



US 20090123530A1

(19) **United States**

(12) **Patent Application Publication**
Betageri et al.

(10) **Pub. No.: US 2009/0123530 A1**

(43) **Pub. Date: May 14, 2009**

(54) **LIPOSOME DRUG DELIVERY**

Related U.S. Application Data

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(60) Division of application No. 10/889,969, filed on Jul.
13, 2004, now Pat. No. 7,387,791, Continuation of
application No. 09/562,207, filed on May 2, 2000, now
Pat. No. 6,761,901.

Publication Classification

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(51) **Int. Cl.**
A61K 9/127 (2006.01)
A61P 35/00 (2006.01)

(52) **U.S. Cl.** **424/450**

(57) **ABSTRACT**

(21) Appl. No.: **12/135,759**

This invention comprises pharmaceutical compositions for administering a biologically active compound to an animal. Particularly provided are proliposomal compositions that are advantageously used to deliver biologically active compounds to the gastrointestinal tract after oral administration.

(22) Filed: **Jun. 9, 2008**

Figure 1A. Thermograms of DMPC-containing Formulations.

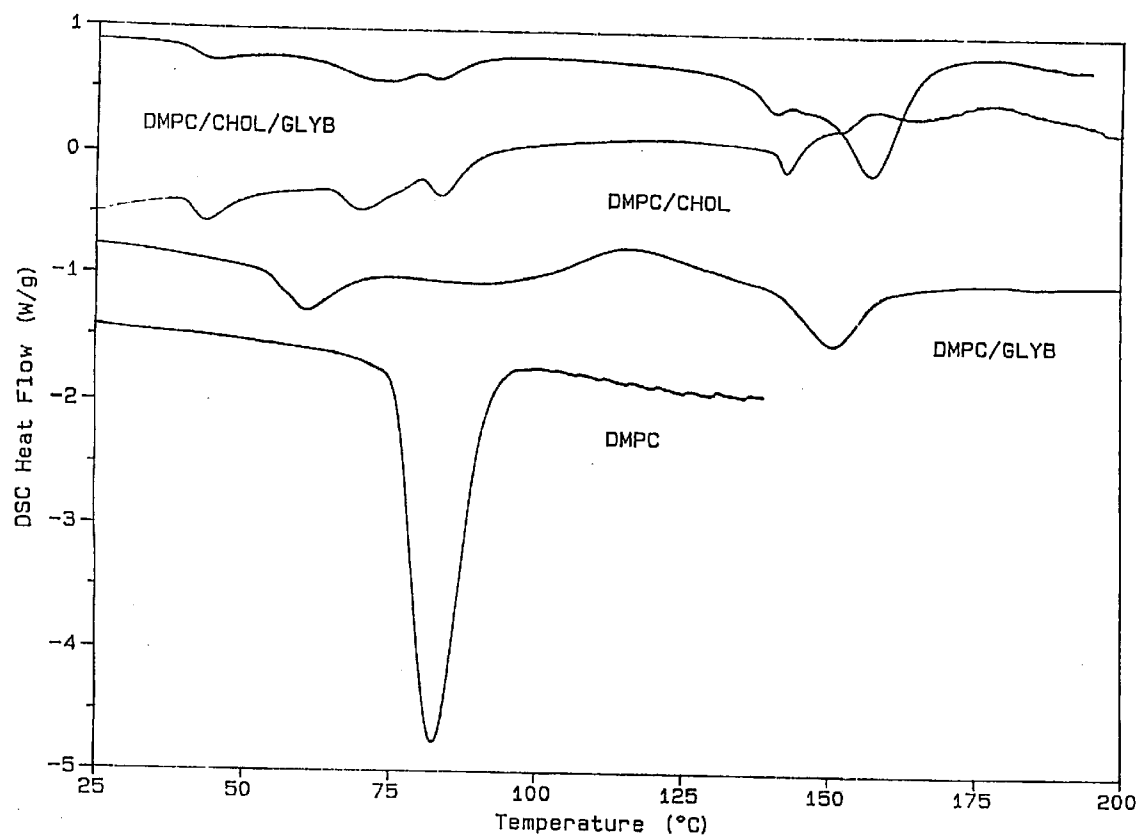


Figure 1B. Thermograms of DSPC-containing Formulations.

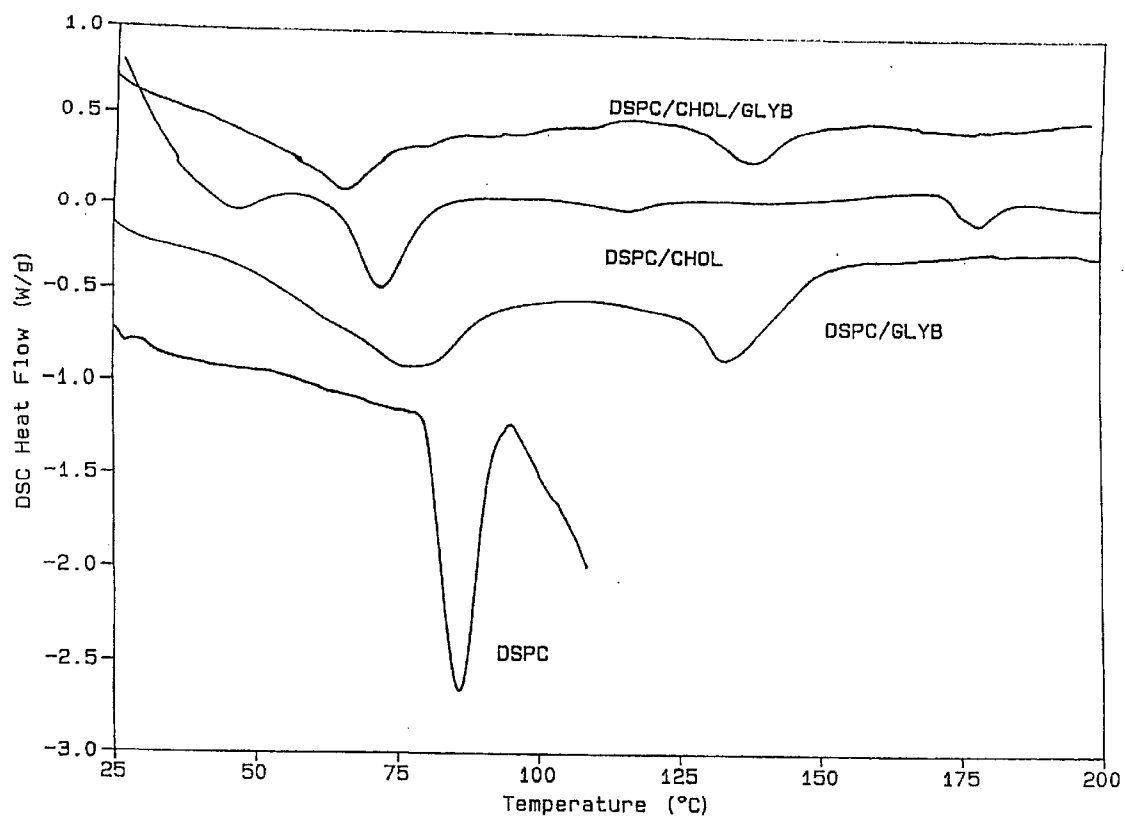


Figure 1C. Thermograms of Glyburide, Cholesterol, and Glyburide/Cholesterol Mixture

