



US 20090139134A1

(19) **United States**

(12) **Patent Application Publication**
Yoshikuni et al.

(10) **Pub. No.: US 2009/0139134 A1**

(43) **Pub. Date: Jun. 4, 2009**

(54) **BIOFUEL PRODUCTION**

Publication Classification

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(51) **Int. Cl.**
C10L 1/18 (2006.01)
C12P 7/06 (2006.01)
C12P 7/04 (2006.01)
C12N 1/21 (2006.01)
C07C 31/08 (2006.01)

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(52) **U.S. Cl. 44/307; 435/161; 435/157; 435/252.3;**
568/840

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(57) **ABSTRACT**

(21) Appl. No.: **12/245,537**

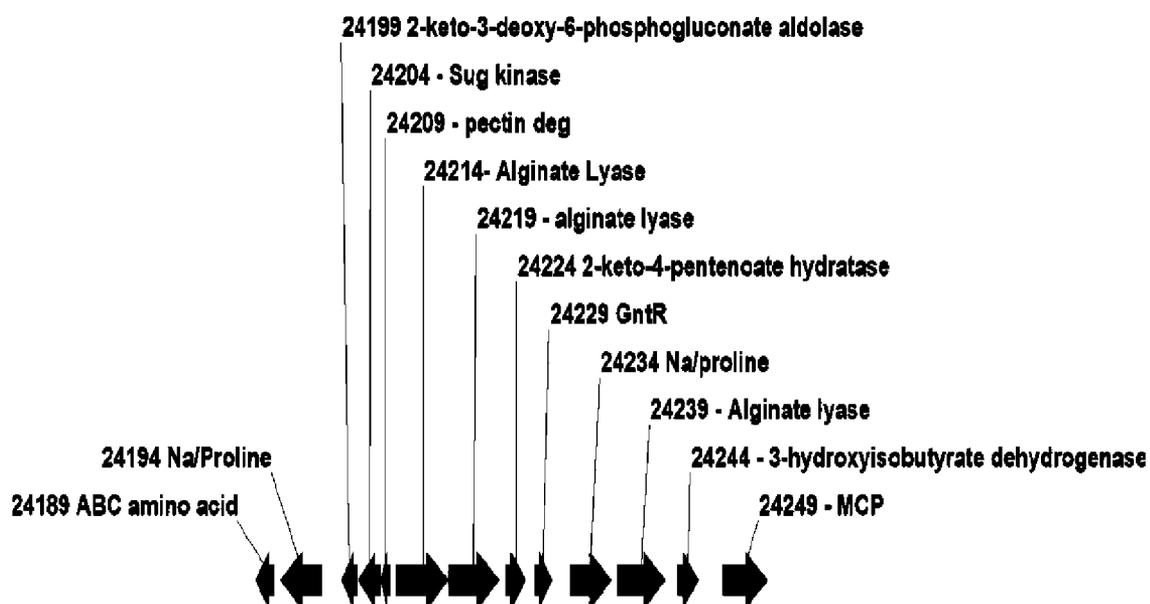
(22) Filed: **Oct. 3, 2008**

Methods, enzymes, recombinant microorganism, and microbial systems are provided for converting polysaccharides, such as those derived from biomass, into suitable monosaccharides or oligosaccharides, as well as for converting suitable monosaccharides or oligosaccharides into commodity chemicals, such as biofuels. Commodity chemicals produced by the methods described herein are also provided. Commodity chemical enriched, refinery-produced petroleum products are also provided, as well as methods for producing the same.

Related U.S. Application Data

(60) Provisional application No. 60/977,628, filed on Oct. 4, 2007.

Figure 1



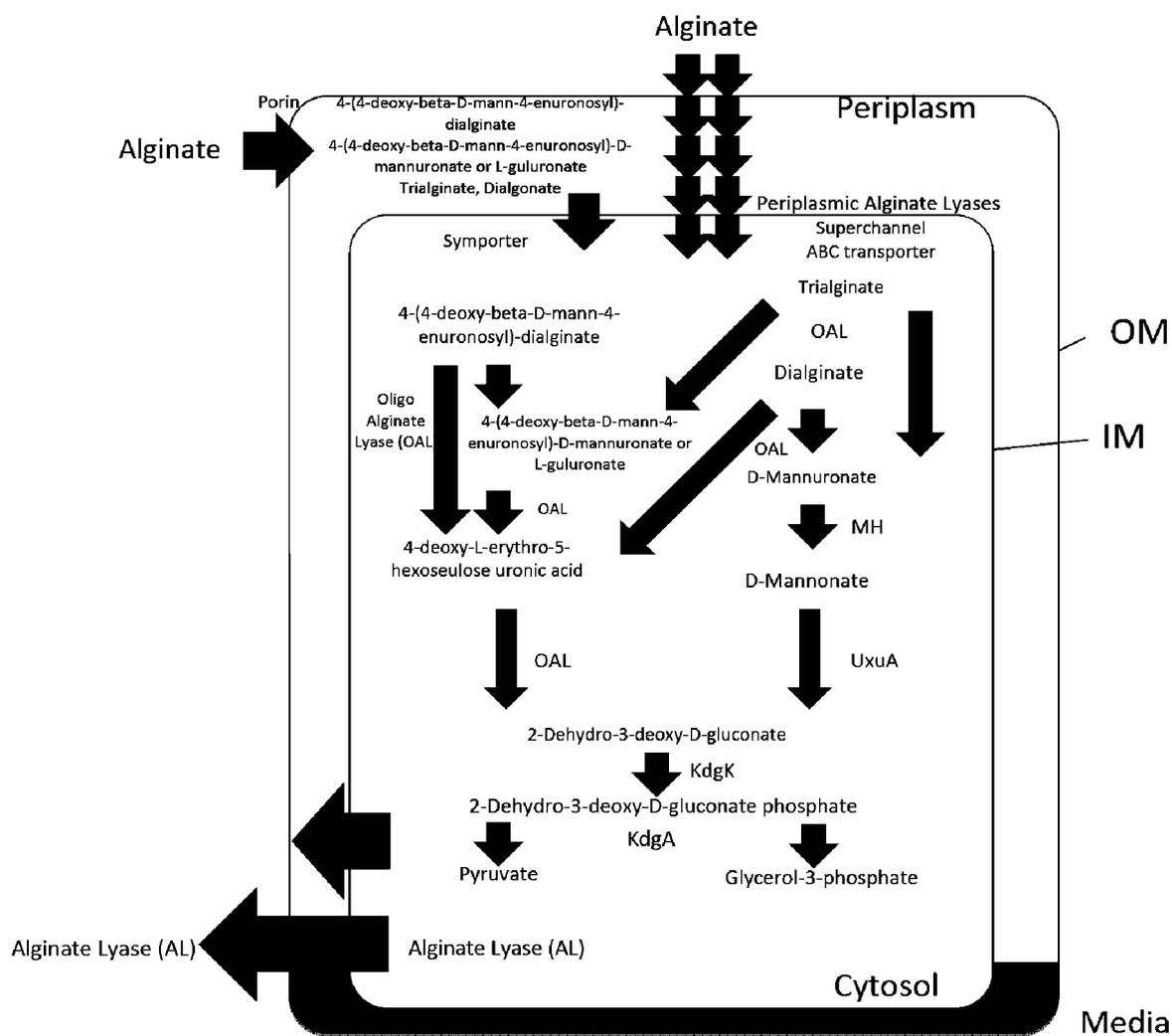
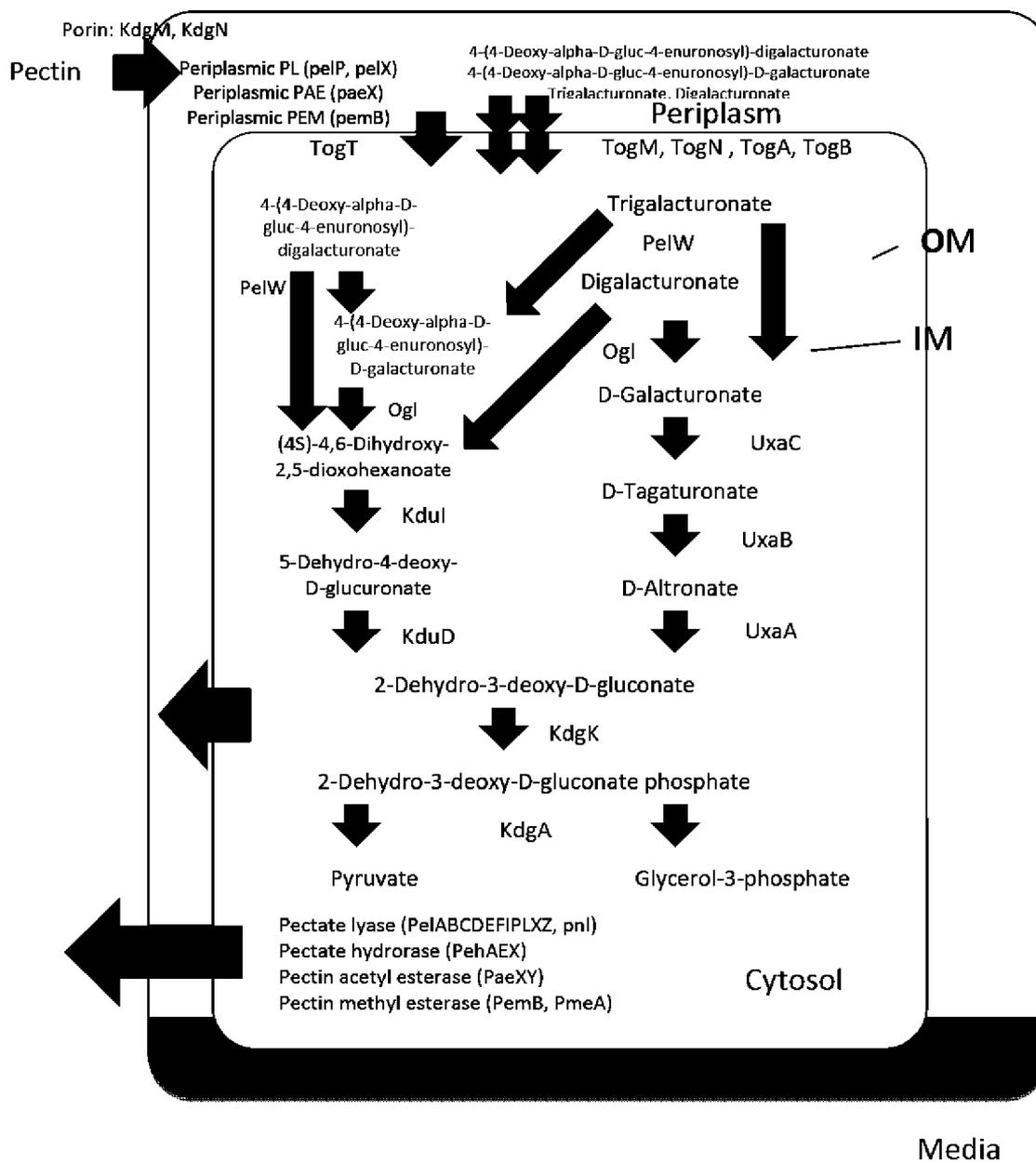


Figure 2

Figure 3



E. coli Growing on Alginate

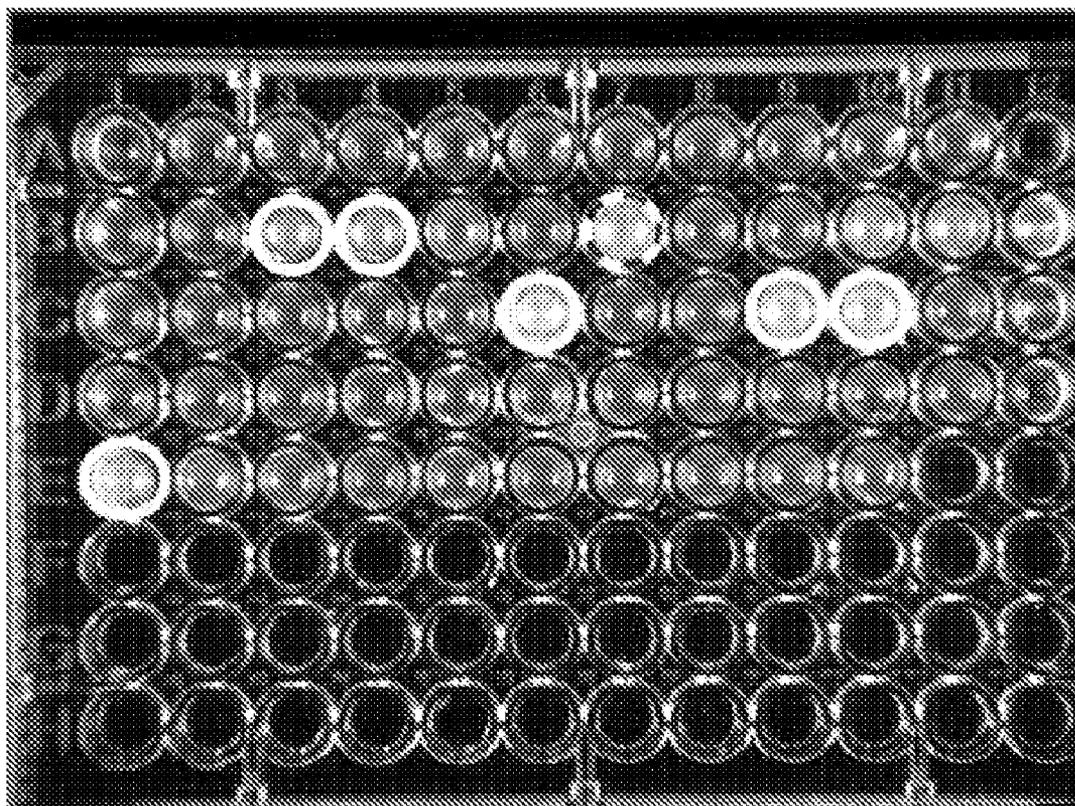


FIG. 4

Figure 5A

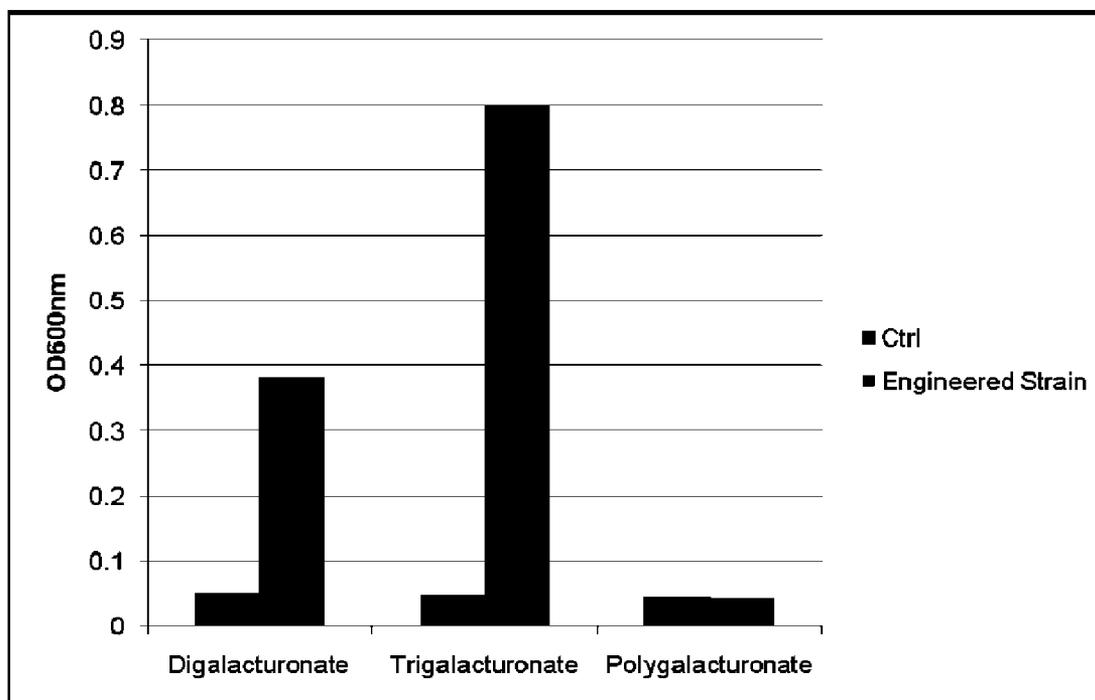
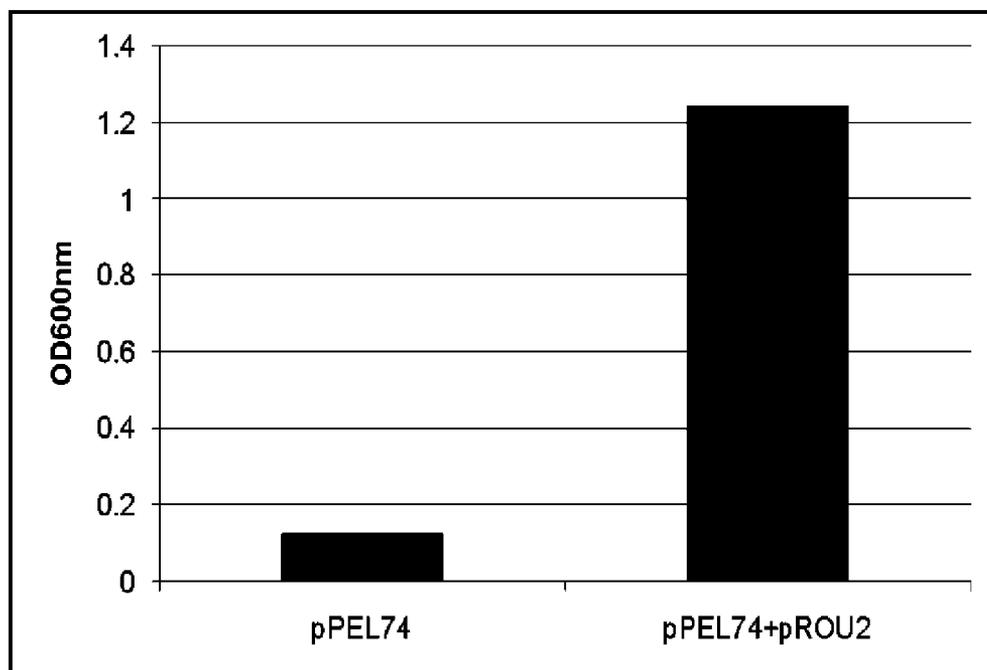


Figure 5B



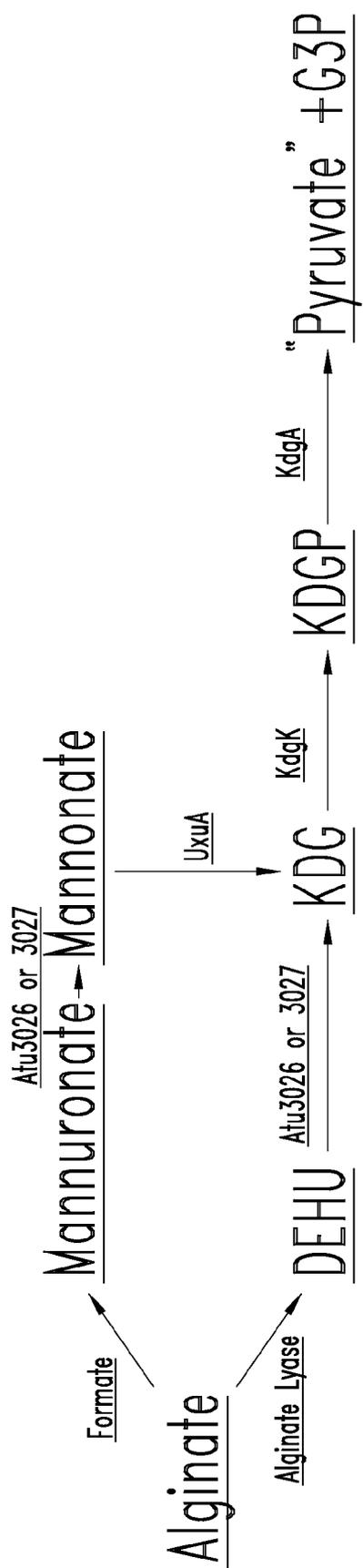


FIG. 6A

Figure 6B

Pyruvate formation from alginate (enzymatic degradation route)

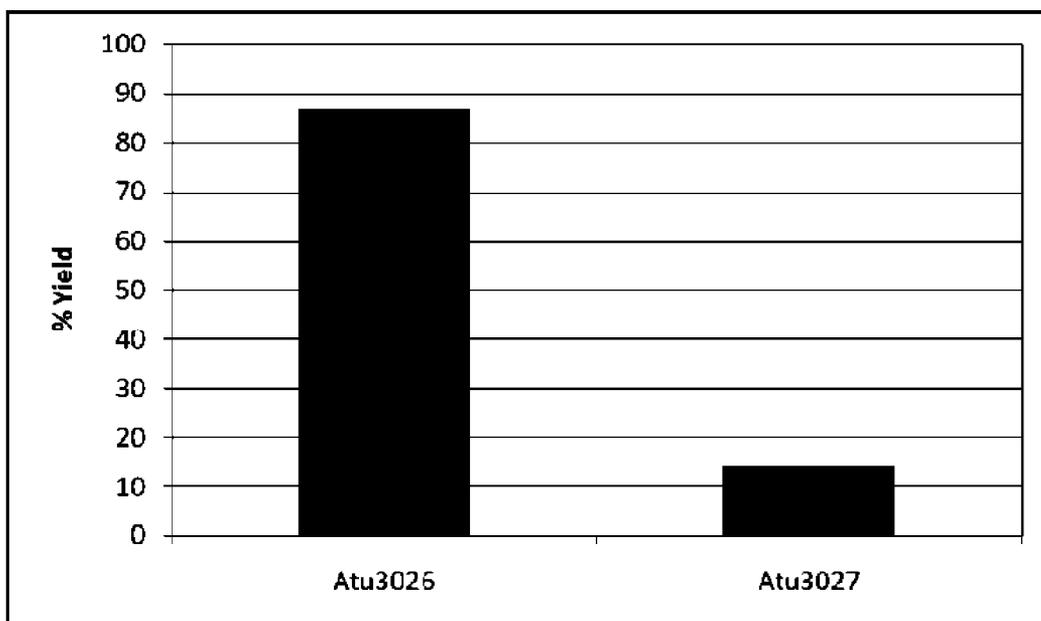
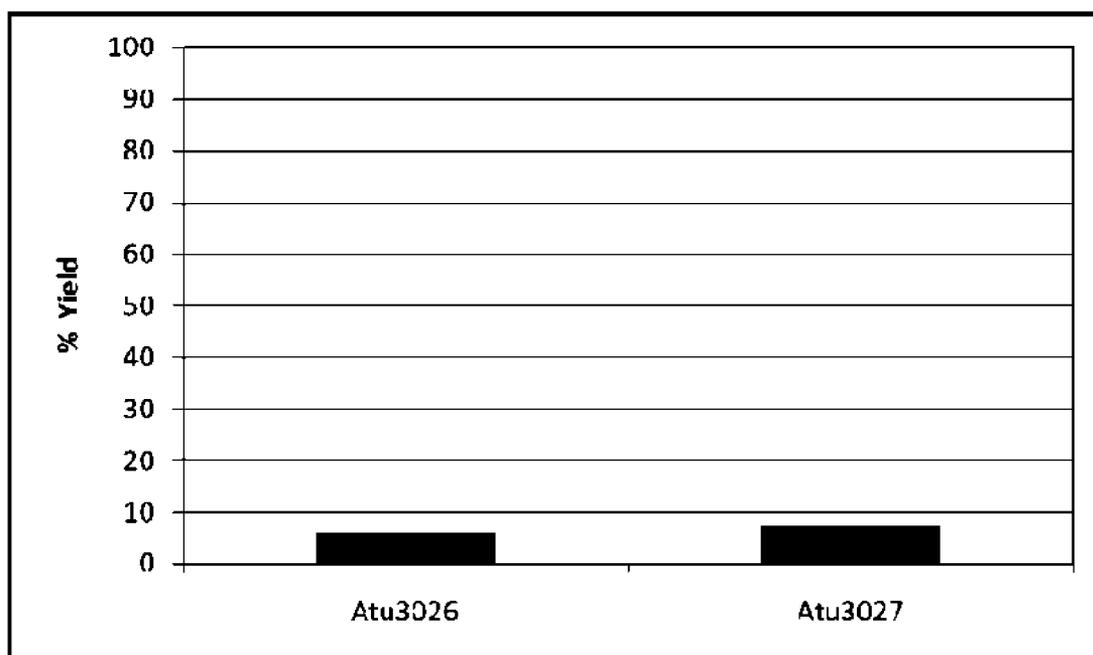


Figure 6C

Pyruvate formation from alginate (chemical degradation route).



GC-MS chromatogram of control

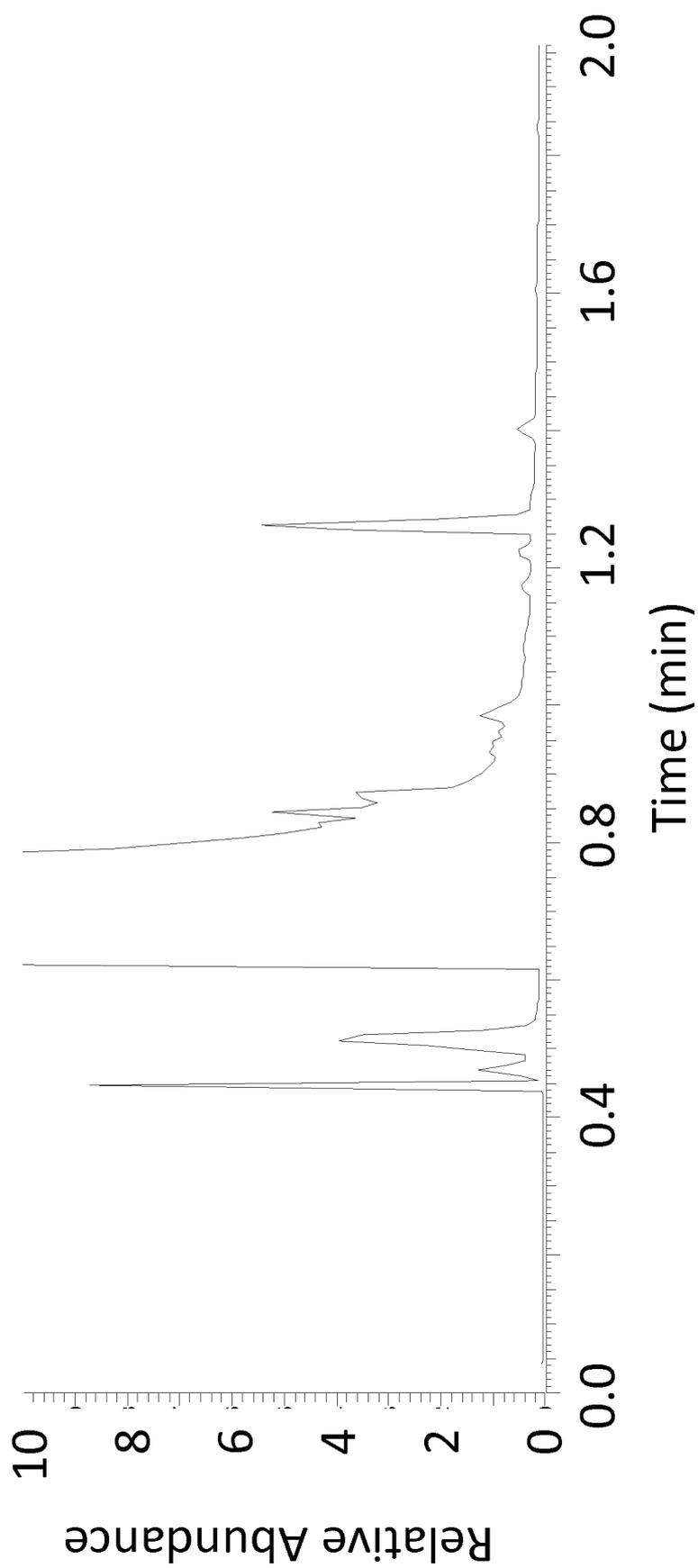


FIG. 8A

GC-MS chromatogram for isobutyraldehyde, 3-methylpentanol, and 2-methylpentanol production from pBADalsS-*ilv*CD-leuABCD2

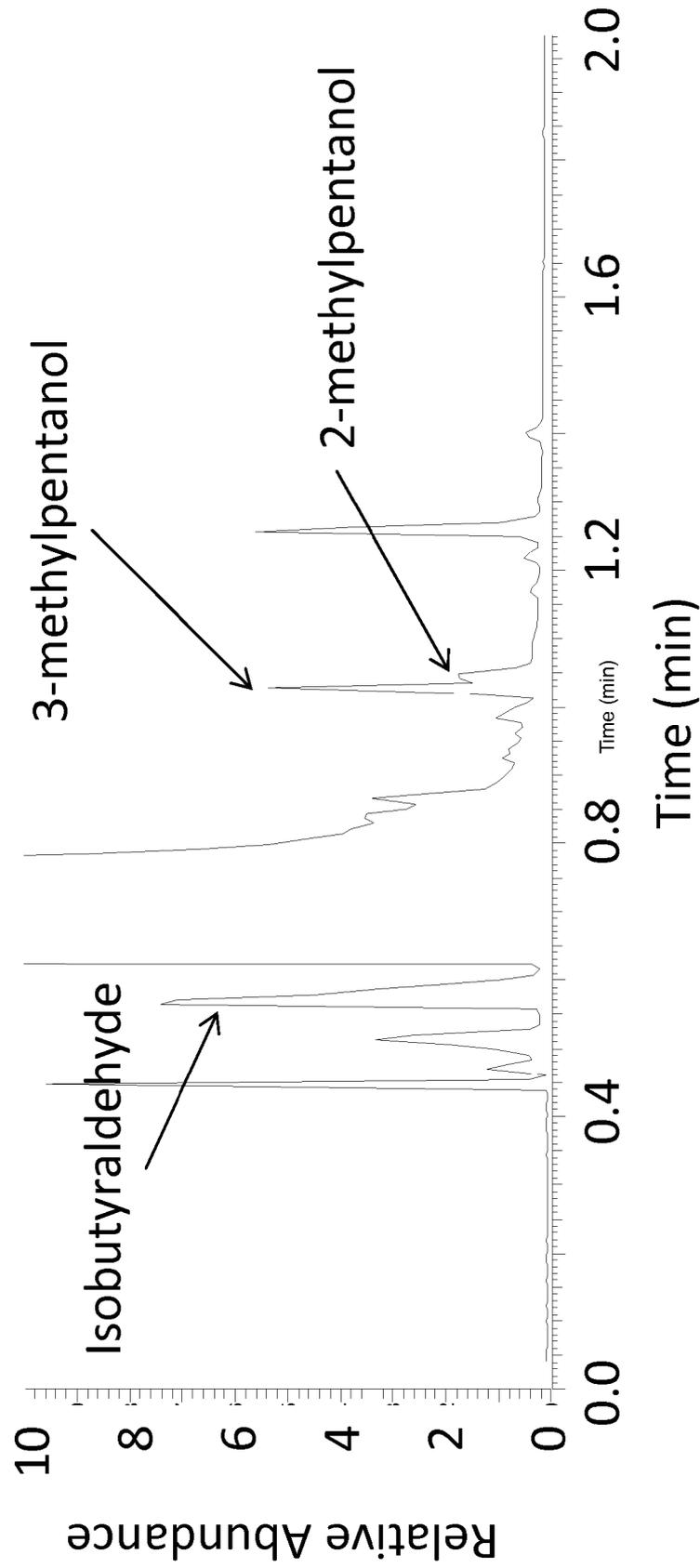


FIG. 8B

Figure 9A
GC-MS chromatogram of Control

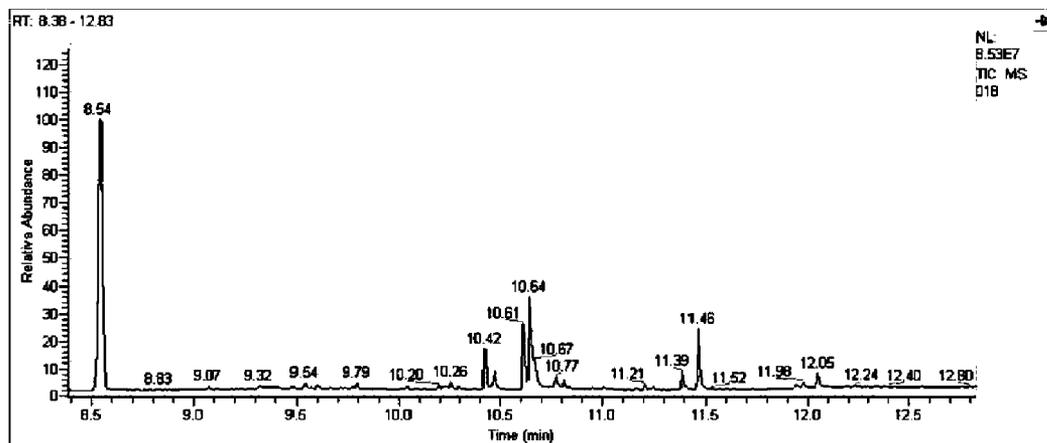


Figure 9B
GC-MS chromatogram for 4-hydroxyphenylethanol and indole-3-ethanol production from pBADtyrA-aroLAC-aroG-tktA-aroBDE and pTrcBALK

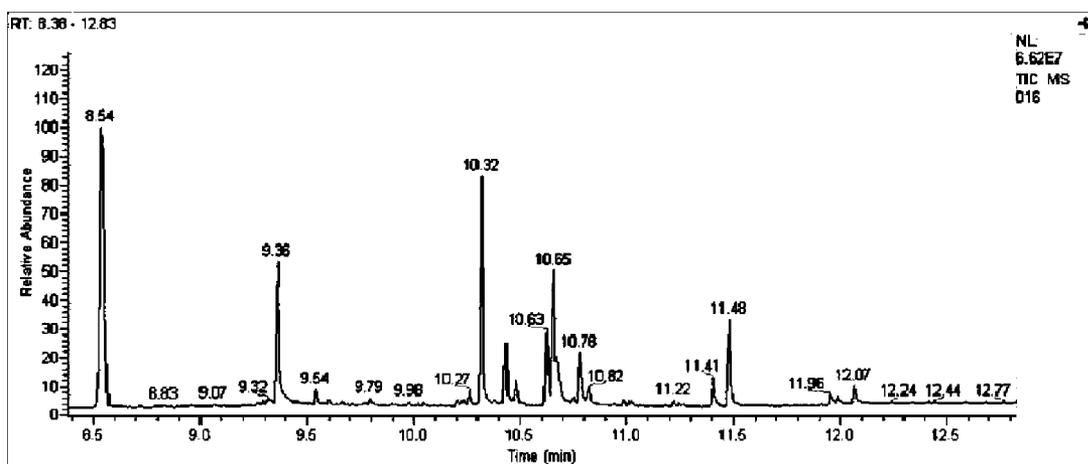


Figure 10A
Mass spectrometry of isobutanal

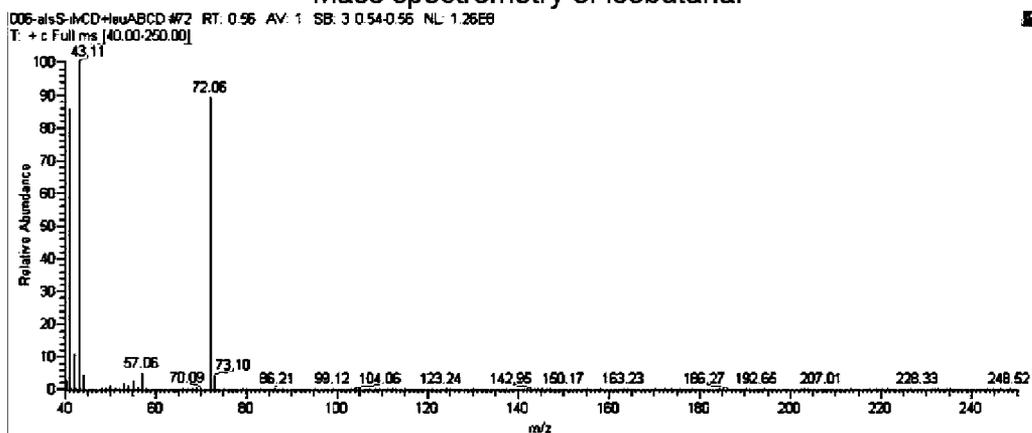


Figure 10B
Mass spectrometry of 3-methylpentanol

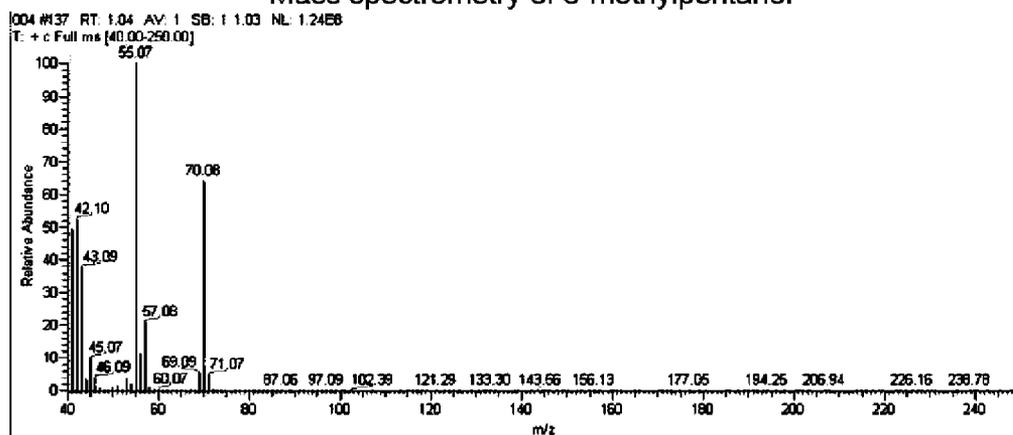


Figure 10C
Mass spectrometry of 2-methylpentanol

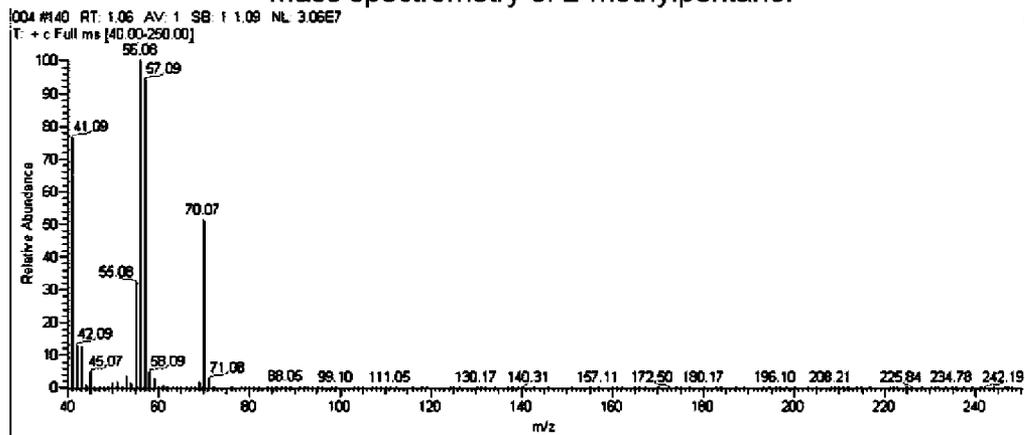


Figure 11A

Mass spectrometry of 2-phenylethanol

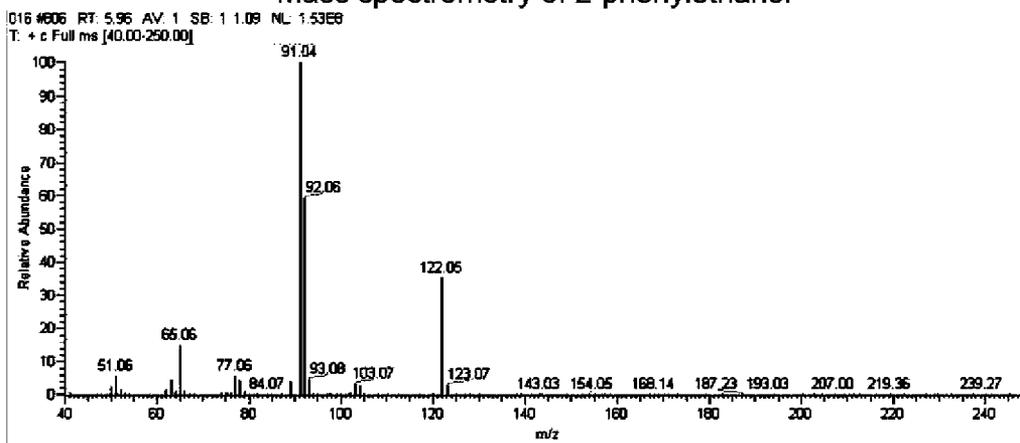


Figure 11B

Mass spectrometry of 4-hydroxyphenylethanol

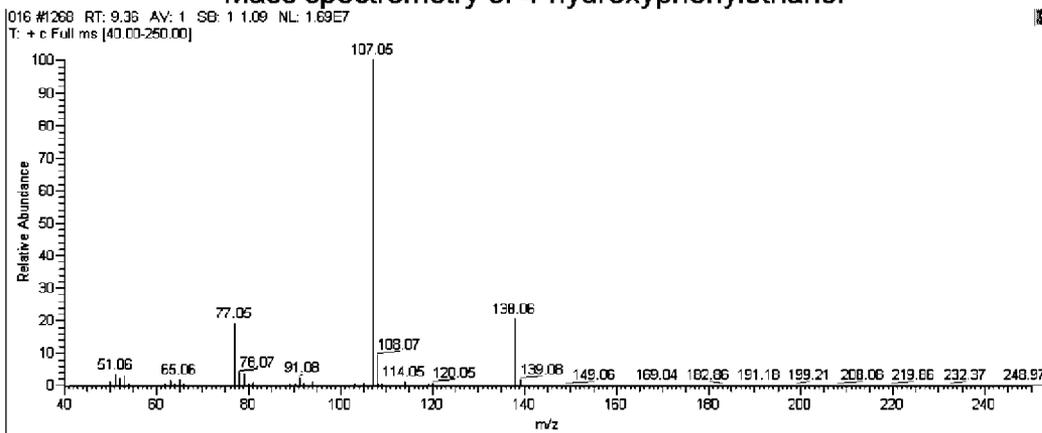


Figure 11C

Mass spectrometry of indole-3-ethanol

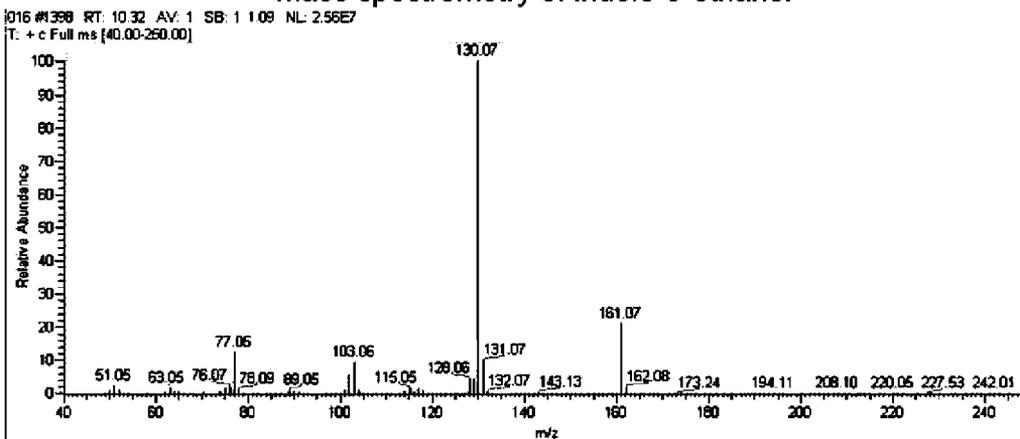


Figure 12A

Reduction of butyoin by *ddh1*, *ddh2*, and *ddh3* monitored by NADH consumption.

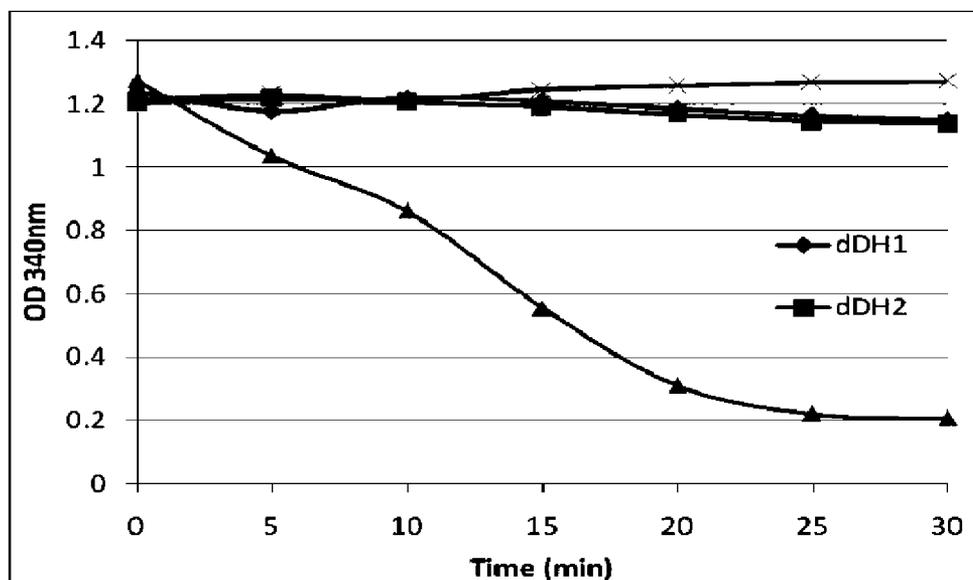


Figure 12B

Oxidation activity of *ddh3* towards 1,2-cyclopentane diol and 1,2-cyclohexane diol as measured by NADH production.

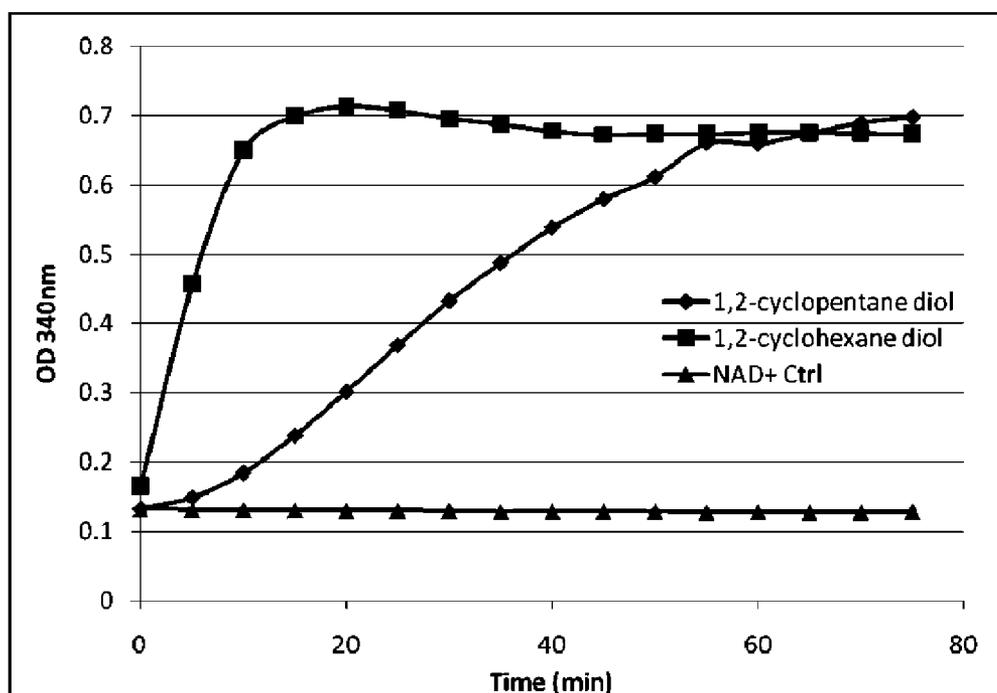


Figure 13

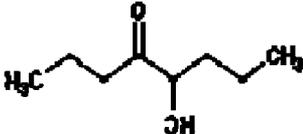
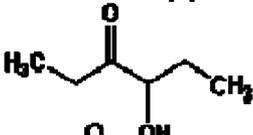
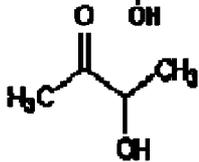
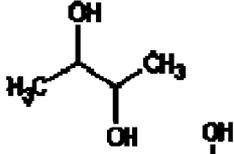
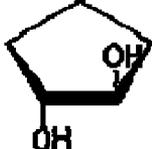
- Butyrolin: $k_{cat}/K_M = 139 \text{ sec}^{-1}\text{M}^{-1} \pm 9$ 
- Propiolin: $k_{cat}/K_M = 709 \text{ sec}^{-1}\text{M}^{-1} \pm 40$ 
- Acetoin: $k_{cat}/K_M = 1990 \text{ sec}^{-1}\text{M}^{-1} \pm 210$ 
- meso-2,3-butanediol: $k_{cat}/K_M = 1450 \text{ sec}^{-1}\text{M}^{-1} \pm 150$ 
- cis-1,2-cyclohexanediol: $k_{cat}/K_M = 108 \text{ sec}^{-1}\text{M}^{-1} \pm 15$ 
- trans-1,2-cyclopentandiol: $k_{cat}/K_M = 19.1 \text{ sec}^{-1}\text{M}^{-1} \pm 4.1$ 

Figure 14A

Nucleotide sequence of diol dehydrogenase DDH1 isolated from *Lactobacillus brevis* ATCC 367

ATGGCATCAAATGGAAAAGTAGCAATGGTTACCGGTGGCGGACAAGGAATTGGTGAAGC
CATCTCGAAACGGTTAGCTAACGACGGCTTTGCTGTGGCAATTGCTGATTTGAACTTGG
ACAATGCCAACAAAGGTCGTTTCTGATATTGAAGCTGCTGGTGGCAAGGCCATTGCGGTC
AAGACCGATGTCTCTGATCGTGATAGCGTGTGTTGCTGCGGTTAATGAAGCGGCCGACAA
GCTGGGCGGCTTTGACGTTATCGTTAATAACGCCGGCCTTGGCCAACCACGCCAATTG
ACACCATCACCCAAGAACAGTTTGATACGGTTTATCACGTTAACGTGGGTGGGGTTCTT
TGGGGCATTCAAGCAGCCCATGCGAAGTTCAAGGAATTGGGTCATGGTGGGAAGATCAT
TTCCGCGACGTCTCAAGCCGGGGTTGTTGGTAACCCGAACCTTAGCTCTGTACAGTGGAA
CTAAGTTTGCCATTCGTGGTGTGACCCAAGTTGCGGCGCGTGACTTAGCCGCTGAAGGT
ATCACGGTCAATGCTTATGCACCCGGGATTGTTAAGACACCAATGATGTTTGACATCGC
TCACAAGGTTGGTCAAAATGCTGGTAAAGACGACGAATGGGGGATGCAAACCTTCTCAA
AGGACATCGCTTTATGTCGATTGTCAGAACCAGAAGATGTGGCTAACGGGGTGGCTTTC
TTAGCCGGTCCCGATTCTAACTACATTACGGGTCAAACACTTGAAGTTGATGGTGGGAT
GCAGTTCCACTAA (SEQ ID NO:97)

Figure 14B

Polypeptide primary sequence of diol dehydrogenase DDH1 isolated from *Lactobacillus brevis* ATCC 367

MASNGKVAMVTGGGQGIGEAISKRLANDGFAVAIADLNLDNANKVVS DIEAAGGKAI AV
KTDVSDRDSVFAAVNEAADKLGDFDIVN NAGLGPTTPI DTTITQE QFDTVYHVNVGGVL
WGIQAAHAKFKELGHGGKII SATSQAGVVG NPNLALYSGTKFAIRGVTQVAARDLAAEG
ITVNAYAPGIVKTPMMFDIAHKVGNAGKDDEWGMQTF SKDIALCRLSEPEDVANGVAF
LAGPDSNYITGQTLEVDGGMQFH (SEQ ID NO:98)

Figure 15A

Nucleotide sequence of diol dehydrogenase DDH2 isolated from *Pseudomonas putida* KT2440

ATGAATGACCTGAGCCACACCCACATGCGCGCGGCCGTCTGGCATGGCCGCCACGATAT
TCGTGTCGAACAGGTACCTTTGCCGGCCGACCTGCGCCGGGCTGGGTGCAGATCAAGG
TGGACTGGTGCGGCATCTGCGGCTCCGACCTGCACGAATATGTTGCCGGCCCCGGTGTTT
ATCCCGGTAGAGGCCCGCACCCGCTGACCGGCATTCAGGGCCAGTGCATCCTCGGCCA
CGAATTCTGCGGCCACATCGCCAAGCTTGGCGAAGGCGTGGAAGGCTATGCCGTAGGCG
ACCCGGTGGCGGCAGACGCGTGCCAGCATTGTGGTACCTGCTATTACTGCACCCATGGC
CTGTACAACATCTGCGAACGCCCTGGCGTTACCCGGCCTGATGAACAACGGTGCCTTCCG
CGAGCTGGTCAACGTGCCCGCCAACCTGCTCTACCCGGCTGCCGCAGGGCTTCCCTGCCG
AAGCCGGGGCACTGATCGAGCCGCTGGCGGTGGGTATGCACGCGGTGAAAAAGCCGGC
AGCCTGCTTGGGCAAACCGTTGTAGTGGTTGGGGCCGGCACCATCGGCCTGTGCACCAT
CATGTGCGCAAGGCTGCAGGTGCGGCACAGGTCATCGCCCTTGAGATGTCTCTGCGC
GCAAAGCCAAGGCCAAGGAAGCGGGCGCCAACGTGGTGGTGGACCCCAGCCAGTGCAT
GCCCTGGCGGAAATCCGCGCACTGACTGCTGGGCTGGGCGCCGATGTGAGTTTTGAGTG
CATCGGCAACAAACATACGGCCAAGCTGGCCATCGACACCATCCGCAAAGCAGGCAAGT
GCGTGCTGGTGGGTATTTTCGAAGAGCCCAGCGAGTTCAACTTCTTCGAGCTGGTGTCC
ACCGAGAAGCAAGTGCTGGGGGCGTTGGCGTACAACGGCGAGTTTGCTGACGTGATTGC
CTTCATTGCTGATGGTCCGGCTGGATATTCGCCCCGCTGGTAACCCGGCCGGATCGGATTGG
AGCAGATTGTCGAGCTGGGCTTCGAGGAAGTGGTGAACAACAAAGAGGAGAACGTGAAG
ATCATCGTTTACCAGGTGTGCGCTGA (SEQ ID NO:99)

Figure 15B

Polypeptide sequence of diol dehydrogenase DDH2 isolated from *Pseudomonas putida* KT2440

MNDLSHTHMRAAVWHGRHDIRVEQVPLPADPAPGWVQIKVDWCGICGSDLHEYVAGPVF
IPVEAPHPLTGIQGQCILGHEFCGHIAKLGEVVEGYAVGDPVAADACQHCGTCTYYCTHG
LYNICERLAFTGLMNGAFAELVNVPANLLYRLPQGFPAEAGALIEPLAVGMHAVKKAG
SLLGQTVVVVAGTIGLCTIMCAKAAGAAQVIALEMSSARKAKAKEAGANVVLDPSQCD
ALAEIRALTAGLGADVSEFCIGNKHTAKLAIDTIRKAGKCVLVGIFEEPSEFNFFELVS
TEKQVLGALAYNGEFADVIAFIADGRDIRPLVTGRIGLEQIVELGFEELVNNKEENVK
IIVSPGVR (SEQ ID NO:100)

Figure 16A

Nucleotide sequence of diol dehydrogenase DDH3 isolated from *Klebsiella pneumoniae* MGH78578

ATGAAAAAAGTCGCACTTGTACC GGCGCCGGCCAGGGGATTGGTAAAGCTATCGCCCT
TCGTCTGGTGAAGGATGGATTTGCCGTGGCCATTGCCGATTATAACGACGCCACCGCCA
AAGCGGTCGCCTCGGAAATCAACCAGGCCGGCGGACACGCCGTGGCGGTGAAAGTGGAT
GTCTCCGACCGCGATCAGGTATTTGCCGCCGTTGAACAGGCGCGCAAACGCTGGGCGG
CTTCGACGTCATCGTCAATAACGCCGGTGTGGCACCGTCTACGCCGATCGAGTCCATTA
CCCCGGAGATTGTCGACAAAGTCTACAACATCAACGTCAAAGGGGTGATCTGGGGTATT
CAGGCGGCGGTTCGAGGCCTTTAAGAAAGAGGGGACCGCGGGAAAATCATCAACGCCTG
TTCCCAGGCCGGCCACGTCGGCAACCCGGAGCTGGCGGTGTATAGCTCCAGTAAATTCG
CGGTACGCGGCTTAACCCAGACCGCCGCTCGCGACCTCGCGCCGCTGGGCATCACGGTC
AACGGCTACTGCCCGGGGATTGTCAAACGCCAATGTGGGCCGAAATTGACCGCCAGGT
GTCCGAAGCCCGCGGTAAACCGCTGGGCTACGGTACCGCCGAGTTCGCCAAACGCATCA
CTCTCGGTCGTCTGTCCGAGCCGGAAGATGTCGCCGCCCTGCGTCTCCTATCTTGCCAGC
CCGATTCTGATTACATGACCGGTCAGTCGTTGCTGATCGACGGCGGGATGGTATTTAA
CTAA (SEQ ID NO:101)

Figure 16B

Polypeptide sequence of diol dehydrogenase DDH3 isolated from *Klebsiella pneumoniae* MGH78578

MKKVALVTGAGQGIGKAIALRLVKDGFVAVAIADYNDATAKAVASEINQAGGHAVAVKVD
VSDRDQVFAAVEQARKTLGGFDVIVNNAGVAPSTPIESITPEIVDKVYNINVKGVIWGI
QAAVEAFKKEGHGGKIINACSQAGHVGNPVELAVYSSSKFAVRGLTQTAARDLAPLGITV
NGYCPGIVKTPMWAEIDRQVSEAAGKPLGYGTAEFAKRITLGRLEPEDVAACVSYLAS
PDSYMTGQSLLIDGGMVFN (SEQ ID NO:102)

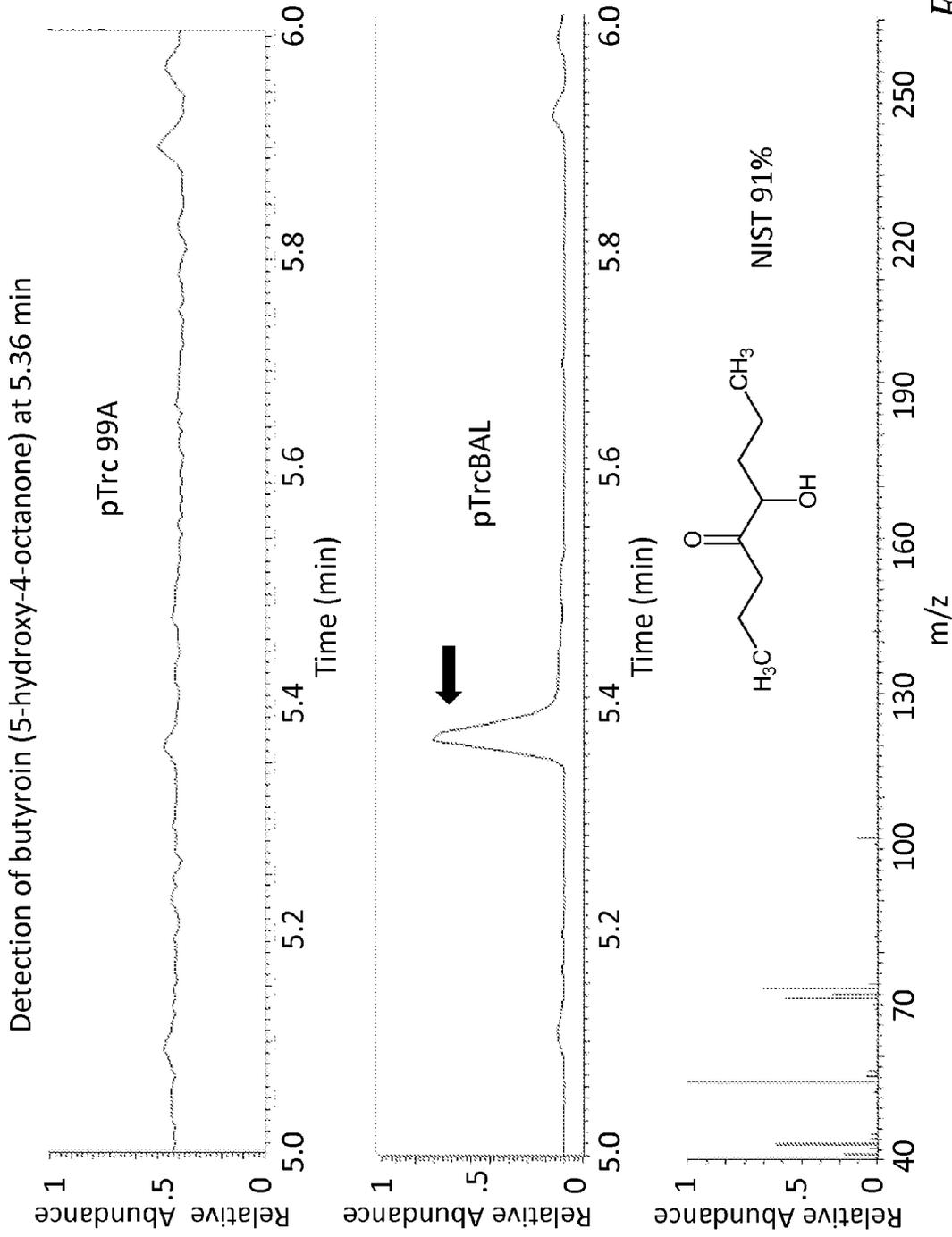


FIG. 17A

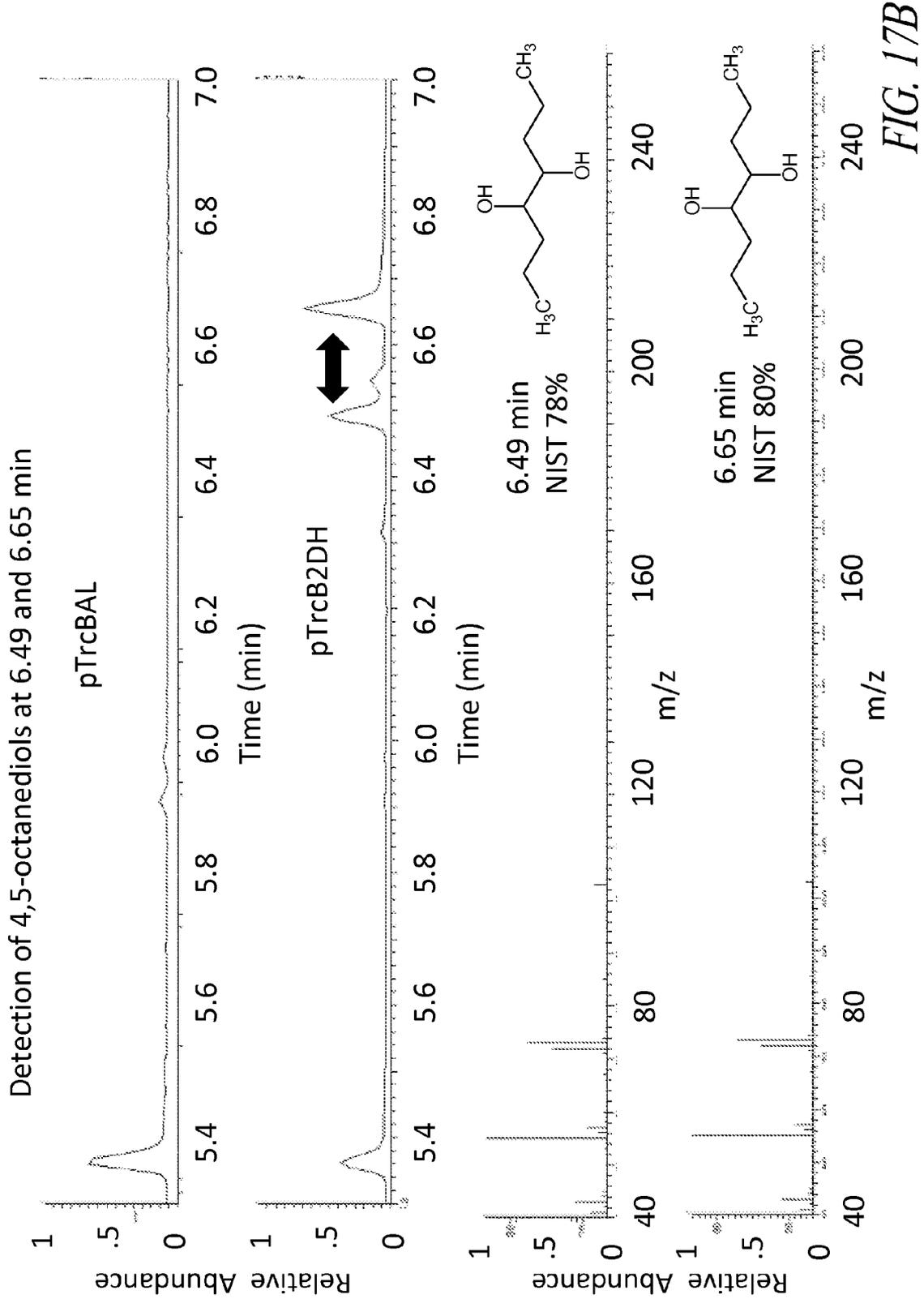


FIG. 17B

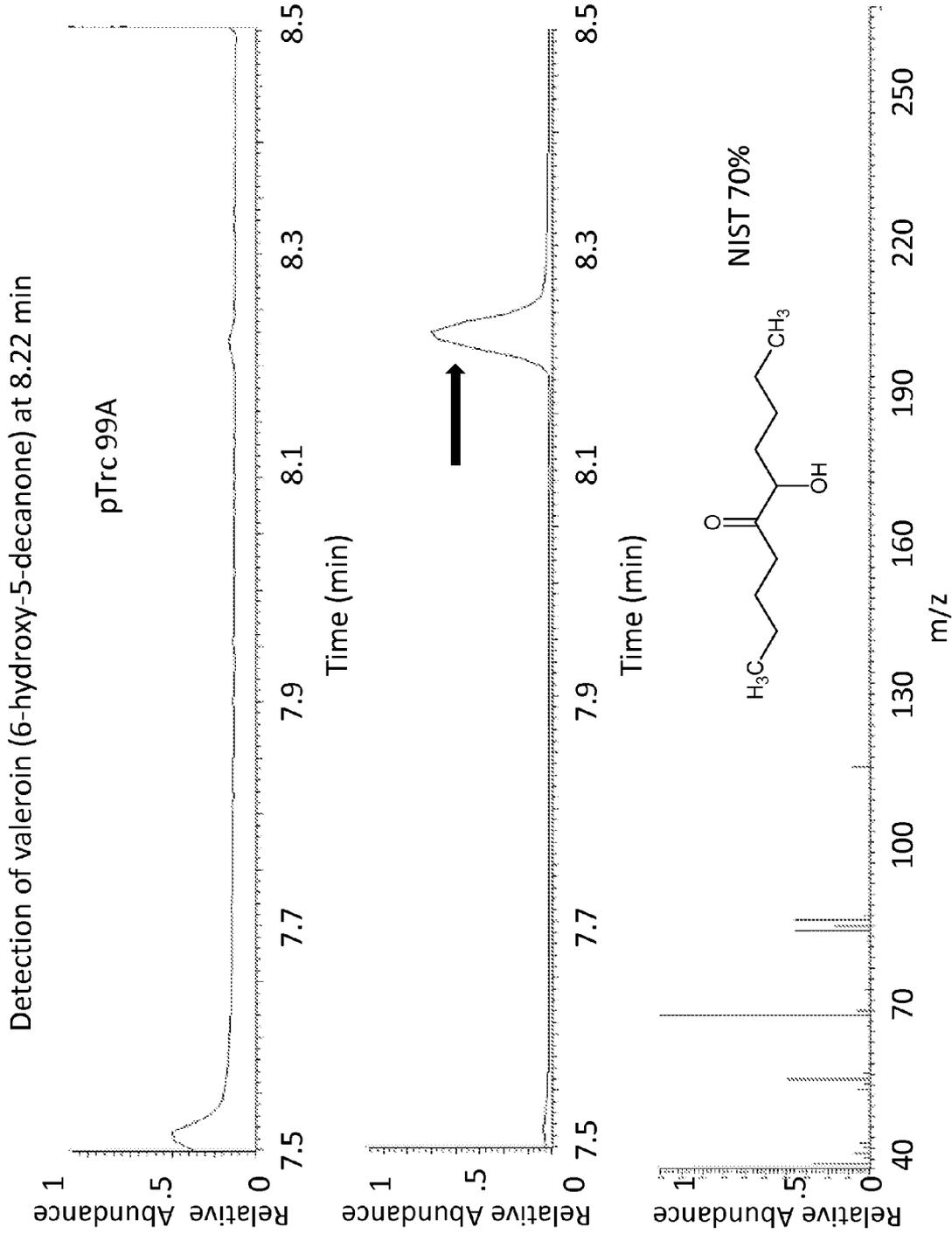


FIG. 18A

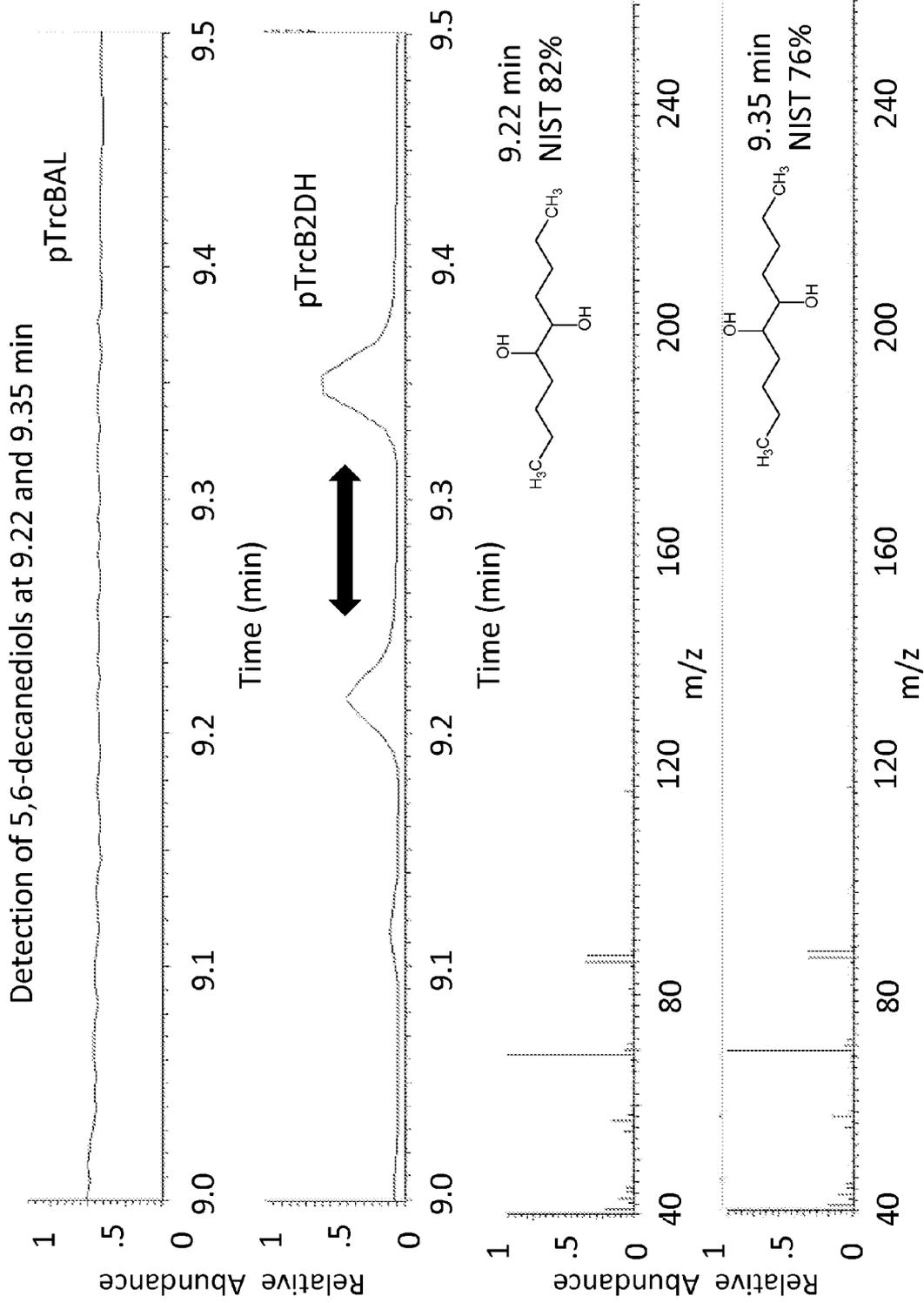


FIG. 18B

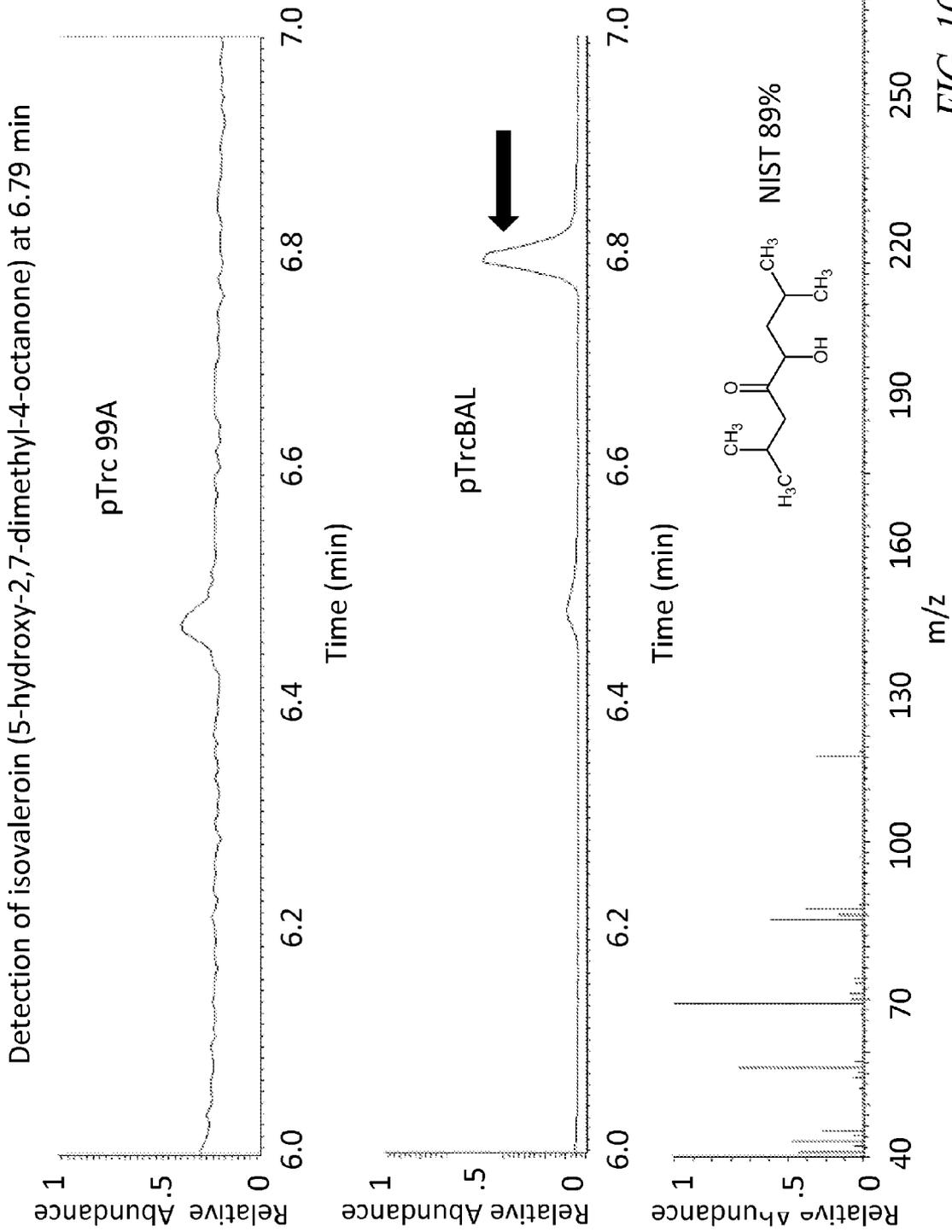


FIG. 19A

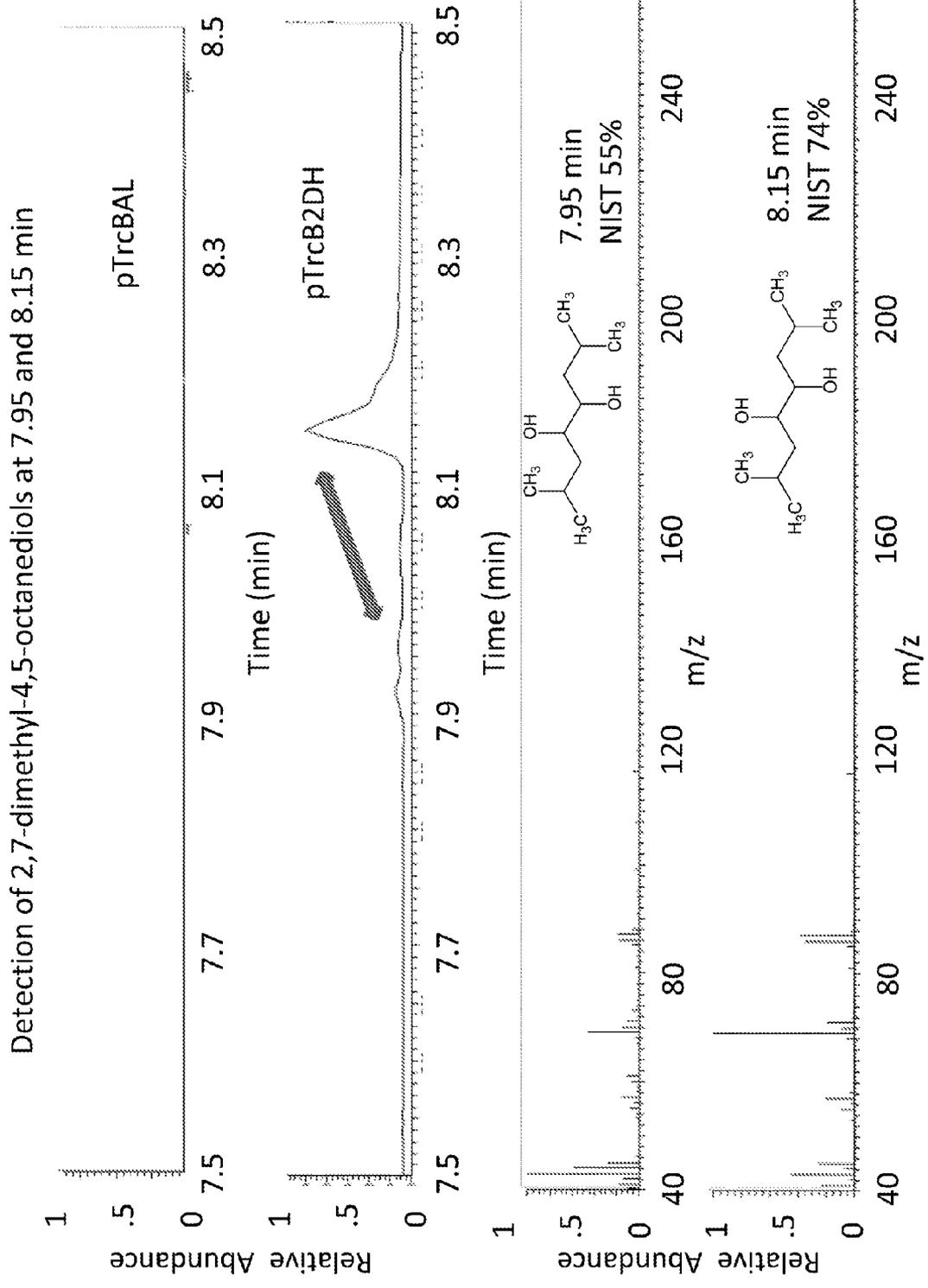


FIG. 19B

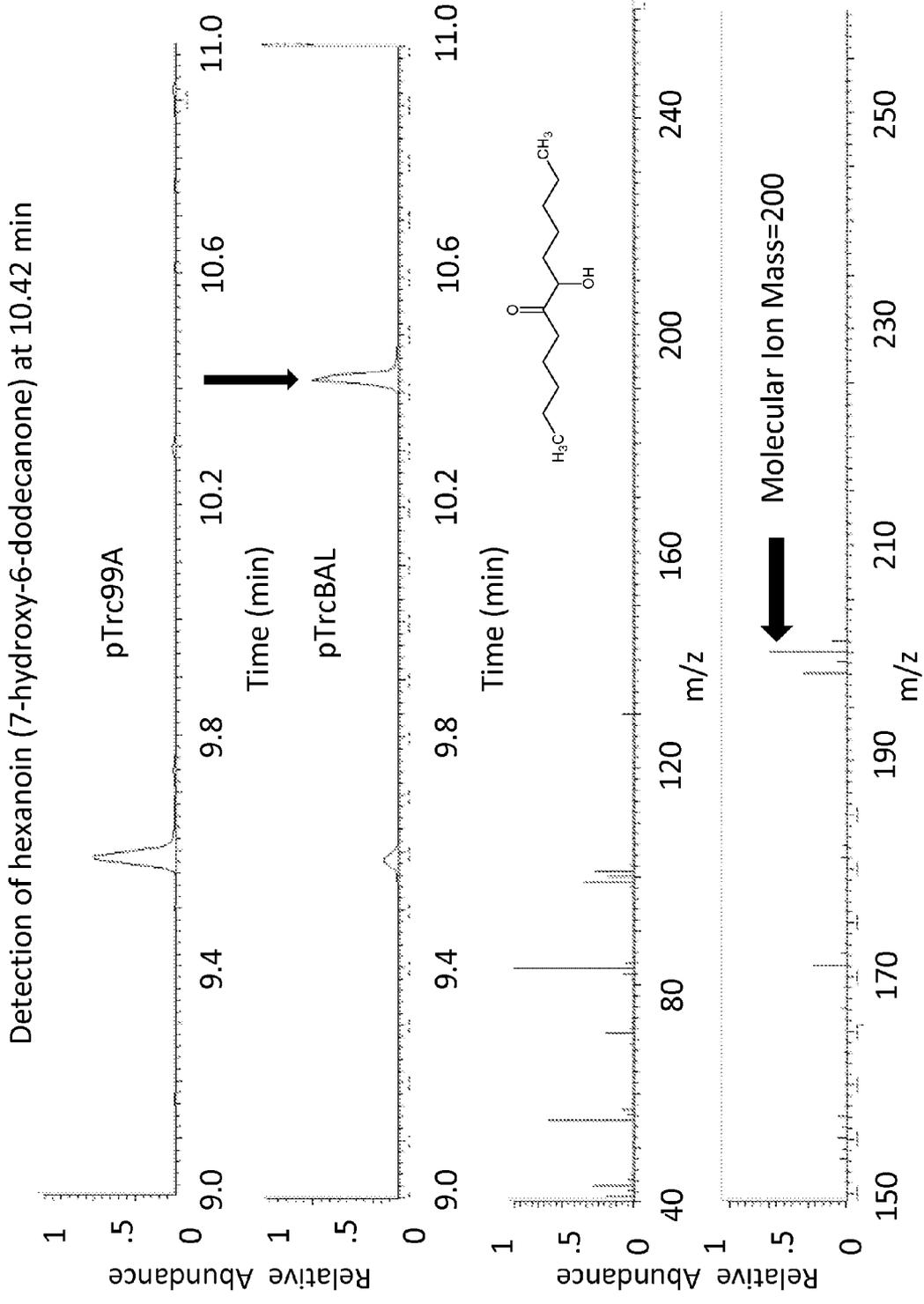


FIG. 20A

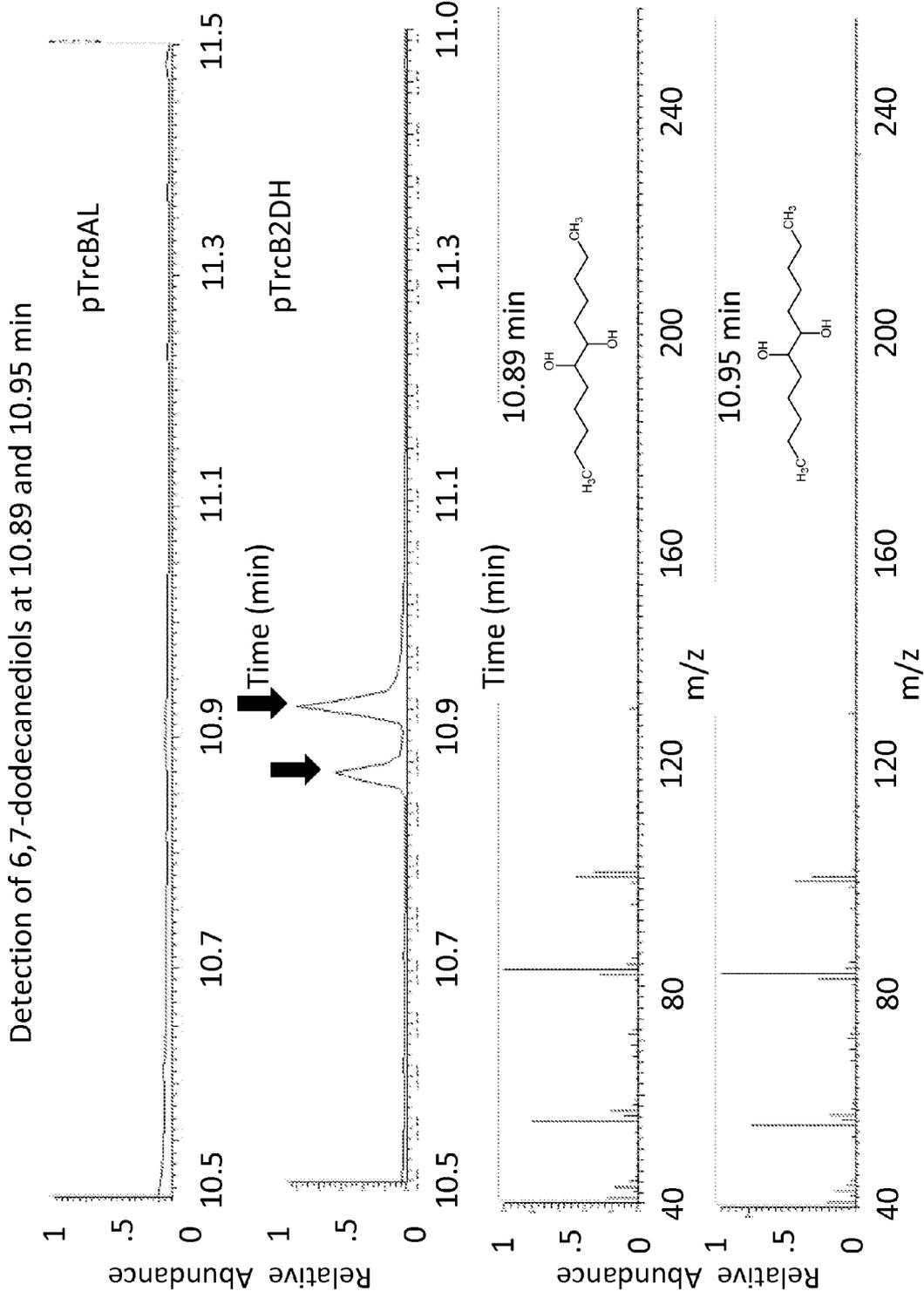


FIG. 20B

Figure 21A
 Detection of isohexanoin (2,9-Dimethyl-6-hydroxy-5-decanone) at 9.45 min

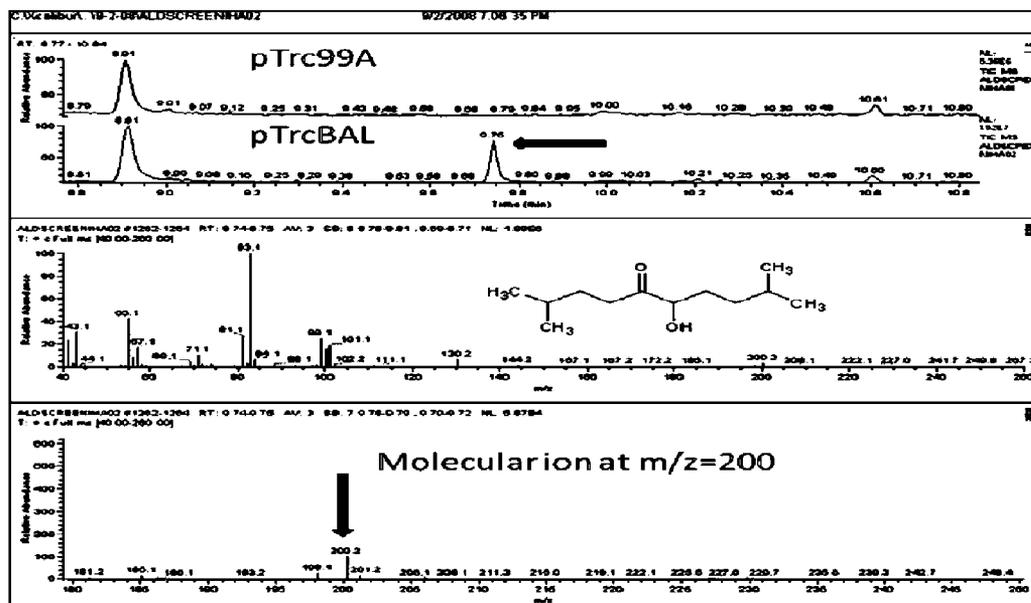


Figure 21B
 Detection of 2,9-dimethyl-5,6-decanediol at 10.38 and 10.44 min

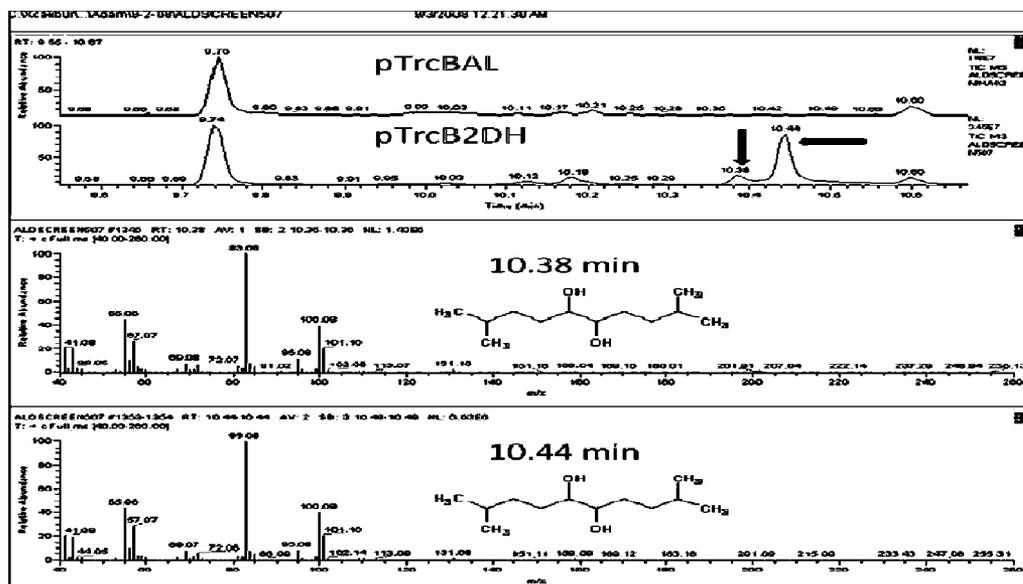


Figure 22
 Detection of octanoin (9-Hydroxy-8- hexadecanone) at 12.35 min

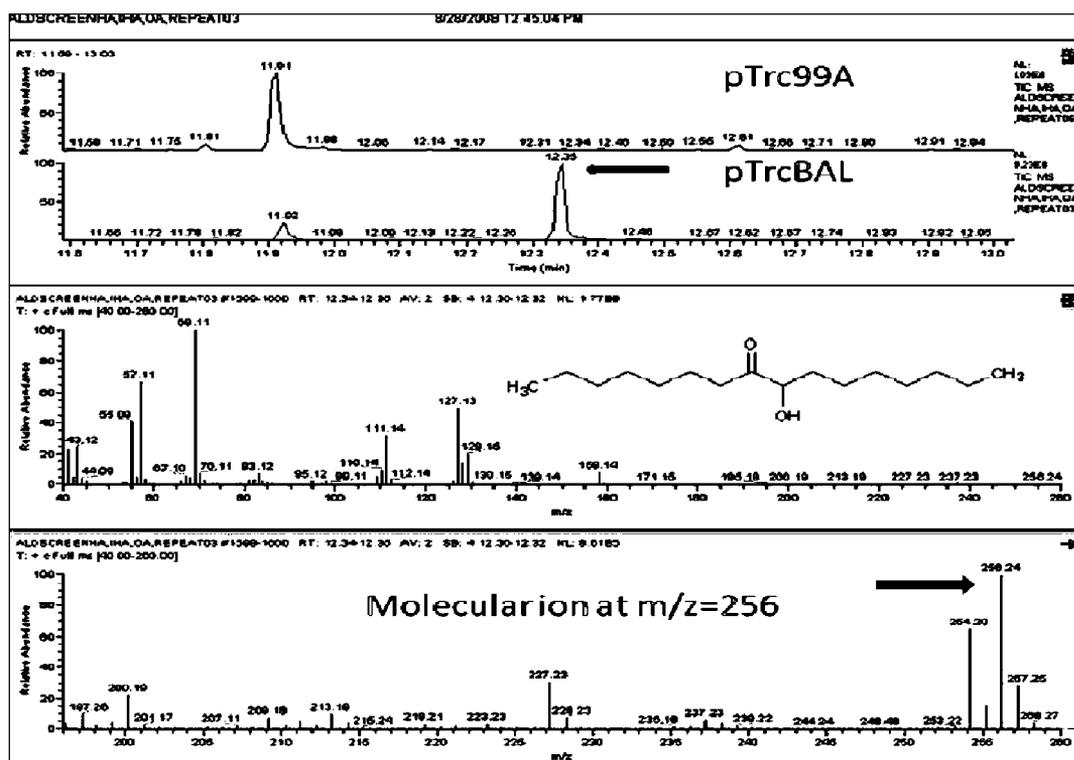
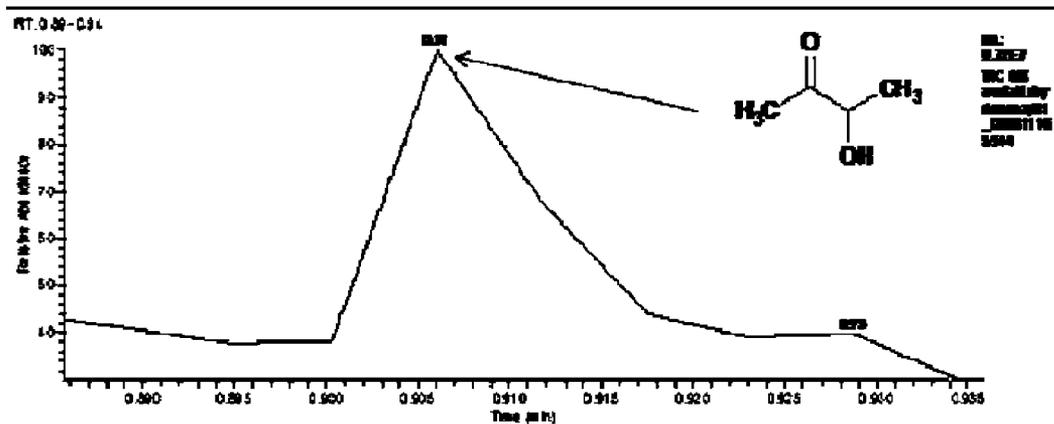


Figure 23

Detection of 3-hydroxy-2-butanone (acetoin) rt=0.91 min

20090709 08:04:29 01_0908111659.L

8-11-2008 4:30:44 PM



20090709 08:04:29 01_0908111659.L MS: 0.91 AB: 1 SE: 15.381-0.050 RL: 3.91E7

T: 0.910 (0.89-0.93)

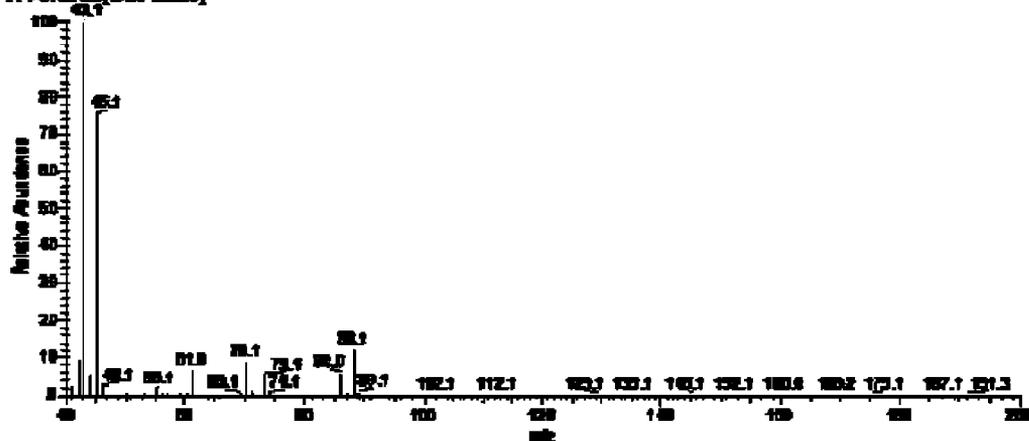


Figure 24A

Detection of 4-hydroxy-3-hexanone (propion) rt=2.62 min

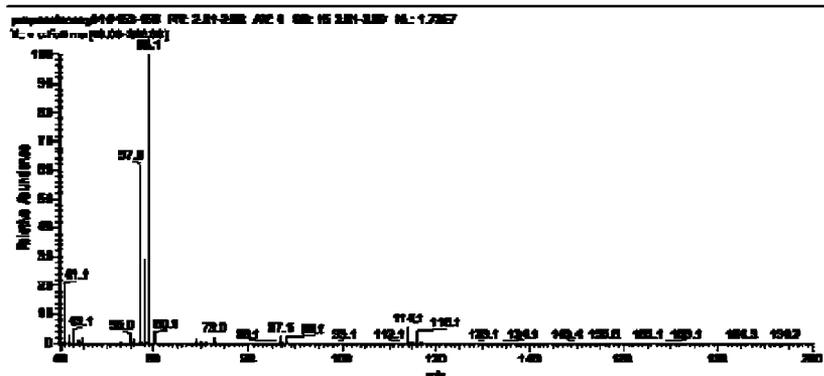
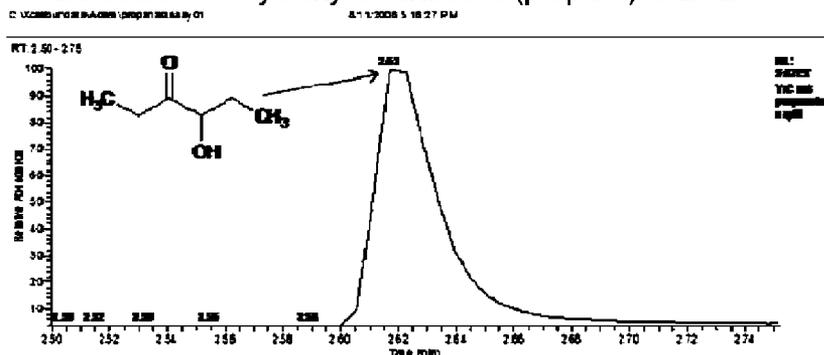


Figure 24B

Detection of 3,4-hexanediol rt=3.79 min

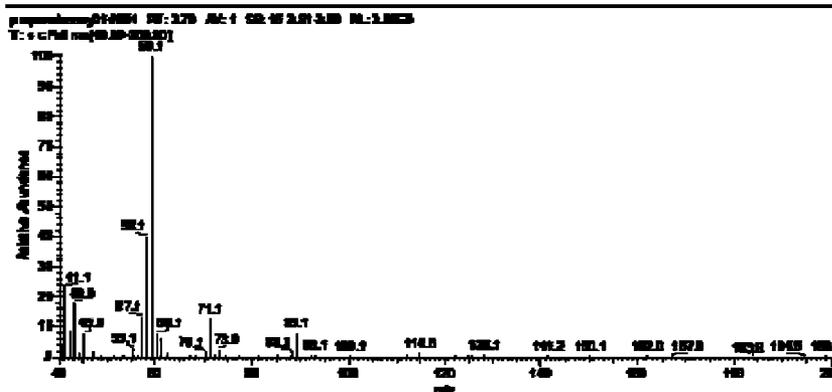
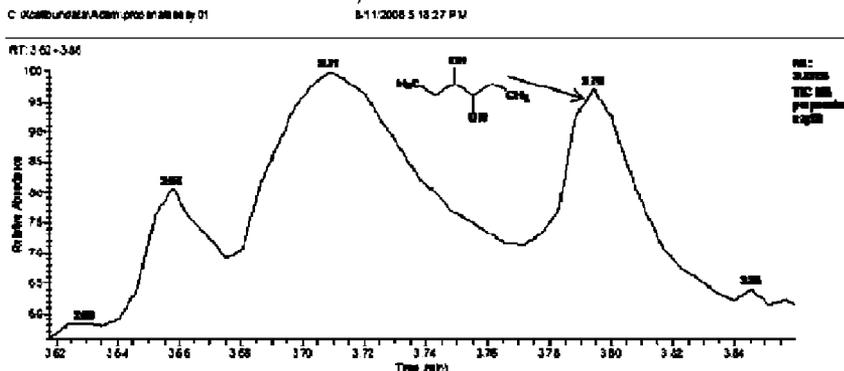
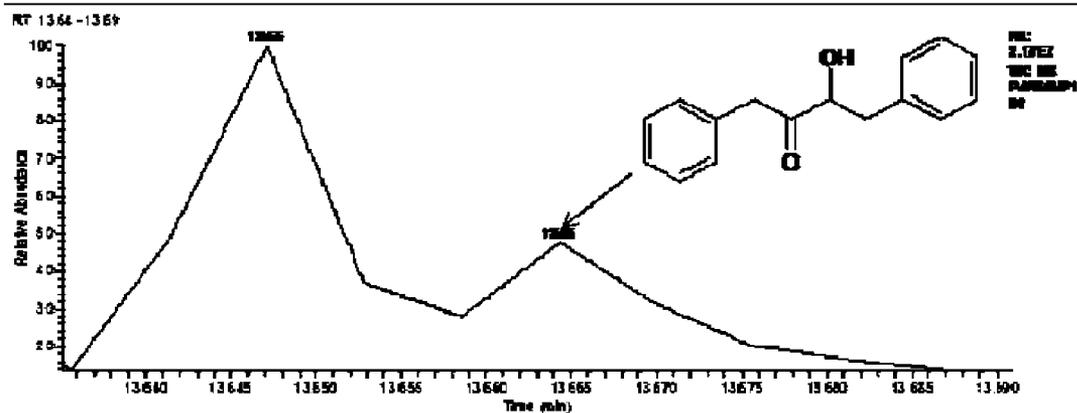


Figure 25

Detection of 1,4-diphenyl-3-hydroxy-2-butanone rt=13.66 min

C:\Xcalibur\data\Acem\FAASAMP101

8-11-2008 10:28:04 AM



FAASAMP101 080808-2208 RT: 13.66-13.67 AC: 2 SR: 15 2.07E2 N: 710E5
T: 0.01 min (0.00-0.01)

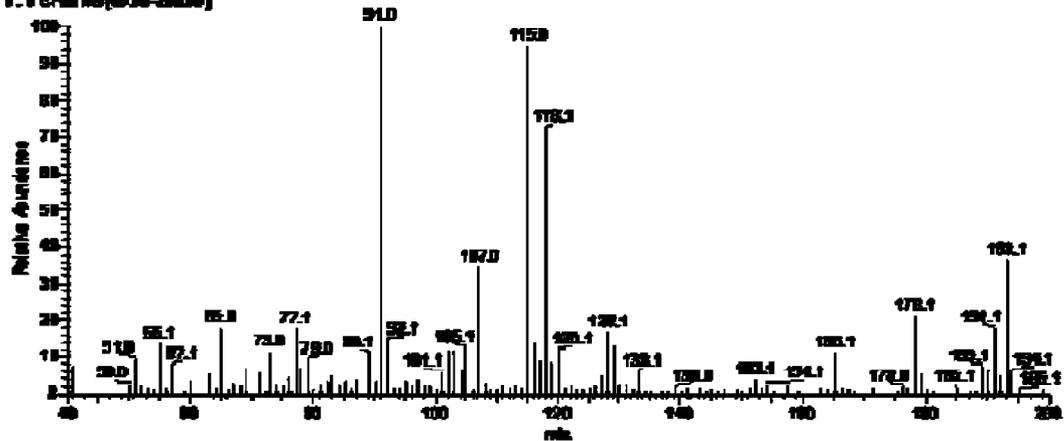


Figure 26A

GC-MS data confirming the presence of 4,5-octanediol in the sample extraction

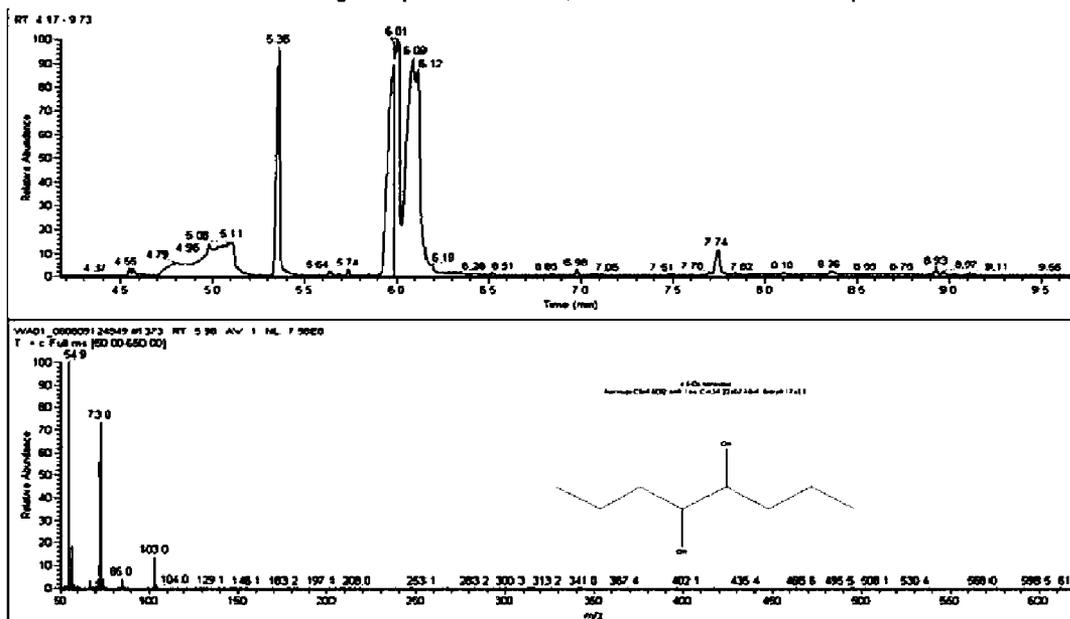
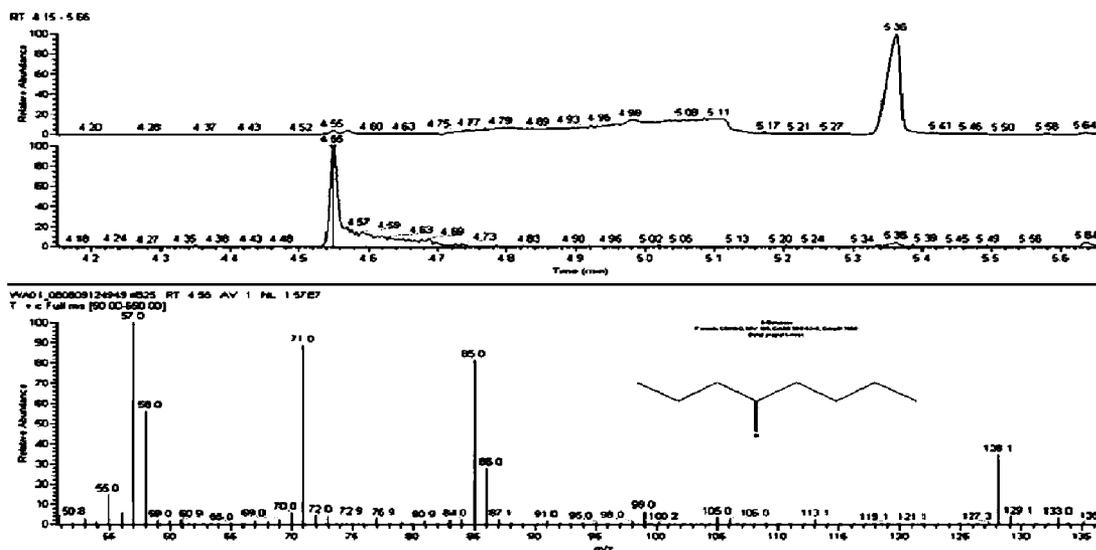


Figure 26B

GC-MS data confirming the presence of 4-octanone in the sample extraction

Sample Solution – Peak at 4.55 Identified as 4-octanone



Comparison between the sample extraction gas chromatograph (top) and the 4-octanone standard gas chromatograph (bottom)

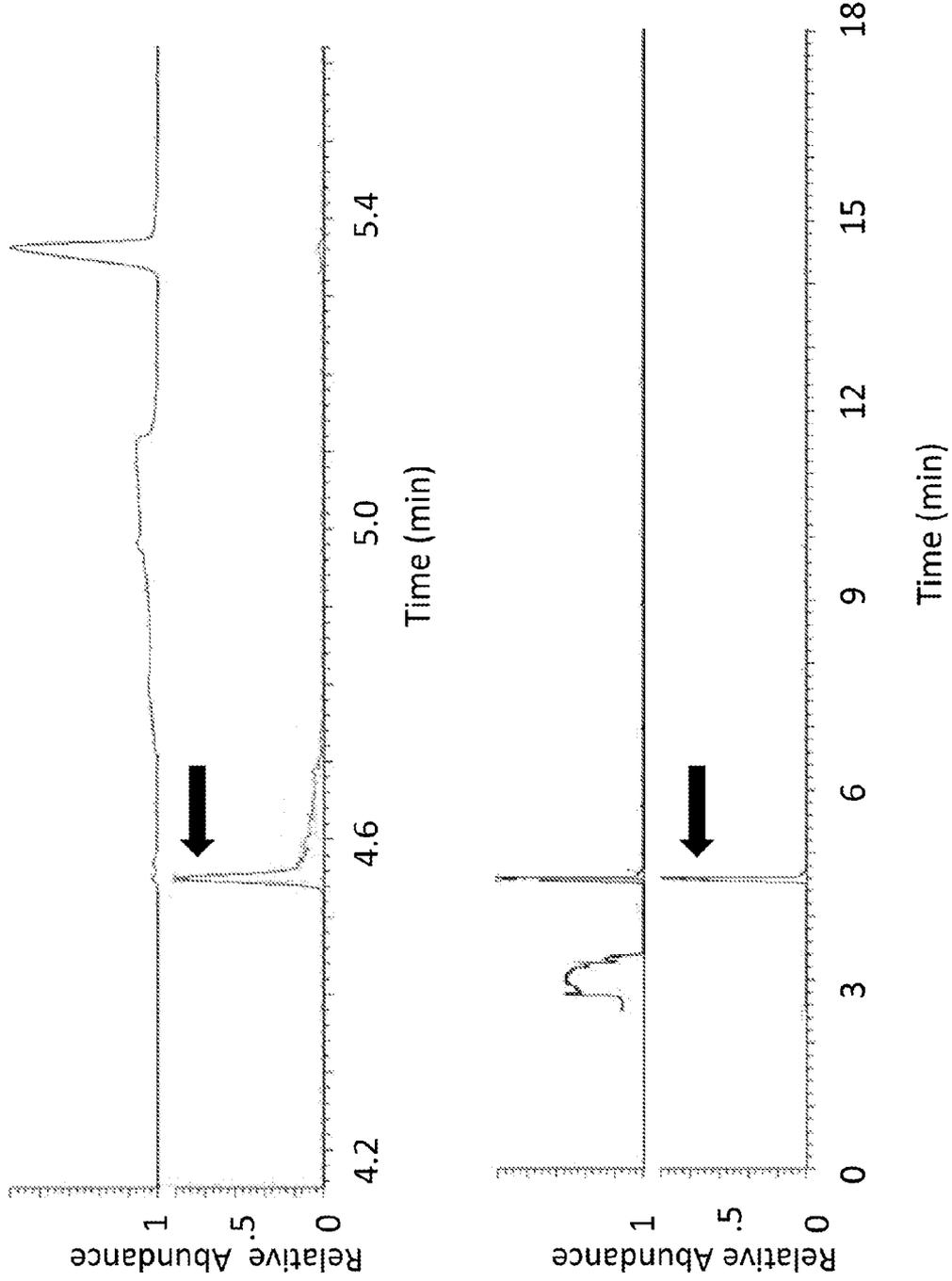
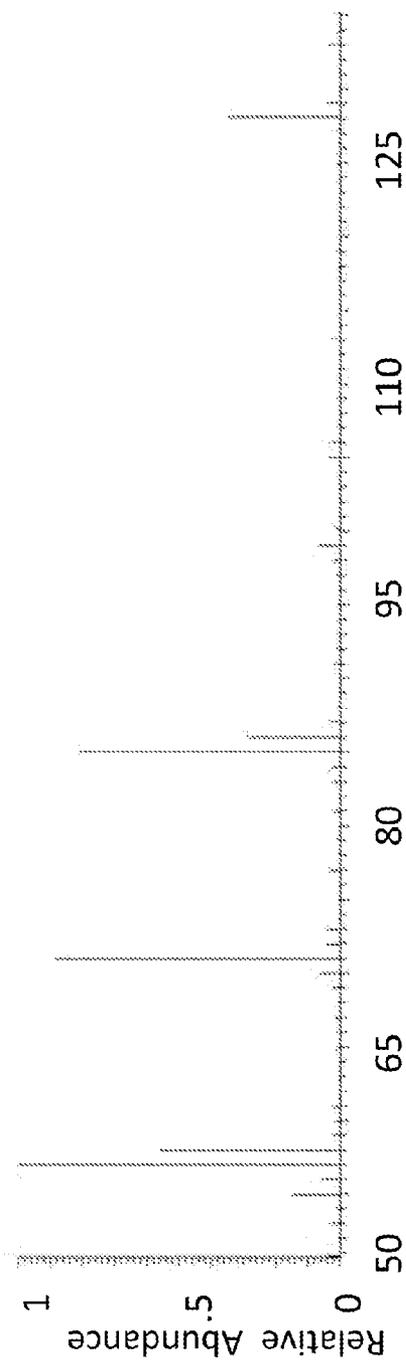
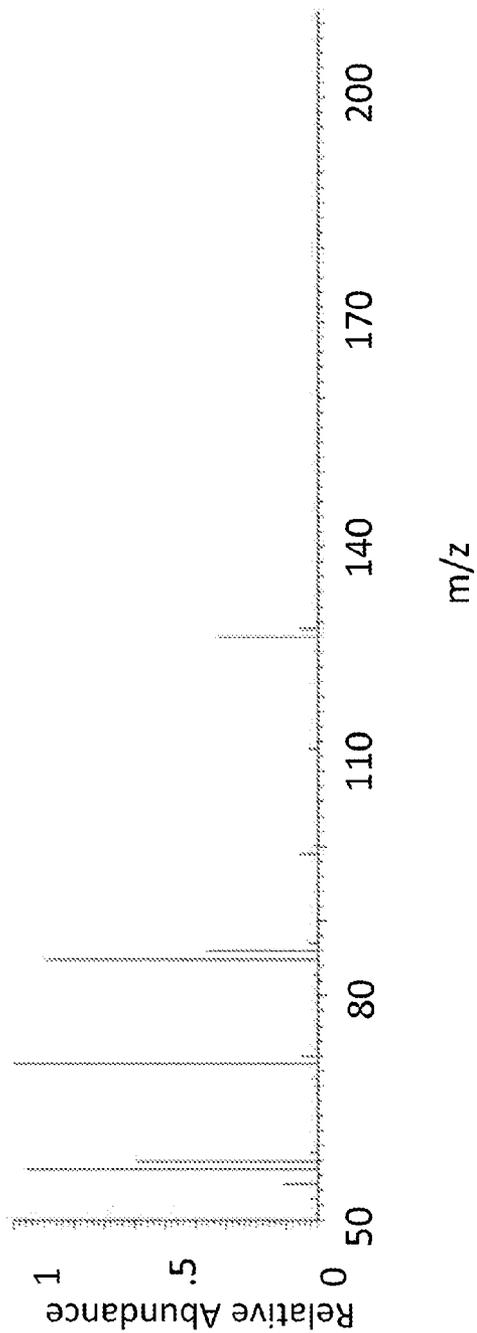


FIG. 27A

Comparison between the sample mass spectrum (top) and the 4-octanone standard mass spectrum (bottom)



m/z



m/z

FIG. 27B

Figure 28A

Nucleotide sequence of diol dehydratase large subunit (*pduC*) isolated from *Klebsiella pneumoniae* MGH78578

ATGAGATCGAAAAGATTTGAAGCACTGGCGAAACGCCCTGTGAATCAGGATGGTTTTCGTTAAGGA
GTGGATTGAAGAGGGCTTTATCGCGATGGAAAGCCCTAACGATCCCAAACCTTCTATCCGCATCG
TCAACGGCGCGGTGACCGAACTCGACGATAAACCGGTTGAGCAGTTCGACCTGATTGACCACTTT
ATCGCGCGCTACGGCATTAACTCTCGCCCGGGCCGAAGAAGTGATGGCCATGGATTTCGGTTAAGCT
CGCCAACATGCTCTGCGACCCGAACGTTAAACGCAGCGACATCGTGCCGCTCACTACCGCGATGA
CCCCGGCGAAAATCGTGGAAGTGGTGTGCGCATATGAACGTGGTCGAGATGATGATGGCGATGCAA
AAAATGCGCGCCCCGCCGACGCCGTCCCAGCAGGCGCATGTCACTAATATCAAAGATAATCCGGT
ACAGATTGCCGCCGACGCCGCTGAAGGCGCATGGCGCGGCTTTGACGAGCAGGAGACCACCGTCG
CCGTGGCGCGCTACGCGCCGTTCAACGCCATCGCCCTGCTGGTCGGTTTACAGGTTGGCCGCCCC
GGCGTCTCACCCAGTGTTCGCTGGAAGAAGCCACCGAGCTGAAACTGGGCATGCTGGGCCACAC
CTGCTATGCCGAAACCAATTTTCGGTATACGGTACGGAACCGGTGTTTACCGATGGCGATGACACCC
CGTGGTCGAAAAGGCTTCCCTCGCCTCCTCCTACGCCCTCGCGCGGCTGAAAATGCGCTTTACCTCC
GGTTCCGGCTCGGAGGTGCAGATGGGCTATGCCGAAGGCAAATCGATGCTTTATCTCGAAGCGCG
CTGCATCTACATCACCAAAGCCGCCGGGGTGAAGGCCTGCAGAAATGGCTCCGTCAGCTGTATCG
GCGTGCCGTCCGCCGTGCCGTCCGGGATCCGCGCCGCTACTGGCGGAAAACCTGATCTGCTCAGCG
CTGGATCTGGAGTGCGCCTCCAGCAACGATCAAACCTTTACCCACTCGGATATGCGGCGTACCGC
GCGTCTGCTGATGCAGTTCCTGCCAGGTACCGACTTTATCTCCTCCGGTTACTCGGCGGTGCCGA
ACTACGACAACATGTTTCGCCGGTTCCAACGAAGATGCCGAAGACTTCGATGACTACAACGTGATC
CAGCGCGACCTGAAGTTCGATGGCGGCTGCGGCCGGTTCGCTGAAGAGGACGTGATCGCCATTCG
CAACAAAGCCGCCCGCGCGCTGCAGGCGGTATTTGCCGGCATGGGTTTGCCGCCATTTACGGATG
AGAAGTAGAAGCCGCCACCTACGCCACGGTTCAAAGATATGCCCTGAGCGCAATATCGTCCGAG
GACATCAAGTTTGCTCAGGAGATCATCAACAAGAACCGCCAACCGCCTGGAGGTGGTGAAGCCCT
GGCGAAAGGCGGCTTCCCCGATGTGCCCCAGGACATGCTCAATAATTCAGAAAGCCAAGCTCACCG
GCGACTACCTGCATACCTCCGCCATCATTTGTTGGCGAGGGCCAGGTGCTCTCGGCCGTGAATGAC
GTGAACGATTATGCCGGTCCGGCAACAGGCTACCGCCTGCAAGGCGAGCGCTGGGAAGAGATTA
AAATATCCCGGGCGCGCTCGATCCCAATGAACTTGGCTAA (SEQ ID NO:103)

Figure 28B

Polypeptide sequence of diol dehydratase large subunit was isolated from *Klebsiella pneumoniae* MGH78578 (*pduC*)

MRSKRFEALAKRPVNQDGFVKEWIEEGFIAMESPNDPKPSIRIVNGAVTELDKPKVEQFDLIDHF
IARYGINLARAEVMMAMDSVKLANMLCDPNVKRSDIVPLTTAMTPAKIVEVVSMMNVVEMMMAMQ
KMRARRTPSQQAHVNTNIKDNVQIAADAAEGAWRGFDEQETTVAVARYAPFNIAIALLVGSQVGRP
GVLTQCSLEEATELKLGLMGLHTCYAETISVYGTPEPVFTDGDPTPWSKGFCLASSYASRGLKMRFTS
GSGSEVQMGYAEGKSMYLEARCIYITKAAGVQGLQNGSVSCI GVP S AVPSGIRAVLAENLCSA
LDLE CASSNDQTFTHSDMRRTARLLMQFLPGTDFISSGYS AVPNYDNMFAGSNEDAEDFDDY NVI
QRDLKVDGGLRPVREEDVIAIRNKAARALQAVFAGMGLPPI TDEEVEAATYAHGSKDMPERNIVE
DIKFAQEIINKNRNGLEVVKALAKGGFPDVAQDMLNIQKAKLTGDYLHTSAILIVGEGQVLSAVND
VNDYAGPATGYRLQGERWEEIKNIPGALDPNELG (SEQ ID NO:104)

Figure 29A

Nucleotide sequence of diol dehydratase medium subunit isolated from *Klebsiella pneumoniae* MGH78578 (*pduD*)

ATGGAAATTAACGAAACGCTGCTGCGCCAGATTATCGAAGAGGTGCTGTCCGAGATGAAATCAGG
CGCAGATAAGCCGGTCTCCTTTAGCGCGCTGCGGCTTCTGTGCGCTCTGCCGCGCCGGTCCGCC
TTGCGCCTGTGTCCGGCGACAGCTTCCTGACGGAATCGGGCAAGCCAAACCCGGCAGCAGCAG
GATGAAGTCATTATTGCCGTCGGGCCAGCGTTTGGTCTGGCGCAAACCGCCAATATCGTCGGCAT
TCCGCATAAAAAATATTCTGCGCGAAGTGATCGCCGGCATTGAGGAAGAAGGCATCAAAGCCCGGG
TGATCCGCTGCTTTAAGTCTTCTGACGTGCGCTTCGTGGCAGTGGAAGGCAACCGCCTGAGCGGC
TCCGGCATCTCGATCGGTATTCAGTCGAAAGGCACCACCGTCATCCACCAGCGCGGCCCTGCCGCC
GCTTTCCAATCTGGAAGTCTTCCCGCAGGCGCCGCTGCTGACGCTGGAAACCTACCGTCAGATTG
GCAAAAACGCCGCGCGCTACGCCAAACGCGAGTCGCCGCGAGCCGGTGCCGACGCTTAACGATCAG
ATGGCTCGTCCCAAATACCAGGCCAAGTCGGCCATTTTGCACATTAAGAGACCCAAATACGTGGT
GACGGGCAAAAACCCGCGAGGAAGTGCAGCGTGGCGCTTTAA (SEQ ID NO:105)

Figure 29B

Polypeptide sequence of diol dehydratase medium subunit isolated from *Klebsiella pneumoniae* MGH78578 (*pduD*)

MEINETLLRQIIEEVLSEMKSGADKPVSFSAAPAASVASAAPVAVAPVSGDSFLTEIGEAKPGTQQ
DEVI IAVGPAFGLAQTANIVGIPHKNILREVIAGIEEEGIKARVIRCFKSSDVAFFVAVEGNRLSG
SGISIGIQSKGTTVIHQRLPPLSNLELFPQAPLLTLETYRQIGKNAARYAKRESPQPVPTLNDQ
MARPKYQAKSAILHIKETKYVVTGKNPQELRVAL (SEQ ID NO:106)

Figure 29C

Nucleotide sequence of diol dehydratase small subunit isolated from *Klebsiella pneumoniae* MGH78578 (*pduE*)

ATGAATACCGACGCAATTGAATCCATGGTACGCGACGTGCTGAGCCGGATGAACAGCCTACAGGA
CGGGATAACGCCCGCGCCAGCCGCGCCGACAAACGACACCGTTCCGCCAGCCAAAAGTTAGCGACT
ACCCGTTAGCGACCCGCCATCCGGAGTGGGTCAAAACCGCTACCAATAAAACGCTCGATGACCTG
ACGCTGGAGAACGTATTAAGCGATCGCGTTACGGCGCAGGACATGCGCATCACTCCGGAACGCT
GCGTATGCAGGCGGCGATCGCCAGGATGCCGGACGCGATCGGCTGGCGATGAACTTTGAGCGGG
CCGCAGAGCTCACCGCGGTTCGCGACCGAATCCTTGAGATCTACAACGCCCTGCGCCATAC
CGTTCCACCCAGGCGGAGCTACTGGCGATCGCTGATGACCTCGAGCATCGCTACCAGGCACGACT
CTGTGCCGCTTTGTTCCGGAAGCGGCGGGCTGTACATCGAGCGTAAGAAGCTGAAAGGCGACG
ATTAA (SEQ ID NO:107)

Figure 29D

Polypeptide sequence of diol dehydratase small subunit isolated from *Klebsiella pneumoniae* MGH78578 (*pduE*)

MNTDAIESMVRDVL SRMNSLQDGI TPAPAAPTNDTVRQPKVSDYPLATRHPWVKTATNKTLDDL
TLENVLSDRVTAQDMRITPETLRMQAAIAQDAGRDLAMNFERAAELTAVPDDRI LEIYNALRPY
RSTQAE LLAIADDDLEHRYQARLCAAFVREAAGLYIERK KLGDD (SEQ ID NO:108)

Figure 31A

Oxidation of 4-octanol monitored by NADH production (2ADH11-18)

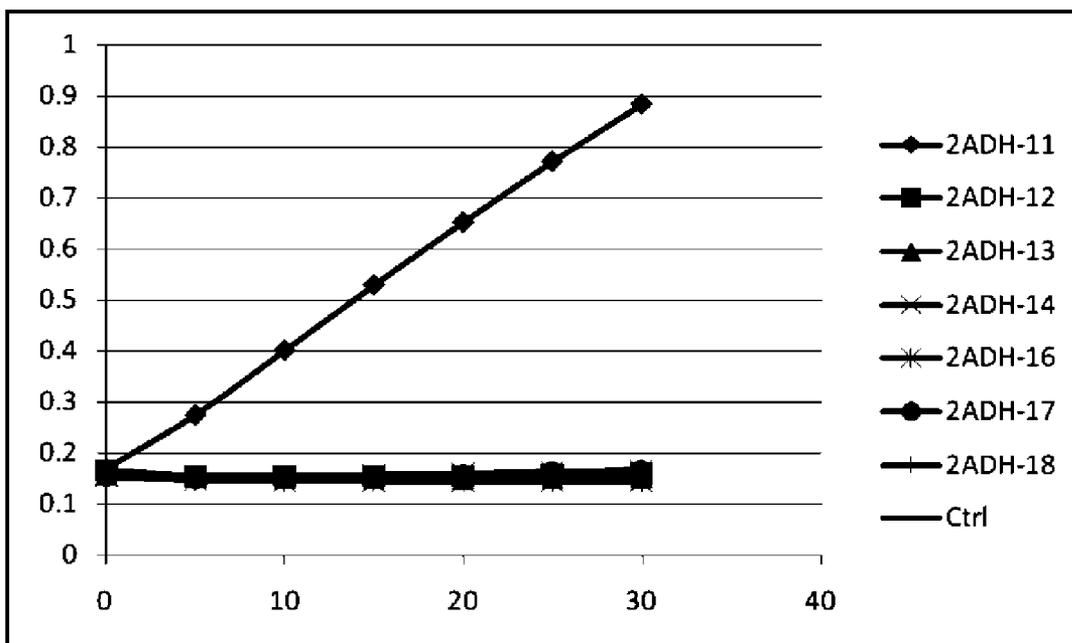


Figure 31B

Oxidation of 4-octanol monitored by NADPH production (2ADH 11-18)

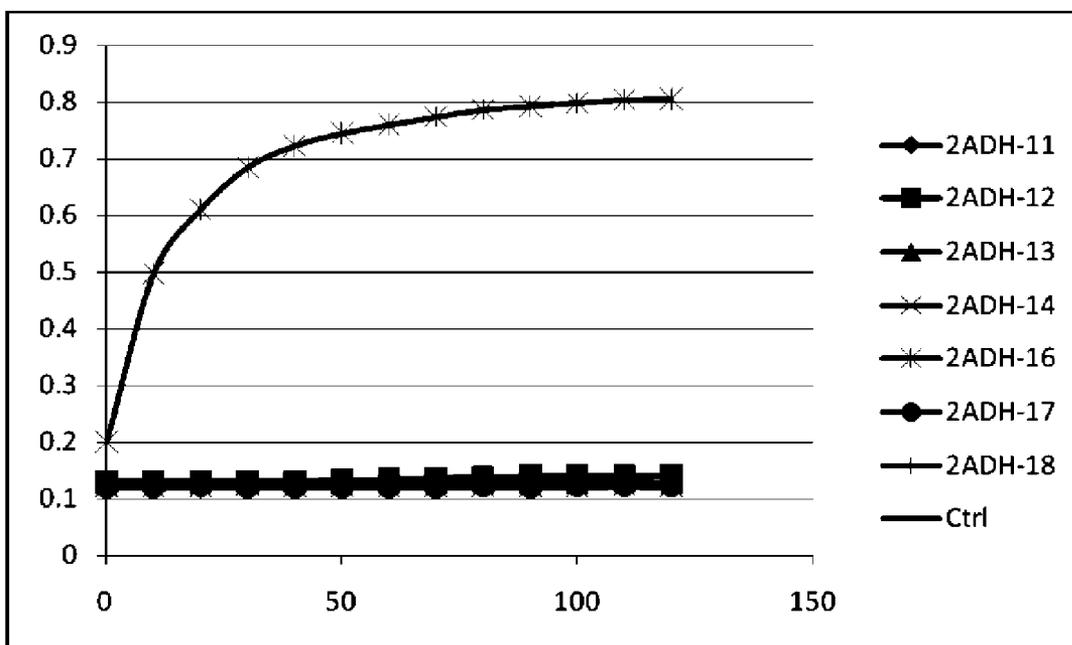


Figure 32A

Oxidation of 2,7-dimethyl octanol monitored by NADH production (2ADH 11-18)

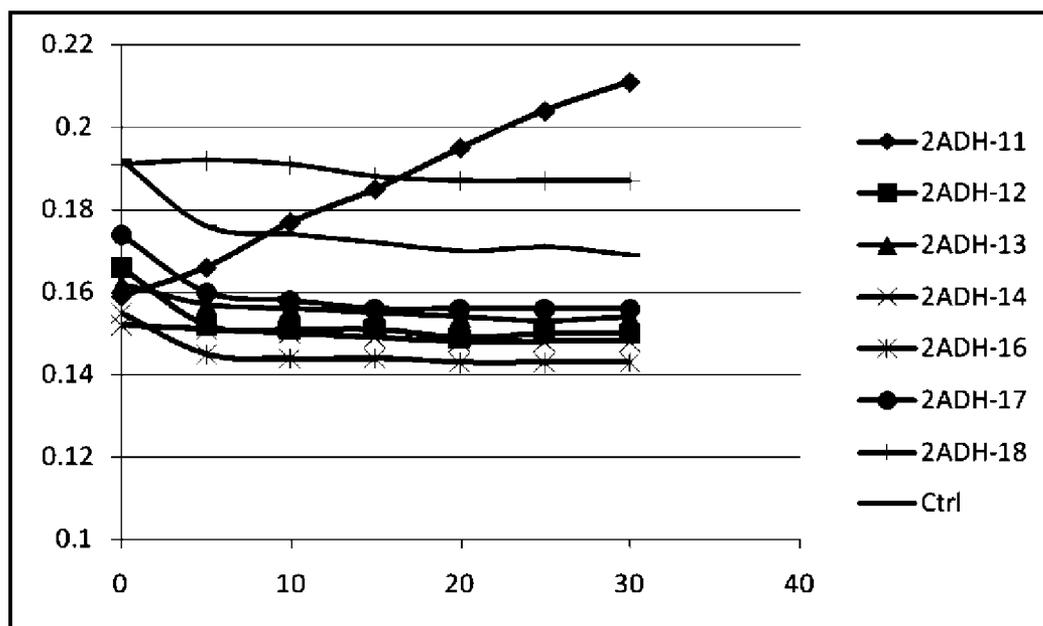


Figure 32B

Oxidation of 2,7-dimethyl octanol monitored by NADPH production (2ADH 11-18)

