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(51) Int.Cl.⁶ A61K 39/395, A61K 35/58, A61K 38/46

(30) 1996/09/11 (60/025,179) US

(54) **FORMULATIONS THERAPEUTIQUES CONTENANT DU
VENIN OU UN ANTISERUM CONTRE LE VENIN, UTILISE
SEUL OU COMBINE POUR LA PROPHYLAXIE
THERAPEUTIQUE ET LE TRAITEMENT DE NEOPLASMES**

(54) **THERAPEUTIC FORMULATIONS CONTAINING VENOM OR
VENOM ANTI-SERUM EITHER ALONE OR IN
COMBINATION FOR THE THERAPEUTIC PROPHYLAXIS
AND THERAPY OF NEOPLASMS**

(57) L'invention concerne une méthode de traitement d'organismes hôtes (l'humain ou l'animal) ayant besoin d'un médicament à activité anti-néoplasique, qui consiste à administrer une dose efficace sur le plan thérapeutique d'antisérum contre le venin seule ou de préférence combiné à un inhibiteur non toxique de la phospholipase C ou un antisérum monoclonal ou polyclonal dirigé contre la phospholipase C ou un vaccin constitué intégralement ou partiellement de venin et/ou d'autres composants d'origine animale, végétale ou provenant d'insectes, présentant une activité de phospholipase A₂ et/ou de phospholipase C. L'invention porte sur des formulations pharmaceutiques contenant du venin d'insecte et/ou de serpent, ou des extraits desdits venins ayant, globalement ou en partie, une activité de phospholipase A₂, seuls ou combinés à la phospholipase A₂ végétale ou animale avec ou sans composés inhibant la phospholipase C, ou un antisérum mono ou polyclonal

(57) The present invention comprises the method of treating host organisms (i.e. human or animal) in need of a drug having anti-neoplastic activity comprising the administration of a therapeutically effective amount of venom anti-serum either alone or preferably in combination with a Phospholipase C inhibitor of non-toxic nature or monoclonal or polyclonal anti-serum to Phospholipase C enzyme or a vaccine containing in whole or in part venom and/or other components of animal, insect or plant origin showing Phospholipase A₂ and/or Phospholipase C activity. This patent presents pharmaceutical formulations containing snake and/or insect venoms, or extracts from such venoms which may contain, total or partial, Phospholipase A₂ enzyme activity alone or in combination with animal or plant Phospholipase A₂ with or without Phospholipase C inhibiting compounds or Phospholipase C mono- or



(21) (A1) **2,265,631**
(86) 1997/09/10
(87) 1998/03/19

dirigé contre la phospholipase C, à utiliser comme vaccin d'intérêt potentiel pour toutes les maladies néoplasiques. Elle se rapporte aussi à des formulations pharmaceutiques thérapeutiques contenant un antisérum dirigé contre les venins de serpents et/ou d'insectes et, de préférence, purifié par affinité à des fins thérapeutiques contre les maladies néoplasiques. Elle concerne encore des formulations pharmaceutiques contenant des molécules d'imitation de polymère organique, générées contre des enzymes ou épitopes de PLA₂ de serpent et/ou d'insecte et/ou de mammifère et/ou de plante, ou un extrait desdits venins ou peptides synthétiques et/ou d'autres molécules présentant l'activité, partielle ou intégrale, de la phospholipase A₂ et C.

polyclonal anti-serum to Phospholipase C enzyme as therapeutic vaccine candidate for all neoplastic diseases. This patent presents therapeutic pharmaceutical formulations containing anti-serum to snake and/or insect venoms wherein the anti-serum is preferably affinity purified for use in treating neoplastic diseases. This patent presents pharmaceutical formulations containing organic polymer mimic molecules generated to snake and/or insect and/or mammalian and/or plant PLA₂ enzymes or epitopes, or extract from such venoms or synthetic peptides and/or other molecules which may contain, total or partial, Phospholipase A₂ and C enzyme activity.



PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : A61K 35/58, 39/395, 38/46, G01N 33/574, A61K 9/127	A1	(11) International Publication Number: WO 98/10776 (43) International Publication Date: 19 March 1998 (19.03.98)
(21) International Application Number: PCT/IB97/01091 (22) International Filing Date: 10 September 1997 (10.09.97) (30) Priority Data: 60/025,179 11 September 1996 (11.09.96) US (71)(72) Applicant and Inventor: SHANAHAN- PRENDERGAST, Elizabeth [IE/IE]; Baybush, Straffan, County Kildare (IE).	(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i>	
(54) Title: THERAPEUTIC FORMULATIONS CONTAINING VENOM OR VENOM ANTI-SERUM EITHER ALONE OR IN COMBINATION FOR THE THERAPEUTIC PROPHYLAXIS AND THERAPY OF NEOPLASMS (57) Abstract <p>The present invention comprises the method of treating host organisms (i.e. human or animal) in need of a drug having anti-neoplastic activity comprising the administration of a therapeutically effective amount of venom anti-serum either alone or preferably in combination with a Phospholipase C inhibitor of non-toxic nature or monoclonal or polyclonal anti-serum to Phospholipase C enzyme or a vaccine containing in whole or in part venom and/or other components of animal, insect or plant origin showing Phospholipase A₂ and/or Phospholipase C activity. This patent presents pharmaceutical formulations containing snake and/or insect venoms, or extracts from such venoms which may contain, total or partial, Phospholipase A₂ enzyme activity alone or in combination with animal or plant Phospholipase A₂ with or without Phospholipase C inhibiting compounds or Phospholipase C mono- or polyclonal anti-serum to Phospholipase C enzyme as therapeutic vaccine candidate for all neoplastic diseases. This patent presents therapeutic pharmaceutical formulations containing anti-serum to snake and/or insect venoms wherein the anti-serum is preferably affinity purified for use in treating neoplastic diseases. This patent presents pharmaceutical formulations containing organic polymer mimic molecules generated to snake and/or insect and/or mammalian and/or plant PLA₂ enzymes or epitopes, or extract from such venoms or synthetic peptides and/or other molecules which may contain, total or partial, Phospholipase A₂ and C enzyme activity.</p>		

THERAPEUTIC FORMULATIONS CONTAINING VENOM OR VENOM ANTI-SERUM EITHER ALONE OR IN COMBINATION FOR THE THERAPEUTIC PROPHYLAXIS AND THERAPY OF NEOPLASMS

5 The present invention comprises the method of treating a host organisms
(man or animal) in need of a drug having direct or prophylactic anti-neoplastic
activity comprising the administration of a therapeutically effective amount of
Phospholipase A₂ targeted venom anti-serum alone or in combination with a
known Phospholipase C anti-serum or a Phospholipase C inhibitory
10 compound. A vaccine containing in whole or in part snake or insect venom
or mammalian PLA₂ components comprising epitopes demonstrating
Phospholipase A₂ activity and/or Phospholipase C enzyme components. This
patent presents therapeutic pharmaceutical formulations containing snake
and/or insect venoms, or extracts from such venoms which contain, total or
15 partial, phospholipase A₂ enzyme activity or PLA₂ epitopes. This patent
presents therapeutic pharmaceutical formulations containing anti-serum to
snake and/or insect venoms and/or mammalian PLA₂ enzymes wherein the
anti-serum has been preferably affinity purified for use in treating patients
suffering from neoplastic disease. This patent presents pharmaceutical
20 formulations containing organic polymer mimic molecules generated to snake
and/or insect venoms or the PLA₂ enzyme components thereof and/or PLA₂
enzymes isolated from insect, mammalian or plant cells, and/or
Phospholipase C enzyme preparation or extract from such venoms which may
contain, total or partial, phospholipase A₂ enzyme activity.

25 In this patent the affinity purified anti-serum to venoms Phospholipase A₂
(PLA₂) and mammalian or plant PLA₂ are shown to be active anti -
proliferative neoplastic agents.

 The present invention comprises the method of treating host organisms
(i.e. human or animal) in need of a drug having anti-neoplastic activity
30 comprising the administration of a therapeutically effective amount of venom
anti-serum either alone or preferably in combination with a Phospholipase C
inhibitor of non-toxic nature or monoclonal or polyclonal anti-serum to
Phospholipase C enzyme or a vaccine containing in whole or in part venom

and/or other components of animal, insect or plant origin showing Phospholipase A₂ and/or Phospholipase C activity. This patent presents pharmaceutical formulations containing snake and/or insect venoms, or extracts from such venoms which may contain, total or partial, Phospholipase A₂ enzyme activity alone or in combination with animal or plant Phospholipase A₂ with or without Phospholipase C inhibiting compounds or Phospholipase C mono or polyclonal anti-serum to Phospholipase C enzyme as therapeutic vaccine candidate for all neoplastic diseases. This patent presents therapeutic pharmaceutical formulations containing anti-serum to snake and/or insect venoms wherein the antiserum is preferably affinity purified for use in treating neoplastic diseases. This patent presents pharmaceutical formulations containing organic polymer mimic molecules generated to snake and/or insect and/or mammalian and/or plant PLA₂ enzymes or epitopes, or extract from such venoms or synthetic peptides and/or other molecules which may contain, total or partial, Phospholipase A₂ and C enzyme activity.

