	oUsoCsoAoCoGoCoGoAoGoCoCoGoAoAoCoGoAoAoCsoAsoAsoAs - Chol	Mir-375
3234	oCoUGGGAAAGoUoCAAGoCoCoCAoUdTsdT - Chol	AR
3235	oCoUGoUGoCAAGoUGoCoCoCAAGAoUdTsdT - Chol	AR
3253	GGAfUfCAfUfCfUfCAAGfUfCfUfUAfCdTsdT - Chol	FVII
3256	ACUGCAGGGUGAAGAAUUAdTsdTs - Chol	Hif-1 α
3257	GCACAUAGGAGAGAUGAGCUsUs - Chol	VEGF
3258	GAACUGUGUGUGAGAGGUCCsUs - Chol	Luc
3264	CCAGGUUUUUUCUUACUUTsTs - Chol	VEGF
3265	UUCCUCAAUCAAUUACCATsTs - Chol	VEGF
3266	GGAAGGCUCCCUUGAUGGAdTsdTs - Chol	VEGF
3268	GACACAGUGUGUUUGAUUUdTsdTs - Chol	Hif-1 α
3269	UGCCAAGCCAGAUUCUCUdTsdTs - Chol	PLGF

3271	CUCAGGAAUUCAGUGCCUdTsdTs-Chol	PLGF
3275	CUGGACUUCAGAGAACAAdTdT-Chol	Lamin A/C
3150	Chol-sGUCAUCACACUGAAUACCAAsU	ApoB
5225	GUCAUCACACUGAAUACCAAU _s -Chol	ApoB
4967	GcACcAUCUUCUUCaAGGACGs-Chol	GFP
5225	GUCAUCACACUGAAUACCAAU _s -Chol	ApoB
5221	AGGUGUAUGGCUUCAACCCUG _s -Chol	ApoB
5255	GUGAUCAGACUCAAUACGAAU _s -Chol	ApoB
5474	GGAAUCoUoUAoUAoUoUGAUCoCAAs-Chol	ApoB
4750	CCACAUGAAGCAGCACGACUU _s -Chol	GFP
3148	GUCAUCACACUGAAUACCAAU _s -Thiochol	ApoB
3208	AUUGGUAUUCAGUGUGAUGAoCsoAsCs-Thiochol	ApoB
3233	AUUGGUAUUCAGUGUGAUGAoCsoAsCs-Thiochol	ApoB
2774	CUUACGCUGAGUACUUCGAdTdT-Thiochol	Luc
2775	UCGAAGUACUCAGCGUAAGdTdT-Thiochol	Luc
3149	Thiochol-sGUCAUCACACUGAAUACCAAsU	ApoB
3207	AUUGGUAUUCAGUGUGAUGAoCsoAsCs-Cholanic acid	ApoB
3231	GUCAUCACACUGAAUACCAAU _s -Lithocholic I	ApoB
3189	GUCAUCACACUGAAUACCAAU _s -Distearyllyceride	ApoB
2767	CUUACGCUGAGUACUUCGAdTdT-Distearyllyceride	Luc
2768	3' Distearyllyceride-dTdTGAAUGCGACUCAUGAAGCU 5'	Luc
3204	Distearyllyceride-sGUCAUCACACUGAAUACCAAsU	ApoB
2918	Distearyllyceride-CUUACGCUGAGUACUUCGAdTdT	ApoB
2919	3' dTdTGAAUGCGACUCAUGAAGCU-Distearyllyceride 5'	Luc
3190	GUCAUCACACUGAAUACCAAU _s -Vitamin E	ApoB
2920	Vitamin E-CUUACGCUGAGUACUUCGA dTdT'	Luc
2921	3' dTdTGAAUGCGACUCAUGAAGCU-Vitamin E 5'	ApoB
3192	Aminoalkyl-sGUCAUCACACUGAAUACCAAU _s -Chol	ApoB

“oN” (N = A, C, G or U) indicates 2'-O-Methyl modified nucleotide; “fN” (N = A, C, G or U) indicates 2'-deoxy-2'-fluoro modified nucleotide, “s” indicates phosphorothioate linkage; “Chol” indicates cholesterol conjugate; “Thiochol” indicates thiocholesterol conjugate; “Cholanic Acid” indicates 5β-cholanic acid conjugate; “Naproxen” indicates Naproxen conjugate; “Lithocholic I” indicates lithocholic acid derivative conjugate; “Distearyllyceride” indicates distearyllyceride conjugate; “Vitamin E” indicates vitamin E conjugate and “Aminoalkyl” indicates amino linker conjugate.

An oligonucleotide agent featured in the invention can include a nucleotide sequence that is substantially identical to a nucleotide sequence of an miRNA, such as an endogenous miRNA. An oligonucleotide sequence that is substantially identical to a second nucleotide sequence is 70%, 80%, 90%, or more identical to the second nucleotide sequence. Preferably, the agent is identical in sequence with an endogenous miRNA. An oligonucleotide agent that is substantially identical to a nucleotide sequence of an miRNA can be delivered to a cell or a human to replace or supplement the activity of an endogenous miRNA, such as when an miRNA deficiency is linked to a disease or disorder. In one embodiment, an oligonucleotide agent featured in the invention can have a nucleotide sequence that is substantially identical to an miRNA known to be down-regulated or lost in certain cancers. For example, an oligonucleotide agent can have a nucleotide sequence that is substantially identical to miR-15 (e.g., miR-15a or miR-15b) or miR-16, miRNAs known to be downregulated or lost in many cases of B cell chronic lymphocytic leukemia, mantle cell lymphoma, multiple myelomas and prostate cancer (Callin *et al.*, *Proc. Natl. Acad. Sci.* 99:15524-15529, 2002). In another example, an oligonucleotide agent can have a nucleotide sequence that is substantially identical to miR-143 or miR-145, miRNAs known to be downregulated in many cases of adenomatous and cancer stages of colorectal neoplasia (Micheal *et al.*, *Mol. Cancer Res.* 1:882-891, 2003). In yet another example, an oligonucleotide agent can have a nucleotide sequence that is substantially identical to let-7, an miRNA known to be downregulated in lung cancer tissue (Johnson *et al.*, *Cell* 120:635-647, 2005).

Oligonucleotide agents that are substantially identical to at least a portion of an miRNA, such as those described above, can be administered to a subject to treat the disease or disorder associated with the downregulation of miRNA expression. Other suitable oligonucleotide agents are substantially identical to miRNAs described, e.g., in John *et al.*, *PLoS Biology* 2:1862-1879, 2004 (correction in *PLoS* 3:1328, 2005), and The microRNA Registry (Griffiths-Jones, *NAR* 32:D109-D111, 2004).

MicroRNA-type Oligonucleotide Agents

Oligonucleotide agents include microRNAs (miRNAs). MicroRNAs are small noncoding RNA molecules that are capable of causing post-transcriptional silencing of specific genes in cells such as by the inhibition of translation or through degradation of the targeted

mRNA. An miRNA can be completely complementary or can have a region of noncomplementarity with a target nucleic acid, consequently resulting in a “bulge” at the region of non-complementarity. The region of noncomplementarity (the bulge) can be flanked by regions of sufficient complementarity, preferably complete complementarity to allow duplex
5 formation. Preferably, the regions of complementarity are at least 8 to 10 nucleotides long (*e.g.*, 8, 9, or 10 nucleotides long). A miRNA can inhibit gene expression by repressing translation, such as when the microRNA is not completely complementary to the target nucleic acid, or by causing target RNA degradation, which is believed to occur only when the miRNA binds its target with perfect complementarity. The invention also can include double-stranded precursors
10 of miRNAs that may or may not form a bulge when bound to their targets.

In a preferred embodiment an oligonucleotide agent featured in the invention can target an endogenous miRNA or pre-miRNA. The oligonucleotide agent featured in the invention can include naturally occurring nucleobases, sugars, and covalent internucleoside (backbone) linkages as well as oligonucleotides having non-naturally-occurring portions that function
15 similarly. Such modified or substituted oligonucleotides are often preferred over native forms because of desirable properties such as, for example, enhanced cellular uptake, enhanced affinity for the endogenous miRNA target, and/or increased stability in the presence of nucleases. An oligonucleotide agent designed to bind to a specific endogenous miRNA has substantial complementarity, *e.g.*, at least 70, 80, 90, or 100% complementary, with at least 10, 20, or 25 or
20 more bases of the target miRNA.

A miRNA or pre-miRNA can be 18-100 nucleotides in length, and more preferably from 18-80 nucleotides in length. Mature miRNAs can have a length of 19-30 nucleotides, preferably 21-25 nucleotides, particularly 21, 22, 23, 24, or 25 nucleotides. MicroRNA precursors can have a length of 70-100 nucleotides and have a hairpin conformation. MicroRNAs can be generated
25 *in vivo* from pre-miRNAs by enzymes called Dicer and Drosha that specifically process long pre-miRNA into functional miRNA. The microRNAs or precursor mi-RNAs featured in the invention can be synthesized *in vivo* by a cell-based system or can be chemically synthesized. MicroRNAs can be synthesized to include a modification that imparts a desired characteristic. For example, the modification can improve stability, hybridization thermodynamics with a target
30 nucleic acid, targeting to a particular tissue or cell-type, or cell permeability, *e.g.*, by an

endocytosis-dependent or -independent mechanism. Modifications can also increase sequence specificity, and consequently decrease off-site targeting. Methods of synthesis and chemical modifications are described in greater detail below.

Given a sense strand sequence (*e.g.*, the sequence of a sense strand of a cDNA molecule),
5 an miRNA can be designed according to the rules of Watson and Crick base pairing. The
miRNA can be complementary to a portion of an RNA, *e.g.*, a miRNA, a pre-miRNA, a pre-
mRNA or an mRNA. For example, the miRNA can be complementary to the coding region or
noncoding region of an mRNA or pre-mRNA, *e.g.*, the region surrounding the translation start
site of a pre-mRNA or mRNA, such as the 5' UTR. An miRNA oligonucleotide can be, for
10 example, from about 12 to 30 nucleotides in length, preferably about 15 to 28 nucleotides in
length (*e.g.*, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 nucleotides in length).

In particular, an miRNA or a pre-miRNA featured in the invention can have a chemical
modification on a nucleotide in an internal (*i.e.*, non-terminal) region having
noncomplementarity with the target nucleic acid. For example, a modified nucleotide can be
15 incorporated into the region of a miRNA that forms a bulge. The modification can include a
ligand attached to the miRNA, *e.g.*, by a linker (*e.g.*, see diagrams OT-I through OT-IV below).
The modification can, for example, improve pharmacokinetics or stability of a therapeutic
miRNA, or improve hybridization properties (*e.g.*, hybridization thermodynamics) of the miRNA
to a target nucleic acid. In some embodiments, it is preferred that the orientation of a
20 modification or ligand incorporated into or tethered to the bulge region of a miRNA is oriented
to occupy the space in the bulge region. For example, the modification can include a modified
base or sugar on the nucleic acid strand or a ligand that functions as an intercalator. These are
preferably located in the bulge. The intercalator can be an aromatic, *e.g.*, a polycyclic aromatic
or heterocyclic aromatic compound. A polycyclic intercalator can have stacking capabilities, and
25 can include systems with 2, 3, or 4 fused rings. The universal bases described below can be
incorporated into the miRNAs. In some embodiments, it is preferred that the orientation of a
modification or ligand incorporated into or tethered to the bulge region of a miRNA is oriented
to occupy the space in the bulge region. This orientation facilitates the improved hybridization
properties or an otherwise desired characteristic of the miRNA.

In one embodiment, an miRNA or a pre-miRNA can include an aminoglycoside ligand, which can cause the miRNA to have improved hybridization properties or improved sequence specificity. Exemplary aminoglycosides include glycosylated polylysine; galactosylated polylysine; neomycin B; tobramycin; kanamycin A; and acridine conjugates of aminoglycosides, such as Neo-N-acridine, Neo-S-acridine, Neo-C-acridine, Tobra-N-acridine, and KanaA-N-acridine. Use of an acridine analog can increase sequence specificity. For example, neomycin B has a high affinity for RNA as compared to DNA, but low sequence-specificity. An acridine analog, neo-S-acridine has an increased affinity for the HIV Rev-response element (RRE). In some embodiments the guanidine analog (the guanidinoglycoside) of an aminoglycoside ligand is tethered to an oligonucleotide agent. In a guanidinoglycoside, the amine group on the amino acid is exchanged for a guanidine group. Attachment of a guanidine analog can enhance cell permeability of an oligonucleotide agent.

In one embodiment, the ligand can include a cleaving group that contributes to target gene inhibition by cleavage of the target nucleic acid. Preferably, the cleaving group is tethered to the miRNA in a manner such that it is positioned in the bulge region, where it can access and cleave the target RNA. The cleaving group can be, for example, a bleomycin (*e.g.*, bleomycin-A₅, bleomycin-A₂, or bleomycin-B₂), pyrene, phenanthroline (*e.g.*, *O*-phenanthroline), a polyamine, a tripeptide (*e.g.*, lys-tyr-lys tripeptide), or metal ion chelating group. The metal ion chelating group can include, *e.g.*, an Lu(III) or EU(III) macrocyclic complex, a Zn(II) 2,9-dimethylphenanthroline derivative, a Cu(II) terpyridine, or acridine, which can promote the selective cleavage of target RNA at the site of the bulge by free metal ions, such as Lu(III). In some embodiments, a peptide ligand can be tethered to a miRNA or a pre-miRNA to promote cleavage of the target RNA, *e.g.*, at the bulge region. For example, 1,8-dimethyl-1,3,6,8,10,13-hexaazacyclotetradecane (cyclam) can be conjugated to a peptide (*e.g.*, by an amino acid derivative) to promote target RNA cleavage. The methods and compositions featured in the invention include miRNAs that inhibit target gene expression by a cleavage or non-cleavage dependent mechanism.

An miRNA or a pre-miRNA can be designed and synthesized to include a region of noncomplementarity (*e.g.*, a region that is 3, 4, 5, or 6 nucleotides long) flanked by regions of

sufficient complementarity to form a duplex (*e.g.*, regions that are 7, 8, 9, 10, or 11 nucleotides long).

For increased nuclease resistance and/or binding affinity to the target, the miRNA sequences can include 2'-O-methyl, 2'-fluorine, 2'-O-methoxyethyl, 2'-O-aminopropyl, 2'-amino, and/or phosphorothioate linkages. Inclusion of locked nucleic acids (LNA), 2-thiopyrimidines (*e.g.*, 2-thio-U), 2-amino-A, G-clamp modifications, and ethylene nucleic acids (ENA), *e.g.*, 2'-4'-ethylene-bridged nucleic acids, can also increase binding affinity to the target. The inclusion of furanose sugars in the oligonucleotide backbone can also decrease endonucleolytic cleavage. An miRNA or a pre-miRNA can be further modified by including a 3' cationic group, or by inverting the nucleoside at the 3'-terminus with a 3'-3' linkage. In another alternative, the 3'-terminus can be blocked with an aminoalkyl group, *e.g.*, a 3' C5-aminoalkyl dT. Other 3' conjugates can inhibit 3'-5' exonucleolytic cleavage. While not being bound by theory, a 3' conjugate, such as naproxen or ibuprofen, may inhibit exonucleolytic cleavage by sterically blocking the exonuclease from binding to the 3' end of oligonucleotide. Even small alkyl chains, aryl groups, or heterocyclic conjugates or modified sugars (D-ribose, deoxyribose, glucose etc.) can block 3'-5'-exonucleases.

The 5' -terminus can be blocked with an aminoalkyl group, *e.g.*, a 5'-O-alkylamino substituent. Other 5' conjugates can inhibit 5'-3' exonucleolytic cleavage. While not being bound by theory, a 5' conjugate, such as naproxen or ibuprofen, may inhibit exonucleolytic cleavage by sterically blocking the exonuclease from binding to the 5' end of oligonucleotide. Even small alkyl chains, aryl groups, or heterocyclic conjugates or modified sugars (D-ribose, deoxyribose, glucose etc.) can block 3'-5'-exonucleases.

In one embodiment, an miRNA or a pre-miRNA includes a modification that improves targeting, *e.g.* a targeting modification described herein. Examples of modifications that target miRNA molecules to particular cell types include carbohydrate sugars such as galactose, N-acetylgalactosamine, mannose; vitamins such as folates; other ligands such as RGDs and RGD mimics; and small molecules including naproxen, ibuprofen or other known protein-binding molecules.

An miRNA or a pre-miRNA can be constructed using chemical synthesis and/or enzymatic ligation reactions using procedures known in the art. For example, an miRNA or a pre-miRNA can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the miRNA or a pre-miRNA and target nucleic acids, *e.g.*, phosphorothioate derivatives and acridine substituted nucleotides can be used. Other appropriate nucleic acid modifications are described herein. Alternatively, the miRNA or pre-miRNA nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (*i.e.*, RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest).

Antisense-type Oligonucleotide Agents

The single-stranded oligonucleotide agents featured in the invention include antisense nucleic acids. An "antisense" nucleic acid includes a nucleotide sequence that is complementary to a "sense" nucleic acid encoding a gene expression product, *e.g.*, complementary to the coding strand of a double-stranded cDNA molecule or complementary to an RNA sequence, *e.g.*, a pre-mRNA, mRNA, miRNA, or pre-miRNA. Accordingly, an antisense nucleic acid can form hydrogen bonds with a sense nucleic acid target.

Given a coding strand sequence (*e.g.*, the sequence of a sense strand of a cDNA molecule), antisense nucleic acids can be designed according to the rules of Watson and Crick base pairing. The antisense nucleic acid molecule can be complementary to a portion of the coding or noncoding region of an RNA, *e.g.*, a pre-mRNA or mRNA. For example, the antisense oligonucleotide can be complementary to the region surrounding the translation start site of a pre-mRNA or mRNA, *e.g.*, the 5' UTR. An antisense oligonucleotide can be, for example, about 10 to 25 nucleotides in length (*e.g.*, 11, 12, 13, 14, 15, 16, 18, 19, 20, 21, 22, 23, or 24 nucleotides in length). An antisense oligonucleotide can also be complementary to a miRNA or pre-miRNA.

An antisense nucleic acid can be constructed using chemical synthesis and/or enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (*e.g.*, an antisense oligonucleotide) can be chemically synthesized using naturally occurring

nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and target nucleic acids, *e.g.*, phosphorothioate derivatives and acridine substituted nucleotides can be used. Other appropriate nucleic acid modifications are described herein. Alternatively, the
5 antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (*i.e.*, RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest).

An antisense agent can include ribonucleotides only, deoxyribonucleotides only (*e.g.*, oligodeoxynucleotides), or both deoxyribonucleotides and ribonucleotides. For example, an
10 antisense agent consisting only of ribonucleotides can hybridize to a complementary RNA, and prevent access of the translation machinery to the target RNA transcript, thereby preventing protein synthesis. An antisense molecule including only deoxyribonucleotides, or deoxyribonucleotides and ribonucleotides, *e.g.*, DNA sequence flanked by RNA sequence at the 5' and 3' ends of the antisense agent, can hybridize to a complementary RNA, and the RNA
15 target can be subsequently cleaved by an enzyme, *e.g.*, RNase H. Degradation of the target RNA prevents translation. The flanking RNA sequences can include 2'-O-methylated nucleotides, and phosphorothioate linkages, and the internal DNA sequence can include phosphorothioate internucleotide linkages. The internal DNA sequence is preferably at least five nucleotides in length when targeting by RNaseH activity is desired.

20 For increased nuclease resistance, an antisense agent can be further modified by inverting the nucleoside at the 3'-terminus with a 3'-3' linkage. In another alternative, the 3'-terminus can be blocked with an aminoalkyl group.

In one embodiment, an antisense oligonucleotide agent includes a modification that improves targeting, *e.g.* a targeting modification described herein.

25 Decoy-type Oligonucleotide Agents

An oligonucleotide agent featured in the invention can be a decoy nucleic acid, *e.g.*, a decoy RNA. A decoy nucleic acid resembles a natural nucleic acid, but is modified in such a way as to inhibit or interrupt the activity of the natural nucleic acid. For example, a decoy RNA

can mimic the natural binding domain for a ligand. The decoy RNA therefore competes with natural binding target for the binding of a specific ligand. The natural binding target can be an endogenous nucleic acid, *e.g.*, a pre-miRNA, miRNA, premRNA, mRNA or DNA. For example, it has been shown that over-expression of HIV trans-activation response (TAR) RNA can act as a
5 "decoy" and efficiently bind HIV tat protein, thereby preventing it from binding to TAR sequences encoded in the HIV RNA.

In one embodiment, a decoy RNA includes a modification that improves targeting, *e.g.* a targeting modification described herein.

The chemical modifications described above for miRNAs and antisense RNAs, and
10 described elsewhere herein, are also appropriate for use in decoy nucleic acids.

Aptamer-type Oligonucleotide Agents

An oligonucleotide agent featured in the invention can be an aptamer. An aptamer binds to a non-nucleic acid ligand, such as a small organic molecule or protein, *e.g.*, a transcription or
15 translation factor, and subsequently modifies (*e.g.*, inhibits) activity. An aptamer can fold into a specific structure that directs the recognition of the targeted binding site on the non-nucleic acid ligand. An aptamer can contain any of the modifications described herein.

In one embodiment, an aptamer includes a modification that improves targeting, *e.g.* a targeting modification described herein.

The chemical modifications described above for miRNAs and antisense RNAs, and
20 described elsewhere herein, are also appropriate for use in decoy nucleic acids.

The details of one or more embodiments of the invention are set forth in the accompanying drawings and the description below. Other features and advantages of the invention will be apparent from the description and drawings, and from the claims. This
25 application incorporates all cited references, patents, and patent applications by references in their entirety for all purposes.

DESCRIPTION OF DRAWINGS

FIG. 1 a general synthetic scheme for incorporation of SRMS monomers into an
30 oligonucleotide.

FIG. 2A is a list of substituents that may be present on silicon in OFG¹.

FIG. 2B is a list of substituents that may be present on the C2'-orthoester group.

FIG. 3 is list of representative cyclic, sugar replacement monomer subunit (SRMS) carriers. Panel 1 shows pyrroline-based SRMSs; panel 2 shows 3-hydroxyproline-based SRMSs; panel 3 shows piperidine-based SRMSs; panel 4 shows morpholine and piperazine-based SRMSs; and panel 5 shows decalin-based SRMSs. R1 is succinate or phosphoramidate and R2 is H or a conjugate ligand.

FIG. 4 is a general reaction scheme for 3' conjugation of peptide into an oligonucleotide agent.

FIG. 5 is a general reaction scheme for 5' conjugation of peptide into an oligonucleotide agent.

FIG. 6 is a general reaction scheme for the synthesis of aza-peptides.

FIG. 7 is a general reaction scheme for the synthesis of N-methyl amino acids and peptides.

FIG. 8 is a general reaction scheme for the synthesis of β -methyl amino acids and Ant and Tat peptides.

FIG. 9 is a general reaction scheme for the synthesis of Ant and Tat oligocarbamates.

FIG. 10 is a a general reaction scheme for the synthesis of Ant and Tat oligoureases.

FIG. 11 is a schematic representation of peptide carriers.

FIG. 12. is a list of representative cholesterol-tethered SRMS monomers.

FIG. 13 shows LCMS data for a 3' cholesterol conjugate after PAGE purification.

FIG. 14 is a denaturing gel analysis of the human serum stability assay for AL-DUP-1000. C is the 4 hour time point for siRNA duplex incubated in PBS buffer alone, OH- is the partial alkaline hydrolysis marker, *s/as represents siRNA duplex containing 5' end-labeled sense RNA and s/*as represents duplex containing 5' end-labeled antisense RNA. Samples were incubated in 90% human serum and time points were assayed at 10 seconds, 5 min, 15 min, 30 min, 1 hour, 2 hours and 4 hours. Black lines to the right of bands indicate exonucleolytic degradation fragments and the red lines highlight a few of the endonucleolytic degradation fragment.

FIG. 15A is a denaturing gel analysis of the human serum stability assay for AL-DUP-1393. C is the 4 hour time point for each siRNA duplex incubated in PBS buffer alone, *s/as represents siRNA duplex containing 5' end-labeled sense RNA and s/*as represents duplex containing 5' end-labeled antisense RNA. Samples were assayed at 10 seconds, 15 min, 30 min, 1 hour, 2 hours and 4 hours.

FIG. 15B is a denaturing gel analysis of the human serum stability assay for AL-DUP-1329. The lanes are labeled and the experiment was performed as described for FIG. 15A.

FIG. 16 is a denaturing gel analysis of AL-DUP-1036, AL-DUP-13ff, and AL-DUP-1363 (see Table 12 for sequences). Black vertical lines highlight regions where exonuclease cleavage is suppressed, stars indicate sites of strong endonucleolytic cleavage in the antisense strand and weaker endonucleolytic cleavage in the sense strand. C is the 4 hour time point for each siRNA duplex incubated in PBS buffer alone, *s/as represents siRNA duplex containing 5' end-labeled sense RNA and s/*as represents duplex containing 5' end-labeled antisense RNA. Samples were assayed at 10 seconds, 15 min, 30 min, 1 hour, 2 hours and 4 hours.

FIG. 17. Human serum stability profile of siRNA duplexes containing cationic modifications. Denaturing gel analysis of AL-DUP-10aa (alkylamino-dT), AL-DUP-1ccc (abasic pyrrolidine cationic), and AL-DUP-1403 (see Table 13 for sequences). Black line highlights region where exonuclease cleavage is suppressed and red star indicates site of strong endonucleolytic cleavage in the antisense strand. C is the 4 hour time point for each siRNA duplex incubated in PBS buffer alone, *s/as represents siRNA duplex containing 5' end-labeled sense RNA and s/*as represents duplex containing 5' end-labeled antisense RNA. Samples were assayed at 10 seconds, 15 min, 30 min, 1 hour, 2 hours and 4 hours.

FIG. 18 is a denaturing gel analysis of the human serum stability assay for AL-DUP-1069. The black vertical line highlights the region where exonuclease cleavage is suppressed. C is the 4 hour time point for each siRNA duplex incubated in PBS buffer alone, *s/as represents siRNA duplex containing 5' end-labeled sense RNA and s/*as represents duplex containing 5' end-labeled antisense RNA. Samples were assayed at 10 seconds, 15 min, 30 min, 1 hour, 2 hours and 4 hours.

FIG. 19A is a panel of Northern blots of total RNA (15 μ g) isolated from mouse liver 24 h after injection of differently modified RNAs (240 mg/kg) targeting miR-122. Samples were separated in 14%-polyacrylamide gels in the absence of formamide, and the membranes were probed for miR-122. Ethidium bromide staining of tRNA is shown as a loading control.

5 FIG. 19B is a panel of Northern blots of total RNA (15 μ g) isolated from mouse liver 24 h after injection of differently modified RNAs (240 mg/kg) against miR-122. Samples were separated in 14%-polyacrylamide gels in the absence of formamide, and the membranes were probed for miR-122, let7, and miR-22 RNAs. Ethidium bromide staining of tRNA is shown as a loading control.

10 FIG. 19C is a panel of Northern blots of total RNA (15 μ g) isolated from mouse liver 24 h after injection of differently modified RNAs (240 mg/kg) against miR-122. Samples were separated in 14%-polyacrylamide gels in the presence of 20% formamide, and the membranes were probed for miR-122. Ethidium bromide staining of tRNA is shown as a loading control.

FIG. 20A is a panel of Northern blots of total RNA (15 μ g) isolated from mouse livers. RNA was isolated 24 h after injection of 80 mg/kg bodyweight antagomir-122 (n=2) on 1, 2, or 15 3 consecutive days as indicated. Membranes were probed for both the endogenous miR-122 and the injected antagomir-122. Ethidium bromide staining of tRNA is shown as a loading control.

FIG. 20B is a panel of Northern blots of total RNA (15 μ g) isolated from mouse livers. RNA was isolated 3, 6, 9, 13, and 23 days after injection of antagomir-122. Membranes were 20 probed for both the endogenous miR-122 and the injected antagomir-122. Ethidium bromide staining of tRNA is shown as a loading control.

FIG. 21A is a panel of Northern blots of total RNA (10-30 μ g) isolated from different mouse tissues 24 h after injection of antagomir-16 (n=3). Membranes were probed for miR-16. The precursor miR-16 transcript was visible on Northern blots of bone marrow and expression 25 was similar in all mice. Ethidium bromide staining of tRNA is shown as a loading control.

FIG. 21B is a panel of Northern blots of total RNA (10-30 μ g) isolated from different mouse tissues 24 h after injection of antagomir-16 (n=3). Total RNA from 3 mice were pooled for the detection of miR-16 and the injected antagomir-16. Ethidium bromide staining of tRNA is shown as a loading control.

DETAILED DESCRIPTION

In a typical embodiment, the subject is a mammal such as a cow, horse, mouse, rat, dog, pig, goat, or a primate. The subject can be a dairy mammal (*e.g.*, a cow, or goat) or other farmed
5 animal (*e.g.*, a chicken, turkey, sheep, pig, fish, shrimp). In a much preferred embodiment, the subject is a human, *e.g.*, a normal individual or an individual that has, is diagnosed with, or is predicted to have a disease or disorder.

Further, because oligonucleotide agent-mediated modulation persists for several days after administering the oligonucleotide agent composition, in many instances it is possible to
10 administer the composition with a frequency of less than once per day, or, for some instances, only once for the entire therapeutic regimen. For example, treatment of some cancer cells may be mediated by a single bolus administration, whereas a chronic viral infection may require regular administration, *e.g.*, once per week or once per month. For example, treatment of diffuse large B cell lymphoma (DLBCL) or Burkitt's lymphoma may be treated with a single bolus
15 administration of a single-stranded oligonucleotide agent, *e.g.*, a single-stranded oligonucleotide agent that targets miR-155.

A number of exemplary routes of delivery are described that can be used to administer an oligonucleotide agent to a subject. In addition, the oligonucleotide agent can be formulated according to an exemplary method described herein.

20 LIGAND-CONJUGATED MONOMER SUBUNITS AND MONOMERS FOR OLIGONUCLEOTIDE SYNTHESIS

Definitions

The term "halo" refers to any radical of fluorine, chlorine, bromine or iodine.

25 The term "alkyl" refers to a hydrocarbon chain that may be a straight chain or branched chain, containing the indicated number of carbon atoms. For example, C₁-C₁₂ alkyl indicates that the group may have from 1 to 12 (inclusive) carbon atoms in it. The term "haloalkyl" refers to an alkyl in which one or more hydrogen atoms are replaced by halo, and includes alkyl moieties in which all hydrogens have been replaced by halo (*e.g.*, perfluoroalkyl). Alkyl and

haloalkyl groups may be optionally inserted with O, N, or S. The terms "aralkyl" refers to an alkyl moiety in which an alkyl hydrogen atom is replaced by an aryl group. Aralkyl includes groups in which more than one hydrogen atom has been replaced by an aryl group. Examples of "aralkyl" include benzyl, 9-fluorenyl, benzhydryl, and trityl groups.

5 The term "alkenyl" refers to a straight or branched hydrocarbon chain containing 2-8 carbon atoms and characterized in having one or more double bonds. Examples of a typical alkenyl include, but not limited to, allyl, propenyl, 2-butenyl, 3-hexenyl and 3-octenyl groups. The term "alkynyl" refers to a straight or branched hydrocarbon chain containing 2-8 carbon atoms and characterized in having one or more triple bonds. Some examples of a typical alkynyl
10 are ethynyl, 2-propynyl, and 3-methylbutynyl, and propargyl. The sp^2 and sp^3 carbons may optionally serve as the point of attachment of the alkenyl and alkynyl groups, respectively.

The terms "alkylamino" and "dialkylamino" refer to $-NH(alkyl)$ and $-NH(alkyl)_2$ radicals respectively. The term "aralkylamino" refers to a $-NH(aralkyl)$ radical. The term "alkoxy" refers to an $-O-alkyl$ radical, and the terms "cycloalkoxy" and "aralkoxy" refer to an -
15 O-cycloalkyl and O-aralkyl radicals respectively. The term "siloxy" refers to a R_3SiO- radical. The term "mercapto" refers to an SH radical. The term "thioalkoxy" refers to an $-S-alkyl$ radical.

The term "alkylene" refers to a divalent alkyl (*i.e.*, $-R-$), *e.g.*, $-CH_2-$, $-CH_2CH_2-$, and $-CH_2CH_2CH_2-$. The term "alkylenedioxo" refers to a divalent species of the structure $-O-R-O-$, in which R represents an alkylene.

20 The term "aryl" refers to an aromatic monocyclic, bicyclic, or tricyclic hydrocarbon ring system, wherein any ring atom can be substituted. Examples of aryl moieties include, but are not limited to, phenyl, naphthyl, anthracenyl, and pyrenyl.

The term "cycloalkyl" as employed herein includes saturated cyclic, bicyclic, tricyclic, or polycyclic hydrocarbon groups having 3 to 12 carbons, wherein any ring atom can be substituted.
25 The cycloalkyl groups herein described may also contain fused rings. Fused rings are rings that share a common carbon-carbon bond or a common carbon atom (*e.g.*, spiro-fused rings). Examples of cycloalkyl moieties include, but are not limited to, cyclohexyl, adamantyl, and norbornyl, and decalin.

The term "heterocyclyl" refers to a nonaromatic 3-10 membered monocyclic, 8-12
30 membered bicyclic, or 11-14 membered tricyclic ring system having 1-3 heteroatoms if monocyclic, 1-6 heteroatoms if bicyclic, or 1-9 heteroatoms if tricyclic, said heteroatoms

selected from O, N, or S (*e.g.*, carbon atoms and 1-3, 1-6, or 1-9 heteroatoms of N, O, or S if monocyclic, bicyclic, or tricyclic, respectively), wherein any ring atom can be substituted. The heterocyclyl groups herein described may also contain fused rings. Fused rings are rings that share a common carbon-carbon bond or a common carbon atom (*e.g.*, spiro-fused rings).

5 Examples of heterocyclyl include, but are not limited to tetrahydrofuranyl, tetrahydropyranyl, piperidinyl, morpholino, pyrrolinyl and pyrrolidinyl.

The term "cycloalkenyl" as employed herein includes partially unsaturated, nonaromatic, cyclic, bicyclic, tricyclic, or polycyclic hydrocarbon groups having 5 to 12 carbons, preferably 5 to 8 carbons, wherein any ring atom can be substituted. The cycloalkenyl groups herein
10 described may also contain fused rings. Fused rings are rings that share a common carbon-carbon bond or a common carbon atom (*e.g.*, spiro-fused rings). Examples of cycloalkenyl moieties include, but are not limited to cyclohexenyl, cyclohexadienyl, or norbornenyl.

The term "heterocycloalkenyl" refers to a partially saturated, nonaromatic 5-10 membered monocyclic, 8-12 membered bicyclic, or 11-14 membered tricyclic ring system
15 having 1-3 heteroatoms if monocyclic, 1-6 heteroatoms if bicyclic, or 1-9 heteroatoms if tricyclic, said heteroatoms selected from O, N, or S (*e.g.*, carbon atoms and 1-3, 1-6, or 1-9 heteroatoms of N, O, or S if monocyclic, bicyclic, or tricyclic, respectively), wherein any ring atom can be substituted. The heterocycloalkenyl groups herein described may also contain fused rings. Fused rings are rings that share a common carbon-carbon bond or a common carbon atom
20 (*e.g.*, spiro-fused rings). Examples of heterocycloalkenyl include but are not limited to tetrahydropyridyl and dihydropyran.

The term "heteroaryl" refers to an aromatic 5-8 membered monocyclic, 8-12 membered bicyclic, or 11-14 membered tricyclic ring system having 1-3 heteroatoms if monocyclic, 1-6 heteroatoms if bicyclic, or 1-9 heteroatoms if tricyclic, said heteroatoms selected from O, N, or S
25 (*e.g.*, carbon atoms and 1-3, 1-6, or 1-9 heteroatoms of N, O, or S if monocyclic, bicyclic, or tricyclic, respectively), wherein any ring atom can be substituted. The heteroaryl groups herein described may also contain fused rings that share a common carbon-carbon bond.

The term "oxo" refers to an oxygen atom, which forms a carbonyl when attached to carbon, an N-oxide when attached to nitrogen, and a sulfoxide or sulfone when attached to sulfur.

The term “acyl” refers to an alkylcarbonyl, cycloalkylcarbonyl, arylcarbonyl, heterocyclylcarbonyl, or heteroarylcarbonyl substituent, any of which may be further substituted by substituents.

The term “substituents” refers to a group “substituted” on an alkyl, cycloalkyl, alkenyl, alkynyl, heterocyclyl, heterocycloalkenyl, cycloalkenyl, aryl, or heteroaryl group at any atom of that group. Suitable substituents include, without limitation, alkyl, alkenyl, alkynyl, alkoxy, halo, hydroxy, cyano, nitro, amino, SO_3H , sulfate, phosphate, perfluoroalkyl, perfluoroalkoxy, methylenedioxy, ethylenedioxy, carboxyl, oxo, thioxo, imino (alkyl, aryl, aralkyl), $\text{S}(\text{O})_n$ alkyl (where n is 0-2), $\text{S}(\text{O})_n$ aryl (where n is 0-2), $\text{S}(\text{O})_n$ heteroaryl (where n is 0-2), $\text{S}(\text{O})_n$ heterocyclyl (where n is 0-2), amine (mono-, di-, alkyl, cycloalkyl, aralkyl, heteroaralkyl, and combinations thereof), ester (alkyl, aralkyl, heteroaralkyl), amide (mono-, di-, alkyl, aralkyl, heteroaralkyl, and combinations thereof), sulfonamide (mono-, di-, alkyl, aralkyl, heteroaralkyl, and combinations thereof), unsubstituted aryl, unsubstituted heteroaryl, unsubstituted heterocyclyl, and unsubstituted cycloalkyl. In one aspect, the substituents on a group are independently any one single, or any subset of the aforementioned substituents.

The terms “adeninyl, cytosinyl, guaninyl, thyminyl, and uracil” and the like refer to radicals of adenine, cytosine, guanine, thymine, and uracil.

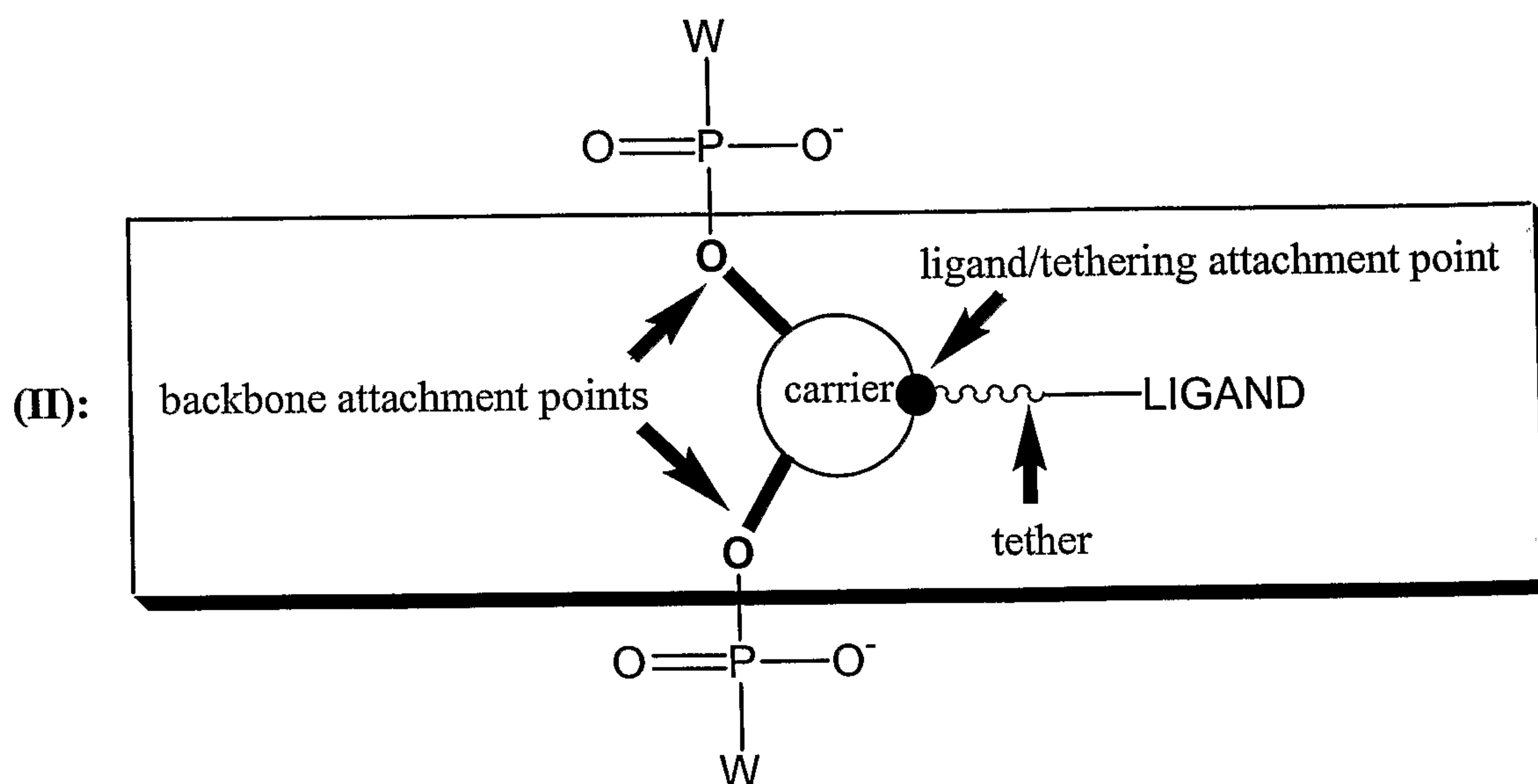
A “protected” moiety refers to a reactive functional group, *e.g.*, a hydroxyl group or an amino group, or a class of molecules, *e.g.*, sugars, having one or more functional groups, in which the reactivity of the functional group is temporarily blocked by the presence of an attached protecting group. Protecting groups useful for the monomers and methods described herein can be found, *e.g.*, in Greene, T.W., *Protective Groups in Organic Synthesis* (John Wiley and Sons: New York), 1981, which is hereby incorporated by reference.

General

An oligonucleotide agent, *e.g.*, a conjugated oligonucleotide agent, containing a preferred, but nonlimiting ligand-conjugated monomer subunit is presented as formula (II) below and in the scheme in FIG. 1. The carrier (also referred to in some embodiments as a “linker”) can be a cyclic or acyclic moiety and includes two “backbone attachment points” (*e.g.*, hydroxyl groups) and a ligand. The ligand can be directly attached (*e.g.*, conjugated) to the carrier or indirectly attached (*e.g.*, conjugated) to the carrier by an intervening tether (*e.g.*, an acyclic chain

of one or more atoms; or a nucleobase, *e.g.*, a naturally occurring nucleobase optionally having one or more chemical modifications, *e.g.*, an unusual base; or a universal base). The carrier therefore also includes a “ligand or tethering attachment point” for the ligand and tether/tethered ligand, respectively.

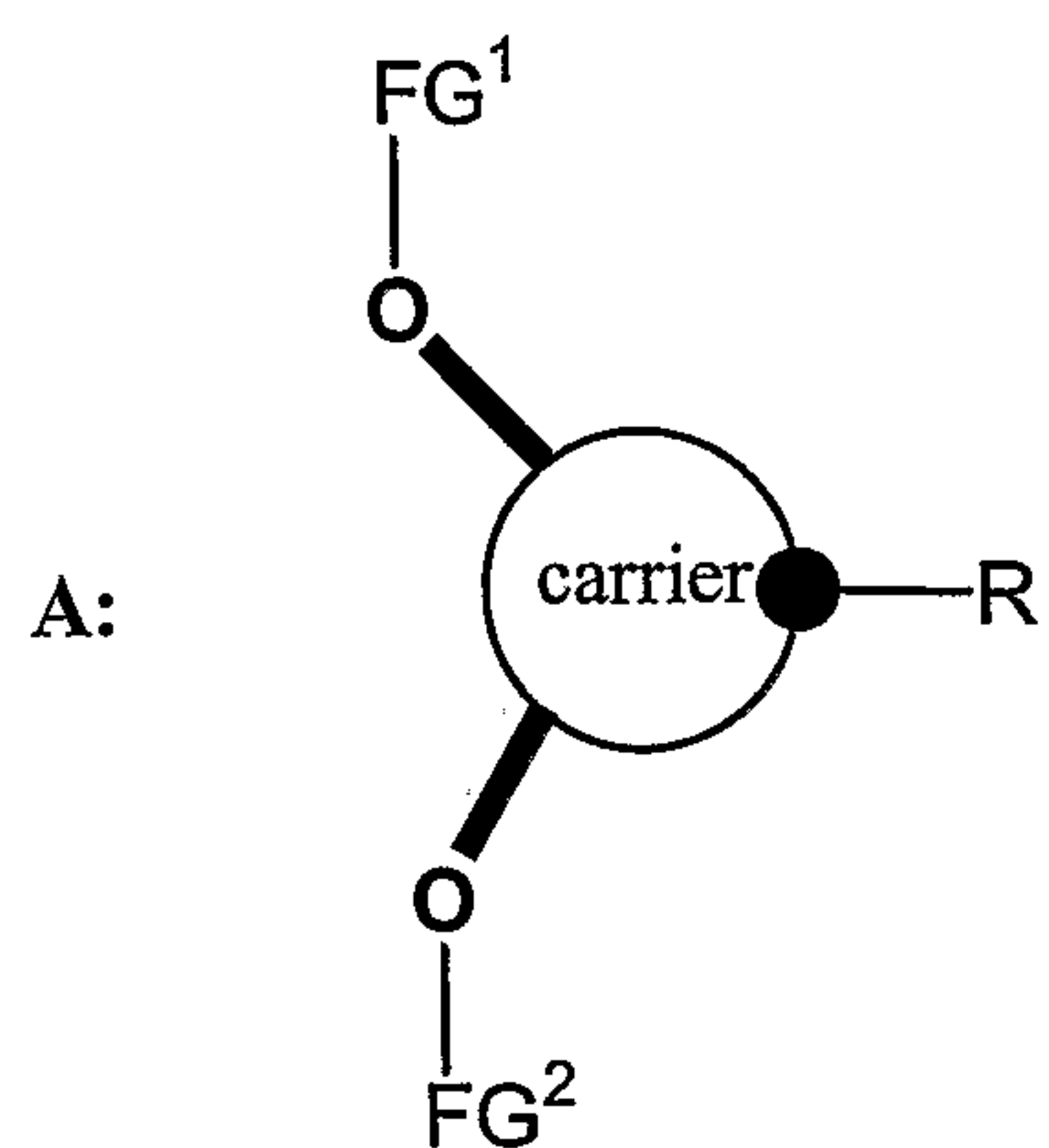
5 The ligand-conjugated monomer subunit may be the 5' or 3' terminal subunit of the RNA molecule, *i.e.*, one of the two “W” groups may be a hydroxyl group, and the other “W” group may be a chain of two or more unmodified or modified ribonucleotides. Alternatively, the ligand-conjugated monomer subunit may occupy an internal position, and both “W” groups may be one or more unmodified or modified ribonucleotides. More than one ligand-conjugated
 10 monomer subunit may be present in a RNA molecule, *e.g.*, an oligonucleotide agent. Preferred positions for inclusion of a tethered ligand-conjugated monomer subunit, *e.g.*, one in which a lipophilic moiety, *e.g.*, cholesterol, is tethered to the carrier are at the 3' terminus, the 5' terminus, or at an internal position.



15

The modified RNA molecule of formula (II) can be obtained using oligonucleotide synthetic methods known in the art. In a preferred embodiment, the modified RNA molecule of formula (II) can be prepared by incorporating one or more of the corresponding monomer compounds (see, *e.g.*, A, B, and C below and in the scheme in FIG. 1) into a growing strand,
 20 utilizing, *e.g.*, phosphoramidite or H-phosphonate coupling strategies.

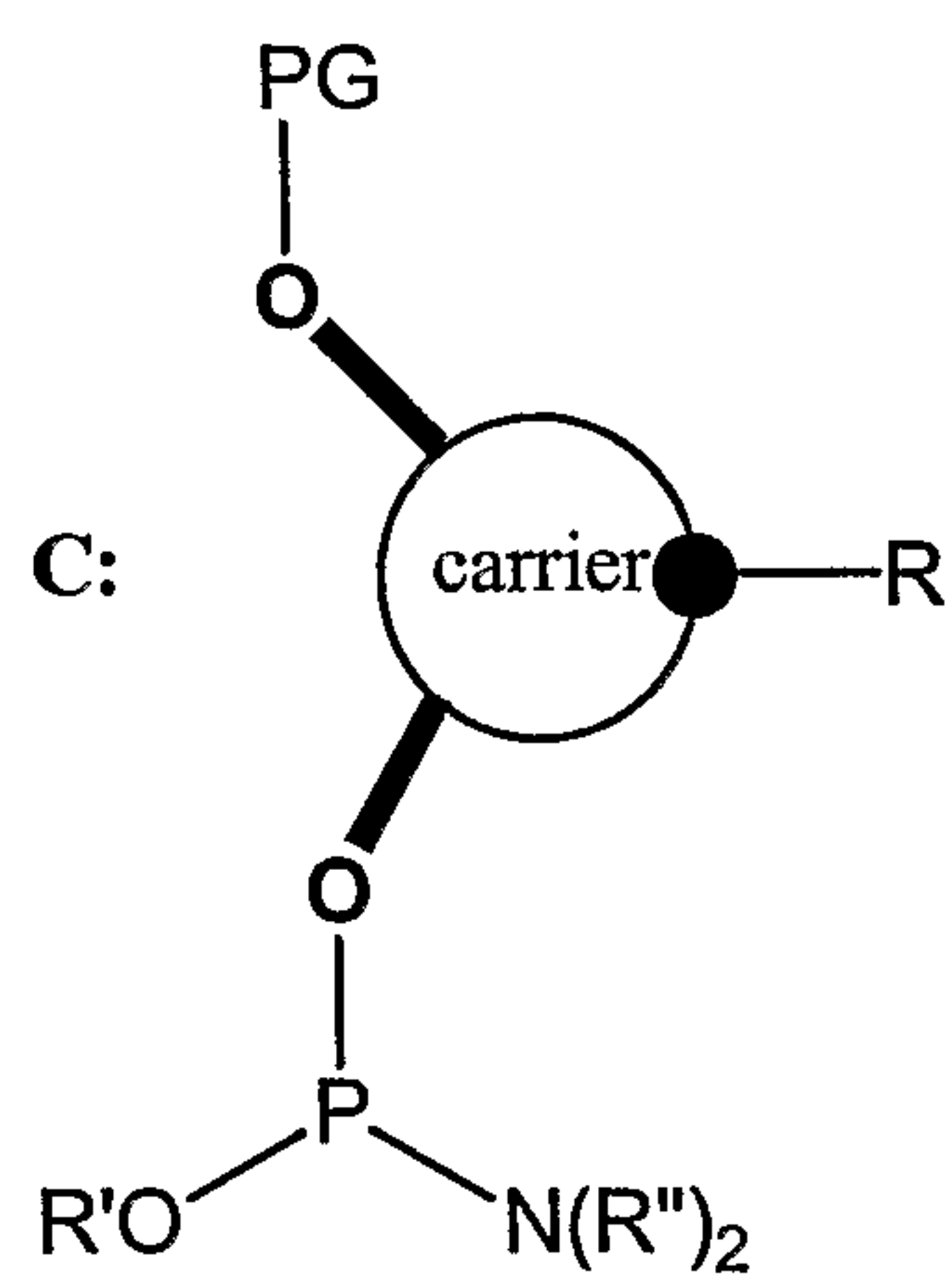
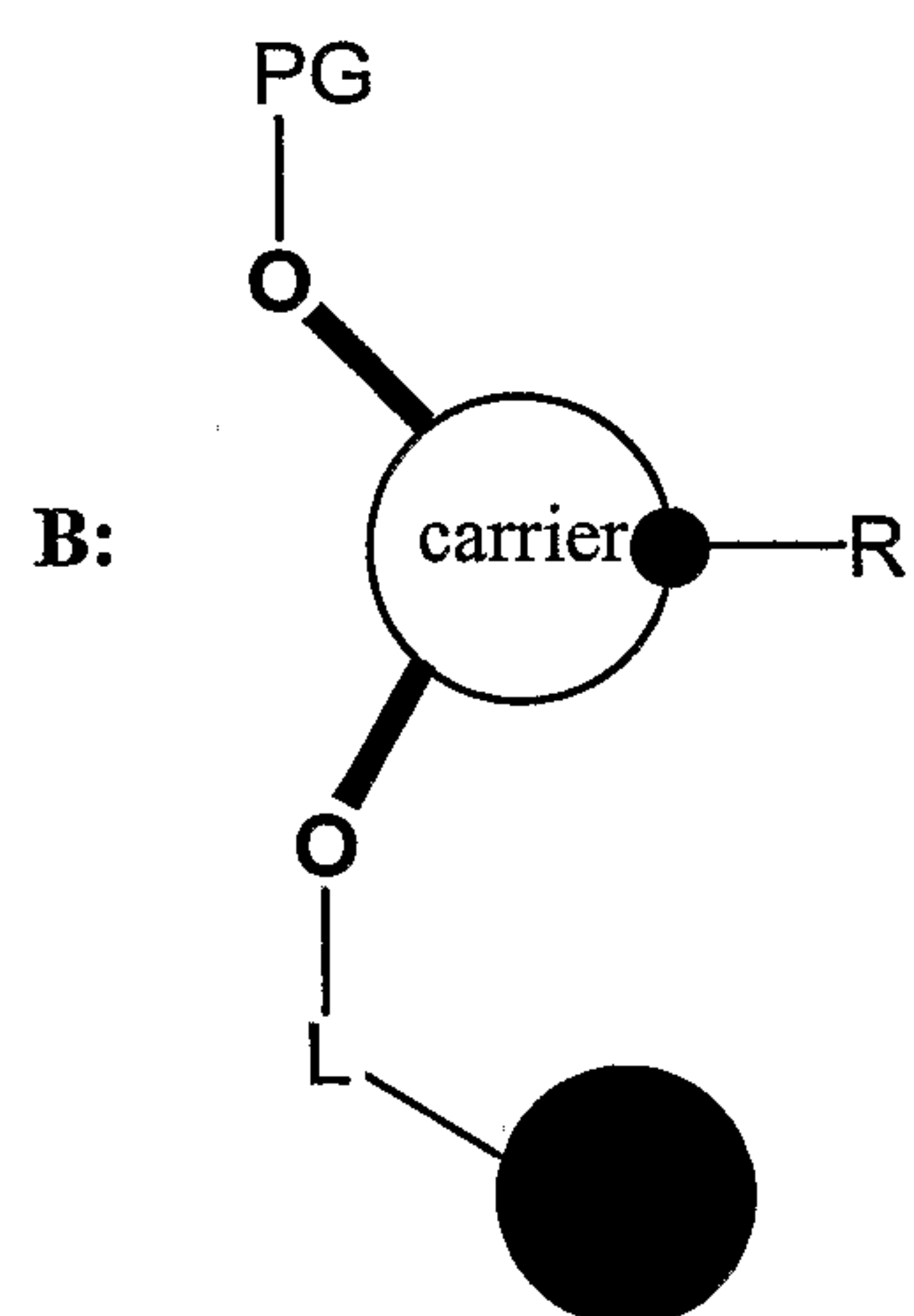
The monomers, *e.g.*, a ligand-conjugated monomers, generally include two differently functionalized hydroxyl groups (OFG¹ and OFG²), which are linked to the carrier molecule (see A below and in FIG. 1), and a ligand/tethering attachment point. As used herein, the term “functionalized hydroxyl group” means that the hydroxyl proton has been replaced by another substituent. As shown in representative structures B and C below and in FIG. 1, one hydroxyl group (OFG¹) on the carrier is functionalized with a protecting group (PG). The other hydroxyl group (OFG²) can be functionalized with either (1) a liquid or solid phase synthesis support reagent (solid circle) directly or indirectly through a linker, L, as in B, or (2) a phosphorus-containing moiety, *e.g.*, a phosphoramidite as in C. The tethering attachment point may be connected to a hydrogen atom, a suitable protecting group, a tether, or a tethered ligand at the time that the monomer is incorporated into the growing strand (see variable “R” in A below). Thus, the tethered ligand can be, but need not be attached to the monomer at the time that the monomer is incorporated into the growing strand. In certain embodiments, the tether, the ligand or the tethered ligand may be linked to a “precursor” ligand-conjugated monomer subunit after a “precursor” ligand-conjugated monomer subunit has been incorporated into the strand. The wavy line used below (and elsewhere herein) refers to a connection, and can represent a direct bond between the moiety and the attachment point or a tethering molecule which is interposed between the moiety and the attachment point. Directly tethered means the moiety is bound directly to the attachment point. Indirectly tethered means that there is a tether molecule interposed between the attachment point and the moiety.



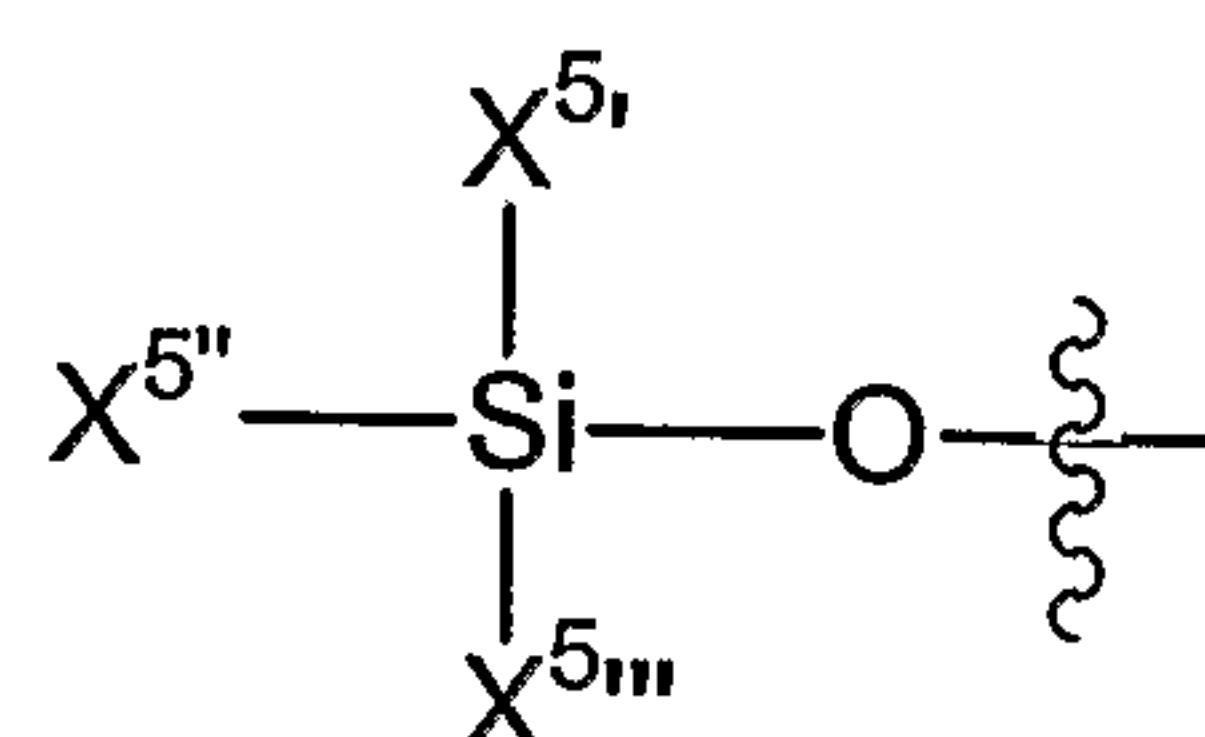
R = H, protecting group;

R = ()

R = () + (LIGAND)



The (OFG¹) protecting group may be selected as desired, *e.g.*, from T.W. Greene and P.G.M. Wuts, *Protective Groups in Organic Synthesis*, 2d. Ed., John Wiley and Sons (1991). The protecting group is preferably stable under amidite synthesis conditions, storage conditions, and oligonucleotide synthesis conditions. Hydroxyl groups, -OH, are nucleophilic groups (*i.e.*, Lewis bases), which react through the oxygen with electrophiles (*i.e.*, Lewis acids). Hydroxyl groups in which the hydrogen has been replaced with a protecting group, *e.g.*, a triarylmethyl group or a trialkylsilyl group, are essentially unreactive as nucleophiles in displacement reactions. Thus, the protected hydroxyl group is useful in preventing *e.g.*, homocoupling of compounds exemplified by structure C during oligonucleotide synthesis. In some embodiments, a preferred protecting group is the dimethoxytrityl group. In other embodiments, a preferred protecting group is a silicon-based protecting group having the formula below:



X^{5'}, X^{5''}, and X^{5'''} can be selected from substituted or unsubstituted alkyl, cycloalkyl, aryl, arakyl, heteroaryl, alkoxy, cycloalkoxy, aralkoxy, aryloxy, heteroaryloxy, or siloxy (*i.e.*, R₃SiO-, the three "R" groups can be any combination of the above listed groups). X^{5'}, X^{5''}, and X^{5'''} may all be the same or different; also contemplated is a combination in which two of X^{5'}, X^{5''}, and X^{5'''} are identical and the third is different. In certain embodiments X^{5'}, X^{5''}, and X^{5'''} include at least one alkoxy or siloxy groups and may be any one of the groups listed in FIG. 2A, a preferred combination includes X^{5'}, X^{5''} = trimethylsiloxy and X^{5'''} = 1, 3-(triphenylmethoxy)-2-propoxy or cyclododecyloxy.

Other preferred combinations of X^{5'}, X^{5''}, and X^{5'''} include those that result in OFG¹ groups that meet the deprotection and stability criteria delineated below. The group is preferably stable under amidite synthesis conditions, storage conditions, and oligonucleotide synthesis conditions. Rapid removal, *i.e.*, less than one minute, of the silyl group from *e.g.*, a support-bound oligonucleotide is desirable because it can reduce synthesis times and thereby reduce exposure time of the growing oligonucleotide chain to the reagents. Oligonucleotide synthesis

can be improved if the silyl protecting group is visible during deprotection, *e.g.*, from the addition of a chromophore silyl substituent.

Selection of silyl protecting groups can be complicated by the competing demands of the essential characteristics of stability and facile removal, and the need to balance these competitive goals. Most substituents that increase stability can also increase the reaction time required for removal of the silyl group, potentially increasing the level of difficulty in removal of the group.

The addition of alkoxy and siloxy substituents to OFG¹ silicon-containing protecting groups increases the susceptibility of the protecting groups to fluoride cleavage of the silylether bonds. Increasing the steric bulk of the substituents preserves stability while not decreasing fluoride lability to an equal extent. An appropriate balance of substituents on the silyl group makes a silyl ether a viable nucleoside protecting group.

Candidate OFG¹ silicon-containing protecting groups may be tested by exposing a tetrahydrofuran solution of a preferred carrier bearing the candidate OFG¹ group to five molar equivalents of tetrahydrofuran at room temperature. The reaction time may be determined by monitoring the disappearance of the starting material by thin layer chromatography.

When the OFG² in **B** includes a linker, *e.g.*, a relatively long organic linker, connected to a soluble or insoluble support reagent, solution or solid phase synthesis techniques can be employed to build up a chain of natural and/or modified ribonucleotides once OFG¹ is deprotected and free to react as a nucleophile with another nucleoside or monomer containing an electrophilic group (*e.g.*, an amidite group). Alternatively, a natural or modified ribonucleotide or oligoribonucleotide chain can be coupled to monomer **C** *via* an amidite group or H-phosphonate group at OFG². Subsequent to this operation, OFG¹ can be deblocked, and the restored nucleophilic hydroxyl group can react with another nucleoside or monomer containing an electrophilic group. R' can be substituted or unsubstituted alkyl or alkenyl. In preferred embodiments, R' is methyl, allyl or 2-cyanoethyl. R'' may a C₁-C₁₀ alkyl group, preferably it is a branched group containing three or more carbons, *e.g.*, isopropyl.

OFG² in **B** can be hydroxyl functionalized with a linker, which in turn contains a liquid or solid phase synthesis support reagent at the other linker terminus. The support reagent can be any support medium that can support the monomers described herein. The monomer can be attached to an insoluble support via a linker, L, which allows the monomer (and the growing chain) to be solubilized in the solvent in which the support is placed. The solubilized, yet

immobilized, monomer can react with reagents in the surrounding solvent; unreacted reagents and soluble by-products can be readily washed away from the solid support to which the monomer or monomer-derived products is attached. Alternatively, the monomer can be attached to a soluble support moiety, *e.g.*, polyethylene glycol (PEG) and liquid phase synthesis techniques can be used to build up the chain. Linker and support medium selection is within skill of the art. Generally the linker may be $-C(O)(CH_2)_qC(O)-$, or $-C(O)(CH_2)_qS-$, in which q can be 0, 1, 2, 3, or 4; preferably, it is oxalyl, succinyl or thioglycolyl. Standard control pore glass solid phase synthesis supports can not be used in conjunction with fluoride labile 5' silyl protecting groups because the glass is degraded by fluoride with a significant reduction in the amount of full-length product. Fluoride-stable polystyrene based supports or PEG are preferred.

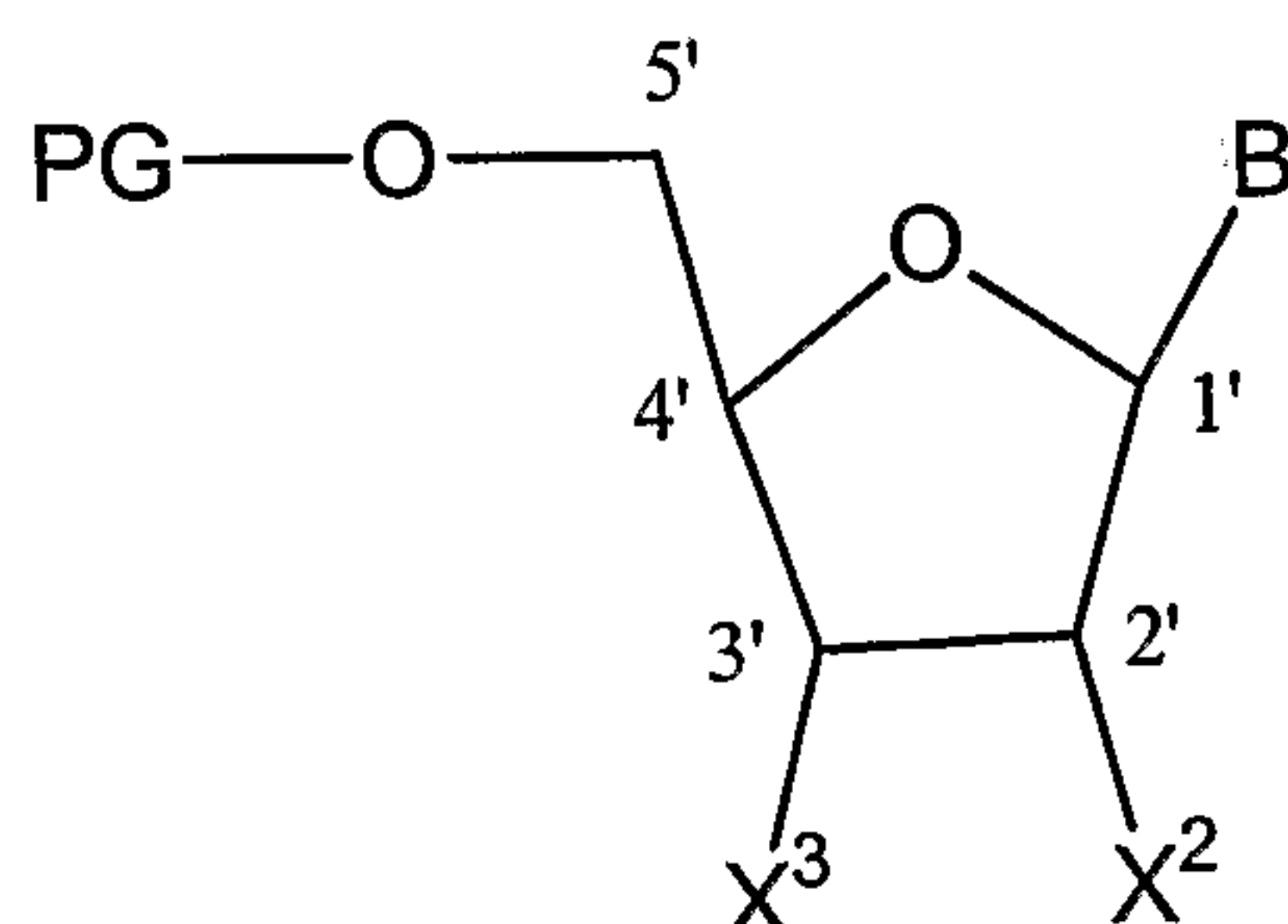
The ligand/tethering attachment point can be any divalent, trivalent, tetravalent, pentavalent or hexavalent atom. In some embodiments, ligand/tethering attachment point can be a carbon, oxygen, nitrogen or sulfur atom. For example, a ligand/tethering attachment point precursor functional group can have a nucleophilic heteroatom, *e.g.*, $-SH$, $-NH_2$, secondary amino, ONH_2 , or NH_2NH_2 . As another example, the ligand/tethering attachment point precursor functional group can be an olefin, *e.g.*, $-CH=CH_2$ or a Diels-Alder diene or dienophile and the precursor functional group can be attached to a ligand, a tether, or tethered ligand using, *e.g.*, transition metal catalyzed carbon-carbon (for example olefin metathesis) processes or cycloadditions (*e.g.*, Diels-Alder). As a further example, the ligand/tethering attachment point precursor functional group can be an electrophilic moiety, *e.g.*, an aldehyde. When the carrier is a cyclic carrier, the ligand/tethering attachment point can be an endocyclic atom (*i.e.*, a constituent atom in the cyclic moiety, *e.g.*, a nitrogen atom) or an exocyclic atom (*i.e.*, an atom or group of atoms attached to a constituent atom in the cyclic moiety).

The carrier can be any organic molecule containing attachment points for OFG^1 , OFG^2 , and the ligand. In certain embodiments, carrier is a cyclic molecule and may contain heteroatoms (*e.g.*, O, N or S). *E.g.*, carrier molecules may include aryl (*e.g.*, benzene, biphenyl, etc.), cycloalkyl (*e.g.*, cyclohexane, *cis* or *trans* decalin, etc.), or heterocyclyl (piperazine, pyrrolidine, etc.). In other embodiments, the carrier can be an acyclic moiety, *e.g.*, based on serinol. Any of the above cyclic systems may include substituents in addition to OFG^1 , OFG^2 , and the ligand.

Sugar-Based Monomers

In some embodiments, the carrier molecule is an oxygen containing heterocycle. Preferably the carrier is a ribose sugar as shown in structure **LCM-I**. In this embodiment, the ligand-conjugated monomer is a nucleoside.

5



LCM-I

“B” represents a nucleobase, *e.g.*, a naturally occurring nucleobase optionally having one or more chemical modifications, *e.g.*, and unusual base; or a universal base.

As used herein, an “unusual” nucleobase can include any one of the following:

10

2-methyladeninyl,

N6-methyladeninyl,

2-methylthio-N6-methyladeninyl,

N6-isopentenyladeninyl,

2-methylthio-N6-isopentenyladeninyl,

15

N6-(*cis*-hydroxyisopentenyl)adeninyl,

2-methylthio-N6-(*cis*-hydroxyisopentenyl) adeninyl,

N6-glycinylcarbamoyladeninyl,

N6-threonylcarbamoyladeninyl,

2-methylthio-N6-threonyl carbamoyladeninyl,

20

N6-methyl-N6-threonylcarbamoyladeninyl,

N6-hydroxynorvalylcarbamoyladeninyl,

2-methylthio-N6-hydroxynorvalyl carbamoyladeninyl,

N6,N6-dimethyladeninyl,

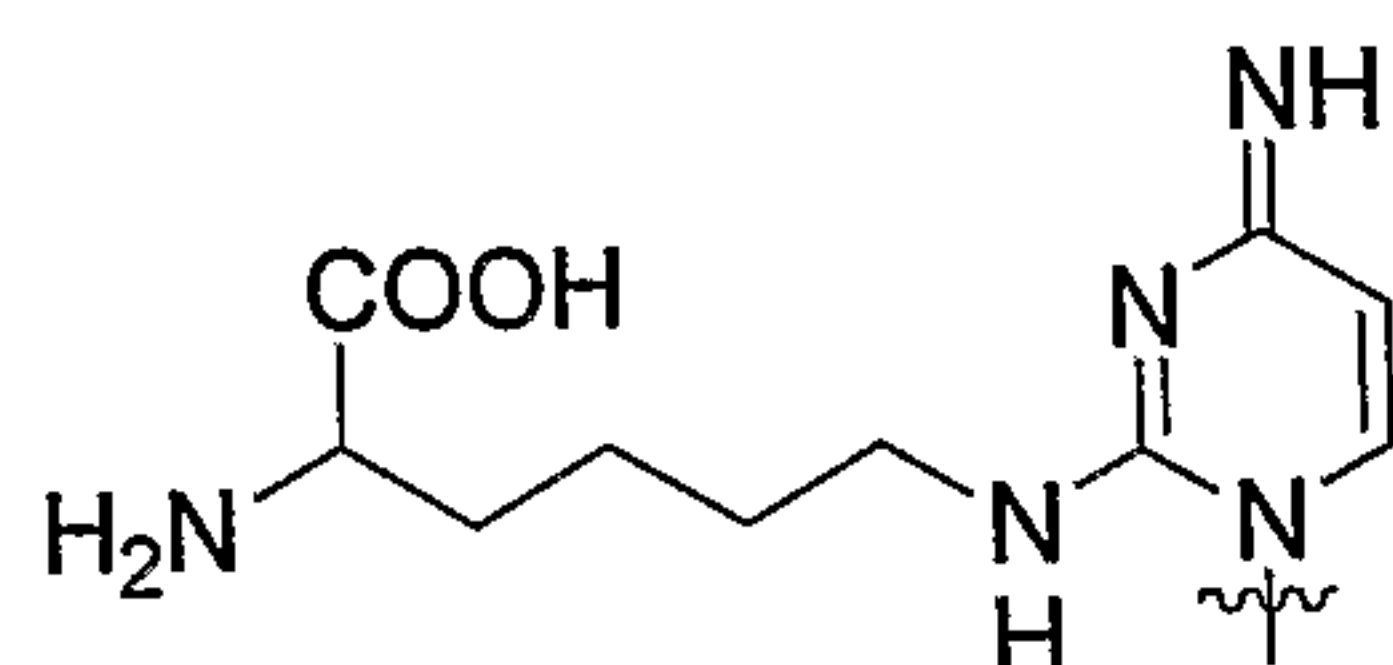
3-methylcytosinyl,

5-methylcytosinyl,

2-thiocytosinyl,

5-formylcytosinyl,

5



,

N4-methylcytosinyl,

10

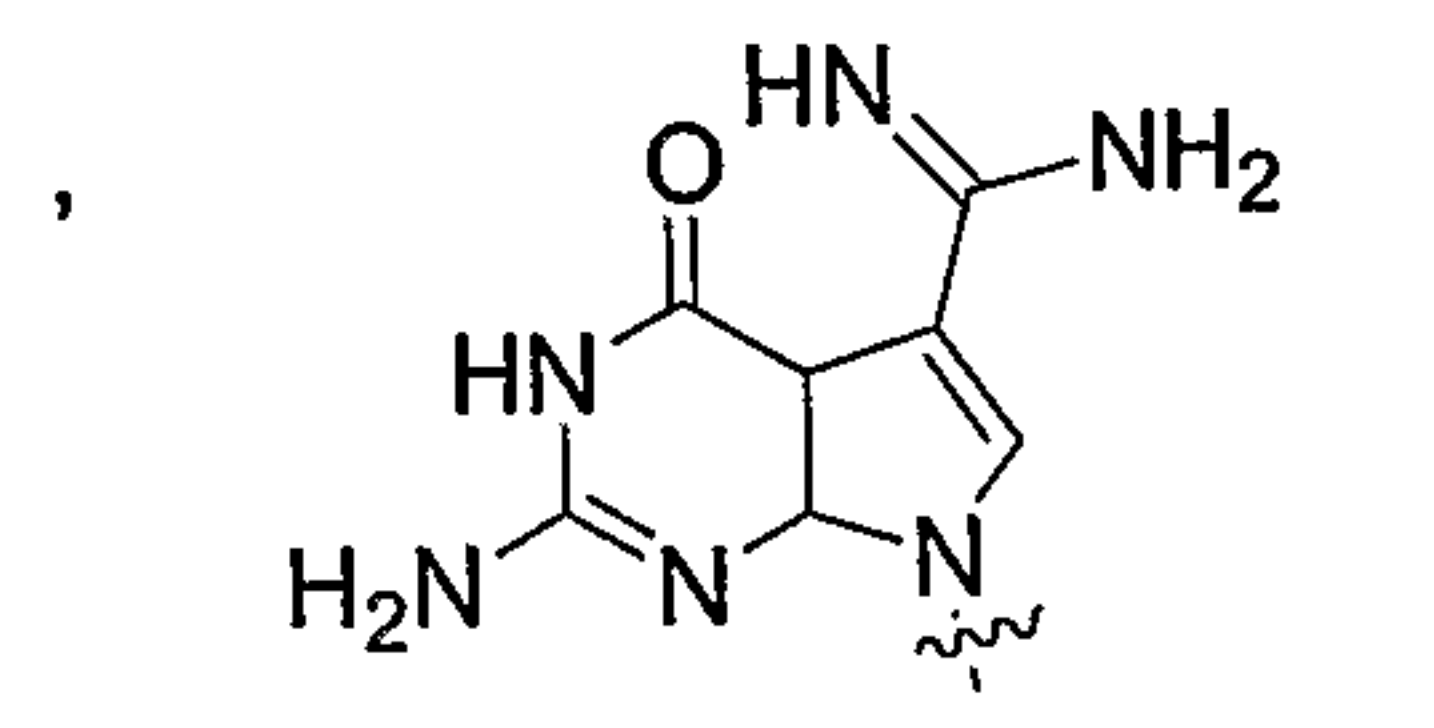
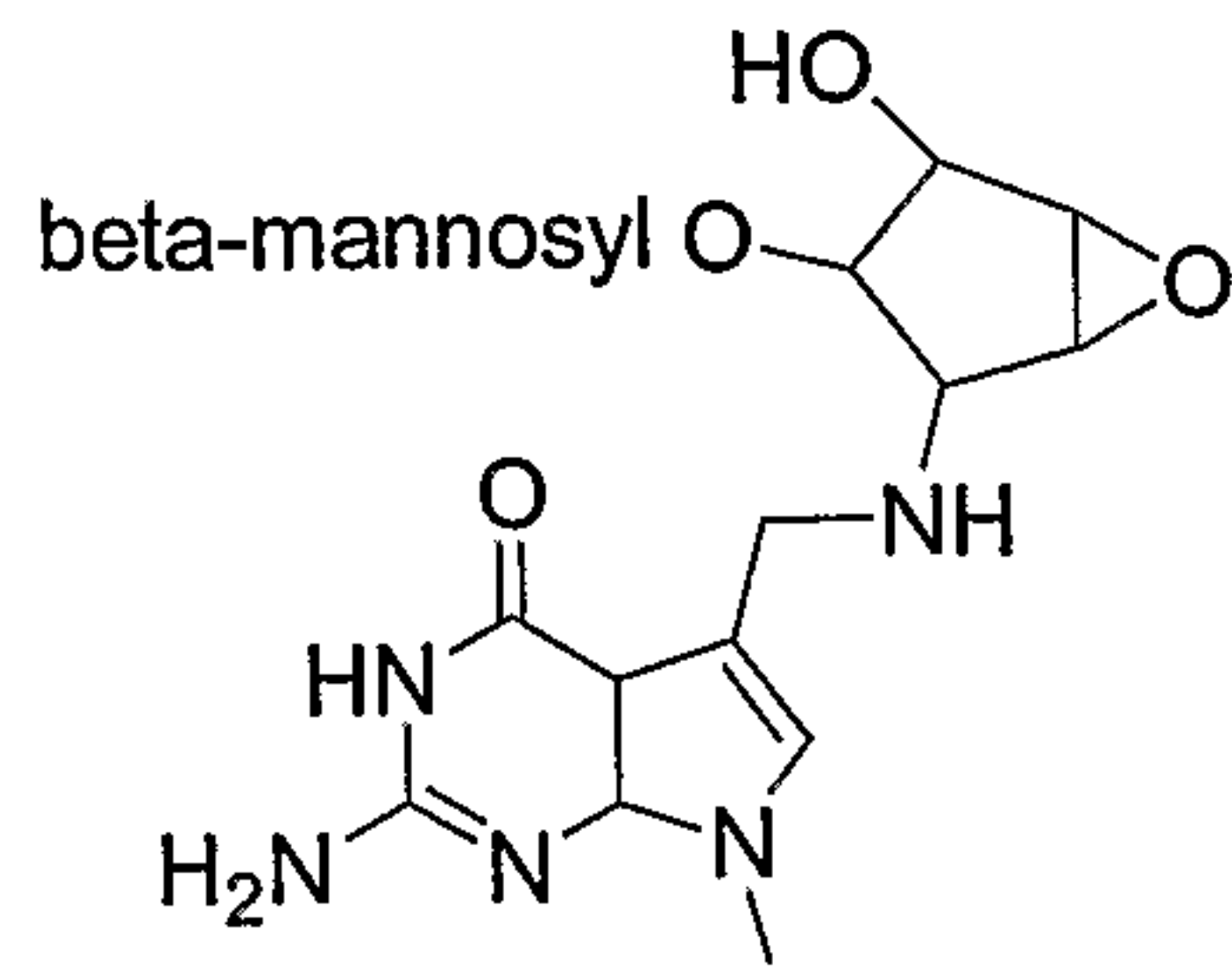
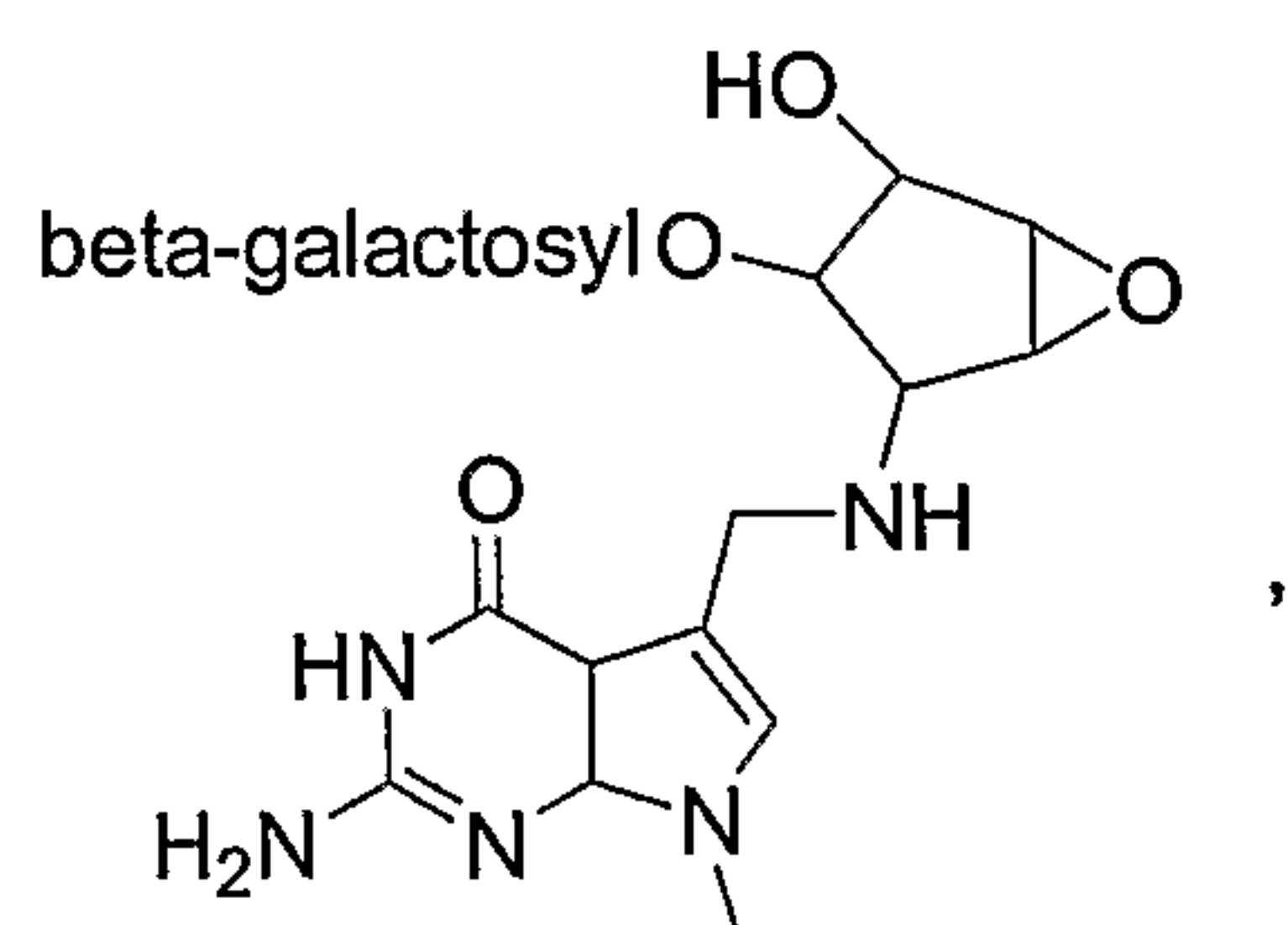
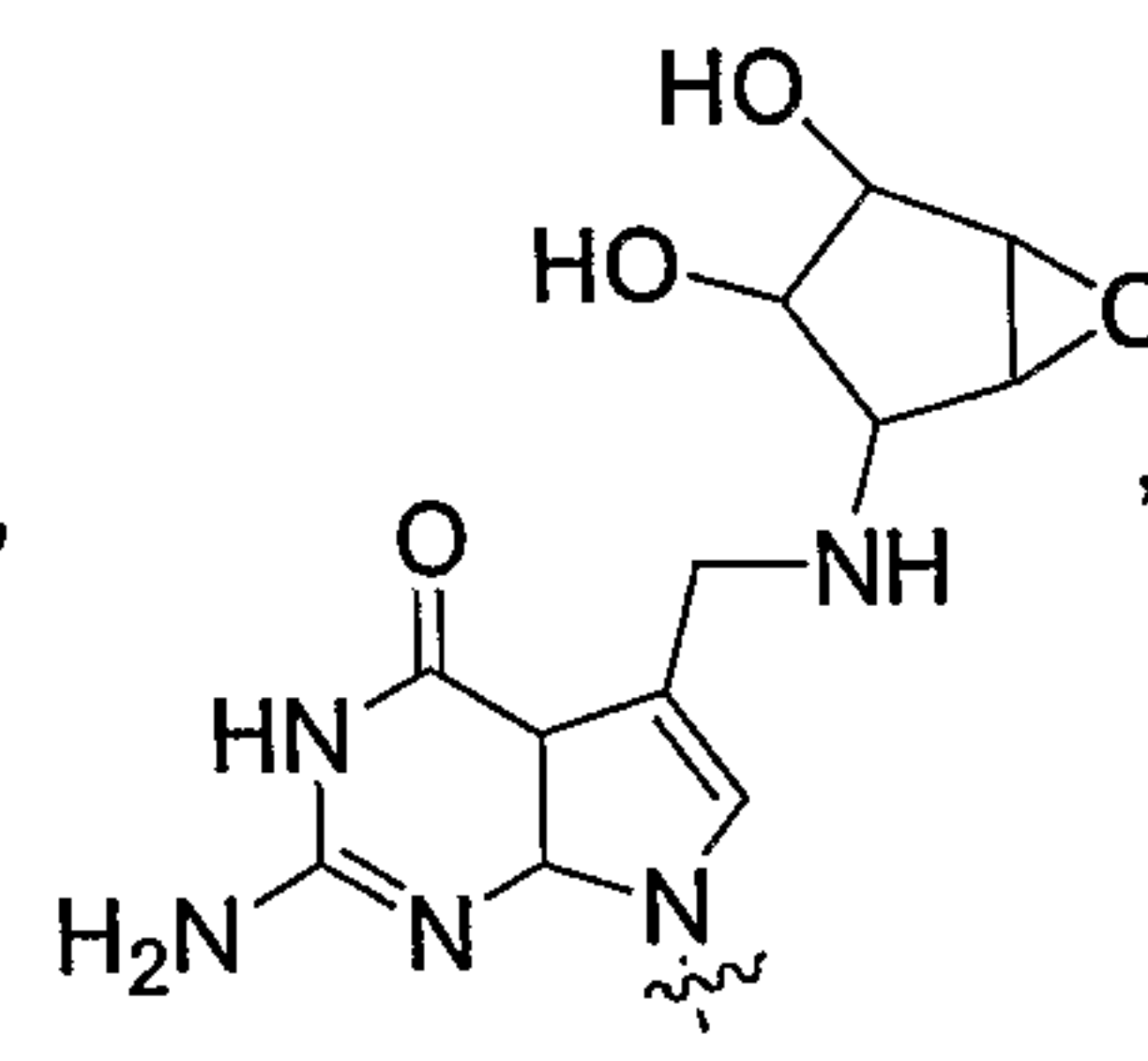
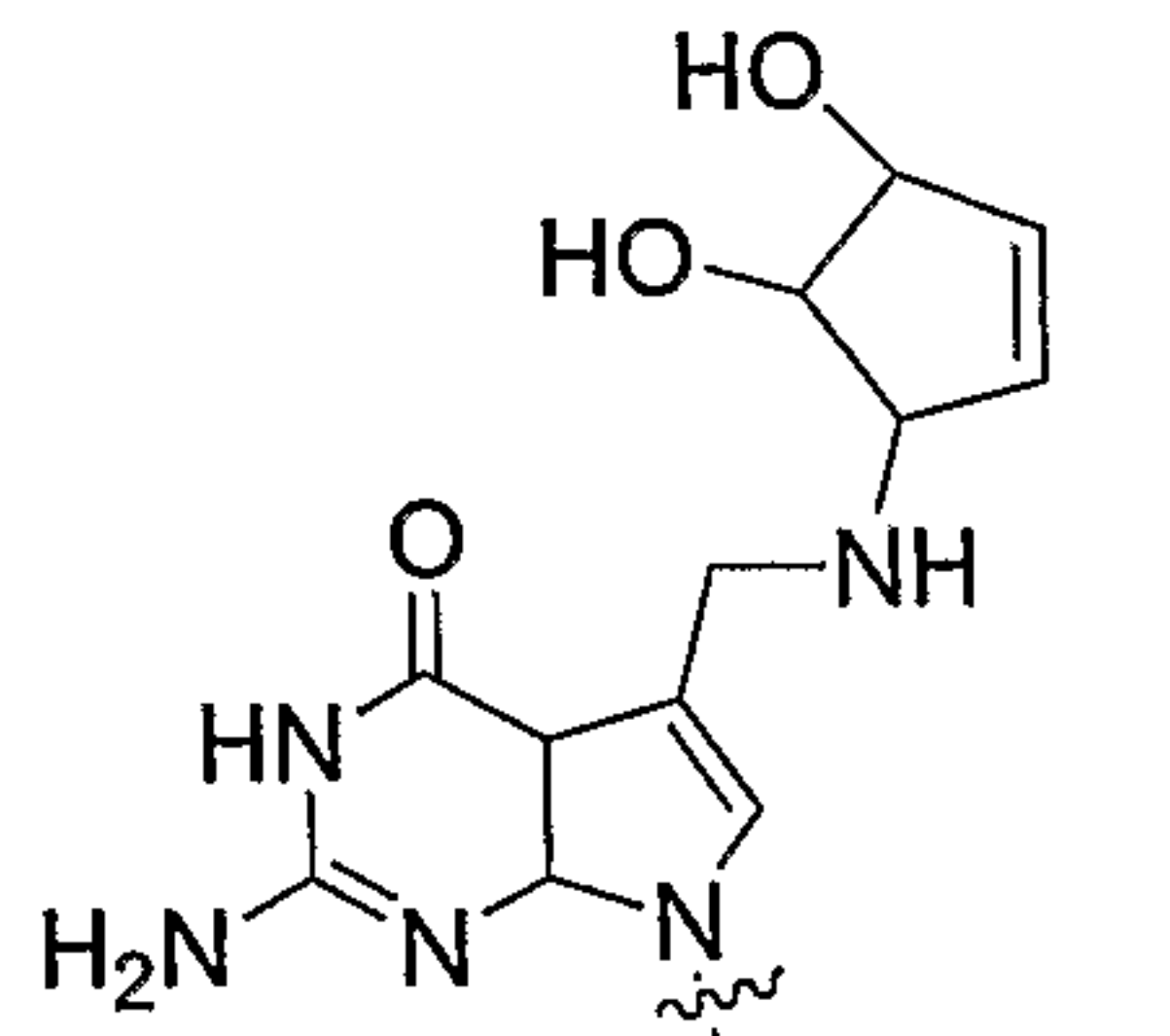
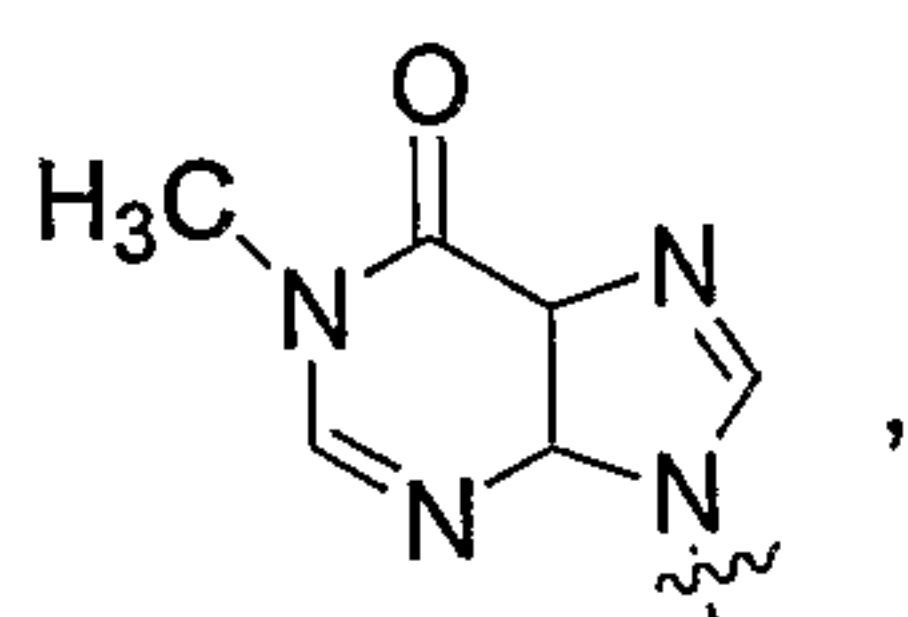
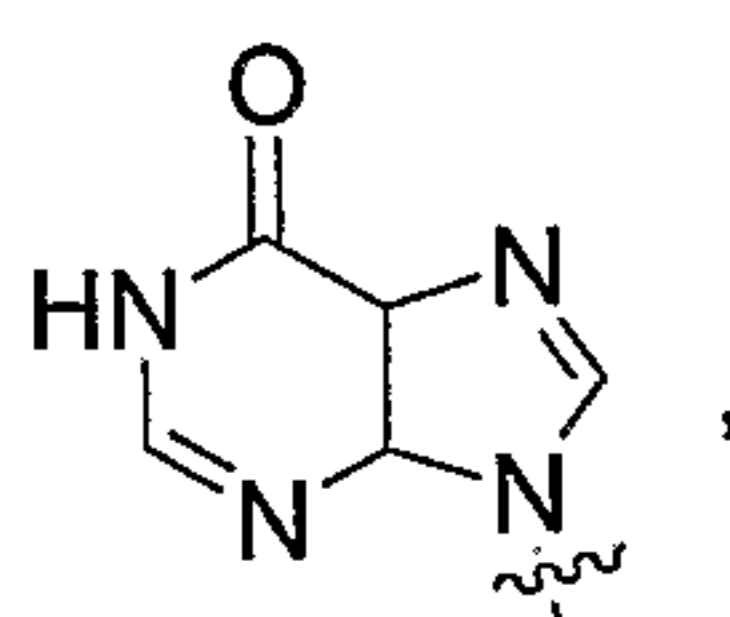
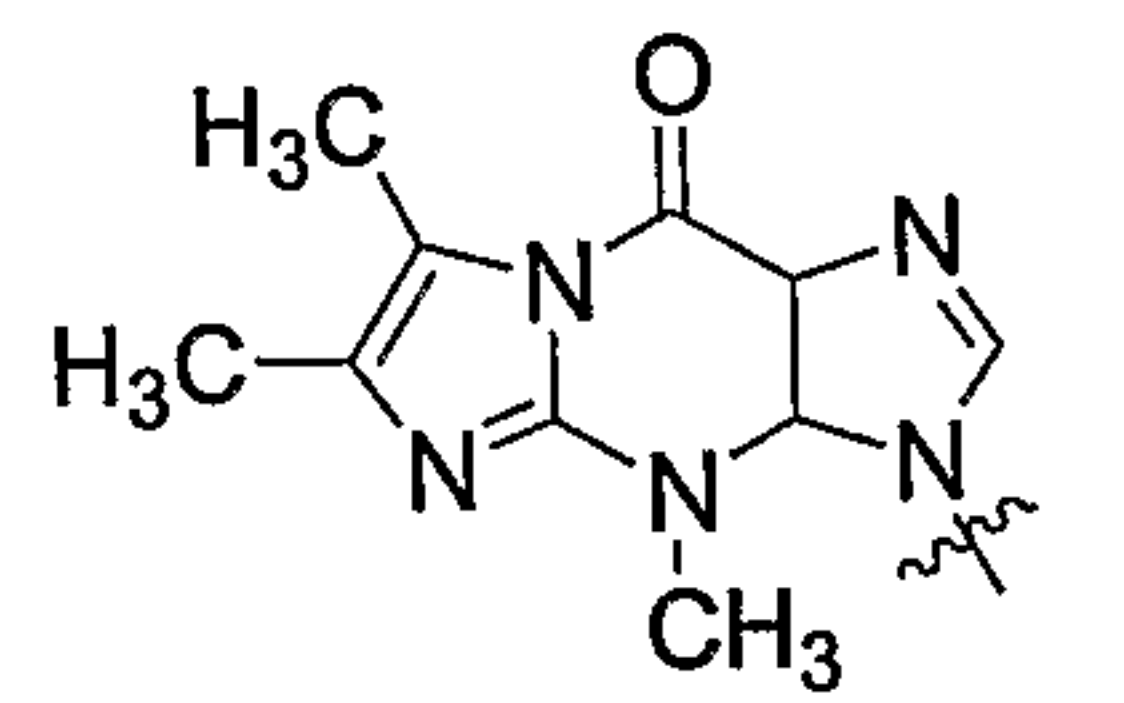
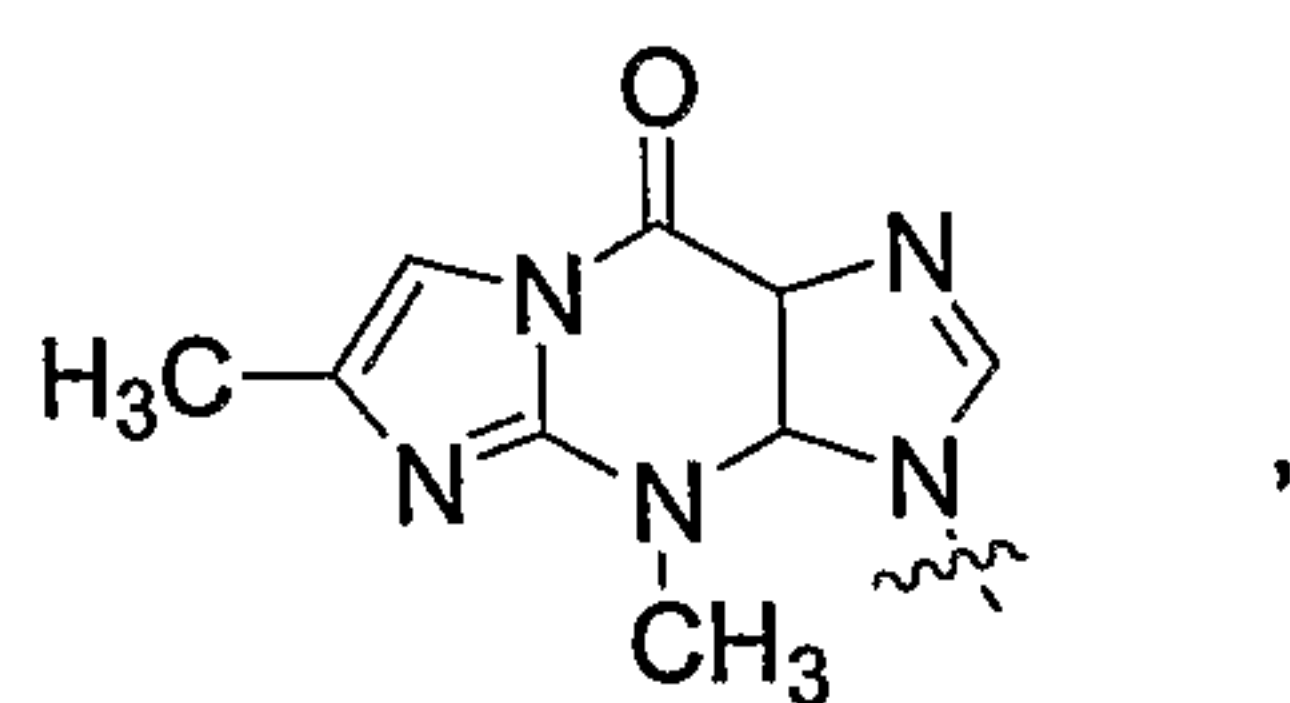
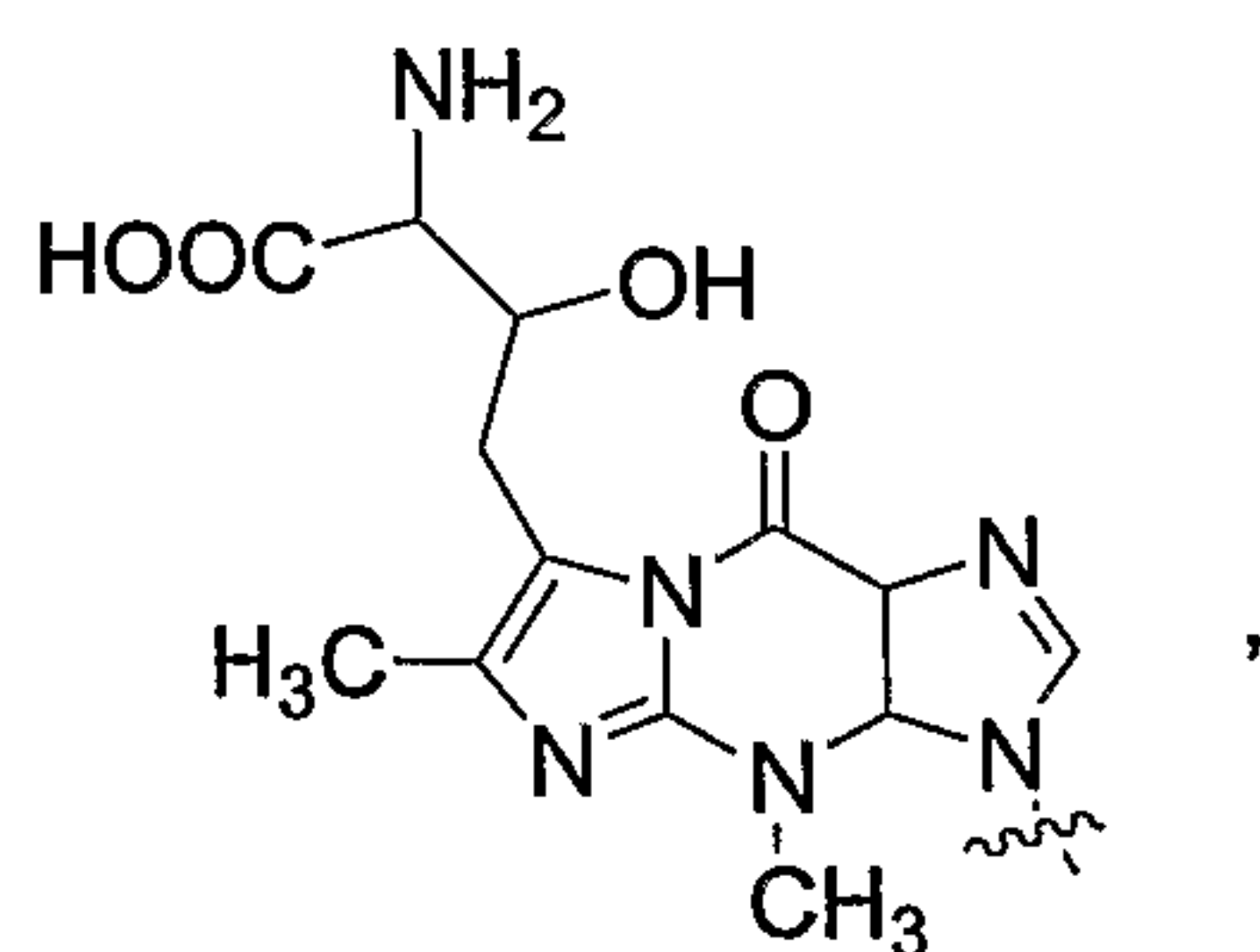
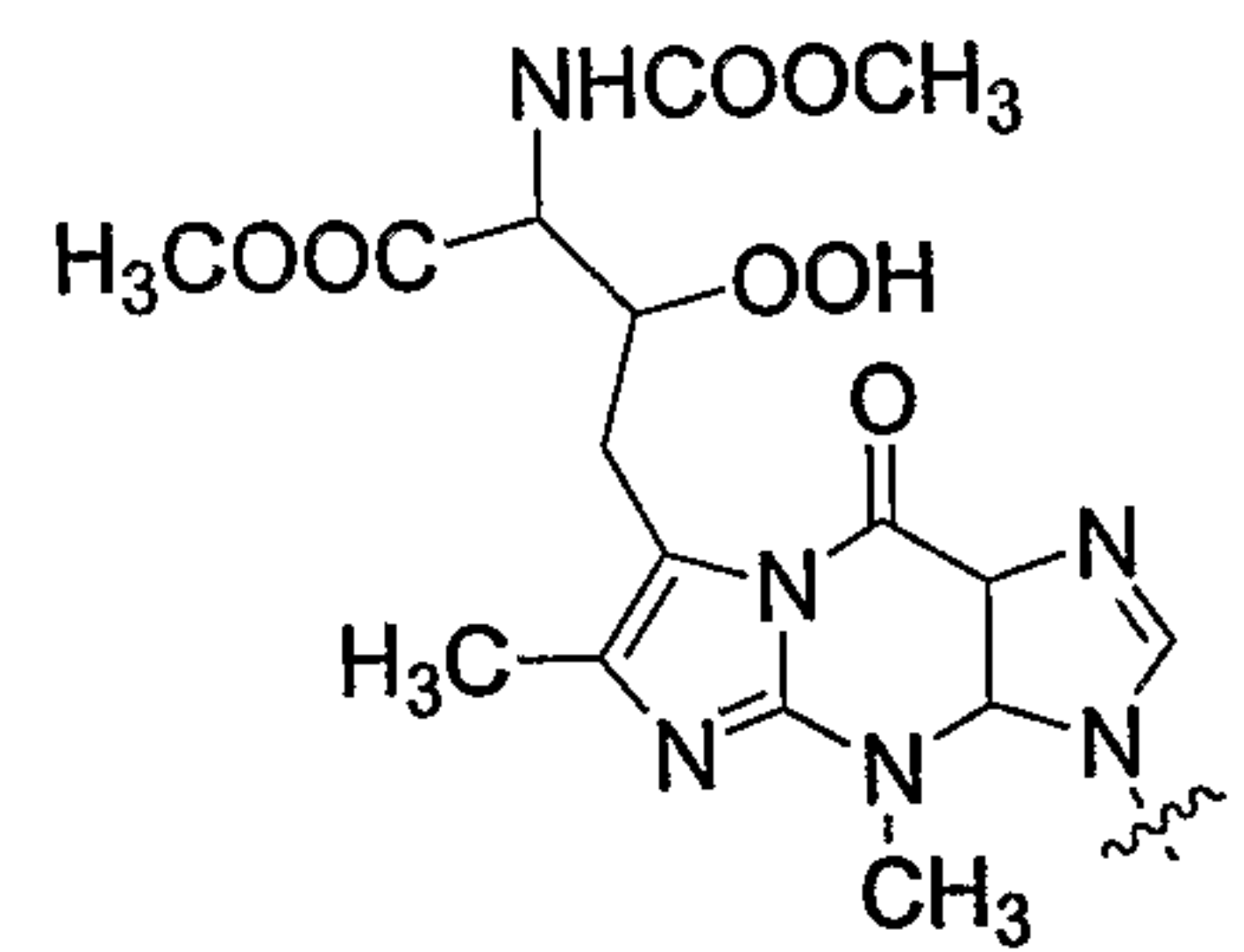
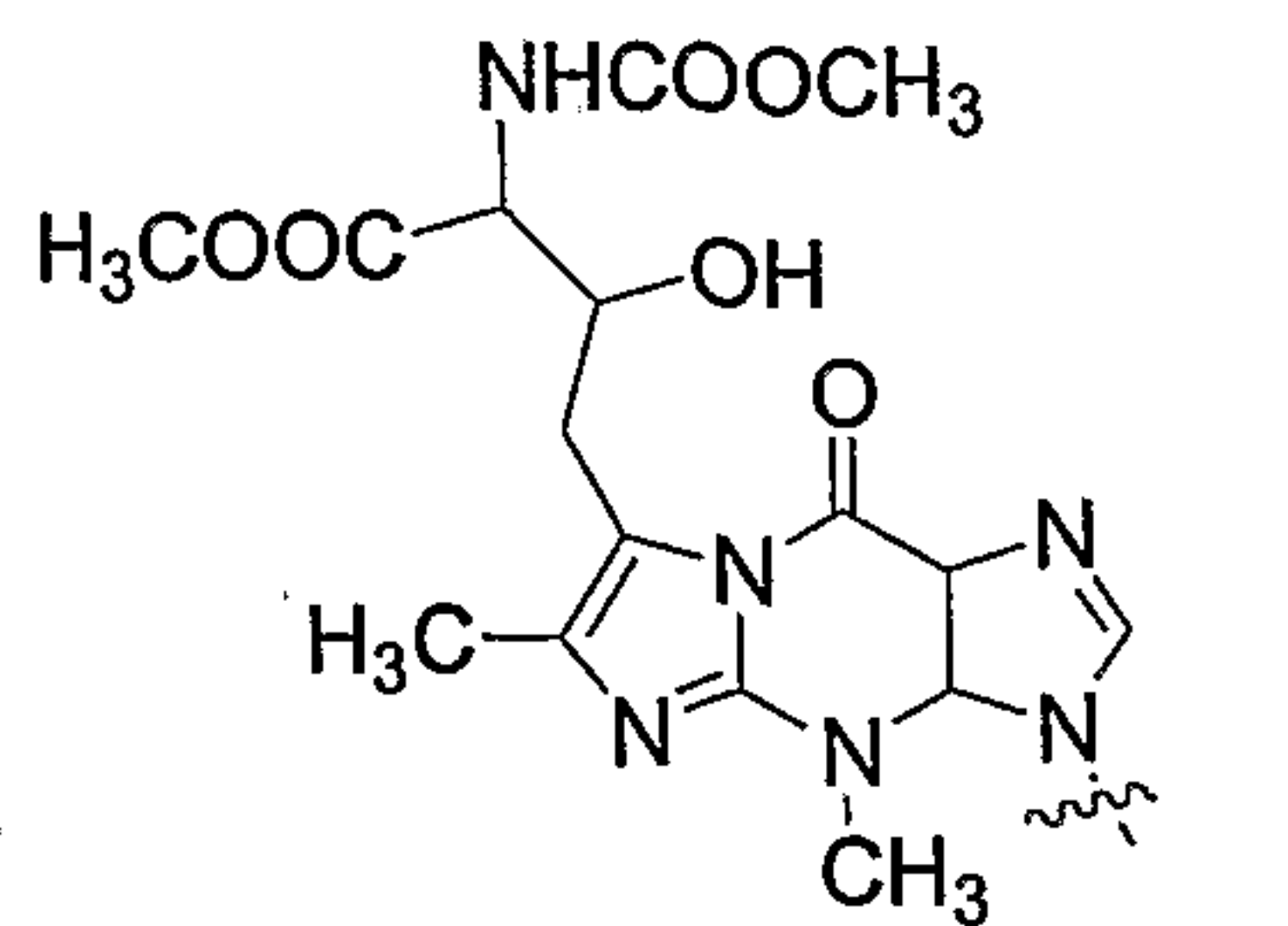
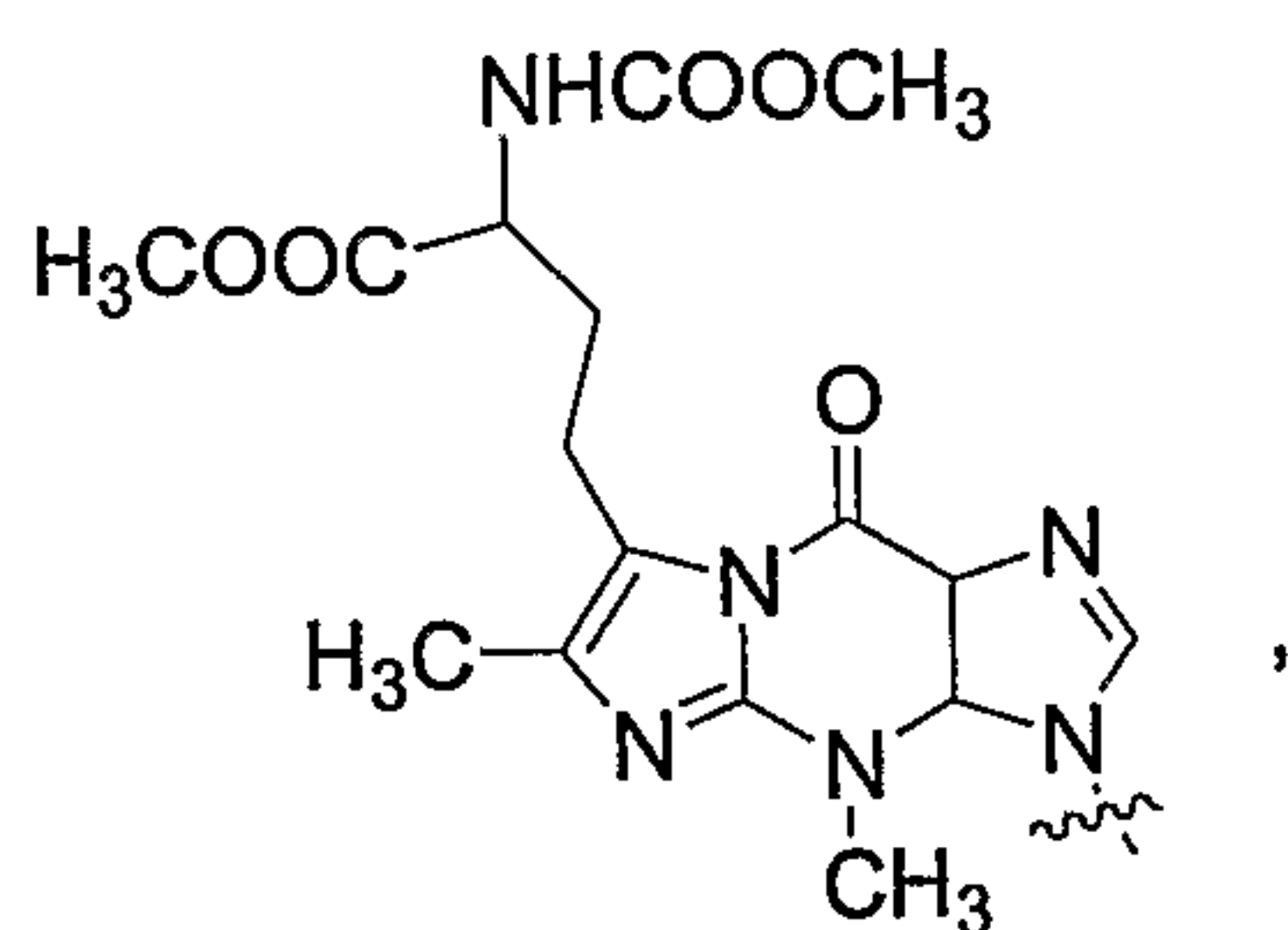
5-hydroxymethylcytosinyl,

1-methylguaninyl,

N2-methylguaninyl,

7-methylguaninyl,

N2,N2-dimethylguaninyl,



N2,7-dimethylguaninyl,
N2,N2,7-trimethylguaninyl,
1-methylguaninyl,
5 7-cyano-7-deazaguaninyl,
7-aminomethyl-7-deazaguaninyl,
pseudouracilyl,
dihydrouracilyl,
5-methyluracilyl,
10 1-methylpseudouracilyl,
2-thiouracilyl,
4-thiouracilyl,
2-thiothyminyl
5-methyl-2-thiouracilyl,
15 3-(3-amino-3-carboxypropyl)uracilyl,
5-hydroxyuracilyl,
5-methoxyuracilyl,
uracilyl 5-oxyacetic acid,
uracilyl 5-oxyacetic acid methyl ester,
20 5-(carboxyhydroxymethyl)uracilyl,
5-(carboxyhydroxymethyl)uracilyl methyl ester,
5-methoxycarbonylmethyluracilyl,
5-methoxycarbonylmethyl-2-thiouracilyl,
5-aminomethyl-2-thiouracilyl,
25 5-methylaminomethyluracilyl,
5-methylaminomethyl-2-thiouracilyl,
5-methylaminomethyl-2-selenouracilyl,
5-carbamoylmethyluracilyl,
5-carboxymethylaminomethyluracilyl,
30 5-carboxymethylaminomethyl-2-thiouracilyl,
3-methyluracilyl,

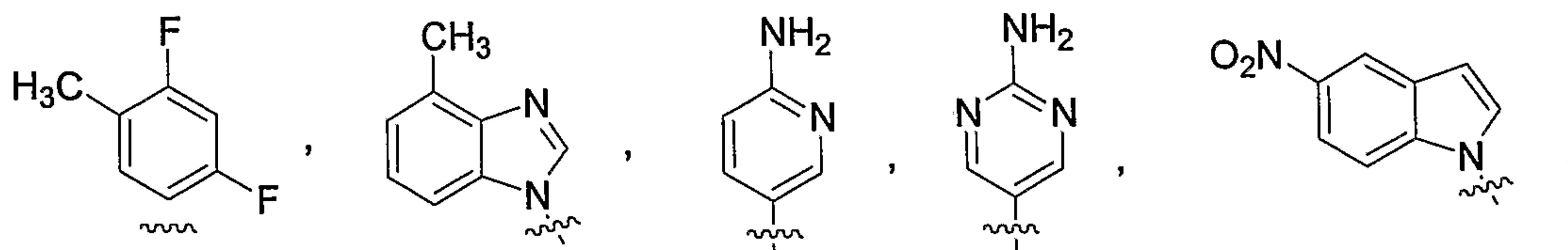
1-methyl-3-(3-amino-3-carboxypropyl) pseudouracilyl,
5-carboxymethyluracilyl,
5-methyldihydrouracilyl, or
3-methylpseudouracilyl.

5

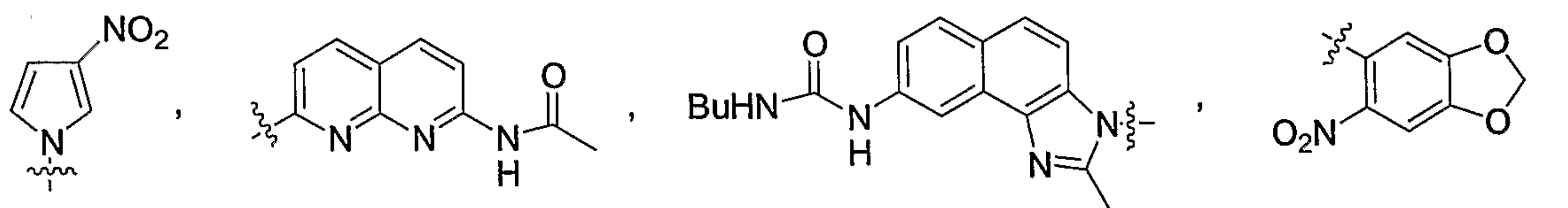
A universal base can form base pairs with each of the natural DNA/RNA bases, exhibiting relatively little discrimination between them. In general, the universal bases are non-hydrogen bonding, hydrophobic, aromatic moieties which can stabilize *e.g.*, duplex RNA or RNA-like molecules, via stacking interactions. A universal base can also include hydrogen

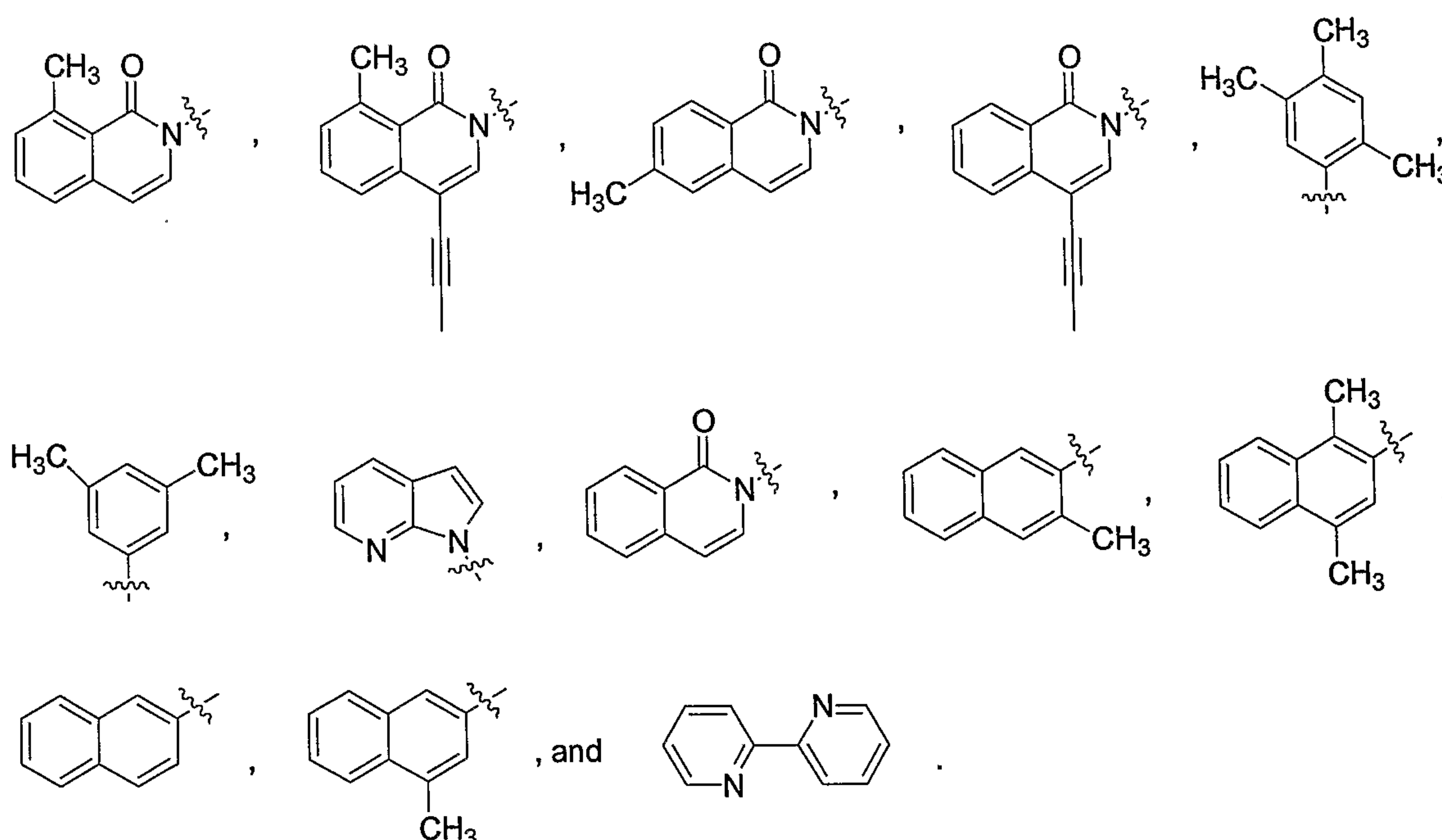
10 bonding substituents.

As used herein, a “universal base” can include anthracenes, pyrenes or any one of the following:



5





In some embodiments, B can form part of a tether that connects a ligand to the carrier. For example, the tether can be B-CH=CH-C(O)NH-(CH₂)₅-NHC(O)-LIGAND. In a preferred embodiment, the double bond is *trans*, and the ligand is a substituted or unsubstituted cholesteryl radical (*e.g.*, attached through the D-ring side chain or the C-3 hydroxyl); an aralkyl moiety having at least one stereogenic center and at least one substituent on the aryl portion of the aralkyl group; or a nucleobase. In certain embodiments, B, in the tether described above, is uracilyl or a universal base, *e.g.*, an aryl moiety, *e.g.*, phenyl, optionally having additional substituents, *e.g.*, one or more fluoro groups. B can be substituted at any atom with the remainder of the tether.

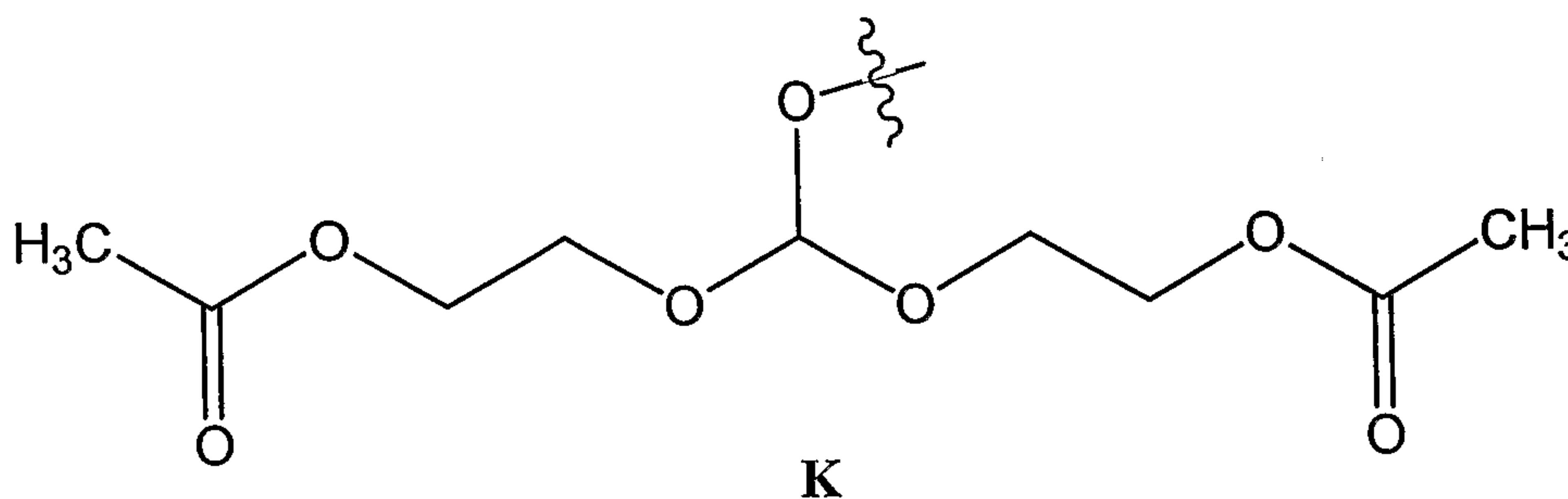
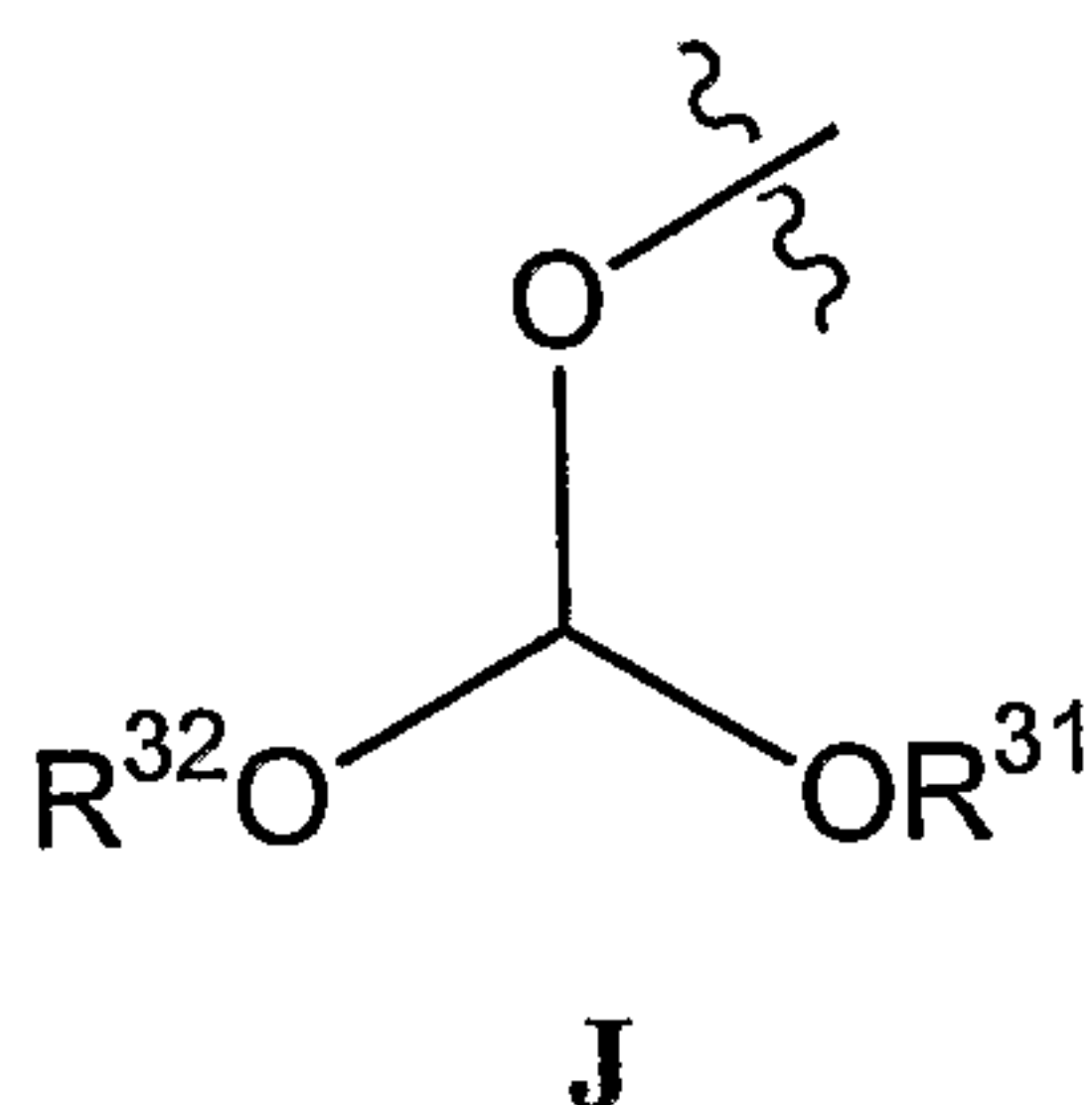
X² can include “oxy” or “deoxy” substituents in place of the 2’-OH or be a ligand or a tethered ligand.

Examples of “oxy”-substituents include alkoxy or aryloxy (OR, *e.g.*, R = H, alkyl, cycloalkyl, aryl, aralkyl, heteroaryl, sugar, or protecting group); polyethyleneglycols (PEG), O(CH₂CH₂O)_nCH₂CH₂OR; “locked” nucleic acids (LNA) in which the 2’ hydroxyl is connected, *e.g.*, by a methylene bridge, to the 4’ carbon of the same ribose sugar; O-PROTECTED AMINE (AMINE = NH₂; alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, or diheteroaryl amino, ethylene diamine, polyamino) and aminoalkoxy,

$O(CH_2)_n$ PROTECTED AMINE, (*e.g.*, AMINE = NH_2 ; alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, or diheteroaryl amino, ethylene diamine, polyamino), and orthoester. Amine protecting groups can include formyl, amido, benzyl, allyl, etc.

Preferred orthoesters have the general formula **J**. The groups R^{31} and R^{32} may be the same or different and can be any combination of the groups listed in FIG. 2B. A preferred orthoester is the “ACE” group, shown below as structure **K**.

1.



10

“Deoxy” substituents include hydrogen (*i.e.* deoxyribose sugars); halo (*e.g.*, fluoro); protected amino (*e.g.* NH_2 ; alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, diheteroaryl amino, or amino acid in which all amino are protected); fully protected polyamino (*e.g.*, $NH(CH_2CH_2NH)_nCH_2CH_2$ -AMINE, wherein AMINE = NH_2 ; alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, or diheteroaryl amino and all amino groups are protected), $-NHC(O)R$ (R = alkyl, cycloalkyl, aryl, aralkyl, heteroaryl or sugar), cyano; alkyl-thio-alkyl; thioalkoxy; and alkyl, cycloalkyl, aryl, alkenyl and alkynyl, which may be optionally substituted with *e.g.*, a protected amino

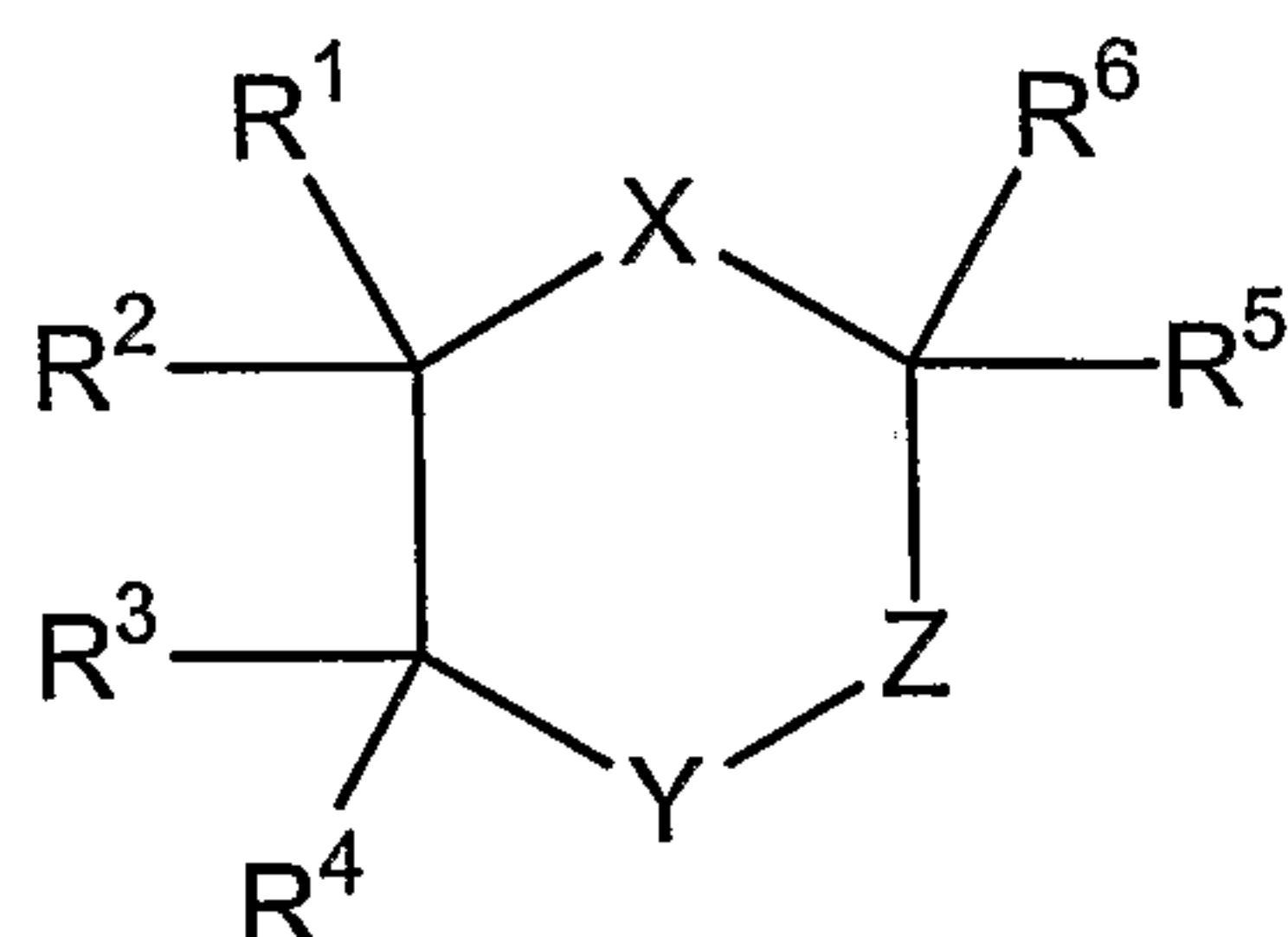
functionality. Preferred substituents are 2'-methoxyethyl, 2'-OCH₃, 2'-O-allyl, 2'-C-allyl, and 2'-fluoro.

X³ is as described for OFG² above.

PG can be a triarylmethyl group (*e.g.*, a dimethoxytrityl group) or Si(X^{5'})(X^{5''})(X^{5'''}) in
 5 which (X^{5'}), (X^{5''}), and (X^{5'''}) are as described elsewhere.

Sugar Replacement-Based Monomers

Cyclic sugar replacement-based monomers, *e.g.*, sugar replacement-based ligand-
 10 conjugated monomers, are also referred to herein as sugar replacement monomer subunit
 (SRMS) monomer compounds. Preferred carriers have the general formula (LCM-2) provided
 below. (In that structure preferred backbone attachment points can be chosen from R¹ or R²; R³
 or R⁴; or R⁹ and R¹⁰ if Y is CR⁹R¹⁰ (two positions are chosen to give two backbone attachment
 points, *e.g.*, R¹ and R⁴, or R⁴ and R⁹). Preferred tethering attachment points include R⁷; R⁵ or R⁶
 15 when X is CH₂. The carriers are described below as an entity, which can be incorporated into a
 strand. Thus, it is understood that the structures also encompass the situations wherein one (in
 the case of a terminal position) or two (in the case of an internal position) of the attachment
 points, *e.g.*, R¹ or R²; R³ or R⁴; or R⁹ or R¹⁰ (when Y is CR⁹R¹⁰), is connected to the phosphate,
 or modified phosphate, *e.g.*, sulfur containing, backbone. *E.g.*, one of the above-named R groups
 20 can be -CH₂-, wherein one bond is connected to the carrier and one to a backbone atom, *e.g.*, a
 linking oxygen or a central phosphorus atom.



(LCM-2)

5

in which,

X is $\text{N}(\text{CO})\text{R}^7$, NR^7 or CH_2 ;

Y is NR^8 , O, S, CR^9R^{10} ;

Z is $\text{CR}^{11}\text{R}^{12}$ or absent;

10 Each of R^1 , R^2 , R^3 , R^4 , R^9 , and R^{10} is, independently, H, OR^a , or $(\text{CH}_2)_n\text{OR}^b$, provided that at least two of R^1 , R^2 , R^3 , R^4 , R^9 , and R^{10} are OR^a and/or $(\text{CH}_2)_n\text{OR}^b$;

Each of R^5 , R^6 , R^{11} , and R^{12} is, independently, a ligand, H, C_1 - C_6 alkyl optionally substituted with 1-3 R^{13} , or $\text{C}(\text{O})\text{NHR}^7$; or R^5 and R^{11} together are C_3 - C_8 cycloalkyl optionally substituted with R^{14} ;

15 R^7 can be a ligand, *e.g.*, R^7 can be R^d , or R^7 can be a ligand tethered indirectly to the carrier, *e.g.*, through a tethering moiety, *e.g.*, C_1 - C_{20} alkyl substituted with NR^cR^d ; or C_1 - C_{20} alkyl substituted with $\text{NHC}(\text{O})\text{R}^d$;

R^8 is H or C_1 - C_6 alkyl;

R^{13} is hydroxy, C_1 - C_4 alkoxy, or halo;

20 R^{14} is NR^cR^7 ;

R^{15} is C_1 - C_6 alkyl optionally substituted with cyano, or C_2 - C_6 alkenyl;

R^{16} is C_1 - C_{10} alkyl;

R^{17} is a liquid or solid phase support reagent;

L is $-\text{C}(\text{O})(\text{CH}_2)_q\text{C}(\text{O})-$, or $-\text{C}(\text{O})(\text{CH}_2)_q\text{S}-$;

25 R^a is a protecting group, *e.g.*, CAr_3 ; (*e.g.*, a dimethoxytrityl group) or $\text{Si}(\text{X}^{5'}) (\text{X}^{5''}) (\text{X}^{5'''})$ in which $(\text{X}^{5'})$, $(\text{X}^{5''})$, and $(\text{X}^{5'''})$ are as described elsewhere.

R^b is $P(O)(O^-)H$, $P(OR^{15})N(R^{16})_2$ or $L-R^{17}$;

R^c is H or C_1-C_6 alkyl;

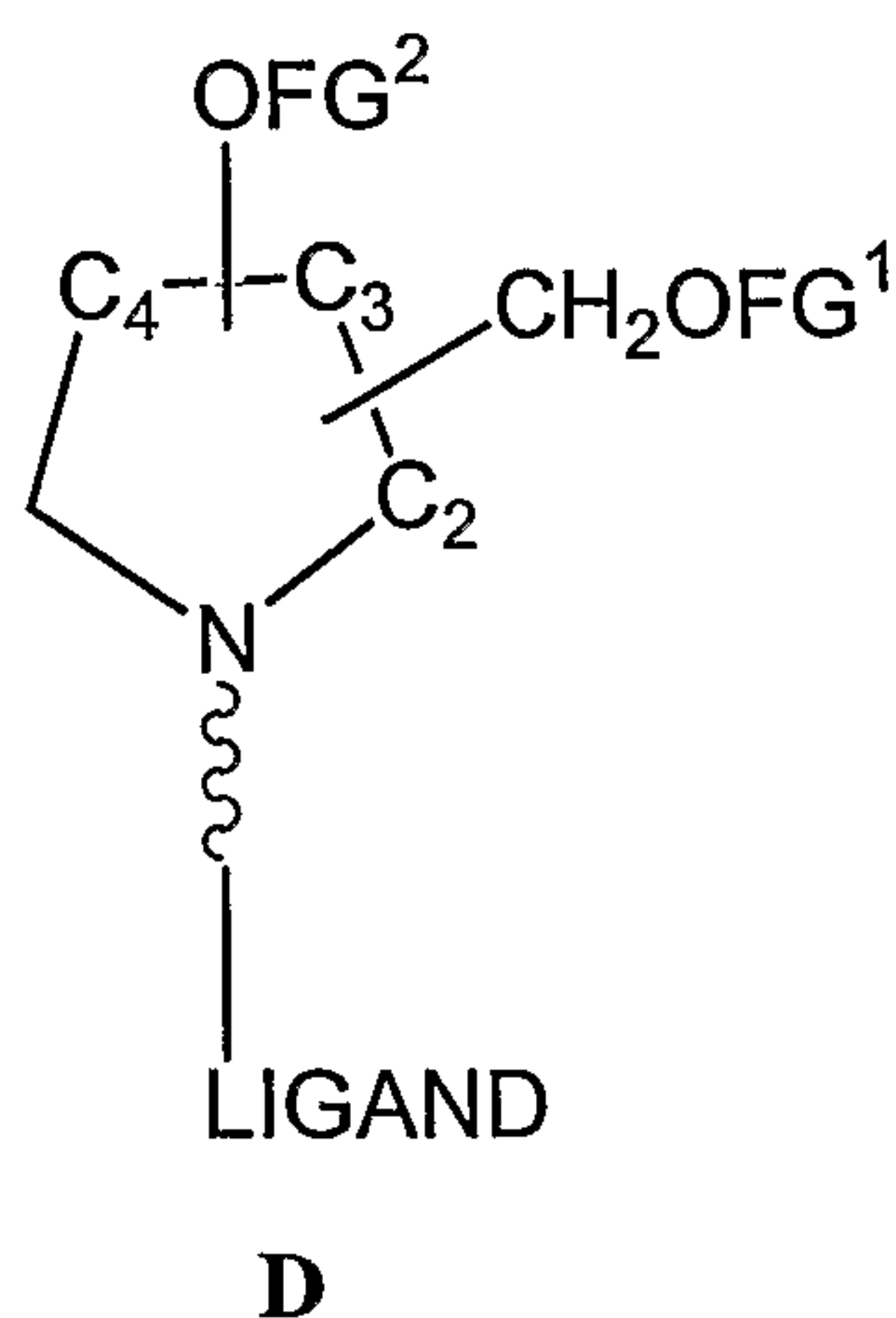
R^d is H or a ligand;

Each Ar is, independently, C_6-C_{10} aryl optionally substituted with C_1-C_4 alkoxy;

n is 1-4; and q is 0-4.