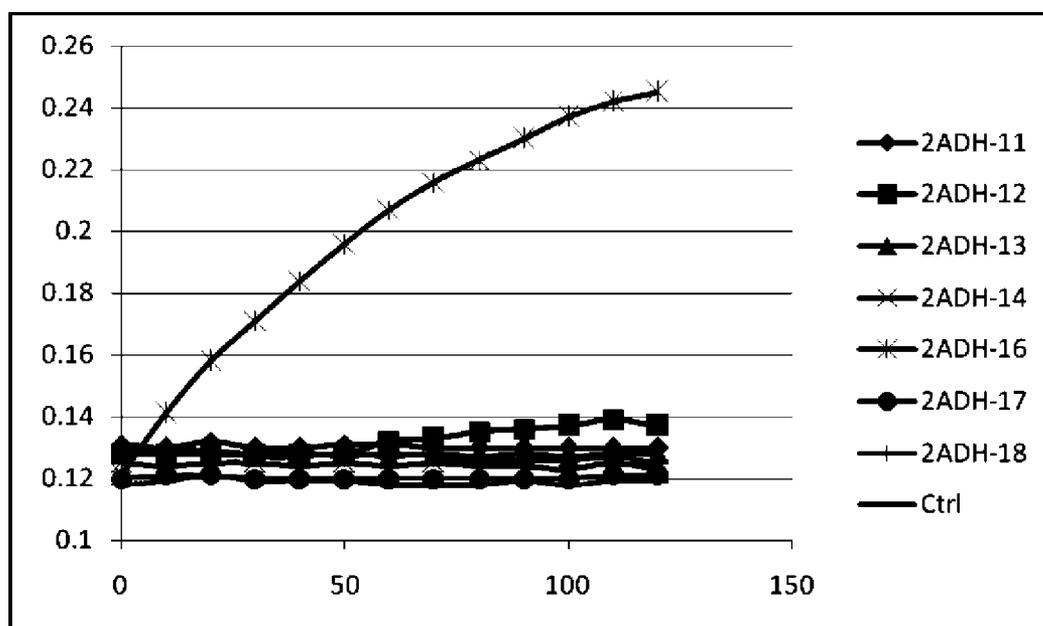


Figure 33A

Reduction of of 2,7-dimethyl octanol monitored by NADPH consumption

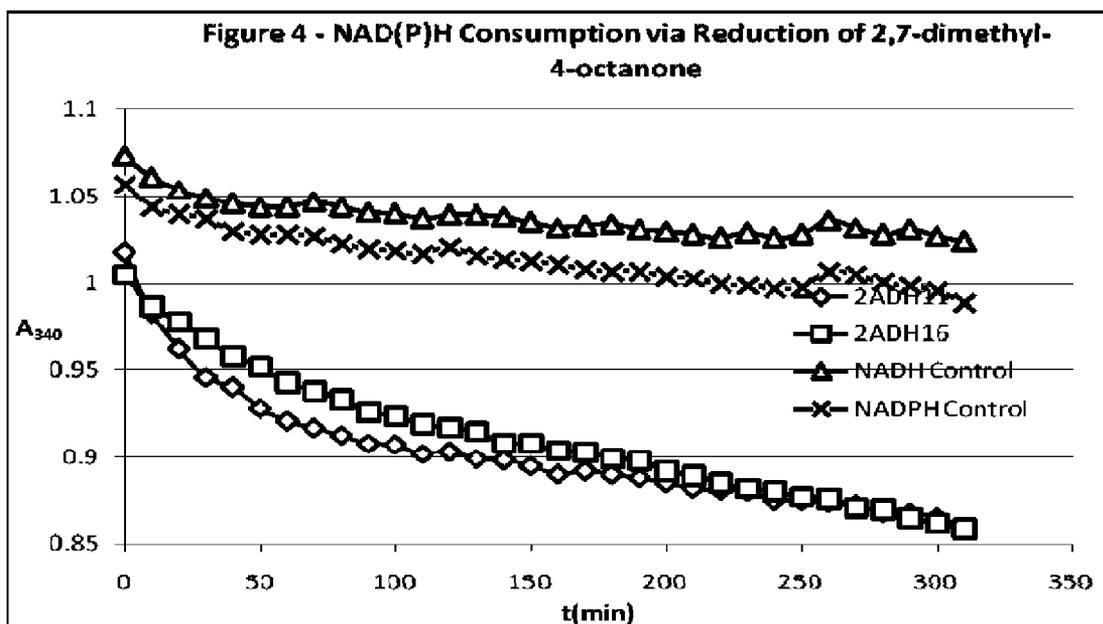


Figure 33B

Activity of 2ADH11 and 2ADH16 Towards Various Substrates

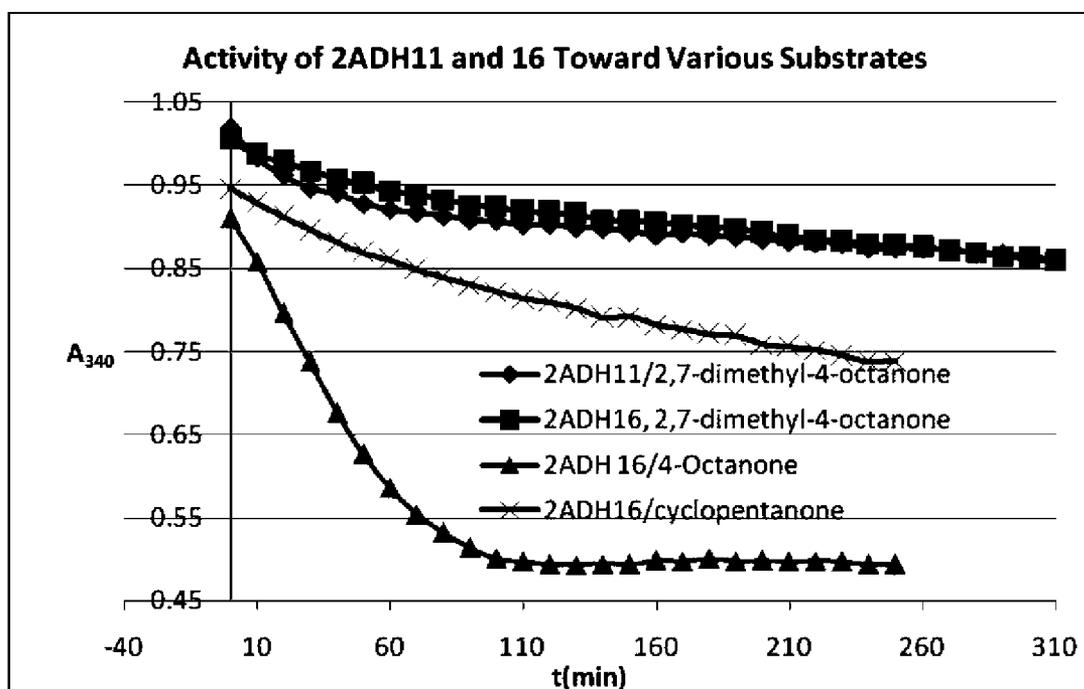


Figure 34A

Oxidation of cyclopentanol catalyzed by 2ADH as monitored by NADH or NADPH formation

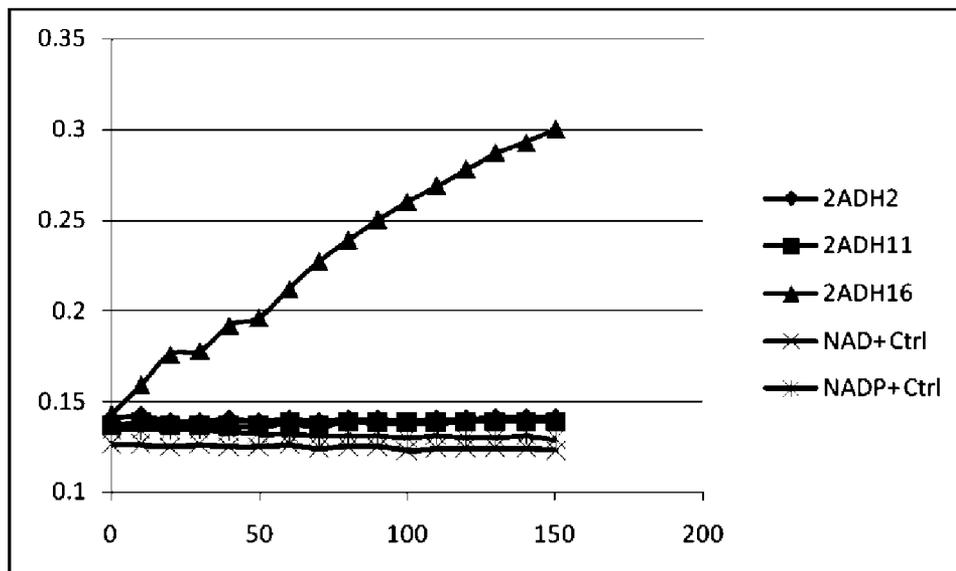


Figure 34B

Reduction of cyclopentanone catalyzed by 2ADH as monitored by NADPH consumption

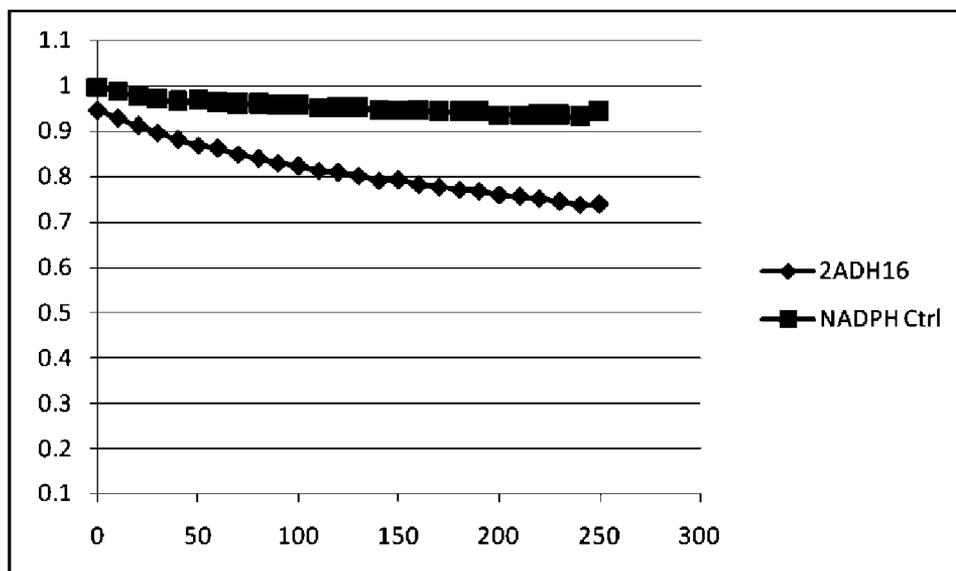
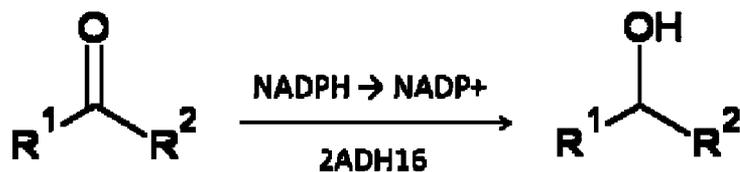
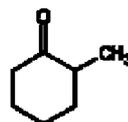


Figure 35
 Calculated Rate Constants for Reduction Reactions of 2ADH16

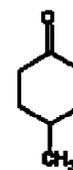


• Reduction Reaction

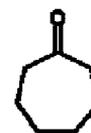
- 2-methylcyclohexanone: $k_{cat}/K_M = 211 \text{ sec}^{-1}\text{M}^{-1} \pm 18$



- 4-methylcyclohexanone: $k_{cat}/K_M = 17.5 \text{ sec}^{-1}\text{M}^{-1} \pm 2.0$



- cycloheptanone: $k_{cat}/K_M = 4.08 \text{ sec}^{-1}\text{M}^{-1} \pm 0.62$



- 4-octanone: $k_{cat}/K_M = 44.7 \text{ sec}^{-1}\text{M}^{-1} \pm 3.7$

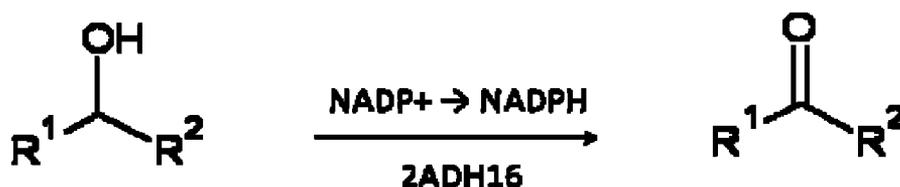


- 4-decanone: $k_{cat}/K_M \text{ approx } 150 \text{ sec}^{-1}\text{M}^{-1}$



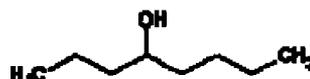
Figure 36

Calculated Rate Constants for Oxidation Reactions of 2ADH16

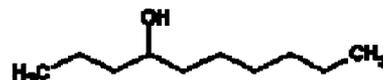


• Oxidation Reaction

- 4-octanol: $k_{cat}/K_M = 1430 \text{ sec}^{-1}\text{M}^{-1} \pm 200$



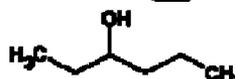
- 4-decanol: $k_{cat}/K_M = 1260 \text{ sec}^{-1}\text{M}^{-1} \pm 290$



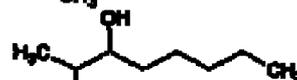
- cycloheptanol: $k_{cat}/K_M = 198 \text{ sec}^{-1}\text{M}^{-1} \pm 19$



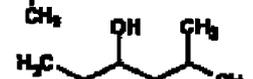
- 3-hexanol: $k_{cat}/K_M = 148 \text{ sec}^{-1}\text{M}^{-1} \pm 16$



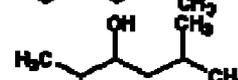
- 2-methyl-3-octanol: $k_{cat}/K_M = 123 \text{ sec}^{-1}\text{M}^{-1} \pm 16$



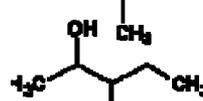
- 5-methyl-3-hexanol: $k_{cat}/K_M = 17.9 \text{ sec}^{-1}\text{M}^{-1} \pm 3.5$



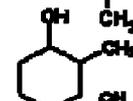
- 2,5-dimethyl-3-hexanol: $k_{cat}/K_M = 1.55 \text{ sec}^{-1}\text{M}^{-1} \pm .24$



- 3-methyl-2-pentanol: $k_{cat}/K_M = 8.54 \text{ sec}^{-1}\text{M}^{-1} \pm 2.04$



- 2-methylcyclohexanol: $k_{cat}/K_M = 97.4 \text{ sec}^{-1}\text{M}^{-1} \pm 14.4$



- 4-methylcyclohexanol: $k_{cat}/K_M = 10.4 \text{ sec}^{-1}\text{M}^{-1} \pm 21.1$

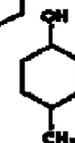


Figure 37A
Alginate Lyases

| Protein | Organism | GenBank/ GenPept | |
|--|---|------------------|--------------------|
| Family 5 | | | |
| alginate lyase (AlgL) | <u>Azotobacter chroococcum</u> <u>ATCC 4412</u> | <u>AJ223605</u> | <u>CAA11481.1</u> |
| | | <u>AF027499</u> | <u>AAC04567.1</u> |
| alginate lyase (AlgL) | <u>Azotobacter vinelandii</u> | <u>AF037600</u> | <u>AAC32313.1</u> |
| alginate lyase (Alg) | <u>Cobetia marina N-1</u> | <u>AB018795</u> | <u>BAA33966.1</u> |
| alginate lyase (AlgL) | <u>Pseudomonas aeruginosa</u> <u>8830</u> | <u>L14597</u> | <u>AAA71990.1</u> |
| alginate lyase (AlgL) | <u>Pseudomonas aeruginosa</u> <u>FRD1</u> | <u>U27829</u> | <u>AAA91127.1</u> |
| alginate lyase (AlgL;PA3547) | <u>Pseudomonas aeruginosa</u> <u>PAO1</u> | <u>AE004775</u> | <u>AAG06935.1</u> |
| | | <u>NC_002516</u> | <u>NP_252237.1</u> |
| alginate lyase (AlgL) | <u>Pseudomonas sp. QD03</u> | <u>AY380832</u> | <u>AAR23929.1</u> |
| alginate lyase (AlgL) | <u>Pseudomonas sp. QDA</u> | <u>AY163384</u> | <u>AAN63147.1</u> |
| alginate lyase (AlgL) | <u>Pseudomonas syringae pv.</u> <u>syringae FF5</u> | <u>AF222020</u> | <u>AAF32371.1</u> |
| alginate lyase (aly;A1-I/ PolyG+PolyM;A1-II/ PolyG;A1-III/PolyM) | <u>Sphingomonas sp. A1</u> | - | <u>2009330A</u> |
| | | <u>AB011415</u> | <u>BAB03312.1</u> |
| Family 6 | | | |
| alginate lyase (AlyP) | <u>Pseudomonas sp. OS-ALG-9</u> | <u>D10336</u> | <u>BAA01182.1</u> |
| Family 7 | | | |
| guluronate lyase (alyPG) | <u>Corynebacterium sp. ALY-1</u> | <u>AB030481</u> | <u>BAA83339.1</u> |
| poly(-L-guluronate) lyase (AlyA) | <u>Klebsiella pneumoniae subsp.</u> <u>aerogenes</u> | <u>L19657</u> | <u>AAA25049.1</u> |
| alginate lyase / poly-mannuronate lyase (AlxM) | <u>Photobacterium sp. ATCC</u> <u>43367</u> | <u>X70036</u> | <u>CAA49630.1</u> |
| alginate lyase (PA1167) | <u>Pseudomonas aeruginosa</u> <u>PAO1</u> | <u>AE004547</u> | <u>AAG04556.1</u> |
| | | <u>NC_002516</u> | <u>NP_249858.1</u> |
| alginate lyase (A1-II') | <u>Sphingomonas sp. A1</u> | <u>AB120939</u> | <u>BAD16656.1</u> |
| alginate lyase (aly;A1-I/ PolyG+PolyM;A1-II/ PolyG;A1-III/PolyM) | <u>Sphingomonas sp. A1</u> | - | <u>2009330A</u> |
| | | <u>AB011415</u> | <u>BAB03312.1</u> |

Figure 37B
Alginate Lyases

| Protein | Organism | GenBank | GenPept |
|--|--|------------------|--------------------|
| Family 7 | | | |
| poly(a-L-guluronate) lyase (AlyVG1;AlyVG1) | <u>Vibrio halioticoli IAM14596T</u> | <u>AF114039</u> | <u>AAF22512.1</u> |
| alginate lyase / poly-mannuronate lyase (AlyVOA) | <u>Vibrio sp. O2</u> | <u>DQ235160</u> | <u>ABB36771.1</u> |
| alginate lyase / poly-mannuronate lyase (AlyVOB) | <u>Vibrio sp. O2</u> | <u>DQ235161</u> | <u>ABB36772.1</u> |
| alginate lyase (AlyVI) | <u>Vibrio sp. QY101</u> | <u>AY221030</u> | <u>AAP45155.1</u> |
| exo-oligoalginate lyase (HdAlex;HdAlex-1) | <u>Haliotis discus hannai</u> | <u>AB234872</u> | <u>BAE81787.1</u> |
| alginate lyase (HdAly) | <u>Haliotis discus hannai</u> | <u>AB110094</u> | <u>BAC87758.1</u> |
| polysaccharide lyase acting on glucuronic acid (vAL-1) | <u>Chlorella virus CVK2</u> | <u>AB044791</u> | <u>BAB19127.1</u> |
| alginate lyase (AlyII) | <u>Pseudomonas sp. OS-ALG-9</u> | <u>AB003330</u> | <u>BAA19848.1</u> |
| Family18 | | | |
| alginate lyase | <u>Pseudoalteromonas sp. 272</u> | | |
| alginate lyase (Aly) | <u>Pseudoalteromonas sp. IAM14594</u> | <u>AF082561</u> | <u>AAD16034.1</u> |
| Family15 | | | |
| exotype alginate lyase (Atu3025) | <u>Agrobacterium tumefaciens str. C58</u> | <u>AE009232</u> | <u>AAL43841.1</u> |
| | | <u>NC_003305</u> | <u>NP_533525.1</u> |
| exotype alginate lyase (AGR_L_3558p) | <u>Agrobacterium tumefaciens str. C58 (Cereon)</u> | <u>AE008381</u> | <u>AAK90358.1</u> |
| | | <u>NC_003063</u> | <u>NP_357573.1</u> |
| oligo alginate lyase (A1-IV) | <u>Sphingomonas sp. A1</u> | <u>AB011415</u> | <u>BAB03319.1</u> |
| alginate lyase (A1-IV') | <u>Sphingomonas sp. A1</u> | <u>AB176667</u> | <u>BAD90006.1</u> |

Figure 38A
Pectate Lyases

| Protein | Organism | GenBank/GenPept | |
|-----------------------------|---|------------------|--------------------|
| Family1 | | | |
| pectate lyase | <u>Bacillus agaradhaerens</u> | - | <u>AAE59745.1</u> |
| | | - | <u>AAS29292.1</u> |
| pectate lyase | <u>Bacillus halodurans</u> | - | <u>AAE59748.1</u> |
| | | - | <u>AAS29295.1</u> |
| pectate lyase (BH3819) | <u>Bacillus halodurans C-125</u> | <u>AP001520</u> | <u>BAB07538.1</u> |
| | | <u>NC 002570</u> | <u>NP 244686.1</u> |
| pectate lyase (BH0698) | <u>Bacillus halodurans C-125</u> | <u>AP001509</u> | <u>BAB04417.1</u> |
| | | <u>NC 002570</u> | <u>NP 241564.1</u> |
| pectate lyase (PelA) | <u>Bacillus licheniformis 14A</u> | - | <u>AAE59746.1</u> |
| | | - | <u>AAN26179.1</u> |
| | | <u>AJ517194</u> | <u>CAD56882.1</u> |
| | | - | <u>AAS29293.1</u> |
| BLi04129 or BL00947 (PelII) | <u>Bacillus licheniformis DSM 13 ATCC 14580</u> | <u>CP000002</u> | <u>AAU25568.1</u> |
| | | <u>AE017333</u> | <u>AAU42942.1</u> |
| pectate lyase (Pel) | <u>Bacillus licheniformis RN1</u> | <u>AB428424</u> | <u>BAG12908.1</u> |
| pectate lyase (PelB) | <u>Bacillus pumilus DKS1</u> | <u>EU652988</u> | <u>ACD11362.1</u> |
| pectate lyase (fragment) | <u>Bacillus sp. KSM-P7</u> | <u>AB015043</u> | <u>BAA76884.1</u> |
| pectate lyase | <u>Bacillus sp. AAI12</u> | - | <u>AAE59747.1</u> |
| | | - | <u>AAS29294.1</u> |
| pectate lyase | <u>Bacillus sp. I534</u> | - | <u>AAE59749.1</u> |
| | | - | <u>AAR65348.1</u> |
| | | - | <u>AAS29296.1</u> |
| pectate lyase (Pel-103) | <u>Bacillus sp. KSM-P103</u> | <u>AB015044</u> | <u>BAA76885.1</u> |
| pectate lyase (Pel-34K) | <u>Bacillus sp. P-2850</u> | <u>AB080666</u> | <u>BAC11008.1</u> |
| pectate lyase (Pel-4A) | <u>Bacillus sp. P-4-N</u> | <u>AB041769</u> | <u>BAA96477.1</u> |
| pectate lyase (Pel-4B) | <u>Bacillus sp. P-4-N</u> | <u>AB042100</u> | <u>BAA96478.1</u> |
| pectate lyase (PI47) | <u>Bacillus sp. TS-47</u> | <u>AB045986</u> | <u>BAB40336.1</u> |
| pectate lyase (PelK) | <u>Bacillus sp. YA-14</u> | <u>D26349</u> | <u>BAA05383.1</u> |
| pectate lyase | <u>Bacillus subtilis</u> | <u>AX601431</u> | <u>CAD67509.1</u> |
| | | <u>AX601436</u> | <u>CAD67510.1</u> |
| | | <u>AX601448</u> | <u>CAD67511.1</u> |
| | | <u>AX951870</u> | <u>CAF05441.1</u> |
| pectate lyase (Pel) | <u>Bacillus subtilis AC327</u> | <u>D86417</u> | <u>BAA22313.1</u> |

Figure 38B
Pectate Lyases

| Protein | Organism | GenBank | GenPept |
|--|---|------------------|--------------------|
| pectin lyase (Ppr) | <u>Bacillus subtilis IFO 3134</u> | <u>D83791</u> | <u>BAA12119.1</u> |
| pectate lyase (Pel) | <u>Bacillus subtilis SO113</u> | <u>X74880</u> | <u>CAA52866.1</u> |
| | | - | <u>AAR45489.1</u> |
| | | <u>D86417</u> | <u>BAA22313.1</u> |
| | | <u>X74880</u> | <u>CAA52866.1</u> |
| pectate lyase (Pel;BSU07560) | <u>Bacillus subtilis subsp. subtilis str. 168</u> | <u>Z99108</u> | <u>CAB12585.1</u> |
| | | <u>NC 000964</u> | <u>NP 388637.1</u> |
| pectate lyase (Pel-1;Pel1) | <u>Erwinia carotovora 71</u> | <u>L32171</u> | <u>AAA73933.1</u> |
| Pel9.5 (fragment) | <u>Erwinia carotovora EC14</u> | <u>X61088</u> | <u>CAA43402.1</u> |
| pectin lyase (Pnl) (probable fragment) | <u>Erwinia carotovora ER</u> | <u>M65057</u> | <u>AAA24857.1</u> |
| | | <u>M18859</u> | <u>AAA24845.1</u> |
| | | <u>S51490</u> | <u>AAC60423.1</u> |
| | <u>Erwinia carotovora ER / IAM1068 / atroseptica EC / atroseptica C18</u> | <u>D00217</u> | <u>BAA00155.1</u> |
| pectate lyase 1 (Pel1;PelI) | | <u>X81847</u> | <u>CAA57439.1</u> |
| | <u>Erwinia carotovora ER / IAM1068 / atroseptica EC / atroseptica C18</u> | <u>M17364</u> | <u>AAA24848.1</u> |
| | | <u>S51475</u> | <u>AAC60422.1</u> |
| pectate lyase 2 (Pel2;PelII) | | <u>X81847</u> | <u>CAA57440.1</u> |
| ECA4067 (PelA) | <u>Erwinia carotovora subsp. atroseptica SCRI1043</u> | <u>BX950851</u> | <u>CAG76964.1</u> |
| pectate lyase (PelZ) | <u>Erwinia chrysanthemi 3937</u> | <u>X97119</u> | <u>CAA65785.1</u> |
| pectate lyase A (PelA) | <u>Erwinia chrysanthemi 3937</u> | <u>M77808</u> | <u>AAA24846.1</u> |
| | | | <u>CAA47821.1</u> |
| pectate lyase B (PelB) | <u>Erwinia chrysanthemi 3937</u> | <u>X67475</u> | <u>S25262</u> |
| pectate lyase (PelD) | <u>Erwinia chrysanthemi 3937</u> | <u>AJ132101</u> | <u>CAA10570.1</u> |
| | | <u>M33584</u> | <u>AAA24854.1</u> |
| pectate lyase E (PelE) | <u>Erwinia chrysanthemi 3937 / B374</u> | <u>X17284</u> | <u>CAA35175.1</u> |
| pectate lyase D (PelD) | <u>Erwinia chrysanthemi B374</u> | <u>X17284</u> | <u>CAA35176.1</u> |
| | | | |
| pectate lyase (PelA) | <u>Erwinia chrysanthemi EC16</u> | <u>M14509</u> | <u>AAA24843.1</u> |
| | | <u>M19411</u> | <u>AAA24849.1</u> |
| | | - | <u>AAR45490.1</u> |
| pectate lyase (PelC) | <u>Erwinia chrysanthemi EC16</u> | - | <u>AAW11900.1</u> |
| pectate lyase (PelB;PIB) | <u>Erwinia chrysanthemi EC16</u> | <u>M14510</u> | <u>AAA24847.1</u> |
| pectate lyase (PelE) | <u>Erwinia chrysanthemi EC16</u> | <u>M14509</u> | <u>AAA24844.1</u> |

Figure 38C
Pectate Lyases

| Protein | Organism | GenBank | GenPept |
|---------------------------------------|--|------------------|--------------------|
| pectate lyase C (PelC) | <u>Erwinia chrysanthemi strain 3937</u> | <u>AJ132325</u> | <u>CAA10642.1</u> |
| pectin lyase (PnIA) | <u>Pectobacterium carotovorum Ecc71</u> | <u>M59909</u> | <u>AAA24856.1</u> |
| pectate lyase III (Pel3;PelC) | <u>Pectobacterium carotovorum Er</u> | <u>D10064</u> | <u>BAA00953.1</u> |
| pectate lyase B (PelB) | <u>Pseudoalteromonas haloplanktis 505</u> | <u>AF278705</u> | <u>AAF86343.1</u> |
| | | <u>AF278705</u> | <u>AAF86343.2</u> |
| pectate lyase A | <u>Pseudoalteromonas haloplanktis ANT/505</u> | <u>AF278706</u> | <u>AAF86344.2</u> |
| pectate lyase (Pel) | <u>Pseudomonas fluorescens CY091</u> | <u>L41673</u> | <u>AAA93535.1</u> |
| | | <u>L38902</u> | <u>AAB46399.1</u> |
| pectin lyase (PnL) (fragment) | <u>Pseudomonas marginalis N6301</u> | <u>M84971</u> | <u>AAA92512.1</u> |
| | | <u>D32121</u> | <u>BAA06847.1</u> |
| pectate lyase (Pel) | <u>Pseudomonas marginalis N6301</u> | <u>S65042</u> | <u>AAC60448.1</u> |
| | | <u>D32122</u> | <u>BAA06848.1</u> |
| pectate lyase P (PelP) | <u>Pseudomonas syringae pv. lachrymans</u> | <u>U75414</u> | <u>AAB17879.1</u> |
| | | <u>L38901</u> | <u>AAB46398.1</u> |
| | | <u>L38574</u> | <u>AAC41521.1</u> |
| | | <u>DQ273695</u> | <u>ABB55454.1</u> |
| pectate lyase (Pel;Pstru-4) | <u>Pseudomonas viridiflava</u> | <u>D44611</u> | <u>BAA08077.1</u> |
| pectate lyase (Pel) | <u>Pseudonocardia sp.</u> | <u>AF002241</u> | <u>AAC38059.1</u> |
| pectate lyase (SCO2821;SCBAC17F8.12c) | <u>Streptomyces coelicolor A3(2)</u> | <u>AL596030</u> | <u>CAC44284.1</u> |
| | | <u>NC 003888</u> | <u>NP 627050.1</u> |
| pectate lyase (SCO1880;SCI39.27c) | <u>Streptomyces coelicolor A3(2)</u> | <u>AL591322</u> | <u>CAC38815.1</u> |
| | | <u>NC 003888</u> | <u>NP 626147.1</u> |
| pectate lyase A (PelA;TM0433) | <u>Thermotoga maritima MSB8</u> | <u>AE001722</u> | <u>AAD35518.1</u> |
| | | <u>NC 000853</u> | <u>NP 228243.1</u> |
| XC_1298 | <u>Xanthomonas campestris pv. campestris str. 8004</u> | <u>CP000050</u> | <u>AAV48367.1</u> |
| XC_3590 | <u>Xanthomonas campestris pv. campestris str. 8004</u> | <u>CP000050</u> | <u>AAV50632.1</u> |
| pectate lyase (Pel;XCC0645) | <u>Xanthomonas campestris pv. campestris str. ATCC 33913</u> | <u>AE012162</u> | <u>AAM39961.1</u> |
| | | <u>NC 003902</u> | <u>NP 636037.1</u> |
| pectate lyase II (PelB;XCC2815) | <u>Xanthomonas campestris pv. campestris str. ATCC 33913</u> | <u>AE012393</u> | <u>AAM42087.1</u> |
| | | <u>NC 003902</u> | <u>NP 638163.1</u> |

Figure 38D
Pectate Lyases

| Protein | Organism | GenBank | GenPept |
|--------------------------------------|---|--------------------------------|-------------------|
| pectate lyase (PelB;PI;Pstru-3) | <u>Xanthomonas campestris pv. malvacearum strain B414</u> | <u>L38573</u> | <u>AAC41522.1</u> |
| pectin lyase (AN2331.2) | <u>Aspergillus nidulans FGSC A4</u> | <u>DQ490478</u> | <u>ABF50854.1</u> |
| | | <u>AACD010000</u> <u>38</u> | <u>EAA64442.1</u> |
| pectin lyase (AN2569.2) | <u>Aspergillus nidulans FGSC A4</u> | <u>AACD010000</u> <u>43</u> | <u>EAA64674.1</u> |
| | | <u>DQ490480</u> | <u>ABF50856.1</u> |
| pectate lyase (PelA;AN0741.2) | <u>Aspergillus nidulans FGSC A4</u> | <u>U05592</u> | <u>AAA80568.1</u> |
| | | <u>DQ490468</u> | <u>ABF50844.1</u> |
| | | <u>EF452421</u> | <u>ABO38859.1</u> |
| | | <u>AACD010000</u> <u>12</u> | <u>EAA65383.1</u> |
| pectate lyase (AN7646.2) | <u>Aspergillus nidulans FGSC A4</u> | <u>AACD010001</u> <u>30</u> | <u>EAA61832.1</u> |
| | | <u>DQ490513</u> | <u>ABF50889.1</u> |
| pectin lyase A (PelA) - PI1A | <u>Aspergillus niger CBS 120.49 / N400</u> | <u>X55784</u> | <u>CAA39305.1</u> |
| | | <u>X60724</u> | <u>CAA43130.1</u> |
| pectin lyase C (PelC) | <u>Aspergillus niger CBS 120.49 / N400</u> | <u>AY839647</u> | <u>AAW03313.1</u> |
| pectin lyase F (PelF) | <u>Aspergillus niger CBS 120.49 / N400</u> | <u>AJ489943</u> | <u>CAD34589.1</u> |
| pectate lyase A (PlyA) | <u>Aspergillus niger CBS 120.49 / N400</u> | <u>AJ276331</u> | <u>CAC33162.1</u> |
| pectin lyase B (PelB) | <u>Aspergillus niger CBS 120.49 / N400</u> | <u>A12248</u> | <u>CAA01023.1</u> |
| | | <u>X65552</u> | <u>CAA46521.1</u> |
| An14g04370 (PelA) | <u>Aspergillus niger CBS 513.88</u> | <u>AM270321</u> | <u>CAK48529.1</u> |
| An03g00190 (PelB) | <u>Aspergillus niger CBS 513.88</u> | <u>AM270043</u> | <u>CAK37997.1</u> |
| An15g07160 (PelF) | <u>Aspergillus niger CBS 513.88</u> | <u>AM270351</u> | <u>CAK48551.1</u> |
| An19g00270 (PelD) | <u>Aspergillus niger CBS 513.88</u> | <u>AM270415</u> | <u>CAK47350.1</u> |
| pectate lyase I (PlyA;An10g00870) | <u>Aspergillus niger CBS 513.88</u> | <u>AM270216</u> | <u>CAK40523.1</u> |
| pectin lyase D (PelD) | <u>Aspergillus niger N756</u> | <u>M55657</u> | <u>AAA32701.1</u> |
| pectin lyase 2 (Pel2) | <u>Aspergillus oryzae KBN616</u> | <u>AB029323</u> | <u>BAB82468.1</u> |
| pectin lyase 1 (Pel1) | <u>Aspergillus oryzae KBN616</u> | <u>AB029322</u> | <u>BAB82467.1</u> |
| pectin lyase 1 (Pel1;AO090010000504) | <u>Aspergillus oryzae RIB 40</u> | <u>EF452419</u> | <u>ABO38857.1</u> |
| | | <u>AP007175</u> | <u>BAE66352.1</u> |
| pectin lyase 2 (Pel2;AO090010000030) | <u>Aspergillus oryzae RIB 40</u> | <u>AP007175</u> | <u>BAE65949.1</u> |

Figure 38E
Pectate Lyases

| Protein | Organism | GenBank | GenPept |
|--------------------------------------|---|-----------------|-------------------|
| pectate lyase (PelB) | <u>Colletotrichum gloeosporioides</u> | <u>AF052632</u> | <u>AAD09857.1</u> |
| pectin lyase (PnlA) | <u>Colletotrichum gloeosporioides</u> | <u>L22857</u> | <u>AAA21817.1</u> |
| pectate lyase 2 (Pel-2) | <u>Colletotrichum gloeosporioides f. sp. malvae</u> | <u>AF156985</u> | <u>AAD43566.1</u> |
| pectin lyase (Pnl1;Pnl-1) | <u>Colletotrichum gloeosporioides f. sp. malvae</u> | <u>AF158256</u> | <u>AAF22244.1</u> |
| pectin lyase 2 (Pnl2;Pnl-2) | <u>Colletotrichum gloeosporioides f. sp. malvae</u> | <u>AF156984</u> | <u>AAD43565.1</u> |
| pectate lyase 1 (Pel-1) | <u>Colletotrichum gloeosporioides f. sp. malvae</u> | <u>AF156983</u> | <u>AAD43564.1</u> |
| pectate lyase (LLP-52) | <u>Lilium longiflorum</u> | <u>L18911</u> | <u>AAA33398.1</u> |
| | | <u>EF026017</u> | <u>ABM68553.1</u> |
| | | <u>Z17328</u> | <u>CAA78976.1</u> |
| pectate lyase (PelI;PI1;MwPI1;Ban17) | <u>Musa acuminata Williams</u> | <u>AF206319</u> | <u>AAF19195.1</u> |
| | | <u>DQ663594</u> | <u>ABG74583.1</u> |
| | | <u>X92943</u> | <u>CAA63496.1</u> |
| pectate lyase | <u>Nicotiana tabacum</u> | <u>X61102</u> | <u>CAA43414.1</u> |
| | | <u>X67158</u> | <u>CAA47630.1</u> |
| | | <u>X67159</u> | <u>CAA47631.1</u> |
| pectate lyase | <u>Zinnia elegans</u> | <u>Y09541</u> | <u>CAA70735.1</u> |
| | | <u>AX005936</u> | <u>CAC05181.1</u> |

Figure 39A
Rhamnogalacturonases

| Protein | Organism | GenBank/GenPept | |
|--|--|---------------------|--------------------|
| rhamnogalacturonate lyase (RhiE) | <u>Erwinia chrysanthemi</u> <u>3937</u> | <u>AJ438339</u> | <u>CAD27359.1</u> |
| rhamnogalacturonan lyase (RhgB) | <u>Aspergillus aculeatus</u> <u>KSM 510</u> | <u>L35500</u> | <u>AAA64368.1</u> |
| rhamnogalacturonan lyase (AN6395.2) | <u>Aspergillus nidulans</u> <u>FGSC A4</u> | <u>AACD01000108</u> | <u>EAA58417.1</u> |
| | | <u>DQ490501</u> | <u>ABF50877.1</u> |
| rhamnogalacturonan lyase (AN7135.2) | <u>Aspergillus nidulans</u> <u>FGSC A4</u> | <u>AACD01000122</u> | <u>EAA61387.1</u> |
| | | <u>DQ490504</u> | <u>ABF50880.1</u> |
| rhamnogalacturonan lyase (YesW;BSU07050) | <u>Bacillus subtilis</u> <u>subsp. subtilis str.</u> <u>168</u> | <u>Z99107</u> | <u>CAB12524.1</u> |
| | | <u>NC 000964</u> | <u>NP 388586.1</u> |
| exo-unsaturated rhamnogalacturonan lyase (YesX;BSU07060) | <u>Bacillus subtilis</u> <u>subsp. subtilis str.</u> <u>168</u> | <u>Z99107</u> | <u>CAB12525.1</u> |
| | | <u>NC 000964</u> | <u>NP 388587.1</u> |
| rhamnogalacturonan lyase - Rgl11A | <u>Cellvibrio japonicus</u> <u>(formerly</u> <u>Pseudomonas</u> <u>cellulosa)</u> | <u>AY026755</u> | <u>AAK20911.1</u> |
| CJA_3559 (rhamnogalacturonan lyase) - Rgl11A | <u>Cellvibrio japonicus</u> <u>Ueda107</u> | <u>CP000934.1</u> | <u>ACE83155.1</u> |
| rhamnogalacturonan lyase Y - Rgl11Y | <u>Clostridium</u> <u>cellulolyticum ATCC</u> <u>35319</u> | <u>AF316823</u> | <u>AAG45161.1</u> |

Figure 39B
Rhamnogalacturonate Hydrolases

| Protein | Organism | GenBank/GenPept | |
|---|---|------------------|--------------------|
| <i>GH family 105</i> | | | |
| unsaturated rhamnogalacturonyl hydrolase (BSU30120; YteR) | <u>Bacillus subtilis</u> <u>subsp. subtilis str.</u> <u>168</u> | <u>Z99119</u> | <u>CAB14990.1</u> |
| unsaturated rhamnogalacturonyl hydrolase (BSU07000; YesR) | <u>Bacillus subtilis</u> <u>subsp. subtilis str.</u> <u>168</u> | <u>Z99107</u> | <u>CAB12519.1</u> |
| | | <u>NC 000964</u> | <u>NP 388581.1</u> |

Figure 40A
Pectate Methyl Esterases

| Protein | Organism | GenBank/GenPept | |
|---|---|---------------------|--------------------|
| Family 8 | | | |
| ECA3253 (PemA) | <u>Erwinia carotovora subsp. atroseptica</u> SCRI1043 | <u>BX950851</u> | <u>CAG76151.1</u> |
| ECA0107 (PmeB) | <u>Erwinia carotovora subsp. atroseptica</u> SCRI1043 | <u>BX950851</u> | <u>CAG73027.1</u> |
| pectin methylesterase b | <u>Erwinia chrysanthemi</u> 3937 | <u>X84665</u> | <u>CAA59151.1</u> |
| pectin methylesterase A (PemA;Pem) | <u>Erwinia chrysanthemi</u> 3937 / B374 | <u>L07644</u> | <u>AAA24852.1</u> |
| | | - | <u>AAR64146.1</u> |
| | | <u>Y00549</u> | <u>CAA68628.1</u> |
| pectate lyase A | <u>Pseudoalteromonas haloplanktis</u> ANT/505 | <u>AF278706</u> | <u>AAF86344.2</u> |
| pectin methylesterase (Pme5; Vgd1;At2g47040/F14M4.13) | <u>Arabidopsis thaliana</u> | <u>AC004411</u> | <u>AAC34240.1</u> |
| | | <u>AY091768</u> | <u>AAM10316.1</u> |
| | | <u>BT001120</u> | <u>AAN64511.1</u> |
| | | <u>AY830948</u> | <u>AAV91508.1</u> |
| | | <u>AJ250430</u> | <u>CAB58974.1</u> |
| | | <u>NM 130272</u> | <u>NP 182227.1</u> |
| pectin methylesterase (Pme1) | <u>Aspergillus aculeatus</u> | <u>U49378</u> | <u>AAB42153.1</u> |
| pectin methyl esterase (AN3390.2) | <u>Aspergillus nidulans</u> FGSC A4 | <u>DQ490489</u> | <u>ABF50865.1</u> |
| | | <u>AACD01000055</u> | <u>EAA63358.1</u> |
| pectin methylesterase (Pme1) | <u>Aspergillus niger</u> RH 5344 | <u>A34997</u> | <u>CAA02198.1</u> |
| | | <u>A35006</u> | <u>CAA02201.1</u> |
| | | <u>A35008</u> | <u>CAA02202.1</u> |
| | | <u>X52902</u> | <u>CAA37084.1</u> |
| | | <u>X54145</u> | <u>CAA38084.1</u> |
| pectin methylesterase (PmeA) | <u>Aspergillus oryzae</u> KBN616 | <u>AB011211</u> | <u>BAA75474.1</u> |
| pectin methylesterase (PmeA;AO090012000749) | <u>Aspergillus oryzae</u> RIB 40 | <u>AP007161</u> | <u>BAE60873.1</u> |
| pectin methylesterase (Bcpme2) | <u>Botryotinia fuckeliana</u> Bd90 | <u>AJ428403</u> | <u>CAD21438.1</u> |
| pectin methyl esterase (Bcpme1) | <u>Botryotinia fuckeliana</u> T4 | <u>AJ309701</u> | <u>CAC29255.1</u> |
| pectin methylesterase 1.1 (PECS-1.1) | <u>Citrus sinensis</u> | <u>U82973</u> | <u>AAB57667.1</u> |
| | | <u>U82976</u> | <u>AAB57670.1</u> |

Figure 40B
Pectate Methyl Esterases

| Protein | Organism | GenBank | GenPept |
|---|--|-----------------|-------------------|
| pectin methylesterase (PME1) | <u>Cochliobolus carbonum</u> | <u>AF159252</u> | <u>AA43340.1</u> |
| pectin methylesterase | <u>Daucus carota</u> | | |
| pectinesterase FaPE1 | <u>Fragaria x ananassa</u> | <u>AY324809</u> | <u>AAQ21124.1</u> |
| pectin methyl-esterase (Pef1) | <u>Medicago truncatula</u> | <u>AJ249611</u> | <u>CAB65291.1</u> |
| pectin methyl-esterase (Per) | <u>Medicago truncatula</u> | <u>AJ249611</u> | <u>CAB65290.2</u> |
| pectin methylesterase | <u>Nicotiana benthamiana</u> | <u>AY238968</u> | <u>AAO85706.1</u> |
| pectin methylesterase | <u>Nicotiana plumbaginifolia</u> | <u>Z71752</u> | <u>CAA96434.1</u> |
| pectin methylesterase (NtPME1) | <u>Nicotiana tabacum</u> | <u>AY772945</u> | <u>AAX13972.1</u> |
| pectin methylesterase | <u>Nicotiana tabacum</u> | <u>AJ401158</u> | <u>CAB95025.1</u> |
| pectin methylesterase (PME1) (fragment) | <u>Orobanche cumana</u> | <u>AY072720</u> | <u>AAL66865.1</u> |
| pectin methylesterase (fragment) | <u>Orobanche cumana</u> | <u>AF333068</u> | <u>AAG49395.1</u> |
| pectin methylesterase | <u>Petunia inflata</u> | <u>L27101</u> | <u>AAA33714.1</u> |
| pectin methyl esterase (PttPME1) | <u>Populus tremula x Populus tremuloides</u> | <u>AJ277547</u> | <u>CAC01624.1</u> |
| pectin methylesterase PME1 (fragment) | <u>Prunus armeniaca</u> | <u>AF184079</u> | <u>AAG12248.1</u> |
| pectin methylesterase (SgPME1) | <u>Salix gilgiana</u> | <u>AB029461</u> | <u>BAA89480.1</u> |
| pectin methylesterase | <u>Sitophilus oryzae</u> | <u>AY841894</u> | <u>AAW28928.1</u> |
| | | <u>U50985</u> | <u>AAB67739.1</u> |
| | | - | <u>AAQ71552.1</u> |
| | | <u>A15983</u> | <u>CAA01257.1</u> |
| | | <u>A17011</u> | <u>CAA01315.1</u> |
| | | <u>X07910</u> | <u>CAA30746.1</u> |
| pectin methylesterase 2 | <u>Solanum lycopersicum</u> | <u>X74639</u> | <u>CAA52704.1</u> |
| | | <u>U49330</u> | <u>AAD09283.1</u> |
| pectin methylesterase (PmeU1) | <u>Solanum lycopersicum</u> | <u>AY046596</u> | <u>AAL02367.1</u> |
| | | <u>U50986</u> | <u>AAB67740.1</u> |
| | | <u>A17010</u> | <u>CAA01314.1</u> |
| pectin methylesterase 1 (PME1.9) | <u>Solanum lycopersicum</u> | <u>X74638</u> | <u>CAA52703.1</u> |
| pectin methyl esterase (Pest1) | <u>Solanum tuberosum</u> | <u>AF152171</u> | <u>AAF23891.1</u> |
| pectin methyl esterase (Pest2) | <u>Solanum tuberosum</u> | <u>AF152172</u> | <u>AAF23892.1</u> |
| pectin methylesterase isoform alpha (PME2) (fragment) | <u>Vigna radiata</u> | <u>AF229849</u> | <u>AAF35897.1</u> |
| pectin methylesterase (PME) | <u>Vitis riparia</u> | <u>AF178989</u> | <u>AAD51853.1</u> |

Figure 41
Pectate Acetyl Esterases

| Protein | Organism | GenBank/GenPept | |
|---|---|---------------------|--------------------|
| Family12 | | | |
| acetyl xylan esterase (Rgae;BH1115) | <u>Bacillus halodurans C-125</u> | <u>AP001511</u> | <u>BAB04834.1</u> |
| cephalosporin C deacetylase | <u>Bacillus sp. KCCM10143</u> | <u>NC 002570</u> | <u>NP 241981.1</u> |
| acetyl xylan esterase (YesT;BSU07020) | <u>Bacillus subtilis subsp. subtilis str. 168</u> | <u>AF184175</u> | <u>AAF25818.1</u> |
| ECA3252 (PaeY) | <u>Erwinia carotovora subsp. atroseptica SCRI1043</u> | <u>Z99107</u> | <u>CAB12521.1</u> |
| pectin acetylerase (PaeY) | <u>Erwinia chrysanthemi 3937</u> | <u>NC 000964</u> | <u>NP 388583.1</u> |
| rhamnogalacturonan acetylerase (Rha1) | <u>Aspergillus aculeatus KSM 510</u> | <u>BX950851</u> | <u>CAG76150.1</u> |
| rhamnogalacturonan acetylerase (AN2528.2) | <u>Aspergillus nidulans FGSC A4</u> | <u>Y09828</u> | <u>CAA70971.1</u> |
| pectin acetylerase | <u>Vigna radiata Wilzeck</u> | <u>X89714</u> | <u>CAA61858.1</u> |
| | | <u>DQ490479</u> | <u>ABF50855.1</u> |
| | | <u>AACD01000043</u> | <u>EAA64633.1</u> |
| | | <u>X99348</u> | <u>CAA67728.1</u> |

Figure 42A

Production of 2-phenyl ethanol (24 hrs)

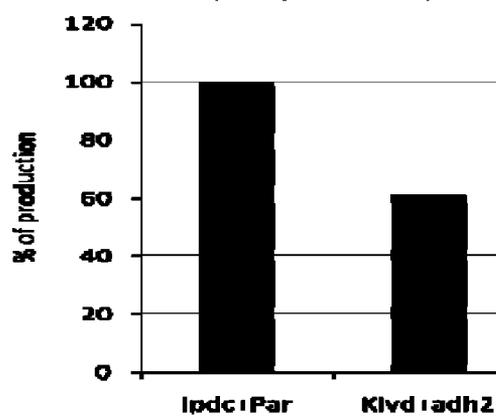


Figure 42B

Production of 2-(4-hydroxyphenyl) ethanol (24 hrs)

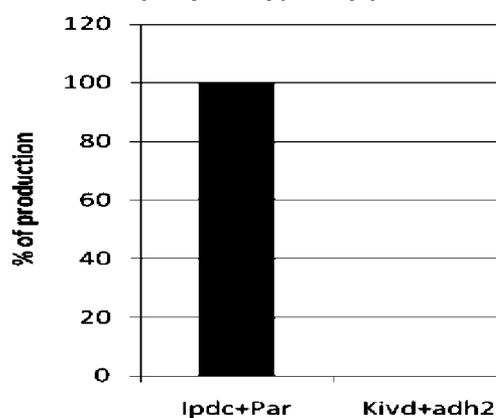


Figure 42C

Production of 2-(indole-3-)ethanol (24 hrs)

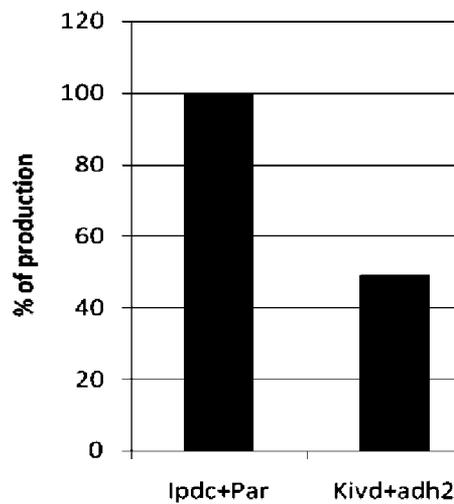


Figure 43A
GC-MS chromatogram for control (pBAD33 and pTrc99A) (one week)

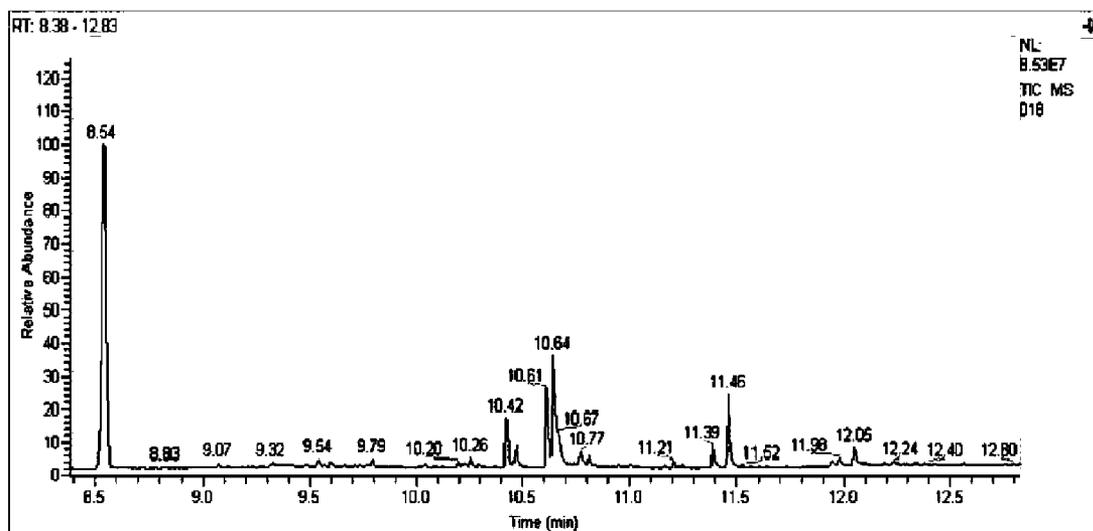


Figure 43B
GC-MS chromatogram for 2-phenylethanol (5.97 min) production from pBADpheA-aroLAC-aroG-*tk*A-aroBDE and pTrcBALK (one week)

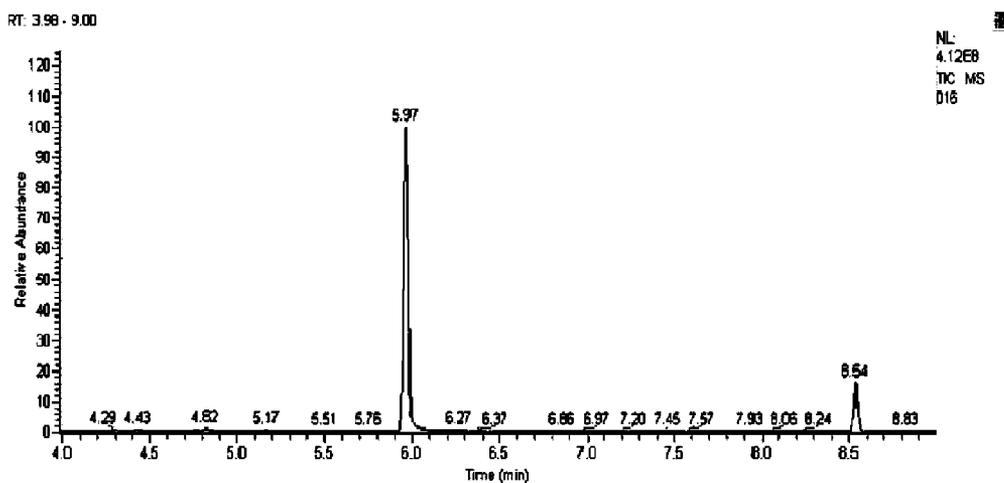
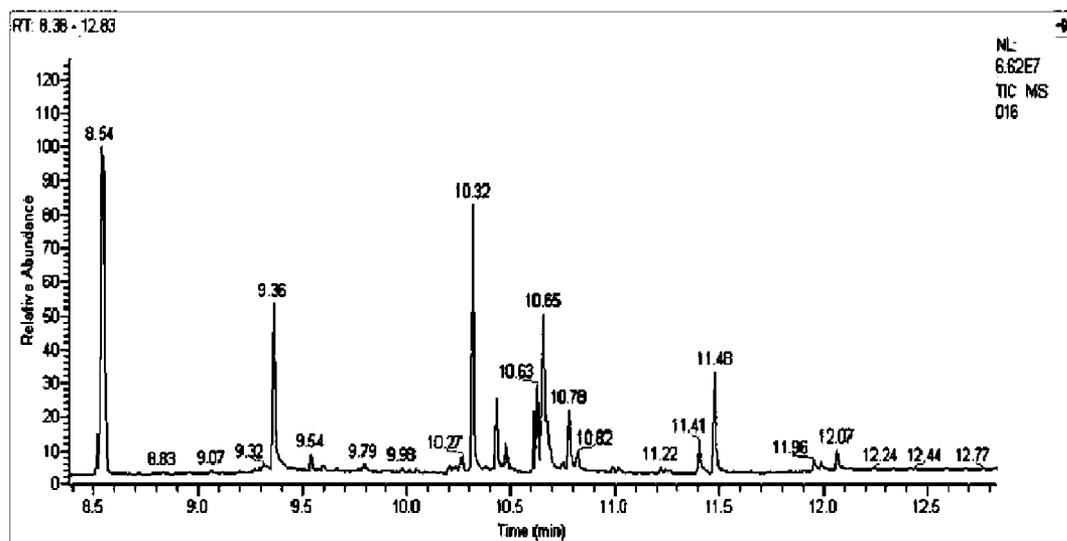


Figure 44

GC-MS chromatogram for 2-(4-hydroxyphenyl) ethanol (9.36 min) and 2-(indole-3) ethanol (10.32 min) production from pBADtyrA-aroLAC-aroG-tktA-aroBDE and pTrcBALK (one week)



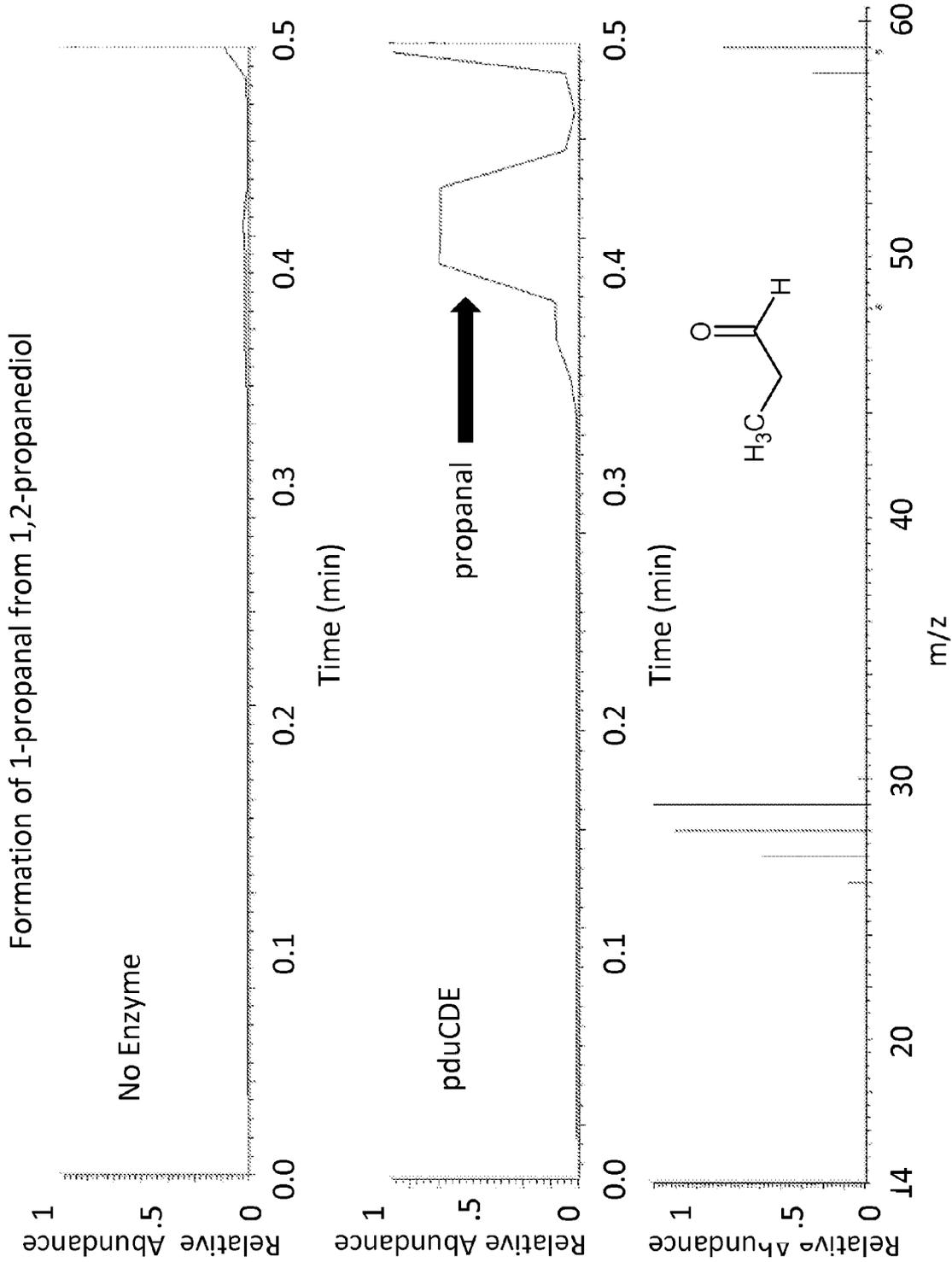


FIG. 45

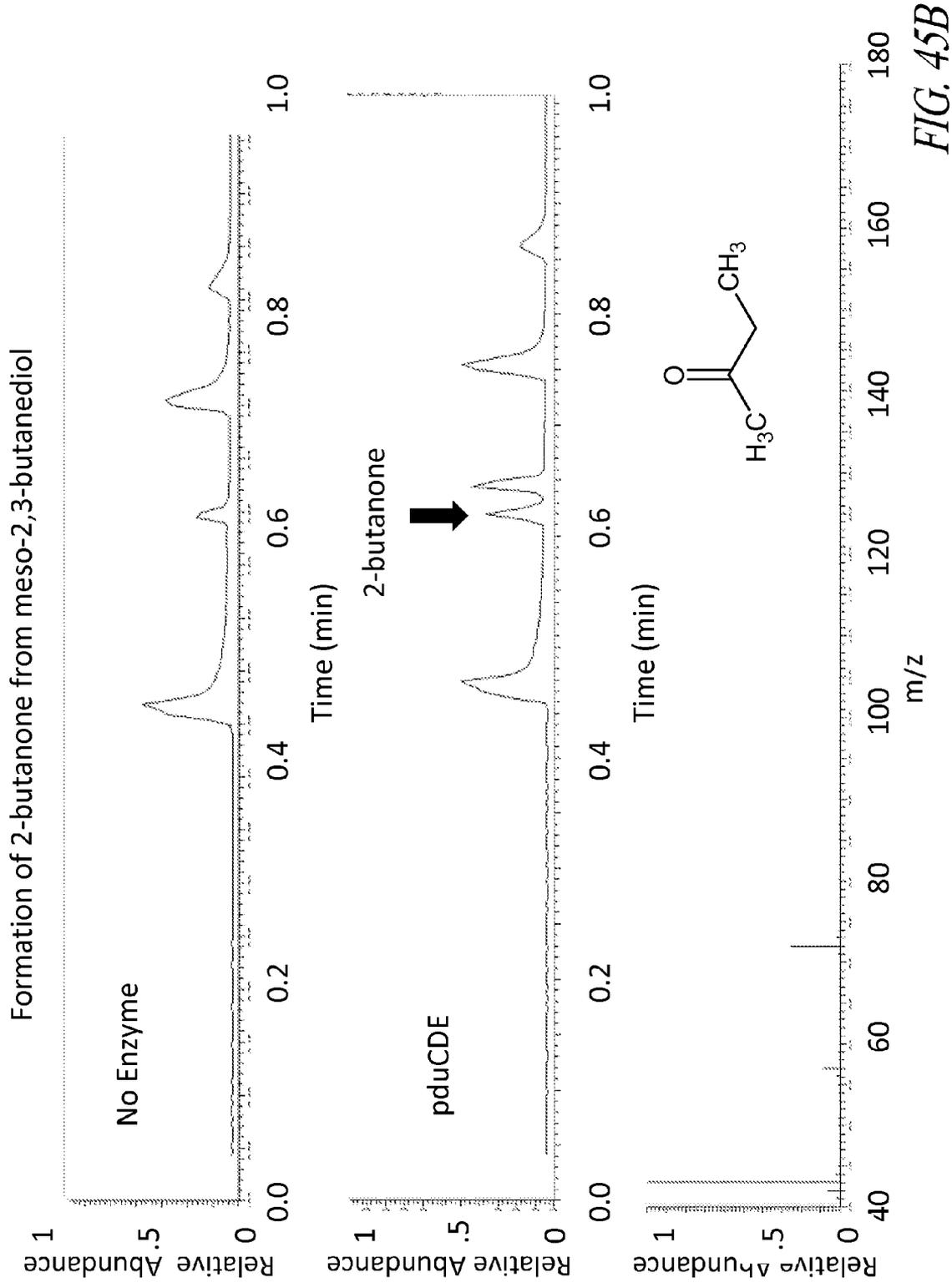
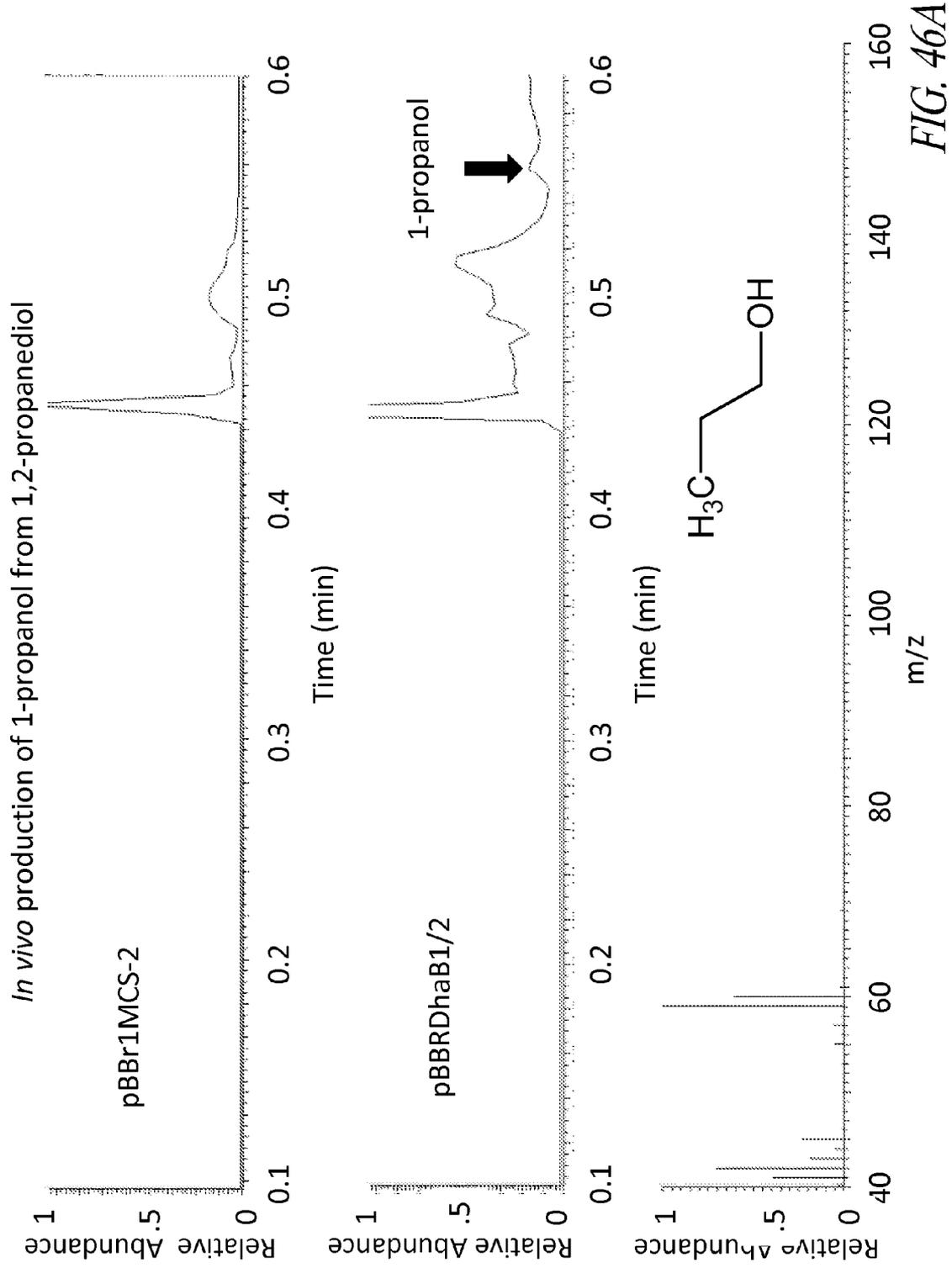
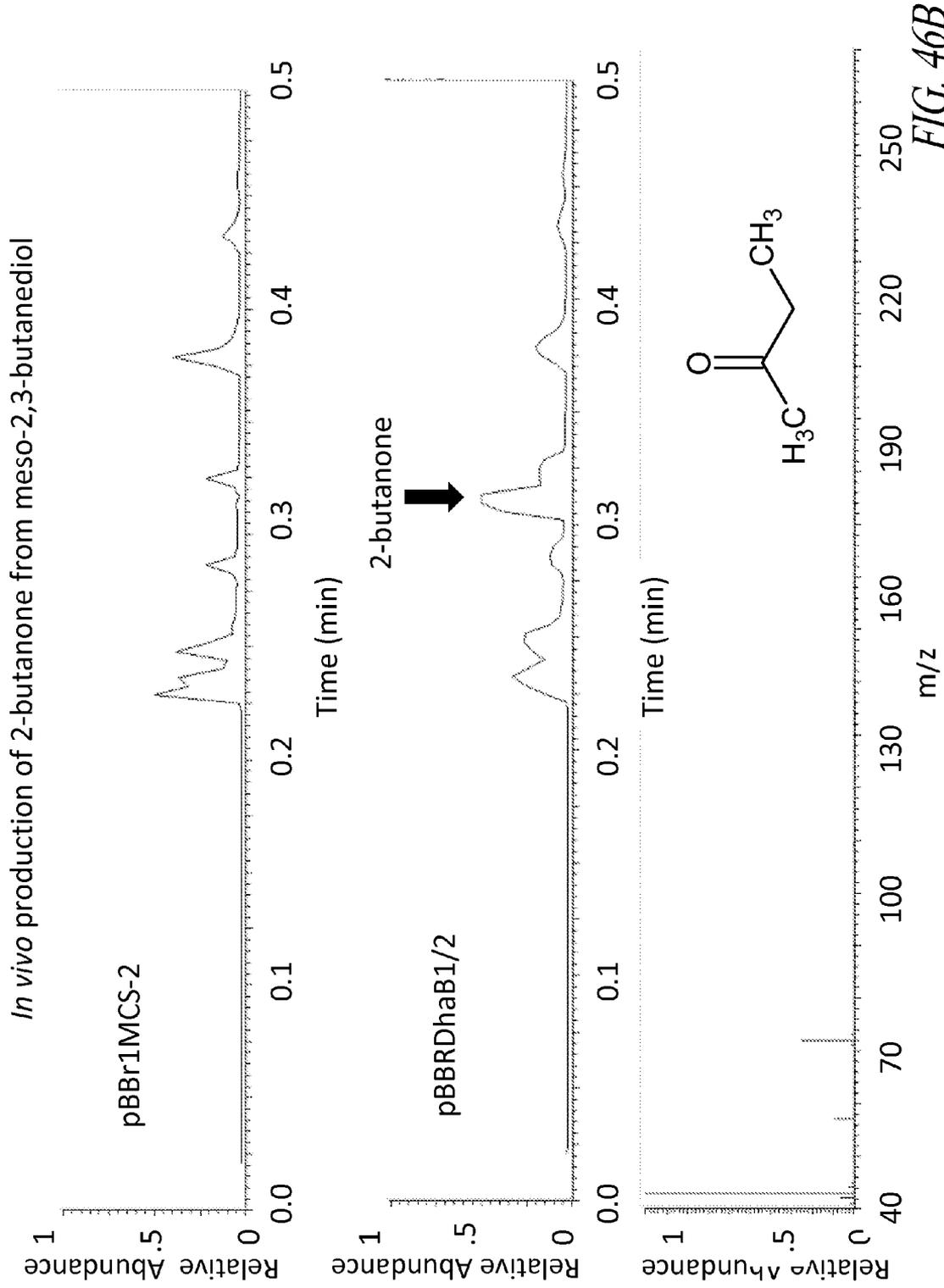
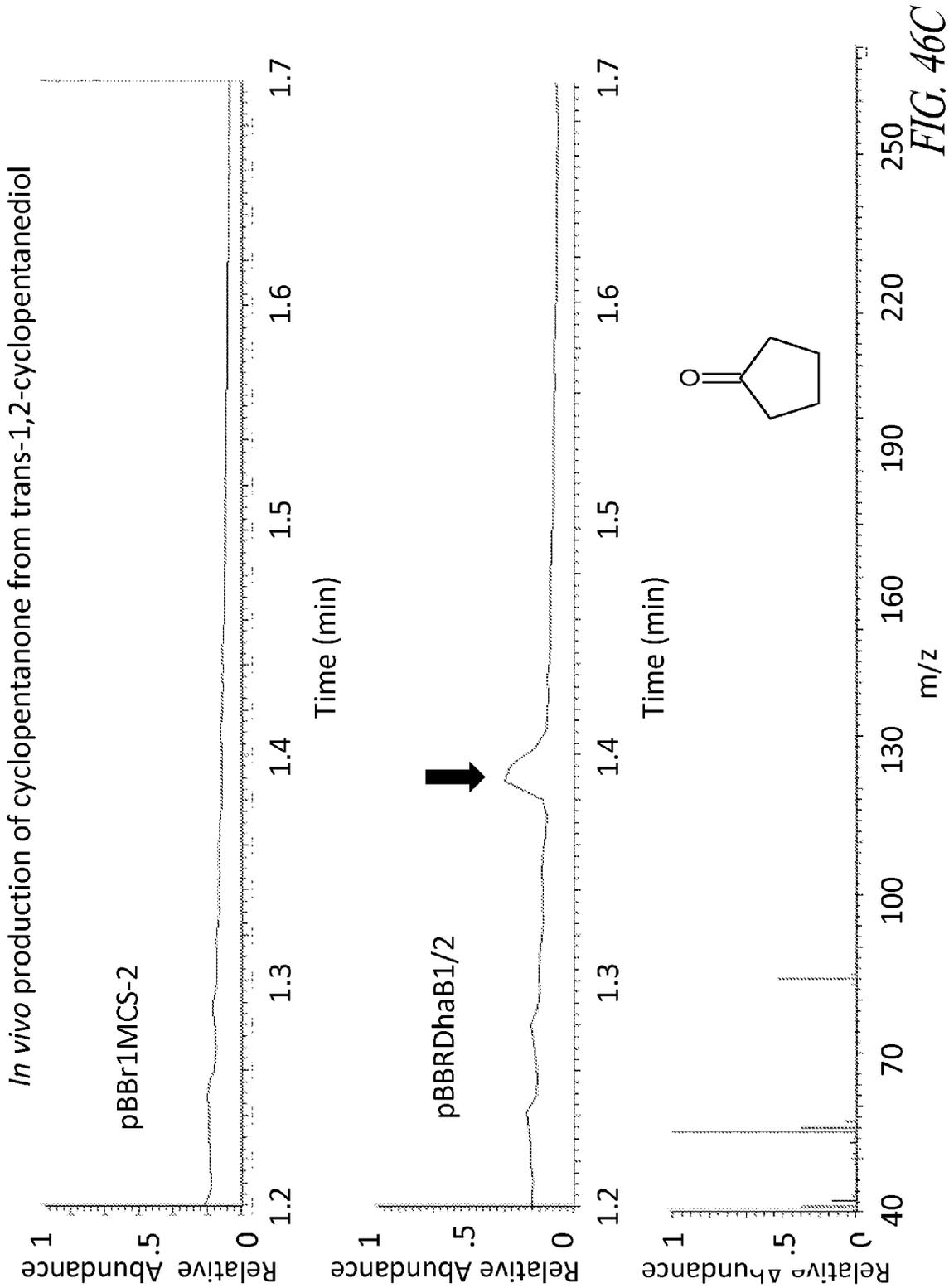


FIG. 45B







Thiobarbituric acid (TBA) Assay

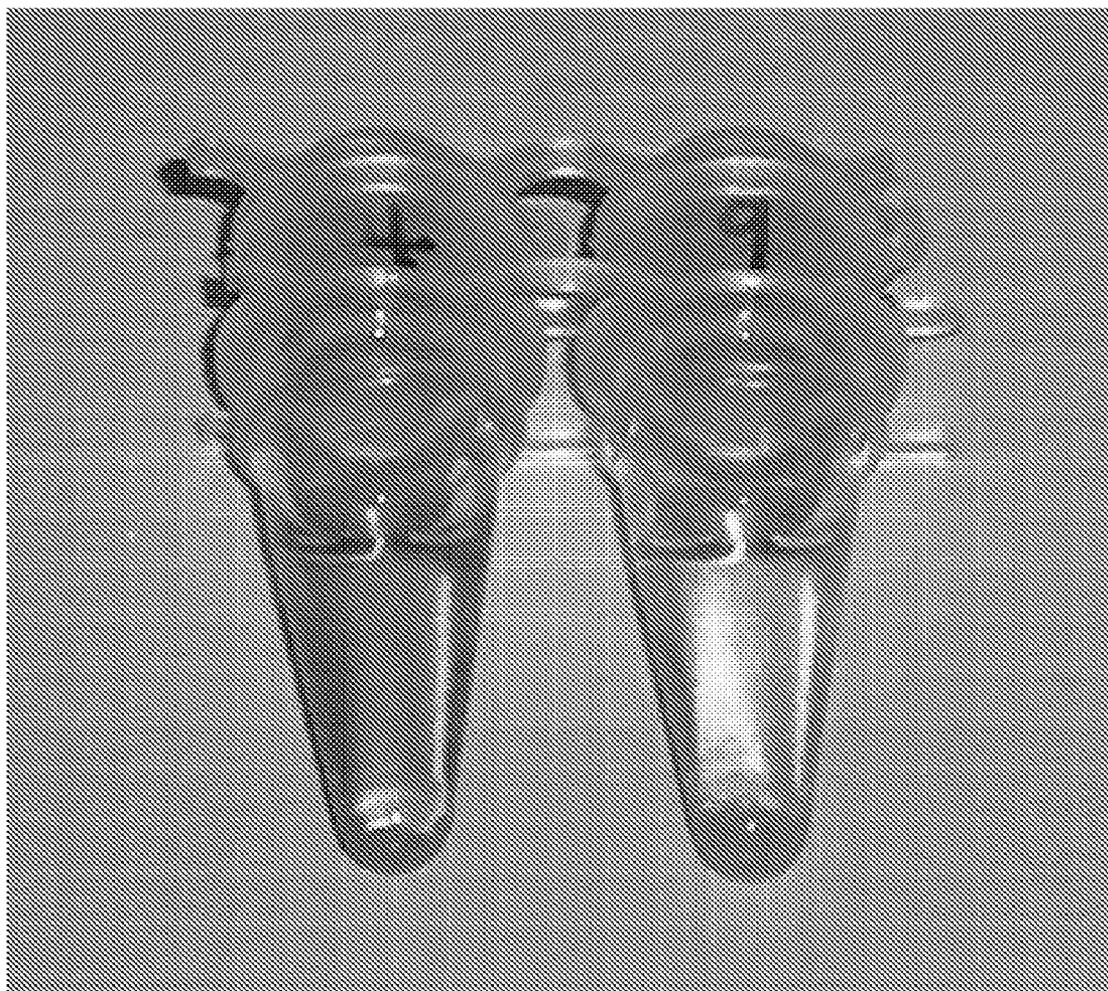


FIG. 47

In vivo production of 3-hydroxy-2-pentanone and 2-hydroxy-3-pentanone from ligation reaction between acetaldehyde and propionaldehyde catalyzed by BAL enzyme

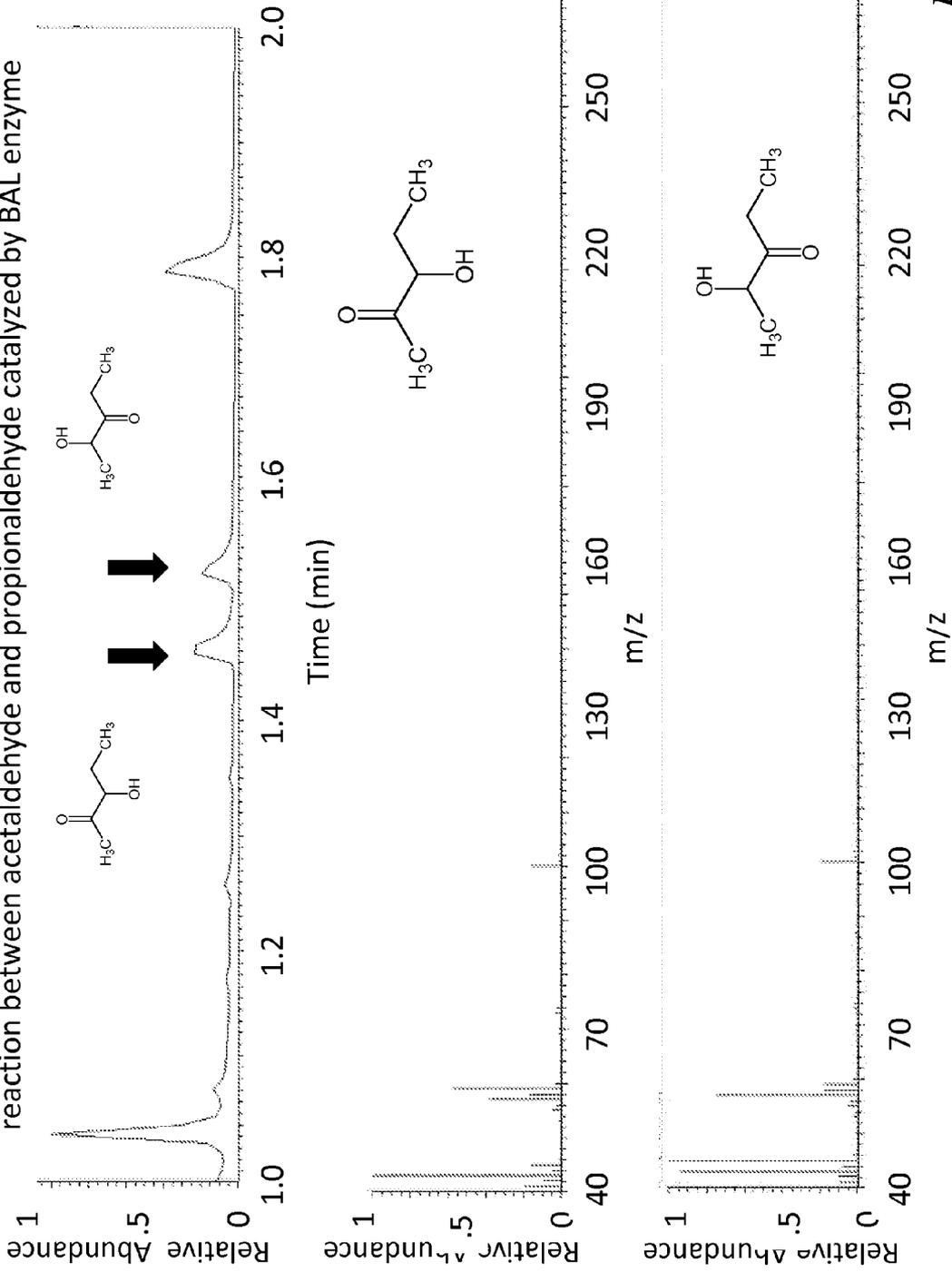
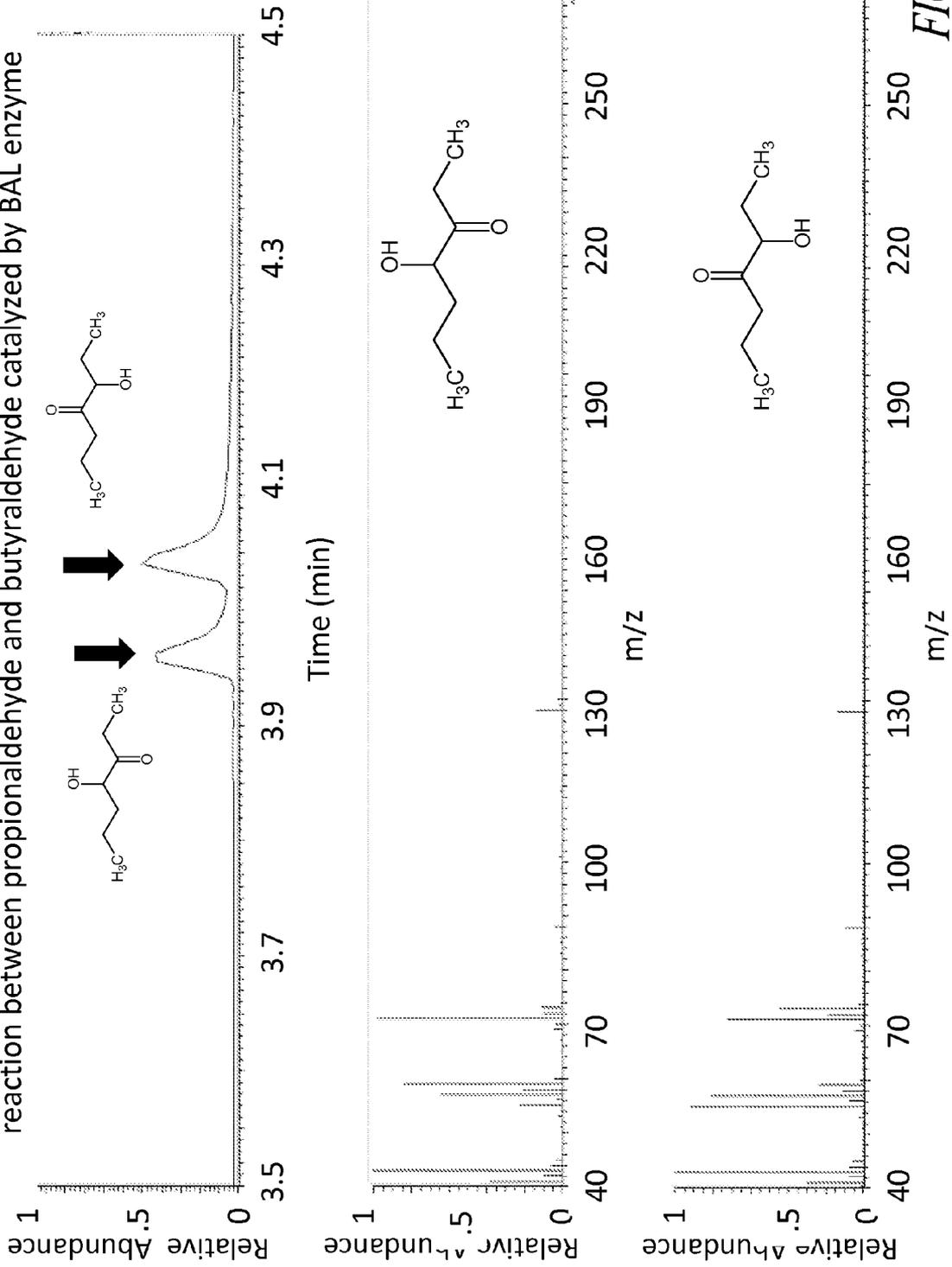


FIG. 48A

In vivo production of 4-hydroxy-3-heptanone and 3-hydroxy-4-heptanone from ligation reaction between propionaldehyde and butyraldehyde catalyzed by BAL enzyme



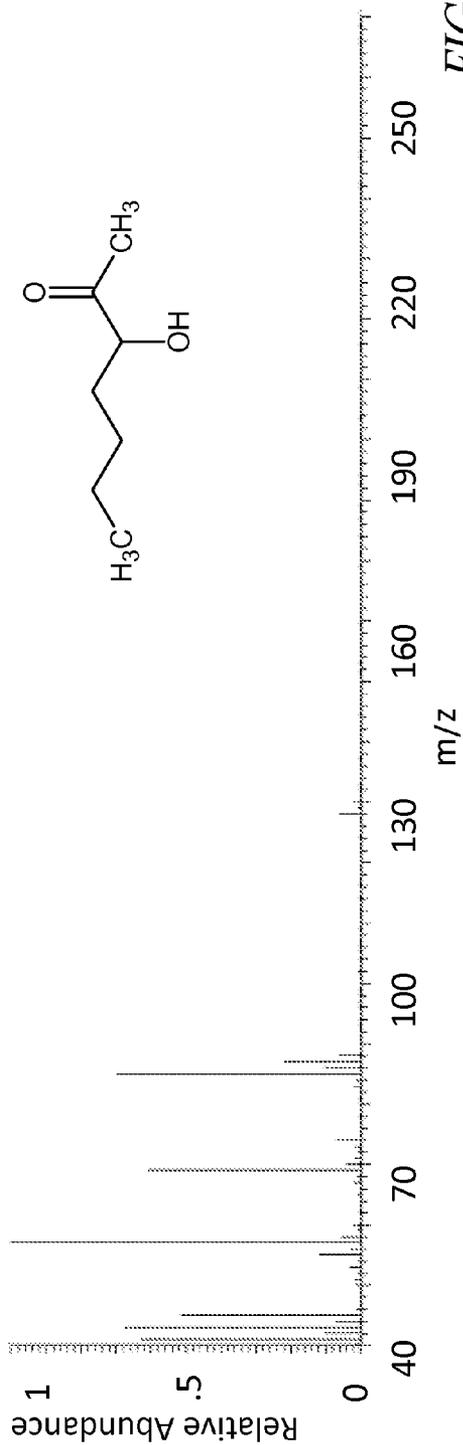
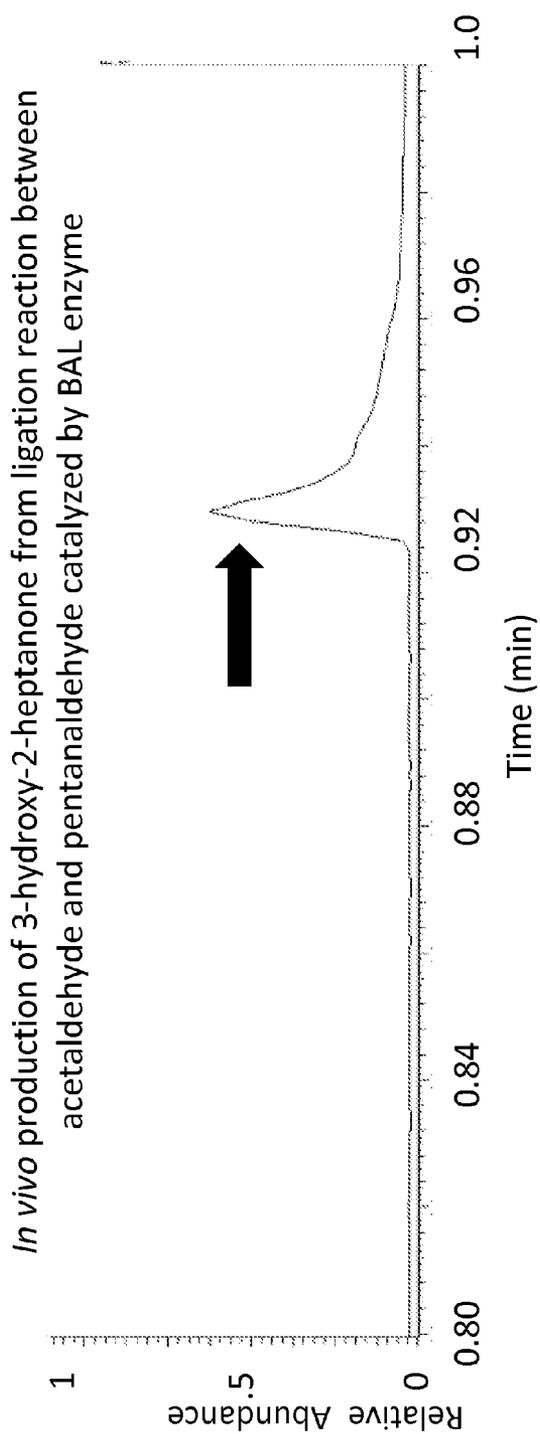


FIG. 49A

In vivo production of 4-hydroxy-3-octanone and 3-hydroxy-4-octanone from ligation reaction between pentanaldehyde and propionaldehyde catalyzed by the BAL enzyme

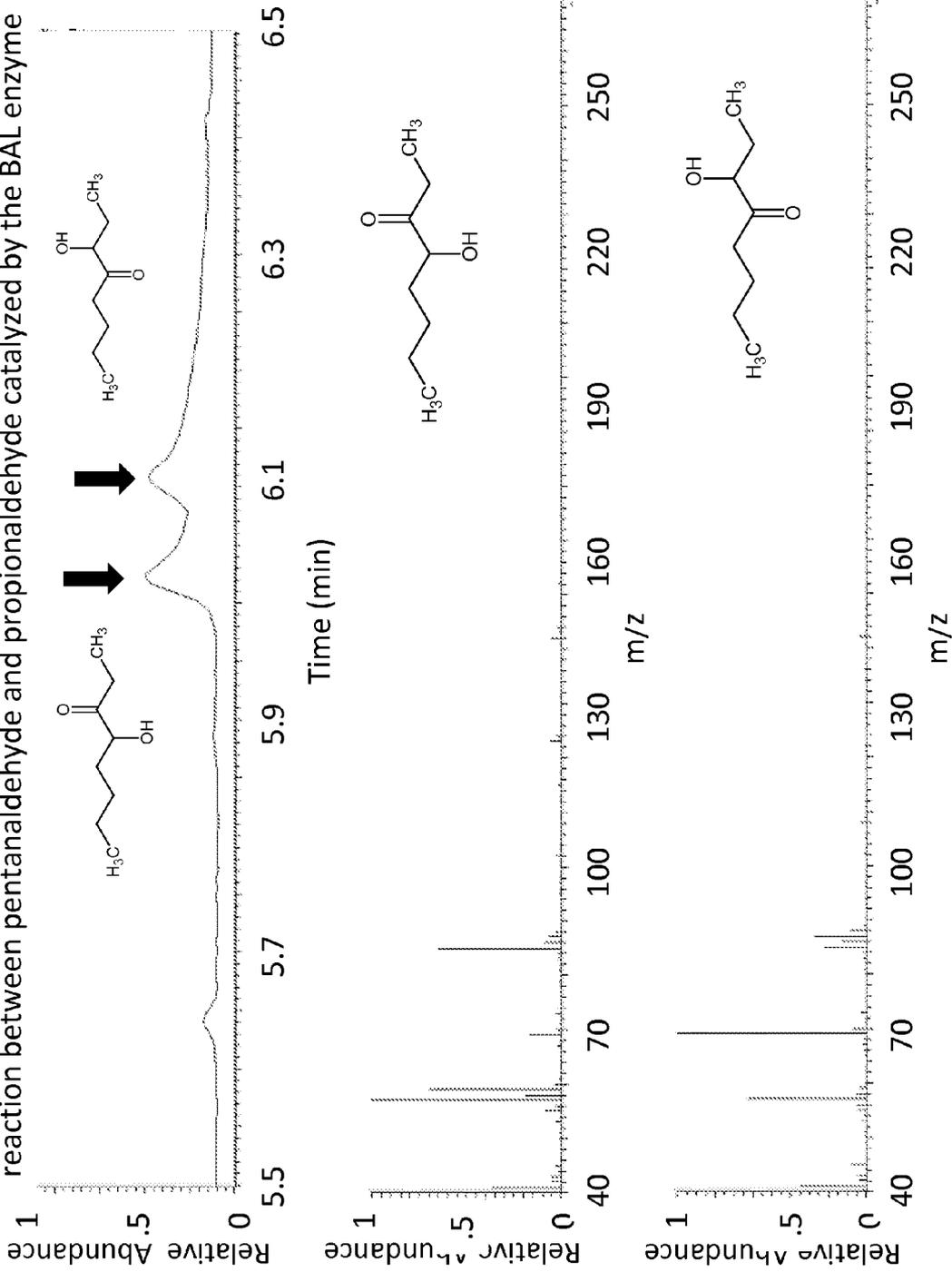


FIG. 49B

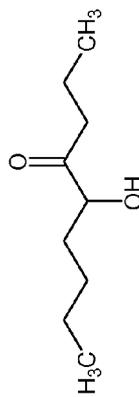
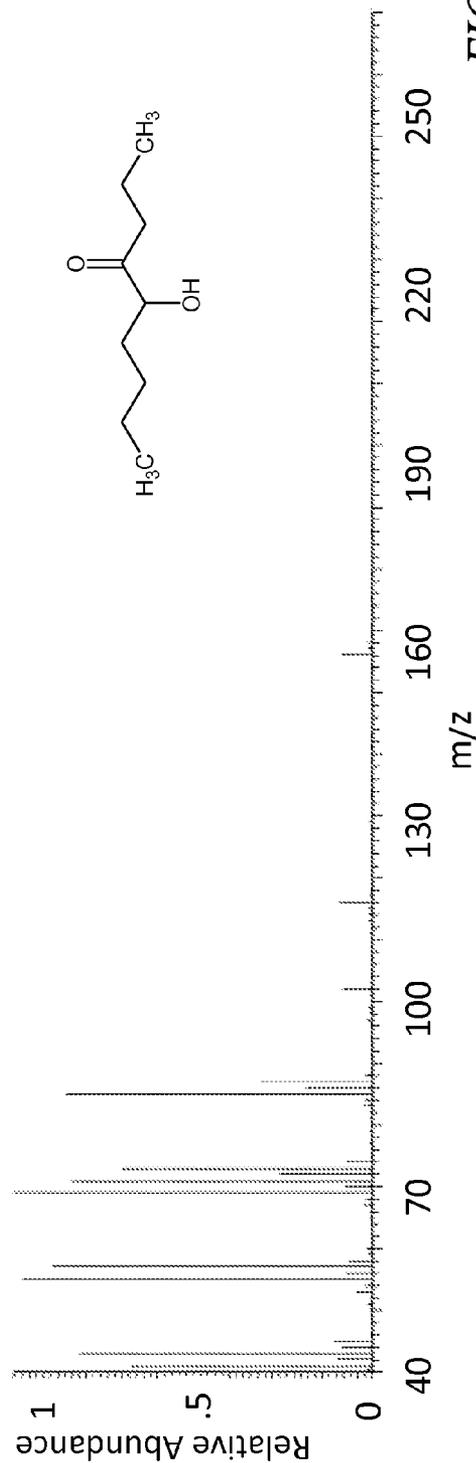
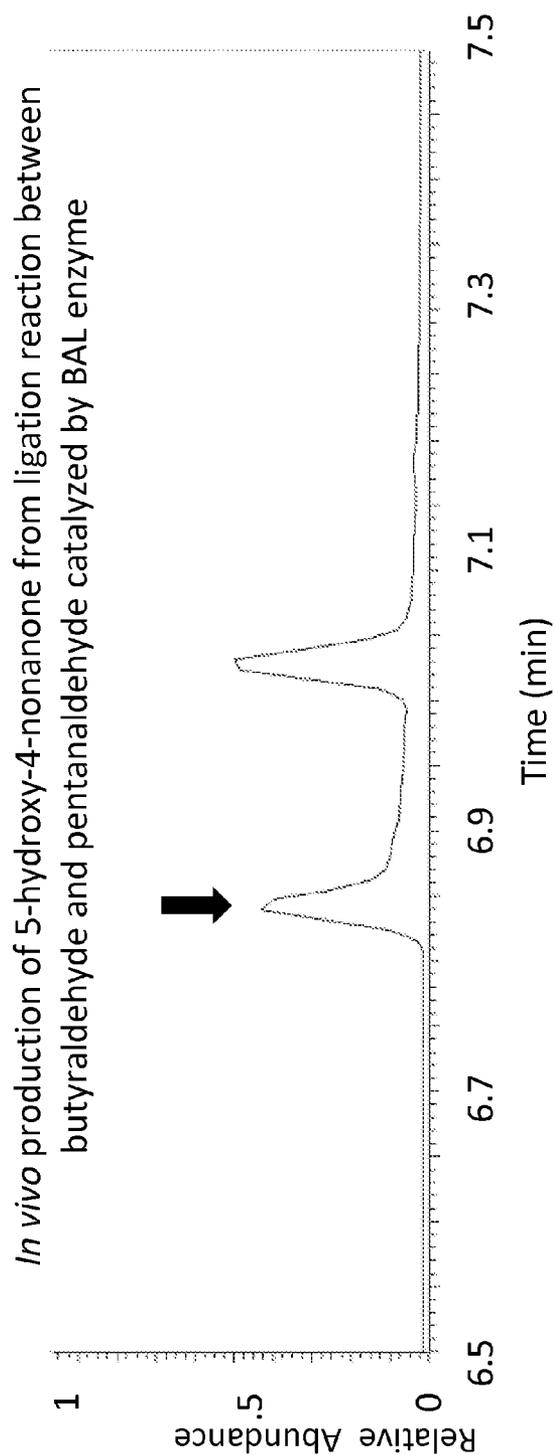


FIG. 50A

In vivo production of 2-methyl-5-hydroxy-4-decanone and 2-methyl-4-hydroxy-5-decanone from ligation reaction between hexanal and 3-methylbutyraldehyde catalyzed by BAL

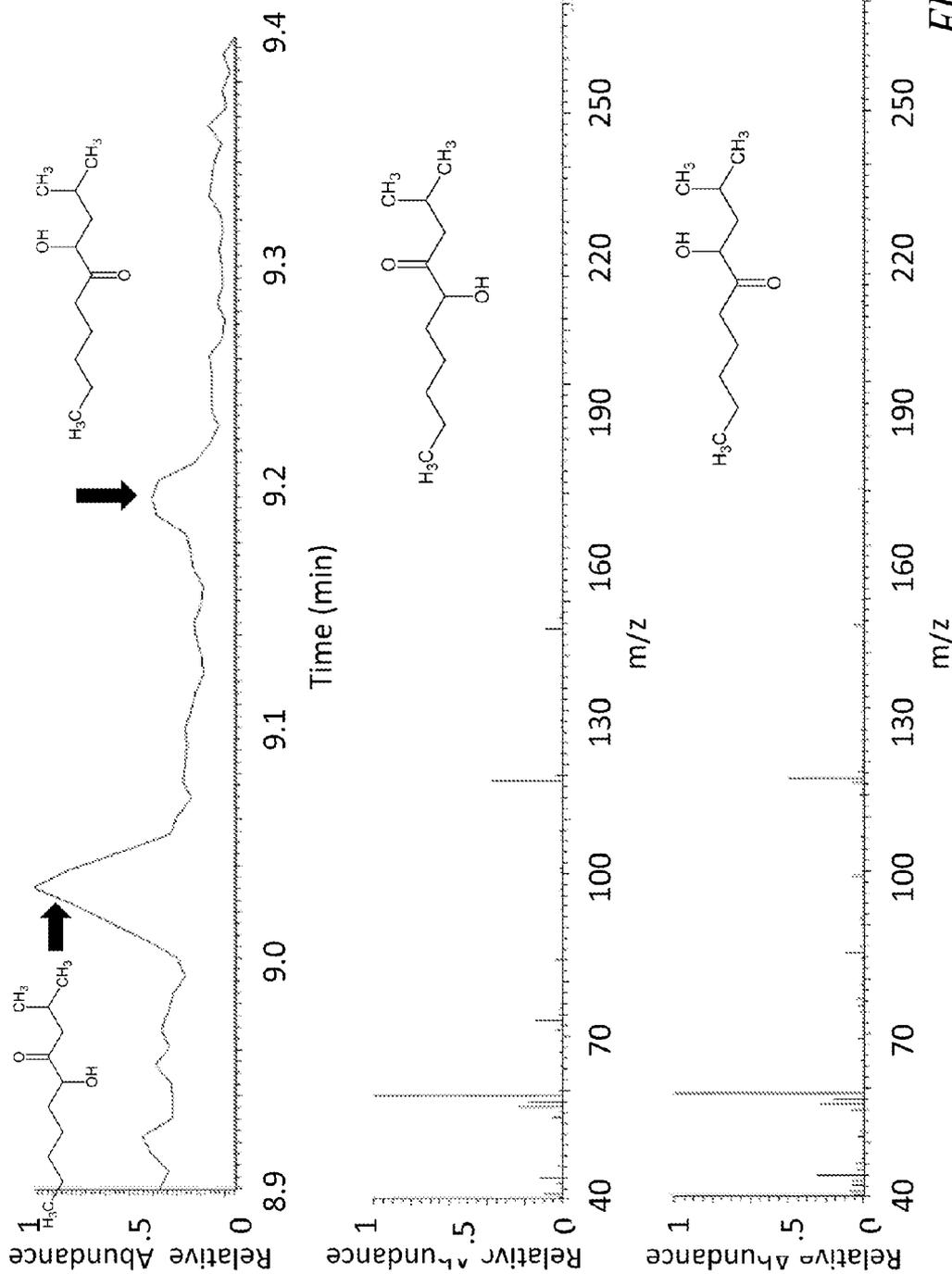


FIG. 50B

In vivo production of 6-methyl-3-hydroxy-2-heptanone from ligation reaction between acetaldehyde and 4-methylhexanaldehyde catalyzed by BAL enzyme

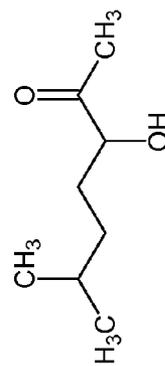
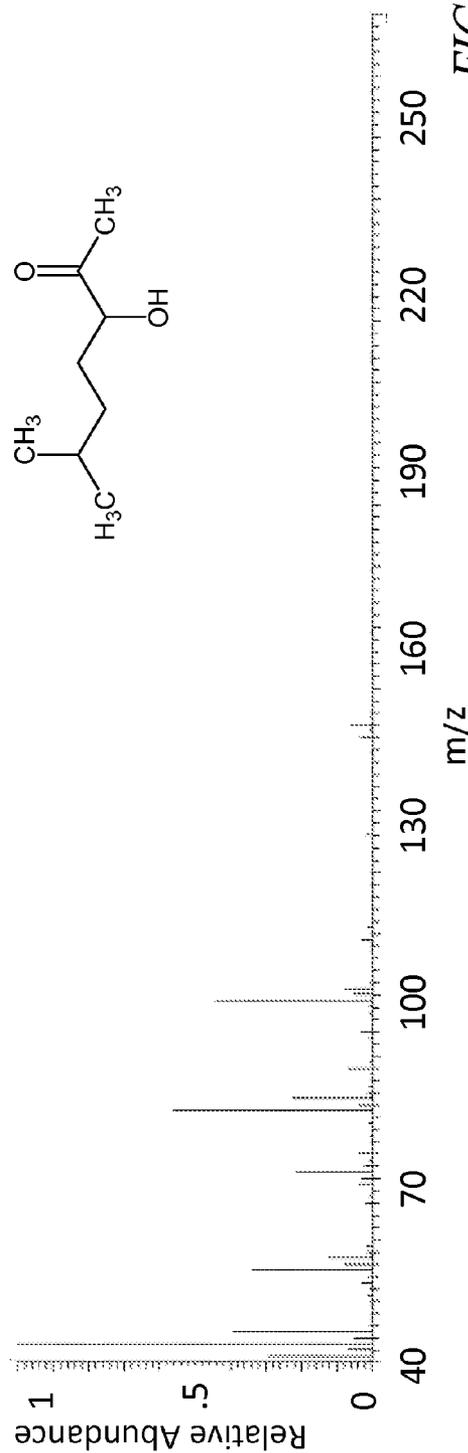
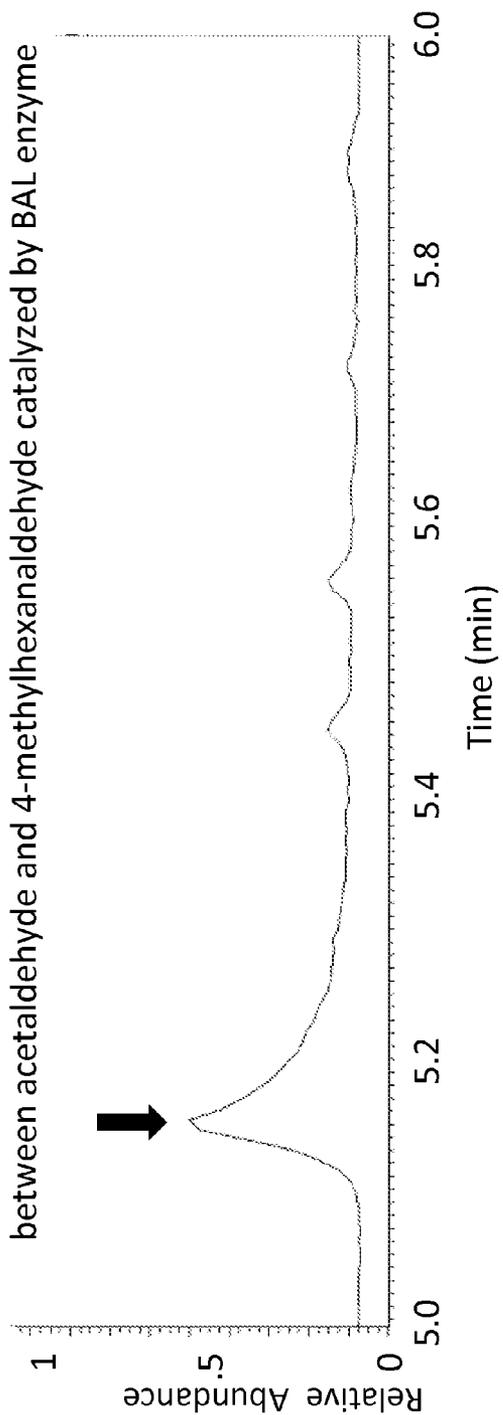


FIG. 51A

In vivo production of 7-methyl-4-hydroxy-3-octanone from ligation reaction between 4-methylhexanaldehyde and propionaldehyde catalyzed by BAL enzyme

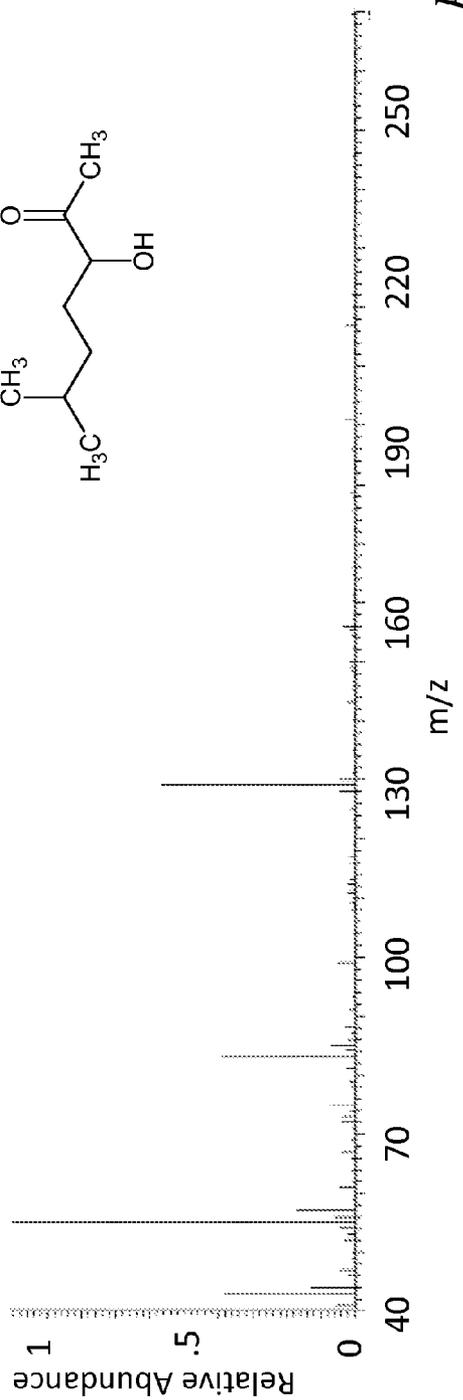
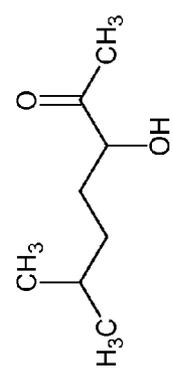
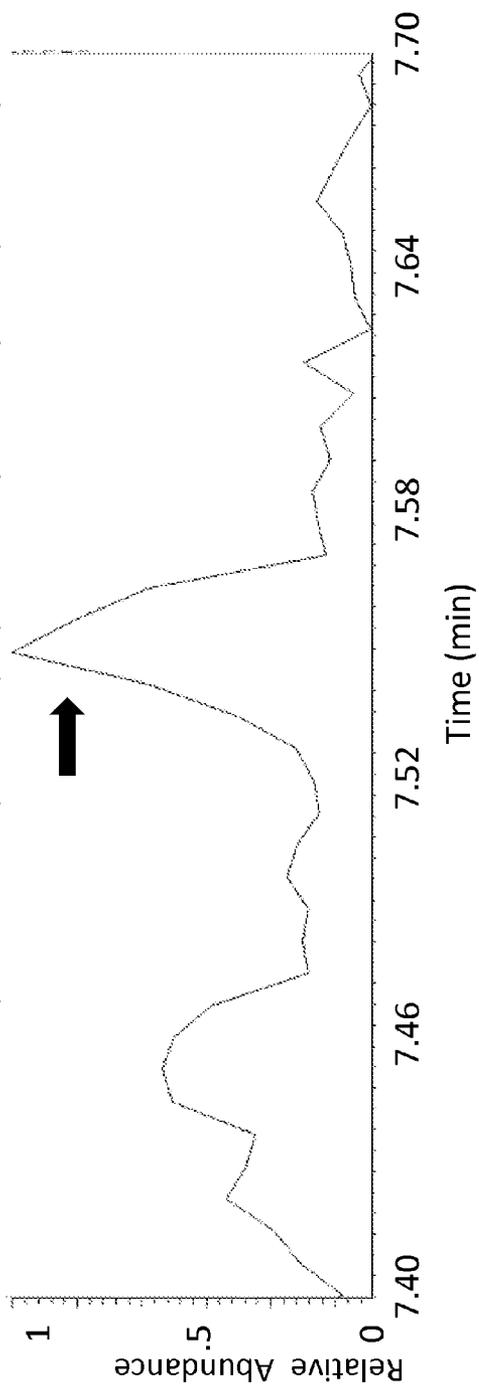


FIG. 51B

In vivo production of 8-methyl-5-hydroxy-4-nonanone from ligation reaction between 4-methylhexanaldehyde and butyraldehyde catalyzed by BAL enzyme

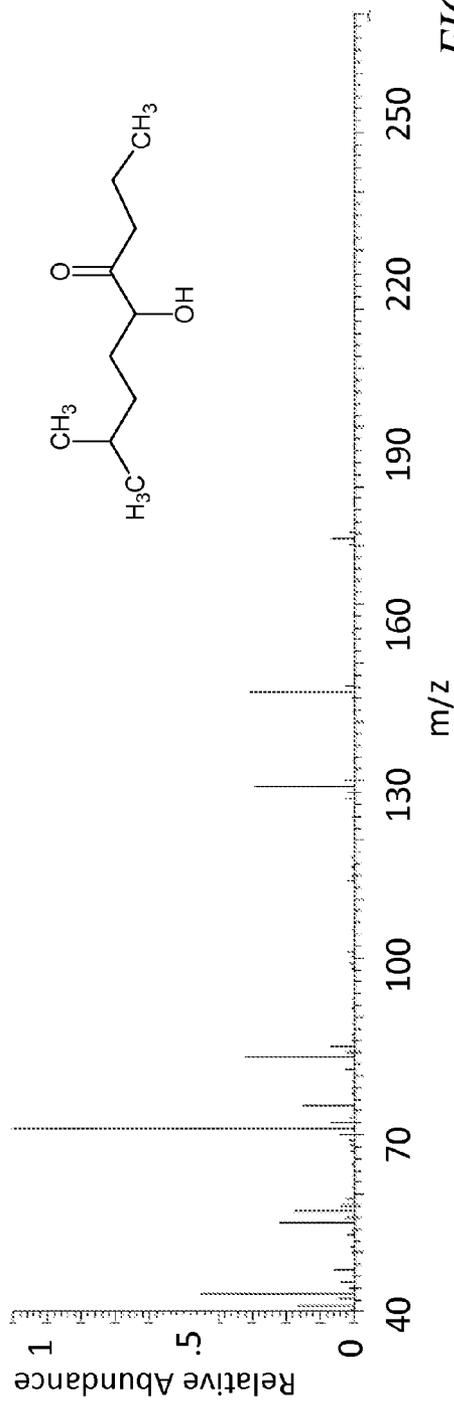
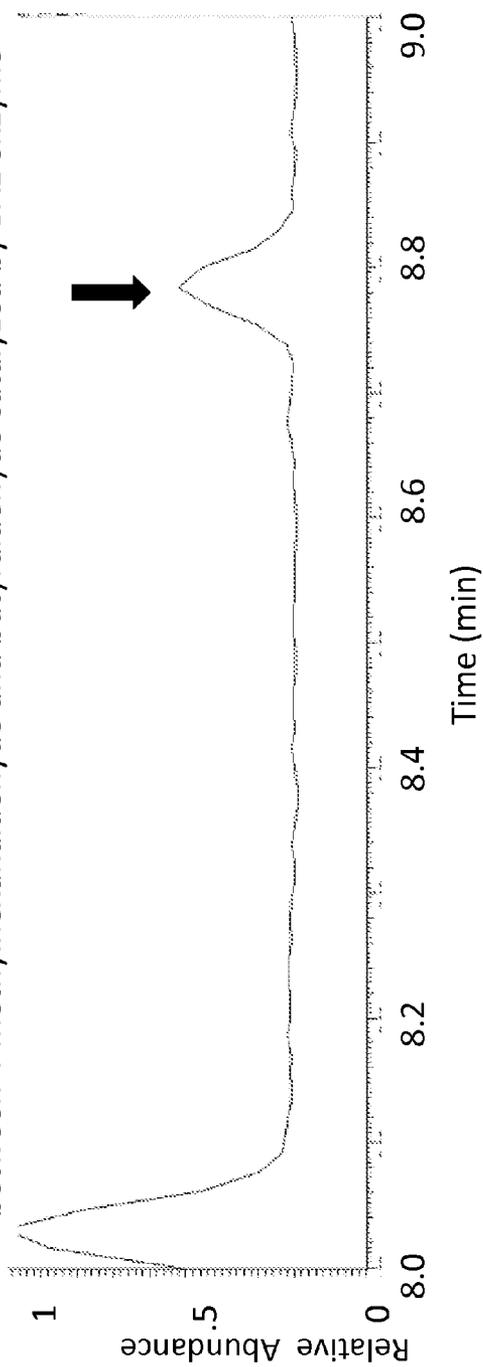


FIG. 52A

In vivo production 3-hydroxy-2-decanone from ligation reaction between acetaldehyde and octanal catalyzed by BAL

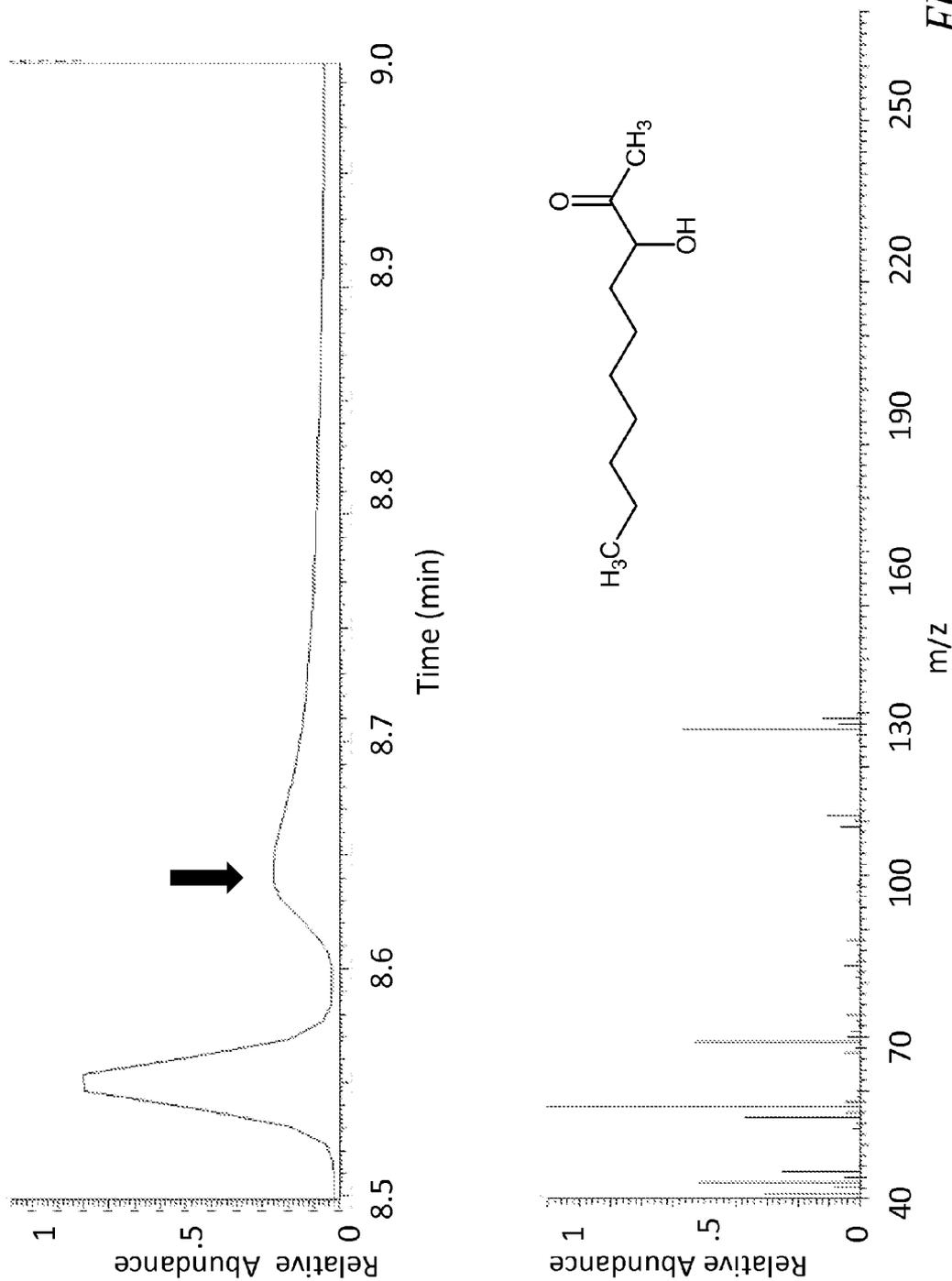


FIG. 52B

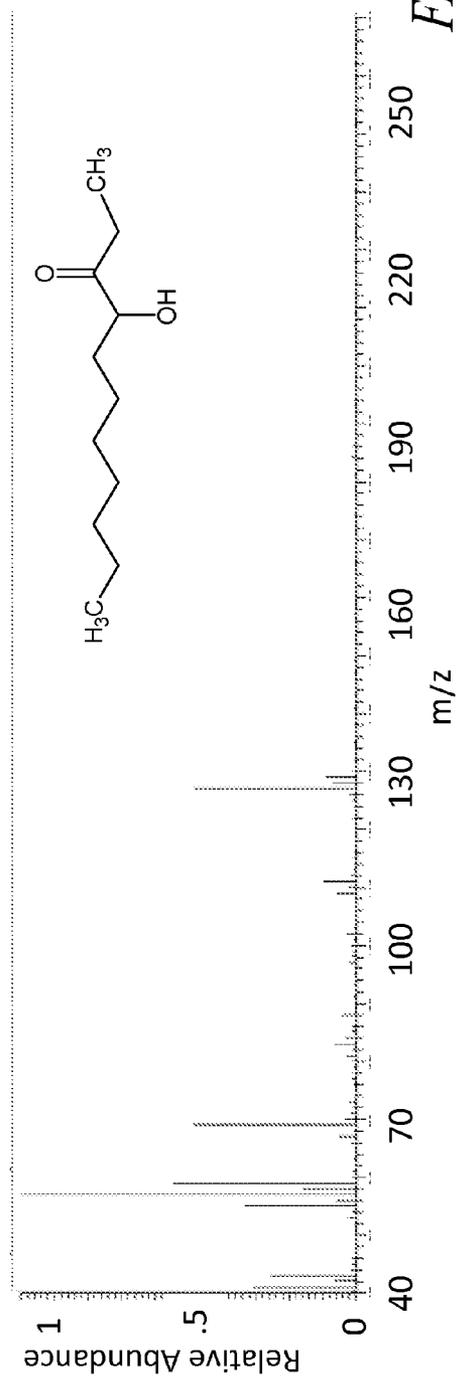
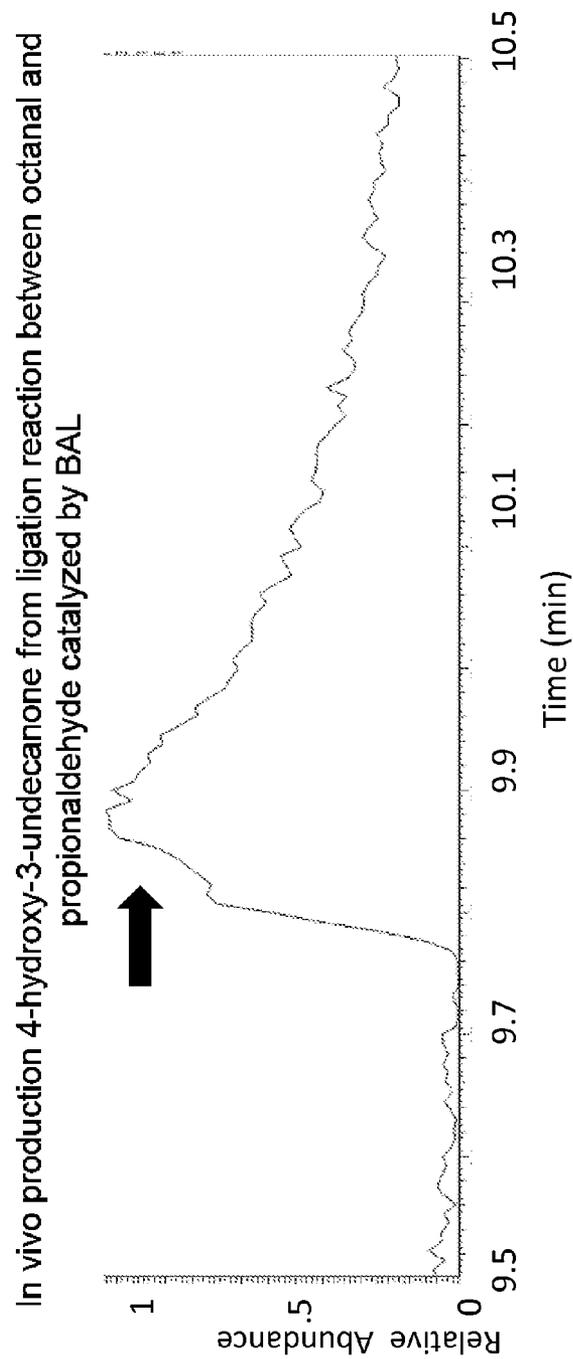


FIG. 53A

In vivo production 5-hydroxy-4-dodecanone from ligation reaction between octanal and butyraldehyde catalyzed by BAL

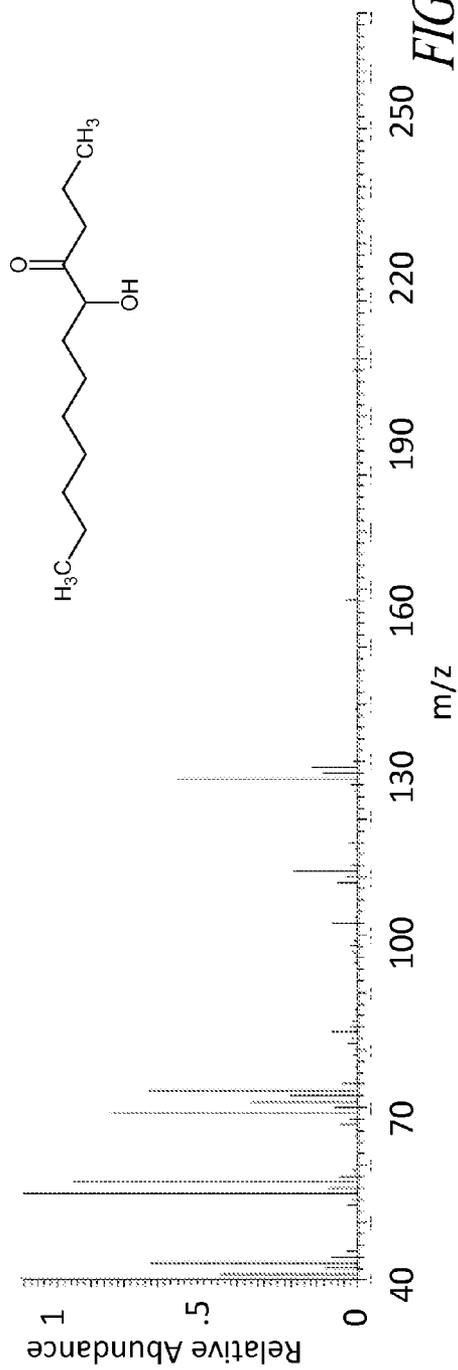
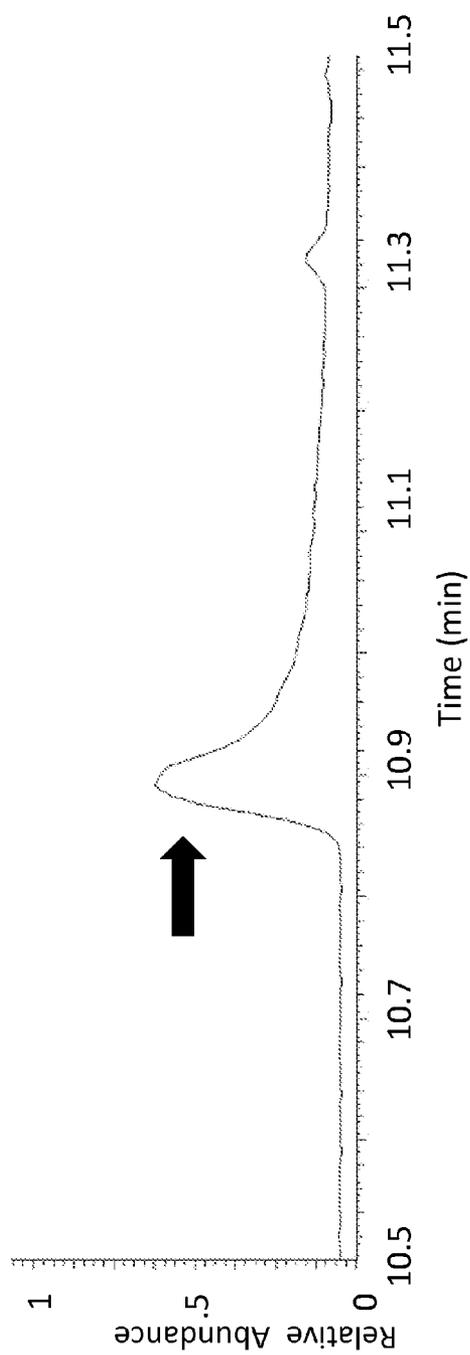


FIG. 53B

In vivo production of 6-hydroxy-5-tridecanone from ligation reaction between octanaldehyde and pentanaldehyde catalyzed by BAL enzyme

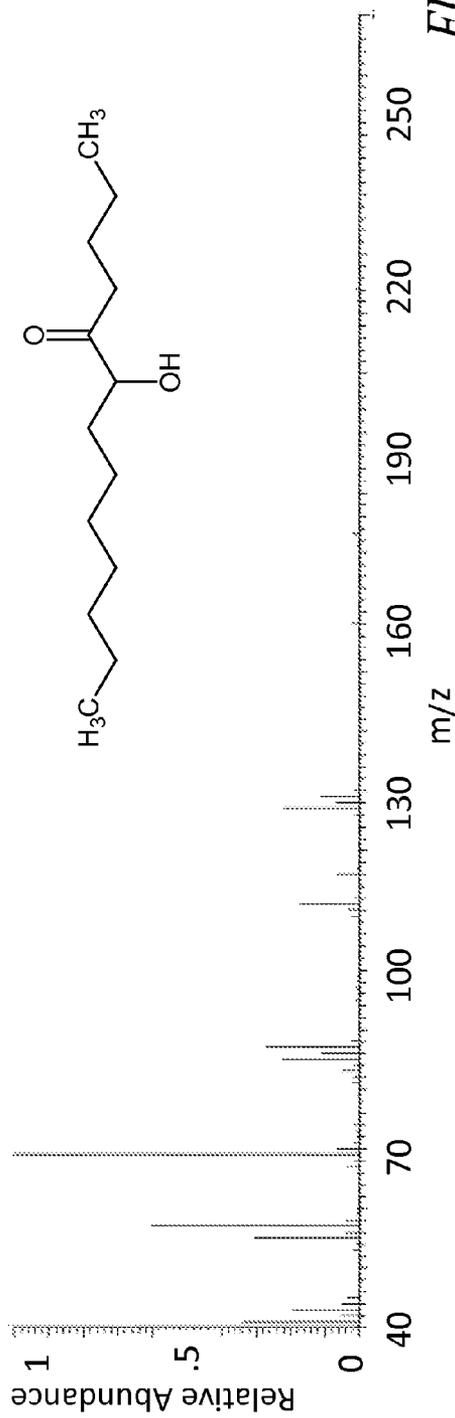
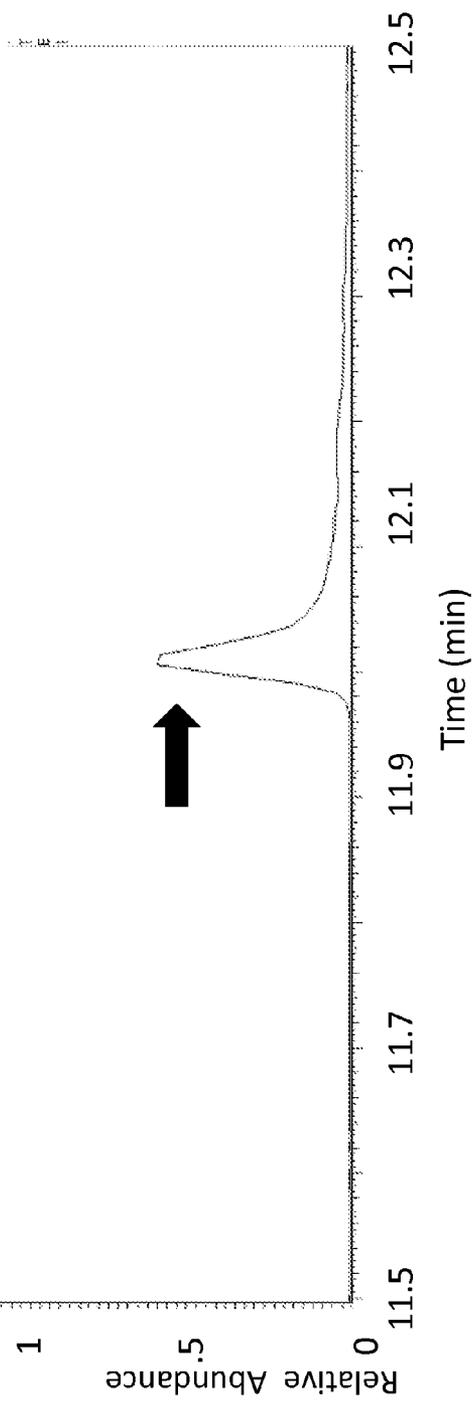


FIG. 54A

In vivo production 2-methyl-5-hydroxy-4-dodecanone and 2-methyl-4-hydroxy-5-decanone from ligation reaction between octanal and 3-methylbutyraldehyde catalyzed by BAL

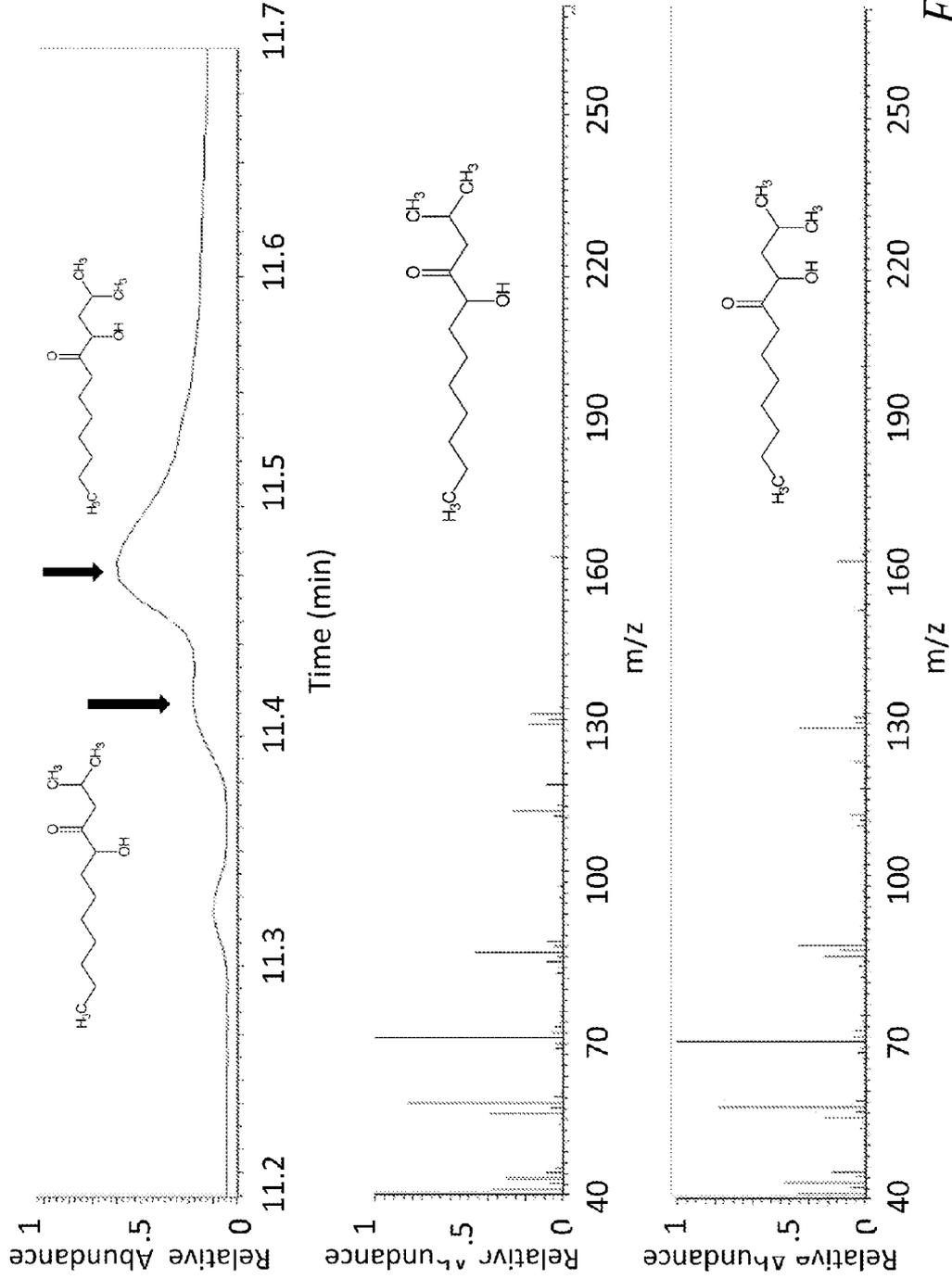


FIG. 54B

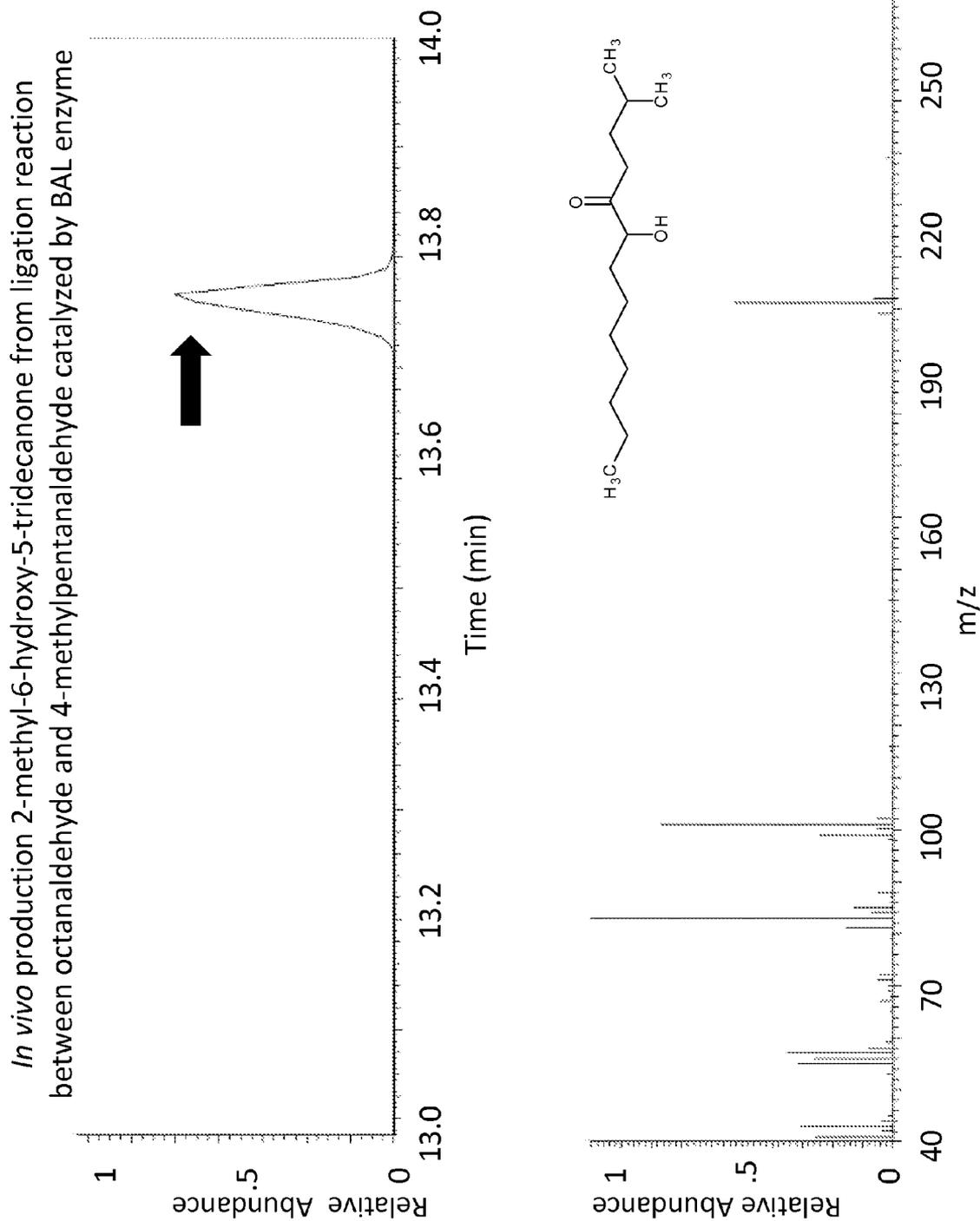


FIG. 55

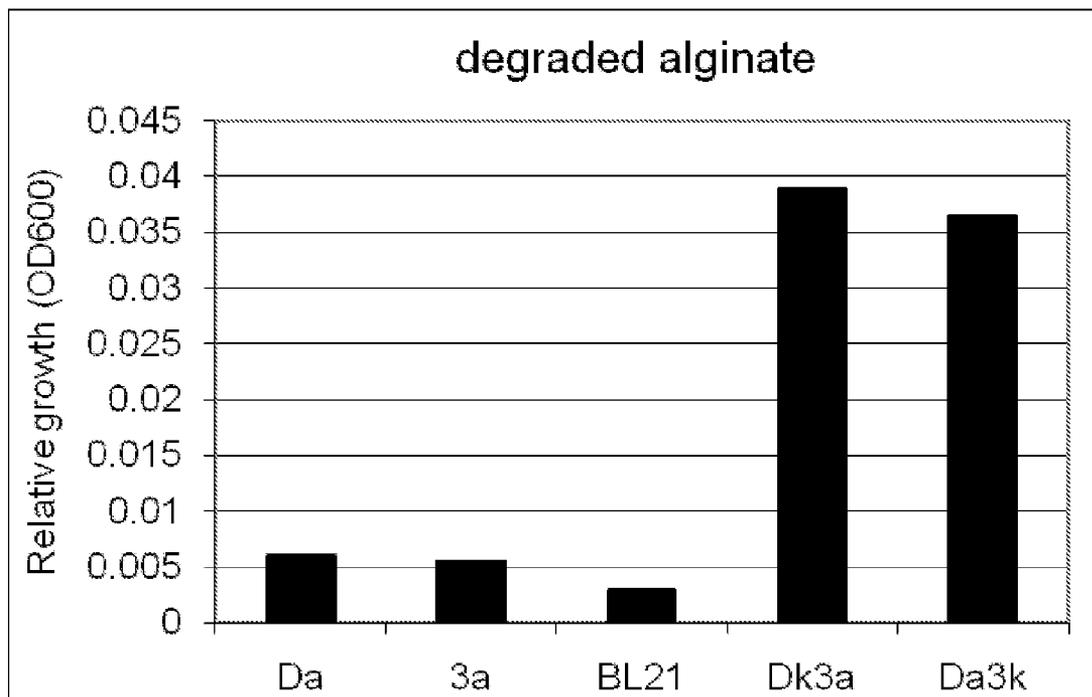


FIG. 56A

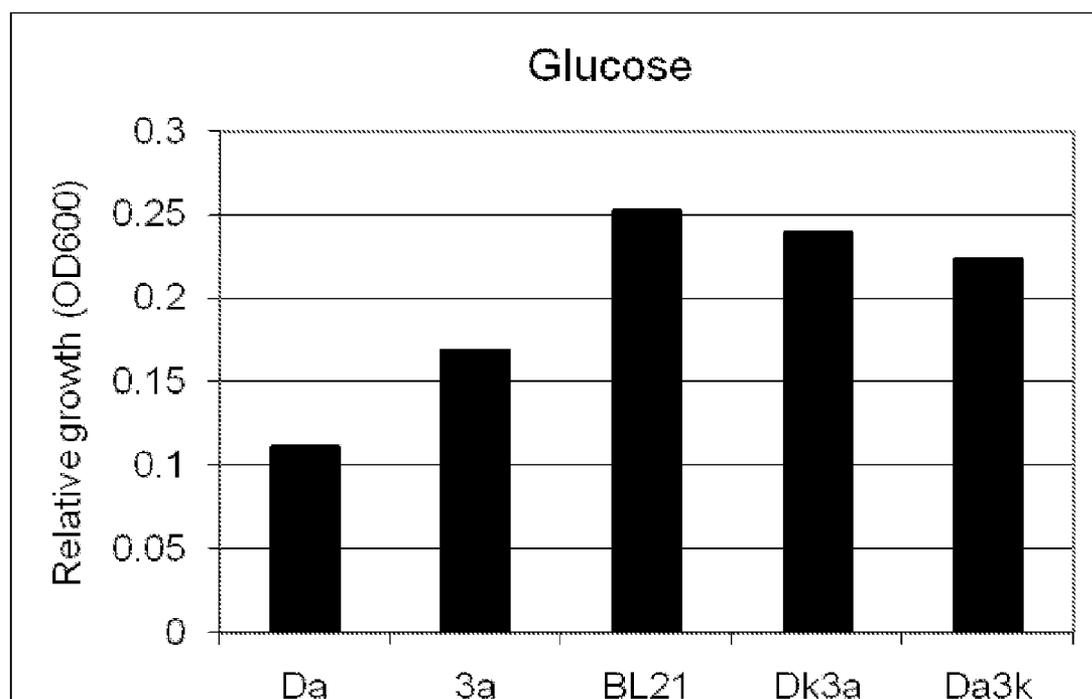
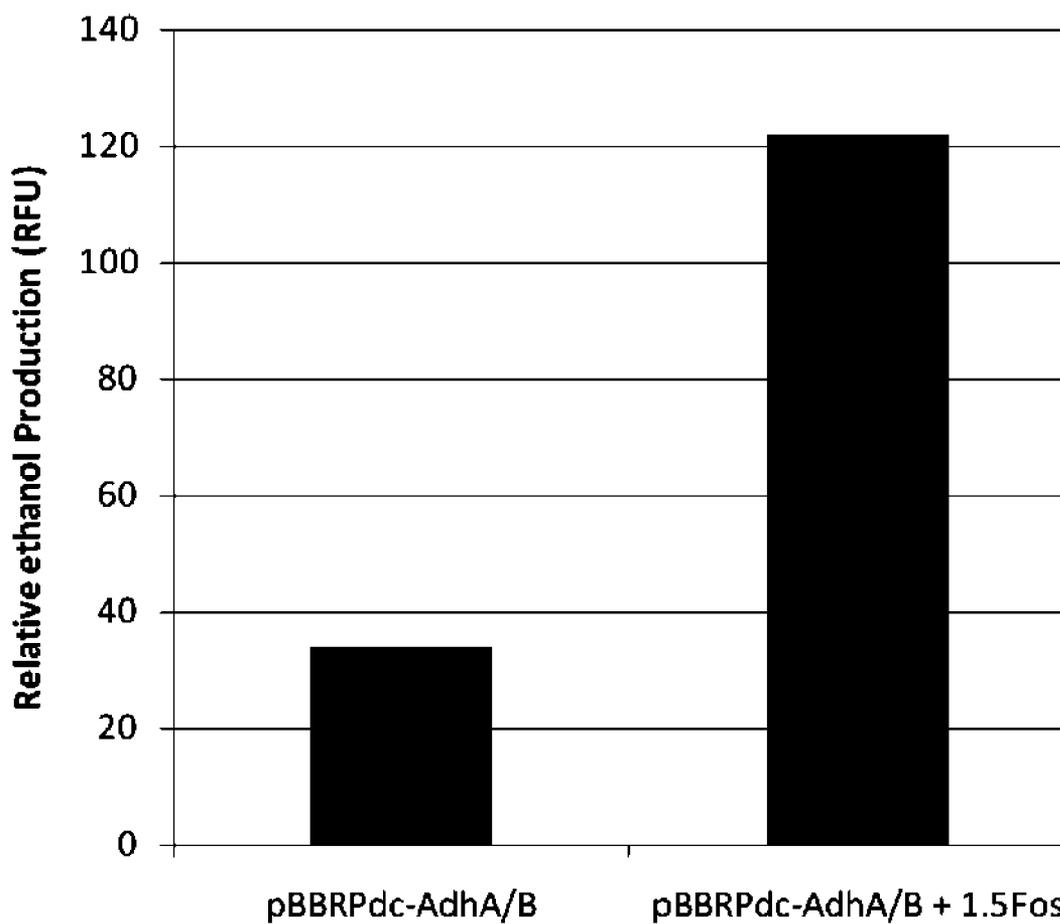


FIG. 56B

Figure 57
Production of Ethanol from Alginate



BIOFUEL PRODUCTION**CROSS-REFERENCE TO RELATED APPLICATION**

[0001] This application claims the benefit under 35 U.S.C. § 119(e) of U.S. Provisional Patent Application No. 60/977,628 filed Oct. 4, 2007, which application is incorporated herein by reference in its entirety.