Phospholipase A₂ are lipolytic enzymes that hydrolyze the sn-2-acyl ester bond in glycerophospholipids. Many forms of PLA₂ exist in nature and have been described and classified into several groups. Types I, II and III PLA₂ are low molecular weight peptides (13-18 kDa) extra-cellular enzymes, including pancreatic and cobra venom PLA₂ (type I), rattle snake and inflammatory PLA₂ (type II) and bee venom type III. Intracellular cytosolic PLA₂ belong to different groups, including the 85 kDa (type IV) and 40-75 kDa enzymes.

Affinity purified anti-serum to venoms, animal or plant tissue demonstrating the ability to bind PLA₂ enzymes are shown herein below, by way of example, to be active in-vitro and in-vivo anti-proliferative neoplastic agents. Accordingly, these affinity purified antisera either alone or in combination with non-toxic Phospholipase C inhibitor or anti-serum to Phospholipase C are useful in the control of proliferation of neoplastic tissue.

BACKGROUND OF THE INVENTION

There is evidence to indicate that Phospholipase A₂ (PLA₂) is involved in the pathogenesis of many diseases. Thus local and circulating levels of Phospholipase A₂ enzyme and enzymatic products are elevated during infection, inflammatory diseases, tissue injury and brain dysfunction and is a very early indication of neoplastic development prior to tumour cell mass being evident by conventional methods of scanning tissue tumours.

Excessive Phospholipase A₂ activity may promote chronic inflammation, allergic reaction, tissue damage and pathophysiological complications. These effects may be the result of accumulating Phospholipase A₂ products (lysophospholipids and free fatty acids, e.g. Arachidonic Acid) and destruction of key structural phospholipid membrane components, but are potentiated by secondary metabolites, such as eicosanoids and platelet-activating factor. Phospholipase A₂ products or lipid mediators derived therefrom have been implicated in numerous activities that are an integral part of cell activation; chemotaxis, adhesion, degranulation, phagocytosis and aggregation.

Phospholipase A₂ secreted excessively at local sites may be responsible for tissue damage common to rheumatic disorders, alveolar epithelial injury of lung disease and reperfusion.

During acute myocardial ischemia, cytosolic Phospholipase A₂ and Phospholipase C activation causes increased intracellular Ca²⁺. Subsequent accumulation of lysophospholipids and free fatty acids promote damage to sarcolemmal membranes leading to irreversible cell injury and eventually cell death.

Altered cytosolic Phospholipase A₂ and Phospholipase C activity or defects in their control and regulation is a predisposing factor to causing tumour cell development.

Prostaglandins and related eicosanoids are important mediators and regulators of both immune and inflammatory responses. Prostaglandin E₂ induces bone resorption and Leukotriene B₄ stimulates vasodilation and chemotaxis. Increased levels of Phospholipase A₂ is noted in Rheumatoid Arthritis (R.A.), osteoarthritis, gout, collagen and vascular diseases.

Phospholipase A_2 induces non specific airway hyperactivity that is evident in asthma. Phospholipase A_2 is also elevated in peritonitis, septic shock, renal failure, pancreatitis, Chrons and Graves Disease.

5 The activity of cell-mediated defence systems is stimulated by consecutive formation of interleukin 1β (IL- 1β), interleukin-2 (IL-2) and interferon γ (IFN γ). The system is inhibited by interleukin-4 (IL-4) and also by prostaglandin E_2 (PGE $_2$) and histamine, which are released when the immune system is activated. The inhibition is strong in cancer patients, because PGE $_2$ is formed in many cancer cells and its formation is stimulated
10 by IL- 1β . PGE $_2$ and histamine are feedback inhibitors of cell mediated immunity.

PGE $_2$ is formed from arachidonic acid in monocytes, macrophages, cancer cells and other cells, when arachidonic acid is released from cellular phospholipids. The formation of PGE $_2$ is stimulated by several compounds,
15 including histamine, IL-1 (α and β) and Tumour Necrosis Factor α (TNF α). PGE $_2$ inhibits the formation and receptor expression of IL-2 by increasing the level of cyclic AMP (cAMP) in helper T cells. This concomitantly decreases the formation of IFN γ .

20 PGE $_2$ inhibits the ability of natural killer cells (NK) to bind with tumour cells by increasing cAMP in Natural Killer Cells. This decreases tumour cell killing.

When the immune system is stimulated to destroy tumour cells, the killing is prevented because IL- 1β stimulates PGE $_2$ formation in tumour cells, which increases cAMP levels in NK cells and prevents the binding of NK and tumour
25 cells.

The activation of the cell-mediated defence is blocked also because PGE $_2$ -increases cAMP in helper T cells and inhibits the formation of IL-2 and IFN γ .

30 Cytotoxic T cells can also produce PGE $_2$ thus inhibiting the activity of NK cells.

A number of human and experimental animal tumours, contain and/or produce large quantities of prostaglandins (PG). Prostaglandins E₂ has been shown to effect significantly cell proliferation in tumour growth and to suppress immune responsiveness.

5 Phosphatidylinositol specific phospholipase C is an important enzyme for intracellular signalling. There are at least three major classes of Phosphatidylinositol specific Phospholipase C (PtdInsPLC: PtdInsPLC β , γ , δ). PtdInsPLCs hydrolyse a minor membrane phospholipid, phosphatidylinositol (4, 5) biphosphate (PtdIns (4,5) P₂) to give the second
10 messengers inositol (1, 4, 5) trisphosphate (Ins (1, 4, 5) P₃), which releases Ca⁺⁺ from intracellular stores to increase the intracellular free CA⁺⁺ concentration, and diacylglycerol which activates the Ca⁺⁺ and phospholipid-dependent protein serine/threonine kinase, protein kinase C. Proteins phosphorylated by protein kinase C include transcription factors. Together,
15 the increase in intracellular free Ca⁺⁺ concentration and the activation of protein kinase C lead to a series of events that culminate in DNA synthesis and cell proliferation in tumour cells.

A number of growth factors and mitogens, including platelet-derived growth factor (PDGF), epidermal growth factor (EGF) and bombesin, act
20 through specific receptors to increase Ptd Ins PLC activity in cells. Continued stimulation of Ptd Ins PLC can lead to cell transformation.

Ptd Ins PLC activity is found to be increased in a number of human tumours. 76% of human breast cancers have detectable Ptd Ins PLC- γ immunoreactive protein compared to only 6% in benign breast tissue.

25 Cytosolic Ptd Ins PLC activity is increased up to >4-fold in human non-small cell lung cancer and renal cell cancer compared to normal tissue.

SUMMARY OF THE INVENTION

The present invention comprises the method of treating mammals including humans in need of a drug to prevent neoplastic tissue growth and
30 spread by the administration of a therapeutically effective amount of venom anti-serum prepared to whole venom or to parts of the venom or components

of plant or animal origin which demonstrate PLA₂ activity. Also enhanced anti-cancer effects both in-vitro and in-vivo have been realised by combining this affinity purified anti-serum to PLA₂ components and/or mammalian PLA₂ with a non-toxic inhibitor of Phospholipase C or with anti-serum (polyclonal or monoclonal) developed to Phospholipase C enzyme.

This patent relates to the administration of one or more compounds which can generally be described as performing their function by either directly or indirectly causing Phospholipase A₂ and/or Phospholipase C enzyme inhibition, wherein the said inhibition is either partial or total. In addition this patent relates to the administration of one or more compounds which can generally be described as performing their function by interaction with the neoplastic cell membrane preventing their growth or spread, thus preventing further disruption of non-involved organs of the body and causing no toxicity to the infected patient or animal being treated.

Additional aspects of the invention relates to pharmaceutical compositions containing the compounds of the invention as active ingredients, modifying unwanted immune responses, and to methods of retarding proliferation of tumour cells using the compounds and compositions of this invention.

The anti-serum to snake venom PLA₂ and to plant, insect, mammalian and/or to PLA₂ epitopes or mimic molecules are shown herein to be active anti-tumour proliferative compounds and immune enhancing. For use in this regard, the compounds of the invention are administered to mammals, including humans, in an effective amount of 0.05 to 5 gms per day per kilogram of body weight. The amount depends, of course, on the condition to be treated, the severity of the condition, the route of administration of the drug, and the nature of the subject. The drugs may be administered IV, orally, parenterally, or by other standard administration routes including targeting with liposomes/RBC.

The therapeutic activity of the compounds of this invention are demonstrated by inhibition of the tumour cell lines in-vitro and in-vivo. The

compounds were tested for toxicity in Scid mice. Results as in Figure 1 [toxicity data].

Toxicity Study

Method

5 Female Scid mice (6-8 weeks of age) were treated with either a Neat or a 1:10 dilution of the anti-serum preparation, subcutaneously (0.1 ml, daily) for a period of 14 days. The weights of the mice were measured daily. At termination, organs were removed and fixed in formalin for histological examination.

10 **Results**

No toxicity, as assessed by animal weights and clinical well-being, was evident (Figure 1).

The compounds of this invention may be combined with other known anti-inflammatory/immunosuppressive or chemotherapeutic agents such as
15 steroids or non-steroidal anti-inflammatory agents in the pharmaceutical compositions and methods described herein.