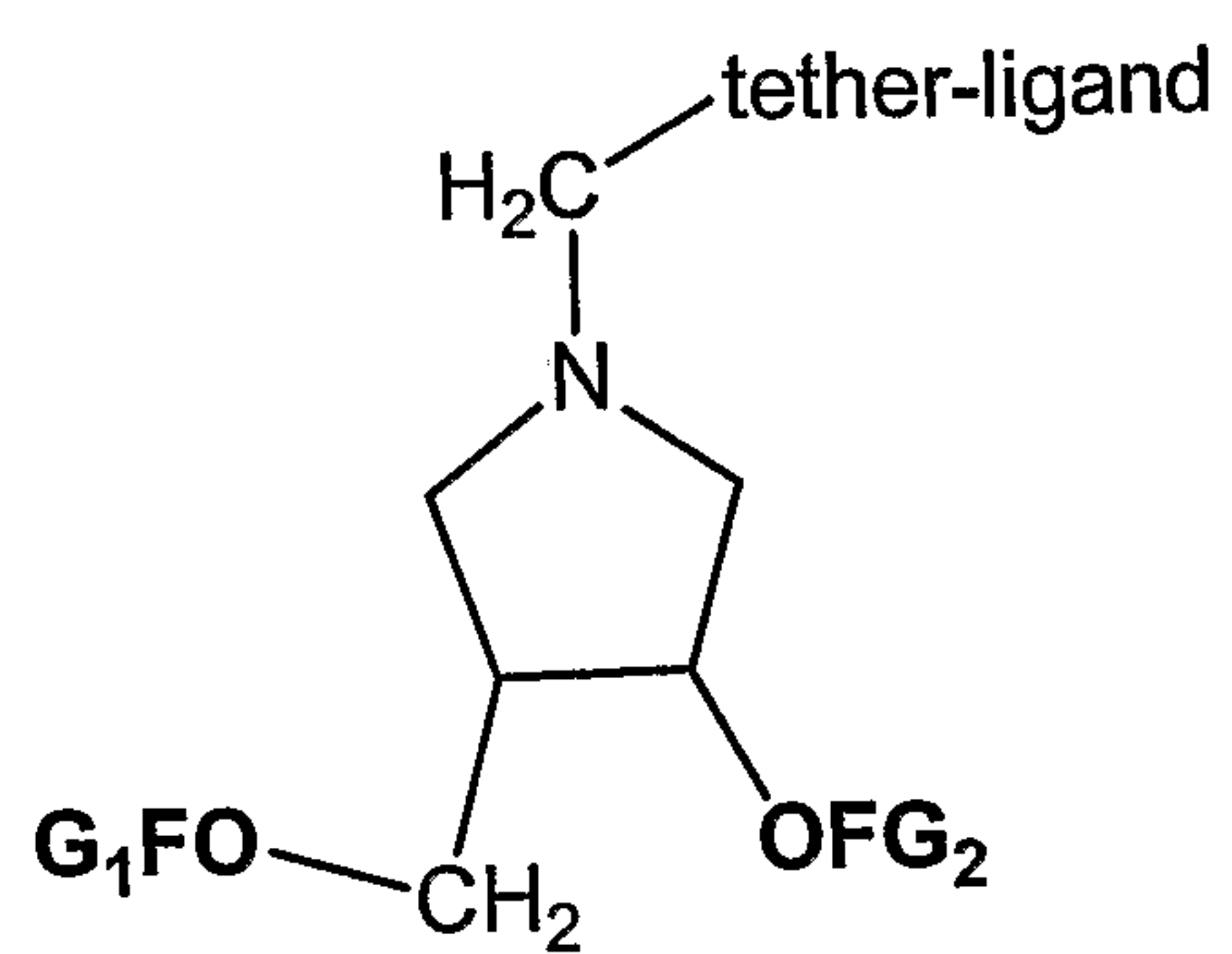
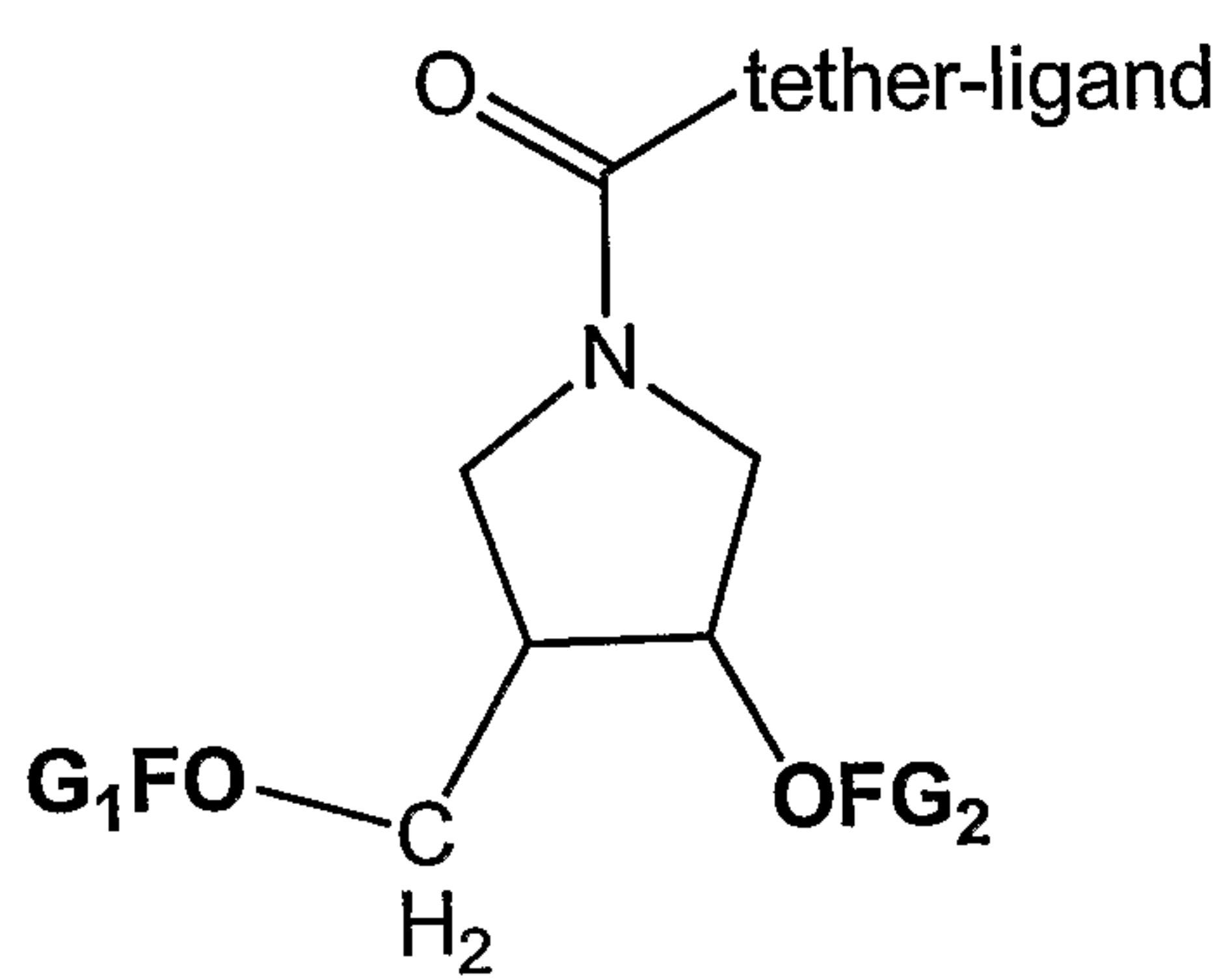
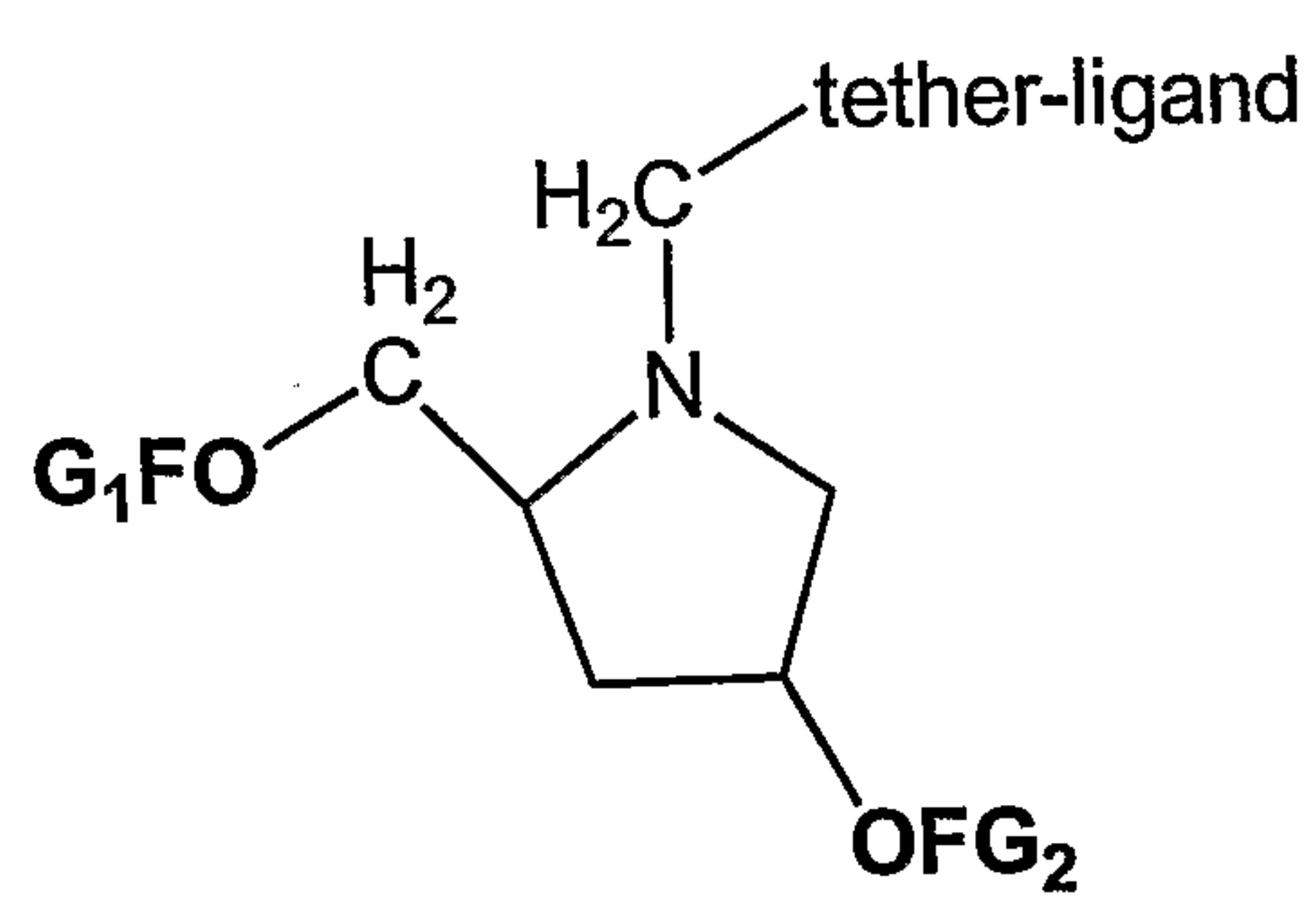
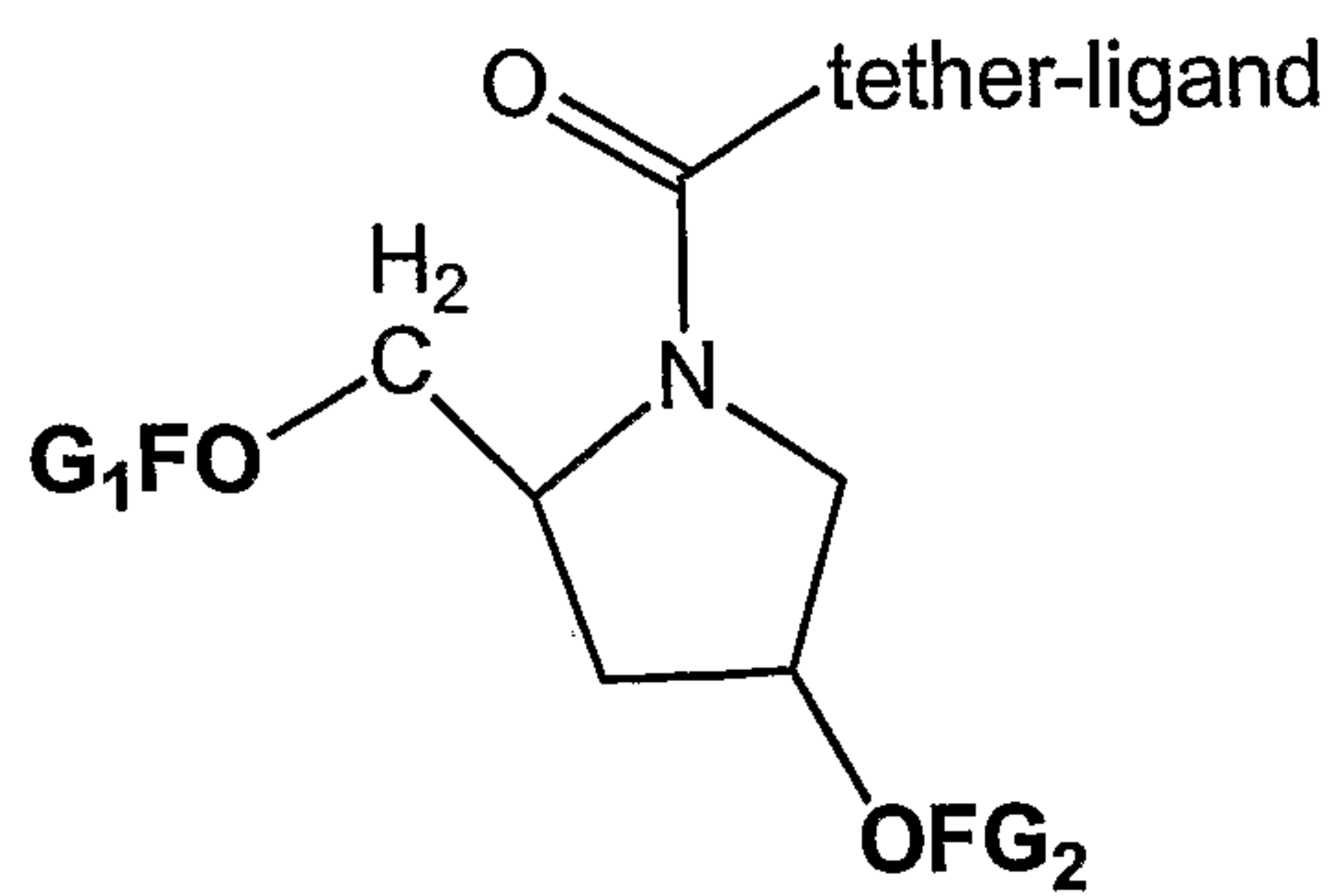
Exemplary carriers include those in which, *e.g.*, X is $N(CO)R^7$ or NR^7 , Y is CR^9R^{10} , and Z is absent; or X is $N(CO)R^7$ or NR^7 , Y is CR^9R^{10} , and Z is $CR^{11}R^{12}$; or X is $N(CO)R^7$ or NR^7 , Y is NR^8 , and Z is $CR^{11}R^{12}$; or X is $N(CO)R^7$ or NR^7 , Y is O, and Z is $CR^{11}R^{12}$; or X is CH_2 ; Y is CR^9R^{10} ; Z is $CR^{11}R^{12}$, and R^5 and R^{11} together form C_6 cycloalkyl (**H**, $z = 2$), or the indane ring system, *e.g.*, X is CH_2 ; Y is CR^9R^{10} ; Z is $CR^{11}R^{12}$, and R^5 and R^{11} together form C_5 cycloalkyl (**H**, $z = 1$).

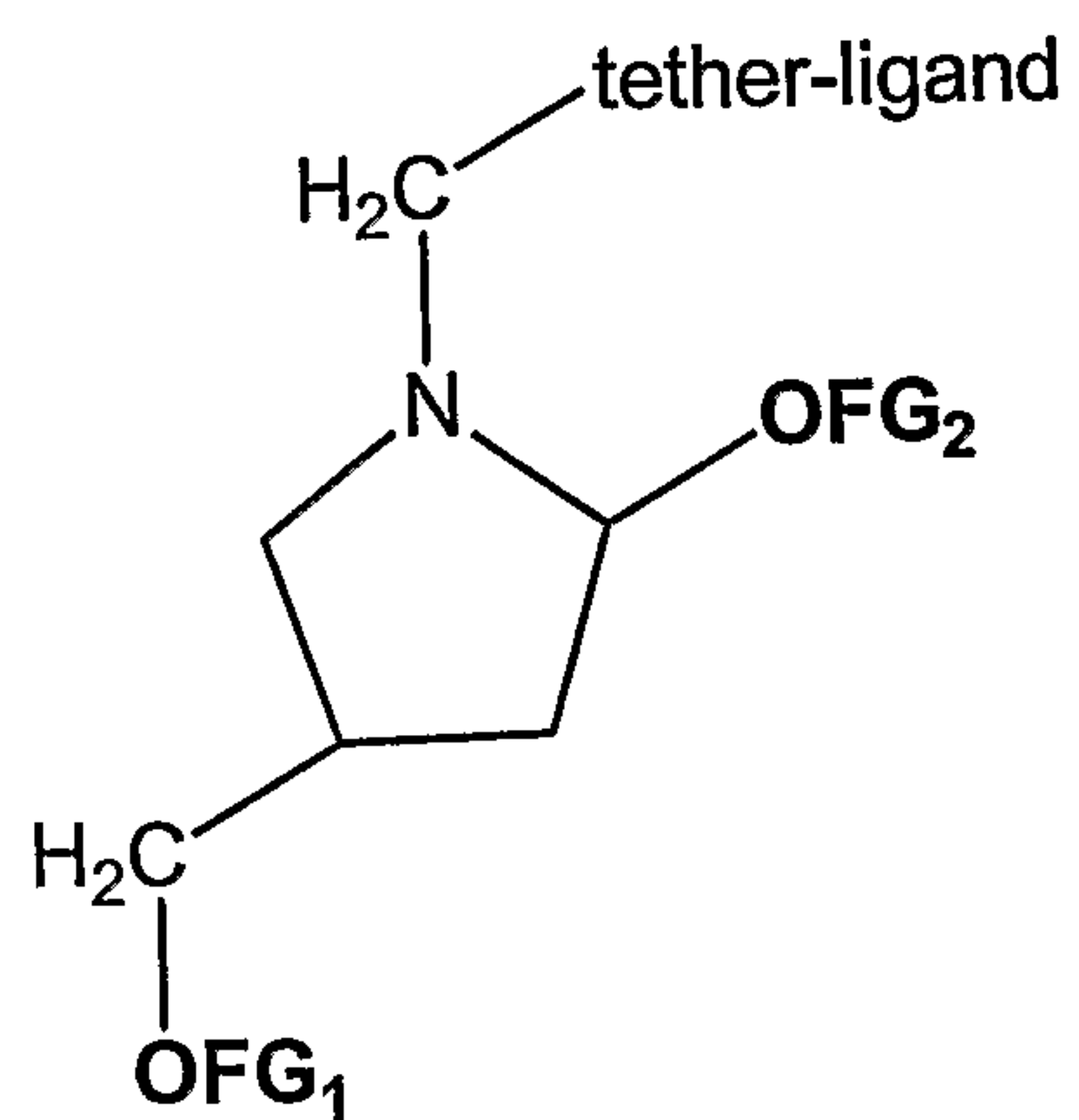
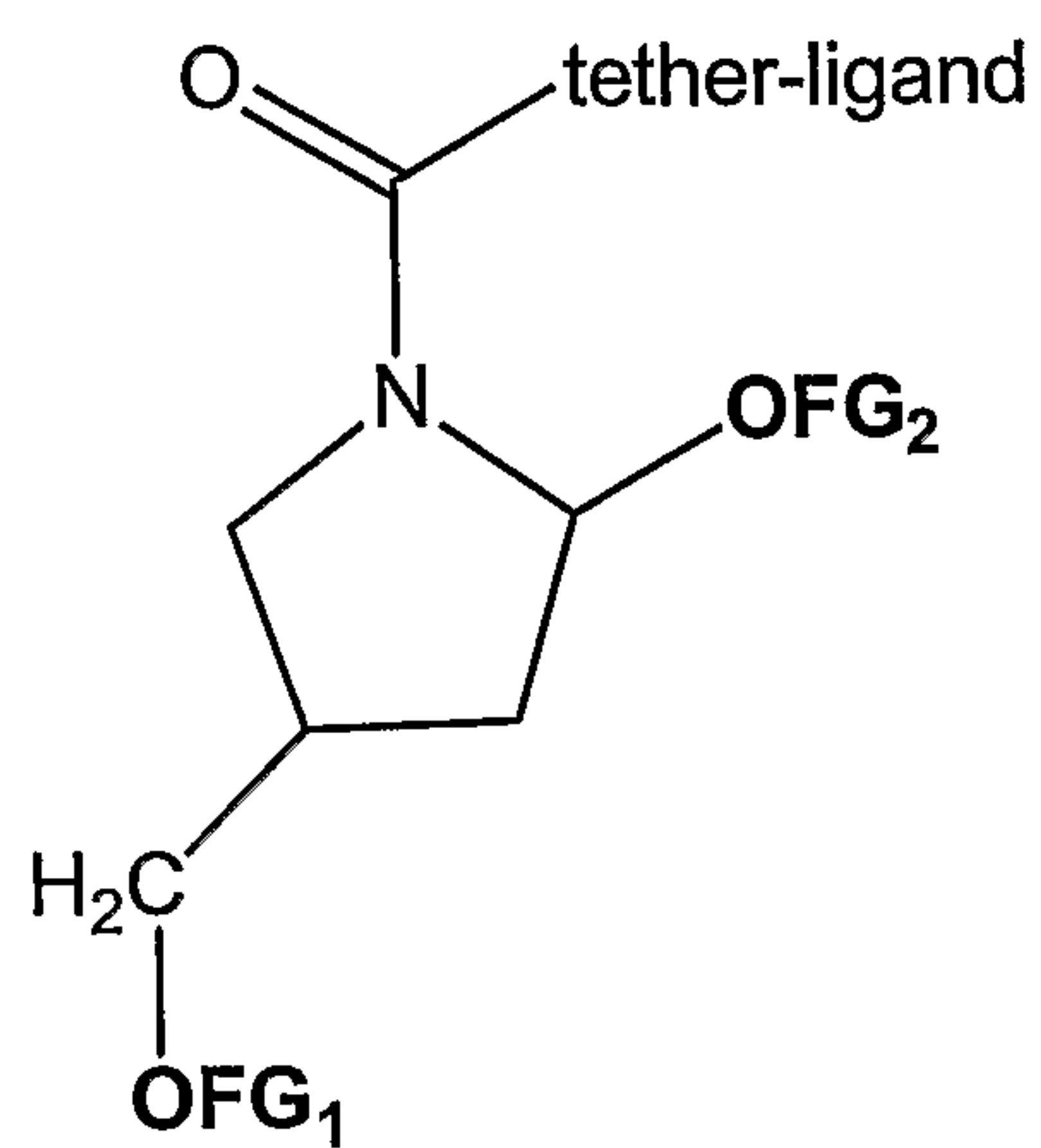
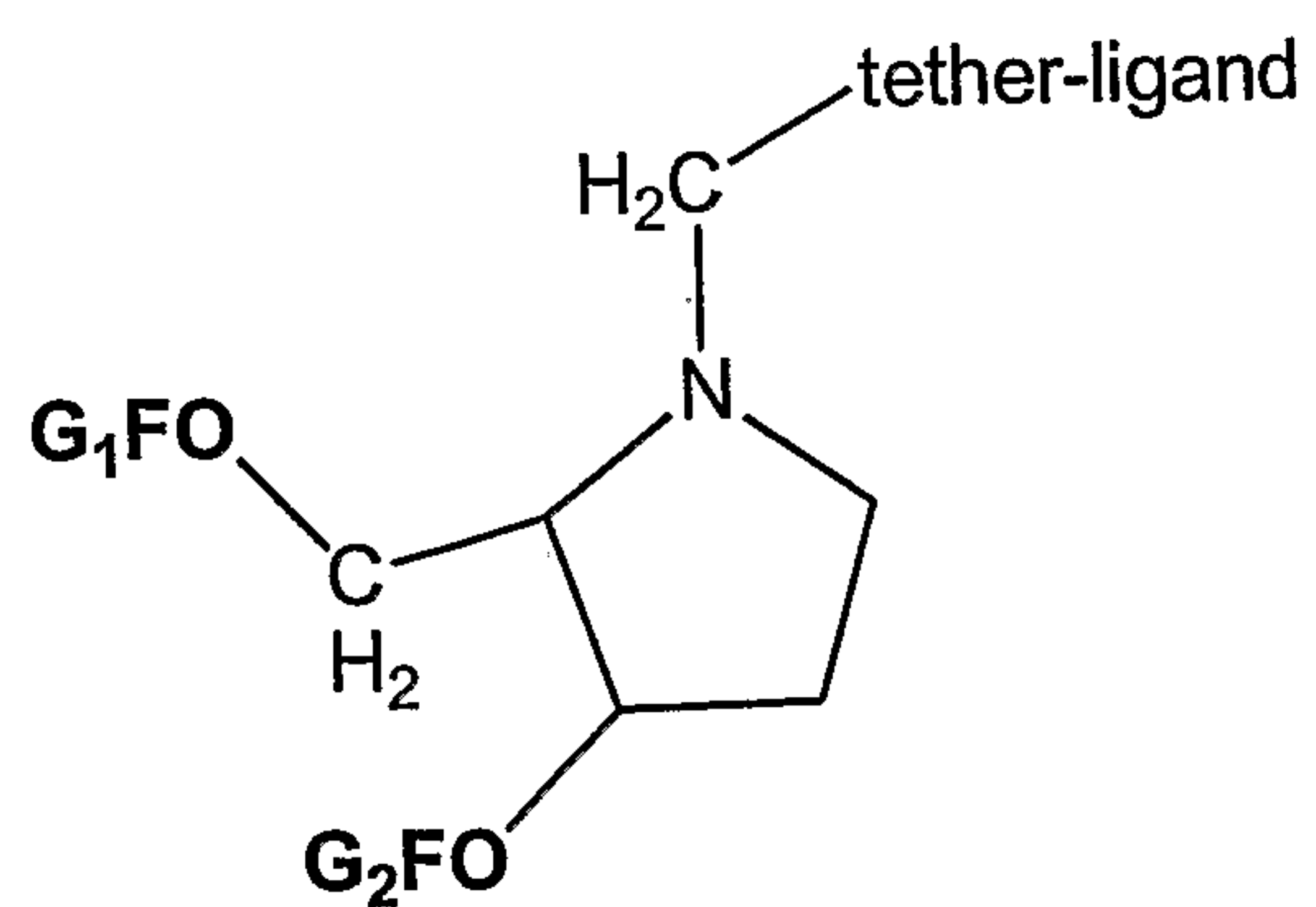
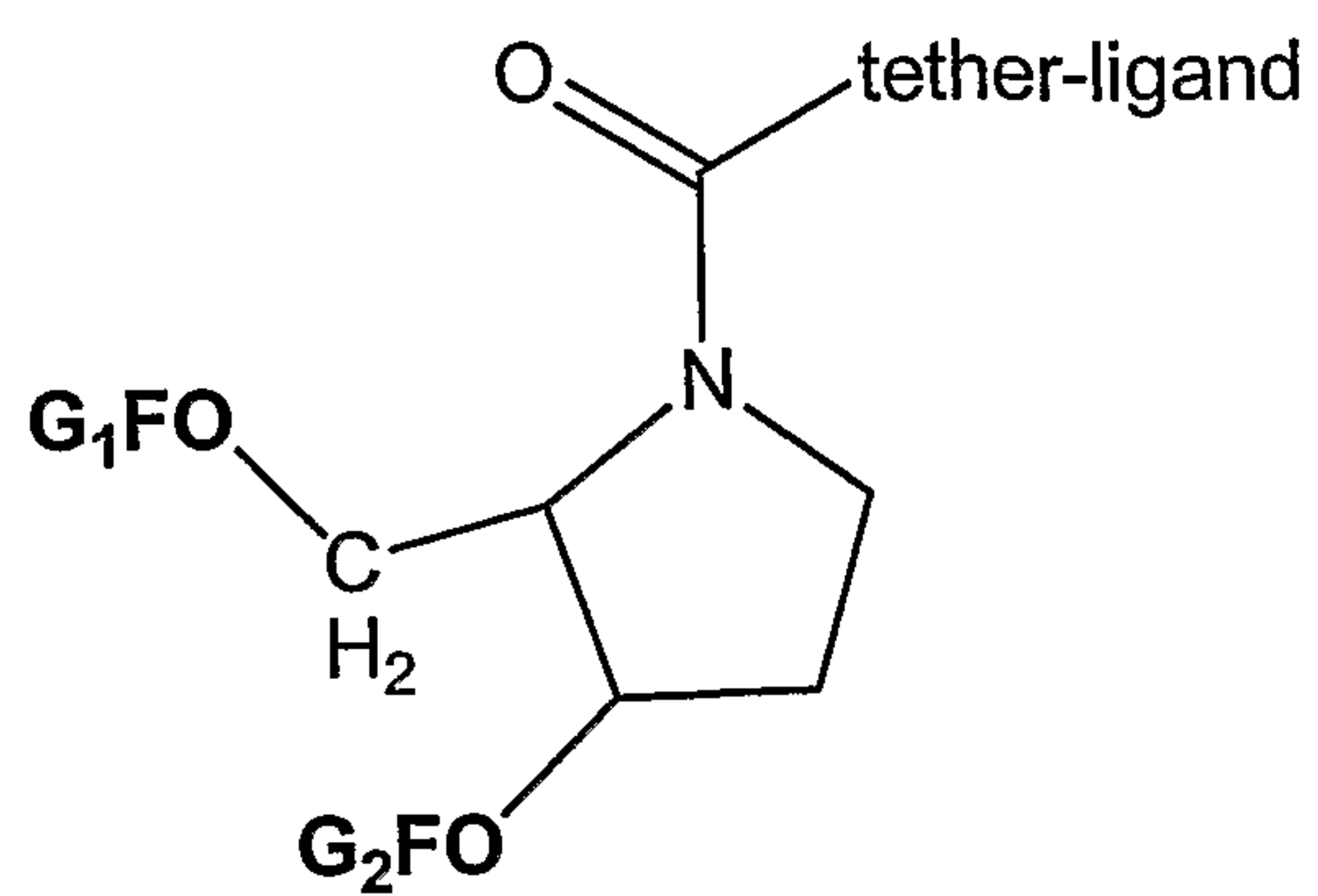
In certain embodiments, the carrier may be based on the pyrroline ring system or the 4-hydroxyproline ring system, *e.g.*, X is $N(CO)R^7$ or NR^7 , Y is CR^9R^{10} , and Z is absent (**D**). OFG^1 is preferably attached to a primary carbon, *e.g.*, an exocyclic alkylene



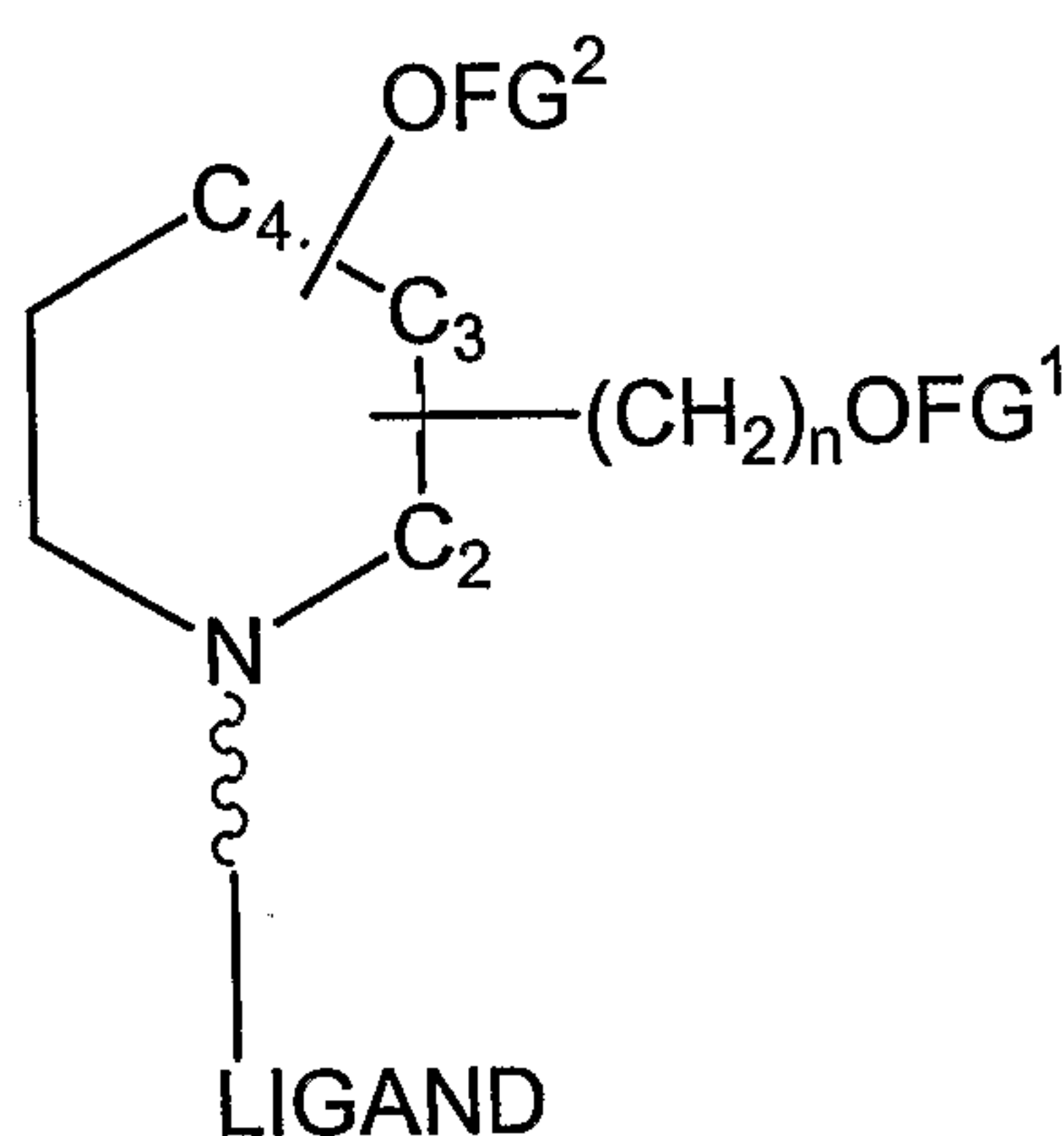
group, *e.g.*, a methylene group, connected to one of the carbons in the five-membered ring ($-CH_2OFG^1$ in **D**). OFG^2 is preferably attached directly to one of the carbons in the five-membered ring ($-OFG^2$ in **D**). For the pyrroline-based carriers, $-CH_2OFG^1$ may be attached to C-2 and OFG^2 may be attached to C-3; or $-CH_2OFG^1$ may be attached to C-3 and OFG^2 may be attached to C-4. In certain embodiments, CH_2OFG^1 and OFG^2 may be geminally substituted to

one of the above-referenced carbons. For the 3-hydroxyproline-based carriers, $-\text{CH}_2\text{OFG}^1$ may be attached to C-2 and OFG^2 may be attached to C-4. The pyrroline- and 4-hydroxyproline-based monomers may therefore contain linkages (*e.g.*, carbon-carbon bonds) wherein bond rotation is restricted about that particular linkage, *e.g.* restriction resulting from the presence of a ring. Thus, CH_2OFG^1 and OFG^2 may be *cis* or *trans* with respect to one another in any of the pairings delineated above. Accordingly, all *cis/trans* isomers are expressly included. The monomers may also contain one or more asymmetric centers and thus occur as racemates and racemic mixtures, single enantiomers, individual diastereomers and diastereomeric mixtures. All such isomeric forms of the monomers are expressly included (*e.g.*, the centers bearing CH_2OFG^1 and OFG^2 can both have the R configuration; or both have the S configuration; or one center can have the R configuration and the other center can have the S configuration and *vice versa*). The tethering attachment point is preferably nitrogen. Preferred examples of carrier **D** include the following:





In certain embodiments, the carrier may be based on the piperidine ring system (**E**), *e.g.*, X is N(CO)R⁷ or NR⁷, Y is CR⁹R¹⁰, and Z is CR¹¹R¹². OFG¹ is preferably



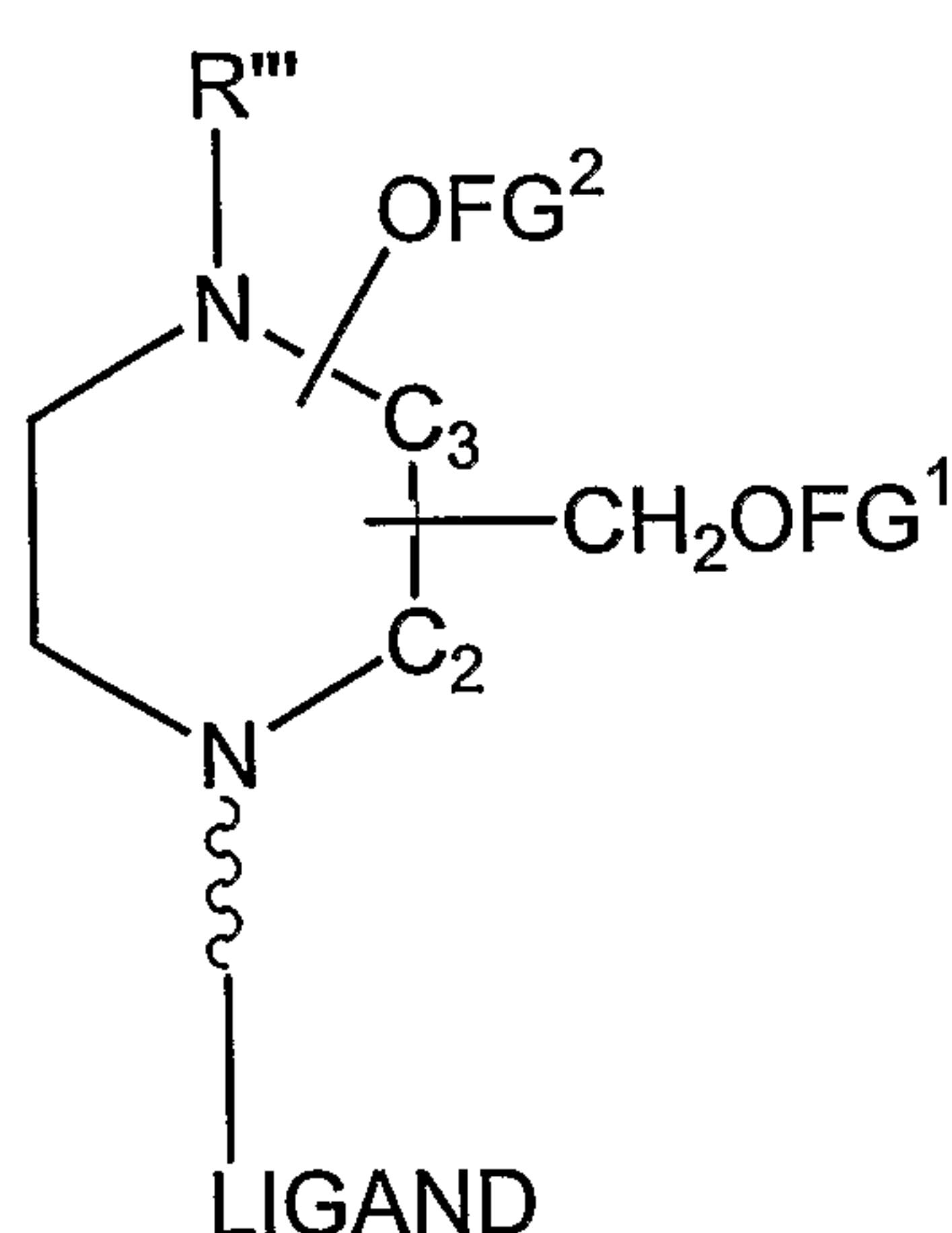
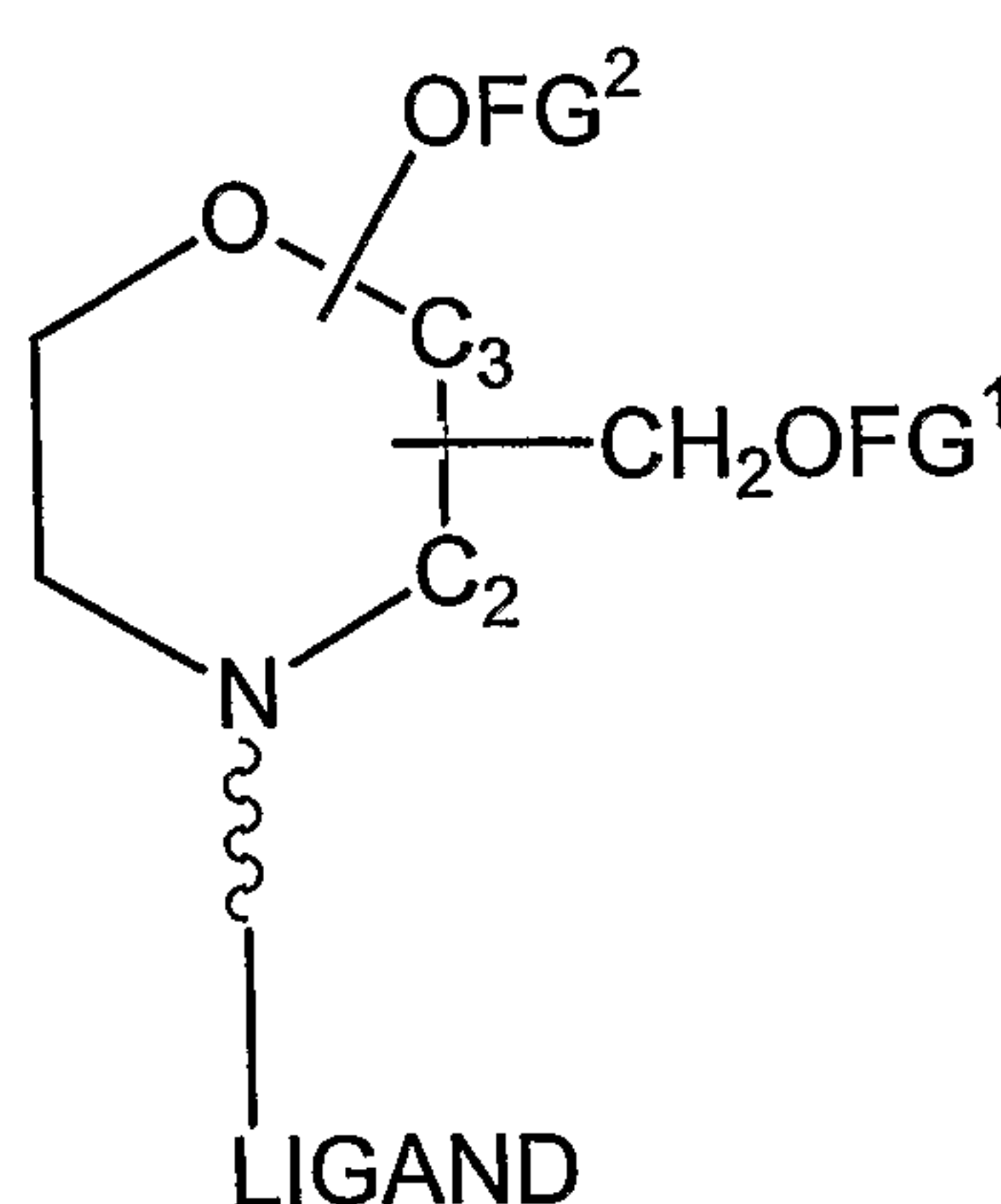
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attached to a primary carbon, *e.g.*, an exocyclic alkylene group, *e.g.*, a methylene group ($n=1$) or ethylene group ($n=2$), connected to one of the carbons in the six-membered ring [$-(CH_2)_nOFG^1$ in **E**]. OFG^2 is preferably attached directly to one of the carbons in the six-membered ring ($-OFG^2$ in **E**). $-(CH_2)_nOFG^1$ and OFG^2 may be disposed in a geminal manner on the ring, *i.e.*, both groups may be attached to the same carbon, *e.g.*, at C-2, C-3, or C-4. Alternatively, $-(CH_2)_nOFG^1$ and OFG^2 may be disposed in a vicinal manner on the ring, *i.e.*, both groups may be attached to adjacent ring carbon atoms, *e.g.*, $-(CH_2)_nOFG^1$ may be attached to C-2 and OFG^2 may be attached to C-3; $-(CH_2)_nOFG^1$ may be attached to C-3 and OFG^2 may be attached to C-2; $-(CH_2)_nOFG^1$ may be attached to C-3 and OFG^2 may be attached to C-4; or $-(CH_2)_nOFG^1$ may be attached to C-4 and OFG^2 may be attached to C-3. The piperidine-based monomers may therefore contain linkages (*e.g.*, carbon-carbon bonds) wherein bond rotation is restricted about that particular linkage, *e.g.* restriction resulting from the presence of a ring. Thus, $-(CH_2)_nOFG^1$ and OFG^2 may be *cis* or *trans* with respect to one another in any of the pairings delineated above. Accordingly, all *cis/trans* isomers are expressly included. The monomers may also contain one or more asymmetric centers and thus occur as racemates and racemic mixtures, single enantiomers, individual diastereomers and diastereomeric mixtures. All such isomeric forms of the monomers are expressly included (*e.g.*, the centers bearing CH_2OFG^1 and OFG^2 can

20

both have the R configuration; or both have the S configuration; or one center can have the R configuration and the other center can have the S configuration and *vice versa*). The tethering attachment point is preferably nitrogen.

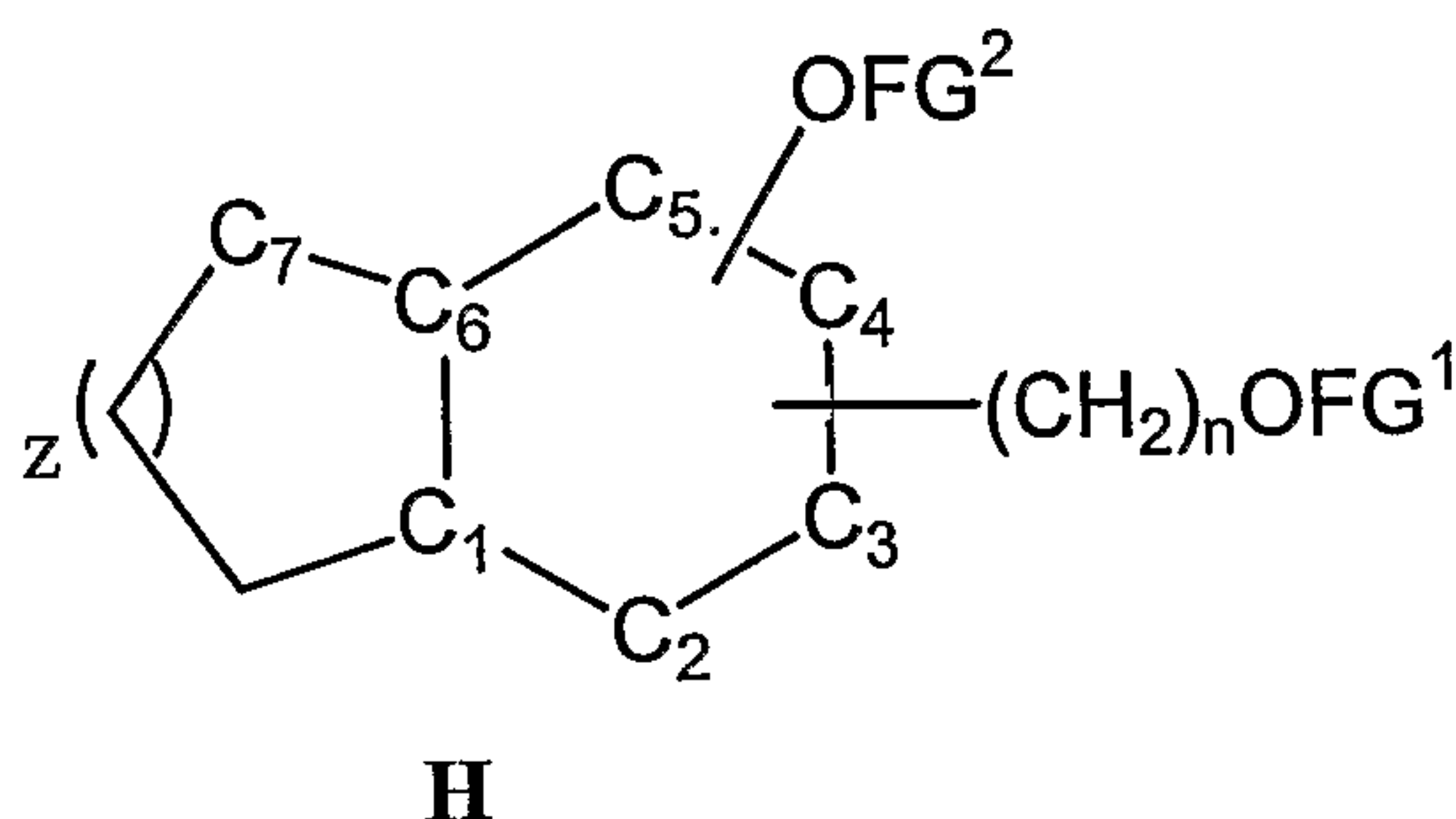
In certain embodiments, the carrier may be based on the piperazine ring system (**F**), *e.g.*,
 5 X is N(CO)R⁷ or NR⁷, Y is NR⁸, and Z is CR¹¹R¹², or the morpholine ring system (**G**), *e.g.*, X is N(CO)R⁷ or NR⁷, Y is O, and Z is CR¹¹R¹². OFG¹ is preferably

**F****G**

10 attached to a primary carbon, *e.g.*, an exocyclic alkylene group, *e.g.*, a methylene group, connected to one of the carbons in the six-membered ring (-CH₂OFG¹ in **F** or **G**). OFG² is preferably attached directly to one of the carbons in the six-membered rings (-OFG² in **F** or **G**). For both **F** and **G**, -CH₂OFG¹ may be attached to C-2 and OFG² may be attached to C-3; or *vice versa*. In certain embodiments, CH₂OFG¹ and OFG² may be geminally substituted to one of the
 15 above-referenced carbons. The piperazine- and morpholine-based monomers may therefore contain linkages (*e.g.*, carbon-carbon bonds) wherein bond rotation is restricted about that particular linkage, *e.g.* restriction resulting from the presence of a ring. Thus, CH₂OFG¹ and OFG² may be *cis* or *trans* with respect to one another in any of the pairings delineated above. Accordingly, all *cis/trans* isomers are expressly included. The monomers may also contain one
 20 or more asymmetric centers and thus occur as racemates and racemic mixtures, single enantiomers, individual diastereomers and diastereomeric mixtures. All such isomeric forms of

the monomers are expressly included (*e.g.*, the centers bearing CH_2OFG^1 and OFG^2 can both have the R configuration; or both have the S configuration; or one center can have the R configuration and the other center can have the S configuration and *vice versa*). R''' can be, *e.g.*, $\text{C}_1\text{-C}_6$ alkyl, preferably CH_3 . The tethering attachment point is preferably nitrogen in both F and G.

In certain embodiments, the carrier may be based on the decalin ring system, *e.g.*, X is CH_2 ; Y is CR^9R^{10} ; Z is $\text{CR}^{11}\text{R}^{12}$, and R^5 and R^{11} together form C_6 cycloalkyl (**H**, $z = 2$), or the indane ring system, *e.g.*, X is CH_2 ; Y is CR^9R^{10} ; Z is $\text{CR}^{11}\text{R}^{12}$, and R^5 and R^{11} together form C_5 cycloalkyl (**H**, $z = 1$). OFG^1 is preferably attached to a primary carbon,

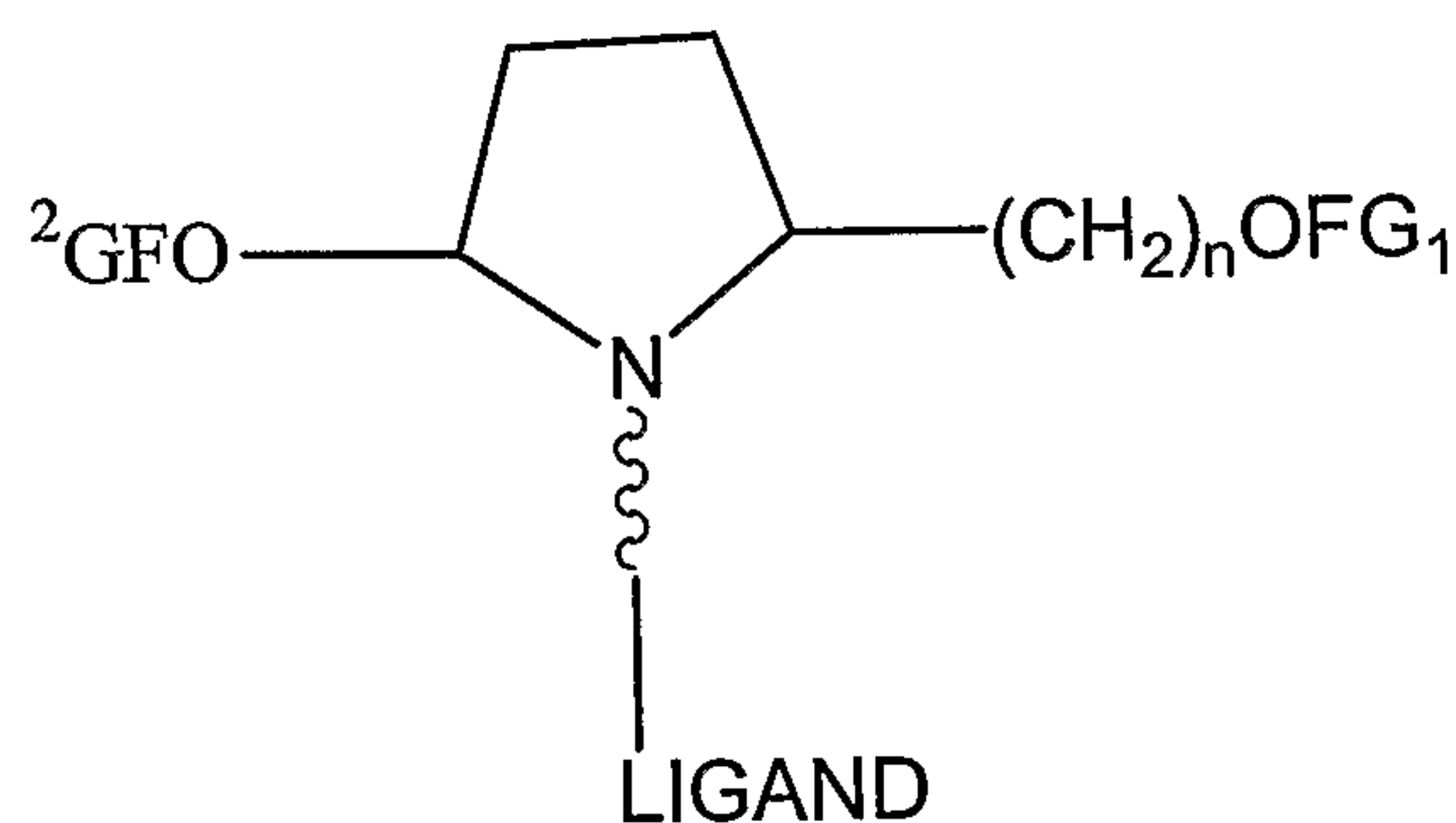


e.g., an exocyclic methylene group ($n=1$) or ethylene group ($n=2$) connected to one of C-2, C-3, C-4, or C-5 [$-(\text{CH}_2)_n\text{OFG}^1$ in **H**]. OFG^2 is preferably attached directly to one of C-2, C-3, C-4, or C-5 ($-\text{OFG}^2$ in **H**). $-(\text{CH}_2)_n\text{OFG}^1$ and OFG^2 may be disposed in a geminal manner on the ring, *i.e.*, both groups may be attached to the same carbon, *e.g.*, at C-2, C-3, C-4, or C-5.

Alternatively, $-(\text{CH}_2)_n\text{OFG}^1$ and OFG^2 may be disposed in a vicinal manner on the ring, *i.e.*, both groups may be attached to adjacent ring carbon atoms, *e.g.*, $-(\text{CH}_2)_n\text{OFG}^1$ may be attached to C-2 and OFG^2 may be attached to C-3; $-(\text{CH}_2)_n\text{OFG}^1$ may be attached to C-3 and OFG^2 may be attached to C-2; $-(\text{CH}_2)_n\text{OFG}^1$ may be attached to C-3 and OFG^2 may be attached to C-4; or $-(\text{CH}_2)_n\text{OFG}^1$ may be attached to C-4 and OFG^2 may be attached to C-3; $-(\text{CH}_2)_n\text{OFG}^1$ may be attached to C-4 and OFG^2 may be attached to C-5; or $-(\text{CH}_2)_n\text{OFG}^1$ may be attached to C-5 and OFG^2 may be attached to C-4. The decalin or indane-based monomers may therefore contain linkages (*e.g.*, carbon-carbon bonds) wherein bond rotation is restricted about that particular linkage, *e.g.* restriction resulting from the presence of a ring. Thus, $-(\text{CH}_2)_n\text{OFG}^1$ and OFG^2 may be *cis* or *trans* with respect to one another in any of the pairings delineated above. Accordingly,

all *cis/trans* isomers are expressly included. The monomers may also contain one or more asymmetric centers and thus occur as racemates and racemic mixtures, single enantiomers, individual diastereomers and diastereomeric mixtures. All such isomeric forms of the monomers are expressly included (*e.g.*, the centers bearing CH_2OFG^1 and OFG^2 can both have the R configuration; or both have the S configuration; or one center can have the R configuration and the other center can have the S configuration and *vice versa*). In a preferred embodiment, the substituents at C-1 and C-6 are *trans* with respect to one another. The tethering attachment point is preferably C-6 or C-7.

Other carriers may include those based on 3-hydroxyproline (J). Thus, $-(\text{CH}_2)_n\text{OFG}^1$ and OFG^2 may be *cis* or *trans* with respect to one another. Accordingly, all *cis/trans* isomers are expressly included. The monomers may also contain one or more asymmetric centers



J

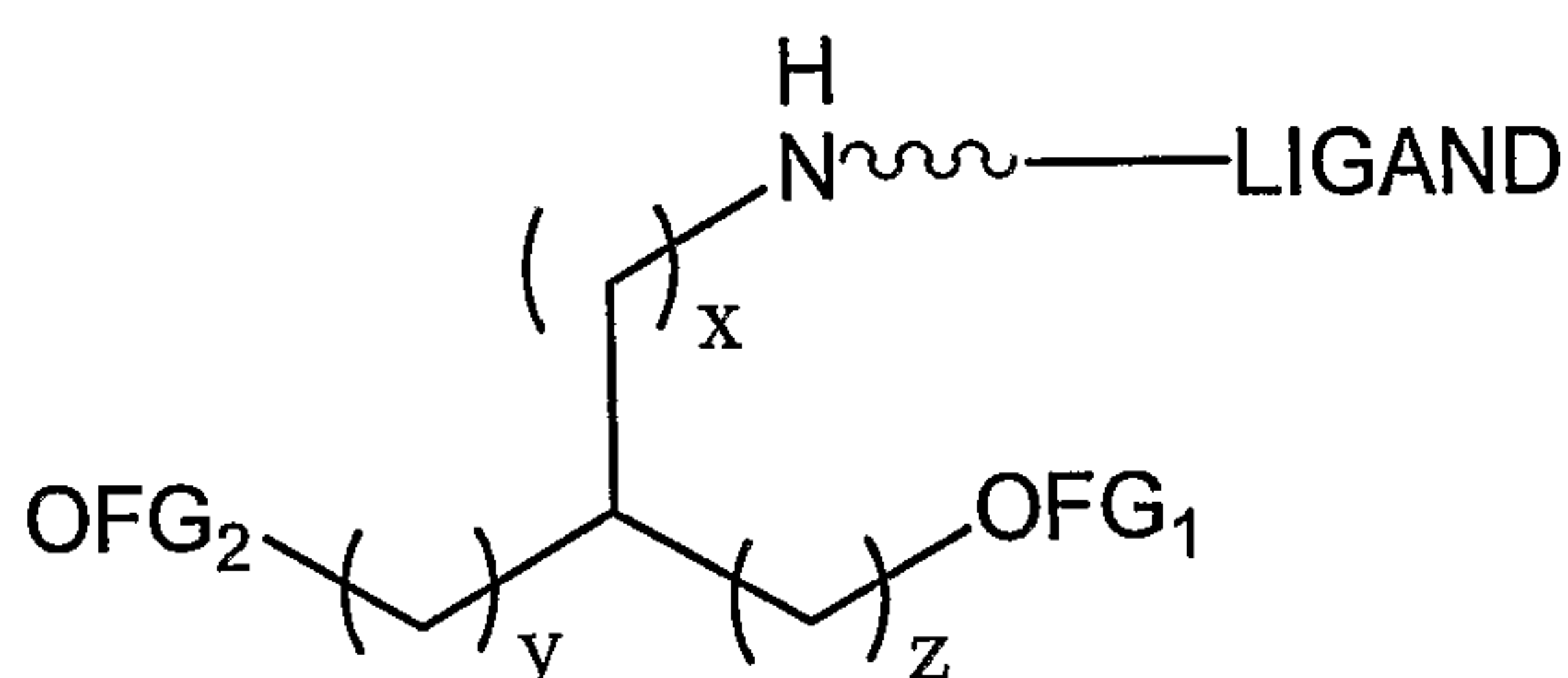
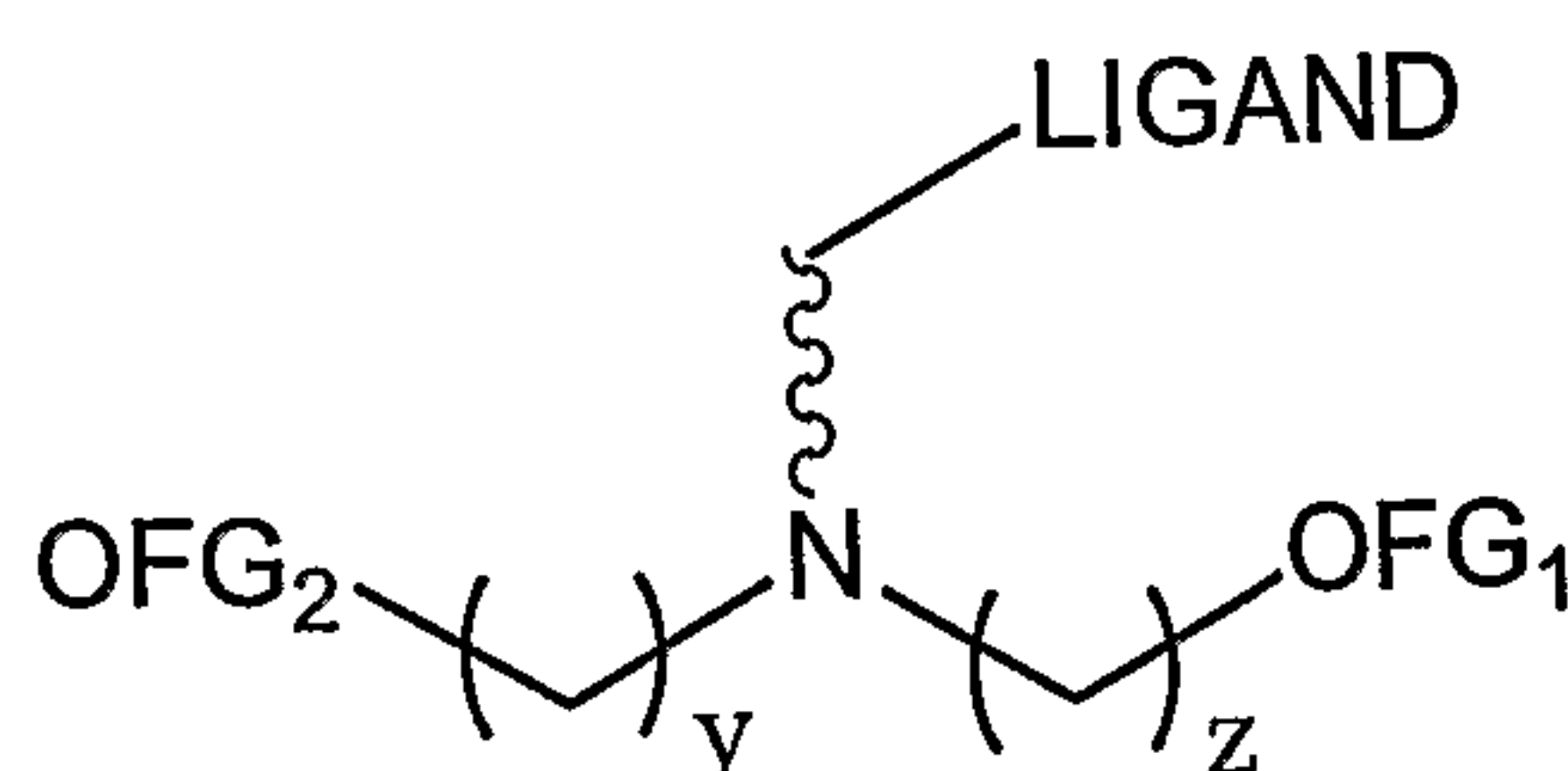
and thus occur as racemates and racemic mixtures, single enantiomers, individual diastereomers and diastereomeric mixtures. All such isomeric forms of the monomers are expressly included (*e.g.*, the centers bearing CH_2OFG^1 and OFG^2 can both have the R configuration; or both have the S configuration; or one center can have the R configuration and the other center can have the S configuration and *vice versa*). The tethering attachment point is preferably nitrogen.

Representative cyclic, sugar replacement-based carriers are shown in FIG. 3.

20 Sugar Replacement-Based Monomers (Acyclic)

Acyclic sugar replacement-based monomers, *e.g.*, sugar replacement-based ligand-conjugated monomers, are also referred to herein as sugar replacement monomer subunit

(SRMS) monomer compounds. Preferred acyclic carriers can have formula **LCM-3** or **LCM-4** below.

**LCM-3****LCM-4**

5 In some embodiments, each of x, y, and z can be, independently of one another, 0, 1, 2, or 3. In formula **LCM-3**, when y and z are different, then the tertiary carbon can have either the R or S configuration. In preferred embodiments, x is zero and y and z are each 1 in formula **LCM-3** (*e.g.*, based on serinol), and y and z are each 1 in formula **LCM-3**. Each of formula **LCM-3** or **LCM-4** below can optionally be substituted, *e.g.*, with hydroxy, alkoxy, perhaloalkyl.

10 Tethers

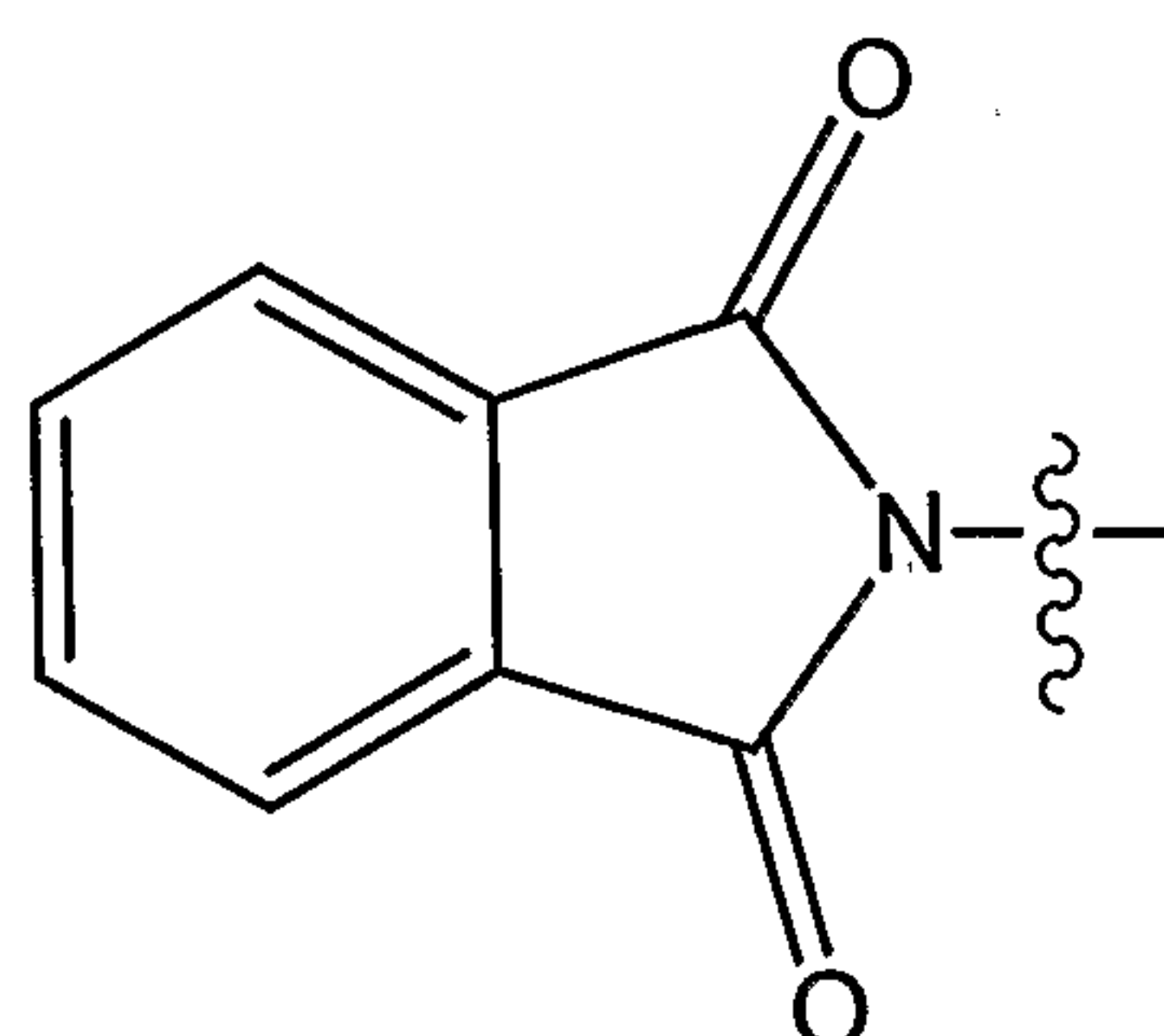
In certain embodiments, a moiety, *e.g.*, a ligand may be connected indirectly to the carrier *via* the intermediacy of an intervening tether. Tethers are connected to the carrier at a tethering attachment point (TAP) and may include any C₁-C₁₀₀ carbon-containing moiety, (*e.g.* C₁-C₇₅, C₁-C₅₀, C₁-C₂₀, C₁-C₁₀; C₁, C₂, C₃, C₄, C₅, C₆, C₇, C₈, C₉, or C₁₀), preferably having at least one
 15 nitrogen atom. In preferred embodiments, the nitrogen atom forms part of a terminal amino or amido (NHC(O)-) group on the tether, which may serve as a connection point for the ligand. Preferred tethers (underlined) include TAP-(CH₂)_nNH-; TAP-C(O)(CH₂)_nNH-; TAP-NR''''(CH₂)_nNH-, TAP-C(O)-(CH₂)_n-C(O)-; TAP-C(O)-(CH₂)_n-C(O)O-; TAP-C(O)-O-; TAP-C(O)-(CH₂)_n-NH-C(O)-; TAP-C(O)-(CH₂)_n-; TAP-C(O)-NH-; TAP-C(O)-; TAP-(CH₂)_n-C(O)-; TAP-(CH₂)_n-C(O)O-; TAP-(CH₂)_n-; or TAP-(CH₂)_n-NH-C(O)-; in which n is 1-20 (*e.g.*, 1, 2, 3,
 20 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20) and R'''' is C₁-C₆ alkyl. Preferably, n is 5, 6, or 11. In other embodiments, the nitrogen may form part of a terminal oxyamino group, *e.g.*, -ONH₂, or hydrazino group, -NHNH₂. The tether may optionally be substituted, *e.g.*, with hydroxy, alkoxy, perhaloalkyl, and/or optionally inserted with one or more additional

heteroatoms, *e.g.*, N, O, or S. Preferred tethered ligands may include, *e.g.*, TAP-
(CH₂)_nNH(LIGAND); TAP-C(O)(CH₂)_nNH(LIGAND); TAP-NR'''(CH₂)_nNH(LIGAND);
TAP-(CH₂)_nONH(LIGAND); TAP-C(O)(CH₂)_nONH(LIGAND); TAP-
NR'''(CH₂)_nONH(LIGAND); TAP-(CH₂)_nNHNH₂(LIGAND), TAP-
5 C(O)(CH₂)_nNHNH₂(LIGAND); TAP-NR'''(CH₂)_nNHNH₂(LIGAND); TAP-C(O)-(CH₂)_n-
C(O)(LIGAND); TAP-C(O)-(CH₂)_n-C(O)O(LIGAND); TAP-C(O)-O(LIGAND); TAP-C(O)-
(CH₂)_n-NH-C(O)(LIGAND); TAP-C(O)-(CH₂)_n(LIGAND); TAP-C(O)-NH(LIGAND); TAP-
C(O)(LIGAND); TAP-(CH₂)_n-C(O)(LIGAND); TAP-(CH₂)_n-C(O)O(LIGAND); TAP-
(CH₂)_n(LIGAND); or TAP-(CH₂)_n-NH-C(O)(LIGAND). In some embodiments, amino
10 terminated tethers (*e.g.*, NH₂, ONH₂, NH₂NH₂) can form an imino bond (*i.e.*, C=N) with the
ligand. In some embodiments, amino terminated tethers (*e.g.*, NH₂, ONH₂, NH₂NH₂) can
acylated, *e.g.*, with C(O)CF₃.

In some embodiments, the tether can terminate with a mercapto group (*i.e.*, SH) or an
olefin (*e.g.*, CH=CH₂). For example, the tether can be TAP-(CH₂)_n-SH, TAP-C(O)(CH₂)_nSH,
15 TAP-(CH₂)_n-(CH=CH₂), or TAP-C(O)(CH₂)_n(CH=CH₂), in which n can be as described
elsewhere. In certain embodiments, the olefin can be a Diels-Alder diene or dienophile. The
tether may optionally be substituted, *e.g.*, with hydroxy, alkoxy, perhaloalkyl, and/or optionally
inserted with one or more additional heteroatoms, *e.g.*, N, O, or S. The double bond can be *cis* or
trans or *E* or *Z*.

In other embodiments the tether may include an electrophilic moiety, preferably at the
terminal position of the tether. Preferred electrophilic moieties include, *e.g.*, an aldehyde, alkyl
halide, mesylate, tosylate, nosylate, or brosylate, or an activated carboxylic acid ester, *e.g.* an
NHS ester, or a pentafluorophenyl ester. Preferred tethers (underlined) include TAP-
(CH₂)_nCHO; TAP-C(O)(CH₂)_nCHO; or TAP-NR'''(CH₂)_nCHO, in which n is 1-6 and R''' is
25 C₁-C₆ alkyl; or TAP-(CH₂)_nC(O)ONHS; TAP-C(O)(CH₂)_nC(O)ONHS; or TAP-NR'''(CH₂)
_nC(O)ONHS, in which n is 1-6 and R''' is C₁-C₆ alkyl; TAP-(CH₂)_nC(O)OC₆F₅; TAP-
C(O)(CH₂)_nC(O)OC₆F₅; or TAP-NR'''(CH₂)_nC(O)OC₆F₅, in which n is 1-11 and R''' is C₁-
C₆ alkyl; or -(CH₂)_nCH₂LG; TAP-C(O)(CH₂)_nCH₂LG; or TAP-NR'''(CH₂)_nCH₂LG, in which n
30 can be as described elsewhere and R''' is C₁-C₆ alkyl (LG can be a leaving group, *e.g.*, halide,
mesylate, tosylate, nosylate, brosylate). Tethering can be carried out by coupling a nucleophilic
group of a ligand, *e.g.*, a thiol or amino group with an electrophilic group on the tether.

In other embodiments, it can be desirable for the ligand-conjugated monomer or a ligand-conjugated monomer to include a phthalimido group (K) at the terminal position of the tether.



K

5 In other embodiments, other protected amino groups can be at the terminal position of the tether, *e.g.*, alloc, monomethoxy trityl (MMT), trifluoroacetyl, Fmoc, or aryl sulfonyl (*e.g.*, the aryl portion can be *ortho*-nitrophenyl or *ortho, para*-dinitrophenyl).

Any of the tethers described herein may further include one or more additional linking groups, *e.g.*, -O-(CH₂)_n-, -(CH₂)_n-SS-, -(CH₂)_n-, or -(CH=CH)-.

10

Tethered Ligands

A wide variety of entities can be tethered to an oligonucleotide agent, *e.g.*, to the carrier of a ligand-conjugated monomer. Examples are described below in the context of a ligand-
15 conjugated monomer but that is only one preferred embodiment. Entities can be coupled at other points to an oligonucleotide agent.

A ligand tethered to an oligonucleotide agent (*e.g.*, an oligonucleotide agent targeting an miRNA) can have a favorable effect on the agent. For example, the ligand can improve stability, hybridization thermodynamics with a target nucleic acid, targeting to a particular tissue or cell-
20 type, or cell permeability, *e.g.*, by an endocytosis-dependent or -independent mechanism. Ligands and associated modifications can also increase sequence specificity and consequently decrease off-site targeting.

A tethered ligand can include one or more modified bases or sugars that can function as intercalators. These are preferably located in an internal region, such as in a bulge of a

miRNA/target duplex. The intercalator can be an aromatic, *e.g.*, a polycyclic aromatic or heterocyclic aromatic compound. A polycyclic intercalator can have stacking capabilities, and can include systems with 2, 3, or 4 fused rings. The universal bases described herein can be included on a ligand.

5 In one embodiment, the ligand can include a cleaving group that contributes to target gene inhibition by cleavage of the target nucleic acid. The cleaving group can be, for example, a bleomycin (*e.g.*, bleomycin-A5, bleomycin-A2, or bleomycin-B2), pyrene, phenanthroline (*e.g.*, O-phenanthroline), a polyamine, a tripeptide (*e.g.*, lys-tyr-lys tripeptide), or metal ion chelating group. The metal ion chelating group can include, *e.g.*, an Lu(III) or EU(III) macrocyclic
10 complex, a Zn(II) 2,9-dimethylphenanthroline derivative, a Cu(II) terpyridine, or acridine, which can promote the selective cleavage of target RNA at the site of the bulge by free metal ions, such as Lu(III). In some embodiments, a peptide ligand can be tethered to a miRNA to promote cleavage of the target RNA, *e.g.*, at the bulge region. For example, 1,8-dimethyl-1,3,6,8,10,13-hexaazacyclotetradecane (cyclam) can be conjugated to a peptide (*e.g.*, by an amino acid
15 derivative) to promote target RNA cleavage.

A tethered ligand can be an aminoglycoside ligand, which can cause an oligonucleotide agent to have improved hybridization properties or improved sequence specificity. Exemplary aminoglycosides include glycosylated polylysine, galactosylated polylysine, neomycin B, tobramycin, kanamycin A, and acridine conjugates of aminoglycosides, such as Neo-N-acridine,
20 Neo-S-acridine, Neo-C-acridine, Tobra-N-acridine, and KanaA-N-acridine. Use of an acridine analog can increase sequence specificity. For example, neomycin B has a high affinity for RNA as compared to DNA, but low sequence-specificity. An acridine analog, neo-S-acridine has an increased affinity for the HIV Rev-response element (RRE). In some embodiments the guanidine analog (the guanidinoglycoside) of an aminoglycoside ligand is tethered to an
25 oligonucleotide agent. In a guanidinoglycoside, the amine group on the amino acid is exchanged for a guanidine group. Attachment of a guanidine analog can enhance cell permeability of an oligonucleotide agent, *e.g.*, an oligonucleotide agent targeting an miRNA or pre-miRNA..

A tethered ligand can be a poly-arginine peptide, peptoid or peptidomimetic, which can enhance the cellular uptake of an oligonucleotide agent.

30 Preferred moieties are ligands, which are coupled, preferably covalently, either directly or indirectly via an intervening tether, to the ligand-conjugated carrier. In preferred embodiments,

the ligand is attached to the carrier *via* an intervening tether. As discussed above, the ligand or tethered ligand may be present on the monomer when the monomer is incorporated into the growing strand. In some embodiments, the ligand may be incorporated into a “precursor” a ligand-conjugated monomer subunit after a “precursor” a ligand-conjugated monomer has been incorporated into the growing strand. For example, a monomer having, *e.g.*, an amino-terminated tether, *e.g.*, TAP-(CH₂)_nNH₂ may be incorporated into a growing oligonucleotide strand. In a subsequent operation, *i.e.*, after incorporation of the precursor monomer into the strand, a ligand having an electrophilic group, *e.g.*, a pentafluorophenyl ester or aldehyde group, can subsequently be attached to the precursor monomer subunit by coupling the electrophilic group of the ligand with the terminal nucleophilic group of the precursor monomer subunit tether.

In preferred embodiments, a ligand alters the distribution, targeting or lifetime of an oligonucleotide agent into which it is incorporated. In preferred embodiments a ligand provides an enhanced affinity for a selected target, *e.g.*, molecule, cell or cell type, compartment, *e.g.*, a cellular or organ compartment, tissue, organ or region of the body, as, *e.g.*, compared to a species absent such a ligand.

Preferred ligands can improve transport, hybridization, and specificity properties and may also improve nuclease resistance of the resultant natural or modified oligoribonucleotide, or a polymeric molecule comprising any combination of monomers described herein and/or natural or modified ribonucleotides.

Ligands in general can include therapeutic modifiers, *e.g.*, for enhancing uptake; diagnostic compounds or reporter groups *e.g.*, for monitoring distribution; cross-linking agents; nuclease-resistance conferring moieties; and natural or unusual nucleobases. General examples include lipophiles, lipids, steroids (*e.g.*, uvaol, hecigenin, diosgenin), terpenes (*e.g.*, triterpenes, *e.g.*, sarsasapogenin, Friedelin, epifriedelanol derivatized lithocholic acid), vitamins (*e.g.*, folic acid, vitamin A, biotin, pyridoxal), carbohydrates, proteins, protein binding agents, integrin targeting molecules, polycationics, peptides, polyamines, and peptide mimics.

Ligands can include a naturally occurring substance, (*e.g.*, human serum albumin (HSA), low-density lipoprotein (LDL), or globulin); carbohydrate (*e.g.*, a dextran, pullulan, chitin, chitosan, inulin, cyclodextrin or hyaluronic acid); amino acid, or a lipid. The ligand may also be a recombinant or synthetic molecule, such as a synthetic polymer, *e.g.*, a synthetic polyamino

acid. Examples of polyamino acids include polyamino acid is a polylysine (PLL), poly L-aspartic acid, poly L-glutamic acid, styrene-maleic acid anhydride copolymer, poly(L-lactide-co-glycolide) copolymer, divinyl ether-maleic anhydride copolymer, N-(2-hydroxypropyl)methacrylamide copolymer (HMPA), polyethylene glycol (PEG), polyvinyl alcohol (PVA), polyurethane, poly(2-ethylacrylic acid), N-isopropylacrylamide polymers, or polyphosphazene. Example of polyamines include: polyethylenimine, polylysine (PLL), spermine, spermidine, polyamine, pseudopeptide-polyamine, peptidomimetic polyamine, dendrimer polyamine, arginine, amidine, protamine, cationic lipid, cationic porphyrin, quaternary salt of a polyamine, or an alpha helical peptide.

Ligands can also include targeting groups, *e.g.*, a cell or tissue targeting agent, *e.g.*, a lectin, glycoprotein, lipid or protein, *e.g.*, an antibody, that binds to a specified cell type such as a kidney cell. A targeting group can be a thyrotropin, melanotropin, lectin, glycoprotein, surfactant protein A, Mucin carbohydrate, multivalent lactose, multivalent galactose, N-acetyl-galactosamine, N-acetyl-glucosamine, multivalent mannose, multivalent fucose, glycosylated polyaminoacids, multivalent galactose, transferrin, bisphosphonate, polyglutamate, polyaspartate, a lipid, cholesterol, a steroid, bile acid, folate, vitamin B12, biotin, or an RGD peptide or RGD peptide mimetic.

Other examples of ligands include dyes, intercalating agents (*e.g.* acridines and substituted acridines), cross-linkers (*e.g.* psoralene, mitomycin C), porphyrins (TPPC4, texaphyrin, Sapphyrin), polycyclic aromatic hydrocarbons (*e.g.*, phenazine, dihydrophenazine, phenanthroline, pyrenes), lys-tyr-lys tripeptide, aminoglycosides, guanidium aminoglycosides, artificial endonucleases (*e.g.* EDTA), lipophilic molecules, *e.g.* cholesterol (and thio analogs thereof), cholic acid, cholanic acid, lithocholic acid, adamantane acetic acid, 1-pyrene butyric acid, dihydrotestosterone, glycerol (*e.g.*, esters (*e.g.*, mono, bis, or tris fatty acid esters, *e.g.*, C₁₀, C₁₁, C₁₂, C₁₃, C₁₄, C₁₅, C₁₆, C₁₇, C₁₈, C₁₉, or C₂₀ fatty acids) and ethers thereof, *e.g.*, C₁₀, C₁₁, C₁₂, C₁₃, C₁₄, C₁₅, C₁₆, C₁₇, C₁₈, C₁₉, or C₂₀ alkyl; *e.g.*, 1,3-bis-O(hexadecyl)glycerol, 1,3-bis-O(octaadecyl)glycerol), geranyloxyhexyl group, hexadecylglycerol, borneol, menthol, 1,3-propanediol, heptadecyl group, palmitic acid, stearic acid (*e.g.*, glyceryl distearate), oleic acid, myristic acid, O3-(oleoyl)lithocholic acid, O3-(oleoyl)cholenic acid, dimethoxytrityl, or phenoxazine) and peptide conjugates (*e.g.*, antennapedia peptide, Tat peptide), alkylating agents, phosphate, amino, mercapto, PEG (*e.g.*, PEG-40K), MPEG, [MPEG]₂, polyamino, alkyl,

substituted alkyl, radiolabeled markers, enzymes, haptens (*e.g.* biotin), transport/absorption facilitators (*e.g.*, aspirin, naproxen, vitamin E, folic acid), synthetic ribonucleases (*e.g.*, imidazole, bisimidazole, histamine, imidazole clusters, acridine-imidazole conjugates, Eu³⁺ complexes of tetraazamacrocycles), dinitrophenyl, HRP, or AP.

5 Ligands can be proteins, *e.g.*, glycoproteins, or peptides, *e.g.*, molecules having a specific affinity for a co-ligand, or antibodies *e.g.*, an antibody, that binds to a specified cell type such as a cancer cell, endothelial cell, or bone cell. Ligands may also include hormones and hormone receptors. They can also include non-peptidic species, such as lipids, lectins, carbohydrates, vitamins, cofactors, multivalent lactose, multivalent galactose, N-acetyl-galactosamine, N-acetyl-
10 gulucosamine multivalent mannose, or multivalent fucose. The ligand can be, for example, a lipopolysaccharide, an activator of p38 MAP kinase, or an activator of NF- κ B.

 The ligand can be a substance, *e.g.*, a drug, which can increase the uptake of the oligonucleotide agent into the cell, for example, by disrupting the cell's cytoskeleton, *e.g.*, by disrupting the cell's microtubules, microfilaments, and/or intermediate filaments. The drug can
15 be, for example, taxon, vincristine, vinblastine, cytochalasin, nocodazole, japlakinolide, latrunculin A, phalloidin, swinholide A, indanocine, or myoservin.

 The ligand can increase the uptake of the oligonucleotide agent into the cell by activating an inflammatory response, for example. Exemplary ligands that would have such an effect include tumor necrosis factor alpha (TNFalpha), interleukin-1 beta, or gamma interferon.

20 In one aspect, the ligand is a lipid or lipid-based molecule. Such a lipid or lipid-based molecule preferably binds a serum protein, *e.g.*, human serum albumin (HSA). An HSA binding ligand allows for distribution of the conjugate to a target tissue, *e.g.*, a non-kidney target tissue of the body. For example, the target tissue can be the liver, including parenchymal cells of the liver. Other molecules that can bind HSA can also be used as ligands. For example, neproxin or
25 aspirin can be used. A lipid or lipid-based ligand can (a) increase resistance to degradation of the conjugate, (b) increase targeting or transport into a target cell or cell membrane, and/or (c) can be used to adjust binding to a serum protein, *e.g.*, HSA.

 A lipid based ligand can be used to modulate, *e.g.*, control the binding of the conjugate to a target tissue. For example, a lipid or lipid-based ligand that binds to HSA more strongly will
30 be less likely to be targeted to the kidney and therefore less likely to be cleared from the body.

A lipid or lipid-based ligand that binds to HSA less strongly can be used to target the conjugate to the kidney.

In a preferred embodiment, the lipid based ligand binds HSA. A lipid-based ligand can bind HSA with a sufficient affinity such that the conjugate will be preferably distributed to a non-kidney tissue. However, it is preferred that the affinity not be so strong that the HSA-ligand binding cannot be reversed.

In another preferred embodiment, the lipid based ligand binds HSA weakly or not at all, such that the conjugate will be preferably distributed to the kidney. Other moieties that target to kidney cells can also be used in place of or in addition to the lipid based ligand.

In another aspect, the ligand is a moiety, *e.g.*, a vitamin, which is taken up by a target cell, *e.g.*, a proliferating cell. These are particularly useful for treating disorders characterized by unwanted cell proliferation, *e.g.*, of the malignant or non-malignant type, *e.g.*, cancer cells. Exemplary vitamins include vitamin A, E, and K. Other exemplary vitamins include are B vitamin, *e.g.*, folic acid, B12, riboflavin, biotin, pyridoxal or other vitamins or nutrients taken up by cancer cells. Also included are HSA and low density lipoprotein (LDL).

In another aspect, the ligand is a cell-permeation agent, preferably a helical cell-permeation agent. Preferably, the agent is amphipathic. An exemplary agent is a peptide such as tat or antennopodia. If the agent is a peptide, it can be modified, including a peptidylmimetic, invertomers, non-peptide or pseudo-peptide linkages, and use of D-amino acids. The helical agent is preferably an alpha-helical agent, which preferably has a lipophilic and a lipophobic phase.

Peptides that target markers enriched in proliferating cells can be used. *E.g.*, RGD containing peptides and peptidomimetics can target cancer cells, in particular cells that exhibit an $\alpha_v\beta_3$ integrin. Thus, one could use RGD peptides, cyclic peptides containing RGD, RGD peptides that include D-amino acids, as well as synthetic RGD mimics. In addition to RGD, one can use other moieties that target the $\alpha_v\beta_3$ integrin ligand. Generally, such ligands can be used to control proliferating cells and angiogenesis. Preferred conjugates of this type include an oligonucleotide agent that targets PECAM-1, VEGF, or other cancer gene, *e.g.*, a cancer gene described herein.

The oligonucleotide agents of the invention are particularly useful when targeted to the liver. For example, a single stranded oligonucleotide agent featured in the invention can target

an miRNA enriched in the liver, and the oligonucleotide agent can include a ligand for enhanced delivery to the liver. An oligonucleotide agent can be targeted to the liver by incorporation of a monomer derivatized with a ligand which targets to the liver. For example, a liver-targeting agent can be a lipophilic moiety. Preferred lipophilic moieties include lipid, cholesterol, oleyl, retinyl, or cholesteryl residues. Other lipophilic moieties that can function as liver-targeting agents include cholic acid, adamantane acetic acid, 1-pyrene butyric acid, dihydrotestosterone, 1,3-Bis-O(hexadecyl)glycerol, geranyloxyhexyl group, hexadecylglycerol, borneol, menthol, 1,3-propanediol, heptadecyl group, palmitic acid, myristic acid, O3-(oleoyl)lithocholic acid, O3-(oleoyl)cholenic acid, dimethoxytrityl, or phenoxazine.

An oligonucleotide agent can also be targeted to the liver by association with a low-density lipoprotein (LDL), such as lactosylated LDL. Polymeric carriers complexed with sugar residues can also function to target oligonucleotide agents to the liver.

A targeting agent that incorporates a sugar, *e.g.*, galactose and/or analogues thereof, is particularly useful. These agents target, in particular, the parenchymal cells of the liver (see Table 2). For example, a targeting moiety can include more than one or preferably two or three galactose moieties, spaced about 15 angstroms from each other. The targeting moiety can alternatively be lactose (*e.g.*, three lactose moieties), which is glucose coupled to a galactose. The targeting moiety can also be N-Acetyl-Galactosamine, N-Ac-Glucosamine. A mannose or mannose-6-phosphate targeting moiety can be used for macrophage targeting.

The ligand can be a peptide or peptidomimetic. A peptidomimetic (also referred to herein as an oligopeptidomimetic) is a molecule capable of folding into a defined three-dimensional structure similar to a natural peptide. The attachment of peptide and peptidomimetics to oligonucleotide agents can affect pharmacokinetic distribution of the iRNA, such as by enhancing cellular recognition and absorption. The peptide or peptidomimetic moiety can be about 5-50 amino acids long, *e.g.*, about 5, 10, 15, 20, 25, 30, 35, 40, 45, or 50 amino acids long (see Table 2, for example).

Table 2. Exemplary Cell Permeation Peptides

Cell Permeation Peptide	Amino acid Sequence	Reference
Penetratin	RQIKIWFQNRRMKWKK (SEQ ID NO:1)	Derossi <i>et al.</i> , J. Biol. Chem. 269:10444,

		1994
Tat fragment (48-60)	GRKKRRQRRRPPQC (SEQ ID NO:2)	Vives <i>et al.</i> , J. Biol. Chem., 272:16010, 1997
Signal Sequence- based peptide	GALFLGWLGAAGSTMGAWSQPKKKRKV (SEQ ID NO:3)	Chaloin <i>et al.</i> , Biochem. Biophys. Res. Commun., 243:601, 1998
PVEC	LLIILRRRIRKQAHAAHSK (SEQ ID NO:4)	Elmqvist <i>et al.</i> , Exp. Cell Res., 269:237, 2001
Transportan	GWTLNSAGYLLKINLKALAALAKKIL (SEQ ID NO:5)	Pooga <i>et al.</i> , FASEB J., 12:67, 1998
Amphiphilic model peptide	KLALKLALKALKAALKLA (SEQ ID NO:6)	Oehlke <i>et al.</i> , Mol. Ther., 2:339, 2000
Arg ₉	RRRRRRRRR (SEQ ID NO:7)	Mitchell <i>et al.</i> , J. Pept. Res., 56:318, 2000
Bacterial cell wall permeating	KFFKFFKFFK (SEQ ID NO:8)	
LL-37	LLGDFFRKSKEKIGKEFKRIVQRIKDFLRN LVPRTES (SEQ ID NO:9)	
Cecropin P1	SWLSKTAKKLENSAKKRISGIAIAIQGGP R (SEQ ID NO:10)	
α -defensin	ACYCRIPACIAGERRYGTCTYQGRLWAFC C (SEQ ID NO:11)	
b-defensin	DHYNCVSSGGQCLYSACPIFTKIQTCTYR GKAKCCK (SEQ ID NO:12)	
Bactenecin	RKCRIVVIRVCR (SEQ ID NO:13)	
PR-39	RRRPRPPYLPRPRPPFFPPRLPPRIPPGFPP RFPPRFPGKR-NH ₂ (SEQ ID NO:14)	
Indolicidin	ILPWKWPWWPWRR-NH ₂ (SEQ ID NO:15)	

A peptide or peptidomimetic can be, for example, a cell permeation peptide, cationic peptide, amphipathic peptide, or hydrophobic peptide (*e.g.*, consisting primarily of Tyr, Trp or Phe). The peptide moiety can be a dendrimer peptide, constrained peptide or crosslinked

peptide. In another alternative, the peptide moiety can include a hydrophobic membrane translocation sequence (MTS). An exemplary hydrophobic MTS-containing peptide is RFGF having the amino acid sequence AAVALLPAVLLALLAP (SEQ ID NO:16). An RFGF analogue (*e.g.*, amino acid sequence AALLPVLLAAP (SEQ ID NO:17)) containing a hydrophobic MTS can also be a targeting moiety. The peptide moiety can be a “delivery” peptide, which can carry large polar molecules including peptides, oligonucleotides, and proteins across cell membranes. For example, sequences from the HIV Tat protein (GRKKRRQRRRPPQ (SEQ ID NO:18)) and the Drosophila Antennapedia protein (RQIKIWFQNRRMKWKK (SEQ ID NO:19)) have been found to be capable of functioning as delivery peptides. A peptide or peptidomimetic can be encoded by a random sequence of DNA, such as a peptide identified from a phage-display library, or one-bead-one-compound (OBOC) combinatorial library (Lam *et al.*, *Nature*, 354:82-84, 1991). Preferably the peptide or peptidomimetic tethered to an oligonucleotide agent via an incorporated monomer unit is a cell targeting peptide such as an arginine-glycine-aspartic acid (RGD)-peptide, or RGD mimic. A peptide moiety can range in length from about 5 amino acids to about 40 amino acids. The peptide moieties can have a structural modification, such as to increase stability or direct conformational properties. Any of the structural modifications described below can be utilized.

An RGD peptide moiety can be used to target a tumor cell, such as an endothelial tumor cell or a breast cancer tumor cell (Zitzmann *et al.*, *Cancer Res.*, 62:5139-43, 2002). An RGD peptide can facilitate targeting of an oligonucleotide agent (*e.g.*, an oligonucleotide agent targeting an miRNA or pre-miRNA) to tumors of a variety of other tissues, including the lung, kidney, spleen, or liver (Aoki *et al.*, *Cancer Gene Therapy* 8:783-787, 2001). Preferably, the RGD peptide will facilitate targeting of an oligonucleotide agent to the kidney. The RGD peptide can be linear or cyclic, and can be modified, *e.g.*, glycosylated or methylated to facilitate targeting to specific tissues. For example, a glycosylated RGD peptide can deliver an oligonucleotide agent to a tumor cell expressing $\alpha_v\beta_3$ (Haubner *et al.*, *Jour. Nucl. Med.*, 42:326-336, 2001).

Peptides that target markers enriched in proliferating cells can be used. *E.g.*, RGD containing peptides and peptidomimetics can target cancer cells, in particular cells that exhibit an $\alpha_v\beta_3$ integrin. Thus, one could use RGD peptides, cyclic peptides containing RGD, RGD

peptides that include D-amino acids, as well as synthetic RGD mimics. In addition to RGD, one can use other moieties that target the $\alpha_v\beta_3$ integrin ligand. Generally, such ligands can be used to control proliferating cells and angiogenesis. Preferred conjugates of this type include an oligonucleotide agent that targets PECAM-1, VEGF, or other cancer gene, *e.g.*, a cancer gene
5 described herein.

A "cell permeation peptide" is capable of permeating a cell, *e.g.*, a microbial cell, such as a bacterial or fungal cell, or a mammalian cell, such as a human cell. A microbial cell-permeating peptide can be, for example, an α -helical linear peptide (*e.g.*, LL-37 or Ceropin P1), a disulfide bond-containing peptide (*e.g.*, α -defensin, β -defensin or bactenecin), or a peptide
10 containing only one or two dominating amino acids (*e.g.*, PR-39 or indolicidin). A cell permeation peptide can also include a nuclear localization signal (NLS). For example, a cell permeation peptide can be a bipartite amphipathic peptide, such as MPG, which is derived from the fusion peptide domain of HIV-1 gp41 and the NLS of SV40 large T antigen (Simeoni *et al.*, *Nucl. Acids Res.* 31:2717-2724, 2003).

15 In one embodiment, a targeting peptide tethered to a ligand-conjugated monomer can be an amphipathic α -helical peptide. Exemplary amphipathic α -helical peptides include, but are not limited to, cecropins, lycotoxins, paradaxins, buforin, CPF, bombinin-like peptide (BLP), cathelicidins, ceratotoxins, *S. clava* peptides, hagfish intestinal antimicrobial peptides (HFIAPs), magainines, brevinins-2, dermaseptins, melittins, pleurocidin, H₂A peptides, *Xenopus* peptides,
20 esculentinis-1, and caerins. A number of factors will preferably be considered to maintain the integrity of helix stability. For example, a maximum number of helix stabilization residues will be utilized (*e.g.*, leu, ala, or lys), and a minimum number of helix destabilization residues will be utilized (*e.g.*, proline, or cyclic monomeric units). The capping residue will be considered (for example Gly is an exemplary N-capping residue) and/or C-terminal amidation can be used to
25 provide an extra H-bond to stabilize the helix. Formation of salt bridges between residues with opposite charges, separated by $i \pm 3$, or $i \pm 4$ positions can provide stability. For example, cationic residues such as lysine, arginine, homo-arginine, ornithine or histidine can form salt bridges with the anionic residues glutamate or aspartate.

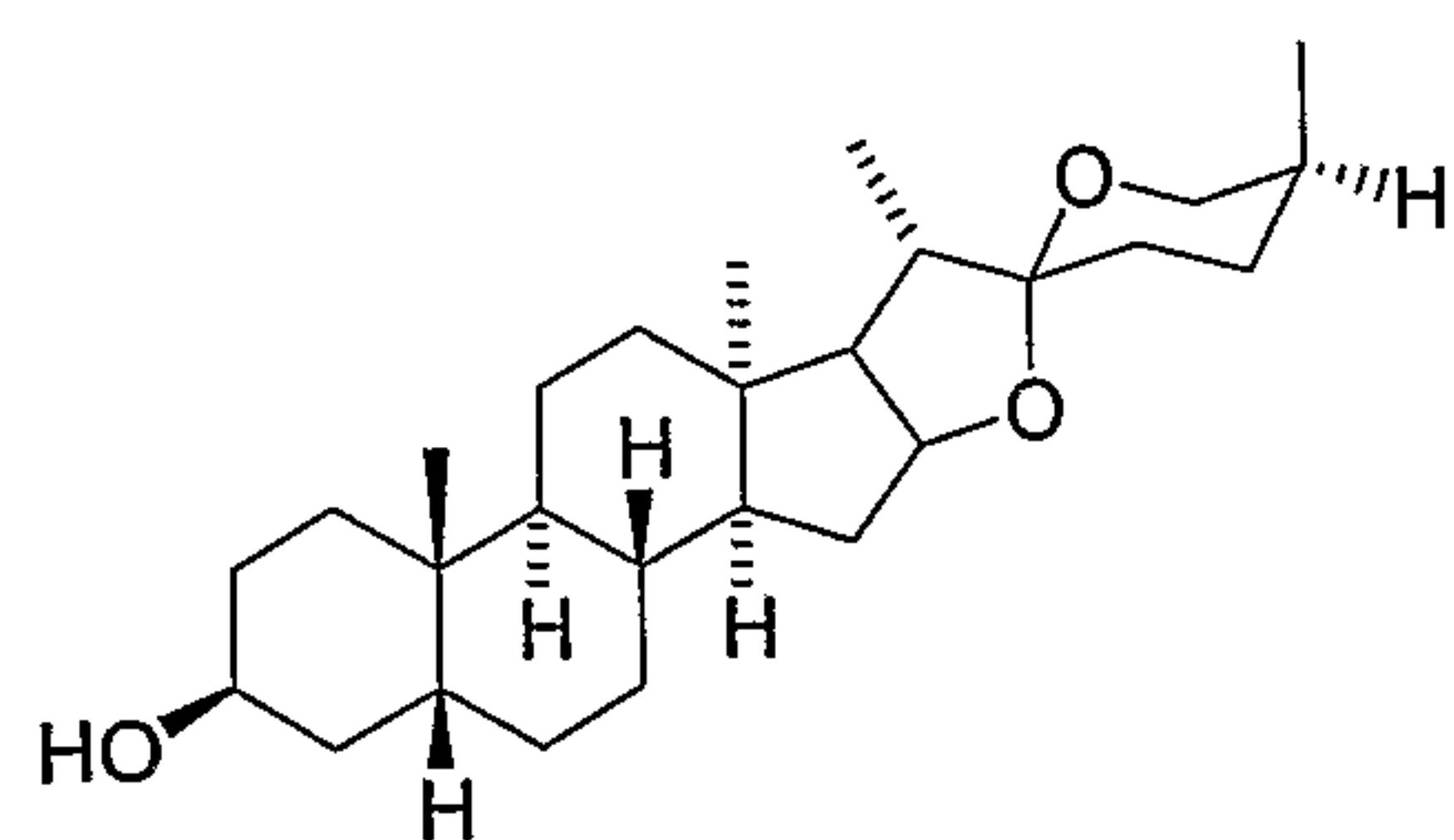
Peptide and peptidomimetic ligands include those having naturally occurring or modified peptides, *e.g.*, D or L peptides; α , β , or γ peptides; N-methyl peptides; azapeptides; peptides having one or more amide, *i.e.*, peptide, linkages replaced with one or more urea, thiourea, carbamate, or sulfonyl urea linkages; or cyclic peptides.

5 In some embodiments, the peptide can have a cationic and/or a hydrophobic moiety.

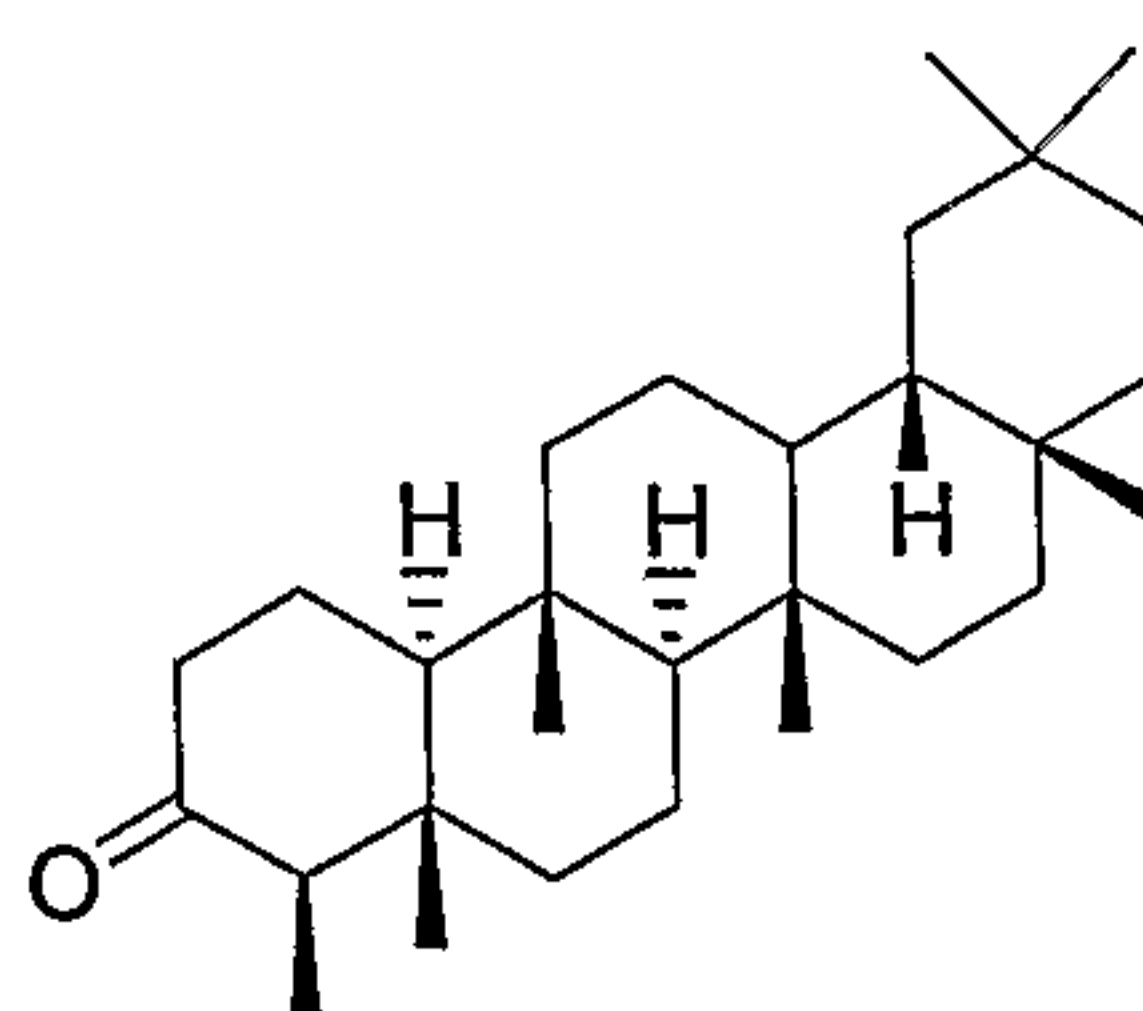
In some embodiments, the ligand can be any of the nucleobases described herein.

In some embodiments, the ligand can be a substituted amine, *e.g.* dimethylamino. In some embodiments, the substituted amine can be quaternized, *e.g.*, by protonation or alkylation, rendering it cationic. In some embodiments, the substituted amine can be at the terminal position
10 of a relatively hydrophobic tether, *e.g.*, alkylene.

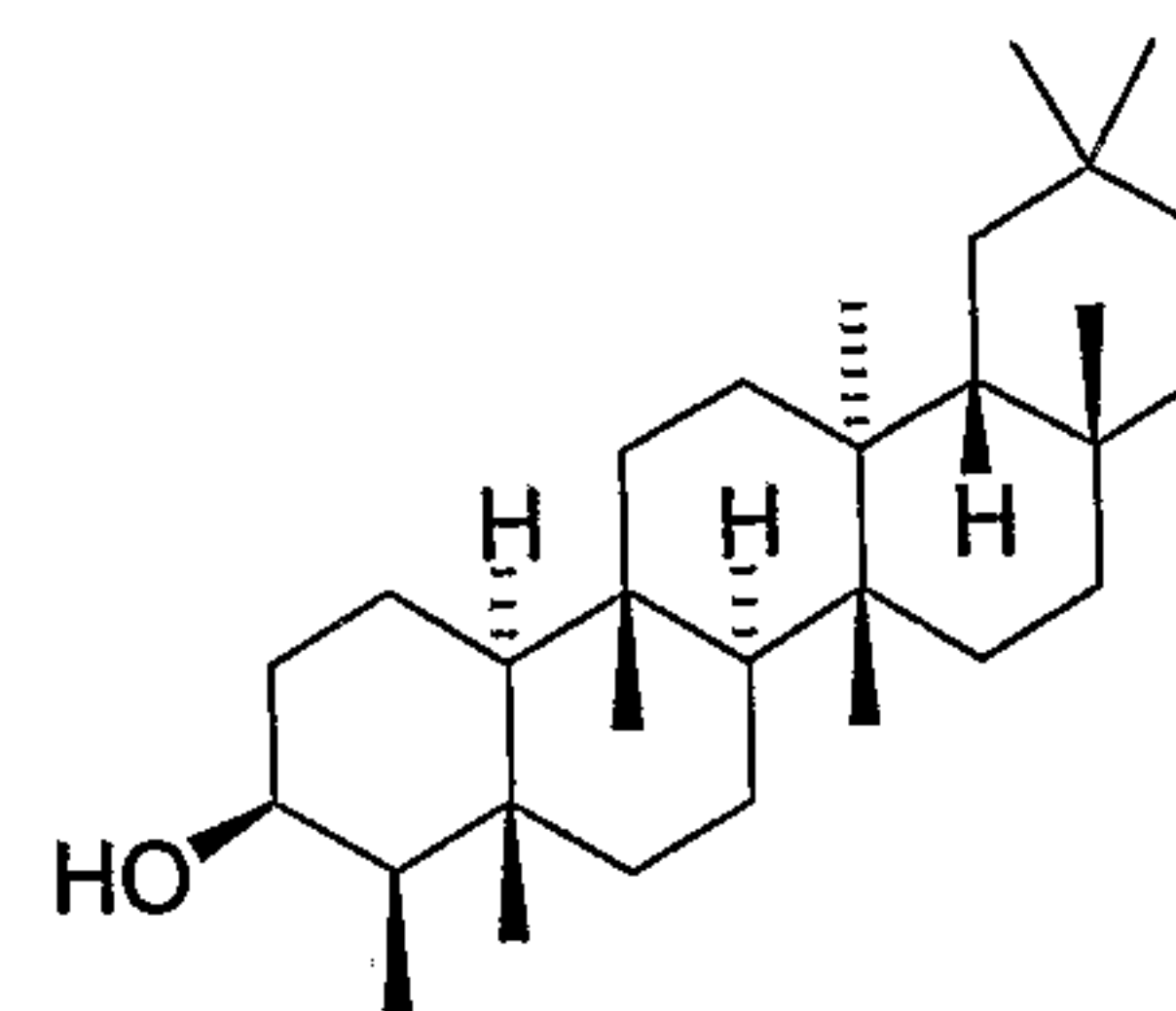
In some embodiments, the ligand can be one of the following triterpenes:



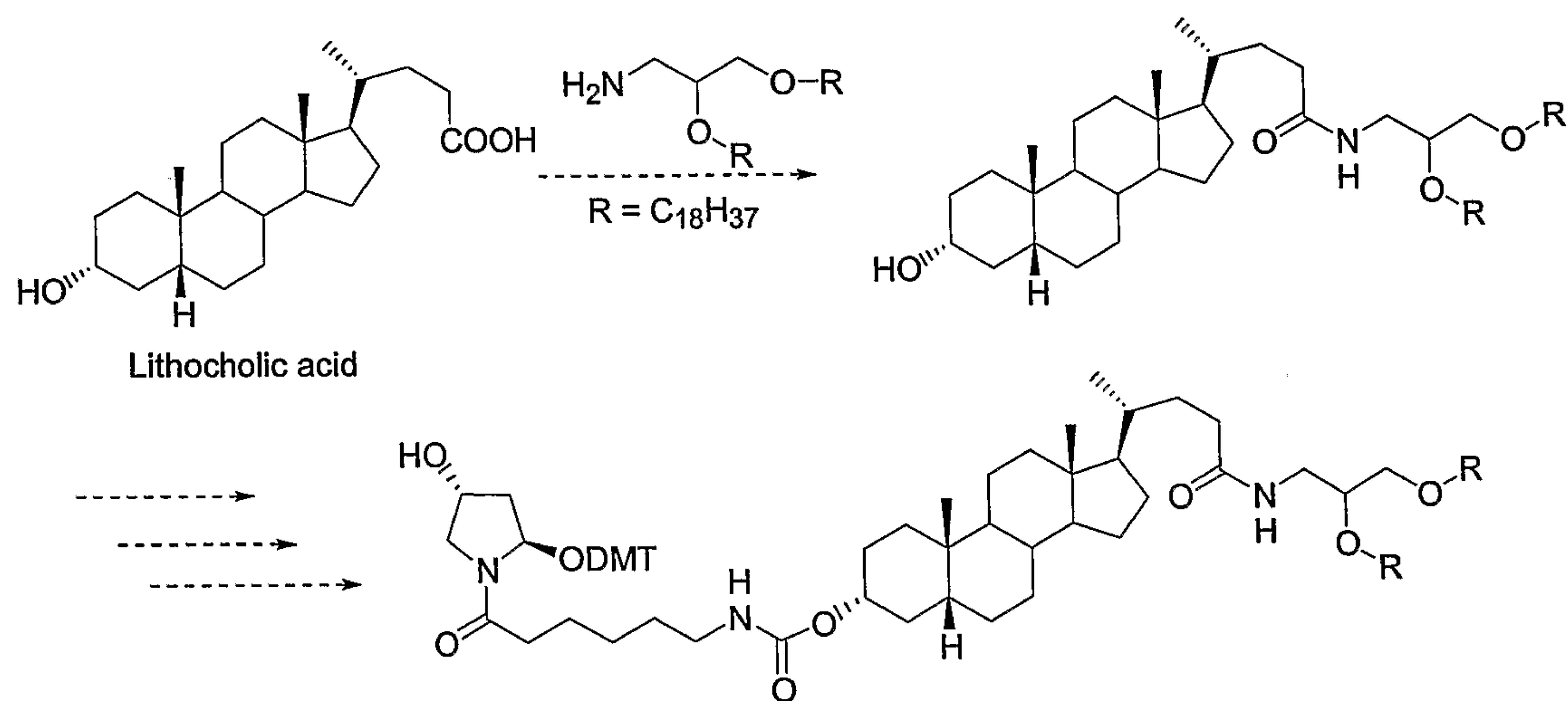
Sarsasapogenin



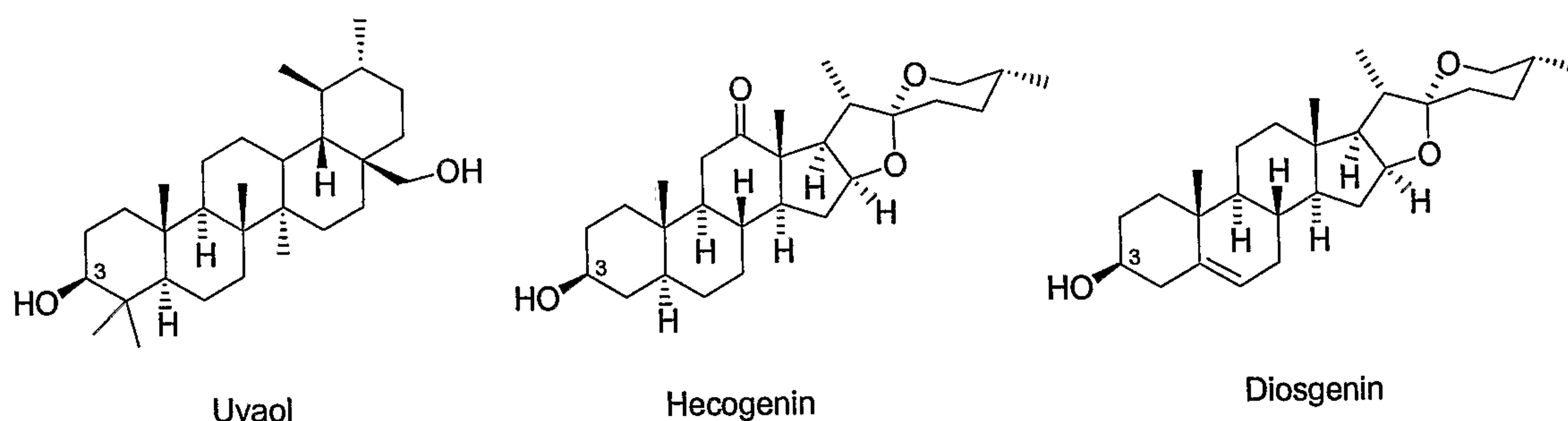
Friedelin



Epifriedelanol



In some embodiments, the ligand can be substituted or unsubstituted cholesterol, or a stereoisomer thereof or one of the following steroids:



5 In some embodiments, a tethered ligand can contain one or more atoms than the corresponding untethered or uncoupled ligand (*e.g.*, one or more protons of a heteroatom-based functional group or an entire heteroatom-based functional group may be displaced from the uncoupled ligand during coupling of a ligand to a carrier or tether). For example, the proton of the 3-hydroxy group of cholesterol can be replaced by a tether (*e.g.*, Chol-3-OH (uncoupled) and
10 Chol-3-O-tether (coupled)) or the entire 3-hydroxy group of cholesterol can be replaced by a sulfur atom (*e.g.*, Chol-3-OH (uncoupled) and Chol-3-S-tether (coupled, *e.g.*, thiocholesterol)).

Methods for making oligonucleotide agents

15 A listing of ribonucleosides containing the unusual bases described herein are described in "The RNA Modification Database" maintained by Pamela F. Crain, Jef Rozenski and James A. McCloskey; Departments of Medicinal Chemistry and Biochemistry, University of Utah, Salt Lake City, UT 84112, USA.

The 5' silyl protecting group can be used in conjunction with acid labile orthoesters at the 2' position of ribonucleosides to synthesize oligonucleotides via phosphoramidite chemistry.
20 Final deprotection conditions are known not to significantly degrade RNA products. Functional groups on the unusual and universal bases are blocked during oligonucleotide synthesis with protecting groups that are compatible with the operations being performed that are described herein. All syntheses can be conducted in any automated or manual synthesizer on large, medium, or small scale. The syntheses may also be carried out in multiple well plates or glass
25 slides.

The 5'-O-silyl group can be removed via exposure to fluoride ions, which can include any source of fluoride ion, *e.g.*, those salts containing fluoride ion paired with inorganic counterions *e.g.*, cesium fluoride and potassium fluoride or those salts containing fluoride ion paired with an organic counterion, *e.g.*, a tetraalkylammonium fluoride. A crown ether catalyst can be utilized
5 in combination with the inorganic fluoride in the deprotection reaction. Preferred fluoride ion source are tetrabutylammonium fluoride or aminehydrofluorides (*e.g.*, combining aqueous HF with triethylamine in a dipolar aprotic solvent, *e.g.*, dimethylformamide).

The choice of protecting groups for use on the phosphite triesters and phosphotriesters can alter the stability of the triesters towards fluoride. Methyl protection of the phosphotriester
10 or phosphitetriester can stabilize the linkage against fluoride ions and improve process yields.

Since ribonucleosides have a reactive 2' hydroxyl substituent, it can be desirable to protect the reactive 2' position in RNA with a protecting group that is compatible with a 5'-O-silyl protecting group, *e.g.* one stable to fluoride. Orthoesters meet this criterion and can be readily removed in a final acid deprotection step that can result in minimal RNA degradation.

15 Tetrazole catalysts can be used in the standard phosphoramidite coupling reaction. Preferred catalysts include *e.g.* tetrazole, S-ethyl-tetrazole, p-nitrophenyltetrazole.

The general process is as follows. Nucleosides are suitably protected and functionalized for use in solid-phase or solution-phase synthesis of RNA oligonucleotides. The 2'-hydroxyl group in a ribonucleotide can be modified using a tris orthoester reagent. The 2'-hydroxyl can be
20 modified to yield a 2'-O-orthoester nucleoside by reacting the ribonucleoside with the tris orthoester reagent in the presence of an acidic catalyst, *e.g.*, pyridinium p-toluene sulfonate. This reaction is known to those skilled in the art. The product can then be subjected to further protecting group reactions (*e.g.*, 5'-O-silylation) and functionalizations (*e.g.*, 3'-O-phosphitylation) to produce a desired reagent (*e.g.*, nucleoside phosphoramidite) for
25 incorporation within an oligonucleotide or polymer by reactions known to those skilled in the art.

Preferred orthoesters include those comprising ethylene glycol ligands which are protected with acyl or ester protecting groups. Specifically, the preferred acyl group is acetyl. The nucleoside reagents may then be used by those skilled in the art to synthesize RNA oligonucleotides on commercially available synthesizer instruments, *e.g.*, Gene Assembler Plus
30 (Pharmacia), 380B (Applied Biosystems). Following synthesis (either solution-phase or solid-phase) of an oligonucleotide or polymer, the product can be subjected to one or more reactions

using non-acidic reagents. One of these reactions may be strong basic conditions, for example, 40% methylamine in water for 10 minutes at 55°C, which will remove the acyl protecting groups from the ethylene glycol ligands but leave the orthoester moiety attached. The resultant orthoester may be left attached when the polymer or oligonucleotide is used in subsequent applications, or it may be removed in a final mildly-acidic reaction, for example, 10 minutes at 55°C in 50 mM acetic acid, pH 3.0, followed by addition of equal volume of 150 mM TRIS buffer for 10 minutes at 55°C.

Universal bases are described in "Survey and Summary: The Applications of Universal DNA base analogues" Loakes, D., *Nucleic Acid Research* **2001**, *29*, 2437, which is incorporated by reference in its entirety. Specific examples are described in the following: Liu, D.; Moran, S.; Kool, E. T. *Chem. Biol.*, **1997**, *4*, 919-926; Morales, J. C.; Kool, E. T. *Biochemistry*, **2000**, *39*, 2626-2632; Matray, T. J.; Kool, E. T. *J. Am. Chem. Soc.*, **1998**, *120*, 6191-6192; Moran, S. Ren, R. X.-F.; Rumney IV, S.; Kool, E. T. *J. Am. Chem. Soc.*, **1997**, *119*, 2056-2057; Guckian, K. M.; Morales, J. C.; Kool, E. T. *J. Org. Chem.*, **1998**, *63*, 9652-9656; Berger, M.; Wu, Y.; Ogawa, A. K.; McMinn, D. L.; Schultz, P. G.; Romesberg, F. E. *Nucleic Acids Res.*, **2000**, *28*, 2911-2914; Ogawa, A. K.; Wu, Y.; McMinn, D. L.; Liu, J.; Schultz, P. G.; Romesberg, F. E. *J. Am. Chem. Soc.*, **2000**, *122*, 3274-3287; Ogawa, A. K.; Wu, Y.; Berger, M.; Schultz, P. G.; Romesberg, F. E. *J. Am. Chem. Soc.*, **2000**, *122*, 8803-8804; Tae, E. L.; Wu, Y.; Xia, G.; Schultz, P. G.; Romesberg, F. E. *J. Am. Chem. Soc.*, **2001**, *123*, 7439-7440; Wu, Y.; Ogawa, A. K.; Berger, M.; McMinn, D. L.; Schultz, P. G.; Romesberg, F. E. *J. Am. Chem. Soc.*, **2000**, *122*, 7621-7632; McMinn, D. L.; Ogawa, A. K.; Wu, Y.; Liu, J.; Schultz, P. G.; Romesberg, F. E. *J. Am. Chem. Soc.*, **1999**, *121*, 11585-11586; Brotschi, C.; Haberli, A.; Leumann, C. *Angew. Chem. Int. Ed.*, **2001**, *40*, 3012-3014; Weizman, H.; Tor, Y. *J. Am. Chem. Soc.*, **2001**, *123*, 3375-3376; Lan, T.; McLaughlin, L. W. *J. Am. Chem. Soc.*, **2000**, *122*, 6512-13.

As discussed above, the monomers and methods described herein can be used in the preparation of modified RNA molecules, or polymeric molecules comprising any combination of monomer compounds described herein and/or natural or modified ribonucleotides in which one or more subunits contain an unusual or universal base. Modified RNA molecules include *e.g.* those molecules containing a chemically or stereochemically modified nucleoside (*e.g.*, having one or more backbone modifications, *e.g.*, phosphorothioate or P-alkyl; having one or more

sugar modifications, *e.g.*, 2'-OCH₃ or 2'-F; and/or having one or more base modifications, *e.g.*, 5-alkylamino or 5-allylamino) or a nucleoside surrogate.

Coupling of 5'-hydroxyl groups with phosphoramidites forms phosphite ester intermediates, which in turn are oxidized *e.g.*, with iodine, to the phosphate diester.

5 Alternatively, the phosphites may be treated with, *e.g.*, sulfur, selenium, amino, and boron reagents to form modified phosphate backbones. Linkages between the monomers described herein and a nucleoside or oligonucleotide chain can also be treated with iodine, sulfur, selenium, amino, and boron reagents to form unmodified and modified phosphate backbones respectively. Similarly, the monomers described herein may be coupled with nucleosides or oligonucleotides
10 containing any of the modifications or nucleoside surrogates described herein.

The synthesis and purification of oligonucleotide peptide conjugates can be performed by established methods. See, for example, Trufert *et al.*, Tetrahedron, 52:3005, 1996; and Manoharan, "Oligonucleotide Conjugates in Antisense Technology," in Antisense Drug Technology, ed. S.T. Crooke, Marcel Dekker, Inc., 2001. Exemplary methods are shown in
15 FIGs. 4 and 5.

In one embodiment of the invention, a peptidomimetic can be modified to create a constrained peptide that adopts a distinct and specific preferred conformation, which can increase the potency and selectivity of the peptide. For example, the constrained peptide can be an azapeptide (Gante, *Synthesis*, 1989, 405-413). An azapeptide is synthesized by replacing the
20 α -carbon of an amino acid with a nitrogen atom without changing the structure of the amino acid side chain. For example, the azapeptide can be synthesized by using hydrazine in traditional peptide synthesis coupling methods, such as by reacting hydrazine with a "carbonyl donor," *e.g.*, phenylchloroformate. A general azapeptide synthesis is shown in FIG. 6.

In one embodiment of the invention, a peptide or peptidomimetic (*e.g.*, a peptide or
25 peptidomimetic tethered to an ligand-conjugated monomer) can be an N-methyl peptide. N-methyl peptides are composed of N-methyl amino acids, which provide an additional methyl group in the peptide backbone, thereby potentially providing additional means of resistance to proteolytic cleavage. N-methyl peptides can be synthesized by methods known in the art (see, for example, Lindgren *et al.*, Trends Pharmacol. Sci. 21:99, 2000; Cell Penetrating Peptides: Processes and Applications, Langel, ed., CRC Press, Boca Raton, FL, 2002; Fische *et al.*,
30 Bioconjugate. Chem. 12: 825, 2001; Wander *et al.*, J. Am. Chem. Soc., 124:13382, 2002). For

example, an Ant or Tat peptide can be an N-methyl peptide. An exemplary synthesis is shown in FIG. 7.

10 In one embodiment of the invention, a peptide or peptidomimetic (*e.g.*, a peptide or peptidomimetic tethered to a ligand-conjugated monomer) can be a β -peptide. β -peptides form
5 stable secondary structures such as helices, pleated sheets, turns and hairpins in solutions. Their cyclic derivatives can fold into nanotubes in the solid state. β -peptides are resistant to degradation by proteolytic enzymes. β -peptides can be synthesized by methods known in the art. For example, an Ant or Tat peptide can be a β -peptide. An exemplary synthesis is shown in FIG. 8.

10 In one embodiment of the invention, a peptide or peptidomimetic (*e.g.*, a peptide or peptidomimetic tethered to a ligand-conjugated monomer) can be an oligocarbamate. Oligocarbamate peptides are internalized into a cell by a transport pathway facilitated by carbamate transporters. For example, an Ant or Tat peptide can be an oligocarbamate. An exemplary synthesis is shown in FIG. 9.

15 In one embodiment of the invention, a peptide or peptidomimetic (*e.g.*, a peptide or peptidomimetic tethered to a ligand-conjugated monomer) can be an oligourea conjugate (or an oligothiourea conjugate), in which the amide bond of a peptidomimetic is replaced with a urea moiety. Replacement of the amide bond provides increased resistance to degradation by
20 proteolytic enzymes, *e.g.*, proteolytic enzymes in the gastrointestinal tract. In one embodiment, an oligourea conjugate is tethered to an oligonucleotide agent for use in oral delivery. The backbone in each repeating unit of an oligourea peptidomimetic can be extended by one carbon atom in comparison with the natural amino acid. The single carbon atom extension can increase peptide stability and lipophilicity, for example. An oligourea peptide can therefore be advantageous when an oligonucleotide agent is directed for passage through a bacterial cell wall,
25 or when an oligonucleotide agent must traverse the blood-brain barrier, such as for the treatment of a neurological disorder. In one embodiment, a hydrogen bonding unit is conjugated to the oligourea peptide, such as to create an increased affinity with a receptor. For example, an Ant or Tat peptide can be an oligourea conjugate (or an oligothiourea conjugate). An exemplary synthesis is shown in FIG. 10.

30 The siRNA peptide conjugates of the invention can be affiliated with, *e.g.*, tethered to, ligand-conjugated monomers occurring at various positions on an oligonucleotide agent. For

example, a peptide can be terminally conjugated, on either the sense or the antisense strand, or a peptide can be bisconjugated (one peptide tethered to each end, one conjugated to the sense strand, and one conjugated to the antisense strand). In another option, the peptide can be internally conjugated, such as in the loop of a short hairpin oligonucleotide agent. In yet another option, the peptide can be affiliated with a complex, such as a peptide-carrier complex.

A peptide-carrier complex consists of at least a carrier molecule, which can encapsulate one or more oligonucleotide agents (such as for delivery to a biological system and/or a cell), and a peptide moiety tethered to the outside of the carrier molecule, such as for targeting the carrier complex to a particular tissue or cell type. A carrier complex can carry additional targeting molecules on the exterior of the complex, or fusogenic agents to aid in cell delivery. The one or more oligonucleotide agents encapsulated within the carrier can be conjugated to lipophilic molecules, which can aid in the delivery of the agents to the interior of the carrier.

A carrier molecule or structure can be, for example, a micelle, a liposome (*e.g.*, a cationic liposome), a nanoparticle, a microsphere, or a biodegradable polymer. A peptide moiety can be tethered to the carrier molecule by a variety of linkages, such as a disulfide linkage, an acid labile linkage, a peptide-based linkage, an oxyamino linkage or a hydrazine linkage. For example, a peptide-based linkage can be a GFLG peptide. Certain linkages will have particular advantages, and the advantages (or disadvantages) can be considered depending on the tissue target or intended use. For example, peptide based linkages are stable in the blood stream but are susceptible to enzymatic cleavage in the lysosomes. A schematic of preferred carriers is shown in FIG. 11.

The protected monomer compounds can be separated from a reaction mixture and further purified by a method such as column chromatography, high pressure liquid chromatography, or recrystallization. As can be appreciated by the skilled artisan, further methods of synthesizing the compounds of the formulae herein will be evident to those of ordinary skill in the art.

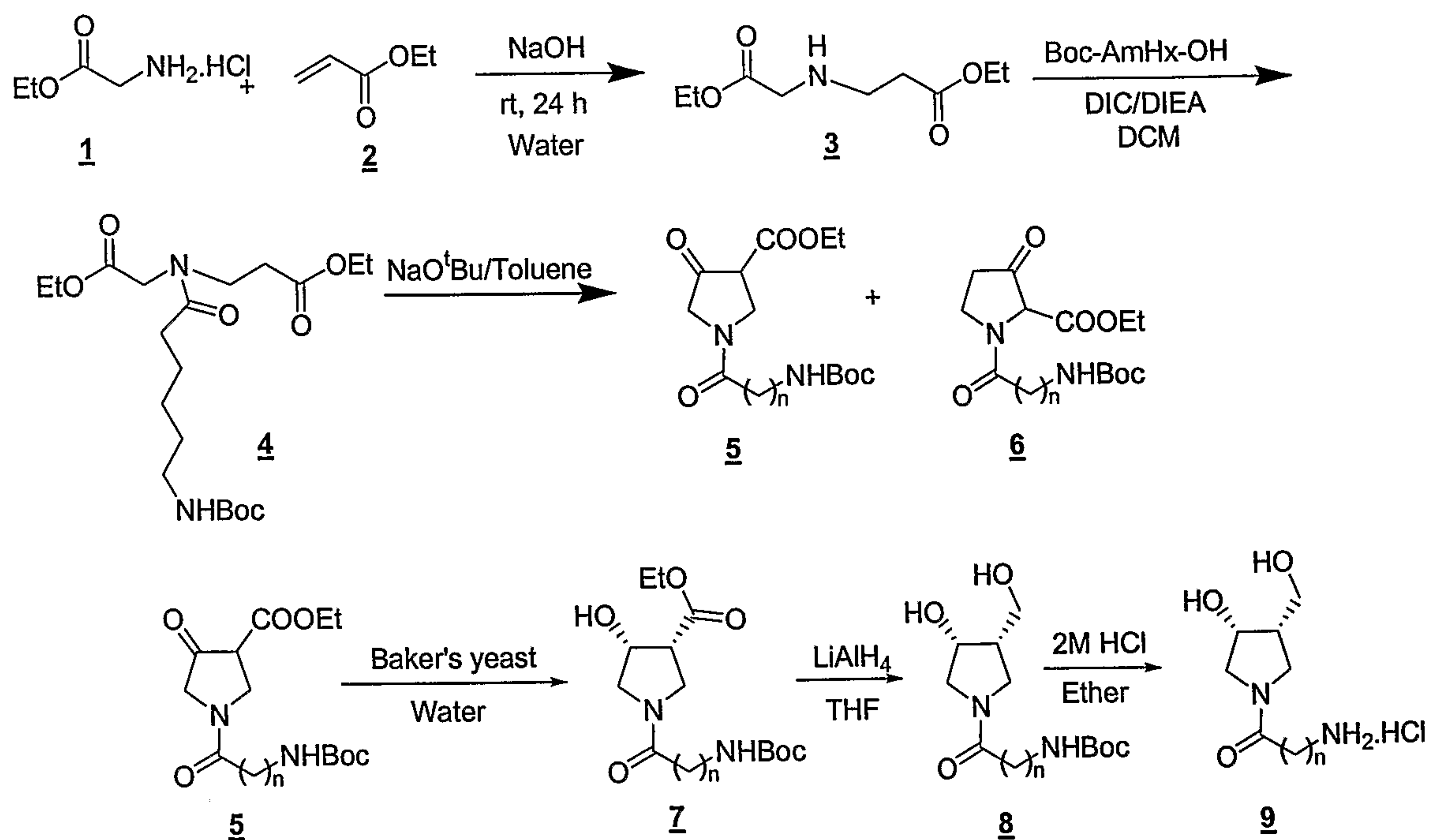
Additionally, the various synthetic steps may be performed in an alternate sequence or order to give the desired compounds. Other synthetic chemistry transformations, protecting groups (*e.g.*, for hydroxyl, amino, etc. present on the bases) and protecting group methodologies (protection and deprotection) useful in synthesizing the compounds described herein are known in the art and include, for example, those such as described in R. Larock, *Comprehensive Organic Transformations*, VCH Publishers (1989); T.W. Greene and P.G.M. Wuts, *Protective Groups in*

Organic Synthesis, 2d. Ed., John Wiley and Sons (1991); L. Fieser and M. Fieser, *Fieser and Fieser's Reagents for Organic Synthesis*, John Wiley and Sons (1994); and L. Paquette, ed., *Encyclopedia of Reagents for Organic Synthesis*, John Wiley and Sons (1995), and subsequent editions thereof.

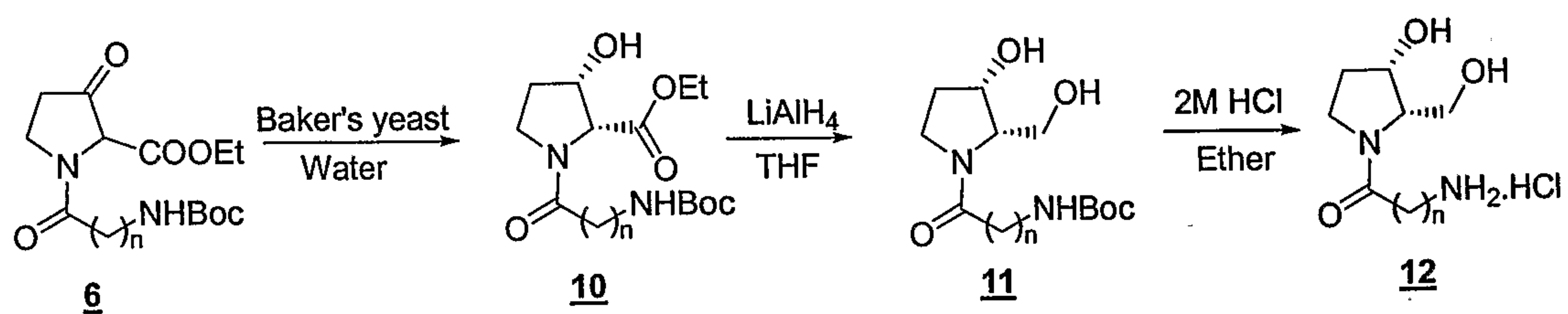
5 The protected monomer compounds of this invention may contain one or more asymmetric centers and thus occur as racemates and racemic mixtures, single enantiomers, individual diastereomers and diastereomeric mixtures. All such isomeric forms of these compounds are expressly included in the present invention. The compounds described herein can also contain linkages (*e.g.*, carbon-carbon bonds, carbon-nitrogen bonds, *e.g.*, amides) or
10 substituents that can restrict bond rotation, *e.g.* restriction resulting from the presence of a ring or double bond. Accordingly, all *cis/trans*, *E/Z* isomers, and rotational isomers (rotamers) are expressly included herein. The compounds of this invention may also be represented in multiple tautomeric forms, in such instances, the invention expressly includes all tautomeric forms of the compounds described herein (*e.g.*, alkylation of a ring system may result in alkylation at
15 multiple sites, the invention expressly includes all such reaction products). All such isomeric forms of such compounds are expressly included in the present invention. All crystal forms of the compounds described herein are expressly included in the present invention.

 Representative ligand-conjugated monomers and typical syntheses for preparing ligand-conjugated monomers and related compounds described herein are provided below. As
20 discussed elsewhere, protecting groups for ligand-conjugated monomer hydroxyl groups, *e.g.*, OFG¹, include but are not limited to the dimethoxytrityl group (DMT). For example, it can be desirable in some embodiments to use silicon-based protecting groups as a protecting group for OFG¹. Silicon-based protecting groups can therefore be used in conjunction with or in place of the DMT group as necessary or desired. Thus, the ligand-conjugated monomers and syntheses
25 delineated below, which feature the DMT protecting group as a protecting group for OFG¹, is not to be construed as limiting in any way to the invention.

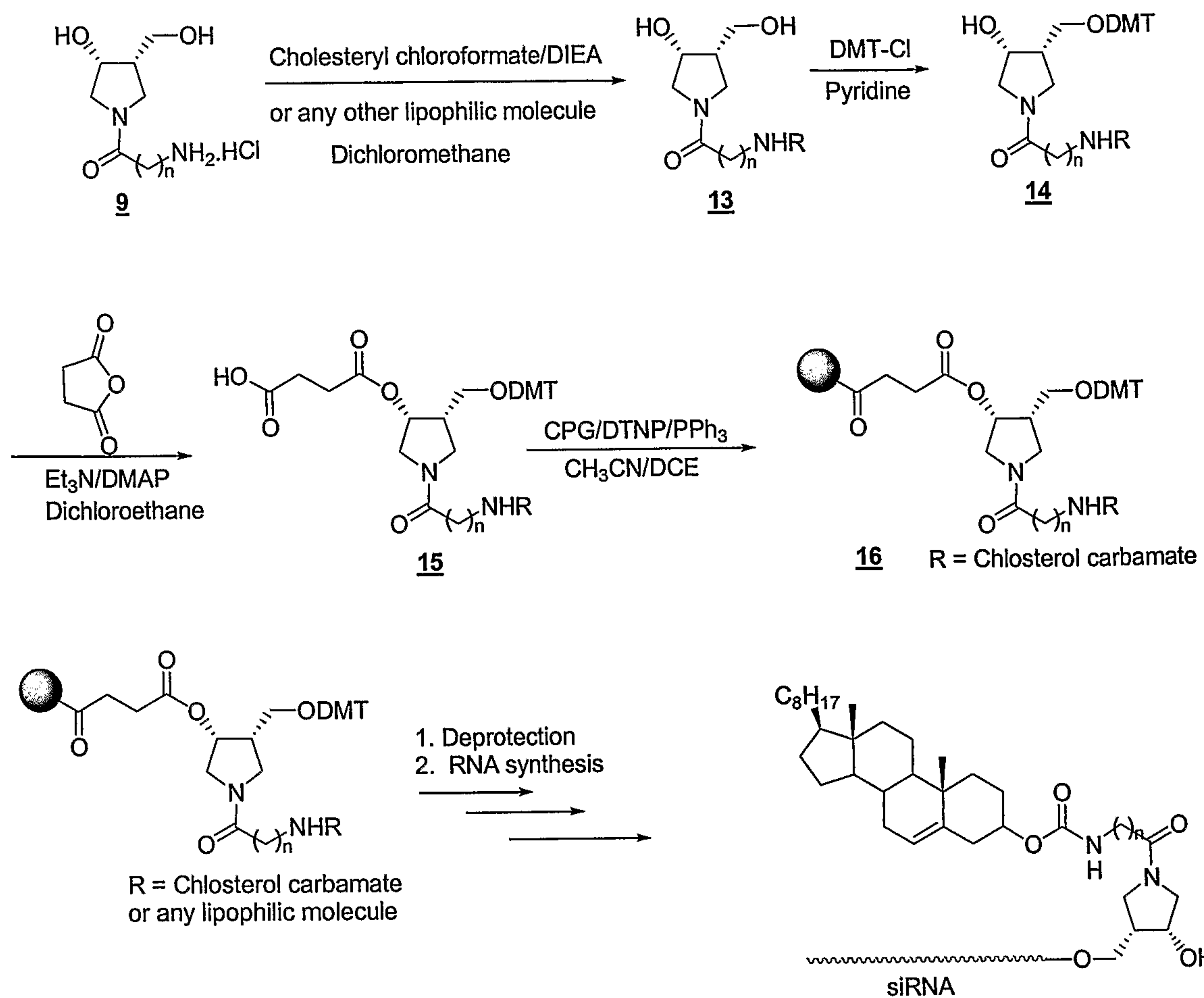
Synthesis of pyrroline carrier

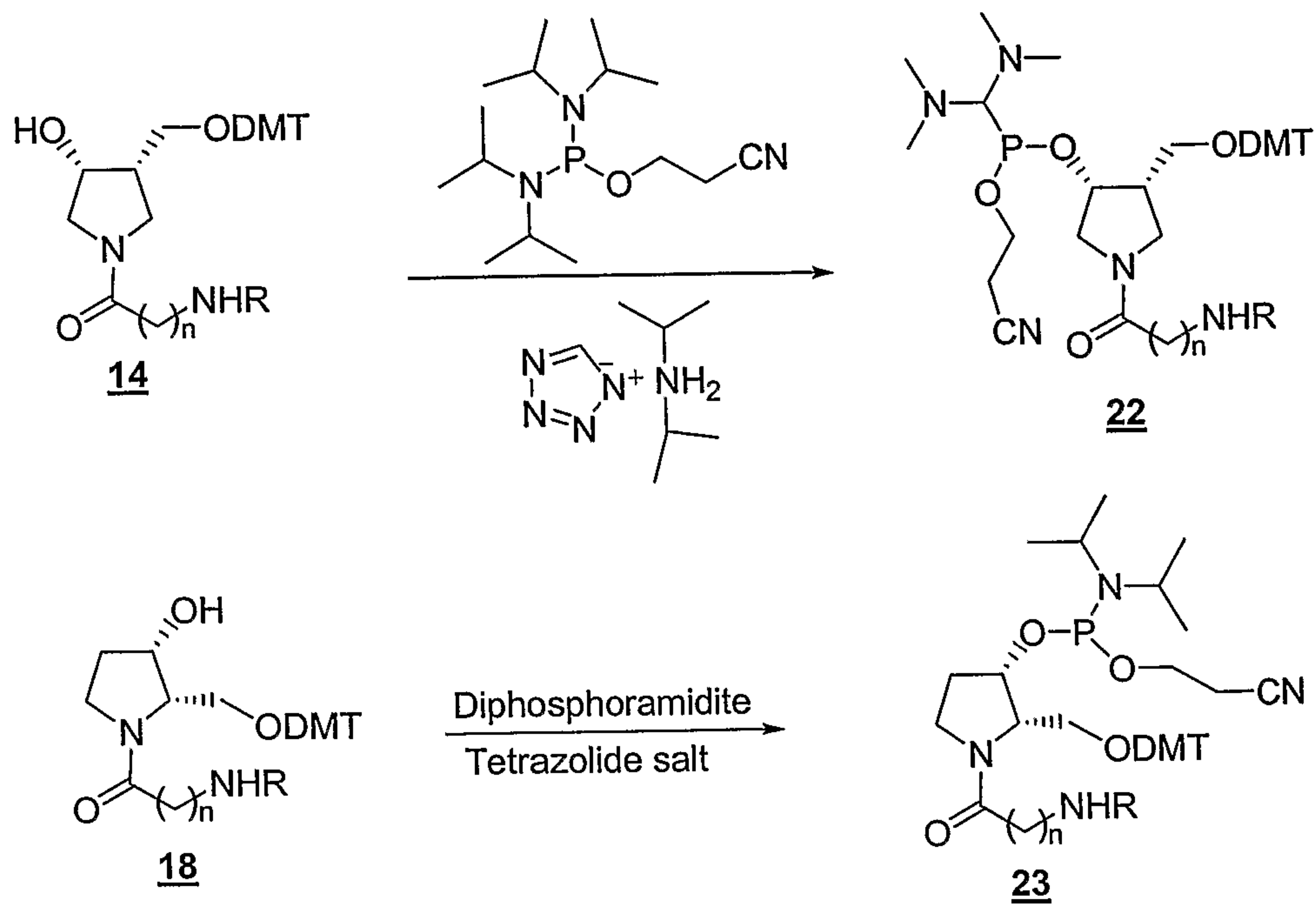
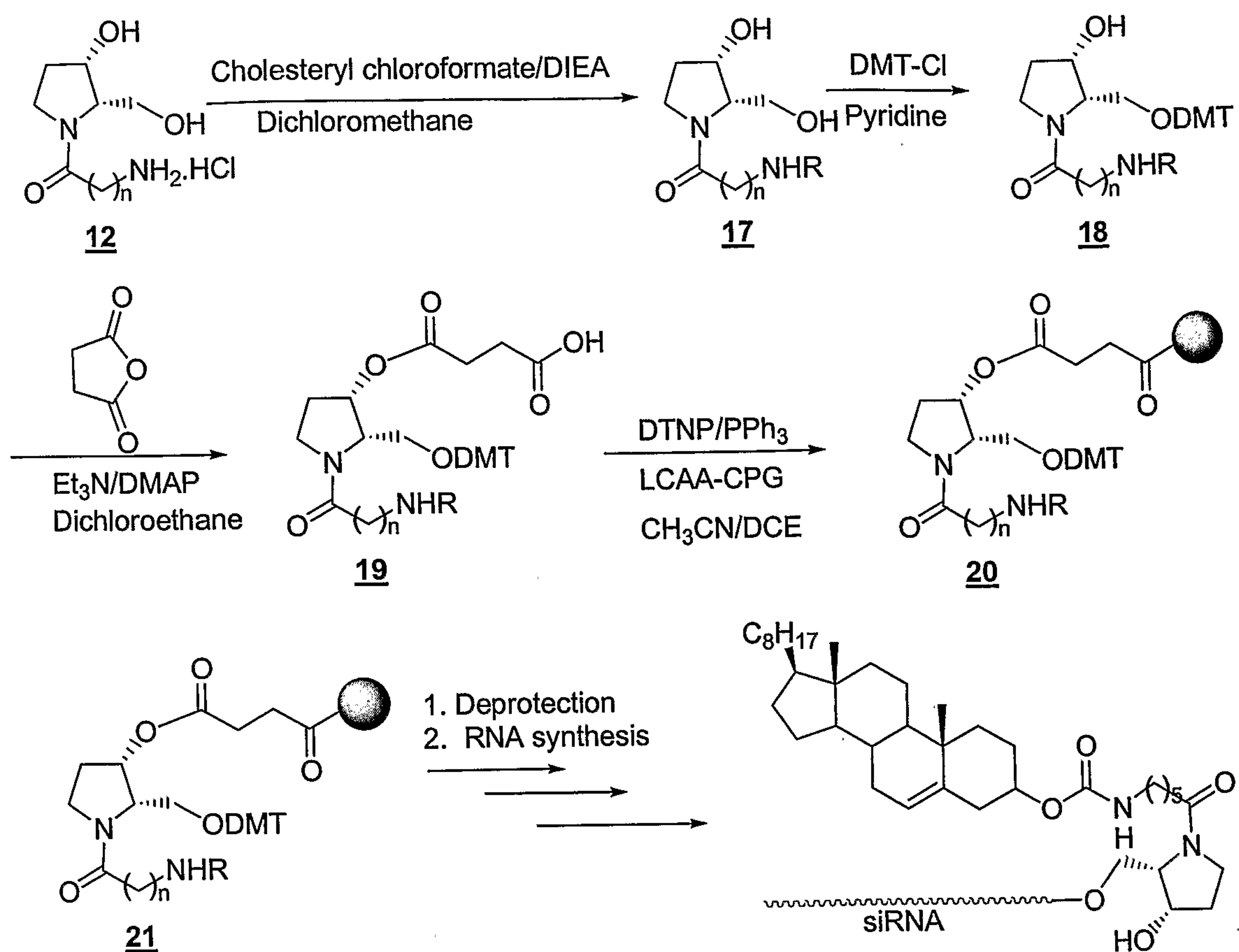


Scheme 1. Synthesis of cis-(-(3S,4R)-pyrrolidine diol

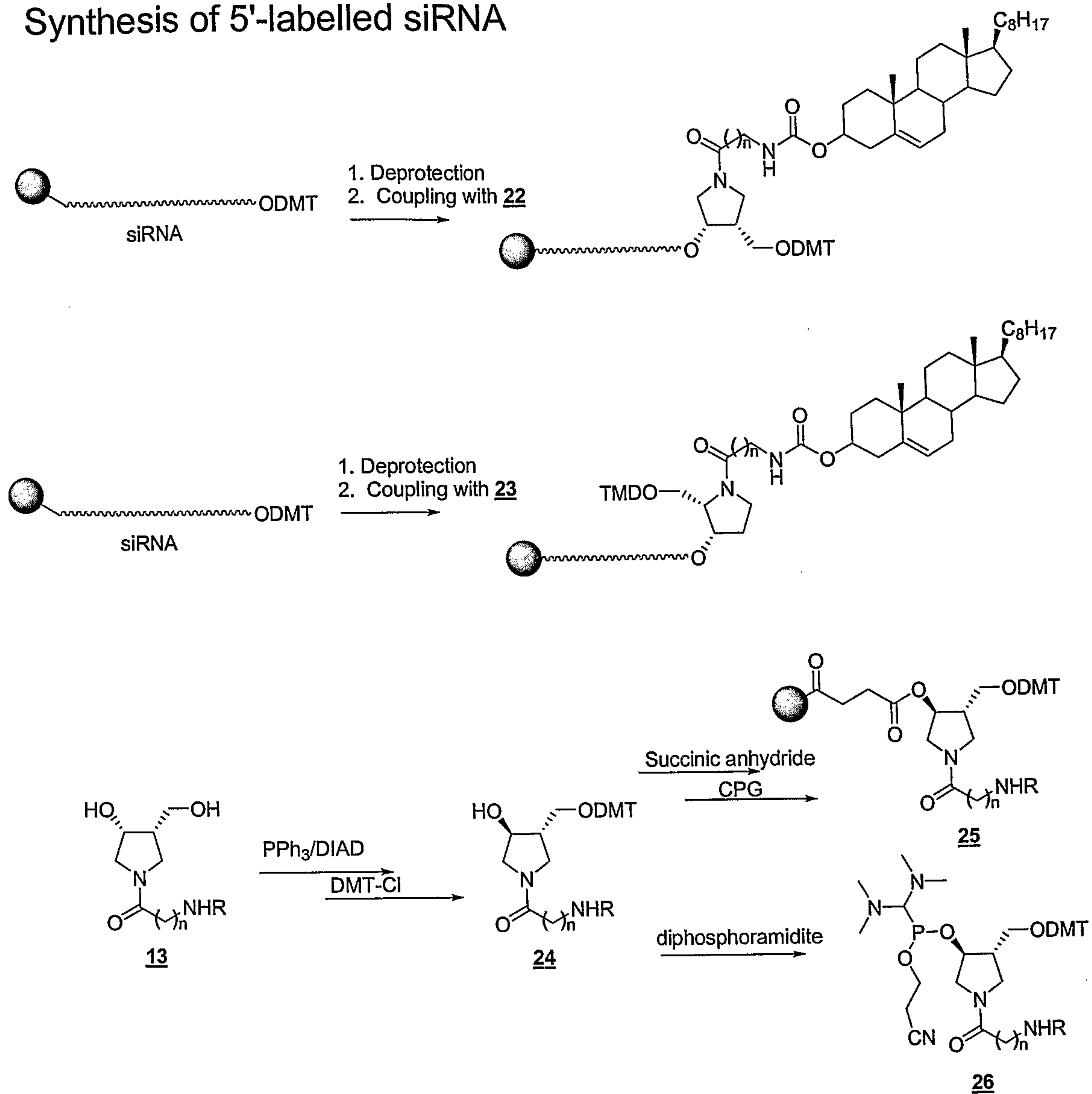


Scheme 2. Synthesis of cis-(-(2R, 3S)-pyrrolidine diol

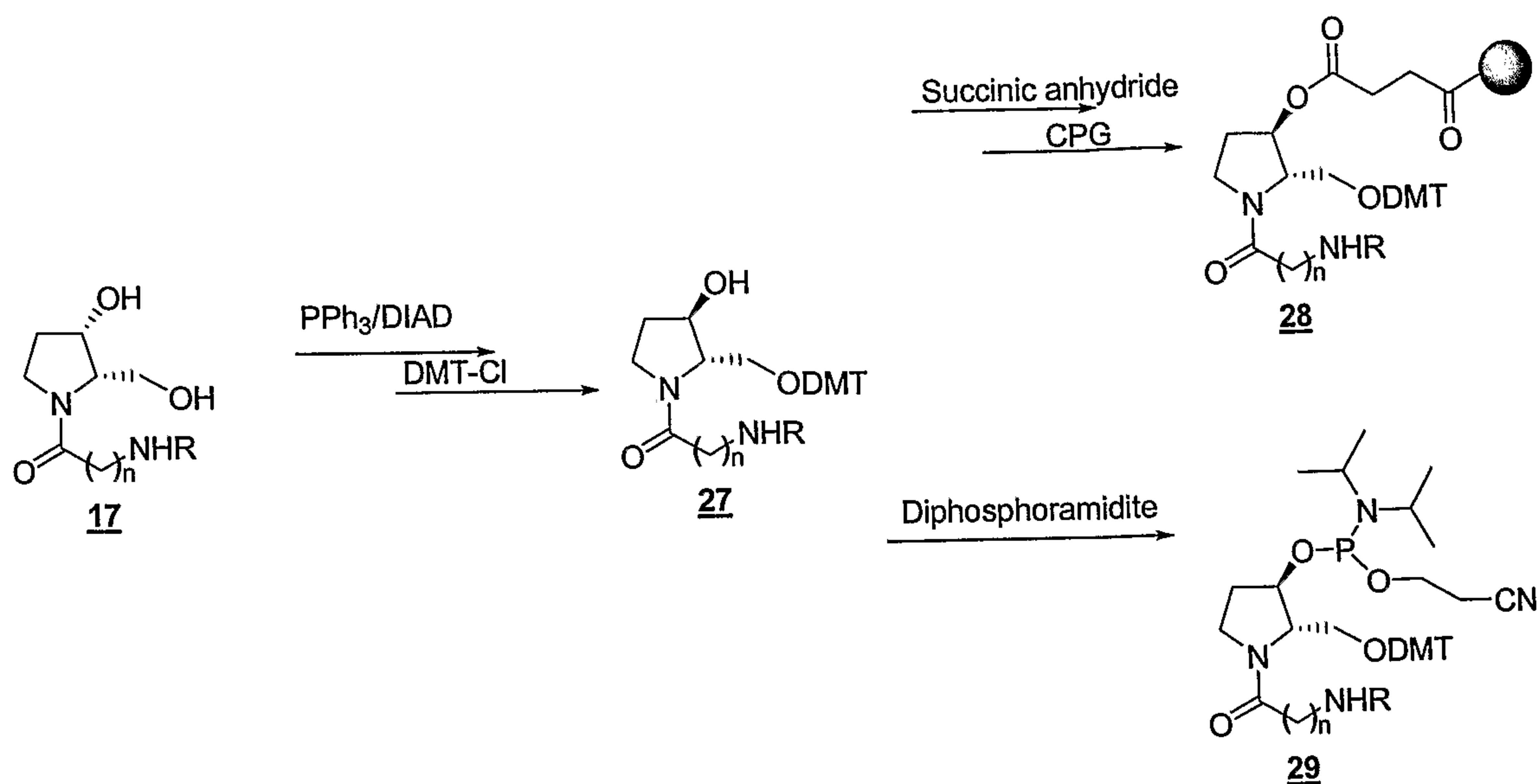




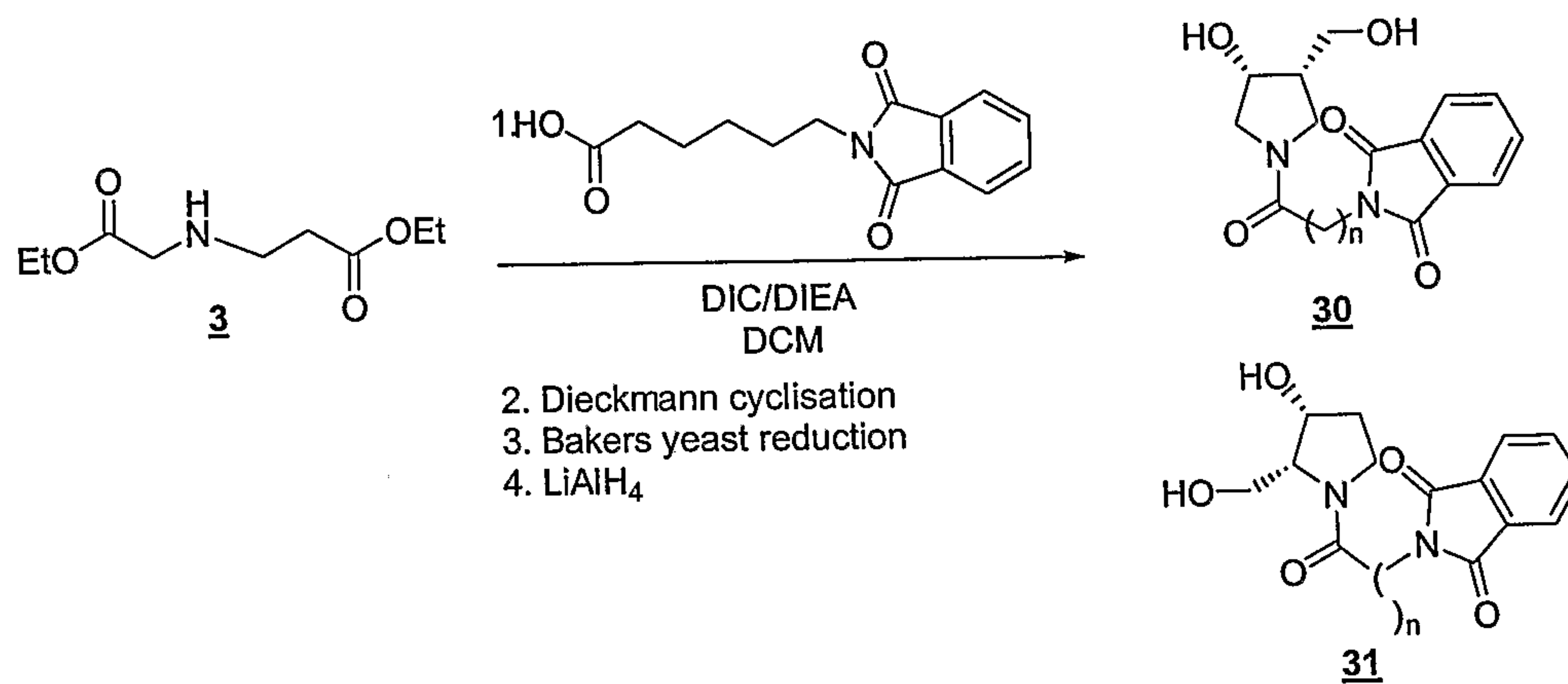
Synthesis of 5'-labelled siRNA



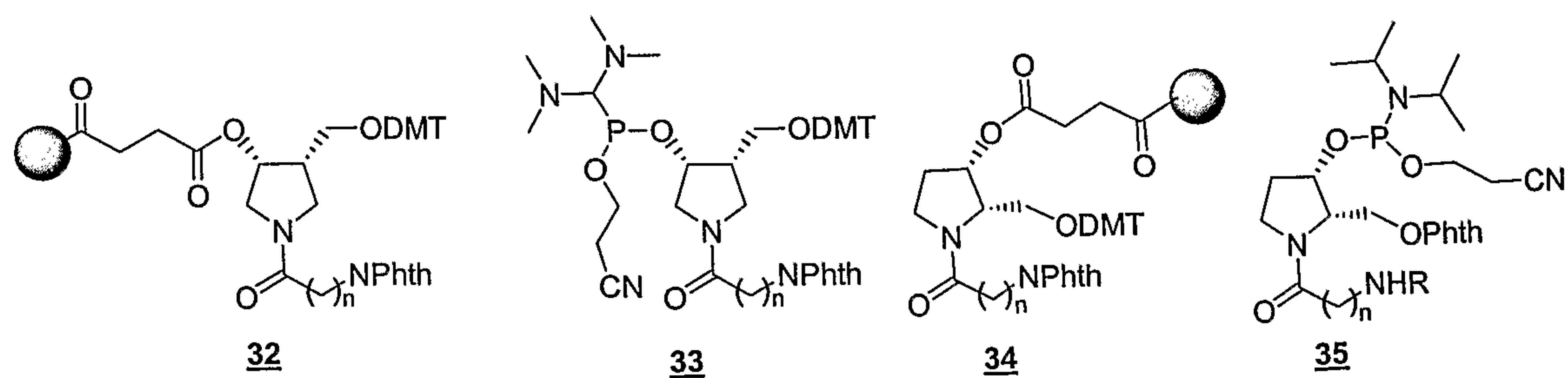
25 & **26** can be used for 3', 5'- conjugation respectively.