STATEMENT REGARDING SEQUENCE LISTING

[0002] The Sequence Listing associated with this application is provided in text format in lieu of a paper copy, and is hereby incorporated by reference into the specification. The name of the text file containing the Sequence Listing is 150097_40101_SEQUENCE_LISTING.txt. The text file is 519 KB, was created on Oct. 3, 2008, and is being submitted electronically via EFS-Web.

TECHNICAL FIELD

[0003] The present application relates generally to the use of microbial and chemical systems to convert biomass to commodity chemicals, such as biofuels/biopetrols.

BACKGROUND

[0004] Petroleum is facing declining global reserves and contributes to more than 30% of greenhouse gas emissions driving global warming. Annually 800 billion barrels of transportation fuel are consumed globally. Diesel and jet fuels account for greater than 50% of global transportation fuels.

[0005] Significant legislation has been passed, requiring fuel producers to cap or reduce the carbon emissions from the production and use of transportation fuels. Fuel producers are seeking substantially similar, low carbon fuels that can be blended and distributed through existing infrastructure (e.g., refineries, pipelines, tankers).

[0006] Due to increasing petroleum costs and reliance on petrochemical feedstocks, the chemicals industry is also looking for ways to improve margin and price stability, while reducing its environmental footprint. The chemicals industry is striving to develop greener products that are more energy, water, and CO₂ efficient than current products. Fuels produced from biological sources, such as biomass, represent one aspect of process.

[0007] Presents method for converting biomass into biofuels focus on the use of lignocellulosic biomass, and there are many problems associated with using this process. Large-scale cultivation of lignocellulosic biomass requires substantial amount of cultivated land, which can be only achieved by replacing food crop production with energy crop production, deforestation, and by recultivating currently uncultivated land. Other problems include a decrease in water availability and quality and an increase in the use of pesticides and fertilizers.

[0008] The degradation of lignocellulosic biomass using biological systems is a very difficult challenge due to its substantial mechanistic strength and the complex chemical components. Approximately thirty different enzymes are required to fully convert lignocellulose to monosaccharides. The only available alternate to this complex approach requires a substantial amount of heat, pressure, and strong

acids. The art therefore needs an economic and technically simple process for converting biomass into hydrocarbons for use as biofuels or biopetrols.

BRIEF DESCRIPTION OF THE DRAWINGS

[0009] FIG. 1 shows the *Vibrio splendidus* genomic region of the fosmid clone described in Example 1. Genes are indicated with orange arrows. Labels show the numerical gene indices and the predicted function of the proteins.

[0010] FIG. 2 illustrates the pathways involved in certain embodiment in which *E. coli* may be engineered to grow on alginate as a sole source of carbon.

[0011] FIG. 3 illustrates the pathways involved in certain embodiment in which *E. coli* may be engineered to grow on pectin as a sole source of carbon.

[0012] FIG. 4 shows the results of engineered or recombinant *E. coli* growing on alginate as a sole source of carbon (see solid circles). *Agrobacterium tumefaciens* cells provide a positive control (see hatched circles). The well to the immediate left of the of the *A. tumefaciens* positive control contains DH10B *E. coli* cells, which provide a negative control.

[0013] FIG. 5 shows the growth of recombinant strain of *E. coli* on galacturonates and pectin. FIG. 5A shows the growth of *E. coli* on various lengths of galacturonate after 24 hr. The recombinant strain in FIG. 5A is the *E. coli* BL21(DE3) strain harboring pTrlogl-kdgR+pBBRGal3P, and the control strain is the BL21(DE3) strain harboring pTrc99A+pBBR1MCS-2, as described in Example 2. FIG. 5B shows the growth of recombinant *E. coli* on pectin after 3-4 days. The recombinant strain in FIG. 5B is *E. coli* DH5a strain containing pPEL74 (Ctrl) and pPEL74 and pROU2, as described in Example 2.

[0014] FIG. 6 shows the degradation of alginate to form pyruvate. FIG. 6A illustrates a simplified metabolic pathway for alginate degradation and metabolism. FIG. 6B shows the results of in vitro degradation of alginate to form pyruvate by an enzymatic degradation route. FIG. 6C shows the results of in vitro degradation of alginate to form pyruvate by a chemical degradation route.

[0015] FIG. 7 shows the biological activity of various alcohol dehydrogenases isolated from *Agrobacterium tumefaciens* C58. FIG. 7A shows DEHU hydrogenase activity as monitored by NADPH consumption, and FIG. 7B shows mannuronate hydrogenase activity as monitored by NADPH consumption.

[0016] FIG. 8 shows the GC-MS chromatogram results for the control sample (FIG. 8A) and for isobutyraldehyde, 3-methylpentanol, and 2-methylpentanol production from pBAD-alsS-ilvCD-leuABCD2 and pTrcBALK (FIG. 8B).

[0017] FIG. 9 shows the GC-MS chromatogram results for the control sample (FIG. 9A) and for 4-hydroxyphenylethanol and indole-3-ethanol production from pBADtyrA-aroLAC-aroG-ktkA-aroBDE and pTrcBALK (FIG. 9B).

[0018] FIG. 10 shows the mass spectrometry results for isobutanol (FIG. 10A), 3-methylpentanol (FIG. 10B), and 2-methylpentanol (FIG. 10C).

[0019] FIG. 11 shows the mass spectrometry results for phenylethanol (FIG. 11A), 4-hydroxyphenylethanol (FIG. 11B), and indole-3-ethanol (FIG. 11C).

[0020] FIG. 12 shows the biological activity of diol dehydratases. FIG. 12A shows the reduction of butyoin by ddh1, ddh2, and ddh3 as monitored by NADH consumption. FIG. 12B shows the oxidation activity of ddh3 towards 1,2-cyclopentanediol and 1,2-cyclohexanediol as measured by NADH production.

[0021] FIG. 13 summarizes the results of kinetic studies for various substrates in the oxidation reactions catalyzed by the DDH polypeptides. These reactions were NAD⁺-dependent.

[0022] FIG. 14 shows the nucleotide sequence (FIG. 14A) (SEQ ID NO:97) and polypeptide sequence (FIG. 14B) (SEQ ID NO:98) of diol dehydrogenase DDH1 isolated from *Lactobacillus brevis* ATCC 367.

[0023] FIG. 15 shows the nucleotide sequence (FIG. 15A) (SEQ ID NO:99) and polypeptide sequence (FIG. 15B) (SEQ ID NO:100) of diol dehydrogenase DDH2 isolated from *Pseudomonas putida* KT2440.

[0024] FIG. 16 shows the nucleotide sequence (FIG. 16A) (SEQ ID NO:101) and polypeptide sequence (FIG. 16B) (SEQ ID NO:102) of diol dehydrogenase DDH3 isolated from *Klebsiella pneumoniae* MGH78578.

[0025] FIG. 17 shows the sequential in vivo biological activity of a benzaldehyde lyase (bal) gene isolated from *Pseudomonas fluorescens* (codon usage was optimized for *E. coli* protein expression) and a ddh gene isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 (DDH3). This reaction illustrates the sequential conversion of butanal into 5-hydroxy-4-octanone and then 4,5-octanediol. FIG. 17A shows the detection of butyrolin (5-hydroxy-4-octanone) at 5.36 minutes, and FIG. 17B shows the detection of 4,5-octanediol at 6.49 and 6.65 minutes.

[0026] FIG. 18 shows the sequential in vivo biological activity of a benzaldehyde lyase (bal) gene isolated from *Pseudomonas fluorescens* (codon usage was optimized for *E. coli* protein expression) and a ddh gene isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 (DDH3). This Figure illustrates the sequential conversion of n-pentanal into 6-hydroxy-5-decanone and then 5,6-decanediol. FIG. 18A shows the detection of valeroin (6-hydroxy-5-decanone) at 8.22 minutes, and FIG. 18B shows the detection of 5,6 decanediol at 9.22 and 9.35 minutes.

[0027] FIG. 19 shows the sequential in vivo biological activity of a benzaldehyde lyase (bal) gene isolated from *Pseudomonas fluorescens* (codon usage was optimized for *E. coli* protein expression) and a ddh gene isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 (DDH3). This Figure illustrates the sequential conversion of 3-methylbutanal into 2,7-dimethyl-5-hydroxy-4-octanone and then 2,7-dimethyl-4,5-octanediol. FIG. 19A shows the detection of isoveraloin (2,7-dimethyl-5-hydroxy-4-octanone) at 6.79 minutes, and FIG. 19B shows the detection of 2,7-dimethyl-4,5-octanediol at 7.95 and 8.15 minutes.

[0028] FIG. 20 shows the sequential in vivo biological activity of a benzaldehyde lyase (bal) gene isolated from *Pseudomonas fluorescens* (codon usage was optimized for *E. coli* protein expression) and a ddh gene isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 (DDH3). This Figure illustrates the sequential conversion of n-hexanal into 7-hydroxy-6-dodecanone and then 6,7-dodecanediol. FIG. 20A shows the detection of hexanoin (7-hydroxy-6-decanone) at 10.42 minutes, and FIG. 20B shows the detection of 6,7 dodecanediol at 10.89 and 10.95 minutes.

[0029] FIG. 21 shows the sequential in vivo biological activity of a benzaldehyde lyase (bal) gene isolated from *Pseudomonas fluorescens* (codon usage was optimized for *E. coli* protein expression) and a ddh gene isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 (DDH3). This Figure illustrates the sequential conversion of 4-methylpentanal into 2,9-dimethyl-6-hydroxy-5-decanone and then 2,9-dimethyl-5,6-decanediol. FIG. 21A shows the detec-

tion of isohexanoin (2,9-Dimethyl-6-hydroxy-5-decanone) at 9.45 minutes, and FIG. 21B shows the detection of 2,9-dimethyl-5,6-decanediol at 10.38 and 10.44 minutes.

[0030] FIG. 22 shows the in vivo biological activity of a benzaldehyde lyase (bal) gene isolated from *Pseudomonas fluorescens* (codon usage was optimized for *E. coli* protein expression) and a ddh gene isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 (DDH3). This Figure illustrates the conversion of n-octanal into 9-hydroxy-8-hexadecanone by showing the detection of octanoin (9-hydroxy-8-hexadecanone) at 12.35 minutes.

[0031] FIG. 23 shows the in vivo biological activity of a benzaldehyde lyase (bal) gene isolated from *Pseudomonas fluorescens* (codon usage was optimized for *E. coli* protein expression) and a ddh gene isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 (DDH3). This Figure illustrates the conversion of acetaldehyde into 3-hydroxy-2-butanone by showing the detection of acetoin (3-hydroxy-2-butanone) at rt=0.91 minutes.

[0032] FIG. 24 shows the sequential in vivo biological activity of a benzaldehyde lyase (bal) gene isolated from *Pseudomonas fluorescens* (codon usage was optimized for *E. coli* protein expression) and a ddh gene isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 (DDH3). This Figure illustrates the sequential conversion of n-propional into 4-hydroxy-3-hexanone and then 3,4-hexanediol. FIG. 24A shows the detection of propioin (4-hydroxy-3-hexanone) at rt=2.62 minutes, and FIG. 24B shows the detection of 3,4-hexanediol at rt=3.79 minutes.

[0033] FIG. 25 the in vivo biological activity of a benzaldehyde lyase (bal) gene isolated from *Pseudomonas fluorescens* (codon usage was optimized for *E. coli* protein expression) and a ddh gene isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 (DDH3). This Figure illustrates the conversion of phenylacetaldehyde into 1,4-diphenyl-3-hydroxy-2-butanone by showing the detection of 1,4-diphenyl-3-hydroxy-2-butanone at rt=13.66 minutes.

[0034] FIG. 26 shows the sequential biological activity of a diol dehydrogenase ddh from *Klebsiella pneumoniae* MGH 78578 (DDH3) and a diol dehydratase pduCDE from *Klebsiella pneumoniae* MGH 78578. FIG. 26A shows GC-MS data which confirms the presence of 4,5-octanediol in the sample extraction, which is the expected product resulting from the reduction of butyrolin by ddh3. FIG. 26B shows GC-MS data confirming the presence of 4-octanone in the sample extraction, which is the expected product resulting from the sequential dehydrogenation of butyrolin and dehydration of 4,5-octanediol by ddh3 and pduCDE, respectively.

[0035] FIG. 27 shows the sequential biological activity of a diol dehydrogenase ddh from *Klebsiella pneumoniae* MGH 78578 (DDH3) and a diol dehydratase pduCDE from *Klebsiella pneumoniae* MGH 78578. FIGS. 27A and 27B show comparisons between the sample extraction gas chromatograph/mass spectrum and the 4-octanone standard gas chromatograph/mass spectrum, confirming that 4-octanone was produced from butyrolin using the enzymes diol dehydrogenase (ddh3) and a diol dehydratase (pduCDE).

[0036] FIG. 28 shows the nucleotide sequence (FIG. 28A) (SEQ ID NO:103) and polypeptide sequence (FIG. 28B) (SEQ ID NO:104) of a diol dehydratase large subunit (pduC) isolated from *Klebsiella pneumoniae* MGH78578.

[0037] FIG. 29 shows the nucleotide sequence (FIG. 29A) (SEQ ID NO:105) and polypeptide sequence (FIG. 29B) (SEQ ID NO:106) of a diol dehydratase medium subunit

isolated from *Klebsiella pneumoniae* MGH78578 (pduD), in addition to the nucleotide sequence (FIG. 29C) (SEQ ID NO:107) and polypeptide sequence (FIG. 29D) (SEQ ID NO:108) of a diol dehydratase small subunit isolated from *Klebsiella pneumoniae* MGH78578 (pduE).

[0038] FIG. 30 shows the oxidation of 4-octanol by secondary alcohol dehydrogenases as monitored by NADH production (FIG. 30A) and NADPH production (FIG. 30B).

[0039] FIG. 31 shows the oxidation of 4-octanol by secondary alcohol dehydrogenases as monitored by NADH production (FIG. 31A) and NADPH production (FIG. 31B).

[0040] FIG. 32 shows the oxidation of 2,7-dimethyl octanol by secondary alcohol dehydrogenases as monitored by NADH production (FIG. 32A) and NADPH production (FIG. 32B).

[0041] FIG. 33 shows the oxidation and reduction activity of 2ADH11 and 2ADH16. FIG. 33A shows the reduction of 2,7-dimethyl-4-octanone as measured by NADPH consumption. FIG. 33B shows the reduction of 2,7-dimethyl-4-octanone, 4-octanone, and cyclopentanone.

[0042] FIG. 34 shows the oxidation and reduction of cyclopentanol by secondary alcohol dehydrogenases. FIG. 34A shows the oxidation of cyclopentanol as monitored by NADH or NADPH formation. FIG. 34B shows the reduction of cyclopentanol as monitored by NADPH consumption.

[0043] FIG. 35 shows the calculated rate constants for the illustrated reduction reactions for each substrate catalyzed by secondary alcohol dehydrogenase ADH-16 (SEQ ID NO:138).

[0044] FIG. 36 shows the calculated rate constants for the illustrated oxidation reactions for each substrate catalyzed by secondary alcohol dehydrogenase ADH-16 (SEQ ID NO:138).

[0045] FIG. 37 shows a list of alginate lyases genes/proteins that may be utilized according to the methods and recombinant microorganisms described herein.

[0046] FIG. 38 shows a list of pectate lyase genes/proteins that may be utilized according to the methods and recombinant microorganisms described herein.

[0047] FIG. 39A shows a list of rhamnogalacturonan lyase genes/proteins that may be utilized according to the methods and recombinant microorganisms described herein. FIG. 39B shows a list of rhamnogalacturonate hydrolase genes/proteins that may be utilized according to the methods and recombinant microorganisms described herein.

[0048] FIG. 40 shows a list of pectin methyl esterase genes/proteins that may be utilized according to the methods and recombinant microorganisms described herein.

[0049] FIG. 41 shows a list of pectin acetyl esterase genes/proteins that may be utilized according to the methods and recombinant microorganisms described herein.

[0050] FIG. 42 shows the production of 2-phenyl ethanol (FIG. 42A), 2-(4-hydroxyphenyl)ethanol (FIG. 42B), and 2-(indole-3-)ethanol (FIG. 42C) at 24 hours from the recombinant microorganisms described in Example 4, which comprise functional 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol, and 2-(indole-3-)ethanol biosynthesis pathways.

[0051] FIG. 43 shows the GC-MS chromatogram results that confirm the production of 2-phenyl ethanol (FIG. 43B) at one week from the recombinant microorganisms described in Example 4 (pBADpheA-aroLAC-aroG-tktA-aroBDE and pTrcBALK). FIG. 43A shows the negative control cells (pBAD33 and pTrc99A).

[0052] FIG. 44 shows the GC-MS chromatogram results that confirm the production of 2-(4-hydroxyphenyl)ethanol (9.36 min) and 2-(indole-3) ethanol (10.32 min) at one week from the recombinant microorganisms described in Example 4 (pBADtyrA-aroLAC-aroG-tktA-aroBDE and pTrcBALK).

[0053] FIG. 45 confirms both the formation of 1-propanal from 1,2-propanediol (FIG. 45A), and the formation of 2-butanone from meso-2,3-butanediol (FIG. 45B), both of which were catalyzed in vitro by an isolated B12 independent diol dehydratase, as described in Example 9.

[0054] FIG. 46A shows the in vivo production of 1-propanol from 1,2-propanediol. FIG. 46B shows the in vivo production of 2-butanol from meso-2,3 butanediol. FIG. 46C shows the in vivo production of cyclopentanone from trans-1,2-cyclopentanediol. These experiments were performed as described in Example 9.

[0055] FIG. 47 shows the results of the TBA assay, as performed in Example 10. The left tube in FIG. 47 represents media taken from an overnight culture of cells expressing Vs24254, showing secretion of an alginate lyase, while the right hand tube shows the TBA reaction using media from cells expressing Vs24259 (negative control). The lack of pink coloration in the negative control indicates that little or no cleavage of the alginate polymer has occurred.

[0056] FIG. 48 shows the in vivo biological activity of a C—C ligase isolated from *Pseudomonas fluorescens* and cloned into *E. coli*. The GC-MS chromatogram results show that codon-optimized benzaldehyde lyase (BAL) catalyzed the in vivo production of 3-hydroxy-2-pentanone and 2-hydroxy-3-pentanone from a ligation reaction between acetaldehyde and propionaldehyde (FIG. 48A), and catalyzed the in vivo production of 4-hydroxy-3-heptanone and 3-hydroxy-4-heptanone from a ligation reaction between propionaldehyde and butyraldehyde (FIG. 48B).

[0057] FIG. 49 shows the in vivo biological activity of a C—C ligase isolated from *Pseudomonas fluorescens* and cloned into *E. coli*. The GC-MS chromatogram results show that codon-optimized BAL catalyzed the in vivo production of 3-hydroxy-2-heptanone from a ligation reaction between acetaldehyde and pentanal (FIG. 49A), and catalyzed the in vivo production of 4-hydroxy-3-octanone and 3-hydroxy-4-octanone from a ligation reaction between pentanal and propionaldehyde (FIG. 49B).

[0058] FIG. 50 shows the in vivo biological activity of a C—C ligase isolated from *Pseudomonas fluorescens* and cloned into *E. coli*. The GC-MS chromatogram results show that codon-optimized BAL catalyzed the in vivo production of 5-hydroxy-4-nonanone from ligation reaction between butyraldehyde and pentanal (FIG. 50A), and catalyzed the in vivo production of 2-methyl-5-hydroxy-4-decanone and 2-methyl-4-hydroxy-5-decanone from ligation reaction between hexanal and 3-methylbutyraldehyde (FIG. 50B).

[0059] FIG. 51 shows the in vivo biological activity of a C—C ligase isolated from *Pseudomonas fluorescens* and cloned into *E. coli*. The GC-MS chromatogram results show that codon-optimized BAL catalyzed the in vivo production of 6-methyl-3-hydroxy-2-heptanone from ligation reaction between acetaldehyde and 4-methylhexanal (FIG. 51A), and catalyzed the in vivo production of 7-methyl-4-hydroxy-3-octanone from a ligation reaction between 4-methylhexanal and propionaldehyde (FIG. 51B).

[0060] FIG. 52 shows the in vivo biological activity of a C—C ligase isolated from *Pseudomonas fluorescens* and cloned into *E. coli*. The GC-MS chromatogram results show

that codon-optimized BAL catalyzed the in vivo production of 8-methyl-5-hydroxy-4-nonanone from ligation reaction between 4-methylhexanal and butyraldehyde (FIG. 52A), and catalyzed the in vivo production of 3-hydroxy-2-decanone from a ligation reaction between acetaldehyde and octanal (FIG. 52B).

[0061] FIG. 53 shows the in vivo biological activity of a C—C ligase isolated from *Pseudomonas fluorescens* and cloned into *E. coli*. The GC-MS chromatogram results show that codon-optimized BAL catalyzed the in vivo production of 4-hydroxy-3-undecanone from ligation reaction between octanal and propionaldehyde (FIG. 53A), and catalyzed the in vivo production of 5-hydroxy-4-dodecanone from a ligation reaction between octanal and butyraldehyde (FIG. 53B).

[0062] FIG. 54 shows the in vivo biological activity of a C—C ligase isolated from *Pseudomonas fluorescens* and cloned into *E. coli*. The GC-MS chromatogram results show that codon-optimized BAL catalyzed the in vivo production of 6-hydroxy-5-tridecanone (FIG. 54A) from ligation reaction between octanal and pentanal, and catalyzed the in vivo production of 2-methyl-5-hydroxy-4-dodecanone and 2-methyl-4-hydroxy-5-decanone from a ligation reaction between octanal and 3-methylbutyraldehyde (FIG. 54B).

[0063] FIG. 55 shows the in vivo biological activity of a C—C ligase isolated from *Pseudomonas fluorescens* and cloned into *E. coli*. The GC-MS chromatogram results show that codon-optimized BAL catalyzed the in vivo production of 2-methyl-6-hydroxy-5-tridecanone from a ligation reaction between octanal and 4-methylpentanal.

[0064] FIG. 56 shows the growth of recombinant *E. coli* on alginate as a sole source of carbon (FIG. 56A), as described in Example 10. Growth on glucose (FIG. 56B) provides a positive control. The cells were transformed with either no plasmid (BL21—negative control), one plasmid (e.g., Da or 3a), or two plasmids (e.g., Dk3a and Da3k). The plasmids are indicated by the lower case letter: “a” refers to the pET-DEST42 plasmid backbone and “k” refers to the pENTR/D/TOPO backbone. “D” indicates that the plasmid contains the genomic region Vs24214-24249, while “3” indicates that the plasmid contains the genomic region Vs24189-24209. Thus, Da would be pET-DEST42-Vs24214-24249, Da3k would be pET-DEST42-Vs24214-24249 and pENTR/D/TOPO-Vs24189-24209 and so on. These results show that the combined genomic regions Vs24214-24249 and Vs24189-24209 are sufficient to confer on *E. coli* the ability to grow on alginate as a sole source of carbon.

[0065] FIG. 57 shows the production of ethanol by *E. coli* growing on alginate, as performed in Example 11. *E. coli* was transformed with either pBBRPdc-AdhA/B or pBBRPdc-AdhA/B+1.5 FOS and allowed to grow in m9 media containing alginate.

BRIEF SUMMARY

[0066] Embodiments of the present invention include methods for converting a polysaccharide to a commodity chemical, comprising (a) contacting the polysaccharide, wherein the polysaccharide is optionally derived from biomass, with a polysaccharide degrading or depolymerizing metabolic system, wherein the metabolic system is selected from; (i) enzymatic or chemical catalysis, and (ii) a microbial system, wherein the microbial system comprises a recombinant microorganism, wherein the recombinant microorganism comprises one or exogenous genes that allow it to grow on the polysaccharide as a sole source of carbon, thereby

converting the polysaccharide to a suitable monosaccharide or oligosaccharide; and (b) contacting the suitable monosaccharide or oligosaccharide with commodity chemical biosynthesis pathway, wherein the commodity chemical biosynthesis pathway comprises an aldehyde or ketone biosynthesis pathway, thereby converting the polysaccharide to the commodity chemical.

[0067] In certain aspects, the biomass is selected from marine biomass and vegetable/fruit/plant biomass. In certain aspects, the marine biomass is selected from kelp, giant kelp, sargasso, seaweed, algae, marine microflora, microalgae, and sea grass. In certain aspects, the vegetable/fruit/plant biomass comprises plant peel or pomace. In certain aspects, the vegetable/fruit/plant biomass is selected from citrus, potato, tomato, grape, gooseberry, carrot, mango, sugar-beet, apple, switchgrass, wood, and stover.

[0068] In certain aspects, the polysaccharide is selected from alginate, agar, carrageenan, fucoidan, pectin, polygalacturonate, cellulose, hemicellulose, xylan, arabinan, and mannan. In certain aspects, the suitable monosaccharide or oligosaccharide is selected from 2-keto-3-deoxy D-gluconate (KDG), D-mannitol, guluronate, mannuronate, mannitol, lyxose, glycerol, xylitol, glucose, mannose, galactose, xylose, arabinose, glucuronate, galacturonates, and rhamnose.

[0069] In certain aspects, the commodity chemical is selected from methane, methanol, ethane, ethene, ethanol, n-propane, 1-propene, 1-propanol, propanal, acetone, propionate, n-butane, 1-butene, 1-butanol, butanal, butanoate, isobutanol, isobutanol, 2-methylbutanal, 2-methylbutanol, 3-methylbutanal, 3-methylbutanol, 2-butene, 2-butanol, 2-butanone, 2,3-butanediol, 3-hydroxy-2-butanone, 2,3-butanedione, ethylbenzene, ethenylbenzene, 2-phenylethanol, phenylacetaldehyde, 1-phenylbutane, 4-phenyl-1-butene, 4-phenyl-2-butene, 1-phenyl-2-butene, 1-phenyl-2-butanol, 4-phenyl-2-butanol, 1-phenyl-2-butanone, 4-phenyl-2-butanone, 1-phenyl-2,3-butanediol, 1-phenyl-2,3-butanone, 4-phenyl-3-hydroxy-2-butanone, 1-phenyl-2,3-butanedione, n-pentane, ethylphenol, ethenylphenol, 2-(4-hydroxyphenyl)ethanol, 4-hydroxyphenylacetaldehyde, 1-(4-hydroxyphenyl)butane, 4-(4-hydroxyphenyl)-1-butene, 4-(4-hydroxyphenyl)-2-butene, 1-(4-hydroxyphenyl)-1-butene, 1-(4-hydroxyphenyl)-2-butanol, 4-(4-hydroxyphenyl)-2-butanol, 1-(4-hydroxyphenyl)-2-butanone, 4-(4-hydroxyphenyl)-2-butanone, 1-(4-hydroxyphenyl)-2,3-butanediol, 1-(4-hydroxyphenyl)-3-hydroxy-2-butanone, 4-(4-hydroxyphenyl)-3-hydroxy-2-butanone, 1-(4-hydroxyphenyl)-2,3-butanedione, indolyethane, indolyethene, 2-(indole-3-)ethanol, n-pentane, 1-pentene, 1-pentanol, pentanal, pentanoate, 2-pentene, 2-pentanol, 3-pentanol, 2-pentanone, 3-pentanone, 4-methylpentanal, 4-methylpentanol, 2,3-pentanediol, 2-hydroxy-3-pentanone, 3-hydroxy-2-pentanone, 2,3-pentanedione, 2-methylpentane, 4-methyl-1-pentene, 4-methyl-2-pentene, 4-methyl-3-pentene, 4-methyl-2-pentanol, 2-methyl-3-pentanol, 4-methyl-2-pentanone, 2-methyl-3-pentanone, 4-methyl-2,3-pentanediol, 4-methyl-2-hydroxy-3-pentanone, 4-methyl-3-hydroxy-2-pentanone, 4-methyl-2,3-pentanedione, 1-phenylpentane, 1-phenyl-1-pentene, 1-phenyl-2-pentene, 1-phenyl-3-pentene, 1-phenyl-2-pentanol, 1-phenyl-3-pentanol, 1-phenyl-2-pentanone, 1-phenyl-3-pentanone, 1-phenyl-2,3-pentanediol, 1-phenyl-2-hydroxy-3-pentanone, 1-phenyl-3-hydroxy-2-pentanone, 1-phenyl-2,3-pentanedione, 4-methyl-1-phenylpentane, 4-methyl-1-phenyl-1-pen-

tene, 4-methyl-1-phenyl-2-pentene, 4-methyl-1-phenyl-3-pentene, 4-methyl-1-phenyl-3-pentanol, 4-methyl-1-phenyl-2-pentanol, 4-methyl-1-phenyl-3-pentanone, 4-methyl-1-phenyl-2-pentanone, 4-methyl-1-phenyl-2,3-pentanediol, 4-methyl-1-phenyl-2,3-pentanedione, 4-methyl-1-phenyl-3-hydroxy-2-pentanone, 4-methyl-1-phenyl-2-hydroxy-3-pentanone, 1-(4-hydroxyphenyl) pentane, 1-(4-hydroxyphenyl)-1-pentene, 1-(4-hydroxyphenyl)-2-pentene, 1-(4-hydroxyphenyl)-3-pentene, 1-(4-hydroxyphenyl)-2-pentanol, 1-(4-hydroxyphenyl)-3-pentanol, 1-(4-hydroxyphenyl)-2-pentanone, 1-(4-hydroxyphenyl)-3-pentanone, 1-(4-hydroxyphenyl)-2,3-pentanedione, 1-(4-hydroxyphenyl)-2-hydroxy-3-pentanone, 1-(4-hydroxyphenyl)-3-hydroxy-2-pentanone, 1-(4-hydroxyphenyl)-2,3-pentanedione, 4-methyl-1-(4-hydroxyphenyl) pentane, 4-methyl-1-(4-hydroxyphenyl)-2-pentene, 4-methyl-1-(4-hydroxyphenyl)-3-pentene, 4-methyl-1-(4-hydroxyphenyl)-1-pentene, 4-methyl-1-(4-hydroxyphenyl)-3-pentanol, 4-methyl-1-(4-hydroxyphenyl)-2-pentanol, 4-methyl-1-(4-hydroxyphenyl)-3-pentanone, 4-methyl-1-(4-hydroxyphenyl)-2-pentanone, 4-methyl-1-(4-hydroxyphenyl)-2,3-pentanedione, 4-methyl-1-(4-hydroxyphenyl)-2,3-pentanedione, 4-methyl-1-(4-hydroxyphenyl)-3-hydroxy-2-pentanone, 4-methyl-1-(4-hydroxyphenyl)-2-hydroxy-3-pentanone, 1-indole-3-pentane, 1-(indole-3)-1-pentene, 1-(indole-3)-2-pentene, 1-(indole-3)-3-pentene, 1-(indole-3)-2-pentanol, 1-(indole-3)-3-pentanol, 1-(indole-3)-2-pentanone, 1-(indole-3)-3-pentanone, 1-(indole-3)-2,3-pentanedione, 1-(indole-3)-2-hydroxy-3-pentanone, 1-(indole-3)-3-hydroxy-2-pentanone, 1-(indole-3)-2,3-pentanedione, 4-methyl-1-(indole-3)pentane, 4-methyl-1-(indole-3)-2-pentene, 4-methyl-1-(indole-3)-3-pentene, 4-methyl-1-(indole-3)-1-pentene, 4-methyl-2-(indole-3)-3-pentanol, 4-methyl-1-(indole-3)-2-pentanol, 4-methyl-1-(indole-3)-3-pentanone, 4-methyl-1-(indole-3)-2-pentanone, 4-methyl-1-(indole-3)-2,3-pentanedione, 4-methyl-1-(indole-3)-3-hydroxy-2-pentanone, 4-methyl-1-(indole-3)-2-hydroxy-3-pentanone, n-hexane, 1-hexene, 1-hexanol, hexanal, hexanoate, 2-hexene, 3-hexene, 2-hexanol, 3-hexanol, 2-hexanone, 3-hexanone, 2,3-hexanediol, 2,3-hexanedione, 3,4-hexanediol, 3,4-hexanedione, 2-hydroxy-3-hexanone, 3-hydroxy-2-hexanone, 3-hydroxy-4-hexanone, 4-hydroxy-3-hexanone, 2-methylhexane, 3-methylhexane, 2-methyl-2-hexene, 2-methyl-3-hexene, 5-methyl-1-hexene, 5-methyl-2-hexene, 4-methyl-1-hexene, 4-methyl-2-hexene, 3-methyl-3-hexene, 3-methyl-2-hexene, 3-methyl-1-hexene, 2-methyl-3-hexanol, 5-methyl-2-hexanol, 5-methyl-3-hexanol, 2-methyl-3-hexanone, 5-methyl-2-hexanone, 5-methyl-3-hexanone, 2-methyl-3,4-hexanediol, 2-methyl-3,4-hexanedione, 5-methyl-2,3-hexanediol, 5-methyl-2,3-hexanedione, 4-methyl-2,3-hexanediol, 4-methyl-2,3-hexanedione, 2-methyl-3-hydroxy-4-hexanone, 2-methyl-4-hydroxy-3-hexanone, 5-methyl-2-hydroxy-3-hexanone, 5-methyl-3-hydroxy-2-hexanone, 4-methyl-2-hydroxy-3-hexanone, 4-methyl-3-hydroxy-2-hexanone, 2,5-dimethylhexane, 2,5-dimethyl-2-hexene, 2,5-dimethyl-3-hexene, 2,5-dimethyl-3-hexanol, 2,5-dimethyl-3-hexanone, 2,5-dimethyl-3,4-hexanediol, 2,5-dimethyl-3,4-hexanedione, 2,5-dimethyl-3-hydroxy-4-hexanone, 5-methyl-1-phenylhexane, 4-methyl-1-phenylhexane, 5-methyl-1-phenyl-1-hexene, 5-methyl-1-phenyl-2-hexene, 5-methyl-1-phenyl-3-hexene, 4-methyl-1-phenyl-1-hexene, 4-methyl-1-phenyl-2-hexene, 4-methyl-1-phenyl-3-hexene, 5-methyl-1-phenyl-2-hexanol, 5-methyl-

1-phenyl-3-hexanol, 4-methyl-1-phenyl-2-hexanol, 4-methyl-1-phenyl-3-hexanol, 5-methyl-1-phenyl-2-hexanone, 5-methyl-1-phenyl-3-hexanone, 4-methyl-1-phenyl-2-hexanone, 4-methyl-1-phenyl-3-hexanone, 5-methyl-1-phenyl-2,3-hexanediol, 4-methyl-1-phenyl-2,3-hexanediol, 5-methyl-1-phenyl-3-hydroxy-2-hexanone, 5-methyl-1-phenyl-2-hydroxy-3-hexanone, 4-methyl-1-phenyl-3-hydroxy-2-hexanone, 4-methyl-1-phenyl-2-hydroxy-3-hexanone, 5-methyl-1-phenyl-2,3-hexanedione, 4-methyl-1-phenyl-2,3-hexanedione, 4-methyl-1-(4-hydroxyphenyl)hexane, 5-methyl-1-(4-hydroxyphenyl)-1-hexene, 5-methyl-1-(4-hydroxyphenyl)-2-hexene, 5-methyl-1-(4-hydroxyphenyl)-3-hexene, 4-methyl-1-(4-hydroxyphenyl)-1-hexene, 4-methyl-1-(4-hydroxyphenyl)-2-hexene, 4-methyl-1-(4-hydroxyphenyl)-3-hexene, 5-methyl-1-(4-hydroxyphenyl)-2-hexanol, 5-methyl-1-(4-hydroxyphenyl)-3-hexanol, 4-methyl-1-(4-hydroxyphenyl)-2-hexanol, 4-methyl-1-(4-hydroxyphenyl)-3-hexanol, 5-methyl-1-(4-hydroxyphenyl)-2-hexanone, 5-methyl-1-(4-hydroxyphenyl)-3-hexanone, 4-methyl-1-(4-hydroxyphenyl)-2-hexanone, 4-methyl-1-(4-hydroxyphenyl)-3-hexanone, 5-methyl-1-(4-hydroxyphenyl)-2,3-hexanediol, 4-methyl-1-(4-hydroxyphenyl)-2,3-hexanediol, 5-methyl-1-(4-hydroxyphenyl)-3-hydroxy-2-hexanone, 5-methyl-1-(4-hydroxyphenyl)-2-hydroxy-3-hexanone, 4-methyl-1-(4-hydroxyphenyl)-3-hydroxy-2-hexanone, 4-methyl-1-(4-hydroxyphenyl)-2-hydroxy-3-hexanone, 5-methyl-1-(4-hydroxyphenyl)-2,3-hexanedione, 4-methyl-1-(indole-3)-1-hexene, 5-methyl-1-(indole-3)-2-hexene, 5-methyl-1-(indole-3)-3-hexene, 4-methyl-1-(indole-3)-1-hexene, 4-methyl-1-(indole-3)-2-hexene, 4-methyl-1-(indole-3)-3-hexene, 5-methyl-1-(indole-3)-2-hexanol, 5-methyl-1-(indole-3)-3-hexanol, 4-methyl-1-(indole-3)-2-hexanol, 4-methyl-1-(indole-3)-3-hexanol, 5-methyl-1-(indole-3)-2-hexanone, 5-methyl-1-(indole-3)-3-hexanone, 4-methyl-1-(indole-3)-2-hexanone, 5-methyl-1-(indole-3)-3-hexanone, 2,3-hexanediol, 4-methyl-1-(indole-3)-2,3-hexanediol, 4-methyl-1-(indole-3)-2,3-hexanedione, 5-methyl-1-(indole-3)-3-hydroxy-2-hexanone, 5-methyl-1-(indole-3)-2-hydroxy-3-hexanone, 4-methyl-1-(indole-3)-3-hydroxy-2-hexanone, 5-methyl-1-(indole-3)-2-hydroxy-3-hexanone, 5-methyl-1-(indole-3)-2,3-hexanedione, 4-methyl-1-(indole-3)-2,3-hexanedione, n-heptane, 1-heptene, 1-heptanol, heptanal, heptanoate, 2-heptene, 3-heptene, 2-heptanol, 3-heptanol, 4-heptanol, 2-heptanone, 3-heptanone, 4-heptanone, 2,3-heptanediol, 2,3-heptanedione, 3,4-heptanediol, 3,4-heptanedione, 2-hydroxy-3-heptanone, 3-hydroxy-2-heptanone, 3-hydroxy-4-heptanone, 4-hydroxy-3-heptanone, 2-methylheptane, 3-methylheptane, 6-methyl-2-heptene, 6-methyl-3-heptene, 2-methyl-3-heptene, 2-methyl-2-heptene, 5-methyl-2-heptene, 5-methyl-3-heptene, 3-methyl-3-heptene, 2-methyl-3-heptanol, 2-methyl-4-heptanol, 6-methyl-3-heptanol, 5-methyl-3-heptanol, 3-methyl-4-heptanol, 2-methyl-3-heptanone, 2-methyl-4-heptanone, 6-methyl-3-heptanone, 5-methyl-3-heptanone, 3-methyl-4-heptanone, 2-methyl-3,4-heptanediol, 2-methyl-3,4-heptanedione, 6-methyl-3,4-heptanediol, 6-methyl-3,4-heptanedione, 5-methyl-3,4-heptanediol, 5-methyl-3,4-heptanedione, 2-methyl-3-hydroxy-4-heptanone, 2-methyl-4-hydroxy-3-heptanone, 6-methyl-3-hydroxy-4-heptanone, 6-methyl-4-hydroxy-3-heptanone, 5-methyl-3-hydroxy-4-heptanone, 5-methyl-4-hydroxy-3-heptanone, 2,6-dimethylheptane, 2,5-dimethylheptane, 2,6-dimethyl-2-heptene, 2,6-dimethyl-3-heptene, 2,5-dimethyl-2-heptene, 2,5-dimethyl-

3-heptene, 3,6-dimethyl-3-heptene, 2,6-dimethyl-3-heptanol, 2,6-dimethyl-4-heptanol, 2,5-dimethyl-3-heptanol, 2,5-dimethyl-4-heptanol, 2,6-dimethyl-3,4-heptanediol, 2,6-dimethyl-3,4-heptanedione, 2,5-dimethyl-3,4-heptanediol, 2,5-dimethyl-3,4-heptanedione, 2,6-dimethyl-3-hydroxy-4-heptanone, 2,6-dimethyl-4-hydroxy-3-heptanone, 2,5-dimethyl-3-hydroxy-4-heptanone, 2,5-dimethyl-4-hydroxy-3-heptanone, n-octane, 1-octene, 2-octene, 1-octanol, octanal, octanoate, 3-octene, 4-octene, 4-octanol, 4-octanone, 4,5-octanediol, 4,5-octanedione, 4-hydroxy-5-octanone, 2-methyl-3-octene, 2-methyl-4-octene, 7-methyl-3-octene, 3-methyl-3-octene, 3-methyl-4-octene, 6-methyl-3-octene, 2-methyl-4-octanol, 7-methyl-4-octanol, 3-methyl-4-octanol, 6-methyl-4-octanol, 2-methyl-4-octanone, 7-methyl-4-octanone, 3-methyl-4-octanone, 6-methyl-4-octanone, 2-methyl-4,5-octanediol, 2-methyl-4,5-octanedione, 3-methyl-4,5-octanediol, 3-methyl-4,5-octanedione, 2-methyl-4-hydroxy-5-octanone, 2-methyl-5-hydroxy-4-octanone, 3-methyl-4-hydroxy-5-octanone, 3-methyl-5-hydroxy-4-octanone, 2,7-dimethyloctane, 2,7-dimethyl-3-octene, 2,7-dimethyl-4-octene, 2,7-dimethyl-4-octanol, 2,7-dimethyl-4-octanone, 2,7-dimethyl-4,5-octanediol, 2,7-dimethyl-4,5-octanedione, 2,7-dimethyl-4-hydroxy-5-octanone, 2,6-dimethyloctane, 2,6-dimethyl-3-octene, 2,6-dimethyl-4-octene, 3,7-dimethyl-3-octene, 2,6-dimethyl-4-octanol, 3,7-dimethyl-4-octanol, 2,6-dimethyl-4-octanone, 3,7-dimethyl-4-octanone, 2,6-dimethyl-4,5-octanediol, 2,6-dimethyl-4,5-octanedione, 2,6-dimethyl-4-hydroxy-5-octanone, 2,6-dimethyl-5-hydroxy-4-octanone, 3,6-dimethyloctane, 3,6-dimethyl-3-octene, 3,6-dimethyl-4-octene, 3,6-dimethyl-4-octanol, 3,6-dimethyl-4-octanone, 3,6-dimethyl-4,5-octanediol, 3,6-dimethyl-4,5-octanedione, 3,6-dimethyl-4-hydroxy-5-octanone, n-nonane, 1-nonene, 1-nonanol, nonanal, nonanoate, 2-methylnonane, 2-methyl-4-nonene, 2-methyl-5-nonene, 8-methyl-4-nonene, 2-methyl-5-nonanol, 8-methyl-4-nonanol, 2-methyl-5-nonanone, 8-methyl-4-nonanone, 8-methyl-4,5-nonanediol, 8-methyl-4,5-nonanedione, 8-methyl-4-hydroxy-5-nonanone, 8-methyl-5-hydroxy-4-nonanone, 2,8-dimethylnonane, 2,8-dimethyl-3-nonene, 2,8-dimethyl-4-nonene, 2,8-dimethyl-5-nonene, 2,8-dimethyl-4-nonanol, 2,8-dimethyl-5-nonanol, 2,8-dimethyl-4-nonanone, 2,8-dimethyl-5-nonanone, 2,8-dimethyl-4,5-nonanediol, 2,8-dimethyl-4,5-nonanedione, 2,8-dimethyl-4-hydroxy-5-nonanone, 2,8-dimethyl-5-hydroxy-4-nonanone, 2,7-dimethylnonane, 3,8-dimethyl-3-nonene, 3,8-dimethyl-4-nonene, 3,8-dimethyl-5-nonene, 3,8-dimethyl-4-nonanol, 3,8-dimethyl-5-nonanol, 3,8-dimethyl-4-nonanone, 3,8-dimethyl-5-nonanone, 3,8-dimethyl-4,5-nonanediol, 3,8-dimethyl-4,5-nonanedione, 3,8-dimethyl-4-hydroxy-5-nonanone, 3,8-dimethyl-5-hydroxy-4-nonanone, n-decane, 1-decene, 1-decanol, decanoate, 2,9-dimethyldecane, 2,9-dimethyl-3-decene, 2,9-dimethyl-4-decene, 2,9-dimethyl-5-decanol, 2,9-dimethyl-5-decanone, 2,9-dimethyl-5,6-decanediol, 2,9-dimethyl-6-hydroxy-5-decanone, 2,9-dimethyl-5,6-decanedione, 1-undecene, 1-undecanol, undecanal, undecanoate, n-dodecane, 1-dodecene, 1-dodecanol, dodecanal, dodecanoate, n-dodecane, 1-decdecene, 1-dodecanol, ddodecanal, dodecanoate, n-tridecane, 1-tridecene, 1-tridecanol, tridecanal, tridecanoate, n-tetradecane, 1-tetradecene, 1-tetradecanol, tetradecanal, tetradecanoate, n-pentadecane, 1-pentadecene, 1-pentadecanol, pentadecanal, pentadecanoate, n-hexadecane, 1-hexadecene, 1-hexadecanol, hexadecanal, hexadecanoate, n-heptadecane, 1-heptadecene, 1-heptadecanol,

heptadecanal, heptadecanoate, n-octadecane, 1-octadecene, 1-octadecanol, octadecanal, octadecanoate, n-nonadecane, 1-nonadecene, 1-nonadecanol, nonadecanal, nonadecanoate, eicosane, 1-eicosene, 1-eicosanol, eicosanal, eicosanoate, 3-hydroxy propanal, 1,3-propanediol, 4-hydroxybutanal, 1,4-butanediol, 3-hydroxy-2-butanone, 2,3-butanediol, 1,5-pentane diol, homocitrate, homoisocitrate, b-hydroxy adipate, glutarate, glutarsemialdehyde, glutaraldehyde, 2-hydroxy-1-cyclopentanone, 1,2-cyclopentanediol, cyclopentanone, cyclopentanol, (S)-2-acetolactate, (R)-2,3-Dihydroxy-isovalerate, 2-oxoisovalerate, isobutyryl-CoA, isobutyrate, isobutyraldehyde, 5-amino pentaldehyde, 1,10-diaminododecane, 1,10-diamino-5-decene, 1,10-diamino-5-hydroxydecane, 1,10-diamino-5-decanone, 1,10-diamino-5,6-decanediol, 1,10-diamino-6-hydroxy-5-decanone, phenylacetaldehyde, 1,4-diphenylbutane, 1,4-diphenyl-1-butene, 1,4-diphenyl-2-butene, 1,4-diphenyl-2-butanol, 1,4-diphenyl-2-butanone, 1,4-diphenyl-2,3-butanediol, 1,4-diphenyl-3-hydroxy-2-butanone, 1-(4-hydroxyphenyl)-4-phenylbutane, 1-(4-hydroxyphenyl)-4-phenyl-1-butene, 1-(4-hydroxyphenyl)-4-phenyl-2-butene, 1-(4-hydroxyphenyl)-4-phenyl-2-butanol, 1-(4-hydroxyphenyl)-4-phenyl-2-butanone, 1-(4-hydroxyphenyl)-4-phenyl-2,3-butanediol, 1-(4-hydroxyphenyl)-4-phenyl-3-butanediol, 1-(indole-3)-4-phenylbutane, 1-(indole-3)-4-phenyl-1-butene, 1-(indole-3)-4-phenyl-2-butene, 1-(indole-3)-4-phenyl-2-butanol, 1-(indole-3)-4-phenyl-2-butanone, 1-(indole-3)-4-phenyl-2,3-butanediol, 1-(indole-3)-4-phenyl-3-hydroxy-2-butanone, 4-hydroxyphenylacetaldehyde, 1,4-di(4-hydroxyphenyl)butane, 1,4-di(4-hydroxyphenyl)-1-butene, 1,4-di(4-hydroxyphenyl)-2-butene, 1,4-di(4-hydroxyphenyl)-2-butanol, 1,4-di(4-hydroxyphenyl)-2-butanone, 1,4-di(4-hydroxyphenyl)-2,3-butanediol, 1,4-di(4-hydroxyphenyl)-3-hydroxy-2-butanone, 1-(4-hydroxyphenyl)-4-(indole-3)-butane, 1-(4-hydroxyphenyl)-4-(indole-3)-1-butene, 1-di(4-hydroxyphenyl)-4-(indole-3)-2-butene, 1-(4-hydroxyphenyl)-4-(indole-3)-2-butanol, 1-(4-hydroxyphenyl)-4-(indole-3)-2-butanone, 1-(4-hydroxyphenyl)-4-(indole-3)-2,3-butanediol, 1-(4-hydroxyphenyl)-4-(indole-3)-3-hydroxy-2-butanone, indole-3-acetaldehyde, 1,4-di(indole-3)-butane, 1,4-di(indole-3)-1-butene, 1,4-di(indole-3)-2-butene, 1,4-di(indole-3)-2-butanol, 1,4-di(indole-3)-2-butanone, 1,4-di(indole-3)-2,3-butanediol, 1,4-di(indole-3)-3-hydroxy-2-butanone, succinate semialdehyde, hexane-1,8-dicarboxylic acid, 3-hexene-1,8-dicarboxylic acid, 3-hydroxy-hexane-1,8-dicarboxylic acid, 3-hexanone-1,8-dicarboxylic acid, 3,4-hexanediol-1,8-dicarboxylic acid, 4-hydroxy-3-hexanone-1,8-dicarboxylic acid, fucoidan, iodine, chlorophyll, carotenoid, calcium, magnesium, iron, sodium, potassium, and phosphate.

[0070] Certain embodiments of the present invention include methods for converting a polysaccharide to a suitable monosaccharide or oligosaccharide, comprising: (a) contacting the polysaccharide, wherein the polysaccharide is optionally obtained from biomass, with a microbial system for a time sufficient to convert the polysaccharide to a suitable monosaccharide or oligosaccharide, wherein the microbial system comprises, (i) at least one gene encoding and expressing an enzyme selected from a lyase and a hydrolase, wherein the lyase and/or hydrolase optionally comprises at least one signal peptide or at least one autotransporter domain; (ii) at least one gene encoding and expressing an enzyme selected from a monosaccharide transporter, a disaccharide trans-

porter, a trisaccharide transporter, an oligosaccharide transporter, a polysaccharide transporter, and a superchannel; and (iii) at least one gene encoding and expressing an enzyme selected from a monosaccharide dehydrogenase, an isomerase, a dehydratase, a kinase, and an aldolase, thereby converting the polysaccharide to a suitable monosaccharide or oligosaccharide.

[0071] Certain embodiments of the present invention include methods for converting a polysaccharide to a suitable monosaccharide or oligosaccharide, comprising: (a) contacting the polysaccharide, wherein the polysaccharide is optionally obtained from biomass, with a chemical or enzymatic catalysis pathway for a time sufficient to convert the polysaccharide to a first monosaccharide or oligosaccharide; and (b) contacting the first monosaccharide or oligosaccharide with a microbial system for a time sufficient to convert the first monosaccharide or oligosaccharide to the suitable monosaccharide or oligosaccharide, wherein the microbial system comprises, (i) at least one gene encoding and expressing an enzyme selected from a lyase and a hydrolase, (ii) at least one gene encoding and expressing an enzyme selected from a monosaccharide transporter, a disaccharide transporter, a trisaccharide transporter, an oligosaccharide transporter, a polysaccharide transporter, and a superchannel; and (ii) at least one gene encoding and expressing an enzyme selected from a monosaccharide dehydrogenase, an isomerase, a dehydratase, a kinase, and an aldolase, thereby converting the polysaccharide to the suitable monosaccharide or oligosaccharide.

[0072] In certain aspects, the lyase is selected from an alginate lyase, a pectate lyase, a polymannuronate lyase, a polygluronate lyase, a polygalacturonate lyase and a rhamnogalacturonate lyase. In certain aspects, the hydrolase is selected from an alginate hydrolase, a rhamnogalacturonate hydrolase, a polymannuronate hydrolase, a pectin hydrolase, and a polygalacturonate hydrolase. In certain aspects, the transporter is selected from an ABC transporter, a symporter, and an outer membrane porin. In certain aspects, the ABC transporter is selected from Atu3021, Atu3022, Atu3023, Atu3024, algM1, algM2, AlgQ1, AlgQ2, AlgS, OG2516_05558, OG2516_05563, OG2516_05568, OG2516_05573, TogM, TogN, TogA, TogB, and functional variants thereof. In certain aspects, the symporter is selected from V12B01_24239 (SEQ ID NO:26), V12B01_24194 (SEQ ID NO:8), and TogT, and functional variants thereof. In certain aspects, the outer membrane porin comprises a porin selected from V12B01_24269, KdgM, and KdgN, and functional variants thereof.

[0073] Certain embodiments include a recombinant microorganism that is capable of growing on a polysaccharide as a sole source of carbon, wherein the polysaccharide is selected from alginate, pectin, tri-galacturonate, di-galacturonate, cellulose, and hemi-cellulose. In certain aspects, the polysaccharide is alginate. In certain aspects, the polysaccharide is pectin. In certain aspects, the polysaccharide is tri-galacturonate.

[0074] Certain embodiments include a recombinant microorganism, comprising (i) at least one gene encoding and expressing an enzyme selected from a lyase and a hydrolase, wherein the lyase or hydrolase optionally comprises at least one signal peptide or at least one autotransporter domain; (ii) at least one gene encoding and expressing an enzyme selected from a monosaccharide transporter, a disaccharide transporter, a trisaccharide transporter, an oligosaccharide transporter, a polysaccharide transporter, and a superchannel; and

(iii) at least one gene encoding and expressing an enzyme selected from a monosaccharide dehydrogenase, an isomerase, a dehydratase, a kinase, and an aldolase. In certain aspects, the microorganism is capable of growing on a polysaccharide as a sole source of carbon. In certain aspects, the polysaccharide is selected from alginate, pectin, and tri-galacturonate.

[0075] Certain embodiments include methods for converting a suitable monosaccharide or oligosaccharide to a first commodity chemical comprising, (a) contacting the suitable monosaccharide or oligosaccharide with a microbial system for a time sufficient to convert to the suitable monosaccharide or oligosaccharide to the commodity chemical, wherein the microbial system comprises a recombinant microorganism, wherein the microorganism comprises a commodity chemical biosynthesis pathway, thereby converting the suitable monosaccharide or oligosaccharide to the first commodity chemical. In certain aspects, the commodity chemical pathway comprises one or more genes encoding an aldehyde or ketone biosynthesis pathway.

[0076] In certain aspects, the aldehyde or ketone biosynthesis pathway is selected from one or more of an acetoaldehyde, a propionaldehyde, a butyraldehyde, an isobutyraldehyde, a 2-methyl-butyraldehyde, a 3-methyl-butyraldehyde, a 2-phenyl acetaldehyde, a 2-(4-hydroxyphenyl)acetaldehyde, a 2-Indole-3-acetoaldehyde, a glutaraldehyde, a 5-amino-pentaldehyde, a succinate semialdehyde, and a succinate 4-hydroxyphenyl acetaldehyde biosynthesis pathway. In certain aspects, the aldehyde or ketone biosynthesis pathway comprises an acetoaldehyde biosynthesis pathway and a biosynthesis pathway selected from a propionaldehyde, butyraldehyde, isobutyraldehyde, 2-methyl-butyraldehyde, 3-methyl-butyraldehyde, a 2-phenyl acetoaldehyde, a 2-(4-hydroxyphenyl)acetaldehyde, and a 2-Indole-3-acetoaldehyde biosynthesis pathway.

[0077] In certain aspects, the aldehyde or ketone biosynthesis pathway comprises a propionaldehyde biosynthesis pathway and a biosynthesis pathway selected from a butyraldehyde, isobutyraldehyde, 2-methyl-butyraldehyde, 3-methyl-butyraldehyde, and phenylacetoaldehyde biosynthesis pathway. In certain aspects, the aldehyde or ketone biosynthesis pathway comprises a butyraldehyde biosynthesis pathway and a biosynthesis pathway selected from an isobutyraldehyde, 2-methyl-butyraldehyde, 3-methyl-butyraldehyde, a 2-phenyl acetoaldehyde, a 2-(4-hydroxyphenyl)acetaldehyde, and a 2-Indole-3-acetoaldehyde biosynthesis pathway. In certain aspects, the aldehyde or ketone biosynthesis pathway comprises an isobutyraldehyde biosynthesis pathway and a biosynthesis pathway selected from a 2-methyl-butyraldehyde, 3-methyl-butyraldehyde, a 2-phenyl acetoaldehyde, a 2-(4-hydroxyphenyl)acetaldehyde, and a 2-Indole-3-acetoaldehyde biosynthesis pathway.

[0078] In certain aspects, the aldehyde or ketone biosynthesis pathway comprises a 2-methyl-butyraldehyde biosynthesis pathway and a biosynthesis pathway selected from a 3-methyl-butyraldehyde, a 2-phenyl acetoaldehyde, a 2-(4-hydroxyphenyl)acetaldehyde, and a 2-Indole-3-acetoaldehyde biosynthesis pathway. In certain aspects, the aldehyde or ketone biosynthesis pathway comprises a 3-methyl-butyraldehyde biosynthesis pathway and a biosynthesis pathway selected from a 2-phenyl acetoaldehyde, a 2-(4-hydroxyphenyl)acetaldehyde, and a 2-Indole-3-acetoaldehyde biosynthesis pathway. In certain aspects, the aldehyde or ketone biosynthesis pathway comprises a 2-phenyl acetoaldehyde

biosynthesis pathway and a biosynthesis pathway selected from a 2-(4-hydroxyphenyl)acetaldehyde and a 2-Indole-3-acetoaldehyde biosynthesis pathway.

[0079] In certain aspects, the aldehyde or ketone biosynthesis pathway comprises a 2-(4-hydroxyphenyl)acetaldehyde biosynthesis pathway and a 2-Indole-3-acetoaldehyde biosynthesis pathway. In certain aspects, the first commodity chemical is further enzymatically and/or chemically reduced and dehydrated to a second commodity chemical.

[0080] Certain embodiments include methods for converting a suitable monosaccharide or oligosaccharide to a commodity chemical comprising, (a) contacting the suitable monosaccharide or oligosaccharide with a microbial system for a time sufficient to convert to the suitable monosaccharide or oligosaccharide to the commodity chemical, wherein the microbial system comprises; (i) one or more genes encoding and expressing an aldehyde biosynthesis pathway, wherein the aldehyde biosynthesis pathway comprises one or more genes encoding and expressing a decarboxylase enzyme; and (ii)

[0081] one or more genes encoding and expressing an aldehyde reductase, thereby converting the suitable monosaccharide or oligosaccharide to the commodity chemical. In certain aspects, the decarboxylase enzyme is an indole-3-pyruvate decarboxylase (IPDC). In certain aspects, the IPDC comprises an amino acid sequence that is at least 80%, 90%, 95%, 98%, or 99% identical to the amino acid sequence set forth in SEQ ID NO: 312. In certain aspects, the aldehyde reductase enzyme is a phenylacetaldehyde reductase (PAR). In certain aspects, the PAR comprises an amino acid sequence that is at least 80%, 90%, 95%, 98%, or 99% identical to the amino acid sequence set forth in SEQ ID NO: 313. In certain aspects, the commodity chemical is selected from 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol, and indole-3-ethanol.

[0082] Certain embodiments include a recombinant microorganism, comprising (i) one or more genes encoding and expressing an aldehyde biosynthesis pathway, wherein the aldehyde biosynthesis pathway comprises one or more genes encoding and expressing a decarboxylase enzyme; and (ii) one or more genes encoding and expressing an aldehyde reductase. In certain aspects, the aldehyde biosynthesis pathway further comprises one or more genes encoding and expressing an enzyme selected from a CoA-linked aldehyde dehydrogenase, an aldehyde dehydrogenase, and an alcohol dehydrogenase. In certain aspects, the decarboxylase enzyme is an indole-3-pyruvate decarboxylase (IPDC). In certain aspects, the aldehyde reductase enzyme is a phenylacetaldehyde reductase (PAR). In certain aspects, the microorganism is capable of converting a suitable monosaccharide or oligosaccharide to a commodity chemical. In certain aspects, the commodity chemical is selected from 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol, and indole-3-ethanol.

[0083] Certain embodiments include a recombinant microorganism, wherein the microorganism comprises reduced ethanol production capability compared to a wild-type microorganism. In certain aspects, the microorganism comprises a reduction or inhibition in the conversion of acetyl-coA to ethanol. In certain aspects, the recombinant microorganism comprises a reduction of an ethanol dehydrogenase, thereby providing a reduced ethanol production capability. In certain aspects, the ethanol dehydrogenase is an adhE, homolog or variant thereof. In certain aspects, the microorganism comprises a deletion or knockout of an adhE, homolog or variant thereof. In certain aspects, the recombinant microorganism

comprises one or more deletions or knockouts in a gene encoding an enzyme selected from an enzyme that catalyzes the conversion of acetyl-coA to ethanol, an enzyme that catalyzes the conversion of pyruvate to lactate, an enzyme that catalyzes the conversion of fumarate to succinate, an enzyme that catalyzes the conversion of acetyl-coA and phosphate to coA and acetyl phosphate, an enzyme that catalyzes the conversion of acetyl-coA and formate to coA and pyruvate, and an enzyme that catalyzes the conversion of alpha-keto acid to branched chain amino acids.

[0084] Certain embodiments include wherein the microbial systems or recombinant microorganisms described herein comprise a microorganism selected from *Acetobacter aceti*, *Achromobacter*, *Acidiphilium*, *Acinetobacter*, *Actinomadura*, *Actinoplanes*, *Aeropyrum pernix*, *Agrobacterium*, *Alcaligenes*, *Ananas comosus* (M), *Arthrobacter*, *Aspergillus niger*, *Aspergillus oryzae*, *Aspergillus melleus*, *Aspergillus pulverulentus*, *Aspergillus saitoi*, *Aspergillus sojae*, *Aspergillus usamii*, *Bacillus alcalophilus*, *Bacillus amyloliquefaciens*, *Bacillus brevis*, *Bacillus circulans*, *Bacillus clausii*, *Bacillus lentus*, *Bacillus licheniformis*, *Bacillus macerans*, *Bacillus stearothermophilus*, *Bacillus subtilis*, *Bifidobacterium*, *Brevibacillus brevis*, *Burkholderia cepacia*, *Candida cylindracea*, *Candida rugosa*, *Carica papaya* (L), *Cellulosimicrobium*, *Cephalosporium*, *Chaetomium erraticum*, *Chaetomium gracile*, *Clostridium*, *Clostridium butyricum*, *Clostridium acetobutylicum*, *Clostridium thermocellum*, *Corynebacterium* (glutamicum), *Corynebacterium efficiens*, *Escherichia coli*, *Enterococcus*, *Erwinia chrysanthemi*, *Gluconobacter*, *Gluconacetobacter*, *Haloarcula*, *Humicola insolens*, *Humicola nsolens*, *Kitasatospora setae*, *Klebsiella*, *Klebsiella oxytoca*, *Kluyveromyces*, *Kluyveromyces fragilis*, *Kluyveromyces lactis*, *Kocuria*, *Lactolactis*, *Lactobacillus*, *Lactobacillus fermentum*, *Lactobacillus sake*, *Lactococcus*, *Lactococcus lactis*, *Leuconostoc*, *Methylocystis*, *Methanobacterium bryantii*, *Methanobacterium imperiale*, *Micrococcus lysodeikticus*, *Microlunatus*, *Mucor javanicus*, *Mycobacterium*, *Myrothecium*, *Nitrobacter*, *Nitrosomonas*, *Nocardia*, *Papaya carica*, *Pediococcus*, *Pediococcus halophilus*, *Penicillium*, *Penicillium camemberti*, *Penicillium citrinum*, *Penicillium emersonii*, *Penicillium roqueforti*, *Penicillium lilactinum*, *Penicillium multicolor*, *Paracoccus pantotrophus*, *Propionibacterium*, *Pseudomonas*, *Pseudomonas fluorescens*, *Pseudomonas denitrificans*, *Pyrococcus*, *Pyrococcus furiosus*, *Pyrococcus horikoshii*, *Rhizobium*, *Rhizomucor miehei*, *Rhizomucor pusillus* Lindt, *Rhizopus*, *Rhizopus delemar*, *Rhizopus japonicus*, *Rhizopus niveus*, *Rhizopus oryzae*, *Rhizopus oligosporus*, *Rhodococcus*, *Sccharomyces cerevisiae*, *Sclerotinia libertina*, *Sphingobacterium multivorum*, *Sphingobium*, *Sphingomonas*, *Streptococcus*, *Streptococcus thermophilus* Y-1, *Streptomyces*, *Streptomyces griseus*, *Streptomyces lividans*, *Streptomyces murinus*, *Streptomyces rubiginosus*, *Streptomyces violaceoruber*, *Streptoverticillium mobarraense*, *Tetragenococcus*, *Thermus*, *Thiosphaera pantotropha*, *Trametes*, *Trichoderma*, *Trichoderma longibrachiatum*, *Trichoderma reesei*, *Trichoderma viride*, *Trichosporon penicillatum*, *Vibrio alginolyticus*, *Xanthomonas*, yeast, *Zygosaccharomyces rouxii*, *Zymomonas*, and *Zymomonas mobilis*.

[0085] Certain embodiments include a commodity chemical produced by the methods described herein. Certain aspects include a blended commodity chemical comprising a commodity chemical produced by the methods provided

herein and a refinery-produced petroleum product. In certain aspects, the commodity chemical is selected from a C10-C12 hydrocarbon, 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol, and indole-3-ethanol. In certain aspects, the C10-C12 hydrocarbon is selected from 2,7-dimethyloctane and 2,9-dimethyldecane. In certain aspects, the refinery-produced petroleum product is selected from jet fuel and diesel fuel.

[0086] Certain embodiments include methods of producing a commodity chemical enriched refinery-produced petroleum product, comprising (a) blending the refinery-produced petroleum product with the commodity chemical produced by the methods described herein, thereby producing the commodity chemical enriched refinery-produced petroleum product.

DETAILED DESCRIPTION

[0087] Embodiments of the present invention relate to the unexpected discovery that microorganisms which are otherwise incapable of growing on certain polysaccharides derived from biomass as a sole source of carbon, can be engineered to grow on these polysaccharides as a sole source of carbon. Such microorganisms can include both prokaryotic and eukaryotic microorganisms, such as bacteria and yeast. In some aspects, certain laboratory and/or wild-type strains of *E. coli* can be engineered to grow on biomass derived from either alginate or pectin as a sole source of carbon to produce suitable monosaccharides or other molecules. Among other uses apparent to a person skilled in the art, the monosaccharides and other molecules produced by the growth of these engineered or recombinant microorganisms on alginate or pectin may be utilized as feedstock in the production of various commodity chemicals, such as biofuels.

[0088] Alginate and pectin provide advantages over other biomass sources in the production of biofuel feedstocks. For example, large-scale aquatic-farming can generate a significant amount of biomass without replacing food crop production with energy crop production, deforestation, and recultivating currently uncultivated land, as most of hydrosphere including oceans, rivers, and lakes remains untapped. As one particular example, the Pacific coast of North America is abundant in minerals necessary for large-scale aqua-farming. Giant kelp, which lives in the area, grows as fast as 1 m/day, the fastest among plants on earth, and grows up to 50 m. Additionally, aqua-farming has other benefits including the prevention of a red tide outbreak and the creation of a fish-friendly environment.

[0089] As an additional advantage, and in contrast to lignocellulosic biomass, biomass derived from aquatic, fruit, plant and/or vegetable sources is easy to degrade. Such biomass typically lacks lignin and is significantly more fragile than lignocellulosic biomass and can thus be easily degraded using either enzymes or chemical catalysts (e.g., formate). As one example, aquatic biomass such as seaweed may be easily converted to monosaccharides using either enzymes or chemical catalysis, as seaweed has significantly simpler major sugar components (Alginate: 30%, Mannitol: 15%) as compared to lignocellulose (Glucose: 24.1-39%, Mannose: 0.2-4.6%, Galactose: 0.5-2.4%, Xylose: 0.4-22.1%, Arabinose 1.5-2.8%, and Uronic acids: 1.2-20.7%, and total sugar contents are corresponding to 36.5-70% of dried weight).

[0090] As an additional example, biomass from plants such as fruit and/or vegetable contains pectin, a heteropolysaccharide derived from the plant cell wall. The characteristic structure of pectin is a linear chain of α -(1-4)-linked D-galacturonic

acid that forms the pectin-backbone, a homogalacturonan. Pectin can be easily converted to oligosaccharides or suitable monosaccharides using either enzymes, chemical catalysis, and/or microbial systems designed to utilize pectin as a source of carbon, as described herein. Saccharification and fermentation using aquatic, fruit, and/or vegetable biomass is much easier than using lignocellulose.

[0091] In this regard, embodiments of the present invention also relate to the surprising discovery that certain microorganisms can be engineered to produce various commodity chemicals, such as biofuels. In certain aspects, these biofuels may include alkanes, such as medium to long chain alkanes, which provide advantages over ethanol based biofuels. In certain aspects, the monosaccharides (e.g., 2-keto-3-deoxy D-gluconate; KDG) and other molecules produced by the growth of various engineered or recombinant microorganisms (e.g., recombinant microorganisms growing on pectin or alginate as a source of carbon) may be useful in the production of commodity chemicals, such as biofuels. As one example, suitable monosaccharides such as KDG may be utilized by recombinant microorganisms to produce alkanes, such as medium to long chain alkanes, among other chemicals. In certain aspects, such recombinant microorganisms may be utilized to produce such commodity chemical as 2,7 dimethyl octane and 2,9 dimethyl decane, among others provided herein and known in the art.

[0092] Such processes produce biofuels with significant advantages over other biofuels. In particular, medium to long chain alkanes provide a number of important advantages over the existing common biofuels such as ethanol and butanol, and are attractive long-term replacements of petroleum-based fuels such as gasoline, diesels, kerosene, and heavy oils in the future. As one example, medium to long chain alkanes and alcohols are major components in all petroleum products and jet fuel in particular, and hence alkanes we produce can be utilized directly by existing engines. By way of further example, medium to long chain alcohols are far better fuels than ethanol, and have a nearly comparable energy density to gasoline.

[0093] As another example, n-alkanes are major components of all oil products including gasoline, diesels, kerosene, and heavy oils. Microbial systems or recombinant microorganisms may be used to produce n-alkanes with different carbon lengths ranging, for example, from C7 to over C20: C7 for gasoline (e.g., motor vehicles), C10-C15 for diesels (e.g., motor vehicles, trains, and ships), and C8-C16 for kerosene (e.g., aviation and ships), and for all heavy oils.

[0094] As one aspect of the invention, the commodity chemicals produced by the methods and recombinant microorganisms described herein may be utilized by existing petroleum refineries for the purposes of blending with petroleum products produced by traditional refinery methods. To this end, as noted above, fuel producers are seeking substantially similar, low carbon fuels that can be blended and distributed through existing infrastructure (refineries, pipelines, tankers). As hydrocarbons, the commodity chemicals produced according to the methods herein are substantially similar to petroleum derived fuels, reduce green house gas emissions by more than 80% from petroleum derived fuels, and are compatible with existing infrastructure in the oil and gas industry. For instance, certain of the commodity chemicals produced herein, including, for example, various C10-C12 hydrocarbons such as 2,7 dimethyloctane, 2,7 dimethyldecane,

among others, are blendable directly into refinery-produced petroleum products, such as jet and diesel fuels. By using such biologically produced commodity chemicals as a blend-stock for jet and diesel fuels, refineries may reduce Green House Gas emissions by more than 80%.

[0095] Accordingly, certain embodiments of the present invention relate generally to methods for converting biomass to a commodity chemical, comprising obtaining a polysaccharide from biomass; contacting the polysaccharide with a polysaccharide degrading or depolymerizing pathway, thereby converting the polysaccharide to a suitable monosaccharide. The suitable monosaccharide obtained from such a process may be used for any desired purpose. For instance, in certain aspects, the suitable monosaccharide may then be converted to a commodity chemical (e.g., biofuel) by contacting the suitable monosaccharide with a biofuel biosynthesis pathway, whether as part of a recombinant microorganism, an in vitro enzymatic or chemical pathway, or a combination thereof, thereby converting the monosaccharide to a commodity chemical.

[0096] In other aspects, in producing a commodity chemical such as a biofuel, a suitable monosaccharide may be obtained directly from any available source and converted to a commodity chemical by contacting the suitable monosaccharide with a biofuel biosynthesis pathway, as described herein. Among other uses apparent to a person skilled in the art, such biofuels may then be blended directly with refinery produced petroleum products, such as jet and diesel fuels, to produce commodity chemical enriched, refinery-produced petroleum products.

DEFINITIONS

[0097] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by those of ordinary skill in the art to which the invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, preferred methods and materials are described. For the purposes of the present invention, the following terms are defined below. All references referred to herein are incorporated by reference in their entirety.

[0098] The articles “a” and “an” are used herein to refer to one or to more than one (i.e. to at least one) of the grammatical object of the article. By way of example, “an element” means one element or more than one element.

[0099] By “about” is meant a quantity, level, value, number, frequency, percentage, dimension, size, amount, weight or length that varies by as much as 30, 25, 20, 15, 10, 9, 8, 7, 6, 5, 4, 3, 2 or 1% to a reference quantity, level, value, number, frequency, percentage, dimension, size, amount, weight or length.

[0100] The term “biologically active fragment”, as applied to fragments of a reference polynucleotide or polypeptide sequence, refers to a fragment that has at least about 0.1, 0.5, 1, 2, 5, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 96, 97, 98, 99, 100, 110, 120, 150, 200, 300, 400, 500, 600, 700, 800, 900, 1000% or more of the activity of a reference sequence.

[0101] The term “reference sequence” refers generally to a nucleic acid coding sequence, or amino acid sequence, of any enzyme having a biological activity described herein (e.g., saccharide dehydrogenase, alcohol dehydrogenase, dehydratase, lyase, transporter, decarboxylase, hydrolase, etc.),

such as a “wild-type” sequence, including those reference sequences exemplified by SEQ ID NOS:1-144, and 308-313. A reference sequence may also include naturally-occurring, functional variants (i.e., orthologs or homologs) of the sequences described herein.

[0102] Included within the scope of the present invention are biologically active fragments of at least about 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 40, 50, 60, 70, 80, 90, 100, 120, 140, 160, 180, 200, 220, 240, 260, 280, 300, 320, 340, 360, 380, 400, 500, 600 or more contiguous nucleotides or amino acid residues in length, including all integers in between, which comprise or encode a polypeptide having an enzymatic activity of a reference polynucleotide or polypeptide. Representative biologically active fragments generally participate in an interaction, e.g., an intra-molecular or an inter-molecular interaction. An inter-molecular interaction can be a specific binding interaction or an enzymatic interaction. Examples of enzymatic interactions or activities include saccharide dehydrogenase activities, alcohol dehydrogenase activities, dehydratase activities, lyase activities, transporter activities, isomerase activities, kinase activities, among others described herein. Biologically active fragments typically comprise one or more active sites or enzymatic/binding motifs, as described herein and known in the art.

[0103] By “coding sequence” is meant any nucleic acid sequence that contributes to the code for the polypeptide product of a gene. By contrast, the term “non-coding sequence” refers to any nucleic acid sequence that does not contribute to the code for the polypeptide product of a gene.

[0104] Throughout this specification, unless the context requires otherwise, the words “comprise”, “comprises” and “comprising” will be understood to imply the inclusion of a stated step or element or group of steps or elements but not the exclusion of any other step or element or group of steps or elements.

[0105] By “consisting of,” is meant including, and limited to, whatever follows the phrase “consisting of” Thus, the phrase “consisting of” indicates that the listed elements are required or mandatory, and that no other elements may be present.

[0106] By “consisting essentially of” is meant including any elements listed after the phrase, and limited to other elements that do not interfere with or contribute to the activity or action specified in the disclosure for the listed elements. Thus, the phrase “consisting essentially of” indicates that the listed elements are required or mandatory, but that other elements are optional and may or may not be present depending upon whether or not they affect the activity or action of the listed elements.

[0107] The terms “complementary” and “complementarity” refer to polynucleotides (i.e., a sequence of nucleotides) related by the base-pairing rules. For example, the sequence “A-G-T;” is complementary to the sequence “T-C-A.” Complementarity may be “partial,” in which only some of the nucleic acids’ bases are matched according to the base pairing rules. Or, there may be “complete” or “total” complementarity between the nucleic acids. The degree of complementarity between nucleic acid strands has significant effects on the efficiency and strength of hybridization between nucleic acid strands.

[0108] By “corresponds to” or “corresponding to” is meant (a) a polynucleotide having a nucleotide sequence that is substantially identical or complementary to all or a portion of a reference polynucleotide sequence or encoding an amino

acid sequence identical to an amino acid sequence in a peptide or protein; or (b) a peptide or polypeptide having an amino acid sequence that is substantially identical to a sequence of amino acids in a reference peptide or protein.

[0109] By “derivative” is meant a polypeptide that has been derived from the basic sequence by modification, for example by conjugation or complexing with other chemical moieties (e.g., pegylation) or by post-translational modification techniques as would be understood in the art. The term “derivative” also includes within its scope alterations that have been made to a parent sequence including additions or deletions that provide for functionally equivalent molecules.

[0110] By “enzyme reactive conditions” it is meant that any necessary conditions are available in an environment (i.e., such factors as temperature, pH, lack of inhibiting substances) which will permit the enzyme to function. Enzyme reactive conditions can be either in vitro, such as in a test tube, or in vivo, such as within a cell.

[0111] As used herein, the terms “function” and “functional” and the like refer to a biological or enzymatic function.

[0112] By “gene” is meant a unit of inheritance that occupies a specific locus on a chromosome and consists of transcriptional and/or translational regulatory sequences and/or a coding region and/or non-translated sequences (i.e., introns, 5' and 3' untranslated sequences).

[0113] “Homology” refers to the percentage number of amino acids that are identical or constitute conservative substitutions. Homology may be determined using sequence comparison programs such as GAP (Deveraux et al., 1984, *Nucleic Acids Research* 12, 387-395) which is incorporated herein by reference. In this way sequences of a similar or substantially different length to those cited herein could be compared by insertion of gaps into the alignment, such gaps being determined, for example, by the comparison algorithm used by GAP.

[0114] The term “host cell” includes an individual cell or cell culture which can be or has been a recipient of any recombinant vector(s) or isolated polynucleotide of the invention. Host cells include progeny of a single host cell, and the progeny may not necessarily be completely identical (in morphology or in total DNA complement) to the original parent cell due to natural, accidental, or deliberate mutation and/or change. A host cell includes cells transfected, transformed, or infected in vivo or in vitro with a recombinant vector or a polynucleotide of the invention. A host cell which comprises a recombinant vector of the invention is a recombinant host cell, recombinant cell, or recombinant microorganism.

[0115] By “isolated” is meant material that is substantially or essentially free from components that normally accompany it in its native state. For example, an “isolated polynucleotide”, as used herein, refers to a polynucleotide, which has been purified from the sequences which flank it in a naturally-occurring state, e.g., a DNA fragment which has been removed from the sequences that are normally adjacent to the fragment. Alternatively, an “isolated peptide” or an “isolated polypeptide” and the like, as used herein, refer to in vitro isolation and/or purification of a peptide or polypeptide molecule from its natural cellular environment, and from association with other components of the cell, i.e., it is not associated with in vivo substances.

[0116] By “increased” or “increasing” is meant the ability of one or more recombinant microorganisms to produce a

greater amount of a given product or molecule (e.g., commodity chemical, biofuel, or intermediate product thereof) as compared to a control microorganism, such as an unmodified microorganism or a differently modified microorganism. An “increased” amount is typically a “statistically significant” amount, and may include an increase that is 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 30 or more times (including all integers and decimal points in between, e.g., 1.5, 1.6, 1.7, 1.8, etc.) the amount produced by an unmodified microorganism or a differently modified microorganism.

[0117] By “obtained from” is meant that a sample such as, for example, a polynucleotide extract or polypeptide extract is isolated from, or derived from, a particular source, such as a desired organism, typically a microorganism. “Obtained from” can also refer to the situation in which a polynucleotide or polypeptide sequence is isolated from, or derived from, a particular organism or microorganism. For example, a polynucleotide sequence encoding a benzaldehyde lyase enzyme may be isolated from a variety of prokaryotic or eukaryotic microorganisms, such as *Pseudomonas*.

[0118] The term “operably linked” as used herein means placing a gene under the regulatory control of a promoter, which then controls the transcription and optionally the translation of the gene. In the construction of heterologous promoter/structural gene combinations, it is generally preferred to position the genetic sequence or promoter at a distance from the gene transcription start site that is approximately the same as the distance between that genetic sequence or promoter and the gene it controls in its natural setting; i.e. the gene from which the genetic sequence or promoter is derived. As is known in the art, some variation in this distance can be accommodated without loss of function. Similarly, the preferred positioning of a regulatory sequence element with respect to a heterologous gene to be placed under its control is defined by the positioning of the element in its natural setting; i.e., the genes from which it is derived. “Constitutive promoters” are typically active, i.e., promote transcription, under most conditions. “Inducible promoters” are typically active only under certain conditions, such as in the presence of a given molecule factor (e.g., IPTG) or a given environmental condition (e.g., CO₂ concentration, nutrient levels, light, heat). In the absence of that condition, inducible promoters typically do not allow significant or measurable levels of transcriptional activity.

[0119] The recitation “polynucleotide” or “nucleic acid” as used herein designates mRNA, RNA, cRNA, rRNA, cDNA or DNA. The term typically refers to polymeric form of nucleotides of at least 10 bases in length, either ribonucleotides or deoxynucleotides or a modified form of either type of nucleotide. The term includes single and double stranded forms of DNA.

[0120] As will be understood by those skilled in the art, the polynucleotide sequences of this invention can include genomic sequences, extra-genomic and plasmid-encoded sequences and smaller engineered gene segments that express, or may be adapted to express, proteins, polypeptides, peptides and the like. Such segments may be naturally isolated, or modified synthetically by the hand of man.

[0121] Polynucleotides may be single-stranded (coding or antisense) or double-stranded, and may be DNA (genomic, cDNA or synthetic) or RNA molecules. Additional coding or non-coding sequences may, but need not, be present within a

polynucleotide of the present invention, and a polynucleotide may, but need not, be linked to other molecules and/or support materials.

[0122] Polynucleotides may comprise a native sequence (i.e., an endogenous sequence) or may comprise a variant, or a biological functional equivalent of such a sequence. Polynucleotide variants may contain one or more substitutions, additions, deletions and/or insertions, as further described below, preferably such that the enzymatic activity of the encoded polypeptide is not substantially diminished relative to the unmodified polypeptide, and preferably such that the enzymatic activity of the encoded polypeptide is improved (e.g., optimized) relative to the unmodified polypeptide. The effect on the enzymatic activity of the encoded polypeptide may generally be assessed as described herein.

[0123] The polynucleotides of the present invention, regardless of the length of the coding sequence itself, may be combined with other DNA sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their overall length may vary considerably. It is therefore contemplated that a polynucleotide fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant DNA protocol.

[0124] The terms “polynucleotide variant” and “variant” and the like refer to polynucleotides that display substantial sequence identity with any of the reference polynucleotide sequences or genes described herein, and to polynucleotides that hybridize with any polynucleotide reference sequence described herein, or any polynucleotide coding sequence of any gene or protein referred to herein, under low stringency, medium stringency, high stringency, or very high stringency conditions that are defined hereinafter and known in the art. These terms also encompass polynucleotides that are distinguished from a reference polynucleotide by the addition, deletion or substitution of at least one nucleotide. Accordingly, the terms “polynucleotide variant” and “variant” include polynucleotides in which one or more nucleotides have been added or deleted, or replaced with different nucleotides. In this regard, it is well understood in the art that certain alterations inclusive of mutations, additions, deletions and substitutions can be made to a reference polynucleotide whereby the altered polynucleotide retains the biological function or activity of the reference polynucleotide, or has increased activity in relation to the reference polynucleotide (i.e., optimized). Polynucleotide variants include, for example, polynucleotides having at least 50% (and at least 51% to at least 99% and all integer percentages in between) sequence identity with a reference polynucleotide described herein.

[0125] The terms “polynucleotide variant” and “variant” also include naturally-occurring allelic variants that encode these enzymes. Examples of naturally-occurring variants include allelic variants (same locus), homologs (different locus), and orthologs (different organism). Naturally occurring variants such as these can be identified and isolated using well-known molecular biology techniques including, for example, various polymerase chain reaction (PCR) and hybridization-based techniques as known in the art. Naturally occurring variants can be isolated from any organism that encodes one or more genes having a suitable enzymatic activ-

ity described herein (e.g., C—C ligase, diol dehydrogenase, pectate lyase, alginate lyase, diol dehydratase, transporter, etc.).

[0126] Non-naturally occurring variants can be made by mutagenesis techniques, including those applied to polynucleotides, cells, or organisms. The variants can contain nucleotide substitutions, deletions, inversions and insertions. Variation can occur in either or both the coding and non-coding regions. In certain aspects, non-naturally occurring variants may have been optimized for use in a given microorganism (e.g., *E. coli*), such as by engineering and screening the enzymes for increased activity, stability, or any other desirable feature. The variations can produce both conservative and non-conservative amino acid substitutions (as compared to the originally encoded product). For nucleotide sequences, conservative variants include those sequences that, because of the degeneracy of the genetic code, encode the amino acid sequence of a reference polypeptide. Variant nucleotide sequences also include synthetically derived nucleotide sequences, such as those generated, for example, by using site-directed mutagenesis but which still encode a biologically active polypeptide. Generally, variants of a particular reference nucleotide sequence will have at least about 30%, 40%, 50%, 55%, 60%, 65%, 70%, generally at least about 75%, 80%, 85%, 90% to 95% or more, and even about 97% or 98% or more sequence identity to that particular nucleotide sequence as determined by sequence alignment programs described elsewhere herein using default parameters.

[0127] As used herein, the term “hybridizes under low stringency, medium stringency, high stringency, or very high stringency conditions” describes conditions for hybridization and washing. Guidance for performing hybridization reactions can be found in Ausubel et al., “Current Protocols in Molecular Biology”, John Wiley & Sons Inc, 1994-1998, Sections 6.3.1-6.3.6. Aqueous and non-aqueous methods are described in that reference and either can be used.

[0128] Reference herein to “low stringency” conditions include and encompass from at least about 1% v/v to at least about 15% v/v formamide and from at least about 1 M to at least about 2 M salt for hybridization at 42° C., and at least about 1 M to at least about 2 M salt for washing at 42° C. Low stringency conditions also may include 1% Bovine Serum Albumin (BSA), 1 mM EDTA, 0.5 M NaHPO₄ (pH 7.2), 7% SDS for hybridization at 65° C., and (i) 2×SSC, 0.1% SDS; or (ii) 0.5% BSA, 1 mM EDTA, 40 mM NaHPO₄ (pH 7.2), 5% SDS for washing at room temperature. One embodiment of low stringency conditions includes hybridization in 6× sodium chloride/sodium citrate (SSC) at about 45° C., followed by two washes in 0.2×SSC, 0.1% SDS at least at 50° C. (the temperature of the washes can be increased to 55° C. for low stringency conditions).

[0129] “Medium stringency” conditions include and encompass from at least about 16% v/v to at least about 30% v/v formamide and from at least about 0.5 M to at least about 0.9 M salt for hybridization at 42° C., and at least about 0.1 M to at least about 0.2 M salt for washing at 55° C. Medium stringency conditions also may include 1% Bovine Serum Albumin (BSA), 1 mM EDTA, 0.5 M NaHPO₄ (pH 7.2), 7% SDS for hybridization at 65° C., and (i) 2×SSC, 0.1% SDS; or (ii) 0.5% BSA, 1 mM EDTA, 40 mM NaHPO₄ (pH 7.2), 5% SDS for washing at 60-65° C. One embodiment of medium

stringency conditions includes hybridizing in 6×SSC at about 45° C., followed by one or more washes in 0.2×SSC, 0.1% SDS at 60° C.

[0130] “High stringency” conditions include and encompass from at least about 31% v/v to at least about 50% v/v formamide and from about 0.01 M to about 0.15 M salt for hybridization at 42° C., and about 0.01 M to about 0.02 M salt for washing at 55° C. High stringency conditions also may include 1% BSA, 1 mM EDTA, 0.5 M NaHPO₄ (pH 7.2), 7% SDS for hybridization at 65° C., and (i) 0.2×SSC, 0.1% SDS; or (ii) 0.5% BSA, 1 mM EDTA, 40 mM NaHPO₄ (pH 7.2), 1% SDS for washing at a temperature in excess of 65° C. One embodiment of high stringency conditions includes hybridizing in 6×SSC at about 45° C., followed by one or more washes in 0.2×SSC, 0.1% SDS at 65° C.

[0131] One embodiment of “very high stringency” conditions includes hybridizing in 0.5 M sodium phosphate, 7% SDS at 65° C., followed by one or more washes in 0.2×SSC, 1% SDS at 65° C.

[0132] Other stringency conditions are well known in the art and a skilled addressee will recognize that various factors can be manipulated to optimize the specificity of the hybridization. Optimization of the stringency of the final washes can serve to ensure a high degree of hybridization. For detailed examples, see Ausubel et al., supra at pages 2.10.1 to 2.10.16 and Sambrook et al., Current Protocols in Molecular Biology (1989), at sections 1.101 to 1.104.

[0133] While stringent washes are typically carried out at temperatures from about 42° C. to 68° C., one skilled in the art will appreciate that other temperatures may be suitable for stringent conditions. Maximum hybridization rate typically occurs at about 20° C. to 25° C. below the T_m for formation of a DNA-DNA hybrid. It is well known in the art that the T_m is the melting temperature, or temperature at which two complementary polynucleotide sequences dissociate. Methods for estimating T_m are well known in the art (see Ausubel et al., supra at page 2.10.8).

[0134] In general, the T_m of a perfectly matched duplex of DNA may be predicted as an approximation by the formula: T_m=81.5+16.6 (log₁₀M)+0.41 (% G+C)-0.63 (% formamide)-(600/length) wherein: M is the concentration of Na⁺, preferably in the range of 0.01 molar to 0.4 molar; % G+C is the sum of guano sine and cytosine bases as a percentage of the total number of bases, within the range between 30% and 75% G+C; % formamide is the percent formamide concentration by volume; length is the number of base pairs in the DNA duplex. The T_m of a duplex DNA decreases by approximately 1° C. with every increase of 1% in the number of randomly mismatched base pairs. Washing is generally carried out at T_m-15° C. for high stringency, or T_m-30° C. for moderate stringency.

[0135] In one example of a hybridization procedure, a membrane (e.g., a nitrocellulose membrane or a nylon membrane) containing immobilized DNA is hybridized overnight at 42° C. in a hybridization buffer (50% deionizer formamide, 5×SSC, 5× Reinhardt’s solution (0.1% fecal, 0.1% polyvinylpyrrolidone and 0.1% bovine serum albumin), 0.1% SDS and 200 mg/mL denatured salmon sperm DNA) containing a labeled probe. The membrane is then subjected to two sequential medium stringency washes (i.e., 2×SSC, 0.1% SDS for 15 min at 45° C., followed by 2×SSC, 0.1% SDS for 15 min at 50° C.), followed by two sequential higher stringency washes (i.e., 0.2×SSC, 0.1% SDS for 12 min at 55° C. followed by 0.2×SSC and 0.1% SDS solution for 12 min at 65-68° C.

[0136] Polynucleotides and fusions thereof may be prepared, manipulated and/or expressed using any of a variety of well established techniques known and available in the art. For example, polynucleotide sequences which encode polypeptides of the invention, or fusion proteins or functional equivalents thereof, may be used in recombinant DNA molecules to direct expression of a selected enzyme in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences that encode substantially the same or a functionally equivalent amino acid sequence may be produced and these sequences may be used to clone and express a given polypeptide.

[0137] As will be understood by those of skill in the art, it may be advantageous in some instances to produce polypeptide-encoding nucleotide sequences possessing non-naturally occurring codons. For example, codons preferred by a particular prokaryotic or eukaryotic host can be selected to increase the rate of protein expression or to produce a recombinant RNA transcript having desirable properties, such as a half-life which is longer than that of a transcript generated from the naturally occurring sequence. Such nucleotides are typically referred to as “codon-optimized.” Any of the nucleotide sequences described herein may be utilized in such a “codon-optimized” form. For example, the nucleotide coding sequence of the benzaldehyde lyase from *Pseudomonas fluorescens* may be codon-optimized for expression in *E. coli*.

[0138] Moreover, the polynucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter polypeptide encoding sequences for a variety of reasons, including but not limited to, alterations which modify the cloning, processing, expression and/or activity of the gene product.

[0139] In order to express a desired polypeptide, a nucleotide sequence encoding the polypeptide, or a functional equivalent, may be inserted into appropriate expression vector, i.e., a vector that contains the necessary elements for the transcription and translation of the inserted coding sequence. Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding a polypeptide of interest and appropriate transcriptional and translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. Such techniques are described in Sambrook et al., Molecular Cloning, A Laboratory Manual (1989), and Ausubel et al., Current Protocols in Molecular Biology (1989).

[0140] “Polypeptide,” “polypeptide fragment,” “peptide” and “protein” are used interchangeably herein to refer to a polymer of amino acid residues and to variants and synthetic analogues of the same. Thus, these terms apply to amino acid polymers in which one or more amino acid residues are synthetic non-naturally occurring amino acids, such as a chemical analogue of a corresponding naturally occurring amino acid, as well as to naturally-occurring amino acid polymers. In certain aspects, polypeptides may include enzymatic polypeptides, or “enzymes,” which typically catalyze (i.e., increase the rate of) various chemical reactions.

[0141] The recitation polypeptide “variant” refers to polypeptides that are distinguished from a reference polypeptide sequence by the addition, deletion or substitution of at least one amino acid residue. In certain embodiments, a

polypeptide variant is distinguished from a reference polypeptide by one or more substitutions, which may be conservative or non-conservative. In certain embodiments, the polypeptide variant comprises conservative substitutions and, in this regard, it is well understood in the art that some amino acids may be changed to others with broadly similar properties without changing the nature of the activity of the polypeptide. Polypeptide variants also encompass polypeptides in which one or more amino acids have been added or deleted, or replaced with different amino acid residues.

[0142] The present invention contemplates the use in the methods described herein of variants of full-length polypeptides having any of the enzymatic activities described herein, truncated fragments of these full-length polypeptides, variants of truncated fragments, as well as their related biologically active fragments. Typically, biologically active fragments of a polypeptide may participate in an interaction, for example, an intra-molecular or an inter-molecular interaction. An inter-molecular interaction can be a specific binding interaction or an enzymatic interaction (e.g., the interaction can be transient and a covalent bond is formed or broken). Biologically active fragments of a polypeptide/enzyme an enzymatic activity described herein include peptides comprising amino acid sequences sufficiently similar to, or derived from, the amino acid sequences of a (putative) full-length reference polypeptide sequence. Typically, biologically active fragments comprise a domain or motif with at least one enzymatic activity, and may include one or more (and in some cases all) of the various active domains. A biologically active fragment of an enzyme can be a polypeptide fragment which is, for example, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, 220, 240, 260, 280, 300, 320, 340, 360, 380, 400, 450, 500, 600 or more contiguous amino acids, including all integers in between, of a reference polypeptide sequence. In certain embodiments, a biologically active fragment comprises a conserved enzymatic sequence, domain, or motif, as described elsewhere herein and known in the art. Suitably, the biologically-active fragment has no less than about 1%, 10%, 25%, 50% of an activity of the wild-type polypeptide from which it is derived.

[0143] The term “exogenous” refers generally to a polynucleotide sequence or polypeptide that does not naturally occur in a wild-type cell or organism, but is typically introduced into the cell by molecular biological techniques, i.e., engineering to produce a recombinant microorganism. Examples of “exogenous” polynucleotides include vectors, plasmids, and/or man-made nucleic acid constructs encoding a desired protein or enzyme. The term “endogenous” refers generally to naturally occurring polynucleotide sequences or polypeptides that may be found in a given wild-type cell or organism. For example, certain naturally-occurring bacterial or yeast species do not typically contain a benzaldehyde lyase gene, and, therefore, do not comprise an “endogenous” polynucleotide sequence that encodes a benzaldehyde lyase. In this regard, it is also noted that even though an organism may comprise an endogenous copy of a given polynucleotide sequence or gene, the introduction of a plasmid or vector encoding that sequence, such as to over-express or otherwise regulate the expression of the encoded protein, represents an “exogenous” copy of that gene or polynucleotide sequence. Any of the pathways, genes, or enzymes described herein may utilize or rely on an “endogenous” sequence, or may be

provided as one or more “exogenous” polynucleotide sequences, and/or may be utilized according to the endogenous sequences already contained within a given microorganism.

[0144] A “recombinant” microorganism typically comprises one or more exogenous nucleotide sequences, such as in a plasmid or vector.

[0145] The recitations “sequence identity” or, for example, comprising a “sequence 50% identical to,” as used herein, refer to the extent that sequences are identical on a nucleotide-by-nucleotide basis or an amino acid-by-amino acid basis over a window of comparison. Thus, a “percentage of sequence identity” may be calculated by comparing two optimally aligned sequences over the window of comparison, determining the number of positions at which the identical nucleic acid base (e.g., A, T, C, G, I) or the identical amino acid residue (e.g., Ala, Pro, Ser, Thr, Gly, Val, Leu, Ile, Phe, Tyr, Trp, Lys, Arg, His, Asp, Glu, Asn, Gln, Cys and Met) occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison (i.e., the window size), and multiplying the result by 100 to yield the percentage of sequence identity.

[0146] Terms used to describe sequence relationships between two or more polynucleotides or polypeptides include “reference sequence”, “comparison window”, “sequence identity”, “percentage of sequence identity” and “substantial identity”. A “reference sequence” is at least 12 but frequently 15 to 18 and often at least 25 monomer units, inclusive of nucleotides and amino acid residues, in length. Because two polynucleotides may each comprise (1) a sequence (i.e., only a portion of the complete polynucleotide sequence) that is similar between the two polynucleotides, and (2) a sequence that is divergent between the two polynucleotides, sequence comparisons between two (or more) polynucleotides are typically performed by comparing sequences of the two polynucleotides over a “comparison window” to identify and compare local regions of sequence similarity. A “comparison window” refers to a conceptual segment of at least 6 contiguous positions, usually about 50 to about 100, more usually about 100 to about 150 in which a sequence is compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned. The comparison window may comprise additions or deletions (i.e., gaps) of about 20% or less as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Optimal alignment of sequences for aligning a comparison window may be conducted by computerized implementations of algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package Release 7.0, Genetics Computer Group, 575 Science Drive Madison, Wis., USA) or by inspection and the best alignment (i.e., resulting in the highest percentage homology over the comparison window) generated by any of the various methods selected. Reference also may be made to the BLAST family of programs as for example disclosed by Altschul et al., 1997, *Nucl. Acids Res.* 25:3389. A detailed discussion of sequence analysis can be found in Unit 19.3 of Ausubel et al., “Current Protocols in Molecular Biology”, John Wiley & Sons Inc, 1994-1998, Chapter 15.

[0147] “Transformation” refers generally to the permanent, heritable alteration in a cell resulting from the uptake and

incorporation of foreign DNA into the host-cell genome; also, the transfer of an exogenous gene from one organism into the genome of another organism.

[0148] By “vector” is meant a polynucleotide molecule, preferably a DNA molecule derived, for example, from a plasmid, bacteriophage, yeast or virus, into which a polynucleotide can be inserted or cloned. A vector preferably contains one or more unique restriction sites and can be capable of autonomous replication in a defined host cell including a target cell or tissue or a progenitor cell or tissue thereof, or be integrable with the genome of the defined host such that the cloned sequence is reproducible. Accordingly, the vector can be an autonomously replicating vector, i.e., a vector that exists as an extra-chromosomal entity, the replication of which is independent of chromosomal replication, e.g., a linear or closed circular plasmid, an extra-chromosomal element, a mini-chromosome, or an artificial chromosome. The vector can contain any means for assuring self-replication. Alternatively, the vector can be one which, when introduced into the host cell, is integrated into the genome and replicated together with the chromosome(s) into which it has been integrated. Such a vector may comprise specific sequences that allow recombination into a particular, desired site of the host chromosome. A vector system can comprise a single vector or plasmid, two or more vectors or plasmids, which together contain the total DNA to be introduced into the genome of the host cell, or a transposon. The choice of the vector will typically depend on the compatibility of the vector with the host cell into which the vector is to be introduced. In the present case, the vector is preferably one which is operably functional in a bacterial cell, such as a cyanobacterial cell. The vector can include a reporter gene, such as a green fluorescent protein (GFP), which can be either fused in frame to one or more of the encoded polypeptides, or expressed separately. The vector can also include a selection marker such as an antibiotic resistance gene that can be used for selection of suitable transformants.

[0149] The terms “wild-type” and “naturally occurring” are used interchangeably to refer to a gene or gene product that has the characteristics of that gene or gene product when isolated from a naturally occurring source. A wild type gene or gene product (e.g., a polypeptide) is that which is most frequently observed in a population and is thus arbitrarily designated the “normal” or “wild-type” form of the gene.

[0150] Examples of “biomass” include aquatic or marine biomass, fruit-based biomass such as fruit waste, and vegetable-based biomass such as vegetable waste, among others. Examples of aquatic or marine biomass include, but are not limited to, kelp, giant kelp, seaweed, algae, and marine microflora, microalgae, sea grass, and the like. In certain aspects, biomass does not include fossilized sources of carbon, such as hydrocarbons that are typically found within the top layer of the Earth’s crust (e.g., natural gas, nonvolatile materials composed of almost pure carbon, like anthracite coal, etc).

[0151] Examples of fruit and/or vegetable biomass include, but are not limited to, any source of pectin such as plant peel and pomace including citrus, orange, grapefruit, potato, tomato, grape, mango, gooseberry, carrot, sugar-beet, and apple, among others.

[0152] Examples of polysaccharides, oligosaccharides, monosaccharides or other sugar components of biomass include, but are not limited to, alginate, agar, carrageenan, fucoidan, pectin, glucuronate, mannuronate, mannitol, lyxose, cellulose, hemicellulose, glycerol, xylitol, glucose, mannose,

galactose, xylose, xylan, mannan, arabinan, arabinose, glucuronate, galacturonate (including di- and tri-galacturonates), rhamnose, and the like.

[0153] Certain examples of alginate-derived polysaccharides include saturated polysaccharides, such as β -D-mannuronate, α -L-glucuronate, dialginate, trialginate, pentalginate, hexylginate, heptalginate, octalginate, nonalginate, decalginate, undecalginate, dodecalginate and polyalginate, as well as unsaturated polysaccharides such as 4-deoxy-L-erythro-5-hexoseulose uronic acid, 4-(4-deoxy-beta-D-mann-4-enuronosyl)-D-mannuronate or L-guluronate, 4-(4-deoxy-beta-D-mann-4-enuronosyl)-dialginate, 4-(4-deoxy-beta-D-mann-4-enuronosyl)-trialginate, 4-(4-deoxy-beta-D-mann-4-enuronosyl)-tetralginate, 4-(4-deoxy-beta-D-mann-4-enuronosyl)-pentalginate, 4-(4-deoxy-beta-D-mann-4-enuronosyl)-hexylginate, 4-(4-deoxy-beta-D-mann-4-enuronosyl)-heptalginate, 4-(4-deoxy-beta-D-mann-4-enuronosyl)-octalginate, 4-(4-deoxy-beta-D-mann-4-enuronosyl)-nonalginate, 4-(4-deoxy-beta-D-mann-4-enuronosyl)-undecalginate, and 4-(4-deoxy-beta-D-mann-4-enuronosyl)-dodecalginate.

[0154] Certain examples of pectin-derived polysaccharides include saturated polysaccharides, such as galacturonate, digalacturonate, trigalacturonate, tetragalacturonate, pentagalacturonate, hexagalacturonate, heptagalacturonate, octagalacturonate, nonagalacturonate, decagalacturonate, dodecagalacturonate, polygalacturonate, and rhamnopolygalacturonate, as well as saturated polysaccharides such as 4-deoxy-L-threo-5-hexosulose uronate, 4-(4-Deoxy-alpha-D-gluc-4-enuronosyl)-D-galacturonate, 4-(4-Deoxy-alpha-D-gluc-4-enuronosyl)-D-digalacturonate, 4-(4-Deoxy-alpha-D-gluc-4-enuronosyl)-D-trigalacturonate, 4-(4-Deoxy-alpha-D-gluc-4-enuronosyl)-D-tetragalacturonate, 4-(4-Deoxy-alpha-D-gluc-4-enuronosyl)-D-pentagalacturonate, 4-(4-Deoxy-alpha-D-gluc-4-enuronosyl)-D-hexagalacturonate, 4-(4-Deoxy-alpha-D-gluc-4-enuronosyl)-D-heptagalacturonate, 4-(4-Deoxy-alpha-D-gluc-4-enuronosyl)-D-octagalacturonate, 4-(4-Deoxy-alpha-D-gluc-4-enuronosyl)-D-nongalacturonate, 4-(4-Deoxy-alpha-D-gluc-4-enuronosyl)-D-decagalacturonate, and 4-(4-Deoxy-alpha-D-gluc-4-enuronosyl)-D-dodecagalacturonate.

[0155] These polysaccharide or oligosaccharide components may be converted into “suitable monosaccharides” or other “suitable saccharides,” such as “suitable oligosaccharides,” by the microorganisms described herein which are capable of growing on such polysaccharides or other sugar components as a source of carbon (e.g., a sole source of carbon).

[0156] A “suitable monosaccharide” or “suitable saccharide” refers generally to any saccharide that may be produced by a recombinant microorganism growing on pectin, alginate, or other saccharide (e.g., galacturonate, cellulose, hemi-cellulose etc.) as a source or sole source of carbon, and also refers generally to any saccharide that may be utilized in a biofuel biosynthesis pathway of the present invention to produce hydrocarbons such as biofuels or biopetrols. Examples of suitable monosaccharides or oligosaccharides include, but are not limited to, 2-keto-3-deoxy D-gluconate (KDG), D-mannitol, gluconate, mannuronate, mannitol, lyxose, glycerol, xylitol, glucose, mannose, galactose, xylose, arabinose, glucuronate, galacturonates, and rhamnose, and the like. As noted herein, a “suitable monosaccharide” or “suitable saccharide” as used herein may be produced by an engineered or

recombinant microorganism of the present invention, or may be obtained from commercially available sources.

[0157] The recitation “commodity chemical” as used herein includes any saleable or marketable chemical that can be produced either directly or as a by-product of the methods provided herein, including biofuels and/or biopetrols. General examples of “commodity chemicals” include, but are not limited to, biofuels, minerals, polymer precursors, fatty alcohols, surfactants, plasticizers, and solvents. The recitation “biofuels” as used herein includes solid, liquid, or gas fuels derived, at least in part, from a biological source, such as a recombinant microorganism.