Anti-serum to snake and/or insect venoms and/or mammalian and/or PLA₂ enzyme or its epitopes can be used as a therapeutic treatment in diseases where elevated levels of Phospholipase A₂ are evident, (e.g.
20 Rheumatoid Arthritis, see Table B). It is also envisaged that this novel therapy with anti-serum to venom PLA₂ (snake or insect) and/or to PLA₂ components (derived from animal or plant) can be applied as a prophylactic therapy by using sub-lethal doses of venoms or the venom PLA₂ enzyme extracts together with mammalian or plant PLA₂ or synthetic peptides
25 demonstrating PLA₂ activity plus adjuvant to stimulate an immunoglobulin response within the patient, see results - Vaccine Efficacy in Balb/c mice. It is also envisaged that a synthetic peptide incorporating the Phospholipase A₂ and/or Phospholipase C activity could be used to generate said anti-serum or therapeutic agent or vaccine. Use may also be made in the generating of
30 this therapeutic vaccine/anti-serum by using the known sequence homology that exists between human Phospholipase A₂ and snake/insect venoms

together with animal PLA₂ used in combination with compounds known to inhibit Phospholipase C activity or anti-serum developed to this enzyme.

Sustained or directed release compositions can be formulated, e.g. liposomes or those wherein the active compound is protected with differentially degradable coatings, e.g. by microencapsulation, multiple coatings, etc.. It is also possible to freeze-dry the new compounds and use the lyophilizates obtained, for example, for the preparation of products for storage and subsequent injection.

EXPERIMENTATION

The compounds of this invention can be identified as anti-serum to snake or insect venoms mammalian or plant PLA₂ or parts thereof or Phospholipase C or mimic molecules generated to venoms or mammalian PLA₂ molecules and/or Phospholipase C or parts thereof also the pharmaceutical use of venoms or parts thereof and/or mammalian PLA₂ or enzyme components as vaccine antigen are incorporated. Non-toxic compounds showing anti-phospholipase C activity can be incorporated with the anti-serum to PLA₂ of any origin, or mimic molecules demonstrating Phospholipase A₂ activity.

In certain applications of this therapy it may be necessary to curtail the ADCC reaction which could cause serum sickness and to ensure that this does not occur the IgG (FC) component is enzymatically cleaved from the affinity purified immunoglobulin so that natural killer cells will not react to the immunoglobulin in the anti-serum.

Ability of anti-serum to snake venom to inhibit Phospholipase A₂ enzyme isolated from human synovial fluid (Table A2).

The inhibition of Phospholipase A₂ enzyme from synovial fluid isolated from a patient with Rheumatoid Arthritis was tested with a range of dilutions of anti-serum to snake venom. Anti-serum to snake venom generated in horse, reconstituted in 10 ml sterile water. The following dilutions were used 1:10, 1:20, 1:40 and 1:60. The method used was as outlined in "Infection and

Immunity, Sept. 1992, p. 3928-3931. Induction of Circulating Group II Phospholipase A₂ Expression in Adults with Malaria.

	Results	(Table A2)
	<u>Dilution</u>	<u>Inhibition</u>
5	1:10	63%
	1:20	50%
	1:40	35%
	1:60	29%

In-vitro testing of un-affinity purified snake venom.

10 A range of tumour cell lines were tested with 3 concentrations of the anti-serum to snake venom by the MTT Assay. This anti-serum was not affinity purified. MTT Assay described by Alley et al, (Cancer Research, 48: 589-601, 1988) See Table B.

SUMMARY OF RESULTS (Table B)

15	Molt 4:	Human T cell Lymphoma Cancer
	Serum-containing	
	Dilution	% of Control
	Neat	48.1
	1:10	53.7
20	1:20	40.8
	Serum-Free	
	Neat	58.7
	1:10	51.2
	1:20	40.6
25	MDA 468:	Human Breast Cancer
	Serum-containing	
	Dilution	% of Control
	Neat	8.0
	1:10	53.7
30	1:20	58.9
	Serum-Free	

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	Neat	15.4
	1:10	48.4
	1:20	58.9
	C17OHM2:	Human Colon Cancer
5	Serum-containing	
	Dilution	% of Control
	Neat	9.3
	1:10	61.4
	1:20	55.6
10	Serum-Free	
	Neat	15.2
	1:10	47.3
	1:20	49.5
	Pan 1:	Human Pancreatic Cancer
15	Serum-Containing	
	Dilution	% of Control
	Neat	9.3
	1:10	47.5
	1:20	49.2
20	Serum-Free	
	Neat	43.1
	1:10	53.2
	1:20	69.4
	841: Human small cell lung cancer	
25	Serum-containing	
	Dilution	% of Control
	Neat	25.2
	1:10	45.5
	1:20	51.1
30	Serum-Free	
	Neat	63.4

1:10 60.1

1:20 59.8

T24: Human Bladder Cancer

Serum-containing

5 **Dilution % of Control**

Neat 68.5

1:10 75.1

1:20 76.2

Serum-Free

10 Neat 84.1

1:10 87.9

1:20 88.4

**Testing un-affinity purified anti-serum to Snake Venom against
B16F1 Melanoma Cell Line.**

15 **Mice**

C57BL/6

Procedure

The mice were inoculated with 0.5×10^6 B16 F1 melanoma cells subcutaneously (sc) into flank region. Once palpable tumours had developed the mice received daily sc injections as follows: -

		number of mice
20	A - Sterile water 100 μ l	6
	B - anti-serum (full strength) 100 μ l	6
25	C - anti-serum (diluted 1:10) 100 μ l	6

The dimensions of the tumours were taken daily using callipers. Once the tumours of the control mice were approximately 1.5 cm or larger in diameter all mice were killed. The tumours were removed and weighed.

Results

30 Small tumours were first discernible by palpitation in all mice 6-7 days after inoculation. The changes in volume as measured by callipers, together with

tumour weights at autopsy. See Fig. 2 [Effect of un-affinity purified anti-serum to snake venom on Melanoma B16F1 Growth] for effect of anti-serum to snake venom on tumour growth retardation.

IN-VITRO SREENING OF THE AFFINITY PURIFIED ANTI-SERUM TO SNAKE VENOM PREPARATION AGAINST A RANGE OF TUMOUR CELL LINES (Illustrated in Fig. 3A [Human colorectal tumour C170HM2], Fig. 3B [Human bladder tumour T24], Fig. 3C [Human lymphoma tumour MOLT 4], Fig. 3D [Human pancreatic tumour PAN 1], Fig. 3E [Human breast tumour MDA 468], Fig. 3F [Human small cell lung tumour 841], Fig. 3G [Human gastric ST24], and Fig. 3H [Human Ovarian OVCAR3])

Introduction

The in-vitro inhibitory effects of the horse generated anti-serum to snake venom preparation, previously evaluated were obscured due to serum enhancement of tumour cell growth. Thus in the following assay, affinity purified anti-serum to snake venom was evaluated.

Method

The cell lines were seeded into 96 well plates at a cell concentration of 10^4 cells per well in both serum free (Hams F12:RPMI 1640 + 0.5% bovine serum albumen) and serum-containing medium (RPMI 1640 + 10% heat inactivated foetal calf serum). The anti-serum preparation was diluted in the corresponding medium and added to the wells, 2-3 hours after the cells (to allow for cell adherence). The plates were incubated at 37 °C in -5% CO₂ for 3 days. The cells were then incubated with 1 mg/ml MTT (methyl thiazol tetrazolium) for 4 hours at 37 °C. The crystals were then solublised with dimethyl sulphoxide and the absorbance measured at 550nm.

Results

The test anti-sera inhibited all of the cell lines at all concentrations examined.

The level of inhibition was statistically significant from the untreated control at all anti-serum dilutions, with all cell lines as assessed by a one way analysis of variance.

IN-VIVO TEST

The effects of affinity purified anti-serum to snake venom on human colorectal C17OHM₂ cell line.

Materials and Methods

5 C170MH₂ cells were injected subcutaneously into the left flank of ten male nude mice. The mice were allocated randomly to two groups.

Group 1 - 100µl anti-serum twice daily intravenously (IV)

Group 2 - 100µl PBS twice daily IV

Tumours were measured twice weekly, using callipers, in two dimensions.

10 Cross-sectional areas were calculated. The mice were also weighed once weekly. The therapy was terminated at day 22.

Results

The cross-sectional areas were measured at increasing time points during the experiment, as shown in Fig. 4 [Effect of affinity purified anti-serum to snake venom on the mean cross-sectional area of C170HM₂ in nude mice].
15 The affinity purified anti-serum preparation induced a slowing in growth when compared to saline controls. An ANOVA was performed on the results in which the treatment was evaluated with respect to time, and shows a significance of P = 0.028.

20 At the termination of the experiment, the tumours were weighed and the results are shown in Fig. 5 [Effect of affinity purified anti-serum to snake venom on the final tumour weight of C170HM₂]. No toxic effect of the affinity purified anti-serum preparation was observed.

In-vitro screen of the affinity purified anti-serum to snake venom preparation in combination with a phospholipase C inhibitor 1-oleoyl-2-acetyl-sn-glycerol (OAG) 5µ molar, on a range of cancer cell lines.
25

Methods

The affinity purified anti-serum to snake venom preparation was diluted 1:2 and 1:10 and was combined with 5 µ molar OAG and added to the wells
30 as previously described for the MTT Assay. The cell lines tested were Human Breast tumour, MDA 468, Human small cell lung tumour 841 and

Human renal TK-10. Results as shown in Fig. 6A [Affinity purified anti-serum to snake venom and (OAG) a Phospholipase C inhibitor combination--Human breast tumour MDA 468], Fig. 6B [Affinity purified anti-serum to snake venom and (OAG) a phospholipase C inhibitor combination--Human small cell lung tumour 841] and 6C [Affinity purified anti-serum to snake venom and (OAG) a phospholipase C inhibitor combination--Human renal TK-10].

In-vivo testing of the combination of affinity purified anti-serum to snake venom and 1-oleoyl-2-acetyl-sn-glycerol (OAG) at 5µm concentration on the growth of MDA 468 cell line.

10 **Method**

MDA 468 tumours were aseptically removed from donor female Scid mice. The tissue was aseptically minced, pooled and implanted into anaesthetised female Scid mice (anaesthetic comprised of a 0.2 ml injection of Hypnorm (Janssen) :Hyonovel (Roche) : distilled water in a 1: 1:5 ratio). Tissue implants consisted of 3-5 mm² pieces and after subcutaneous transplantation into the left flank, the incision was clipped. The Scid mice were then randomised into 2 groups of 10 animals. They were treated daily with a 0.2 ml subcutaneous injection (in the opposite flank to the tumour graft) of a combination of affinity purified anti-serum to snake venom and 5µm molar of (OAG) dilution of the anti-serum preparation. The control animals received 0.2 ml phosphate buffered saline, pH 7.6. All animals were terminated on day 63, and the tumours were dissected out, weighed and processed for histology. Results are in Fig. 7 [Effect of the affinity purified anti-serum to venom in combination with the Phospholipase C inhibitor (OAG) 5 µm].

25 **Vaccine Efficacy in Balb/c mice after challenge with WEHI-3 cell.**

The objective of study is to demonstrate the efficacy of sub-lethal levels of Russell's viper venom entrapped in liposomes and porcine phospholipase A₂ enzyme entrapped in liposomes working in combination to confer a sustained and protective antibody response to a challenge by Leukaemia cells (WEHI-3 cells)

30

The Russell's viper venom was toxoided with 2% osmium tetroxide and entrapped in liposomes (egg phosphocholine and cholesterol). The liposomes were sterilised.

5 The Porcine Phospholipase A₂ enzyme was entrapped in liposomes (egg phosphocholine, and cholesterol) and were sterilised.

Immunisation of mice consisted of an initial subcutaneous injection of 0.25 mls (containing 250 µg of venom) and 3 days later the mice were injected subcutaneously with 0.25 mls of porcine PLA₂ (containing 250 µg of porcine PLA₂). Boosters of each vaccine were given at 3 week intervals.

10 Control mice were injected with 0.25 mls of sterile physiological saline on days corresponding to test mice inoculations.

Animals

Balb/c mice (20-25 g) were used in the study. 15 mice were used in each group.

15 Group I - test mice
Group II - control mice

Challenge

20 The immunised mice and controls were challenged by intravenous injection into tail vein with approximately 5×10^5 leukemic cells (WEHI-3 cells) on day 30 of study.

Test mice are observed for extended life span after the death of the control mice after approximately 24 days.

Results Obtained

25 All control mice died of leukaemia within the allotted time span of 24 days. The venom combination inoculation protected the vaccinated group from the cancer cell challenge and there was a 100% survival rate at day 35 when the experiment was terminated.

30 Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilise the present invention to its fullest extent. The preferred specific embodiments are, therefore, to be construed as merely illustrative, and not limitative of the disclosure in any way whatsoever.

I Claim

1. A method of treating neoplasm in a mammal in need of such treatment, comprising administering to said mammal a therapeutic agent comprising
5 venom and/or mammalian, plant or insect anti-serum reactive with at least one Phospholipase A₂ enzyme.
2. A method according to claim 1 wherein the anti-serum is reactive with two or more Phospholipase A₂ type enzymes.
3. A method according to claim 1 wherein the at least one Phospholipase
10 A₂ Type enzyme is Type I, Type II, Type III or Type IV.
4. A method according to claim 1 wherein the anti-serum is either polyclonal or monoclonal.
5. A method of treating a mammal prophylactically to prevent neoplastic development, comprising administering to said mammal a therapeutic vaccine
15 containing venom and/or mammalian, plant or insect PLA₂ enzymes or part thereof as the principal antigen component.
6. A pharmaceutical formulation containing venom and/or mammalian plant or insect anti-serum to PLA₂ enzyme or part thereof in combination with anti-serum to Phospholipase C enzyme or part thereof or inhibitory compounds
20 to Phospholipase C for use as a therapeutic agent for the therapy of a neoplastic condition in a human or animal.
7. A method according to claim 6 wherein the inhibitory compounds to Phospholipase C is one or more of EDTA, Phenanthroline, Chloromercuribenzoic Acid, Iodoacetic Acid, and 1-oleoyl-2-acetyl-sn-glycerol(OAG).
25
8. A pharmaceutical formulation containing one or more venoms or venom components as antigen and/or mammalian, plant or insect PLA₂ enzyme as antigen in combination with Phospholipase C enzyme.
9. A method according to Claim 8 wherein the phospholipase C enzyme
30 inhibitor is used in combination with the therapeutic agents of Claim I to enhance anti neoplastic and anti metastatic activity.

10. A method according to any one of Claims 1, 5, 6 and 8, wherein the administration is part of a combination therapy with other therapeutically effective agents.
11. A method according to Claims 1, 5, 6 and 8 wherein the administration is in combination with adjuvants.
12. A method according to Claims 1, 5, 6 and 8 wherein the venom is that of snake and/or insect.
13. A method according to Claims 1, 5, 6 and 8 wherein the Phospholipase A₂ enzyme showing Phospholipase A₂ activity is obtained from more than one species of snake and/or insect, mammal or plant.
14. A method according to Claims 1, 5, 6 and 8 wherein the therapeutic agent is administered as an anti-inflammatory agent.
15. A method according to Claims 1, 5, 6 and 8 wherein the therapeutic agent is administered to prevent the occurrence of immunosuppression.
16. A method according to Claims 1, 5, 6 and 8 wherein the therapeutic agent is administered in the treating of allergic contact dermatitis, Asthma and Psoriasis and bronchitis.
17. A method according to Claims 1, 5, 6 and 8 wherein the anti-serum is administered for the treatment of physiological condition resultant from elevated levels of phospholipase A₂ products and/or metabolites.
18. A method according to claim 17 wherein the physiological condition is Schizophrenia.
19. A method according to Claims 1, 5, 6, 8 and 17 wherein the anti-serum to Phospholipase A₂ and/or C are produced synthetically by molecular imprinting of template organic molecules using these enzymes.
20. Therapeutic agents according to Claims 1, 5, 6 and 8 for treating one or more of the following:- Rheumatoid arthritis, osteoarthritis, gout, rheumatic carditis and autoimmune diseases, allergic diseases, bronchial asthma, septic shock, renal failure, pancreatitis, myasthenia gravis and ocular and dermal inflammatory diseases, psoriasis, splenomegaly, cancer, metastatic spread of neoplasm, collagen vascular disease, myocardial ischemia, cellular

chemotaxis, depression, erythema, vascular permeability resultant from enhanced production of PGE₂, acne, atopic diseases, malaria, allergic conjunctivitis, schizophrenia, reiters syndrome, raynaud's phenomenon, lupus, Chron's and Graves disease.

- 5 21. A method according to Claims 1, 5, 6, 8 and 17 wherein the Fc receptor of the antibody to either Phospholipase A₂ and C used in this therapeutic method is either totally or partially removed.
22. A method according to Claims 6, 8, 19 and 21 wherein a non-toxic compound demonstrating inhibiting activity against Phospholipase C enzymes
10 may be utilised in conjunction with the PLA₂ anti-serum to enhance its anti-neoplastic (tumour) and anti-metastatic activity.
23. A method according to Claims 1, 5, 6, 8,17 and 19 wherein the anti-serum is generated to human Phospholipase A₂ enzyme either in a mono and/or polyclonal form.
- 15 24. A method according to Claims 1, 5, 6, 8 and 17 wherein the anti-serum to Phospholipase A₂ enzyme is generated in eggs, producing antibodies which do not react with the human Compliment system.
25. A method according to Claims 1, 5, 6, 8 and 17 wherein the anti-serum to venom, mammalian, plant or insect Phospholipase A₂ is generated in
20 mammals and extracted from the colostrum and preferably but not essentially affinity purified for use in oral administration to patients either alone or in combination with anti-serum similarly produced to human Phospholipase C enzyme components.
26. A method of inoculation of human or animal with a combination of two
25 or more phospholipase A₂ enzymes types.
27. A method according to claim 26 where the antibody response to the inoculation confers prophylactic and/or therapeutic benefit to patient.
28. A method according to claim 27 wherein the patient is in need of a treatment for a neoplastic condition.
- 30 29. A method according to claims 26, 27 and 28 wherein the phospholipase A₂ type is Type I, Type II, Type III or Type IV.

30. A method according to claim 29 wherein the Phospholipase A₂ is obtained from venom.
31. A method according to claim 29 wherein the Phospholipase A₂ is obtained from animal or plant species.
- 5 32. A method according to claim 1, 5, 6, 8 and 26 wherein the phospholipase A₂ is synthetically produced or cloned.
33. A method of early detection of neoplastic disease by utilising the detection of enhanced PLA₂ levels in patients.
34. A method according to claim 33 wherein the detection of enhanced
10 PLA₂ is established by the use of Lipose Diagnostic Kit.
35. A method according to claims 2, 26, 27 and 28 wherein Phospholipase A₂ type enzyme has a size of between 40-80 kDa.
36. A method of targeting cancer cells by use of Type I and/or Type II PLA₂ as targeting agent with hydrophilic tail.
- 15 37. A method according to claim 36 wherein the targeting agent is a liposome containing anti-serum to PLA₂ or conventional chemotherapy drugs.
38. A method treating parasitic and bacterial infections in mammals by the administration of a therapeutic agent containing venom and/or mammalian, plant or insect anti-serum reactive with Phospholipase A₂ enzymes
- 20 39. A method according to Claim 38 wherein the anti-serum is reactive with one or more Phospholipase A₂ type enzymes
40. A method according to Claim 39 wherein the Phospholipase A₂ Type enzymes is one of Type I, Type II, Type III or Type IV.
41. A method according to Claim 38 wherein said parasite is an
25 haemoflagellate parasite.
42. A method as recited in Claim 41 wherein said parasite is a member of the group of haemoflagellate parasites consisting of Leishmania, Trypanosomia and Toxoplasma.

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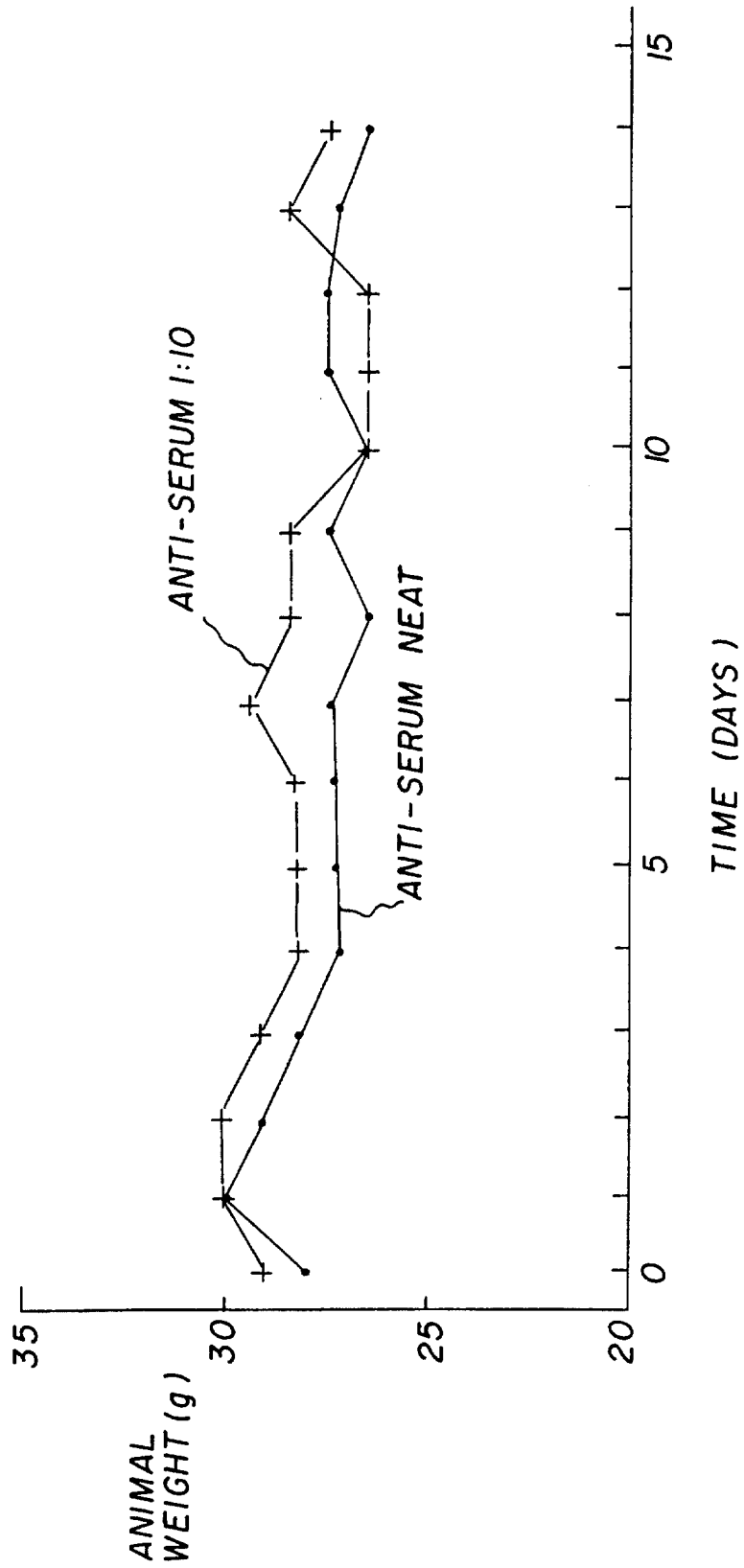


Fig. 1

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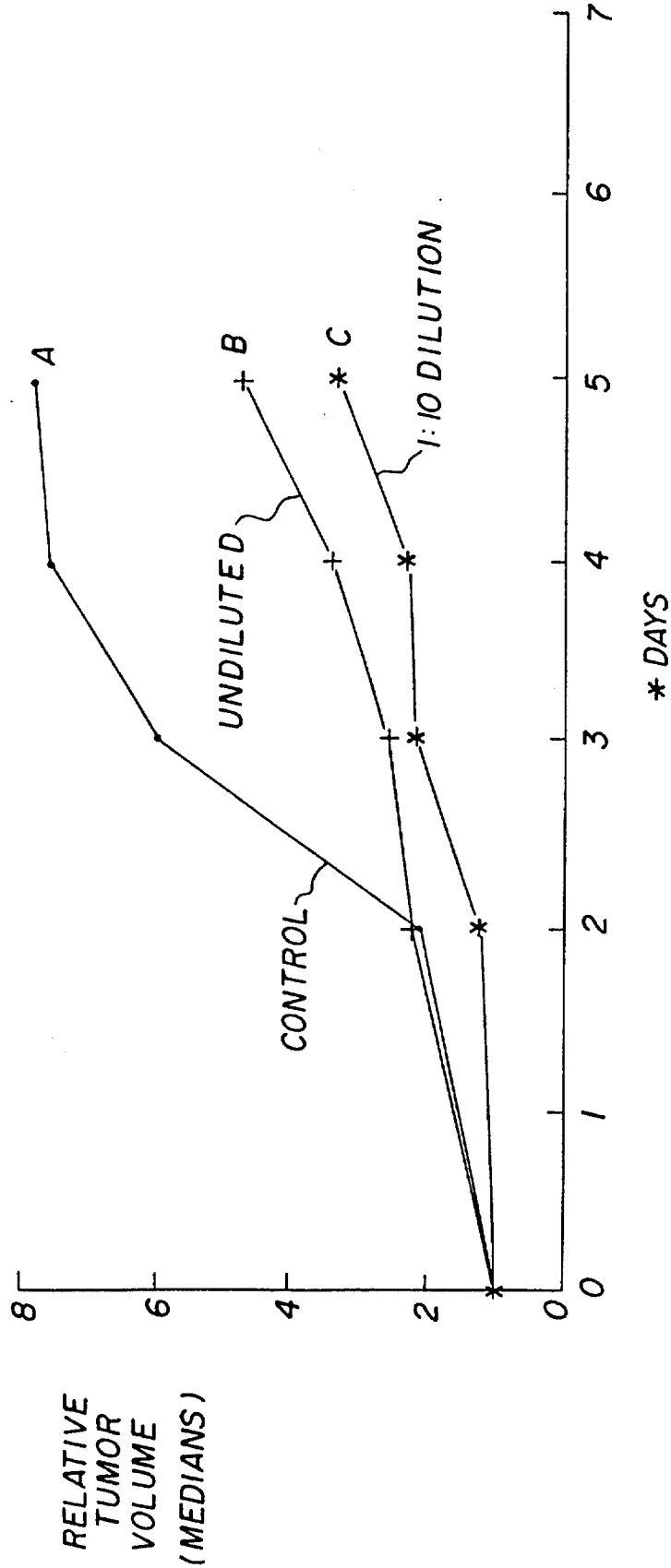


Fig. 2

*Days are measured with day zero taken as day 7 past tumour inoculation.

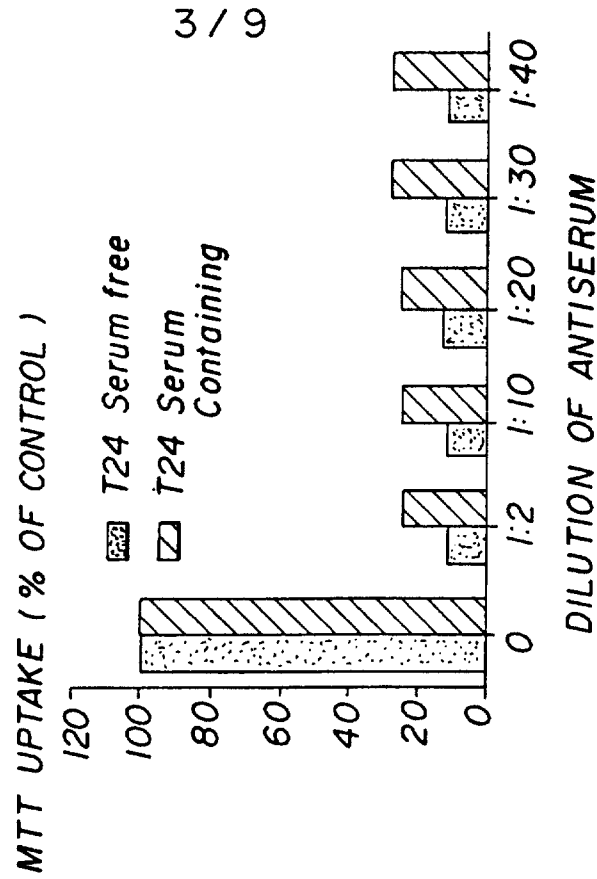


Fig. 3B

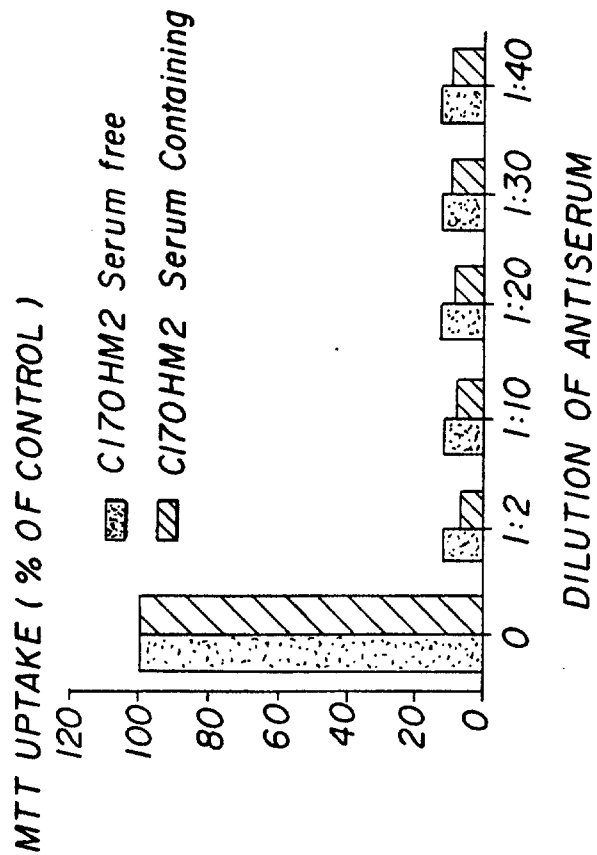


Fig. 3A

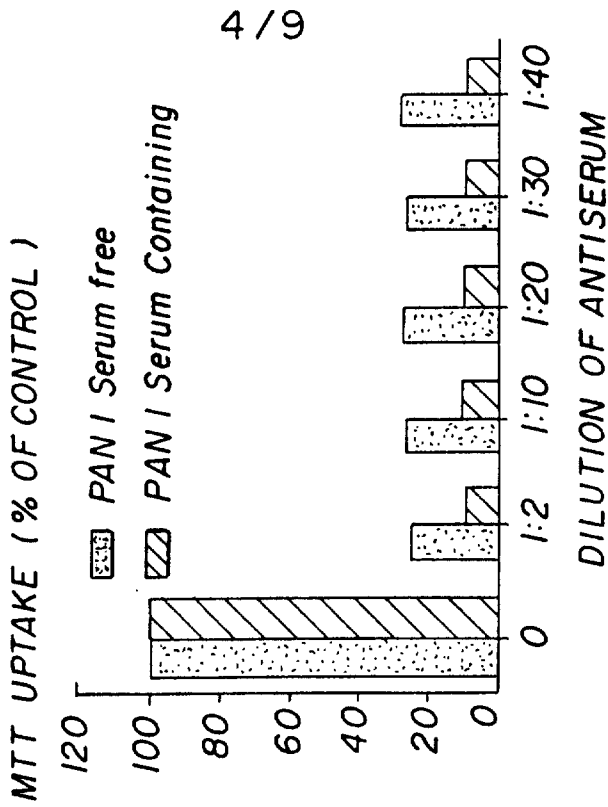


Fig. 3D

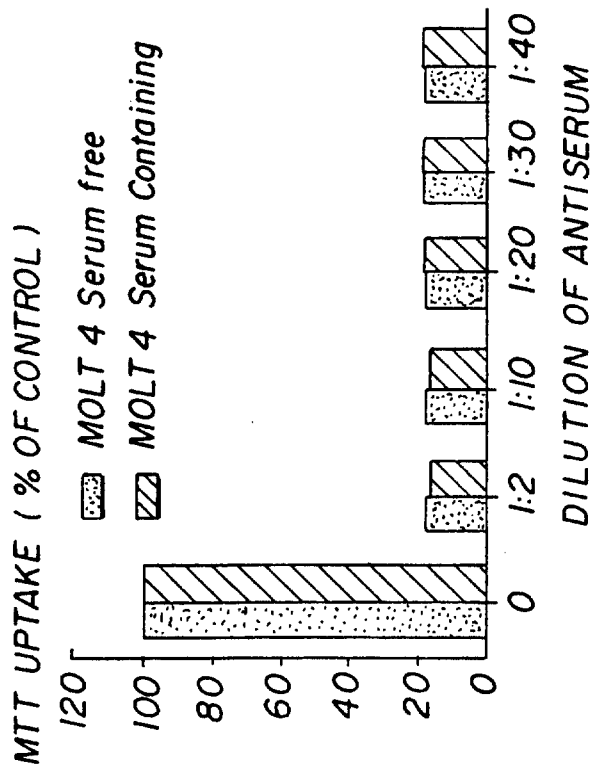


Fig. 3C

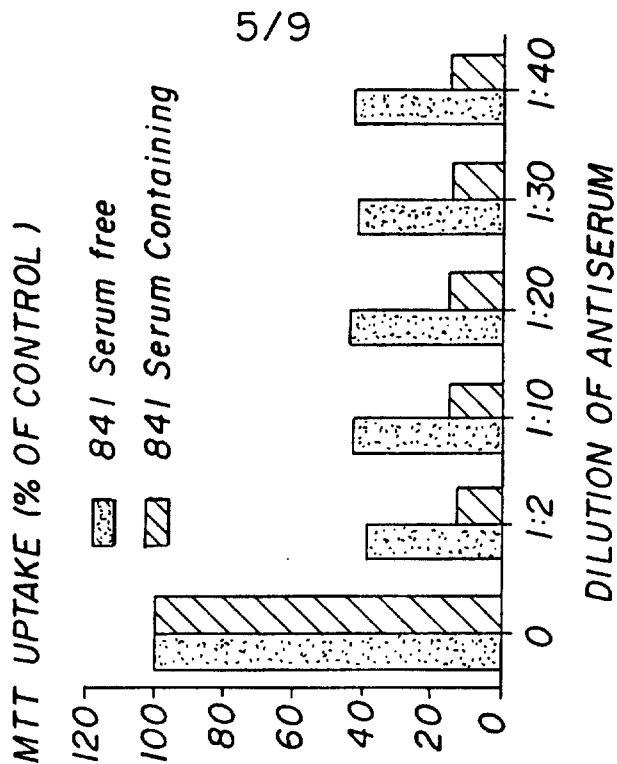


Fig. 3F

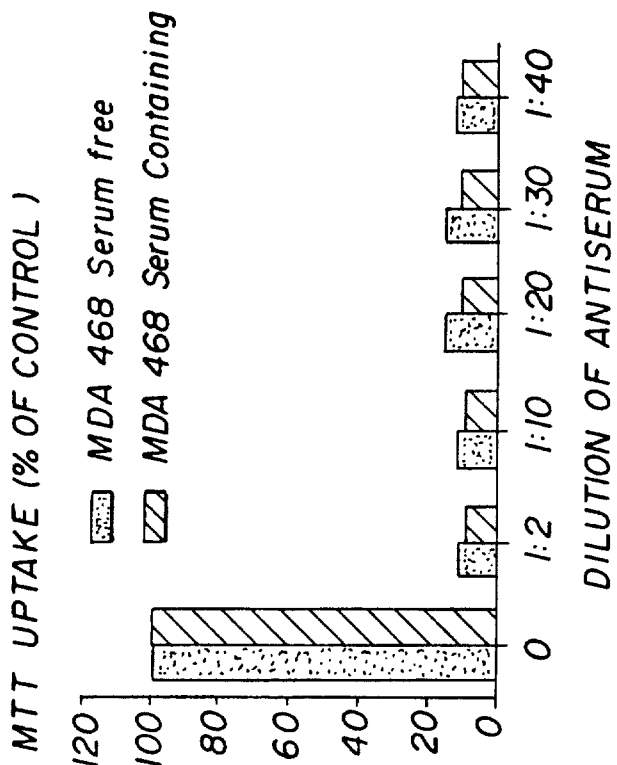


Fig. 3E

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Human Ovarian OVCAR3

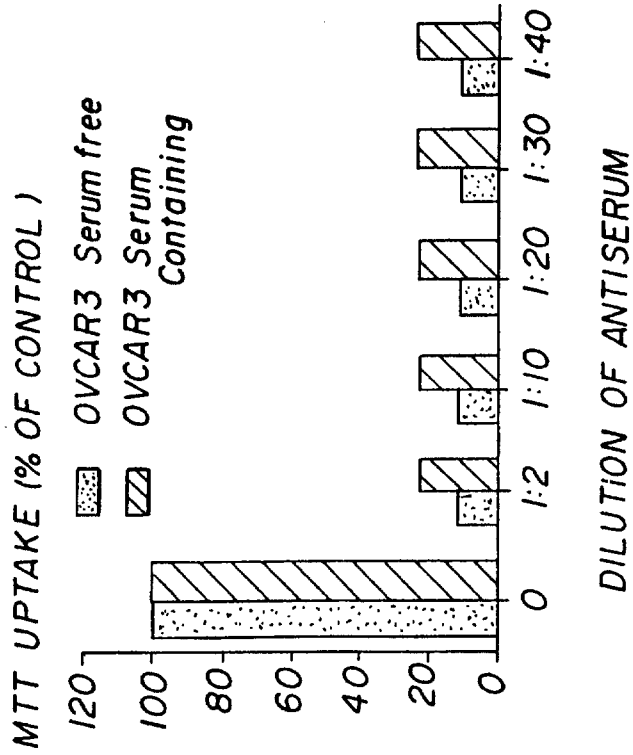


Fig. 3H

Human gastric ST24

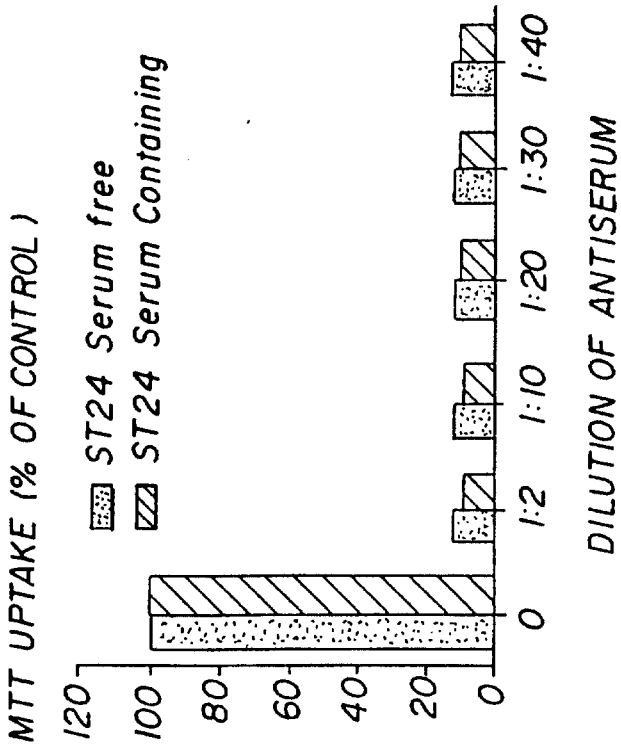


Fig. 3G

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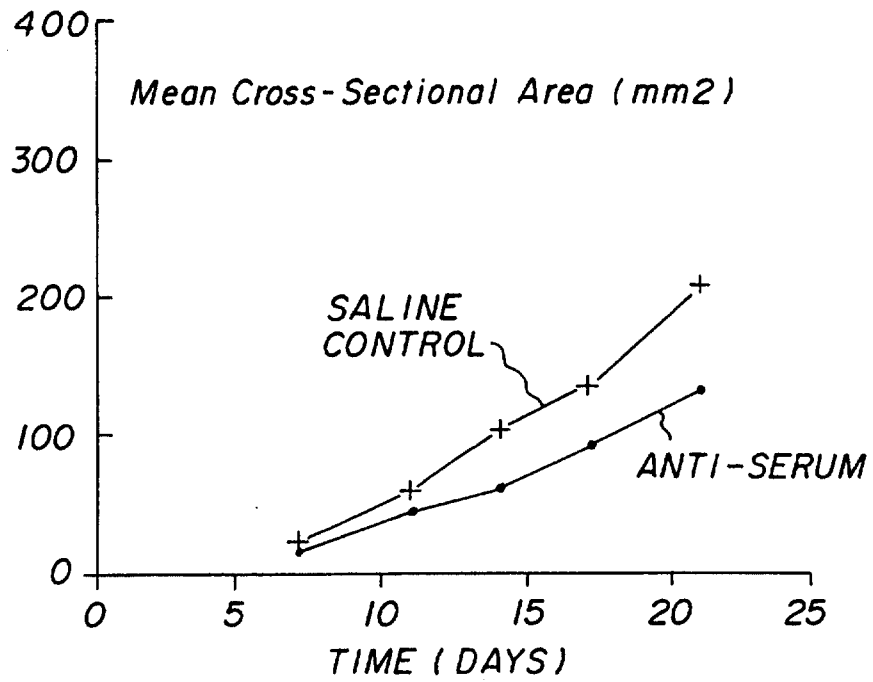