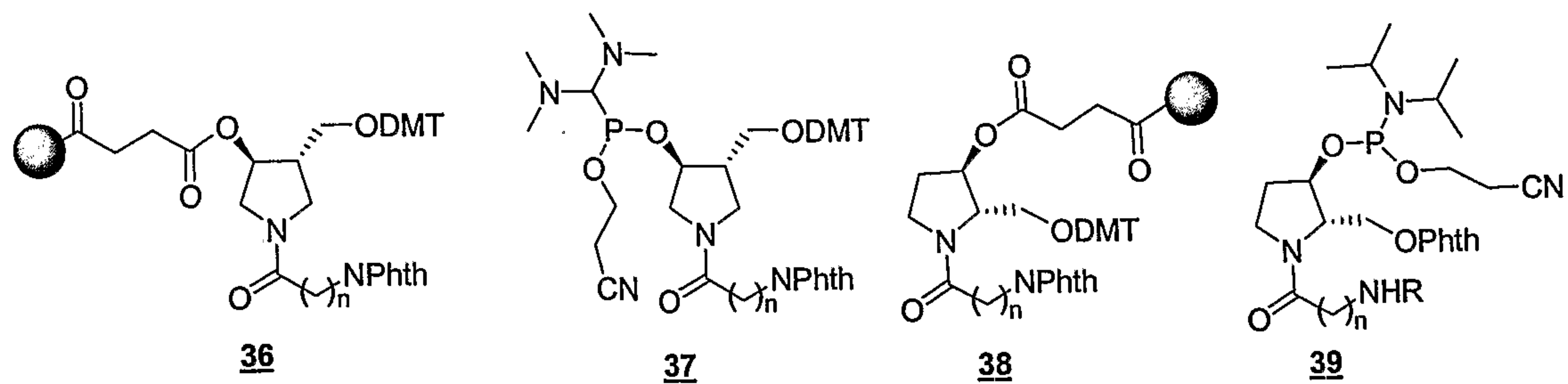


Synthesis of phthalimido derivative

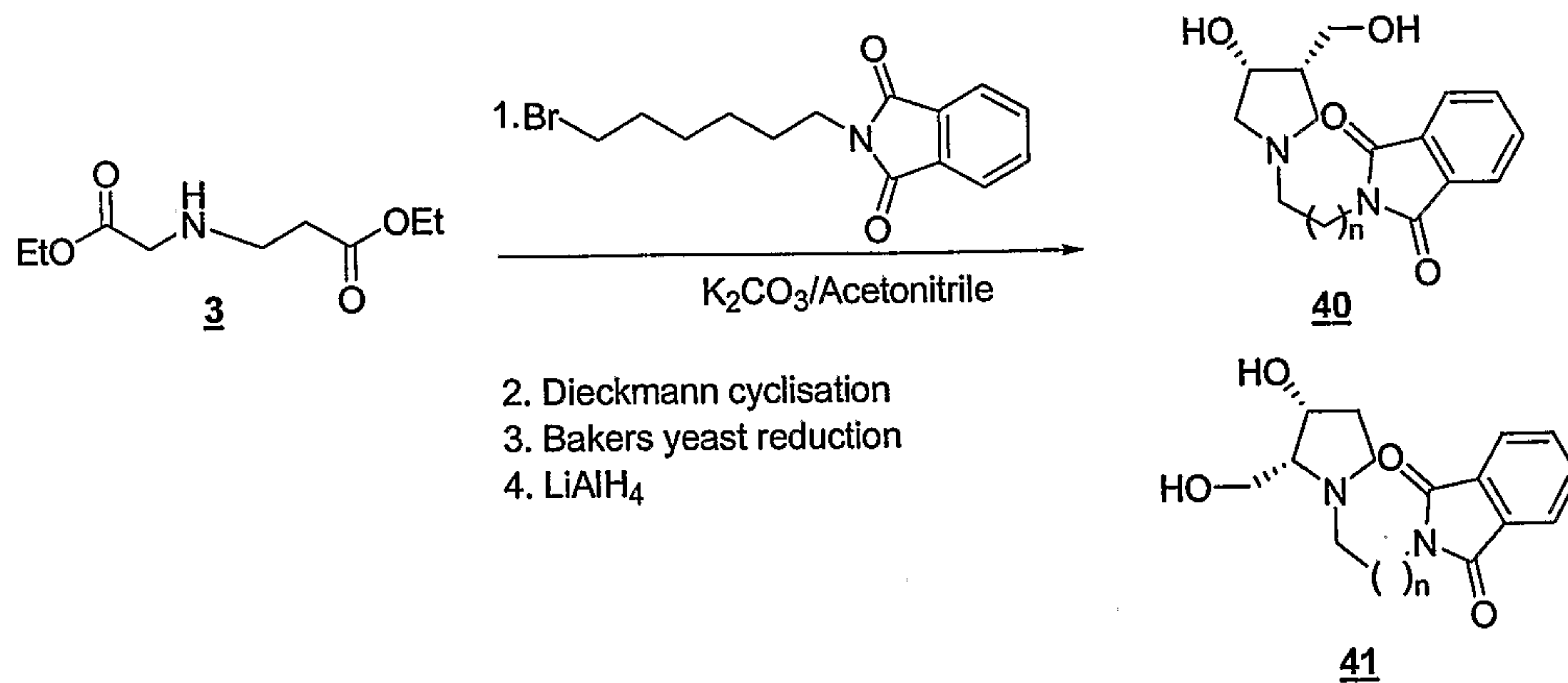


30 and **31** can be converted to similar derivatives as shown in schemes 2-4 for 3' and 5' conjugation of siRNA

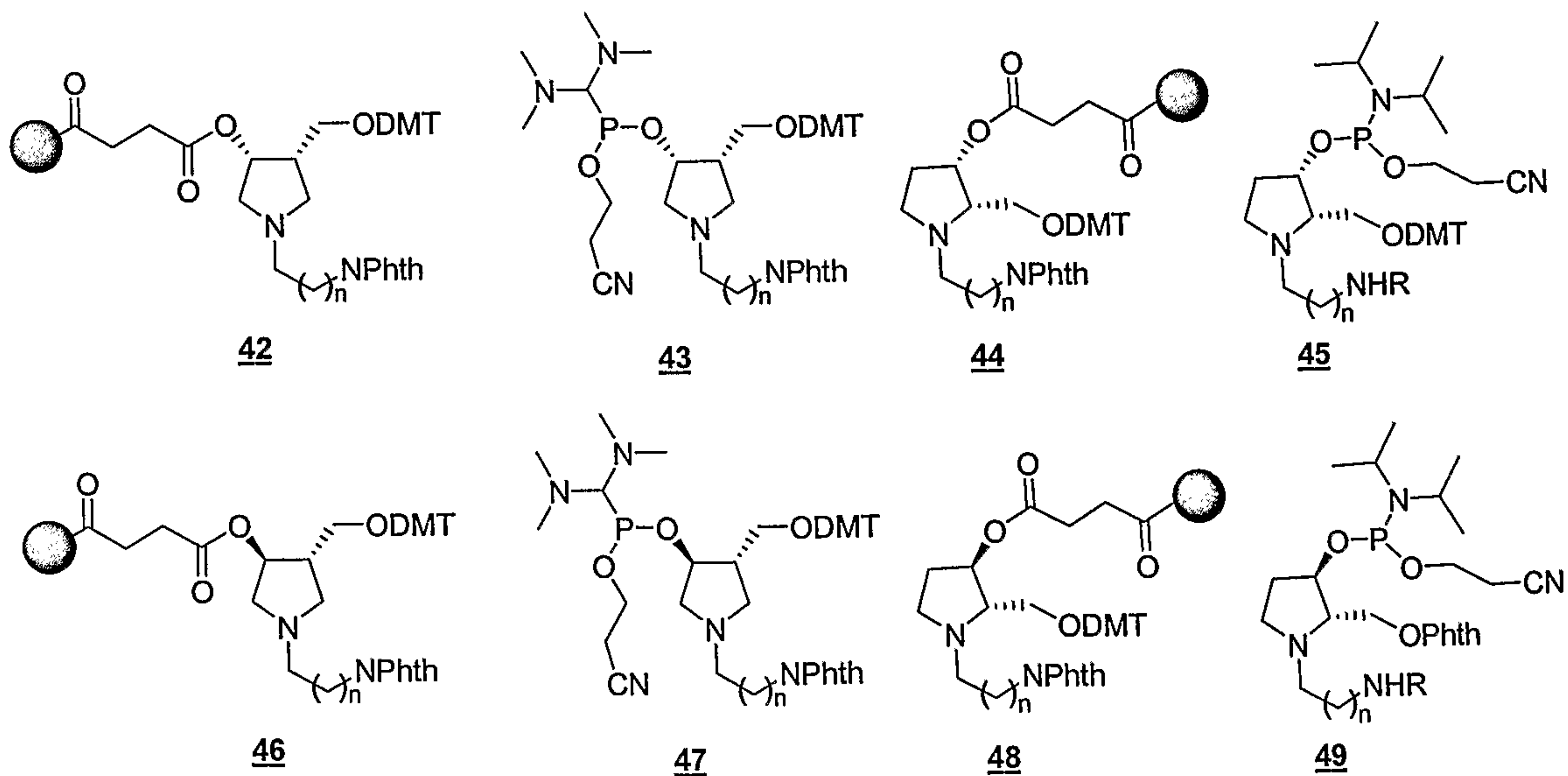




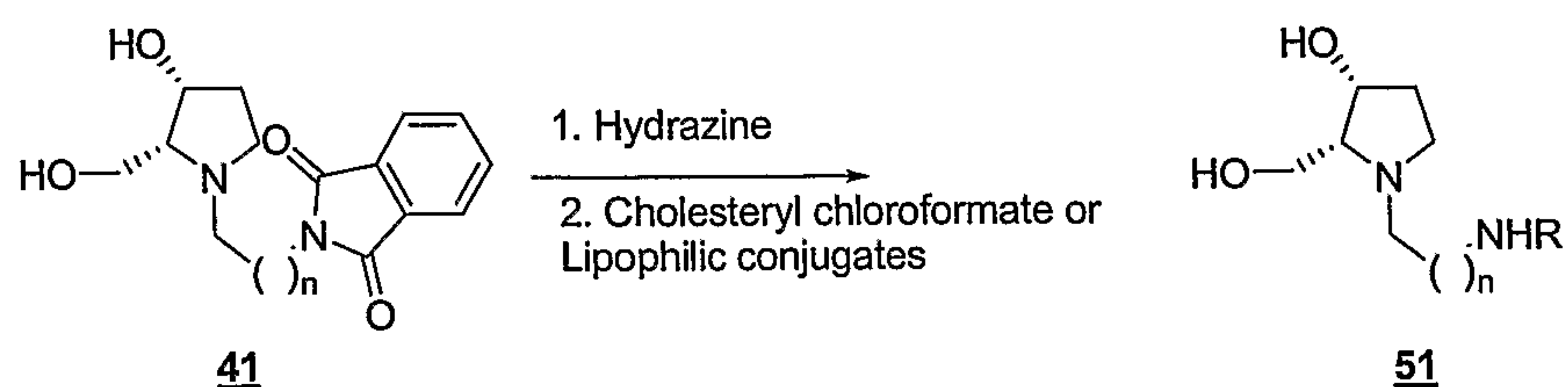
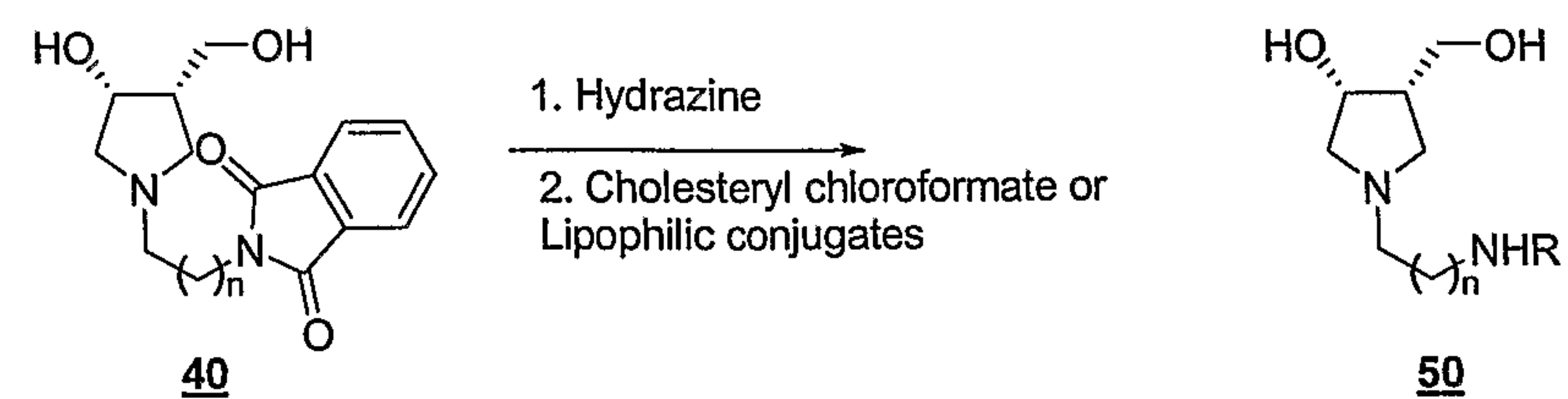
Synthesis of thalimido derivative



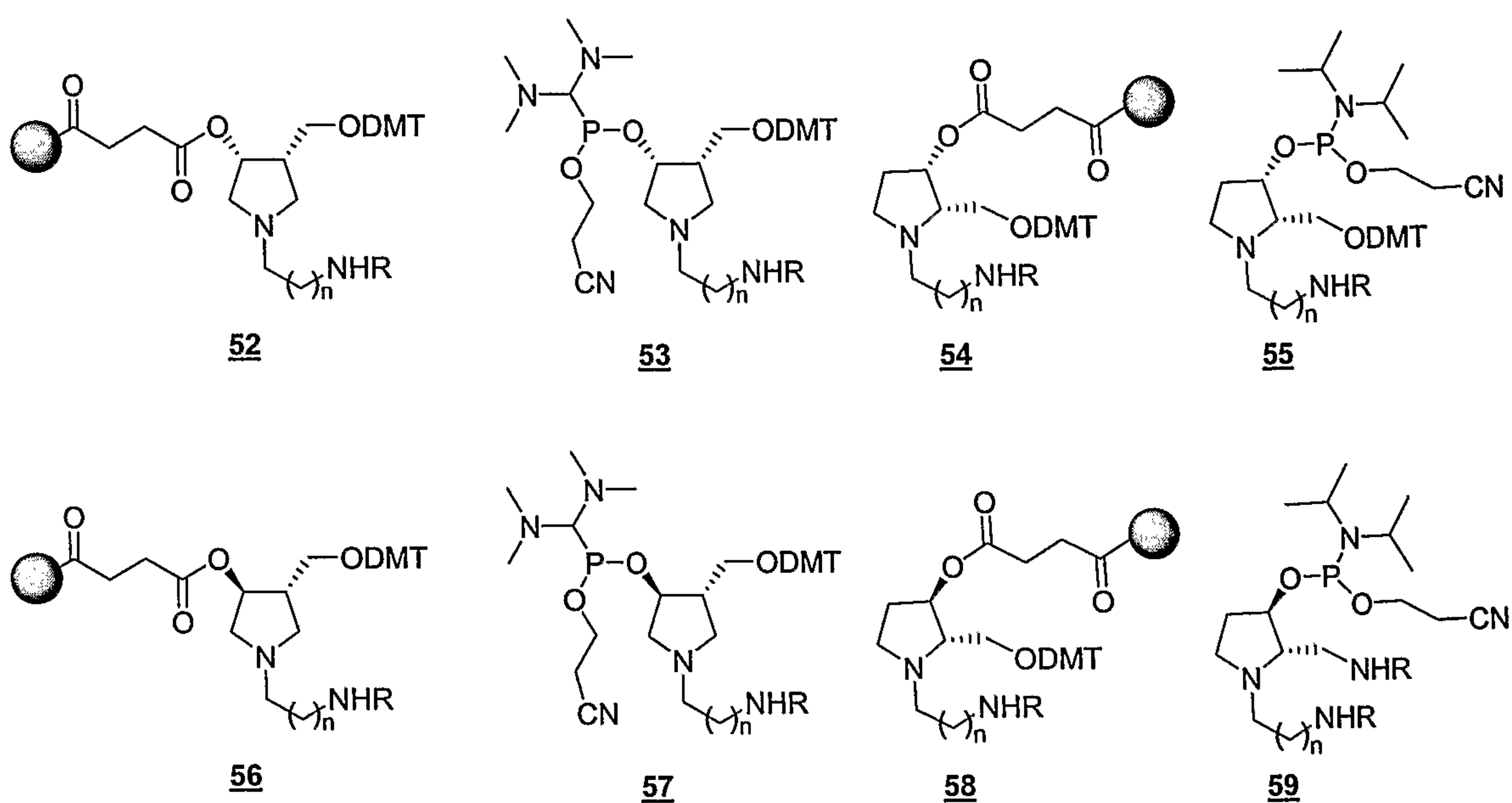
40 and **41** can be converted to similar derivatives as shown in schemes 2-4 for 3' and 5' cpnjugation of siRNA



Synthesis of N-alkyl pyrroline derivatives

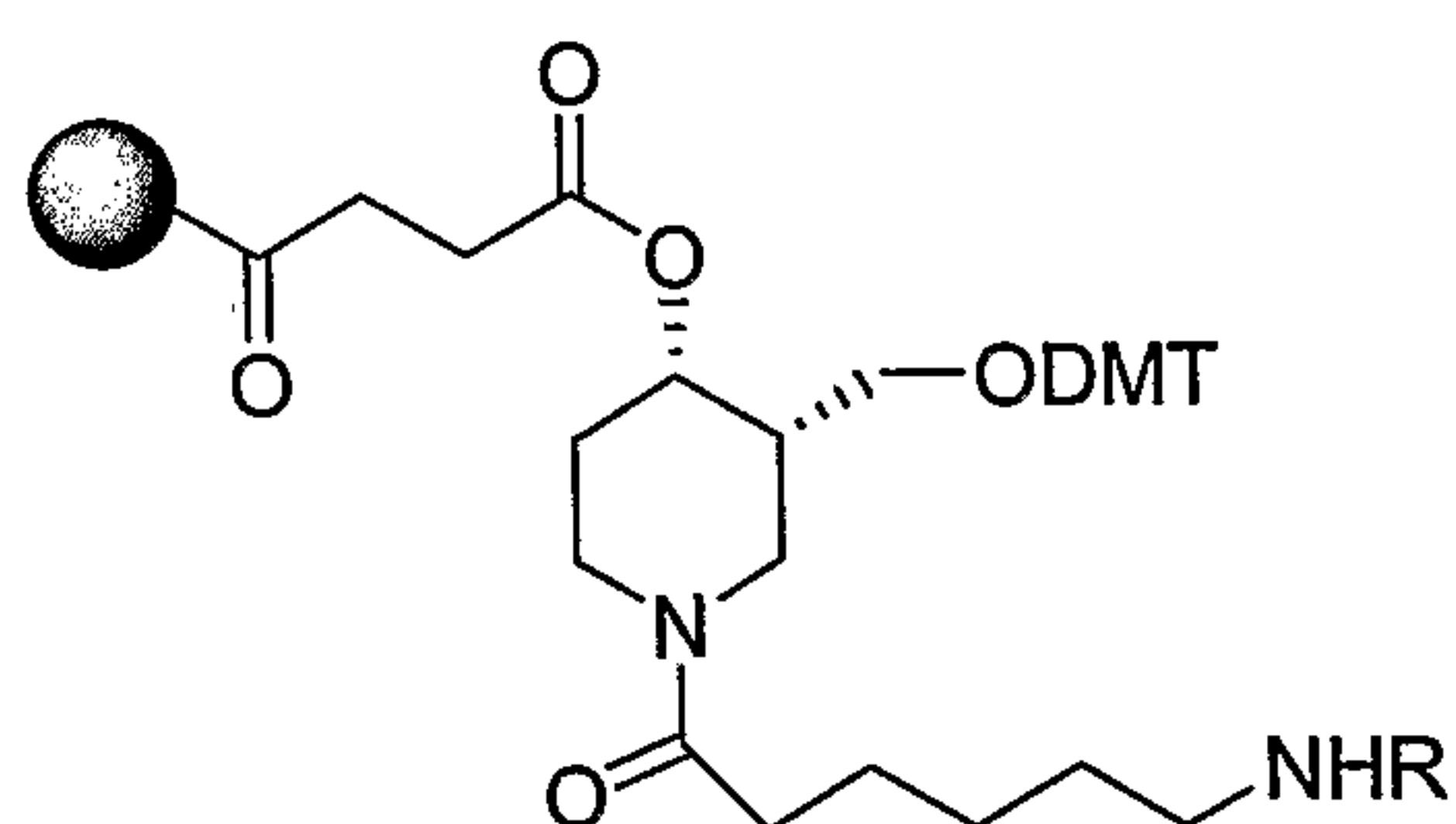


Intermediates **50** and **51** can be converted to analogs which could be conjugated with siRNA using similar reactions

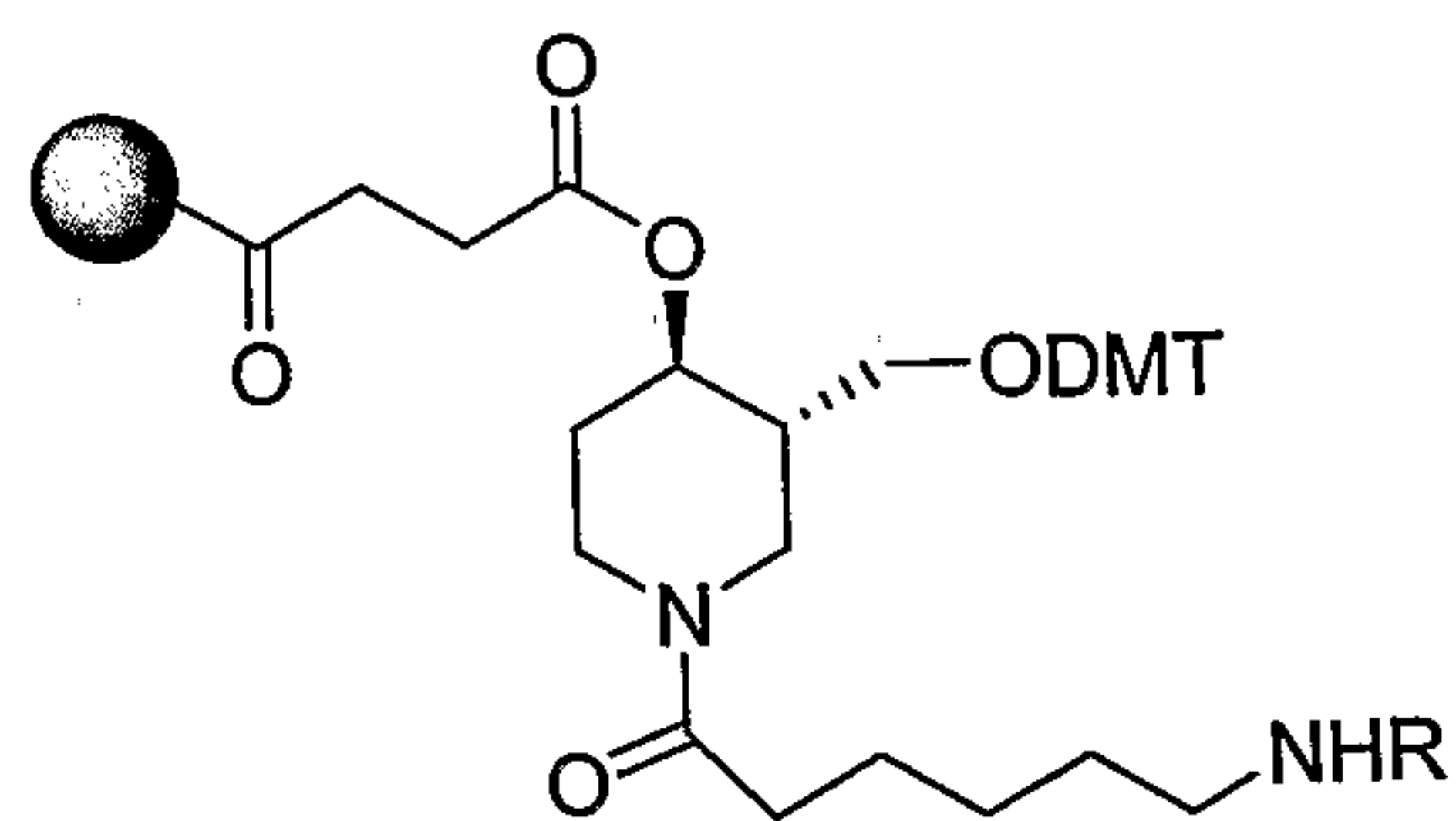


Piperidine series ligands:

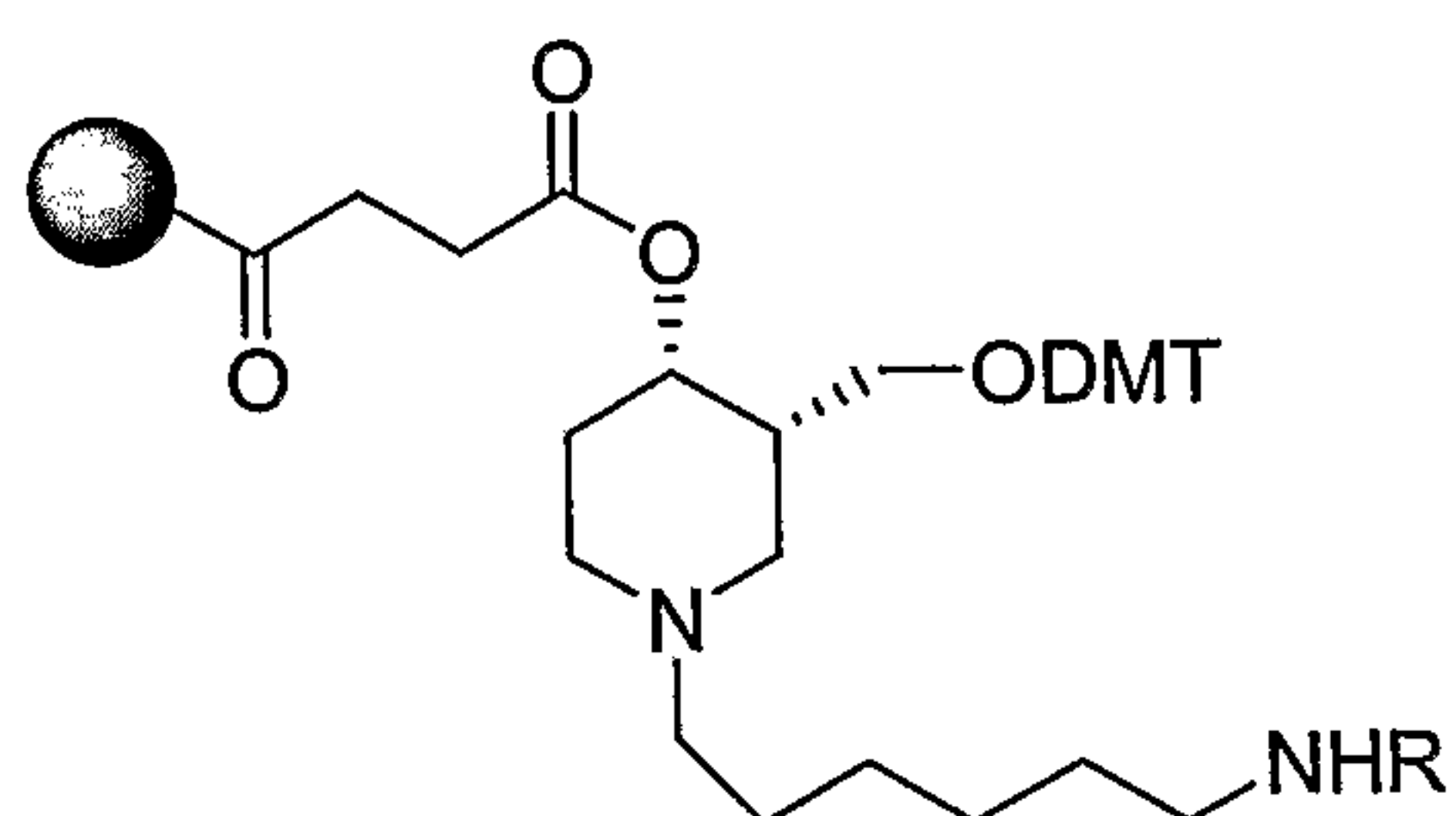
Similar to pyrroline series piperidine series can be synthesised



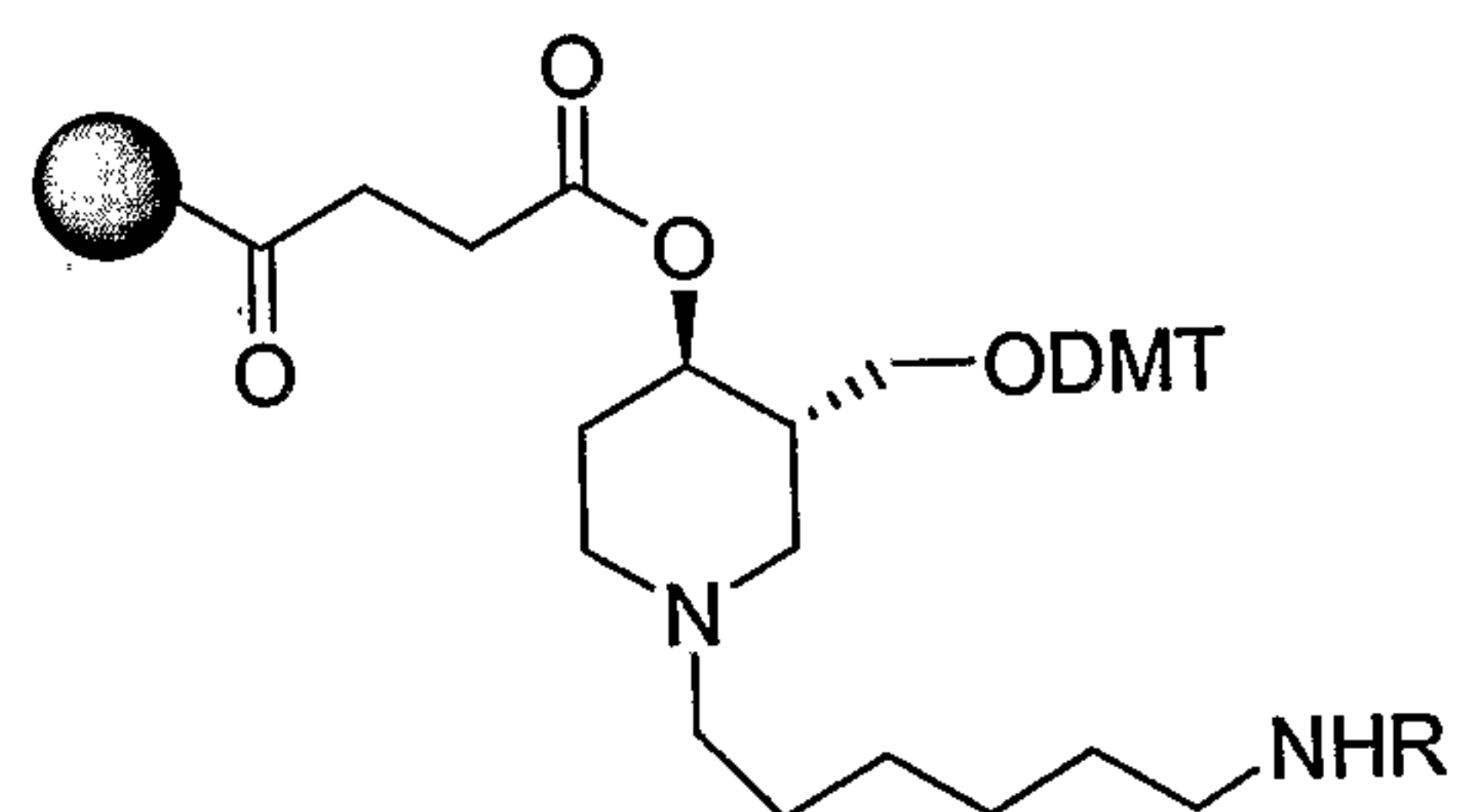
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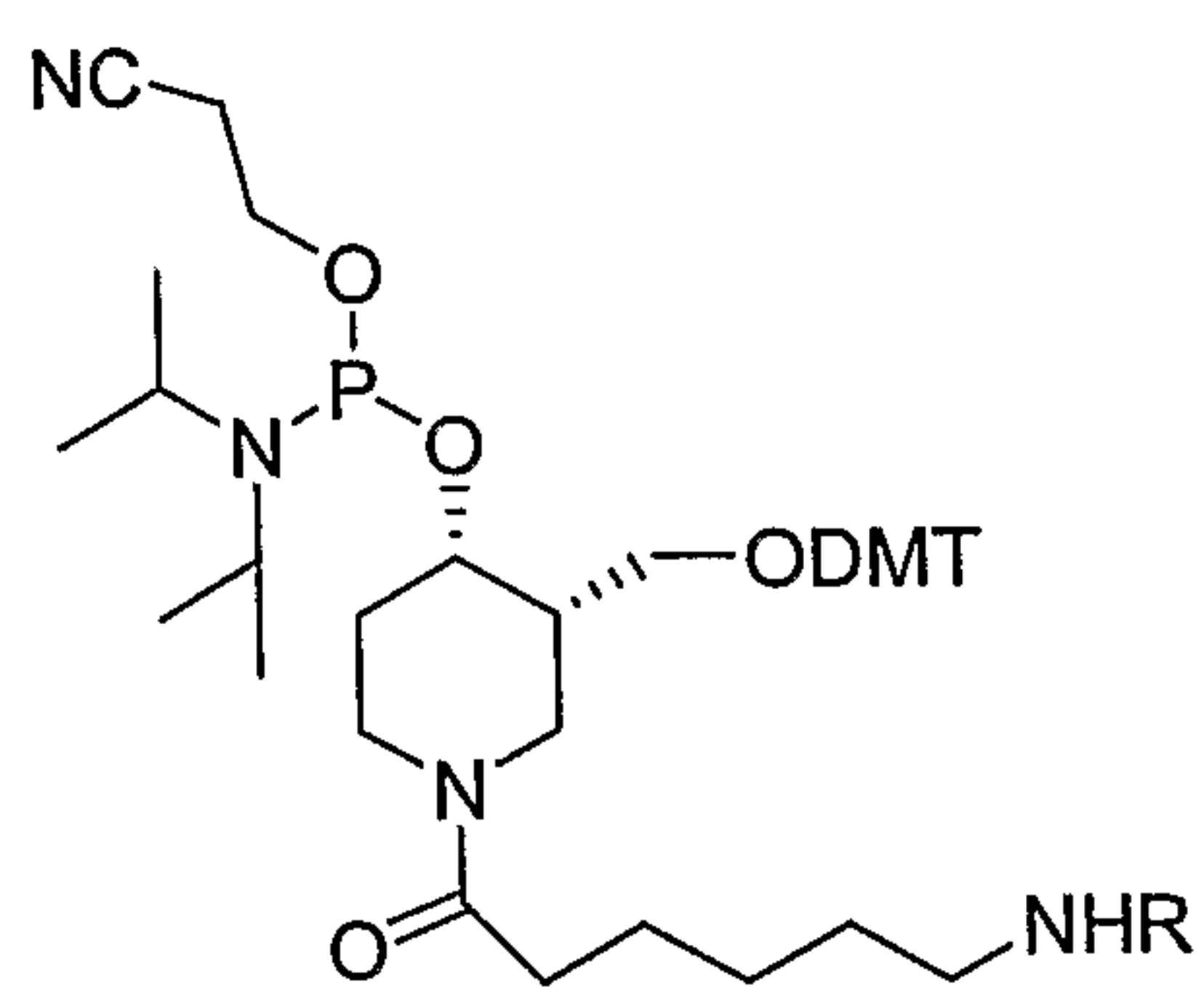
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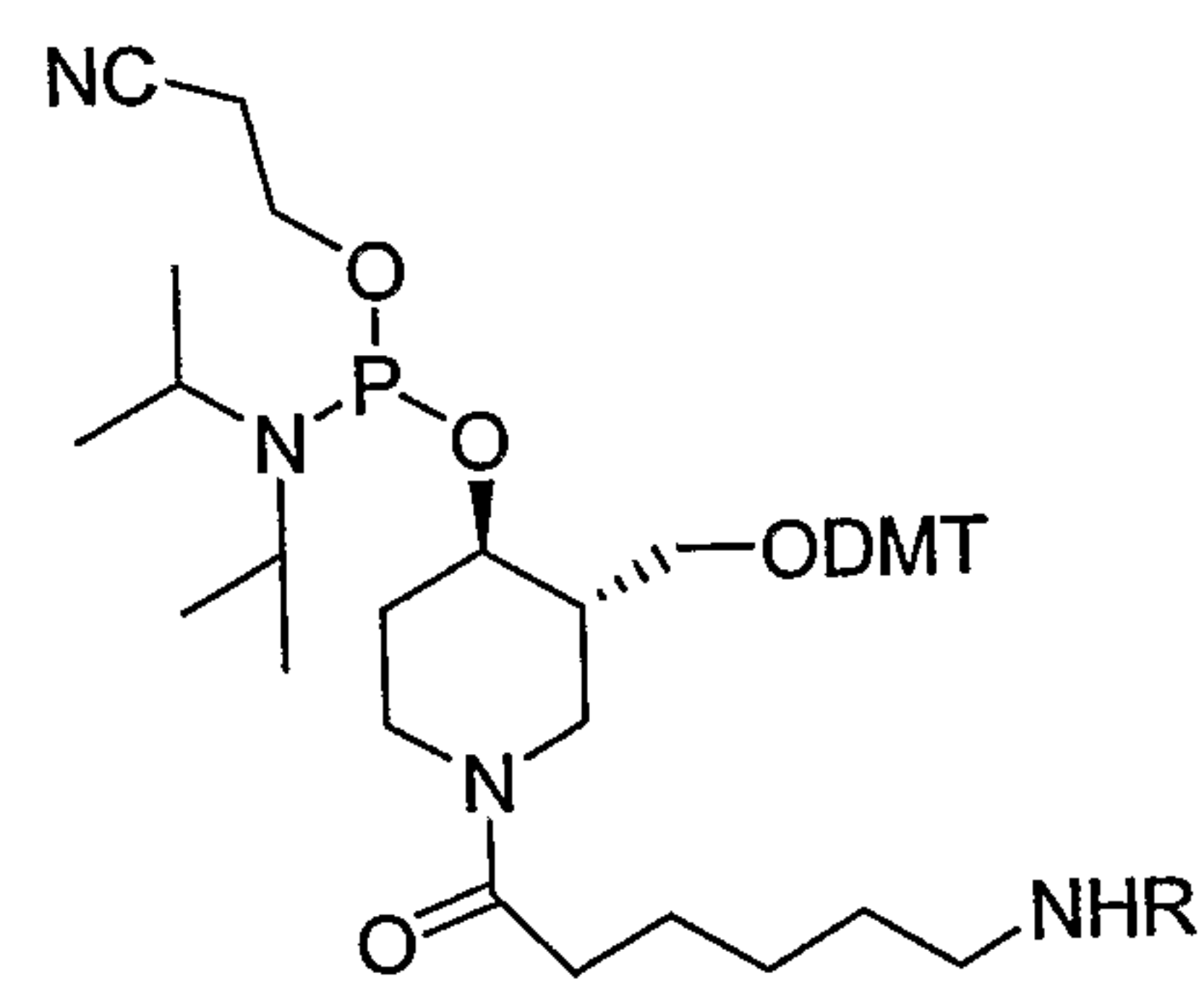
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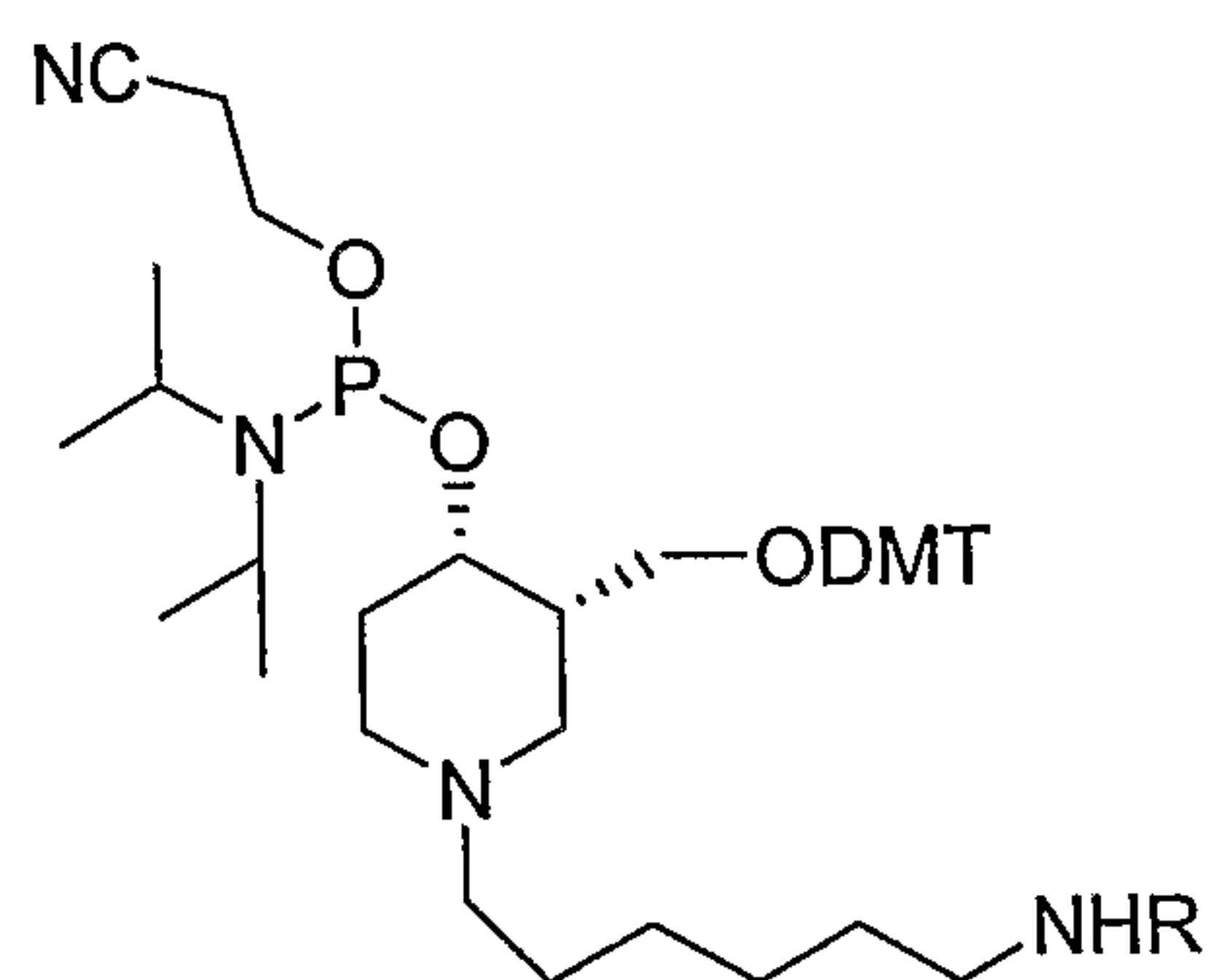
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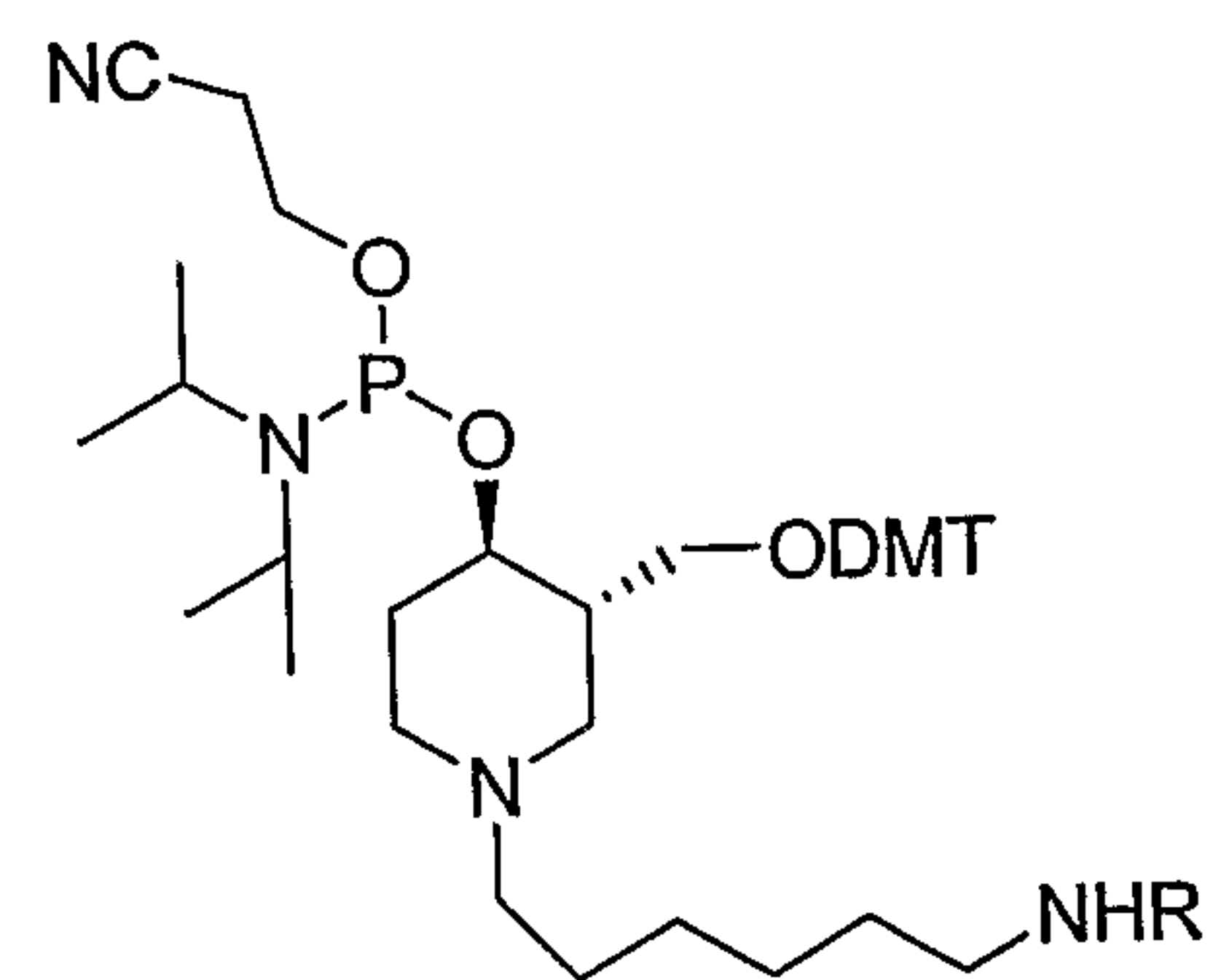
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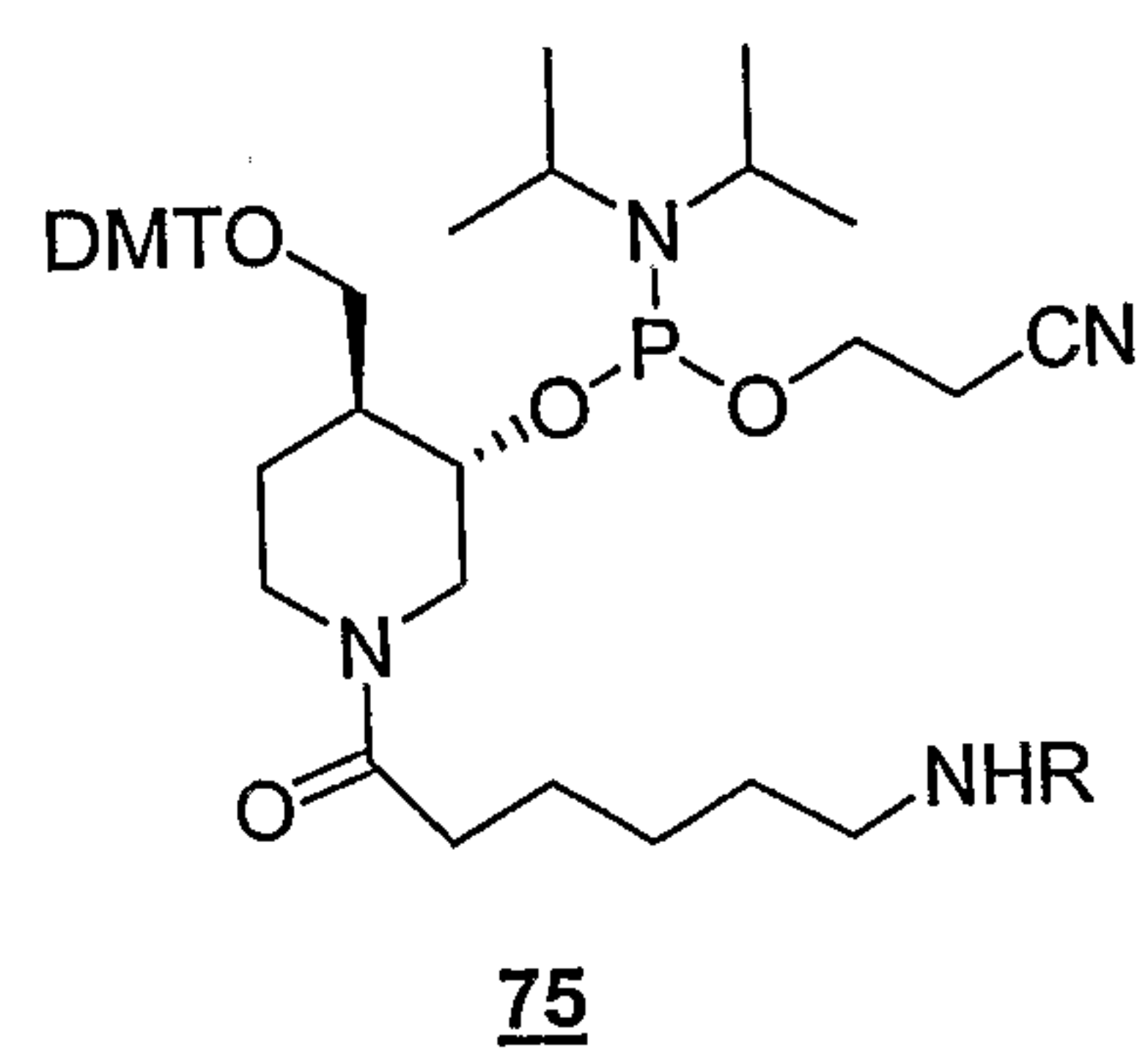
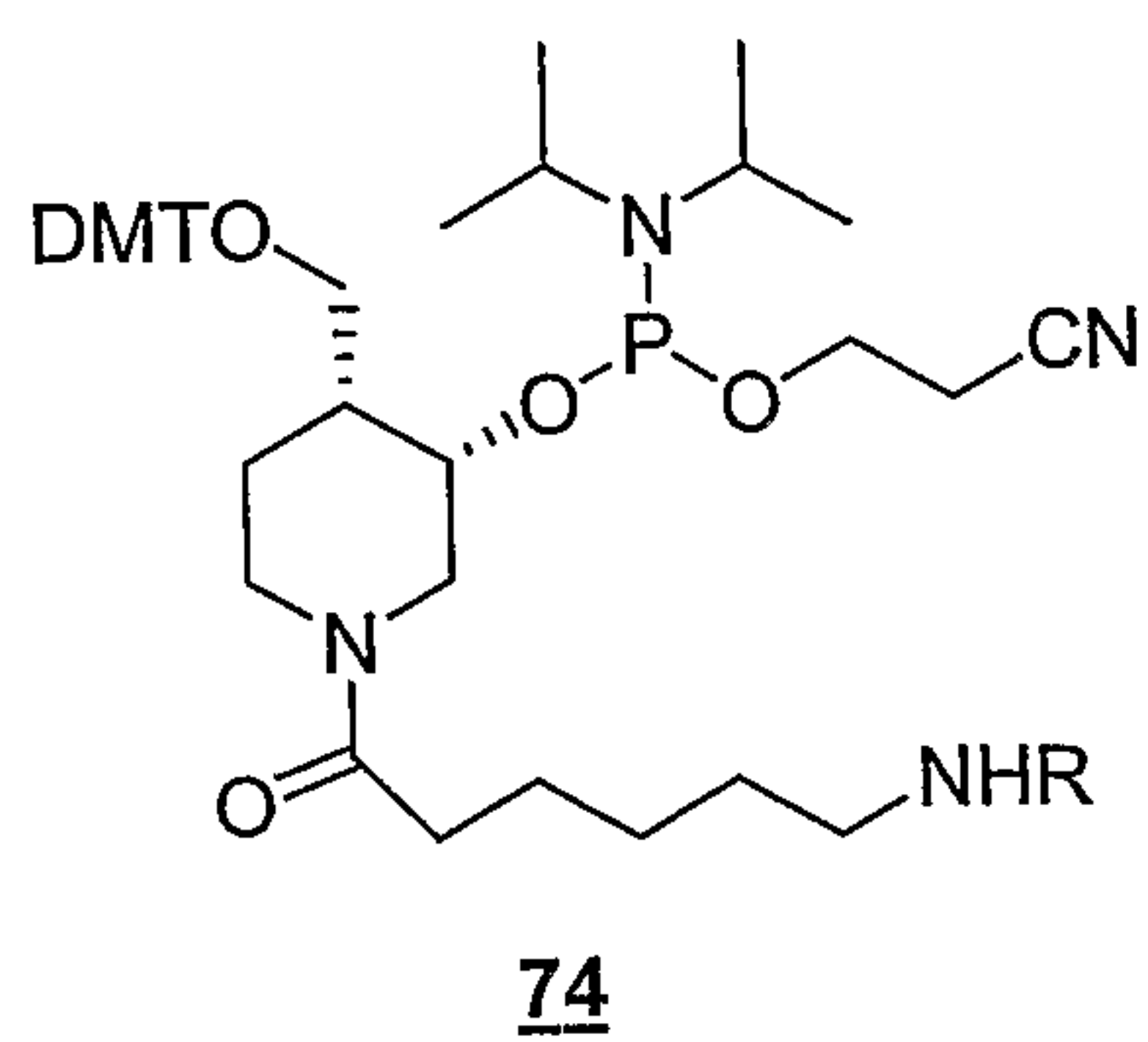
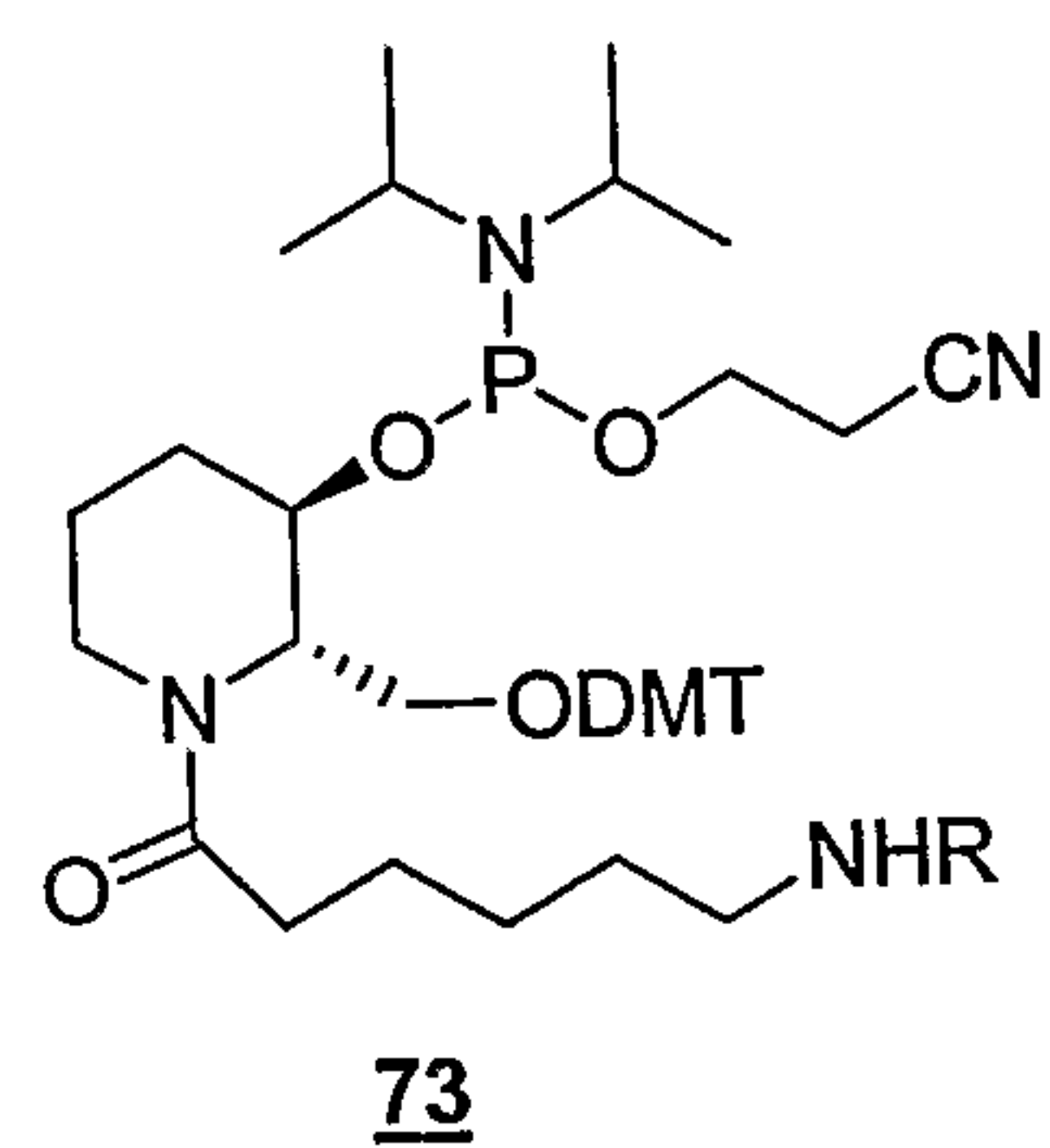
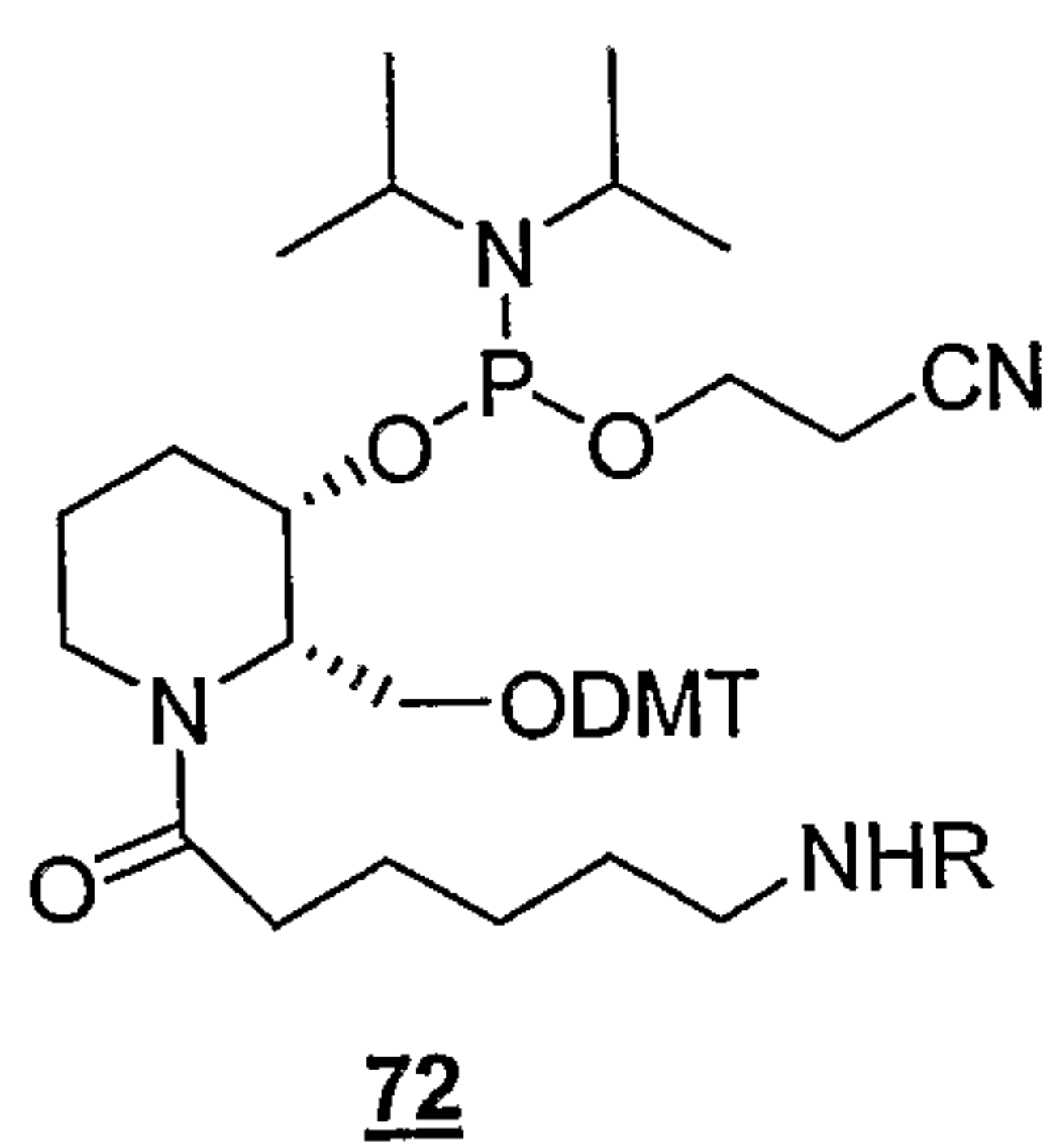
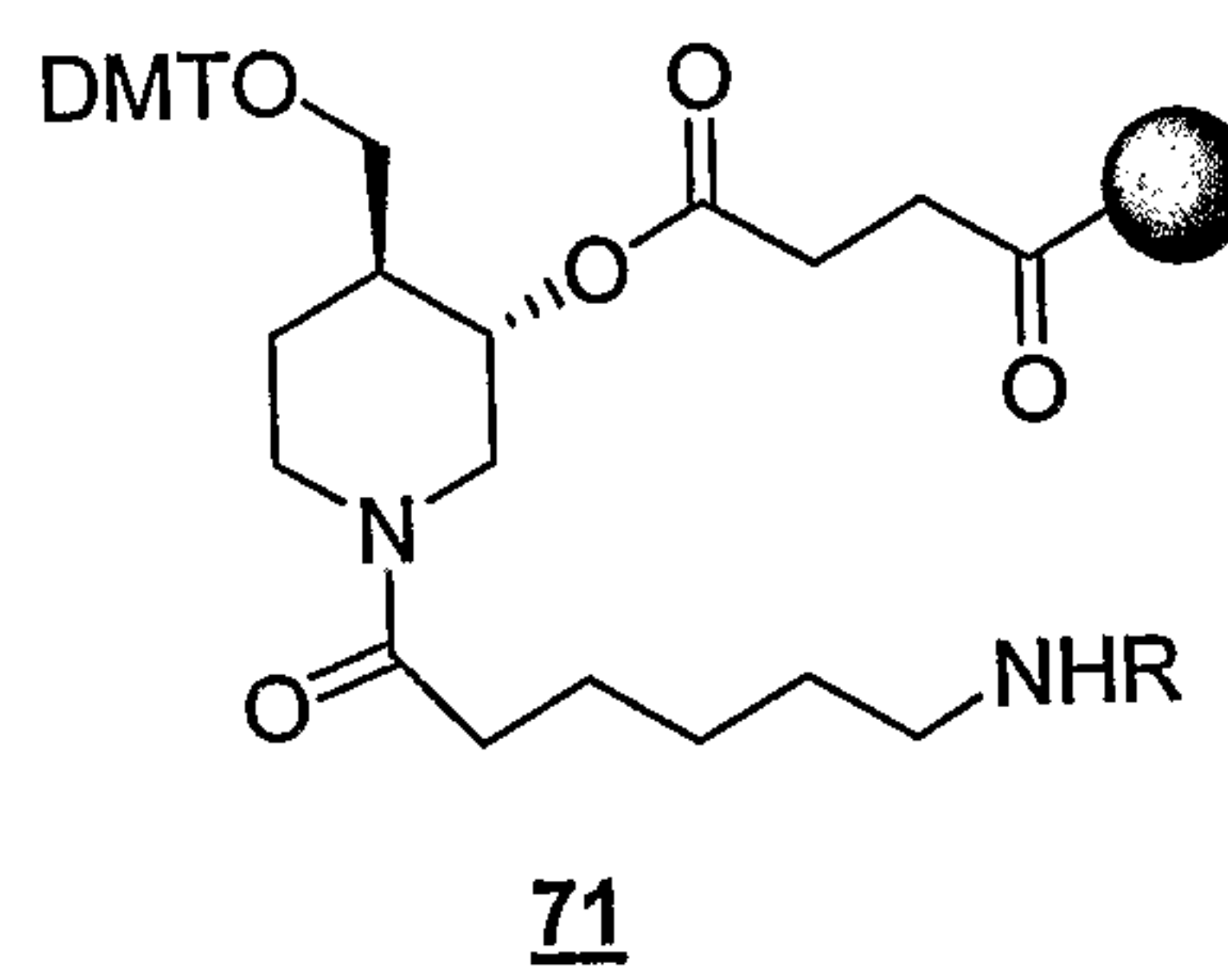
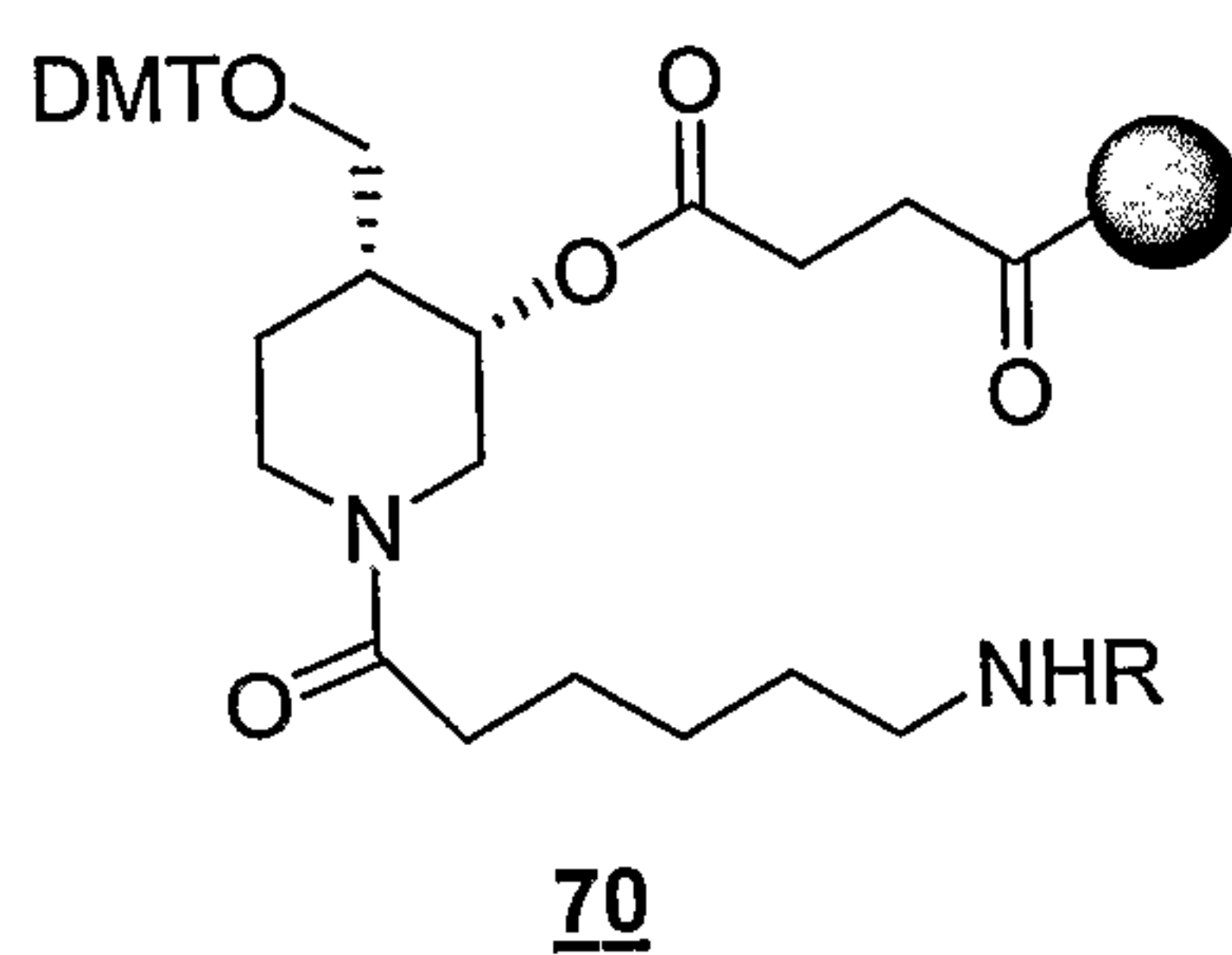
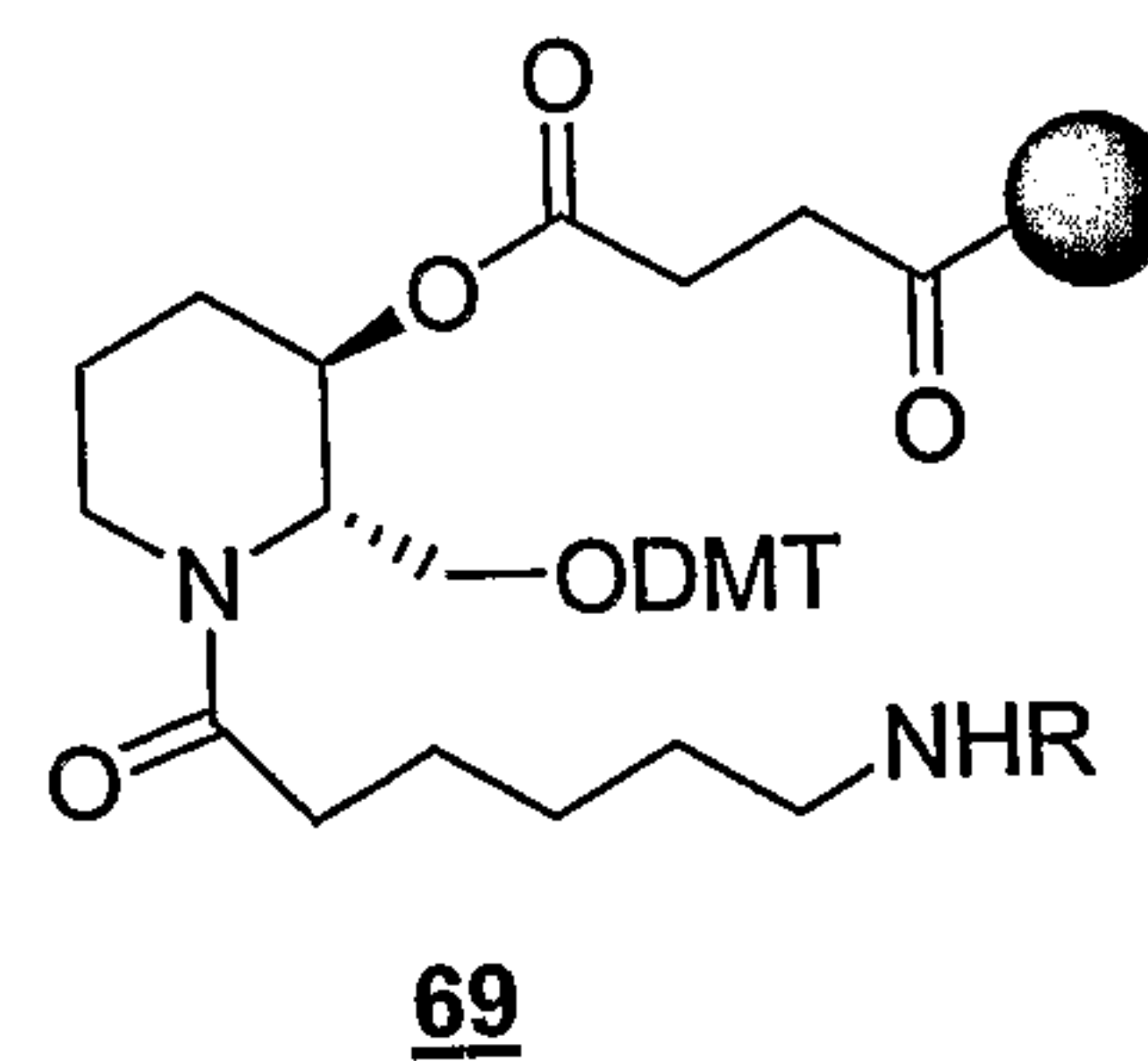
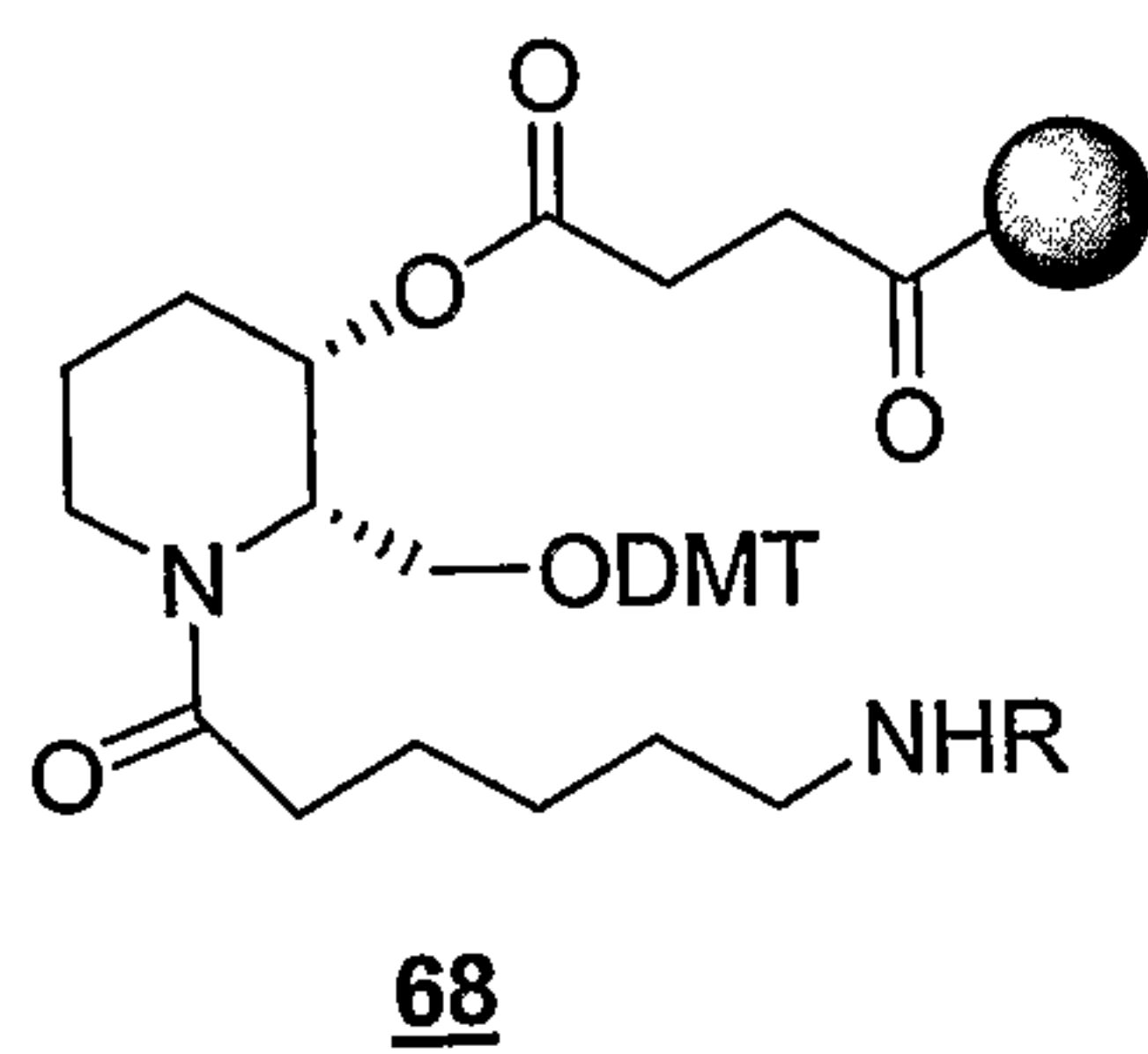
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67

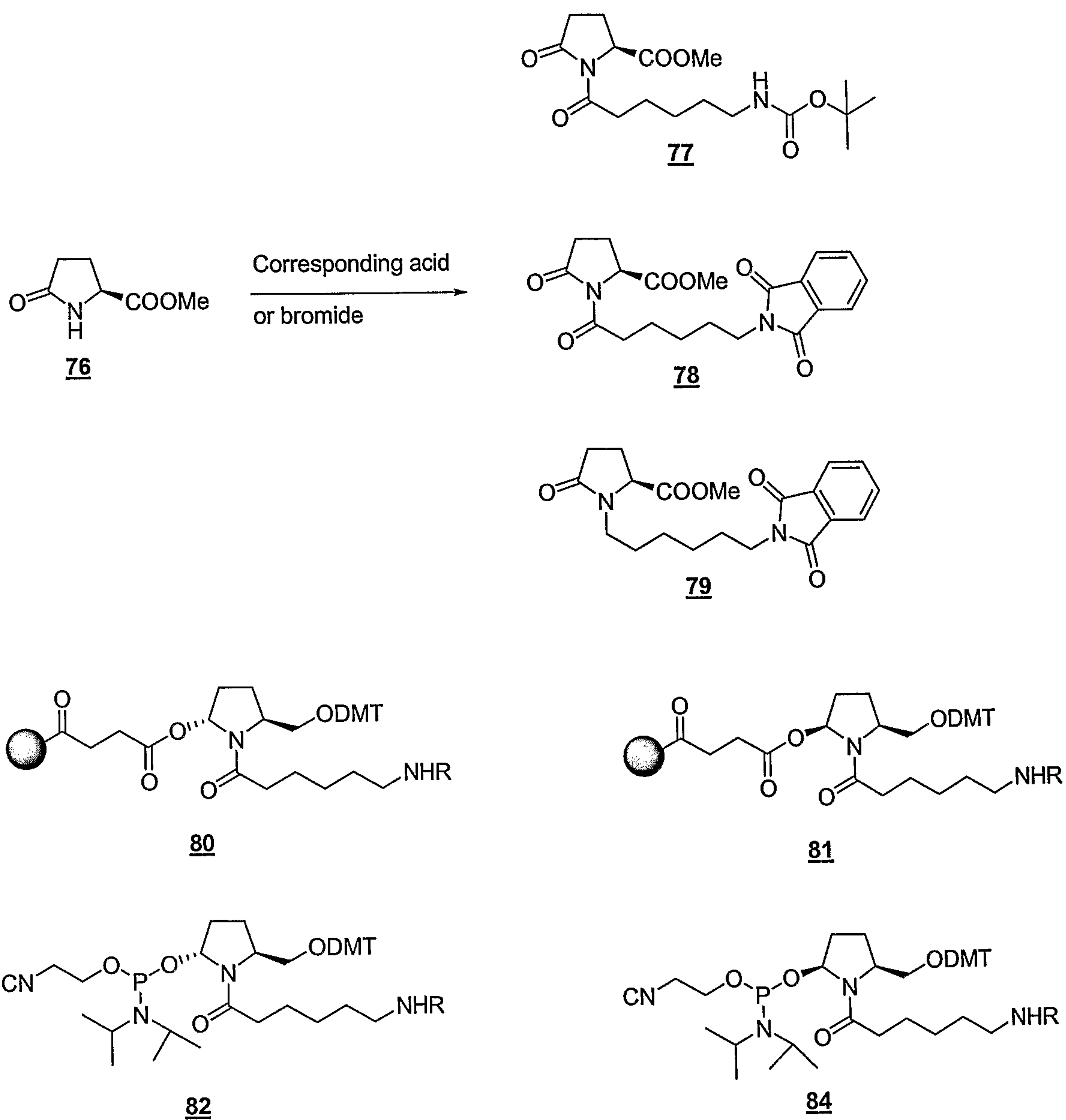
Piperidine series ligands:

Similar to pyrroline series piperidine series can be synthesised

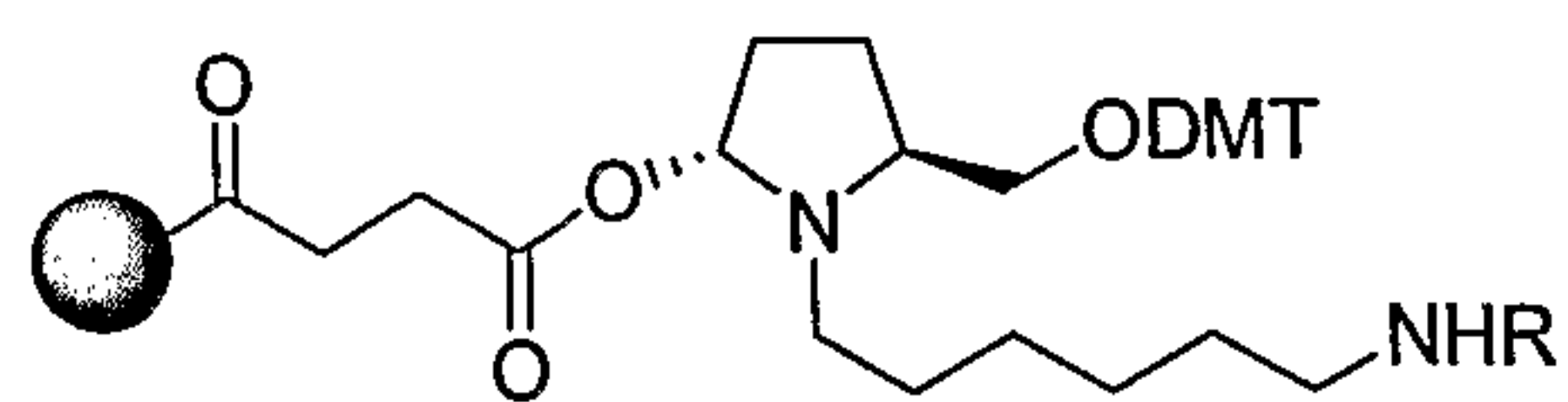
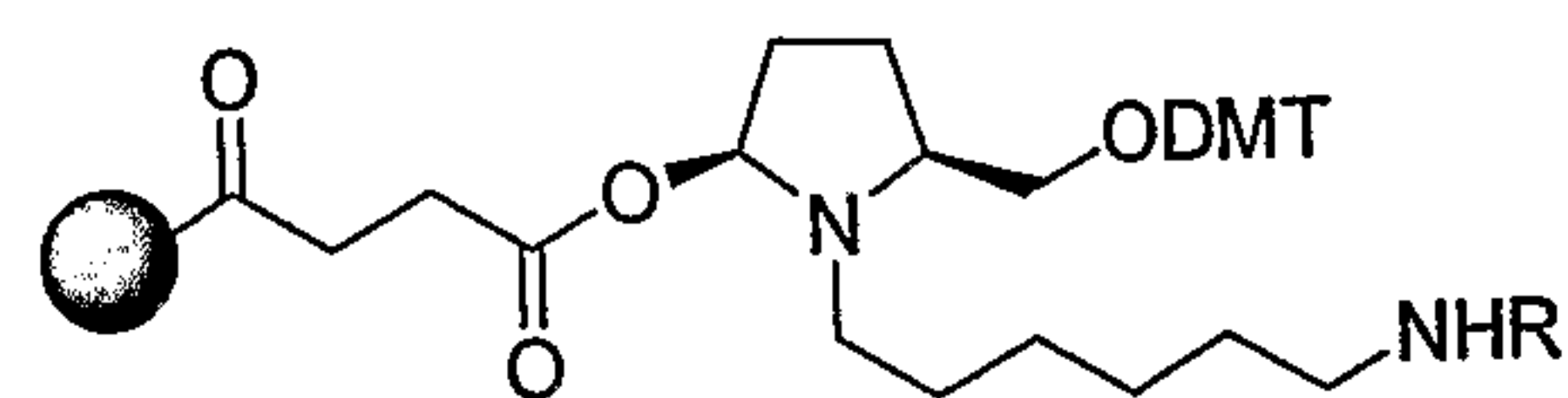
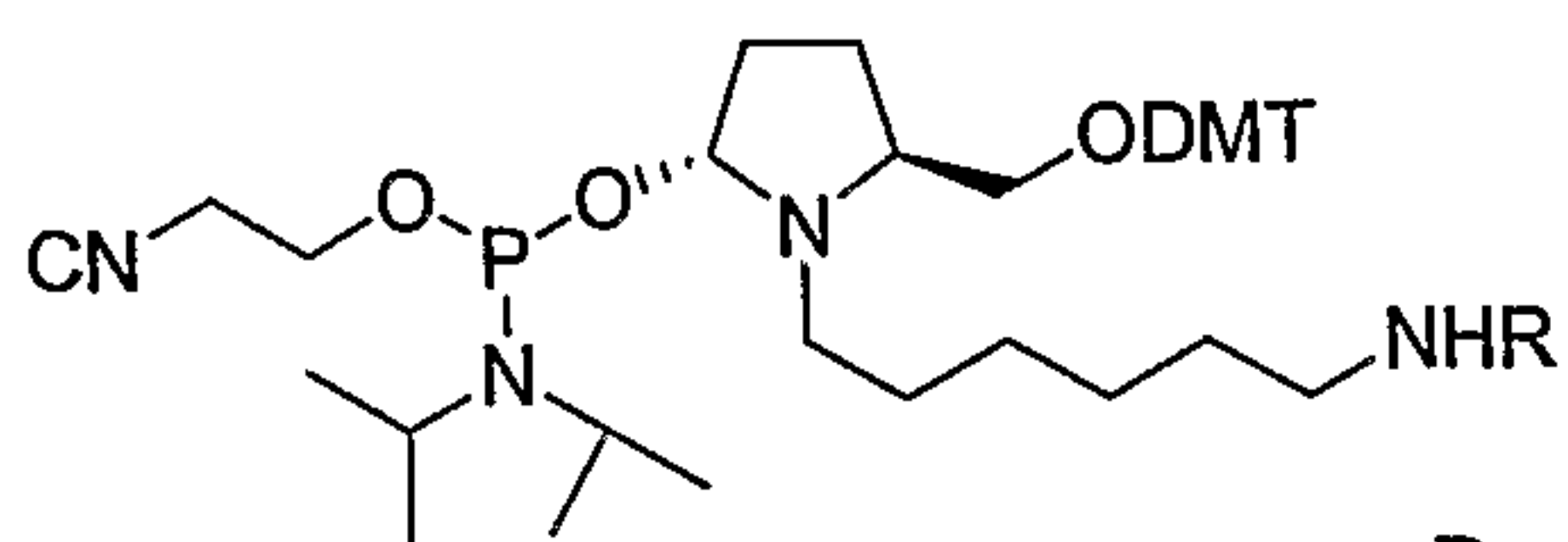


Hydroxy proline series linkers:

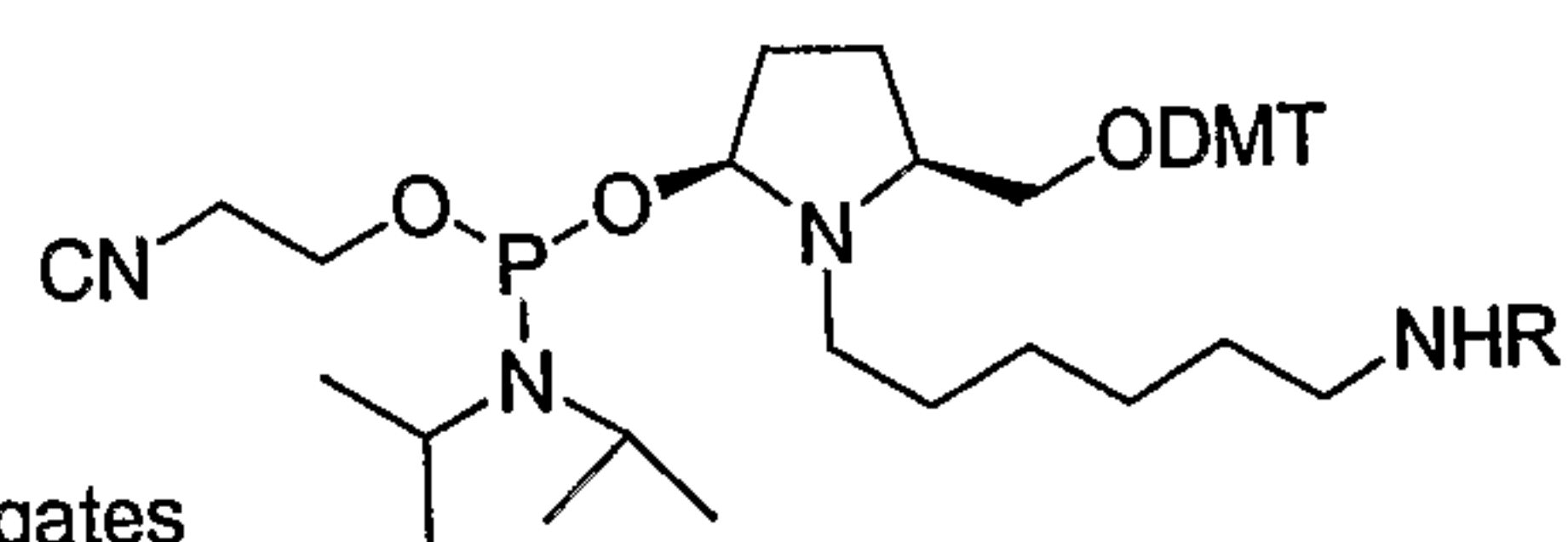
From commercially available cis-3-hydroxy proline and (s)-pyrrolidone carboxylate



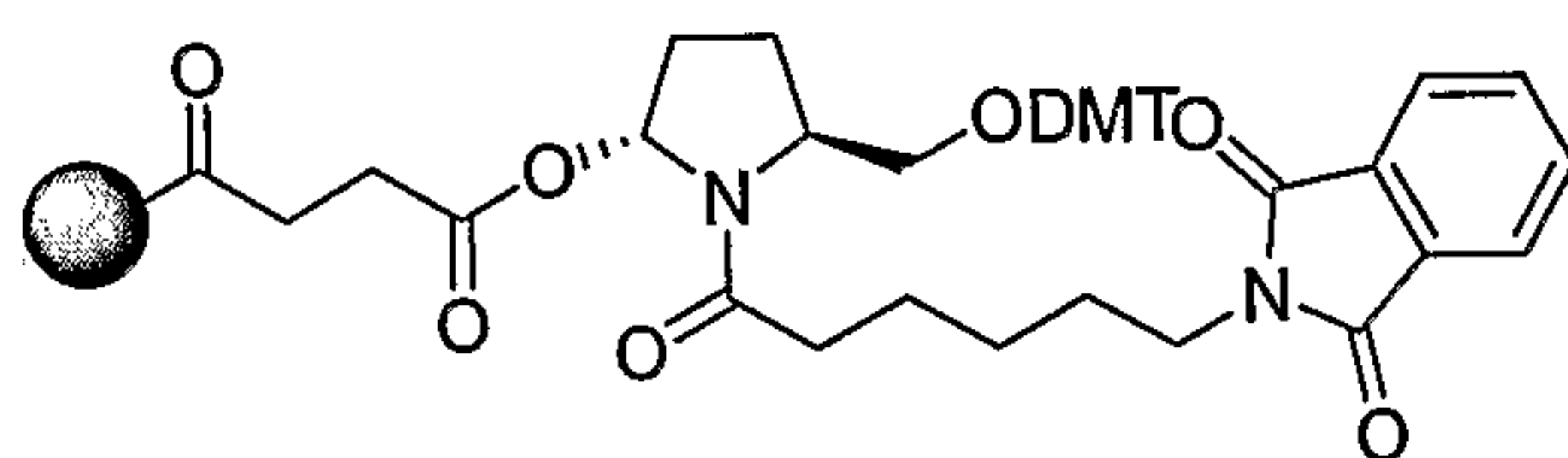
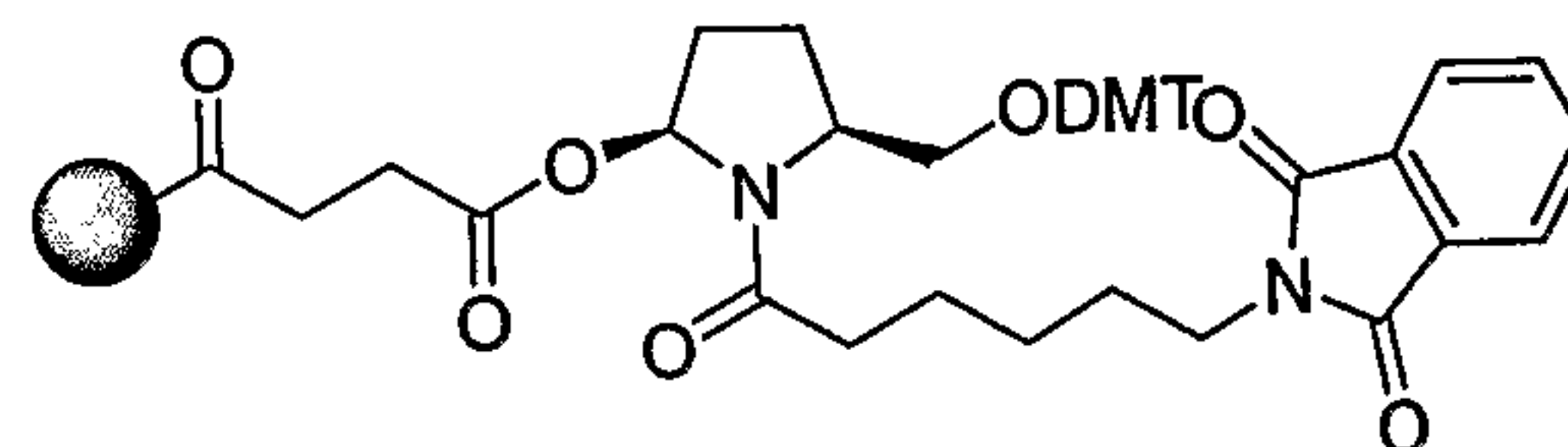
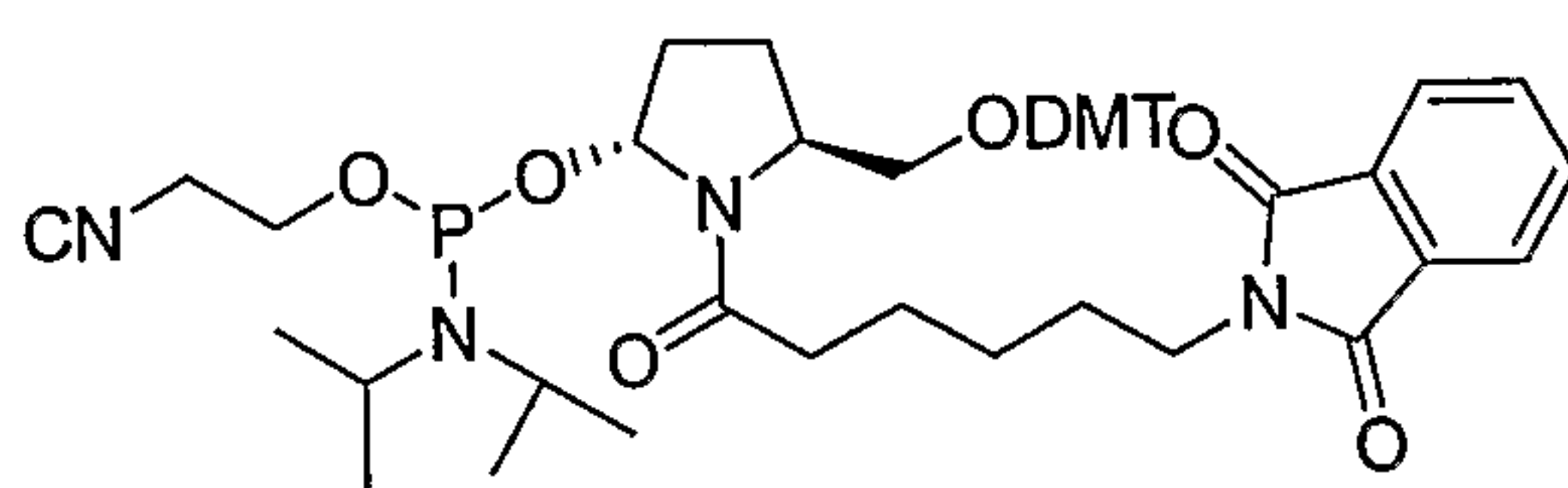
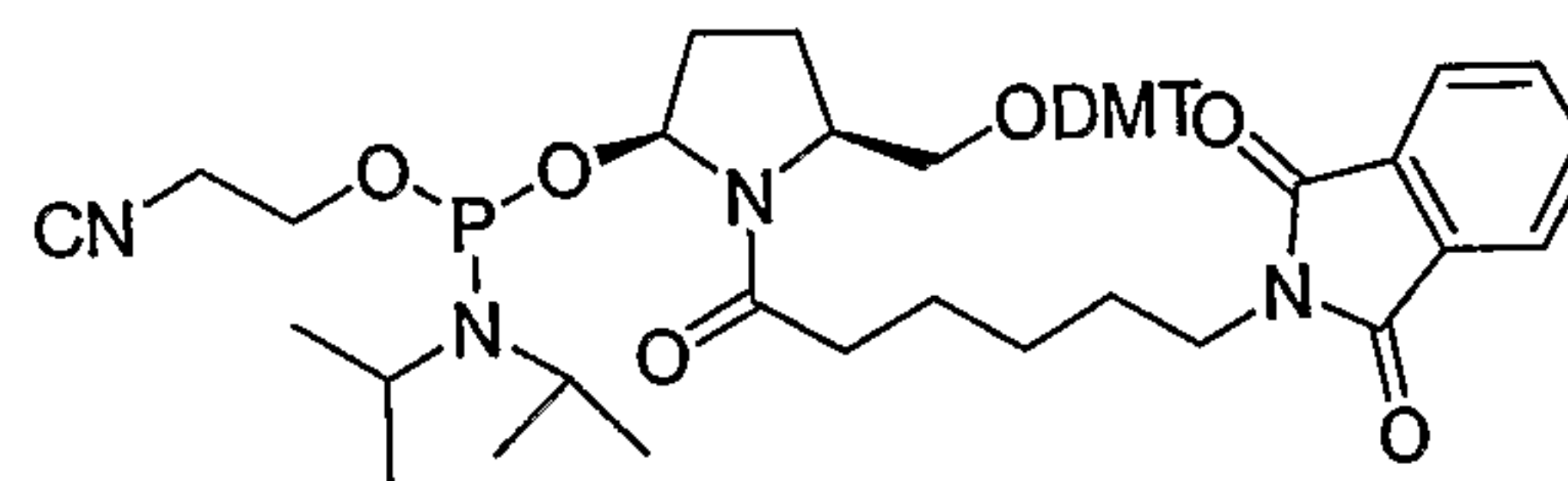
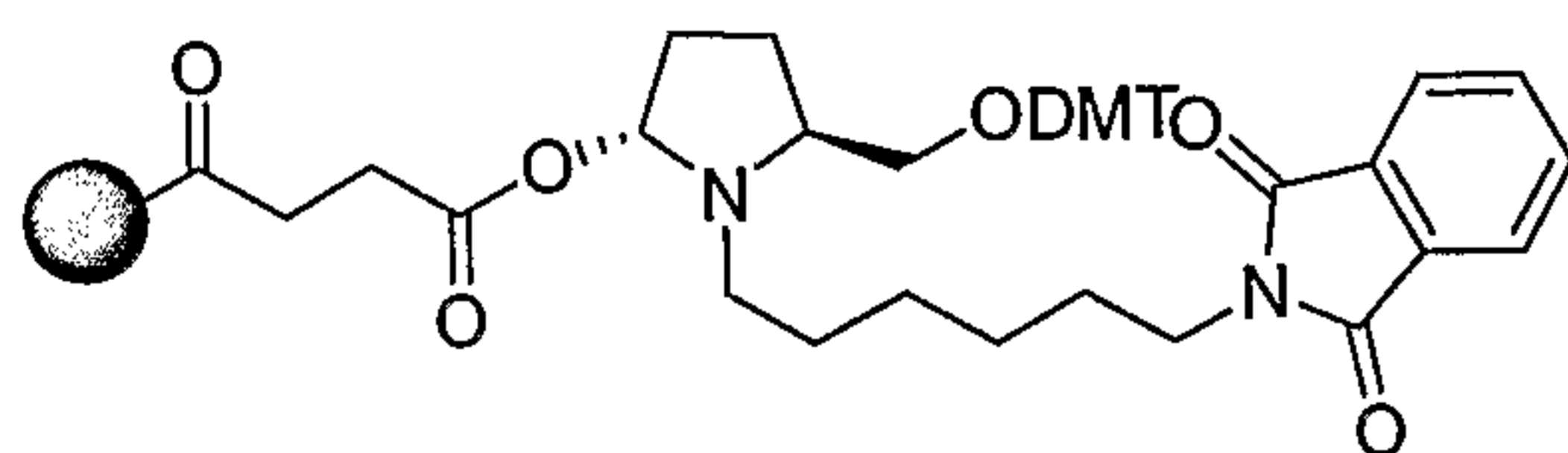
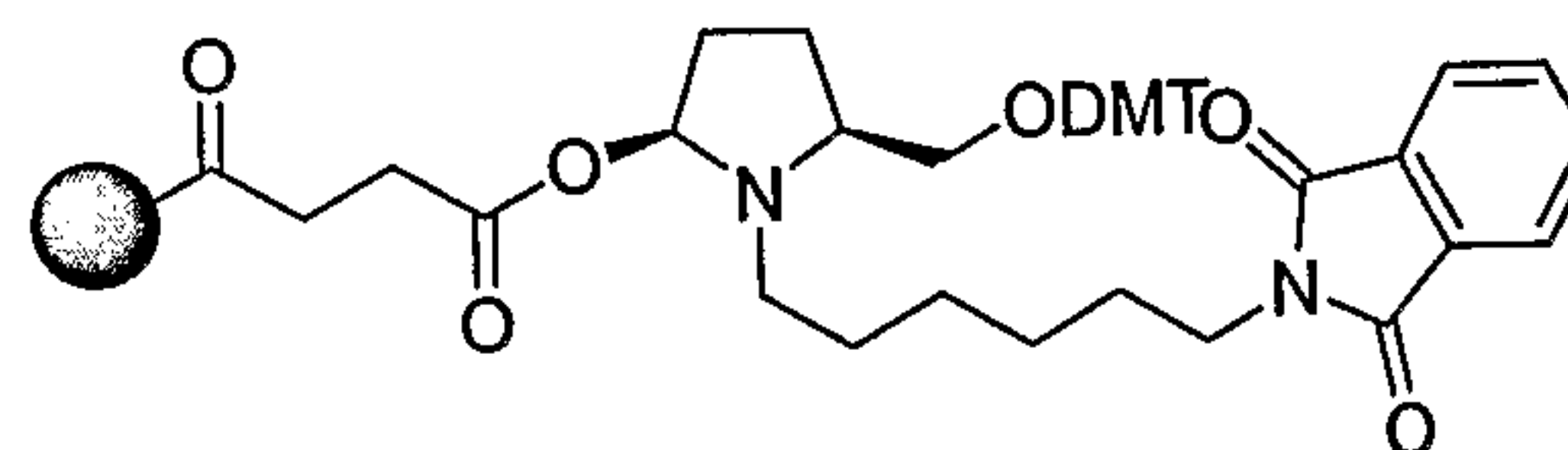
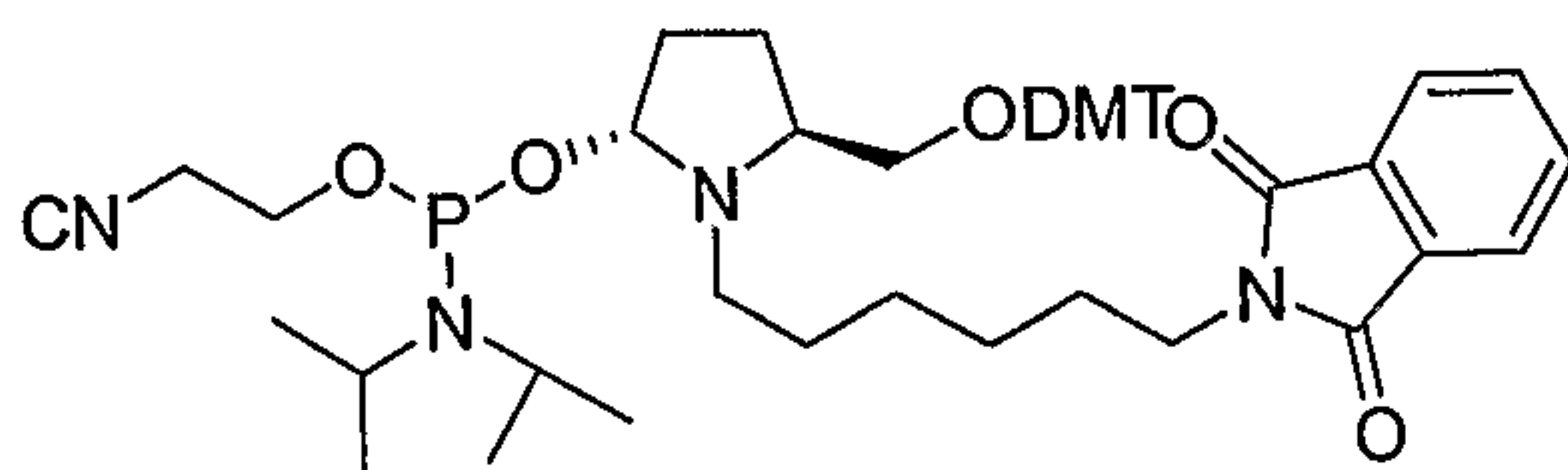
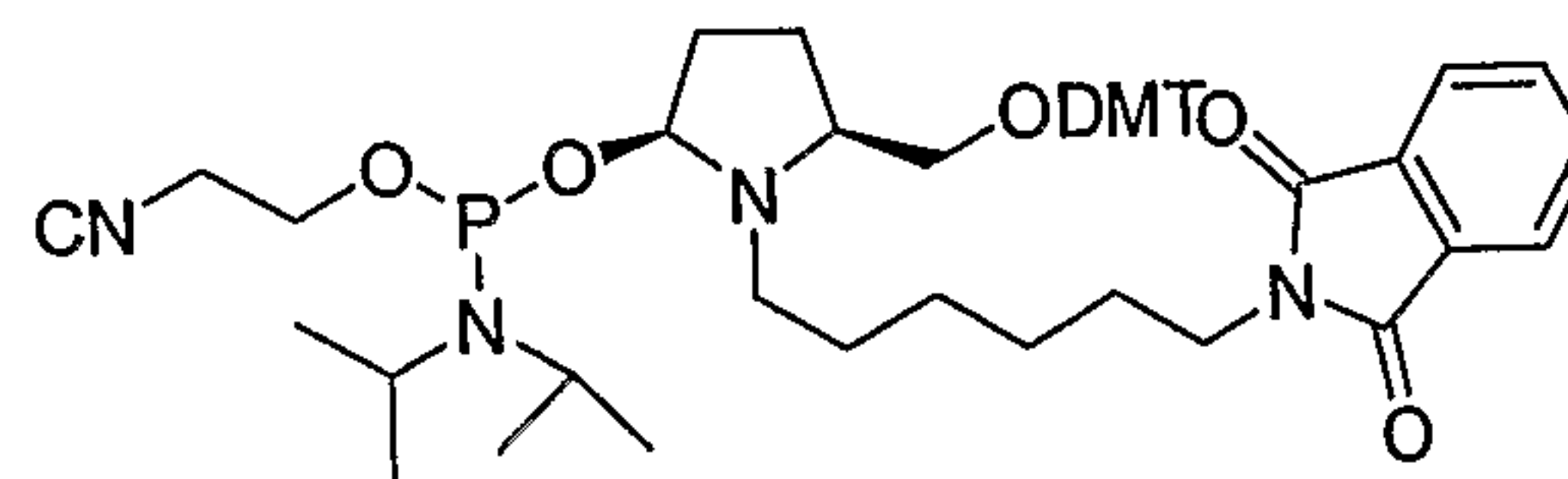
R = Lipophilic conjugates

858687

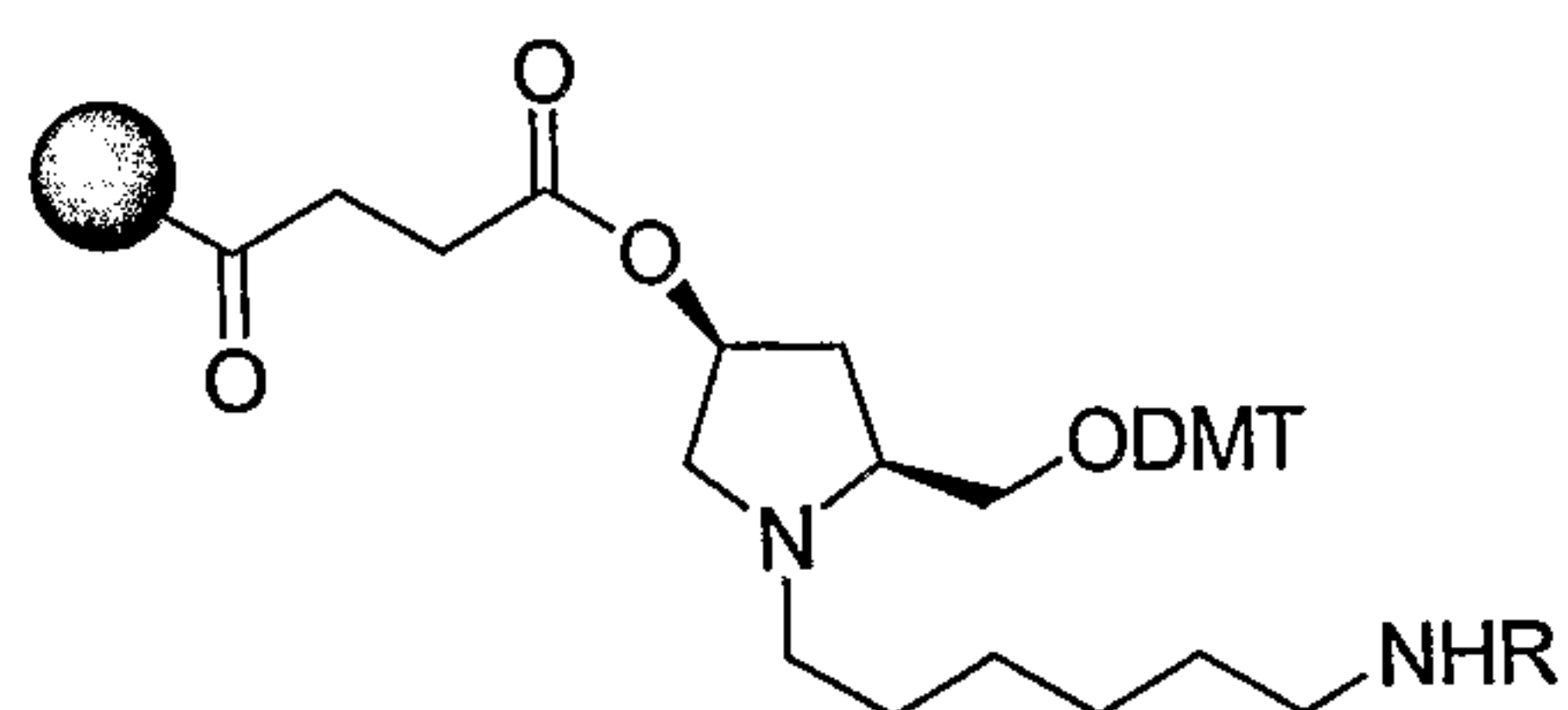
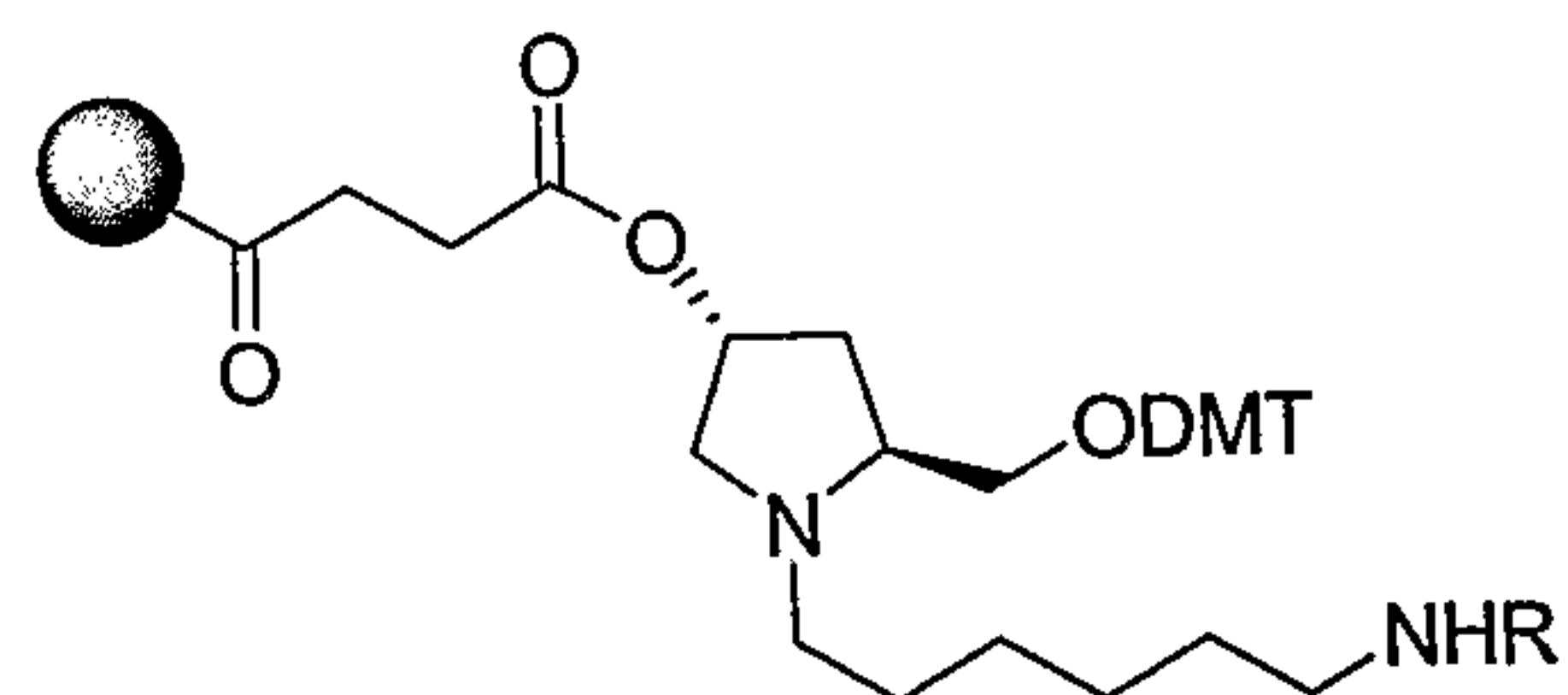
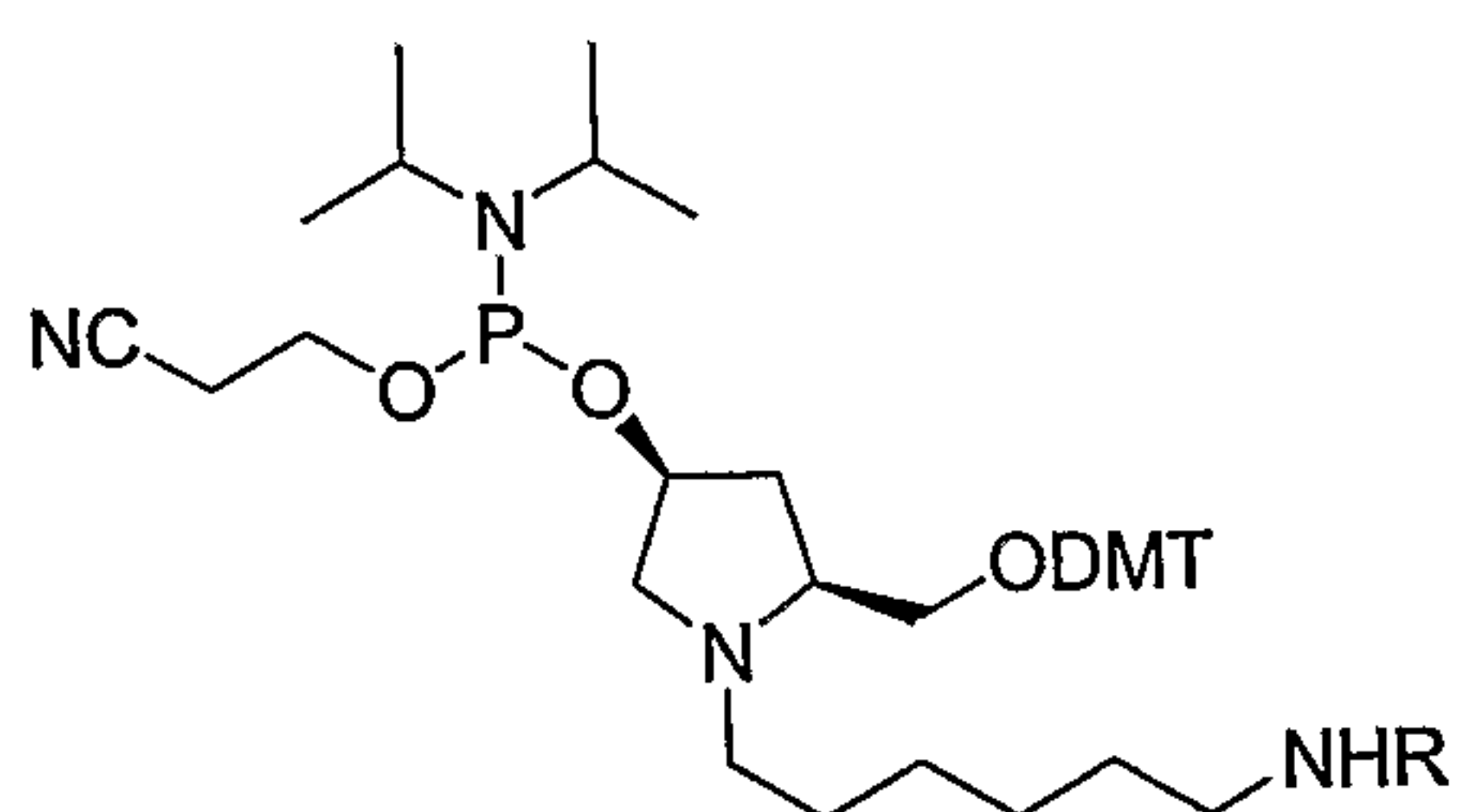
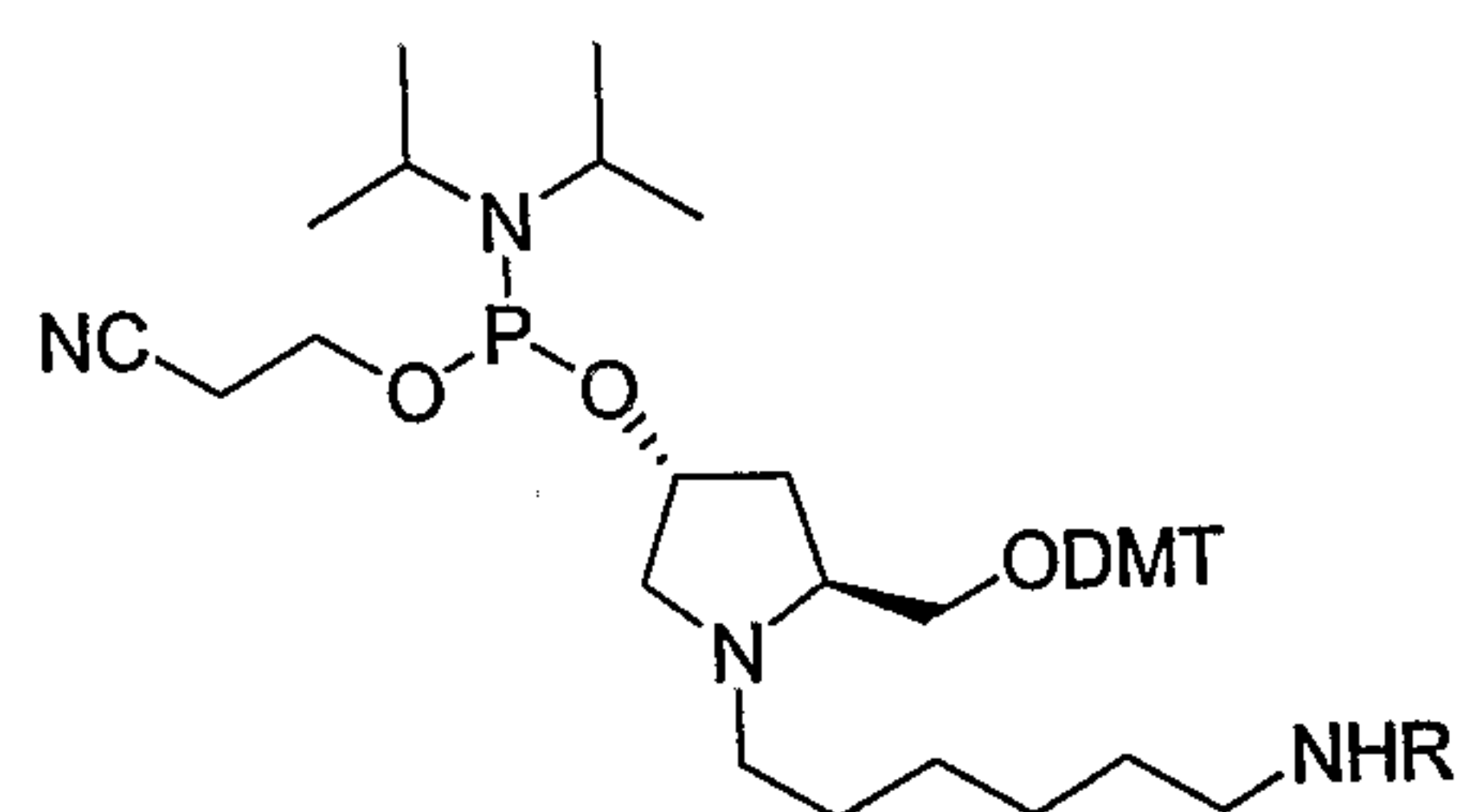
R = Lipophilic conjugates

88

Phthalimide derivative to stabilise siRNA

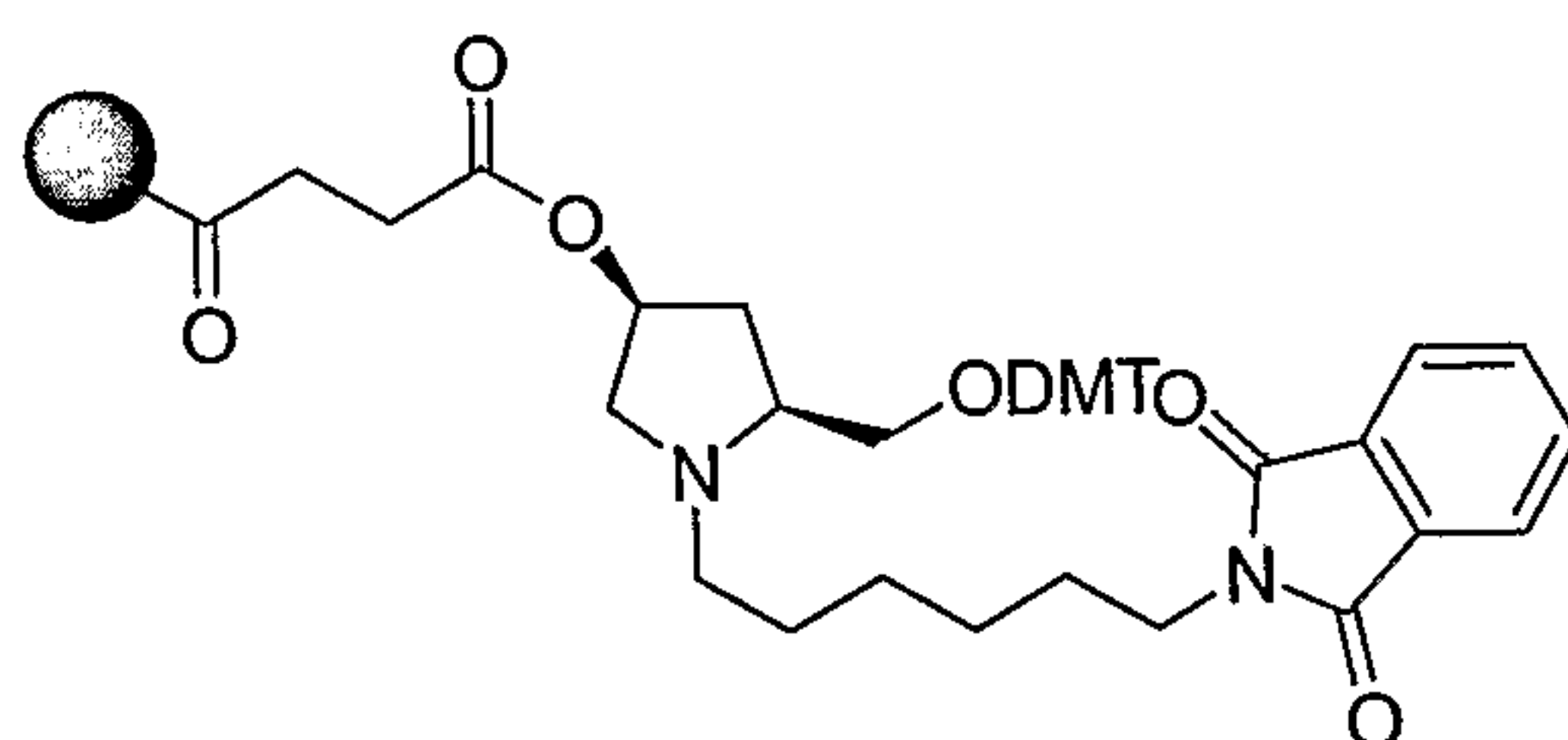
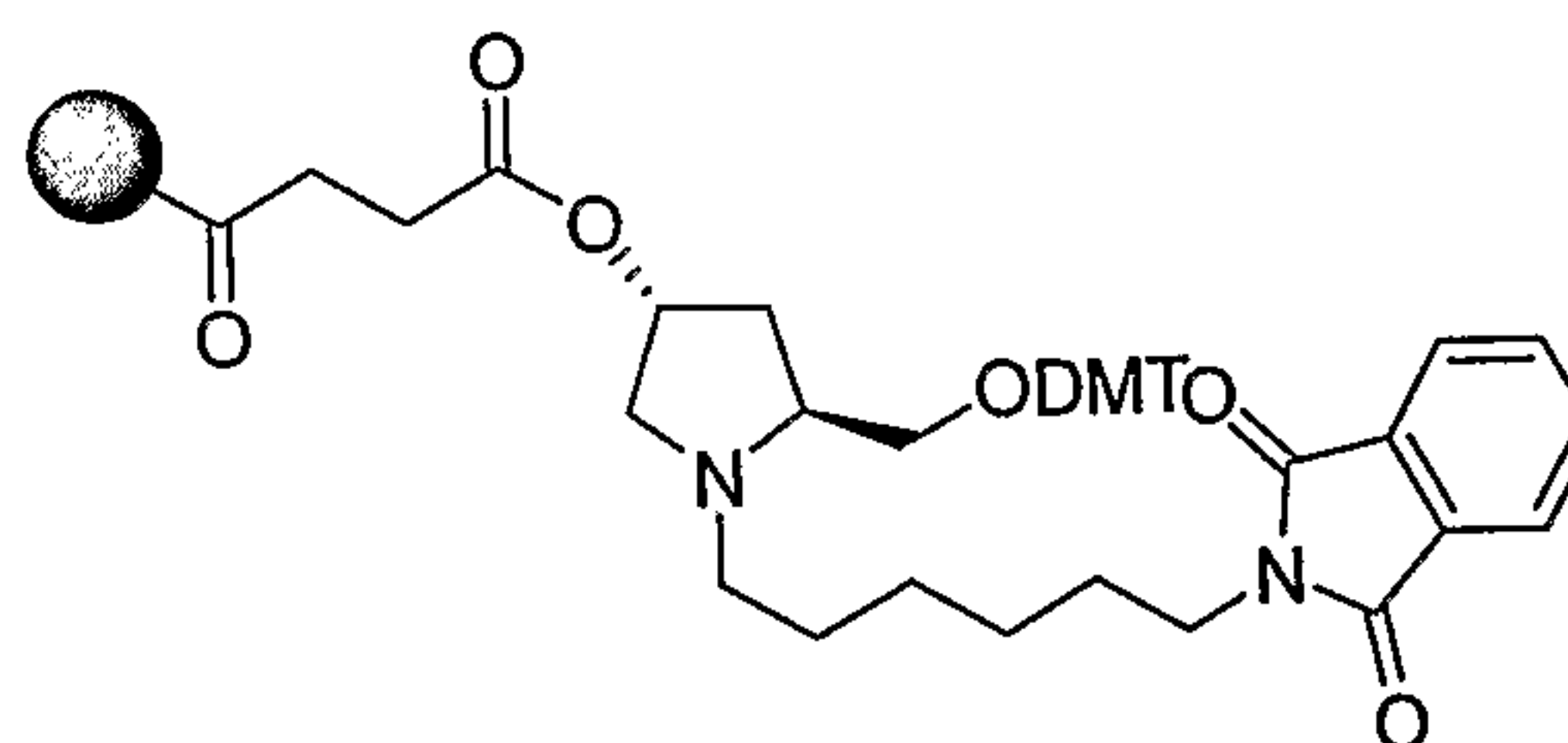
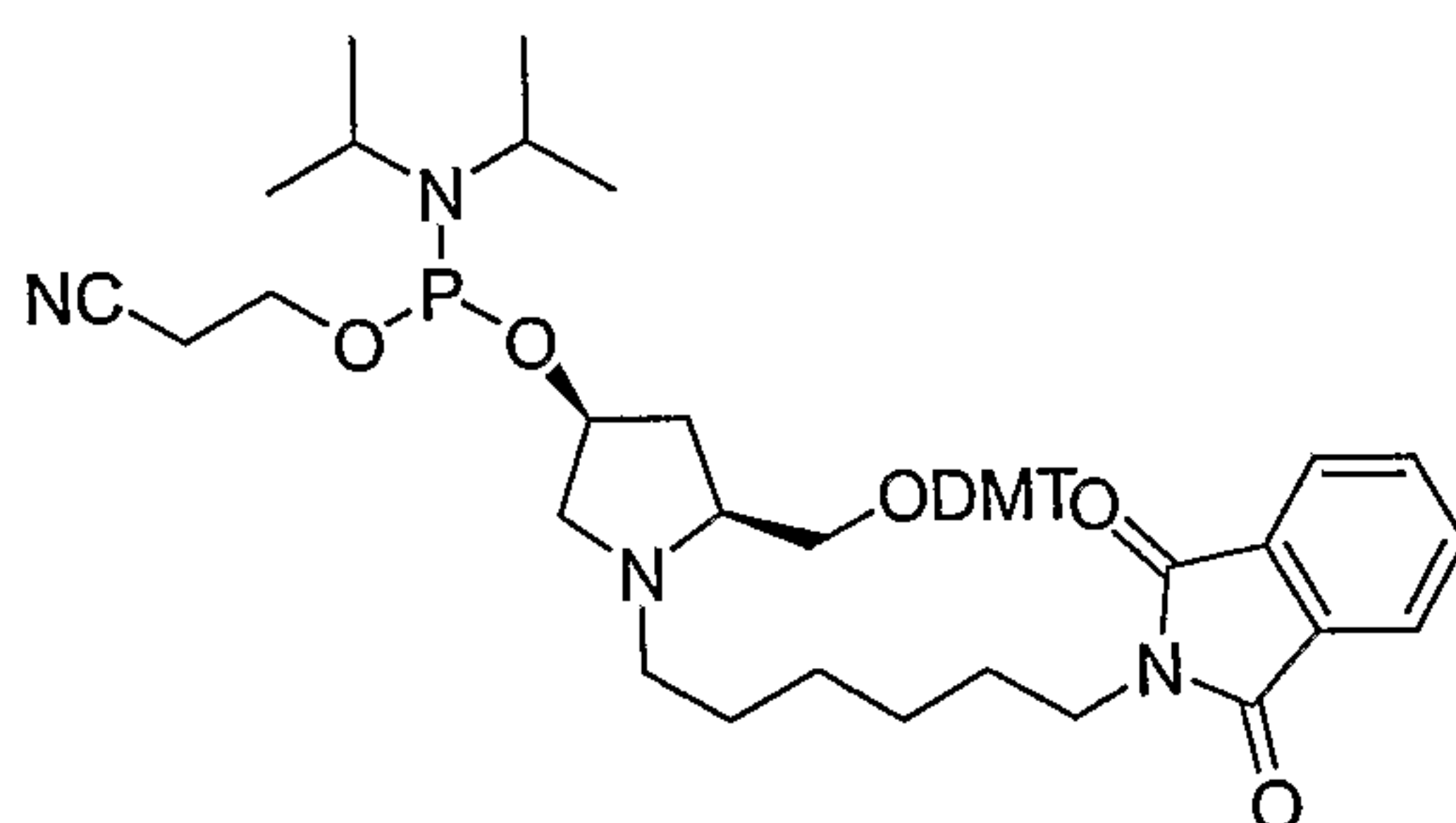
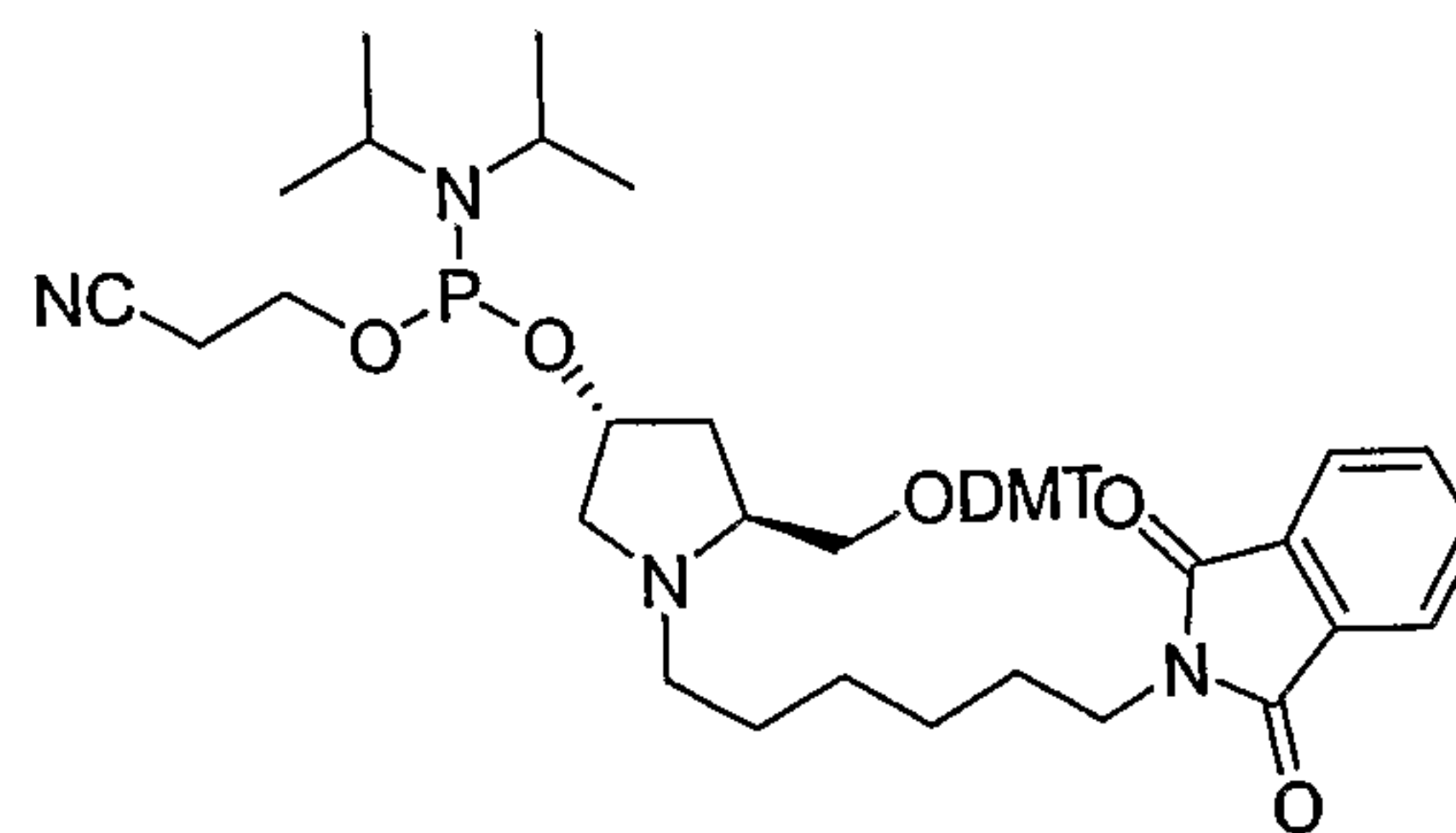
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4-hydroxy proline derivatives

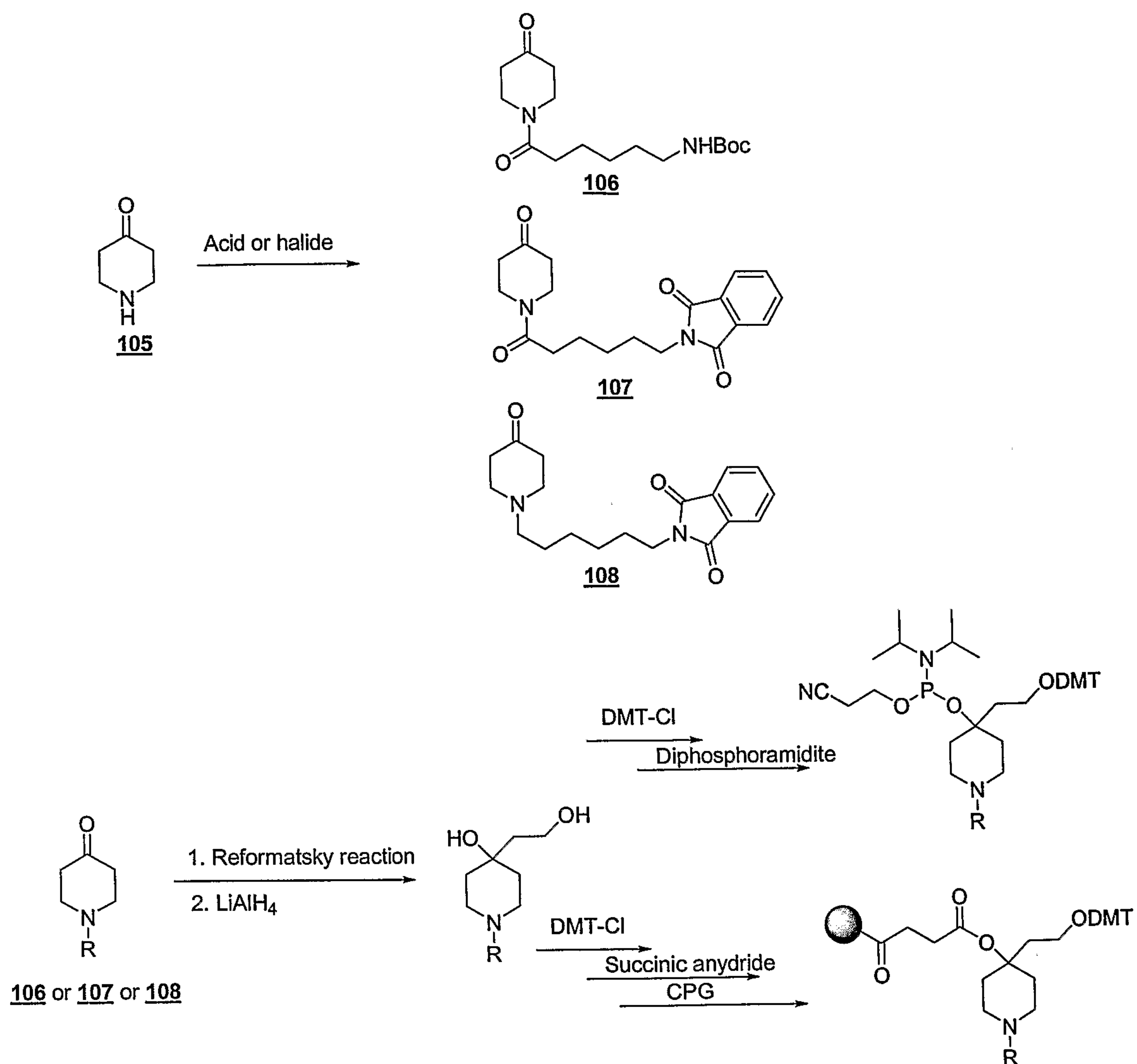
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R = Lipophilic conjugates