[0158] Examples of commodity chemicals include, but are not limited to, methane, methanol, ethane, ethene, ethanol, n-propane, 1-propene, 1-propanol, propanal, acetone, propionate, n-butane, 1-butene, 1-butanol, butanal, butanoate, isobutanol, isobutanol, 2-methylbutanal, 2-methylbutanol, 3-methylbutanal, 3-methylbutanol, 2-butene, 2-butanol, 2-butanone, 2,3-butanediol, 3-hydroxy-2-butanone, 2,3-butanedione, ethylbenzene, ethenylbenzene, 2-phenylethanol, phenylacetaldehyde, 1-phenylbutane, 4-phenyl-1-butene, 4-phenyl-2-butene, 1-phenyl-2-butene, 1-phenyl-2-butanol, 4-phenyl-2-butanol, 1-phenyl-2-butanone, 4-phenyl-2-butanone, 1-phenyl-2,3-butanediol, 1-phenyl-3-hydroxy-2-butanone, 4-phenyl-3-hydroxy-2-butanone, 1-phenyl-2,3-butanedione, n-pentane, ethylphenol, ethenylphenol, 2-(4-hydroxyphenyl)ethanol, 4-hydroxyphenylacetaldehyde, 1-(4-hydroxyphenyl) butane, 4-(4-hydroxyphenyl)-1-butene, 4-(4-hydroxyphenyl)-2-butene, 1-(4-hydroxyphenyl)-1-butene, 1-(4-hydroxyphenyl)-2-butanol, 4-(4-hydroxyphenyl)-2-butanol, 1-(4-hydroxyphenyl)-2-butanone, 4-(4-hydroxyphenyl)-2-butanone, 1-(4-hydroxyphenyl)-2,3-butanediol, 1-(4-hydroxyphenyl)-3-hydroxy-2-butanone, 4-(4-hydroxyphenyl)-3-hydroxy-2-butanone, 1-(4-hydroxyphenyl)-2,3-butanedione, indolyethane, indolyethene, 2-(indole-3)ethanol, n-pentane, 1-pentene, 1-pentanol, pentanal, pentanoate, 2-pentene, 2-pentanol, 3-pentanol, 2-pentanone, 3-pentanone, 4-methylpentanal, 4-methylpentanol, 2,3-pentanediol, 2-hydroxy-3-pentanone, 3-hydroxy-2-pentanone, 2,3-pentanedione, 2-methylpentane, 4-methyl-1-pentene, 4-methyl-2-pentene, 4-methyl-3-pentene, 4-methyl-2-pentanol, 2-methyl-3-pentanol, 4-methyl-2-pentanone, 2-methyl-3-pentanone, 4-methyl-2,3-pentanediol, 4-methyl-2-hydroxy-3-pentanone, 4-methyl-3-hydroxy-2-pentanone, 4-methyl-2,3-pentanedione, 1-phenylpentane, 1-phenyl-1-pentene, 1-phenyl-2-pentene, 1-phenyl-3-pentene, 1-phenyl-2-pentanol, 1-phenyl-3-pentanol, 1-phenyl-2-pentanone, 1-phenyl-3-pentanone, 1-phenyl-2,3-pentanediol, 1-phenyl-2-hydroxy-3-pentanone, 1-phenyl-3-hydroxy-2-pentanone, 1-phenyl-2,3-pentanedione, 4-methyl-1-phenylpentane, 4-methyl-1-phenyl-1-pentene, 4-methyl-1-phenyl-2-pentene, 4-methyl-1-phenyl-3-pentene, 4-methyl-1-phenyl-2-pentanol, 4-methyl-1-phenyl-3-pentanol, 4-methyl-1-phenyl-2-pentanol, 4-methyl-1-phenyl-3-pentanol, 4-methyl-1-phenyl-2-pentanone, 4-methyl-1-phenyl-3-pentanone, 4-methyl-1-phenyl-2,3-pentanediol, 4-methyl-1-phenyl-2,3-pentanedione, 4-methyl-1-phenyl-3-hydroxy-2-pentanone, 4-methyl-1-phenyl-2-hydroxy-3-pentanone, 1-(4-hydroxyphenyl) pentane, 1-(4-hydroxyphenyl)-1-pentene, 1-(4-hydroxyphenyl)-2-pentene, 1-(4-hydroxyphenyl)-3-pentene, 1-(4-hydroxyphenyl)-2-pentanol, 1-(4-hydroxyphenyl)-3-pentanol, 1-(4-hydroxyphenyl)-2-pentanone, 1-(4-hydroxyphenyl)-3-pentanone, 1-(4-hydroxyphenyl)-2,3-pentanediol, 1-(4-hydroxyphenyl)-2-hydroxy-3-pentanone, 1-(4-

hydroxyphenyl)-3-hydroxy-2-pentanone, 1-(4-hydroxyphenyl)-2,3-pentanedione, 4-methyl-1-(4-hydroxyphenyl) pentane, 4-methyl-1-(4-hydroxyphenyl)-2-pentene, 4-methyl-1-(4-hydroxyphenyl)-3-pentene, 4-methyl-1-(4-hydroxyphenyl)-1-pentene, 4-methyl-1-(4-hydroxyphenyl)-3-pentanol, 4-methyl-1-(4-hydroxyphenyl)-2-pentanol, 4-methyl-1-(4-hydroxyphenyl)-3-pentanone, 4-methyl-1-(4-hydroxyphenyl)-2-pentanone, 4-methyl-1-(4-hydroxyphenyl)-2,3-pentanediol, 4-methyl-1-(4-hydroxyphenyl)-2,3-pentanedione, 4-methyl-1-(4-hydroxyphenyl)-3-hydroxy-2-pentanone, 4-methyl-1-(4-hydroxyphenyl)-2-hydroxy-3-pentanone, 1-indole-3-pentane, 1-(indole-3)-1-pentene, 1-(indole-3)-2-pentene, 1-(indole-3)-3-pentene, 1-(indole-3)-2-pentanol, 1-(indole-3)-3-pentanol, 1-(indole-3)-2-pentanone, 1-(indole-3)-3-pentanone, 1-(indole-3)-2,3-pentanediol, 1-(indole-3)-2-hydroxy-3-pentanone, 1-(indole-3)-3-hydroxy-2-pentanone, 1-(indole-3)-2,3-pentanedione, 4-methyl-1-(indole-3)pentane, 4-methyl-1-(indole-3)-2-pentene, 4-methyl-1-(indole-3)-3-pentene, 4-methyl-1-(indole-3)-1-pentene, 4-methyl-2-(indole-3)-3-pentanol, 4-methyl-1-(indole-3)-2-pentanol, 4-methyl-1-(indole-3)-3-pentanone, 4-methyl-1-(indole-3)-2-pentanone, 4-methyl-1-(indole-3)-2,3-pentanediol, 4-methyl-1-(indole-3)-2,3-pentanedione, 4-methyl-1-(indole-3)-3-hydroxy-2-pentanone, 4-methyl-1-(indole-3)-3-hydroxy-2-pentanone, 4-methyl-1-(indole-3)-3-hydroxy-2-pentanone, 4-methyl-1-(indole-3)-3-hydroxy-2-pentanone, 4-methyl-1-(indole-3)-3-hydroxy-2-pentanone, n-hexane, 1-hexene, 1-hexanol, hexanal, hexanoate, 2-hexene, 3-hexene, 2-hexanol, 3-hexanol, 2-hexanone, 3-hexanone, 2,3-hexanediol, 2,3-hexanedione, 3,4-hexanediol, 3,4-hexanedione, 2-hydroxy-3-hexanone, 3-hydroxy-2-hexanone, 3-hydroxy-4-hexanone, 4-hydroxy-3-hexanone, 2-methylhexane, 3-methylhexane, 2-methyl-2-hexene, 2-methyl-3-hexene, 5-methyl-1-hexene, 5-methyl-2-hexene, 4-methyl-1-hexene, 4-methyl-2-hexene, 3-methyl-3-hexene, 3-methyl-2-hexene, 3-methyl-1-hexene, 2-methyl-3-hexanol, 5-methyl-2-hexanol, 5-methyl-3-hexanol, 2-methyl-3-hexanone, 5-methyl-2-hexanone, 5-methyl-3-hexanone, 2-methyl-3,4-hexanediol, 2-methyl-3,4-hexanedione, 5-methyl-2,3-hexanediol, 5-methyl-2,3-hexanedione, 4-methyl-2,3-hexanediol, 4-methyl-2,3-hexanedione, 2-methyl-3-hydroxy-4-hexanone, 2-methyl-4-hydroxy-3-hexanone, 5-methyl-2-hydroxy-3-hexanone, 5-methyl-3-hydroxy-2-hexanone, 4-methyl-2-hydroxy-3-hexanone, 4-methyl-3-hydroxy-2-hexanone, 2,5-dimethylhexane, 2,5-dimethyl-2-hexene, 2,5-dimethyl-3-hexene, 2,5-dimethyl-3-hexanol, 2,5-dimethyl-3-hexanone, 2,5-dimethyl-3,4-hexanediol, 2,5-dimethyl-3,4-hexanedione, 2,5-dimethyl-3-hydroxy-4-hexanone, 5-methyl-1-phenylhexane, 4-methyl-1-phenylhexane, 5-methyl-1-phenyl-1-hexene, 5-methyl-1-phenyl-2-hexene, 5-methyl-1-phenyl-3-hexene, 4-methyl-1-phenyl-1-hexene, 4-methyl-1-phenyl-2-hexene, 4-methyl-1-phenyl-3-hexene, 5-methyl-1-phenyl-2-hexanol, 5-methyl-1-phenyl-3-hexanol, 4-methyl-1-phenyl-2-hexanol, 4-methyl-1-phenyl-3-hexanol, 5-methyl-1-phenyl-2-hexanone, 5-methyl-1-phenyl-3-hexanone, 4-methyl-1-phenyl-2-hexanone, 4-methyl-1-phenyl-3-hexanone, 5-methyl-1-phenyl-2,3-hexanediol, 4-methyl-1-(4-hydroxyphenyl)hexane, 5-methyl-1-(4-hydroxyphenyl)-1-hexene, 5-methyl-1-(4-hydroxyphenyl)-2-hexene, 5-methyl-1-(4-hydroxyphenyl)-3-hexene, 4-methyl-1-(4-hydroxyphenyl)-1-hexene, 4-methyl-

1-(4-hydroxyphenyl)-2-hexene, 4-methyl-1-(4-hydroxyphenyl)-3-hexene, 5-methyl-1-(4-hydroxyphenyl)-2-hexanol, 5-methyl-1-(4-hydroxyphenyl)-3-hexanol, 4-methyl-1-(4-hydroxyphenyl)-2-hexanol, 4-methyl-1-(4-hydroxyphenyl)-2-hexanone, 5-methyl-1-(4-hydroxyphenyl)-3-hexanone, 4-methyl-1-(4-hydroxyphenyl)-2-hexanone, 4-methyl-1-(4-hydroxyphenyl)-3-hexanone, 5-methyl-1-(4-hydroxyphenyl)-2,3-hexanediol, 4-methyl-1-(4-hydroxyphenyl)-2,3-hexanediol, 5-methyl-1-(4-hydroxyphenyl)-3-hydroxy-2-hexanone, 5-methyl-1-(4-hydroxyphenyl)-2-hydroxy-3-hexanone, 4-methyl-1-(4-hydroxyphenyl)-3-hydroxy-2-hexanone, 4-methyl-1-(4-hydroxyphenyl)-2-hydroxy-3-hexanone, 5-methyl-1-(4-hydroxyphenyl)-2,3-hexanedione, 4-methyl-1-(4-hydroxyphenyl)-2,3-hexanedione, 4-methyl-1-(indole-3)-hexane, 5-methyl-1-(indole-3)-1-hexene, 5-methyl-1-(indole-3)-2-hexene, 5-methyl-1-(indole-3)-3-hexene, 4-methyl-1-(indole-3)-1-hexene, 4-methyl-1-(indole-3)-2-hexanol, 4-methyl-1-(indole-3)-3-hexanol, 5-methyl-1-(indole-3)-2-hexanol, 4-methyl-1-(indole-3)-3-hexanol, 5-methyl-1-(indole-3)-2-hexanone, 5-methyl-1-(indole-3)-3-hexanone, 4-methyl-1-(indole-3)-2-hexanone, 4-methyl-1-(indole-3)-3-hexanone, 5-methyl-1-(indole-3)-2,3-hexanedione, 4-methyl-1-(indole-3)-2,3-hexanedione, 5-methyl-1-(indole-3)-2,3-hexanedione, 5-methyl-1-(indole-3)-3-hydroxy-2-hexanone, 5-methyl-1-(indole-3)-2-hydroxy-3-hexanone, 4-methyl-1-(indole-3)-3-hydroxy-2-hexanone, 4-methyl-1-(indole-3)-2-hydroxy-3-hexanone, 5-methyl-1-(indole-3)-2,3-hexanedione, 4-methyl-1-(indole-3)-2,3-hexanedione, n-heptane, 1-heptene, 1-heptanol, heptanal, heptanoate, 2-heptene, 3-heptene, 2-heptanol, 3-heptanol, 4-heptanol, 2-heptanone, 3-heptanone, 4-heptanone, 2,3-heptanediol, 2,3-heptanedione, 3,4-heptanediol, 3,4-heptanedione, 2-hydroxy-3-heptanone, 3-hydroxy-2-heptanone, 3-hydroxy-4-heptanone, 4-hydroxy-3-heptanone, 2-methylheptane, 3-methylheptane, 6-methyl-2-heptene, 6-methyl-3-heptene, 2-methyl-3-heptene, 2-methyl-2-heptene, 5-methyl-2-heptene, 5-methyl-3-heptene, 3-methyl-3-heptene, 2-methyl-3-heptanol, 2-methyl-4-heptanol, 6-methyl-3-heptanol, 5-methyl-3-heptanol, 3-methyl-4-heptanol, 2-methyl-3-heptanone, 2-methyl-4-heptanone, 6-methyl-3-heptanone, 5-methyl-3-heptanone, 3-methyl-4-heptanone, 2-methyl-3,4-heptanediol, 2-methyl-3,4-heptanedione, 6-methyl-3,4-heptanediol, 6-methyl-3,4-heptanedione, 5-methyl-3,4-heptanediol, 5-methyl-3,4-heptanedione, 2-methyl-3-hydroxy-4-heptanone, 2-methyl-4-hydroxy-3-heptanone, 6-methyl-3-hydroxy-4-heptanone, 6-methyl-4-hydroxy-3-heptanone, 5-methyl-3-hydroxy-4-heptanone, 5-methyl-4-hydroxy-3-heptanone, 2,6-dimethylheptane, 2,5-dimethylheptane, 2,6-dimethyl-2-heptene, 2,6-dimethyl-3-heptene, 2,5-dimethyl-2-heptene, 2,5-dimethyl-3-heptene, 3,6-dimethyl-3-heptene, 2,6-dimethyl-3-heptanol, 2,6-dimethyl-4-heptanol, 2,5-dimethyl-3-heptanol, 2,5-dimethyl-4-heptanol, 2,6-dimethyl-3,4-heptanediol, 2,6-dimethyl-3,4-heptanedione, 2,5-dimethyl-3,4-heptanediol, 2,5-dimethyl-3,4-heptanedione, 2,6-dimethyl-3-hydroxy-4-heptanone, 2,6-dimethyl-4-hydroxy-3-heptanone, 2,5-dimethyl-3-hydroxy-4-heptanone, 2,5-dimethyl-4-hydroxy-3-heptanone, n-octane, 1-octene, 2-octene, 1-octanol, octanal, octanoate, 3-octene, 4-octene, 4-octanol, 4-octanone, 4,5-octanediol, 4,5-octanedione, 4-hydroxy-5-octanone, 2-methyl-3-octene, 2-methyl-4-octene, 7-methyl-3-octene, 3-methyl-3-octene, 3-methyl-4-octene, 6-methyl-3-octene, 2-methyl-4-octanol, 7-methyl-4-octanol,

3-methyl-4-octanol, 6-methyl-4-octanol, 2-methyl-4-octanone, 7-methyl-4-octanone, 3-methyl-4-octanone, 6-methyl-4-octanone, 2-methyl-4,5-octanediol, 2-methyl-4,5-octanedione, 3-methyl-4,5-octanediol, 3-methyl-4,5-octanedione, 2-methyl-4-hydroxy-5-octanone, 2-methyl-5-hydroxy-4-octanone, 3-methyl-4-hydroxy-5-octanone, 3-methyl-5-hydroxy-4-octanone, 2,7-dimethyloctane, 2,7-dimethyl-3-octene, 2,7-dimethyl-4-octene, 2,7-dimethyl-4-octanol, 2,7-dimethyl-4-octanone, 2,7-dimethyl-4,5-octanediol, 2,7-dimethyl-4,5-octanedione, 2,7-dimethyl-4-hydroxy-5-octanone, 2,6-dimethyloctane, 2,6-dimethyl-3-octene, 2,6-dimethyl-4-octene, 3,7-dimethyl-3-octene, 2,6-dimethyl-4-octanol, 3,7-dimethyl-4-octanol, 2,6-dimethyl-4-octanone, 3,7-dimethyl-4-octanone, 2,6-dimethyl-4,5-octanediol, 2,6-dimethyl-4,5-octanedione, 2,6-dimethyl-4-hydroxy-5-octanone, 2,6-dimethyl-5-hydroxy-4-octanone, 3,6-dimethyloctane, 3,6-dimethyl-3-octene, 3,6-dimethyl-4-octene, 3,6-dimethyl-4-octanol, 3,6-dimethyl-4-octanone, 3,6-dimethyl-4,5-octanedione, 3,6-dimethyl-4-hydroxy-5-octanone, n-nonane, 1-nonene, 1-nonanol, nonanal, nonanoate, 2-methylnonane, 2-methyl-4-nonene, 2-methyl-5-nonene, 8-methyl-4-nonene, 2-methyl-5-nonanol, 8-methyl-4-nonanol, 2-methyl-5-nonanone, 8-methyl-4-nonanone, 8-methyl-4,5-nonanediol, 8-methyl-4,5-nonanedione, 8-methyl-4-hydroxy-5-nonanone, 8-methyl-5-hydroxy-4-nonanone, 2,8-dimethylnonane, 2,8-dimethyl-3-nonene, 2,8-dimethyl-4-nonene, 2,8-dimethyl-5-nonene, 2,8-dimethyl-4-nonanol, 2,8-dimethyl-5-nonanol, 2,8-dimethyl-4-nonanone, 2,8-dimethyl-5-nonanone, 2,8-dimethyl-4,5-nonanediol, 2,8-dimethyl-4,5-nonanedione, 2,8-dimethyl-4-hydroxy-5-nonanone, 3,8-dimethyl-5-hydroxy-4-nonanone, 2,7-dimethylnonane, 3,8-dimethyl-3-nonene, 3,8-dimethyl-4-nonene, 3,8-dimethyl-5-nonene, 3,8-dimethyl-4-nonanol, 3,8-dimethyl-5-nonanol, 3,8-dimethyl-4-nonanone, 3,8-dimethyl-5-nonanone, 3,8-dimethyl-4,5-nonanediol, 3,8-dimethyl-4,5-nonanedione, 3,8-dimethyl-4-hydroxy-5-nonanone, 3,8-dimethyl-5-hydroxy-4-nonanone, n-decane, 1-decene, 1-decanol, decanoate, 2,9-dimethyldecane, 2,9-dimethyl-3-decene, 2,9-dimethyl-4-decene, 2,9-dimethyl-5-decanol, 2,9-dimethyl-5-decanone, 2,9-dimethyl-5,6-decanediol, 2,9-dimethyl-6-hydroxy-5-decanone, 2,9-dimethyl-5,6-decanedione, 1-undecene, 1-undecanol, undecanal, undecanoate, n-dodecane, 1-dodecene, 1-dodecanol, dodecanal, dodecanoate, n-dodecane, 1-decdecene, 1-dodecanol, ddodecanal, dodecanoate, n-tridecane, 1-tridecene, 1-tridecanol, tridecanal, tridecanoate, n-tetradecane, 1-tetradecene, 1-tetradecanol, tetradecanal, tetradecanoate, n-pentadecane, 1-pentadecene, 1-pentadecanol, pentadecanal, pentadecanoate, n-hexadecane, 1-hexadecene, 1-hexadecanol, hexadecanal, hexadecanoate, n-heptadecane, 1-heptadecene, 1-heptadecanol, heptadecanal, heptadecanoate, n-octadecane, 1-octadecene, 1-octadecanol, octadecanal, octadecanoate, n-nonadecane, 1-nonadecene, 1-nonadecanol, nonadecanal, nonadecanoate, eicosane, 1-eicosene, 1-eicosanol, eicosanal, eicosanoate, 3-hydroxy propanal, 1,3-propanediol, 4-hydroxybutanal, 1,4-butanediol, 3-hydroxy-2-butanone, 2,3-butanediol, 1,5-pentane diol, homocitrate, homoisocitrate, b-hydroxy adipate, glutarate, glutarsemialdehyde, glutaraldehyde, 2-hydroxy-1-cyclopentanone, 1,2-cyclopentanediol, cyclopentanone, cyclopentanol, (S)-2-acetolactate, (R)-2,3-Dihydroxy-isovalerate, 2-oxoisovalerate, isobutyryl-CoA, isobutyrate, isobutyraldehyde, 5-amino pentaldehyde, 1,10-diaminododecane, 1,10-diamino-5-decene, 1,10-diamino-5-

hydroxydecane, 1,10-diamino-5-decanone, 1,10-diamino-5,6-decanediol, 1,10-diamino-6-hydroxy-5-decanone, phenylacetaldehyde, 1,4-diphenylbutane, 1,4-diphenyl-1-butene, 1,4-diphenyl-2-butene, 1,4-diphenyl-2-butanol, 1,4-diphenyl-2-butanone, 1,4-diphenyl-2,3-butanediol, 1,4-diphenyl-3-hydroxy-2-butanone, 1-(4-hydroxyphenyl)-4-phenylbutane, 1-(4-hydroxyphenyl)-4-phenyl-1-butene, 1-(4-hydroxyphenyl)-4-phenyl-2-butene, 1-(4-hydroxyphenyl)-4-phenyl-2-butanol, 1-(4-hydroxyphenyl)-4-phenyl-2-butanone, 1-(4-hydroxyphenyl)-4-phenyl-2,3-butanediol, 1-(4-hydroxyphenyl)-4-phenyl-3-hydroxy-2-butanone, 1-(indole-3)-4-phenylbutane, 1-(indole-3)-4-phenyl-1-butene, 1-(indole-3)-4-phenyl-2-butene, 1-(indole-3)-4-phenyl-2-butanol, 1-(indole-3)-4-phenyl-2-butanone, 1-(indole-3)-4-phenyl-2,3-butanediol, 1-(indole-3)-4-phenyl-3-hydroxy-2-butanone, 4-hydroxyphenylacetaldehyde, 1,4-di(4-hydroxyphenyl)butane, 1,4-di(4-hydroxyphenyl)-1-butene, 1,4-di(4-hydroxyphenyl)-2-butene, 1,4-di(4-hydroxyphenyl)-2-butanol, 1,4-di(4-hydroxyphenyl)-2-butanone, 1,4-di(4-hydroxyphenyl)-2,3-butanediol, 1,4-di(4-hydroxyphenyl)-3-hydroxy-2-butanone, 1-(4-hydroxyphenyl)-4-(indole-3)-butane, 1-(4-hydroxyphenyl)-4-(indole-3)-1-butene, 1-di(4-hydroxyphenyl)-4-(indole-3)-2-butene, 1-(4-hydroxyphenyl)-4-(indole-3)-2-butanol, 1-(4-hydroxyphenyl)-4-(indole-3)-2-butanone, 1-(4-hydroxyphenyl)-4-(indole-3)-2,3-butanediol, 1-(4-hydroxyphenyl)-4-(indole-3)-3-hydroxy-2-butanone, indole-3-acetaldehyde, 1,4-di(indole-3)butane, 1,4-di(indole-3)-1-butene, 1,4-di(indole-3)-2-butene, 1,4-di(indole-3)-2-butanol, 1,4-di(indole-3)-2-butanone, 1,4-di(indole-3)-2,3-butanediol, 1,4-di(indole-3)-3-hydroxy-2-butanone, succinate semialdehyde, hexane-1,8-dicarboxylic acid, 3-hexene-1,8-dicarboxylic acid, 3-hydroxy-hexane-1,8-dicarboxylic acid, 3-hexanone-1,8-dicarboxylic acid, 3,4-hexanediol-1,8-dicarboxylic acid, 4-hydroxy-3-hexanone-1,8-dicarboxylic acid, fucoidan, iodine, chlorophyll, carotenoid, calcium, magnesium, iron, sodium, potassium, phosphate, and the like.

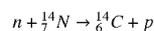
[0159] The recitation “optimized” as used herein refers to a pathway, gene, polypeptide, enzyme, or other molecule having an altered biological activity, such as by the genetic alteration of a polypeptide’s amino acid sequence or by the alteration/modification of the polypeptide’s surrounding cellular environment, to improve its functional characteristics in relation to the original molecule or original cellular environment (e.g., a wild-type sequence of a given polypeptide or a wild-type microorganism). Any of the polypeptides or enzymes described herein may be optionally “optimized,” and any of the genes or nucleotide sequences described herein may optionally encode an optimized polypeptide or enzyme. Any of the pathways described herein may optionally contain one or more “optimized” enzymes, or one or more nucleotide sequences encoding for an optimized enzyme or polypeptide.

[0160] Typically, the improved functional characteristics of the polypeptide, enzyme, or other molecule relate to the suitability of the polypeptide or other molecule for use in a biological pathway (e.g., a biosynthesis pathway, a C—C ligation pathway) to convert a monosaccharide or oligosaccharide into a biofuel. Certain embodiments, therefore, contemplate the use of “optimized” biological pathways. An exemplary “optimized” polypeptide may contain one or more alterations or mutations in its amino acid coding sequence (e.g., point mutations, deletions, addition of heterologous sequences) that facilitate improved expression and/or stabil-

ity in a given microbial system or microorganism, allow regulation of polypeptide activity in relation to a desired substrate (e.g., inducible or repressible activity), modulate the localization of the polypeptide within a cell (e.g., intracellular localization, extracellular secretion), and/or effect the polypeptide’s overall level of activity in relation to a desired substrate (e.g., reduce or increase enzymatic activity). A polypeptide or other molecule may also be “optimized” for use with a given microbial system or microorganism by altering one or more pathways within that system or organism, such as by altering a pathway that regulates the expression (e.g., up-regulation), localization, and/or activity of the “optimized” polypeptide or other molecule, or by altering a pathway that minimizes the production of undesirable by-products, among other alterations. In this manner, a polypeptide or other molecule may be “optimized” with or without altering its wild-type amino acid sequence or original chemical structure. Optimized polypeptides or biological pathways may be obtained, for example, by direct mutagenesis or by natural selection for a desired phenotype, according to techniques known in the art.

[0161] In certain aspects, “optimized” genes or polypeptides may comprise a nucleotide coding sequence or amino acid sequence that is 50% to 99% identical (including all integers in between) to the nucleotide or amino acid sequence of a reference (e.g., wild-type) gene or polypeptide. In certain aspects, an “optimized” polypeptide or enzyme may have about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 20, 30, 40, 50, 100 (including all integers and decimal points in between e.g., 1.2, 1.3, 1.4, 1.5, 5.5, 5.6, 5.7, 60, 70, etc.), or more times the biological activity of a reference polypeptide.

[0162] Certain aspects of the invention also include a commodity chemical, such as a biofuel, that is produced according to the methods and recombinant microorganisms described herein. Such a biofuel (e.g., medium to long chain alkane) may be distinguished from other fuels, such as those fuels produced by traditional refinery from crude carbon sources, by radio-carbon dating techniques. For instance, carbon has two stable, nonradioactive isotopes: carbon-12 (^{12}C), and carbon-13 (^{13}C). In addition, there are trace amounts of the unstable isotope carbon-14 (^{14}C) on Earth. Carbon-14 has a half-life of 5730 years, and would have long ago vanished from Earth were it not for the unremitting impact of cosmic rays on nitrogen in the Earth’s atmosphere, which create more of this isotope. The neutrons resulting from the cosmic ray interactions participate in the following nuclear reaction on the atoms of nitrogen molecules (N_2) in the atmospheric air:



[0163] Plants and other photosynthetic organisms take up atmospheric carbon dioxide by photosynthesis. Since many plants are ingested by animals, every living organism on Earth is constantly exchanging carbon-14 with its environment for the duration of its existence. Once an organism dies, however, this exchange stops, and the amount of carbon-14 gradually decreases over time through radioactive beta decay.

[0164] Most hydrocarbon-based fuels, such as crude oil and natural gas derived from mining operations, are the result of compression and heating of ancient organic materials (i.e., kerogen) over geological time. Formation of petroleum typically occurs from hydrocarbon pyrolysis, in a variety of

mostly endothermic reactions at high temperature and/or pressure. Today's oil formed from the preserved remains of prehistoric zooplankton and algae, which had settled to a sea or lake bottom in large quantities under anoxic conditions (the remains of prehistoric terrestrial plants, on the other hand, tended to form coal). Over geological time the organic matter mixed with mud, and was buried under heavy layers of sediment resulting in high levels of heat and pressure (known as diagenesis). This process caused the organic matter to chemically change, first into a waxy material known as kerogen which is found in various oil shales around the world, and then with more heat into liquid and gaseous hydrocarbons in a process known as catagenesis. Most hydrocarbon based fuels derived from crude oil have been undergoing a process of carbon-14 decay over geological time, and, thus, will have little to no detectable carbon-14. In contrast, certain biofuels produced by the living microorganisms of the present invention will comprise carbon-14 at a level comparable to all other presently living things (i.e., an equilibrium level). In this manner, by measuring the carbon-12 to carbon-14 ratio of a hydrocarbon-based biofuel of the present invention, and comparing that ratio to a hydrocarbon based fuel derived from crude oil, the biofuels produced by the methods provided herein can be structurally distinguished from typical sources of hydrocarbon based fuels.

[0165] Embodiments of the present invention include methods for converting a polysaccharide to a suitable monosaccharide comprising, (a) obtaining the polysaccharide; and (b) contacting the polysaccharide with a recombinant microorganism or microbial system comprising such a microorganism for a time sufficient to convert the polysaccharide to a suitable monosaccharide, wherein the microbial system comprises, (i) at least one gene encoding and expressing an enzyme selected from a lyase and a hydrolase, wherein the lyase and/or hydrolase optionally comprises at least one signal peptide or at least one autotransporter domain; (ii) at least one gene encoding and expressing an enzyme selected from a monosaccharide transporter, a disaccharide transporter, a trisaccharide transporter, an oligosaccharide transporter, and a polysaccharide transporter; and (iii) at least one gene encoding and expressing an enzyme selected from a monosaccharide dehydrogenase, an isomerase, a dehydratase, a kinase, and an aldolase, thereby converting the polysaccharide to a suitable monosaccharide.

[0166] Alternatively, certain aspects may include methods for converting a polysaccharide to a suitable monosaccharide comprising, (a) obtaining the polysaccharide; and (b) contacting the polysaccharide with a microbial system for a time sufficient to convert the polysaccharide to a suitable monosaccharide, wherein the microbial system comprises, (i) at least one gene encoding and expressing an enzyme selected from a lyase and a hydrolase; (ii) at least one gene encoding and expressing a superchannel; and (iii) at least one gene encoding and expressing an enzyme selected from a monosaccharide dehydrogenase, an isomerase, a dehydratase, a kinase, and an aldolase, thereby converting the polysaccharide to a suitable monosaccharide.

[0167] In certain embodiments, a microbial system or isolated microorganism is capable of growing using a polysaccharide (e.g., alginate, pectin, etc.) as a sole source of carbon and/or energy. A "sole source of carbon" refers generally to the ability to grow on a given carbon source as the only carbon source in a given growth medium.

[0168] With regard to alginate, approximately 50 percent of seaweed dry-weight comprises various sugar components, among which alginate and mannitol are major components corresponding to 30 and 15 percent of seaweed dry-weight, respectively. With regard to pectin, although microorganisms such as *E. coli* are generally considered as a host organisms in synthetic biology, and although such microorganism are able to metabolize mannitol, they completely lack the ability to degrade and metabolize alginate. In this regard, many laboratory or wild-type microorganisms, such as *E. coli*, are unable to grow on alginate as a sole source of carbon. Similarly, many organisms such as *E. coli* are unable to degrade and metabolize pectin, a polysaccharide found in many food waste products, and, thus are unable to grown on pectin as a sole source of carbon. Accordingly, embodiments of the present application include engineered microorganisms, such as *E. coli*, or microbial systems containing such engineered microorganisms, that are capable of using polysaccharides, such as alginate and pectin, as a sole source of carbon and/or energy.

[0169] Alginate is a block co-polymer of β -D-mannuronate (M) and α -D-gluronate (G) (M and G are epimeric about the C5-carboxyl group). Each alginate polymer comprises regions of all M (polyM), all G (polyG), and/or the mixture of M and G (polyMG). To utilize alginate to produce one or more suitable monosaccharides, certain aspects of the present invention provide an engineered or recombinant microorganism or microbial system that is able to degrade or depolymerize alginate and to use it as a source of carbon and/or energy. As one means of accomplishing this purpose, such recombinant microorganisms may incorporate a set of polysaccharide degrading or depolymerizing enzymes such as alginate lyases (ALs) to the microbial system.

[0170] ALs are mainly classified into two distinctive sub-families depending on their acts of catalysis: endo-(EC 4.2.2.3) and exo-acting (EC 4.2.2.-) ALs. Endo-acting ALs are further classified based on their catalytic specificity; M specific and G specific ALs. The endo-acting ALs randomly cleave alginate via a 1-elimination mechanism and mainly depolymerize alginate to di-, tri- and tetrasaccharides. The uronate at the non-reducing terminus of each oligosaccharide are converted to unsaturated sugar uronate, 4-deoxy- α -L-erythro-hex-4-ene pyranosyl uronates. The exo-acting ALs catalyze further depolymerization of these oligosaccharides and release unsaturated monosaccharides, which may be non-enzymatically converted to monosaccharides, including α -keto acid, 4-deoxy- α -L-erythro-hexoselulose uronate (DEHU). Certain embodiments of an engineered microbial system or isolated, engineered microorganism may include endoM-, endoG- and exo-acting ALs to degrade or depolymerize aquatic or marine-biomass polysaccharides such as alginate to a monosaccharide such as DEHU.

[0171] Embodiments of the present invention may also include lyases such as alginate lyases isolated from various sources, including, but not limited to, marine algae, mollusks, and wide varieties of microbes such as genus *Pseudomonas*, *Vibrio*, and *Sphingomonas*. Many alginate lyases are endo-acting M specific, several are G specific, and few are exo-acting. For example, ALs isolated from *Sphingomonas* sp. strain Al include five endo-acting ALs, Al-I, Al-II, Al-II', Al-III, and Al-IV' and an exo-acting AL, Al-IV.

[0172] Typically, Al-I, Al-II, and Al-III have molecular weights of 66 kDa, 25 kDa, and 40 kDa, respectively. Al-II and Al-III are self-splicing products of Al-1. Al-II may be

more specific to G and Al-III may be specific to M. Al-I may have high activity for both M and G. Al-IV has molecular weight of about 85 kDa and catalyzes exo-lytic depolymerization of oligoalginate. Although both Al-II' and Al-IV' are functional homologues of Al-II and Al-IV. Al-II' has endolytic activity and may have no preference to M or G. Al-IV has primarily endo-lytic activity. In addition to these ALs, exolytic AL Atu3025 derived from *Agrobacterium tumefaciens* has high activity for depolymerization of oligoalginate, and may be used in certain embodiments of the present invention. Certain embodiments may incorporate into the microbial system or isolated microorganism the genes encoding Al-I, Al-II', Al-IV, and Atu3025, and may include optimal codon usage for the suitable host organisms, such as *E. coli*.

[0173] Certain examples of alginate lyases or oligoalginate lyases that may be utilized herein include enzymes or polypeptides sharing at least 60%, 70%, 80%, 90%, 95%, 98%, or more sequence identity (including all integers in between) to SEQ ID NOS:67-68, which show the nucleotide (SEQ ID NO:67) and polypeptide (SEQ ID NO:68) sequences of oligoalginate lyase Atu3025 isolated from *Agrobacterium tumefaciens*. Certain examples of alginate lyases that may be utilized herein include enzymes or polypeptides sharing at least 60%, 70%, 80%, 90%, 95%, 98%, or more sequence identity (including all integers in between) to the alginate lyase enzymes described in FIG. 37, as well as the secreted alginate lyase encoded by Vs24254 from *Vibrio splendidus*.

[0174] In certain embodiments, a microbial system or recombinant microorganism may be engineered to secrete or display the lyases or alginate lyases (ALs) to the culture media, such as by incorporating a signal peptide or autotransporter domain into the lyase. In this regard, it is typically understood that bacteria have at least four different types of protein secretion machinery (type I, II, III and IV). For example, in *E. coli*, the type II secretion machinery is used for the secretion of recombinant proteins. The type II secretion machinery may comprise a two-step process: the translocation of premature proteins tagged with signal peptides to the periplasm fraction and processing to the mature proteins followed by secretion to media.

[0175] The first process may proceed by any of three different pathways: secB-dependent pathway, signal recognition particle (SRP) pathway, or twin-arginine translocation (TAT) pathway. Recombinant proteins may be secreted into periplasm fraction. The fates of the mature proteins vary dependent on the type of proteins. For example, some proteins are secreted spontaneously by diffusion or passively by a secretion apparatus named secretion that consists of 12-16 proteins, and others stay in periplasm fraction and are eventually degraded.

[0176] Some proteins may also be secreted by an autotransporter apparatus, such as by utilizing an autotransporter domain. The proteins secreted by autotransporter domains typically comprise an N-terminal signal peptide that plays a role in translocation to the periplasm, which may be mediated by secB or SRP pathways, passenger domain, and/or C-terminal translocation unit (UT) having a characteristic β -barrel structure. The β -barrel portion of the UT builds an aqueous pore channel across the outer membrane and helps the transportation of passenger domain to media. Autodisplayed passenger proteins are often cleaved by the autotransporter and set free to media.

[0177] The type I secretion machinery may also be used for the secretion of recombinant proteins in *E. coli*. The type I

secretion machinery may be used for the secretion of high-molecular-weight toxins and exoenzymes. The type I secretion machinery consist of two inner membrane proteins (HlyB and HlyD) that are the member of the ATP binding cassette (ABC) transporter family, and an endogenous outer membrane protein (TolC). The secretion of recombinant proteins based on type I secretion machinery may utilize the C-terminal region of α -haemolysin (HlyA) as a signal sequence. The recombinant proteins may readily pass through the inner membrane, periplasm, and outer membrane through the type I secretion machinery.

[0178] Depending on the types of linker and signal peptides utilized by various embodiments of the present application, both autotransporter and type I secretion machinery can be altered to the cell surface display machinery. Alternatively, a system specific to cell surface display may be used. For example, in this system, target proteins may be fused to PgsA protein (a poly- γ -glutamate synthetase complex) that is natively displayed on the surface of *Bacillus subtilis*.

[0179] Certain embodiments may include lyases such as alginate lyases fused with various signal peptides and/or autotransporter domains found in proteins secreted by both type I and type II secretion machinery. Other embodiments may include lyases such as alginate lyases fused with any combination of signal peptides and or autotransporter domains found in proteins secreted transport machinery as described herein or known to a person skilled in the art. Embodiments may also include signal peptides or autotransporter domains that are experimentally redesigned to maximize the secretion of lyases such as alginate lyases to the culture media, and may also include the use of many different linker sequences that fuse signal peptides, lyases, and autotransporters that improve the efficiency of secretion or the cell surface presentation of lyases.

[0180] Certain embodiments may include a microbial system or isolated microorganism that comprise saccharide transporters, which are able to transport monosaccharides (e.g., DEHU) and oligosaccharides from the media to the cytosol to efficiently utilize these monosaccharides as a source of carbon and/or energy. For instance, genes encoding monosaccharide permeases (i.e., monosaccharide transporters) such as DEHU permeases may be isolated from bacteria that grow on polysaccharides such as alginate as a source of carbon and/or energy, and may be incorporated into embodiments of the present microbial system or isolated microorganism. As an additional example, embodiments may also include redesigned native permeases or transporters with altered specificity for monosaccharide (e.g., DEHU) transportation.

[0181] In this regard, *E. coli* contains several permeases able to transport monosaccharides, which include, but are not limited to, KdgT for 2-keto-3-deoxy-D-gluconate (KDG) transporter, ExuT for aldohexuronates such as D-galacturonate and D-glucuronate transporter, GntT, GntU, GntP, and GntT for gluconate transporter, and KgtP for proton-driven α -ketoglutarate transporter. Microbial systems or recombinant microorganisms described herein may comprise any of these permeases, in addition to those permeases known to a person of skill in the art and not mentioned herein, and may also include permease enzymes redesigned to transport other monosaccharides, such as DEHU.

[0182] A microbial system or recombinant microorganism according to the present invention may also comprise permeases/transporters/superchannels/porins that catalyze the

transport of monosaccharides (e.g., D-mannuronate and D-lyxose) from media to the periplasm or cytosol of a microorganism. For example, genes encoding the permeases of D-mannuronate in soil *Aeromonas* may be incorporated into a microbial system as described herein.

[0183] As one alternative example, a microbial system or microorganism may comprise native permeases/transporters that are redesigned to alter their specificity for efficient monosaccharide transportation, such as for D-mannuronate and D-lyxose transportation. For instance, *E. coli* contains several permeases that are able to transport monosaccharides or sugars such as D-mannuronate and D-lyxose, including KdgT for 2-keto-3-deoxy-D-gluconate (KDG) transporter, ExuT for aldohexuronates such as D-galacturonate and D-gluconate transporter, GntPTU for gluconate/fructuronate transporter, uidB for glucuronide transporter, fucP for L-fucose transporter, galP for galactose transporter, yghK for glycolate transporter, dgoT for D-galactonate transporter, uhpT for hexose phosphate transporter, dctA for orotate/citrate transporter, gntUT for gluconate transporter, malEGF for maltose transporter: alsABC for D-allose transporter, idnT for L-idonate/D-gluconate transporter, KgtP for proton-driven α -ketoglutarate transporter, lacY for lactose/galactose transporter, xylEFGH for D-xylose transporter, araEFGH for L-arabinose transporter, and rbsABC for D-ribose transporter. In certain embodiments, a microbial system or recombinant microorganism may comprise permeases or transporters as described above, including those that are re-designed or optimized for improved transport of certain monosaccharides, such as D-mannuronate, DEHU, and D-lyxose.

[0184] Certain aspects may employ a recombinant microorganism that comprises a "superchannel," by which aquatic or marine-biomass polysaccharides such as alginate polymers, or fruit or vegetable biomass such as pectin polymers, may be directly incorporated into the cytosol and degraded inside the microbial system. For instance, a group of bacteria characterized as Sphingomonads have a wide range in capability of degrading environmentally hazardous compounds such as polychlorinated polycyclic aromatics (dioxin). These bacteria contain characteristic large pleat-like molecules on their cell surfaces. In this regard, certain Sphingomonads have structures characterized as "superchannels" that enable the bacteria to directly take up macromolecules.

[0185] As one particular example of a microorganism comprising a superchannel, *Sphingomonas* sp. strain A1 directly incorporates polysaccharides such as alginate through a superchannel. Such superchannels may consist of a pit on the outer membrane (e.g., AlgR), alginate-binding proteins in the periplasm (e.g., AlgQ1 and Alg Q2), and an ATP-binding cassette (ABC) transporter (e.g., AlgM1, AlgM2, and AlgS). Incorporated polysaccharides such as alginate may be readily depolymerized by lyases such as alginate lyases produced in the cytosol. Thus, certain embodiments may incorporate genes encoding a superchannel (e.g., ccpA, algS, algM1, algM2, algQ1, algQ2) to introduce this ability to the microbial system or recombinant microorganism. Other embodiments may include microorganisms such as *Sphingomonas subarctica* IFO 16058^T, which harbor the plasmid containing genes that encode a superchannel, and which have significantly improved ability to utilize marine or aquatic biomass polysaccharides such as alginate as a source of carbon and/or energy. Certain recombinant microorganisms may employ these superchannel encoding plasmid sequences contained within *Sphingomonas subarctica* IFO 16058^T.

[0186] Certain examples of alginate ABC transporters that may be utilized herein, include ABC transporters Atu3021, Atu3022, Atu3023, Atu3024, algM1, algM2, AlgQ1, AlgQ2, AlgS, OG2516_05558, OG2516_05563, OG2516_05568, and OG2516_05573, including functional variants thereof. Certain examples of alginate symporters that may be utilized herein include symporters V12B01_24239 and V12B01_24194, among others, including functional variants thereof. One additional example of an alginate porin includes V12B01_24269, and variants thereof.

[0187] As noted above, certain embodiments may include recombinant microorganisms that comprise one or more monosaccharide dehydrogenases, isomerases, dehydratases, kinases, and aldolases. With regard to monosaccharide dehydrogenases, certain microbial systems or recombinant microorganism may incorporate enzymes that reduce various monosaccharides (e.g., DEHU, mannuronate) to a monosaccharide that is suitable for biofuel biosynthesis, such as 2-keto-3-deoxy-D-gluconate (KDG) or D-mannitol. Such exemplary enzymes, include, for example, DEHU hydrogenases and mannuronate hydrogenases, in addition to various alcohol dehydrogenases having DEHU hydrogenase and/or mannuronate dehydrogenase activity, such as the novel ADH1 through ADH12 enzymes isolated from *Agrobacterium tumefaciens* C58 (see, e.g., SEQ ID NOS:69-92).

[0188] For more detail on the ADH1 through ADH12 enzymes, SEQ ID NO:69 shows the nucleotide and SEQ ID NO:70 shows the polypeptide sequence of ADH1 Atu1557 isolated from *Agrobacterium tumefaciens* C58. SEQ ID NO:71 shows the nucleotide and SEQ ID NO:72 shows the polypeptide sequence of ADH2 Atu2022 isolated from *Agrobacterium tumefaciens* C58. SEQ ID NO:73 shows the nucleotide and SEQ ID NO:74 shows the polypeptide sequence of ADH3 Atu0626 isolated from *Agrobacterium tumefaciens* C58.

[0189] SEQ ID NO:75 shows the nucleotide and SEQ ID NO:76 shows the polypeptide sequence of ADH4 Atu5240 isolated from *Agrobacterium tumefaciens* C58. SEQ ID NO:77 shows the nucleotide and SEQ ID NO:78 shows the polypeptide sequence of ADH5 Atu3163 isolated from *Agrobacterium tumefaciens* C58. SEQ ID NO:79 shows the nucleotide and SEQ ID NO:80 shows the polypeptide sequence of ADH6 Atu2151 isolated from *Agrobacterium tumefaciens* C58.

[0190] SEQ ID NO:81 shows the nucleotide and SEQ ID NO:82 shows the polypeptide sequence of ADH7 Atu2814 isolated from *Agrobacterium tumefaciens* C58. SEQ ID NO:83 shows the nucleotide and SEQ ID NO:84 shows the polypeptide sequence of ADH8 Atu5447 isolated from *Agrobacterium tumefaciens* C58. SEQ ID NO:85 shows the nucleotide and SEQ ID NO:86 shows the polypeptide sequence of ADH9 Atu4087 isolated from *Agrobacterium tumefaciens* C58.

[0191] SEQ ID NO:87 shows the nucleotide and SEQ ID NO:88 shows the polypeptide sequence of ADH10 Atu4289 isolated from *Agrobacterium tumefaciens* C58.

[0192] SEQ ID NO:89 shows the nucleotide and SEQ ID NO:90 shows the polypeptide sequence of ADH11 Atu3027 isolated from *Agrobacterium tumefaciens* C58. SEQ ID NO:91 shows the nucleotide and SEQ ID NO:92 shows the polypeptide sequence of ADH12 Atu3026 isolated from *Agrobacterium tumefaciens* C58.

[0193] Further examples of enzymes having dehydrogenase activity include Atu3026, Atu3027, OG2516_05543,

OG2516_05538 and V12B01_24244. The microorganisms and methods of the present invention may also utilize biologically active fragments and variants of these hydrogenase enzymes, including optimized variants thereof.

[0194] As a further example, *Pseudomonas* grown using alginate as a sole source of carbon and energy comprises a DEHU hydrogenase enzyme that uses NADPH as a co-factor, is more stable when NADP⁺ is present in the solution, and is active at ambient pH. Thus, certain embodiments of a microbial system or a recombinant microorganism as described herein may incorporate genes encoding hydrogenases such as DEHU or mannuronate hydrogenase derived or obtained from various microbes, in which these microbes may be capable of growing on polysaccharides such as alginate or pectin as a source of carbon and/or energy.

[0195] Certain embodiments may incorporate components of a microbial system or isolated microorganism that is capable of efficiently growing on monosaccharides such as D-mannuronate or D-lyxose as a source of carbon and energy. For instance, both *Aeromonas* and *Aerobacter aerogenes* PRL-R3 comprise genes encoding monosaccharide dehydrogenases such as D-mannuronate hydrogenase and D-lyxose isomerase. Thus, certain microbial systems or recombinant microorganisms may comprise monosaccharide dehydrogenases such as D-mannuronate hydrogenase and D-lyxose isomerase from *Aeromonas*, *Aerobacter aerogenes* PRL-R3, or various other suitable microorganisms, including those microorganisms capable of growing on D-mannuronate or D-lyxose as a source of carbon and energy.

[0196] Certain embodiments may include a microbial system or isolated microorganism with enhanced efficiency for converting monosaccharides such as D-mannuronate and D-xylulose into monosaccharides suitable for a biofuel biosynthesis pathway such as KDG. Merely by way of explanation, D-mannuronate and D-xylulose are metabolites in microbes such as *E. coli*. D-mannuronate is converted by a D-mannuronate dehydratase to KDG. D-xylulose enters the pentose phosphate pathway. Thus, to increase conversion of D-mannuronate to KDG, an exogenous or endogenous D-mannuronate dehydratase (e.g., *uxuA*) gene may be over-expressed in a recombinant microorganism of the invention. Similarly, in other embodiments, suitable endogenous or exogenous genes such as kinases (e.g., *kdgK*), *nad*, as well as KDG aldolases (e.g., *kdgA* and *eda*) may be either incorporated or overexpressed in a given recombinant microorganism (see SEQ ID NOS:93-96), including biologically active variants or fragments thereof, such as optimized variants of these genes. SEQ ID NO:93 shows the nucleotide sequence and SEQ ID NO:94 shows the polypeptide sequence of a 2-keto-deoxy gluconate kinase (*KdgK*) from *Escherichia coli* DH10B. SEQ ID NO:95 shows the nucleotide sequence and SEQ ID NO:96 shows the polypeptide sequence of a 2-keto-deoxy gluconate-6-phosphate aldorase (*KdgA*) from *Escherichia coli* DH10B.

[0197] In certain aspects, as noted above, a recombinant microorganism that is capable of growing on alginate or pectin as a sole source of carbon may utilize a naturally-occurring or endogenous copy of a dehydratase, kinase, and/or aldolase. For instance, *E. coli* contains endogenous dehydratases, kinases, and aldolases that are capable of catalyzing the appropriate steps in the conversion of polysaccharides to a suitable monosaccharide. In these and other related aspects, the naturally-occurring dehydratase or kinase may also be over-expressed, such as by providing an exogenous copy of

the naturally-occurring dehydratase, kinase or aldolase operable linked to a highly constitutive or inducible promoter.

[0198] As one exemplary source of enzymes for engineering a recombinant microorganism to grow on alginate as a sole source of carbon, *Vibrio splendidus* is known to be able to metabolize alginate to support growth. For example, SEQ ID NO:1 shows a secretome region carrying certain *Vibrio splendidus* genes (V12B01_02425 to V12B01_02480), which encodes a type II secretion apparatus. SEQ ID NO:2 shows the nucleotide sequence of an entire genomic region between V12B01_24189 to V12B01_24249, which was derived from *Vibrio splendidus*, and which when transformed into *E. coli* as a fosmid clone was sufficient to confer the ability to grow on alginate as a sole source of carbon. SEQ ID NOS:3-64 show the individual putative genes contained within SEQ ID NO:2. Thus, in certain aspects, a recombinant microorganism (e.g., *E. coli*) that is able to grow on alginate as a sole source of carbon and/or energy may comprise one or more nucleotide or polypeptide reference sequences described in SEQ ID NOS:1-64, including biologically active fragments or variants thereof, such as optimized variants.

[0199] In certain aspects, a recombinant microorganism that is able to grow on alginate as a sole source of carbon may contain certain coding nucleotide or polypeptide sequences contained within SEQ ID NO:2, such as the sequences in SEQ ID NOS:3-64, or biologically active fragments or variants thereof, including optimized variants. These sequences are described in further detail below.

[0200] SEQ ID NO:3 shows the nucleotide coding sequence of the putative protein V12B01_24184. This putative coding sequence is contained within the polynucleotide sequence of SEQ ID NO:2, and encodes a polypeptide that is similar to an autotransporter adhesion or type I secretion target *ggxgxdxxx* (SEQ ID NO:145) repeat. SEQ ID NO:4 shows the polypeptide sequence of putative protein V12B01_24184, encoded by the polynucleotide of SEQ ID NO:3. This putative polypeptide is similar to autotransporter adhesion or type I secretion target *ggxgxdxxx* (SEQ ID NO:145) repeat.

[0201] SEQ ID NO:5 shows the nucleotide sequence that encodes the putative protein V12B01_24189. SEQ ID NO:6 shows the polypeptide sequence of the putative protein V12B01_24189, which is similar to cyclohexadienyl dehydratase.

[0202] SEQ ID NO:7 shows the nucleotide sequence that encodes the putative protein V12B01_24194. SEQ ID NO:8 shows the polypeptide sequence of the putative protein V12B01_24194, which is similar to a Na/proline transporter.

[0203] SEQ ID NO:9 shows the nucleotide sequence that encodes the putative protein V12B01_24199. SEQ ID NO:10 shows the polypeptide sequence of the putative protein V12B01_24199, which is similar to a keto-deoxy-phosphogluconate aldolase.

[0204] SEQ ID NO:11 shows the nucleotide sequence that encodes the putative protein V12B01_24204. SEQ ID NO:12 shows the polypeptide sequence of the putative protein V12B01_24204, which is similar to 2-dehydro-3-deoxy-gluconokinase.

[0205] SEQ ID NO:13 shows the nucleotide sequence that encodes the putative protein V12B01_24209. SEQ ID NO:14 shows the polypeptide sequence of the putative protein V12B01_24209.

[0206] SEQ ID NO:15 shows the nucleotide sequence that encodes the putative protein V12B01_24214. SEQ ID

NO:16 shows the polypeptide sequence of the putative protein V12B01_24214, which is similar to a chondroitin AC/alginase lyase.

[0207] SEQ ID NO:17 shows the nucleotide sequence that encodes the putative protein V12B01_24219. SEQ ID NO:18 shows the polypeptide sequence of the putative protein V12B01_24219, which is similar to a chondroitin AC/alginase lyase.

[0208] SEQ ID NO:19 shows the nucleotide sequence that encodes the putative protein V12B01_24224. SEQ ID NO:20 shows the polypeptide sequence of the putative protein V12B01_24224, which is similar to a 2-keto-4-pentenoate hydratase/2-oxohepta-3-ene-1,7-dioic acid hydratase.

[0209] SEQ ID NO:21 shows the nucleotide sequence that encodes the putative protein V12B01_24229. SEQ ID NO:22 shows the polypeptide sequence of the putative protein V12B01_24229, which is similar to a GntR-family transcriptional regulator.

[0210] SEQ ID NO:23 shows the nucleotide sequence that encodes the putative protein V12B01_24234. SEQ ID NO:24 shows the polypeptide sequence of the putative protein V12B01_24234, which is similar to a Na⁺/proline symporter.

[0211] SEQ ID NO:25 shows the nucleotide sequence that encodes the putative protein V12B01_24239. SEQ ID NO:26 shows the polypeptide sequence of the putative protein V12B01_24239, which is similar to an oligoalginase lyase.

[0212] SEQ ID NO:27 shows the nucleotide sequence that encodes the putative protein V12B01_24244. SEQ ID NO:28 shows the polypeptide sequence of putative protein V12B01_24244, which is similar to a 3-hydroxyisobutyrate dehydrogenase.

[0213] SEQ ID NO:29 shows the nucleotide sequence that encodes the putative protein V12B01_24249. SEQ ID NO:30 shows the polypeptide sequence of the putative protein V12B01_24249, which is similar to a methyl-accepting chemotaxis protein.

[0214] SEQ ID NO:31 shows the nucleotide sequence that encodes the putative protein V12B01_24254. SEQ ID NO:32 shows the polypeptide sequence of putative protein V12B01_24254, which is similar to an alginase lyase.

[0215] SEQ ID NO:33 shows the nucleotide sequence that encodes the putative protein V12B01_24259. SEQ ID NO:34 shows the polypeptide sequence of putative protein V12B01_24259, which is similar to an alginase lyase.

[0216] SEQ ID NO:35 shows the nucleotide sequence that encodes the putative protein V12B01_24264. SEQ ID NO:36 shows the polypeptide sequence of putative protein V12B01_24264.

[0217] SEQ ID NO:37 shows the nucleotide sequence that encodes the putative protein V12B01_24269. SEQ ID NO:38 shows the polypeptide sequence of putative protein V12B01_24269, which is similar to a putative oligogalacturonate specific porin.

[0218] SEQ ID NO:39 shows the nucleotide sequence that encodes the putative protein V12B01_24274. SEQ ID NO:40 shows the polypeptide sequence of putative protein V12B01_24274, which is similar to an alginase lyase.

[0219] FIG. 32 shows the nucleotide coding sequence and polypeptide sequence of putative protein V12B01_02425. FIG. 32A shows the nucleotide sequence that encodes the putative protein V12B01_02425 (SEQ ID NO:41). FIG. 32B

shows the polypeptide sequence of putative protein V12B01_02425 (SEQ ID NO:42), which is similar to a type II secretory pathway component EpsC.

[0220] SEQ ID NO:43 shows the nucleotide sequence that encodes the putative protein V12B01_02430. SEQ ID NO:44 shows the polypeptide sequence of putative protein V12B01_02430, which is similar to a type II secretory pathway component EpsD.

[0221] SEQ ID NO:45 shows the nucleotide sequence that encodes the putative protein V12B01_02435. SEQ ID NO:46 shows the polypeptide sequence of putative protein V12B01_02435, which is similar to a type II secretory pathway component EpsE.

[0222] SEQ ID NO:47 shows the nucleotide sequence that encodes the putative protein V12B01_02440. SEQ ID NO:48 shows the polypeptide sequence of putative protein V12B01_02440, which is similar to a type II secretory pathway component EpsF.

[0223] SEQ ID NO:49 shows the nucleotide sequence that encodes the putative protein V12B01_02445. SEQ ID NO:50 shows the polypeptide sequence of putative protein V12B01_02445, which is similar to a type II secretory pathway component EpsG.

[0224] SEQ ID NO:51 shows the nucleotide sequence that encodes the putative protein V12B01_02450. SEQ ID NO:52 shows the polypeptide sequence of putative protein V12B01_02450, which is similar to a type II secretory pathway component EpsH.

[0225] SEQ ID NO:53 shows the nucleotide sequence that encodes the putative protein V12B01_02455. SEQ ID NO:54 shows the polypeptide sequence of putative protein V12B01_02455, which is similar to a type II secretory pathway component EpsI.

[0226] SEQ ID NO:55 shows the nucleotide sequence that encodes the putative protein V12B01_02460. SEQ ID NO:56 shows the polypeptide sequence of putative protein V12B01_02460, which is similar to a type II secretory pathway component EpsJ.

[0227] SEQ ID NO:57 shows the nucleotide sequence that encodes the putative protein V12B01_02465. SEQ ID NO:58 shows the polypeptide sequence of putative protein V12B01_02465, which is similar to a type II secretory pathway component EpsK.

[0228] SEQ ID NO:59 shows the nucleotide sequence that encodes the putative protein V12B01_02470. SEQ ID NO:60 shows the polypeptide sequence of putative protein V12B01_02470, which is similar to a type II secretory pathway component EpsL.

[0229] SEQ ID NO:61 shows the nucleotide sequence that encodes the putative protein V12B01_02475. SEQ ID NO:62 shows the polypeptide sequence of putative protein V12B01_02475, which is similar to a type II secretory pathway component EpsM.

[0230] SEQ ID NO:63 shows the nucleotide sequence that encodes the putative protein V12B01_02480. SEQ ID NO:64 shows the nucleotide sequence that encodes the putative protein V12B01_02480, which is similar to a type II secretory pathway component EpsC.

[0231] As a further exemplary source of enzymes for engineering a microorganism to grow on alginate, *Agrobacterium tumefaciens* C58 is able to metabolize relatively small sizes of alginate molecules (1000 mers) as a sole source of carbon and energy. Since *A. tumefaciens* C58 has long been used for plant biotechnology, the genetics of this organism has been rela-

tively well studied, and many genetic tools are available and compatible with other gram-negative bacteria such as *E. coli*. Thus, certain aspects may employ this microbe, or the genes therein, for the production of suitable monosaccharides. For instance, as noted above, the present disclosure provides a series of novel ADH genes having both DEHU and mannuronate hydrogenase activity that were obtained from *Agrobacterium tumefaciens* C58 (see SEQ ID NOS: 67-92).

[0232] As noted above, certain aspects may include a recombinant microorganism or microbial system that is capable of growing on pectin as a sole source of carbon and/or energy. Pectin is a linear chain of α -(1-4)-linked D-galacturonic acid that forms the pectin-backbone, a homogalacturonan. Into this backbone, there are regions where galacturonic acid is replaced by (1-2)-linked L-rhamnose. From rhamnose, side chains of various neutral sugars typically branch off. This type of pectin is called rhamnogalacturonan I. Over all, about up to every 25th galacturonic acid in the main chain is exchanged with rhamnose. Some stretches consisting of alternating galacturonic acid and rhamnose—“hairy regions”, others with lower density of rhamnose—“smooth regions.” The neutral sugars mainly comprise D-galactose, L-arabinose and D-xylose; the types and proportions of neutral sugars vary with the origin of pectin. In nature, around 80% of carboxyl groups of galacturonic acid are esterified with methanol. Some plants, like sugar-beet, potatoes and pears, contain pectins with acetylated galacturonic acid in addition to methyl esters. Acetylation prevents gel-formation but increases the stabilising and emulsifying effects of pectin. Certain pectin degradation and metabolic pathways are exemplified in FIG. 3.

[0233] In addition to the genes, enzymes, and biological pathways described above, certain recombinant microorganisms may incorporate features that are useful for growth on pectin as a sole source of carbon. For instance, to degrade and metabolize pectin as a sole source of carbon, pectin methyl and acetyl esterases first catalyze the hydrolysis of methyl and acetyl esters on pectin. Examples of pectin methyl esterases include, but are not limited to, *pemA* and *pmeB*. Examples of pectin acetyl esterases include, but are not limited to, *PaeX* and *PaeY*. Further examples of pectin methyl esterases that may be utilized herein include enzymes or polypeptides sharing at least 60%, 70%, 80%, 90%, 95%, 98%, or more sequence identity (including all integers in between) to the pectate methyl esterases in FIG. 40. Further examples of pectate acetyl esterases that may be utilized herein include enzymes or polypeptides sharing at least 60%, 70%, 80%, 90%, 95%, 98%, or more sequence identity (including all integers in between) to the pectate acetyl esterases described in FIG. 41.

[0234] Further to this end, pectate lyases and hydrolases may catalyze the endolytic cleavage of pectate via β -elimination and hydrolysis, respectively, to produce oligopectates. Other enzymes that may be utilized to metabolize pectin include Examples of pectate lyases include, but are not limited to, *PelA*, *PelB*, *PelC*, *PelD*, *PelE*, *PelF*, *PelI*, *PelL*, and *PelZ*. Examples of pectate hydrolases include, but are not limited to, *PehA*, *PehN*, *PehV*, *PehW*, and *PehX*. Further examples of pectate lyases include polypeptides or enzymes sharing at least 60%, 70%, 80%, 90%, 95%, 98%, or more sequence identity (including all integers in between) to the pectate lyases described in FIG. 38.

[0235] Polygalacturonases, rhamnogalacturonan lyases, and rhamnogalacturonan hydrolases may also be utilized

herein to degrade and metabolize pectin. Examples of rhamnogalacturonan lyases include polypeptides or enzymes sharing at least 60%, 70%, 80%, 90%, 95%, 98%, or more sequence identity (including all integers in between) to the rhamnogalacturonan lyases (i.e., rhamnogalacturonases) described in FIG. 39A. Examples of rhamnogalacturonate hydrolases include polypeptides or enzymes sharing at least 60%, 70%, 80%, 90%, 95%, 98%, or more sequence identity (including all integers in between) to the rhamnogalacturonate hydrolases described in FIG. 39B.

[0236] Thus, to degrade and metabolize pectin, certain of the recombinant microorganisms and methods of the present invention may incorporate one or more of the above noted methyl and acetyl esterases, lyases, and/or hydrolases, among others known in the art. These may enzymes may be encoded and expressed by endogenous or exogenous genes, and may also include biologically active fragments or variants thereof, such as homologs, orthologs, and/or optimized variants of these enzymes.

[0237] To further metabolize the degradation products of pectin, oligopectates may be transported into the periplasm fraction of gram-negative bacteria by outer membrane porins, where they are further degraded into such components as di- and tri-galacturonates. Examples of outer membrane porins include that can transport oligopectates into the periplasm include, but are not limited to, *kdgN* and *kdgM*. Certain recombinant microorganism may incorporate these or similar genes.

[0238] Di- and tri-galacturonates may then be transported into the cytosol for further degradation. Bacteria contain at least two different transporter systems responsible for di- and tri-galacturonate transportation, including symporter and ABC transporter (e.g., *TogT* and *TogMNAB*, respectively). Thus, certain of the recombinant microorganisms provided herein may comprise one or more a di- or tri-galacturonate transporter systems, such as *TogT* and/or *TogMNAB*.

[0239] Once di- and trigalacturonate are incorporated into the cytosol, short pectate or galacturonate lyases, break them down to D-galacturonate and (4S)-4,6-dihydroxy-2,5-dioxohexuronate. Examples of short pectate or galacturonate lyases include, but are not limited to, *PelW* and *Ogl*, which genes may be either endogenously or exogenously incorporated into certain recombinant microorganisms provided herein. D-galacturonate and (4S)-4,6-dihydroxy-2,5-dioxohexuronate are then converted to 5-dehydro-4-deoxy-D-glucuronate and further to KDG, which steps may be catalyzed by *KduI* and *KduD*, respectively. The *KduI* enzyme has an isomerase activity, and the *KduD* enzyme has a dehydrogenase activity, such as a 2-deoxy-D-gluconate 3-dehydrogenase activity. Accordingly, certain recombinant microorganisms provided herein may comprise one or more short pectate or galacturonate lyases, such as *PelW* and/or *Ogl*, and may optionally comprise one or more isomerases, such as *KduI*, as well as one or more dehydrogenases, such as *KduD*, to convert di- and trigalacturonates into a suitable monosaccharide, such as KDG.

[0240] In certain aspects, a recombinant microorganism, such as *E. coli*, that is able to grown on pectin or tri-galacturonate as a sole source of carbon and/or energy may comprise one or more of the gene sequences contained within SEQ ID NOS:65 and 66, including biologically active fragments or variants thereof, such as optimized variants. SEQ ID NO:65 shows the nucleotide sequence of the *kdgF*-*PaeX* region from *Erwinia carotovora* subsp. *Atroseptica* SCRI1043. SEQ ID

NO:66 shows the nucleotide sequence of ogl-kdgR from *Erwinia carotovora* subsp. *Atroseptica* SCRI1043.

[0241] In certain aspects, a recombinant microorganism, such as *E. coli*, that is able to grow on pectin or tri-galacturonate as a sole source of carbon and/or energy may comprise one or more genomic regions of *Erwinia chrysanthemi*, comprising several genes (kdgF, kduI, kduD, pelW, togM, togN, togA, togB, kdgM, paeX, ogl, and kdgR) encoding enzymes (kduI, kduD, ogl, pelW, and paeX), transporters (togM, togN, togA, togB, and kdgM), and regulatory proteins (kdgR) responsible for degradation of di- and trigalacturonate, as well as several genes (pelA, pelE, paeY, and pem) encoding pectate lyases (pelA and pelE), pectin acetylsterases (paeY), and pectin methylsterase (pem) (see Example 2).

[0242] Additional examples of isomerases that may be utilized herein include gluconate isomerases, such as those in the family uxaC, as well as 4-deoxy-L-threo-5-hexylose uronate isomerases, such as those in the family KduI. Additional examples of reductases that may be utilized herein include tagaturonate reductases, such as those in the family uxaB. Additional examples of dehydratases that may be utilized herein include altronate dehydratases, such as those in the family uxaA. Additional examples of dehydrogenases that may be utilized herein include 2-deoxy-D-gluconate 3-dehydrogenases, such as those in the family kduD.

[0243] Certain aspects may also utilize recombinant microorganisms engineered to enhance the efficiency of the KDG degradation pathway. For instance, in bacteria, KDG is a common metabolic intermediate in the degradation of hexuronates such as D-glucuronate and D-galacturonate and enters into Entner Doudoroff pathway where it is converted to pyruvate and glyceraldehyde-3-phosphate (G3P). In this pathway, KDG is first phosphorylated by KDG kinase (KdgK) followed by its cleavage into pyruvate and glyceraldehyde-3-phosphate (G3P) using 2-keto-3-deoxy-D-6-phosphate-gluconate (KDPG) aldolase (KdgA). The expression of these enzymes concurrently with KDG permease (e.g., KdgT) is negatively regulated by KdgR and is almost none at basal level. The expression is dramatically (3-5-fold) induced upon the addition of hexuronates, and a similar result has been reported in *Pseudomonas* grown on alginate. Hence, to increase the conversion of KDG to pyruvate and G3P, the negative regulator KdgR may be removed. To further improve the pathway efficiency, exogenous copies of KdgK and KdgA may also be incorporated into a given recombinant microorganism.

[0244] In certain aspects, a recombinant microorganism that is able to grow on a polysaccharide (e.g., alginate, pectin, etc) as a sole source of carbon may be capable of producing an increased amount of a given commodity chemical (e.g., ethanol) while growing on that polysaccharide. For example, *E. coli* engineered to grow on alginate may be engineered to produce an increased amount of ethanol from alginate as compared to *E. coli* that is not engineered to grow on alginate (see Example 11). Thus, certain aspects include a recombinant microorganism that is capable of growing on alginate or pectin as a sole source carbon, and that is capable of producing an increased amount of ethanol, such as by comprising one or more genes encoding and expressing a pyruvate decarboxylase (pdc) and/or an alcohol dehydrogenase, including functional variants thereof. In certain aspects, such a recombinant microorganism may comprise a pyruvate decarboxylase (pdc) and two alcohol dehydrogenases (adhA and adhB) obtained from *Zymomonas mobilis*.

[0245] Embodiments of the present invention also include methods for converting polysaccharide to a suitable monosaccharide comprising, (a) obtaining a polysaccharide; (b) contacting the polysaccharide with a chemical catalysis or enzymatic pathway, thereby converting the polysaccharide to a first monosaccharide or oligosaccharide; and (c) contacting the first monosaccharide with a microbial system for a time sufficient to convert the first monosaccharide or oligosaccharide to the suitable monosaccharide, wherein the microbial system comprises, (i) at least one gene encoding and expressing an enzyme selected from a monosaccharide transporter, a disaccharide transporter, a trisaccharide transporter, an oligosaccharide transporter, and a polysaccharide transporter; and (ii) at least one gene encoding and expressing an enzyme selected from a monosaccharide dehydrogenase, an isomerase, a dehydratase, a kinase, and an aldolase, thereby converting the polysaccharide to a suitable monosaccharide.

[0246] In certain aspects of the present invention, aquatic or marine-biomass polysaccharides such as alginate may be chemically degraded using chemical catalysts such as acids. Similarly, biomass-derived pectin may be chemically degraded. For instance, the reaction catalyzed by chemical catalysts is typically through hydrolysis, as opposed to the β -elimination type of reactions catalyzed by enzymatic catalysts. Thus, certain embodiments may include boiling alginate or pectin with strong mineral acids to liberate carbon dioxide from D-mannuronate, thereby forming D-lyxose, a common sugar metabolite utilized by many microorganisms. Such embodiments may use, for example, formate, hydrochloric acid, sulfuric acid, in addition to other suitable acids known in the art as chemical catalysts.

[0247] An enzymatic pathway may utilize one or more enzymes described herein that are capable of catalyzing the degradation of polysaccharides, such as alginate or pectin.

[0248] Other embodiments may use variations of chemical catalysis similar to those described herein or known to a person skilled in the art, including improved or redesigned methods of chemical catalysis suitable for use with biomass related polysaccharides. Certain embodiments include those wherein the resulting monosaccharide uronate is D-mannuronate.

[0249] As noted above, the suitable monosaccharides or suitable oligosaccharides produced by the recombinant microorganisms and microbial systems of the present invention may be utilized as a feedstock in the production of commodity chemicals, such as biofuels, as well as commodity chemical intermediates. Thus, certain embodiments of the present invention relate generally to methods for converting a suitable monosaccharide or oligosaccharide to a commodity chemical, such as a biofuel, comprising, (a) obtaining a suitable monosaccharide or oligosaccharide; (b) contacting the suitable monosaccharide or oligosaccharide with a microbial system for a time sufficient to convert to the suitable monosaccharide to the biofuel, thereby converting the suitable monosaccharide to the biofuel.

[0250] Certain aspects include methods for converting a suitable monosaccharide to a first commodity chemical such as a biofuel, comprising, (a) obtaining a suitable monosaccharide; (b) contacting the suitable monosaccharide with a microbial system for a time sufficient to convert to the suitable monosaccharide to the first commodity chemical, wherein the microbial system comprises one or more genes

encoding a aldehyde or ketone biosynthesis pathway, thereby converting the suitable monosaccharide to the first commodity chemical.

[0251] In these and other related aspects, depending on the particular ketone or aldehyde biosynthesis pathway employed, the first commodity chemical may be further enzymatically and/or chemically reduced and dehydrated to a second commodity chemical. Examples of such second commodity chemicals include, but are not limited to, butene or butane; 1-phenylbutene or 1-phenylbutane; pentene or pentane; 2-methylpentene or 2-methylpentane; 1-phenylpentene or 1-phenylpentane; 1-phenyl-4-methylpentene or 1-phenyl-4-methylpentane; hexene or hexane; 2-methylhexene or 2-methylhexane; 3-methylhexene or 3-methylhexane; 2,5-dimethylhexene or 2,5-dimethylhexane; 1-phenylhexene or 1-phenylhexane; 1-phenyl-4-methylhexene or 1-phenyl-4-methylhexane; 1-phenyl-5-methylhexene or 1-phenyl-5-methylhexane; heptene or heptane; 2-methylheptene or 2-methylheptane; 3-methylheptene or 3-methylheptane; 2,6-dimethylheptene or 2,6-dimethylheptane; 3,6-dimethylheptene or 3,6-dimethylheptane; 3-methyloctene or 3-methyloctane; 2-methyloctene or 2-methyloctane; 2,6-dimethyloctene or 2,6-dimethyloctane; 2,7-dimethyloctene or 2,7-dimethyloctane; 3,6-dimethyloctene or 3,6-dimethyloctane; and cyclopentane or cyclopentene.

[0252] Certain embodiments of the present invention may also include methods for converting a suitable monosaccharide or oligosaccharide to a commodity chemical comprising (a) obtaining a suitable monosaccharide or oligosaccharide; (b) contacting the suitable monosaccharide or oligosaccharide with a microbial system for a time sufficient to convert to the suitable monosaccharide or oligosaccharide to the commodity chemical, wherein the microbial system comprises; (i) one or more genes encoding a biosynthesis pathway; (ii) one or more genes encoding and expressing a C—C ligation pathway; and (iii) one or more genes encoding and expressing a reduction and dehydration pathway, comprising a diol dehydrogenase, a diol dehydratase, and a secondary alcohol dehydrogenase, thereby converting the suitable monosaccharide or oligosaccharide to the commodity chemical.

[0253] Certain aspects also include recombinant microorganism that comprise (i) one or more genes encoding a biosynthesis pathway; (ii) one or more genes encoding and expressing a C—C ligation pathway; and (iii) one or more genes encoding and expressing a reduction and dehydration pathway, comprising a diol dehydrogenase, a diol dehydratase, and a secondary alcohol dehydrogenase. Certain aspects also include recombinant microorganisms that comprise the above pathways individually or in certain combinations, such as recombinant microorganism that comprises one or more genes encoding a biosynthesis pathway, as described herein. Certain aspects may also include recombinant microorganisms that comprise one or more genes encoding and expressing a C—C ligation pathway, as described herein. Certain aspects may also include recombinant microorganisms that comprise one or more genes encoding and expressing a reduction and dehydration pathway, comprising a diol dehydrogenase, a diol dehydratase, and a secondary alcohol dehydrogenase, as described herein.

[0254] As for recombinant microorganisms that comprise combinations of the above-noted pathways, certain aspects may include recombinant microorganisms that comprise (i) one or more genes encoding a biosynthesis pathway; and (ii) one or more genes encoding and expressing a C—C ligation

pathway. Certain aspects may also include recombinant microorganisms that comprise (i) one or more genes encoding and expressing a C—C ligation pathway; and (ii) one or more genes encoding and expressing a reduction and dehydration pathway, comprising a diol dehydrogenase, a diol dehydratase, and a secondary alcohol dehydrogenase.

[0255] Certain aspects may also include recombinant microorganisms that comprise one or more individual components of a dehydration and reduction pathway, such as a recombinant microorganism that comprises a diol dehydrogenase, a diol dehydratase, or a secondary alcohol dehydrogenase. These and other microorganisms may be utilized, for example, to convert a suitable polysaccharide to a first commodity chemical, or an intermediate thereof, or to convert a first commodity chemical, or an intermediate thereof, to a second commodity chemical.

[0256] Merely by way of illustration, a recombinant microorganism comprising a C—C ligation pathway may be utilized to convert butanal into a first commodity chemical, or an intermediate thereof, such as 5-hydroxy-4-octanone, which can then be converted into a second commodity chemical, or intermediate thereof, by any suitable pathway. As a further example, a recombinant microorganism comprising a C—C ligation pathway and a diol hydrogenase may be utilized for the sequential conversion of butanal into 5-hydroxy-4-octanone and then 4,5-octanediol. Examples of recombinant microorganisms that comprise these and other various combinations of the individual pathways described herein, as well as various combinations of the individual components of those pathways, will be apparent to those skilled in the art, and may also be found in the Examples.

[0257] Also included are methods of converting a polysaccharide to a first commodity chemical, or an intermediate thereof, such as by utilizing a recombinant microorganism that comprises an aldehyde or ketone biosynthesis pathway. Also included are methods of converting a first commodity chemical, or intermediate thereof, to a second commodity chemical, such as by utilizing a recombinant microorganism that optionally comprises a biosynthesis pathway, optionally comprises C—C ligation pathway and/or optionally comprises one or more of the individual components of a dehydration and reduction pathway. Merely by way of illustration, a recombinant microorganism comprising an exogenous C—C ligase (e.g., benzaldehyde lyase from *Pseudomonas fluorescens*) could be utilized in a method to convert a first commodity chemical such as 3-methylbutanal to a second commodity chemical such as 2,7-dimethyl-5-hydroxy-4-octanone. Along this line of illustration, the same or different recombinant microorganism comprising a diol dehydrogenase could be utilized in a method to convert 2,7-dimethyl-5-hydroxy-4-octanone to another commodity chemical such as 2,7-dimethyl-4,5-octanediol (see Table 2 for other examples). As an additional illustrative example, a recombinant microorganism comprising an exogenous secondary alcohol dehydrogenase could be utilized in a method to convert a first commodity chemical such as 2,7-dimethyl-4-octanone to a second commodity chemical such as 2,7-dimethyloctanol.

[0258] Embodiments of a microbial system or isolated microorganism of the present application may include a naturally-occurring biosynthesis pathway, and/or an engineered, reconstructed, or re-designed biosynthesis pathway that has been optimized for improved functionality.

[0259] Embodiments of a microbial system or recombinant microorganism of the present invention may include a natural or reconstructed biosynthesis pathway, such as a butyraldehyde biosynthesis pathway, as found in such microorganisms as *Clostridium acetobutylicum* and *Streptomyces coelicolor*. In explanation, butyrate and butanol are the common fermentation products of certain bacterial species such as *Clostridia*, in which the production of butyrate and butanol is mediated by a synthetic thiolase dependent pathway characteristically similar to fatty acid degradation pathway. Such pathways may be initiated with the condensation of two molecules of acetyl-CoA to acetoacetyl-CoA, which is catalyzed by thiolase. Acetoacetyl-CoA is then reduced to β -hydroxy butyryl-CoA, which is catalyzed by NAD(P)H dependent β -hydroxy butyryl-CoA dehydrogenase (HBDH). Crotonase catalyzes dehydration from β -hydroxy butyryl-CoA to form crotonyl-CoA. Further reduction catalyzed by NADH-dependent butyryl-CoA dehydrogenase (BCDH) saturates the double bond at C2 of crotonyl-CoA to form butyryl-CoA.

[0260] In certain embodiments, thiolase, the first enzyme in this pathway, may be overexpressed to maximize production. In certain embodiments, thiolase may over-expressed in *E. coli*. In this regard, all three enzymes (e.g., HBDH, crotonase, and BCDH) catalyzing the following reaction steps are found in *Clostridium acetobutylicum* ATCC824. In certain embodiments, BDH, crotonase, and BCDH may be expressed or over-expressed in a suitable microorganism such as *E. coli*. Alternatively, a short-chain aliphatic acyl-CoA dehydrogenase derived from *Pseudomonas putida* KT2440 may be utilized in other embodiments of a microbial system or isolated microorganism of the present application.

[0261] Further to this end, butyryl-CoA in *Clostridia* may be readily converted to butanol and/or butyrate by at least a few different pathways. In one pathway, butyryl-CoA is directly reduced to butyraldehyde catalyzed by NADH dependent CoA-acylating aldehyde dehydrogenase (ALDH). Butyraldehyde may be further reduced to butanol by NADH-dependent butanol dehydrogenase. Although CoA-acylating ALDH catalyzes the one step reduction of butyryl-CoA to butyraldehyde, the incorporation of CoA-acylating ALDH to the microbial system may result in acetoaldehyde formation because of its promiscuous acetyl-CoA deacylating activity. In certain embodiments, the formation of acetoaldehyde may be minimized by functionally redesigning the relevant enzyme(s).

[0262] Butyryl-CoA in other biosynthesis pathways is deacylated to form butyryl phosphate catalyzed by phosphotransbutyrylase. Butyryl phosphate is then hydrolyzed by reversible butyryl phosphate kinase to form butyrate. This reaction is coupled with ATP generation from ADP. The butyrate formation through these enzymes is known to be significantly more specific. Certain embodiments may comprise phosphotransbutyrylase and butyryl phosphate kinase to the microbial system. In other embodiments, butyrate may be directly formed from butyryl-CoA by short chain acyl-CoA thioesterase.

[0263] Butyrate in *Clostridia* may also be sequentially reduced to butanol, which is catalyzed by a single alcohol/aldehyde dehydrogenase. Certain embodiments may comprise short chain aldehyde dehydrogenase from other bacteria such as *Pseudomonas putida* to complement the production of butyraldehyde in the microbial system. One potential concern in using short chain aldehyde dehydrogenase involves the possible formation of acetoaldehyde from acetate. Certain

embodiments may be directed to minimizing the acetate formation in the microbial system, for example, by deleting several genes encoding enzymes involved in the acetate production.

[0264] Moreover, there are multiple routes in *E. coli* to form acetate, one of which is mediated by pyruvate oxygenase (POXB) from pyruvate, whereas another is mediated by phosphotransacetylase (PTA) and acetyl phosphate kinase (ACKA) from acetyl-CoA. The acetate production from *E. coli* mutant strains with $poxB^-$, pta^- , and $ackA^-$ are significantly diminished. In addition, incorporation of acetyl-CoA synthase (ACS) which catalyses the acetyl-CoA formation from acetate is also known to significantly reduce the accumulation of acetate. Certain embodiments may comprise a microbial system or isolated microorganism with deleted POXB, PTA, and/or ACKA genes, and other embodiments may also comprise, separately or together with the deleted genes, one or more genes encoding and expressing ACS.

[0265] A microbial system or recombinant microorganism provided herein may also comprise a glutaraldehyde biosynthesis pathway. As one example, *Saccharomyces cerevisiae* has a lysine biosynthetic pathway in which acetyl-CoA is initially condensed to α -ketoglutarate, a common metabolite in citric acid cycle, to form homocitrate. This reaction is catalyzed by homocitrate synthase derived from Yeast, *Thermus thermophilus*, or *Deinococcus radiodurans*. Homocitrate derived from Yeast, *Thermus thermophilus*, or *Deinococcus radiodurans* catalyzes the conversion between homocitrate and homoisocitrate. Homoisocitrate is then oxidatively decarboxylated to form 2-ketoadipate, which is catalyzed by homoisocitrate dehydrogenase derived from Yeast, *Thermus thermophilus*, or *Deinococcus radiodurans*. Homoisocitrate is also oxidatively decarboxylated to form glutaryl-CoA, which may be catalyzed by homoisocitrate dehydrogenase. Thus, certain embodiments may comprise a homocitrate synthase, a homoaconitase, and/or a homoisocitrate dehydrogenase.

[0266] Further to this end, in synthesizing 2-keto-adipic-semialdehyde, 2-ketoadipate is reduced to 2-keto-adipic-semialdehyde. This reaction can be catalyzed by dialdehyde dehydrogenase, which, for example, may be isolated from *Agrobacterium tumefaciens* C58. Thus, certain embodiments may incorporate dialdehyde dehydrogenases into a microbial system or recombinant microorganism.

[0267] In synthesizing glutaraldehyde, Acyl-CoA thioesterases (ACOT) may also catalyze the hydrolysis of glutaryl-CoA. The genes encoding (β -carboxylic acyl-CoA specific peroxisomal ACOTs) are found in many mammalian species; both ACOT4 and ACOT8 derived from mice have been previously expressed in *E. coli* and shown that both enzymes are highly active on the hydrolysis of glutaryl-CoA to form glutarate. Certain embodiments may comprise one or more Acyl-CoA thioesterases.

[0268] Glutarate is sequentially reduced to glutaraldehyde. This reaction can be catalyzed by glutaraldehyde dehydrogenase (CpnE), which, for example, may be isolated from *Comomonas* sp. Strain NCIMB 9872. Certain embodiments may incorporate glutaraldehyde dehydrogenases such as CpnE into a microbial system or isolated microorganism. Other embodiments may comprise both ACOT and CpnE enzymes. Other embodiments may comprise CpnE enzymes redesigned to catalyze the reduction of 1-hydroxy propanoate and succinate to 1-hydroxy propanal and succinaldehyde.

[0269] In certain aspects, the biosynthesis pathway may include an aldehyde biosynthesis pathway, a ketone biosynthesis pathway, or both. In certain aspects, the biosynthesis pathway may include one or more of an acetoaldehyde, propionaldehyde, butyraldehyde, isobutyraldehyde, 2-methyl-butyraldehyde, 3-methyl-butyraldehyde, 4-methyl-pentaldehyde, phenylacetaldehyde, 2-phenyl acetoaldehyde, 2-(4-hydroxyphenyl)acetaldehyde, 2-Indole-3-acetoaldehyde, glutaraldehyde, 5-amino-pentaldehyde, succinate semialdehyde, and/or succinate 4-hydroxyphenyl acetaldehyde biosynthesis pathway, including various combinations thereof.

[0270] With regard to combinations of biosynthesis pathways, a biosynthesis pathway may comprise an acetoaldehyde biosynthesis pathway in combination with at least one of a propionaldehyde, butyraldehyde, isobutyraldehyde, 2-methyl-butyraldehyde, 3-methyl-butyraldehyde, or phenylacetoaldehyde biosynthesis pathway. In certain aspects, a biosynthesis pathway may comprise a propionaldehyde biosynthesis pathway in combination with at least one of a butyraldehyde, isobutyraldehyde, 2-methyl-butyraldehyde, 3-methyl-butyraldehyde, or phenylacetoaldehyde biosynthesis pathway. In certain aspects, a biosynthesis pathway may comprise an isobutyraldehyde biosynthesis pathway in combination with at least one of a 2-methyl-butyraldehyde, 3-methyl-butyraldehyde, or phenylacetoaldehyde biosynthesis pathway. In certain aspects, a biosynthesis pathway may comprise a 2-methyl-butyraldehyde biosynthesis pathway in combination with at least one of a 3-methyl-butyraldehyde or a phenylacetoaldehyde biosynthesis pathway. In certain aspects, a biosynthesis pathway may comprise a 3-methyl-butyraldehyde biosynthesis pathway in combination with a phenylacetoaldehyde biosynthesis pathway.

[0271] In certain aspects, a propionaldehyde biosynthesis pathway may comprise a threonine deaminase (ilvA) gene from an organism such as *Escherichia coli* and a keto-isovalerate decarboxylase (kivd) gene from an organism such as *Lactococcus lactis*, and/or functional variants of these enzymes, including homologs or orthologs thereof, as well as optimized variants. These enzymes may be utilized generally to convert L-threonine to propionaldehyde.

[0272] In certain aspects, a butyraldehyde biosynthesis pathway may comprise at least one of a thiolase (atoB) gene from an organism such as *E. coli*, a β -hydroxy butyryl-CoA dehydrogenase (hbd) gene, a crotonase (crt) gene, a butyryl-CoA dehydrogenase (bcd) gene, an electron transfer flavoprotein A (etfA) gene, and/or an electron transfer flavoprotein B (etfB) gene from an organism such as *Clostridium acetobutyricum* (e.g., ATCC 824), as well as a coenzyme A-linked butyraldehyde dehydrogenase (ald) gene from an organism such as *Clostridium beijerinckii acetobutyricum* ATCC 824. In certain aspects, a coenzyme A-linked alcohol dehydrogenase (adhE2) gene from an organism such as *Clostridium acetobutyricum* ATCC 824 may be used as an alternative to an ald gene.

[0273] In certain aspects, an isobutyraldehyde biosynthetic pathway may comprise an acetolactate synthase (alsS) from an organism such as *Bacillus subtilis* or an als gene from an organism such as *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 (codon usage may be optimized for *E. coli*

protein expression). Such a pathway may also comprise acetolactate reductoisomerase (ilvC) and/or 2,3-dihydroxyisovalerate dehydratase (ilvD) genes from an organism such as *E. coli*, as well as a keto-isovalerate decarboxylase (kivd) gene from an organism such as *Lactococcus lactis*.

[0274] In certain aspects, a 3-methylbutyraldehyde and 2-methylbutyraldehyde biosynthesis pathway may comprise an acetolactate synthase (alsS) gene from an organism such as *Bacillus subtilis* or an (als) gene from an organism such as *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 (codon usage may be optimized for *E. coli* protein expression). Certain aspects of such a pathway may also comprise acetolactate reductoisomerase (ilvC), 2,3-dihydroxyisovalerate dehydratase (ilvD), isopropylmalate synthase (LeuA), isopropylmalate isomerase (LeuC and LeuD), and 3-isopropylmalate dehydrogenase (LeuB) genes from an organism such as *E. coli*, as well as a keto-isovalerate decarboxylase (kivd) from an organism such as *Lactococcus lactis*.

[0275] In certain aspects, a phenylacetoaldehyde and 4-hydroxyphenylacetoaldehyde biosynthesis pathway may comprise one or more of 3-deoxy-7-phosphoheptulonate synthase (aroF, aroG, and aroH), 3-dehydroquinone synthase (aroB), a 3-dehydroquinone dehydratase (aroD), dehydroshikimate reductase (aroE), shikimate kinase II (aroL), shikimate kinase I (aroK), 5-enolpyruvylshikimate-3-phosphate synthetase (aroA), chorismate synthase (aroC), fused chorismate mutase P/prephenate dehydratase (pheA), and/or fused chorismate mutase T/prephenate dehydrogenase (tyrA) genes from an organism such as *E. coli*, as well as a keto-isovalerate decarboxylase (kivd) from an organism such as *Lactococcus lactis*.

[0276] In certain aspects, such as for the ultimate production of 1,10-diamino-5-decanol and 1,10-dicarboxylic-5-decanol, a biosynthesis pathway may comprise one or more homocitrate synthase, homoaconitate hydratase, homoisocitrate dehydrogenase, and/or homoisocitrate dehydrogenase genes from an organism such as *Deinococcus radiodurans* and/or *Thermus thermophilus*, as well as a keto-adipate decarboxylase gene, a 2-aminoadipate transaminase gene, and a L-2-Aminoadipate-6-semialdehyde: NAD+6-oxidoreductase gene. Such a biosynthesis pathway would be able to convert α -ketoglutarate to 5-aminopentaldehyde.

[0277] In certain aspects, such as for one step in cyclopentanol production, a α -keto adipate semialdehyde biosynthesis pathway may comprise homocitrate synthase (hcs), homoaconitate hydratase, and homoisocitrate dehydrogenase genes from an organism such as *Deinococcus radiodurans* and/or *Thermus thermophilus*, and an α -keto adipate semialdehyde dehydrogenase gene. Such a biosynthesis pathway would be able to convert acetyl-CoA and α -ketoglutarate to α -keto adipate semialdehyde.

[0278] For the production of certain commodity chemicals, such as 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol, and indole-3-ethanol, among other similar chemicals, a biosynthesis pathway (e.g., aldehyde biosynthesis pathway) may optionally or further comprise one or more genes encoding a carboxylase enzyme, such as an indole-3-pyruvate decarboxylase (IPDC). An IPDC may be obtained, for example, from such microorganisms as *Azospirillum brasilense* and *Paenibacillus polymyxa* E681. In this regard, an IPDC may be utilized to more efficiently catalyze the decarboxylation of various carboxylic acids to form the corresponding aldehyde, which can be further converted to a commodity chemical by a reductase or dehydrogenase, as detailed herein.

[0279] In certain aspects, a 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol, and 2-(indole-3-yl)ethanol biosynthesis pathway may comprise a transketolase (tktA), a 3-deoxy-7-phosphoheptulonate synthase (aroF, aroG, and aroH), 3-dehydroquinate synthase (aroB), a 3-dehydroquinate dehydratase (aroD), a dehydroshikimate reductase (aroE), a shikimate kinase II (aroL), a shikimate kinase I (aroK), a 5-enolpyruvylshikimate-3-phosphate synthetase (aroA), a chorismate synthase (aroC), a fused chorismate mutase P/prephenate dehydratase (pheA), and a fused chorismate mutase T/prephenate dehydrogenase (tyrA) genes from *E. coli*, keto-isovalerate decarboxylase (kivd) from *Lactococcus lactis*, alcohol dehydrogenase (adh2) from *Saccharomyces cerevisiae*, Indole-3-pyruvate decarboxylase (ipdc) from *Azospirillum brasilense*, phenylethanol reductase (par) from *Rhodococcus* sp. ST-10, and abenzaldehyde lyase (bal) from *Pseudomonas fluorescens*.

[0280] As for all other pathways described herein, the components for each of the biosynthesis pathways described herein may be present in a recombinant microorganism either endogenously or exogenously. To improve the efficiency of a given biosynthesis pathway, endogenous genes, for example, may be up-regulated or over-expressed, such as by introducing an additional (i.e., exogenous) copy of that endogenous gene into the recombinant microorganism. Such pathways may also be optimized by altering via mutagenesis the endogenous version of a gene to improve functionality, followed by introduction of the altered gene into the microorganism. The expression of endogenous genes may be up or down-regulated, or even eliminated, according to known techniques in the art and described herein. Similarly, the expression levels of exogenously provided genes may be regulated as desired, such as by using various constitutive or inducible promoters. Such genes may also be "codon-optimized," as described herein and known in the art. Also included are functional naturally-occurring variants of the genes and enzymes described herein, including homologs or orthologs thereof.

[0281] Certain embodiments of a microbial system or isolated microorganism may comprise a CC-ligation pathway. In certain aspects, a CC-ligation pathway may comprise a ThDP-dependent enzyme, such as a C—C ligase, or an optimized C—C ligase. For example, eight-carbon unit molecules (butyrosins) may be made from condensing together two four-carbon unit molecules (butyraldehydes). ThDP-dependent enzymes are a group of enzymes known to catalyze both breaking and formation of C—C bonds and have been utilized as catalysts in chemoenzymatic syntheses. The spectrum of chemical reactions that these enzymes catalyze ranges from decarboxylation of α -keto acids, oxidative decarboxylation, carboligation, and to the cleavage of C—C bonds.

[0282] To provide a few examples, benzaldehyde lyase (BAL) from *Pseudomonas fluorescens*, benzoylformate decarboxylase (BFD) from *Pseudomonas putida*, and pyruvate decarboxylase (PDC) from *Zymomonas mobilis* may catalyze a carboligation reaction between two aldehydes. BAL accepts the broadest spectrum of aldehydes as substrates among these three enzymes ranging from substituted benzaldehyde to acetaldehyde, among others, as shown herein. BAL catalyzes stereospecific carboligation reaction between two aldehydes and forms α -hydroxy ketone with over 99% ee for R-configuration. The benzoin formation from two benzaldehyde molecules is a favored reaction catalyzed by BAL and proceeds as fast as 320 μmol (benzoin) mg (protein)⁻¹

min^{-1} . The formation of α -hydroxy ketone may be carried out using many different aldehydes, including butyraldehyde.

[0283] BFD and PCD may also catalyze the carboligation reactions between two aldehyde molecules. BFD and PCD accept relatively larger and smaller aldehyde molecules, respectively. With the presence of benzaldehyde and acetaldehyde, BFD catalyzes the formation of benzoin and (S)- α -hydroxy phenylpropanone (2S-HPP), whereas PCD catalyzes the formation of (R)- α -hydroxy phenylpropanone (2R-HPP) and (R)- α -hydroxy 2-butanone (acetoin). As detailed below, certain microbial systems or isolated microorganisms of the present application may comprise natural or optimized C—C ligases (ThDP-dependent enzymes) selected from benzaldehyde lyase (BAL) from *Pseudomonas fluorescens*, benzoylformate decarboxylase (BFD) from *Pseudomonas putida*, and pyruvate decarboxylase (PDC) from *Zymomonas mobilis*. Other embodiments may comprise a benzaldehyde lyase (BAL) from *Pseudomonas fluorescens* (see SEQ ID NOS:143-144, showing the nucleotide and polypeptide sequences, respectively) including biologically active variants thereof, such as optimized variants.

[0284] A C—C ligation pathway of the present invention typically comprises one or more C—C ligases, such as a lyase enzyme. Exemplary lyases include, but are not limited to, acetaldehyde lyases, propionaldehyde lyases, butyraldehyde lyases, isobutyraldehyde lyases, 2-methyl-butyraldehyde lyases, 3-methyl-butyraldehyde lyases (isovaldehyde), phenylacetaldehyde lyases, α -keto adipate carboxylases, pentaldehyde lyases, 4-methyl-pentaldehyde lyases, hexyldehyde lyases, heptaldehyde lyases, octaldehyde lyases, 4-hydroxyphenylacetaldehyde lyases, indoleacetaldehyde lyases, indolephenylacetaldehyde lyases. In certain aspects, a selected CC-ligase or lyase enzyme may have one or more of the above exemplified lyase activities, such as acetaldehyde lyase activity, a propionaldehyde lyase activity, a butyraldehyde lyase activity, and/or an isobutyraldehyde lyase activity, among others.

[0285] As noted above, a C—C ligase may comprise a benzaldehyde lyase, such as a benzaldehyde lyase isolated from *Pseudomonas fluorescens* (SEQ ID NOS:143-144), as well as biologically active fragments or variants of this reference sequence, such as optimized variants of a benzaldehyde lyase. In this regard, certain aspects may comprise nucleotide sequences or polypeptide sequences having 80%, 85%, 90%, 95%, 97%, 98%, 99% sequence identity to SEQ ID NOS:143-144, and which are capable of catalyzing a carboligation reaction, or which possess C—C lyase activity, as described herein. In certain aspects, a BAL enzyme will comprise one or more conserved amino acid residues, including G27, E50, A57, G155, P162, P234, D271, G277, G422, G447, D448, and/or G512.

[0286] *Pseudomonas fluorescens* is able to grow on R-benzoin as the sole carbon and energy source because it harbours the enzyme benzaldehyde lyase that cleaves the acyloln linkage using thiamine diphosphate (ThDP) as a cofactor. In the reverse reaction, as utilized herein, benzaldehyde lyase catalyzes the carboligation of two aldehydes with high substrate and stereospecificity. Structure-based comparisons with other proteins show that benzaldehyde lyase belongs to a group of closely related ThDP-dependent enzymes. The ThDP cofactors of these enzymes are fixed at their two ends in separate domains, suspending a comparatively mobile thiazolium ring between them. While the residues binding the two ends of ThDP are well conserved, the lining of the active

centre pocket around the thiazolium moiety varies greatly within the group. The active sites for BAL have been described, for example, in Kneen et al (*Biochimica et Biophysica Acta* 1753:263-271, 2005) and Brandt et al. (*Biochemistry* 47:7734-43, 2008). Benzaldehyde lyase derived from *Pseudomonas fluorescens* has been demonstrated herein to at least have an acetaldehyde lyase activity, a propionaldehyde lyase activity, a butyraldehyde lyase activity, a 3-methyl-butyraldehyde lyase activity, a pentaldehyde lyase activity, a 4-methylpentaldehyde lyase activity, a hexyldehyde lyase activity, a phenylacetaldehyde lyase activity, and an octaldehyde lyase activity (see Table 2), among other in vivo lyase activities (see FIGS. 48-55).

[0287] In certain aspects, a C—C ligase, such as BAL derived from *Pseudomonas fluorescens*, BFD derived from *Pseudomonas putida*, or PDC derived from *Zymomonas mobilis* may comprise a lyase with a combination of lyase activities, such as a lyase having both a propionaldehyde lyase activity and a 3-methyl-butyraldehyde lyase activity, among other combinations and activities, such as those exemplary combinations detailed herein. Merely by way of illustration, a lyase having a combination of lyase activities may be referred to herein as a propionaldehyde/3-methyl-butyraldehyde lyase.

[0288] A dehydration and reduction pathway, comprising a diol dehydrogenase, a diol dehydratase, and a secondary alcohol dehydrogenase, may be utilized to further convert an aldehyde, ketone, or corresponding alcohol, to a commodity chemical, such as a biofuel.

[0289] To this end, a dehydration and reduction pathway may comprise one or more diol dehydrogenases. A “diol dehydrogenase” refers generally to an enzyme that catalyzes the reversible reduction and oxidation of a α -hydroxy ketone and/or its corresponding diol. Certain embodiments of a microbial system or isolated microorganism may comprise genes encoding a diol dehydrogenase that specifically catalyzes the reduction of α -hydroxy-ketones, including, for example, a 4, 5, octanediol dehydrogenase. Diol dehydrogenases, such as 4, 5, octanediol dehydrogenase, may be isolated from a variety of organisms and incorporated into a microbial system or isolated microorganism. A particular group of alcohol dehydrogenases has a characteristic ability to oxidize various α -hydroxy alcohols and reduce various α -hydroxy ketones and α -keto ketones. As such, the recitation “diol dehydrogenase” may also encompass such alcohol dehydrogenases.

[0290] By way of example regarding diol dehydrogenases from exemplary organisms, glycerol dehydrogenase isolated from *Hansenula ofunaensis* has broad substrate specificity and is capable of catalyzing the oxidation of various α -hydroxy alcohols, including 1,2-octane, as well as the reduction of various α -hydroxy ketones and α -keto ketones, including 3-hydroxy-2-butanone and 3,4-hexanedione, with the activity comparable to its native substrates, glycerol and dihydroxyacetone, respectively (40-200%). As one further example, glycerol dehydrogenase discovered in *Hansenula polymorpha* DI-1 works similarly. In certain embodiments, a microbial system or recombinant microorganism may comprise a glycerol dehydrogenase gene isolated from *Hansenula ofunaensis*, a glycerol dehydrogenase isolated from *Hansenula polymorpha* DI-1 and/or a meso-2,3-butane diol dehydrogenase from *Klebsiella pneumoniae*. In other embodiments, a microbial system or isolated microorganism may comprise a 4, 5, octanediol dehydrogenase, among oth-

ers detailed herein. Diol dehydrogenases may also be obtained from *Lactobacillus brevis* ATCC 367, *Pseudomonas putida* KT2440, and *Klebsiella pneumoniae* MGH78578), as described herein (see Example 5).

[0291] Exemplary diol dehydrogenases include, but are not limited to, 2,3-butanediol dehydrogenase, 3,4-hexanediol dehydrogenase, 4,5-octanediol dehydrogenase, 5,6-decanediol dehydrogenase, 6,7-dodecanediol dehydrogenase, 7,8-tetradecanediol dehydrogenase, 8,9-hexadecanediol dehydrogenase, 2,5-dimethyl-3,4-hexanediol dehydrogenase, 3,6-dimethyl-4,5-octanediol dehydrogenase, 2,7-dimethyl-4,5-octanediol dehydrogenase, 2,9-dimethyl-5,6-decanediol dehydrogenase, 1,4-diphenyl-2,3-butanediol dehydrogenase, bis-1,4-(4-hydroxyphenyl)-2,3-butanediol dehydrogenase, 1,4-diindole-2,3-butanediol dehydrogenase, 1,2-cyclopentanediol dehydrogenase, 2,3-pentanediol dehydrogenase, 2,3-hexanediol dehydrogenase, 2,3-heptanediol dehydrogenase, 2,3-octanediol dehydrogenase, 2,3-nonanediol dehydrogenase, 4-methyl-2,3-pentanediol dehydrogenase, 4-methyl-2,3-hexanediol dehydrogenase, 5-methyl-2,3-hexanediol dehydrogenase, 6-methyl-2,3-heptanediol dehydrogenase, 1-phenyl-2,3-butanediol dehydrogenase, 1-(4-hydroxyphenyl)-2,3-butanediol dehydrogenase, 1-indole-2,3-butanediol dehydrogenase, 3,4-heptanediol dehydrogenase, 3,4-octanediol dehydrogenase, 3,4-nonanediol dehydrogenase, 3,4-decanediol dehydrogenase, 3,4-undecanediol dehydrogenase, 2-methyl-3,4-hexanediol dehydrogenase, 5-methyl-3,4-heptanediol dehydrogenase, 6-methyl-3,4-heptanediol dehydrogenase, 7-methyl-3,4-octanediol dehydrogenase, 1-phenyl-2,3-pentanediol dehydrogenase, 1-(4-hydroxyphenyl)-2,3-pentanediol dehydrogenase, 1-indole-2,3-pentanediol dehydrogenase, 4,5-nonanediol dehydrogenase, 4,5-decanediol dehydrogenase, 4,5-undecanediol dehydrogenase, 4,5-dodecanediol dehydrogenase, 2-methyl-3,4-heptanediol dehydrogenase, 3-methyl-4,5-octanediol dehydrogenase, 2-methyl-4,5-octanediol dehydrogenase, 8-methyl-4,5-nonanediol dehydrogenase, 1-phenyl-2,3-hexanediol dehydrogenase, 1-(4-hydroxyphenyl)-2,3-hexanediol dehydrogenase, 1-indole-2,3-hexanediol dehydrogenase, 5,6-undecanediol dehydrogenase, 5,6-undecanediol dehydrogenase, 5,6-tridecanediol dehydrogenase, 2-methyl-3,4-octanediol dehydrogenase, 3-methyl-4,5-nonanediol dehydrogenase, 2-methyl-4,5-nonanediol dehydrogenase, 2-methyl-5,6-decanediol dehydrogenase, 1-phenyl-2,3-heptanediol dehydrogenase, 1-(4-hydroxyphenyl)-2,3-heptanediol dehydrogenase, 1-indole-2,3-heptanediol dehydrogenase, 6,7-tridecanediol dehydrogenase, 6,7-tetradecanediol dehydrogenase, 2-methyl-3,4-decanediol dehydrogenase, 3-methyl-4,5-decanediol dehydrogenase, 2-methyl-4,5-decanediol dehydrogenase, 2-methyl-5,6-undecanediol dehydrogenase, 1-phenyl-2,3-octanediol dehydrogenase, 1-(4-hydroxyphenyl)-2,3-octanediol dehydrogenase, 1-indole-2,3-octanediol dehydrogenase, 7,8-pentadecanediol dehydrogenase, 2-methyl-3,4-decanediol dehydrogenase, 3-methyl-4,5-undecanediol dehydrogenase, 2-methyl-4,5-undecanediol dehydrogenase, 2-methyl-5,6-dodecanediol dehydrogenase, 1-phenyl-2,3-nonanediol dehydrogenase, 1-(4-hydroxyphenyl)-2,3-nonanediol dehydrogenase, 1-indole-2,3-nonanediol dehydrogenase, 2-methyl-3,4-undecanediol dehydrogenase, 3-methyl-4,5-dodecanediol dehydrogenase, 2-methyl-4,5-dodecanediol dehydrogenase, 2-methyl-5,6-tridecanediol dehydrogenase, 1-phenyl-2,3-decanediol dehydrogenase, 1-(4-hydroxyphenyl)-2,3-decanediol dehydroge-

nase, 1-indole-2,3-decanediol dehydrogenase, 2,5-dimethyl-3,4-heptanediol dehydrogenase, 2,6-dimethyl-3,4-heptanediol dehydrogenase, 2,7-dimethyl-3,4-octanediol dehydrogenase, 1-phenyl-4-methyl-2,3-pentanediol dehydrogenase, 1-(4-hydroxyphenyl)-4-methyl-2,3-pentanediol dehydrogenase, 1-indole-4-methyl-2,3-pentanediol dehydrogenase, 2,6-dimethyl-4,5-octanediol dehydrogenase, 3,8-dimethyl-4,5-nonanediol dehydrogenase, 1-phenyl-4-methyl-2,3-hexanediol dehydrogenase, 1-(4-hydroxyphenyl)-4-methyl-2,3-hexanediol dehydrogenase, 1-indole-4-methyl-2,3-hexanediol dehydrogenase, 2,8-dimethyl-4,5-nonanediol dehydrogenase, 1-phenyl-5-methyl-2,3-hexanediol dehydrogenase, 1-(4-hydroxyphenyl)-5-methyl-2,3-hexanediol dehydrogenase, 1-indole-5-methyl-2,3-hexanediol dehydrogenase, 1-phenyl-6-methyl-2,3-heptanediol dehydrogenase, 1-(4-hydroxyphenyl)-6-methyl-2,3-heptanediol dehydrogenase, 1-indole-6-methyl-2,3-heptanediol dehydrogenase, 1-(4-hydroxyphenyl)-4-phenyl-2,3-butanediol dehydrogenase, 1-indole-4-phenyl-2,3-butanediol dehydrogenase, 1-indole-4-(4-hydroxyphenyl)-2,3-butanediol dehydrogenase, 1,10-diamino-5,6-decanediol dehydrogenase, 1,4-di(4-hydroxyphenyl)-2,3-butanediol, 2,3-hexanediol-1,6-dicarboxylic acid dehydrogenase, and the like.

[0292] In certain aspects, a selected diol dehydrogenase enzyme may have one or more of the above exemplified diol dehydrogenase activities, such as a 2,3-butanediol dehydrogenase activity, a 3,4-hexanediol dehydrogenase activity, and/or a 4,5-octanediol dehydrogenase activity, among others.

[0293] In certain aspects, a recombinant microorganism may comprise a diol dehydrogenase encoded by a nucleotide reference sequence selected from SEQ ID NO:97, 99, and 101, or an enzyme having a polypeptide sequence selected from SEQ ID NO:98, 100, and 102, including biologically active fragments or variants thereof, such as optimized variants. Certain aspects may also comprises nucleotide sequences or polypeptide sequences having 80%, 85%, 90%, 95%, 97%, 98%, 99% sequence identity to SEQ ID NOS:97-102.

[0294] Other embodiments may comprise re-designed diol dehydrogenases for reduction of 1-hydroxy propanal, succinaldehyde, and glutaraldehyde to 1,3-propanediol, 1,4-butanediol, and 1,5 pentanediol, respectively, among others.

[0295] A dehydration and reduction pathway, as described herein, may comprise one or more diol dehydratases. A "diol dehydratase" refers generally to an enzyme that catalyzes the irreversible dehydration of diols. For instance, this enzyme may serve to dehydrate octanediol to form 4-octane. It has been recognized that there are at least two different types of diol dehydratases: a group dependent on and independent of coenzyme B12 for its catalysis. Coenzyme B12 dependent diol dehydratases are known to catalyze a radical mediated dehydration reaction from α -hydroxy alcohol to aldehydes or ketones. For example, a diol dehydratase from *Klebsiella pneumoniae* catalyzes the dehydration of glycerol to form β -hydroxypropyl aldehyde, accepts 2,3-butanediol as a substrate, and catalyzes the dehydration reaction to form 2-butanone.

[0296] As a further example, *Clostridium butylicum* contains coenzyme B12 independent diol dehydratases. FIG. 46 shows the in vivo biological activity of coenzyme B12 independent diol dehydratase (dhaB1) and activator (dhaB2) isolated from *Clostridium butylicum* (see Example 9). 46A shows the in vivo production of 1-propanol from 1,2-pro-

panediol, FIG. 46B shows the in vivo production of 2-butanol from meso-2,3 butanediol, and FIG. 46C shows the in vivo production of cyclopentanone from trans-1,2-cyclopentanediol.

[0297] Thus, certain embodiments of the present invention may comprise optimized or redesigned diol dehydratases that accommodate various substrates, such as 4,5-octanediol as a substrate, and may include diol dehydratases isolated and/or optimized from *Klebsiella pneumoniae* and *Clostridium butylicum*, among other organisms described herein and known in the art.

[0298] Exemplary diol dehydratases include, but are not limited to, 2,3-butanediol dehydratase, 3,4-hexanediol dehydratase, 4,5-octanediol dehydratase, 5,6-decanediol dehydratase, 6,7-dodecanediol dehydratase, 7,8-tetradecanediol dehydratase, 8,9-hexadecanediol dehydratase, 2,5-dimethyl-3,4-hexanediol dehydratase, 3,6-dimethyl-4,5-octanediol dehydratase, 2,7-dimethyl-4,5-octanediol dehydratase, 2,9-dimethyl-5,6-decanediol dehydratase, 1,4-diphenyl-2,3-butanediol dehydratase, bis-1,4-(4-hydroxyphenyl)-2,3-butanediol dehydratase, 1,4-diindole-2,3-butanediol dehydratase, 1,2-cyclopentanediol dehydratase, 2,3-pentanediol dehydratase, 2,3-hexanediol dehydratase, 2,3-heptanediol dehydratase, 2,3-octanediol dehydratase, 2,3-nonanediol dehydratase, 4-methyl-2,3-pentanediol dehydratase, 4-methyl-2,3-hexanediol dehydratase, 5-methyl-2,3-hexanediol dehydratase, 6-methyl-2,3-heptanediol dehydratase, 1-phenyl-2,3-butanediol dehydratase, 1-(4-hydroxyphenyl)-2,3-butanediol dehydratase, 1-indole-2,3-butanediol dehydratase, 3,4-heptanediol dehydratase, 3,4-octanediol dehydratase, 3,4-nonanediol dehydratase, 3,4-decanediol dehydratase, 3,4-undecanediol dehydratase, 2-methyl-3,4-hexanediol dehydratase, 5-methyl-3,4-heptanediol dehydratase, 6-methyl-3,4-heptanediol dehydratase, 7-methyl-3,4-octanediol dehydratase, 1-phenyl-2,3-pentanediol dehydratase, 1-(4-hydroxyphenyl)-2,3-pentanediol dehydratase, 1-indole-2,3-pentanediol dehydratase, 4,5-nonanediol dehydratase, 4,5-decanediol dehydratase, 4,5-undecanediol dehydratase, 4,5-dodecanediol dehydratase, 2-methyl-3,4-heptanediol dehydratase, 3-methyl-4,5-octanediol dehydratase, 2-methyl-4,5-octanediol dehydratase, 8-methyl-4,5-nonanediol dehydratase, 1-phenyl-2,3-hexanediol dehydratase, 1-(4-hydroxyphenyl)-2,3-hexanediol dehydratase, 1-indole-2,3-hexanediol dehydratase, 5,6-undecanediol dehydratase, 5,6-undecanediol dehydratase, 5,6-tridecanediol dehydratase, 2-methyl-3,4-octanediol dehydratase, 3-methyl-4,5-nonanediol dehydratase, 2-methyl-4,5-nonanediol dehydratase, 2-methyl-5,6-decanediol dehydratase, 1-phenyl-2,3-heptanediol dehydratase, 1-(4-hydroxyphenyl)-2,3-heptanediol dehydratase, 1-indole-2,3-heptanediol dehydratase, 6,7-tridecanediol dehydratase, 6,7-tetradecanediol dehydratase, 2-methyl-3,4-nonanediol dehydratase, 3-methyl-4,5-decanediol dehydratase, 2-methyl-4,5-decanediol dehydratase, 2-methyl-5,6-undecanediol dehydratase, 1-phenyl-2,3-octanediol dehydratase, 1-(4-hydroxyphenyl)-2,3-octanediol dehydratase, 1-indole-2,3-octanediol dehydratase, 7,8-pentadecanediol dehydratase, 2-methyl-3,4-decanediol dehydratase, 3-methyl-4,5-undecanediol dehydratase, 2-methyl-4,5-undecanediol dehydratase, 2-methyl-5,6-dodecanediol dehydratase, 1-phenyl-2,3-nonanediol dehydratase, 1-(4-hydroxyphenyl)-2,3-nonanediol dehydratase, 1-indole-2,3-nonanediol dehydratase, 2-methyl-3,4-undecanediol dehydratase, 3-methyl-4,5-dodecanediol dehydratase, 2-methyl-4,5-dode-

canediol dehydratase, 2-methyl-5,6-tridecanediol dehydratase, 1-phenyl-2,3-decanediol dehydratase, 1-(4-hydroxyphenyl)-2,3-decanediol dehydratase, 1-indole-2,3-decanediol dehydratase, 2,5-dimethyl-3,4-heptanediol dehydratase, 2,6-dimethyl-3,4-heptanediol dehydratase, 2,7-dimethyl-3,4-octanediol dehydratase, 1-phenyl-4-methyl-2,3-pentanediol dehydratase, 1-(4-hydroxyphenyl)-4-methyl-2,3-pentanediol dehydratase, 1-indole-4-methyl-2,3-pentanediol dehydratase, 2,6-dimethyl-4,5-octanediol dehydratase, 3,8-dimethyl-4,5-nonanediol dehydratase, 1-phenyl-4-methyl-2,3-hexanediol dehydratase, 1-(4-hydroxyphenyl)-4-methyl-2,3-hexanediol dehydratase, 1-indole-4-methyl-2,3-hexanediol dehydratase, 2,8-dimethyl-4,5-nonanediol dehydratase, 1-phenyl-5-methyl-2,3-hexanediol dehydratase, 1-(4-hydroxyphenyl)-5-methyl-2,3-hexanediol dehydratase, 1-indole-5-methyl-2,3-hexanediol dehydratase, 1-phenyl-6-methyl-2,3-heptanediol dehydratase, 1-(4-hydroxyphenyl)-6-methyl-2,3-heptanediol dehydratase, 1-indole-6-methyl-2,3-heptanediol dehydratase, 1-(4-hydroxyphenyl)-4-phenyl-2,3-butanediol dehydratase, 1-indole-4-phenyl-2,3-butanediol dehydratase, 1-indole-4-(4-hydroxyphenyl)-2,3-butanediol dehydratase, 1,10-diamino-5,6-decanediol dehydratase, 1,4-di(4-hydroxyphenyl)-2,3-butanediol, 2,3-hexanediol-1,6-dicarboxylic acid dehydratase, and the like.

[0299] In certain aspects, a selected diol dehydratase enzyme may have one or more of the above exemplified diol dehydratase activities, such as a 2,3-butanediol dehydratase activity, a 3,4-hexanediol dehydratase activity, and/or a 4,5-octanediol dehydratase activity, among others.

[0300] In certain aspects, diol dehydratases may be obtained from *Klebsiella pneumoniae* MGH 78578, including from the pduCDE gene of this and other microorganisms. In certain aspects, a recombinant microorganism may comprise one or more diol dehydratases encoded by a nucleotide reference sequence selected from SEQ ID NO:103, 105, and 107, or an enzyme having a polypeptide sequence selected from SEQ ID NO:104, 106, and 108, including biologically active fragments or variants thereof, such as optimized variants. Certain aspects may also comprise nucleotide sequences or polypeptide sequences having 80%, 85%, 90%, 95%, 97%, 98%, 99% sequence identity to SEQ ID NOS:103-108. In certain aspects, polypeptides of SEQ ID NO:104 may comprise certain conserved amino acid residues, including those chosen from D149, P151, A155, A159, G165, E168, E170, A183, G189, G196, Q200, E208, G215, Y219, E221, T222, S224, Y226, G227, T228, F232, G235, D236, D237, T238, P239, S241, L245, Y249, S251, R252, G253, K255, R257, S260, E265, M268, G269, S275, Y278, L279, E280, C283, G291, Q293, G294, Q296, N297, G298, G312, E329, S341, R344, G356, D371, N372, F374, S377, R392, D393, R412, L477, A486, G499, D500, S516, N522, D523, Y524, G526, and G530.

[0301] In certain aspects, a diol dehydratase may include a polypeptide that comprises an amino acid sequence having 0%, 85%, 90%, 95%, 97%, 98%, 99% sequence identity to SEQ ID NOS:308-311. SEQ ID NO:308 shows the polypeptide sequence of PduG, a diol dehydratase reactivation large subunit derived from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578. SEQ ID NO:309 shows the polypeptide sequence of PduH, diol dehydratase reactivation small subunit derived from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578. SEQ ID NO:310 shows the polypeptide sequence of a B12-independent glycerol dehydratase from

Clostridium Butyricum. SEQ ID NO:311 shows the polypeptide sequence of a glycerol dehydratase activator from *Clostridium Butyricum*. In certain aspects, a B12-independent glycerol dehydratase may comprise conserved amino acid residues, such as T36, G74, P87, E88, E97, W126, R221, A263, Q265, R287, D289, E309, R317, G335, G345, G346, N356, P374, R379, G399, G401, P403, D408, G432, C433, N452, C529, G533, G539, G540, S559, G603, N₆₀₄, A654, G658, R659, D676, N₇₀₂, Q735, N737, A747, P751, R760, V761, A762, G763, Q776, I780, and/or R782. In certain aspects, a B12-independent glycerol dehydratase activator may comprise certain conserved amino acid residues, including D19, G20, G22, R24, F28, G31, C32, C36, W38, C39, N41, P42, C58, C64, C96, G129, T132, G135, G136, D185, R187, N₂₀₈, R222, and/or R264.

[0302] A dehydration and reduction pathway, as described herein, may comprise one or more alcohol dehydrogenases or secondary alcohol dehydrogenases. An "alcohol dehydrogenase" or "secondary alcohol dehydrogenase" that is part of a dehydration and reduction pathway refers generally to an enzyme that catalyzes the conversion of aldehyde or ketone substituents to alcohols. For instance, 4-octanone may be reduced to 4-octanol by a secondary alcohol dehydrogenase one enzymatic step for the conversion of butyrolin to a biofuel. *Pseudomonads* express at least one secondary alcohol dehydrogenase that oxidizes 4-octanol to 4-octanone using NAD⁺ as a co-factor. As another example, *Rhodococcus erythropolis* ATCC4277 catalyzes oxidation of medium to long chain secondary fatty alcohols using NADH as a co-factor, using an enzyme that also catalyzes the oxidation of 3-decanol and 4-decanol. In addition, *Norcadia fusca* AKU2123 contains an (S)-specific secondary alcohol dehydrogenase.

[0303] Genes encoding secondary alcohol dehydrogenases may be isolated from these and other organisms according to known techniques in the art and incorporated into the microbial systems recombinant organisms as described herein. In certain embodiments, a microbial system or isolated microorganism may comprise natural or optimized secondary alcohol dehydrogenases from *Pseudomonads*, *Rhodococcus erythropolis* ATCC4277, *Norcadia fusca* AKU2123, or other suitable organisms.

[0304] Examples of secondary alcohol dehydrogenases include, but are not limited to, 2-butanol dehydrogenase, 3-hexanol dehydrogenase, 4-octanol dehydrogenase, 5-decanol dehydrogenase, 6-dodecanol dehydrogenase, 7-tetradecanol dehydrogenase, 8-hexadecanol dehydrogenase, 2,5-dimethyl-3-hexanol dehydrogenase, 3,6-dimethyl-4-octanol dehydrogenase, 2,7-dimethyl-4-octanol dehydrogenase, 2,9-dimethyl-4-decanol dehydrogenase, 1,4-diphenyl-2-butanol dehydrogenase, bis-1,4-(4-hydroxyphenyl)-2-butanol dehydrogenase, 1,4-diindole-2-butanol dehydrogenase, cyclopentanol dehydrogenase, 2(or 3)-pentanol dehydrogenase, 2(or 3)-hexanol dehydrogenase, 2(or 3)-heptanol dehydrogenase, 2(or 3)-octanol dehydrogenase, 2(or 3)-nonanol dehydrogenase, 4-methyl-2(or 3)-pentanol dehydrogenase, 4-methyl-2(or 3)-hexanol dehydrogenase, 5-methyl-2(or 3)-hexanol dehydrogenase, 6-methyl-2(or 3)-heptanol dehydrogenase, 1-phenyl-2(or 3)-butanol dehydrogenase, 1-(4-hydroxyphenyl)-2(or 3)-butanol dehydrogenase, 1-indole-2(or 3)-butanol dehydrogenase, 3(or 4)-heptanol dehydrogenase, 3(or 4)-octanol dehydrogenase, 3(or 4)-nonanol dehydrogenase, 3(or 4)-decanol dehydrogenase, 3(or 4)-undecanol dehydrogenase, 2-methyl-3(or 4)-hexanol dehydrogenase, 5-methyl-3(or 4)-heptanol dehydrogenase,

6-methyl-3 (or 4)-heptanol dehydrogenase, 7-methyl-3(or 4)-octanol dehydrogenase, 1-phenyl-2(or 3)-pentanol dehydrogenase, 1-(4-hydroxyphenyl)-2(or 3)-pentanol dehydrogenase, 1-indole-2(or 3)-pentanol dehydrogenase, 4(or 5)-nonanol dehydrogenase, 4(or 5)-decanol dehydrogenase, 4(or 5)-undecanol dehydrogenase, 4(or 5)-dodecanol dehydrogenase, 2-methyl-3(or 4)-heptanol dehydrogenase, 3-methyl-4(or 5)-octanol dehydrogenase, 2-methyl-4(or 5)-octanol dehydrogenase, 8-methyl-4(or 5)-nonanol dehydrogenase, 1-phenyl-2(or 3)-hexanol dehydrogenase, 1-(4-hydroxyphenyl)-2(or 3)-hexanol dehydrogenase, 1-indole-2(or 3)-hexanol dehydrogenase, 4(or 5)-undecanol dehydrogenase, 5(or 6)-undecanol dehydrogenase, 5(or 6)-tridecanol dehydrogenase, 2-methyl-3(or 4)-octanol dehydrogenase, 3-methyl-4(or 5)-nonanol dehydrogenase, 2-methyl-4(or 5)-nonanol dehydrogenase, 2-methyl-5(or 6)-decanol dehydrogenase, 1-phenyl-2(or 3)-heptanol dehydrogenase, 1-(4-hydroxyphenyl)-2(or 3)-heptanol dehydrogenase, 1-indole-2(or 3)-heptanol dehydrogenase, 6(or 7)-tridecanol dehydrogenase, 6(or 7)-tetradecanol dehydrogenase, 2-methyl-3(or 4)-nonanol dehydrogenase, 3-methyl-4(or 5)-decanol dehydrogenase, 2-methyl-4(or 5)-decanol dehydrogenase, 2-methyl-5(or 6)-undecanol dehydrogenase, 1-phenyl-2(or 3)-octanol dehydrogenase, 1-(4-hydroxyphenyl)-2(or 3)-octanol dehydrogenase, 1-indole-2(or 3)-octanol dehydrogenase, 7(or 8)-pentadecanol dehydrogenase, 2-methyl-3(or 4)-decanol dehydrogenase, 3-methyl-4(or 5)-undecanol dehydrogenase, 2-methyl-4(or 5)-undecanol dehydrogenase, 2-methyl-5(or 6)-tridecanol dehydrogenase, 1-phenyl-2(or 3)-decanol dehydrogenase, 1-(4-hydroxyphenyl)-2 (or 3)-nonanol dehydrogenase, 1-indole-2(or 3)-nonanol dehydrogenase, 2-methyl-3(or 4)-undecanol dehydrogenase, 3-methyl-4(or 5)-dodecanol dehydrogenase, 2-methyl-4(or 5)-dodecanol dehydrogenase, 2-methyl-5(or 6)-tridecanol dehydrogenase, 1-phenyl-2(or 3)-decanol dehydrogenase, 1-(4-hydroxyphenyl)-2 (or 3)-decanol dehydrogenase, 1-indole-2(or 3)-decanol dehydrogenase, 2,5-dimethyl-3(or 4)-heptanol dehydrogenase, 2,6-dimethyl-3(or 4)-heptanol dehydrogenase, 2,7-dimethyl-3(or 4)-octanol dehydrogenase, 1-phenyl-4-methyl-2(or 3)-pentanol dehydrogenase, 1-(4-hydroxyphenyl)-4-methyl-2(or 3)-pentanol dehydrogenase, 1-indole-4-methyl-2(or 3)-pentanol dehydrogenase, 2,6-dimethyl-4(or 5)-octanol dehydrogenase, 3,8-dimethyl-4(or 5)-nonanol dehydrogenase, 1-phenyl-4-methyl-2(or 3)-hexanol dehydrogenase, 1-(4-hydroxyphenyl)-4-methyl-2 (or 3)-hexanol dehydrogenase, 1-indole-4-methyl-2(or 3)-hexanol dehydrogenase, 2,8-dimethyl-4(or 5)-nonanol dehydrogenase, 1-phenyl-5-methyl-2(or 3)-hexanol dehydrogenase, 1-(4-hydroxyphenyl)-5-methyl-2(or 3)-hexanol dehydrogenase, 1-indole-5-methyl-2(or 3)-hexanol dehydrogenase, 1-phenyl-6-methyl-2(or 3)-heptanol dehydrogenase, 1-(4-hydroxyphenyl)-6-methyl-2(or 3)-heptanol dehydrogenase, 1-indole-6-methyl-2(or 3)-heptanol dehydrogenase, 1-(4-hydroxyphenyl)-4-phenyl-2(or 3)-butanol dehydrogenase, 1-indole-4-phenyl-2(or 3)-butanol dehydrogenase, 1-indole-4-(4-hydroxyphenyl)-2(or 3)-butanol dehydrogenase, 1,10-diamino-5-decanol dehydrogenase, 1,4-di(4-hydroxyphenyl)-2-butanol dehydrogenase, 2-hexanol-1,6-dicarboxylic acid dehydrogenase, phenylethanol dehydrogenase, 4-hydroxyphenylethanol dehydrogenase, Indole-3-ethanol dehydrogenase, and the like.

[0305] In certain aspects, a selected alcohol dehydrogenase or secondary alcohol dehydrogenase may have one or more of the above exemplified alcohol dehydrogenase activities, such

as a 2-butanol dehydrogenase activity, 3-hexanol dehydrogenase activity, and/or a 4-octanol dehydrogenase activity, among others.

[0306] In certain aspects, a recombinant microorganism may comprise one or more secondary alcohol dehydrogenases encoded by a nucleotide reference sequence selected from SEQ ID NO:109, 111, 113, 115, 117, 119, 121, 123, 125, 127, 129, 131, 133, 135, 137, 139, and 141, or an enzyme having a polypeptide sequence selected from SEQ ID NO:110, 112, 114, 116, 118, 120, 122, 124, 126, 128, 130, 132, 134, 136, 138, 140, and 142, including biologically active fragments or variants thereof, such as optimized variants. Certain aspects may also comprises nucleotide sequences or polypeptide sequences having 80%, 85%, 90%, 95%, 97%, 98%, 99% sequence identity to SEQ ID NOS:109-142.

[0307] For the secondary alcohol dehydrogenase sequences referred to above, SEQ ID NO:109 is the nucleotide sequence and SEQ ID NO:110 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-1: PP_1946) isolated from *Pseudomonas putida* KT2440. SEQ ID NO:111 is the nucleotide sequence and SEQ ID NO:112 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-2: PP_1817) isolated from *Pseudomonas putida* KT2440.

[0308] SEQ ID NO:113 is the nucleotide sequence and SEQ ID NO:114 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-3: PP_1953) isolated from *Pseudomonas putida* KT2440. SEQ ID NO:115 is the nucleotide sequence and SEQ ID NO:116 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-4: PP_3037) isolated from *Pseudomonas putida* KT2440.

[0309] SEQ ID NO:117 is the nucleotide sequence and SEQ ID NO:118 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-5: PP_1852) isolated from *Pseudomonas putida* KT2440. SEQ ID NO:119 is the nucleotide sequence and SEQ ID NO:120 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-6: PP_2723) isolated from *Pseudomonas putida* KT2440.

[0310] SEQ ID NO:121 is the nucleotide sequence and SEQ ID NO:122 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-7: PP_2002) isolated from *Pseudomonas putida* KT2440. SEQ ID NO:123 is the nucleotide sequence and SEQ ID NO:124 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-8: PP_1914) isolated from *Pseudomonas putida* KT2440.

[0311] SEQ ID NO:125 is the nucleotide sequence and SEQ ID NO:126 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-9: PP_1914) isolated from *Pseudomonas putida* KT2440. SEQ ID NO:127 is the nucleotide sequence and SEQ ID NO:128 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-10: PP_3926) isolated from *Pseudomonas putida* KT2440.

[0312] SEQ ID NO:129 is the nucleotide sequence and SEQ ID NO:130 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-11: PFL_1756) isolated from *Pseudomonas fluorescens* Pf-5. SEQ ID NO:131 is the nucleotide sequence and SEQ ID NO:132 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-12: KPN_01694) isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578.

[0313] SEQ ID NO:133 is the nucleotide sequence and SEQ ID NO:134 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-13: KPN_02061) isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH

78578. SEQ ID NO:135 is the nucleotide sequence and SEQ ID NO:136 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-14: KPN_00827) isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578.

[0314] SEQ ID NO:137 is the nucleotide sequence and SEQ ID NO:138 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-16: KPN_01350) isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578. SEQ ID NO:139 is the nucleotide sequence and SEQ ID NO:140 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-17: KPN_03369) isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578. SEQ ID NO:141 is the nucleotide sequence and SEQ ID NO:142 is the polypeptide sequence of a secondary alcohol dehydrogenase (2adh-18: KPN_03363) isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578.

[0315] In certain aspects, an alcohol dehydrogenase (e.g., DEHU hydrogenase), a secondary alcohol dehydrogenase (2ADH), a fragment, variant, or derivative thereof, or any other enzyme that utilizes such an active site, may comprise at least one of a nicotinamide adenine dinucleotide (NAD⁺), NADH, nicotinamide adenine dinucleotide phosphate (NADP⁺), or NADPH binding motif. In certain embodiments, the NAD⁺, NADH, NADP⁺, or NADPH binding motif may be selected from the group consisting of Y-X-G-G-X-Y, Y-X-X-G-G-X-Y, Y-X-X-X-G-G-X-Y, Y-X-G-X-X-Y, Y-X-X-G-G-X-X-Y, Y-X-X-X-G-X-X-Y, Y-X-G-X-Y, Y-X-X-G-X-Y, Y-X-X-X-G-X-Y, and Y-X-X-X-X-G-X-Y; wherein Y is independently selected from alanine, glycine, and serine, wherein G is glycine, and wherein X is independently selected from a genetically encoded amino acid.

[0316] As one example of a step in a reduction and dehydration pathway, α -hydroxy cyclopentanone may be reduced to 1,2-cyclopentanediol. For example, the glycerol dehydrogenase isolated from *Hansenula ofumaensis* favors the reduction of α -hydroxy ketones and α -keto ketones, and has very broad substrate specificity. The similar alcohol dehydrogenase derived from *Hansenula polymorpha* and meso-2,3-butanediol dehydrogenase has similar properties. Certain embodiments may incorporate a 1,2-cyclopentanediol dehydrogenase to the microbial system or isolated microorganism. Other embodiments may incorporate a glycerol dehydrogenase from *Hansenula ofumaensis*, *Hansenula polymorpha*, *Klebsiella pneumoniae*, or any other suitable organism.

[0317] By way of example, a chemical or hydrocarbon such as 1,2-cyclopentanediol may be dehydrated to form cyclopentanone as one enzymatic step in a reduction and dehydration pathway. There are at least two different types of diol dehydratases that may catalyze dehydration of chemicals such as 1,2-cyclopentanediol. Certain embodiments of microbial system comprising a reduction and dehydration pathway will comprise diol dehydratases such as 1,2-cyclopentanediol dehydratase.

[0318] In the last enzymatic step for a reduction and dehydration pathway, the conversion of such exemplary chemicals as α -hydroxy cyclopentanone to cyclopentanol may include the reduction of cyclopentanone to cyclopentanol. This step may be catalyzed by cyclopentanol dehydrogenase, which is found in *Comomonas* sp. strain NCIMB 9872 and its gene (cpnA) has been isolated. Certain embodiments of a microbial system or isolated microorganism may comprise a cyclopentanol dehydrogenase, such as that expressed by cpnA in *Comomonas* sp. strain NCIMB 9872, among others described herein.

[0319] As detailed below, in certain embodiments, selected C—C ligation pathways may be utilized in combination with selected components or enzymes of a reduction and dehydration pathway to produce a commodity chemical, or intermediate thereof.

[0320] For example, certain embodiments include a method wherein the C—C ligation pathway may comprise an acetaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 2,3-butanediol dehydrogenase, a 2,3-butanediol dehydratase, and a 2-butanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise a propionaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 3,4-hexanediol dehydrogenase, a 3,4-hexanediol dehydratase, and a 3-hexanol dehydrogenase.

[0321] Additional embodiments include a method wherein the C—C ligation pathway may comprise a butyraldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 4,5-octanediol dehydrogenase, a 4,5-octanediol dehydratase, and a 4-octanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise a butyraldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 5,6-decanediol dehydrogenase, a 5,6-decanediol dehydratase, and a 5-decanol dehydrogenase.

[0322] Additional embodiments include a method wherein the C—C ligation pathway may comprise a butyraldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 6,7-dodecanediol dehydrogenase, a 6,7-dodecanediol dehydratase, and a 6-dodecanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise a butyraldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 7,8-tetradecanediol dehydrogenase, a 7,8-tetradecanediol dehydratase, and a 7-tetradecanol dehydrogenase.

[0323] Additional embodiments include a method wherein the C—C ligation pathway may comprise a butyraldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 8,9-hexadecanediol dehydrogenase, a 8,9-hexadecanediol dehydratase, and a 8-hexadecanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise an isobutyraldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 2,5-dimethyl-3,4-hexanediol dehydrogenase, a 2,5-dimethyl-3,4-hexanediol dehydratase, and a 2,5-dimethyl-3-hexanol dehydrogenase.

[0324] Additional embodiments include a method wherein the C—C ligation pathway may comprise a 2-methyl-butyraldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 3,6-dimethyl-4,5-octanediol dehydrogenase, a 3,6-dimethyl-4,5-octanediol dehydratase, and a 3,6-dimethyl-4-octanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise a 3-methyl-butyraldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 2,7-dimethyl-4,5-octanediol dehydrogenase, a 2,7-dimethyl-4,5-octanediol dehydratase, and a 2,7-dimethyl-4-octanol dehydrogenase.

[0325] Additional embodiments include a method wherein the C—C ligation pathway may comprise a 3-methyl-butyraldehyde lyase and wherein the reduction and dehydration

pathway may comprise at least one of a 2,9-dimethyl-5,6-decanediol dehydrogenase, a 2,9-dimethyl-4,5-decanediol dehydratase, and a 2,9-dimethyl-4-decanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise a phenylacetaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 1,4-diphenyl-2,3-butanediol dehydrogenase, a 1,4-diphenyl-2,3-butanediol dehydratase, and a 1,4-diphenyl-2-butanol dehydrogenase.

[0326] Additional embodiments include a method wherein the C—C ligation pathway may comprise a phenylacetaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a bis-1,4-(4-hydroxyphenyl)-2,3-butanediol dehydrogenase, a bis-1,4-(4-hydroxyphenyl)-2,3-butanediol dehydratase, and a bis-1,4-(4-hydroxyphenyl)-2-butanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise a phenylacetaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 1,4-diindole-2,3-butanediol dehydrogenase, a 1,4-diindole-2,3-butanediol dehydratase, and a 1,4-diindole-2-butanol dehydrogenase.

[0327] Additional embodiments include a method wherein the C—C ligation pathway may comprise an α -keto adipate carboxylase, and wherein the reduction and dehydration pathway may comprise at least one of a 1,2-cyclopentenediol dehydrogenase, a 1,2-cyclopentenediol dehydratase, and a cyclopentanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of an acetaldehyde/propionaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 2,3-pentenediol dehydrogenase, a 2,3-pentenediol dehydratase, and a 2(or 3)-pentanol dehydrogenase.

[0328] Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of an acetaldehyde/butyraldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 2,3-hexanediol dehydrogenase, a 2,3-hexanediol dehydratase, and a 2(or 3)-hexanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of an acetaldehyde/pentaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 2,3-heptanediol dehydrogenase, a 2,3-heptanediol dehydratase, and a 2(or 3)-heptanol dehydrogenase.

[0329] Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of an acetaldehyde/hexylaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 2,3-octanediol dehydrogenase, a 2,3-octanediol dehydratase, and a 2(or 3)-octanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of an acetaldehyde/octaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 2,3-nonanediol dehydrogenase, a 2,3-nonanediol dehydratase, and a 2(or 3)-nonanol dehydrogenase.

[0330] Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of an acetaldehyde/isobutyraldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 4-methyl-2,3-pentenediol dehydrogenase, a 4-methyl-2,3-pentenediol dehydratase, and a 4-methyl-2(or 3)-pentanol

dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of an acetaldehyde/2-methyl-butyraldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 4-methyl-2,3-hexanediol dehydrogenase, a 4-methyl-2,3-hexanediol dehydratase, and a 4-methyl-2(or 3)-hexanol dehydrogenase.

[0331] Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of an acetaldehyde/3-methyl-butyraldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 5-methyl-2,3-hexanediol dehydrogenase, a 5-methyl-2,3-hexanediol dehydrogenase, and a 5-methyl-2(or 3)-hexanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of an acetaldehyde/4-methyl-pentaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 6-methyl-2,3-heptanediol dehydrogenase, a 6-methyl-2,3-heptanediol dehydratase, and a 6-methyl-2(or 3)-heptanol dehydrogenase.

[0332] Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of an acetaldehyde/phenylacetaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 1-phenyl-2,3-butanediol dehydrogenase, a 1-phenyl-2,3-butanediol dehydratase, and a 1-phenyl-2(or 3)-butanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of an acetaldehyde/4-hydroxyphenylacetaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 1-(4-hydroxyphenyl)-2,3-butanediol dehydrogenase, a 1-(4-hydroxyphenyl)-2,3-butanediol dehydratase, and a 1-(4-hydroxyphenyl)-2(or 3)-butanol dehydrogenase.

[0333] Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of an acetaldehyde/indoleacetaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 1-indole-2,3-butanediol dehydrogenase, a 1-indole-2,3-butanediol dehydratase, and a 1-indole-2(or 3)-butanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of a propionaldehyde/butyraldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 3,4-heptanediol dehydrogenase, a 3,4-heptanediol dehydratase, and a 3(or 4)-heptanol dehydrogenase.

[0334] Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of a propionaldehyde/pentaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 3,4-octanediol dehydrogenase, a 3,4-octanediol dehydratase, and a 3(or 4)-octanol dehydrogenase. Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of a propionaldehyde/hexylaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 3,4-nonanediol dehydrogenase, a 3,4-nonanediol dehydratase, and a 3(or 4)-nonanol dehydrogenase.

[0335] Additional embodiments include a method wherein the C—C ligation pathway may comprise at least one of a propionaldehyde/heptaldehyde lyase and wherein the reduction and dehydration pathway may comprise at least one of a 3,4-decanediol dehydrogenase, a 3,4-decanediol dehydratase, and a 3(or 4)-decanol dehydrogenase. Additional

dehydration pathway may comprise at least one of a 2,3-hexanediol-1,6-dicarboxylic acid dehydrogenase, a 2,3-hexanediol-1,6-dicarboxylic acid dehydratase, and a 2-hexanol-1,6-dicarboxylic dehydrogenase.

[0375] Certain embodiments of a microbial system or recombinant microorganism may comprise genes encoding enzymes that are able to catalyze (e.g., reduction and dehydration) the conversion of 4-octanol to octene or octane. Other embodiments may comprise redesigned or de novo designed enzymes for this reduction and dehydration pathway. For example, three redesigned enzymes could convert 4-octanone to either 3- and 4-octene. The first step could be catalyzed by redesigned isocitrate dehydrogenase. This enzyme could catalyze the formation of 4-hydroxy-3(or 5)-carboxylic octane. The 4-hydroxy group could be phosphorylated by redesigned kinase. Finally, redesigned mevalonate diphosphate decarboxylase catalyzes the formation of 3(or 4)-octene.

[0376] In other embodiments, several redesigned enzymes could convert 4-octanone to octane. For example, the 4-hydroxy-3(or 5)-carboxylic octane is sequentially reduced and dehydrated to form 3(or 5)-carboxylic octane. Redesigned enzymes involved in fatty acid metabolism can catalyze these reactions. The 3(or 5)-carboxylic octane can be reduced to corresponding aldehyde by aldehyde dehydrogenase and the product may be decarbonylated to form octane catalyzed by a redesigned decarbonylase.

[0377] As noted above, for the production of certain commodity chemicals, such as 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol, and indole-3-ethanol, among other similar chemicals, a biosynthesis pathway (e.g., aldehyde biosynthesis pathway) may optionally or further comprise one or more genes encoding a decarboxylase enzyme, such as an indole-3-pyruvate decarboxylase (IPDC), to produce an aldehyde. In certain aspects, an IPDC may comprise an amino acid sequence that is at least 80%, 90%, 95%, 98%, or 99% identical to the amino acid sequence set forth in SEQ ID NO:312. An IPDC enzyme may comprise certain conserved amino acid residues, such as G24, D25, E48, A55, R60, G75, E89, H113, G252, G405, G413, G428, G430, and/or N456.

[0378] In these and other embodiments, a recombinant microorganism may comprise an aldehyde reductase, such as a phenylacetaldehyde reductase (PAR), to convert an aldehyde to a commodity chemical. In certain aspects, a PAR may comprise an amino acid sequence that is at least 80%, 90%, 95%, 98%, or 99% identical to the amino acid sequence set forth in SEQ ID NO:313, which shows the sequence of a PAR enzymed derived from *Rhodococcus* sp. ST-10. In certain aspects, a PAR enzyme may comprise at least one of a nicotinamide adenine dinucleotide (NAD⁺), NADH, nicotinamide adenine dinucleotide phosphate (NADP⁺), or NADPH binding motif. In certain embodiments, the NAD⁺, NADH, NADP⁺, or NADPH binding motif may be selected from the group consisting of Y-X-G-G-X-Y, Y-X-X-G-G-X-Y, Y-X-X-G-G-X-Y, Y-X-G-X-X-Y, Y-X-X-G-G-X-X-Y, Y-X-X-X-G-X-X-Y, Y-X-G-X-Y, Y-X-X-G-X-Y, Y-X-X-X-G-X-Y, and Y-X-X-X-G-X-Y; wherein Y is independently selected from alanine, glycine, and serine, wherein G is glycine, and wherein X is independently selected from a genetically encoded amino acid.

[0379] In certain embodiments, such a recombinant microorganism may also or alternatively comprise a secondary alcohol dehydrogenase having an activity selected from at least one of a phenylethanol dehydrogenase activity, a 4-hy-

droxyphenylethanol dehydrogenase activity, and an Indole-3-ethanol dehydrogenase activity, to reduce the aldehyde to its corresponding alcohol (e.g. 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol, and indole-3-ethanol).

[0380] Embodiments of the present invention also include methods for converting a suitable monosaccharide to a commodity chemical comprising, (a) obtaining a suitable monosaccharide; (b) contacting the suitable monosaccharide with a microbial system for a time sufficient to convert to the suitable monosaccharide to the biofuel, wherein the microbial system comprises, (i) one or more genes encoding and expressing a fatty acid biosynthesis pathway, an amino acid biosynthetic pathway, and/or a short chain alcohol biosynthetic pathway; (ii) one or more genes encoding and expressing a keto-acid decarboxylase, aldehyde dehydrogenase, and/or alcohol dehydrogenase; and (iii) an enzymatic reduction pathway selected from (1) an enzymatic long chain alcohol reduction pathway, (2) an enzymatic decarbonylation pathway, (3) an enzymatic decarboxylation pathway, and (4) an enzymatic reduction pathway comprising (1), (2), and/or (3), thereby converting the suitable monosaccharide to the commodity chemical.

[0381] Embodiments of the present invention may comprise one or more genes encoding and expressing enzymes in a fatty acid synthesis pathway, which may be used, as one example, to produce biofuels in the form of alkanes, such as medium to long chain alkanes. In certain embodiments, the specificity of the fatty acid biosynthesis pathway in the microbial system may be recalibrated or redesigned. Merely by way of example, microorganisms generally produce a mixture of long chain fatty acids (e.g., *E. coli* naturally produce large quantities of long chain fatty acids (C16-C19: <95% in whole cells) and small quantity of medium chain fatty acids (C12: 2% and C14: 5% in whole cells)).

[0382] In certain embodiments, the recalibration or re-engineering may be directed to increasing production of medium chain alkanes, including, but not limited to, caprylate (C8), caprate (C10), laurate (C12), myristate (C14), and palmitate (C16), as alkanes produced from these fatty acids are major components of gasoline, diesels, and kerosene. In addition to these fatty acids, other embodiments may be directed to increased production of long chain fatty acids, including, but not limited to, stearate (C18), arachidonate (C20), behenate (C22) and longer fatty acids, as n-alkanes produced from these fatty acids are one of major components in heavy oils.

[0383] For example, *Cuphea* mainly accumulate medium chain fatty acids as major components in their seed oils, and these compositions alter depending on species. In particular, *Cuphea pulcherrima* accumulates caprylate (C8:0) 96%, *Cuphea koehneana* accumulates caprate (C10:0) 95.3%, and *Cuphea polymorpha* accumulates laurate (C12:0) 80.1%. Embodiments of the microbial systems or isolated microorganisms according to the present application may incorporate genes from various *Cuphea* species encoding enzymes involved in a fatty acid biosynthesis pathway, and these microorganisms may be directed in part to the production of middle chain fatty acids.

[0384] In other embodiments, acyl-acyl carrier protein (ACP) thioesterases (TEs) derived from various species including *Cuphea hookeriana*, *Cuphea palustris*, *Umbellularia californica*, and *Cinnamomum camphorum* may be over-expressed in such microorganisms as *E. coli*, wherein the specific activity for the formation of each medium chain

fatty acids, caprylate (C8), caprate (C10), laurate (C12), myristate (C14), and palmitate (C16) is improved over the wild type. Certain embodiments may include other enzyme components involved in fatty acid biosynthesis as known to a person skilled in the arts, including, but not limited to, ACP and β -ketoacyl ACP synthase (KAS) IV.

[0385] Microbial systems and isolated microorganisms of the present application may also incorporate fatty aldehyde dehydrogenases to reduce fatty acids to fatty aldehydes. Merely by way of explanation, the conversion of fatty acids to fatty aldehydes may be catalyzed by medium and/or long chain fatty aldehyde dehydrogenases isolated from various suitable organisms. Certain embodiments may incorporate, for example, a fatty aldehyde dehydrogenase derived from *Vibrio harveyi*.

[0386] Microbial systems and isolated microorganisms of the present application may also incorporate one or more enzymes that catalyze the conversion of fatty aldehydes to biofuels such as n-alkanes, including, for example, enzymes comprising an enzymatic long chain alcohol reduction pathway. Certain embodiments may incorporate genes from various other sources that encode enzymes capable of catalyzing the reduction and dehydration of fatty acids to biofuels, such as alkanes. For example, bacterial strain HD-1 is able to produce biofuels, such as n-alkanes, with various chain lengths, and also produces both odd and even numbered alkanes. Certain embodiments of the microbial systems and recombinant microorganisms provided herein may incorporate the HD-1 genes encoding the enzymes involved in this pathway.

[0387] Other embodiments may incorporate redesigned or de novo designed enzymes for this reduction pathway. For example, embodiments of the present invention may include a redesigned isocitrate dehydrogenase, which may catalyze the formation of 2-carboxy-1-alcohols. In certain embodiments, the 2-carboxy-1-alcohols may be sequentially reduced and dehydrated to form 2-carboxy-alkanes, which may be catalyzed by redesigned enzymes involved in fatty acid metabolism. The 2-carboxy-alkanes can be reduced to corresponding aldehyde by aldehyde dehydrogenase and then decarbonylated to form n-alkanes catalyzed by the redesigned decarbonylase as discussed below. Certain embodiments of these microbial systems may produce either even numbered n-alkanes, odd numbered n-alkanes, or both.

[0388] Certain embodiments of the present application may incorporate the genes encoding enzymes catalyzing decarbonylation, or an enzymatic decarbonylation pathway. Merely by way of example, green colonial alga *Botryococcus braunii*, race A, produces linear odd-numbered C27, C29, and C31 hydrocarbons that total up to 32% of the alga's dry weight. Microsomal preparations of this organism have decarbonylation activity. This decarbonylase from *B. braunii* culture is a cobalt-protoporphyrin IX containing enzyme. Certain microbial systems of isolated microorganisms may incorporate the gene encoding fatty aldehyde decarbonylase from *Botryococcus braunii*.

[0389] Other embodiments may include redesigned decarbonylase enzymes, for example, wherein the N-terminal membrane sequence is substituted. By way of explanation, the functional activity of a similar enzyme, cytochrome P450 containing Fe-protoporphyrin IX (heme), is improved by substituting N-terminal membrane associated sequence, and the

functional activity of decarbonylases of the present microbial systems may comprise similar substitutions or improvements.

[0390] Other embodiments may incorporate the genes encoding a Co-porphyrin synthase. In explanation, decarbonylase enzymes may use Co-protoporphyrin IX as a co-factor, and *Clostridium tetranomorphum* is able to incorporate cobalt into incubated protoporphyrin IX. Certain embodiments may incorporate the Co-porphyrin synthase from *Clostridium tetranomorphum*, or from other suitable microorganisms. Other embodiments may incorporate de novo designed decarbonylation enzymes using inorganic metals such as Co^{2+} , Fe^{2+} , and Ni^{2+} as catalysts.

[0391] Certain embodiments may comprise genes encoding the enzymes responsible for the formation of alkenes, or an enzymatic decarboxylation pathway. These genes may be derived or isolated from various sources, such as higher plants and insects. For example, higher plants such as germinating safflower (*Carthamus tinctorius* L.) produce a number of odd numbered 1-alkenes, including 1-pentadecene, 1-heptadecene, 1,8-heptadecadiene and 1,8,11-heptadecatriene besides about 80-90% 1,8,11,14-heptadecatetraene by decarboxylation from their corresponding fatty acids. Certain embodiments may incorporate the genes from higher plants such as *Carthamus tinctorius*.

[0392] Other embodiments may incorporate the genes encoding the enzymes responsible for the formation of alkenes (e.g., an enzymatic decarboxylation pathway) from microorganisms, including, but not limited to, such as bacterial strain DH-1. By way of explanation, bacterial strain DH-1 produces n-alkenes in addition to n-alkanes.

[0393] Other embodiments may incorporate the genes from de novo designed enzymes for an enzymatic decarboxylation pathway. For example, these redesigned enzymes convert β -hydroxy fatty acids to n-alkenes. The first step is catalyzed by a redesigned kinase, which catalyzes the phosphorylation of a β -hydroxy group. A redesigned mevalonate diphosphate decarboxylase then catalyzes the formation of n-alkenes, such as n-1-alkene.

[0394] Any microorganism may be utilized according to the present invention. In certain aspects, a microorganism is a eukaryotic or prokaryotic microorganism. In certain aspects, a microorganism is a yeast, such as *S. cerevisiae*. In certain aspects, a microorganism is a bacteria, such as a gram-positive bacteria or a gram-negative bacteria. Given its rapid growth rate, well-understood genetics, the variety of available genetic tools, and its capability in producing heterologous proteins, genetically modified *E. coli* may be used in certain embodiments of a microbial system as described herein, whether for the degradation and metabolism of a polysaccharide, such as alginate or pectin, or the formation or biosynthesis of commodity chemicals, such as biofuels.

[0395] Other microorganisms may be used according to the present invention, based in part on the compatibility of enzymes and metabolites to host organisms. For example, other organisms such as *Acetobacter aceti*, *Achromobacter*, *Acidiphilium*, *Acinetobacter*, *Actinomadura*, *Actinoplanes*, *Aeropyrum pernix*, *Agrobacterium*, *Alcaligenes*, *Ananas comosus* (M), *Arthrobacter*, *Aspergillus niger*, *Aspergillus oryzae*, *Aspergillus melleus*, *Aspergillus pulverulentus*, *Aspergillus saitoi*, *Aspergillus sojae*, *Aspergillus usamii*, *Bacillus alcalophilus*, *Bacillus amyloliquefaciens*, *Bacillus brevis*, *Bacillus circulans*, *Bacillus clausii*, *Bacillus lentus*, *Bacillus licheniformis*, *Bacillus macerans*, *Bacillus stearo-*

thermophilus, *Bacillus subtilis*, *Bifidobacterium*, *Brevibacillus brevis*, *Burkholderia cepacia*, *Candida cylindracea*, *Candida rugosa*, *Carica papaya* (L), *Cellulosimicrobium*, *Cephalosporium*, *Chaetomium erraticum*, *Chaetomium gracile*, *Clostridium*, *Clostridium butyricum*, *Clostridium acetobutylicum*, *Clostridium thermocellum*, *Corynebacterium* (glutamicum), *Corynebacterium efficiens*, *Escherichia coli*, *Enterococcus*, *Erwina chrysanthemi*, *Gliconobacter*, *Gluconacetobacter*, *Haloarcula*, *Humicola insolens*, *Humicola insolens*, *Kitasatospora setae*, *Klebsiella*, *Klebsiella oxytoca*, *Kluyveromyces*, *Kluyveromyces fragilis*, *Kluyveromyces lactis*, *Kocuria*, *Lactolactis*, *Lactobacillus*, *Lactobacillus fermentum*, *Lactobacillus sake*, *Lactococcus*, *Lactococcus lactis*, *Leuconostoc*, *Methylocystis*, *Methanobolus siciliae*, *Methanogenium organophilum*, *Methanobacterium bryantii*, *Microbacterium imperiale*, *Micrococcus lysodeikticus*, *Microlunatus*, *Mucor javanicus*, *Mycobacterium*, *Myrothecium*, *Nitrobacter*, *Nitrosomonas*, *Nocardia*, *Papaya carica*, *Pediococcus*, *Pediococcus halophilus*, *Penicillium*, *Penicillium camemberti*, *Penicillium citrinum*, *Penicillium emersonii*, *Penicillium roqueforti*, *Penicillium lilactinum*, *Penicillium multicolor*, *Paracoccus pantotrophus*, *Propionibacterium*, *Pseudomonas*, *Pseudomonas fluorescens*, *Pseudomonas denitrificans*, *Pyrococcus*, *Pyrococcus furiosus*, *Pyrococcus horikoshii*, *Rhizobium*, *Rhizomucor miehei*, *Rhizomucor pusillus* Lindt, *Rhizopus*, *Rhizopus delemar*, *Rhizopus japonicus*, *Rhizopus niveus*, *Rhizopus oryzae*, *Rhizopus oligosporus*, *Rhodococcus*, *Sccharomyces cerevisiae*, *Sclerotinia libertina*, *Sphingobacterium multivorum*, *Sphingobium*, *Sphingomonas*, *Streptococcus*, *Streptococcus thermophilus* Y-1, *Streptomyces*, *Streptomyces griseus*, *Streptomyces lividans*, *Streptomyces murinus*, *Streptomyces rubiginosus*, *Streptomyces violaceoruber*, *Streptoverticillium mobaraense*, *Tetragenococcus*, *Thermus*, *Thiosphaera pantotropha*, *Trametes*, *Trichoderma*, *Trichoderma longibrachiatum*, *Trichoderma reesei*, *Trichoderma viride*, *Trichosporon penicillatum*, *Vibrio alginolyticus*, *Xanthomonas*, yeast, *Zygosaccharomyces rouxii*, *Zymomonas*, and *Zymomonas mobilis*, may be utilized as recombinant microorganisms provided herein, and, thus, may be utilized according to the various methods of the present invention.

[0396] The following Examples are offered by way of illustration, not limitation.

EXAMPLES

Example 1

Engineering *E. Coli* to Grow on Alginate as a Sole Source of Carbon

[0397] Wild type *E. coli* cannot use alginate polymer or degraded alginate as its sole carbon source (see FIG. 4). *Vibrio splendidus*, however, is known to be able to metabolize alginate to support growth. To generate recombinant *E. coli* that use degraded alginate as its sole carbon source, a *Vibrio splendidus* fosmid library was constructed and cloned into *E. coli*.

[0398] To prepare the *Vibrio splendidus* fosmid library, genomic DNA was isolated from *Vibrio Splendidus* B01 (gift from Dr. Martin Polz, MIT) using the DNeasy Blood and Tissue Kit (Qiagen, Valencia, Calif.). A fosmid library was then constructed using Copy Control Fosmid Library Production Kit (Epicentre, Madison, Wis.). This library consisted of random genomic fragments of approximately 40 kb inserted into the vector pCC1 FOS (Epicentre, Madison, Wis.).

[0399] The fosmid library was packaged into phage, and *E. coli* DH10B cells harboring a pDONR221 plasmid (Invitrogen, Carlsbad, Calif.) carrying certain *Vibrio splendidus* genes (V12B01_02425 to V12B01_02480; encoding a type II secretion apparatus; see SEQ ID NO:1) were transfected with the phage library. This secretome region encodes a type II secretion apparatus derived from *Vibrio splendidus*, which was cloned into a pDONR221 plasmid and introduced into *E. coli* strain DH10B (see Example 1).

[0400] Transformants were selected for chloroamphenicol resistance and then screened for their ability to grow on degraded alginate. The resultant transformants were screened for growth on degraded alginate media. Degraded alginate media was prepared by incubating 2% Alginate (Sigma-Aldrich, St. Louis, Mo.) 10 mM Na-Phosphate buffer, 50 mM KCl, 400 mM NaCl with alginate lyase from *Flavobacterium* sp. (Sigma-Aldrich, St. Louis, Mo.) at room temperature for at least one week. This degraded alginate was diluted to a concentration of 0.8% to make growth media that had a final concentration of 1xM9 salts, 2 mM MgSO₄, 100 μM CaCl₂, 0.007% Leucine, 0.01% casamino acids, 1.5% NaCl (this includes all sources of sodium: M9, diluted alginate and added NaCl).

[0401] One fosmid-containing *E. coli* clone was isolated that grew well on this media. The fosmid DNA from this clone was isolated and prepared using FosmidMAX DNA Purification Kit (Epicentre, Madison, Wis.). This isolated fosmid was transferred back into DH10B cells, and these cells were tested for the ability to grown on alginate.

[0402] The results are illustrated in FIG. 4, which shows that certain fosmid-containing *E. coli* clones are capable of growing on alginate as a sole source of carbon. *Agrobacterium tumefaciens* provides a positive control (see hatched circles). As a negative control, *E. coli* DH10B cells are not capable of growing on alginate (see immediate left of positive control).

[0403] These results also demonstrate that the sequences contained within this *Vibrio splendidus* derived fosmid clone are sufficient to confer on *E. coli* the ability to grow on degraded alginate as a sole source of carbon. Accordingly, the type II secretion machinery sequences contained within the pDONR221 vector (i.e., SEQ ID NO:1), which was harbored by the original DH10B cells, were not necessary for growth on degraded alginate.

[0404] The isolated fosmid sufficient to confer growth alginate as a sole source of carbon was sequenced by Elim Biopharmaceuticals (Hayward, Calif.) using the following primers:

Uni R3—GGGCGGCCGCAAGGGGTTTCGCGTTGGCCGA (SEQ ID NO:147) and PCC1FOS_uni_F—GGAGAAAATACCGCATCAGGCG (SEQ ID NO:148). Sequencing showed that the vector contained a genomic DNA section that contained the full length genes V12B01_24189 to V12B01_24249 (see SEQ ID NOS:2-64). SEQ ID NO:2 shows the nucleotide sequence of entire region between V12B01_24189 to V12B01_24249. SEQ ID NOS:3-64 show the individual putative genes contained within SEQ ID NO:2. In this sequence, there is a large gene before V12B01_24189 that is truncated in the fosmid clone. The large gene V12B01_24184 is a putative protein with similarity to autotransporters and belongs to COG3210, which is a cluster of orthologous proteins that include large exoproteins involved in heme utilization or adhesion. In the fosmid clone, V12B01_24184 is N-terminally truncated such that the first

5893 bp are missing from the predicted open reading frame (which is predicted to contain 22889 bp in total).

Example 2

Engineering *E. Coli* to Grow on Pectin as a Sole Source of Carbon

[0405] Wild type *E. coli* is not capable of growing on pectin, di-, or tri-galacturonates as a sole source of carbon. To identify the minimal components to confer on *E. coli* the capability of growing on pectin, di- and/or tri-galacturonates as a sole source of carbon, an *E. coli* strain BL21(DE3) harboring both the pBBRGal3P plasmid and the pTrcogl-kdgR plasmid was engineered and tested for the ability to grow on these polysaccharides.

[0406] The pBBRGal3P plasmid was engineered to contain certain genomic region of *Erwinia carotovora* subsp. *Atroseptica* SCR11043, comprising several genes (kdgF, kduI, kduD, pelW, togM, togN, toga, togB, kdgM, and paeX) encoding certain enzymes (kduI, kduD, ogl, pelW and paeX), transporters (togM, togN, togA, togB, and kdgM), and regulatory proteins (kdgR) responsible for the degradation of di- and trigalacturonate. SEQ ID NO:65 shows the nucleotide sequence of the kdgF-PaeX region from *Erwinia carotovora* subsp. *Atroseptica* SCR11043.

[0407] To construct this plasmid, the DNA sequence encoding kdgF, kduI, kduD, pelW, togM, togN, toga, togB, kdgM, paeX, ogl, and kdgR of *Erwinia carotovora* subsp. *Atroseptica* SCR11043 was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 6 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CGGGATCC AAGTTGCAGGATATGACGAAAGCG-3') (SEQ ID NO:149) and reverse (5'-GCTCTAGA AGATTATCCCTGTCTGCGGAAGCGG-3') (SEQ ID NO:150) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Erwinia carotovora* subsp. *Atroseptica* SCR11043 genome (ATCC) in 50 μl.

[0408] The vector pBBR1MCS-2 was then amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 2.5 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-GCTCTAGA GGGGTGCCTAATGAGTGAGCTAAC-3') (SEQ ID NO:151) and reverse (5'-CGGGATCC GCGTAAATATTTGTAAAATTCGC-3') (SEQ ID NO:152) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng pBBR1MCS-2 in 50 μl. Both amplified DNA fragments were digested with BamHI and XbaI and ligated.

[0409] The pTrcogl-kdgR plasmid was engineered to contain certain genomic regions of *Erwinia carotovora* subsp. *Atroseptica* SCR11043, comprising two genes (ogl and kdgR) encoding an enzyme (ogl) and a regulatory protein (kdgR) responsible for degradation of di- and trigalacturonate. SEQ ID NO:66 shows the nucleotide sequence of ogl-kdgR from *Erwinia carotovora* subsp. *Atroseptica* SCR11043.

[0410] To prepare this construct, the DNA sequence encoding ogl and kdgR of *Erwinia carotovora* subsp. *Atroseptica* SCR11043 was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 4 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-GCTCTAGA GTTATGTCGCACCCGCCGTTGG-3')

(SEQ ID NO:153) and reverse (5'-CCCAAGC TTA-GAAAGGGAAATTGTGGTAGCCC-3') (SEQ ID NO:154) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Erwinia carotovora* subsp. *Atroseptica* SCR11043 genome (ATCC) in 50 μl. The amplified DNA fragment was digested with XbaI and HindIII and ligated into pTrc99A pre-digested with the same restriction enzymes.

[0411] The plasmids pBBRGal3P and pTrcogl-kdgR were co-transformed into *E. coli* strain BL21(DE3). A single colony was inoculated into LB media containing 50 μg/ml kanamycin and 100 μg/ml ampicillin, and the culture was grown in incubation shaker with 200 rpm at 37 C. When culture reached OD 600 nm of 0.6, 500 μl of culture was transferred to eppendorf tube and centrifuged to pellet the cells. The cells were resuspended into 50 μl of M9 media containing 2 mM MgSO₄, 100 μM CaCl₂, 0.4% di- or trigalacturonate, and 5 μl of this solution was inoculated into 500 μl of fresh M9 media containing 2 mM MgSO₄, 100 μM CaCl₂, 0.4% di- or trigalacturonate. The culture was grown in incubation shaker with 200 rpm at 37 C.

[0412] The results in FIG. 5A show that these two plasmids were sufficient to provide *E. coli* ability to grow on di- and trigalacturonate as sole source of carbon, but not pectin. In particular, these results show that the regions kdgF-paeX and ogl-kdgR were sufficient to confer this ability on *E. coli*.

[0413] Based on the information obtained from the above experiments, it was considered whether the introduction of pectate lyase, pectate acetyltransferase, and methyltransferase might confer *E. coli* capability of growing on pectin. To test this hypothesis, *E. coli* strain DH5α bacterial cells were engineered to contain both the pROU2 plasmid and the pPEL74 plasmid.

[0414] The pROU2 plasmid contains certain genomic regions of *Erwinia chrysanthemi*, comprising several genes (kdgF, kduI, kduD, pelW, togM, togN, toga, togB, kdgM, paeX, ogl, and kdgR) encoding enzymes (kduI, kduD, ogl, pelW, and paeX), transporters (togM, togN, toga, togB, and kdgM), and regulatory proteins (kdgR) responsible for degradation of di- and trigalacturonate.

[0415] The pPEL74 plasmid contains certain genomic regions of *Erwinia chrysanthemi*, comprising several genes (pela, pelE, paeY, and pem) encoding pectate lyases (pela and pelE), pectin acetyltransferases (paeY), and pectin methyltransferase (pem).

[0416] As shown in FIG. 5B, *E. coli* DH5α engineered with pROU2 and pPEL74 was able to grow on pectin as a sole source of carbon, showing that the genes contained within these plasmids are sufficient to confer this property on an organism that is otherwise incapable of growing on pectin as a sole source of carbon.

Example 3

In Vitro Conversion of Alginate to Pyruvate and Glycerinaldehyde-3-Phosphate

[0417] The ability of an enzyme mixture containing all required enzymes for alginate degradation and metabolism was investigated for its ability to produce pyruvate from alginate. In addition, various novel alcohol dehydrogenases (ADHs), such as ADH1-12 (see SEQ ID NOS:69-92), isolated from *Agrobacterium tumefaciens*, were tested for their ability to catalyze either DEHU or mannuronate hydrogenation.

[0418] A simplified metabolic pathway for alginate degradation and metabolism is shown in FIG. 2. Alginate can be degraded by at least two different methodologies: enzymatic and chemical methodologies.

[0419] In enzymatic degradation, the degradation of alginate is catalyzed by a family of enzymes called alginate lyases. For this experiment, Atu3025 was used. Atu3025 is an exolytically acting enzyme and yields DEHU from alginate polymer. DEHU is converted to the common hexuronate metabolite, KDG. This reaction is catalyzed by alcohol dehydrogenases (e.g., DEHU hydrogenases).

[0420] Chemical degradation catalyzed by acid solution, such as formate, yields a monosaccharide mannuronate. Mannuronate is then converted to mannonate, which is catalyzed by enzymes with mannonate dehydrogenase (mannuronate reductase) activity. In bacteria, mannonate dehydratase (UxuA) catalyzes dehydration from mannuronate to form KDG.

[0421] KDG is readily metabolized to form of pyruvate and glyceraldehydes-3-phosphate (G3P). KDG is first phosphorylated to KDG-6-phosphate (KDGP), which is catalyzed by KDG kinase, and then broken down to pyruvate and G3P, which is catalyzed by KDGP aldolase.

[0422] Preparation of oligoalginate lyase Atu3025 derived from *Agrobacterium tumefaciens* C58. pETAtu3025 was constructed based on pET29 plasmid backbone (Novagen). The oligoalginate lyase Atu3025 was amplified by PCR: 98° C. for 10 sec, 55° C. for 15 sec, and 72° C. for 60 sec, repeated for 30 times. The reaction mixture contained 1× Phusion buffer, 2 mM dNTP, 0.5 μM forward (5'-GGAATTCATATGCGTCCCTCTGCCCGGCC-3') (SEQ ID NO:155) and reverse (5'-CGGGATCCTTAGAACTGCTTGGGAAGG-GAG-3') (SEQ ID NO:156) primers, 2.5 U Phusion DNA polymerase (Finezyme), and an aliquot of *Agrobacterium tumefaciens* C58 (gift from Professor Eugene Nester, University of Washington) cells as a template in total volume of 100

μl. The amplified fragment was digested with NdeI and BamHI and ligated into pET29 pre-digested with the same enzymes using T4 DNA ligase to form pETAtu3025. The constructed plasmid was sequenced (Elim Biopharmaceuticals) and the DNA sequence of the insert was confirmed. The nucleotide sequence of the Atu3025 insert is provided in SEQ ID NO:67. The polypeptide sequence encoded by the Atu3025 insert is provided in SEQ ID NO:68.

[0423] The pETAtu3025 was transformed into *Escherichia coli* strain BL21(DE3). A colony of BL21(DE3) containing pETAtu3025 was inoculated into 50 ml of LB media containing 50 μg/ml kanamycin (Km⁵⁰). This strain was grown in an orbital shaker with 200 rpm at 37° C. The 0.2 mM IPTG was added to the culture when the OD_{600 nm} reached 0.6, and the induced culture was grown in an orbital shaker with 200 rpm at 20° C. 24 hours after the induction, the cells were harvested by centrifugation at 4,000 rpm×g for 10 min and the pellet was resuspended into 2 ml of Bugbuster (Novagen) containing 10 μl of Lysonase™ Bioprocessing Reagent (Novagen). The solution was again centrifuged at 4,000 rpm×g for 10 min and the supernatant was obtained.

[0424] Construction of pETADH1 through pETADH12. DNA sequences of ADH1-12 of *Agrobacterium tumefaciens* C58 were amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (Table 1) and reverse (Table 1) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Agrobacterium tumefaciens* C58 genome in 50 μl. Amplified DNA fragment was digested with NdeI and BamHI and ligated into pET28 pre-digested with the same restriction enzymes. For DNA sequences with internal NdeI or BamHI site, front and bottom half sequences of each ADH were first amplified using described method. The resulting two DNA fragments were gel purified and spliced by overlapping PCR.

TABLE 1

| Primers used to amplify ADH1-12 from <i>Agrobacterium tumefaciens</i> C58. | | | |
|--|------------------------------|--|---|
| Name | <i>A. tumefaciens</i> C58 | Forward Primer | Reverse Primer |
| ADH1 | Atu1557 | GGAATTCATATGTTTACAACGTCCGCCTA (SEQ ID NO:276) | GCTTGACGGCCATGTGGCCGAGGCCGC (SEQ ID NO:277) |
| | | GCGGCCTCGGCCACATGGCCGTCAAGC (SEQ ID NO:278) | CGGGATCCTTAGCGCCCTTCTGGCGCG (SEQ ID NO:279) |
| ADH2 | Atu2022 | GGAATTCATATGGCTATTGCAAGAGGTTA (SEQ ID NO:280) | CGGGATCCTTAAGCGTCGAGCGAGGCCA (SEQ ID NO:281) |
| ADH3 | Atu0626 | GGAATTCATATGACTAAAACAATGAAGGC (SEQ ID NO:282) | CACCGGGCCGGGTCCGGTATTGCCA (SEQ ID NO:283) |
| | | TGGCAATACCGGACCCCGCCCGGTG (SEQ ID NO:284) | CGGGATCCTTAGCGCGGATCCACGA (SEQ ID NO:285) |
| ADH4 | Atu5240 | GGAATTCATATGACCGGGCGAACCAGCC (SEQ ID NO:286) | ATAGCCGCTCATACGCCCTCGGTTGCCT (SEQ ID NO:287) |
| | | AGGCAACCGAGGCGTATGAGCGGCTAT (SEQ ID NO:288) | CGGGATCCTTAAGCGCCGTCCGGAAGGA (SEQ ID NO:289) |
| ADH5 | Atu3163 | GGAATTCATATGACCATGCATGCCATTCA (SEQ ID NO:290) | CGGGATCCTTATTCGGCTGCAAATTGCA (SEQ ID NO:291) |
| ADH6 | Atu2151 | GGAATTCATATGCGCGCCTTTATTACGA (SEQ ID NO:292) | CGGGATCCTTATTCGAACCGGTCGATGA (SEQ ID NO:293) |

TABLE 1-continued

| Primers used to amplify ADH1-12 from <i>Agrobacterium tumefaciens</i> C58. | | | |
|--|------------------------------|--|---|
| Name | <i>A. tumefaciens</i> C58 | Forward Primer | Reverse Primer |
| ADH7 | Atu2814 | GGAATTCATATGCTGGCGATTTTCTGTGA (SEQ ID NO:294) | CGGGATCCTTATGCGACCTCCACCATGC (SEQ ID NO:295) |
| ADH8 | Atu5447 | GGAATTCATATGAAAGCCTTCGTCTCGA (SEQ ID NO:296) | CGGGATCCTTAGGATGCGTATGTAACCA (SEQ ID NO:297) |
| ADH9 | Atu4087 | GGAATTCATATGAAAGCGATTGTCCGCCA (SEQ ID NO:298) | CGGGATCCTTAGGAAAAGGCGATCTGCA (SEQ ID NO:299) |
| ADH10 | Atu4289 | GGAATTCATATGCCGATGGCGCTCGGGCA (SEQ ID NO:300) | CGGGATCCTTAGAATTCGATGACTTGCC (SEQ ID NO:301) |
| ADH11 | Atu3027 | GGAATTCATATGAAACATTCTCAGGACAA (SEQ ID NO:302) | GGGCGCCGATCATGTGGTTCGTTTCCG (SEQ ID NO:303) |
| | | CGGAAACGCACCACATGATCGGCGCCC (SEQ ID NO:304) | CGGGATCCTTATGCCATACGTTCCATAT (SEQ ID NO:305) |
| ADH12 | Atu3026 | GGAATTCATATGCAGCGTTTTACCAACAG (SEQ ID NO:306) | CGGGATCCTTAGGAAAACAGGACGCCGC (SEQ ID NO:307) |

Expression and Purification of ADH 1-10.

[0425] All plasmids were transformed into *Escherichia coli* strain BL21(DE3). The single colonies of BL21(DE3) containing respective alcohol dehydrogenase (ADH) genes were inoculated into 50 ml of LB media containing 50 µg/ml kanamycin (Km⁵⁰). These strains were grown in an orbital shaker with 200 rpm at 37° C. The 0.2 mM IPTG was added to each culture when the OD_{600 nm} reached 0.6, and the induced culture was grown in an orbital shaker with 200 rpm at 20° C. 24 hours after the induction, the cells were harvested by centrifugation at 4,000 rpm×g for 10 min and the pellet was resuspended into 2 ml of Bugbuster (Novagen) containing 10 µl of Lysonase™ Bioprocessing Reagent (Novagen). The solution was again centrifuged at 4,000 rpm×g for 10 min and the supernatant was obtained.

Preparation of ~2% DEHU Solution by Enzymatic Degradation.

[0426] DEHU solution was enzymatically prepared. A 2% alginate solution was prepared by adding 10 g of low viscosity alginate into the 500 ml of 20 mM Tris-HCl (pH7.5) solution. An approximately 10 mg of alginate lyase derived from *Flavobacterium* sp. (purchased from Sigma-aldrich) was added to the alginate solution. 250 ml of this solution was then transferred to another bottle and the *E. coli* cell lysate containing Atu3025 prepared above section was added. The alginate degradation was carried out at room temperature over night. The resulting products were analyzed by thin layer chromatography, and DEHU formation was confirmed.

Preparation of D-Mannuronate Solution by Chemical Degradation.

[0427] D-mannuronate solution was chemically prepared based on the protocol previously described by Spoehr (Archive of Biochemistry, 14: pp 153-155). Fifty milligram of alginate was dissolved into 800 µL of ninety percent formate. This solution was incubated at 100° C. for over night. Formate was then evaporated and the residual substances were washed with absolute ethanol twice. The residual substance

was again dissolved into absolute ethanol and filtrated. Ethanol was evaporated and residual substances were resuspended into 20 mL of 20 mM Tris-HCl (pH 8.0) and the solution was filtrated to make a D-mannuronate solution. This D-mannuronate solution was diluted 5-fold and used for assay.

Assay for DEHU Hydrogenase.

[0428] To identify DEHU hydrogenase, a NADPH dependent DEHU hydrogenation assay was performed. 20 µl of prepared cell lysate containing each ADH was added to 160 µl of 20-fold deluted DEHU solution prepared in the above section. 20 µl of 2.5 mg/ml of NADPH solution (20 mM Tris-HCl, pH 8.0) was added to initiate the hydrogenation reaction, as a preliminary study using cell lysate of *A. tumefaciens* C58 have shown that DEHU hydrogenation requires NADPH as a co-factor. The consumption of NADPH was monitored an absorbance at 340 nm for 30 min using the kinetic mode of ThermoMAX 96 well plate reader (Molecular Devises). *E. coli* cell lysate containing alcohol dehydrogenase (ADH) 10 lacking a portion of N-terminal domain was used in a control reaction mixture.

Assay for D-Mannuronate Hydrogenase.

[0429] To identify D-mannuronate hydrogenase, a NADPH dependent D-mannuronate hydrogenation assay was performed. 20 µl of prepared cell lysate containing each ADH was added to 160 µl of D-mannuronate solution prepared in the above section. 20 µl of 2.5 mg/ml of NADPH solution (20 mM Tris-HCl, pH 8.0) was added to initiate the hydrogenation reaction. The consumption of NADPH was monitored an absorbance at 340 nm for 30 min using the kinetic mode of ThermoMAX 96 well plate reader (Molecular Devises). *E. coli* cell lysate containing alcohol dehydrogenase (ADH) 10 lacking a portion of N-terminal domain was used in a control reaction mixture.

Construction of pETkdgK.

[0430] DNA sequence of kdgK of *Escherichia coli* encoding 2-keto-deoxy gluconate kinase was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction

mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-AGGTACGGTCAAATAA AGGAGG ATATACATATGTCCAAAAAGATTGCCGT-3') (SEQ ID NO:157) and reverse (5'-TTTTCCTTTT GCGGCCGCCCGCTGGCATCGCCTCAC-3') (SEQ ID NO:158) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Escherichia coli* DH10B genome in 50 μl. Amplified DNA fragment was digested with NdeI and NotI and ligated into pET29 pre-digested with the same restriction enzymes.

Construction of pETkd2A.

[0431] DNA sequence of *kdgA* *Escherichia coli* encoding 2-keto-deoxy gluconate-6-phosphate aldolase was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-GGCGATGCCAGCGTAA AGGAGG ATATA CATATGAAAACTGGAAAACAAG-3') (SEQ ID NO:159) and reverse (5'-TTTTCCTTTT GCGGCCGCCCGCTTAGCGCCTTCTA-3') (SEQ ID NO:160) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Escherichia coli* DH10B genome in 50 μl. Amplified DNA fragment was digested with NdeI and NotI and ligated into pET29 pre-digested with the same restriction enzymes.

Protein Expression and Purification.

[0432] All plasmids (pETAtu3025, pETADH11, pETADH12, pETkdgA, pETkdgK, and pETuxuA) were transformed into *Escherichia coli* strain BL21(DE3). The single colonies of BL21(DE3) containing respective plasmids were inoculated into 50 ml of LB media containing 50 μg/ml kanamycin (KM⁵⁰). These strains were grown in an orbital shaker with 200 rpm at 37° C. The 0.2 mM IPTG was added to each culture when the OD_{600 nm} reached 0.6, and the induced culture was grown in an orbital shaker with 200 rpm at 20° C. 24 hours after the induction, the cells were harvested by centrifugation at 4,000 rpm×g for 10 min and the pellet was resuspended into 2 ml of Bugbuster (Novagen) containing 10 μl of Lysozyme™ Bioprocessing Reagent (Novagen) and suggested amount of protease inhibitor cocktail (SIGMA). The solution was again centrifuged at 4,000 rpm×g for 10 min and the supernatant was obtained. The supernatant was applied to Nickel-NTA spin column (Qiagen) to purify His-tagged proteins.

[0433] The results of the assays for DEHU hydrogenase activity and D-mannuronate hydrogenase activity of ADH1-10 are shown in FIGS. 7A and 7B. These results demonstrate that the novel enzymes ADH1 and ADH2 showed significant DEHU hydrogenase activity (FIG. 7A), and that the novel enzymes ADH3, ADH4, and ADH9 showed significant mannuronate hydrogenase activity (FIG. 7B).

In Vitro Pyruvate Formation.

[0434] The reaction mixture contained 1% alginate or ~0.5% mannuronate, ~5 μg of purified Atu3026 (ADH12) or Atu3027 (ADH11), and ~5 μg of purified oligoalginate lyase (Atu3025), UxuA, KdgK, and KdgA, 2 mM of ATP, and 0.6 mM of NADPH in 20 mM Tris-HCl pH7.0. The reaction was carried out over night and the pyruvate formation was monitored by the pyruvate assay kit (BioVision, Inc).

[0435] The results of in vitro pyruvate formation from alginate mediated by enzymatic and chemical degradation are shown in FIG. 6B and FIG. 6C, respectively. As can be seen in these figures, alginate was converted to pyruvate via the isolated enzymes. These results also show that each of Atu3026 (ADH12) and Atu3027 (ADH11) are capable of catalyzing both DEHU hydrogenase and mannuronate hydrogenase reactions.

Example 4

Construction and Biological Activity of Biosynthesis Pathways

Construction of Pathways:

[0436] A propionaldehyde biosynthetic pathway comprising a threonine deaminase (*ilvA*) gene from *Escherichia coli* and keto-isovalerate decarboxylase (*kivd*) from *Lactococcus lactis* is constructed and tested for the ability to convert L-threonine to propionaldehyde.

[0437] A butyraldehyde biosynthetic pathway comprising a thiolase (*atoB*) gene from *E. coli*, β-hydroxy butyryl-CoA dehydrogenase (*hbd*), crotonase (*crt*), butyryl-CoA dehydrogenase (*bcd*), electron transfer flavoprotein A (*etfA*), and electron transfer flavoprotein B (*etfB*) genes from *Clostridium acetobutyricum* ATCC 824, and a coenzyme A-linked butyraldehyde dehydrogenase (*ald*) gene from *Clostridium beijerinckii acetobutyricum* ATCC 824 was constructed in *E. coli* and tested for the ability to produce butyraldehyde. Also, a coenzyme A-linked alcohol dehydrogenase (*adhE2*) gene from *Clostridium acetobutyricum* ATCC 824 was used as an alternative to *ald* and tested for the ability to produce butanol.

[0438] An isobutyraldehyde biosynthetic pathway comprising an acetolactate synthase (*alsS*) from *Bacillus subtilis* or (*als*) from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 (codon usage was optimized for *E. coli* protein expression) and acetolactate reductoisomerase (*ilvC*) and 2,3-dihydroxyisovalerate dehydratase (*ilvD*), genes from *E. coli* and keto-isovalerate decarboxylase (*kivd*) from *Lactococcus lactis* was constructed and tested for the ability to produce isobutyraldehyde, as measured by isobutanal production.

[0439] 3-methylbutyraldehyde and 2-methylbutyraldehyde biosynthesis pathways comprising an acetolactate synthase (*alsS*) from *Bacillus subtilis* or (*als*) from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 (codon usage was optimized for *E. coli* protein expression), acetolactate reductoisomerase (*ilvC*), 2,3-dihydroxyisovalerate dehydratase (*ilvD*), isopropylmalate synthase (*LeuA*), isopropylmalate isomerase (*LeuC* and *LeuD*), and 3-isopropylmalate dehydrogenase (*LeuB*) genes from *E. coli* and keto-isovalerate decarboxylase (*kivd*) from *Lactococcus lactis* were constructed and tested for the ability to produce 3-isovaleraldehyde and 2-isovaleraldehyde.

[0440] Phenylacetaldehyde and 4-hydroxyphenylacetaldehyde biosynthesis pathways comprising a transketolase (*tktA*), a 3-deoxy-7-phosphoheptulonate synthase (*aroF*, *aroG*, and *aroH*), 3-dehydroquininate synthase (*aroB*), a 3-dehydroquininate dehydratase (*aroD*), a dehydroshikimate reductase (*aroE*), a shikimate kinase II (*aroL*), a shikimate kinase I (*aroK*), a 5-enolpyruvylshikimate-3-phosphate synthetase (*aroA*), a chorismate synthase (*aroC*), a fused chorismate mutase P/prephenate dehydratase (*pheA*), and a fused chorismate mutase T/prephenate dehydrogenase (*tyrA*) genes from

E. coli, keto-isovalerate decarboxylase (kivd) from *Lactococcus lactis* were constructed and tested for the ability to produce phenylacetaldehyde and/or 4-hydroxyphenylacetaldehyde.

[0441] A 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol, and 2-(indole-3-)-ethanol biosynthesis pathway comprising a transketolase (tktA), a 3-deoxy-7-phosphoheptulonate synthase (aroF, aroG, and aroH), 3-dehydroquinase synthase (aroB), a 3-dehydroquinase dehydratase (aroD), a dehydroshikimate reductase (aroE), a shikimate kinase II (aroL), a shikimate kinase I (aroK), a 5-enolpyruvylshikimate-3-phosphate synthetase (aroA), a chorismate synthase (aroC), a fused chorismate mutase P/prephenate dehydratase (pheA), and a fused chorismate mutase T/prephenate dehydrogenase (tyrA) genes from *E. coli*, keto-isovalerate decarboxylase (kivd) from *Lactococcus lactis*, alcohol dehydrogenase (adh2) from *Saccharomyces cerevisiae*, Indole-3-pyruvate decarboxylase (ipdc) from *Azospirillum brasilense*, phenylethanol reductase (par) from *Rhodococcus* sp. ST-10, and benzaldehyde lyase (bal) from *Pseudomonas fluorescens* was constructed and tested for the ability to produce 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol and/or 2-(indole-3)ethanol.

Construction of pBADButP.

[0442] The DNA sequence encoding hbd, crt, bcd, etfA, and etfB of *Clostridium acetobutyricum* ATCC 824 was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 3 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CCC GAGCTCTTAGGAGGATTAGTCATGGAAC-3') (SEQ ID NO:161) and reverse (5'-GCTCTAGA TTATTTTGAATAATCGTAGAAACC-3') (SEQ ID NO:162) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Clostridium acetobutyricum* ATCC 824 genome (ATCC) in 50 μl. Amplified DNA fragment was digested with BamHI and XbaI and ligated into pBAD33 pre-digested with the same restriction enzymes.

Construction of pBADButP-atoB.

[0443] The DNA sequence encoding atoB of *Escherichia coli* DH10B was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-GC TCTAGAGGAGGATATATATATGAAAAATTGTGTCATC GTC-3') (SEQ ID NO:163) and reverse (5'-AA CTGCAGTTAATTCAACCGTTCAATCACC-3') (SEQ ID NO:164) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Escherichia coli* DH10B genome in 50 μl. Amplified DNA fragment was digested with XbaI and PstI and ligated into pBADButP pre-digested with the same restriction enzymes.

Construction of pBADatoB-ald.

[0444] The DNA sequence encoding atoB of *Escherichia coli* DH10B and ald from *Clostridium beijerinckii* were amplified separately by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CGAGCTC AGGAGGATATATATGAAAAATTGTGTCATCGTCAGTG-3') (SEQ ID NO:165) for atoB and 5'-GGTTGAATTAAGGAGGATATATATAT-GAATAAAGACACTAATACCTAC-3' for ald (SEQ ID NO:166) and reverse (5'-GTCTTTATTCATATATATATC-

CTCCTTAATCAACCGTTCAATCACCATC-3' (SEQ ID NO:146) for atoB and 5'-CCCAAGCTTAGCCGGCAAG-TACACATCTTC-3' for ald (SEQ ID NO:167) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Escherichia coli* DH10B and *Clostridium beijerinckii* genome (ATCC) in 50 μl, respectively. The amplified DNA fragments were gel purified and eluted into 30 ul of EB buffer (Qiagen). 5 ul from each DNA solution was combined and each DNA fragment was spliced by another round of PCR: 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 2 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CGAGCTC AGGAGGATATATATGAAAAATTGTGTCATCGTCAGTG-3') (SEQ ID NO:168) and reverse (5'-CCCAAGCTTAGCCGGCAAGTACACATCTTC-3') (SEQ ID NO:169) primers, 1 U Phusion High Fidelity DNA polymerase (NEB). The spliced fragment was digested with SacI and HindIII and ligated into pBADButP pre-digested with the same restriction enzymes.

Construction of pBADButP-atoB-ALD.

[0445] The DNA fragment 1 encoding chloramphenicol acetyltransferase (CAT), P15 origin of replication, araBAD promoter, atoB of *Escherichia coli* DH10B and ald of *Clostridium beijerinckii* and the DNA fragment 2 encoding araBAD promoter, hbd, crt, bcd, etfA, and etfB of *Clostridium acetobutyricum* ATCC 824 were amplified separately by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 4 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-AAGGAAAAA GCGGCCGCCCTGAACCGACGACCGGGTTCG-3') (SEQ ID NO:170) for fragment 1 and 5'-CGG GGTACCCTTTTCATACTCCCGCCATTCAG-3' (SEQ ID NO:274) for fragment 2, and reverse (5'-CGG GGTACC GCGGATACATATTTGAATGTATTTAG-3') (SEQ ID NO:171) for fragment 1 and (5'-AAGGAAAAA GCGGCCGCGCGGATACATATTTGAATGTATTTAG-3') (SEQ ID NO:172) for fragment 2) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng pBADatoB-ald and pBADButP in 50 μl, respectively. Amplified DNA fragments were digested with NotI and KpnI and ligated each other.

Construction of pBADilvCD.

[0446] The DNA fragments encoding ilvC and ilvD of *Escherichia coli* DH10B were amplified separately by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-GCTCTAGAGGAGGATATATATAT GGCTAACTACTTCAATACAC-3') (SEQ ID NO:173) for ilvC and 5'-TGCTGTTCGGGGTTAAGGAG-GATATATATATGCCTAAGTACCGTTCCGCC-3' for ilvD) (SEQ ID NO:174) and reverse (5'-AACGGTACTTAG-CATATATATATATCCTTCCTTAACCCGCAA-CAGCAATACG-3') (SEQ ID NO:175) for ilvC and 5'-AC ATGCAITGCTTAACCCCGAGTTTCGATT-3') (SEQ ID NO:176) for ilvD) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Escherichia coli* DH10B genome (ATCC) in 50 μl. The amplified DNA fragments were gel purified and eluted into 30 ul of EB buffer (Qiagen). 5 ul from each DNA solution was combined and each DNA fragment was spliced by another round of PCR: 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 2 min, repeated 30 times. The

reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-GCTCTAGAGGAGGATATATA TATGGCTAACTACTTCAATACAC-3') (SEQ ID NO:177) and reverse (5'-AC ATGCATGCTTAAACCCCCAGTTTTGATT-3') (SEQ ID NO:178) primers, 1 U Phusion High Fidelity DNA polymerase (NEB). The spliced fragment was digested with XbaI and SphI and ligated into pBAD33 pre-digested with the same restriction enzymes.

Construction of pBADals-*ilvCD*.

[0447] The DNA fragment encoding als of *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 of its codon usage optimized for over-expression in *E. coli* was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CCCGAGCTCAGGAGGATATATATAT GGATAAACAGTATCCGGT-3') (SEQ ID NO:179) and reverse (5'-GC TCTAGATTACAGAATTTGACTCAGGT-3') (SEQ ID NO:180) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng pETals in 50 μl. The amplified DNA fragment was digested with SacI and XbaI and ligated into pBAD*ilvCD* pre-digested with the same restriction enzymes. Construction of pBADalsS-*ilvCD*.

[0448] The DNA fragments encoding front and bottom halves of alsS of *Bacillus subtilis* B26 were amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 0.5 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CCCGAGCTCAGGAGGATATATATATG TTGACAAAAGCAACAAAAG-3') (SEQ ID NO:181) for front and 5'-CGGTACCCTTCCAGAGATTTAGAG-3' (SEQ ID NO:275) for back halves, and reverse (5'-CTCTAAATCTCTGGAAAGGGTACCG-3') (SEQ ID NO:182) for front and (5'-GC TCTAGATTAGAGAGCTTTTCGTTTTTCATG-3' for back halves) (SEQ ID NO:183) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Bacillus subtilis* B26 genome (ATCC) in 50 μl. The amplified DNA fragments were gel purified and eluted into 30 ul of EB buffer (Qiagen). 5 ul from each DNA solution was combined and each DNA fragment was spliced by another round of PCR: 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CCCGAGCTCAGGAGGAT ATATATATGTTGACAAAAGCAACAAAAG-3') (SEQ ID NO:184) and reverse (5'-GC TCTAGATTAGAGAGCTTTTCGTTTTTCATG-3') (SEQ ID NO:185) primers, 1 U Phusion High Fidelity DNA polymerase (NEB). The spliced fragment was internal XbaI site free and thus was digested with SacI and XbaI and ligated into pBAD*ilvCD* pre-digested with the same restriction enzymes. Construction of pBADLeuABCD.

[0449] The DNA fragment encoding leuA, leuB, leuC, and leuD of *Escherichia coli* BL21(DE3) was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 3 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CGAGCTCAGGAGGATATATATAG CCGCAAGTCATTATTTTCG-3') (SEQ ID NO:186) and reverse (5'-AAAA CTGCAGCGTTTGATGACGTGGACGATAGCGG-3') (SEQ ID NO:187) primers, 1 U Phusion High Fidelity DNA

polymerase (NEB), and 50 ng *Escherichia coli* BL21(DE3) genome in 50 μl. The amplified DNA fragment was digested with SacI and XbaI and ligated into pBAD33 pre-digested with the same restriction enzymes.

Construction of pBADLeuABCD2.

[0450] The DNA fragment 1 encoding leuA and leuB and the DNA fragment 2 encoding leuC and leuD of *Escherichia coli* BL21 (DE3) were amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CG AGCTCAGGAGGATATATATGAGCCAGCAAGTCAT TATTTTCG-3') (SEQ ID NO:188) for fragment 1 and (5'-AGGGGTGTAAGGAGGATATATATATG-GCTAAGACGTTATACGAAAAATTG-3') (SEQ ID NO:189) for fragment 2 and reverse (5'-CGTCTTAGC-CATATATATATCCTCCTTACACCCCT-TCTGCTACATAGCGG-3') (SEQ ID NO:190) for fragment 1 and (5'-AAAA CTGCAGCGTTTGATGACGTGGACGATAGCGG-3') (SEQ ID NO:191) for fragment 2 primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Escherichia coli* BL21(DE3) genome in 50 μl, respectively. The amplified DNA fragments were gel purified and eluted into 30 ul of EB buffer (Qiagen). 5 ul from each DNA solution was combined and each DNA fragment was spliced by another round of PCR: 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 3 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CG AGCTCAGGAGGATATATATATGAGCCAGCAAGTCAT TATTTTCG-3') (SEQ ID NO:192) and reverse (5'-AAAA CTGCAGCGTTTGATGACGTGGACGATAGCGG-3') (SEQ ID NO:193) primers, 1 U Phusion High Fidelity DNA polymerase (NEB). The spliced fragment was digested with SacI and XbaI and ligated into pBAD33 pre-digested with the same restriction enzymes.

Construction of pBADLeuABCD4.

[0451] The DNA fragments encoding leuA, leuB, leuC and leuD of *Escherichia coli* BL21(DE3) were amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CGAGCTCAGGAGGATATATATAG CCGCAAGTCATTATTTTCG-3') (SEQ ID NO:194) for leuA, (5'-GAAACCGTGTGAGGAGGATATATATAT-GTCAAGAATTACCATATTGCCG-3') (SEQ ID NO:195) for leuB, (5'-AGGGGTGTAAGGAGGATATATATATG-GCTAAGACGTTATACGAAAAATTG-3') (SEQ ID NO:196) for leuC, and (5'-ACATTAATAAGGAG-GATATATATGCGAGAGAAATTTATCAAACACAC-3') (SEQ ID NO:197) for leuD and reverse (5'-ATTCTTCGA-CATATATATATCCTCCTCACACGGTTTC-CTTGTTGTTTTTCG-3') (SEQ ID NO:198) for leuA, (5'-CGTCTTAGCCATATATATATCCTCCTTACACCCCTTCT GCTACATAGCGG-3') (SEQ ID NO:199) for leuB, (5'-TTTCTGCCATATATATATCTCTCT-TATTAAATGTTGCGAATGTCCGCG-3') (SEQ ID NO:200) for leuC, and (5'-AAAAGTGCAGCGTTTGAT-GACGTGGACGATAGCGG-3') (SEQ ID NO:201) for leuD primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Escherichia coli* BL21(DE3) genome in 50 μl, respectively. The amplified DNA fragments were gel purified and eluted into 30 ul of EB buffer (Qiagen). 5 ul from each DNA solution was combined and each DNA fragment was spliced by another round of PCR: 98° C. for 10 sec, 60° C. for 15 sec, and

72° C. for 3 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CGAGCTCAGGAGGATATATATATGAGCCA GCAAGTCATTATTTTCG-3') (SEQ ID NO:202) and reverse (5'-AAAA CTGCAGCGTTTGATGACGTGGACGATAGCGG-3') (SEQ ID NO:203) primers, 1 U Phusion High Fidelity DNA polymerase (NEB). The spliced fragment was digested with SacI and XbaI and ligated into pBAD33 pre-digested with the same restriction enzymes.

Construction of pBADals-ilvCD-leuABCD, pBADals-ilvCD-leuABCD2 pBADals-ilvCD-leuABCD4 pBADalsS-ilvCD-leuABCD, pBADalsS-ilvCD-leuABCD2 pBADalsS-ilvCD-leuABCD4.

[0452] The DNA fragments 1 (for als) and 2 (for alsS) encoding chloramphenicol acetyltransferase (CAT), P15 origin of replication, araBAD promoter, als of *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 of its codon usage optimized for over-expression in *E. coli* or alsS of *Bacillus subtilis* B26 and ilvC and ilvD of *E. coli* DH 10B were amplified separately by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 4 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-AAGGAAAAA GCGGCCGCCCTGAACCGACGACCGGGTTCG-3') (SEQ ID NO:204) and reverse (5'-CGG GGTACCGCGGATACATATTTGAATGTATTTAG-3') (SEQ ID NO:205) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng pBADals-ilvCD and pBADalsS-ilvCD in 50 μl, respectively.

[0453] To remove an internal SphI restriction enzyme site from leuC, overlap PCR was carried out. The front and bottom halves of DNA fragment 3 (for leuABCD), fragment 4 (for leuABCD2), and fragment 5 (for leuABCD4) encoding araBAD promoter, leuA, leuB, leuC, and leuD of *E. coli* BL21(DE3) were amplified separately by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 4 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-AAGGAAAAA GCGGCCGCACTTTTCATACTCCCGCCATTTCAG-3') (SEQ ID NO:206) for front and (5'-CAAAGGCCGTCTG-CACGCGCCGAAAGGCAAA-3') (SEQ ID NO:207) for back halves) and reverse (5'-TTTGCCTTCGGCGCGTG-CAGACGGCCTTTG-3') (SEQ ID NO:208) for front and (5'-AC ATGCATGCCGTTTGATGACGTGGACGATAGCGG-3') (SEQ ID NO:209) for bottom halves, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng pBADleuABCD, pBADleuABCD2, and pBADleuABCD4 in 50 μl, respectively. The amplified DNA fragments were gel purified and eluted into 30 ul of EB buffer (Qiagen). 5 ul from each DNA solution was combined and each DNA fragment was spliced by another round of PCR: 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 4 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-AAGGAAAAA GCGGCCGCACTTTTCATACTCCCGCCATTTCAG-3') (SEQ ID NO:210) and reverse (5'-AC ATGCATGCCGTTTGATGACGTGGACGATAGCGG-3') (SEQ ID NO:211) primers, 1 U Phusion High Fidelity DNA polymerase (NEB). The resulting fragment 3, 4, and 5 were

digested with SphI and NotI and ligated into both fragment 1 and 2 pre-digested with the same restriction enzymes.

Construction of pBADaroG-tktA-aroBDE.

[0454] The DNA fragments encoding aroG, tktA, aroB, aroD, and aroE of *Escherichia coli* BL21(DE3) were amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CCC GAGCTCAGGAGGATATAT ATGAATTATCAGAAC-GACGATTTAC-3') (SEQ ID NO:212) for aroG, (5'-GCGTCGCGGGTAAGGAGGAAAATTTTAT-GTCCTCACGTAAAGAGCTTGCC-3') (SEQ ID NO:213) for tktA, (5'-GAACTGCTGTAAGGAGGTTAAAATTATG-GAGAGGATTGCTGTTACTCTCG-3') (SEQ ID NO:214) for aroB, (5'-CAATCAGCGTAAGGAGGTATATATAAT-GAAAACCGTAACTGTAAAAGATC-3') (SEQ ID NO:215) for aroD, and (5'-TACACCAGGCATAAGGAG-GAATTAATTATGGAAACCTATGCTGTTTTTGG-3') (SEQ ID NO:216) for aroE and reverse (5'-TACGTGAGGA-CATAAAATTTTCTCCTTACCCGC-GACGCGCTTTTACTGC-3') (SEQ ID NO:217) for aroG, (5'-CAATCCTCTCCATAATTTTAACTCCT-TACAGCAGTTCTTTTGCTTTCGC-3') (SEQ ID NO:218) for tktA, (5'-CAATCAGCGTAAGGAGGTATATATAAT-GAAAACCGTAACTGTAAAAGATC-3') (SEQ ID NO:219) for aroB, (5'-TACGGTTTTTATTATATACCTC-CTTACCGTGATTGACAATCGGCAATG-3') (SEQ ID NO:220) for aroD, and (5'-AC ATGCATGCTTACGCGGACAATTCCTCCTGCAA-3') (SEQ ID NO:221) for aroE, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Escherichia coli* BL21(DE3) genome in 50 μl, respectively. The amplified DNA fragments were gel purified and eluted into 30 ul of EB buffer (Qiagen). 5 ul from each DNA solution was combined and each DNA fragment was spliced by another round of PCR: 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 3 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CCCGAGCTCAG GAGGATATATATATGAATTATCAGAACGACGATTTAC-3') (SEQ ID NO:222) and reverse (5'-AC ATGCATGCTTACGCGGACAATTCCTCCTGCAA-3') (SEQ ID NO:223) primers, 1 U Phusion High Fidelity DNA polymerase (NEB). The spliced fragment was digested with SacI and SphI and ligated into pBAD33 pre-digested with the same restriction enzymes.

Construction of pBADpheA-aroLAC.

[0455] The DNA fragments encoding pheA, aroL, aroA, and aroC of *Escherichia coli* DH10 were amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward (5'-CCCGAGCTCAGGAGGATATATATG ACATCGGAAAACCCGTTACTGG-3') (SEQ ID NO:224) for pheA, (5'-GATCCAACCTAAGGAGGAAAATTTTAT-GACACAACCTCTTTTTCTGATCG-3') (SEQ ID NO:225) for aroL, (5'-GATCAATTGTTAAGGAGG-TATATATAATGGAATCCCTGACGTTACAACCC-3') (SEQ ID NO:226) for aroA, and (5'-CAGGCAGCCTAAG-GAGGAATTAATTATGGCTGGAACA-CAATTGGACAAC-3') (SEQ ID NO:227) for aroC and reverse (5'-AGGTTGTGTCATAAAATTTTCTCCT-TAGGTTGGATCAACAGGCACTACG-3') (SEQ ID NO:228) for pheA, (5'-CAGGGATTCCATTATATATAC-

CTCCTTAACAATTGATCGTCTGTGCCAGG-3') (SEQ ID NO:229) for aroL, (5'-GTTTCCAGCCATAATTAATTCCTCCTTAGGCTGCCTGGCTAATCCGCGCC-3') (SEQ ID NO:230) for aroA, and (5'-ACATGCAATGCTTACCAGCGTGGAATATCAGTCTTC-3') (SEQ ID NO:231) for aroC primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Escherichia coli* BL21(DE3) genome in 50 μ l, respectively. The amplified DNA fragments were gel purified and eluted into 30 μ l of EB buffer (Qiagen). 5 μ l from each DNA solution was combined and each DNA fragment was spliced by another round of PCR: 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 4 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-CCC GAGCTCAGGAGGATATATATGACATCGGAAAACC CGTACTGG-3') (SEQ ID NO:232) and reverse (5'-ACATGCAATGCTTACCAGCGTGGAATATCAGTCTTC-3') (SEQ ID NO:233) primers, 1 U Phusion High Fidelity DNA polymerase (NEB). The spliced fragment was digested with SacI and SphI and ligated into pBAD33 pre-digested with the same restriction enzymes.

Construction of pBADtyrA-aroLAC.

[0456] The DNA fragments encoding pheA, aroL, aroA, and aroC of *Escherichia coli* DH10 were amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-CCCAGCTCAGGAGGATATATATGGTTGCTGAATTGACCGCATTAC-3') (SEQ ID NO:234) for tyrA, (5'-AATCGCCAGTAAAGGAGGAAAATTTTATGACACAACCTCTTTTTCTGATCG-3') (SEQ ID NO:235) for aroL, (5'-GATCAATTGTTAAGGAGG-TATATATAATGGAATCCCTGACGTTACAACCC-3') (SEQ ID NO:236) for aroA, and (5'-CAGGCAGCCTAAGGAGGAATTAATATGCTGGAAACA-CAATTGGACAAC-3') (SEQ ID NO:237) for aroC, and reverse (5'-GAGGTTGTGTCATAAAATTTTCTCCTTACTGGCGATTGTCATTCGCTG-3') (SEQ ID NO:238) for tyrA, (5'-CAGGGATTCCATTATATATACCTCTTAACAATTGATCGTCTGTGCCAGG-3') (SEQ ID NO:239) for aroL, (5'-GTTTCCAGCCATAATTAATTCCTCCTTAGGCTGCCTGGCTAATCCGCGCC-3') (SEQ ID NO:240) for aroA, and (5'-ACATGCAATGCTTACCAGCGTGGAATATCAGTCTTC-3') (SEQ ID NO:241) for aroC, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Escherichia coli* BL21(DE3) genome in 50 μ l, respectively. The amplified DNA fragments were gel purified and eluted into 30 μ l of EB buffer (Qiagen). 5 μ l from each DNA solution was combined and each DNA fragment was spliced by another round of PCR: 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 4 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-CCCAGCTCAGGAGGATATATATGGTTGCTGAATTGACCGCATTAC-3') (SEQ ID NO:242) and reverse (5'-ACATGCAATGCTTACCAGCGTGGAATATCAGTCTTC-3') (SEQ ID NO:243) primers, 1 U Phusion High Fidelity DNA polymerase (NEB). The spliced fragment was digested with SacI and SphI and ligated into pBAD33 pre-digested with the same restriction enzymes.

Construction of pBADpheA-aroLAC-aroG-*tk*tA-aroBDE and pBADtyrA-aroLAC-aroG-*tk*tA-aroBDE.

[0457] A DNA fragment 1 (for pheA) and 2 (for tyrA) encoding chloramphenicol acetyltransferase (CAT), P15 ori-

gin of replication, araBAD promoter, pheA or tyrA, aroL, aroA, aroC of *Escherichia coli* DH10B and a DNA fragment 3 encoding araBAD promoter, aroG, *tk*tA, aroB, aroD, and aroE of *Escherichia coli* DH10B were amplified separately by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 4 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-AAGGAAAAA GCGGCCGCCCCTGAACCGACGACCGGGTCG-3') (SEQ ID NO:244) for fragment 1 and 2 and (5'-GCTCTAGAACTTTTCATACTCCCGCCATTCAG-3') (SEQ ID NO:245) for fragment 3, and reverse (5'-GCTCTAGAGCGGATACATATTTGAATGTATTTAG-3') (SEQ ID NO:246) for fragment 1 and 2 and (5'-AAGGAAAA GCGGCCGCGCGGATACATATTTGAATGTATTTAG-3') (SEQ ID NO:247) for fragment 3, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng pBADpheA-aroLAC, pBADtyrA-aroLAC, and pBADaroG-*tk*tA-aroBDE in 50 μ l, respectively. Amplified DNA fragments 1 and 2 were digested with NotI and XbaI and ligated into fragment 3 pre-digested with the same restriction enzymes.

Construction of pTrcBAL.

[0458] A DNA sequence encoding benzaldehyde lyase (bal) of *Pseudomonas fluorescens* of its codon usage optimized for over-expression in *E. coli* was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-CATGCCATGGCTATGATTACTGGTGG-3') (SEQ ID NO:248) and reverse (5'-CCCCGAGCTCTTACGCGCCGGATTGGAAATACA-3') (SEQ ID NO:249) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng pETBAL in 50 μ l. Amplified DNA fragment was digested with NcoI and SacI and ligated into pTrc99A pre-digested with the same restriction enzymes.

Construction of pTrcAdhE2.

[0459] A DNA sequence encoding Co-A linked alcohol/aldehyde dehydrogenase (adhE2) of *Clostridium acetobutyricum* ATCC824 was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-CATGCCATGGCCAAAGTTACAAATCAAAAAG-3') (SEQ ID NO:250) and reverse (5'-CAGCTCTTAAAATGATTTTATATAGATATCC-3') (SEQ ID NO:251) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Clostridium acetobutyricum* ATCC824 genome in 50 μ l. Amplified DNA fragment was digested with NcoI and SacI and ligated into pTrc99A pre-digested with the same restriction enzymes.

Construction of pTrcAdh2.

[0460] A DNA sequence encoding alcohol dehydrogenase (adh2) of *Saccharomyces cerevisiae* was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-CATGCCATGGGTATTCCAGAACTCAAAAAG-3') (SEQ ID NO:252) and reverse (5'-CCC GAGCTCTTATTTAGAAGTGTCAACAACG-3') (SEQ ID NO:253) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng genome of *Saccharomyces cerevi-*

siae in 50 μ l. Amplified DNA fragment was digested with NcoI and SacI and ligated into pTrc99A pre-digested with the same restriction enzymes.

Construction of pTrcBALD.

[0461] A DNA sequence encoding CoA-linked aldehyde dehydrogenase (ald) of *Clostridium beijerinckii* was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-CCCCGAGCTCAGGAGGATATACATATGAATAAAGACACACTAATACC-3') (SEQ ID NO:254) and reverse (5'-CCC AAGCTTAGCCGGCAAGTACACATCTTC-3') (SEQ ID NO:255) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng pETBAL in 50 μ l. Amplified DNA fragment was digested with SacI and HndIII and ligated into pTrcBAL pre-digested with the same restriction enzymes.

Construction of pTrcBALK.

[0462] A DNA sequence encoding ketoisovalerate decarboxylase (kivd) of *Lactococcus lactis* was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-CCCGAGCTCAGGAGGATATATATGTATACAGTAGGAGATTACC-3') (SEQ ID NO:256) and reverse (5'-GCTCTAGATTATGATTTATTTGTTTCAGCAAAT-3') (SEQ ID NO:257) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng pETBAL in 50 μ l. Amplified DNA fragment was digested with SacI and XbaI and ligated into pTrcBAL pre-digested with the same restriction enzymes.

Construction of pTrcAdh-Kivd.

[0463] A DNA sequence encoding ketoisovalerate decarboxylase (kivd) of *Lactococcus lactis* was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-CCCGAGCTCAGGAGGATATATATGTATACAGTAGGAGATTACC-3') (SEQ ID NO:258) and reverse (5'-GCTCTAGATTATGATTTATTTGTTTCAGCAAAT-3') (SEQ ID NO:259) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng pETBAL in 50 μ l. Amplified DNA fragment was digested with SacI and XbaI and ligated into pTrcAdh2 pre-digested with the same restriction enzymes.

Construction of pTrcBAL-DDH-2ADH.

[0464] To remove internal NcoI site, overlap PCR was carried out. DNA fragments encoding front and bottom halves of meso-2,3-butanediol dehydrogenase (ddh) of *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 and secondary alcohol dehydrogenase (2adh) of *Pseudomonas fluorescens* were amplified separately by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-C GAGCTCAGGAGGATATATATATGAAAAAGTCGCAC TTGTTACCG-3') (SEQ ID NO:260) for front half of ddh, (5'-GGCCGGCGCCGCGCGATGGCGGTGAAAGTG-3') (SEQ ID NO:261) for bottom half of ddh, (5'-AAC-TAATCTAGAGGAGGATATATATATGAG-CATGACGTTTTCCGGCCAGG-3') (SEQ ID NO:262) for front half of 2adh, and (5'-CCTTGCGGAGGGCTCGATG-GATGAGTTTCGAC-3') (SEQ ID NO:263) for bottom half of 2adh, and reverse (5'-CACTTTCACCGCCATCGCGCGGC-

CGCCGGCC-3') (SEQ ID NO:264) for front half of ddh, (5'-GCTCATATATATATCCTCCTCTAGATT-AGTTAAACACCATCCCGCCGTCG-3') (SEQ ID NO:265) for bottom half of ddh, (5'-GTCGAACTCATCCATCGAGC-CCTCCGCAAGG-3') (SEQ ID NO:266) for front half of 2adh, and (5'-CCC AAGCTTAGATCGCGGTGGCCCCGCGTCG-3') (SEQ ID NO:267) for bottom half of 2adh, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 for ddh and *Pseudomonas fluorescens* genome for 2adh in 50 μ l, respectively. The amplified DNA fragments were gel purified and eluted into 30 μ l of EB buffer (Qiagen). 5 μ l from each DNA solution was combined and each DNA fragment was spliced by another round of PCR: 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 2 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-CGAGCTCAGGAGGATATATATATGAAAA AAGTCGCACTTGTTACCG-3') (SEQ ID NO:268) and reverse (5'-CCC AAGCTTAGATCGCGGTGGCCCCGCGTCG-3') (SEQ ID NO:269) primers, 1 U Phusion High Fidelity DNA polymerase (NEB). The spliced fragment was digested with SacI and HindIII and ligated into pTrcBAL pre-digested with the same restriction enzymes.

Construction of pBBRPduCDEGH.

[0465] A DNA sequence encoding propanediol dehydratase medium (pduD) and small (pdue) subunits and propanediol dehydratase reactivation large (pduG) and small (pduH) subunits of *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 2 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-GC TCTAGAGGAGGATTTAAAATGGAAATTAACGAAA CGCTGC-3') (SEQ ID NO:270) and reverse (5'-TCC CCGCGGTTAAGCATGGCGATCCCCGAAATGGAATCC CTTTGAC-3') (SEQ ID NO:271) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 in 50 μ l. Amplified DNA fragment was digested with SacII and XbaI and ligated into pTrc99A pre-digested with the same restriction enzymes to form pBBRPduDEGH.

[0466] A DNA sequence encoding propanediol dehydratase large subunit (pduC) of *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction mixture contained 1 \times Phusion buffer (NEB), 2 mM dNTP, 0.5 μ M forward (5'-CCGCTCGAGGAGGATATATATATGAGATCGA AAAGATTTGAAGC-3') (SEQ ID NO:272) and reverse (5'-GCTCTAGATTAGCCAAGTTCATGGGATCG-3') (SEQ ID NO:273) primers, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 in 50 μ l. Amplified DNA fragment was digested with XhoI and XbaI and ligated into pBBRPduDEGH pre-digested with the same restriction enzymes.

Construction of pTrcIpdC-Par.

[0467] A DNA sequence encoding indole-3-pyruvate (ipdc) of *Azospirillum brasilense* and phenylethanol reductase (par) of *Rhodococcus* sp. ST-10 were amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 1 min, repeated 30 times. The reaction

mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 μM forward primers (5'-CATG CCAATGGGACTGGCTGAGGCACTGCTGC-3' (SEQ ID NO:314) for ipdc and 5'-CGAGCTCAGGAGGATATAT ATATGAAAGCTATCCAGTACACCCGTAT-3' (SEQ ID NO:315) for par, and reverse primers (5'-CGAGCTCTTAT-TCGCGCGGTGCCGCGTGCAGG-3' (SEQ ID NO:316) for ipdc and 5'-GC TCTAGATTACAGGCCCGGAACCACAACGGCGC-3' (SEQ ID NO:317) for par, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng pTrcIpdc and pTrcPar, respectively, in 50 μl. Amplified DNA fragment of ipdc and par were digested with NcoI/SacI and SacI/XbaI, respectively, and were ligated into pTrc99A pre-digested with NcoI and XbaI.

Testing and Results:

[0468] To test the butyraldehyde biosynthesis pathway, DH10B harboring pBADButP-atoB/pTrcBALD and pBAD-ButP-atoB-ALD/pTrcB2DH/pBBRpduCDEGH were grown overnight in LB media containing 50 ug/ml chloramphenicol (Cm⁵⁰) and 100 ug/ml ampicillin (Amp¹⁰⁰) at 37 C, 200 rpm. An aliquot of each seed culture was inoculated into fresh TB media containing Cm⁵⁰ and Amp¹⁰⁰ and was grown in incubation shaker at 37 C, 200 rpm. Three hours after inoculation, the cultures were induced with 13.3 mM arabinose and 1 mM IPTG and were grown for overnight. 700 ul of this culture was extracted with equal volume of ethylacetate and analyzed by GC-MS.

[0469] To test the isobutyraldehyde biosynthesis pathway, DH10B cells harboring pBADals-ilvCD/pTrcBALK or pBADalsS-ilvCD/pTrcBALK were grown overnight in LB media containing 50 ug/ml chloramphenicol (Cm⁵⁰) and 100 ug/ml ampicillin (Amp¹⁰⁰) at 37 C, 200 rpm. An aliquot of each seed culture was inoculated into fresh TB media containing Cm⁵⁰ and Amp¹⁰⁰ and was grown in incubation shaker at 37 C, 200 rpm. Three hours after inoculation, the cultures were induced with 13.3 mM arabinose and 1 mM IPTG and were grown for overnight. 700 ul of this culture was extracted with equal volume of ethylacetate and analyzed by GC-MS for the production of isobutyraldehyde. FIG. 8B shows the production of isobutanol from these cultures.

[0470] To test the 3-methylbutyraldehyde and 2-methylbutyraldehyde biosynthesis pathways, DH10B harboring pBADals-ilvCD-LeuABCD/pTrcBALK, pBADals-ilvCD-LeuABCD2/pTrcBALK, pBADals-ilvCD-LeuABCD/pTrcBALK4, pBADalsS-LeuABCD/pTrcBALK, pBADalsS-LeuABCD2/pTrcBALK, or pBADalsS-LeuABCD4/pTrcBALK were grown overnight in LB media containing 50 ug/ml chloramphenicol (Cm⁵⁰) and 100 ug/ml ampicillin (Amp¹⁰⁰) at 37 C, 200 rpm. An aliquot of each seed culture was inoculated into fresh TB media containing Cm⁵⁰ and Amp¹⁰⁰ and was grown in incubation shaker at 37 C, 200 rpm. Three hours after inoculation, the cultures were induced with 13.3 mM arabinose and 1 mM IPTG and were grown for overnight. 700 ul of this culture was extracted with equal volume of ethylacetate and analyzed by GC-MS. The production of 2-isovaleralcohol (2-methylpentanol) and 3-isovaleralcohol (3-methylpentanal) was monitored because 3-isovaleraldehyde and 2-isovaleraldehyde are spontaneously converted to their corresponding alcohols. FIG. 8B shows the production of 2-methylpentanol and 3-methylpentanal from these cultures.

[0471] To test the phenylacetaldehyde and 4-hydroxyphenylacetaldehyde biosynthesis pathways, DH10B cells har-

boring pBADpheA-aroLAC/pTrcBALK, pBADtyrA-aroLAC/pTrcBALK, pBADaroG-tktA-aroBDE/pTrcBALK, pBADpheA-aroLAC-aroG-tktA-aroBDE/pTrcBALK, and pBADpheA-aroLAC-aroG-tktA-aroBDE/pTrcBALK were grown overnight in LB media containing 50 ug/ml chloramphenicol (Cm⁵⁰) and 100 ug/ml ampicillin (Amp¹⁰⁰) at 37 C, 200 rpm. An aliquot of each seed culture was inoculated into fresh TB media containing Cm⁵⁰ and Amp¹⁰⁰ and was grown in incubation shaker at 37 C, 200 rpm. Three hours after inoculation, the cultures were induced with 13.3 mM arabinose and 1 mM IPTG and were grown for overnight. 700 ul of this culture was extracted with equal volume of ethylacetate and analyzed by GC-MS. The production of phenylacetaldehyde, 4-hydroxyphenylaldehyde and their corresponding alcohols were monitored using GC-MS. FIG. 9B shows the production of 4-hydroxyphenylethanol from these cultures.

[0472] To test the 2-phenylethanol, 2-(4-hydroxyphenyl) ethanol, and 2-(indole-3) ethanol biosynthesis pathways, DH10B harboring pBADpheA-aroLAC-aroG-tktA-aroBDE/pTrcBALK, pBADpheA-aroLAC-aroG-tktA-aroBDE/pTrcBALK, pBADpheA-aroLAC-aroG-tktA-aroBDE/pTrcAdh2-Kivd, pBADpheA-aroLAC-aroG-tktA-aroBDE/pTrcAdh2-Kivd, pBADpheA-aroLAC-aroG-tktA-aroBDE/pTrcIpdc-Par, and pBADpheA-aroLAC-aroG-tktA-aroBDE/pTrcIpdc-Par were grown overnight in LB media containing 50 ug/ml chloramphenicol (Cm⁵⁰) and 100 ug/ml ampicillin (Amp¹⁰⁰) at 37 C, 200 rpm. An aliquot of each seed culture was inoculated into fresh TB media containing Cm⁵⁰ and Amp¹⁰⁰ and was grown in incubation shaker at 37 C, 200 rpm. Three hours after inoculation, the cultures were induced with 13.3 mM arabinose and 1 mM IPTG and were grown for overnight to a week. 700 ul of this culture was extracted with equal volume of ethylacetate and analyzed by GC-MS. The results are detailed below.

[0473] The production of 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol and/or 2-(indole-3)ethanol was monitored using GC-MS. FIG. 42A shows the production of 2-phenylethanol from these cultures at 24 hours. FIG. 42B shows the production of 2-(4-hydroxyphenyl)ethanol from these cultures at 24 hours. FIG. 42C shows the production of 2-(indole-3)ethanol from these cultures at 24 hours.

[0474] FIG. 43A shows the GC-MS chromatogram for control (pBAD33 and pTrc99A) at one week. FIG. 43B shows the GC-MS chromatogram for 2-phenylethanol (5.97 min) production from pBADpheA-aroLAC-aroG-tktA-aroBDE and pTrcBALK at one week. FIG. 44 shows the GC-MS chromatogram for 2-(4-hydroxyphenyl)ethanol (9.36 min) and 2-(indole-3) ethanol (10.32 min) production from pBADtyrA-aroLAC-aroG-tktA-aroBDE and pTrcBALK at one week.

Example 5

Isolation and Biological Activity of Diol Dehydrogenases

[0475] Available substrates such as 3-hydroxy-2-butanone (acetoin), 4-hydroxy-3-hexanone (propioin), 5-hydroxy-4-octanone (butyroin), 6-hydroxy-5-decanone (valeroin), and 1,2-cyclopentanediol were used to measure the ability of diol dehydrogenases (ddh) to catalyze the reduction of large saturated α-hydroxyketones to produce a diol. All reagents were purchased from Sigma-Aldrich Co. and TCI America, unless otherwise stated.

[0476] For cloning and isolation of DDH polypeptides, genomic DNA from several species of bacteria were obtained from ATCC (*Lactobacillus brevis* ATCC 367, *Pseudomonas putida* KT2440, and *Klebsiella pneumoniae* MGH78578), PCR-amplified (using Phusion with polymerase with 1× Phusion buffer, 0.2 mM dNTP, 0.5 μL Phusion enzyme, 1.5 μM primers, and 20 pg template DNA in a 50 μL reaction) utilizing the following protocol: 30 cycles, 98° C./10 secs (denaturing), 60° C./15 secs (annealing), 72° C./30 secs (elongation). Polymerase chain reaction products were then digested using restriction enzymes NdeI and BamHI, then ligated into NdeI/BamHI digested pET28 vectors. Vectors containing ddh clones were transformed into BL21 (DE3) competent cells for protein expression. Single colony was inoculated into LB media, and expression of 6×His-tagged proteins of interest was induced at OD₆₀₀=0.6 with 0.1 mM IPTG. Expression was allowed to proceed for 15 hours at 22° C. The 6×His-tagged enzymes were purified using Ni-NTA spin columns following suggested protocols by QIAGEN, yielding purified protein concentrations in the range of 1.1-6.5 mg/mL (determined by Bradford assay).

[0477] Diol dehydrogenase ddh 1 was isolated from *Lactobacillus brevis* ATCC 367, diol dehydrogenase ddh2 was isolated from *Pseudomonas putida* KT2440, and diol dehydrogenase ddh3 was isolated from *Klebsiella pneumoniae* MGH78578. The nucleotide sequence encoding and polypeptide sequence of ddh 1 are shown in SEQ ID NOS:97 and 98, respectively; nucleotide sequence encoding and polypeptide sequence of ddh2 are shown in SEQ ID NOS:99 and 100, respectively; and nucleotide sequence encoding and polypeptide sequence of ddh3 are shown in SEQ ID NOS:101 and 102, respectively.

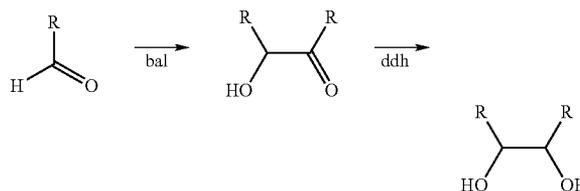
[0478] Reactions to measure biological activity of DDH polypeptides were performed in a final volume of 200 μL as follows: 25 mM substrate, 0.04 mg/mL DDH polypeptide, 0.25 mg/mL nicotinamide cofactor, 200 mM imidazole, 14 mM Tris-HCl, and 1.5% by volume DMSO. Biological activity was assayed using a Molecular Devices Thermomax 96 well plate reader, monitoring absorbance at 340 nm, which corresponds to NADH or NADPH concentration. For the kinetic studies, 0.04 mg/mL DDH polypeptide, 0.25 mg/mL NADH, 20 mM Tris HCl Buffer pH 6.5(red) or 9.0(ox), T=25 C, 100 uL total volume was used.

[0479] FIG. 12A shows the biological activity of ddh1, ddh2, and ddh3 using butyoin as a substrate (triangles represent ddh3 activity). FIG. 12B shows the oxidation activity of ddh3 towards 1,2-cyclopentenediol and 1,2-cyclohexanediol as measured by NADH production. FIG. 13 summarizes the results of kinetic studies for various substrates in the oxidation reactions catalyzed by the DDH polypeptides. These reactions were NAD⁺ dependent.

Example 6

Sequential In Vivo Biological Activity of CC-Ligases (Lyases) and Diol Dehydrogenases

[0480] The ability of a C—C lyase and a diol hydrogenase to perform the following sequential reaction was tested in *E. coli*:



[0481] For α-hydroxyketone and diol production, a pathway comprising a benzaldehyde lyase (bal) gene isolated from *Pseudomonas fluorescens* (codon usage was optimized for *E. coli* protein expression) and meso-2,3-butanediol dehydrogenase (ddh) gene isolated from *Klebsiella pneumoniae* subsp. *pneumoniae* MGH 78578 was constructed in *E. coli* and tested for its ability to condensate the substrates detailed below in Table 2 (e.g., acetaldehyde, propionaldehyde, butyraldehyde, isobutyraldehyde, 2-methyl-butylaldehyde, 3-methyl-butylaldehyde, phenylacetaldehyde, and 4-hydroxyphenylacetaldehyde, or their corresponding alcohols) to form α-hydroxyketone and the corresponding diol in vivo. The production of various α-hydroxyketones and diols was monitored by gas chromatography-mass spectrometry (GC-MS).

TABLE 2

| Summary of substrates and products. | | | |
|-------------------------------------|-----------------------------------|-----------------------------|---------|
| Substrate | Produced α-hydroxyketone | Produced diol | Figures |
| Butanal | 5-Hydroxy-4-octanone | 4,5-Octanediol | 17A & B |
| n-Pentanal | 6-Hydroxy-5-decanone | 5,6-Decanediol | 18A & B |
| 3-Methylbutanal | 2,7-Dimethyl-5-hydroxy-4-octanone | 2,7-Dimethyl-4,5-octanediol | 19A & B |
| n-Hexanal | 7-Hydroxy-6-dodecanone | 6,7-dodecanediol | 20A & B |
| 4-Methylpentanal | 2,9-Dimethyl-6-hydroxy-5-decanone | 2,9-Dimethyl-5,6-decanediol | 21A & B |
| n-Octanal | 9-Hydroxy-8-hexadecanone | 8,9-hexadecanediol | 22 |
| Acetaldehyde | 3-Hydroxy-2-butanone | 2,3-Butanediol | 23 |
| n-Propanal | 4-Hydroxy-3-hexanone | 3,4-Hexanediol | 24A & B |
| Phenylacetaldehyde | 1,4-Diphenyl-3-hydroxy-2-butanone | 1,4-Diphenyl-2,3-butanediol | 25 |

For Analysis of \cong C10.

[0482] *E. coli* harboring pTrcBAL-DDH-2ADH was grown for overnight in LB media containing 50 ug/ml Kanamycine (Km). This seed culture was inoculated into M9 media containing 3% (v/v) glycerol, 0.5% (g/v) and 50 ug/ml Km. 10 mL cultures were grown to O.D.₆₀₀=0.7, then cultures were induced with 0.5 mM IPTG. The cells were allowed to express the enzymes of interest for 3 hours before various aldehydes were added to a concentration of 5-10 mM. After addition of aldehydes, the cultures were capped and incubated at 37° C. with shaking for 72 hours. Cultures were extracted with 2 mL ethyl acetate, and analyzed on GC-MS using the following protocol:

- [0483] 1 μ L injection w/50:1 split
- [0484] Inlet temperature—150° C.
- [0485] Initial oven temperature—50° C.
- [0486] Temperature Ramp 1—10° C./min to 150° C.
- [0487] Temperature Ramp 2—50° C./min to 300° C.
- [0488] GC to MS transfer temp—250° C.
- [0489] MS detection—full scan MW 35-200

For Analysis of \cong C12.

[0490] *E. coli* DH10B strains harboring pTrc99A (Ctrl vector) or pTrcBAL were inoculated into 0.75xM9/0.5% LB containing 0.1 mM CaCl₂, 2 mM MgSO₄, 1 mM KCl, 1% galacturonate, 5 μ g/mL thiamine, Amp. The cultures were grown up to an optical density (600 nm) of 0.8 and induced with 0.25 mM IPTG. The cells were allowed to express the proteins for 2.5 hours at 37° C., then aldehyde substrate was added to a concentration of 5 mM, the culture vial was capped tightly and incubated for 72 hours at 37° C. w/shaking 200 rpm. 1 mL of the final culture was extracted with 0.75 mL of ethyl acetate, centrifuged facilitate phase separation, then analyzed via GCMS using the following method.

- [0491] 1 μ L injection w/50:1 split
- [0492] Inlet temperature—250° C.
- [0493] Initial oven temperature—50° C.
- [0494] Temperature Ramp 1—10° C./min to 125° C.
- [0495] Temperature Ramp 2—30° C./min to 300° C.
- [0496] Final Temperature 300° C.—1 minute
- [0497] GC to MS transfer temp—250° C.
- [0498] MS detection—full scan MW 40-260.
- [0499] The results are depicted in FIGS. 17 through 25. FIG. 17 shows the sequential conversion of butanal into 5-hydroxy-4-octanone and then 4,5-octanediol. FIG. 18 shows the sequential conversion of n-pentanal into 6-hydroxy-5-decanone and then 5,6-decanediol. FIG. 19 shows the conversion of 3-methylbutanal into 2,7-dimethyl-5-hydroxy-4-octanone and then 2,7-Dimethyl-4,5-octanediol. FIG. 20 shows the sequential conversion of n-hexanal into 7-hydroxy-6-dodecanone and then 6,7-dodecanediol. FIG. 21 shows the conversion of 4-methylpentanal into 2,9-dimethyl-6-hydroxy-5-decanone and then 2,9-dimethyl-5,6-decanediol. FIG. 22 shows the conversion of n-octanal into 9-hydroxy-8-hexadecanone. FIG. 23 shows the conversion of acetaldehyde into 3-hydroxy-2-butanone. FIG. 24 shows the sequential conversion of n-propanal into 4-hydroxy-3-hexanone and then 3,4-hexanediol. FIG. 25 shows the conversion of phenylacetaldehyde into 1,4-diphenyl-3-hydroxy-2-butanone.
- [0500] Similar to above, a pathway comprising a benzaldehyde lyase (bal) gene isolated from *Pseudomonas fluorescens* (codon usage was optimized for *E. coli* protein expression) was constructed in *E. coli* and tested for its ability to catalyze

the production of various α -hydroxyketones. The results, which show the broad spectrum of C—C ligase activity for the bal gene tested, are set forth in FIG. 48 through FIG. 55.

Example 7

Sequential Biological Activity of Diol Dehydrogenases and Diol Dehydratases

[0501] To test the sequential biological activity of diol dehydrogenases and diol dehydratases in a dehydration and reduction pathway, butyrolin was used as a substrate in a sequential reaction to produce 4-octanone. The enzyme diol dehydrogenase (e.g., ddh) catalyzes the reversible reduction and oxidation of α -hydroxy ketones and its corresponding diol, such as 5-hydroxy-4-octanone and 4,5-octanediol, and the enzyme diol dehydratase (e.g., pduCDE) catalyzes the irreversible dehydration of diols, such as 4,5-octanediol.

[0502] Diol dehydrogenase ddh from *Kiebsiella pneumoniae* MGH 78578 and diol dehydratase pduCDE from *Kiebsiella pneumoniae* MGH 78578 were cloned into a bacterial expression vector and expressed and purified on a Ni-NTA column, as described in Example X except that 1 mM of 1,2-propanediol was added at all time during the expression and purification of diol dehydratase. The large, medium, and small subunits of the pduCDE polypeptide are encoded by the nucleotide sequences of SEQ ID NOs: 103, 105, and 107, respectively, and the polypeptide sequence are set forth in SEQ ID NOs: 104, 106, and 108, respectively.

[0503] The ddh3 and pduCDE polypeptides were incubated with butyrolin and their appropriate cofactors, then assayed using gas chromatography-mass spectrometry (GC-MS) for their ability to perform sequential reactions resulting in the product 4-octanone. Reaction conditions are given in Table 3 below. The reaction mixture was incubated at 37° C. for 40 hours in a 0.6 mL eppendorf tube with minimal head space. The reaction product was extracted with an equivalent volume of ethyl acetate, stored in a glass vial, and sent to Thermo Fischer Scientific Instruments Division for compositional analysis by GC-MS.

TABLE 3

| Reaction Conditions | |
|---|-----------------------|
| Rxn Component | Concentration |
| 5-hydroxy-4-octanone (butyrolin) | 8.4 mM |
| Adenosylcobalamin (coenzyme B ₁₂) | 33.5 μ M |
| KCl | 9.6 mM |
| NADH | 18 mM |
| ddh3 enzyme | 0.19 mg/mL |
| dDOH1 enzyme mix | 0.15 mg/mL |
| Reaction Buffer | 10 mM Tris HCl pH 7.0 |

[0504] FIG. 26A shows GC-MS data which confirms the presence of 4,5-octanediol in the sample extraction. The mass-spectra of the peaks, retention time, at 5.36 was identified as butyrolin (substrate), and at 6.01, 6.09, and 6.12 min were identified as different isomers of 4,5-octanediol. This compound is the expected product resulting from the reduction of butyrolin by ddh3.

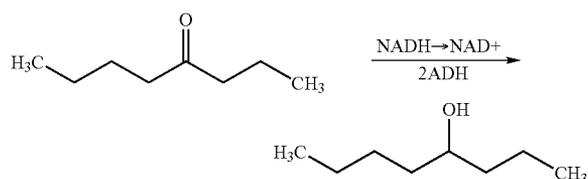
[0505] FIG. 26B shows GC-MS data confirming the presence of 4-octanone in the sample extraction. The mass-spectra of the peak, retention time, at 4.55 was identified as 4-octanone. This compound is the expected product resulting

from the sequential dehydrogenation of butyoin and dehydrogenation of 4,5-octanediol by ddh3 and pduCDE, respectively. [0506] FIGS. 27A and 27B show comparisons between the sample extraction gas chromatograph/mass spectrum and the 4-octanone standard gas chromatograph/mass spectrum. These results demonstrate that 4-octanone was produced from butyoin using the enzymes diol dehydrogenase (ddh3) and a diol dehydratase (pduCDE). GC-MS analysis of the incubated reaction mixture confirmed starting material, intermediate and product, demonstrating that these enzymes can be reapropriated for these specific substrates.

Example 8

Isolation and Biological Activity of Secondary Alcohol Dehydrogenases

[0507] Substrates such as 4-octanone, 2,7-dimethyl-4-octanone, cyclopentanone and corresponding alcohols were utilized to measure the ability of secondary alcohol dehydrogenases (2ADHs) to catalyze the reduction of large saturated ketones to secondary alcohols. An example of a reaction catalyzed by secondary alcohol dehydrogenases is illustrated below (reduction of 4-octanone to 4-octanol is shown):



[0508] All enzymes and reagents were purchased from New England Biolabs and Sigma, respectively, unless otherwise stated.

[0509] Various secondary alcohol dehydrogenases (2ADHs) were isolated from *Pseudomonas putida* KT2440, *Pseudomonas fluorescens* Pf-5, and *Klebsiella pneumoniae* MGH 78578. All vectors were transformed in BL21(DE3) competent cells and expression of the genes encoding the proteins of interest was induced with IPTG (via the T7 promoter). The cells were lysed, proteins were extracted and then purified on Ni-NTA columns. Final protein concentration in the Ni-NTA eluate was diluted to 0.15 mg/mL prior to assays.

[0510] NADPH/NADP consumption and production assays were performed using a THERMOMax microplate reader in the kinetic mode, monitoring the NADPH absorbance peak at 340 nm until the reaction reached equilibrium. In the assay described in Table 2, 2ADH-2, 2ADH-5, 2ADH-8, and 2ADH-10 were tested for their ability to either catalyze the oxidation of 4-octanol or catalyze the reduction of 4-octanone. These reaction conditions are found in Table 4 below.

TABLE 4

| Reaction Conditions for Various Enzyme Assays | |
|---|---------------------|
| Reaction Component | Final Concentration |
| NADH Production Assay (30° C.) | |
| 2ADH enzyme | Approx. 0.058 µg/µL |
| 4-octanol | 5.55 mM |
| NAD ⁺ | Approx. 1.4 µg/µL |
| Imidazole (from Elution Buffer) | Approx. 280 mM |

TABLE 4-continued

| Reaction Conditions for Various Enzyme Assays | |
|---|---------------------|
| Reaction Component | Final Concentration |
| NADH Consumption Assay (30° C.) | |
| 2ADH enzyme | Approx. 0.075 µg/µL |
| 4-octanone | 5.0 mM |
| NADH | Approx. 0.25 µg/µL |
| Imidazole (from Elution Buffer) | Approx. 250 mM |
| NADPH Production Assay (30° C.) | |
| 2ADH enzyme | Approx. 0.058 µg/µL |
| 4-octanol | 5.55 mM |
| NADP ⁺ | Approx. 1.4 µg/µL |
| Imidazole (from Elution Buffer) | Approx. 280 mM |

[0511] Further testing was performed, as described in Tables 5 below, in which 2ADH-2, 2ADH-11, 2ADH-12, 2ADH-13, 2ADH-14, 2ADH-15, 2ADH-16, 2ADH-17, and 2ADH-18 were tested for their ability to either catalyze the oxidation of 4-octanol, 2,7-dimethyl-4-octanol, or cyclopentanone, or catalyze the reduction of 4-octanone, 2,7-dimethyl-4-octanone, or cyclopentanone.

TABLE 5

| Rxn Component | Final Concentration |
|--|---------------------|
| Rxn Components for NADPH Consumption Assays (Reduction) | |
| Substrate | 25 mM |
| Enzyme | 0.04 mg/mL |
| Nicotinamide cofactor | 0.25 mg/mL |
| Imidazole | 200 mM |
| Tris HCl | 14 mM |
| DMSO | 1.5% by volume |
| Total Volume | 200 µL |
| Rxn Components for NAD(P)H Production Assays (Oxidation) | |
| Substrate | 5 mM |
| Enzyme | 0.04 mg/mL |
| Nicotinamide cofactor | 0.25 mg/mL |
| Imidazole | 200 mM |
| Tris HCl | 14 mM |
| Rxn Components for NAD(P)H Production Assay using 2,7-dimethyl-4-octanone as a substrate | |
| Substrate | 50 mM |
| Enzyme | 0.08 mg/mL |
| Nicotinamide cofactor | 0.25 mg/mL |
| Imidazole | 200 mM |
| Tris HCl | 14 mM |
| DMSO | 3% by volume |

[0512] FIG. 30A shows the results from the NADH Production Assay of Table 3, in which 2ADH-2 catalyzes the oxidation of 4-octanol in the presence of NAD⁺, as measured by NADH production. FIG. 30B shows the results of the NADPH Production Assay of Table 3, in which 2ADH-5, 2ADH-8, and 2ADH-10 catalyze the oxidation of 4-octanol in the presence of NADP⁺, as measured by NADPH production.

[0513] FIG. 31 shows the oxidation of 4-octanol by 2ADH-11 (FIG. 31A) and 2ADH-16 (FIG. 31B), as measured by NADH and NADPH production, respectively. FIG. 32 shows the oxidation of 2,7-dimethyloctanol by 2ADH-11 and others (FIG. 32A) and 2ADH-16 (FIG. 32B), as measured by NADH and NADPH production, respectively.

[0514] FIG. 33A shows the reduction of 2,7-dimethyl octanol by 2ADH 11 and 2ADH16 as monitored by NADPH consumption. FIG. 33B shows the reduction activity of both 2ADH11 and 2ADH16 towards various substrates. FIG. 34 shows the oxidation (FIG. 34A) and reduction (FIG. 34B) of cyclopentanol by 2ADH-16.

[0515] Similar to above, kinetic testing for both oxidation and reduction reactions was performed on various substrates using 2ADH-16. The conditions for these studies were as follows: 0.04 mg/mL enzyme, 0.25 mg/mL cofactor, 20 mM Tris HCl Buffer pH 6.5(red) or 9.0(ox), T=25 C, 100 uL total volume was used. The calculated rate constants for the reduction reactions, along with the structures of the substrates, are summarized in FIG. 35. The calculated rate constants for the oxidation reactions, along with the structures of the substrates, are summarized in FIG. 36. These results show that 2ADH-16 is capable of catalyzing both the oxidation and reduction of a wide variety of substrates.

Example 9

Isolation and In Vitro and In Vivo Activity of Coenzyme B 12 Independent Diol Dehydratases

[0516] Substrates such as 1,2-propanediol, meso-2,3-butanediol, and trans-1,2-cyclopentane diol were utilized to test both the in vitro and in vivo biological activity of a B12 independent diol dehydratase in a dehydration and reduction pathway. Diol dehydratases catalyze the irreversible dehydration of diols, such as 1,2-propanediol.

[0517] For in vitro activity, *E. coli* BL21(DE3) harboring pETPduCDE (diol dehydratase subunits) was inoculated into 100 mL LB media, grown to OD₆₀₀=0.7, induced with 0.15 mM IPTG, and incubated for 22 hours at 22° C. The cells were lysed and proteins of interest were purified on a Ni-NTA spin column. Purification of all three dehydratase subunits was accomplished by adding 5 mM 1,2-propanediol to the lysis and wash buffers. The Ni-NTA purification yielded approximately 660 µL of protein mixture at a concentration of 2.2 mg/mL. Protein concentration assays were conducted using a Bradford reagent protocol.

[0518] The purified PduCDE was used to set up in vitro diol dehydratase reactions. Three assays were conducted with 1,2-propanediol and meso-2,3-butanediol. Control reactions were also set up with elution buffer added in place of purified PduCDE. In vitro reactions were conducted under semi-anaerobic conditions in 2 mL screw cap glass vials. Reaction components and concentrations are given in Table 6.

TABLE 6

| Reaction conditions for B ₁₂ dependent DDOH in vitro assay | |
|---|-----------------------|
| Rxn Component | Concentration |
| Diol substrate | 10 mM |
| Adenosylcobalamin (B ₁₂) | 100 µg/mL |
| KCl | 10 mM |
| dOH1 enzyme mix | 0.08 mg/mL |
| Reaction Buffer | 10 mM Tris HCl pH 7.5 |

[0519] After 48 hours, 1 mL of the reaction mixture was extracted with 0.5 mL of either ethylacetate or hexanol and analyzed by GCMS.

[0520] The following GCMS protocol was used for all experiments:

[0521] 1 µL injection w/50:1 split

[0522] Inlet temperature—250° C.

[0523] Initial oven temperature—50° C.

[0524] Temperature Ramp 1—10° C./min to 125° C.

[0525] Temperature Ramp 2—30° C./min to 300° C.

[0526] Final Temperature 300° C.—1 minute

[0527] GC to MS transfer temp—250° C.

[0528] MS detection—full scan MW 40-260

[0529] The results are shown in FIG. 45. FIG. 45A confirms the formation of 1-propanal from 1,2-propanediol, and FIG. 45B confirms the formation of 2-butanone from meso-2,3-butanediol, both of which were catalyzed by B12 independent diol dehydratase.

[0530] For in vivo activity, the pBBRDhaB1/2 plasmid was constructed as follows: the DNA sequence encoding B12-independent glycerol dehydratase (dhaB1) and activator (dhaB2) of *Clostridium butyricum* was amplified by polymerase chain reaction (PCR): 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 2 min for dhaB1 and 1 min for dhaB2, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 µM forward primers (5'-CCGCTCGAGGAGGATATATATGATTTCTAAAGGCTTTAGCACCC-3' (SEQ ID NO:318) for dhaB1 and 5'-ACGTGATGTAATCTAGAGGAGGATATATATGAGCAAAGAAATTAAGG-3' (SEQ ID NO:319) for dhaB2, and reverse primers (5'-TCTTGTCTCATATATATCCTCCTCTAGATTACATCACGTGTTTCAGTAC-3' (SEQ ID NO:320) for dhaB1 and 5'-C GAGCTCTTATTCGGCGCCAATGGTGCACGGG-3' (SEQ ID NO:321) for dhaB2, 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng pETdhaB1 and pETdhaB2, respectively, in 50 µL. Amplified fragments were gel purified and spliced by another round of PCR: 98° C. for 10 sec, 60° C. for 15 sec, and 72° C. for 2.5 min, repeated 30 times. The reaction mixture contained 1× Phusion buffer (NEB), 2 mM dNTP, 0.5 µM forward (5'-CCGCTCGAGGAGGATATATATGATTTCTAAAGGCTTTAGCACCC-3' (SEQ ID NO:322) and reverse primers (5'-C GAGCTCTTATTCGGCGCCAATGGTGCACGGG-3' (SEQ ID NO:323), 1 U Phusion High Fidelity DNA polymerase (NEB), and 50 ng each fragment in 50 µL. Amplified DNA fragment was digested with XhoI and SacI and ligated into pBBR1MCS-2 pre-digested with the same restriction enzymes.

[0531] Two strains of *E. coli* DH10B harboring pBBR1MCS-2 or pBBRDhaB1/2 into TB media without glycerol were inoculated. Cultures were grown to OD₆₀₀=0.5 and the substrates 1,2-propanediol, meso-2,3-butanediol, and trans-1,2-cyclopentane diol were added to separate cultures to a concentration of 10 mM. 5 µg/ml of coenzyme S-adenosylmethionine was added before the culture is transferred to anaerobic environment. The cultures were incubated at 37 C for 48 hrs.

[0532] After 48 hours, 1 mL of culture was extracted with 0.5 mL of ethylacetate or hexanol and analyzed by GCMS, as described above. The results are shown in FIG. 46. FIG. 46A shows the in vivo production of 1-propanol from 1,2-propanediol. FIG. 46B shows the in vivo production of 2-butanone from meso-2,3 butanediol. FIG. 46C shows the in vivo production of cyclopentanone from trans-1,2-cyclopentane diol.

Example 10

Identification of Secreted Alginate Lyase and Genomic Regions Sufficient for Growth on Alginate as a Sole Source of Carbon

[0533] To identify secreted or external alginate lyases, and to identify genomic regions from *Vibrio splendidus* that are sufficient to confer growth in alginate as a sole source of carbon, the following clones were made using the gateway

system from Invitrogen (Carlsbad, Calif.). First, entry vectors were made by TOPO cloning PCR fragments into pENTR/D/TOPO. PCR fragments were generated using *Vibrio splendidus* B01 genomic DNA as a template and amplified with the following primer pairs:

[0534] Vs24214-24249: genomic region corresponding to gene id between V12B01_24214 and V12B01_24249 (see Example 1).

TABLE 7

| | | |
|---------|-----------------------------|-----------------|
| 24214 F | cacc caagcgatagtttatatagcgt | (SEQ ID NO:324) |
| 24249R | gaaatgaacggatattacgt | (SEQ ID NO:325) |

[0535] Vs24189-24209: genomic region corresponding to gene id between V12B01_24189 and V12B01_24209 (see Example 1).

TABLE 8

| | | |
|---------|----------------------------|-----------------|
| 24189 R | cggaacaggtgattgtggt | (SEQ ID NO:326) |
| 24209 F | cacc gccacttcaagatgaagctgt | (SEQ ID NO:327) |

[0536] Vs24214-24239: genomic region corresponding to gene id between V12B01_24214 and V12B01_24239 (see Example 1).

TABLE 9

| | | |
|-----------|-----------------------------|-----------------|
| 24214 F | cacc caagcgatagtttatatagcgt | (SEQ ID NO:328) |
| 24239 R_1 | gtggctaagtacatgccggt | (SEQ ID NO:329) |

[0537] The entry vectors were recombined with the destination vector pET-DEST42 (Invitrogen) using the LR recombinase enzyme (Invitrogen). These destination vectors were then put into electrocompetent DH10B or BL21 cells.

[0538] The alginate lyase clones were then made by digesting (using enzymes Nde I and Bam HI) the PCR products that were generated using *Vibrio splendidus* 12B01 genomic DNA as a template and amplified with the following primer pairs:

TABLE 10

| | |
|------------------------------------|-----------------|
| GGAATTC CAT | (SEQ ID NO:330) |
| atgacaaagaatatgacgactaaac | |
| for | |
| 24214 ndeF | |
| forward primer for V12B01_24214 | |
| CG GGATCC ttattatttccctgcccctgcagt | (SEQ ID NO:331) |
| for reverse primer | |
| 24214 bamR | |
| for V12B01_24214 | |
| 24219 ndeF | |
| GGAATTC CAT atgagctatcaaccacttttac | (SEQ ID NO:332) |
| for forward | |
| primer for V12B01_24219 | |

TABLE 10-continued

| | |
|---------------------------------|-----------------|
| CG GGATCC ttacagttgagcaaatgatcc | (SEQ ID NO:333) |
| for reverse primer | |
| 24219 bamR | |
| for V12B01_24219 | |

[0539] The digested PCR products were then ligated into cut pET28 vector. Certain of the cloned genomic regions of *Vibrio splendidus* B01 were tested for the presence of secreted alginate lyases, and the above-described constructs were tested in various combinations for the ability to confer growth on alginate as a sole source of carbon.

[0540] The Vs24254 (SEQ ID NO: 32) region of *Vibrio splendidus* encodes a functional external alginate lyase. BL21 cells expressing Vs24254 from the pET28 vector were capable of breaking down alginate in the growth medium. When grown on LB+2% alginate+0.1 mM Isopropyl β -D-1-thiogalactopyranoside (IPTG), only cells expressing the Vs24254 gene give a positive TBA assay result of pink color. This assay was performed by spinning down an overnight culture grown on the above mentioned media. The media was then mixed in a 1:1 ratio with 0.8% thiobarbituric acid (TBA), heated for 10 min at 99 degrees Celsius, and assayed for pink coloration. FIG. 47 shows the results of this assay. The left tube in FIG. 47 represents media taken from an overnight culture of cells expressing Vs24254, while the right hand tube shows the TBA reaction using media from cells expressing Vs24259 (negative control). The lack of pink coloration in the negative control indicates that little or no cleavage of the alginate polymer has occurred. Wildtype *E. coli* cells not expressing any recombinant proteins show the same coloration as the negative control Vs24259 (data not shown).

[0541] To test the ability of recombinant *E. coli* to grow on alginate as a sole source of carbon, transformed cells were grown for 19 hours at 30 degrees Celsius with mild shaking in a 96-well plate. Each well held 222 μ l of minimal media (see growth conditions for explanation of minimal media) with the 0.66% carbon source in the form of either degraded alginate or glucose (positive control for growth). All cells were either BL21 with no plasmid (BL21—negative control), one plasmid (Da or 3a), or two plasmids (Dk3a and Da3k). The plasmids are indicated by the lower case letter: “a” refers to the plasmid backbone pET-DEST42 and “k” refers to the pENTR/D/TOPO backbone. “D” indicates that the plasmid contains the genomic region Vs24214-24249, while “3” indicates that the plasmid contains the genomic region Vs24189-24209. Thus, Da would be pET-DEST42-Vs24214-24249, Da3k would be pET-DEST42-Vs24214-24249 and pENTR/D/TOPO-Vs24189-24209 and so on.

[0542] As shown in FIG. 56A, the two vector-constructs pET-DEST42-Vs24214-24249 and pENTR/D/TOPO-Vs24189-24209 when combined in *E. coli* confer growth on degraded alginate as the sole carbon source. This same result is observed when these genomic inserts are switched into the opposite vector (pET-DEST42-Vs24189-24209 and pENTR/D/TOPO-Vs24214-24249). FIG. 56B shows growth on glucose as a positive control. Thus, the combined genomic regions of Vs24214-24249 and Vs24189-24209 from *Vibrio splendidus* were sufficient to confer on *E. coli* the ability to grow on alginate as a sole source of carbon.

Example 11

Production of Ethanol from Alginate

[0543] The ability of recombinant *E. coli* to produce ethanol by growing on alginate on a source of carbon was tested.

To generate recombinant *E. coli*, DNA sequences encoding pyruvate decarboxylase (pdc), and two alcohol dehydrogenase (adhA and adhB) of *Zymomonas mobilis* were amplified by polymerase chain reaction (PCR). These amplified fragments were gel purified and spliced together by another round of PCR. The final amplified DNA fragment was digested with BamHI and XbaI ligated into pBBR1MCS-2 pre-digested with the same restriction enzymes. The resulting plasmid is referred to as pBBRPdc-AdhA/B.

[0544] *E. coli* was transformed with either pBBRPdc-AdhA/B or pBBRPdc-AdhA/B+1.5 Fos (fosmid clone containing genomic region between VI 2B01_24189 and V12B01_24249; these sequences confer on *E. coli* the ability to use alginate as a sole source of carbon, see Examples 1 and 10), grown in m9 media containing alginate, and tested for the production of ethanol. The results are shown in FIG. 57, which demonstrates that the strain harboring pBBRPdc-AdhA/B+1.5 FOS showed significantly higher ethanol production when growing on alginate. These results indicate that the pBBRPdc-AdhA/B+1.5 FOS was able to utilize alginate as a source of carbon in the production of ethanol.

[0545] The various embodiments described above can be combined to provide further embodiments. All of the U.S. patents, U.S. patent application publications, U.S. patent applications, foreign patents, foreign patent applications and non-patent publications referred to in this specification and/or listed in the Application Data Sheet, are incorporated herein by reference, in their entirety. Aspects of the embodiments can be modified, if necessary to employ concepts of the various patents, applications and publications to provide yet further embodiments.

[0546] These and other changes can be made to the embodiments in light of the above-detailed description. In general, in the following claims, the terms used should not be construed to limit the claims to the specific embodiments disclosed in the specification and the claims, but should be construed to include all possible embodiments along with the full scope of equivalents to which such claims are entitled. Accordingly, the claims are not limited by the disclosure.

[0547] The following publications are herein incorporated by reference in their entirety.

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SEQUENCE LISTING

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 Ile Pro Glu Thr Leu Ser Leu Ala Ser Leu Leu Ser Glu Leu Asn Ser
 225 230 235 240
 Asp Ile Thr Ser Ser Gly Gln Ser Val Ile Phe Thr Tyr Asp Ala Thr
 245 250 255
 Thr Asn Ser Ile Val Gly Val Gln Asp Thr Asp Glu Val Leu Arg Ile
 260 265 270
 Asp Ile Asp Ala Val Ser Val Gly Asn Asn Ile Glu Leu Ser Leu Thr
 275 280 285
 Thr Thr Ile Ser Gln Pro Ile Asp His Val Pro Ser Val Gly Gly Gly
 290 295 300
 Gln Val Ser Tyr Thr Gly Asp Gln Ile Asp Ile Ala Phe Asp Ile Gln
 305 310 315 320
 Gly Glu Asp Thr Ala Gly Asn Pro Leu Ala Thr Pro Val Asn Ala Gln
 325 330 335
 Val Ser Val Phe Asp Gly Ile Asp Pro Ser Val Glu Ser Val Asn Ile
 340 345 350
 Thr Asn Val Glu Thr Ser Ser Ala Ala Ile Glu Gly Thr Phe Ser Asn
 355 360 365
 Ile Gly Ser Asp Asn Leu Gln Ser Ala Val Phe Asp Ala Ser Ala Leu
 370 375 380
 Asp Gln Phe Asp Gly Leu Leu Ser Asp Asn Gln Asn Thr Leu Ala Arg
 385 390 395 400
 Leu Ser Asp Asp Gly Thr Thr Ile Thr Leu Ser Ile Gln Gly Arg Gly
 405 410 415
 Glu Val Val Leu Thr Ile Ser Leu Asp Thr Asp Gly Thr Tyr Lys Phe
 420 425 430
 Glu Gln Ser Asn Pro Ile Glu Gln Val Gly Thr Asp Ser Leu Thr Phe
 435 440 445
 Ala Leu Pro Ile Thr Ile Thr Asp Phe Asp Gln Asp Val Val Thr Asn
 450 455 460
 Thr Ile Asn Ile Ala Ile Thr Asp Gly Asp Ser Pro Val Ile Thr Asn
 465 470 475 480
 Val Asp Ser Ile Asp Val Asp Glu Ala Gly Ile Val Gly Gly Ser Gln
 485 490 495
 Glu Gly Thr Ala Pro Val Ser Gly Thr Gly Gly Ile Thr Ala Asp Ile
 500 505 510
 Phe Glu Ser Asp Ile Ile Asp His Tyr Glu Leu Glu Pro Thr Glu Phe
 515 520 525
 Asn Thr Asn Gly Thr Leu Val Ser Asn Gly Glu Ala Val Leu Leu Glu
 530 535 540
 Leu Ile Asp Glu Thr Asn Gly Val Arg Thr Tyr Glu Gly Tyr Val Glu
 545 550 555 560
 Val Asn Gly Ser Arg Ile Thr Val Phe Asp Val Lys Ile Asp Ser Pro
 565 570 575

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| 980 | | | | 985 | | | | 990 | | | | | | | |
|-----|------|------|------|------|------|-----|------|-----|------|------|------|------|-----|------|------|
| Asp | His | Ser | Val | Ser | Glu | Asp | Ile | Val | Lys | Ser | Ile | Val | Val | Thr | Ser |
| | 995 | | | | | | 1000 | | | | | 1005 | | | |
| Ser | Asp | Phe | Asp | Asn | Asp | Pro | Val | Thr | Ser | Thr | Ile | Thr | Leu | Thr | Ile |
| | 1010 | | | | | | 1015 | | | | | 1020 | | | |
| Thr | Asp | Gly | Asp | Asn | Pro | Thr | Ile | Asp | Val | Ile | Pro | Ser | Val | Thr | Leu |
| | 1025 | | | | 1030 | | | | | | 1035 | | | | 1040 |
| Ser | Glu | Ile | Asn | Leu | Ser | Asp | Gly | Ser | Ala | Pro | Ser | Gly | Ser | Ala | Val |
| | | | | 1045 | | | | | | 1050 | | | | 1055 | |
| Ser | Ser | Thr | Gln | Thr | Ile | Thr | Phe | Thr | Asn | Gln | Ser | Asp | Asp | Val | Val |
| | | | 1060 | | | | | | 1065 | | | | | 1070 | |
| Arg | Phe | Arg | Ile | Glu | Ser | Thr | Glu | Phe | Asn | Thr | Asn | Asp | Asp | Leu | Lys |
| | | 1075 | | | | | 1080 | | | | | 1085 | | | |
| Ser | Asn | Gly | Leu | Ala | Val | Glu | Leu | Arg | Glu | Asp | Pro | Ala | Gly | Ser | Gly |
| | 1090 | | | | | | 1095 | | | | 1100 | | | | |
| Asp | Tyr | Ile | Gly | Phe | Thr | Thr | Ser | Ala | Thr | Asn | Val | Glu | Thr | Pro | Val |
| | 1105 | | | | | | 1110 | | | | 1115 | | | | 1120 |
| Phe | Thr | Leu | Ser | Phe | Asn | Ser | Gly | Ser | Leu | Gly | Glu | Tyr | Thr | Phe | Thr |
| | | | | 1125 | | | | | | 1130 | | | | 1135 | |
| Leu | Ile | Glu | Ala | Leu | Asp | His | Gln | Asp | Ala | Arg | Gly | Asn | Asn | Asp | Leu |
| | | | 1140 | | | | | | 1145 | | | | | 1150 | |
| Ser | Phe | Asp | Leu | Pro | Val | Tyr | Ala | Val | Asp | Ser | Asp | Gly | Asp | Asp | Ser |
| | | 1155 | | | | | 1160 | | | | | 1165 | | | |
| Leu | Val | Ser | Pro | Leu | Asn | Val | Thr | Ile | Gly | Asp | Asp | Val | Gln | Ile | Met |
| | 1170 | | | | | | 1175 | | | | 1180 | | | | |
| Gln | Asp | Ser | Thr | Leu | Asp | Ile | Val | Glu | Pro | Thr | Val | Ala | Asp | Leu | Ala |
| | 1185 | | | | 1190 | | | | | | 1195 | | | | 1200 |
| Ala | Gly | Thr | Val | Thr | Thr | Asn | Thr | Ile | Asp | Val | Met | Pro | Asn | Gln | Ser |
| | | | 1205 | | | | | | 1210 | | | | | 1215 | |
| Ala | Asp | Gly | Ala | Thr | Val | Thr | Gln | Phe | Thr | Tyr | Asp | Gly | Gln | Leu | Arg |
| | | | 1220 | | | | | | 1225 | | | | | 1230 | |
| Thr | Leu | Asp | Gln | Asn | Asp | Asn | Gly | Glu | Gln | Gln | Phe | Ser | Phe | Thr | Glu |
| | | 1235 | | | | | 1240 | | | | | 1245 | | | |
| Gly | Glu | Leu | Phe | Ile | Thr | Leu | Gln | Gly | Asp | Val | Arg | Phe | Glu | Pro | Asn |
| | 1250 | | | | | | 1255 | | | | 1260 | | | | |
| Arg | Asn | Leu | Asp | His | Thr | Leu | Ser | Glu | Asp | Ile | Val | Lys | Ser | Ile | Val |
| | 1265 | | | | 1270 | | | | | | 1275 | | | | 1280 |
| Val | Thr | Ser | Ser | Asp | Ser | Asp | Asn | Asp | Val | Leu | Thr | Ser | Thr | Val | Thr |
| | | | | 1285 | | | | | 1290 | | | | | 1295 | |
| Leu | Thr | Ile | Thr | Asp | Gly | Asp | Ile | Pro | Thr | Ile | Asp | Asn | Val | Pro | Thr |
| | | | 1300 | | | | | | 1305 | | | | | 1310 | |
| Val | Asn | Leu | Ser | Glu | Thr | Asn | Leu | Ser | Asp | Gly | Ser | Ala | Pro | Ser | Gly |
| | | 1315 | | | | | 1320 | | | | | 1325 | | | |
| Ser | Ala | Val | Ser | Ser | Thr | Gln | Thr | Ile | Thr | Tyr | Thr | Thr | Gln | Ser | Asp |
| | | 1330 | | | | | 1335 | | | | | 1340 | | | |
| Asp | Val | Thr | Ser | Phe | Arg | Ile | Glu | Pro | Thr | Glu | Phe | Asn | Val | Gly | Gly |
| | 1345 | | | | 1350 | | | | | | 1355 | | | | 1360 |
| Ala | Leu | Thr | Ser | Asn | Gly | Leu | Ala | Val | Glu | Leu | Lys | Ala | Asp | Pro | Thr |
| | | | | 1365 | | | | | 1370 | | | | | 1375 | |
| Thr | Pro | Gly | Gly | Tyr | Ile | Gly | Phe | Val | Thr | Asp | Gly | Ser | Asn | Val | Glu |
| | | | 1380 | | | | | | 1385 | | | | | 1390 | |

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Thr Asn Val Phe Thr Ile Ser Phe Ser Asp Thr Asn Leu Gly Gln Tyr
 1395 1400 1405

Thr Phe Thr Leu Leu Glu Ala Leu Asp His Val Asp Gly Leu Ala Asn
 1410 1415 1420

Asn Asp Leu Thr Phe Asp Leu Pro Val Tyr Ala Val Asp Ser Asp Gly
 1425 1430 1435 1440

Asp Asp Ser Leu Val Ser Gln Leu Asn Val Thr Ile Gly Asp Asp Val
 1445 1450 1455

Gln Ile Met Gln Gly Gly Thr Leu Asp Ile Thr Glu Pro Asn Leu Ala
 1460 1465 1470

Asp Gly Thr Ile Thr Thr Asn Thr Ile Asp Val Met Pro Glu Gln Ser
 1475 1480 1485

Ala Asp Gly Ala Thr Ile Thr Gln Phe Thr Tyr Asp Gly Gln Val Arg
 1490 1495 1500

Thr Leu Asp Gln Thr Asp Asn Gly Glu Gln Gln Phe Ser Phe Thr Glu
 1505 1510 1515 1520

Gly Glu Leu Phe Ile Thr Leu Gln Gly Asp Val Arg Phe Glu Pro Asn
 1525 1530 1535

Arg Asn Leu Asp His Thr Ala Ser Glu Asp Ile Val Lys Ser Ile Val
 1540 1545 1550

Val Thr Ser Ser Asp Leu Asp Asn Asp Val Val Thr Ser Thr Val Thr
 1555 1560 1565

Leu Thr Ile Thr Asp Gly Asp Ile Pro Thr Ile Asp Ala Val Pro Ser
 1570 1575 1580

Val Thr Leu Ser Glu Ile Asn Leu Ser Asp Gly Ser Ala Pro Ser Gly
 1585 1590 1595 1600

Thr Ala Val Ser Gln Thr Glu Thr Ile Thr Phe Thr Asn Gln Ser Asp
 1605 1610 1615

Asp Val Thr Ser Phe Arg Ile Glu Pro Ile Glu Phe Asn Val Gly Gly
 1620 1625 1630

Ala Leu Lys Ser Asn Gly Phe Ala Val Glu Ile Lys Glu Asp Ser Ala
 1635 1640 1645

Asn Pro Gly Thr Tyr Ile Gly Phe Ile Thr Asn Gly Ser Gly Ala Glu
 1650 1655 1660

Ile Pro Val Phe Thr Ile Ala Phe Ser Thr Ser Ser Leu Gly Glu Tyr
 1665 1670 1675 1680

Thr Phe Thr Leu Leu Glu Ala Leu Asp His Val Asp Gly Leu Asp Lys
 1685 1690 1695

Asn Asp Leu Ser Phe Asp Leu Pro Val Tyr Ala Val Asp Thr Asp Gly
 1700 1705 1710

Asp Asp Ser Leu Val Ser Gln Leu Asn Val Thr Ile Gly Asp Asp Val
 1715 1720 1725

Gln Ile Met Gln Asp Gly Thr Leu Asp Ile Ile Glu Pro Asn Leu Ala
 1730 1735 1740

Asp Gly Thr Ile Thr Thr Ser Thr Ile Asp Val Met Pro Asn Gln Ser
 1745 1750 1755 1760

Ala Asp Gly Ala Thr Ile Thr Gln Phe Thr Tyr Asp Gly Gln Leu Arg
 1765 1770 1775

Thr Leu Asp Gln Asn Asp Thr Gly Glu Gln Gln Phe Ser Phe Thr Glu
 1780 1785 1790

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Gly Glu Leu Phe Ile Thr Leu Glu Gly Glu Val Arg Phe Glu Pro Asn
 1795 1800 1805
 Arg Asp Leu Asp His Thr Ala Ser Glu Asp Ile Val Lys Ser Ile Val
 1810 1815 1820
 Val Thr Ser Ser Asp Phe Asp Asn Asp Ser Leu Thr Ser Thr Val Thr
 1825 1830 1835 1840
 Leu Thr Ile Thr Asp Gly Asp Asn Pro Thr Ile Asp Val Ile Pro Ser
 1845 1850 1855
 Val Thr Leu Ser Glu Thr Asn Leu Ser Asp Gly Ser Ala Pro Ser Gly
 1860 1865 1870
 Ser Ala Val Ser Ser Thr Gln Thr Ile Thr Phe Thr Asn Gln Ser Asp
 1875 1880 1885
 Asp Val Val Arg Phe Arg Ile Glu Pro Thr Glu Phe Asn Thr Asn Asp
 1890 1895 1900
 Asp Leu Lys Ser Asn Gly Leu Ala Val Glu Leu Arg Glu Asp Pro Ala
 1905 1910 1915 1920
 Gly Ser Gly Asp Tyr Ile Gly Phe Thr Thr Ser Ala Thr Asn Val Glu
 1925 1930 1935
 Thr Thr Val Phe Thr Leu Ser Phe Ser Ser Thr Thr Leu Gly Glu Tyr
 1940 1945 1950
 Thr Phe Thr Leu Leu Glu Ala Leu Asp His Gln Asp Ala Arg Gly Asn
 1955 1960 1965
 Asn Asp Leu Ser Phe Glu Leu Pro Val Tyr Ala Val Asp Ser Asp Gly
 1970 1975 1980
 Asp Asp Ser Leu Met Ser Pro Leu Asn Val Thr Ile Gly Asp Asp Val
 1985 1990 1995 2000
 Gln Ile Met Gln Asp Gly Thr Leu Asp Ile Val Glu Pro Thr Val Ala
 2005 2010 2015
 Asp Leu Ala Ala Gly Ile Val Thr Thr Asn Thr Ile Asp Val Met Pro
 2020 2025 2030
 Asn Gln Ser Ala Asp Gly Ala Thr Ile Thr Gln Phe Thr Tyr Asp Gly
 2035 2040 2045
 Gln Leu Arg Thr Leu Asp Gln Asn Asp Asn Gly Glu Gln Gln Phe Ser
 2050 2055 2060
 Phe Thr Glu Gly Glu Leu Phe Ile Thr Leu Glu Gly Glu Val Arg Phe
 2065 2070 2075 2080
 Glu Pro Asn Arg Asn Leu Asp His Thr Leu Asn Glu Asp Ile Val Lys
 2085 2090 2095
 Ser Ile Val Val Thr Ser Ser Asp Ser Asp Asn Asp Val Leu Thr Ser
 2100 2105 2110
 Thr Val Thr Leu Thr Ile Thr Asp Gly Asp Ile Pro Thr Ile Asp Asn
 2115 2120 2125
 Val Pro Thr Val Ser Leu Ser Glu Thr Ser Leu Ser Asp Gly Ser Ser
 2130 2135 2140
 Pro Ser Gly Ser Ala Val Ser Ser Thr Gln Thr Ile Thr Tyr Thr Thr
 2145 2150 2155 2160
 Gln Ser Asp Asp Val Thr Ser Phe Arg Ile Glu Pro Thr Glu Phe Asn
 2165 2170 2175
 Val Gly Gly Ala Leu Lys Ser Asn Gly Leu Ala Val Glu Leu Lys Ala
 2180 2185 2190
 Asp Pro Thr Thr Pro Gly Gly Tyr Ile Gly Phe Val Thr Asp Gly Ser

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| 2195 | | | | 2200 | | | | 2205 | | | | | | | |
|------|-----|-----|-----|------|-----|-----|-----|------|-----|-----|-----|------|-----|-----|-----|
| Asn | Val | Glu | Thr | Asn | Val | Phe | Thr | Ile | Ser | Phe | Ser | Asp | Thr | Asn | Leu |
| 2210 | | | | 2215 | | | | 2220 | | | | | | | |
| Gly | Gln | Tyr | Thr | Phe | Thr | Leu | Leu | Glu | Ala | Leu | Asp | His | Ala | Asp | Ser |
| 2225 | | | | 2230 | | | | 2235 | | | | 2240 | | | |
| Leu | Ala | Asn | Asn | Asp | Leu | Ser | Phe | Asp | Leu | Pro | Val | Tyr | Ala | Val | Asp |
| 2245 | | | | 2250 | | | | 2255 | | | | | | | |
| Ser | Asp | Gly | Asp | Asp | Ser | Leu | Val | Ser | Gln | Leu | Asn | Val | Thr | Ile | Gly |
| 2260 | | | | 2265 | | | | 2270 | | | | | | | |
| Asp | Asp | Val | Gln | Ile | Met | Gln | Gly | Gly | Thr | Leu | Asp | Ile | Thr | Glu | Pro |
| 2275 | | | | 2280 | | | | 2285 | | | | | | | |
| Asn | Leu | Ala | Asp | Gly | Thr | Thr | Thr | Thr | Asn | Thr | Ile | Asp | Val | Met | Pro |
| 2290 | | | | 2295 | | | | 2300 | | | | | | | |
| Glu | Gln | Ser | Ala | Asp | Gly | Ala | Thr | Ile | Thr | Gln | Phe | Thr | Tyr | Asp | Gly |
| 2305 | | | | 2310 | | | | 2315 | | | | 2320 | | | |
| Gln | Val | Arg | Thr | Leu | Asp | Gln | Thr | Asp | Asn | Gly | Glu | Gln | Gln | Phe | Ser |
| 2325 | | | | 2330 | | | | 2335 | | | | | | | |
| Phe | Thr | Glu | Gly | Glu | Leu | Phe | Ile | Thr | Leu | Gln | Gly | Asp | Val | Arg | Phe |
| 2340 | | | | 2345 | | | | 2350 | | | | | | | |
| Glu | Pro | Asn | Arg | Asn | Leu | Asp | His | Thr | Ala | Ser | Glu | Asp | Ile | Val | Lys |
| 2355 | | | | 2360 | | | | 2365 | | | | | | | |
| Ser | Ile | Val | Val | Thr | Ser | Ser | Asp | Ser | Asp | Asn | Asp | Val | Val | Thr | Ser |
| 2370 | | | | 2375 | | | | 2380 | | | | | | | |
| Thr | Val | Thr | Leu | Thr | Ile | Thr | Asp | Gly | Asp | Leu | Pro | Thr | Ile | Asp | Ala |
| 2385 | | | | 2390 | | | | 2395 | | | | 2400 | | | |
| Val | Pro | Ser | Val | Thr | Leu | Ser | Glu | Thr | Asn | Leu | Ser | Asp | Gly | Ser | Ala |
| 2405 | | | | 2410 | | | | 2415 | | | | | | | |
| Pro | Ser | Gly | Ser | Ala | Val | Ser | Gln | Thr | Glu | Thr | Ile | Thr | Phe | Thr | Asn |
| 2420 | | | | 2425 | | | | 2430 | | | | | | | |
| Gln | Ser | Asp | Asp | Val | Ala | Ser | Phe | Arg | Ile | Glu | Pro | Thr | Glu | Phe | Asn |
| 2435 | | | | 2440 | | | | 2445 | | | | | | | |
| Val | Gly | Gly | Ala | Leu | Lys | Ser | Asn | Gly | Phe | Ala | Val | Glu | Ile | Lys | Glu |
| 2450 | | | | 2455 | | | | 2460 | | | | | | | |
| Asp | Ser | Ala | Asn | Pro | Gly | Thr | Tyr | Ile | Gly | Phe | Ile | Ala | Asn | Gly | Ser |
| 2465 | | | | 2470 | | | | 2475 | | | | 2480 | | | |
| Ser | Ala | Glu | Ile | Pro | Val | Phe | Thr | Ile | Ala | Phe | Ser | Thr | Ser | Thr | Leu |
| 2485 | | | | 2490 | | | | 2495 | | | | | | | |
| Gly | Glu | Tyr | Thr | Phe | Thr | Leu | Leu | Glu | Ala | Leu | Asp | His | Ala | Asp | Gly |
| 2500 | | | | 2505 | | | | 2510 | | | | | | | |
| Leu | Asp | Lys | Asn | Asp | Leu | Ser | Phe | Glu | Leu | Pro | Val | Tyr | Ala | Val | Asp |
| 2515 | | | | 2520 | | | | 2525 | | | | | | | |
| Thr | Asp | Gly | Asp | Asp | Ser | Leu | Val | Ser | Gln | Leu | Asn | Val | Thr | Ile | Gly |
| 2530 | | | | 2535 | | | | 2540 | | | | | | | |
| Asp | Asp | Val | Gln | Ile | Met | Gln | Asp | Gly | Thr | Leu | Asp | Val | Ile | Glu | Pro |
| 2545 | | | | 2550 | | | | 2555 | | | | 2560 | | | |
| Asn | Leu | Ala | Asp | Gly | Thr | Ile | Thr | Thr | Asn | Thr | Ile | Asp | Val | Met | Pro |
| 2565 | | | | 2570 | | | | 2575 | | | | | | | |
| Glu | Gln | Ser | Ala | Asp | Gly | Ala | Thr | Ile | Thr | Gln | Phe | Thr | Tyr | Asp | Gly |
| 2580 | | | | 2585 | | | | 2590 | | | | | | | |
| Gln | Leu | Arg | Thr | Leu | Asp | Gln | Asn | Asp | Thr | Gly | Glu | Gln | Gln | Phe | Ser |
| 2595 | | | | 2600 | | | | 2605 | | | | | | | |

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Phe Thr Glu Gly Glu Leu Phe Ile Thr Leu Glu Gly Glu Val Arg Phe
 2610 2615 2620
 Glu Pro Asn Arg Asp Leu Asp His Ser Val Ser Glu Asp Ile Val Lys
 2625 2630 2635 2640
 Ser Ile Val Val Thr Ser Ser Asp Phe Asp Asn Asp Pro Val Thr Ser
 2645 2650 2655
 Ala Ile Thr Leu Thr Ile Thr Asp Gly Asp Asn Pro Thr Ile Asp Ser
 2660 2665 2670
 Val Pro Ser Val Val Leu Glu Glu Ala Asp Leu Thr Asp Gly Ser Ser
 2675 2680 2685
 Pro Ser Gly Ser Ala Val Ser Gln Thr Glu Thr Ile Thr Phe Thr Asn
 2690 2695 2700
 Gln Ser Asp Asp Val Glu Lys Phe Arg Leu Glu Pro Ser Glu Phe Asn
 2705 2710 2715 2720
 Thr Asn Asn Ala Leu Lys Ser Asp Gly Leu Ile Ile Glu Ile Arg Glu
 2725 2730 2735
 Glu Pro Thr Gly Ser Gly Asn Tyr Ile Gly Phe Thr Thr Asp Ile Ser
 2740 2745 2750
 Asn Val Glu Thr Thr Val Phe Thr Leu Asp Phe Ser Ser Thr Thr Leu
 2755 2760 2765
 Gly Glu Tyr Thr Phe Thr Leu Leu Glu Ala Ile Asp His Thr Pro Val
 2770 2775 2780
 Gln Gly Asn Asn Asp Leu Thr Phe Asn Leu Pro Val Tyr Ala Val Asp
 2785 2790 2795 2800
 Ser Asp Gly Asp Asp Ser Leu Met Ser Ser Leu Ser Val Thr Ile Thr
 2805 2810 2815
 Asp Asp Val Gln Val Met Val Ser Gly Ser Leu Ser Ile Glu Glu Pro
 2820 2825 2830
 Thr Val Ala Asp Leu Ala Ala Gly Thr Pro Thr Thr Ser Val Phe Asp
 2835 2840 2845
 Val Leu Thr Ser Ala Ser Ala Asp Gly Ala Thr Ile Thr Gln Phe Thr
 2850 2855 2860
 Tyr Asp Gly Gly Ala Val Leu Thr Leu Asp Gln Asn Asp Thr Gly Glu
 2865 2870 2875 2880
 Gln Lys Phe Val Val Ala Asp Gly Ala Leu Tyr Ile Thr Leu Gln Gly
 2885 2890 2895
 Asp Ile Arg Phe Glu Pro Ser Arg Asn Leu Asp His Thr Gly Gly Asp
 2900 2905 2910
 Ile Val Lys Ser Ile Val Val Thr Ser Ser Asp Ser Asp Ser Asp Leu
 2915 2920 2925
 Val Ser Ser Thr Val Thr Leu Thr Ile Thr Asp Gly Asp Ile Pro Thr
 2930 2935 2940
 Ile Asp Thr Val Pro Ser Val Thr Leu Ser Glu Thr Asn Leu Ser Asp
 2945 2950 2955 2960
 Gly Ser Ala Pro Asn Ala Ser Ala Val Ser Ser Thr Gln Thr Ile Thr
 2965 2970 2975
 Phe Thr Asn Gln Ser Asp Asp Val Thr Ser Phe Arg Ile Glu Pro Thr
 2980 2985 2990
 Asp Phe Asn Val Gly Gly Ala Leu Lys Ser Asn Gly Leu Ala Val Glu
 2995 3000 3005

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Leu Lys Ala Asp Pro Thr Thr Pro Gly Gly Tyr Ile Gly Phe Val Thr
 3010 3015 3020
 Asp Gly Ser Asn Val Glu Thr Asn Val Phe Thr Ile Ser Phe Ser Asp
 3025 3030 3035 3040
 Thr Asn Leu Gly Gln Tyr Thr Phe Thr Leu Leu Glu Ala Leu Asp His
 3045 3050 3055
 Val Asp Gly Leu Val Lys Asn Asp Leu Thr Phe Asp Leu Pro Val Tyr
 3060 3065 3070
 Ala Val Asp Ser Asp Gly Asp Asp Ser Leu Val Ser Gln Leu Asn Val
 3075 3080 3085
 Thr Ile Gly Asp Asp Val Gln Val Met Gln Asn Gln Ala Leu Asn Ile
 3090 3095 3100
 Ile Glu Pro Thr Val Ala Asp Leu Ala Ala Gly Thr Pro Thr Thr Ala
 3105 3110 3115 3120
 Thr Val Asp Val Met Pro Ser Gln Ser Ala Asp Gly Ala Thr Ile Thr
 3125 3130 3135
 Gln Phe Thr Tyr Asp Gly Gly Ala Ala Ile Thr Leu Asp Gln Asn Asp
 3140 3145 3150
 Thr Gly Glu Gln Lys Phe Val Phe Thr Glu Gly Ser Leu Phe Ile Thr
 3155 3160 3165
 Leu Gln Gly Glu Val Arg Phe Glu Pro Asn Arg Asn Leu Asn His Thr
 3170 3175 3180
 Ala Ser Glu Asp Ile Val Lys Ser Ile Val Val Thr Ser Ser Asp Leu
 3185 3190 3195 3200
 Asp Asn Asp Val Leu Thr Ser Thr Val Thr Leu Thr Ile Thr Asp Gly
 3205 3210 3215
 Asp Ile Pro Thr Ile Asp Ala Val Pro Ser Val Thr Leu Ser Glu Thr
 3220 3225 3230
 Asn Leu Ser Asp Gly Ser Ala Pro Ser Ser Ser Ala Val Ser Gln Thr
 3235 3240 3245
 Glu Thr Ile Thr Phe Ile Asn Gln Ser Asp Asp Val Ala Ser Phe Arg
 3250 3255 3260
 Ile Glu Pro Thr Glu Phe Asn Val Gly Gly Ala Leu Lys Ser Asn Gly
 3265 3270 3275 3280
 Phe Ala Val Glu Ile Lys Glu Asp Ser Ala Asn Pro Gly Thr Tyr Ile
 3285 3290 3295
 Gly Phe Ile Thr Asp Gly Ser Asn Thr Glu Val Pro Val Phe Thr Ile
 3300 3305 3310
 Ala Phe Ser Thr Ser Thr Leu Gly Glu Tyr Thr Phe Thr Leu Leu Glu
 3315 3320 3325
 Ala Leu Asp His Ala Asn Gly Leu Asp Lys Asn Asp Leu Ser Phe Asp
 3330 3335 3340
 Leu Pro Val Tyr Ala Val Asp Ser Asp Gly Asp Asp Ser Leu Val Ser
 3345 3350 3355 3360
 Gln Leu Asn Val Thr Ile Gly Asp Asp Val Gln Ile Met Gln Asp Gly
 3365 3370 3375
 Thr Leu Asp Ile Thr Glu Pro Asn Leu Ala Asp Gly Thr Ile Thr Thr
 3380 3385 3390
 Asn Thr Ile Asp Val Met Pro Asn Gln Ser Ala Asp Gly Ala Thr Ile
 3395 3400 3405
 Thr Glu Phe Ser Phe Gly Gly Ile Val Lys Thr Leu Asp Gln Ser Ile

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| 3410 | | | | | 3415 | | | | | 3420 | | | | |
|---|---|------|--|--|------|--|--|------|--|------|--|--|--|------|
| Val Gly Glu Gln Gln | Phe Ser Phe Thr Glu Gly Glu Leu Phe Ile Thr | | | | | | | | | | | | | |
| 3425 | | 3430 | | | | | | 3435 | | | | | | 3440 |
| Leu Gln Gly Gln Val Arg Phe Glu Pro Asn Arg Asp Leu Asp His Ser | | | | | | | | | | | | | | |
| | | 3445 | | | | | | 3450 | | | | | | 3455 |
| Ala Ser Glu Asp Ile Val Lys Ser Ile Val Val Thr Ser Ser Asp Phe | | | | | | | | | | | | | | |
| | | 3460 | | | | | | 3465 | | | | | | 3470 |
| Asp Asn Asp Pro Val Thr Ser Thr Val Thr Leu Thr Ile Thr Asp Gly | | | | | | | | | | | | | | |
| | | 3475 | | | | | | 3480 | | | | | | 3485 |
| Asp Ile Pro Thr Ile Asp Ala Val Pro Ser Val Thr Leu Ser Glu Thr | | | | | | | | | | | | | | |
| | | 3490 | | | | | | 3495 | | | | | | 3500 |
| Asn Leu Ala Asp Gly Ser Ala Pro Ser Gly Ser Ala Val Ser Gln Thr | | | | | | | | | | | | | | |
| | | 3505 | | | | | | 3510 | | | | | | 3520 |
| Glu Thr Ile Thr Phe Thr Asn Gln Ser Asp Asp Val Val Arg Phe Arg | | | | | | | | | | | | | | |
| | | 3525 | | | | | | 3530 | | | | | | 3535 |
| Leu Glu Pro Thr Glu Phe Asn Thr Asn Asp Ala Leu Lys Ser Asn Gly | | | | | | | | | | | | | | |
| | | 3540 | | | | | | 3545 | | | | | | 3550 |
| Leu Ala Val Glu Leu Arg Glu Glu Pro Gln Gly Ser Gly Gln Tyr Ile | | | | | | | | | | | | | | |
| | | 3555 | | | | | | 3560 | | | | | | 3565 |
| Gly Phe Thr Thr Ser Ser Ser Asn Val Glu Thr Thr Val Phe Thr Leu | | | | | | | | | | | | | | |
| | | 3570 | | | | | | 3575 | | | | | | 3580 |
| Asp Phe Asn Ser Gly Thr Leu Gly Glu Tyr Thr Phe Thr Leu Ile Glu | | | | | | | | | | | | | | |
| | | 3585 | | | | | | 3590 | | | | | | 3600 |
| Ala Leu Asp His Gln Asp Ala Arg Gly Asn Asn Asp Leu Ser Phe Asn | | | | | | | | | | | | | | |
| | | 3605 | | | | | | 3610 | | | | | | 3615 |
| Leu Pro Val Tyr Ala Val Asp Ser Asp Gly Asp Asp Ser Leu Val Ser | | | | | | | | | | | | | | |
| | | 3620 | | | | | | 3625 | | | | | | 3630 |
| Gln Leu Gly Val Thr Ile Gly Asp Asp Val Gln Leu Met Gln Asp Gly | | | | | | | | | | | | | | |
| | | 3635 | | | | | | 3640 | | | | | | 3645 |
| Thr Ile Thr Ser Arg Glu Pro Ala Ala Ser Val Glu Thr Ser Asn Thr | | | | | | | | | | | | | | |
| | | 3650 | | | | | | 3655 | | | | | | 3660 |
| Phe Asp Val Met Pro Asn Gln Ser Ala Asp Gly Ala Lys Val Thr Ser | | | | | | | | | | | | | | |
| | | 3665 | | | | | | 3670 | | | | | | 3680 |
| Phe Val Phe Asp Gly Lys Thr Ala Glu Ser Leu Asp Leu Asn Val Asn | | | | | | | | | | | | | | |
| | | 3685 | | | | | | 3690 | | | | | | 3695 |
| Gly Glu Gln Glu Phe Val Phe Thr Glu Gly Ser Val Phe Ile Thr Thr | | | | | | | | | | | | | | |
| | | 3700 | | | | | | 3705 | | | | | | 3710 |
| Glu Gly Glu Ile Arg Phe Glu Pro Val Arg Asn Gln Asn His Ala Gly | | | | | | | | | | | | | | |
| | | 3715 | | | | | | 3720 | | | | | | 3725 |
| Gly Asp Ile Thr Lys Ser Ile Glu Val Thr Ser Val Asp Leu Asp Gly | | | | | | | | | | | | | | |
| | | 3730 | | | | | | 3735 | | | | | | 3740 |
| Asp Ile Val Thr Ser Thr Val Thr Leu Lys Ile Val Asp Gly Asp Leu | | | | | | | | | | | | | | |
| | | 3745 | | | | | | 3750 | | | | | | 3760 |
| Pro Thr Ile Asp Leu Val Pro Gly Ile Thr Leu Ser Glu Val Asp Leu | | | | | | | | | | | | | | |
| | | 3765 | | | | | | 3770 | | | | | | 3775 |
| Ala Asp Gly Ser Val Pro Thr Gly Asn Pro Val Thr Met Thr Gln Thr | | | | | | | | | | | | | | |
| | | 3780 | | | | | | 3785 | | | | | | 3790 |
| Ile Thr Tyr Thr Ala Gly Ser Asp Asp Val Ser His Phe Arg Ile Asp | | | | | | | | | | | | | | |
| | | 3795 | | | | | | 3800 | | | | | | 3805 |
| Pro Thr Gln Phe Asn Thr Ser Gly Val Leu Lys Ser Asn Gly Leu Asp | | | | | | | | | | | | | | |
| | | 3810 | | | | | | 3815 | | | | | | 3820 |

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Val Glu Ile Lys Glu Gln Pro Ala Asn Ser Gly Asn Tyr Ile Gly Phe
 3825 3830 3835 3840
 Val Lys Asp Gly Ser Asn Val Glu Thr Asn Val Phe Thr Ile Ser Phe
 3845 3850 3855
 Ser Thr Ser Asn Leu Gly Gln Tyr Thr Phe Thr Leu Leu Glu Ala Leu
 3860 3865 3870
 Asp His Val Asp Gly Leu Gln Asn Asn Ile Leu Ser Phe Asp Val Pro
 3875 3880 3885
 Val Leu Ala Val Asp Ala Asp Gly Asp Asp Ser Ala Met Ser Pro Met
 3890 3895 3900
 Thr Val Ala Ile Thr Asp Asp Val Gln Gly Val Gln Asp Gly Thr Leu
 3905 3910 3915 3920
 Ser Ile Thr Glu Pro Ser Leu Ala Asp Leu Ala Ser Gly Thr Pro Pro
 3925 3930 3935
 Thr Thr Ala Ile Ile Asp Val Met Pro Thr Gln Ser Ala Asp Gly Ala
 3940 3945 3950
 Lys Val Thr Gln Phe Thr Tyr Asp Gly Gly Thr Ala Val Thr Leu Asp
 3955 3960 3965
 Pro Ser Ile Ala Thr Glu Gln Val Phe Thr Val Thr Asp Gly Leu Leu
 3970 3975 3980
 Tyr Ile Thr Ile Glu Gly Glu Val Arg Phe Glu Pro Ser Arg Asp Leu
 3985 3990 3995 4000
 Asp His Ser Ser Gly Asp Ile Val Arg Thr Ile Val Val Thr Thr Ser
 4005 4010 4015
 Asp Phe Asp Asn Asp Thr Asp Thr Ala Asp Val Thr Leu Thr Ile Lys
 4020 4025 4030
 Asp Gly Ile Asn Pro Val Ile Asn Val Val Pro Asp Val Asn Leu Ser
 4035 4040 4045
 Glu Val Asn Leu Ala Asp Gly Ser Thr Pro Ser Gly Ser Ala Val Ser
 4050 4055 4060
 Ser Thr His Thr Ile Thr Tyr Thr Glu Gly Ser Asp Asp Phe Ser His
 4065 4070 4075 4080
 Phe Arg Ile Ala Thr Asn Glu Phe Asn Pro Gly Asp Leu Leu Lys Ser
 4085 4090 4095
 Ser Gly Leu Val Val Gln Leu Lys Glu Asp Pro Ala Ser Ala Gly Asp
 4100 4105 4110
 Tyr Ile Gly Tyr Thr Asp Asp Gly Met Gly Asn Val Thr Asp Val Phe
 4115 4120 4125
 Thr Ile Ser Phe Asp Ser Ala Asn Lys Ala Gln Phe Thr Phe Thr Leu
 4130 4135 4140
 Ile Glu Ala Leu Asp His Leu Asp Gly Val Leu Tyr Asn Asp Leu Thr
 4145 4150 4155 4160
 Phe Arg Leu Pro Ile Tyr Ala Val Asp Thr Asp Asp Ser Glu Ser Thr
 4165 4170 4175
 Lys Arg Asp Val Val Val Thr Ile Glu Asp Asp Ile Gln Gln Met Gln
 4180 4185 4190
 Asp Gly Phe Leu Thr Ile Thr Glu Pro Asn Ser Gly Thr Pro Thr Thr
 4195 4200 4205
 Thr Thr Val Asp Val Met Pro Ile Pro Ser Ala Asp Gly Ala Thr Ile
 4210 4215 4220

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Thr Gln Phe Thr Tyr Asp Gly Gly Ser Pro Ile Thr Leu Asn Gln Ser
 4225 4230 4235 4240
 Ile Ser Gly Glu Gln Glu Phe Val Phe Thr Glu Gly Ser Leu Phe Val
 4245 4250 4255
 Thr Leu Asp Gly Asp Val Arg Phe Glu Pro Asn Arg Asn Leu Asp His
 4260 4265 4270
 Ser Ala Gly Asp Ile Val Lys Ser Ile Val Phe Thr Ser Ser Asp Phe
 4275 4280 4285
 Asp Asn Asp Ile Phe Ser Ser Lys Val Thr Leu Thr Ile Val Asp Gly
 4290 4295 4300
 Asp Gly Pro Thr Ile Asp Val Val Pro Gly Val Ala Leu Ser Glu Ser
 4305 4310 4315 4320
 Leu Leu Ala Asp Gly Ser Thr Pro Ser Val Asn Pro Val Ser Met Thr
 4325 4330 4335
 Gln Thr Ile Thr Ser Leu Ala Ser Ser Asp Asp Ile Ala Glu Ile Val
 4340 4345 4350
 Val Glu Val Gly Leu Phe Asn Thr Asn Gly Ala Leu Lys Ser Asp Gly
 4355 4360 4365
 Leu Ser Leu Ser Leu Arg Glu Asp Pro Val Asn Ser Gly Asp Tyr Ile
 4370 4375 4380
 Ala Phe Thr Thr Asn Gly Ser Gly Val Glu Lys Val Ile Phe Thr Leu
 4385 4390 4395 4400
 Asp Phe Asp Asp Thr Asn Pro Ser Gln Tyr Thr Phe Thr Leu Leu Glu
 4405 4410 4415
 Arg Leu Asp His Val Asp Gly Leu Gly Asn Asn Asp Leu Ser Phe Asp
 4420 4425 4430
 Leu Ser Val Tyr Ala Glu Asp Thr Asp Gly Asp Ile Ser Ala Ser Lys
 4435 4440 4445
 Pro Leu Thr Val Thr Ile Thr Asp Asp Val Gln Leu Met Gln Ser Gly
 4450 4455 4460
 Ala Leu Asn Ile Thr Glu Pro Thr Thr Gly Thr Pro Thr Thr Ala Val
 4465 4470 4475 4480
 Phe Asp Val Met Pro Ala Gln Ser Ala Asp Gly Ala Thr Ile Thr Lys
 4485 4490 4495
 Phe Thr Tyr Gly Ser Gln Pro Glu Glu Ser Leu Val Gln Thr Val Thr
 4500 4505 4510
 Gly Glu Gln Glu Phe Val Phe Thr Glu Gly Ser Leu Phe Ile Asn Leu
 4515 4520 4525
 Glu Gly Asp Val Arg Phe Glu Pro Asn Arg Asn Leu Asp His Ser Gly
 4530 4535 4540
 Gly Asn Ile Val Lys Thr Ile Thr Val Thr Ser Glu Asp Lys Asp Gly
 4545 4550 4555 4560
 Asp Ile Val Thr Ser Thr Val Thr Leu Thr Ile Val Asp Gly Ala Pro
 4565 4570 4575
 Pro Val Ile Asp Thr Val Pro Thr Val Ala Leu Glu Glu Ala Asn Leu
 4580 4585 4590
 Val Asp Gly Ser Ser Pro Gly Leu Pro Val Ser Gln Thr Glu Ile Ile
 4595 4600 4605
 Thr Phe Thr Ala Gly Ser Asp Asp Val Ser His Phe Arg Ile Asp Pro
 4610 4615 4620
 Ala Gln Phe Asn Thr Ser Gly Asp Leu Lys Ala Asp Gly Leu Val Val

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| | | | |
|---|------|------|------|
| 4625 | 4630 | 4635 | 4640 |
| Gln Leu Lys Glu Asp Pro Leu Asn Ser Asp Asn Tyr Ile Gly Tyr Val | 4645 | 4650 | 4655 |
| Glu Ser Gly Gly Val Gln Thr Asp Ile Phe Thr Ile Thr Phe Ser Ser | 4660 | 4665 | 4670 |
| Val Val Leu Gly Glu Tyr Thr Phe Thr Leu Leu Glu Glu Leu Asp His | 4675 | 4680 | 4685 |
| Leu Pro Val Gln Gly Asn Asn Asp Gln Ile Phe Thr Leu Pro Val Ile | 4690 | 4695 | 4700 |
| Ala Val Asp Lys Asp Asn Thr Asp Ser Ala Val Lys Pro Leu Thr Val | 4705 | 4710 | 4715 |
| Thr Ile Thr Asp Asp Val Pro Thr Ile Thr Asp Thr Thr Gly Ala Ser | 4725 | 4730 | 4735 |
| Thr Phe Val Val Asp Glu Asp Asp Leu Gly Thr Leu Ala Gln Ala Thr | 4740 | 4745 | 4750 |
| Gly Ser Phe Val Thr Thr Glu Gly Ala Asp Gln Val Glu Val Tyr Glu | 4755 | 4760 | 4765 |
| Leu Arg Asn Ile Ser Thr Leu Glu Ala Thr Leu Ser Ser Gly Ser Glu | 4770 | 4775 | 4780 |
| Gly Ile Lys Ile Thr Glu Ile Thr Gly Ala Ala Asn Thr Thr Thr Tyr | 4785 | 4790 | 4795 |
| Gln Gly Ala Thr Asp Pro Ser Gly Thr Pro Ile Phe Thr Leu Val Leu | 4805 | 4810 | 4815 |
| Thr Asp Asp Gly Ala Tyr Thr Phe Thr Leu Leu Gly Pro Leu Asn His | 4820 | 4825 | 4830 |
| Ala Thr Thr Pro Ser Asn Leu Asp Thr Leu Thr Ile Pro Phe Asp Val | 4835 | 4840 | 4845 |
| Val Ala Val Asp Gly Asp Gly Asp Asp Ser Asn Gln Tyr Val Leu Pro | 4850 | 4855 | 4860 |
| Ile Glu Val Leu Asp Asp Val Pro Val Met Thr Ala Pro Thr Gly Glu | 4865 | 4870 | 4875 |
| Thr Val Val Asp Glu Asp Asp Leu Thr Gly Ile Gly Ser Asp Gln Ser | 4885 | 4890 | 4895 |
| Glu Asp Thr Ile Ile Asn Gly Leu Phe Thr Val Asp Glu Gly Ala Asp | 4900 | 4905 | 4910 |
| Gly Val Val Leu Tyr Glu Leu Val Asp Glu Asp Leu Val Leu Thr Gly | 4915 | 4920 | 4925 |
| Leu Thr Ser Asp Gly Glu Ser Leu Glu Trp Leu Ala Val Ser Gln Asn | 4930 | 4935 | 4940 |
| Gly Thr Thr Phe Thr Tyr Val Ala Gln Thr Ala Thr Ser Asn Glu Ala | 4945 | 4950 | 4955 |
| Val Phe Glu Ile Ile Phe Asp Thr Ser Asp Asn Ser Tyr Gln Phe Glu | 4965 | 4970 | 4975 |
| Leu Phe Lys Pro Leu Lys His Pro Asp Gly Ala Asn Glu Asn Ala Ile | 4980 | 4985 | 4990 |
| Asp Leu Asp Phe Ser Ile Val Ala Glu Asp Phe Asp Gln Asp Gln Ser | 4995 | 5000 | 5005 |
| Asp Ala Ile Gly Leu Lys Ile Thr Val Thr Asp Asp Val Pro Leu Val | 5010 | 5015 | 5020 |
| Thr Thr Gln Ser Ile Thr Arg Leu Glu Gly Gln Gly Tyr Gly Asn Ser | 5025 | 5030 | 5035 |
| | | | 5040 |

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Lys Val Asp Met Phe Ala Asn Ala Thr Asp Val Gly Ala Asp Gly Ala
 5045 5050 5055
 Val Leu Ser Arg Ile Glu Gly Ile Ser Asn Asn Gly Ala Asp Ile Val
 5060 5065 5070
 Phe Arg Ser Gly Asn Asn Gly Pro Tyr Ser Ser Gly Phe Asp Leu Asn
 5075 5080 5085
 Ser Gly Ser Gln Gln Val Arg Val Tyr Glu Gln Thr Asn Gly Gly Ala
 5090 5095 5100
 Asp Thr Arg Glu Leu Gly Arg Leu Arg Ile Asn Ser Asn Gly Glu Val
 5105 5110 5115 5120
 Glu Phe Arg Ala Asn Gly Tyr Leu Asp His Asp Gly Asp Asp Thr Ile
 5125 5130 5135
 Asp Phe Ser Ile Asn Val Ile Ala Thr Asp Gly Asp Leu Asp Thr Ser
 5140 5145 5150
 Glu Thr Pro Leu Asp Ile Thr Ile Thr Asp Arg Asp Ser Thr Arg Ile
 5155 5160 5165
 Ala Leu Lys Val Thr Thr Phe Glu Asp Ala Gly Arg Asp Ser Thr Ile
 5170 5175 5180
 Pro Tyr Ala Thr Gly Asp Glu Pro Thr Leu Glu Asn Val Gln Asp Asn
 5185 5190 5195 5200
 Gln Asn Gly Leu Pro Asn Ala Pro Ala Gln Val Ala Leu Gln Val Ser
 5205 5210 5215
 Leu Tyr Asp Gln Asp Asn Ala Glu Ser Ile Gly Gln Leu Thr Ile Lys
 5220 5225 5230
 Ser Pro Asn Gly Gly Asp Ser His Gln Gly Thr Phe Tyr Tyr Phe Asp
 5235 5240 5245
 Gly Ala Asp Tyr Ile Glu Leu Val Pro Glu Ser Asn Gly Ser Ile Ile
 5250 5255 5260
 Phe Gly Ser Pro Glu Leu Glu Gln Ser Phe Ala Pro Asn Pro Ser Glu
 5265 5270 5275 5280
 Pro Arg Gln Thr Ile Ala Thr Ile Asp Asn Leu Phe Phe Val Pro Asp
 5285 5290 5295
 Gln His Ala Ser Ser Asp Glu Thr Gly Gly Arg Val Arg Tyr Glu Leu
 5300 5305 5310
 Glu Ile Glu Lys Asn Gly Ser Thr Asp His Thr Val Asn Ser Asn Phe
 5315 5320 5325
 Arg Ile Glu Ile Glu Ala Val Ala Asp Ile Ala Thr Trp Asp Asp Ser
 5330 5335 5340
 Asn Ser Thr Tyr Gln Tyr Gln Val Asn Glu Asp Glu Asp Asn Val Thr
 5345 5350 5355 5360
 Leu Gln Leu Asn Ala Glu Ser Gln Asp Asn Ser Asn Thr Glu Thr Ile
 5365 5370 5375
 Thr Tyr Glu Leu Glu Ala Val Gln Gly Asp Gly Lys Phe Glu Leu Leu
 5380 5385 5390
 Asp Gln Asn Gly Asn Val Leu Thr Pro Val Asn Gly Val Tyr Ile Ile
 5395 5400 5405
 Ala Ser Ala Asp Ile Asn Ser Thr Val Val Asn Pro Ile Asp Asn Phe
 5410 5415 5420
 Ser Gly Gln Ile Glu Phe Lys Ala Thr Ala Ile Thr Glu Glu Thr Leu
 5425 5430 5435 5440

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| | | | | | | | | | | | | | | | |
|-----|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|
| Asn | Pro | Tyr | Asp | Asp | Ser | Asp | Asn | Gly | Gly | Ala | Asn | Asp | Lys | Thr | Thr |
| | | | 5445 | | | | | | 5450 | | | | | 5455 | |
| Ala | Arg | Ser | Val | Glu | Gln | Ser | Ile | Val | Ile | Asp | Val | Thr | Ala | Asp | Ala |
| | | | 5460 | | | | | | 5465 | | | | | 5470 | |
| Asp | Pro | Gly | Thr | Phe | Ser | Val | Ser | Arg | Ile | Gln | Ile | Asn | Glu | Asp | Asn |
| | | 5475 | | | | | | 5480 | | | | | 5485 | | |
| Ile | Asp | Asp | Pro | Asp | Tyr | Val | Gly | Pro | Leu | Asp | Asn | Lys | Asp | Ala | Phe |
| | 5490 | | | | | 5495 | | | | | | 5500 | | | |
| Thr | Leu | Asp | Glu | Val | Ile | Thr | Met | Thr | Gly | Ser | Val | Asp | Ser | Asp | Ser |
| | 5505 | | | | | 5510 | | | | 5515 | | | | | 5520 |
| Ser | Glu | Glu | Leu | Phe | Val | Arg | Ile | Ser | Asn | Val | Thr | Glu | Gly | Ala | Val |
| | | | | 5525 | | | | | | 5530 | | | | | 5535 |
| Leu | Tyr | Phe | Leu | Gly | Thr | Thr | Thr | Val | Val | Pro | Thr | Ile | Thr | Ile | Asn |
| | | | 5540 | | | | | | | 5545 | | | | 5550 | |
| Gly | Val | Asp | Tyr | Gln | Glu | Ile | Ala | Tyr | Ser | Asp | Leu | Ala | Asn | Val | Glu |
| | | | 5555 | | | | | 5560 | | | | | | 5565 | |
| Val | Val | Pro | Thr | Lys | His | Ser | Asn | Val | Asp | Phe | Thr | Phe | Asp | Val | Thr |
| | | | 5570 | | | | 5575 | | | | | 5580 | | | |
| Gly | Val | Val | Lys | Asp | Thr | Ala | Asn | Leu | Ser | Thr | Gly | Ala | Gln | Ile | Asp |
| | | | 5585 | | | 5590 | | | | | 5595 | | | | 5600 |
| Glu | Glu | Ile | Leu | Gly | Thr | Lys | Thr | Val | Asn | Val | Glu | Val | Lys | Gly | Val |
| | | | | 5605 | | | | | | 5610 | | | | | 5615 |
| Ala | Asp | Thr | Pro | Tyr | Gly | Gly | Thr | Asn | Gly | Thr | Ala | Trp | Ser | Ala | Ile |
| | | | | 5620 | | | | | | 5625 | | | | 5630 | |
| Thr | Asp | Gly | Thr | Thr | Ser | Gly | Val | Gln | Thr | Thr | Ile | Gln | Glu | Ser | Gln |
| | | 5635 | | | | | | 5640 | | | | | 5645 | | |
| Asn | Gly | Asp | Thr | Phe | Ala | Glu | Leu | Asp | Phe | Thr | Val | Leu | Ser | Gly | Glu |
| | | 5650 | | | | | 5655 | | | | | 5660 | | | |
| Arg | Arg | Pro | Asp | Thr | Gly | Thr | Thr | Pro | Leu | Ala | Asp | Asp | Gly | Ser | Glu |
| | | | | | 5670 | | | | | | 5675 | | | | 5680 |
| Ser | Ile | Thr | Val | Ile | Leu | Ser | Gly | Ile | Pro | Asp | Gly | Val | Val | Leu | Glu |
| | | | | | 5685 | | | | | 5690 | | | | | 5695 |
| Asp | Gly | Asp | Gly | Thr | Val | Ile | Asp | Leu | Asn | Phe | Val | Gly | Tyr | Glu | Thr |
| | | | 5700 | | | | | | 5705 | | | | | 5710 | |
| Gly | Pro | Gly | Gly | Ser | Pro | Asp | Leu | Ser | Lys | Pro | Ile | Tyr | Glu | Ala | Asn |
| | | | 5715 | | | | | 5720 | | | | | | 5725 | |
| Ile | Thr | Glu | Ala | Gly | Lys | Thr | Ser | Gly | Ile | Arg | Ile | Arg | Pro | Val | Asp |
| | | 5730 | | | | | | 5735 | | | | | 5740 | | |
| Ser | Ser | Thr | Glu | Asn | Ile | His | Ile | Gln | Gly | Lys | Val | Ile | Val | Thr | Glu |
| | | | 5745 | | | | 5750 | | | | 5755 | | | | 5760 |
| Asn | Asp | Gly | His | Thr | Leu | Thr | Phe | Asp | Gln | Glu | Ile | Arg | Val | Leu | Val |
| | | | | 5765 | | | | | | 5770 | | | | | 5775 |
| Ile | Pro | Arg | Ile | Asp | Thr | Ser | Ala | Thr | Tyr | Val | Asn | Thr | Thr | Asn | Gly |
| | | | | 5780 | | | | | 5785 | | | | | 5790 | |
| Asp | Glu | Asp | Thr | Ala | Ile | Asn | Ile | Asp | Trp | His | Pro | Glu | Gly | Thr | Asp |
| | | | 5795 | | | | | 5800 | | | | | | 5805 | |
| Tyr | Ile | Asp | Asp | Asp | Glu | His | Phe | Thr | Lys | Ile | Thr | Ile | Asn | Gly | Ile |
| | | | 5810 | | | | | 5815 | | | | | | 5820 | |
| Pro | Leu | Gly | Val | Thr | Ala | Val | Val | Asn | Gly | Asp | Val | Thr | Val | Asp | Asp |
| | | | | | 5825 | | | | | | 5835 | | | | 5840 |
| Ser | Thr | Pro | Gly | Thr | Leu | Ile | Ile | Thr | Pro | Lys | Asp | Ala | Ser | Gln | Thr |

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| 5845 | | | | | 5850 | | | | | 5855 | | | | | |
|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|
| Pro | Glu | Gln | Phe | Thr | Gln | Ile | Ala | Leu | Ala | Asn | Asn | Phe | Ile | Gln | Met |
| | | | 5860 | | | | | 5865 | | | | | 5870 | | |
| Thr | Pro | Pro | Ala | Asp | Ser | Ser | Ala | Asp | Phe | Thr | Leu | Thr | Thr | Glu | Leu |
| | | | 5875 | | | | 5880 | | | | | 5885 | | | |
| Lys | Met | Glu | Glu | Arg | Asp | His | Glu | Tyr | Thr | Ser | Ser | Gly | Leu | Glu | Asp |
| | 5890 | | | | 5895 | | | | | | | 5900 | | | |
| Glu | Asp | Gly | Gly | Tyr | Val | Glu | Ala | Asp | Pro | Asp | Ile | Thr | Gly | Ile | Ile |
| 5905 | | | | | 5910 | | | | | 5915 | | | | | 5920 |
| Asn | Val | Gln | Val | Arg | Pro | Val | Val | Glu | Pro | Gly | Asp | Ala | Asp | Asn | Lys |
| | | | | 5925 | | | | | 5930 | | | | | 5935 | |
| Ile | Val | Val | Ser | Asn | Glu | Asp | Gly | Ser | Gly | Asp | Leu | Thr | Thr | Ile | Thr |
| | | | 5940 | | | | | 5945 | | | | | | 5950 | |
| Ala | Asp | Ala | Asn | Gly | Val | Ile | Lys | Phe | Thr | Thr | Asn | Ser | Asp | Asn | Gln |
| | | 5955 | | | | | 5960 | | | | | 5965 | | | |
| Thr | Thr | Asp | Thr | Asn | Gly | Asp | Glu | Ile | Trp | Asp | Gly | Glu | Tyr | Val | Val |
| | 5970 | | | | | 5975 | | | | | 5980 | | | | |
| Arg | Tyr | Gln | Glu | Thr | Asp | Leu | Ser | Thr | Val | Glu | Glu | Gln | Val | Asp | Glu |
| 5985 | | | | | 5990 | | | | | 5995 | | | | 6000 | |
| Val | Ile | Val | Gln | Leu | Thr | Asn | Thr | Asp | Gly | Ser | Ala | Leu | Ser | Asp | Asp |
| | | | | 6005 | | | | | 6010 | | | | | 6015 | |
| Ile | Leu | Gly | Gln | Leu | Leu | Val | Thr | Gly | Ala | Ser | Tyr | Glu | Gly | Gly | Gly |
| | | | 6020 | | | | | 6025 | | | | | 6030 | | |
| Arg | Trp | Val | Val | Thr | Asn | Glu | Asp | Ala | Phe | Ser | Val | Ser | Ala | Pro | Asn |
| | | 6035 | | | | | 6040 | | | | | 6045 | | | |
| Gly | Leu | Asp | Phe | Thr | Pro | Ala | Asn | Asp | Ala | Asp | Asp | Val | Ala | Thr | Asp |
| | 6050 | | | | | | 6055 | | | | 6060 | | | | |
| Phe | Asn | Asp | Ile | Lys | Met | Thr | Ile | Phe | Thr | Leu | Val | Ser | Asp | Pro | Gly |
| 6065 | | | | | 6070 | | | | | 6075 | | | | 6080 | |
| Asp | Ala | Asn | Asn | Glu | Thr | Ser | Ala | Gln | Val | Gln | Arg | Thr | Gly | Glu | Val |
| | | | | 6085 | | | | | 6090 | | | | | 6095 | |
| Thr | Leu | Ser | Tyr | Pro | Glu | Val | Leu | Thr | Ala | Pro | Asp | Lys | Val | Ala | Ala |
| | | | 6100 | | | | | 6105 | | | | | 6110 | | |
| Asp | Ile | Ala | Ile | Val | Pro | Asp | Ser | Val | Ile | Asp | Ala | Val | Glu | Asp | Thr |
| | | 6115 | | | | | 6120 | | | | | 6125 | | | |
| Gln | Leu | Asp | Leu | Gly | Ala | Ala | Leu | Asn | Gly | Ile | Leu | Ser | Leu | Thr | Gly |
| | 6130 | | | | | | 6135 | | | | | 6140 | | | |
| Arg | Asp | Asp | Ser | Thr | Asp | Gln | Val | Thr | Val | Ile | Ile | Asp | Gly | Thr | Leu |
| 6145 | | | | | 6150 | | | | | 6155 | | | | 6160 | |
| Val | Ile | Asp | Ala | Thr | Thr | Ser | Phe | Pro | Ile | Ser | Leu | Ser | Gly | Thr | Ser |
| | | | | 6165 | | | | | 6170 | | | | | 6175 | |
| Asp | Val | Asp | Phe | Val | Asn | Gly | Lys | Tyr | Val | Tyr | Glu | Thr | Thr | Val | Glu |
| | | | 6180 | | | | | 6185 | | | | | 6190 | | |
| Gln | Gly | Val | Ala | Val | Asp | Ser | Ser | Gly | Leu | Leu | Leu | Asn | Leu | Pro | Pro |
| | | 6195 | | | | | 6200 | | | | | 6205 | | | |
| Asn | Tyr | Ser | Gly | Asp | Phe | Arg | Leu | Pro | Met | Thr | Ile | Val | Thr | Lys | Asp |
| | 6210 | | | | | 6215 | | | | | 6220 | | | | |
| Leu | Gln | Ser | Gly | Asp | Glu | Lys | Thr | Leu | Val | Thr | Glu | Val | Ile | Ile | Lys |
| 6225 | | | | | 6230 | | | | | 6235 | | | | 6240 | |
| Val | Ala | Pro | Asp | Ala | Glu | Thr | Asp | Pro | Thr | Ile | Glu | Val | Asn | Val | Val |
| | | | | 6245 | | | | | 6250 | | | | | 6255 | |

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Gly Ser Leu Asp Asp Ala Phe Asn Pro Val Asp Thr Asp Gly Gln Ala
 6260 6265 6270

Gly Gln Asp Pro Val Gly Tyr Glu Asp Thr Tyr Ile Gln Leu Asp Phe
 6275 6280 6285

Asn Ser Thr Ile Ser Asp Gln Val Ser Gly Val Glu Gly Gly Gln Glu
 6290 6295 6300

Ala Phe Thr Ser Ile Thr Leu Thr Leu Asp Asp Pro Ser Ile Gly Ala
 6305 6310 6315 6320

Phe Tyr Asp Asn Thr Gly Thr Ser Leu Gly Thr Ser Val Thr Phe Asn
 6325 6330 6335

Gln Ala Glu Ile Ala Ala Gly Ala Leu Asp Asn Val Leu Phe Arg Ala
 6340 6345 6350

Ile Glu Asn Tyr Pro Thr Gly Asn Asp Ile Asn Gln Val Gln Val Asn
 6355 6360 6365

Val Ser Gly Thr Val Thr Asp Thr Ala Thr Tyr Asn Asp Pro Ala Ser
 6370 6375 6380

Pro Ala Gly Thr Ala Thr Asp Ser Asp Thr Phe Ser Thr Ser Val Ser
 6385 6390 6395 6400

Phe Glu Val Val Pro Val Val Asp Asp Val Ser Val Thr Gly Pro Gly
 6405 6410 6415

Ser Asp Pro Asp Val Ile Glu Ile Thr Gly Asn Glu Asp Gln Leu Ile
 6420 6425 6430

Ser Leu Ser Gly Thr Gly Pro Val Ser Ile Ala Leu Thr Asp Leu Asp
 6435 6440 6445

Gly Ser Glu Gln Phe Val Ser Ile Lys Phe Thr Asp Val Pro Asp Gly
 6450 6455 6460

Phe Gln Met Arg Ala Asp Ala Gly Ser Thr Tyr Thr Val Lys Asn Asn
 6465 6470 6475 6480

Gly Asn Gly Glu Trp Ser Val Gln Leu Pro Gln Ala Ser Gly Leu Ser
 6485 6490 6495

Phe Asp Leu Ser Glu Ile Ser Ile Leu Pro Pro Lys Asn Phe Ser Gly
 6500 6505 6510

Thr Ala Glu Phe Gly Val Glu Val Phe Thr Gln Glu Ser Leu Leu Gly
 6515 6520 6525

Val Pro Thr Ala Ala Ala Asn Leu Pro Ser Phe Lys Leu His Val Val
 6530 6535 6540

Pro Val Gly Asp Asp Val Asp Thr Asn Pro Thr Asp Ser Val Thr Gly
 6545 6550 6555 6560

Asn Glu Gly Gln Asn Ile Asp Ile Glu Ile Asn Ala Thr Ile Leu Asp
 6565 6570 6575

Lys Glu Leu Ser Ala Thr Gly Ser Gly Thr Tyr Thr Glu Asn Ala Pro
 6580 6585 6590

Glu Thr Leu Arg Val Glu Val Ala Gly Val Pro Gln Asp Ala Ser Ile
 6595 6600 6605

Phe Tyr Pro Asp Gly Thr Thr Leu Ala Ser Tyr Asp Pro Ala Thr Gln
 6610 6615 6620

Leu Trp Thr Leu Asp Val Pro Ala Gln Ser Leu Asp Lys Ile Val Phe
 6625 6630 6635 6640

Asn Ser Gly Glu His Asn Ser Asp Thr Gly Asn Val Leu Gly Ile Asn
 6645 6650 6655

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Gly Pro Leu Gln Ile Thr Val Arg Ser Val Asp Thr Asp Ala Asp Asn
 6660 6665 6670
 Thr Glu Tyr Leu Gly Thr Pro Thr Ser Phe Asp Val Asp Leu Val Ile
 6675 6680 6685
 Asp Pro Ile Asn Asp Gln Pro Ile Phe Val Asn Val Thr Asn Ile Glu
 6690 6695 6700
 Thr Ser Glu Asp Ile Ser Val Ala Ile Asp Asn Phe Ser Ile Tyr Asp
 6705 6710 6715 6720
 Val Asp Ala Asn Phe Asp Asn Pro Asp Ala Pro Tyr Glu Leu Thr Leu
 6725 6730 6735
 Lys Val Asp Gln Thr Leu Pro Gly Ala Gln Gly Val Phe Glu Phe Thr
 6740 6745 6750
 Ser Ser Pro Asp Val Thr Phe Val Leu Gln Pro Asp Gly Ser Leu Val
 6755 6760 6765
 Ile Thr Gly Lys Glu Ala Asp Ile Asn Thr Ala Leu Thr Asn Gly Ala
 6770 6775 6780
 Val Thr Phe Lys Pro Asp Pro Asp Gln Asn Tyr Leu Asn Gln Thr Gly
 6785 6790 6795 6800
 Leu Val Thr Ile Asn Ala Thr Leu Asp Asp Gly Gly Asn Asn Gly Leu
 6805 6810 6815
 Ile Asp Ala Val Asp Pro Asn Thr Ala Gln Thr Asn Gln Thr Thr Phe
 6820 6825 6830
 Thr Ile Lys Val Thr Glu Val Asn Asp Ala Pro Val Ala Thr Asn Val
 6835 6840 6845
 Asp Leu Gly Ser Ile Ala Glu Asp Ala Gln Ile Val Ile Val Glu Ser
 6850 6855 6860
 Asp Leu Ile Ala Ala Ser Ser Asp Leu Glu Asn His Asn Leu Thr Val
 6865 6870 6875 6880
 Thr Gly Val Thr Leu Thr Gln Gly Gln Gly Gln Leu Thr Arg Tyr Glu
 6885 6890 6895
 Asn Ala Gly Gly Ala Asp Asp Ala Ala Ile Thr Gly Pro Phe Trp Ile
 6900 6905 6910
 Phe Ile Ala Asp Asn Asp Phe Asn Gly Asp Val Lys Phe Asn Tyr Ser
 6915 6920 6925
 Ile Ile Asp Asp Gly Thr Thr Asn Gly Val Asp Asp Phe Lys Thr Asp
 6930 6935 6940
 Ser Ala Glu Ile Ser Leu Val Val Thr Glu Val Asn Asp Gln Pro Val
 6945 6950 6955 6960
 Ala Ser Asn Ile Asp Leu Gly Thr Met Leu Glu Glu Gly Gln Leu Val
 6965 6970 6975
 Ile Lys Glu Glu Asp Leu Ile Ser Ala Thr Thr Asp Pro Glu Asn Asp
 6980 6985 6990
 Thr Ile Thr Val Asn Ser Leu Val Leu Asp Gln Gly Gln Gly Gln Leu
 6995 7000 7005
 Gln Arg Phe Glu Asn Val Gly Gly Ala Asp Asp Ala Thr Ile Thr Gly
 7010 7015 7020
 Pro Tyr Trp Val Phe Thr Ala Ala Asn Glu Tyr Asn Gly Asp Val Lys
 7025 7030 7035 7040
 Phe Thr Tyr Thr Val Glu Asp Asp Gly Thr Thr Asn Gly Ala Asp Asp
 7045 7050 7055
 Phe Leu Thr Asp Thr Gly Glu Ile Ser Val Val Val Thr Glu Val Asn

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| 7060 | | | | | 7065 | | | | | 7070 | | | | | |
|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|
| Asp | Gln | Pro | Val | Ala | Thr | Asp | Ile | Asp | Leu | Gly | Asn | Ile | Leu | Glu | Glu |
| | 7075 | | | | | | 7080 | | | | | 7085 | | | |
| Gly | Gln | Leu | Ile | Ile | Lys | Glu | Glu | Asp | Leu | Ile | Ala | Ala | Thr | Ser | Asp |
| | 7090 | | | | | 7095 | | | | | 7100 | | | | |
| Pro | Glu | Asn | Asp | Thr | Ile | Thr | Val | Thr | Asn | Leu | Val | Leu | Asp | Glu | Gly |
| | 7105 | | | | | 7110 | | | | | 7115 | | | | 7120 |
| Gln | Gly | Gln | Leu | Gln | Arg | Phe | Glu | Asn | Val | Gly | Gly | Ala | Asp | Asp | Ala |
| | | | | 7125 | | | | | 7130 | | | | | 7135 | |
| Met | Ile | Thr | Gly | Pro | Tyr | Trp | Ile | Phe | Thr | Ala | Ala | Asp | Glu | Tyr | Asn |
| | | | 7140 | | | | | 7145 | | | | | 7150 | | |
| Gly | Asn | Val | Lys | Phe | Thr | Tyr | Thr | Val | Glu | Asp | Asp | Gly | Thr | Thr | Asn |
| | | 7155 | | | | | 7160 | | | | | 7165 | | | |
| Gly | Ala | Asn | Asp | Phe | Leu | Thr | Asp | Thr | Ala | Glu | Ile | Thr | Ala | Ile | Val |
| | 7170 | | | | | 7175 | | | | | 7180 | | | | |
| Asp | Gly | Val | Asn | Asp | Thr | Pro | Val | Val | Asn | Gly | Asp | Ser | Val | Thr | Thr |
| | 7185 | | | | | 7190 | | | | | 7195 | | | | 7200 |
| Ile | Val | Asp | Glu | Asp | Ala | Gly | Gln | Leu | Leu | Ser | Gly | Ile | Asn | Val | Ser |
| | | | | 7205 | | | | | 7210 | | | | | 7215 | |
| Asp | Pro | Asp | Tyr | Val | Asp | Ala | Phe | Ser | Asn | Asp | Leu | Met | Thr | Val | Thr |
| | | | 7220 | | | | 7225 | | | | | | 7230 | | |
| Leu | Thr | Val | Asp | Tyr | Gly | Thr | Leu | Asn | Val | Ser | Leu | Pro | Ala | Val | Thr |
| | | 7235 | | | | | 7240 | | | | | 7245 | | | |
| Thr | Val | Met | Val | Asn | Gly | Asn | Asn | Thr | Gly | Ser | Val | Ile | Leu | Val | Gly |
| | | 7250 | | | | 7255 | | | | | 7260 | | | | |
| Thr | Leu | Ser | Asp | Leu | Asn | Ala | Leu | Ile | Asp | Thr | Pro | Thr | Ser | Pro | Asn |
| | 7265 | | | | | 7270 | | | | | 7275 | | | | 7280 |
| Gly | Val | Tyr | Leu | Asp | Ala | Ser | Leu | Ser | Pro | Thr | Asn | Ser | Ile | Gly | Leu |
| | | | | 7285 | | | | | 7290 | | | | | 7295 | |
| Glu | Val | Ile | Ala | Lys | Asp | Ser | Gly | Asn | Pro | Ser | Gly | Ile | Ala | Ile | Glu |
| | | | 7300 | | | | | 7305 | | | | | 7310 | | |
| Thr | Ala | Pro | Val | Val | Tyr | Asn | Ile | Ala | Val | Thr | Pro | Val | Ala | Asn | Ala |
| | | 7315 | | | | | 7320 | | | | | 7325 | | | |
| Pro | Thr | Leu | Ser | Ile | Asp | Pro | Ala | Phe | Asn | Tyr | Val | Arg | Asn | Ile | Thr |
| | | 7330 | | | | 7335 | | | | | 7340 | | | | |
| Thr | Ser | Ser | Ser | Val | Val | Ala | Asn | Ser | Gly | Val | Ala | Leu | Val | Gly | Ile |
| | | | | 7345 | | 7350 | | | | | 7355 | | | | 7360 |
| Val | Ala | Ala | Leu | Thr | Asp | Ile | Thr | Glu | Glu | Leu | Thr | Leu | Lys | Ile | Ser |
| | | | | 7365 | | | | | 7370 | | | | | 7375 | |
| Asp | Val | Pro | Asp | Gly | Val | Asp | Val | Thr | Ser | Asp | Val | Gly | Thr | Val | Ser |
| | | | 7380 | | | | | 7385 | | | | | 7390 | | |
| Leu | Val | Gly | Asp | Thr | Trp | Ile | Ala | Thr | Ala | Asp | Ala | Ile | Asp | Ser | Leu |
| | | 7395 | | | | | 7400 | | | | | 7405 | | | |
| Arg | Leu | Val | Glu | Gln | Ser | Ser | Leu | Gly | Lys | Pro | Leu | Thr | Pro | Gly | Asn |
| | | 7410 | | | | | 7415 | | | | | 7420 | | | |
| Tyr | Thr | Leu | Lys | Val | Glu | Ala | Leu | Ser | Glu | Glu | Thr | Asp | Asn | Asn | Asp |
| | | | | 7425 | | 7430 | | | | | 7435 | | | | 7440 |
| Ile | Ala | Ile | Ser | Gln | Asn | Ile | Asp | Leu | Asn | Leu | Asn | Ile | Val | Ala | Asn |
| | | | | 7445 | | | | | 7450 | | | | | 7455 | |
| Pro | Ile | Asp | Leu | Asp | Leu | Ser | Ser | Glu | Thr | Asp | Asp | Val | Gln | Leu | Leu |
| | | | 7460 | | | | | 7465 | | | | | 7470 | | |

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Ala Ser Asn Phe Asp Thr Asn Leu Thr Gly Gly Thr Gly Asn Asp Arg
 7475 7480 7485
 Leu Val Gly Gly Ala Gly Asp Asp Thr Leu Val Gly Gly Asp Gly Asn
 7490 7495 7500
 Asp Thr Leu Ile Gly Gly Gly Ser Asp Ile Leu Thr Gly Gly Asn
 7505 7510 7515 7520
 Gly Met Asp Ser Phe Val Trp Leu Asn Ile Glu Asp Gly Val Glu Asp
 7525 7530 7535
 Thr Ile Thr Asp Phe Ser Leu Ser Glu Gly Asp Gln Ile Asp Leu Arg
 7540 7545 7550
 Glu Val Leu Pro Glu Leu Lys Asn Thr Ser Pro Asp Met Ser Ala Leu
 7555 7560 7565
 Leu Gln Gln Ile Asp Ala Lys Val Glu Gly Asp Asp Ile Glu Leu Thr
 7570 7575 7580
 Ile Lys Ser Asp Gly Leu Gly Thr Thr Glu Gln Val Ile Val Val Glu
 7585 7590 7595 7600
 Asp Leu Ala Pro Gln Leu Thr Leu Ser Gly Thr Met Pro Ser Asp Ile
 7605 7610 7615
 Leu Asp Ala Leu Val Gln Gln Asn Val Ile Thr His Gly
 7620 7625

<210> SEQ ID NO 5
 <211> LENGTH: 765
 <212> TYPE: DNA
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 5

```

atgaaaaaaaa catcactatt acttgcttcc attactctgg cactttctgg tgtagtacaa    60
gctgaccagc tagaagacat tcaaaaatca ggcacacttc gcgctcggcac cacagcgac    120
tacaaaacct tttcttactt cgacggcaaa acctactctg gttatgacat tgacgtagcc    180
aaacatggtt cagagcagtt gggcggtgaa ttacagattg ttcgtaccac atggaaagat    240
ctactgaccg atctagacag cgataaatat gacatcgcga tgggcggtat cacgcgtaaa    300
atgcagcgtc agttaaacgc agaacaaact caaggttaca tgaccttgg caagtgtttc    360
ttagttgcga aaggcaaagc agaacaatc aacagcattg agaaagtga cctctcttct    420
gtgctgtgtt gcgtcaatat cgggtgggact aatgagatgt ttgcggatgc taacttgcaa    480
gacgcgagct ttacgcgtta cgagaacaac ctacagcttc cgcaagcgtg tgcggaaggt    540
aaagttgatg taatggtgac agaaactcct gaaggtctgt tctatcaagt gacggacgaa    600
cgtcttgaag cggcacgctg tgaaacaccg tttaccaaca gtcaattcgg ttacctgata    660
ccaaaagggt aacaacgctt gttgaacaca gtgaacttca ttatggatga gatgaaattg    720
aaaggcgtcg aagaagagtt cctgatccac aactctctta agtaa                    765
    
```

<210> SEQ ID NO 6
 <211> LENGTH: 254
 <212> TYPE: PRT
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 6

Met Lys Lys Thr Ser Leu Leu Leu Ala Ser Ile Thr Leu Ala Leu Ser
 1 5 10 15

-continued

Gly Val Val Gln Ala Asp Gln Leu Glu Asp Ile Gln Lys Ser Gly Thr
 20 25 30

Leu Arg Val Gly Thr Thr Gly Asp Tyr Lys Pro Phe Ser Tyr Phe Asp
 35 40 45

Gly Lys Thr Tyr Ser Gly Tyr Asp Ile Asp Val Ala Lys His Val Ala
 50 55 60

Glu Gln Leu Gly Val Glu Leu Gln Ile Val Arg Thr Thr Trp Lys Asp
 65 70 75 80

Leu Leu Thr Asp Leu Asp Ser Asp Lys Tyr Asp Ile Ala Met Gly Gly
 85 90 95

Ile Thr Arg Lys Met Gln Arg Gln Leu Asn Ala Glu Gln Thr Gln Gly
 100 105 110

Tyr Met Thr Phe Gly Lys Cys Phe Leu Val Ala Lys Gly Lys Ala Glu
 115 120 125

Gln Tyr Asn Ser Ile Glu Lys Val Asn Leu Ser Ser Val Arg Val Gly
 130 135 140

Val Asn Ile Gly Gly Thr Asn Glu Met Phe Ala Asp Ala Asn Leu Gln
 145 150 155 160

Asp Ala Ser Phe Thr Arg Tyr Glu Asn Asn Leu Asp Val Pro Gln Ala
 165 170 175

Val Ala Glu Gly Lys Val Asp Val Met Val Thr Glu Thr Pro Glu Gly
 180 185 190

Leu Phe Tyr Gln Val Thr Asp Glu Arg Leu Glu Ala Ala Arg Cys Glu
 195 200 205

Thr Pro Phe Thr Asn Ser Gln Phe Gly Tyr Leu Ile Pro Lys Gly Glu
 210 215 220

Gln Arg Leu Leu Asn Thr Val Asn Phe Ile Met Asp Glu Met Lys Leu
 225 230 235 240

Lys Gly Val Glu Glu Glu Phe Leu Ile His Asn Ser Leu Lys
 245 250

<210> SEQ ID NO 7
 <211> LENGTH: 765
 <212> TYPE: DNA
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 7

```

atgaaaaaaaa catcactatt acttgcttcc attactctgg cactttctgg tgtagtacaa      60
gctgaccagc tagaagacat tcaaaaatca ggcacacttc gcgtcggcac cacaggcgac      120
tacaaacctt tttcttactt cgacggcaaa acctactctg gttatgacat tgacgtagcc      180
aaacatgttg cagagcagtt gggcgttgaa ttacagattg ttcgtaccac atggaaagat      240
ctactgaccg atctagacag cgataaatc gacatcgcga tgggcggtat cacgcgtaaa      300
atgcagcgtc agttaaacgc agaacaaact caaggttaca tgacctttgg caagtgtttc      360
ttagttgcga aaggcaaagc agaacaatac aacagcattg agaaagtga cctctcttct      420
gtgctgtggt gcgtcaatat cgggtgggact aatgagatgt ttgcggatgc taacttgcaa      480
gacgcgagct ttacgcggtta cgagaacaac ctacagcttc cgcaagccgt tgcggaaggt      540
aaagttgatg taatggtgac agaaactcct gaaggtctgt tctatcaagt gacggacgaa      600
cgtcttgaag cggcacgctg tgaaacaccc tttaccaaca gtcaattcgg ttacctgata      660
ccaaaagggtg aacaacgctt gttgaacaca gtgaacttca ttatggatga gatgaaattg      720
  
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-continued

aaaggcgtcg aagaagagtt cctgatccac aactctctta agtaa

765

<210> SEQ ID NO 8

<211> LENGTH: 588

<212> TYPE: PRT

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 8

Met Thr Ile Asp Thr Phe Val Val Leu Ala Tyr Phe Phe Phe Leu Ile
 1 5 10 15

Ala Ile Gly Trp Met Phe Arg Lys Phe Thr Thr Ser Thr Ser Asp Tyr
 20 25 30

Phe Arg Gly Gly Gly Lys Met Leu Trp Trp Met Val Gly Ala Thr Ala
 35 40 45

Phe Met Thr Gln Phe Ser Ala Trp Thr Phe Thr Gly Ala Ala Gly Arg
 50 55 60

Ala Phe Asn Asp Gly Phe Val Ile Val Ile Leu Phe Leu Ala Asn Ala
 65 70 75 80

Phe Gly Tyr Phe Met Asn Tyr Met Tyr Phe Ala Pro Lys Phe Arg Gln
 85 90 95

Leu Arg Val Val Thr Ala Ile Glu Ala Ile Arg Gln Arg Phe Gly Lys
 100 105 110

Thr Ser Glu Gln Phe Phe Thr Trp Ala Gly Met Pro Asp Ser Leu Ile
 115 120 125

Ser Ala Gly Ile Trp Leu Asn Gly Leu Ala Ile Phe Val Ala Ala Val
 130 135 140

Phe Asn Ile Pro Met Glu Ala Thr Ile Val Val Thr Gly Met Val Leu
 145 150 155 160

Val Leu Met Ala Val Thr Gly Gly Ser Trp Ala Val Val Ala Ser Asp
 165 170 175

Phe Met Gln Met Leu Val Ile Met Ala Val Thr Ile Thr Cys Ala Val
 180 185 190

Ala Ala Tyr Phe His Gly Gly Gly Leu Thr Asn Ile Val Ala Asn Phe
 195 200 205

Asp Gly Asp Phe Met Leu Gly Asn Asn Leu Asn Tyr Met Ser Ile Phe
 210 215 220

Val Leu Trp Val Val Phe Ile Phe Val Lys Gln Phe Gly Val Met Asn
 225 230 235 240

Asn Ser Ile Asn Ala Tyr Arg Tyr Leu Cys Ala Lys Asp Ser Glu Asn
 245 250 255

Ala Arg Lys Ala Ala Gly Leu Ala Cys Ile Leu Met Val Val Gly Pro
 260 265 270

Leu Ile Trp Phe Leu Pro Pro Trp Tyr Val Ser Ala Phe Met Pro Asp
 275 280 285

Phe Ala Leu Glu Tyr Ala Ser Met Gly Asp Lys Ala Gly Asp Ala Ala
 290 295 300

Tyr Leu Ala Phe Val Gln Asn Val Met Pro Ala Gly Met Val Gly Leu
 305 310 315 320

Leu Met Ser Ala Met Phe Ala Ala Thr Met Ser Ser Met Asp Ser Gly
 325 330 335

Leu Asn Arg Asn Ala Gly Ile Phe Val Met Asn Phe Tyr Ser Pro Ile
 340 345 350

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Leu Arg Gln Asn Ala Thr Gln Lys Glu Leu Val Ile Val Ser Lys Leu
 355 360 365
 Thr Thr Ile Met Met Gly Ile Ile Ile Ala Ile Gly Leu Phe Ile
 370 375 380
 Asn Ser Leu Arg His Leu Ser Leu Phe Asp Ile Val Met Asn Val Gly
 385 390 395 400
 Ala Leu Ile Gly Phe Pro Met Leu Ile Pro Val Leu Leu Gly Met Trp
 405 410 415
 Ile Arg Lys Thr Pro Asp Trp Ala Gly Trp Ser Thr Leu Ile Val Gly
 420 425 430
 Gly Phe Val Ser Tyr Ile Phe Gly Ile Ser Leu Gln Ala Glu Asp Ile
 435 440 445
 Glu His Leu Phe Gly Met Glu Thr Ala Leu Thr Gly Arg Glu Trp Ser
 450 455 460
 Asp Leu Lys Val Gly Leu Ser Leu Ala Ala His Val Val Phe Thr Gly
 465 470 475 480
 Gly Tyr Phe Ile Leu Thr Ser Arg Phe Tyr Lys Gly Leu Ser Pro Glu
 485 490 495
 Arg Glu Lys Glu Val Asp Gln Leu Phe Thr Asn Trp Asn Thr Pro Leu
 500 505 510
 Val Ala Glu Gly Glu Glu Gln Gln Asn Leu Asp Thr Lys Gln Arg Ser
 515 520 525
 Met Leu Gly Lys Leu Ile Ser Thr Ala Gly Phe Gly Ile Leu Ala Met
 530 535 540
 Ala Leu Ile Pro Asn Glu Pro Thr Gly Arg Leu Leu Phe Leu Leu Cys
 545 550 555 560
 Gly Ser Met Val Leu Thr Val Gly Ile Leu Val Asn Ala Ser Lys
 565 570 575
 Ala Pro Ala Lys Met Asn Asn Glu Ser Val Ala Lys
 580 585

<210> SEQ ID NO 9

<211> LENGTH: 627

<212> TYPE: DNA

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 9

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atgacgacat taaatgaaca actagcaaac ctaaaagtaa ttcctgtaat cgcgatcaac   60
cgtgctgaag acgctatccc tctaggtaaa gcgctggttg aaaatggcat gccatgtgca  120
gaaattacac tacgtacaga atgtgcaatc gaagcgattc gcatcatgcg taaagaattc   180
ccagacatgc taatcggttc aggtactgta ctgactaacg agcaagttga cgcactatc   240
gaagctggty ttgatttcat cgtaagccca ggttttaacc cacgtactgt tcaatactgt   300
atcgataaag gtattgcaat cgtaccgggt gttaacaacc caagcctagt tgagcaagca   360
atggaaatgg gtcttcgca cgttgaagttc ttcctgctg agccttcagg cggactaggc   420
atgcttaaag cactaacagc agtttaccct gttaaattca tgcctactgg tggcgtaagc   480
ttgaagaatg ttgatgaata cctatogac ccttctgttc ttgcgtgtgg cggactaggc   540
atggttccaa ctaaccttat cgatgaaggt aagtgggacg aactaggcaa gcttgttcgt   600
gacgcagttg atcacgttaa cgcttaa

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<210> SEQ ID NO 10
 <211> LENGTH: 208
 <212> TYPE: PRT
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 10

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Met Thr Thr Leu Asn Glu Gln Leu Ala Asn Leu Lys Val Ile Pro Val
1          5          10          15

Ile Ala Ile Asn Arg Ala Glu Asp Ala Ile Pro Leu Gly Lys Ala Leu
20          25          30

Val Glu Asn Gly Met Pro Cys Ala Glu Ile Thr Leu Arg Thr Glu Cys
35          40          45

Ala Ile Glu Ala Ile Arg Ile Met Arg Lys Glu Phe Pro Asp Met Leu
50          55          60

Ile Gly Ser Gly Thr Val Leu Thr Asn Glu Gln Val Asp Ala Ser Ile
65          70          75          80

Glu Ala Gly Val Asp Phe Ile Val Ser Pro Gly Phe Asn Pro Arg Thr
85          90          95

Val Gln Tyr Cys Ile Asp Lys Gly Ile Ala Ile Val Pro Gly Val Asn
100         105         110

Asn Pro Ser Leu Val Glu Gln Ala Met Glu Met Gly Leu Arg Thr Leu
115         120         125

Lys Phe Phe Pro Ala Glu Pro Ser Gly Gly Thr Gly Met Leu Lys Ala
130         135         140

Leu Thr Ala Val Tyr Pro Val Lys Phe Met Pro Thr Gly Gly Val Ser
145         150         155         160

Leu Lys Asn Val Asp Glu Tyr Leu Ser Ile Pro Ser Val Leu Ala Cys
165         170         175

Gly Gly Thr Trp Met Val Pro Thr Asn Leu Ile Asp Glu Gly Lys Trp
180         185         190

Asp Glu Leu Gly Lys Leu Val Arg Asp Ala Val Asp His Val Asn Ala
195         200         205

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<210> SEQ ID NO 11
 <211> LENGTH: 933
 <212> TYPE: DNA
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 11

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atgaaatcat taaacatcgc ggtcattggc gagtgcattg ttgagctaca aaagaaacaa    60
gacgggctta agcaaagttt tgggtggcgat acgctgaata ctgcacttta cttgtcacgc    120
ttaacaaaag agcaagatat caacacgagc tacgtaactg cactaggcac tgaccattc    180
agtaccgaca tgttaaaaaa ttggcaagcg gaaggtatcg acacgagctt aattgctcag    240
ctggaccaca aacaaccagg gctttactac atcgagaccg atgaaactgg tgaacgcagt    300
ttccactact ggcgtagtga tgctgcagcg aagttcatgt ttgatcagga agacacgcct    360
gctcttcttg ataagctggt ctcttttgac gcgatttact taagtggat tacgctggca    420
atcttgacag aaaatggtcg cacgcagcta ttcaacttct tagacaaatt caaagtcaa    480
ggcggccaag tattcttcga caataactac cgacctaac tttgggaaag ccaacaagaa    540
gcgatttctt ggtacttgaa aatgcttaag tacacagata cggetctgct gacgtttgat    600
gatgagcaag agctatacgg cgacgaaagc attgaacaat gtattacag tacgtcagag    660

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tctggtgtga aagagatcgt cattaacgt ggcgcgaaag actgcttagt ggttgaaagc 720
caaagcgctc aatacgttgc acccaaccct gtagacaaca tcgttgatac gactgccgct 780
ggcgactcgt tcagtgccagg cttcttgccc aagcgcttga gcggcggttag tgctcgtgat 840
gctgcatttg caggtcatat tgtggcagga accgtgattc agcatccagg tgctatcatt 900
cctctagaag cgacgcctga tctgtctcta taa 933

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<210> SEQ ID NO 12
<211> LENGTH: 310
<212> TYPE: PRT
<213> ORGANISM: Vibrio splendidus

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<400> SEQUENCE: 12

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Met Lys Ser Leu Asn Ile Ala Val Ile Gly Glu Cys Met Val Glu Leu
1           5           10           15
Gln Lys Lys Gln Asp Gly Leu Lys Gln Ser Phe Gly Gly Asp Thr Leu
20          25          30
Asn Thr Ala Leu Tyr Leu Ser Arg Leu Thr Lys Glu Gln Asp Ile Asn
35          40          45
Thr Ser Tyr Val Thr Ala Leu Gly Thr Asp Pro Phe Ser Thr Asp Met
50          55          60
Leu Lys Asn Trp Gln Ala Glu Gly Ile Asp Thr Ser Leu Ile Ala Gln
65          70          75          80
Leu Asp His Lys Gln Pro Gly Leu Tyr Tyr Ile Glu Thr Asp Glu Thr
85          90          95
Gly Glu Arg Ser Phe His Tyr Trp Arg Ser Asp Ala Ala Ala Lys Phe
100         105         110
Met Phe Asp Gln Glu Asp Thr Pro Ala Leu Leu Asp Lys Leu Phe Ser
115         120         125
Phe Asp Ala Ile Tyr Leu Ser Gly Ile Thr Leu Ala Ile Leu Thr Glu
130         135         140
Asn Gly Arg Thr Gln Leu Phe Asn Phe Leu Asp Lys Phe Lys Ala Gln
145         150         155         160
Gly Gly Gln Val Phe Phe Asp Asn Asn Tyr Arg Pro Lys Leu Trp Glu
165         170         175
Ser Gln Gln Glu Ala Ile Ser Trp Tyr Leu Lys Met Leu Lys Tyr Thr
180         185         190
Asp Thr Ala Leu Leu Thr Phe Asp Asp Glu Gln Glu Leu Tyr Gly Asp
195         200         205
Glu Ser Ile Glu Gln Cys Ile Thr Arg Thr Ser Glu Ser Gly Val Lys
210         215         220
Glu Ile Val Ile Lys Arg Gly Ala Lys Asp Cys Leu Val Val Glu Ser
225         230         235         240
Gln Ser Ala Gln Tyr Val Ala Pro Asn Pro Val Asp Asn Ile Val Asp
245         250         255
Thr Thr Ala Ala Gly Asp Ser Phe Ser Ala Gly Phe Leu Ala Lys Arg
260         265         270
Leu Ser Gly Gly Ser Ala Arg Asp Ala Ala Phe Ala Gly His Ile Val
275         280         285
Ala Gly Thr Val Ile Gln His Pro Gly Ala Ile Ile Pro Leu Glu Ala
290         295         300

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Thr Pro Asp Leu Ser Leu
305 310

<210> SEQ ID NO 13
<211> LENGTH: 336
<212> TYPE: DNA
<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 13

atgaactcct tctttatcct agatgaaaat ccatgggaag aacttggtgg cgcattaag 60
cgtaaaatcg ttgcttacac tgacgatcta atggcagtac acctatgctt tgataagggc 120
gcgattggcc accctcatac tcacgaaatt cagcaccaaa tcggttatgt tgttcgtggt 180
agcttcgaag ctgaaatcga cggcgagaag aaagtgctta aagaaggcga tgcttacttc 240
gctcgtaaac acatgatgca cgggtcagtt gctctagaac aagacagcat ccttcttgat 300
atcttcaatc ctgcgctgta agatttccta aaataa 336

<210> SEQ ID NO 14
<211> LENGTH: 111
<212> TYPE: PRT
<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 14

Met Asn Ser Phe Phe Ile Leu Asp Glu Asn Pro Trp Glu Glu Leu Gly
1 5 10 15
Gly Gly Ile Lys Arg Lys Ile Val Ala Tyr Thr Asp Asp Leu Met Ala
20 25 30
Val His Leu Cys Phe Asp Lys Gly Ala Ile Gly His Pro His Thr His
35 40 45
Glu Ile His Asp Gln Ile Gly Tyr Val Val Arg Gly Ser Phe Glu Ala
50 55 60
Glu Ile Asp Gly Glu Lys Lys Val Leu Lys Glu Gly Asp Ala Tyr Phe
65 70 75 80
Ala Arg Lys His Met Met His Gly Ala Val Ala Leu Glu Gln Asp Ser
85 90 95
Ile Leu Leu Asp Ile Phe Asn Pro Ala Arg Glu Asp Phe Leu Lys
100 105 110

<210> SEQ ID NO 15
<211> LENGTH: 2208
<212> TYPE: DNA
<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 15

atgacgacta aaccagtatt gttgactgaa gotgaaatcg aacagcttca tcttgaagtg 60
ggccgttcta gcttaatggg caaaaccatt gcagcgaacg cgaaagacct agaagcattc 120
atgcttttac ctattgatgt tccaggtcac ggtgaagctg ggggttacga acataaccgc 180
cacaagcaaa attacacgta catgaaccta gotggtcgca tgttcttgat cactaaagag 240
caaaaatcag ctgactttgt tacagaatta ctagaagagt acgcagacaa atatctaacy 300
tttgattacc acgtacagaa aaacaccaac ccaacaggtc gtttgttoca ccaaatccta 360
aacgaacact getggttaat gttctcaagc ttagcttatt cttgtgttgc ttcaacactg 420
acacaagatc agcgtgacaa tattgagtct cgcatttttg aacctatgct agaaatgttc 480

-continued

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acggttaaat acgcacacga cttcgaccgt attcacaatc acggtatttg ggcagtagcc 540
gctgtgggta tctgtggtct tgctttaggc aaacgtgaat acctagaaat gtcagtgtac 600
ggcatcgacc gtaatgatac tggcgggttc ctacgcgaag tttctcagct atttgcacct 660
tctggctact acatggaagg tccttactac catcgttatg cgattcgccc aacgtgtgtg 720
ttcgctgaag tgattcaccg tcatatgcoct gaagttgata tctacaacta caaaggcggc 780
gtgattggta acacagtaca agctatgctt gcgacagcgt acccgaacgg cgagttcccc 840
gctctgaatg atgcttctcg tactatgggt atcacagaca tgggtgttca ggttgcggtc 900
agtgtttaca gtaagcatta ctcttctgaa aacgggttag accaaaacat tctgggtatg 960
gcgaagattc aagacgcagt atggatgcat ccatgtggtc ttgagctatc taaagcatac 1020
gaagccgcac ctgcagagaa agaaatcggc atgcctttct ggccaagtgt tgaattgaat 1080
gaaggccctc aaggtcacia cggcgcgcaa ggctttatcc gtatgcagga taagaaaggc 1140
gacgtttctc aacttgtgat gaactacggc caacacggca tgggtcacgg caactttgat 1200
acgctgggta tttctttctt taaccgcggg caagaagtgc tacgtgaata cggcttctgt 1260
cgttgggta acgttgagcc aaaattcggc ggccgttacc tagacgaaaa caaatcttac 1320
gctcgtcaaa cgattgctca caatgcagtt acgattgatg aaaaatgtca gaacaacttt 1380
gacgttgaac gtgcagactc agtacatggt ttacctcact tctttaaagt agaagacgat 1440
caaatcaacg gtatgagtgc atttgctaac gatcattacc aaggcttga catgcaacgc 1500
agcgtgttca tgctaaatct tgaagaatta gaatctcctg tattgttaga cctataccgc 1560
ttagattcta caaaaggcgg cgaaggcgag caccaatacg actattcaca ccaatatgcg 1620
ggtcagattg ttgcactaa cttcgaatac caagcgaaca aagagctaaa cactctaggt 1680
gacgatttcg gttaccaaca tctatggaac gtcgcaagcg gtgaagtga gggcacagca 1740
attgtaagtt ggctacaaaa caacacctac tacacatggc taggtgcaac gtctaacgat 1800
aatgctgaag taatatttac tcgcactggc gctaacgacc caagtttcaa tctacgttca 1860
gagcctgcgt tcattctacg cagcaaaagg gaaacaacac tgtttgcttc tgttgtgaa 1920
acgcacgggt atttcaacga agaattcgag caatctgtca atgcacgtgg tgttgtgaa 1980
gacatcaaaq tcgtggctca caccaatgtc ggttcggtag ttgagatcac cacagagaaa 2040
tcaaactgta cagtgatgat cagcaaccaa cttggcgcga ctgacagcac tgaacacaaa 2100
gtagaactga acggcaaagt atacagctgg aaaggcttct actcagtaga gacaacttta 2160
caagaaacga attcagaaga acttagcact gcagggcagg gaaaataa 2208

```

<210> SEQ ID NO 16

<211> LENGTH: 735

<212> TYPE: PRT

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 16

```

Met Thr Thr Lys Pro Val Leu Leu Thr Glu Ala Glu Ile Glu Gln Leu
1           5           10           15

```

```

His Leu Glu Val Gly Arg Ser Ser Leu Met Gly Lys Thr Ile Ala Ala
20           25           30

```

```

Asn Ala Lys Asp Leu Glu Ala Phe Met Arg Leu Pro Ile Asp Val Pro
35           40           45

```

```

Gly His Gly Glu Ala Gly Gly Tyr Glu His Asn Arg His Lys Gln Asn

```


-continued

Ala Asp Ser Val His Gly Leu Pro His Phe Phe Lys Val Glu Asp Asp
465 470 475 480

Gln Ile Asn Gly Met Ser Ala Phe Ala Asn Asp His Tyr Gln Gly Phe
485 490 495

Asp Met Gln Arg Ser Val Phe Met Leu Asn Leu Glu Glu Leu Glu Ser
500 505 510

Pro Leu Leu Leu Asp Leu Tyr Arg Leu Asp Ser Thr Lys Gly Gly Glu
515 520 525

Gly Glu His Gln Tyr Asp Tyr Ser His Gln Tyr Ala Gly Gln Ile Val
530 535 540

Arg Thr Asn Phe Glu Tyr Gln Ala Asn Lys Glu Leu Asn Thr Leu Gly
545 550 555 560

Asp Asp Phe Gly Tyr Gln His Leu Trp Asn Val Ala Ser Gly Glu Val
565 570 575

Lys Gly Thr Ala Ile Val Ser Trp Leu Gln Asn Asn Thr Tyr Tyr Thr
580 585 590

Trp Leu Gly Ala Thr Ser Asn Asp Asn Ala Glu Val Ile Phe Thr Arg
595 600 605

Thr Gly Ala Asn Asp Pro Ser Phe Asn Leu Arg Ser Glu Pro Ala Phe
610 615 620

Ile Leu Arg Ser Lys Gly Glu Thr Thr Leu Phe Ala Ser Val Val Glu
625 630 635 640

Thr His Gly Tyr Phe Asn Glu Glu Phe Glu Gln Ser Val Asn Ala Arg
645 650 655

Gly Val Val Lys Asp Ile Lys Val Val Ala His Thr Asn Val Gly Ser
660 665 670

Val Val Glu Ile Thr Thr Glu Lys Ser Asn Val Thr Val Met Ile Ser
675 680 685

Asn Gln Leu Gly Ala Thr Asp Ser Thr Glu His Lys Val Glu Leu Asn
690 695 700

Gly Lys Val Tyr Ser Trp Lys Gly Phe Tyr Ser Val Glu Thr Thr Leu
705 710 715 720

Gln Glu Thr Asn Ser Glu Glu Leu Ser Thr Ala Gly Gln Gly Lys
725 730 735

<210> SEQ ID NO 17

<211> LENGTH: 2154

<212> TYPE: DNA

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 17

```

atgagctatc aaccactttt acttaacttt gatgaagcag ctgaacttcg taaagaactt    60
ggcaaggata gcctattagg taacgcactg actcgcgaca ttaacaaac tgacgcttac    120
atggctgaag ttggcattga agtaccaggt cacggtgaag gcgcggtta cgagcacaac    180
cgtcataagc aaaactacat ccatatggat ctagcaggcc gtttgttct tactactgag    240
gaaacaaaaa accgagatta catcgttgat atgctaacag cgtacgcgac ggtataccca    300
acacttgaaa gcaacgtaag cegtgactct aacctcegg gtaagctggt ccaccaaacg    360
ttgaacgaga acatgtggat gctttacgct tcttgtgcgt acagctgcat ctaccacacg    420
atctctgaag agcaaaagcg tctgatcgaa gacgatcttc ttaagcaaat gatcgaaatg    480

```

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```

ttcgttgga cttacgcaca cgacttcgat atcgtacaca accacggcctt atgggcagtg 540
gcagcagtag gtatctgtgg ttacgcaatc aacgatcaag agtctgtaga caaagcacta 600
tacggcctga aactagacaa agtcagcggc ggtttcttag cgcaactaga ccaactgttt 660
tcgccagacg gctactacat ggaaggtcct tactaccacc gtttctctct gcgtccaatc 720
tacctgttcg cagaagcgat tgaacgtcgt cagcctgaag ttggtatcta tgaattcaac 780
gattcagtag tcaagacaac gtcttactct gtattcaaaa cggcattccc agacggtaca 840
ttgctgctc tgaacgattc atcgaagaca atctctatca acgatgaagg cgttatcatg 900
gcaacgtctg tgtgttacca ccggttacgag caaactgaaa ctctacttgg tatggctaac 960
caccagcaaa acgtttgggt tcatgcttca ggtaaaacac tgtctgacgc ggttgatgca 1020
gcagacgaca tcaaagcatt caactggggg agcctgtttg taaccgacgg cctgaagggc 1080
gaaaaaggcg gcgtaagcat ccttcgtcac cgtgacgaac aagatgacga cacgatggcg 1140
ttgatctggt ttggtaacaa cggttctgat caccagtacc actctgctct agaccacggt 1200
cactacgatg gctgcacct aagcgtatct aaccgtggcc acgaagtgct gcacgatttc 1260
ggcttcggtc gctgggtaaa cgttgagcct aagtttgccg gtcgttacat cccagagaac 1320
aagtcttact gtaagcagac gggtgctcac aacacagtaa cggttgatca gaaaacgcag 1380
aacaacttca acacagcatt ggctgagctc aagtttggtc agaagcactt cttcgtagca 1440
gacgaccagt ctctacaagg catgagcggc acaatttctg agtactacac tggcgtagac 1500
atgcaacgca gcgtgattct tgctgaactt cctgagttcg agaagccact tgtaatcgac 1560
gtataccgca tcgaagctga cgctgaacac cagtacgacc tacccgttca ccactctggt 1620
cagatcatcc gtactgactt cgattacaac atggaaaaaa cgcttaagcc gctaggtgaa 1680
gacaacgggt acccagcact atggaacgtg gcttcaggca aagtgaacga agaaggttct 1740
ctagtaagct ggctacatga cagcagctac tacagcctag taaccagcgc gaatgcgggc 1800
agcgaagtga tttttgctcg cactggtgct aacgatccag acttcaacct taagagttag 1860
cctgcgttca tcttacgtca gtctggtcaa aaccacgtgt ttgcttctgt actagaaacg 1920
catggttact ttaacgagtc tatcgaagcc tctgtaggag ctcgtggtct agttaaata 1980
gtatctggtg tgggccataa cagtgtcggg actgtgttcc gcattcagac tacttctggc 2040
aacacttacc actacggtat ctcaaacc aa gctgaagaca cgcagcaagc aactcacat 2100
gttgagttcg cgggtgagac atactcgtgg gaaggatcat ttgctcaact gtaa 2154

```

<210> SEQ ID NO 18

<211> LENGTH: 717

<212> TYPE: PRT

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 18

```

Met Ser Tyr Gln Pro Leu Leu Leu Asn Phe Asp Glu Ala Ala Glu Leu
1           5           10           15
Arg Lys Glu Leu Gly Lys Asp Ser Leu Leu Gly Asn Ala Leu Thr Arg
20          25          30
Asp Ile Lys Gln Thr Asp Ala Tyr Met Ala Glu Val Gly Ile Glu Val
35          40          45
Pro Gly His Gly Glu Gly Gly Tyr Glu His Asn Arg His Lys Gln
50          55          60

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| | | | | | | | | | | | | | | | |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Asn | Tyr | Ile | His | Met | Asp | Leu | Ala | Gly | Arg | Leu | Phe | Leu | Ile | Thr | Glu |
| 65 | | | | | 70 | | | | | 75 | | | | | 80 |
| Glu | Thr | Lys | Tyr | Arg | Asp | Tyr | Ile | Val | Asp | Met | Leu | Thr | Ala | Tyr | Ala |
| | | | | 85 | | | | | 90 | | | | | 95 | |
| Thr | Val | Tyr | Pro | Thr | Leu | Glu | Ser | Asn | Val | Ser | Arg | Asp | Ser | Asn | Pro |
| | | | 100 | | | | | 105 | | | | | 110 | | |
| Pro | Gly | Lys | Leu | Phe | His | Gln | Thr | Leu | Asn | Glu | Asn | Met | Trp | Met | Leu |
| | | 115 | | | | | 120 | | | | | 125 | | | |
| Tyr | Ala | Ser | Cys | Ala | Tyr | Ser | Cys | Ile | Tyr | His | Thr | Ile | Ser | Glu | Glu |
| | 130 | | | | | 135 | | | | | 140 | | | | |
| Gln | Lys | Arg | Leu | Ile | Glu | Asp | Asp | Leu | Leu | Lys | Gln | Met | Ile | Glu | Met |
| 145 | | | | | 150 | | | | | 155 | | | | 160 | |
| Phe | Val | Val | Thr | Tyr | Ala | His | Asp | Phe | Asp | Ile | Val | His | Asn | His | Gly |
| | | | | 165 | | | | | 170 | | | | | 175 | |
| Leu | Trp | Ala | Val | Ala | Ala | Val | Gly | Ile | Cys | Gly | Tyr | Ala | Ile | Asn | Asp |
| | | 180 | | | | | 185 | | | | | | 190 | | |
| Gln | Glu | Ser | Val | Asp | Lys | Ala | Leu | Tyr | Gly | Leu | Lys | Leu | Asp | Lys | Val |
| | | 195 | | | | | 200 | | | | | 205 | | | |
| Ser | Gly | Gly | Phe | Leu | Ala | Gln | Leu | Asp | Gln | Leu | Phe | Ser | Pro | Asp | Gly |
| | 210 | | | | 215 | | | | | | 220 | | | | |
| Tyr | Tyr | Met | Glu | Gly | Pro | Tyr | Tyr | His | Arg | Phe | Ser | Leu | Arg | Pro | Ile |
| 225 | | | | | 230 | | | | | 235 | | | | | 240 |
| Tyr | Leu | Phe | Ala | Glu | Ala | Ile | Glu | Arg | Arg | Gln | Pro | Glu | Val | Gly | Ile |
| | | | 245 | | | | | | 250 | | | | | 255 | |
| Tyr | Glu | Phe | Asn | Asp | Ser | Val | Ile | Lys | Thr | Thr | Ser | Tyr | Ser | Val | Phe |
| | | | 260 | | | | | 265 | | | | | 270 | | |
| Lys | Thr | Ala | Phe | Pro | Asp | Gly | Thr | Leu | Pro | Ala | Leu | Asn | Asp | Ser | Ser |
| | | 275 | | | | | 280 | | | | | 285 | | | |
| Lys | Thr | Ile | Ser | Ile | Asn | Asp | Glu | Gly | Val | Ile | Met | Ala | Thr | Ser | Val |
| | 290 | | | | 295 | | | | | | 300 | | | | |
| Cys | Tyr | His | Arg | Tyr | Glu | Gln | Thr | Glu | Thr | Leu | Leu | Gly | Met | Ala | Asn |
| 305 | | | | | 310 | | | | | 315 | | | | | 320 |
| His | Gln | Gln | Asn | Val | Trp | Val | His | Ala | Ser | Gly | Lys | Thr | Leu | Ser | Asp |
| | | | | 325 | | | | | 330 | | | | | 335 | |
| Ala | Val | Asp | Ala | Ala | Asp | Asp | Ile | Lys | Ala | Phe | Asn | Trp | Gly | Ser | Leu |
| | | | 340 | | | | | 345 | | | | | 350 | | |
| Phe | Val | Thr | Asp | Gly | Pro | Glu | Gly | Glu | Lys | Gly | Gly | Val | Ser | Ile | Leu |
| | | 355 | | | | | 360 | | | | | 365 | | | |
| Arg | His | Arg | Asp | Glu | Gln | Asp | Asp | Asp | Thr | Met | Ala | Leu | Ile | Trp | Phe |
| | 370 | | | | 375 | | | | | | 380 | | | | |
| Gly | Gln | His | Gly | Ser | Asp | His | Gln | Tyr | His | Ser | Ala | Leu | Asp | His | Gly |
| 385 | | | | | 390 | | | | | 395 | | | | | 400 |
| His | Tyr | Asp | Gly | Leu | His | Leu | Ser | Val | Phe | Asn | Arg | Gly | His | Glu | Val |
| | | | | 405 | | | | | 410 | | | | | 415 | |
| Leu | His | Asp | Phe | Gly | Phe | Gly | Arg | Trp | Val | Asn | Val | Glu | Pro | Lys | Phe |
| | | 420 | | | | | | 425 | | | | | 430 | | |
| Gly | Gly | Arg | Tyr | Ile | Pro | Glu | Asn | Lys | Ser | Tyr | Cys | Lys | Gln | Thr | Val |
| | | 435 | | | | | 440 | | | | | 445 | | | |
| Ala | His | Asn | Thr | Val | Thr | Val | Asp | Gln | Lys | Thr | Gln | Asn | Asn | Phe | Asn |
| | 450 | | | | | 455 | | | | | 460 | | | | |
| Thr | Ala | Leu | Ala | Glu | Ser | Lys | Phe | Gly | Gln | Lys | His | Phe | Phe | Val | Ala |

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| | | | | | | | | | | | | | | | |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| 465 | | | | 470 | | | | | 475 | | | | 480 | | |
| Asp | Asp | Gln | Ser | Leu | Gln | Gly | Met | Ser | Gly | Thr | Ile | Ser | Glu | Tyr | Tyr |
| | | | | 485 | | | | | 490 | | | | 495 | | |
| Thr | Gly | Val | Asp | Met | Gln | Arg | Ser | Val | Ile | Leu | Ala | Glu | Leu | Pro | Glu |
| | | | 500 | | | | | 505 | | | | | 510 | | |
| Phe | Glu | Lys | Pro | Leu | Val | Ile | Asp | Val | Tyr | Arg | Ile | Glu | Ala | Asp | Ala |
| | | 515 | | | | | 520 | | | | | 525 | | | |
| Glu | His | Gln | Tyr | Asp | Leu | Pro | Val | His | His | Ser | Gly | Gln | Ile | Ile | Arg |
| | 530 | | | | | | 535 | | | | 540 | | | | |
| Thr | Asp | Phe | Asp | Tyr | Asn | Met | Glu | Lys | Thr | Leu | Lys | Pro | Leu | Gly | Glu |
| 545 | | | | | 550 | | | | | 555 | | | | | 560 |
| Asp | Asn | Gly | Tyr | Gln | His | Leu | Trp | Asn | Val | Ala | Ser | Gly | Lys | Val | Asn |
| | | | | 565 | | | | | 570 | | | | | 575 | |
| Glu | Glu | Gly | Ser | Leu | Val | Ser | Trp | Leu | His | Asp | Ser | Ser | Tyr | Tyr | Ser |
| | | | 580 | | | | | 585 | | | | | 590 | | |
| Leu | Val | Thr | Ser | Ala | Asn | Ala | Gly | Ser | Glu | Val | Ile | Phe | Ala | Arg | Thr |
| | | 595 | | | | | 600 | | | | | 605 | | | |
| Gly | Ala | Asn | Asp | Pro | Asp | Phe | Asn | Leu | Lys | Ser | Glu | Pro | Ala | Phe | Ile |
| | 610 | | | | | 615 | | | | | 620 | | | | |
| Leu | Arg | Gln | Ser | Gly | Gln | Asn | His | Val | Phe | Ala | Ser | Val | Leu | Glu | Thr |
| 625 | | | | | 630 | | | | 635 | | | | | | 640 |
| His | Gly | Tyr | Phe | Asn | Glu | Ser | Ile | Glu | Ala | Ser | Val | Gly | Ala | Arg | Gly |
| | | | | 645 | | | | | 650 | | | | | 655 | |
| Leu | Val | Lys | Ser | Val | Ser | Val | Val | Gly | His | Asn | Ser | Val | Gly | Thr | Val |
| | | | 660 | | | | | 665 | | | | | 670 | | |
| Val | Arg | Ile | Gln | Thr | Thr | Ser | Gly | Asn | Thr | Tyr | His | Tyr | Gly | Ile | Ser |
| | | 675 | | | | | 680 | | | | | 685 | | | |
| Asn | Gln | Ala | Glu | Asp | Thr | Gln | Gln | Ala | Thr | His | Thr | Val | Glu | Phe | Ala |
| | 690 | | | | | 695 | | | | | 700 | | | | |
| Gly | Glu | Thr | Tyr | Ser | Trp | Glu | Gly | Ser | Phe | Ala | Gln | Leu | | | |
| 705 | | | | | 710 | | | | 715 | | | | | | |

<210> SEQ ID NO 19
 <211> LENGTH: 825
 <212> TYPE: DNA
 <213> ORGANISM: *Vibrio splendidus*
 <400> SEQUENCE: 19

```

atgaagtggg tattgccaat agttgcgatg tctggtgtcg cattggcggc agaaaataag    60
aatgttgagg tgagcagtga gcatttcgtc cgttatcaat accaagacaa aatcagctat    120
ggaaagctag acaatgacgc agtgttaccg gtcagcggcg atctctttgg cgaatattcg    180
gtagcaaaaa attcgatecc gttagagtgc gttgaggtgt tactaccgac aaaaccagag    240
aaagtcttcg ccgctcggat gaacttcgct agccacttag cctcacctgc cgatgcacca    300
ccgcccagatg ttcttaaact tccttcttct ttgattctca cgggcgaagt gattcaagtg    360
ccacaaaaag caagaaatgt tcattttgaa ggcgagctgg tggttgatgat tggtagagag    420
ctcagtcaag ccagtgaaga agaagccgaa caagcagatc ttggcgtcac ggtgggcaac    480
gatattactg aaagaagttg gcaaggcgcc gatttacaat ggctccgagc gaaagcttcc    540
gatggttttg gcccggttgg caacacaatt gtgcgaggca ttgattacaa caatattgag    600
ttaaccactc gtgtaaccg taaagtgggt caacaagaaa atacttcggt catgatccac    660
    
```

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```

aagccaagaa aagtcgtgag ctatttgagc tattatttta ccctcaaacc gggcgatcta 720
atattcatgg gcacgccagg tagaacttat gctctgtccg acaaagatca agtgagtgtc 780
acgattgaag gggtaggac tgtggtaaat gaagtgcggt tctga 825

```

```

<210> SEQ ID NO 20
<211> LENGTH: 274
<212> TYPE: PRT
<213> ORGANISM: Vibrio splendidus

```

```

<400> SEQUENCE: 20

```

```

Met Lys Trp Leu Leu Ala Ile Val Ala Met Ser Gly Val Ala Leu Ala
 1           5           10          15
Ala Glu Asn Lys Asn Val Glu Val Ser Ser Glu His Phe Val Arg Tyr
          20          25          30
Gln Tyr Gln Asp Lys Ile Ser Tyr Gly Lys Leu Asp Asn Asp Ala Val
          35          40          45
Leu Pro Val Ser Gly Asp Leu Phe Gly Glu Tyr Ser Val Ala Lys Asn
          50          55          60
Ser Ile Pro Leu Glu Ser Val Glu Val Leu Leu Pro Thr Lys Pro Glu
          65          70          75          80
Lys Val Phe Ala Val Gly Met Asn Phe Ala Ser His Leu Ala Ser Pro
          85          90          95
Ala Asp Ala Pro Pro Pro Met Phe Leu Lys Leu Pro Ser Ser Leu Ile
          100         105         110
Leu Thr Gly Glu Val Ile Gln Val Pro Pro Lys Ala Arg Asn Val His
          115         120         125
Phe Glu Gly Glu Leu Val Val Val Ile Gly Arg Glu Leu Ser Gln Ala
          130         135         140
Ser Glu Glu Glu Ala Glu Gln Ala Ile Phe Gly Val Thr Val Gly Asn
          145         150         155         160
Asp Ile Thr Glu Arg Ser Trp Gln Gly Ala Asp Leu Gln Trp Leu Arg
          165         170         175
Ala Lys Ala Ser Asp Gly Phe Gly Pro Val Gly Asn Thr Ile Val Arg
          180         185         190
Gly Ile Asp Tyr Asn Asn Ile Glu Leu Thr Thr Arg Val Asn Gly Lys
          195         200         205
Val Val Gln Gln Glu Asn Thr Ser Phe Met Ile His Lys Pro Arg Lys
          210         215         220
Val Val Ser Tyr Leu Ser Tyr Tyr Phe Thr Leu Lys Pro Gly Asp Leu
          225         230         235         240
Ile Phe Met Gly Thr Pro Gly Arg Thr Tyr Ala Leu Ser Asp Lys Asp
          245         250         255
Gln Val Ser Val Thr Ile Glu Gly Val Gly Thr Val Val Asn Glu Val
          260         265         270
Arg Phe

```

```

<210> SEQ ID NO 21
<211> LENGTH: 717
<212> TYPE: DNA
<213> ORGANISM: Vibrio splendidus

```

```

<400> SEQUENCE: 21

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```

atggctagca cttttaattc aatttcgggc tcgaagcgta gcctgcacgt gcaagtagca    60
cgcgaaatcg ctcgaggaat tttgtctggt gatctgccgc aaggttctat tattcctggt    120
gaaatggcgt tgtgtgaaca gtttggtatc agccgaacgg cacttcgtga agcagttaa    180
ctactgacct ctaaaggctc gtttagagtct cgcctaaaaa ttggtactcg cgtagtgcac    240
cgcgcatact ggaacttcct tgatcctcaa ctgattgaat ggatggacgg actaaccgac    300
gtagaccaat tctgttctca gtttttaggc cttcgccgtg cgatcgagcc tgaagcgtgt    360
gcaactggcg caaaatttgc gacagctgaa caacgtatcg agctttcaga gatcttccaa    420
aagatggtcg aagtggatga agctgaagtg tttgaccaag aacgttggac agacattgat    480
actcgtttcc atagcttgat cttcaatcgc accggtaacg acttctatct accgttcggt    540
aatattctga ctactatggt cgttaacttc atagtgcatt cttctgaaga ggaagcaca    600
tgcatcaatg aacaccgcag aatctatgaa gctatcatgg ccggtgattg tgacaaggct    660
agaattgctt ctgctgttca cttgcaagat gcccaaccacc gtttggcaac agcataa    717

```

<210> SEQ ID NO 22

<211> LENGTH: 238

<212> TYPE: PRT

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 22

```

Met Ala Ser Thr Phe Asn Ser Ile Ser Gly Ser Lys Arg Ser Leu His
 1                               5 10 15
Val Gln Val Ala Arg Glu Ile Ala Arg Gly Ile Leu Ser Gly Asp Leu
 20 25 30
Pro Gln Gly Ser Ile Ile Pro Gly Glu Met Ala Leu Cys Glu Gln Phe
 35 40 45
Gly Ile Ser Arg Thr Ala Leu Arg Glu Ala Val Lys Leu Leu Thr Ser
 50 55 60
Lys Gly Leu Leu Glu Ser Arg Pro Lys Ile Gly Thr Arg Val Val Asp
 65 70 75 80
Arg Ala Tyr Trp Asn Phe Leu Asp Pro Gln Leu Ile Glu Trp Met Asp
 85 90 95
Gly Leu Thr Asp Val Asp Gln Phe Cys Ser Gln Phe Leu Gly Leu Arg
 100 105 110
Arg Ala Ile Glu Pro Glu Ala Cys Ala Leu Ala Ala Lys Phe Ala Thr
 115 120 125
Ala Glu Gln Arg Ile Glu Leu Ser Glu Ile Phe Gln Lys Met Val Glu
 130 135 140
Val Asp Glu Ala Glu Val Phe Asp Gln Glu Arg Trp Thr Asp Ile Asp
 145 150 155 160
Thr Arg Phe His Ser Leu Ile Phe Asn Ala Thr Gly Asn Asp Phe Tyr
 165 170 175
Leu Pro Phe Gly Asn Ile Leu Thr Thr Met Phe Val Asn Phe Ile Val
 180 185 190
His Ser Ser Glu Glu Gly Ser Thr Cys Ile Asn Glu His Arg Arg Ile
 195 200 205
Tyr Glu Ala Ile Met Ala Gly Asp Cys Asp Lys Ala Arg Ile Ala Ser
 210 215 220
Ala Val His Leu Gln Asp Ala Asn His Arg Leu Ala Thr Ala
 225 230 235

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-continued

<210> SEQ ID NO 23
<211> LENGTH: 1779
<212> TYPE: DNA
<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 23

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atggaactca acacgattat tgtcggcatt tatttcctat tcttgattgc gataggttgg      60
atgttttagaa catttacaag tactactagt gactacttcc gcgggggcgg taacatgttg     120
tgggtgattgg ttgggtcaac cgcctttatg acccagttta gtgcatggac attcaccggt     180
gcagcaggtg aagcgtataa cgatggtttc gctgtagcgg tcactctcgt agccaacgca     240
tttggttact teatgaacta cgcgtacttc gcgccgaaat tccgtcaact tcgcgtgtgt     300
acggtaatec aagcgattcg tatgcgtttt ggtgcgacca acgaacaagt attcacttgg     360
tcttcaatgc caaactcagt ggtatctgcy ggtgtgtggt taaacgcatt ggcaatcacc     420
gcttcgggta tcttcggttt cgacatgaac atgactatct gggtgactgg cctagtggta     480
ttggcaatgt cggtaacagg tggttcatgg gcggaatcg catctgactt catgcagatg     540
ggtatcatca tggcggtaac ggtaacttgt gcggtttag cggttgttca agtgggcggg     600
gttggtgaga ttgtaacaa cttcccagta caagatggtg gttcgttcc tgggggcaac     660
aacatcaact acctaagcat ctttacgatt tgggcattct tcactctcgt taagcagttc     720
tcaatcacga acaacatgct taactcttac cgttacctag cggctaaga ctcaaagaac     780
gctaagaaag ctgcactgct tgcttgtgtg ttgatgttgt gtggtgtggt tatttggttc     840
atgccttctt ggttcattgc aggccaaggt gttgatttat cagcggctta cccgaatgca     900
ggtaaaaaag cgggtgactt tgcttaccta tacttctgac aagagtacat gccagcaggt     960
atggttggtc tattagtgtc cgcgatgttt gcagcgacaa tgtcttcaat ggactcaggt    1020
ctaaaccgta actcaggtat ttttgtaag aacttctacg aaacaatcgt tcgtaaggt     1080
caagcatcag agaaagagct agtaaccgta tctaaaatta cttcagcggg atttggtttc    1140
gctattatcc taatcgcaac gttcatcaac tcattaaaag gcttaagcct gtttgatacg    1200
atgatgtacg taggtgcggt aatcggcttc cctatgacga ttctgcatt ccttggtttc    1260
ttcatcaaga agactccgga ctgggctggt tggggaacgc tagttgttgg tggatcgtgta    1320
tcttatgtgg ttggttttgt tatcaacgcg gagatggtag cagcggcgtt tggctctgat    1380
actctaacag gacgtgaatg gtctgatggt aaagttgcga ttggtctgat tgetcacatc    1440
acgtaaccg gtggcttctt cgtactatct acgatgttct acaagcctct atcaaaagaa    1500
cgtaacgagg atggtgataa gttctttggc aacttagata cccattagt agctgaatcg    1560
gcagagcaaa aagtggtgga taacaacaaa cgtcaaatgc ttggtaaact gattgaggta    1620
gcgggtgttg gtattatgct gatggctctt ctgactaacc caatgtgggg gcgctagtc     1680
ttcatcttat gtggtgtgat agtgggtggt gtcggtattc tacttgtgaa agcggtcgat    1740
gacggcggca agcaagcgaa agcagtaacc gaaagctaa                                1779
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<210> SEQ ID NO 24
<211> LENGTH: 592
<212> TYPE: PRT
<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 24

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Met Glu Leu Asn Thr Ile Ile Val Gly Ile Tyr Phe Leu Phe Leu Ile
 1 5 10 15
 Ala Ile Gly Trp Met Phe Arg Thr Phe Thr Ser Thr Thr Ser Asp Tyr
 20 25 30
 Phe Arg Gly Gly Asn Met Leu Trp Trp Met Val Gly Ala Thr Ala
 35 40 45
 Phe Met Thr Gln Phe Ser Ala Trp Thr Phe Thr Gly Ala Ala Gly Lys
 50 55 60
 Ala Tyr Asn Asp Gly Phe Ala Val Ala Val Ile Phe Val Ala Asn Ala
 65 70 75 80
 Phe Gly Tyr Phe Met Asn Tyr Ala Tyr Phe Ala Pro Lys Phe Arg Gln
 85 90 95
 Leu Arg Val Val Thr Val Ile Glu Ala Ile Arg Met Arg Phe Gly Ala
 100 105 110
 Thr Asn Glu Gln Val Phe Thr Trp Ser Ser Met Pro Asn Ser Val Val
 115 120 125
 Ser Ala Gly Val Trp Leu Asn Ala Leu Ala Ile Ile Ala Ser Gly Ile
 130 135 140
 Phe Gly Phe Asp Met Asn Met Thr Ile Trp Val Thr Gly Leu Val Val
 145 150 155 160
 Leu Ala Met Ser Val Thr Gly Gly Ser Trp Ala Val Ile Ala Ser Asp
 165 170 175
 Phe Met Gln Met Val Ile Ile Met Ala Val Thr Val Thr Cys Ala Val
 180 185 190
 Val Ala Val Val Gln Gly Gly Gly Val Gly Glu Ile Val Asn Asn Phe
 195 200 205
 Pro Val Gln Asp Gly Gly Ser Phe Leu Trp Gly Asn Asn Ile Asn Tyr
 210 215 220
 Leu Ser Ile Phe Thr Ile Trp Ala Phe Phe Ile Phe Val Lys Gln Phe
 225 230 235 240
 Ser Ile Thr Asn Asn Met Leu Asn Ser Tyr Arg Tyr Leu Ala Ala Lys
 245 250 255
 Asp Ser Lys Asn Ala Lys Lys Ala Ala Leu Leu Ala Cys Val Leu Met
 260 265 270
 Leu Cys Gly Val Phe Ile Trp Phe Met Pro Ser Trp Phe Ile Ala Gly
 275 280 285
 Gln Gly Val Asp Leu Ser Ala Ala Tyr Pro Asn Ala Gly Lys Lys Ala
 290 295 300
 Gly Asp Phe Ala Tyr Leu Tyr Phe Val Gln Glu Tyr Met Pro Ala Gly
 305 310 315 320
 Met Val Gly Leu Leu Val Ala Ala Met Phe Ala Ala Thr Met Ser Ser
 325 330 335
 Met Asp Ser Gly Leu Asn Arg Asn Ser Gly Ile Phe Val Lys Asn Phe
 340 345 350
 Tyr Glu Thr Ile Val Arg Lys Gly Gln Ala Ser Glu Lys Glu Leu Val
 355 360 365
 Thr Val Ser Lys Ile Thr Ser Ala Val Phe Gly Phe Ala Ile Ile Leu
 370 375 380
 Ile Ala Gln Phe Ile Asn Ser Leu Lys Gly Leu Ser Leu Phe Asp Thr
 385 390 395 400

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Met Met Tyr Val Gly Ala Leu Ile Gly Phe Pro Met Thr Ile Pro Ala
 405 410 415

Phe Leu Gly Phe Phe Ile Lys Lys Thr Pro Asp Trp Ala Gly Trp Gly
 420 425 430

Thr Leu Val Val Gly Gly Ile Val Ser Tyr Val Val Gly Phe Val Ile
 435 440 445

Asn Ala Glu Met Val Ala Ala Ala Phe Gly Leu Asp Thr Leu Thr Gly
 450 455 460

Arg Glu Trp Ser Asp Val Lys Val Ala Ile Gly Leu Ile Ala His Ile
 465 470 475 480

Thr Leu Thr Gly Gly Phe Phe Val Leu Ser Thr Met Phe Tyr Lys Pro
 485 490 495

Leu Ser Lys Glu Arg Gln Ala Asp Val Asp Lys Phe Phe Gly Asn Leu
 500 505 510

Asp Thr Pro Leu Val Ala Glu Ser Ala Glu Gln Lys Val Leu Asp Asn
 515 520 525

Lys Gln Arg Gln Met Leu Gly Lys Leu Ile Ala Val Ala Gly Val Gly
 530 535 540

Ile Met Leu Met Ala Leu Leu Thr Asn Pro Met Trp Gly Arg Leu Val
 545 550 555 560

Phe Ile Leu Cys Gly Val Ile Val Gly Gly Val Gly Ile Leu Leu Val
 565 570 575

Lys Ala Val Asp Asp Gly Gly Lys Gln Ala Lys Ala Val Thr Glu Ser
 580 585 590

<210> SEQ ID NO 25
 <211> LENGTH: 2079
 <212> TYPE: DNA
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 25

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atgagcgacc aaaaatctct tgatgcaatc aggaagatga agctggaaaa cgatacttca    60
gcaggaatc ttgtagacct actccctatc gaagttcaaa cacgtgactt cgacctatca    120
ttcctagaca ccttgagcga agcacgtccg cgtcttcttg ttcaagctga tcagctagaa    180
gaattcaaag caaaagtga agctgatcaa gctcactgta tgtttgatga tttctacaac    240
aactctaccg ttaagttcct tgagactgct cctttcgaag agcctcaagc gtaccagct    300
gagacggtag gtaaagcttc tctatggcgt ccttattggc gtcaaagtga cgttgattgc    360
caaatggcac tgaacgcgac acgtaaccta gcgattgctg gtgttgtaaa agaagacgaa    420
gcgctcattg cgaaagcaaa agcctggact ctaaaactgt ctacgtacga tccagaaggc    480
gtgacttctc gtggctataa cgatgaagcg gctttccgtg ttatcgctgc tatggcttgg    540
ggttacgatt ggctacacgg ctacttcacc gatgaagaac gccagcaagt tcaagatgct    600
ttgattgagc gtctagacga aatcatgcac cacctgaaag tgacggttga tctattgaac    660
aaccactaa atagccacgg tgttcgttct atctcttctg ctatcatocc aacgtgtatc    720
gcgctttacc acgatcaccc gaaagcagcg gagtacattg catacgcgct agaatactac    780
gcagtacatt acccaccatg gggcggtgta gacggcggtt gggctgaagg tcttgattac    840
tggaacacgc aaactgcatt cctagggcaa gcattcgacc tattgaaagc atactgtggt    900
gtagacatgt ttaacaaaac attctacgaa aacacaggtg atttcccgtt ttactgcatg    960
    
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ccagttcact ctaagcgcgc gagcttctgt gaccagtctt caatcggcga tttcccaggt 1020
ttaaactgg cttacaacat caagcactac gcagggtgta accagaagcc tgagtacgtt 1080
tggtactata accagcttaa aggccgtgat actgaagcac acaccaaatt ctacaacttc 1140
ggttggtggg acttcgggta tgacgatctt cgttttaact tcctttggga tgcacctgaa 1200
gagaaagccc catcgaacga tccactgttg aaagtattcc caatcacggg ttgggtgca 1260
ttccacaaca agatgactga gcgtgataac catattcaca tggtattcaa atgttctccg 1320
tttggtcaa tcagccactc tcacgggtgac caaaacgcat ttacgcttca cgcatttggg 1380
gaaacgctag cgtcagtaac aggttactat ggtggtttcg gtgtagacat gcacacgaaa 1440
tggcgtcgtc aaacgttctc taaaaacctg ccactatttg gcggtaaagg tcagtacggc 1500
gagaacaaga acacaggcta cgaaaaccac caagatcgtt tttgtatcga agcggggcggc 1560
actatctctg acttcgacac tgaatctgat gtgaagatgg ttgaaggtga tgcaacggca 1620
tcttacaagt acttcgttcc tgaaatcgaa tcttacaagc gtaaagtctg gttcgttcaa 1680
ggtaaagtct tcgtaatgca agacaaggca acgctttctg aagagaaaga catgacttgg 1740
ctaatgcaca caactttcgc aaacgaagtg gcagacaagt ctttactat cegtggcgaa 1800
gttgcgcacc tagacgtaaa cttcatcaac gagtctgctg ataacatcac gtcagttaag 1860
aacgttgaag gctttggcga agttgaccca tacgagttca aagatcttga gatccaccgt 1920
cacgtggaag tggaattcaa gccatcgaaa gagcacaaca tcctgacgct tcttgttctt 1980
aataagaatg aaggcgagca agttgaagtg tttcacaagc ttgaaggcaa cacgctactg 2040
ctaatgttg acggcgaaac ggtttcaatc gaactgtaa 2079

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<210> SEQ ID NO 26

<211> LENGTH: 692

<212> TYPE: PRT

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 26

```

Met Ser Asp Gln Lys Ser Leu Asp Ala Ile Arg Lys Met Lys Leu Glu
1          5          10          15
Asn Asp Thr Ser Ala Gly Asn Leu Val Asp Leu Leu Pro Ile Glu Val
20        25        30
Gln Thr Arg Asp Phe Asp Leu Ser Phe Leu Asp Thr Leu Ser Glu Ala
35        40        45
Arg Pro Arg Leu Leu Val Gln Ala Asp Gln Leu Glu Glu Phe Lys Ala
50        55        60
Lys Val Lys Ala Asp Gln Ala His Cys Met Phe Asp Asp Phe Tyr Asn
65        70        75        80
Asn Ser Thr Val Lys Phe Leu Glu Thr Ala Pro Phe Glu Glu Pro Gln
85        90        95
Ala Tyr Pro Ala Glu Thr Val Gly Lys Ala Ser Leu Trp Arg Pro Tyr
100       105       110
Trp Arg Gln Met Tyr Val Asp Cys Gln Met Ala Leu Asn Ala Thr Arg
115       120       125
Asn Leu Ala Ile Ala Gly Val Val Lys Glu Asp Glu Ala Leu Ile Ala
130       135       140
Lys Ala Lys Ala Trp Thr Leu Lys Leu Ser Thr Tyr Asp Pro Glu Gly
145       150       155       160

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| | | | | | | | | | | | | | | | |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Val | Thr | Ser | Arg | Gly | Tyr | Asn | Asp | Glu | Ala | Ala | Phe | Arg | Val | Ile | Ala |
| | | | | 165 | | | | | 170 | | | | | 175 | |
| Ala | Met | Ala | Trp | Gly | Tyr | Asp | Trp | Leu | His | Gly | Tyr | Phe | Thr | Asp | Glu |
| | | | 180 | | | | | 185 | | | | | 190 | | |
| Glu | Arg | Gln | Gln | Val | Gln | Asp | Ala | Leu | Ile | Glu | Arg | Leu | Asp | Glu | Ile |
| | | 195 | | | | | 200 | | | | | 205 | | | |
| Met | His | His | Leu | Lys | Val | Thr | Val | Asp | Leu | Leu | Asn | Asn | Pro | Leu | Asn |
| | 210 | | | | | 215 | | | | | 220 | | | | |
| Ser | His | Gly | Val | Arg | Ser | Ile | Ser | Ser | Ala | Ile | Ile | Pro | Thr | Cys | Ile |
| 225 | | | | | 230 | | | | | 235 | | | | | 240 |
| Ala | Leu | Tyr | His | Asp | His | Pro | Lys | Ala | Gly | Glu | Tyr | Ile | Ala | Tyr | Ala |
| | | | | 245 | | | | | 250 | | | | | 255 | |
| Leu | Glu | Tyr | Tyr | Ala | Val | His | Tyr | Pro | Pro | Trp | Gly | Gly | Val | Asp | Gly |
| | | | 260 | | | | | 265 | | | | | 270 | | |
| Gly | Trp | Ala | Glu | Gly | Pro | Asp | Tyr | Trp | Asn | Thr | Gln | Thr | Ala | Phe | Leu |
| | | 275 | | | | | 280 | | | | | 285 | | | |
| Gly | Glu | Ala | Phe | Asp | Leu | Leu | Lys | Ala | Tyr | Cys | Gly | Val | Asp | Met | Phe |
| | 290 | | | | | 295 | | | | 300 | | | | | |
| Asn | Lys | Thr | Phe | Tyr | Glu | Asn | Thr | Gly | Asp | Phe | Pro | Leu | Tyr | Cys | Met |
| 305 | | | | | 310 | | | | | 315 | | | | | 320 |
| Pro | Val | His | Ser | Lys | Arg | Ala | Ser | Phe | Cys | Asp | Gln | Ser | Ser | Ile | Gly |
| | | | | 325 | | | | | 330 | | | | | 335 | |
| Asp | Phe | Pro | Gly | Leu | Lys | Leu | Ala | Tyr | Asn | Ile | Lys | His | Tyr | Ala | Gly |
| | | 340 | | | | | | 345 | | | | | 350 | | |
| Val | Asn | Gln | Lys | Pro | Glu | Tyr | Val | Trp | Tyr | Tyr | Asn | Gln | Leu | Lys | Gly |
| | | 355 | | | | | 360 | | | | | 365 | | | |
| Arg | Asp | Thr | Glu | Ala | His | Thr | Lys | Phe | Tyr | Asn | Phe | Gly | Trp | Trp | Asp |
| | 370 | | | | | 375 | | | | | 380 | | | | |
| Phe | Gly | Tyr | Asp | Asp | Leu | Arg | Phe | Asn | Phe | Leu | Trp | Asp | Ala | Pro | Glu |
| 385 | | | | | 390 | | | | | 395 | | | | | 400 |
| Glu | Lys | Ala | Pro | Ser | Asn | Asp | Pro | Leu | Leu | Lys | Val | Phe | Pro | Ile | Thr |
| | | | 405 | | | | | 410 | | | | | | 415 | |
| Gly | Trp | Ala | Ala | Phe | His | Asn | Lys | Met | Thr | Glu | Arg | Asp | Asn | His | Ile |
| | | 420 | | | | | | 425 | | | | | 430 | | |
| His | Met | Val | Phe | Lys | Cys | Ser | Pro | Phe | Gly | Ser | Ile | Ser | His | Ser | His |
| | 435 | | | | | | 440 | | | | | 445 | | | |
| Gly | Asp | Gln | Asn | Ala | Phe | Thr | Leu | His | Ala | Phe | Gly | Glu | Thr | Leu | Ala |
| | 450 | | | | | 455 | | | | | 460 | | | | |
| Ser | Val | Thr | Gly | Tyr | Tyr | Gly | Gly | Phe | Gly | Val | Asp | Met | His | Thr | Lys |
| 465 | | | | 470 | | | | | | 475 | | | | | 480 |
| Trp | Arg | Arg | Gln | Thr | Phe | Ser | Lys | Asn | Leu | Pro | Leu | Phe | Gly | Gly | Lys |
| | | | 485 | | | | | | 490 | | | | | 495 | |
| Gly | Gln | Tyr | Gly | Glu | Asn | Lys | Asn | Thr | Gly | Tyr | Glu | Asn | His | Gln | Asp |
| | | 500 | | | | | | 505 | | | | | 510 | | |
| Arg | Phe | Cys | Ile | Glu | Ala | Gly | Gly | Thr | Ile | Ser | Asp | Phe | Asp | Thr | Glu |
| | | 515 | | | | | | 520 | | | | 525 | | | |
| Ser | Asp | Val | Lys | Met | Val | Glu | Gly | Asp | Ala | Thr | Ala | Ser | Tyr | Lys | Tyr |
| | 530 | | | | | 535 | | | | | 540 | | | | |
| Phe | Val | Pro | Glu | Ile | Glu | Ser | Tyr | Lys | Arg | Lys | Val | Trp | Phe | Val | Gln |
| 545 | | | | | 550 | | | | | 555 | | | | | 560 |
| Gly | Lys | Val | Phe | Val | Met | Gln | Asp | Lys | Ala | Thr | Leu | Ser | Glu | Glu | Lys |

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gccgatatac gctttgacga tttgaaaacg cagcttgaac agctggacag catctacgaa 540
gccaaaaggca ttatcatcga cgaaaagggg atggtggctg cttcaacaat cgaaaacgtg 600
cttccgcaaa ccaatatatc ttctgcagac actcaaatga aactcaacag tgccattgaa 660
cagcctgatc aattcattga ggggtgtgatt gatggtaacc agagaatctt gatggccaag 720
aaagtggata ttggcagcca gaaagagtgg ttcattgatc ccagtattga ccctgaactc 780
gcgctcaatc agctgaatgg cgtgatgctg agtgccgcga tccttatcgt cgcttggtga 840
cttggctcgg tgatattgat gattttactt ctgaatcgtt tctaccgccc aatcgtgtca 900
ctgcgcaaaa tcgtccacga tctatcacia ggtaacggag acctcactca aaggcttgct 960
gagaagggga atgatgactt agggcatatc gccaaagaca tcaacttggt cattatcggc 1020
ttacaagaga tggttaagga tgtgaaatc aagaactcgg atctcgatac caaggtactg 1080
agtattcgcg aaggttgtaa agaaaccagc gatgtactga aagttcatac tgatgaaacg 1140
gttcaagtgg tctctcgcgat taacggcttg tctgaagcat caaacgaagt agagaagagt 1200
tctcagtcgg cggcagaagc agcaagagag gccgctgtgt tcagtgatga gacgaaacag 1260
attaacacgg tgacggaaac ctatatcagt gatcttgaga agcaagtctg caccacttct 1320
gatgacattc gctcaatggc caatgaaacg cagagcatcc agtctatcgt gtctgtgatt 1380
ggcggaaatt cggaacaaac taatttgctg gcattgaatg cgtcaattga ageggcgagg 1440
gcggtggaac atggtcaggg tttcgcggtg gttgctgatg aagtcctgac gctagccaac 1500
cgaacgcaaa tcagtacctc tgaaattgat gaagcgttat ctggcttgca gtctaaatca 1560
gatggtttgg ttaaatctat tgagttgacc aaaagtaact gtgaactgac tcgcgctcaa 1620
gttgttcaag ctgtaaacat gttggcgaag ctaaccgagc agatggaac agtaagtctg 1680
ttaaataatg acatttcggg ttcgtctgtt gagcaaaaac cccttattca gagcattgct 1740
aagaacatgc ataagattga aagctttggt gaggagctta ataaactaag ccaagatcag 1800
ttaaactaat cagcagaaat caaaacactt aacggtagcg ttagtgaatt gatgagcagc 1860
ttaaaggttt aa 1872

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<210> SEQ ID NO 30

<211> LENGTH: 623

<212> TYPE: PR

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 30

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Met Val Ala Val Val Ser Ser Ser Ala Leu Ala Phe Thr Asn Trp Phe
1           5           10           15
Thr Leu Asn Leu Ala Thr Glu Gln Val Asn Gln Thr Ile Tyr Asn Glu
20          25          30
Ile Asp His Ser Leu Thr Ile Glu Ile Asn Gln Ile Glu Ser Thr Val
35          40          45
Gln Arg Thr Ile Asp Thr Val Asn Ser Val Ala Gln Glu Phe Met Lys
50          55          60
Ser Pro Tyr Gln Val Pro Asn Glu Ala Leu Met His Tyr Ala Ala Lys
65          70          75          80
Leu Gly Gly Ile Asp Lys Ile Val Val Gly Phe Asp Asp Gly Arg Ser
85          90          95
Tyr Thr Ser Arg Pro Ser Glu Ser Phe Pro Asn Gly Val Gly Ile Lys

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| 100 | | | | | 105 | | | | | 110 | | | | | |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Glu | Lys | Tyr | Asn | Pro | Thr | Thr | Arg | Pro | Trp | Tyr | Gln | Gln | Ala | Lys | Leu |
| | 115 | | | | | | 120 | | | | | 125 | | | |
| Lys | Ser | Gly | Leu | Ser | Phe | Ser | Gly | Leu | Phe | Phe | Thr | Lys | Ser | Thr | Gln |
| | 130 | | | | | | 135 | | | | 140 | | | | |
| Val | Pro | Met | Ile | Gly | Val | Thr | Tyr | Ser | Tyr | Gln | Asp | Arg | Val | Ile | Met |
| 145 | | | | | 150 | | | | | 155 | | | | | 160 |
| Ala | Asp | Ile | Arg | Phe | Asp | Asp | Leu | Glu | Thr | Gln | Leu | Glu | Gln | Leu | Asp |
| | | | | 165 | | | | | | 170 | | | | | 175 |
| Ser | Ile | Tyr | Glu | Ala | Lys | Gly | Ile | Ile | Ile | Asp | Glu | Lys | Gly | Met | Val |
| | | | 180 | | | | | | | 185 | | | | 190 | |
| Val | Ala | Ser | Thr | Ile | Glu | Asn | Val | Leu | Pro | Gln | Thr | Asn | Ile | Ser | Ser |
| | | 195 | | | | | 200 | | | | | | 205 | | |
| Ala | Asp | Thr | Gln | Met | Lys | Leu | Asn | Ser | Ala | Ile | Glu | Gln | Pro | Asp | Gln |
| | 210 | | | | | | 215 | | | | | 220 | | | |
| Phe | Ile | Glu | Gly | Val | Ile | Asp | Gly | Asn | Gln | Arg | Ile | Leu | Met | Ala | Lys |
| 225 | | | | | 230 | | | | | 235 | | | | | 240 |
| Lys | Val | Asp | Ile | Gly | Ser | Gln | Lys | Glu | Trp | Phe | Met | Ile | Ser | Ser | Ile |
| | | | | 245 | | | | | | 250 | | | | | 255 |
| Asp | Pro | Glu | Leu | Ala | Leu | Asn | Gln | Leu | Asn | Gly | Val | Met | Ser | Ser | Ala |
| | | | 260 | | | | | | 265 | | | | | 270 | |
| Arg | Ile | Leu | Ile | Val | Ala | Cys | Val | Leu | Gly | Ser | Val | Ile | Leu | Met | Ile |
| | | 275 | | | | | 280 | | | | | | 285 | | |
| Leu | Leu | Leu | Asn | Arg | Phe | Tyr | Arg | Pro | Ile | Val | Ser | Leu | Arg | Lys | Ile |
| | 290 | | | | | | 295 | | | | | 300 | | | |
| Val | His | Asp | Leu | Ser | Gln | Gly | Asn | Gly | Asp | Leu | Thr | Gln | Arg | Leu | Ala |
| 305 | | | | | 310 | | | | | | 315 | | | | 320 |
| Glu | Lys | Gly | Asn | Asp | Asp | Leu | Gly | His | Ile | Ala | Lys | Asp | Ile | Asn | Leu |
| | | | | 325 | | | | | | 330 | | | | | 335 |
| Phe | Ile | Ile | Gly | Leu | Gln | Glu | Met | Val | Lys | Asp | Val | Lys | Tyr | Lys | Asn |
| | | | 340 | | | | | | 345 | | | | | 350 | |
| Ser | Asp | Leu | Asp | Thr | Lys | Val | Leu | Ser | Ile | Arg | Glu | Gly | Cys | Lys | Glu |
| | | 355 | | | | | 360 | | | | | | 365 | | |
| Thr | Ser | Asp | Val | Leu | Lys | Val | His | Thr | Asp | Glu | Thr | Val | Gln | Val | Val |
| | 370 | | | | | | 375 | | | | | | 380 | | |
| Ser | Ala | Ile | Asn | Gly | Leu | Ser | Glu | Ala | Ser | Asn | Glu | Val | Glu | Lys | Ser |
| 385 | | | | | 390 | | | | | 395 | | | | | 400 |
| Ser | Gln | Ser | Ala | Ala | Glu | Ala | Ala | Arg | Glu | Ala | Ala | Val | Phe | Ser | Asp |
| | | | | 405 | | | | | 410 | | | | | | 415 |
| Glu | Thr | Lys | Gln | Ile | Asn | Thr | Val | Thr | Glu | Thr | Tyr | Ile | Ser | Asp | Leu |
| | | | 420 | | | | | | 425 | | | | | 430 | |
| Glu | Lys | Gln | Val | Cys | Thr | Thr | Ser | Asp | Asp | Ile | Arg | Ser | Met | Ala | Asn |
| | | 435 | | | | | | 440 | | | | | 445 | | |
| Glu | Thr | Gln | Ser | Ile | Gln | Ser | Ile | Val | Ser | Val | Ile | Gly | Gly | Ile | Ala |
| | 450 | | | | | | 455 | | | | | | 460 | | |
| Glu | Gln | Thr | Asn | Leu | Leu | Ala | Leu | Asn | Ala | Ser | Ile | Glu | Ala | Ala | Arg |
| 465 | | | | | 470 | | | | | 475 | | | | | 480 |
| Ala | Gly | Glu | His | Gly | Arg | Gly | Phe | Ala | Val | Val | Ala | Asp | Glu | Val | Arg |
| | | | | 485 | | | | | 490 | | | | | | 495 |
| Ala | Leu | Ala | Asn | Arg | Thr | Gln | Ile | Ser | Thr | Ser | Glu | Ile | Asp | Glu | Ala |
| | | | 500 | | | | | | 505 | | | | | 510 | |

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Leu Ser Gly Leu Gln Ser Lys Ser Asp Gly Leu Val Lys Ser Ile Glu
 515 520 525
 Leu Thr Lys Ser Asn Cys Glu Leu Thr Arg Ala Gln Val Val Gln Ala
 530 535 540
 Val Asn Met Leu Ala Lys Leu Thr Glu Gln Met Glu Thr Val Ser Arg
 545 550 555 560
 Phe Asn Asn Asp Ile Ser Gly Ser Ser Val Glu Gln Asn Ala Leu Ile
 565 570 575
 Gln Ser Ile Ala Lys Asn Met His Lys Ile Glu Ser Phe Val Glu Glu
 580 585 590
 Leu Asn Lys Leu Ser Gln Asp Gln Leu Thr Glu Ser Ala Glu Ile Lys
 595 600 605
 Thr Leu Asn Gly Ser Val Ser Glu Leu Met Ser Ser Phe Lys Val
 610 615 620

<210> SEQ ID NO 31

<211> LENGTH: 1743

<212> TYPE: DNA

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 31

```

gtgaataagc caatctttgt cgctgtactc gcttcgctta cgtatggctg cggtggaagc   60
agctccagtg actctagtga ccctctgat accaataact caggagcadc ttatggtggt   120
gttgctccct atgatattgc caagatcaa aacatccttt ccagctcaga tcttcagggt   180
tctgatccta atggagagga gggcaataaa acctctgaag tcaaagatgg taacttcgat   240
ggttatgtca gtgattatgt ttatgctgac gaagagacgg aaaatctgat cttcaaaatg   300
gcgaactaca agatgcgctc tgaagtctgt gaaggagaaa acttcgatat caatgaagca   360
ggcgtaagac gcagctctaca tgcggaaata agcctacctg atattgagca tghtaatggcg   420
agttctcccg cagatcacga tgaagtgacc gtgctacaga tccacaataa aggtacagac   480
gagagtggca cgggttatat ccctcatccg ctattgctgt tggtttggga gcaagaacga   540
gatggcctca caggctacta ctgggcagtc atgaaaaata atgccattga ctgtagcagt   600
gccgctgact cttcggattg ttatgccact tcatataatc gctacgattt gggagaggcg   660
gatctcgata acttcaccaa gtttgatctt tctgtttatg aaaataccct ttcgatcaaa   720
gtgaacgatg aagttaaagt cgacgaagac atcacctact ggcagcatct actgagttac   780
tttaaagcgg gtatctacaa tcaatttgaa aatggtgaag ccacggctca ctttcaggca   840
ctgcgataca ccaccacaca ggtcaacggc tcaaacgatt gggatattaa tgattggaag   900
ttgacgattc ctgcgagtaa agacacttgg tatggaagtg ggggtgacag tgcggctgaa   960
ctagaacctg agcgtctgca atcgagcaaa gaccttctcg ccaacgacag tgatgtctac  1020
gacagcgata ttggtctttc ttatttcaat accgatgaag ggagagtgca ctttagagcg   1080
gatatgggat atggcacctc taccgaaaat tctagctata ttcgctctga gctcaggagg   1140
ttgtatcaaa gcagtgttca accggattgt agcaccagcg atgaagatac aagttggtat   1200
ttggacgaca ctagaacgaa cgctaccagt cacgagttaa ccgcaagctt acgaattgaa   1260
gactaccgca acattaataa ccaagaccgg aaagtgggtc ttgggcaaat acacggttgg   1320
aagatcaatc aagcattggt gaagtgtgta tgggaaggcg agagtaagcc agtaagagtg   1380

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atactgaact ctgattttga gcgcaacaac caagactgta accattgtga cccgttcagt 1440
gtcaggttag gtacttattc ggcaagtga gagtggcgat atacgattcg agccaatcaa 1500
gacggtatct acttagcgac tcatgattta gatggaacta atacggtttc tcatttaatc 1560
ccttggggac aagattacac agataaagat ggggacacgg tctcgttgac gtcagattgg 1620
acatcgacag acatcgcttt ctatttcaaa gcgggcatct acccacaatt taagcctgat 1680
agcgactatg cgggtgaagt gtttgatgtg agctttagtt ctctaagagc agagcataac 1740
tga 1743

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<210> SEQ ID NO 32
<211> LENGTH: 580
<212> TYPE: PRT
<213> ORGANISM: Vibrio splendidus

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<400> SEQUENCE: 32

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```

Met Asn Lys Pro Ile Phe Val Val Val Leu Ala Ser Leu Thr Tyr Gly
1 5 10 15
Cys Gly Gly Ser Ser Ser Ser Asp Ser Ser Asp Pro Ser Asp Thr Asn
20 25 30
Asn Ser Gly Ala Ser Tyr Gly Val Val Ala Pro Tyr Asp Ile Ala Lys
35 40 45
Tyr Gln Asn Ile Leu Ser Ser Ser Asp Leu Gln Val Ser Asp Pro Asn
50 55 60
Gly Glu Glu Gly Asn Lys Thr Ser Glu Val Lys Asp Gly Asn Phe Asp
65 70 75 80
Gly Tyr Val Ser Asp Tyr Phe Tyr Ala Asp Glu Glu Thr Glu Asn Leu
85 90 95
Ile Phe Lys Met Ala Asn Tyr Lys Met Arg Ser Glu Val Arg Glu Gly
100 105 110
Glu Asn Phe Asp Ile Asn Glu Ala Gly Val Arg Arg Ser Leu His Ala
115 120 125
Glu Ile Ser Leu Pro Asp Ile Glu His Val Met Ala Ser Ser Pro Ala
130 135 140
Asp His Asp Glu Val Thr Val Leu Gln Ile His Asn Lys Gly Thr Asp
145 150 155 160
Glu Ser Gly Thr Gly Tyr Ile Pro His Pro Leu Leu Arg Val Val Trp
165 170 175
Glu Gln Glu Arg Asp Gly Leu Thr Gly His Tyr Trp Ala Val Met Lys
180 185 190
Asn Asn Ala Ile Asp Cys Ser Ser Ala Ala Asp Ser Ser Asp Cys Tyr
195 200 205
Ala Thr Ser Tyr Asn Arg Tyr Asp Leu Gly Glu Ala Asp Leu Asp Asn
210 215 220
Phe Thr Lys Phe Asp Leu Ser Val Tyr Glu Asn Thr Leu Ser Ile Lys
225 230 235 240
Val Asn Asp Glu Val Lys Val Asp Glu Asp Ile Thr Tyr Trp Gln His
245 250 255
Leu Leu Ser Tyr Phe Lys Ala Gly Ile Tyr Asn Gln Phe Glu Asn Gly
260 265 270
Glu Ala Thr Ala His Phe Gln Ala Leu Arg Tyr Thr Thr Thr Gln Val
275 280 285

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Asn Gly Ser Asn Asp Trp Asp Ile Asn Asp Trp Lys Leu Thr Ile Pro
 290 295 300
 Ala Ser Lys Asp Thr Trp Tyr Gly Ser Gly Gly Asp Ser Ala Ala Glu
 305 310 315 320
 Leu Glu Pro Glu Arg Cys Glu Ser Ser Lys Asp Leu Leu Ala Asn Asp
 325 330 335
 Ser Asp Val Tyr Asp Ser Asp Ile Gly Leu Ser Tyr Phe Asn Thr Asp
 340 345 350
 Glu Gly Arg Val His Phe Arg Ala Asp Met Gly Tyr Gly Thr Ser Thr
 355 360 365
 Glu Asn Ser Ser Tyr Ile Arg Ser Glu Leu Arg Glu Leu Tyr Gln Ser
 370 375 380
 Ser Val Gln Pro Asp Cys Ser Thr Ser Asp Glu Asp Thr Ser Trp Tyr
 385 390 395 400
 Leu Asp Asp Thr Arg Thr Asn Ala Thr Ser His Glu Leu Thr Ala Ser
 405 410 415
 Leu Arg Ile Glu Asp Tyr Pro Asn Ile Asn Asn Gln Asp Pro Lys Val
 420 425 430
 Val Leu Gly Gln Ile His Gly Trp Lys Ile Asn Gln Ala Leu Val Lys
 435 440 445
 Leu Leu Trp Glu Gly Glu Ser Lys Pro Val Arg Val Ile Leu Asn Ser
 450 455 460
 Asp Phe Glu Arg Asn Asn Gln Asp Cys Asn His Cys Asp Pro Phe Ser
 465 470 475 480
 Val Glu Leu Gly Thr Tyr Ser Ala Ser Glu Glu Trp Arg Tyr Thr Ile
 485 490 495
 Arg Ala Asn Gln Asp Gly Ile Tyr Leu Ala Thr His Asp Leu Asp Gly
 500 505 510
 Thr Asn Thr Val Ser His Leu Ile Pro Trp Gly Gln Asp Tyr Thr Asp
 515 520 525
 Lys Asp Gly Asp Thr Val Ser Leu Thr Ser Asp Trp Thr Ser Thr Asp
 530 535 540
 Ile Ala Phe Tyr Phe Lys Ala Gly Ile Tyr Pro Gln Phe Lys Pro Asp
 545 550 555 560
 Ser Asp Tyr Ala Gly Glu Val Phe Asp Val Ser Phe Ser Ser Leu Arg
 565 570 575
 Ala Glu His Asn
 580

<210> SEQ ID NO 33

<211> LENGTH: 1569

<212> TYPE: DNA

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 33

```

atgaaacaaa ttactctaaa aactttactc gcttcttcta ttctacttgc ggttggttgt    60
gcgagcacga gcacgcctac tgctgatttt ccaaataaca aagaaactgg tgaagcgctt    120
ctgacgccag ttgctgtttc cgctagtagc catgatggta acggacctga tcgtctcgtt    180
gaccaagacc taactacacg ttggtcacct cggggtgacg gcgagtgggc aacgctagac    240
tatggttcag tacaggagtt tgacgcgggt caggecatett tcagtaaagg taatcagcgc    300
caatctaaat ttgatatcca agtgagtgtt gatggcgaac gctggacaac ggtactagaa    360

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aaccaactaa gctcaggtaa agcgatcggc ctagagcggt tccaatttga gccagtagtg 420
caagcacgct acgtaagata cgttgggtcac ggtaacacca aaaacggttg gaacagtgtg 480
actggattag cggcgggttaa ctgtagcatt aacgcatgtc ctgctagcca tatcatcact 540
tcagacgtgg ttgcagcaga agccgtgatt attgctgaaa tgaaaagcggc agaaaaagca 600
cgtaaagatg cgcgcaaaga tctacgctct ggtaacttcg gtgtagcagc ggtttaccct 660
tgtgagacga ccggtgaatg tgacactcgc agtgcacttc cagttccgac aggccctgcca 720
gcgacaccag ttgcaggtaa ctgcgcaagc gaaaactttg acatgacgca ttggtacctt 780
tctcaacat ttgaccatga caaaaatggc aaacctgatg atgtgtctga gtggaacctt 840
gaaaacgggt accaacaccc tgaaatcttc tacacagctg atgacggcgg cctagtattc 900
aaagcttacg tgaagggtgt acgtacctct aaaaactata agtacgcgcg tacagagctt 960
cgtgaaatga tgcgctcgtg tgatcagtct attagcacta aagtggttaa taagaataac 1020
tgggtattct caagcgctcc tgaatctgac ttagagtcgg cagcgggtat tgacggcgtt 1080
ctagaagcga cgttggaaat cgaccatgca acaacgacgg gtaatgcgaa tgaagtaggt 1140
cgctttatca ttggtcagat tcacgatcaa aacgatgaac caattcgttt gtactaccgt 1200
aaactgccaa accaagaaac gggtcggtt tacttcgcac atgaaagcca agacgcaact 1260
aaagaggact tctacctct agtggcgac atgacggctg aagtgggtga cgatggtatc 1320
gcgcttggcg aagtgttcag ctaccgtatt gacgttaaag gcaacacgat gactgtaacg 1380
ctaatacgtg aaggcaaaga cgatgttgta caagtggttg atatgagcaa cagcggctac 1440
gagcagggcg gcaagtacat gtacttcaaa gccggtgttt acaacaaaa catcagcggc 1500
gacntagacg attactcaca agcgacttcc taccagctag atgtatcgca cgatcaatac 1560
aaaaagtaa 1569

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<210> SEQ ID NO 34

<211> LENGTH: 522

<212> TYPE: PRT

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 34

```

Met Lys Gln Ile Thr Leu Lys Thr Leu Leu Ala Ser Ser Ile Leu Leu
 1           5           10          15
Ala Val Gly Cys Ala Ser Thr Ser Thr Pro Thr Ala Asp Phe Pro Asn
          20           25           30
Asn Lys Glu Thr Gly Glu Ala Leu Leu Thr Pro Val Ala Val Ser Ala
          35           40           45
Ser Ser His Asp Gly Asn Gly Pro Asp Arg Leu Val Asp Gln Asp Leu
          50           55           60
Thr Thr Arg Trp Ser Ser Ala Gly Asp Gly Glu Trp Ala Thr Leu Asp
 65           70           75           80
Tyr Gly Ser Val Gln Glu Phe Asp Ala Val Gln Ala Ser Phe Ser Lys
          85           90           95
Gly Asn Gln Arg Gln Ser Lys Phe Asp Ile Gln Val Ser Val Asp Gly
          100          105          110
Glu Ser Trp Thr Thr Val Leu Glu Asn Gln Leu Ser Ser Gly Lys Ala
          115          120          125
Ile Gly Leu Glu Arg Phe Gln Phe Glu Pro Val Val Gln Ala Arg Tyr

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| 130 | | | 135 | | | 140 | | | | | | | | | |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Val | Arg | Tyr | Val | Gly | His | Gly | Asn | Thr | Lys | Asn | Gly | Trp | Asn | Ser | Val |
| 145 | | | | | 150 | | | | | 155 | | | | 160 | |
| Thr | Gly | Leu | Ala | Ala | Val | Asn | Cys | Ser | Ile | Asn | Ala | Cys | Pro | Ala | Ser |
| | | | 165 | | | | | | 170 | | | | | 175 | |
| His | Ile | Ile | Thr | Ser | Asp | Val | Val | Ala | Ala | Glu | Ala | Val | Ile | Ile | Ala |
| | | | 180 | | | | | 185 | | | | | 190 | | |
| Glu | Met | Lys | Ala | Ala | Glu | Lys | Ala | Arg | Lys | Asp | Ala | Arg | Lys | Asp | Leu |
| | | 195 | | | | | 200 | | | | | 205 | | | |
| Arg | Ser | Gly | Asn | Phe | Gly | Val | Ala | Ala | Val | Tyr | Pro | Cys | Glu | Thr | Thr |
| | 210 | | | | | 215 | | | | | 220 | | | | |
| Val | Glu | Cys | Asp | Thr | Arg | Ser | Ala | Leu | Pro | Val | Pro | Thr | Gly | Leu | Pro |
| 225 | | | | | 230 | | | | | 235 | | | | 240 | |
| Ala | Thr | Pro | Val | Ala | Gly | Asn | Ser | Pro | Ser | Glu | Asn | Phe | Asp | Met | Thr |
| | | | 245 | | | | | | 250 | | | | | 255 | |
| His | Trp | Tyr | Leu | Ser | Gln | Pro | Phe | Asp | His | Asp | Lys | Asn | Gly | Lys | Pro |
| | | | 260 | | | | | 265 | | | | | 270 | | |
| Asp | Asp | Val | Ser | Glu | Trp | Asn | Leu | Ala | Asn | Gly | Tyr | Gln | His | Pro | Glu |
| | | 275 | | | | | 280 | | | | | 285 | | | |
| Ile | Phe | Tyr | Thr | Ala | Asp | Asp | Gly | Gly | Leu | Val | Phe | Lys | Ala | Tyr | Val |
| | 290 | | | | | 295 | | | | | 300 | | | | |
| Lys | Gly | Val | Arg | Thr | Ser | Lys | Asn | Thr | Lys | Tyr | Ala | Arg | Thr | Glu | Leu |
| 305 | | | | | 310 | | | | | 315 | | | | 320 | |
| Arg | Glu | Met | Met | Arg | Arg | Gly | Asp | Gln | Ser | Ile | Ser | Thr | Lys | Gly | Val |
| | | | 325 | | | | | | 330 | | | | | 335 | |
| Asn | Lys | Asn | Asn | Trp | Val | Phe | Ser | Ser | Ala | Pro | Glu | Ser | Asp | Leu | Glu |
| | | 340 | | | | | | 345 | | | | | 350 | | |
| Ser | Ala | Ala | Gly | Ile | Asp | Gly | Val | Leu | Glu | Ala | Thr | Leu | Lys | Ile | Asp |
| | | 355 | | | | | 360 | | | | | 365 | | | |
| His | Ala | Thr | Thr | Thr | Gly | Asn | Ala | Asn | Glu | Val | Gly | Arg | Phe | Ile | Ile |
| | 370 | | | | | 375 | | | | | 380 | | | | |
| Gly | Gln | Ile | His | Asp | Gln | Asn | Asp | Glu | Pro | Ile | Arg | Leu | Tyr | Tyr | Arg |
| 385 | | | | | 390 | | | | | 395 | | | | 400 | |
| Lys | Leu | Pro | Asn | Gln | Glu | Thr | Gly | Ala | Val | Tyr | Phe | Ala | His | Glu | Ser |
| | | | 405 | | | | | | 410 | | | | | 415 | |
| Gln | Asp | Ala | Thr | Lys | Glu | Asp | Phe | Tyr | Pro | Leu | Val | Gly | Asp | Met | Thr |
| | | 420 | | | | | | 425 | | | | | 430 | | |
| Ala | Glu | Val | Gly | Asp | Asp | Gly | Ile | Ala | Leu | Gly | Glu | Val | Phe | Ser | Tyr |
| | | 435 | | | | 440 | | | | | | 445 | | | |
| Arg | Ile | Asp | Val | Lys | Gly | Asn | Thr | Met | Thr | Val | Thr | Leu | Ile | Arg | Glu |
| | 450 | | | | | 455 | | | | | 460 | | | | |
| Gly | Lys | Asp | Asp | Val | Val | Gln | Val | Val | Asp | Met | Ser | Asn | Ser | Gly | Tyr |
| 465 | | | | | 470 | | | | | 475 | | | | 480 | |
| Asp | Ala | Gly | Gly | Lys | Tyr | Met | Tyr | Phe | Lys | Ala | Gly | Val | Tyr | Asn | Gln |
| | | | 485 | | | | | | 490 | | | | | 495 | |
| Asn | Ile | Ser | Gly | Asp | Leu | Asp | Asp | Tyr | Ser | Gln | Ala | Thr | Phe | Tyr | Gln |
| | | 500 | | | | | | 505 | | | | | 510 | | |
| Leu | Asp | Val | Ser | His | Asp | Gln | Tyr | Lys | Lys | | | | | | |
| | | 515 | | | | | 520 | | | | | | | | |

-continued

<211> LENGTH: 1230

<212> TYPE: DNA

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 35

```

atgcaaattt ctaaagtgc tacagctgct gctctttcga caggtttatt atttggtgt 60
aacagtgatg gtttacctat tccaacagat ccaggcggaa cagaccctgt tgaacctgtt 120
gaagtttact ctatagaaaa cgtctattgg gatctgacag gtggtgctgt tgctgcacag 180
tcactcagcg gaacttcacc atatcgcttt gataataatg aggaaggtag tcgtgctcta 240
agcatttaca gtggagacgt agctaattgg ttcacttttg agagttcaat atatactgct 300
gaagaagaag gtggtgtttc ctttgaaggt aaggactgta cttacacagt gactgagcaa 360
cagctagata tgacctgtga aaaagatgac gtagaacag cttactcagc aacagagatt 420
acagatgaat ctgttataac tgcattagaa aatgccgatg atggaaaacc taaatcagtc 480
gatgatgtga acgctcggat tgcactcagc gaagatggcg cgattattga tttatcatct 540
gaaggtacgt ttgataccgg tgttattgag ctaaataaag ctgtcacaat tgatggtgct 600
ggttttagcaa ccattaccgg agatgcttgt attgatgtca ctgcaccggg tgcaggtatc 660
aaaaacatga cttttgctaa cgacaatttg gccgggtggt ttggtaggga gtcagctggt 720
acttcagata atgaaactgg tgcgatcgtt attggtaaaa ttggtaaaga ttcagatcct 780
gtagcacttg aaaacctaaa gttcgatgca aacggcatta ccgaagatga tctaggtact 840
aaaaaagcaa gttggttatt ctctcgaggt tactttacat tagacaatag cgaatttgtc 900
ggtttaagtg gcagtttcca aaataatgca attcgtatta actgtagtag tgacaacggg 960
cgatttggtt cacaaatcac aaataataga ttcactatta actctggtgg tagtgatgtg 1020
ggcggaaatta aagttggtga ttctagcagt gccgtcataa agaatagtag tgataacctt 1080
ggctgtaatg tcactattga aagcaatagc ttcaatggtt acaaaaccct actttcagct 1140
gacaacggta aagatataag aaatacagcc atctacgcac aaccatctgc agtgaacact 1200
gcggcaggta aagaaaaat cttgaactaa 1230

```

<210> SEQ ID NO 36

<211> LENGTH: 409

<212> TYPE: PRT

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 36

```

Met Gln Ile Ser Lys Val Ala Thr Ala Val Ala Leu Ser Thr Gly Leu
1           5           10           15

Leu Phe Gly Cys Asn Ser Asp Gly Leu Pro Ile Pro Thr Asp Pro Gly
20           25           30

Gly Thr Asp Pro Val Glu Pro Val Glu Val Tyr Ser Ile Glu Asn Val
35           40           45

Tyr Trp Asp Leu Thr Gly Gly Ala Val Ala Ala Gln Ser Leu Ser Gly
50           55           60

Thr Ser Pro Tyr Arg Phe Asp Asn Asn Glu Glu Gly Thr Arg Ala Leu
65           70           75           80

Ser Ile Tyr Ser Gly Asp Val Ala Asn Gly Phe Thr Phe Glu Ser Ser
85           90           95

Ile Tyr Thr Ala Glu Glu Glu Gly Val Val Ser Phe Glu Gly Lys Asp
100          105          110

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Cys Thr Tyr Thr Val Thr Glu Gln Gln Leu Asp Met Thr Cys Glu Lys
 115 120 125
 Asp Asp Val Glu Thr Ala Tyr Ser Ala Thr Glu Ile Thr Asp Glu Ser
 130 135 140
 Val Ile Thr Ala Leu Glu Asn Ala Asp Asp Gly Lys Pro Lys Ser Val
 145 150 155 160
 Asp Asp Val Asn Ala Ala Ile Ala Ser Ala Glu Asp Gly Ala Ile Ile
 165 170 175
 Asp Leu Ser Ser Glu Gly Thr Phe Asp Thr Gly Val Ile Glu Leu Asn
 180 185 190
 Lys Ala Val Thr Ile Asp Gly Ala Gly Leu Ala Thr Ile Thr Gly Asp
 195 200 205
 Ala Cys Ile Asp Val Thr Ala Pro Gly Ala Gly Ile Lys Asn Met Thr
 210 215 220
 Phe Ala Asn Asp Asn Leu Ala Gly Cys Phe Gly Arg Glu Ser Ala Gly
 225 230 235 240
 Thr Ser Asp Asn Glu Thr Gly Ala Ile Val Ile Gly Lys Ile Gly Lys
 245 250 255
 Asp Ser Asp Pro Val Ala Leu Glu Asn Leu Lys Phe Asp Ala Asn Gly
 260 265 270
 Ile Thr Glu Asp Asp Leu Gly Thr Lys Lys Ala Ser Trp Leu Phe Ser
 275 280 285
 Arg Gly Tyr Phe Thr Leu Asp Asn Ser Glu Phe Val Gly Leu Ser Gly
 290 295 300
 Ser Phe Gln Asn Asn Ala Ile Arg Ile Asn Cys Ser Ser Asp Asn Gly
 305 310 315 320
 Arg Phe Gly Ser Gln Ile Thr Asn Asn Thr Phe Thr Ile Asn Ser Gly
 325 330 335
 Gly Ser Asp Val Gly Gly Ile Lys Val Gly Asp Ser Ser Ser Ala Val
 340 345 350
 Ile Lys Asn Ser Asp Asp Asn Leu Gly Cys Asn Val Thr Ile Glu Ser
 355 360 365
 Asn Thr Phe Asn Gly Tyr Lys Thr Leu Leu Ser Ala Asp Asn Gly Lys
 370 375 380
 Asp Ile Arg Asn Thr Ala Ile Tyr Ala Gln Pro Ser Ala Val Asn Thr
 385 390 395 400
 Ala Ala Gly Lys Glu Asn Ile Leu Asn
 405

<210> SEQ ID NO 37

<211> LENGTH: 861

<212> TYPE: DNA

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 37

```

atgaattctg ttacaaaaat tgctgcagct gttgcatgta ctcttttagc gggcacagct    60
gctggtgcat ctcttgatta tcgttacgag tatcgtgctg cgacggatta tacaaagact    120
aatggtgata cggctcacgt agacgctcgc catcaacacc gagttaagct aggtgaaagc    180
ttaaagctgt cagacaagtg gaagcactct actggtctag aacttaagtt ccacggtgat    240
gactcttact atgatgaaga ttcaggttct gttaaatcag caaacagcca gagtttttac    300

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gatggcaatt ggtacatcta tggatggag atcgataaca ctgcgacata caaaatagac 360
aataattggt atctacaaat gggatgacct attgcttggg attgggatga gcctaagtct 420
aacgatggcg actggaagat gaaaaaggtt acgtttaaac ctcagttccg cgttggctat 480
aaagcagata tgggtttaac aactgctatt cgttaccgtc atgaatatgc tgacttccgt 540
aaccacacac aatttggcga caaagattct gaaactggcg agcgtttaga atcagctcaa 600
aagtctaaag ttacactgac gggctcttac aaaattgaat ctctacctaa gcttggcctt 660
tcttacgaag caaactatgt aaaatctttg gataacgtac ttctttataa tagtgatgac 720
tgggaatggg atgctggcct aaaggtaaac tacaagttcg gttcttggaa accttttgct 780
gaaatctggt cttctgatat cagttcatct tcaaaagatc gtgaagctaa ataccgtggt 840
ggtattgctt actcattcta a 861

```

<210> SEQ ID NO 38

<211> LENGTH: 286

<212> TYPE: PRT

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 38

```

Met Asn Ser Val Thr Lys Ile Ala Ala Ala Val Ala Cys Thr Leu Leu
1      5      10      15
Ala Gly Thr Ala Ala Gly Ala Ser Leu Asp Tyr Arg Tyr Glu Tyr Arg
20     25     30
Ala Ala Thr Asp Tyr Thr Lys Thr Asn Gly Asp Thr Ala His Val Asp
35     40     45
Ala Arg His Gln His Arg Val Lys Leu Gly Glu Ser Phe Lys Leu Ser
50     55     60
Asp Lys Trp Lys His Ser Thr Gly Leu Glu Leu Lys Phe His Gly Asp
65     70     75     80
Asp Ser Tyr Tyr Asp Glu Asp Ser Gly Ser Val Lys Ser Ala Asn Ser
85     90     95
Gln Ser Phe Tyr Asp Gly Asn Trp Tyr Ile Tyr Gly Met Glu Ile Asp
100    105    110
Asn Thr Ala Thr Tyr Lys Ile Asp Asn Asn Trp Tyr Leu Gln Met Gly
115    120    125
Met Pro Ile Ala Trp Asp Trp Asp Glu Pro Asn Ala Asn Asp Gly Asp
130    135    140
Trp Lys Met Lys Lys Val Thr Phe Lys Pro Gln Phe Arg Val Gly Tyr
145    150    155    160
Lys Ala Asp Met Gly Leu Thr Thr Ala Ile Arg Tyr Arg His Glu Tyr
165    170    175
Ala Asp Phe Arg Asn His Thr Gln Phe Gly Asp Lys Asp Ser Glu Thr
180    185    190
Gly Glu Arg Leu Glu Ser Ala Gln Lys Ser Lys Val Thr Leu Thr Gly
195    200    205
Ser Tyr Lys Ile Glu Ser Leu Pro Lys Leu Gly Leu Ser Tyr Glu Ala
210    215    220
Asn Tyr Val Lys Ser Leu Asp Asn Val Leu Leu Tyr Asn Ser Asp Asp
225    230    235    240
Trp Glu Trp Asp Ala Gly Leu Lys Val Asn Tyr Lys Phe Gly Ser Trp
245    250    255

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Lys Pro Phe Ala Glu Ile Trp Ser Ser Asp Ile Ser Ser Ser Ser Lys
 260 265 270

Asp Arg Glu Ala Lys Tyr Arg Val Gly Ile Ala Tyr Ser Phe
 275 280 285

<210> SEQ ID NO 39
 <211> LENGTH: 1038
 <212> TYPE: DNA
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 39

```

atgtttaaga aaaacatatt agcagtggcg ttattagcga ctgtgccaat ggttactttc    60
gcaaataaacg gtgtttctta ccccgtaact gccgataaat tcgatatgca taattggaaa    120
ataaccatac cttcagatat taatgaagat ggtcgcgttg atgaaataga aggggtcgct    180
atgatgagct actcacatag tgatttcttc catcttgata aagacggcaa ccttgtat    240
gaagtgcaga accaagcgat tacgacgaaa aactcgaaga atgcgcgttc tgagttacgc    300
cagatgccaa gaggcgcaga tttctctatc gatacggctg ataaaggaaa ccagtgggca    360
ctgtcgagtc acccagcggc tagtgaatac agtgctgtgg gcggaacatt agaagcgaca    420
ttaaaagtga atcacgtctc agttaacgct aagttcccag aaaaaatccc agctcattct    480
gttgtgggtg gtcagattca tgctaaaaaa cacaacgagc taatcaaagc tgaaccgggt    540
tatgggcatg gtaatgaacc actaaagatc ttctataaga agtttcctga ccaagaaatg    600
ggttcagtat tctggaacta tgaacgtaac ctagagaaaa aagatcctaa ccgtgccgat    660
atcgcttate cagtgtgggg taacacgtgg gaaaaccctg cagagccggg tgaagccgggt    720
attgctcttg gtgaagagtt tagctacaaa gtggaagtga aaggcaccat gatgtacct    780
acgtttgaaa ccgagcgtca cgataccggt aagtatgaaa tcgacctgag taagggcatt    840
gatgaacttg actcaccaac gggctatgct gaagatgatt tttactacaa agcgggcgca    900
tacggccaat gtagcgtgag cgattctcac cctgtatggg ggcctggttg tggcggtact    960
ggcgatttcg ctgtcgataa aaagaatggc gattacaaca gtgtgacttt ctctgcgctt   1020
aagttaaacg gtaaatag                                     1038
  
```

<210> SEQ ID NO 40
 <211> LENGTH: 345
 <212> TYPE: PRT
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 40

```

Met Phe Lys Lys Asn Ile Leu Ala Val Ala Leu Leu Ala Thr Val Pro
1                  5                                  10                                  15
Met Val Thr Phe Ala Asn Asn Gly Val Ser Tyr Pro Val Pro Ala Asp
                  20                                  25                                  30
Lys Phe Asp Met His Asn Trp Lys Ile Thr Ile Pro Ser Asp Ile Asn
          35                                  40                                  45
Glu Asp Gly Arg Val Asp Glu Ile Glu Gly Val Ala Met Met Ser Tyr
          50                                  55                                  60
Ser His Ser Asp Phe Phe His Leu Asp Lys Asp Gly Asn Leu Val Phe
65                  70                                  75                                  80
Glu Val Gln Asn Gln Ala Ile Thr Thr Lys Asn Ser Lys Asn Ala Arg
          85                                  90                                  95
  
```

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Ser Glu Leu Arg Gln Met Pro Arg Gly Ala Asp Phe Ser Ile Asp Thr
 100 105 110

Ala Asp Lys Gly Asn Gln Trp Ala Leu Ser Ser His Pro Ala Ala Ser
 115 120 125

Glu Tyr Ser Ala Val Gly Gly Thr Leu Glu Ala Thr Leu Lys Val Asn
 130 135 140

His Val Ser Val Asn Ala Lys Phe Pro Glu Lys Tyr Pro Ala His Ser
 145 150 155 160

Val Val Val Gly Gln Ile His Ala Lys Lys His Asn Glu Leu Ile Lys
 165 170 175

Ala Gly Thr Gly Tyr Gly His Gly Asn Glu Pro Leu Lys Ile Phe Tyr
 180 185 190

Lys Lys Phe Pro Asp Gln Glu Met Gly Ser Val Phe Trp Asn Tyr Glu
 195 200 205

Arg Asn Leu Glu Lys Lys Asp Pro Asn Arg Ala Asp Ile Ala Tyr Pro
 210 215 220

Val Trp Gly Asn Thr Trp Glu Asn Pro Ala Glu Pro Gly Glu Ala Gly
 225 230 235 240

Ile Ala Leu Gly Glu Glu Phe Ser Tyr Lys Val Glu Val Lys Gly Thr
 245 250 255

Met Met Tyr Leu Thr Phe Glu Thr Glu Arg His Asp Thr Val Lys Tyr
 260 265 270

Glu Ile Asp Leu Ser Lys Gly Ile Asp Glu Leu Asp Ser Pro Thr Gly
 275 280 285

Tyr Ala Glu Asp Asp Phe Tyr Tyr Lys Ala Gly Ala Tyr Gly Gln Cys
 290 295 300

Ser Val Ser Asp Ser His Pro Val Trp Gly Pro Gly Cys Gly Gly Thr
 305 310 315 320

Gly Asp Phe Ala Val Asp Lys Lys Asn Gly Asp Tyr Asn Ser Val Thr
 325 330 335

Phe Ser Ala Leu Lys Leu Asn Gly Lys
 340 345

<210> SEQ ID NO 41
 <211> LENGTH: 897
 <212> TYPE: DNA
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 41

```

atggataact ctccggtgct gagccgattt ttagagaatg gatttttact ccagcagaaa    60
ctgagccttg ttctttgttg tgtgttgatc gcagcttctg catggatttt aggacagctt    120
gcatggttta ttgaacctgc tgagcaaacc gtcgtgccat ggacagcaac ggcttcctcg    180
tcttcaacgc ctcaatcgac tcttgatc tcttcttgc agcagagcaa catgtttggt    240
gcttataacc caaccacgcc tgctgtggtt gagcagcaag ttatccaaga tgcgcaaag    300
acgcgactga acctcgtttt agtgggtgca gtagccagtt ctaatccaaa gctgagcttg    360
gctgtgattg ccaatcgcg cacacaagca acctacggca ttaatgaaga gatcgaaggt    420
acgcgagcta agttaaagc ggtattagtc gatcgctga ttattgataa ctcaggtcga    480
gacgaaacct tgatgcttga aggcattgag tacaagcgtt tgtctgtatc agcacctgcg    540
ccacctcgta cctcttcttc tgtgcgtggc aacaaccag cttctgcaga agagaagcta    600
    
```

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gatgaaatta aagcgaagat aatgaaagat ccgcaacaaa tcttccaata tgttcgactg   660
tctcaggtga aacgcgacga taaagtgatt ggttatcgtg tgagccctgg caaagattca   720
gaacttttta actctgttgg gctccaaaac ggagatattg ccaactcagtt aaatggacaa   780
gacctgacag accctgctgc tatgggcaac atattccggt ctatctcaga gctgacagag   840
ctaaacctcg tcgtcgagag agatgggtcaa caacatgaag tgtttattga attttag    897

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<210> SEQ ID NO 42

<211> LENGTH: 298

<212> TYPE: PRT

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 42

```

Met Asp Asn Ser Pro Val Leu Ser Arg Phe Leu Glu Asn Gly Phe Leu
 1          5          10          15
Leu Gln Gln Lys Leu Ser Leu Val Leu Cys Cys Val Leu Ile Ala Ala
 20          25          30
Ser Ala Trp Ile Leu Gly Gln Leu Ala Trp Phe Ile Glu Pro Ala Glu
 35          40          45
Gln Thr Val Val Pro Trp Thr Ala Thr Ala Ser Ser Ser Ser Thr Pro
 50          55          60
Gln Ser Thr Leu Asp Ile Ser Ser Leu Gln Gln Ser Asn Met Phe Gly
 65          70          75          80
Ala Tyr Asn Pro Thr Thr Pro Ala Val Val Glu Gln Gln Val Ile Gln
 85          90          95
Asp Ala Pro Lys Thr Arg Leu Asn Leu Val Leu Val Gly Ala Val Ala
100          105          110
Ser Ser Asn Pro Lys Leu Ser Leu Ala Val Ile Ala Asn Arg Gly Thr
115          120          125
Gln Ala Thr Tyr Gly Ile Asn Glu Glu Ile Glu Gly Thr Arg Ala Lys
130          135          140
Leu Lys Ala Val Leu Val Asp Arg Val Ile Ile Asp Asn Ser Gly Arg
145          150          155          160
Asp Glu Thr Leu Met Leu Glu Gly Ile Glu Tyr Lys Arg Leu Ser Val
165          170          175
Ser Ala Pro Ala Pro Pro Arg Thr Ser Ser Ser Val Arg Gly Asn Asn
180          185          190
Pro Ala Ser Ala Glu Glu Lys Leu Asp Glu Ile Lys Ala Lys Ile Met
195          200          205
Lys Asp Pro Gln Gln Ile Phe Gln Tyr Val Arg Leu Ser Gln Val Lys
210          215          220
Arg Asp Asp Lys Val Ile Gly Tyr Arg Val Ser Pro Gly Lys Asp Ser
225          230          235          240
Glu Leu Phe Asn Ser Val Gly Leu Gln Asn Gly Asp Ile Ala Thr Gln
245          250          255
Leu Asn Gly Gln Asp Leu Thr Asp Pro Ala Ala Met Gly Asn Ile Phe
260          265          270
Arg Ser Ile Ser Glu Leu Thr Glu Leu Asn Leu Val Val Glu Arg Asp
275          280          285
Gly Gln Gln His Glu Val Phe Ile Glu Phe
290          295

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<210> SEQ ID NO 43
<211> LENGTH: 2025
<212> TYPE: DNA
<213> ORGANISM: Vibrio splendidus

<400> SEQUENCE: 43
gtgaagcatt ggtttaagaa aagtgcattg ttattggcag gaagcttaat ctgcacaccc   60
gcagccatcg cgagtgattt tagtgccagc ttaaaggca ctgatattca agagtttatt   120
aatattgttg gtcgtaacct agagaagacg atcatcgttg acccttcggg gcgcgaaaa   180
atcgatgtac gcagctacga cgtactcaat gaagagcaat actacagctt cttcctaaac   240
gtattggaag tgtatggcta cgcggtgtgc gaaatggact cgggtgttct taagatcatc   300
aaggccaaag attcgaanaa atcggcaatt ccagtcgttg gagacagtga cagatcaaaa   360
ggcgacaatg tgggtgacag tgttgtagc gttcgtaatg tctcgggtgc tgaactttct   420
cctctgcttc gtcaactaaa cgacaatgca ggcgaggta acgttgtgca ctacgacca   480
gccaacatca tccttattac aggccgagcg gcggtagtaa accgtttagc tgaatcatc   540
aagcgtgttg accaagcggg tgataaagag attgaagtgc ttgagctaaa gaatgcttct   600
gcgcgagaaa tggtagctat cgttgatgcy ttaagcaaaa cactgatgc gaaaaacaca   660
cctgcatttc tacaacctaa attagttgcc gatgaacgta ccaatgcgat tcttatctca   720
ggcgacccta aagtacgtag ccgtttaaga aggctgattg aacagcttga tgttgaatg   780
gcaaccaag gcaataacca agttatttac cttaaatatg caaaagccga agatctagtt   840
gatgtgctga aaggcgtgtc ggacaaccta caatcagaga agcagacatc aaccaaagga   900
agttcatcgc agcgtaacca agtgatgatc tcagctcaca gtgacaccaa ctctttagtg   960
attaccgcac agccggacat catgaatgcy cttcaagatg tgatcgaca gctggatatt  1020
cgtcgtgctc aagtattgat tgaagcactg attgtcgaag tggccgaagg tgacggcgtt  1080
aaccttggty tgcagtgggg taaccttgaa acgggtgcca tgattcagta cagcaacct  1140
ggcgttcca ttggcgtgtg gatggttggg ttagaagaag cgaagacag cgaaacgaca  1200
accgctgttt atgattcaga cggtaaatc ttacgtaatg aaaccacgac ggaagaaggt  1260
gactattcaa cattagcttc cgcactttct ggtgttaatg gtgcggcaat gagtgtggta  1320
atgggtgact ggaccgctt gatcagtgca gtagcgaccg attcaaatc aatatccta  1380
tcttctcaa gtatcacctg gatggataac ggcgaagcgt cattcattgt gggatgaagag  1440
gtgctgttcc taaccggttc tacagcagcc tcaagtaacg acaaccatt ccaaacagtt  1500
gaacgtaaa gagtgggat caagcttaaa gtggtgccgc aaatcaatga aggtgattcg  1560
gttcaactgc aatagaaca agaagtatcg aacgtattag gcgcaatgg tgcggttgat  1620
gtgcttttg ctaagcgaca gctaaataca tcagtgattg ttcaagacgg tcaaatgctg  1680
gtgttgggtg gcttgattga cgagcgagca ttgaaagtg aatctaaggt gccgttcttg  1740
ggagatattc ctgtgcttgg acacttgttc aaatcaacca gtactcaggt tgagaaaaag  1800
aacctaatgg tcttcatcaa accaaccatt attcgtgatg gtatgacagc cgatggatc  1860
acgcagccta aatacaactt catccgtgct gagcagttgt acaaggctga gcaaggactg  1920
aagttaatgg cagacgataa catcccagta ttgcctaaat ttggtgccga catgaatcac  1980
ccgctgaaa ttcaagcctt catcgatcaa atggaacaag aataa 2025

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<210> SEQ ID NO 44
<211> LENGTH: 674
<212> TYPE: PRT
<213> ORGANISM: Vibrio splendidus

<400> SEQUENCE: 44

Met Lys His Trp Phe Lys Lys Ser Ala Trp Leu Leu Ala Gly Ser Leu
1          5          10          15
Ile Cys Thr Pro Ala Ala Ile Ala Ser Asp Phe Ser Ala Ser Phe Lys
20          25          30
Gly Thr Asp Ile Gln Glu Phe Ile Asn Ile Val Gly Arg Asn Leu Glu
35          40          45
Lys Thr Ile Ile Val Asp Pro Ser Val Arg Gly Lys Ile Asp Val Arg
50          55          60
Ser Tyr Asp Val Leu Asn Glu Glu Gln Tyr Tyr Ser Phe Phe Leu Asn
65          70          75          80
Val Leu Glu Val Tyr Gly Tyr Ala Val Val Glu Met Asp Ser Gly Val
85          90          95
Leu Lys Ile Ile Lys Ala Lys Asp Ser Lys Thr Ser Ala Ile Pro Val
100         105         110
Val Gly Asp Ser Asp Thr Ile Lys Gly Asp Asn Val Val Thr Arg Val
115         120         125
Val Thr Val Arg Asn Val Ser Val Arg Glu Leu Ser Pro Leu Leu Arg
130         135         140
Gln Leu Asn Asp Asn Ala Gly Ala Gly Asn Val Val His Tyr Asp Pro
145         150         155         160
Ala Asn Ile Ile Leu Ile Thr Gly Arg Ala Ala Val Val Asn Arg Leu
165         170         175
Ala Glu Ile Ile Lys Arg Val Asp Gln Ala Gly Asp Lys Glu Ile Glu
180         185         190
Val Val Glu Leu Lys Asn Ala Ser Ala Ala Glu Met Val Arg Ile Val
195         200         205
Asp Ala Leu Ser Lys Thr Thr Asp Ala Lys Asn Thr Pro Ala Phe Leu
210         215         220
Gln Pro Lys Leu Val Ala Asp Glu Arg Thr Asn Ala Ile Leu Ile Ser
225         230         235         240
Gly Asp Pro Lys Val Arg Ser Arg Leu Arg Arg Leu Ile Glu Gln Leu
245         250         255
Asp Val Glu Met Ala Thr Lys Gly Asn Asn Gln Val Ile Tyr Leu Lys
260         265         270
Tyr Ala Lys Ala Glu Asp Leu Val Asp Val Leu Lys Gly Val Ser Asp
275         280         285
Asn Leu Gln Ser Glu Lys Gln Thr Ser Thr Lys Gly Ser Ser Ser Gln
290         295         300
Arg Asn Gln Val Met Ile Ser Ala His Ser Asp Thr Asn Ser Leu Val
305         310         315         320
Ile Thr Ala Gln Pro Asp Ile Met Asn Ala Leu Gln Asp Val Ile Ala
325         330         335
Gln Leu Asp Ile Arg Arg Ala Gln Val Leu Ile Glu Ala Leu Ile Val
340         345         350
Glu Met Ala Glu Gly Asp Gly Val Asn Leu Gly Val Gln Trp Gly Asn
355         360         365

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Leu Glu Thr Gly Ala Met Ile Gln Tyr Ser Asn Thr Gly Ala Ser Ile
 370 375 380
 Gly Gly Val Met Val Gly Leu Glu Glu Ala Lys Asp Ser Glu Thr Thr
 385 390 395 400
 Thr Ala Val Tyr Asp Ser Asp Gly Lys Phe Leu Arg Asn Glu Thr Thr
 405 410 415
 Thr Glu Glu Gly Asp Tyr Ser Thr Leu Ala Ser Ala Leu Ser Gly Val
 420 425 430
 Asn Gly Ala Ala Met Ser Val Val Met Gly Asp Trp Thr Ala Leu Ile
 435 440 445
 Ser Ala Val Ala Thr Asp Ser Asn Ser Asn Ile Leu Ser Ser Pro Ser
 450 455 460
 Ile Thr Val Met Asp Asn Gly Glu Ala Ser Phe Ile Val Gly Glu Glu
 465 470 475 480
 Val Pro Val Leu Thr Gly Ser Thr Ala Gly Ser Ser Asn Asp Asn Pro
 485 490 495
 Phe Gln Thr Val Glu Arg Lys Glu Val Gly Ile Lys Leu Lys Val Val
 500 505 510
 Pro Gln Ile Asn Glu Gly Asp Ser Val Gln Leu Gln Ile Glu Gln Glu
 515 520 525
 Val Ser Asn Val Leu Gly Ala Asn Gly Ala Val Asp Val Arg Phe Ala
 530 535 540
 Lys Arg Gln Leu Asn Thr Ser Val Ile Val Gln Asp Gly Gln Met Leu
 545 550 555 560
 Val Leu Gly Gly Leu Ile Asp Glu Arg Ala Leu Glu Ser Glu Ser Lys
 565 570 575
 Val Pro Phe Leu Gly Asp Ile Pro Val Leu Gly His Leu Phe Lys Ser
 580 585 590
 Thr Ser Thr Gln Val Glu Lys Lys Asn Leu Met Val Phe Ile Lys Pro
 595 600 605
 Thr Ile Ile Arg Asp Gly Met Thr Ala Asp Gly Ile Thr Gln Arg Lys
 610 615 620
 Tyr Asn Phe Ile Arg Ala Glu Gln Leu Tyr Lys Ala Glu Gln Gly Leu
 625 630 635 640
 Lys Leu Met Ala Asp Asp Asn Ile Pro Val Leu Pro Lys Phe Gly Ala
 645 650 655
 Asp Met Asn His Pro Ala Glu Ile Gln Ala Phe Ile Asp Gln Met Glu
 660 665 670
 Gln Glu

<210> SEQ ID NO 45

<211> LENGTH: 1503

<212> TYPE: DNA

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 45

atggctgaat tggtaggggc ggcacgtact taccagcgct tgccgttag ctttgccaat 60
 cgctacaaga tgggtgtgga ataccaacat ccagagcgcg caccgatact ttattatggt 120
 gagccactga aatcgggcgc gatcattgaa gtgagtcgtg ttgtgaaaaa tggtttcacg 180
 ccacaagcga ttactctcga tgagtttgat aaaaaactaa ccgatgctta tcagcgtgac 240
 tcgtcagaag ctgcgcagct catggaagac attggtgctg atagtatga tttcttctca 300

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ctagcggaag aactgcctca agacgaagac ttacttgaat cagaagatga tgcaccaatc 360
atcaagttaa tcaatgcgat gctgggtgag gcgatcaaag aggggtgcttc ggatatacac 420
atcgaaacct ttgaaaagtc actttgtatc cgtttccgag ttgatgggtg gctgcgtgat 480
gttctagcgc caagccgtaa actggctccg ctattggttt cagctgtcaa ggttatggct 540
aaactggata ttgcggaaaa acgctgtcca caagatggtc gtatttctct gcgtattggt 600
ggccgagcgg ttgatgttcg tgtttcaacc atgccttctt cgcattggtga gcgtgtggta 660
atgctctctg tggacaaaaa tgccactcgt ctgacttgc acagtttagg tatgacagcc 720
gaaaaccatg aaaacttccg taagctgatt cagcgcaccac atggcattat cttggtgacc 780
ggcccacag gttcaggtaa atcgaagacc ttgtacgag gtctgcaaga actcaacagc 840
aatgaacgaa acattttaac cgttgaagac ccaatcgaat tcgatatcga tggcattggt 900
caaacacaag tgaaccctaa ggttgatag acctttgcgc gtggtttacg tgccattctt 960
cgtaagatc ctgatgttgt tatgattggt gagatccgtg acttgagac cgcagagatt 1020
gctgtccagg cctctttgac aggtcactta gttatgtcga ctctgcatac caatactgcc 1080
gtcgtgctga ttacacgtct acgtgatag ggcattgaac ctttcttgat ctcttcttcg 1140
ctgctgggtg ttttggtcga gcgcttggtt cgtactttat gtaacgaatg taaagaacct 1200
tatgaagccg ataaagagca gaagaaactg tttgggttga agaagaaaga aagcttgacg 1260
ctttaccatg ccaaaggttg tgaagagtgt ggccataagg gttatcgagg tcgtacgggt 1320
attcatgagc tgttgatgat tgatgattca gtacaagagc tgattcacag tgaagcgggt 1380
gagcaggcga ttgataaagc aattcgtggc acaacaccaa gtattcgaga tgatggcttg 1440
agcaaagttc tgaaggggtt aacgtcccta gaagaagtga tgcgcgtgac caaggaagtc 1500
tag 1503

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<210> SEQ ID NO 46
<211> LENGTH: 500
<212> TYPE: PRT
<213> ORGANISM: Vibrio splendidus

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<400> SEQUENCE: 46

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```

Met Ala Glu Leu Val Gly Ala Ala Arg Thr Tyr Gln Arg Leu Pro Phe
 1             5             10             15
Ser Phe Ala Asn Arg Tyr Lys Met Val Leu Glu Tyr Gln His Pro Glu
 20             25             30
Arg Ala Pro Ile Leu Tyr Tyr Val Glu Pro Leu Lys Ser Ala Ala Ile
 35             40             45
Ile Glu Val Ser Arg Val Val Lys Asn Gly Phe Thr Pro Gln Ala Ile
 50             55             60
Thr Leu Asp Glu Phe Asp Lys Lys Leu Thr Asp Ala Tyr Gln Arg Asp
 65             70             75             80
Ser Ser Glu Ala Arg Gln Leu Met Glu Asp Ile Gly Ala Asp Ser Asp
 85             90             95
Asp Phe Phe Ser Leu Ala Glu Glu Leu Pro Gln Asp Glu Asp Leu Leu
 100            105            110
Glu Ser Glu Asp Asp Ala Pro Ile Ile Lys Leu Ile Asn Ala Met Leu
 115            120            125
Gly Glu Ala Ile Lys Glu Gly Ala Ser Asp Ile His Ile Glu Thr Phe

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-continued

| 130 | 135 | 140 |
|--|-----|-----|
| Glu Lys Ser Leu Cys Ile Arg Phe Arg Val Asp Gly Val Leu Arg Asp 145 150 155 160 | | |
| Val Leu Ala Pro Ser Arg Lys Leu Ala Pro Leu Leu Val Ser Arg Val 165 170 175 | | |
| Lys Val Met Ala Lys Leu Asp Ile Ala Glu Lys Arg Val Pro Gln Asp 180 185 190 | | |
| Gly Arg Ile Ser Leu Arg Ile Gly Gly Arg Ala Val Asp Val Arg Val 195 200 205 | | |
| Ser Thr Met Pro Ser Ser His Gly Glu Arg Val Val Met Arg Leu Leu 210 215 220 | | |
| Asp Lys Asn Ala Thr Arg Leu Asp Leu His Ser Leu Gly Met Thr Ala 225 230 235 240 | | |
| Glu Asn His Glu Asn Phe Arg Lys Leu Ile Gln Arg Pro His Gly Ile 245 250 255 | | |
| Ile Leu Val Thr Gly Pro Thr Gly Ser Gly Lys Ser Thr Thr Leu Tyr 260 265 270 | | |
| Ala Gly Leu Gln Glu Leu Asn Ser Asn Glu Arg Asn Ile Leu Thr Val 275 280 285 | | |
| Glu Asp Pro Ile Glu Phe Asp Ile Asp Gly Ile Gly Gln Thr Gln Val 290 295 300 | | |
| Asn Pro Lys Val Asp Met Thr Phe Ala Arg Gly Leu Arg Ala Ile Leu 305 310 315 320 | | |
| Arg Gln Asp Pro Asp Val Val Met Ile Gly Glu Ile Arg Asp Leu Glu 325 330 335 | | |
| Thr Ala Glu Ile Ala Val Gln Ala Ser Leu Thr Gly His Leu Val Met 340 345 350 | | |
| Ser Thr Leu His Thr Asn Thr Ala Val Gly Ala Ile Thr Arg Leu Arg 355 360 365 | | |
| Asp Met Gly Ile Glu Pro Phe Leu Ile Ser Ser Ser Leu Leu Gly Val 370 375 380 | | |
| Leu Ala Gln Arg Leu Val Arg Thr Leu Cys Asn Glu Cys Lys Glu Pro 385 390 395 400 | | |
| Tyr Glu Ala Asp Lys Glu Gln Lys Lys Leu Phe Gly Leu Lys Lys Lys 405 410 415 | | |
| Glu Ser Leu Thr Leu Tyr His Ala Lys Gly Cys Glu Glu Cys Gly His 420 425 430 | | |
| Lys Gly Tyr Arg Gly Arg Thr Gly Ile His Glu Leu Leu Met Ile Asp 435 440 445 | | |
| Asp Ser Val Gln Glu Leu Ile His Ser Glu Ala Gly Glu Gln Ala Ile 450 455 460 | | |
| Asp Lys Ala Ile Arg Gly Thr Thr Pro Ser Ile Arg Asp Asp Gly Leu 465 470 475 480 | | |
| Ser Lys Val Leu Lys Gly Val Thr Ser Leu Glu Glu Val Met Arg Val 485 490 495 | | |
| Thr Lys Glu Val 500 | | |

<210> SEQ ID NO 47

<211> LENGTH: 1221

<212> TYPE: DNA

<213> ORGANISM: *Vibrio splendidus*

-continued

<400> SEQUENCE: 47

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atggcggcat ttgaatacaa agcactggat gccaaaggca aaagtaaaaa aggctcaatt    60
gaagcagata atgctcgtca ggctcgccaa agaataaaag agcttggtct gatgccggtt    120
gagatgaccg aggctaaagc aaaaacagca aaagggtgctc agccatcgac cagctttaa    180
cgcgcatca gtacgcctga tcttgcgctt attactcgtc aaatatccac gctcgttcaa    240
tctggtatgc cgctagaaga gtgtttgaaa gccgttgccg aacagtctga gaaacctcgt    300
attcgcacca tgctactcgc ggtgagatct aaggtgactg aaggttattc gttagcagac    360
agcttgctctg attatcccca tatcttcgat gagctattca gagccatggt tgctgctggt    420
gagaagtcag ggcactctaga tgcggtattg gaacgattgg ctgactacgc agaaaaccgt    480
cagaagatgc gttctaagtt gctgcaagcg atgatctacc ccacgtgctt ggtgggtgtt    540
gcggtgacga ttgtgtcgtt cctactggca acggtagtgc cgaagatcgt tgagcctatt    600
atccaaatgg gacaagagct ccctcagtcg acacaatttt tattagcatc gagtgaattt    660
atccagaatt ggggcatcca attactggtg ttgaccattg gtgtgattgt gttggttaag    720
actgcgctga aaaagccggg cgcttcgatg agctgggacg gcaaattatt gagcatcccg    780
ctgataggca agatagcgaa agggatcaac acctctcgtt ttgcacgaac actttctatc    840
tgtacctcta gtgcgattcc tatccttgaa gggatgaagg tcgcggtaga tgtgatgtcg    900
aatcatcacg tgaacaaca agtattacag gcacagata gcgttagaga aggggcaagc    960
ctgcgtaaag cgcttgatca aaccaaactc tttccccga tgatgctgca tatgatcgcc    1020
agtggtgagc agagtggcca attggaacag atgctgacaa gagcggcaga taatcaggat    1080
caaagctttg aatcgaccgt taatcgcgct ttaggcattt ttaccccagc gcttattgctg    1140
ttgatggctg gcttagtgct gtttatcgtg atggcgacgc tgatgccaat gcttgaaatg    1200
aacaatttaa tgagtggta a                                     1221

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<210> SEQ ID NO 48

<211> LENGTH: 406

<212> TYPE: PRT

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 48

```

Met Ala Ala Phe Glu Tyr Lys Ala Leu Asp Ala Lys Gly Lys Ser Lys
1           5           10           15
Lys Gly Ser Ile Glu Ala Asp Asn Ala Arg Gln Ala Arg Gln Arg Ile
20          25          30
Lys Glu Leu Gly Leu Met Pro Val Glu Met Thr Glu Ala Lys Ala Lys
35          40          45
Thr Ala Lys Gly Ala Gln Pro Ser Thr Ser Phe Lys Arg Gly Ile Ser
50          55          60
Thr Pro Asp Leu Ala Leu Ile Thr Arg Gln Ile Ser Thr Leu Val Gln
65          70          75          80
Ser Gly Met Pro Leu Glu Glu Cys Leu Lys Ala Val Ala Glu Gln Ser
85          90          95
Glu Lys Pro Arg Ile Arg Thr Met Leu Leu Ala Val Arg Ser Lys Val
100         105         110
Thr Glu Gly Tyr Ser Leu Ala Asp Ser Leu Ser Asp Tyr Pro His Ile
115        120        125

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-continued

Phe Asp Glu Leu Phe Arg Ala Met Val Ala Ala Gly Glu Lys Ser Gly
 130 135 140

His Leu Asp Ala Val Leu Glu Arg Leu Ala Asp Tyr Ala Glu Asn Arg
 145 150 155 160

Gln Lys Met Arg Ser Lys Leu Leu Gln Ala Met Ile Tyr Pro Ile Val
 165 170 175

Leu Val Val Phe Ala Val Thr Ile Val Ser Phe Leu Leu Ala Thr Val
 180 185 190

Val Pro Lys Ile Val Glu Pro Ile Ile Gln Met Gly Gln Glu Leu Pro
 195 200 205

Gln Ser Thr Gln Phe Leu Leu Ala Ser Ser Glu Phe Ile Gln Asn Trp
 210 215 220

Gly Ile Gln Leu Leu Val Leu Thr Ile Gly Val Ile Val Leu Val Lys
 225 230 235 240

Thr Ala Leu Lys Lys Pro Gly Val Arg Met Ser Trp Asp Arg Lys Leu
 245 250 255

Leu Ser Ile Pro Leu Ile Gly Lys Ile Ala Lys Gly Ile Asn Thr Ser
 260 265 270

Arg Phe Ala Arg Thr Leu Ser Ile Cys Thr Ser Ser Ala Ile Pro Ile
 275 280 285

Leu Glu Gly Met Lys Val Ala Val Asp Val Met Ser Asn His His Val
 290 295 300

Lys Gln Gln Val Leu Gln Ala Ser Asp Ser Val Arg Glu Gly Ala Ser
 305 310 315 320

Leu Arg Lys Ala Leu Asp Gln Thr Lys Leu Phe Pro Pro Met Met Leu
 325 330 335

His Met Ile Ala Ser Gly Glu Gln Ser Gly Gln Leu Glu Gln Met Leu
 340 345 350

Thr Arg Ala Ala Asp Asn Gln Asp Gln Ser Phe Glu Ser Thr Val Asn
 355 360 365

Ile Ala Leu Gly Ile Phe Thr Pro Ala Leu Ile Ala Leu Met Ala Gly
 370 375 380

Leu Val Leu Phe Ile Val Met Ala Thr Leu Met Pro Met Leu Glu Met
 385 390 395 400

Asn Asn Leu Met Ser Gly
 405

<210> SEQ ID NO 49
 <211> LENGTH: 444
 <212> TYPE: DNA
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 49

atgaaaata aatgaaaaa acaatcaggc tttaccctat tagaagtcac ggttgtgtgc 60

ggtatccttg gtgttctagc aagttttggt gtacctaacc tgttgggcaa caaagagaag 120

gcgatcaac aaaaagccat cactgatatt gtggcgctag agaacgcgct cgacatgtac 180

aaactggata acagcgttta cccaacaacg gatcaaggcc tggacggggt ggtgacaaaag 240

ccaagcagtc cagagcctcg taactaccga gacggcgggt acatcaagcg tctacctaac 300

gaccatggg gcaatgagta ccaataccta agtctcgggtg ataacggcac aattgatatc 360

ttcactcttg gcgcagatgg tcaagaaggt ggtgaaggta ttgctgcaga tatcggaac 420

-continued

 tggaacatgc aggacttcca ataa

444

<210> SEQ ID NO 50
 <211> LENGTH: 146
 <212> TYPE: PRT
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 50

Lys Asn Lys Met Lys Lys Gln Ser Gly Phe Thr Leu Leu Glu Val Met
 1 5 10 15
 Val Val Val Val Ile Leu Gly Val Leu Ala Ser Phe Val Val Pro Asn
 20 25 30
 Leu Leu Gly Asn Lys Glu Lys Ala Asp Gln Gln Lys Ala Ile Thr Asp
 35 40 45
 Ile Val Ala Leu Glu Asn Ala Leu Asp Met Tyr Lys Leu Asp Asn Ser
 50 55 60
 Val Tyr Pro Thr Thr Asp Gln Gly Leu Asp Gly Leu Val Thr Lys Pro
 65 70 75 80
 Ser Ser Pro Glu Pro Arg Asn Tyr Arg Asp Gly Gly Tyr Ile Lys Arg
 85 90 95
 Leu Pro Asn Asp Pro Trp Gly Asn Glu Tyr Gln Tyr Leu Ser Pro Gly
 100 105 110
 Asp Asn Gly Thr Ile Asp Ile Phe Thr Leu Gly Ala Asp Gly Gln Glu
 115 120 125
 Gly Gly Glu Gly Ile Ala Ala Asp Ile Gly Asn Trp Asn Met Gln Asp
 130 135 140
 Phe Gln
 145

<210> SEQ ID NO 51
 <211> LENGTH: 594
 <212> TYPE: DNA
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 51

gtgaaaacta agcaaacaca gccaggtttc accttgattg agattctttt ggtggtggta 60
 ttactgtcag tatcgcggtg cgcggtgatc tcgaccatcc ctaccaatag caaagatggt 120
 gctaaaaaat acgctcaaag cttttatcag cgaattcagc tactcaatga agaggctatt 180
 ttgagtggct tagatttttg tgctcgtggt gatgaaaaaa aatcgactta cgttctgatg 240
 actttgaagt ctgatggctg gcaagaaacg gagttcgaaa agatcccttc ttcaactgaa 300
 ttaccggaag aactggcact gtcgctgaca ttagtggtg ggcggtggga agacgatgat 360
 cggttgttca atccaggaag cttatttgat gaagatatgt ttgctgatct tgaagaggaa 420
 aagaagccga aaccaccaca gatctacatc ttgtcgagtg ctgaaatgac gccatttgta 480
 ctgtcgtttt acccaataac cggtgacaca atacaagatg tttggcgcac tcgagtattg 540
 gataatgggtg tgattcgatt actcgagcgc ggagaagaag atgaagaaga ataa 594

<210> SEQ ID NO 52
 <211> LENGTH: 197
 <212> TYPE: PRT
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 52

-continued

Met Lys Thr Lys Gln Thr Gln Pro Gly Phe Thr Leu Ile Glu Ile Leu
 1 5 10 15
 Leu Val Leu Val Leu Leu Ser Val Ser Ala Val Ala Val Ile Ser Thr
 20 25 30
 Ile Pro Thr Asn Ser Lys Asp Val Ala Lys Lys Tyr Ala Gln Ser Phe
 35 40 45
 Tyr Gln Arg Ile Gln Leu Leu Asn Glu Glu Ala Ile Leu Ser Gly Leu
 50 55 60
 Asp Phe Gly Val Arg Val Asp Glu Lys Lys Ser Thr Tyr Val Leu Met
 65 70 75 80
 Thr Leu Lys Ser Asp Gly Trp Gln Glu Thr Glu Phe Glu Lys Ile Pro
 85 90 95
 Ser Ser Thr Glu Leu Pro Glu Glu Leu Ala Leu Ser Leu Thr Leu Gly
 100 105 110
 Gly Gly Ala Trp Glu Asp Asp Asp Arg Leu Phe Asn Pro Gly Ser Leu
 115 120 125
 Phe Asp Glu Asp Met Phe Ala Asp Leu Glu Glu Glu Lys Lys Pro Lys
 130 135 140
 Pro Pro Gln Ile Tyr Ile Leu Ser Ser Ala Glu Met Thr Pro Phe Val
 145 150 155 160
 Leu Ser Phe Tyr Pro Asn Thr Gly Asp Thr Ile Gln Asp Val Trp Arg
 165 170 175
 Ile Arg Val Leu Asp Asn Gly Val Ile Arg Leu Leu Glu Pro Gly Glu
 180 185 190
 Glu Asp Glu Glu
 195

<210> SEQ ID NO 53
 <211> LENGTH: 396
 <212> TYPE: DNA
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 53
 atgaagaaga ataaccgttc tccttatcgt tctcgcggta tgcctcttgg ttctcgagga 60
 atgactctgc ttgaagtatt ggttgogctg gctatcttcg ctacggcggc gatcagtggtg 120
 attcgtgctg tcaccacgca catcaatacg ctcagttatc tcgaagaaaa aaccttcgcg 180
 gcgatggctg ttgataatca aatggcccta gtcatgctac atcctgagat gcttaaaaaa 240
 gcgcagggca cgcaagagtt agcgggaaga gaatggttct ggaaggtgac tcccatcgat 300
 accacgata atttatataa ggcgtttgat gtgagtgcgg caaccagtaa gaaagcgtct 360
 ccagtcgtta cggtgcgcag ttatgtggtt aattaa 396

<210> SEQ ID NO 54
 <211> LENGTH: 131
 <212> TYPE: PRT
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 54
 Met Lys Lys Asn Asn Arg Ser Pro Tyr Arg Ser Arg Gly Met Pro Leu
 1 5 10 15
 Gly Ser Arg Gly Met Thr Leu Leu Glu Val Leu Val Ala Leu Ala Ile
 20 25 30

-continued

Phe Ala Thr Ala Ala Ile Ser Val Ile Arg Ala Val Thr Gln His Ile
 35 40 45

Asn Thr Leu Ser Tyr Leu Glu Glu Lys Thr Phe Ala Ala Met Val Val
 50 55 60

Asp Asn Gln Met Ala Leu Val Met Leu His Pro Glu Met Leu Lys Lys
 65 70 75 80

Ala Gln Gly Thr Gln Glu Leu Ala Gly Arg Glu Trp Phe Trp Lys Val
 85 90 95

Thr Pro Ile Asp Thr Ser Asp Asn Leu Leu Lys Ala Phe Asp Val Ser
 100 105 110

Ala Ala Thr Ser Lys Lys Ala Ser Pro Val Val Thr Val Arg Ser Tyr
 115 120 125

Val Val Asn
 130

<210> SEQ ID NO 55
 <211> LENGTH: 804
 <212> TYPE: DNA
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 55

atgtggtaa ttaagagaat gtggcaatt aagagcatgt tattaattaa gaacagctcg 60

ctaactaaga gcgtgtcgct aactaagagc atgtcggaaa ataagcgta ggcgcgtaaa 120

caaggctcac cttcaaaagg gagaggcttt accttaattg aagtcttggc ctcgattgct 180

atctttgcca cgctaagat ggcggcttat caggtgggta atcaggtgca gcgaagcaac 240

gagatctcta ttgagcgcag tgctcgtttg aaccaactgc aacgcagttt agtcatttta 300

gataatgatt ttcgccagat ggcggtgcca aaatttcgta ccaacgggta agaagcatca 360

tctaagctga tcttaatgaa agagtattta ttggactccg acagtgtagg catcatgttt 420

actcgtctag gttggcaca cccacaacag cagtttctc gcggtgaagt cacgaagggt 480

ggctaccgta ttaaagaaga aaccttgag cgtgtatggt ggcgttatcc cgatacacct 540

tcaggccaag aaggtgtgat taccctctg cttgatgatg ttgaaagctt ggaattcgag 600

ttttatgacg gaagccgctg ggggaaagag tggcaaaccg ataaatcact gccgaaagcg 660

gtgaggctta agctgacact gaaagactat ggtgagatag agcgtgttta tctcactccc 720

ggtggcacc tagatcagc cgatgattct tcaaacagtg actcttcagg cagtagtgag 780

gggaataatg actcatcgaa ctaa 804

<210> SEQ ID NO 56
 <211> LENGTH: 267
 <212> TYPE: PRT
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 56

Met Trp Leu Ile Lys Arg Met Trp Ser Ile Lys Ser Met Leu Leu Ile
 1 5 10 15

Lys Asn Ser Ser Leu Thr Lys Ser Val Ser Leu Thr Lys Ser Met Ser
 20 25 30

Glu Asn Lys Arg Thr Pro Arg Lys Gln Gly Leu Pro Ser Lys Gly Arg
 35 40 45

Gly Phe Thr Leu Ile Glu Val Leu Val Ser Ile Ala Ile Phe Ala Thr
 50 55 60

-continued

Leu Ser Met Ala Ala Tyr Gln Val Val Asn Gln Val Gln Arg Ser Asn
 65 70 75 80
 Glu Ile Ser Ile Glu Arg Ser Ala Arg Leu Asn Gln Leu Gln Arg Ser
 85 90 95
 Leu Val Ile Leu Asp Asn Asp Phe Arg Gln Met Ala Val Arg Lys Phe
 100 105 110
 Arg Thr Asn Gly Glu Glu Ala Ser Ser Lys Leu Ile Leu Met Lys Glu
 115 120 125
 Tyr Leu Leu Asp Ser Asp Ser Val Gly Ile Met Phe Thr Arg Leu Gly
 130 135 140
 Trp His Asn Pro Gln Gln Gln Phe Pro Arg Gly Glu Val Thr Lys Val
 145 150 155 160
 Gly Tyr Arg Ile Lys Glu Glu Thr Leu Glu Arg Val Trp Trp Arg Tyr
 165 170 175
 Pro Asp Thr Pro Ser Gly Gln Glu Gly Val Ile Thr Pro Leu Leu Asp
 180 185 190
 Asp Val Glu Ser Leu Glu Phe Glu Phe Tyr Asp Gly Ser Arg Trp Gly
 195 200 205
 Lys Glu Trp Gln Thr Asp Lys Ser Leu Pro Lys Ala Val Arg Leu Lys
 210 215 220
 Leu Thr Leu Lys Asp Tyr Gly Glu Ile Glu Arg Val Tyr Leu Thr Pro
 225 230 235 240
 Gly Gly Thr Leu Asp Gln Ala Asp Asp Ser Ser Asn Ser Asp Ser Ser
 245 250 255
 Gly Ser Ser Glu Gly Asn Asn Asp Ser Ser Asn
 260 265

<210> SEQ ID NO 57

<211> LENGTH: 1050

<212> TYPE: DNA

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 57

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atgactcadc gaactaataa gcgcttagcg acaaggtcag ccttgggacg taaacaacgt    60
gggtgcgcgc tgatcattat tttgatgcta ttggcgatca tggcaacccat tgctggcagc    120
atgtccgagc gtttgtttac gcaattcaag cgcgttgcta accaactgaa ttaccaacag    180
gcttactggt acagcattgg tgtggaagcg cttgtgcaaa acggtattag gcaaagtac    240
aaagacagtg ataccgtgaa cctaagccaa ccatgggcgt tagaagagca ggtataccca    300
ttggattatg gccaagttaa gggccgcatt gttgatgctc aggcattgtt taatcttaat    360
gccttagccc gagtggcgac cacttcaagt aaccagactc cttatntaat cacggtttgg    420
caaaccttat tggaaaacca agacgttgag ccttatcagg ctgaggttat cgcaaattca    480
acgtgggaat ttgttgatgc ggatacacga accacctett cgtctggtgt agaagacagc    540
acgtatgaag cgatgaagcc ctcttatttg gggcgcaatg gcttaatggc cgatgaatcc    600
gagctacgag cggtttatca agtcactggt gaagtgatga ataaggttcg cccctttggt    660
tgcgctctgc caaccgatga tttccgcttg aatgtgaata ctctcacgga aaaacaagca    720
ccgttatttg aagcgatggt tgccgacgag ttaagtgaat cggatgcca acagctgata    780
gataaacgcc cattedgatgg ctgggatacg gtagatgctt tcatggctga acctgccatt    840

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-continued

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gttgggtgtaa gtgccgaagt cagcaagaaa gcgaaagcat atttaactgt agatagcgcc 900
tattttgagc tagatgcaga ggtattagtt gagcagtcac gtgtacgtat acggacgctt 960
ttctatagta gtaatcgaga aacagtgcgc gtagtacgcc gtcgttttgg aggaatcagt 1020
gagcgagttt ctgaccgttc gactgagtag 1050

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<210> SEQ ID NO 58
<211> LENGTH: 349
<212> TYPE: PRT
<213> ORGANISM: Vibrio splendidus

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<400> SEQUENCE: 58

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Met Thr His Arg Thr Asn Lys Arg Leu Ala Thr Arg Ser Ala Leu Gly
1      5      10     15
Arg Lys Gln Arg Gly Val Ala Leu Ile Ile Leu Met Leu Leu Ala
20     25     30
Ile Met Ala Thr Ile Ala Gly Ser Met Ser Glu Arg Leu Phe Thr Gln
35     40     45
Phe Lys Arg Val Gly Asn Gln Leu Asn Tyr Gln Gln Ala Tyr Trp Tyr
50     55     60
Ser Ile Gly Val Glu Ala Leu Val Gln Asn Gly Ile Arg Gln Ser Tyr
65     70     75     80
Lys Asp Ser Asp Thr Val Asn Leu Ser Gln Pro Trp Ala Leu Glu Glu
85     90     95
Gln Val Tyr Pro Leu Asp Tyr Gly Gln Val Lys Gly Arg Ile Val Asp
100    105    110
Ala Gln Ala Cys Phe Asn Leu Asn Ala Leu Ala Gly Val Ala Thr Thr
115    120    125
Ser Ser Asn Gln Thr Pro Tyr Leu Ile Thr Val Trp Gln Thr Leu Leu
130    135    140
Glu Asn Gln Asp Val Glu Pro Tyr Gln Ala Glu Val Ile Ala Asn Ser
145    150    155    160
Thr Trp Glu Phe Val Asp Ala Asp Thr Arg Thr Thr Ser Ser Ser Gly
165    170    175
Val Glu Asp Ser Thr Tyr Glu Ala Met Lys Pro Ser Tyr Leu Ala Ala
180    185    190
Asn Gly Leu Met Ala Asp Glu Ser Glu Leu Arg Ala Val Tyr Gln Val
195    200    205
Thr Gly Glu Val Met Asn Lys Val Arg Pro Phe Val Cys Ala Leu Pro
210    215    220
Thr Asp Asp Phe Arg Leu Asn Val Asn Thr Leu Thr Glu Lys Gln Ala
225    230    235    240
Pro Leu Leu Glu Ala Met Phe Ala Pro Gly Leu Ser Glu Ser Asp Ala
245    250    255
Lys Gln Leu Ile Asp Lys Arg Pro Phe Asp Gly Trp Asp Thr Val Asp
260    265    270
Ala Phe Met Ala Glu Pro Ala Ile Val Gly Val Ser Ala Glu Val Ser
275    280    285
Lys Lys Ala Lys Ala Tyr Leu Thr Val Asp Ser Ala Tyr Phe Glu Leu
290    295    300
Asp Ala Glu Val Leu Val Glu Gln Ser Arg Val Arg Ile Arg Thr Leu
305    310    315    320

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-continued

Phe Tyr Ser Ser Asn Arg Glu Thr Val Thr Val Val Arg Arg Arg Phe
 325 330 335

Gly Gly Ile Ser Glu Arg Val Ser Asp Arg Ser Thr Glu
 340 345

<210> SEQ ID NO 59
 <211> LENGTH: 1248
 <212> TYPE: DNA
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 59

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gtgagcagagt ttctgaccgt tcgactgagt agcgaaccac aaagccctgt gcagtggtta      60
gtttggtcga caagccaaca agaagtgata gcaagcggtg aactgtctag ctgggaacag      120
cttgacgagt taacgcctta cgctgaaaag cgcagctgta tcgctttatt gccgggaagt      180
gaatgcttaa ttaagcgtgt tgagatcccg aaaggtgctg ctcgccagtt tgattctatg      240
ctgccgttct tattagaaga cgaagtcgca caagatatcg aagacttaca cctgactatt      300
ttagataaag atgccactca cgctaccgtg tgtggtgtgg atcgtgaatg gctaaaacaa      360
gctttagacc tgtttcgcga agccaatata atcttccgta aggtgctacc agatacacta      420
gccgtgcctt ttgaagaaca aggcacagtg gcgttgacga tagatcagca ttggttattg      480
cgccaaggtc actctcaacg tcaaggtcac tatcaagccg tatcgatcag tgaagcatgg      540
ttaccgatgt ttttgcaaag tgattggggt gtcgctggtg aggaagagca agcgacgact      600
atcttcagct ataccgcgat gccgagcgac gacgttcaac agcaaagcgg cctcgagtgg      660
caagcaaagc ctgcggaatt ggtgatgtct ttattgagtc agcaagcgat cacaagcggc      720
gtaaatttac tgactggcac ctttaaaacc aatcttcat tcagtaata ttggcgtggt      780
tggcagaaag tggcgattgc tgcttgtttg ctggtggccg tgattgtgac tcagcaagtg      840
ttgaagggtc agcaatacga agcgcaagca caagcctacc gcatggagag tgagcgtatc      900
tttagagctg tgctgcctgg caaacaacgc attccgaccg tgagttacct caagcgtcag      960
atgaatgatg aagctaagaa ataccggtgt tcaggcgaag gtgattcttt acttggttgg     1020
ttagctttgc tgctgaaac cttagggcaa gtgaagacga tcgaagtga aagcattcgc     1080
tacgatggca accgttctga ggttcgactg caggetaaaa gttctgactt ccaacacttt     1140
gagaccgcaa gggtagaagt cgaagagaag tttgtcgttg agcaagggcc attgaaccgt     1200
aatggcgatg ccgtatttgg cagttttact cttaaaccce atcaataa     1248

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<210> SEQ ID NO 60
 <211> LENGTH: 415
 <212> TYPE: PRT
 <213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 60

Met Ser Glu Phe Leu Thr Val Arg Leu Ser Ser Glu Pro Gln Ser Pro
 1 5 10 15

Val Gln Trp Leu Val Trp Ser Thr Ser Gln Gln Glu Val Ile Ala Ser
 20 25 30

Gly Glu Leu Ser Ser Trp Glu Gln Leu Asp Glu Leu Thr Pro Tyr Ala
 35 40 45

Glu Lys Arg Ser Cys Ile Ala Leu Leu Pro Gly Ser Glu Cys Leu Ile
 50 55 60

-continued

Lys Arg Val Glu Ile Pro Lys Gly Ala Ala Arg Gln Phe Asp Ser Met
 65 70 75 80
 Leu Pro Phe Leu Leu Glu Asp Glu Val Ala Gln Asp Ile Glu Asp Leu
 85 90 95
 His Leu Thr Ile Leu Asp Lys Asp Ala Thr His Ala Thr Val Cys Gly
 100 105 110
 Val Asp Arg Glu Trp Leu Lys Gln Ala Leu Asp Leu Phe Arg Glu Ala
 115 120 125
 Asn Ile Ile Phe Arg Lys Val Leu Pro Asp Thr Leu Ala Val Pro Phe
 130 135 140
 Glu Glu Gln Gly Ile Ser Ala Leu Gln Ile Asp Gln His Trp Leu Leu
 145 150 155 160
 Arg Gln Gly His Ser Gln Arg Gln Gly His Tyr Gln Ala Val Ser Ile
 165 170 175
 Ser Glu Ala Trp Leu Pro Met Phe Leu Gln Ser Asp Trp Val Val Ala
 180 185 190
 Gly Glu Glu Glu Gln Ala Thr Thr Ile Phe Ser Tyr Thr Ala Met Pro
 195 200 205
 Ser Asp Asp Val Gln Gln Gln Ser Gly Leu Glu Trp Gln Ala Lys Pro
 210 215 220
 Ala Glu Leu Val Met Ser Leu Leu Ser Gln Gln Ala Ile Thr Ser Gly
 225 230 235 240
 Val Asn Leu Leu Thr Gly Thr Phe Lys Thr Lys Ser Ser Phe Ser Lys
 245 250 255
 Tyr Trp Arg Val Trp Gln Lys Val Ala Ile Ala Ala Cys Leu Leu Val
 260 265 270
 Ala Val Ile Val Thr Gln Gln Val Leu Lys Val Gln Gln Tyr Glu Ala
 275 280 285
 Gln Ala Gln Ala Tyr Arg Met Glu Ser Glu Arg Ile Phe Arg Ala Val
 290 295 300
 Leu Pro Gly Lys Gln Arg Ile Pro Thr Val Ser Tyr Leu Lys Arg Gln
 305 310 315 320
 Met Asn Asp Glu Ala Lys Lys Tyr Gly Gly Ser Gly Glu Gly Asp Ser
 325 330 335
 Leu Leu Gly Trp Leu Ala Leu Leu Pro Glu Thr Leu Gly Gln Val Lys
 340 345 350
 Thr Ile Glu Val Glu Ser Ile Arg Tyr Asp Gly Asn Arg Ser Glu Val
 355 360 365
 Arg Leu Gln Ala Lys Ser Ser Asp Phe Gln His Phe Glu Thr Ala Arg
 370 375 380
 Val Lys Leu Glu Glu Lys Phe Val Val Glu Gln Gly Pro Leu Asn Arg
 385 390 395 400
 Asn Gly Asp Ala Val Phe Gly Ser Phe Thr Leu Lys Pro His Gln
 405 410 415

<210> SEQ ID NO 61

<211> LENGTH: 489

<212> TYPE: DNA

<213> ORGANISM: *Vibrio splendidus*

<400> SEQUENCE: 61

atgagaaata tgattgaacc actccaagcg tgggtgggctt caataagtca gcggaacaa 60

-continued

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cgattagtca ttggttggtc tattttattg atactgggcg ttgtctattg gggattaata 120
caaccactta gccaacgagc cgagcttgca caaagccgca ttcaaagtga gaagcaactt 180
ctggcttggg taacggacaa agcgaatcaa gtggttgaac tacgaggcag tggtggcac 240
agtgccagtc agcctttgaa ccaatctgtg cctgcttcta tgcgccgttt taacatcgag 300
ctgatacgcg tgcaaccacg cggtgagatg ctgcaagttt ggattaagcc tgtgccattt 360
aataagtteg ttgactggct gacatacctg aaagaaaagc aggggtgtga ggttgagttt 420
atggatattg atcgctctga tagccctggg gttattgaga tcaaccgact acagtttaaa 480
cgaggtaaa 489

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<210> SEQ ID NO 62
<211> LENGTH: 162
<212> TYPE: PRT
<213> ORGANISM: Vibrio splendidus

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<400> SEQUENCE: 62

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```

Met Arg Asn Met Ile Glu Pro Leu Gln Ala Trp Trp Ala Ser Ile Ser
 1             5             10            15
Gln Arg Glu Gln Arg Leu Val Ile Gly Cys Ser Ile Leu Leu Ile Leu
          20             25             30
Gly Val Val Tyr Trp Gly Leu Ile Gln Pro Leu Ser Gln Arg Ala Glu
          35             40             45
Leu Ala Gln Ser Arg Ile Gln Ser Glu Lys Gln Leu Leu Ala Trp Val
          50             55             60
Thr Asp Lys Ala Asn Gln Val Val Glu Leu Arg Gly Ser Gly Gly Ile
 65             70             75             80
Ser Ala Ser Gln Pro Leu Asn Gln Ser Val Pro Ala Ser Met Arg Arg
          85             90             95
Phe Asn Ile Glu Leu Ile Arg Val Gln Pro Arg Gly Glu Met Leu Gln
          100            105            110
Val Trp Ile Lys Pro Val Pro Phe Asn Lys Phe Val Asp Trp Leu Thr
          115            120            125
Tyr Leu Lys Glu Lys Gln Gly Val Glu Val Glu Phe Met Asp Ile Asp
          130            135            140
Arg Ser Asp Ser Pro Gly Val Ile Glu Ile Asn Arg Leu Gln Phe Lys
          145            150            155            160
Arg Gly

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<210> SEQ ID NO 63
<211> LENGTH: 780
<212> TYPE: DNA
<213> ORGANISM: Vibrio splendidus

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<400> SEQUENCE: 63

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gtgaaacgcg gtttatcttt caaatacggc ctgttattca gcgtcatttt tatcgttttt 60
ttctcggtaa gcttgtgtgct gcatttgctt gccgcttttg ctctcaagca tgcaccgctc 120
gtgcgtgggt taagcattga aggcgttgag ggcaccgttt ggcaaggctc cgctaacaat 180
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<210> SEQ ID NO 64
<211> LENGTH: 259
<212> TYPE: PRT
<213> ORGANISM: Vibrio splendidus

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<400> SEQUENCE: 64

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20          25          30
Phe Ala Leu Lys His Ala Pro Val Val Arg Gly Leu Ser Ile Glu Gly
35          40          45
Val Glu Gly Thr Val Trp Gln Gly Arg Ala Asn Asn Ile Ala Trp Gln
50          55          60
Arg Val Asn Tyr Gly Ser Val Gln Trp Asp Phe Gln Phe Ser Lys Leu
65          70          75          80
Phe Gln Ala Lys Ala Glu Leu Ala Val Arg Phe Gly Arg Asn Ser Asp
85          90          95
Met Asn Leu Ser Gly Lys Gly Arg Val Gly Tyr Ser Met Ser Gly Ala
100         105        110
Tyr Ala Glu Asn Leu Val Ala Ser Met Pro Ala Ser Asn Val Met Lys
115        120        125
Tyr Ala Pro Ala Ile Pro Val Pro Val Ser Ile Ala Gly Gln Val Glu
130        135        140
Leu Thr Ile Lys His Ala Val His Ala Gln Pro Trp Cys Gln Ser Gly
145        150        155        160
Glu Gly Thr Leu Ala Trp Ser Gly Ala Ala Val Asp Ser Pro Val Gly
165        170        175
Ser Leu Asp Leu Gly Pro Val Ile Ala Asp Ile Thr Cys Glu Asp Ser
180        185        190
Thr Ile Ala Ala Lys Gly Thr Gln Lys Ser Asp Gln Val Asp Ser Glu
195        200        205
Phe Ser Ala Ser Val Thr Pro Asn Gln Arg Tyr Thr Ser Ala Ala Trp
210        215        220
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<210> SEQ ID NO 65
<211> LENGTH: 10967
<212> TYPE: DNA

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<213> ORGANISM: *Erwinia carotovora* subsp. *Atroseptica* SCRI1043

<400> SEQUENCE: 65

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<210> SEQ ID NO 66

<211> LENGTH: 2582

<212> TYPE: DNA

<213> ORGANISM: *Erwinia carotovora* subsp. *Atroseptica* SCRI1043

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| tgtttctata ataccgatat gacaggcgtc tcgctgaga tttgtggcct gatttttgaa | 300 |
| caaccggtgt cggggtgacc gattcgtcgg acgttcagta atgtcagggt atcgaagcgt | 360 |
| atgctgtgtt ggctgcaaat tcttcatgat aagttctaag gatttacgga tggccaaagg | 420 |
| taataagatc cccctaactt ttcataccta ccaggatgca gcaaccggca ccgaagtgtt | 480 |
| gcgtttaacc cgcctcgatg ttatctgcca ccggaattat ttctaccaga agtgtttctt | 540 |
| caatgacggt agcaactgct tgtttggcgc tgcatttgat ggcccatgga actactatct | 600 |
| gctgatttta aaagagcaga acgccacaca gttgacggaa ggcaaaggcg acaatacttt | 660 |
| tggtggtttc ctgtctccga atgacgatgc gctatattac gttaaaaata cccgtaattt | 720 |
| gatgctgtgc gatctgacta cgctggaaga gaaaacgatt tatcagggtc ctgacgattg | 780 |
| ggtcggctac ggtacttggg ttgccaaact cgattgcacc aaaatggtcg gtattgagat | 840 |
| caagaaagaa gactggaagc cactgaccga ttggaaaaaa ttcaggagt tctactcac | 900 |
| taatccttgc tgtcgtctga ttccgctcga tttggtaacg ggcaagcgg agactatcct | 960 |
| tcaggaaaac cagtggctgg gtcacccaat ctaccgtcca ggtgatgaca acacggttgc | 1020 |
| ttctgtcac gaaggcccgc atgacctggt tgatgctcgt atgtggttca tcaacgaaga | 1080 |
| tggcaccaac atgcccgaag tgaaagagca tgcagaaggc gaaagctgca cccacgaatt | 1140 |
| ttgggtgccg gatggctccg cgatgattta tgtctcttat cttaaagacg ataccaaccg | 1200 |
| ttatattcgc agcatcgatc ccgttacgct ggaagatcgc caactgcgtg taatgccgcc | 1260 |
| gtgttctcac ctgatgagta actatgatgg cactctgttg gtcggtgatg gttccgatgc | 1320 |
| accggtcgac gtgcaggatg atggtggcta caaaattgag aacgatccgt tctgtatgt | 1380 |
| tttcaacctg aaaactggca aagaacatcg tattgcccag cacaatacat cctgggaagt | 1440 |
| gttggaaggg gaccgtcagg tcaactcacc gcaccctct ttcacgccgg ataataaaca | 1500 |
| agttctgttt acttctgacg tagatggaac acctgcgttg tatctggcga aggttctctga | 1560 |
| ttcagtctgg aactaataat actaataaat ccgctcacg tttcatggcg cggattat | 1620 |
| taaaatattt acttacatat tattttatta agtctctgac gcggttattt ctcaaaacta | 1680 |
| acttgattat cgttgtgtgt ccattgccat aatcaaagcg ttcctttat actaaaacca | 1740 |
| ttgttctatt ttttttaaaa caaaaaaac tgagtagggt aaccacaaaa atggctagt | 1800 |
| cagatttaga taaacaaccc gattccgtgt cgtccgtttt aaagggtttt ggtattttgc | 1860 |
| aggeattagg tgaagagaga gaaattggta ttaccgagct ttctcagcga gtcgatgt | 1920 |
| ctaagagtac cgtttaccgt ttcttgcaga cgatgaaatc cctgggctat gtcgcccagg | 1980 |
| aagggtgaate agagaagat tcgctaaccg tcaagttggt tgaactgggt gcaaaagcat | 2040 |
| tgcagaacgt agacttaatc cgcagtgcgg atatacagat gcgaggttg tctgtgctga | 2100 |
| cgcggaaac gattcacctt ggcgcgttgg atgaagcgg catcgtttat atccacaaga | 2160 |
| ttgattctat gtataacctg cgtatgtatt cgcgcategg tcgcccgaat ccaactacaca | 2220 |
| gtaccgcaat tggtaaagtg ttgctggctt ggcgcgatcg cggtgaagtg gaagaggttc | 2280 |
| tgctgactgt cgaattcacg cgtagtacgc cacacacatt gtgtactgct gaagatcttc | 2340 |

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| | |
|--|------|
| tcaatcaact ggatgtcgtg cgtgagcaag gctacgggga agataaagaa gagcaggaag | 2400 |
| aagggtcgcg ttgtatcgct gtgccagtat tcgatcgttt tgggtgtggtg attgccggcc | 2460 |
| tcagtatttc ctccccaacg attcgttttt cagaagaaaa caaacacgaa tatgtggcca | 2520 |
| tgctgcacac cgcagctaga aatatctctg agcaaatggg ctaccacaat ttccctttct | 2580 |
| ga | 2582 |

<210> SEQ ID NO 67

<211> LENGTH: 2331

<212> TYPE: DNA

<213> ORGANISM: Agrobacterium tumefaciens

<400> SEQUENCE: 67

| | |
|--|------|
| atgcgccctc ctgccccggc catctccaga cagacacttc tcgatgaacc ccgccccggc | 60 |
| tcattgacca ttggctacga gccgagcgaa gaagcacaac cgacggagaa ccctccgcgc | 120 |
| ttttcatggc taccgcgat tgaacgagc gcgcgttacg tgctgcgcat ttccgaccgat | 180 |
| cccgttttta cagacaaaaa aacgctcgtc ttccgaggatc tcgctcggaa tttcttcacc | 240 |
| ccggatgaag cactgcgcga cggccattat cactggtggt atgctgatg ggatcagaaa | 300 |
| tccgcaacag cgcattccaa ctggagcacc gtacgcagtt tcgagatcag tgaagcactg | 360 |
| ccgaaaaacg cgtctcccg caggtctgcc cgccatgctg ccgcgcaaac cagccaccct | 420 |
| cggtctgtgc tcaactccga gcaattgagt gccttcgccc atgcccgttc gaaggacccc | 480 |
| aaccattgtg gctgggccc gttttacgaa aaatcggtcg agccgtggct cgagcggccg | 540 |
| gtcatgccgg aaccgcagcc ctatcccaac aacacgcgtg tcgccacgct ctggcggcag | 600 |
| atgtatatag actgccagga agtgatctat gcgatccggc acctggccat tgcgggccgc | 660 |
| gtgctcggac ggcagcact tctcgatgca tcccgcaaat ggctgctggc cgtcgcggcc | 720 |
| tgggacacga aaggtcggac ctcacgcgcc tataatgacg aggcgggggt ccgctcgtc | 780 |
| gtcgcactcg cctgggggta tgactggctg tacgaccatc tgagcgaaga cgaacgcagg | 840 |
| accgtgcgat ccgtttctt cgaacggagc cgggaagtgg ccgatcatgt catcgcacac | 900 |
| gcccgcattc acgtctttcc ctatgacagc catgcggtgc gctcgtttc ggctgtattg | 960 |
| acgcccgcct gcatcgcact tcaggagaaa agcgcagagg ctggcgaatg gctcgaactat | 1020 |
| accgtcgaat tccttgccc gctctattct ccctgggccc gaaccgatgg tggttggggc | 1080 |
| gaaggtcccg attactggat gaccggcatg gcctatctca tcgaggccgc caatctgatc | 1140 |
| cgctctata ttggttatga cctctatcaa cggccgtttt tccagaatac cggctcgttc | 1200 |
| ccgctttaca ccaaggccc gggaaaccgc cgcgccaact tcggcgacga ctccaccctt | 1260 |
| ggcgaccttc ccggcctgaa gctgggatac aacgtccggc aattcgcggc cgtcacccgc | 1320 |
| aatggccatt accagtggta tttcgatcac atcaaggccg atgacgacagg cacggaaatg | 1380 |
| gccttttaca attacggctg gtgggacctc aacttcgacg atctcgtota tcgccacgat | 1440 |
| taccgcaggg tggaaagcgt gtctccgcc gacctgccgg cactcgcgtt ttccgatgat | 1500 |
| attggttggg cgaccatcca aaaagacatg gaagaccggc accggcacct gcagttcgtc | 1560 |
| ttcaaatcca gcccttacgg ttcgctcagc cacagtcaag gcgaccagaa tgcctttgtg | 1620 |
| ctttatgcc atggcgagga tctggogac cagtcgggtt attacgtggc gttcaattcg | 1680 |
| cagatgcac tgaattggcg gcgtcagaca cggctcgaata atgcccgtct gatcggcggc | 1740 |

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aaaggccaat atgcggaaaa ggacaaggcg cttgcacgcc gcgccgccgg cgcacatcgtc 1800
tcggtggagg aacagcccgg ccattgttcgt atcgtcggcg atgcaaccgc cgctaccag 1860
gttgcaaac cgctggttca aaagtgctg cgcgaaaccc acttcgtaa tgacagctat 1920
ttctgtattg tcgacgaagt cgaatgttcg gaaccccagg aactgcaatg gctttgccat 1980
acactcggag cgccgcagac cggcaggta agcttccgct acaatggccg gaaagccggt 2040
ttctacggac agttcgttta ctcttcgggc ggcacgccgc aaatcagcgc cgtggagggt 2100
tttcccgata tcgacccgaa agaattcgaa gggctcgaca tacaccacca tgtctgcgcc 2160
acggttccgg ccgccaccgc gcatcgcctt gtcacccttc tgggtgcctta cagcctgaag 2220
gagccgaagc gcatcttcag ctctcagat gatcagggtt tttccaccga catctacttc 2280
agtgatgtcg atgacgagcg tttcaagctc tcccttccca agcagttcta a 2331

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<210> SEQ ID NO 68

<211> LENGTH: 776

<212> TYPE: PRT

<213> ORGANISM: Agrobacterium tumefaciens

<400> SEQUENCE: 68

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Met Arg Pro Ser Ala Pro Ala Ile Ser Arg Gln Thr Leu Leu Asp Glu
1          5          10          15
Pro Arg Pro Gly Ser Leu Thr Ile Gly Tyr Glu Pro Ser Glu Glu Ala
20        25        30
Gln Pro Thr Glu Asn Pro Pro Arg Phe Ser Trp Leu Pro Asp Ile Asp
35        40        45
Asp Gly Ala Arg Tyr Val Leu Arg Ile Ser Thr Asp Pro Gly Phe Thr
50        55        60
Asp Lys Lys Thr Leu Val Phe Glu Asp Leu Ala Trp Asn Phe Phe Thr
65        70        75        80
Pro Asp Glu Ala Leu Pro Asp Gly His Tyr His Trp Cys Tyr Ala Leu
85        90        95
Trp Asp Gln Lys Ser Ala Thr Ala His Ser Asn Trp Ser Thr Val Arg
100       105       110
Ser Phe Glu Ile Ser Glu Ala Leu Pro Lys Thr Pro Leu Pro Gly Arg
115       120       125
Ser Ala Arg His Ala Ala Ala Gln Thr Ser His Pro Arg Leu Trp Leu
130       135       140
Asn Ser Glu Gln Leu Ser Ala Phe Ala Asp Ala Val Ala Lys Asp Pro
145       150       155       160
Asn His Cys Gly Trp Ala Glu Phe Tyr Glu Lys Ser Val Glu Pro Trp
165       170       175
Leu Glu Arg Pro Val Met Pro Glu Pro Gln Pro Tyr Pro Asn Asn Thr
180       185       190
Arg Val Ala Thr Leu Trp Arg Gln Met Tyr Ile Asp Cys Gln Glu Val
195       200       205
Ile Tyr Ala Ile Arg His Leu Ala Ile Ala Gly Arg Val Leu Gly Arg
210       215       220
Asp Asp Leu Leu Asp Ala Ser Arg Lys Trp Leu Leu Ala Val Ala Ala
225       230       235       240
Trp Asp Thr Lys Gly Ala Thr Ser Arg Ala Tyr Asn Asp Glu Ala Gly
245       250       255

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Phe Arg Val Val Val Ala Leu Ala Trp Gly Tyr Asp Trp Leu Tyr Asp
 260 265 270
His Leu Ser Glu Asp Glu Arg Arg Thr Val Arg Ser Val Leu Leu Glu
 275 280 285
Arg Thr Arg Glu Val Ala Asp His Val Ile Ala His Ala Arg Ile His
 290 295 300
Val Phe Pro Tyr Asp Ser His Ala Val Arg Ser Leu Ser Ala Val Leu
305 310 315 320
Thr Pro Ala Cys Ile Ala Leu Gln Gly Glu Ser Asp Glu Ala Gly Glu
 325 330 335
Trp Leu Asp Tyr Thr Val Glu Phe Leu Ala Thr Leu Tyr Ser Pro Trp
 340 345 350
Ala Gly Thr Asp Gly Gly Trp Ala Glu Gly Pro His Tyr Trp Met Thr
 355 360 365
Gly Met Ala Tyr Leu Ile Glu Ala Ala Asn Leu Ile Arg Ser Tyr Ile
 370 375 380
Gly Tyr Asp Leu Tyr Gln Arg Pro Phe Phe Gln Asn Thr Gly Arg Phe
385 390 395 400
Pro Leu Tyr Thr Lys Ala Pro Gly Thr Arg Arg Ala Asn Phe Gly Asp
 405 410 415
Asp Ser Thr Leu Gly Asp Leu Pro Gly Leu Lys Leu Gly Tyr Asn Val
 420 425 430
Arg Gln Phe Ala Gly Val Thr Gly Asn Gly His Tyr Gln Trp Tyr Phe
 435 440 445
Asp His Ile Lys Ala Asp Ala Thr Gly Thr Glu Met Ala Phe Tyr Asn
 450 455 460
Tyr Gly Trp Trp Asp Leu Asn Phe Asp Asp Leu Val Tyr Arg His Asp
465 470 475 480
Tyr Pro Gln Val Glu Ala Val Ser Pro Ala Asp Leu Pro Ala Leu Ala
 485 490 495
Val Phe Asp Asp Ile Gly Trp Ala Thr Ile Gln Lys Asp Met Glu Asp
 500 505 510
Pro Asp Arg His Leu Gln Phe Val Phe Lys Ser Ser Pro Tyr Gly Ser
 515 520 525
Leu Ser His Ser His Gly Asp Gln Asn Ala Phe Val Leu Tyr Ala His
 530 535 540
Gly Glu Asp Leu Ala Ile Gln Ser Gly Tyr Tyr Val Ala Phe Asn Ser
545 550 555 560
Gln Met His Leu Asn Trp Arg Arg Gln Thr Arg Ser Lys Asn Ala Val
 565 570 575
Leu Ile Gly Gly Lys Gly Gln Tyr Ala Glu Lys Asp Lys Ala Leu Ala
 580 585 590
Arg Arg Ala Ala Gly Arg Ile Val Ser Val Glu Glu Gln Pro Gly His
 595 600 605
Val Arg Ile Val Gly Asp Ala Thr Ala Ala Tyr Gln Val Ala Asn Pro
 610 615 620
Leu Val Gln Lys Val Leu Arg Glu Thr His Phe Val Asn Asp Ser Tyr
625 630 635 640
Phe Val Ile Val Asp Glu Val Glu Cys Ser Glu Pro Gln Glu Leu Gln
 645 650 655
Trp Leu Cys His Thr Leu Gly Ala Pro Gln Thr Gly Arg Ser Ser Phe

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| | | | | | | | | | | | | | | | | | | | |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|--|--|--|--|
| | 660 | | 665 | | 670 | | | | | | | | | | | | | | |
| Arg | Tyr | Asn | Gly | Arg | Lys | Ala | Gly | Phe | Tyr | Gly | Gln | Phe | Val | Tyr | Ser | | | | |
| | 675 | | | | | | 680 | | | | | 685 | | | | | | | |
| Ser | Gly | Gly | Thr | Pro | Gln | Ile | Ser | Ala | Val | Glu | Gly | Phe | Pro | Asp | Ile | | | | |
| | 690 | | | | | 695 | | | | | 700 | | | | | | | | |
| Asp | Pro | Lys | Glu | Phe | Glu | Gly | Leu | Asp | Ile | His | His | His | Val | Cys | Ala | | | | |
| 705 | | | | | 710 | | | | | 715 | | | | 720 | | | | | |
| Thr | Val | Pro | Ala | Ala | Thr | Arg | His | Arg | Leu | Val | Thr | Leu | Leu | Val | Pro | | | | |
| | | | | 725 | | | | | 730 | | | | | 735 | | | | | |
| Tyr | Ser | Leu | Lys | Glu | Pro | Lys | Arg | Ile | Phe | Ser | Phe | Ile | Asp | Asp | Gln | | | | |
| | | | 740 | | | | | 745 | | | | | 750 | | | | | | |
| Gly | Phe | Ser | Thr | Asp | Ile | Tyr | Phe | Ser | Asp | Val | Asp | Asp | Glu | Arg | Phe | | | | |
| | | 755 | | | | | 760 | | | | | 765 | | | | | | | |
| Lys | Leu | Ser | Leu | Pro | Lys | Gln | Phe | | | | | | | | | | | | |
| | 770 | | | | | 775 | | | | | | | | | | | | | |

<210> SEQ ID NO 69
 <211> LENGTH: 1068
 <212> TYPE: DNA
 <213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 69

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atgttcacaa cgtccgccta tgcctgcat gacggctctt cgccgatgaa gctcgcgacc 60
atcaggcgcc gcatgcccg tccgcgcat gtcgaaatcg agatagaatt ctgtggcgtc 120
tgccactcgg acatccatac ggcccgcagc gaatggccgg gctccctcta cccttgcgtc 180
cccggccaag aatcgctcgg ccgtgtcggg cgggtggggc cgcaagtca cgggttcaag 240
acgggtgacc gcgtcggtgt cggtgtatc gtcgatagct gcccgcaatg cgcaagctgc 300
gccgaagggc tggagcaata ttgcgaaaac ggcatgaccg gcaacctataa ctcccctgac 360
aaggcgatgg gggcgggcgc gcatacgctt ggccgctatt ccgcccattg ggtggtggat 420
gaccgctatg tgctcaatat tcccgaagg ctcgatccgg cggcagcagc accgctactc 480
tgcgctggta tcaccaccta ctgcgctg cgccactgga atgccggccc cggcaaacgc 540
gtcggcgtcg tcggtctggg cggcctcggc catatggccg tcaagctcgc caatgccatg 600
ggtgcgactg tcgtgatgat caccacctcg cccggcaagg cggaggatgc caaaaaactc 660
ggcgcacacg aggtgatcat ctcccgcgat gcggagcaga tgaagaaggc tacctcgagc 720
ctcgatctca tcatcgatgc tgcgcccgc gaccacgaca tcgacgcta tctggcgtg 780
ctgaaacgcg atggcgcgct ggtgcagggt ggcgcgccgg aaaagccact ttcggtgatg 840
gccttcagcc tcatccccgg ccgcaagacc tttgccggct cgatgatcgg cggatttccc 900
gagactcagg aatcgctgga tttctgcgcc gaaaaaggca tcgccggcga aatcgagatg 960
atcgatatcg atcagatcaa tgacgcttat gaacgatga taaaagcga tgtgcgttat 1020
cgtttcgtca ttgatatgaa gagcctgccg cgccagaagg ccgctcga 1068
    
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<210> SEQ ID NO 70
 <211> LENGTH: 355
 <212> TYPE: PRN
 <213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 70

Met Phe Thr Thr Ser Ala Tyr Ala Cys Asp Asp Gly Ser Ser Pro Met

-continued

| 1 | 5 | 10 | 15 |
|-----|-----|-----|-----|
| Lys | Leu | Ala | Thr |
| | 20 | Arg | Arg |
| | | Arg | Asp |
| | | | 25 |
| | | Pro | Gly |
| | | | Pro |
| | | | Arg |
| | | | Asp |
| | | | 30 |
| | | | Val |
| | | | Glu |
| Ile | Glu | Ile | Glu |
| | 35 | Phe | Cys |
| | | Gly | Val |
| | | | 40 |
| | | Cys | His |
| | | Ser | Asp |
| | | | 45 |
| | | Ile | His |
| | | Thr | Ala |
| Arg | Ser | Glu | Trp |
| | 50 | Pro | Gly |
| | | Ser | Leu |
| | | | 55 |
| | | Tyr | Pro |
| | | Cys | Val |
| | | | 60 |
| | | Pro | Gly |
| | | His | Glu |
| Ile | Val | Gly | Arg |
| | 65 | Val | Gly |
| | | Arg | Val |
| | | | 70 |
| | | Ala | Gln |
| | | | 75 |
| | | Val | Thr |
| | | Arg | Phe |
| | | | 80 |
| | | Lys | |
| Thr | Gly | Asp | Arg |
| | | | 85 |
| | | Val | Gly |
| | | Val | Gly |
| | | | 90 |
| | | Cys | Ile |
| | | | 95 |
| | | Val | Asp |
| | | Ser | Cys |
| | | | 95 |
| Cys | Ala | Ser | Cys |
| | 100 | Ala | Glu |
| | | Gly | Leu |
| | | | 105 |
| | | Gln | Tyr |
| | | Cys | Glu |
| | | | 110 |
| | | Asn | Gly |
| | | | Met |
| Thr | Gly | Thr | Tyr |
| | 115 | Asn | Ser |
| | | | 120 |
| | | Pro | Asp |
| | | Lys | Ala |
| | | | Met |
| | | | 125 |
| | | Gly | Gly |
| | | Gly | Ala |
| | | | His |
| Thr | Leu | Gly | Gly |
| | 130 | Tyr | Ser |
| | | | 135 |
| | | Ala | His |
| | | Val | Val |
| | | | Val |
| | | | 140 |
| | | Asp | Asp |
| | | Arg | Tyr |
| | | | Val |
| Leu | Asn | Ile | Pro |
| | 145 | Glu | Gly |
| | | Leu | Asp |
| | | | 150 |
| | | Pro | Ala |
| | | | 155 |
| | | Ala | Ala |
| | | Ala | Pro |
| | | | Leu |
| | | | Leu |
| | | | 160 |
| Cys | Ala | Gly | Ile |
| | | | 165 |
| | | Thr | Thr |
| | | Tyr | Ser |
| | | | 170 |
| | | Pro | Leu |
| | | | Arg |
| | | | His |
| | | | Trp |
| | | | Asn |
| | | | Ala |
| | | | Gly |
| | | | 175 |
| Pro | Gly | Lys | Arg |
| | | | 180 |
| | | Val | Gly |
| | | Val | Val |
| | | | 185 |
| | | Gly | Leu |
| | | | Gly |
| | | | Gly |
| | | | Leu |
| | | | Gly |
| | | | His |
| | | | Met |
| | | | 190 |
| Ala | Val | Lys | Leu |
| | | | 195 |
| | | Ala | Asn |
| | | Ala | Met |
| | | | 200 |
| | | Gly | Ala |
| | | Thr | Val |
| | | | Val |
| | | | Met |
| | | | Ile |
| | | | Thr |
| | | | 205 |
| Thr | Ser | Pro | Gly |
| | 210 | Lys | Ala |
| | | | Glu |
| | | | 215 |
| | | Asp | Ala |
| | | Lys | Lys |
| | | | Leu |
| | | | Gly |
| | | | Ala |
| | | | His |
| | | | Glu |
| | | | 220 |
| Val | Ile | Ile | Ser |
| | 225 | Arg | Asp |
| | | | Ala |
| | | Glu | Gln |
| | | | Met |
| | | | Lys |
| | | | Lys |
| | | | Ala |
| | | | Thr |
| | | | Ser |
| | | | Ser |
| | | | 240 |
| Leu | Asp | Leu | Ile |
| | | | 245 |
| | | Ile | Asp |
| | | Ala | Val |
| | | | Ala |
| | | | Ala |
| | | | 250 |
| | | Asp | His |
| | | | Asp |
| | | | Ile |
| | | | Asp |
| | | | Ala |
| | | | 255 |
| Tyr | Leu | Ala | Leu |
| | | | 260 |
| | | Leu | Lys |
| | | Arg | Asp |
| | | | Gly |
| | | | Ala |
| | | | Leu |
| | | | Val |
| | | | Gln |
| | | | Val |
| | | | Gly |
| | | | Ala |
| | | | 270 |
| Pro | Glu | Lys | Pro |
| | | | 275 |
| | | Leu | Ser |
| | | | Val |
| | | | Met |
| | | | 280 |
| | | Ala | Phe |
| | | | Ser |
| | | | Leu |
| | | | Ile |
| | | | Pro |
| | | | Glu |
| | | | Thr |
| | | | Gln |
| | | | Glu |
| | | | 290 |
| Met | Leu | Asp | Phe |
| | 305 | Cys | Ala |
| | | | Glu |
| | | | Lys |
| | | | Gly |
| | | | Ile |
| | | | Ala |
| | | | Gly |
| | | | Glu |
| | | | Ile |
| | | | Glu |
| | | | Met |
| | | | 320 |
| Ile | Asp | Ile | Asp |
| | | | Gln |
| | | | Ile |
| | | | Asn |
| | | | Asp |
| | | | Ala |
| | | | Tyr |
| | | | Glu |
| | | | Arg |
| | | | Met |
| | | | Ile |
| | | | Lys |
| | | | Ser |
| | | | 335 |
| Asp | Val | Arg | Tyr |
| | | | 340 |
| | | Arg | Phe |
| | | Val | Ile |
| | | | Asp |
| | | | Met |
| | | | Lys |
| | | | Ser |
| | | | Leu |
| | | | Pro |
| | | | Arg |
| | | | Gln |
| | | | 350 |
| Lys | Ala | Ala | |
| | | | 355 |

<210> SEQ ID NO 71

<211> LENGTH: 1047

<212> TYPE: DNA

<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 71

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atggctattg caagagggta tgctgcgacc gacgcgtcga agccgcttac cccgttcacc   60
ttcgaacgcc gcgagccgaa tgatgacgac gtcgtcatcg atatcaaata tgcgggcac   120
tgccactcgg acatccacac cgtccgcaac gaatggcaca atgccgttta cccgatcgtt   180
ccgggccacg aaatcgccgg tgctgtcggg gccgttggtt ccaaggtcac gcggttcaag   240
gtcggcgacc atgtcggcgt cggctgcttt gtcgattcct gcgttggetg cgcaccccg   300
gatgtcgaca atgagcagta tatgccgggt ctcgtcgaga cctacaattc cgttgaacgg   360
gacggcaaga gcgcgacca gggcggttat tccgaccata tcgtggtcag ggaagactac   420
gtcctgtcca tcccggacaa cctgcogctc gatgcctcgg cgcgcttct ctgcgcccgg   480
atcacgctct attcgcgct gcagcactgg aatgcaggcc cggcaagaa agtggctatc   540
gtcggcatgg gtggccttgg ccacatgggc gtgaagatcg gtcgggcat gggcgctgat   600
atcaccgttc tctcgcagac gctgtcgaag aaggaagacg gcctcaagct cggcgcgaag   660
gaatattaeg ccaccagcga cgcctcgacc tttgagaaac tcgcccggcac cttcgacctg   720
atcctgtgca cagtctcggc cgaaatcgac tggaaagcct acctcaacct gctcaaggtc   780
aacggcacga tggttctgct cggcgtgccc gaacatgcca tcccggtgca cgcattctcg   840
gtcattcccg ccgcgcttc gctcgcgggt tcgatgatcg gctcgatcaa ggaaccag   900
gaaatgctgg atttctcggc caagcacgac atcgtttcgg aaatcgaaac gatcggcacc   960
aaggacgtca acgaagccta tgagcgcgtg ctgaagagcg acgtgcgtta ccgcttcgct   1020
atcgacatgg cctcgcgca cgcttga                                     1047

```

<210> SEQ ID NO 72

<211> LENGTH: 348

<212> TYPE: PRT

<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 72

```

Met Ala Ile Ala Arg Gly Tyr Ala Ala Thr Asp Ala Ser Lys Pro Leu
 1           5           10           15
Thr Pro Phe Thr Phe Glu Arg Arg Glu Pro Asn Asp Asp Asp Val Val
 20           25           30
Ile Asp Ile Lys Tyr Ala Gly Ile Cys His Ser Asp Ile His Thr Val
 35           40           45
Arg Asn Glu Trp His Asn Ala Val Tyr Pro Ile Val Pro Gly His Glu
 50           55           60
Ile Ala Gly Val Val Arg Ala Val Gly Ser Lys Val Thr Arg Phe Lys
 65           70           75           80
Val Gly Asp His Val Gly Val Gly Cys Phe Val Asp Ser Cys Val Gly
 85           90           95
Cys Ala Thr Arg Asp Val Asp Asn Glu Gln Tyr Met Pro Gly Leu Val
100          105          110
Gln Thr Tyr Asn Ser Val Glu Arg Asp Gly Lys Ser Ala Thr Gln Gly
115          120          125
Gly Tyr Ser Asp His Ile Val Val Arg Glu Asp Tyr Val Leu Ser Ile
130          135          140
Pro Asp Asn Leu Pro Leu Asp Ala Ser Ala Pro Leu Leu Cys Ala Gly
145          150          155          160
Ile Thr Leu Tyr Ser Pro Leu Gln His Trp Asn Ala Gly Pro Gly Lys
165          170          175

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-continued

Lys Val Ala Ile Val Gly Met Gly Gly Leu Gly His Met Gly Val Lys
 180 185 190

Ile Gly Ser Ala Met Gly Ala Asp Ile Thr Val Leu Ser Gln Thr Leu
 195 200 205

Ser Lys Lys Glu Asp Gly Leu Lys Leu Gly Ala Lys Glu Tyr Tyr Ala
 210 215 220

Thr Ser Asp Ala Ser Thr Phe Glu Lys Leu Ala Gly Thr Phe Asp Leu
 225 230 235 240

Ile Leu Cys Thr Val Ser Ala Glu Ile Asp Trp Asn Ala Tyr Leu Asn
 245 250 255

Leu Leu Lys Val Asn Gly Thr Met Val Leu Leu Gly Val Pro Glu His
 260 265 270

Ala Ile Pro Val His Ala Phe Ser Val Ile Pro Ala Arg Arg Ser Leu
 275 280 285

Ala Gly Ser Met Ile Gly Ser Ile Lys Glu Thr Gln Glu Met Leu Asp
 290 295 300

Phe Cys Gly Lys His Asp Ile Val Ser Glu Ile Glu Thr Ile Gly Ile
 305 310 315 320

Lys Asp Val Asn Glu Ala Tyr Glu Arg Val Leu Lys Ser Asp Val Arg
 325 330 335

Tyr Arg Phe Val Ile Asp Met Ala Ser Leu Asp Ala
 340 345

<210> SEQ ID NO 73
 <211> LENGTH: 1029
 <212> TYPE: DNA
 <213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 73

```

atgactaaaa caatgaaggc ggcggttgtc cgcgcatctg gaaaaccgct gaccatcgag    60
gaagtggcaa taccgatgcc cggccccggt gaaattctca tcaactacaa ggcgacgggc    120
gtttgcaca ccgacctgca cgccgcaacg ggggattggc cggtaagcc caaccgcccc    180
ttcattcccg gacatgaagg tgcaggttac gtcgccaaga tcggcgctgg cgtcaccggc    240
atcaaggagg gcgaccgcgc cggcacgccc tggctctaca ccgcctgagg atgctgeatt    300
cctgccgta ccggctggga aaccctgtgc ccgagccaga agaactcagg ttattccgtc    360
aacggcagct ttgccgaata tggccttgcc gatccgaaat tcgtcgcccg cctgectgac    420
aatctcgatt tcggcccagc cgcacccgtg ctctgcgccc gcgttacagt ctataagggc    480
ctgaaggaaa ccgaagtcag gcccggtgaa tgggtggtca tttcaggcat tggcgggctt    540
ggccacatgg ccgtgcaata tgcgaaagcc atgggcatgc atgtggttgc cgcgatatt    600
ttcgacgaca agctggcgct tgccaaaaag ctcgagccg acgtcgtcgt caacggccgc    660
gcgctgacg cgggtggagca agtgcaaaa gcaaccggcg gcgtccatgg cgcgctggtg    720
acggcggttt caccgaaggc catggagcag gottatggt tcctcgcttc caagggcacg    780
atggcgcttg tcggtctgcc gccgggcttc atctccatc cggtggtcga cacggtgctg    840
aagcgcatca cggtcgctgg ctccatcgtc ggcacgccc aggatctgga ggaggcgttg    900
accttcgccc gtgaaggcaa ggtggccgcc cacttctcgt gggacaagct cgaaaacatc    960
aatgatctct tccatcgcat ggaagaggcc aagatcgacg gccgtatcgt cgtggatctc   1020
    
```

-continued

gccgcctga

1029

<210> SEQ ID NO 74

<211> LENGTH: 342

<212> TYPE: PRT

<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 74

```

Met Thr Lys Thr Met Lys Ala Ala Val Val Arg Ala Phe Gly Lys Pro
1      5      10      15
Leu Thr Ile Glu Glu Val Ala Ile Pro Asp Pro Gly Pro Gly Glu Ile
20     25     30
Leu Ile Asn Tyr Lys Ala Thr Gly Val Cys His Thr Asp Leu His Ala
35     40     45
Ala Thr Gly Asp Trp Pro Val Lys Pro Asn Pro Pro Phe Ile Pro Gly
50     55     60
His Glu Gly Ala Gly Tyr Val Ala Lys Ile Gly Ala Gly Val Thr Gly
65     70     75     80
Ile Lys Glu Gly Asp Arg Ala Gly Thr Pro Trp Leu Tyr Thr Ala Cys
85     90     95
Gly Cys Cys Ile Pro Cys Arg Thr Gly Trp Glu Thr Leu Cys Pro Ser
100    105    110
Gln Lys Asn Ser Gly Tyr Ser Val Asn Gly Ser Phe Ala Glu Tyr Gly
115    120    125
Leu Ala Asp Pro Lys Phe Val Gly Arg Leu Pro Asp Asn Leu Asp Phe
130    135    140
Gly Pro Ala Ala Pro Val Leu Cys Ala Gly Val Thr Val Tyr Lys Gly
145    150    155    160
Leu Lys Glu Thr Glu Val Arg Pro Gly Glu Trp Val Val Ile Ser Gly
165    170    175
Ile Gly Gly Leu Gly His Met Ala Val Gln Tyr Ala Lys Ala Met Gly
180    185    190
Met His Val Val Ala Ala Asp Ile Phe Asp Asp Lys Leu Ala Leu Ala
195    200    205
Lys Lys Leu Gly Ala Asp Val Val Val Asn Gly Arg Ala Pro Asp Ala
210    215    220
Val Glu Gln Val Gln Lys Ala Thr Gly Gly Val His Gly Ala Leu Val
225    230    235    240
Thr Ala Val Ser Pro Lys Ala Met Glu Gln Ala Tyr Gly Phe Leu Arg
245    250    255
Ser Lys Gly Thr Met Ala Leu Val Gly Leu Pro Pro Gly Phe Ile Ser
260    265    270
Ile Pro Val Phe Asp Thr Val Leu Lys Arg Ile Thr Val Arg Gly Ser
275    280    285
Ile Val Gly Thr Arg Gln Asp Leu Glu Glu Ala Leu Thr Phe Ala Gly
290    295    300
Glu Gly Lys Val Ala Ala His Phe Ser Trp Asp Lys Leu Glu Asn Ile
305    310    315    320
Asn Asp Ile Phe His Arg Met Glu Glu Gly Lys Ile Asp Gly Arg Ile
325    330    335
Val Val Asp Leu Ala Ala
340

```

-continued

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<210> SEQ ID NO 75
<211> LENGTH: 1008
<212> TYPE: DNA
<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 75
atgaccgggg cgaaccagcc ttgggaggtt caagaggttc cgttccgaa ggcagagcca    60
ggacttgtec ttgttaaaat ccacgcctcc ggcatgtgct acacggacgt gtgggagacg    120
cagggtgccc gtggcgacat ctatccgcag acccccggcc atgaggttgt cggcgagatc    180
atcgaggtcg gcgcgggcgt tcatacgcgc aaggtgggag accgggtcgg caccacctgg    240
gtgcagtcct cttgtggaag atgctcctac tgccgccaga accgtccgtt gaccggccag    300
acagccatga actgcgattc acccaggaca acgggggttcg cgacgcaagg cgggcacgca    360
gagtacatcg cgatctctgc tgaaggcaca gtgttattac ccgacgggct cgactacacg    420
gatgccgcac ccatgatgtg cgcaggctac acgacctgga gcggccttgcg cgacgccgag    480
cccaaacctg gtgacagaat tgcggtactt ggcatcggcg ggctggggca cgtcgccgtg    540
cagttctcca aagccttggg gtttgagacc atcgcgatca cgcattcacc cgacaagcac    600
aagttggcca ccgatcttgg tgcagacatc gtcgtcgcgg atggcaaaga gttattggag    660
gccggcgggtg cggacgttct tctggttacg accaacgact tcgacaccgc cgaaaaagcg    720
atggcgggcg taaggcctga cgggcgcacg gttctttgcg cgctcgactt cagcaagccg    780
ttctcgatcc cgtccgacgg caagccgttc cacatgatgc gccaacgcgt ggttgggtcc    840
acgatggcg gacagcacta tctcgccgaa atcctcgatc tcgccgcaa gggcaaggtc    900
aagccgattg tcgagacctt cgccctcgag caggcaaccg aggcatatga gcggctatcc    960
accgggaaga tgcgcttcgg gggcgtgttc cttccgcacg gcgcttga    1008

```

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<210> SEQ ID NO 76
<211> LENGTH: 335
<212> TYPE: PRT
<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 76
Met Thr Gly Ala Asn Gln Pro Trp Glu Val Gln Glu Val Pro Val Pro
1          5          10          15
Lys Ala Glu Pro Gly Leu Val Leu Val Lys Ile His Ala Ser Gly Met
20          25          30
Cys Tyr Thr Asp Val Trp Ala Thr Gln Gly Ala Gly Gly Asp Ile Tyr
35          40          45
Pro Gln Thr Pro Gly His Glu Val Val Gly Glu Ile Ile Glu Val Gly
50          55          60
Ala Gly Val His Thr Arg Lys Val Gly Asp Arg Val Gly Thr Thr Trp
65          70          75          80
Val Gln Ser Ser Cys Gly Arg Cys Ser Tyr Cys Arg Gln Asn Arg Pro
85          90          95
Leu Thr Gly Gln Thr Ala Met Asn Cys Asp Ser Pro Arg Thr Thr Gly
100         105         110
Phe Ala Thr Gln Gly Gly His Ala Glu Tyr Ile Ala Ile Ser Ala Glu
115         120         125
Gly Thr Val Leu Leu Pro Asp Gly Leu Asp Tyr Thr Asp Ala Ala Pro
130         135         140

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-continued

Met Met Cys Ala Gly Tyr Thr Thr Trp Ser Gly Leu Arg Asp Ala Glu
 145 150 155 160

Pro Lys Pro Gly Asp Arg Ile Ala Val Leu Gly Ile Gly Gly Leu Gly
 165 170 175

His Val Ala Val Gln Phe Ser Lys Ala Leu Gly Phe Glu Thr Ile Ala
 180 185 190

Ile Thr His Ser Pro Asp Lys His Lys Leu Ala Thr Asp Leu Gly Ala
 195 200 205

Asp Ile Val Val Ala Asp Gly Lys Glu Leu Leu Glu Ala Gly Gly Ala
 210 215 220

Asp Val Leu Leu Val Thr Thr Asn Asp Phe Asp Thr Ala Glu Lys Ala
 225 230 235 240

Met Ala Gly Val Arg Pro Asp Gly Arg Ile Val Leu Cys Ala Leu Asp
 245 250 255

Phe Ser Lys Pro Phe Ser Ile Pro Ser Asp Gly Lys Pro Phe His Met
 260 265 270

Met Arg Gln Arg Val Val Gly Ser Thr His Gly Gly Gln His Tyr Leu
 275 280 285

Ala Glu Ile Leu Asp Leu Ala Ala Lys Gly Lys Val Lys Pro Ile Val
 290 295 300

Glu Thr Phe Ala Leu Glu Gln Ala Thr Glu Ala Tyr Glu Arg Leu Ser
 305 310 315 320

Thr Gly Lys Met Arg Phe Arg Gly Val Phe Leu Pro His Gly Ala
 325 330 335

<210> SEQ ID NO 77
 <211> LENGTH: 1017
 <212> TYPE: DNA
 <213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 77

```

atgaccatgc atgccattca attcgtcgag aagggacgcg cegtgtctggc ggaactcccc 60
gtcgccgatc tgcgcgcggg ccatgogctc gtgcgggtca aggcttcggg gctttgccat 120
accgatatcg acgtgctgca tgcgcgttat ggcgacggtg cgttccccgt cattccgggg 180
catgaatatg ctggcgaagt cgcagccgtg gcttccgatg tgacagtctt caaggctggc 240
gaccgggttg tgcgcatcc caatctgccc tgtggcacct gcgccagctg caggaaaggg 300
ctgaccaacc tttgcagcac attgaaagct tacggcgttt ccacaaatgg cggctttgcg 360
gagttcagtg tgggtgcgtgc cgatcacctg cacggtatcg gttcgatgcc ctatcacgtc 420
gcggcgctgg ctgagccgct tgcctgtggt gtcaatggca tgcagagtgc gggatttggc 480
gagagtggcg tgggtgcgga gaatgcgctt gttttcggtg ctgggcccac cggcctgctg 540
cttgcctctg cgctgaaatc acgcggcatt gcgacggtga cgatggccga tatcaatgaa 600
agcaggctgg cctttgcccc ggacctcggg cttcagacgg cggtatccgg ctcggaagcg 660
ctctcgcggc agcggaagga gttcgatttc gtggccgatg cgaagggtat tgccccggtc 720
gccgaggcga tgatccccgt ggttgcggat ggcggcacgg cgctattctt cggcgtctgc 780
gcgccggatg cccgtatttc ggtggcaccc tttgaaatct tccggcgcca gctgaaactt 840
gtcggtctgc attcgtgaa ccgcaacata ccgcaggcgc ttgccattct ggagacggat 900
ggcgaggtca tggcgcgctt cgtttgcgac cgcttgccgc tttcgagat gctgccgttc 960
    
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 tttagcaaaa aaccgtctga tccggcgagc atgaaagtgc aattgcagc cgaatga 1017

<210> SEQ ID NO 78

<211> LENGTH: 338

<212> TYPE: PRT

<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 78

Met Thr Met His Ala Ile Gln Phe Val Glu Lys Gly Arg Ala Val Leu
 1 5 10 15
 Ala Glu Leu Pro Val Ala Asp Leu Pro Pro Gly His Ala Leu Val Arg
 20 25 30
 Val Lys Ala Ser Gly Leu Cys His Thr Asp Ile Asp Val Leu His Ala
 35 40 45
 Arg Tyr Gly Asp Gly Ala Phe Pro Val Ile Pro Gly His Glu Tyr Ala
 50 55 60
 Gly Glu Val Ala Ala Val Ala Ser Asp Val Thr Val Phe Lys Ala Gly
 65 70 75 80
 Asp Arg Val Val Val Asp Pro Asn Leu Pro Cys Gly Thr Cys Ala Ser
 85 90 95
 Cys Arg Lys Gly Leu Thr Asn Leu Cys Ser Thr Leu Lys Ala Tyr Gly
 100 105 110
 Val Ser His Asn Gly Gly Phe Ala Glu Phe Ser Val Val Arg Ala Asp
 115 120 125
 His Leu His Gly Ile Gly Ser Met Pro Tyr His Val Ala Ala Leu Ala
 130 135 140
 Glu Pro Leu Ala Cys Val Val Asn Gly Met Gln Ser Ala Gly Ile Gly
 145 150 155 160
 Glu Ser Gly Val Val Pro Glu Asn Ala Leu Val Phe Gly Ala Gly Pro
 165 170 175
 Ile Gly Leu Leu Leu Ala Leu Ser Leu Lys Ser Arg Gly Ile Ala Thr
 180 185 190
 Val Thr Met Ala Asp Ile Asn Glu Ser Arg Leu Ala Phe Ala Gln Asp
 195 200 205
 Leu Gly Leu Gln Thr Ala Val Ser Gly Ser Glu Ala Leu Ser Arg Gln
 210 215 220
 Arg Lys Glu Phe Asp Phe Val Ala Asp Ala Thr Gly Ile Ala Pro Val
 225 230 235 240
 Ala Glu Ala Met Ile Pro Leu Val Ala Asp Gly Gly Thr Ala Leu Phe
 245 250 255
 Phe Gly Val Cys Ala Pro Asp Ala Arg Ile Ser Val Ala Pro Phe Glu
 260 265 270
 Ile Phe Arg Arg Gln Leu Lys Leu Val Gly Ser His Ser Leu Asn Arg
 275 280 285
 Asn Ile Pro Gln Ala Leu Ala Ile Leu Glu Thr Asp Gly Glu Val Met
 290 295 300
 Ala Arg Leu Val Ser His Arg Leu Pro Leu Ser Glu Met Leu Pro Phe
 305 310 315 320
 Phe Thr Lys Lys Pro Ser Asp Pro Ala Thr Met Lys Val Gln Phe Ala
 325 330 335
 Ala Glu

-continued

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<210> SEQ ID NO 79
<211> LENGTH: 1044
<212> TYPE: DNA
<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 79
atgCGcgcgc tttattacga acgattcggc gagaccctg tagtCGcgtc cctgcctgat    60
ccggcaccga gCGatggcgg cgtggtgatt gCGgtgaagg caaccggcct ctgccgcagc    120
gactggcatg gctggatggg acatgacacg gatatcgtc tgccgcatgt gcccggccac    180
gagttcgccg gCGtcatctc cgcagtcggc agaaacgtca cccgcttcaa gacgggtgat    240
cgcggtaccg tgcctttcgt ctccggctgc ggccattgcc atgagtgcCG ctccggcaat    300
cagcaggctc gCGaaacgca gttccagccc ggcttcaccc attggggttc cttcgccgaa    360
tatgtcgcca tCGactatgc cGatcagaac ctCGtgCacc tgccggaatc gatgagttac    420
gccaccgcCG cCGgctcCGg ttgcccgttc gccacctct tccgggCGgt gacggatcag    480
ggacgcctga agggcggcga atggctggct gtccatggct gCGgCGgtgt cGgtctctcc    540
gccatcatga tCGgCGccg cctCGgCGca caggtcgtCG ccatcgatat tCGcgaagac    600
aagctCGaac tCGccCGgca actgggtgca accCGaacca tcaacagcCG ctccgttgcc    660
gatgtCGccg aagCGgtgCG cGacatcacc ggtggCGgCG cGcatgtgtc ggtggatgCG    720
cttggccatc cCGagacctg ctGcaattcc atcagcaacc tCGcCGgCG cGgacCGcat    780
gtcaggtgCG ggctgatgct gCGagacct gccatgCGg ccatcccat ggccCGgtg    840
atCGctcatg agctggagat ctatggcagc cagCGcatgc agCGatggCG ttacgaggac    900
atGctggcca tGatCGaaag cGgCaggctt gCGccGaaa agctgattgg cGccatatac    960
tCGtgaccg aagCGgCGct cGccctgccc ggaatggata ggttcagga gagCGgcatc    1020
agcatcatCG accggttCGa atag                                         1044

