

FORM 1

COMMONWEALTH OF AUSTRALIA

PATENTS ACT 1952

APPLICATION FOR A STANDARD PATENT

I\We,

BOARD OF REGENTS, THE
UNIVERSITY OF TEXAS SYSTEM

of

201 WEST SEVENTH STREET
AUSTIN
TEXAS 78701
USA

LODGED AT SUB-OFFICE

26 MAR 1986

Melbourne

hereby apply for the grant of a standard patent for an
invention entitled:

MAGNETIC ALTERATION OF CELLULOSE
DURING ITS BIOSYNTHESIS

which is described in the accompanying complete specification

Details of basic application(s):

Number of basic application	Name of Convention country in which basic application was filed	Date of basic application
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719505

US

03 APR 85

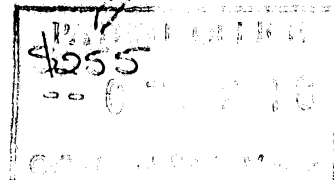
My/our address for service is care of CLEMENT HACK & CO., Patent
Attorneys, 601 St. Kilda Road, Melbourne 3004, Victoria,
Australia.

DATED this 26th day of March 1986

BOARD OF REGENTS, THE
UNIVERSITY OF TEXAS SYSTEM

CLEMENT HACK & CO.

TO: The Commissioner of Patents.



APPLICATION ACCEPTED AND AMENDMENTS
ALLOWED 29-11-89

Australia Patent Declaration Form

Forms 7 and 8

AUSTRALIA

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Patents Act 1952

DECLARATION IN SUPPORT OF A CONVENTION OR NON-CONVENTION
APPLICATION FOR A PATENT OR PATENT OF ADDITIONName(s) of
Applicant(s)In support of the application made by BOARD OF REGENTS,
THE UNIVERSITY OF TEXAS SYSTEM

Title

for a patent for an invention entitled
MAGNETIC ALTERATION OF CELLULOSE DURING ITS BIOSYNTHESISName(s) and
address(es)
of person(s)
making
declarationI/~~we~~, Hans Mark, Chancellor for the BOARD OF REGENTS, THE UNIVERSITY
OF TEXAS SYSTEM, a state institution established according to the
Constitution of the state of Texas having a place of business at
201 West 7th Street, Austin, Texas 78701 United States of America
do solemnly and sincerely declare as follows:-

1. I am/~~we~~ the applicant(s) for the patent, or
am/are authorised by the abovementioned applicant
to make this declaration on its behalf.
2. The basic application(s) as defined by Section 141
of the Act was/were made in the following country
or countries on the following date(s) by the
following applicant(s) namely:-

Country, filing
date and name
of Applicant(s)
for the or
each basic
applicationin United States of America on April 3 19 85
by R. Malcolm Brown, Jr, Debra S. Brown and Michael R. Gletz
in _____ on _____ 19 ____
by _____

3. The said basic application(s) was/were the first
application(s) made in a Convention country in respect
of the invention the subject of the application.
4. The actual inventor(s) of the said invention is/are
R. Malcolm Brown, Jr. - 3509 Bonnie, Austin, Texas 78731 U.S.A.
Debra S. Brown - P.O. Box 157, Pasadena, California 91102 U.S.A.
Michael R. Gletz - 3524 Greystone Drive, Austin, Texas 78731 U.S.A.
5. The facts upon which the applicant(s) is/are entitled
to make this application are as follows:-
USSN: 719,505 Filed: April 3, 1985
The applicant is the assignee of the actual inventors.

Name(s) and
address(es)
of the or
each actual
inventorSee reverse
side of this
form for
guidance in
completing
this partDECLARED at austin, TX this 7 day of March 19 86Hans Mark
Hans Mark, Chancellor

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MAGNETIC ALTERATION OF CELLULOSE BIOSYNTHESIS

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(56) Prior Art Documents
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(57) Claim

1. A method of producing amorphous or disordered cellulose comprising:
incubating a microorganism actively producing cellulose in the presence of a magnetic field greater than 500 gauss.

16. Cellulose produced by a cellulose-producing microorganism subjected to a magnetic field greater than 500 gauss, wherein the cellulose is amorphous or disordered.