Phthalimido derivatives

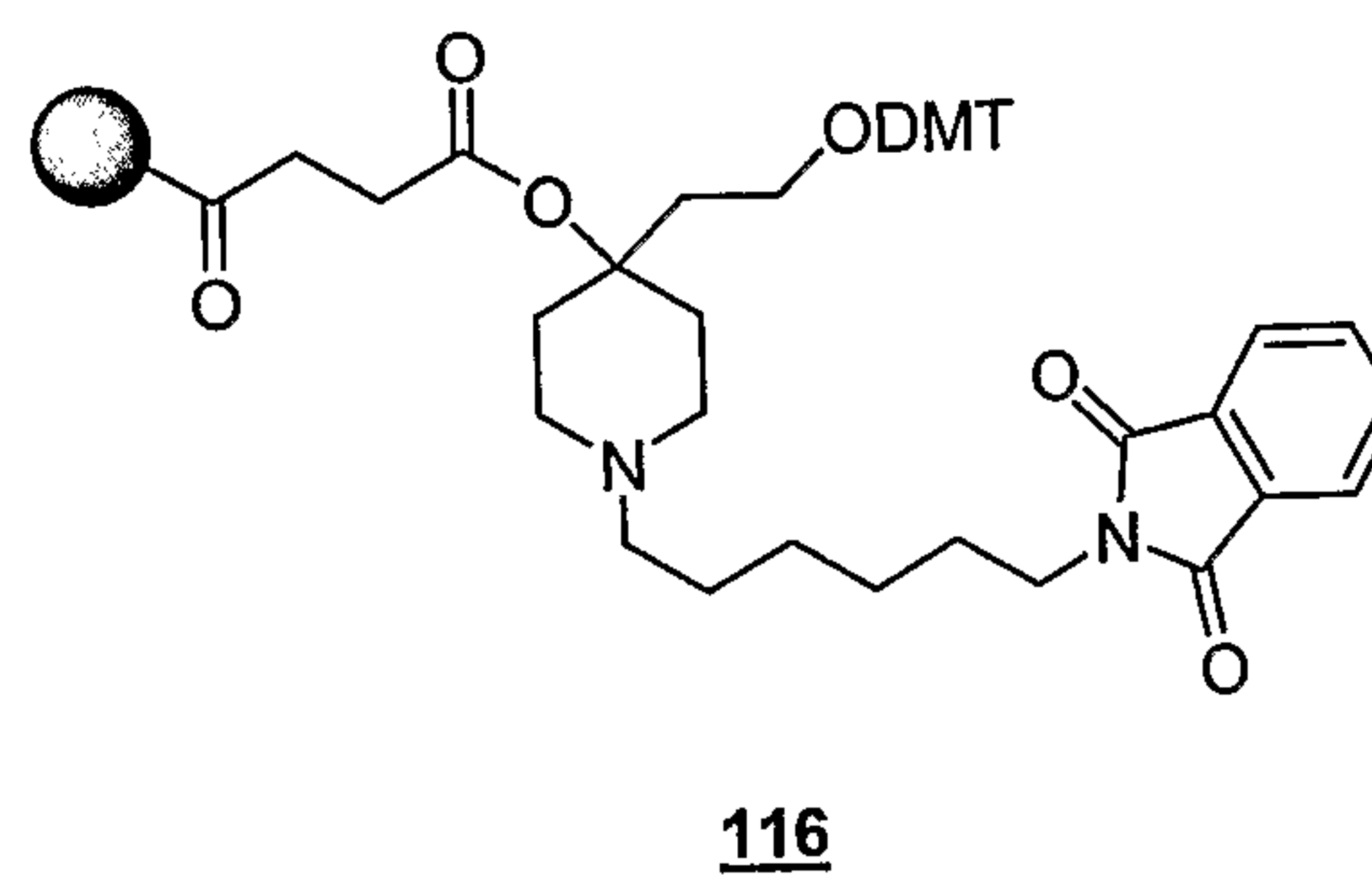
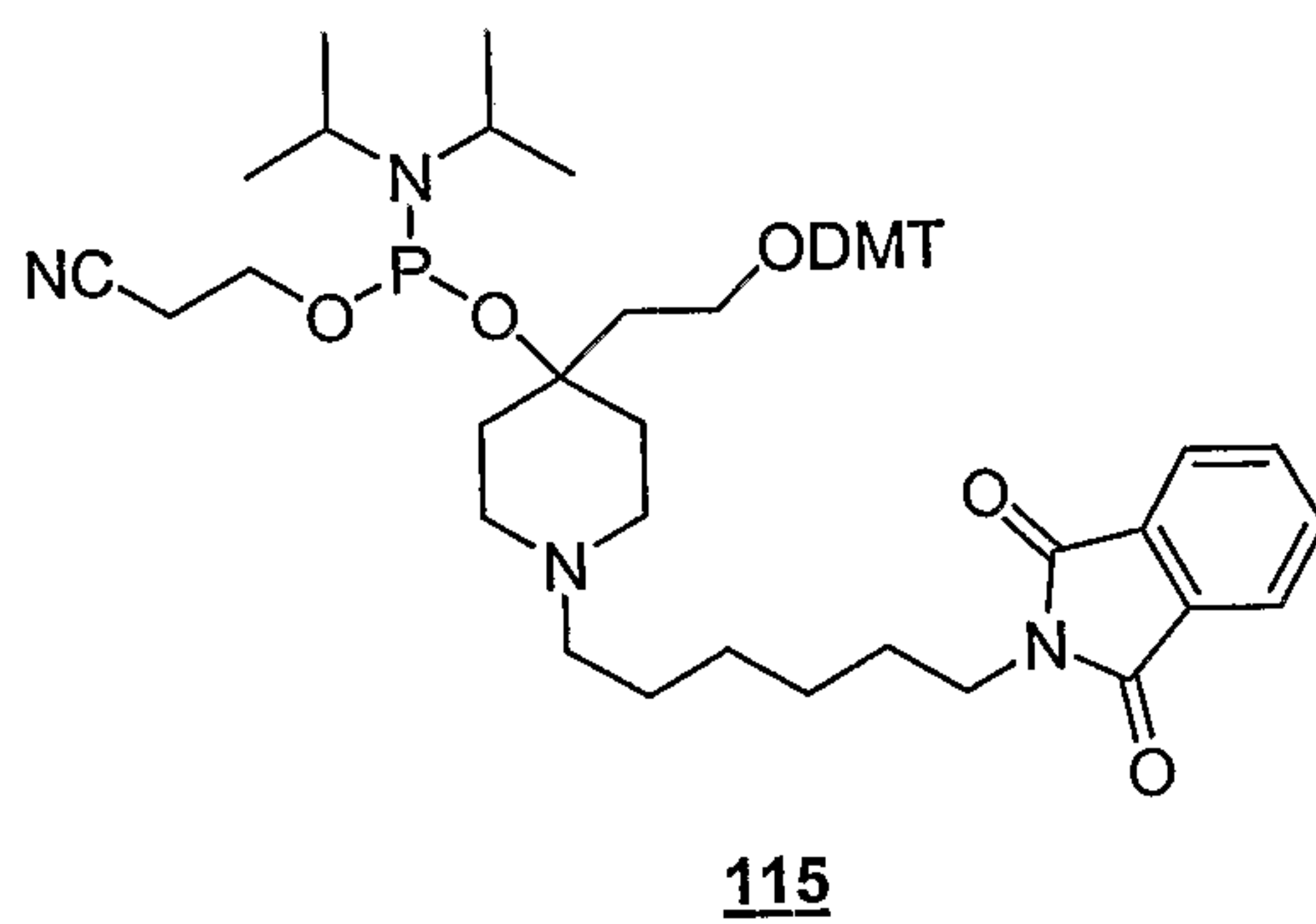
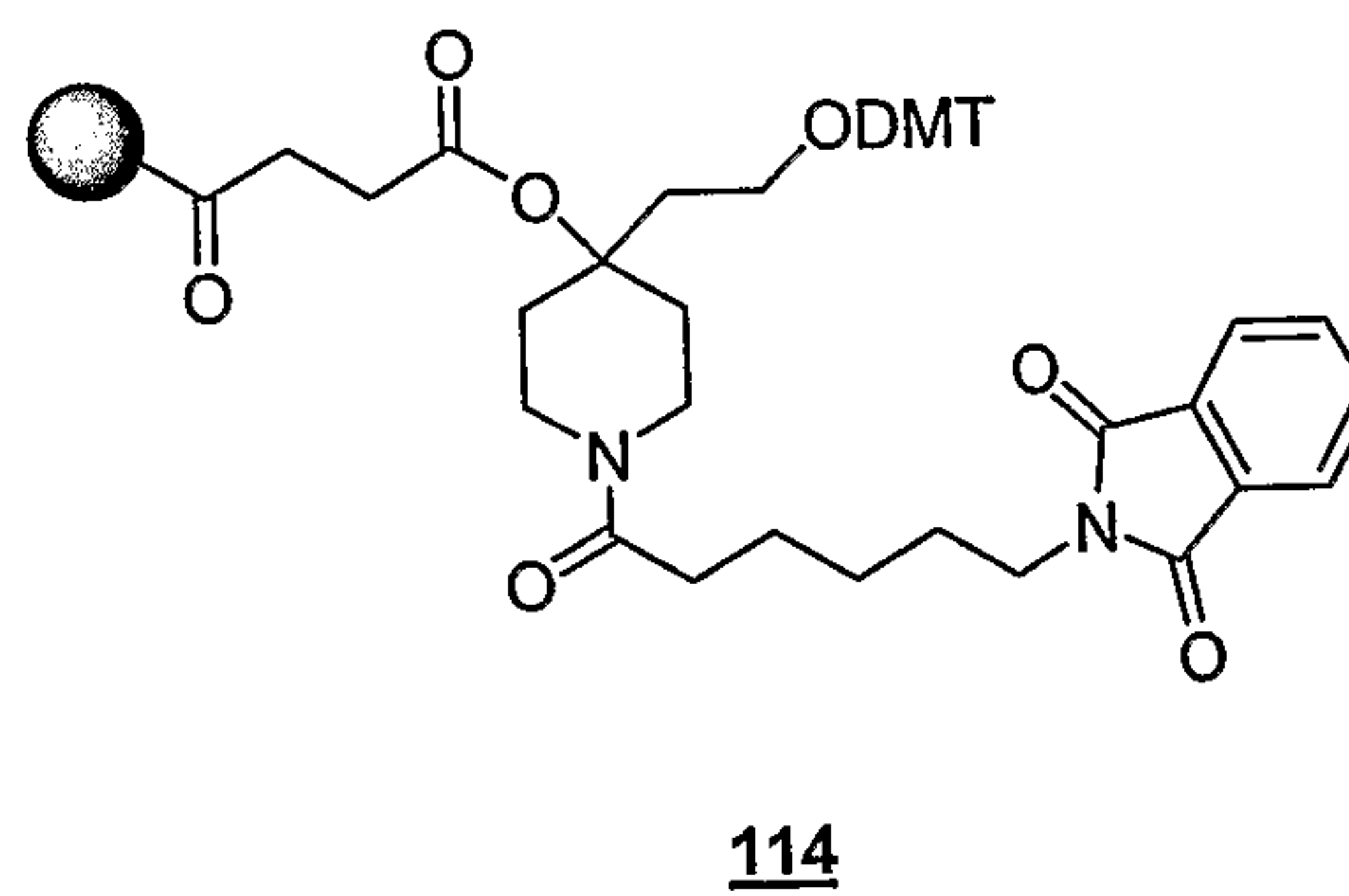
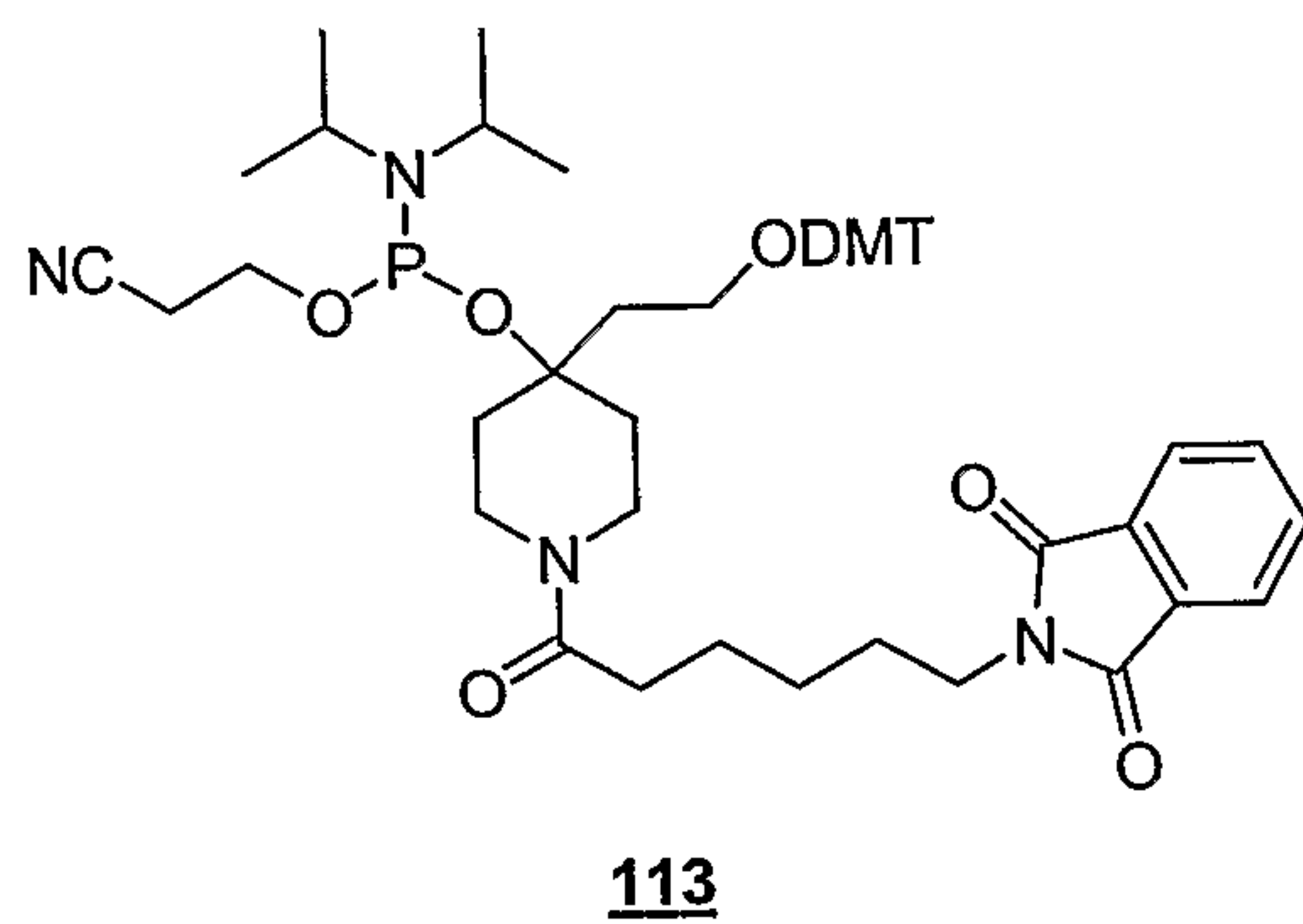
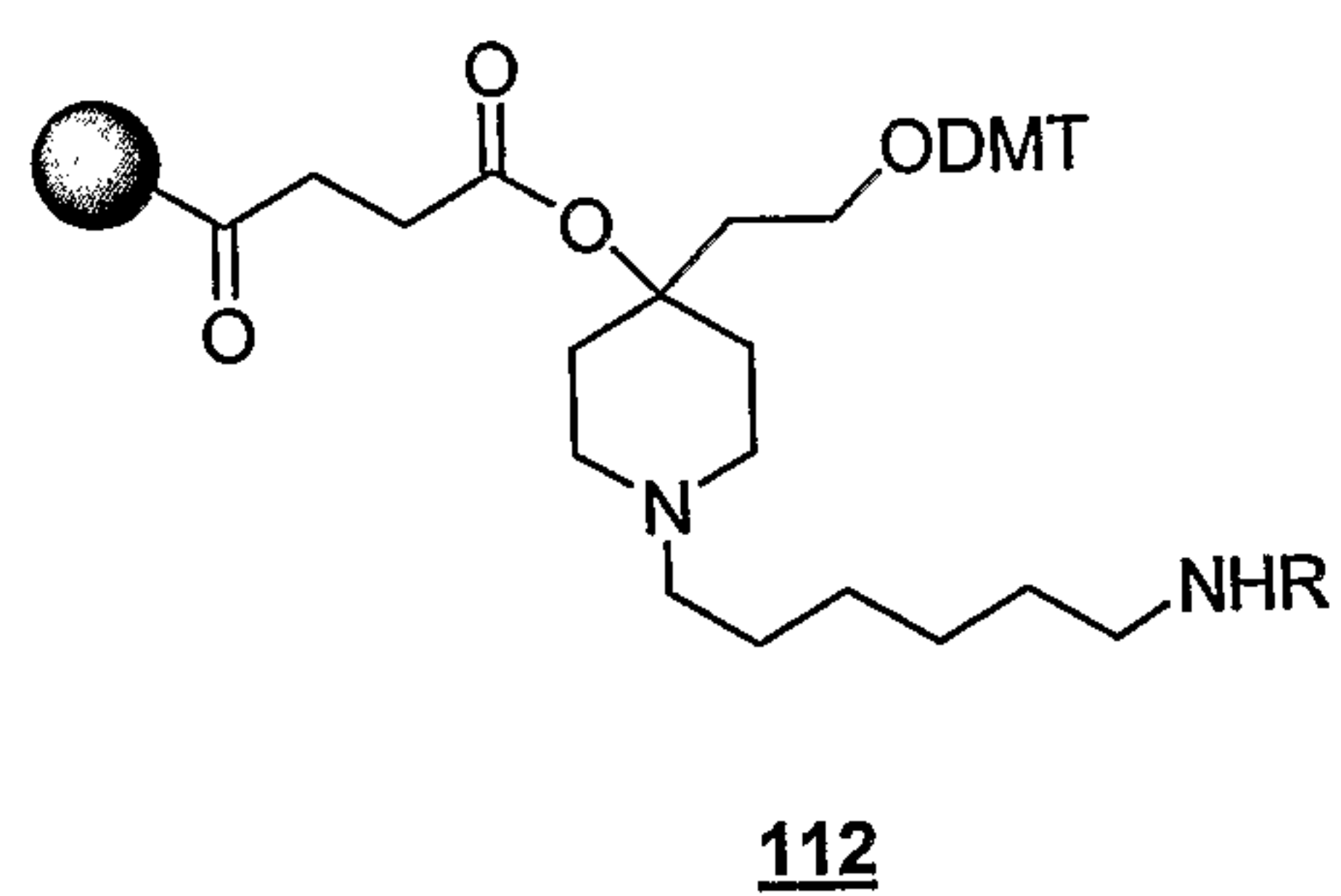
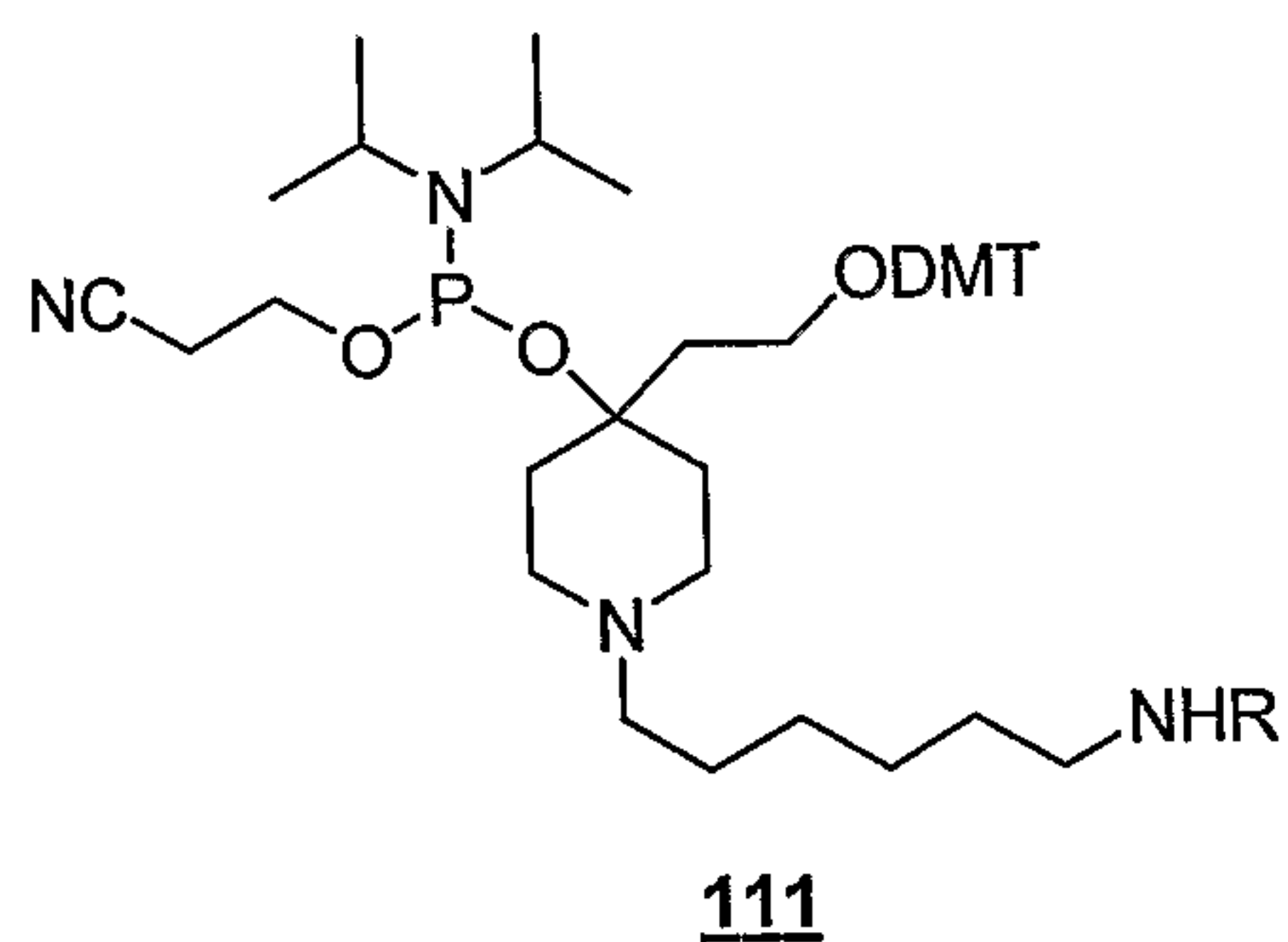
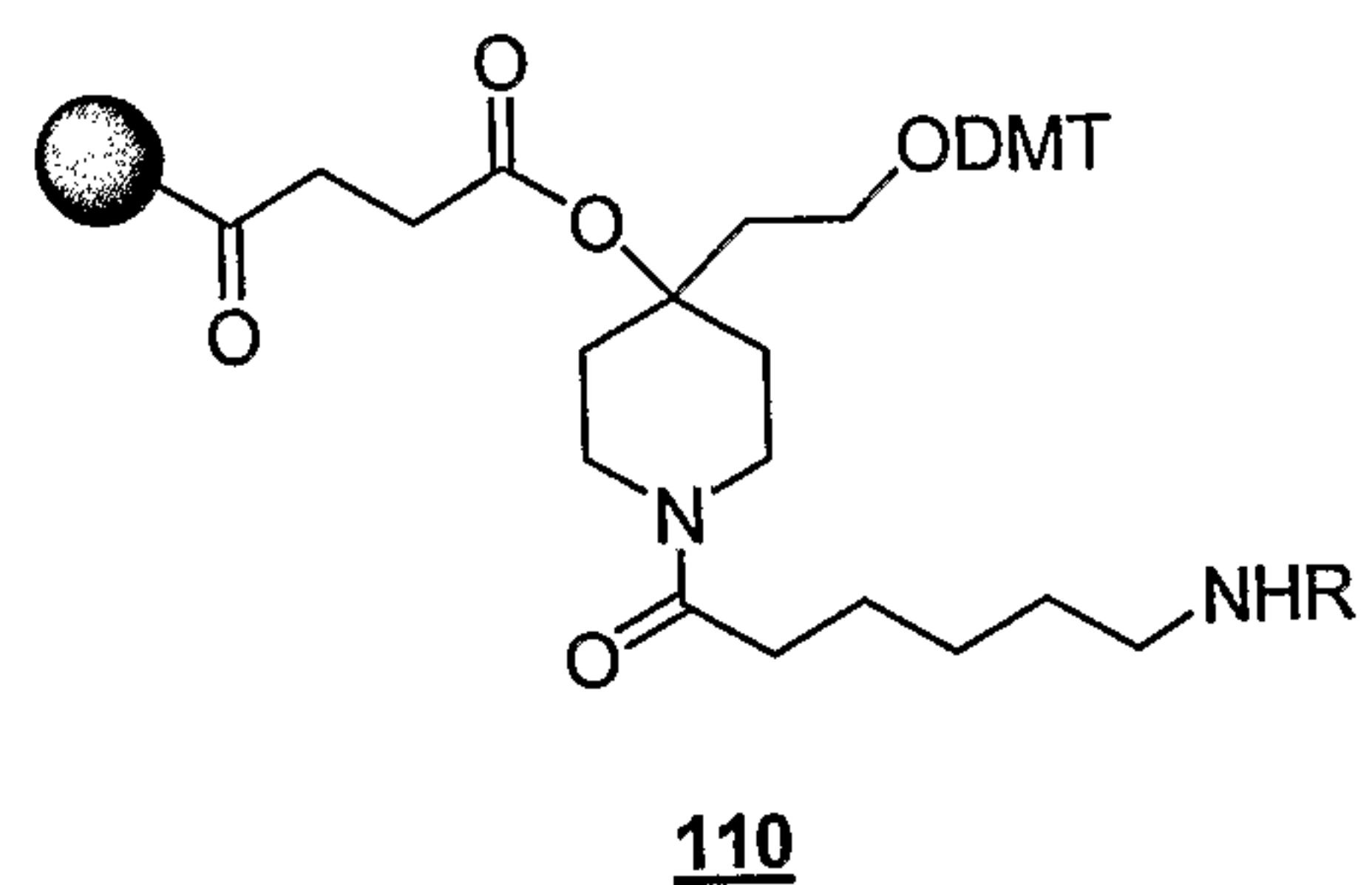
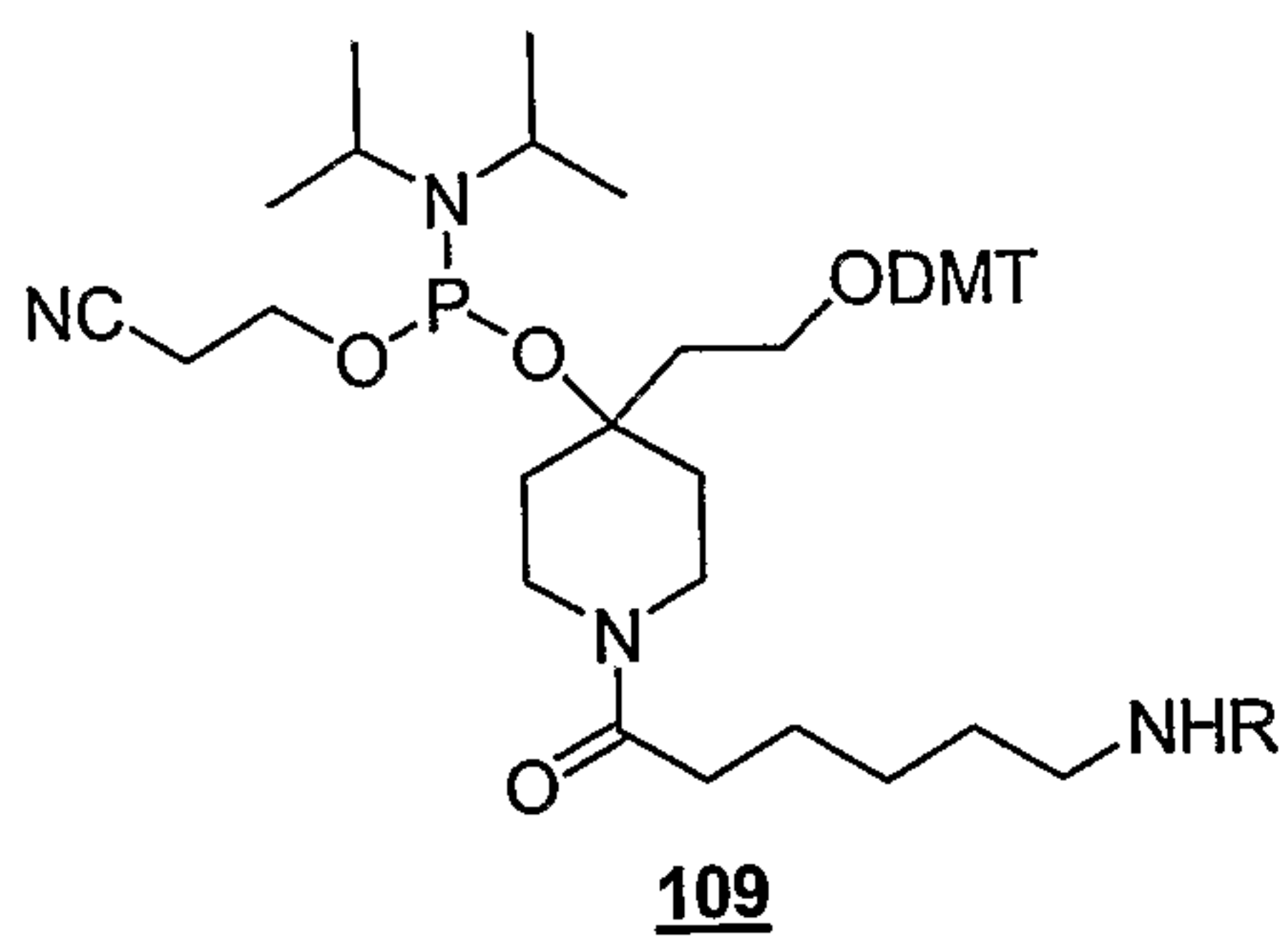
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Synthesis of 6-membered linker

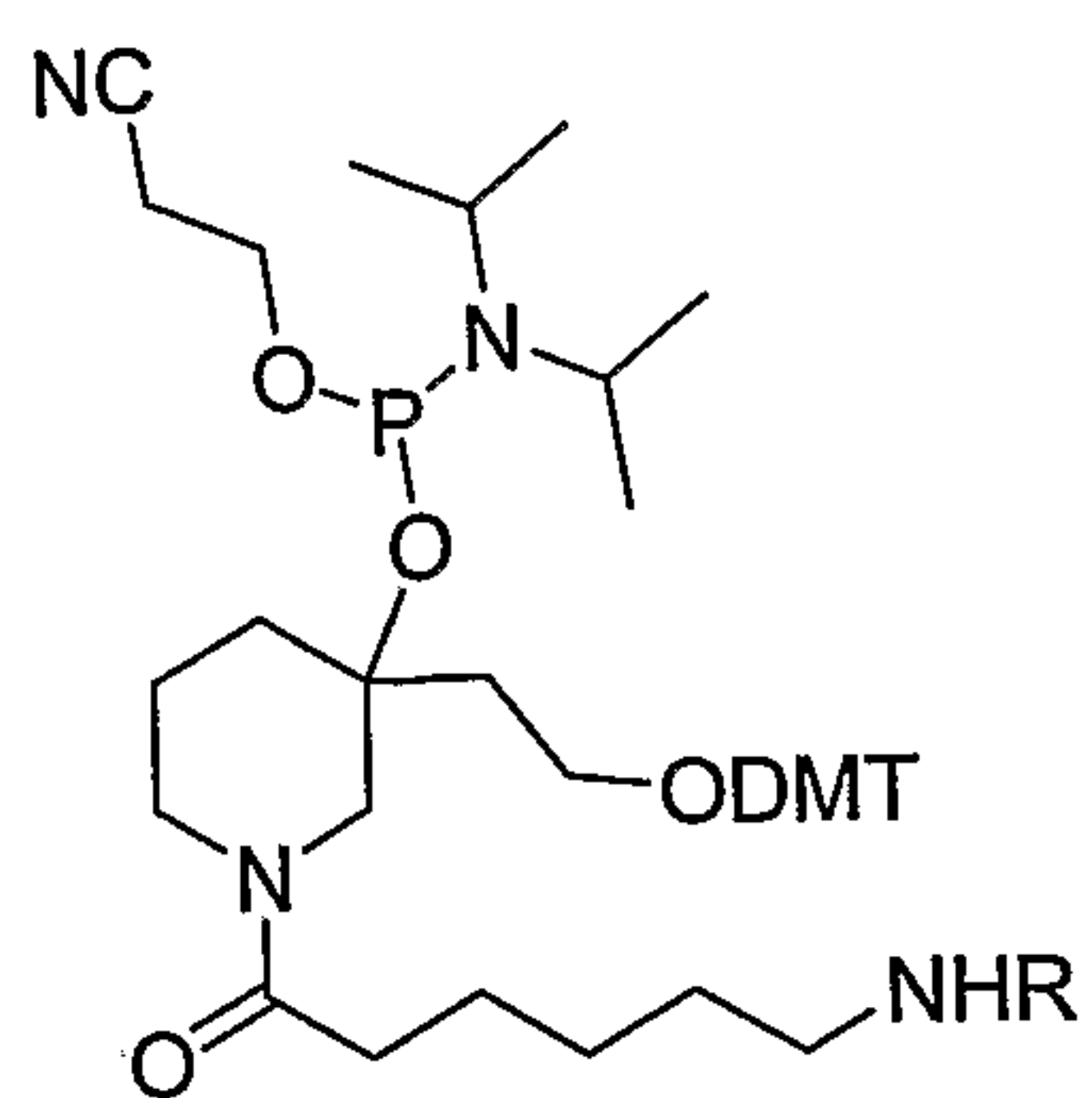
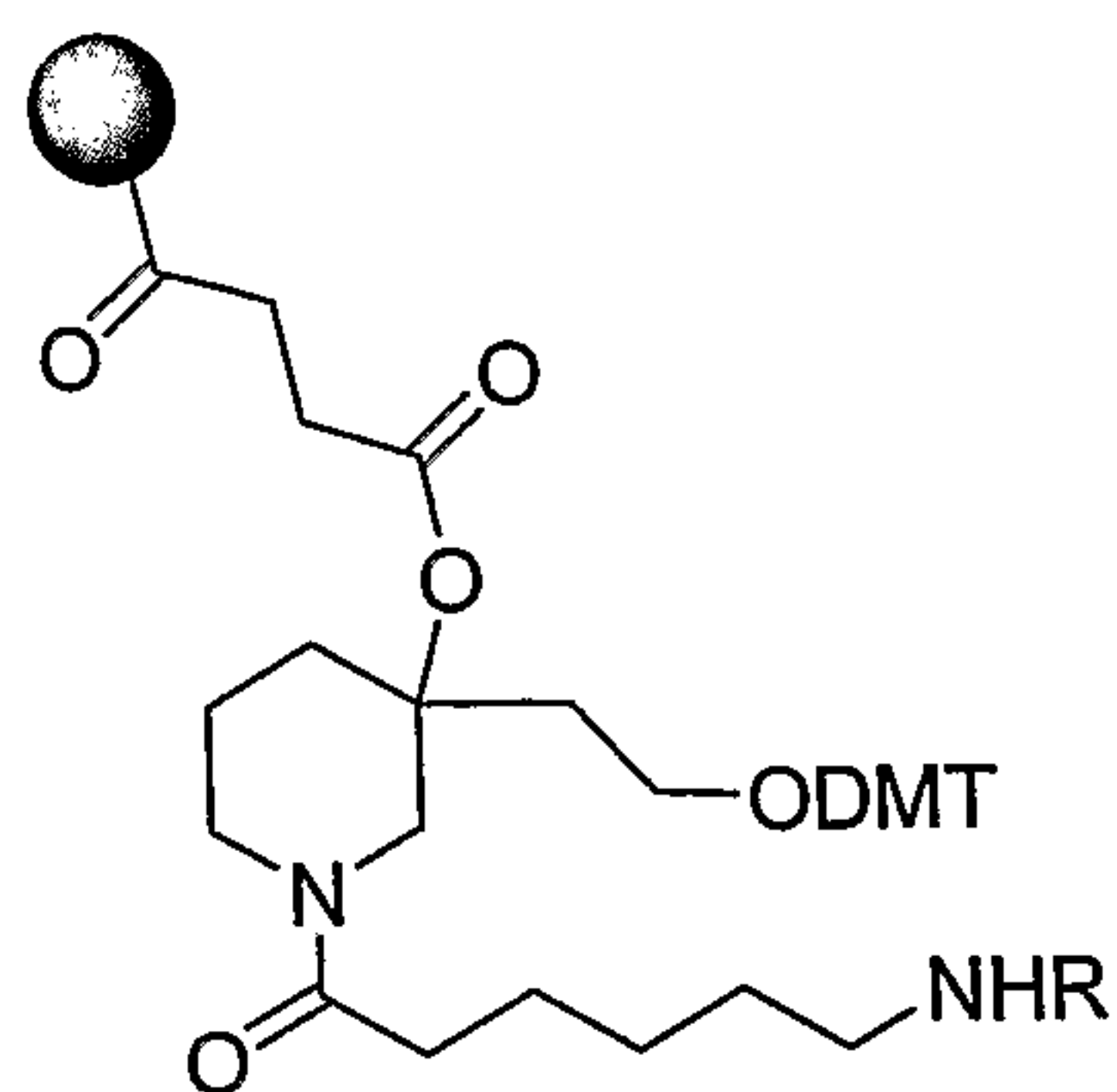
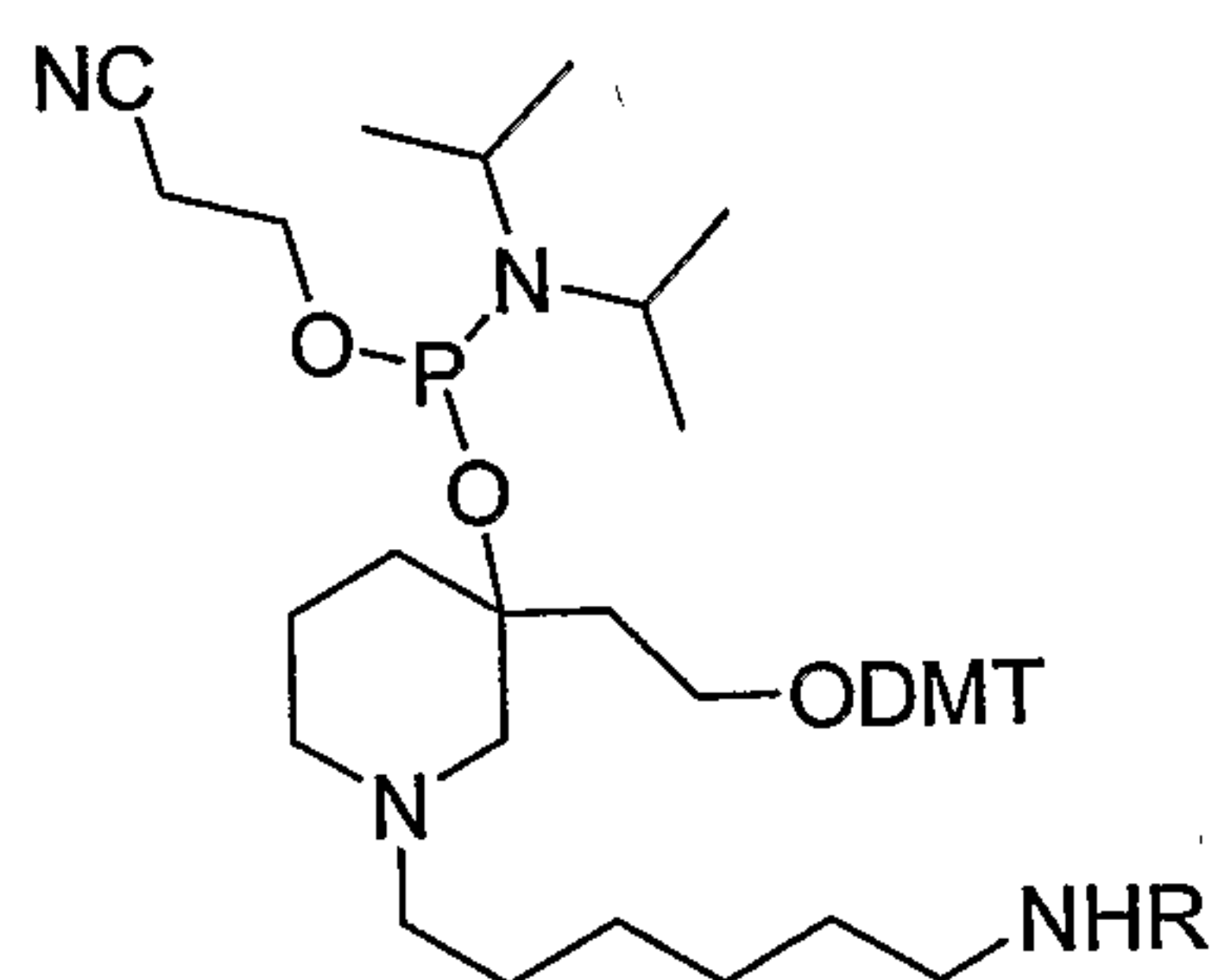
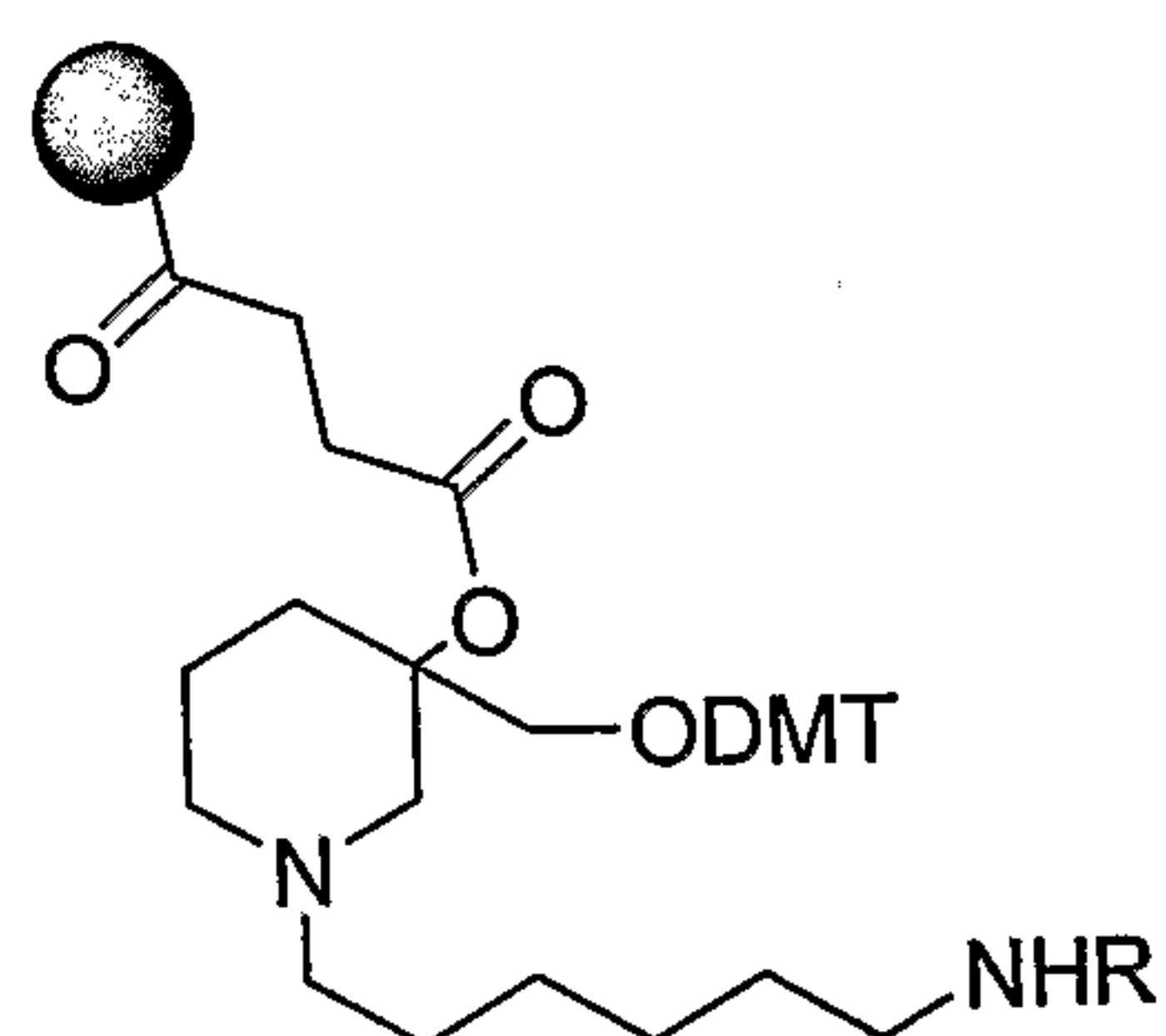
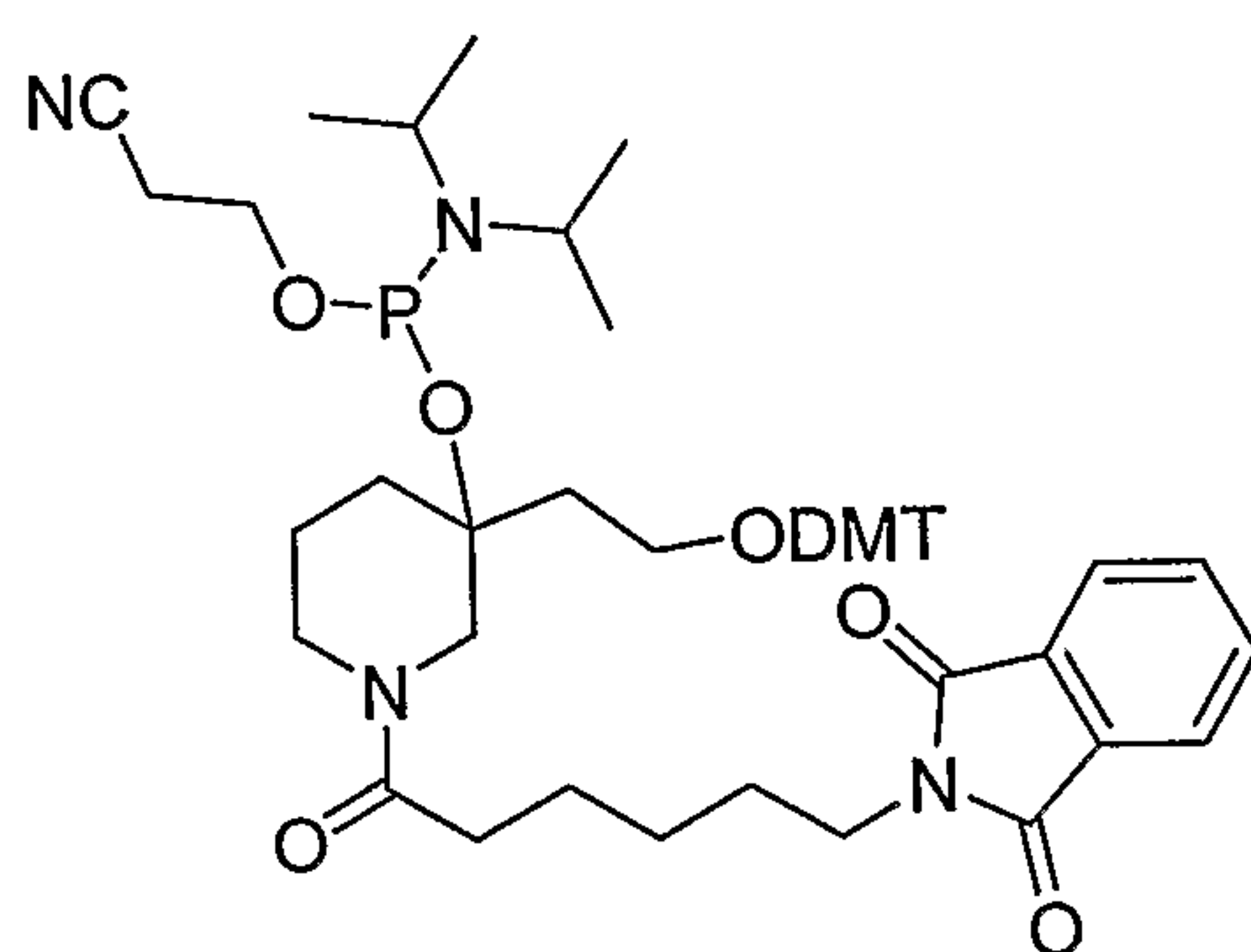
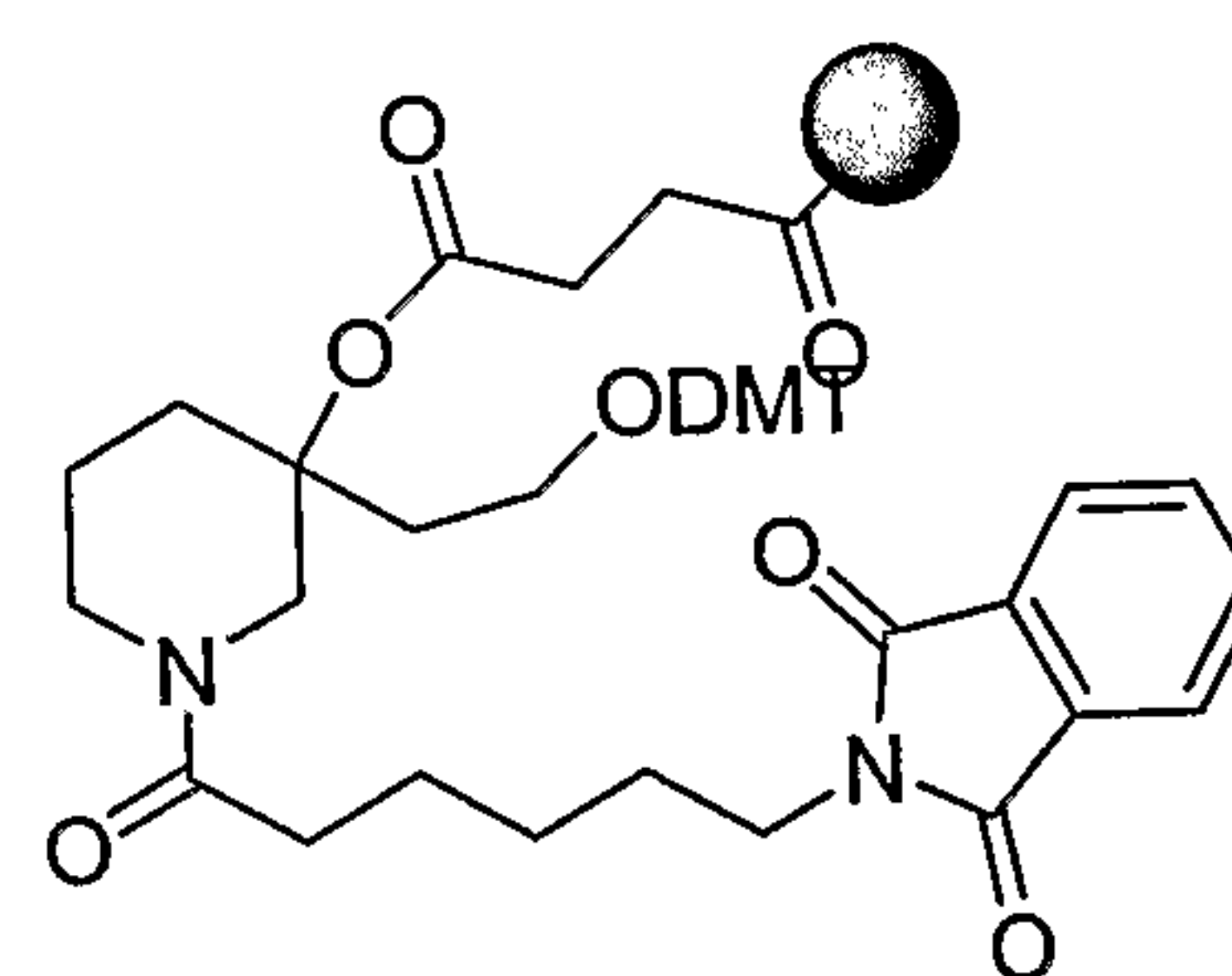
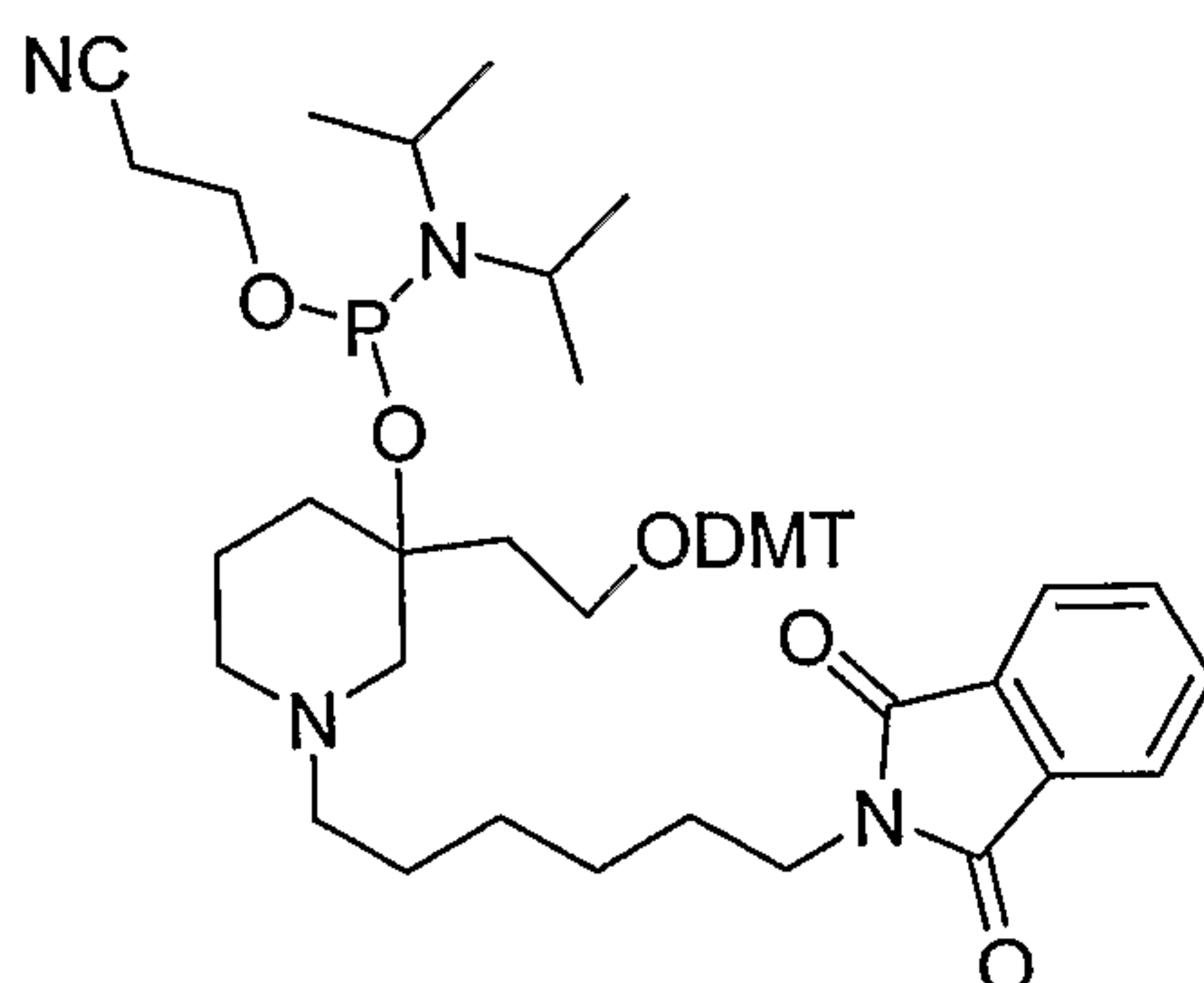
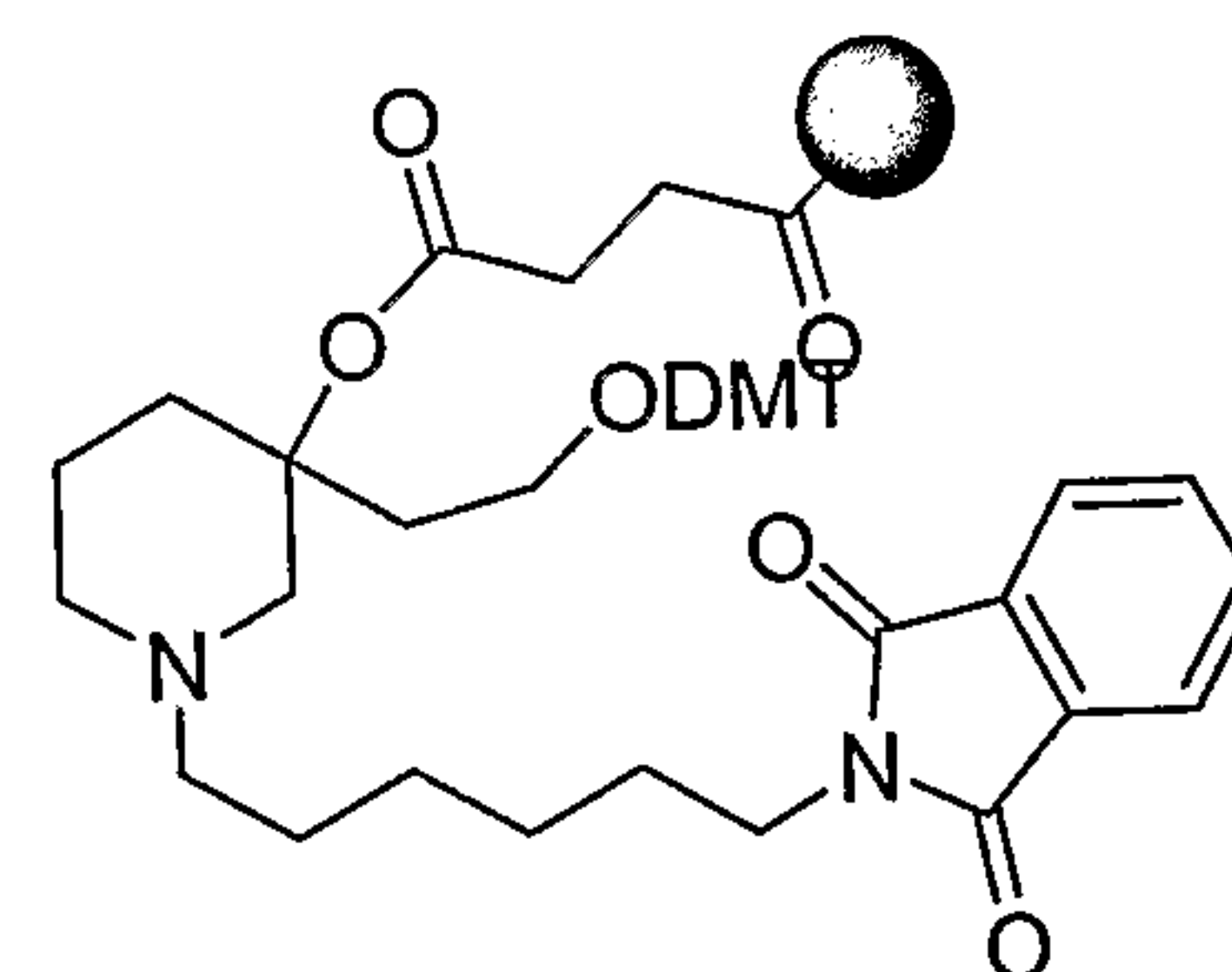


Similar reaction can be carried out with 2-piperidone and 3-piperidone

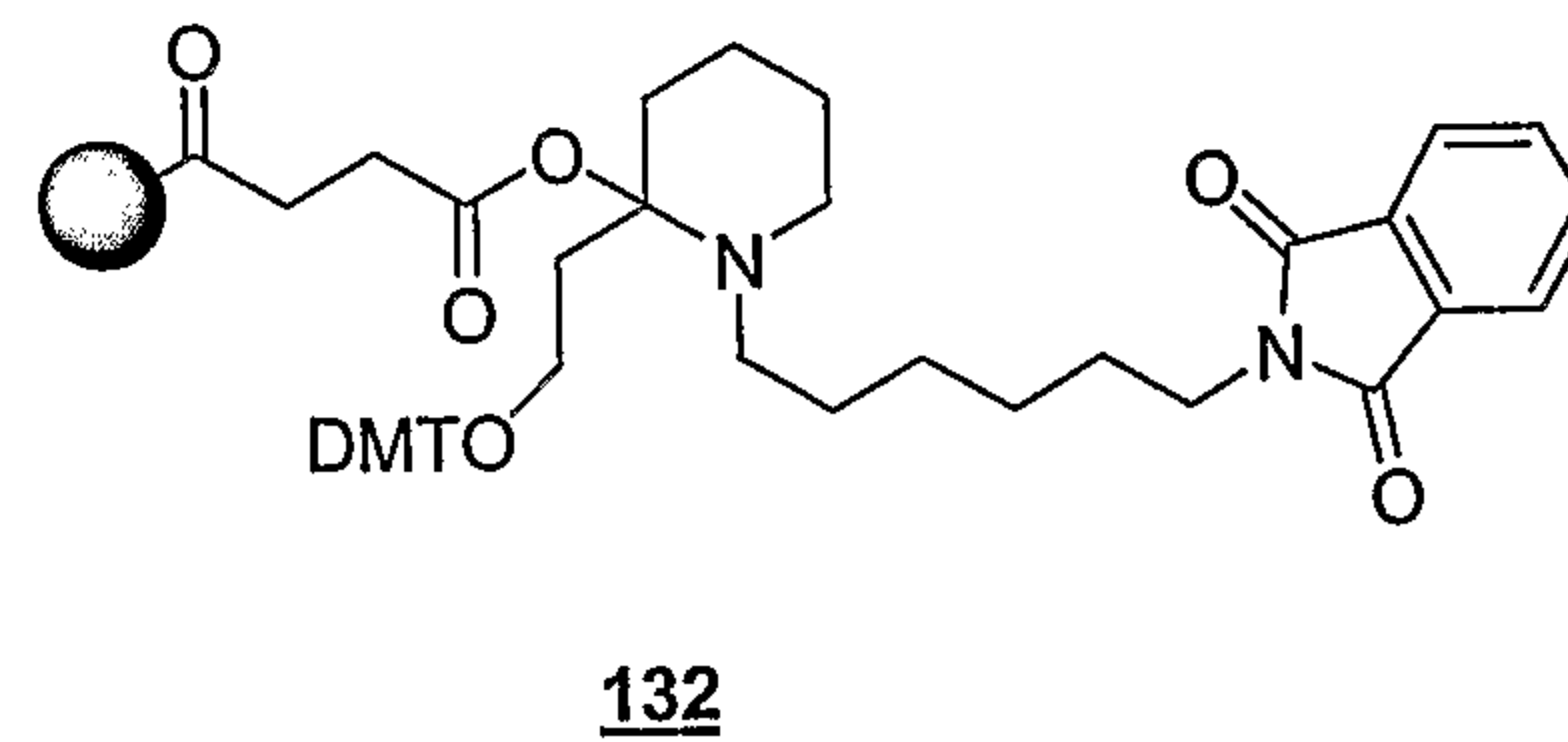
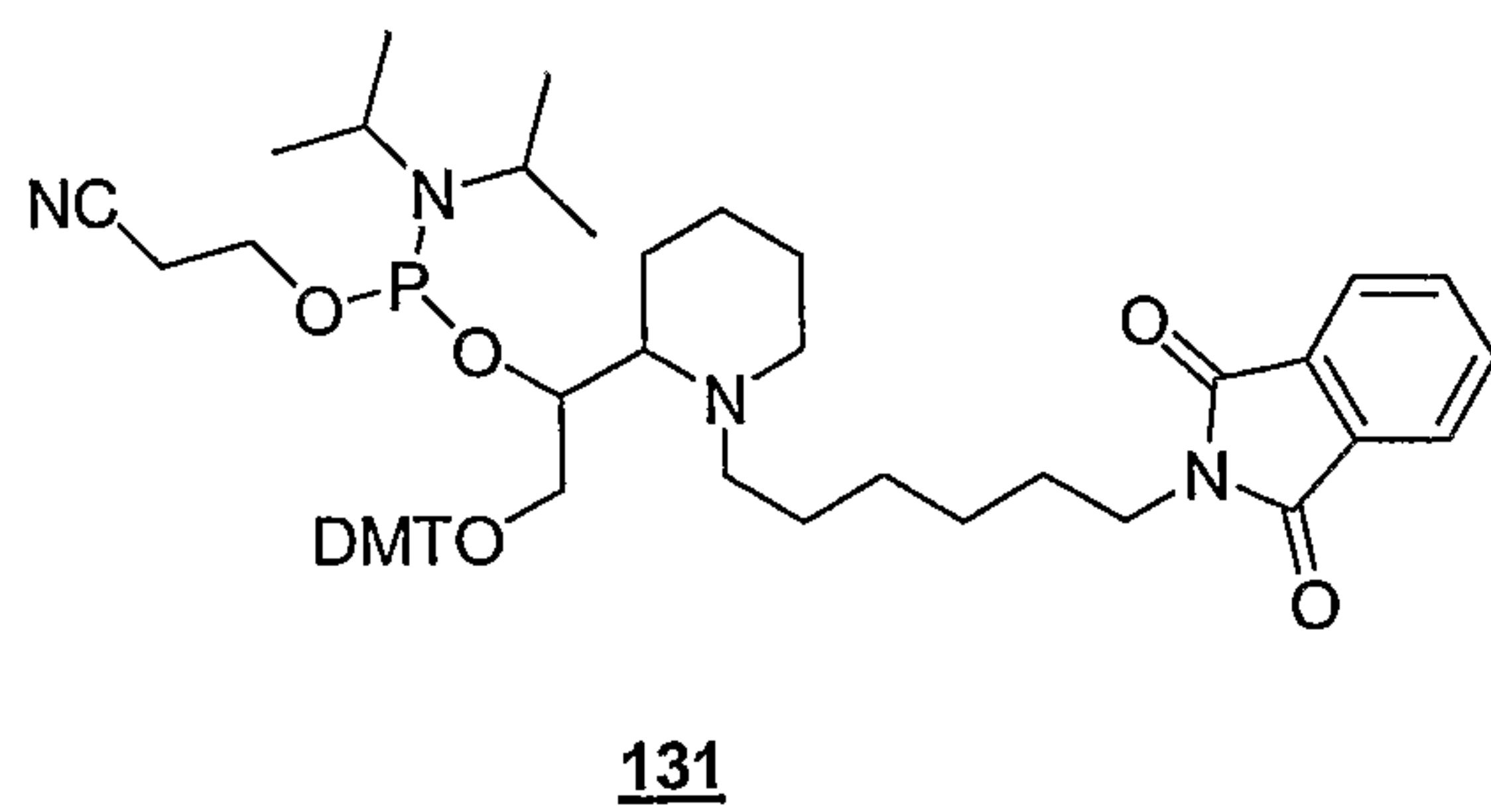
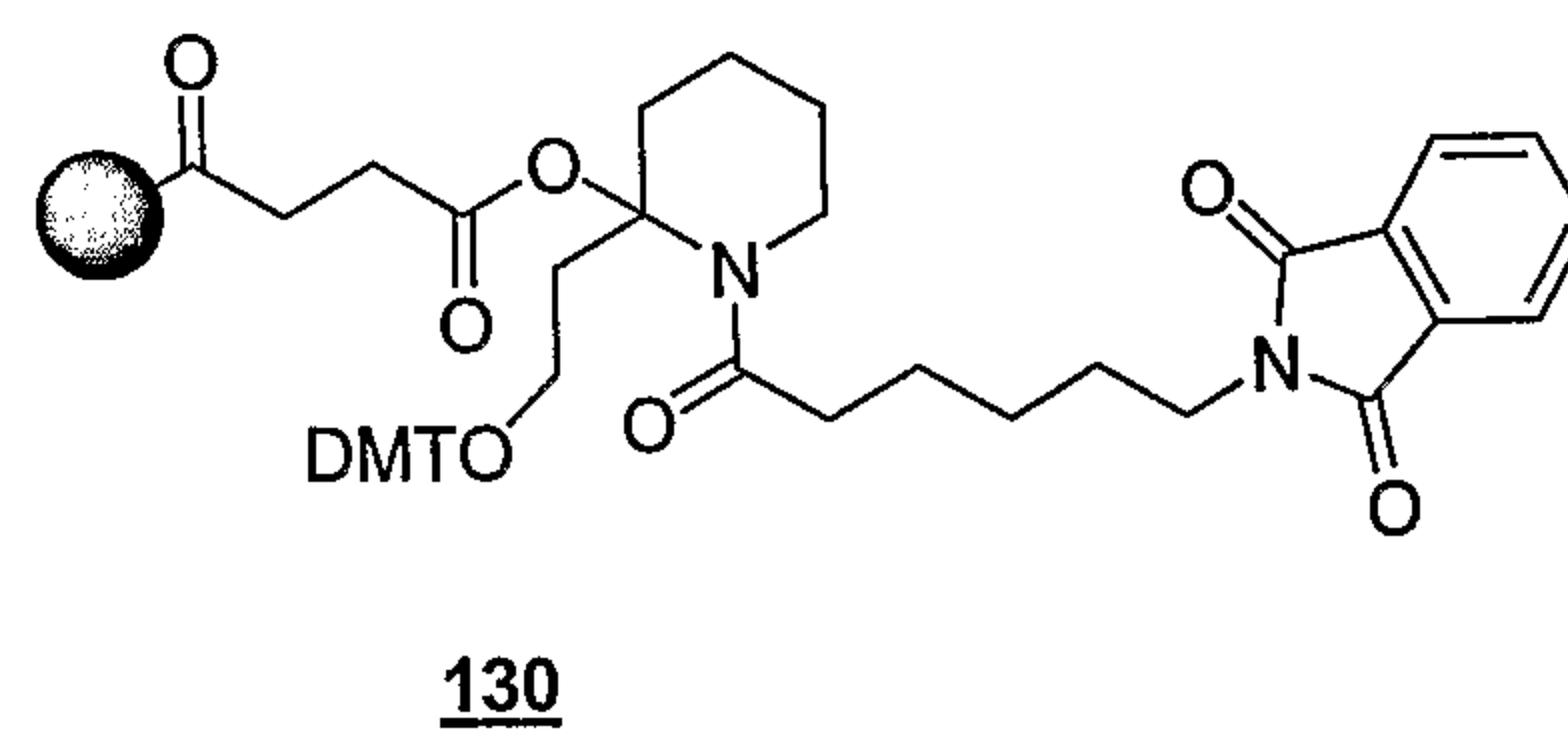
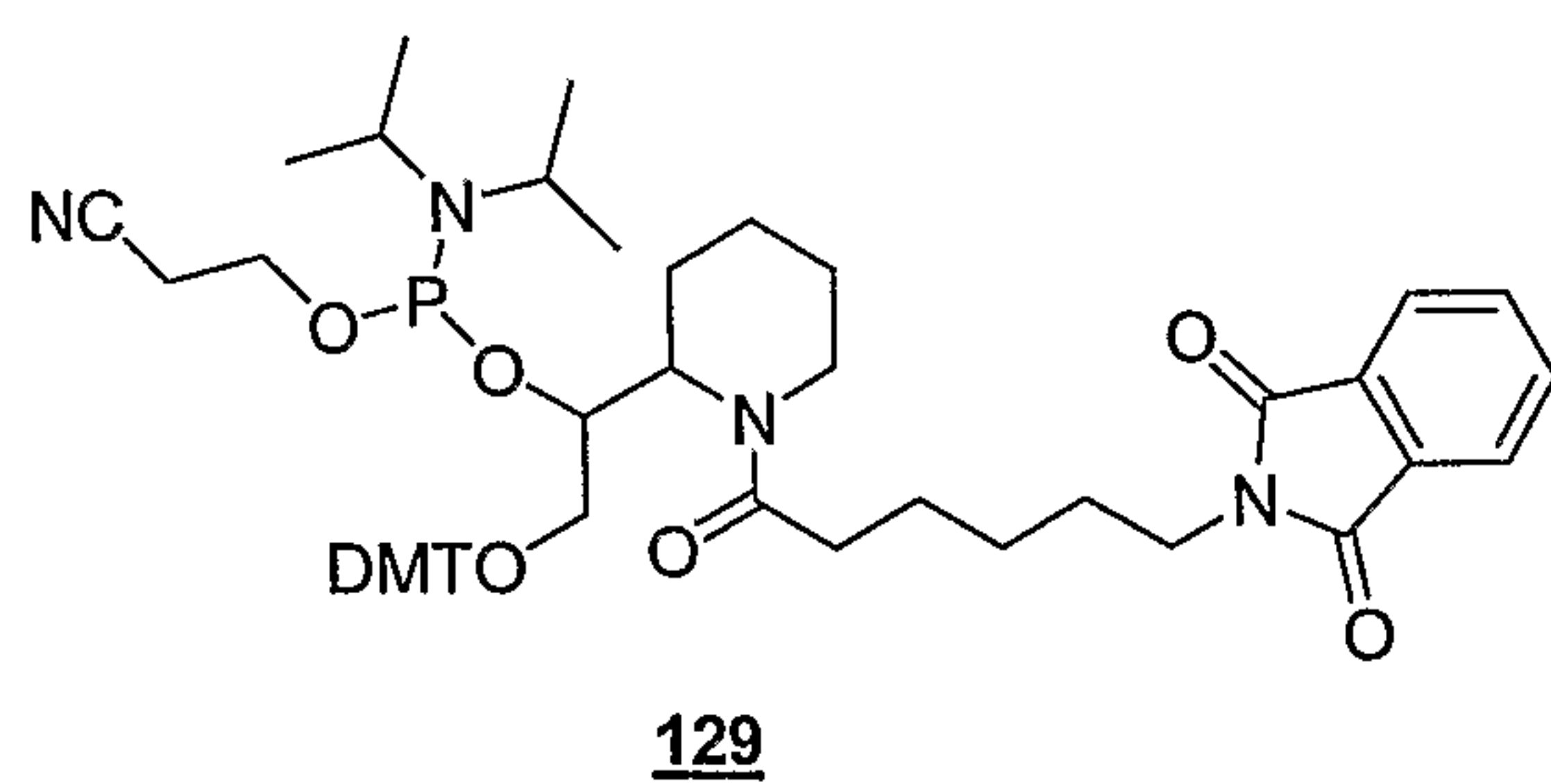
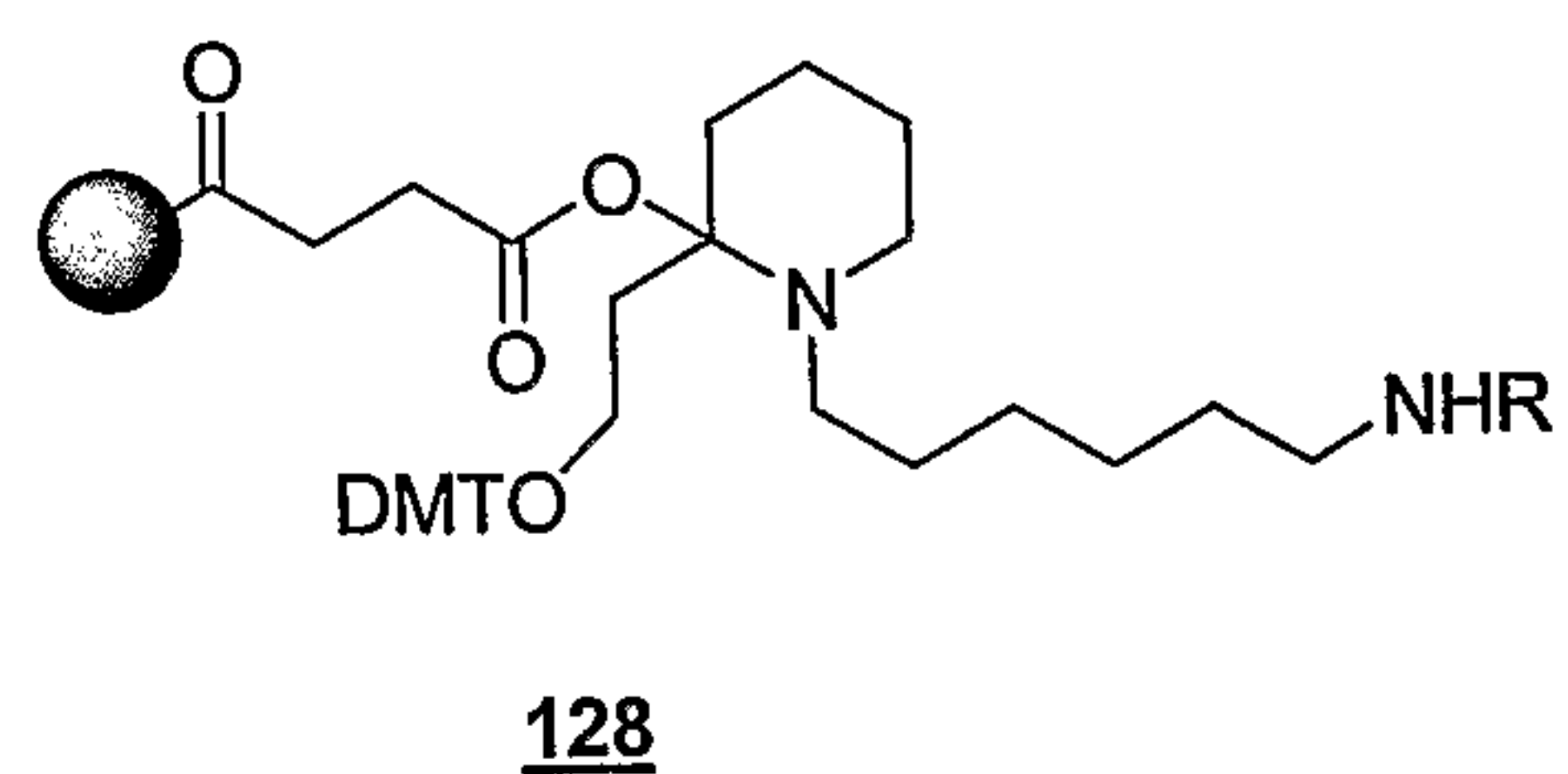
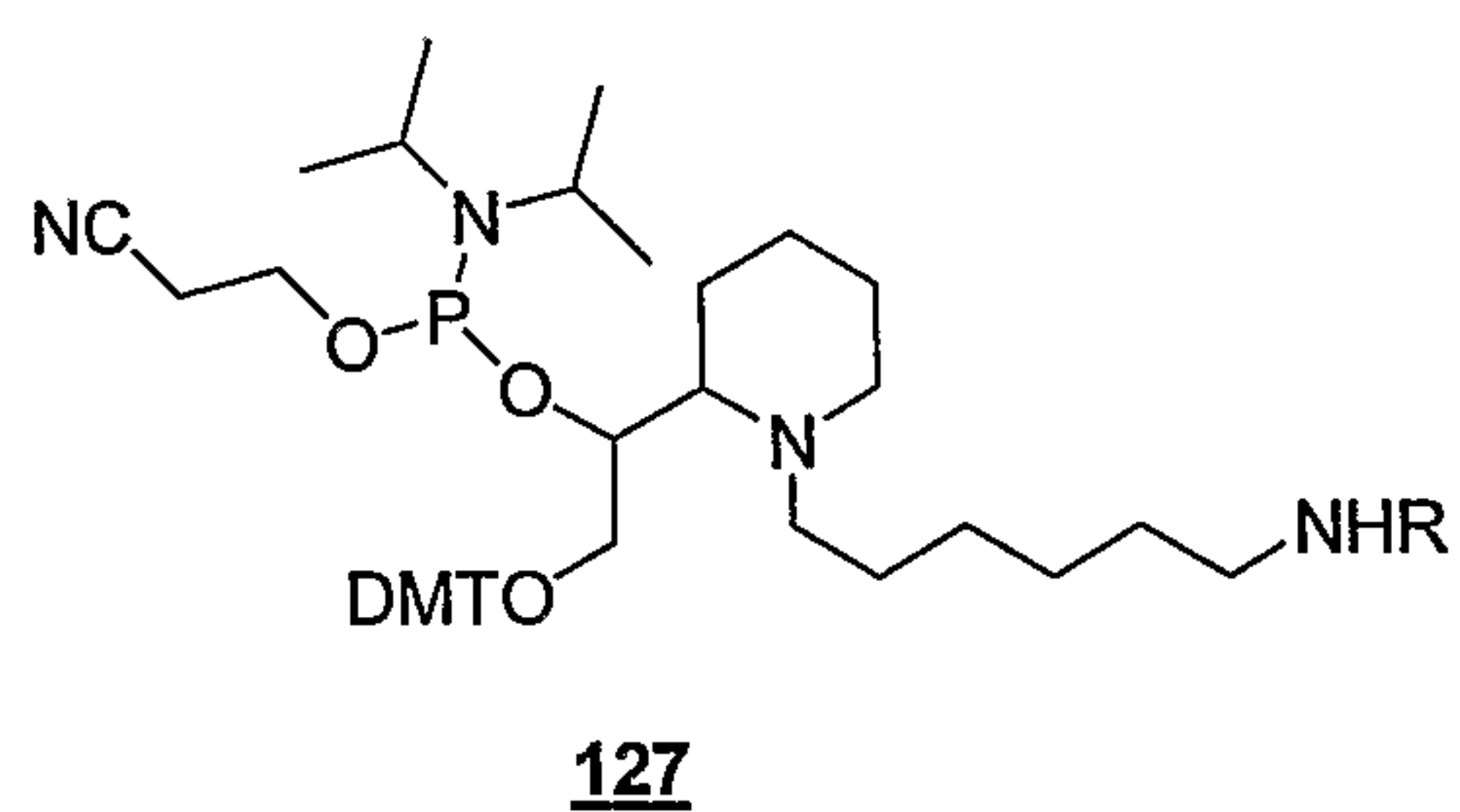
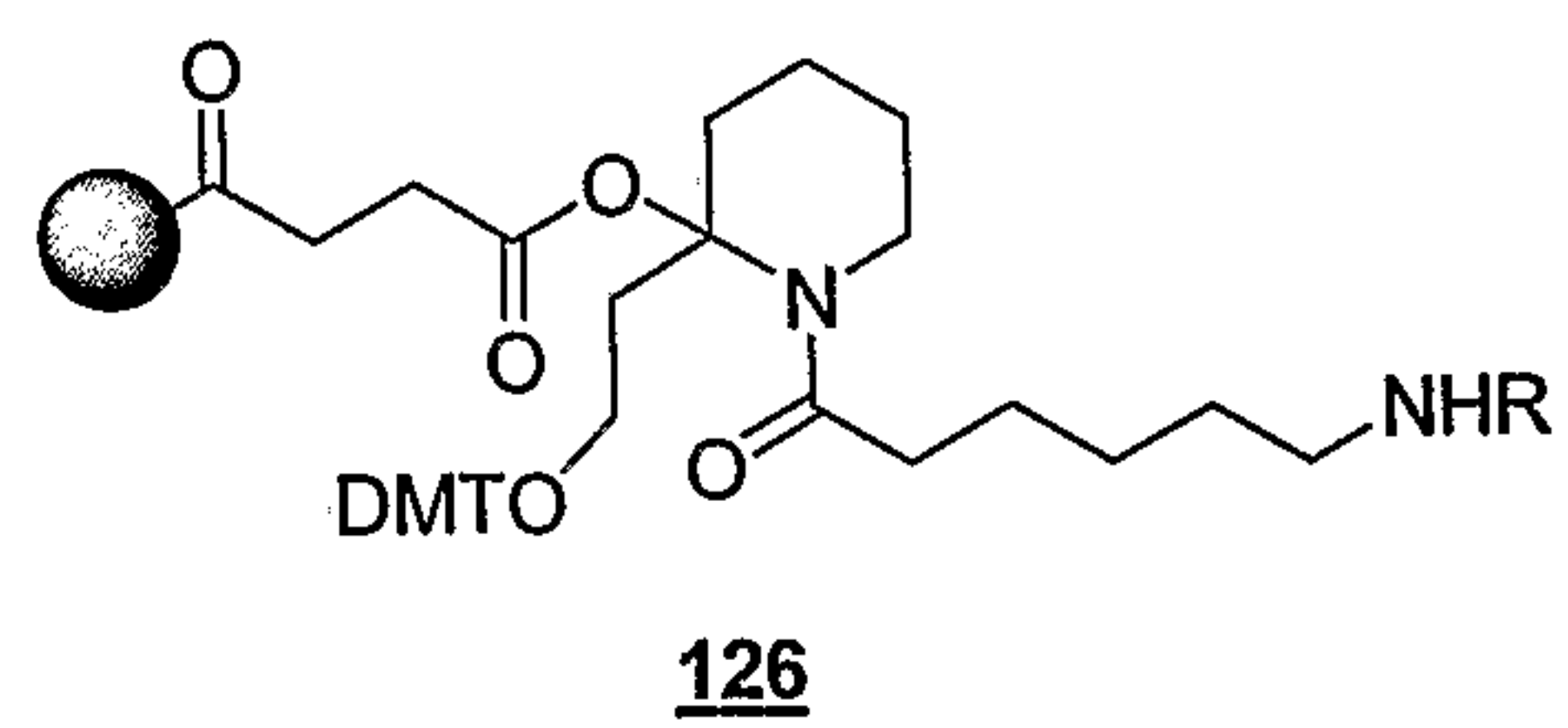
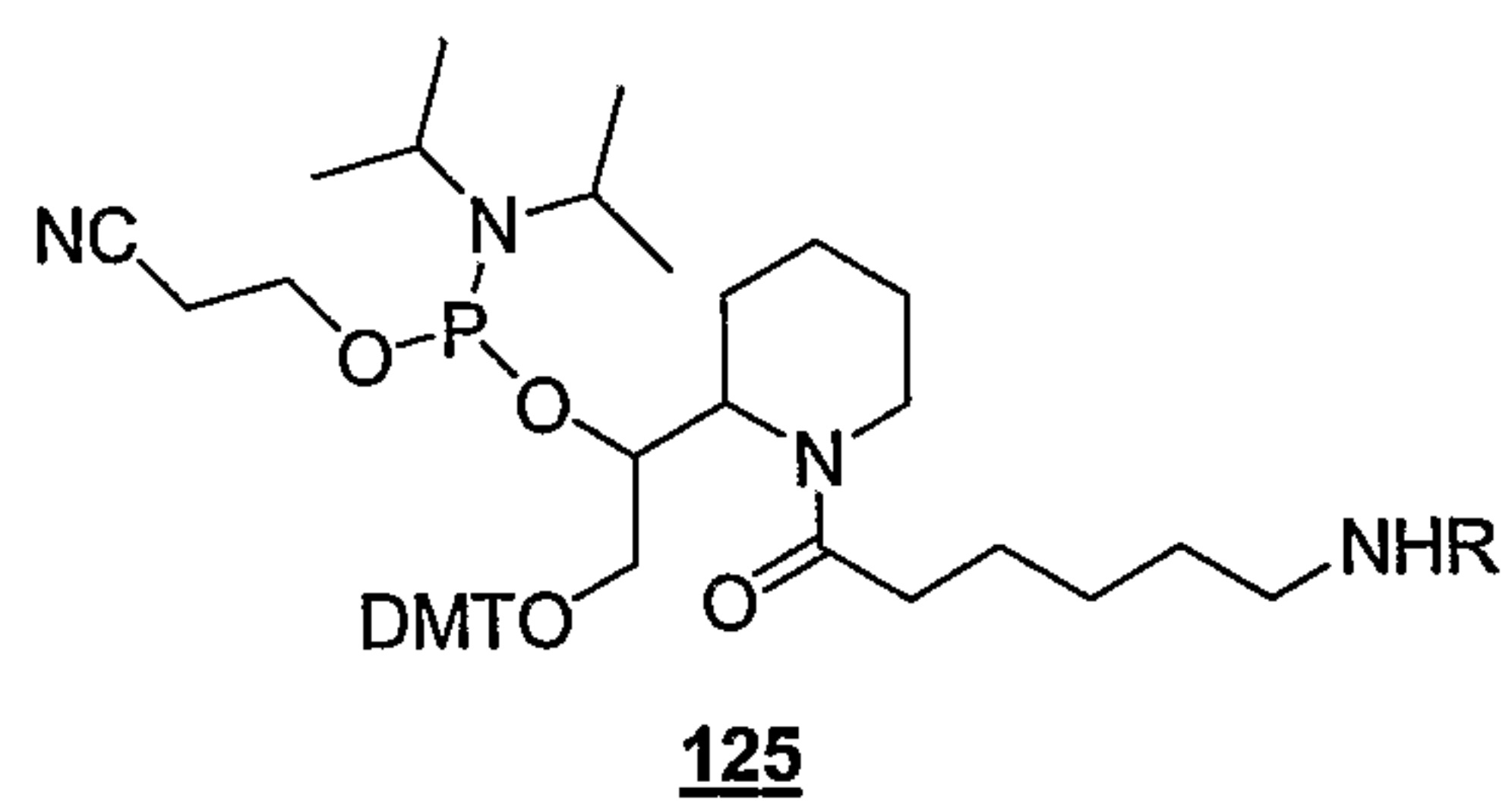
Linkers from 4-piperidone



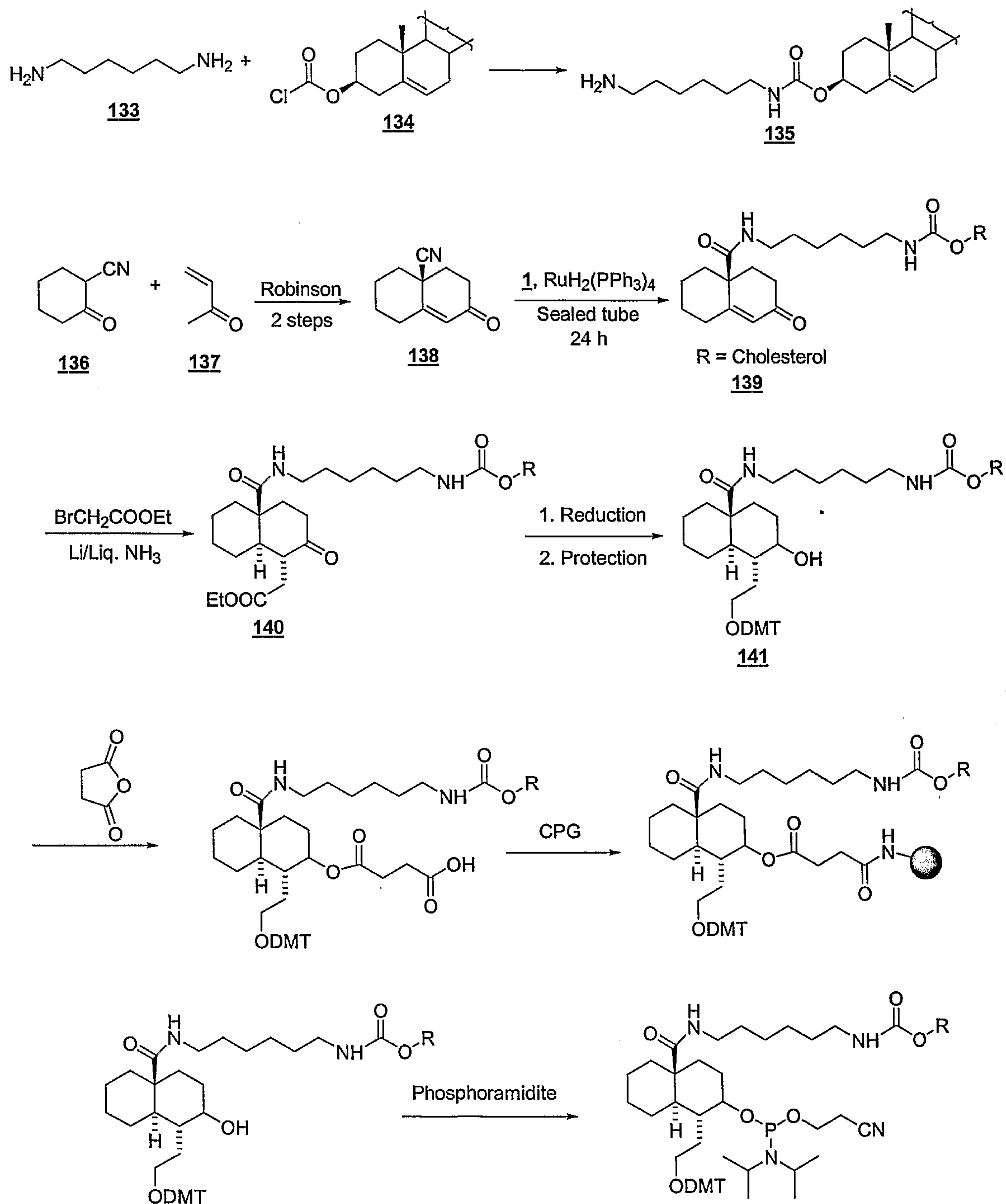
Linkers from 3-piperidone

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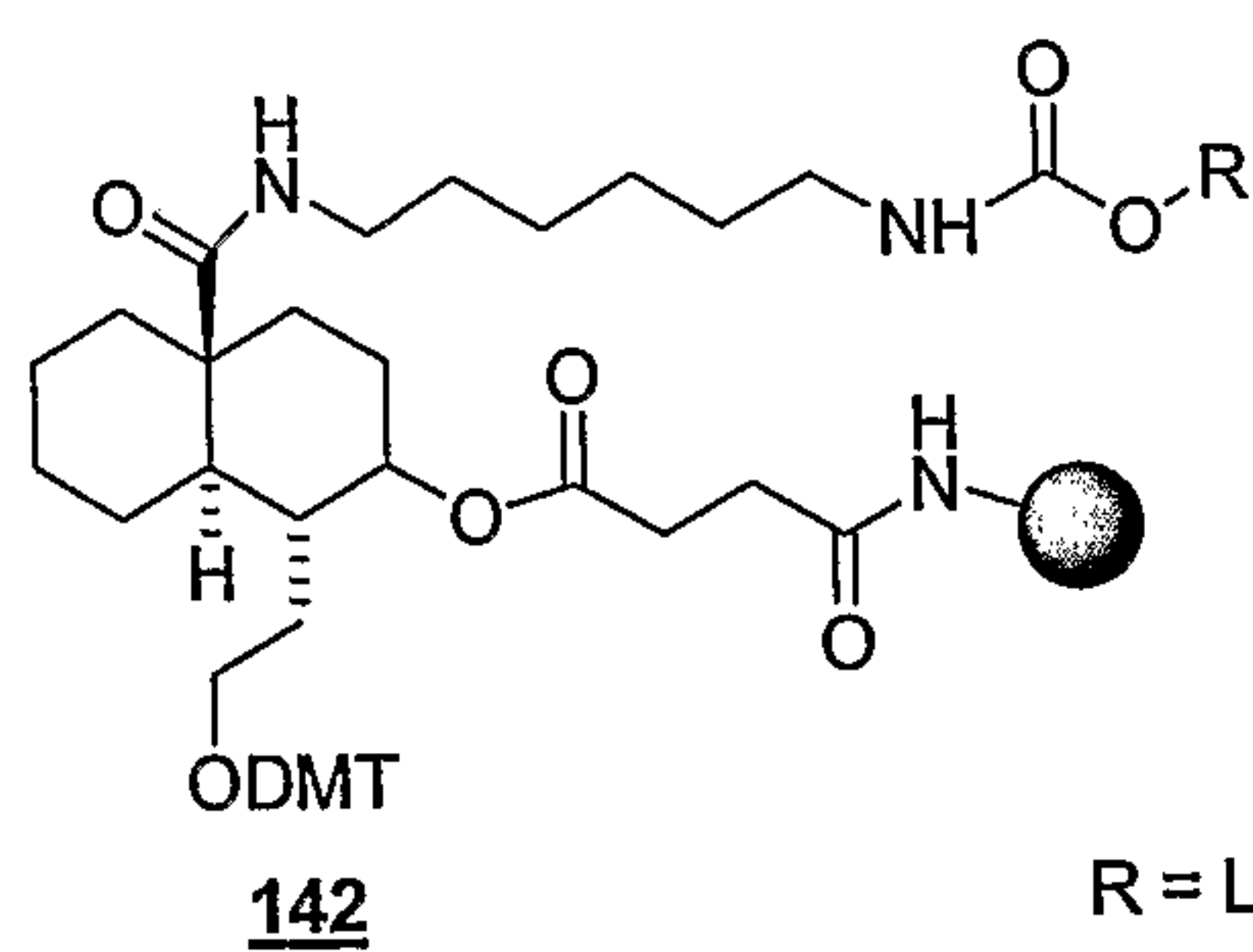
Linkers from 2-piperidone



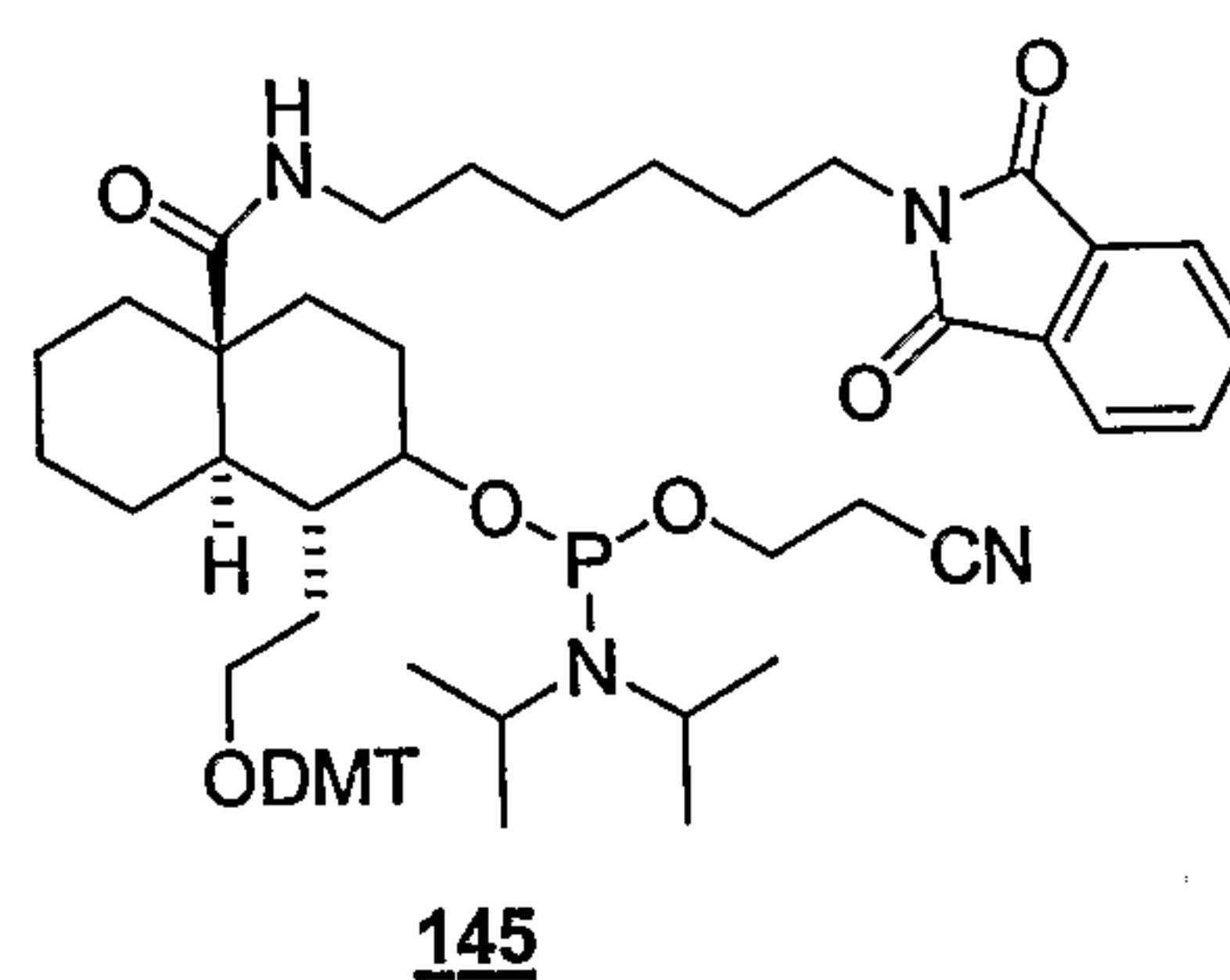
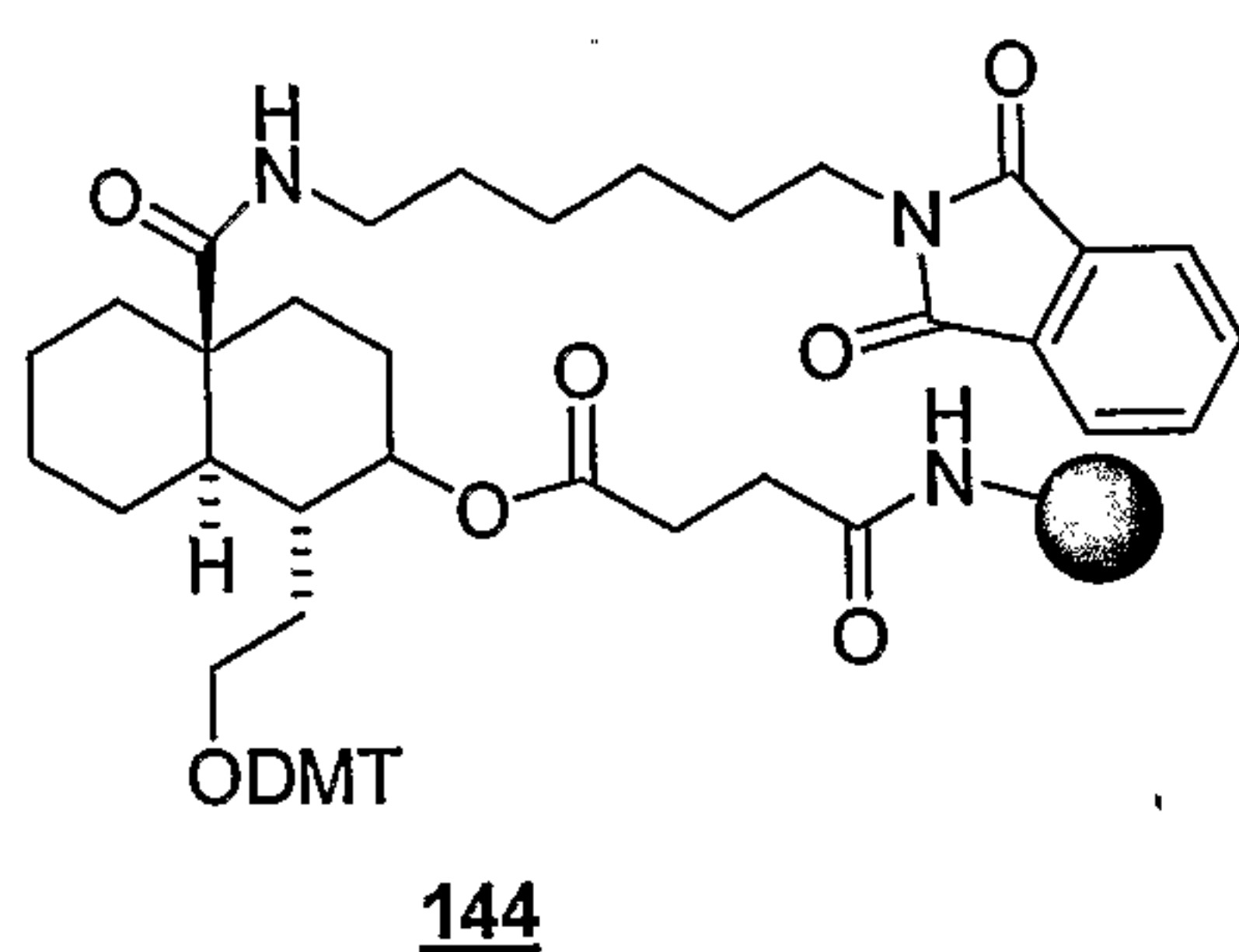
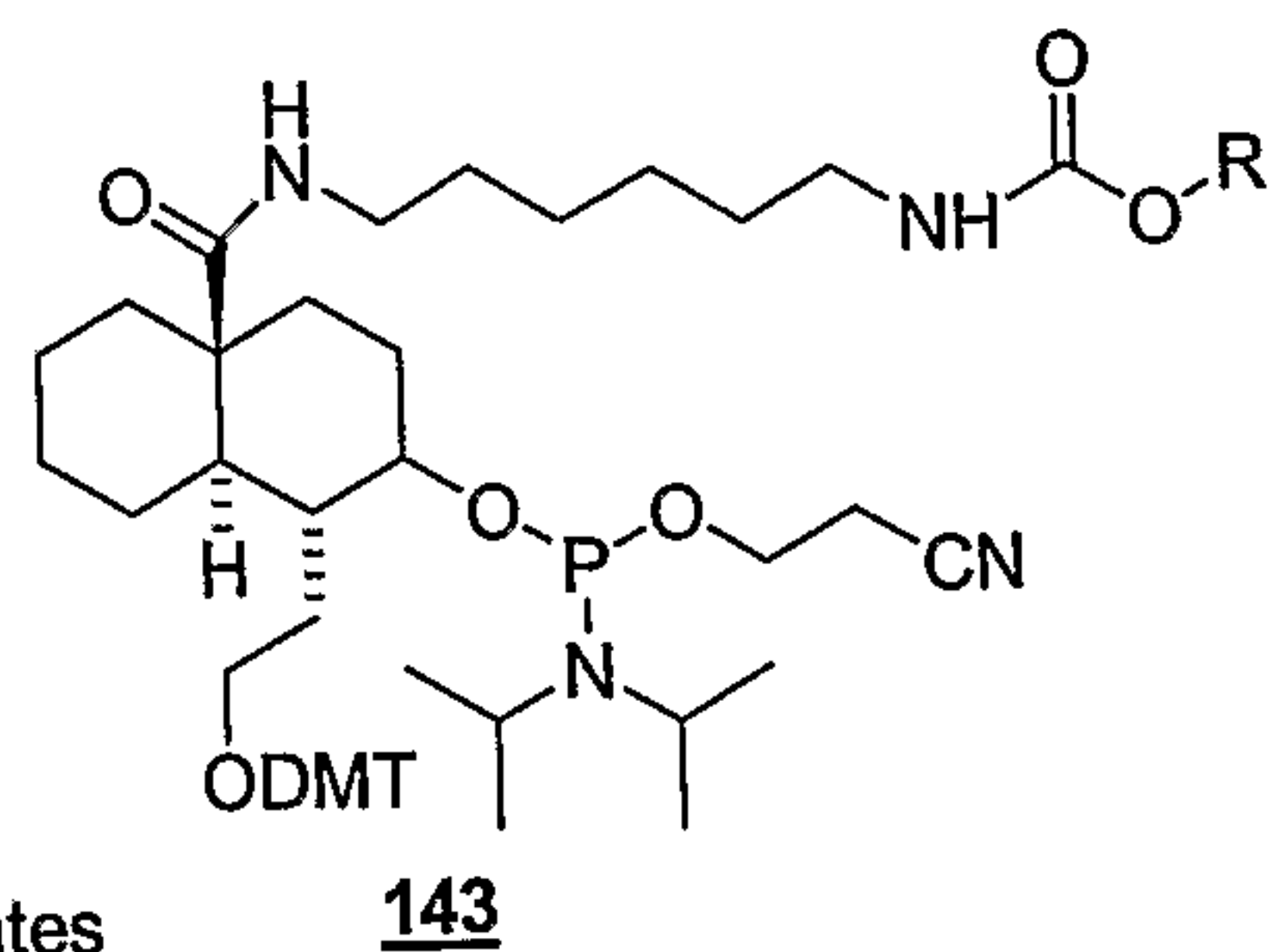
Conjugation through decalin system



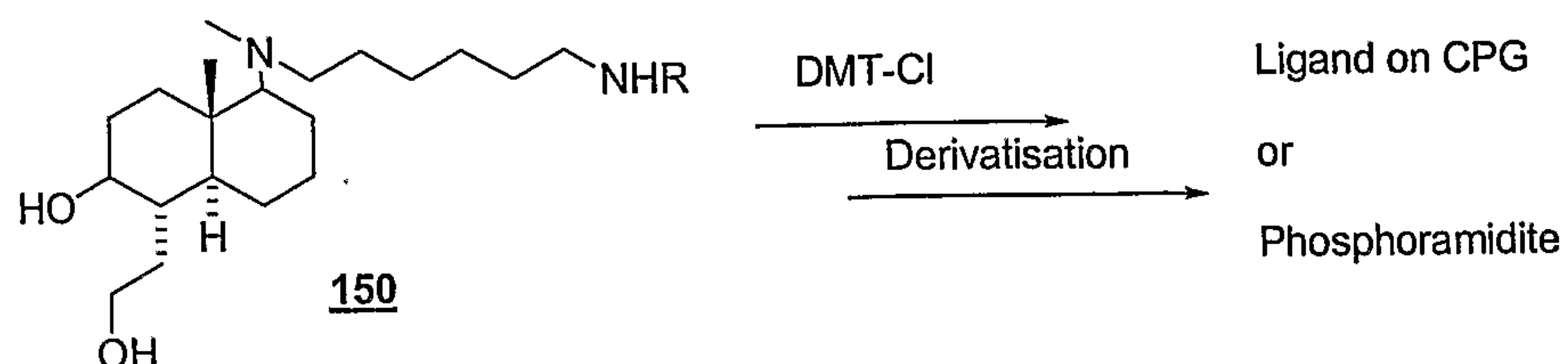
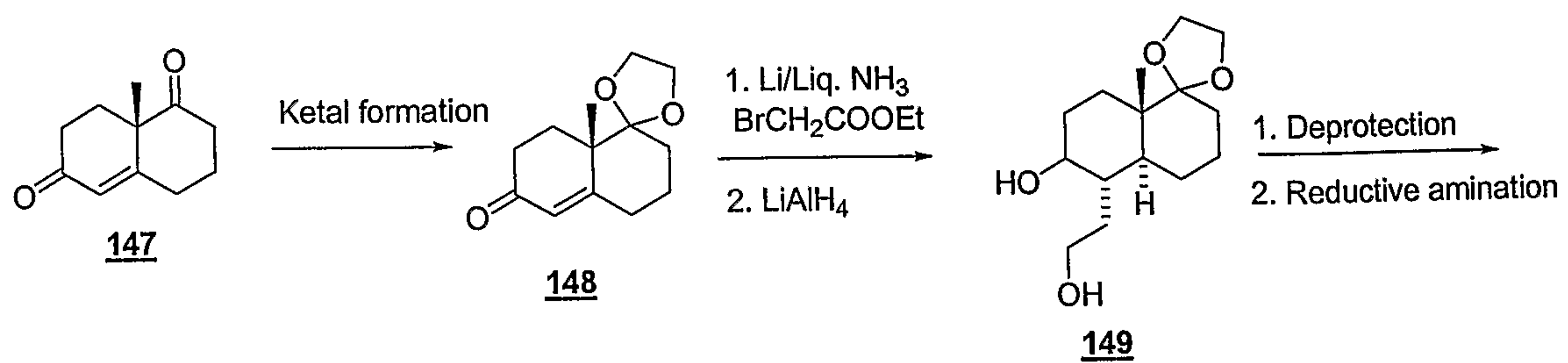
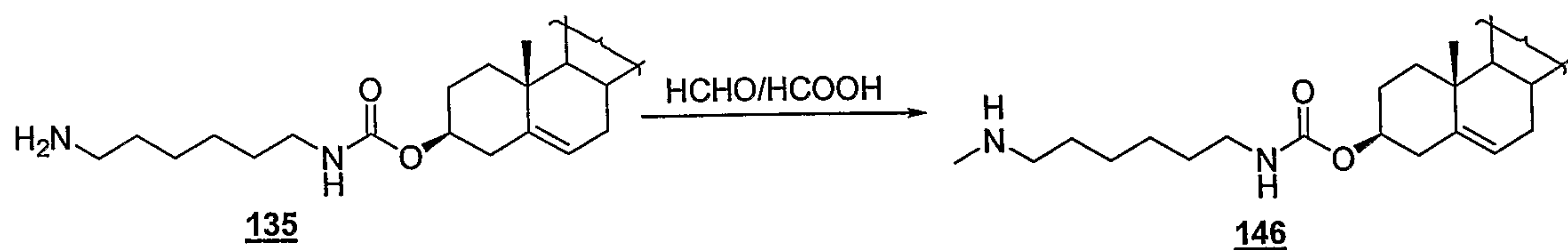
Conjugates from decalin system:

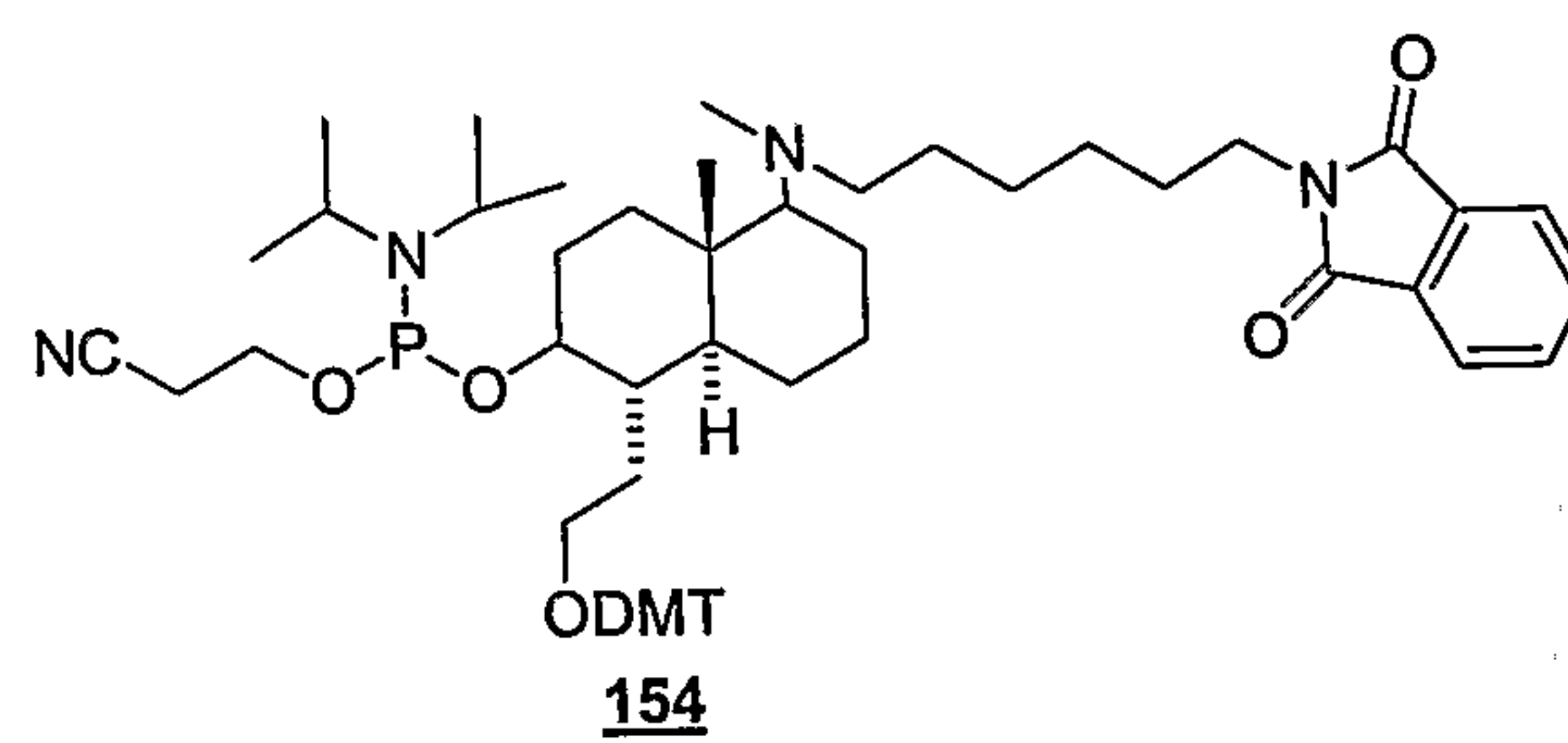


R = Lipophilic conjugates

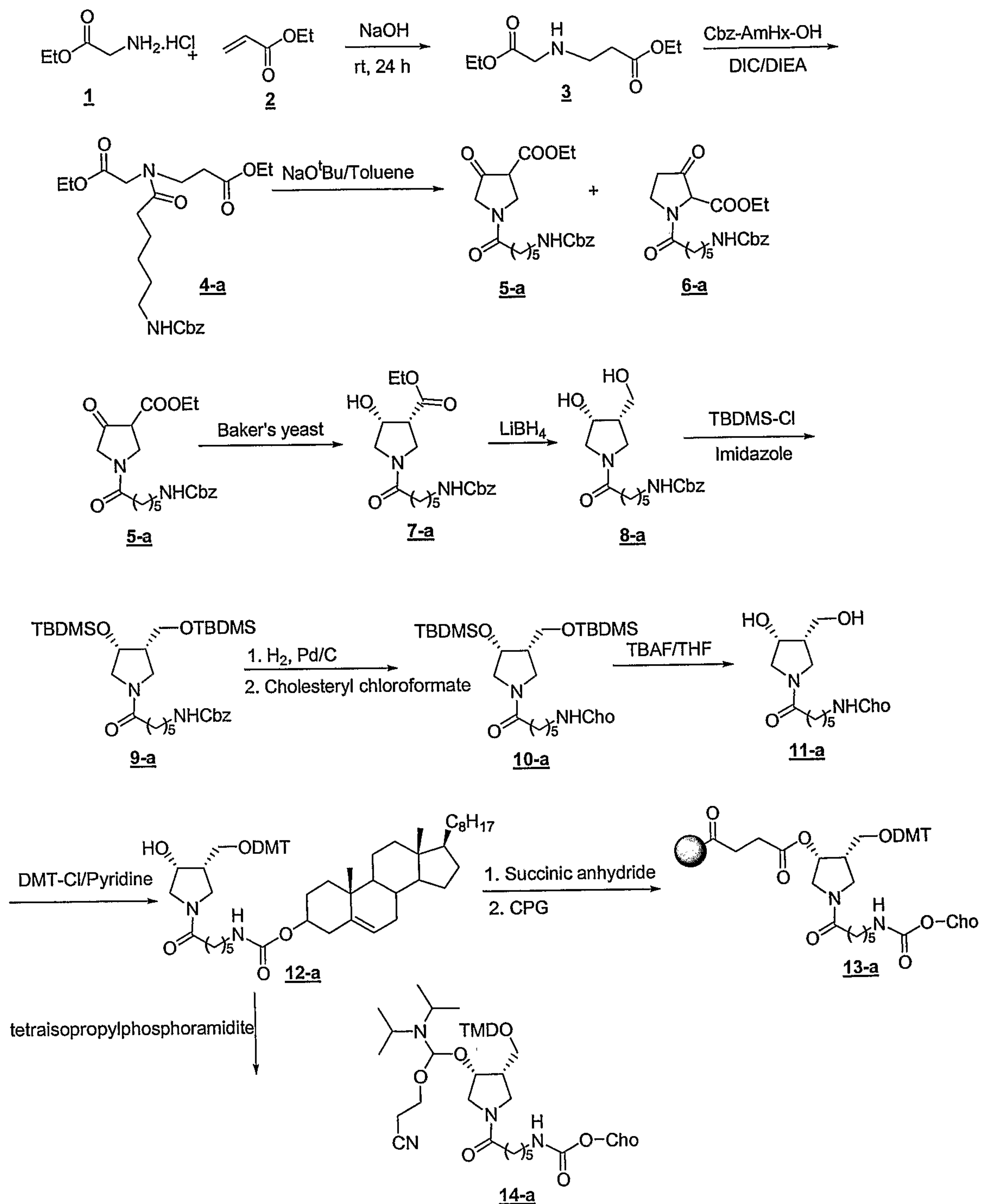


Decalin linker from Wieland-Miescher ketone

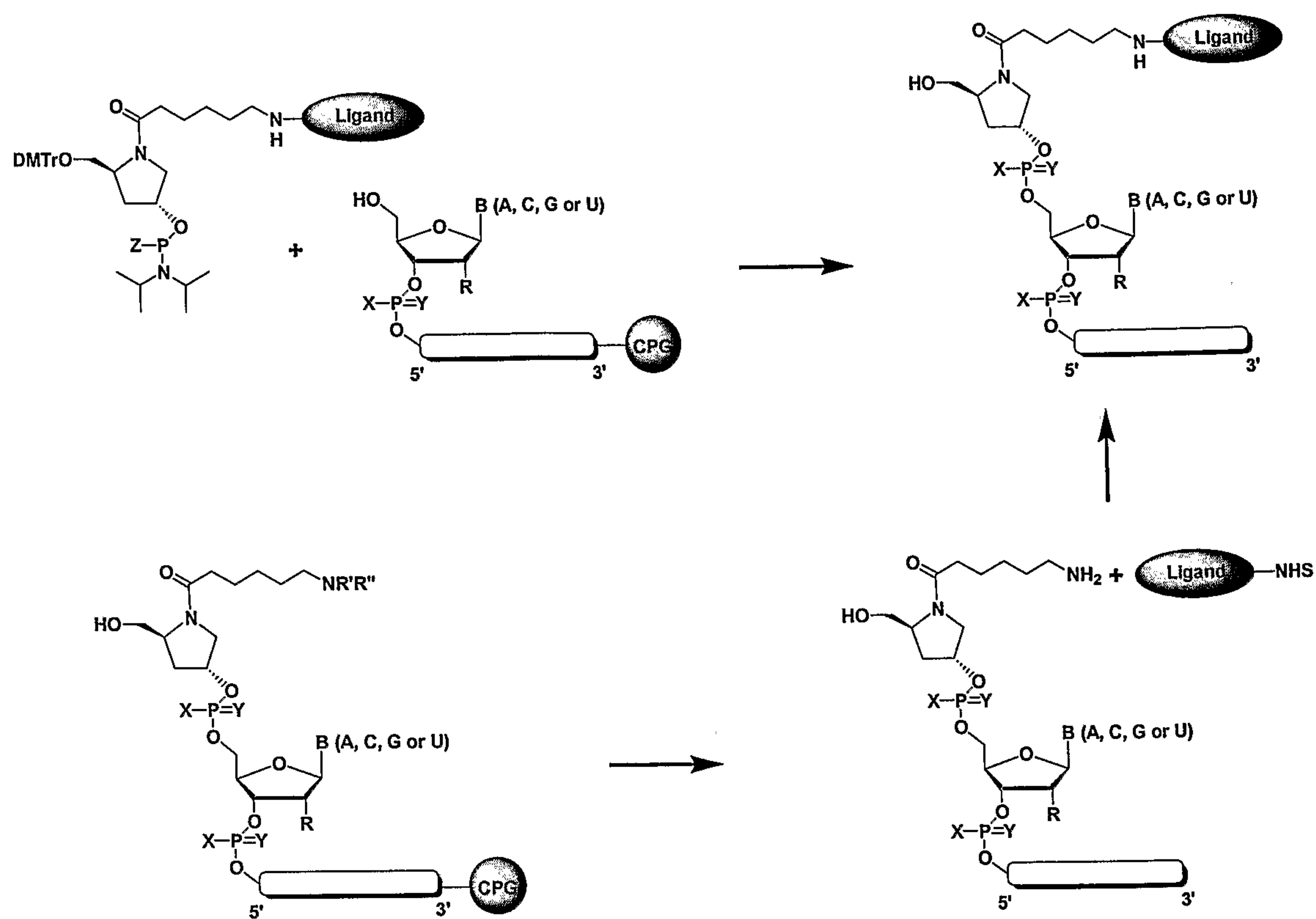




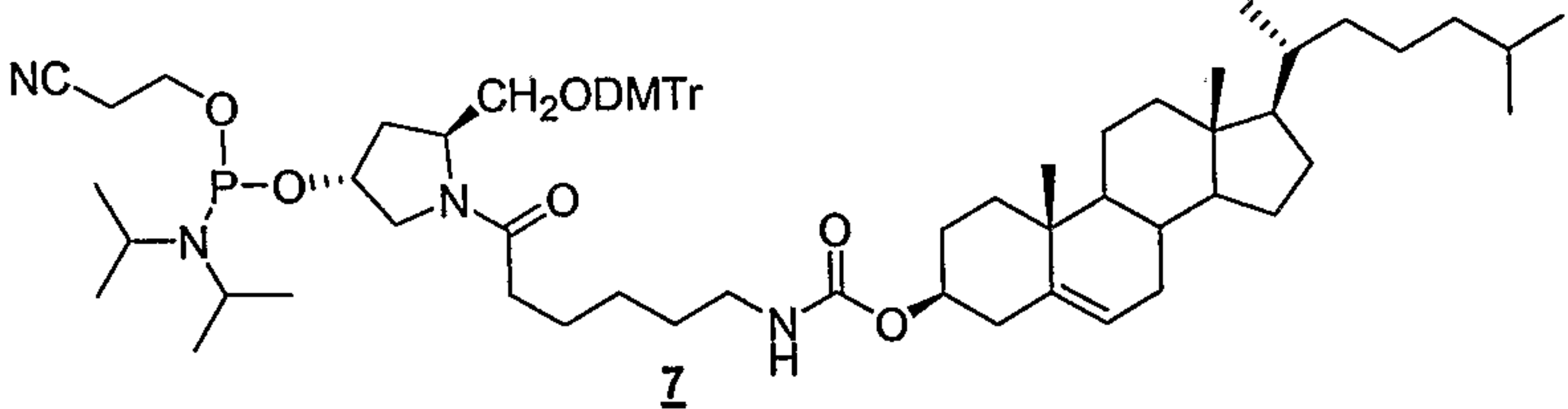
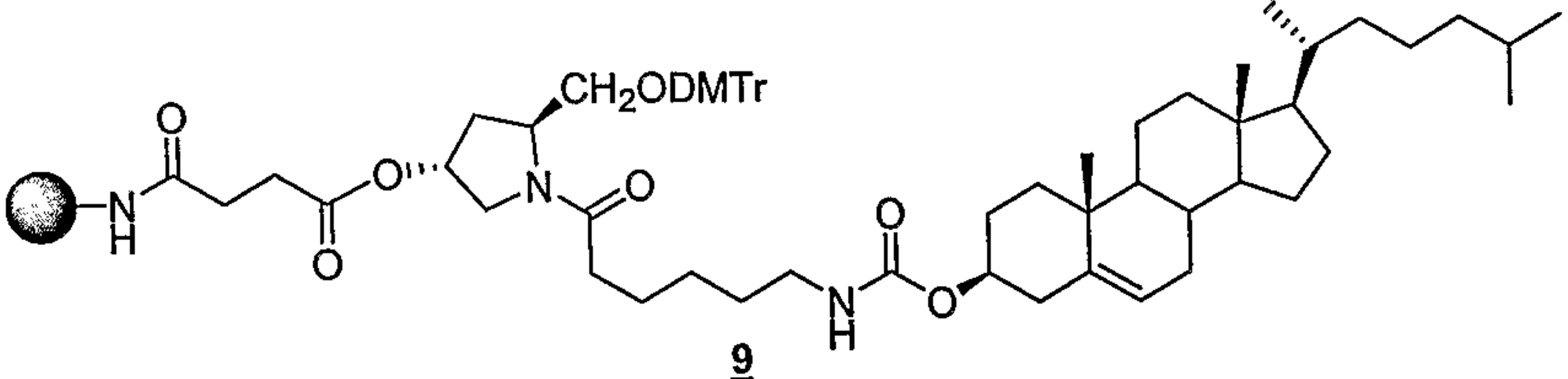
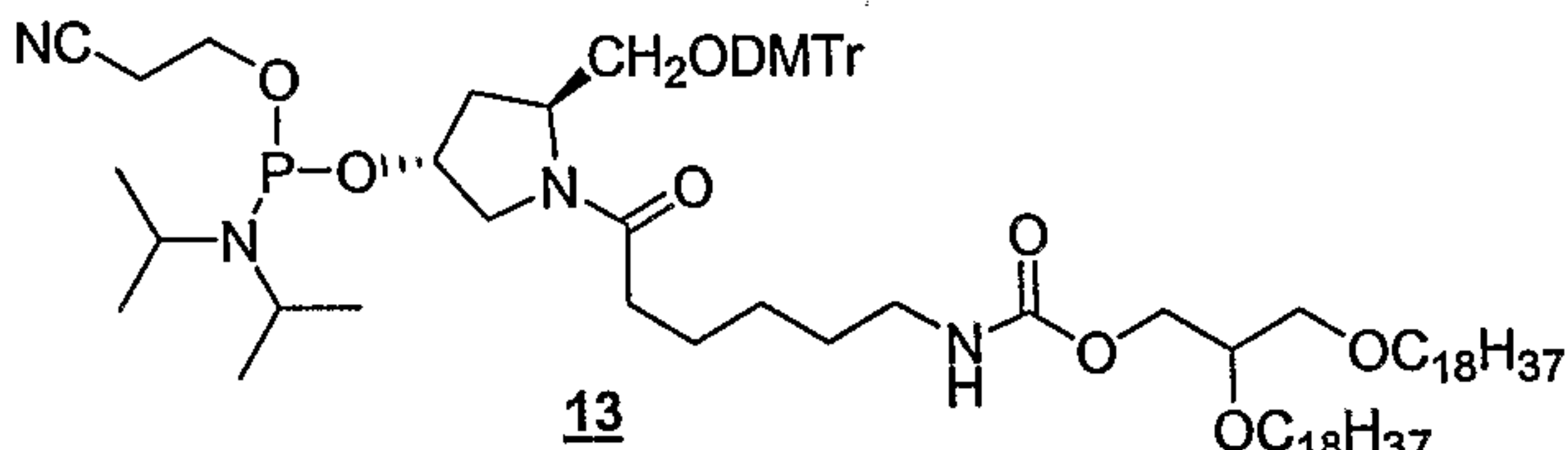
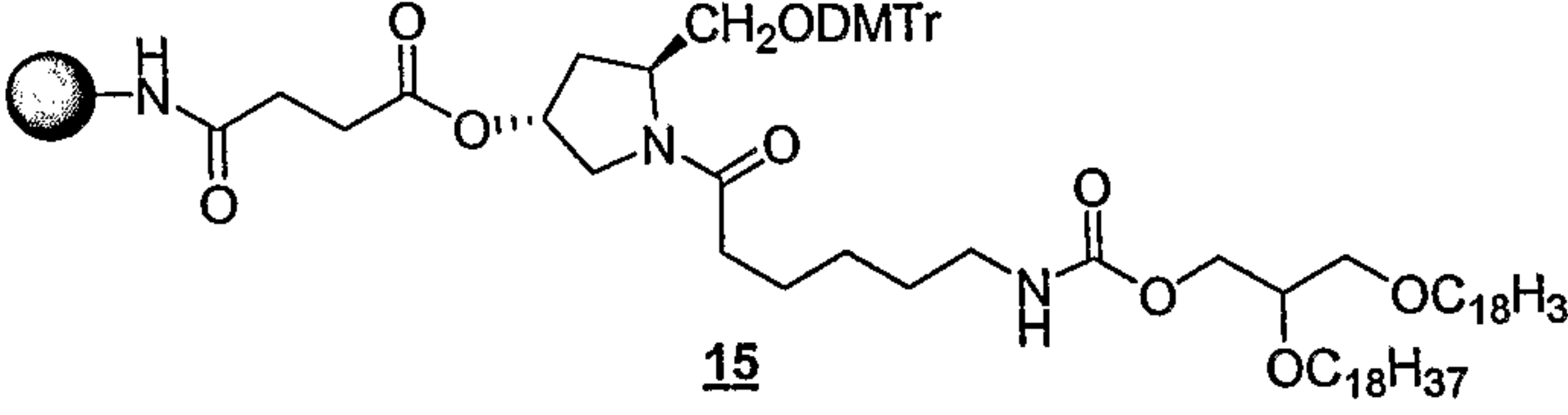
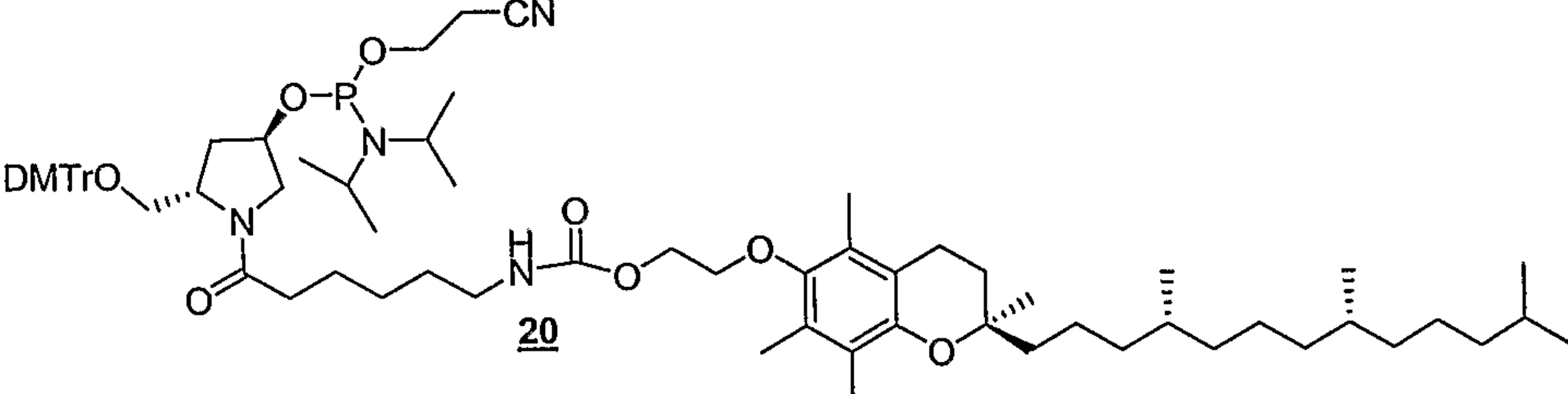
Synthesis of pyrroline linker:



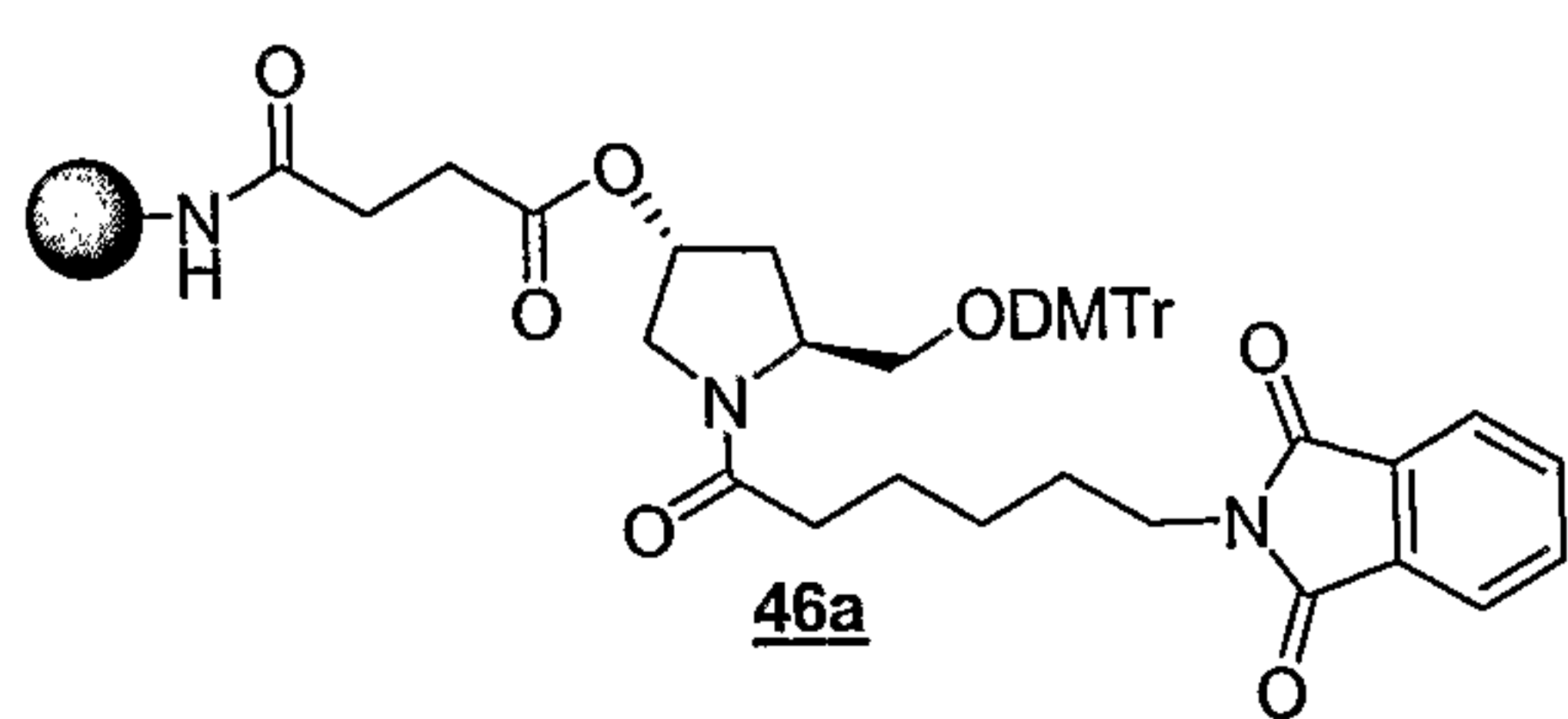
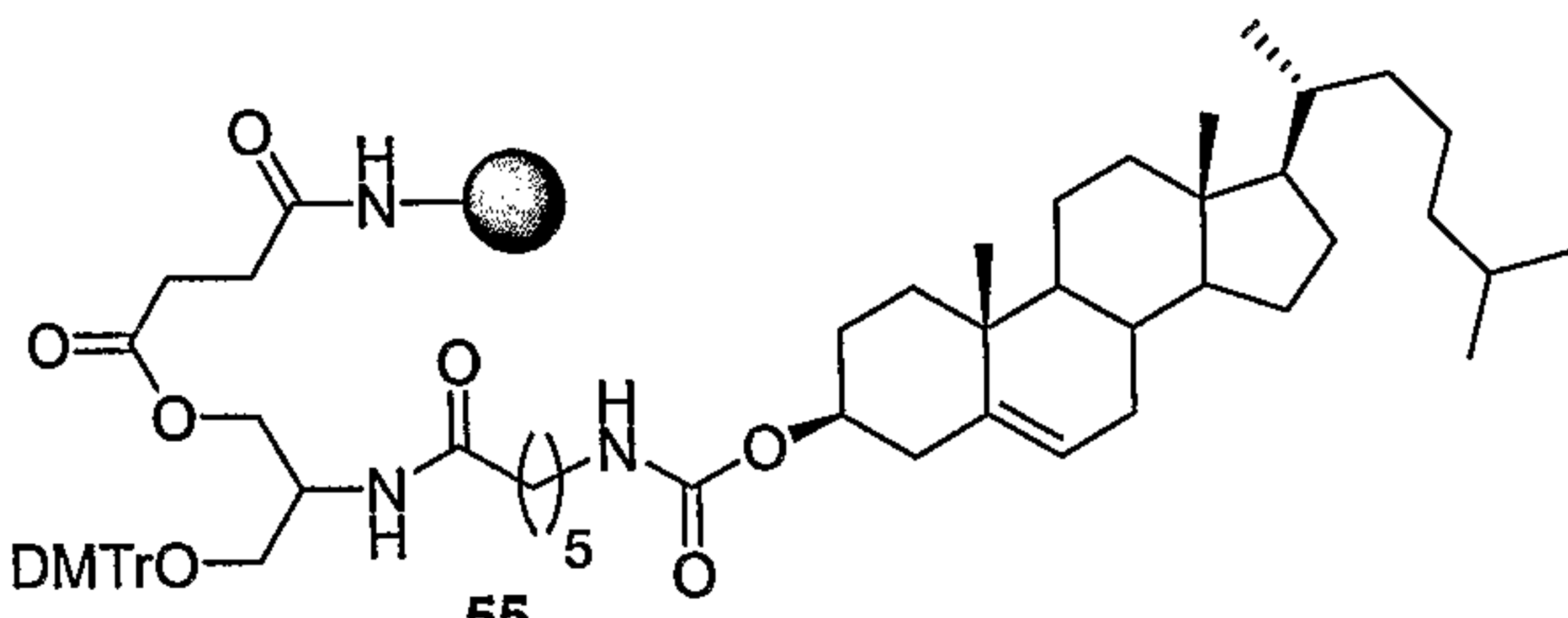
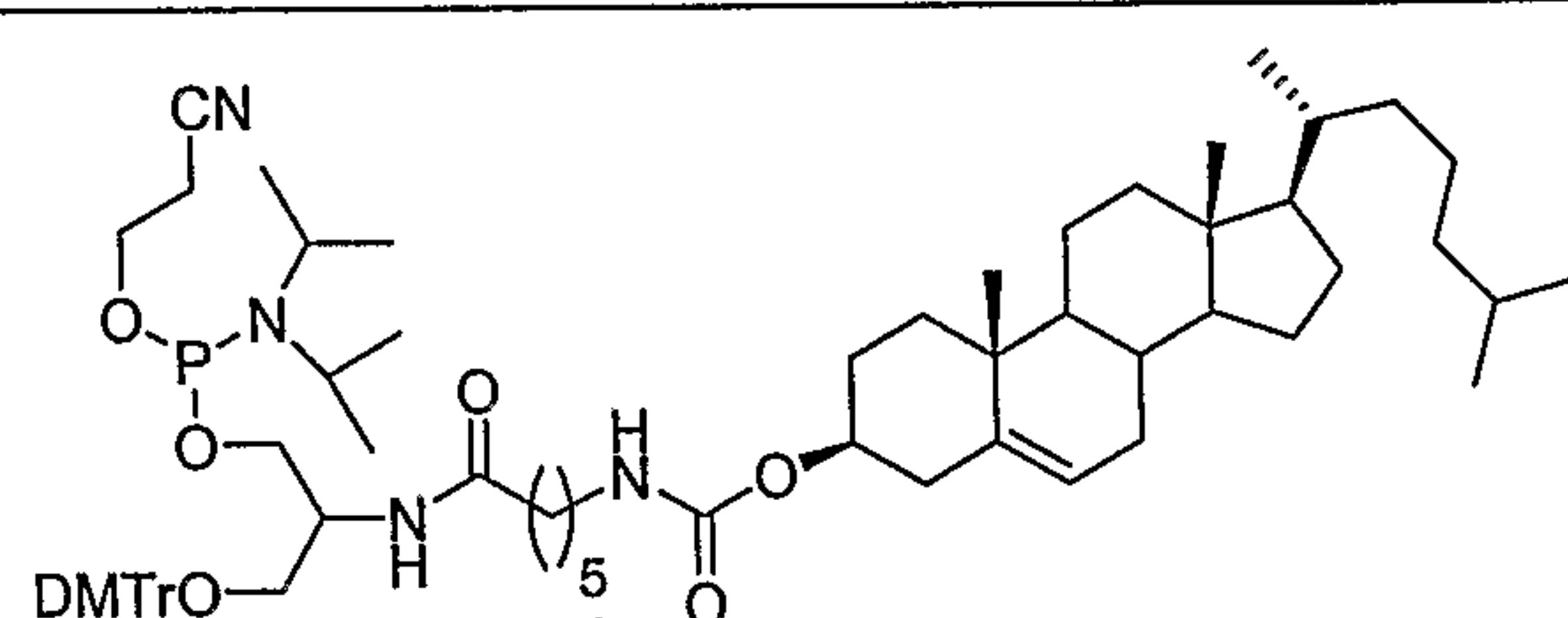
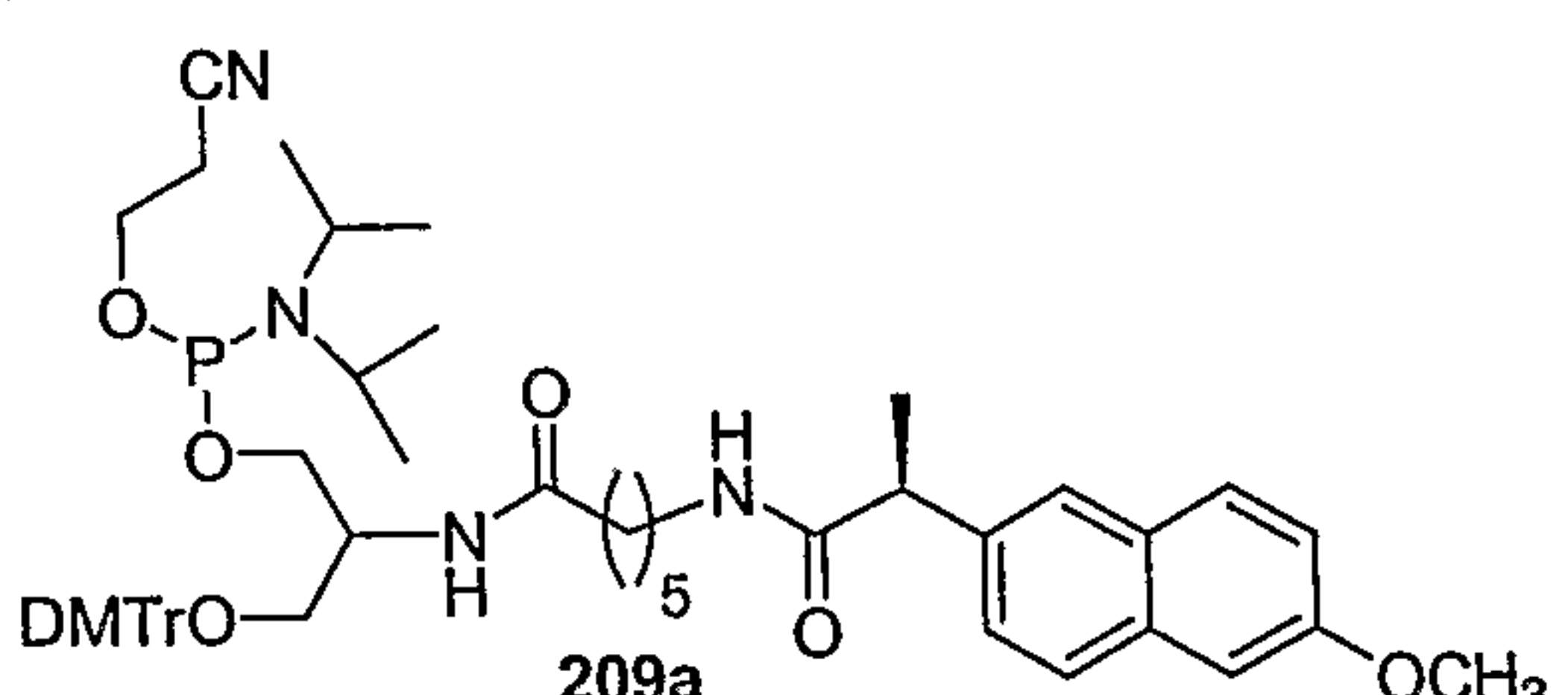
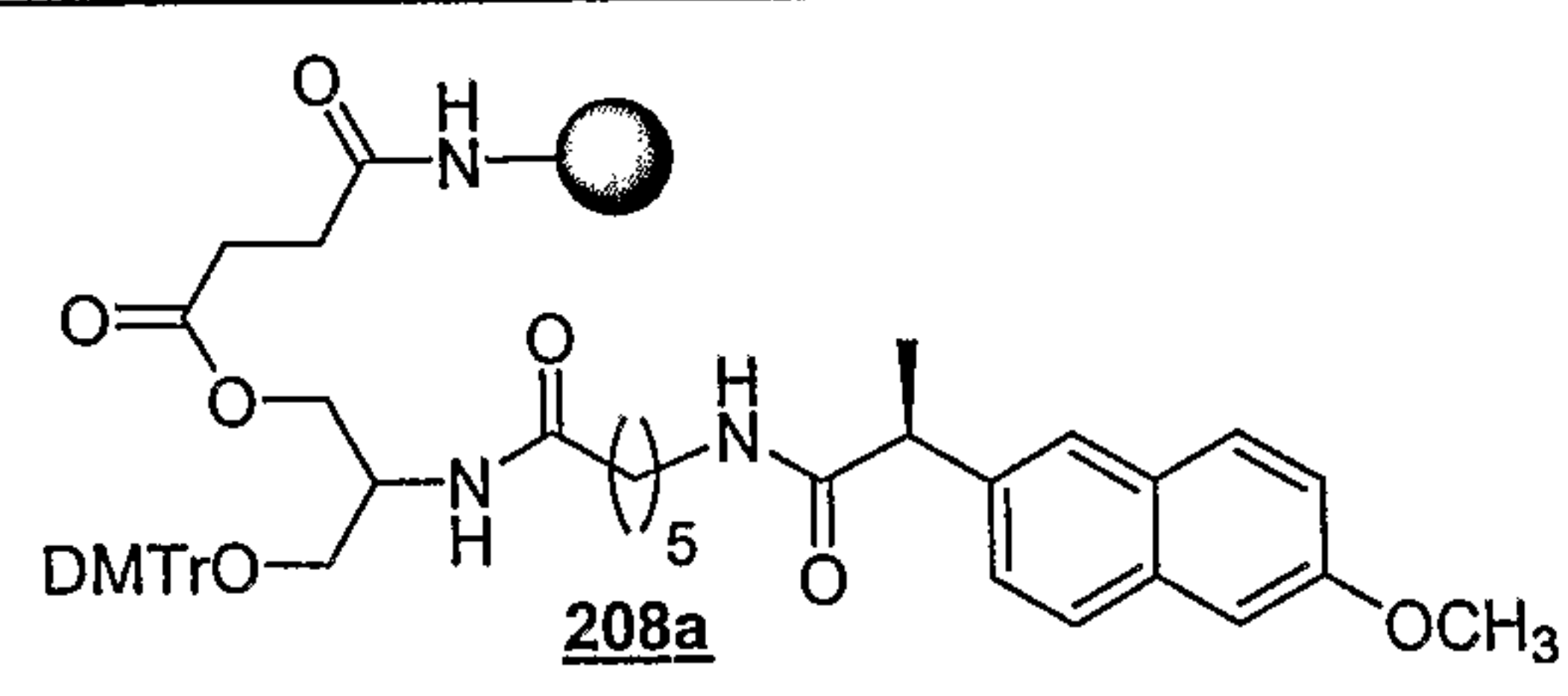
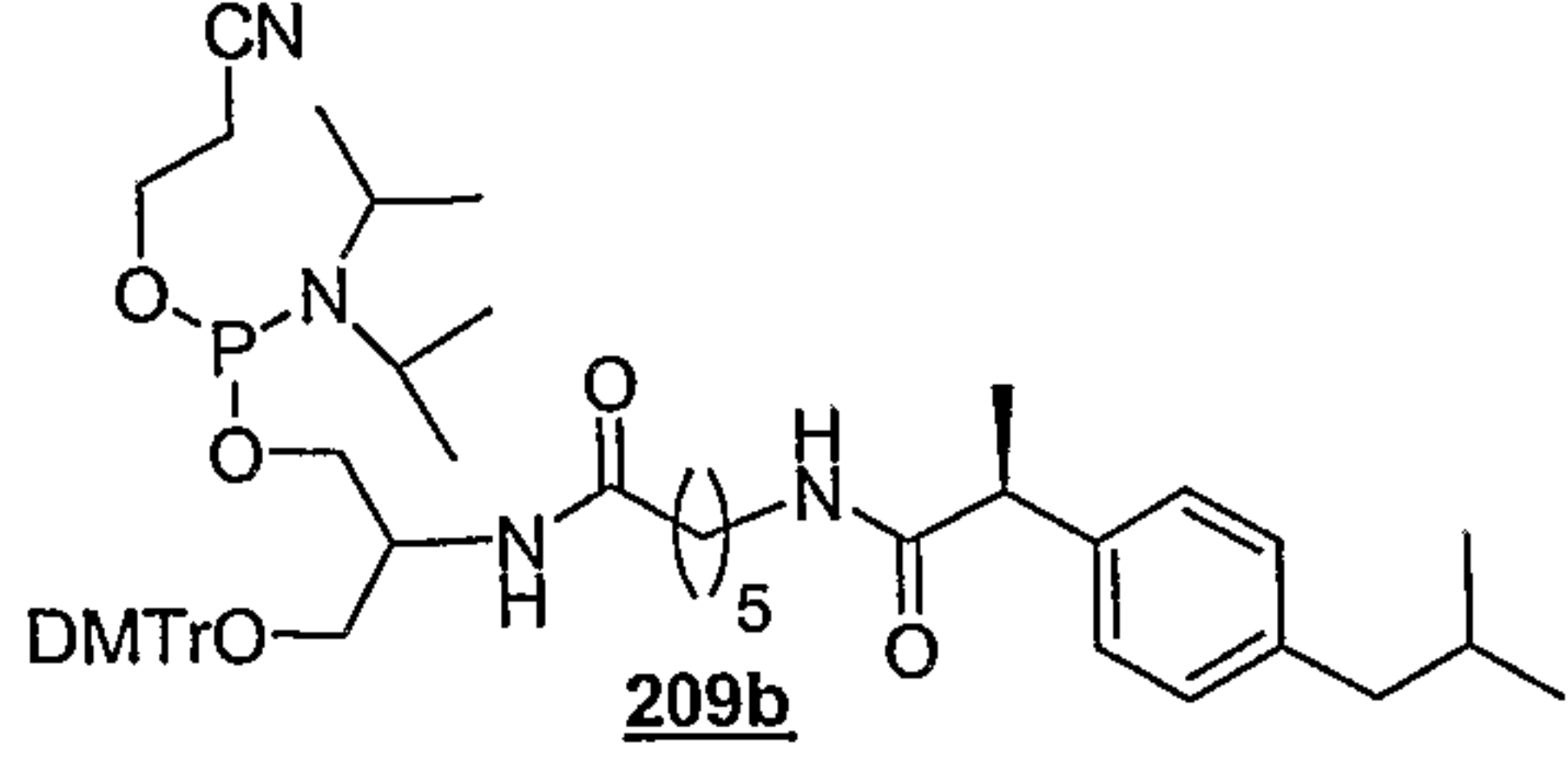
Solid Phase Synthesis and Post-Synthesis Conjugation:

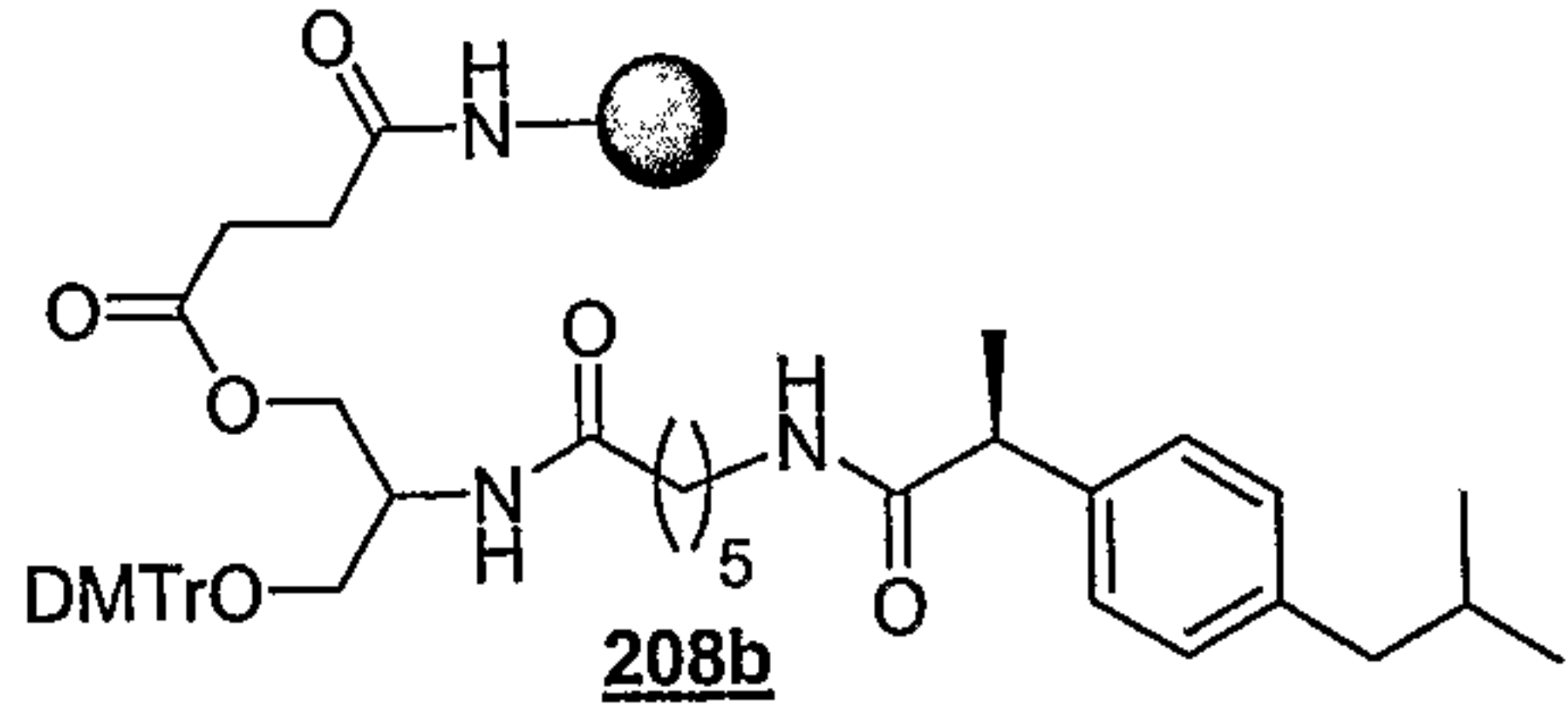
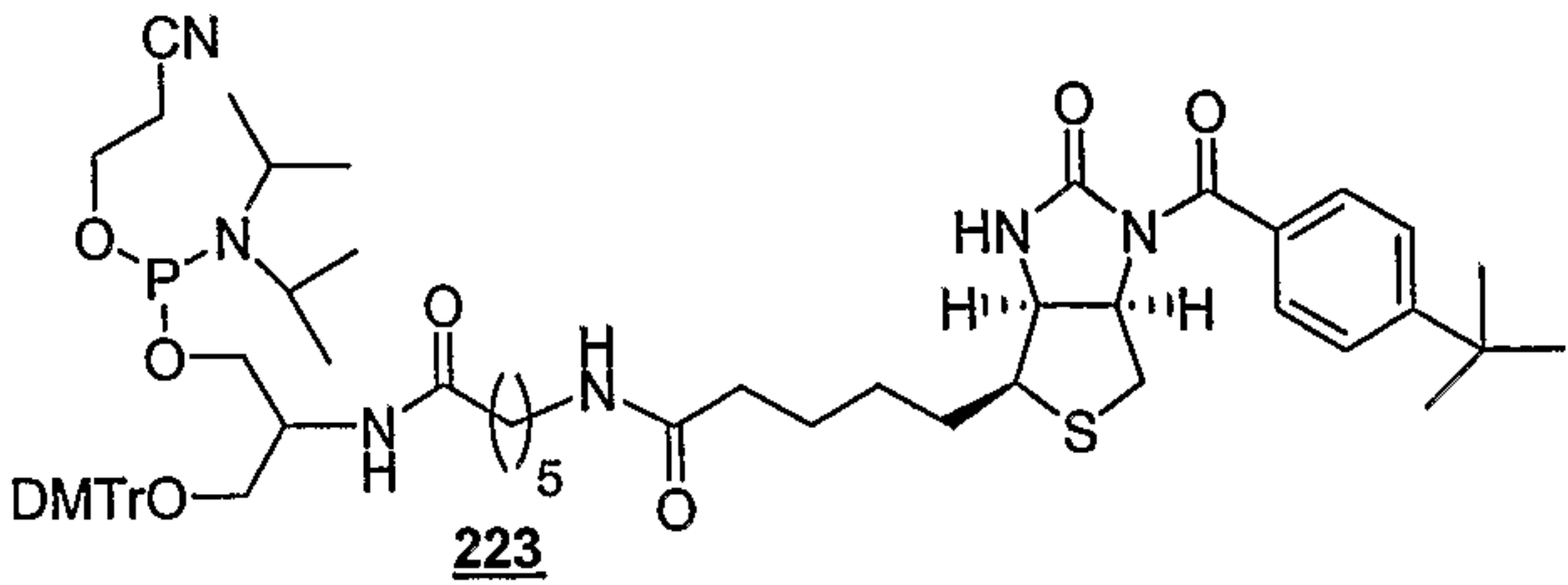
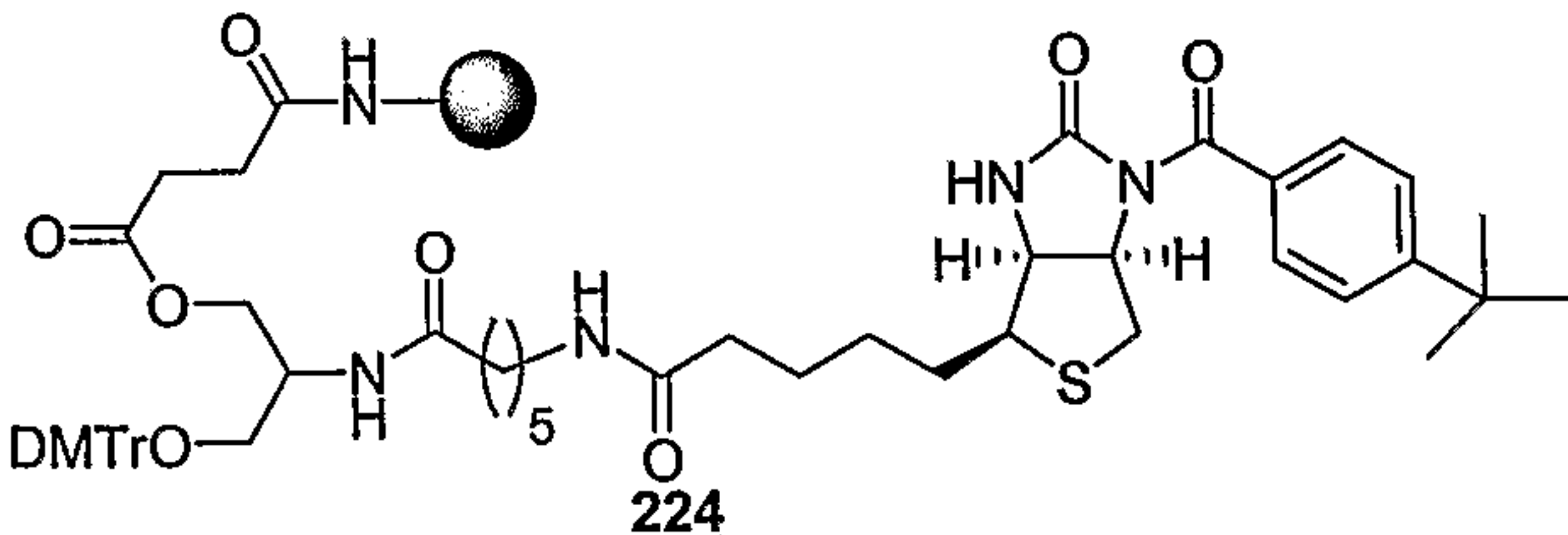
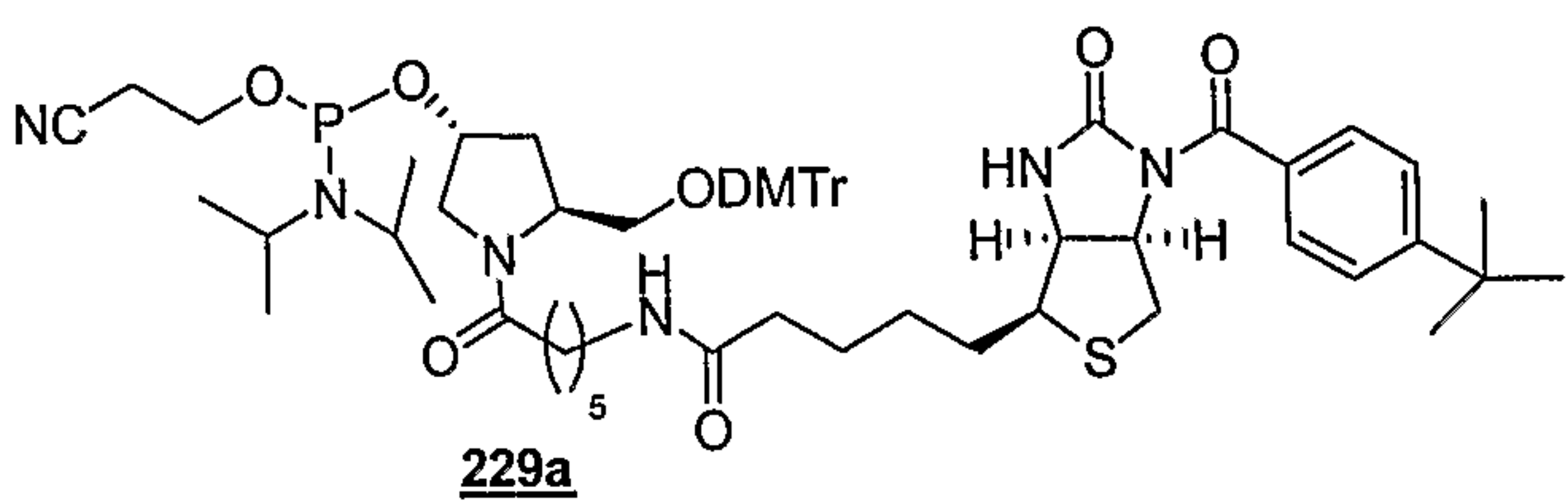
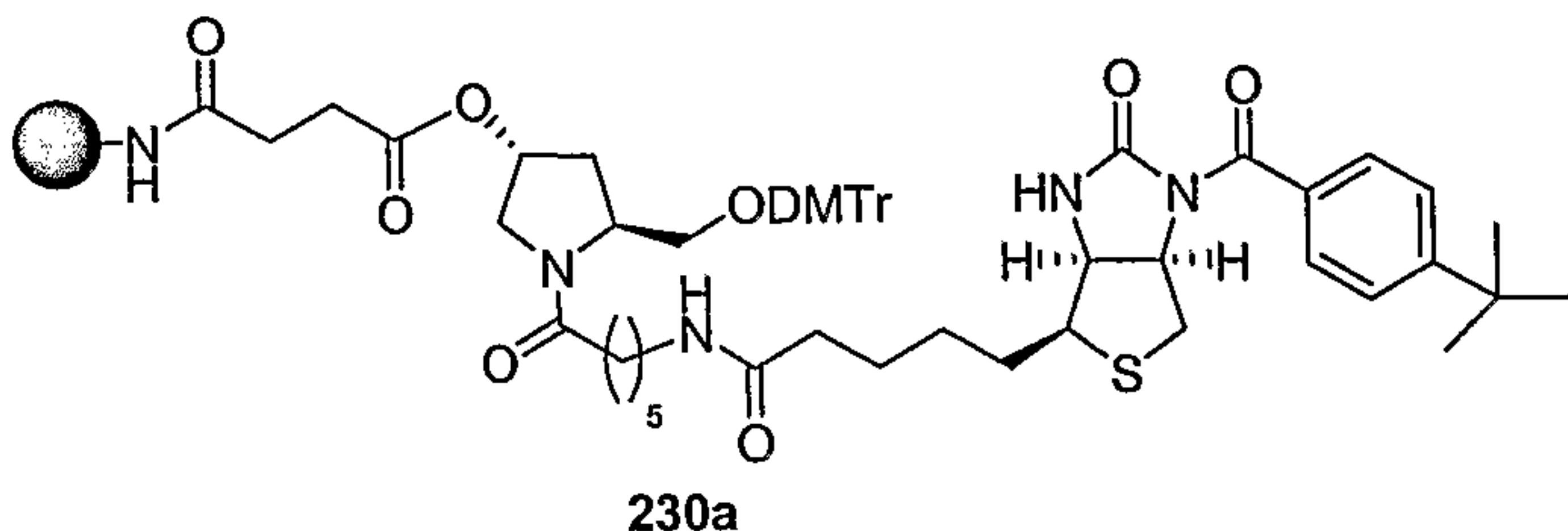
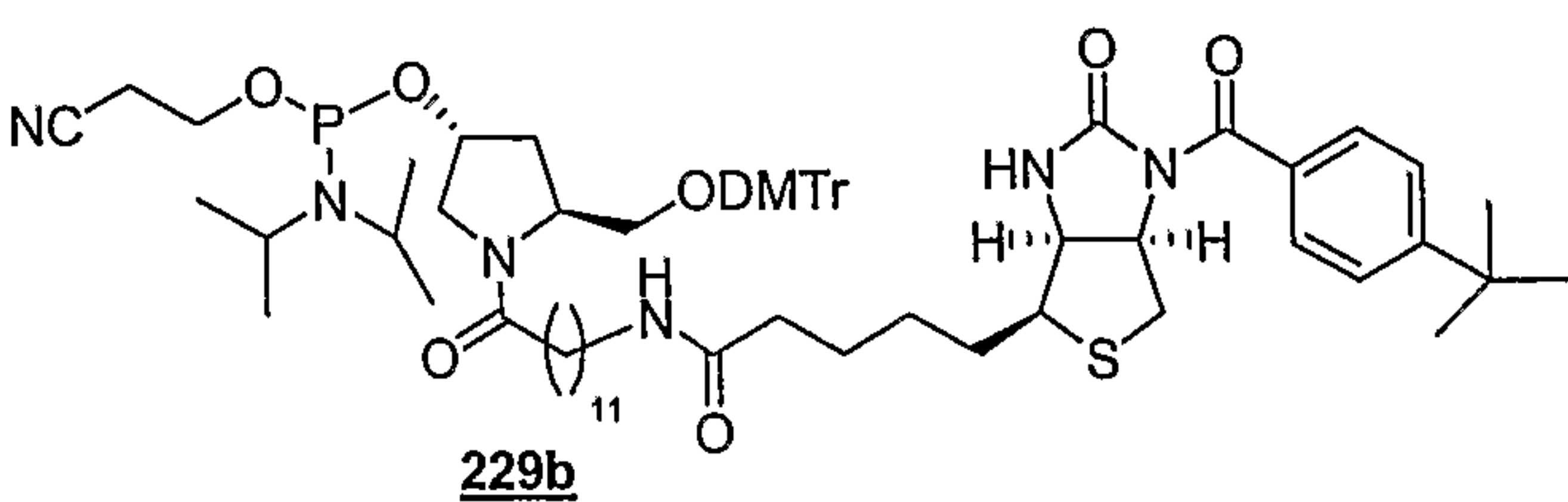


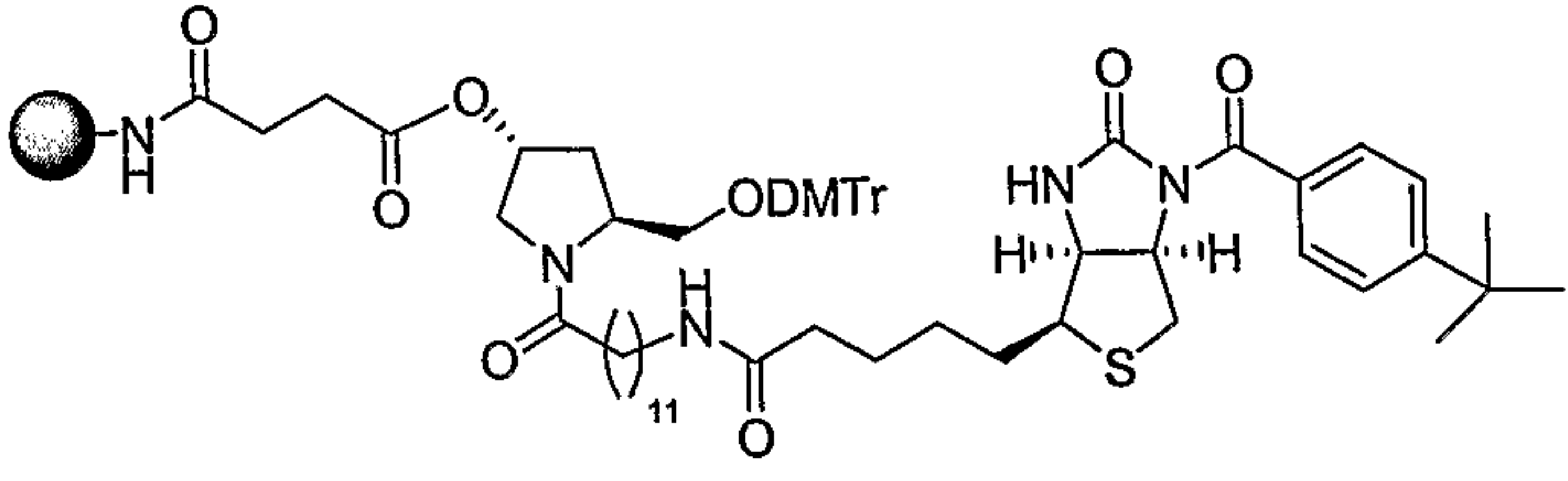
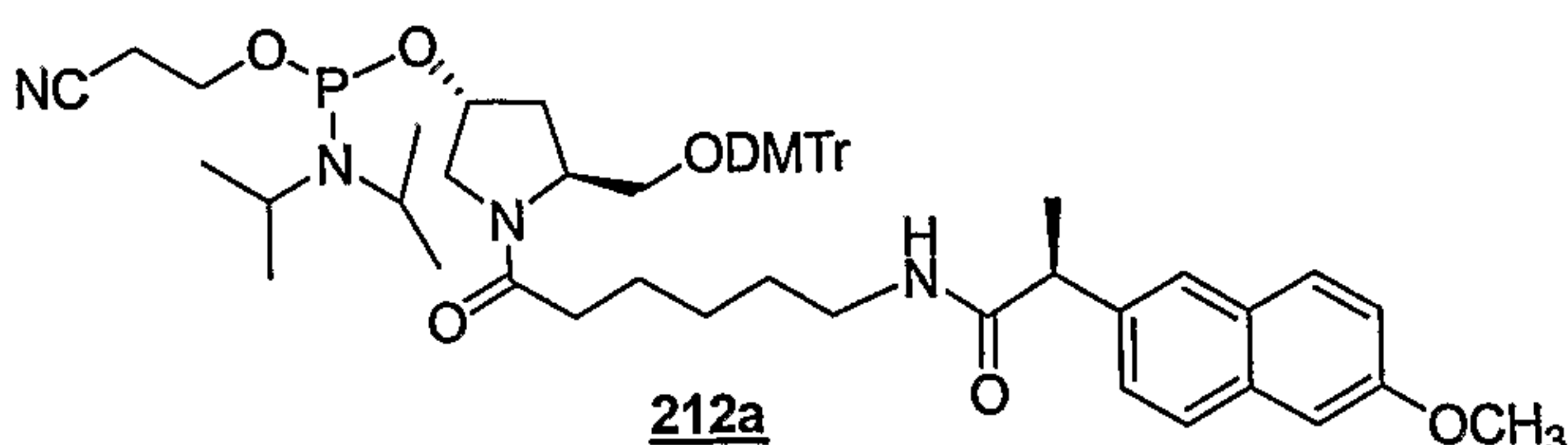
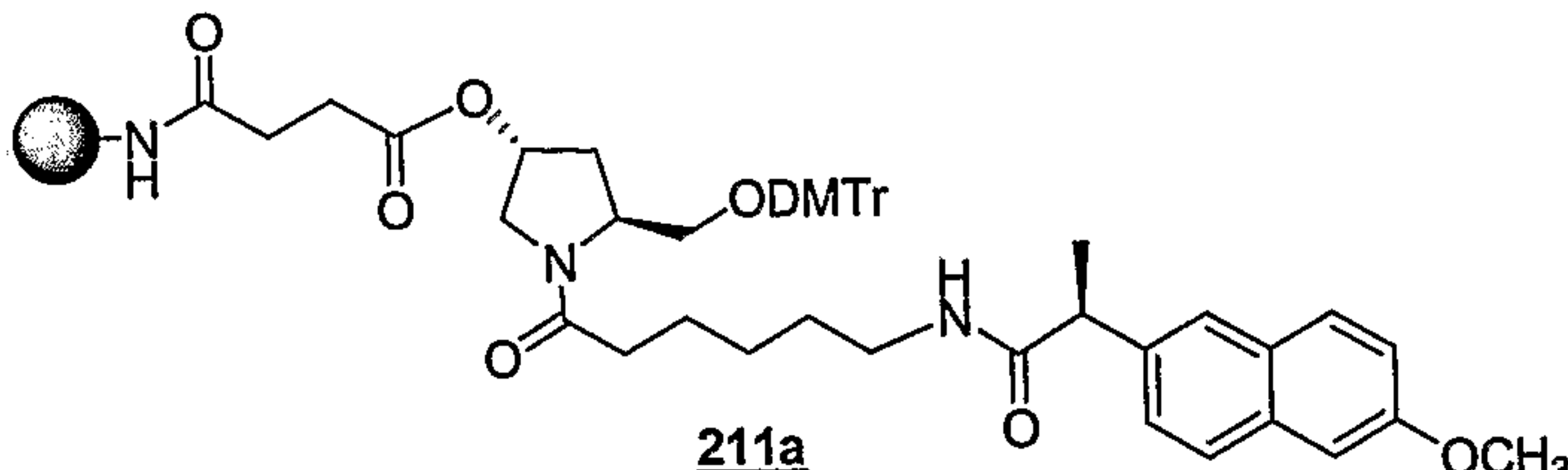
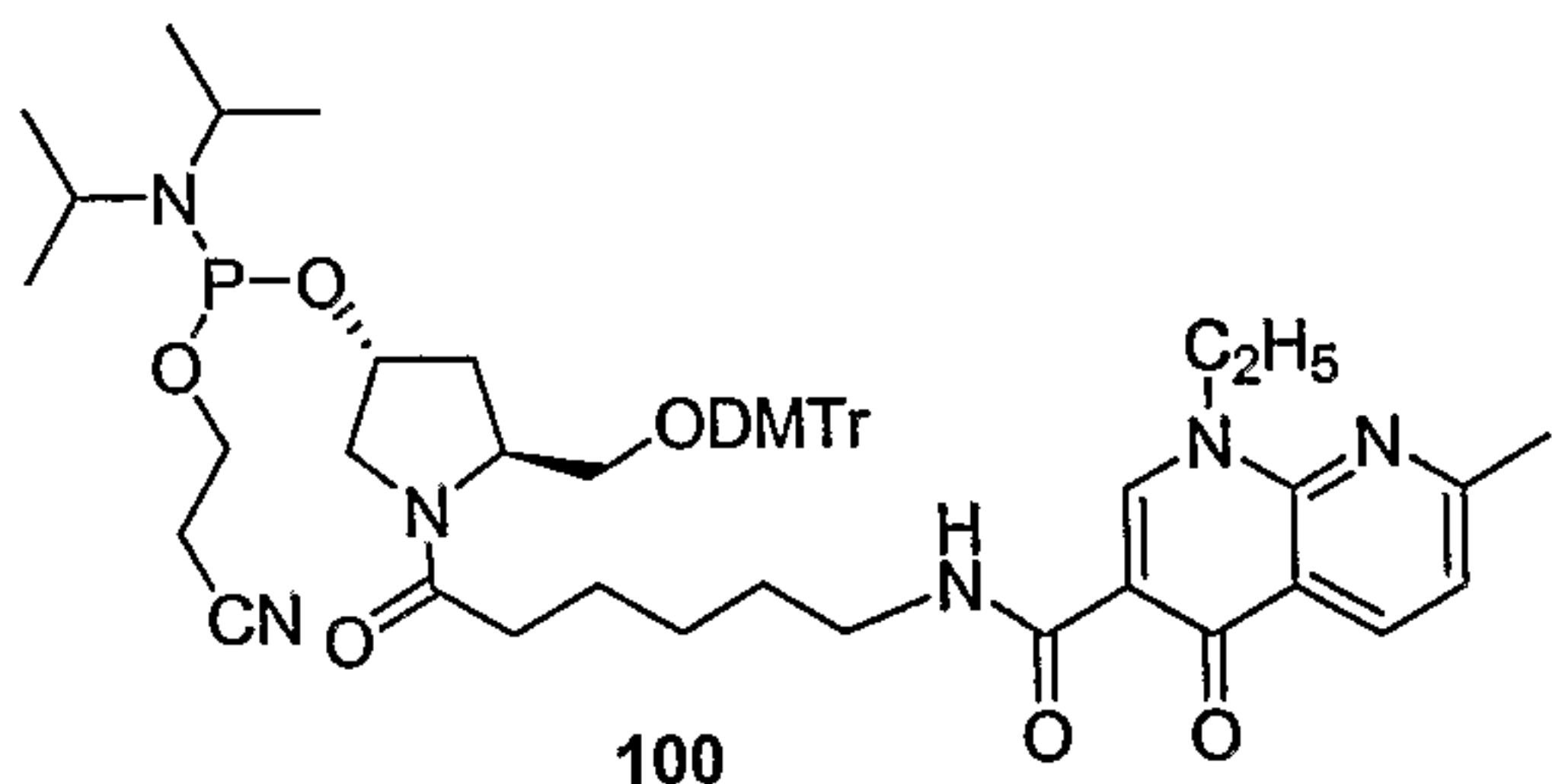
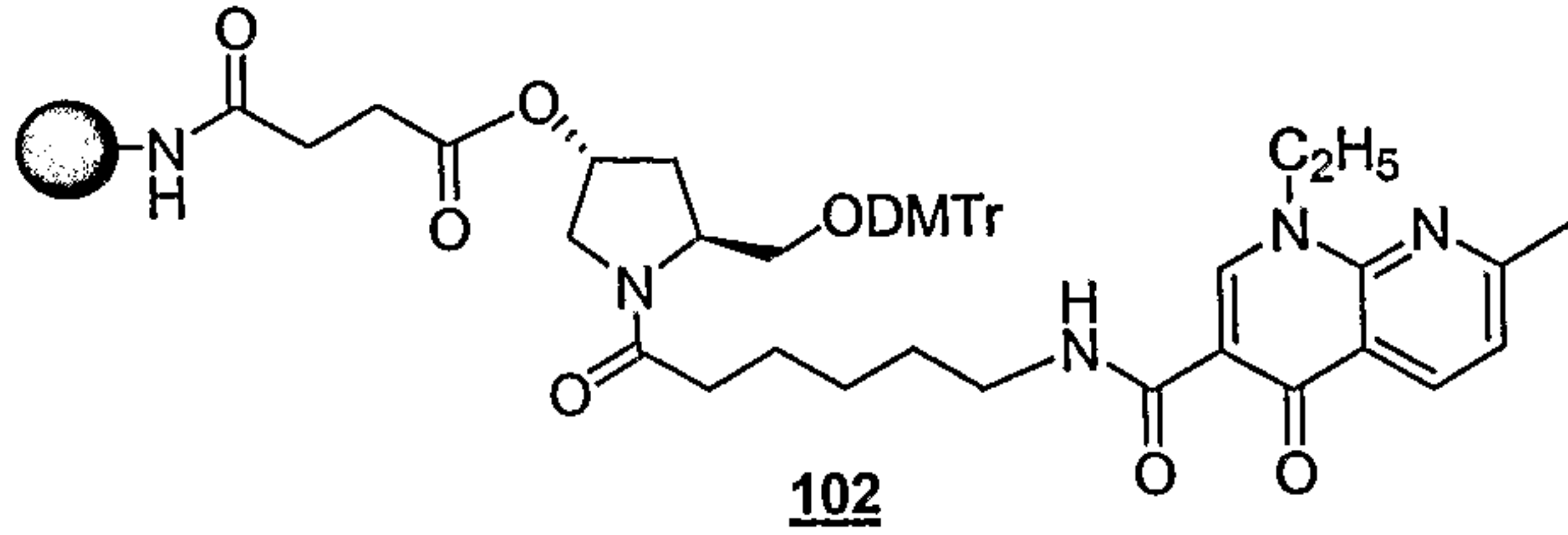
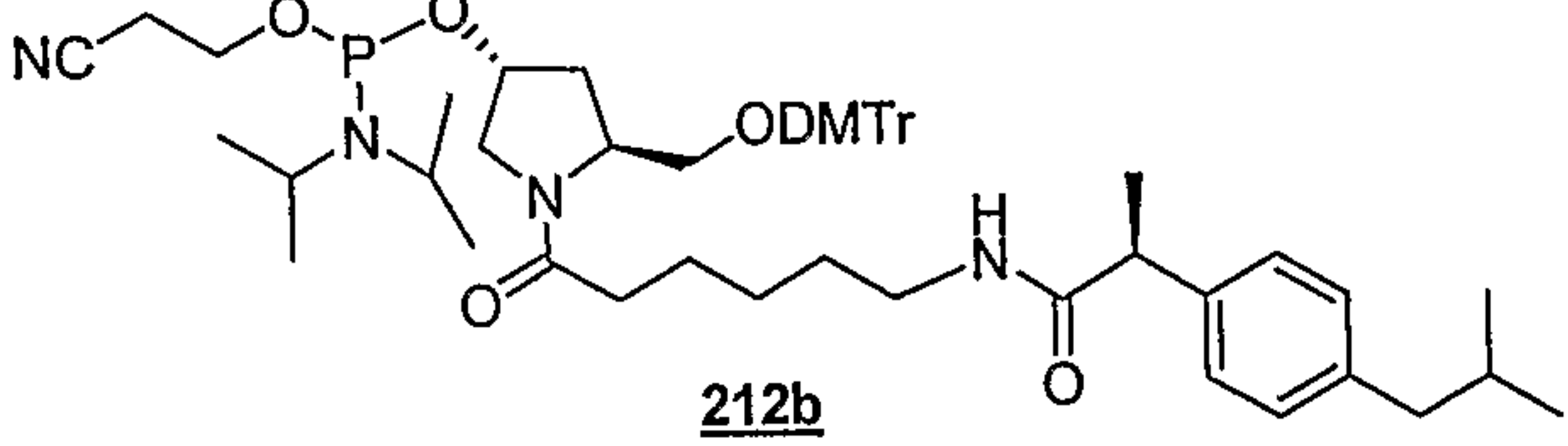
Exemplary Ligand Conjugated MonomersLCM-*E.g.-*

1	 <u>7</u>
2	 <u>9</u>
3	 <u>13</u>
4	 <u>15</u>
5	 <u>20</u>

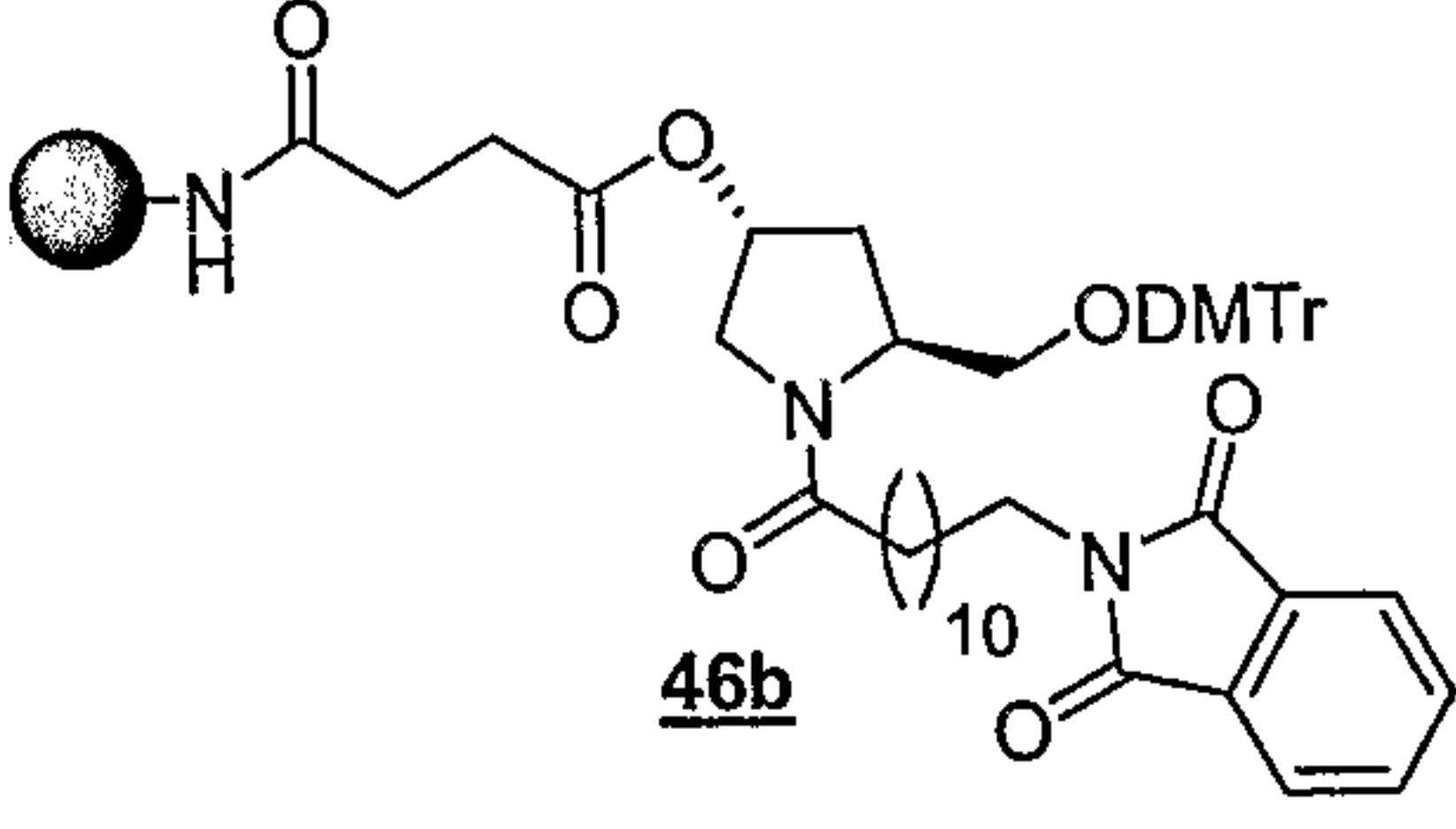
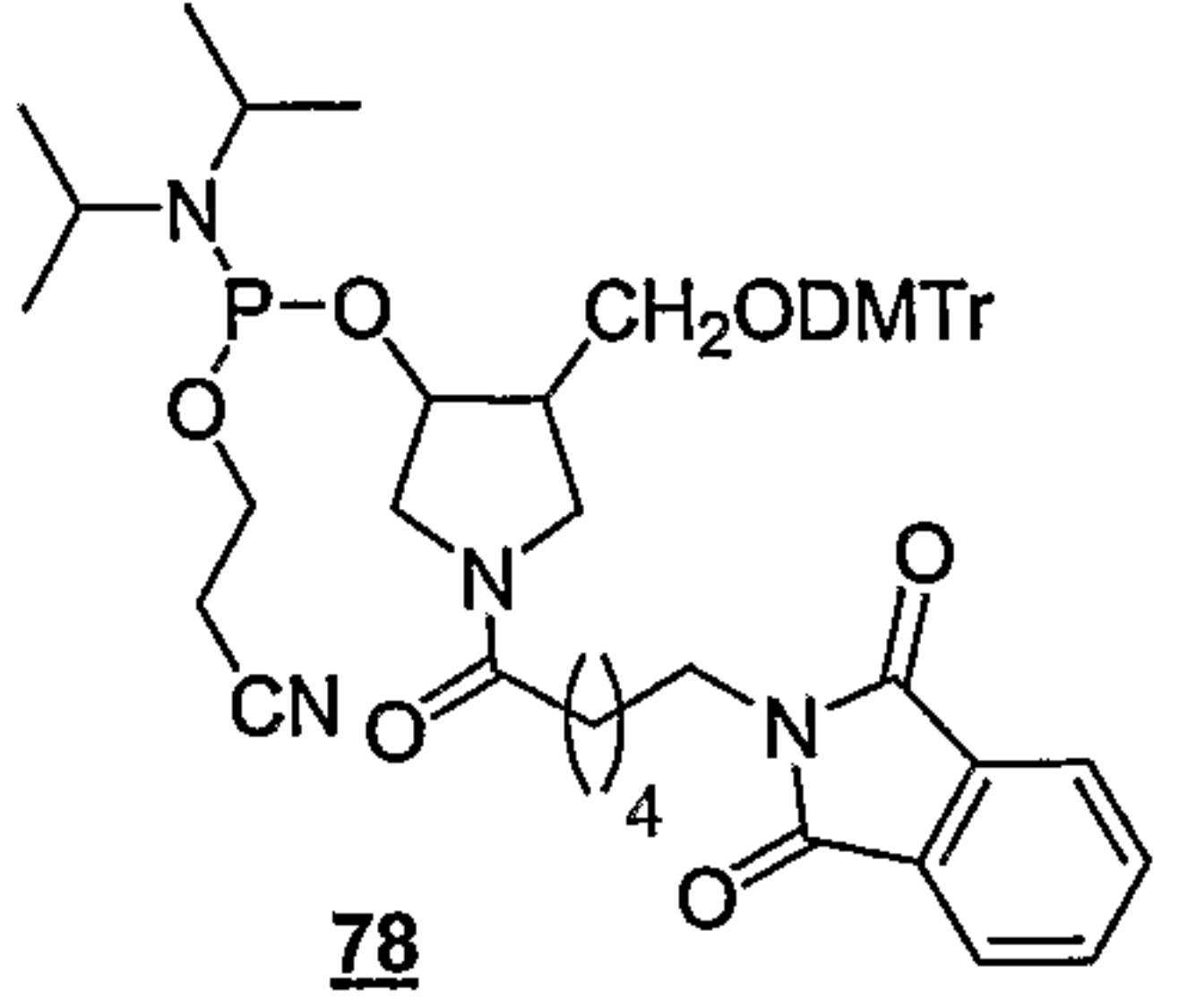
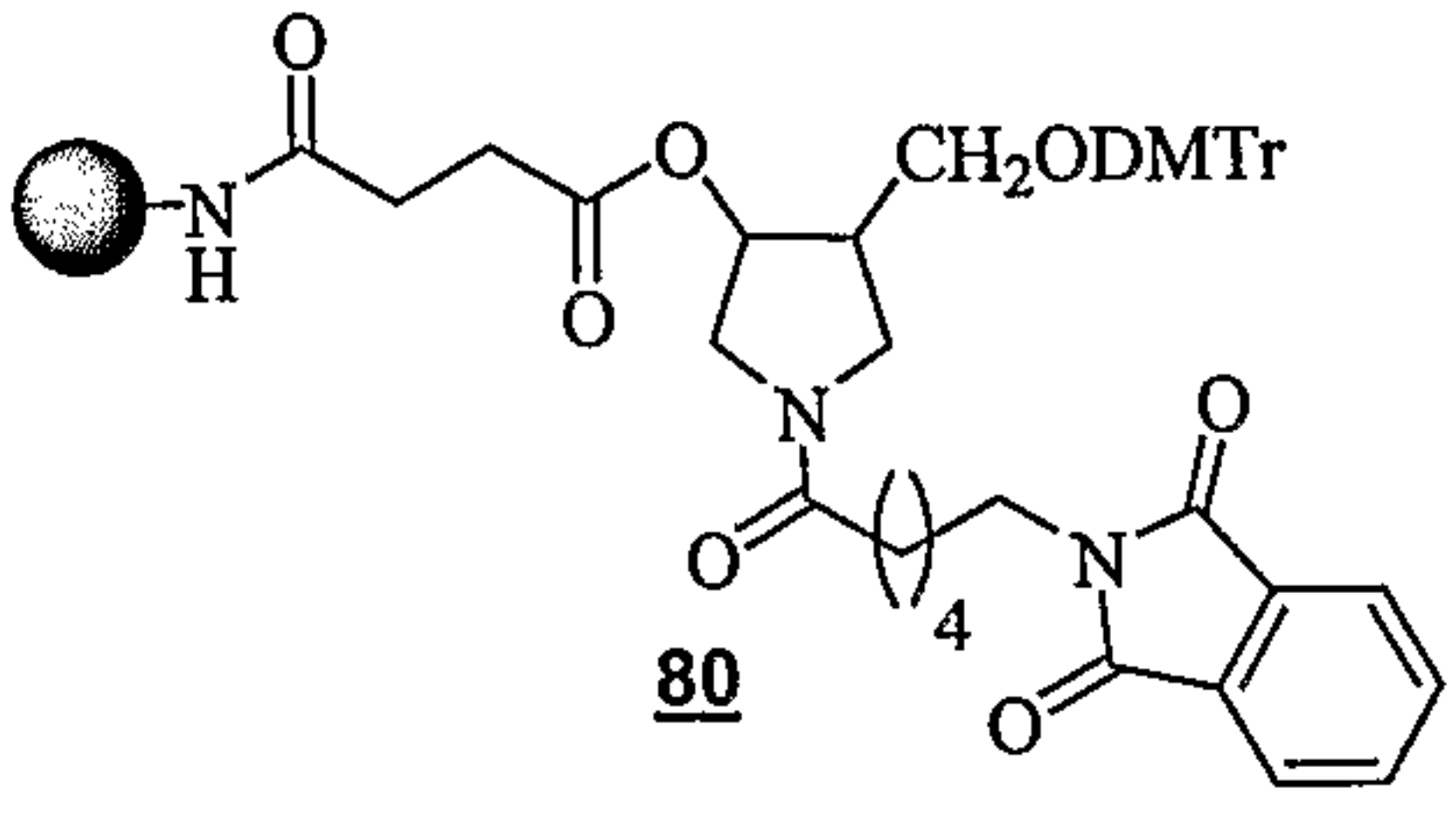
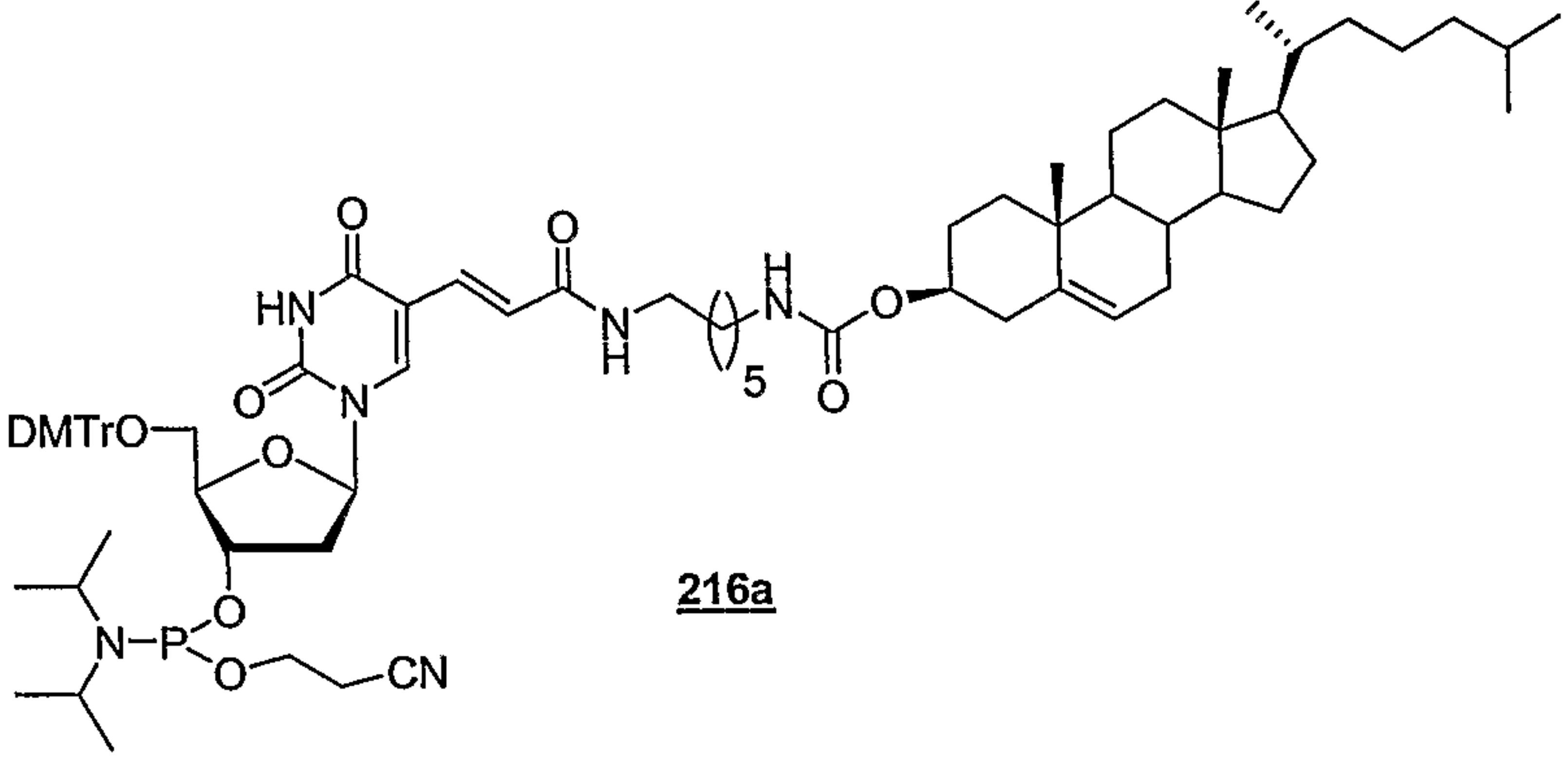
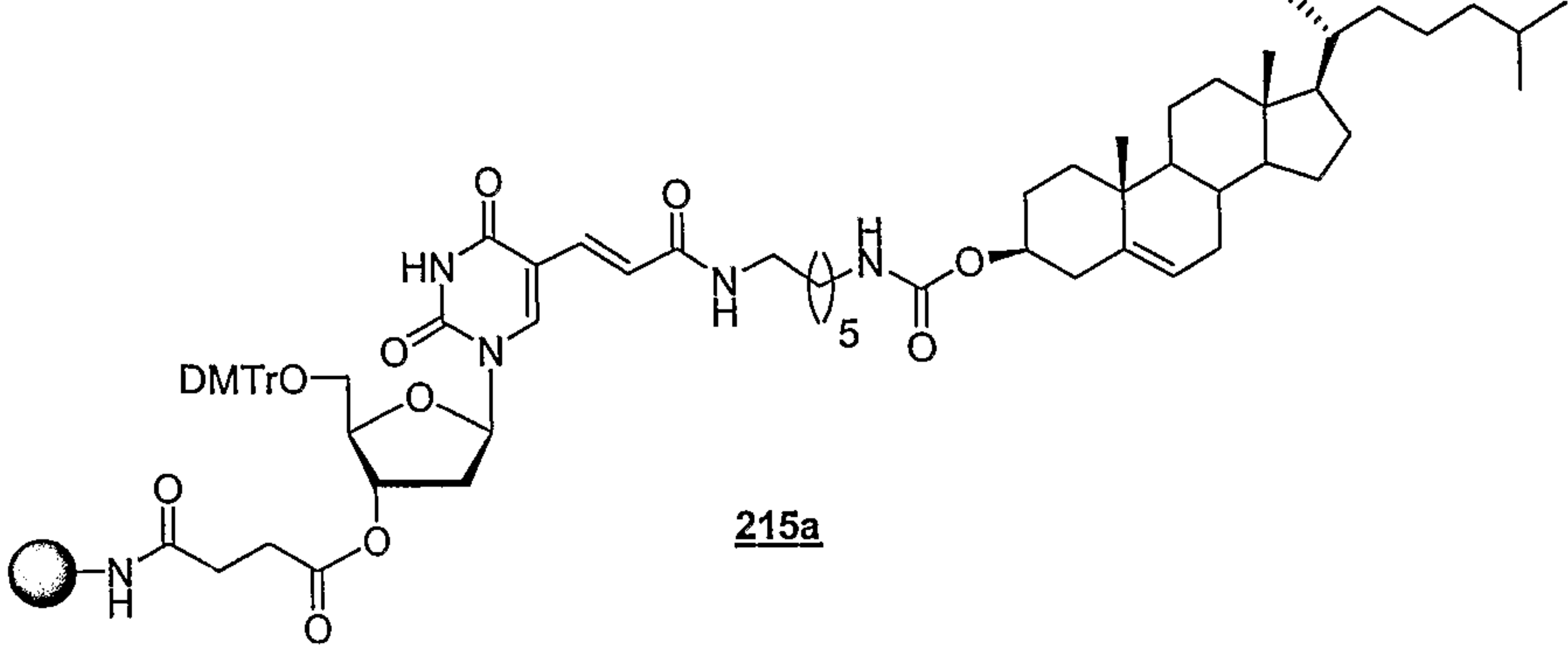
6	Chemical structure 22: A complex molecule featuring a DMTrO-protected pyrrolidine ring connected via an ester linkage to a long chain containing a thioether and a steroid-like core.
7	Chemical structure 26: A molecule with a DMTrO-protected pyrrolidine ring, a phosphonate group, and a steroid-like core.
8	Chemical structure 28: A molecule with a DMTrO-protected pyrrolidine ring, a thioether, and a steroid-like core.
9	Chemical structure 33: A molecule with a phosphonate group, a DMTrO-protected pyrrolidine ring, and a steroid-like core.
10	Chemical structure 35: A molecule with a DMTrO-protected pyrrolidine ring, a thioether, and a steroid-like core.
11	Chemical structure 45a: A molecule with a phosphonate group, a DMTrO-protected pyrrolidine ring, and a benzimidazole-like core.

12	 46a
13	 55
14	 56
15	 209a
16	 208a
17	 209b

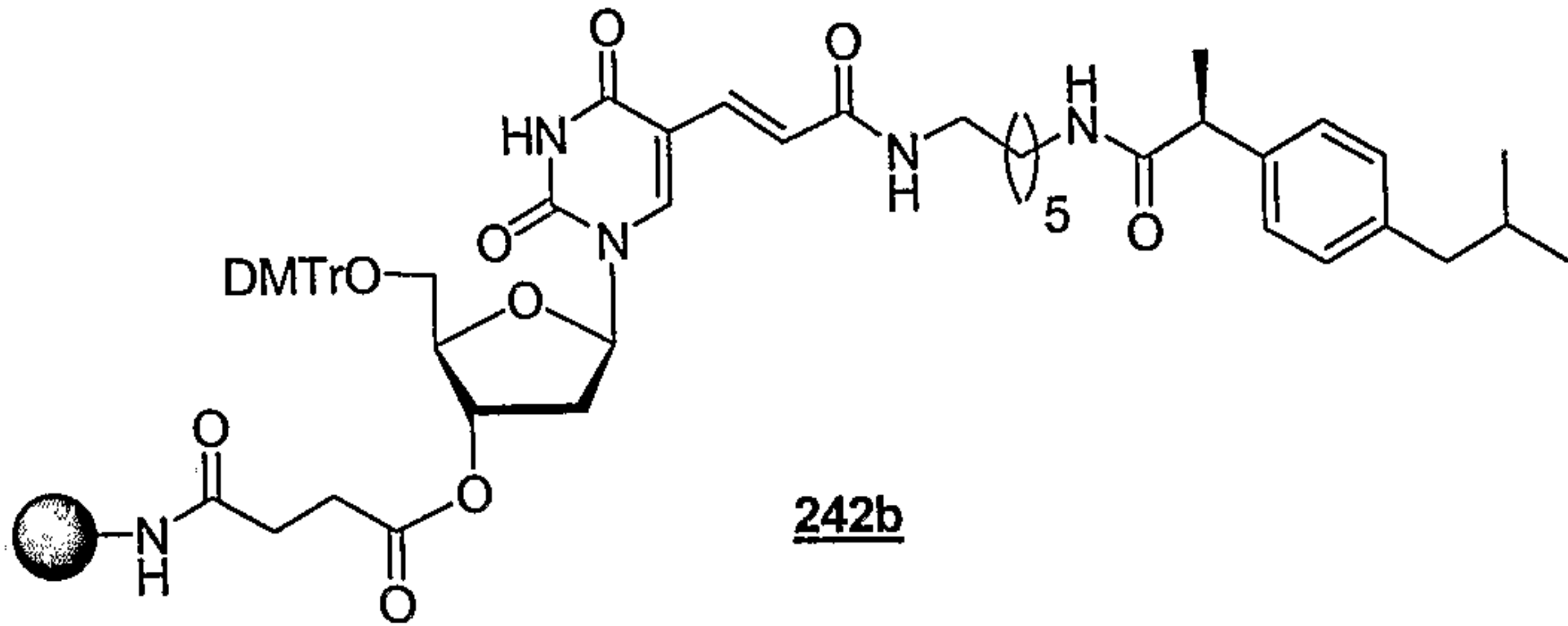
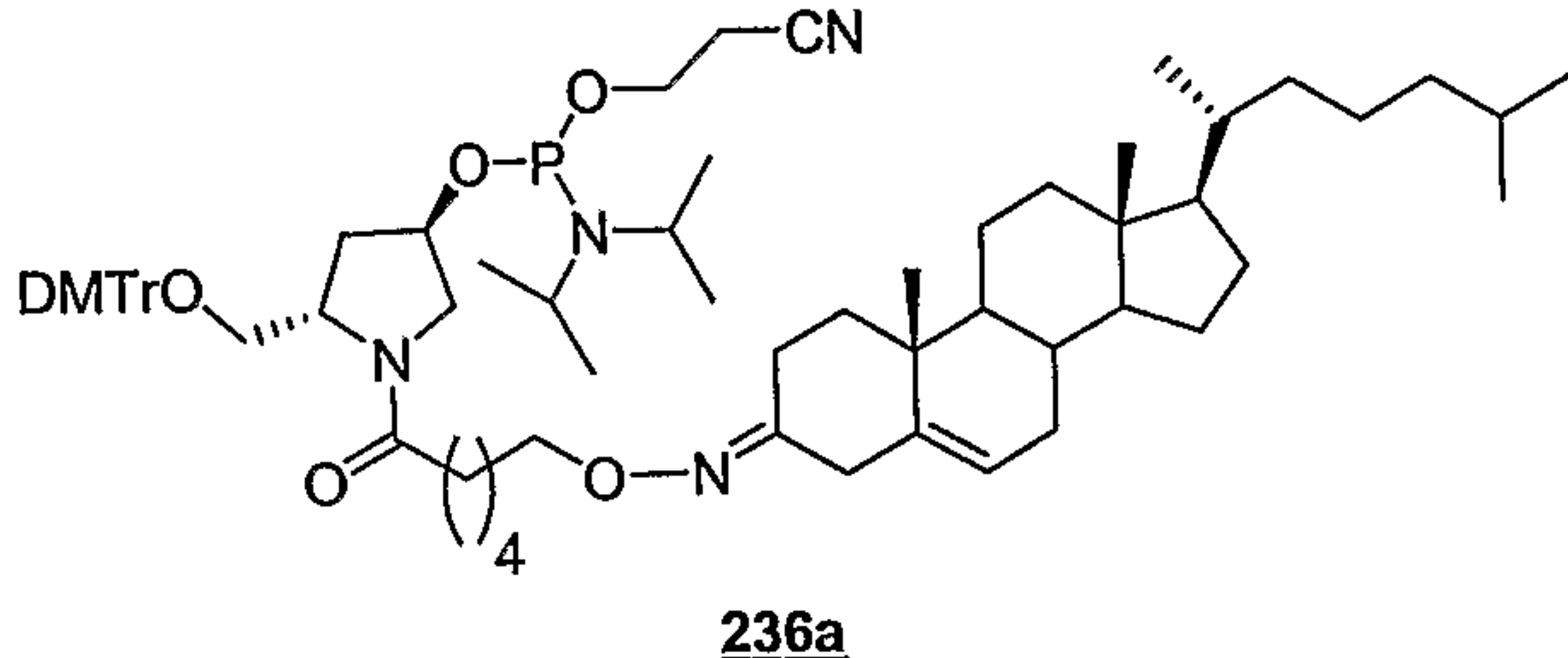
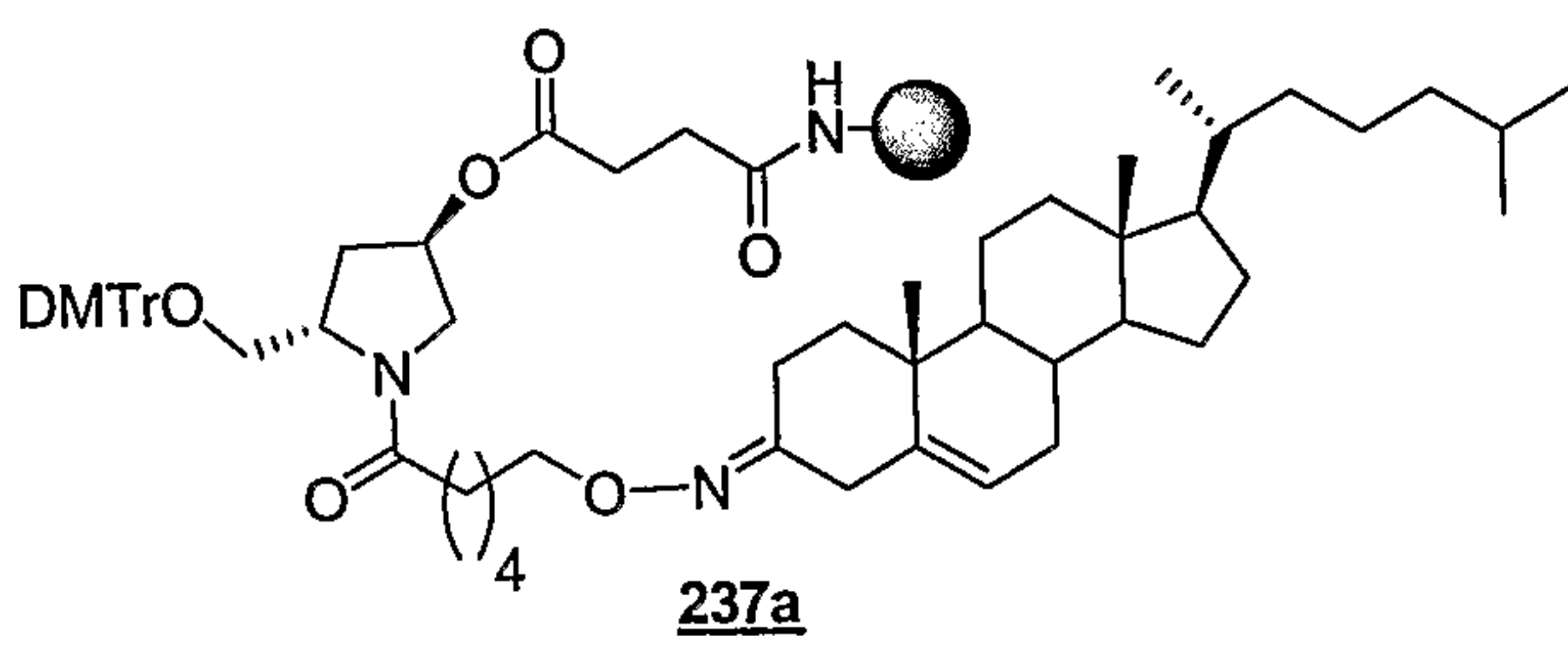
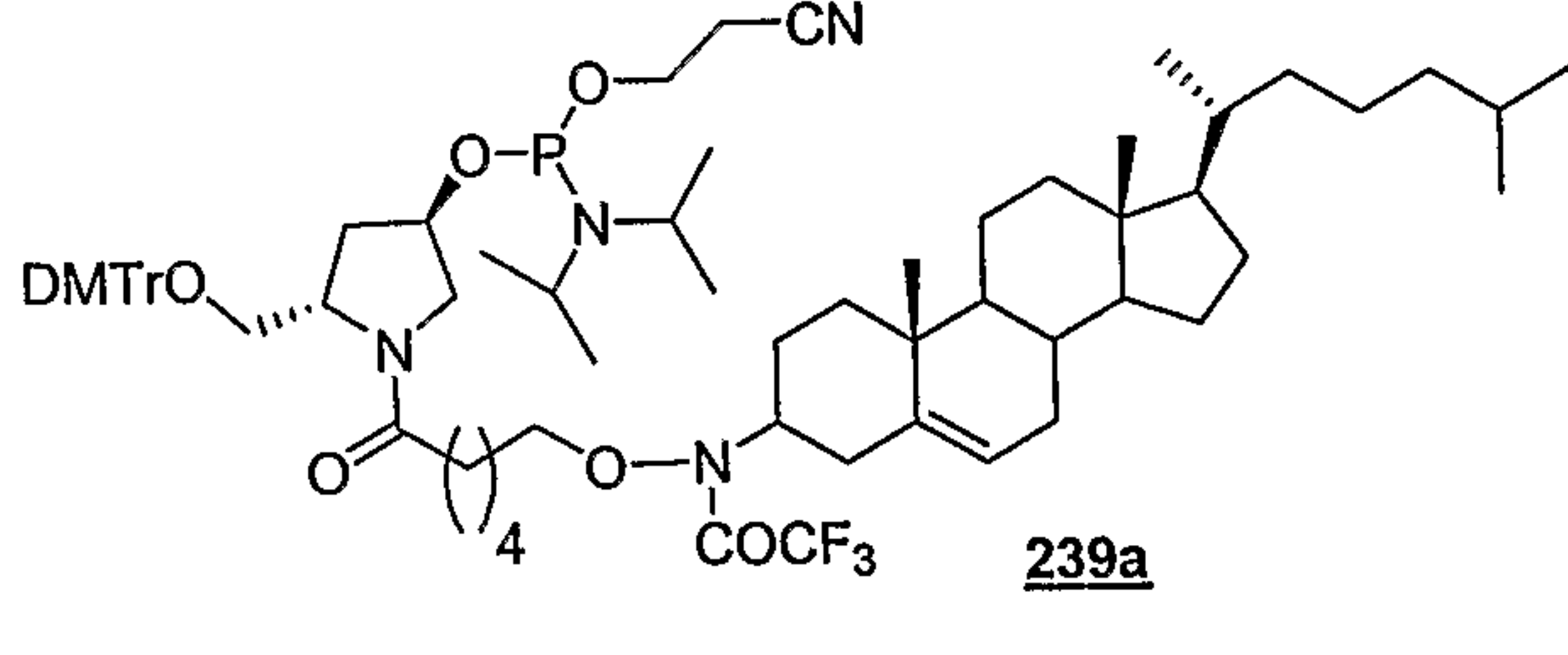
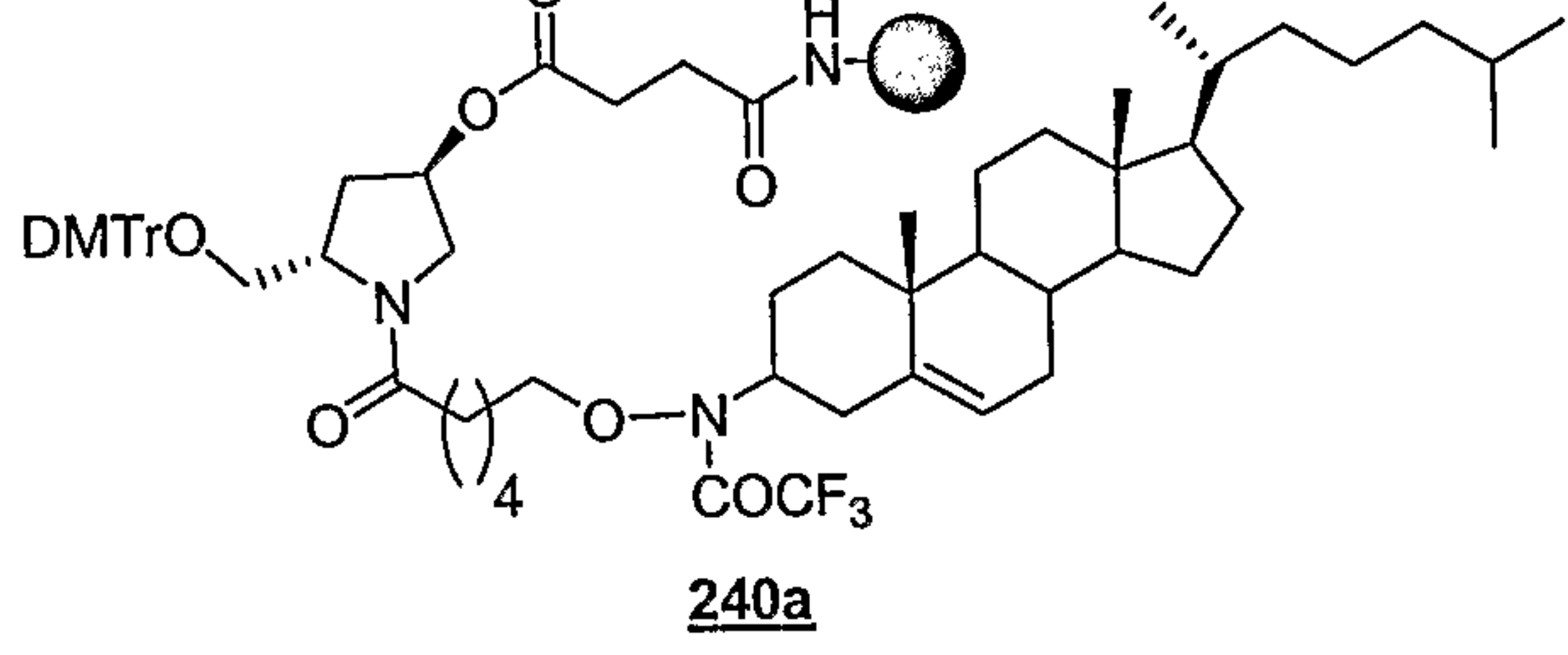
18	 208b
19	 223
20	 224
21	 229a
22	 230a
23	 229b

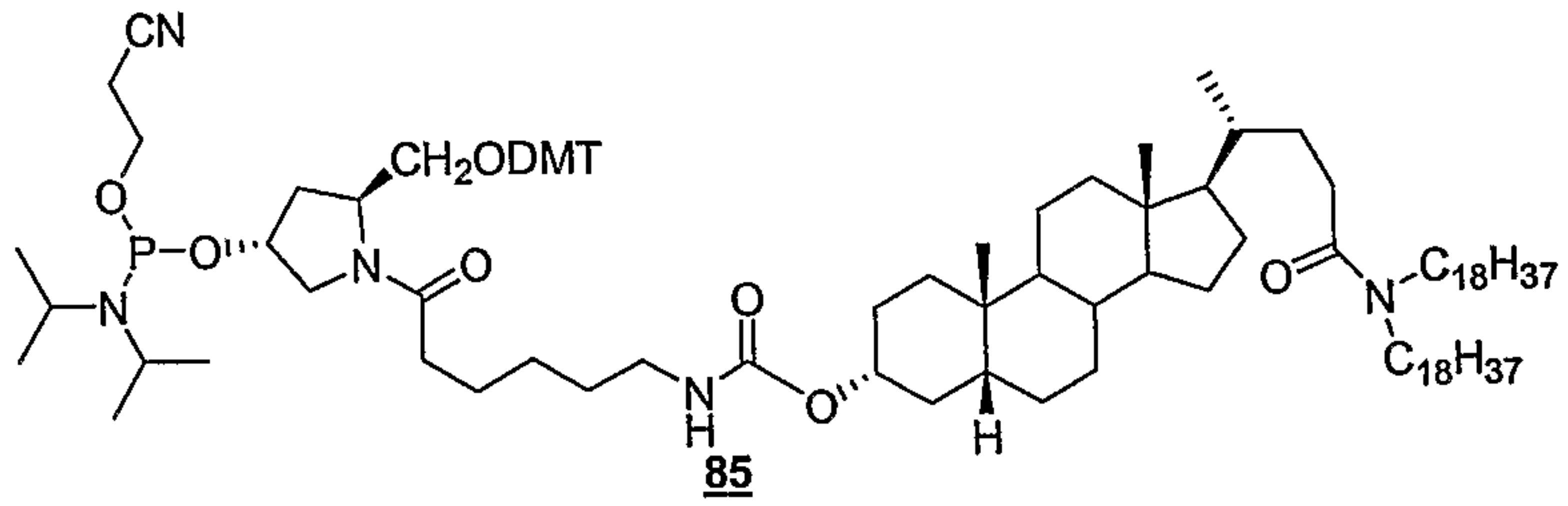
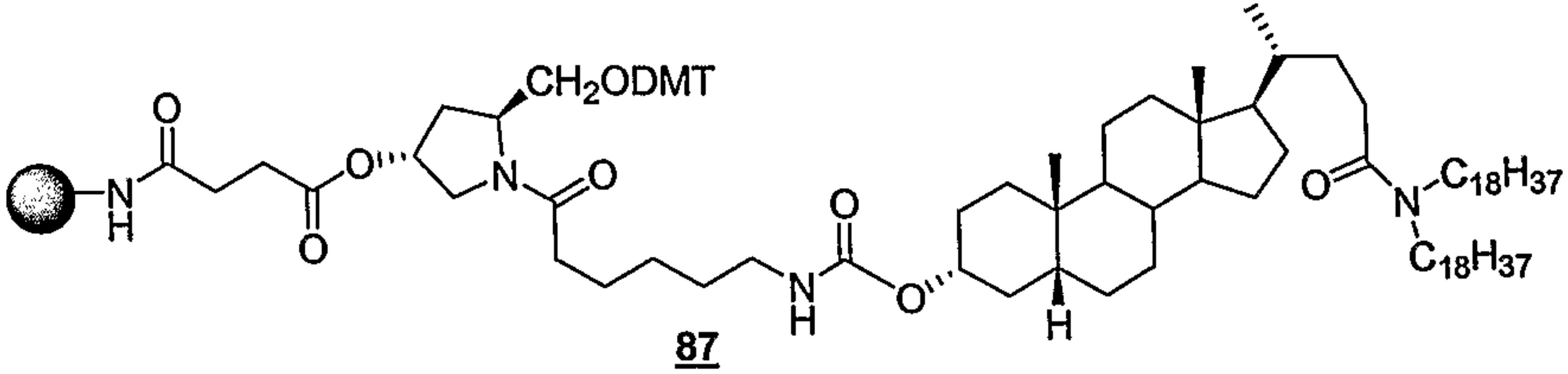
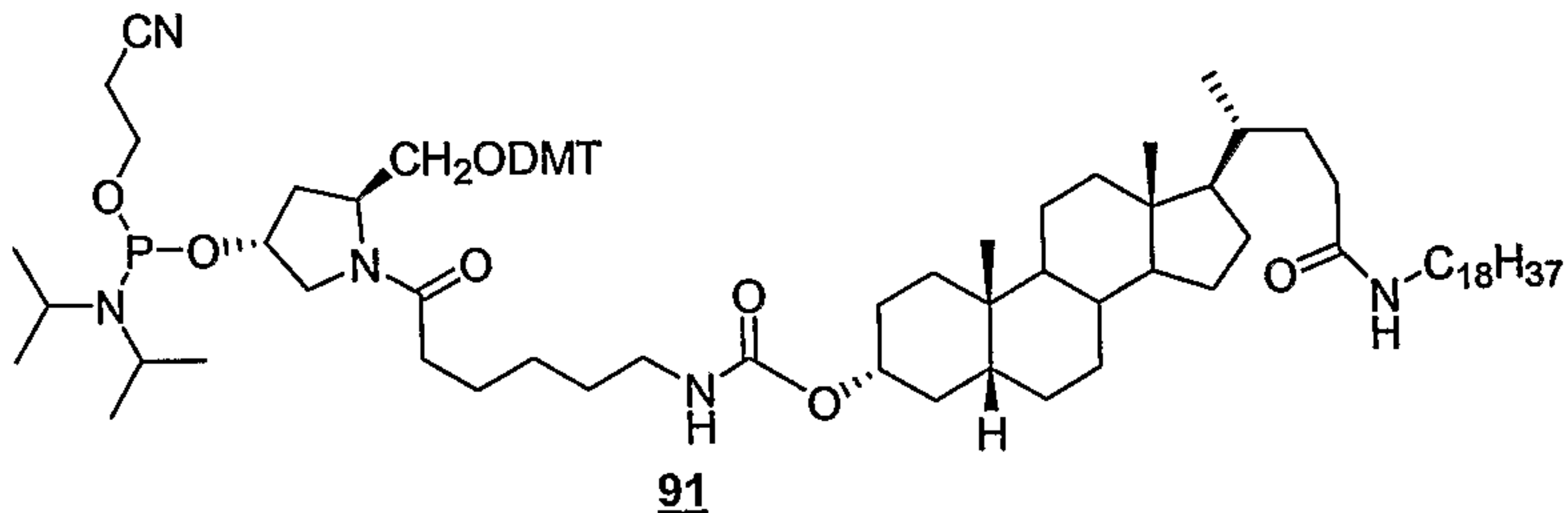
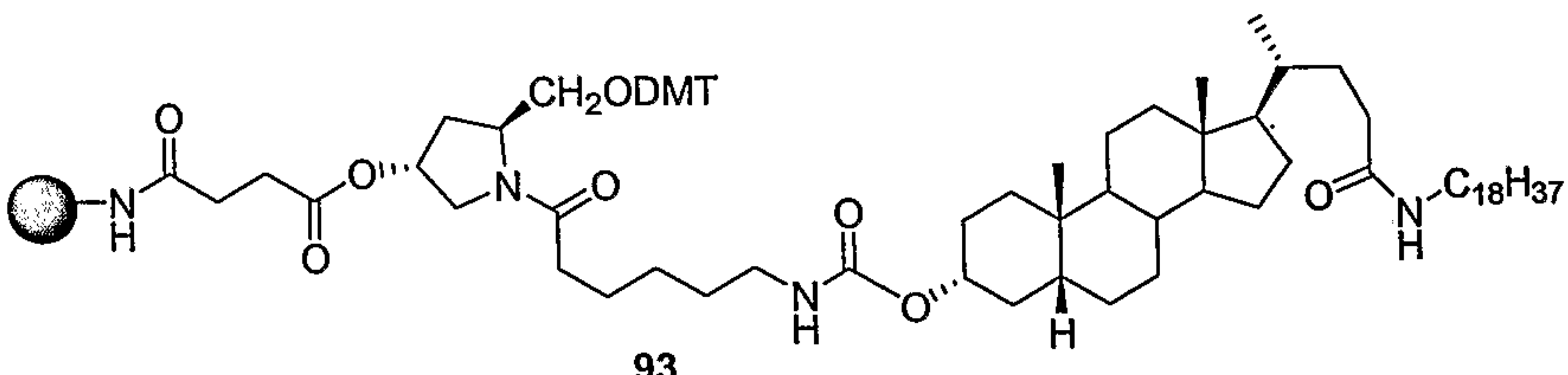
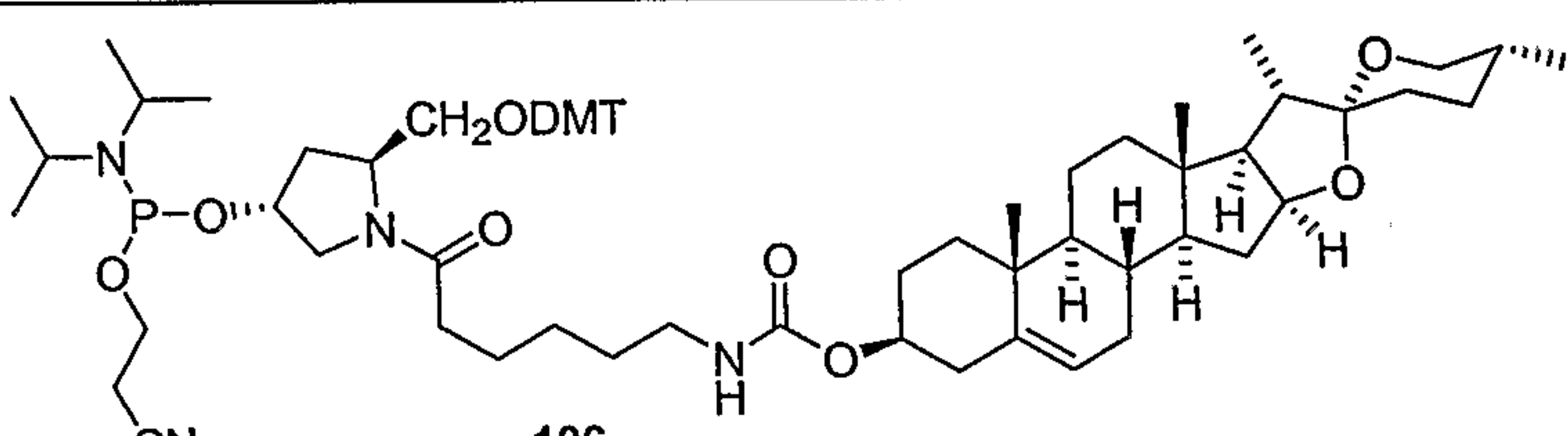
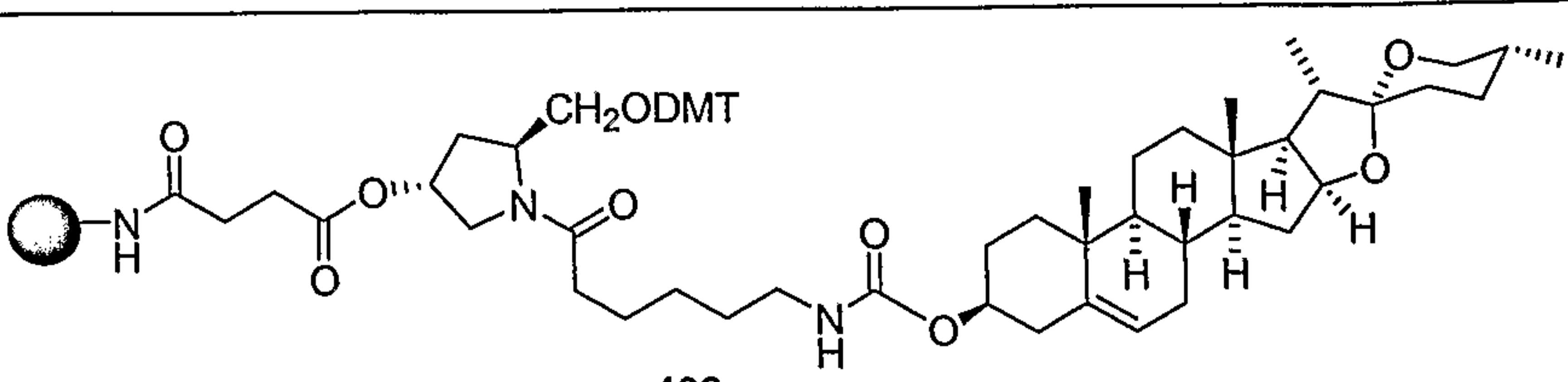
24	 <u>230b</u>
25	 <u>212a</u>
26	 <u>211a</u>
27	 <u>100</u>
28	 <u>102</u>
29	 <u>212b</u>

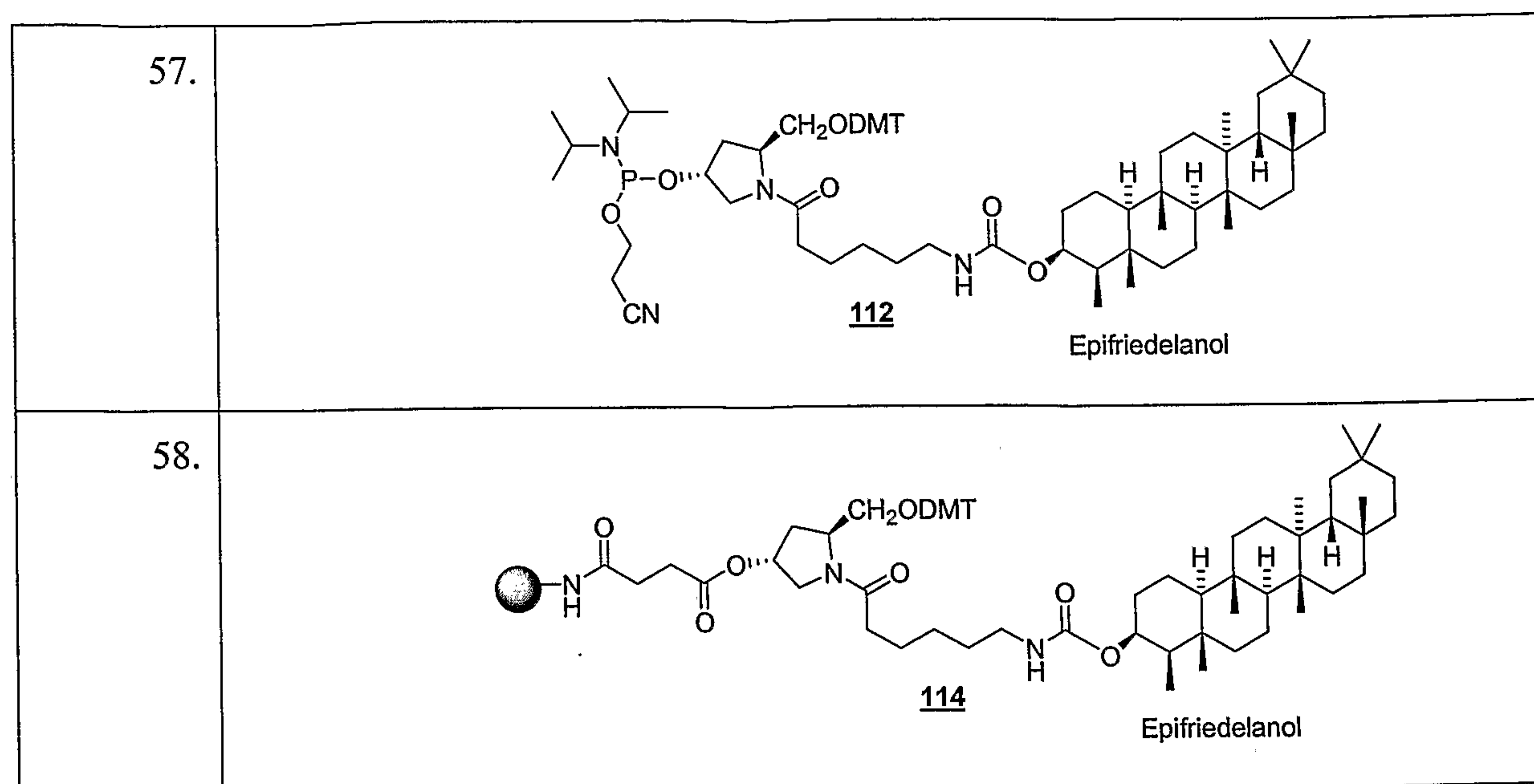
30	211b
31	67
32	69
33	95
34	97
35	45b

36	 46b
37	 78
38	 80
39	 216a
40	 215a

41	216b
42	215b
43	243a
44	242a
45	243b

46	 242b
47	 236a
48	 237a
49	 239a
50	 240a

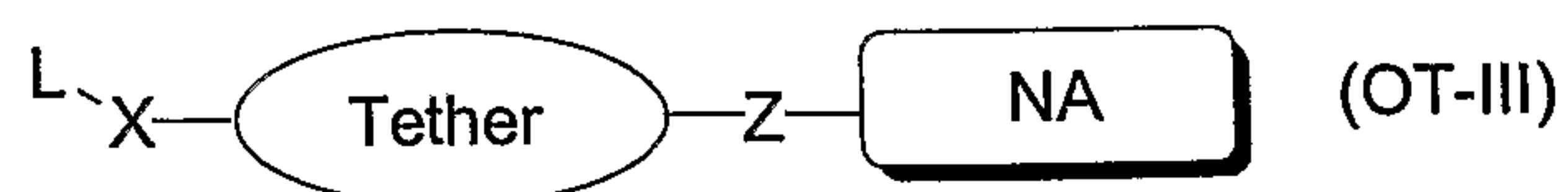
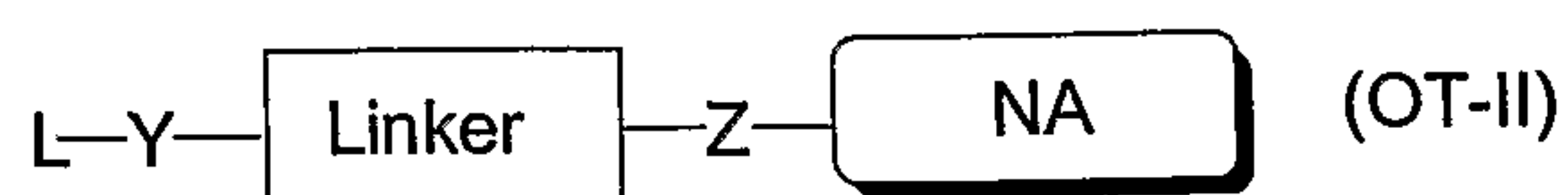
51	 <u>85</u>
52	 <u>87</u>
53	 <u>91</u>
54	 <u>93</u>
55.	 <u>106</u> Diosgenin
56.	 <u>108</u> Diosgenin



Conjugation of Ligands to oligonucleotide agents

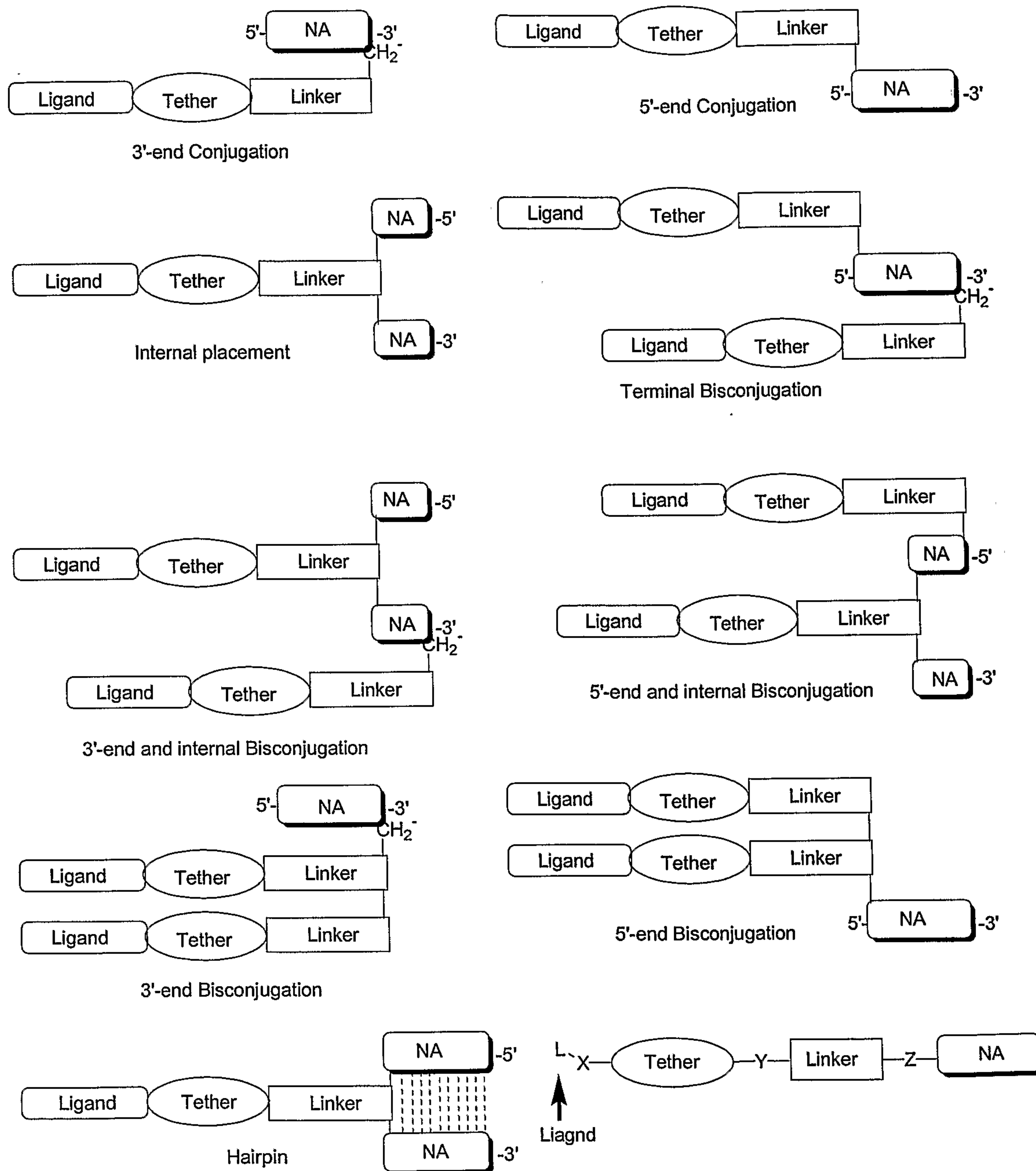
5 The conjugation of a ligand to an oligonucleotide agent, *e.g.*, an oligonucleotide agent that targets an miRNA or pre-miRNA can have a favorable effect on the modulating effect of the agent. For example, the agent can improve pharmacokinetics, stability, and/or tissue specificity.

In some embodiments, an oligonucleotide agent (referred to as “NA” in formula OT-I through OT-IV below, *e.g.*, RNA, DNA, chimeric RNA-DNA, DNA-RNA, RNA-DNA-RNA, or DNA-RNA-DNA) can be chemically modified by conjugating a moiety that includes a ligand having one or more chemical linkages for attachment of the ligand (L) to the oligonucleotide or nucleic acid. The ligand of an oligonucleotide agent can be coupled by one or both of a tether and linker. In the diagram below, exemplary chemical linkages are represented as X, Y, and Z. These can be part of the tether or linker.



- 5 Ligands can be attached at one or both of the 3' end, the 5' end, and internal positions. In certain embodiments, the oligonucleotide agent can be chemically modified by conjugating one or more moieties having formula OT-I. Table 3, shows a variety of conjugates.

Table 3.



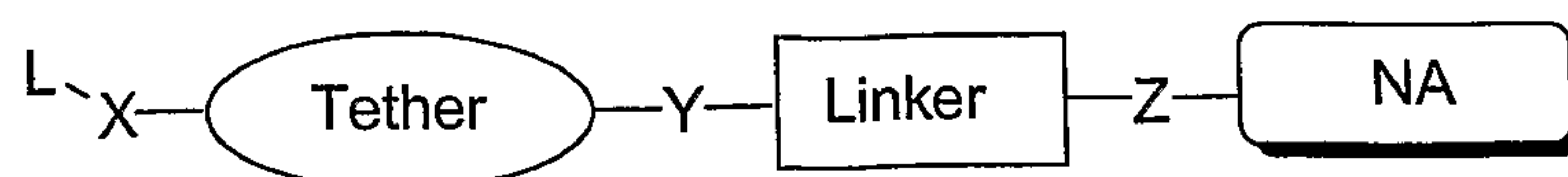
Exemplary ligands are listed in Table 4 and are discussed elsewhere herein. The
 5 exemplary ligands (L) shown in Table 4 are preferred.

Table 4.

L =	Cholesterol Thiocholesterol 5β-Cholanic Acid Cholic acid Lithocholic acid Biotin Vitamin E Naproxen Ibuprofen Amines (mono, di, tri, tetraalkyl or aryl) Folate Sugar (<i>N</i>-Acetylgalactosamine, galactosamine, galactose, Mannose) -(CH₂)_nNQ₁Q₂, where n = 0-40, Q₁, Q₂ = H, Me or Et; Q₁ = H, Q₂ = H, Me, Et or aryl -(CH₂)_pCH=CH(CH₂)_qNQ₁Q₂, where p and/or q = 0-40, Q₁, Q₂ = H, Me or Et; Q₁ = H, Q₂ = H, Me, Et or aryl with E and/or Z configuration -(CH₂)_pCH$\equiv$CH(CH₂)_qNQ₁Q₂, where p and/or q = 0-40, Q₁, Q₂ = H, Me or Et; Q₁ = H, Q₂ = H, Me, Et or aryl -(CH₂)_pCH=CH(CH₂)_qCH=CH(CH₂)_rNQ₁Q₂, where p, q and/or r = 0-40, Q₁, Q₂ = H, Me or Et; Q₁ = H, Q₂ = H, Me, Et or aryl with E and/or Z configuration -O(CH₂)_m(OCH₂CH₂)_n-OR, where m, n = 0-40 and R = H, Me, NQ₁Q₂, -C(O)NR'R'' -C(S)NR'R'' -NH(CH₂)_m(OCH₂CH₂)_n-OR, where m, n = 0-40 and R = H, Me, NQ₁Q₂, -C(O)NR'R'' -C(S)NR'R'' -O(CH₂)_m(NHCH₂CH₂)_n-R, where m, n = 0-40 and R = H, OH, Me, NQ₁Q₂, -C(O)NR'R'' -C(S)NR'R'' -NH(CH₂)_m(NHCH₂CH₂)_n-R, where m, n = 0-40 and R = H, OH, Me, NQ₁Q₂, -C(O)NR'R'' -C(S)NR'R'' Dialkylglycerol (<i>sn</i>3, <i>sn</i>1, <i>sn</i>2 and racemic) with number of methylene varies from 0-40 Diacylglycerol (<i>sn</i>3, <i>sn</i>1, <i>sn</i>2 and racemic) with number of methylene varies from 0-40 Dialkylglycerol (<i>sn</i>3, <i>sn</i>1, <i>sn</i>2 and racemic) with number of methylene varies from 0-40 and the alkyl chain contains one or more double bonds with E and/or Z isomers Diacylglycerol (<i>sn</i>3, <i>sn</i>1, <i>sn</i>2 and racemic) with number of methylene varies from 0-40 and the alkyl chain contains one or more double bonds with E and/or Z isomers Lipids

Exemplary X, Y, and Z moieties are shown in in Table 5. The X, Y, and Z moieties can
 5 be selected independently of one another.

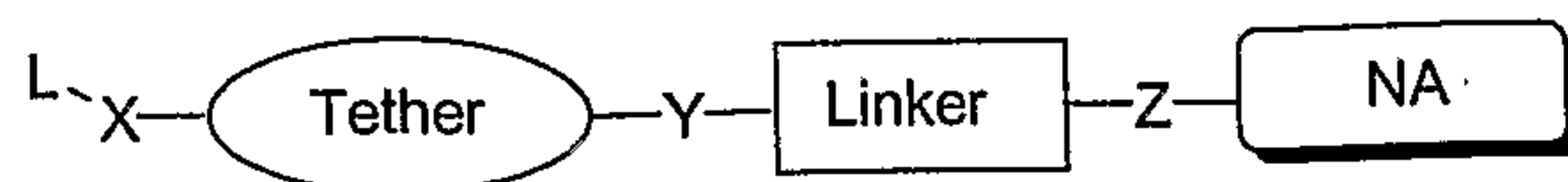
Table 5.

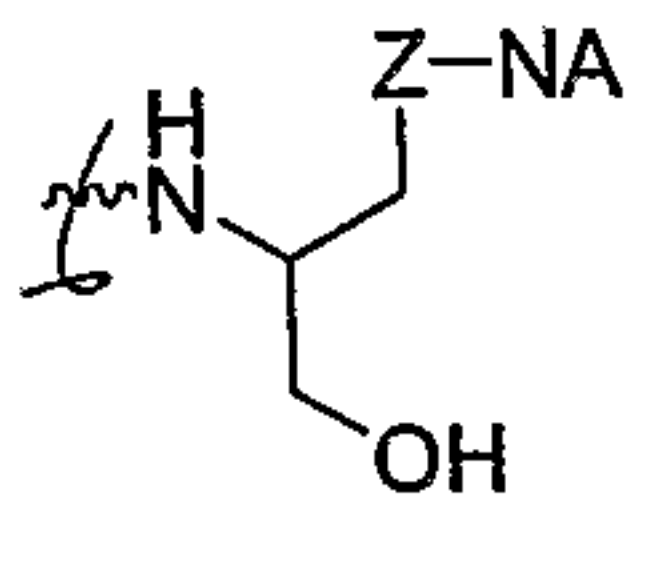
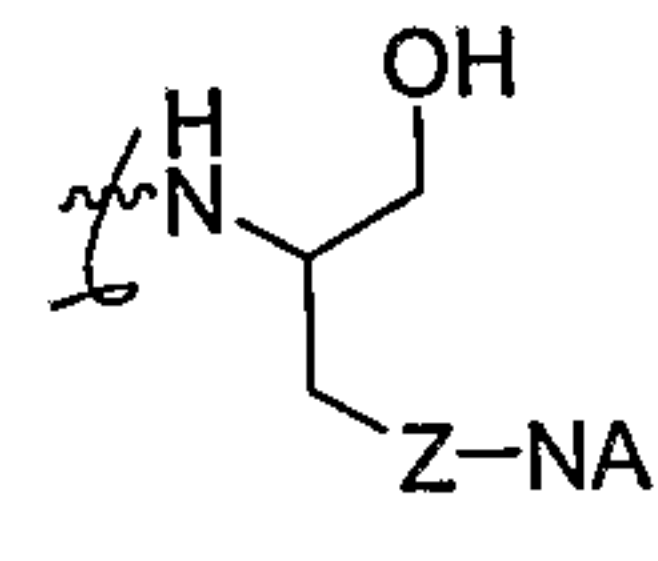
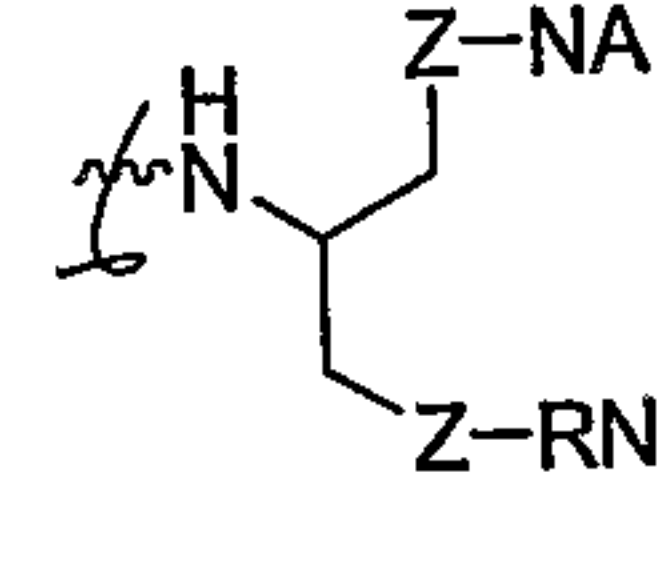
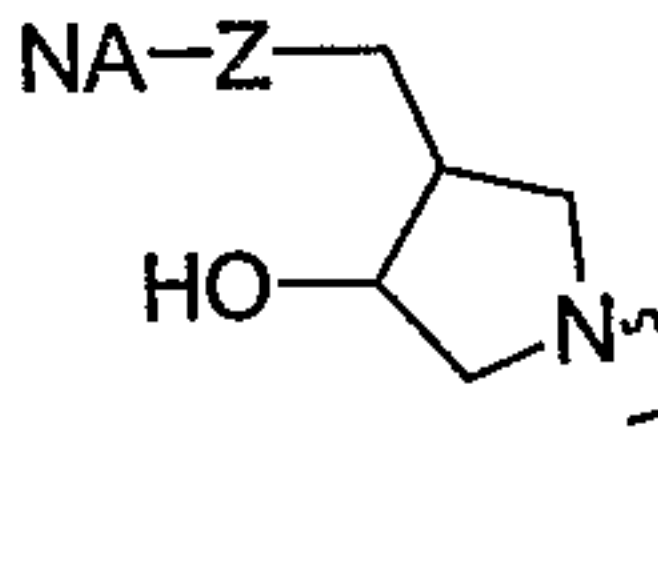
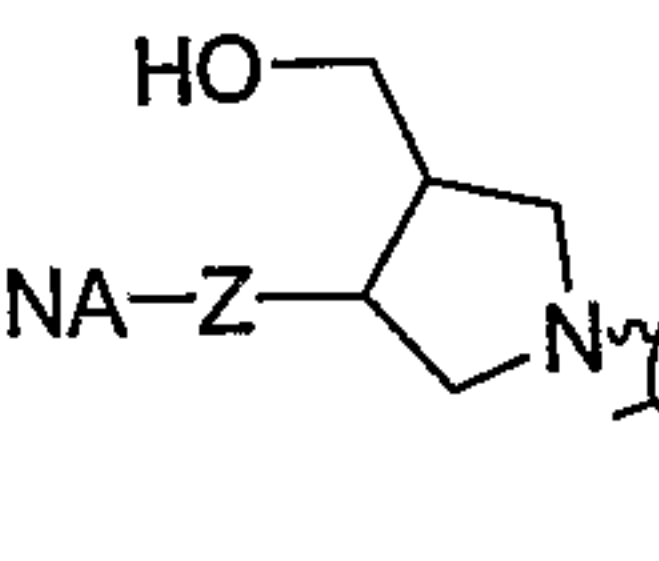
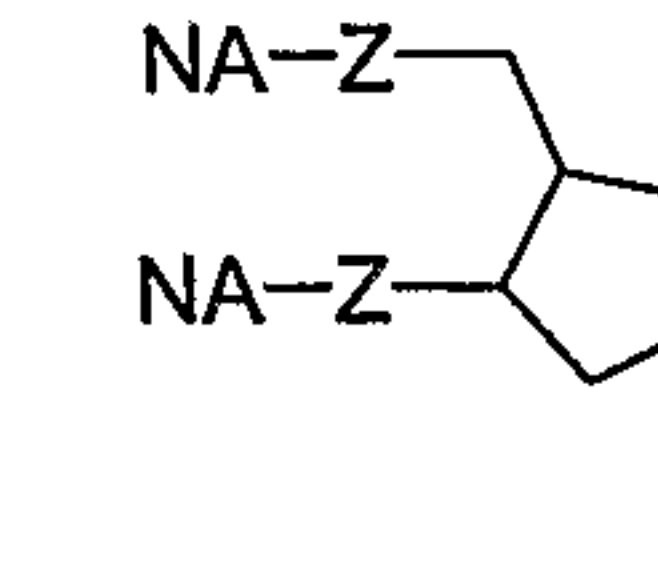
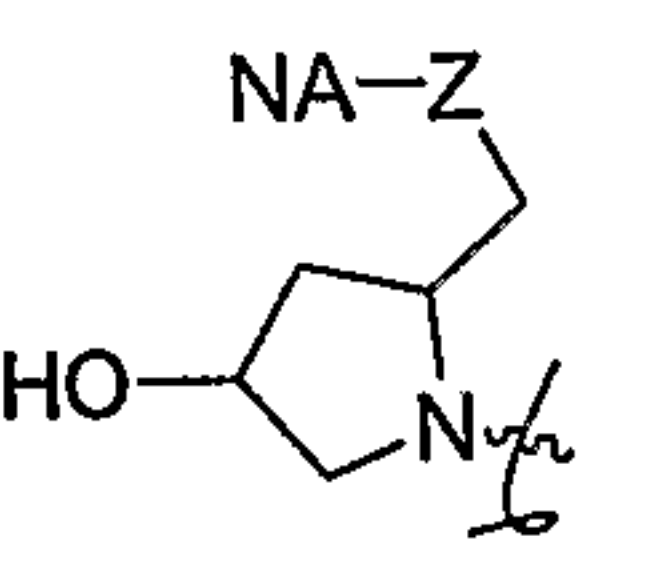
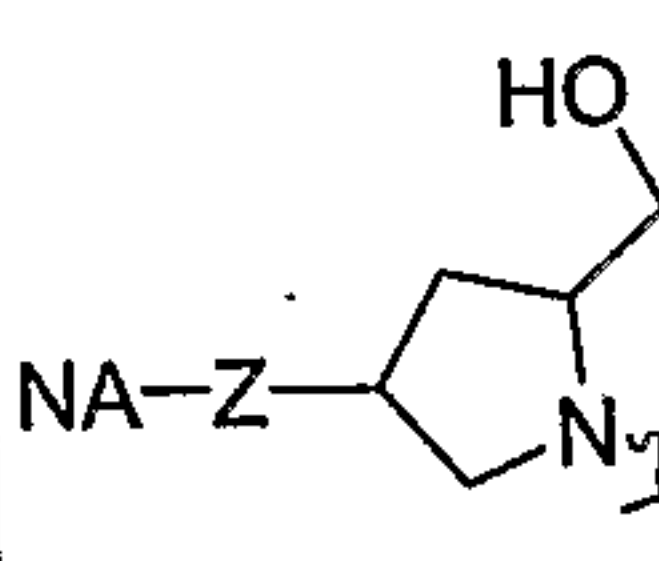
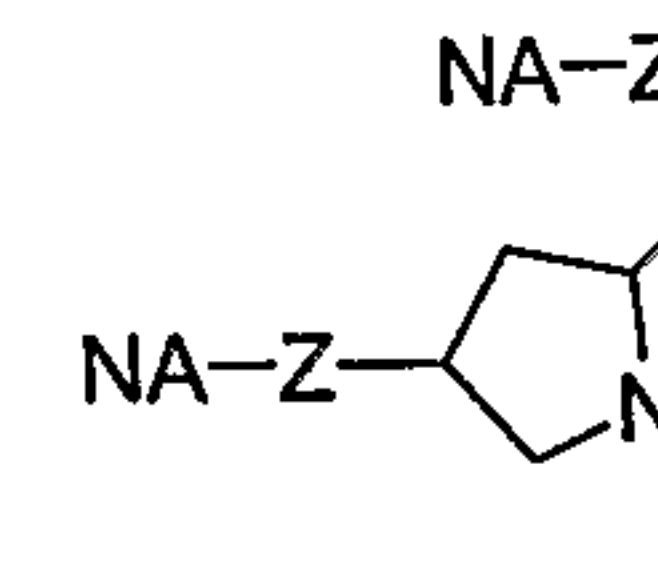
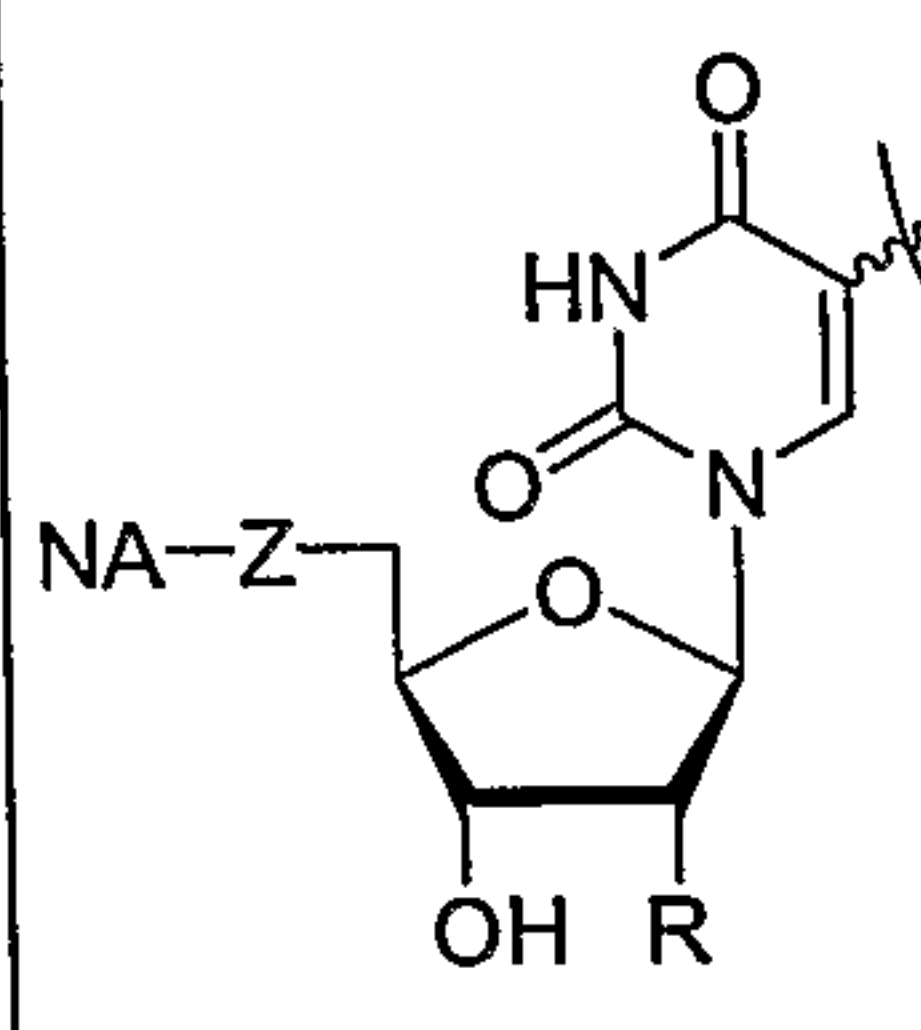
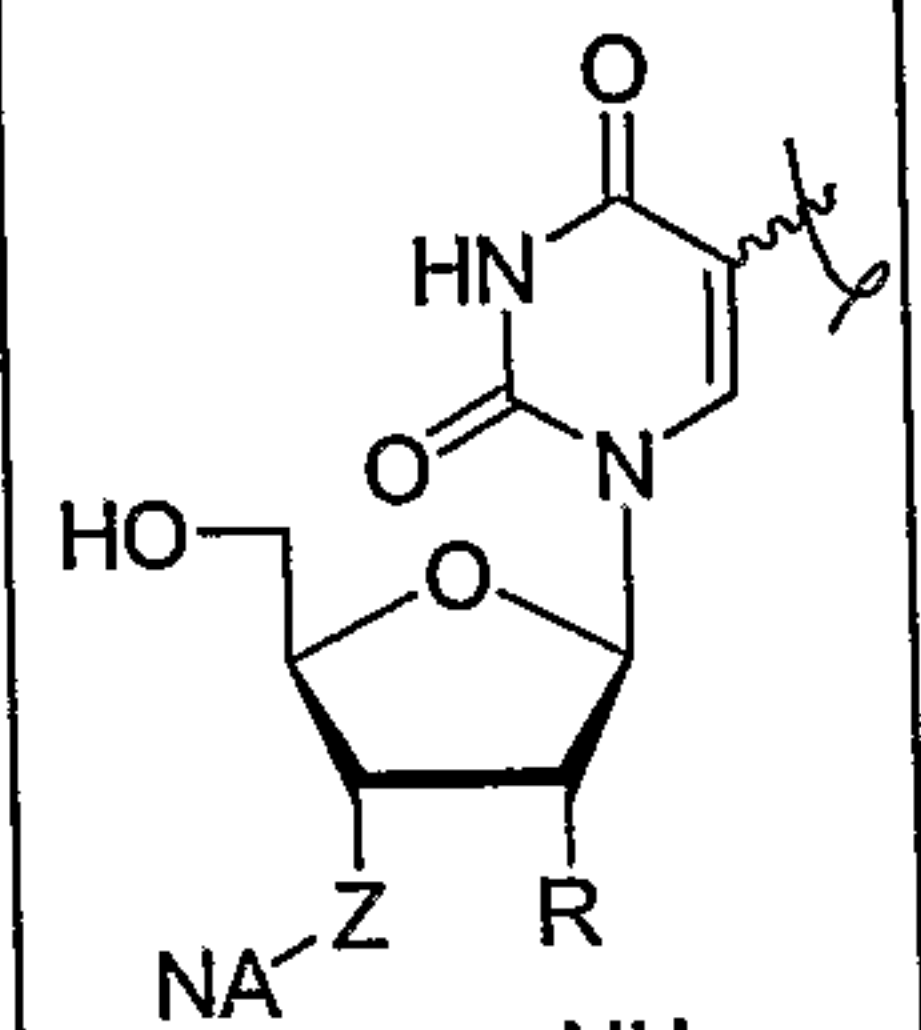
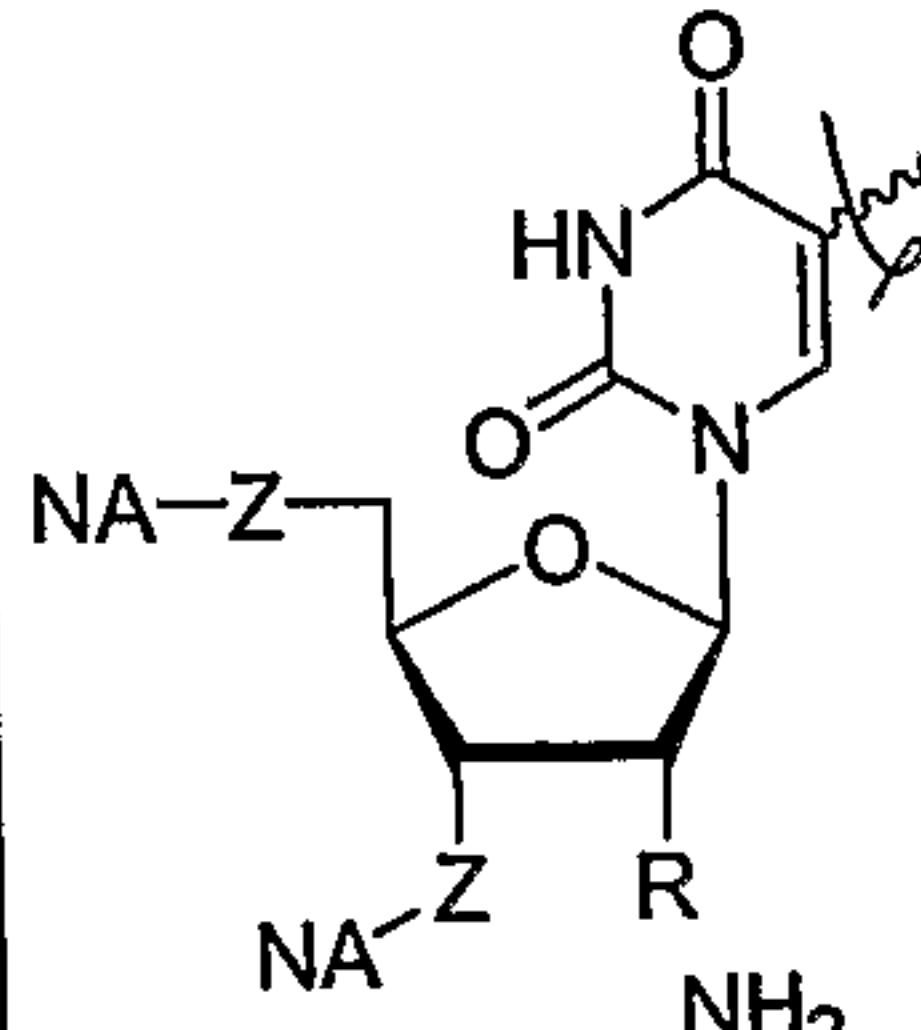
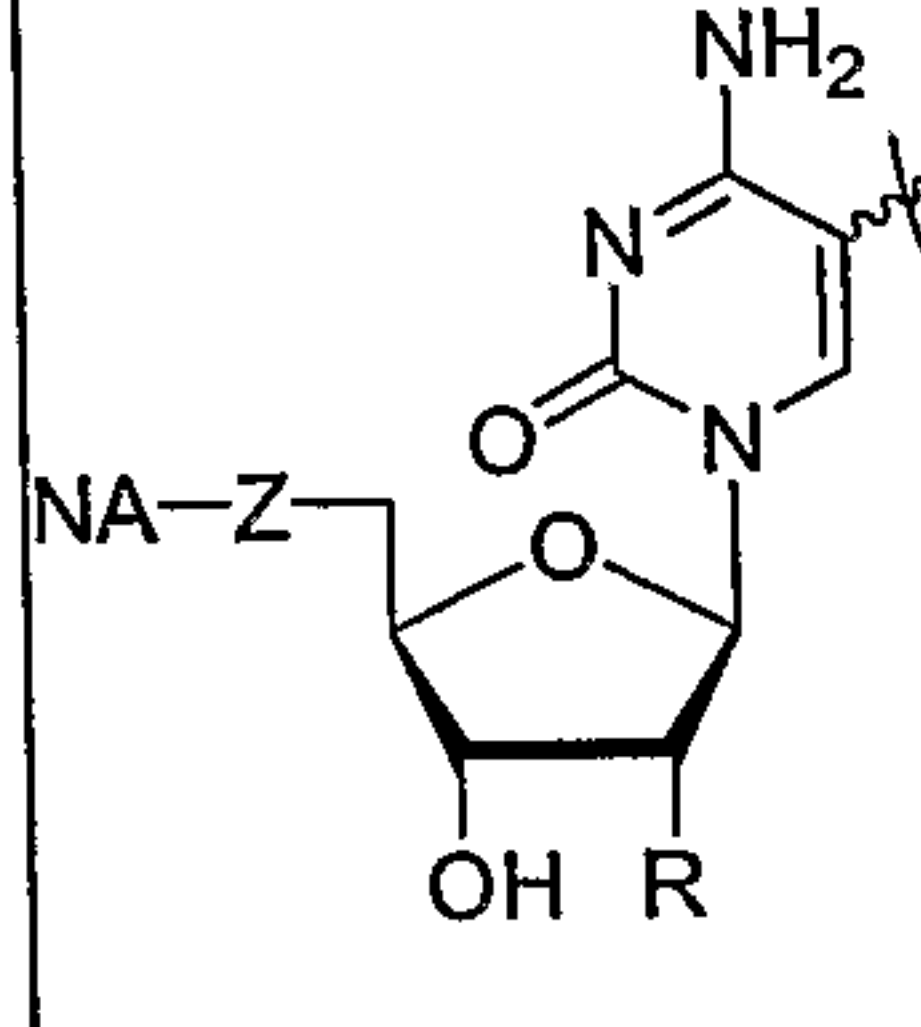
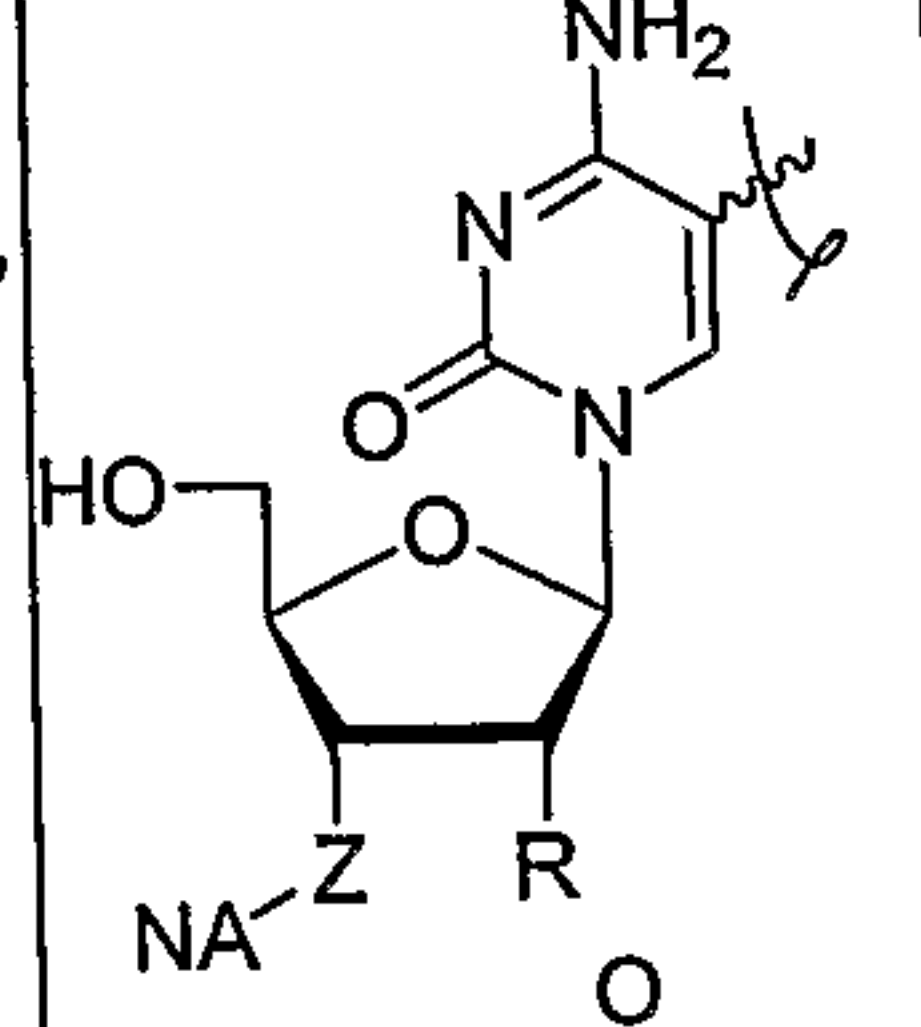
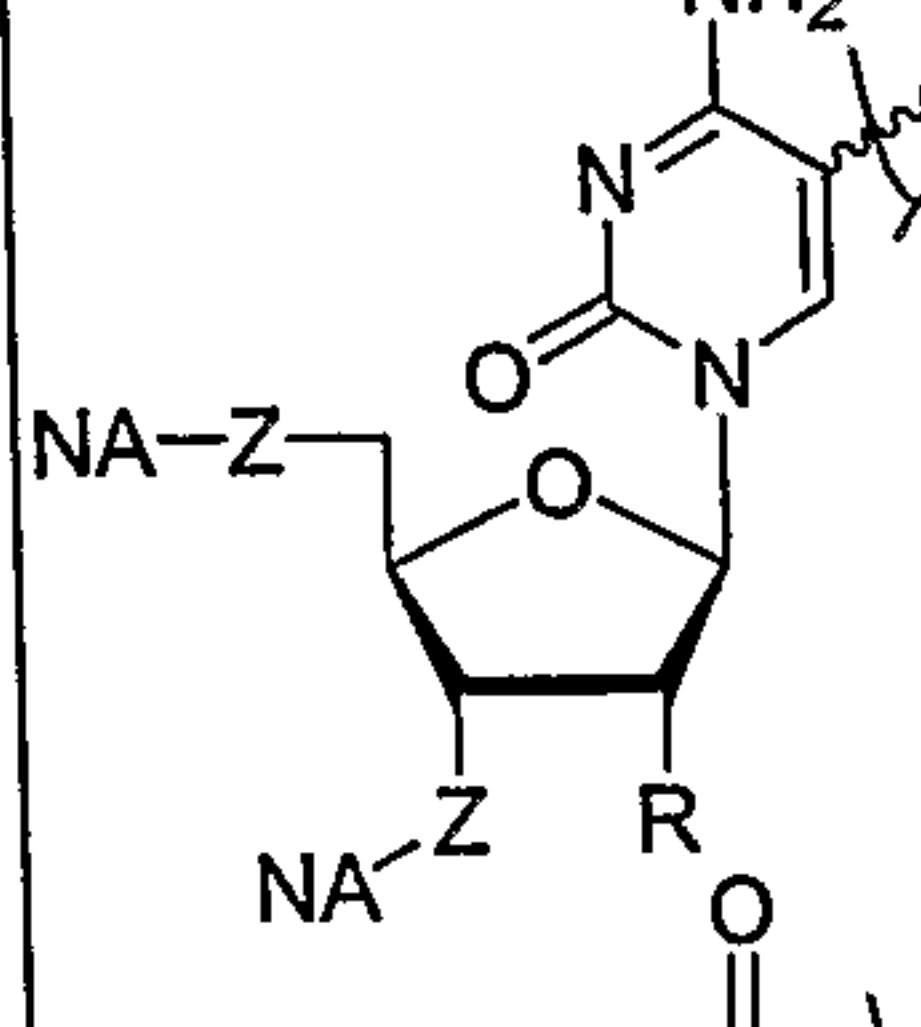
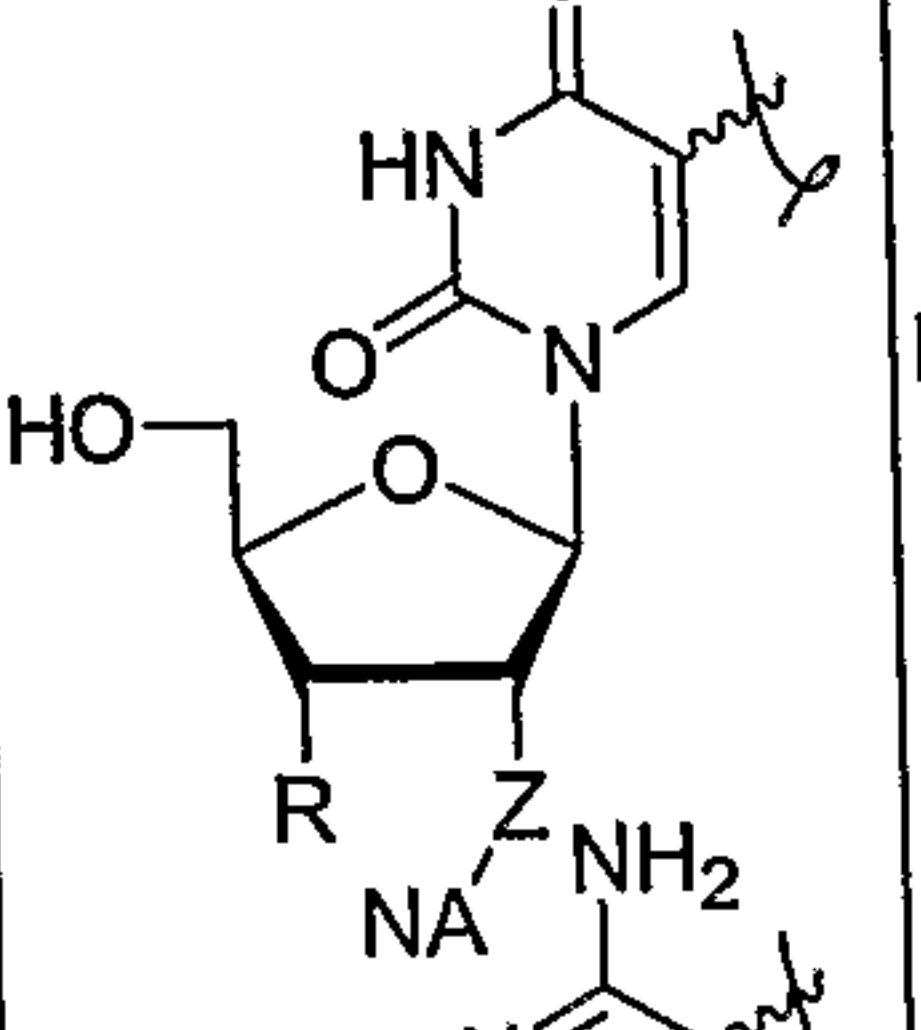
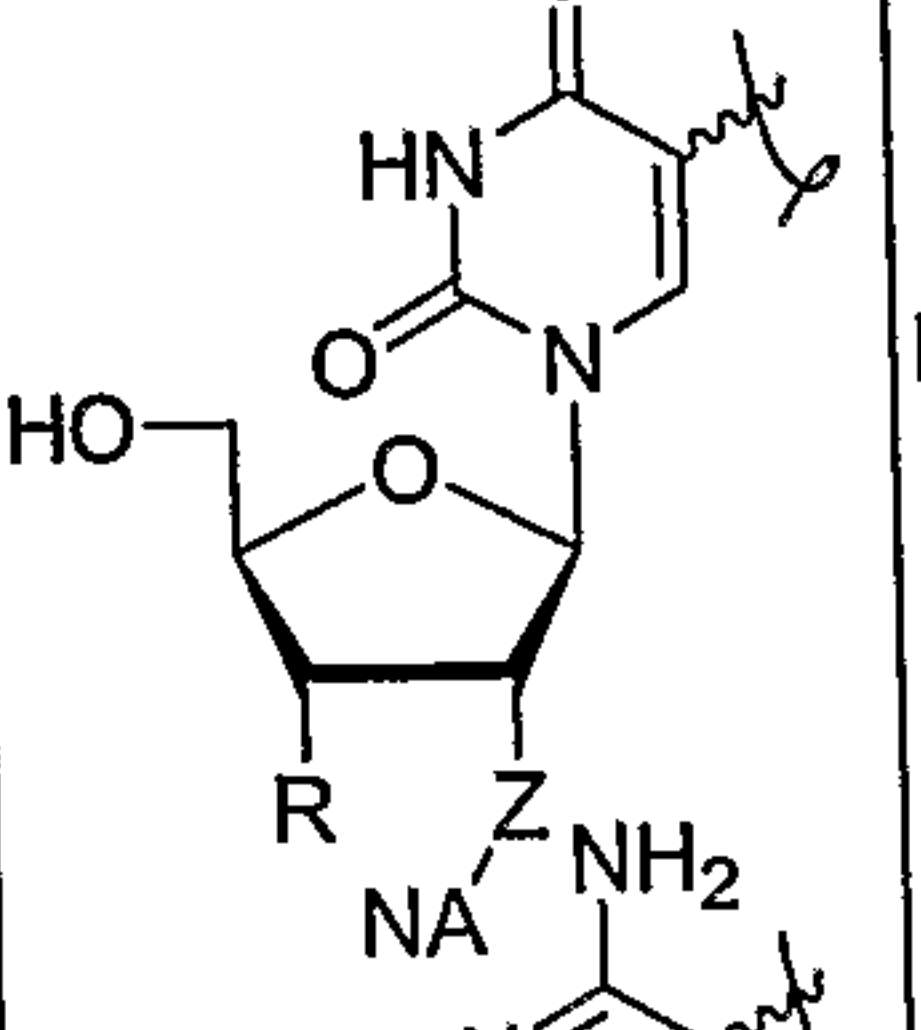
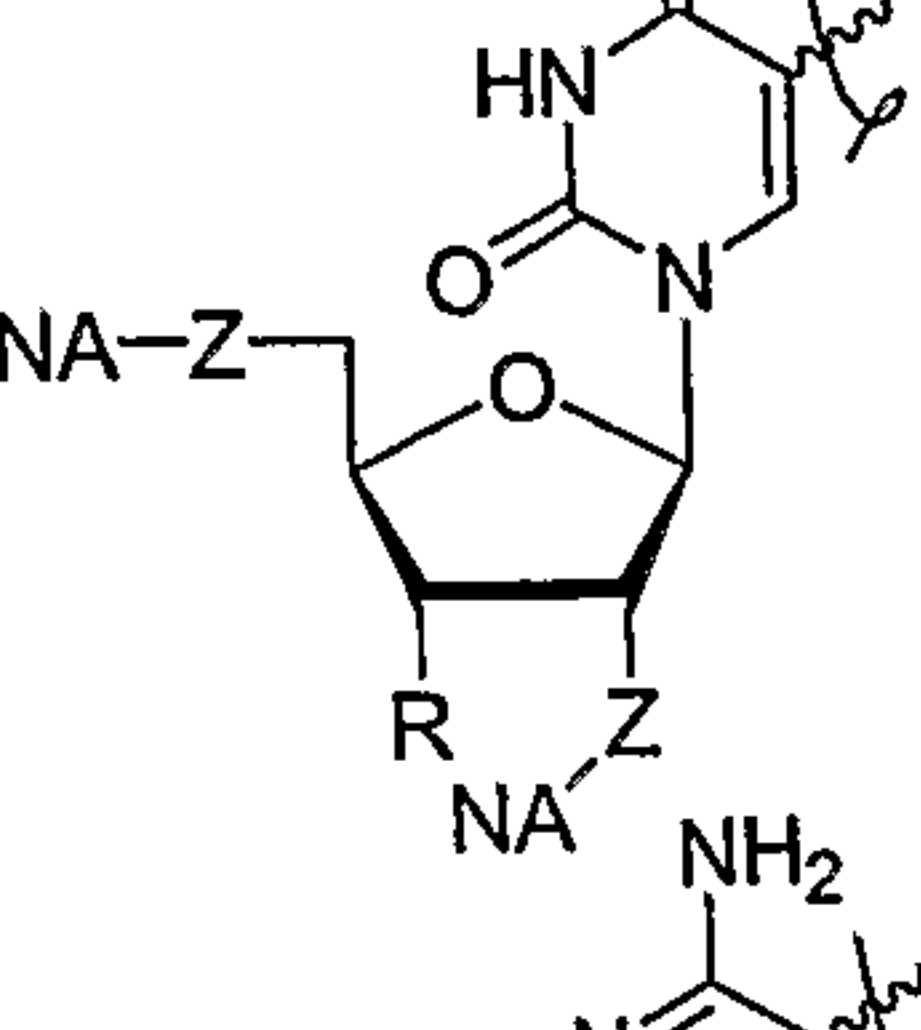
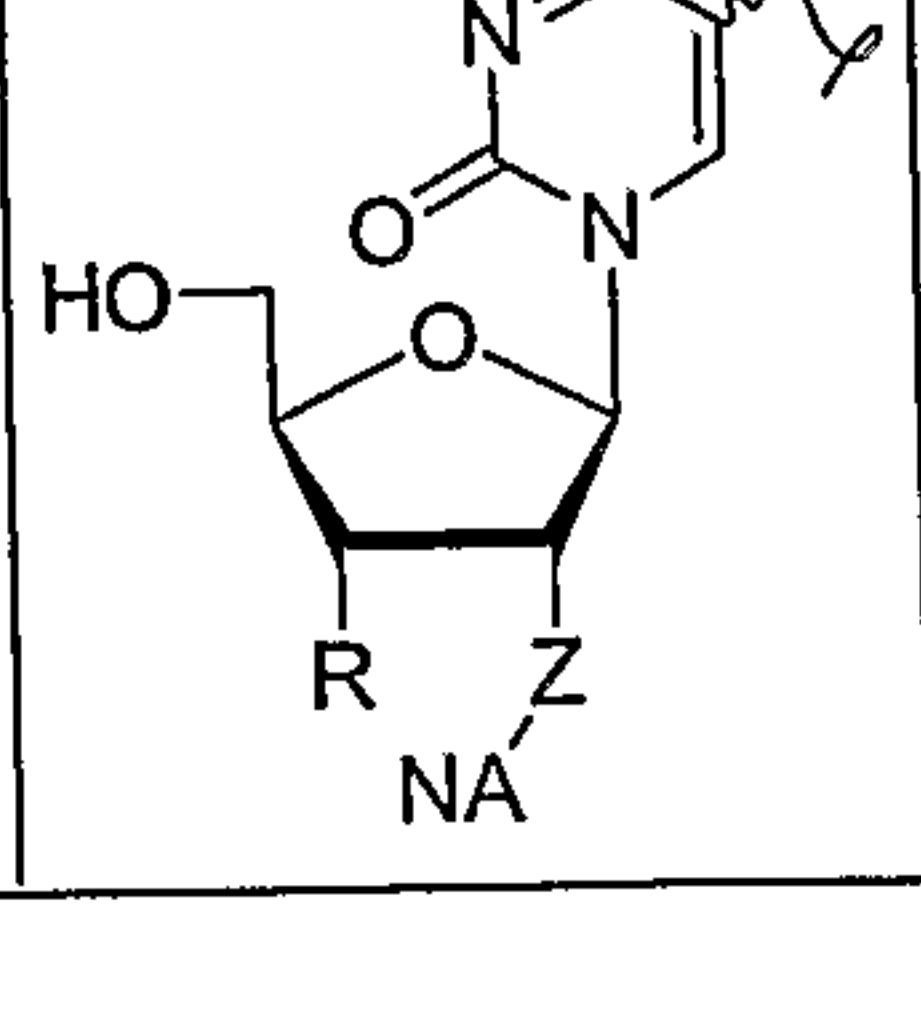
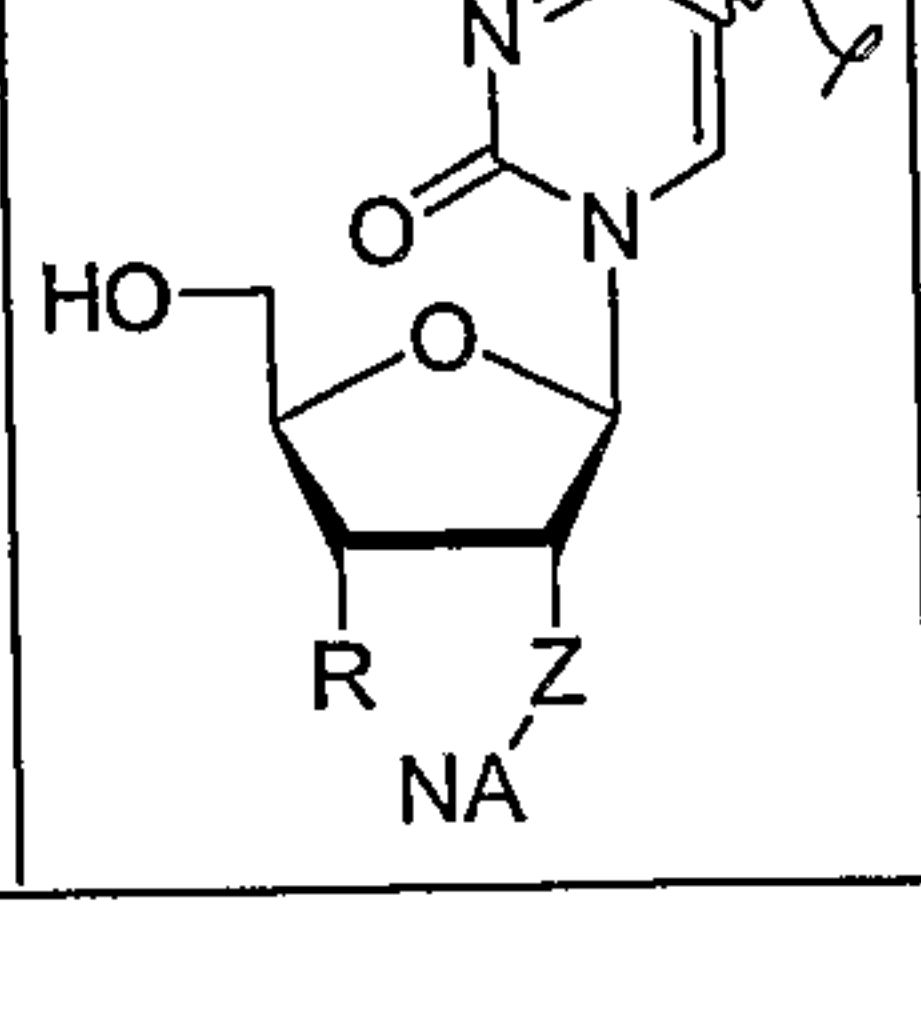
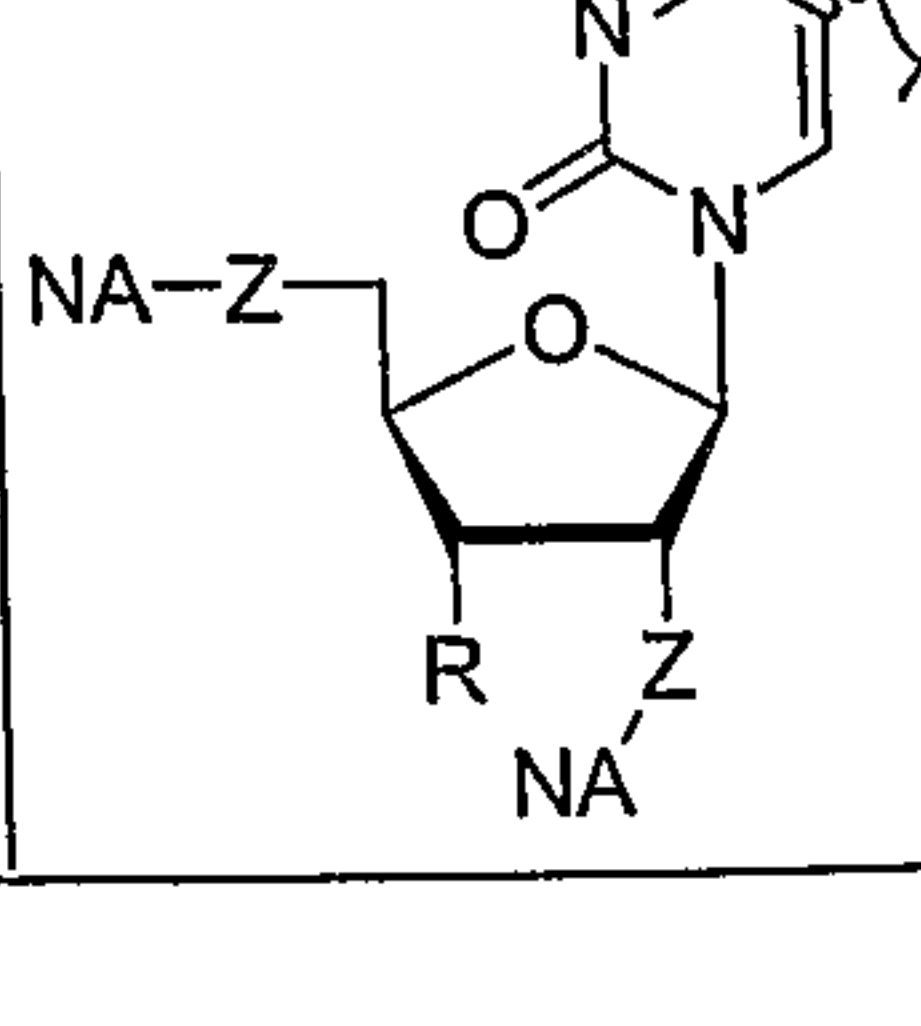


X =	Y =	Z =
-NHC(O)-	-NHC(O)-	-NHC(O)-
-C(O)NH-	-C(O)NH-	-C(O)NH-
-OC(O)NH-	-OC(O)NH-	-OC(O)NH-
-NHC(O)O-	-NHC(O)O-	-NHC(O)O-
-O-	-O-	-O-
-S-	-S-	-S-
-SS-	-SS-	-SS-
-S(O)-	-S(O)-	-S(O)-
-S(O ₂)-	-S(O ₂)-	-S(O ₂)-
-NHC(O)NH-	-NHC(O)NH-	-NHC(O)NH-
-NHC(S)NH-	-NHC(S)NH-	-NHC(S)NH-
-C(O)O-	-C(O)O-	-C(O)O-
-OC(O)-	-OC(O)-	-OC(O)-
-NHC(S)-	-NHC(S)-	-NHC(S)-
-NHC(S)O-	-NHC(S)O-	-NHC(S)O-
-C(S)NH-	-C(S)NH-	-C(S)NH-
-OC(S)NH-	-OC(S)NH-	-OC(S)NH-
-NHC(S)O-	-NHC(S)O-	-NHC(S)O-
-CH ₂ -	-CH ₂ -	-CH ₂ -
-CH ₂ CH=CH-	-CH ₂ CH=CH-	-CH ₂ CH=CH-
-C(O)CH=CH-	-C(O)CH=CH-	-C(O)CH=CH-
-NH-CH ₂ CH=CH-	-NH-CH ₂ CH=CH-	-NH-CH ₂ CH=CH-
-O-P(O)(OH)-O-	-O-P(O)(OH)-O-	-O-P(O)(OH)-O-
-O-P(S)(OH)-O-	-O-P(S)(OH)-O-	-O-P(S)(OH)-O-
-O-P(S)(SH)-O-	-O-P(S)(SH)-O-	-O-P(S)(SH)-O-
-S-P(O)(OH)-O-	-S-P(O)(OH)-O-	-S-P(O)(OH)-O-
-O-P(O)(OH)-S-	-O-P(O)(OH)-S-	-O-P(O)(OH)-S-
-S-P(O)(OH)-S-	-S-P(O)(OH)-S-	-S-P(O)(OH)-S-
-O-P(S)(OH)-S-	-O-P(S)(OH)-S-	-O-P(S)(OH)-S-
-S-P(S)(OH)-O-	-S-P(S)(OH)-O-	-S-P(S)(OH)-O-
-O-P(O)(R)-O-	-O-P(O)(R)-O-	-O-P(O)(R)-O-
-O-P(S)(R)-O-	-O-P(S)(R)-O-	-O-P(S)(R)-O-
-S-P(O)(R)-O-	-S-P(O)(R)-O-	-S-P(O)(R)-O-
-S-P(S)(R)-O-	-S-P(S)(R)-O-	-S-P(S)(R)-O-
-S-P(O)(R)-S-	-S-P(O)(R)-S-	-S-P(O)(R)-S-
-O-P(S)(R)-S-	-O-P(S)(R)-S-	-O-P(S)(R)-S-

R = Alkyl, fluoroalkyl, aryl or aralkyl

Exemplary tethers are shown in Table 7.

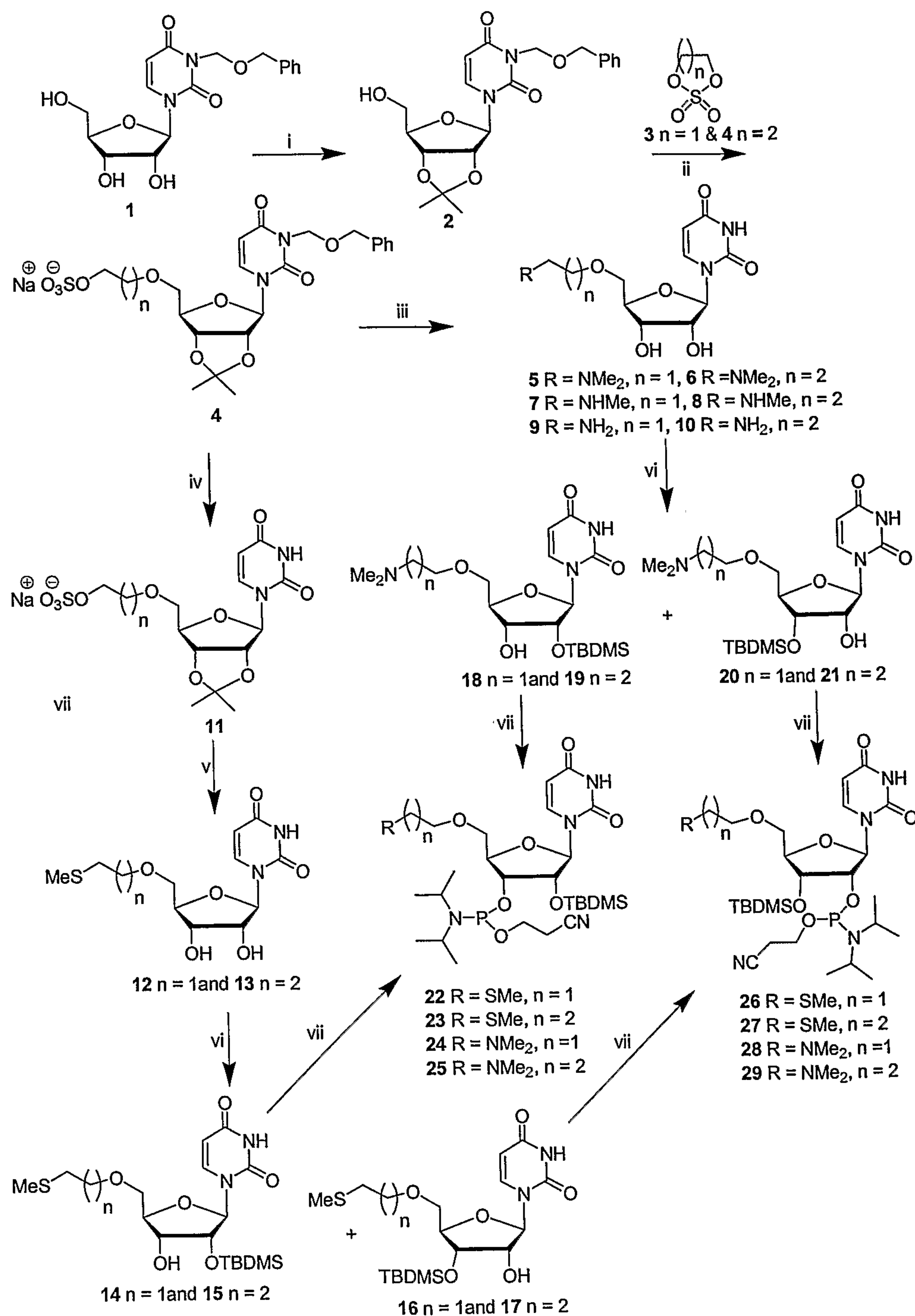


Tether:	Linker =		
	3'-end	5'-end	interior
-(CH ₂) _n -, where n = 1-40 -(CH ₂ -CH ₂ O) _n -, where n = 1-20 -O(CH ₂ -CH ₂ O) _n -, where n = 1-20 -(CH ₂ -CH ₂ NH) _n -, where n = 1-20 -NH(CH ₂ -CH ₂ NH) _n -, where n = 1-20 -(CH ₂) _l [(CH=CH) _m (CH ₂) _n] _p (CH=CH) _q (CH ₂) _r -, where l, m, n, p, q and/or r = 0-20 -(CH ₂) _l [(C≡C) _m (CH ₂) _n] _p (C≡C) _q (CH ₂) _r -, where l, m, n, p, q and/or r = 0-20			
			
			
			
			
			
			
			
			

Compounds described herein can be prepared by methods described herein or by conventional methods from commercially available reagents and starting materials.

5

Compound 1 is prepared as reported by Fraser *et al.* (Tetrahedron Lett. 41:1523, 2000). Steps (ii), (iii) (a), (iii) (c), (iv), (v) and (vii) are performed according to literature procedure (Fraser *et al.*, Tetrahedron Lett. 41:1523, 2000). Step (iii) (b) and (v) (b) are performed as reported in the literature (Bioorg. Med. Chem. Lett. 13:1713, 2003). Step (iv) is performed as
10 reported in the literature (Corey and Venkateswarlu, J. Am. Chem. Soc. 94:6190, 1972).

