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(57) Abstract			

The present invention relates to an antibody or functional fragment thereof which binds to a mammalian (e.g., human) CC-chemokine receptor (2) (CCR2) or a portion of the receptor and blocks binding of a ligand to the receptor. The invention further relates to a method of inhibiting the interaction of a cell bearing mammalian CCR2 with a ligand thereof, and to use of the antibodies and fragments in therapeutic, prophylactic and diagnostic methods.



ANTI-CCR2 ANTIBODIES AND METHODS OF USE THEREFOR

BACKGROUND OF THE INVENTION

Over the past several years a growing family of 5 leukocyte chemoattractant/activating factors, termed chemokines, has been described (Oppenheim, J.J. et al., *Annu. Rev. Immunol.*, 9:617-648 (1991); Schall and Bacon, *Curr. Opin. Immunol.*, 6:865-873 (1994); Baggio, M., et al., *Adv. Immunol.*, 55:97-179 (1994)). Members of this 10 family are produced and secreted by many cell types in response to early inflammatory mediators such as IL-1 β or TNF α . The chemokine superfamily comprises two main branches: the α -chemokines (or CXC chemokines) and the β -chemokines (CC chemokines). The α -chemokine branch includes 15 proteins such as IL-8, neutrophil activating peptide-2 (NAP-2), melanoma growth stimulatory activity (MGSA/gro or GRO α), and ENA-78, each of which have attracting and activating effects predominantly on neutrophils. The members of the β -chemokine branch affect other cell types such as monocytes, 20 lymphocytes, basophils, and eosinophils (Oppenheim, J.J. et al., *Annu. Rev. Immunol.*, 9:617-648 (1991); Baggio, M., et al., *Adv. Immunol.*, 55:97-179 (1994); Miller and Krangel, *Crit. Rev. Immunol.*, 12:17-46 (1992); Jose, P.J., et al., *J. Exp. Med.*, 179:881-887 (1994); Ponath, P.D., et al., *J. 25 Clin. Invest.*, 97:604-612 (1996)), and include proteins such as monocyte

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chemotactic proteins 1-4 (MCP-1, MCP-2, MCP-3, and MCP-4), RANTES, and macrophage inflammatory proteins (MIP-1 α , MIP-1 β). Recently a new class of membrane-bound chemokines designated CX3C chemokines has been identified (Bazan, J.F., et al., *Nature* 385:640-644 (1997)). Chemokines can mediate a range of pro-inflammatory effects on leukocytes, such as triggering of chemotaxis, degranulation, synthesis of lipid mediators, and integrin activation (Oppenheim, J.J. et al., *Annu. Rev. Immunol.*, 9:617-648 (1991); Bagioli, M., et al., *Adv. Immunol.*, 55:97-179 (1994); Miller, M.D. and Krangel, M.S., *Crit. Rev. Immunol.*, 12:17-46 (1992)). Lately, certain β -chemokines have been shown to suppress HIV-1 infection of human T cell lines *in vitro* (Cocchi, F., et al., *Science (Wash. DC)*, 270:1811-1815 (1995)).

Chemokines bind to 7 transmembrane spanning (7TMS) G protein-coupled receptors (Murphy, P.M., *Annu. Rev. Immunol.*, 12:593-633 (1994)). Some known receptors for the CC or β chemokines include CCR1, which binds MIP-1 α and RANTES (Neote, K., et al., *Cell*, 72:415-425 (1993); Gao, J.L., *J. Exp. Med.*, 177:1421-1427 (1993)); CCR2, which binds chemokines including MCP-1, MCP-2, MCP-3 and MCP-4 (Charo, I.F., et al., *Proc. Natl. Acad. Sci. USA*, 91:2752-2756 (1994); Myers, S.J., et al., *J. Biol. Chem.*, 270:5786-5792 (1995); Gong et al., *J. Biol. Chem.* 272:11682-11685 (1997); Garcia-Zepeda et al., *J. Immunol.* 157:5613-5626 (1996)); CCR3, which binds chemokines including eotaxin, RANTES and MCP-3 (Ponath, P.D., et al., *J. Exp. Med.*, 183:2437-2448 (1996)); CCR4, which has been found to signal in response to MCP-1, MIP-1 α , and RANTES (Power, C.A., et al., *J. Biol. Chem.*, 270:19495-19500 (1995)); and CCR5, which has been shown to signal in response to MIP-1 α , MIP-1 β and RANTES (Boring, L., et al., *J. Biol. Chem.*, 271 (13):7551-7558 (1996); Raport, C.J., *J. Biol. Chem.*, 271:17161-17166

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(1996); and Samson, M. et al., *Biochemistry*, 35:3362-3367 (1996)).

CCR2 is expressed on the surface of several leukocyte subsets, and appears to be expressed in two slightly different forms (CCR2a and CCR2b) due to alternative splicing of the mRNA encoding the carboxy-terminal region (Charo et al., *Proc. Natl. Acad. Sci. USA* 91:2752-2756 (1994)). MCP-1 acts upon monocytes, lymphocytes and basophils, inducing chemotaxis, granule release, respiratory burst and histamine and cytokine release. Studies have suggested that MCP-1 is implicated in the pathology of diseases such as rheumatoid arthritis, atherosclerosis, granulomatous diseases and multiple sclerosis (Koch, *J. Clin. Invest.* 90:772-79 (1992); Hosaka et al., *Clin. Exp. Immunol.* 97:451-457 (1994); Schwartz et al., *Am. J. Cardiol.* 71(6):9B-14B (1993); Schimmer et al., *J. Immunol.* 160:1466-1471 (1998); Flory et al., *Lab. Invest.* 69:396-404 (1993); Gong et al., *J. Exp. Med.* 186:131-137 (1997)). Additionally, CCR2 can act as a co-receptor for HIV (Connor et al., *J. Exp. Med.* 185:621-628 (1997)). Thus, CCR2 receptor antagonists may represent a new class of important therapeutic agents.

SUMMARY OF THE INVENTION

The present invention relates to an antibody (immunoglobulin) or functional fragment thereof (e.g., an antigen-binding fragment) which binds to a mammalian CC-chemokine receptor 2 (also referred to as CCR2, CKR-2, MCP-1RA or MCP-1RB) or portion of the receptor (anti-CCR2). In one embodiment, the antibody of the present invention or fragment thereof has specificity for human or rhesus CCR2 or a portion thereof. In another embodiment, the antibody or fragment of the invention blocks binding of a ligand (e.g., MCP-1, MCP-2, MCP-3, MCP-4) to the receptor and inhibits function associated

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with binding of the ligand to the receptor (e.g., leukocyte trafficking). For example, as described herein, antibodies and fragments thereof of the present invention which bind human or rhesus CCR2 or a portion 5 thereof, can block binding of a chemokine (e.g., MCP-1, MCP-2, MCP-3, MCP-4) to the receptor and inhibit function associated with binding of the chemokine to the receptor. In one embodiment, the antibody is monoclonal antibody (mAb) LS132.1D9 (1D9) or an antibody which can 10 compete with 1D9 for binding to human CCR2 or a portion of human CCR2. Functional fragments of the foregoing antibodies are also envisioned.

In another embodiment, the antibody or functional fragment of the present invention binds human CCR2 or a 15 portion thereof, and inhibits human immunodeficiency virus (HIV) binding to the receptor, thereby inhibiting function associated with binding of HIV to the receptor (e.g., HIV antigen release and infectivity). In one embodiment, the antibody is monoclonal antibody 1D9 or 20 an antibody which can compete with 1D9 for binding to human CCR2 or a portion of human CCR2.

The present invention also relates to an antibody or functional fragment thereof (e.g., an antigen-binding fragment) which binds to a mammalian CCR2 or portion of 25 the receptor and provides increased fluorescent staining intensity of CCR2 or compositions comprising CCR2 relative to other anti-CCR2 antibodies. In one embodiment, the antibody is monoclonal antibody 1D9 or LS132.8G2 (8G2) or an antibody which can compete with 30 1D9 or 8G2 for binding to human CCR2 or a portion of human CCR2.

The present invention further relates to a method of inhibiting the interaction of a cell bearing mammalian (e.g., human, non-human primate or murine) 35 CCR2 with a ligand thereof, comprising contacting the cell with an effective amount of an antibody or

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functional fragment thereof which binds to a mammalian CCR2 or a portion of CCR2. Suitable cells include granulocytes, leukocytes, such as monocytes, macrophages, basophils and eosinophils, mast cells, and 5 lymphocytes including T cells (e.g., CD8+ cells, CD4+ cells, CD25+ cells, CD45RO+ cells), and other cells expressing CCR2 such as a recombinant cell expressing CCR2 (e.g., transfected cells). In a particular embodiment, the antibody is 1D9 or an antibody which can 10 compete with 1D9 for binding to human CCR2 or a portion of human CCR2.

Another embodiment of the invention relates to a method of inhibiting the interaction of a cell bearing mammalian CCR2 with a chemokine, comprising contacting 15 said cell with an effective amount of an antibody or functional fragment thereof which binds to CCR2 or a portion of said receptor. In one embodiment of the method, the antibody or functional fragment thereof is any one or more of 1D9, an antigen-binding fragment of 20 1D9 or an antibody or fragment thereof having an epitopic specificity which is the same as or similar to that of 1D9. Furthermore, the invention relates to a method of inhibiting a function associated with binding 25 of a chemokine to CCR2, comprising administering an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 protein or a portion of said receptor. In one aspect of the method, the antibody or functional fragment thereof is any one or more of 1D9, an antigen-binding fragment of 1D9 or an 30 antibody or fragment thereof having an epitopic specificity which is the same as or similar to that of 1D9.

Another aspect of the invention is a method of identifying expression of a mammalian CCR2 or portion of 35 the receptor by a cell. According to the method, a composition comprising a cell or fraction thereof (e.g.,

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a membrane fraction) is contacted with an antibody or functional fragment thereof (e.g., 1D9 or 8G2) which binds to a mammalian CCR2 protein or portion of the receptor under conditions appropriate for binding of the 5 antibody thereto, and the formation of a complex between said antibody or fragment and said protein or portion thereof is detected. Detection of the complex, directly or indirectly, indicates the presence of the receptor on the cell. The present invention also relates to a kit 10 for use in detecting the presence of CCR2 or a portion thereof in a biological sample, comprising an antibody or functional fragment thereof which binds to a mammalian CC-chemokine receptor 2 or a portion of said receptor, and one or more ancillary reagents suitable 15 for detecting the presence of a complex between said antibody or fragment and said protein or portion thereof.

Also encompassed by the present invention are methods of identifying additional ligands or other 20 substances which bind a mammalian CCR2 protein, including inhibitors and/or promoters of mammalian CCR2 function. For example, agents having the same or a similar binding specificity as that of an antibody of the present invention or functional fragment thereof can 25 be identified by a competition assay with said antibody or fragment. Thus, the present invention also encompasses methods of identifying ligands or other substances which bind the CCR2 receptor, including inhibitors (e.g., antagonists) or promoters (e.g., 30 agonists) of receptor function. In one embodiment, cells which naturally express CCR2 receptor protein or suitable host cells which have been engineered to express a CCR2 receptor or variant encoded by a nucleic acid introduced into said cells are used in an assay to 35 identify and assess the efficacy of ligands, inhibitors or promoters of receptor function. Such cells are also

useful in assessing the function of the expressed receptor protein or polypeptide.

Thus, the invention also relates to a method of detecting or identifying an agent which binds a 5 mammalian CCR2 or ligand binding variant thereof, comprising combining an agent to be tested, an antibody or antigen-binding fragment of the present invention (e.g., monoclonal antibody 1D9, an antibody having an epitopic specificity which is the same as or 10 similar to that of 1D9, antigen-binding fragments of 1D9, monoclonal antibody 8G2, an antibody having an epitopic specificity which is the same as or similar to that of 8G2, and antigen-binding fragments of 8G2) and a composition comprising a mammalian CCR2 protein or a 15 ligand binding variant thereof. The foregoing components can be combined under conditions suitable for binding of the antibody or antigen-binding fragment to mammalian CCR2 protein or a ligand binding variant thereof, and binding of the antibody or fragment to the 20 mammalian CCR2 protein or ligand binding variant is detected or measured, either directly or indirectly, according to methods described herein or other suitable methods. A decrease in the amount of complex formed relative to a suitable control (e.g., in the absence of 25 the agent to be tested) is indicative that the agent binds said receptor or variant. The composition comprising a mammalian CCR2 protein or a ligand binding variant thereof can be a membrane fraction of a cell bearing recombinant CCR2 protein or ligand binding 30 variant thereof. The antibody or fragment thereof can be labeled with a label such as a radioisotope, spin label, antigen label, enzyme label, fluorescent group and chemiluminescent group. These and similar assays can be used to detect agents, including ligands (e.g., 35 chemokines which interact with CCR2) or other substances, including inhibitors or promoters of

receptor function, which can bind CCR2 and compete with the antibodies described herein for binding to the receptor.

According to the present invention, ligands, 5 inhibitors or promoters of receptor function can be identified in a suitable assay, and further assessed for therapeutic effect. Inhibitors of receptor function can be used to inhibit (reduce or prevent) receptor activity, and ligands and/or promoters can be used to 10 induce (trigger or enhance) normal receptor function where indicated. The present invention also provides a method of treating inflammatory diseases, autoimmune diseases, atherosclerosis, and graft rejection, or HIV infection, comprising administering an inhibitor of 15 receptor function (e.g., chemokine binding or HIV binding) to an individual (e.g., a mammal, such as a human). The present invention further provides a method of stimulating receptor function by administering a novel ligand or promoter to an individual, providing a 20 new approach to selective stimulation of leukocyte function, which is useful, for example, in the treatment of infectious diseases and cancer.

Another aspect of the invention relates to a method of inhibiting HIV infection of a cell which expresses a 25 mammalian CCR2 or portion thereof, comprising contacting the cell with an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or portion of the receptor and inhibits HIV binding and infection. In a particular embodiment of the 30 invention, the antibody or functional fragment thereof is any of 1D9, an antibody having an epitopic specificity which is the same as or similar to that of 1D9, an antibody which can compete with 1D9 for binding to human CCR2, and antigen-binding fragments thereof.

35 Also encompassed by the present invention is a method of inhibiting (e.g., treating) HIV in a patient,

comprising administering to the patient an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or a portion of said receptor and inhibits HIV binding to the CCR2 receptor.

5 The anti-CCR2 antibody or fragment can be administered alone or in combination with one or more additional therapeutic agents, e.g., one or more antibodies which bind a co-receptor for HIV infection and inhibit binding to said co-receptor, such as an anti-CCR3, anti-CCR5, 10 and/or anti-CXCR4 antibody.

Another aspect of the invention also relates to a method of preventing or inhibiting HIV infection in an individual, comprising administering to the individual an effective amount of an antibody or functional

15 fragment thereof which binds to CCR2 and inhibits HIV binding to CCR2. According to the method, preventing HIV infection includes treatment in order to prevent (reduce or eliminate) infection of new cells in an infected individual or in order to prevent infection in 20 an individual who may be, may have been or has been exposed to HIV. For example, individuals such as an HIV infected individual, a fetus of an HIV infected female, or a health care worker can be treated according to the method of the present invention.

25 The present invention also encompasses a method of inhibiting leukocyte trafficking in a patient, comprising administering to the patient an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or portion of said 30 receptor and inhibits function associated with binding of a ligand to the receptor.

The present invention also relates to a method of inhibiting or treating CCR2-mediated disorders, such as inflammatory disorders, comprising administering to a 35 patient an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or

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portion of said receptor and inhibits CCR2-mediated function. For example, the invention relates to a method of inhibiting or treating stenosis or restenosis of the vasculature comprising administering to a patient 5 an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or portion of said receptor and inhibits CCR2-mediated function.

The present invention further relates to an 10 antibody or fragment thereof as described herein (e.g., monoclonal antibody 1D9 or an antigen-binding fragment thereof) for use in therapy (including prophylaxis) or diagnosis, and to the use of such an antibody or fragment for the manufacture of a medicament for the 15 treatment of a CCR2-mediated disorder, or other disease or inflammatory condition as described herein.

BRIEF DESCRIPTION OF THE FIGURES

Figures 1A-10 are fluorescence activated cell scanning (FACS) histogram profiles illustrating that 20 mAbs 1D9 and 8G2 stain CCR2 transfectants but not CCR5 or CCR1 transfectants. L1/2 (also referred to herein as L1.2) murine pre-B lymphoma host cells were transfected with CCR2, CCR5 and CCR1 as indicated, and stained with antibodies with different receptor specificities. 25 Staining was analyzed by flow cytometry.

Figures 2A-2L are FACS dot plots showing expression of CCR2 on most monocytes, a subpopulation of 30 lymphocytes and a small subset of granulocytes. Whole blood cells were stained with one of three anti-CCR2 mAbs (5A11, generated using a peptide consisting of the first 32 amino acids of the CCR2 amino-terminus as an immunogen, and 1D9 and 8G2 generated as described herein using CCR2b L1/2 cell transfectants as the immunogen). 35 Staining was analyzed by flow cytometry, and the lymphocyte, granulocyte and monocyte populations were

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gated using the forward and side light scatter. The X-axis represents forward light scatter (a measure of cell size), and the Y-axis fluorescence intensity of staining for CCR2. The level of negative control staining is
5 indicated by a line.

Figures 3A-3I are FACS dot plots showing that mAb 1D9 stains an IgE positive population in peripheral blood (basophils) using two-color staining for IgE and CCR2. Whole blood cells were first stained with either
10 a negative control antibody (anti-Flag), anti-CCR2 antibody 1D9, or an anti-CXCR1 antibody, as indicated, and detected by an anti-mouse-FITC conjugate. A second staining was done using either PBS or a biotinylated antibody specific for IgE or CD16, as indicated, and
15 detected with a streptavidin-phycoerythrin. Staining was analyzed by flow cytometry.

Figure 4 illustrates that mAb 1D9 inhibits [¹²⁵I]MCP-1 binding to THP-1 cell membranes. 3.0 µg of THP-1 membrane protein was incubated with 0.1 nM
20 [¹²⁵I]MCP-1 in the presence of various concentrations of 1D9 or the isotype-matched anti-CXCR3 antibody 1C6. The amount of bound tracer was determined by separation of free from bound by filtration and scintillation counting. The data was analyzed to determine the IC₅₀,
25 value by non-linear regression using a 4-parameter logistic equation with KaleidaGraph software.

Figure 5 illustrates that mAb 1D9 inhibits [¹²⁵I]MCP-1 binding to fresh human PBMC. Freshly isolated peripheral blood mononuclear cells (500,000)
30 were incubated with 0.1 nM [¹²⁵I]MCP-1 in the presence of various concentrations of 1D9 or the isotype-matched anti-CXCR3 antibody 1C6. The amount of bound tracer was determined by separation of free from bound by filtration and scintillation counting. The data was
35 analyzed to determine the IC₅₀ value as for Figure 4.

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Figures 6A and 6B are graphs demonstrating that mAb 1D9 inhibits MCP-1-induced chemotaxis, but not RANTES-induced chemotaxis, of fresh PBMC. Figure 6A shows the results of chemotaxis assays of PBMC to 10 nM MCP-1 with 5 no antibody, or 0.1 or 10 μ g/ml of 1D9 or nonspecific murine IgG2a. The spontaneous nonspecific migration is also indicated. Figure 6B shows the results of chemotaxis assays of PBMC to 10 nM RANTES with no antibody, 10 μ g/ml 1D9 or 10 μ g/ml nonspecific murine 10 IgG2a. The spontaneous nonspecific migration in the absence of RANTES is also indicated.

DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to an antibody (anti-CCR2) or functional fragment thereof which binds 15 mammalian CC-chemokine receptor 2 (CCR2, CKR-2, MCP-1RA or MCP-1RB) or a portion of CCR2. In one embodiment, the antibody has specificity for human or rhesus CCR2 or portion thereof. In one embodiment, the antibodies (immunoglobulins) are raised against an isolated and/or 20 recombinant mammalian CCR2 or portion thereof (e.g., peptide) or against a host cell which expresses mammalian CCR2. In a preferred embodiment, the antibodies specifically bind human CCR2 receptor(s) (e.g., CCR2a and/or CCR2b) or a portion thereof, and in 25 a particularly preferred embodiment the antibodies have specificity for a naturally occurring or endogenous human CCR2. As used herein, "CC-chemokine receptor 2" ("CCR2") refers to CC-chemokine receptor 2a and/or CC-chemokine receptor 2b. Antibodies or functional 30 fragments thereof which can inhibit one or more functions characteristic of a mammalian CCR2, such as a binding activity (e.g., ligand, inhibitor and/or promoter binding), a signaling activity (e.g., activation of a mammalian G protein, induction of a 35 rapid and transient increase in the concentration of

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cytosolic free calcium $[Ca^{2+}]_i$), and/or stimulation of a cellular response (e.g., stimulation of chemotaxis, exocytosis or inflammatory mediator release by leukocytes, integrin activation) are also encompassed by 5 the present invention, such as an antibody which can inhibit binding of a ligand (i.e., one or more ligands) to CCR2 and/or one or more functions mediated by CCR2 in response to a ligand. For example, in one aspect, the antibodies or functional fragments thereof can inhibit 10 (reduce or prevent) the interaction of receptor with a natural ligand, such as MCP-1, MCP-2, MCP-3 and/or MCP-4. In another aspect, an antibody or functional fragment thereof that binds to CCR2 can inhibit binding of MCP-1, MCP-2, MCP-3 and/or MCP-4 and/or HIV to 15 mammalian CCR2 (e.g., human CCR2, non-human primate CCR2, murine CCR2). The antibodies or functional fragments thereof of the present invention can inhibit functions mediated by human CCR2, including leukocyte trafficking, HIV entry into a cell, T cell activation, 20 inflammatory mediator release and/or leukocyte degranulation. Preferably, the antibodies or fragments can bind CCR2 with an affinity of at least about 0.1×10^{-9} M, preferably at least about 1×10^{-9} M, and more preferably at least about 3×10^{-9} M. In a particular 25 embodiment, antibodies or functional fragments thereof demonstrate inhibition of chemokine-induced (e.g., MCP-1-induced) chemotaxis of cells (e.g., PBMC) at less than about 150 μ g/ml, preferably less than about 100 μ g/ml, more preferably less than about 50 μ g/ml, and even more 30 preferably less than about 20 μ g/ml.

In a further embodiment of the invention, the antibodies or functional fragments thereof of the invention can inhibit binding of a CCR2 ligand (e.g., a chemokine) to CCR2 with an IC_{50} of less than about 1.0 35 μ g/ml, preferably less than about 0.05 μ g/ml, and more preferably less than about 0.005 μ g/ml.

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Murine monoclonal antibodies specific for CCR2, designated 1D9 and 8G2, were produced as described herein. In a preferred embodiment, the antibodies of the present invention bind human CCR2, and have an epitopic specificity which is the same as or similar to that of murine 1D9 or 8G2 antibody described herein. Antibodies with an epitopic specificity which is the same as or similar to that of murine 1D9 monoclonal antibody can be identified by their ability to compete with murine 1D9 monoclonal antibody for binding to human CCR2 (e.g., to cells bearing human CCR2, such as transfectants bearing CCR2, CD8+ cells, CD4+ cells, CDR45RO+ cells, CD25+ cells, monocytes, dendritic cells, macrophages and basophils). Similarly, antibodies with an epitopic specificity which is the same as or similar to that of murine 8G2 monoclonal antibody can be identified by their ability to compete with murine 8G2 monoclonal antibody for binding to human CCR2. Using receptor chimeras (Rucker et al., *Cell* 87:437-446 (1996)), the binding site of mAbs 1D9 and 8G2 has been mapped to the amino-terminal domain of human CC-chemokine receptor 2, specifically to an epitope comprising from about amino acid 1 to about amino acid 30 of the protein. Using these or other suitable techniques, antibodies having an epitopic specificity which is the same as or similar to that of an antibody of the present invention can be identified. mAbs 1D9 and 8G2 have epitopic specificity for the amino-terminal domain of the CCR2 receptor, e.g., from about amino acid number 1 to about amino acid number 30 of the receptor protein. Thus, the invention pertains to an antibody or functional portion thereof which binds to the amino-terminal domain or portion thereof of mammalian CC-chemokine receptor 2, and particularly to an epitope comprising from about amino acid 1 to about amino acid 30 of mammalian CC-chemokine receptor 2.

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The invention also relates to a bispecific antibody, or functional fragment thereof (e.g., $F(ab')_2$), which has the same or similar epitopic specificity as at least two of the antibodies described herein (see, e.g., 5 U.S. Patent No. 5,141,736 (Iwasa *et al.*), U.S. Patent Nos. 4,444,878, 5,292,668, 5,523,210 (all to Paulus *et al.*) and U.S. Patent No. 5,496,549 (Yamazaki *et al.*)). For example, a bispecific antibody of the present 10 invention can have the same or similar epitopic specificity as mAb 1D9 and 8G2, e.g., binds the amino terminal domain, or portion thereof, of mammalian CCR2 protein.

Hybridoma cell lines producing antibodies according to the present invention were deposited on July 17, 15 1998, on behalf of LeukoSite, Inc., 215 First Street, Cambridge, MA 02142, U.S.A., at the American Type Culture Collection, 10801 University Boulevard, Manassas, Virginia 20110, U.S.A., under Accession Nos. HB-12549 (1D9) and HB-12550 (8G2). The present invention 20 also pertains to the hybridoma cell lines deposited under ATCC Accession No. HB-12549 and ATCC Accession No. HB-12550, as well as to the monoclonal antibodies produced by the hybridoma cell lines deposited under ATCC Accession Nos. HB-12549 and HB-12550.

25 The antibodies of the present invention can be polyclonal or monoclonal, and the term "antibody" is intended to encompass both polyclonal and monoclonal antibodies. Furthermore, it is understood that methods described herein which utilize 8G2 can also utilize 30 functional fragments (e.g., antigen-binding fragments) of 8G2, antibodies which have the same or similar epitopic specificity as 8G2, and combinations thereof, optionally in combination with antibodies or fragments having an epitopic specificity which is not the same as 35 or similar to 8G2; similarly, methods described as utilizing 1D9 can also utilize functional fragments of

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1D9, antibodies which have the same or similar epitopic specificity as 1D9, and combinations thereof, optionally in combination with antibodies or fragments having an epitopic specificity which is not the same as or similar 5 to 1D9. Antibodies of the present invention can be raised against an appropriate immunogen, such as isolated and/or recombinant mammalian CCR2 protein or portion thereof, or synthetic molecules, such as synthetic peptides. In a preferred embodiment, cells 10 which express receptor, such as transfected cells, can be used as immunogens or in a screen for antibody which binds receptor.

The antibodies of the present invention, and fragments thereof, are useful in therapeutic, diagnostic 15 and research applications as described herein. The present invention encompasses an antibody or functional portion thereof of the present invention (e.g., mAb 1D9 or 8G2, or antigen-binding fragments thereof) for use in therapy (including prophylaxis) or diagnosis (e.g., of 20 particular diseases or conditions as described herein), and use of such antibodies or functional portions thereof for the manufacture of a medicament for use in treatment of diseases or conditions as described herein.

Preparation of immunizing antigen, and polyclonal 25 and monoclonal antibody production can be performed as described herein, or using other suitable techniques. A variety of methods have been described (see e.g., Kohler et al., *Nature*, 256: 495-497 (1975) and *Eur. J. Immunol.* 6: 511-519 (1976); Milstein et al., *Nature* 266: 550-552 30 (1977); Koprowski et al., U.S. Patent No. 4,172,124; Harlow, E. and D. Lane, 1988, *Antibodies: A Laboratory Manual*, (Cold Spring Harbor Laboratory: Cold Spring Harbor, NY); *Current Protocols In Molecular Biology*, Vol. 2 (Supplement 27, Summer '94), Ausubel, F.M. et 35 al., Eds., (John Wiley & Sons: New York, NY), Chapter 11, (1991)). Generally, a hybridoma can be produced by

fusing a suitable immortal cell line (e.g., a myeloma cell line such as SP2/0) with antibody producing cells. The antibody producing cell, preferably those of the spleen or lymph nodes, are obtained from animals 5 immunized with the antigen of interest. The fused cells (hybridomas) can be isolated using selective culture conditions, and cloned by limiting dilution. Cells which produce antibodies with the desired binding properties can be selected by a suitable assay (e.g., 10 ELISA).

Other suitable methods of producing or isolating antibodies which bind CCR2, including human or artificial antibodies, can be used, including, for example, methods which select recombinant antibody 15 (e.g., single chain Fv or Fab) from a library, or which rely upon immunization of transgenic animals (e.g., mice) capable of producing a repertoire of human antibodies (see e.g., Jakobovits *et al.*, *Proc. Natl. Acad. Sci. USA*, 90: 2551-2555 (1993); Jakobovits *et al.*, 20 *Nature*, 362: 255-258 (1993); Lonberg *et al.*, U.S. Patent No. 5,545,806; Surani *et al.*, U.S. Patent No. 5,545,807).

Single chain antibodies, and chimeric, humanized or primatized (CDR-grafted) antibodies, as well as chimeric 25 or CDR-grafted single chain antibodies, and the like, comprising portions derived from different species, are also encompassed by the present invention and the term "antibody". The various portions of these antibodies can be joined together chemically by conventional 30 techniques, or can be prepared as a contiguous protein using genetic engineering techniques. For example, nucleic acids encoding a chimeric or humanized chain can be expressed to produce a contiguous protein. See, e.g., Cabilly *et al.*, U.S. Patent No. 4,816,567; Cabilly 35 *et al.*, European Patent No. 0,125,023 B1; Boss *et al.*, U.S. Patent No. 4,816,397; Boss *et al.*, European Patent

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No. 0,120,694 B1; Neuberger, M.S. et al., WO 86/01533; Neuberger, M.S. et al., European Patent No. 0,194,276 B1; Winter, U.S. Patent No. 5,225,539; Winter, European Patent No. 0,239,400 B1; and Queen et al., U.S. Patent 5 Nos. 5,585,089, 5,698,761 and 5,698,762. See also, Newman, R. et al., *BioTechnology*, 10: 1455-1460 (1992), regarding primatized antibody, and Ladner et al., U.S. Patent No. 4,946,778 and Bird, R.E. et al., *Science*, 242: 423-426 (1988) regarding single chain antibodies.

10 In addition, functional fragments of antibodies, including fragments of chimeric, humanized, primatized or single chain antibodies, can also be produced. Functional fragments of the foregoing antibodies retain at least one binding function and/or modulation function 15 of the full-length antibody from which they are derived. Preferred functional fragments retain an antigen-binding function of a corresponding full-length antibody (e.g., the ability to bind a mammalian CCR2). Particularly preferred functional fragments retain the ability to 20 inhibit one or more functions characteristic of a mammalian CCR2, such as a binding activity, a signaling activity, and/or stimulation of a cellular response. For example, in one embodiment, a functional fragment can inhibit the interaction of CCR2 with one or more of 25 its ligands (e.g., MCP-1, MCP-2, MCP-3 and/or MCP-4) and/or can inhibit one or more receptor-mediated functions, such as leukocyte trafficking, HIV entry into cells, T cell activation, inflammatory mediator release and/or leukocyte degranulation.

30 For example, antibody fragments capable of binding to a mammalian CCR2 receptor or portion thereof, including, but not limited to, Fv, Fab, Fab' and F(ab'), fragments are encompassed by the invention. Such fragments can be produced by enzymatic cleavage or by 35 recombinant techniques, for example. For instance, papain or pepsin cleavage can generate Fab or F(ab'),

fragments, respectively. Antibodies can also be produced in a variety of truncated forms using antibody genes in which one or more stop codons has been introduced upstream of the natural stop site. For 5 example, a chimeric gene encoding a F(ab'), heavy chain portion can be designed to include DNA sequences encoding the CH₁ domain and hinge region of the heavy chain.

The term "humanized immunoglobulin" as used herein 10 refers to an immunoglobulin comprising portions of immunoglobulins of different origin, wherein at least one portion is of human origin. Accordingly, the present invention relates to a humanized immunoglobulin which binds mammalian CCR2 (e.g., human CCR2, murine 15 CCR2), said immunoglobulin comprising an antigen-binding region of nonhuman origin (e.g., rodent) and at least a portion of an immunoglobulin of human origin (e.g., a human framework region, a human constant region or portion thereof). For example, the humanized antibody 20 can comprise portions derived from an immunoglobulin of nonhuman origin with the requisite specificity, such as a mouse, and from immunoglobulin sequences of human origin (e.g., a chimeric immunoglobulin), joined together chemically by conventional techniques (e.g., 25 synthetic) or prepared as a contiguous polypeptide using genetic engineering techniques (e.g., DNA encoding the protein portions of the chimeric antibody can be expressed to produce a contiguous polypeptide chain). Another example of a humanized immunoglobulin of the 30 present invention is an immunoglobulin containing one or more immunoglobulin chains comprising a CDR of nonhuman origin (e.g., one or more CDRs derived from an antibody of nonhuman origin) and a framework region derived from a light and/or heavy chain of human origin (e.g., 35 CDR-grafted antibodies with or without framework changes). In one embodiment, the humanized

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immunoglobulin can compete with murine 1D9 or 8G2 monoclonal antibody for binding to human CCR2. In a preferred embodiment, the antigen-binding region of the humanized immunoglobulin (a) is derived from 1D9 5 monoclonal antibody (e.g., as in a humanized immunoglobulin comprising CDR1, CDR2 and CDR3 of the 1D9 light chain and CDR1, CDR2 and CDR3 of the 1D9 heavy chain) or (b) is derived from 8G2 monoclonal antibody (e.g., as in a humanized immunoglobulin comprising CDR1, 10 CDR2 and CDR3 of the 8G2 light chain and CDR1, CDR2 and CDR3 of the 8G2 heavy chain). Chimeric or CDR-grafted single chain antibodies are also encompassed by the term humanized immunoglobulin.

Such humanized immunoglobulins can be produced 15 using synthetic and/or recombinant nucleic acids to prepare genes (e.g., cDNA) encoding the desired humanized chain. For example, nucleic acid (e.g., DNA) sequences coding for humanized variable regions can be constructed using PCR mutagenesis methods to alter DNA 20 sequences encoding a human or humanized chain, such as a DNA template from a previously humanized variable region (see e.g., Kamman, M., et al., *Nucl. Acids Res.*, 17: 5404 (1989); Sato, K., et al., *Cancer Research*, 53: 851-856 (1993); Daugherty, B.L. et al., *Nucleic Acids 25 Res.*, 19(9): 2471-2476 (1991); and Lewis, A.P. and J.S. Crowe, *Gene*, 101: 297-302 (1991)). Using these or other suitable methods, variants can also be readily produced. In one embodiment, cloned variable regions can be 30 mutagenized, and sequences encoding variants with the desired specificity can be selected (e.g., from a phage library; see e.g., Krebber et al., U.S. 5,514,548; Hoogenboom et al., WO 93/06213, published April 1, 1993)).

Anti-idiotypic antibodies are also provided. Anti- 35 idiotypic antibodies recognize antigenic determinants associated with the antigen-binding site of another

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antibody. Anti-idiotypic antibodies can be prepared against second antibody by immunizing an animal of the same species, and preferably of the same strain, as the animal used to produce the second antibody. See e.g., 5 U.S. Patent No. 4,699,880.

The present invention also pertains to the hybridoma cell lines deposited under ATCC Accession Nos. HB-12549 and HB-12550, as well as to the monoclonal antibodies produced by the hybridoma cell lines 10 deposited under ATCC Accession Nos. HB-12549 and HB-12550. The cell lines of the present invention have uses other than for the production of the monoclonal antibodies. For example, the cell lines of the present invention can be fused with other cells (such as 15 suitably drug-marked human myeloma, mouse myeloma, human-mouse heteromyeloma or human lymphoblastoid cells) to produce additional hybridomas, and thus provide for the transfer of the genes encoding the monoclonal antibodies. In addition, the cell lines can be used as 20 a source of nucleic acids encoding the anti-CCR2 immunoglobulin chains, which can be isolated and expressed (e.g., upon transfer to other cells using any suitable technique (see e.g., Cabilly et al., U.S. Patent No. 4,816,567; Winter, U.S. Patent No. 25 5,225,539)). For instance, clones comprising a rearranged anti-CCR2 light or heavy chain can be isolated (e.g., by PCR) or cDNA libraries can be prepared from mRNA isolated from the cell lines, and cDNA clones encoding an anti-CCR2 immunoglobulin chain 30 can be isolated. Thus, nucleic acids encoding the heavy and/or light chains of the antibodies or portions thereof can be obtained and used in accordance with recombinant DNA techniques for the production of the specific immunoglobulin, immunoglobulin chain, or 35 variants thereof (e.g., humanized immunoglobulins) in a variety of host cells or in an *in vitro* translation

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system. For example, the nucleic acids, including cDNAs, or derivatives thereof encoding variants such as a humanized immunoglobulin or immunoglobulin chain, can be placed into suitable prokaryotic or eukaryotic 5 vectors (e.g., expression vectors) and introduced into a suitable host cell by an appropriate method (e.g., transformation, transfection, electroporation, infection), such that the nucleic acid is operably linked to one or more expression control elements (e.g., 10 in the vector or integrated into the host cell genome). For production, host cells can be maintained under conditions suitable for expression (e.g., in the presence of inducer, suitable media supplemented with appropriate salts, growth factors, antibiotic, 15 nutritional supplements, etc.), whereby the encoded polypeptide is produced. If desired, the encoded protein can be recovered and/or isolated (e.g., from the host cells, medium, milk). It will be appreciated that the method of production encompasses expression in a 20 host cell of a transgenic animal (see e.g., WO 92/03918, GenPharm International, published March 19, 1992).

As described herein, antibodies and functional fragments thereof of the present invention can block (inhibit) binding of a ligand to CCR2 and/or inhibit 25 function associated with binding of the ligand to the CCR2. As discussed below various methods can be used to assess inhibition of binding of a ligand to CCR2 and/or function associated with binding of the ligand to the receptor.

30 Binding Assays

As used herein "mammalian CCR2 protein" refers to naturally occurring or endogenous mammalian CCR2 proteins and to proteins having an amino acid sequence which is the same as that of a naturally occurring or 35 endogenous corresponding mammalian CCR2 protein (e.g.,

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recombinant proteins). Accordingly, as defined herein, the term includes mature receptor protein, polymorphic or allelic variants, and other isoforms of a mammalian CCR2 (e.g., produced by alternative splicing or other 5 cellular processes), and modified or unmodified forms of the foregoing (e.g., glycosylated, unglycosylated). Mammalian CCR2 proteins can be isolated and/or recombinant proteins (including synthetically produced proteins). Naturally occurring or endogenous mammalian 10 CCR2 proteins include wild type proteins such as mature CCR2, polymorphic or allelic variants and other isoforms which occur naturally in mammals (e.g., humans, non-human primates), such as the CCR2a and CCR2b forms of the receptor protein which are produced by 15 alternative splicing of the carboxy-terminus of the protein. Such proteins can be recovered or isolated from a source which naturally produces mammalian CCR2, for example. These proteins and mammalian CCR2 proteins having the same amino acid sequence as a naturally 20 occurring or endogenous corresponding mammalian CCR2, are referred to by the name of the corresponding mammal. For example, where the corresponding mammal is a human, the protein is designated as a human CCR2 protein (e.g., a recombinant human CCR2 produced in a suitable host 25 cell).

"Functional variants" of mammalian CCR2 proteins include functional fragments, functional mutant proteins, and/or functional fusion proteins (e.g., produced via mutagenesis and/or recombinant techniques).

30 Generally, fragments or portions of mammalian CCR2 proteins include those having a deletion (i.e., one or more deletions) of an amino acid (i.e., one or more amino acids) relative to the mature mammalian CCR2 protein (such as N-terminal, C-terminal or internal 35 deletions). Fragments or portions in which only contiguous amino acids have been deleted or in which

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non-contiguous amino acids have been deleted relative to mature mammalian CCR2 protein are also envisioned.

Generally, mutants of mammalian CCR2 proteins include natural or artificial variants of a mammalian CCR2 protein differing by the addition, deletion and/or substitution of one or more contiguous or non-contiguous amino acid residues (e.g., receptor chimeras). Such mutations can be in a conserved region or nonconserved region (compared to other CXC and/or CC chemokine receptors), extracellular, cytoplasmic, or transmembrane region, for example.

Generally, fusion proteins encompass polypeptides comprising a mammalian CCR2 (e.g., human CCR2) as a first moiety, linked via a peptide bond to a second moiety not occurring in the mammalian CCR2 as found in nature. Thus, the second moiety can be an amino acid, oligopeptide or polypeptide. The first moiety can be in an N-terminal location, C-terminal location or internal to the fusion protein. In one embodiment, the fusion protein comprises an affinity ligand (e.g., an enzyme, an antigen, epitope tag) as the first moiety, and a second moiety comprising a linker sequence and human CCR2 or a portion thereof.