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Form 10

COMPLETE SPECIFICATION

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Complete Specification for the invention entitled:
MAGNETIC ALTERATION OF CELLULOSE
DURING ITS BIOSYNTHESIS

The following statement is a full description of this invention
including the best method of performing it known to me:-

5

10

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MAGNETIC ALTERATION OF
CELLULOSE DURING ITS BIOSYNTHESIS

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The present invention relates generally to amorphous cellulose and disordered cellulose compositions and to methods of making the same. The invention more particularly concerns a method of making altered cellulose by the action of a magnetic field upon cellulose-producing

25

organisms.

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Cellulose, a natural polysaccharide found throughout the plant world, is used for many purposes. Cellulosic products are usually obtained in various forms from plants such as trees, after different types of processing. The specific types of processing depend upon the uses to which the final cellulosic product is to be put.

35

Cellulose may also be obtained from cellulose-producing microorganisms. Such microbial cellulose, similar to plant derived cellulose in chemical structure, may be utilized for many analogous purposes.

The structure of microbial cellulose membranes has been studied by Purz et al (Faserforschung und Textil-technik V. 28(4) pp. 155-163, 1977 and V. 27(11) pp. 561-570, 1976) and determined to be an interwoven and dis-
5 ordered mesh of fibrillar strands with diameters of 50 mm to 100 mm.

Microbial cellulose has been produced in altered form by chemical alterations on the addition of fluorescent
10 brightening agents or direct cellulose dyes.

Magnetic fields have been implicated as having effects on many living systems. The growth of winter wheat seeds, for example, has been found to be stimulated
15 in a magnetic field of 100 gauss, this stimulation not being further increased by magnetic fields up to 1500 gauss (Cope, Physiol.: Chem. & Physics (1981) V. 13, pp. 567-568). Geacintov et al (Biochem. et Biophys. Acta (1972) V. 267, pp. 65-79 has shown a magnetic orientation
20 effect on chloroplasts and whole Chlorella cells. McKenzie and Pittman (Can. J. Plant Sci. (1980) V. 60, pp. 87-90) demonstrated that magnetotropic root growth is a plant characteristic inheritable through plant cytoplasm. A growth response of Lepidium seedlings exposed to mag-
25 netic field gradient was shown by Audus (Nature V. 185: pp. 132-134, 1960) and oriented growth of pollen tubes in a strong magnetic field has been demonstrated (Sperber and Dransfeld, Naturwissenschaften V. 68; pp. 40-41, 1981).

30 Adey (Physiological Review (1981) V. 61, pp. 435-514) has presented a general review of tissue interactions with nonionizing electromagnetic fields. In this review Adey (ibid, p. 463) cites evidence that agarose, a bacterial polysaccharide, is affected in orientation by magnetic
35 fields during an agarose gelation process. Agarose gelled in a 1.0T magnetic field (apparently about 16,667 gauss)

was about 7% more permeable to electrophoretically migrating bacterial DNA than was agarose gel formed at about 0.5 gauss.

5 Several polymers have been shown to adopt a particular orientation with respect to externally applied magnetic fields. Precipitation of collagen in a field of approximately 20 kilogauss resulted in highly ordered arrays of fibrils with their axes at right angles to the
10 field (Murthy, Biopolymers V. 23; pp. 1261-1267, 1984). Increasing the field beyond 20 kilogauss increased the order of the fibrils. Skeletal muscle actin has also been shown to adopt a preferred orientation in strong magnetic fields (Torbet and Dickens, FEBS Letts V. 173 p. 403,
15 1984).

 In U.S. Patent No. 4,020,590, Davis describes an apparatus and method for exposing seeds to a magnetic field and thereby altering seed germination and plant growth therefrom. In U.S. Patent No. 4,065,386 Rigby
20 describes a method of algae growth control wherein water is passed through a magnetic field. Amburn in U.S. Patent Nos. 3,910,233 and 3,991,714 describes methods and an apparatus for magnetically increasing the incidence of fertilization by fowl sperm and magnetically inducing
25 greater hatching rates of fertilized eggs.