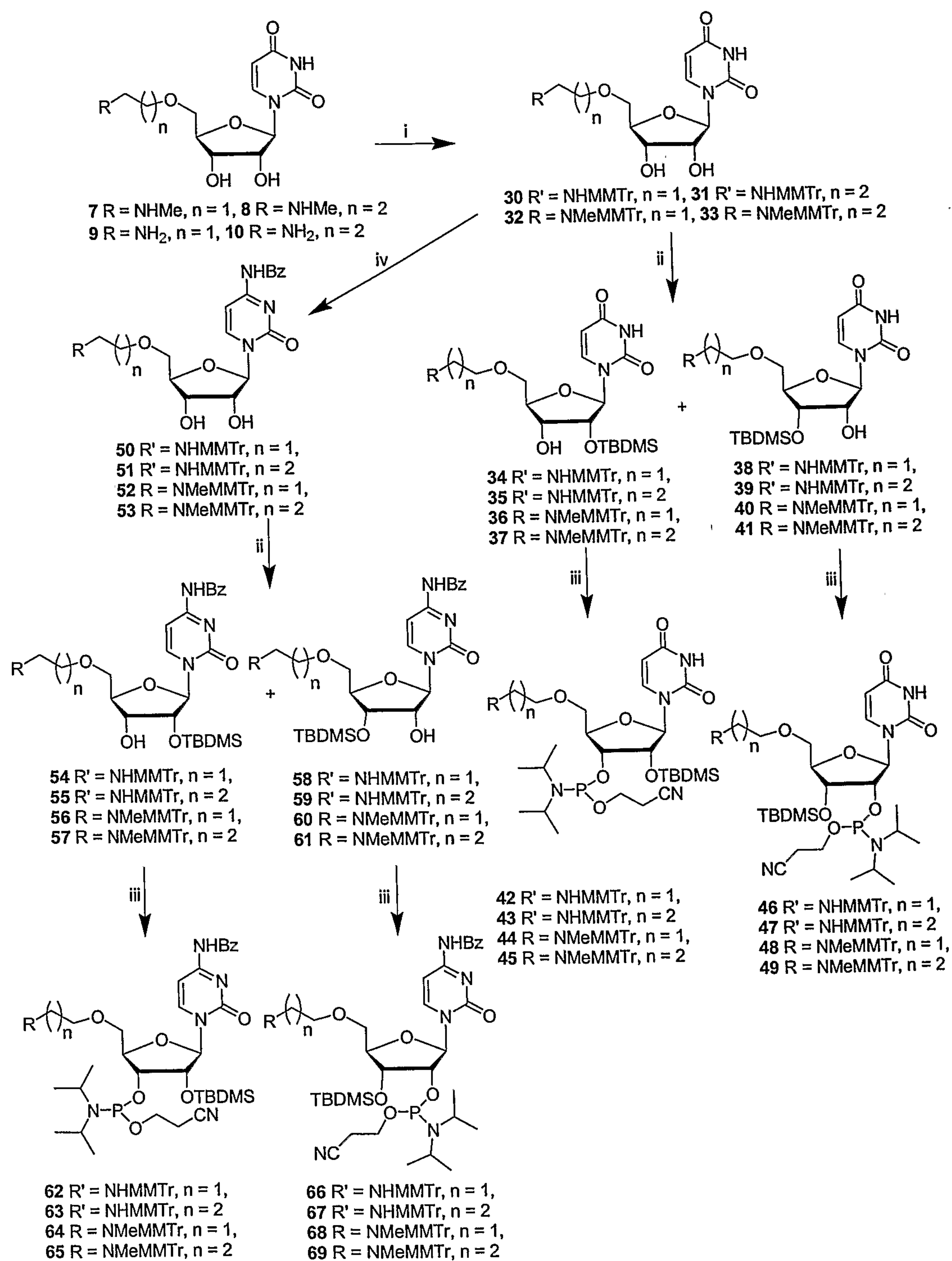
Scheme 1^a

^a (i) 2,2-Di-O-methylpropane, PTSA; (ii) NaH/DMF, 3 or 4, -45 °C to rt; (iii) (a) NH₃, NH₂Me or NHMe₂,

5 THF, autoclave, (b) HCOOH-H₂O and (c) Pd(OH), EtOH, AcOH, H₂ at 55 psi; (iv) Pd(OH), EtOH, AcOH, H₂ at 55

psi; (v) NaSMe/DMF, 80 °C and (b) HCOOH-H₂O (vi) TBDMS-Cl, Imidazole/Py; (vii) diisopropylamine tetrazolide, 2-cyanoethyl-*N,N,N',N'*-tetraisopropylphosphoramidite/CH₂Cl₂

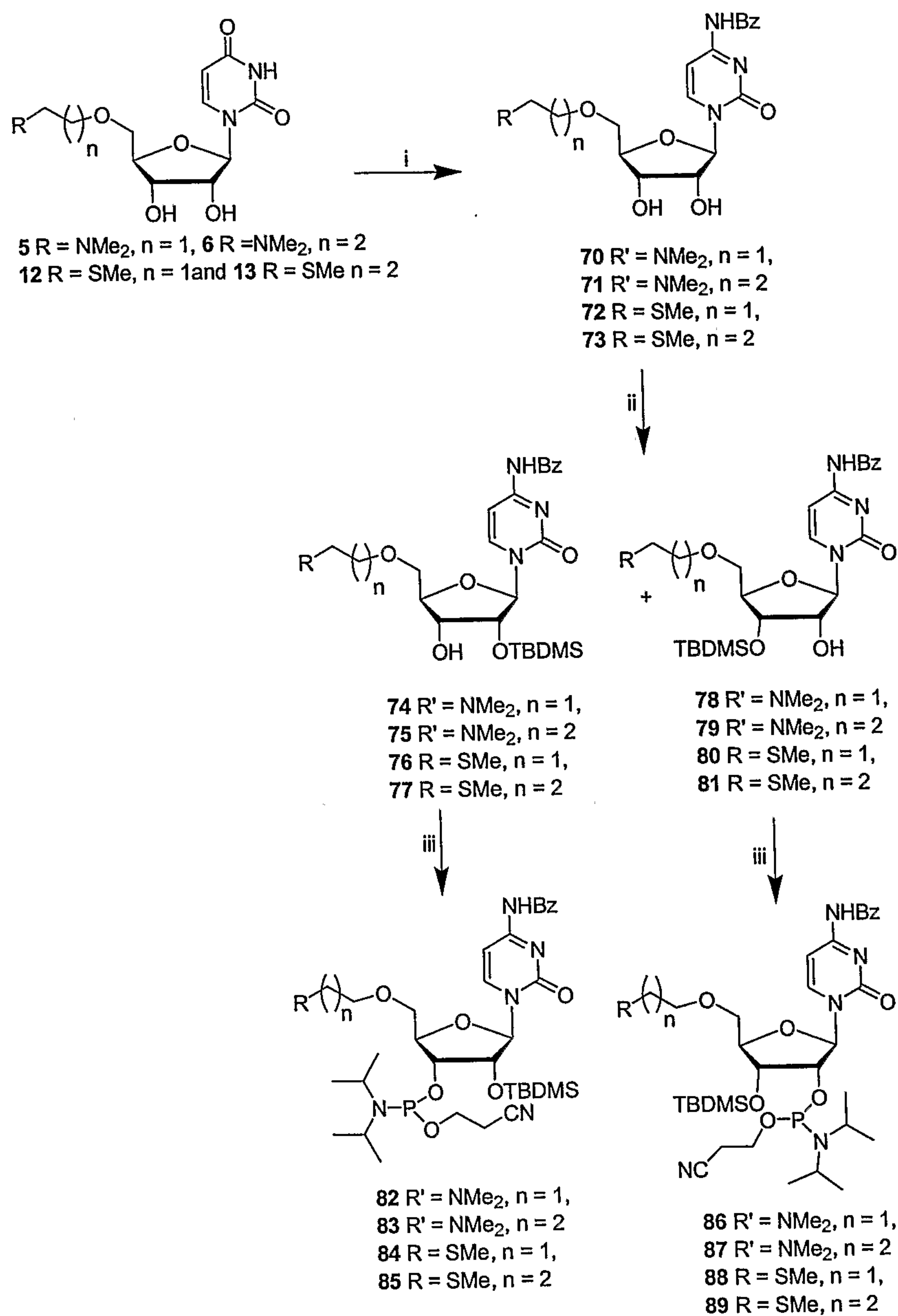
The synthesis of certain compounds is described in scheme 2, below. Step (i) is performed as reported in Dubowchik and Radia (Tetrahedron Lett., 38:5257, 1997); step (ii) is
5 performed as reported in Corey and Venkateswarlu (J. Am. Chem. Soc. 94:6190, 1972); step (iii) is performed as reported in Fraser *et al.* (Tetrahedron Lett. 41:1523, 2000) and step (iv) is performed as described in Miller *et al.* (Current Protocol in Nucleic Acids Chemistry, 2000, 2.5.1-2.5.36, John Wiley and Sons, Inc.).

Scheme 2^a

^a (i) MMTTr-Cl, TEA/CH₂Cl₂; (ii) TBDMS-Cl, Imidazole/Py; (iii) diisopropylamine tetrazolide, 2-cyanoethyl-*N,N,N',N'*-tetraisopropylphosphoramidite/CH₂Cl₂; (iv) (a) Ac₂O/Py, (b) Triazole, TEA, 4-chlorophenyl dichlorophosphate/MeCN, (c) NH₄OH and (d) Pentafluorophenyl benzoate/Py

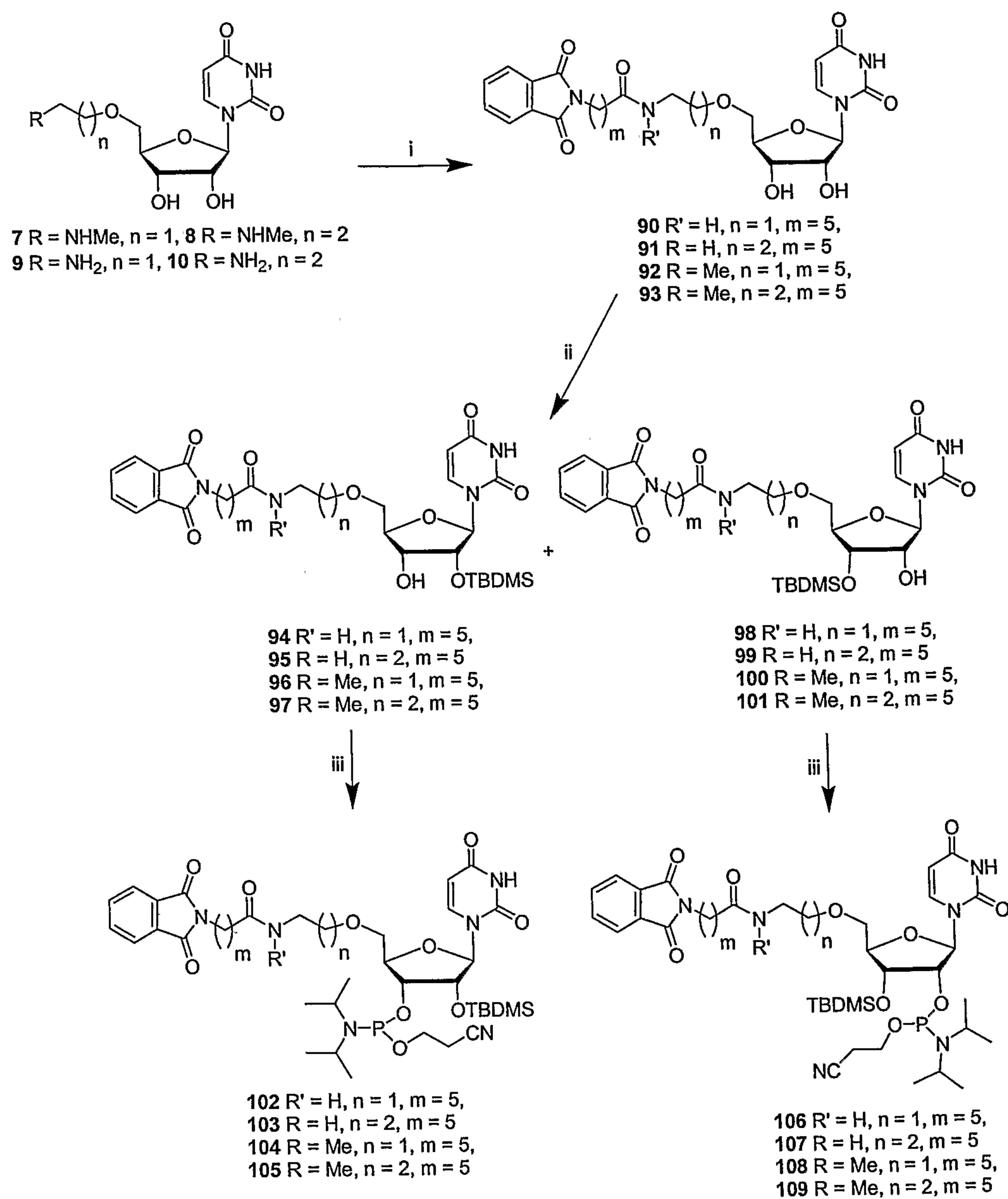
5 The synthesis of certain compounds is performed as described in Scheme 3, below. Step (i) is performed as described in Miller *et al.* (Current Protocol in Nucleic Acids Chemistry, 2000, 2.5.1-2.5.36, John Wiley and Sons, Inc.); step (ii) is performed as reported in the Corey and Venkateswarlu (J. Am. Chem. Soc. 94:6190, 1972) and step (iii) is performed as reported by Fraser *et al.* (Tetrahedron Lett. 41:1523, 2000).

10

Scheme 3^a

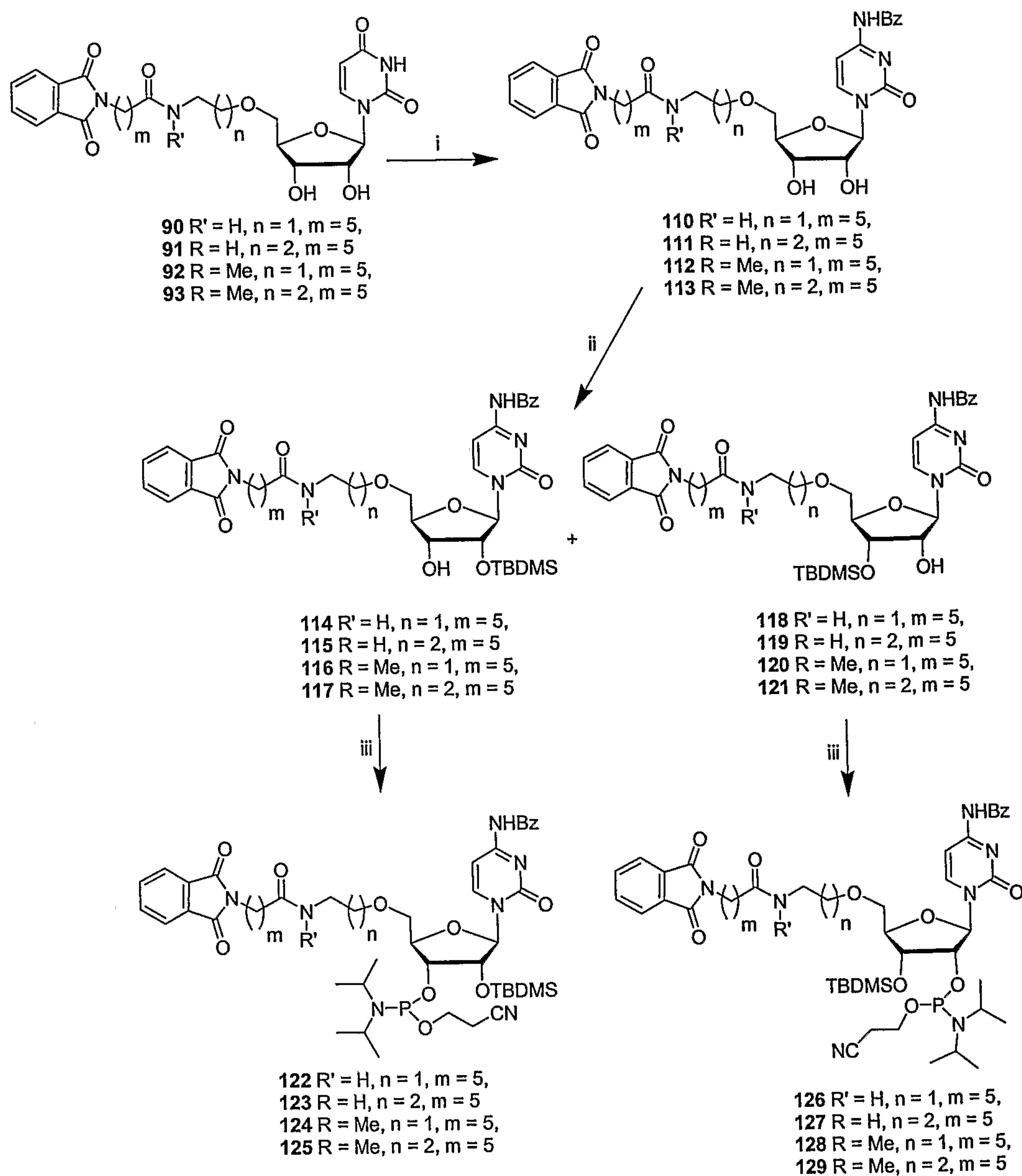
- 5 ^a (i) (a) Ac₂O/Py, (b) Triazole, TEA, 4-chlorophenyl dichlorophosphate/MeCN, (c) NH₄OH and (d) Pentafluorophenyl benzoate/Py (ii) TBDMS-Cl, Imidazole/Py; (iii) diisopropylamine tetrazolide, 2-cyanoethyl-*N,N,N',N'*-tetraisopropylphosphoramidite/CH₂Cl₂

The synthesis of certain compounds is performed as described in Scheme 4 below. Step (ii) is performed as reported in Corey and Venkateswarlu (J. Am. Chem. Soc. 94:6190, 1972) and step (iii) is performed as reported by Fraser *et al.* (Tetrahedron Lett. 41:1523, 2000).

Scheme 4^a

^a (i) *N*-Phthalimido-6-aminocaproic acid, DCC, DMAP, HOBT; (ii) TBDMS-Cl, Imidazole/Py; (iii) diisopropylamine tetrazolide, 2-cyanoethyl-*N,N,N',N'*-tetraisopropylphosphoramidite/CH₂Cl₂

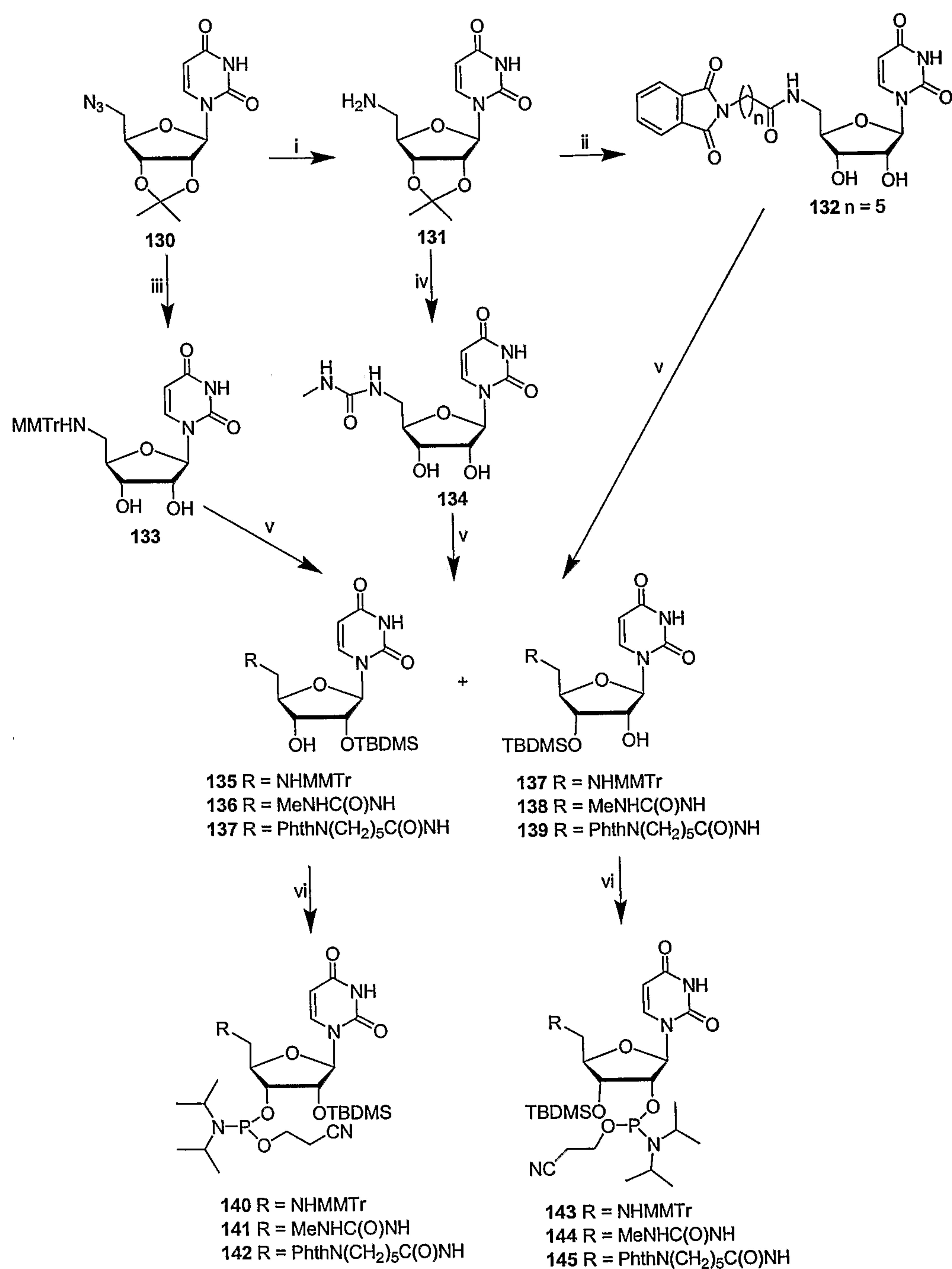
The synthesis of certain compounds is described in Scheme 5, below. Step (i) is performed as described in Miller *et al.* (Current Protocol in Nucleic Acids Chemistry, 2000, 2.5.1-2.5.36, John Wiley and Sons, Inc.); step (ii) is performed as described in Corey and Venkateswarlu (J. Am. Chem. Soc. 94:6190, 1972) and step (iii) is performed as reported by
5 Fraser *et al.* (Tetrahedron Lett. 41:1523, 2000).

Scheme 5^a

5 ^a (i) (a) Ac_2O/Py , (b) Triazole, TEA, 4-chlorophenyl dichlorophosphate/MeCN, (c) NH_4OH and (d) Pentafluorophenyl benzoate/Py (ii) TBDMS-Cl, Imidazole/Py; (iii) diisopropylamine tetrazolide, 2-cyanoethyl- N,N,N',N' -tetraisopropylphosphoramidite/ CH_2Cl_2

10 The synthesis of certain compounds is described in Scheme 6, below. Compound 130, shown in Scheme 6, is obtained as reported in Liu and Austin, J. Org. Chem. 66:8643, 2001).

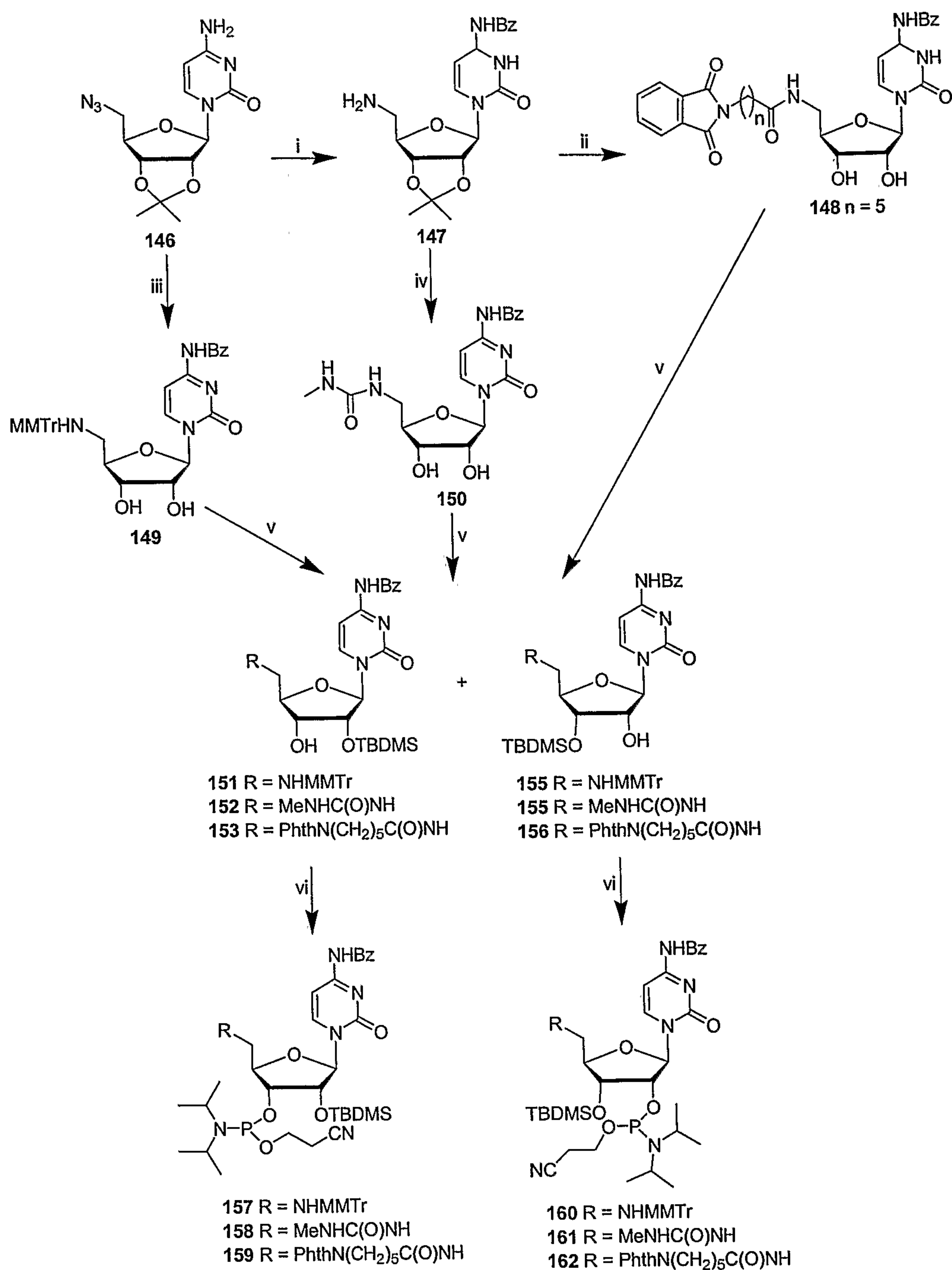
Step (i) and (iii) (b) are performed as reported in the literature (Chem. Rev., 1954, 54, 1); step (ii) (a) is performed according to literature procedures (J. Org. Chem., 1993, 58, 2334); step (ii) (b), (iii) (a) and (iv) (b) are performed as reported in the literature (Bioorg. Med. Chem. Lett., 2003, 13, 1713); step (iii) (c) is performed as reported in Dubowchik and Radia (Tetrahedron Lett. 38:5257, 1997); step (iv) (a) is performed as reported in the literature (Organic Lett., 2001, 3, 1809); step (v) is performed as reported in Corey and Venkateswarlu (J. Am. Chem. Soc. 94:6190, 1972) and step (vi) is performed as reported by Fraser *et al.* (Tetrahedron Lett. 41:1523, 2000).

Scheme 6^a

- 5 ^a (i) H₂, Pd-C (10 %)/MeOH 1 atm; (ii) (a) *N*-Phthalimido-6-aminocaproic acid, DCC, DMAP, HOBT and (b) HCOOH-H₂O; (iii) (a) HCOOH-H₂O, (b) H₂, Pd-C (10 %)/MeOH 1 atm and (c) MMTTr-Cl, TEA/CH₂Cl₂; (iv) (a) CDI (carbonyldiimidazole)/THF, MeNH₂ or *p*-Nitrophenylchloroformate, DMAP/Py, MeNH₂ and (b) HCOOH-H₂O

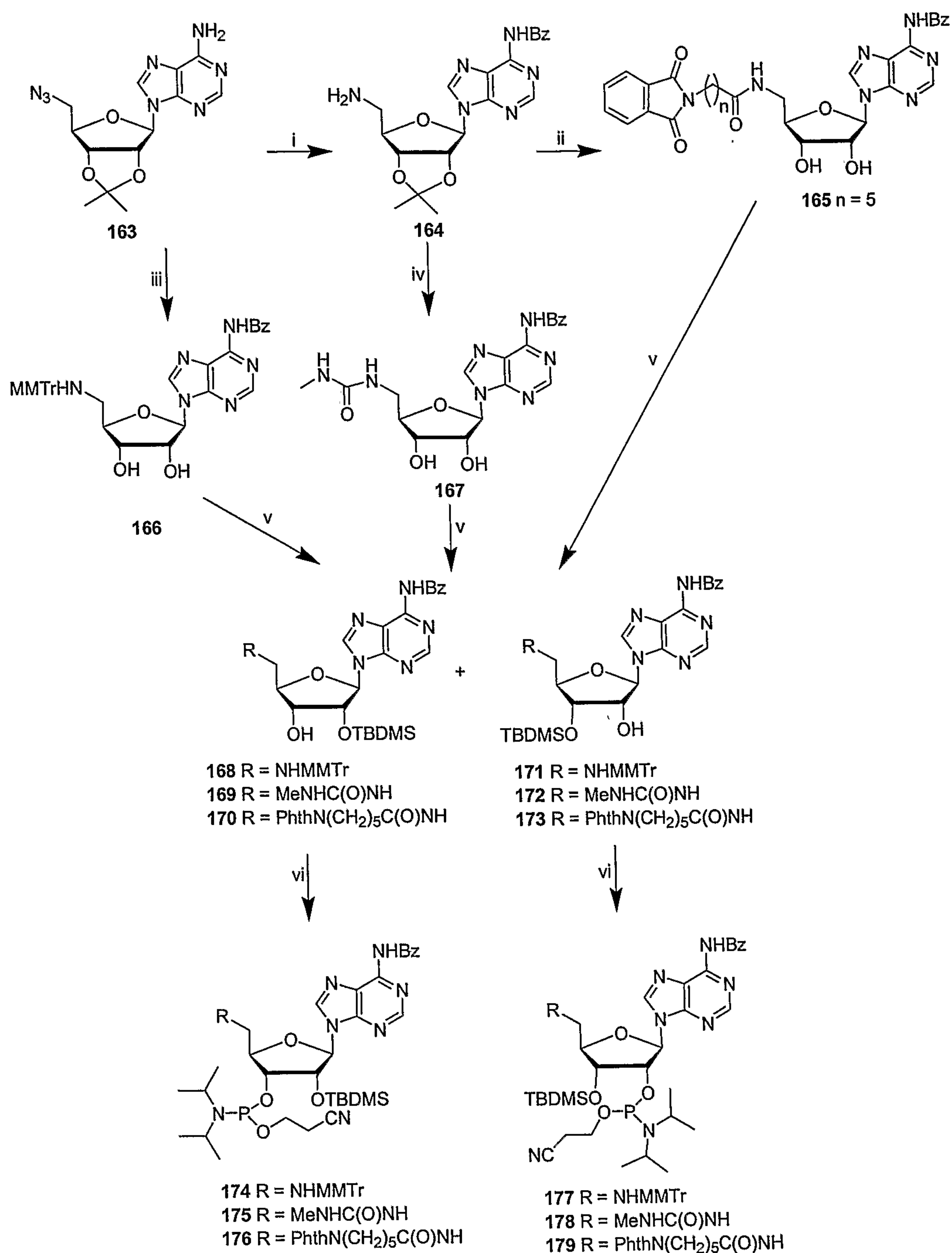
(v) TBDMS-Cl, Imidazole/Py; (vi) diisopropylamine tetrazolide, 2-cyanoethyl-*N,N,N',N'*-tetraisopropylphosphoramidite/ CH_2Cl_2

5 The synthesis of certain compounds is described in Scheme 7, below. Compound 146 is
obtained as reported in Liu and Austin (J. Org. Chem., 2001, 66, 8643). Step (i) (b) and (iii) (c)
are performed as reported in the literature (Chem. Rev., 1954, 54, 1); step (ii) (a) is performed
according to literature procedures (J. Org. Chem., 1993, 58, 2334); step (ii) (b), (iii) (b) and (iv)
(b) are performed as reported in the literature (Bioorg. Med. Chem. Lett., 2003, 13, 1713); step
10 (iii) (d) is performed as reported in Dubowchik and Radia (Tetrahedron Lett., 1997, 38, 5257);
step (iv) (a) is performed as reported in the literature (Organic Lett., 2001, 3, 1809); step (v) is
performed as reported in Corey and Venkateswarlu (J. Am. Chem. Soc., 1972, 94, 6190) and step
(vi) is performed as reported by Fraser *et al.* (Tetrahedron Lett., 2000, 41, 1523)

Scheme 7^a

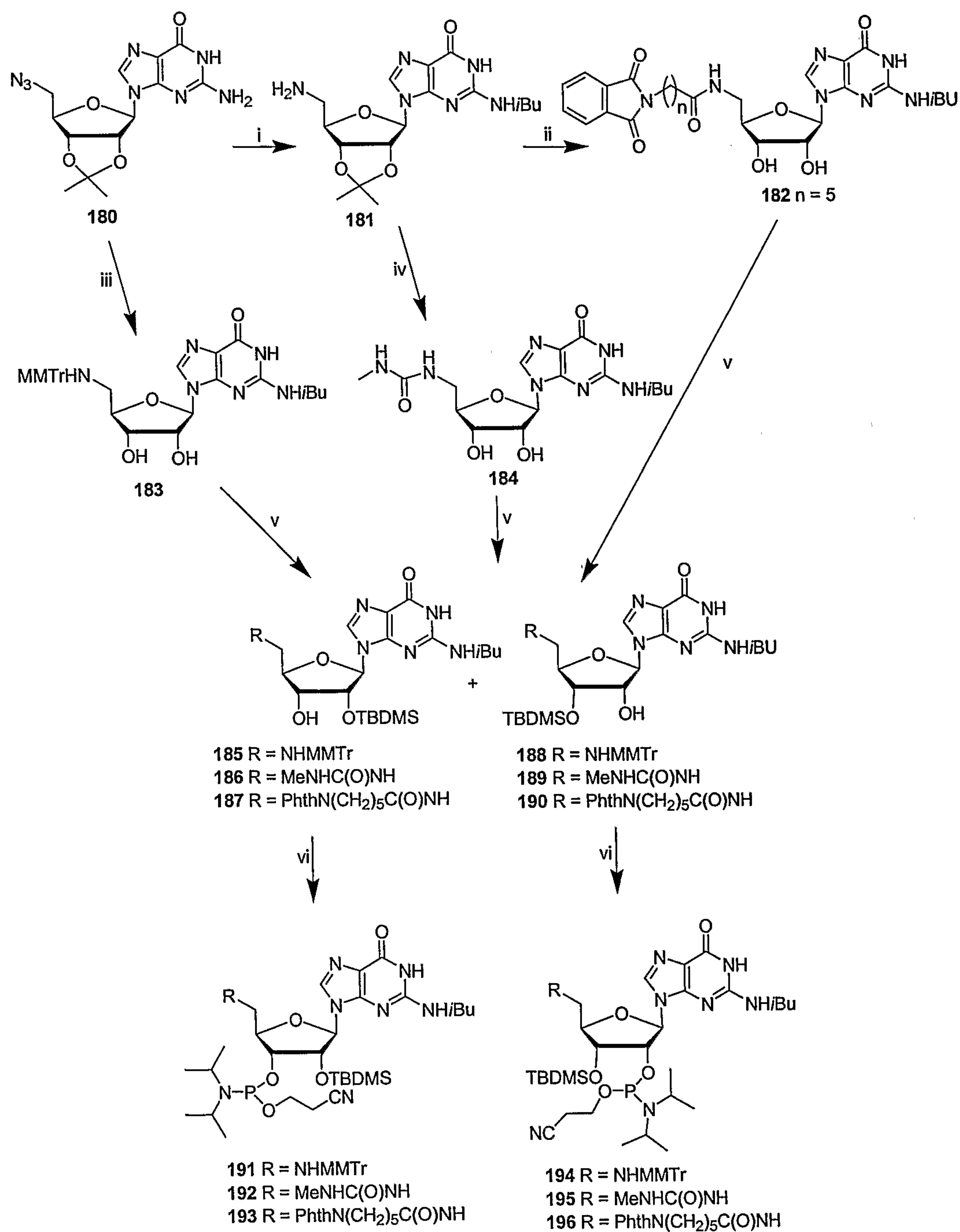
- ^a (i) (a) Bz₂O/Py and (b) H₂, Pd-C (10 %)/MeOH 1 atm; (ii) (a) *N*-Phthalimido-6-aminocaproic acid, DCC, DMAP, HOBT and (b) HCOOH-H₂O; (iii) (a) Bz₂O/Py, (b) HCOOH-H₂O, (c) H₂, Pd-C (10 %)/MeOH 1 atm and (d) MMTTr-Cl/Py; (iv) (a) CDI (carbonyldiimidazole)/THF, MeNH₂ or *p*-Nitrophenylchloroformate, DMAP/Py, MeNH₂ and (b) HCOOH-H₂O (v) TBDMS-Cl, Imidazole/Py; (vi) diisopropylamine tetrazolide, 2-cyanoethyl-*N,N,N',N'*-tetraisopropylphosphoramidite/CH₂Cl₂

The synthesis of certain compounds is described in Scheme 8, below. Compound 163 is obtained as reported in Liu and Austin (J. Org. Chem., 2001, 66, 8643).

Scheme 8^a

- ^a (i) (a) Bz₂O/Py and (b) H₂, Pd-C (10 %)/MeOH 1 atm; (ii) (a) *N*-Phthalimido-6-aminocaproic acid, DCC, DMAP, HOBT and (b) HCOOH-H₂O; (iii) (a) Bz₂O/Py, (b) HCOOH-H₂O, (c) H₂, Pd-C (10 %)/MeOH 1 atm and (d) MMTr-Cl/Py; (iv) (a) CDI (carbonyldiimidazole)/THF, MeNH₂ or *p*-Nitrophenylchloroformate, DMAP/Py, MeNH₂ and (b) HCOOH-H₂O (v) TBDMS-Cl, Imidazole/Py; (vi) diisopropylamine tetrazolide, 2-cyanoethyl-*N,N,N',N'*-tetraisopropylphosphoramidite/CH₂Cl₂

The synthesis of certain compounds is described in Scheme 9, below. Compound 180 is obtained as reported in Liu and Austin (J. Org. Chem., 2001, 66, 8643).

Scheme 9^a

^a (i) (a) *i*BuCOCl/Py and (b) H₂, Pd-C (10 %)/MeOH 1 atm; (ii) (a) *N*-Phthalimido-6-aminocaproic acid, DCC, DMAP, HOBT and (b) HCOOH-H₂O; (iii) (a) *i*BuCOCl/Py, (b) HCOOH-H₂O, (c) H₂, Pd-C (10 %)/MeOH 1 atm and (d) MMTTr-Cl/Py; (iv) (a) CDI (carbonyldiimidazole)/THF, MeNH₂ or *p*-Nitrophenylchloroformate, DMAP/Py, MeNH₂ and (b) HCOOH-H₂O (v) TBDMS-Cl, Imidazole/Py; (vi) diisopropylamine tetrazolide, 2-cyanoethyl-*N,N,N',N'*-tetraisopropylphosphoramidite/CH₂Cl₂

Targeting

The oligonucleotide agents featured in the invention, *e.g.*, oligonucleotide agents that target miRNAs or pre-miRNAs, can be targeted to particular tissues or cell types. For example, an oligonucleotide agent can be targeted to the liver, kidney, a cell of the nervous system, or a muscle cell. For targeting to the liver, for example, an oligonucleotide agent can include an SRMS containing a ligand that targets the liver, *e.g.*, a lipophilic moiety. Lipophilic moieties include lipid, cholesterol, oleyl, retinyl, or cholesteryl residues. Other lipophilic moieties that can function as liver-targeting agents include cholic acid, adamantane acetic acid, 1-pyrene butyric acid, dihydrotestosterone, 1,3-Bis-O(hexadecyl)glycerol, geranyloxyhexyl group, hexadecylglycerol, borneol, menthol, 1,3-propanediol, heptadecyl group, palmitic acid, myristic acid, O3-(oleoyl)lithocholic acid, O3-(oleoyl)cholenic acid, dimethoxytrityl, or phenoxazine. An oligonucleotide agent can also be targeted to the liver by association with a low-density lipoprotein (LDL), such as lactosylated LDL. Polymeric carriers complexed with sugar residues can also function to target oligonucleotide agents to the liver.

Conjugation of an oligonucleotide agent with a serum albumin (SA), such as human serum albumin, can also be used to target the oligonucleotide agent to a non-kidney tissue.

An oligonucleotide agent targeted to a tissue by an SRMS targeting moiety described herein can target a gene expressed in the tissue. For example, an oligonucleotide agent targeted to the liver, can target p21(WAF1/DIP1), P27(KIP1), beta-catenin, or c-MET, such as for treating a cancer of the liver. In another embodiment, the oligonucleotide agent can target apoB-100, such as for the treatment of an HDL/LDL cholesterol imbalance; dyslipidemias, *e.g.*, familial combined hyperlipidemia (FCHL), or acquired hyperlipidemia; hypercholesterolemia; statin-resistant hypercholesterolemia; coronary artery disease (CAD); coronary heart disease (CHD); or atherosclerosis.

A targeting agent that incorporates a sugar, *e.g.*, galactose and/or analogues thereof, can be useful. These agents target, for example, the parenchymal cells of the liver. For example, a

targeting moiety can include more than one or preferably two or three galactose moieties, spaced about 15 angstroms from each other. The targeting moiety can alternatively be lactose (*e.g.*, three lactose moieties), which is glucose coupled to a galactose. The targeting moiety can also be N-Acetyl-Galactosamine, N-Ac-Glucosamine. A mannose or mannose-6-phosphate targeting moiety can be used for macrophage targeting.

The oligonucleotide agents of the invention can also be targeted to the kidney, *e.g.*, by incorporation of an SRMS containing a ligand that targets the kidney.

An oligonucleotide agent targeted to the kidney by an SRMS targeting moiety described herein can target a gene expressed in the kidney.

Ligands on SRMSs can include folic acid, glucose, cholesterol, cholic acid, Vitamin E, Vitamin K, or Vitamin A.

Conjugation with ligands to promote entry into cells

Oligonucleotide agents, such as an miRNA oligonucleotide agents targeting miRNAs or pre-miRNAs, can be modified so as to enhance entry into cells, *e.g.*, an endocytic or non-endocytic mechanism. A ligand that increases cell permeability can be attached to an oligonucleotide agent in a number of ways, such as by attachment to an SRMS, *e.g.*, pyrroline-based SRMS.

In one embodiment, an oligonucleotide can be conjugated to a polyarginine that will enhance uptake into a wide range of cell-types. While not being bound by theory, the enhanced uptake is believed to be by a nonendocytic route. In another embodiment, an oligonucleotide can be conjugated to a guanidium analog of an aminoglycoside to promote cell permeability.

In another embodiment, an oligonucleotide can be conjugated with a lipophilic moiety. The lipophilic moiety can be attached at the nitrogen atom of a pyrroline-based SRMS. Examples of lipophilic moieties include cholesterol, lipid, oleyl, retinyl, or cholesteryl residues. Other lipophilic moieties include cholic acid, adamantane acetic acid, 1-pyrene butyric acid, dihydrotestosterone, 1,3-Bis-O(hexadecyl)glycerol, geranyloxyhexyl group, hexadecylglycerol, borneol, menthol, 1,3-propanediol, heptadecyl group, palmitic acid, myristic acid, O3-(oleoyl)lithocholic acid, O3-(oleoyl)cholenic acid, dimethoxytrityl, or phenoxazine. Cholesterol is a particularly preferred example.

The ligand that enhances cell permeability can be attached at the 3' terminus, the 5' terminus, or internally. The ligand can be attached to an SRMS, *e.g.*, a pyrroline-based SRMS at the 3' terminus, the 5' terminus, or at an internal linkage. The attachment can be direct or through a tethering molecule. Tethers, spacers or linkers discussed herein can be used to attach the moiety to the SRMS.

An oligonucleotide agent to which one or more cell-permeability ligands is conjugated (called an "OA-cell permeability conjugate") can be delivered *in vivo*, *e.g.*, to a cell, such as a cell of a tissue in a subject, such as a mammalian subject (*e.g.*, a human or mouse). Alternatively, or in addition, the oligonucleotide agent can be delivered *in vitro*, *e.g.*, to a cell in a cell line. Cell lines can be, for example, from a vertebrate organism, such as a mammal (*e.g.*, a human or a mouse). Delivery of an OA-cell permeability conjugate to a cell line can be in the absence of other transfection reagents. For example, delivery of an OA-cell permeability conjugate to a cell can be in the absence of, or optionally, in the presence of, LipofectamineTM (Invitrogen, Carlsbad, CA), Lipofectamine 2000TM, TransIT-TKOTM (Mirus, Madison, WI), FuGENE 6 (Roche, Indianapolis, IN), polyethylenimine, X-tremeGENE Q2 (Roche, Indianapolis, IN), DOTAP, DOSPER, or MetafecteneTM (Biontex, Munich, Germany), or another transfection reagent. Exemplary cell lines can be provided by the American Type Culture Collection (ATCC) (Manassus, Virginia). An OA-cell permeability conjugate can be delivered to a cell line, such as any cell line described herein, to target a specific gene for downregulation.

In one example, an oligonucleotide agent-lipophilic conjugate can be delivered to a primary cell line, *e.g.*, a synoviocyte (such as type B), cardiac myocyte, keratinocyte, hepatocyte, smooth muscle cell, endothelial cell, or dermal fibroblast cell line.

OLIGONUCLEOTIDE AGENT STRUCTURE

An oligonucleotide agent that is NAT ("nucleic acid targeting") includes a region of sufficient complementarity to the target gene, and is of sufficient length in terms of nucleotides, such that the oligonucleotide agent forms a duplex with the target nucleic acid. The oligonucleotide agent can modulate the function of the targeted molecule. For example, when the

targeted molecule is an mRNA or pre-mRNA, the NAT can inhibit gene expression; when the target is an miRNA, the NAT will inhibit the miRNA function and will thus up-regulate expression of the mRNAs targeted by the particular miRNA; when the target is a region of a pre-mRNA that affects splicing, the NAT can alter the choice of splice site and thus the mRNA sequence; when the NAT functions as an miRNA, expression of the targeted mRNA is inhibited. For ease of exposition the term nucleotide or ribonucleotide is sometimes used herein in reference to one or more monomeric subunits of an oligonucleotide agent. It will be understood herein that the usage of the term "ribonucleotide" or "nucleotide" herein can, in the case of a modified RNA or nucleotide surrogate, also refer to a modified nucleotide, or surrogate replacement moiety at one or more positions.

A NAT oligonucleotide agent is, or includes, a region that is at least partially, and in some embodiments fully, complementary to the target RNA. It is not necessary that there be perfect complementarity between the oligonucleotide agent and the target, but the correspondence must be sufficient to enable the oligonucleotide agent, or a cleavage product thereof, to modulate (*e.g.*, inhibit) target gene expression.

An oligonucleotide agent will preferably have one or more of the following properties:

- (1) it will be of the Formula 1, 2, 3, or 4 described below;
- (2) it will have a 5' modification that includes one or more phosphate groups or one or more analogs of a phosphate group;
- (3) it will, despite modifications, even to a very large number of bases specifically base pair and form a duplex structure with a homologous target RNA of sufficient thermodynamic stability to allow modulation of the activity of the targeted RNA;
- (4) it will, despite modifications, even to a very large number, or all of the nucleosides, still have "RNA-like" properties, *i.e.*, it will possess the overall structural, chemical and physical properties of an RNA molecule, even though not exclusively, or even partly, of ribonucleotide-based content. For example, all of the nucleotide sugars can contain *e.g.*, 2'OMe, 2' fluoro in place of 2' hydroxyl. This deoxyribonucleotide-containing agent can still be

expected to exhibit RNA-like properties. While not wishing to be bound by theory, the electronegative fluorine prefers an axial orientation when attached to the C2' position of ribose. This spatial preference of fluorine can, in turn, force the sugars to adopt a C_{3'}-*endo* pucker. This is the same puckering mode as observed in RNA molecules and gives rise to the RNA-
 5 characteristic A-family-type helix. Further, since fluorine is a good hydrogen bond acceptor, it can participate in the same hydrogen bonding interactions with water molecules that are known to stabilize RNA structures. (Generally, it is preferred that a modified moiety at the 2' sugar position will be able to enter into hydrogen-bonding which is more characteristic of the 2'-OH moiety of a ribonucleotide than the 2'-H moiety of a deoxyribonucleotide. A preferred
 10 oligonucleotide agent will: exhibit a C_{3'}-*endo* pucker in all, or at least 50, 75, 80, 85, 90, or 95 % of its sugars; exhibit a C_{3'}-*endo* pucker in a sufficient amount of its sugars that it can give rise to a the RNA-characteristic A-family-type helix; will have no more than 20, 10, 5, 4, 3, 2, or 1 sugar which is not a C_{3'}-*endo* pucker structure.

Preferred 2'-modifications with C_{3'}-*endo* sugar pucker include:

15 2'-OH, 2'-O-Me, 2'-O-methoxyethyl, 2'-O-aminopropyl, 2'-F, 2'-O-CH₂-CO-NHMe, 2'-O-CH₂-CH₂-O-CH₂-CH₂-N(Me)₂, LNA

(5) regardless of the nature of the modification, and even though the oligonucleotide agent can contain deoxynucleotides or modified deoxynucleotides, it is preferred that DNA molecules, or any molecule in which more than 50, 60, or 70 % of the nucleotides in the
 20 molecule are deoxyribonucleotides, or modified deoxyribonucleotides which are deoxy at the 2' position, are excluded from the definition of oligonucleotide agent.

Preferred 2'-modifications with a C_{2'}-*endo* sugar pucker include:

2'-H, 2'-Me, 2'-S-Me, 2'-Ethyne, 2'-ara-F.

Sugar modifications can also include L-sugars and 2'-5'-linked sugars.

25 As used herein, "specifically hybridizable" and "complementary" are terms that are used to indicate a sufficient degree of complementarity such that stable and specific binding occurs between a compound of the invention and a target RNA molecule in the case of NAT

oligonucleotides agents that bind target RNAs. Specific binding requires a sufficient lack of complementarity to non-target sequences under conditions in which specific binding is desired, *i.e.*, under physiological conditions in the case of *in vivo* assays or therapeutic treatment, or in the case of *in vitro* assays, under conditions in which the assays are performed. It has been shown
5 that a single mismatch between targeted and non-targeted sequences are sufficient to provide discrimination for siRNA targeting of an mRNA (Brummelkamp *et al.*, *Cancer Cell*, 2002, 2:243).

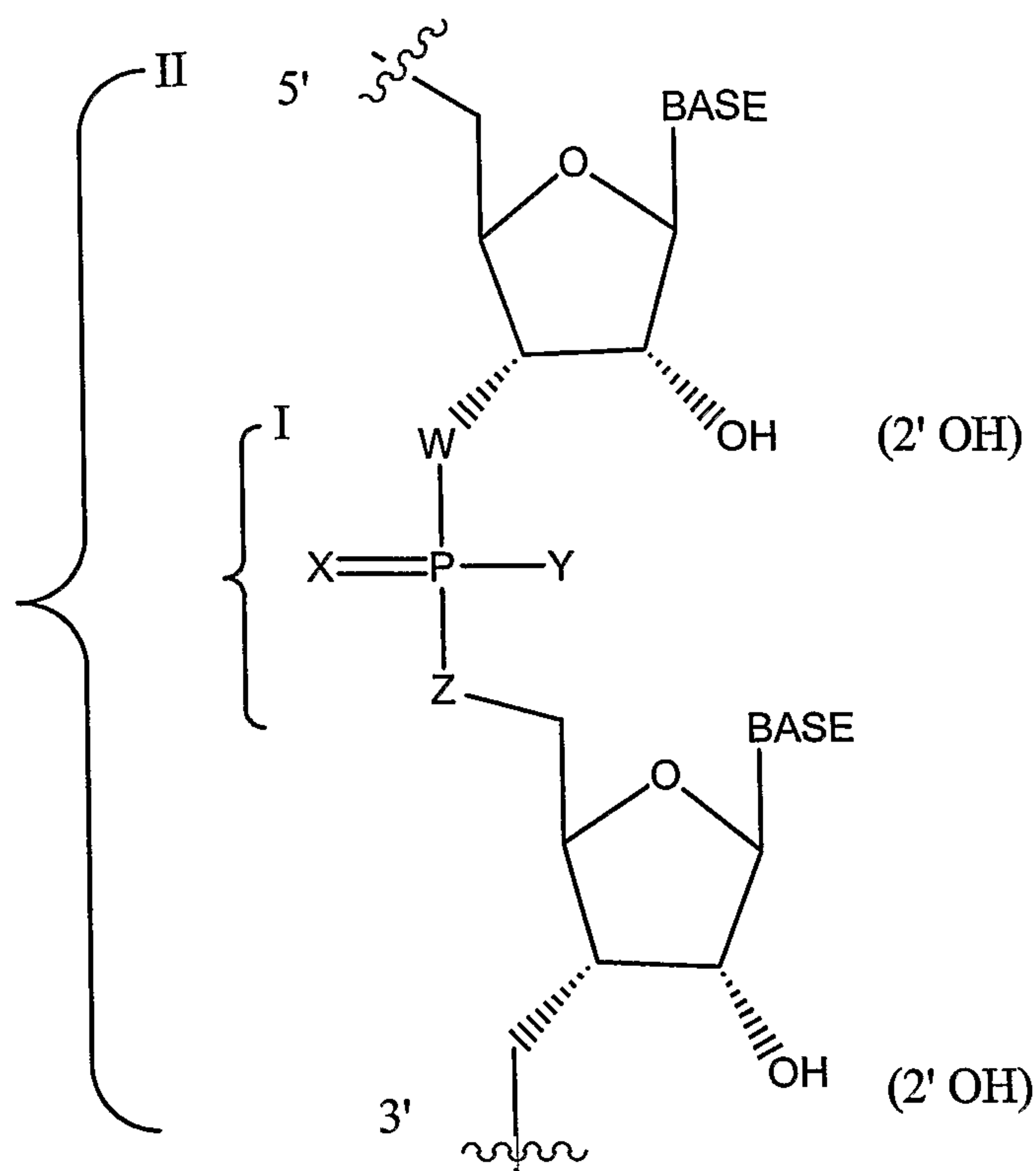
In one embodiment, a NAT oligonucleotide agent is “sufficiently complementary” to a target RNA, such that the oligonucleotide agent inhibits production of protein encoded by the
10 target mRNA. The target RNA can be, *e.g.*, a pre-mRNA, mRNA, or miRNA endogenous to the subject. In another embodiment, the oligonucleotide agent is “exactly complementary” (excluding the SRMS containing subunit(s)) to a target RNA, *e.g.*, the target RNA and the oligonucleotide agent can anneal to form a hybrid made exclusively of Watson-Crick base pairs in the region of exact complementarity. A “sufficiently complementary” target RNA can include
15 a region (*e.g.*, of at least 7 nucleotides) that is exactly complementary to a target RNA. Moreover, in some embodiments, the oligonucleotide agent specifically discriminates a single-nucleotide difference. In this case, the oligonucleotide agent only down-regulates gene expression if exact complementarity is found in the region the single-nucleotide difference.

Oligonucleotide agents discussed herein include otherwise unmodified RNA and DNA as
20 well as RNA and DNA that have been modified, *e.g.*, to improve efficacy, and polymers of nucleoside surrogates. Unmodified RNA refers to a molecule in which the components of the nucleic acid, namely sugars, bases, and phosphate moieties, are the same or essentially the same as that which occur in nature, preferably as occur naturally in the human body. The art has referred to rare or unusual, but naturally occurring, RNAs as modified RNAs, see, *e.g.*, Limbach
25 *et al.* (*Nucleic Acids Res.*, 1994, 22:2183-2196). Such rare or unusual RNAs, often termed modified RNAs, are typically the result of a post transcriptional modification and are within the term unmodified RNA as used herein. Modified RNA, as used herein, refers to a molecule in which one or more of the components of the nucleic acid, namely sugars, bases, and phosphate moieties, are different from that which occur in nature, preferably different from that which
30 occurs in the human body. While they are referred to as “modified RNAs” they will of course,

because of the modification, include molecules that are not, strictly speaking, RNAs. Nucleoside
surrogates are molecules in which the ribophosphate backbone is replaced with a non-
ribophosphate construct that allows the bases to be presented in the correct spatial relationship
such that hybridization is substantially similar to what is seen with a ribophosphate backbone,
5 *e.g.*, non-charged mimics of the ribophosphate backbone. Examples of all of the above are
discussed herein.

As nucleic acids are polymers of subunits or monomers, many of the modifications
described below occur at a position which is repeated within a nucleic acid, *e.g.*, a modification
of a base, or a phosphate moiety, or a non-linking O of a phosphate moiety. In some cases the
10 modification will occur at all of the subject positions in the nucleic acid but in many, and in fact
in most cases it will not. By way of example, a modification may only occur at a 3' or 5'
terminal position, may only occur in a terminal region, *e.g.* at a position on a terminal
nucleotide or in the last 2, 3, 4, 5, or 10 nucleotides of a strand. The ligand can be attached at
the 3' end, the 5' end, or at an internal position, or at a combination of these positions. For
15 example, the ligand can be at the 3' end and the 5' end; at the 3' end and at one or more internal
positions; at the 5' end and at one or more internal positions; or at the 3' end, the 5' end, and at
one or more internal positions. *E.g.*, a phosphorothioate modification at a non-linking O position
may only occur at one or both termini, or may only occur in a terminal region, *e.g.*, at a position
on a terminal nucleotide or in the last 2, 3, 4, 5, or 10 nucleotides of the oligonucleotide. The 5'
20 end can be phosphorylated.

Modifications and nucleotide surrogates are discussed below.

FORMULA 1

The scaffold presented above in Formula 1 represents a portion of a ribonucleic acid. The basic components are the ribose sugar, the base, the terminal phosphates, and phosphate internucleotide linkers. Where the bases are naturally occurring bases, *e.g.*, adenine, uracil, guanine or cytosine, the sugars are the unmodified 2' hydroxyl ribose sugar (as depicted) and W, X, Y, and Z are all O, Formula 1 represents a naturally occurring unmodified oligoribonucleotide.

Unmodified oligoribonucleotides may be less than optimal in some applications, *e.g.*, unmodified oligoribonucleotides can be prone to degradation by *e.g.*, cellular nucleases. Nucleases can hydrolyze nucleic acid phosphodiester bonds. However, chemical modifications to one or more of the above RNA components can confer improved properties, and, *e.g.*, can render oligoribonucleotides more stable to nucleases. Unmodified oligoribonucleotides may also be less than optimal in terms of offering tethering points for attaching ligands or other moieties to an oligonucleotide agent.

Modified nucleic acids and nucleotide surrogates can include one or more of:

- (i) alteration, *e.g.*, replacement, of one or both of the non-linking (X and Y) phosphate oxygens and/or of one or more of the linking (W and Z) phosphate oxygens (When the phosphate is in the terminal position, one of the positions W or Z will not link the phosphate to an additional element in a naturally occurring ribonucleic acid. However, for simplicity of terminology, except where otherwise noted, the W position at the 5' end of a nucleic acid and the terminal Z position at the 3' end of a nucleic acid, are within the term "linking phosphate oxygens" as used herein.);
- 10 (ii) alteration, *e.g.*, replacement, of a constituent of the ribose sugar, *e.g.*, of the 2' hydroxyl on the ribose sugar, or wholesale replacement of the ribose sugar with a structure other than ribose, *e.g.*, as described herein;
- (iii) wholesale replacement of the phosphate moiety (bracket I) with "dephospho" linkers;
- (iv) modification or replacement of a naturally occurring base;
- 15 (v) replacement or modification of the ribose-phosphate backbone (bracket II);
- (vi) modification of the 3' end or 5' end of the RNA, *e.g.*, removal, modification or replacement of a terminal phosphate group or conjugation of a moiety, *e.g.* a fluorescently labeled moiety, to either the 3' or 5' end of RNA.

The terms replacement, modification, alteration, and the like, as used in this context, do not imply any process limitation, *e.g.*, modification does not mean that one must start with a reference or naturally occurring ribonucleic acid and modify it to produce a modified ribonucleic acid but rather modified simply indicates a difference from a naturally occurring molecule.

It is understood that the actual electronic structure of some chemical entities cannot be adequately represented by only one canonical form (*i.e.* Lewis structure). While not wishing to be bound by theory, the actual structure can instead be some hybrid or weighted average of two or more canonical forms, known collectively as resonance forms or structures. Resonance structures are not discrete chemical entities and exist only on paper. They differ from one another only in the placement or "localization" of the bonding and nonbonding electrons for a particular chemical entity. It can be possible for one resonance structure to contribute to a greater extent to the hybrid than the others. Thus, the written and graphical descriptions of the embodiments of the present invention are made in terms of what the art recognizes as the

predominant resonance form for a particular species. For example, any phosphoroamidate (replacement of a nonlinking oxygen with nitrogen) would be represented by $X = O$ and $Y = N$ in the above figure.

Specific modifications are discussed in more detail below.

5

The Phosphate Group

The phosphate group is a negatively charged species. The charge is distributed equally over the two non-linking oxygen atoms (*i.e.*, X and Y in Formula 1 above). However, the phosphate group can be modified by replacing one of the oxygens with a different substituent.

10 One result of this modification to RNA phosphate backbones can be increased resistance of the oligoribonucleotide to nucleolytic breakdown. Thus while not wishing to be bound by theory, it can be desirable in some embodiments to introduce alterations which result in either an uncharged linker or a charged linker with unsymmetrical charge distribution.

Examples of modified phosphate groups include phosphorothioate, phosphoroselenates, 15 borano phosphates, borano phosphate esters, hydrogen phosphonates, phosphoroamidates, alkyl or aryl phosphonates and phosphotriesters. Phosphorodithioates have both non-linking oxygens replaced by sulfur. Unlike the situation where only one of X or Y is altered, the phosphorus center in the phosphorodithioates is achiral which precludes the formation of oligoribonucleotides diastereomers. Diastereomer formation can result in a preparation in which 20 the individual diastereomers exhibit varying resistance to nucleases. Further, the hybridization affinity of RNA containing chiral phosphate groups can be lower relative to the corresponding unmodified RNA species. Thus, while not wishing to be bound by theory, modifications to both X and Y which eliminate the chiral center, *e.g.* phosphorodithioate formation, may be desirable in that they cannot produce diastereomer mixtures. Thus, X can be any one of S, Se, B, C, H, N, 25 or OR (R is alkyl or aryl). Thus Y can be any one of S, Se, B, C, H, N, or OR (R is alkyl or aryl). Replacement of X and/or Y with sulfur is preferred.

The phosphate linker can also be modified by replacement of a linking oxygen (*i.e.*, W or Z in Formula 1) with nitrogen (bridged phosphoroamidates), sulfur (bridged phosphorothioates) and carbon (bridged methylenephosphonates). The replacement can occur at a terminal oxygen 30 (position W (3') or position Z (5')). Replacement of W with carbon or Z with nitrogen is preferred.

Candidate agents can be evaluated for suitability as described below.

The Sugar Group

A modified RNA can include modification of all or some of the sugar groups of the
 5 ribonucleic acid. *E.g.*, the 2' hydroxyl group (OH) can be modified or replaced with a number of
 different "oxy" or "deoxy" substituents. While not being bound by theory, enhanced stability is
 expected since the hydroxyl can no longer be deprotonated to form a 2' alkoxide ion. The 2'
 alkoxide can catalyze degradation by intramolecular nucleophilic attack on the linker phosphorus
 atom. Again, while not wishing to be bound by theory, it can be desirable to some embodiments
 10 to introduce alterations in which alkoxide formation at the 2' position is not possible.

Examples of "oxy"-2' hydroxyl group modifications include alkoxy or aryloxy (OR, *e.g.*,
 R = H, alkyl, cycloalkyl, aryl, aralkyl, heteroaryl or sugar); polyethyleneglycols (PEG),
 $O(CH_2CH_2O)_nCH_2CH_2OR$; "locked" nucleic acids (LNA) in which the 2' hydroxyl is connected,
e.g., by a methylene bridge or ethylene bridge (*e.g.*, 2'-4'-ethylene bridged nucleic acid (ENA)),
 15 to the 4' carbon of the same ribose sugar; O-AMINE (AMINE = NH_2 ; alkylamino, dialkylamino,
 heterocyclyl, arylamino, diaryl amino, heteroaryl amino, or diheteroaryl amino, ethylene
 diamine, polyamino) and aminoalkoxy, $O(CH_2)_nAMINE$, (*e.g.*, AMINE = NH_2 ; alkylamino,
 dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, or diheteroaryl amino,
 ethylene diamine, polyamino). It is noteworthy that oligonucleotides containing only the
 20 methoxyethyl group (MOE), $(OCH_2CH_2OCH_3)$, a PEG derivative), exhibit nuclease stabilities
 comparable to those modified with the robust phosphorothioate modification.

"Deoxy" modifications include hydrogen (*i.e.* deoxyribose sugars); halo (*e.g.*, fluoro);
 amino (*e.g.* NH_2 ; alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl
 amino, diheteroaryl amino, or amino acid); $NH(CH_2CH_2NH)_nCH_2CH_2-AMINE$ (AMINE = NH_2 ;
 25 alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, or
 diheteroaryl amino), $-NHC(O)R$ (R = alkyl, cycloalkyl, aryl, aralkyl, heteroaryl or sugar), cyano;
 mercapto; alkyl-thio-alkyl; thioalkoxy; and alkyl, cycloalkyl, aryl, alkenyl and alkynyl, which
 may be optionally substituted with *e.g.*, an amino functionality. Preferred substituents are 2'-
 methoxyethyl, 2'-OCH₃, 2'-O-allyl, 2'-C-allyl, and 2'-fluoro.

The sugar group can also contain one or more carbons that possess the opposite stereochemical configuration than that of the corresponding carbon in ribose. Thus, a modified RNA can include nucleotides containing *e.g.*, arabinose, as the sugar.

Modified RNAs can also include “abasic” sugars, which lack a nucleobase at C-1'. These
5 abasic sugars can also be further contain modifications at one or more of the constituent sugar atoms.

To maximize nuclease resistance, the 2' modifications can be used in combination with one or more phosphate linker modifications (*e.g.*, phosphorothioate). The so-called “chimeric” oligonucleotides are those that contain two or more different modifications.

10 The modification can also entail the wholesale replacement of a ribose structure with another entity at one or more sites in the oligonucleotide agent. These modifications are described in section entitled Sugar Replacements for SRMSs.

Candidate modifications can be evaluated as described below.

15 Replacement of the Phosphate Group

The phosphate group can be replaced by non-phosphorus containing connectors (*cf.* Bracket I in Formula 1 above). While not wishing to be bound by theory, it is believed that since the charged phosphodiester group is the reaction center in nucleolytic degradation, its replacement with neutral structural mimics should impart enhanced nuclease stability. Again,
20 while not wishing to be bound by theory, it can be desirable, in some embodiment, to introduce alterations in which the charged phosphate group is replaced by a neutral moiety.

Examples of moieties which can replace the phosphate group include siloxane, carbonate, carboxymethyl, carbamate, amide, thioether, ethylene oxide linker, sulfonate, sulfonamide, thioformacetal, formacetal, oxime, methyleneimino, methylenemethylimino, methylenehydrazo,
25 methylenedimethylhydrazo and methyleneoxymethylimino. Preferred replacements include the methylenecarbonylamino and methylenemethylimino groups.

Candidate modifications can be evaluated as described below.

Replacement of Ribophosphate Backbone

30 Oligonucleotide- mimicking scaffolds can also be constructed wherein the phosphate linker and ribose sugar are replaced by nuclease resistant nucleoside or nucleotide surrogates

(see Bracket II of Formula 1 above). While not wishing to be bound by theory, it is believed that the absence of a repetitively charged backbone diminishes binding to proteins that recognize polyanions (*e.g.* nucleases). Again, while not wishing to be bound by theory, it can be desirable in some embodiment, to introduce alterations in which the bases are tethered by a neutral surrogate backbone.

Examples include the morpholino, cyclobutyl, pyrrolidine and peptide nucleic acid (PNA) nucleoside surrogates. A preferred surrogate is a PNA surrogate.

Candidate modifications can be evaluated as described below.

Terminal Modifications

The 3' and 5' ends of an oligonucleotide can be modified. Such modifications can be at the 3' end, 5' end or both ends of the molecule. They can include modification or replacement of an entire terminal phosphate or of one or more of the atoms of the phosphate group. *E.g.*, the 3' and 5' ends of an oligonucleotide can be conjugated to other functional molecular entities such as labeling moieties, *e.g.*, fluorophores (*e.g.*, pyrene, TAMRA, fluorescein, Cy3 or Cy5 dyes) or protecting groups (based *e.g.*, on sulfur, silicon, boron or ester). The functional molecular entities can be attached to the sugar through a phosphate group and/or a spacer. The terminal atom of the spacer can connect to or replace the linking atom of the phosphate group or the C-3' or C-5' O, N, S or C group of the sugar. Alternatively, the spacer can connect to or replace the terminal atom of a nucleotide surrogate (*e.g.*, PNAs). These spacers or linkers can include *e.g.*, $-(CH_2)_n-$, $-(CH_2)_nN-$, $-(CH_2)_nO-$, $-(CH_2)_nS-$, $O(CH_2CH_2O)_nCH_2CH_2OH$ (*e.g.*, $n = 3$ or 6), abasic sugars, amide, carboxy, amine, oxyamine, oximine, thioether, disulfide, thiourea, sulfonamide, or morpholino, or biotin and fluorescein reagents. While not wishing to be bound by theory, it is believed that conjugation of certain moieties can improve transport, hybridization, and specificity properties. Again, while not wishing to be bound by theory, it may be desirable to introduce terminal alterations that improve nuclease resistance. Other examples of terminal modifications include dyes, intercalating agents (*e.g.* acridines), cross-linkers (*e.g.* psoralene, mitomycin C), porphyrins (TPPC4, texaphyrin, Sapphyrin), polycyclic aromatic hydrocarbons (*e.g.*, phenazine, dihydrophenazine), artificial endonucleases (*e.g.* EDTA), lipophilic carriers (*e.g.*, cholesterol, cholic acid, adamantane acetic acid, 1-pyrene butyric acid, dihydrotestosterone,

1,3-Bis-O(hexadecyl)glycerol, geranyloxyhexyl group, hexadecylglycerol, borneol, menthol, 1,3-propanediol, heptadecyl group, palmitic acid, myristic acid, O3-(oleoyl)lithocholic acid, O3-(oleoyl)cholenic acid, dimethoxytrityl, or phenoxazine) and peptide conjugates (*e.g.*, antennapedia peptide, Tat peptide), alkylating agents, phosphate, amino, mercapto, PEG (*e.g.*,
 5 PEG-40K), MPEG, [MPEG]₂, polyamino, alkyl, substituted alkyl, radiolabeled markers, enzymes, haptens (*e.g.* biotin), transport/absorption facilitators (*e.g.*, aspirin, vitamin E, folic acid), synthetic ribonucleases (*e.g.*, imidazole, bisimidazole, histamine, imidazole clusters, acridine-imidazole conjugates, Eu³⁺ complexes of tetraazamacrocycles).

Terminal modifications can be added for a number of reasons, including as discussed
 10 elsewhere herein to modulate activity or to modulate resistance to degradation. Preferred modifications include the addition of a methylphosphonate at the 3'-most terminal linkage; a 3' C5-aminoalkyl-dT; 3' cationic group; or another 3' conjugate to inhibit 3'-5' exonucleolytic degradation.

Terminal modifications useful for modulating activity include modification of the 5' end
 15 with phosphate or phosphate analogs. *E.g.*, in preferred embodiments oligonucleotide agents are 5' phosphorylated or include a phosphoryl analog at the 5' terminus. 5'-phosphate modifications include those which are compatible with RISC mediated gene silencing. Suitable modifications include: 5'-monophosphate ((HO)₂(O)P-O-5'); 5'-diphosphate ((HO)₂(O)P-O-P(HO)(O)-O-5'); 5'-triphosphate ((HO)₂(O)P-O-(HO)(O)P-O-P(HO)(O)-O-5'); 5'-guanosine cap (7-methylated or
 20 non-methylated) (7m-G-O-5'-(HO)(O)P-O-(HO)(O)P-O-P(HO)(O)-O-5'); 5'-adenosine cap (A_{ppp}), and any modified or unmodified nucleotide cap structure (N-O-5'-(HO)(O)P-O-(HO)(O)P-O-P(HO)(O)-O-5'); 5'-monothiophosphate (phosphorothioate; (HO)₂(S)P-O-5'); 5'-monodithiophosphate (phosphorodithioate; (HO)(HS)(S)P-O-5'), 5'-phosphorothiolate ((HO)₂(O)P-S-5'); any additional combination of oxygen/sulfur replaced monophosphate,
 25 diphosphate and triphosphates (*e.g.* 5'-alpha-thiotriphosphate, 5'-gamma-thiotriphosphate, etc.), 5'-phosphoramidates ((HO)₂(O)P-NH-5', (HO)(NH₂)(O)P-O-5'), 5'-alkylphosphonates (R=alkyl=methyl, ethyl, isopropyl, propyl, etc., *e.g.* RP(OH)(O)-O-5'-, (OH)₂(O)P-5'-CH₂-), 5'-alkyletherphosphonates (R=alkylether=methoxymethyl (MeOCH₂-), ethoxymethyl, etc., *e.g.* RP(OH)(O)-O-5'-).

30 Terminal modifications can also be useful for monitoring distribution, and in such cases the preferred groups to be added include fluorophores, *e.g.*, fluorescein or an Alexa dye, *e.g.*,

Alexa 488. Terminal modifications can also be useful for enhancing uptake, useful modifications for this include cholesterol. Terminal modifications can also be useful for cross-linking an oligonucleotide agent to another moiety; modifications useful for this include mitomycin C.

5 Candidate modifications can be evaluated as described below.

The Bases

Adenine, guanine, cytosine and uracil are the most common bases found in RNA. These bases can be modified or replaced to provide RNA's having improved properties. *E.g.*, nuclease
10 resistant oligoribonucleotides can be prepared with these bases or with synthetic and natural nucleobases (*e.g.*, inosine, thymine, xanthine, hypoxanthine, nebularine, isoguanisine, or tubercidine) and any one of the above modifications. Alternatively, substituted or modified analogs of any of the above bases, *e.g.*, "unusual bases" and "universal bases" described herein, can be employed. Examples include without limitation 2-aminoadenine, 6-methyl and other
15 alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 5-halouracil and cytosine, 5-propynyl uracil and cytosine, 6-azo uracil, cytosine and thymine, 5-uracil (pseudouracil), 4-thiouracil, 5-halouracil, 5-(2-aminopropyl)uracil, 5-amino allyl uracil, 8-halo, amino, thiol, thioalkyl, hydroxyl and other 8-substituted adenines and guanines, 5-trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine, 5-
20 substituted pyrimidines, 6-azapyrimidines and N-2, N-6 and O-6 substituted purines, including 2-aminopropyladenine, 5-propynyluracil and 5-propynylcytosine, dihydrouracil, 3-deaza-5-azacytosine, 2-aminopurine, 5-alkyluracil, 7-alkylguanine, 5-alkyl cytosine, 7-deazaadenine, N6, N6-dimethyladenine, 2,6-diaminopurine, 5-amino-allyl-uracil, N3-methyluracil, substituted 1,2,4-triazoles, 2-pyridinone, 5-nitroindole, 3-nitropyrrole, 5-methoxyuracil, uracil-5-oxyacetic
25 acid, 5-methoxycarbonylmethyluracil, 5-methyl-2-thiouracil, 5-methoxycarbonylmethyl-2-thiouracil, 5-methylaminomethyl-2-thiouracil, 3-(3-amino-3-carboxypropyl)uracil, 3-methylcytosine, 5-methylcytosine, N⁴-acetyl cytosine, 2-thiocytosine, N6-methyladenine, N6-isopentyladenine, 2-methylthio-N6-isopentenyladenine, N-methylguanines, or O-alkylated bases. Further purines and pyrimidines include those disclosed in U.S. Pat. No. 3,687,808, those
30 disclosed in the Concise Encyclopedia Of Polymer Science And Engineering, pages 858-859,

Kroschwitz, J. I., ed. John Wiley & Sons, 1990, and those disclosed by Englisch *et al.*, *Angewandte Chemie, International Edition*, 1991, 30, 613.

Generally, base changes are less preferred for promoting stability, but they can be useful for other reasons, *e.g.*, some, *e.g.*, 2,6-diaminopurine and 2 amino purine (*e.g.*, 2-amino adenine),
5 are fluorescent. Modified bases can reduce target specificity. This should be taken into consideration in the design of oligonucleotide agents.

Candidate modifications can be evaluated as described below.

Evaluation of Candidate Oligonucleotide agents

10 One can evaluate a candidate oligonucleotide agent, *e.g.*, a modified oligonucleotide agent, for a selected property by exposing the agent or modified molecule and a control molecule to the appropriate conditions and evaluating for the presence of the selected property. For example, resistance to a degradant can be evaluated as follows. A candidate modified RNA (and preferably a control molecule, usually the unmodified form) can be exposed to degradative
15 conditions, *e.g.*, exposed to a milieu, which includes a degradative agent, *e.g.*, a nuclease. *E.g.*, one can use a biological sample, *e.g.*, one that is similar to a milieu, which might be encountered, in therapeutic use, *e.g.*, blood or a cellular fraction, *e.g.*, a cell-free homogenate or disrupted cells. The candidate and control could then be evaluated for resistance to degradation by any of a number of approaches. For example, the candidate and control could be labeled, preferably
20 prior to exposure, with, *e.g.*, a radioactive or enzymatic label, or a fluorescent label, such as Cy3 or Cy5. Control and modified RNA's can be incubated with the degradative agent, and optionally a control, *e.g.*, an inactivated, *e.g.*, heat inactivated, degradative agent. A physical parameter, *e.g.*, size, of the modified and control molecules are then determined. They can be determined by a physical method, *e.g.*, by polyacrylamide gel electrophoresis or a sizing column,
25 to assess whether the molecule has maintained its original length, or assessed functionally. Alternatively, Northern blot analysis can be used to assay the length of an unlabeled modified molecule.

A functional assay can also be used to evaluate the candidate agent. A functional assay can be applied initially or after an earlier non-functional assay, (*e.g.*, assay for resistance to
30 degradation) to determine if the modification alters the ability of the molecule to inhibit gene expression. For example, a cell, *e.g.*, a mammalian cell, such as a mouse or human cell, can be

co-transfected with a plasmid expressing a fluorescent protein, *e.g.*, GFP, and a candidate oligonucleotide agent homologous to the transcript encoding the fluorescent protein (see, *e.g.*, WO 00/44914). For example, a modified oligonucleotide agent homologous to the GFP mRNA can be assayed for the ability to inhibit GFP expression by monitoring for a decrease in cell fluorescence, as compared to a control cell, in which the transfection did not include the candidate oligonucleotide agent, *e.g.*, controls with no agent added and/or controls with a non-modified RNA added. Efficacy of the candidate agent on gene expression can be assessed by comparing cell fluorescence in the presence of the modified and unmodified oligonucleotide agent. In an alternative functional assay, a candidate oligonucleotide agent homologous to an endogenous mouse gene, preferably a maternally expressed gene, such as *c-mos*, can be injected into an immature mouse oocyte to assess the ability of the agent to inhibit gene expression *in vivo* (see, *e.g.*, WO 01/36646). A phenotype of the oocyte, *e.g.*, the ability to maintain arrest in metaphase II, can be monitored as an indicator that the agent is inhibiting expression. For example, cleavage of *c-mos* mRNA by an oligonucleotide agent would cause the oocyte to exit metaphase arrest and initiate parthenogenetic development (Colledge *et al.* Nature 370: 65-68, 1994; Hashimoto *et al.* Nature, 370:68-71, 1994). The effect of the modified agent on target RNA levels can be verified by Northern blot to assay for a decrease in the level of target RNA, or by Western blot to assay for a decrease in the level of target protein, as compared to a negative control. Controls can include cells in which with no agent is added and/or cells in which a non-modified RNA is added.

An oligonucleotide agent that targets an miRNA ore pre-miRNA can be assayed by monitoring expression of the transcript targeted by the miRNA. For example, an oligonucleotide agent designed to bind an miRNA that targets GFP can be assessed by monitoring for an increase in cell fluorescence, as compared to a control cell, in which the transfection did not include the candidate oligonucleotide agent, *e.g.*, controls with no agent added and/or controls with a non-modified RNA added. In another example, an oligonucleotide agent designed to bind an miRNA that targets an endogenous enzyme can be assessed by monitoring for an increase in enzyme activity, as compared to a control cell. The effect of the modified oligonucleotide agent on target miRNA levels can be verified by Northern blot to assay for a decrease in the level of the target miRNA.

References

General References

The oligoribonucleotides and oligoribonucleosides used in accordance with this invention may be with solid phase synthesis, see for example "Oligonucleotide synthesis, a practical approach", Ed. M. J. Gait, IRL Press, 1984; "Oligonucleotides and Analogues, A Practical Approach", Ed. F. Eckstein, IRL Press, 1991 (especially Chapter 1, Modern machine-aided methods of oligodeoxyribonucleotide synthesis, Chapter 2, Oligoribonucleotide synthesis, Chapter 3, 2'-O-Methyloligoribonucleotide- s: synthesis and applications, Chapter 4, Phosphorothioate oligonucleotides, Chapter 5, Synthesis of oligonucleotide phosphorodithioates, Chapter 6, Synthesis of oligo-2'-deoxyribonucleoside methylphosphonates, and. Chapter 7, Oligodeoxynucleotides containing modified bases. Other particularly useful synthetic procedures, reagents, blocking groups and reaction conditions are described in Martin, P., *Helv. Chim. Acta*, **1995**, 78, 486-504; Beaucage, S. L. and Iyer, R. P., *Tetrahedron*, **1992**, 48, 2223-2311 and Beaucage, S. L. and Iyer, R. P., *Tetrahedron*, **1993**, 49, 6123-6194, or references referred to therein.

Modification described in WO 00/44895, WO01/75164, or WO02/44321 can be used herein.

The disclosure of all publications, patents, and published patent applications listed herein are hereby incorporated by reference.

Phosphate Group References

The preparation of phosphinate oligoribonucleotides is described in U.S. Pat. No. 5,508,270. The preparation of alkyl phosphonate oligoribonucleotides is described in U.S. Pat. No. 4,469,863. The preparation of phosphoramidite oligoribonucleotides is described in U.S. Pat. No. 5,256,775 or U.S. Pat. No. 5,366,878. The preparation of phosphotriester oligoribonucleotides is described in U.S. Pat. No. 5,023,243. The preparation of borano phosphate oligoribonucleotide is described in U.S. Pat. Nos. 5,130,302 and 5,177,198. The preparation of 3'-Deoxy-3'-amino phosphoramidate oligoribonucleotides is described in U.S. Pat. No. 5,476,925. 3'-Deoxy-3'-methylenephosphonate oligoribonucleotides is described in An, H, *et al. J. Org. Chem.* **2001**, 66, 2789-2801. Preparation of sulfur bridged nucleotides is described in Sproat *et al. Nucleosides Nucleotides* **1988**, 7,651 and Crosstick *et al. Tetrahedron Lett.* **1989**, 30, 4693.

Sugar Group References

Modifications to the 2' modifications can be found in Verma, S. *et al. Annu. Rev. Biochem.* **1998**, *67*, 99-134 and all references therein. Specific modifications to the ribose can be found in the following references: 2'-fluoro (Kawasaki *et. al., J. Med. Chem.*, **1993**, *36*, 831-841), 2'-MOE (Martin, P. *Helv. Chim. Acta* **1996**, *79*, 1930-1938), "LNA" (Wengel, J. *Acc. Chem. Res.* **1999**, *32*, 301-310).

Replacement of the Phosphate Group References

Methylenemethylimino linked oligoribonucleosides, also identified herein as MMI linked oligoribonucleosides, methylenedimethylhydrazo linked oligoribonucleosides, also identified herein as MDH linked oligoribonucleosides, and methylenecarbonylamino linked oligonucleosides, also identified herein as amide-3 linked oligoribonucleosides, and methyleneaminocarbonyl linked oligonucleosides, also identified herein as amide-4 linked oligoribonucleosides as well as mixed backbone compounds having, as for instance, alternating MMI and PO or PS linkages can be prepared as is described in U.S. Pat. Nos. 5,378,825, 5,386,023, 5,489,677 and in published PCT applications PCT/US92/04294 and PCT/US92/04305 (published as WO 92/20822 WO and 92/20823, respectively). Formacetal and thioformacetal linked oligoribonucleosides can be prepared as is described in U.S. Pat. Nos. 5,264,562 and 5,264,564. Ethylene oxide linked oligoribonucleosides can be prepared as is described in U.S. Pat. No. 5,223,618. Siloxane replacements are described in Cormier, J.F. *et al. Nucleic Acids Res.* **1988**, *16*, 4583. Carbonate replacements are described in Tittensor, J.R. *J. Chem. Soc. C* **1971**, 1933. Carboxymethyl replacements are described in Edge, M.D. *et al. J. Chem. Soc. Perkin Trans. 1* **1972**, 1991. Carbamate replacements are described in Stirchak, E.P. *Nucleic Acids Res.* **1989**, *17*, 6129.

Replacement of the Phosphate-Ribose Backbone References

Cyclobutyl sugar surrogate compounds can be prepared as is described in U.S. Pat. No. 5,359,044. Pyrrolidine sugar surrogate can be prepared as is described in U.S. Pat. No. 5,519,134. Morpholino sugar surrogates can be prepared as is described in U.S. Pat. Nos. 5,142,047 and 5,235,033, and other related patent disclosures. Peptide Nucleic Acids (PNAs) are

known per se and can be prepared in accordance with any of the various procedures referred to in Peptide Nucleic Acids (PNA): Synthesis, Properties and Potential Applications, Bioorganic & Medicinal Chemistry, 1996, 4, 5-23. They may also be prepared in accordance with U.S. Pat. No. 5,539,083.

5

Terminal Modification References

Terminal modifications are described in Manoharan, M. *et al. Antisense and Nucleic Acid Drug Development* 12, 103-128 (2002) and references therein.

10

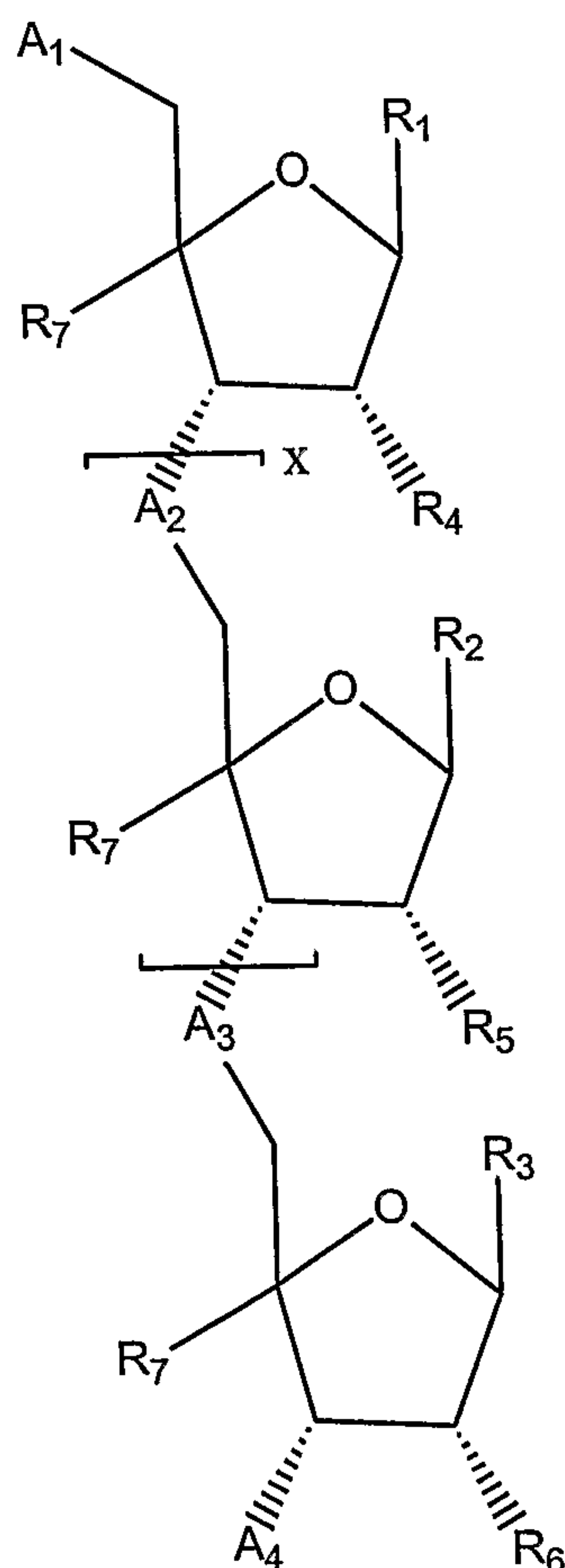
Bases References

N-2 substituted purine nucleoside amidites can be prepared as is described in U.S. Pat. No. 5,459,255. 3-Deaza purine nucleoside amidites can be prepared as is described in U.S. Pat. No. 5,457,191. 5,6-Substituted pyrimidine nucleoside amidites can be prepared as is described in U.S. Pat. No. 5,614,617. 5-Propynyl pyrimidine nucleoside amidites can be prepared as is described in U.S. Pat. No. 5,484,908. Additional references can be disclosed in the above section on base modifications.

15

Preferred oligonucleotide agents

Preferred oligonucleotide agents have the following structure (see Formula 2 below):

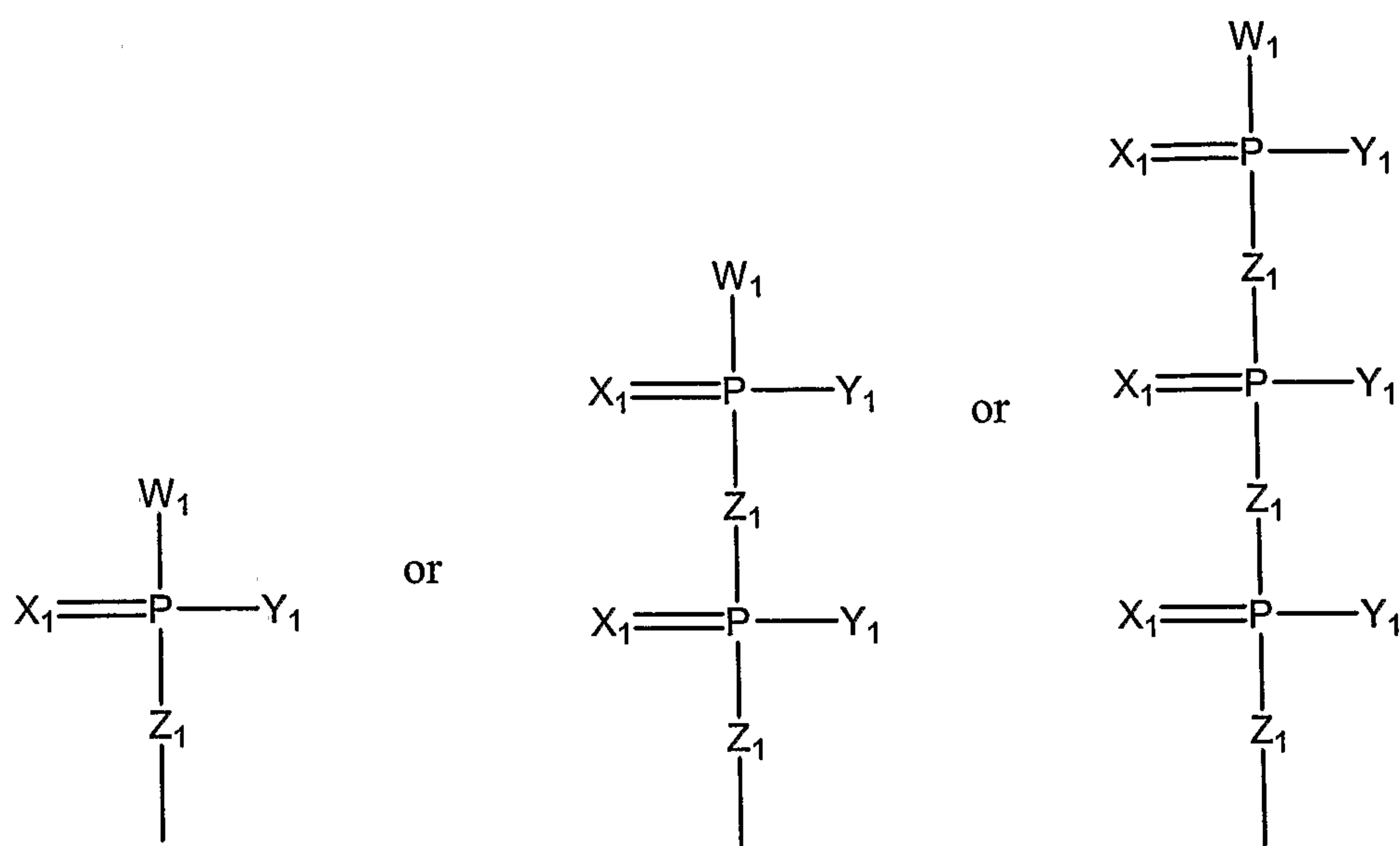
FORMULA 2

5 Referring to Formula 2 above, R^1 , R^2 , and R^3 are each, independently, H, (*i.e.* abasic nucleotides), adenine, guanine, cytosine and uracil, inosine, thymine, xanthine, hypoxanthine, nubularine, tubercidine, isoguanisine, 2-aminoadenine, 6-methyl and other alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 5-halouracil and cytosine, 5-propynyl uracil and cytosine, 6-azo uracil, cytosine and thymine, 5-uracil
 10 (pseudouracil), 4-thiouracil, 5-halouracil, 5-(2-aminopropyl)uracil, 5-amino allyl uracil, 8-halo, amino, thiol, thioalkyl, hydroxyl and other 8-substituted adenines and guanines, 5-

trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine, 5-substituted pyrimidines, 6-azapyrimidines and N-2, N-6 and O-6 substituted purines, including 2-aminopropyladenine, 5-propynyluracil and 5-propynylcytosine, dihydrouracil, 3-deaza-5-azacytosine, 2-aminopurine, 5-alkyluracil, 7-alkylguanine, 5-alkyl cytosine, 7-deazaadenine, 7-deazaguanine, N6, N6-dimethyladenine, 2,6-diaminopurine, 5-amino-allyl-uracil, N3-methyluracil, substituted 1,2,4-triazoles, 2-pyridinone, 5-nitroindole, 3-nitropyrrole, 5-methoxyuracil, uracil-5-oxyacetic acid, 5-methoxycarbonylmethyluracil, 5-methyl-2-thiouracil, 5-methoxycarbonylmethyl-2-thiouracil, 5-methylaminomethyl-2-thiouracil, 3-(3-amino-3carboxypropyl)uracil, 3-methylcytosine, 5-methylcytosine, N⁴-acetyl cytosine, 2-thiocytosine, N6-methyladenine, N6-isopentyladenine, 2-methylthio-N6-isopentenyladenine, N-methylguanines, or O-alkylated bases.

R⁴, R⁵, and R⁶ are each, independently, OR⁸, O(CH₂CH₂O)_mCH₂CH₂OR⁸; O(CH₂)_nR⁹; O(CH₂)_nOR⁹, H; halo; NH₂; NHR⁸; N(R⁸)₂; NH(CH₂CH₂NH)_mCH₂CH₂NHR⁹; NHC(O)R⁸; ; cyano; mercapto, SR⁸; alkyl-thio-alkyl; alkyl, aralkyl, cycloalkyl, aryl, heteroaryl, alkenyl, alkynyl, each of which may be optionally substituted with halo, hydroxy, oxo, nitro, haloalkyl, alkyl, alkaryl, aryl, aralkyl, alkoxy, aryloxy, amino, alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, diheteroaryl amino, acylamino, alkylcarbamoyl, arylcarbamoyl, aminoalkyl, alkoxycarbonyl, carboxy, hydroxyalkyl, alkanesulfonyl, alkanesulfonamido, arenesulfonamido, aralkylsulfonamido, alkylcarbonyl, acyloxy, cyano, or ureido; or R⁴, R⁵, or R⁶ together combine with R⁷ to form an [-O-CH₂-] covalently bound bridge between the sugar 2' and 4' carbons.

A¹ is:

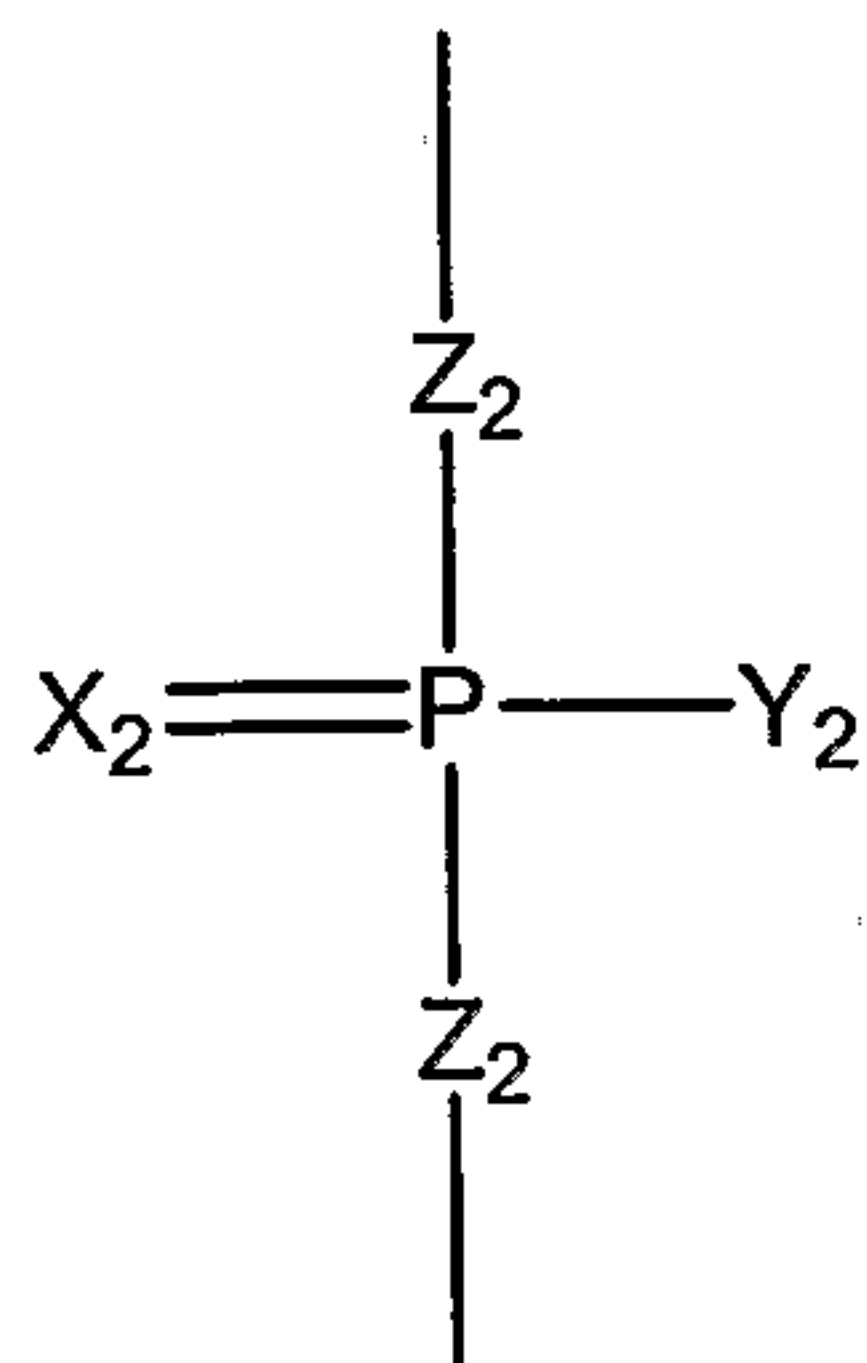


5 ; H; OH; OCH₃; W¹; an abasic nucleotide; or absent;

(a preferred A¹, especially with regard to anti-sense strands, is chosen from 5'-monophosphate ((HO)₂(O)P-O-5'), 5'-diphosphate ((HO)₂(O)P-O-P(HO)(O)-O-5'), 5'-triphosphate ((HO)₂(O)P-O-(HO)(O)P-O-P(HO)(O)-O-5'), 5'-guanosine cap (7-methylated or non-methylated) (7m-G-O-5'-(HO)(O)P-O-(HO)(O)P-O-P(HO)(O)-O-5'), 5'-adenosine cap (Appp), and any modified or unmodified nucleotide cap structure (N-O-5'-(HO)(O)P-O-(HO)(O)P-O-P(HO)(O)-O-5'), 5'-monothiophosphate (phosphorothioate; (HO)₂(S)P-O-5'), 5'-monodithiophosphate (phosphorodithioate; (HO)(HS)(S)P-O-5'), 5'-phosphorothiolate ((HO)₂(O)P-S-5'); any additional combination of oxygen/sulfur replaced monophosphate, diphosphate and triphosphates (*e.g.* 5'-alpha-thiotriphosphate, 5'-gamma-thiotriphosphate, etc.), 5'-phosphoramidates ((HO)₂(O)P-NH-5', (HO)(NH₂)(O)P-O-5'), 5'-alkylphosphonates (R=alkyl=methyl, ethyl, isopropyl, propyl, etc., *e.g.* RP(OH)(O)-O-5'-, (OH)₂(O)P-5'-CH₂-), 5'-

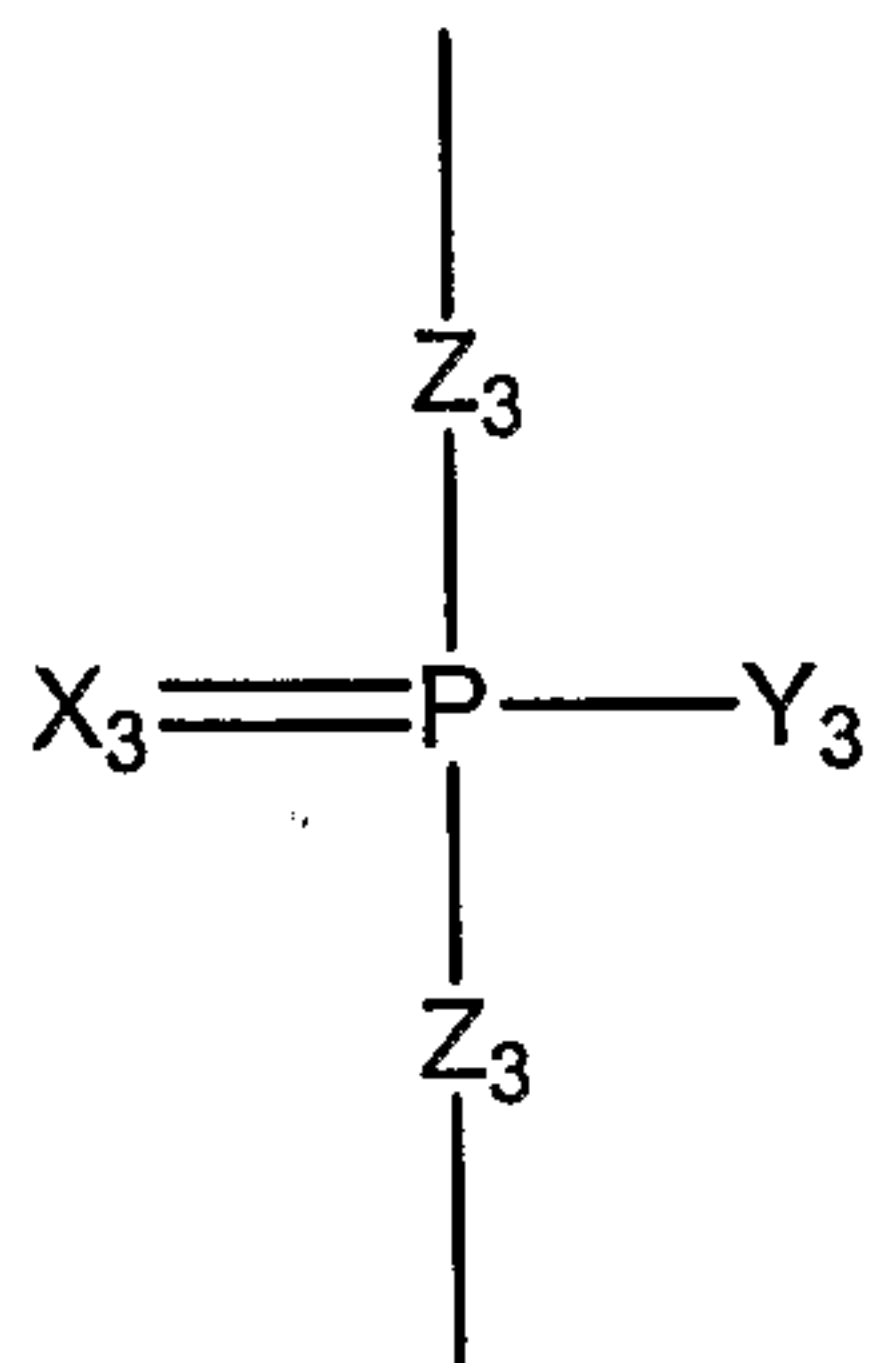
alkyletherphosphonates (R=alkylether=methoxymethyl (MeOCH₂-), ethoxymethyl, etc., *e.g.* RP(OH)(O)-O-5'-)).

A² is:



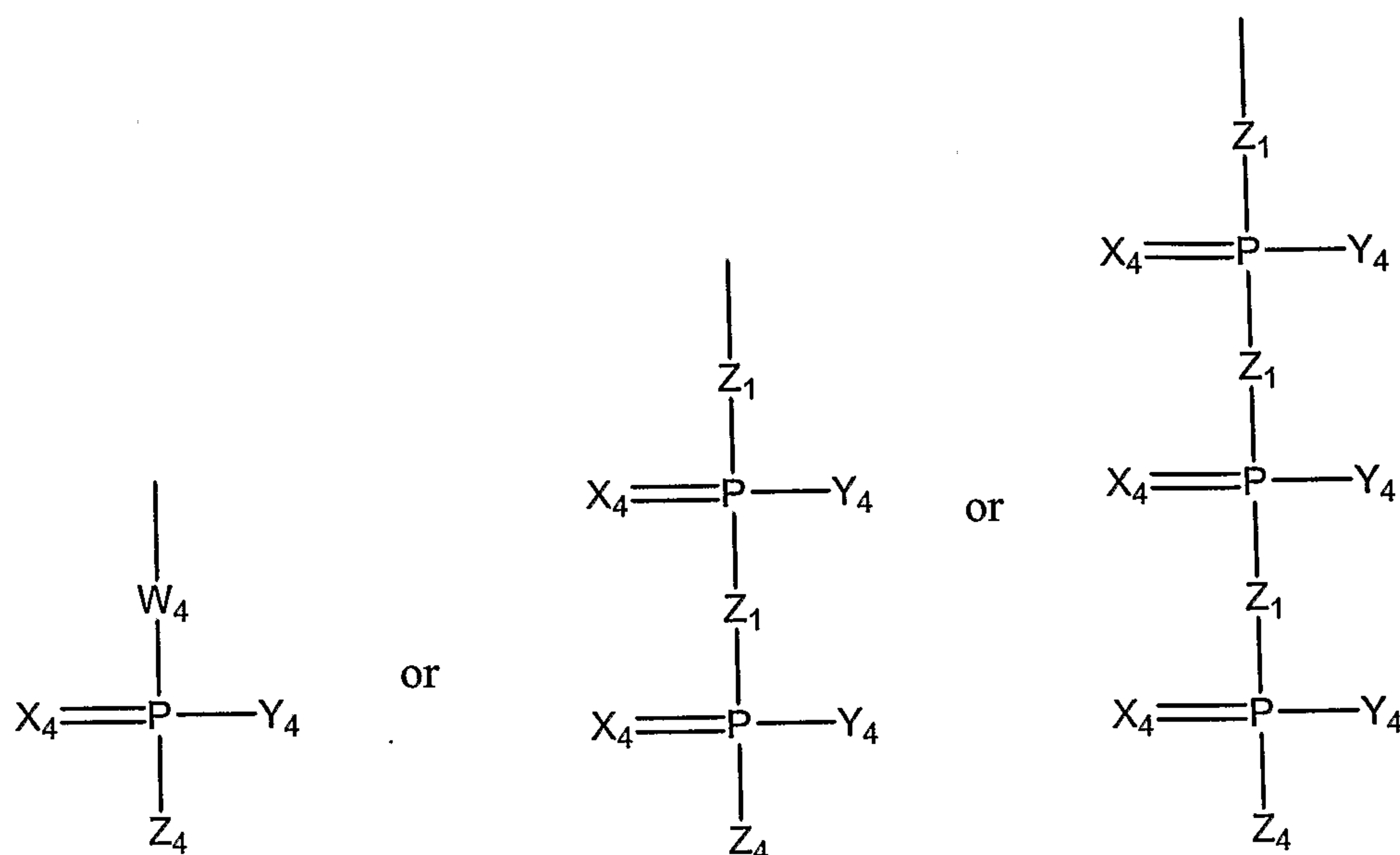
5

A³ is:



; and

A⁴ is:



5 ; H; Z⁴; an inverted nucleotide; an abasic nucleotide; or absent.

W¹ is OH, (CH₂)_nR¹⁰, (CH₂)_nNHR¹⁰, (CH₂)_nOR¹⁰, (CH₂)_nSR¹⁰; O(CH₂)_nR¹⁰;
 O(CH₂)_nOR¹⁰, O(CH₂)_nNR¹⁰, O(CH₂)_nSR¹⁰; O(CH₂)_nSS(CH₂)_nOR¹⁰, O(CH₂)_nC(O)OR¹⁰,
 NH(CH₂)_nR¹⁰; NH(CH₂)_nNR¹⁰; NH(CH₂)_nOR¹⁰, NH(CH₂)_nSR¹⁰; S(CH₂)_nR¹⁰, S(CH₂)_nNR¹⁰,
 S(CH₂)_nOR¹⁰, S(CH₂)_nSR¹⁰ O(CH₂CH₂O)_mCH₂CH₂OR¹⁰; O(CH₂CH₂O)_mCH₂CH₂NHR¹⁰,
 10 NH(CH₂CH₂NH)_mCH₂CH₂NHR¹⁰; Q-R¹⁰, O-Q-R¹⁰ N-Q-R¹⁰, S-Q-R¹⁰ or -O-. W⁴ is O, CH₂,
 NH, or S.

X¹, X², X³, and X⁴ are each, independently, O or S.

Y¹, Y², Y³, and Y⁴ are each, independently, OH, O⁻, OR⁸, S, Se, BH₃⁻, H, NHR⁹, N(R⁹)₂
 alkyl, cycloalkyl, aralkyl, aryl, or heteroaryl, each of which may be optionally substituted.

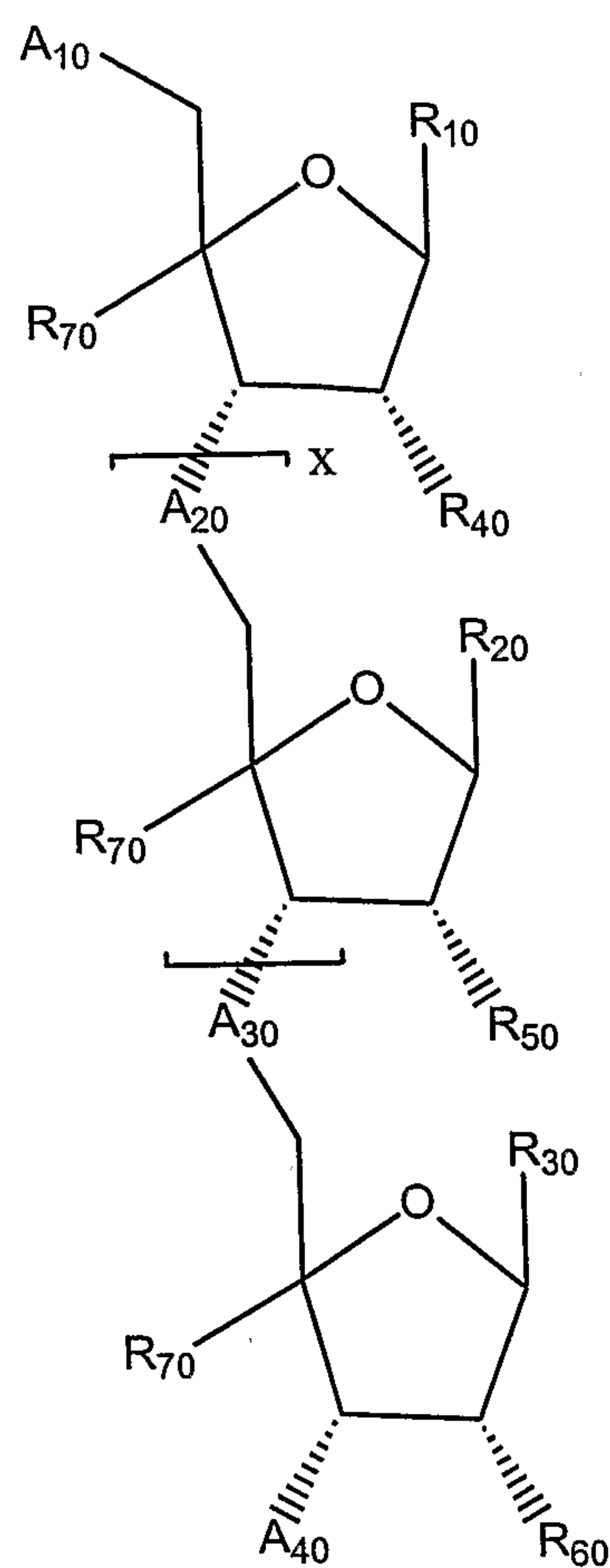
Z^1 , Z^2 , and Z^3 are each independently O, CH₂, NH, or S. Z^4 is OH, (CH₂)_nR¹⁰, (CH₂)_nNHR¹⁰, (CH₂)_nOR¹⁰, (CH₂)_nSR¹⁰; O(CH₂)_nR¹⁰; O(CH₂)_nOR¹⁰, O(CH₂)_nNR¹⁰, O(CH₂)_nSR¹⁰, O(CH₂)_nSS(CH₂)_nOR¹⁰, O(CH₂)_nC(O)OR¹⁰; NH(CH₂)_nR¹⁰; NH(CH₂)_nNR¹⁰; NH(CH₂)_nOR¹⁰, NH(CH₂)_nSR¹⁰; S(CH₂)_nR¹⁰, S(CH₂)_nNR¹⁰, S(CH₂)_nOR¹⁰, S(CH₂)_nSR¹⁰,
 5 O(CH₂CH₂O)_mCH₂CH₂OR¹⁰, O(CH₂CH₂O)_mCH₂CH₂NHR¹⁰, NH(CH₂CH₂NH)_mCH₂CH₂NHR¹⁰; Q-R¹⁰, O-Q-R¹⁰, N-Q-R¹⁰, S-Q-R¹⁰.

X is 5-100, chosen to comply with a length for an oligonucleotide agent described herein.

R⁷ is H; or is together combined with R⁴, R⁵, or R⁶ to form an [-O-CH₂-] covalently bound bridge between the sugar 2' and 4' carbons.

10 R⁸ is alkyl, cycloalkyl, aryl, aralkyl, heterocyclyl, heteroaryl, amino acid, or sugar; R⁹ is NH₂, alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, diheteroaryl amino, or amino acid; and R¹⁰ is H; fluorophore (pyrene, TAMRA, fluorescein, Cy3 or Cy5 dyes); sulfur, silicon, boron or ester protecting group; intercalating agents (*e.g.* acridines), cross-linkers (*e.g.* psoralene, mitomycin C), porphyrins (TPPC4, texaphyrin, Sapphyrin),
 15 polycyclic aromatic hydrocarbons (*e.g.*, phenazine, dihydrophenazine), artificial endonucleases (*e.g.* EDTA), lipophilic carriers (cholesterol, cholic acid, adamantane acetic acid, 1-pyrene butyric acid, dihydrotestosterone, 1,3-Bis-O(hexadecyl)glycerol, geranyloxyhexyl group, hexadecylglycerol, borneol, menthol, 1,3-propanediol, heptadecyl group, palmitic acid, myristic acid, O3-(oleoyl)lithocholic acid, O3-(oleoyl)cholenic acid, dimethoxytrityl, or phenoxazine) and
 20 peptide conjugates (*e.g.*, antennapedia peptide, Tat peptide), alkylating agents, phosphate, amino, mercapto, PEG (*e.g.*, PEG-40K), MPEG, [MPEG]₂, polyamino; alkyl, cycloalkyl, aryl, aralkyl, heteroaryl; radiolabelled markers, enzymes, haptens (*e.g.* biotin), transport/absorption facilitators (*e.g.*, aspirin, vitamin E, folic acid), synthetic ribonucleases (*e.g.*, imidazole, bisimidazole, histamine, imidazole clusters, acridine-imidazole conjugates, Eu³⁺ complexes of
 25 tetraazamacrocycles); or an oligonucleotide agent. M is 0-1,000,000, and n is 0-20. Q is a spacer selected from the group consisting of abasic sugar, amide, carboxy, oxyamine, oxyimine, thioether, disulfide, thiourea, sulfonamide, or morpholino, biotin or fluorescein reagents.

Preferred oligonucleotide agents in which the entire phosphate group has been replaced have the following structure (see Formula 3 below):

FORMULA 3

5 Referring to Formula 3, A¹⁰-A⁴⁰ is L-G-L; A¹⁰ and/or A⁴⁰ may be absent, in which L is a linker, wherein one or both L may be present or absent and is selected from the group consisting of CH₂(CH₂)_g; N(CH₂)_g; O(CH₂)_g; S(CH₂)_g. G is a functional group selected from the group consisting of siloxane, carbonate, carboxymethyl, carbamate, amide, thioether, ethylene oxide linker, sulfonate, sulfonamide, thioformacetal, formacetal, oxime, methyleneimino, methylenemethylimino, methylenehydrazo, methylenedimethylhydrazo and methyleneoxymethylimino.

10

R^{10} , R^{20} , and R^{30} are each, independently, H, (*i.e.* abasic nucleotides), adenine, guanine, cytosine and uracil, inosine, thymine, xanthine, hypoxanthine, nubularine, tubercidine, isoguanisine, 2-aminoadenine, 6-methyl and other alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 5-halouracil and cytosine, 5-propynyl
 5 uracil and cytosine, 6-azo uracil, cytosine and thymine, 5-uracil (pseudouracil), 4-thiouracil, 5-halouracil, 5-(2-aminopropyl)uracil, 5-amino allyl uracil, 8-halo, amino, thiol, thioalkyl, hydroxyl and other 8-substituted adenines and guanines, 5-trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine, 5-substituted pyrimidines, 6-azapyrimidines and N-2, N-6 and O-6 substituted purines, including 2-aminopropyladenine, 5-propynyluracil
 10 and 5-propynylcytosine, dihydrouracil, 3-deaza-5-azacytosine, 2-aminopurine, 5-alkyluracil, 7-alkylguanine, 5-alkyl cytosine, 7-deazaadenine, 7-deazaguanine, N6, N6-dimethyladenine, 2,6-diaminopurine, 5-amino-allyl-uracil, N3-methyluracil substituted 1,2,4-triazoles, 2-pyridinone, 5-nitroindole, 3-nitropyrrole, 5-methoxyuracil, uracil-5-oxyacetic acid, 5-methoxycarbonylmethyluracil, 5-methyl-2-thiouracil, 5-methoxycarbonylmethyl-2-thiouracil, 5-
 15 methylaminomethyl-2-thiouracil, 3-(3-amino-3carboxypropyl)uracil, 3-methylcytosine, 5-methylcytosine, N⁴-acetyl cytosine, 2-thiocytosine, N6-methyladenine, N6-isopentyladenine, 2-methylthio-N6-isopentenyladenine, N-methylguanines, or O-alkylated bases.

R^{40} , R^{50} , and R^{60} are each, independently, OR^8 , $O(CH_2CH_2O)_mCH_2CH_2OR^8$; $O(CH_2)_nR^9$; $O(CH_2)_nOR^9$, H; halo; NH_2 ; NHR^8 ; $N(R^8)_2$; $NH(CH_2CH_2NH)_mCH_2CH_2R^9$; $NHC(O)R^8$; cyano; mercapto, SR^7 ; alkyl-thio-alkyl; alkyl, aralkyl, cycloalkyl, aryl, heteroaryl, alkenyl, alkynyl, each
 20 of which may be optionally substituted with halo, hydroxy, oxo, nitro, haloalkyl, alkyl, alkaryl, aryl, aralkyl, alkoxy, aryloxy, amino, alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, diheteroaryl amino, acylamino, alkylcarbamoyl, arylcarbamoyl, aminoalkyl, alkoxycarbonyl, carboxy, hydroxyalkyl, alkanesulfonyl, alkanesulfonamido, arenesulfonamido, aralkylsulfonamido, alkylcarbonyl, acyloxy, cyano, and ureido groups; or R^{40} ,
 25 R^{50} , or R^{60} together combine with R^{70} to form an $[-O-CH_2-]$ covalently bound bridge between the sugar 2' and 4' carbons.

X is 5-100 or chosen to comply with a length for an oligonucleotide agent described herein.

R^{70} is H; or is together combined with R^{40} , R^{50} , or R^{60} to form an $[-O-CH_2-]$ covalently bound bridge between the sugar 2' and 4' carbons.

R^8 is alkyl, cycloalkyl, aryl, aralkyl, heterocyclyl, heteroaryl, amino acid, or sugar; and R^9 is NH_2 , alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, 5 diheteroaryl amino, or amino acid. M is 0-1,000,000, n is 0-20, and g is 0-2.

Preferred nucleoside surrogates have the following structure (see Formula 4 below):



FORMULA 4

10

S is a nucleoside surrogate selected from the group consisting of morpholino, cyclobutyl, pyrrolidine and peptide nucleic acid. L is a linker and is selected from the group consisting of $CH_2(CH_2)_g$; $N(CH_2)_g$; $O(CH_2)_g$; $S(CH_2)_g$; $-C(O)(CH_2)_n$ -or may be absent. M is an amide bond; sulfonamide; sulfinate; phosphate group; modified phosphate group as described herein; or may 15 be absent.

R^{100} , R^{200} , and R^{300} are each, independently, H (*i.e.*, abasic nucleotides), adenine, guanine, cytosine and uracil, inosine, thymine, xanthine, hypoxanthine, nucleobase, tubercidine, isoguanisine, 2-aminoadenine, 6-methyl and other alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 5-halouracil and cytosine, 5-propynyl 20 uracil and cytosine, 6-azo uracil, cytosine and thymine, 5-uracil (pseudouracil), 4-thiouracil, 5-halouracil, 5-(2-aminopropyl)uracil, 5-amino allyl uracil, 8-halo, amino, thiol, thioalkyl, hydroxyl and other 8-substituted adenines and guanines, 5-trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine, 5-substituted pyrimidines, 6-azapyrimidines and N-2, N-6 and O-6 substituted purines, including 2-aminopropyladenine, 5-propynyluracil 25 and 5-propynylcytosine, dihydrouracil, 3-deaza-5-azacytosine, 2-aminopurine, 5-alkyluracil, 7-alkylguanine, 5-alkyl cytosine, 7-deazaadenine, 7-deazaguanine, N6, N6-dimethyladenine, 2,6-

diaminopurine, 5-amino-allyl-uracil, N3-methyluracil substituted 1, 2, 4,-triazoles, 2-pyridinones, 5-nitroindole, 3-nitropyrrole, 5-methoxyuracil, uracil-5-oxyacetic acid, 5-methoxycarbonylmethyluracil, 5-methyl-2-thiouracil, 5-methoxycarbonylmethyl-2-thiouracil, 5-methylaminomethyl-2-thiouracil, 3-(3-amino-3carboxypropyl)uracil, 3-methylcytosine, 5-methylcytosine, N⁴-acetyl cytosine, 2-thiocytosine, N6-methyladenine, N6-isopentyladenine, 2-methylthio-N6-isopentenyladenine, N-methylguanines, or O-alkylated bases.

X is 5-100, or chosen to comply with a length for an oligonucleotide agent described herein; and g is 0-2.

Nuclease resistant monomers

The monomers and methods described herein can be used to prepare an oligonucleotide agent, that incorporates a nuclease resistant monomer (NRM).

An oligonucleotide agent can include monomers which have been modified so as to inhibit degradation, *e.g.*, by nucleases, *e.g.*, endonucleases or exonucleases, found in the body of a subject. These monomers are referred to herein as NRMs, or nuclease resistance promoting monomers or modifications. In many cases these modifications will modulate other properties of the oligonucleotide agent as well, *e.g.*, the ability to interact with a protein, *e.g.*, a transport protein, *e.g.*, serum albumin, or a member of the RISC (RNA-induced Silencing Complex), or the ability of the first and second sequences to form a duplex with one another or to form a duplex with another sequence, *e.g.*, a target molecule.

While not wishing to be bound by theory, it is believed that modifications of the sugar, base, and/or phosphate backbone in an oligonucleotide agent can enhance endonuclease and exonuclease resistance, and can enhance interactions with transporter proteins and one or more of the functional components of the RISC complex. Preferred modifications are those that increase exonuclease and endonuclease resistance and thus prolong the half-life of the oligonucleotide agent prior to interaction with the RISC complex, but at the same time do not render the oligonucleotide agent resistant to endonuclease activity in the RISC complex. Again, while not wishing to be bound by any theory, it is believed that placement of the modifications at or near the 3' and/or 5' end of the oligonucleotide agent can result in agents that meet the preferred nuclease resistance criteria delineated above.

Modifications that can be useful for producing oligonucleotide agents that meet the preferred nuclease resistance criteria delineated above can include one or more of the following chemical and/or stereochemical modifications of the sugar, base, and/or phosphate backbone:

5 (i) chiral (S_P) thioates. Thus, preferred NRMs include nucleotide dimers with an enriched for or having a pure chiral form of a modified phosphate group containing a heteroatom at the nonbridging position, *e.g.*, S_P or R_P , at the position X, where this is the position normally occupied by the oxygen. The atom at X can also be S, Se, Nr_2 , or Br_3 . When X is S, enriched or chirally pure S_P linkage is preferred. Enriched means at least 70, 80, 90, 95, or 99% of the preferred form. Such NRMs are discussed in more detail below;

10 (ii) attachment of one or more cationic groups to the sugar, base, and/or the phosphorus atom of a phosphate or modified phosphate backbone moiety. Thus, preferred NRMs include monomers at the terminal position derivatized at a cationic group. As the 5' end of an oligonucleotide agent should have a terminal -OH or phosphate group, this NRM is preferably not used at the 5' end of the agent. The group should be attached at a position on the base which
15 minimizes interference with H bond formation and hybridization, *e.g.*, away from the face which interacts with the complementary base on the other strand, *e.g.*, at the 5' position of a pyrimidine or a 7-position of a purine. These are discussed in more detail below;

(iii) nonphosphate linkages at the termini. Thus, preferred NRMs include Non-phosphate linkages, *e.g.*, a linkage of 4 atoms which confers greater resistance to cleavage than does a
20 phosphate bond. Examples include 3' $CH_2-NCH_3-O-CH_2-5'$ and 3' $CH_2-NH-(O=)-CH_2-5'$;

(iv) 3'-bridging thiophosphates and 5'-bridging thiophosphates. Thus, preferred NRM's can included these structures;

(v) L-RNA, 2'-5' linkages, inverted linkages, α -nucleosides. Thus, other preferred NRM's include: L nucleosides and dimeric nucleotides derived from L-nucleosides; 2'-5'
25 phosphate, non-phosphate and modified phosphate linkages (*e.g.*, thiophosphates, phosphoramidates and boronophosphates); dimers having inverted linkages, *e.g.*, 3'-3' or 5'-5' linkages; monomers having an α linkage at the 1' site on the sugar, *e.g.*, the structures described herein having an α linkage;

(vi) conjugate groups. Thus, preferred NRM's can include *e.g.*, a targeting moiety or a
30 conjugated ligand described herein, *e.g.*, conjugated with the monomer, *e.g.*, through the sugar, base, or backbone;

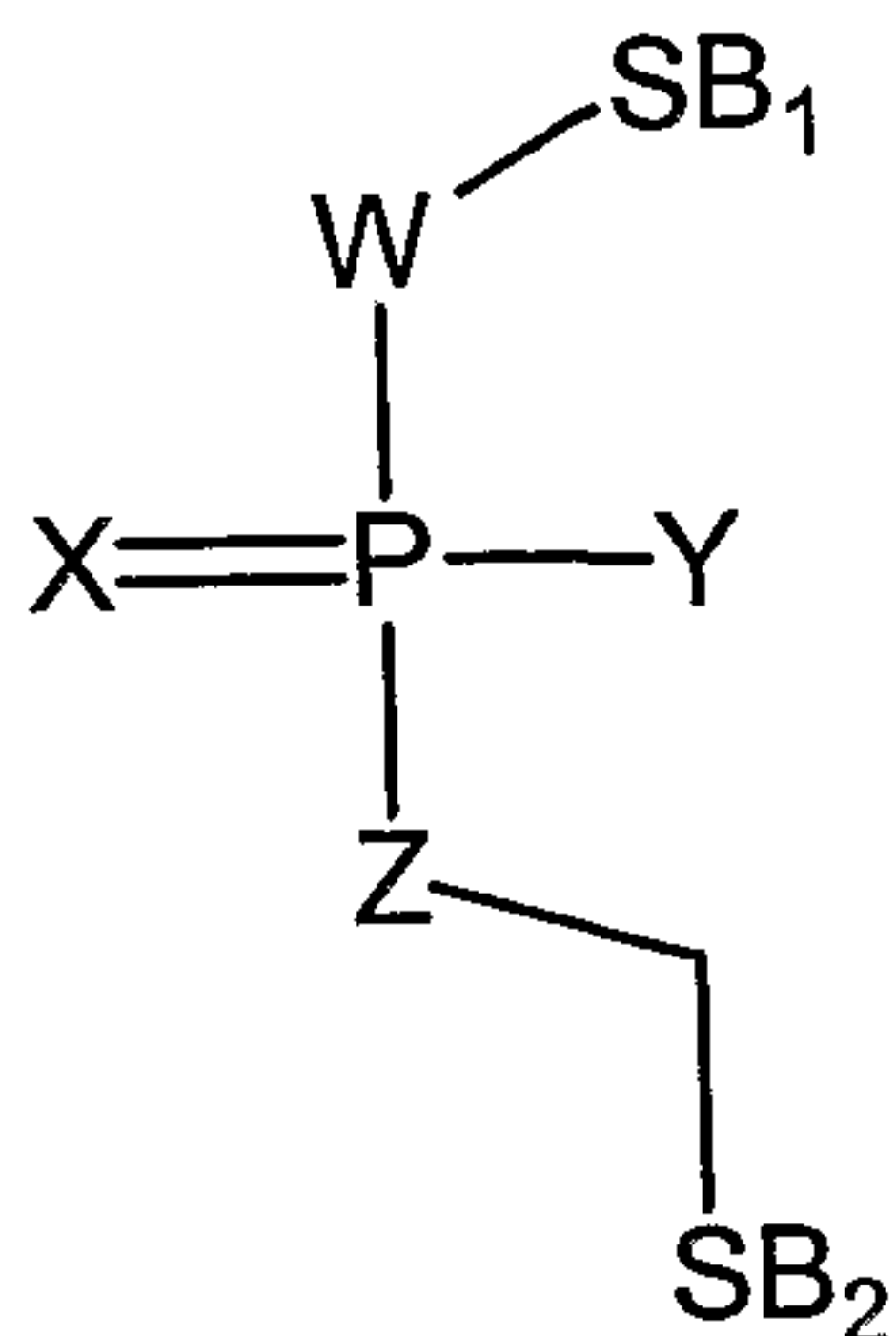
(vi) abasic linkages. Thus, preferred NRM's can include an abasic monomer, *e.g.*, an abasic monomer as described herein (*e.g.*, a nucleobaseless monomer); an aromatic or heterocyclic or polyheterocyclic aromatic monomer as described herein; and

5 (vii) 5'-phosphonates and 5'-phosphate prodrugs. Thus, preferred NRM's include monomers, preferably at the terminal position, *e.g.*, the 5' position, in which one or more atoms of the phosphate group are derivatized with a protecting group, which protecting group or groups, are removed as a result of the action of a component in the subject's body, *e.g.*, a carboxyesterase or an enzyme present in the subject's body. *E.g.*, a phosphate prodrug in which a carboxy esterase cleaves the protected molecule resulting in the production of a thioate anion
10 which attacks a carbon adjacent to the O of a phosphate and resulting in the production of an unprotected phosphate.

One or more different NRM modifications can be introduced into an oligonucleotide agent or into a sequence of an oligonucleotide agent. An NRM modification can be used more than once in a sequence or in an oligonucleotide agent. As some NRM's interfere with
15 hybridization the total number incorporated, should be such that acceptable levels of oligonucleotide agent/target RNA duplex formation are maintained.

Chiral S_P Thioates

A modification can include the alteration, *e.g.*, replacement, of one or both of the non-
20 linking (X and Y) phosphate oxygens and/or of one or more of the linking (W and Z) phosphate oxygens. Formula X below depicts a phosphate moiety linking two sugar/sugar surrogate-base moieties, SB₁ and SB₂.



FORMULA X

5

In certain embodiments, one of the non-linking phosphate oxygens in the phosphate backbone moiety (X and Y) can be replaced by any one of the following: S, Se, BR₃ (R is hydrogen, alkyl, aryl, etc.), C (i.e., an alkyl group, an aryl group, etc.), H, NR₂ (R is hydrogen, alkyl, aryl, etc.), or OR (R is alkyl or aryl). The phosphorus atom in an unmodified phosphate group is achiral. However, replacement of one of the non-linking oxygens with one of the above atoms or groups of atoms renders the phosphorus atom chiral; in other words a phosphorus atom in a phosphate group modified in this way is a stereogenic center. The stereogenic phosphorus atom can possess either the “R” configuration (herein R_P) or the “S” configuration (herein S_P). Thus if 60% of a population of stereogenic phosphorus atoms have the R_P configuration, then the remaining 40% of the population of stereogenic phosphorus atoms have the S_P configuration.

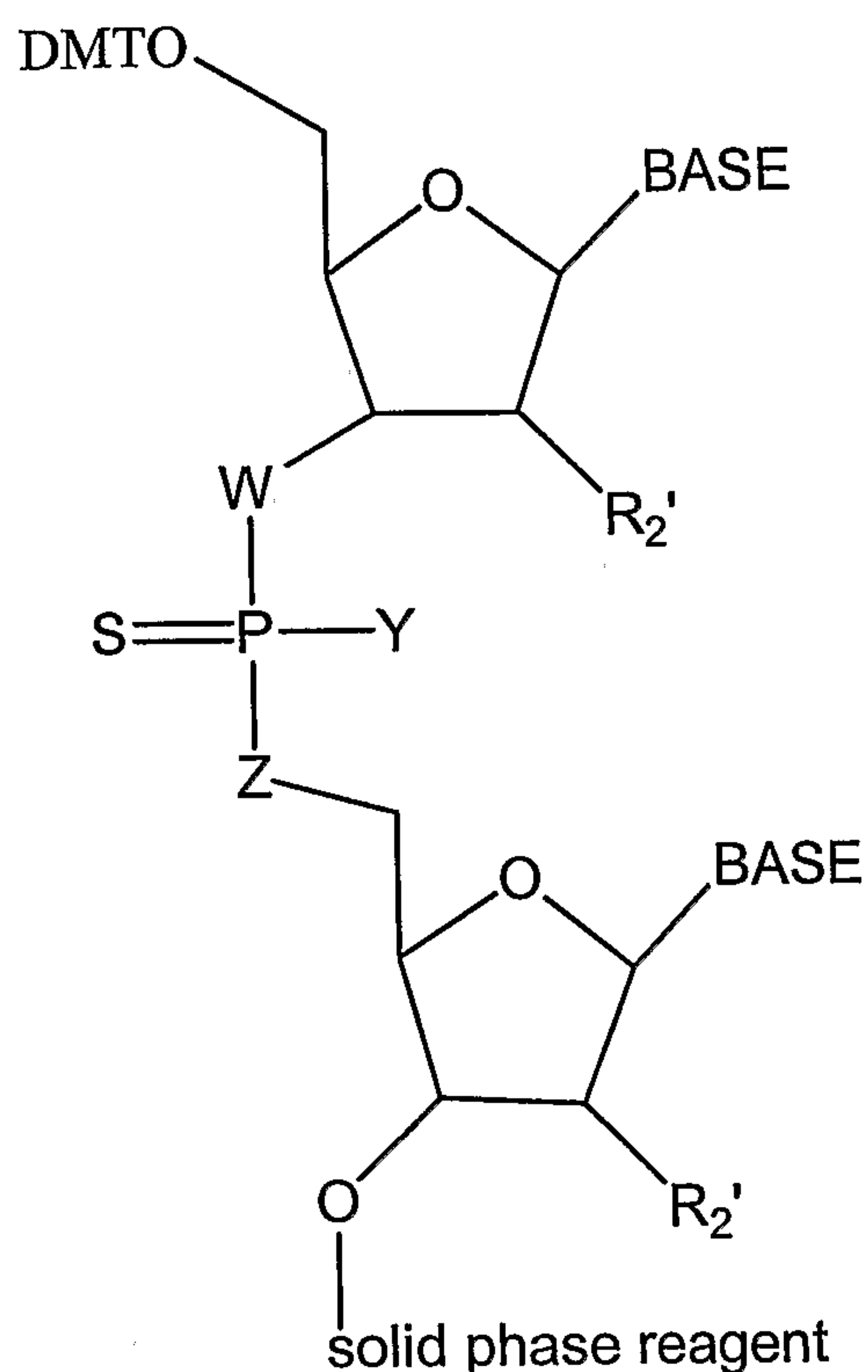
In some embodiments, oligonucleotide agents have phosphate groups in which a phosphate non-linking oxygen has been replaced by another atom or group of atoms, may contain a population of stereogenic phosphorus atoms in which at least about 50% of these atoms (e.g., at least about 60% of these atoms, at least about 70% of these atoms, at least about 80% of these atoms, at least about 90% of these atoms, at least about 95% of these atoms, at least about 98% of these atoms, at least about 99% of these atoms) have the S_P configuration. Alternatively, oligonucleotide agents having phosphate groups in which a phosphate non-linking oxygen has been replaced by another atom or group of atoms may contain a population of stereogenic phosphorus atoms in which at least about 50% of these atoms (e.g., at least about 60% of these

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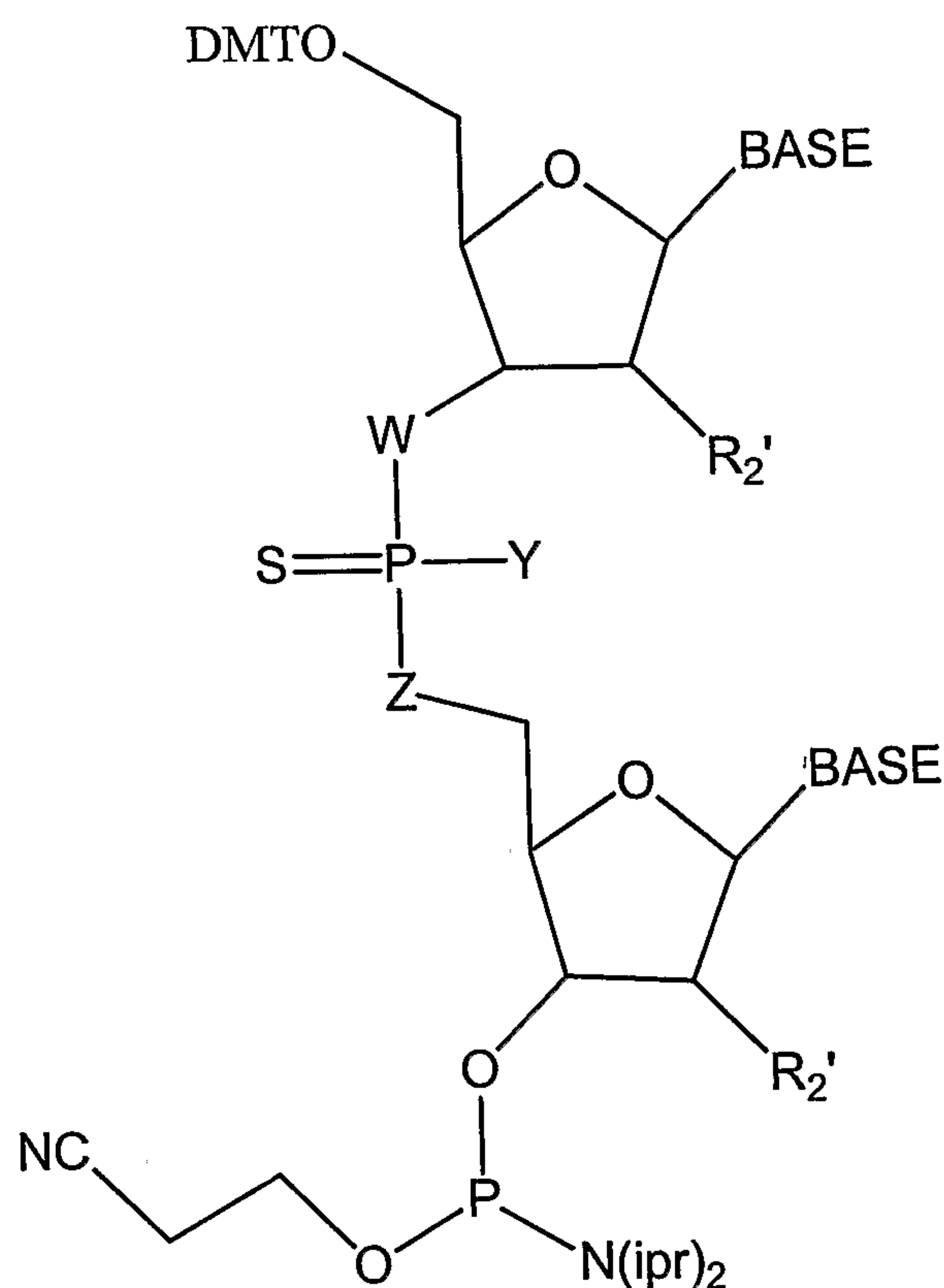
atoms, at least about 70% of these atoms, at least about 80% of these atoms, at least about 90% of these atoms, at least about 95% of these atoms, at least about 98% of these atoms, at least about 99% of these atoms) have the R_P configuration. In other embodiments, the population of stereogenic phosphorus atoms may have the S_P configuration and may be substantially free of stereogenic phosphorus atoms having the R_P configuration. In still other embodiments, the population of stereogenic phosphorus atoms may have the R_P configuration and may be substantially free of stereogenic phosphorus atoms having the S_P configuration. As used herein, the phrase “substantially free of stereogenic phosphorus atoms having the R_P configuration” means that moieties containing stereogenic phosphorus atoms having the R_P configuration cannot be detected by conventional methods known in the art (chiral HPLC, ^1H NMR analysis using chiral shift reagents, etc.). As used herein, the phrase “substantially free of stereogenic phosphorus atoms having the S_P configuration” means that moieties containing stereogenic phosphorus atoms having the S_P configuration cannot be detected by conventional methods known in the art (chiral HPLC, ^1H NMR analysis using chiral shift reagents, etc.).

In a preferred embodiment, modified oligonucleotide agents contain a phosphorothioate group, i.e., a phosphate groups in which a phosphate non-linking oxygen has been replaced by a sulfur atom. In an especially preferred embodiment, the population of phosphorothioate stereogenic phosphorus atoms may have the S_P configuration and be substantially free of stereogenic phosphorus atoms having the R_P configuration.

Phosphorothioates may be incorporated into oligonucleotide agents using dimers *e.g.*, formulas X-1 and X-2. The former can be used to introduce phosphorothioate



X-1



X-2

5

at the 3' end of a strand, while the latter can be used to introduce this modification at the 5' end or at a position that occurs *e.g.*, 1, 2, 3, 4, 5, or 6 nucleotides from either end of the strand. In the above formulas, Y can be 2-cyanoethoxy, W and Z can be O, R₂' can be, *e.g.*, a substituent that can impart the C-3 endo configuration to the sugar (*e.g.*, OH, F, OCH₃), DMT is dimethoxytrityl, and "BASE" can be a natural, unusual, or a universal base.

10

X-1 and X-2 can be prepared using chiral reagents or directing groups that can result in phosphorothioate-containing dimers having a population of stereogenic phosphorus atoms having essentially only the R_P configuration (*i.e.*, being substantially free of the S_P configuration) or only the S_P configuration (*i.e.*, being substantially free of the R_P configuration). Alternatively, dimers can be prepared having a population of stereogenic phosphorus atoms in which about

15

50% of the atoms have the R_P configuration and about 50% of the atoms have the S_P configuration. Dimers having stereogenic phosphorus atoms with the R_P configuration can be identified and separated from dimers having stereogenic phosphorus atoms with the S_P configuration using *e.g.*, enzymatic degradation and/or conventional chromatography techniques.

5 Cationic Groups

Modifications can also include attachment of one or more cationic groups to the sugar, base, and/or the phosphorus atom of a phosphate or modified phosphate backbone moiety. A cationic group can be attached to any atom capable of substitution on a natural, unusual or universal base. A preferred position is one that does not interfere with hybridization, *i.e.*, does not interfere with the hydrogen bonding interactions needed for base pairing. A cationic group can be attached *e.g.*, through the C2' position of a sugar or analogous position in a cyclic or acyclic sugar surrogate. Cationic groups can include *e.g.*, protonated amino groups, derived from *e.g.*, O-AMINE (AMINE = NH_2 ; alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, or diheteroaryl amino, ethylene diamine, polyamino); aminoalkoxy, *e.g.*, $O(CH_2)_n$ AMINE, (*e.g.*, AMINE = NH_2 ; alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, or diheteroaryl amino, ethylene diamine, polyamino); amino (*e.g.* NH_2 ; alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, diheteroaryl amino, or amino acid); or $NH(CH_2CH_2NH)_nCH_2CH_2$ -AMINE (AMINE = NH_2 ; alkylamino, dialkylamino, heterocyclyl, arylamino, diaryl amino, heteroaryl amino, or diheteroaryl amino).

Nonphosphate Linkages

Modifications can also include the incorporation of nonphosphate linkages at the 5' and/or 3' end of a strand. Examples of nonphosphate linkages which can replace the phosphate group include methyl phosphonate, hydroxylamino, siloxane, carbonate, carboxymethyl, carbamate, amide, thioether, ethylene oxide linker, sulfonate, sulfonamide, thioformacetal, formacetal, oxime, methyleneimino, methylenemethylimino, methylenehydrazo, methylenedimethylhydrazo and methyleneoxymethylimino. Preferred replacements include the methyl phosphonate and hydroxylamino groups.

3'-bridging thiophosphates and 5'-bridging thiophosphates; locked-RNA, 2'-5' linkages, inverted linkages, α -nucleosides; conjugate groups; abasic linkages; and 5'-phosphonates and 5'-phosphate prodrugs are also linkages that can be included in oligonucleotide agents.

Referring to formula X above, modifications can include replacement of one of the
5 bridging or linking phosphate oxygens in the phosphate backbone moiety (W and Z). Unlike the situation where only one of X or Y is altered, the phosphorus center in the phosphorodithioates is achiral which precludes the formation of oligonucleotide agents containing a stereogenic phosphorus atom.

Modifications can also include linking two sugars via a phosphate or modified phosphate
10 group through the 2' position of a first sugar and the 5' position of a second sugar. Also contemplated are inverted linkages in which both a first and second sugar are each linked through the respective 3' positions. Modified RNA's can also include "abasic" sugars, which lack a nucleobase at C-1'. The sugar group can also contain one or more carbons that possess the opposite stereochemical configuration than that of the corresponding carbon in ribose. Thus, a
15 modified oligonucleotide agent can include nucleotides containing *e.g.*, arabinose, as the sugar. In another subset of this modification, the natural, unusual, or universal base may have the α -configuration. Modifications can also include L-RNA.

Modifications can also include 5'-phosphonates, *e.g.*, $P(O)(O^-)_2-X-C^{5'}$ -sugar ($X=CH_2$, CF_2 , CHF and 5'-phosphate prodrugs, *e.g.*, $P(O)[OCH_2CH_2SC(O)R]_2CH_2C^{5'}$ -sugar. In the
20 latter case, the prodrug groups may be decomposed *via* reaction first with carboxy esterases. The remaining ethyl thiolate group via intramolecular S_N2 displacement can depart as episulfide to afford the underivatized phosphate group.

Modification can also include the addition of conjugating groups described elsewhere herein, which are preferably attached to an oligonucleotide agent through any amino group
25 available for conjugation.

Nuclease resistant modifications include some which can be placed only at the terminus and others which can go at any position. Generally, these modifications can inhibit hybridization so it is preferably to use them only in terminal regions, and preferable to not use them at the cleavage site or in the cleavage region of a sequence.

30 Modifications which interfere with or inhibit endonuclease cleavage should not be inserted in the region of an oligonucleotide agent which is subject to RISC mediated cleavage,

e.g., the cleavage site or the cleavage region. As used herein cleavage site refers to the nucleotide on either side of the cleavage site on the target or on the oligonucleotide agent strand which hybridizes to it. Cleavage region means an nucleotide with 1, 2, or 3 nucleotides of the cleavage site, in either direction.)

5 Such modifications can be introduced into the terminal regions, *e.g.*, at the terminal position or with 2, 3, 4, or 5 positions of the terminus.

 An oligonucleotide agent can have the following:

 an NRM modification at or within 1, 2, 3, 4, 5, or 6 positions from the 3' end;

 an NRM modification at or within 1, 2, 3, 4, 5, or 6 positions from the 5' end (5' end

10 NRM modifications are preferentially not at the terminus but rather at a position 1, 2, 3, 4, 5, or 6 away from the 5' terminus of the oligonucleotide agent);

 an NRM modification at or within 1, 2, 3, 4, 5, or 6 positions from the 3' end and which has a NRM modification at or within 1, 2, 3, 4, 5, or 6 positions from the 5' end;

 an NRM modification at the cleavage site or in the cleavage region;

15 an NRM modification at the cleavage site or in the cleavage region and one or more of an NRM modification at or within 1, 2, 3, 4, 5, or 6 positions from the 3' end, an NRM modification at or within 1, 2, 3, 4, 5, or 6 positions from the 5' end, or NRM modifications at or within 1, 2, 3, 4, 5, or 6 positions from both the 3' and the 5' end (5' end NRM modifications are preferentially not at the terminus but rather at a position 1, 2, 3, 4, 5, or 6 away from the 5' terminus of the oligonucleotide agent).

Ribose Mimics

 The monomers and methods described herein can be used to prepare an oligonucleotide agent, that incorporates a ribose mimic.

