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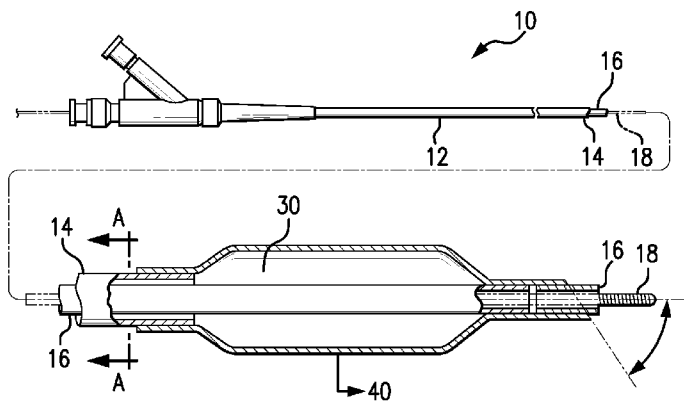


FIG. 1a

(57) Abstract: A drug delivery balloon is provided, the a balloon having an outer surface, and a tunable coating disposed on at least a length of the balloon surface. The tunable coating includes a first therapeutic agent and a first excipient, and can include a second therapeutic agent and a second excipient. The first and second therapeutic agents have different dissolution rates during balloon inflation and therefore provide a coating that is tunable.

COATINGS WITH TUNABLE MOLECULAR ARCHITECTURE FOR DRUG-COATED BALLOON

CROSS REFERENCE TO RELATED APPLICATIONS

This application claims priority to U.S. Provisional Patent Application
5 Serial No. 12/636,079 filed December 11, 2009, the contents of which is hereby
incorporated within by reference in its entirety.

FIELD OF THE INVENTION

The disclosed subject matter is related to the delivery of drugs from an
insertable medical device. More particularly, the disclosed subject matter relates to a
10 medical device including a balloon for delivery of a therapeutic agent, the balloon
having a tunable, durable coating disposed on its outer surface.

BACKGROUND OF THE INVENTION

Atherosclerosis is a syndrome affecting arterial blood vessels. A
hallmark of atherosclerosis is a chronic inflammatory response in the walls of arteries,
15 in large part due to the accumulation of lipid, cholesterol, leucocytes, and other
inflammatory cells and the formation of plaque on the arterial wall. Atherosclerosis is
commonly referred to as hardening of the arteries. Angioplasty is a vascular
interventional technique involving mechanically widening an obstructed blood vessel,
typically caused by atherosclerosis.

20 During angioplasty, a catheter having a tightly folded balloon is
inserted into the vasculature of the patient and is passed to the narrowed location of
the blood vessel at which point the balloon is inflated to a fixed size using an inflation
fluid, typically angiographic contrast media. Percutaneous coronary intervention
(PCI), commonly known as coronary angioplasty, is a therapeutic procedure to treat
25 the stenotic coronary arteries of the heart, often found in coronary heart disease.

In contrast, peripheral angioplasty, commonly known as percutaneous
transluminal angioplasty (PTA), refers to the use of mechanical widening of blood
vessels other than the coronary arteries. PTA is most commonly used to treat
narrowing of the leg arteries, especially, the iliac, external iliac, superficial femoral
30 and popliteal arteries. PTA can also treat narrowing of veins, and other blood vessels.

Although the blood vessel is often successfully widened by angioplasty, sometimes the treated wall of the blood vessel experienced abrupt closure after balloon inflation or dilatation due to acute recoil or vasospasm. One solution to such collapse is stenting the blood vessel to prevent collapse. A stent is a device,
5 typically a metal tube or scaffold, that is inserted into the blood vessel following angioplasty, in order to hold the blood vessel open.

While the advent of stents eliminated many of the complications of abrupt vessel closure after angioplasty procedures, within about six months of stenting, a re-narrowing of the blood vessel can form, a condition known as
10 restenosis. Restenosis was discovered to be a response to the injury of the angioplasty procedure and is characterized by a growth of smooth muscle cells—analogueous to a scar forming over an injury. As a solution, drug eluting stents were developed to address the reoccurrence of the narrowing of blood vessels. One example of a drug
15 eluting stent is a metal stent that has been coated with a drug that is known to interfere with the process of restenosis. A potential drawback of certain drug eluting stents is known as late stent thrombosis, which is an event in which blood clots inside the stent.

Drug eluting balloons are believed to be a viable alternative to drug eluting stents in the treatment of atherosclerosis. In a study which evaluated
20 restenosis and the rate of major adverse cardiac events such as heart attack, bypass, repeat stenosis, or death in patients treated with drug eluting balloons and drug eluting stents, the patients treated with drug eluting balloons experienced only 3.7 percent restenosis and 4.8% MACE (major adverse coronary events) as compared to patients treated with drug eluting stents, in which restenosis was 20.8 percent and 22.0 percent
25 MACE rate. (See, PEPCAD II study, Rotenburg, Germany).

Although drug eluting balloons are a viable alternative, and in some cases appear to have greater efficacy than drug eluting stents as suggested by the PEPCAD II study, drug eluting balloons present challenges due to the very short period of contact between the drug coated balloon surface and the blood vessel wall.
30 In particular, the balloon can only be inflated for less than one minute, and is often inflated for only thirty seconds. Therefore, an efficacious, therapeutic amount of drug must be transferred to the vessel wall within a thirty-second to one-minute time

period. For the peripheral vasculature, the allowable inflation times can be greater than one minute, but are still measured in minutes. Thus, there are challenges specific to drug delivery via a drug coated balloon because of the necessity of a short inflation time, and therefore time for drug or coating transfer--a challenge not presented by a
5 drug eluting stent, which remains in the patient's vasculature once implanted.

Other considerations are the current theories about the mechanism by which a drug coated balloon transfers drug to the vessel wall. One theory, for example, is that upon balloon expansion, drug mechanically fractures or dissolves from the coating, diffuses to the vessel wall and then permeates into the vessel wall.
10 A second theory is that upon balloon expansion the balloon coating is transferred to the vessel wall, and then drug permeates into the vessel wall from the coating adhered to the vessel wall. Another theory is that the balloon expansion creates tears and microfissures in the vessel wall, and a portion of the coating inserts into the tears and microfissures. Drug then permeates into the vessel wall from the coating within the
15 tears and fissures. Yet another theory is that upon balloon expansion, a layer of dissolved drug and coating excipients is formed at a high concentration on the vessel wall as a boundary layer. The drug diffuses and permeates from this boundary layer into the vessel wall. In most of these theories, the drug transfers from the balloon to the circulation or the vascular wall tissue upon fracture of the coating due to inflation
20 of the balloon and occurs within one minute, and preferably within 30 seconds. Therefore, a need exists for a drug coated balloon having efficient drug transfer to a vessel wall.

Various embodiments of drug-coated balloons have been proposed, including balloons with a therapeutic agent disposed directly on the balloon surface
25 and balloons having various protective sheaths. However, not all embodiments result in an efficacious response in reducing restenosis after balloon and bare metal stent trauma.

Therefore, a need exists for a drug eluting balloon and more particularly, a balloon coated with a therapeutic agent that provides for effective
30 delivery kinetics of the therapeutic agent from the surface of the balloon.

SUMMARY OF INVENTION

The purpose and advantages of the disclosed subject matter will be set forth in and apparent from the description that follows, as well as will be learned by practice of the disclosed subject matter. Additional advantages of the disclosed subject matter will be realized and attained by the methods and systems particularly pointed out in the written description and claims hereof, as well as from the appended drawings.

In accordance with an aspect of the disclosed subject matter, a method is provided for coating a medical device including a body having an outer surface. For example, the medical device is a drug delivery balloon or a balloon catheter. The method includes selecting a cytostatic therapeutic agent, selecting at least one excipient, and blending the cytostatic agent and excipient to define a coating. In one embodiment, the cytostatic therapeutic agent and at least one excipient has a weight ratio of about 20:1 to about 1:20, and defines a coating, which provides increased efficiency of therapeutic transfer to a body lumen. In another embodiment, the cytostatic therapeutic agent and polymeric excipient define a coating in which at least one polymeric excipient has a polydispersity index from about 1.05 to about 10, and provides increased efficiency of therapeutic transfer to a body lumen.

The method can include adding or blending a plasticizer with the cytostatic therapeutic agent and the excipient. For example, but not limitation, the plasticizer includes glycerol. In one embodiment, the excipient to plasticizer weight ratio is from about 20:1 to about 1:20 and the cytostatic therapeutic agent and the at least one excipient have a weight ratio of about 20:1 to about 1:20.

In another embodiment, the medical device is a drug delivery balloon having a coating tuned to have a dissolution rate of about 10 seconds to about 1h. For the therapeutic agent, delivery with balloon inflation occurs in less than 60 seconds and preferably less than 30 seconds.

In yet another aspect of the disclosed subject matter, a method is provided for tuning the solubility of a coating for application to a medical device. In one embodiment, the excipient is modified prior to blending the cytostatic therapeutic agent and excipient to achieve desired delivery kinetics. In one embodiment, the excipient is modified by positively charging the excipient. In another embodiment,

the excipient includes a cyclic and aliphatic carbon chain and the excipient is modified by adjusting the ratio of cyclic chain to aliphatic chain. Advantageously, the adjusted chain ratio results in reduced elasticity and/or reduced release rate of the therapeutic agent. For example but not limitation, the excipient can be
5 poly(vinylpyrrolidone), poly(ethylene glycol), or poly(ester amide) polymer.

The excipient can be modified by grafting a low molecular weight polyethylene glycol molecule to the excipient. In this regard, the modified excipient exhibits increased adhesion to a vessel wall. In another embodiment, the excipient is modified by increasing the crystallinity of the excipient. For example and not
10 limitation, the excipient is poly(L-lactide-co-caprolactone) polyester, and the crystallinity of the excipient is modified by adjusting the content of L-lactide. In this regard, the content of L-lactide can be increased, thereby defining a coating having greater storage stability

The coatings of the disclosed subject matter can be applied to any
15 surface of the medical device. In one embodiment, it is applied to the outer surface of the medical device. In accordance with one embodiment, the coating is applied to the surface of the medical device by direct fluid application. The method can further include applying heat to the coated medical device to dry the coating.

It is to be understood that both the foregoing description is exemplary
20 and is intended to provide further explanation of the disclosed subject matter claimed to a person of ordinary skill in the art. The accompanying drawings are included to illustrate various embodiments of the disclosed subject matter to provide a further understanding of the disclosed subject matter. The exemplified embodiments of the disclosed subject matter are not intended to limit the scope of the claims.

25 **BRIEF DESCRIPTION OF DRAWINGS**

The disclosed subject matter will now be described in conjunction with the accompanying drawings in which:

FIG. 1A depicts a representative planar view of a medical device of the disclosed subject matter, which is shown as a balloon catheter for illustration and
30 not limitation.

FIG. 1B is a cross-sectional view taken along lines A-A in FIG. 1A in accordance with one embodiment of the disclosed subject matter.

FIG. 2 is a graph illustrating percent drug release as a function of drug, excipient and plasticizer ratio (D:E:P) in accordance with one embodiment of the disclosed subject matter.

FIG. 3 are optical micrographs (100X magnification) demonstrating the results of scratch tests of zotarolimus:PVP:glycerol coatings on glass slides at 10:1:0.4 (left panel) and 2:1:0.4 (right panel) in accordance with one embodiment of the disclosed subject matter.

FIG. 4 are optical micrographs (100X magnification) demonstrating the results of scratch tests of glass slide coatings of zotarolimus:PVP:Tween 20, 2:1:0.67 (left panel) and zotarolimus:PVP:Tween 80, 2:1:0.4 (right panel) in accordance with one embodiment of the disclosed subject matter.

FIG. 5 are optical micrographs (50X magnification) demonstrating the results of scratch tests of glass slide coatings of zotarolimus:PEG-PE, 2:1 (left panel) and zotarolimus:PEG-PE, 1:1 (right panel) in accordance with one embodiment of the disclosed subject matter.

FIG. 6 are optical micrographs demonstrating the results of scratch tests of glass slide coatings of zotarolimus:PEG-PE, 1:2 under brightfield (left panel, 50X magnification) or crossed polarizers (right panel, 200X magnification) in accordance with one embodiment of the disclosed subject matter.

DETAILED DESCRIPTION

Reference will now be made in detail to the various aspects of the disclosed subject matter. The method of the disclosed subject matter will be described in conjunction with the detailed description of the device, the figures and examples provided herein.

As disclosed herein, the devices and methods presented can be used for delivery within and/or treating of the lumen of a patient. In particular, the disclosed subject matter is particularly suited for treatment of the cardiovascular system of a patient, such as performance of angioplasty and delivery of a balloon expandable medical device, such as a stent, filter and coil.

As disclosed herein, a balloon catheter is provided for delivery of a therapeutic agent, the balloon including an outer surface having a tunable and durable coating disposed on at least a length of the outer surface. The tunable and durable coating includes a therapeutic agent and an excipient. The solubility of the coating *in vivo*, the biosolubility, of the coating is tunable based on the substances and concentrations chosen for the therapeutic agent and excipient.

Referring to Figure 1, for purposes of illustration and not limitation, an exemplary embodiment of balloon catheter device in accordance with the disclosed subject matter is shown schematically in Figures 1A and 1B. As depicted in Figures 1A and 1B, the balloon catheter device 10 generally includes an elongated catheter shaft 12 having a proximal end and having a distal end and an expandable balloon 30 located proximate to the distal end of the catheter shaft. The expandable balloon has an outer surface and an inner surface disposed at the distal end portion of the catheter shaft. In accordance with the disclosed subject matter, a tunable coating 40 is applied to at least one length of the balloon catheter, the tunable coating including a first therapeutic agent and a first excipient, and can include a second therapeutic agent and a second excipient, wherein the first and second therapeutic agents have different dissolution rates during balloon inflation. In accordance with a preferred embodiment, as illustrated by way of example and not limitation in Figure 1A, the coating is applied to at least one length of the outer surface of the balloon catheter.

The elongated catheter shaft 12 comprises an outer tubular member 14 and an inner tubular member 16. The outer tubular member 14 defines an inflation lumen 20 that can be disposed between the proximal end portion and the distal end portion of the catheter shaft 12. Specifically, as illustrated in Figure 1B, the coaxial relationship between the inner tubular member 16 and the outer tubular member 14 defines an annular inflation lumen 20. The expandable member 30 is placed in fluid communication with the inflation lumen 20. The inflation lumen can supply fluid under pressure, and establish negative pressure to the expandable member. The expandable member 30 can thus be inflated and deflated. The elongated catheter is sized and configured for delivery through a tortuous anatomy, and can further include a guidewire lumen 22 that permits it to be delivered over a guidewire 18. As illustrated in Figure 1B, the inner tubular member 16 defines the guidewire lumen 22

for the guidewire 18. Although Figures 1A and 1B illustrate the guidewire lumen as having an over-the-wire (OTW) construction, the guidewire lumen can be configured as a rapid-exchange (RX) construction, as is well known in the art.

As disclosed herein, the coating is tunable with respect to its solubility.

5 Therefore, the drug delivery balloon is able to provide the desired delivery kinetics as a result of its tunability. The choice of excipient is key in determining efficacy factors such as, retaining of the therapeutic agent during delivery, releasing of the therapeutic agent during deployment, minimizing systemic dosing, maximizing agent delivery efficiency and therapeutic effect, and preventing particulate generation and related
10 thromboses, among other factors.

As used in accordance with the disclosed subject matter, “tunable” refers to the ability to be tuned or adjusted for desired functioning. Accordingly, a tunable coating refers to a coating that can be adjusted according to various parameter discussed herein.

15 As disclosed herein, the balloon includes a tunable coating that comprises a therapeutic agent and an excipient. In accordance with one embodiment, the tunable coating includes a first therapeutic agent and a first excipient, and a second therapeutic agent and a second excipient. The coating has a biosolubility that is tunable based on the substances and concentrations chosen for each of the
20 therapeutic agent and excipient. Preferably, the therapeutic agents have different dissolution rates. The coating can include additional therapeutic agents and excipients.

In accordance with the disclosed subject matter, the solubility of the coating can be adjusted by modifying a number of factors, including excipient type,
25 composition and molecular weight of the excipient, modulation of excipient or polymer properties such as aqueous solubility, octanol/water partition coefficient, HLB (hydrophile-lipophile balance) number, glass transition temperature, degree of amorphous versus crystalline polymer, and orientation. Furthermore, the solubility or dissolution rates of the coating can be adjusted by varying the therapeutic agent
30 concentration, therapeutic agent to excipient ratio, or coating thickness. Accordingly, these factors can be varied in order to provide a coating with the desired solubility and drug delivery kinetics.

The tunable coating provides for dissolution rates during balloon inflation that can be characterized generally as ranging from fast, soluble, intermediate, slow, extra slow, and non-soluble. Depending on the target tissue or vasculature where the therapeutic agent is to be delivered, the coating can be tuned such that the dissolution rate provides for effective drug delivery and uptake. A “fast” coating dissolution rate will typically have a dissolution time of less than 1 minute. A “soluble” coating dissolution rate will typically have a dissolution time ranging from about 1 minute to about 1 hour. An “intermediate” coating dissolution rate will typically have a dissolution time ranging from about 1 hour to about 2 weeks. A “slow” coating dissolution rate will typically have a dissolution time ranging from about 2 weeks to about 3 months. An “extra slow” coating dissolution rate will typically have a dissolution time ranging from about 3 months to 2 years. A “non-soluble” coating dissolution rate will typically have a dissolution time greater than 2 years. However, it shall be noted that the specific dissolution of a coating composition is dependent upon an interplay between input factors and that the dissolution rates provided herein are, therefore, recited as ranges.

The excipients include various oil-based, biosoluble, and durable or biodurable substances that are suitable for the delivery of a therapeutic agent. Biosolubility indicates solubility in a relevant biological media, such as blood. A substance which is not intended to degrade in the body, or which degrades only very slowly, is biodurable.

In accordance with a preferred embodiment, the excipients of the disclosed subject matter are water soluble. The excipients can include non-ionic hydrophilic polymers. Non-ionic hydrophilic polymers include, but are not limited to, poly(vinyl pyrrolidone) (PVP, povidone), silk-elastin like polymer, poly(vinyl alcohol), poly(ethylene glycol) (PEG), pluronics (PEO-PPO-PEO), poly(vinyl acetate), poly(ethylene oxide) (PEO), PVP-vinyl acetate (copovidone), polysorbate 80 (Tween 80), and polysorbate 20 (Tween 20). Preferably, the molecular weight of non-ionic hydrophilic polymers can be less than 50 kDa for fast solubility. The excipient can also include fatty acids. Further, the excipient can be a lubricious material which improves spreading and uniformity of coating.

In accordance with one embodiment, the excipient consists of a biocompatible plasticizer. Alternatively, the plasticizer can be added to the excipient to keep it soft and pliable. Plasticizers can allow for greater coating flexibility and elongation to prevent coating cracking during inflation or brittleness. Plasticizers include, but are not limited to, glycerol, ethanol, dimethylsulfoxide, ethyl lactate, benzyl alcohol, benzyl benzoate, Cremophor EL, Vitamin E, tocopherol, liquid PEG (MW<1000), triethyl citrate, tributyl citrate, acetyl tributyl citrate, acetyl triethyl citrate, dibutyl phthalate, dibutyl sebacate, dimethyl phthalate, triacetin, propylene glycol, glycerin, 2-pyrrolidone, and combinations thereof. Preferably, a biocompatible plasticizer is used.

In accordance with yet another embodiment, sugars, polysaccharides or cellulose, can be used as binders for the particles. Polysaccharides include, but are not limited to, dextran, sulfonated dextran, hydrogenated dextran, chondroitin sulfate, sodium hyaluronate, hyaluronic acid, hyaluronan, chitosan, sodium alginate, sucrose, pectin, mannitol, carboxymethyl cellulose (CMC) sodium, methyl cellulose, ethyl cellulose, hydroxyethyl cellulose, hydroxypropyl cellulose, and hydroxypropylmethylcellulose. Certain negative charged polysaccharides will provide a mucoadhesive effect to enhance tissue drug retention. Furthermore, sugars such as mannitol will provide a decreased hygroscopic effect when blended with more moisture-sensitive active ingredients such as cytostatic drugs or moisture sensitive excipients. Water soluble cellulosic materials can enhance coating strength or brittleness.

In accordance with yet another embodiment, anti-coagulants can be used as an excipient. For example, heparin based polysaccharides can provide a minimally thrombogenic surface to prevent blood clotting on the balloon surface or minimize platelet activation induced by the procedure. Heparin based polysaccharides include, but are not limited to, heparin, heparin sulfate, heparin disaccharides, heparin fraction 1, heparin fraction 2, low molecular weight heparin, heparin ammonium, heparin calcium, heparin lithium, heparin lithium, and heparin zinc lithium. Low molecular weight heparin includes centaxarin, periodate-oxidized heparin, heparin sodium end-amidated, heparin sodium, and nitrous acid delaminated.

In accordance with a preferred embodiment of the disclosed subject matter, the excipient possesses a mucoadhesive property. This mucoadhesive property of the binder will lead to longer drug retention within the coating adhered to the vessel wall. In particular, positively charged excipients such as chitosan, negatively charged excipients such as some polysaccharides (e.g. carboxymethylcellulose, sodium hyaluronate, sodium alginate) and some non-ionic hydrophilic polymers exhibit mucoadhesive properties. Any of the above carboxylated materials can also be lightly activated with esters such as nitrophenolate or NHS-esters (N-hydroxy succinimide) for increased mucoadhesiveness. Alternatively, any above materials can be lightly thiolated for increased mucoadhesiveness and continued solubility.

Additionally or alternatively, the excipient is or includes a contrast agent, including but not limited to, Iopromide (Ultravist), Ioxaglate (Hexabrix), Ioversol (Optiray), Iopamidol (Isovue), Diatrizoate (Conray), Iodixanol (Visipaque), Iohexol (Omnipaque), and Iotrolan. At an intermediate coating thickness, a lower molecular weight (< 1 kDa) hydrophilic contrast agent such as Iopromide (Ultravist) would enable faster therapeutic release and a slightly higher viscous coating of the vessel wall as compared with drug alone. The contrast agents are lipophilic and can aid in drug uptake and retention into the tissue wall. In accordance with one embodiment, Ultravist and Optiray can be used given their more benign history of effects to smooth muscle and endothelial cells.

In accordance with yet another embodiment, excipients can consist of carboxylated aromatics similar in molecular structure to the structure used in contrast agents but without iodide substituents. These negatively charged carboxylated aromatic structures can be alkylated (C2-C12) to optimize drug tissue uptake, or halogenated with fluoride, chloride or bromide for the same reason. The negatively charged structures are beneficial for tissue adhesiveness.

Table 1 provides non-limiting examples of the solubility data for excipients that can be used in accordance with the disclosed subject matter:

Table 1: Solubility Enhancement of a Therapeutic Agent with Select Excipients

Solution (5% w/w)	Zotarolimus Solubility (µg/ml, n=3)
Phosphate buffered saline	0.53
PVP C-17	5.6 ± 1.6
Hydroxypropyl-β-cyclodextrin	11.6 ± 3.1
PEG 400	31.5 ± 3.5
Glycerol	43.2 ± 30.1
5% γ-Cyclodextrin	55.3 ± 34.3
Vitamin E TPGS	512 ± 49.5
Tween 20	732 ± 94.7
18:0 PEG2000 PE (PEG-PE)*	1020 ± 417

*1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine-N-[methoxy(polyethyleneglycol)-2000] (ammonium salt)

5

As illustrated in Table 1, the excipients provide for increased solubility for the cytostatic drug, zotarolimus, as compared to saline alone. The excipients Vitamin E TPGS, Tween 20, and PEG-PE demonstrate the largest increase in zotarolimus solubility.

10

Table 2 provides non-limiting examples of coating dissolution rates during balloon inflation and representative excipient examples.

Table 2: Examples of Delivery Kinetics and Expected Variable Ranges for Balloon Coatings

Coating Dissolution Rate (during balloon inflation)	Coating Dissolution Time	Representative Excipient Example
Fast	< 1 minute	Poly(vinylpyrrolidone) (PVP) (MW < 60 kDa) or Poly(ethylene glycol) (PEG) (lower MW < 35 kDa)
Soluble	1 min to 1 hour	Poly(vinylpyrrolidone) (PVP) (MW > 60 kDa) or Polyethylene oxide (PEO) (higher MW > 100

		kDa)
Intermediate	1 hour to 2 weeks	Silk-elastin like protein polymers
Slow	2 weeks – 3 months	Biodegradable polymer such as Poly(D,L-lactide-co-glycolide) (PLGA) (50:50)
Extra Slow	3 months - 2 years	Biodegradable polymer such as Poly(L-lactide-co-ε-caprolactone) (PLLA:PCL) (70:30)
Non-Soluble	> 2 years	Durable polymer such as Poly(vinylidene fluoride-co-hexafluoropropylene)

As illustrated in Table 2 above, for a “fast” coating dissolution rate, representative excipient examples include, without limitation, polyvinylpyrrolidone (PVP) with a molecular weight less than about 60 kDa, or polyethylene glycol (PEG) having a molecular weight less than about 35 kDa. The drug delivery mechanism and kinetics expected with this representative example include the release of the therapeutic agent with the coating during inflation. Further, the potential mucoadhesive polymer increases drug retention time on tissue or vasculature. Alternatively, or additionally, the lipophilic additive increases drug uptake in tissue.

As illustrated in Table 2 above, for a “soluble” coating dissolution rate, representative excipient examples include, without limitation, poly(vinylpyrrolidone) (PVP) having a molecular weight greater than about 60 kDa, or poly(ethylene glycol) (PEG) having a molecular weight greater than about 100 kDa. The drug delivery mechanism and kinetics expected with this representative example are similar to that of the “fast” coating dissolution rate, however, the slightly slower dissolution time allows for less drug wash off during balloon delivery before inflation.

As illustrated in Table 2 above, for an “intermediate” coating dissolution rate, representative excipient examples include, without limitation, silk-elastin like protein polymers. The drug delivery mechanism and kinetics expected with this representative example provides for enhanced systemic drug loss protection and absence of short-term solubility, therefore allowing for enhanced particulate safety. For an “intermediate” dissolution rate, the therapeutic agent is not released together with the coating but from the coating. The therapeutic agent release kinetics and transfer to tissue are significantly enhanced by mechanical action during balloon inflation.

Typically, these type of coating materials can be hydrophilic and can swell to some extent upon hydration to aid in fast drug release.

As illustrated in Table 2 above, for a “slow” coating dissolution rate, representative excipient examples include, without limitation, biodegradable polymers such as Poly(D,L-lactide-co-glycolide) (PLGA) (50:50). The coatings from biodegradable hydrophobic polymers will offer enhanced systemic drug loss protection and a better particulate safety profile. The therapeutic agent is not released together with the coating but from the coating. Drug release kinetics and transfer to tissue are significantly enhanced by mechanical action during balloon inflation. Techniques such as using a thin coating, a polymer with a low glass transition temperature (T_g), and amorphous material or low crystalline material can provide for a more rapid drug release profile when using a biodegradable polymer.

As illustrated in Table 2 above, for an “extra slow” coating dissolution rate, representative excipient examples include, without limitation, biodegradable polymers such as poly(L-lactide-co-ε-caprolactone) (PLLA:PCL) (70:30). The drug delivery mechanism and kinetics are similar to a “slow” coating dissolution rate, however the degradation time is significantly extended. These coatings will have more long term degradation and mechanical stability under storage.

As illustrated above, for a “non-soluble” coating dissolution rate, representative excipient examples include, without limitation, durable polymers such as poly(vinylidene fluoride-co-hexafluoropropylene). The drug delivery mechanism and kinetics are similar to both a “slow” and “extra slow” coating dissolution rate, however the material is non-biodegradable. These non-soluble coatings will have the most chemical and mechanical stability under storage than other types.

In accordance with one embodiment, the excipient is a durable or biodurable excipient. Certain representative examples of durable excipients are provided in Table 3.

Table 3. Examples of durable excipients

Excipient	Specific Description
Hydroxypropylmethylcellulose phthalate	Coating agent, can improve coating strength and be combined with plasticizer to reduce brittleness
PVP/PEO + acrylates + photoinitiator + PVP crosslinker	UV curable coating would retain drug during delivery and rapidly release drug after balloon expansion, lubricious
Polyethylene vinyl acetate phthalate	Coating agent, can improve coating strength and be combined with plasticizer to reduce brittleness
Poly(ϵ -caprolactone) (PCL); Poly (caprolactone) co-polymerized with lactide and/or glycolide	flexible material, low Tg and low crystallinity material can provide for rapid drug release
Poly(DL-lactide)	Flexible material, soluble in various solvents for spray processing, can improve coating strength and be combined with plasticizer to reduce brittleness
Poly(L-lactide-co- ϵ -caprolactone)(PLCL) (50:50)	Flexible material, soluble in various solvents for spray processing, can improve coating strength and be combined with plasticizer to reduce brittleness
PCL-PEG (di and tri-block copolymers); Poly(trimethylenecarbonate) copolymerized with caprolactone, lactide, glycolide and/or other comonomers; Poly(lactide)-PEG(di- and tri-block copolymers)lactide copolymerized with caprolactone, glycolide and/or other comonomers	Flexible material, can modulate PEG length for varying degrees of water absorption and release rates
Polyethylene vinyl acetate	Flexible material; biocompatible
PVDF and PVDF copolymers (e.g., Solef)	Blood compatible, low thrombogenicity, flexible materials with good temperature stability, soluble in acetone for spray processing
PEA based on leucine, alanine, phenylalanine, any other amino acid, or combinations thereof	Polymer structure can be fine tuned to modulate drug miscibility and release profile, soluble in various solvents for spray processing
Phosphorylcholine derivatized polymers including acrylates, methacrylates, PEA and aliphatic polyesters	Non-thrombogenic polymer structure that can be fine tuned to modulate drug miscibility and release profile, soluble in various solvents for spray processing
Silk-elastin like polymers	Thermally crosslinked, biocompatible

PEG-based thiol-ene crosslinked gels	Enhanced mucoadhesiveness, low thrombogenicity
Polyacrylic acid (Carbomer)	Mucoadhesive, swells but does not dissolve in aqueous media
PBMA copolymers; HPMA; PHEMA-co-PAAm	Good adhesion
Hydrophilic polyurethane containing a PEO or other hydrophilic soft segment	Blood compatible, flexible material, can modulate water absorption and drug release rate with soft segment MW and content

The durable excipients in accordance with the disclosed subject matter bind the therapeutic agent to the balloon or device surface and protect the agent during delivery to a treatment location. For example, after expansion of the balloon at the treatment site, the balloon will contact the vessel wall and the therapeutic agent will be rapidly released to cause the desired beneficial effect. During and after inflation, the biodurable excipient will remain on the balloon with no particulate loss. To release the drug at a sufficient rate from these polymers, it is necessary to have high drug concentrations above percolation, with drug to polymer ratios at about 1:1 or higher.

In accordance with one embodiment, a primer coating is necessary to allow for good and adequate adhesion such that, for example, a stent and coating can be removed in a safe manner.

In accordance with the disclosed subject matter, the outer surface of the balloon has a tunable coating that is disposed on at least a length of the outer surface. Preferably, the tunable coating includes a first therapeutic agent and a first excipient and a second therapeutic agent and a second excipient. In accordance with a preferred embodiment, the first and second therapeutic agents have different dissolution rates during balloon inflation. Thus, the desired coating dissolution rates can be tunable and achieved as desired for either drug kinetics or safety profile. The delivery of the therapeutic agents can be modified and optimized to meet the therapeutic need. Furthermore, depending on the excipients used, the therapeutic agents can be released from the excipient or coating or with the excipient or coating. In accordance with one embodiment, the first therapeutic agent is released from the coating, and the second therapeutic agent is released with the coating.

In one embodiment, the first therapeutic agent is different than the second therapeutic agent. Alternatively, however, the therapeutic agents can be the same.

5 In accordance with another embodiment, the coating can also include a third therapeutic agent and a third excipient. The therapeutic agents and excipients can be applied simultaneously to the balloon surface or they can be applied separately.

In accordance with yet another embodiment, the disclosed subject matter includes a balloon having a the tunable coating including a cytostatic drug and at least one excipient, wherein in the coating at least one polymeric component has a
10 polydispersity index from about 1.05 to about 10, more preferably from 1.05 to 5. The polydispersity index (PDI), is a measure of the distribution of molecular mass in a given polymer sample. The PDI calculated is the weight average molecular weight divided by the number average molecular weight. It indicates the distribution of individual molecular masses in a batch of polymers. A smaller PDI value provides a
15 more consistent dissolution rate among the polymeric excipient molecules.

It has been found that coatings can be tunable to achieve desirable dissolution and drug delivery kinetics. In this regard, the choice of an excipient or modified excipient can be important to define coatings exhibiting efficacy factors such as but not limited to: how the therapeutic agent is retained during delivery, how
20 the agent is released during balloon inflation, minimizing systemic drug dosing, maximizing agent delivery efficiency and therapeutic effect, and preventing particulate loss, related thromboses and embolic events.

Accordingly, in one aspect of the disclosed subject matter, a method is provided for coating a medical device, such as a drug coated balloon. The coating
25 includes a cytostatic therapeutic agent and an excipient having a tunable molecular architecture. As used herein the phrase "tunable molecular architecture" means selection of an appropriate excipient composition, molecular structure, functionality, and morphology including appropriate modifications to yield the desired coating dissolution and drug delivery kinetics. The molecular architecture can be tuned
30 through the design of input variables such as monomer/polymer composition, aromaticity, hydrophilicity, molecular charge, neutrality, aliphatic chain length, density of functional groups, molecular weight, aqueous solubility, octanol/water

partition coefficient, HLB number, glass transition temperature, and percent crystallinity. The method of the disclosed subject matter advantageously is capable of providing desired delivery kinetics as a result of its tunability.

In accordance with the disclosed subject matter, the method includes selecting a cytostatic therapeutic agent and an excipient, and blending or mixing the cytostatic agent and excipient to define a coating. The method can further include tuning the molecular structure of the coating such that specific characteristics of the coating that are important to product performance can be adjusted and optimized. Some of these characteristics include coating solubility, coating hydrophilicity, coating adhesion and cohesion, coating stability under sterilization and storage, drug release kinetics, drug solubility and stability, and safety profile including particulate hazard and re-endothelialization.

For example and not limitation, the molecular architecture of the excipient can be modified and tuned through the adjustment of several input parameters, as described in Table 4 below.

Table 4: Inputs that Affect Molecular Architecture and Excipient or Coating Characteristics

Excipient Modification	Effect(s) of Increase	Effect(s) of Decrease	Example
Monomer/Polymer Composition	Choice of excipient or coating composition has a large effect on all coating characteristics.		Poly(vinylpyrrolidone) (PVP) or contrast agent for soluble coating agent. Thin poly(vinylidene fluoride-co-hexafluoropropylene) (PVDF-HFP) for durable coating.
Monomer/Polymer Composition - Intramolecular attraction: H Bonding versus Steric Hindrance	A high interaction between polymer chains can provide for increased coating mechanical stability, cohesion, higher crystallinity and greater molecular packing density.	More spacing between chains can provide more amorphous content and lower crystallinity for faster solubility and decreased stability of material.	Increased L-lactide content in poly(L-lactide-co-caprolactone) copolymers can lead to higher crystallinity. Polyurethane ureas exhibit increased H-bonding and mechanical stability.

Hydrophilicity	Will provide faster water absorption, faster drug release, and faster coating dissolution.	Slower water absorption, slower drug release, and slower coating dissolution.	Non-ionic hydrophilic polymers such as PVP and polyethylene glycol (PEG) are water soluble and provide for fast coating dissolution and drug delivery. Can increase or decrease PEG content as a soft segment in polyurethanes to increase or decrease hydrophilicity. Non-ionic hydrophobic polymers such as PVDF-HFP or PCL are non-water soluble.
Molecular Charge or Neutrality	Charged molecules tend to possess mucoadhesive properties.	Lower charge or more neutral species can exhibit less mucoadhesive properties.	Charged polysaccharides such as carboxymethylcellulose, sodium hyaluronate, chitosan, and sodium alginate can exhibit mucoadhesive properties as well as complex and/or physically bind negatively charged molecules.
Cyclic Chain	Cyclic chain will increase chain rigidity (higher Tg), therefore slow down the release rate. The elasticity will also be reduced as a result.	Lower chain rigidity (lower Tg), faster release rate and higher % elongation.	Adjusting the ratio of cyclic to aliphatic chain in poly(ester amide) (PEA) polymers.
Aliphatic Chain Length	Increased carbon chain length will lend towards increased flexibility (lower Tg) of material for higher	Decreased carbon chain lengths will lend towards increased stiffness	Adjusting length of hydrocarbon chains in poly(ester amide) (PEA) polymers.

	% elongation and slower drug release.	(higher Tg) of material for lower % elongation and faster drug release.	
Density of Functional Groups	Increase density of grafted signaling or other molecules such as RGD sequences for cell attachment.	Lower density of functional groups and attached ligands can allow for decreased steric hindrance and increased bioavailability.	RGD grafted low MW PEG can provide for increased adhesion of released coating to the vascular endothelial cell wall.
Molecular Weight	Higher molecular weight material will tend towards higher spray viscosity and lower solubility in various spray solvents and in aqueous media. In general, a higher molecular weight for provide for increased mechanical strength and cohesion of a coating to a certain cut-off value. Higher MW will also translate to slower release rate.	Lower molecular weight will tend towards lower spray viscosity and increased solubility in various spray solvents and in aqueous media. Lower Mw will also translate to faster release rate.	Lower molecular weight (< 60 kDa). poly(vinylpyrrolidone) is easily spray coated due to its low spray viscosity and high solubility in aqueous and organic media.
Glass Transition Temperature	Increased EtO sterilization stability and slower drug release. Less flexible coating for decreased handling and catheter processing performance.	Decreased EtO sterilization stability leading to coating flow and faster drug release. More flexible coating for enhanced catheter processing performance.	Certain low Tg Poly(ϵ -caprolactone) based materials can provide a very fast burst release of drug up to 99%+ release at 1 day. Adding a plasticizer such as glycerol to non-ionic hydrophilic polymers such as poly(vinylpyrrolidone) can lower the dry

			coating Tg to increase coating flexibility.
% Crystallinity	Higher storage stability, slower drug release, less flexible coating.	Lower storage stability, faster solubility, faster drug release, more flexible coating.	Increasing or decreasing L-lactide) content in poly(L-lactide-co-caprolactone) polyester copolymers. Adding HFP to PVDF copolymer to increase coating flexibility and drug release.

In accordance with the disclosed subject matter, the coating can be applied to the medical device by processes such as dip-coating, pipette coating, syringe coating, air assisted spraying, electrostatic spraying, piezoelectric spraying, spray drying, pneumatic spray, ultrasonic spray, spray with patterning, electrospinning, direct fluid application, or other means as known to those skilled in the art. The coating can be applied over at least a length or the entirety of the balloon or medical device. By way of example, and not limitation, certain coating processes that can be used with the instant disclosed subject matter are described in U.S. Patent No. 6,669,980 to Hansen; U.S. Patent No. 7,241,344 to Worsham; and U.S. Publication No. 20040234748 to Stenzel, the entire disclosures of which are hereby incorporated by reference. In accordance with one embodiment of the disclosed subject matter, the medical device is a balloon catheter and the coating can be applied to either a folded or inflated balloon. Furthermore, the coating can be directly applied into the folds of the folded balloons. The coating characteristics are affected by process variables. For example, for dip-coating process, coating quality and thickness can vary as an effect of variables such as number, rate, and depth of dips along with drying time and temperature.

In accordance with one embodiment, the balloon can be sprayed with therapeutic agent encapsulated in the durable excipient solution. Spray solvents can consist of the following class III solvents including but not limited to acetone, anisole, 1-butanol, 2-butanol, butyl acetate, tert-butylmethyl ether, cumene, dimethyl sulfoxide, ethanol, ethyl acetate, ethyl ether, ethyl formate, heptane, hexane,

cyclohexane, isobutyl acetate, isopropyl acetate, methyl acetate, 3-methyl-1-butanol, methylethyl ketone, methylisobutyl ketone, cyclohexanone, 2-methyl-1-propanol, pentanol, 1-pentanol, 1-propanol, and propyl acetate, or blends thereof.

Additional spray solvents that can be used or blended with class III solvents include class II spray solvents. The class II spray solvents include but are not limited to, acetonitrile, chloroform, 1,2-dichloroethane, dichloromethane, 1,2-dimethoxyethene, N,N-dimethylacetamide, N,N-dimethylformamide, 1,4-dioxane, 2-ethoxyethanol, ethylene glycol, formamide, hexane, methanol, 2-methoxyethanol, methyl butyl ketone, methylcyclohexane, N-methylpyrrolidone, nitromethane, pyridine, sulfolane, tetrahydrofuran, tetralin, toluene, 1,1,2-trichloroethene, and xylene.

In accordance with the disclosed subject matter, the excipient and therapeutic agent coating process can occur aseptically or be followed with terminal sterilization methods such as E-beam, gamma irradiation, or ethylene oxide sterilization.

In accordance with the disclosed subject matter, excipients are utilized together with the therapeutic agent in the coating at ratios ranging from 1:20 to 20:1 excipient:drug by weight, preferably from 1:10 to 10:1, more preferably from 1:2 to 2:1. Preferably, the coating includes a plasticizer. In this regards, the excipient to plasticizer weight ratio is from about 20:1 to about 1:20, more preferably from 10:1 to 1:1.

In accordance with another embodiment of the disclosed subject matter, the coating includes various layers. In one embodiment, the coating includes first and second layers adsorbed to the surface of the balloon. The first layer typically consists of one therapeutic agent and one excipient and the second layer typically consists of a second therapeutic agent and second excipient. The drug coated balloon is designed such that the first and second layers each have a dissolution rate. Preferably, the dissolution profile of the first layer is different than the dissolution profile of the second layer. Providing layers with various dissolution profiles allows the coating to be tuned to an optimized range.

In accordance with yet another embodiment, the disclosed subject matter includes a method of increasing the efficiency of therapeutic transfer to a body

lumen by implanting or inserting a medical device in a body lumen. The medical device includes an expandable member having an outer surface and a coating disposed on the outer surface of the medical device, the coating including a therapeutic agent and an excipient.

5 For example and not limitation, the therapeutic agent or drug can include anti-proliferative, anti-inflammatory, antineoplastic, antiplatelet, anti-coagulant, anti-fibrin, antithrombotic, antimitotic, antibiotic, antiallergic and antioxidant compounds. Thus, the therapeutic agent can be, again without limitation, a synthetic inorganic or organic compound, a protein, a peptide, a polysaccharides and
10 other sugars, a lipid, DNA and RNA nucleic acid sequences, an antisense oligonucleotide, an antibody, a receptor ligands, an enzyme, an adhesion peptide, a blood clot agent including streptokinase and tissue plasminogen activator, an antigen, a hormone, a growth factor, a ribozyme, and a retroviral vector. Preferably, however, the therapeutic agents include a cytostatic drug. The term "cytostatic" as
15 used herein means a drug that mitigates cell proliferation, allows cell migration, and does not induce cell toxicity. These cytostatic drugs, include for the purpose of illustration and without limitation, macrolide antibiotics, rapamycin, everolimus, zotarolimus, biolimus, novolimus, myolimus, temsirolimus, deforolimus, structural derivatives and functional analogues of rapamycin, structural derivatives and
20 functional analogues of everolimus, structural derivatives and functional analogues of zotarolimus and any macrolide immunosuppressive drugs. The term "antiproliferative" as used herein means a drug used to inhibit cell growth, such as chemotherapeutic drugs. Some non-limiting examples of antiproliferative drugs include taxanes, paclitaxel, and protaxel.

25 Therefore, in accordance with a preferred embodiment, a balloon for delivery of a cytostatic drug is provided. The outer surface of the balloon includes a tunable coating, the tunable coating including a first cytostatic drug and a first excipient and a second cytostatic drug and a second excipient. The first and second cytostatic drugs preferably have different dissolution rates during balloon inflation.
30 The various dissolution rates allow for more effective and efficient delivery of the therapeutic agent.

With reference to the balloon construction, a polymeric expandable balloon material is preferred. For example, the polymeric material utilized to form the balloon body can be compliant, non-compliant or semi-compliant polymeric material or polymeric blends.

5 In one embodiment, the polymeric material is compliant such as but not limited to a polyamide/polyether block copolymer (commonly referred to as PEBA or polyether-block-amide). Preferably, the polyamide and polyether segments of the block copolymers can be linked through amide or ester linkages. The polyamide block can be selected from various aliphatic or aromatic polyamides
10 known in the art. Preferably, the polyamide is aliphatic. Some non-limiting examples include nylon 12, nylon 11, nylon 9, nylon 6, nylon 6/12, nylon 6/11, nylon 6/9, and nylon 6/6. Preferably, the polyamide is nylon 12. The polyether block can be selected from various polyethers known in the art. Some non-limiting examples of polyether segments include poly(tetramethylene ether), tetramethylene ether,
15 polyethylene glycol, polypropylene glycol, poly(pentamethylene ether) and poly(hexamethylene ether). Commercially available PEBA material can also be utilized such as for example, PEBAX® materials supplied by Arkema (France). Various techniques for forming a balloon from polyamide/polyether block copolymer is known in the art. One such example is disclosed in U.S. Patent No. 6,406,457 to
20 Wang, the disclosure of which is incorporated by reference.

 In another embodiment, the balloon material is formed from polyamides. Preferably, the polyamide has substantial tensile strength, be resistant to pin-holing even after folding and unfolding, and be generally scratch resistant, such as those disclosed in U.S. Patent No. 6,500,148 to Pinchuk, the disclosure of which is
25 incorporated herein by reference. Some non-limiting examples of polyamide materials suitable for the balloon include nylon 12, nylon 11, nylon 9, nylon 69 and nylon 66. Preferably, the polyamide is nylon 12. Other suitable materials for constructing non-compliant balloons are polyesters such as poly(ethylene terephthalate) (PET), Hytrel thermoplastic polyester, and polyethylene.

30 In another embodiment, the balloon is formed of a polyurethane material, such as TECOTHANE® (Thermedics). TECOTHANE® is a thermoplastic, aromatic, polyether polyurethane synthesized from methylene diisocyanate (MDI),

polytetramethylene ether glycol (PTMEG) and 1,4 butanediol chain extender. TECOTHANE® grade 1065D is presently preferred, and has a Shore durometer of 65D, an elongation at break of about 300%, and a high tensile strength at yield of about 10,000 psi. However, other suitable grades can be used, including
5 TECOTHANE® 1075D, having a Shore D hardness of 75. Other suitable compliant polymeric materials include ENGAGE® (DuPont Dow Elastomers (an ethylene alpha-olefin polymer) and EXACT® (Exxon Chemical), both of which are thermoplastic polymers. Other suitable compliant materials include, but are not limited to, elastomeric silicones, latexes, and urethanes.

10 The compliant material can be cross linked or uncrosslinked, depending upon the balloon material and characteristics required for a particular application. The presently preferred polyurethane balloon materials are not crosslinked. However, other suitable materials, such as the polyolefinic polymers ENGAGE® and EXACT®, are preferably crosslinked. By crosslinking the balloon
15 compliant material, the final inflated balloon size can be controlled. Conventional crosslinking techniques can be used including thermal treatment and E-beam exposure. After crosslinking, initial pressurization, expansion, and preshrinking, the balloon will thereafter expand in a controlled manner to a reproducible diameter in response to a given inflation pressure, and thereby avoid overexpanding the stent (if
20 used in a stent delivery system) to an undesirably large diameter.

 In one embodiment, the balloon is formed from a low tensile set polymer such as a silicone-polyurethane copolymer. Preferably, the silicone-polyurethane is an ether urethane and more specifically an aliphatic ether urethane such as PURSIL AL 575A and PURSIL AL10, (Polymer Technology Group), and
25 ELAST-EON 3-70A, (Elastomedics), which are silicone polyether urethane copolymers, and more specifically, aliphatic ether urethane cosiloxanes. In an alternative embodiment, the low tensile set polymer is a diene polymer. A variety of suitable diene polymers can be used such as, but not limited to, an isoprene such as an AB and ABA poly(styrene-block-isoprene), a neoprene, an AB and ABA
30 poly(styrene-block-butadiene) such as styrene butadiene styrene (SBS) and styrene butadiene rubber (SBR), and 1,4-polybutadiene. Preferably, the diene polymer is an isoprene including isoprene copolymers and isoprene block copolymers such as

poly(styrene-block-isoprene). A presently preferred isoprene is a styrene-isoprene-styrene block copolymer, such as Kraton 1161K available from Kraton, Inc. However, a variety of suitable isoprenes can be used including HT 200 available from Apex Medical, Kraton R 310 available from Kraton, and isoprene (i.e., 2-methyl-1,3-butadiene) available from Dupont Elastomers. Neoprene grades useful in the disclosed subject matter include HT 501 available from Apex Medical, and neoprene (i.e., polychloroprene) available from Dupont Elastomers, including Neoprene G, W, T and A types available from Dupont Elastomers.

In accordance with another aspect of the disclosed subject matter, the outer surface of the balloon is modified. In this regard, the balloon surface can include a textured surface, roughened surface, voids, spines, channels, dimples, pores, or microcapsules or a combination thereof, as will be described below.

In accordance with the disclosed subject matter, the balloon does not include a stent or is free of a stent. However, a stent can be mounted onto the coated balloon. The stent will not detrimentally affect coating integrity or drug delivery. The type of stent that can be used includes, but is not limited to, bare metal stent, balloon expandable stent, self expanding stent, drug eluting stent, prohealing stent, and self-expanding vulnerable plaque implant. The balloon can be coated independently of the stent or in conjunction with the stent coating process. The stent coating can contain the same or different therapeutic agents from the balloon catheter or expandable member. However, the particular coating on the balloon catheter or expandable member preferably has distinct release kinetics from the therapeutic coating on the stent.

In one embodiment of the disclosed subject matter, the balloon is formed of a porous elastomeric material having at least one void formed in the wall of the balloon surface. For example, the entire cross section of the balloon can contain a plurality of voids. Alternatively, the plurality of void can be distributed along select lengths of the balloon outer surface. For example and not limitation, the plurality of voids can be distributed only along only the working section of the balloon. The voids define an open space within the outer surface of the balloon. Preferably, the therapeutic agent is dispersed within the space defined by the plurality of voids across the cross section of the balloon outer surface.

In operation, the therapeutic agent is released or is expelled from the pores upon inflation of the balloon. In this regard, the durometer of the polymeric material of the balloon surface and in particular the depression of the void is sufficiently flexible to allow for expulsion of the therapeutic agent and/or coating
5 contained within the plurality of voids upon inflation of the balloon. The expelled coating with therapeutic agent is released into the vessel lumen or into the tissue surrounding and contacting the inflated balloon.

In another embodiment, the balloon includes protrusions configured to contact or penetrate the arterial wall of a vessel upon inflation of the balloon. A
10 coating containing therapeutic agent is disposed on the protrusions and when inflated the coating and/or therapeutic agent coats the tissue of the arterial wall. Alternatively, the balloon can include two concentric balloons in a nesting configuration. The coating with therapeutic agent is disposed between the two concentric balloons. Thus, the space between the two concentric balloons; one being an interior balloon and the
15 other being an exterior balloon, acts as a reservoir. In this regard, the protrusions can include apertures for expulsion of the coating and/or therapeutic agent upon inflation of the interior and exterior concentric balloons. For example, as described in U.S. Patent No. 6,991,617 to Hektner, the disclosure of which is incorporated herein by reference thereto. In another embodiment, the balloon can include longitudinal
20 protrusions configured to form ridges on the balloon surface. As described in U.S. Patent No. 7,273,417 to Wang, the entire disclosure of which is incorporated herein by reference, the ridges can be formed of filaments spaced equidistantly apart around the circumference of the balloon. However, a larger or smaller number of ridges can alternatively be used. The longitudinal ridges can be fully or partially enveloped by
25 the polymeric material of the balloon.

In yet another embodiment of the disclosed subject matter, the balloon can include microcapsules on its outer surface. In this regard, the microcapsules are configured to encompass the coating and/or therapeutic agent. Upon inflation of the balloon the microcapsules located on the surface of the balloon contact the tissue of
30 the arterial wall. Alternatively, the microcapsules can be formed in the wall of the balloon surface. The coating and/or therapeutic agent can be released from the microcapsules by fracturing of the microcapsules and/or diffusion from the

microcapsule into the arterial wall. The microcapsules can be fabricated in accordance with the methods disclosed in U.S. Patent No. 5,102,340 to Dror or U.S. Patent No. 6,129,705 to Grantz and the patents referenced therein, each of which is incorporated herein by reference in its entirety.

5 In accordance with another aspect of the disclosed subject matter, if desired, a protective sheath can be utilized to protect the coating from being rubbed off of the balloon during the movement of the coated balloon through the body lumen. The sheath is preferably made from an elastic and resilient material which conforms to the shape of the balloon and in particular is capable of expanding upon inflation of
10 the balloon. The sheath preferably includes apertures along a length thereof. In operation, the inflation of the balloon causes the apertures of the sheath to widen for release of the coating and/or therapeutic agent to the tissue of the arterial wall. Preferably, the sheath has a thickness less than 10 mils. However, other thicknesses are possible.

15 In another embodiment, the sheath has at least one longitudinal line of weakness allowing the sheath to rupture upon inflation of the balloon and the release of the coating and/or therapeutic agent onto the tissue of the arterial wall of the vessel. Preferably, the sheath is formed from polymeric material known to be suitable for use in balloon catheters. Preferably, the sheath material is an elastomeric material which
20 will also spring back when it splits to expose more of the body lumen to the coating. The line of weakness could be provided by various techniques known in the art. However, one non-limiting examples include perforating the sheath material. In operation, the sheath is placed over the coated balloon while in the deflated state. When the coated balloon is inflated, the sheath is expanded to the extent that it
25 exceeds its elastic limit at the line of weakness and bursts to expose and therefore release the coating and/or therapeutic agent to the tissue of the arterial wall or vessel lumen. For example, see U.S. Patent No. 5,370,614 to Amundson, the entire disclosure of which is incorporated by reference.

 In accordance with another embodiment, an outer fibrous coating can
30 be electrospun or stretched onto the medical device or balloon catheter to prevent drug loss during delivery. During balloon inflation, the coating is stretched and allows for coating dissolution and release. The fiber diameters and material

properties can be fine tuned for optimal pore size and to release the particles containing the therapeutic agent. Fibrous coatings on expandable members are described in U.S. Patent Application Serial No. 12/237,998 to R. von Oepen and U.S. Patent Application Serial No. 12/238,026 to K. Ehrenreich, the disclosures of which
5 are incorporated by reference in their entirety.

It is to be noted that the term “a” entity or “an” entity refers to one or more of that entity. For example, a protein refers to one or more proteins or at least one protein. As such, the terms “a”, “an”, “one or more”, and “at least one” can be used interchangeably herein. The terms “comprising,” “including,” and “having”
10 can also be used interchangeably. In addition, the terms “amount” and “level” are also interchangeable and can be used to describe a concentration or a specific quantity. Furthermore, the term “selected from the group consisting of” refers to one or more members of the group in the list that follows, including mixtures (i.e. combinations) of two or more members.

The term “about” or “approximately” means within an acceptable error range for the particular value as determined by one of ordinary skill in the art, which will depend in part on how the value is measured or determined, i.e., the limitations of the measurement system. For example, “about” can mean within 3 or more than 3 standard deviations, per the practice in the art. Alternatively, “about”
20 can mean a range of up to +/-20%, preferably up to +/-10%, more preferably up to +/-5%, and more preferably still up to +/-1% of a given value. Alternatively, particularly with respect to biological systems or processes, the term can mean within an order of magnitude, preferably within 5-fold, and more preferably within 2-fold, of a value. With reference to pharmaceutical compositions, the term “about” refers to a range that
25 is acceptable for quality control standards of a product approved by regulatory authorities.

EXAMPLES

The present application is further described by means of the examples, presented below. The use of such examples is illustrative only and in no way limits
30 the scope and meaning of the disclosed subject matter or of any exemplified term.

EXAMPLE A

To simulate drug release from a drug coated balloon, a three step in-vitro release method was developed. This method consists of a sequential dip release in 37°C porcine serum for 1 min, inflation release in 37°C porcine serum for 1 min and extraction release in 50% acetonitrile solution designed to mimic the balloon release during delivery to the lesion, drug delivery on inflation and the remaining drug on the balloon respectively. The resulting zotarolimus concentrations in the porcine serum supernatant are measured by liquid chromatography mass spectrometry (LCMS) and drug from the extraction measured by high performance liquid chromatography (HPLC).

This in-vitro release method was used to evaluate the drug release from zotarolimus (Zot):poly(vinylpyrrolidone) (PVP):glycerol drug coated balloons as a function of drug:excipient:plasticizer ratio (D:E:P) and PVP K-value. For the combined dip release and inflation release that simulates coating dissolution rate and drug delivery from a drug coated balloon, it is shown in Figure 2 that a higher drug to excipient ratio such as D:E:P, 20:1:0.4 (w/w) resulted in a “soluble” coating dissolution rate with a dissolution time in the range of 1min to 1h releasing less than 5% of drug in 2 min. For lower D:E:P ratios and increasing amounts of plasticizer, the Zot:PVP:glycerol formulation demonstrated a “fast” dissolution rate, i.e. less than 1 min releasing up to 90% of drug in 2 min. For a lower molecular weight or PVP K-value such as PVP C-15, the coating dissolution rate and drug release during the dip release was further increased to 30% as compared to the PVP C-30 coating at the same 1:1:0.4, D:E:P ratio which demonstrated less than 5% dip release. The K-Values of C-15 and C-30 designate PVP K value for low endotoxin grade.

EXAMPLE B

Scratch tests of coatings on glass slides were used to qualitatively evaluate coating mechanical properties in terms of hardness and tackiness. Figures 3 through 6 are optical micrographs of drug delivery balloon coating formulations coated onto glass slides. To produce the optical micrographs, the coating formulations were pipetted onto glass slides, and then the slides were baked at 50°C for one hour. While observing under an optical microscope, the coatings were scratched with a steel mandrel to assess their hardness, brittleness, and resistance to

fracture in the dry state. Stickiness was assessed by placing another clean glass slide onto the coatings, pressing them together and noting the force, if any, required to pull them apart.

It was observed that increasing the drug to excipient ratio for zotarolimus:PVP:glycerol to 10:1:0.4 (Figure. 3 left panel) from 2:1:0.4 (Figure 3 right panel) resulted in increased hardness and reduced tackiness. Representative optical micrographs of these coatings after scratching are shown in Figure 3. Coating with the 10:1:0.4 ratio (left panel) is much harder than the 2:1:0.4 ratio (right panel), which was waxy.

10

EXAMPLE C

The qualitative mechanical properties of zotarolimus:PVP:tween 20 coatings were evaluated by scratch tests of coatings deposited on glass slides. It was shown with representative optical micrographs in Figure 4 (left panel) that zotarolimus:PVP:tween 20 at a ratio of 2:1:0.67 exhibited a soft and waxy coating. With zotarolimus:PVP:tween 80 at a ratio of 2:1:0.4 as shown in Figure 4 (right panel) a slightly brittle, weak and chunky coating resulted.

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EXAMPLE D

The qualitative mechanical properties of zotarolimus:PEG-PE coatings were evaluated by scratch tests of coatings deposited on glass slides. It was shown with representative optical micrographs in Figure 5 (left panel) that zotarolimus:PEG-PE, 2:1 exhibited a soft and waxy coating. With zotarolimus:PEG-PE, 1:1 as shown in Figure 5 (right panel) a soft, waxy and tacky coating resulted. This coating underwent visible flow after scratching. Zotarolimus:PEG-PE, 1:2 as shown in Figure 6 (left panel) exhibited a hard and waxy coating and was birefringence when observed under polarized light (right panel). This drug:excipient system exhibited eutectic behavior with the 1:1 ratio coating being the softest and compositions greater in either component being harder. Furthermore, at the lower drug:excipient ratio of 1:2 the birefringence indicates crystallization of the PEG-PE component.

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The disclosed subject matter can be embodied in other specific forms without departing from its spirit or essential characteristics. The described embodiments are to be considered in all respects only as illustrative and not

restrictive. Thus, it is intended that the disclosed subject matter include modifications and variations that are within the scope of the appended claims and their equivalents. All references recited herein are incorporated herein in their entirety by specific reference.

CLAIMS

WHAT IS CLAIMED IS:

1. A method of coating a medical device having an outer surface, the method comprising:
 - 5 selecting a cytostatic therapeutic agent;
 - selecting at least one excipient;
 - blending the cytostatic agent and excipient to define a coating; and
 - applying the coating to the outer surface of the medical device, wherein the cytostatic therapeutic agent and the at least one excipient have a weight ratio of about
 - 10 20:1 to about 1:20, and further wherein the coating provides increased efficiency of therapeutic transfer to a body lumen.
2. The method of claim 1, further including:
 - blending a plasticizer with the cytostatic therapeutic agent and the excipient.
3. The method of claim 2, wherein the excipient to plasticizer weight ratio is
- 15 from about 20:1 to about 1:20.
4. The method of claim 1, wherein the cytostatic therapeutic agent and the at least one excipient has a weight ratio of about 2:1 to about 1:2.
5. The method of claim 1, wherein the cytostatic therapeutic agent is zotarolimus.
- 20 6. The method of claim 1, wherein the excipient is poly(vinylpyrrolidone).
7. The method of claim 2, wherein the plasticizer is glycerol.
8. The method of claim 1, wherein the coating is applied to the outer surface of the medical device by direct fluid application.
9. The method of claim 1, further including applying heat to the coated medical
- 25 device to dry the coating.
10. The method of claim 1, wherein the medical device is a balloon catheter.

11. A method of coating a medical device having an outer surface, the method comprising:
- selecting a cytostatic therapeutic agent;
 - selecting at least one polymeric excipient;
 - 5 blending the cytostatic therapeutic agent and at least one polymeric excipient to define a coating; and
 - applying the coating to at least one length of the outer surface of the medical device, wherein the at least one polymeric excipient has a polydispersity index from about 1.05 to about 10 and provides increased efficiency of therapeutic transfer to a
 - 10 body lumen.
12. The method of claim 11, further including:
- blending a plasticizer with the cytostatic therapeutic agent and excipient. .
13. The method of claim 11, wherein cytostatic therapeutic agent is zotarolimus.
14. The method of claim 11, wherein the excipient is poly(vinylpyrrolidone).
- 15 15. The method of claim 12, wherein the plasticizer is glycerol.
16. The method of claim 11, wherein the medical device is a balloon catheter.
17. A method of coating a medical device having an outer surface, the method comprising:
- selecting a cytostatic therapeutic agent;
 - 20 selecting at least one excipient;
 - blending the cytostatic therapeutic agent and at least one excipient to define a coating;
 - applying the coating to at least one length of the outer surface of the medical device; and
 - 25 inserting or implanting the coated medical device in a body lumen, wherein the coating has a dissolution rate of about 10 seconds to about 1 hour and provides an increased efficiency of therapeutic transfer to a body lumen.
18. The method of claim 17, further including:

- blending a plasticizer with the cytostatic therapeutic agent and excipient. .
19. The method of claim 17, wherein cytostatic therapeutic agent is zotarolimus.
20. The method of claim 17, wherein the excipient is poly(vinylpyrrolidone).
21. The method of claim 18, wherein the plasticizer is glycerol.
- 5 22. The method of claim 17, wherein the therapeutic transfer to the body lumen occurs in less than 5 minutes.
23. The method of claim 17, wherein the therapeutic transfer to the body lumen occurs in 30 seconds or less.
24. The method of claim 17, wherein the medical device is a balloon catheter
- 10 25. A method of tuning the solubility of a coating for a medical device, the method comprising:
- selecting a first cytostatic therapeutic agent;
- selecting a first excipient;
- blending the first cytostatic agent and the first excipient to define a first
- 15 coating, the first coating having a first dissolution rate;
- selecting a second cytostatic therapeutic agent;
- selecting a second excipient;
- blending the second cytostatic agent and second excipient to define a second
- 20 coating, the second coating having a second dissolution rate; wherein the first and second dissolution rates are different; and
- applying the first and second coatings to at least one length of the outer surface of the medical device, the different dissolution rates defining a tunable solubility.
26. The method of claim 25, wherein the first excipient is a polymer having a
- 25 molecular weight of less than about 35 kDalton.
27. The method of claim 25, wherein the second excipient is a polymer having a molecular weight greater than about 100 kDalton.

28. The method of claim 25, wherein the first excipient is poly(vinylpyrrolidone) or poly(ethylene glycol).
29. The method of claim 25, wherein the second excipient is poly(vinylpyrrolidone) or poly(ethylene glycol).
- 5 30. The method of claim 25, wherein the medical device is a balloon catheter
31. A method of coating a medical device, the method comprising:
- selecting a therapeutic agent;
- selecting at least one excipient;
- modifying the excipient;
- 10 blending the therapeutic agent and the charged excipient to define a coating;
- and
- applying the modified excipient to an expandable member, wherein the coating provides desired delivery kinetics.
32. The method of claim 31, further including charging the excipient with a
- 15 positive charge.
33. The method of claim 31, further including charging the excipient with a negative charge.
34. The method of claim 31, wherein the excipient includes cyclic and aliphatic carbon chains, and further wherein the excipient is modified by adjusting cyclic chain
- 20 to aliphatic chain ratio.
35. The method of claim 34, wherein the adjusted ratio results in reduced elasticity.
36. The method of claim 34, wherein the modified excipient includes a therapeutic agent and further wherein the adjusted ratio results in reduced release rate of the
- 25 therapeutic agent.
37. The method of claim 31, wherein the excipient is poly(ester amide) polymer.

38. The method of claim 31, wherein the excipient is modified by grafting a low molecular weight polyethylene glycol molecule to the excipient, and further wherein modified excipient exhibits increased adhesion to a vessel wall.
39. The method of claim 31, wherein the modified excipient has increased
5 crystallinity.
40. The method of claim 39, wherein the excipient is poly(L-lactide-co-caprolactone) polyester, and the crystallinity of the excipient is modified by adjusting the content of L-lactide.
41. The method of claim 40, wherein the content of L-lactide is increased and the
10 excipient exhibits greater storage stability.
42. The method of claim 31, wherein the coating is applied to a balloon.

1/6

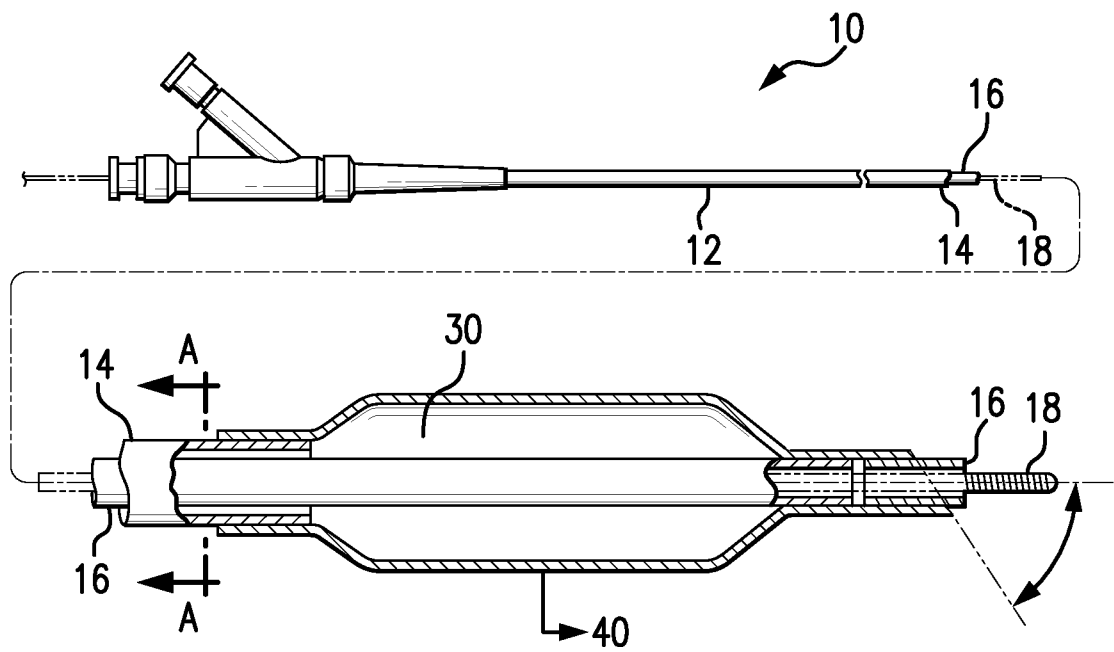


FIG. 1a

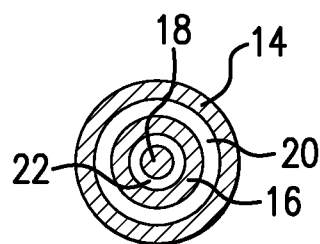


FIG. 1b

2/6

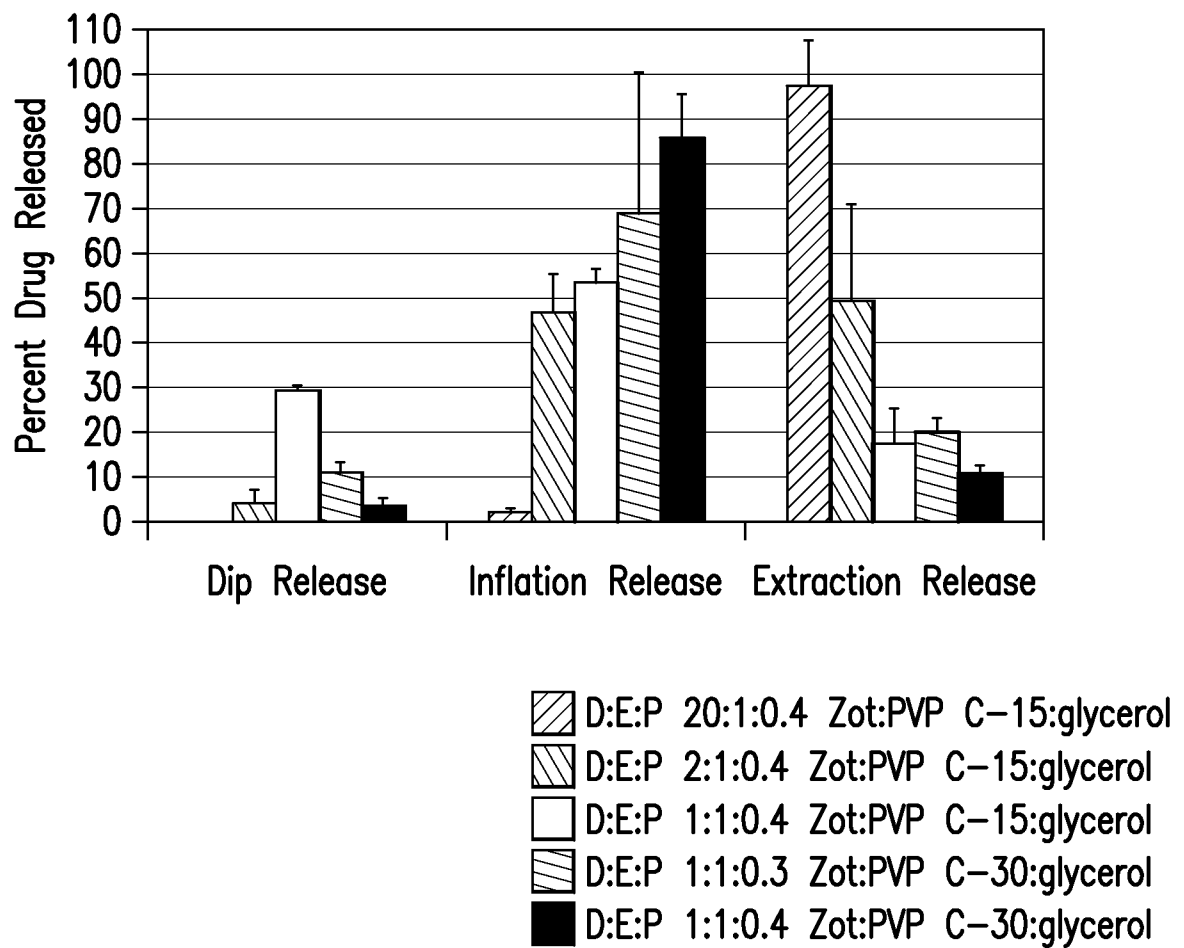


FIG. 2

3/6

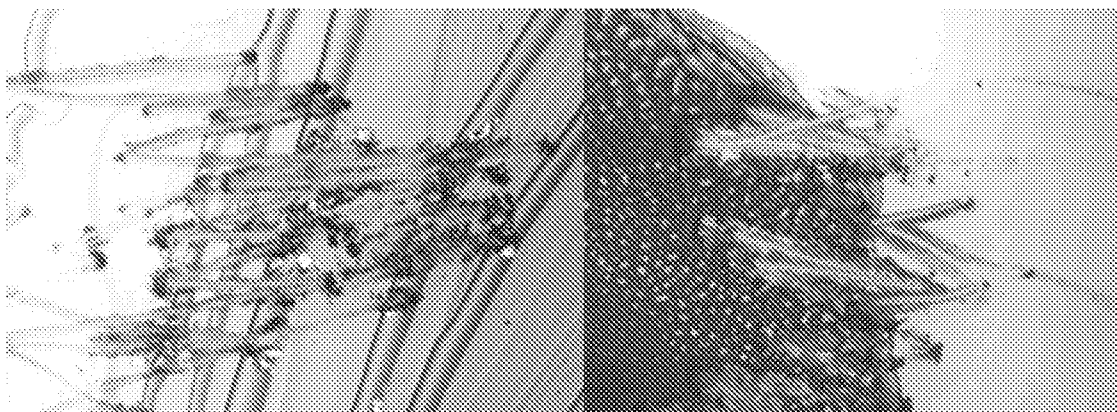


FIG. 3

4/6

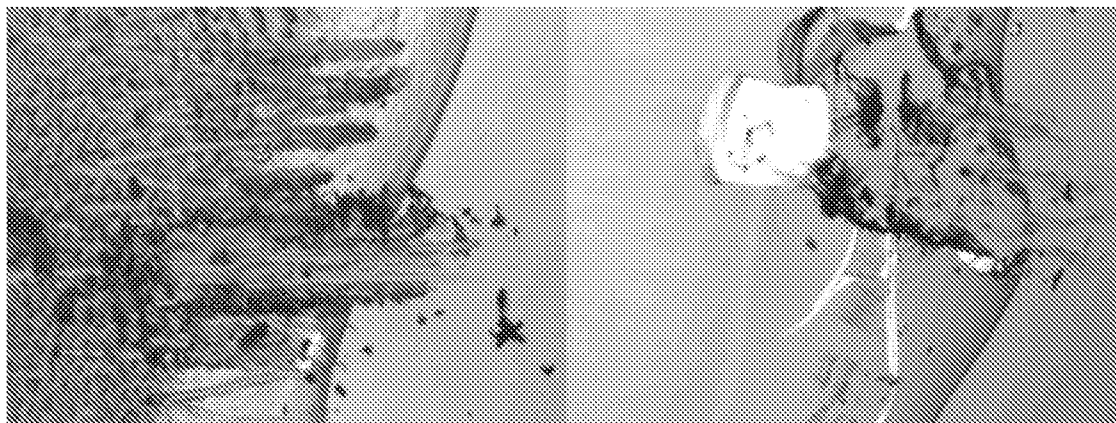


FIG. 4

5/6

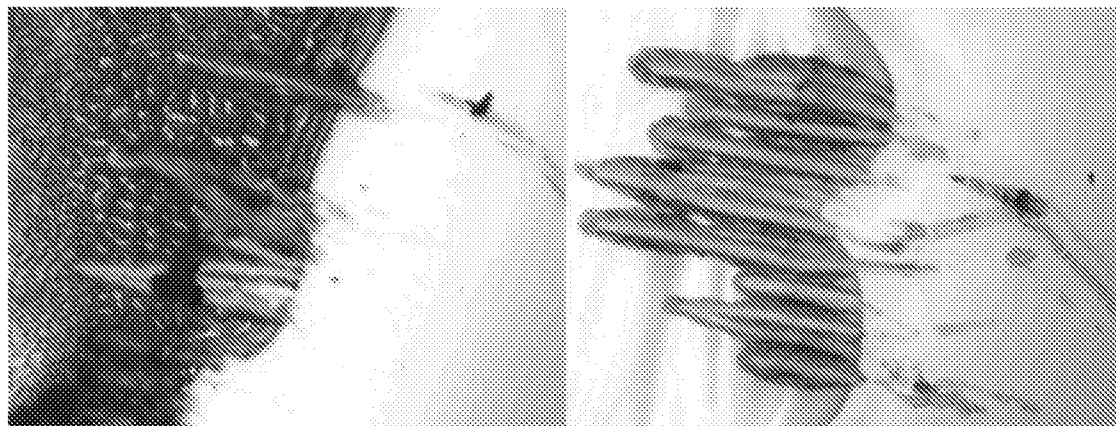


FIG. 5

6/6

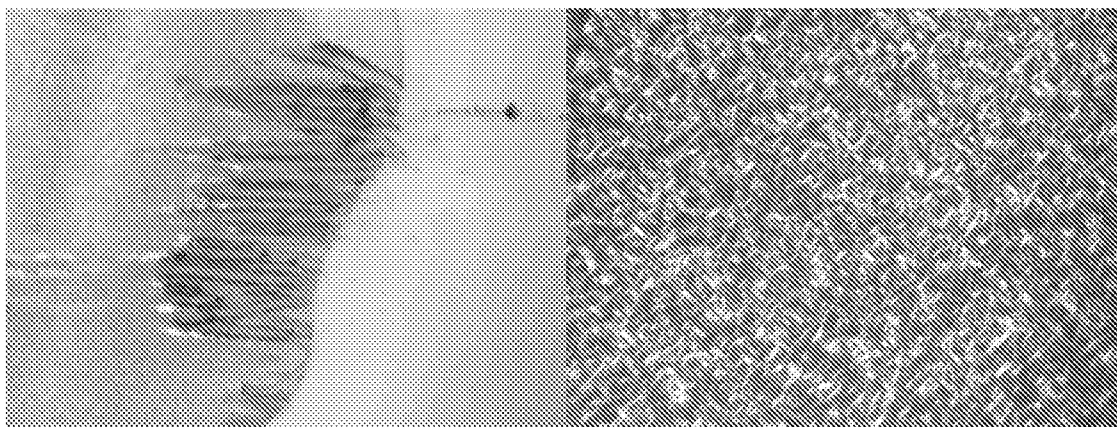


FIG. 6

INTERNATIONAL SEARCH REPORT

International application No
PCT/US2010/055806

A. CLASSIFICATION OF SUBJECT MATTER INV. A61L29/08 A61L29/16 C23C2/00 A61F2/84 ADD.		
According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) A61L C23C A61F		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the international search (name of data base and, where practical, search terms used) EPO-Internal, WPI Data		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 2009/285974 A1 (KERRIGAN CAMERON K [US] ET AL) 19 November 2009 (2009-11-19) claim 10 paragraphs [0016], [0036], [0038]	1-3, 5-22, 24, 31, 32, 37, 42
X	DE 10 2007 034991 A1 (BIOTRONIK VI PATENT AG [CH]) 29 January 2009 (2009-01-29) claims 1,3 <div style="text-align: center;">----- -/-</div>	1,8-11, 16
<div style="display: flex; justify-content: space-between;"> <input checked="" type="checkbox"/> Further documents are listed in the continuation of Box C. <input checked="" type="checkbox"/> See patent family annex. </div>		
* Special categories of cited documents : <div style="display: flex;"> <div style="flex: 1;"> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> </div> <div style="flex: 1;"> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"&" document member of the same patent family</p> </div> </div>		
Date of the actual completion of the international search <div style="text-align: center; font-size: 1.2em;">14 March 2011</div>		Date of mailing of the international search report <div style="text-align: center; font-size: 1.2em;">21/03/2011</div>
Name and mailing address of the ISA/ European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Fax: (+31-70) 340-3016		Authorized officer <div style="text-align: center; font-size: 1.2em;">Siebum, Bastiaan</div>

INTERNATIONAL SEARCH REPORT

International application No

PCT/US2010/055806

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

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X	----- WO 2006/024492 A2 (INTERSTITIAL THERAPEUTICS [CH]; POPOWSKI YOURI [CH]) 9 March 2006 (2006-03-09)	25, 28-31, 39,42
Y	claims 10, 46,47,53 page 29, line 30 - page 30, line 14	23,26,27
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