 The present invention comprises a method of producing cellulose altered during its biosynthesis. Both eucaryotic and procaryotic cellulose-producing organisms, when
30 subjected to a magnetic field substantially greater than 0.5 gauss, produce altered cellulose. The altered cellulose produced by procaryotic cells is amorphous, has an increased absorptivity for water and greater chemical reactivity. The procaryotic cells are preferably in a
35 liquid medium comprising bacterial nutrients when subjected to the magnetic field. On the other hand, plant

tissue, (eucaryotic cells), when exposed to a magnetic field, generates altered cellulose microfibril patterns. The disordered arrangement of cellulose microfibrils affects the mechanical strength and other physical characteristics of the cell wall.

Cellulose-producing microorganisms useful in the present invention include members of the *Acetobacter*, *Rhizobium*, *Agrobacterium*, *Pseudomonas* and *Alcaligenes* genera (Brown et al. J. Applied Polymer Science Appl. Polymer Symp. (1983) V. 37 pp.33-78). The growth of such cellulose-producing microorganisms and the synthesis of cellulose occur generally in a suitable nutrient medium having a pH between about 3.5 and about 7.0 when incubated under aerobic conditions and at temperatures between about 20°C and about 35°C. A use of *Acetobacter xylinum* to coat synthetic fibers with microbial cellulose is disclosed in U.S. 4,378,431, issued to Brown, which is incorporated by reference herein. Many varieties of cellulose-producing microorganisms, particularly *Acetobacter xylinum*, exist and are virtually ubiquitous in natural surroundings such as damp forests, for example.

According to Bergey's Manual of Systematic Bacteriology Vol. 1, (ed. N.R. Krieg pp. 268-274, Williams and Wilkins, Baltimore, Md. 1984) cellulose synthesizing *Acetobacter* strains which were formerly classified as *Acetobacter aceti*, subspecies *xylinum* are now classified as subspecies of *Acetobacter pasteurianus* and *Acetobacter hansenii*. The strain of *Acetobacter xylinum* utilized herein is synonymous with *Acetobacter aceti*, subspecies *xylinum*. The Bergey reference cited above also refers to studies indicating that static culture favors cellulose synthesizing cells while shake cultures favor cellulose-free mutant cells.

The plant tissue considered here includes specifically the grass, Avena. Avena is a typical vascular plant, and as such, has the capacity to synthesize cell walls of cellulose (Raven, Evert, and Curtis, "Biology of Plants", 5 Worth Publishers, New York, 1976), hereafter referred to as "plant" cellulose.

The present invention comprises a process for altering microbial and plant cellulose during the production thereof by organisms. When cellulose-producing organisms are subjected to a magnetic field greater than that of the earth's magnetic field (i.e. about 0.5 gauss) during a period of cellulose production, the nature of their cellulose product is markedly altered. When such cellulose-producing microorganisms are removed from the magnetic field, their cellulose then produced is of a normal configuration and a return to the magnetic field results in a resumption of altered cellulose production.

Microbial cellulose produced under the influence of a magnetic field has an apparent crystallinity less than that normally found with microbial cellulose. The individual glucan chains interassociate with less force and order when formed in the presence of an effective magnetic field as judged by the ease of physically disrupting the resultant microfibril coalescence and the apparent decreased density and lack of microfibril bundle formation. The magnetically affected cellulose assumes a relatively amorphous configuration as compared to the normal and more orderly bundle or ribbon of microfibrils.

The magnetically altered form of microbial cellulose, having a much greater surface to volume ratio than that non-magnetically produced, has certain properties desirable for particular uses. For example, magnetically altered microbial cellulose has properties such as: A) a

greater capacity for water absorption; B) greater chemical reactivity; C) lower density; D) lower tensile strength; E) greater surface area; F) and greater potential for graft copolymerization. Since the altered or amorphous
5 cellulose glucan chains are less completely interasso-
ciated, the carbohydrate constituents thereof exhibit
greater chemical reactivity in general. Such properties
are of particular use in the development of effective
absorbents and textile products having a variety of
10 textures.

Plants exposed to magnetic fields significantly
higher than 0.5 gauss do not exhibit normal cellulose
microfibril deposition. Cellulose microfibrils in mag-
15 netic field treated tissue are deposited in considerable
disarray. The highly ordered nature of deposition of
plant cellulose is one of the major characteristics that
determine the physical properties of cellulosic plant
products (Preston, "The Physical Biology of Plant Cell
20 Walls", Chapman & Hall, 1974). Magnetic field induced
disruption of ordered deposition in plants provides a
method for development of wood and textile products with
different properties such as: materials with lower den-
sity, lower tensile strength, greater elasticity, or
25 greater accessibility to chemical treatment or modifi-
cations. Random and disarrayed microfibrill or patterns
result in the organism's cell walls having a lowered
density and tensile strength.

