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(54) Titre : PROCEDE ET KIT DESTINES A EXPRIMER UNE PROTEINE SOUS REGULATION DE L'EXPRESSION A PARTIR D'UNE SEQUENCE REPETEE FORMEE PAR UNE AMPLIFICATION DE GENES ET UN TRANSFORMANT

(54) Title: METHOD AND KIT FOR EXPRESSING PROTEIN UNDER REGULATION OF THE EXPRESSION FROM REPEATED SEQUENCE FORMED BY GENE AMPLIFICATION AND TRANSFORMANT

(57) **Abrégé/Abstract:**

It is intended to provide a method of releasing the transcriptional regulation caused by a repeated sequence in a gene, a kit therefor and so on to thereby establish a system capable of producing a protein in a large amount. The above-described method can be achieved by any one or more of the following methods: (a) in the amplification of a gene encoding a target protein, co-amplifying a polynucleotide of 10kbp or more such as a λ -phage DNA or an insulator sequence; (b) selecting by culturing cells having undergone gene amplification in media containing a drug with a gradual increase in concentration; (c) elevating the promoter activity of inducing the expression of a gene encoding a target protein; (d) excising an amplified gene region from a chromosome with the use of Cre-LoxP System; (e) treating cells having undergone gene amplification with 5-aza-2'-deoxycytidine to thereby lower the methylation degree of DNA; and (f) selecting the mammalian cells having undergone gene amplification on double minute chromosomes.



ABSTRACT

It is intended to provide a method of releasing the transcriptional regulation caused by a repeated sequence in a gene, a kit therefor and so on to thereby establish a system capable of producing a protein in a large amount. The above-described method can be achieved by any one or more of the following methods: (a) in the amplification of a gene encoding a target protein, co-amplifying a polynucleotide of 10kbp or more such as a λ -phage DNA or an insulator sequence; (b) selecting by culturing cells having undergone gene amplification in media containing a drug with a gradual increase in concentration; (c) elevating the promoter activity of inducing the expression of a gene encoding a target protein; (d) excising an amplified gene region from a chromosome with the use of Cre-LoxP System; (e) treating cells having undergone gene amplification with 5-aza-2'-deoxycytidine to thereby lower the methylation degree of DNA; and (f) selecting the mammalian cells having undergone gene amplification on double minute chromosomes.

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DESCRIPTION

METHOD AND KIT FOR EXPRESSING PROTEIN UNDER
REGULATION OF THE EXPRESSION FROM REPEATED SEQUENCE
5 FORMED BY GENE AMPLIFICATION, AND TRANSFORMANT

TECHNICAL FIELD

The present invention relates to a method of expressing a protein,
whose expression is repressed, from a repeated sequence formed in
10 mammalian cells in which gene amplification is induced.

BACKGROUND ART

The inventor of the present invention has found that it becomes
possible to copy a gene (target gene), which encodes a target protein, in an
15 intercellular copy number increased to approximately 100,000 copies simply
by plasmid conduction into human-derived cancer cells (COLO 320 colon
cancer cell line, and HeLa cell line) by lipofection, the plasmid (hereinafter,
"IR/MAR plasmid") having mammalian copying initiation region (IR;
initiation region) and a matrix attachment region (MAR; matrix attachment
20 region), and selecting by utilizing a gene tolerant to a chemical (Blasticidine
or Neomycine), and that the mass amplification of the target gene can be

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attained regardless of whether the target gene has a gene structure identical to that of the IR/MAR plasmid (i.e., the target gene has a cis gene structure) or the target gene has a gene structure different from that of the IR/MAR plasmid (i.e., the target gene has a trans gene structure) (see Patent Document 1, Patent Document 2, Non-Patent Document 1, and Non-Patent Document 2).

A cell line in which an IR/MAR plasmid and a target gene were transfected, was analyzed quantitatively as to a transcription amount of mRNA from the target gene. This analysis found that the transcription amount of mRNA was not increased while the copy number of the target gene was increased. It was deduced that the transcription was repressed due to a repeated sequence produced by the mass amplification of a region including the target gene.

One known method of releasing the transcription repression caused by the repeated gene sequence is, for example, treating the cells with histone acetylating enzyme inhibitor such as trichostatin A (see Non-Patent Document 3).

[Patent Document 1]

Japanese Unexamined Patent Application Publication, *Tokukai*, No. 2003-245083 (published on September 2, 2003)

[Patent Document 2]

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Japanese Unexamined Patent Application Publication, *Tokukai*, No.
2004-337066 (published on December 2, 2004)

[Non-Patent Document 1]

Noriaki Shimizu, et al. (2001) Plasmids with a Mammalian
5 Replication Origin and a Matrix Attachment Region Initiate the Event
Similar to Gene Amplification. *Cancer Research* vol.61, no.19, p6987-6990.

[Non-Patent Document 2]

Noriaki Shimizu, et al (2003) Amplification of plasmids containing a
mammalian replication initiation region is mediated by controllable conflict
10 between replication and transcription. *Cancer Research*, vol.63, no.17,
p5281-5290.

[Non-Patent Document 3]

McBurney, M. W. et al, *Exp Cell Res* (2002), vol 274, p1-8

15 Therefore, there is such a problem in that even if a polynucleotide
containing the amplified target gene is amplified in order to mass-produce a
protein via the amplification of the target gene, the transcription of the target
gene is repressed once the repeated sequence is created thereby failing to
express the protein of the final target.

20 Sole application of the method of treating the cells with histone
acetylating enzyme inhibitor such as trichostatin A as disclosed in Non-

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Patent Document 3 could not sufficiently release the transcription repression caused by the repeated sequence of the gene amplified to several thousand to approximately 10 thousand copies.

An object of the present invention is to provide a method and kit etc.
5 for releasing the transcription repression caused by the repeated sequence of the gene, and to establish a system for mass production of useful protein by gene amplification.

DISCLOSURE OF INVENTION

10 In order to attain the object, the inventor of the present invention diligently worked to find a method capable of amplifying a target gene by using the IR/MAR plasmid, and expressing a protein without transcription repression, even in case where repeated sequence occurs. The inventor accomplished the present invention based on the result of the diligent work.

15 In order to attain the object, a method according to the present invention is a method of expressing a protein from a repeated sequence formed in mammalian cells in which gene amplification is induced, the protein having been under expression repression, the method including:
transfecting a first polynucleotide and a second polynucleotide
20 simultaneously into the mammalian cells, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that

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function in eukaryotic cells, and the second polynucleotide encodes a protein to be expressed, the method further comprising: transfecting at least one of a third polynucleotide and a fourth polynucleotide into the mammalian cells in transfecting the first polynucleotide and the second polynucleotide into the
5 mammalian cells, where the third polynucleotide has a length of 10kbp or more, and the fourth polynucleotide includes an insulator sequence.

Moreover, in order to attain the object, a method according to the present invention is a method of expressing a protein from a repeated sequence formed in mammalian cells in which gene amplification is induced,
10 the protein having been under expression repression, the method including: transfecting a first polynucleotide and a fifth polynucleotide simultaneously in the mammalian cells, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the fifth polynucleotide includes a second polynucleotide and a
15 chemical tolerant gene, the second polynucleotide encoding a protein to be expressed, the method further comprising: culturing the mammalian cells sequentially in a medium of increasing concentrations of a chemical.

Moreover, in order to attain the object, a method according to the present invention is a method of expressing a protein from a repeated
20 sequence formed in mammalian cells in which gene amplification is induced, the protein having been under expression repression, the method including:

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transfecting a first polynucleotide and a sixth polynucleotide simultaneously
in the mammalian cells where the first polynucleotide includes an origin of
replication and a nucleic matrix attachment region that function in eukaryotic
cells, and the sixth polynucleotide includes a promoter region and a second
5 polynucleotide controllably linked with each other, the second polynucleotide
encoding a protein to be expressed, the method further comprising:
transfecting into the mammalian cells a seventh polynucleotide, which
encodes a transcription activation factor of the promoter, and expressing the
transcription activation factor.

10 Moreover, in order to attain the object, a method according to the
present invention is a method of expressing a protein from a repeated
sequence formed in mammalian cells in which gene amplification is induced,
the protein having been under expression repression, the method including:
transfecting a first polynucleotide and an eighth polynucleotide
15 simultaneously in the mammalian cells, where the first polynucleotide
includes an origin of replication and a nucleic matrix attachment region that
function in eukaryotic cells, and the eighth polynucleotide includes LoxP
gene and a second polynucleotide encoding a protein to be expressed, the
method further comprising: transfecting into the mammalian cells a ninth
20 polynucleotide, which includes Cre Recombinase gene, and expressing the
Cre Recombinase gene.

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Moreover, in order to attain the object, a method according to the present invention is a method of expressing a protein from a repeated sequence formed in mammalian cells in which gene amplification is induced, the protein having been under expression repression, the method including:

5 transfecting a first polynucleotide and a second polynucleotide simultaneously into the mammalian cells, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the second polynucleotide encodes a protein to be expressed, the method further comprising: treating the mammalian cells

10 with 5-aza-2'-deoxycytidine.

Moreover, in order to attain the object, a method according to the present invention is a method of expressing a protein from a repeated sequence formed in mammalian cells in which gene amplification is induced, the protein having been under expression repression, the method comprising:

15 treating the mammalian cells with 5-aza-2'-deoxycytidine.

Moreover, in order to attain the object, a method according to the present invention is a method of expressing a protein from a repeated sequence formed in mammalian cells in which gene amplification is induced, the protein having been under expression repression, the method including:

20 transfecting a first polynucleotide and a second polynucleotide simultaneously into the mammalian cells, where the first polynucleotide

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includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the second polynucleotide encodes a protein to be expressed, the method further comprising: selecting mammalian cells that the gene amplification occurs on a double minute chromosome.

5 Moreover, in order to attain the object, a method according to the present invention is a method of expressing a protein from a repeated sequence formed in mammalian cells in which gene amplification is induced, the protein having been under expression repression, the method comprising: selecting mammalian cells that the gene amplification occurs on a double
10 minute chromosome.

 Moreover, in order to attain the object, a method according to the present invention is a method of expressing a protein from a repeated sequence formed in mammalian cells in which gene amplification is induced, the protein having been under expression repression, the method comprising:
15 transfecting a first polynucleotide and a sixth polynucleotide simultaneously in the mammalian cells, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the sixth polynucleotide includes a promoter region and a second polynucleotide controllably linked with each other, the second polynucleotide
20 encoding a protein to be expressed, the method further comprising: transfecting a seventh polynucleotide into the mammalian cells

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simultaneously in transfecting the first and sixth polynucleotides into the mammalian cells, the seventh polynucleotide encoding a transcription activation factor of the promoter.

Moreover, in order to attain the object, a method according to the present invention is a method of expressing a protein from a repeated sequence formed in mammalian cells in which gene amplification is induced, the protein having been under expression repression, the method including: transfecting a first polynucleotide and a second polynucleotide simultaneously into the mammalian cells, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the second polynucleotide encodes a protein to be expressed, the method further comprising: selecting mammalian cells that the gene amplification occurs on a double minute chromosome; and treating the mammalian cells with 5-aza-2'-deoxycytidine.

Moreover, in order to attain the object, a method according to the present invention is a method of expressing a protein from a repeated sequence formed in mammalian cells in which gene amplification is induced, the protein having been under expression repression, the method including: transfecting a first polynucleotide and a sixth polynucleotide simultaneously in the mammalian cells where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic

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cells, and the sixth polynucleotide includes a promoter region and a second polynucleotide controllably linked with each other, the second polynucleotide encoding a protein to be expressed, the method further comprising: transfecting into the mammalian cells a seventh polynucleotide simultaneously in transfecting the first polynucleotide and the second polynucleotide into the mammalian cells, the seventh polynucleotide encoding a transcription activation factor of the promoter; and treating the mammalian cells with 5-aza-2'-deoxycytidine.

The methods according to the present invention may be arranged such that the origin of replication is derived from an origin of replication of c-myc locus, dihydrofolate reductase locus, or β -globin locus.

The methods according to the present invention may be arranged such that the nucleic matrix attachment region is derived from a nucleic matrix attachment region of Ig κ locus, SV40 initial region, or dihydrofolate reductase locus.

On the other hand, a kit according to the present invention is a kit for expressing a protein from a repeated sequence formed in mammalian cells in which gene amplification is induced, the protein having been under expression repression, the kit comprising: a first polynucleotide including an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells; and at least one of a third polynucleotide and a fourth

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polynucleotide where the third polynucleotide has a length of 10kbp or more,
and the fourth polynucleotide includes an insulator sequence.

Moreover, a kit according to the present invention is a kit for
expressing a protein from a repeated sequence formed in mammalian cells in
5 which gene amplification is induced, the protein having been under
expression repression, the kit comprising: a first polynucleotide including an
origin of replication and a nucleic matrix attachment region that function in
eukaryotic cells; a polynucleotide including a promoter region; and a seventh
polynucleotide encoding a transcription activation factor of the promoter.

10 Moreover, a kit according to the present invention is a kit for
expressing a protein from a repeated sequence formed in mammalian cells in
which gene amplification is induced, the protein having been under
expression repression, the kit comprising: a first polynucleotide including an
origin of replication and a nucleic matrix attachment region that function in
15 eukaryotic cells; a polynucleotide including LoxP gene; and a ninth
polynucleotide including Cre Recombinase gene.

The kits according to the present invention preferably further includes
5-aza-2'-deoxycytidine.

Moreover, the kits according to the present invention may be arranged
20 such that the origin of replication is derived from an origin of replication of
c-myc locus, dihydrofolate reductase locus, or β -globin locus.

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Moreover, the kits according to the present invention may be arranged such that the nucleic matrix attachment region is derived from a nucleic matrix attachment region of Igk locus, SV40 initial region, or dihydrofolate reductase locus.

5 Meanwhile, a transformant according to the present invention is a transformant prepared by inserting into mammalian cells: a first polynucleotide including an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells; a second polynucleotide encoding a protein to be expressed; and at least one of a third polynucleotide
10 and a fourth polynucleotide, where the third polynucleotide has a length of 10kbp or more, and the fourth polynucleotide includes an insulator sequence.

Moreover, a transformant according to the present invention is a transformant prepared by inserting into mammalian cells: a first polynucleotide including an origin of replication and a nucleic matrix
15 attachment region that function in eukaryotic cells; a sixth polynucleotide includes a promoter region and a second polynucleotide controllably linked with each other, the second polynucleotide encoding a protein to be expressed; and a seventh polynucleotide encoding a transcription activation factor of the promoter.

20 Moreover, a transformant according to the present invention is a transformant prepared by transfecting into mammalian cells: a first

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polynucleotide including an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells; an eighth polynucleotide including LoxP gene and a second polynucleotide encoding a protein to be expressed; and a ninth polynucleotide including Cre Recombinase gene, and
5 expressing the Cre Recombinase gene.

Moreover, the transformants according to the present invention may be arranged such that the origin of replication is derived from an origin of replication of c-myc locus, dihydrofolate reductase locus, or β -globin locus.

Moreover, the transformants according to the present invention may
10 be arranged such that the nucleic matrix attachment region is derived from a nucleic matrix attachment region of Igk locus, SV40 initial region, or dihydrofolate reductase locus.

Furthermore, the transformants according to the present invention may be arranged such that the mammalian cells are cells selected from the group
15 consisting of COLO 320DM cells, COLO 320HSR cells, Hela cells, and CHO cells.

According to the present invention, a protein that has been under expression repression can be expressed from a repeated sequence formed in mammalian cells in which gene amplification is induced. Therefore, the
20 present invention makes it possible to establish a mass production system for a useful protein by employing a gene amplification technique.

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For a fuller understanding of the nature and advantages of the invention, reference should be made to the ensuing detailed description taken in conjunction with the accompanying drawings.

5 BRIEF DESCRIPTION OF DRAWINGS

Fig. 1(a) is a view illustrating a fluorescence intensity of a control of Example 1, where only pSFVdhfr (GAP-GFP), but not λ -phage DNA, was transfected into cells.

Fig. 1(b) is a view illustrating a fluorescence intensity of a case of
10 Example 1, where a mixture of pSFVdhfr (GAP-GFP) and λ -phage DNA was transfected into cells (cotransfection).

Fig. 2(a) is a view illustrating a fluorescence intensity of a control of Example 2, where only pSFVdhfr/d2EGFP was transfected into cells.

Fig. 2(b) is a view illustrating a fluorescence intensity of a case of
15 Example 2, where only pSFVdhfr/d2EGFP was transfected into cells, and the cells were selected with 80 μ g/ml of Blasticidine.

Fig. 2(c) is a view illustrating a fluorescence intensity of a case of Example 2, where a mixture of pSFVdhfr/d2EGFP and a plasmid DNA having an Aves-derived HS4 insulator sequence was transfected into cells.

20 Fig. 2(d) is a view illustrating a fluorescence intensity of a case of Example 2, where a mixture of pSFVdhfr/d2EGFP and a plasmid DNA

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having the Aves-derived HS4 insulator sequence was transfected into cells,
and the cells were selected with 80µg/ml of Blastcidine.

Fig. 3(a) is a view illustrating a fluorescence intensity of cells selected
with 5µg/ml of Blastcidine in Example 3.

5 Fig. 3(b) is a view illustrating a fluorescence intensity of cells selected
with 10µg/ml of Blastcidine in Example 3.

Fig. 3(c) is a view illustrating a fluorescence intensity of cells selected
with 40µg/ml of Blastcidine in Example 3.

10 Fig. 3(d) is a view illustrating a fluorescence intensity of cells selected
with 160µg/ml of Blastcidine in Example 3.

Fig. 3(e) is a view illustrating a fluorescence intensity of cells selected
with 320µg/ml of Blastcidine in Example 3.

Fig. 4(a) is a phase contrast microscopic image (×200) of cells in
which normal expression of Tet-ON gene occurred in Example 4.

15 Fig. 4(b) is a phase contrast microscopic image (×200) of cells in
which high expression of Tet-ON gene occurred in Example 4.

Fig. 4(c) is a fluorescence microscopic image (×200) of cells in which
normal expression of Tet-ON gene occurred in Example 4.

20 Fig. 4(d) is a fluorescence microscopic image (×200) of cells in which
high expression of Tet-ON gene occurred in Example 4.

Fig. 5(a) is a view illustrating a fluorescence intensity of cells in

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which pSFVdhfr/d2EGFP was transfected and normal expression of Tet-ON protein occurred in Example 4.

Fig. 5(b) is a view illustrating a fluorescence intensity of cells in which pSFVdhfr/d2EGFP was transfected and high expression of Tet-ON
5 protein occurred in Example 4.

Fig. 5(c) is a view illustrating a fluorescence intensity of cells in which the Aves-derived HS4 insulator sequence and pSFVdhfr/d2EGFP were coamplified and normal expression of Tet-ON protein occurred in Example 4.

Fig. 5(d) is a view illustrating a fluorescence intensity of cells in
10 which the Aves-derived HS4 insulator sequence and pSFVdhfr/d2EGFP were coamplified and high expression of Tet-ON protein occurred in Example 4.

Fig. 6(a) is a phase contrast microscopic image ($\times 200$) of cells before transfection of Cre Recombinase gene in Example 5.

Fig. 6(b) is a phase contrast microscopic image ($\times 200$) of cells after
15 the transfection of the Cre Recombinase gene in Example 5.

Fig. 6(c) is a fluorescence microscopic image ($\times 200$) of cells before the transfection of the Cre Recombinase gene in Example 5.

Fig. 6(d) is a fluorescence microscopic image ($\times 200$) of cells after the transfection of the Cre Recombinase gene in Example 5.

20 Fig. 7(a) is a phase contrast microscopic image ($\times 200$) of cells before treatment with 5-aza-2'-deoxycytidine (5-aza) in Example 6.

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Fig. 7(b) is a phase contrast microscopic image ($\times 200$) of cells after treatment with $1\mu\text{M}$ of 5-aza in Example 6.

Fig. 7(c) is a fluorescence microscopic image ($\times 200$) of cells before treatment with 5-aza in Example 6.

5 Fig. 7(d) is a fluorescence microscopic image ($\times 200$) of cells after treatment with $1\mu\text{M}$ of 5-aza in Example 6.

Fig. 8(a) is a view illustrating a fluorescence intensity of cells before treatment with 5-aza-2'-deoxycytidine (5-aza) in Example 6.

Fig. 8(b) is a view illustrating a fluorescence intensity of cells after
10 treatment with 20nM of Trichostatin A (TSA) in Example 6.

Fig. 8(c) is a view illustrating a fluorescence intensity of cells after treatment with $1\mu\text{M}$ of 5-aza in Example 6.

Fig. 8(d) is a view illustrating a fluorescence intensity of cells after treatment with $1\mu\text{M}$ of 5-aza and 20nM of TSA in Example 6.

15 Fig. 8(e) is a view illustrating a fluorescence intensity of cells after treatment with $3\mu\text{M}$ of 5-aza in Example 6.

Fig. 8(f) is a view illustrating a fluorescence intensity of cells after treatment with $3\mu\text{M}$ of 5-aza and 20nM of TSA in Example 6.

Fig. 9 is a view illustrating the result of analysis of a level of
20 expression of d2EGFP in a polyclonal population of Tet-ON cells by using a cell sorter, where pSFVdhfr/d2EGFP was transfected in the Tet-ON cells and

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the expression of d2EGFP was induced with Doxycycline in Example 7.

Fig. 10(a) is a fluorescence microscopic image illustrating the result of detection of a base sequence of pSFVdhfr/d2EGFP plasmid in clone 4 by FISH method, where clone 4 was selected from among the polyclonal population of Tet-ON cells in which pSFVdhfr/d2EGFP was transfected in Example 7.

Fig. 10(b) is a fluorescence microscopic image illustrating the result of detection of a base sequence of pSFVdhfr/d2EGFP plasmid in clone 5 by FISH method, where clone 5 was selected from among the polyclonal population of Tet-ON cells in which pSFVdhfr/d2EGFP was transfected in Example 7.

Fig. 10(c) is a fluorescence microscopic image illustrating the result of detection of a base sequence of pSFVdhfr/d2EGFP plasmid in clone 6 by FISH method, where clone 6 was selected from among the polyclonal population of Tet-ON cells in which pSFVdhfr/d2EGFP was transfected in Example 7.

Fig. 10(d) is a fluorescence microscopic image illustrating the result of detection of a base sequence of pSFVdhfr/d2EGFP plasmid in clone 9 by FISH method, where clone 9 was selected from among the polyclonal population of Tet-ON cells in which pSFVdhfr/d2EGFP was transfected in Example 7.

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Fig. 11(a) is a view illustrating the result of analysis on the level of the expression of d2EGFP in clone 4 after the expression of d2EGFP was induced with Doxycycline and one-week incubation without adding 5-aza in a medium where clone 4 was obtained in Example 7 and the analysis was
5 carried out by using a cell sorter.

Fig. 11(b) is a view illustrating the result of analysis on the level of the expression of d2EGFP in clone 4 after the expression of d2EGFP was induced with Doxycycline and one-week incubation with 5-aza (final concentration of 1 μ M) added in a medium where clone 4 was obtained in
10 Example 7 and the analysis was carried out by using a cell sorter.

Fig. 11(c) is a view illustrating the result of analysis on the level of the expression of d2EGFP in clone 4 after the expression of d2EGFP was induced with Doxycycline and one-week incubation with 5-aza (final concentration of 3 μ M) added in a medium where clone 4 was obtained in
15 Example 7 and the analysis was carried out by using a cell sorter.

Fig. 12(a) is a view illustrating the result of analysis on the level of the expression of d2EGFP in clone 5 after the expression of d2EGFP was induced with Doxycycline and one-week incubation without adding 5-aza in a medium where clone 5 was obtained in Example 7 and the analysis was
20 carried out by using a cell sorter.

Fig. 12(b) is a view illustrating the result of analysis on the level of

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the expression of d2EGFP in clone 5 after the expression of d2EGFP was induced with Doxycycline and one-week incubation with 5-aza (final concentration of 1 μ M) added in a medium where clone 5 was obtained in Example 7 and the analysis was carried out by using a cell sorter.

5 Fig. 12(c) is a view illustrating the result of analysis on the level of the expression of d2EGFP in clone 5 after the expression of d2EGFP was induced with Doxycycline and one-week incubation with 5-aza (final concentration of 3 μ M) added in a medium where clone 5 was obtained in Example 7 and the analysis was carried out by using a cell sorter.

10 Fig. 13(a) is a view illustrating the result of analysis on the level of the expression of d2EGFP in clone 6 after the expression of d2EGFP was induced with Doxycycline and one-week incubation without adding 5-aza in a medium where clone 6 was obtained in Example 7 and the analysis was carried out by using a cell sorter.

15 Fig. 13(b) is a view illustrating the result of analysis on the level of the expression of d2EGFP in clone 6 after the expression of d2EGFP was induced with Doxycycline and one-week incubation with 5-aza (final concentration of 1 μ M) added in a medium where clone 6 was obtained in Example 7 and the analysis was carried out by using a cell sorter.

20 Fig. 13(c) is a view illustrating the result of analysis on the level of the expression of d2EGFP in clone 6 after the expression of d2EGFP was

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induced with Doxycycline and one-week incubation with 5-aza (final concentration of 3 μ M) added in a medium where clone 6 was obtained in Example 7 and the analysis was carried out by using a cell sorter.

Fig. 14(a) is a view illustrating the result of analysis on the level of
5 the expression of d2EGFP in clone 9 after the expression of d2EGFP was induced with Doxycycline and one-week incubation without adding 5-aza in a medium where clone 9 was obtained in Example 7 and the analysis was carried out by using a cell sorter.

Fig. 14(b) is a view illustrating the result of analysis on the level of
10 the expression of d2EGFP in clone 9 after the expression of d2EGFP was induced with Doxycycline and one-week incubation with 5-aza (final concentration of 1 μ M) added in a medium where clone 9 was obtained in Example 7 and the analysis was carried out by using a cell sorter.

Fig. 14(c) is a view illustrating the result of analysis on the level of
15 the expression of d2EGFP in clone 9 after the expression of d2EGFP was induced with Doxycycline and one-week incubation with 5-aza (final concentration of 3 μ M) added in a medium where clone 9 was obtained in Example 7 and the analysis was carried out by using a cell sorter.

Figs. 15 are phase contrast microscopic images and fluorescence
20 microscopic images of a polyclonal population of transformed cells before and after induction of the expression of d2EGFP with Doxycycline, where the

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transformed cells were obtained by transfecting pSFVdhfr/d2EGFP and pTet-ON plasmid simultaneously in human colon cancer COLO 320DM cells in Example 8. Fig. 15(a) is a phase contrast microscopic image (×200) before the addition of Doxycycline. Fig. 15(b) is a fluorescence microscopic image (×200) before the addition of Doxycycline. Fig. 15(c) is a phase contrast
5 microscopic image (×200) after the addition of Doxycycline. Fig. 15(d) is a fluorescence microscopic image (×200) after the addition of Doxycycline.

Fig. 16(a) is a view illustrating the result of analysis on the level of the expression of d2EGFP in the polyclonal population of transformed cells before induction of the expression of d2EGFP with Doxycycline, where the
10 transformed cells were obtained by transfecting pSFVdhfr/d2EGFP and pTet-ON plasmid simultaneously in the human colon cancer COLO 320DM cells and the analysis was carried out by using the cell sorter in Example 8.

Fig. 16(b) is a view illustrating the result of analysis on the level of the expression of d2EGFP in the polyclonal population of transformed cells after 1-hour induction of the expression of d2EGFP with Doxycycline, where the transformed cells were obtained by transfecting pSFVdhfr/d2EGFP and pTet-ON plasmid simultaneously in the human colon cancer COLO 320DM cells and the analysis was carried out by using the cell sorter in Example 8.

20 Fig. 16(c) is a view illustrating the result of analysis on the level of the expression of d2EGFP in the polyclonal population of transformed cells

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after 3-hour induction of the expression of d2EGFP with Doxycycline, where the transformed cells were obtained by transfecting pSFVdhfr/d2EGFP and pTet-ON plasmid simultaneously in the human colon cancer COLO 320DM cells and the analysis was carried out by using the cell sorter in Example 8.

5 Fig. 16(d) is a view illustrating the result of analysis on the level of the expression of d2EGFP in the polyclonal population of transformed cells after 6-hour induction of the expression of d2EGFP with Doxycycline, where the transformed cells were obtained by transfecting pSFVdhfr/d2EGFP and pTet-ON plasmid simultaneously in the human colon cancer COLO 320DM
10 cells and the analysis was carried out by using the cell sorter in Example 8.

Fig. 16(e) is a view illustrating the result of analysis on the level of the expression of d2EGFP in the polyclonal population of transformed cells after 15-hour induction of the expression of d2EGFP with Doxycycline, where the transformed cells were obtained by transfecting pSFVdhfr/d2EGFP
15 and pTet-ON plasmid simultaneously in the human colon cancer COLO 320DM cells and the analysis was carried out by using the cell sorter in Example 8.

Fig. 16(f) is a view illustrating the result of analysis on the level of the expression of d2EGFP in the polyclonal population of transformed cells
20 after 24-hour induction of the expression of d2EGFP with Doxycycline, where the transformed cells were obtained by transfecting pSFVdhfr/d2EGFP

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and pTet-ON plasmid simultaneously in the human colon cancer COLO 320DM cells and the analysis was carried out by using the cell sorter in Example 8.

Fig. 16(g) is a view illustrating the result of analysis on the level of
5 the expression of d2EGFP in the polyclonal population of transformed cells after 48-hour induction of the expression of d2EGFP with Doxycycline, where the transformed cells were obtained by transfecting pSFVdhfr/d2EGFP and pTet-ON plasmid simultaneously in the human colon cancer COLO 320DM cells and the analysis was carried out by using the cell sorter in
10 Example 8.

Fig. 16(h) is a view illustrating the result of analysis on the level of the expression of d2EGFP in the polyclonal population of transformed cells after 120-hour induction of the expression of d2EGFP with Doxycycline, where the transformed cells were obtained by transfecting pSFVdhfr/d2EGFP
15 and pTet-ON plasmid simultaneously in the human colon cancer COLO 320DM cells and the analysis was carried out by using the cell sorter in Example 8.

Fig. 16(i) is a view illustrating the result of analysis on the level of the expression of d2EGFP in the polyclonal population of transformed cells after
20 2-week induction of the expression of d2EGFP with Doxycycline, where the transformed cells were obtained by transfecting pSFVdhfr/d2EGFP and pTet-

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ON plasmid simultaneously in the human colon cancer COLO 320DM cells and the analysis was carried out by using the cell sorter in Example 8.

Fig. 16(j) is a view illustrating the result of analysis on the level of the expression of d2EGFP in a polyclonal population of a negative control after
5 induction of the expression of d2EGFP with Doxycycline in Example 8, where the analysis was carried out by using the cell sorter.

Fig. 17 is a chart in which averages of the levels of the expression of d2EGFP and living cell ratios are plotted against time for the polyclonal population of the transformed cells obtained by simultaneously transfecting
10 pSFVdhfr/d2EGFP and the pTet-ON plasmid into the human colon cancer COLO 320DM cells in Example 8.

Fig. 18(a) is a view illustrating the result of analysis on the level of the expression of d2EGFP in clone A before the induction of the expression in Example 8, where clone A is a given clone selected from among the
15 polyclonal population of the transformed cells obtained by transfecting pSFVdhfr/d2EGFP and pTet-ON plasmid simultaneously in the human colon cancer COLO 320DM cells and the analysis was carried out by using the cell sorter.

Fig. 18(b) is a view illustrating the result of analysis on the level of
20 the expression of d2EGFP in clone A after the induction of the expression in Example 8, where clone A is a given clone selected from among the

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polyclonal population of the transformed cells obtained by transfecting pSFVdhfr/d2EGFP and pTet-ON plasmid simultaneously in the human colon cancer COLO 320DM cells and the analysis was carried out by using the cell sorter.

5 Fig. 18(c) is a view illustrating the result of analysis on the level of the expression of d2EGFP in clone B before the induction of the expression in Example 8, where clone B is a given clone selected from among the polyclonal population of the transformed cells obtained by transfecting pSFVdhfr/d2EGFP and pTet-ON plasmid simultaneously in the human colon
10 cancer COLO 320DM cells and the analysis was carried out by using the cell sorter.

 Fig. 18(d) is a view illustrating the result of analysis on the level of the expression of d2EGFP in clone B after the induction of the expression in Example 8, where clone B is a given clone selected from among the
15 polyclonal population of the transformed cells obtained by transfecting pSFVdhfr/d2EGFP and pTet-ON plasmid simultaneously in the human colon cancer COLO 320DM cells and the analysis was carried out by using the cell sorter.

 Fig. 18(e) is a view illustrating the result of analysis on the level of
20 the expression of d2EGFP in clone C before the induction of the expression in Example 8, where clone C is a given clone selected from among the

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polyclonal population of the transformed cells obtained by transfecting pSFVdhfr/d2EGFP and pTet-ON plasmid simultaneously in the human colon cancer COLO 320DM cells and the analysis was carried out by using the cell sorter.

5 Fig. 18(f) is a view illustrating the result of analysis on the level of the expression of d2EGFP in clone C after the induction of the expression in Example 8, where clone C is a given clone selected from among the polyclonal population of the transformed cells obtained by transfecting pSFVdhfr/d2EGFP and pTet-ON plasmid simultaneously in the human colon
10 cancer COLO 320DM cells and the analysis was carried out by using the cell sorter.

Fig. 19 is a fluorescence microscopic image illustrating that implementation of the method according to Example 8 activated transcription of RNA from an amplified gene on heterochromatized HSR.

15 Fig. 20(a) is a fluorescence microscopic image illustrating the result of detection of a base sequence of pSFVdhfr/d2EGFP plasmid transfected in a HSR clone isolated in Example 9, where the detection was carried out by the FISH method.

Fig. 20(b) is a fluorescence microscopic image illustrating the result
20 of detection of a base sequence of pSFVdhfr/d2EGFP plasmid transfected in a DM clone isolated in Example 9, where the detection was carried out by the

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FISH method.

Fig. 21(a) is a fluorescence microscopic image illustrating the result of analysis on the expression of d2EGFP in the isolated HSR clone after 2-day induction of the expression of d2EGFP with Doxycycline in Example 9, where the analysis was carried out by the cell sorter.

Fig. 21(b) is a fluorescence microscopic image illustrating the result of analysis on the expression of d2EGFP in the isolated DM clone (not treated with 5-aza) after 2-day induction of the expression of d2EGFP with Doxycycline in Example 9, where the analysis was carried out by the cell sorter.

Fig. 21(c) is a fluorescence microscopic image illustrating the result of analysis on the expression of d2EGFP in the isolated DM clone (treated with 5-aza) after 2-day induction of the expression of d2EGFP with Doxycycline in Example 9, where the analysis was carried out by the cell sorter.

Fig. 21(d) is a bar graph illustrating accumulated fluorescence intensities in a negative control, the HSR clone, and DM clones (treated and not treated with 5-aza) 2 days after the addition of Doxycycline.

Fig. 22(a) is a fluorescence microscopic image illustrating the result of detection of a base sequence of pSFVdhfr/d2EGFP plasmid transfected in an isolated HSR clone in CHO cells in Example 10, where the detection was

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carried out by the FISH method.

Fig. 22(b) is a fluorescence microscopic image illustrating the result of detection of a base sequence of pSFVdhfr/d2EGFP plasmid transfected in an isolated DM clone in CHO cells in Example 10, where the detection was
5 carried out by the FISH method.

Fig. 23(a) is a view illustrating the result of analysis on the expression of d2EGFP in the isolated max clone (treated with 5-aza) after 2-day induction of the expression of d2EGFP with Doxycycline in Example 10, where the analysis was carried out by the cell sorter.

10 Fig. 23(b) is a bar graph illustrating accumulated fluorescence intensities of a polyclonal population of transformed cells as a negative control, a polyclonal population of transformed cells of Example 10, and max clone in Example 10.

Fig. 23(c) is a fluorescence microscopic image illustrating the result
15 of detection of a base sequence of pSFVdhfr/d2EGFP plasmid transfected in the max clone isolated in Example 10, where the detection was carried out by the FISH method.

BEST MODE FOR CARRYING OUT THE INVENTION

20 Embodiments of the present invention are described below. It should be noted that the present invention is not limited thereto.

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The present invention is directed to a method of expressing a protein from a repeated sequence formed in mammalian cells in which gene amplification is induced, the protein having been under expression repression. Here, what is meant by the term "repeated sequence" is a base
5 sequence in which one unit base sequence is repeated. It is known that there is a case that the presence of such a repeated sequence causes heterochromatization in a gene region in which it is present, thereby repressing the gene transcription.

As described above, the inventor of the present invention has found
10 that it becomes possible to copy a gene (target gene), which encodes a target protein, in an intercellular copy number increased to approximately 100,000 copies simply by plasmid conduction into human-derived cancer cells (COLO 320 colon cancer cell line, and HeLa cell line) by lipofection, the plasmid (hereinafter, "IR/MAR plasmid") having mammalian copying initiation
15 region (IR; initiation region) and a matrix attachment region (MAR; matrix attachment region), and selecting by utilizing a chemical-tolerant gene (Blasticidine or Neomycine), and that the mass amplification of the target gene can be attained regardless of whether the target gene has a gene structure identical to that of the IR/MAR plasmid (i.e., the target gene has a
20 cis gene structure) or the target gene has a gene structure different from that of the IR/MAR plasmid (i.e., the target gene has a trans gene structure) (see

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Patent Document 1, Patent Document 2, Non-Patent Document 1, and Non-Patent Document 2).

Therefore, it was found that a mass gene amplification system does not show an increase in transcription of Blasticidine resistant gene (target gene) in mRNA even if a copy number of the Blasticidine resistant gene is increased. It is deduced that this phenomenon was due to transcription repression by the repeated sequence produced as a result of the mass amplification of a gene region including the target gene. The present invention is accomplished to express an expression-repressed protein by (a) releasing the transcription repression caused by such a repeated sequence, or (b) selecting cells (clones) in which the transcription repression does not occur (or a degree of the transcription repression is low), out of a cell population (clone population) in the majority of which the transcription repression occurs.

Therefore, in the present invention, the term "method of expressing a protein that has been under expression repression" encompasses not only methods of expressing an expression-repressed protein by releasing transcription repression caused by a repeated sequence, but also methods of expressing a desired protein by selecting cells (clones) in which the transcription repression does not occur (or a degree of the transcription repression is low), out of a cell population (clone population) in the majority

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of which the transcription repression occurs.

In the present invention, what is meant by the term "mammalian cells in which gene amplification is induced" is mammalian cells in which amplification of a gene (polynucleotide) is induced. One example of such mammalian cells is mammalian cells in which the above-described IR/MAR plasmid is conducted. The mammalian cells are not particularly limited. For example, the mammalian cells may be CHO-K1 cells (available as e.g., ATCC CCL-61, RIKEN RCB0285, RIKEN RCB0403, etc.). However, it is especially preferable that the mammalian cells be tumor cells having an infinite proliferation ability. Examples of such tumor cells encompass HeLa cells (available as e.g., ATCC CCL-2, ATCC CCL-2.2, RIKEN RCB0007, RIKEN RCB0191 etc.), human colon cancer COLO 320DM cells (derived from, e.g., ATCC CCL-220), human colon cancer COLO 320HSR cells (available as e.g., ATCC CCL-220.1), NSO cells (available as e.g., RIKEN RCB0213), and the like.

[Embodiment 1]

A method according to the present embodiment includes transfecting a first polynucleotide and a second polynucleotide simultaneously into mammalian cells in which the gene amplification is induced, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the second

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polynucleotide encodes a protein to be expressed, the method further including transfecting at least one of a third polynucleotide and a fourth polynucleotide into the mammalian cells in transfecting the first polynucleotide and the second polynucleotide into the mammalian cells, 5 where the third polynucleotide has a length of 10kbp or more, and the fourth polynucleotide includes an insulator sequence.

The first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in the eukaryotic cells. The origin of replication included in the first polynucleotide is not particularly limited, 10 provided that the origin of replication functions in eukaryotic cells. Examples of the origin of replication encompass c-myc locus, dihydrofolate reductase locus, β -globin locus, etc. The c-myc locus is described for example in "McWhinney, C. et al., Nucleic Acids Res. vol. 18, p1233-1242 (1990)". The dihydrofolate reductase locus is described for example in "Dijkwel, P.A. et 15 al., Mol. Cell. Biol. vol.8, p5398-5409 (1988)". β -globin locus is described for example in "Aladjem, M. et al., Science vol. 281, p1005-1009 (1998)".

The nucleic matrix attachment region included in the first polynucleotide is not particularly limited, provided that the nucleic matrix attachment region functions in eukaryotic cells. Examples of the nucleic 20 matrix attachment region encompass sequences derived from a nucleic matrix attachment region of a locus such as Igk locus, SV40 initial region,

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dihydrofolate reductase locus, or the like. The nucleic matrix attachment region of the Igk locus is described for example in Tsutsui, K. et al., J. Biol. Chem. vol. 268, p12886-12894 (1993). The nucleic matrix attachment region of the SV40 initial region is described for example in Pommier, Y. et al., J. Virol., vol 64, p419-423 (1990). The nucleic matrix attachment region of dihydrofolate reductase locus is described for example in Shimizu N. et al., Cancer Res. vol. 61, p6987-6990.

If necessary for its purposes, the first polynucleotide may include a sequence necessary for cloning in *Escherichia coli*, or a selection marker such as a chemical-tolerant gene (Blasticidine resistant gene, Neomycine resistant gene, etc.), green fluorescent protein gene or the like, as a marker protein, in addition of the origin of replication and nucleic matrix attachment region. The selection marker can be used as an indicator to select the cells in which the first polynucleotide is transfected.

Meanwhile, the second polynucleotide is a polynucleotide for encoding the protein to be expressed (such a polynucleotide is referred to as a "gene (for) encoding a target protein" where appropriate). The second polynucleotide is not particularly limited and a polynucleotide encoding a desired protein can be appropriately selected to employ. The polynucleotide can be obtained based on its basic sequence information by using a known method such as PCR. In the explanation of the present invention, the "protein

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to be expressed" is referred to as the "target protein".

By simultaneously transfecting, into the mammalian cells, the first polynucleotide and the second polynucleotide that encodes the protein to be expressed, the "mass gene amplification system" disclosed in Patent Document 1 etc. by the inventor of the present invention can be constructed, and the cells can amplify the second polynucleotide. The mammalian cells in which the first and second nucleotides are simultaneously transfected are "mammalian cells in which the gene amplification is induced" as termed in the present invention. The mammalian cells to which the first and second polynucleotides are to be transfected is not particularly limited, and may be CHO cells or the like. It is however especially preferable that the mammalian cells be tumor cells that have infinite proliferation ability. Examples of the tumor cells encompass HeLa cells, human colon cancer COLO 320DM cells, human colon cancer COLO 320HSR cells, NS0 cells, etc. The human colon cancer COLO 320DM cells and human colon cancer COLO 320HSR cells are described for example in Quinn, L.A., Moore, G.E., Morgan, R.T., and Woods, L.K. Cell lines from human colon carcinoma with unusual cell products, double minutes, and homogeneously staining regions. Cancer Res. 1979 39(12):4914-4924. and in Shimizu, N., Kanda, T., and Wahl, G. M. Selective capture of acentric fragments by micronuclei provides a rapid method of purifying extrachromosomally amplified DNA. Nat. Genet., 12:

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65-71, 1996.

The transfection of the first and second polynucleotide into the cell is not particularly limited, provided that both the polynucleotides are thereby transfected simultaneously into the mammalian cells. Both the
5 polynucleotides may be transfected therein as one gene structure in which both the polynucleotides are linked, or as different gene structures. The form of the gene structure may be a plasmid or a cosmid.

The first and second polynucleotides may be transfected into the cells by any method, which may be a known method such as lipofection,
10 electroporation, particle-gun, etc.

In such mammalian cells in which the first and second polynucleotides are transfected, the amplification of the second polynucleotide produces a repeated sequence, which causes transcription repression, resulting in repression of protein expression. The inventor of the present invention
15 compared this phenomenon with a repeated sequence that occurs in gene amplification of carcinogenesis process ("Reference Example 1" described later). The comparison showed that the transcription of mRNA having a gene amplification region caused in the carcinogenesis process was not repressed. A repeated sequence produced by a plasmid (pSFVdhfr) having an IR and an
20 MAR, which was shown in Comparative Example 1, is different from the repeated sequence produced in the carcinogenesis process in that the

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sequence repeated in the latter was long (usually, 100 to 200kbp), and the repeated sequence of the latter was complicated as a result of recombination and the like. Therefore, the results of Comparative Example 1 and Reference Example 1 suggest that a longer repeating unit and more complicated
5 repeated structure would increase transcription of the repeated sequence produced by the plasmid having IR and MAR to mRNA.

A method according to the present embodiment was established based on this study. More specifically, the method according to the present embodiment includes transfecting at least one of the third polynucleotide and
10 the fourth polynucleotide into the mammalian cells in transfecting the first nucleotide and the second nucleotide into the mammalian cells, where the third polynucleotide has a length of 10kbp or more, and the fourth polynucleotide includes an insulator sequence.

By transfecting the third polynucleotide into the mammalian cells
15 simultaneously with the transfection of the first and second polynucleotides therein, the polynucleotide encoding the protein to be expressed and the third polynucleotide are coamplified. Consequently, the repeated sequence is formed with a long repeating unit or a complicated structure by the amplification of the polynucleotide (gene). This releases the transcription
20 repression caused by the repeated sequence, thereby making it possible to express the protein as much as the polynucleotides are copied.

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The length of the third polynucleotide is preferably 10kbp or longer, more preferably 50kbp or more, and most preferably 100kbp or more. The third polynucleotide is not particularly limited in terms of its basic sequence, provided that the third polynucleotide satisfies the requirement of the length.

5 The third polynucleotide may be a polynucleotide encoding a protein or a polynucleotide not encoding a protein. The third polynucleotide may be prepared appropriately by synthesis or the other method, or may be selected appropriately from among known polynucleotides such as λ -phage DNA, human genome DNA, etc.

10 On the other hand, by transfecting the fourth polynucleotide into the mammalian cells simultaneously with the transfection of the first and second polynucleotides therein, the polynucleotide encoding the protein to be expressed and the polynucleotide having the insulator sequence are coamplified. Consequently, the repeated sequence is formed with a long
15 repeating unit or a complicated structure by the amplification of the polynucleotide (gene). Further, heterochromatization will not be spread around. This releases the transcription repression caused by the repeated sequence, thereby making it possible to express the protein as much as the polynucleotides are copied.

20 Here, the "insulator sequence" is a boundary of chromatin, and it is known that the insulator sequence is a DNA sequence independent of the

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gene expression. Examples of the insulator sequence encompass HS4
insulator sequence derived from Aves (1210bp, see "Recillas-Targa, F. et al.,
Proc. Natl. Acad. Sci. U.S.A., vol. 99, p6883-6888, (2002)"), ARS insulator
sequence derived from *Echinoidea* (575bp, "Akasaka K, et al., Cell. Mol.
5 Biol. vol. 45, p555-565 (1999).").

Into the mammalian cells, the third or fourth polynucleotide and the
first or second polynucleotide may be transfected as a gene structure or
different gene structures. Because the third or fourth polynucleotide is
relatively large in size, and the transfection of one gene structure including
10 the third or fourth polynucleotide and first or second polynucleotide into the
mammalian cells further requires transfecting the third or fourth
polynucleotide into a gene structure including the first or second
polynucleotide, it is preferable that the third or fourth polynucleotide and the
first or second polynucleotide be transfected as different gene structures. For
15 attaining greater releasing of the transcription repression, it is preferable to
transfect both of the third and fourth polynucleotides into the mammalian
cells, albeit it is sufficient that the third or fourth polynucleotide is
transfected therein. As to the form and transfecting method of the gene
structure into the mammalian cells, the third and the fourth polynucleotides
20 can be described in the same way as the first and second polynucleotides are
described.

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The later-described Examples 1 and 2 discuss how effective the method according to the present embodiment is. In Examples 1 and 2, it was confirmed that transcription repression of polynucleotides encoding proteins to be expressed was released whereby the expression of target protein was
5 increased.

The present invention covers a transformant obtained by the method according to the present embodiment.

[Embodiment 2]

A method according to the present embodiment includes transfecting a
10 first polynucleotide and a fifth polynucleotide simultaneously in the mammalian cells in which the gene amplification is induced, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the fifth polynucleotide includes a second polynucleotide and a chemical tolerant
15 gene, the second polynucleotide encoding a protein to be expressed, the method further including culturing the mammalian cells sequentially in a medium of increasing concentrations of a chemical.

The fifth polynucleotide is a polynucleotide includes the second polynucleotide encoding a protein to be expressed, and a chemical tolerant
20 gene. Examples of the chemical tolerant gene encompass Blasticidine resistant gene, Neomycine resistant gene, Hygromycine tolerant gene, and the

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like. The transfection of the fifth polynucleotide can be carried out by
transfecting into the mammalian cells one gene structure including the fifth
polynucleotide and the first or fifth polynucleotide, or by transfecting into
the mammalian cells the fifth polynucleotide and the first polynucleotide as
5 different gene structures. As to the form and transfecting method of the gene
structure into the mammalian cells, the fifth polynucleotides can be described
in the same way as the first and second polynucleotides are described.

The present embodiment includes the mammalian cells, in which the
first and fifth polynucleotides are transfected, sequentially in the medium of
10 increasing concentrations of the chemical. In the mammalian cells in which
the first and the fifth polynucleotides are transfected, the polynucleotide
encoding the protein to be expressed and the chemical tolerant gene are
coamplified. By culturing such mammalian cells sequentially in the medium
of increasing concentrations of the chemical, mammalian cells in which the
15 transcription repression caused by the repeated sequence does not occur (or a
degree of the transcription repression is low) can be selected using the
chemical tolerance as an indicator. Therefore, it is possible to select the
mammalian cells in which the transcription repression caused by the repeated
sequence does not occur (or a degree of the transcription repression is low)
20 for the polynucleotide encoding the protein to be expressed. Thereby, it
becomes possible to express the protein as much as this polynucleotide is

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copied.

The increasing the concentration of the chemicals for the cell culturing can be done in any way. The cells may be passaged in the medium of increasing concentrations of the chemical. As an alternative, the chemical
5 may be added to the medium consequently. More specifically, the former case is preferably arranged such that the cells are incubated in the medium of one concentration for 3 to 7 days and then incubated to the medium of a next concentration. Moreover, the former case is preferably arranged such that an increasing rate of the chemical concentration is in a range of 30 to 100%. The
10 latter case is preferably arranged such that the chemical is added to the medium to increase its concentration by 10% to 30% per 24 hours. In the later-described Example 3, the concentration of a chemical (Blasticidine) was doubled in intervals of 3 to 5 days.

There is no limitation as to an upper limit of the chemical to be add to
15 the medium, because the upper limit is dependent on the kind of the chemical, kind of the cells, and conditions, etc. The upper limit should be determined as appropriate.

The later-described Example 3 discusses how effective the method according to the present embodiment. In Example 3, it was confirmed that
20 transcription repression of a polynucleotide encoding a protein to be expressed was released whereby the expression of the target protein was

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increased.

[Embodiment 3]

A method according to the present embodiment includes transfecting a first polynucleotide and a sixth polynucleotide simultaneously in the mammalian cells in which the gene amplification is induced, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the sixth polynucleotide includes a promoter region and a second polynucleotide controllably linked with each other, the second polynucleotide encoding a protein to be expressed, the method further including transfecting into the mammalian cells a seventh polynucleotide, which encodes a transcription activation factor of the promoter, and expressing the transcription activation factor.

The sixth polynucleotide is a polynucleotide including a promoter region and the second polynucleotide controllably linked with each other. The second polynucleotide is a polynucleotide encoding a protein to be expressed. The promoter region is not particularly limited, provided that the promoter region can function in the mammalian cells in which the sixth polypeptide is to be transfected, and transcription of the promoter region can be controlled by the transcription activation factor. For example, the promoter region may be a TRE promoter (Clontech Laboratories Inc.), T-

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REX promoter (Invitrogen Corp.), or the like. The transfection of the sixth polynucleotide into the mammalian cells having a gene amplification ability may be carried out by transfecting one gene structure including the sixth polynucleotide and the first polynucleotide, or by transfecting the sixth
5 polynucleotide and the first polynucleotide as different gene structures. As to the form and transfecting method of the gene structure into the mammalian cells, the sixth polynucleotides can be described in the same way as the first and second polynucleotides are described.

The present embodiment includes transfecting into the mammalian
10 cells the seventh polynucleotide, which encodes a transcription activation factor of the promoter, and expressing the transcription activation factor. By this step, it is possible to encourage the activation of the promoter that controls the expression of the protein to be expressed. This makes it possible to express the protein that has been under the expression repression by the
15 repeated sequence. The seventh polynucleotide is a polynucleotide that encodes the transcription activation factor of the promoter included in the sixth polynucleotide. For example, if the promoter was a TRE promoter, the seventh polynucleotide might be Tet-ON gene (Clontech Laboratories Inc.).

In transfecting the seventh polynucleotide into the mammalian cells
20 and expressing the transcription activation factor, the transfection of the seventh polynucleotide may be carried out by one gene structure into the

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mammalian cells, the gene structure constructed by controllably linking the seventh polynucleotide with a promoter that promotes the expression of the seventh polynucleotide. The promoter for promoting the expression of the seventh polynucleotide is not particularly limited, provided that the promoter
5 can function in mammalian cells. The promoter may be inducible or non-inducible. Moreover, timing of the transfecting the gene structure including the seventh polynucleotide into the mammalian cells may be after or before the transfection of the first and sixth polynucleotides. As to the form and transfecting method of the gene structure into the mammalian cells, the
10 seventh polynucleotides can be described in the same way as the first and second polynucleotides are described.

The later-described Example 4 discusses how effective the method according to the present embodiment. In Example 4, it was confirmed that transcription repression of a polynucleotide encoding a protein to be
15 expressed was released whereby the expression of the target protein was increased.

The present invention covers a transformant obtained by the method according to the present embodiment.

[Embodiment 4]

20 A method according to the present embodiment includes transfecting a first polynucleotide and an eighth polynucleotide simultaneously in the

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mammalian cells in which the gene amplification is induced, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the eighth polynucleotide includes LoxP gene and a second polynucleotide encoding a protein to be expressed, the method further including transfecting into the mammalian cells a ninth polynucleotide, which includes Cre Recombinase gene, and expressing the Cre Recombinase gene.

The eighth polynucleotide is a polynucleotide, which includes the second polynucleotide and the LoxP gene. The LoxP gene is a gene relating to Cre-LoxP system (see "Molecular Cloning----a laboratory manual 3rd Ed.", by J. Sambrook and D.W. Russell, Cold Spring Harbor Laboratory Press (2001), page 4.82-4.85.) The LoxP gene may be provided with a restriction endonuclease recognizing sequence or a restriction endonuclease cleaved sequence on both terminals (5' terminal and 3' terminal).

The ninth polynucleotide is a polynucleotide, which includes Cre Recombinase gene that relates to Cre-LoxP System. The ninth polynucleotide preferably includes a promoter for promoting the expression of the Cre Recombinase gene.

The transfection of the first, eighth, and ninth polynucleotides into the mammalian cells causes amplification of the LoxP gene and the polynucleotide for encoding the protein to be expressed. As a result, Cre

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Recombinase expressed in the mammalian cells recognizes the LoxP gene and extrachromosomally cleaves the polynucleotide for encoding the protein to be expressed. The cleaved-away polynucleotide forms a cyclic molecule. The extrachromosomal cyclic molecule is hardly susceptible to the transcription repression or the like. Therefore, this avoids the transcriptions
5 caused by the repeated sequence. Thereby, it becomes possible to express the protein as much as this polynucleotide is copied.

As to the transfecting method of the polynucleotides into the mammalian cells, the polynucleotides can be described in the same way as
10 the first and second polynucleotides are described. Moreover, timing of the transfecting, the transfection of the ninth polynucleotide may be after or before the transfection of the first and eighth polynucleotides.

The later-described Example 5 discusses how effective the method according to the present embodiment. In Example 5, it was confirmed that
15 transcription repression of a polynucleotide encoding a protein to be expressed was released whereby the expression of the target protein was increased.

The present invention covers a transformant obtained by the method according to the present embodiment.

20 [Embodiment 5]

A method according to the present embodiment includes transfecting a

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first polynucleotide and a second polynucleotide simultaneously into mammalian cells in which the gene amplification is induced, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the second
5 polynucleotide encodes a protein to be expressed, the method further including treating the mammalian cells with 5-aza-2'-deoxycytidine.

Silencing (transcription repression) of DNA by RNAi is accompanied with methylation of DNA. On this account, the inventor of the present invention uniquely arrived to an idea that the transcription repression can be
10 released by lowering a degree of the metylation of DNA. Because it is known that 5-aza-2'-deoxycytidine lowers the degree of the metylation of DNA, the inventor of the present invention employs 5-aza-2'-deoxycytidine. 5-aza-2'-deoxycytidine may be a commercially available one. For example, 5-aza-2'-deoxycytidine may be purchased from Sigma.

15 The treatment of the mammalian cells with 5-aza-2'-deoxycytidine may be carried out in any way, provided that the mammalian cells can be in contact with 5-aza-2'-deoxycytidine. For example, the mammalian cells may be cultured in a medium containing 5-aza-2'-deoxycytidine. In this arrangement, differenet kinds and conditions of the cells have different
20 optimum conditions. Thus, the concentration of 5-aza-2'-deoxycytidine may be set as appropriate. In general, the concentration of 5-aza-2'-deoxycytidine

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is preferably in a range of 0.2 μ M to 10 μ M, and more preferably in a range of 1 μ M to 2 μ M. A concentration of 5-aza-2'-deoxycytidine above the preferable ranges would be toxic to the mammalian cells, while a concentration of 5-aza-2'-deoxycytidine below the preferable ranges could not sufficiently
5 release the transcription repression. The medium to which 5-aza-2'-deoxycytidine is added is not particularly limited, provided that the mammalian cells can be cultured in the medium. For example, the medium may be Dulbecco's modified Eagle's medium (DEM medium: Invitrogen Corp.), RPMI1640 medium (Nissui Pharmaceutical Co., Ltd.), or the like
10 medium. The culturing is carried out with the medium to which 10% fetal bovine serum is added.

Treatment period of the mammalian cells is not particularly limited, because the treatment period is dependent on the concentration of 5-aza-2'-deoxycytidine, the kinds and conditions of the cells. However, it is preferable
15 that the mammalian cells be treated for a period in a range of 3 days to 10 days.

The later-described Example 6 discusses how effective the method according to the present embodiment. In Example 6, it was confirmed that transcription repression of a polynucleotide encoding a protein to be
20 expressed was released whereby the expression of the target protein was increased.

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The method including treating the mammalian cells with 5-aza-2'-deoxycytidine is not only applicable to the mammalian cells in which the first and second polynucleotides are transfected, but also to a wide range of mammalian cells in which the gene amplification is induced. For example, this method is also applicable to mammalian cells prepared by simultaneously transfecting (a) a gene encoding a protein and (b) DHFR (Dihydrofolate reductase) gene into CHO (Chinese Hamster Ovary) cells so that the gene amplification becomes possible.

[Embodiment 6]

10 A method according to the present embodiment includes transfecting a first polynucleotide and a second polynucleotide simultaneously into mammalian cells in which the gene amplification is induced, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the second
15 polynucleotide encodes a protein to be expressed, the method further including selecting mammalian cells that the gene amplification occurs on a double minute chromosome.

The inventor of the present invention isolated clones expressing a target protein, from among stable transformant cell population of mammalian
20 cells in which a polynucleotide (gene) encoding the target protein and an IR/MAR plasmid. Then, the inventor compared the clones in terms of an

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amount of the target protein expressed. The comparison demonstrated that the expression of the target protein was increased in a clone in which the amplification of the polynucleotide (gene) encoding the target protein occurred on the extrachromosomal double minute chromosome (hereinafter, referred to as "DM") than in a clone in which the amplification of the polynucleotide (gene) encoding the target protein occurred on a homogeneously staining regeon (hereinafter, referred to as "HSR") of a chromosome. That is, a clone in which the transcription repression caused by the repeated sequence is not occurred can be selected by selecting a clone in which the amplification of the polynucleotide (gene) encoding a target protein is occurred on DM, from among mammalian cells in which the polynucleotide (gene) is highly amplified by using a mass gene amplification system.

The selecting of the mammalian cells (clone) in which the gene amplification occurs on DM may be carried out by any method. One example of the selecting method is a method including performing a well-known FISH method (fluorescence in situ hybridization) on chromosomes in mitotic phase, detecting any one of the first to ninth polynucleotides transfected in the mammalian cells, and isolating the mammalian cells (clone) having fluorescing DM. HSR is a region on a chromosome, while DM is present out of the chromosome. Thus, mammalian cells (clone) having a

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fluorescing extrachromosomal region may be select by fluorescence microscope observation. The FISH method is not specifically limited and may be performed in any way. The FISH method may be carried out in a well-known fashion appropriately selected.

5 As an alternative, another example of the selecting method is a method including embedding Lactose Operator (LacO) sequence in any one of the first to ninth polynucleotides, amplifying the polynucleotide containing Lactose Operator (LacO), and visualizing the polynucleotide containing Lactose Operator (LacO) by expressing Lactose Repressor
10 (LacR)-Green Fluorescence Protein (GFP) fusion gene (see "Kanda, T., and G.M. Wahl. 2000. The dynamics of acentric chromosomes in cancer cells revealed by GFP-based chromosome labeling strategies. *J Cell Biochem. Suppl*:107-114."; "Li, G., G. Sudlow, and A.S. Belmont. 1998. Interphase cell cycle dynamics of a late-replicating, heterochromatic homogeneously
15 staining region: Precise choreography of condensation/decondensation and nuclear positioning. *J Cell Biol.* 140:975-989."; and "Shimizu, N., K. Shingaki, Y. Kaneko-Sasaguri, T. Hashizume, and T. Kanda. 2005. When, where and how the bridge breaks: anaphase bridge breakage plays a crucial role in gene amplification and HSR generation. *Exp Cell Res.* 302:233-
20 243.").

The followings are known for DM and HSR. Gene amplification is a

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main system for attaining infinite proliferation -restricting amplification and anti-chemical property of tumor cells (see "Benner, S. E., Wahl, G. M., and Von Hoff, D. D. Double minute chromosomes and homogeneously staining regions in tumors taken directly from patients versus in human tumor cell
5 lines. *Anti-Cancer Drugs*, 2: 11-25, 1991"). Cytogenetically, the amplified gene is most frequently found on the extrachromosomal double minute chromosome (DM) in vivo. Long in vitro passage, however, dominantly amplifies cells having the amplified gene in a homogeneously staining region (HSR) of a chromosome in general. Previous researches showed that the
10 tumor cells in tumor expressing and cell differentiating states are restored to a normal state by removing the amplified gene on DM therefrom (see "Shimizu, N., Nakamura, H., Kadota, T., Kitajima, K., Oda, T., Hirano, T., and Utiyama, H. Loss of amplified c-myc genes in the spontaneously differentiated HL-60 cells. *Cancer Res.*, 54: 3561-3567, 1994.", "Von Hoff,
15 D. D., McGill, J. R., Forseth, B. J., Davidson, K. K., Bradley, T. P., Van Devanter, D. R., and Wahl, G. M. Elimination of extrachromosomally amplified MYC genes from human tumor cells reduces their tumorigenicity. *Proc. Natl. Acad. Sci. USA*, 89: 8165-8169, 1992.", and "Eckhardt, S. G., Dai, A., Davidson, K. K., Forseth, B. J., Wahl, G. M., and Von Hoff, D. D.
20 Induction of differentiation in HL60 cells by the reduction of extrachromosomally amplified c-myc. *Proc. Natl. Acad. Sci. USA*, 91: 6674 -

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6678, 1994."). This kind of removing step is mediated by selectively
 uptaking DM into microcells discharged from dividing cells. (see "Von Hoff,
 D. D., McGill, J. R., Forseth, B. J., Davidson, K. K., Bradley, T. P., Van
 Devanter, D. R., and Wahl, G. M. Elimination of extrachromosomally
 5 amplified MYC genes from human tumor cells reduces their tumorigenicity.
 Proc. Natl. Acad. Sci. USA, 89: 8165-8169, 1992.", "Shimizu, N., Kanda, T.,
 and Wahl, G. M. Selective capture of acentric fragments by micronuclei
 provides a rapid method of purifying extrachromosomally amplified DNA.
 Nat. Genet., 12: 65-71, 1996.", and "Shimizu, N., Shimura, T., and Tanaka,
 10 T. Selective elimination of acentric double minutes from cancer cells through
 the extrusion of micronuclei. Mutat. Res., 448: 81-90, 2000."). It has been
 understood that the micronucleation closely relates to behavior of DM in the
 cells in cell cycle (see "Tanaka, T., and Shimizu, N. Induced detachment of
 acentric chromatin from mitotic chromosomes leads to their cytoplasmic
 15 localization at G1 and the micronucleation by lamin reorganization at S
 phase. J. Cell Sci., 113: 697 - 707, 2000."). DM is constituted by acentic
 cyclic DNAs in various sizes (see "Levan, A., and Levan, G. Have double
 minutes functioning centromeres? Hereditas, 88: 81-92, 1978."). Albeit it is
 acentic, DM can be divided and descended stably to daughter cells by
 20 attaching to mitotically dividing chromosomes (see "Tanaka, T., and
 Shimizu, N. Induced detachment of acentric chromatin from mitotic

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chromosomes leads to their cytoplasmic localization at G1 and the
micronucleation by lamin reorganization at S phase. *J. Cell Sci.*, 113: 697-
707, 2000.", "Levan, A., and Levan, G. Have double minutes functioning
centromeres? *Hereditas*, 88: 81-92, 1978.", and "Kanda, T., Sullivan, K. F.,
5 and Wahl, G. M. Histone-GFP fusion protein enables sensitive analysis of
chromosome dynamics in living mammalian cells. *Curr. Biol.*, 8: 377-385,
1998."). Recent important relevant findings showed that similar systems are
used in mitosis of some viral nuclear plasmids such as bovine papilloma
virus (see "Lehman, C. W., and Botchan, M. R. Segregation of viral plasmids
10 depends on tethering to chromosomes and is regulated by phosphorylation.
Proc. Natl. Acad. Sci. USA, 95: 4338-4343, 1998."), EB virus (see
"Marechal, V., Dehee, A., Chikhi-Brachet, R., Piolot, T., Coppey-Moisan,
M., and Nicolas, J. C. Mapping EBNA-1 domains involved in binding
to metaphase chromosomes. *J. Virol.*, 73: 4385-4392, 1999."), Kaposi's
15 sarcoma-associated herpes virus (see "Ballestas, M. E., Chatis, P. A., and
Kaye, K. M. Efficient persistence of extrachromosomal KSHV DNA
mediated by latency-associated nuclear antigen. *Science (Wash. DC)*, 284:
641-644, 1999."), and SV40 (see "Baiker, A., Maercker, C., Piechaczek, C.,
Schmidt, S. B., Bode, J., Benham, C., and Lipps, H. J. Mitotic stability of an
20 episomal vector containing a human scaffold/ matrix-attached region is
provided by association with nuclear matrix. *Nature Cell Biol.*, 2: 182 - 184,

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2000.”). Furthermore, a recent interesting research found that a plasmid having EB virus replicon can be embedded into DM in tumor cells (see “Kanda, T., Otter, M., and Wahl, G. M. Mitotic segregation of viral and cellular acentric extrachromosomal molecules by chromosome tethering. J. Cell Sci., 114: 49-58, 2001.).

As to the method of transfecting the first and second polynucleotides into the mammalian cells, and as to the gene structure for transfecting the polynucleotides into the mammalian cells, the present embodiment may be described in the same way as Embodiment 1.

10 The later-described Example 7 discusses how effective the method according to the present embodiment. In Example 7, it was confirmed that transcription repression of a polynucleotide encoding a protein to be expressed was released whereby the expression of the target protein was increased.

15 The method according to the present embodiment is not only applicable to the mammalian cells in which the first and second polynucleotides are transfected, but also to a wide range of mammalian cells in which the gene amplification is induced. For example, this method is also applicable to mammalian cells prepared by simultaneously transfecting (a) a
20 gene encoding a protein and (b) DHFR (Dihydrofolate reductase) gene into CHO (Chinese Hamster Ovary) cells so that the gene amplification becomes

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possible.

[Embodiment 7]

A method according to the present embodiment includes transfecting a first polynucleotide and a sixth polynucleotide simultaneously in the mammalian cells in which the gene amplification is induced, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the sixth polynucleotide includes a promoter region and a second polynucleotide controllably linked with each other, the second polynucleotide encoding a protein to be expressed, the method further including transfecting a seventh polynucleotide into the mammalian cells simultaneously in transfecting the first and sixth polynucleotides into the mammalian cells, the seventh polynucleotide encoding a transcription activation factor of the promoter.

The sixth polynucleotide is a polynucleotide including a promoter region and the second polynucleotide controllably linked with each other. The second polynucleotide is a polynucleotide encoding a protein to be expressed. The promoter region is not particularly limited, provided that the promoter region can function in the mammalian cells in which the sixth polypeptide is to be transfected, and transcription of the promoter region can be controlled by the transcription activation factor. For example, the promoter region may be a TRE promoter (Clontech Laboratories Inc.), T-

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REX promoter (Invitrogen Corp.), or the like. The transfection of the sixth polynucleotide into the mammalian cells having a gene amplification ability may be carried out by transfecting one gene structure including the sixth polynucleotide and the first polynucleotide, or by transfecting the sixth
5 polynucleotide and the first polynucleotide as different gene structures. As to the form and transfecting method of the gene structure into the mammalian cells, the sixth polynucleotides can be described in the same way as the first and second polynucleotides are described.

The present invention includes transfecting the seventh polynucleotide
10 into the mammalian cells simultaneously in transfecting the first and sixth polynucleotides into the mammalian cells, the seventh polynucleotide encoding a transcription activation factor of the promoter. This step improves the activity of the promoter that controls the expression of the target protein, thereby allowing the expression of the target protein that has been under the
15 expression repression caused by the repeated sequence. The seventh polynucleotide is a polynucleotide that encodes the transcription activation factor of the promoter included in the sixth polynucleotide. For example, if the promoter was a TRE promoter, the seventh polynucleotide might be Tet-
ONTM Gene (Clontech Laboratories Inc.).

20 In transfecting the seventh polynucleotide into the mammalian cells and expressing the transcription activation factor, the transfection of the

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seventh polynucleotide may be carried out by one gene structure into the mammalian cells, the gene structure constructed by controllably linking the seventh polynucleotide with the promoter that promotes the expression of the seventh polynucleotide. The promoter for promoting the expression of the seventh polynucleotide is not particularly limited, provided that the promoter can function in mammalian cells. The promoter may be inducible or non-inducible. Moreover, timing of the transfecting the gene structure including the seventh polynucleotide into the mammalian cells is concurrent with the transfecting the first and sixth polynucleotides (cotransfection). In one simple method, a mixture of (a) the gene structure including the first and sixth polynucleotides and (b) the gene structure including the seventh polynucleotide may be used to transfect the polynucleotides into the mammalian cells by lipofection or the like. As an alternative, a mixture of (i) a gene structure including the first polynucleotide, (ii) a gene structure including the sixth polynucleotide, and (iii) a gene structure including the seventh polynucleotide may be used to transfect the polynucleotides into the mammalian cells by lipofection or the like. As to the form and transfecting method of the gene structure into the mammalian cells, the polynucleotides can be described in the same way as the first and second polynucleotides are described.

The later-described Example 8 discusses how effective the method

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according to the present embodiment is. In Example 8, it was confirmed that transcription repression of polynucleotides encoding proteins to be expressed was released whereby the expression of the target protein was increased.

The present invention covers a transformant obtained by the method
5 according to the present embodiment.

The steps described in Embodiments 1 to 7 may be appropriately combined. Such combinations of these steps are more effective in releasing the transcription repression caused by the repeated sequence, whereby the expression of the protein having been under the expression repression can be
10 further increased.

[Embodiment 8]

The present invention encompasses a kit for expressing a protein having been under expression repression, from a repeated sequence formed in mammalian cells in which gene amplification is induced, the kit comprising
15 any ones of the polynucleotides described in Embodiments 1 to 7, an agent, and/or the like in appropriate combinations. The kit according to the present invention makes it possible to easily express a protein having been under expression repression, from a repeated sequence formed in mammalian cells in which gene amplification is induced.

20 There is no particular limitation in the kit according to the present invention. One example of the kit is a kit including the first polynucleotide

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and at least one of the third and fourth polynucleotides, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, the third polynucleotide has a length of 10kbp or more, and the fourth polynucleotide includes an
5 insulator sequence. The kit may be used according to the method described in Embodiment 1. Moreover, the first and the third and/or fourth polynucleotide may be constituted as one gene structure.

Moreover, one exemplary kit according to the present invention includes the first polynucleotide and the seventh polynucleotide, where the
10 first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the seventh polynucleotide encodes a transcription activation factor of the promoter. After the sixth polynucleotide is prepared by linking the promoter region and target gene, this kit is used according to the method described in Embodiment
15 3 or 7. The first polynucleotide and the polynucleotide including the promoter region may be constituted as one gene structure.

Another exemplary kit according to the present invention includes the first polynucleotide, a polynucleotide including LoxP gene, and the ninth polynucleotide, where the first polynucleotide includes an origin of
20 replication and a nucleic matrix attachment region that function in eukaryotic cells, and the ninth polynucleotide includes Cre Recombinase gene. After the

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eighth polynucleotide is prepared by linking, with the target gene, the polynucleotide including the LoxP gene, this kit is used according to the method described in Embodiment 4. The first polynucleotide and the polynucleotide including the LoxP gene may be constituted as one gene
5 structure.

Moreover, the kit according to the present invention may include 5-aza-2'-deoxycytidine, which increases the level of the expression of the protein as described above.

The kit according to the present invention may include an agent, tool,
10 and/or the like necessary for carrying out the method according the present invention described in Embodiments 1 to 7. For example, the kit according to the present invention may include an reagent and a tool necessary for transfecting the polynucleotides into the mammalian cells. Furthermore, the kit may include mammalian cells and a medium or the like for culturing the
15 mammalian cells.

In the following, examples are described referring to the drawing attached herewith, in order to explain the embodiments of the present invention more specifically. Needless to say, the present invention is not limited to these examples and may be modified in various ways in details.
20 Furthermore, the present invention is not limited to the embodiments described above and may be modified within the scope of claims. The

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technical scope of the present invention also includes embodiments obtained by appropriately combining the disclosed technical means.

Moreover, all the academic documents and patent documents cited in the Description of the present application are incorporated herein by
5 reference.

[Examples]

Materials and methods employed in Comparative Examples 1, Reference Example 1, and Examples 1 to 10 are described below.

(Plasmid)

10 pSFVdhfr (11.0kbp) was kindly provided from Dr. John Kolman and Dr. Geoffrey M. Wahl (The Salk Institute, San Diego, CA). pSFVdhfr has a 4.6kbp fragment including Ori β derived from 3'-downstream region with respect to hydrofolate reductase (see "Dijkwel, P. A., and Hamlin, J. L. Matrix attachment regions are positioned near replication initiation sites,
15 genes, and an interamplicon junction in the amplified dihydrofolate reductase domain of Chinese hamster ovary cells. Mol. Cell Biol., 8: 5398-5409, 1988.").

All sequences derived from dihydrofolate reductase is deleted from pSFVdhfr by NotI digestion so as to construct pSFV-V plasmid (6.4kbp)
20 lacking the origin of replication.

Further, pSFVdhfr(GAP-GFP) was constructed by deleting

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Hygromycine tolerant gene expressing unit of pSFVdhfr, and embedding, at
the position where the expressing unit had been present, a GAP-GFP
expressing unit cleaved out from pEPBG plasmid (kindly provided from Dr.
Geoffrey M. Wahl, Salk Institute) so that transcription would proceed toward
5 IR.

Moreover, pSFVdhfr/d2EGFP was constructed by deleting
Hygromycine tolerant gene-expressing unit of pSFVdhfr, and embedding, at
the locus where the expressing unit had been present, a TRE promoter,
d2EGFP structuring gene, SV40 poly A sequence (derived from pTRE-
10 d2EGFP (Clontech Laboratories Inc.)) in this order, so that transcription
would proceed toward IR.

(Cells)

Human colorectal COLO 320DM and COLO 320HSR tumor cell line
(human colon cancer COLO 320DM cell line and human colon cancer COLO
15 320HSR cell line) were obtained and maintained according to the description
in "Shimizu, N., Kanda, T., and Wahl, G. M. Selective capture of
acentricfragments by micronuclei provides a rapid method of purifying
extrachromosomally amplified DNA. Nat. Genet., 12: 65-71, 1996."

Moreover, culturing of the cells and transformed cells thereof was
20 carried out according to this document. In short, the cell lines were cultured
at 37°C in the presence of 5% CO₂ in a RPMI 1640 medium (Nissui

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Pharmaceutical Co., Ltd.) to which 10% fetal bovine serum was added.

(Lipofection)

All the plasmids were purified by using Qiagen purification kit (Qiagen Inc., Valencia CA), and transfected into cells by Gene Porter 2
5 lipofection kit (Gene Therapy Systems, San Diego, CA).

(Analysis by Cell Sorter)

Fluorescence intensity of these cells were measured by using a cell sorter. The measurement was carried out by sorting 20,000 cells by setting living cells at a gate. The cell sorter used was made by Becton Dickinson,
10 and operated under conditions as described in the manual attached thereto.

[Comparative Example 1] Gene Amplification using Plasmid having IR and MAR, and Measurement of Transcription Amount of mRNA

(Method)

Into human colon cancer COLO 320 DM cell line, a plasmid
15 (pSFVdhfr) having IR and MAR derived from human DHFR (dihydrofolate reductase) gene was transfected by lipofection. Moreover, as a control, a vector plasmid (pSFV) not having IR and MAR derived from human DHFR (dihydrofolate reductase) gene was transfected into human colon cancer COLO 320 DM cell line by lipofection.

20 Both the plasmids had a Blasticidine resistant gene. Therefore, transformed cells were selected with 5µg/ml Blasticidine (Funakoshi Corp.).

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(Result)

From the cells transformed with pSFVdhfr, a clone (clone 12) in which gene amplification of the plasmid sequence occurred on double minute (DM), and a clone (clone 22) in which the gene amplification of the plasmid sequence occurred on Homogeneously staining region (HSR) were obtained. Moreover, the cells transformed with the vector plasmid pSFV-V did not increase the gene copy number because of the lack of IR and MAR, but gave transfected cells in which many copies were transfected.

These cells were quantitatively analyzed by Competitive PCR in terms of an amount of DNA of the Blasticidine resistant gene, and an amount of transcribed mRNA. The results are shown in Table 1.

Table 1

	Gene Copy Number	Level of Expression of RNA (relative ratio)
pSFV-V	12	1
pSFVdhfr clone 12	653	3.3
pSFVdhfr clone 22	3273	0.87

15

As shown in Table 1, it was found that, due to the plasmid (pSFVdhfr) having the IR and MAR, the increase in the copy number of the Blasticidine

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resistant gene did not lead to the increase in the transcription of the
Blasticidine resistant gene to mRNA. It was deduced that repeated sequence
was produced as a result of the mass amplification of the gene region
including the target gene and the repeated sequence caused transcription
5 repression.

[Reference Example 1] Transcription from Gene Amplification Region
caused in Carcinogenesis Process to mRNA

Regarding human colon cancer cell line COLO 320DM cell line
(ATCC CCL220), it is known that c-myc cancer gene is amplified in
10 carcinogenesis process in patients and locally present on DM or HSR (see
"Alitalo, Kari, Schwab, Manfred, Lin, C.C., Varmus, Harold, Bishop, J.
Michael, Homogenously staining chromosomal regions contain amplified
copies of an abundantly expressed cellular oncogene(c-myc) in malignant
neuroendocrine cells from a human colon carcinoma. Proc Natl Acad Sci U S
15 A. v80., p1701-1711"). Thus, c-myc cancer gene was quantitatively analyzed
by competitive PCR in terms of its copy number and mRNA transcription
amount. The competitive PCR was carried out following a method described
in "shimizu et al., nature genetics 1996".

The results are shown in Table 2.

20

Table 2

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	Gene Copy Number	Level of Expression of RNA (relative ratio)
WI-38	1	1
COLO 320 DM	60	65
COLO 320 HSR	24	33

As shown in Table 2, mRNA transcription in proportion with the copy number of c-my cancer gene was detected in human colon cancer cell line COLO 320DM and human colon cancer cell line COLO 320HSR. The control
5 was normal human diploid fibrocyte line WI-38.

The results showed that the transcription from the gene amplified gene caused in carcinogenesis process to mRNA was not repressed.

A repeated sequence caused in carcinogenesis process is different from the repeated sequence produced using the plasmid (pSFVdhfr) having
10 IR and MAR in that the repeated sequence caused in carcinogenesis process is long (usually, 100 to 200kbp) and it is complicated by recombination etc. The results of Comparative Example 1 and Reference Example 1 suggested that a longer repeating unit and more complicated repeating structure would increase the mRNA transcription amount from the repeated sequence
15 produced using the plasmid having IR and MAR.

[Example 1] Effect of Transfecting λ -Phage DNA and Target Gene into Host Cells

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A plasmid (pSFVdhfr (GAP-GFP)) and λ -phage DNA were mixed at a ratio of 1:2 by weight. The plasmid had IR and MAR derived from human DHFR (dihydrofolate reductase) locus, and expresses a fused protein from GFP (Green Fluorescence Protein) gene and GAP (G-associated polypeptide) gene. The mixture of the plasmid and λ -phage DNA was transfected to human colon cancer cells COLO 320DM cells by lipofection. From 2 days from the gene transfection, the cells were selected using 5 μ g/ml of Blasticidine (Funakoshi Corp.) that was selective to Blasticidine tolerant gene on the plasmid. After 3 to 4 weeks, many colonies of transformed cells were obtained.

Fig. 1 shows results of fluorescence intensity analysis of GFP by using a cell sorter. Fig. 1(a) shows the result of a case where only pSFVdhfr (GAP-GFP), but not λ -phage DNA, was transfected into the cells. Fig. 1(b) shows a result of a case where pSFVdhfr (GAP-GFP) and λ -phage DNA was transfected into the cells (cotransfection). In parenthesis in Figs. 1, averages of fluorescence intensity is shown.

Fluorescence intensity in the case where the mixture of pSFVdhfr (GAP-GFP) and λ -phage DNA was transfected into the cells (cotransfection) was 85.27, while the control had a fluorescence intensity of 65.51. Thus, this indicated that cotransfection clearly increases expression of GFP (target protein). In other words, it was found that coamplification of (a) a

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polynucleotide encoding a target protein and (b) λ -phage DNA releases the repeated sequence-caused expression repression on the protein.

[Example 2] Effect of Transfection of Insulator Sequence and Target Gene into Host Cells

5 (Method)

A plasmid DNA having HS4 insulator sequence derived from Aves (see "Recillas-Targa, F. et al., Proc. Natl. Acad. Sci. U.S.A., vol. 99, p6883-6888, (2002)") and a plasmid (pSFVdhfr/d2EGFP) were mixed and transfected into human colon cancer cell COLO 320DM cells by lipofection.

10 The plasmid (pSFVdhfr/d2EGFP) was a plasmid having (a) IR and MAR derived from human DHFR gene locus, (b) Blasticidine tolerant gene, and (c) d2EGFP gene that was under control of TRE-promoter (Tetracycline inducing promoter). The plasmid DNA having the Aves-derived HS4 insulator sequence was a plasmid having 5' HS4 insulator sequence of 1.2
15 kbp described in the paper of Recillas-Targa, F. et al. From 2 days from the gene transfection, the cells were selected using 5 μ g/ml of Blasticidine (Funakoshi Corp.) that was selective to Blasticidine tolerant gene on the plasmid. After 3 to 4 weeks, many colonies of transformed cells were obtained.

20 From 2 days from the gene transfection, the cells were selected using 5 μ g/ml of Blasticidine (Funakoshi Corp.) that was selective to Blasticidine

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tolerant gene on the plasmid. After 3 to 4 weeks, many colonies of transformed cells were obtained. After that, the transformed cells were selected by passaging in media of increasing concentration of Blasticidine (up to final concentration of 320µg/ml) with passage intervals of 3 to 6 days.

5 (Results)

Fig. 2 shows results of fluorescence intensity analysis of GFP by using a cell sorter.

Fig. 2(a) shows a result of control where only pSFVdhfr/d2EGFP was transfected into cells. Fig. 2(b) shows a result of a case where only
10 pSFVdhfr/d2EGFP was transfected into cells and the cells were then selected with 80µg/ml of Blasticidine. Fig. 2(c) shows a result of a case where pSFVdhfr/d2EGFP and the plasmid DNA having the Aves-derived HS4 insulator sequence were mixed and transfected into the cells. Fig. 2(d) shows a result of a case where pSFVdhfr/d2EGFP and the plasmid DNA having the
15 Aves-derived HS4 insulator sequence were mixed and transfected into the cells, and the cells were then selected with 80µg/ml of Blasticidine. In parenthesis in Figs. 2, averages of fluorescence intensity is shown.

In this method for detecting the fluorescence intensity, detection limit was 1, and all fluorescence intensities of 1 or less would be detected as 1. In
20 view of this, a product of a ratio (cell %) of cells of fluorescence intensities of 2 or more in all living cells (10,000 cells) and an average of the

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fluorescence intensities of the cells having fluorescence intensities of 2 or more was calculated out for each experimental plot. This product is referred to as "accumulated fluorescence intensity" hereinafter and in later Examples and shown in boxed letter in the figures.

5 While the control shown in Fig. 2(a) had an average fluorescence intensity of 2.73 and an accumulated fluorescence intensity of 61.5, the case shown in Fig. 2(c) where pSFVdhfr/d2EGFP and the plasmid DNA having the Aves-derived HS4 insulator sequence were mixed and transfected into the cells had an average fluorescence intensity of 7.14 and an accumulated
10 fluorescence intensity of 312. This clearly demonstrated that the expression of d2EGFP, which was the target protein, was increased as a result of the cotransfection subsequent coamplification of the target protein-encoding gene and the Aves-derived HS4 insulator sequence.

Moreover, the average fluorescence intensity was 6.09 and the
15 accumulated fluorescence intensity was 542 in the case shown in Fig. 2(b) where only pSFVdhfr/d2EGFP was transfected into cells and the cells were then selected with 80µg/ml of Blastidine, while the control shown in Fig. 2(a) had the average fluorescence intensity of 2.73 and the accumulated fluorescence intensity of 61.5. the average fluorescence intensity was 9.65
20 and the accumulated fluorescence intensity was 941 in the case shown in Fig. 2(d) where a result of a case where pSFVdhfr/d2EGFP and the plasmid DNA

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having the Aves-derived HS4 insulator sequence were mixed and transfected into the cells, and the cells were then selected with 80µg/ml of Blasticidine, while the case shown in Fig. 2(c) where pSFVdhfr/d2EGFP and the plasmid DNA having the Aves-derived HS4 insulator sequence were mixed and
5 transfected into the cells had an average fluorescence intensity of 7.14 and an accumulated fluorescence intensity of 312.

This demonstrated that the expression of the target protein could be increased by selecting, with Blasticidine of stepwisely increasing concentrations, the cells in which the gene amplification occurred. Moreover,
10 this showed that the expression of the target protein, d2EGFP, was further increased by performing cotransfection and subsequent coamplification of the target protein-encoding gene and the Aves-derived HS4 insulator sequence, and then selecting the cells with Blasticidine of stepwisely increasing concentrations. In other words, the combination of means for increasing
15 expression of the target proteins makes it possible to attain further higher expression of the target protein.

[Example 3] Effect of Selecting Transformed Cells with Selective
Chemical of Stepwisely Increasing Concentrations

(Method)

20 A plasmid (pSFVdhfr/d2EGFP) was transfected into human colon cancer cell COLO 320DM cells by lipofection. The plasmid

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(pSFVdhfr/d2EGFP) was a plasmid having (a) IR and MAR derived from human DHFR gene locus, (b) Blasticidine tolerant gene, and (c) d2EGFP gene that was under control of TRE-promoter (Tetracycline inducing promoter). The transformed cells were selected with 5µg/ml of Blasticidine
5 (Funakoshi Corp.) thereby obtaining many transformed clones. After the colons were mixed, the culturing was continued. Passaging was carried out subsequently with media having Blasticidine concentrations doubled (from 5µg/ml to 320µg/ml) every passage intervals of 3 to 5 days. Cells select with the respective Blasticidine concentrations were treated with 1µg/ml of
10 Doxycycline (Clontech Laboratories Inc.) added to the media so as to activate TRE-promoter and thereby inducing expression of d2EGFP (whose half-life is much shorter than general EGFP).

(Result)

Fig. 3 shows a result of analysis on the level of the expression of
15 d2EGFP in the cells of each case using cell sorter.

Fig. 3(a) shows a result of the cells selected with 5µg/ml of Blasticidine. Fig. 3(b) shows a result of the cells selected with 10µg/ml of Blasticidine. Fig. 3(c) shows a result of the cells selected with 40µg/ml of Blasticidine. Fig. 3(d) shows a result of the cells selected with 160µg/ml of
20 Blasticidine. Fig. 3(e) shows a result of the cells selected with 320µg/ml of Blasticidine. In Figs. 3, the value in parenthesis is an average of fluorescence

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intensities (i.e., average fluorescence intensity), and boxed value is an accumulated fluorescence intensity.

As shown in Fig. 3(a), the cells select with 5 μ g/ml of Blastcidine had an average fluorescence intensity of 2.94 and an accumulated fluorescence
5 intensity of 222. As shown in Fig. 3(b), the cells select with 10 μ g/ml of Blastcidine had an average fluorescence intensity of 3.77 and an accumulated fluorescence intensity of 322. As shown in Fig. 3(c), the cells select with 40 μ g/ml of Blastcidine had an average fluorescence intensity of 6.67 and an accumulated fluorescence intensity of 640. As shown in Fig.
10 3(d), the cells select with 160 μ g/ml of Blastcidine had an average fluorescence intensity of 9.47 and an accumulated fluorescence intensity of 929. As shown in Fig. 3(e), the cells select with 320 μ g/ml of Blastcidine had an average fluorescence intensity of 18.55 and an accumulated fluorescence intensity of 1850.

15 This demonstrated that the selecting the transformed cells with the stepwisely increasing chemical concentrations can increase the expression of the target protein.

[Example 4] Effect of Promoter for Controlling Expression of Target Protein

20 By lipofection, pTet-ON plasmid (Clontech Laboratories Inc.) having Tet-ON gene encoding a transcription activation factor (Tet-ON protein) for

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activating TRE-promoter was transfected into human cancer COLO 320DM cells. Then, stable transformed cells were selected with hygromycin. Into the cells thus obtained (hereinafter referred to as "Tet-ON cells"), a mixture of a plasmid (pSFVdhfr/d2EGFP) and the plasmid having the Aves-derived HS4
5 insulator sequence was transfected by lipofection. The plasmid (pSFVdhfr/d2EGFP) was a plasmid having (a) IR and MAR derived from human DHFR gene locus, (b) Blasticidine tolerant gene, and (c) d2EGFP gene that was under control of TRE-promoter (Tetracycline inducing promoter). Then, stable transformant was selected with Blasticidine. The
10 plasmid having the Aves-derived HS4 insulator sequence was identical with the one described in Example 2.

Next, by lipofection the pTet-ON plasmid was introduced again into the cells thus obtained. Two days later when transient expression occurred, a level of the expression of d2EGFP was measured.

15 In general, Tet-ON protein (rtTA) is expressed at a low expression level in the stable transformed cells, while the expression is high level of expression in the transient expression. Therefore, the level of the expression of d2EGFP in the stable transformed cells was regarded as a result of normal expression of Tet-ON protein, while the level of the expression of d2EGFP in
20 the case of the transient expression was regarded as a result of high expression of Tet-ON protein.

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(Result)

Figs. 4 illustrate phase contrast microscopic images and fluorescent
microscopic images of the cells of each case. Moreover, Figs. 5 illustrate
results of analysis of the level of the expression of d2EGFP in the cells of
5 each case by using a cell sorter.

Fig. 4(a) is a phase contrast microscopic image (×200) of cells in the
normal expression of Tet-ON protein. Fig. 4(b) is a phase contrast
microscopic image (×200) of cells in the high expression of Tet-ON protein.
Fig. 4(c) is a fluorescent microscopic image (×200) of cells in the normal
10 expression of Tet-ON protein. Fig. 4(d) is a fluorescent microscopic image
(×200) of cells in the high expression of Tet-ON protein.

Moreover, Fig. 5(a) illustrates fluorescence intensity of the normal
expression of Tet-ON protein of the cells in which the plasmid prepared by
transfecting the d2EGFP gene under control of the TRE-promoter into pSFV-
15 V plasmid not having an origin of replication nor a nucleic matrix attachment
region. Fig. 5(b) illustrates fluorescence intensity of the high expression of
Tet-ON protein of the cells in which the plasmid prepared by transfecting the
d2EGFP gene under control of the TRE-promoter into the pSFV-V plasmid.
Fig. 5(c) illustrates fluorescence intensity of the normal expression of Tet-
20 ON protein of the cells in which the Aves-derived HS4 insulator sequence
was amplified simultaneously with pSFVdhfr/d2EGFP1. Fig. 5(d) illustrates

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fluorescence intensity of the high expression of Tet-ON protein of the cells in which the Aves-derived HS4 insulator sequence was amplified simultaneously with pSFVdhfr/d2EGFP1. In Figs. 5, the value in parenthesis is an average of fluorescence intensities (i.e., average fluorescence intensity), and boxed value is an accumulated fluorescence intensity.

From Figs 4(a) to 4(d), it was clear that the fluorescence intensity of the cells with the high expression of Tet-ON protein was higher than that of the stable transformed cells with the normal expression of Tet-ON protein (especially, see Figs. 4(c) and 4(d)).

Moreover, the average fluorescence intensity was 1.02 and the accumulated fluorescence intensity was 0.787 in Fig. 5(a), while the average fluorescence intensity was 1.21 and the accumulated fluorescence intensity was 18.1 in Fig. 5(b). Further, the average fluorescence intensity was 5.80 and the accumulated fluorescence intensity was 532 in Fig. 5(c), while the average fluorescence intensity was 8.15 and the accumulated fluorescence intensity was 766 in Fig. 5(d). This indicates that the fluorescence intensity increased, that is, the level of the expression of the target protein increased in the case of high expression of Tet-ON protein regardless of the copy number of the gene amplification. Further, the fluorescence intensity (Fig. 5(d)) of the cells in which the Aves-derived HS4 insulator sequence was amplified simultaneously with pSFVdhfr/d2EGFP1 and the high expression of Tet-ON

5 [Example 5] Effect of Extrachromosomally Cleaving Out Gene
 Encoding Target Protein in Amplification Region

A plasmid was constructed by removing Hygromycine tolerant gene from the pSFVdhfr/d2EGFP plasmid and embedding synthetic loxP sequence

GCGCGGCCGCATAACTTCGTATAGCATACATTATACGAAGTTATGCG
GCCGCGC-3' (SEQ. ID. NO. 1), by using a well-known DNA synthesizer.

The plasmid thus prepared was transfected into human colon cancer cells COLO 320DM cells by lipofection. The cells thus transformed was selected with 5µg/ml of Blasticidine. The transformed cells thus prepared was transformed with a plasmid having Cre Recombnase gene (kindly provided from Dr. Rolf Springer (Max-Planck-Institut, Germany). see

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"Shimshek, D.R., J. Kim, M.R. Hubner, D.J. Spergel, F. Buchholz, E. Casanova, A.F. Stewart, P.H. Seeburg, and R. Springer. 2002. Codon-improved Cre recombinase (iCre) expression in the mouse. *Genesis*. 32:19-26") and then cultured for 1 week.

5 (Result)

Figs. 6 illustrates phase contrast microscopic images and fluorescence microscopic images of the cells of each case. Fig. 6(a) is a phase contrast microscopic image (×200) of the cells before the transfection of the Cre Recombinase gene. Fig. 6(b) is a phase contrast microscopic image (×200) of
10 cells after the transfection of the Cre Recombinase gene. Fig. 6(c) is a fluorescent microscopic image (×200) of cells before the transfection of the Cre Recombinase gene. Fig. 6(d) is a fluorescent microscopic image (×200) of cells after the transfection of the Cre Recombinase gene.

In Figs. 6(a) to 6(d) it was clearly showed that the fluorescence
15 intensity of the cells after the transfection of the Cre Recombinase gene was higher than before the transfection (see especially, Figs. 6(c) and 6(d)). Therefore, it was demonstrated that the extrachromosomal cleavage of the gene region causing the repeated sequence by the gene amplification makes it possible to express the target protein that has been under expression
20 repression.

[Example 6] Treatment of Cells with 5-aza-2'-deoxycytidine

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(Method)

A mixture of a plasmid (pSFVdhfr/d2EGFP) and the plasmid DNA having the Aves-derived HS4 insulator sequence was transfected to human colon cancer cells COLO 320DM cells by lipofection. The plasmid (pSFVdhfr/d2EGFP) was a plasmid having (a) IR and MAR derived from human DHFR gene locus, (b) Blasticidine tolerant gene, and (c) d2EGFP gene that was under control of TRE-promoter (Tetracycline inducing promoter). The plasmid DNA having the Aves-derived HS4 insulator sequence was identical with the one described in Example 2. From 2 days later since the gene transfection, the cells were selected with 5µg/ml of Blasticidine (Funakoshi Corp.). After 3 to 4 weeks, many colonies of transformed cells were obtained.

Into a culture solution of the transformed cells, of 1µM or 3µM of 5-aza-2'deoxyctidine (hereinafter referred to as "5-aza" where appropriate; Sigma) was added. Thereby, the cells were treated with 5-aza for 3 days. For comparison purposes, 20nM of Trichostatin A (hereinafter referred to as "TSA" where appropriate; histone deacetylation inhibitor, Sigma), which is known as a material for increasing gene expression from the repeated sequence, was added to some culture solutions of the transformed cells and analyzed.

(Result)

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Figs. 7 illustrate phase contrast microscopic images and fluorescence microscopic images of the cases of each case. Figs. 8 illustrate results of analysis of the level of the expression of d2EGFP in the cells of each case by using a cell sorter.

5 Fig. 7(a) illustrates a phase contrast microscopic image ($\times 200$) of the cells before the treatment with 5-aza. Fig. 7(b) illustrates a phase contrast microscopic image ($\times 200$) of the cells after the treatment with 5-aza. Fig. 7(c) illustrates a fluorescence microscopic image ($\times 200$) of the cells before the treatment with 5-aza. Fig. 7(d) illustrates a fluorescence microscopic
10 image ($\times 200$) of the cells after the treatment with 5-aza.

Furthermore, Fig. 8(a) illustrates the fluorescence intensity of the cells before the 5-aza treatment. Fig. 8(b) illustrates the fluorescence intensity of the cells after the treatment with 20nM of TSA. Fig. 8(c) illustrates the fluorescence intensity of the cells after the treatment with 1 μ M of 5-aza. Fig.
15 8(d) illustrates the fluorescence intensity of the cells after the treatment with 1 μ M of 5-aza and 20nM of TSA. Fig. 8(e) illustrates the fluorescence intensity of the cells after the treatment with 3 μ M of 5-aza. Fig. 8(f) illustrates the fluorescence intensity of the cells after the treatment with 3 μ M of 5-aza and 20nM of TSA. In Figs. 8, the value in parenthesis is an average
20 of fluorescence intensities (i.e., average fluorescence intensity), and boxed value is an accumulated fluorescence intensity.

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In Figs. 7(a) to 7(d), it was clearly demonstrated that the fluorescence intensity of the cells after the treatment with 5-aza was higher than that of the cells before the treatment with 5-aza (especially, see Figs 7(c) and 7(d)).

Moreover, the average fluorescence intensity was 4.29 and the
5 accumulated fluorescence intensity was 364 in the cells before the treatment with 5-aza illustrated in Fig. 8(a), while the average fluorescence intensity was 9.48 and the accumulated fluorescence intensity was 895 in the cells after the treatment with 1 μ M of 5-aza illustrated in Fig. 8(c). This showed that the expression of d2EGFP, which was the target protein, was increased
10 by treating the cells with 5-aza. As illustrated in Fig. 8(e), the cells after the treatment with 3 μ M of 5-aza showed the average fluorescence intensity of 10.93 and the accumulated fluorescence intensity of 1040 which were higher than in the cells after the treatment with 1 μ M of 5-aza. This demonstrated that the treatment with 5-aza of higher concentration further increases the
15 expression of the target protein.

On the other hand, the average fluorescence intensity was 3.02 and the accumulated fluorescence intensity was 228 in the cells after the treatment with 20nM of TSA illustrated in Fig. 8(b), while the average fluorescence intensity was 4.29 and the accumulated fluorescence intensity was 364 in the
20 cells before the treatment with 5-aza illustrated in Fig. 8(a). This showed that the treatment with 20nM of TSA cannot increase the expression of the target

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protein. Moreover, the average fluorescence intensity was 9.40 and the accumulated fluorescence intensity was 878 in the cells after the treatment with 1 μ M of 5-aza and 20nM of TSA illustrated in Fig. 8(d), while the average fluorescence intensity was 9.48 and the accumulated fluorescence intensity was 895 in the cells after the treatment with 1 μ M of 5-aza illustrated in Fig. 8(c). Further, the average fluorescence intensity was 11.09 and the accumulated fluorescence intensity was 1050 in the cells after the treatment with 3 μ M of 5-aza and 20nM of TSA illustrated in Fig. 8(f), while the cells after the treatment with 3 μ M of 5-aza showed the average fluorescence intensity of 10.93 and the accumulated fluorescence intensity of 1040 illustrated in Fig. 8(e). This explained that the combination of 5-aza and TSA cannot increase the expression of the target protein.

[Example 7] Effect of Selecting Clone in which Gene Amplification Occurred on DM

(Method)

By lipofection, the pTet-ON plasmid (Clontech Laboratories Inc.) having Tet-ON gene encoding a transcription activating factor (Tet-ON protein) for promoting TRE-promoter was transfected into human colon cancer COLO 320DM. Transformed cells (hereinafter referred to as "Tet-ON cells") which stably expressed Tet-ON protein, were selected with

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Hygromycine.

Next, the pSFVdhfr/d2EGFP plasmid was transfected into the Tet-ON cells by lipofection. The cells thus obtained was selected with 5µg/ml of Blasticidine for approximately 3 weeks, thereby obtaining a polyclonal population of the stable transformed cells.

The expression of d2EGFP was induced in the polyclonal population by activating TRE-promoter by adding 5µg/ml of Doxycycline (Clontech Laboratories Inc.) in the medium. Fig. 9 illustrates a result of analysis of the level of the expression of d2EGFP in this case by using a cell sorter. In Fig. 9, the value in parenthesis is an average of fluorescence intensities (i.e., average fluorescence intensity), and boxed value is an accumulated fluorescence intensity. Based on the result of Fig. 9, it was confirmed that the polyclonal population was a population for expressing d2EGFP (average fluorescence intensity: 6.25, accumulated fluorescence intensity: 536).

From the polyclonal population, given 30 colons were obtained by a limiting dilution method that was carried out by inoculating a clone to 6 well plates with such a low cell concentration of 30 cells/ml, and incubating the plates for 10 days to 2 weeks thereby obtaining colonies thereon.

From among the clones, 4 clones having high levels of expression of d2EGFP (Clone 4, Clone 5, Clone 6, and Clone 9) were selected by fluorescence microscopic observation. Mitotic chromosome samples of the 4

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clones were prepared. By FISH method, detection for a base sequence derived from the transfected pSFVdhfr/d2EGFP plasmid was carried out. The preparation of mitotic chromosome samples and the FISH method were carried out as described in Shimizu, N., Kanda, T., and Wahl, G. M. Selective
5 capture of acentric fragments by micronuclei provides a rapid method of purifying extrachromosomally amplified DNA. *Nature Genet.*, 12: 65-71, 1996.

Figs. 10 illustrates the result of the detection of the base sequence derived from the pSFVdhfr/d2EGFP plasmid, by FISH method. Fig. 10(a)
10 shows the result of clone 4. Fig. 10(b) shows the result of clone 5. Fig. 10(c) shows the result of clone 6. Fig. 10(d) shows the result of clone 9. The arrows in Figs. 10 are pointing the base sequence derived from the pSFVdhfr/d2EGFP plasmid.

The results shown in Figs. 10 showed that the base sequence derived
15 from the pSFVdhfr/d2EGFP plasmid was amplified on HSR in clones 4, 5, and 6, but the base sequence was amplified on DM in clone 9.

For 4 clones, induction with Doxycycline (Clontech Laboratories Inc.; 1µg/ml) was carried out and 5-aza (0µM, 1µM or 3µM) was added in the medium. After 1 week culturing, the level of expression of d2EGFP in each
20 clone was analyzed by using a cell sorter.

The results are illustrated in Figs. 11, 12, 13, and 14. Figs. 11

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illustrates the result of clone 4. Fig. 11(a) illustrates a result of a case where 5-aza was not added (0 μ M). Fig. 11(b) illustrates a result of a case where 1 μ M of 5-aza was added (1 μ M). Fig. 11(c) illustrates a result of a case where 3 μ M of 5-aza was added (3 μ M).

5 Figs. 12 illustrates the result of clone 5. Fig. 12(a) illustrates a result of a case where 5-aza was not added (0 μ M). Fig. 12(b) illustrates a result of a case where 1 μ M of 5-aza was added (1 μ M). Fig. 12(c) illustrates a result of a case where 3 μ M of 5-aza was added (3 μ M).

10 Figs. 13 illustrates the result of clone 6. Fig. 13(a) illustrates a result of a case where 5-aza was not added (0 μ M). Fig. 13(b) illustrates a result of a case where 1 μ M of 5-aza was added (1 μ M). Fig. 13(c) illustrates a result of a case where 3 μ M of 5-aza was added (3 μ M).

15 Figs. 14 illustrates the result of clone 9. Fig. 14(a) illustrates a result of a case where 5-aza was not added (0 μ M). Fig. 14(b) illustrates a result of a case where 1 μ M of 5-aza was added (1 μ M). Fig. 14(c) illustrates a result of a case where 3 μ M of 5-aza was added (3 μ M).

In Figs. 11 to 13, the value in parenthesis is an average of fluorescence intensities (i.e., average fluorescence intensity), and boxed value is an accumulated fluorescence intensity.

20 Figs. 11 illustrate clone 4 as follows: an average fluorescence intensity and an accumulate fluorescence intensity indicating a level of

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expression in the case of no addition of 5-aza in the medium (0 μ M) were 1.89 and 90.0 respectively; an average fluorescence intensity and an accumulate fluorescence intensity in the case of the addition of 1 μ M of 5-aza in the medium were 5.46 and 453 respectively; and an average fluorescence intensity and an accumulate fluorescence intensity in the case of the addition of 3 μ M of 5-aza in the medium were 23.76 and 2300 respectively.

Figs. 12 illustrate clone 5 as follows: an average fluorescence intensity and an accumulate fluorescence intensity indicating a level of expression in the case of no addition of 5-aza in the medium (0 μ M) were 1.02 and 1.11 respectively; an average fluorescence intensity and an accumulate fluorescence intensity in the case of the addition of 1 μ M of 5-aza in the medium were 2.23 and 126 respectively; and an average fluorescence intensity and an accumulate fluorescence intensity in the case of the addition of 3 μ M of 5-aza in the medium were 10.44 and 962 respectively.

Figs. 13 illustrate clone 6 as follows: an average fluorescence intensity and an accumulate fluorescence intensity indicating a level of expression in the case of no addition of 5-aza in the medium (0 μ M) were 1.16 and 8.84 respectively; an average fluorescence intensity and an accumulate fluorescence intensity in the case of the addition of 1 μ M of 5-aza in the medium were 2.09 and 111 respectively; and an average fluorescence intensity and an accumulate fluorescence intensity in the case of the addition

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of 3 μ M of 5-aza in the medium were 10.87 and 1000 respectively.

Figs. 14 illustrate clone 9 as follows: an average fluorescence intensity and an accumulate fluorescence intensity indicating a level of expression in the case of no addition of 5-aza in the medium (0 μ M) were
5 41.46 and 4130 respectively; an average fluorescence intensity and an accumulate fluorescence intensity in the case of the addition of 1 μ M of 5-aza in the medium were 131.86 and 13200 respectively; and an average fluorescence intensity and an accumulate fluorescence intensity in the case of the addition of 3 μ M of 5-aza in the medium were 248.60 and 24900
10 respectively.

From the results illustrated in Figs. 11 to 14, it was demonstrated that the level of the expression of d2EGFP was very low in clones (clone 4, clone 5, and clone 6) in which the base sequence derived from the pSFVdhfr/d2EGFP plasmid was amplified on HSR, but that this level of the
15 expression of d2EGFP could be increased by adding 5-aza to the medium. On the other hand, it was found that the addition of 5-aza in the medium was not necessary to attain a high level of the expression of d2EGFP (average fluorescence intensity: 41.46, accumulate fluorescence intensity: 4130) in the clone (clone 9) in which the base sequence derived from the
20 pSFVdhfr/d2EGFP plasmid was amplified on DM, but that the addition of 5-aza in the medium dramatically increased the level of expression in such a

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clone (average fluorescence intensity: 131.86 or 248.60, accumulate
fluorescence intensity: 13200 or 24900).

Therefore, it was shown that a high level of expression of a target
protein can be attained by selecting a clone in which gene amplification
5 occurs on DM.

[Example 8] Effect of Cotransfection to attaining High Expression of
Transcription Activating Factor for Activation Promoter for Controlling
Expression of Target Protein.

(Method and Result)

10 An equiamout mixture of a plasmid (pSFVdhfr/d2EGFP) and pTet-ON
plasmid (Clontech Laboratories Inc.) was simultaneously transfected into
human colon cancer COLO 320DM cells. The plasmid (pSFVdhfr/d2EGFP)
was a plasmid having (a) IR and MAR derived from human DHFR gene
locus, (b) Blasticidine tolerant gene, and (c) d2EGFP gene that was under
15 control of TRE-promoter (Tetracycline inducing promoter). Meanwhile, the
pTet-ON plasmid was a plasmid having Tet-ON gene encoding the
transcription activating factor (Tet-ON protein) of TRE-promoter. The cells
were selected with 5 μ g/ml of Blasticidine thereby obtaining a polyclonal
population of transformed cells. TRE-promoter was activated by adding
20 1 μ g/ml of Doxycycline (Clontech Laboratories Inc.) in the medium of a cell
culturing solution of the polyclonal population, thereby inducing the

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expression of d2EGFP.

Fig. 15(a) illustrates a phase contrast microscopic image ($\times 200$) of the polyclonal population before the addition of Doxycycline, while Fig. 15(b) illustrates a fluorescence microscopic image ($\times 200$) thereof. Fig. 15(c) illustrates a phase contrast microscopic image ($\times 200$) of the polyclonal population after the addition of Doxycycline, while Fig. 15(d) illustrates a fluorescence microscopic image ($\times 200$) thereof.

According to Figs. 15(a) to (d), fluorescence intensity of d2EGFP was very weak before the addition of Doxycycline, but the fluorescence intensity of d2EGFP was increased by the addition of Doxycycline. Thus, it was confirmed that the addition of Doxycycline (Clontech Laboratories Inc.) in the medium activated TRE-promoter and thereby inducing the expression of d2EGFP in the polyclonal population of the transformed cells in which pSFVdhfr/d2EGFP and the pTet-ON plasmid were simultaneously transfected.

Next, after the expression of d2EGFP was induced by Doxycycline, the level of the expression of d2EGFP was analyzed by using a cell sorter so as to find a change in the level of the expression of d2EGFP over time. The results are shown in Figs. 16. Fig. 16(a) shows the reading before the addition of Doxycycline. Fig. 16(b) shows the reading 1 hour after the addition of Doxycycline. Fig. 16(c) shows the reading 3 hours after the

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addition of Doxycycline. Fig. 16(d) shows the reading 6 hours after the
addition of Doxycycline. Fig. 16(e) shows the reading 15 hours after the
addition of Doxycycline. Fig. 16(f) shows the reading 24 hours (1 day) after
the addition of Doxycycline. Fig. 16(g) shows the reading 48 hours (2 days)
5 after the addition of Doxycycline. Fig. 16(h) shows the reading 120 hours (5
days) after the addition of Doxycycline. Fig. 16(i) shows the reading
approximately two weeks from the addition of Doxycycline. In Figs. 16, the
value in parenthesis is an average of fluorescence intensities (i.e., average
fluorescence intensity), and boxed value is an accumulated fluorescence
10 intensity.

Fig. 17 illustrates a graph plotting the averages (i.e., average
fluorescence intensities: indicated by filled circles in Fig. 17) and living cell
ratio (a ratio of living cells over total cell number: indicated by open circles)
of the level of the expression of d2EGFP against time. According to Fig. 17,
15 the polyclonal population thus obtained showed very low level of the
expression of d2EGFP before the addition of Doxycycline, the level of the
expression was peaked in 1 to 2 days after the addition of Doxycycline. The
level was maintained high even after two weeks lapsed (labeled as "2wk" in
Fig. 17). Moreover, in the polyclonal population, no decrease in the survival
20 ratio of the cells due to the expression of d2EGFP induced by Doxycycline
and the cells were proliferated normally expressing d2EGFP at a high level.

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The result of analysis on expression of d2EGFP of a negative control by using a cell sorter is shown in Fig. 16(j). The negative control was prepared as follows. Into the pSFV-V plasmid having no origin of replication and no nucleic matrix attachment region, the d2EGFP gene under control of the TRE-promoter was embedded thereby constructing a plasmid. The plasmid thus constructed was transfected in Tet-ON cells thereby obtaining a polyclonal population of transformed cells. In the polyclonal population, the expression of d2EGFP was induced by Doxycycline. According to Fig. 16(j), the negative control had a level of expression of d2EGFP which had an average fluorescence intensity of 1.02 and accumulated fluorescence intensity of 0.787.

Meanwhile, an average fluorescence intensity was 2.99 and accumulated fluorescence intensity was 216 one hour after the addition of Doxycycline (Fig. 16(b)). An average fluorescence intensity was 3.72 and accumulated fluorescence intensity was 293 three hours after the addition of Doxycycline (Fig. 16(c)). An average fluorescence intensity was 4.22 and accumulated fluorescence intensity was 341 six hours after the addition of Doxycycline (Fig. 16(d)). An average fluorescence intensity was 9.24 and accumulated fluorescence intensity was 841 fifteen hours after the addition of Doxycycline (Fig. 16(e)). An average fluorescence intensity was 28.77 and accumulated fluorescence intensity was 2800 twenty four hours after the

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addition of Doxycycline (Fig. 16(f)). An average fluorescence intensity was 53.97 and accumulated fluorescence intensity was 5320 forty eight hours (2 days) after the addition of Doxycycline (Fig. 16(g)). An average fluorescence intensity was 39.56 and accumulated fluorescence intensity was 3880 one
5 hundred twenty hours (5 days) after the addition of Doxycycline (Fig. 16(h)). An average fluorescence intensity was 35.32 and accumulated fluorescence intensity was 3500 two weeks after the addition of Doxycycline (Fig. 16(i)).

From these results, it was found that the polyclonal populations obtained by the method according to the present Example showed a very high
10 level of expression after the induction of the expression, and the level was approximately 300 times to 7000 times greater than the negative control in terms of the accumulated fluorescence intensity.

Furthermore, given 5 clones selected from among the polyclonal population thus obtained by the method according to the present Example
15 were analyzed by a cell sorter so as to find the level of the expression of d2EGFP after the expression was induced. Typical examples of the results (three clones) are shown in Figs. 18. Figs. 18(a) and 18(b) show the results of clone A, where Fig. 18(a) shows the result before the induction of the expression and Fig. 18(b) shows the result after the induction of the
20 expression. Figs. 18(c) and 18(d) show the results of clone B, where Fig. 18(c) shows the result before the induction of the expression and Fig. 18(d)

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shows the result after the induction of the expression. Figs. 18(e) and 18(f) show the results of clone C, where Fig. 18(e) shows the result before the induction of the expression and Fig. 18(f) shows the result after the induction of the expression. In Figs. 18, the value in parenthesis is an average of
5 fluorescence intensities (i.e., average fluorescence intensity), and boxed value is an accumulated fluorescence intensity.

From Figs. 18, it was understood that it was possible to easily attain a clone having a very high level of expression of the target protein (d2EGFP) from the polyclonal population obtained by the method according to the
10 present Example.

Furthermore, the method according to the present Example is expected to attain a high expression of the target protein (d2EGFP) from the amplified gene on HSR. This can be confirmed by the following experiment: A mixture of pSFVdhfr and pECMS2Beta was cotransfected to human colon cancer
15 COLO 320DM cells by lipofection, where pSFVdhfr was a plasmid having an origin of replication and a nucleic matrix attachment region, and pECMS2Beta was a plasmid having, in the downstream of LacO (lactose operator)-repeat sequence and TRE-promoter, an MS2 junction sequence (pECMS2Beta was kindly provided from Dr. Susan Janicki and Dr. David
20 Spector (cold Spring Harbor Laboratory); see "Janicki, S.M., T. Tsukamoto, S.E. Salghetti, W.P. Tansey, R. Sachidanandam, K.V. Prasanth, T. Ried, Y.

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Shav-Tal, E. Bertrand, R.H. Singer, and D.L. Spector. 2004. From silencing to gene expression: real-time analysis in single cells. *Cell*. 116:683-698.”).

The human colon cancer COLO 320DM cells used here were cloned cells which became Neomycine tolerant by transfecting the pLacR-CFP
5 plasmid by lipofection in advance, where the pLacR-CFP plasmid was a plasmid that expressed a fused protein of LacR (lactose repressor) and CFP (cyan fluorescence protein), and was kindly provided from Dr. Susan Janicki and Dr. David Spector (cold Spring Harbor Laboratory); see “Janicki, S.M., T. Tsukamoto, S.E. Salghetti, W.P. Tansey, R. Sachidanandam, K.V.
10 Prasanth, T. Ried, Y. Shav-Tal, E. Bertrand, R.H. Singer, and D.L. Spector. 2004. From silencing to gene expression: real-time analysis in single cells. *Cell*. 116:683-698.”

As a result, a clone in which HSR was formed by coamplification of both the plasmids. In the clone, HSR showed cyan fluorescence because
15 LacR-CFP is bound with the LacO sequence in HSR.

Moreover, the pTet-ON plasmid (Clontech Laboratories Inc.) and a plasmid for expressing a fused protein of MS2 and YFP (yellow fluorescecnce protein) (which was kindly provided from Dr. Susan Janicki and Dr. David Spector (cold Spring Harbor Laboratory); see “Janicki, S.M.,
20 T. Tsukamoto, S.E. Salghetti, W.P. Tansey, R. Sachidanandam, K.V. Prasanth, T. Ried, Y. Shav-Tal, E. Bertrand, R.H. Singer, and D.L. Spector.

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2004. From silencing to gene expression: real-time analysis in single cells.
Cell. 116:683-698.") were simultaneously transfected in the clone by the
electroporation. After 2.5 hours, 1µg/ml of Doxycycline (Clontech
Laboratories Inc.) was added to the medium. At this stage, the expressed Tet-
5 ON protein and Doxycycline activated the TRE-promoter in HSR, thereby
causing transcription of RNA for encoding the fused protein of MS2 and YFP
(yellow fluorescence protein). This RNA has a MS2 junctional sequence.
Because of this, the fused protein of MS2 and YFP is bound thereto thereby
showing yellow fluorescence.

10 A typical photographs showing the result are shown in Figs. 19. In
Figs. 19, A, B and C are fluorescence microscopic images (labeled as "-Dox"
on the left) of the clones before the induction of the expression by
Doxycycline, while D and E are fluorescence microscopic images (labeled as
"+Dox 1.5hr" on the left) of the clones 1.5 hours after the induction of the
15 expression by Doxycycline. Moreover, the images labeled as "LaminB" on
the top are images in which nuclear lamina is visualized by indirect
fluorescent antibody technique using anti-Lamin B antibody (Santa Cruz
Biotechnologies Inc.). The images labeled as "CFP-HSR" on the top are
images in which HSR is visualized using cyan fluorescence protein. The
20 images labeled as "YFP-MS2 RNA" on the top are images in which RNA is
visualized using YFP (yellow fluorescence protein). The images labeled as

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"Merge" on the top are images in which the three images are overlapped.

Before the induction of the expression with Doxycycline, HSR (indicated by the arrows in Figs. 19) visualized with cyan color of CFP (cyan fluorescence protein) was heterochromatized and thus having small and
5 shrunk spherical structures. It was clearly observed that the induction of the expression with Doxycycline caused HSR in cyan color to be loosened up largely and RNA (indicated by being circled) visualized in yellow fluorescence was coming out therefrom.

These results showed that the activation of the TRE-promoter with the
10 Tet-ON protein and Doxycycline looses heterochromatized HSR formed by the plasmid having IR and MAR, thereby activating the RNA transcription.

[Example 9] Effect of Cotransfection for High Expression of Transcription Activation Factor of Promoter for Controlling Expression of Target Protein, and Treating Cells with 5-aza-2'-deoxycytidine.

15 (Method and Result)

An equiamout mixture of a plasmid (pSFVdhfr/d2EGFP) and pTet-ON plasmid (Clontech Laboratories Inc.) was simultaneously transfected into human colon cancer COLO 320DM cells. The plasmid (pSFVdhfr/d2EGFP) was a plasmid having (a) IR and MAR derived from human DHFR gene
20 locus, (b) Blasticidine tolerant gene, and (c) d2EGFP gene that was under control of TRE-promoter (Tetracycline inducing promoter). Meanwhile, the

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pTet-ON plasmid was a plasmid having Tet-ON gene encoding the transcription activating factor (Tet-ON protein) of TRE-promoter. The cells were selected with 5µg/ml of Blasticidine thereby obtaining a polyclonal population of transformed cells. From the polyclonal population, given 10
5 clones were selected by limiting dilution. Then, from among the 10 clones a clone in which a base sequence derived from the pSFVdhfr/d2EGFP plasmid was amplified on HSR (this clone is referred to as "HSR clone" hereinafter for easy explanation), and a clone in which the base sequence is amplified on DM (this clone is referred to as "DM clone" hereinafter for easy explanation)
10 were screened. Figs. 20(a) and 20(b) respectively illustrate the results of detection of the base sequence derived from the transfected pSFVdhfr/d2EGFP plasmid for HSR clone and DM clone. In Figs. 20, the detected base sequence derived from the pSFVdhfr/d2EGFP plasmid is indicated by the arrow. The limiting dilution and the screening was carried
15 out as described in Example 7.

In each culture solution of the two clones, 3µM of 5-aza was added and thereby the clones were treated with 5-aza for 3 days. Then, Doxycycline (Clontech Laboratories Inc.; 1µg/ml) was added in the culture solutions and thereby the expression of d2EGFP was induced for 2 days. After the
20 culturing, the level of the expression of d2EGFP in each clone was analyzed by a cell sorter. Moreover, as a negative control a polyclonal population of

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transformed cells was prepared in the same way except that pSFV-V plasmid having no origin of replication and no nucleic matrix attachment region was used in replacement of pSFVdhfr/d2EGFP. This polyclonal population of transformed cells was subjected to the induction of expression of d2EGFP with Doxycycline and a level of the expression of d2EGFP therein was analyzed by using the cell sorter in the same way as described above. The results are illustrated in Figs. 21(a) to 21(c). Fig. 21(a) illustrates the result of the fluorescence intensity measurements of HSR clone two days after the addition of Doxycycline. Fig. 21(b) illustrates the result of the fluorescence intensity measurements of DM clone (not treated with 5-aza) two days after the addition of Doxycycline. Fig. 21(c) illustrates the result of the fluorescence intensity measurements of DM clone (treated with 5-aza) two days after the addition of Doxycycline. Figs. 21, the value in parenthesis is an average of fluorescence intensities (i.e., average fluorescence intensity), and boxed value is an accumulated fluorescence intensity. Moreover, Fig. 21(d) is a bar graph showing the accumulated fluorescence intensities two days after the addition of Doxycycline for the negative control, HSR clone, and DM clones (treated and not treated with 5-aza).

According to Fig. 21(d), the accumulated fluorescence intensity of the negative control (labeled as "N" in Fig. 21(d)) was 1826, the accumulated fluorescence intensity of the HSR clone (labeled as "HSR" in Fig. 21(d)) was

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22119, the accumulated fluorescence intensity of the DM clone (not treated with 5-aza; labeled as "DM(-)" in Fig. 21(d)) was 75099, and the accumulated fluorescence intensity of DM clone (treated with 5-aza; labeled as "DM(+)" in Fig. 21(d)) was 238113. This showed that the HSR clone and
5 DM clones had significantly higher levels of the expression of d2EGFP than the negative control clone, and that the 5-aza treatment improved the level of the expression of d2EGFP.

[Example 10] Effect of the Present Invention in CHO Cells.

(Method and Result)

10 The present embodiment was carried out in the same way as described in Example 9, except that the cells to which the plasmids were transfected were CHO-K1 cells (kindly provided from Cell Research Center for Biomedical Research, Institute of Development, Aging and Cancer, Tohoku University.

15 As a result, a clone in which a base sequence derived from the pSFVdhfr/d2EGFP plasmid was amplified on HSR (this clone is referred to as "HSR clone" hereinafter for easy explanation), and a clone in which the base sequence is amplified on DM (this clone is referred to as "DM clone" hereinafter for easy explanation) were successfully isolated from the CHO
20 cells. This proved that the present invention is applicable to cells other than tumor cells such as COLO 320 DM cells. Fig. 22(a) illustrates the result of

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the detection of the base sequence of the transfected pSFVdhfr/d2EGFP plasmid in the isolated HSR clone of the CHO cells by the FISH method. Fig. 22(b) illustrates the result of the detection of the DM clone of the CHO cells. The detected base sequence of the transfected pSFVdhfr/d2EGFP plasmid is
5 indicated by the arrow in Figs. 22.

Fig. 23(a) illustrates the result of fluorescence intensity measurement of that clone two days after the addition of Doxycycline, which had the highest level of the expression of d2EGFP according to the cell sorter (this clone is referred to as "max clone" hereinafter for easy explanation).
10 Moreover, Fig. 23(b) illustrates accumulated fluorescence intensities (2 days after the addition of Doxycycline) of a polyclonal population of transformed cells in a negative control, a polyclonal population of transformed cells in the present embodiment, and the max clone.

According to Fig. 23(b), the polyclonal population (labeled as "P" in
15 Fig. 23(b)) of the transformed cells in the present embodiment had the accumulated fluorescence intensity of 14022, and the max clone (labeled as "M" in Fig. 23(b)) of the transformed cells in the present embodiment had the accumulated fluorescence intensity of 167865, while the polyclonal population (labeled as "NP" in Fig. 23(b)) of the transformed cells in the
20 negative control had the accumulated fluorescence intensity of 296. This demonstrated that the polyclonal population of the transformed cells in the

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present embodiment and the max clone had significantly higher levels of the expression of d2EGFP than the polyclonal population of the transformed cells in the negative control. Especially, the max clone isolated from the polyclonal population had further higher levels of the expression of d2EGFP
5 than the negative control.

Fig. 23(c) illustrates the result of detection of the base sequence of the transfected pSFVdhfr/d2EGFP plasmid in the max clone by the FISH method. The detected base sequence of the pSFVdhfr/d2EGFP plasmid is indicated by the arrow in Fig. 23(c). From Fig. 23(c), it can be understood that the max
10 clone is a clone in which the base sequence derived the pSFVdhfr/d2EGFP plasmid was amplified on HSR (relatively short HSR).

INDUSTRIAL APPLICABILITY

As described above, a method and means for releasing transcription repression caused by a repeated gene sequence are provided by a method and
15 a kit according to the present invention. As a result, mass production of useful protein by gene amplification can be done by the method and the kit according to the present invention. Therefore, the present invention is applicable to protein-producing industries such as pharmaceutical, chemical, food, cosmetic, textile industries, etc.

CLAIMS

1. A method of expressing a target protein under expression repression from a repeated sequence formed in mammalian cells in which gene amplification is induced by transfecting a first polynucleotide and a second polynucleotide simultaneously into the mammalian cells *in vitro*, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the second polynucleotide encodes a target protein to be expressed, the method comprising transfecting into the mammalian cells *in vitro*:

- (i) the first polynucleotide including an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells;
- (ii) a polynucleotide obtained by controllably linking a promoter region with the second polynucleotide encoding a protein to be expressed; and
- (iii) a polynucleotide encoding a transcription activation factor of the promoter,

wherein the first polynucleotide of (i), the polynucleotide of (ii), and the polynucleotide of (iii) may be transfected on the same or on separate nucleic acid molecules.

2. The method as set forth in claim 1 wherein:
the polynucleotide of (ii) includes a chemical tolerant gene, and
the method further comprises:
culturing the mammalian cells sequentially in a medium of increasing concentrations of a chemical, the chemical tolerant gene being tolerant to the chemical.

3. The method as set forth in claim 1, comprising:
selecting mammalian cells that the gene amplification occurs on a double minute chromosome.

4. The method as set forth in claim 1, comprising:

treating the mammalian cells with 5-aza-2'-deoxycytidine.

5. The method as set forth in claim 1, wherein the origin of replication is derived from an origin of replication of c-myc locus, dihydrofolate reductase locus, or β -globin locus.

6. The method as set forth in claim 1, wherein the nucleic matrix attachment region is derived from a nucleic matrix attachment region of Igk locus, SV40 initial region, or dihydrofolate reductase locus.

7. A kit for expressing a target protein under expression repression from a repeated sequence formed in mammalian cells in which gene amplification is induced by transfecting a first polynucleotide and a second polynucleotide simultaneously into the mammalian cells, where the first polynucleotide includes an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells, and the second polynucleotide encodes a target protein to be expressed, the kit comprising:

- (i) the first polynucleotide including an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells,
- (ii) a polynucleotide obtained by controllably linking a promoter region with the second polynucleotide encoding a protein to be expressed;
- (iii) a polynucleotide encoding a transcription activation factor of the promoter;

8. The kit as set forth in claim 7, the kit further comprising 5-aza-2'-deoxycytidine.

9. The kit as set forth in claim 7, wherein the origin of replication is derived from an origin of replication of c-myc locus, dihydrofolate reductase locus, or β -globin locus.

10. The kit as set forth in claim 7, wherein the nucleic matrix attachment region is derived from a nucleic matrix attachment region of Igk locus, SV40 initial region, or dihydrofolate reductase locus.

11. A transformant of a mammalian cell produced *in vitro* from cells in culture prepared by inserting into mammalian cells *in vitro*:

- (i) a first polynucleotide including an origin of replication and a nucleic matrix attachment region that function in eukaryotic cells;
- (ii) a polynucleotide obtained by controllably linking a promoter region with a second polynucleotide encoding a protein to be expressed; and
- (iii) a polynucleotide encoding a transcription activation factor of the promoter,

the target protein being expressed under no expression repression from a repeated sequence formed by gene amplification.

12. The transformant as set forth in claim 11, wherein the origin of replication is derived from an origin of replication of c-myc locus, dihydrofolate reductase locus, or β -globin locus.

13. The transformant as set forth in claim 11, wherein the nucleic matrix attachment region is derived from a nucleic matrix attachment region of Igk locus, SV40 initial region, or dihydrofolate reductase locus.

14. The transformant as set forth in claim 11, wherein the mammalian cells are cells selected from the group consisting of COLO 320DM cells, COLO 320HSR cells, HeLa cells, and CHO cells.

Application number/numéro de demande: 2604398

Figures: 4a, 4b, 4c, 4d, 6a, 6b, 6c, 6d, 7a, 7b, 7c, 7d, 10b,
10c, 19, 20a, 20b, 22a, 22b, 23c,
10a, 10d, 15

Pages: 7/46 - 10/46, 13/46 - 20/46, 24/46, 40/46 - 41/46
44/46, 46/46

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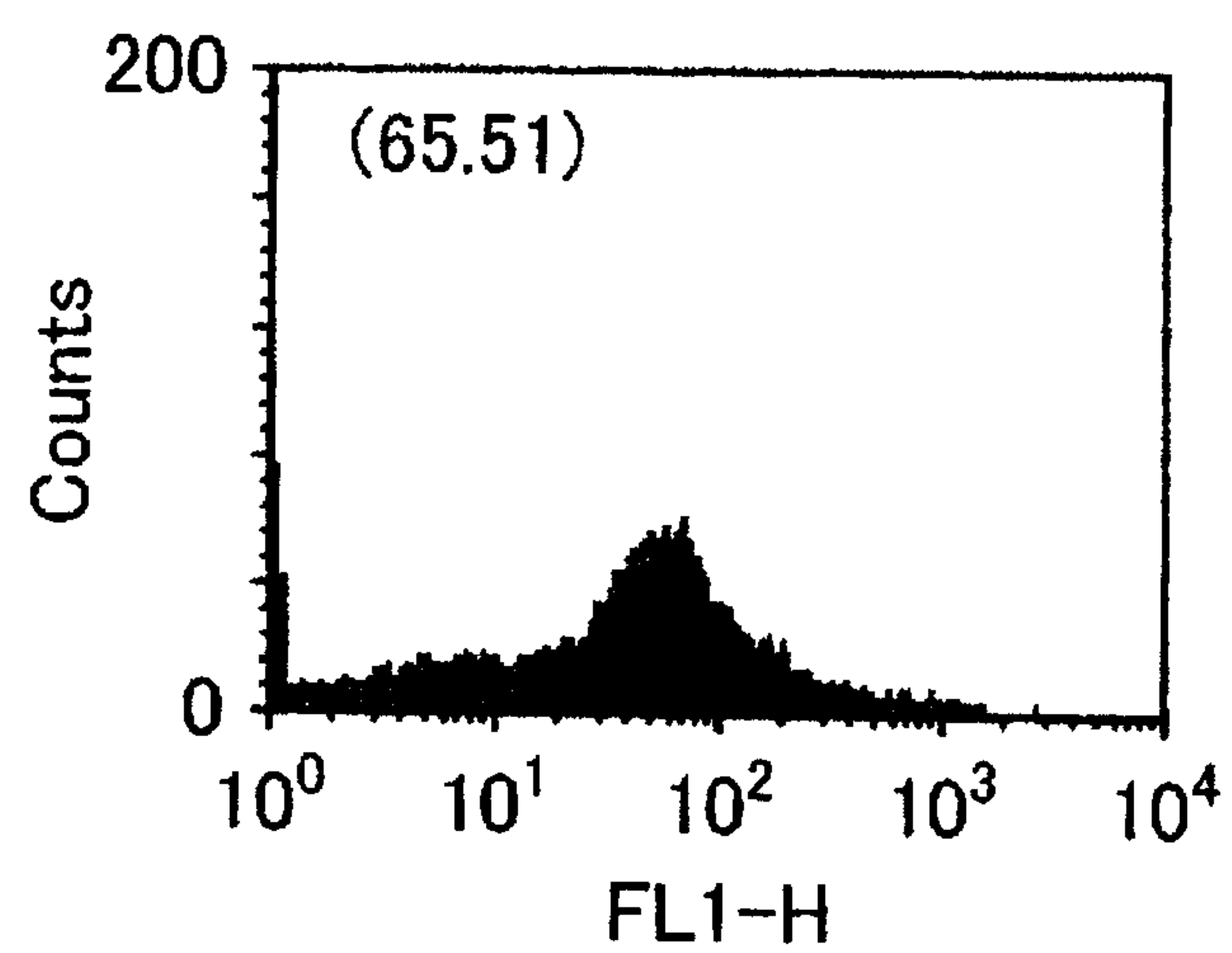
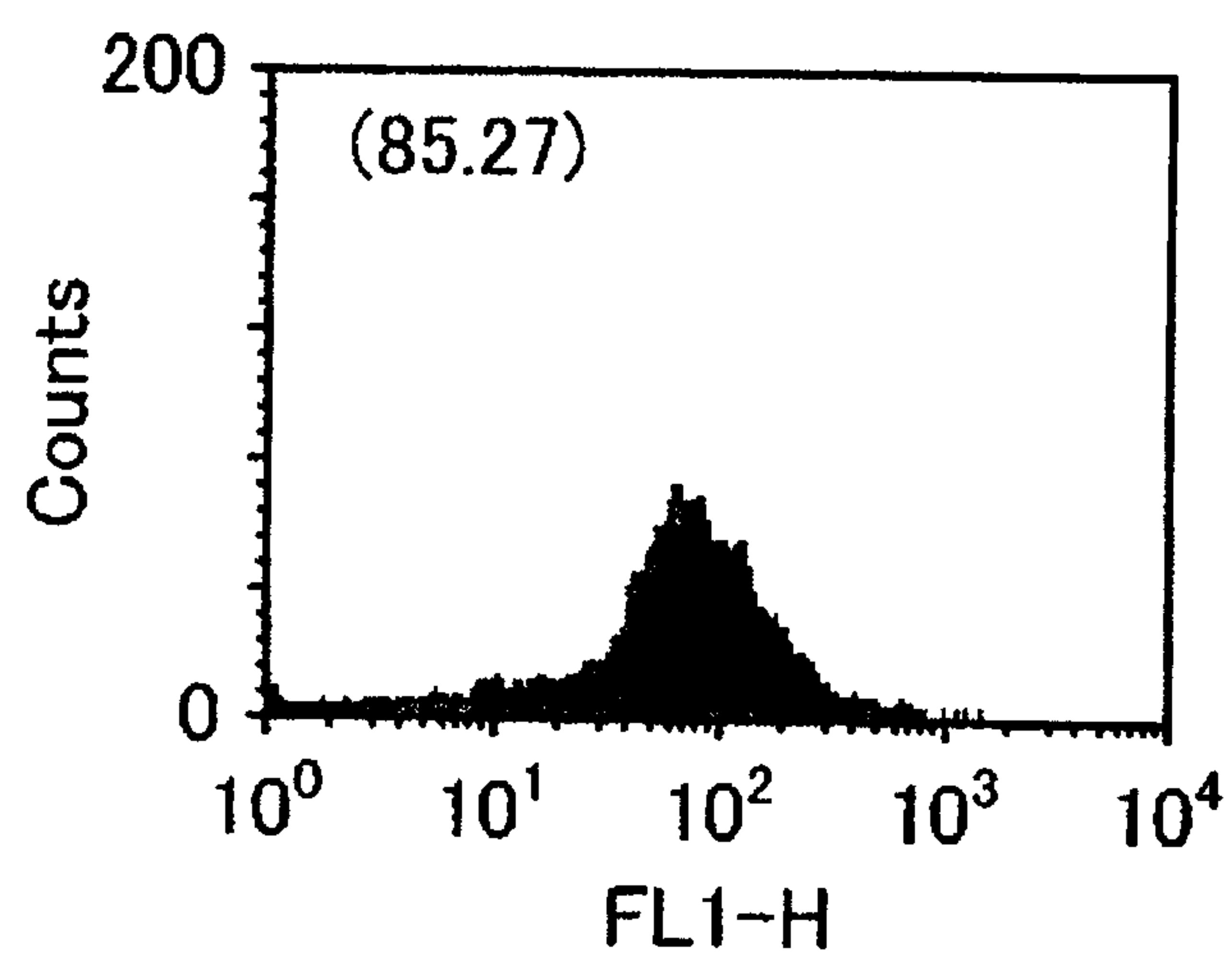
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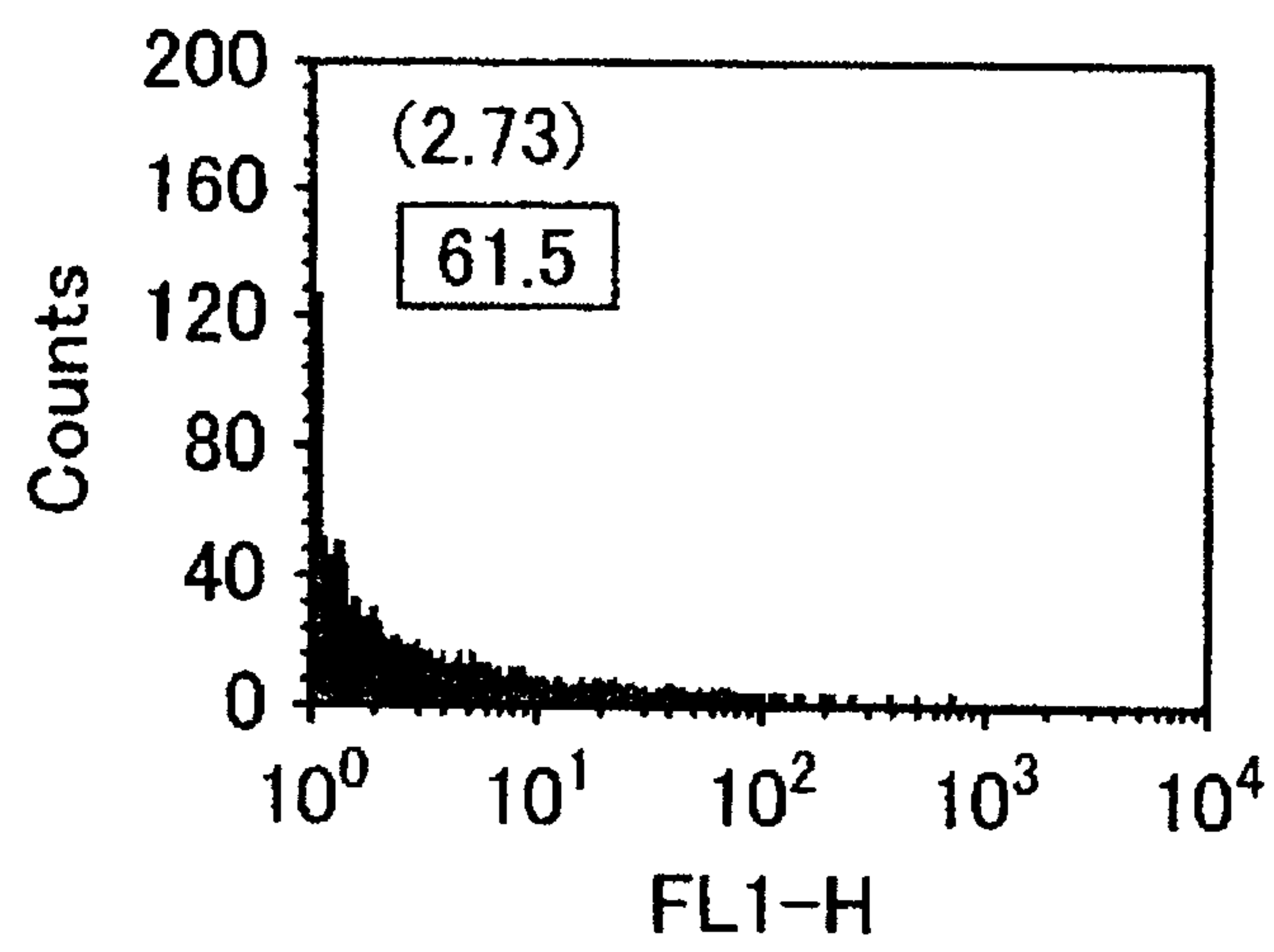
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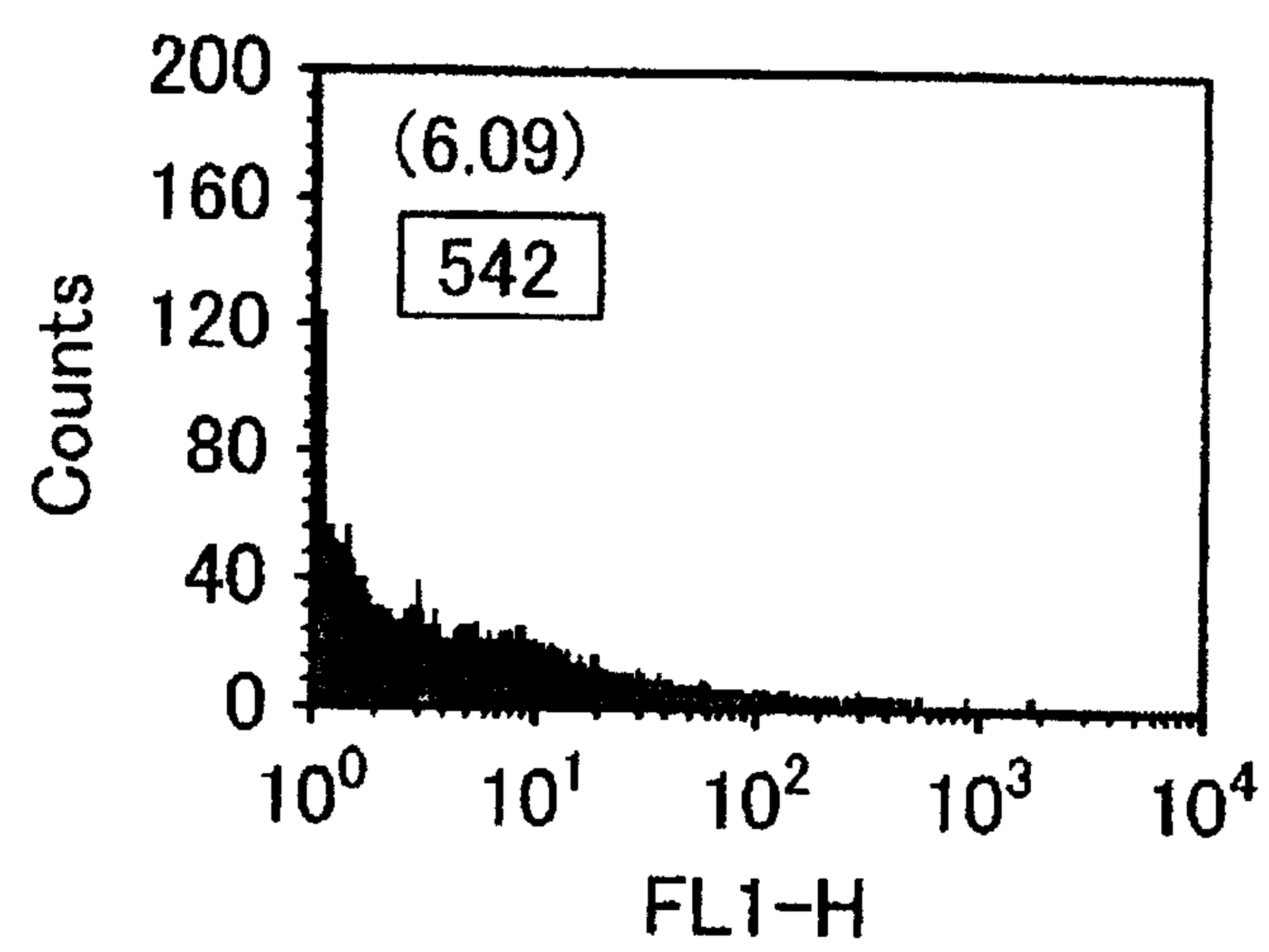
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[ 1(a)][ 1(b)]

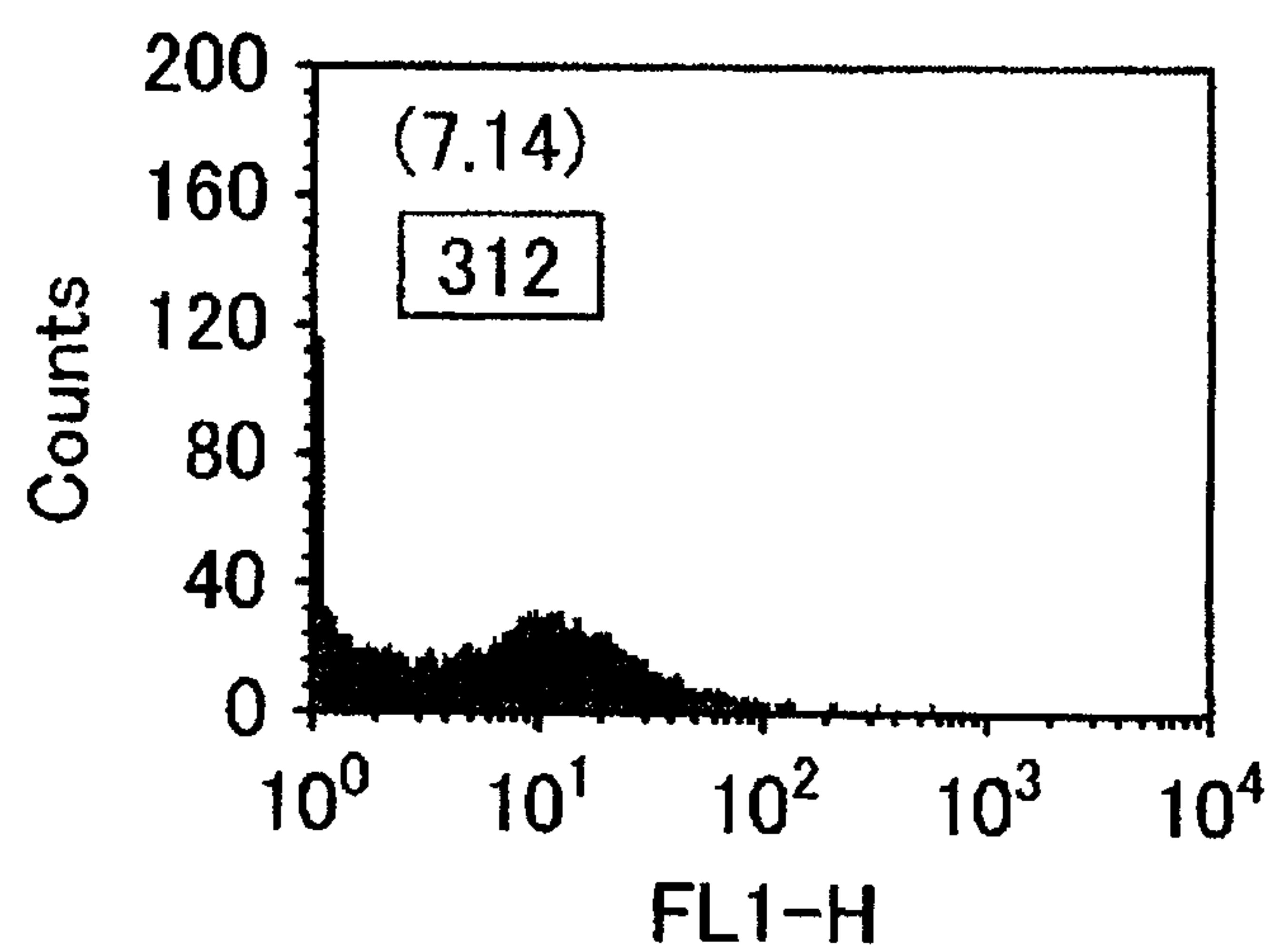
[図2(a)]



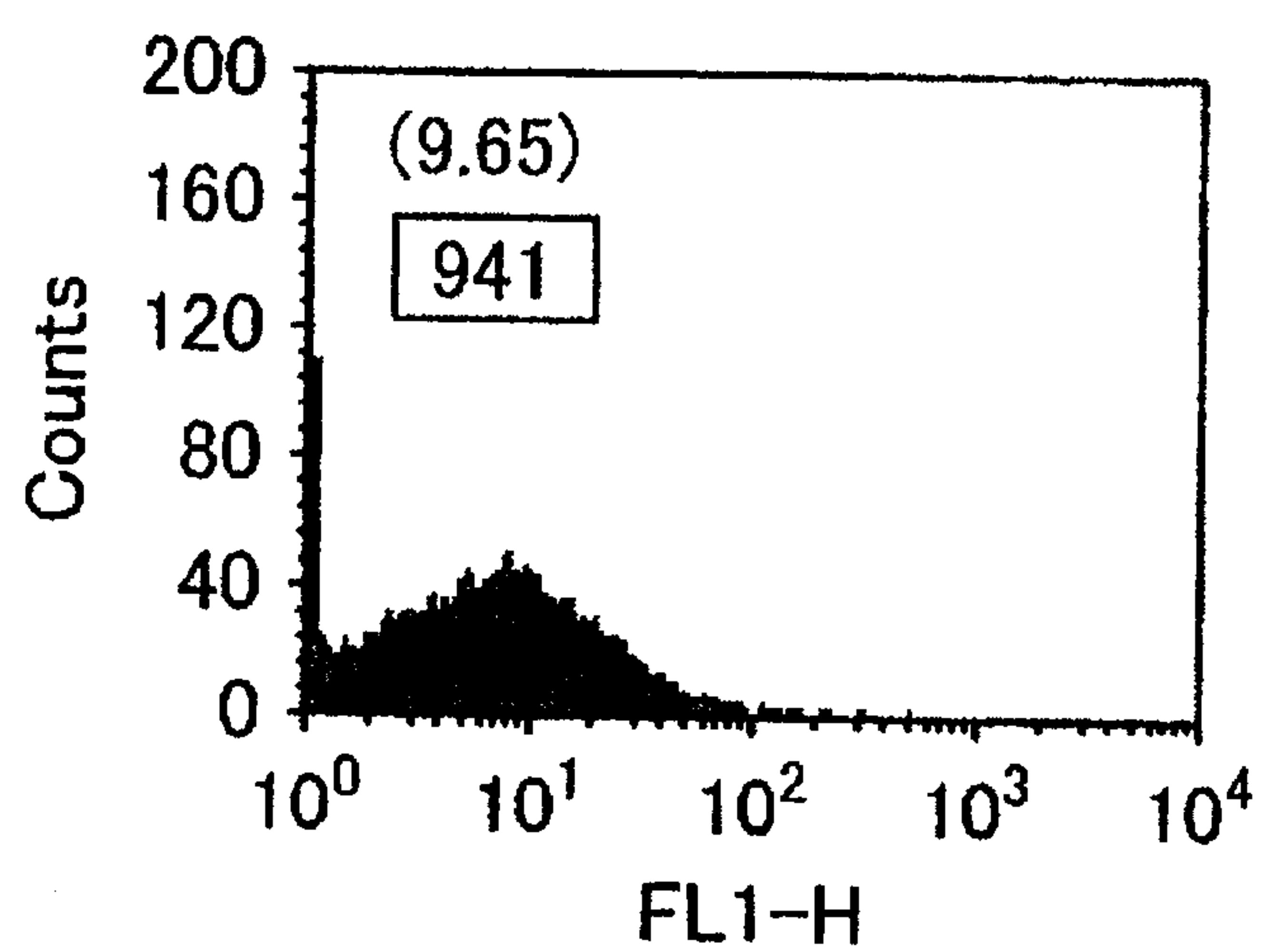
[図2(b)]



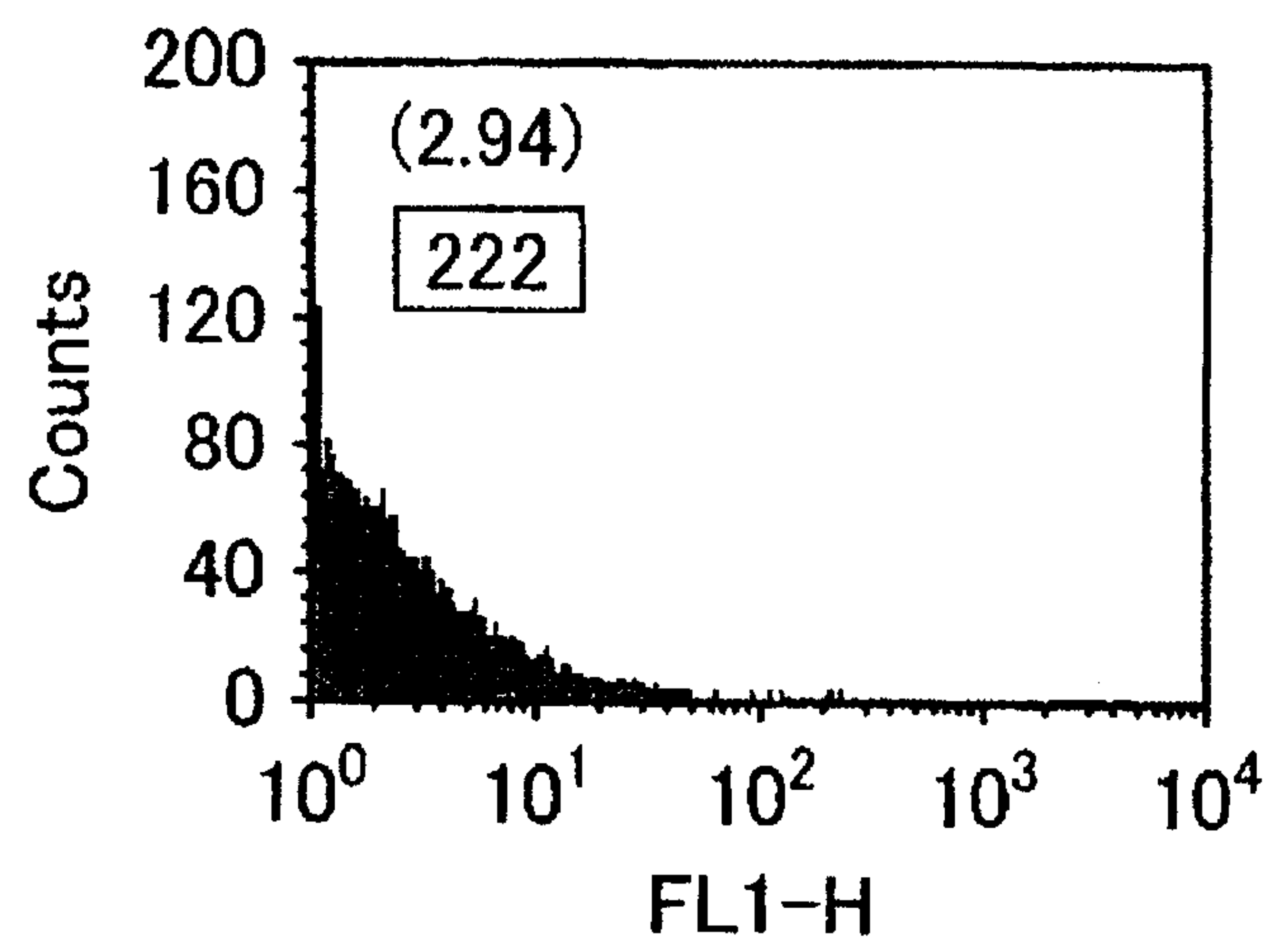
[圖2(c)]



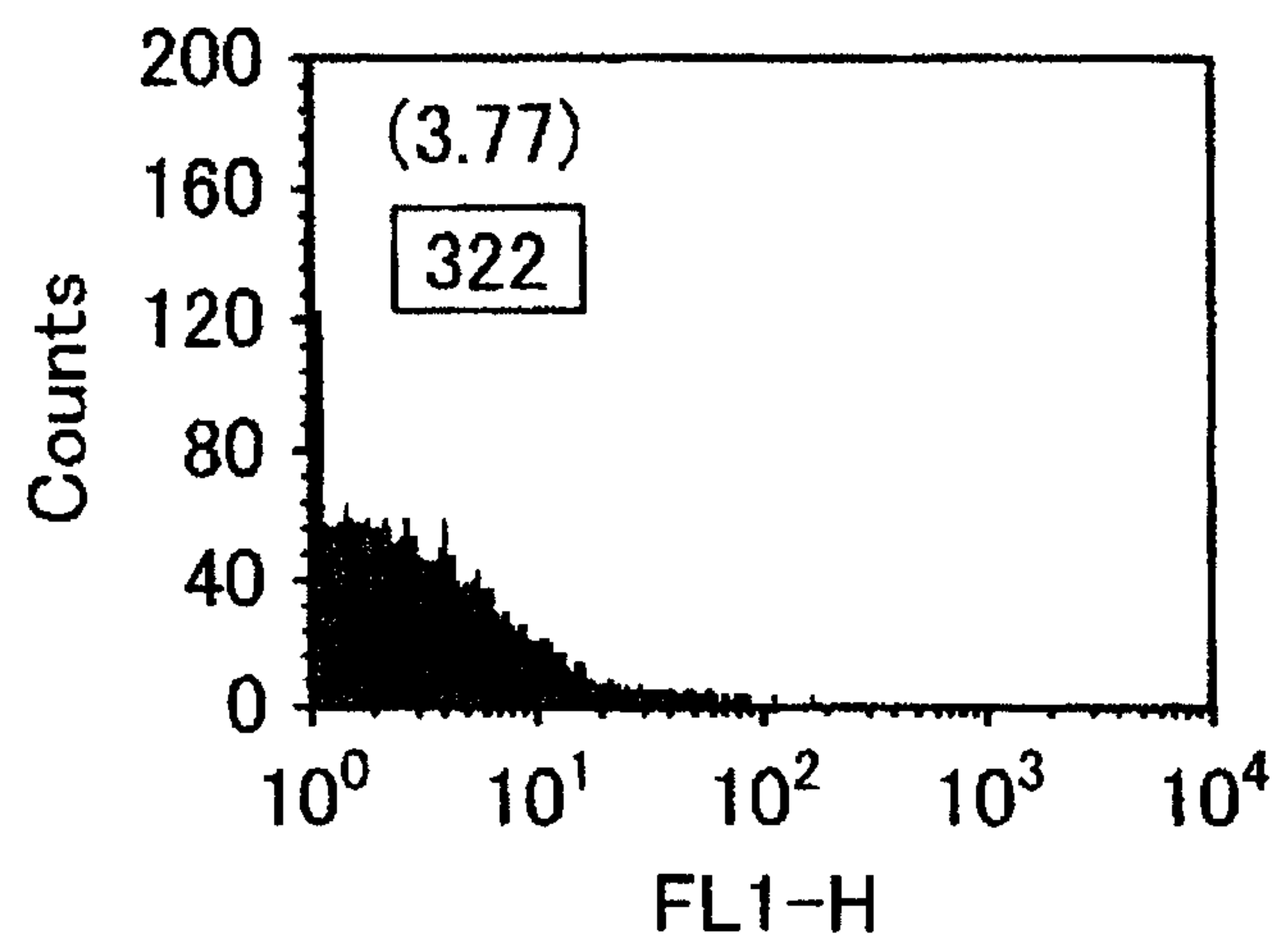
[圖2(d)]



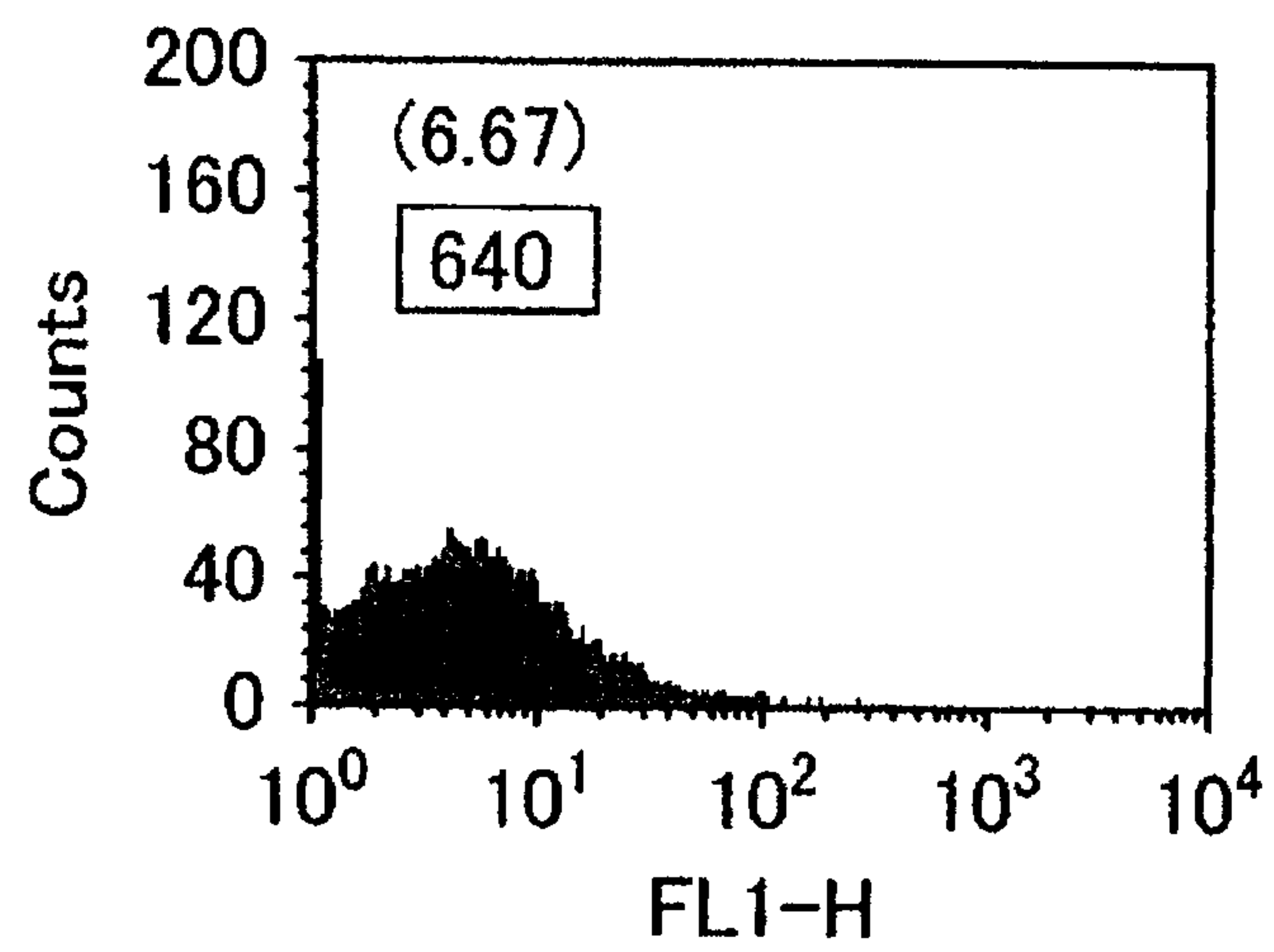
[図3(a)]



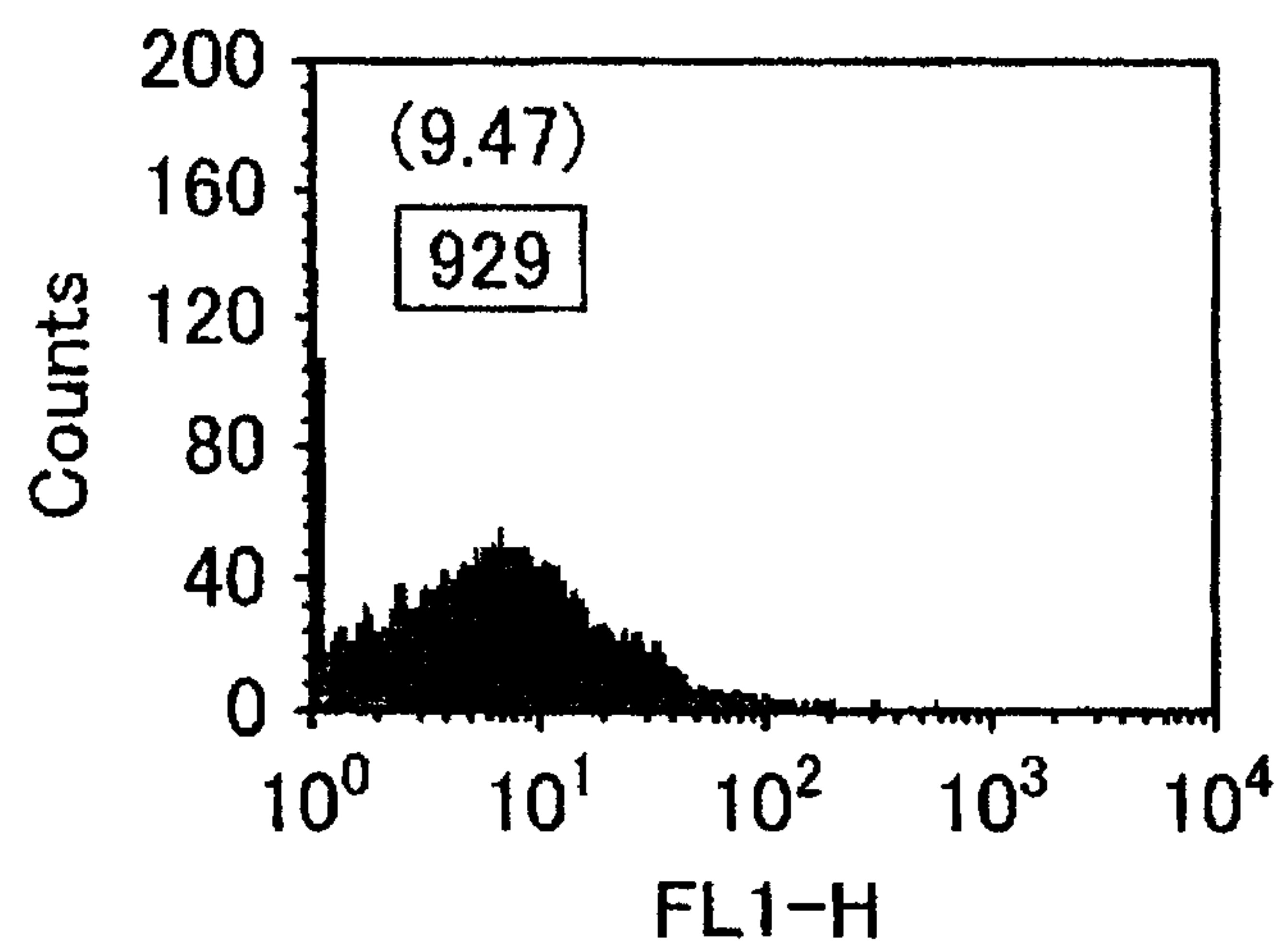
[図3(b)]



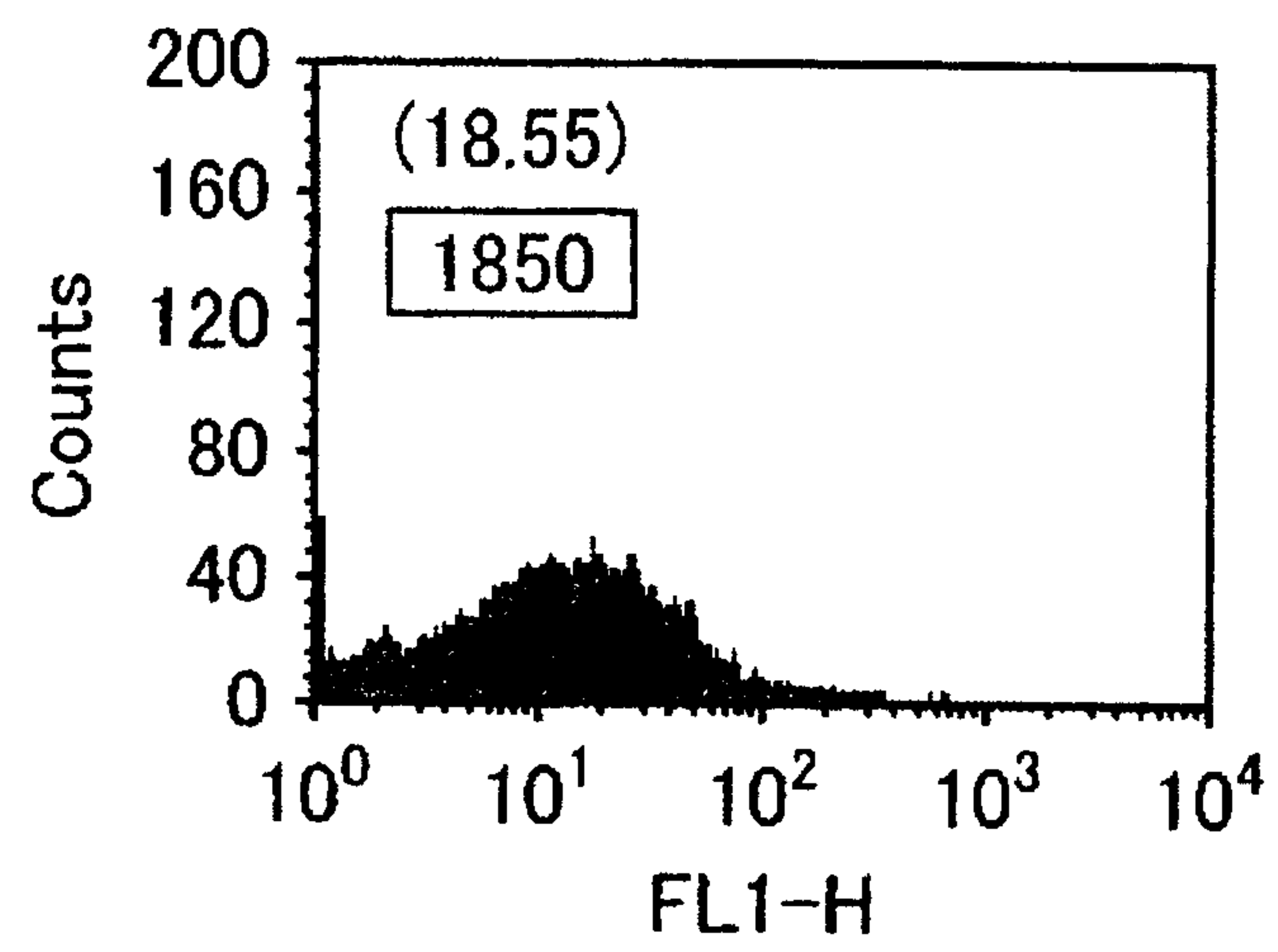
[图3(c)]



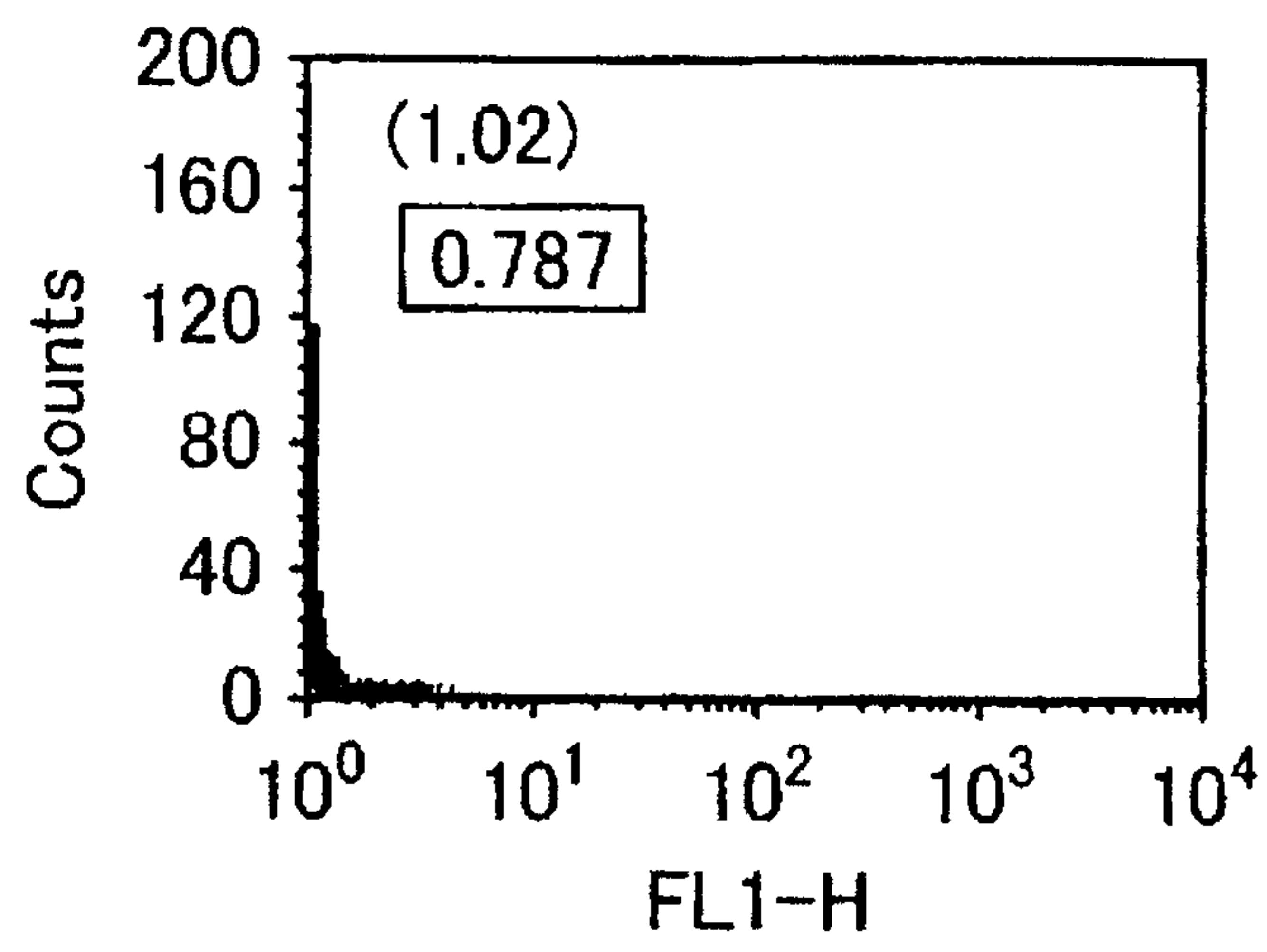
[图3(d)]



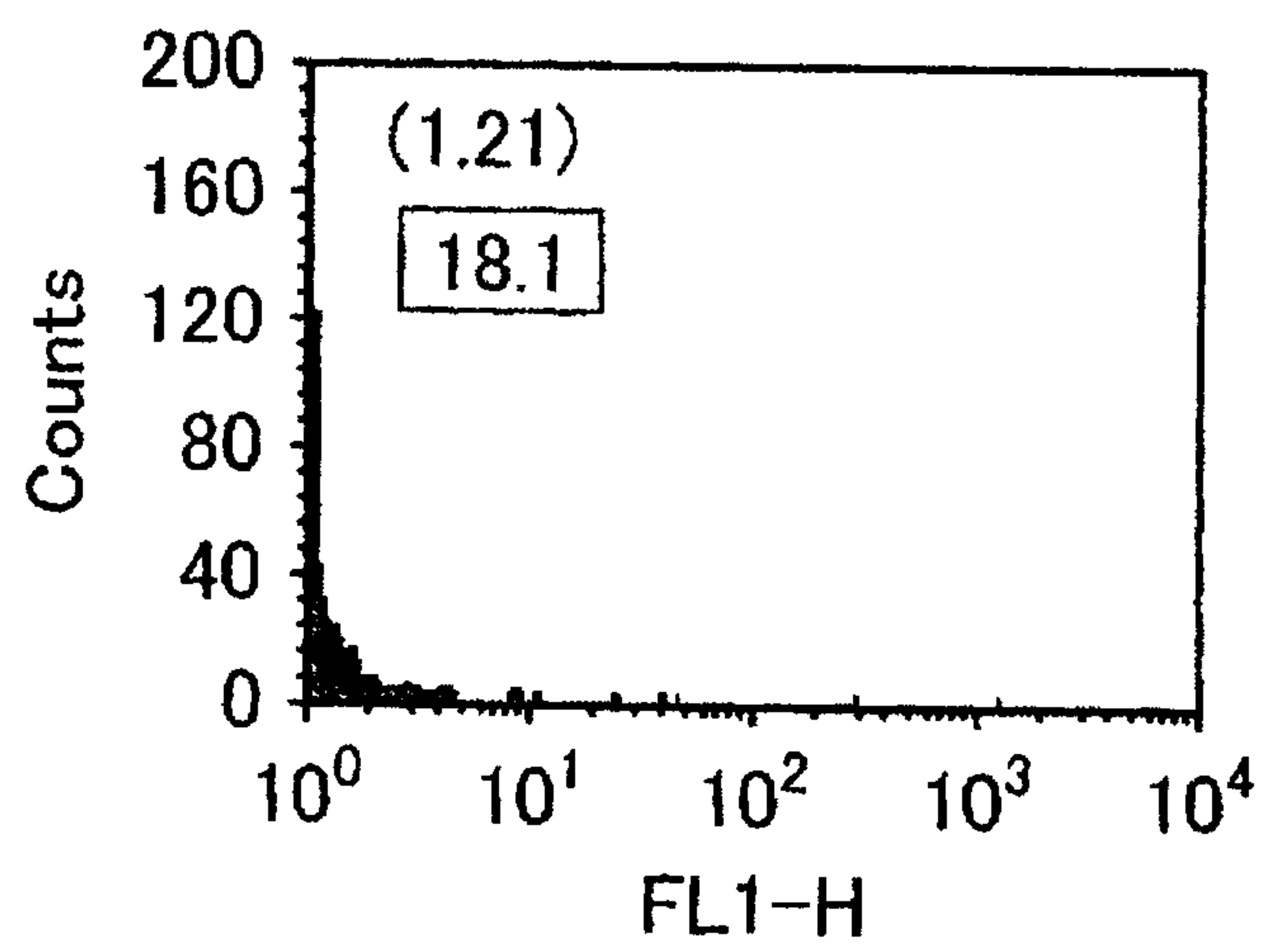
[圖3(e)]



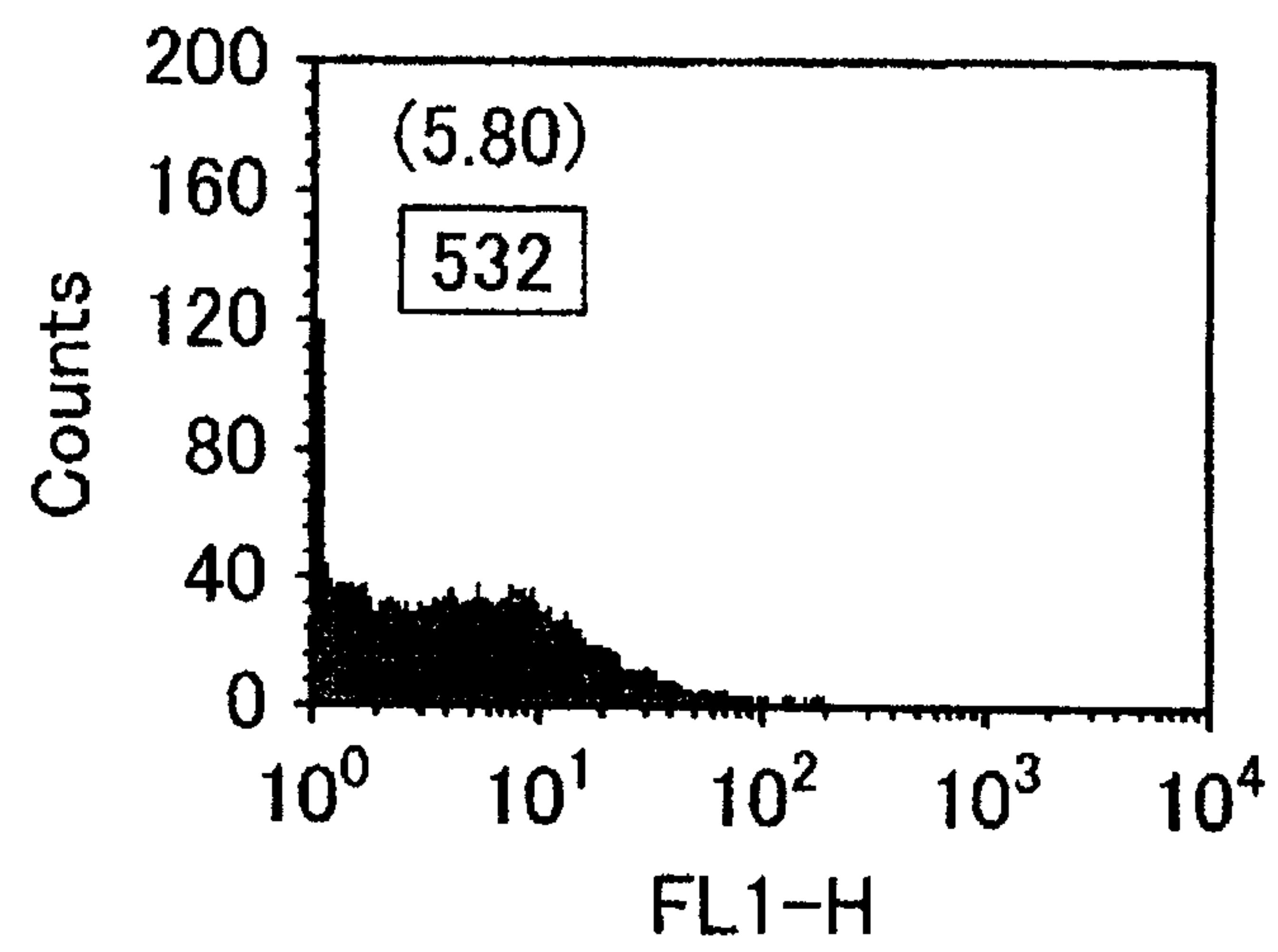
[圖5(a)]



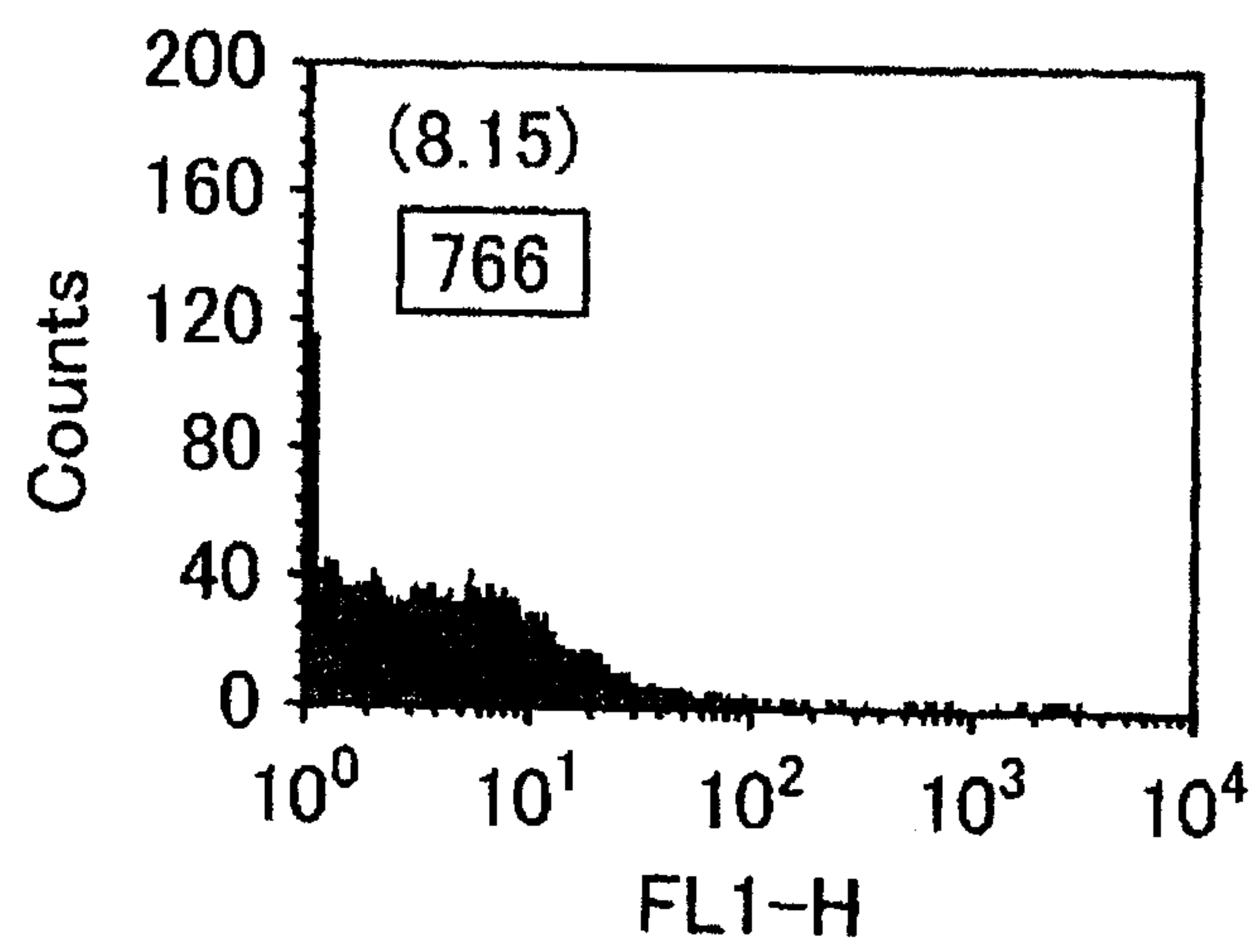
[圖5(b)]



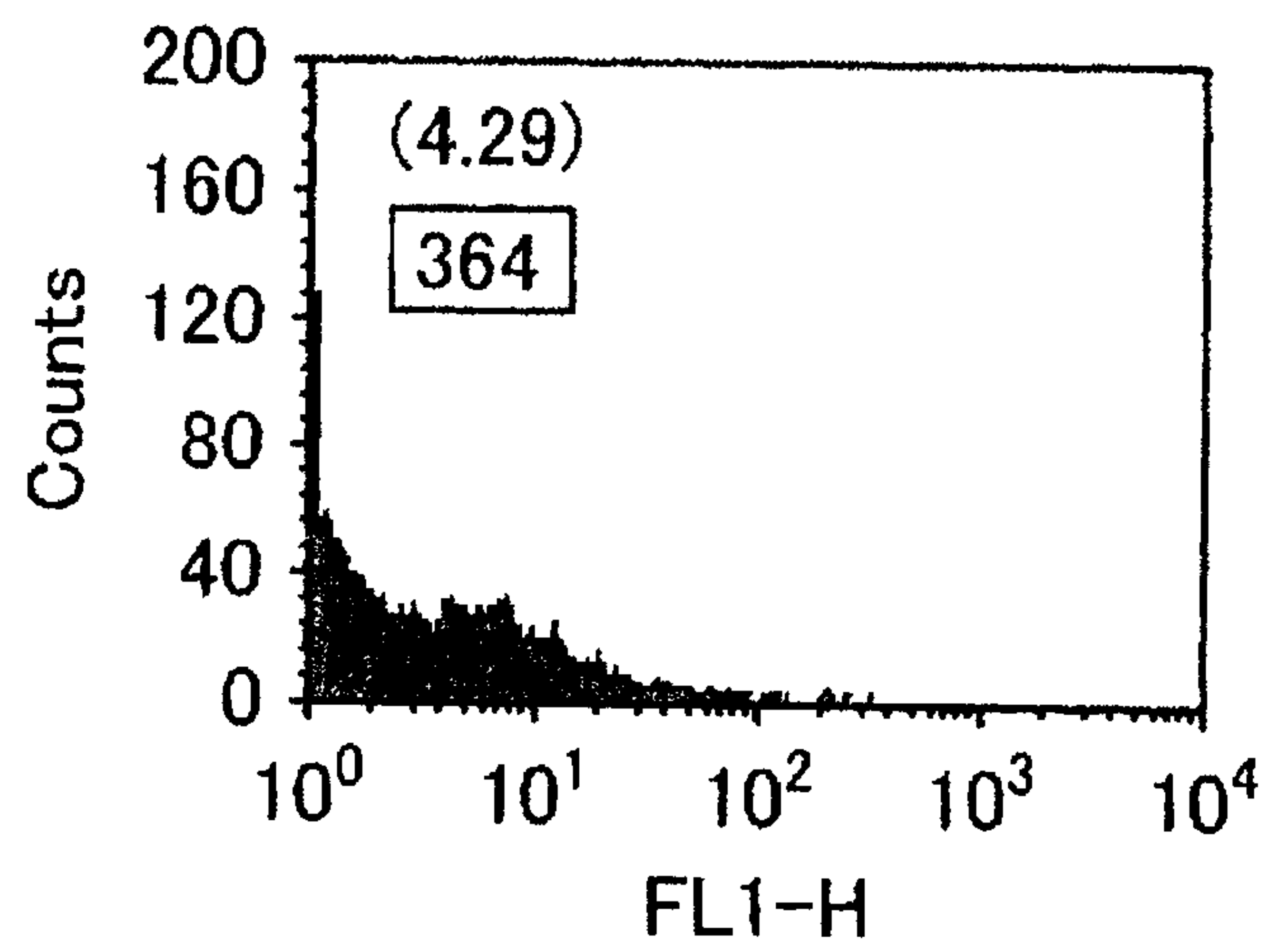
[図5(c)]



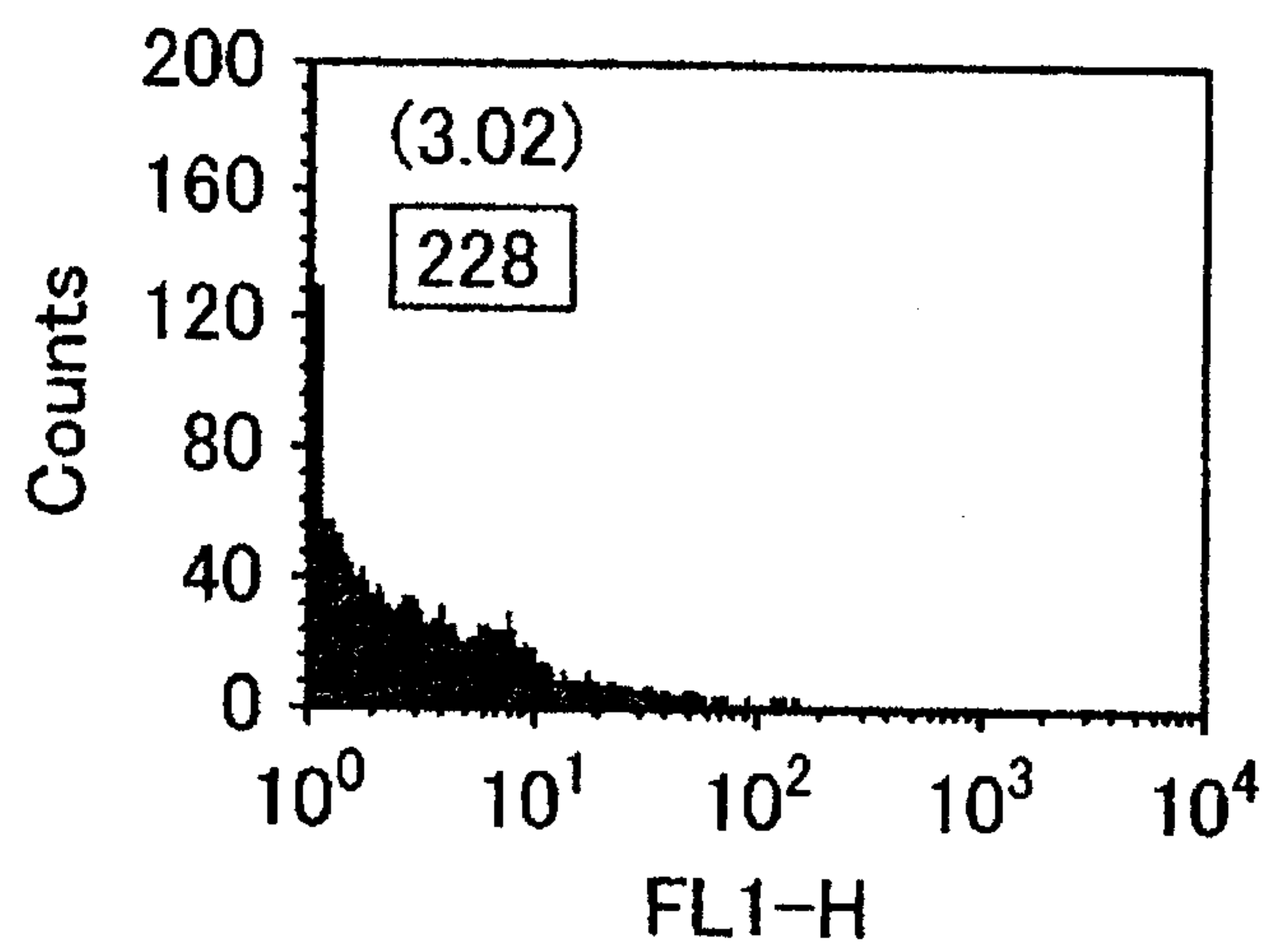
[図5(d)]



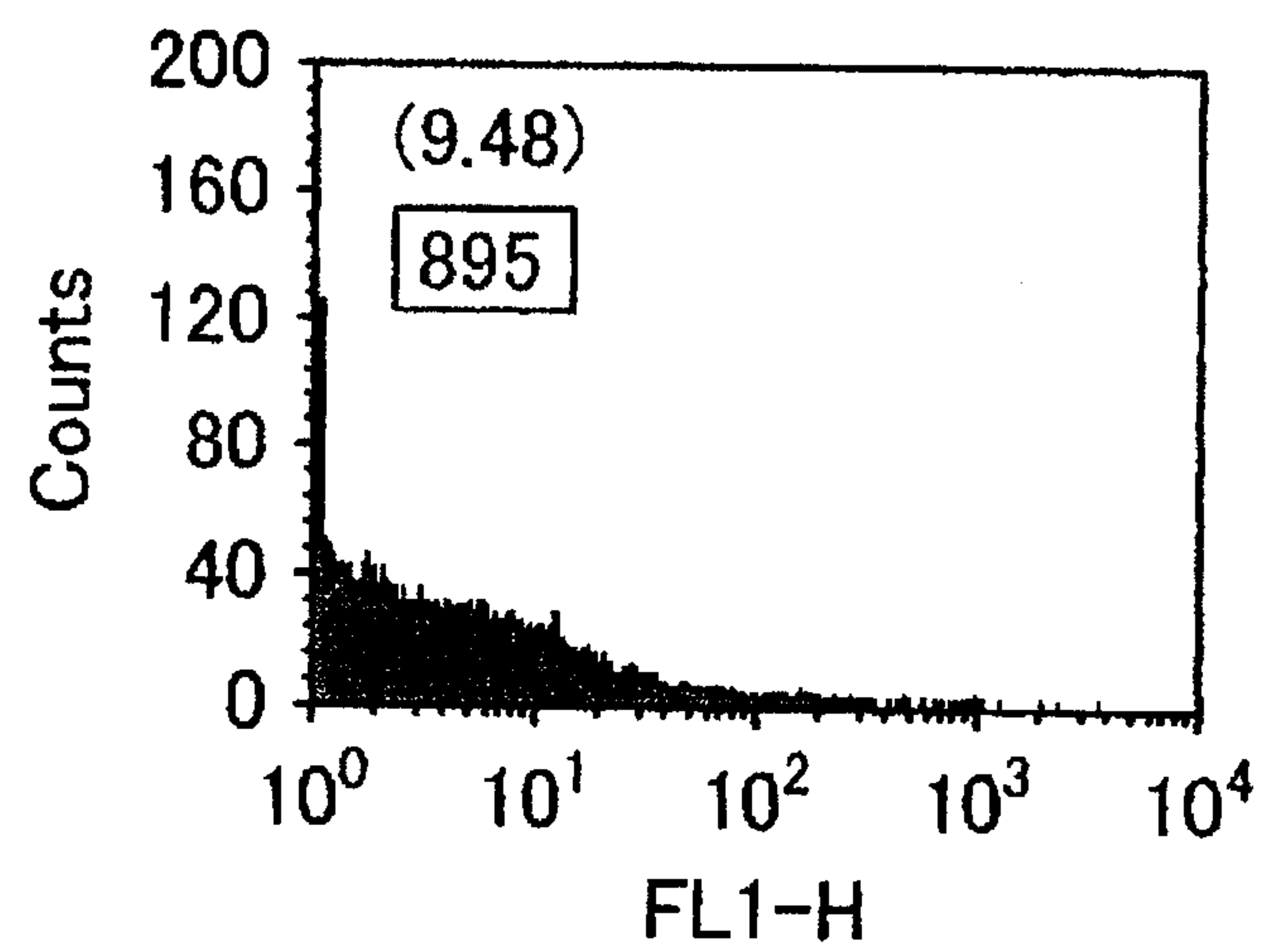
[図8(a)]



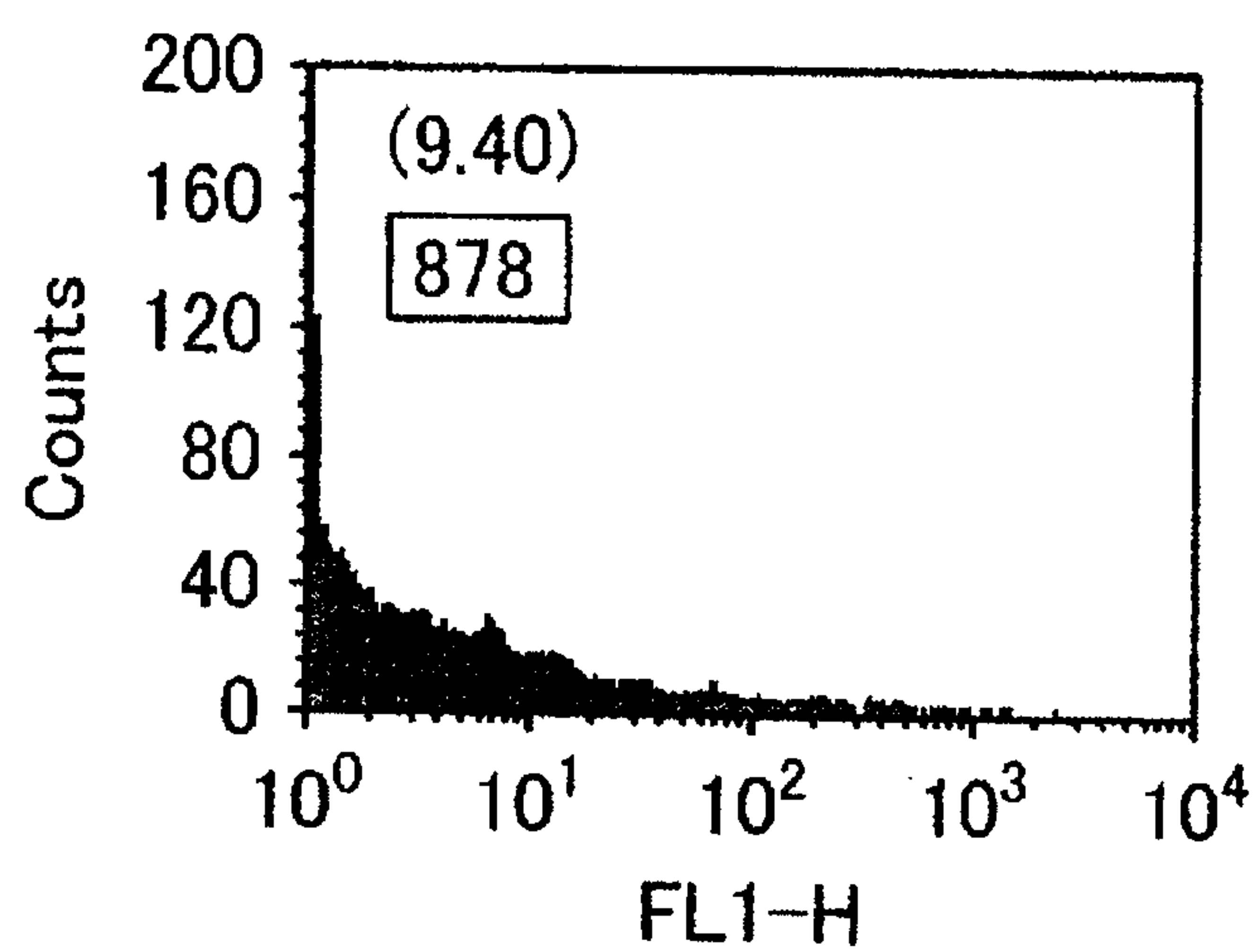
[図8(b)]



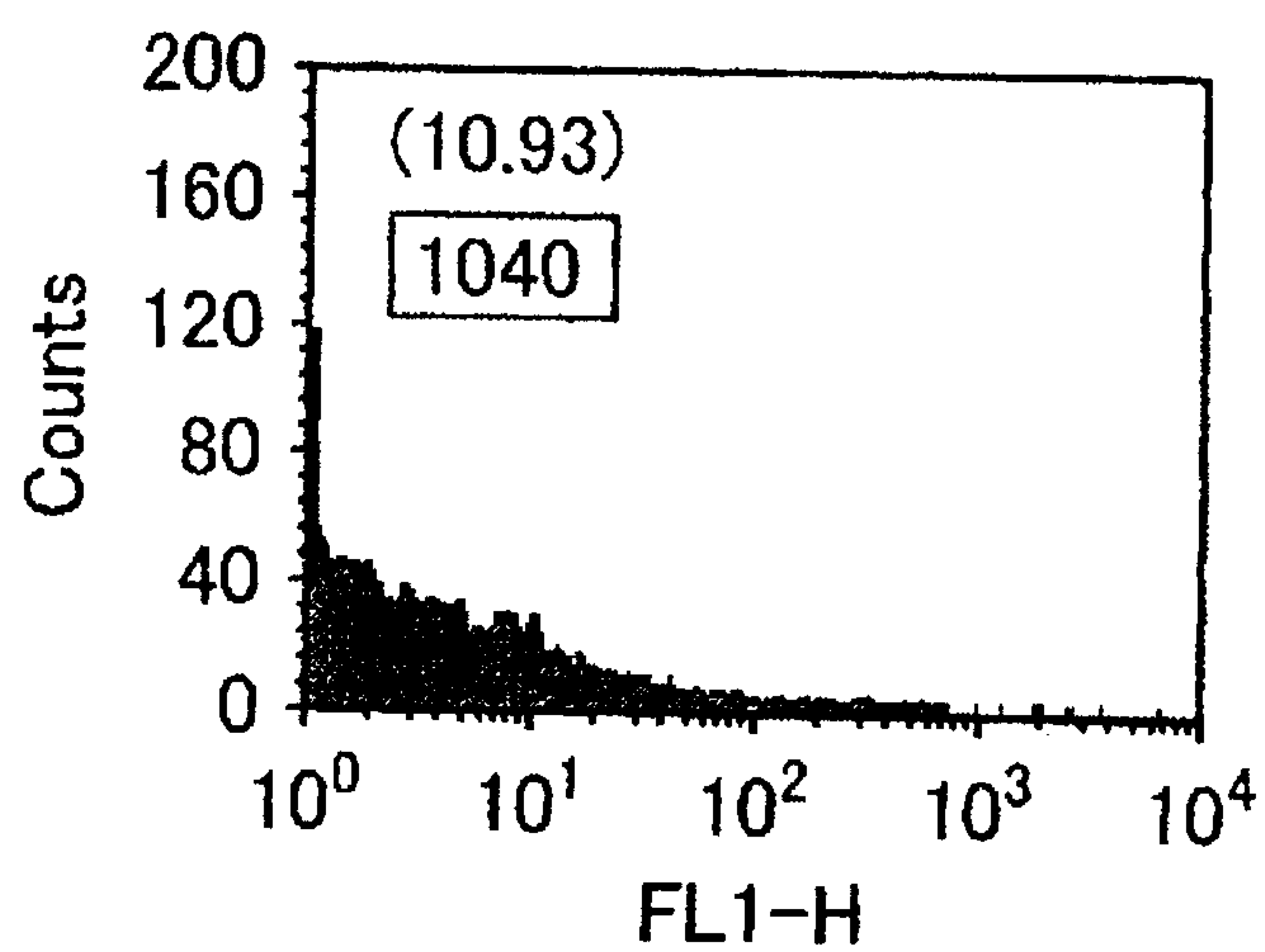
[図8(c)]



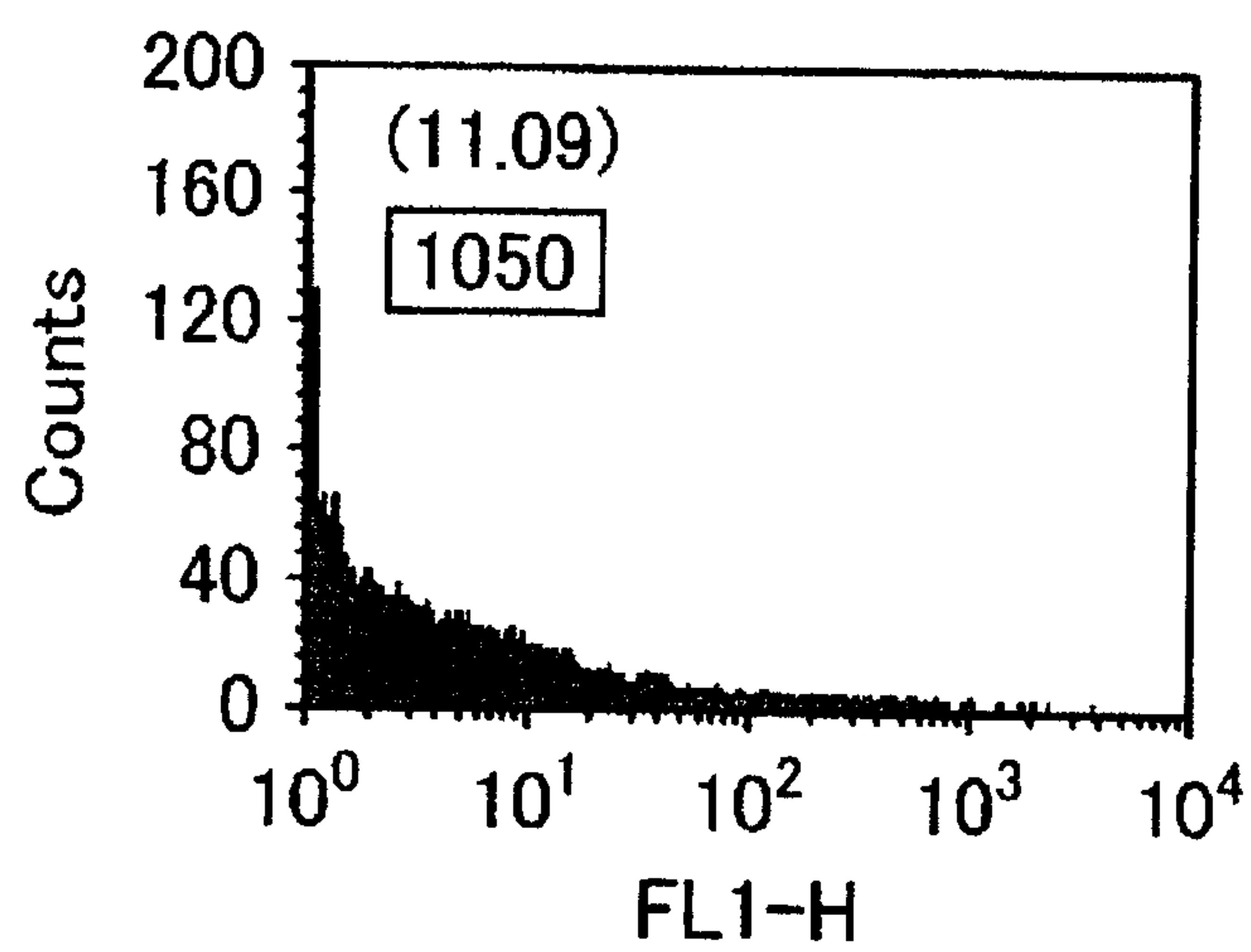
[[8(d)]]



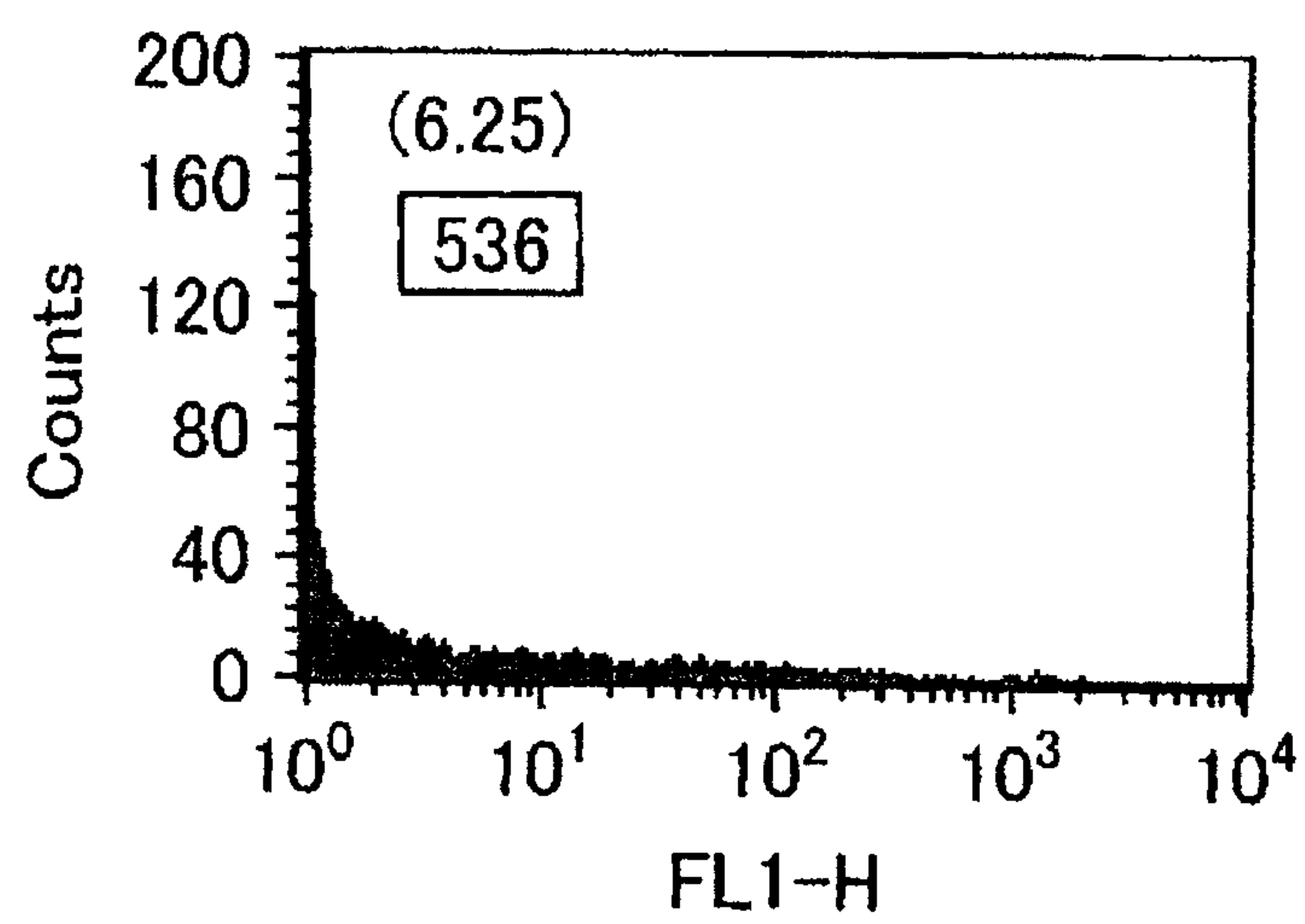
[[8(e)]]



[[8(f)]]



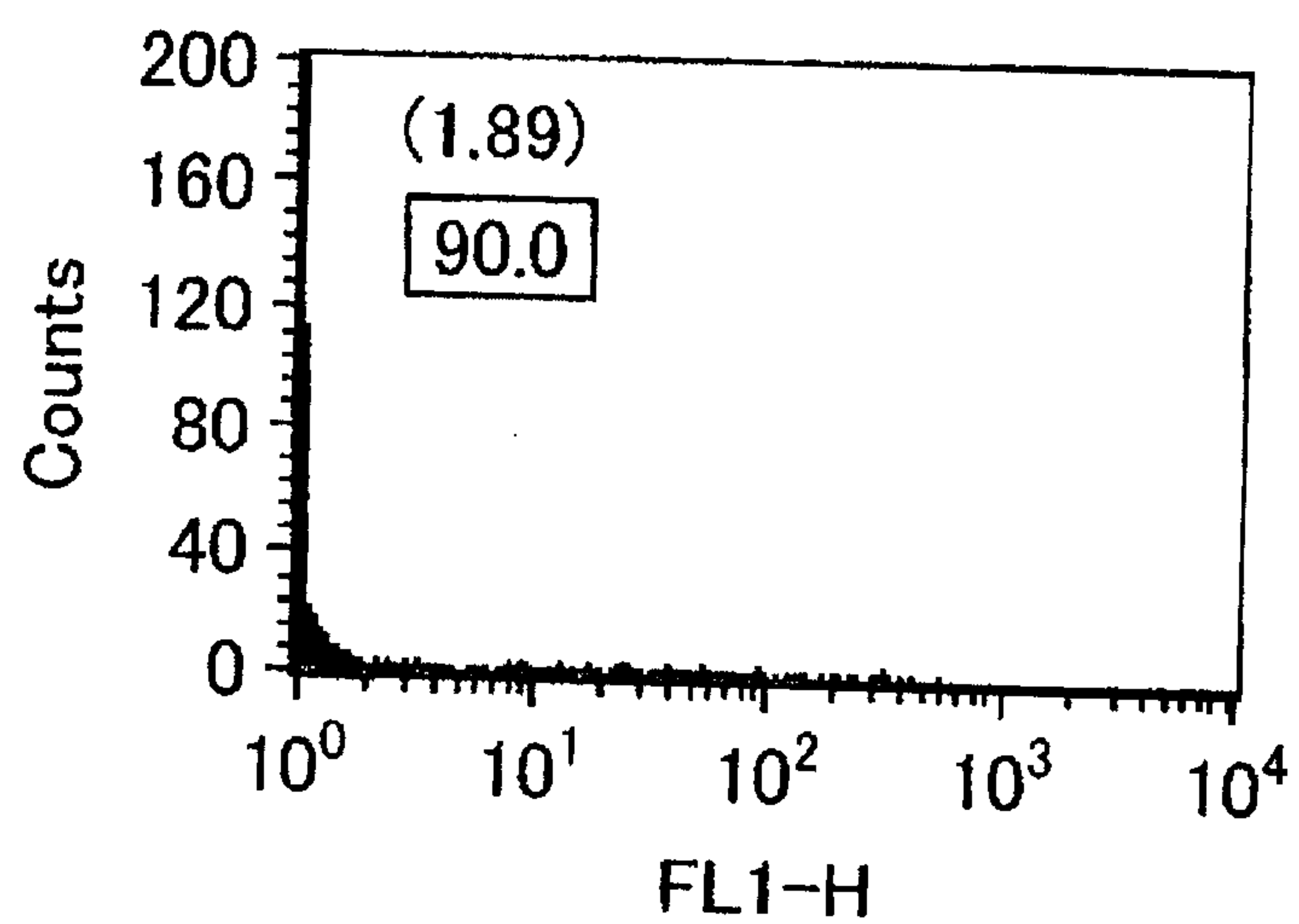
[X]9]



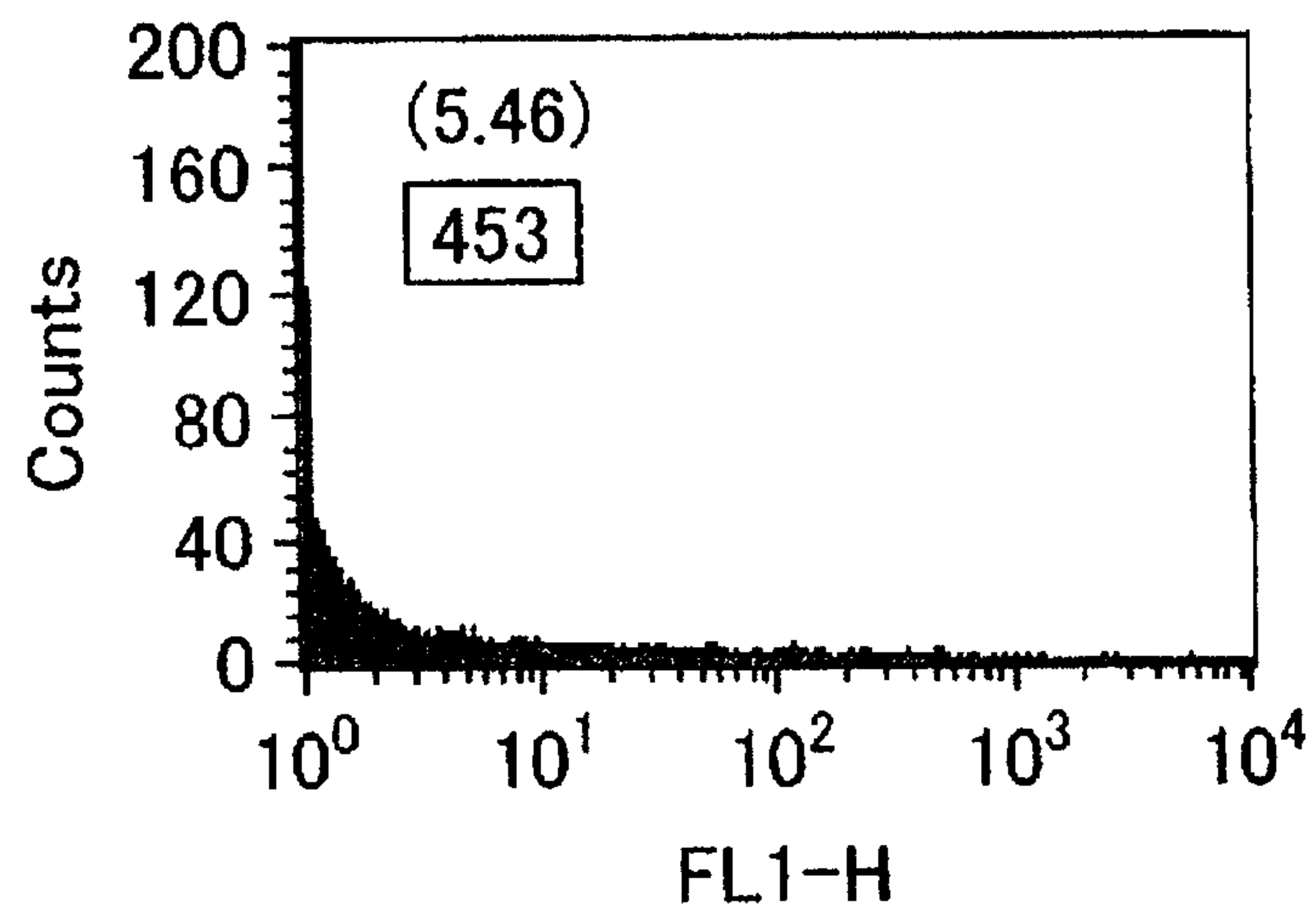
[X]10(a)]

[10(d)]

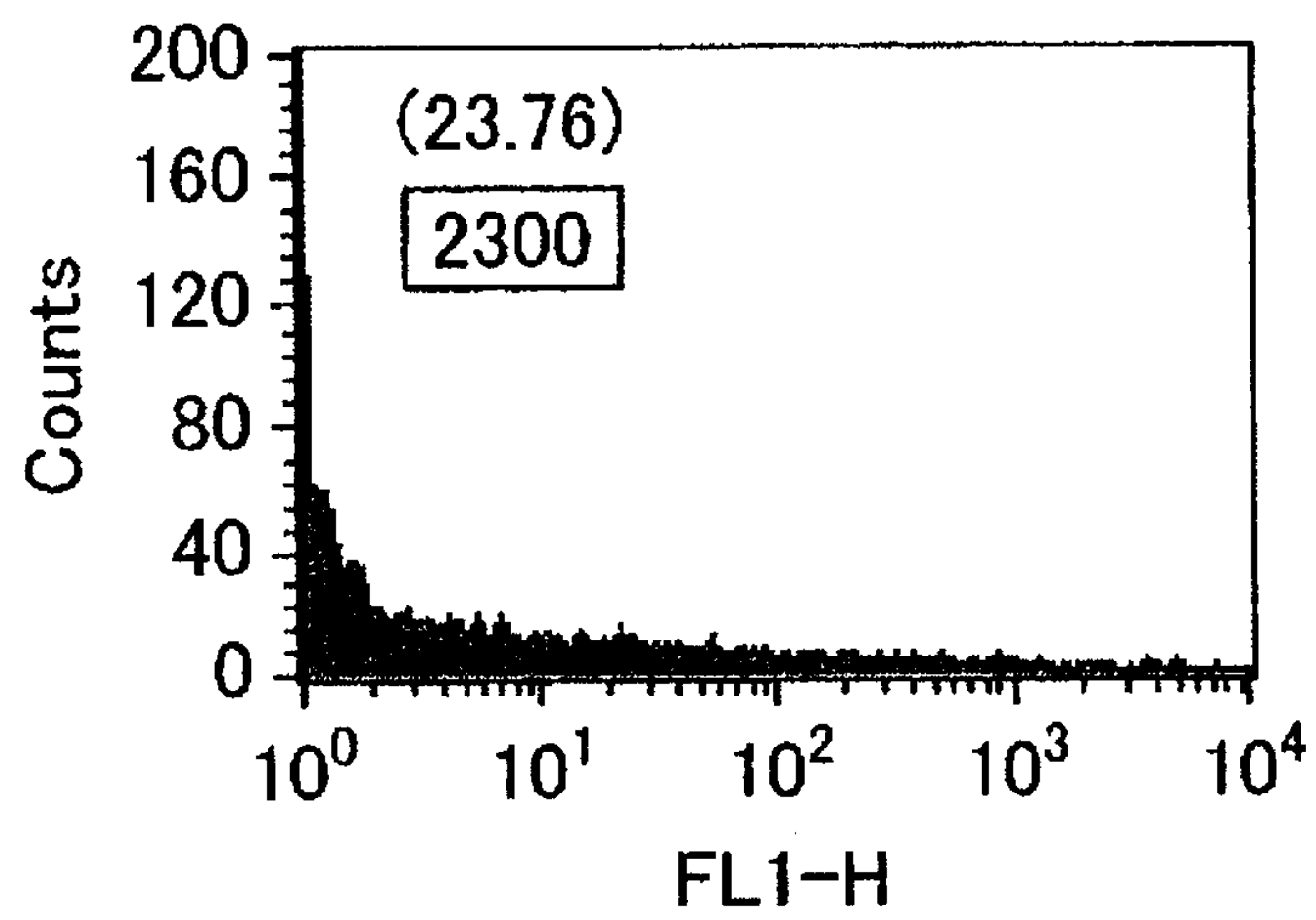
[REDACTED]

[11(a)]

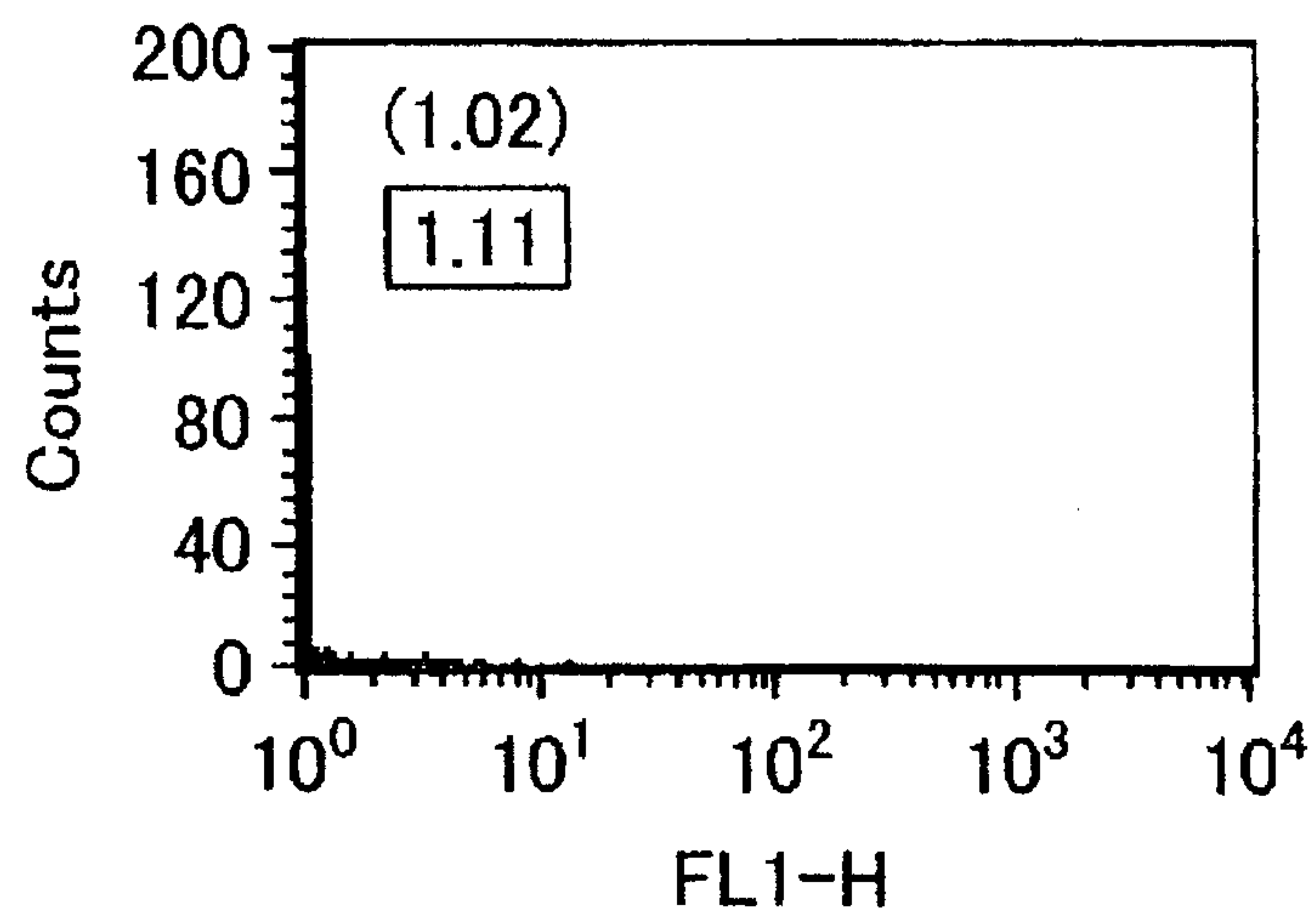
[図11(b)]



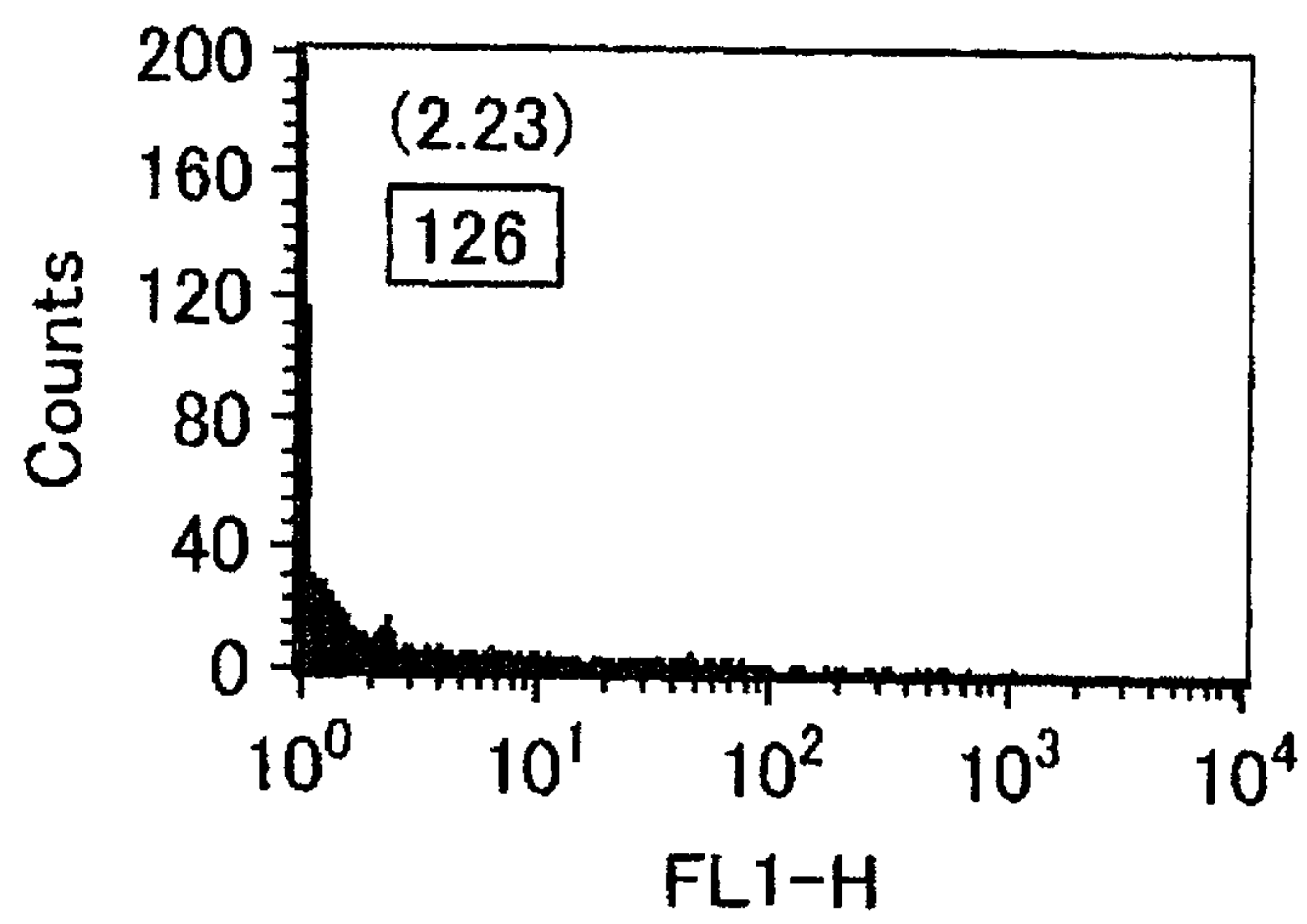
[図11(c)]



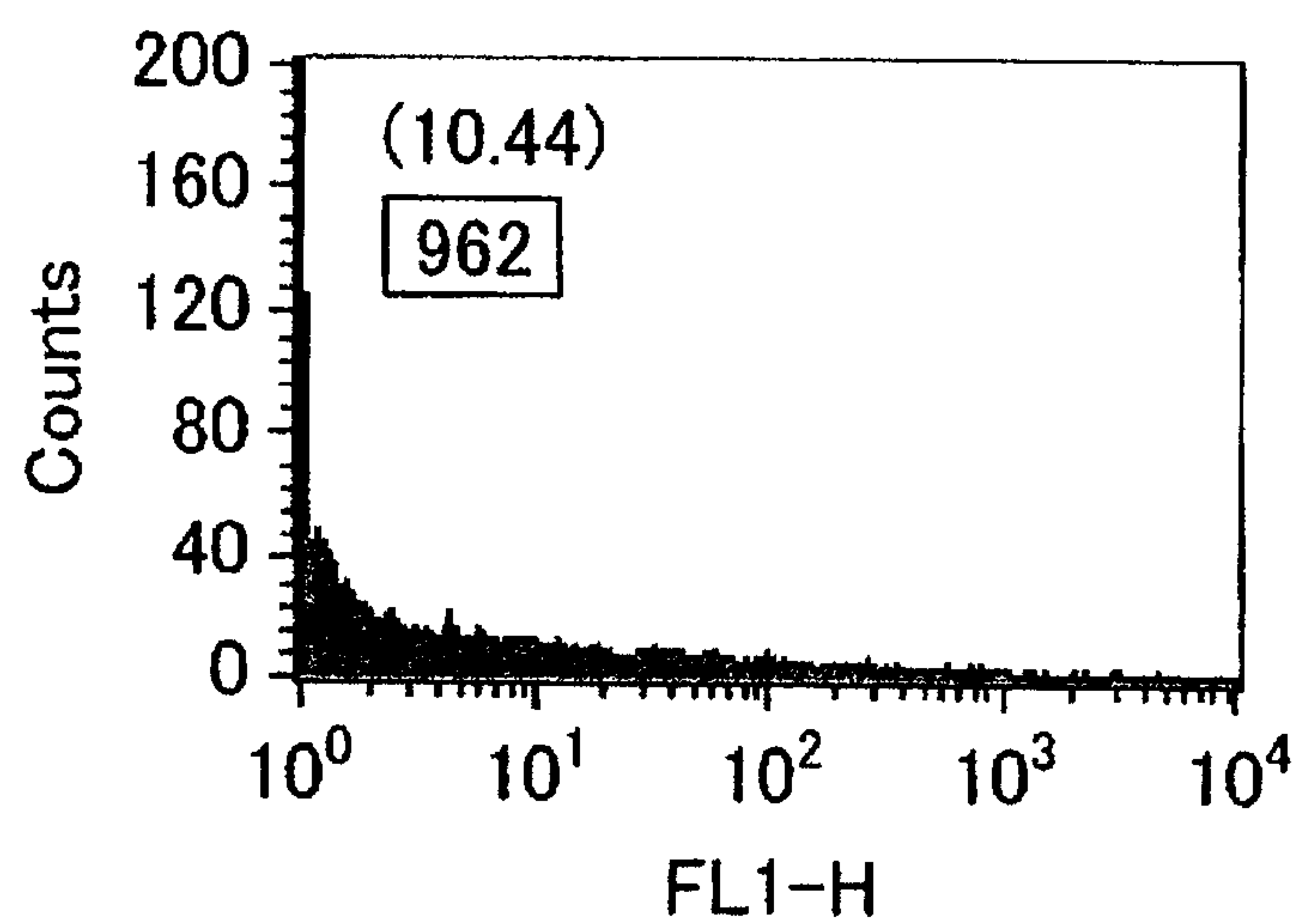
[図12(a)]



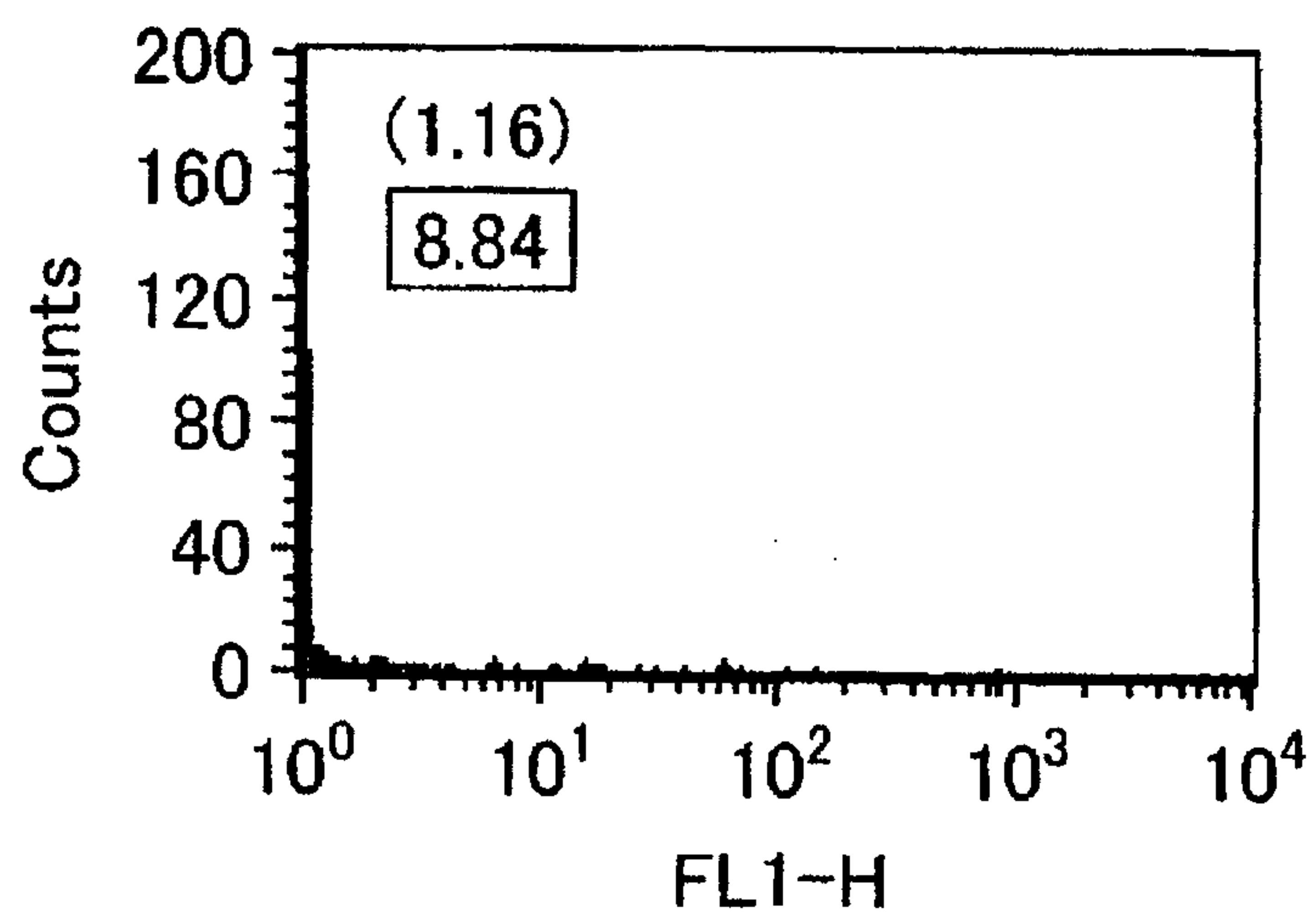
[図12(b)]



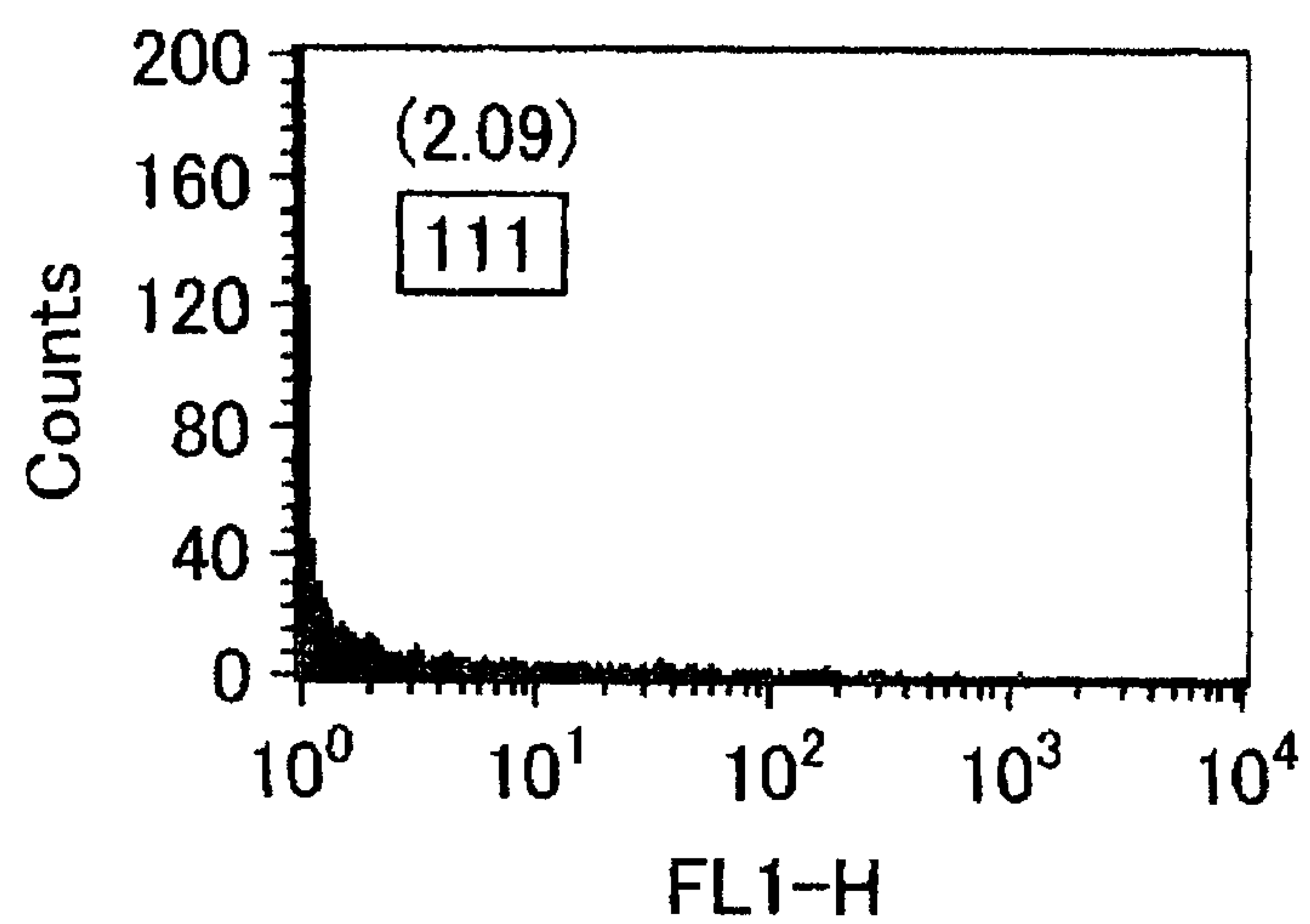
[図12(c)]



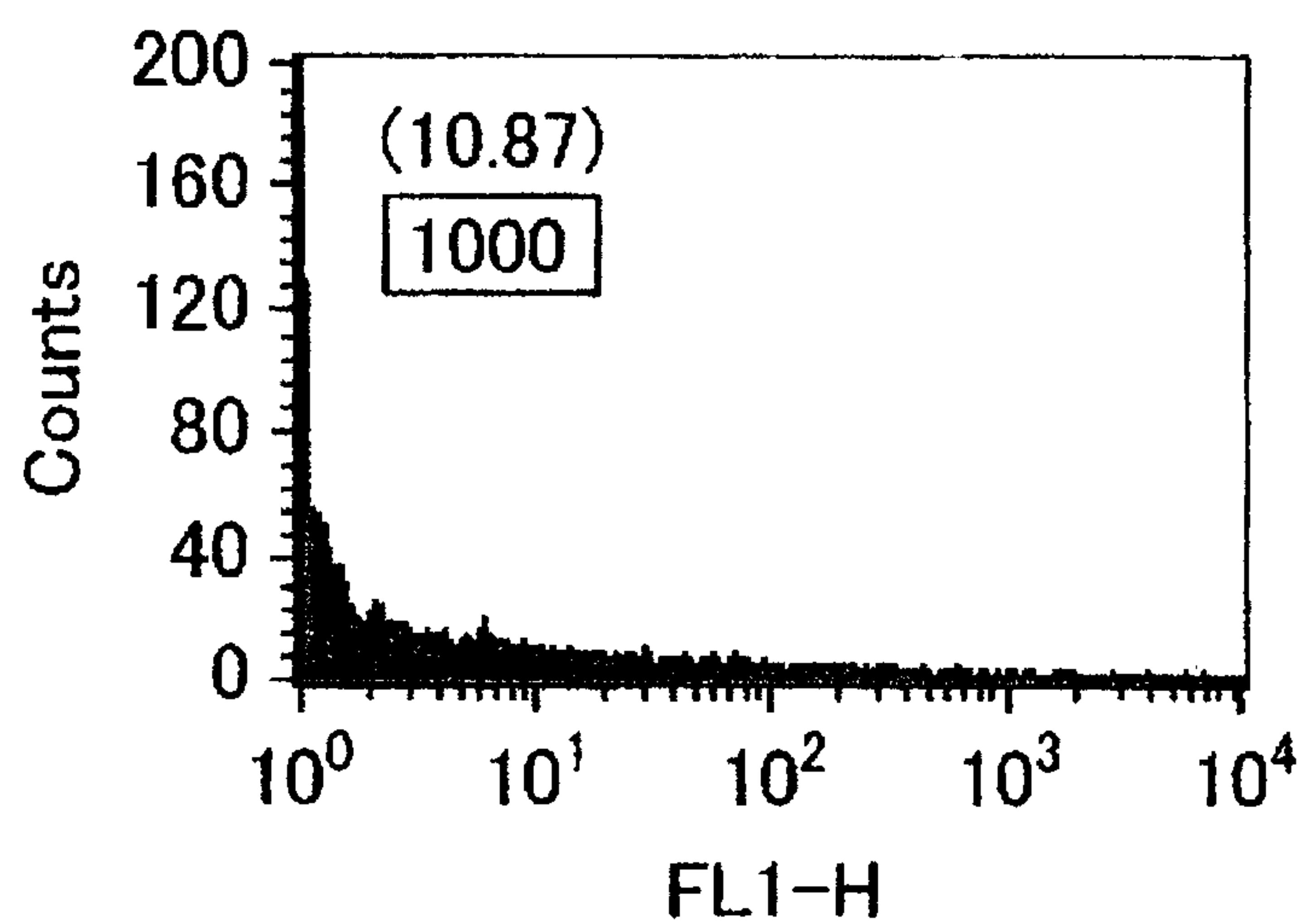
[図13(a)]



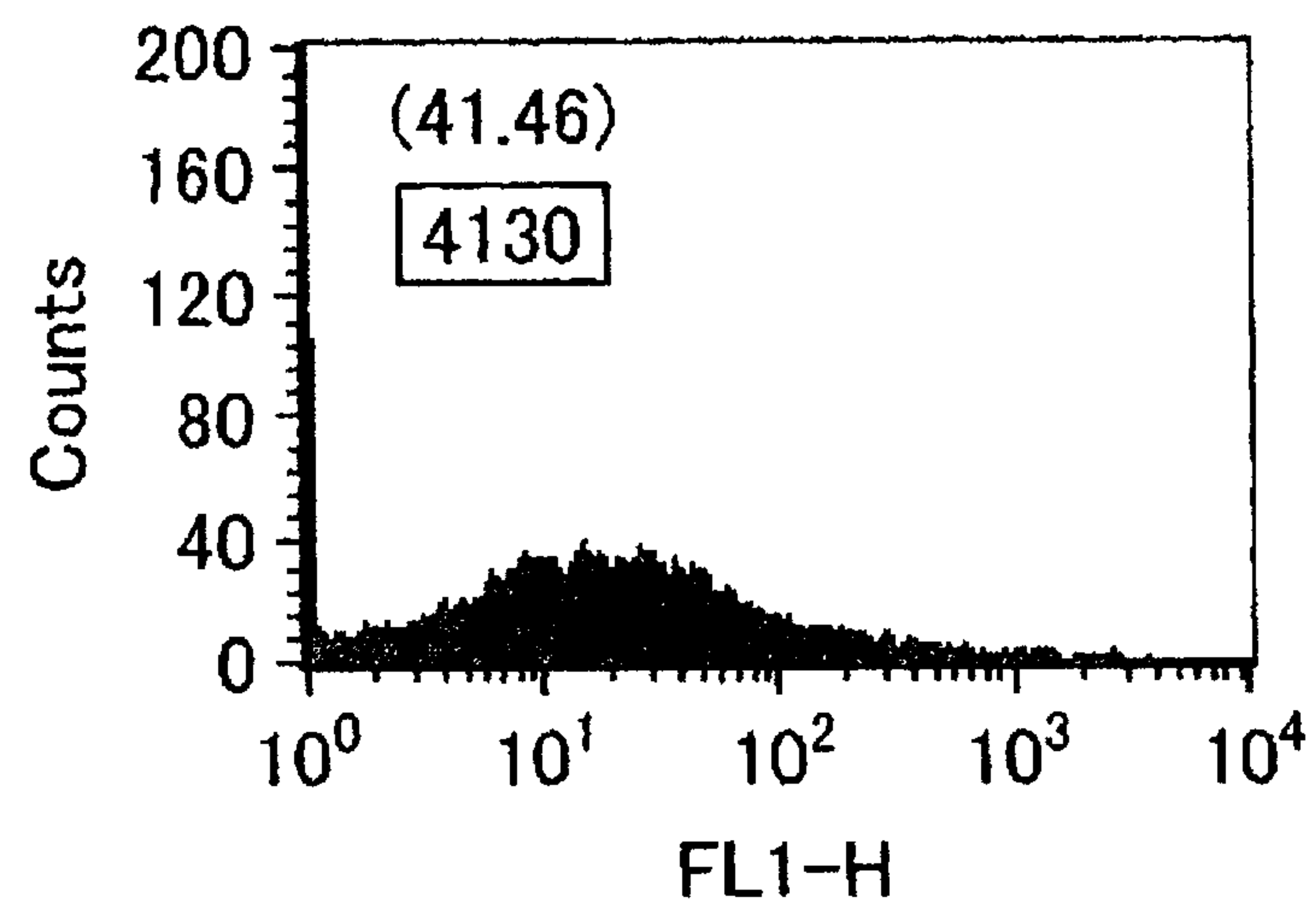
[圖13(b)]



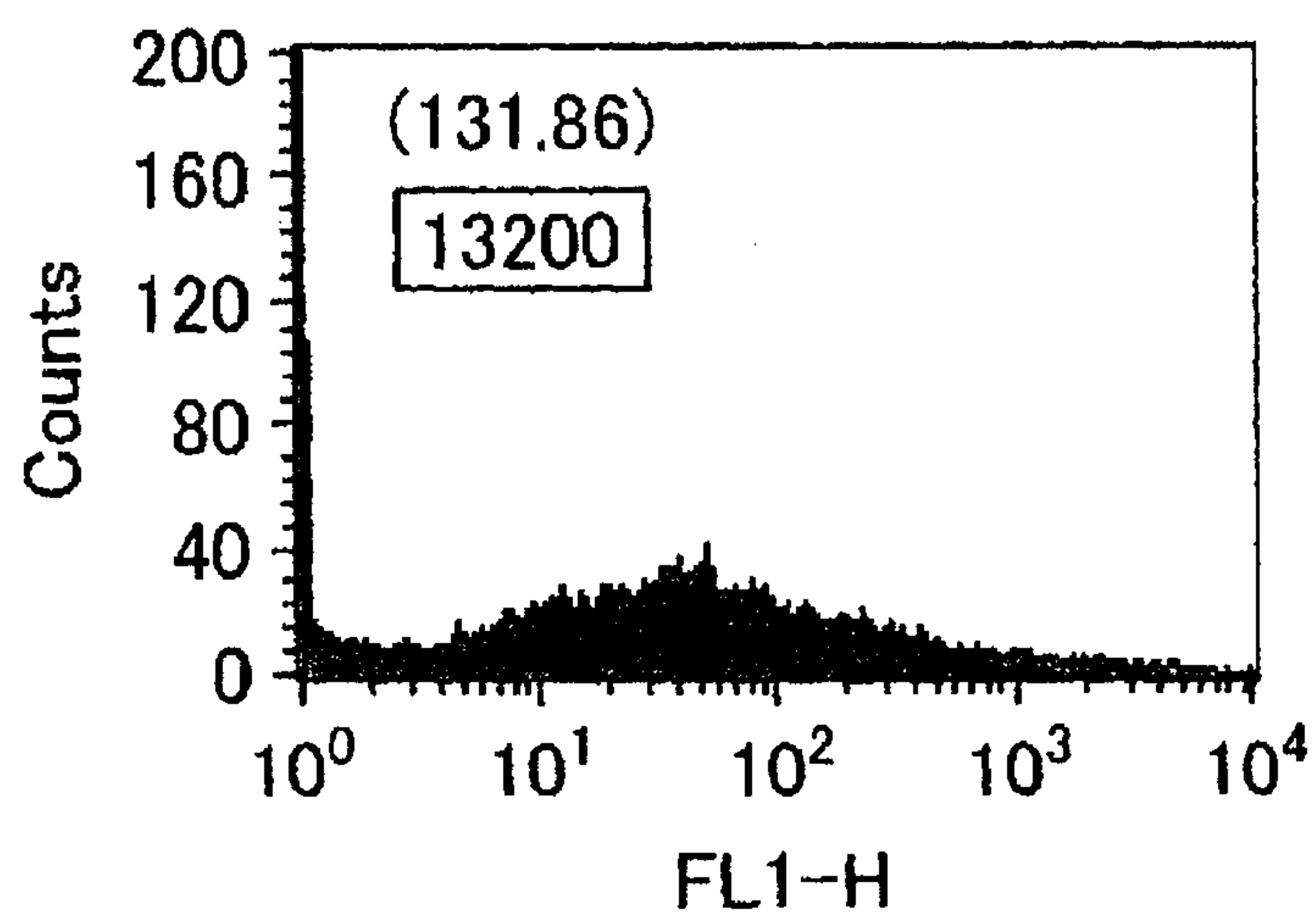
[圖13(c)]



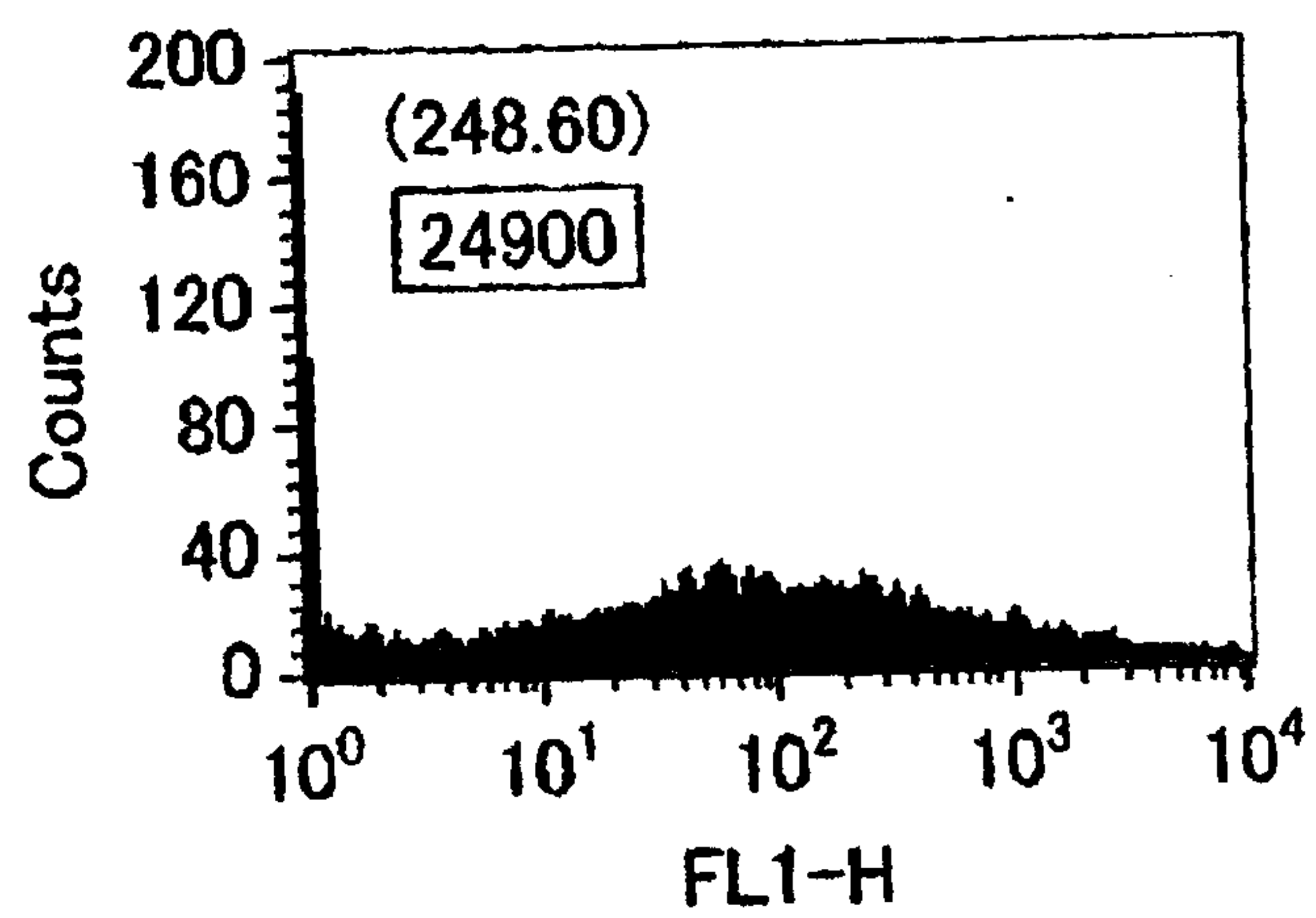
[圖14(a)]



[圖14(b)]

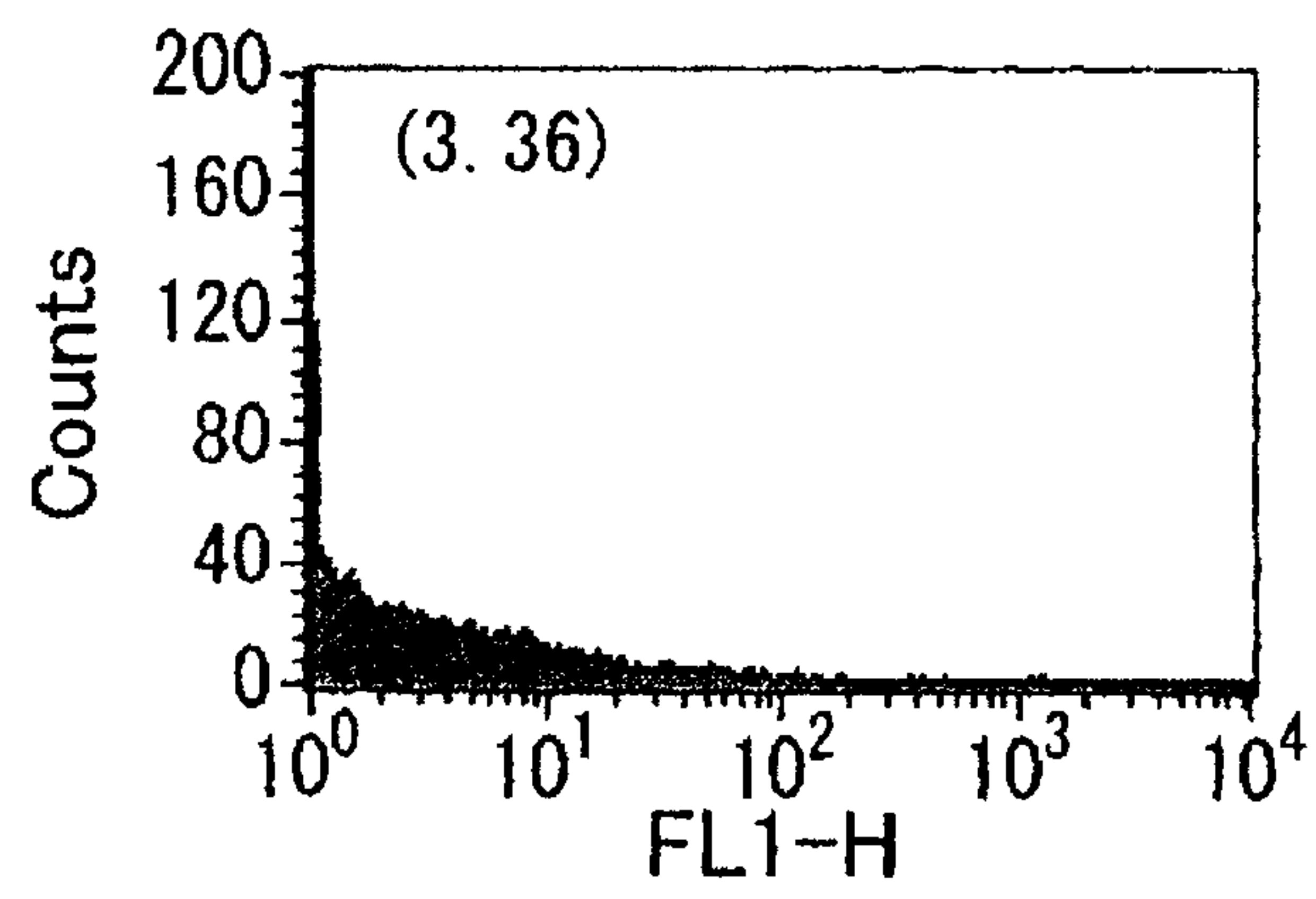


[図14(c)]

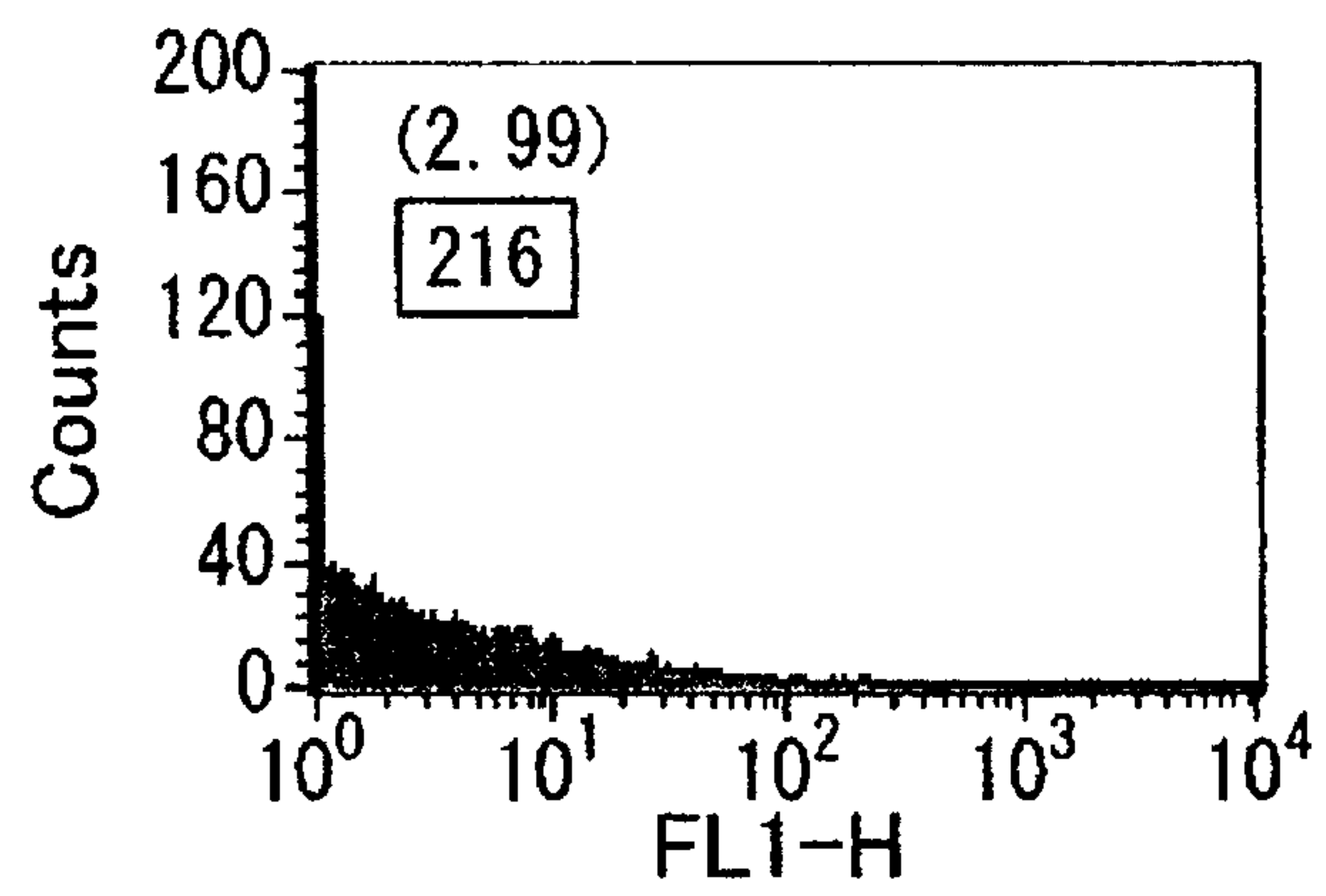


[図15]

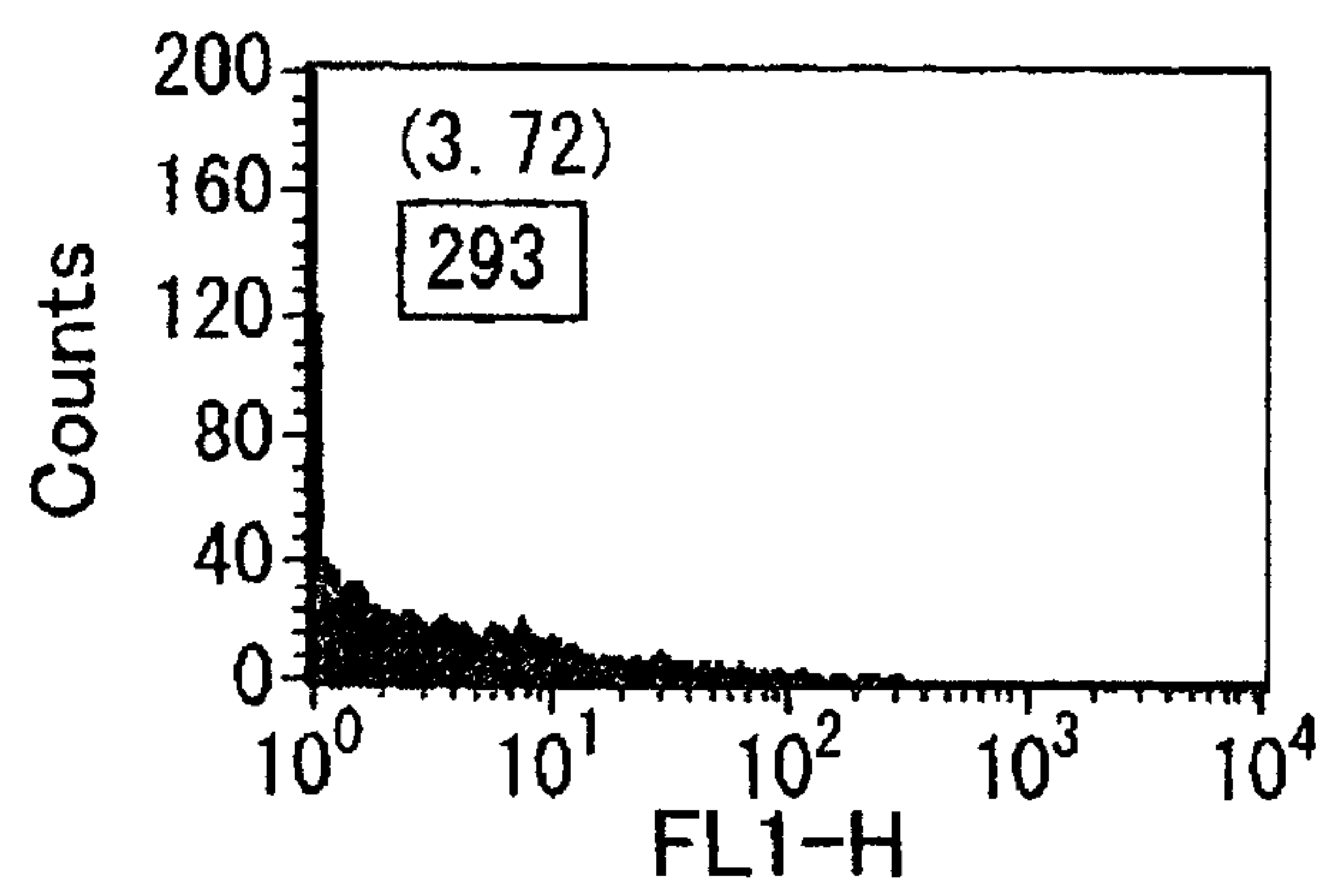
[圖]16(a)



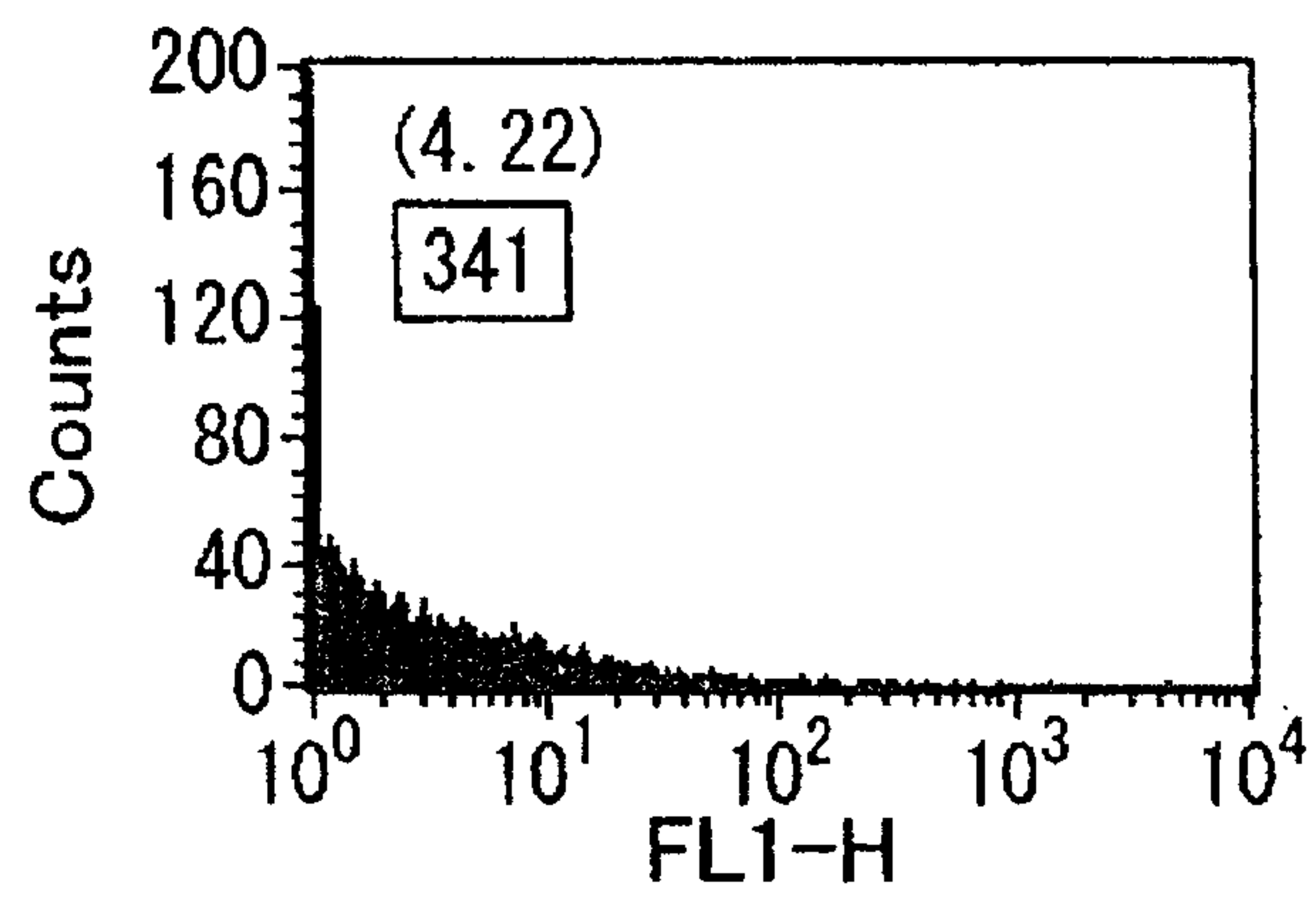
[圖]16(b)



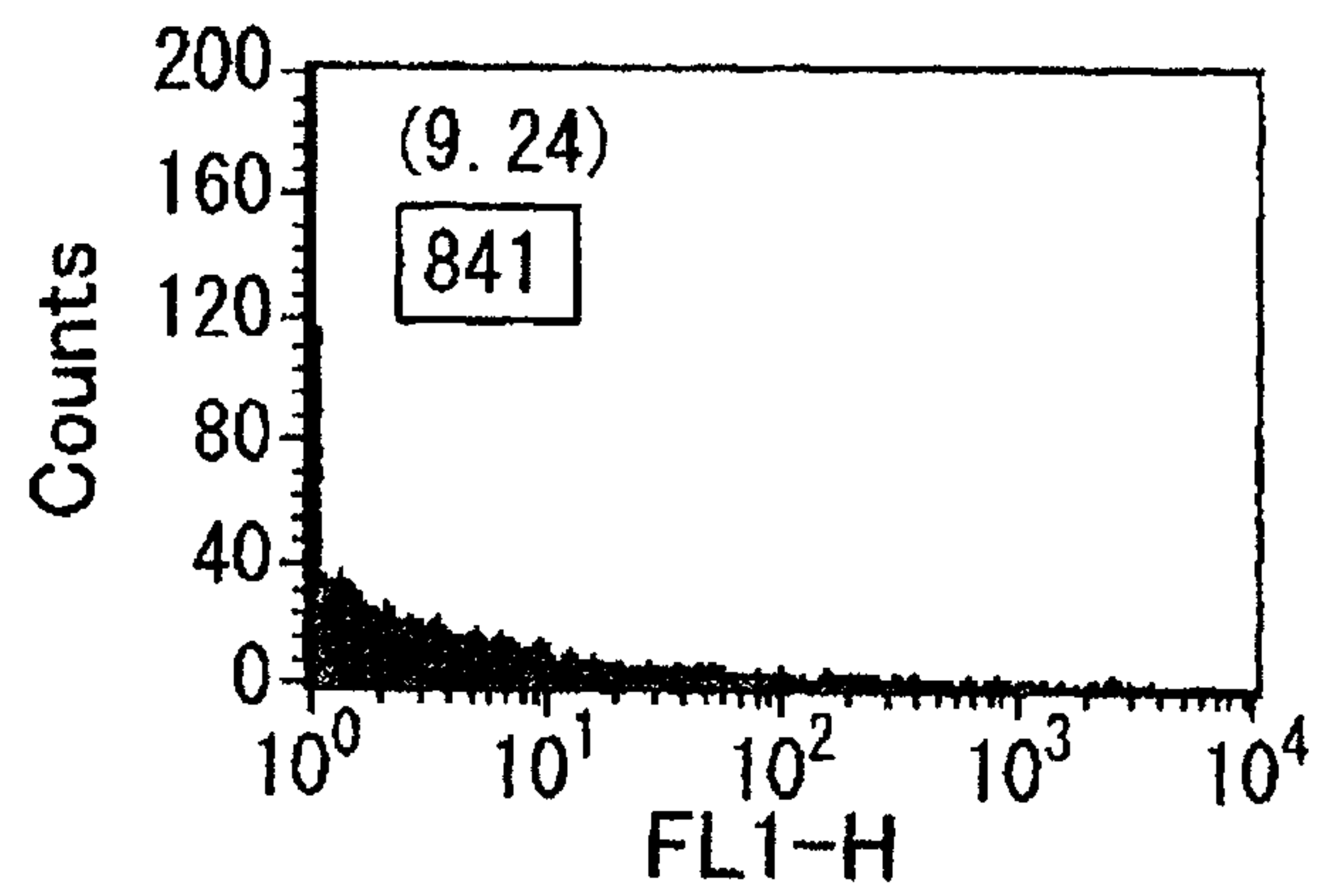
[圖]16(c)



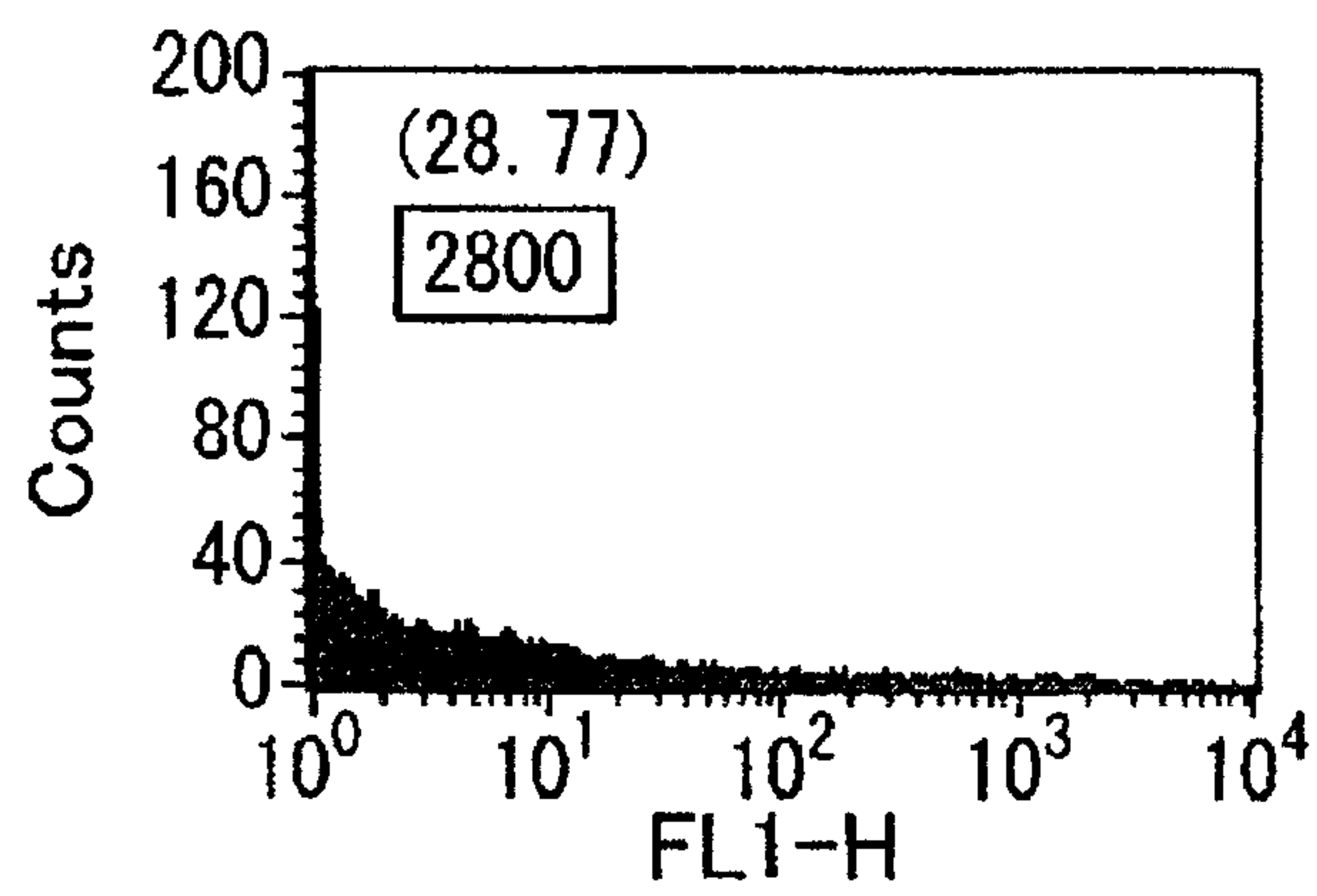
[図16(d)]



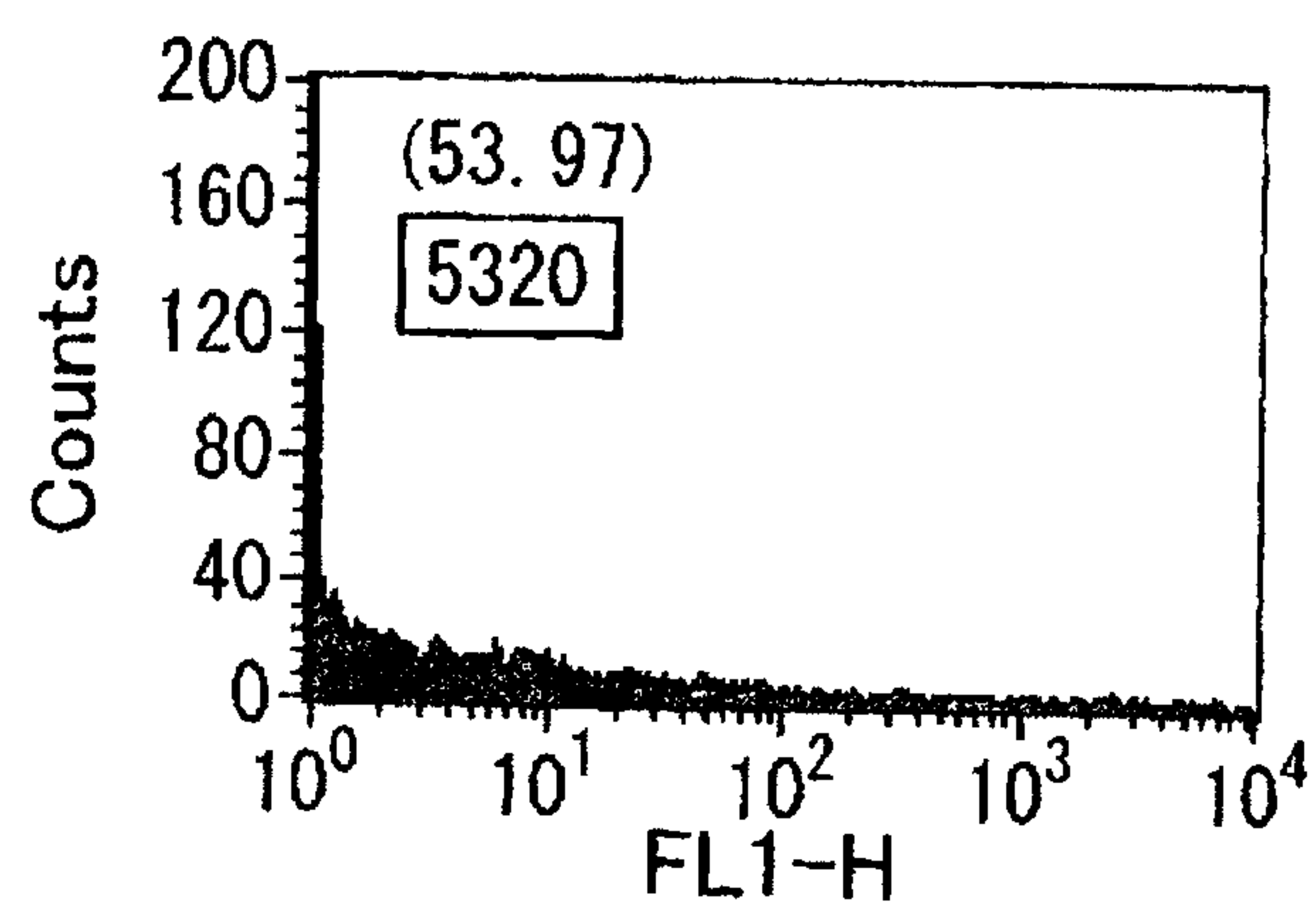
[図16(e)]



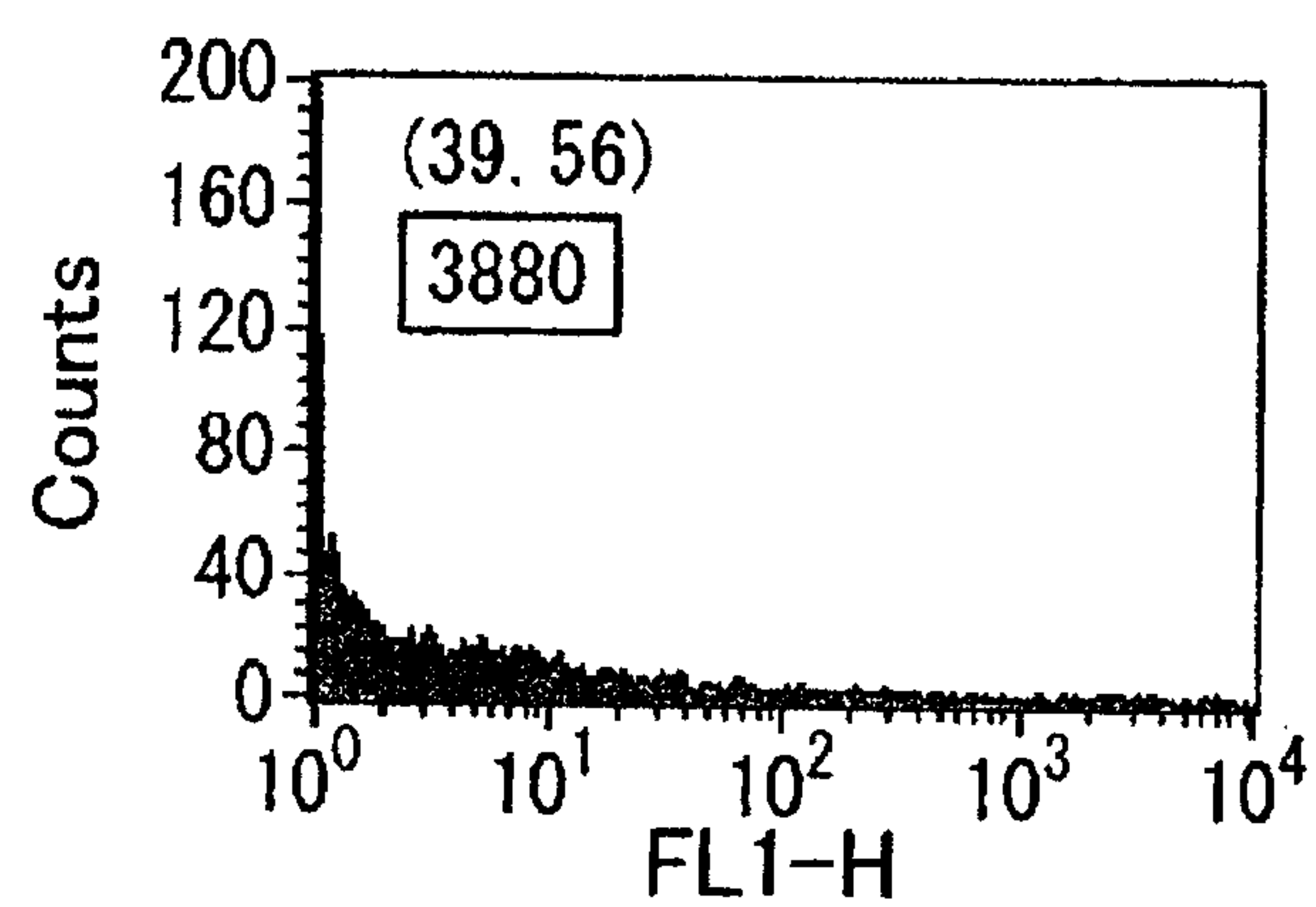
[図16(f)]



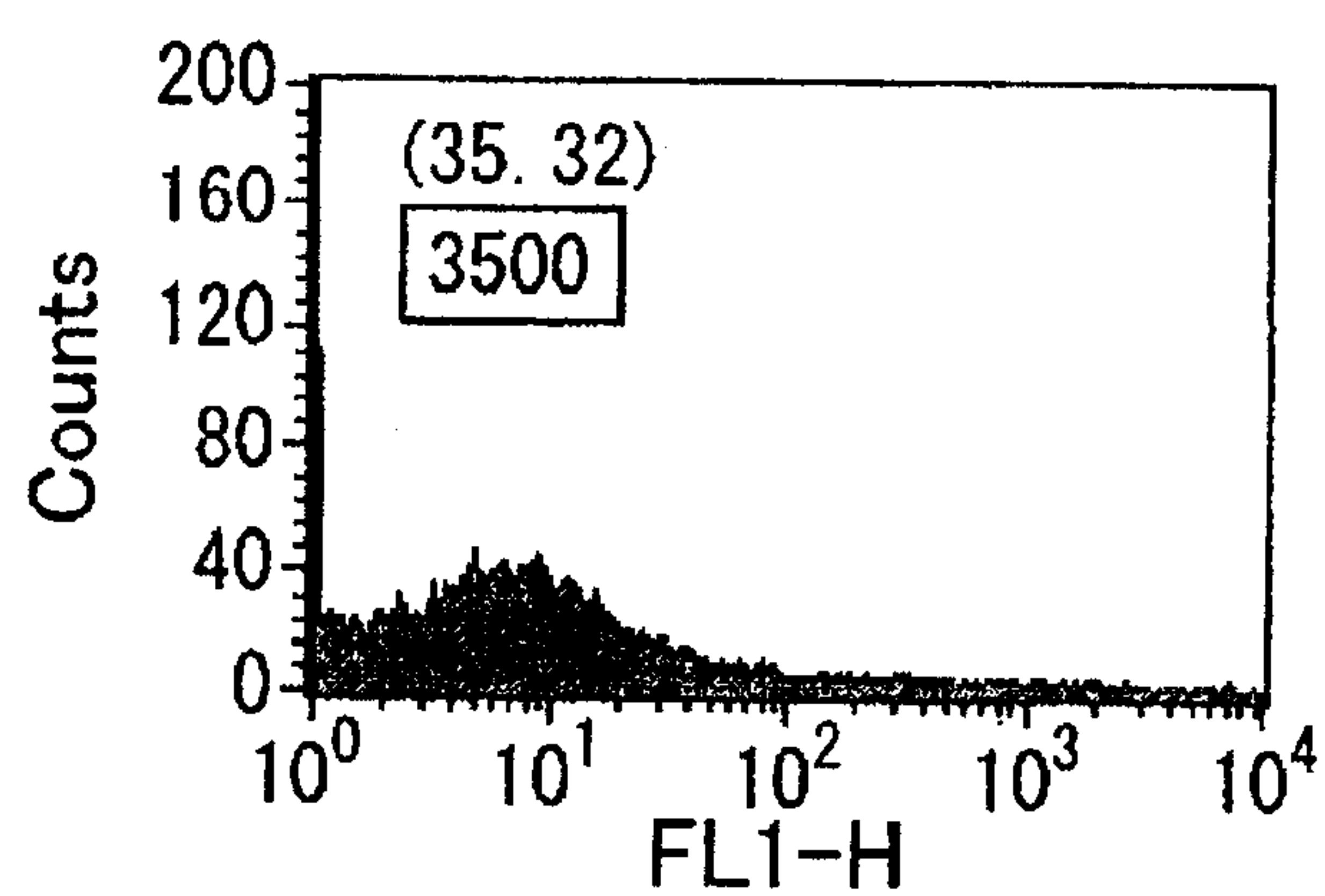
[圖16(g)]



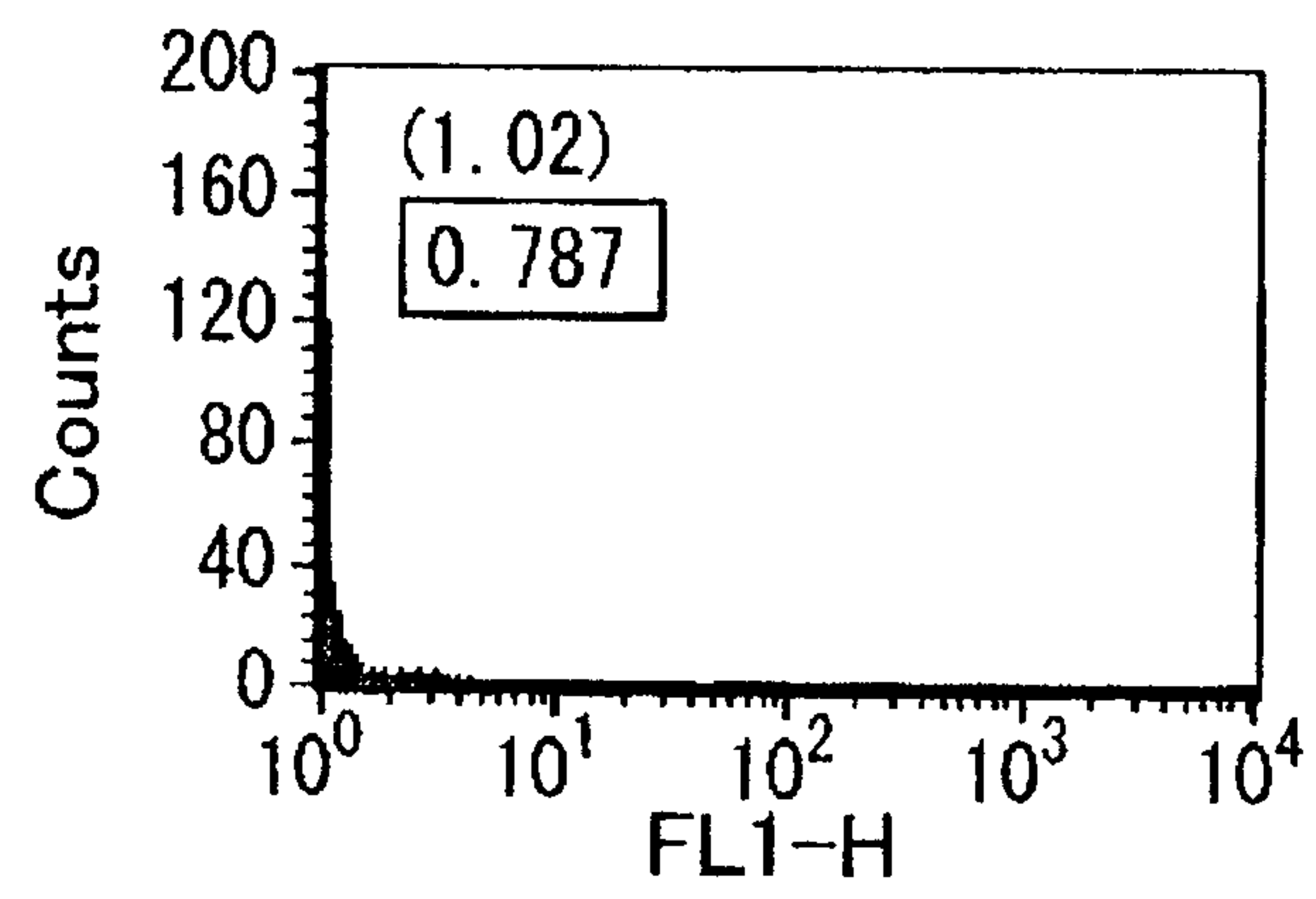
[圖16(h)]



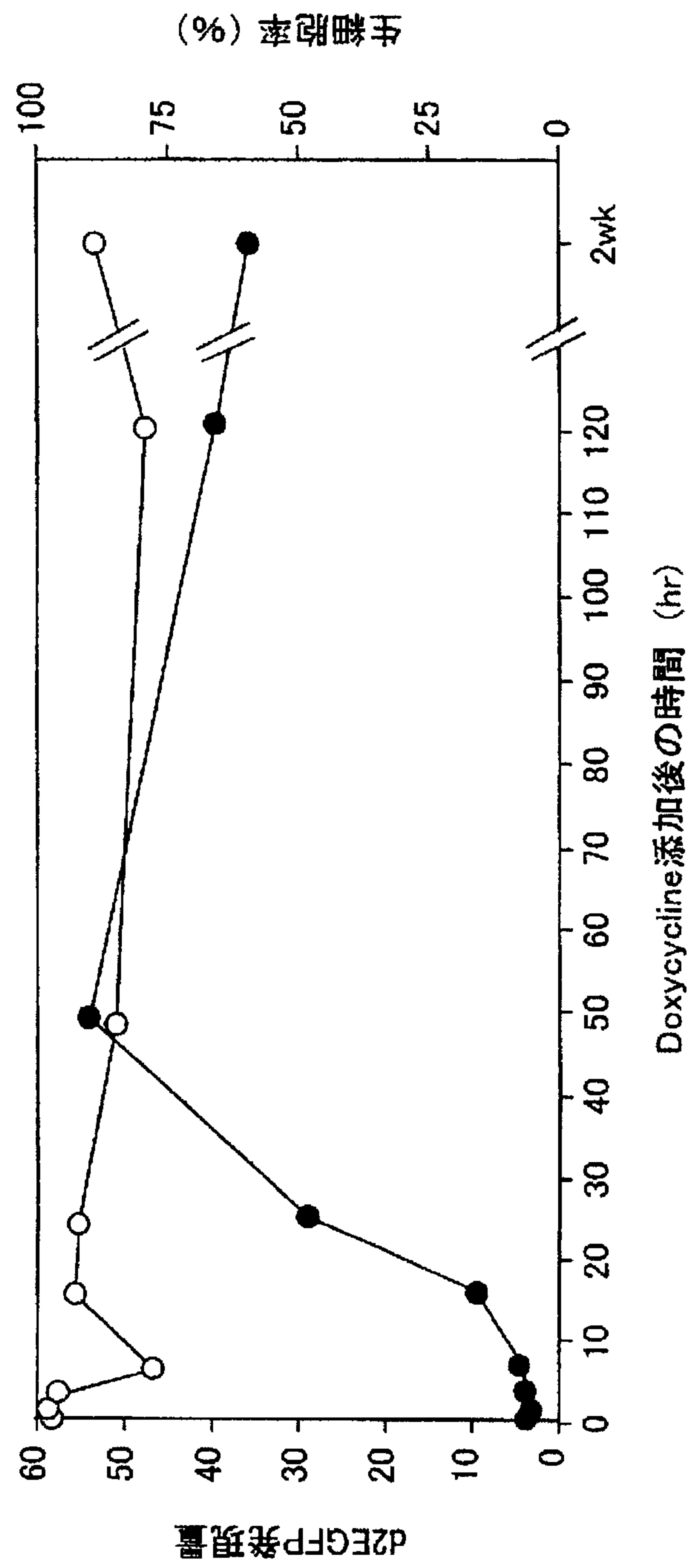
[圖16(i)]



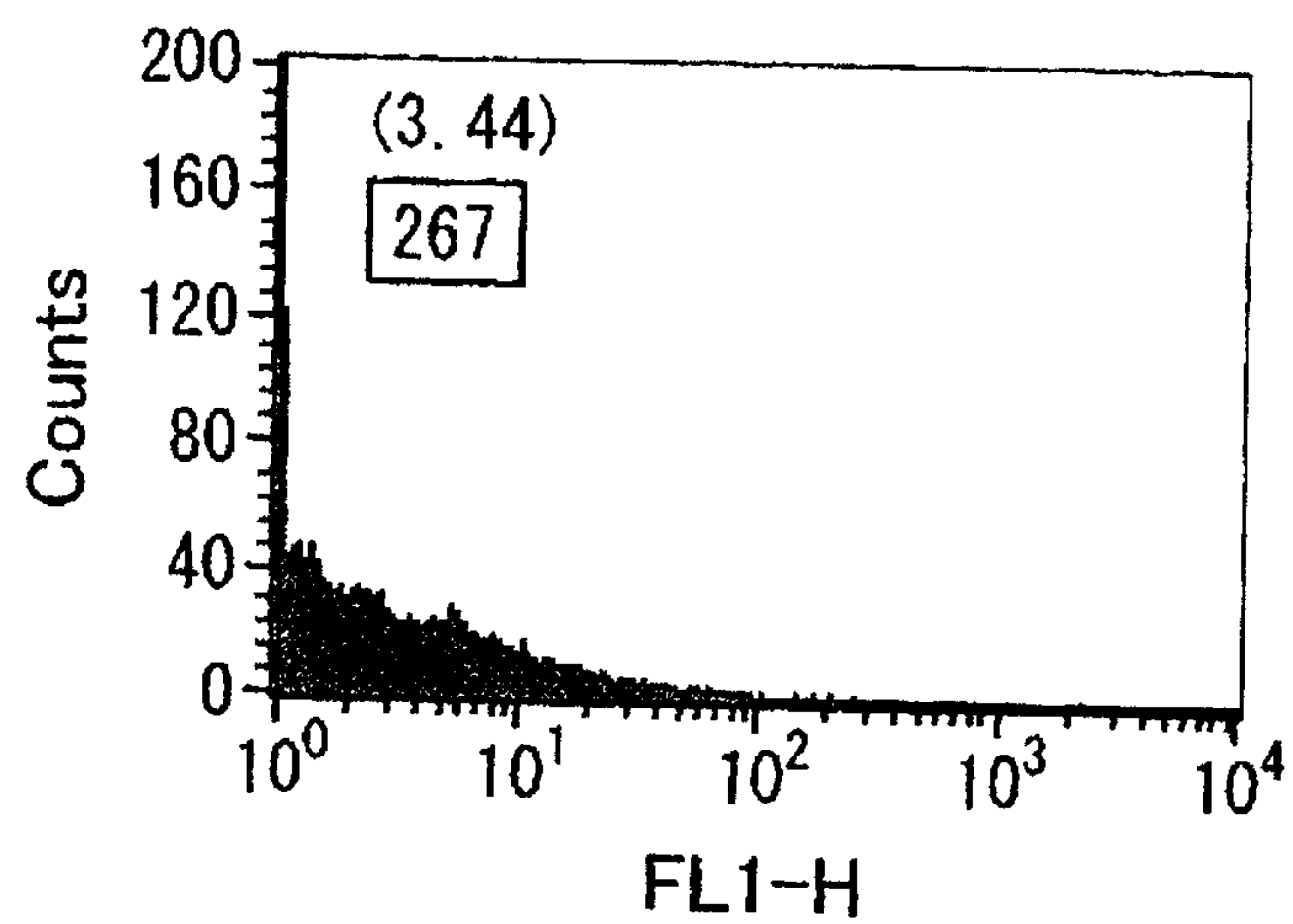
[16(j)]



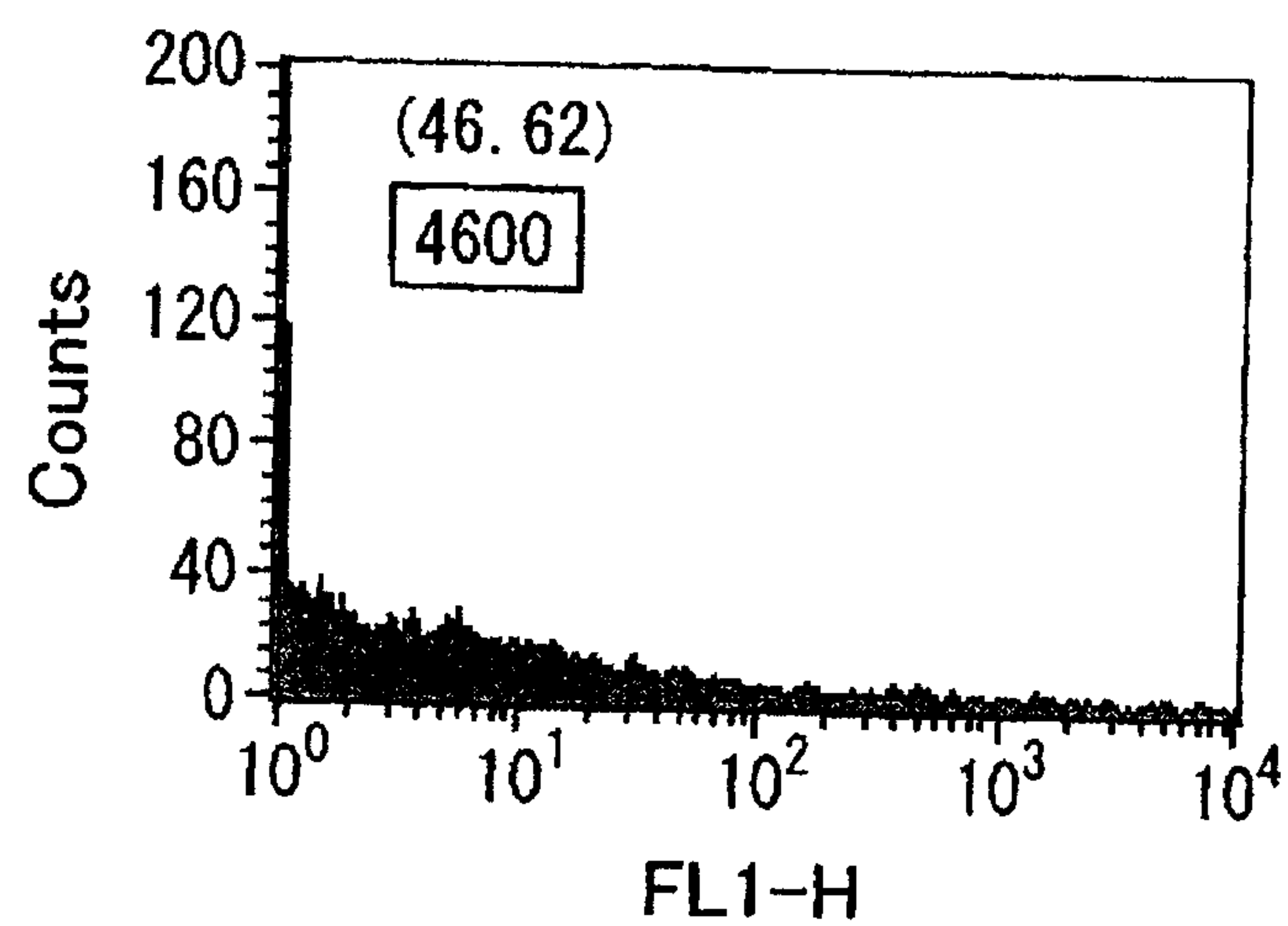
[図17]

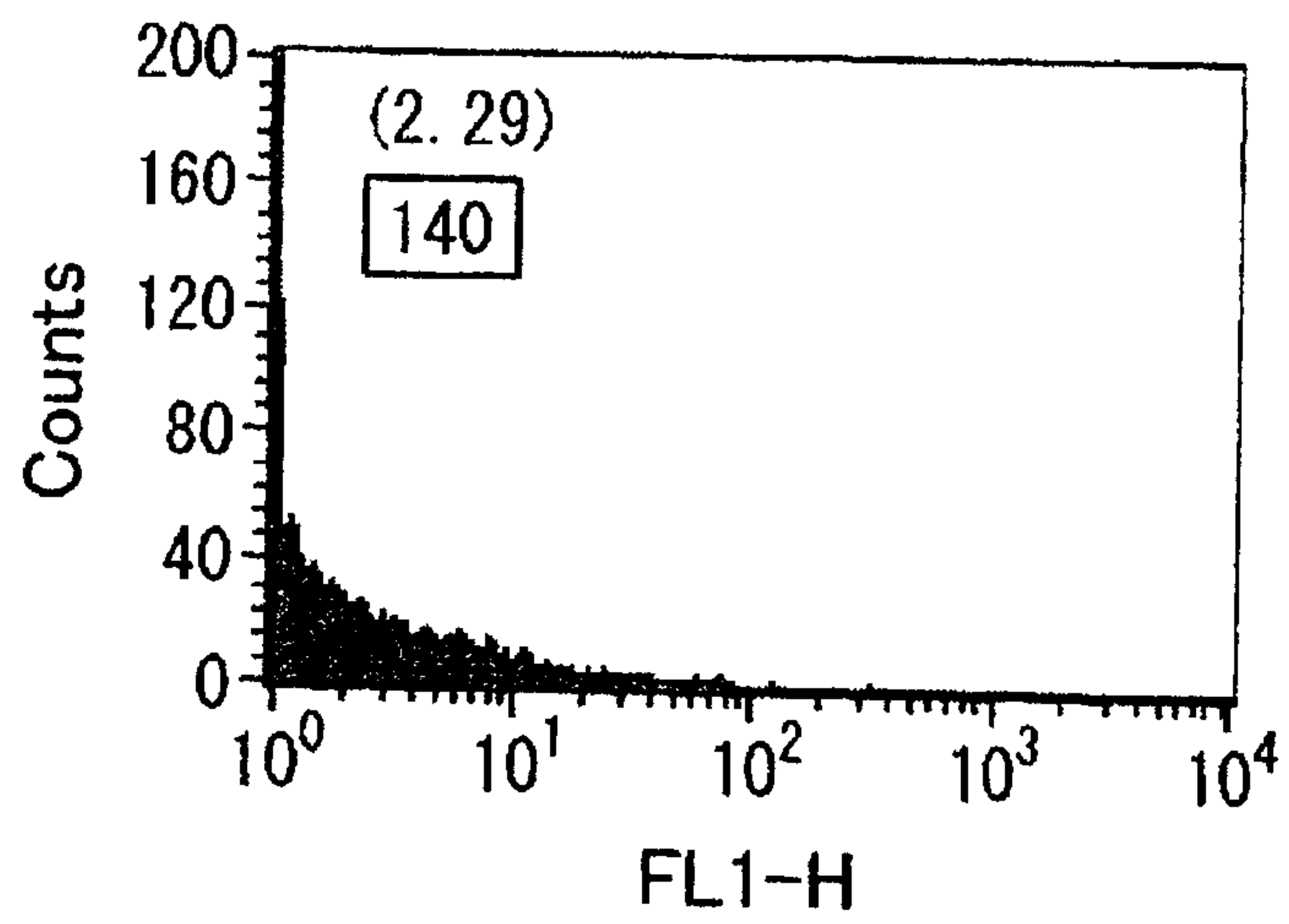
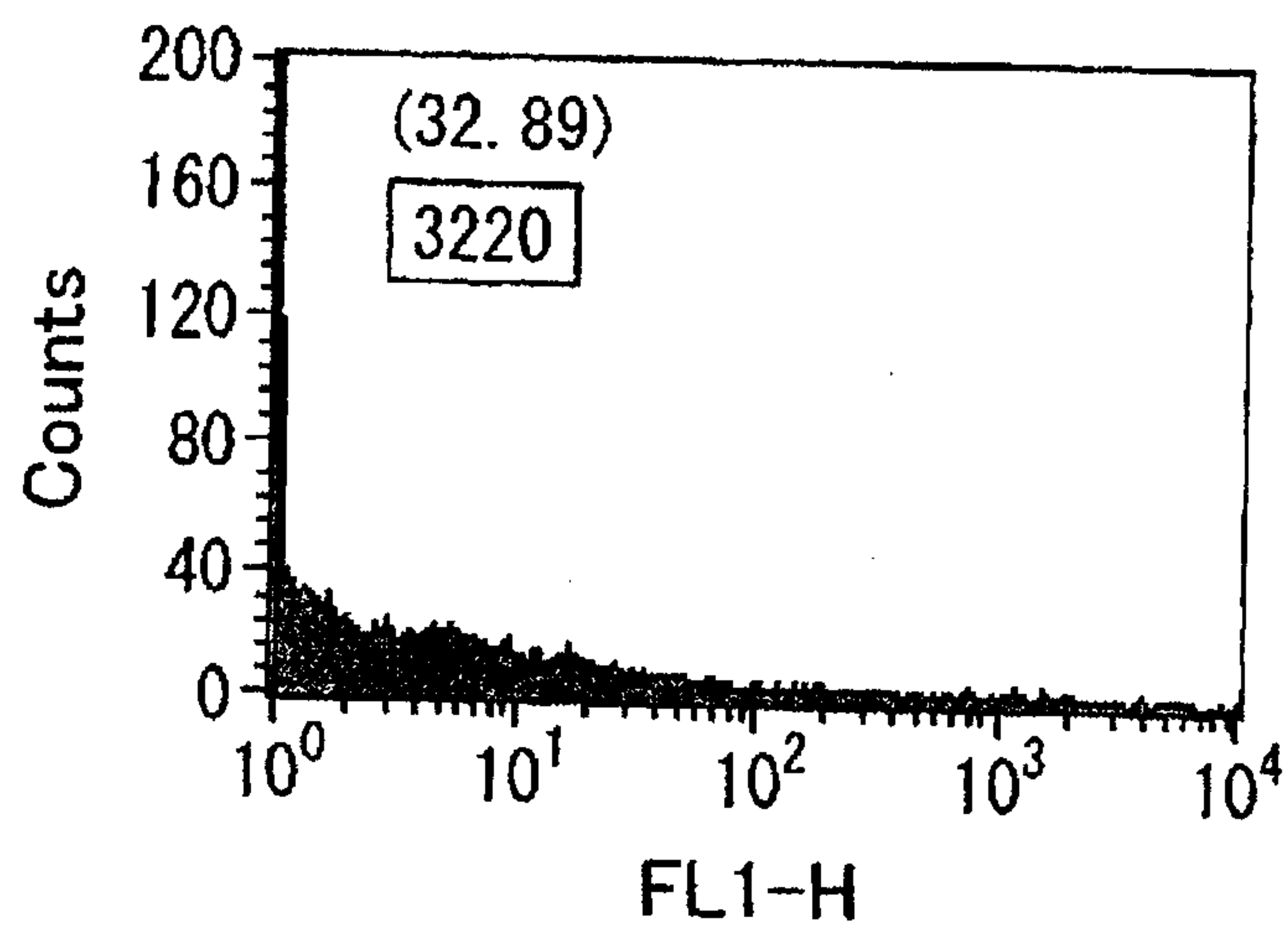


[図18(a)]

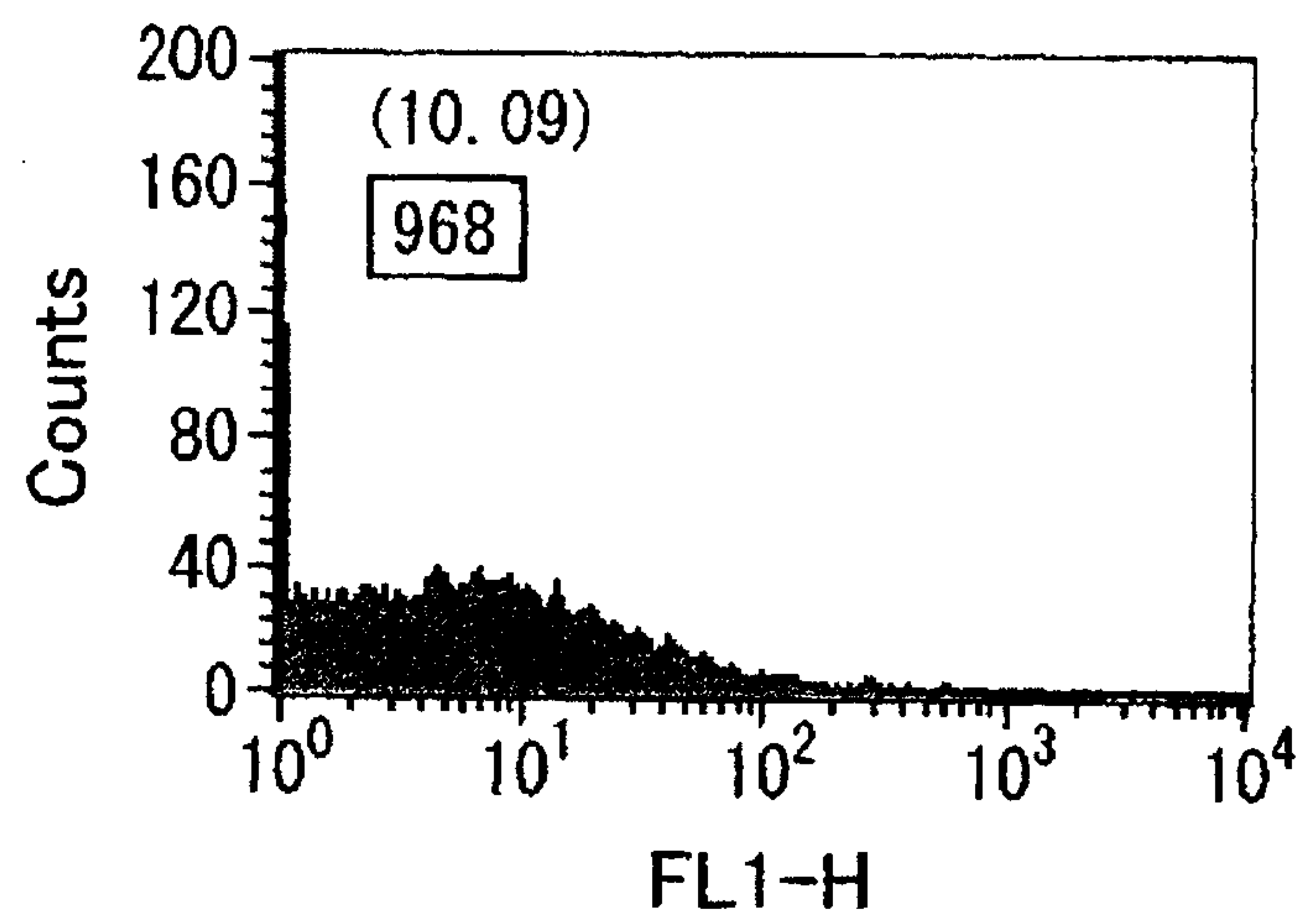


[図18(b)]

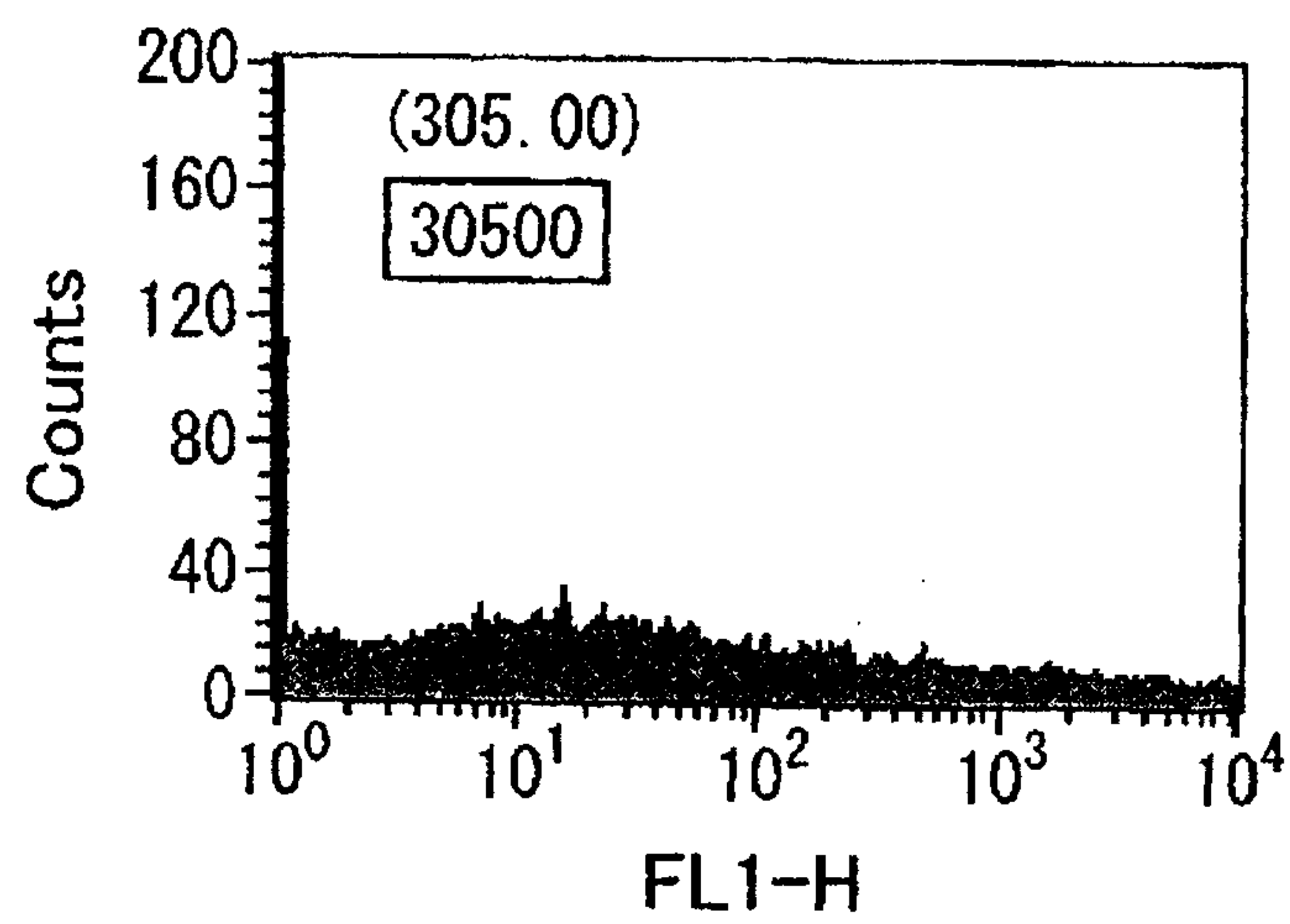


[18(c)][18(d)]

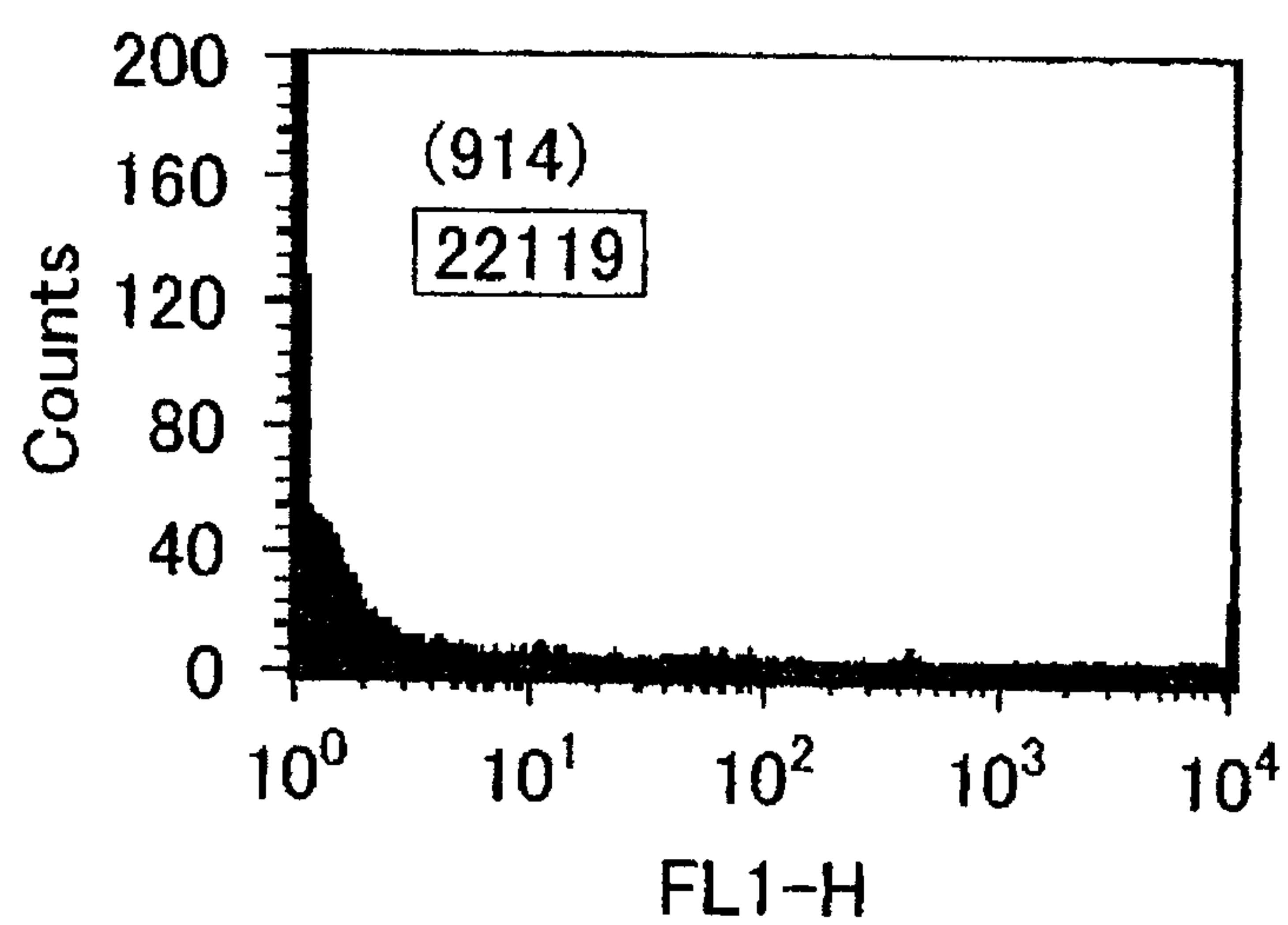
[圖18(e)]



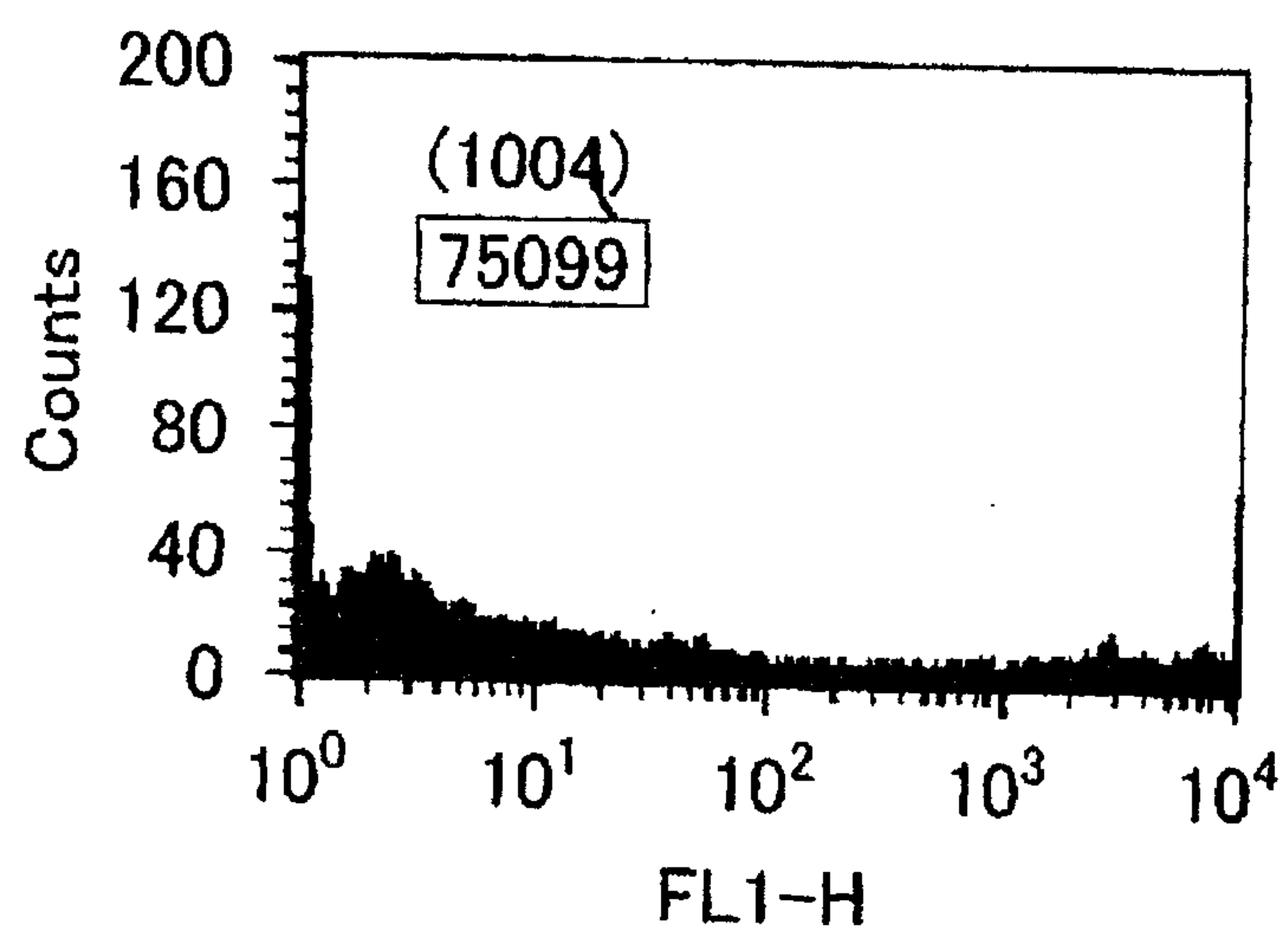
[圖18(f)]



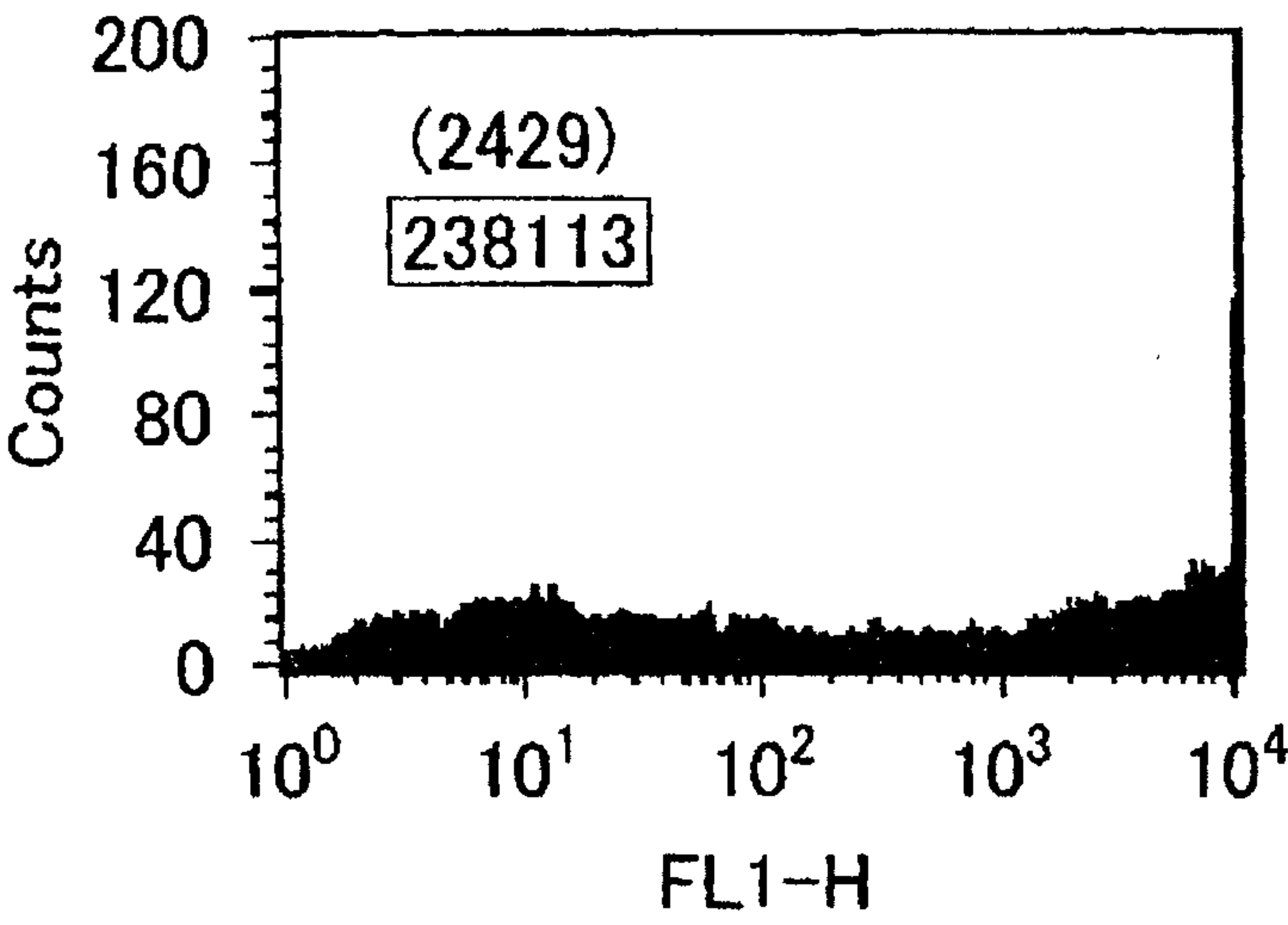
[图21(a)]



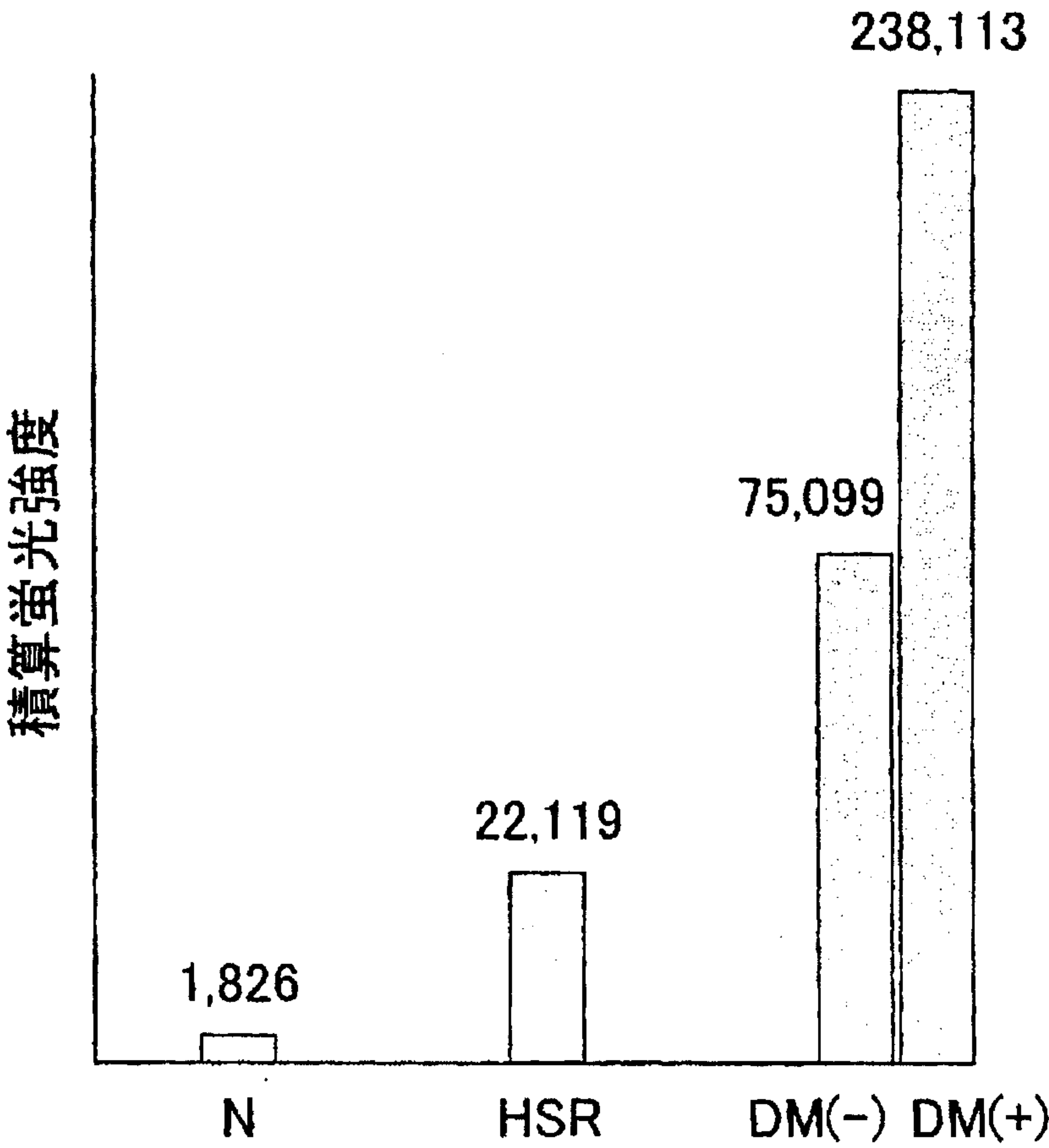
[图21(b)]



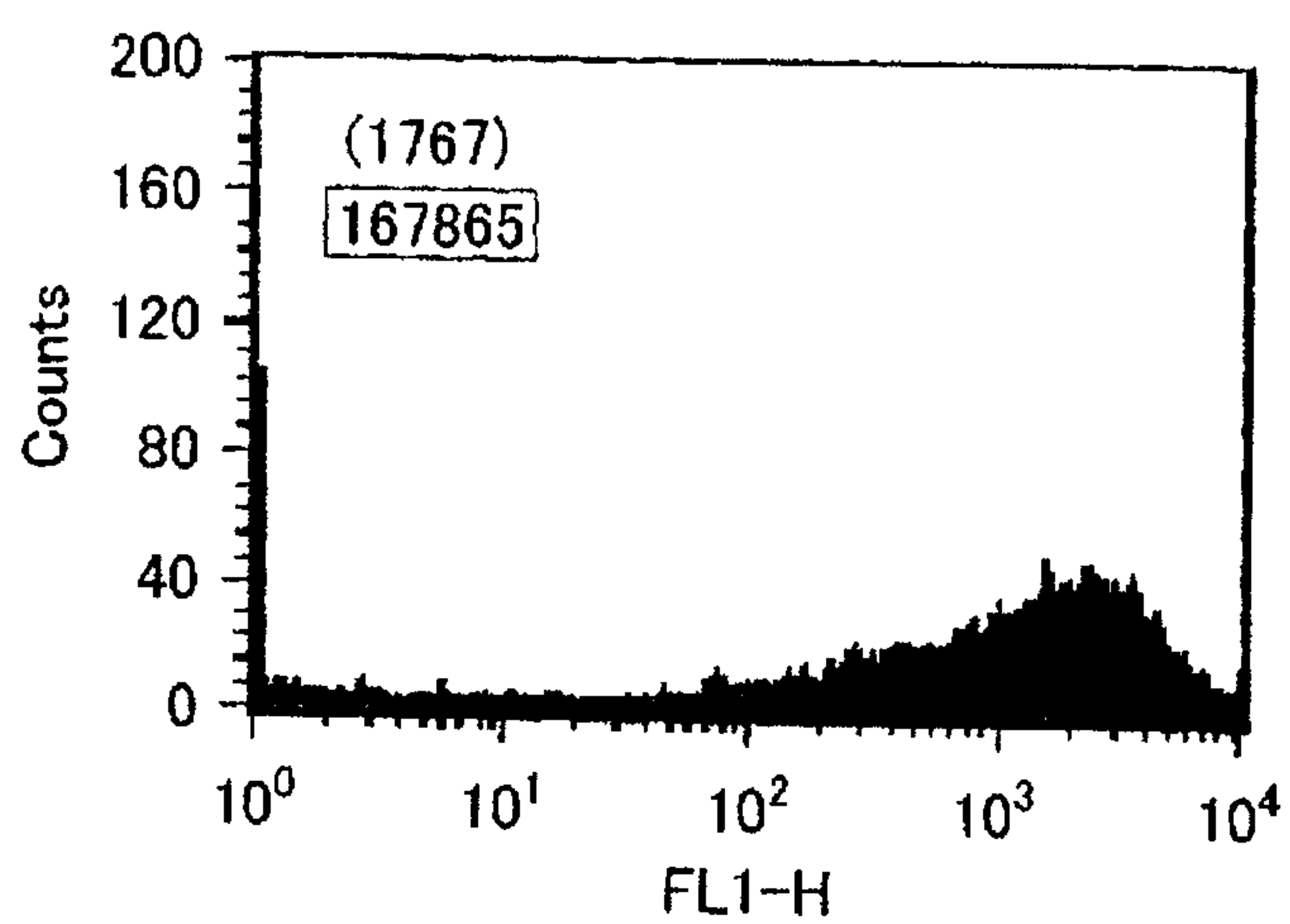
[図21(c)]



[図21(d)]



[図23(a)]



[図23(b)]