A "functional fragment or portion", "functional mutant" and/or "functional fusion protein" of a mammalian CCR2 protein refers to an isolated and/or recombinant protein or polypeptide which has at least one function characteristic of a mammalian CCR2 protein as described herein, such as a binding activity, a signaling activity and/or ability to stimulate a cellular response. Preferred functional variants can bind a ligand (i.e., one or more ligands such as MCP-1, MCP-2, MCP-3 and/or MCP-4), and are referred to herein as "ligand binding variants".

In one embodiment, a functional variant of mammalian CCR2 shares at least about 85% sequence

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identity with said mammalian CCR2, preferably at least about 90% sequence identity, and more preferably at least about 95% sequence identity with said mammalian CCR2. The nucleic acid and amino acid sequences of 5 human CCR2a and CCR2b are described in U.S. Patent No. 5,707,815. Sequence identity can be determined using a suitable program, such as the Blastx program (Version 1.4), using appropriate parameters, such as default parameters. In one embodiment, parameters for Blastx 10 search are scoring matrix BLOSUM62, W=3. In another embodiment, a functional variant comprises a nucleic acid sequence which is different from the naturally-occurring nucleic acid molecule but which, due to the degeneracy of the genetic code, encodes mammalian CCR2 15 or a portion thereof.

A composition comprising an isolated and/or recombinant mammalian CCR2 or functional variant thereof can be maintained under conditions suitable for binding, the mammalian CCR2 or variant is contacted with an 20 antibody or fragment to be tested, and binding is detected or measured directly or indirectly. In one embodiment, cells which naturally express CCR2 or cells comprising a recombinant nucleic acid sequence which encodes a mammalian CCR2 or variant thereof are used. 25 The cells are maintained under conditions appropriate for expression of receptor. The cells are contacted with an antibody or fragment under conditions suitable for binding (e.g., in a suitable binding buffer), and binding is detected by standard techniques. To 30 determine binding, the extent of binding can be determined relative to a suitable control (e.g., compared with background determined in the absence of antibody, compared with binding of a second antibody (i.e., a standard), compared with binding of antibody to 35 untransfected cells). A cellular fraction, such as a

membrane fraction, containing receptor or liposomes comprising receptor can be used in lieu of whole cells.

In one embodiment, the antibody is labeled with a suitable label (e.g., fluorescent label, isotope label, 5 antigen or epitope label, enzyme label), and binding is determined by detection of the label. In another embodiment, bound antibody can be detected by labeled second antibody. Specificity of binding can be assessed by competition or displacement, for example, using 10 unlabeled antibody or a ligand as competitor.

Binding inhibition assays can also be used to identify antibodies or fragments thereof which bind CCR2 and inhibit binding of another compound such as a ligand (e.g., MCP-1, MCP-2, MCP-3 and/or MCP-4) to CCR2 or a 15 functional variant. For example, a binding assay can be conducted in which a reduction in the binding of a ligand of CCR2 (in the presence of an antibody), as compared to binding of the ligand in the absence of the antibody, is detected or measured. A composition 20 comprising an isolated and/or recombinant mammalian CCR2 or functional variant thereof can be contacted with the ligand and antibody simultaneously, or one after the other, in either order. A reduction in the extent of binding of the ligand in the presence of the antibody, 25 is indicative of inhibition of binding by the antibody. For example, binding of the ligand could be decreased or abolished.

In one embodiment, direct inhibition of the binding 30 of a ligand (e.g., a chemokine such as MCP-1) to a mammalian CCR2 or variant thereof by an antibody or fragment is monitored. For example, the ability of an antibody to inhibit the binding of ¹²⁵I-labeled MCP-1, ¹²⁵I-labeled MCP-2, ¹²⁵I-labeled MCP-3 or ¹²⁵I-labeled MCP-4 to mammalian CCR2 can be monitored. Such an assay can 35 be conducted using suitable cells bearing CCR2 or a functional variant thereof, such as isolated blood cells

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(e.g., T cells, PBMC) or a suitable cell line naturally expressing CCR2, or a cell line containing nucleic acid encoding a mammalian CCR2, or a membrane fraction from said cells, for instance.

5 Other methods of identifying the presence of an antibody which binds CCR2 are available, such as other suitable binding assays, or methods which monitor events which are triggered by receptor binding, including signaling function and/or stimulation of a cellular response (e.g., leukocyte trafficking).

10 It will be understood that the inhibitory effect of antibodies of the present invention can be assessed in a binding inhibition assay. Competition between antibodies for receptor binding can also be assessed in 15 the method. Antibodies which are identified in this manner can be further assessed to determine whether, subsequent to binding, they act to inhibit other functions of CCR2 and/or to assess their therapeutic utility.

20 **Signaling Assays**

The binding of a ligand or promoter, such as an agonist, to CCR2 can result in signaling by this G protein-coupled receptor, and the activity of G proteins as well as other intracellular signaling molecules is 25 stimulated. The induction of signaling function by a compound (e.g., an antibody or fragment thereof) can be monitored using any suitable method. Such an assay can be used to identify antibody agonists of CCR2. The inhibitory activity of an antibody or functional 30 fragment thereof can be determined using a ligand or promoter in the assay, and assessing the ability of the antibody to inhibit the activity induced by ligand or promoter.

35 G protein activity, such as hydrolysis of GTP to GDP, or later signaling events triggered by receptor

binding, such as induction of rapid and transient increase in the concentration of intracellular (cytosolic) free calcium $[Ca^{2+}]_i$, can be assayed by methods known in the art or other suitable methods (see 5 e.g., Neote, K. et al., *Cell*, 72: 415-425 (1993); Van Riper et al., *J. Exp. Med.*, 177: 851-856 (1993); Dahinden, C.A. et al., *J. Exp. Med.*, 179: 751-756 (1994)).

For example, the functional assay of Sledziewski et 10 al. using hybrid G protein coupled receptors can be used to monitor the ability a ligand or promoter to bind receptor and activate a G protein (Sledziewski et al., U.S. Patent No. 5,284,746, the teachings of which are incorporated herein by reference).

15 Such assays can be performed in the presence of the antibody or fragment thereof to be assessed; and the ability of the antibody or fragment to inhibit the activity induced by the ligand or promoter is determined using known methods and/or methods described herein.

20 **Chemotaxis and Assays of Cellular Stimulation**

Chemotaxis assays can also be used to assess the 25 ability of an antibody or functional fragment thereof to block binding of a ligand to mammalian CCR2 or functional variant thereof and/or inhibit function associated with binding of the ligand to the receptor. These assays are based on the functional migration of 30 cells *in vitro* or *in vivo* induced by a compound. Chemotaxis can be assessed as described in the Examples, e.g., in an assay utilizing a 96-well chemotaxis plate, 35 or using other art-recognized methods for assessing chemotaxis. For example, the use of an *in vitro* transendothelial chemotaxis assay is described by Springer et al. (Springer et al., WO 94/20142, published September 15, 1994, the teachings of which are incorporated herein by reference; see also Berman et

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al., *Immunol. Invest.* 17: 625-677 (1988)). Migration across endothelium into collagen gels has also been described (Kavanaugh et al., *J. Immunol.*, 146: 4149-4156 (1991)). Stable transfectants of mouse L1-2 pre-B cells 5 or of other suitable host cells capable of chemotaxis can be used in chemotaxis assays, for example.

Generally, chemotaxis assays monitor the directional movement or migration of a suitable cell (such as a leukocyte (e.g., lymphocyte, eosinophil, 10 basophil)) into or through a barrier (e.g., endothelium, a filter), toward increased levels of a compound, from a first surface of the barrier toward an opposite second surface. Membranes or filters provide convenient barriers, such that the directional movement or 15 migration of a suitable cell into or through a filter, toward increased levels of a compound, from a first surface of the filter toward an opposite second surface of the filter, is monitored. In some assays, the membrane is coated with a substance to facilitate 20 adhesion, such as ICAM-1, fibronectin or collagen. Such assays provide an *in vitro* approximation of leukocyte "homing".

For example, one can detect or measure inhibition 25 of the migration of cells in a suitable container (a containing means), from a first chamber into or through a microporous membrane into a second chamber which contains an antibody to be tested, and which is divided from the first chamber by the membrane. A suitable membrane, having a suitable pore size for monitoring 30 specific migration in response to compound, including, for example, nitrocellulose, polycarbonate, is selected. For example, pore sizes of about 3-8 microns, and preferably about 5-8 microns can be used. Pore size can be uniform on a filter or within a range of suitable 35 pore sizes.

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To assess migration and inhibition of migration, the distance of migration into the filter, the number of cells crossing the filter that remain adherent to the second surface of the filter, and/or the number of cells 5 that accumulate in the second chamber can be determined using standard techniques (e.g., microscopy). In one embodiment, the cells are labeled with a detectable label (e.g., radioisotope, fluorescent label, antigen or epitope label), and migration can be assessed in the 10 presence and absence of the antibody or fragment by determining the presence of the label adherent to the membrane and/or present in the second chamber using an appropriate method (e.g., by detecting radioactivity, fluorescence, immunoassay). The extent of migration 15 induced by an antibody agonist can be determined relative to a suitable control (e.g., compared to background migration determined in the absence of the antibody, compared to the extent of migration induced by a second compound (i.e., a standard), compared with 20 migration of untransfected cells induced by the antibody).

In one embodiment, particularly for T cells, monocytes or cells expressing a mammalian CCR2, transendothelial migration can be monitored. In this 25 embodiment, transmigration through an endothelial cell layer is assessed. To prepare the cell layer, endothelial cells can be cultured on a microporous filter or membrane, optionally coated with a substance such as collagen, fibronectin, or other extracellular 30 matrix proteins, to facilitate the attachment of endothelial cells. Preferably, endothelial cells are cultured until a confluent monolayer is formed. A variety of mammalian endothelial cells can be available for monolayer formation, including for example, vein, 35 artery or microvascular endothelium, such as human umbilical vein endothelial cells (Clonetics Corp, San

Diego, CA). To assay chemotaxis in response to a particular mammalian receptor, endothelial cells of the same mammal are preferred; however endothelial cells from a heterologous mammalian species or genus can also 5 be used.

Generally, the assay is performed by detecting the directional migration of cells into or through a membrane or filter, in a direction toward increased levels of a compound, from a first surface of the filter 10 toward an opposite second surface of the filter, wherein the filter contains an endothelial cell layer on a first surface. Directional migration occurs from the area adjacent to the first surface, into or through the membrane, towards a compound situated on the opposite 15 side of the filter. The concentration of compound present in the area adjacent to the second surface, is greater than that in the area adjacent to the first surface.

In one embodiment used to test for an antibody 20 inhibitor, a composition comprising cells capable of migration and expressing a mammalian CCR2 receptor can be placed in the first chamber. A composition comprising one or more ligands or promoters capable of inducing chemotaxis of the cells in the first chamber 25 (having chemoattractant function) is placed in the second chamber. Preferably shortly before the cells are placed in the first chamber, or simultaneously with the cells, a composition comprising the antibody to be tested is placed, preferably, in the first chamber. 30 Antibodies or functional fragments thereof which can bind receptor and inhibit the induction of chemotaxis, by a ligand or promoter, of the cells expressing a mammalian CCR2 in this assay are inhibitors of receptor function (e.g., inhibitors of stimulatory function). A 35 reduction in the extent of migration induced by the ligand or promoter in the presence of the antibody or

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fragment is indicative of inhibitory activity. Separate binding studies (see above) could be performed to determine whether inhibition is a result of binding of the antibody to receptor or occurs via a different

5 mechanism.

In vivo assays which monitor leukocyte infiltration of a tissue, in response to injection of a compound (e.g., chemokine or antibody) in the tissue, are described below (see Models of Inflammation). These

10 models of *in vivo* homing measure the ability of cells to respond to a ligand or promoter by emigration and chemotaxis to a site of inflammation and to assess the ability of an antibody or fragment thereof to block this emigration.

15 In addition to the methods described, the effects of an antibody or fragment on the stimulatory function of CCR2 can be assessed by monitoring cellular responses induced by active receptor, using suitable host cells containing receptor.

20 **Identification of Additional Ligands, Inhibitors and/or Promoters of Mammalian CCR2 Function**

The assays described above, which can be used to assess binding and function of the antibodies and fragments of the present invention, can be adapted to

25 identify additional ligands or other substances which bind a mammalian CCR2 or functional variant thereof, as well as inhibitors and/or promoters of mammalian CCR2 function. For example, agents having the same or a similar binding specificity as that of an antibody of

30 the present invention or functional portion thereof can be identified by a competition assay with said antibody or portion thereof. Thus, the present invention also encompasses methods of identifying ligands of the receptor or other substances which bind a mammalian CCR2

35 protein, as well as inhibitors (e.g., antagonists) or

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- promoters (e.g., agonists) of receptor function. In one embodiment, cells bearing a mammalian CCR2 protein or functional variant thereof (e.g., leukocytes, cell lines or suitable host cells which have been engineered to
- 5 express a mammalian CCR2 protein or functional variant encoded by a nucleic acid introduced into said cells) are used in an assay to identify and assess the efficacy of ligands or other substances which bind receptor, including inhibitors or promoters of receptor function.
- 10 Such cells are also useful in assessing the function of the expressed receptor protein or polypeptide.
- According to the present invention, ligands and other substances which bind receptor, inhibitors and promoters of receptor function can be identified in a
- 15 suitable assay, and further assessed for therapeutic effect. Inhibitors of receptor function can be used to inhibit (reduce or prevent) receptor activity, and ligands and/or promoters can be used to induce (trigger or enhance) normal receptor function where indicated.
- 20 Thus, the present invention provides a method of treating inflammatory diseases, including autoimmune disease and graft rejection, comprising administering an inhibitor of receptor function to an individual (e.g., a mammal). The present invention further provides a
- 25 method of stimulating receptor function by administering a novel ligand or promoter of receptor function to an individual, providing a new approach to selective stimulation of leukocyte function, which is useful, for example, in the treatment of infectious diseases and
- 30 cancer.

As used herein, a "ligand" of a mammalian CCR2 protein refers to a particular class of substances which bind to a mammalian CCR2 protein, including natural ligands and synthetic and/or recombinant forms of

35 natural ligands. Infectious agents having a tropism for mammalian CCR2-positive cells (e.g., viruses such as

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HIV) can also bind to a mammalian CCR2 protein. A natural ligand of a selected mammalian receptor is of a mammalian origin which is the same as that of the mammalian CCR2 protein (e.g., a chemokine such as MCP-1, 5 MCP-2, MCP-3 and/or MCP-4). In a preferred embodiment, ligand binding of a mammalian CCR2 protein occurs with high affinity.

As used herein, an "inhibitor" is a substance which inhibits (decreases or prevents) at least one function 10 characteristic of a mammalian CCR2 protein (e.g., a human CCR2), such as a binding activity (e.g., ligand binding, promoter binding, antibody binding), a signaling activity (e.g., activation of a mammalian G protein, induction of rapid and transient increase in 15 the concentration of cytosolic free calcium $[Ca^{2+}]_i$), and/or cellular response function (e.g., stimulation of chemotaxis, exocytosis or inflammatory mediator release by leukocytes). An inhibitor is also a substance which inhibits HIV entry into a cell. The term inhibitor 20 refers to substances including antagonists which bind receptor (e.g., an antibody, a mutant of a natural ligand, small molecular weight organic molecules, other competitive inhibitors of ligand binding), and substances which inhibit receptor function without 25 binding thereto (e.g., an anti-idiotypic antibody).

As used herein, a "promoter" is a substance which promotes (induces, causes, enhances or increases) at least one function characteristic of a mammalian CCR2 protein (e.g., a human CCR2), such as a binding activity 30 (e.g., ligand, inhibitor and/or promoter binding), a signaling activity (e.g., activation of a mammalian G protein, induction of rapid and transient increase in the concentration of cytosolic free calcium $[Ca^{2+}]_i$), and/or a cellular response function (e.g., stimulation 35 of chemotaxis, exocytosis or inflammatory mediator release by leukocytes). The term promoter refers to

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substances including agonists which bind receptor (e.g., an antibody, a homolog of a natural ligand from another species), and substances which promote receptor function without binding thereto (e.g., by activating an 5 associated protein). In a preferred embodiment, the agonist is other than a homolog of a natural ligand.

Thus, the invention also relates to a method of detecting or identifying an agent which binds a mammalian CC-chemokine receptor 2 or ligand binding 10 variant thereof, including ligands, inhibitors, promoters, and other substances which bind a mammalian CCR2 receptor or functional variant. According to the method, an agent to be tested, an antibody or antigen-binding fragment of the present invention (e.g., 8G2, 15 1D9, an antibody having an epitopic specificity which is the same as or similar to that of 8G2 or 1D9, and antigen-binding fragments thereof) and a composition comprising a mammalian CC-chemokine receptor 2 or a ligand binding variant thereof can be combined. The 20 foregoing components are combined under conditions suitable for binding of the antibody or antigen-binding fragment to mammalian CC-chemokine receptor 2 or a ligand binding variant thereof, and binding of the antibody or fragment to the mammalian CC-chemokine 25 receptor 2 or ligand binding variant is detected or measured, either directly or indirectly, according to methods described herein or other suitable methods. A decrease in the amount of complex formed relative to a suitable control (e.g., in the absence of the agent to 30 be tested) is indicative that the agent binds said receptor or variant. The composition comprising a mammalian CC-chemokine receptor 2 or a ligand binding variant thereof can be a membrane fraction of a cell bearing recombinant chemokine receptor 2 protein or 35 ligand binding variant thereof. The antibody or fragment thereof can be labeled with a label such as a

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radioisotope, spin label, antigen or epitope label, enzyme label, fluorescent group and chemiluminescent group.

In one embodiment, the invention relates to a
5 method of detecting or identifying an agent which binds a mammalian CC-chemokine receptor 2 or a ligand binding variant thereof, comprising combining an agent to be tested, an antibody or antigen-binding fragment of the present invention (e.g., 1D9, 8G2, an antibody having an
10 epitopic specificity which is the same as or similar to that of 1D9 or 8G2, or antigen-binding fragments thereof) and a cell bearing a mammalian CC-chemokine receptor 2 or a ligand binding variant thereof. The foregoing components are combined under conditions
15 suitable for binding of the antibody or antigen-binding fragment to the CCR2 protein or ligand binding variant thereof, and binding of the antibody or fragment to the mammalian CC-chemokine receptor 2 or variant is detected or measured, either directly or indirectly, by methods
20 described herein and or other suitable methods. A decrease in the amount of complex formed relative to a suitable control is indicative that the agent binds the receptor or variant. The antibody or fragment thereof can be labeled with a label selected from the group
25 consisting of a radioisotope, spin label, antigen or epitope label, enzyme label, fluorescent group and chemiluminescent group. These and similar assays can be used to detect agents, including ligands (e.g., chemokines or strains of HIV which interact with CCR2)
30 or other substances, including inhibitors or promoters of receptor function, which can bind CCR2 and compete with the antibodies described herein for binding to the receptor.

The assays described above can be used, alone or in
35 combination with each other or other suitable methods, to identify ligands or other substances which bind a

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mammalian CCR2 protein, and inhibitors or promoters of a mammalian CCR2 protein or variant. The *in vitro* methods of the present invention can be adapted for high-throughput screening in which large numbers of samples

5 are processed (e.g., a 96-well format). Cells expressing mammalian CCR2 (e.g., human CCR2) at levels suitable for high-throughput screening can be used, and thus, are particularly valuable in the identification and/or isolation of ligands or other substances which

10 bind receptor, and inhibitors or promoters of mammalian CCR2 proteins. Expression of receptor can be monitored in a variety of ways. For instance, expression can be monitored using antibodies of the present invention which bind receptor or a portion thereof. Also,

15 commercially available antibodies can be used to detect expression of an antigen- or epitope-tagged fusion protein comprising a receptor protein or polypeptide (e.g., FLAG tagged receptors), and cells expressing the desired level can be selected.

20 Nucleic acid encoding a mammalian CCR2 protein or functional variant thereof can be incorporated into an expression system to produce a receptor protein or polypeptide. An isolated and/or recombinant mammalian CCR2 protein or variant, such as a receptor expressed in

25 cells stably or transiently transfected with a construct comprising a recombinant nucleic acid encoding a mammalian CCR2 protein or variant, or in a cell fraction containing receptor (e.g., a membrane fraction from transfected cells, liposomes incorporating receptor),

30 can be used in tests for receptor function. The receptor can be further purified if desired. Testing of receptor function can be carried out *in vitro* or *in vivo*.