30 The magnetically induced disarray of microfibrillar
patterns will have further benefits as yet incompletely
defined. One example of these further benefits is the
production of houseplants whose normal patterns of growth
and conformation have been modified by exposure to a mag-
35 netic field of at least 500 gauss. These plants of
altered conformation will at least be of decorative use

and interest to those studying magnetic influence on plant growth patterns.

The culture of cellulose-producing organisms in
5 liquid nutrient medium and in a magnetic field substantially greater than 0.5 gauss may be performed to produce harvestable amounts of amorphous microbial cellulose. Said cellulose may be collected as a pellicle formed on a culture surface or by other means known by those skilled
10 in the art. The particular amorphous structure of the cellulose produced, although microscopically apparent, is as yet incompletely defined.

The following specific examples are presented to more
15 fully describe the present invention and are not meant to be limiting unless otherwise specifically designated in the accompanying claims.

Example 1

20

Acetobacter xylinum Cellulose Production
Under an Intense Magnetic Field

Bacterial cells of the species Acetobacter xylinum,
25 American Type Culture Collection (ATCC) No. 23769 were taken from a single colony and inoculated in 5 ml of Schramm & Hestrin culture medium (J. General Biology V. 11, pp. 1233-129, 1954) containing: glucose, 10 g/l; peptone, 5 g/l; yeast extract, 5 g/l; anhydrous dibasic
30 sodium phosphate, 2.7 g/l; and citric acid monohydrate, 1.15 g/l. The pH of the culture medium was adjusted to pH 6.0 and may be adjusted to between pH 3.5 and about pH 7.0 by addition of hydrochloric acid. The inoculated culture medium was aerobically incubated in an Erlenmeyer flask
35 under static conditions for a period of five days. A pellicle comprising microbial cellulose was formed near the

surface of the culture medium. The cellulosic pellicle was removed from the flask and washed with 50 mM pH 7.0 phosphate buffer containing 1M glucose. Acetobacter xylinum cells were removed from the washed pellicle by physical disruption of the pellicle in pH 7.0 phosphate buffer containing 1M glucose. The cells thus removed were placed in 10 mm thin walled sample tubes used for nuclear magnetic resonance studies. One tube of cells was inoculated at 31°C as a control, and another tube of cells was placed in the 1.8 tesla (1.8×10^4 gauss) magnetic field of a Varian FT80A nuclear magnetic spectrometer (Varian Inc.) The cells were incubated in the magnetic field at 31°C for a total period of 30 minutes.

Samples were withdrawn after magnetic and control incubation periods of 12 minutes and 30 minutes. The 30 minute magnetically-incubated cells were removed from the magnetic field, incubated for a 10 minute period at about 31°C in the absence of an artificial magnetic field, and then reintroduced into the magnetic field for a final 10 minute incubation, again at about 31°C. Cell samples were withdrawn after each of these 10 minute incubations.

Acetobacter xylinum cells and accompanying attached microbial cellulose in the withdrawn samples were subjected to negative staining with uranyl acetate and examined by electron microscopy. The results found with cells obtained after a 12 minute or 30 minute control incubation or magnetic incubation are summarized in Table 1.

With the control an electron micrograph showed uranyl acetate-stained Acetobacter xylinum producing a cellulose ribbon with normal, orderly fasciation of microfibrils in the absence of artificial magnetic fields. An electron micrograph showed uranyl acetate-stained Acetobacter

xylinum producing, under the influence of a magnetic field (18,000 gauss for 30 minutes at 31°C), cellulose as an amorphous mass. The amorphous cellulose mass is secreted from the cells as a web or blanket formation, rather than the normal orderly ribbon shown with the control.