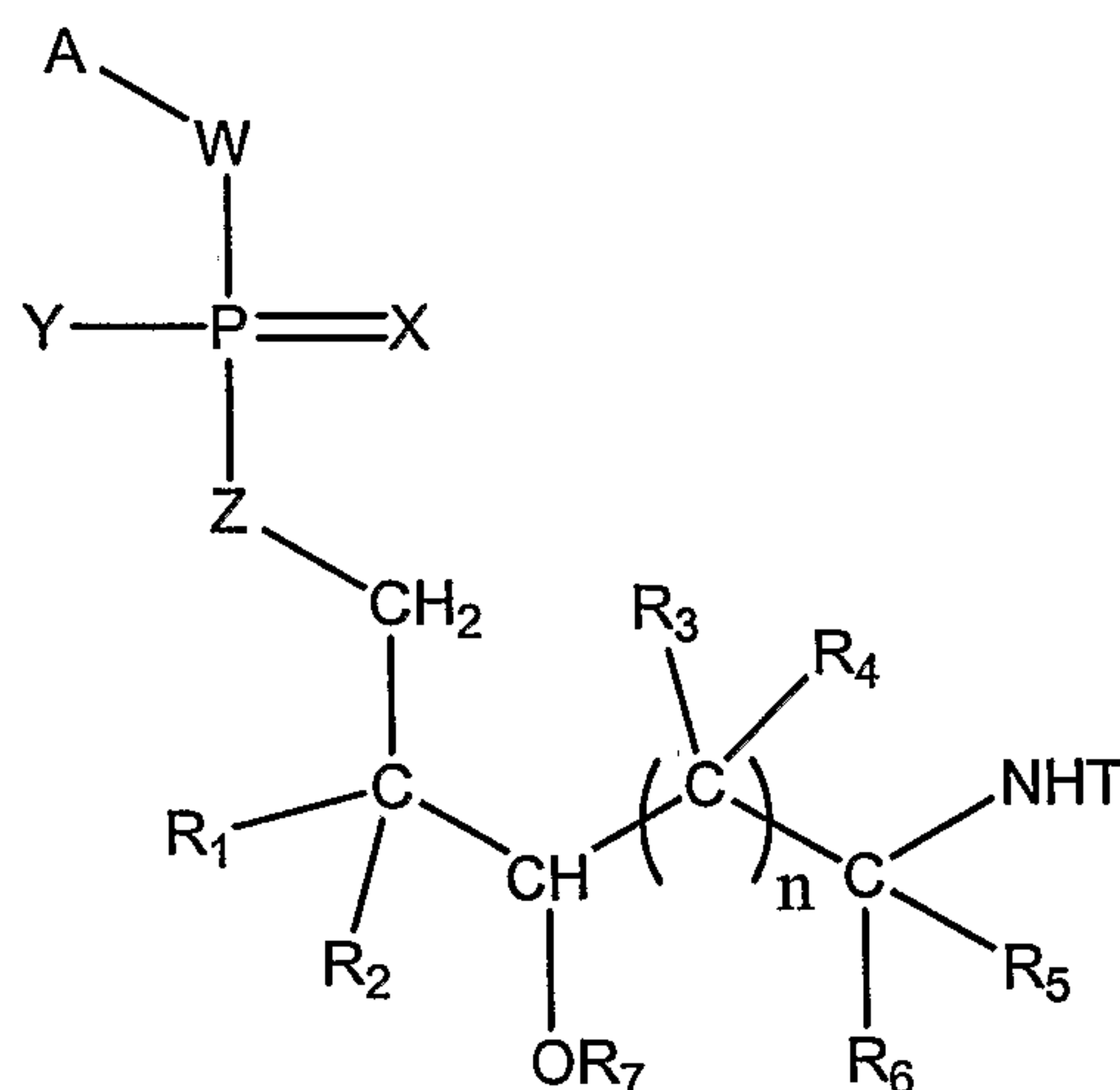
25 Thus, an aspect of the invention features an oligonucleotide agent that includes a secondary hydroxyl group, which can increase efficacy and/or confer nuclease resistance to the agent. Nucleases, *e.g.*, cellular nucleases, can hydrolyze nucleic acid phosphodiester bonds, resulting in partial or complete degradation of the nucleic acid. The secondary hydroxy group confers nuclease resistance to an oligonucleotide agent by rendering the oligonucleotide agent less prone to nuclease degradation relative to an oligonucleotide agent that lacks the

30 modification. While not wishing to be bound by theory, it is believed that the presence of a

secondary hydroxyl group on the oligonucleotide agent can act as a structural mimic of a 3' ribose hydroxyl group, thereby causing it to be less susceptible to degradation.

The secondary hydroxyl group refers to an "OH" radical that is attached to a carbon atom substituted by two other carbons and a hydrogen. The secondary hydroxyl group that confers
5 nuclease resistance as described above can be part of any acyclic carbon-containing group. The hydroxyl may also be part of any cyclic carbon-containing group, and preferably one or more of the following conditions is met (1) there is no ribose moiety between the hydroxyl group and the terminal phosphate group or (2) the hydroxyl group is not on a sugar moiety which is coupled to a base. The hydroxyl group is located at least two bonds (*e.g.*, at least three bonds away, at least
10 four bonds away, at least five bonds away, at least six bonds away, at least seven bonds away, at least eight bonds away, at least nine bonds away, at least ten bonds away, etc.) from the terminal phosphate group phosphorus of the oligonucleotide agent. In preferred embodiments, there are five intervening bonds between the terminal phosphate group phosphorus and the secondary hydroxyl group.

Preferred oligonucleotide agent delivery modules with five intervening bonds between the terminal phosphate group phosphorus and the secondary hydroxyl group have the following structure (see formula Y below):



(Y)

5

Referring to formula Y, A is an oligonucleotide agent, including any oligonucleotide agent described herein. The oligonucleotide agent may be connected directly or indirectly (e.g., through a spacer or linker) to “W” of the phosphate group. These spacers or linkers can include e.g., $-(CH_2)_n-$, $-(CH_2)_nN-$, $-(CH_2)_nO-$, $-(CH_2)_nS-$, $O(CH_2CH_2O)_nCH_2CH_2OH$ (e.g., $n = 3$ or 6), abasic sugars, amide, carboxy, amine, oxyamine, oxyimine, thioether, disulfide, thiourea, sulfonamide, or morpholino, or biotin and fluorescein reagents.

The oligonucleotide agents can have a terminal phosphate group that is unmodified (e.g., W, X, Y, and Z are O) or modified. In a modified phosphate group, W and Z can be independently NH, O, or S; and X and Y can be independently S, Se, BH_3^- , C_1-C_6 alkyl, C_6-C_{10} aryl, H, O, O^- , alkoxy or amino (including alkylamino, arylamino, etc.). Preferably, W, X and Z are O and Y is S.

R_1 and R_3 are each, independently, hydrogen; or C_1-C_{100} alkyl, optionally substituted with hydroxyl, amino, halo, phosphate or sulfate and/or may be optionally inserted with N, O, S, alkenyl or alkynyl.

20

R_2 is hydrogen; C_1 - C_{100} alkyl, optionally substituted with hydroxyl, amino, halo, phosphate or sulfate and/or may be optionally inserted with N, O, S, alkenyl or alkynyl; or, when n is 1, R_2 may be taken together with R_4 or R_6 to form a ring of 5-12 atoms.

5 R_4 is hydrogen; C_1 - C_{100} alkyl, optionally substituted with hydroxyl, amino, halo, phosphate or sulfate and/or may be optionally inserted with N, O, S, alkenyl or alkynyl; or, when n is 1, R_4 may be taken together with R_2 or R_5 to form a ring of 5-12 atoms.

R_5 is hydrogen, C_1 - C_{100} alkyl optionally substituted with hydroxyl, amino, halo, phosphate or sulfate and/or may be optionally inserted with N, O, S, alkenyl or alkynyl; or, when n is 1, R_5 may be taken together with R_4 to form a ring of 5-12 atoms.

10 R_6 is hydrogen, C_1 - C_{100} alkyl, optionally substituted with hydroxyl, amino, halo, phosphate or sulfate and/or may be optionally inserted with N, O, S, alkenyl or alkynyl, or, when n is 1, R_6 may be taken together with R_2 to form a ring of 6-10 atoms;

R_7 is hydrogen, C_1 - C_{100} alkyl, or $C(O)(CH_2)_qC(O)NHR_9$; T is hydrogen or a functional group; n and q are each independently 1-100; R_8 is C_1 - C_{10} alkyl or C_6 - C_{10} aryl; and R_9 is
15 hydrogen, C_1 - C_{10} alkyl, C_6 - C_{10} aryl or a solid support agent.

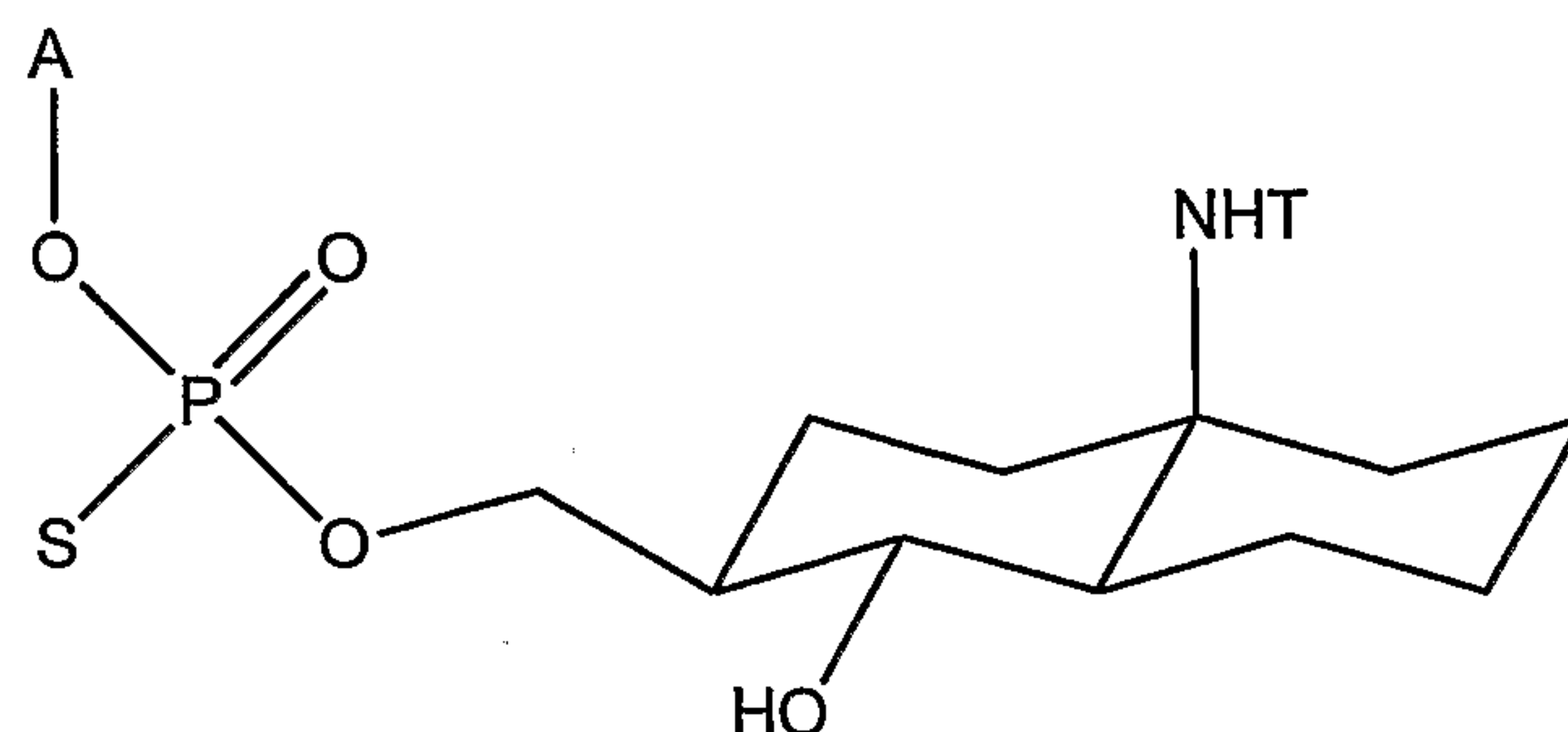
Preferred embodiments may include one of more of the following subsets of oligonucleotide agent delivery modules.

In one subset of oligonucleotide agent delivery modules, A can be connected directly or indirectly through a terminal 3' or 5' ribose sugar carbon of the oligonucleotide agent.

20 In another subset of Oligonucleotide agent delivery modules, X , W , and Z are O and Y is S.

In still yet another subset of oligonucleotide agent delivery modules, n is 1, and R_2 and R_6 are taken together to form a ring containing six atoms and R_4 and R_5 are taken together to form a ring containing six atoms. Preferably, the ring system is a *trans*-decalin. For example, the Oligonucleotide agent delivery module of this subset can include a compound of Formula (Y-1):

5



The functional group can be, for example, a targeting group (*e.g.*, a steroid or a carbohydrate), a reporter group (*e.g.*, a fluorophore), or a label (an isotopically labelled moiety). The targeting group can further include protein binding agents, endothelial cell targeting groups (*e.g.*, RGD peptides and mimetics), cancer cell targeting groups (*e.g.*, folate Vitamin B12, Biotin), bone cell targeting groups (*e.g.*, bisphosphonates, polyglutamates, polyaspartates), multivalent mannose (for *e.g.*, macrophage testing), lactose, galactose, N-acetyl-galactosamine, monoclonal antibodies, glycoproteins, lectins, melanotropin, or thyrotropin.

As can be appreciated by the skilled artisan, methods of synthesizing the compounds of the formulae herein will be evident to those of ordinary skill in the art. The synthesized compounds can be separated from a reaction mixture and further purified by a method such as column chromatography, high pressure liquid chromatography, or recrystallization. Additionally, the various synthetic steps may be performed in an alternate sequence or order to give the desired compounds. Synthetic chemistry transformations and protecting group methodologies (protection and deprotection) useful in synthesizing the compounds described herein are known in the art and include, for example, those such as described in R. Larock, *Comprehensive Organic Transformations*, VCH Publishers (1989); T.W. Greene and P.G.M. Wuts, *Protective Groups in Organic Synthesis*, 2d. Ed., John Wiley and Sons (1991); L. Fieser and M. Fieser, *Fieser and Fieser's Reagents for Organic Synthesis*, John Wiley and Sons (1994);

and L. Paquette, ed., *Encyclopedia of Reagents for Organic Synthesis*, John Wiley and Sons (1995), and subsequent editions thereof.

Delivery Module

5 The monomers and methods described herein can be used to prepare an oligonucleotide agent, *e.g.*, conjugated oligonucleotide agent described herein, that can be used with a drug delivery conjugate or module.

10 The oligonucleotide agents can be complexed to a delivery agent that features a modular complex. The complex can include a carrier agent linked to one or more of (preferably two or more, more preferably all three of): (a) a condensing agent (*e.g.*, an agent capable of attracting, *e.g.*, binding, a nucleic acid, *e.g.*, through ionic or electrostatic interactions); (b) a fusogenic agent (*e.g.*, an agent capable of fusing and/or being transported through a cell membrane, *e.g.*, an endosome membrane); and (c) a targeting group, *e.g.*, a cell or tissue targeting agent, *e.g.*, a lectin, glycoprotein, lipid or protein, *e.g.*, an antibody, that binds to a specified cell type such as a
15 kidney cell.

 An oligonucleotide agent can be linked, *e.g.*, coupled or bound, to the modular complex. The oligonucleotide agent can interact with the condensing agent of the complex, and the complex can be used to deliver an oligonucleotide agent to a cell, *e.g.*, *in vitro* or *in vivo*. For example, the complex can be used to deliver an oligonucleotide agent to a subject in need
20 thereof, *e.g.*, to deliver an oligonucleotide agent to a subject having a disease or disorder.

 The fusogenic agent and the condensing agent can be different agents or the one and the same agent. For example, a polyamino chain, *e.g.*, polyethyleneimine (PEI), can be the fusogenic and/or the condensing agent.

 The delivery agent can be a modular complex. For example, the complex can include a
25 carrier agent linked to one or more of (preferably two or more, more preferably all three of):

 (a) a condensing agent (*e.g.*, an agent capable of attracting, *e.g.*, binding, a nucleic acid, *e.g.*, through ionic interaction),

 (b) a fusogenic agent (*e.g.*, an agent capable of fusing and/or being transported through a cell membrane, *e.g.*, an endosome membrane), and

30 (c) a targeting group, *e.g.*, a cell or tissue targeting agent, *e.g.*, a lectin, glycoprotein, lipid or protein, *e.g.*, an antibody, that binds to a specified cell type such as a kidney cell. A targeting

group can be a thyrotropin, melanotropin, lectin, glycoprotein, surfactant protein A, Mucin carbohydrate, multivalent lactose, multivalent galactose, N-acetyl-galactosamine, N-acetyl-gulucosamine multivalent mannose, multivalent fucose, glycosylated polyaminoacids, multivalent galactose, transferrin, bisphosphonate, polyglutamate, polyaspartate, a lipid, cholesterol, a steroid, bile acid, folate, vitamin B12, biotin, Neproxin, or an RGD peptide or RGD peptide mimetic.

Carrier agents

The carrier agent of a modular complex described herein can be a substrate for attachment of one or more of: a condensing agent, a fusogenic agent, and a targeting group. The carrier agent would preferably lack an endogenous enzymatic activity. The agent would preferably be a biological molecule, preferably a macromolecule. Polymeric biological carriers are preferred. It would also be preferred that the carrier molecule be biodegradable.

The carrier agent can be a naturally occurring substance, such as a protein (*e.g.*, human serum albumin (HSA), low-density lipoprotein (LDL), or globulin); carbohydrate (*e.g.*, a dextran, pullulan, chitin, chitosan, inulin, cyclodextrin or hyaluronic acid); or lipid. The carrier molecule can also be a recombinant or synthetic molecule, such as a synthetic polymer, *e.g.*, a synthetic polyamino acid. Examples of polyamino acids include polylysine (PLL), poly L-aspartic acid, poly L-glutamic acid, styrene-maleic acid anhydride copolymer, poly(L-lactide-co-glycolide) copolymer, divinyl ether-maleic anhydride copolymer, N-(2-hydroxypropyl)methacrylamide copolymer (HMPA), polyethylene glycol (PEG), polyvinyl alcohol (PVA), polyurethane, poly(2-ethylacrylic acid), N-isopropylacrylamide polymers, or polyphosphazine. Other useful carrier molecules can be identified by routine methods.

A carrier agent can be characterized by one or more of: (a) is at least 1 Da in size; (b) has at least 5 charged groups, preferably between 5 and 5000 charged groups; (c) is present in the complex at a ratio of at least 1:1 carrier agent to fusogenic agent; (d) is present in the complex at a ratio of at least 1:1 carrier agent to condensing agent; (e) is present in the complex at a ratio of at least 1:1 carrier agent to targeting agent.

Fusogenic agents

A fusogenic agent of a modular complex described herein can be an agent that is responsive to, *e.g.*, changes charge depending on, the pH environment. Upon encountering the pH of an endosome, it can cause a physical change, *e.g.*, a change in osmotic properties which
5 disrupts or increases the permeability of the endosome membrane. Preferably, the fusogenic agent changes charge, *e.g.*, becomes protonated, at pH lower than physiological range. For example, the fusogenic agent can become protonated at pH 4.5-6.5. The fusogenic agent can serve to release the oligonucleotide agent into the cytoplasm of a cell after the complex is taken up, *e.g.*, via endocytosis, by the cell, thereby increasing the cellular concentration of the
10 oligonucleotide agent in the cell.

In one embodiment, the fusogenic agent can have a moiety, *e.g.*, an amino group, which, when exposed to a specified pH range, will undergo a change, *e.g.*, in charge, *e.g.*, protonation. The change in charge of the fusogenic agent can trigger a change, *e.g.*, an osmotic change, in a vesicle, *e.g.*, an endocytic vesicle, *e.g.*, an endosome. For example, the fusogenic agent, upon
15 being exposed to the pH environment of an endosome, will cause a solubility or osmotic change substantial enough to increase the porosity of (preferably, to rupture) the endosomal membrane.

The fusogenic agent can be a polymer, preferably a polyamino chain, *e.g.*, polyethyleneimine (PEI). The PEI can be linear, branched, synthetic or natural. The PEI can be, *e.g.*, alkyl substituted PEI, or lipid substituted PEI.

20 In other embodiments, the fusogenic agent can be polyhistidine, polyimidazole, polypyridine, polypropyleneimine, mellitin, or a polyacetal substance, *e.g.*, a cationic polyacetal. In some embodiment, the fusogenic agent can have an alpha helical structure. The fusogenic agent can be a membrane disruptive agent, *e.g.*, mellittin.

A fusogenic agent can have one or more of the following characteristics: (a) is at least
25 1Da in size; (b) has at least 10 charged groups, preferably between 10 and 5000 charged groups, more preferably between 50 and 1000 charged groups; (c) is present in the complex at a ratio of at least 1:1 fusogenic agent to carrier agent; (d) is present in the complex at a ratio of at least 1:1 fusogenic agent to condensing agent; (e) is present in the complex at a ratio of at least 1:1 fusogenic agent to targeting agent.

30 Other suitable fusogenic agents can be tested and identified by a skilled artisan. The ability of a compound to respond to, *e.g.*, change charge depending on, the pH environment can

be tested by routine methods, *e.g.*, in a cellular assay. For example, a test compound is combined or contacted with a cell, and the cell is allowed to take up the test compound, *e.g.*, by endocytosis. An endosome preparation can then be made from the contacted cells and the endosome preparation compared to an endosome preparation from control cells. A change, *e.g.*, a decrease, in the endosome fraction from the contacted cell vs. the control cell indicates that the test compound can function as a fusogenic agent. Alternatively, the contacted cell and control cell can be evaluated, *e.g.*, by microscopy, *e.g.*, by light or electron microscopy, to determine a difference in endosome population in the cells. The test compound can be labeled. In another type of assay, a modular complex described herein is constructed using one or more test or putative fusogenic agents. The modular complex can be constructed using a labeled nucleic acid instead of the oligonucleotide. The ability of the fusogenic agent to respond to, *e.g.*, change charge depending on, the pH environment, once the modular complex is taken up by the cell, can be evaluated, *e.g.*, by preparation of an endosome preparation, or by microscopy techniques, as described above. A two-step assay can also be performed, wherein a first assay evaluates the ability of a test compound alone to respond to, *e.g.*, change charge depending on, the pH environment; and a second assay evaluates the ability of a modular complex that includes the test compound to respond to, *e.g.*, change charge depending on, the pH environment.

Condensing agent

The condensing agent of a modular complex described herein can interact with (*e.g.*, attracts, holds, or binds to) an oligonucleotide agent and act to (a) condense, *e.g.*, reduce the size or charge of the oligonucleotide agent and/or (b) protect the oligonucleotide agent, *e.g.*, protect the oligonucleotide agent against degradation. The condensing agent can include a moiety, *e.g.*, a charged moiety, that can interact with a nucleic acid, *e.g.*, an oligonucleotide agent, *e.g.*, by ionic interactions. The condensing agent would preferably be a charged polymer, *e.g.*, a polycationic chain. The condensing agent can be a polylysine (PLL), spermine, spermidine, polyamine, pseudopeptide-polyamine, peptidomimetic polyamine, dendrimer polyamine, arginine, amidine, protamine, cationic lipid, cationic porphyrin, quarternary salt of a polyamine, or an alpha helical peptide.

A condensing agent can have the following characteristics: (a) at least 1Da in size; (b) has at least 2 charged groups, preferably between 2 and 100 charged groups; (c) is present in the

complex at a ratio of at least 1:1 condensing agent to carrier agent; (d) is present in the complex at a ratio of at least 1:1 condensing agent to fusogenic agent; (e) is present in the complex at a ratio of at least 1:1 condensing agent to targeting agent.

Other suitable condensing agents can be tested and identified by a skilled artisan, *e.g.*, by evaluating the ability of a test agent to interact with a nucleic acid, *e.g.*, an oligonucleotide agent. The ability of a test agent to interact with a nucleic acid, *e.g.*, an oligonucleotide agent, *e.g.*, to condense or protect the oligonucleotide agent, can be evaluated by routine techniques. In one assay, a test agent is contacted with a nucleic acid, and the size and/or charge of the contacted nucleic acid is evaluated by a technique suitable to detect changes in molecular mass and/or charge. Such techniques include non-denaturing gel electrophoresis, immunological methods, *e.g.*, immunoprecipitation, gel filtration, ionic interaction chromatography, and the like. A test agent is identified as a condensing agent if it changes the mass and/or charge (preferably both) of the contacted nucleic acid, compared to a control. A two-step assay can also be performed, wherein a first assay evaluates the ability of a test compound alone to interact with, *e.g.*, bind to, *e.g.*, condense the charge and/or mass of, a nucleic acid; and a second assay evaluates the ability of a modular complex that includes the test compound to interact with, *e.g.*, bind to, *e.g.*, condense the charge and/or mass of, a nucleic acid.

Amphipathic Delivery Agents

The oligonucleotide agents described herein can be used with an amphipathic delivery conjugate or module, such as those described herein and those described in copending, co-owned United States Provisional Application Serial No. 60/455,050, filed on March 13, 2003, and International Application Serial No. PCT/US04/07070, filed March 8, 2004, which is hereby incorporated by reference.

OLIGONUCLEOTIDE PRODUCTION

An oligonucleotide agent can be produced, *e.g.*, in bulk, by a variety of methods. Exemplary methods include: organic synthesis and RNA cleavage, *e.g.*, *in vitro* cleavage.

Organic Synthesis

A large bioreactor, *e.g.*, the OligoPilot II from Pharmacia Biotec AB (Uppsala Sweden), can be used to produce a large amount of an oligonucleotide agent. The OligoPilotII reactor can efficiently couple a nucleotide using only a 1.5 molar excess of a phosphoramidite nucleotide. To make an RNA strand, ribonucleotides amidites are used. Standard cycles of monomer
5 addition can be used to synthesize the oligonucleotide agent.

Organic synthesis can be used to produce a discrete oligonucleotide agent species. The complementary of the species to a particular target gene can be precisely specified. For example, the species may be complementary to a region that includes a polymorphism, *e.g.*, a single nucleotide polymorphism. Further the location of the polymorphism can be precisely defined.
10 In some embodiments, the polymorphism is located in an internal region, *e.g.*, at least 4, 5, 7, or 9 nucleotides from one or both of the termini.

An oligonucleotide agent preparation can be prepared in a solution (*e.g.*, an aqueous and/or organic solution) that is appropriate for formulation. For example, the oligonucleotide agent preparation can be precipitated and redissolved in pure double-distilled
15 water, and lyophilized. The dried oligonucleotide agent can then be resuspended in a solution appropriate for the intended formulation process.

Synthesis of modified and nucleotide surrogate oligonucleotide agents is discussed below.

FORMULATION

20 The oligonucleotide agents described herein can be formulated for administration to a subject.

For ease of exposition the formulations, compositions and methods in this section are discussed largely with regard to unmodified oligonucleotide agents. It should be understood, however, that these formulations, compositions and methods can be practiced with other
25 oligonucleotide agents, *e.g.*, modified oligonucleotide agents, and such practice is within the invention.

A formulated oligonucleotide agent composition can assume a variety of states. In some examples, the composition is at least partially crystalline, uniformly crystalline, and/or anhydrous (*e.g.*, less than 80, 50, 30, 20, or 10% water). In another example, the oligonucleotide
30 agent is in an aqueous phase, *e.g.*, in a solution that includes water.

The aqueous phase or the crystalline compositions can, *e.g.*, be incorporated into a delivery vehicle, *e.g.*, a liposome (particularly for the aqueous phase) or a particle (*e.g.*, a microparticle as can be appropriate for a crystalline composition). Generally, the oligonucleotide agent composition is formulated in a manner that is compatible with the intended method of administration (see, below).

In particular embodiments, the composition is prepared by at least one of the following methods: spray drying, lyophilization, vacuum drying, evaporation, fluid bed drying, or a combination of these techniques; or sonication with a lipid, freeze-drying, condensation and other self-assembly.

An oligonucleotide agent preparation can be formulated in combination with another agent, *e.g.*, another therapeutic agent or an agent that stabilizes an oligonucleotide agent, *e.g.*, a protein that complexes with an oligonucleotide agent. Still other agents include chelators, *e.g.*, EDTA (*e.g.*, to remove divalent cations such as Mg^{2+}), salts, RNase inhibitors (*e.g.*, a broad specificity RNase inhibitor such as RNAsin) and so forth.

In one embodiment, the oligonucleotide agent preparation includes a second oligonucleotide agent, *e.g.*, a second oligonucleotide agent that can modulate gene expression with respect to a second gene, or with respect to the same gene. Still other preparation can include at least three, five, ten, twenty, fifty, or a hundred or more different oligonucleotide agent species. Such oligonucleotide agents can modulate gene expression with respect to a similar number of different genes.

In one embodiment, the oligonucleotide agent preparation includes at least a second therapeutic agent (*e.g.*, an agent other than an RNA or a DNA). For example, an oligonucleotide agent composition for the treatment of a viral disease, *e.g.* HIV, might include a known antiviral agent (*e.g.*, a protease inhibitor or reverse transcriptase inhibitor). In another example, an oligonucleotide agent composition for the treatment of a cancer might further comprise a chemotherapeutic agent.

Oligonucleotide agents described herein can be formulated for delivery in a membranous molecular assembly, *e.g.*, a liposome or a micelle. In one embodiment, a preparation including an oligonucleotide agent can be formulated as an emulsion that includes a surfactant.

Targeting

For ease of exposition the formulations, compositions and methods in this section are discussed largely with regard to unmodified oligonucleotide agents. It should be understood, however, that these formulations, compositions and methods can be practiced with other
5 oligonucleotide agents, *e.g.*, modified oligonucleotide agents, and such practice is within the invention.

In some embodiments, an oligonucleotide agent, or a DNA which encodes an oligonucleotide agent, or precursor thereof, is targeted to a particular cell. For example, a liposome or particle or other structure that includes an oligonucleotide agent can also include a
10 targeting moiety that recognizes a specific molecule on a target cell. The targeting moiety can be a molecule with a specific affinity for a target cell. Targeting moieties can include antibodies directed against a protein found on the surface of a target cell, or the ligand or a receptor-binding portion of a ligand for a molecule found on the surface of a target cell. For example, the targeting moiety can recognize a cancer-specific antigen of the kidney (*e.g.*, G250, CA15-3,
15 CA19-9, CEA, or HER2/neu) or a viral antigen, thus delivering the oligonucleotide agent to a cancer cell or a virus-infected cell. Exemplary targeting moieties include antibodies (such as IgM, IgG, IgA, IgD, and the like, or a functional portions thereof), ligands for cell surface receptors (*e.g.*, ectodomains thereof).

Table 6 provides a number of antigens which can be used to target an oligonucleotide
20 agent to a selected cell, such as when targeting of the oligonucleotide agent to a tissue other than the kidney is desired.

Table 6. Targeting Antigens

ANTIGEN	Exemplary tumor tissue
CEA (carcinoembryonic antigen)	colon, breast, lung
PSA (prostate specific antigen)	prostate cancer
CA-125	ovarian cancer
CA 15-3	breast cancer
CA 19-9	breast cancer
HER2/neu	breast cancer
α -feto protein	testicular cancer, hepatic cancer
β -HCG (human chorionic gonadotropin)	testicular cancer, choriocarcinoma
MUC-1	breast cancer
Estrogen receptor	breast cancer, uterine cancer
Progesterone receptor	breast cancer, uterine cancer
EGFr (epidermal growth factor receptor)	bladder cancer

In one embodiment, the targeting moiety is attached to a liposome. For example, US
5 Patent No. 6,245,427 describes a method for targeting a liposome using a protein or peptide. In
another example, a cationic lipid component of the liposome is derivatized with a targeting
moiety. For example, WO 96/37194 describes converting N-glutaryldioleoylphosphatidyl
ethanolamine to a N-hydroxysuccinimide activated ester. The product was then coupled to an
RGD peptide.

10

TARGETING TO THE KIDNEY

The kidney is an important site of gene expression. Aspects of the invention relate to
silencing genes expressed in the kidney, *e.g.*, to treat disorders of or related to the kidney.
Accordingly, the invention includes compositions and methods for delivering oligonucleotide
15 agents to the kidney. The invention also includes compositions and methods for minimizing
delivery of oligonucleotide agents to the kidney.

An oligonucleotide agent composition of the invention can be one that has been modified
to alter distribution in favor of the kidney. A composition of the invention includes an
oligonucleotide agent, *e.g.*, an oligonucleotide agent described herein.

20 One aspect of the invention provides a method for treating a human having or at risk for
having a disorder of the kidney. The method of treatment includes administering an

oligonucleotide agent to the human, wherein the oligonucleotide agent targets a nucleic acid, *e.g.*, an RNA expressed in the kidney. In one embodiment, the human is suffering from a disorder characterized by elevated or otherwise unwanted expression of a nucleic acid, *e.g.*, elevated gene expression levels or elevated RNA levels, in the kidney. The unwanted expression
5 levels can correspond to a gene encoding a chemokine, such as RANTES, MCP1 or osteopontin; or a gene encoding a complement factor or a growth factor (*e.g.*, Transforming growth factor-beta (TGFbeta), Platelet derived growth factor (PDGF), IGF-1, IGF-2 or Vascular endothelial growth factor (VEGF)). In another embodiment, the gene can encode an inflammatory cytokine, such as IL1alpha or TNFalpha; a fibrogenic cytokine; a vasoactive protein, such as angiotensin II
10 or ET1; or a growth factor receptor, such as KDR (VEGF receptor), an epidermal growth factor receptor, or a fibroblast growth factor receptor.

In one embodiment, the oligonucleotide agent targets an miRNA expressed in the kidney. In another embodiment, the human is suffering from a disorder characterized by overexpression or accumulation of the miRNA in the kidney, or decreased expression of a nucleic acid that is the
15 target of the miRNA expressed in the kidney. Administration of the oligonucleotide agent to the subject, or to a cell of the kidney of the subject, can result in the pairing of the oligonucleotide agent with the target miRNA and the subsequent downregulation of the miRNA.

In one embodiment, the oligonucleotide agent is substantially identical to an miRNA normally expressed in kidney tissue, and in another embodiment, the human is suffering from a
20 disorder characterized by decreased expression of the miRNA in the kidney. Administration of the oligonucleotide agent to the subject, or to a cell of the kidney, at least partially rescues the function of the downregulated miRNA.

In one embodiment, the human has or is at risk for having renal vascular hypertension, a ureter obstruction, diabetes, diabetic nephropathy, glomerular sclerosis, glomerular nephritis,
25 systemic lupus erythematosus, HIV-associated nephropathy, renal fibrosis, proteinuria, renal carcinoma, Fanconi's syndrome or Bartter's syndrome. In another embodiment, an oligonucleotide agent targeting the kidney can be administered to a subject in shock, or the agent can be administered before, during, and/or following a kidney transplant.

In one embodiment, the oligonucleotide agent targets a growth factor, such as TGFbeta, or a growth factor receptor, and the human has or is at risk for having diabetic nephropathy, progressive renal disease, chronic tissue injury, or glomerulosclerosis. In one embodiment, the oligonucleotide agent targets a growth factor, such as TGFbeta, and the human has had or is
5 going to have a kidney transplant, or has been identified as a candidate for a kidney transplant.

In one embodiment, the oligonucleotide agent targets PDGF and the human has had or is going to have a kidney transplant, or has been identified as a candidate for a kidney transplant.

In one embodiment, the oligonucleotide agent targets a vasoconstrictor, such as angiotensin II, or a vasoconstrictor receptor, such as angiotensin receptor I, and the human has or
10 is at risk for having angiotensin II-dependent hypertension or type II diabetes, or the human is in a hyperglycemic state.

In one embodiment, the oligonucleotide agent targets a vasoconstrictor, such as endothelin-1 (ET-1), or an ET-1 receptor, such as ETA or ETB, and the human has or is at risk for having an autosomal-dominant polycystic kidney disease and/or chronic renal disease. For
15 example, the human can have an autosomal-dominant polycystic kidney disease, and in one embodiment, the patient's condition has progressed to a chronic renal disease.

In one embodiment, the oligonucleotide agent targets a transcription factor, such as a ligand-activated transcription factor, *e.g.*, the nuclear hormone receptor peroxisome proliferator-activated receptor (PPAR), and the human has or is at risk for having diabetic nephropathy, a
20 kidney tumor, or glomerulosclerosis. In one embodiment the oligonucleotide agent targets PPAR-alpha, PPAR beta/delta, or PPAR gamma.

In one embodiment, the oligonucleotide agent targets a growth factor receptor, such as an IGF receptor (*e.g.*, IGFR1), the VEGF receptor KDR, an epidermal growth factor receptor, or a fibroblast growth factor receptor, and the human has or is at risk for having a renal cell
25 carcinoma, diabetic nephropathy, renal hypertrophy, glomerular enlargement, increased urinary albumin excretion, and/or diabetes.

In one embodiment, the oligonucleotide agent targets a costimulatory molecule, *e.g.*, B7-1, B7-2, ICOS, CD40, and/or CD154, and the human has or is at risk for having an autoimmune disease or transplant rejection.

In one embodiment, the oligonucleotide agent targets a chemokine, such as MCP-1, RANTES and/or osteopontin, and the human has or is at risk for having systemic hypertension, renal parenchymal injury, an acute or chronic rejection of a kidney allograft, or chronic hypoxia-induced hypertension.

One aspect of the invention provides an oligonucleotide agent that targets a complement component, such as complement factor C3, C4, C5 or B. An oligonucleotide that targets a complement component can be desirable, *e.g.*, to inhibit the immune response.

In one aspect, the invention provides for a method of delivering an oligonucleotide agent to the kidney of a subject, *e.g.*, a mammalian subject, such as a mouse or a human. In one embodiment, the oligonucleotide agent can be delivered to a cell or cells in the glomerulus of the kidney, *e.g.*, glomerular endothelial cells, glomerular epithelial cells, mesangial cells, and the like; and/or the oligonucleotide agent can be delivered to the proximal tubular cells of the kidney. For example, an oligonucleotide agent can be delivered to the proximal tubular cells of the kidney for treatment of shock, ureter obstruction, diabetes, proteinuria, renal carcinoma, or a tubular defect disease, such as Fanconi or Bartter's syndrome. An oligonucleotide agent directed to the treatment of a renal transplant patient can also be directed to the proximal tubular cells of the kidney. In one embodiment, an oligonucleotide directed to the proximal tubular cells of the kidney will further be delivered to the interstitium and other downstream cells. It is preferable that the oligonucleotide agent silences a target gene at the target site within the kidney.

An oligonucleotide agent delivered to the kidney, *e.g.*, the proximal tubular cells of the kidney, can be an unmodified oligonucleotide agent. In one embodiment, the oligonucleotide agent can be stabilized with phosphodiester linkages. In another embodiment, the 3' end of the oligonucleotide agent can be modified by a cationic group, *e.g.*, an alkyl amine (such as an 2'-O-alkyl amine), polyamine, cationic peptide, or cationic amino acid. The modification can be an external or terminal cationic residue. In another embodiment, the oligonucleotide agent can be modified with a sugar, *e.g.*, a glycoconjugate or alkylglycoside component, *e.g.*, glucose,

mannose, 2-deoxy-glucose, or an analog thereof. In another embodiment, the oligonucleotide agent can be conjugated to an enzyme substrate, *e.g.*, a substrate for which the relative enzyme is present in a higher amount, as compared to the enzyme level in other tissues of the body. For example, the oligonucleotide agent can be conjugated to a substrate of γ -glutamyl transferase or
5 n-acetyl- γ -glutamyl transferase.

In one embodiment, the oligonucleotide agent can be conjugated to a folic acid or folic acid derivative, *e.g.*, γ -folate, α -folate, 5-methyl tetrahydrofolic acid, a pteridine analog, or an alternative analog thereof.

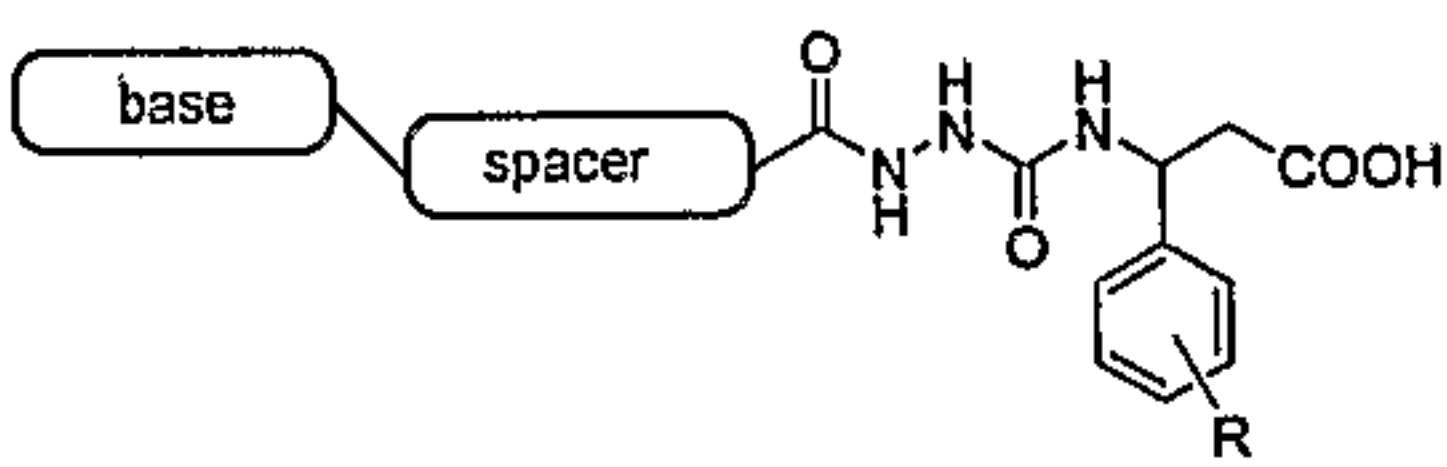
In one embodiment, the oligonucleotide agent of the invention can be conjugated to a
10 protein that will accumulate in the kidney when administered systematically. For example, the oligonucleotide agent can be conjugated to a lysozyme, cytochrome-c or aprotinin protein. In one embodiment, the oligonucleotide agent can be conjugated to a lysine residue of the protein.

In one embodiment, an oligonucleotide agent targeted to the kidney can be conjugated to a low molecular weight polyethylene glycol (PEG) molecule, or guanidium group, and in another
15 embodiment, the oligonucleotide agent can be conjugated to an RGD peptide, peptide analog, or peptide mimetic or derivative thereof. An oligonucleotide conjugated to an RGD peptide, peptide analog, or peptide mimetic can bind to an $\alpha_v\beta_3$ integrin.

Synthetic Integrin Inhibitors

Ref: Goodman, S. L.; Hölzemann, G.; Sulyok, G. A. G.; Kessler, H. *J. Med. Chem.* **2002**, *45*, 1045-1051.

Table 8

	R	base	spacer
	(Carboxamide) ^a	guanidine	<i>m</i> -C ₆ H ₄ -
	none	guanidine	ω-C ₄ H ₈ -
	none	guanidine	<i>m</i> -C ₆ H ₄ -
	4-F	guanidine	<i>m</i> -C ₆ H ₄ -
	4-Cl	guanidine	<i>m</i> -C ₆ H ₄ -
	4-Br	guanidine	<i>m</i> -C ₆ H ₄ -
	4-OCH ₃	guanidine	<i>m</i> -C ₆ H ₄ -
	4-OCF ₃	guanidine	guanidine
	(1-naphthyl) ^a	guanidine	<i>m</i> -C ₆ H ₄ -
	3-Cl,5-Cl	guanidine	<i>m</i> -C ₆ H ₄ -
	(H) ^a	2-NH ₂ -pyridine	ω-C ₄ H ₈ -
	none	2-NH ₂ -pyridine	ω-C ₄ H ₈ -
	4-F	2-NH ₂ -pyridine	ω-C ₄ H ₈ -
	4-Cl	2-NH ₂ -pyridine	ω-C ₄ H ₈ -
	4-Br	2-NH ₂ -pyridine	ω-C ₄ H ₈ -
	4-OCH ₃	2-NH ₂ -pyridine	ω-C ₄ H ₈ -
	4-OCF ₃	2-NH ₂ -pyridine	ω-C ₄ H ₈ -
	(1-naphthyl) ^a	2-NH ₂ -pyridine	ω-C ₄ H ₈ -

	3-Cl,5-Cl	2-NH ₂ -pyridine	ω -C ₄ H ₈ -
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^a Instead of substituted phenyl ring

Ref: Sulyok, G. A. G.; Gibson, C.; Goodman, S. L.; Hölzemann, G.; Wiesner, M.; Kessler H. *J. Med. Chem.* **2001**, *44*, 1938-1950

In a preferred embodiment, at least 30%, 40%, 50%, 60%, 70%, 80%, 90% or more of the oligonucleotide agent administered to the subject is successfully targeted to the kidney. In a preferred embodiment between 30-90%, 40-80% or 50-70% 50-80%, or 50-90% of the oligonucleotide agent administered to the subject is successfully targeted to the kidney.

In any of the embodiments described above, the oligonucleotide agent/conjugate can have additional modifications, such as a stabilizing modification. For example, a linker molecule can tether a protein, PEG or RGD peptide to the oligonucleotide agent. Exemplary linkers are described *infra*, and can include amino linkers (*e.g.*, aminooxy linkers), thiol linkers, carboxyl linkers, aldehyde linkers, haloacetyl linkers, and the like.

In another aspect, the invention features an conjugate oligonucleotide agent. The conjugate includes an oligonucleotide agent coupled to, *e.g.*, linked to, a ligand or therapeutic agent. The oligonucleotide agent is optionally coupled to the ligand or therapeutic agent by a linker (*e.g.*, a peptide linker or other linker described herein). The ligand can function to, *e.g.*, affect the distribution of the oligonucleotide agent in the body and/or to target the oligonucleotide agent to a particular tissue or cell.

The ligand can be placed at an end of the oligonucleotide agent, preferably at the 3' end of an oligonucleotide agent. The ligand can also be placed at the 5' end, or within the middle of the oligonucleotide agent. In some embodiments, more than one ligand can be coupled to the oligonucleotide agent. For example, a ligand can be coupled to the 3' end of an oligonucleotide agent; a ligand can be coupled to an end, *e.g.*, a 3' end, and to the middle of an oligonucleotide agent; a ligand can be coupled to the 3' end and the 5' of an oligonucleotide agent; a ligand can be coupled to the 3' end, the 5' end, and to one or more internal positions of an oligonucleotide agent.

In one embodiment, the ligand of a conjugated oligonucleotide agent is a lipid or lipid-based molecule. Such a lipid or lipid-based molecule preferably binds a serum protein, *e.g.*, human serum albumin (HSA). An HSA binding ligand allows for distribution of the conjugate to a target tissue, *e.g.*, a non-kidney target tissue of the body. For example, the target tissue can be the liver, including, but not limited to parenchymal cells of the liver. Other molecules that can bind HSA can also be used as ligands. For example, neproxin or aspirin can be used. A lipid or lipid-based ligand can (a) increase resistance to degradation of the conjugate, (b) increase targeting or transport into a target cell or cell membrane, and/or (c) can be used to adjust binding to a serum protein, *e.g.*, HSA.

A lipid based ligand can be used to modulate, *e.g.*, control the binding of the conjugate to a target tissue. For example, a lipid or lipid-based ligand that binds to HSA more strongly will be less likely to be targeted to the kidney and therefore less likely to be cleared from the body. A lipid or lipid-based ligand that binds to HSA less strongly can be used to target the conjugate to the kidney.

In a preferred embodiment, the lipid based ligand binds HSA. Preferably, it binds HSA with a sufficient affinity such that the conjugate will be preferably distributed to a non-kidney tissue. However, it is preferred that the affinity not be so strong that the HSA-ligand binding cannot be reversed.

In another preferred embodiment, the lipid based ligand binds HSA weakly or not at all, such that the conjugate will be preferably distributed to the kidney. Other moieties that target to kidney cells can also be used in place of or in addition to the lipid based ligand.

In a preferred embodiment, the lipid or lipid based ligand is a phosphorothioate. In this embodiment, it is preferred that the number of sulfurs on the phosphorothioate not be so prevalent that they interfere with binding to a serum protein, *e.g.*, HSA.

In another embodiment, the ligand is a peptide or peptoid. Peptoids, in particular amphipathic species, such as Antennapedia or tat, are preferred.

In another embodiment, the ligand is a polyethylene glycol (PEG) or derivatives thereof. A PEG can, *e.g.*, allow the agent to be kept in circulation. A PEG is intrinsically amphipathic, and can promote stability, particularly if coupled at the 3' end of the oligonucleotide agent.

5 In another embodiment, the ligand is a charged group or moiety, *e.g.*, a polyamine or cationic group or moiety. This type of linker moiety, *e.g.*, because of its charge, *e.g.*, its negative charge, can help overcome the resistance of entry of the oligonucleotide agent into a cell. Preferably, these are conjugated at the 3' end, but they can also be at the 5' end or within the middle of the oligonucleotide molecule. Exemplary polyamines include polyarginine, polylysine, polyhistidine, polypreproline, or polymorpholinos, polyornithine.

10 In another embodiment, the ligand is a vitamin or other moiety that is taken up by a target cell, *e.g.*, a proliferating cell. These are particularly useful for treating disorders characterized by unwanted cell proliferation, *e.g.*, of the malignant or non-malignant type, *e.g.*, cancer cells. Exemplary vitamins are B vitamin, *e.g.*, folic acid, B12, riboflavin, biotin, pyridoxal or other vitamins or nutrients taken up by cancer cells. Also included are HSA and low density
15 lipoprotein (LDL).

In another embodiment, the ligand is a cell-permeation agent, preferably a helical cell-permeation agent. Preferably, the agent is amphipathic. An exemplary agent is a peptide such as tat or Antennapodia. If the agent is a peptide, it can be modified, including a peptidylmimetic, invertomers, non-peptide or pseudo-peptide linkages, and use of D-amino acids. The helical
20 agent is preferably an alpha-helical agent, which preferably has a lipophilic and a lipophobic phase.

The ligand can be a targeting agent. The targeting agent can be a sugar, a peptide, *e.g.*, an RGD containing peptide.

Another useful targeting agent is one that incorporates a sugar, *e.g.*, galactose and/or
25 analogs thereof. These are useful because they target the liver, in particular, the parenchymal cells of the liver. In a preferred embodiment, the targeting agent includes more than one galactose moiety, preferably two or three. Preferably, the targeting agent includes 3 galactose moieties, *e.g.*, spaced about 15 angstroms from each other. The targeting agent can be lactose.

Lactose is a glucose coupled to a galactose. Preferably, the targeting agent includes three lactoses. The targeting agent can also be N-Acetyl-Galactosamine, N-Ac-Glucosamine. A mannose, or mannose-6-phosphate targeting agent can be used for macrophage targeting.

Peptides that target markers enriched in proliferating cells can be used. *E.g.*, RGD
5 containing peptides and peptidomimetics can target cancer cells, in particular cells that exhibit an $\alpha_v\beta_3$ integrin. Thus, one could use RGD peptides, cyclic peptides containing RGD, RGD peptides that include D-amino acids, as well as synthetic RGD mimics. In addition to RGD, one can use other moieties that target the $\alpha_v\beta_3$ integrin ligand. Generally, such ligands can be used to control proliferating cells and angiogenesis. Preferred conjugates of this type include an
10 oligonucleotide agent that targets PECAM-1, VEGF, or other cancer gene, *e.g.*, a cancer gene described herein.

In one embodiment, an oligonucleotide agent is linked, *e.g.*, directly linked, *e.g.*, covalently, or non-covalently linked, to the targeting agent, *e.g.*, a targeting agent described herein. This is referred to as a “conjugation” approach. In another embodiment, the targeting
15 agent (*e.g.*, the same targeting agent) is simply mixed with the oligonucleotide agent. This is referred to as a “complexing” approach. In a complexing approach, the oligonucleotide agent can be mixed with, *e.g.*, a cationic molecule, *e.g.*, a cationic lipid, *e.g.*, with or without a targeting group, *e.g.*, with or without a sugar or an RGD construct described herein. In some embodiments, the oligonucleotide agent is mixed with a polymer-based system, *e.g.*, with or
20 without a targeting group. In other embodiments, the oligonucleotide agent is mixed with a nanoparticle.

The conjugate oligonucleotide agents described herein can include a targeting agent that targets the oligonucleotide agent to a desired target cell or tissue. The target cell or tissue can be a cancer cell, a cell of the vasculature, *e.g.*, tumor vasculature, an angiogenic cell, *e.g.*, a tumor
25 angiogenic cell, or an endosome. A preferred target is the kidney. In another embodiment, the liver *e.g.*, the parenchymal cells of the liver, is a preferred target.

The methods and compositions of the invention, *e.g.*, the conjugates described herein, can be used with any of the oligonucleotide agents described herein. In addition, the methods and

compositions of the invention can be used for the treatment of any disease or disorder described herein, and for the treatment of any subject, *e.g.*, any animal, any mammal, such as any human.

The methods and compositions of the invention, *e.g.*, the conjugates described herein, can be used with any dosage and/or formulation described herein, as well as with any route of
5 administration described herein.

As used herein, “conjugated” means two entities are associated, *e.g.*, with sufficient affinity that the therapeutic benefit of the association between the two entities is realized. Conjugated can include covalent or noncovalent bonding as well as other forms of association, such as entrapment, *e.g.*, of one entity on or within the other, or of either or both entities on or
10 within a third entity, such as a micelle. Particularly preferred forms of conjugation are by covalent bonding, *e.g.*, those described herein. An entity can be conjugated to an oligonucleotide agent, *e.g.*, at the 3’ or 5’ terminus or internally. It is preferred that an entity is conjugated to the oligonucleotide agent in such a way as to preserve the ability of the oligonucleotide agent to mediate inhibition of gene expression.

15 TREATMENT OF OCULAR DISEASE

The oligonucleotide agents featured in the invention can be used to treat an ocular disease, such as age-related macular degeneration (AMD), glaucoma, cataract, optic nerve atrophy, diabetic retinopathy (DR), or retinitis pigmentosa. Table 9 summarizes gene targets for oligonucleotide agents described herein for the treatment of an ocular disease.

20 **Table 9.** Potential gene targets for treatment of ocular disease.

Disease	Gene	OMIM number	Reference
AMD, DME	VEGF	192240	Husain <i>et al.</i> , 2002
AMD, DME, glaucoma	PKC-beta	176970	Clark & Yario, 2003
AMD, DR	ACE	106180	Sjolie & Chaturvedi., 2002

AMD, DR	angiotensin II receptor	106165	Sjolie & Chaturvedi., 2002
AMD, DR	MMPs-2, -9, -13 and -14	120360, 120361, 600108, 600754	Scatena, 2000
AMD, DR	Integrins alpha v beta 3 and alpha v beta 5	193210, 173470, 147561	Friedlander <i>et al.</i> , 1996
AMD, DR	GH1	139250	Smith <i>et al.</i> , 1997
AMD, DR	IGF-I	147440	Smith <i>et al.</i> , 1997
AMD, DR, glaucoma	C-raf kinase (Raf-1)	164760	Danis <i>et al.</i> 2003, Hecquet <i>et al.</i> 2002
Glaucoma, DR	Ras		Hecquet <i>et al.</i> 2002
Glaucoma, DR	cyclin D1	168461	Hecquet <i>et al.</i> 2002
Glaucoma	Carbonic anhydrases XII (CA12), II (CA2), IV (CA4)	603263, 259730, 114760	Liao <i>et al.</i> , 2003
Glaucoma	myosin light chain kinase	600922	Clark & Yorio, 2003
Glaucoma, DR	nitric oxide synthase	163731	Wilson, 1999; Clark & Yorio, 2003
Glaucoma, DR	TNF- α	191160	Wilson, 1999; Clark & Yorio, 2003
Glaucoma, DR	TNF- α receptor	191190	Wilson, 1999; Clark & Yorio, 2003
Glaucoma, DR	Bcl-2, Bcl-XL		Wilson, 1999; Clark & Yorio, 2003

In one embodiment, an oligonucleotide agent targets VEGF or protein kinase C for the treatment of, *e.g.*, AMD and diabetic macular edema (DME).

- 5 In another embodiment, an oligonucleotide agent is used as a therapy for the treatment of AMD or DR, by targeting, *e.g.*, angiotensin converting enzyme (ACE), angiotensin II receptor, growth factors (*e.g.*, c-Abl, c-Kit, platelet-derived growth factor (PDGF), epidermal growth factor (EGF), insulin-like growth factor (IGF), growth hormone 1 (GH1), insulin-like growth factor-I (IGF-I), or c-raf kinase (raf-1), or their receptors), an inhibitor of matrix
- 10 metalloproteinases (MMPs) -2, -9, -13 or -14, integrin alpha v beta 3, or alpha v beta 5.

In one embodiment, an oligonucleotide agent targets a caspase, Bcl family member, nitric oxide synthase, endothelin, TNF- α or TNF- α receptor, ERK2, MEK1/2, or cyclin D1. These oligonucleotide agents can be useful for the treatment of, *e.g.*, glaucoma, retinitis pigmentosa, cataract formation, retinoblastoma, retinal ischemia, DR, or an ocular diseases involving the posterior segment, such as a viral infection or vitreoretinopathy.

In one embodiment, an oligonucleotide agent targets a carbonic anhydrase (CA2, CA4, and CA12) or protein kinase, *e.g.*, protein kinase C or myosin light chain kinase. These oligonucleotide agents can be useful for the treatment of, *e.g.*, glaucoma.

In one embodiment, an oligonucleotide agent targets an adhesion molecule, cytokine, chemokine, MMPs, or tissue inhibitor of metalloproteinase (TIMPs). These oligonucleotide agents can be useful for the treatment of, *e.g.*, immunologic challenge, wounding, infection, genetic disease, diabetes, or vitamin A deficiency.

In one embodiment, an oligonucleotide agent targets NOS-II. These oligonucleotide agents can be useful for the treatment of, *e.g.*, uveitis.

In one embodiment, an oligonucleotide agent targets cyclin D1, *e.g.*, to decrease cell proliferation in cells of the eye, *e.g.*, retinal pigment epithelium cells.

In one embodiment, the oligonucleotide agent targets an miRNA expressed in the eye. In another embodiment, the human is suffering from a disorder characterized by increased expression or accumulation of the miRNA in the eye, or decreased expression of a nucleic acid that is the target of the miRNA expressed in the eye. Administration of the oligonucleotide agent to the subject, *e.g.*, to the eye of the subject, results in the pairing of the oligonucleotide agent with the target miRNA and the subsequent downregulation of the miRNA.

In one embodiment, the oligonucleotide agent is substantially identical to an miRNA normally expressed in the eye, and in another embodiment, the human is suffering from a disorder characterized by decreased expression of the miRNA in the eye. Administration of the oligonucleotide agent to the subject, *e.g.*, to the eye of the subject, at least partially rescues the function of the downregulated miRNA.

An oligonucleotide agent useful for treating an ocular disease can be delivered to the eye by, *e.g.*, iontophoresis (*e.g.*, transcorneoscleral iontophoresis), topically (*e.g.*, by a patch or disk,

or by eye drops), or by intravitreal injection. The oligonucleotide agent can be formulated in sterically stabilized liposomes.

TARGETING TO THE LIVER

5 Aspects of the invention relate to silencing genes expressed in the liver, or to upregulating genes that are regulated by one or more endogenous miRNAs. Accordingly, the invention includes compositions and methods for delivering oligonucleotide agents to the liver, *e.g.*, to treat disorders of or related to the liver.

10 An oligonucleotide agent composition of the invention can be one that has been modified to alter distribution in favor of the liver. A composition of the invention includes an oligonucleotide agent, *e.g.*, an oligonucleotide agent described herein.

An oligonucleotide agent directed to the liver can target apoB-100 to treat a disorder characterized by elevated or otherwise unwanted expression of apoB-100, elevated or otherwise unwanted levels of cholesterol, and/or dysregulation of lipid metabolism. The oligonucleotide agent can be administered to an individual at risk for the disorder to delay onset of the disorder or a symptom of the disorder. These disorders include HDL/LDL cholesterol imbalance; dyslipidemias, *e.g.*, familial combined hyperlipidemia (FCHL), acquired hyperlipidemia; hypercholesterolemia; statin-resistant hypercholesterolemia; coronary artery disease (CAD) coronary heart disease (CHD) atherosclerosis. In one embodiment, the oligonucleotide agent that targets apoB-100 is administered to a subject diagnosed as having statin-resistant hypercholesterolemia.

25 The apoB-100 oligonucleotide agent can be administered in an amount sufficient to reduce levels of serum LDL-C and/or HDL-C and/or total cholesterol in a subject. In one embodiment, the oligonucleotide agent is administered in an amount sufficient to reduce the risk of myocardial infarction the subject.

In one embodiment, expression levels of apoB-100 are decreased in the liver following administration of the apoB-100 oligonucleotide agent. For example, the oligonucleotide agent

can be complexed with a moiety that targets the liver, *e.g.*, an antibody or ligand that binds a receptor on the liver.

In other embodiments, an oligonucleotide agent targeted to the liver can modulate expression of, *e.g.*, beta-catenin or glucose-6-phosphatase RNA, to treat a liver-related disorder.

5 In another embodiment, the oligonucleotide agent targets an miRNA or pre-miRNA expressed in the liver. In another embodiment, the human is suffering from a disorder characterized by overexpression or accumulation of the miRNA in the liver, or decreased expression of a nucleic acid that is the target of the miRNA expressed in the liver. Administration of the oligonucleotide agent to the subject, or to a cell of the lung of the subject,
10 can result in the pairing of the oligonucleotide agent with the target miRNA and the subsequent downregulation of the miRNA.

In one embodiment, the oligonucleotide agent is substantially identical to an miRNA normally expressed in liver tissue, and in another embodiment, the human is suffering from a disorder characterized by decreased expression of the miRNA in the liver. Administration of the
15 oligonucleotide agent to the subject, or to a cell of the liver, at least partially rescues the function of the downregulated miRNA.

TREATMENT OF PULMONARY DISEASE

An oligonucleotide agent featured in the invention can be used for treating a patient
20 diagnosed as having a pulmonary disease, *e.g.*, chronic bronchitis (including chronic bronchitis), emphysema, asthma (including pediatric asthma), chronic obstructive pulmonary disease (COPD), lung cancer, or a respiratory infection.

In one embodiment, an oligonucleotide agent targets, *e.g.*, a stress kinase (such as JNK, MAPK, or p38), a redox sensitive transcription factor (such as NF-kappa B, KJE, or AP-1),
25 interleukin-5 (IL-5) or the IL-5 receptor, phosphodiesterase 4, ICAM-1, CD11/CD18, E-selectin, interleukin-10, stem cell factor (SCF), MUC5AC, or adenosine A1 receptor. These oligonucleotide agents can be useful for the treatment of, *e.g.*, COPD or asthma, such as bronchial asthma.

Other gene targets for the treatment of COPD include beta-2 adrenergic receptor,
30 leukotriene D₄, 5'-lipoxygenase, interleukin-8, MCP-1, TNF-alpha, epidermal growth factor

receptor, tyrosine kinase, MUC4, MUC8, and matrix-degrading proteinases, such as serine elastase, ELA2, OMIM 130130 and neutrophil elastase.

Table 10. Potential gene targets for treatment of pulmonary disease.

5

Disease	Gene	OMIM number	Reference
asthma	adenosine A1 receptor	102755	Ball <i>et al.</i> , 2003
Cancer	Bcl family		Koty <i>et al.</i> , 2002
asthma and COPD	Beta-2 adrenergic receptor	109690	Barnes, 1999
asthma and COPD	CD11/CD18	600065	Barnes, 1999
asthma and COPD	E-selectin	131210	Barnes, 1999
Cancer	farnesyl transferase	134635, 134636	Scharovsky <i>et al.</i> , 2000
asthma and COPD	ICAM-1	147840	Barnes, 1999
asthma	IL-5	147850	Blumchen <i>et al.</i> , 2001
asthma	IL-5 receptor		Blumchen <i>et al.</i> , 2001
asthma and COPD	interleukin-10	124092	Barnes, 1999
asthma and COPD	interleukin-8	146930	Barnes, 1999
asthma and COPD	leukotriene D ₄		Barnes, 1999
asthma and COPD	MAP-kinase		Barnes, 1999
asthma and COPD	matrix metalloproteinases		Shapiro, 2002
asthma and COPD	MCP-1	158105	Barnes, 1999
COPD	MUC-4 and MUC-8	158372, 601932	Barnes, 1999
asthma	MUC5AC	158373	Fahy, 2002
asthma and	neutrophil elastase	130130	Wright <i>et al.</i> , 2002

COPD			
asthma and COPD	Phosphodiesterase 4		Barnes, 1999
Cancer	polo-like kinase-1	602098	Spankuch-Schmitt <i>et al.</i> , 2002
Cancer	R2 small subunit component of human ribonucleotide reductase	180390	Lee <i>et al.</i> , 2003
Cancer	ras		Scharovsky <i>et al.</i> , 2000
asthma	SCF	184745	Finotto <i>et al.</i> , 2001
asthma and COPD	serine elastase		Wright <i>et al.</i> , 2002
asthma and COPD	serine proteinases		Shapiro, 2002
Cancer	Stat3	102582	Song <i>et al.</i> , 2003
asthma and COPD	TNF-alpha	191160	Barnes, 1999
Cancer	VEGF receptors	191306	Pavco <i>et al.</i> , 2003
Viral diseases	viral RNA polymerase genes		Paddle, 2003; Mizuta <i>et al.</i> , 1999

Oligonucleotide agents described herein can be useful in the treatment of pathogenic infection. For example, a human infected with influenza A can be administered an oligonucleotide agent that targets influenza A PB2 or PA genes. Oligonucleotide agents that target genes of pathogens such as influenza A can also be useful in the treatment of victims of biowarfare attack.

An oligonucleotide agent can target a ras family gene or farnesyl transferase, Stat3, the R2 small subunit component of human ribonucleotide reductase, a Bcl gene (*e.g.*, Bcl-2), polo-like kinase-1 (PLK1), a VEGF receptors, anti-Flt-1 (VEGFR-1) or anti-KDR (VEGFR-2). These oligonucleotide agents would be useful for treating a lung cancer.

An oligonucleotide agent can target SCR, SCF, or the p65 subunit of NF-kappa B for the inhibition or treatment of a pulmonary reaction to an allergen. For example, these oligonucleotide agents can be used to inhibit or treat lung inflammation.

An oligonucleotide agent that targets a gene encoding the p65 subunit of NF-kappa B can be used to treat pneumonitis.

In one embodiment, the oligonucleotide agent targets an miRNA expressed in a tissue of the pulmonary system, *e.g.*, in the lung. In another embodiment, the human is suffering from a disorder characterized by overexpression or accumulation of the miRNA in the lung, or decreased expression of a nucleic acid that is the target of the miRNA expressed in the lung. Administration of the oligonucleotide agent to the subject, or to a cell of the lung of the subject, can result in the pairing of the oligonucleotide agent with the target miRNA and the subsequent downregulation of the miRNA.

An oligonucleotide agent useful for the treatment of asthma can be delivered directly to the lung, *e.g.*, via an inhaled aerosol. A liposomal delivery agent can be used to deliver an oligonucleotide agent to the lung. Oligonucleotide agents, and oligonucleotide agents that include phosphorothioate linkages in particular, can be administered via inhalation, and can subsequently localize to the bronchiolar and alveolar epithelium and endothelium. An oligonucleotide agent can be administered via inhalation at a dosage less than about 15 mg/kg, *e.g.*, less than 12 mg/kg, 10 mg/kg, 8 mg/kg, 6 mg/kg, 5 mg/kg, 4 mg/kg, 3 mg/kg, 2 mg/kg, 1 mg/kg, 0.1 mg/kg, or 0.001 mg/kg.

An oligonucleotide agent can be administered by intra-tracheal instillation, intranasally, or intravenously for the treatment of a pulmonary disorder.

TREATMENT OF VIRAL INFECTION

An oligonucleotide agent, *e.g.*, a conjugated oligonucleotide agent, can be used to target a viral gene, *e.g.*, a gene of a hepatitis C virus (HCV), hepatitis B virus (HBV), hepatitis A virus (HAV), HIV, Epstein-Barr virus, Respiratory syncytial virus, cytomegalovirus (CMV), human papilloma virus (HPV), or influenza, *e.g.*, influenza A.

An oligonucleotide agent can be used to treat a viral infection or a cancer. For example, an oligonucleotide agent can target an HPV, *e.g.*, HPV 16, 18, 31, 33, or 45, for the treatment of cervical neoplasia. An oligonucleotide agent can target, *e.g.*, E6, E7, or MCP-1 of HPV. In addition, or in an alternative, an oligonucleotide agent can target nucleolin in the host subject.

An oligonucleotide agent for the treatment of an HIV infection can target, *e.g.*, the gag, tat, vpr, rev, env, nef, pol, vir, or gp120 gene of the HIV virus.

An oligonucleotide agent that targets a viral gene can be delivered to a subject in the form of a recombinant adeno-vector or by retroviral-delivery. Delivery can be directly to a tumor tissue, for example.

In one embodiment, the oligonucleotide agent targets an miRNA expressed by a pathogen, *e.g.*, a viral pathogen or bacterial pathogen. For example, an oligonucleotide agent can target an miRNA expressed in a virus, such as a herpesvirus. The herpesvirus can be, for example, an Epstein-Barr virus (EBV or HHV4), a gammaherpesvirus (*e.g.*, a Kaposi sarcoma-associated virus (KSHV or HHV8), a mouse gammaherpesvirus 68 (MHV68)), or a betaherpesvirus (*e.g.*, a human cytomegalovirus (HCMV or HHV5)) (see Pfeffer *et al.*, *Science* 304:734-736, 2004, and Pfeffer *et al.*, *Nature Methods* 2:269-276, 2005). Administration of the oligonucleotide agent to the subject results in the pairing of the oligonucleotide agent with the target miRNA and the subsequent decrease in the interaction with the pathogenic miRNA with its target, *e.g.*, a target miRNA endogenous to the pathogen or to the host subject. This interaction results in a decrease in infection of the host by the pathogen.

OTHER EXEMPLARY THERAPEUTIC GENE TARGETS

An oligonucleotide agent, *e.g.*, a conjugated oligonucleotide agent described herein, can inhibit a gene involved in cellular adhesion, *e.g.*, ICAM-1, VCAM-1, or ELAM-1.

An oligonucleotide agent can inhibit a gene required for the regulation of cellular proliferation, *e.g.*, c-myc, vascular endothelial growth factor (VEGF), Ha-ras, A-raf kinase, c-raf kinase, or MRP.

An oligonucleotide agent can inhibit a gene involved in the pathogenesis of disease, *e.g.*, beta-thalassemia.

An oligonucleotide agent can inhibit a gene involved in the pathogenesis of disease caused by a pathogen. For example, a plasmodium can cause malaria PS1 or PSII, or a shistosoma can cause bloodfluke infections. An oligonucleotide agent that targets a gene of the pathogen can be used to treat the disease.

In one embodiment, the oligonucleotide agent binds an miRNA involved in the pathogenesis of disease.

An oligonucleotide agent can also be administered to human at risk for or afflicted with a neurological disease or disorder, *e.g.*, Alzheimer Disease or Parkinson Disease. For example, an

oligonucleotide agent can target an amyloid-family gene, *e.g.*, APP; a presenilin gene, *e.g.*, PSEN1 and PSEN2, or α -synuclein.

An oligonucleotide agent can be administered to a human to treat a neurodegenerative trinucleotide repeat disorder, *e.g.*, Huntington disease (HD), dentatorubral pallidoluysian atrophy (DRPLA) or a spinocerebellar ataxia, *e.g.*, SCA1, SCA2, SCA3 (Machado-Joseph disease), SCA7 or SCA8. For example, an oligonucleotide agent can decrease expression of an HD, DRPLA, SCA1, SCA2, MJD1, CACNL1A4, SCA7, or SCA8 gene.

Oligonucleotide agents featured in the invention can be used to treat a disease of the pancreas, *e.g.* pancreatitis, pancreatic cancer, diabetes or hyperglycemia. For example, an oligonucleotide agent that targets Ras, JNK, or survivin can be useful for treatment of pancreatic cancer. In one embodiment, the oligonucleotide agent can target an miRNA expressed in the pancreas, *e.g.*, miR-375, such as for the treatment of diabetes. In another embodiment, the oligonucleotide agent is substantially identical to an miRNA expressed in the pancreas, *e.g.*, miR-375.

Oligonucleotide agents featured in the invention can be used to treat a disease of the gut, *e.g.* gastroenteritis.

ROUTE OF DELIVERY

The oligonucleotide agents described herein can be administered by various routes of delivery, *e.g.*, by ocular, pulmonary, intravenous, topical, rectal, anal, or vaginal, delivery, *e.g.* as described in International Application Serial No. PCT/US2004/11829, filed April 16, 2004. The contents of this reference are incorporated herein in their entirety.

DOSAGE

In one aspect, the invention features a method of administering an oligonucleotide agent, to a subject (*e.g.*, a human subject). The method includes administering a unit dose of the oligonucleotide agent, *e.g.*, a microRNA, antisense RNA, decoy RNA or aptamer, that targets an RNA, *e.g.*, an miRNA, or protein in the subject (*e.g.*, an endogenous or pathogen target RNA or protein). In one embodiment, the unit dose is less than 1.4 mg per kg of bodyweight, or less than 10, 5, 2, 1, 0.5, 0.1, 0.05, 0.01, 0.005, 0.001, 0.0005, 0.0001, 0.00005 or 0.00001 mg per kg of bodyweight, and less than 200 nmole of oligonucleotide agent (*e.g.* about 4.4×10^{16} copies) per

kg of bodyweight, or less than 1500, 750, 300, 150, 75, 15, 7.5, 1.5, 0.75, 0.15, 0.075, 0.015, 0.0075, 0.0015, 0.00075, 0.00015 nmole of oligonucleotide agent per kg of bodyweight.

The defined amount can be an amount effective to treat or prevent a disease or disorder, *e.g.*, a disease or disorder associated with the target RNA, such as an RNA present in the kidney.

5 The unit dose, for example, can be administered by injection (*e.g.*, intravenous or intramuscular), an inhaled dose, or a topical application. Particularly preferred dosages are less than 2, 1, or 0.1 mg/kg of body weight.

In a preferred embodiment, the unit dose is administered less frequently than once a day, *e.g.*, less than every 2, 4, 8 or 30 days. In another embodiment, the unit dose is not administered
10 with a frequency (*e.g.*, not a regular frequency). For example, the unit dose may be administered a single time.

In one embodiment, the effective dose is administered with other traditional therapeutic modalities. In one embodiment, the subject has a viral infection and the modality is an antiviral agent other than an oligonucleotide agent. In another embodiment, the subject has
15 atherosclerosis and the effective dose of an oligonucleotide agent is administered in combination with, *e.g.*, after surgical intervention, *e.g.*, angioplasty.

In one embodiment, a subject is administered an initial dose and one or more maintenance doses of an oligonucleotide agent, or a precursor, *e.g.*, a larger oligonucleotide agent which can be processed into an oligonucleotide agent, or a DNA which encodes an
20 oligonucleotide agent, or precursor thereof. The maintenance dose or doses are generally lower than the initial dose, *e.g.*, one-half less of the initial dose. A maintenance regimen can include treating the subject with a dose or doses ranging from 0.01 μ g to 1.4 mg/kg of body weight per day, *e.g.*, 10, 1, 0.1, 0.01, 0.001, or 0.00001 mg per kg of bodyweight per day. The maintenance doses are preferably administered no more than once every 5, 10, or 30 days. Further, the
25 treatment regimen may last for a period of time, which will vary depending upon the nature of the particular disease, its severity and the overall condition of the patient. In preferred embodiments the dosage may be delivered no more than once per day, *e.g.*, no more than once per 24, 36, 48, or more hours, *e.g.*, no more than once for every 5 or 8 days. Following treatment, the patient can be monitored for changes in his condition and for alleviation of the symptoms of
30 the disease state. The dosage of the compound may either be increased in the event the patient does not respond significantly to current dosage levels, or the dose may be decreased if an

alleviation of the symptoms of the disease state is observed, if the disease state has been ablated, or if undesired side-effects are observed.

The effective dose can be administered in a single dose or in two or more doses, as desired or considered appropriate under the specific circumstances. If desired to facilitate
5 repeated or frequent infusions, implantation of a delivery device, *e.g.*, a pump, semi-permanent stent (*e.g.*, intravenous, intraperitoneal, intracisternal or intracapsular), or reservoir may be advisable.

In one embodiment, the oligonucleotide agent pharmaceutical composition includes a plurality of oligonucleotide agent species. In another embodiment, the oligonucleotide agent
10 species has sequences that are non-overlapping and non-adjacent to another species with respect to a naturally occurring target sequence. In another embodiment, the plurality of oligonucleotide agent species is specific for different naturally occurring target genes. In another embodiment, the oligonucleotide agent is allele specific.

In some cases, a patient is treated with an oligonucleotide agent in conjunction with other
15 therapeutic modalities. For example, a patient being treated for a kidney disease, *e.g.*, early stage renal disease, can be administered an oligonucleotide agent specific for a target gene known to enhance the progression of the disease in conjunction with a drug known to inhibit activity of the target gene product. For example, a patient who has early stage renal disease can be treated with an oligonucleotide agent that targets an SGLT2 RNA, in conjunction with the small molecule
20 phlorizin, which is known to block sodium-glucose cotransport and to subsequently reduce single nephron glomerular filtration rate. In another example, a patient being treated for a cancer of the kidney can be administered an oligonucleotide agent specific for a target essential for tumor cell proliferation in conjunction with a chemotherapy.

Following successful treatment, it may be desirable to have the patient undergo
25 maintenance therapy to prevent the recurrence of the disease state, wherein the compound of the invention is administered in maintenance doses, ranging from 0.01 μ g to 100 g per kg of body weight (see US 6,107,094).

The concentration of the oligonucleotide agent composition is an amount sufficient to be effective in treating or preventing a disorder or to regulate a physiological condition in humans.
30 The concentration or amount of oligonucleotide agent administered will depend on the parameters determined for the agent and the method of administration, *e.g.* nasal, buccal,

pulmonary. For example, nasal formulations tend to require much lower concentrations of some ingredients in order to avoid irritation or burning of the nasal passages. It is sometimes desirable to dilute an oral formulation up to 10-100 times in order to provide a suitable nasal formulation.

Certain factors may influence the dosage required to effectively treat a subject, including
5 but not limited to the severity of the disease or disorder, previous treatments, the general health and/or age of the subject, and other diseases present. Moreover, treatment of a subject with a therapeutically effective amount of an oligonucleotide agent, *e.g.*, a double-stranded oligonucleotide agent or a DNA which encodes an oligonucleotide agent, or precursor thereof) can include a single treatment or, preferably, can include a series of treatments. It will also be
10 appreciated that the effective dosage of an oligonucleotide agent used for treatment may increase or decrease over the course of a particular treatment. Changes in dosage may result and become apparent from the results of diagnostic assays as described herein. For example, the subject can be monitored after administering an oligonucleotide agent composition. Based on information from the monitoring, an additional amount of the oligonucleotide agent composition can be
15 administered.

Dosing is dependent on severity and responsiveness of the disease condition to be treated, with the course of treatment lasting from several days to several months, or until a cure is effected or a diminution of disease state is achieved. Optimal dosing schedules can be calculated from measurements of drug accumulation in the body of the patient. Persons of ordinary skill can
20 easily determine optimum dosages, dosing methodologies and repetition rates. Optimum dosages may vary depending on the relative potency of individual compounds, and can generally be estimated based on EC50s found to be effective in *in vitro* and *in vivo* animal models. In some embodiments, the animal models include transgenic animals that express a human gene, *e.g.* a gene that produces a target RNA. The transgenic animal can be deficient for the corresponding
25 endogenous RNA. In another embodiment, the composition for testing includes an oligonucleotide agent that is complementary, at least in an internal region, to a sequence that is conserved between the target RNA in the animal model and the target RNA in a human.

In one aspect, the invention features a method that includes: administering a first amount of a composition that comprises an oligonucleotide agent or a DNA which encodes an
30 oligonucleotide agent, *e.g.*, a double-stranded oligonucleotide agent or precursor thereof) to a subject, wherein the oligonucleotide agent is substantially complementary to a target nucleic

acid; evaluating an activity associated with a protein encoded by the target nucleic acid; wherein the evaluation is used to determine if a second amount should be administered. In a preferred embodiment the method includes administering a second amount of the composition, wherein the timing of administration or dosage of the second amount is a function of the evaluating. The method can include other features described herein.

In another aspect, the invention features a method of administering a source of an oligonucleotide agent to a subject. The method includes administering or implanting a source of an oligonucleotide agent. In one embodiment, the source releases the oligonucleotide agent over time, *e.g.* the source is a controlled or a slow release source, *e.g.*, a microparticle that gradually releases the oligonucleotide agent. In another embodiment, the source is a pump, *e.g.*, a pump that includes a sensor or a pump that can release one or more unit doses.

In one aspect, the invention features a pharmaceutical composition that includes a NAT oligonucleotide agent or a DNA which encodes an oligonucleotide agent, including a nucleotide sequence sufficiently complementary to a target RNA to allow duplex formation with a target nucleic acid. The target RNA can be a transcript of an endogenous human gene. In one embodiment, the oligonucleotide agent (a) is about 5 to about 100 nucleobases long, *e.g.*, about 8 to about 75, *e.g.*, about 8 to about 50 nucleotides long, *e.g.*, about 15 to about 30 nucleotides long, *e.g.*, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 nucleotides; and (b) is complementary to an endogenous target RNA. In one embodiment, the pharmaceutical composition can be an emulsion, microemulsion, cream, jelly, or liposome.

In certain other aspects, the invention provides kits that include a suitable container containing a pharmaceutical formulation of an oligonucleotide agent or a DNA which encodes an oligonucleotide agent or a precursor of an oligonucleotide agent). In certain embodiments the individual components of the pharmaceutical formulation may be provided in one container. Alternatively, it may be desirable to provide the components of the pharmaceutical formulation separately in two or more containers, *e.g.*, one container for an oligonucleotide agent preparation, and at least another for a carrier compound. The kit may be packaged in a number of different configurations such as one or more containers in a single box. The different components can be combined, *e.g.*, according to instructions provided with the kit. The components can be combined according to a method described herein, *e.g.*, to prepare and administer a pharmaceutical composition. The kit can also include a delivery device.

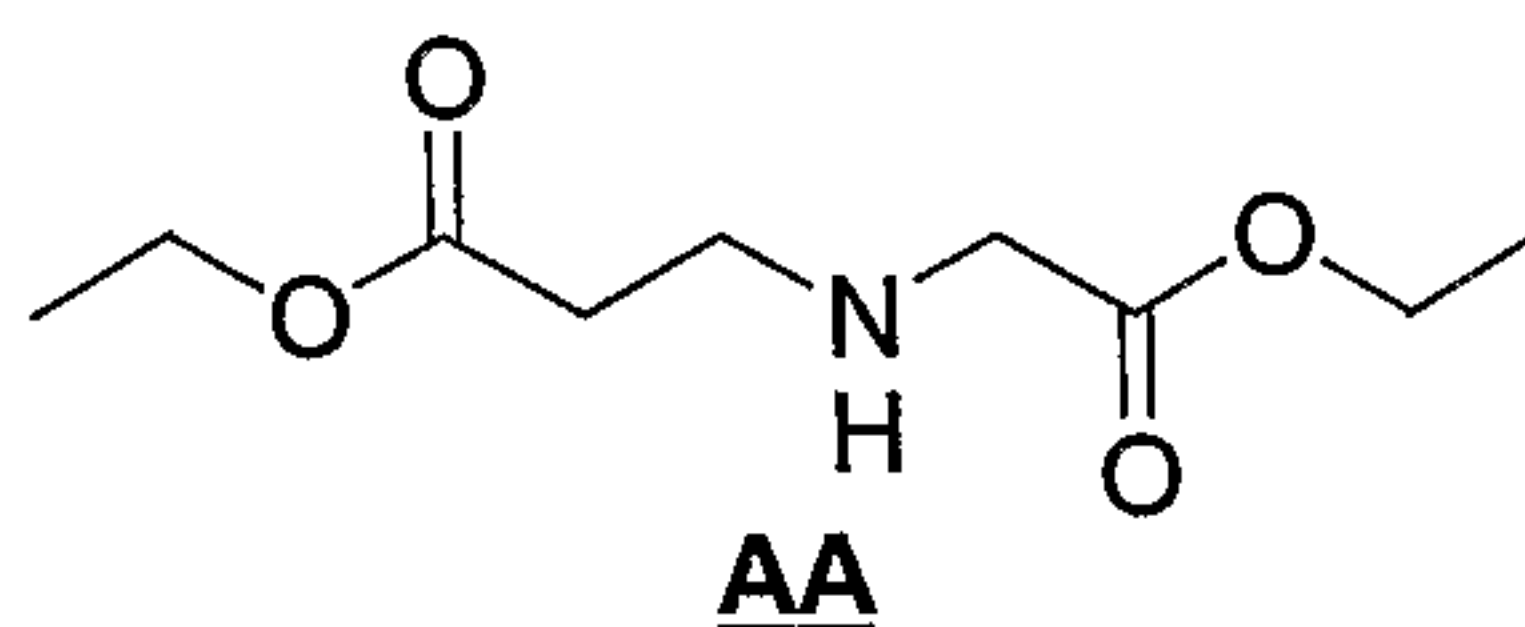
In another aspect, the invention features a device, *e.g.*, an implantable device, wherein the device can dispense or administer a composition that includes an oligonucleotide agent, or a precursor, *e.g.*, a larger oligonucleotide agent which can be processed into an oligonucleotide agent, or a DNA which encodes an oligonucleotide agent. The oligonucleotide agent can inhibit
5 expression of an endogenous transcript. In one embodiment, the device is coated with the composition. In another embodiment the oligonucleotide agent is disposed within the device. In another embodiment, the device includes a mechanism to dispense a unit dose of the composition. In other embodiments the device releases the composition continuously, *e.g.*, by diffusion. Exemplary devices include stents, catheters, pumps, artificial organs or organ
10 components (*e.g.*, artificial heart, a heart valve, etc.), and sutures.

The invention is further illustrated by the following examples, which should not be construed as further limiting.

EXAMPLES

Example 1:

Diethyl2-azabutane-1,4-dicarboxylate AA



5

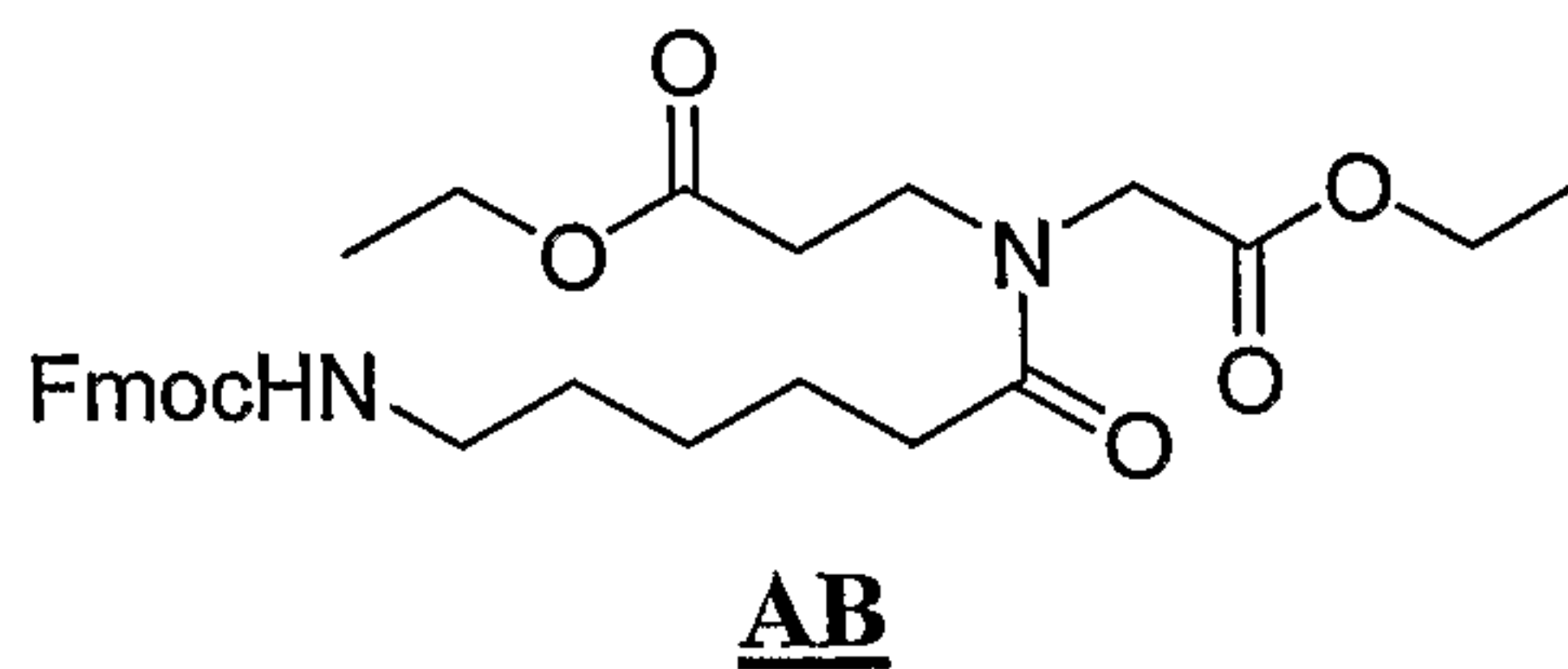
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A 4.7M aqueous solution of sodium hydroxide (50 mL) was added into a stirred, ice-cooled solution of ethyl glycinate hydrochloride (32.19 g, 0.23 mole) in water (50 mL). Then, ethyl acrylate (23.1 g, 0.23 mole) was added and the mixture was stirred at room temperature until the completion of reaction was ascertained by TLC (19 h). After 19 h which it was partitioned with dichloromethane (3 x 100 mL). The organic layer was dried with anhydrous sodium sulfate, filtered and evaporated. The residue was distilled to afford AA (28.8 g, 61%).

15

Example 2:

3-{Ethoxycarbonylmethyl-[6-(9H-fluoren-9-ylmethoxycarbonyl-amino)-hexanoyl]-amino}-propionic acid ethyl ester AB



20

25

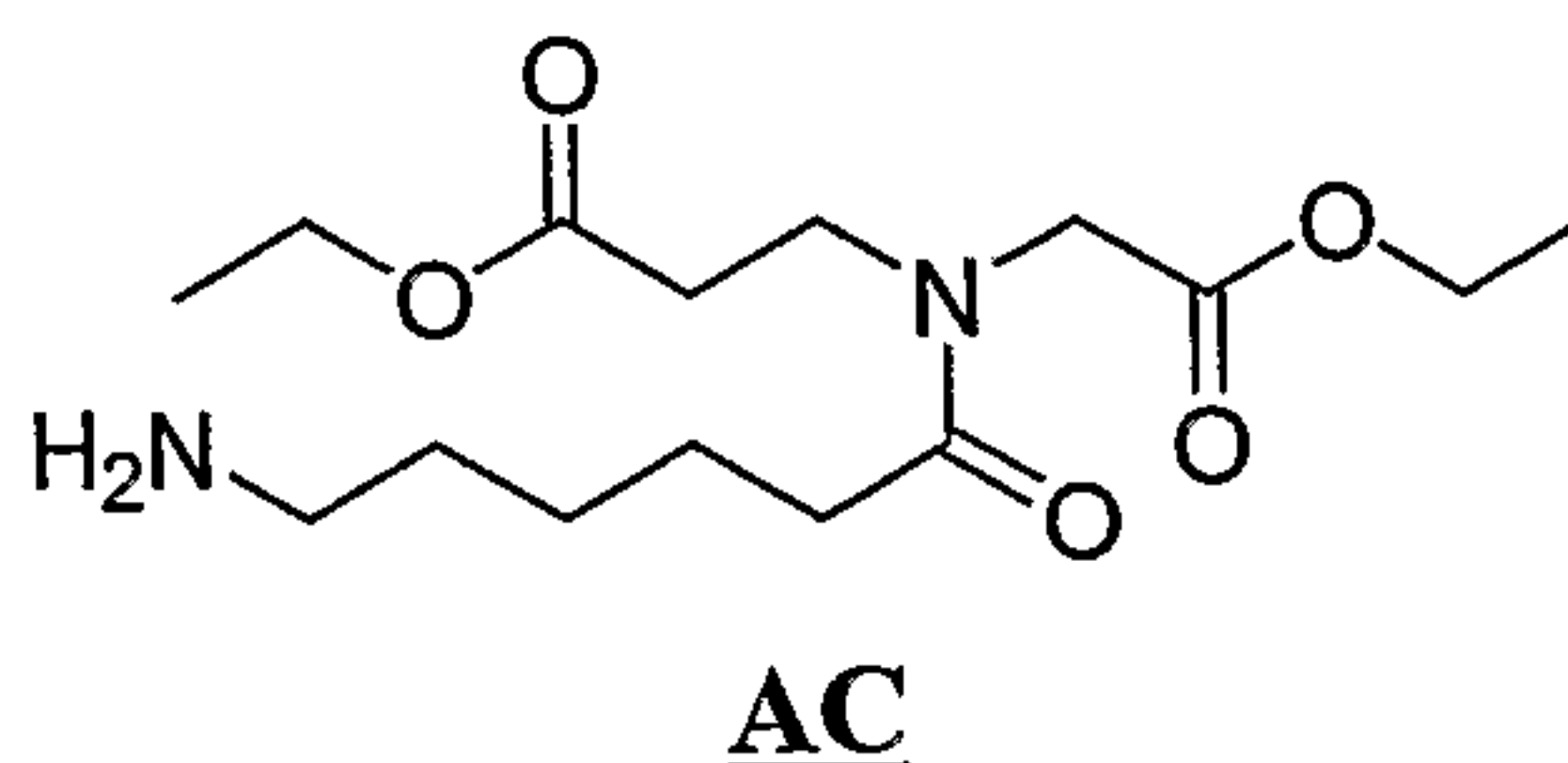
Fmoc-6-amino-hexanoic acid (9.12 g, 25.83 mmol) was dissolved in dichloromethane (50 mL) and cooled with ice. Diisopropylcarbodiimide (3.25 g, 3.99 mL, 25.83 mmol) was added to the solution at 0 °C. It was then followed by the addition of Diethyl2-azabutane-1,4-dicarboxylate (5 g, 24.6 mmol) and dimethylamino pyridine (0.305 g, 2.5 mmol). The solution was brought to room temperature and stirred further for 6 h. the completion of the reaction was ascertained by TLC. The reaction mixture was concentrated in vacuum and to the ethylacetate was added to precipitate diisopropyl urea. The suspension was filtered. The filtrate was washed

with 5% aqueous hydrochloric acid, 5% sodium bicarbonate and water. The combined organic layer was dried over sodium sulfate and concentrated to give the crude product which was purified by column chromatography (50 % EtOAC/Hexanes) to yield 11.87 g (88%) of **AB**

5

Example 3:

**3-[(6-Amino-hexanoyl)-ethoxycarbonylmethyl-amino]-
propionic acid ethyl ester AC**



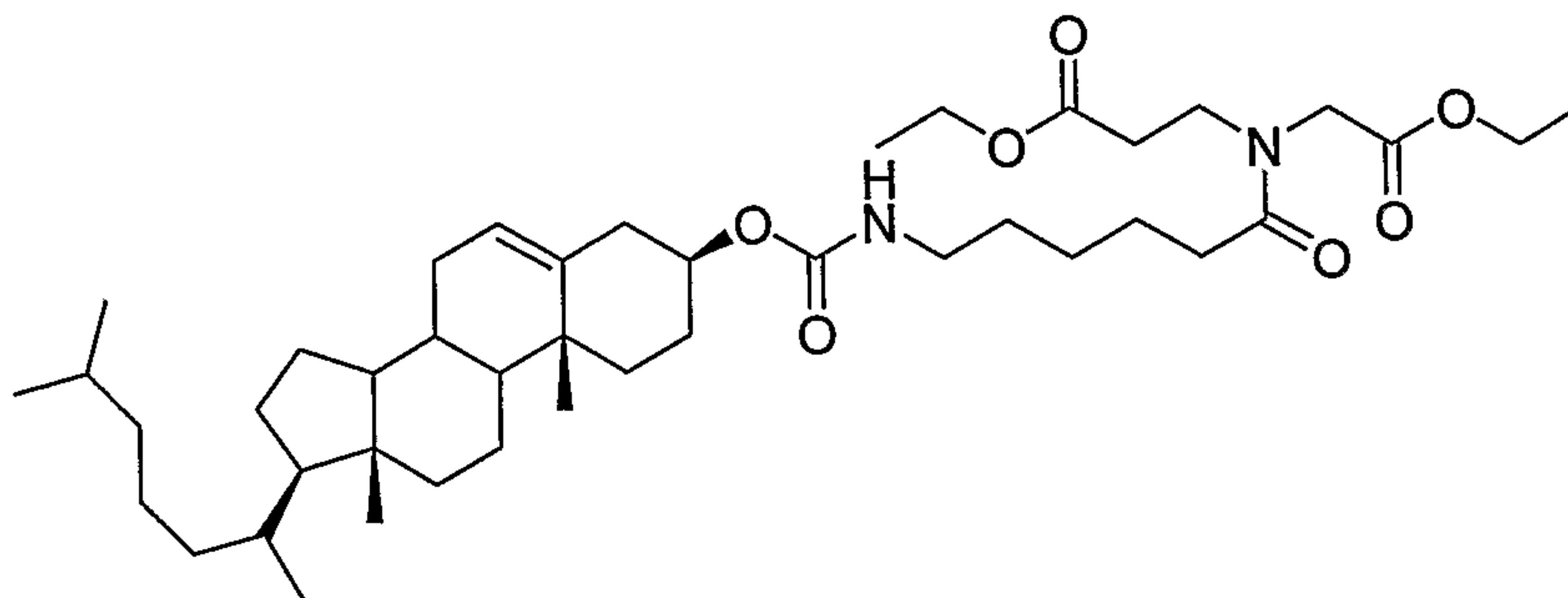
10

3-{Ethoxycarbonylmethyl-[6-(9H-fluoren-9-ylmethoxycarbonylamino)-hexanoyl]-amino}-propionic acid ethyl ester **AB** (11.5 g, 21.3 mmol) was dissolved in 20% piperidine in dimethylformamide at 0 °C. The solution was continued stirring for 1 h. The reaction mixture was concentrated in vacuum and the residue water was added and the product was extracted with ethyl acetate. The crude product was purified by converting into hydrochloride salt.

15

Example 4:

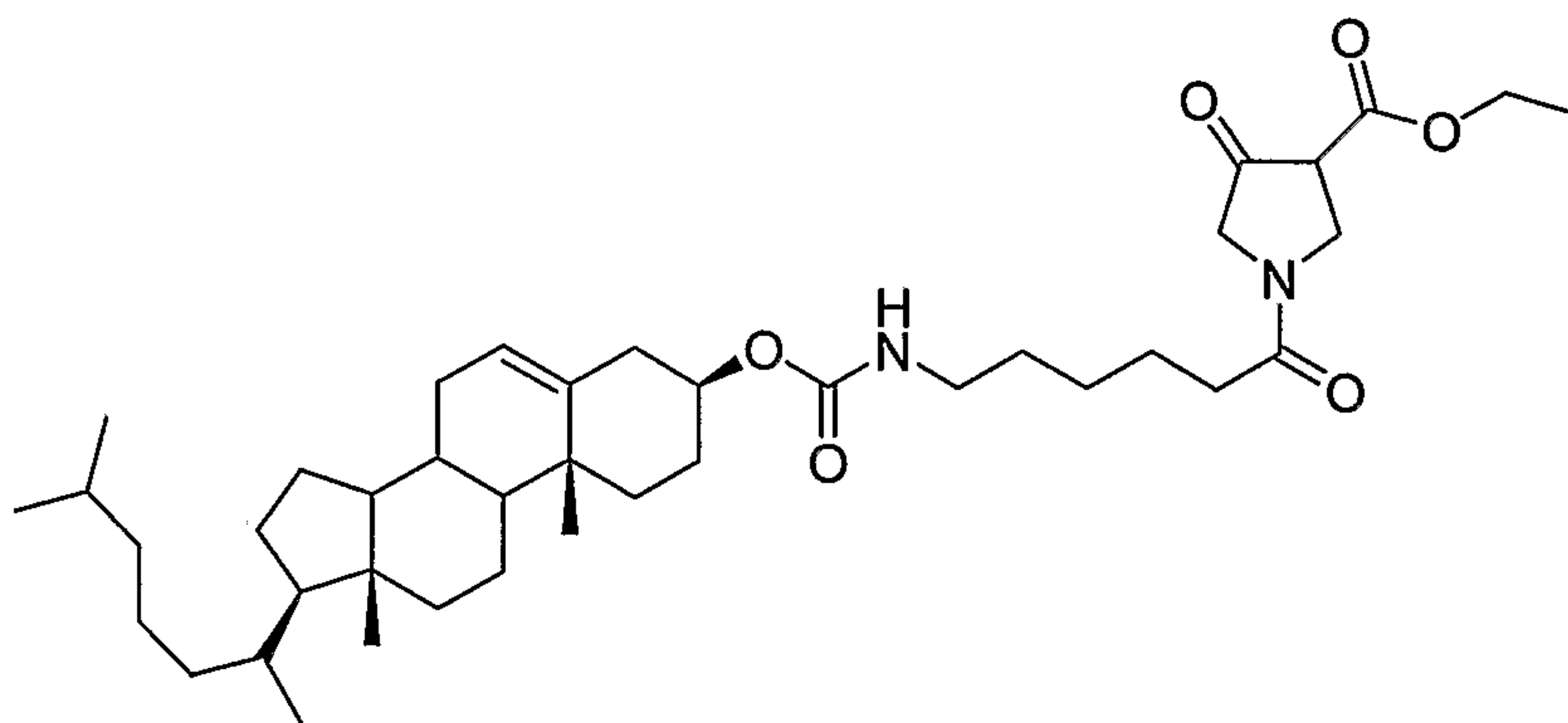
3-({6-[17-(1,5-Dimethyl-hexyl)-10,13-dimethyl-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1H-cyclopenta[a]phenanthren-3-yloxy-carbonylamino]-hexanoyl}ethoxycarbonylmethyl-amino)-propionic acid ethyl ester **AD**

**AD**

Hydrochloride salt of 3-[(6-Amino-hexanoyl)-ethoxycarbonylmethyl-amino]-propionic acid ethyl ester **AC** (4.7 g, 14.8 mmol) was taken in dichloromethane. The suspension was cooled to 0 °C with ice. To the suspension diisopropylethylamine (3.87 g, 5.2 mL, 30 mmol) was added. To the resulting solution cholesteryl chloroformate (6.675 g, 14.8 mmol) was added. The reaction mixture was stirred overnight. The reaction mixture was diluted with dichloromethane and washed with 10% hydrochloric acid. The product was purified flash chromatography (10.3 g, 92%).

Example 5:

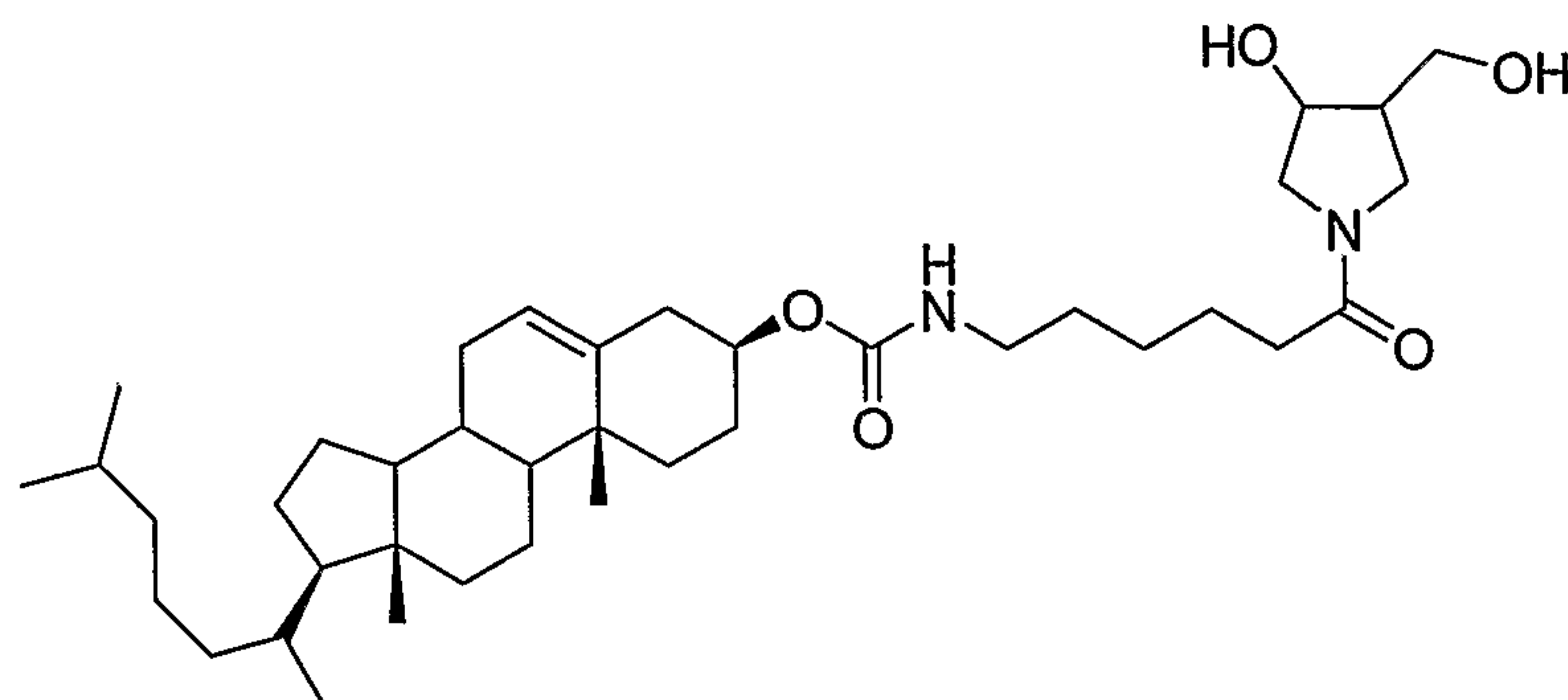
1-{6-[17-(1,5-Dimethyl-hexyl)-10,13-dimethyl-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1H-cyclopenta[a] phenanthren-3-yloxy-carbonylamino]-hexanoyl}-4-oxo-pyrrolidine-3-carboxylic acid ethyl ester AE

**AE**

Potassium t-butoxide (1.1 g, 9.8 mmol) was slurried in 30 mL of dry toluene. The mixture was cooled to 0 °C and 5 g (6.6 mmol) of diester was added slowly with stirring within 20 mins. The temperature was kept below 5 °C during the addition. The stirring was continued for 30 mins at 0 °C and 1 mL of glacial acetic acid was added, immediately followed by 4 g of NaH₂PO₄·H₂O in 40 mL of water. The resultant mixture was extracted with two 100 mL of dichloromethane and the combined organic extracts were washed twice with 10 mL of phosphate buffer, dried, and evaporated to dryness. The residue was dissolved in 60 mL of toluene, cooled to 0 °C and extracted with three 50 mL portions of cold pH 9.5 carbonate buffer. The aqueous extracts were converted to pH 3 with phosphoric acid, and extracted with five 40 mL portions of chloroform which were combined, dried and evaporated to a residue. The residue was purified by column chromatography using 25% ethylacetate/hexanes to afford 1.9 g of β-ketoester was obtained (39 %).

Example 6

[6-(3-Hydroxy-4-hydroxymethyl-pyrrolidin-1-yl)-6-oxo-hexyl]-carbamic acid 17-(1,5-dimethyl-hexyl)-10,13-dimethyl-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1H-cyclopenta[a]phenanthren-3-yl ester AF

AF

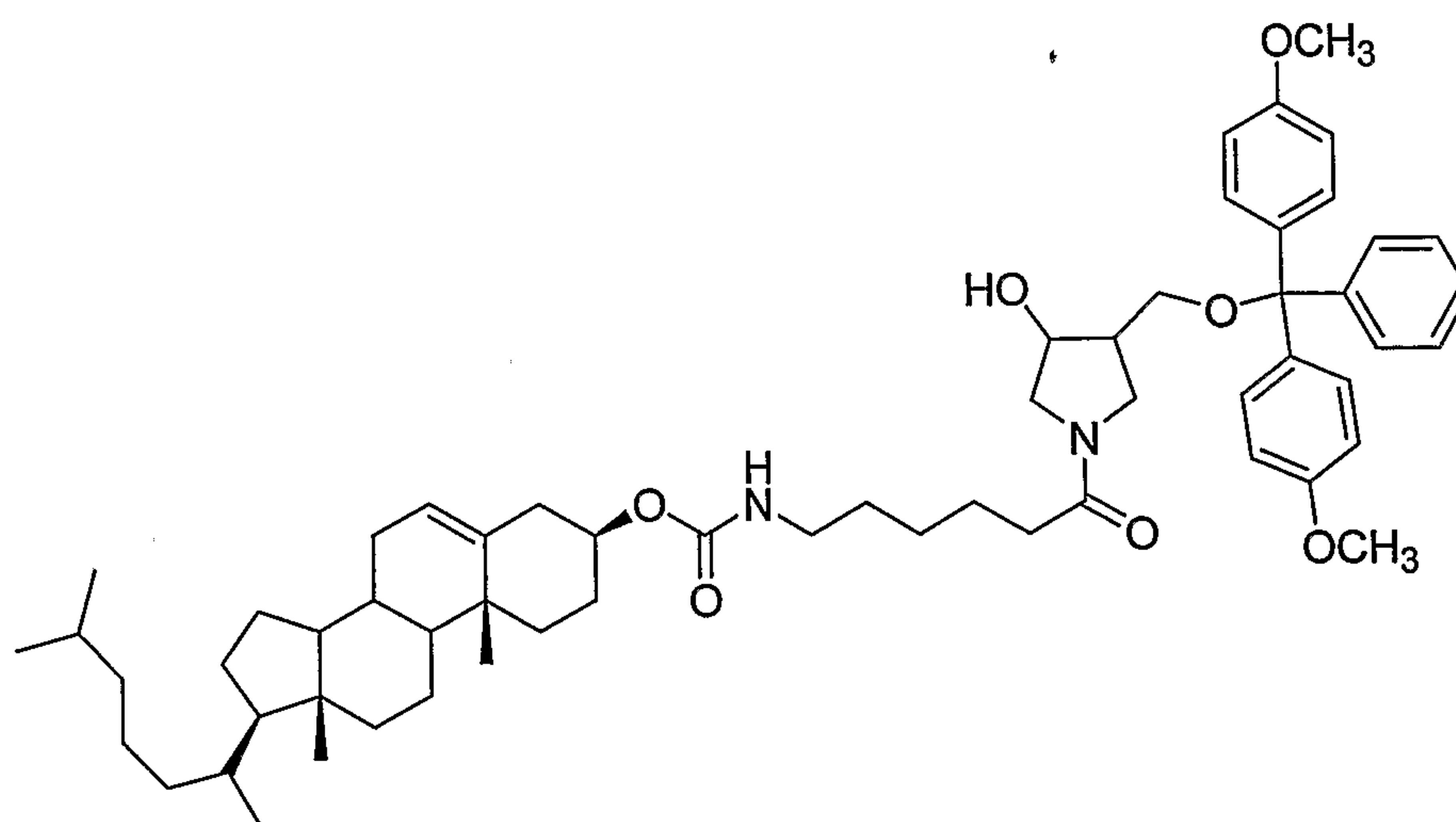
Methanol (2 mL) was added dropwise over a period of 1 h to a refluxing mixture of ketoester AE (1.5 g, 2.2 mmol) and sodium borohydride (0.226 g, 6 mmol) in tetrahydrofuran (10 mL). Stirring is continued at reflux temperature for 1 h. After cooling to room temperature, 1 N HCl (12.5 mL) was added, the mixture was extracted with ethylacetate (3 x 40 mL). The combined ethylacetate layer was dried over anhydrous sodium sulfate and concentrated in vacuum to yield the product which purified by column chromatography (10% MeOH/CHCl₃). (89%).

Example 7:

(6-{3-[Bis-(4-methoxy-phenyl)-phenyl-methoxymethyl]-4-hydroxy-pyrrolidin-1-yl}-
6-oxo-hexyl)-carbamic acid 17-(1,5-dimethyl-hexyl)-10,13-dimethyl-

5 2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1H-cyclopenta[a]phenanthren-3-yl ester

AG



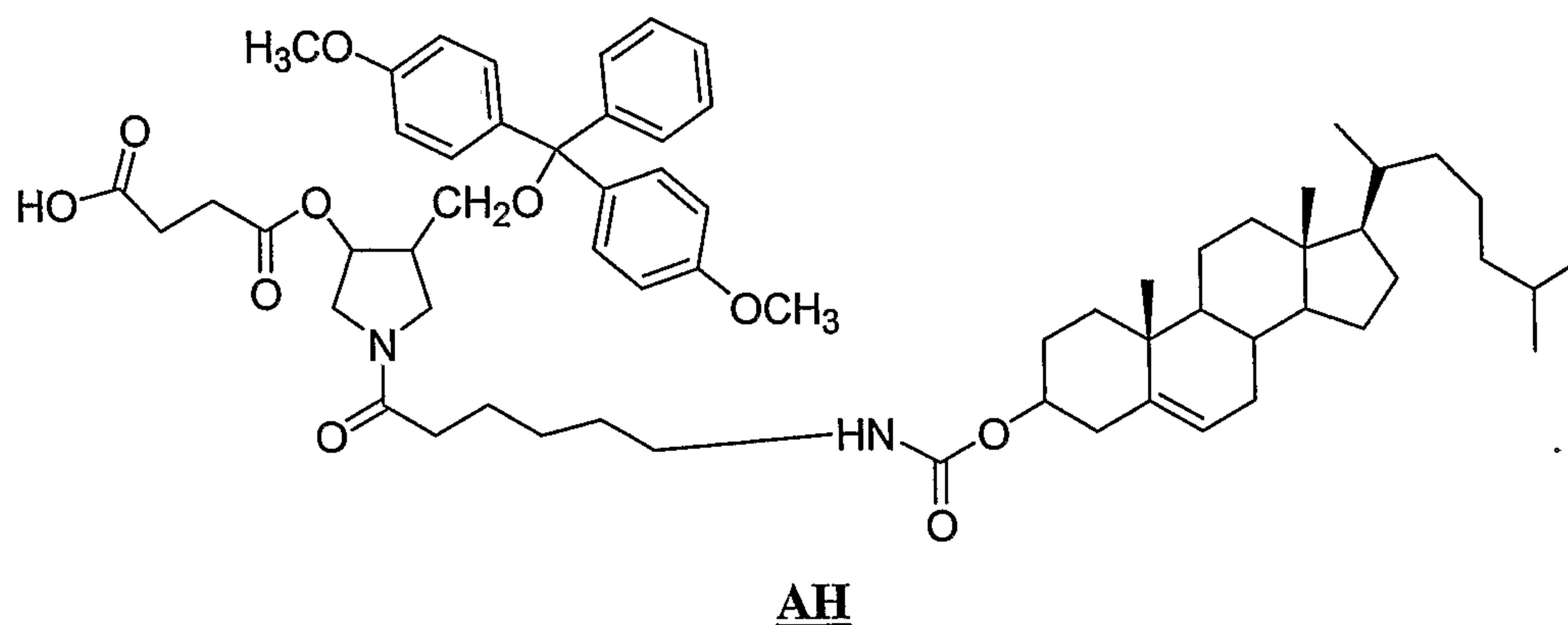
AG

10 Diol **AF** (1.25 gm 1.994 mmol) was dried by evaporating with pyridine (2 x 5 mL) in vacuo. Anhydrous pyridine (10 mL) and 4,4'-dimethoxytritylchloride (0.724 g, 2.13 mmol) were added with stirring. The reaction was carried out at room temperature for overnight. The reaction was quenched by the addition of methanol. The reaction mixture was concentrated in vacuum and to the residue dichloromethane (50 mL) was added. The organic layer was washed with 1M
15 aqueous sodium bicarbonate. The organic layer was dried over anhydrous sodium sulfate, filtered and concentrated. The residual pyridine was removed by evaporating with toluene. The crude product was purified by column chromatography (2% MeOH/Chloroform, $R_f = 0.5$ in 5% MeOH/ CHCl_3). (1.75 g, 95%)

Example 8

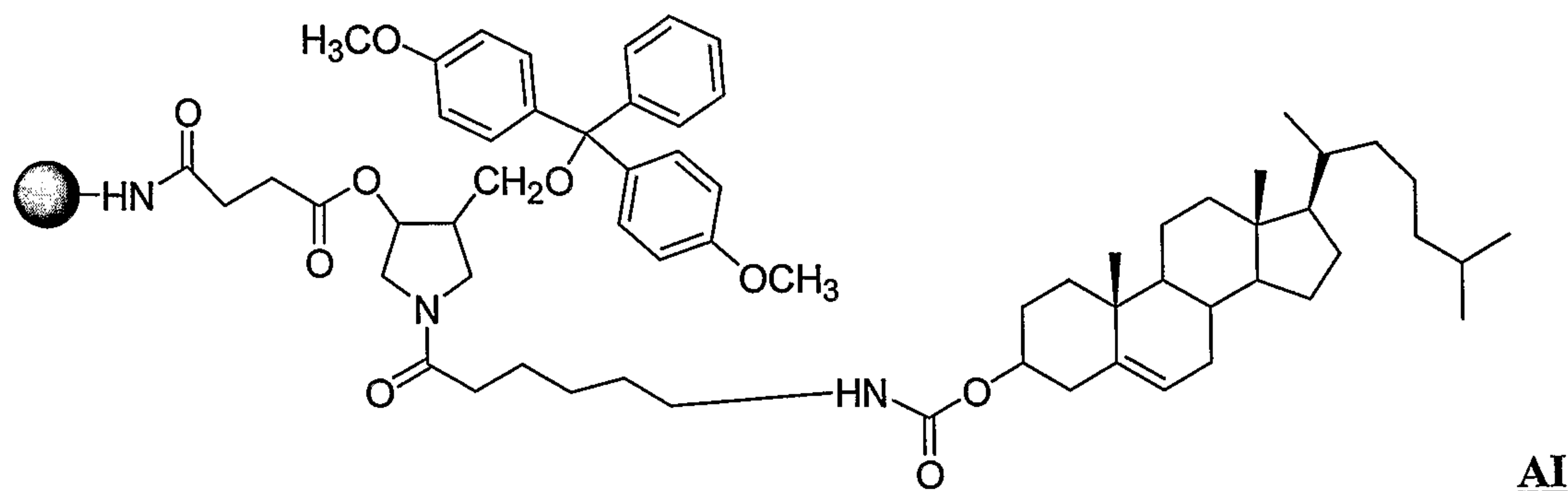
Succinic acid mono-(4-[bis-(4-methoxy-phenyl)-phenyl-methoxymethyl]-1-{6-[17-(1,5-dimethyl-hexyl)-10,13-dimethyl 2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1H cyclopenta[a]phenanthren-3-yloxy-carbonylamino]-hexanoyl}-pyrrolidin-3-yl) ester AH

5



Compound AG (1.0 g, 1.05 mmol) was mixed with succinic anhydride (0.150 g, 1.5 mmol) and DMAP (0.073 g, 0.6 mmol) and dried in a vacuum at 40 °C overnight. The mixture was dissolved in anhydrous dichloroethane (3 mL), triethylamine (0.318 g, 0.440 mL, 3.15 mmol) was added and the solution was stirred at room temperature under argon atmosphere for 16 h. It was then diluted with dichloromethane (40 mL) and washed with ice cold aqueous citric acid (5 wt%, 30 mL) and water (2 X 20 mL). The organic phase was dried over anhydrous sodium sulfate and concentrated to dryness. The residue was used as such for the next step.

Example 9

cholesterol derivatised CPG AI

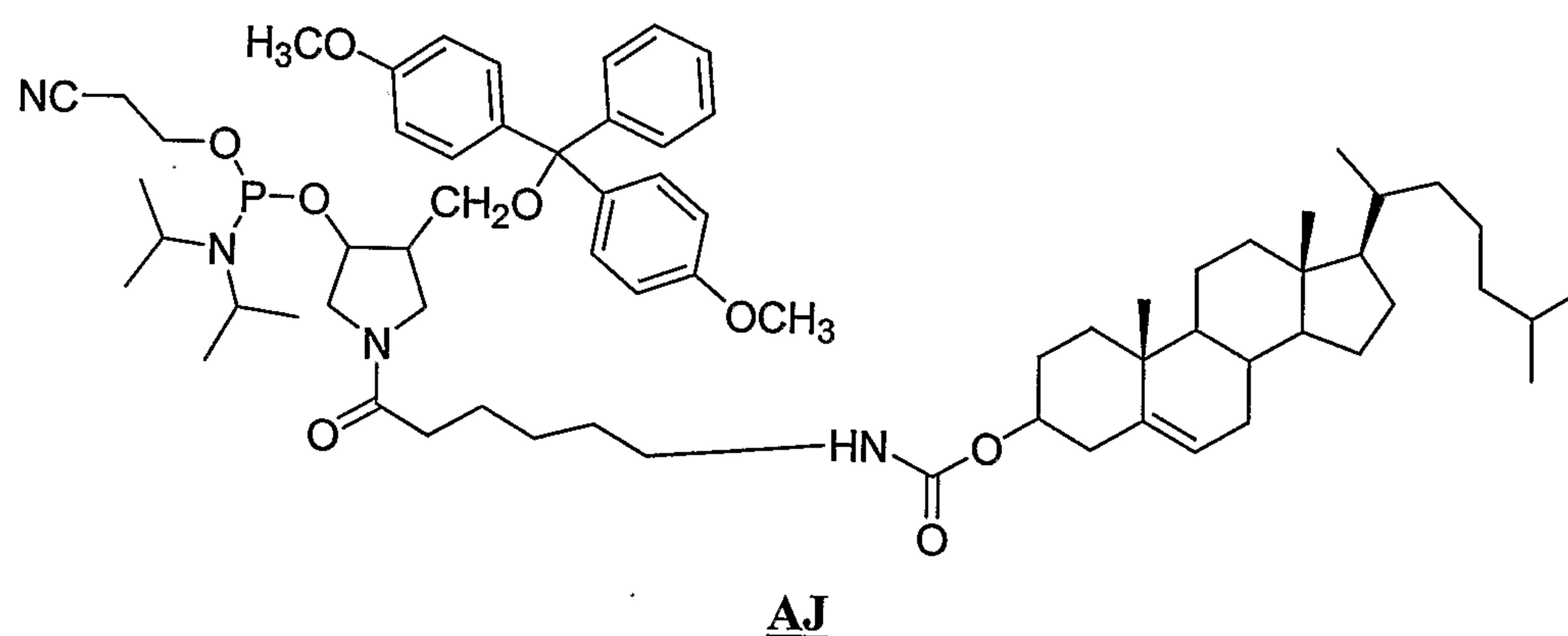
5

Succinate AH (0.254 g, 0.242 mmol) was dissolved in mixture of dichloromethane/acetonitrile (3:2, 3 mL). To that solution DMAP (0.0296 g, 0.242 mmol) in acetonitrile (1.25 mL), 2,2'-Dithio-bis(5-nitropyridine) (0.075 g, 0.242 mmol) in acetonitrile/dichloroethane (3:1, 1.25 mL) were added successively. To the resulting solution triphenylphosphine (0.064 g, 0.242 mmol) in acetonitrile (0.6 ml) was added. The reaction mixture turned bright orange in color. The solution was agitated briefly using wrist-action shaker (5 mins). Long chain alkyl amine-CPG (LCAA-CPG) (1.5 g, 61 $\mu\text{m/g}$) was added. The suspension was agitated for 2 h. The CPG was filtered through a sintered funnel and washed with acetonitrile, dichloromethane and ether successively. Unreacted amino groups were masked using acetic anhydride/pyridine. The loading capacity of the CPG was measured by taking UV measurement. (37 $\mu\text{M/g}$).

15

Example 10

(4-[bis-(4-methoxy-phenyl)-phenyl-methoxymethyl]-1-{6-[17-(1,5-dimethyl-hexyl)-
10,13-dimethyl 2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1H
cyclopenta[a]phenanthren-3-yloxycarbonylamino]-hexanoyl}-pyrrolidin-3-yl)
phosphoramidite AJ



Compound AG (0.15 g, 0.158 mmol) was coevaporated with toluene (5 mL). To the residue N,N-tetraisopropylammonium tetrazolide (0.0089 g, 0.079 mmol) was added and the mixture was dried over P₂O₅ in a vacuum oven for overnight at 40 °C. The reaction mixture was dissolved in the mixture of anhydrous acetonitrile/dichloromethane (2;1, 1 mL) and 2-cyanoethyl-N,N,N',N'-tetraisopropylphosphoramidite (0.0714 g, 0.0781 mL, 0.237 mmol) was added. The reaction mixture was stirred at ambient temperature for overnight. The completion of the reaction was ascertained by TLC (1;1 ethyl acetate:hexane). The solvent was removed under reduced pressure and the residue was dissolved in ethyl acetate (10 mL) and washed with 5% NaHCO₃ (4 mL) and brine (4 mL). The ethyl acetate layer was dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The resulting mixture was chromatographed (50:49:1, EtOAc:Hexane:triethylamine) to afford AJ as white foam (0.152 g, 84%).

Example 11

RNA Synthesis, Deprotection and Purification Protocol

1. Synthesis:

The RNA molecules were synthesized on a 394 ABI machine using the standard 93 step cycle written by the manufacturer with modifications to a few wait steps as described below. The solid support was controlled pore glass (CPG, 1 μ mole, 500Å, Glen Research, Sterling VA) and the monomers were RNA phosphoramidites with standard protecting groups (N⁶-benzoyl-5'-O-dimethoxytrityladenosine-2'-*t*butyldimethylsilyl- 3'-O-N,N'-diisopropyl-2-cyanoethylphosphoramidite, 5'-O-dimethoxytrityluridine-2'-*t*butyldimethylsilyl- 3'-O-N,N'-diisopropyl-2-cyanoethylphosphoramidite, N²-isobutyryl-5'-O-dimethoxytritylguanosine-2'-*t*butyldimethylsilyl, 3'-O-N,N'-diisopropyl-2-cyanoethylphosphoramidite, and N⁴-benzoyl-5'-O-dimethoxytritylcytidine-2'-*t*butyldimethylsilyl-3'-O-N,N'-diisopropyl-2-cyanoethylphosphoramidite from Chemgenes Corp MA) used at a concentration of 0.15M in acetonitrile (CH₃CN) and a coupling time of 7.5min. The activator was thiotetrazole (0.25M), For the PO-oxidation Iodine/Water/Pyridine was used and the PS-oxidation Beaucage reagent 0.5M solution in acetomitrile was used. All reagents for synthesis were also from Glen Research.

2. Deprotection- I (Oligomer cleavage, Base and Phosphate deprotection)

After completion of synthesis the controlled pore glass (CPG) was transferred to a screw cap vial (Fisher, catalog number 03-340-5N) or a screw cap RNase free microfuge tube. The oligonucleotide was cleaved from the CPG with simultaneous deprotection of base and phosphate groups with 1.0 mL of a mixture of ethanolic ammonia [ammonia: ethanol (3:1)] for 6 hours to overnight at 55°C. The vial was cooled briefly on ice and then the ethanolic ammonia mixture was transferred to a new microfuge tube. The CPG was washed with 3 x 0.25 mL portions of 50% acetonitrile (70% CH₃CN for cholesterol and such hydrophobic conjugated oligomers). The approximate 1.75 mL of solution is best divided equally into two microfuge tubes, capped tightly and then cooled at -80°C for 15 min, before drying in a speed vac/lyophilizer for about 90min.

3. Deprotection-II (Removal of 2' TBDMS group)

The white residue obtained was resuspended in 200 μ L of triethylamine trihydrofluoride (TEA.3HF, Aldrich) and heated at 65°C for 1.5h to remove the tertbutyldimethylsilyl (TBDMS) groups at the 2' position. The reaction was then quenched with 400 μ L of
5 isopropoxytrimethylsilane (*i*PrOMe₃Si Aldrich) and further incubated on the heating block leaving the caps open for 15min; (This causes the volatile isopropoxytrimethylsilylfluoride adduct to vaporize). The residual quenching reagent was removed by drying in a speed vac. The oligomer was then precipitated in anhydrous methanol (MeOH, 800 μ L). The liquid was removed very carefully after spinning in a centrifuge for 5 minutes on the highest speed
10 available. Residual methanol was removed by drying briefly in a speed vac after freezing at -80°C. The crude RNA was obtained as a white fluffy material in the microfuge tube.

4. Quantitation of Crude Oligomer or Raw Analysis

Samples were dissolved in 50% aqueous acetonitrile (0.5mL) and quantitated as follows:
15 Blanking was first performed with 50% aqueous acetonitrile alone (1mL).

5 μ L of sample and 995 μ L of 50% acetonitrile, were mixed well in a microfuge tube, transferred to cuvette and absorbance reading obtained at 260 nm. The crude material is dried down and stored at -20°C.

20 5. Purification of Oligomers

The crude oligomers were analyzed and purified by HPLC (Mono Q Pharmacia Biotech 5/50). The buffer system is A = 100mM Tris HCl 10% HPLC grade acetonitrile pH = 8, B = 100mM Tris-HCl pH 8, 10% HPLC grade acetonitrile 1 M NaCl, flow 1.0 mL/min, wavelength 260 nm. For the unmodified RNA 21mer a gradient of 0-0.6M NaCl is usually adequate. One can
25 purify a small amount of material (~5 OD) and analyze by CGE or MS. Once the identity of this material is confirmed the crude oligomer can then be purified using a larger amount of material. i.e 40 OD's per run, flow rate of 1mL/min and a less sensitive wavelength of 280 nm to avoid saturation of the detector. Fractions containing the full length oligonucleotides are then pooled together, evaporated and finally desalted as described below.

6. Desalting of Purified Oligomer

The purified dry oligomer was then desalted using either C-18 Sepak cartridges (Waters) or Sephadex G-25M (Amersham Biosciences). The cartridge was conditioned with 10 mL each of acetonitrile, followed 50% acetonitrile, 100 mM buffer (this can be triethylammonium acetate, sodium acetate or ammonium acetate). Finally the purified oligomer dissolved thoroughly in 10mL RNase free water was applied to the cartridge with very slow dropwise elution. The cartridge was washed with water (10mL) to remove salts. And finally the salt free oligomer was eluted with 50% acetonitrile or 50% methanol directly into a screw cap vial.

7. Capillary Gel Electrophoresis (CGE) and Electrospray LC/MS

1 μ L of approximately 0.04 OD oligomer is first dried down, redissolved in water (2 μ L) and then pipetted in special vials for CGE and LC/MS analysis. In general, desalting should be carried out prior to analysis.

Example 14**5' CHOLESTEROL-CUUACGCUGAGUACUUCGAdTdT-3'****(SEQ ID NO: 55)**

Compound 14-a was used to synthesize oligonucleotide conjugates where cholesterol was conjugated at the 5' end of RNA molecules.

The phosphoramidite 14-a was dissolved in acetomitrile/methylene chloride 1:1 solution to give a 0.2M solution. This was used for the terminal coupling during the oligonucleotide synthesis. For the PO-oxidation Iodine/Water/Pyridine was used and the PS-oxidation Beaucage reagent 0.5M solution in acetomitrile was used. The diamathoxy triotyl group was removed in the synthesizer.

EXAMPLE 15**siRNA Modifications Enhanced Duplex Stability**

Radiolabel method for monitoring serum stability of siRNA duplexes: siRNA duplexes were prepared at a stock concentration of 1 μ M in which either the sense (S) or antisense strand (AS) contained a trace amount of 5'-³²P labeled material (*e.g.* ³²P-S/AS and S/³²P-AS). The presence of the end-labeled sense or antisense strand allowed for monitoring of the individual strand within the context of the siRNA duplex. Therefore, two duplex preparations were made for each siRNA sequence tested. siRNA duplexes were incubated in 90% human serum at a final concentration of 100nM duplex. Samples were removed and quenched in a stop mix at appropriate times. For a typical time course, 10 seconds, 15 minutes, 30 minutes, 1 hour, 2 hours and 4 hours time points were taken. Samples were analyzed by denaturing polyacrylamide gel electrophoresis along with a control sample (4 hour buffer-alone incubation) and a partial alkaline hydrolysis ladder of the labeled sense or antisense strand as a marker. The gel was imaged using a Fuji phosphorimager to detect the full length sense and antisense strands along with any degradation fragments that were generated by serum nucleases during incubation.

Since there is the possibility of losing the 5' phosphate label due to phosphatase activity in the serum, an alternative to 5' end labeling is to place an internal ³²P or ³³P label within either

the sense or antisense strand. This labeling method is much more laborious than 5' end labeling and currently we have no evidence that dephosphorylation occurs during serum incubation.

A series of chemical modifications that fall into the following categories; backbone modification, sugar modification, nucleobase modification and 3' conjugate, were tested and showed enhanced serum stability as compared to a unmodified siRNA duplex. A description of each modification, its location within the siRNA duplex, and the serum stability data follows.

Serum stability of unmodified parent duplex: The unmodified parent duplex, AL-DUP-1000, was used to establish the serum stability baseline for evaluating the effect of chemical modifications on nuclease resistance.

AL-DUP-1000

5' -CUUACGCUGAGUACUUCGAdTdT-3' ALN-SEQ-1000 SEQ ID NO:
3' dTdTGA AUGCGACUCAUGAAGCU-5' ALN-SEQ-1001 SEQ ID NO:

AL-DUP-1000 was subjected to the serum stability assay to evaluate its inherent nuclease resistance and to define its degradation pattern (FIG. 14). Denaturing gel electrophoresis was used to analyze AL-DUP-1000 in a human serum stability assay. An siRNA duplex containing 5' end-labeled sense RNA (*s/as) and a duplex containing 5' end-labeled antisense RNA (as/s*) were each incubated in 90% human serum and time points were assayed at 10 seconds, 5min, 15min, 30min, 1hour, 2hours and 4 hours. The control was a 4 hour time point for siRNA duplex incubated in PBS buffer alone, OH⁻ was the partial alkaline hydrolysis marker. This unmodified duplex was observed to be degraded by both 3'-5' exonucleases and endonucleases (FIG. 14).

Cleavage of the 3' end of both the sense and antisense strands by 3'-5' exonucleases occurs within the first 5 minutes of incubation resulting in the loss of the 3' terminal dT residues (top vertical lines in s*/as and s/as* panels of FIG. 14). In addition to exonuclease degradation, both strands were cleaved by endonucleases. There was a major endonuclease site at position sixteen of the antisense strand (bottom vertical lines in s*/as and s/as* panels of FIG. 14) that appears as early as 10 seconds. Very little full length sense or antisense strand was remaining after 1 hour in human serum. Chemical modifications were introduced within the context of the parent duplex to evaluate their effect on nuclease resistance. These chemical modifications fall

within one of the following classes: backbone modification, sugar modification, nucleobase modification, cationic modification and conjugate.

Backbone modifications enhanced nuclease resistance: Specific phosphodiester linkages of the siRNA duplex were replaced by either phosphorothioate or methylphosphonate and their stability was evaluated in the human serum stability assay. Table 11 contains the sequences of the duplexes tested. Substitution of the phosphodiester linkage at the 3' end of both the sense and antisense strands inhibited exonucleolytic degradation of the 3' overhangs (FIGs. 15A and 15B) as compared to the unmodified parent duplex (refer to FIG. 14). Full length starting material was present for four hours for both the sense and antisense strands. The endonucleolytic cleavage pattern seen in the unmodified duplex was unchanged. Similar results were obtained for duplexes that contained additional phosphorothioates at their 3' ends (data not shown). The placement of phosphorothioates at the endonucleolytic cleavage sites (duplexes 1419, 1420 and 1421) did not inhibit endonucleolytic cleavage at these sites (data not shown). In summary, a single phosphorothioate or methylphosphonate between the two 3' terminal nucleotides was sufficient to protect the 3' ends from exonuclease degradation. Additional phosphorothioates at the 3' ends appear to enhance this effect, which may be necessary for long term exposure to serum nucleases.

Table 1 siRNA duplexes containing backbone modifications. (* = phosphorothioate, mp = methylphosphonate)		
Alnylam Duplex	Duplex Sequence	Alnylam Sequence
AL-DUP-1393	5' - CUUACGCUGAGUACUUCGAdT*dT-3' 3' - dT*dTGAAUGCGACUCAUGAAGCU-5'	AL-SEQ-1026 AL-SEQ-1027
AL-DUP-1394	5' - CUUACGCUGAGUACUUCGA*dT*dT-3' 3' - dT*dT*GAAUGCGACUCAUGAAGCU-5'	AL-SEQ-1028 AL-SEQ-1029
AL-DUP-1395	5' - CUUACGCUGAGUACUUCG*A*dT*dT-3' 3' - dT*dT*G*AAUGCGACUCAUGAAGCU-5'	AL-SEQ-1030 AL-SEQ-1031
AL-DUP-1396	5' - CUUACGCUGAGUACUUC*G*A*dT*dT-3' 3' - dT*dT*G*A*AUGCGACUCAUGAAGCU-5'	AL-SEQ-1032 AL-SEQ-1033
AL-DUP-1419	5' - CUUACGCUGAGU*ACUUCGAdTdT-3' 3' - dTdTGAAUGCGACUCA*UGAAGCU-5'	AL-SEQ-2182 AL-SEQ-2184
AL-DUP-1420	5' - CUU*ACGCUGAGU*ACUUCGAdTdT-3' 3' - dTdTGAA*UGCGACUCA*UGAAGCU-5'	AL-SEQ-2183 AL-SEQ-2185
AL-DUP-1421	5' - CUU*ACGCUGAGU*ACUUCGAdT*dT-3' 3' - dT*dTGAA*UGCGACUCA*UGAAGCU-5'	AL-SEQ-2186 AL-SEQ-2188
AL-DUP-1329	5' - CUUACGCUGAGUACUUCGAdTmPdT-3' 3' - dTmPdTGAAUGCGACUCAUGAAGCU-5'	AL-SEQ-1038 AL-SEQ-1039

Sugar modifications enhanced nuclease resistance: The effect of replacing the 2'OH with 2'OMe was evaluated at the sites of endonucleolytic cleavage as well as at the 3' ends of the siRNA duplex. The duplexes tested in the human serum stability assay are shown in Table 12. Some of these duplexes also contained phosphorothioate linkages to evaluate whether the combination of the two modifications enhance nuclease resistance more significantly. Substitution of the terminal dT residues with 2'OMe-U (AL-DUP-1027) reduced 3'-5' exonuclease degradation slightly over the unmodified parent duplex (data not shown); however, the extent of exonuclease protection by 2'OMe-U was far less than that achieved by placing a

Table 4 siRNA duplexes containing 2'OMe substitutions. (U = 2'OMe-uridine, *= phosphorothioate)		
Alnylam Duplex	Duplex Sequence	Alnylam Sequence
AL-DUP-1027	5' - CUUACGCUGAGUACUUCGAUU - 3' 3' - UUGAAUGCGACUCAUGAAGCU - 5'	AL-SEQ-1006
		AL-SEQ-1007
AL-DUP-1036	5' - C*UUACGCUGAGUACUUCGAU*U - 3' 3' - U*UGAAUGCGACUCAUGAAGC*U - 5'	AL-SEQ-1008
		AL-SEQ-1009
AL-DUP-13ff	5' - C*UUACGCUGAGUACUUCGAU*U - 3' 3' - U*UGAAUGCGACUCAUGAAGC*U - 5'	AL-SEQ-gggg
		AL-SEQ-hhhh
AL-DUP-1363	5' - C*UUACGCUGAGUACUUCGAU*U - 3' 3' - U*UGAAUGCGACUCAUGAAGC*U - 5'	AL-SEQ-1162
		AL-SEQ-1163

phosphorothioate between the two terminal dT residues (see FIG. 15A). Addition of a single phosphorothioate between the two terminal 2'OMe-uridine residues effectively inhibited 3'-5' exonucleolytic cleavage as seen in FIG. 16 for duplexes AL-DUP-1036, AL-DUP-13ff, and AL-DUP-1363. 2'OMe substitution on its own was much more effective at protecting from endonucleolytic cleavage when placed at the internal cleavage sites. The parent duplex was cleaved 3' of U at two UpA sites within the duplex. Both strands are cleaved due to the symmetry of this dinucleotide repeat and mapping data was used to confirm the sites of cleavage (data not shown). Placement of 2'OMe at the strong endonucleolytic site ((FIG. 16, star in s/* as gel, AL-DUP-13ff) resulted in inhibition of cleavage at this site. The second, weaker endonucleolytic site (FIG. 16, black star in *s/as), however, was slightly enhanced when the strong site was protected with 2'OMe (FIG. 16, compare AL-DUP-13ff to AL-DUP-1036). Protection of both sites with 2'OMe (AL-DUP-1363) resulted in reduced endonucleolytic

cleavage at both sites (FIG. 16). The inhibitory effect of the 2'OMe substitution is consistent with the mechanism of endonucleolytic cleavage, which requires the 2'OH as a nucleophile in the cleavage reaction. 2'OMe modification will also be an effective means to protect the 3' overhang of single overhang siRNA duplexes where the 3' overhang is composed of

5 ribonucleotides. In this situation, 2'OMe substitution can be used to block the possible loss of the terminal two nucleotides by endonucleolytic cleavage and phosphorothioate can be used to protect from exonuclease degradation.

Cationic modifications enhanced nuclease resistance: The effect of three different cationic chemical modifications on nuclease resistance was tested and compared to the parent

10 unmodified duplex. The structures of the three cationic modifications tested are shown below.

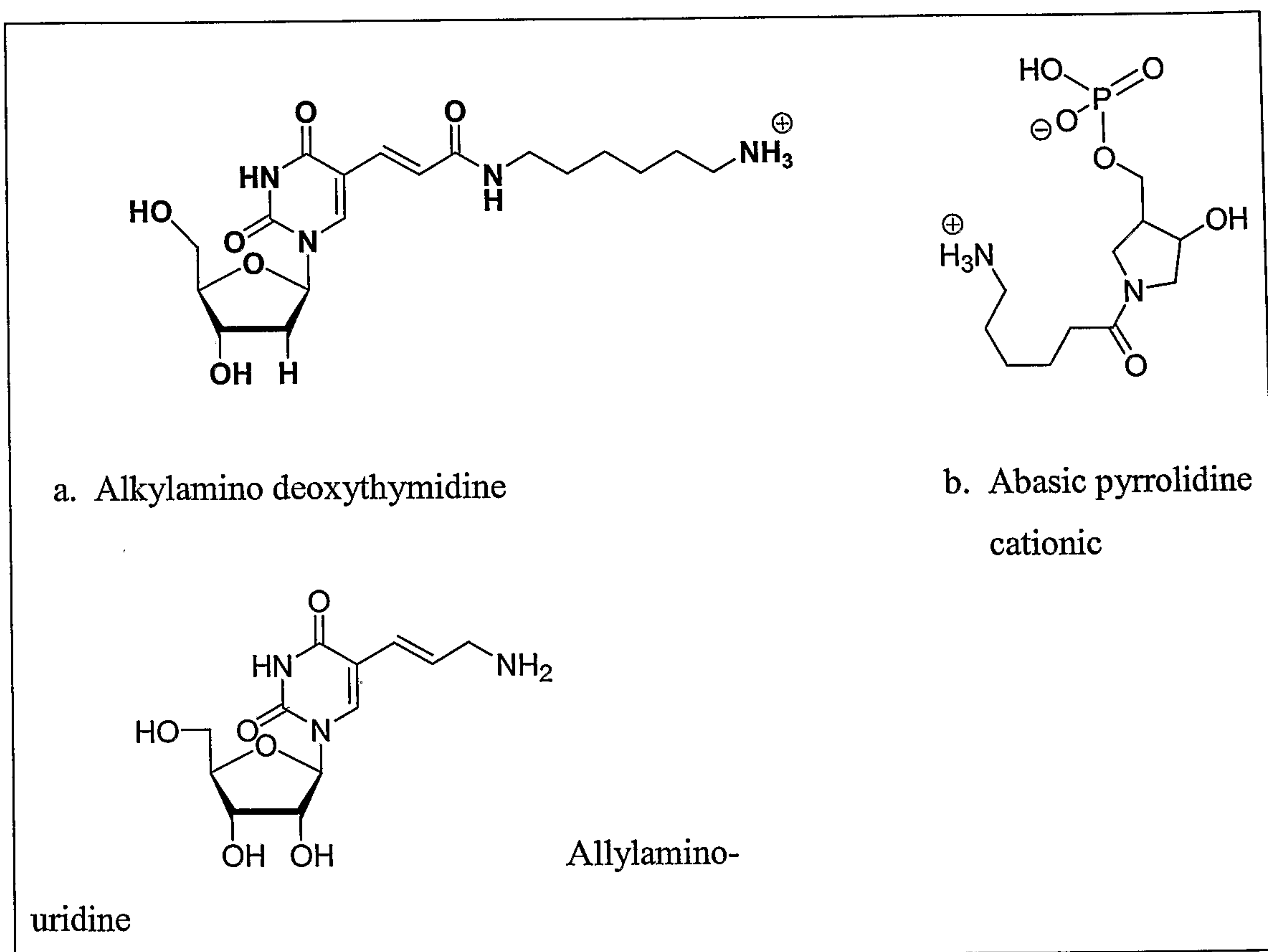


Table 3 siRNA duplexes containing cationic substitutions. (aadT = alkylamine-dT, AbP = abasic pyrrolidine cationic, aaU = allylamino-U, * = phosphorothioate, <u>U</u> = 2'OMe-U)		
Alnylam Duplex	Duplex Sequence	Alnylam Sequence
AL-DUP-10aa	5' - CUUACGCUGAGUACUUCGAdTaadT - 3' 3' - aadTdTGAAUGCGACUCAUGAAGCU - 5'	AL-SEQ-1017
		AL-SEQ-1018
AL-DUP-10bb	5' - CUUACGCUGAGUACUUCGAaadTaadT - 3' 3' - aadTaadTGAAUGCGACUCAUGAAGCU - 5'	AL-SEQ-1015
		AL-SEQ-1016
AL-DUP-1ccc	5' - CUUACGCUGAGUACUUCGAdTdTABP - 3' 3' - AbPdTdTGAAUGCGACUCAUGAAGCU - 5'	AL-SEQ-dddd
		AL-SEQ-eeee
AL-DUP-1403	5' - C*UaaUACGCUGAGUACUUCGA <u>U</u> * <u>U</u> - 3' 3' - <u>U</u> * <u>U</u> GAAaaUGCGACUCAUGAAGC*U - 5'	AL-SEQ-2080
		AL-SEQ-2081
AL-DUP-1406	5' - C*UaaUACGCUGAGaaUACUUCGA <u>U</u> * <u>U</u> - 3' 3' - <u>U</u> * <u>U</u> GAAaaUGCGACUCAaaUGAAGC*U - 5'	AL-SEQ-2082
		AL-SEQ-2083

The sequences of the duplexes assayed in the human serum stability assay are shown in Table 13. Both alkylamino-dT and abasic pyrrolidine cationic modifications were placed at the 3' terminal overhang to evaluate their effect on 3'-5' exonuclease degradation. Allylamino-uridines were placed at the internal endonucleolytic cleavage sites to evaluate their ability to inhibit endonucleolytic cleavage. As seen in FIG. 17, replacing the 3' terminal dT residue with a single alkylamino-dT efficiently inhibited 3'-5' exonucleolytic degradation (FIG. 17, AL-DUP-10aa, left gel image). Replacement of both dT residues in the overhang with alkylamino-dT resulted in a similar extent of inhibition (data not shown). Addition of an abasic pyrrolidine cationic modification at the 3' terminus of each strand also protected against exonucleolytic degradation (FIG. 17, middle gel image). Both the alkylamino-dT and abasic pyrrolidine modifications protected from 3'-5' exonucleolytic cleavage up to 23 hours (data not shown). Placement of allylamino-U at the internal cleavage sites inhibited endonucleolytic cleavage as shown in FIG. 17 for duplex AL-DUP-1403. The ends of this duplex were stabilized from exonucleolytic degradation by 2'OMe-U and phosphorothioate substitutions in order to separate the two different cleavage events. Endonucleolytic cleavage was inhibited at both internal cleavage sites by allylamino-U substitution for AL-DUP-1406 (data not shown).

3' conjugates enhanced nuclease resistance: Conjugation of naproxen and ibuprofen to the 3' end of the siRNA were tested for their ability to inhibit 3'-5' exonucleolytic degradation. The structure of naproxen is shown in below:

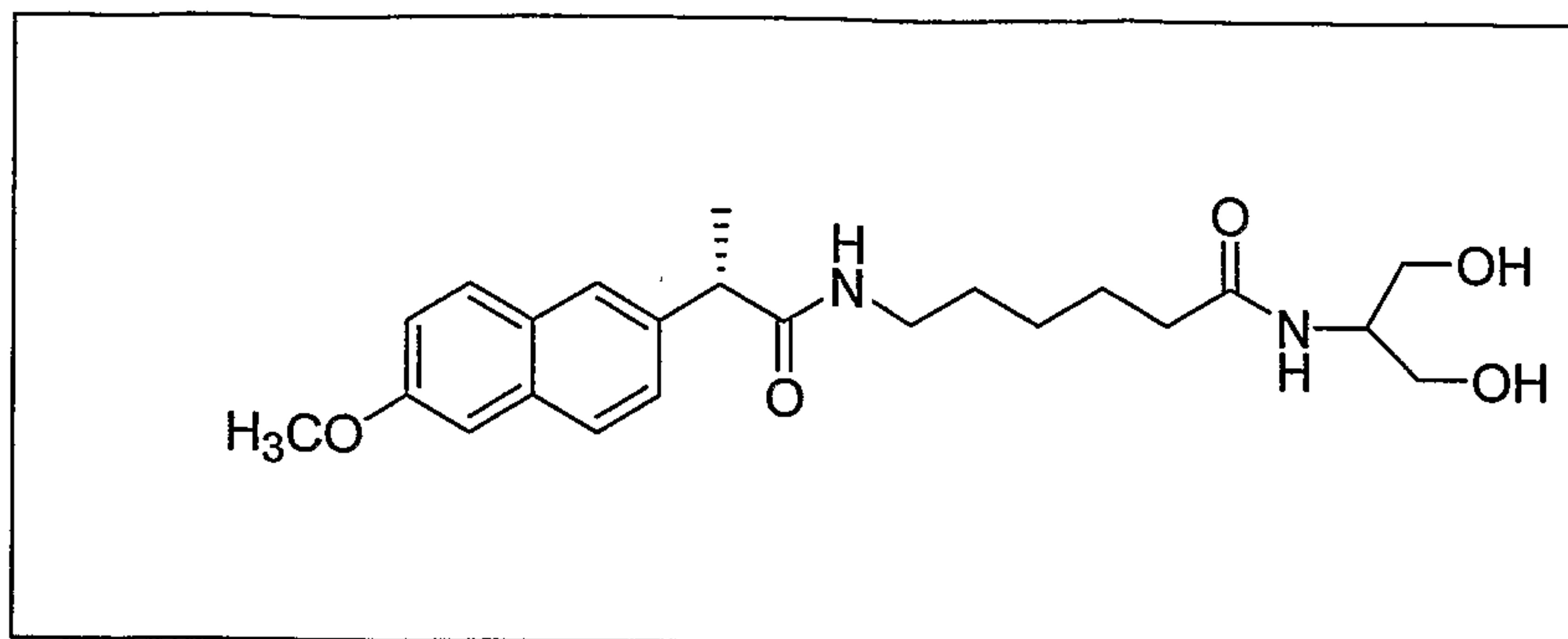


Table 14 lists the siRNAs that were tested in the human serum stability assay.

Conjugation of either naproxen or ibuprofen to the 3' end inhibited exonucleolytic degradation.

- 5 FIG. 18 shows the serum stability data for the naproxen modified duplex (AL-DUP-1069) and similar results were obtained for AL-DUP1413. Presumably the conjugates inhibit exonucleolytic cleavage by sterically blocking the exonuclease from binding to the 3' end of the siRNA duplex. Similar data was also obtained for AL-DUP-1069 in pooled mouse serum.

Table 4 siRNA duplexes containing 3' conjugates. (Nap = Naproxen, Ibu = Ibuprofen)		
Alnylam Duplex	Duplex Sequence	SEQ ID NO:
AL-DUP-1069	5' - CUUACGCUGAGUACUUCGAdTdTNap - 3' 3' - NapdTdTGA AUGCGACUCAUGAAGCU - 5'	
AL-DUP-1413	5' - CUUACGCUGAGUACUUCGAdTdTIBu - 3' 3' - NapdTdTGA AUGCGACUCAUGAAGCU - 5'	

Additional examples describing monomer synthesis and conjugation strategies are described in Appendix I, which is expressly included as part of the disclosure of this application.