An isolated and/or recombinant mammalian CCR2

35 protein or functional variant thereof, such as a human CCR2, can be used in the present method, in which the

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- effect of a compound is assessed by monitoring receptor function as described herein or using other suitable techniques. For example, stable or transient transfectants (e.g., baculovirus infected Sf9 cells, 5 stable transfectants of mouse L1/2 pre-B cells), can be used in binding assays. Stable transfectants of Jurkat cells or of other suitable cells capable of chemotaxis can be used (e.g., mouse L1/2 pre-B cells) in chemotaxis assays, for example.
- 10 According to the method of the present invention, compounds can be individually screened or one or more compounds can be tested simultaneously according to the methods herein. Where a mixture of compounds is tested, the compounds selected by the processes described can be 15 separated (as appropriate) and identified by suitable methods (e.g., PCR, sequencing, chromatography, mass spectroscopy). The presence of one or more compounds (e.g., a ligand, inhibitor, promoter) in a test sample can also be determined according to these methods.
- 20 Large combinatorial libraries of compounds (e.g., organic compounds, recombinant or synthetic peptides, "peptoids", nucleic acids) produced by combinatorial chemical synthesis or other methods can be tested (see e.g., Zuckerman, R.N. et al., *J. Med. Chem.*, 37: 2678- 25 2685 (1994) and references cited therein; see also, Ohlmeyer, M.H.J. et al., *Proc. Natl. Acad. Sci. USA* 90:10922-10926 (1993) and DeWitt, S.H. et al., *Proc. Natl. Acad. Sci. USA* 90:6909-6913 (1993), relating to tagged compounds; Rutter, W.J. et al. U.S. Patent No. 30 5,010,175; Huebner, V.D. et al., U.S. Patent No. 5,182,366; and Geysen, H.M., U.S. Patent No. 4,833,092). Where compounds selected from a combinatorial library by the present method carry unique tags, identification of individual compounds by chromatographic methods is 35 possible.

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In one embodiment, phage display methodology is used. For example, a mammalian CCR2 protein or functional variant, an antibody or functional portion thereof of the present invention, and a phage (e.g., a 5 phage or collection of phage such as a library) displaying a polypeptide, can be combined under conditions appropriate for binding of the antibody or portion thereof to the mammalian CCR2 protein or variant (e.g., in a suitable binding buffer). Phage which can 10 compete with the antibody or portion thereof and bind to the mammalian CCR2 protein or variant can be detected or selected using standard techniques or other suitable methods. Bound phage can be separated from receptor using a suitable elution buffer. For example, a change 15 in the ionic strength or pH can lead to a release of phage. Alternatively, the elution buffer can comprise a release component or components designed to disrupt binding of compounds (e.g., one or more compounds which can disrupt binding of the displayed peptide to the 20 receptor, such as a ligand, inhibitor, and/or promoter which competitively inhibits binding). Optionally, the selection process can be repeated or another selection step can be used to further enrich for phage which bind receptor. The displayed polypeptide can be 25 characterized (e.g., by sequencing phage DNA). The polypeptides identified can be produced and further tested for binding, and for inhibitor or promoter function. Analogs of such peptides can be produced which will have increased stability or other desirable 30 properties.

In one embodiment, phage expressing and displaying fusion proteins comprising a coat protein with an N-terminal peptide encoded by random sequence nucleic acids can be produced. Suitable host cells expressing a 35 mammalian CCR2 protein or variant and an anti-CCR2 antibody or functional portion thereof, are combined

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with the phage, bound phage are selected, recovered and characterized. (See e.g., Doorbar, J. and G. Winter, *J. Mol. Biol.*, 244: 361 (1994) discussing a phage display procedure used with a G protein-coupled receptor).

5 Other sources of potential ligands or other substances which bind to, or inhibitors and/or promoters of, mammalian CCR2 proteins include, but are not limited to, variants of CCR2 ligands, including naturally occurring, synthetic or recombinant variants of MCP-1, 10 MCP-2, MCP-3 and/or MCP-4, substances such as other chemoattractants or chemokines, variants thereof, low molecular weight organic molecules, other inhibitors and/or promoters (e.g., anti-CCR2 antibodies, antagonists, agonists), other G protein-coupled receptor 15 ligands, inhibitors and/or promoters (e.g., antagonists or agonists), and soluble portions of a mammalian CCR2 receptor, such as a suitable receptor peptide or analog which can inhibit receptor function (see e.g., Murphy, R.B., WO 94/05695).

20 **Models of Inflammation**

In vivo models of inflammation are available which can be used to assess the effects of antibodies and fragments of the invention in vivo as therapeutic agents. For example, leukocyte infiltration upon 25 intradermal injection of a chemokine and an antibody or fragment thereof reactive with mammalian CCR2 into a suitable animal, such as rabbit, mouse, rat, guinea pig or rhesus macaque can be monitored (see e.g., Van Damme, J. et al., *J. Exp. Med.*, 176: 59-65 (1992); Zachariae, 30 C.O.C. et al., *J. Exp. Med.* 171: 2177-2182 (1990); Jose, P.J. et al., *J. Exp. Med.* 179: 881-887 (1994)). In one embodiment, skin biopsies are assessed histologically for infiltration of leukocytes (e.g., eosinophils, 35 granulocytes). In another embodiment, labeled cells (e.g., stably transfected cells expressing a mammalian

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- CCR2, labeled with ^{111}In for example) capable of chemotaxis and extravasation are administered to the animal. For example, an antibody or fragment to be assessed can be administered, either before,
5 simultaneously with or after ligand or agonist is administered to the test animal. A decrease of the extent of infiltration in the presence of antibody as compared with the extent of infiltration in the absence of inhibitor is indicative of inhibition.

10 **Diagnostic and Therapeutic Applications**

- The antibodies and fragments of the present invention are useful in a variety of applications, including research, diagnostic and therapeutic applications. In one embodiment, the antibodies are
15 labeled with a suitable label (e.g., fluorescent label, chemiluminescent label, isotope label, antigen or epitope label or enzyme label). For instance, they can be used to isolate and/or purify receptor or portions thereof, and to study receptor structure (e.g.,
20 conformation) and function.

In addition, the various antibodies of the present invention can be used to detect CCR2 or to measure the expression of receptor, for example, on T cells (e.g., CD8+ cells, CD45RO+ cells), monocytes and/or on cells
25 transfected with a receptor gene. Thus, they also have utility in applications such as cell sorting (e.g., flow cytometry, fluorescence activated cell sorting), for diagnostic or research purposes.

The anti-CCR2 antibodies of the present invention
30 have value in diagnostic applications. An anti-CCR2 antibody or fragment thereof can be used to monitor expression of this receptor in HIV infected individuals, similar to the way anti-CD4 has been used as a diagnostic indicator of disease stage.

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Typically, diagnostic assays entail detecting the formation of a complex resulting from the binding of an antibody or fragment thereof to CCR2. For diagnostic purposes, the antibodies or antigen-binding fragments 5 can be labeled or unlabeled. The antibodies or fragments can be directly labeled. A variety of labels can be employed, including, but not limited to, radionuclides, fluorescers, enzymes, enzyme substrates, enzyme cofactors, enzyme inhibitors and ligands (e.g., 10 biotin, haptens). Numerous appropriate immunoassays are known to the skilled artisan (see, for example, U.S. Patent Nos. 3,817,827; 3,850,752; 3,901,654 and 4,098,876). When unlabeled, the antibodies or fragments can be detected using suitable means, as in 15 agglutination assays, for example. Unlabeled antibodies or fragments can also be used in combination with another (i.e., one or more) suitable reagent which can be used to detect antibody, such as a labeled antibody (e.g., a second antibody) reactive with the first 20 antibody (e.g., anti-idiotype antibodies or other antibodies that are specific for the unlabeled immunoglobulin) or other suitable reagent (e.g., labeled protein A).

In one embodiment, the antibodies or fragments of 25 the present invention can be utilized in enzyme immunoassays, wherein the subject antibody or fragment, or second antibodies, are conjugated to an enzyme. When a biological sample comprising a mammalian CCR2 protein is combined with the subject antibodies, binding occurs 30 between the antibodies and CCR2 protein. In one embodiment, a sample containing cells expressing a mammalian CCR2 protein, such as human blood, is combined with the subject antibodies, and binding occurs between the antibodies and cells bearing a human CCR2 protein 35 comprising an epitope recognized by the antibody. These bound cells can be separated from unbound reagents and

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the presence of the antibody-enzyme conjugate specifically bound to the cells can be determined, for example, by contacting the sample with a substrate of the enzyme which produces a color or other detectable 5 change when acted on by the enzyme. In another embodiment, the subject antibodies can be unlabeled, and a second, labeled antibody can be added which recognizes the subject antibody.

Kits for use in detecting the presence of a 10 mammalian CCR2 protein in a biological sample can also be prepared. Such kits will include an antibody or functional fragment thereof which binds to a mammalian CC-chemokine receptor 2 or portion of said receptor, as well as one or more ancillary reagents suitable for 15 detecting the presence of a complex between the antibody or fragment and CCR2 or portion thereof. The antibody compositions of the present invention can be provided in lyophilized form, either alone or in combination with additional antibodies specific for other epitopes. The 20 antibodies, which can be labeled or unlabeled, can be included in the kits with adjunct ingredients (e.g., buffers, such as Tris, phosphate and carbonate, stabilizers, excipients, biocides and/or inert proteins, e.g., bovine serum albumin). For example, the 25 antibodies can be provided as a lyophilized mixture with the adjunct ingredients, or the adjunct ingredients can be separately provided for combination by the user. Generally these adjunct materials will be present in less than about 5% weight based on the amount of active 30 antibody, and usually will be present in a total amount of at least about 0.001% weight based on antibody concentration. Where a second antibody capable of binding to the monoclonal antibody is employed, such antibody can be provided in the kit, for instance in a 35 separate vial or container. The second antibody, if present, is typically labeled, and can be formulated in

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an analogous manner with the antibody formulations described above.

Similarly, the present invention also relates to a method of detecting and/or quantitating expression of a 5 mammalian CCR2 or a portion of the receptor by a cell, in which a composition comprising a cell or fraction thereof (e.g., membrane fraction) is contacted with an antibody or functional fragment thereof (e.g., 1D9 and/or 8G2) which binds to a mammalian CCR2 or portion 10 of the receptor under conditions appropriate for binding of the antibody or fragment thereto, and binding is monitored. Detection of the antibody, indicative of the formation of a complex between antibody and CCR2 or a portion thereof, indicates the presence of the receptor. 15 Binding of antibody to the cell can be determined as described above under the heading "Binding Assays", for example. The method can be used to detect expression of CCR2 on cells from an individual (e.g., in a sample, such as a body fluid, such as blood, saliva or other 20 suitable sample). The level of expression of CCR2 on the surface of T cells or monocytes can also be determined, for instance, by flow cytometry, and the level of expression (e.g., staining intensity) can be correlated with disease susceptibility, progression or 25 risk.

Chemokine receptors function in the migration of leukocytes throughout the body, particularly to inflammatory sites. Inflammatory cell emigration from the vasculature is regulated by a three-step process 30 involving interactions of leukocyte and endothelial cell adhesion proteins and cell specific chemoattractants and activating factors (Springer, T.A., *Cell*, 76:301-314 (1994); Butcher, E.C., *Cell*, 67:1033-1036 (1991); Butcher, E.C. and Picker, L.J., *Science (Wash. DC)*, 35 272:60-66 (1996)). These are: (a) a low affinity interaction between leukocyte selectins and endothelial

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cell carbohydrates; (b) a high-affinity interaction between leukocyte chemoattractant receptors and chemoattractant/activating factors; and (c) a tight-binding between leukocyte integrins and endothelial cell adhesion proteins of the immunoglobulin superfamily. 5 Different leukocyte subsets express different repertoires of selectins, chemoattractant receptors and integrins. Additionally, inflammation alters the expression of endothelial adhesion proteins and the 10 expression of chemoattractant and leukocyte activating factors. As a consequence, there is a great deal of diversity for regulating the selectivity of leukocyte recruitment to extravascular sites. The second step is crucial in that the activation of the leukocyte 15 chemoattractant receptors is thought to cause the transition from the selectin-mediated cell rolling to the integrin-mediated tight binding. This results in the leukocyte being ready to transmigrate to perivascular sites. The chemoattractant/chemoattractant 20 receptor interaction is also crucial for transendothelial migration and localization within a tissue (Campbell, J.J., et al., *J. Cell Biol.*, 134:255-266 (1996); Carr, M.W., et al., *Immunity*, 4:179-187 (1996)). This migration is directed by a concentration 25 gradient of chemoattractant leading towards the inflammatory focus.

CCR2 has an important role in leukocyte trafficking. It is likely that CCR2 is a key chemokine receptor for T cell or T cell subset or monocyte 30 migration to certain inflammatory sites, and so anti-CCR2 mAbs can be used to inhibit (reduce or prevent) T cell or monocyte migration, particularly that associated with T cell dysfunction, such as autoimmune disease, or allergic reactions or with monocyte-mediated disorders 35 such as atherosclerosis. Accordingly, the antibodies and fragments thereof of the present invention can also

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be used to modulate receptor function in research and therapeutic applications. For instance, the antibodies and functional fragments described herein can act as inhibitors to inhibit (reduce or prevent) (a) binding 5 (e.g., of a ligand, an inhibitor or a promoter) to the receptor, (b) a receptor signaling function, and/or (c) a stimulatory function. Antibodies which act as inhibitors of receptor function can block ligand or promoter binding directly or indirectly (e.g., by 10 causing a conformational change). For example, antibodies can inhibit receptor function by inhibiting binding of a ligand, or by desensitization (with or without inhibition of binding of a ligand). Antibodies which bind receptor can also act as agonists of receptor 15 function, triggering or stimulating a receptor function, such as a signaling and/or a stimulatory function of a receptor (e.g., leukocyte trafficking) upon binding to receptor.

Thus, the present invention provides a method of 20 inhibiting leukocyte trafficking in a mammal (e.g., a human patient), comprising administering to the mammal an effective amount of an antibody or functional fragment of the present invention. Administration of an antibody or fragment of the present invention can result 25 in amelioration or elimination of the disease state.

The antibody of the present invention, or a functional fragment thereof, can also be used to treat disorders in which activation of the CCR2 receptor by binding of chemokines is implicated. For example, the 30 antibodies or functional fragments thereof (e.g., 1D9 and/or 8G2 or functional fragments thereof) can be used to treat allergy, atherogenesis, anaphylaxis, malignancy, chronic and acute inflammation, histamine and IgE-mediated allergic reactions, shock, and 35 rheumatoid arthritis, atherosclerosis, multiple sclerosis, allograft rejection, fibrotic disease, asthma, and inflammatory glomerulopathies.

AMENDED SHEET

Diseases or conditions of humans or other species which can be treated with inhibitors of CCR2 receptor function (including antibodies or suitable fragments thereof), include, but are not limited to:

- 5 • inflammatory or allergic diseases and conditions, including respiratory allergic diseases such as asthma, allergic rhinitis, hypersensitivity lung diseases, hypersensitivity pneumonitis, interstitial lung diseases (ILD) (e.g., idiopathic pulmonary fibrosis, or ILD
- 10 10 associated with rheumatoid arthritis, systemic lupus erythematosus, ankylosing spondylitis, systemic sclerosis, Sjogren's syndrome, polymyositis or dermatomyositis); anaphylaxis or hypersensitivity responses, drug allergies (e.g., to penicillin, 15 cephalosporins), insect sting allergies; inflammatory bowel diseases, such as Crohn's disease and ulcerative colitis; spondyloarthropathies; scleroderma; psoriasis and inflammatory dermatoses such as dermatitis, eczema, atopic dermatitis, allergic contact dermatitis,
- 20 20 urticaria; vasculitis (e.g., necrotizing, cutaneous, and hypersensitivity vasculitis);
 - autoimmune diseases, such as arthritis (e.g., rheumatoid arthritis, psoriatic arthritis), multiple sclerosis, systemic lupus erythematosus, myasthenia 25 gravis, juvenile onset diabetes, nephritides such as glomerulonephritis, autoimmune thyroiditis, Behcet's disease;
 - graft rejection (e.g., in transplantation), including allograft rejection or graft-versus-host
- 30 30 disease, and organ transplant-associated arteriosclerosis;
 - atherosclerosis;
 - cancers with leukocyte infiltration of the skin or organs;
- 35 35 • stenosis or restenosis of the vasculature, particularly of the arteries, e.g., the coronary artery,

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such as stenosis or restenosis which results from vascular intervention (e.g., surgical, therapeutic or mechanical intervention), as well as neointimal hyperplasia. For example, restenosis, which typically 5 produces a narrowing of the luminal opening of the vessel, can result from vascular injury including, but not limited to, that produced by vascular graft procedures, angioplasty, including angioplasty performed by balloon, atherectomy, laser or other suitable method 10 (e.g., percutaneous transluminal coronary angioplasty (PTCA)), stent placement (e.g., mechanical or biological endovascular stent placement), vascular bypass procedures or combinations thereof, as well as other procedures used to treat stenotic or occluded blood 15 vessels;

• other diseases or conditions (including CCR2-mediated diseases or conditions), in which undesirable inflammatory responses are to be inhibited can be treated, including, but not limited to, reperfusion 20 injury, certain hematologic malignancies, cytokine-induced toxicity (e.g., septic shock, endotoxic shock), polymyositis, dermatomyositis, and granulomatous diseases including sarcoidosis.

Diseases or conditions of humans or other species 25 which can be treated with promoters of CCR2 receptor function (including antibodies or fragments thereof), include, but are not limited to:

• immunosuppression, such as that in individuals with immunodeficiency syndromes such as AIDS, 30 individuals undergoing radiation therapy, chemotherapy, therapy for autoimmune disease or other drug therapy (e.g., corticosteroid therapy), which causes immunosuppression; and immunosuppression due congenital deficiency in receptor function or other causes.

35 Anti-CCR2 antibodies of the present invention can block the binding of one or more chemokines, thereby

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blocking the downstream cascade of one or more events leading to the above disorders.

Antibodies and functional fragments thereof which are antagonists of CCR2 can be used as therapeutics for 5 AIDS, as well as certain inflammatory diseases. HIV-1 and HIV-2 are the etiologic agents of acquired immunodeficiency syndrome (AIDS) in humans. AIDS results in part from the depletion of CD4+ T lymphocytes in HIV infected individuals. HIV-1 infects primarily T 10 lymphocytes, monocytes/macrophages, dendritic cells and, in the central nervous system, microglia. All of these cells express the CD4 glycoprotein, which serves as a receptor for HIV-1 and HIV-2. Efficient entry of HIV into target cells is dependent upon binding of the viral 15 exterior envelope glycoprotein, gp120, to the amino-terminal CD4 domain. After virus binding, the HIV-1 envelope glycoproteins mediate the fusion of viral and host cell membranes to complete the entry process. Membrane fusion directed by HIV-1 envelope glycoproteins 20 expressed on the infected cell surface leads to cell-cell fusion, resulting in syncytia.

Recently, host cell factors in addition to CD4 have been suggested to determine the efficiency of HIV-1 envelope glycoprotein-mediated membrane fusion. The 7 25 transmembrane receptor (7TMR) termed HUMSTR, LESTR, or "fusin" has been shown to allow a range of CD4-expressing cells to support infection and cell fusion mediated by laboratory-adapted HIV-1 envelope glycoproteins (Feng, Y., et al., *Science (Wash. DC)*, 30 272:872-877 (1996)). Antibodies to HUMSTR blocked cell fusion and infection by laboratory-adapted HIV-1 isolates but not by macrophage-tropic primary viruses *in vitro* (Feng, Y., et al., *Science (Wash. DC)*, 272:872-877 (1996)).

35 The ability of chemokine receptors and related molecules to facilitate the infection of primary

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clinical HIV-1 isolates has been reported recently by several groups (see e.g., Bates, P., *Cell*, 86:1-3 (1996); Choe, H., et al., *Cell*, 85:1135-1148 (1996); Doranz et al., *Cell* 85:1149-1158 (1996)). These studies 5 indicated that involvement of various members of the chemokine receptor family in the early stages of HIV-1 infection helps to explain viral tropism and β -chemokine inhibition of primary HIV-1 isolates.