Table 1

	<u>Control</u>		<u>Magnetically incubated</u>	
	<u>12 min</u>	<u>30 min</u>	<u>12 min</u>	<u>30 min</u>
Percent cells without visible cellulose product	45	71	69	74
Percent cells with normal cellulose ribbon attached	54	28	1	1
Percent cells with amorphous cellulose attached	1	1	30	25

A normal cellulose ribbon is produced by an orderly fasciation of cellulose microfibrils biosynthesized and secreted by Acetobacter cells. When cellulose biosynthesis and secretion occurred in the magnetic field, the microfibrils were not subject to orderly fasciation and formed an amorphous mass with little in the way of crystalline structure.

It was further determined that orderly fasciation resumed when the Acetobacter cells were removed from the magnetic field, the amorphous cellulose mass now becoming attached to the cells by a normal cellulose ribbon. Upon reintroduction into the magnetic field, amorphous cellulose production resumed resulting in cells with a chain of cellulose attachments, i.e., amorphous mass; cellulose ribbon; amorphous mass.

The amorphous nature of the magnetically induced cellulose product was found with Acetobacter cultures between about pH 4.8 and pH 7.0. It was also found that intermittent subjection of the cellulose-producing organisms to the magnetic field resulted in an increase in the incidence of cellulose microfibrillar disorder or amorphousness resulting from the magnetic field.

EXAMPLE 2

10

Structure of Amorphous Acetobacter Cellulose.

The procedures and components described in Example 1 were utilized, but with the following modification. The Acetobacter xylinum cells were isolated in 50 mM, pH 7.0 phosphate buffer from the cellulosic pellicle and all incubations were at 32°C. The magnetic field was that of a Varian E-9 EPR Spectrometer, with a constant flux density of 6,000 gauss. Cells were incubated in succession for ten minutes in the magnetic field, ten minutes in the absence of the magnetic field, ten more minutes in the field, and finally ten minutes without the field. Samples of the cells were then subjected to treatment with colloidal gold particles or with the enzyme, cellobiohydrolase with adherent colloidal gold particles according to the method of Chanzy, et al. (1984, FEBS Letts, V. 172, pp. 193-197). An electron micrograph demonstrated the adherence of the gold-cellobiohydrolase to the amorphous cellulose. This adherence indicates the integrity of beta 1,4-linked glucan bonds in the amorphous cellulose. An electron micrograph demonstrated that colloidal gold particles alone have no specific adherence for the amorphous cellulose.

EXAMPLE 3

Amorphous Cellulose Production
at 500 gauss and 6,000 gauss.

5

Utilizing the instrumentation and procedures generally described in Example 2, *Acetobacter xylinum* cells were subjected to magnetic fields with the following modification. In one case, cells were subjected to a 500
10 gauss field for twenty minutes, followed by a ten minute 'rest' period without any artificial magnetism. Control cells were incubated for forty minutes in the absence of an artificial magnetic field. Electron micrographs of the various cells were obtained and examined. The data
15 resulting from this examination are shown in Table 2.

Table 2. Effects of a 500 G magnetic field and intermittent exposure to 6000 G field on Acetobacter xylinum cellulose production.

Conditions of Treatment	500 G field 20 minutes	6000 G field for 10 minutes (2 exposures) with a 10 minute "recovery period" between exposures	40 minutes
Percent of total cells exhibiting no cellulose production	41	63	89
Percent of total cells producing a normal cellulose ribbon	13	6	9
Percent of total cells producing atypical/ amorphous cellulose	46	31	3

TABLE 2

- 12 -

*All cells were isolated 50mM phosphate buffer, pH 7.0, and incubated in this buffer at 32 C during all treatments. Magnetic field treatments were followed by a 10 minute "recovery period" during which no field was applied.

As shown in Table 2, the extent of amorphous cellulose production is enhanced by both the 500 gauss magnetic field and the 6,000 gauss magnetic field. In addition, there is an overall stimulatory effect of magnetic fields on cellulose production, as yet incompletely understood.

EXAMPLE 4

10 Three day old seedlings of *Avena sativa* CV Garry, (Agway, Syracuse, N.Y.) were grown in the dark at 25°C and were transferred to 10mm thin-walled glass NMR tubes with a small amount of growth medium. Coleoptiles were oriented perpendicular to the field lines in a Varian FT80
15 spectrometer magnet (18,000 gauss) and exposed to the field for 30 minutes. Control plants were incubated at 31°C for 30 minutes outside of the applied magnetic field. Coleoptiles were sliced into small segments directly after 30 minutes, immediately mounted on gold holders and frozen
20 in freon cooled with liquid nitrogen. Specimens were fractured in a Balzers BA360 Freeze Fracture instrument (Balzers, Lichtenstein) and replicas were examined in a Philips 420 transmission electron microscope (Eindhoven, Netherlands).