EXAMPLE 16

Silencing of microRNAs with single stranded iRNA agents

Chemically-stabilized, cholesterol-conjugated single-stranded RNAs complementary to miRNAs were designed and synthesized. These single-stranded modified RNAs are referred to herein as "antagomirs" (see below). To explore the potential of these synthetic RNAs to silence endogenous miRNAs, antagomir-122 was designed to target miR-122, an miRNA expressed in

the liver. The sequence of antagomir-122 is shown in Table 15. Antagomir-122 was administered to mice by intravenous injection in a small volume (0.2 ml, 80 mg/kg, 3 consecutive days) and normal pressure. Administration of antagomir-122 resulted in a striking reduction of endogenous miR-122 levels as detected by Northern blot analysis (FIG.19A).

- 5 Administration of unmodified single-stranded RNA (anti-122) had no effect on hepatic miR-122 expression levels (FIG.19A), while injection of unconjugated, but chemically-stabilized single-stranded RNAs with partial (pS) or complete (fS) phosphorothioate backbone and 2'-O-methyl sugar modifications (anti-122fS, anti-122pS, see Table 15) led to an incomplete effect (Fig. 19A). The effects of antagomir-122 were found to be specific as animals injected with a control
- 10 antagomir-122 derivative that harbored four mismatch mutations (mm-antagomir-122) had no effect on miR-122 expression in the liver. Furthermore, expression levels of miR-let7 and miR-22 were unaffected in antagomir-122 and mm-antagomir-122 treated mice, suggesting that silencing was miRNA-specific (FIG. 19B). The structure of the single stranded RNAs injected into mice is described in Table 15.

15

Table 15. Antagomirs

RNA	Sequence	AL-SQ NO:
anti-122	5'-ACAAACACCAUUGUCACACUCCA-3'	3033
anti-122pS	5'-a _s c _s aaacaccauugucacac _s u _s c _s c _s a-3'	3226
anti-122fS	5'-a _s c _s a _s a _s a _s c _s a _s c _s c _s a _s u _s u _s g _s u _s c _s a _s c _s a _s c _s u _s c _s c _s a-3'	3037
antagomir-122	5'-a _s c _s aaacaccauugucacacu _s c _s c _s a _s -Chol-3'	3038
mm-antagomir-122	5'-a _s c _s acacaacacugucacauu _s c _s c _s a _s -Chol-3'	3040
antagomir-122(I)	5'-u _s g _s gagugugacaaugguguu _s u _s g _s u _s -Chol-3'	3223
antagomir-122(II)	5'-u _s g _s gaaggugacaguguuguu _s u _s g _s u _s -Chol-3'	3224
antagomir-122(III)	5'-u _s c _s acgcgagccgaacgaac _s a _s a _s -Chol-3'	3230
antagomir-16	5'-c _s g _s ccaauauuuacgugcug _s c _s u _s a _s -Chol-3'	3227
antagomir-192	5'-g _s g _s cugucaauuauaggu _s c _s a _s g _s -Chol-3'	3228
antagomir-194	5'-u _s c _s cacauggaguugcuguu _s a _s c _s a _s -Chol-3'	3229

lower case letters represent 2'-O-methyl modified nucleotides;
 subscript 's' represent phosphorothioate linkage;
 "Chol" indicates cholesterol conjugate

5

MiR-122 is expressed at high levels in hepatocytes with over 50,000 copies per cell (Chang J. *et al.*, *RNA Biology* 1:2, 106-113, 2004). To determine whether the silencing of miR-122 following antagomir treatment was caused by stoichiometric duplex formation between miR-122 and antagomir-122 or by catalytic degradation of miR-122, total RNA from livers of mice treated with unconjugated single-stranded anti-miR-122 RNAs (anti-122fS, anti-122pS) or antagomir-122 were examined under stringent, formamide-containing denaturing conditions (FIG. 19C). No difference in miR-122 levels could be detected between PBS and unconjugated anti-miR-122 RNA-treated livers, showing that the decrease in miR-122 levels observed under non-stringent conditions was not caused by degradation, but instead by the formation of miR-122/RNA duplexes. In contrast, miR-122 remained undetectable in livers of mice treated with antagomir-122. These data suggest that the silencing of miRNA-122 in livers of mice treated with antagomir-122 was due to degradation of the miRNA, and the ability of antagomir-122, but not unconjugated anti-122 RNAs, to result in miR-122 degradation may be due to efficient delivery of antagomirs to hepatocytes.

10

15

To determine the dose of antagomir-122 that can completely silence miR-122, mice were injected with 80, 160 or 240 mg/kg bodyweight antagomir-122 and miR-122 expression levels were measured. The highest dose (240 mg/kg bodyweight) resulted in a complete loss of miR-122 signal and was subsequently used for all other experiments (FIG. 20A).

5 The duration of silencing with antagomir-122 was also measured. Levels of miR-122 were undetectable for as long as 23 days post-injection (FIG. 20B), indicating that silencing of miRNAs using antagomirs is long lasting. The injected antagomirs were well tolerated even during the course of the prolonged treatment; no alterations in bodyweight or serum markers of liver toxicity (alanine aminotransferase) were detected. To test the bioavailability of antagomirs
10 *in vivo* and their ability to silence miRNA expression in different tissues, mice were injected with antagomir-16 directed to miR-16, which is abundantly expressed in all tissues (miR-16 is predicted to target one or both of *Activin type II receptor* gene, which is involved in TGFbeta signaling, and *Hox-A5I* (John *et al.*, *PLoS Biology* 2:1862-1878, 2004; correction in *PLoS Biology* 3:1328, 2005)). Tissues were harvested one day after the final injection, and miRNA
15 expression levels were compared to PBS-injected mice. Northern blot analysis revealed that expression of miR-16 was efficiently silenced in all tissues tested except brain (FIG. 21A). Antagomir-16 did not affect the expression of the 89 nt precursor of miR-16 as detected in bone marrow. The bioavailability of antagomir-16 was also assessed by Northern blotting in the above mentioned tissue samples. In concordance with the ability to silence miR-16 levels, significant
20 levels of antagomir-16 were detected in all tissues except brain (FIG. 21B). Together, these data demonstrate that antagomirs achieve broad biodistribution and can efficiently silence miRNAs in most tissues *in vivo*.

Many miRNA genes have been found to be located in close proximity and to be coordinately transcribed. These polycistronic miRNA genes are transcribed to generate long
25 primary transcripts (pri-miRNAs), which are processed by multiple enzymes in the nucleus and cytoplasm to generate the mature miRNA. To investigate if antagomirs targeting polycistronic miRNAs retain their target specificity with no effect on the expression of neighboring miRNAs, mice were injected with antagomirs targeting either miR-192 or miR-194 of the bicistronic cluster miR-192/194. Administration of antagomir-192 into mice resulted in silencing of miR-
30 192 in liver and kidney, with no effect on the expression levels of miR-194. Conversely, injection of antagomir-194 into mice abolished miR-194 expression but had no demonstrable

effect on the miR-192 levels compared to PBS-injected mice. These data demonstrate that antagomirs have the ability to differentially silence specific miRNAs that derive from the same primary transcript.

MicroRNAs are believed to bind to imperfect complementary target sites in 3'UTRs of mRNAs and interfere with translation and possibly mRNA stability. Therefore, silencing of miRNAs would be expected to result in a corresponding increase in target protein and possibly mRNA levels. To test this prediction, the expression of aldolase A, a gene that is repressed in hepatocytes and predicted to be the target of miR-122, was examined. The aldolase-A mRNA has a conserved nucleus with perfect sequence complementarity to miR-122 between nucleotides 29 and 36 downstream of the open reading frame. Aldolase-A expression was increased 4-5 fold in livers of mice injected with antagomir-122 compared to scrambled control (mm- antagomir-122). This regulation was observed in multiple experiments and different time points after injection. The target was also independently confirmed by cloning the 3'UTR Aldolase-A downstream of the luciferase open reading frame and cotransfecting this vector with control miRNAs (miR-124 (5'-UAAGGCACGCGGUGAAUGCCA-3; see Krek *et al.*, *Nature Genetics* 37:495-500, 2005, and Lim *et al.*, *Nature* 433:769-773, 2005) and miR-192) and miR-122 into HEK293 cells, which lack miR-122 expression. Cotransfection of miR-122 resulted in a significant reduction in luciferase activity compared to miR-124 and miR-192 transfected cells. Together, these data indicate that aldolase-A is a physiological target of miR-122.

The upregulation of aldolase-A in mice treated with antagomir-122 demonstrates functional silencing of this miRNA. Aldolase-A is a housekeeping gene expressed in all cells. This gene is produced in large amounts in muscle where it can be as much as 5% of total cellular protein. In adult liver, aldolase-A expression is repressed and aldolase-B is produced. Conversely, dedifferentiated hepatocytes and transformed liver cells have increased aldolase-A expression levels and can even replace aldolase-B. Expression of miR-122 shows an inverse relationship with aldolase-A expression, with highest levels in differentiated adult hepatocytes and complete absence in undifferentiated cells such as HepG2. In contrast, the mRNA levels of aldolase-B, which lacks miR-122 target sites, was unaffected by antagomir-122. These findings provide non-genetic, pharmacologic evidence in mammals that microRNAs define tissue-specific gene expression.

The data demonstrate that antagomirs, single-stranded cholesterol-conjugated RNAs complementary to specific miRNAs, are effective inhibitors of miRNAs *in vivo*. The results also suggest that this process is highly specific and likely to occur in the cytosol since it does not affect the miRNA precursor or non-targeted miRNAs of common polycistronic precursors.

5 Methods

Synthesis of antagomirs RNAs were synthesized using commercially available 5'-O-(4,4'-dimethoxytrityl)-2'-O-methyl-3'-O-(2-cyanoethyl-N,N-diisopropyl) RNA phosphoramidite monomers of 6-N-benzoyladenosine (ABz), 4-N-benzoylcytidine (CBz), 2-N-isobutyrylguanosine (GiBu), and uridine (U), according to standard solid phase oligonucleotide synthesis protocols (Damha and Ogilvie, *Methods Mol. Biol.* 20:81-114, 1993). For antagomirs, i.e., cholesterol conjugated RNAs, the synthesis started from a controlled-pore glass solid support carrying a cholesterol- hydroxyprolinol linker (Manoharan *et al.*, U.S. Pat. Appl. Publ. 20050107325). Antagomirs with phosphorothioate backbone at a given position were achieved by oxidation of phosphite with phenylacetyl disulfide (PADS) during oligonucleotide synthesis (Cheruvallath *et al.*, *Nucleosides Nucleotides* 18:485-492, 1999). After cleavage and de-
10 protection, antagomirs were purified by reverse-phase high-performance liquid chromatography, while the unconjugated RNA oligonucleotides were purified by anion-exchange high-performance liquid chromatography. Purified oligonucleotides were characterized by ES mass spectrometry and capillary gel electrophoresis.

20 *Animals.* All animal models were maintained in C57Bl/6J background on a 12 hours light/dark cycle in a pathogen-free animal facility at Rockefeller University. Six week old mice received, on one to three consecutive days, tail vein injections of saline or different RNAs. RNAs were administered at doses of 80 mg/kg body weight in 0.2 ml per injection. Measurements of miRNA levels in tissues were performed 24 h after the last injection unless
25 indicated otherwise. Tissues were harvested, snap frozen and stored at -80°C.

Northern blotting analysis. Total RNA was isolated using the Trizol reagent (Invitrogen, Carlsbad, CA) and ethanol precipitation. RNA was separated at 45 mA on 14%-polyacrylamide gels that contained 8 M urea and 20% formamide. Antisense probes were designed according to the "microRNA registry" (Griffiths-Jones, *NAR* 32:D109-D111, 2004).

30 *RT-PCR.* Extraction of total RNA, synthesis of cDNA, and PCR were carried out as described in Shih *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 99:3818-3823, 2002.

Assay of luciferase activity. The mouse full length adolase-A 3'UTR was PCR-amplified using the following primers: 5' d-(CCAGAGCTGAACTAAGGCTGCTCCA)-3' and 5' d-(CCCCTTAAATAGTTGTTTAT TGGCA)-3' and cloned downstream of the stop codon in pRL-TK (Promega). HEK293 cells were cultured in 24-well plates and each transfected with 50
5 ng of pRL-TK (Rr-luc), 50 ng of pGL3 control vector (Pp-luc) (Promega) and 200 ng of double-stranded siRNA (Dharmacon). Cells were harvested and assayed 24-30 h post-transfection.

Statistical analysis. Results are given as mean $\pm$ s.d. Statistical analyses were performed by using Student's t-test, and the null hypothesis was rejected at the 0.05 level.

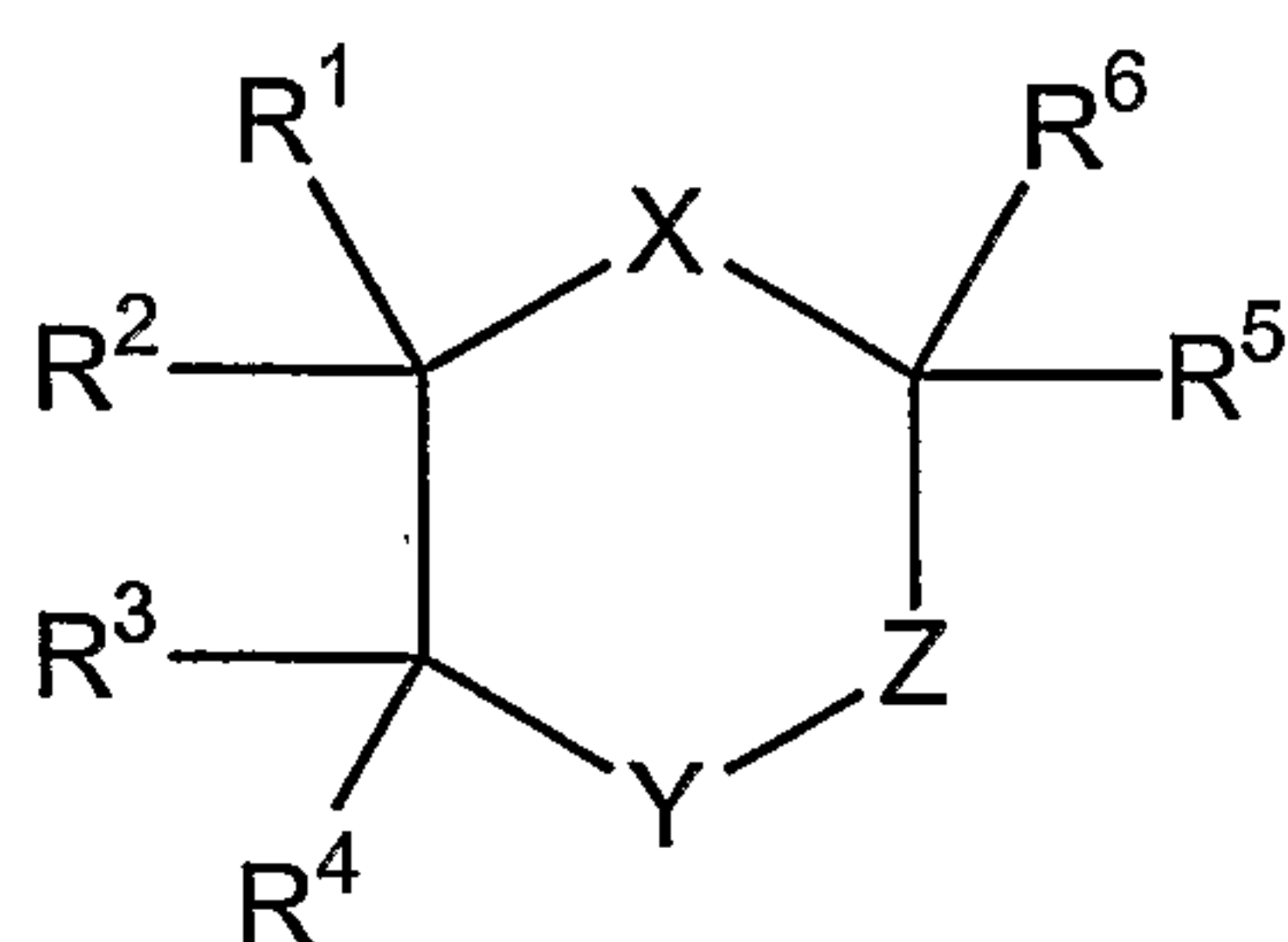
10

OTHER EMBODIMENTS

A number of embodiments of the invention have been described. Nevertheless, it will be understood that various modifications may be made without departing from the spirit and scope of the invention. Accordingly, other embodiments are within the scope of the following claims.

WHAT IS CLAIMED IS:

1. A conjugate comprising an oligonucleotide agent and a ligand, wherein at least one subunit having a formula (I) is incorporated into said oligonucleotide agent:



5

(I)

wherein:

X is N(CO)R⁷, NR⁷ or CH₂;

10

Y is NR⁸, O, S, CR⁹R¹⁰, or absent;

Z is CR¹¹R¹² or absent;

Each of R¹, R², R³, R⁴, R⁹, and R¹⁰ is, independently, H, OR^a, OR^b, (CH₂)_nOR^a, or (CH₂)_nOR^b, provided that at least one of R¹, R², R³, R⁴, R⁹, and R¹⁰ is OR^a or OR^b and that at least one of R¹, R², R³, R⁴, R⁹, and R¹⁰ is (CH₂)_nOR^a, or (CH₂)_nOR^b;

15

Each of R⁵, R⁶, R¹¹, and R¹² is, independently, H, C₁-C₆ alkyl optionally substituted with 1-3 R¹³, or C(O)NHR⁷; or R⁵ and R¹¹ together are C₃-C₈ cycloalkyl optionally substituted with R¹⁴;

R⁷ is R^d; or C₁-C₂₀ alkyl substituted with NR^cR^d or NHC(O)R^d;

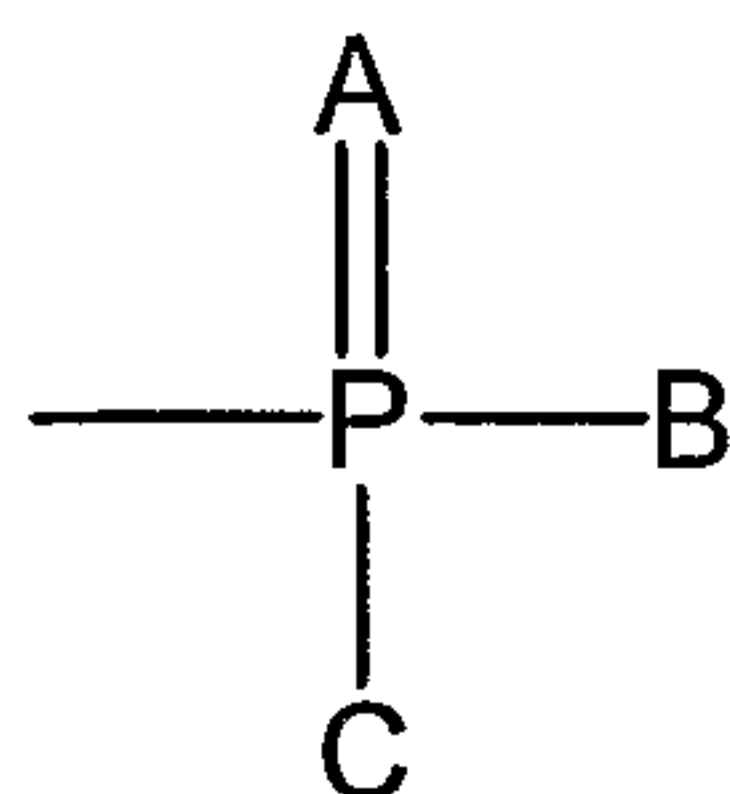
R⁸ is C₁-C₆ alkyl;

20

R¹³ is hydroxy, C₁-C₄ alkoxy, or halo;

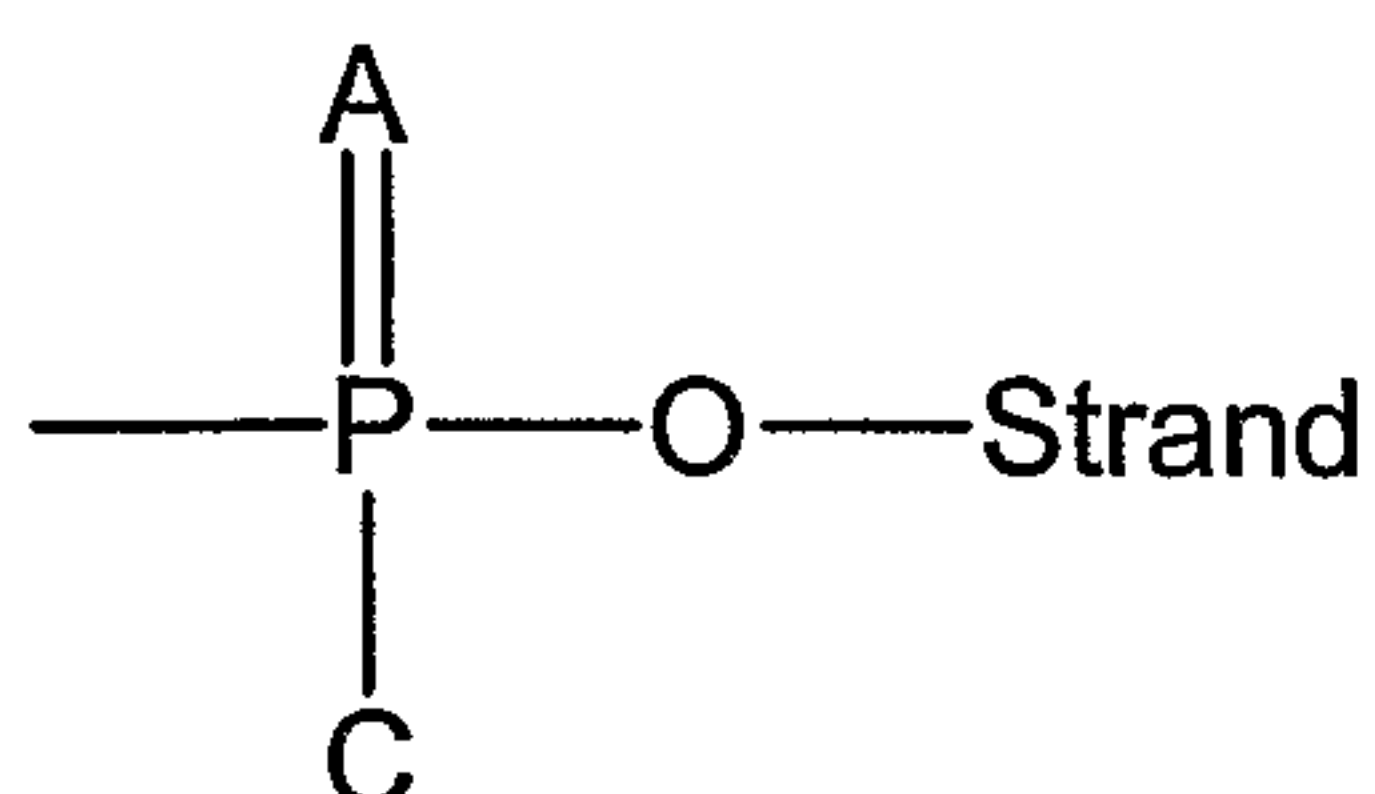
R¹⁴ is NR^cR⁷;

R^a is:



;

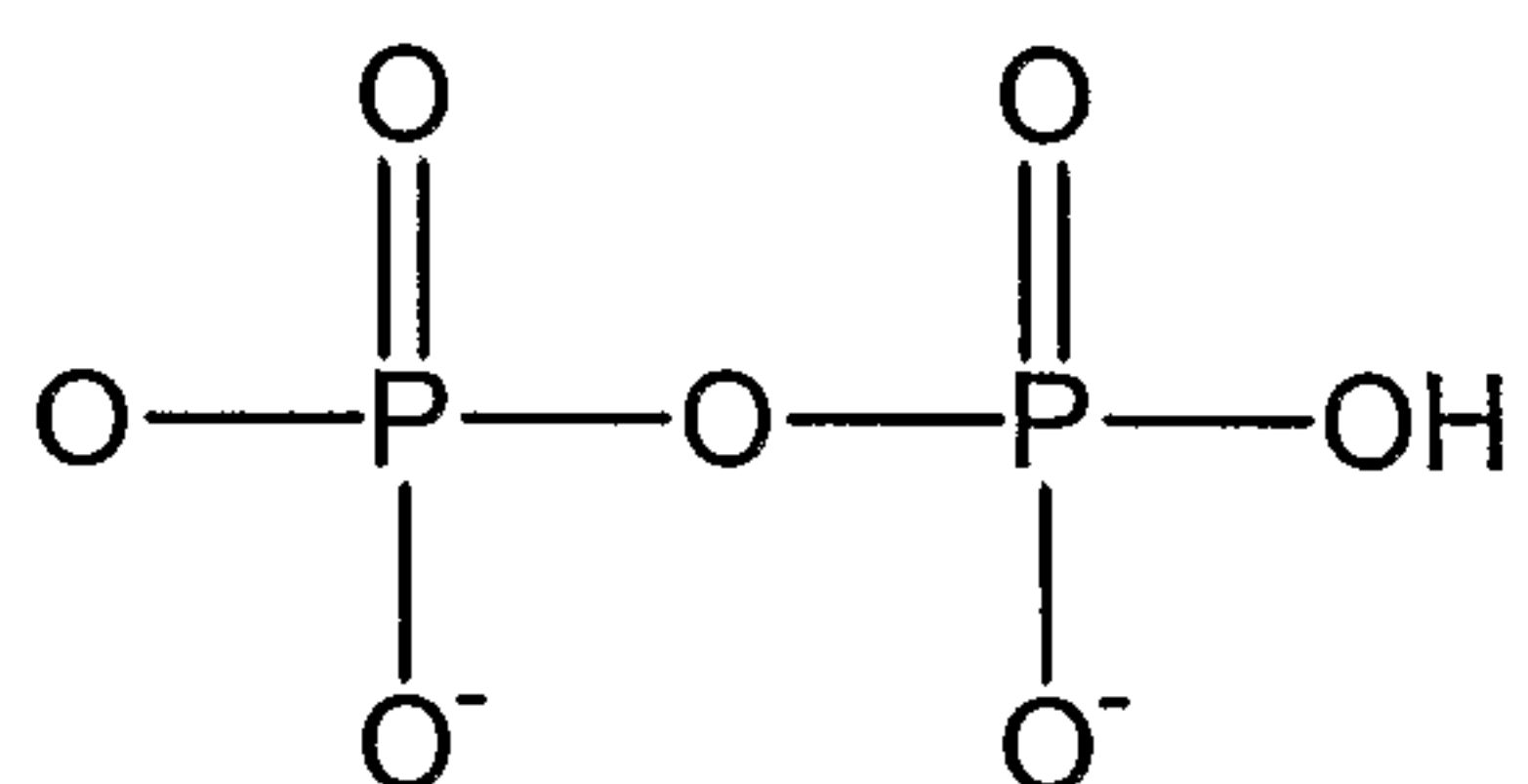
R^b is



;

Each of A and C is, independently, O or S;

B is OH, O^- , or



R^c is H or C_1 - C_6 alkyl;

R^d is H or a ligand; and

n is 1-4.

2. The conjugate of claim 1, wherein X is $N(CO)R^7$ or NR^7 , Y is CR^9R^{10} , and Z is absent.

3. The conjugate of claim 2, wherein R^7 is $(CH_2)_5NHR^d$ or $(CH_2)_5NHC(O)R^d$.

4. The conjugate of claim 3, wherein R^d is chosen from the group consisting of a folic acid radical; a cholesterol radical; a carbohydrate radical; a vitamin A radical; a vitamin E radical; a vitamin K radical; and a vitamin B12 radical.

5. The conjugate of claim 4, wherein R^d is a cholesterol radical.

6. The conjugate of claim 2, wherein R^1 is $(CH_2)_nOR^b$ and R^9 is OR^a .

7. The conjugate of claim 6, wherein R^1 and R^9 are *trans*.

8. The conjugate of claim 6, wherein n is 1.

9. The conjugate of claim 2, wherein R^1 is $(CH_2)_nOR^b$ and R^9 is OR^b .

10. The conjugate of claim 2, wherein R^1 is $(CH_2)_nOR^a$ and R^9 is OR^b .

11. The conjugate of claim 2, wherein R^3 is $(CH_2)_nOR^b$ and R^9 is OR^a .

12. The conjugate of claim 2, wherein R^3 is $(CH_2)_nOR^b$ and R^9 is OR^b .

13. The conjugate of claim 2, wherein R^3 is $(CH_2)_nOR^a$ and R^9 is OR^b .

14. The conjugate of claim 2, wherein R^3 is OR^a and R^9 is $(CH_2)_nOR^b$.

15. The conjugate of claim 2, wherein R^3 is OR^b and R^9 is $(CH_2)_nOR^b$.

16. The conjugate of claim 2, wherein R^3 is OR^b and R^9 is $(CH_2)_nOR^a$.

17. The conjugate of claim 1, wherein X is $\text{N}(\text{CO})\text{R}^7$ or NR^7 , Y is CR^9R^{10} , and Z is $\text{CR}^{11}\text{R}^{12}$.

18. The conjugate of claim 1, wherein X is $\text{N}(\text{CO})\text{R}^7$ or NR^7 , Y is NR^8 , and Z is $\text{CR}^{11}\text{R}^{12}$.

19. The conjugate of claim 1, wherein X is $\text{N}(\text{CO})\text{R}^7$ or NR^7 , Y is O, and Z is $\text{CR}^{11}\text{R}^{12}$.

20. The conjugate of claim 1, wherein X is CH_2 ; Y is CR^9R^{10} ; Z is $\text{CR}^{11}\text{R}^{12}$, and R^5 and R^{11} together form C_6 cycloalkyl.

21. The conjugate of claim 1, wherein X is CH_2 ; Y is CR^9R^{10} ; Z is $\text{CR}^{11}\text{R}^{12}$, and R^5 and R^{11} together form C_5 cycloalkyl.

22. The conjugate of claim 1, wherein the ligand comprises a lipophilic moiety chosen from the group consisting of a lipid, cholesterol, oleyl, retinyl, cholesteryl residues, cholic acid, adamantane acetic acid, 1-pyrene butyric acid, dihydrotestosterone, 1,3-Bis-O(hexadecyl)glycerol, geranyloxyhexyl group, hexadecylglycerol, borneol, menthol, 1,3-propanediol, heptadecyl group, palmitic acid, myristic acid, O3-(oleoyl)lithocholic acid, O3-(oleoyl)cholenic acid, dimethoxytrityl, or phenoxazine.

23. The conjugate of claim 1, wherein the ligand is attached to a 3' terminal subunit.

24. The conjugate of claim 1, wherein the ligand is attached to a 5' terminal subunit.

25. The conjugate of claim 1, wherein the ligand is attached an internal subunit.

26. A method of inhibiting a target gene comprising providing a conjugate of claim 1 to a cell.

27. The method of claim 26, wherein said conjugate is administered to an organism.

28. The method of claim 26, wherein said cell is outside an organism.

5 29. The method of claim 26, wherein said cell is a cell of a cell line.

30. A method of modulating expression of a target gene in a subject, the method comprising administering an conjugate of claim 1 to a subject.

10 31. A pharmaceutical composition comprising an iRNA agent of claims 1 and a pharmaceutically acceptable carrier.

32. A kit comprising an conjugate of claim 1, a sterile container in which the conjugate is enclosed, and instructions for use.

15 33. A monomer suitable for synthesizing an oligonucleotide agent of a conjugate of claim 1.

34. A method of using a conjugate of claim 1 to treat a disorder of the kidney.

20 35. A method of using a conjugate of claim 1 to treat an ocular disorder.

36. A method of using a conjugate of claim 1 to treat a viral disorder.

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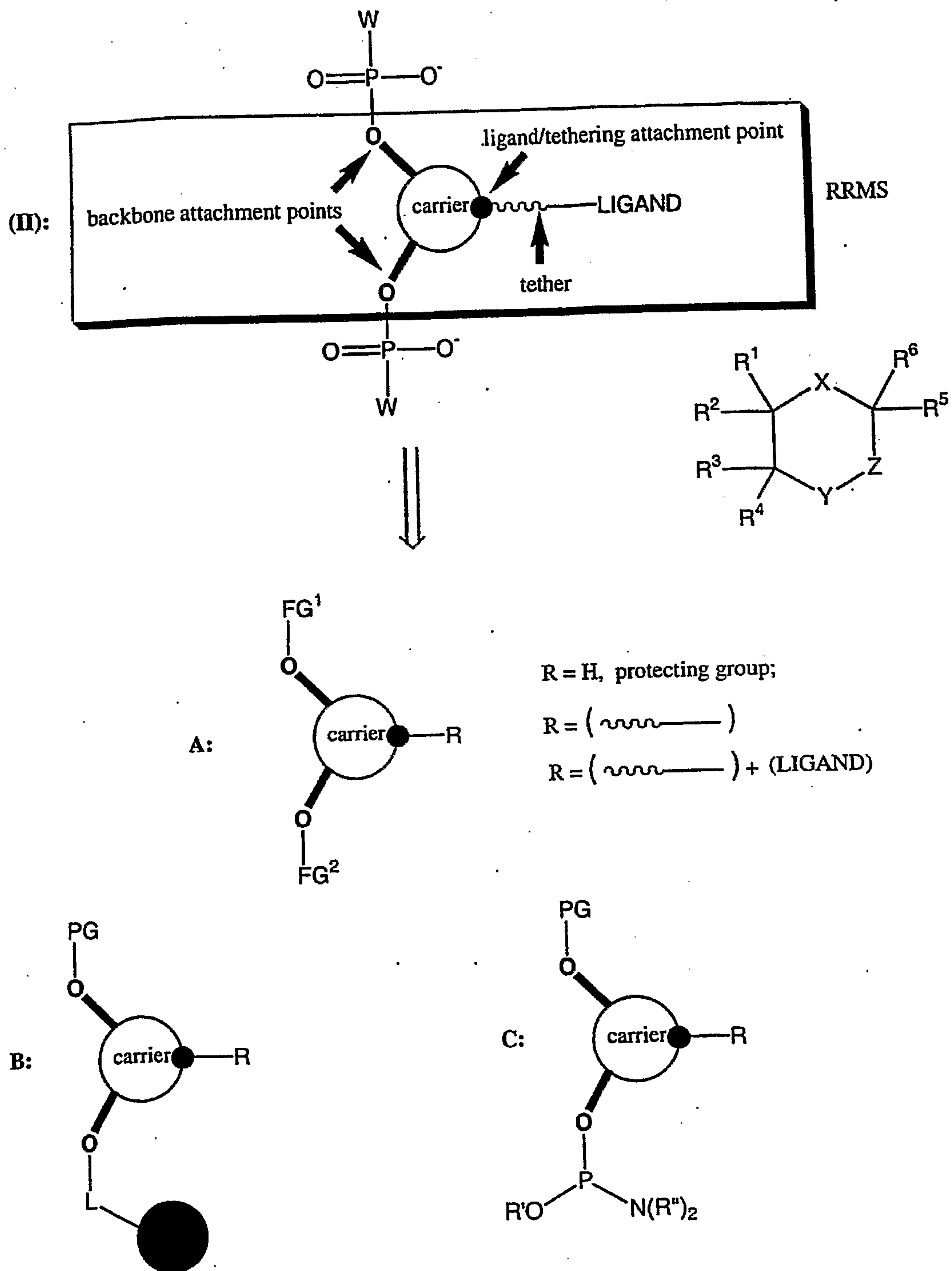


FIG. 1

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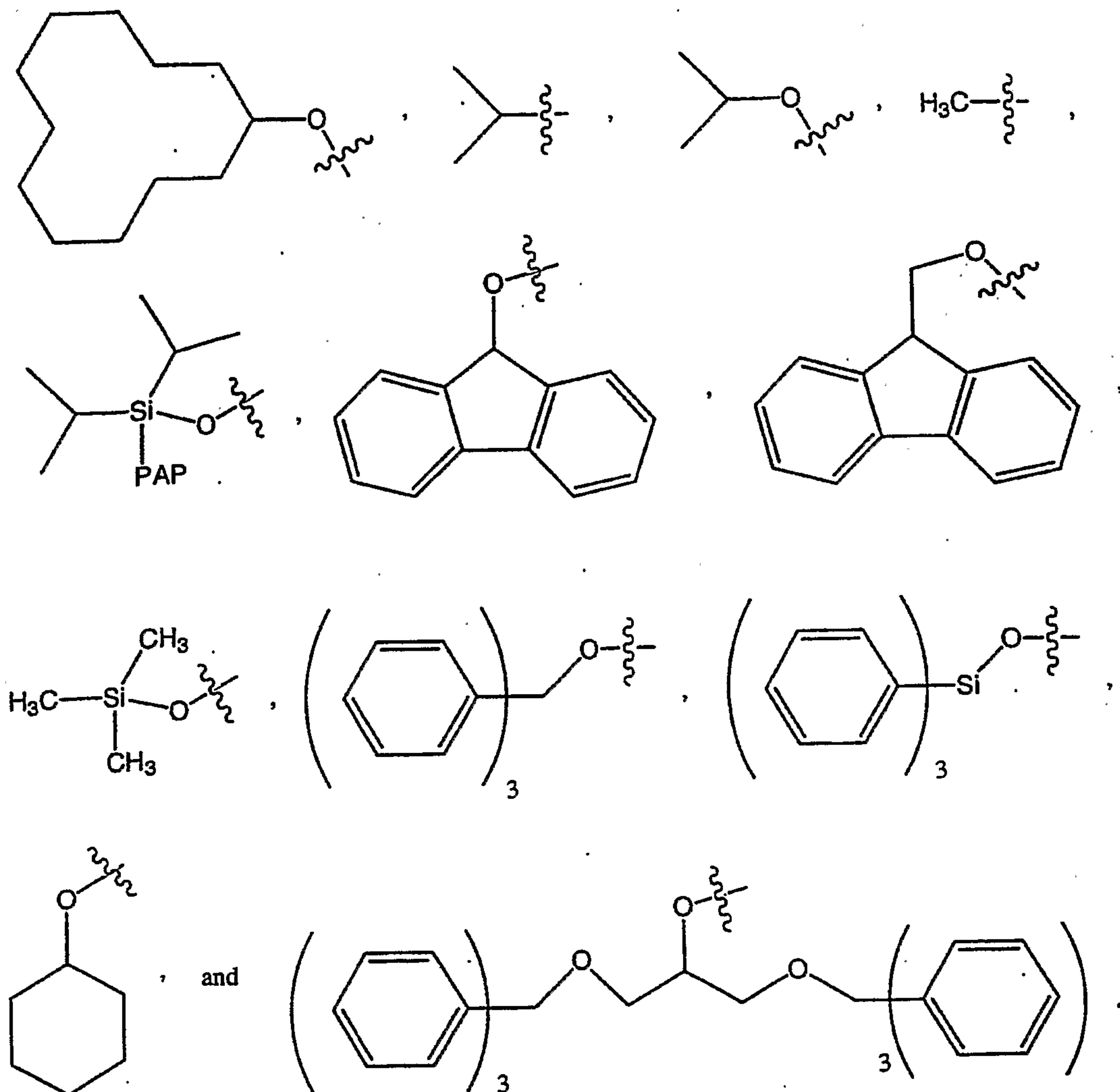


Fig. 2A

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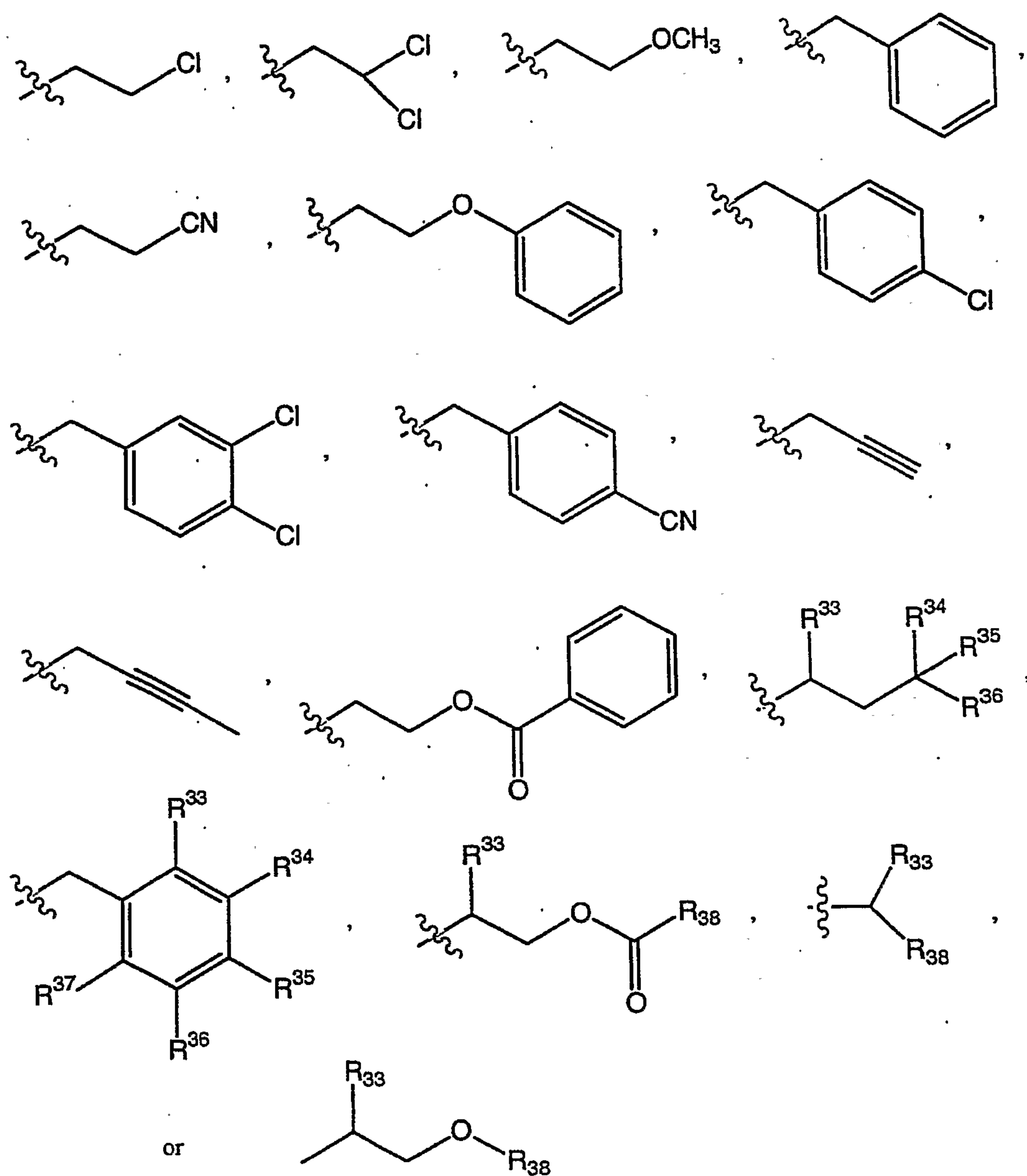


Fig. 2B

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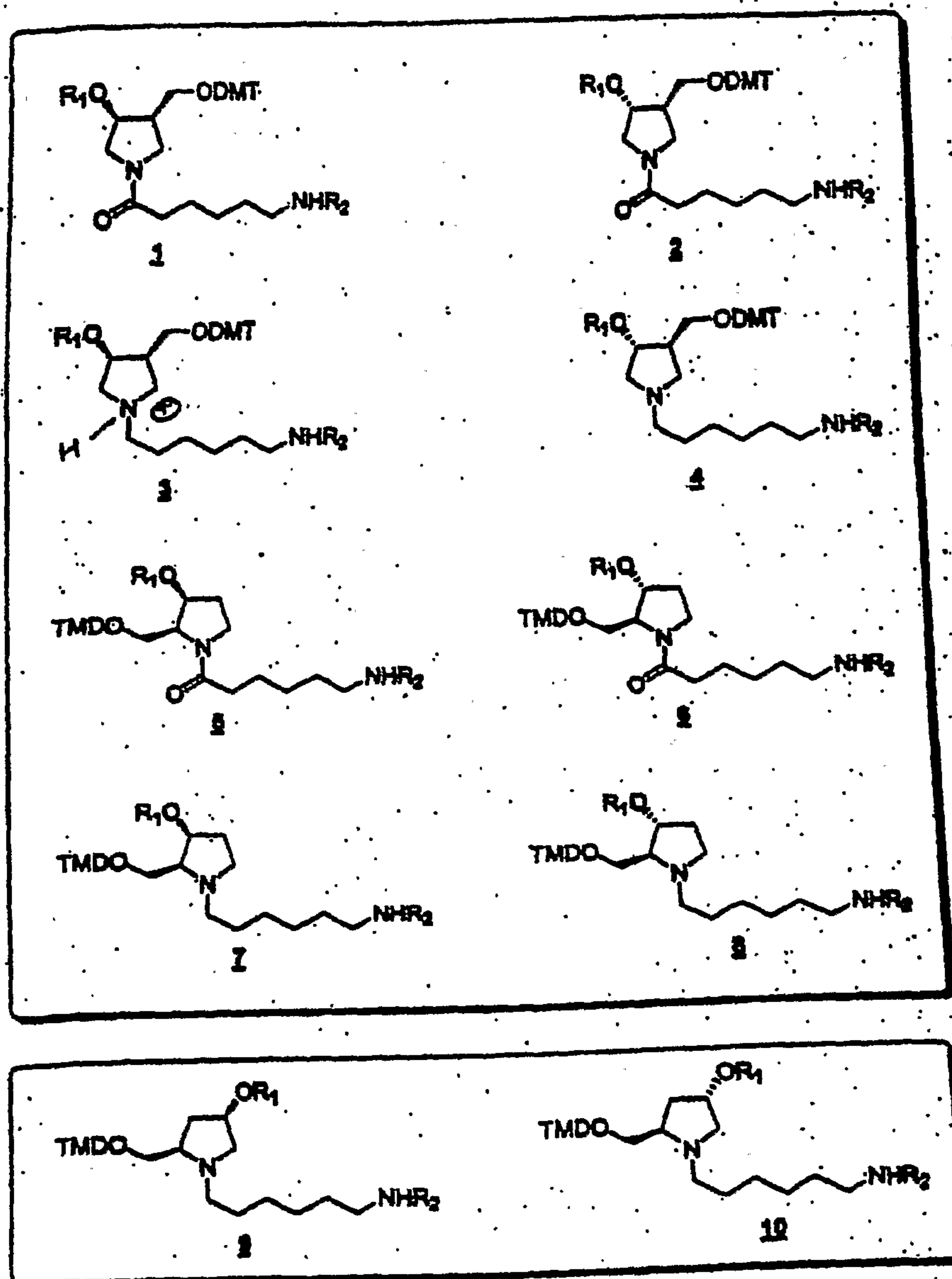


FIG. 3

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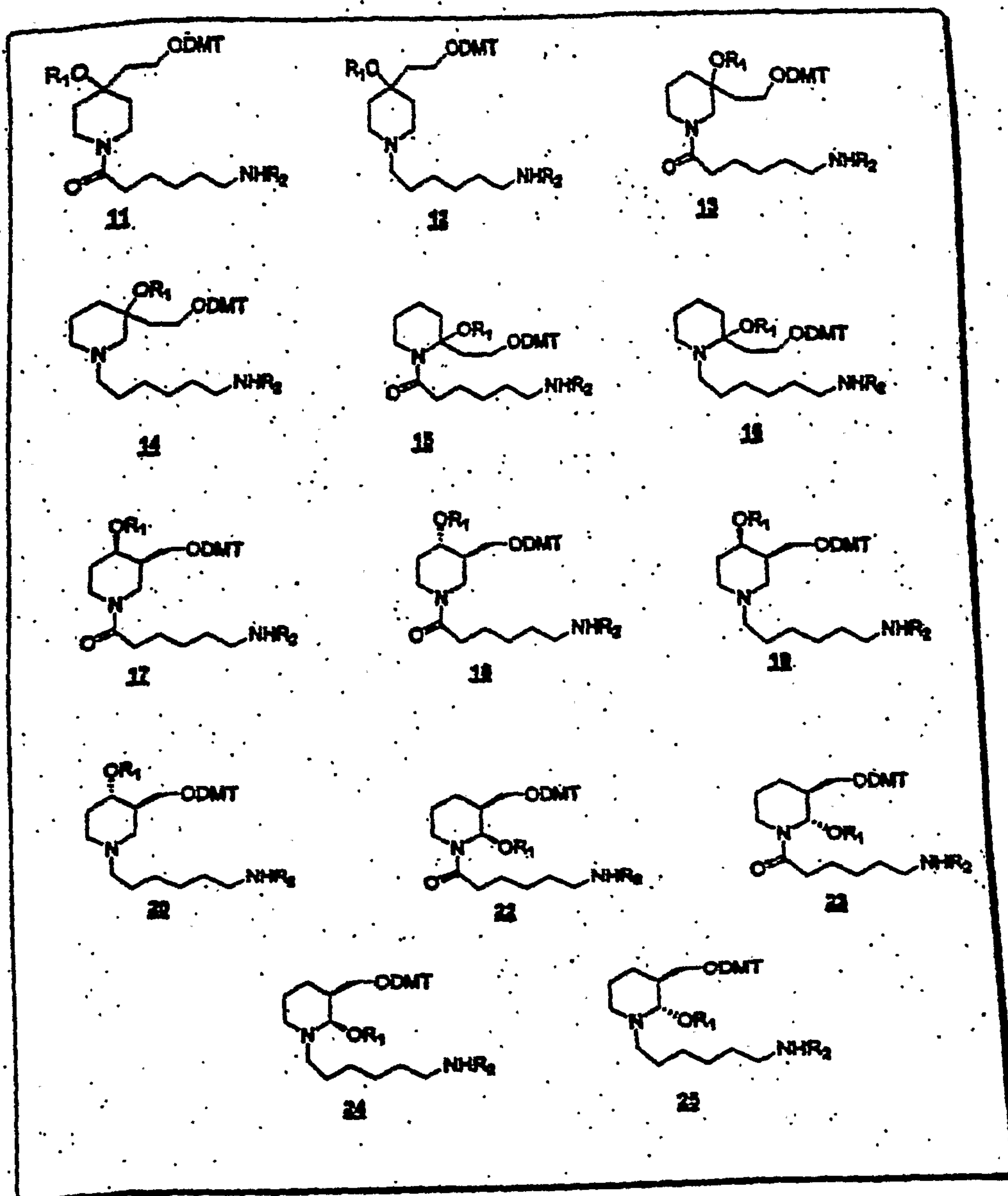


FIG. 3 (Cont'd)

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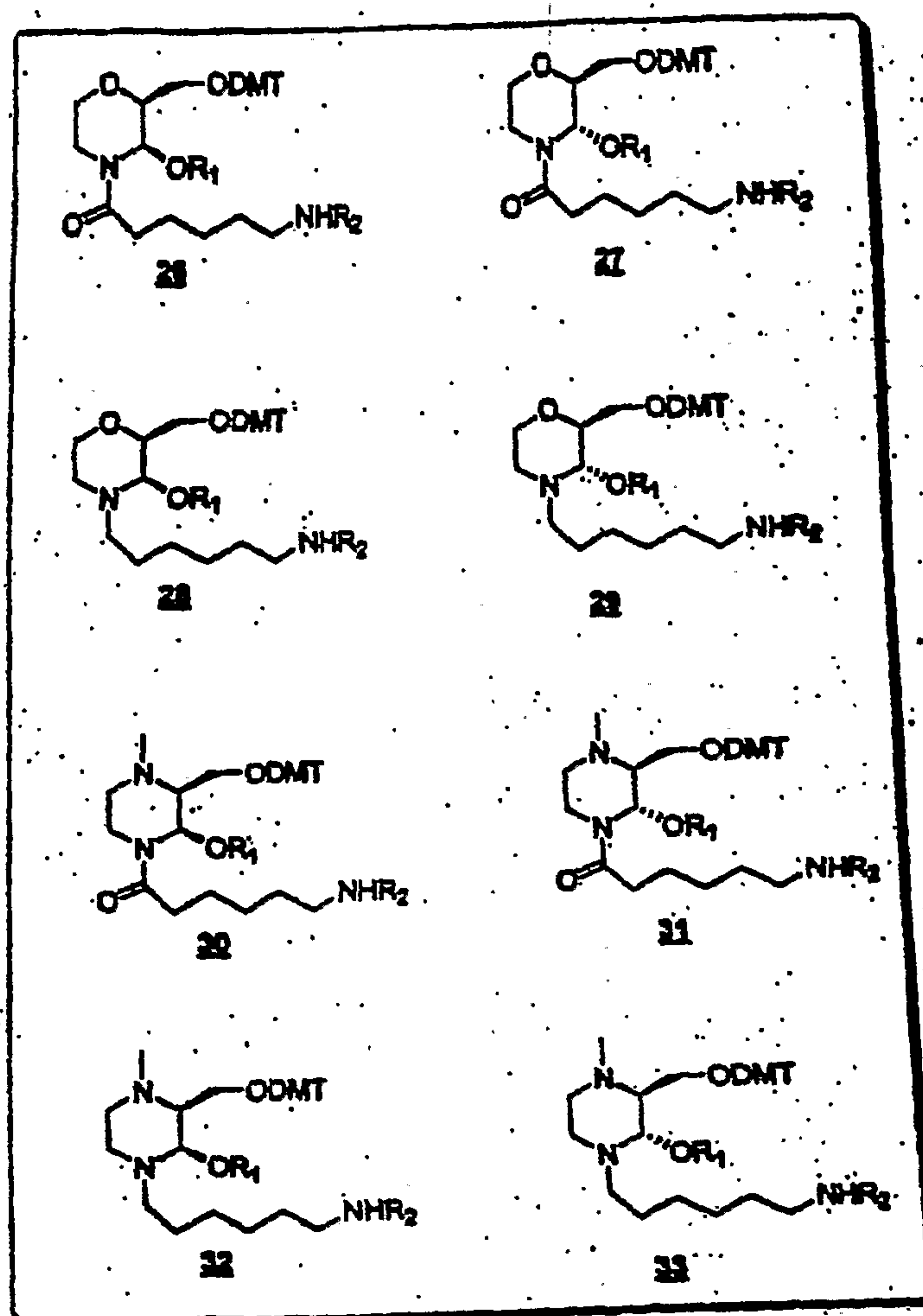


FIG. 3 (Cont'd)

7/25

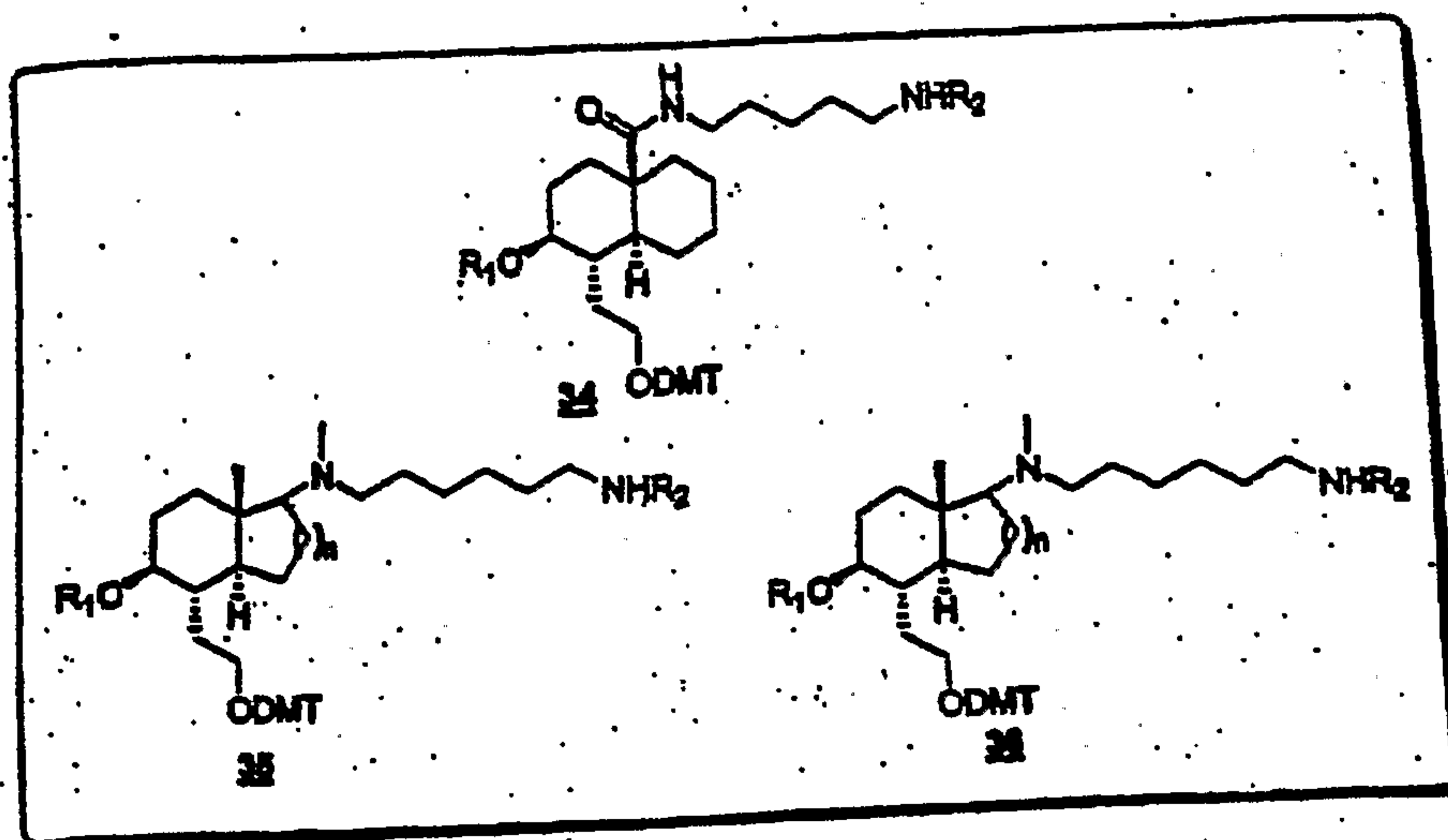
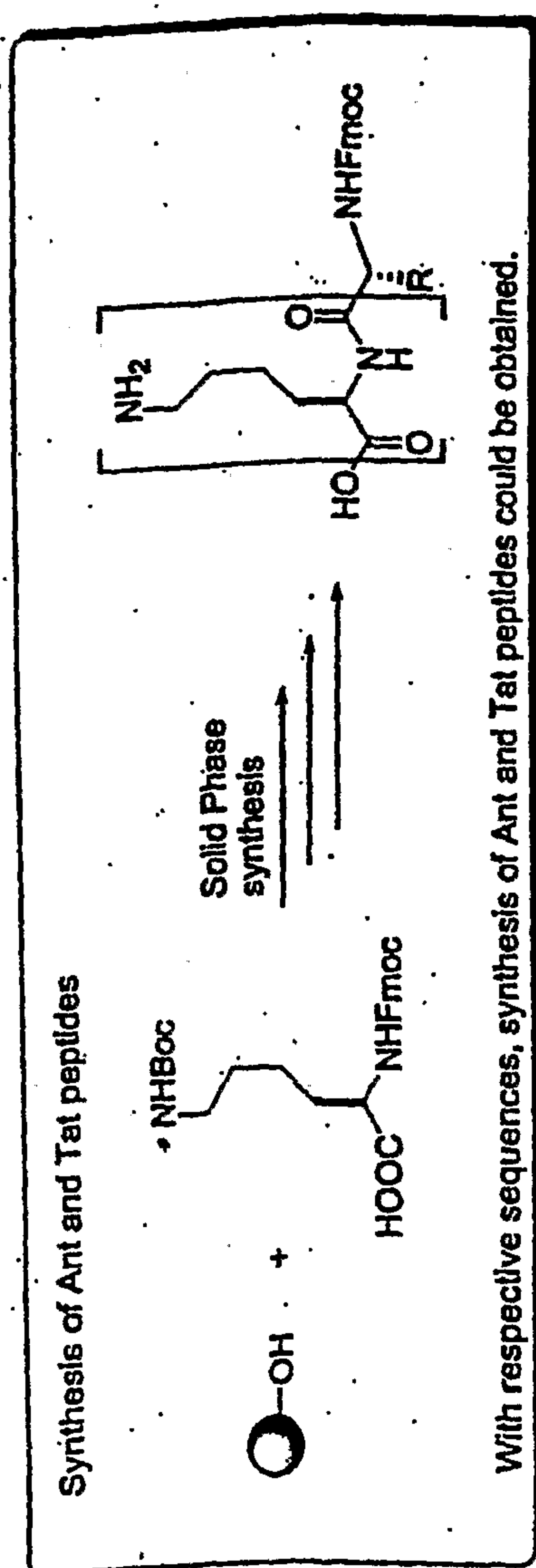


FIG. 3 (Cont'd)

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3' conjugation of peptide into siRNA

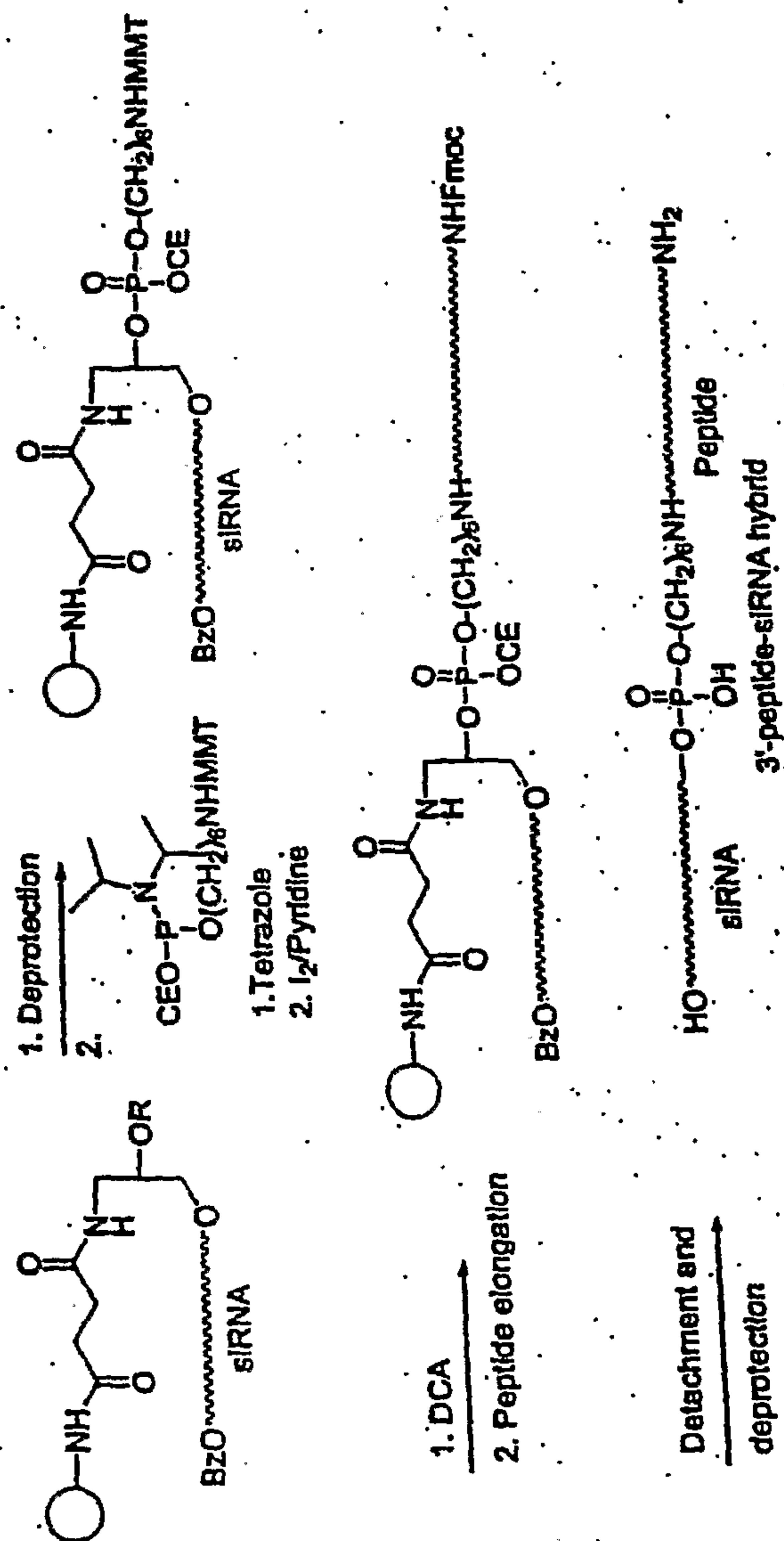
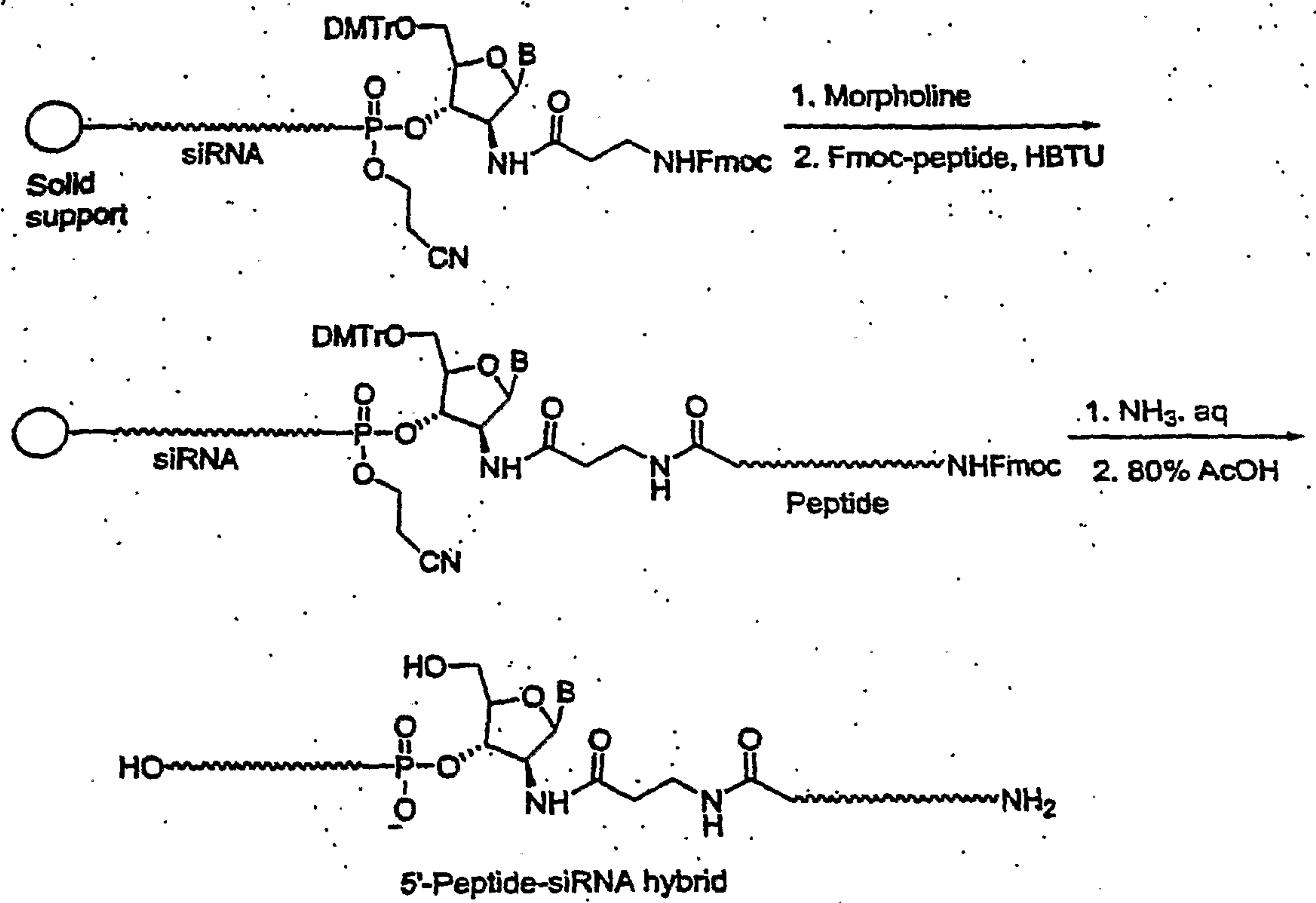


FIG. 4

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5' conjugation of peptide into siRNA**FIG. 5**

10/25

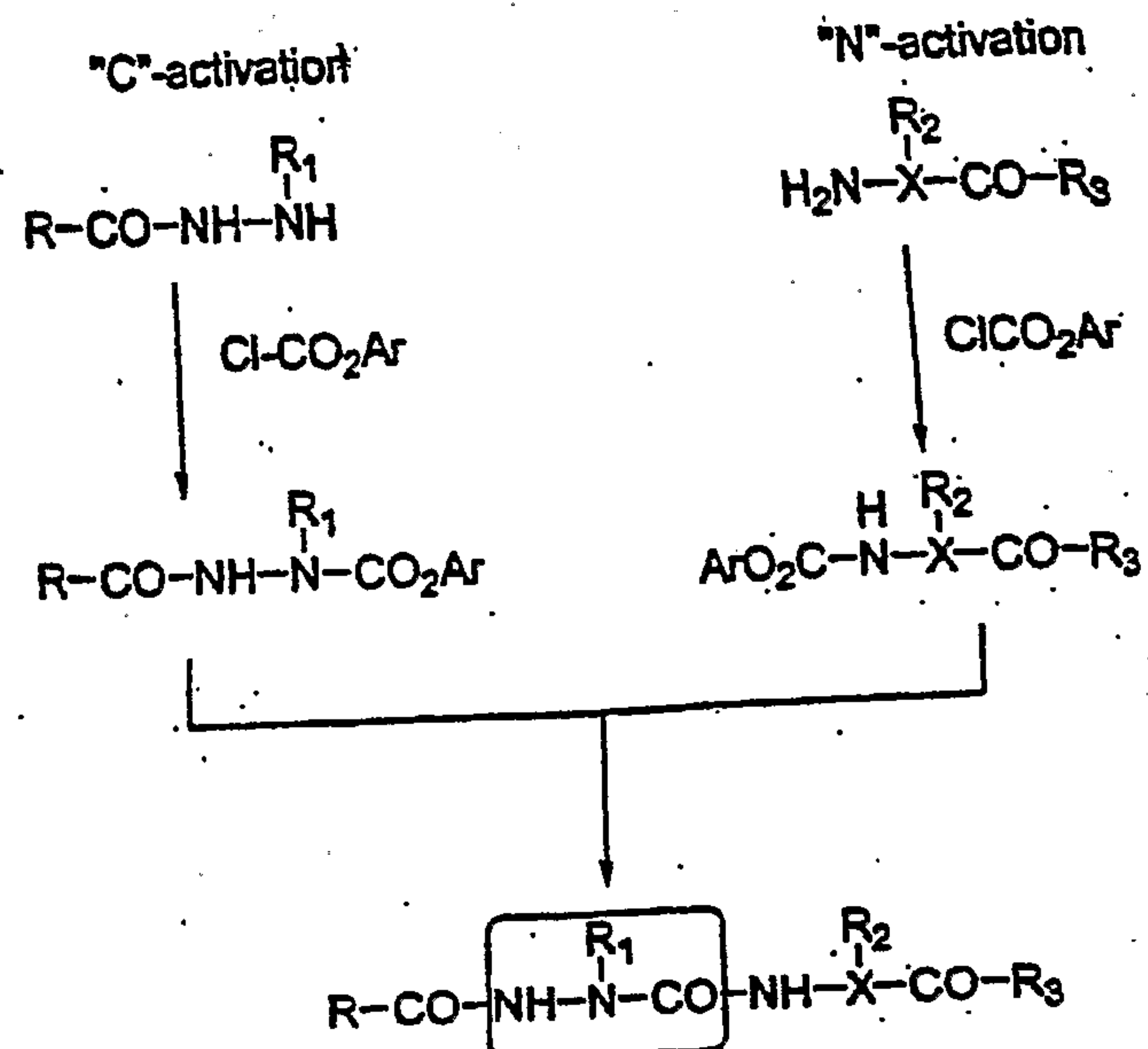


FIG. 6

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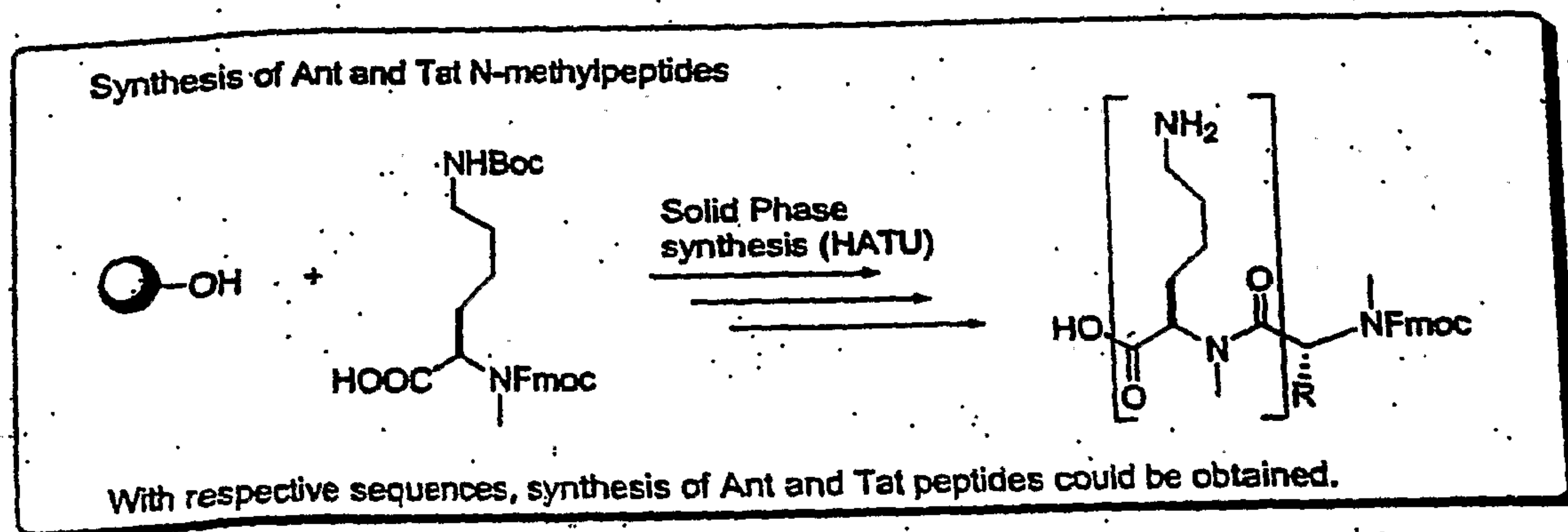
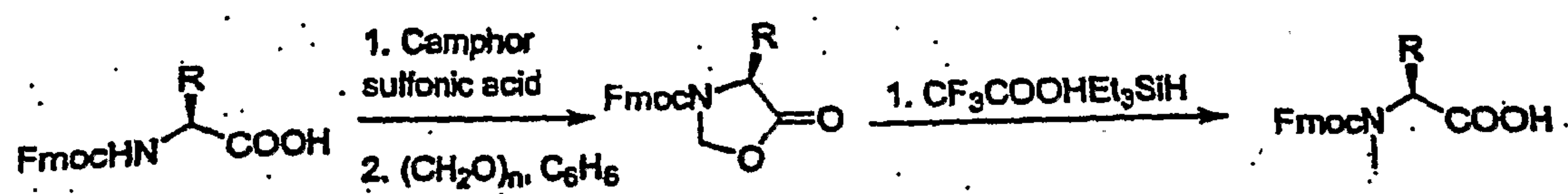


FIG. 7

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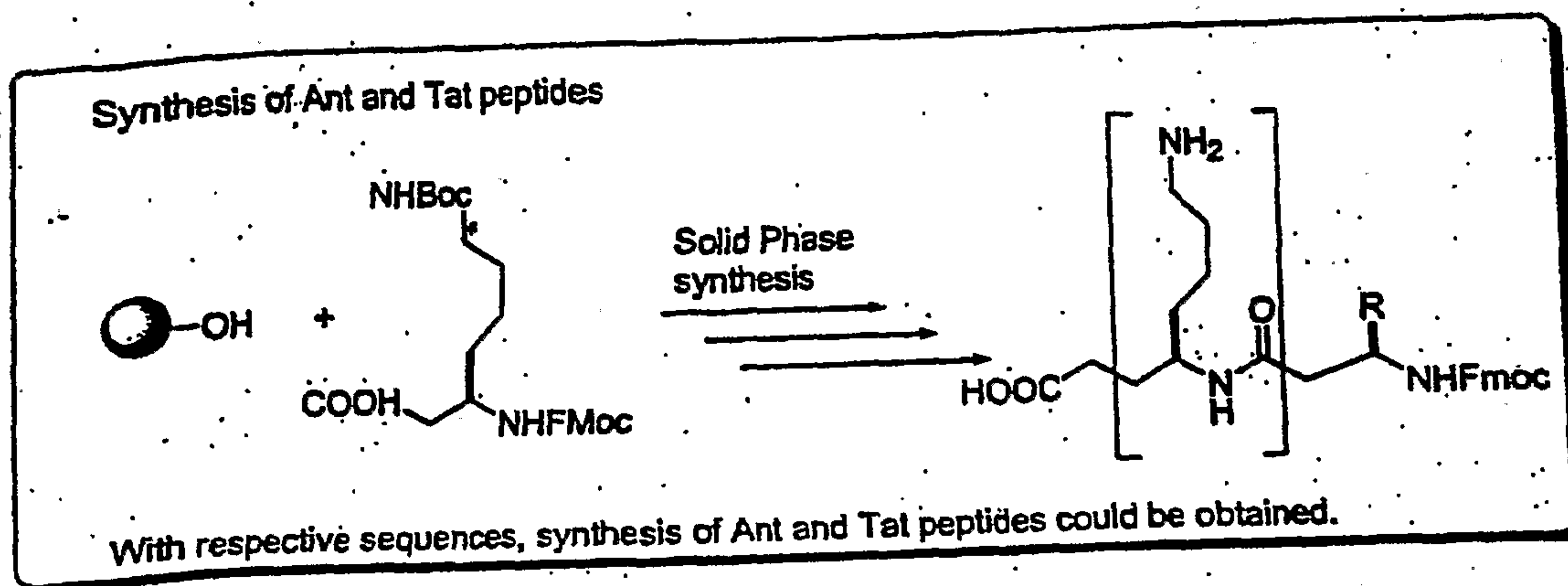
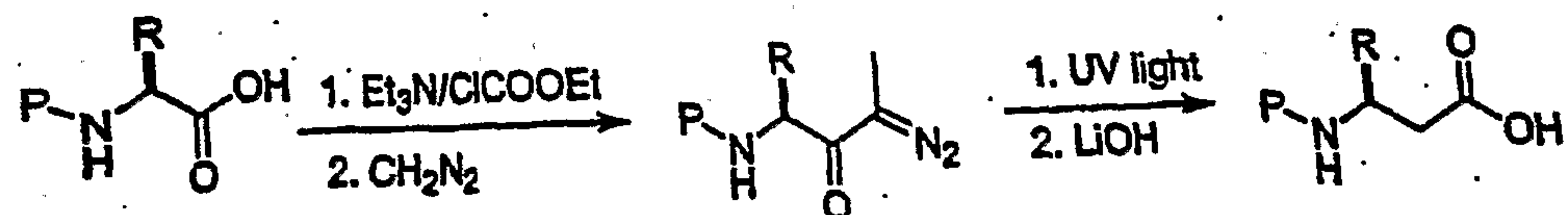


FIG. 8

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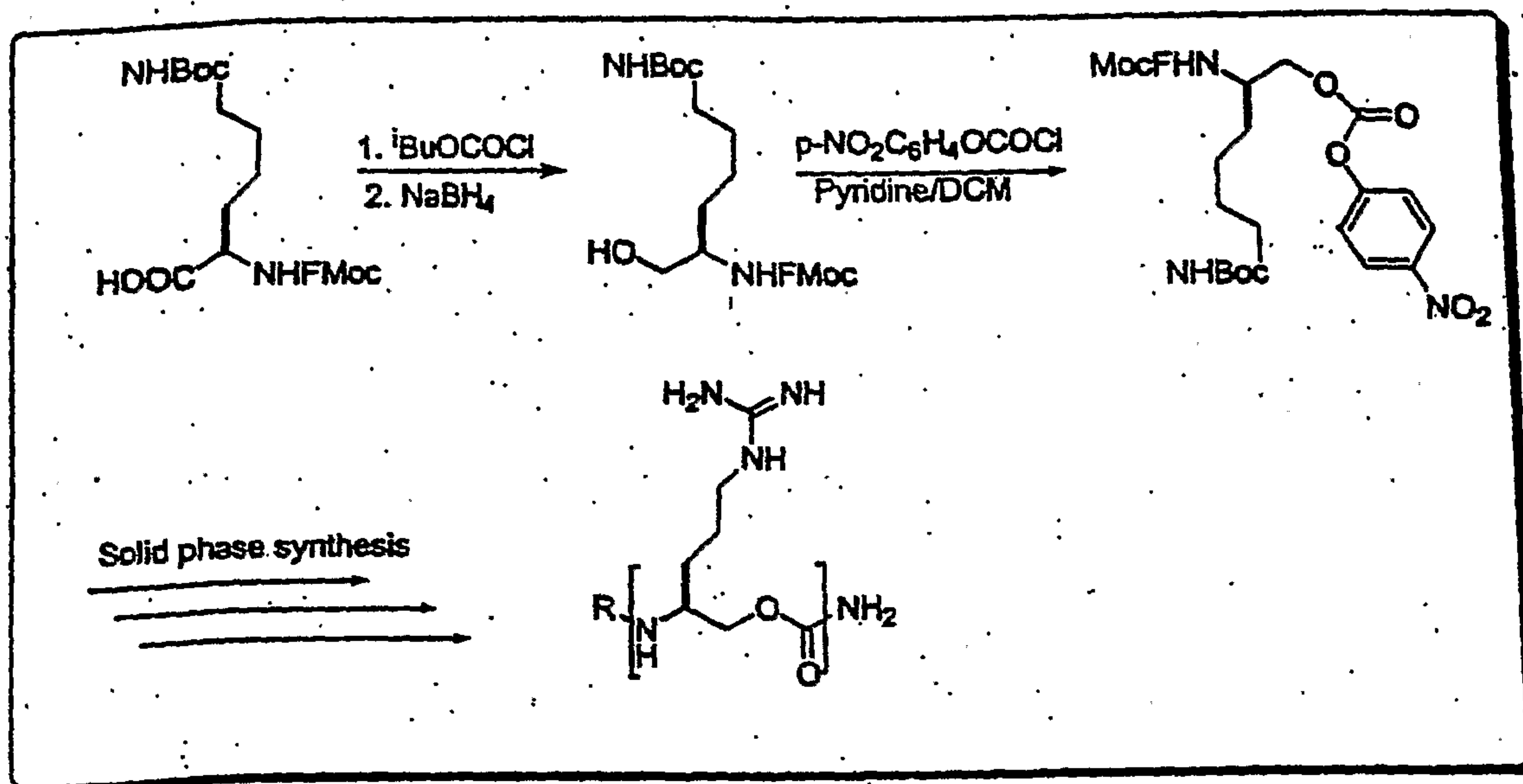


FIG. 9

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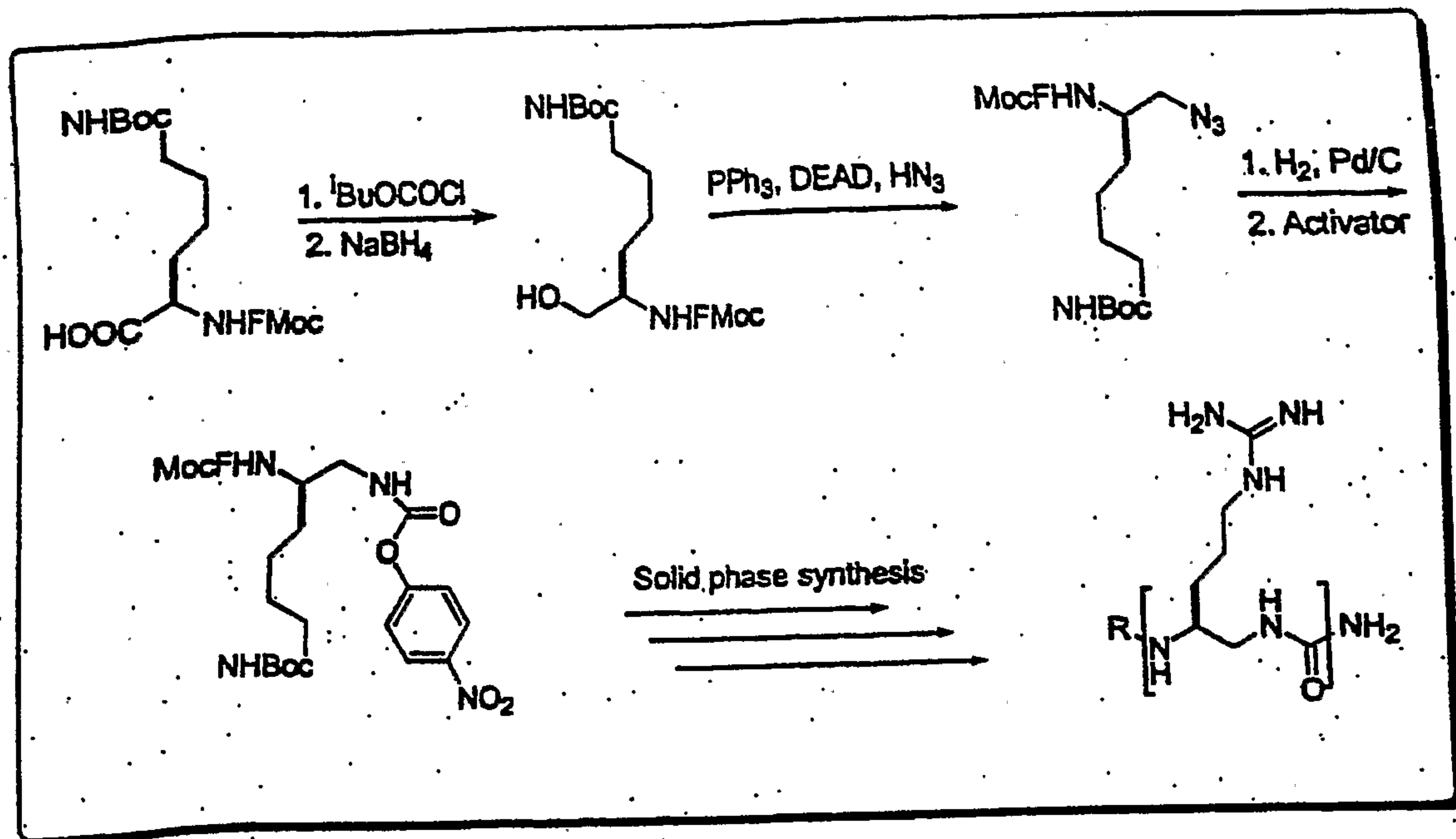


FIG. 10

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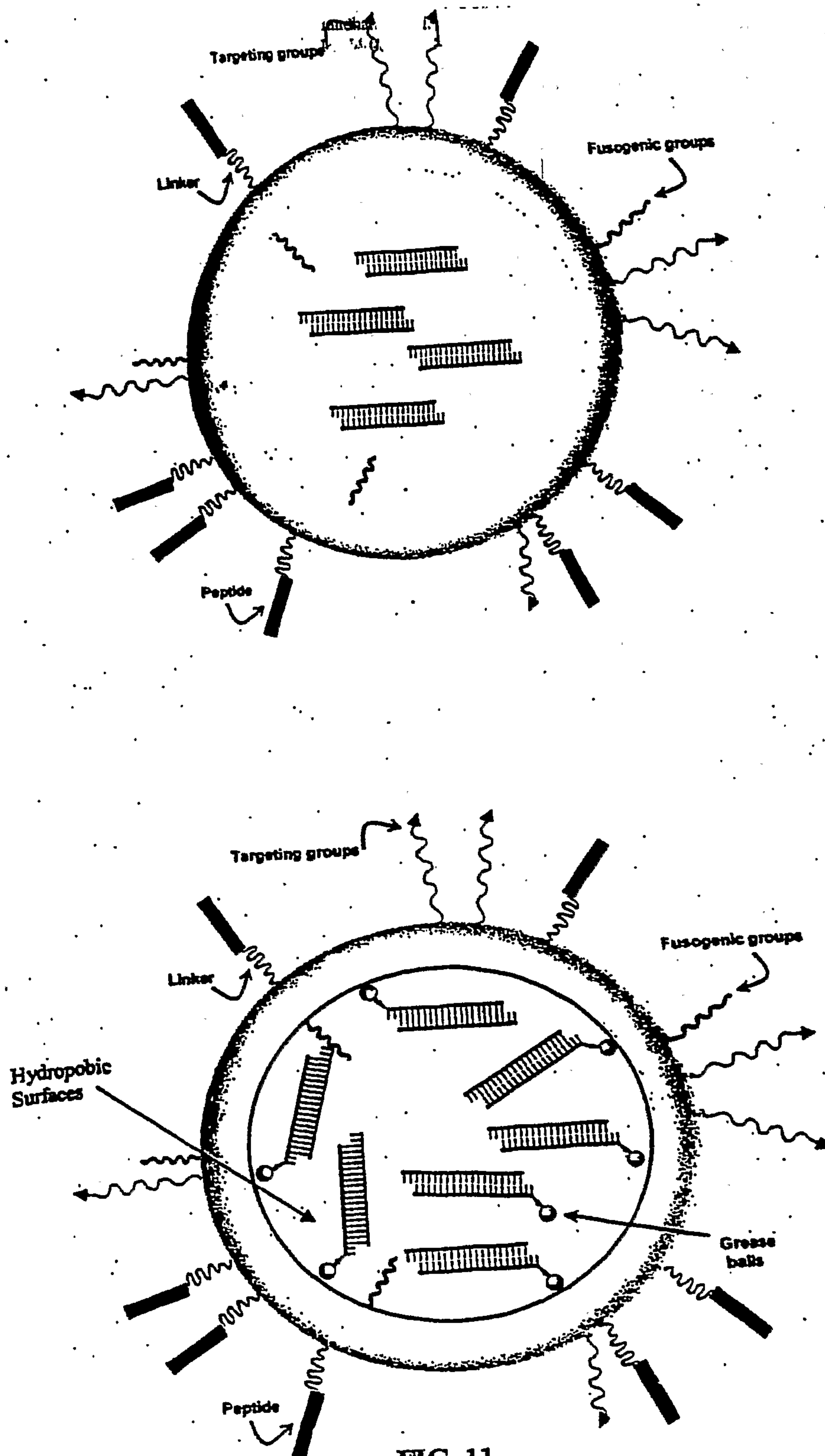


FIG. 11

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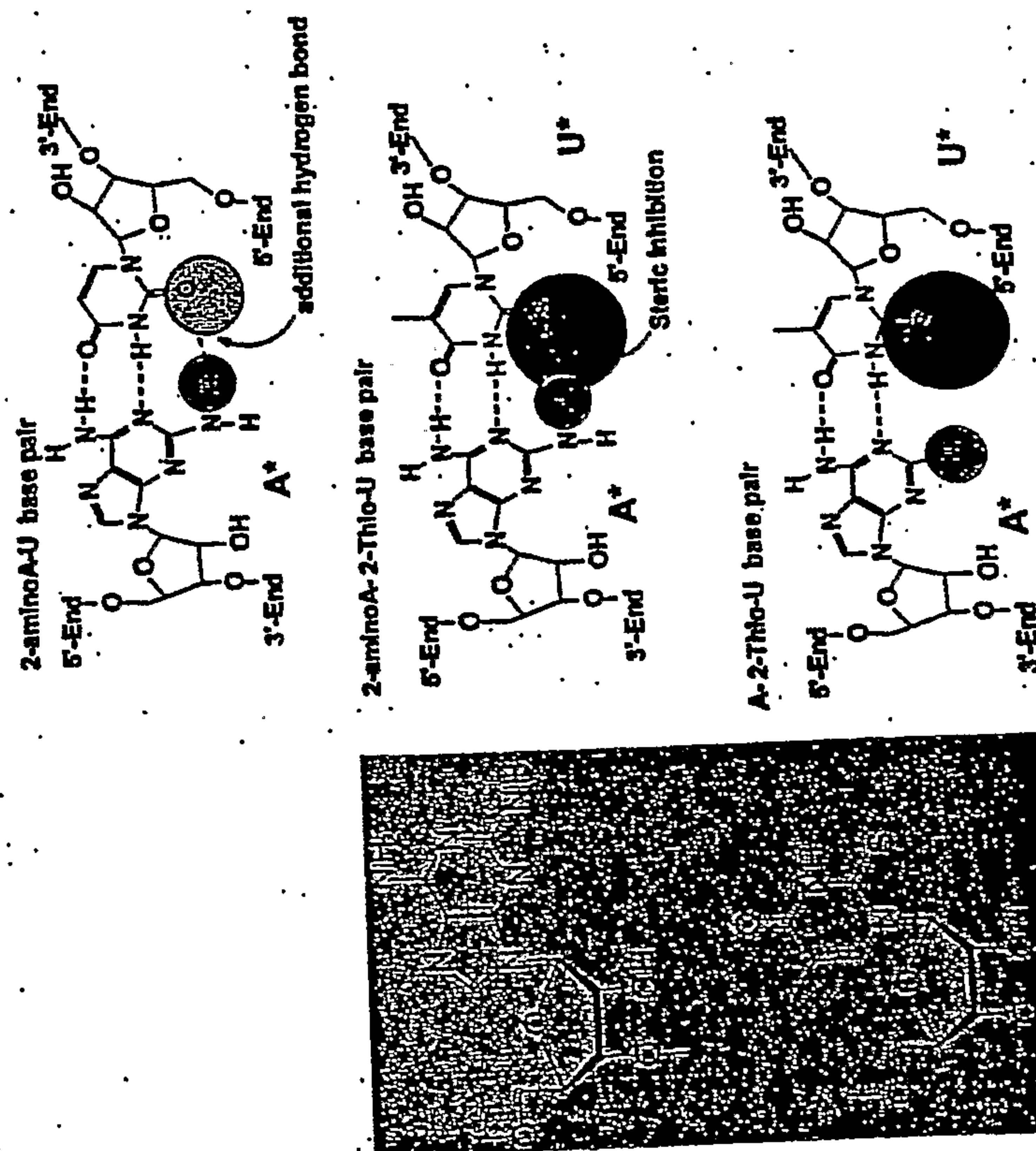


FIG. 12

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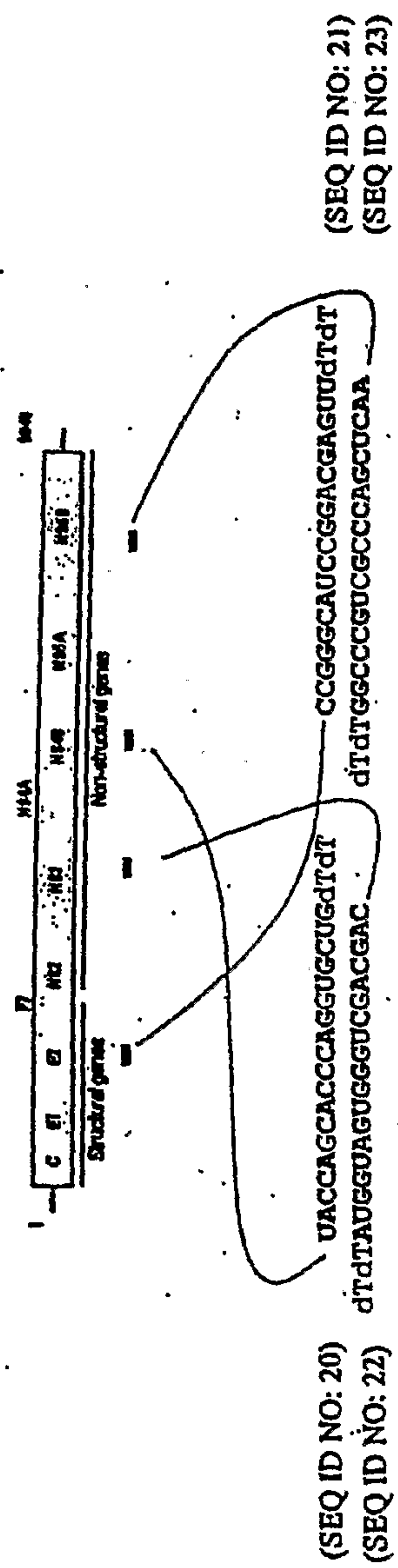


FIG. 13

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AL-DUP-1000

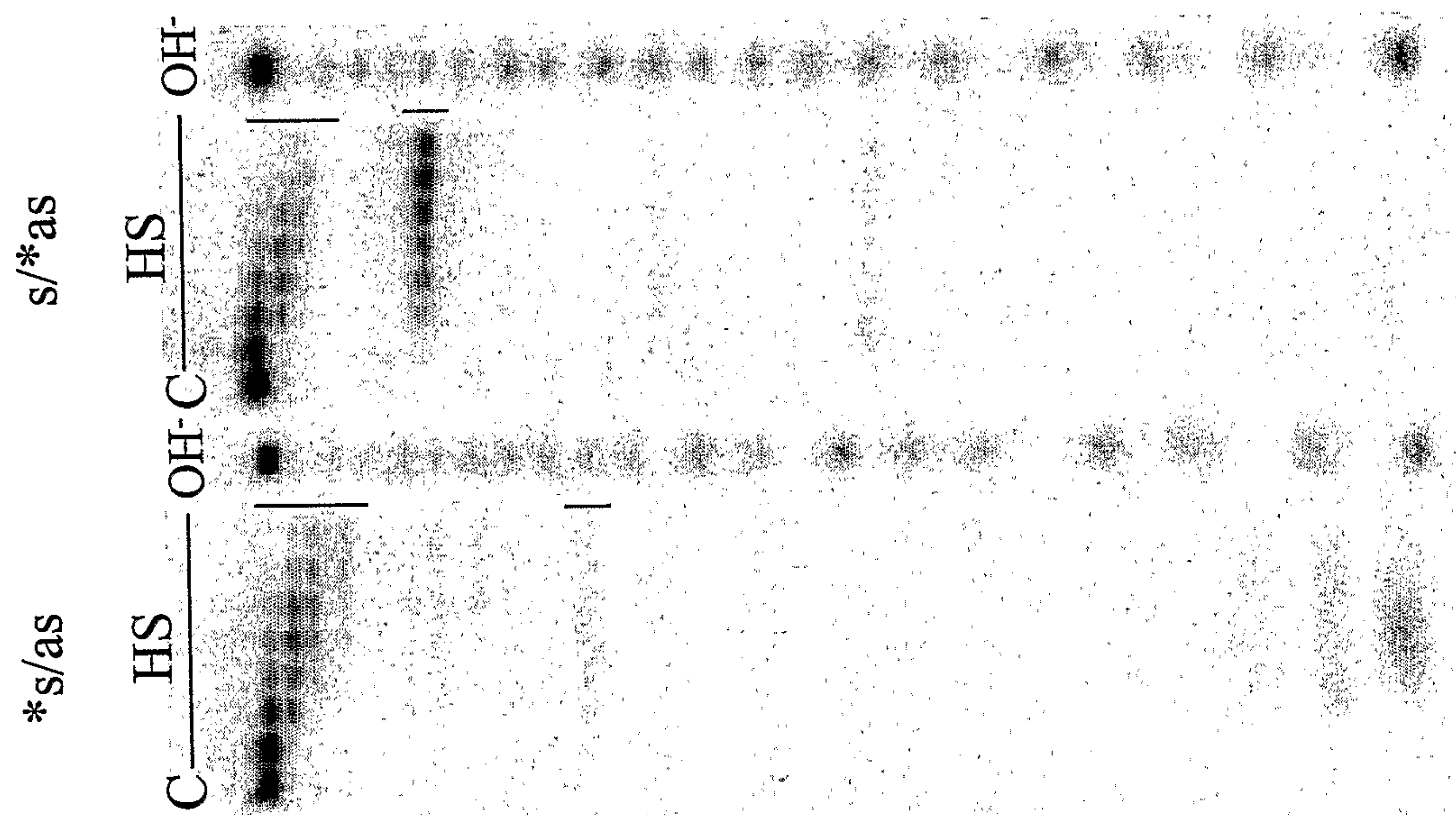


FIG. 14

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AL-DUP-1329

*s/as s/*as

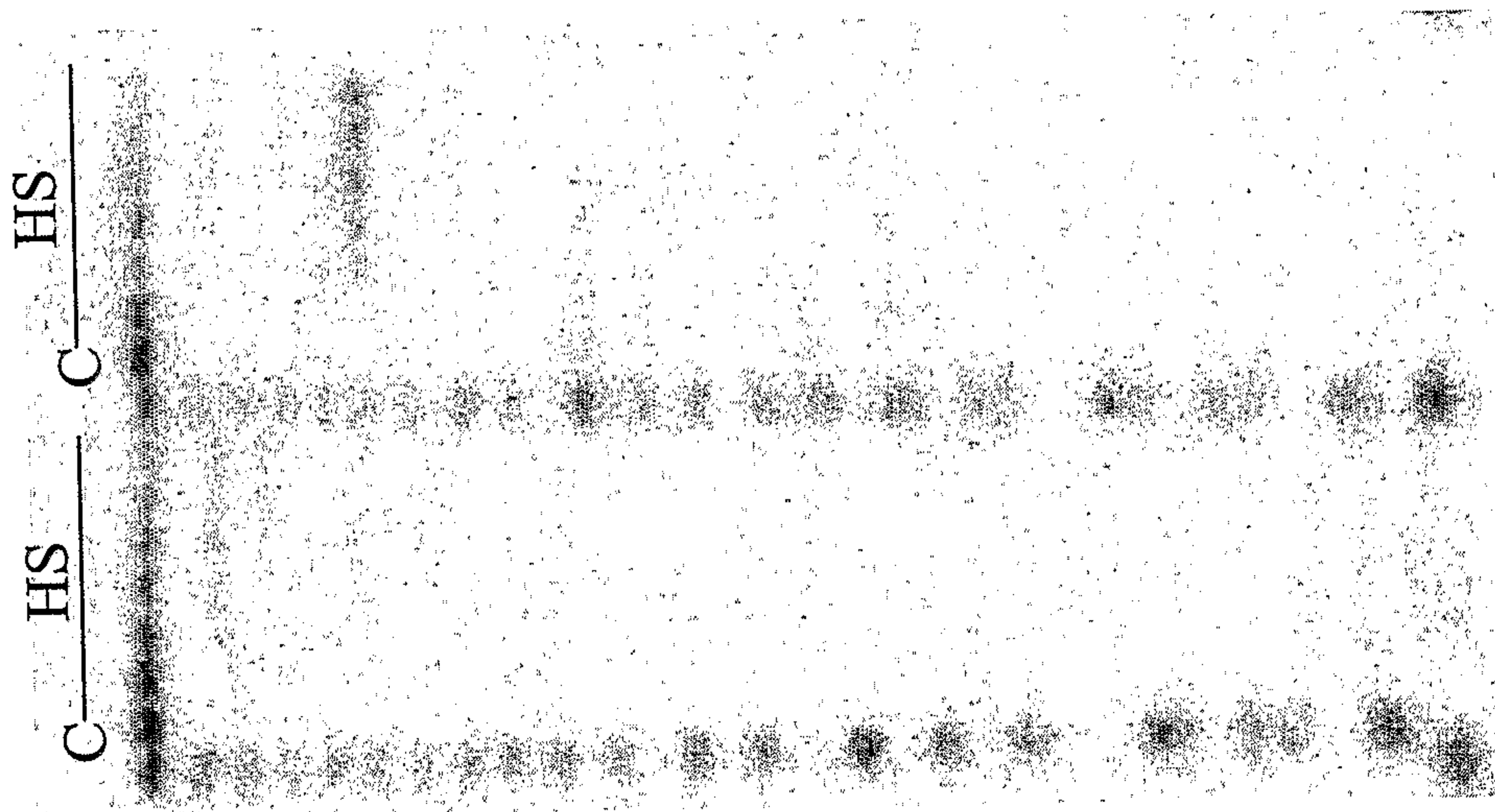


FIG. 15B

AL-DUP-1393

*s/as s/*as

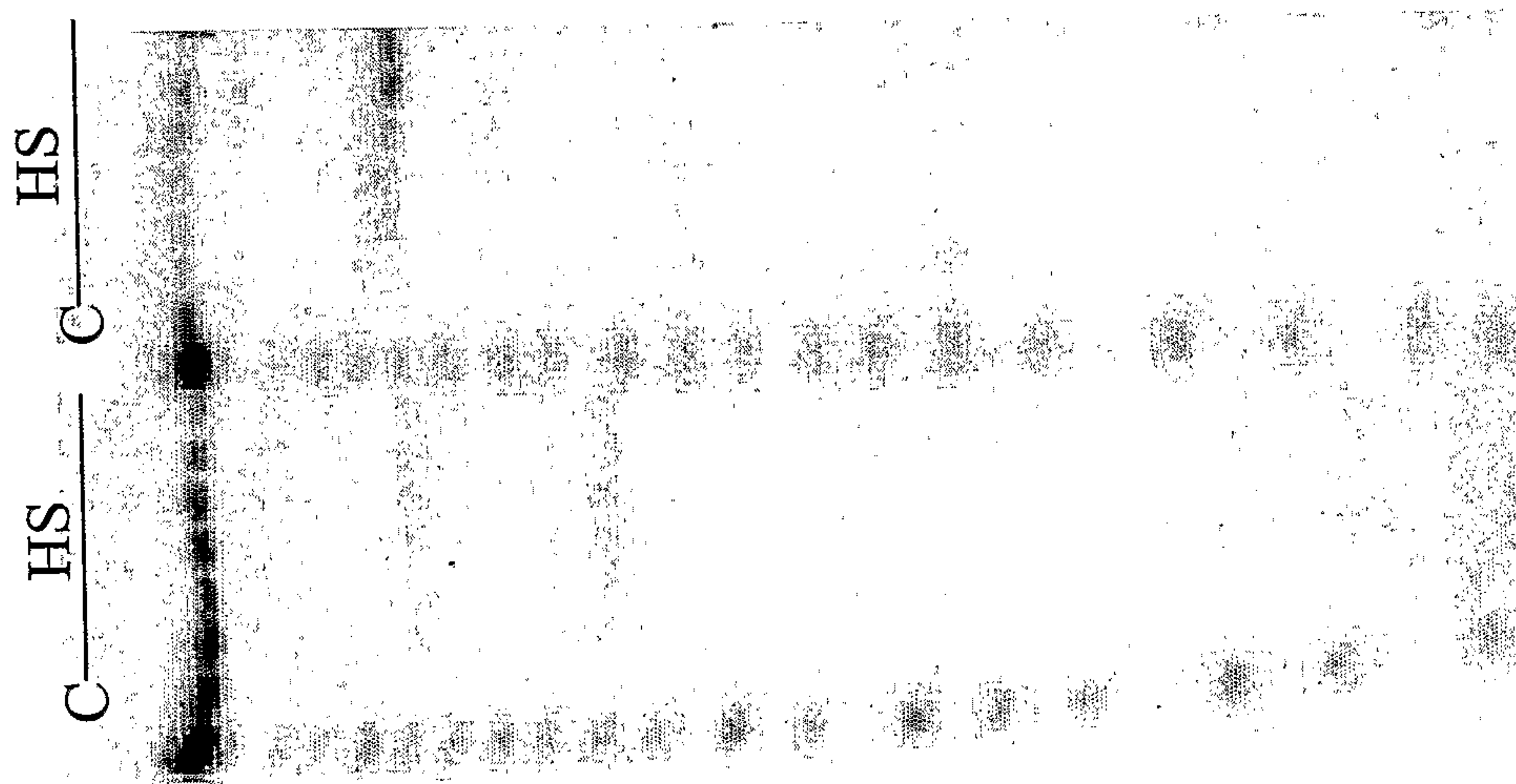
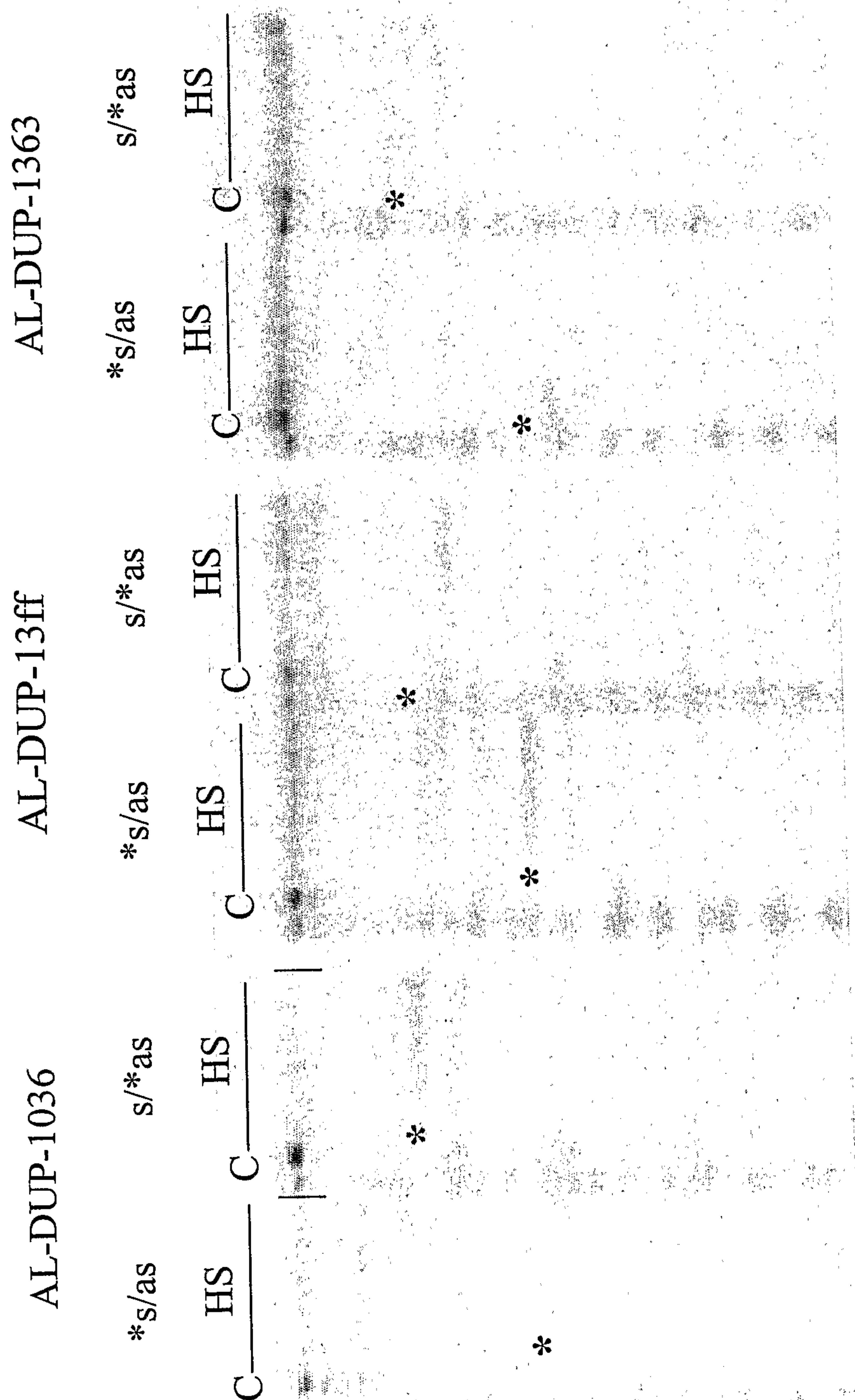


FIG. 15A

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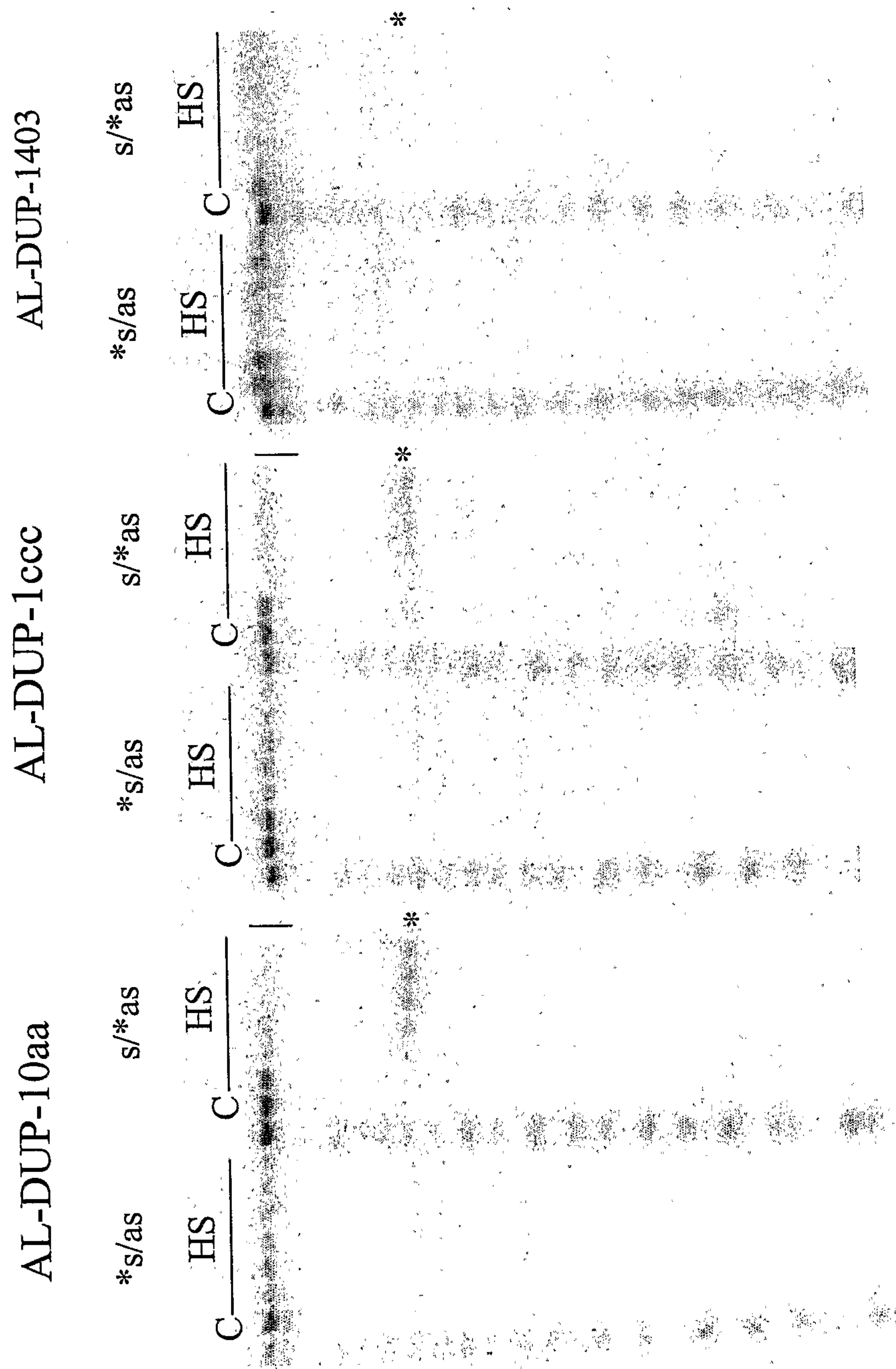


FIG. 17

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AL-DUP-1069

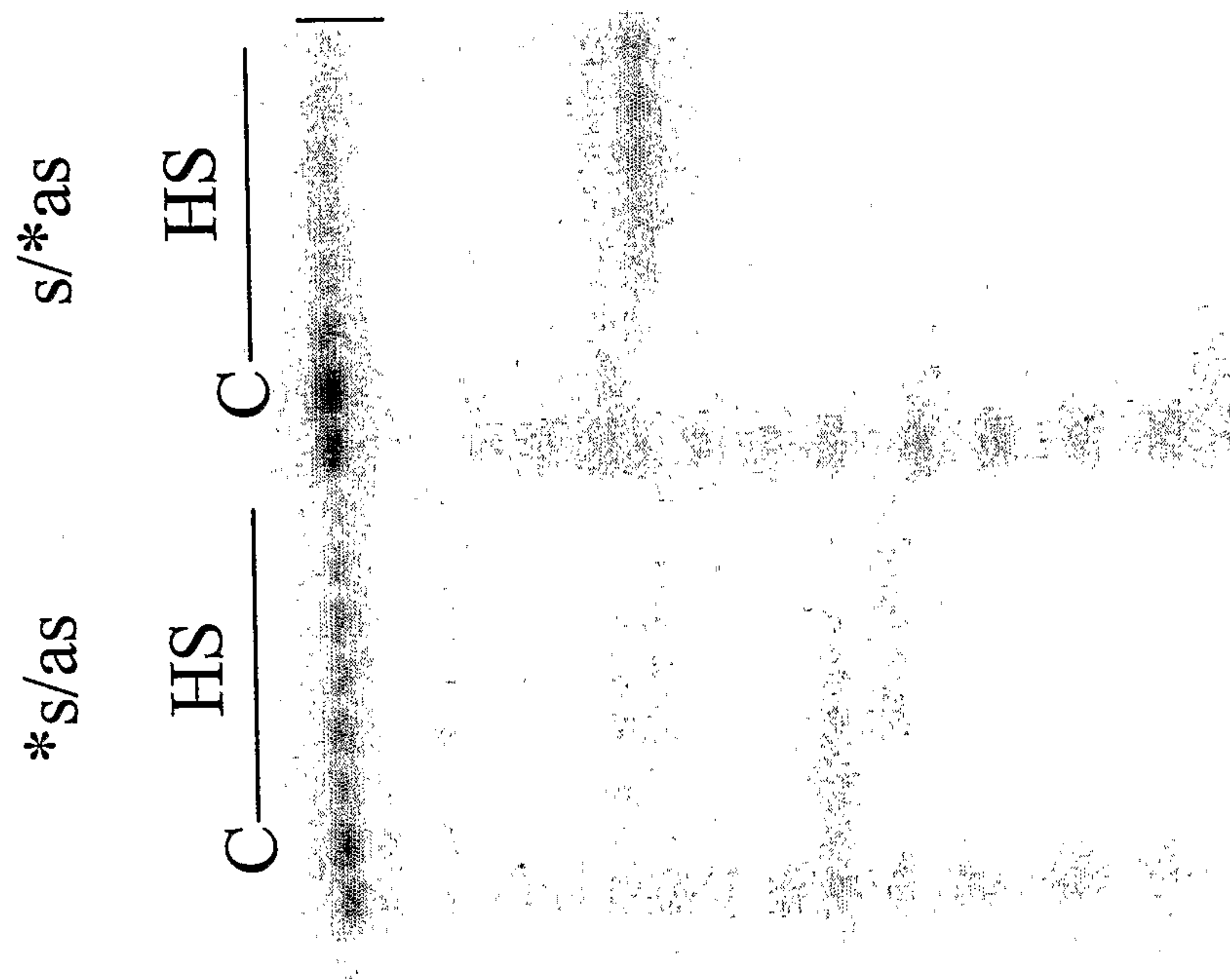


FIG. 18

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FIG. 19A

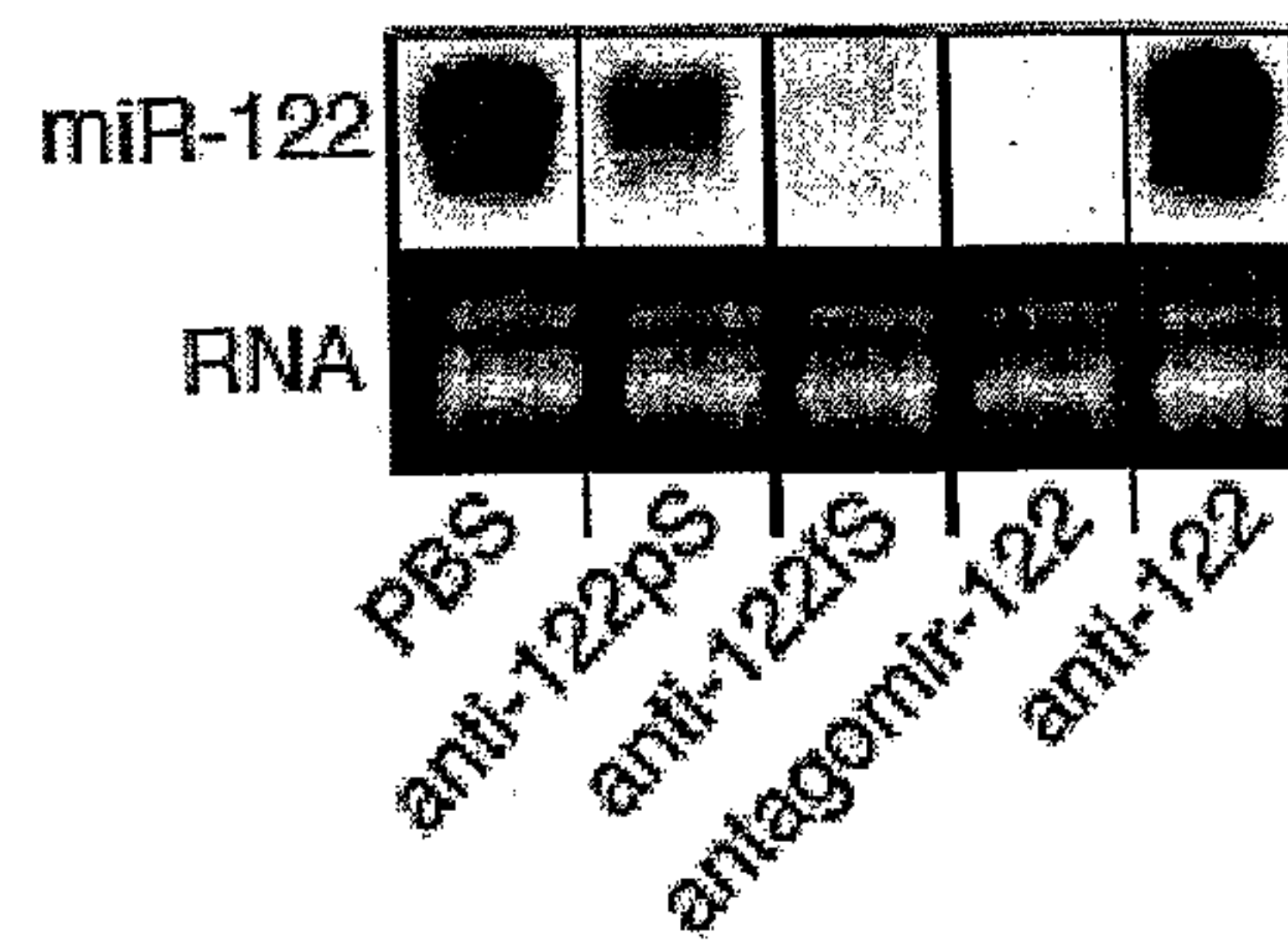


FIG. 19B

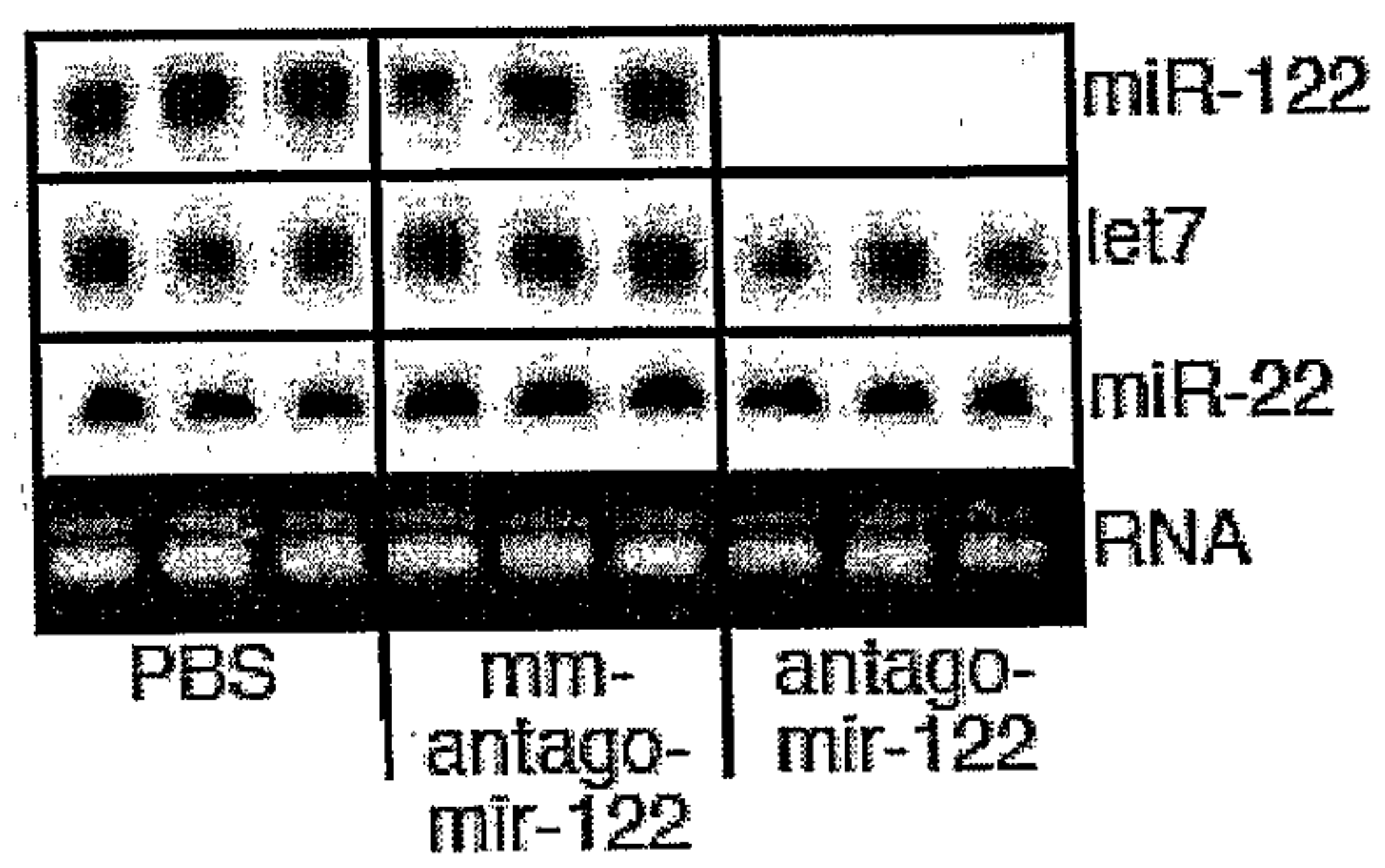
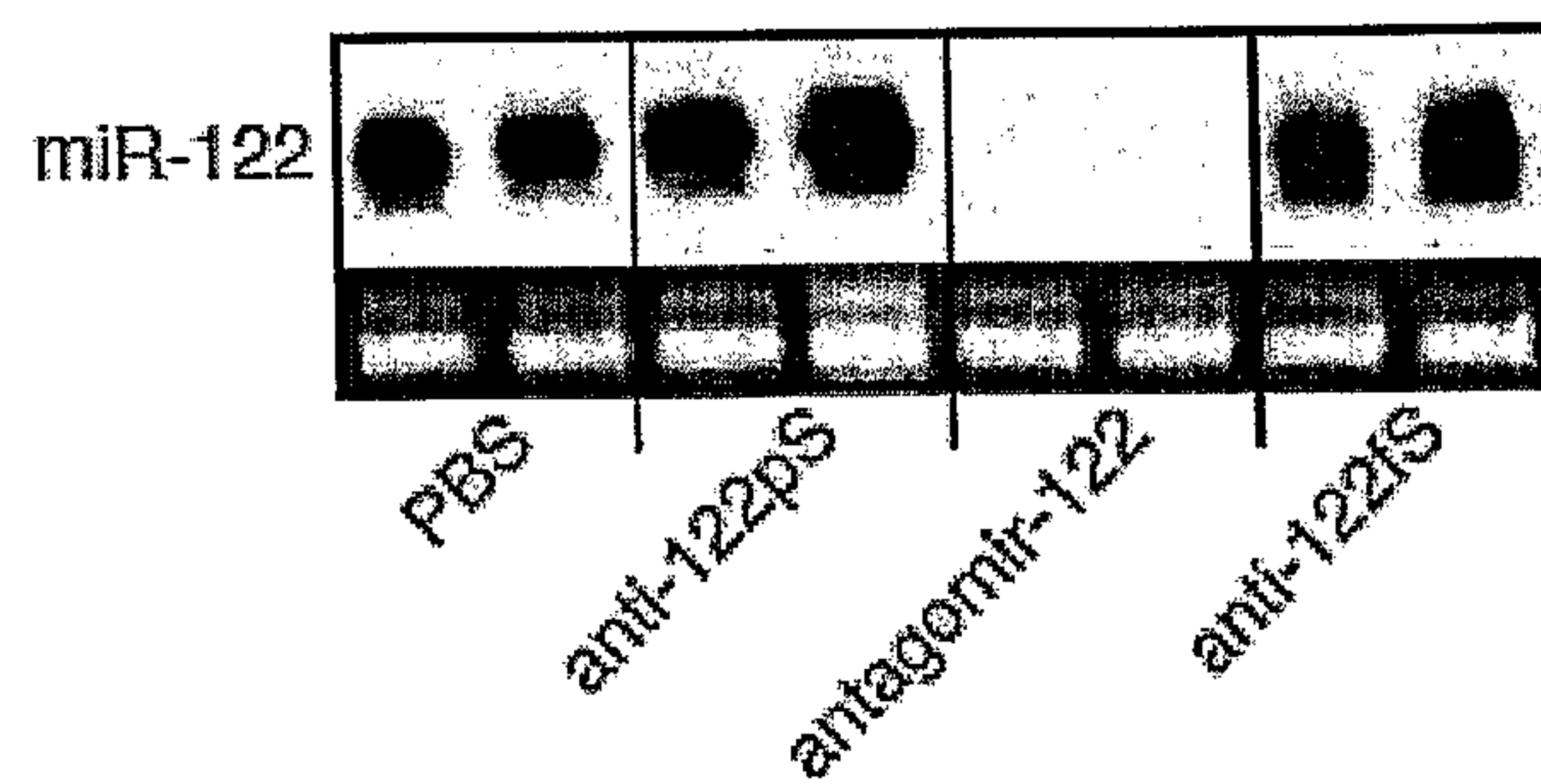


FIG. 19C



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FIG. 20A

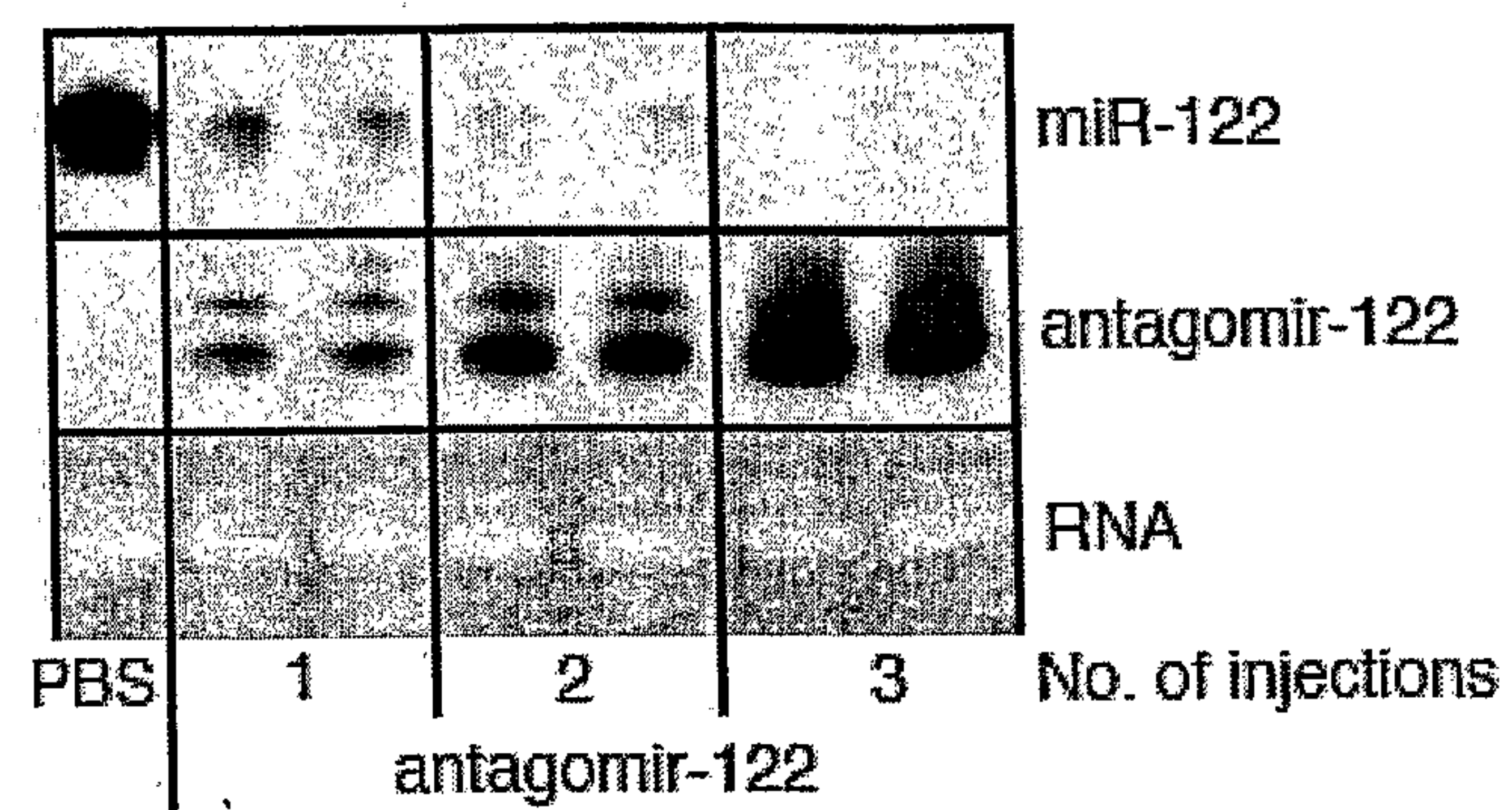


FIG. 20B

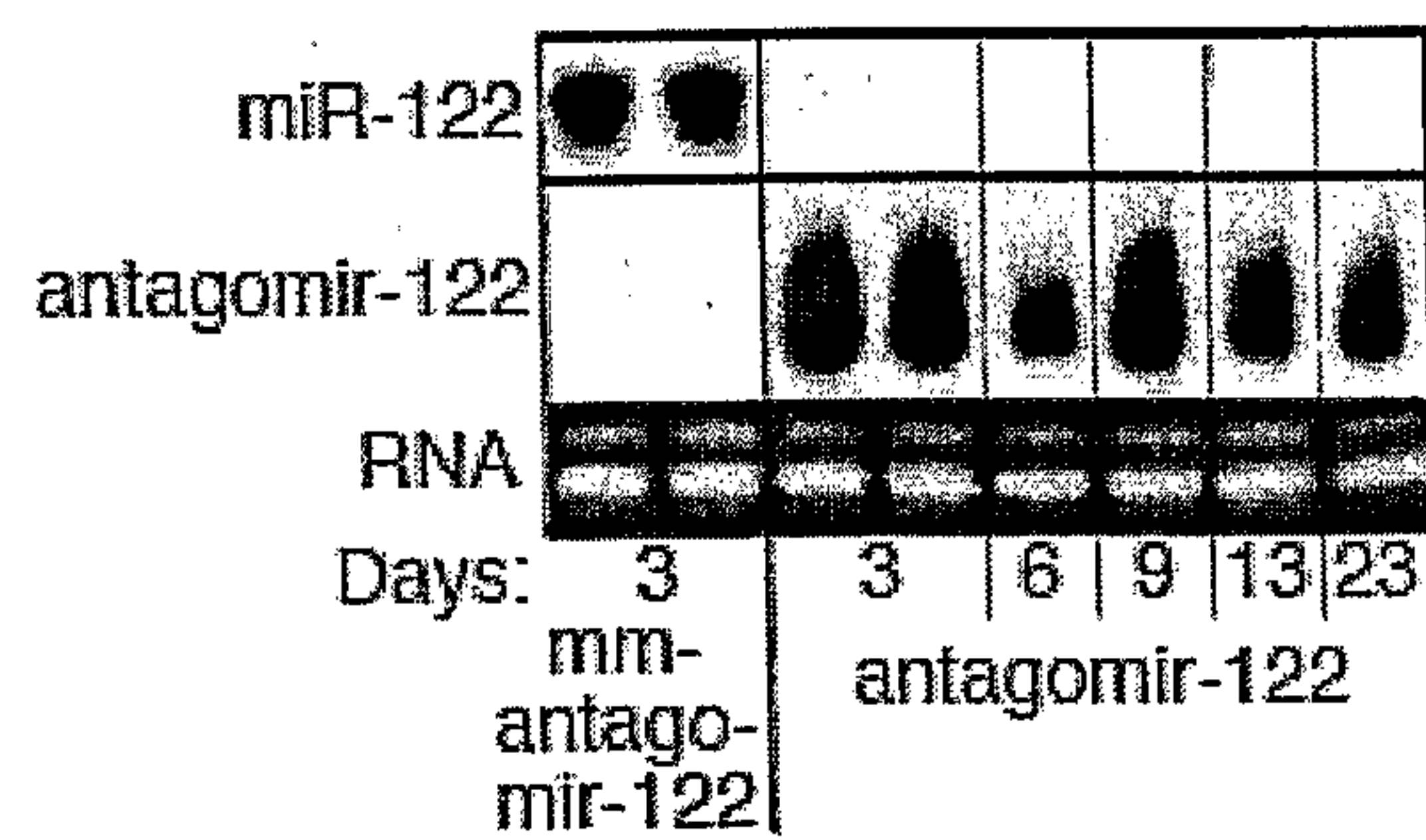


FIG. 21A

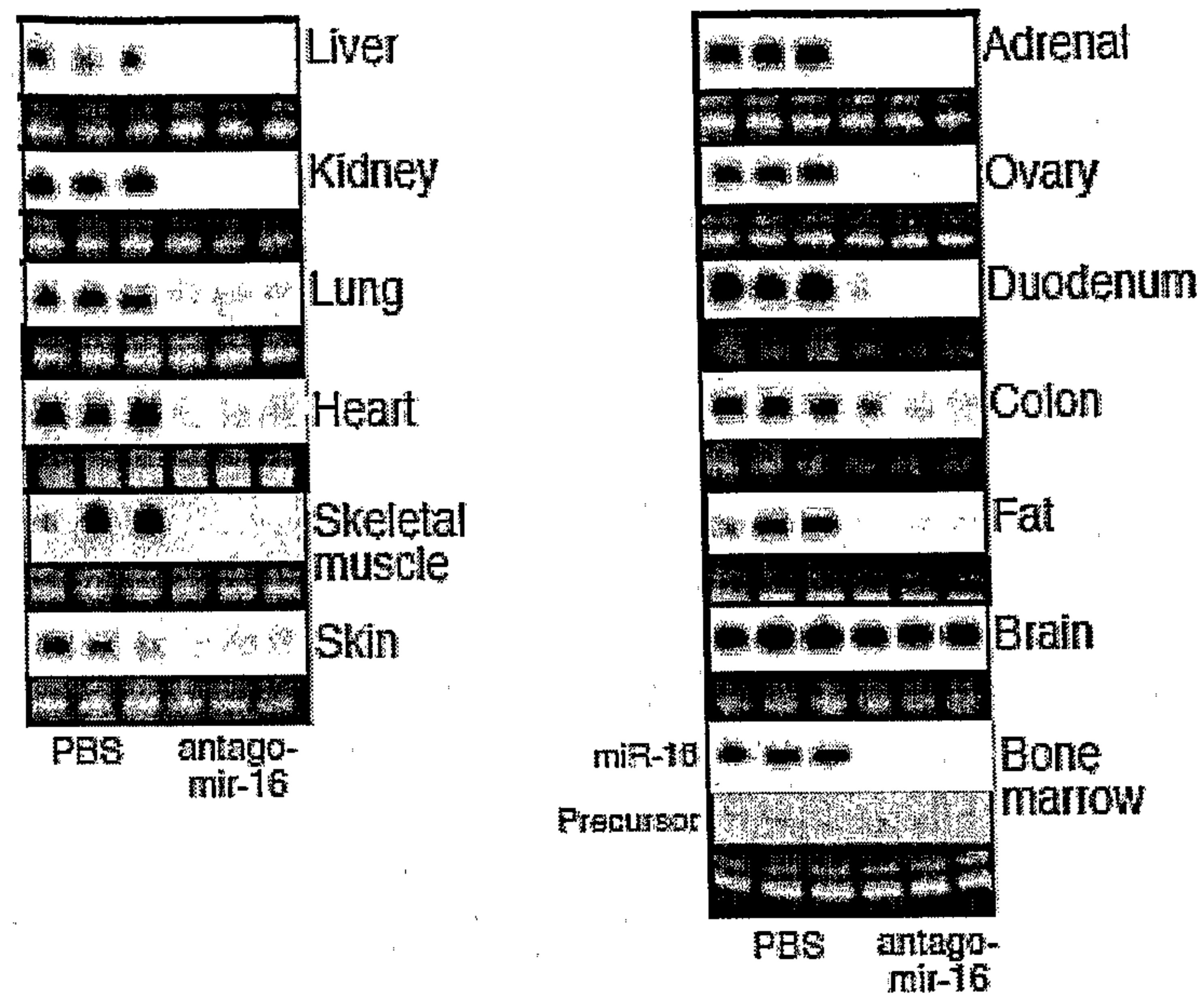
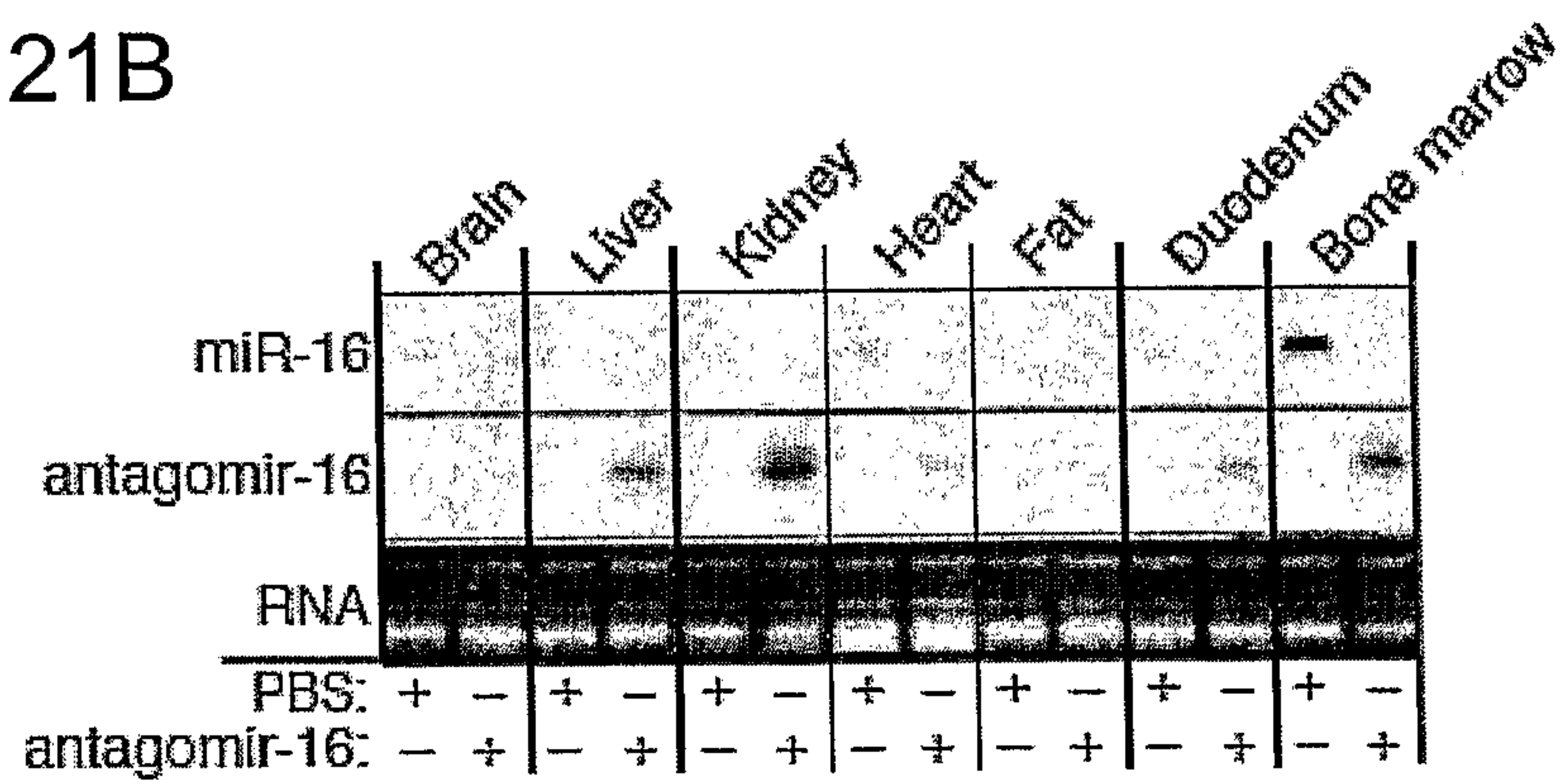
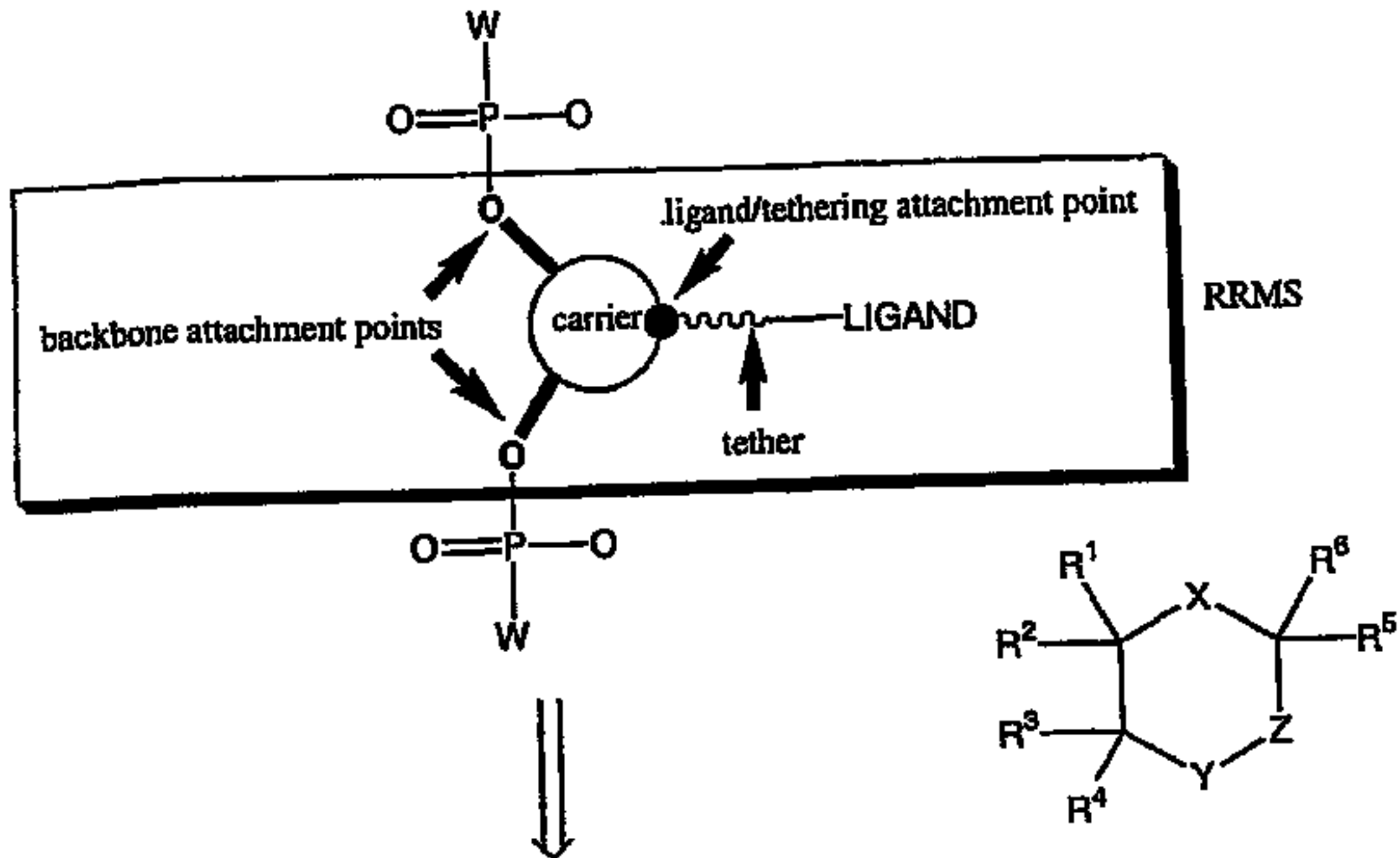


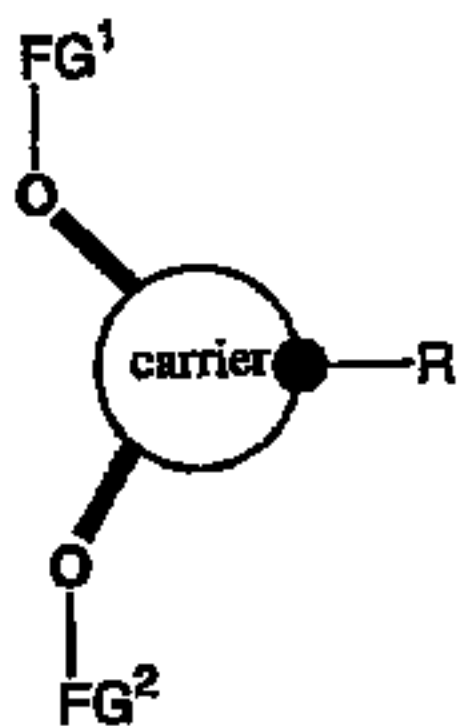
FIG. 21B



(II):



A:

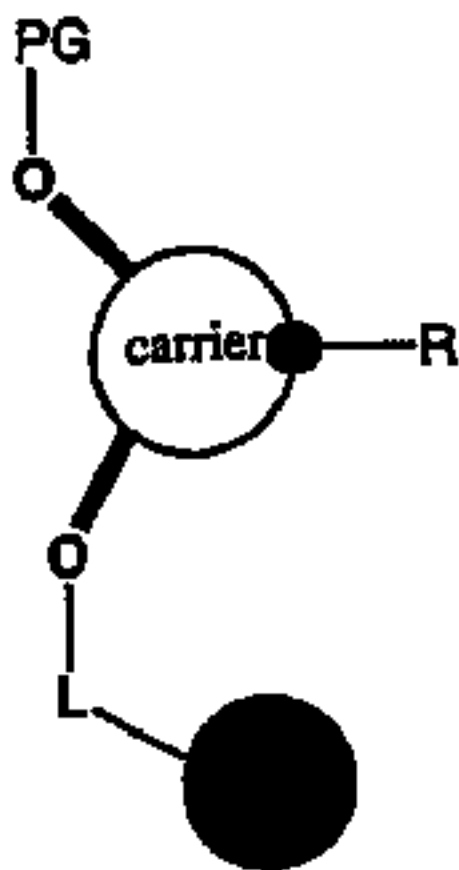


R = H, protecting group;

R = (~~~~~)

R = (~~~~~) + (LIGAND)

B:



C:

