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- (54) **Madárinfluenza kiméra VLP-k**

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(54) **Avian influenza chimeric VLPS**

Vogel-Influenza-VLP-Chimären

Pseudo-particules virales de la grippe aviaire chimère

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EP 2 540 312 B1

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Description**BACKGROUND OF THE INVENTION**

5 **[0001]** Influenza virus is a member of Orthomyxoviridae family (for review, see Murphy and Webster, 1996). There are three subtypes of influenza viruses designated A, B, and C. The influenza virion contains a segmented negative-sense RNA genome. The influenza virion includes the following proteins: hemagglutinin (HA), neuraminidase (NA), matrix (M1), proton ion-channel protein (M2), nucleoprotein (NP), polymerase basic protein 1 (PB1), polymerase basic protein 2 (PB2), polymerase acidic protein (PA), and nonstructural protein 2 (NS2) proteins. The HA, NA, M1, and M2
 10 are membrane associated, whereas NP, PB1, PB2, PA, and NS2 are nucleocapsid associated proteins. The NS1 is the only nonstructural protein not associated with virion particles but specific for influenza-infected cells. The M1 protein is the most abundant protein in influenza particles. The HA and NA proteins are envelope glycoproteins, responsible for virus attachment and penetration of the viral particles into the cell, and the sources of the major immunodominant epitopes for virus neutralization and protective immunity. Both HA and NA proteins are considered the most important components
 15 for prophylactic influenza vaccines because they are highly immunogenic.

[0002] To date, all commercially available influenza vaccines for non-pandemic strains in the United States have been propagated in embryonated hen's eggs. Although influenza virus grows well in hen's eggs, production of vaccine is dependent on the availability of eggs. Supplies of eggs must be organized, and strains for vaccine production selected months in advance of the next flu season, limiting the flexibility of this approach, and often resulting in delays and
 20 shortages in production and distribution. Unfortunately, some influenza vaccine strains, do not replicate well in embryonated chicken eggs, and have to be isolated by cell culture in a costly and time consuming procedure.

[0003] Systems for producing influenza viruses in cell culture have also been developed in recent years (See, e.g., Furminger. Vaccine Production, in Nicholson et al. (eds) Textbook of Influenza pp. 324-332; Merten et al. (1996) Production of influenza virus in cell cultures for vaccine preparation, in Cohen & Shafferman (eds) Novel Strategies in Design and Production of Vaccines pp. 141-151). Typically, these methods involve the infection of suitable immortalized host
 25 cells with a selected strain of virus. While eliminating many of the difficulties related to vaccine production in hen's eggs, not all pathogenic strains of influenza grow well and can be produced according to established tissue culture methods. In addition, many strains with desirable characteristics, e.g., attenuation, temperature sensitivity and cold adaptation, suitable for production of live attenuated vaccines, have not been successfully grown in tissue culture using established
 30 methods. In addition, live attenuated viruses have not been accepted by the general public due to fears reversion to a virulent virus.

[0004] Virus like particles mimic the overall structure of a virus particle without the requirement of containing infectious material. VLPs lack a viral DNA or RNA genome, but retain the three-dimensional structure of an authentic virus. VLPs have the ability to stimulate B-cell mediated responses, CD4 proliferative responses and cytotoxic T lymphocytes responses (see, Schirmbeck et al. (1996) Eur. J. Immunol., 26, 2812-2822). In addition, virus like particles induce MHC
 35 class I-restricted T-cell responses.

SUMMARY OF THE INVENTION

40 **[0005]** The present invention comprises a method of increasing the efficiency of influenza VLP production comprising expressing an avian influenza M1 and seasonal influenza HA and NA proteins in a host cell. The avian influenza M1 protein is an A/Indonesia/5/05 influenza M1 protein. Said HA or NA may have hemagglutinin and neuraminidase activity, respectively.

[0006] The present application also describes a chimeric VLP comprising an avian influenza M1 protein and at least one non-avian influenza protein. In one embodiment, said non-avian influenza protein is HA and/or NA from a non-avian influenza virus. In another embodiment, said non-avian influenza protein is a seasonal influenza protein. In another
 45 embodiment, said HA or NA has hemagglutinin or neuraminidase activity, respectively. In another embodiment, said HA and/or NA are chimeric proteins. In another embodiment, chimeric proteins comprise external domains of non-avian influenza HA and/or NA protein sequences fused to the transmembrane and/or cytoplasmic-terminal domains of the avian or heterologous influenza HA and/or NA. In another embodiment, said non-avian influenza protein is from an infectious agent.
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[0007] The present application also describes an antigenic formulation comprising a chimeric VLP comprising an avian influenza M1 protein and at least one non-avian influenza protein. In one embodiment, said VLP comprises HA and/or NA from a non-avian influenza virus. In another embodiment, said HA or NA have hemagglutinin and neuraminidase activity, respectively. In another embodiment, said HA and/or NA are chimeric proteins.
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[0008] The present application also describes vaccines comprising a chimeric VLP comprising an avian influenza M1 protein and at least one non-avian influenza protein. In one embodiment, said VLP comprises HA and/or NA from a non-avian influenza virus. In another embodiment, said HA or NA have hemagglutinin and neuraminidase activity, respectively.

In another embodiment, said HA and/or NA are chimeric proteins.

[0009] The present application also describes a method of inducing immunity in a vertebrate comprising administering to said vertebrate a chimeric VLP comprising an avian influenza M1 protein and at least one non-avian influenza protein. In one embodiment, said immune response is a humoral immune response. In one embodiment, said immune response is a cellular immune response.

[0010] The present application also describes a method of preventing and/or reducing a viral infection or symptom thereof, comprising administering to a vertebrate a chimeric VLP comprising an avian influenza M1 protein and at least one non-avian influenza protein.

[0011] The present application also describes a method of reducing the severity of influenza in a population, comprising administering the a chimeric VLP comprising an avian influenza M1 protein and at least one non-avian influenza protein to enough individuals in said population in order to prevent or decrease the chance influenza virus transmission to another individual in said population.

BRIEF DESCRIPTION OF THE FIGURES

[0012]

Figure 1 depicts a stained SDS-PAGE gel derived from VLPs made from different constructs after isolation from a sucrose gradient.

Figure 2 depicts a stained western blot derived from VLPs made from different constructs after isolation from a sucrose gradient.

Figure 3 is a stained SDS-PAGE gel derived from VLPs made from wild type or hybrids of A/Indonesia/5/05 M1 and A/Fujian/411/2002 HA and NA.

Figure 4 depicts a stained western blot derived from VLPs made from wild type or hybrids of A/Indonesia/5/05 M1 and A/Fujian/411/2002 HA and NA.

Figure 5 depicts the amino acids sequence of SARS S protein with Indonesia H5N1 HA transmembrane and carboxyl terminal domain (underlined).

Figure 6 depicts the amino acids sequence of Indonesia H5N1 M1 protein.

Figure 7 depicts pFastBac 1 vector containing coding sequences for SARS S with Indonesia H5N1 HA TM/CT domain and Indonesia H5N1 M1 protein.

Figure 8 depicts the purified SARS S/Indo M1 chimeric VLPs. Lane 1 is Coomassie blue stain. Lane 2 is western blot, top panel: anti SARS S; bottom panel: anti influenza M1.

Figure 9 depicts purified wild type SARS VLPs composed of SARS S, M and E proteins. A) Coomassie blue stain; B) Western blot, top panel: anti SARS S; bottom panel: anti SARS M.

Figure 10 depicts particle size analysis result for SARS S/Indo M1 chimeric VLPs with Malvern Zetasizer.

Figure 11A-C depicts electron microscope (EM) negative stain of SARS S/Indo M1 chimeric VLPs. A) EM image for buffer control; B) Selected EM images for VLPs; C) Selected EM images for VLPs at higher magnitude.

Figure 12A-C depicts Published EM images for SARS-CoV and coronavirus.

Figure 13 depicts expression constructs for production of B/Florida/4/06 VLPs in Sf9 insect cells. Shown are the location of HA, NA, and M1 genes, as well as locations of polyhedron promoter. Also shown are the constructs for individual expression of HA and NA genes for reagent purposes.

Figure 14 depicts expression levels of influenza B/Florida/4/06 VLPs by Coomassie staining (left panel) and HA/NA assays (right panel). Lane 1. Sample of B/Florida/4/06 VLPs containing B/Florida/4/06 M1, Lane 2. Sample of B/Florida/4/06 VLPs containing B/Ann Arbor/1/1986 M1, Lane 3 Sample of B/Florida/4/06 VLPs containing A/Indonesia/5/05 (H5N1) M1. The right panels shows HA and NA activity by the hemagglutination and neuraminidase enzyme activity assays.

Figure 15 depicts Electron microscopy of purified VLPs. Negative staining transmission electron microscopy of influenza B/Florida/4/06 VLPs containing M1 from A/Indonesia/5/05 (H5N1) (left), B/Ann Arbor/1/1986 (middle), and B/Florida/4/06 (right).

DETAILED DESCRIPTION

[0013] As use herein, the term "antigenic formulation" or "antigenic composition" refers to a preparation which, when administered to a vertebrate, especially a bird or a mammal, will induce an immune response.

[0014] As used herein the term "adjuvant" refers to a compound that, when used in combination with a specific immunogen (e.g. a VLP) in a formulation, augments or otherwise alters or modifies the resultant immune response. Modification of the immune response includes intensification or broadening the specificity of either or both antibody and cellular immune responses. Modification of the immune response can also mean decreasing or suppressing certain antigen-

specific immune responses.

[0015] As used herein the term "avian influenza virus" refers to influenza viruses found chiefly in birds but that can also infect humans or other animals. In some instances, avian influenza viruses may be transmitted or spread from one human to another. An avian influenza virus that infects humans has the potential to cause an influenza pandemic, i.e., morbidity and/or mortality in humans. A pandemic occurs when a new strain of influenza virus (a virus in which human have no natural immunity) emerges, spreading beyond individual localities, possibly around the globe, and infecting many humans at once.

[0016] As used herein, the term "chimeric protein" refers a constructs that links at least two heterologous proteins into a single macromolecule (fusion protein).

[0017] As used herein, the term "chimeric VLP" refers to a virus-like particle that comprises an avian M1 protein and at least one protein, or portion thereof, that is not from an avian influenza virus.

[0018] As used herein an "effective dose" generally refers to that amount of the VLP of the invention sufficient to induce immunity, to prevent and/or ameliorate influenza virus infection or to reduce at least one symptom of influenza infection and/or to enhance the efficacy of another dose of a VLP. An effective dose may refer to the amount of the VLP sufficient to delay or minimize the onset of an influenza infection. An effective dose may also refer to the amount of the VLP that provides a therapeutic benefit in the treatment or management of influenza infection. Further, an effective dose is the amount with respect to the VLPs of the invention alone, or in combination with other therapies, that provides a therapeutic benefit in the treatment or management of an influenza viral infection. An effective dose may also be the amount sufficient to enhance a subject's (e.g., a human's) own immune response against a subsequent exposure to influenza virus. Levels of immunity can be monitored, e.g., by measuring amounts of neutralizing secretory and/or serum antibodies, e.g., by plaque neutralization, complement fixation, enzyme-linked immunosorbent, or microneutralization assay. In the case of a vaccine, an "effective dose" is one that prevents disease or reduces the severity of symptoms.

[0019] As used herein, the term "external domain" when referring to membrane associated proteins refer to the domain(s) of the protein that are external to the cell and/or cytosol and/or a lumen. The external domain of a protein is also known as an ectodomain.

[0020] As used herein, the term "influenza VLP" refers to a VLP comprising at least one influenza protein. Said VLPs can comprise additional influenza and/or non-influenza proteins.

[0021] As used herein, the term "hemagglutinin activity" refers to the ability of HA-containing proteins, VLPs, or portions thereof to bind and agglutinate red blood cells (erythrocytes).

[0022] As used herein, the term "neuraminidase activity" refers to the enzymatic activity of NA-containing proteins, VLPs, or portions thereof to cleave sialic acid residues from substrates including proteins such as fetuin.

[0023] As use herein, the term "infectious agent" refers to microorganisms that cause an infection in a vertebrate. Usually, the organisms are viruses, bacteria, parasites and/or fungi. The term also refers to different antigenic variations of the same infectious agent.

[0024] As used herein the term "immune stimulator" refers to a compound that enhances an immune response *via* the body's own chemical messengers (cytokines). These molecules comprise various cytokines, lymphokines and chemokines with immunostimulatory, immunopotentiating, and pro-inflammatory activities, such as interleukins (e.g., IL-1, IL-2, IL-3, IL-4, IL-12, IL-13); growth factors (e.g., granulocyte-macrophage (GM)-colony stimulating factor (CSF)); and other immunostimulatory molecules, such as macrophage inflammatory factor, Flt3 ligand, B7.1; B7.2, etc. The immune stimulator molecules can be administered in the same formulation as the influenza VLPs, or can be administered separately. Either the protein or an expression vector encoding the protein can be administered to produce an immunostimulatory effect.

[0025] As used herein the term "immunity" refers to induction of the immune system of a vertebrate wherein said induction results in the prevention, amelioration, and/or reduction of at least one symptom of an infection in said vertebrate. Immunity may also refer to a haemagglutination inhibition (HI) titer of ≥ 40 when VLPs of the invention have been administered to a vertebrate and said VLPs have induced an immune response against a HA of an influenza virus.

[0026] As used herein the term "non-avian influenza protein" refers to a protein that is heterologous to an avian influenza virus. Said non-avian influenza protein may be recombinantly expressed from an expression vector and may be heterologous to the expression vector.

[0027] As used herein the term "seasonal influenza virus" refers to the influenza viral strains that have been determined to be passing within the human population for a given influenza season based on epidemiological surveys conducted by National Influenza Centers worldwide. These epidemiological studies, and some isolated influenza viruses, are sent to one of four World Health Organization (WHO) reference laboratories, one of which is at the Centers for Disease Control and Prevention (CDC) in Atlanta for detailed testing. These laboratories test how well antibodies made to the current vaccine react to the circulating virus and new flu viruses. This information, along with information about flu activity, is summarized and presented to an advisory committee of the U.S. Food and Drug Administration (FDA) and at a WHO meeting. These meetings result in the selection of three viruses (two subtypes of influenza A viruses and one influenza B virus) to go into flu vaccines for the following fall and winter. The selection occurs in February for the northern hemisphere

and in September for the southern hemisphere. Usually, one or two of the three virus strains in the vaccine changes each year.

[0028] As used herein, the term "vaccine" refers to a preparation of dead or weakened pathogens, or of derived antigenic determinants that is used to induce formation of antibodies or immunity against the pathogen. A vaccine is given to provide immunity to the disease, for example, influenza, which is caused by influenza viruses. In addition, the term "vaccine" also refers to a suspension or solution of an immunogen (e.g. VLP) that is administered to a vertebrate to produce protective immunity, i.e., immunity that prevents or reduces the severity of disease associated with infection. The present invention provides for vaccine compositions that are immunogenic and may provide protection against a disease associated with infection.

[0029] As use herein, the term "vertebrate" or "subject" or "patient" refers to any member of the subphylum cordata, including, without limitation, humans and other primates, including non-human primates such as chimpanzees and other apes and monkey species. Farm animals such as cattle, sheep, pigs, goats and horses; domestic mammals such as dogs and cats; laboratory animals including rodents such as mice, rats and guinea pigs; birds, including domestic, wild and game birds such as chickens, turkeys and other gallinaceous birds, ducks, geese, and the like are also non-limiting examples. The terms "mammals" and "animals" are included in this definition. Both adult and newborn individuals are intended to be covered.

[0030] As used herein, the term "virus-like particle" (VLP) refers to a structure that in at least one attribute resembles a virus but which has not been demonstrated to be infectious. Virus-like particle in accordance with the invention do not carry genetic information encoding for the proteins of virus-like particles. In general, virus-like particles lack a viral genome and, therefore, are noninfectious. In addition, virus-like particles can often be produced in large quantities by heterologous expression and can be easily purified.

VLPs of the invention and methods of making VLPs

[0031] In general, virus like particles (VLPs) lack a viral genome and, therefore, are noninfectious. In addition, virus-like particles can often be produced by heterologous expression and can be easily purified. Most VLPs comprise at least a viral core protein. This core protein usually drives budding and release of particles from a host cell. Examples of such proteins comprise RSV M, influenza M1, HIV gag and vesicular stomatis virus (VSV) M protein. In general, VLPs are useful for preparing antigenic formulation and/or vaccines against infectious agents, e.g. influenza.

[0032] However, VLP production has not been particularly efficient. One goal of VLP production is the optimization of culture conditions to obtain the greatest possible productivity. Even incremental increases in productivity can be economically significant and can save lives. The inventors of the present invention have unexpectedly discovered that expressing avian M1 is a host cell significantly enhances production of VLPs from host cells.

[0033] Thus, the invention described herein comprises methods of producing chimeric VLPs comprising an avian A/Indonesia/5/05 influenza M1 protein and seasonal influenza HA and/or NA. In one embodiment, said HA or NA seasonal influenza are A/Wisconsin/67/2005 and/or A/Fujian/411/02. In another embodiment, said HA or NA has hemagglutinin or neuraminidase activity, respectively.

[0034] Chimeric VLPs are useful for preparing vaccines and immunogenic compositions. One important feature of said chimeric VLPs is the ability to express proteins on the surface of said VLPs so that the immune system of a vertebrate can induce an immune response against said protein. However, not all proteins can be expressed on the surface of VLPs. There may be many reasons why certain proteins are not expressed, or poorly expressed, on the surface of VLPs. One reason is that said protein is not directed to the membrane of a host cell or that said protein does not have a transmembrane domain. Sequences near the carboxyl terminus of influenza hemagglutinin may be important for incorporation of HA into the lipid bilayer of the mature influenza enveloped nucleocapsids and for the assembly of HA trimer interaction with the influenza core protein M1 (Ali, et al., (2000) J. Virol. 74, 8709-19). Thus, one method of overcoming the inability of expressing non-avian influenza proteins on the surface of VLPs, and/or increasing the expression of said proteins, is to fuse the cytoplasmic and/or the transmembrane domains of influenza HA and/or NA to a non-avian influenza protein thus creating a chimeric protein.

[0035] Thus, described herein are chimeric VLPs comprising at least one chimeric protein. Said chimeric protein may comprise at least one external domain (ectodomain) of non-avian influenza HA and/or NA protein sequences fused to the transmembrane and/or cytoplasmic-terminal domains of a heterologous HA and/or NA. In another embodiment, said heterologous transmembrane and/or cytoplasmic-terminal domains HA and/or NA is from seasonal influenza and/or avian influenza virus. In another embodiment, said non-avian influenza HA and/or NA are from a seasonal influenza strain A/Wisconsin/67/2005 and HA and/or NA transmembrane and/or cytoplasmic-terminal domains are from an avian influenza strain. In another embodiment, said non-avian influenza HA and/or NA are from influenza strain A/Fujian/411/02 and HA and/or NA transmembrane and/or cytoplasmic-terminal domains are from an avian influenza strain. Said HA and/or NA transmembrane and/or cytoplasmic-terminal domains from avian influenza can be derived from the group consisting of influenza virus H9N2 and/or influenza virus H5N1.

[0036] Said HA and/or NA from H9N2 influenza strain can be isolated from any one of the influenza virus from the group consisting of A/quail/Hong Kong/G1/97, A/Hong Kong/1073/99, A/Hong Kong/2108/03, Duck/HK/Y280/97, CK/HK/G9/97, Gf/HK/SSP607/03, Ph/HK/CSW1323/03, WDK/ST/4808/01, CK/HK/NT142/03, CK/HK/WF126/03, SCK/HK/WF285/03, CK/HK/YU463/03, CK/HK/YU577/03, SCK/HK/YU663/03, Ck/HK/CSW161/03, and GF/HK/NT101/03. In one embodiment, said H9N2 influenza strain is A/Hong Kong/1073/99. In another embodiment, said HA and/or NA from influenza strain H5N1 can be from clade 1 and/or clade 2. In another embodiment, said H5N1 is from clade 1. In another embodiment, said H5N1 is from clade 2. In another embodiment, said H5N1 is selected from the group consisting of A/Vietnam/1194/04, A/Vietnam/1203/04, A/Hongkong/213/03, A/Indonesia/2/2005, A/Bar headed goose/Quinghai/1A/2005, A/Anhui/1/2005, and A/Indonesia/5/05. In another embodiment, said H5N1 strain is A/Indonesia/5/05.

[0037] Chimeric VLPs described herein comprise an avian influenza M1 protein. Said M1 protein can be derived from influenza strain H9N2 or H5N1. Said H9N2 influenza M1 can be isolated from any one of the influenza virus from the group consisting of A/quail/Hong Kong/G1/97, A/Hong Kong/1073/99, A/Hong Kong/2108/03, Duck/HK/Y280/97, CK/HK/G9/97, Gf/HK/SSP607/03, Ph/HK/CSW1323/03, WDK/ST/4808/01, CK/HK/NT142/03, CK/HK/WF126/03, SCK/HK/WF285/03, CK/HK/YU463/03, CK/HK/YU577/03, SCK/HK/YU663/03, Ck/HK/CSW161/03, and GF/HK/NT101/03. In one embodiment, said H9N2 influenza strain is A/Hong Kong/1073/99. In another embodiment, said M1 can be from influenza strain H5N1. In another embodiment, said H5N1 is selected from the group consisting of A/Vietnam/1194/04, A/Vietnam/1203/04, A/Hongkong/213/03, A/Indonesia/2/2005, A/Bar headed goose/Quinghai/1A/2005, A/Anhui/1/2005, and A/Indonesia/5/05. In VLPs produced by methods of the invention, said H5N1 strain is A/Indonesia/5/05.

[0038] In another embodiment, said chimeric VLPs comprise chimeric proteins from influenza B viruses. In one embodiment, said chimeric proteins comprise external domains of influenza B HA and/or NA protein sequences fused to the transmembrane and/or cytoplasmic-terminal domains of a heterologous HA and/or NA cytoplasmic and/or transmembrane region. In another embodiment, said heterologous HA and/or NA is from seasonal influenza A/Wisconsin/67/2005 and/or A/Fujian/411/02 and/or avian influenza A/Indonesia/5/05. In another embodiment, said influenza B viruses are from B/Shanghai/361/2002 and/or B/Hong Kong/330/2001.

[0039] Also described herein are chimeric VLPs comprising an avian M1 with a protein from another infectious agent (non-avian influenza protein). Said protein from another infectious agent can be a type 1 and/or a type 2 protein. A type I protein has a C-terminus located in the cytosol (the transmembrane domain is located near the C-terminus), whereas a type II protein has an N-terminus that is located in the cytosol (the transmembrane domain is located near the N-terminus). In another embodiment, said protein may comprise epitopes that can induce an immune response against said protein when administered to a vertebrate. In another embodiment, said protein can associate with avian influenza M1 directly or indirectly. In another embodiment, said protein is expressed on the surface of the VLP. In another embodiment, said protein, or portion thereof, can be fused to a heterologous protein creating a chimeric protein. For example, the external domains of proteins from infective agents, such as non-avian influenza virus, coronavirus, VZV, Dengue, or yellow fever and/or other agents can be used to generate chimeric proteins by fusing said proteins from infective agents with a protein that associates with avian influenza M1. In one embodiment, said protein that associates with avian influenza M1 is an influenza protein. In another embodiment, said protein that associates with avian M1 is a HA and/or NA from influenza. In another embodiment, said HA and/or NA is from a seasonal influenza virus. In another embodiment, said HA and/or NA is from an avian influenza virus. In another embodiment, said avian influenza virus is H5N1. In another embodiment, said H5N1 strain is A/Indonesia/5/05.

[0040] Also described herein is a VLP comprising a chimeric protein comprising the transmembrane domain and/or cytoplasmic tail of influenza HA and/or influenza NA fused to a protein from an infective agent. In another embodiment, the transmembrane domain and/or cytoplasmic tail of the HA and/or NA protein extends from the N or C-terminus to approximately 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 to about 50 amino acids past the transmembrane domain and is fused to said protein from another infectious agent. In another embodiment, the portion of the protein from another infectious agent that comprises the cytoplasmic and the transmembrane domain is replaced with a cytoplasmic and/or transmembrane domain from an influenza protein (*i.e.* avian and/or seasonal influenza NA and/or HA). In another embodiment, said seasonal influenza HA and/or NA are from influenza strain A/Wisconsin/67/2005 and/or A/Fujian/411/02 and/or avian influenza A/Indonesia/5/05. In another embodiment, said M1 is from an avian influenza strain H5N1. In another embodiment, said M1 is from influenza strain A/Indonesia/5/05. In another embodiment, said M1 is from influenza strain H9N2. In another embodiment, said M1 is from influenza strain A/Hong Kong/1073/99. In another embodiment, the transmembrane domain and/or cytoplasmic tail of A/Wisconsin/67/2005 HA and/or NA is fused to a protein from an infectious agent. In another embodiment, the transmembrane domain and/or cytoplasmic tail of A/Fujian/411/02 HA and/or NA is fused to a protein from an infectious agent. In another embodiment, the transmembrane domain and/or cytoplasmic tail of A/Indonesia/5/05 HA and/or NA is fused to a protein from an infectious agent.

[0041] In another embodiment, the transmembrane domain and/or cytoplasmic tail of influenza HA and/or influenza NA fused to a protein from an infective agent comprises a spacer sequence between the protein segments. Said space

sequences can be any amino acid not in the protein. This spacer sequence may be important for expressing said protein from an infective agent on the surface of the VLP. Examples of spacer sequences include a poly-G amino acids. Said spacer can be from 1 to about 100 amino acids long.

[0042] VLPs produced according to the invention may comprise more than one protein from an infectious agent. In this embodiment, said VLPs are multivariant VLPs capable of inducing an immune response to several proteins from infectious agents. In one embodiment said VLPs comprise proteins from at least two different influenza viruses. For example said multivariant VLPs can comprise a HA and/or NA from a seasonal influenza virus A and/or B. This embodiment also comprises the presentation of HA and/or NA of the three influenza viruses (two subtypes of influenza A viruses and one influenza B virus) that are chosen by WHO and the CDC (see above) to be in the flu vaccines for the fall and winter in a single VLP. In another embodiment, said multivariant VLPs comprise proteins from several viruses, bacteria and/or parasites. For example, said VLPs comprise proteins from influenza and RSV, influenza, RSV and parainfluenza. In another embodiment, said proteins are chimeric proteins wherein each protein comprises the HA and/or NA from an influenza virus. In embodiment according to the present invention, said multivalent VLPs comprise an A/Indonesia/5/05 M1 protein and seasonal influenza HA and NA.

[0043] Also described herein are VLPs comprising chimeric proteins comprising a fusion between the influenza HA with the protein, or a portion thereof, from an infectious agent. In another embodiment, said chimeric proteins comprise a fusion between the proteins, or a portion thereof, of two infectious agents or antigenic variations of the same agent. Said fusion protein will comprise antigenic agents from each protein from said infectious agent. In another embodiment, said chimeric protein comprises an amino acid linker between the proteins. An example of this embodiment is a fusion between the influenza HA and the RSV F protein. An example of this embodiment is a fusion between the influenza HA and the RSV F1 protein (e.g. SEQ ID NO 12). In another embodiment, said chimeric protein comprises the HA and/or NA transmembrane and/or cytoplasmic domain from an avian influenza virus. In another embodiment, said multivalent VLPs comprise an avian influenza M1 protein. In another embodiment, said avian influenza is A/Indonesia/5/05.

[0044] In another embodiment, the chimeric genes (as describe above), which may be codon optimized, are synthesized and cloned through a series of steps into a bacmid construct followed by rescue of recombinant baculovirus by plaque isolation and expression analyses. The VLPs for each of these targets can then be rescued by co-infection with the use of two recombinant baculoviruses (1) expressing the avian M1, and (2) expressing the chimeric protein from an infectious agent (e.g. VZV, RSV, Dengue, yellow fever) with cytoplasmic and/or transmembrane domain from HA and/or NA from a seasonal and/or avian influenza virus. In another embodiment, the VLPs can be rescued by infection with the use of a recombinant baculovirus expressing the avian M1 and the chimeric protein from an infectious agent (e.g. VZV, RSV, Dengue, yellow fever) with cytoplasmic and transmembrane domain from influenza HA and/or NA.

[0045] Infectious agents can be viruses, bacteria, fungi and/or parasites. A protein that may be expressed on the surface of chimeric VLPs of the invention can be derived from viruses, bacteria, fungi and/or parasites. In other embodiments, the proteins expressed on the surface of said chimeric VLPs may be tumor or cancer antigens. The proteins derived from viruses, bacteria, fungi and/or parasites can induce an immune response (cellular and/or humoral) in a vertebrate that which will prevent, treat, manage and/or ameliorate an infectious disease in said vertebrate.

[0046] Non-limiting examples of viruses from which said infectious agent proteins can be derived from are the following: coronavirus (e.g. the agent that causes SARS), hepatitis viruses A, B, C, D & E3, human immunodeficiency virus (HIV), herpes viruses 1, 2, 6 & 7, cytomegalovirus, varicella zoster, papilloma virus, Epstein Barr virus, parainfluenza viruses, respiratory syncytial virus (RSV), human metapneumovirus, adenoviruses, bunya viruses (e.g. hanta virus), coxsackie viruses, picorna viruses, rotaviruses, rhinoviruses, rubella virus, mumps virus, measles virus, Rubella virus, polio virus (multiple types), adeno virus (multiple types), parainfluenza virus (multiple types), avian influenza (various types), shipping fever virus, Western and Eastern equine encephalomyelitis, Japanese encephalomyelitis, fowl pox, rabies virus, slow brain viruses, rous sarcoma virus, Papovaviridae, Parvoviridae, Picomaviridae, Poxviridae (such as Smallpox or Vaccinia), Reoviridae (e.g., Rotavirus), Retroviridae (HTLV-I, HTLV-II, Lentivirus), Togaviridae (e.g., Rubivirus), Newcastle disease virus, West Nile fever virus, Tick borne encephalitis, yellow fever, chikungunya virus, and dengue virus (all serotypes).

[0047] In another embodiment, the specific proteins from viruses may comprise: HA and/or NA from influenza virus (including avian), S protein from coronavirus, gp160, gp140 and/or gp41 from HIV, gp I to IV and Vp from varicella zoster, E and preM/M from yellow fever virus, Dengue (all serotypes) or any flavivirus. Also included are any proteins from a virus that can induce an immune response (cellular and/or humoral) in a vertebrate that can prevent, treat, manage and/or ameliorate an infectious disease in said vertebrate.

[0048] Non-limiting examples of bacteria from which said infectious agent proteins can be derived from are the following: *B. pertussis*, *Leptospira pomona*, *S. paratyphi* A and B, *C. diphtheriae*, *C. tetani*, *C. botulinum*, *C. perfringens*, *C. fescer* and other gas gangrene bacteria, *B. anthracis*, *P. pestis*, *P. multocida*, *Neisseria meningitidis*, *N. gonorrhoeae*, *Hemophilus influenzae*, *Actinomyces* (e.g., *Nocardia*), *Acinetobacter*, *Bacillaceae* (e.g., *Bacillus anthracis*), *Bacteroides* (e.g., *Bacteroides fragilis*), *Blastomycosis*, *Bordetella*, *Borrelia* (e.g., *Borrelia burgdorferi*), *Brucella*, *Campylobacter*, *Chlamydia*, *Coccidioides*, *Corynebacterium* (e.g., *Corynebacterium diphtheriae*), *E. coli* (e.g., Enterotoxigenic *E. coli* and Enterohemor-

rhagic *E. coli*), Enterobacter (e.g. *Enterobacter aerogenes*), Enterobacteriaceae (Klebsiella, Salmonella (e.g., *Salmonella typhi*, *Salmonella enteritidis*, Serratia, Yersinia, Shigella), Erysipelothrix, Haemophilus (e.g., *Haemophilus influenza* type B), Helicobacter, Legionella (e.g., *Legionella pneumophila*), Leptospira, Listeria (e.g., *Listeria monocytogenes*), Mycoplasma, Mycobacterium (e.g., *Mycobacterium leprae* and *Mycobacterium tuberculosis*), Vibrio (e.g., *Vibrio cholerae*), Pasteurellaceae, Proteus, Pseudomonas (e.g., *Pseudomonas aeruginosa*), Rickettsiaceae, Spirochetes (e.g., *Treponema* spp., *Leptospira* spp., *Borrelia* spp.), Shigella spp., Meningococcus, Pneumococcus and Streptococcus (e.g., *Streptococcus pneumoniae* and Groups A, B, and C Streptococci), Ureaplasmas. *Treponema pallidum*, *Staphylococcus aureus*, *Pasteurella haemolytica*, *Corynebacterium diphtheriae* toxoid, Meningococcal polysaccharide, *Bordetella pertussis*, *Streptococcus pneumoniae*, *Clostridium tetani* toxoid, and *Mycobacterium bovis*.

[0049] Non-limiting examples of parasites from which said infectious agent proteins can be derived from are the following: leishmaniasis (*Leishmania tropica mexicana*, *Leishmania tropica*, *Leishmania major*, *Leishmania aethiopica*, *Leishmania braziliensis*, *Leishmania donovani*, *Leishmania infantum*, *Leishmania chagasi*), trypanosomiasis (*Trypanosoma brucei gambiense*, *Trypanosoma brucei rhodesiense*), toxoplasmosis (*Toxoplasma gondii*), schistosomiasis (*Schistosoma haematobium*, *Schistosoma japonicum*, *Schistosoma mansoni*, *Schistosoma mekongi*, *Schistosoma intercalatum*), malaria (*Plasmodium virax*, *Plasmodium falciparum*, *Plasmodium malariae* and *Plasmodium ovale*) Amebiasis (*Entamoeba histolytica*), Babesiosis (*Babesiosis microti*), Cryptosporidiosis (*Cryptosporidium parvum*), Dientamoebiasis (*Dientamoeba fragilis*), Giardiasis (*Giardia lamblia*), Helminthiasis and Trichomonas (*Trichomonas vaginalis*).

[0050] Non-limiting examples of fungi from which said glycoproteins can be derived are from the following: Absidia (e.g. *Absidia corymbifera*), Ajellomyces (e.g. *Ajellomyces capsulatus*, *Ajellomyces dermatitidis*), Arthroderma (e.g. *Arthroderma benhamiae*, *Arthroderma fulvum*, *Arthroderma gypseum*, *Arthroderma incurvatum*, *Arthroderma otae*, *Arthroderma vanbreuseghemii*), Aspergillus (e.g. *Aspergillus fumigatus*, *Aspergillus niger*), Candida (e.g. *Candida albicans*, *Candida albicans* var. *stellatoidea*, *Candida dublinensis*, *Candida glabrata*, *Candida guilliermondii* (*Pichia guilliermondii*), *Candida krusei* (*Issatschenkia orientalis*), *Candida parapsilosis*, *Candida pelliculosa* (*Pichia anomala*), *Candida tropicalis*), Coccidioides (e.g. *Coccidioides immitis*), Cryptococcus (e.g. *Cryptococcus neoformans* (*Filobasidiella neoformans*), *Histoplasma* (e.g. *Histoplasma capsulatum* (*Ajellomyces capsulatus*), *Microsporium* (e.g. *Microsporium canis* (*Arthroderma otae*), *Microsporium fulvum* (*Arthroderma fulvum*), *Microsporium gypseum*, Genus *Pichia* (e.g. *Pichia anomala*, *Pichia guilliermondii*), *Pneumocystis* (e.g. *Pneumocystis jirovecii*), *Cryptosporidium*, *Malassezia furfur*, *Paracoccidioides*.

[0051] The above lists are meant to be illustrative and by no means are meant to limit the invention to those particular bacterial, viral or parasitic organisms.

[0052] The invention also encompasses variants of the said proteins expressed on or in the chimeric VLPs of the invention. The variants may contain alterations in the amino acid sequences of the constituent proteins. The term "variant" with respect to a protein refers to an amino acid sequence that is altered by one or more amino acids with respect to a reference sequence. The variant can have "conservative" changes, wherein a substituted amino acid has similar structural or chemical properties, e.g., replacement of leucine with isoleucine. Alternatively, a variant can have "nonconservative" changes, e.g., replacement of a glycine with a tryptophan. Analogous minor variations can also include amino acid deletion or insertion, or both. Guidance in determining which amino acid residues can be substituted, inserted, or deleted without eliminating biological or immunological activity can be found using computer programs well known in the art, for example, DNASTAR software.

[0053] Natural variants can occur due to mutations in the proteins. These mutations may lead to antigenic variability within individual groups of infectious agents, for example influenza. Thus, a person infected with an influenza strain develops antibody against that virus, as newer virus strains appear, the antibodies against the older strains no longer recognize the newer virus and reinfection can occur. The invention encompasses all antigenic and genetic variability of proteins from infectious agents for making chimeric VLPs.

[0054] General texts which describe molecular biological techniques, which are applicable to the present invention, such as cloning, mutation, cell culture and the like, include Berger and Kimmel, Guide to Molecular Cloning Techniques, Methods in Enzymology volume 152 Academic Press, Inc., San Diego, Calif. (Berger); Sambrook et al., Molecular Cloning--A Laboratory Manual (3rd Ed.), Vol. 1-3, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., 2000 ("Sambrook") and Current Protocols in Molecular Biology, F. M. Ausubel et al., eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc., ("Ausubel"). These texts describe mutagenesis, the use of vectors, promoters and many other relevant topics related to, e.g., the cloning and mutating HA, NA and/or proteins from infectious agents, etc. Thus, the invention also encompasses using known methods of protein engineering and recombinant DNA technology to improve or alter the characteristics of the proteins expressed on or in the VLPs of the invention. Various types of mutagenesis can be used to produce and/or isolate variant nucleic acids that encode for protein molecules and/or to further modify/mutate the proteins in or on the VLPs of the invention. They include but are not limited to site-directed, random point mutagenesis, homologous recombination (DNA shuffling), mutagenesis using uracil containing templates, oligonucleotide-directed mutagenesis, phosphorothioate-modified DNA mutagenesis, mutagenesis using gapped duplex DNA or the like. Additional suitable methods include point mismatch repair, mutagenesis using repair-deficient host strains, restriction-selection and restriction-purification, deletion mutagenesis, mutagenesis

by total gene synthesis, double-strand break repair, and the like. Mutagenesis, e.g., involving chimeric constructs, is also included in the present invention. In one embodiment, mutagenesis can be guided by known information of the naturally occurring molecule or altered or mutated naturally occurring molecule, e.g., sequence, sequence comparisons, physical properties, crystal structure or the like.

[0055] The invention further comprises protein variants which show substantial biological activity, e.g., able to elicit an effective antibody response when expressed on or in VLPs of the invention. Such variants include deletions, insertions, inversions, repeats, and substitutions selected according to general rules known in the art so as to have little effect on activity.

[0056] Methods of cloning said proteins are known in the art. For example, the gene encoding a specific virus protein can be isolated by RT-PCR from polyadenylated mRNA extracted from cells which had been infected with a virus (DNA or RNA virus) or PCR from cells which had been infected with a DNA virus. The resulting product gene can be cloned as a DNA insert into a vector. The term "vector" refers to the means by which a nucleic acid can be propagated and/or transferred between organisms, cells, or cellular components. Vectors include plasmids, viruses, bacteriophages, proviruses, phagemids, transposons, artificial chromosomes, and the like, that replicate autonomously or can integrate into a chromosome of a host cell. A vector can also be a naked RNA polynucleotide, a naked DNA polynucleotide, a polynucleotide composed of both DNA and RNA within the same strand, a poly-lysine-conjugated DNA or RNA, a peptide-conjugated DNA or RNA, a liposome-conjugated DNA, or the like, that is not autonomously replicating. In many, but not all, common embodiments, the vectors of the present invention are plasmids or bacmids.

[0057] Thus, described herein are nucleotides that encode proteins, including chimeric molecules, cloned into an expression vector that can be expressed in a cell that induces the formation of VLPs of the invention. An "expression vector" is a vector, such as a plasmid that is capable of promoting expression, as well as replication of a nucleic acid incorporated therein. Typically, the nucleic acid to be expressed is "operably linked" to a promoter and/or enhancer, and is subject to transcription regulatory control by the promoter and/or enhancer. In one embodiment, said nucleotides encode for a non-avian influenza protein and/or chimeric protein (as discussed above). In another embodiment, said vector comprises nucleotides that encode for a non-avian influenza protein and/or chimeric protein and an avian influenza M1. In another embodiment, said vector comprises nucleotides that encode a chimeric protein comprising the cytoplasmic and/or the transmembrane domain of HA and/or NA from avian and/or seasonal influenza protein. In another embodiment, said seasonal influenza HA and/or NA are from influenza strain A/Wisconsin/67/2005 and said avian influenza HA and/or NA are from influenza strain A/Indonesia/5/05. In another embodiment, said vector comprises nucleotides that encode M1 from influenza strain A/Indonesia/5/05 and a chimeric protein comprising the A/Wisconsin/67/2005 (seasonal influenza) cytoplasmic and/or the transmembrane from HA and/or NA. In another embodiment, said vector comprises nucleotides that encode M1 from influenza strain A/Indonesia/5/05 and a chimeric protein comprising the A/Indonesia/5/05 (avian influenza) cytoplasmic and/or the transmembrane from HA and/or NA.

[0058] In some embodiments, said proteins may comprise mutations containing alterations that produce silent substitutions, additions, or deletions, but do not alter the properties or activities of the encoded protein or how the proteins are made. Nucleotide variants can be produced for a variety of reasons, e.g., to optimize codon expression for a particular host (change codons in the human mRNA to those preferred by insect cells such as Sf9 cells). See U.S. patent publication 2005/0118191.

[0059] In addition, the nucleotides can be sequenced to ensure that the correct coding regions were cloned and do not contain any unwanted mutations. The nucleotides can be subcloned into an expression vector (e.g. baculovirus) for expression in any cell. The above is only one example of how the influenza proteins (including chimeric proteins) can be cloned. A person with skill in the art understands that additional methods are available and are possible.

[0060] The application also describes constructs and/or vectors that comprise nucleotides that encode for avian M1 and non-avian influenza proteins and/or chimeric proteins (as described above). The constructs and/or vectors that comprise avian M1 and non-avian influenza proteins and/or chimeric proteins, should be operatively linked to an appropriate promoter, such as the AcMNPV polyhedrin promoter (or other baculovirus), phage lambda PL promoter, the *E. coli* lac, phoA and tac promoters, the SV40 early and late promoters, and promoters of retroviral LTRs are non-limiting examples. Other suitable promoters will be known to the skilled artisan depending on the host cell and/or the rate of expression desired. The expression constructs will further contain sites for transcription initiation, termination, and, in the transcribed region, a ribosome-binding site for translation. The coding portion of the transcripts expressed by the constructs will preferably include a translation initiating codon at the beginning and a termination codon appropriately positioned at the end of the polypeptide to be translated.

[0061] Expression vectors will preferably include at least one selectable marker. Such markers include dihydrofolate reductase, G418 or neomycin resistance for eukaryotic cell culture and tetracycline, kanamycin or ampicillin resistance genes for culturing in *E. coli* and other bacteria. Among vectors preferred are virus vectors, such as baculovirus, poxvirus (e.g., vaccinia virus, avipox virus, canarypox virus, fowlpox virus, raccoonpox virus, swinepox virus, etc.), adenovirus (e.g., canine adenovirus), herpesvirus, and retrovirus. Other vectors that can be used with the invention comprise vectors for use in bacteria, which comprise pQE70, pQE60 and pQE-9, pBluescript vectors, Phagescript vectors, pNH8A, pNH16a, pNH18A, pNH46A, ptrc99a, pKK223-3, pKK233-3, pDR540, pRIT5. Among preferred eukaryotic vectors are

pFastBac1 pWINEO, pSV2CAT, pOG44, pXT1 and pSG, pSVK3, pBPV, pMSG, and pSVL. Other suitable vectors will be readily apparent to the skilled artisan.

[0062] Next, the recombinant constructs mentioned above could be used to transfect, infect, or transform and can express avian M1 and a non-avian influenza protein and/or chimeric proteins, into eukaryotic cells and/or prokaryotic cells. Thus, host cells may comprise a vector (or vectors) that contain nucleic acids which code for avian M1 and chimeric proteins, and permit the expression of said constructs in said host cell under conditions which allow the formation of VLPs.

[0063] Among eukaryotic host cells are yeast, insect, avian, plant, *C. elegans* (or nematode) and mammalian host cells. Non limiting examples of insect cells are, *Spodoptera frugiperda* (Sf) cells, e.g. Sf9, Sf21, *Trichoplusia ni* cells, e.g. High Five cells, and *Drosophila* S2 cells. Examples of fungi (including yeast) host cells are *S. cerevisiae*, *Kluyveromyces lactis* (*K. lactis*), species of *Candida* including *C. albicans* and *C. glabrata*, *Aspergillus nidulans*, *Schizosaccharomyces pombe* (*S. pombe*), *Pichia pastoris*, and *Yarrowia lipolytica*. Examples of mammalian cells are COS cells, baby hamster kidney cells, mouse L cells, LNCaP cells, Chinese hamster ovary (CHO) cells, human embryonic kidney (HEK) cells, and African green monkey cells, CV1 cells, HeLa cells, MDCK cells, Vero and Hep-2 cells. *Xenopus laevis* oocytes, or other cells of amphibian origin, may also be used. Prokaryotic host cells include bacterial cells, for example, *E. coli*, *B. subtilis*, and mycobacteria.

[0064] Vectors, e.g., vectors comprising polynucleotides of avian M1 and non-avian influenza proteins and/or chimeric proteins, can be transfected into host cells according to methods well known in the art. For example, introducing nucleic acids into eukaryotic cells can be by calcium phosphate co-precipitation, electroporation, microinjection, lipofection, and transfection employing polyamine transfection reagents. In one embodiment, said vector is a recombinant baculovirus. In another embodiment, said recombinant baculovirus is transfected into a eukaryotic cell. In a preferred embodiment, said cell is an insect cell. In another embodiment, said insect cell is a Sf9 cell.

[0065] In another embodiment, said vector and/or host cell comprise nucleotides that encode avian M1 and non-avian influenza proteins and/or chimeric proteins. In another embodiment, said vector and/or host cell consists essentially of avian M1 and non-avian influenza proteins and/or chimeric proteins. In a further embodiment, said vector and/or host cell consists of avian M1 and non-avian influenza proteins and/or chimeric proteins. These vector and/or host cell contain avian M1 and non-avian influenza proteins and/or chimeric proteins, and may contain additional markers, such as an origin of replication, selection markers, etc.

[0066] The invention also provides for methods that will further increase the efficiency of VLPs production. For example, the addition of leader sequences to the avian M1 and non-avian influenza proteins and/or chimeric proteins, can improve the efficiency of protein transporting within the cell. For example, a heterologous signal sequence can be fused to avian M1 and non-avian influenza proteins and/or chimeric proteins. In one embodiment, the signal sequence can be derived from the gene of an insect preprotein and fused to avian M1 and non-avian influenza proteins and/or chimeric proteins. In another embodiment, the signal peptide is the chitinase signal sequence, which works efficiently in baculovirus expression systems.

[0067] The invention comprises a method of increasing the efficiency of producing chimeric VLPs comprising expressing an avian influenza M1 and seasonal influenza HA and/or NA. In one embodiment, said HA or NA have hemagglutinin and neuraminidase activity, respectfully. Said M1 is from influenza strain A/Indonesia/5/05. In one embodiment, said HA and/or NA is from influenza strain A/Wisconsin/67/2005.

[0068] In another embodiment of the invention, the increase in VLP production, for chimeric or non-chimeric VLPs, is about 2 fold, about 4 fold, about 8 fold, about 16 fold, about 20 fold, about 25 fold, about 30 fold, about 35 fold, about 40 fold, about 45 fold, about 50 fold, about 55 fold, about 60 fold, about 65 fold, about 70 fold, about 75 fold, about 80 fold, about 85 fold, about 90 fold, about 95 fold, about 100 fold, or more when compared to VLP production comprising a non-avian influenza M1 protein under similar conditions, for instance seasonal influenza M1. In one embodiment, the efficiency of producing influenza VLPs is increase by about 10%, about 20% about 30%, about 40%, about 50% about 60%, about 70% about 80%, about 90%, about 100%, about 150%, about 200%, about 250%, about 300%, about 350%, about 400%, about 450%, about 500%, about 550%, about 600%, about 650%, about 700%, about 750%, about 800% about 850% about 900% about 950% about 1000% when compared to VLP production comprising a non-avian influenza M1 protein under similar conditions. A preferred M1 is from A/indonesia/5/05 (SEQ ID NO. 3).

[0069] The invention provides for methods of producing VLPs, said methods comprising expressing an avian M1 according to the claims and a seasonal HA and NA under conditions that allow the formation of VLPs. Depending on the expression system and host cell selected, VLPs are produced by growing host cells transformed by an expression vector under conditions whereby the recombinant proteins including avian M1 and seasonal HA and NA are expressed and VLPs are formed. The selection of the appropriate growth conditions is within the skill or a person with skill of one of ordinary skill in the art.

[0070] Methods to grow cells engineered to produce VLPs of the invention include, but are not limited to, batch, batch-fed, continuous and perfusion cell culture techniques. Cell culture means the growth and propagation of cells in a bioreactor (a fermentation chamber) where cells propagate and express protein (e.g. recombinant proteins) for purification and isolation. Typically, cell culture is performed under sterile, controlled temperature and atmospheric conditions in a

bioreactor. A bioreactor is a chamber used to culture cells in which environmental conditions such as temperature, atmosphere, agitation and/or pH can be monitored. In one embodiment, said bioreactor is a stainless steel chamber. In another embodiment, said bioreactor is a pre-sterilized plastic bag (e.g. Cellbag®, Wave Biotech, Bridgewater, NJ). In other embodiment, said pre-sterilized plastic bags are about 50 L to 1000 L bags.

[0071] VLPs are then isolated using methods that preserve the integrity thereof, such as by gradient centrifugation, e.g., cesium chloride, sucrose and iodixanol, as well as standard purification techniques including, e.g., ion exchange and gel filtration chromatography.

[0072] The following is an example of how VLPs of the invention can be made, isolated and purified. Usually VLPs are produced from recombinant cell lines engineered to create a VLP when said cells are grown in cell culture (see above). A person of skill in the art would understand that there are additional methods that can be utilized to make and purify VLPs of the invention, thus the invention is not limited to the method described.

[0073] Production of VLPs of the invention can start by seeding Sf9 cells (non-infected) into shaker flasks, allowing the cells to expand and scaling up as the cells grow and multiply (for example from a 125-ml flask to a 50 L Wave bag). The medium used to grow the cell is formulated for the appropriate cell line (preferably serum free media, e.g. insect medium ExCell-420, JRH). Next, said cells are infected with recombinant baculovirus at the most efficient multiplicity of infection (e.g. from about 1 to about 3 plaque forming units per cell). Once infection has occurred, the avian influenza M1 and at least one avian influenza heterologous protein are expressed from the virus genome, self assemble into VLPs and are secreted from the cells approximately 24 to 72 hours post infection. Usually, infection is most efficient when the cells are in mid-log phase of growth ($4-8 \times 10^6$ cells/ml) and are at least about 90% viable.

[0074] VLPs of the invention can be harvested approximately 48 to 96 hours post infection, when the levels of VLPs in the cell culture medium are near the maximum but before extensive cell lysis. The Sf9 cell density and viability at the time of harvest can be about 0.5×10^6 cells/ml to about 1.5×10^6 cells/ml with at least 20% viability, as shown by dye exclusion assay. Next, the medium is removed and clarified. NaCl can be added to the medium to a concentration of about 0.4 to about 1.0 M, preferably to about 0.5 M, to avoid VLP aggregation. The removal of cell and cellular debris from the cell culture medium containing VLPs of the invention can be accomplished by tangential flow filtration (TFF) with a single use, pre-sterilized hollow fiber 0.5 or 1.00 μ m filter cartridge or a similar device.

[0075] Next, VLPs in the clarified culture medium can be concentrated by ultrafiltration using a disposable, pre-sterilized 500,000 molecular weight cut off hollow fiber cartridge. The concentrated VLPs can be diafiltered against 10 volumes pH 7.0 to 8.0 phosphate-buffered saline (PBS) containing 0.5 M NaCl to remove residual medium components.

[0076] The concentrated, diafiltered VLPs can be further purified on a 20% to 60% discontinuous sucrose gradient in pH 7.2 PBS buffer with 0.5 M NaCl by centrifugation at $6,500 \times g$ for 18 hours at about 4°C to about 10°C. Usually VLPs will form a distinctive visible band between about 30% to about 40% sucrose or at the interface (in a 20% and 60% step gradient) that can be collected from the gradient and stored. This product can be diluted to comprise 200 mM of NaCl in preparation for the next step in the purification process. This product contains VLPs and may contain intact baculovirus particles.

[0077] Further purification of VLPs can be achieved by anion exchange chromatography, or 44% isopycnic sucrose cushion centrifugation. In anion exchange chromatography, the sample from the sucrose gradient (see above) is loaded into column containing a medium with an anion (e.g. Matrix Fractogel EMD TMAE) and eluted via a salt gradient (from about 0.2 M to about 1.0 M of NaCl) that can separate the VLP from other contaminants (e.g. baculovirus and DNA/RNA). In the sucrose cushion method, the sample comprising the VLPs is added to a 44% sucrose cushion and centrifuged for about 18 hours at 30,000 g. VLPs form a band at the top of 44% sucrose, while baculovirus precipitates at the bottom and other contaminating proteins stay in the 0% sucrose layer at the top. The VLP peak or band is collected.

[0078] The intact baculovirus can be inactivated, if desired. Inactivation can be accomplished by chemical methods, for example, formalin or β -propyl lactone (BPL). Removal and/or inactivation of intact baculovirus can also be largely accomplished by using selective precipitation and chromatographic methods known in the art, as exemplified above. Methods of inactivation comprise incubating the sample containing the VLPs in 0.2% of BPL for 3 hours at about 25°C to about 27°C. The baculovirus can also be inactivated by incubating the sample containing the VLPs at 0.05% BPL at 4°C for 3 days, then at 37°C for one hour.

[0079] After the inactivation/removal step, the product comprising VLPs can be run through another diafiltration step to remove any reagent from the inactivation step and/or any residual sucrose, and to place the VLPs into the desired buffer (e.g. PBS). The solution comprising VLPs can be sterilized by methods known in the art (e.g. sterile filtration) and stored in the refrigerator or freezer.

[0080] The above techniques can be practiced across a variety of scales. For example, T-flasks, shake-flasks, spinner bottles, up to industrial sized bioreactors. The bioreactors can comprise either a stainless steel tank or a pre-sterilized plastic bag (for example, the system sold by Wave Biotech, Bridgewater, NJ). A person with skill in the art will know what is most desirable for their purposes.

[0081] Expansion and production of baculovirus expression vectors and infection of cells with recombinant baculovirus to produce recombinant influenza VLPs can be accomplished in insect cells, for example Sf9 insect cells as previously

described. In a preferred embodiment, the cells are Sf9 infected with recombinant baculovirus engineered to produce VLPs of the invention.

Pharmaceutical or Vaccine Formulations and Administration

[0082] The application describes an antigenic formulation comprising a chimeric VLP comprising an avian influenza M1 protein and at least one non-avian influenza protein. In one embodiment, said VLP comprises HA and/or NA from a non-avian influenza virus. In another embodiment, said HA or NA have hemagglutinin and neuraminidase activity, respectfully. In another embodiment, said HA and/or NA are chimeric proteins. In another embodiment, said chimeric proteins comprise external domains of non-avian influenza HA and/or NA protein sequences fused to the transmembrane and/or cytoplasmic-terminal domains of the avian HA and/or NA cytoplasmic region. In another embodiment, said non-avian influenza HA and/or NA are from influenza strain A/Wisconsin/67/2005 and said avian influenza HA and/or NA are from influenza strain A/Indonesia/5/05. In another embodiment, said M1 is from influenza strain A/Indonesia/5/05. In another embodiment, said HA and/or NA is from influenza strain A/Wisconsin/67/2005.

[0083] The application describes a vaccine comprising a chimeric VLP comprising an avian influenza M1 protein and at least one non-avian influenza protein. In one embodiment, said VLP comprises HA and/or NA from a non-avian influenza virus. In another embodiment, said HA or NA have hemagglutinin and neuraminidase activity, respectfully. In another embodiment, said HA and/or NA are chimeric proteins. In another embodiment, said chimeric proteins comprise external domains of non-avian influenza HA and/or NA protein sequences fused to the transmembrane and/or cytoplasmic-terminal domains of the avian HA and/or NA cytoplasmic region. In another embodiment, said non-avian influenza HA and/or NA are from influenza strain A/Wisconsin/67/2005 and said avian influenza HA and/or NA are from influenza strain A/Indonesia/5/05. In another embodiment, said M1 is from influenza strain A/Indonesia/5/05. In another embodiment, said HA and/or NA is from influenza strain A/Wisconsin/67/2005.

[0084] The pharmaceutical compositions useful herein contain a pharmaceutically acceptable carrier, including any suitable diluent or excipient, which includes any pharmaceutical agent that does not itself induce the production of an immune response harmful to the vertebrate receiving the composition, and which may be administered without undue toxicity and a VLP of the invention. As used herein, the term "pharmaceutically acceptable" means being approved by a regulatory agency of the Federal or a state government or listed in the U.S. Pharmacopia, European Pharmacopia or other generally recognized pharmacopia for use in mammals, and more particularly in humans. These compositions can be useful as a vaccine and/or antigenic compositions for inducing a protective immune response in a vertebrate.

[0085] The application describes an antigenic formulation comprising a chimeric VLP comprising an avian influenza M1 protein and at least one non-avian influenza protein. In one embodiment, said VLP comprises HA and/or NA from a non-avian influenza virus. In another embodiment, said HA or NA have hemagglutinin and neuraminidase activity, respectfully. In another embodiment, said HA and/or NA are chimeric proteins. In another embodiment, said chimeric proteins comprise external domains of non-avian influenza HA and/or NA protein sequences fused to the transmembrane and/or Cytoplasmic-terminal domains of the avian HA and/or NA cytoplasmic region. In another embodiment, said non-avian influenza HA and/or NA are from influenza strain A/Wisconsin/67/2005 and said avian influenza HA and/or NA are from influenza strain A/Indonesia/5/05. In another embodiment, said M1 is from influenza strain A/Indonesia/5/05. In another embodiment, said HA and/or NA is from influenza strain A/Wisconsin/67/2005.

[0086] The application describes a vaccine comprising a chimeric VLP comprising an avian influenza M1 protein and at least one non-avian influenza protein. In one embodiment, said VLP comprises HA and/or NA from a non-avian influenza virus. In another embodiment, said HA or NA have hemagglutinin and neuraminidase activity, respectfully. In another embodiment, said HA and/or NA are chimeric proteins. In another embodiment, said chimeric proteins comprise external domains of non-avian influenza HA and/or NA protein sequences fused to the transmembrane and/or cytoplasmic-terminal domains of the avian HA and/or NA cytoplasmic region. In another embodiment, said non-avian influenza HA and/or NA are from influenza strain A/Wisconsin/67/2005 and said avian influenza HA and/or NA are from influenza strain A/Indonesia/5/05. In another embodiment, said M1 is from influenza strain A/Indonesia/5/05. In another embodiment, said HA and/or NA is from influenza strain A/Wisconsin/67/2005.

[0087] Said formulations comprise a formulation comprising VLPs comprising an avian M1 protein and at least one protein from a non-avian influenza protein (e.g. a protein from an infectious agent described above) and a pharmaceutically acceptable carrier or excipient. Pharmaceutically acceptable carriers include but are not limited to saline, buffered saline, dextrose, water, glycerol, sterile isotonic aqueous buffer, and combinations thereof. A thorough discussion of pharmaceutically acceptable carriers, diluents, and other excipients is presented in Remington's Pharmaceutical Sciences (Mack Pub. Co. N.J. current edition). The formulation should suit the mode of administration. In another embodiment, the formulation is suitable for administration to humans, preferably is sterile, non-particulate and/or non-pyrogenic.

[0088] The composition, if desired, can also contain minor amounts of wetting or emulsifying agents, or pH buffering agents. The composition can be a solid form, such as a lyophilized powder suitable for reconstitution, a liquid solution, suspension, emulsion, tablet, pill, capsule, sustained release formulation, or powder. Oral formulation can include stand-

ard carriers such as pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate, etc.

[0089] The application also describes a pharmaceutical pack or kit comprising one or more containers filled with one or more of the ingredients of the vaccine formulations of the invention. In one embodiment, the kit comprises two containers, one containing VLPs and the other containing an adjuvant. In another embodiment, the kit comprises two containers, one containing freeze dried VLPs and the other containing a solution to resuspend said VLPs. Associated with such container(s) can be a notice in the form prescribed by a governmental agency regulating the manufacture, use or sale of pharmaceuticals or biological products, which notice reflects approval by the agency of manufacture, use or sale for human administration.

[0090] The application also envisages that the VLP formulation be packaged in a hermetically sealed container such as an ampoule or sachette indicating the quantity of composition. In one embodiment, the VLP composition is supplied as a liquid, in another embodiment, as a dry sterilized lyophilized powder or water free concentrate in a hermetically sealed container and can be reconstituted, e.g., with water or saline to the appropriate concentration for administration to a subject. In one embodiment, said container comprises at least about 50 $\mu\text{g/ml}$, more preferably at least about 100 $\mu\text{g/ml}$, at least about 200 $\mu\text{g/ml}$, at least 500 $\mu\text{g/ml}$, or at least 1 mg/ml of an antigen associated with VLPs of the invention.

[0091] In an alternative embodiment, the VLP composition is supplied in liquid form in a hermetically sealed container indicating the quantity and concentration of the VLP composition. The liquid form of the VLP composition is supplied in a hermetically sealed container at least about 50 $\mu\text{g/ml}$, more preferably at least about 100 $\mu\text{g/ml}$, at least about 200 $\mu\text{g/ml}$, at least 500 $\mu\text{g/ml}$, or at least 1 mg/ml of an antigen associated with VLPs of the invention.

[0092] Generally, VLPs produced according to the invention are administered in an effective amount or quantity (as defined above) sufficient to stimulate an immune response against one or more infectious agents. Preferably, administration of the VLP of the invention elicits immunity against an infectious agent. Typically, the dose can be adjusted within this range based on, e.g., age, physical condition, body weight, sex, diet, time of administration, and other clinical factors. The prophylactic vaccine formulation is systemically administered, e.g., by subcutaneous or intramuscular injection using a needle and syringe, or a needle-less injection device. Alternatively, the vaccine formulation is administered intranasally, either by drops, large particle aerosol (greater than about 10 microns), or spray into the upper respiratory tract. While any of the above routes of delivery results in an immune response, intranasal administration confers the added benefit of eliciting mucosal immunity at the site of entry of many viruses, including RSV and influenza.

[0093] Thus, the application also describes a method of formulating a vaccine or antigenic composition that induces immunity to an infection or at least one symptom thereof to a mammal, comprising adding to said formulation an effective dose of VLPs.

[0094] Methods of administering a composition comprising VLPs (vaccine and/or antigenic formulations) include, but are not limited to, parenteral administration (e.g., intradermal, intramuscular, intravenous and subcutaneous), epidural, and mucosal (e.g., intranasal and oral or pulmonary routes or by suppositories). In a specific embodiment, compositions of the present invention are administered intramuscularly, intravenously, subcutaneously, transdermally or intradermally. The compositions may be administered by any convenient route, for example by infusion or bolus injection, by absorption through epithelial or mucocutaneous linings (e.g., oral mucous, colon, conjunctiva, nasopharynx, oropharynx, vagina, urethra, urinary bladder and intestinal mucosa, etc.) and may be administered together with other biologically active agents. In some embodiments, intranasal or other mucosal routes of administration of a composition comprising VLPs may induce an antibody or other immune response that is substantially higher than other routes of administration. In another embodiment, intranasal or other mucosal routes of administration of a composition comprising VLPs may induce an antibody or other immune response that will induce cross protection against other strains or organisms that cause infection. For example, a VLP comprising influenza protein, when administered to a vertebrate, can induce cross protection against several influenza strains. Administration can be systemic or local.

[0095] In yet another embodiment, the vaccine and/or antigenic formulation is administered in such a manner as to target mucosal tissues in order to elicit an immune response at the site of immunization. For example, mucosal tissues such as gut associated lymphoid tissue (GALT) can be targeted for immunization by using oral administration of compositions which contain adjuvants with particular mucosal targeting properties. Additional mucosal tissues can also be targeted, such as nasopharyngeal lymphoid tissue (NALT) and bronchial-associated lymphoid tissue (BALT).

[0096] Vaccines and/or antigenic formulations may also be administered on a dosage schedule, for example, an initial administration of the vaccine composition with subsequent booster administrations. In particular embodiments, a second dose of the composition is administered anywhere from two weeks to one year, preferably from about 1, about 2, about 3, about 4, about 5 to about 6 months, after the initial administration. Additionally, a third dose may be administered after the second dose and from about three months to about two years, or even longer, preferably about 4, about 5, or about 6 months, or about 7 months to about one year after the initial administration. The third dose may be optionally administered when no or low levels of specific immunoglobulins are detected in the serum and/or urine or mucosal secretions of the subject after the second dose. In a preferred embodiment, a second dose is administered about one month after the first administration and a third dose is administered about six months after the first administration. In

another embodiment, the second dose is administered about six months after the first administration. In another embodiment, said VLPs can be administered as part of a combination therapy. For example, VLPs can be formulated with other immunogenic compositions, antivirals and/or antibiotics.

[0097] The dosage of the pharmaceutical formulation can be determined readily by the skilled artisan, for example, by first identifying doses effective to elicit a prophylactic or therapeutic immune response, e.g., by measuring the serum titer of virus specific immunoglobulins or by measuring the inhibitory ratio of antibodies in serum samples, or urine samples, or mucosal secretions. Said dosages can be determined from animal studies. A non-limiting list of animals used to study the efficacy of vaccines include the guinea pig, hamster, ferrets, chinchilla, mouse and cotton rat. Most animals are not natural hosts to infectious agents but can still serve in studies of various aspects of the disease. For example, any of the above animals can be dosed with a vaccine candidate, e.g. VLPs, to partially characterize the immune response induced, and/or to determine if any neutralizing antibodies have been produced. For example, many studies have been conducted in the mouse model because mice are small size and their low cost allows researchers to conduct studies on a larger scale.

[0098] In addition, human clinical studies can be performed to determine the preferred effective dose for humans by a skilled artisan. Such clinical studies are routine and well known in the art. The precise dose to be employed will also depend on the route of administration. Effective doses may be extrapolated from dose-response curves derived from *in vitro* or animal test systems.

[0099] As also well known in the art, the immunogenicity of a particular composition can be enhanced by the use of non-specific stimulators of the immune response, known as adjuvants. Adjuvants have been used experimentally to promote a generalized increase in immunity against unknown antigens (e.g., U.S. Pat. No. 4,877,611). Immunization protocols have used adjuvants to stimulate responses for many years, and as such, adjuvants are well known to one of ordinary skill in the art. Some adjuvants affect the way in which antigens are presented. For example, the immune response is increased when protein antigens are precipitated by alum. Emulsification of antigens also prolongs the duration of antigen presentation. The inclusion of any adjuvant described in Vogel et al., "A Compendium of Vaccine Adjuvants and Excipients (2nd Edition)," is envisioned within the scope of this invention.

[0100] Exemplary, adjuvants include complete Freund's adjuvant (a non-specific stimulator of the immune response containing killed *Mycobacterium tuberculosis*), incomplete Freund's adjuvants and aluminum hydroxide adjuvant. Other adjuvants comprise GMCSF, BCG, aluminum hydroxide, MDP compounds, such as thur-MDP and nor-MDP, CGP (MTP-PE), lipid A, and monophosphoryl lipid A (MPL). RIBI, which contains three components extracted from bacteria, MPL, trehalose dimycolate (TDM) and cell wall skeleton (CWS) in a 2% squalene/Tween 80 emulsion also is contemplated. MF-59, Novasomes®, MHC antigens may also be used.

[0101] In one embodiment the adjuvant is a paucilamellar lipid vesicle having about two to ten bilayers arranged in the form of substantially spherical shells separated by aqueous layers surrounding a large amorphous central cavity free of lipid bilayers. Paucilamellar lipid vesicles may act to stimulate the immune response several ways, as non-specific stimulators, as carriers for the antigen, as carriers of additional adjuvants, and combinations thereof. Paucilamellar lipid vesicles act as non-specific immune stimulators when, for example, a vaccine is prepared by intermixing the antigen with the preformed vesicles such that the antigen remains extracellular to the vesicles. By encapsulating an antigen within the central cavity of the vesicle, the vesicle acts both as an immune stimulator and as a carrier for the antigen. In another embodiment, the vesicles are primarily made of nonphospholipid vesicles. In another embodiment, the vesicles are Novasomes. Novasomes® are paucilamellar nonphospholipid vesicles ranging from about 100 nm to about 500 nm. They comprise Brij 72, cholesterol, oleic acid and squalene. Novasomes have been shown to be an effective adjuvant for influenza antigens (see, U.S. Patents 5,629,021, 6,387,373, and 4,911,928).

[0102] Another method of inducing an immune response can be accomplished by formulating the VLPs of the invention with "immune stimulators." These are the body's own chemical messengers (cytokines) to increase the immune system's response. Immune stimulators include, but not limited to, various cytokines, lymphokines and chemokines with immunostimulatory, immunopotentiating, and pro-inflammatory activities, such as interleukins (e.g., IL-1, IL-2, IL-3, IL-4, IL-12, IL-13); growth factors (e.g., granulocyte-macrophage (GM)-colony stimulating factor (CSF)); and other immunostimulatory molecules, such as macrophage inflammatory factor, Flt3 ligand, B7.1; B7.2, etc. The immunostimulatory molecules can be administered in the same formulation as the RSV VLPs, or can be administered separately. Either the protein or an expression vector encoding the protein can be administered to produce an immunostimulatory effect. Thus in one embodiment, the application describes antigenic and vaccine formulations comprising an adjuvant and/or an immune stimulator.

[0103] Thus, described herein is a formulation comprising a chimeric VLP comprising an avian M1 and at least one non-avian influenza protein (or at least one protein from an infectious agent) and adjuvant and/or an immune stimulator. In another embodiment, said adjuvant are Novasomes. In another embodiment, said formulation is suitable for human administration. In another embodiment, the formulation is administered to a vertebrate orally, intradermally, intranasally, intramuscularly, intraperitoneally, intravenously or subcutaneously. In another embodiment, different chimeric VLPs are blended together to create a multivalent formulation. These VLPs may comprise VLPs HA and/or NA from different

strains of influenza virus (e.g. influenza A and/or influenza B) or protein from different infectious agents (e.g. RSV, coronavirus, HIV).

[0104] While stimulation of immunity with a single dose is preferred, additional dosages can be administered by the same or different route to achieve the desired effect. In neonates and infants, for example, multiple administrations may be required to elicit sufficient levels of immunity. Administration can continue at intervals throughout childhood, as necessary to maintain sufficient levels of protection against infections. Similarly, adults who are particularly susceptible to repeated or serious infections, such as, for example, health care workers, day care workers, family members of young children, the elderly, and individuals with compromised cardiopulmonary function may require multiple immunizations to establish and/or maintain protective immune responses. Levels of induced immunity can be monitored, for example, by measuring amounts of neutralizing secretory and serum antibodies, and dosages adjusted or vaccinations repeated as necessary to elicit and maintain desired levels of protection.

Methods of Stimulating an Immune Response

[0105] As mentioned above, the VLPs produced by the invention are useful for preparing compositions that stimulate an immune response that confers immunity to infectious agents. Both mucosal and cellular immunity may contribute to immunity to infectious agents and disease. Antibodies secreted locally in the upper respiratory tract are a major factor in resistance to natural infection. Secretory immunoglobulin A (sIgA) is involved in protection of the upper respiratory tract and serum IgG in protection of the lower respiratory tract. The immune response induced by an infection protects against reinfection with the same virus or an antigenically similar viral strain. For example, influenza undergoes frequent and unpredictable changes; therefore, after natural infection, the effective period of protection provided by the host's immunity may only be a few years against the new strains of virus circulating in the community.

[0106] VLPs produced by the invention can induce an immunity in a vertebrate (e.g. a human) when administered to said vertebrate. The immunity results from an immune response against VLPs of the invention that protects or ameliorates infection or at least reduces a symptom of infection in said vertebrate. In some instances, if the said vertebrate is infected, said infection will be asymptomatic. The response may be not a fully protective response. In this case, if said vertebrate is infected with an infectious agent, the vertebrate will experience reduced symptoms or a shorter duration of symptoms compared to a non-immunized vertebrate.

[0107] The application describes a method of inducing immunity in a vertebrate comprising administering to said vertebrate a VLP comprising an avian influenza M1 protein and at least one non-avian influenza protein. In one embodiment, said immune response is a humoral immune response. In another embodiment, said immune response is a cellular immune response. In another embodiment, said non-avian influenza protein is HA and/or NA from a non-avian influenza virus. In another embodiment, said non-avian influenza protein is a seasonal influenza protein. In another embodiment, said HA or NA has hemagglutinin or neuraminidase activity, respectfully. In one embodiment, said HA and/or NA are chimeric proteins. In another embodiment, said chimeric proteins comprise external domains of non-avian influenza HA and/or NA protein sequences fused to the transmembrane and/or cytoplasmic-terminal domains of the avian HA and/or NA cytoplasmic region. In another embodiment, said non-avian influenza HA and/or NA are from influenza strain A/Wisconsin/67/2005 and said avian influenza HA and/or NA transmembrane and/or cytoplasmic-terminal domains are from influenza strain A/Indonesia/5/05. In another embodiment, said M1 is from influenza strain A/Indonesia/5/05. In another embodiment, said HA and/or NA is from influenza strain A/Wisconsin/67/2005.

[0108] As used herein, an "antibody" is a protein comprising one or more polypeptides substantially or partially encoded by immunoglobulin genes or fragments of immunoglobulin genes. The recognized immunoglobulin genes include the kappa, lambda, alpha, gamma, delta, epsilon and mu constant region genes, as well as myriad immunoglobulin variable region genes. Light chains are classified as either kappa or lambda. Heavy chains are classified as γ , μ , α , δ , or ϵ , which in turn define the immunoglobulin classes, IgG, IgM, IgA, IgD and IgE, respectively. A typical immunoglobulin (antibody) structural unit comprises a tetramer. Each tetramer is composed of two identical pairs of polypeptide chains, each pair having one "light" (about 25 kD) and one "heavy" chain (about 50-70 kD). The N-terminus of each chain defines a variable region of about 100 to 110 or more amino acids primarily responsible for antigen recognition. Antibodies exist as intact immunoglobulins or as a number of well-characterized fragments produced by digestion with various peptidases.

[0109] In another embodiment, the application describes a method of inducing a protective cellular response to an infection or at least one symptom thereof in a subject, comprising administering at least one effective dose of VLPs of the invention, wherein said VLPs comprise an avian influenza M1 protein and at least one non-avian influenza protein. Cell-mediated immunity also plays a role in recovery from infection and may prevent additional complication and contribute to long term immunity.

[0110] As mentioned above, the VLPs produced by the invention can prevent or reduce at least one symptom of an infection in a subject when administered to said subject. Most symptoms of most infections are well known in the art. Thus, the method comprises the prevention or reduction of at least one symptom associated with an infection. A reduction in a symptom may be determined subjectively or objectively, e.g., self assessment by a subject, by a clinician's assessment

or by conducting an appropriate assay or measurement (e.g. body temperature), including, e.g., a quality of life assessment, a slowed progression of an infection or additional symptoms, reduced severity of symptoms or suitable assays (e.g. antibody titer and/or T-cell activation assay). The objective assessment comprises both animal and human assessments.

[0111] The application describes a method of preventing and/or reducing an infection or symptom thereof, comprising administering to said vertebrate a chimeric VLP comprising an avian influenza M1 protein and at least one non-avian influenza protein. In one embodiment said infection is a viral infection. In another embodiment, said viral infection is an influenza infection.

[0112] A strategy for the control of infectious diseases during an outbreak, e.g. influenza, is the universal vaccination of healthy individuals, including children. For example, vaccination with current influenza vaccines of approximately 80% of schoolchildren in a community has decreased respiratory illnesses in adults and excess deaths in the elderly (Reichert *et al.*, 2001). This concept is known as community immunity or "herd immunity" and is thought to play an important part of protecting the community against diseases. Because vaccinated people have antibodies that neutralize and infectious agent, e.g. influenza virus, they are much less likely to transmit said agent to other people. Thus, even people who have not been vaccinated (and those whose vaccinations have become weakened or whose vaccines are not fully effective) often can be shielded by the herd immunity because vaccinated people around them are not getting sick. Herd immunity is more effective as the percentage of people vaccinated increases. It is thought that approximately 95% of the people in the community must be protected by a vaccine to achieve herd immunity. People who are not immunized increase the chance that they and others will get the disease.

[0113] Thus, also described herein is a method of reducing the severity of an infectious disease in a population, comprising administering a VLP comprising an avian influenza M1 protein and at least one non-avian influenza protein to enough individuals in said population in order to prevent or decrease the chance of transmission to another individual in said population. In one embodiment, said infectious disease is caused by influenza virus. The application also describes a method of inducing immunity to an infectious agent to a population or a community in order to reduce the incidence of infections among immunocompromised individuals or non-vaccinated individual by administering VLPs of the invention to a population in a community. In one embodiment, most school-aged children are immunized by administering the VLPs of the invention. In another embodiment, most healthy individuals in a community are immunized by administering the VLPs of the invention. In another embodiment, VLPs of the invention are part of a "dynamic vaccination" strategy. Dynamic vaccination is the steady production of a low-efficacy vaccine that is related to an emerging pandemic strain, but due to an antigenic drift may not provide complete protection in a mammal (see Germann *et al.*, 2006).

[0114] This invention is further illustrated by the following examples that should not be construed as limiting.

EXAMPLES

Example 1

Expressing Seasonal and Avian VLPs from two baculovirus vectors

[0115] Seasonal and avian influenza M1 and HA proteins were cloned and expressed in a baculovirus expression system. In this example, the A/Indonesia/5/05 was cloned into a one baculovirus and the HA and/or NA was cloned in another baculovirus vector. Both viruses were co infected into Sf9 insect cells and grown under conditions that allow VLP formation. Cells comprising either seasonal HA and M1, avian HA and M1 or a combination of seasonal and avian HA and M1 were grown under conditions that allow formation of VLPs. The seasonal influenza strains used for these experiments were A/Fujian/411/2002 and A/Wisconsin/67/2005 and the avian influenza strain used was A/Indonesia/5/05.

[0116] Next, the VLPs were harvested and isolated from the supernatant by centrifugation and by a discontinuous sucrose step gradient. The fraction comprising the VLPs was collected from the top of the gradient. The VLPs isolated from the sucrose gradient were analyzed by SDS-PAGE and western immunoblot. These data are on illustrated on Figure 1 and 2.

[0117] Figure 1 is a stained SDS-PAGE gel. The lanes in the gel comprise the following: 1 to 5, A/Fujian M1 with 4 different HAs or alone; 6 to 10, A/Indo/ M1 with 4 different HAs or alone; 11 to 14, various controls.

[0118] Comparing the bands on the gel, the lanes that comprise VLPs comprising avian M1 have stronger bands of M1 and HA in the same lanes, while the lanes that comprise seasonal influenza do not. M1 and HA bands in the same lane is indicative of HA associating with M1. This association is indicative of VLP formation comprising HA and M1. These data provide evidence that avian influenza proteins form VLPs more efficiently than seasonal influenza M1 either with homologous or heterologous envelopes. These data also show that M1 from avian influenza is strongly expressed and stable when compared to seasonal influenza M1.

[0119] Figure 2 is a western blot showing M1 expression. This blot shows that avian influenza M1 is strongly expressed

as compared to seasonal M1. The intensity of the bands indicate that there is more M1, and thus, more VLPs.

Example 2

Expressing Seasonal and Avian VLPs from one baculovirus vector

[0120] Seasonal and avian influenza M1 and HA proteins were cloned and expressed in a baculovirus expression system. This example, the A/Indonesia/5/05 M1 and A/Fujian/411/2002 HA and NA was cloned into a one baculovirus baculovirus vector. The recombinant virus was infected into Sf9 insect cells and grown under conditions that allow VLP formation. Cells comprising either seasonal HA and M1, avian HA and M1 or a combination of seasonal and avian HA and M1 were grown under conditions that allow formation of VLPs. The seasonal influenza strains used for these experiments were A/Fujian/411/2002 and A/Wisconsin/67/2005 and the avian influenza strain used was A/Indonesia/5/05.

[0121] Next, VLPs were harvested and isolated from the supernatant by centrifugation and by a discontinuous sucrose step gradient. The fraction comprising the VLPs was collected from the top of the gradient. The VLPs isolated from the sucrose gradient were analyzed by SDS-PAGE and western immunoblot. These data are on illustrated on Figure 3 and 4.

[0122] Figure 3 is a stained SDS-PAGE gel. The lanes in the gel comprise the following: 1 and 2 is A/Fujian VLPs (M1, HA and NA) and lane 3 comprises, A/Indo/ M1 with A/Fujian HA and NA.

[0123] Comparing the bands on the gel, the lane that comprise VLPs from A/Indo/ M1 has stronger bands of M1 and HA in the same lanes, while the lanes that comprise A/Fujian do not. M1, HA and NA bands in the same lane is indicative of HA and NA associating with M1. This association is indicative of VLP formation comprising HA, NA and M1. These data provide evidence that avian influenza proteins form VLPs with greater efficiency than seasonal M1 influenza based VLPs. These data also show that M1 from avian influenza is strongly expressed and stable when compared to seasonal influenza M1.

[0124] Figure 4 is a western blot showing M1 expression. This blot shows that VLPs comprising endo A/Indo/ M1 and A/Fujian HA, NA are strongly expressed as compared to A/Fujian VLPs. The intensity of the bands indicate that there is more M1, HA and NA in lanes with avian M1 VLPs, and thus, more VLPs.

Example 3

Expressing Chimeric influenza B HA and NA constructs using common A/Indonesia/5/05 matrix protein to assemble VLPs

[0125] The sequences below depict the transmembrane and terminal sequences derived from A/Indonesia/5/05 HA and NA (underlined). The transmembrane and terminal sequences of HA and NA molecules can be determined using software prediction by GCG/Accelrys or similar software, as well as by other methods. The exact location of junctions for Indonesia/5/05 sequences can vary.

[0126] The sequences below are examples of a chimeric B strain HA with an A/Indonesia/5/05 HA end as well as a chimeric B strain NA with an A/Indo NA substitution of the endodomain and transmembrane regions. These sequences are co-expressed in a baculovirus expression system with an avian influenza M1 protein to produce chimeric VLPs that express influenza B antigens on the surface of VLPs.

Hemagglutinin, HA, from Influenza B virus (B/Hong Kong/557/2000) ABL76892 (SEQ ID NO. 1)

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1  mkaiivllmv vtsnadriect gitssnsphv vktatqgevn vtgviplttt ptkshfanlk
61  gtrtrgklcp dclnctdlav algrpmcvgt tpsakasilh evrpvtsgcf pimhdrtkir
121 qlpnllrgye nirlstqnv daekapggpy rlgtsqscpn atsksgffat mawavpkdnn
181 knatnplte vpyvcteged qitvwgfhsd nktqmknlyg dsnpgkftss angvtthyvs
241 qiggfpdqte dgglpqsgri vvdymvqkpg ktgtivyqrg vllpqkvwca sgrskvikgs
301 lpligeadcl hekygglnks kpyytgehak aigncpwvk tplklangtk yrppakllke
361 rgffgaiagf leggwegmia gwhgytshga hgvavaadlk stqeainkit knlnslsele
421 vknlqrlsga mdelhneile ldekvdldra dtissqiela vllsnegiin sedehllale
481 rklkmlgps avdigngcfe tkhkcngtcl driaagtfna gefslptfde linitaaslnd
541 dgldnhtQIL SIYSTVASSL ALAIMMAGLS LWMCSNGSLQ CRICI

```

Neuraminidase, NA, from Flu B/Shanghai/361/02 ISDN129538 (SEQ ID NO. 2)

MNPNQKIITIGSICMVGIVSLMLQIGNMISSDILLKFSTTEITAPTMLDCANASNVQAVNRSATKGVTLILLPEPE
 WTYPRLSCPGSTFQKALLISPHRFGETKGNAPLI IREPFIACGPKECKHFALTHYAAQPGGYNGTREDNRNKLRLH
 ISVKLGKIPTVENSIFHMAAWSGSACHDGKEWTYIGVDGPDNALLKI KYGEAYTDTYHSYANNILRTQESACNCIG
 GNCYLMITDGSASGISSECRFLKIREGRIIKEIFPTGRVKHTEECTCGFASNKTI ECACRDNSYTAKRPFVKLNVEDT
 5 TAEIRLMCTETYLDTPRPDDGSITGPCESENGNKGSGGIKGGFVHQRMASKIGRWYSRTMSKTKRMGMGLYVKYDGD
 WIDSDALALSGVMVSMEEPGWYSFGFEIKDKKCDVPCIGIEMVHDGGKETWHSAAATAYCLMSGGQLLWDTVTGVDM
 AL

M1 from A/Indonesia (SEQ ID NO. 3)

MSLLTEVETY VLSIIPSGPL KAEIAQKLED VFAGKNTDLE ALMEWLKTRP
 ILSPLTKGIL GFVFTLTVPS ERGLQRRRFV QNALNGNGDP NNMDRAVKLY
 KKLKREITFH GAKEVSLSYS TGALASCMGL IYNRMGTVT EVAFGLVCAT
 CEQIADSQHR SHRQMATITN PLIRHENRMV LASTTAKAME QMAGSSEQAA

EAMEVANQAR QMVQAMRTIG THPNSSAGLR DNLLLENLQAY QKRMGVQMQR
 FK

Example 4

Making chimeric VLPs with coronavirus S protein

Materials and Methods

[0127] *Spodoptera frugiperda* Sf9 insect cells (ATCC CRL-1711) were maintained as suspension in HyQ-SFX insect serum free medium (HyClone, Logan, UT) at 28°C. A Bac-to-Bac baculovirus expression system (Invitrogen, Carlsbad, CA) was used with pFastBac 1 transfer vector in *E. coli* DH10Bac cells for the generation of recombinant baculovirus vectors expressing SARS S and Influenza M1 genes.

[0128] SARS coronavirus (SARS-CoV) Urbani strain spike (S) protein amino acids sequence was obtained from NCBI access number AAP13441. The hemagglutinin amino acids sequence of influenza A virus (A/Indonesia/5/05(H5N1)) was obtained from from NCBI access number ABP51969. To construct the chimeric SARS S protein, the transmembrane and carboxyl terminal domain (TM/CT) of S protein (aa 1196-1255) was removed, and the TM/CT from Indonesia H5N1 HA (aa 531-568) was added after amino acid 1195 of S protein. The amino acids sequence of the chimeric S-HA protein is shown in Figure 5 (SEQ ID NO. 10). The matrix protein 1 (M1) amino acids sequence of influenza Indonesia H5N1 was obtained from NCBI access number ABW06359 (Figure 6).

[0129] The codon optimized DNA sequences of M1 and chimeric S for expression in insect cells were synthesized by Geneart (Germany) and subcloned into BamHI and HindIII sites of pFastBac 1 individually. The SnaBI/PvuI fragment containing M1 coding sequence of pFastBac1-M1 was cut and inserted into the HpaI/PvuI fragment containing S coding sequence from pFastBac1-S. The result tandem vector that expresses two proteins is shown in Figure 7. This vector was used to transform DH10Bac to obtain the bacmid which was tranfected into Sf9 cell to obtain the recombinant baculovirus.

VLPs expression, purification and characterizations

[0130] Sf9 insect cells were infected for 64 hours at a cell density of 2×10^6 cells/ml with recombinant baculoviruses that express both chimeric SARS S and Indo M1 at a MOI=1. Culture supernatants were harvest by centrifuge at 4000 g. The cell free supernatants were concentrated by ultrafiltration (UF) with a 500 kDa MWCO hollow fiber filter (GE healthcare). The retentate was buffer exchanged with diafiltration (DF) to 25 mM TrisCl pH 8.0, 300 mM NaCl. The UF/DF retentate was loaded on an ion exchange column (Fractogel TMAE, EMD) equilibrium in the same buffer. VLPs passed through from the column while baculovirus and DNA bound to the column. The flow through fractions containing VLPs were further concentrated with ultrafiltration before load to a Sephacryl S500 size exclusion column (GE healthcare).

[0131] The pool of VLPs peak from size exclusion column was analyzed with SDS-PAGE (4-12% Bis-Tris NuPage, Invitrogen) and densitometry for purity. The VLPs were also analyzed with particle size analyzer (Malvern Zetasizer NanoSeries NanoZS) and electron microscopy. The antibodies used in this study were from the following vendors: rabbit anti-SARS S and normal anti-rabbit IgG (IMGNEX), rabbit anti-SARS M (Abgent), mouse anti-influenza M1 (Serotec).

Results

[0132] Purified SARS S/Indo M1 chimeric VLPs were analyzed by SDS-PAGE, densitometry and western blot (Figure 8). The purity for SARS S protein was 13.7% and purity for Indo M1 protein was 67.6%. The combined purity for the S and M1 is 81.3%. The western blot confirmed the identity of S and M1 (Figure 8, lane 2).

[0133] Recombinant baculovirus that expressed SARS spike (S), membrane (M) and envelope (E) proteins in a tandem manner were also expressed. We expressed and purified the wild type SARS VLPs with the same protocol that was used to purify chimeric VLPs. The purity of wild type SARS VLPs (no influenza proteins) were analyzed by SDS-PAGE and western blot (Figure 9). The S and M proteins can hardly be seen in the coomassie stained gel and the contaminant proteins were much more prominent. The data indicate that wild type SARS VLPs are insufficient to form in the baculovirus insect cell expression system while the SARS S/Indo M1 chimeric VLPs an greatly improve the yield and purity of the product VLPs.

[0134] Next, we analyzed the average particle size of purified chimeric VLPs to be 159.2 nm (Figure 10). The chimeric VLPs were imaged with electron microscopy (EM) negative stain (Figure 11). The size and morphology of chimeric VLPs are very similar to the published EM images of SARS coronavirus (Figure 12). They are about 100 nm diameter with corona structure on the outer rim. The immuno-gold EM with anti-SARS S antibody confirmed that SARS S proteins were located on the surface of chimeric VLPs (Figure 12).

[0135] The inventors have engineered a chimeric VLP comprising the major spike (S) gene of coronavirus (CoV) that causes SARS. A CoV S chimeric envelope glycoprotein was made by replacing the transmembrane and C-terminus (endodomain) with analogous sequences from the avian influenza HA (A/Indonesia/5/05 H5N1 strain). Unexpected high levels of SARS VLPs were produced in Sf9 insect cells infected with a baculovirus expressing the chimeric SARS S glycoprotein and the avian M1 matrix protein. Chimeric VLPs comprising S protein have the morphology that is nearly identical to the wild type CoV with the recombinant, chimeric S spike protein forming a corona (crown) - envelope in a lipid envelope on spherical particles with an avian influenza M1 core. These recombinant chimeric SARS-avian flu VLPs are efficiently produced in insect cells and were purified as described above.

[0136] These data provide an excellent example that avian M1, e.g. Indonesia H5N1 M1 protein, can form chimeric VLPs with surface antigen from other virus such as SARS-CoV. The chimeric VLPs with avian influenza protein as backbone can be purified through a manufacturing friendly procedure that requires only two steps of chromatography. The size and morphology of the chimeric VLPs are similar to the wild type viruses that carry the same surface antigen.

Example 5

Chimeric influenza B VLPs

[0137] Influenza B virus antigen is an important component of seasonal influenza vaccines. The expression levels of influenza B antigen are critically important for ensuring timely delivery of sufficient number of influenza vaccine doses, otherwise vaccine shortages can occur. Influenza B VLPs for B/Florida/4/06 consist of three proteins, HA (SEQ ID NO 8), NA (SEQ ID NO. 9), and M1 (matrix), which are assembled into VLP structure. HA and NA genes were obtained by RT-PCR from the influenza B/Florida/4/06 virus. In order to improve expression levels of influenza B VLPs, VLPs using three different M1 proteins were made. One M1 protein is derived from the B/Florida/4/06 virus. The second M1 gene is derived from influenza B/Ann Arbor/1/1986 strain, which is often used for preparation of live reassortant influenza B viruses in current influenza vaccine industry. The third M1 is derived from avian influenza A/Indonesia/5/05 (H5N1) virus. Thus, three types of influenza B/Florida/4/06 VLPs have been produced in Sf9 cells, and expression levels have been compared.

[0138] Methods. Baculoviruses were engineered to express full length HA, NA, and M1 genes of influenza. HA and NA genes were obtained by RT-PCR from the influenza B/Florida/4/06 virus. M1 gene has been also generated by RT-PCR from the influenza B/Florida/4/06 virus. Alternatively, M1 gene of B/Ann Arbor/1/1986 was synthesized (GeneArt, Germany) and M1 gene of influenza A/Indonesia/5/05 (H5N1) was also synthesized (GeneArt, Germany). Each gene was cloned into a pFastBac1 vector under the control of the baculovirus polyhedrin promoter (Invitrogen). Then, HA, NA, and M1 genes were combined into tandem vectors as shown on Figure 13. Then, tandem gene constructs were transferred to an AcMNPV baculovirus Bacmid vectors (Invitrogen), the Bacmid DNAs were purified and used to transfect Sf9 insect cells. The resulting recombinant baculoviruses were plaque-purified and virus stocks prepared in Sf9 cells.

[0139] About 30 ml of Sf9 cells, at about 2×10^6 cells/ml in a 125mL shaker flasks, were infected with recombinant baculoviruses expressing HA, NA, and M1 genes at a multiplicity of infection (MOI) of 1 - 3 infectious particles per ml (pfu), incubated at 27°C with constant shaking, then harvested at 66- 72 hours post infection. The media was removed by low speed centrifugation. Then, media were clarified using filtration through 0.45 μ M filters and the media were subjected to ultracentrifugation for 1 hour at 26,000 rpm through 30% sucrose layer. Pellets were resuspended in 200 mL of PBS and analyzed by SDS-PAGE and western blot (Figure 14). Resuspended pellets were also analyzed for

ability to agglutinate guinea pig red blood cells *in vitro*. The data are shown on Figure 14. The resuspended pellets have been also analyzed by negative staining transmission electron microscopy.

[0140] Results. M1 derived from influenza A/Indonesia/5/05 (H5N1) showed significantly higher expression levels by Coomassie gel staining (Figure 14, lane 3) compared to VLPs made using B/Florida/4/06 M1 or B/Ann Arbor/1/1986 M1. Also, HA titers of VLPs containing influenza A/Indonesia/5/05 (H5N1) M1, were 4-8 times higher as compared to the other two VLP types. Electron microscopy of VLPs containing influenza A/Indonesia/5/05 (H5N1) M1 had higher concentration of VLP and more regular spherical shape as compared to the other two VLPs (Figure 15).

Example 6

Making chimeric VLPs with RSV F1 protein

[0141] *Spodoptera frugiperda* Sf9 insect cells are maintained and grown as essentially described above. The codon optimized DNA sequences of influenza M1 (SEQ ID NO: 3) and chimeric RSV F1 (HA TM/CY (SEQ ID NO: 12)) for expression in insect cells are synthesized and subcloned into pFastBac 1. The result vector expresses both proteins. This vector is used to transform DH10Bac to obtain the bacmid which is tranfected into Sf9 cell to obtain the recombinant baculovirus.

[0142] Sf9 insect cells are infected for 64 hours at a cell density of 2×10^6 cells/ml with recombinant baculoviruses that express both chimeric RSV F1 and Indo M1 (SEQ ID NO: 3) at a MOI=1. Culture supernatants are harvest by centrifuge at 4000 g. The cell free supernatants are concentrated by ultrafiltration (UF) with a 500 kDa MWCO hollow fiber filter (GE healthcare). The retentate is buffer exchanged with diafiltration (DF) to 25 mM TrisCl pH 8.0, 300 mM NaCl. The UF/DF retentate is loaded on an ion exchange column (Fractogel TMAE, EMD). VLPs pass through from the column while baculovirus and DNA binds to the column. The flow through fractions containing VLPs are further concentrated with ultrafiltration before loading onto a Sephacryl S500 size exclusion column (GE healthcare).

[0143] The pool of VLPs peak from size exclusion column is analyzed with SDS-PAGE (4-12% Bis-Tris NuPage, Invitrogen) and densitometry for purity. The VLPs are also analyzed with particle size analyzer (Malvern Zetasizer NanoSeries NanoZS), SDS PAGE, western blot analysis, and electron microscopy.

[0144] All patents, publications and patent applications herein are incorporated by reference to the same extent as if each individual patent, publication or cited patent application was specifically and individually indicated to be incorporated by reference.

[0145] The foregoing detailed description has been given for clearness of understanding only and no unnecessary limitations should be understood therefrom as modifications will be obvious to those skilled in the art. It is not an admission that any of the information provided herein is prior art or relevant to the presently claimed inventions, or that any publication specifically or implicitly referenced is prior art.

[0146] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs.

Claims

1. A method of increasing the efficiency of influenza virus-like particle (VLP) production in a host cell comprising expressing an avian influenza matrix (M1) protein and seasonal human influenza hemagglutinin (HA) and neuraminidase (NA) proteins in a host cell, wherein the avian influenza M1 protein is an A/Indonesia/5/05 influenza M1 protein, and wherein said increased efficiency of VLP production is relative to a host cell expressing said HA and NA proteins with a seasonal influenza M1 protein.
2. The method of claim 1, wherein the influenza HA protein has hemagglutinin activity.
3. The method of claim 1, wherein the influenza NA protein has neuraminidase activity.
4. The method of claim 1, wherein the host cell is selected from the group consisting of: a yeast cell, an insect cell, an amphibian cell, an avian cell, and a mammalian cell.
5. The method of claim 4 wherein the host cell is an insect cell selected from the group consisting of: an Sf9 cell, an Sf21 cell, and a *Trichoplusia ni* cell.
6. The method of claim 5, wherein the insect cell is an Sf9 cell.

Patentansprüche

1. Verfahren zum Erhöhen der Wirksamkeit einer Produktion eines Influenzavirus-ähnlichen Partikels (VLP) in einer Wirtszelle, das das Exprimieren eines Vogelgrippematrix- (M1-) Proteins und von menschlichen saisonalen Influenza-Hämagglutinin- (HA-) und Neuraminidase- (NA-) Proteinen in einer Wirtszelle umfasst, worin das Vogelgrippe-M1-Protein ein A/Indonesien/5/05-Influenza-M1-Protein ist und worin die erhöhte Wirksamkeit der VLP-Produktion relativ zu einer Wirtszelle ist, die die HA- und NA-Proteine mit einem saisonalen Influenza-M1-Protein exprimiert.
2. Verfahren nach Anspruch 1, worin das Influenza-HA-Protein eine Hämagglutinin-Aktivität aufweist.
3. Verfahren nach Anspruch 1, worin das Influenza-NA-Protein eine Neuraminidase-Aktivität aufweist.
4. Verfahren nach Anspruch 1, worin die Wirtszelle aus der Gruppe bestehend aus einer Hefezelle, einer Insektenzelle, einer Amphibienzelle, einer Vogelzelle und einer Säugetierzelle ausgewählt ist.
5. Verfahren nach Anspruch 4, worin die Wirtszelle eine Insektenzelle ist, die aus der Gruppe bestehend aus einer Sf9-Zelle, einer Sf21-Zelle und einer *Trichoplusiani*-Zelle ausgewählt ist.
6. Verfahren nach Anspruch 5, worin die Insektenzelle eine Sf9-Zelle ist.

Revendications

1. Méthode pour augmenter l'efficacité d'une production de particules semblables au virus de la grippe (VLP) dans une cellule hôte comprenant l'expression d'une protéine de la matrice de grippe aviaire (M1) et des protéines d'hémagglutinine (HA) et de neuraminidase (NA) de la grippe saisonnière humaine dans une cellule hôte, où la protéine de grippe aviaire M1 est une protéine de grippe M1 de A/Indonésie/5/05, et où la plus grande efficacité de la production de VLP est relative à une cellule hôte exprimant lesdites protéines HA et NA avec une protéine M1 de la grippe saisonnière.
2. Méthode selon la revendication 1, où la protéine HA de grippe possède une activité d'hémagglutinine.
3. Méthode selon la revendication 1, où la protéine NA de grippe possède une activité de neuraminidase.
4. Méthode selon la revendication 1, où la cellule hôte est sélectionnée dans le groupe consistant en :
une cellule de levure, une cellule d'insecte, une cellule d'amphibien, une cellule aviaire et une cellule mammalienne.
5. Méthode selon la revendication 4, où la cellule hôte est une cellule d'insecte sélectionnée dans le groupe consistant en : une cellule Sf9, une cellule Sf21 et une *Trichoplusia ni* cellule.
6. Méthode selon la revendication 5, où la cellule d'insecte est une cellule Sf9.

FIGURE 1

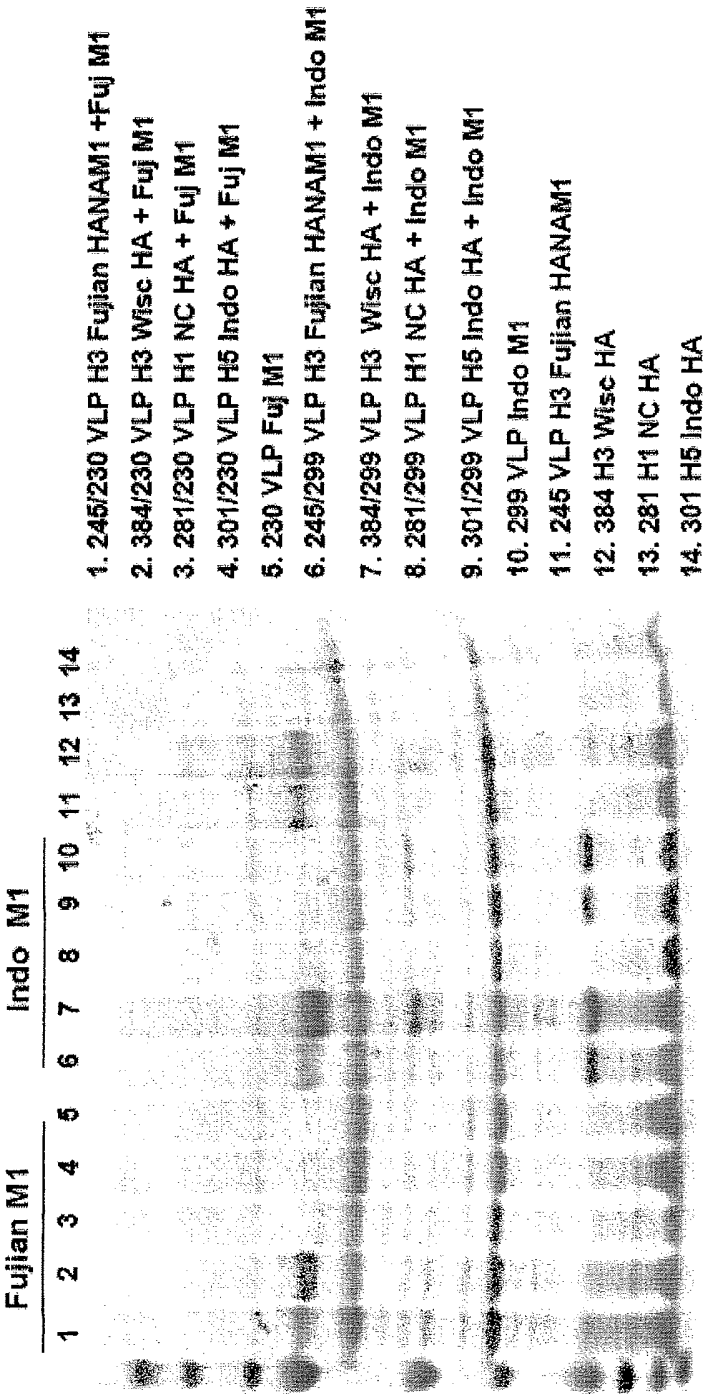


FIGURE 2

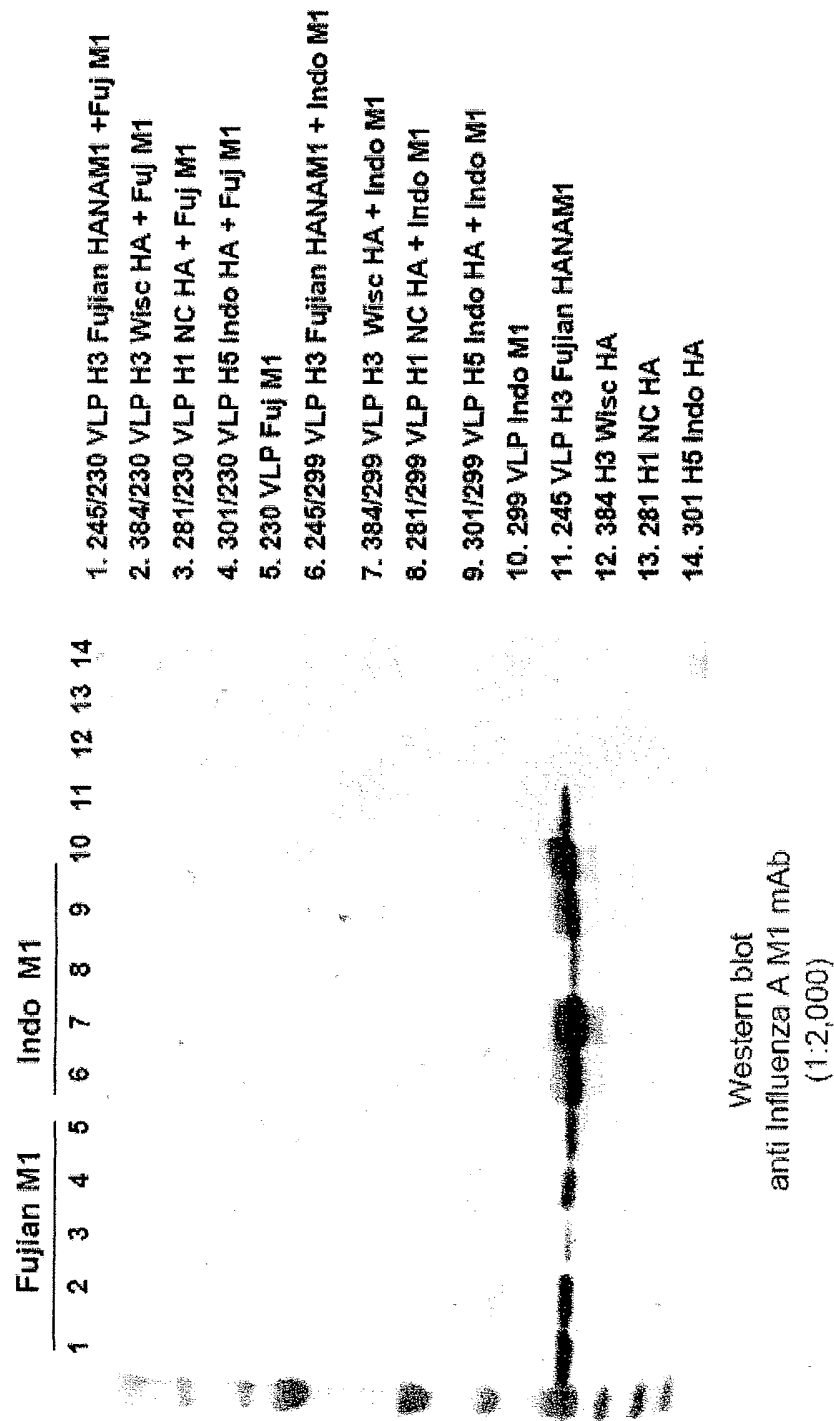


FIGURE 3

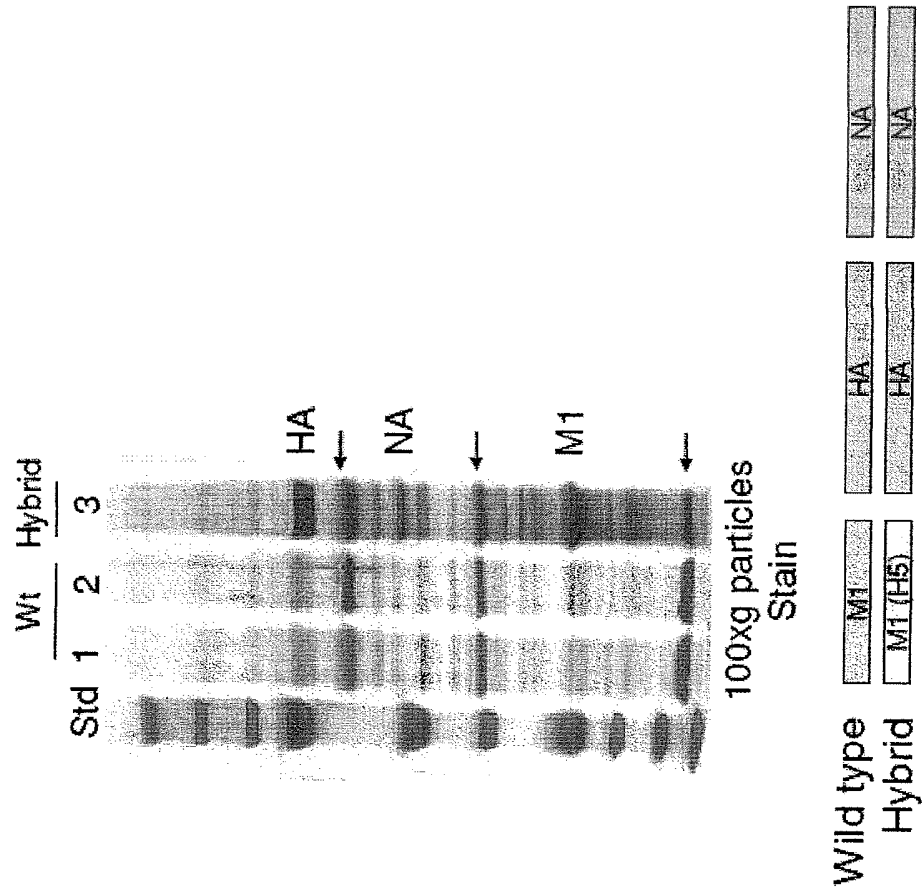


Figure 4

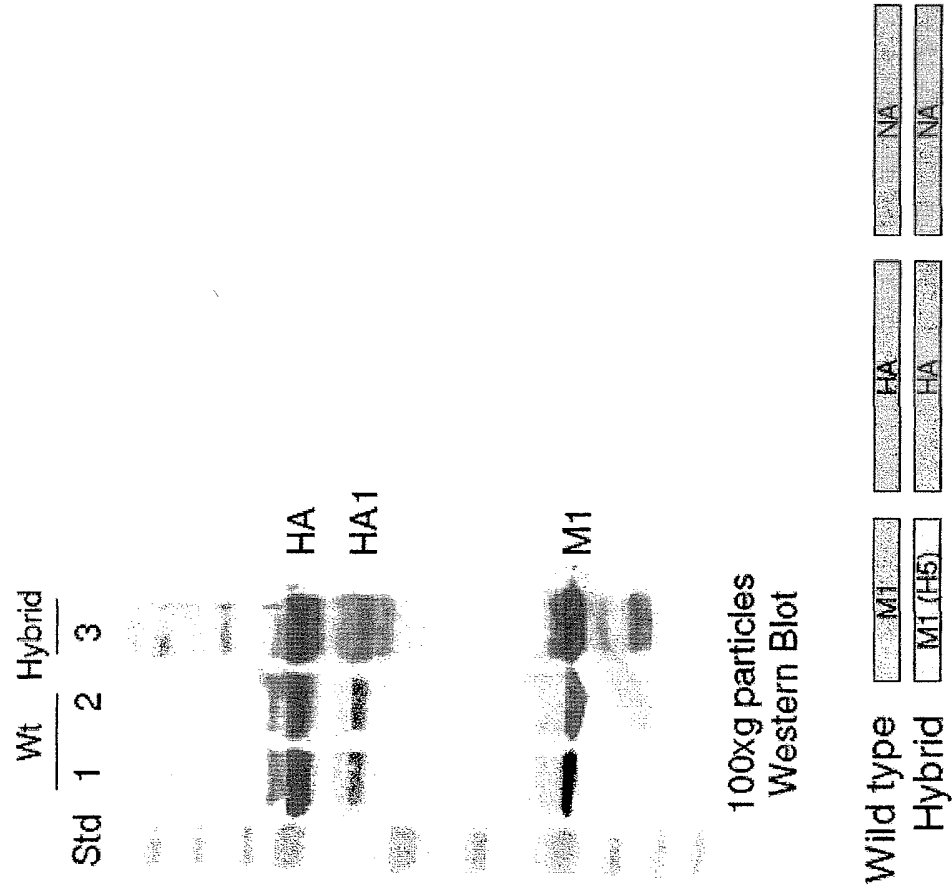


Figure 5

1 MFIFLLFLTL TSGSDLDRCT TFDDVQAPNY TQHTSSMRGV YYPDEIFRSD TLYLTQDLFL
 61 PFYSNVTFGH TINHTFGNPV IPFKDGIYFA ATEKSNVVRG WVFSGTMNNK SQSVIIINNS
 121 TNVVIRACNF ELCDNPFPAV SKPMGTQTHF MIFDNAFNCT FEYISDAFSL DVSEKSGNFK
 181 HLREFVFNK DGFLVYKGY QPIDVVRDLF SGFTILKPIF KLPLGINITN FRAILTAESP
 241 AQDIWGTSA AEFVGYLKPT TFMILKYDENG TITDAVDCSQ NPLAELKCSV KSFEIDKGIY
 301 QTSNFRVPS GDVVRFPNIT NLCPFGEVFN ATKFPFSVIAM ERKISNCVA DYSVLNSTF
 361 FSTFKCYGVS ATKINDLCFS NVYADSFVK GDDVRQIAPG QTGVIAADNY KLPDDFMGCV
 421 LAWNRNIDA TSTGNVNYKY RYLHKGKLR FERDISNVPF SPDGKPCPPP ALNCYWPLND
 481 YGFYTTTGIG YQPYRVVWLS FELLNAPATV CGPKLSTDLI KNQCVNFNFN GLTGTGVLTP
 541 SSKRFQPFQQ FGRDVSDFTD SVRDPKTSEI LDISPCSEFG VSVITPGTNA SSEVAVLYQD
 601 VNCTDVSTAI HADQLTPAWR IYSTGNVVFQ TQAGCLIGAE HVDTSYECDI FIGAGICASY
 661 HTVSLLRSTS QKSIVAYTMS LGADSSIAYS NNTIAIPTNF SISITTEVMP VSMAKTSVDC
 721 NMYICGDSTE CANLLQYGS FCTQINRALS GIAAEQDRNT REVEAQVKQM YKTPTLKYFG
 781 GFNFSQILPD PLKPTKRSEI EDLLFNKVTI ADAGFMKQYG ECLGDINARD LICAQKFNGI
 841 TVLPPLLTD MIAAYTAALV SGTATAGWTF GAGALQIPF AMQMYRFRNG IGVTONVLYE
 901 NQKQIANQFN KAISQIQESL TTTSTALGKL QDVVNQNAQA LNTLVKQLSS NFGAISSVLN
 961 DILSRLDKVE AEVQIDRLIT GRQLSLQTYV TQQLIRAAEI RASANLAATK MSECVLGQSK
 1021 RVDFCGKGYH LMSFPQAAPH GVVELHVTYV PSQERNFTTA PAICHEGKAY FPREGVVFEN
 1081 GTSWFITQRN FFSQIITTD NTFVSGNCDV VIGINNNTVY DPLQFELDSF KEELDKYFKN
 1141 HTSPDVLGD ISGINASVFN IQKEIDRLNE VAKNINESLI DLQELGKYEQ YIKWPQILSI
 1201 YSTVASSLAL AIMMAGLSLW MCSNGSLQCR ICI (SEQ ID NO. 10)

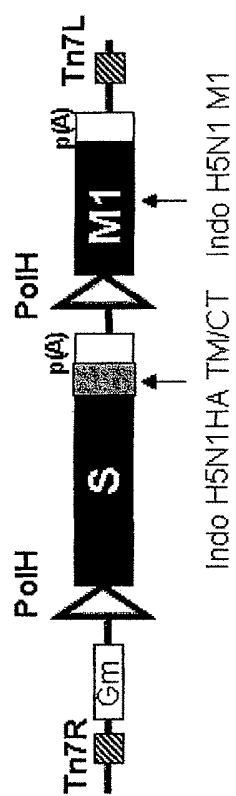
Figure 6

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1  MSLLTEVETYVLSIIPSGPIKAEIAQKLEDVFAGKNTDLEALMEWLKTRP
51  ILSPLTKGILGFVFTLTVPSEGLQRRRFVQNALNGDPNNMDRAVKLY
101 KKLKREITTFHGAKESLSYSTGALASCMGLIYNRMGTVTTEVAFGLVCAT
151 CEQIADSQHRSHRQMATITNPLIRHENRMVLASTTAKAMEQMGASSEQAA
201 EAMEVANQARQMVQAMRTIGTHPNSSAGLRDNLLENLQAYQKRMGVQMQR
251 FK (SEQ ID NO. 3)

```

Figure 7



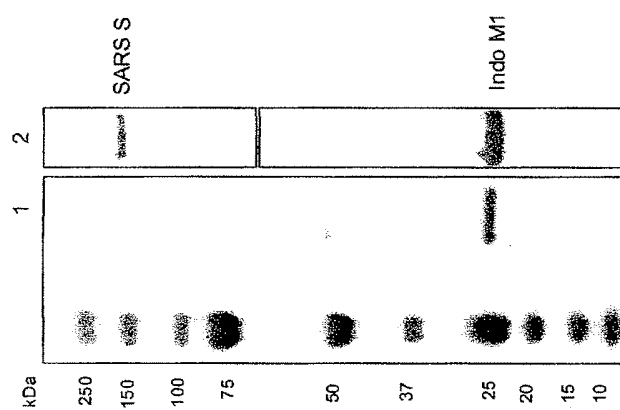


Figure 8

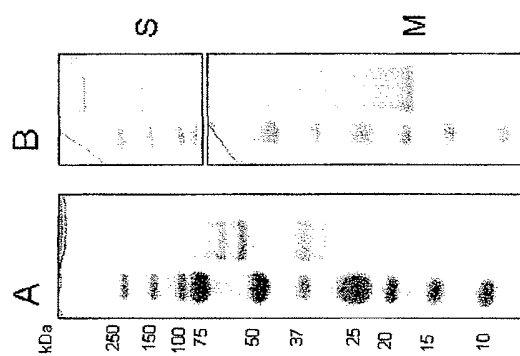


Figure 9

Figure 10

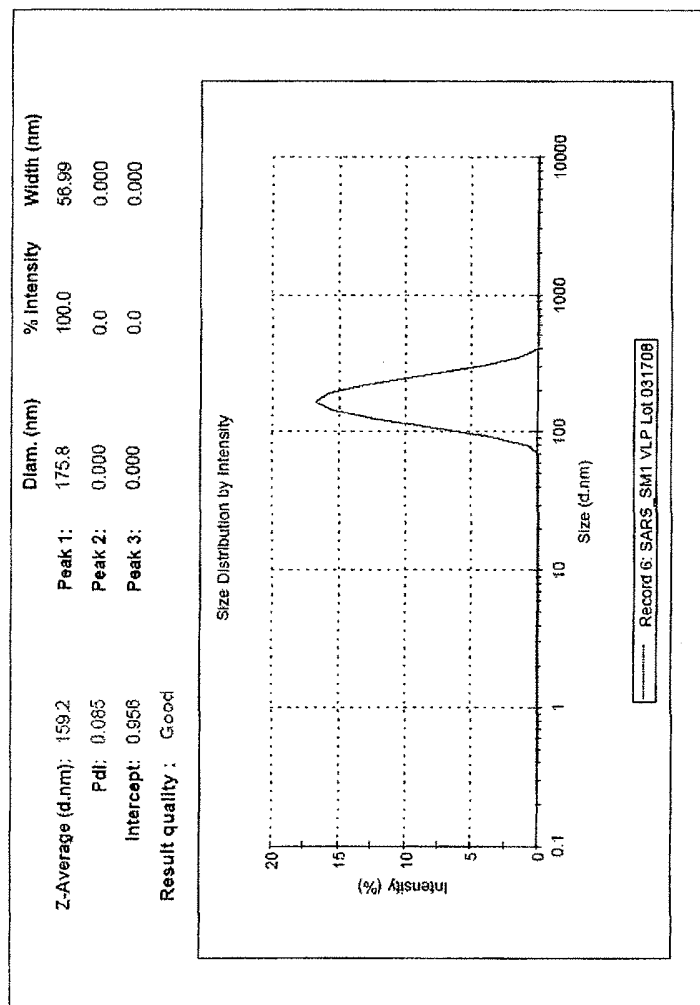


Figure 11

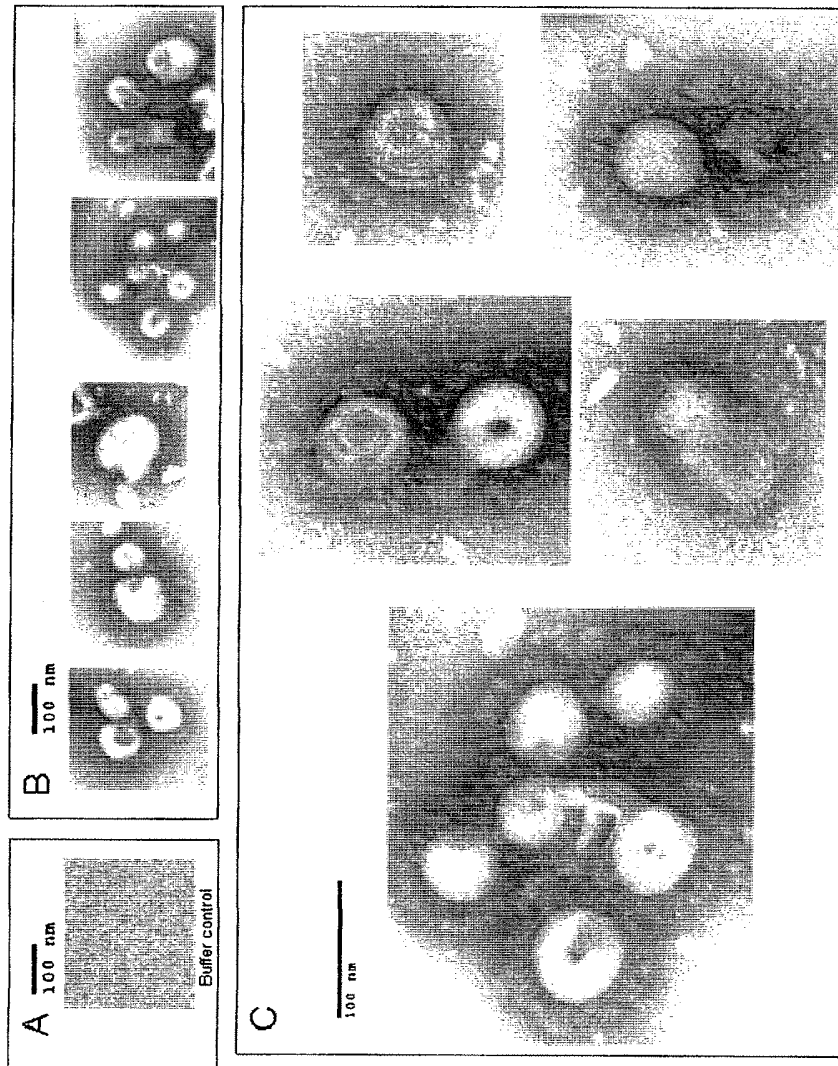


Figure 12

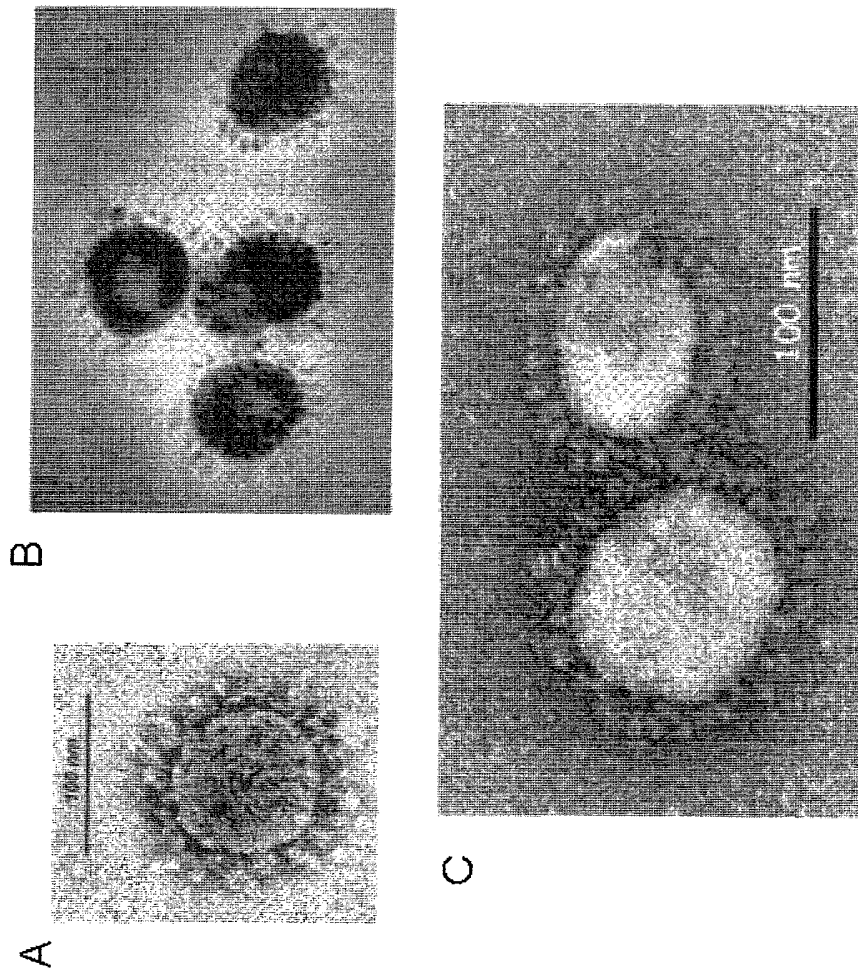


Figure 13

Influenza B/Florida/4/06 VLP Constructs

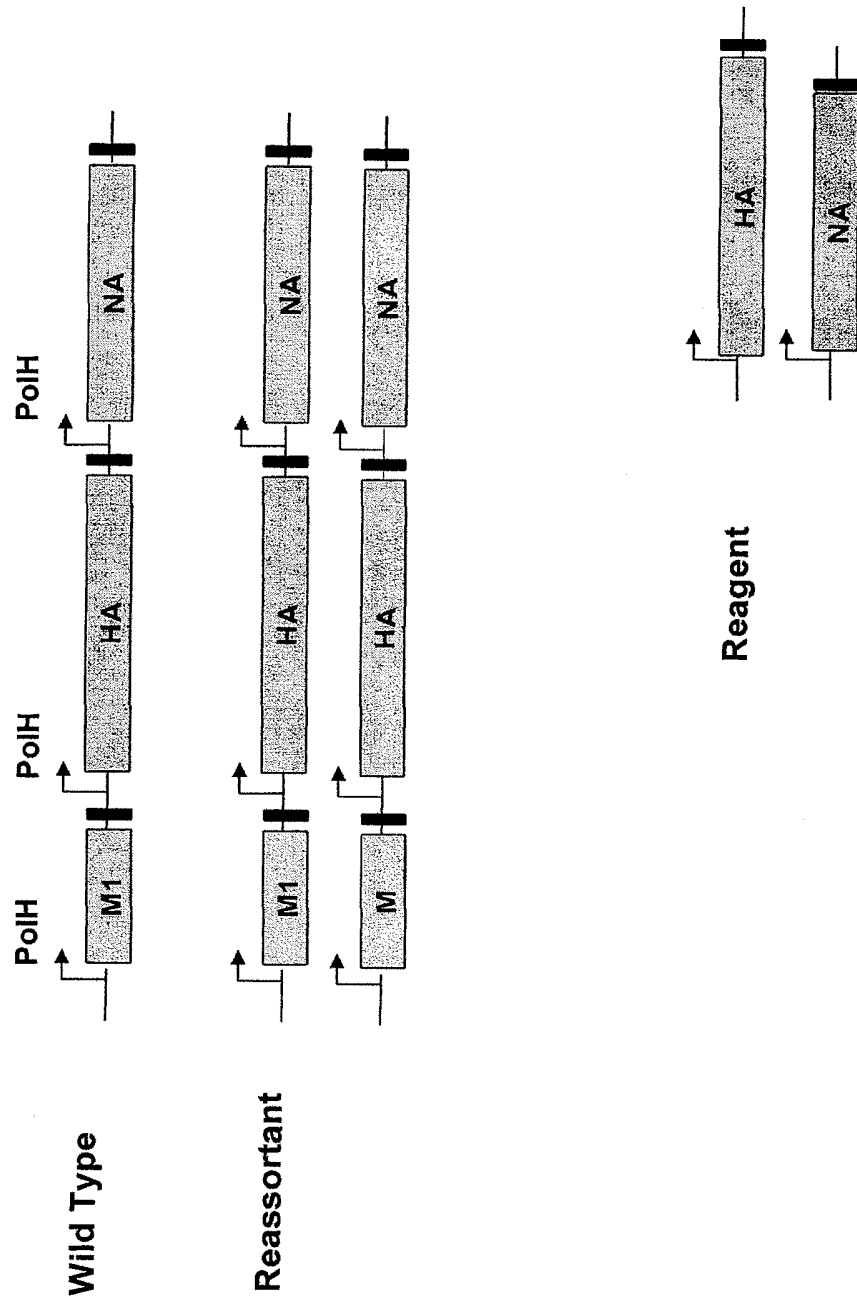
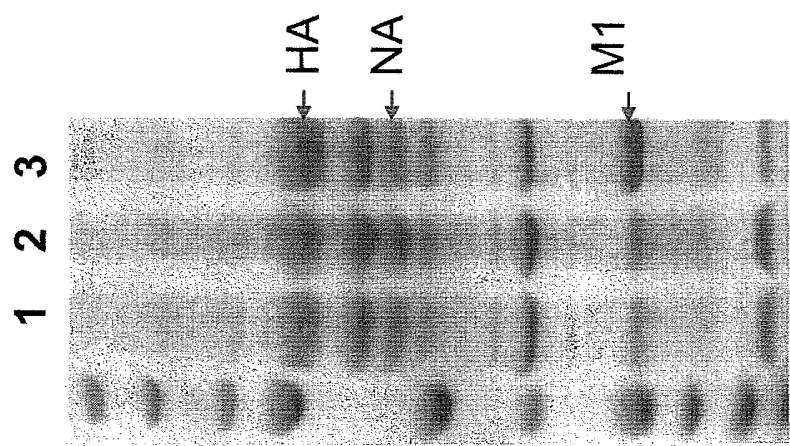


Figure 14

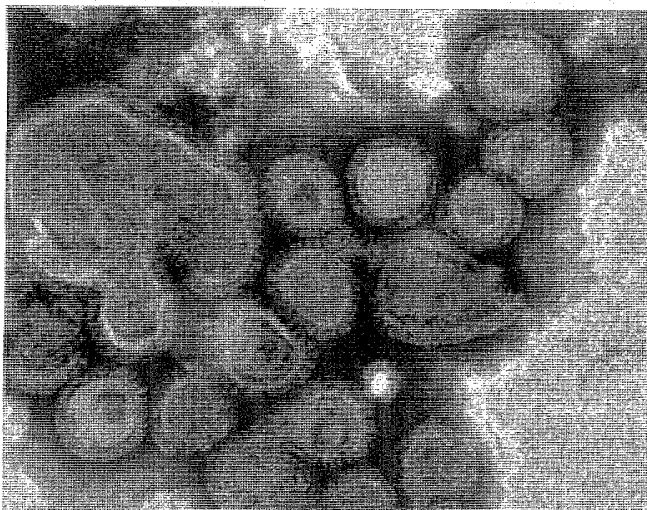


lane	Strain	HA ^G Pig	NA mu/mg
1	540 Inf B Fla WT M	4096	2055
2	539 Inf B Fla AA M	2048	1604
3	538 Inf B Fla Indo M1	16,384	1785

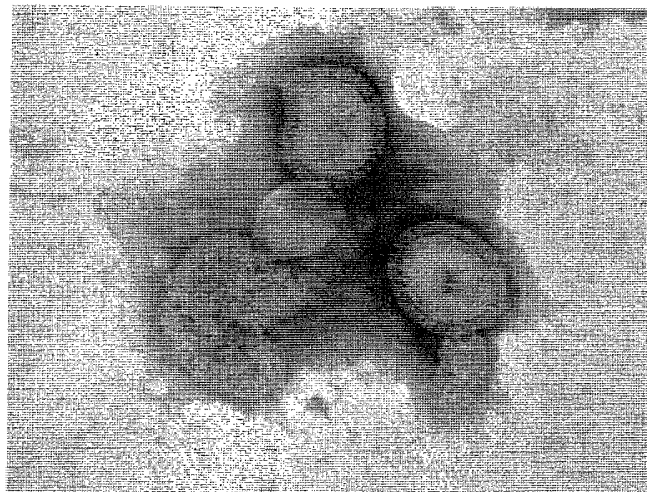
Figure 15

Influenza B/Florida/4/06

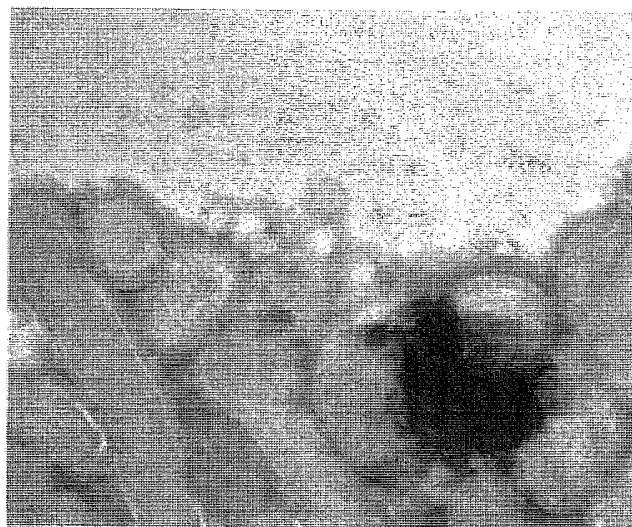
Avian M1 (BV538)



B/AA M1 (BV539)



wt M1 (BV540)



100 nm

Direct Mag: 120000x

REFERENCES CITED IN THE DESCRIPTION

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9459D

EP2540312

Madárinfluenza kiméra VLP-k

Szabadalmi igénypontok

1. Eljárás influenzavírus-szerű részecske (VLP)-termelés hatékonyságának növelésére gazdasejtben, amelynek során expresszáltatunk madárinfluenza mátrixfehérjét (M1) és szezonális humán influenza hemagglutinin (HA) és neuraminidáz (NA) fehérjét gazdasejtben, ahol a madárinfluenza M1 fehérje egy A/Indonesia/5/05 influenza M1 fehérje, és ahol a VLP-termelés megnövelt hatékonysága olyan gazdasejthez van viszonyítva, amely egy szezonális influenza M1 fehérjével együtt expresszálja a HA és NA fehérjéket.
2. Az 1. igénypont szerinti eljárás, amelynél az influenza HA fehérje rendelkezik hemagglutinin aktivitással.
3. Az 1. igénypont szerinti eljárás, amelynél az influenza NA fehérje rendelkezik neuraminidáz aktivitással.
4. Az 1. igénypont szerinti eljárás, amelyben a gazdasejtet a következők által alkotott csoportból választjuk: élesztősejt, rovarsejt, kéléltű-eredetű sejt, madársejt és emlőssejt.
5. A 4. igénypont szerinti eljárás, amelyben a gazdasejt a következők által alkotott csoportból választott rovarsejt: Sf9 sejt, Sf21 sejt és *Trichoplusia ni* sejt.
6. Az 5. igénypont szerinti eljárás, amelyben a rovarsejt Sf9 sejt.