25 Cellulose microfibrils of *Avena* synthesized in the magnetic field were randomly organized and loosely associated with one another. This disarray was accompanied by extensive disruption of intramembranous particle distributions in the plasma membrane. Inner wall layers of
30 control plants showed highly ordered, closely associated arrays of microfibrils that exhibited wall patterns typically found in *Avena* coleoptiles. Disruption in patterns of microfibril deposition in *Avena* was found to occur in magnetic fields with a field strength as low as 5,000
35 gauss.

An electron micrograph of freeze-fractured control Avena coleoptile plasma membrane outer leaflet indicated ordered and uniform cellulose microfibril impressions. The cellulose microfibril impressions reflected that
5 cellulose microfibrils demonstrate a random and disarrayed pattern which may lead to a lowered density of the material tensile strength and resulting from the applications of the artificial magnetic field.

10 Upon examination of the presently disclosed invention, it will be apparent to those skilled in the art that changes in the order of procedural steps and particular composition elements may be made without departing from the spirit and scope of the invention as described in the
15 following claims.

THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:

1. A method of producing amorphous or disordered cellulose comprising:
incubating a microorganism actively producing cellulose in the presence of a magnetic field greater than 500 gauss.
2. The method of Claim 1 wherein the cellulose producing microorganism is defined further as being a procaryotic microorganism.
3. The method of Claim 2 wherein the procaryotic microorganism is in a liquid culture medium having a pH between pH 3.5 and pH 7.0.
4. The method of Claim 1 wherein the amorphous cellulose produced in the magnetic field is defined further as being microbial cellulose.
5. The method of Claim 1 wherein the amorphous cellulose is defined further as having a water-absorptive capacity greater than cellulose produced in the absence of an artificial magnetic field.
6. The method of Claim 1 wherein the amorphous cellulose is defined further as being more chemically reactive than cellulose produced in the absence of an artificial magnetic field.
7. The method of Claim 1 wherein the magnetic field is defined further as being 6,000 gauss.
8. The method of Claim 1 wherein the magnetic field is defined further as being 18,000 gauss.
9. The method of Claim 1 wherein the incubating step is conducted in an intermittent magnetic field greater than 500 gauss.



10. The method of Claim 2 wherein the procaryotic microorganism is of a genus selected from the group consisting of Acetobacter, Rhizobium, Agrobacterium, Pseudomonas or Alcaligene.

11. The method of Claim 1 wherein the cellulose-producing microorganism is defined further as being Acetobacter xylinum, ATCC number 23769.

12. The method of Claim 2 wherein the liquid culture medium is defined further as comprising bacterial nutrients.

13. The method of Claim 1 wherein the microorganism is defined further as being an eucaryotic microorganism, and the cellulose is defined further as being microfibrillar extracellular product having a random and disarrayed pattern.

14. The method of Claim 13, wherein the random and disarrayed pattern is defined further as resulting in microorganism cell walls having a lowered density and tensile strength compared to cellulose produced by plants not exposed to a magnetic field of greater than 0.5 gauss.

15. The method of Claim 13 wherein the cellulose is defined further as being less dense than and as having less tensile strength than cellulose produced in the absence of an artificial magnetic field.

16. Cellulose produced by a cellulose-producing microorganism subjected to a magnetic field greater than 500 gauss, wherein the cellulose is amorphous or disordered.



14.
~~18.~~ The cellulose of Claim 16 defined further as being produced by procaryotic microorganisms of a genus selected from the group consisting of Acetobacter, Rhizobium, Agrobacterium, Pseudomonas or Alcaligenes.

DATED this 25th day of May 1989

BOARD OF REGENTS, THE UNIVERSITY OF TEXAS SYSTEMS
By Their Patent Attorneys:

GRIFFITH HACK & CO
Fellows Institute of Patent
Attorneys of Australia