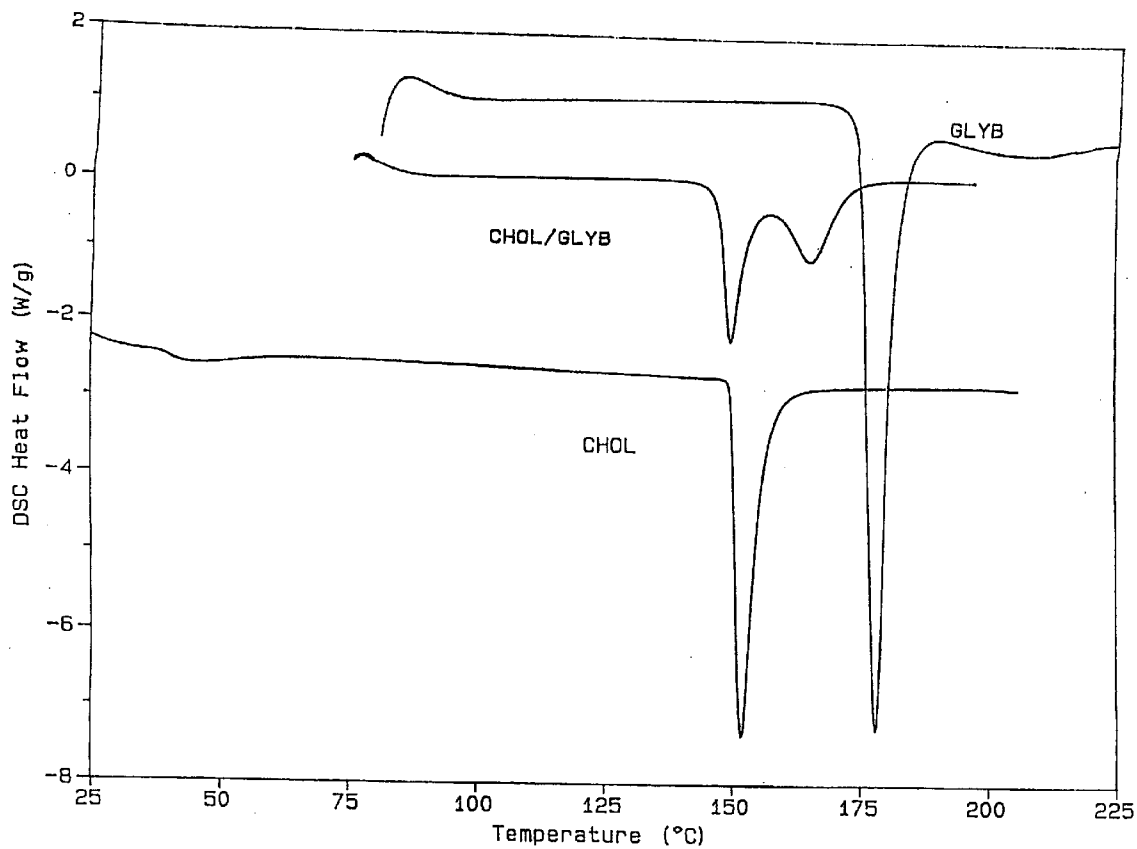


Figure 2. Glyburide flux across Caco-2 cells in DSPC formulations

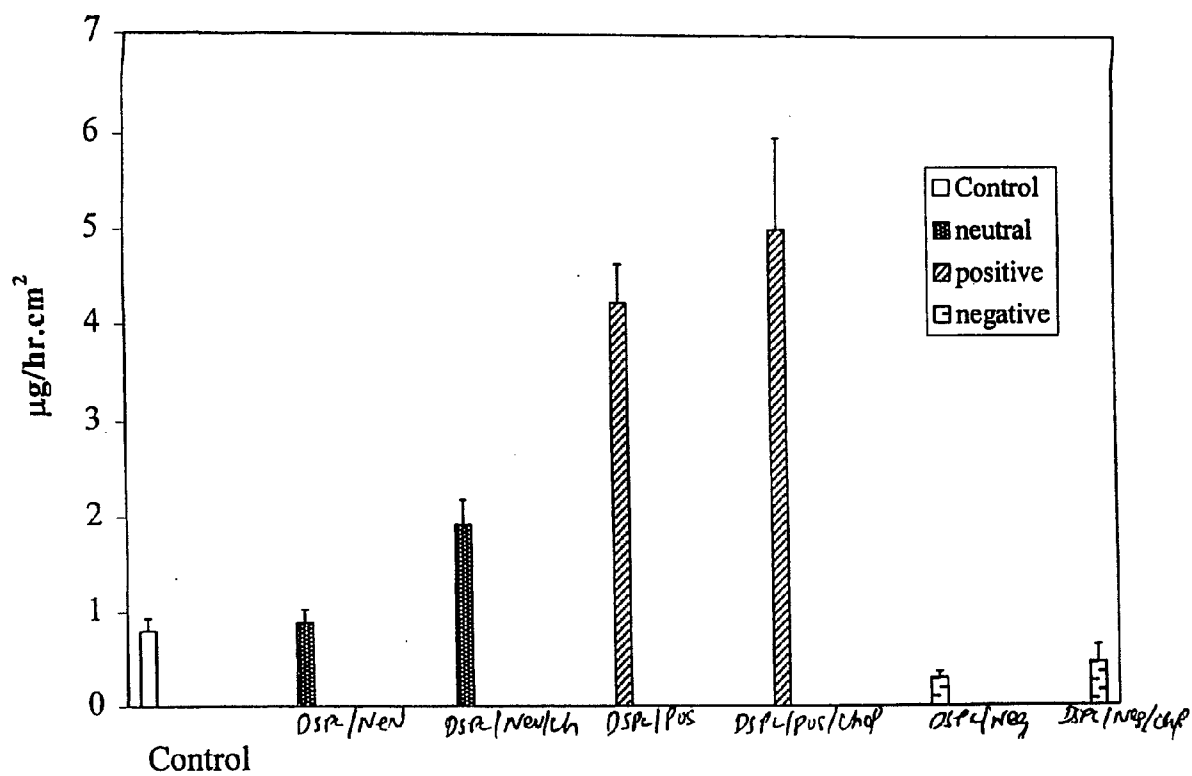


Figure-3. Glyburide Flux across Caco-2 cells in DMPC Formulations

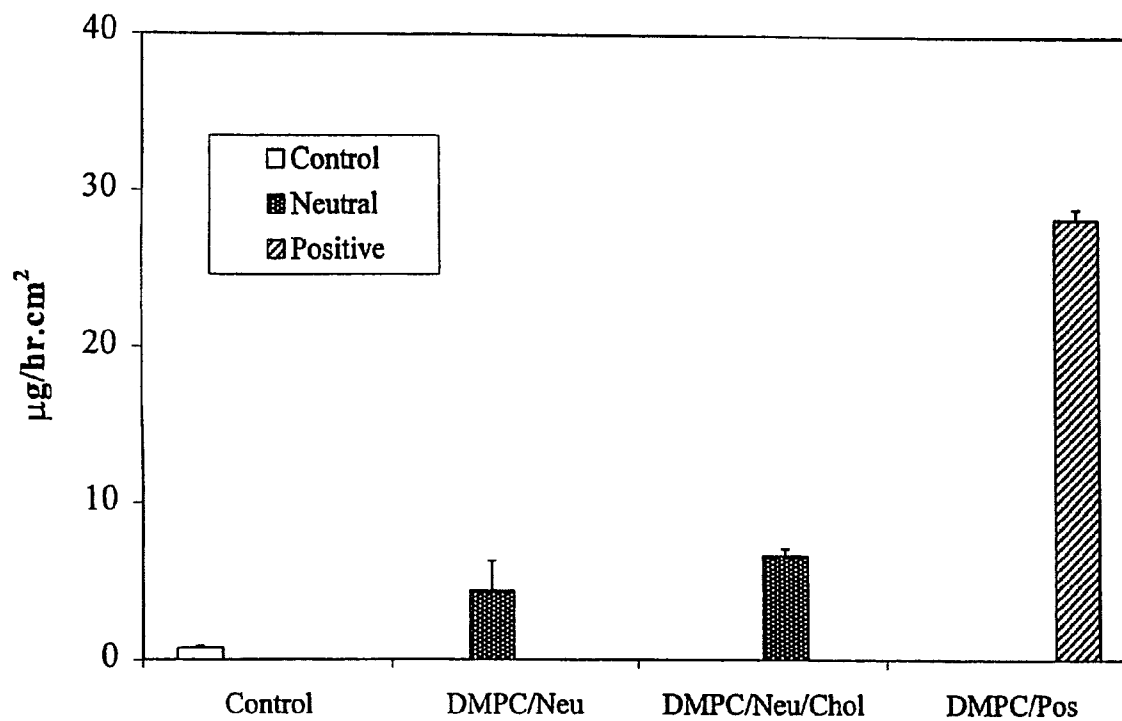


Figure 4. Cumulative amount of glyburide transported across Caco-2 cells with DSPC formulations

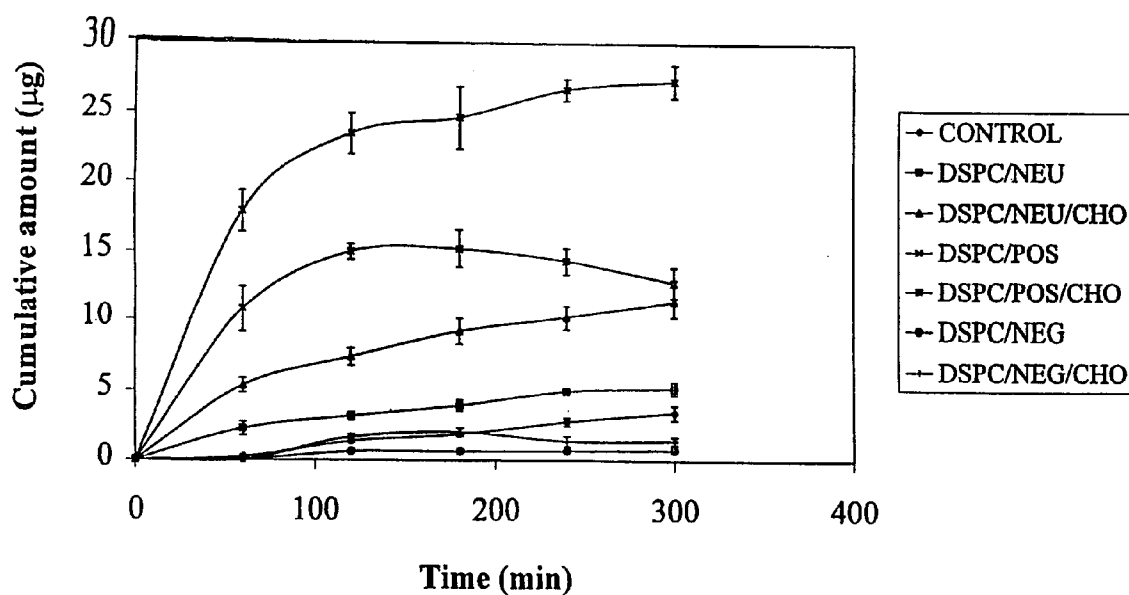
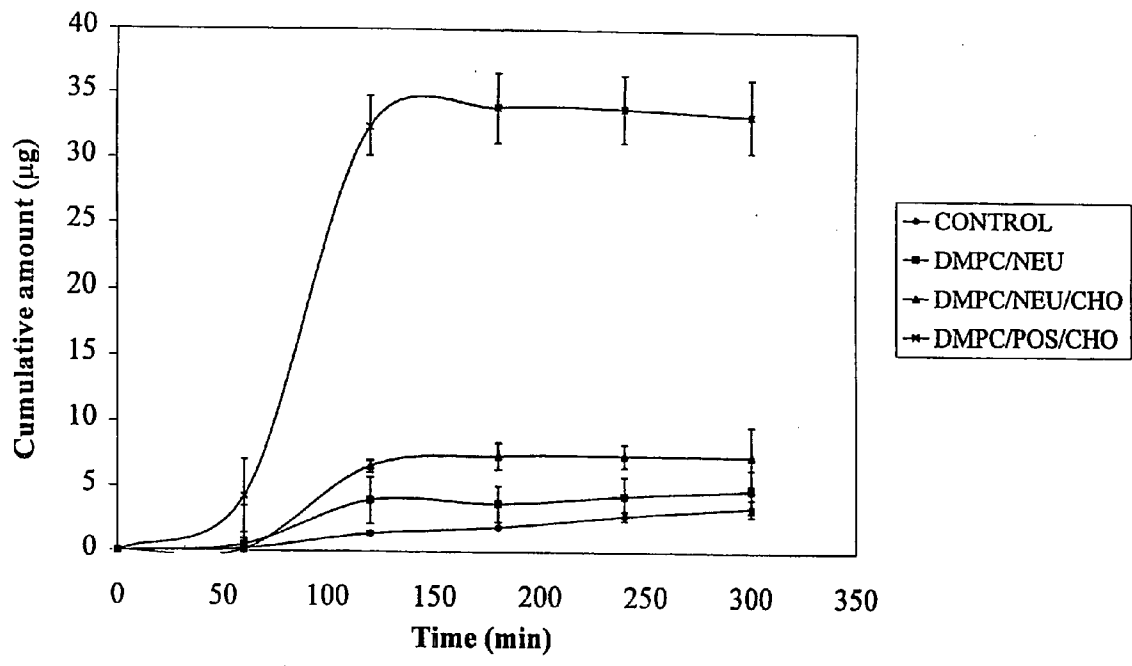


Figure 5. Cumulative Amount of Glyburide Transported across Caco-2 Cells with DMPC formulations



LIPOSOME DRUG DELIVERY

BACKGROUND OF THE INVENTION

[0001] 1. Field of the Invention

[0002] This invention relates to delivery of drugs, nutrients and other compounds to a biological organism. In particular, the invention relates to liposomes and formulations of drugs, nutrients and other compounds into liposomes to improve or effect delivery of such beneficial compounds to cells and tissues in an organism. Specifically, the invention provides such liposome compositions of drugs, nutrients and other compounds in formulations advantageously administered orally to an animal.

[0003] 2. Background of the Related Art

[0004] A major goal in the pharmacological arts has been the development of reagents and methods that reduce the necessity of administering therapeutic compounds, drugs and other agents invasively (i.e., such as by injection). Most preferably, it has been a consistent goal in the art to develop therapeutic compounds, drugs and agents and formulations thereof that permit oral administration (see, for example U.S. Pat. No. 4,963,526 to Ecanow issued Oct. 16, 1990), although other reduced-invasiveness formulations such as suppositories have also been developed. Among the various routes of drug administration, the oral intake of drugs is undoubtedly preferred because of its versatility, safety and patient comfort.

[0005] In addition, it has been a goal in the nutritional arts to develop preparations that increase transit of certain nutrients through the gastrointestinal tract to increase uptake and delivery of such nutrients into the bloodstream. In particular, such preparations have been developed to permit chemically-labile nutrients (such as vitamins and other sensitive compounds) to pass through the chemically-hostile environment **[text missing or illegible when filed]** stomach for absorption in the intestines (see, for example, U.S. Pat. No. 5,958,450 to **[text missing or illegible when filed]** issued Sep. 28, 1999). Preparations having enhanced intestinal uptake have also been deemed desirable.

[0006] One approach known in the prior art for improving efficiency of delivery of therapeutic compounds, drugs and other agents has been to envelop such compounds in a lipid structure termed a liposome (see, for example, U.S. Pat. No. 4,744,989 to Payne et al. issued May 17, 1988). Liposomes generically comprise an enclosed lipid droplet having a core, typically an aqueous core, containing the compound. In certain embodiments, the compound is chemically conjugated to a lipid component of the liposome. In other embodiments, the compound is simply contained within the aqueous compartment inside the liposome.

[0007] Certain liposome formulations are known in the art.

[0008] U.S. Pat. No. 5,223,263, issued Jun. 29, 1993 to Hostetler et al. discloses conjugates between antiviral nucleoside analogues and polar lipids for inclusion in liposomes.

[0009] U.S. Pat. No. 5,466,468 to Schneider et al. issued Nov. 14, 1995 discloses parenterally administrable liposome formulation comprising synthetic lipids.

[0010] U.S. Pat. No. 5,484,809, issued Jan. 16, 1996 to Hostetler et al. discloses taxol and taxol derivatives conjugated to phospholipids.

[0011] U.S. Pat. No. 5,580,571, issued Dec. 3, 1996 to Hostetler et al. discloses nucleoside analogues conjugated to phospholipids.

[0012] U.S. Pat. No. 5,626,869 to Nyqvist et al. issued May 6, 1997 discloses pharmaceutical compositions wherein the

pharmaceutically active compound is heparin or a fragment thereof contained in a defined lipid system comprising at least one amphiphatic and polar lipid component and at least one nonpolar lipid component.

[0013] U.S. Pat. No. 5,744,461, issued Apr. 28, 1998 to Hostetler et al. discloses nucleoside analogues conjugated to phosphonoacetic acid lipid derivatives.

[0014] U.S. Pat. No. 5,744,592, issued Apr. 28, 1998 to Hostetler et al. discloses nucleoside analogues conjugated to phospholipids.

[0015] U.S. Pat. No. 5,756,116, issued May 26, 1998 to Hostetler et al. discloses nucleoside analogues conjugated to phospholipids.

[0016] U.S. Pat. No. 5,843,509 to Calvo Salve et al. issued Dec. 1, 1998 discloses stabilization of colloidal systems through the formation of lipid-polysaccharide complexes comprising a water soluble and positively charged polysaccharide and a negatively charged phospholipid.

[0017] International Patent Application Publication Number WO89/02733, published April 1989 to Vical discloses conjugates between antiviral nucleoside analogues and polar lipids.

[0018] European Patent Application Publication Number 0350287A2 to Vical discloses conjugates between antiviral nucleoside analogues and polar lipids.

[0019] International Patent Application Publication Number WO93/00910 to Vical discloses conjugates between antiviral nucleoside analogues and polar lipids.

[0020] Rahman et al., 1982, *Life Sci.* 31: 2061-71 found that liposomes which contained galactolipid as part of the lipid appeared to have a higher affinity for parenchymal cells than liposomes which lacked galactolipid.

[0021] Gregoriadis, 1995, *Trends in Biotechnology* 13: 527-537 reviews the progress and problems associated with using liposomes for targeted drug delivery.

[0022] Ledley, 1995, *Human Gene Therapy* 6: 1129-1144 reviews the use of liposomes for gene therapy.

[0023] Mickisch, 1995, *World J. Urology* 13: 178-185 reviews the use of liposomes for gene therapy of renal cell carcinoma.

[0024] Yang et al. 1997, *J. Neurotrauma* 14: 281-297 review the use of cationic liposomes for gene therapy directed to the central nervous system.

[0025] Storm & Crommelin, 1997, *Hybridoma* 16: 119-125 review the preliminary use of liposomes for targeting chemotherapeutic drugs to tumor sites.

[0026] Manusama et al., 1998, *Semin. Surg. Oncol.* 14: 232-237 reported on preclinical and clinical trials of liposome-encapsulated tumor necrosis factor for cancer treatments.

[0027] Although liposomes have conventionally been administered parenterally (see, for example, U.S. Pat. No. 5,466,468), reports of oral administration of liposome-related formulations have appeared in the art.

[0028] U.S. Pat. No. 4,921,757 to Wheatley et al. issued May 1, 1990 discloses controlled release of biologically active substances, such as drugs and hormones entrapped in liposomes which are protected from the biological environment by encapsulation within semi-permeable microcapsules or a permeable polymeric matrix.

[0029] U.S. Pat. No. 5,043,165 to Radhakrishnan to Aug. 27, 1991 disclosed a liposome composition for sustained release of steroidal drugs.

[0030] U.S. Pat. No. 5,762,904 to Okada et al. issued Jun. 9, 1998 discloses oral delivery of vaccines using polymerized liposomes.

[0031] U.S. Pat. No. 5,955,451 to Lichtenberger et al. issued Sep. 21, 1999 discloses compositions comprising non-steroid anti-inflammatory drugs (NSAID's) complexed with either zwitterionic or neutral phospholipids, or both, having reduced gastrointestinal irritating effects and enhanced anti-pyretic, analgesic, and anti-inflammatory activity.

[0032] Proliposomes are an alternative to conventional liposomal formulations. Proliposomes are dry, free-flowing granular products, which, on addition of water, disperse to form a multi-lamellar liposomal suspension. The stability problems associated with conventional liposomes such as aggregation, susceptibility to hydrolysis and/or oxidation are avoided by using proliposomes.

[0033] U.S. Pat. No. 5,635,206 to Ganter et al. issued Jun. 3, 1997 discloses a process for preparing liposomes or proliposomes.

[0034] Proliposomes of indomethacin were prepared using effervescent granules, which upon hydration yielded liposomes of high encapsulation efficiency and increased anti-inflammatory activity with decreased ulcerogenic index (see, for example, Katare et al., 1991, *J. Microencapsulation* 81: 1-7).

[0035] The proliposomal concept has been extended to administer drugs through various routes and also to the food industry wherein enzyme immobilization is essential for various food processing regimes. A typical example is the immobilization of the enzyme, chymotrypsin, in liposomes obtained from proliposomes.

[0036] There remains a need in the art for a general, inexpensive and effective means for delivering biologically-active compounds, including drugs, hormones, enzymes, genetic material, antigens including vaccines, and nutrients, to an animal by oral administration. Advantageous embodiments of such delivery means are formulated to efficiently deliver biologically-active compounds to the appropriate portion of the gastrointestinal tract for efficient absorption.

SUMMARY OF THE INVENTION

[0037] The present invention is directed to an improved method for delivering biologically-active compounds, particularly drugs, hormones, enzymes, genetic material, antigens including vaccines, and nutrients, to an animal by oral administration. This delivery system achieves specific delivery of such biologically-active compounds through associating the compounds with liposomes and proliposome components.

[0038] In preferred embodiments, the biologically active compound is formulated as a proliposomal composition that can be reconstituted in vivo to provide a liposomal preparation. Preferably, the invention provides pharmaceutical compositions comprising the biologically active compound and a lipid formulated as a proliposomal preparation. In more preferred embodiments, the pharmaceutical compositions of the invention are formulated for oral administration. Most preferably, the pharmaceutical compositions of the invention formulated for oral administration comprise an enteric coating sufficient to prevent dissolution of the composition in the stomach of an animal. In alternative embodiments, the pharmaceutical compositions also comprise a protective coating between the enteric coating and the core of the composition comprising the proliposomal components thereof. Additional

advantageous components of said orally-administrable pharmaceutical compositions further comprise the pharmaceutical compositions as will be understood by those with skill in the art.

[0039] Specific preferred embodiments of the present invention will become evident from the following more detailed description of certain preferred embodiments and the claims.

BRIEF DESCRIPTION OF THE DRAWINGS

[0040] FIGS. 1A through 1C depict thermograms produced by differential scanning calorimetry as set forth in Example 1.

[0041] FIGS. 2 and 3 depict transfer rates of glyburide through a Caco-2 cellular monolayer using the liposomal compositions of the invention, as set forth in Example 2.

[0042] FIGS. 4 and 5 depict total accumulation of glyburide in the receiving chamber of a transwell comprising a Caco-2 cellular monolayer using the liposomal compositions of the invention, as set forth in Example 2.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

[0043] The present invention provides compositions of matter and methods for facilitating the delivery of biologically-active compounds to the tissues of an animal after oral administration. For the purposes of this invention, the term "biologically-active compound" is intended to encompass all naturally-occurring or synthetic compounds capable of eliciting a biological response or having an effect, either beneficial or cytotoxic, on biological systems, particularly cells and cellular organelles. These compounds are intended to include but are not limited to all varieties of drugs, including but not limited to antibiotic, antibacterial, antiviral, antimycotic, antiproliferative and antineoplastic drugs; hormones, including peptide hormones and steroid hormones, and most particularly including endocrine and exocrine gland hormones; genes, recombinant nucleic acids, oligonucleotides or other nucleic acids encoding all or a portion of a mammalian gene, a viral gene or a gene from a microorganism; antigens, particularly in the form of vaccines; enzymes, particularly digestive enzymes and most particularly enzymes involved in processing, modifying, converting or degrading a nutrient into a form more easily absorbed by an animal's gastrointestinal tract; nutrients, and most preferably vitamins and minerals; and most particularly any biologically active compound, including particularly nutrients, that inefficiently transits the gastrointestinal tract or is unstable in a compartment thereof.

[0044] Pharmaceutical compositions comprising the biologically active compounds of the invention are preferably provided as proliposomal compositions that can be reconstituted, most preferably in vivo, to produce liposomal compositions of the biologically active compounds. As used herein, the term "proliposome" and "proliposomal" are intended to encompass dry, free-flowing granular products, which, on addition of water, disperse to form multi-lamellar liposomal suspensions comprising the biologically active compounds of the invention. Advantageously, the stability problems associated with the conventional liposomes (such as aggregation, susceptibility to hydrolysis and oxidation) are avoided by using proliposomes

[0045] The proliposomal compositions provided by the invention are reconstituted, particularly in vivo, to provide liposomal compositions wherein the biologically active com-

pounds of the invention are encapsulated in said liposomes. In preparing the proliposomal compositions of the invention, lipid components including neutral lipids, positively-charged lipids, negatively-charged lipids, amphoteric lipids such as phospholipids, and cholesterol are advantageously used. As defined herein, the "lipid component" of the proliposomal compositions of the invention are intended to encompass a single species of lipid (such as a particular phospholipid) or combinations of such lipids, either of one type such as combinations of phospholipids (for example, phosphatidylcholine plus phosphatidyl ethanolamine) or of different types (such as a phospholipid plus a charged lipid or a neutral lipid). Combinations comprising a multiplicity of different lipid types are also advantageously encompassed by the proliposomal compositions of the invention (see, Lehninger, 1975, *Biochemistry*, 2d ed., Chapters 11 & 24, Worth Publishers: New York; and Small, 1986, "From alkanes to phospholipids," *Handbook of Lipid Research: Physical Chemistry of Lipids*, Volume 4, Chapters 4 and 12, Plenum Press: New York).

[0046] Biologically active compounds that are unstable in the stomach, such as proteins and peptides, vitamins and other small molecule nutrients, or biologically active compounds that irritate the stomach, such as various analgesics like aspirin, and those compounds that are preferentially absorbed in the small intestine are preferred biological compounds useful with the liposomal formulations of the invention. In preferred embodiments, said compounds include but are not limited to aspirin, ibuprofen, erythromycin, vasopressin, insulin, dideoxyinosine (ddI), cyclosporine, taxol, heparin, halofantrine, ethopropazine, griseofulvin, propofol, furosemide, carbamazepine, diazepam, candesartan and cilexetil.

[0047] The proliposomal preparations comprising the biologically active compounds of the invention are preferably provided in a form that can be orally administered, including but not limited to syrups, elixirs, capsules, tablets, and emulsions. Preferred forms are tablets or capsules, most preferably comprising an enteric coating to prevent premature dissolution under the chemically harsh environment of the stomach. Enteric coatings are prepared as will be understood by one having skill in the art, and preferably include coatings including but not limited to eudragit and cellulose acetate phthalate.

[0048] In alternative embodiments, the tablets or capsules of the invention comprise a protective coating between the enteric coating and the core of the capsule or tablet comprising the proliposomal preparations of the invention. In such embodiments, the protective coating is prepared as will be understood by one having skill in the art, and preferably include coatings including but not limited to hydroxypropyl methylcellulose, polyethylene glycol and ethylcellulose. In additional embodiments, the protective coating further comprises a plasticizing agent, including but not limited to triethylcitrate and polyvinyl pyrrolidone.

[0049] The tablets, capsules and other like embodiments of the proliposomal preparations and pharmaceutical compositions of the invention further advantageously comprise particle lubricants that minimize the tendency of the granular proliposomal compositions to agglomerate. By "particle lubricant" as used herein is meant the class of materials used in the manufacturing of pharmaceutical tablets as lubricants to improve the flowability and prevent agglomeration of an active agent during the tableting process. Examples of particle lubricants include talc, lactose, corn starch, ethyl cellu-

lose, fatty acid salts such as magnesium stearate, agar pectin, fatty acids such as stearic acid, gelatin and acacia.

[0050] The invention specifically provides methods for preparing and administering the proliposomal compositions of the invention as disclosed in the Examples below, and pharmaceutical compositions comprising the proliposomal preparations of biologically active compounds.

[0051] Animals to be treated with the proliposomal preparations and pharmaceutical compositions of the invention are intended to include all vertebrate animals, preferably domesticated animals, such as cattle, horses, goats, sheep, fowl, fish, household pets, and others, as well as wild animals, and most preferably humans.

[0052] One advantage of orally-administered liposomal formulations over parenterally-administered formulations is that oral administration reduces uptake of liposomes by the liver, thus reducing liver toxicity (which is a particular liability of parenterally-administered liposomal formulations). Oral formulations are targeted to deliver biologically active compounds to the intestine, which is a large surface for absorption and results in slow release of the administered compound. Finally, oral administration avoids transport-mediated saturation of drugs like dideoxyinosine.

[0053] The formulations of the invention are advantageously used for treating diseases that cause or result in malabsorption, including but not limited to Crohn's disease, irritable bowel syndrome, celiac sprue, diverticulitis, immunoproliferative small intestine disease, liver disease, diseases and disorders of the gall bladder (including those disorders that are consequent to surgical removal of the gall bladder), pancreatitis, Schwachman's syndrome, steatorrhea, Whipple's disease, parasitic infection, malabsorption as a consequence of chronic laxative use or abuse, pancreatic enzyme deficiency, disaccharidase deficiency, or defects in fat absorption consequent to surgical gastrectomy or other surgical interventions in the gastrointestinal tract.

[0054] The following Examples illustrate certain aspects of the above-described method and advantageous results. The following examples are shown by way of illustration and not by way of limitation.

EXAMPLE 1

[0055] Proliposomal formulations useful for oral administration were developed using an in vitro model system. Human Caco-2 cells (colon adenocarcinoma cells), grown on semipermeable filters, provide a simple and reliable in vitro model for studying drug transport across the intestinal mucosa. Caco-2 cells are recognized in the art for yielding useful predictions on oral absorption of new drug formulations.

[0056] In order to assay the proliposomal tablets of the invention, glyburide (glybenclamide), an oral blood-glucose-lowering drug of the sulfonylurea class, was used as model drug, because uptake in the CaCo-2 system can be monitored by measuring transport across monolayers formed by this cell line.

[0057] Proliposomal tablets were prepared as follows. The identities and amounts of each of the reagents used to prepare the tablets of the invention are shown in Table I. Phospholipids DMPC and DSPC were obtained from Avanti Polar Lipids (Alabaster, Ala.); glyburide, cholesterol, stearylamine, dicylphosphate and all tissue culture reagents were obtained from Sigma Chemical Co. (St. Louis, Mo.); purified talc and anhydrous lactose were obtained from J.T. Baker (Phillips-

burg, N.J.) and Quest, Int'l. (Hoffman Estates, Ill.); chloroform, methanol and ethanol were obtained from Fisher Scientific (Fairlawn, N.J.); Caco-2 cells were obtained from the American Type Culture Collection (Manassas, Va.; Accession No. HTB 37); and transwell culture chambers were obtained from Costar (Cambridge, Mass.).

[0058] Glyburide, lipid and cholesterol were dissolved at room temperature in 10 mL chloroform. Lactose (25 mg/tablet) was suspended in the organic mixture and the suspension evaporated to dryness at 60° C. in a conventional coating pan. The solid residue was collected and sifted through a #60 mesh screen. The sifted residue was then mixed with Explotab® (3 mg/tablet), lactose (50 mg/tablet) and talc (2 mg/tablet) and compressed into tablets using a Manesty B3B 16 station press. The tablets were then coated with a solution of hydroxypropyl methylcellulose in ethyl alcohol (3% w/v) containing triethyl citrate (15% of polymer weight) as a plasticizer. Eudragit L30 D-55 (7% w/w) was then applied on the coated tablets.

TABLE I

Formulary for Preparing Proliposome Tablets						
Quantity of each ingredient used (mg/tablet)						
Formulation	Glyburide	DSPC	DMPC	CHO	STA	DCP
DSCP/Neu	5.0	10.0	—	—	—	—
DSPC.Neu.Cho	5.0	10.0	—	2.45	—	—
DSPC/Pos	5.0	10.0	—	—	0.35	—
DSPC/Pos/Cho	5.0	10.0	—	2.45	0.35	—
DSPC/Neg	5.0	10.0	—	—	—	0.69
DSPC/Neg/Cho	5.0	10.0	—	2.45	—	0.69
DMPC/Neu	5.0	—	10.0	—	—	—
DMPC/Neu/Cho	5.0	—	10.0	2.85	—	—
DMPC/Pos/Cho	5.0	—	10.0	2.85	0.40	—

DSPC = distearylphosphatidylcholine
 DMPC = dimyristylphosphatidylcholine
 STA = stearylamine (Pos: positively charged lipid)
 CHO = cholesterol (Neu: neutral lipid)
 DCP = dicetylphosphate (Neg: negatively charged lipid)

[0059] The purity of the reagents used to make the proliposome tablets of the invention described herein was tested using differential scanning calorimetry. Samples were prepared by dissolving lipid with glyburide and cholesterol separately at a ratio of 1:1 (w/w) in an excess of chloroform. The organic layer was removed and thermograms obtained using a differential scanning calorimeter (TA Instruments, New Castle, Del., Model 2910). Each component was scanned both individually and using a mixture comprising glyburide, lipid and cholesterol at a ratio of 1:1:1 (w:w:w). 2-5 mg of sample was scanned at a rate of 20° C. per minute over a suitable temperature range (25-225° C.) in a hermetically-sealed aluminum pan. The peak transition temperatures of the dispersion were compared with the pure compounds. The results of these experiments are shown in FIGS. 1A through 1C.

[0060] FIG. 1A shows a thermogram of DMPC alone compared with mixtures of DMPC and cholesterol (DMPC/CHOL), DMPC and glyburide (DMPC/GLYB) and DMPC, cholesterol and glyburide (DMPC/CHOL/GLYB). Peak transition temperatures are shown in the Figure. In contrast to the

simple and easily-recognizable peak transition temperature obtained for DMPC, the mixtures are heterogeneous, having more than one localized peak region where a thermal transition occurs.

[0061] FIG. 1B shows a thermogram of DSPC alone compared with mixtures of DSPC and cholesterol (DSPC/CHOL), DSPC and glyburide (DMPC/GLYB) and DSPC, cholesterol and glyburide (DSPC/CHOL/GLYB). Peak transition temperatures are shown in the Figure. A similar pattern is observed herein, where there is a simple and easily-recognizable peak transition temperature obtained for DSPC, but the mixtures are heterogeneous, having more than one localized peak region where a thermal transition occurs.

[0062] Thermograms were also obtained individually and in mixtures for glyburide and cholesterol, and these results are shown in FIG. 1C. From these thermograms, it is evident that the presence of cholesterol acts as an "impurity" in the drug, lowering its melting point. The same effect is observed in mixtures of the drug and lipid. In the presence of both cholesterol and lipid, the melting point of glyburide is further decreased, demonstrating a synergistic effect. These results also indicate that the amount of heat required to melt the drug in a pure state is far higher than the amount needed when the drug is combined with cholesterol or lipid. This explains the increased solubility of the drug when prepared in a solid dispersion of lipid and/or cholesterol.

[0063] Liposomes were reconstituted from proliposomal tablets by adding one tablet to 1 mL phosphate buffered saline in a sterile glass vial. The tablet was allowed to stand at 37° C. for 1 hour with shaking, which was sufficient to dissolve the tablet and reconstitute the liposomal preparation.

[0064] Reconstituted liposomes were characterized for size distribution by large-angle dynamic light scattering using a particle size analyzer (Brookhaven Instruments, Model BI-90). Each preparation was diluted with filtered saline to an appropriate concentration to achieve a medium viscosity of 0.089 centipoise and a medium relative refractive index of 1.332 at room temperature. Measurements obtained under these condition are shown in Table II. These results indicated that the particle size of the resulting liposomes varied both with the presence or absence of cholesterol and with the identity of the phospholipid component. The mean diameter of the liposomes was greater in neutral liposome embodiments than in charged liposome embodiments, and can be explained by the greater propensity of neutral liposomes to aggregate or fuse with one another. Encapsulation efficiency, defined as the percentage of the glyburide encapsulated in liposomes, was determined using the protamine-induced aggregation method as described in Kulkarni et al. (1995, *Pharm. Sci.* 1: 359-362). Briefly, each tablet was disintegrated in 1 mL of phosphate-buffered saline (PBS, pH 7.4) to give a concentration of 10 mg/mL of lipid. To 100 µL of the preparation, equal quantities of a protamine solution (50 mg/mL) in PBS was added and vortexed for about 1 min. The mixture was then incubated for about 12 hours at room temperature. After incubation, the mixture was centrifuged at about 16,000×g for about 5 minutes. 100 µL of the supernatant was removed and the pellet was dissolved in about 1 mL of reagent-grade alcohol (95% ethanol) and sonicated for 5 minutes.

TABLE II

Liposome Particle Size (nm) of Different Tablet Formulations		
Formulation/Charge	Lipid Type	
	DSPC	DMPC
Neutral	1413	1825
Neutral/Cholesterol	1035	748
Positive	1059	N.D.
Positive/Cholesterol	867	629
Negative	3633	N.D.
Negative/Cholesterol	800	N.D.

N.D.: not determined

[0065] The quantity of glyburide in the pellet and the supernatant was determined by HPLC analysis using the Star® 9010 solvent system and Star 9095® variable-wavelength ultraviolet/visible spectrum spectrophotometric detector (Varian Associates, Walnut Creek, Calif.) and the data analyzed by a Dynamax® MacIntegrator (Rainin Instrument Co., Woburn, Mass.). HPLC analysis was performed using a C18 column (Phenomenex®) packed with 5 µm particles and having dimensions of 250 mm in length and an internal diameter of 4.6 mm. The mobile phase was a solution of methanol in 0.1M phosphate buffer, pH 3.5 at a ratio of 75:25 by volume. Column flow rate was 1.0 mL/min and the output was scanned at a wavelength of 225 nm.

[0066] The results of these characterization experiments are shown in Table III.

TABLE III

Drug Encapsulation Efficiency (% ± s.d.)		
Formulation	Lipid Type	
	DSPC	DMPC
Neutral	81.6 ± 0.4	86.7 ± 2.7
Neutral/Cholesterol	80.4 ± 0.6	88.8 ± 1.2
Positive	78.4 ± 0.7	N.D.
Positive/Cholesterol	81.0 ± 1.2	87.6 ± 0.6
Negative	81.2 ± 0.1	N.D.
Negative/Cholesterol	80.4 ± 0.4	N.D.

N.D.: Not determined

[0067] These results demonstrated that a slightly higher percentage of the drug was encapsulated in DMPC. These results are consistent with a slightly higher amount of the drug being encapsulated in "fluid" liposomes (i.e., those comprising DMPC) than liposomes in a gel state (i.e., those comprising DSPC) at 37° C.

EXAMPLE 2

[0068] Caco-2 cell cultures were prepared as monolayers on polycarbonate transwells having a membrane pore size of 4 nm. Caco-2 cells were first grown in T-150 flasks (Falcon, Lincoln Park, N.J.) at 37° C. under an atmosphere of 5% CO₂ and 95% air in Dulbecco's modified Eagle's medium (pH 7.2, Sigma Chemical Co., St. Louis, Mo.), with conventional supplements. The medium was changed every other day until the monolayers reached about 90% confluency. Media was removed and the cells were washed with Hank's balanced salt solution (HBSS, Sigma). The cells were trypsinized by adding 0.5 mL of a 0.25% trypsin solution containing 1 mM EDTA to each flask and incubating the monolayers for 10 min

at 37° C. The separated cells were removed from the flasks and collected into centrifuge tubes, centrifuged at 200×g for 10 min, the supernatant removed and the pellet resuspended in a sufficient amount of Dulbecco's modified Eagle medium to yield a suspension that would produce about 60,000 cells/cm² on plating. The Caco-2 cells were then seeded into Transwell semipermeable membrane inserts having 4 µm pore size. In the transwells, media was changed every other day until the cells were used for the transport studies described below.

[0069] Caco-2 cell cultures on transwell membranes prepared as described above were used for transport studies about 17 days after plating. Proliposome tablets were dissolved as described above by incubation for 1 h with shaking at 37° C. in 2 mL HBSS. As a control, pure glyburide treated with chloroform was compressed into tablet form with lactose and Explotab®; all controls were treated exactly as experimental.

[0070] The medium from the transwell plates was gently removed using a micropipette. 0.5 mL of the reconstituted liposomal suspension was gently added to the donor compartment of the transwell and 1.5 mL of HBSS was added to the receiver compartment. 100 µL of FITC-Dextran was then added to the donor compartment to a final concentration of 10 µg/mL of FITC-Dextran in the donor side. FITC-Dextran was used as a marker to test for the presence of leaks, if any, on the monolayers covering the semipermeable transwell membranes. Samples (300 µL) were carefully withdrawn from the receiver side at 50, 120, 180, 240, 300 minutes after addition, and the receiver side was replenished with 300 µL of fresh HBSS each time the sample was taken. Cells were incubated at 37° C. in a 5% CO₂/95% air atmosphere at all times during these assays. Sampling was done under aseptic conditions in a laminar air-flow hood.

[0071] The amount of glyburide transported during each sampling interval was determined by injecting 90 µL of the sample onto the HPLC system described above in Example 1 and peak areas were recorded. These experiments were performed in triplicate and the average of the results was reported. The results of the experiments are shown in FIGS. 2 through 5.

[0072] FIG. 2 shows the results of glyburide transit across Caco-2 cell monolayers in formulations containing distearylphosphatidylcholine (DSPC). Control experiments performed in the absence of DSPC had a flow rate of almost 1 µg/hr•cm². Formulations of glyburide with DSPC (a "neutral" lipid at physiological pH) showed a similar level of flux across the monolayer, although the addition of cholesterol to these formulations increased the flux about two-fold. Formulations of glyburide with negatively-charged lipid, on the other hand, in either the presence or absence of cholesterol were transported across the monolayer at an even lower rate. In contrast, formulations of glyburide with positively-charged lipid were transported across the membranes at a rate about fourfold higher than control, and the addition of cholesterol increased this to a rate of about fivefold higher than control.

[0073] FIG. 3 shows the results of parallel experiments using dimyristylphosphatidylcholine as the lipid component. A similar pattern of glyburide flux was seen in these experiments; however, the degree of enhancement of transit across the Caco-2 cell monolayer was much higher for formulations containing DMPC. For example, glyburide formulations containing DMPC and positively-charged lipid had a transit rate

almost thirty-fold higher than control. Formulations of neutral lipid were elevated to a lesser degree; in the presence of cholesterol such formulations had a transit rate about eight-fold higher than control, and in the absence of cholesterol this rate was about fivefold higher than control.

[0074] FIGS. 4 and 5 show the cumulative amount of transported glyburide using DSPC- and DMPC-containing formulations over a five hour period. FIG. 4 shows DSPC-containing formulations, wherein the highest accumulation levels were achieved with glyburide formulations containing DSPC and positively-charged lipid (about 27 μg). Similar formulations additionally containing cholesterol had lower total amounts (about 13 μg). DSPC formulations containing neutral lipid and cholesterol showed slower kinetics but achieved essentially the same total accumulation as DSPC/positive lipid/cholesterol formulations. Formulations containing DSPC and neutral lipids in the absence of cholesterol showed the same total accumulation as control (about 2.5 μg), while DSPC formulations with negatively-charged lipid (in the presence or absence of cholesterol) showed lower total accumulation amounts.

[0075] FIG. 5 shows the results of similar experiments performed with DMPC formulations. Total accumulation levels were noticeably higher than control only for formulations containing DMPC, positively-charged lipid and cholesterol (about 34 μg), while DMPC formulations with neutral lipid (in the presence or absence of cholesterol) resulted in total accumulation at levels equivalent to control (about 2-5 μg).

[0076] These results demonstrated that liposomes can be successfully prepared for oral administration in the form of enteric-coated proliposome tablets. The presence of cholesterol reduces the particle size of the formulation. Proliposomes provide a stable system of production of liposomes for oral administration. Degradation of proliposome contents of the tablet in the stomach can be effectively avoided by administering the proliposomes as enteric-coated tablets. Enhanced transport of glyburide across Caco-2 cells was observed with such liposomal formulations. Although the transport of glyburide with DMPC formulations is higher than transport in the DSPC formulation *in vitro*, DSPC formulations are better suited for *in vivo* conditions because of the rigidity and increased stability of the membrane against the attack of bile salts and enzymes of the intestine. Since *in vitro* transport across Caco-2 cells is an indication of bioavailability, an increased transport with the liposome formulation suggests an increased bioavailability of compounds that are poorly absorbed otherwise. For example, using a suitable polymer coating for the proliposomal tablets of the invention, colonic delivery of drugs, especially peptides may be possible.

[0077] Proliposomes are ideally suited for lipophilic compounds, since the majority of such a biologically active compound will partition into the lipid phase. These results also have implications for developing formulations that stabilize the encapsulated drug.

[0078] It should be understood that the foregoing disclosure emphasizes certain specific embodiments of the invention and that all modifications or alternatives equivalent thereto are within the spirit and scope of the invention as set forth in the appended claims.

1-13. (canceled)

14. A pharmacological composition comprising a proliposomal preparation of a biologically active compound, where the biologically active compound is a nutrient, hormone, nucleic acid, antibiotic drug, enzyme, antigen, antiviral drug,

antineoplastic, antiproliferative, peptide or a protein, in a capsule or tablet comprising an enteric coating, a particle lubricant, and a protective coating in between the proliposomal preparation and the enteric coating, wherein the proliposomal preparation comprises a positively charged phospholipid and a neutral lipid, and wherein the protective coating further comprises a plasticizer.

15. A pharmacological composition according to claim 14 wherein the enteric coating is cellulose acetate phthalate or a poly(acrylate, methacrylate) copolymer.

16. A pharmacological composition according to claim 14 wherein the protective coating is hydroxypropyl methylcellulose, polyethylene glycol or ethylcellulose.

17. A pharmacological composition according to claim 14 wherein the neutral lipid is cholesterol.

18. A pharmacological composition according to claim 14 wherein the positively charged phospholipid is a phosphatidylcholine, sphingosine, ceramide, or stearylamine.

19. A pharmacological composition according to claim 14 wherein the plasticizer is triethylcitrate or polyvinyl pyrrolidone.

20. A pharmacological composition according to claim 14 wherein the particle lubricant is talc, lactose, corn starch, ethyl cellulose, fatty acids or salts thereof, agar, pectin, gelatin or acacia.

21. A pharmacological composition according to claim 14 wherein the biologically active compound is aspirin, ibuprofen, erythromycin, vasopressin insulin, dideoxyinosine, cyclosporine, taxol, heparin, halofantrine, ethopropazine, griseofulvin, propofol, furosemide, carbamazepine, diazepam, candesartan or cilexetil.

22. A pharmacological composition according to claim 14 wherein the neutral lipid is cholesterol and the positively charged phospholipid is a phosphatidylcholine, a phosphatidylethanolamine, sphingosine, ceramide, or stearylamine.

23. A pharmacological composition according to claim 22 wherein the phosphatidylcholine is distearylphosphatidylcholine, dimyristylphosphatidylcholine or a mixture thereof.

24. A method for increasing the bioavailability of a biologically active compound, said method comprising orally administering to a human in need thereof a pharmaceutical composition of claim 14.

25. A method according to claim 24 wherein the biologically active compound is aspirin, ibuprofen, erythromycin, vasopressin insulin, dideoxyinosine, cyclosporine, taxol, heparin, halofantrine, ethopropazine, griseofulvin, propofol, furosemide, carbamazepine, diazepam, candesartan or cilexetil; the enteric coating is cellulose acetate phthalate or a poly(acrylate, methacrylate) copolymer; the protective coating is hydroxypropyl methylcellulose, polyethylene glycol or ethylcellulose; the plasticizer is triethylcitrate or polyvinyl pyrrolidone; and wherein the particle lubricant is talc, lactose, corn starch, ethyl cellulose, fatty acids or salts thereof, agar, pectin, gelatin or acacia.

26. A method of treating Crohn's disease, irritable bowel syndrome, celiac sprue, diverticulitis, immunoproliferative small intestine disease, liver disease, diseases and disorders of the gall bladder, disorders that are consequent to the removal of the gall bladder, pancreatitis, Schwachman's syndrome, steatorrhea, Whipple's disease, parasitic infection, malabsorption as a consequence of chronic laxative use or abuse, pancreatic enzyme deficiency, disaccharidase deficiency, or defects in fat absorption consequent to surgical

gastrectomy or other surgical interventions in the gastrointestinal tract, said method comprising administering a composition of claim 14 to human in need thereof.

27. A method according to claim 26 wherein the biologically active compound is aspirin, ibuprofen, erythromycin, vasopressin insulin, dideoxyinosine, cyclosporine, taxol, heparin, halofantrine, ethopropazine, griseofulvin, propofol, furosemide, carbamazepine, diazepam, candesartan or cilexetil.

28. A method according to claim 27, wherein the enteric coating is cellulose acetate phthalate or a poly(acrylate, methacrylate) copolymer; the protective coating is hydroxypropyl methylcellulose, polyethylene glycol or ethylcellulose; the plasticizer is triethylcitrate or polyvinyl pyrrolidone; and wherein the particle lubricant is talc, lactose, corn starch, ethyl cellulose, fatty acids or salts thereof, agar, pectin, gelatin or acacia.

29. A method according to claim 28 wherein the neutral lipid is cholesterol and the positively charged phospholipid is a phosphatidylcholine, a phosphatidylethanolamine, sphingosine, ceramide, or stearylamine.

30. A method according to claim 29, wherein the phosphatidylcholine is distearylphosphatidylcholine, dimyristylphosphatidylcholine or a mixture thereof.

31. A method for delivering a biologically active compound to the intestine or colon, said method comprising orally administering to human in need thereof a composition of claim 14.

32. A method according to claim 31 wherein the biologically active compound is aspirin, ibuprofen, erythromycin, vasopressin insulin, dideoxyinosine, cyclosporine, taxol, heparin, halofantrine, ethopropazine, griseofulvin, propofol, furosemide, carbamazepine, diazepam, candesartan or cilexetil.

33. A method according to claim 32, wherein the enteric coating is cellulose acetate phthalate or a poly(acrylate, methacrylate) copolymer; the protective coating is hydroxypropyl methylcellulose, polyethylene glycol or ethylcellulose; and the plasticizer is triethylcitrate or polyvinyl pyrrolidone; and wherein the particle lubricant is talc, lactose, corn starch, ethyl cellulose, fatty acids or salts thereof, agar, pectin, gelatin or acacia.

34. A method according to claim 33 wherein the neutral lipid is cholesterol and the positively charged phospholipid is a phosphatidylcholine, a phosphatidylethanolamine, sphingosine, ceramide, or stearylamine.

35. A method according to claim 34, wherein the phosphatidylcholine is distearylphosphatidylcholine, dimyristylphosphatidylcholine or a mixture thereof.

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