Fig. 4

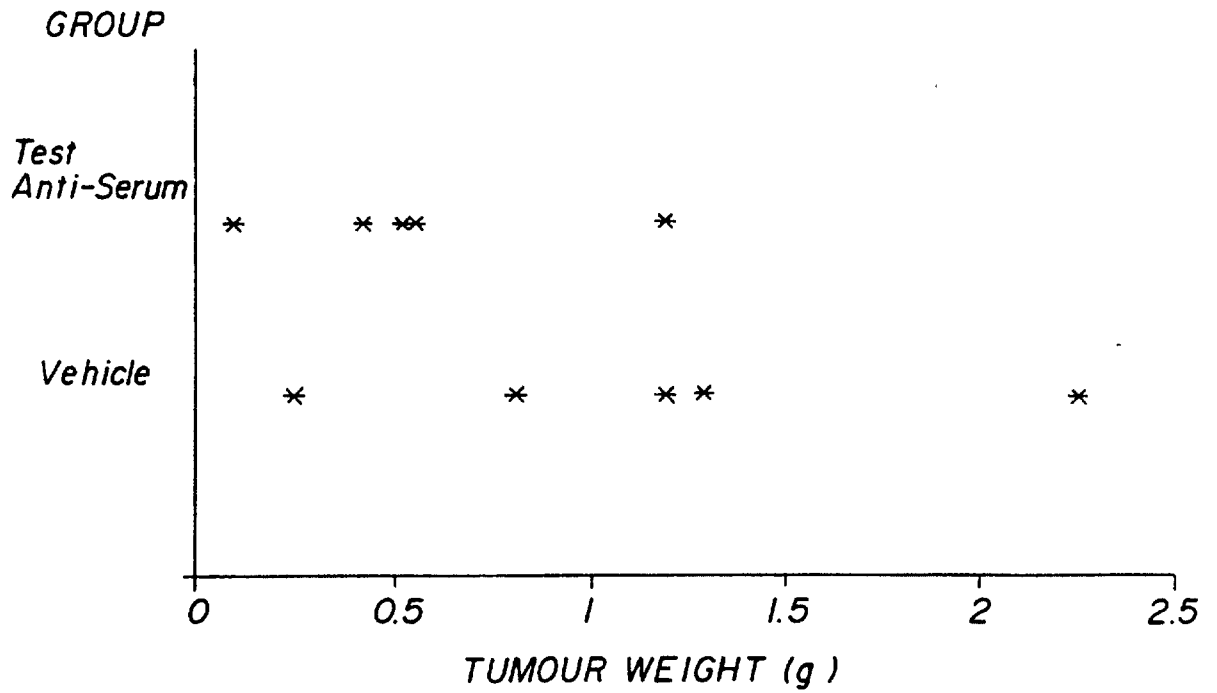
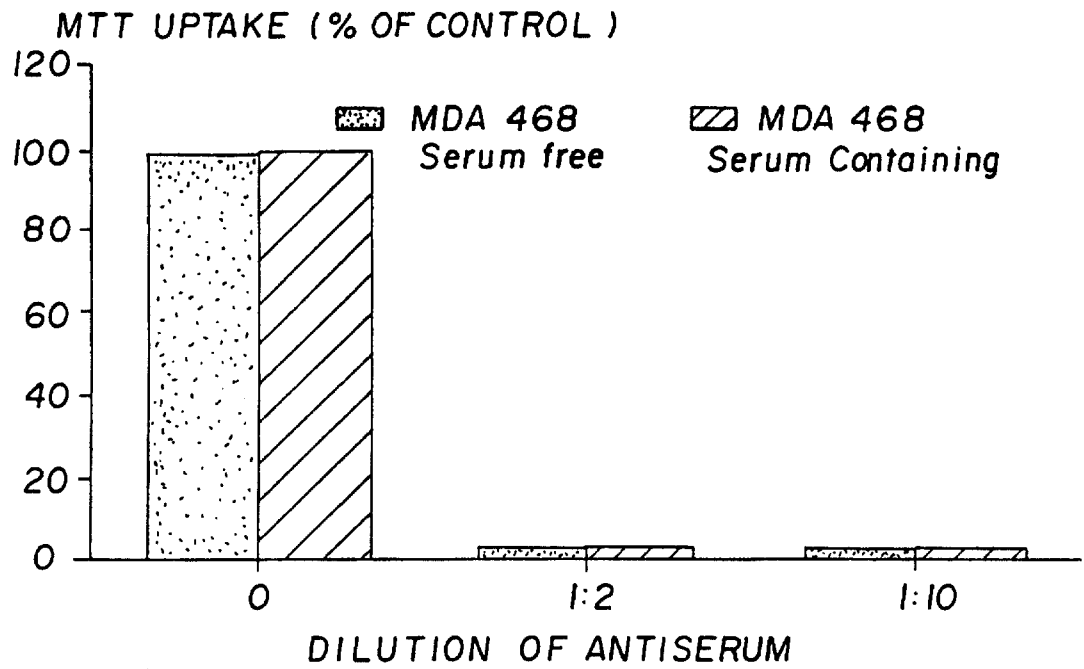
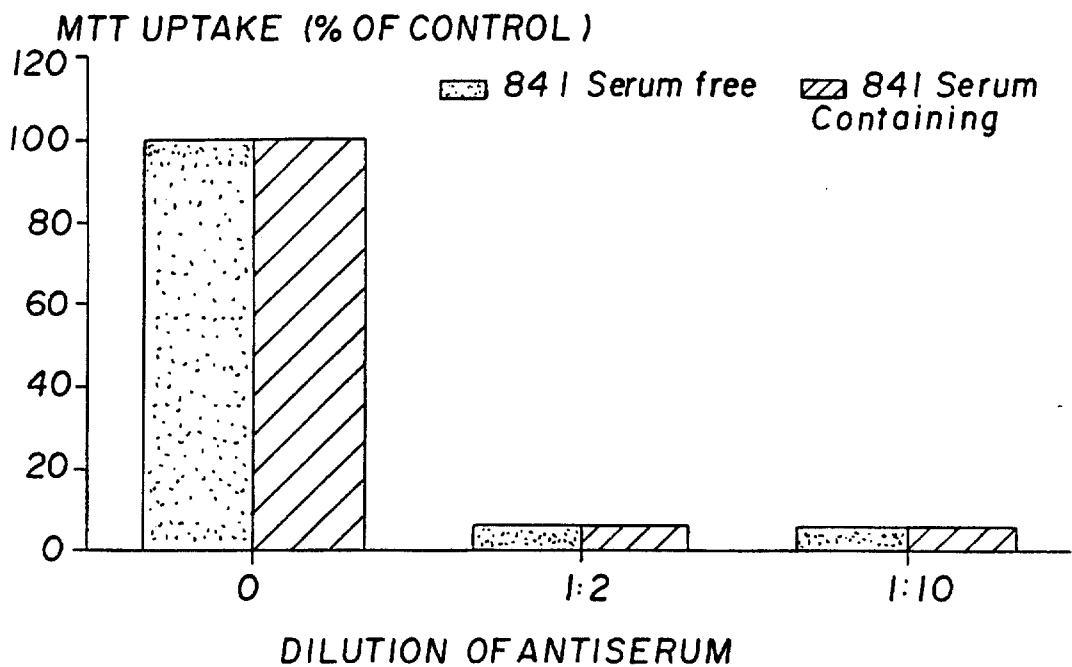


Fig. 5

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*Fig. 6A**Fig. 6B*

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MTT UPTAKE (% OF CONTROL)

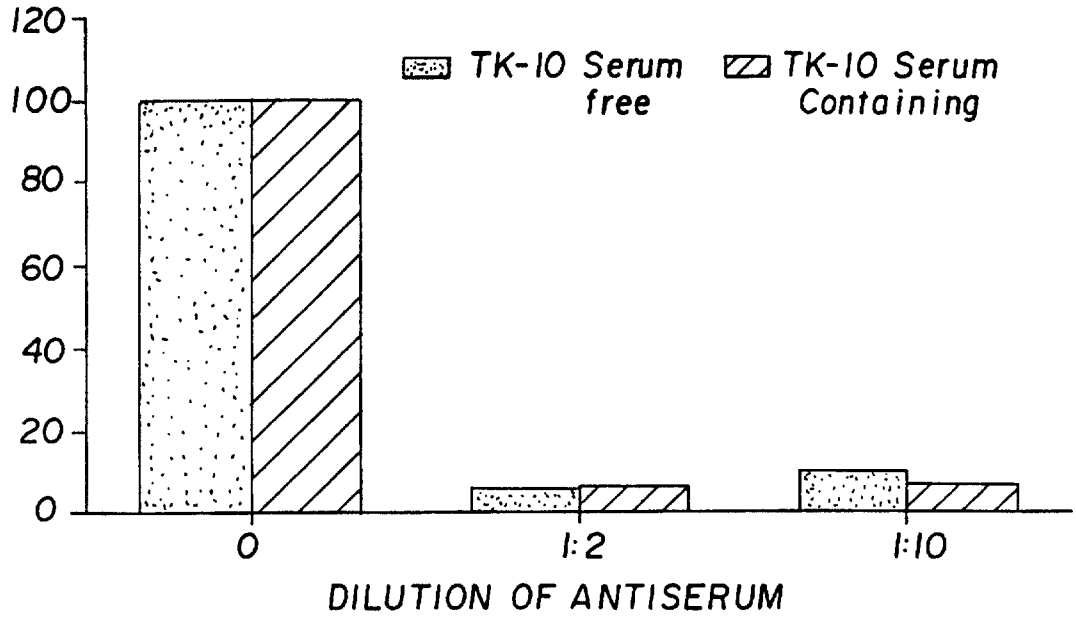


Fig. 6C

TUMOUR CROSS-SECTIONAL AREA

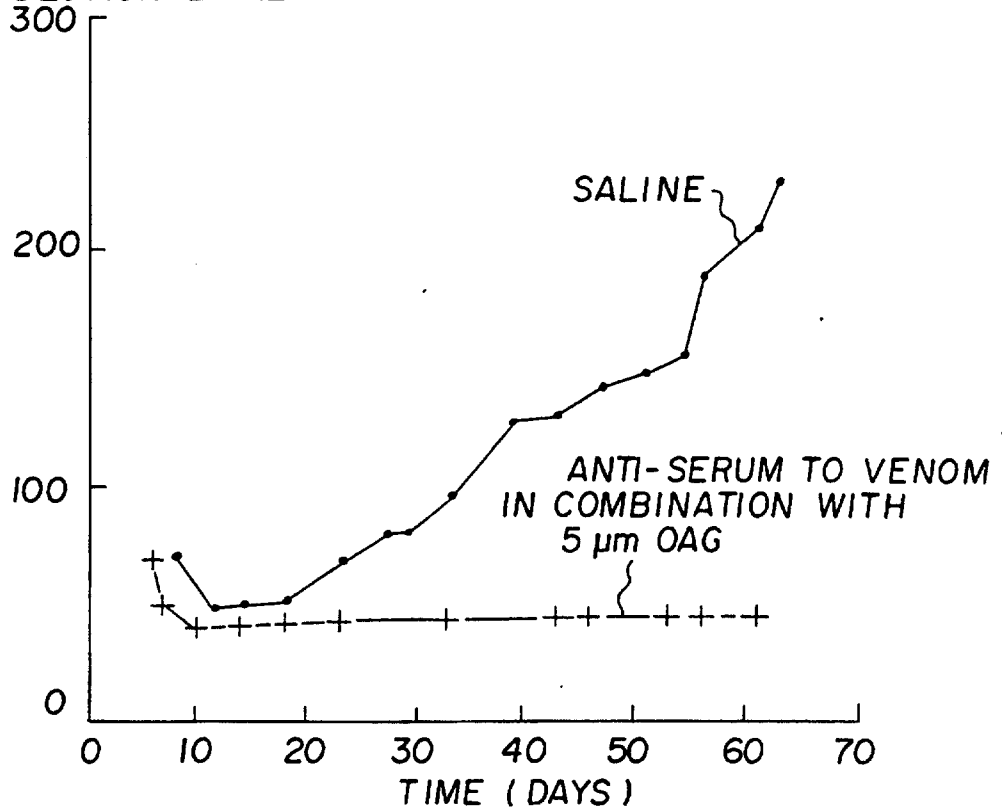


Fig. 7