The present invention also provides a method of 10 inhibiting HIV infection of a cell (e.g., new infection and/or syncytium formation) which expresses a mammalian CCR2 or portion thereof, comprising contacting the cell with a composition comprising an effective amount of an antibody or functional fragment thereof which binds to a 15 mammalian CCR2 or portion of said receptor. The composition can also comprise one or more additional agents effective against HIV, including, but not limited to, anti-CCR3 antibodies, anti-CCR5 antibodies, and anti-fusin antibodies.

20 Various methods can be used to assess binding of HIV to a cell and/or infection of a cell by HIV in the presence of the antibodies of the present invention. For example, assays which assess binding of gp120 or a portion thereof to the receptor, HIV infection and 25 syncytium formation can be used (see, for example, Choe, H., et al., *Cell*, 85:1135-1148 (1996)). The ability of the antibody of the present invention to inhibit these processes can be assessed using these or other suitable methods.

30 In addition, the present invention provides a method of treating HIV in a patient, comprising administering to the patient a composition comprising an effective amount of an antibody or functional fragment thereof which binds to a mammalian CCR2 or portion of 35 said receptor. Again, the composition can also comprise one or more additional agents effective against HIV,

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including, but not limited to, anti-CCR3 antibodies, anti-CCR5 antibodies, and anti-fusin antibodies. Therapeutic use of antibody to treat HIV includes prophylactic use (e.g., for treatment of a patient who 5 may be or who may have been exposed to HIV). For example, health care providers who may be exposed or who have been exposed to HIV (e.g., by needle-stick) can be treated according to the method. Another example is the treatment of a patient exposed to virus after 10 unprotected sexual contact or failure of protection.

In AIDS, multiple drug treatment appears the most promising. An anti-chemokine receptor antagonist that inhibits HIV infection can be added to the drug treatment regimen, in particular by blocking virus 15 infection of new cells. Thus, administration of an antibody or fragment of the present invention in combination with one or more other therapeutic agents such as nucleoside analogues (e.g., AZT, 3TC, ddI) and/or protease inhibitors is envisioned, and provides 20 an important addition to an HIV treatment regimen. In one embodiment, a humanized anti-CCR2 mAb is used in combination with a (i.e., one or more) therapeutic agent to reduce viral load from patients, by preventing fusion and/or infection of new cells. Such an antibody can 25 also be useful in preventing perinatal infection.

Another aspect of the invention relates to a method of preventing HIV infection in an individual, comprising administering to the individual an effective amount of an antibody or functional fragment thereof which binds 30 to CCR2. According to the method, preventing HIV infection includes treatment in order to prevent (reduce or eliminate) infection of new cells in an infected individual or in order to prevent infection in an individual who may be, may have been, or has been, 35 exposed to HIV. For example, individuals such as an HIV infected individual, a fetus of an HIV infected female,

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or a health care worker may be treated according to the method of the present invention.

Modes of Administration

One or more antibodies or fragments of the present invention can be administered to an individual by an appropriate route, either alone or in combination with (before, simultaneous with, or after) another drug or agent, or before, simultaneous with or after surgical, mechanical or therapeutic intervention. For example, 5 the antibodies of the present invention can also be used in combination with other monoclonal or polyclonal antibodies (e.g., in combination with antibodies which bind other chemokine receptors, including, but not limited to, CCR3 and CCR5) or with existing blood plasma 10 products, such as commercially available gamma globulin and immune globulin products used in prophylactic or therapeutic treatments. The antibodies or fragments of the present invention can be used as separately administered compositions given in conjunction with 15 antibiotics and/or antimicrobial agents.

An effective amount of an antibody or fragment (i.e., one or more antibodies or fragments) is administered. An effective amount is an amount sufficient to achieve the desired therapeutic (including 20 prophylactic) effect, under the conditions of administration, such as an amount sufficient for inhibition of a CCR2 function, and thereby, inhibition 25 of an inflammatory response or HIV infection, or an amount sufficient for promotion of a CCR2 function, as indicated.

A variety of routes of administration are possible including, but not necessarily limited to, oral, dietary, topical, parenteral (e.g., intravenous, intraarterial, intramuscular, subcutaneous injection or 30 infusion), inhalation (e.g., intrabronchial,

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intraocular, intranasal or oral inhalation, intranasal drops), depending on the disease or condition to be treated. Other suitable methods of administration can also include rechargeable or biodegradable devices and 5 slow release polymeric devices. The pharmaceutical compositions of this invention can also be administered as part of a combinatorial therapy with other agents.

Formulation of an antibody or fragment to be administered will vary according to the route of 10 administration and formulation (e.g., solution, emulsion, capsule) selected. An appropriate pharmaceutical composition comprising an antibody or functional fragment thereof to be administered can be prepared in a physiologically acceptable vehicle or 15 carrier. A mixture of antibodies and/or fragments can also be used. For solutions or emulsions, suitable carriers include, for example, aqueous or alcoholic/aqueous solutions, emulsions or suspensions, including saline and buffered media. Parenteral 20 vehicles can include sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride, lactated Ringer's or fixed oils. A variety of appropriate aqueous carriers are known to the skilled artisan, including water, buffered water, buffered saline, 25 polyols (e.g., glycerol, propylene glycol, liquid polyethylene glycol), dextrose solution and glycine. Intravenous vehicles can include various additives, preservatives, or fluid, nutrient or electrolyte replenishers (See, generally, *Remington's Pharmaceutical 30 Science*, 16th Edition, Mack, Ed. 1980). The compositions can optionally contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions such as pH adjusting and buffering agents and toxicity adjusting 35 agents, for example, sodium acetate, sodium chloride, potassium chloride, calcium chloride and sodium lactate.

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The antibodies and fragments of this invention can be lyophilized for storage and reconstituted in a suitable carrier prior to use according to art-known lyophilization and reconstitution techniques. The 5 optimum concentration of the active ingredient(s) in the chosen medium can be determined empirically, according to procedures well known to the skilled artisan, and will depend on the ultimate pharmaceutical formulation desired. For inhalation, the antibody or fragment can 10 be solubilized and loaded into a suitable dispenser for administration (e.g., an atomizer, nebulizer or pressurized aerosol dispenser).

The present invention will now be illustrated by the following Examples, which are not intended to be 15 limiting in any way. The teachings of all references cited herein are incorporated herein by reference.

EXAMPLES

Materials:

The following materials were obtained from the 20 indicated sources:
PE-conjugated anti-CD16, PE-conjugated streptavidin, and biotinylated anti-human IgE were from Pharmingen (San Diego, CA). FITC-conjugated goat anti-mouse IgG was from Jackson Immunoresearch Laboratories 25 (West Grove, PA). FACS Lysing Buffer was from Becton Dickenson (Mountain View, CA) and [¹²⁵I]-MCP-1 was from NEN (Boston, MA).

Cells, Cell Lines, and Tissue Culture

The murine pre-B lymphoma cell line L1/2 was 30 maintained in RPMI-1640 supplemented with 10% Fetal Clone I (Gibco BRL, Gaithersburg, MD) 50 Units/mL penicillin (Gibco BRL), 50 µg/mL streptomycin (Gibco BRL), 2 mM L-Glutamine (Gibco BRL), and 55 µM β-mercaptoethanol (Gibco BRL). Other cell lines included

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transfectants of L1/2 cells expressing either CCR1
(Campbell, J. et al. (1996) *J. Cell Biol.*, 134:255-266),
CCR5 (Wu et al., *Nature* 384:179-183 (1996)) grown in the
above culture medium supplemented with 800 µg/ml active
5 G418. THP-1 cells (ATCC No. TIB202) were grown in
accordance with ATCC instructions. PBMC were purified
from heparinized blood as described in Ponath et al., *J.*
Clin. Invest., 97:604-612 (1996).

Preparation of CCR2b Expression Construct and Stable
10 Transfectants

The coding region for the human CCR2b (Charo et al.
(1994) *Proc. Natl. Acad. Sci. USA*, 91:2752) was obtained
by RT-PCR amplification as described (Qin, S. et al.
(1996) *Eur. J. Immunol.*, 26:640-647). cDNA was made
15 using oligo(dT)-priming, and amplification of the CCR2b
coding region was achieved by nested PCR with the
following sets of primers which correspond to the
positions of the CCR2b sequence (GenBank Accession No.
U03905; Charo et al., *Proc. Natl. Acad. Sci. USA*
20 91:2752-2756 (1994)) as indicated:

- 1) 5' primer: 5'-TGAGACAAGCCACAAGCTGAAC-3'
(nucleotides 11 to 32; SEQ ID NO: 1);
3' Primer: 5'-TCTGTATTAGTACACACAGCCC-3'
(nucleotides 1301 to 1280; SEQ ID NO: 2);
- 25 2) 5' Primer: 5'-ATGCTGTCCACATCTCGTTCTCGG-3'
(nucleotides 81 to 104; SEQ ID NO: 3);
3' Primer: 5'-TTATAAACCGCCGAGACTCCTGCTC-3'
(nucleotides 1164 to 1137; SEQ ID NO: 4).

The CCR2B cDNA coding region was modified to
30 contain the CD5 signal peptide leader sequence (Aruffo
et al., *Cell* 61:1303-1313 (1990)). The predicted amino
acid sequence of this peptide is:

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NH₂-Met-Pro-Met-Gly-Ser-Leu-Gln-Pro-Leu-Ala-
Thr-Leu-Tyr-Leu-Leu-Gly-Met-Leu-Val-Ala-Ser-
Val-Leu-Ala... (SEQ ID NO: 5)

Using PCR with the CCR2b cDNA as template and two
5 overlapping 5' primers that contain a BamHI restriction
site, encode the CD5 signal peptide sequence and the
amino terminal sequence of CCR2b, and a 3' primer
located internally in the CCR2b coding region.

10 5' CD5 Seq1 primer
5'- GGGGATCCAGAAACCATGCCATGGGTCTCTGCAACCGCTGGCCA
CCTTGTACCTGCTG-3' (SEQ ID NO: 6)

5' CD5 Seq2 primer
5'- GCCACCTTGTACCTGCTGGGATGCTGGTCGCTCCGTGCTAGCGA
TGCTGTCCACATCTCGTTC-3' (SEQ ID NO: 7)

15 3' CCR2AB2 primer - 5' - GACGACCAGCATGTTGCC-3'
(SEQ ID NO: 8; U03905 nucleotides 272 to 255)

The 278 base pair amplified fragment was digested
with BamHI and Apal and the resulting 209 base pair
fragment was inserted at the Apal site at position 206
20 of the CCR2b cDNA (GenBank Accession No. U03905) to
replace the endogenous 5' base pair fragment of CCR2.
The resulting sequence that encodes a CCR2b with the CD5
signal peptide leader sequence immediately preceding the
receptor initiator methionine was inserted into the
25 BamHI and Xhol sites of pcDNA3 (Invitrogen, San Diego,
CA) to create the mammalian expression plasmid
pCD5MCPRB. The CD5-CCR2b fragment was subcloned into
the BamH I-Not I site of pCDF3 (Goldman et al., (1996)
Biotechniques 21:1013-1015), and this construct was
30 designated CCR2bDEF3. In this expression vector, the

expression of the inserted gene is driven by the EF-1 α promoter.

5 Fifty milliliters of L1/2 cells were seeded at 4 x 10⁵ cells/mL the day before the electroporation. On the day of the electroporation, the cells, which had grown up to a density of 1 x 10⁶/mL, were centrifuged out of their medium and resuspended in 800 μ l room temperature electroporation buffer (Zajac et al., *DNA* 7:509-513).
10 120 mM L-Glutamic Acid (Sigma), 7 mM Mg Acetate (EM Science), 4.3 mM Glucose (Sigma), 17 mM K Pipes, pH 6.9 (Sigma), 1 mM EGTA (Sigma), 5 mM ATP, pH 7.0 (Sigma). Twenty-five micrograms Sca I linearized, phenol/chloroform/isoamyl alcohol extracted and isopropanol precipitated CCR2bDEF3 plasmid DNA was
15 placed in an 0.4 cm gap electroporation curvette. The resuspended cells were added to the curvette, and a single pulse applied at 450 volts, 960 μ Fd. The cells were then transferred from the curvette to a T-75 flask containing 15 mL L1/2 growth medium (described above),
20 and grown for three days, at which time the cells were centrifuged out of their medium and resuspended in L1/2 growth medium additionally supplemented with 1 mM sodium pyruvate (Gibco BRL) and 0.8 mg/mL active G418 (Gibco BRL).
25 Selection of Cells Expressing CCR2b by Chemotaxis

The transfected cells were allowed to grow for eleven days, at which point they were split 1:20 into fresh growth medium. On the sixteenth day, the cells were selected by chemotaxis. 600 μ L 1 nM MCP-1 in RPMI 30 1640 supplemented with 0.5% BSA (RPMI/BSA) was placed in the lower chamber and 1 x 10⁶ CCR2bDEF3 cells in 100 μ l of RPMI/BSA were placed in the upper chamber of a 3.0 micron pore 24-well chemotaxis plate (Becton Dickinson). The cells were allowed to chemotax for four hours and
35 twenty minutes in a 37° C, 5% CO₂, humidified incubator,

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at which time the upper chamber was removed. This incubation time was chosen at the time of the experiment because it was sufficiently long for cells responding to the MCP-1 to chemotax, but short enough to keep the 5 background low.

Secondary Selection of CCR2b Expressing-Cells by FACS Sorting

The cells which had chemotaxed through the membrane and into the lower chamber were grown up, and further 10 purified by sterile FACS sorting. Ten million CCR2bDEF3 cells were centrifuged out of their medium, resuspended in 2.5 mL PBS(+Ca, Mg) supplemented with 1% heat-inactivated Fetal Calf Serum ("HI FCS") (Gibco BRL) and 2.5 mL sterile filtered anti-CCR2b amino-terminal 15 peptide antibody supernatant 5A11. The cells and the antibody were mixed and allowed to incubate on ice for thirty minutes. The cells were then washed twice with PBS (+) (Gibco BRL), and resuspended in 5 mL of a sterile filtered, 1:250 dilution of FITC-conjugated, 20 affinity-purified F(ab¹), goat anti-mouse IgG (Jackson ImmunoResearch Laboratories) in PBS (+) supplemented with 1% HI FCS. The cells were incubated for thirty minutes on ice in the dark, and then washed twice with PBS(+) (GIBCO BRL). The cells were sorted on the 25 FACSCalibur® and the brightest 4% of cells were collected. (FL1 $\geq 3 \times 10^3$).

The sorted cells were allowed to grow up, and they were resorted using the same protocol as above. The brightest 1% of cells were collected. (FL1 $\geq 3 \times 10^3$).

30 Monoclonal Antibody Production

To produce mAbs to CCR2b, transfectants were continually monitored to ensure that levels of expression did not drift downward. FACS staining was performed periodically to ascertain receptor expression

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on the transfecants using the anti CCR2b antibody supernatant 5A11 with goat anti-mouse IgG FITC as the secondary antibody.

Twenty million CCR2bDEF3.L1/2 cells were washed in 5 RPMI 1640 (Gibco BRL) and incubated in RPMI 1640 plus 0.2 mg/mL Mitomycin C for 30 minutes at 37°C. The cells were then washed twice with PBS (+) and 2×10^7 cells in 0.5 mL PBS (+) were injected intraperitoneally into a C57 BL/6 female mouse. This was repeated two more times 10 at two week intervals. The fourth time, 2×10^7 cells were resuspended in 0.25 mL and injected intravenously. Three days after the intravenous injection, the mouse was sacrificed and the spleen removed and the cells fused with the SP2/0 cell line as described (*Current 15 Protocols in Immunology*, John Wiley and Sons, New York, 1992).

This set of mice had previously been immunized many times with 2 different cell lines as well as a synthetic peptide, but no antibodies that stained CCR2 positive 20 cells were generated from several fusions. The above four immunizations with the CCR2bDEF3.L1/2 cell line expressing high levels of CCR2b were critical to obtain the described antibody.

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Selecting Single Cell Clone of CCR2 Transfectants by
Limiting Dilution

After the mouse received the last injection, the twice sorted cells were allowed to grow up again, and 5 then they were further purified by limiting dilution. The cells were plated at 1 and 0.5 cell per well in 96 well plates. Subcloned cells from the 0.5 cell per well dilution were grown up and tested for CCR2b expression by indirect immunofluorescent FACS analysis using the 10 anti-CCR2b antibody supernatant 5A11 with goat anti-mouse IgG FITC as the secondary antibody. The procedure was the same as described above, except that the staining volume was 100 μ l. Four positives were selected and frozen down.

15 Identification of Positive Monoclonal Antibodies

Immunofluorescent staining analysis using a FACScan® (Becton Dickinson & Co., Mountain View, CA) was used to identify the monoclonal antibodies which were reactive with the CCR2b receptor. Hybridoma culture 20 supernatants were assayed in a 96-well format using goat anti-mouse IgG FITC as the secondary antibody. CCR2bDEF3.L1/2 cells were used to identify monoclonal antibodies reactive with CCR2b, and untransfected L1/2 cells were used to eliminate monoclonal antibodies 25 reactive with other cell surface proteins.

FACS Staining - Cultured Cells

For the staining of cultured transfectant cell lines 0.5×10^6 cells in 50 μ l were resuspended in PBS + 1% FCS in a 96 well polystyrene V-bottom plate. 50 μ l 30 of primary antibody supernatants or HT medium (negative control) were added, and the samples were incubated at 4°C for 30 min. 100 μ l of PBS were added and the cells were pelleted by centrifugation and washed once with PBS. The pellet was resuspended in 100 μ l PBS + 1% FCS

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containing FITC-conjugated goat anti-mouse IgG antibody (a 1:250 dilution) and incubated for thirty minutes at 4°C in the dark. The cells were washed twice with PBS, resuspended in PBS, and analyzed by flow cytometry with 5 a FacScan cytometer using the CellQuest software (Becton-Dickenson) Cells were fixed with PBS/1% formaldehyde if they were not to be analyzed the same day. Monoclonal antibodies 1D9 and 8G2 stain CCR2 transfectants but not CCR1 or CCR5 transfectants 10 (Figures 1A-1O).

FACS Staining - Whole Blood

100 µl whole blood was mixed with 100 µL of 1D9 antibody hybridoma supernatants or HT medium (negative control) and incubated at 4°C for 30 min. After one 15 wash with PBS, 100 µL FITC-conjugated goat anti-mouse IgG antibody (a 1:250 dilution) was added to each sample and incubated for 30 min. at 4°C in the dark. Samples were then washed once with PBS if a second color staining is to be done, otherwise washed twice more in 20 PBS. For two color staining 5 µl of mouse serum was added to the cell pellets after the single wash, mixed, and incubated for five minutes at 4°C in the dark. Second primary antibodies (or PBS as a negative control) 25 were added (10 µl anti-CD16, 100 µl 1:200 dilution of anti-IgE) and incubated for thirty minutes at 4°C in the dark. Samples were then washed one time with PBS and resuspended in 100 µL streptavidin PE (1:200 PBS + 1% BSA) and incubated for fifteen minutes at 4°C in the dark. Erythrocytes were lysed by adding 2 ml of FACS 30 Lysing Buffer to each sample and incubating at room temperature in the dark for fifteen minutes or until samples were clear. The cells were pelleted by centrifugation and all but 200 µl of the supernatant was aspirated. The samples were analyzed by flow cytometry 35 on a FacScan cytometer using the CellQuest software

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(Becton-Dickenson). CCR2b is expressed on most monocytes, a subpopulation of lymphocytes and a subset of granulocytes (Figures 2A-2L). CCR2b is expressed on an IgE-positive population in peripheral blood 5 (basophils) (Figures 3A-3I).

MCP-1 Binding Assays

MCP-1 binding was performed in a final volume of 0.1 ml of 50 mM Hepes pH 7.4, 1 mM CaCl₂, 5 mM MgCl₂, 0.02% sodium azide, 0.5% BSA (HBB), containing either 10 2.5 µg THP-1 membrane protein or 500,000 PBMC and 0.1 nM of [¹²⁵I]-MCP-1. Competition binding experiments were performed by including variable concentrations of unlabeled MCP-1, 1D9 antibody, or a negative control IgG2a. Nonspecific binding was determined following the 15 addition of a 2500-fold excess of unlabeled MCP-1. Samples were incubated for 60 minutes at room temperature, and bound and free tracer were separated by filtration through 96-well GF/B filterplates presoaked in 0.3% polyethylenimine. The filters were washed in 20 HBB further supplemented with 0.5 M NaCl, dried, and the amount of bound radioactivity determined by liquid scintillation counting. mAb 1D9 inhibits [¹²⁵I]MCP-1 binding to THP-1 cell membranes with an IC₅₀ of about 0.004 µg/ml (approximately 0.02 nM; Figure 4) and to 25 fresh PBMC with an IC₅₀ of 0.04 µg/ml (approximately 0.2 nM; Figure 5).

Chemotaxis of PBMC

Chemotaxis was assayed using a 3 µm pore size 96-well chemotaxis plate (Neuroprobe, Cabin John, MD). 30 PBMC isolated by standard methods using Ficoll-Hypaque density gradient centrifugation were washed with PBS/0.5%BSA and then resuspended in chemotaxis assay media (HBSS/10mM HEPES/0.5% Fatty acid free BSA) to a final concentration of 10 x 10⁶ cells/ml. Cells were

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principulated in chemotaxis assay media at room temperature for 20 min. with various concentrations of the anti-CCR2 antibody, 1D9, or nonspecific murine IgG2a. The same dilutions of antibody were mixed with chemokine and 30 μ l of 5 the mixture was added to each of the bottom wells of the chemotaxis plate. The bottom wells are covered with the membrane, and 25 μ l of the cell and antibody mixture are added to the top of the filter. The plates are incubated at 37°C in 5% CO₂ incubator for approximately 80 min. At the 10 completion of the migration, the membrane is removed and the plate with the bottom wells is incubated -80C for 30 minutes to freeze the contents. The plates are thawed at 37°C for 10 minutes. 6 μ l of a 1:400 dilution of CyQuant reagent (Molecular Probes, Eugene, OR) in a lysis buffer provided by 15 the supplier is added to each well, and the cell migration is quantified as indicated by fluorescence intensity determined using a CytoFlour fluorescence plate reader at 485ex/535em. mAB 1D9 inhibits MCP-1-induced chemotaxis, but not RANTES-induced chemotaxis, of fresh PBMC (Figures 6A and 20 6B). Inhibition of MCP-1-induced chemotaxis of fresh PBMC has been demonstrated with 10 μ g/ml (\approx 40 nM).

The reference to any prior art in this specification is not, and should not be taken as, an acknowledgment or any form of suggestion that that prior art forms part of the 25 common general knowledge in Australia.

Throughout this specification and the claims which follow, unless the context requires otherwise, the word "comprise", and variations such as "comprises" and "comprising", will be understood to imply the inclusion of a 30 stated integer or step or group of integers or steps but not the exclusion of any other integer or step or group of integers or steps.

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While this invention has been particularly shown and described with reference to preferred embodiments thereof, it will be understood by those skilled in the art that various changes in form and details may be made therein 5 without departing from the spirit and scope of the invention as defined by the appending claims.

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EDITORIAL NOTE

APPLICATION NUMBER - 52208/99

**The following Sequence Listing pages 1 to 2 are part of the
description. The claims pages follow on pages 65 to 96.**

SEQUENCE LISTING

<110> LeukoSite, Inc.
LaRosa, Gregory J.
Horvath, Christopher
Newman, Walter

<120> ANTI-CCR2 ANTIBODIES AND METHODS OF USE
THEREFOR

<130> LKS98-04A

<150> 09/121,781
<151> 1998-07-23

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THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:

1. An antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor.
 - 10 2. An antibody or antigen-binding fragment thereof according to Claim 1 wherein the antibody or antigen-binding fragment binds the mammalian CC-chemokine receptor amino-terminal domain from about amino acid 1 to about amino acid 30.
 - 15 3. An antibody or antigen-binding fragment thereof according to Claim 1 wherein the antibody or antigen-binding fragment thereof is selected from the group consisting of:
 - 20 a) monoclonal antibody 1D9;
 - b) an antibody having the epitopic specificity of 1D9;
 - c) monoclonal antibody 8G2;
 - d) an antibody having the epitopic specificity of 8G2; and
 - e) antigen-binding fragments of any one of (a) through (d) which bind to a mammalian CC-chemokine receptor 2 amino-terminal domain.
 - 25 30 4. An antibody or antigen-binding fragment thereof according to Claim 1 wherein the chemokine is selected

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from the group consisting of MCP-1, MCP-2, MCP-3, MCP-4 and combinations thereof.

5. The hybridoma cell line deposited under ATCC Accession No. HB-12549.
 6. The hybridoma cell line deposited under ATCC Accession No. HB-12550.
 - 10 7. A monoclonal antibody produced by the hybridoma cell line according to Claim 5 or antigen-binding fragment thereof.
 - 15 8. A monoclonal antibody produced by the hybridoma cell line according to Claim 6 or antigen-binding fragment thereof.
 - 20 9. A test kit for use in detecting the presence of a mammalian CC-chemokine receptor 2 or a portion of said receptor comprising an amino-terminal domain thereof, in a biological sample comprising
 - a) at least one antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and wherein said antibody or antigen-binding fragment thereof is selected from the group consisting of:
 - i) monoclonal antibody 1D9;
 - 25 30

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- ii) an antibody having the epitopic specificity of 1D9;

iii) monoclonal antibody 8G2;

iv) an antibody having the epitopic specificity of 8G2; and

v) antigen-binding fragments of any one of (i) through (iv) which bind to the amino-terminal domain of CC-chemokine receptor 2; and

b) one or more ancillary reagents suitable for detecting the presence of a complex between said antibody or antigen-binding fragment thereof and said mammalian CC-chemokine receptor 2 or a portion thereof.

15 10. A method of inhibiting the interaction of a cell bearing mammalian CC-chemokine receptor 2 with a corresponding chemokine, comprising contacting said cell with an effective amount of an antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, inhibits binding of said chemokine to the receptor, and inhibits one or more functions associated with binding of the chemokine to the receptor.

20 25 11. A method according to Claim 10 wherein the cell is selected from the group consisting of lymphocytes, monocytes, granulocytes, T cells, basophils, and cells comprising a recombinant nucleic acid encoding CCR2 or portion thereof.

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12. A method according to Claim 11 wherein the T cells are selected from the group consist of CD8+ cells, CD25+ cells, CD4+ cells and CD45RO+ cells.

5 13. A method according to Claim 10 wherein the chemokine is selected from the group consisting of MCP-1, MCP-2, MCP-3, MCP-4 and combinations thereof.

10 14. A method according to Claim 10 wherein the antibody or antigen-binding fragment thereof is selected from the group consisting of:

a) monoclonal antibody 1D9;
b) an antibody having the epitopic specificity of 1D9;

15 c) an antibody which can compete with 1D9 for binding to mammalian CC-chemokine receptor 2;

d) monoclonal antibody 8G2;

e) an antibody having the epitopic specificity of 8G2;

20 f) an antibody which can compete with 8G2 for binding to mammalian CC-chemokine receptor 2; and

g) antigen-binding fragments of any one of (a) through (f) which bind to mammalian CC-chemokine receptor 2 amino-terminal domain.

25 15. A method of detecting expression of mammalian CC-chemokine receptor 2 or a portion of said receptor comprising an amino-terminal domain thereof, by a cell, comprising:

30 a) contacting a composition comprising a cell to be tested with an antibody or antigen-binding fragment thereof which binds to a mammalian CC-

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- chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and wherein said antibody or antigen-binding fragment thereof is selected from the group consisting of:
- 5 i) monoclonal antibody 1D9;
- 10 ii) an antibody having the epitopic specificity of 1D9;
- iii) monoclonal antibody 8G2;
- iv) an antibody having the epitopic specificity of 8G2;
- 15 v) antigen-binding fragments of any one of (i) through (iv) which bind mammalian CC-chemokine receptor 2 amino-terminal domain; and
- 20 vi) combinations of (i) to (v), under conditions appropriate for binding of said antibody or antigen-binding fragment thereof thereto; and
- b) detecting binding of said antibody or antigen-binding fragment thereof, wherein the binding of said antibody or antigen-binding fragment thereof indicates the presence of said receptor or portion of said receptor comprising an amino-terminal domain thereof, on said cell.
- 25
16. The method of Claim 15 wherein the composition is a
- 30 sample comprising human cells.

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17. A method of detecting a mammalian CC-chemokine receptor 2 or portion of said receptor comprising an amino-terminal domain thereof, comprising:
- 5 a) contacting a sample to be tested with an antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and wherein said antibody or antigen-binding fragment thereof is selected from the group consisting of:
- 10 i) monoclonal antibody 1D9;
- 15 ii) an antibody having the epitopic specificity of 1D9;
- iii) monoclonal antibody 8G2;
- iv) an antibody having the epitopic specificity of 8G2;
- 20 v) antigen-binding fragments of any one of (i) through (iv) which bind to mammalian CC-chemokine receptor 2 amino-terminal domain; and
- 25 vi) combinations of (i) to (v), under conditions appropriate for binding of said antibody or fragment thereof thereto; and
- 26 b) detecting or measuring binding of said antibody or antigen-binding fragment thereof,
- 27 wherein the binding of said antibody or antigen-binding fragment thereof to material in said sample is
- 28 30 indicative of the presence of a mammalian CC-chemokine

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receptor 2 or portion of said receptor comprising an amino-terminal domain thereof, in said sample.

18. A method according to Claim 17, wherein the sample is a
5 cellular fraction which, in normal individuals,
comprises a mammalian CC-chemokine receptor 2 or
portion of said receptor comprising an amino-terminal
domain.

10 19. A method of inhibiting a function associated with
binding of a chemokine to the amino-terminal domain of
mammalian CC-chemokine receptor 2 or a functional
portion comprising an amino-terminal domain of said
receptor, comprising contacting a composition
15 comprising the receptor or portion with an effective
amount of an antibody or antigen-binding fragment
thereof which binds to a mammalian CC-chemokine
receptor 2 amino-terminal domain, wherein said antibody
inhibits binding of said chemokine to mammalian
20 CC-chemokine receptor 2 and inhibits one or more
functions associated with binding of the chemokine to
the receptor.

25 20. A method according to Claim 19 wherein the chemokine is
selected from the group consisting of MCP-1, MCP-2,
MCP-3, MCP-4 and combinations thereof.

30 21. A method according to Claim 19 wherein the antibody or
antigen-binding fragment is selected from the group
consisting of:
a) monoclonal antibody 1D9;

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- b) an antibody having the epitopic specificity of 1D9;
- c) an antibody which can compete with 1D9 for binding to mammalian CC-chemokine receptor 2; and
- 5 d) antigen-binding fragments of any of (a) through (c) which bind to mammalian CC-chemokine receptor 2 amino-terminal domain.

22. A method of detecting or identifying an agent which
- 10 binds a mammalian CC-chemokine receptor 2 or ligand-binding variant thereof, comprising combining
- a) an agent to be tested;
 - b) an antibody or antigen-binding fragment which binds to a mammalian CC-chemokine receptor 2 amino-

15 terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and wherein said antibody or antigen-binding fragment thereof is selected from the group

20 consisting of:

 - i) monoclonal antibody 1D9;
 - 15 ii) an antibody having the epitopic specificity of 1D9;
 - iii) monoclonal antibody 8G2;
 - iv) an antibody having the epitopic specificity of 8G2;
 - v) antigen-binding fragments of any of (i) through (iv) which bind to mammalian

25 CC-chemokine receptor 2 amino-terminal domain; and

 - vi) combinations of (i) to (v); and

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c) a composition comprising a mammalian CC-chemokine receptor 2 or a ligand-binding variant thereof, under conditions suitable for binding of said antibody or antigen-binding fragment to said mammalian CC-chemokine receptor 2 or ligand-binding variant thereof, and detecting or measuring binding of said antibody or antigen-binding fragment to said mammalian CC-chemokine receptor 2 or ligand-binding variant thereof.

10

23. A method according to Claim 22 wherein the formation of a complex between said antibody or antigen-binding fragment and said mammalian CC-chemokine receptor 2 or ligand-binding variant is monitored, and wherein a decrease in the amount of complex formed relative to a suitable control is indicative that the agent binds said receptor or ligand-binding variant thereof.

15

24. A method according to Claim 22 wherein the composition comprising a mammalian CC-chemokine receptor 2 or a ligand-binding variant thereof is a cell bearing recombinant CC-chemokine receptor 2 or ligand-binding variant thereof.

20

25. A method according to Claim 24, wherein the composition comprising a mammalian CC-chemokine receptor 2 or a ligand-binding variant thereof is a membrane fraction of said cell bearing recombinant CC-chemokine receptor 2 or ligand-binding variant thereof.

25

26. A method according to Claim 22 wherein the antibody or antigen-binding fragment thereof is labeled with a

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label selected from the group consisting of a radioisotope, spin label, antigen label, enzyme label, fluorescent group and chemiluminescent group.

- 5 27. A method according to Claim 22 wherein the agent is an
antibody having specificity for a mammalian CC-
chemokine receptor 2 or antigen-binding fragment
thereof.

10 28. A method of inhibiting leukocyte trafficking in a
patient, comprising administering to the patient a
composition comprising an effective amount of an
antibody or antigen-binding fragment thereof which
binds to a mammalian CC-chemokine receptor 2 amino-
15 terminal domain, inhibits binding of a ligand to the
receptor, and inhibits one or more functions associated
with binding of the ligand to the receptor.

20 29. A method of according to Claim 28 wherein the ligand is
a chemokine.

25 30. A method according to Claim 29 wherein the chemokine is
selected from the group consisting of MCP-1, MCP-2,
MCP-3, MCP-4 and combinations of the foregoing.

31. A method according to Claim 28 wherein the antibody or
antigen-binding fragment thereof is selected from the
group consisting of:
a) monoclonal antibody 1D9;
b) an antibody having the epitopic specificity of
1D9;

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- c) an antibody which can compete with 1D9 for binding to mammalian CC-chemokine receptor 2; and
 - d) antigen-binding fragments of any of (a) through (c) which bind to mammalian CC-chemokine receptor 2 amino-terminal domain.
32. A composition comprising an antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and an optional physiologically acceptable vehicle.
33. An antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor with an IC₅₀ of less than about 1.0 µg/ml and inhibits one or more functions associated with binding of the chemokine to the receptor.
34. An antibody or antigen-binding fragment thereof according to Claim 33 wherein the IC₅₀ is less than about 0.05 µg/ml.
35. An antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a

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chemokine to the receptor, and wherein the antibody or antigen-binding fragment binds the receptor with an affinity of at least about 0.1×10^{-9} M and inhibits one or more functions associated with chemokine binding to the receptor.

36. An antibody or antigen-binding fragment thereof according to Claim 35, wherein the affinity is at least about 1×10^{-9} M.

10

37. An antibody or antigen-binding fragment thereof according to Claim 36, wherein the affinity is at least about 3×10^{-9} M.

15 38. A method of treating a CC-chemokine receptor 2-mediated disorder in a patient, comprising administering to the patient an effective amount of an antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits chemokine binding to the receptor and inhibits one or more functions associated with chemokine binding to the receptor.

25 39. A method according to Claim 38 wherein the disorder is an inflammatory disorder.

40. A method according to Claim 38, wherein said CC-chemokine receptor 2-mediated disorder is an autoimmune disorder.

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41. A method according to Claim 40, wherein the autoimmune disorder is selected from the group consisting of multiple sclerosis and rheumatoid arthritis.
- 5 42. A method according to Claim 41, wherein the autoimmune disorder is multiple sclerosis.
- 10 43. A method according to Claim 41, wherein the CC-chemokine receptor 2-mediated disorder is selected from the group consisting of psoriatic arthritis, scleroderma, atherogenesis and atherosclerosis.
- 15 44. A method of inhibiting restenosis in a patient, comprising administering to the patient an effective amount of an antibody or antigen-binding fragment thereof which binds to mammalian CC-chemokine receptor 2, wherein said antibody or antigen-binding fragment thereof inhibits binding of a ligand to the receptor.
- 20 45. A method according to Claim 44, wherein the antibody or antigen-binding fragment thereof is selected from the group consisting of:
 - a) monoclonal antibody 1D9;
 - b) an antibody having the epitopic specificity of monoclonal antibody 1D9;
 - 25 c) an antibody which can compete with monoclonal antibody 1D9 for binding to mammalian CC-chemokine receptor 2;
 - d) monoclonal antibody 8G2;
 - 30 e) an antibody having the epitopic specificity of monoclonal antibody 8G2;

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f) an antibody which can compete with monoclonal antibody 8G2 for binding to mammalian CC-chemokine receptor 2;

5 g) antigen-binding fragments of any one of (a) through (f) which bind to mammalian CC-chemokine receptor 2 or a portion thereof; and

h) combinations of (a) to (g).

46. A method of inhibiting restenosis of a vessel in a
10 mammal, comprising administering to said mammal an effective amount of an antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2, wherein said antibody or antigen-binding fragment thereof inhibits binding of a ligand
15 to the receptor.

47. A method according to Claim 46 wherein the antibody or antigen-binding fragment thereof is selected from the group consisting of:

20 a) monoclonal antibody 1D9;
b) an antibody having the epitopic specificity of monoclonal antibody 1D9;
c) an antibody which can compete with monoclonal antibody 1D9 for binding to mammalian CC-chemokine
25 receptor 2;
d) monoclonal antibody 8G2;
e) an antibody having the epitopic specificity of monoclonal antibody 8G2;
f) an antibody which can compete with monoclonal
30 antibody 8G2 for binding to mammalian CC-chemokine receptor 2;

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- g) antigen-binding fragments of any one of (a) through (g) which bind to mammalian CC-chemokine receptor 2 or a portion thereof; and
 - h) combinations of (a) to (g).

5

48. A method according to Claim 46, wherein said restenosis is associated with vascular intervention in said mammal.

10

- 10 49. A method according to Claim 48, wherein said vascular intervention comprises angioplasty.

50. A method according to Claim 48, wherein said vascular intervention comprises stent placement.

15

51. A method according to Claim 48, wherein said vascular intervention comprises angioplasty and stent placement.

52. A method of inhibiting narrowing of the lumen of a
20 vessel in a mammal, comprising administering to said
mammal an effective amount of an antibody or antigen-
binding fragment thereof which binds to a mammalian CC-
chemokine receptor 2, wherein said antibody or antigen-
binding fragment thereof inhibits binding of a ligand
25 to the receptor.

53. A method according to Claim 52 wherein the antibody or antigen-binding fragment thereof is selected from the group consisting of

 - a) monoclonal antibody 1D9;
 - b) an antibody having the epitopic specificity of monoclonal antibody 1D9.

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- c) an antibody which can compete with monoclonal antibody 1D9 for binding to mammalian CC-chemokine receptor 2;
 - d) monoclonal antibody 8G2;
 - 5 e) an antibody having the epitopic specificity of monoclonal antibody 8G2;
 - f) an antibody which can compete with monoclonal antibody 8G2 for binding to mammalian CC-chemokine receptor 2;
 - 10 g) antigen-binding fragments of any one of (a) through (g) which bind to mammalian CC-chemokine receptor 2 or a portion thereof; and
 - h) combinations of (a) to (g).
- 15 54. A method of inhibiting neointimal hyperplasia of a vessel in a mammal, comprising administering to said mammal an effective amount of an antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2, wherein said antibody or antigen-binding fragment thereof inhibits binding of a ligand to the receptor.
- 20
55. A method according to Claim 54 wherein the antibody or antigen-binding fragment thereof is selected from the group consisting of:
- 25 a) monoclonal antibody 1D9;
 - b) an antibody having the epitopic specificity of monoclonal antibody 1D9;
 - c) an antibody which can compete with monoclonal antibody 1D9 for binding to mammalian CC-chemokine receptor 2;
 - 30 d) monoclonal antibody 8G2;

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- e) an antibody having the epitopic specificity of monoclonal antibody 8G2;
- f) an antibody which can compete with monoclonal antibody 8G2 for binding to mammalian CC-chemokine receptor 2;
- 5 g) antigen-binding fragments of any one of (a) through (f) which bind to mammalian CC-chemokine receptor 2; and
- h) combinations of (a) to (g).

10

56. A method according to Claim 54, wherein said neointimal hyperplasia is associated with vascular intervention in said mammal.

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57. A method according to Claim 56, wherein said vascular intervention comprises angioplasty.

58. A method according to Claim 56, wherein said vascular intervention comprises stent placement.

20

59. A method according to Claim 56, wherein said vascular intervention comprises angioplasty and stent placement.

25

60. An antibody or antigen-binding fragment according to Claim 1, wherein said antibody or antigen-binding fragment is a monoclonal antibody or fragment thereof.

30

61. An antibody or antigen-binding fragment according to Claim 1, wherein said antibody or antigen-binding fragment is a chimeric antibody or fragment thereof.

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62. An antibody or antigen-binding fragment according to Claim 1, wherein said antibody or antigen-binding fragment is a human antibody or fragment thereof.
 63. An antibody or antigen-binding fragment according to Claim 1, wherein said antibody or antigen-binding fragment is a humanized antibody or fragment thereof.
 64. An antibody or antigen-binding fragment according to Claim 1, wherein said antigen-binding fragment is selected from the group consisting of an Fv fragment, an Fab fragment, an Fab' fragment and an F(ab')₂ fragment.
 65. A humanized antibody or antigen-binding fragment thereof which binds to a human CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and wherein said humanized antibody comprises one or more antigen-binding regions of monoclonal antibody 1D9.
 66. A humanized antibody or antigen-binding fragment thereof which binds to a human CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and wherein said humanized antibody

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comprises one or more complementarity-determining regions of monoclonal antibody 1D9.

67. A humanized antibody or antigen binding fragment thereof according to Claim 66, which comprises six complementarity-determining regions of monoclonal antibody 1D9.

68. A composition comprising a humanized 1D9 antibody or antigen-binding fragment thereof which binds to a human CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and an optional physiologically acceptable vehicle.

69. A pharmaceutical composition comprising a humanized 1D9 antibody or antigen-binding fragment thereof which binds to a human CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and a physiologically acceptable vehicle.

70. A recombinant antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to

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the receptor, and wherein said recombinant antibody comprises one or more complementarity-determining regions of monoclonal antibody 1D9.

- 5 71. A recombinant antibody or antigen-binding fragment according to Claim 70, wherein said recombinant antibody comprises six complementarity-determining regions of monoclonal antibody 1D9.

10 72. An antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to said receptor and inhibits one or more functions associated with binding of the chemokine to said receptor, and wherein said antibody or antigen-binding fragment thereof can compete with monoclonal antibody 1D9 for binding to said receptor.

15 73. An antibody or antigen-binding fragment thereof according to Claim 72, wherein said mammalian CC-chemokine receptor 2 is a human CC-chemokine receptor 2.

20 74. An antibody or antigen-binding fragment thereof according to Claim 72, wherein the chemokine is selected from the group consisting of MCP-1, MCP-2, MCP-3, MCP-4 and combinations thereof.

25 75. An antibody or antigen-binding fragment according to Claim 72, wherein said antibody or fragment is a monoclonal antibody or fragment thereof.

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76. An antibody or antigen-binding fragment according to
Claim 72, wherein said antibody or antigen-binding
fragment is a chimeric antibody or fragment thereof.

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77. An antibody or antigen-binding fragment according to
Claim 72, wherein said antibody or antigen-binding
fragment is a human antibody or fragment thereof.

10 78. An antibody or antigen-binding fragment according to
Claim 72, wherein said antibody or antigen-binding
fragment is a humanized antibody or fragment thereof.

15 79. An antibody or antigen-binding fragment according to
Claim 72, wherein said antigen-binding fragment is
selected from the group consisting of an Fv fragment,
an Fab fragment, an Fab' fragment and an Fab(ab')₂
fragment.

20 80. A composition comprising an antibody or antigen-binding
fragment thereof which binds to a mammalian CC-
chemokine receptor 2 amino-terminal domain, wherein
said antibody or antigen-binding fragment thereof
inhibits binding of a chemokine to said receptor and
inhibits one or more functions associated with binding
of the chemokine to said receptor, and wherein said
antibody or antigen-binding fragment thereof can
compete with monoclonal antibody 1D9 for binding to
said receptor, and an optional physiologically
acceptable vehicle.

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81. A pharmaceutical composition comprising an antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to said receptor and inhibits one or more functions associated with binding of the chemokine to said receptor, and wherein said antibody or antigen-binding fragment thereof can compete with monoclonal antibody 1D9 for binding to said receptor, and a physiologically acceptable vehicle.
- 5
82. A humanized antibody or antigen-binding fragment thereof which binds to a human CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and wherein said humanized antibody
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- 15
- 20
- 25
- 30
83. A humanized antibody or antigen-binding fragment thereof which binds to a human CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and wherein said humanized antibody comprises one or more complementarity-determining regions of monoclonal antibody 8G2.

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84. A humanized antibody or antigen binding fragment thereof according to Claim 83, which comprises six complementarity-determining regions of monoclonal antibody 8G2:

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85. A composition comprising a humanized 8G2 antibody or antigen-binding fragment thereof which binds to a human CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and an optional physiologically acceptable vehicle.

15 86. A pharmaceutical composition comprising a humanized 8G2 antibody or antigen-binding fragment thereof which binds to a human CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and a physiologically acceptable vehicle.

20 87. A recombinant antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and wherein said recombinant antibody comprises one or more complementarity-determining regions of monoclonal antibody 8G2.

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88. A recombinant antibody or antigen-binding fragment according to Claim 87, wherein said recombinant antibody comprises six complementarity-determining regions of monoclonal antibody 8G2.

89. An antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to said receptor and inhibits one or more functions associated with binding of the chemokine to said receptor, and wherein said antibody or antigen-binding fragment thereof can compete with monoclonal antibody 8G2 for binding to said receptor.

90. An antibody or antigen-binding fragment thereof according to Claim 89, wherein said mammalian CC-chemokine receptor 2 is a human CC-chemokine receptor 2.

91. An antibody or antigen-binding fragment thereof according to Claim 89, wherein the chemokine is selected from the group consisting of MCP-1, MCP-2, MCP-3, MCP-4 and combinations thereof.

92. An antibody or antigen-binding fragment according to Claim 89, wherein said antibody or antigen-binding fragment is a monoclonal antibody or fragment thereof.

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93. An antibody or antigen-binding fragment according to
Claim 89, wherein said antibody or antigen-binding
fragment is a chimeric antibody or fragment thereof.

5 94. An antibody or antigen-binding fragment according to
Claim 89, wherein said antibody or antigen-binding
fragment is a human antibody or fragment thereof.

10 95. An antibody or antigen-binding fragment according to
Claim 89, wherein said antibody or antigen-binding
fragment is a humanized antibody or fragment thereof.

15 96. An antibody or antigen-binding fragment according to
Claim 89, wherein said antigen-binding fragment is
selected from the group consisting of an Fv fragment,
an Fab fragment, an Fab' fragment and an F(ab')₂
fragment.

20 97. A composition comprising an antibody or antigen-binding
fragment thereof which binds to a mammalian CC-
chemokine receptor 2, wherein said antibody or antigen-
binding fragment thereof inhibits binding of a
chemokine to said receptor and inhibits one or more
functions associated with binding of the chemokine to
said receptor, and wherein said antibody or antigen-
binding fragment thereof can compete with monoclonal
antibody 8G2 for binding to said receptor, and an
optional physiologically acceptable vehicle.

25 30 98. A pharmaceutical composition comprising an antibody or
antigen-binding fragment thereof which binds to a
mammalian CC-chemokine receptor 2, wherein said

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antibody or antigen-binding fragment thereof inhibits binding of a chemokine to said receptor and inhibits one or more functions associated with binding of the chemokine to said receptor, and wherein said antibody or antigen-binding fragment thereof can compete with monoclonal antibody 8G2 for binding to said receptor, and a physiologically acceptable vehicle.

5 99. An antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a ligand to the receptor and inhibits one or more functions associated with binding of the ligand to the receptor

10 15 at a concentration of less than about 10 µg/ml.

100. An antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a ligand to the receptor and inhibits one or more functions associated with binding of the ligand to the receptor

20 25 at a concentration of less than about 0.1 µg/ml.

25 101. An antibody or antigen-binding fragment thereof which binds to a human CC-chemokine receptor 2, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to said receptor and inhibits one or more functions associated with binding of the chemokine to said receptor, and wherein said antibody or antigen-binding fragment thereof binds the amino-terminal domain of said receptor.

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102. An antibody or antigen-binding fragment according to
Claim 101, wherein said antibody or antigen-binding
fragment is a monoclonal antibody or fragment thereof.

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103. An antibody or antigen-binding fragment according to Claim 101, wherein said antibody or antigen-binding fragment is a chimeric antibody or fragment thereof.

10 104. An antibody or antigen-binding fragment according to
Claim 101, wherein said antibody or antigen-binding
fragment is a human antibody or fragment thereof.

105. An antibody or antigen-binding fragment according to
15 Claim 101, wherein said antibody or antigen-binding
fragment is a humanized antibody or fragment thereof.

106. An antibody or antigen-binding fragment thereof
according to 'Claim 101, wherein the chemokine is
20 selected from the group consisting of MCP-1, MCP-2,
MCP-3, MCP-4 and combinations thereof.

107. A composition comprising an antibody or antigen-binding
fragment thereof which binds to a human CC-chemokine
25 receptor 2, wherein said antibody or antigen-binding
fragment thereof inhibits binding of a chemokine to
said receptor and inhibits one or more functions
associated with binding of the chemokine to said
receptor, and wherein said antibody or antigen-binding
30 fragment thereof binds to the amino-terminal domain of
said receptor, and an optional physiologically
acceptable vehicle.

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108. A pharmaceutical composition comprising an antibody or antigen-binding fragment thereof which binds to a human CC-chemokine receptor 2, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to said receptor and inhibits one or more functions associated with binding of the chemokine to said receptor, and wherein said antibody or antigen-binding fragment thereof binds to the amino-terminal domain of said receptor, and a physiologically acceptable vehicle.
109. A pharmaceutical composition comprising an antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to said receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and a physiologically acceptable vehicle.
110. A method according to Claim 38 wherein the disorder is asthma.
111. The recombinant antibody or antigen-binding fragment thereof according to Claim 70, wherein said mammalian CC-chemokine receptor 2 is a human CC-chemokine receptor 2.
112. The recombinant antibody or antigen-binding fragment thereof according to Claim 87, wherein said mammalian

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CC-chemokine receptor 2 is a human CC-chemokine receptor 2.

113. A recombinant antibody or antigen-binding fragment
5 thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and wherein said recombinant antibody comprises one or more antigen-binding regions of monoclonal antibody 1b9.

114. A recombinant antibody or antigen-binding fragment
15 thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor, and wherein said recombinant antibody comprises one or more antigen-binding regions of
20 monoclonal antibody 8G2.

115. A test kit for use in detecting the presence of a
25 mammalian CC-chemokine receptor 2 or a portion of said
receptor comprising an amino-terminal domain thereof,
in a biological sample comprising
a) at least one antibody or antigen-binding fragment
thereof which binds to the amino-terminal domain
30 of mammalian CC-chemokine receptor 2, wherein said
antibody or antigen-binding fragment thereof
inhibits binding of a chemokine to the receptor

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and inhibits one or more functions associated with binding of the chemokine to the receptor; and

- 5 b) one or more ancillary reagents suitable for detecting the presence of a complex between said antibody or antigen-binding fragment thereof and said mammalian CC-chemokine receptor 2 or a portion of said receptor comprising an amino-terminal domain thereof.

10 116. A method of detecting expression of mammalian CC-chemokine receptor 2 or portion of said receptor comprising an amino-terminal domain thereof by a cell, comprising:

- 15 a) contacting a composition comprising a cell to be tested with an antibody or antigen-binding fragment thereof which binds to a mammalian CC-chemokine receptor 2 amino-terminal domain, wherein said antibody or antigen-binding fragment thereof inhibits binding of a chemokine to the receptor and inhibits one or more functions associated with binding of the chemokine to the receptor; and

- 20 b) detecting binding of said antibody or antigen-binding fragment thereof, wherein the binding of said antibody or antigen-binding fragment thereof indicates the presence of said receptor or portion of said receptor comprising an amino-terminal domain thereof on said cell.

25 117. The method according to Claim 44, wherein the antibody or antigen-binding fragment thereof binds to the mammalian CC-chemokine receptor 2 amino-terminal

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domain, or a portion of mammalian CC-chemokine receptor 2 comprising the amino terminal domain.

118. The method according to Claim 46, wherein the antibody
5 or antigen-binding fragment thereof binds to the
mammalian CC-chemokine receptor 2 amino-terminal
domain, or a portion of mammalian CC-chemokine receptor
2 comprising the amino terminal domain.

10 119. The method according to Claim 52, wherein the antibody or antigen-binding fragment thereof binds to the mammalian CC-chemokine receptor 2 amino-terminal domain, or a portion of mammalian CC-chemokine receptor 2 comprising the amino terminal domain.

15

120. The method according to Claim 54, wherein the antibody or antigen-binding fragment thereof binds to the mammalian CC-chemokine receptor 2 amino-terminal domain, or a portion of mammalian CC-chemokine receptor 2 comprising the amino terminal domain.

121. The composition according to Claim 97, wherein the antibody or antigen-binding fragment thereof binds to the mammalian CC-chemokine receptor 2 amino-terminal domain, or a portion of mammalian CC-chemokine receptor 2 comprising the amino terminal domain.

122. The pharmaceutical composition according to Claim 98, wherein the antibody or antigen-binding fragment thereof binds to the mammalian CC-chemokine receptor 2 amino-terminal domain, or a portion of mammalian CC-

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chemokine receptor 2 comprising the amino terminal domain.

123. An antibody or antigen-binding fragment according to
5 any one of claims 1 to 4, 33 to 37, 60 to 67, 70 to 79,
82 to 84, 87 to 96, 99 to 106 or 111 to 114, a
hybridoma cell line according to any one of claims 5 to
10 6, a monoclonal antibody according to any one of claims
7 to 8, a test kit according to claim 11 or 115, a
method according to any one of claims 10 to 31, 38 to
59, 110 or 116 to 120 or a composition according to
claim 32, 68 to 69, 80 to 81, 85 to 86, 97 to 98, 107
to 109 or 121 to 122 substantially as hereinbefore
15 described with reference to the Figures and/or
Examples.

DATED this 14th day of July 2003

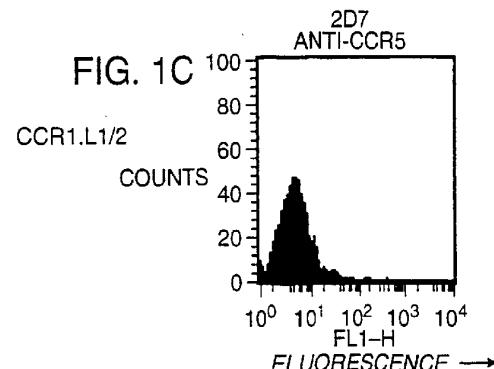
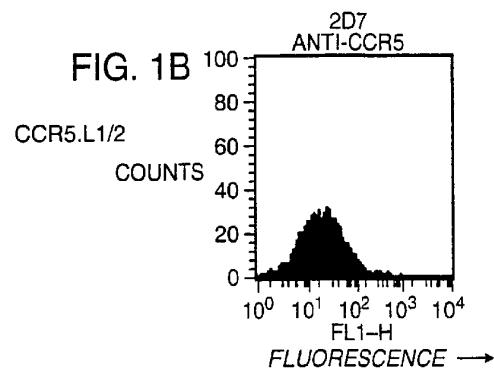
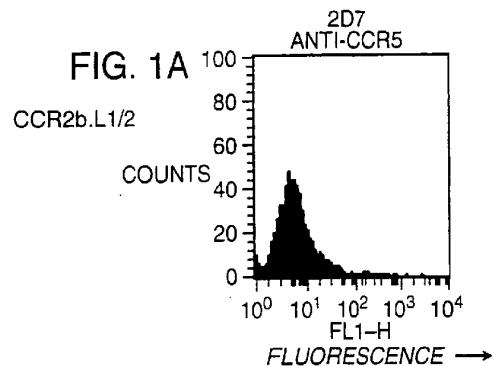
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by DAVIES COLLISON CAVE

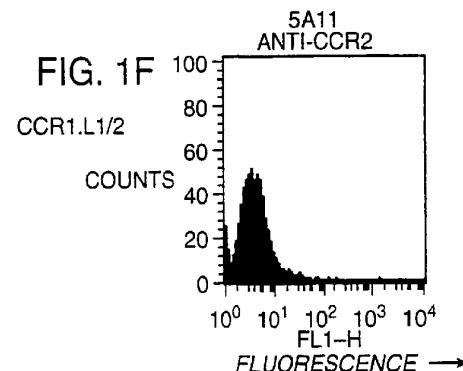
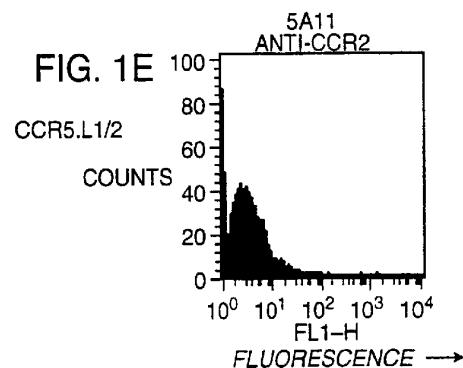
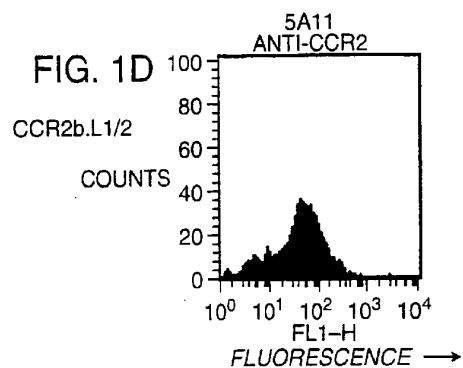
20 Patent Attorneys for the Applicants

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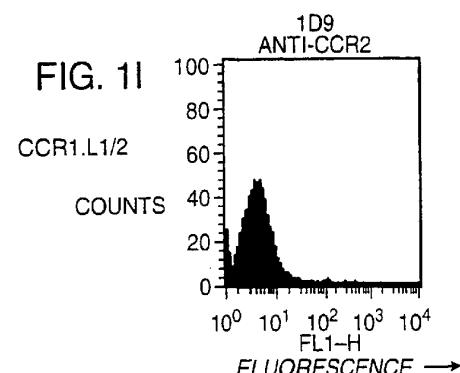
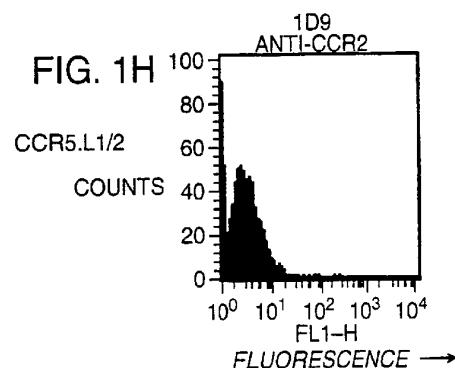
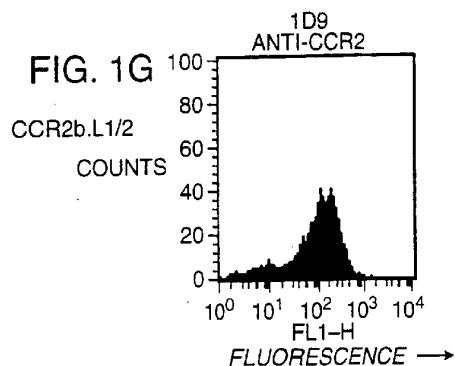
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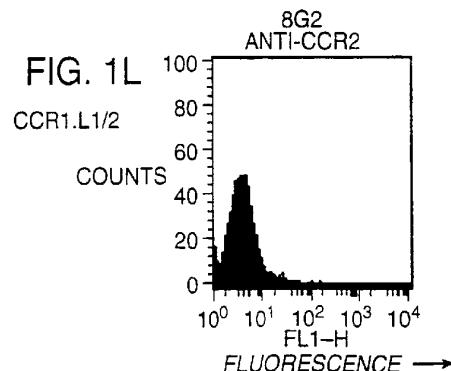
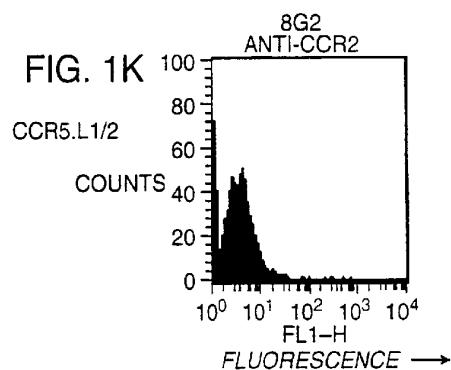
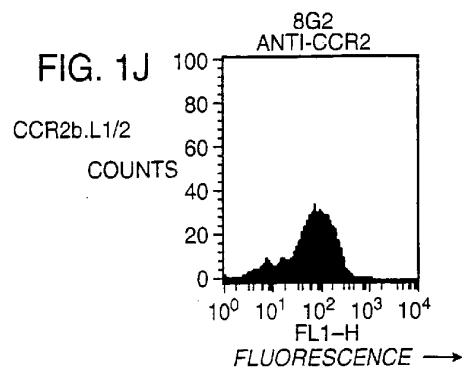
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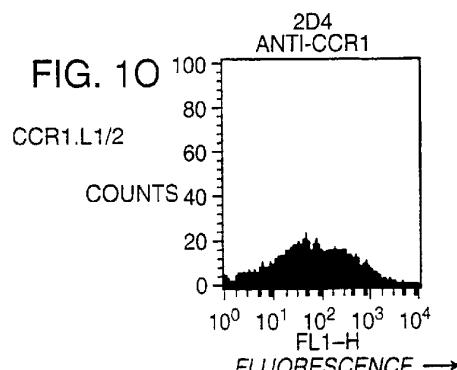
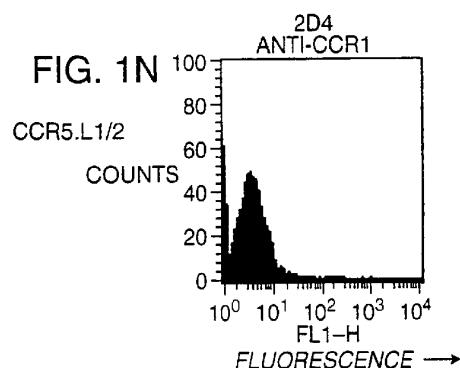
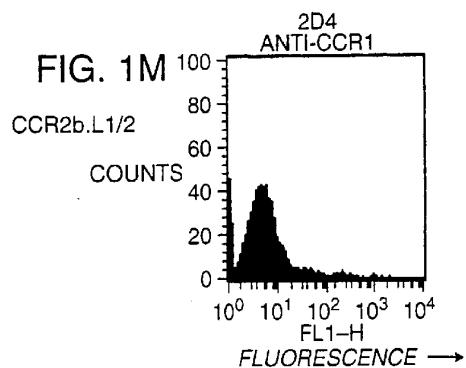


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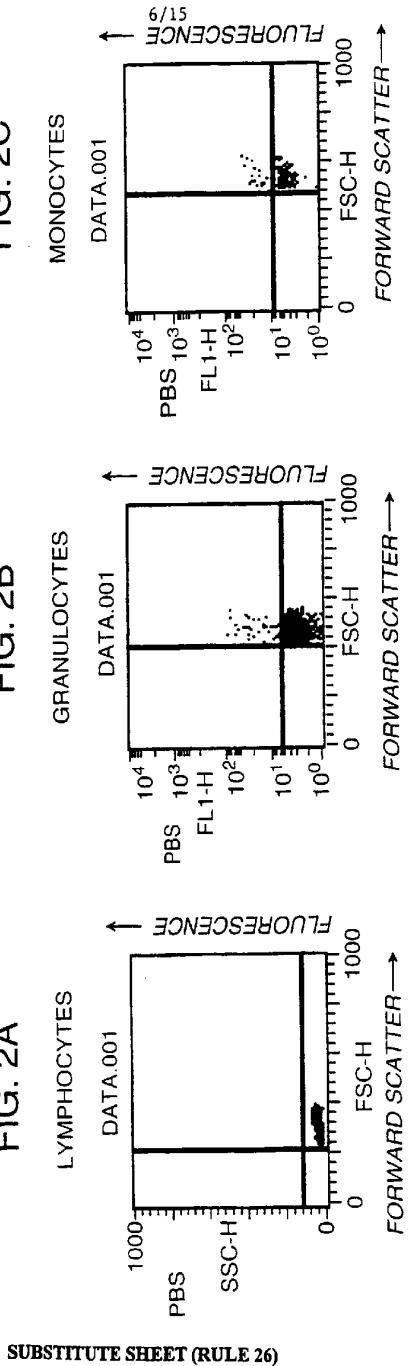


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FIG. 2A
FIG. 2B
FIG. 2C

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FIG. 2F

FIG. 2E

FIG. 2D

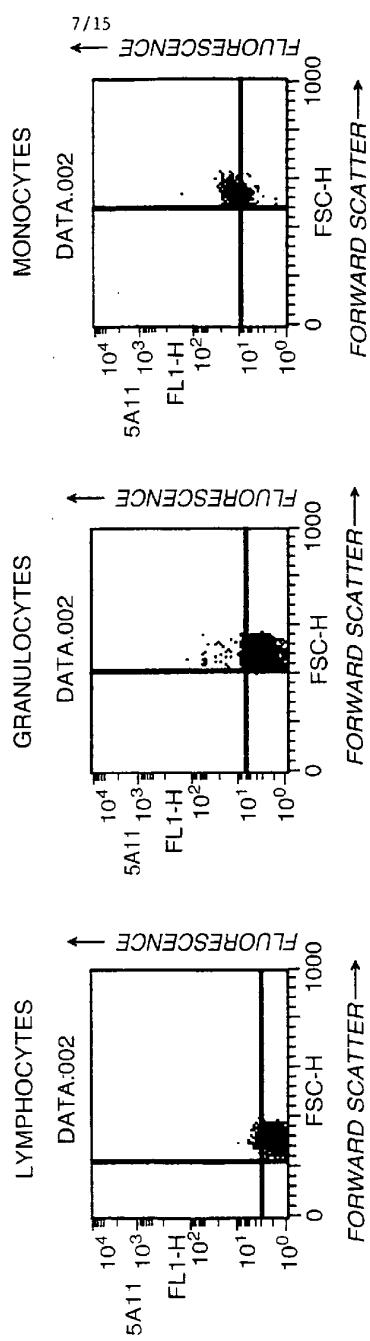


FIG. 2I

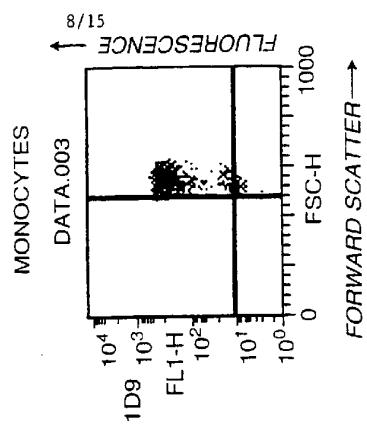


FIG. 2H

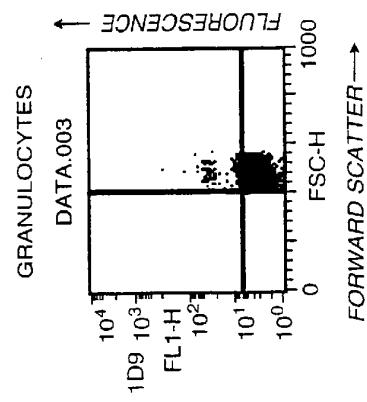
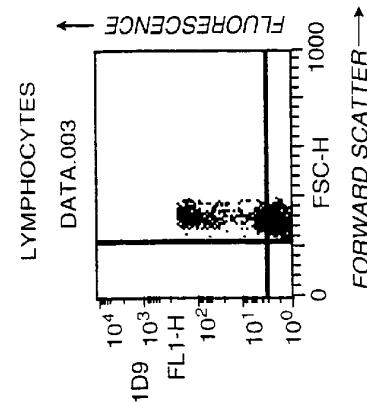


FIG. 2G

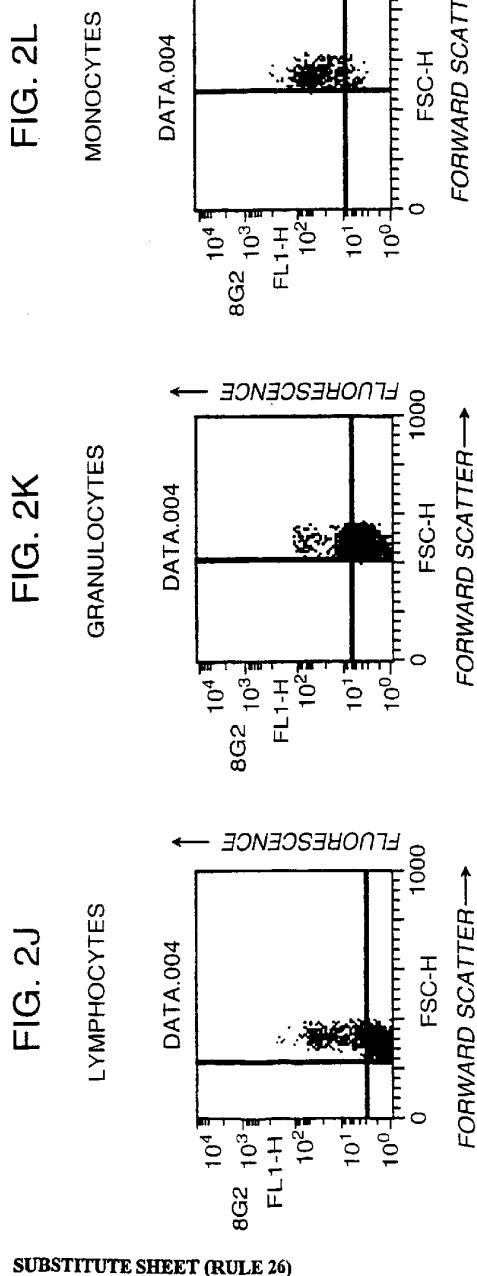


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FIG. 2L

FIG. 2K

FIG. 2J



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FIG. 3A

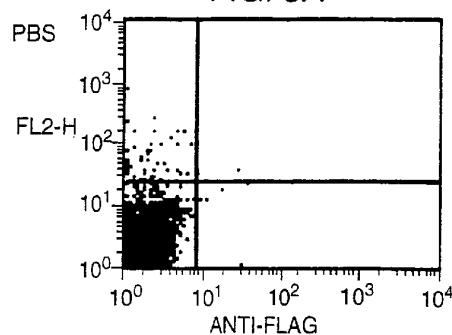


FIG. 3B

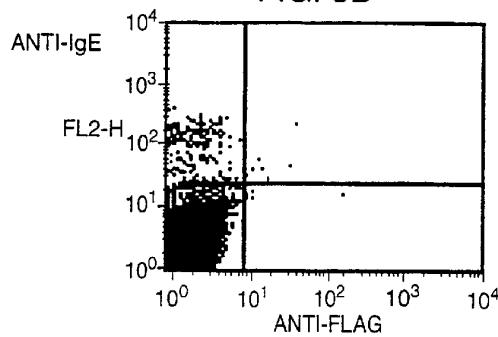
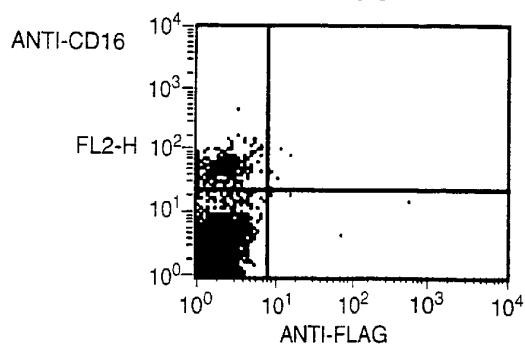


FIG. 3C



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FIG. 3D

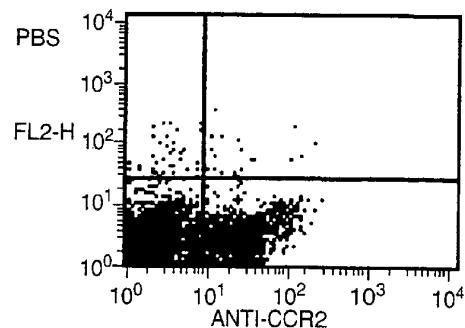


FIG. 3E

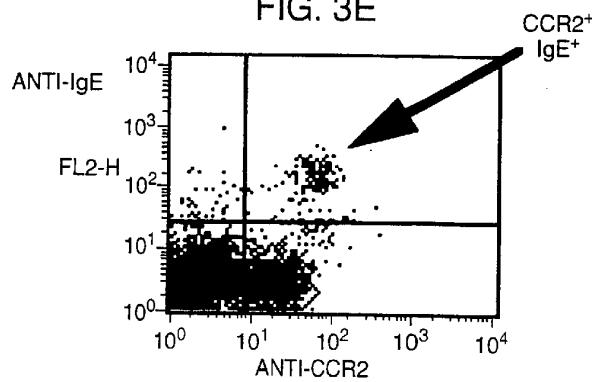


FIG. 3F

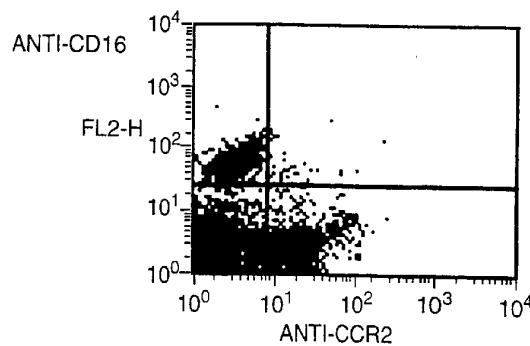


FIG. 3G

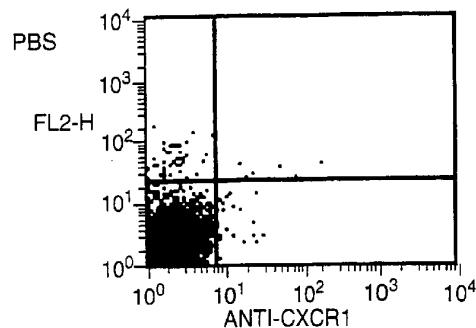


FIG. 3H

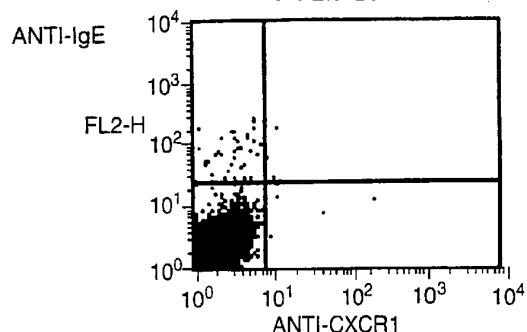
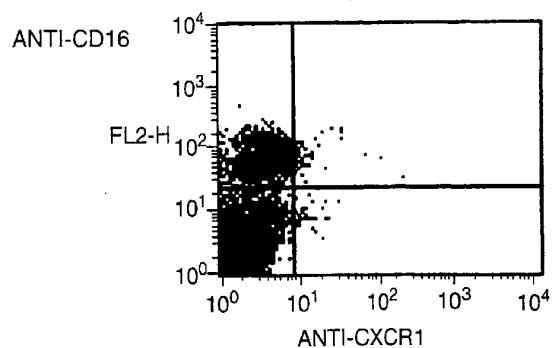


FIG. 3I



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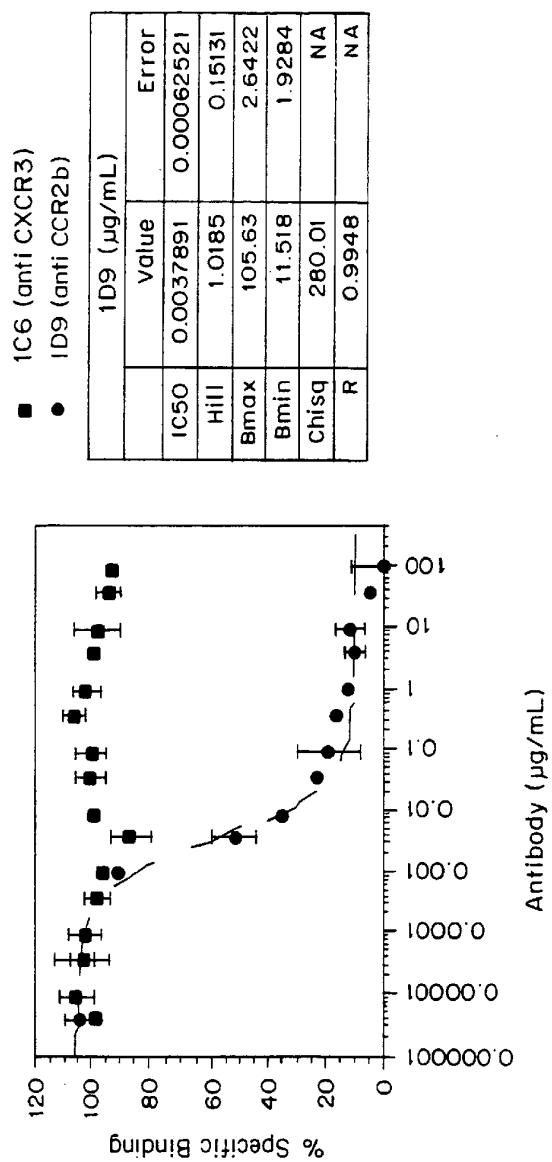


FIG. 4

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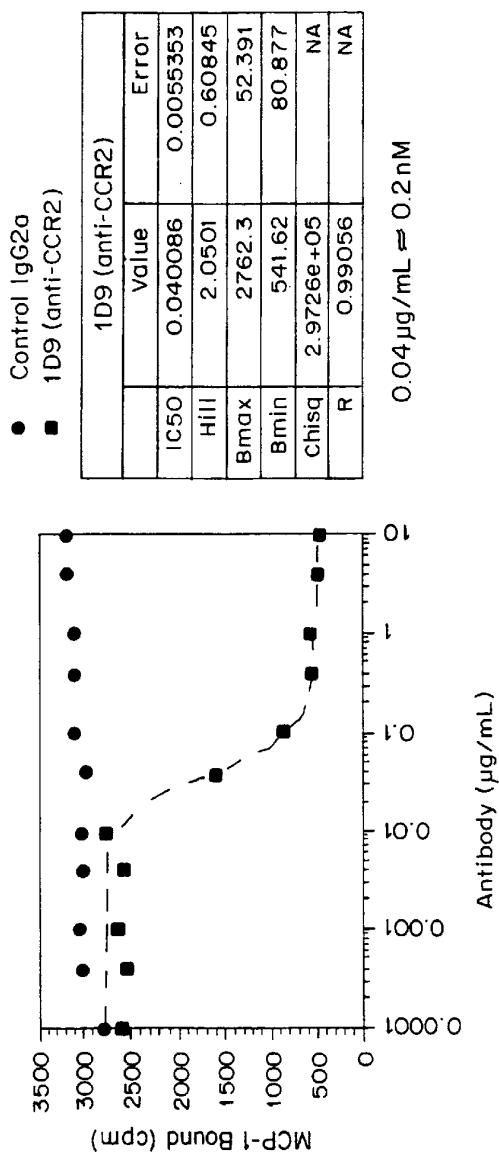
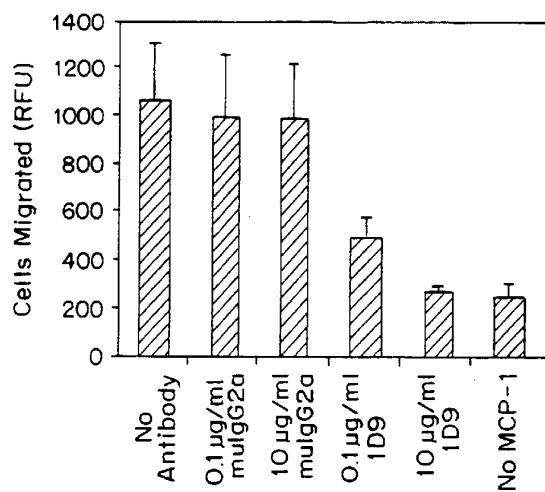
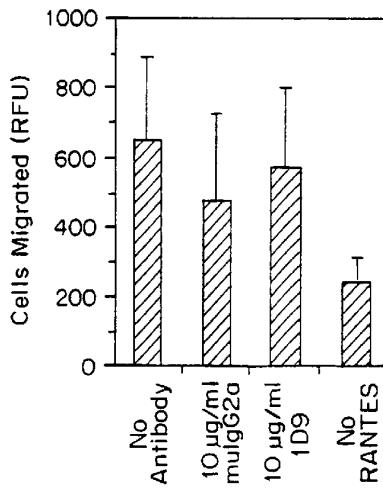


FIG. 5

**FIG. 6A****FIG. 6B****SUBSTITUTE SHEET (RULE 26)**