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<210> SEQ ID NO 80
<211> LENGTH: 357
<212> TYPE: PRT
<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 80
Met Asn Leu Arg Thr Asn Asp Glu Ala Met Met Arg Ala Leu Tyr Tyr
1          5          10          15
Glu Arg Phe Gly Glu Thr Pro Val Val Ala Ser Leu Pro Asp Pro Ala
20        25        30
Pro Ser Asp Gly Gly Val Val Ile Ala Val Lys Ala Thr Gly Leu Cys
35        40        45
Arg Ser Asp Trp His Gly Trp Met Gly His Asp Thr Asp Ile Arg Leu
50        55        60
Pro His Val Pro Gly His Glu Phe Ala Gly Val Ile Ser Ala Val Gly
65        70        75        80
Arg Asn Val Thr Arg Phe Lys Thr Gly Asp Arg Val Thr Val Pro Phe
85        90        95
Val Ser Gly Cys Gly His Cys His Glu Cys Arg Ser Gly Asn Gln Gln
100       105       110
Val Cys Glu Thr Gln Phe Gln Pro Gly Phe Thr His Trp Gly Ser Phe
115      120      125

```

-continued

Ala Glu Tyr Val Ala Ile Asp Tyr Ala Asp Gln Asn Leu Val His Leu
130 135 140

Pro Glu Ser Met Ser Tyr Ala Thr Ala Ala Gly Leu Gly Cys Arg Phe
145 150 155 160

Ala Thr Ser Phe Arg Ala Val Thr Asp Gln Gly Arg Leu Lys Gly Gly
165 170 175

Glu Trp Leu Ala Val His Gly Cys Gly Gly Val Gly Leu Ser Ala Ile
180 185 190

Met Ile Gly Ala Gly Leu Gly Ala Gln Val Val Ala Ile Asp Ile Ala
195 200 205

Glu Asp Lys Leu Glu Leu Ala Arg Gln Leu Gly Ala Thr Ala Thr Ile
210 215 220

Asn Ser Arg Ser Val Ala Asp Val Ala Glu Ala Val Arg Asp Ile Thr
225 230 235 240

Gly Gly Gly Ala His Val Ser Val Asp Ala Leu Gly His Pro Gln Thr
245 250 255

Cys Cys Asn Ser Ile Ser Asn Leu Arg Arg Arg Gly Arg His Val Gln
260 265 270

Val Gly Leu Met Leu Ala Asp His Ala Met Pro Ala Ile Pro Met Ala
275 280 285

Arg Val Ile Ala His Glu Leu Glu Ile Tyr Gly Ser His Gly Met Gln
290 295 300

Ala Trp Arg Tyr Glu Asp Met Leu Ala Met Ile Glu Ser Gly Arg Leu
305 310 315 320

Ala Pro Glu Lys Leu Ile Gly Arg His Ile Ser Leu Thr Glu Ala Ala
325 330 335

Val Ala Leu Pro Gly Met Asp Arg Phe Gln Glu Ser Gly Ile Ser Ile
340 345 350

Ile Asp Arg Phe Glu
355

<210> SEQ ID NO 81

<211> LENGTH: 1011

<212> TYPE: DNA

<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 81

```

atgctggcga ttttctgtga cactcccgtt caattaaccg ccaaggatct gccgaacccc   60
gtgcgcgggc aaggtgaagt cctggtacgt attcgccgga ttggcgtttg cggcacggat   120
ctgcacatct ttaccggcaa ccagccctat ctttctatc cgcggatcat gggtcacgaa   180
ctttccggca cggttgagga ggcacccgct ggcagccacc tttccgctgg cgatgtggtg   240
accataatc cctatatgtc ctgcgggaaa tgcaatgect gcctgaaggg taagagcaat   300
tgctgccgca atatcggtgt gcttggcggt catcgcgatg gcggcatggt ggaatatctg   360
agcgtgcccg agcaattcgt gctgaaggcg gaggggctga gcctcgacca ggcagccatg   420
acggaatttc tggcgatcgg tgcccacgcg gtgcgtcgcg gtgccgtcga aaaagggcaa   480
aaggtcctga tcgtcgtgtc cggcccgatc ggcattggcg ttgctgtctt tgcggttctc   540
gatggcacgg aagtgcagat gatcgacggt cgcaccgacc ggctggattt ctgcaaggac   600
cacctcgggt tcgctcatac agtcgccctc ggcgacggtg acaagatcg tctgtccgac   660
attaccgggt gcaatttctt cgatgoggtg tttgatgcga ccggcaatcc gaaagccatg   720

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gagcgcgggtt tctccttcgt cggtcacggc ggctcctatg ttctgggtgc catcgtcgcc 780
agcgatatca gcttcaacga cccggaattt cacaagcgtg agacgacgct gctcggcagc 840
cgcaacgcga cggctgatga tttcgagcgg gtgcttcgcg ccttgccgca agggaaagtg 900
ccggaggcac taatcaccca tcgcatgaca cttgccgatg ttcctcgaa gttcgcgggc 960
ctgaccgatc cgaaagccgg agtcatcaag ggcgatggtg aggtcgcgatg a 1011

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<210> SEQ ID NO 82
<211> LENGTH: 336
<212> TYPE: PRT
<213> ORGANISM: Agrobacterium tumefaciens C58

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```

<400> SEQUENCE: 82

```

```

Met Leu Ala Ile Phe Cys Asp Thr Pro Gly Gln Leu Thr Ala Lys Asp
1 5 10 15
Leu Pro Asn Pro Val Arg Gly Glu Gly Val Leu Val Arg Ile Arg
20 25 30
Arg Ile Gly Val Cys Gly Thr Asp Leu His Ile Phe Thr Gly Asn Gln
35 40 45
Pro Tyr Leu Ser Tyr Pro Arg Ile Met Gly His Glu Leu Ser Gly Thr
50 55 60
Val Glu Glu Ala Pro Ala Gly Ser His Leu Ser Ala Gly Asp Val Val
65 70 75 80
Thr Ile Ile Pro Tyr Met Ser Cys Gly Lys Cys Asn Ala Cys Leu Lys
85 90 95
Gly Lys Ser Asn Cys Cys Arg Asn Ile Gly Val Leu Gly Val His Arg
100 105 110
Asp Gly Gly Met Val Glu Tyr Leu Ser Val Pro Gln Gln Phe Val Leu
115 120 125
Lys Ala Glu Gly Leu Ser Leu Asp Gln Ala Ala Met Thr Glu Phe Leu
130 135 140
Ala Ile Gly Ala His Ala Val Arg Arg Gly Ala Val Glu Lys Gly Gln
145 150 155 160
Lys Val Leu Ile Val Gly Ala Gly Pro Ile Gly Met Ala Val Ala Val
165 170 175
Phe Ala Val Leu Asp Gly Thr Glu Val Thr Met Ile Asp Gly Arg Thr
180 185 190
Asp Arg Leu Asp Phe Cys Lys Asp His Leu Gly Val Ala His Thr Val
195 200 205
Ala Leu Gly Asp Gly Asp Lys Asp Arg Leu Ser Asp Ile Thr Gly Gly
210 215 220
Asn Phe Phe Asp Ala Val Phe Asp Ala Thr Gly Asn Pro Lys Ala Met
225 230 235 240
Glu Arg Gly Phe Ser Phe Val Gly His Gly Gly Ser Tyr Val Leu Val
245 250 255
Ser Ile Val Ala Ser Asp Ile Ser Phe Asn Asp Pro Glu Phe His Lys
260 265 270
Arg Glu Thr Thr Leu Leu Gly Ser Arg Asn Ala Thr Ala Asp Asp Phe
275 280 285
Glu Arg Val Leu Arg Ala Leu Arg Glu Gly Lys Val Pro Glu Ala Leu
290 295 300

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-continued

Ile Thr His Arg Met Thr Leu Ala Asp Val Pro Ser Lys Phe Ala Gly
305 310 315 320

Leu Thr Asp Pro Lys Ala Gly Val Ile Lys Gly Met Val Glu Val Ala
325 330 335

<210> SEQ ID NO 83
<211> LENGTH: 1005
<212> TYPE: DNA
<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 83

```

gtgaaagcct tcgtcgtcga caagtacaag aagaagggcc cgctgcgtct ggcgcacatg    60
cccaatccgg tcatcgggcg caatgatgtg ctggttcgca tccatgccac tgccatcaat    120
cttctcgaact ccaaggtgcg cgacggggaa ttcaagctgt tcctgccta tcgtcctccc    180
ttcattctcg gtcgatgatc ggccggaacg gtcacccgcg tcggcgcgaa tgtacggcag    240
ttcaagacag ggcagcaggt ttctgctcgc ccgctgata accgggtcgg aaccttcgca    300
gaaatgattg cggctgatgc cgcagacctt gcgctgaagc caacgagcct gtcctatggag    360
caggcagcgt cgatcccgcg cgtcggactg actgcctggc aggcgcttat cgaggttggc    420
aaggtaaagt ccggccagaa ggttttcctc caggccggtt ccggcggtgt cggcaccttc    480
gccatccagc ttgccaaaga tctcggcgct accgtggcca cgaccaccag cgccgcgaat    540
gccgaactgg tcaaaagcct cggcgcagat gtggtgatcg actacaagac gcaggacttc    600
gaacaggtgc tgtccggcta cgatctcgtc ctgaacagcc aggatgcca gacgctggaa    660
aagtcgttga acgtgctgag accggggcga aagctcattt cgatctccgg tccgccggat    720
gttgcccttg ccagatcgtt gaaactgaat ccgctcctgc gttttgctgt cagaatgctg    780
agccgtgggt tctgaaaaa ggcaagcaga cgcggtgctg attactcttt cctgttcatg    840
cgcgccgaag gtcagcaatt gcatgagatc gccgaactga tcgatgccgg caccatccgt    900
ccggtcgtcg acaaggtgtt tcaatttgcg cagacgcccg acgccctggc ctatgtcgag    960
accggacggg caaggggcaa ggttgtggtt acatagcat cctag                               1005

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<210> SEQ ID NO 84
<211> LENGTH: 359
<212> TYPE: PRT
<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 84

```

Met Pro Ser Leu Cys Arg Lys Pro Trp Leu Ser Ser Leu Pro Asp Leu
1      5      10      15

Ile Asn Val Ser His Trp Arg Lys Pro Val Lys Ala Phe Val Val Asp
20      25      30

Lys Tyr Lys Lys Lys Gly Pro Leu Arg Leu Ala Asp Met Pro Asn Pro
35      40      45

Val Ile Gly Ala Asn Asp Val Leu Val Arg Ile His Ala Thr Ala Ile
50      55      60

Asn Leu Leu Asp Ser Lys Val Arg Asp Gly Glu Phe Lys Leu Phe Leu
65      70      75      80

Pro Tyr Arg Pro Pro Phe Ile Leu Gly His Asp Leu Ala Gly Thr Val
85      90      95

Ile Arg Val Gly Ala Asn Val Arg Gln Phe Lys Thr Gly Asp Glu Val
100     105     110

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Phe Ala Arg Pro Arg Asp His Arg Val Gly Thr Phe Ala Glu Met Ile
 115 120 125
 Ala Val Asp Ala Ala Asp Leu Ala Leu Lys Pro Thr Ser Leu Ser Met
 130 135 140
 Glu Gln Ala Ala Ser Ile Pro Leu Val Gly Leu Thr Ala Trp Gln Ala
 145 150 155 160
 Leu Ile Glu Val Gly Lys Val Lys Ser Gly Gln Lys Val Phe Ile Gln
 165 170 175
 Ala Gly Ser Gly Gly Val Gly Thr Phe Ala Ile Gln Leu Ala Lys His
 180 185 190
 Leu Gly Ala Thr Val Ala Thr Thr Thr Ser Ala Ala Asn Ala Glu Leu
 195 200 205
 Val Lys Ser Leu Gly Ala Asp Val Val Ile Asp Tyr Lys Thr Gln Asp
 210 215 220
 Phe Glu Gln Val Leu Ser Gly Tyr Asp Leu Val Leu Asn Ser Gln Asp
 225 230 235 240
 Ala Lys Thr Leu Glu Lys Ser Leu Asn Val Leu Arg Pro Gly Gly Lys
 245 250 255
 Leu Ile Ser Ile Ser Gly Pro Pro Asp Val Ala Phe Ala Arg Ser Leu
 260 265 270
 Lys Leu Asn Pro Leu Leu Arg Phe Val Val Arg Met Leu Ser Arg Gly
 275 280 285
 Val Leu Lys Lys Ala Ser Arg Arg Gly Val Asp Tyr Ser Phe Leu Phe
 290 295 300
 Met Arg Ala Glu Gly Gln Gln Leu His Glu Ile Ala Glu Leu Ile Asp
 305 310 315 320
 Ala Gly Thr Ile Arg Pro Val Val Asp Lys Val Phe Gln Phe Ala Gln
 325 330 335
 Thr Pro Asp Ala Leu Ala Tyr Val Glu Thr Gly Arg Ala Arg Gly Lys
 340 345 350
 Val Val Val Thr Tyr Ala Ser
 355

<210> SEQ ID NO 85

<211> LENGTH: 1032

<212> TYPE: DNA

<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 85

```

atgaaagcga ttgtcgccca cggggcaaa gatgtgcgca tcgaagaccg gccggaggaa    60
aagccgggtc cgggagaggt gcggtccct ctggcgaggg gcgggatctg cggcagtgat    120
ctgcattatt acaatcatgg cggtttcggc gccgtgcggc ttcgtgaacc catggtgctg    180
ggccatgagg tttcccgct catcgaggaa ctgggcaag gcgttgaggg gctgaagatc    240
ggcggctcgg tggcggttcc gccgtgcgca ccatgccgaa cctgccgctt ctgccaggag    300
ggtctgcaca atcagtgcct caacatcgcg ttttatggca gcgccatgcc tttcccgcct    360
atcaggggcg cgttccggga aattctggtg gggacgccc tgcaatgcgt gccggccgat    420
ggtctcagcg ccggggaagc cgccatggcg gaaccgctgg cggtgacgct gcatgccaca    480
cgccgggccc gcgatttgcg gggaaaacgt gtgctcgtca cgggttgctg ccccatcggc    540
attctctcca ttctggtcgc gcgccgggcg ggtgctgctg aaatcgtcgc caccgacctt    600

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tccgatttca cgctcgcaaa ggcgcgtgaa gcgggggccc accgtgtcat caacagcaag 660
gatgagcccc atgcgctcgc cgcttatggt gaaaacaagg gaaccttcga cattctctat 720
gaatgctcgg gtgcccggct ggcgcttgcc ggcggcatta cggcactgcg gccgcgccc 780
atcatcgtcc agctcgggct cggcggcgat atgagcctgc cgatgatggc gatcacagcc 840
aaggaactcg acctgcgtgg ttcctttcgc ttccacgagg aattcgccac cggcgtcgag 900
ctgatgcgca agggcctgat cgacgtcaaa cccttcatca cccagaccgt cgatcttgcc 960
gacgccatct cggccttcga attcgcctcg gatcgagcc gcgccatgaa ggtgcagatc 1020
gccttttct aa 1032

```

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<210> SEQ ID NO 86
<211> LENGTH: 343
<212> TYPE: PRT
<213> ORGANISM: Agrobacterium tumefaciens C58

```

```

<400> SEQUENCE: 86

```

```

Met Lys Ala Ile Val Ala His Gly Ala Lys Asp Val Arg Ile Glu Asp
1 5 10 15
Arg Pro Glu Glu Lys Pro Gly Pro Gly Glu Val Arg Leu Arg Leu Ala
20 25 30
Arg Gly Gly Ile Cys Gly Ser Asp Leu His Tyr Tyr Asn His Gly Gly
35 40 45
Phe Gly Ala Val Arg Leu Arg Glu Pro Met Val Leu Gly His Glu Val
50 55 60
Ser Ala Val Ile Glu Glu Leu Gly Glu Gly Val Glu Gly Leu Lys Ile
65 70 75 80
Gly Gly Leu Val Ala Val Ser Pro Ser Arg Pro Cys Arg Thr Cys Arg
85 90 95
Phe Cys Gln Glu Gly Leu His Asn Gln Cys Leu Asn Met Arg Phe Tyr
100 105 110
Gly Ser Ala Met Pro Phe Pro His Ile Gln Gly Ala Phe Arg Glu Ile
115 120 125
Leu Val Ala Asp Ala Leu Gln Cys Val Pro Ala Asp Gly Leu Ser Ala
130 135 140
Gly Glu Ala Ala Met Ala Glu Pro Leu Ala Val Thr Leu His Ala Thr
145 150 155 160
Arg Arg Ala Gly Asp Leu Leu Gly Lys Arg Val Leu Val Thr Gly Cys
165 170 175
Gly Pro Ile Gly Ile Leu Ser Ile Leu Ala Ala Arg Arg Ala Gly Ala
180 185 190
Ala Glu Ile Val Ala Thr Asp Leu Ser Asp Phe Thr Leu Gly Lys Ala
195 200 205
Arg Glu Ala Gly Ala Asp Arg Val Ile Asn Ser Lys Asp Glu Pro Asp
210 215 220
Ala Leu Ala Ala Tyr Gly Ala Asn Lys Gly Thr Phe Asp Ile Leu Tyr
225 230 235 240
Glu Cys Ser Gly Ala Ala Val Ala Leu Ala Gly Gly Ile Thr Ala Leu
245 250 255
Arg Pro Arg Gly Ile Ile Val Gln Leu Gly Leu Gly Gly Asp Met Ser
260 265 270

```

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Leu Pro Met Met Ala Ile Thr Ala Lys Glu Leu Asp Leu Arg Gly Ser
 275 280 285

Phe Arg Phe His Glu Glu Phe Ala Thr Gly Val Glu Leu Met Arg Lys
 290 295 300

Gly Leu Ile Asp Val Lys Pro Phe Ile Thr Gln Thr Val Asp Leu Ala
 305 310 315 320

Asp Ala Ile Ser Ala Phe Glu Phe Ala Ser Asp Arg Ser Arg Ala Met
 325 330 335

Lys Val Gln Ile Ala Phe Ser
 340

<210> SEQ ID NO 87
 <211> LENGTH: 939
 <212> TYPE: DNA
 <213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 87

```

atgccgatgg cgctcgggca cgaagcggcg ggcgtcgtcg aggcattggg cgaagcgtg    60
cgcgatcttg agcccgggca tcatgtggtc atggtcttca tgcccagttg cggacattgc    120
ctgccctgtg cggaaggcag gcccgctctg tgcgagccgg gcgccgccgc caatgcagca    180
ggcaggctgt tgggtggcgc caccgcctg aactatcatg gcgaggctgt ccatcatcac    240
cttgggtgtg cggcctttgc cgaatatgcc gtggtgtcgc gcaattcgtt ggtcaagatc    300
gaccgcgata ttccatttgt cgaggcggca ctcttcggct gcgcggttct caccggcgtc    360
ggcgccgtcg tgaatacggc aagggtcagg accggctcga ctgcggtcgt catcggactt    420
ggcggtgtgg gccttgccgc ggttctcgga gcccgggcgg ccggtgccag caagatcgtc    480
gccgtcgacc ttctcgagga aaagcttgca ctgcaccagc aactgggcgc gaccgccatc    540
gtgaacggac gcgatgagga tgccgtcgag caggtccgag agctcacttc cggcggtgcc    600
gattatgcct tcgagatggc agggtctatt cgcgccctcg aaaacgcctt caggatgacc    660
aaacgtggcg gcaccaccgt taccgccggt ctgccaccgc cgggtgcggc cctgccgctc    720
aacgtcgtgc agctcgtcgg cgaggagcgg aactcaagg gcagctatat cggcacctgt    780
gtgctctccc gggatattcc gcgcttcac gccctttatc gcgacggccg gttgccgggtg    840
aaccccttc tgagcggaa gctgaagcta gaagacatca atgaagggtt cgaccgcctg    900
cacgacggaa gcgccgttcg gcaagtcac gaattctga                               939

```

<210> SEQ ID NO 88
 <211> LENGTH: 312
 <212> TYPE: PRT
 <213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 88

Met Pro Met Ala Leu Gly His Glu Ala Ala Gly Val Val Glu Ala Leu
 1 5 10 15

Gly Glu Gly Val Arg Asp Leu Glu Pro Gly Asp His Val Val Met Val
 20 25 30

Phe Met Pro Ser Cys Gly His Cys Leu Pro Cys Ala Glu Gly Arg Pro
 35 40 45

Ala Leu Cys Glu Pro Gly Ala Ala Ala Asn Ala Ala Gly Arg Leu Leu
 50 55 60

Gly Gly Ala Thr Arg Leu Asn Tyr His Gly Glu Val Val His His His

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| | | | | | | |
|---|---|-----|--|-----|--|-----|
| 65 | | 70 | | 75 | | 80 |
| Leu Gly Val Ser | Ala Phe Ala Glu Tyr Ala Val Val Ser Arg Asn Ser | 85 | | 90 | | 95 |
| Leu Val Lys Ile Asp Arg Asp Leu Pro Phe Val Glu Ala Ala Leu Phe | | 100 | | 105 | | 110 |
| Gly Cys Ala Val Leu Thr Gly Val Gly Ala Val Val Asn Thr Ala Arg | | 115 | | 120 | | 125 |
| Val Arg Thr Gly Ser Thr Ala Val Val Ile Gly Leu Gly Gly Val Gly | | 130 | | 135 | | 140 |
| Leu Ala Ala Val Leu Gly Ala Arg Ala Ala Gly Ala Ser Lys Ile Val | | 145 | | 150 | | 155 |
| Ala Val Asp Leu Ser Gln Glu Lys Leu Ala Leu Ala Ser Glu Leu Gly | | 165 | | 170 | | 175 |
| Ala Thr Ala Ile Val Asn Gly Arg Asp Glu Asp Ala Val Glu Gln Val | | 180 | | 185 | | 190 |
| Arg Glu Leu Thr Ser Gly Gly Ala Asp Tyr Ala Phe Glu Met Ala Gly | | 195 | | 200 | | 205 |
| Ser Ile Arg Ala Leu Glu Asn Ala Phe Arg Met Thr Lys Arg Gly Gly | | 210 | | 215 | | 220 |
| Thr Thr Val Thr Ala Gly Leu Pro Pro Pro Gly Ala Ala Leu Pro Leu | | 225 | | 230 | | 235 |
| Asn Val Val Gln Leu Val Gly Glu Glu Arg Thr Leu Lys Gly Ser Tyr | | 245 | | 250 | | 255 |
| Ile Gly Thr Cys Val Pro Leu Arg Asp Ile Pro Arg Phe Ile Ala Leu | | 260 | | 265 | | 270 |
| Tyr Arg Asp Gly Arg Leu Pro Val Asn Arg Leu Leu Ser Gly Arg Leu | | 275 | | 280 | | 285 |
| Lys Leu Glu Asp Ile Asn Glu Gly Phe Asp Arg Leu His Asp Gly Ser | | 290 | | 295 | | 300 |
| Ala Val Arg Gln Val Ile Glu Phe | | 305 | | 310 | | |

<210> SEQ ID NO 89

<211> LENGTH: 1035

<212> TYPE: DNA

<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 89

```

atgaaacatt ctcaggacaa accacgcctg ctgattgcca tgcgtagcga gcttccagaa    60
ggcttcttcg gtccgcgcga atgggcaagg ctgaatgccg tagcggacat tattccgggc    120
tttcccata cggatttcga cacggcgaac ggtgccgagg ctctcgccga agcggatatt    180
ctgctcgctg cctggggtac gccatccctg acacgcgaac gactttcacg cgcgccgagg    240
ctgaaaaatc tggcctatgc ggcatcatcg gtgcggatgg ttgcgccgcg agaattctgg    300
gagacgtcgg atattctggt cacgacagca gcttccgcca tggccgtgcc ggttgccgaa    360
ttcacctatg cggcaatcat catgtgcggc aaggatgtgt ttcgattgcg ggatgaacat    420
agaacagagc gcggcaccgg cgtttttggc agcaggcgcg gcagaagcct gccctatctt    480
ggcaatcatg cccgcaaggt tggcattgtc ggcgcctcgc gcacggggcg gctgggtgatg    540
gagatgctgg cgcgcggcac attcgagatt gccgtttacg atccctttct gtcggcgaaa    600
gaggccgcat cccttgccgc gaagaaagcc gaactggacg agcttctcgc atggtccgat    660

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gtggtctcgc tgcaecgcgc gatcctgccc gaaacgcacc atatgatcgg cgcccgcgaa 720
ctggcgctga tggcggacca tgccatcttc atcaacacgg cgcggggctg gctggctcgc 780
cacgatgcat tgctgactga agcgatttcc ggacggctgc gcattctgat tgacacgccc 840
gaaccggagc cctgcccac ggacagcccg ttttacgac tgcccattgt cgttctaacc 900
ccccatatag ccggggcgct gggcaatgaa ttgcgcgcac tttccgatct ggccattacc 960
gaaattgaac gtttcgtggc gggacttgcg cccctccacc cggtcacaaa gcaggatag 1020
gaacgtatgg catga 1035

```

<210> SEQ ID NO 90

<211> LENGTH: 331

<212> TYPE: PRT

<213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 90

```

Met Arg Ser Glu Leu Pro Glu Gly Phe Phe Gly Pro Arg Glu Trp Ala
1 5 10 15
Arg Leu Asn Ala Val Ala Asp Ile Ile Pro Gly Phe Pro His Thr Asp
20 25 30
Phe Asp Thr Ala Asn Gly Ala Glu Ala Leu Ala Glu Ala Asp Ile Leu
35 40 45
Leu Ala Ala Trp Gly Thr Pro Ser Leu Thr Arg Glu Arg Leu Ser Arg
50 55 60
Ala Pro Arg Leu Lys Met Leu Ala Tyr Ala Ala Ser Ser Val Arg Met
65 70 75 80
Val Ala Pro Ala Glu Phe Trp Glu Thr Ser Asp Ile Leu Val Thr Thr
85 90 95
Ala Ala Ser Ala Met Ala Val Pro Val Ala Glu Phe Thr Tyr Ala Ala
100 105 110
Ile Ile Met Cys Gly Lys Asp Val Phe Arg Leu Arg Asp Glu His Arg
115 120 125
Thr Glu Arg Gly Thr Gly Val Phe Gly Ser Arg Arg Gly Arg Ser Leu
130 135 140
Pro Tyr Leu Gly Asn His Ala Arg Lys Val Gly Ile Val Gly Ala Ser
145 150 155 160
Arg Ile Gly Arg Leu Val Met Glu Met Leu Ala Arg Gly Thr Phe Glu
165 170 175
Ile Ala Val Tyr Asp Pro Phe Leu Ser Ala Glu Glu Ala Ala Ser Leu
180 185 190
Gly Ala Lys Lys Ala Glu Leu Asp Glu Leu Leu Ala Trp Ser Asp Val
195 200 205
Val Ser Leu His Ala Pro Ile Leu Pro Glu Thr His His Met Ile Gly
210 215 220
Ala Arg Glu Leu Ala Leu Met Ala Asp His Ala Ile Phe Ile Asn Thr
225 230 235 240
Ala Arg Gly Trp Leu Val Asp His Asp Ala Leu Leu Thr Glu Ala Ile
245 250 255
Ser Gly Arg Leu Arg Ile Leu Ile Asp Thr Pro Glu Pro Glu Pro Leu
260 265 270
Pro Thr Asp Ser Pro Phe Tyr Asp Leu Pro Asn Val Val Leu Thr Pro
275 280 285

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His Ile Ala Gly Ala Leu Gly Asn Glu Leu Arg Ala Leu Ser Asp Leu
 290 295 300

Ala Ile Thr Glu Ile Glu Arg Phe Val Ala Gly Leu Ala Pro Leu His
 305 310 315 320

Pro Val His Lys Gln Asp Met Glu Arg Met Ala
 325 330

<210> SEQ ID NO 91
 <211> LENGTH: 750
 <212> TYPE: DNA
 <213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 91

```

atgcagcgtt ttaccaacag aaccatcgtt gtcgccgggg cggcccgga tatcgccgg 60
gcatgcgcca tccgtttcgc acaggaaggc gccaatgtcg ttcttaccta taatggcgcg 120
gcagaggggc cggccacagc cgttgccgaa atcgaaaagc ttggtcgttc ggctctggcg 180
atcaaggcgg atctcacaaa cgccgccgaa gtcgaggctg ccatatctgc ggctgaggac 240
aagtttgggg agatccacgg cctcgtccat gttgccggcg gcctgatcgc ccgcaagaca 300
atgcagaaa tggatgaagc cttctggcat caggtcctcg acgtcaatct gacatcgctg 360
ttctgacgg ccaagaccgc attgccgaag atggccaagg gcggcgcgat cgtcaacttc 420
tcgtcgcagg cgggcccgtga tggcgggcgc cggggcgctc ttgcctatgc cacttccaag 480
ggtgcccgtga tgaccttcac ccgcggaact gccaaagaag tcggcccaa aatccgcgtc 540
aacgcccgtt gccccggtat gatctccacc accttccacg ataccttcac caagccggag 600
gtgcgcgaac gggtgccggc gcgcagctcg ctcaagcgcg aagggtcgag cgaagacgtc 660
gcccgtctgg tggccttctc cgcgtctgac gatgccgctt atgtcacgg cgectgetac 720
gacatcaatg gcggcgtcct gttttctga 750

```

<210> SEQ ID NO 92
 <211> LENGTH: 249
 <212> TYPE: PRT
 <213> ORGANISM: Agrobacterium tumefaciens C58

<400> SEQUENCE: 92

```

Met Gln Arg Phe Thr Asn Arg Thr Ile Val Val Ala Gly Ala Gly Arg
1 5 10 15
Asp Ile Gly Arg Ala Cys Ala Ile Arg Phe Ala Gln Glu Gly Ala Asn
20 25 30
Val Val Leu Thr Tyr Asn Gly Ala Ala Glu Gly Ala Ala Thr Ala Val
35 40 45
Ala Glu Ile Glu Lys Leu Gly Arg Ser Ala Leu Ala Ile Lys Ala Asp
50 55 60
Leu Thr Asn Ala Ala Glu Val Glu Ala Ala Ile Ser Ala Ala Ala Asp
65 70 75 80
Lys Phe Gly Glu Ile His Gly Leu Val His Val Ala Gly Gly Leu Ile
85 90 95
Ala Arg Lys Thr Ile Ala Glu Met Asp Glu Ala Phe Trp His Gln Val
100 105 110
Leu Asp Val Asn Leu Thr Ser Leu Phe Leu Thr Ala Lys Thr Ala Leu
115 120 125

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Pro Lys Met Ala Lys Gly Gly Ala Ile Val Thr Phe Ser Ser Gln Ala
 130 135 140

Gly Arg Asp Gly Gly Gly Pro Gly Ala Leu Ala Tyr Ala Thr Ser Lys
 145 150 155 160

Gly Ala Val Met Thr Phe Thr Arg Gly Leu Ala Lys Glu Val Gly Pro
 165 170 175

Lys Ile Arg Val Asn Ala Val Cys Pro Gly Met Ile Ser Thr Thr Phe
 180 185 190

His Asp Thr Phe Thr Lys Pro Glu Val Arg Glu Arg Val Ala Gly Ala
 195 200 205

Thr Ser Leu Lys Arg Glu Gly Ser Ser Glu Asp Val Ala Gly Leu Val
 210 215 220

Ala Phe Leu Ala Ser Asp Asp Ala Ala Tyr Val Thr Gly Ala Cys Tyr
 225 230 235 240

Asp Ile Asn Gly Gly Val Leu Phe Ser
 245

<210> SEQ ID NO 93
 <211> LENGTH: 930
 <212> TYPE: DNA
 <213> ORGANISM: Escherichia coli DH10B

<400> SEQUENCE: 93

```

atgtccaaaa agattgccgt gattggcgaa tgcattgattg agctttccga gaaaggcgcg      60
gacgttaagc gcggtttcgg cggcgatacc ctgaacactt ccgtctatat cgccccgcag      120
gtcgcacctg cggcattaac cgttcattac gtaacggcgc tgggaacgga cagtttttagc      180
cagcagatgc tggacgcctg gcacggcgag aacgttgata cttccctgac ccaacggatg      240
gaaaaccgtc tgccgggctt ttactacatt gaaaccgaca gcaccggcga gcgtagcttc      300
tactactggc ggaacgaagc cgccgccaaa ttctggctgg agagtgagca gctgcccggc      360
atttgccaag agctggcgaa ttctgattat ctctacctga gcgggattag cctggcgatc      420
ttaagcccga ccagccgcga aaagtgtctt tccctgctgc gcgaatgccg cgccaacggc      480
ggaaaagtga ttttcgacaa taactatcgt ccgcccctgt gggccagcaa agaagagaca      540
cagcaggtgt accaacaagt gctggaatgc acggatatcg ccttccctgac gctggaacgac      600
gaagacgcgc tgtgggggtc acagccgggtg gaagacgtca ttgcgcgcac ccataacgcg      660
ggcgtgaaa gagtgggtgt gaaacgcggg gcggtattctt gcctggtgtc cattgctggc      720
gaagggttag tggatgttcc ggcggtgaaa ctgccgaaag aaaaagtgat cgataccacc      780
gcagctggcg actctttcag tgccggttat ctggcgggtac gtctgacagg cggcagcgcg      840
gaagacgcgg cgaacgtggg gcacctgacc gcaagtaccg ttattcagta tcgcccgcgcg      900
attatccccg gtgaggcgat gccagcgtaa      930
    
```

<210> SEQ ID NO 94
 <211> LENGTH: 309
 <212> TYPE: PRT
 <213> ORGANISM: Escherichia coli DH10B

<400> SEQUENCE: 94

Met Ser Lys Lys Ile Ala Val Ile Gly Glu Cys Met Ile Glu Leu Ser
 1 5 10 15

Glu Lys Gly Ala Asp Val Lys Arg Gly Phe Gly Gly Asp Thr Leu Asn

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```

ctgatgctgg gtatggacta cggtttgaaa gagttcaaat tcttcccggc tgaagctaac 420
ggcggcgctga aagccctgca ggcgatcgcg ggtccgttct cccaggtccg tttctgcccg 480
acgggtggta tttctccggc taactaccgt gactacctgg cgctgaaaag cgtgctgtgc 540
atcggtggtt cctggctggt tccggcagat gcgctggaag cgggcgatta cgaccgcatt 600
actaagctgg cgcgtgaagc tgtagaaggc gctaagctgt aa 642

```

```

<210> SEQ ID NO 96
<211> LENGTH: 213
<212> TYPE: PRT
<213> ORGANISM: Escherichia coli DH10B

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```

<400> SEQUENCE: 96

```

```

Met Lys Asn Trp Lys Thr Ser Ala Glu Ser Ile Leu Thr Thr Gly Pro
1           5           10          15
Val Val Pro Val Ile Val Val Lys Lys Leu Glu His Ala Val Pro Met
20          25          30
Ala Lys Ala Leu Val Ala Gly Gly Val Arg Val Leu Glu Val Thr Leu
35          40          45
Arg Thr Glu Cys Ala Val Asp Ala Ile Arg Ala Ile Ala Lys Glu Val
50          55          60
Pro Glu Ala Ile Val Gly Ala Gly Thr Val Leu Asn Pro Gln Gln Leu
65          70          75          80
Ala Glu Val Thr Glu Ala Gly Ala Gln Phe Ala Ile Ser Pro Gly Leu
85          90          95
Thr Glu Pro Leu Leu Lys Ala Ala Thr Glu Gly Thr Ile Pro Leu Ile
100         105         110
Pro Gly Ile Ser Thr Val Ser Glu Leu Met Leu Gly Met Asp Tyr Gly
115         120         125
Leu Lys Glu Phe Lys Phe Phe Pro Ala Glu Ala Asn Gly Gly Val Lys
130         135         140
Ala Leu Gln Ala Ile Ala Gly Pro Phe Ser Gln Val Arg Phe Cys Pro
145         150         155         160
Thr Gly Gly Ile Ser Pro Ala Asn Tyr Arg Asp Tyr Leu Ala Leu Lys
165         170         175
Ser Val Leu Cys Ile Gly Gly Ser Trp Leu Val Pro Ala Asp Ala Leu
180         185         190
Glu Ala Gly Asp Tyr Asp Arg Ile Thr Lys Leu Ala Arg Glu Ala Val
195         200         205
Glu Gly Ala Lys Leu
210

```

```

<210> SEQ ID NO 97
<211> LENGTH: 780
<212> TYPE: DNA
<213> ORGANISM: Lactobacillus brevis ATCC 367

```

```

<400> SEQUENCE: 97

```

```

atggcatcaa atgaaaaagt agcaatggtt accggtggcg gacaaggaat tggtagaagc 60
atctgaaaac ggtagctaa cgacggcttt gctgtggcaa ttgctgattt gaacttggac 120
aatgccaaca agtgcgtttc tgatattgaa gctgctggtg gcaaggccat tgcggtcaag 180
accgatgtct ctgatectga tagcgtgttt gctgcggtta atgaagcggc cgacaagctg 240

```

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ggcggtttg acgttatcgt taataacgcc ggccttgccc caaccacgcc aattgacacc 300
atcacccaag aacagtttga tacggtttat cacgttaacg tgggtgggggt tctttggggc 360
attcaagcag cccatgcgaa gttcaaggaa ttgggtcatg gtgggaagat catttccgcg 420
acgtctcaag cgggggttgt tggttaacccg aacttagctc tgtacagtgg aactaagttt 480
gccattcgtg gtgtgaccca agttgcggcg cgtgacttag ccgctgaagg tatcacggtc 540
aatgcttatg caccgggat tgtaagaca ccaatgatgt ttgacatcgc tcacaagggt 600
ggtcaaaatg ctggtaaaga cgacgaatgg gggatgcaaa ccttctcaaa ggacatcgtc 660
ttatgtcgat tgtcagaacc agaagatgtg gctaacgggg tggctttctt agccggtccc 720
gattctaact acattacggg tcaaacactt gaagttgatg gtgggatgca gttccactaa 780

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<210> SEQ ID NO 98

<211> LENGTH: 259

<212> TYPE: PRT

<213> ORGANISM: *Lactobacillus brevis* ATCC 367

<400> SEQUENCE: 98

```

Met Ala Ser Asn Gly Lys Val Ala Met Val Thr Gly Gly Gly Gln Gly
1           5           10          15
Ile Gly Glu Ala Ile Ser Lys Arg Leu Ala Asn Asp Gly Phe Ala Val
20          25          30
Ala Ile Ala Asp Leu Asn Leu Asp Asn Ala Asn Lys Val Val Ser Asp
35          40          45
Ile Glu Ala Ala Gly Gly Lys Ala Ile Ala Val Lys Thr Asp Val Ser
50          55          60
Asp Arg Asp Ser Val Phe Ala Ala Val Asn Glu Ala Ala Asp Lys Leu
65          70          75          80
Gly Gly Phe Asp Val Ile Val Asn Asn Ala Gly Leu Gly Pro Thr Thr
85          90          95
Pro Ile Asp Thr Ile Thr Gln Glu Gln Phe Asp Thr Val Tyr His Val
100         105         110
Asn Val Gly Gly Val Leu Trp Gly Ile Gln Ala Ala His Ala Lys Phe
115         120         125
Lys Glu Leu Gly His Gly Gly Lys Ile Ile Ser Ala Thr Ser Gln Ala
130         135         140
Gly Val Val Gly Asn Pro Asn Leu Ala Leu Tyr Ser Gly Thr Lys Phe
145         150         155         160
Ala Ile Arg Gly Val Thr Gln Val Ala Ala Arg Asp Leu Ala Ala Glu
165         170         175
Gly Ile Thr Val Asn Ala Tyr Ala Pro Gly Ile Val Lys Thr Pro Met
180         185         190
Met Phe Asp Ile Ala His Lys Val Gly Gln Asn Ala Gly Lys Asp Asp
195         200         205
Glu Trp Gly Met Gln Thr Phe Ser Lys Asp Ile Ala Leu Cys Arg Leu
210         215         220
Ser Glu Pro Glu Asp Val Ala Asn Gly Val Ala Phe Leu Ala Gly Pro
225         230         235         240
Asp Ser Asn Tyr Ile Thr Gly Gln Thr Leu Glu Val Asp Gly Gly Met
245         250         255
Gln Phe His

```

-continued

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<210> SEQ ID NO 99
<211> LENGTH: 1089
<212> TYPE: DNA
<213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 99
atgaatgacc tgagccacac ccacatgcgc gcggccgtct ggcattggccg ccacgatatt    60
cgtgtcgaac aggtaccttt gccggccgac cctgcgccgg gctgggtgca gatcaagggtg    120
gactggtgcg gcatctgcgg ctccgacctg cacgaatatg ttgccggccc ggtgttcate    180
ccggtagagg ccccgcaccc gctgaccggc attcagggcc agtgcaccc cggccacgaa    240
ttctgcggcc acatcgccaa gcttggcgaa ggcgtggaag gctatgccgt aggcgacccg    300
gtggcggcag acgctgcca gcattgtggt acctgctatt actgcaccca tggcctgtac    360
aacatctgcg aacgcctggc gttcaccggc ctgatgaaca acggtgcctt cgccgagctg    420
gtcaactgtc ccgccaacct gctctaccgg ctgccgcagg gcttccctgc cgaagccggg    480
gcactgatcg agccgctggc ggtgggtatg cacgcggtga aaaaggccgg cagcctgctt    540
gggcaaaccg ttgtagtggt tggggccggc accatcggcc tgtgcacccat catgtgcgcc    600
aaggctgcag gtgcggcaca ggtcatcgcc cttgagatgt cctctgcgcg caaagccaag    660
gccaaaggaag cgggcgccaa cgtggtgctg gacccagacc agtgcgatgc cctggcgga    720
atccgcgcac tgactgctgg gctggggccc gatgtgagtt ttgagtgcac cggcaacaaa    780
catacggcca agctggccat cgacaccatc cgcaaagcag gcaagtgcgt gctggtgggt    840
atthtcgaag agcccagcga gttcaacttc ttcgagctgg tgtccaccga gaagcaagtg    900
ctgggggctg tggcgtacaa cggcgagttt gctgacgtga ttgccttcat tgetgatggt    960
cggctggata ttcgcccctg ggtaaccggc cggatcggat tggagcagat tgtcgagctg    1020
ggcttcgagg aactggtgaa caacaagag gagaacgtga agatcatcgt ttcaccaggt    1080
gtgcgctga                                         1089

```

```

<210> SEQ ID NO 100
<211> LENGTH: 362
<212> TYPE: PRT
<213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 100
Met Asn Asp Leu Ser His Thr His Met Arg Ala Ala Val Trp His Gly
1          5          10         15
Arg His Asp Ile Arg Val Glu Gln Val Pro Leu Pro Ala Asp Pro Ala
20         25         30
Pro Gly Trp Val Gln Ile Lys Val Asp Trp Cys Gly Ile Cys Gly Ser
35         40         45
Asp Leu His Glu Tyr Val Ala Gly Pro Val Phe Ile Pro Val Glu Ala
50         55         60
Pro His Pro Leu Thr Gly Ile Gln Gly Gln Cys Ile Leu Gly His Glu
65         70         75         80
Phe Cys Gly His Ile Ala Lys Leu Gly Glu Gly Val Glu Gly Tyr Ala
85         90         95
Val Gly Asp Pro Val Ala Ala Asp Ala Cys Gln His Cys Gly Thr Cys
100        105        110
Tyr Tyr Cys Thr His Gly Leu Tyr Asn Ile Cys Glu Arg Leu Ala Phe

```


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```
ctgtccgagc cggaagatgt cgccgctgc gtctcctatc ttgccagccc ggattctgat 720
tacatgaccg gtcagtcggt gctgatcgac ggccggatgg tatttaacta a 771
```

```
<210> SEQ ID NO 102
<211> LENGTH: 256
<212> TYPE: PRT
<213> ORGANISM: Klebsiella pneumoniae MGH78578
```

```
<400> SEQUENCE: 102
```

```
Met Lys Lys Val Ala Leu Val Thr Gly Ala Gly Gln Gly Ile Gly Lys
1 5 10 15
Ala Ile Ala Leu Arg Leu Val Lys Asp Gly Phe Ala Val Ala Ile Ala
20 25 30
Asp Tyr Asn Asp Ala Thr Ala Lys Ala Val Ala Ser Glu Ile Asn Gln
35 40 45
Ala Gly Gly His Ala Val Ala Val Lys Val Asp Val Ser Asp Arg Asp
50 55 60
Gln Val Phe Ala Ala Val Glu Gln Ala Arg Lys Thr Leu Gly Gly Phe
65 70 75 80
Asp Val Ile Val Asn Asn Ala Gly Val Ala Pro Ser Thr Pro Ile Glu
85 90 95
Ser Ile Thr Pro Glu Ile Val Asp Lys Val Tyr Asn Ile Asn Val Lys
100 105 110
Gly Val Ile Trp Gly Ile Gln Ala Ala Val Glu Ala Phe Lys Lys Glu
115 120 125
Gly His Gly Gly Lys Ile Ile Asn Ala Cys Ser Gln Ala Gly His Val
130 135 140
Gly Asn Pro Glu Leu Ala Val Tyr Ser Ser Ser Lys Phe Ala Val Arg
145 150 155 160
Gly Leu Thr Gln Thr Ala Ala Arg Asp Leu Ala Pro Leu Gly Ile Thr
165 170 175
Val Asn Gly Tyr Cys Pro Gly Ile Val Lys Thr Pro Met Trp Ala Glu
180 185 190
Ile Asp Arg Gln Val Ser Glu Ala Ala Gly Lys Pro Leu Gly Tyr Gly
195 200 205
Thr Ala Glu Phe Ala Lys Arg Ile Thr Leu Gly Arg Leu Ser Glu Pro
210 215 220
Glu Asp Val Ala Ala Cys Val Ser Tyr Leu Ala Ser Pro Asp Ser Asp
225 230 235 240
Tyr Met Thr Gly Gln Ser Leu Leu Ile Asp Gly Gly Met Val Phe Asn
245 250 255
```

```
<210> SEQ ID NO 103
<211> LENGTH: 1665
<212> TYPE: DNA
<213> ORGANISM: Klebsiella pneumoniae MGH78578
```

```
<400> SEQUENCE: 103
```

```
atgagatcga aaagatttga agcactggcg aaacgcctcg tgaatcagga tggtttcggt 60
aaggagtgga ttgaagaggg ctttatcgcg atggaaagcc ctaacgatcc caaaccttct 120
atccgcatcg tcaacggcgc ggtgaccgaa ctcgacgata aaccggttga gcagttcgac 180
ctgattgacc accttatcgc gcgctacggc attaactctc cccgggcccga agaagtgatg 240
```

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```

gccatggatt cggttaagct cgccaacatg ctctgcgacc cgaacgttaa acgcagcgac 300
atcgtgccgc tcaactaccgc gatgaaccccg gcgaaaatcg tggaaagtgg gtcgcatatg 360
aacgtggtcg agatgatgat ggcgatgcaa aaaatgcgcg cccgccgcac gccgtcccag 420
caggcgcacg tcaactaatat caaagataat ccggtacaga ttgccgccga cgccgctgaa 480
ggcgcacatg gcggctttga cgagcaggag accaccgtcg ccgtggcgcg ctacgcgcgc 540
ttcaacgcca tcgccctgct ggtcggttca caggttgcc gccccggcgt cctcaccag 600
tgttcgctgg aagaagccac cgagctgaaa ctgggcatgc tgggccacac ctgctatgcc 660
gaaaccattt cggatatacgg tacggaaccg gtgtttaccg atggcgatga cccccgtgg 720
tcgaaaggct tectcgctc ctcctacgcc tcgcgcgcc tgaaaatgcg ctttacctcc 780
ggttcgggct cggaggtgca gatgggctat gccgaaggca aatcgatgct ttatctcgaa 840
gcgcgctgca tctacatcac caaagccgcc ggggtgcaag gcctgcagaa tggctccgtc 900
agctgtatcg gcgtgcccgc cgccgtgccc tccgggatcc gcgccgtact ggcggaaaac 960
ctgatctgct cagcgtgga tctggagtgc gcctccagca acgatcaaac ctttaccac 1020
tcggatagc ggcgtaccgc gcgtctgctg atgcagttcc tgcaggtac cgactttatc 1080
tcctccggtt actcggcggc gccgaactac gacaacatgt tcgccggttc caacgaagat 1140
gccgaagact tcgatgacta caacgtgatc cagcgcgacc tgaaggtcga tggcggcctg 1200
cggccggtgc gtgaagagga cgtgatgcc attcgcaaca aagccgcccg cgcgctgcag 1260
gcggtatttg ccggcatggg tttgcgcct attacggatg aagaagtaga agccgccacc 1320
tacgccacg gttcaaaaga tatgcctgag cgcaatatcg tcgaggacat caagtttget 1380
caggagatca tcaacaagaa ccgcaacggc ctggaggtgg tgaagccct ggcgaaaggc 1440
ggcttccccg atgtcgccca ggacatgctc aatattcaga aagccaagct caccgcgac 1500
tacctgata cctccgccat cattgttggc gagggccagg tgctctcggc cgtgaatgac 1560
gtgaacgatt atgccggtcc ggcaacagc taccgcctgc aagcgagcg ctgggaagag 1620
attaanaata tcccgggcgc gctcgatccc aatgaacttg gctaa 1665

```

<210> SEQ ID NO 104

<211> LENGTH: 554

<212> TYPE: PR

<213> ORGANISM: Klebsiella pneumoniae MGH78578

<400> SEQUENCE: 104

```

Met Arg Ser Lys Arg Phe Glu Ala Leu Ala Lys Arg Pro Val Asn Gln
1           5           10          15
Asp Gly Phe Val Lys Glu Trp Ile Glu Glu Gly Phe Ile Ala Met Glu
20          25          30
Ser Pro Asn Asp Pro Lys Pro Ser Ile Arg Ile Val Asn Gly Ala Val
35          40          45
Thr Glu Leu Asp Asp Lys Pro Val Glu Gln Phe Asp Leu Ile Asp His
50          55          60
Phe Ile Ala Arg Tyr Gly Ile Asn Leu Ala Arg Ala Glu Glu Val Met
65          70          75          80
Ala Met Asp Ser Val Lys Leu Ala Asn Met Leu Cys Asp Pro Asn Val
85          90          95
Lys Arg Ser Asp Ile Val Pro Leu Thr Thr Ala Met Thr Pro Ala Lys

```

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| 100 | | | | 105 | | | | 110 | | | | | | | | |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Ile | Val | Glu | 115 | Val | Val | Ser | His | Met | Asn | Val | Val | Glu | Met | Met | Met | Ala |
| Met | Gln | Lys | 130 | Met | Arg | Ala | Arg | Arg | Thr | Pro | Ser | Gln | Gln | Ala | His | Val |
| Thr | Asn | Ile | 145 | Lys | Asp | Asn | Pro | Val | Gln | Ile | Ala | Ala | Asp | Ala | Ala | Glu |
| Gly | Ala | Trp | 165 | Arg | Gly | Phe | Asp | Glu | Gln | Glu | Thr | Thr | Val | Ala | Val | Ala |
| Arg | Tyr | Ala | 180 | Pro | Phe | Asn | Ala | Ile | Ala | Leu | Leu | Val | Gly | Ser | Gln | Val |
| Gly | Arg | Pro | 195 | Gly | Val | Leu | Thr | Gln | Cys | Ser | Leu | Glu | Glu | Ala | Thr | Glu |
| Leu | Lys | Leu | 210 | Gly | Met | Leu | Gly | His | Thr | Cys | Tyr | Ala | Glu | Thr | Ile | Ser |
| Val | Tyr | Gly | 225 | Thr | Glu | Pro | Val | Phe | Thr | Asp | Gly | Asp | Asp | Thr | Pro | Trp |
| Ser | Lys | Gly | 245 | Phe | Leu | Ala | Ser | Ser | Tyr | Ala | Ser | Arg | Gly | Leu | Lys | Met |
| Arg | Phe | Thr | 260 | Ser | Gly | Ser | Gly | Ser | Glu | Val | Gln | Met | Gly | Tyr | Ala | Glu |
| Gly | Lys | Ser | 275 | Met | Leu | Tyr | Leu | Glu | Ala | Arg | Cys | Ile | Tyr | Ile | Thr | Lys |
| Ala | Ala | Gly | 290 | Val | Gln | Gly | Leu | Gln | Asn | Gly | Ser | Val | Ser | Cys | Ile | Gly |
| Val | Pro | Ser | 305 | Ala | Val | Pro | Ser | Gly | Ile | Arg | Ala | Val | Leu | Ala | Glu | Asn |
| Leu | Ile | Cys | 325 | Ser | Ala | Leu | Asp | Leu | Glu | Cys | Ala | Ser | Ser | Asn | Asp | Gln |
| Thr | Phe | Thr | 340 | His | Ser | Asp | Met | Arg | Arg | Thr | Ala | Arg | Leu | Leu | Met | Gln |
| Phe | Leu | Pro | 355 | Gly | Thr | Asp | Phe | Ile | Ser | Ser | Gly | Tyr | Ser | Ala | Val | Pro |
| Asn | Tyr | Asp | 370 | Asn | Met | Phe | Ala | Gly | Ser | Asn | Glu | Asp | Ala | Glu | Asp | Phe |
| Asp | Asp | Tyr | 385 | Asn | Val | Ile | Gln | Arg | Asp | Leu | Lys | Val | Asp | Gly | Gly | Leu |
| Arg | Pro | Val | 405 | Arg | Glu | Glu | Asp | Val | Ile | Ala | Ile | Arg | Asn | Lys | Ala | Ala |
| Arg | Ala | Leu | 420 | Gln | Ala | Val | Phe | Ala | Gly | Met | Gly | Leu | Pro | Pro | Ile | Thr |
| Asp | Glu | Glu | 435 | Val | Glu | Ala | Ala | Thr | Tyr | Ala | His | Gly | Ser | Lys | Asp | Met |
| Pro | Glu | Arg | 450 | Asn | Ile | Val | Glu | Asp | Ile | Lys | Phe | Ala | Gln | Glu | Ile | Ile |
| Asn | Lys | Asn | 465 | Arg | Asn | Gly | Leu | Glu | Val | Val | Lys | Ala | Leu | Ala | Lys | Gly |
| Gly | Phe | Pro | 485 | Asp | Val | Ala | Gln | Asp | Met | Leu | Asn | Ile | Gln | Lys | Ala | Lys |
| Leu | Thr | Gly | 500 | Asp | Tyr | Leu | His | Thr | Ser | Ala | Ile | Ile | Val | Gly | Glu | Gly |

-continued

Gln Val Leu Ser Ala Val Asn Asp Val Asn Asp Tyr Ala Gly Pro Ala
 515 520 525

Thr Gly Tyr Arg Leu Gln Gly Glu Arg Trp Glu Glu Ile Lys Asn Ile
 530 535 540

Pro Gly Ala Leu Asp Pro Asn Glu Leu Gly
 545 550

<210> SEQ ID NO 105
 <211> LENGTH: 690
 <212> TYPE: DNA
 <213> ORGANISM: Klebsiella pneumoniae MGH78578

<400> SEQUENCE: 105

```

atggaaatta acgaaacgct gctgcccag attatcgaag aggtgctgtc ggagatgaaa    60
tcaggcgcag ataagccggt ctccttagc gcgcctgcgg cttctgtcgc ctctgccgcg    120
ccggtcgcgc ttgcgcctgt gtccggcgac agcttcctga cggaaatcgg cgaagccaaa    180
cccggcacgc agcaggatga agtcattatt gccgtcgggc cagcgtttgg tctggcgcaa    240
accgccaata tcgtcggcat tccgcataaa aatattctgc gcgaagtgat cgcgcgcatt    300
gaggaagaag gcatcaaagc ccgggtgatc cgctgcttta agtcttctga cgtcgccttc    360
gtggcagtgg aaggcaaccg cctgagcggc tccggcatct cgatcggtat tcagtcgaaa    420
ggcaccaccg tcatccacca gcgcggcctg ccgccgcttt ccaatctgga actcttcccg    480
caggcgcgcc tgctgacgct ggaaacctac cgtcagattg gcaaaaacgc cgcgcgctac    540
gccaaaacgc agtcgcgcga gccggtgccg acgcttaacg atcagatggc tcgtcccaaa    600
taccaggcga agtcggccat tttgcacatt aaagagacca aatacgtggt gacgggcaaa    660
aaccgcagg aactgcgcgt ggcgctttaa    690

```

<210> SEQ ID NO 106
 <211> LENGTH: 229
 <212> TYPE: PRT
 <213> ORGANISM: Klebsiella pneumoniae MGH78578

<400> SEQUENCE: 106

```

Met Glu Ile Asn Glu Thr Leu Leu Arg Gln Ile Ile Glu Glu Val Leu
1           5           10           15

Ser Glu Met Lys Ser Gly Ala Asp Lys Pro Val Ser Phe Ser Ala Pro
20           25           30

Ala Ala Ser Val Ala Ser Ala Ala Pro Val Ala Val Ala Pro Val Ser
35           40           45

Gly Asp Ser Phe Leu Thr Glu Ile Gly Glu Ala Lys Pro Gly Thr Gln
50           55           60

Gln Asp Glu Val Ile Ile Ala Val Gly Pro Ala Phe Gly Leu Ala Gln
65           70           75           80

Thr Ala Asn Ile Val Gly Ile Pro His Lys Asn Ile Leu Arg Glu Val
85           90           95

Ile Ala Gly Ile Glu Glu Glu Gly Ile Lys Ala Arg Val Ile Arg Cys
100          105          110

Phe Lys Ser Ser Asp Val Ala Phe Val Ala Val Glu Gly Asn Arg Leu
115          120          125

Ser Gly Ser Gly Ile Ser Ile Gly Ile Gln Ser Lys Gly Thr Thr Val
130          135          140

```

-continued

Ile His Gln Arg Gly Leu Pro Pro Leu Ser Asn Leu Glu Leu Phe Pro
 145 150 155 160

Gln Ala Pro Leu Leu Thr Leu Glu Thr Tyr Arg Gln Ile Gly Lys Asn
 165 170 175

Ala Ala Arg Tyr Ala Lys Arg Glu Ser Pro Gln Pro Val Pro Thr Leu
 180 185 190

Asn Asp Gln Met Ala Arg Pro Lys Tyr Gln Ala Lys Ser Ala Ile Leu
 195 200 205

His Ile Lys Glu Thr Lys Tyr Val Val Thr Gly Lys Asn Pro Gln Glu
 210 215 220

Leu Arg Val Ala Leu
 225

<210> SEQ ID NO 107
 <211> LENGTH: 525
 <212> TYPE: DNA
 <213> ORGANISM: Klebsiella pneumoniae MGH78578

<400> SEQUENCE: 107

```
atgaataccg acgcaattga atccatggta cgcgacgtgc tgagccggat gaacagccta 60
caggacggga taacgcccgc gccagcccg cgcacaaaacg acaccgttcg ccagccaaaa 120
gttagcgact acccgttagc gaccggccat ccggagtggg tcaaaaccgc taccaataaa 180
acgctcgatg acctgacgct ggagaacgta ttaagcgatc gcgttacggc gcaggacatg 240
cgcatcactc cggaaacgct gcgtatgcag cggcgcatcg cccaggatgc cggacgcgat 300
cggctggcga tgaactttga gcgggcccga gagctcaccg cggttcccga cgaccgaatc 360
cttgagatct acaacgcctt gcgccatac cgttccaccc aggcggagct actggcgatc 420
gctgatgacc tcgagcatcg ctaccagcga cgactctgtg ccgcctttgt tcgggaagcg 480
gccgggctgt acatcgagcg taagaagctg aaaggcgcg attaa 525
```

<210> SEQ ID NO 108
 <211> LENGTH: 174
 <212> TYPE: PRT
 <213> ORGANISM: Klebsiella pneumoniae MGH78578

<400> SEQUENCE: 108

Met Asn Thr Asp Ala Ile Glu Ser Met Val Arg Asp Val Leu Ser Arg
 1 5 10 15

Met Asn Ser Leu Gln Asp Gly Ile Thr Pro Ala Pro Ala Ala Pro Thr
 20 25 30

Asn Asp Thr Val Arg Gln Pro Lys Val Ser Asp Tyr Pro Leu Ala Thr
 35 40 45

Arg His Pro Glu Trp Val Lys Thr Ala Thr Asn Lys Thr Leu Asp Asp
 50 55 60

Leu Thr Leu Glu Asn Val Leu Ser Asp Arg Val Thr Ala Gln Asp Met
 65 70 75 80

Arg Ile Thr Pro Glu Thr Leu Arg Met Gln Ala Ala Ile Ala Gln Asp
 85 90 95

Ala Gly Arg Asp Arg Leu Ala Met Asn Phe Glu Arg Ala Ala Glu Leu
 100 105 110

Thr Ala Val Pro Asp Asp Arg Ile Leu Glu Ile Tyr Asn Ala Leu Arg
 115 120 125

-continued

Pro Tyr Arg Ser Thr Gln Ala Glu Leu Leu Ala Ile Ala Asp Asp Leu
 130 135 140

Glu His Arg Tyr Gln Ala Arg Leu Cys Ala Ala Phe Val Arg Glu Ala
 145 150 155 160

Ala Gly Leu Tyr Ile Glu Arg Lys Lys Leu Lys Gly Asp Asp
 165 170

<210> SEQ ID NO 109
 <211> LENGTH: 789
 <212> TYPE: DNA
 <213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 109

```

atgacagtca attatgattt ttccggaaaa gtcgtgctgg ttaccggcgc tggctctggt    60
attggccgtg ccaactgcgt tgccttcgcy cagtcgggcy catccgttgc ggtcgcagac   120
atctcgacty accacggttt gaaaaccgta gagttggtca aagccgaagg aggcgagggc   180
accttcttcc atgtcgatgt aggctctgaa cccagcgtcc agtcgatgct ggctggtgtc   240
gtggcgcatt acggcggcct ggacattgcy cacaacaacg ccggcattga ggccaatata   300
gtgccgtggy ccgagctgga ctccgacaac tggcgtcgtg tcatcgatgt gaacctttcc   360
tcggtgttct attgcctgaa aggtgaaatc cctctgatgc tgaaaaagggg cggcggcgcc   420
attgtgaata ccgcatcgcy ctccgggctg attggcggct atcgcctttc cgggtatacc   480
gcccaagaag acggcgtagt ggggctgact aaggctgctg ctatcgatta tgcaaaccag   540
aatatccgga ttaatgcctg gtgcctggtt ccagttgact cccattcctt ggctgacatg   600
ccgcaacca tgcgcgatcg acttctcttt ggcaactcaa ttggacgatt ggccaccgca   660
gaggagatcg cgcgttcggt tctgtggctg tgttctgacg atgcaaaata cgtggtgggg   720
cattcgatgt cagtcgacgg tggcgtggca gtgactgcyg ttggtactcg aatggatgat   780
ctcttttaa                                     789

```

<210> SEQ ID NO 110
 <211> LENGTH: 262
 <212> TYPE: PRT
 <213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 110

Met Thr Val Asn Tyr Asp Phe Ser Gly Lys Val Val Leu Val Thr Gly
 1 5 10 15

Ala Gly Ser Gly Ile Gly Arg Ala Thr Ala Leu Ala Phe Ala Gln Ser
 20 25 30

Gly Ala Ser Val Ala Val Ala Asp Ile Ser Thr Asp His Gly Leu Lys
 35 40 45

Thr Val Glu Leu Val Lys Ala Glu Gly Gly Glu Ala Thr Phe Phe His
 50 55 60

Val Asp Val Gly Ser Glu Pro Ser Val Gln Ser Met Leu Ala Gly Val
 65 70 75 80

Val Ala His Tyr Gly Gly Leu Asp Ile Ala His Asn Asn Ala Gly Ile
 85 90 95

Glu Ala Asn Ile Val Pro Leu Ala Glu Leu Asp Ser Asp Asn Trp Arg
 100 105 110

Arg Val Ile Asp Val Asn Leu Ser Ser Val Phe Tyr Cys Leu Lys Gly

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Lys Val Val Val Ala Asp Leu Asp Pro Val Gly Gly Glu Ala Thr Val
 35 40 45
 Ala Gln Ile His Ala Ala Gly Gly Glu Ala Leu Phe Ile Ala Cys Asp
 50 55 60
 Val Thr Arg Asp Ala Glu Val Arg Gln Leu His Glu Arg Leu Met Ala
 65 70 75 80
 Ala Tyr Gly Arg Leu Asp Tyr Ala Phe Asn Asn Ala Gly Ile Glu Ile
 85 90 95
 Glu Gln His Arg Leu Ala Glu Gly Ser Glu Ala Glu Phe Asp Ala Ile
 100 105 110
 Met Gly Val Asn Val Lys Gly Val Trp Leu Cys Met Lys Tyr Gln Leu
 115 120 125
 Pro Leu Leu Leu Ala Gln Gly Gly Gly Ala Ile Val Asn Thr Ala Ser
 130 135 140
 Val Ala Gly Leu Gly Ala Ala Pro Lys Met Ser Ile Tyr Ser Ala Ser
 145 150 155 160
 Lys His Ala Val Ile Gly Leu Thr Lys Ser Ala Ala Ile Glu Tyr Ala
 165 170 175
 Lys Lys Gly Ile Arg Val Asn Ala Val Cys Pro Ala Val Ile Asp Thr
 180 185 190
 Asp Met Phe Arg Arg Ala Tyr Gln Ala Asp Pro Arg Lys Ala Glu Phe
 195 200 205
 Ala Ala Ala Met His Pro Val Gly Arg Ile Gly Lys Val Glu Glu Ile
 210 215 220
 Ala Ser Ala Val Leu Tyr Leu Cys Ser Asp Gly Ala Ala Phe Thr Thr
 225 230 235 240
 Gly His Cys Leu Thr Val Asp Gly Gly Ala Thr Ala Ile
 245 250

<210> SEQ ID NO 113

<211> LENGTH: 810

<212> TYPE: DNA

<213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 113

```

atgtcttttc aaaacaaat cgttggtgctc acaggcgcag cttctggcat cggcaaagcg      60
acagcacagc tgctagtgga gcaggggccc catgtggttg ccatggatct taaaagcgac      120
ttgcttcaac aagcattcgg cagtgaggag cacgttctgt gcattccctac cgacgtcagc      180
gatagcgaag ccgtgcgagc cgccttcag gcagtggacg cgaaatttgg ccgtgtcgac      240
gtgattatta acgcccgggg catcaacgca cctacgcgag aagccaacca gaaaatggtt      300
gatgccaacg tcgctgccct cgatgccatg aagagcgggc gggcgccccc tttcgacttc      360
ctggccgata cctcggatca ggatttccgg cgcgtaatgg aagtcaattt gttcagccag      420
ttttactgca ttcgagaggg tgttcogctg atgcgccgag cgggtggcgg cagcatcgtc      480
aacatctcca gcgtggcagc gctcctgggc gtggcaatgc cactttacta ccccgctccc      540
aaggcggcgg tgctgggccc caccctgcca gggcagctg agttggcacc ttacaacatt      600
cgtgtgaaatg ccatcgctcc aggctctgtc gacacacatc tgatgcatga gcaaccaccg      660
gaagtcgttc agttcctggt cagcatgcaa cccatcaagc ggctggccca acccgaggag      720
cttgcccaaa gcatcctggt ccttgccggt gagcattcgt ctttcatcac cggacagacg      780

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-continued

ctttctccca acggcgggat gcacatgtaa

810

<210> SEQ ID NO 114

<211> LENGTH: 269

<212> TYPE: PRT

<213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 114

Met Ser Phe Gln Asn Lys Ile Val Val Leu Thr Gly Ala Ala Ser Gly
 1 5 10 15

Ile Gly Lys Ala Thr Ala Gln Leu Leu Val Glu Gln Gly Ala His Val
 20 25 30

Val Ala Met Asp Leu Lys Ser Asp Leu Leu Gln Gln Ala Phe Gly Ser
 35 40 45

Glu Glu His Val Leu Cys Ile Pro Thr Asp Val Ser Asp Ser Glu Ala
 50 55 60

Val Arg Ala Ala Phe Gln Ala Val Asp Ala Lys Phe Gly Arg Val Asp
 65 70 75 80

Val Ile Ile Asn Ala Ala Gly Ile Asn Ala Pro Thr Arg Glu Ala Asn
 85 90 95

Gln Lys Met Val Asp Ala Asn Val Ala Ala Leu Asp Ala Met Lys Ser
 100 105 110

Gly Arg Ala Pro Thr Phe Asp Phe Leu Ala Asp Thr Ser Asp Gln Asp
 115 120 125

Phe Arg Arg Val Met Glu Val Asn Leu Phe Ser Gln Phe Tyr Cys Ile
 130 135 140

Arg Glu Gly Val Pro Leu Met Arg Arg Ala Gly Gly Gly Ser Ile Val
 145 150 155 160

Asn Ile Ser Ser Val Ala Ala Leu Leu Gly Val Ala Met Pro Leu Tyr
 165 170 175

Tyr Pro Ala Ser Lys Ala Ala Val Leu Gly Leu Thr Arg Ala Ala Ala
 180 185 190

Ala Glu Leu Ala Pro Tyr Asn Ile Arg Val Asn Ala Ile Ala Pro Gly
 195 200 205

Ser Val Asp Thr Pro Leu Met His Glu Gln Pro Pro Glu Val Val Gln
 210 215 220

Phe Leu Val Ser Met Gln Pro Ile Lys Arg Leu Ala Gln Pro Glu Glu
 225 230 235 240

Leu Ala Gln Ser Ile Leu Phe Leu Ala Gly Glu His Ser Ser Phe Ile
 245 250 255

Thr Gly Gln Thr Leu Ser Pro Asn Gly Gly Met His Met
 260 265

<210> SEQ ID NO 115

<211> LENGTH: 771

<212> TYPE: DNA

<213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 115

atgacccttg aaggcaaac tgcaactcgtc accggttcca ccagcggcat tggcctgggc 60

atcgcccagg tattggcccg ggctggcgcc aacatcgtgc tcaacggctt tggtgaccgg 120

ggccccgcca tggcggaat tgccccgcac ggggtgaagg ttgtgcacca cccggccgac 180

-continued

```

ctgtcggatg tgggtccagat cgaggctttg ttcaacctgg ccgaacgcga gttcggcggc 240
gtcgacatcc tgggtcaaca cgccggatc cagcatgtgg caccgggtga gcagttccc 300
ccagaaagct gggacaagat catcgccctg aacctgtcgg ccgtattcca tggcacgcgc 360
ctggcgctgc cgggcatgcg cacgcgcaac tgggggcgca tcataaatat cgcttcggtg 420
catggcctgg tcggctcgat tggcaaggca gcctactgtg cagccaagca tggcgtgatc 480
ggcctgacca aggtggctcg cctggaaacc gccaccagtc atgtcacctg caatgccata 540
tgcccgggct ggggtgctgac accgctgggt caaaagcaga tcgacgatcg tgcggccaag 600
ggtggcgatc ggctgcaagc gcagcacgat ctgctggcag aaaagcaacc gtcgctggct 660
ttcgtcacc ccgaacacct cgggtgagctg gtactcttcc tgtgcagcga ggcggtagc 720
caggttcgcg gcgcccctg gaacgtcgat ggtggctggt tggcccagtg a 771

```

<210> SEQ ID NO 116

<211> LENGTH: 256

<212> TYPE: PRT

<213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 116

```

Met Thr Leu Glu Gly Lys Thr Ala Leu Val Thr Gly Ser Thr Ser Gly
1      5      10     15
Ile Gly Leu Gly Ile Ala Gln Val Leu Ala Arg Ala Gly Ala Asn Ile
20     25     30
Val Leu Asn Gly Phe Gly Asp Pro Gly Pro Ala Met Ala Glu Ile Ala
35     40     45
Arg His Gly Val Lys Val Val His His Pro Ala Asp Leu Ser Asp Val
50     55     60
Val Gln Ile Glu Ala Leu Phe Asn Leu Ala Glu Arg Glu Phe Gly Gly
65     70     75     80
Val Asp Ile Leu Val Asn Asn Ala Gly Ile Gln His Val Ala Pro Val
85     90     95
Glu Gln Phe Pro Pro Glu Ser Trp Asp Lys Ile Ile Ala Leu Asn Leu
100    105    110
Ser Ala Val Phe His Gly Thr Arg Leu Ala Leu Pro Gly Met Arg Thr
115    120    125
Arg Asn Trp Gly Arg Ile Ile Asn Ile Ala Ser Val His Gly Leu Val
130    135    140
Gly Ser Ile Gly Lys Ala Ala Tyr Val Ala Ala Lys His Gly Val Ile
145    150    155    160
Gly Leu Thr Lys Val Val Gly Leu Glu Thr Ala Thr Ser His Val Thr
165    170    175
Cys Asn Ala Ile Cys Pro Gly Trp Val Leu Thr Pro Leu Val Gln Lys
180    185    190
Gln Ile Asp Asp Arg Ala Ala Lys Gly Gly Asp Arg Leu Gln Ala Gln
195    200    205
His Asp Leu Leu Ala Glu Lys Gln Pro Ser Leu Ala Phe Val Thr Pro
210    215    220
Glu His Leu Gly Glu Leu Val Leu Phe Leu Cys Ser Glu Ala Gly Ser
225    230    235    240
Gln Val Arg Gly Ala Ala Trp Asn Val Asp Gly Gly Trp Leu Ala Gln
245    250    255

```

-continued

<210> SEQ ID NO 117

<211> LENGTH: 750

<212> TYPE: DNA

<213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 117

```

atgtccaagc aacttacact cgaaggcaaa gtggccctgg ttcagggcgg tccccgaggc    60
attggcgcgag ctatcgtaag gcgcctggcc cgcaaggcgg cgcaagtggc cttcacctat    120
gtcagctctg ccggccccgc tgaagaactg gctcgggaaa ttaccgagaa cggcggcaaa    180
gccttgggccc tgcgggctga cagcgtgat gccgcggccg tgcagctggc ggttgatgac    240
accgagaaaag ccttggggcgg gctggatata ctggtcaaca acgccgtgtg gctggcagtg    300
gccccagtga cagagttcga cctggccgac ttgatcata tgctggccgt gaacgtacgc    360
agcgtgttcg tcgccagcca ggccgcggca cgctatatgg gccagggcgg tcgtatcatc    420
aacattggca gcaccaacgc cgagcgcgat ccgtttgccg gtggtgcacc gtacgccatg    480
agcaagtggg cactggttgg tctgacctgc ggcatggcac gcgacctcgg gccgcagggc    540
attaccgtga acaacgtgca gcccgggccc gtggacaccg acatgaaccc ggccagtggc    600
gagtttgccc agagcctgat tccgctgatg gccattgggc gatatggcga gccggaggag    660
attgccagct tcgtggctta cctggcaggg cctgaagccg ggtatatcac cggggccagc    720
ctgactgtag atggtggggt tgcagcctga                                750

```

<210> SEQ ID NO 118

<211> LENGTH: 249

<212> TYPE: PRT

<213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 118

```

Met Ser Lys Gln Leu Thr Leu Glu Gly Lys Val Ala Leu Val Gln Gly
 1           5           10          15
Gly Ser Arg Gly Ile Gly Ala Ala Ile Val Arg Arg Leu Ala Arg Glu
 20          25          30
Gly Ala Gln Val Ala Phe Thr Tyr Val Ser Ser Ala Gly Pro Ala Glu
 35          40          45
Glu Leu Ala Arg Glu Ile Thr Glu Asn Gly Gly Lys Ala Leu Ala Leu
 50          55          60
Arg Ala Asp Ser Ala Asp Ala Ala Val Gln Leu Ala Val Asp Asp
 65          70          75          80
Thr Glu Lys Ala Leu Gly Arg Leu Asp Ile Leu Val Asn Asn Ala Gly
 85          90          95
Val Leu Ala Val Ala Pro Val Thr Glu Phe Asp Leu Ala Asp Phe Asp
100         105         110
His Met Leu Ala Val Asn Val Arg Ser Val Phe Val Ala Ser Gln Ala
115         120         125
Ala Ala Arg Tyr Met Gly Gln Gly Gly Arg Ile Ile Asn Ile Gly Ser
130         135         140
Thr Asn Ala Glu Arg Met Pro Phe Ala Gly Gly Ala Pro Tyr Ala Met
145         150         155         160
Ser Lys Ser Ala Leu Val Gly Leu Thr Arg Gly Met Ala Arg Asp Leu
165         170         175
Gly Pro Gln Gly Ile Thr Val Asn Asn Val Gln Pro Gly Pro Val Asp

```


-continued

| | | | | | | | | | | | | | | | |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| | 85 | | 90 | | 95 | | | | | | | | | | |
| Leu | Pro | Gly | Asp | Leu | Ala | Gln | Lys | Gln | His | Cys | His | Asp | Ile | Val | Asp |
| | | | 100 | | | | | 105 | | | | | 110 | | |
| Lys | Thr | Val | Ala | Gln | Phe | Gly | Arg | Ile | Asp | Ile | Leu | Val | Asn | Asn | Ala |
| | | 115 | | | | | 120 | | | | 125 | | | | |
| Ala | Phe | Gln | Met | Ala | His | Glu | Ser | Leu | Asp | Asp | Ile | Asp | Asp | Asp | Glu |
| | 130 | | | | | 135 | | | | | 140 | | | | |
| Trp | Val | Lys | Thr | Phe | Asp | Thr | Asn | Ile | Thr | Ala | Ile | Phe | Arg | Ile | Cys |
| 145 | | | | | 150 | | | | | 155 | | | | | 160 |
| Gln | Arg | Ala | Leu | Pro | Ser | Met | Pro | Lys | Gly | Gly | Ser | Ile | Ile | Asn | Thr |
| | | | | 165 | | | | | 170 | | | | | 175 | |
| Ser | Ser | Val | Asn | Ser | Asp | Asp | Pro | Ser | Pro | Ser | Leu | Leu | Ala | Tyr | Ala |
| | | | 180 | | | | | 185 | | | | | | 190 | |
| Ala | Thr | Lys | Gly | Ala | Ile | Ala | Asn | Phe | Thr | Ala | Gly | Leu | Ala | Gln | Leu |
| | | 195 | | | | | 200 | | | | | 205 | | | |
| Leu | Gly | Lys | Gln | Gly | Ile | Arg | Val | Asn | Ser | Val | Ala | Pro | Gly | Pro | Ile |
| | 210 | | | | | 215 | | | | | 220 | | | | |
| Trp | Thr | Pro | Leu | Ile | Pro | Ala | Thr | Met | Pro | Asp | Glu | Ala | Val | Arg | Asn |
| 225 | | | | | 230 | | | | | 235 | | | | | 240 |
| Phe | Gly | Ser | Gly | Tyr | Pro | Met | Gly | Arg | Pro | Gly | Gln | Pro | Val | Glu | Val |
| | | | | 245 | | | | | 250 | | | | | 255 | |
| Ala | Pro | Ile | Tyr | Val | Leu | Leu | Gly | Ser | Asp | Glu | Ala | Ser | Tyr | Ile | Ser |
| | | | 260 | | | | | 265 | | | | | 270 | | |
| Gly | Ser | Arg | Tyr | Ala | Val | Thr | Gly | Gly | Lys | Pro | Ile | Leu | | | |
| | | 275 | | | | | 280 | | | | | 285 | | | |

<210> SEQ ID NO 121
 <211> LENGTH: 774
 <212> TYPE: DNA
 <213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 121

```

atgatcgaaa tcagcggcag caccocgggc cacaatggcc gggtagcctt ggtcacgggc      60
gccgcccgcg gcatcggtct gggcattgcc gcatggctga tctgcaagg ctggcaagtg      120
gtgctgagtg atctggaccg ccagcgtggt accaaagtgg ccaaggcgtt gggcgacaac      180
gectggttca tcaccatgga cgttgccgac gaggcccagg tcagtgccgg cgtgtccgaa      240
gtgctcgggc agttcggccg gctggacgcg ctggtgtgca atgcggccat tgccaaccg      300
cacaaccaga cgctggaaag cctgagcctg gcacaatgga accgggtgct gggggtcAAC      360
ctcagcggcc ccatgctgct ggccaagcat tgtgcgccgt acctgcgtgc gcacaatggg      420
gcatcgtca acctgacctc taccctgctc cggcagtcgg aaccggacac cgaggcttac      480
gcggaagca agggcggcct ggtggctttg acccatgccc tggccatgag cctgggcccg      540
gagattcgcg tcaatgcggg gagcccgggc tggatcgatg cccgtgatcc gtcgcagcgc      600
cgtgccgagc cgttgagcga agctgaccat gccagcatc caacgggcag ggtaggggacc      660
gtggaagatg tcgcccgcct ggttgccctg ttgctgtcac gccaggcggc atttgtcacc      720
ggccaggagt ttgtggtcga tggcggcatg acccgcaaga tgatctatac ctga      774
    
```

<210> SEQ ID NO 122
 <211> LENGTH: 257
 <212> TYPE: PRT

-continued

<213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 122

```

Met Ile Glu Ile Ser Gly Ser Thr Pro Gly His Asn Gly Arg Val Ala
1      5      10      15
Leu Val Thr Gly Ala Ala Arg Gly Ile Gly Leu Gly Ile Ala Ala Trp
20     25     30
Leu Ile Cys Glu Gly Trp Gln Val Val Leu Ser Asp Leu Asp Arg Gln
35     40     45
Arg Gly Thr Lys Val Ala Lys Ala Leu Gly Asp Asn Ala Trp Phe Ile
50     55     60
Thr Met Asp Val Ala Asp Glu Ala Gln Val Ser Ala Gly Val Ser Glu
65     70     75     80
Val Leu Gly Gln Phe Gly Arg Leu Asp Ala Leu Val Cys Asn Ala Ala
85     90     95
Ile Ala Asn Pro His Asn Gln Thr Leu Glu Ser Leu Ser Leu Ala Gln
100    105    110
Trp Asn Arg Val Leu Gly Val Asn Leu Ser Gly Pro Met Leu Leu Ala
115    120    125
Lys His Cys Ala Pro Tyr Leu Arg Ala His Asn Gly Ala Ile Val Asn
130    135    140
Leu Thr Ser Thr Arg Ala Arg Gln Ser Glu Pro Asp Thr Glu Ala Tyr
145    150    155    160
Ala Ala Ser Lys Gly Gly Leu Val Ala Leu Thr His Ala Leu Ala Met
165    170    175
Ser Leu Gly Pro Glu Ile Arg Val Asn Ala Val Ser Pro Gly Trp Ile
180    185    190
Asp Ala Arg Asp Pro Ser Gln Arg Arg Ala Glu Pro Leu Ser Glu Ala
195    200    205
Asp His Ala Gln His Pro Thr Gly Arg Val Gly Thr Val Glu Asp Val
210    215    220
Ala Ala Met Val Ala Trp Leu Leu Ser Arg Gln Ala Ala Phe Val Thr
225    230    235    240
Gly Gln Glu Phe Val Val Asp Gly Gly Met Thr Arg Lys Met Ile Tyr
245    250    255

```

Thr

<210> SEQ ID NO 123

<211> LENGTH: 741

<212> TYPE: DNA

<213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 123

```

atgagcctgc aaggtaaagt tgcaactggt accggcgcca gccgtggcat tggccaggcc      60
atgccctcag agctgggccc ccaggggcgg accgtgatcg gtaccgccac gteggcgtec      120
ggtgccgagc gcatcgctgc caccctgaaa gaacacggca ttaccggcac tggcatggag      180
ctgaacgtga ccagcgccga atcggttgaa gccgtactgg ccgccattgg cgagcagttc      240
ggcgcgccgg ccattctggt caacaatgcc ggtatcacc cgcacaacct catgctgcgc      300
atgaaagacg acgagtgtgt tgatgtcatc gacaccaacc tgaacagcct ctaccgtctg      360
tccaagggcg tgctgcgtgg catgaccaag ggcggttggg gtcgtatcat cagcatcggc      420

```

-continued

```

tcggtcggtg gtgccatggg taacgcaggt caggccaact acgcggtgc caaggccggt 480
ctggaaggtt tcagccgcgc cctggcgcgt gaagtgggtt cgcgtggtat caccgtcaac 540
tcggtgaccc caggcttcat cgataccgac atgacccgcg agctgccaga agctcagcgc 600
gaagccctgc agaccagat tccgtggggc cgctggggcc aggctgacga aattgccaag 660
gtggtttcgt tcttggcatc cgacggcgcc gcctactga cggcgctac cgtgcccgtc 720
aacggcggga tgtacatgta a 741

```

```

<210> SEQ ID NO 124
<211> LENGTH: 246
<212> TYPE: PRT
<213> ORGANISM: Pseudomonas putida KT2440

```

```

<400> SEQUENCE: 124

```

```

Met Ser Leu Gln Gly Lys Val Ala Leu Val Thr Gly Ala Ser Arg Gly
1          5          10          15
Ile Gly Gln Ala Ile Ala Leu Glu Leu Gly Arg Gln Gly Ala Thr Val
20          25          30
Ile Gly Thr Ala Thr Ser Ala Ser Gly Ala Glu Arg Ile Ala Ala Thr
35          40          45
Leu Lys Glu His Gly Ile Thr Gly Thr Gly Met Glu Leu Asn Val Thr
50          55          60
Ser Ala Glu Ser Val Glu Ala Val Leu Ala Ala Ile Gly Glu Gln Phe
65          70          75          80
Gly Ala Pro Ala Ile Leu Val Asn Asn Ala Gly Ile Thr Arg Asp Asn
85          90          95
Leu Met Leu Arg Met Lys Asp Asp Glu Trp Phe Asp Val Ile Asp Thr
100         105         110
Asn Leu Asn Ser Leu Tyr Arg Leu Ser Lys Gly Val Leu Arg Gly Met
115         120         125
Thr Lys Ala Arg Trp Gly Arg Ile Ile Ser Ile Gly Ser Val Val Gly
130         135         140
Ala Met Gly Asn Ala Gly Gln Ala Asn Tyr Ala Ala Ala Lys Ala Gly
145         150         155         160
Leu Glu Gly Phe Ser Arg Ala Leu Ala Arg Glu Val Gly Ser Arg Gly
165         170         175
Ile Thr Val Asn Ser Val Thr Pro Gly Phe Ile Asp Thr Asp Met Thr
180         185         190
Arg Glu Leu Pro Glu Ala Gln Arg Glu Ala Leu Gln Thr Gln Ile Pro
195         200         205
Leu Gly Arg Leu Gly Gln Ala Asp Glu Ile Ala Lys Val Val Ser Phe
210         215         220
Leu Ala Ser Asp Gly Ala Ala Tyr Val Thr Gly Ala Thr Val Pro Val
225         230         235         240
Asn Gly Gly Met Tyr Met
245

```

```

<210> SEQ ID NO 125
<211> LENGTH: 738
<212> TYPE: DNA
<213> ORGANISM: Pseudomonas putida KT2440

```

```

<400> SEQUENCE: 125

```

-continued

```

atgactcaga aaatagctgt cgtgaccggc ggcagtcgcg gcattggcaa gtccatcgtg    60
ctggccctgg cggcgcgggg ttatcaggtt gccttcagtt atgtccgtga cgaggcgtca    120
gccgtgcct  tgcaggcgca ggtcgaaggg ctcggccggg actgcctggc cgtgcagtgt    180
gatgtcaagg aagcgccgag cattcaggcg ttttttgaac gggtcgagca acgtttcgag    240
cgtatcgact tgttggtcaa caacgccgtt attacccgtg acggtttgct cgccacgcaa    300
tcgttgaacg acatcaccga ggtcatccag accaacctgg tcggcacggt gttgtgctgt    360
cagcaggtgc tgcctgcat gatgcgcaa cgcagcgggt gcatcgtcaa cctcagttcg    420
gtggccgcbc aaaagcccgg caagggccag agcaactacg ccgcccgaaggcggtgta    480
gaagcattga cacgcgcact ggcggtggag ttggcgccgc gcaacatccg ggtcaacgcg    540
gtggcgcccg gcacgtcag caccgacatg agccaagccc tggtcggcgc ccatgagcag    600
gaaatccagt cgcggtggt gatcaaacgg ttcgcccggc ctgaagaaat tgcgacgcg    660
gtgctgtatc tggccgagcg cggcctgtac atcacgggcg aagtcctgtc cgtcaacggc    720
ggattgaaaa tgccatga                                                    738

```

<210> SEQ ID NO 126

<211> LENGTH: 245

<212> TYPE: PRT

<213> ORGANISM: Pseudomonas putida KT2440

<400> SEQUENCE: 126

```

Met Thr Gln Lys  Ile Ala Val Val Thr Gly Gly Ser Arg Gly Ile Gly
1          5          10          15
Lys Ser Ile Val Leu Ala Leu Ala Gly Ala Gly Tyr Gln Val Ala Phe
20        25        30
Ser Tyr Val Arg Asp Glu Ala Ser Ala Ala Ala Leu Gln Ala Gln Val
35        40        45
Glu Gly Leu Gly Arg Asp Cys Leu Ala Val Gln Cys Asp Val Lys Glu
50        55        60
Ala Pro Ser Ile Gln Ala Phe Phe Glu Arg Val Glu Gln Arg Phe Glu
65        70        75        80
Arg Ile Asp Leu Leu Val Asn Asn Ala Gly Ile Thr Arg Asp Gly Leu
85        90        95
Leu Ala Thr Gln Ser Leu Asn Asp Ile Thr Glu Val Ile Gln Thr Asn
100       105       110
Leu Val Gly Thr Leu Leu Cys Cys Gln Gln Val Leu Pro Cys Met Met
115       120       125
Arg Gln Arg Ser Gly Cys Ile Val Asn Leu Ser Ser Val Ala Ala Gln
130       135       140
Lys Pro Gly Lys Gly Gln Ser Asn Tyr Ala Ala Lys Gly Gly Val
145       150       155       160
Glu Ala Leu Thr Arg Ala Leu Ala Val Glu Leu Ala Pro Arg Asn Ile
165       170       175
Arg Val Asn Ala Val Ala Pro Gly Ile Val Ser Thr Asp Met Ser Gln
180       185       190
Ala Leu Val Gly Ala His Glu Gln Glu Ile Gln Ser Arg Leu Leu Ile
195       200       205
Lys Arg Phe Ala Arg Pro Glu Glu Ile Ala Asp Ala Val Leu Tyr Leu
210       215       220

```


-continued

| | | | |
|---|---------------------|-----------------------------|-----|
| 145 | 150 | 155 | 160 |
| Tyr Ser Val Thr | Lys Ala Ala Val Ile | Asn Met Thr Lys Val Phe Ala | |
| | 165 | 170 | 175 |
| Lys Glu Cys Ala Pro Phe Gly Ile Arg Cys Asn Ala Leu Leu Pro Gly | | | |
| | 180 | 185 | 190 |
| Leu Thr Asp Thr Lys Phe Ala Ser Ala Leu Val Lys Asn Glu Ala Ile | | | |
| | 195 | 200 | 205 |
| Leu Asn Ala Ala Leu Gln Gln Ile Pro Leu Lys Arg Val Ala Asp Pro | | | |
| | 210 | 215 | 220 |
| Lys Glu Met Ala Gly Ala Val Leu Tyr Leu Ala Ser Asp Ala Ser Ser | | | |
| | 225 | 230 | 235 |
| 240 | | | |
| Tyr Thr Thr Gly Thr Thr Leu Asn Val Asp Gly Gly Phe Leu Ser | | | |
| | 245 | 250 | 255 |

<210> SEQ ID NO 129
 <211> LENGTH: 762
 <212> TYPE: DNA
 <213> ORGANISM: Pseudomonas fluorescens Pf-5

<400> SEQUENCE: 129

```

atgagcatga cgttttccgg ccaggtggcc ctagtgaccg gcgagccaa tggatcggc 60
cgcgccaccg cccaggcatt tgccgcacaa ggcttgaagg tgggtgtggc ggacctggac 120
acggcggggg gcgagggcac cgtggcgctg atccgcgagg ccggtggcga ggcattgttc 180
gtgccgtgca acgttaccct ggaggcggat gtgcaaagcc tcatggcccg caccatcgaa 240
gcctatgggc gcctggatta cgccttaac aatgccgcta tcgagatcga aaagggccgc 300
cttgccgagg gctccatgga tgagttcgac gccatcatgg gggtaacgt caaaggggtc 360
tggctgtgca tgaagtacca gttgcccgtg ctgctggccc agggcgggtg ggcgatcgtc 420
aacaccgct cggtggcggg cctggggcgc ggcgcaaga tgagatcta tgcggcctcc 480
aagcatgctg tgatcggcct gaccaagtgc gcggccatcg aatatgcaaa gaagaaaatc 540
cgcgtgaaac cggtatgccc gccggtgatc gacaccgaca tgttccgccg tgctacgag 600
gcggaaccga agaagggcga gttcggcgcg gccatgcacc ccggtggggcg catcggaag 660
gtcgaggaga tcgccagtgc ggtgctctac ctgtgcagcg atggcgcggc ctttaccacc 720
ggccatgcac tggcggtcga cggcggggcc accgcatct ga 762
    
```

<210> SEQ ID NO 130
 <211> LENGTH: 253
 <212> TYPE: PRT
 <213> ORGANISM: Pseudomonas fluorscens Pf-5

<400> SEQUENCE: 130

| | | | | | | | | | | | | | | |
|---|--|--|----|--|--|--|----|--|--|--|--|----|--|----|
| Met Ser Met Thr Phe Ser Gly Gln Val Ala Leu Val Thr Gly Ala Ala | | | | | | | | | | | | | | |
| 1 | | | 5 | | | | 10 | | | | | 15 | | |
| Asn Gly Ile Gly Arg Ala Thr Ala Gln Ala Phe Ala Ala Gln Gly Leu | | | 20 | | | | 25 | | | | | 30 | | |
| Lys Val Val Val Ala Asp Leu Asp Thr Ala Gly Gly Glu Gly Thr Val | | | 35 | | | | 40 | | | | | 45 | | |
| Ala Leu Ile Arg Glu Ala Gly Gly Glu Ala Leu Phe Val Pro Cys Asn | | | 50 | | | | 55 | | | | | 60 | | |
| Val Thr Leu Glu Ala Asp Val Gln Ser Leu Met Ala Arg Thr Ile Glu | | | 65 | | | | 70 | | | | | 75 | | 80 |

-continued

Ala Tyr Gly Arg Leu Asp Tyr Ala Phe Asn Asn Ala Gly Ile Glu Ile
 85 90 95
 Glu Lys Gly Arg Leu Ala Glu Gly Ser Met Asp Glu Phe Asp Ala Ile
 100 105 110
 Met Gly Val Asn Val Lys Gly Val Trp Leu Cys Met Lys Tyr Gln Leu
 115 120 125
 Pro Leu Leu Leu Ala Gln Gly Gly Gly Ala Ile Val Asn Thr Ala Ser
 130 135 140
 Val Ala Gly Leu Gly Ala Ala Pro Lys Met Ser Ile Tyr Ala Ala Ser
 145 150 155 160
 Lys His Ala Val Ile Gly Leu Thr Lys Ser Ala Ala Ile Glu Tyr Ala
 165 170 175
 Lys Lys Lys Ile Arg Val Asn Ala Val Cys Pro Ala Val Ile Asp Thr
 180 185 190
 Asp Met Phe Arg Arg Ala Tyr Glu Ala Asp Pro Lys Lys Ala Glu Phe
 195 200 205
 Ala Ala Ala Met His Pro Val Gly Arg Ile Gly Lys Val Glu Glu Ile
 210 215 220
 Ala Ser Ala Val Leu Tyr Leu Cys Ser Asp Gly Ala Ala Phe Thr Thr
 225 230 235 240
 Gly His Ala Leu Ala Val Asp Gly Gly Ala Thr Ala Ile
 245 250

<210> SEQ ID NO 131
 <211> LENGTH: 735
 <212> TYPE: DNA
 <213> ORGANISM: Klebsiella pneumoniae subsp. pneumoniae MGH78578

<400> SEQUENCE: 131
 atgaaacttg ccagtaaac cgccattgtc accggcgccg cacgcggtat cggctttggc 60
 attgcccagg tgcttgccgc ggaaggcgcg cgagtgatta tcgccgatcg tgatgcacac 120
 ggcgaagccg ccgcccttc cctgcgcgaa tcgggcgcac aggcgctggt tatcagctgc 180
 aatatcgctg aaaaaacgca ggtcgaagcc ctgtattccc aggccgaaga ggcggttggc 240
 ccggtagaca ttctggtgaa taacgccgga atcaaccgcg acgccatgct gcacaaatta 300
 acggaagcgg actgggacac ggttatcgac gttaacctga aaggcacttt cctctgtatg 360
 cagcaggccg ctatccgcat gcgcgagcgc ggtgcgggcc gcattatcaa tatcgcttcc 420
 gccagttggc ttggcaacgt cgggcaaacc aactattcgg cgtcaaaagc cggcgtggtg 480
 ggaatgacca aaaccgctg ccgcgaactg gcgaaaaaag gtgtcacggt gaatgccatc 540
 tgcccgggct ttatcgatac cgacatgacg cgcggcgctac cggaaaaagt ctggcaaatc 600
 atggtcagca aaattcccgc gggttacgcc ggcgaggcga aagacgtcgg cgagtgtgtg 660
 gcgtttctgg cgtccgatgg cgccgcgtat atcaatggtg aagtgattaa cgtcggcggc 720
 ggcattggtg tgtaa 735

<210> SEQ ID NO 132
 <211> LENGTH: 253
 <212> TYPE: PRT
 <213> ORGANISM: Klebsiella pneumoniae subsp. pneumoniae MGH78578
 <400> SEQUENCE: 132

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| | | | | | | | | | | | | | | | |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Met | Ser | Met | Thr | Phe | Ser | Gly | Gln | Val | Ala | Leu | Val | Thr | Gly | Ala | Ala |
| 1 | | | | 5 | | | | | 10 | | | | | 15 | |
| Asn | Gly | Ile | Gly | Arg | Ala | Thr | Ala | Gln | Ala | Phe | Ala | Ala | Gln | Gly | Leu |
| | | | 20 | | | | | 25 | | | | | 30 | | |
| Lys | Val | Val | Val | Ala | Asp | Leu | Asp | Thr | Ala | Gly | Gly | Glu | Gly | Thr | Val |
| | | 35 | | | | | 40 | | | | | 45 | | | |
| Ala | Leu | Ile | Arg | Glu | Ala | Gly | Gly | Glu | Ala | Leu | Phe | Val | Pro | Cys | Asn |
| | 50 | | | | | 55 | | | | | 60 | | | | |
| Val | Thr | Leu | Glu | Ala | Asp | Val | Gln | Ser | Leu | Met | Ala | Arg | Thr | Ile | Glu |
| 65 | | | | | 70 | | | | | 75 | | | | | 80 |
| Ala | Tyr | Gly | Arg | Leu | Asp | Tyr | Ala | Phe | Asn | Asn | Ala | Gly | Ile | Glu | Ile |
| | | | | 85 | | | | | 90 | | | | | 95 | |
| Glu | Lys | Gly | Arg | Leu | Ala | Glu | Gly | Ser | Met | Asp | Glu | Phe | Asp | Ala | Ile |
| | | | 100 | | | | | 105 | | | | | 110 | | |
| Met | Gly | Val | Asn | Val | Lys | Gly | Val | Trp | Leu | Cys | Met | Lys | Tyr | Gln | Leu |
| | | 115 | | | | | 120 | | | | | 125 | | | |
| Pro | Leu | Leu | Leu | Ala | Gln | Gly | Gly | Gly | Ala | Ile | Val | Asn | Thr | Ala | Ser |
| | 130 | | | | | 135 | | | | | 140 | | | | |
| Val | Ala | Gly | Leu | Gly | Ala | Ala | Pro | Lys | Met | Ser | Ile | Tyr | Ala | Ala | Ser |
| 145 | | | | | 150 | | | | | 155 | | | | | 160 |
| Lys | His | Ala | Val | Ile | Gly | Leu | Thr | Lys | Ser | Ala | Ala | Ile | Glu | Tyr | Ala |
| | | | | 165 | | | | | 170 | | | | | 175 | |
| Lys | Lys | Lys | Ile | Arg | Val | Asn | Ala | Val | Cys | Pro | Ala | Val | Ile | Asp | Thr |
| | | | 180 | | | | | 185 | | | | | 190 | | |
| Asp | Met | Phe | Arg | Arg | Ala | Tyr | Glu | Ala | Asp | Pro | Lys | Lys | Ala | Glu | Phe |
| | | 195 | | | | | 200 | | | | | 205 | | | |
| Ala | Ala | Ala | Met | His | Pro | Val | Gly | Arg | Ile | Gly | Lys | Val | Glu | Glu | Ile |
| | 210 | | | | | 215 | | | | | 220 | | | | |
| Ala | Ser | Ala | Val | Leu | Tyr | Leu | Cys | Ser | Asp | Gly | Ala | Ala | Phe | Thr | Thr |
| 225 | | | | | 230 | | | | | 235 | | | | | 240 |
| Gly | His | Ala | Leu | Ala | Val | Asp | Gly | Gly | Ala | Thr | Ala | Ile | | | |
| | | | 245 | | | | | | 250 | | | | | | |

<210> SEQ ID NO 133

<211> LENGTH: 750

<212> TYPE: DNA

<213> ORGANISM: Klebsiella pneumoniae subsp. pneumoniae MGH78578

<400> SEQUENCE: 133

```

atgttattga aagataaagt cgccattatt actggcgcgg cctccgcacg cggtttgggc    60
ttcgcgactg cgaattattt cgccgaaaac ggcgcgaaag tggtcattat cgacctcaat    120
ggcgaagcca gtaaaaccgc cgccggcgca ttaggcgaag accatctcgg cctggcggcc    180
aacgtcgctg atgaagtgca ggtgcagcgc gccatcgaac agatcctggc gaaatacggc    240
cgggttgatg tactggtcaa taacgccggg attaccacgc cgctgaagct gatggatgc    300
aagcgcgcca actatgacgc ggtgcttgat gttagcctgc gcggcacgct gctgatgctg    360
caggcgggta tccccacat gcgggcgcaa aaatccggca gcatcgtctg catctcgtcc    420
gtctccgccc agcgcggcgg cggtattttc ggccgaccgc actacagcgc ggcaaaagcc    480
ggggtgctgg gtctggcgcg ggcgatggcg cgcgagcttg gcccgataa cgtccgcggt    540
aactgcatca ccccgggggt gattcagacc gacattaccg ccggcaagct gactgatgac    600

```

-continued

```

atgacggcca acattcttgc cggcattccg atgaaccgcc ttggcgacgc gatagacatc   660
gcgcgcgcgc cgctgttctc cggcagcgat ctttctctct actccaccgg catcaccttg   720
gacgttaacg gcggcatggt aattcactaa                                     750

```

```

<210> SEQ ID NO 134
<211> LENGTH: 249
<212> TYPE: PRT
<213> ORGANISM: Klebsiella pneumoniae subsp. pneumoniae MGH78578

```

```

<400> SEQUENCE: 134

```

```

Met Leu Leu Lys Asp Lys Val Ala Ile Ile Thr Gly Ala Ala Ser Ala
 1           5           10          15
Arg Gly Leu Gly Phe Ala Thr Ala Lys Leu Phe Ala Glu Asn Gly Ala
 20          25          30
Lys Val Val Ile Ile Asp Leu Asn Gly Glu Ala Ser Lys Thr Ala Ala
 35          40          45
Ala Ala Leu Gly Glu Asp His Leu Gly Leu Ala Ala Asn Val Ala Asp
 50          55          60
Glu Val Gln Val Gln Ala Ala Ile Glu Gln Ile Leu Ala Lys Tyr Gly
 65          70          75          80
Arg Val Asp Val Leu Val Asn Asn Ala Gly Ile Thr Gln Pro Leu Lys
 85          90          95
Leu Met Asp Ile Lys Arg Ala Asn Tyr Asp Ala Val Leu Asp Val Ser
 100         105         110
Leu Arg Gly Thr Leu Leu Met Ser Gln Ala Val Ile Pro Thr Met Arg
 115         120         125
Ala Gln Lys Ser Gly Ser Ile Val Cys Ile Ser Ser Val Ser Ala Gln
 130         135         140
Arg Gly Gly Gly Ile Phe Gly Gly Pro His Tyr Ser Ala Ala Lys Ala
 145         150         155         160
Gly Val Leu Gly Leu Ala Arg Ala Met Ala Arg Glu Leu Gly Pro Asp
 165         170         175
Asn Val Arg Val Asn Cys Ile Thr Pro Gly Leu Ile Gln Thr Asp Ile
 180         185         190
Thr Ala Gly Lys Leu Thr Asp Asp Met Thr Ala Asn Ile Leu Ala Gly
 195         200         205
Ile Pro Met Asn Arg Leu Gly Asp Ala Ile Asp Ile Ala Arg Ala Ala
 210         215         220
Leu Phe Leu Gly Ser Asp Leu Ser Ser Tyr Ser Thr Gly Ile Thr Leu
 225         230         235         240
Asp Val Asn Gly Gly Met Leu Ile His
 245

```

```

<210> SEQ ID NO 135
<211> LENGTH: 750
<212> TYPE: DNA
<213> ORGANISM: Klebsiella pneumoniae subsp. pneumoniae MGH78578

```

```

<400> SEQUENCE: 135

```

```

atgttattga aagataaagt cgccattatt actggcgcgg cctccgcacg cggtttgggc   60
ttcgcgactg cgaaattatt cgccgaaaac ggcgcgaaag tggtcattat cgacctcaat   120
ggcgaagcca gtaaaaccgc cgcggcggca ttaggcgaag accatctcgg cctggcggcc   180

```

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```

aacgtcgcctg atgaagtgca ggtgcaggcg gccatcgaac agatcctggc gaaatacggg 240
cgggttgatg tactggtcaa taacgccggg attacccagc cggtgaagct gatggatac 300
aagcgcgcga actatgacgc ggtgcttgat gttagcctgc gcggcacgct gctgatgctg 360
caggcgggta tccccacat gggggcgcaa aaatccggca gcacgtctg catctcgtcc 420
gtctccgccc agcgcggcgg cggtattttc ggcggaccgc actacagcgc ggcaaaagcc 480
ggggtgctgg gtctggcgcg ggcgatggcg cgcgagcttg gcccgataa cgtccgcggt 540
aactgatca cccccgggct gattcagacc gacattaccg ccggcaagct gactgatgac 600
atgacggcca acattcttgc cggcattccg atgaaccgcc ttggcgacgc gatagacatc 660
gcgcgcgcgc cgctgttctc cggcagcgat ctttctctct actccaecgg catcacctg 720
gacgttaacg gcggcatggt aattcactaa 750

```

<210> SEQ ID NO 136

<211> LENGTH: 249

<212> TYPE: PRT

<213> ORGANISM: Klebsiella pneumoniae subsp. pneumoniae MGH78578

<400> SEQUENCE: 136

```

Met Leu Leu Lys Asp Lys Val Ala Ile Ile Thr Gly Ala Ala Ser Ala
1      5      10      15
Arg Gly Leu Gly Phe Ala Thr Ala Lys Leu Phe Ala Glu Asn Gly Ala
20     25     30
Lys Val Val Ile Ile Asp Leu Asn Gly Glu Ala Ser Lys Thr Ala Ala
35     40     45
Ala Ala Leu Gly Glu Asp His Leu Gly Leu Ala Ala Asn Val Ala Asp
50     55     60
Glu Val Gln Val Gln Ala Ala Ile Glu Gln Ile Leu Ala Lys Tyr Gly
65     70     75     80
Arg Val Asp Val Leu Val Asn Asn Ala Gly Ile Thr Gln Pro Leu Lys
85     90     95
Leu Met Asp Ile Lys Arg Ala Asn Tyr Asp Ala Val Leu Asp Val Ser
100    105   110
Leu Arg Gly Thr Leu Leu Met Ser Gln Ala Val Ile Pro Thr Met Arg
115   120   125
Ala Gln Lys Ser Gly Ser Ile Val Cys Ile Ser Ser Val Ser Ala Gln
130   135   140
Arg Gly Gly Gly Ile Phe Gly Gly Pro His Tyr Ser Ala Ala Lys Ala
145   150   155   160
Gly Val Leu Gly Leu Ala Arg Ala Met Ala Arg Glu Leu Gly Pro Asp
165   170   175
Asn Val Arg Val Asn Cys Ile Thr Pro Gly Leu Ile Gln Thr Asp Ile
180   185   190
Thr Ala Gly Lys Leu Thr Asp Asp Met Thr Ala Asn Ile Leu Ala Gly
195   200   205
Ile Pro Met Asn Arg Leu Gly Asp Ala Ile Asp Ile Ala Arg Ala Ala
210   215   220
Leu Phe Leu Gly Ser Asp Leu Ser Ser Tyr Ser Thr Gly Ile Thr Leu
225   230   235   240
Asp Val Asn Gly Gly Met Leu Ile His
245

```

-continued

```

<210> SEQ ID NO 137
<211> LENGTH: 714
<212> TYPE: DNA
<213> ORGANISM: Klebsiella pneumoniae subsp. pneumoniae MGH78578

<400> SEQUENCE: 137
atgacagcgt ttcacaaca atcagtgctg gttttaggcg ggagtcgggg aattggcgcg    60
gcgatcgtea ggcgttttgt cgccgatggc gcgtcggtgg tgtttagcta ttccggttcg    120
ccggaagcgg ccgagcggct ggcggcagag accggcagca cggcggtgca ggcggacagc    180
gccgatcgcg atgcggtgat aagcctggtc cgcgacagcg gcccgctgga cgtgtagtg    240
gtcaatgcgc ggatcgcgct ttccggtgac gctctcgagc aggacagcga tgcaatcgat    300
cgctgtttcc acatcaatat tcacgcccc taccatgcct ccgtcgaagc ggcgcccgc    360
atgccggaag gcgggcgcac tattgtcatc ggctcagtea atggcgatcg catgccgttg    420
ccgggaatgg cggcctatgc gctcagcaaa tcggccctgc aggggctggc ggcgcccctg    480
gcgcgggatt ttggcccgcg cggcatcacg gtcaacgtcg tccagcccgg cccaattgat    540
accgacgcca acccgagaa cggcccgatg aaagagctga tgcacagctt tatggccatt    600
aagcgccatg gccgtccgga agagtgggc ggaatggtgg cgtggctggc cggtccggag    660
gcgtcgtttg tcaactggcg catgcacacc atcgacggag cgtttggcgc ctga      714

```

```

<210> SEQ ID NO 138
<211> LENGTH: 237
<212> TYPE: PRT
<213> ORGANISM: Klebsiella pneumoniae subsp. pneumoniae MGH78578

<400> SEQUENCE: 138
Met Thr Ala Phe His Asn Lys Ser Val Leu Val Leu Gly Gly Ser Arg
1      5      10      15
Gly Ile Gly Ala Ala Ile Val Arg Arg Phe Val Ala Asp Gly Ala Ser
20     25     30
Val Val Phe Ser Tyr Ser Gly Ser Pro Glu Ala Ala Glu Arg Leu Ala
35     40     45
Ala Glu Thr Gly Ser Thr Ala Val Gln Ala Asp Ser Ala Asp Arg Asp
50     55     60
Ala Val Ile Ser Leu Val Arg Asp Ser Gly Pro Leu Asp Val Leu Val
65     70     75     80
Val Asn Ala Gly Ile Ala Leu Phe Gly Asp Ala Leu Glu Gln Asp Ser
85     90     95
Asp Ala Ile Asp Arg Leu Phe His Ile Asn Ile His Ala Pro Tyr His
100    105    110
Ala Ser Val Glu Ala Ala Arg Arg Met Pro Glu Gly Gly Arg Ile Ile
115    120    125
Val Ile Gly Ser Val Asn Gly Asp Arg Met Pro Leu Pro Gly Met Ala
130    135    140
Ala Tyr Ala Leu Ser Lys Ser Ala Leu Gln Gly Leu Ala Arg Gly Leu
145    150    155    160
Ala Arg Asp Phe Gly Pro Arg Gly Ile Thr Val Asn Val Val Gln Pro
165    170    175
Gly Pro Ile Asp Thr Asp Ala Asn Pro Glu Asn Gly Pro Met Lys Glu
180    185    190

```

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Leu Met His Ser Phe Met Ala Ile Lys Arg His Gly Arg Pro Glu Glu
 195 200 205
 Val Ala Gly Met Val Ala Trp Leu Ala Gly Pro Glu Ala Ser Phe Val
 210 215 220
 Thr Gly Ala Met His Thr Ile Asp Gly Ala Phe Gly Ala
 225 230 235

<210> SEQ ID NO 139
 <211> LENGTH: 750
 <212> TYPE: DNA
 <213> ORGANISM: Klebsiella pneumoniae subsp. pneumoniae MGH78578

<400> SEQUENCE: 139

```

atgaacggcc tgctaaacgg taaacgtatt gtcgtcaccg gtgcggcgcg cggctctcggg    60
taccactttg ccgaagcctg cgccgctcag ggcgcgacgg tggatgatgtg cgacatcctg    120
cagggagagc tggcgaaaag cgctcatcgc ctgcagcaga agggctatca ggtcgaatct    180
cacgceatcg atcttgccag tcaagcatcg atcgagcagg tcttcagcgc catcgcgcg    240
caggggtcta tcgatggctt agtcaataac gcagcgatgg ccaccggcgt cggcggaaaa    300
aatatgatcg attacgatcc ggatctgtgg gatcgggtaa tgacgggtcaa cgttaaaggc    360
acctggttgg tgaccgcgcg ggcggtaccg ctgctgcgcg aagggcgggc gatcgtcaac    420
gtegcttcgg ataccgcgct gtggggcgcg ccgcggtga tggcctatgt cgccagtaag    480
ggcgcggtga ttgcgatgac ccgctccatg gcccgcgagc tgggtgaaaa gcggatccgt    540
atcaacgcca tcgcgccggg actgaccgcg gttgaggcca cggaatacgt tcccgcgag    600
cgtcatcagc tgtatgagaa cggccgcgcg ctcagcggcg cgcagcagcc ggaagatgtc    660
accggcagcg tggctctgct gctgagcgat ctttcgcgct ttatcaccgg ccaactgatc    720
cgggtcaacg gcggttttgt cttaactaa    750

```

<210> SEQ ID NO 140
 <211> LENGTH: 249
 <212> TYPE: PRT
 <213> ORGANISM: Klebsiella pneumoniae subsp. pneumoinae MGH78578

<400> SEQUENCE: 140

```

Met Asn Gly Leu Leu Asn Gly Lys Arg Ile Val Val Thr Gly Ala Ala
  1          5          10          15
Arg Gly Leu Gly Tyr His Phe Ala Glu Ala Cys Ala Ala Gln Gly Ala
  20          25          30
Thr Val Val Met Cys Asp Ile Leu Gln Gly Glu Leu Ala Glu Ser Ala
  35          40          45
His Arg Leu Gln Gln Lys Gly Tyr Gln Val Glu Ser His Ala Ile Asp
  50          55          60
Leu Ala Ser Gln Ala Ser Ile Glu Gln Val Phe Ser Ala Ile Gly Ala
  65          70          75          80
Gln Gly Ser Ile Asp Gly Leu Val Asn Asn Ala Ala Met Ala Thr Gly
  85          90          95
Val Gly Gly Lys Asn Met Ile Asp Tyr Asp Pro Asp Leu Trp Asp Arg
  100         105         110
Val Met Thr Val Asn Val Lys Gly Thr Trp Leu Val Thr Arg Ala Ala
  115         120         125
Val Pro Leu Leu Arg Glu Gly Ala Ala Ile Val Asn Val Ala Ser Asp

```


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| | | | | | | | | | | | | | | | |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Ala | Ala | Leu | Gln | Asn | Glu | Tyr | Pro | Glu | Gly | Glu | Val | Phe | Ser | Trp | Arg |
| | 50 | | | | | 55 | | | | | 60 | | | | |
| Cys | Asp | Val | Leu | Asn | Glu | Ala | Glu | Val | Glu | Ala | Phe | Ala | Ala | Ala | Val |
| 65 | | | | 70 | | | | | | 75 | | | | | 80 |
| Ala | Ala | Arg | Phe | Gly | Gly | Val | Asp | Met | Leu | Ile | Asn | Asn | Ala | Gly | Gln |
| | | | 85 | | | | | 90 | | | | | | 95 | |
| Gly | Tyr | Val | Ala | His | Phe | Ala | Asp | Thr | Pro | Arg | Glu | Ala | Trp | Leu | His |
| | | | 100 | | | | | 105 | | | | | | 110 | |
| Glu | Ala | Glu | Leu | Lys | Leu | Phe | Gly | Val | Ile | Asn | Pro | Val | Lys | Ala | Phe |
| | | | 115 | | | | 120 | | | | | | | 125 | |
| Gln | Ser | Leu | Leu | Glu | Ala | Ser | Asp | Ile | Ala | Ser | Ile | Thr | Cys | Val | Asn |
| | | 130 | | | | | 135 | | | | | 140 | | | |
| Ser | Leu | Leu | Ala | Leu | Gln | Pro | Glu | Glu | His | Met | Ile | Ala | Thr | Ser | Ala |
| 145 | | | | | 150 | | | | | | 155 | | | | 160 |
| Ala | Arg | Ala | Ala | Leu | Leu | Asn | Met | Thr | Leu | Thr | Leu | Ser | Lys | Glu | Leu |
| | | | | 165 | | | | | 170 | | | | | | 175 |
| Val | Asp | Lys | Gly | Ile | Arg | Val | Asn | Ser | Ile | Leu | Leu | Gly | Met | Val | Glu |
| | | | 180 | | | | | | 185 | | | | | 190 | |
| Ser | Gly | Gln | Trp | Gln | Arg | Arg | Phe | Glu | Ser | Arg | Ser | Asp | Lys | Ser | Gln |
| | | 195 | | | | | 200 | | | | | | 205 | | |
| Ser | Trp | Gln | Gln | Trp | Thr | Ala | Asp | Ile | Ala | Arg | Lys | Arg | Gly | Ile | Pro |
| | | 210 | | | | | 215 | | | | | 220 | | | |
| Met | Ala | Arg | Leu | Gly | Lys | Pro | Gln | Glu | Pro | Ala | Gln | Ala | Leu | Leu | Phe |
| 225 | | | | | 230 | | | | | | 235 | | | | 240 |
| Leu | Ala | Ser | Pro | Leu | Ala | Ser | Phe | Thr | Thr | Gly | Ala | Ala | Leu | Asp | Val |
| | | | | 245 | | | | | 250 | | | | | | 255 |
| Ser | Gly | Gly | Phe | Cys | Arg | His | Leu | | | | | | | | |
| | | | 260 | | | | | | | | | | | | |

<210> SEQ ID NO 143

<211> LENGTH: 1795

<212> TYPE: DNA

<213> ORGANISM: Pseudomonas fluorescens

<400> SEQUENCE: 143

```

cgccaagcaa tcgggctttg gggcagaatt gggtcgcgaa gggcttgagg agtttgccca    60
gtccaagatc atcaacgccg cgctataaat taaaggatcc cccatggcga tgattacagg    120
cggcgaactg gttgttcgca ccctaataaa ggctggggtc gaacatctgt tcggcctgca    180
cggcgcgcgc atcgatacga tttttcaagc ctgtctcgat catgatgtgc cgatcatcga    240
caccgcgcgc gaggcgcgcg cagggcatgc ggccgagggc tatgcccgcg ctggcgccaa    300
gctggggcgt gctggtcacg gcgggcgggg gatttaccac tgcggtcacg cccattgcca    360
acgcttggct ggatcgcaag gccggtgat tcctcaccgc ggatcgggcg cgctgcgtga    420
tgatgaaacc aacacgttgc aggcggggat tgatcaggtc gccatggcgg cgcgccattc    480
caaatgggcg catcgggtga tggcaaccga gcatatccca cggctggtga tgcaggcgat    540
ccgcgcgcgc ttgagcgcgc cacgcggggc ggtgttgctg gatctgccgt gggatattct    600
gatgaaccag attgatgagg atagegtcat tatcccgat ctggtcttgt ccgcgcattg    660
ggccagaccg gaccctgcgc atctggatca ggctctcgcg cttttgcgca agcgggagcg    720
gccggtcacg gtgctcggct cagaagcctc gcggacagcg cgcaagacgg cgcttagcgc    780

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cttcgtggcg gcgactggcg tgccggtggt tgccgattat gaagggctaa gcatgctctc 840
ggggctgccc gatgctatgc ggggcgggct ggtgcaaac ctctattctt ttgccaaagc 900
cgatgcgcgc ccagatctcg tgctgatgct gggggcgcgc tttggcctta acaccgggca 960
tggatctggg cagttgatcc cccatagcgc gcaggtcatt caggtcgacc ctgatgctg 1020
cgagctggga cgctgcagg gcatcgctct gggcattgtg gccgatgtgg gtgggacct 1080
cgaggctttg gcgcaggcca ccgcgcaaga tgccgcttgg ccggatcgcg gcgactggtg 1140
cgccaaagtg acggatctgg cgcaagagcg ctatgccagc atcgctcgga aatcgagcag 1200
cgagcatgcg ctccaccct ttcacgcctc gcaggtcatt gccaaacacg tcgatgcagg 1260
ggtgacgggtg gtageggatg gtgcgctgac ctatctctgg ctgtccgaag tgatgagccg 1320
cgtgaaaccc ggcggttttc tctgccacgg ctatctaggc tcgatgggcg tgggcttcgg 1380
cacggcgctg ggcgcgcaag tggccgatct tgaagcaggc cgccgcacga tccttgtgac 1440
cggcgatggc tcggtgggct atagcatcgg tgaatttgat acgctggtgc gcaacaatt 1500
gccgctgac gtcatcatca tgaacaacca aagctggggg gcgacattgc atttccagca 1560
attggccgct gccccaate gcgtgacggg caccgcttg gaaaatggct cctatcacgg 1620
ggtggccgcc gctttgggcg cggatggcta tcatgctgac agtgtggaga gcttttctgc 1680
ggctctggcc caagcgcctc cccataatcg ccccgctgc atcaatgtcg cggtcgcgct 1740
cgatccgatc ccgccgaag aactcattct gatcggcatg gacccttcg catga 1795

```

<210> SEQ ID NO 144

<211> LENGTH: 563

<212> TYPE: PRT

<213> ORGANISM: Pseudomonas fluorescens

<400> SEQUENCE: 144

```

Met Ala Met Ile Thr Gly Gly Glu Leu Val Val Arg Thr Leu Ile Lys
1           5           10          15
Ala Gly Val Glu His Leu Phe Gly Leu His Gly Ala His Ile Asp Thr
20          25          30
Ile Phe Gln Ala Cys Leu Asp His Asp Val Pro Ile Ile Asp Thr Arg
35          40          45
His Glu Ala Ala Ala Gly His Ala Ala Glu Gly Tyr Ala Arg Ala Gly
50          55          60
Ala Lys Leu Gly Val Ala Gly His Gly Gly Arg Gly Ile Tyr Gln Cys
65          70          75          80
Gly His Ala His Cys Gln Arg Leu Ala Gly Ser Gln Gly Arg Cys Ile
85          90          95
Pro His Pro Gly Ser Gly Ala Leu Arg Asp Asp Glu Thr Asn Thr Leu
100         105         110
Gln Ala Gly Ile Asp Gln Val Ala Met Ala Ala Pro Ile Thr Lys Trp
115         120         125
Ala His Arg Val Met Ala Thr Glu His Ile Pro Arg Leu Val Met Gln
130         135         140
Ala Ile Arg Ala Ala Leu Ser Ala Pro Arg Gly Pro Val Leu Leu Asp
145         150         155         160
Leu Pro Trp Asp Ile Leu Met Asn Gln Ile Asp Glu Asp Ser Val Ile
165         170         175
Ile Pro Asp Leu Val Leu Ser Ala His Gly Ala Arg Pro Asp Pro Ala

```

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| 180 | | | | | 185 | | | | | 190 | | | | | |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Asp | Leu | Asp | Gln | Ala | Leu | Ala | Leu | Leu | Arg | Lys | Ala | Glu | Arg | Pro | Val |
| | 195 | | | | | | 200 | | | | | 205 | | | |
| Ile | Val | Leu | Gly | Ser | Glu | Ala | Ser | Arg | Thr | Ala | Arg | Lys | Thr | Ala | Leu |
| | 210 | | | | | 215 | | | | | 220 | | | | |
| Ser | Ala | Phe | Val | Ala | Ala | Thr | Gly | Val | Pro | Val | Phe | Ala | Asp | Tyr | Glu |
| | 225 | | | | | 230 | | | | | 235 | | | | 240 |
| Gly | Leu | Ser | Met | Leu | Ser | Gly | Leu | Pro | Asp | Ala | Met | Arg | Gly | Gly | Leu |
| | | | | 245 | | | | | 250 | | | | | 255 | |
| Val | Gln | Asn | Leu | Tyr | Ser | Phe | Ala | Lys | Ala | Asp | Ala | Ala | Pro | Asp | Leu |
| | | | 260 | | | | | 265 | | | | | 270 | | |
| Val | Leu | Met | Leu | Gly | Ala | Arg | Phe | Gly | Leu | Asn | Thr | Gly | His | Gly | Ser |
| | | 275 | | | | | 280 | | | | | 285 | | | |
| Gly | Gln | Leu | Ile | Pro | His | Ser | Ala | Gln | Val | Ile | Gln | Val | Asp | Pro | Asp |
| | 290 | | | | | 295 | | | | | 300 | | | | |
| Ala | Cys | Glu | Leu | Gly | Arg | Leu | Gln | Gly | Ile | Ala | Leu | Gly | Ile | Val | Ala |
| | 305 | | | | | 310 | | | | | 315 | | | | 320 |
| Asp | Val | Gly | Gly | Thr | Ile | Glu | Ala | Leu | Ala | Gln | Ala | Thr | Ala | Gln | Asp |
| | | | | 325 | | | | | 330 | | | | | 335 | |
| Ala | Ala | Trp | Pro | Asp | Arg | Gly | Asp | Trp | Cys | Ala | Lys | Val | Thr | Asp | Leu |
| | | | 340 | | | | | 345 | | | | | 350 | | |
| Ala | Gln | Glu | Arg | Tyr | Ala | Ser | Ile | Ala | Ala | Lys | Ser | Ser | Ser | Glu | His |
| | | 355 | | | | | 360 | | | | | 365 | | | |
| Ala | Leu | His | Pro | Phe | His | Ala | Ser | Gln | Val | Ile | Ala | Lys | His | Val | Asp |
| | 370 | | | | | 375 | | | | | 380 | | | | |
| Ala | Gly | Val | Thr | Val | Val | Ala | Asp | Gly | Ala | Leu | Thr | Tyr | Leu | Trp | Leu |
| | 385 | | | | | 390 | | | | | 395 | | | | 400 |
| Ser | Glu | Val | Met | Ser | Arg | Val | Lys | Pro | Gly | Gly | Phe | Leu | Cys | His | Gly |
| | | | | 405 | | | | | 410 | | | | | 415 | |
| Tyr | Leu | Gly | Ser | Met | Gly | Val | Gly | Phe | Gly | Thr | Ala | Leu | Gly | Ala | Gln |
| | | | 420 | | | | 425 | | | | | | 430 | | |
| Val | Ala | Asp | Leu | Glu | Ala | Gly | Arg | Arg | Thr | Ile | Leu | Val | Thr | Gly | Asp |
| | | 435 | | | | | 440 | | | | | 445 | | | |
| Gly | Ser | Val | Gly | Tyr | Ser | Ile | Gly | Glu | Phe | Asp | Thr | Leu | Val | Arg | Lys |
| | 450 | | | | | 455 | | | | | 460 | | | | |
| Gln | Leu | Pro | Leu | Ile | Val | Ile | Ile | Met | Asn | Asn | Gln | Ser | Trp | Gly | Ala |
| | 465 | | | 470 | | | | | 475 | | | | | | 480 |
| Thr | Leu | His | Phe | Gln | Gln | Leu | Ala | Val | Gly | Pro | Asn | Arg | Val | Thr | Gly |
| | | | | 485 | | | | | 490 | | | | | 495 | |
| Thr | Arg | Leu | Glu | Asn | Gly | Ser | Tyr | His | Gly | Val | Ala | Ala | Ala | Phe | Gly |
| | | | 500 | | | | 505 | | | | | | 510 | | |
| Ala | Asp | Gly | Tyr | His | Val | Asp | Ser | Val | Glu | Ser | Phe | Ser | Ala | Ala | Leu |
| | | 515 | | | | | 520 | | | | | 525 | | | |
| Ala | Gln | Ala | Leu | Ala | His | Asn | Arg | Pro | Ala | Cys | Ile | Asn | Val | Ala | Val |
| | 530 | | | | | 535 | | | | | 540 | | | | |
| Ala | Leu | Asp | Pro | Ile | Pro | Pro | Glu | Glu | Leu | Ile | Leu | Ile | Gly | Met | Asp |
| | 545 | | | 550 | | | | | 555 | | | | | 560 | |
| Pro | Phe | Ala | | | | | | | | | | | | | |

<210> SEQ ID NO 145

<211> LENGTH: 9

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<212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: A polypeptide that is similar to an
 autotransporter adhesion or type I secretion
 target repeat.
 <220> FEATURE:
 <221> NAME/KEY: VARIANT
 <222> LOCATION: 3, 5, 7, 8, 9
 <223> OTHER INFORMATION: Xaa = Any Amino Acid

<400> SEQUENCE: 145

Gly Gly Xaa Gly Xaa Asp Xaa Xaa Xaa
 1 5

<210> SEQ ID NO 146
 <211> LENGTH: 50
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 146

gtctttattc atatatatat cctccttaat tcaaccgttc aatcaccatc 50

<210> SEQ ID NO 147
 <211> LENGTH: 30
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 147

gggcggccgc aaggggttcg cgttggccga 30

<210> SEQ ID NO 148
 <211> LENGTH: 22
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 148

ggagaaaata cgcacacagg cg 22

<210> SEQ ID NO 149
 <211> LENGTH: 32
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 149

cgggatccaa gttgcaggat atgacgaaag cg 32

<210> SEQ ID NO 150
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 150

gctctagaag attatccctg tctgcggaag cgg 33

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<210> SEQ ID NO 151
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 151

gctctagagg ggtgcctaat gagtgagcta ac 32

<210> SEQ ID NO 152
<211> LENGTH: 33
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 152

cgggatccgc gttaatattt tgttaaaatt cgc 33

<210> SEQ ID NO 153
<211> LENGTH: 31
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 153

gctctagagt ttatgtcgca cccgcggtg g 31

<210> SEQ ID NO 154
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 154

cccaagctta gaaagggaaa ttgtggtagc cc 32

<210> SEQ ID NO 155
<211> LENGTH: 31
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 155

ggaattccat atgcgtccct ctgccccggc c 31

<210> SEQ ID NO 156
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 156

cgggatcctt agaactgctt gggaaggag 30

<210> SEQ ID NO 157
<211> LENGTH: 50
<212> TYPE: DNA

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<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 157

aggtacggtg aaataaagga g gatatacat atgtccaaaa agattgccgt 50

<210> SEQ ID NO 158
<211> LENGTH: 37
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 158

ttttcctttt gggccgccc cgtggcatc gctcac 37

<210> SEQ ID NO 159
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 159

ggc gatgcca gcgtaaagga g gatatacat atgaaaaact ggaaaacaag 50

<210> SEQ ID NO 160
<211> LENGTH: 37
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 160

ttttcctttt gggccgccc cagcttagcg ccttcta 37

<210> SEQ ID NO 161
<211> LENGTH: 31
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 161

cccgagctct taggaggatt agtcatggaa c 31

<210> SEQ ID NO 162
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 162

gctctagatt attttgaata atcgtagaaa cc 32

<210> SEQ ID NO 163
<211> LENGTH: 42
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

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<400> SEQUENCE: 163

gctctagagg aggatatata tatgaaaaat tgtgtcatcg tc 42

<210> SEQ ID NO 164

<211> LENGTH: 30

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 164

aactgcagtt aattcaaccg ttcaatcacc 30

<210> SEQ ID NO 165

<211> LENGTH: 46

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 165

cgagctcagg aggatatata tatgaaaaat tgtgtcatcg tcagtg 46

<210> SEQ ID NO 166

<211> LENGTH: 50

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 166

ggttgaatta aggaggatat atatatgaat aaagacacac taatacctac 50

<210> SEQ ID NO 167

<211> LENGTH: 30

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 167

cccaagctta gccggcaagt acacatcttc 30

<210> SEQ ID NO 168

<211> LENGTH: 46

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 168

cgagctcagg aggatatata tatgaaaaat tgtgtcatcg tcagtg 46

<210> SEQ ID NO 169

<211> LENGTH: 30

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 169

cccaagctta gccggcaagt acacatcttc 30

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<210> SEQ ID NO 170
<211> LENGTH: 40
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 170

aaggaaaaaa gcgggccgcc ctgaaccgac gaccgggtcg 40

<210> SEQ ID NO 171
<211> LENGTH: 35
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 171

cggggtaccg cggatacata ttggaatgta tttag 35

<210> SEQ ID NO 172
<211> LENGTH: 44
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 172

aaggaaaaaa gcgggccgdc ggatacatat ttggaatgat ttg 44

<210> SEQ ID NO 173
<211> LENGTH: 43
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 173

gctctagagg aggatatata tatggctaac tacttcaata cac 43

<210> SEQ ID NO 174
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 174

tgctgttgcg ggtaaggag gatatatata tgcctaagta ccgttccgcc 50

<210> SEQ ID NO 175
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 175

aacggtactt aggcataat atactctct taaccgcaa cagcaatagc 50

<210> SEQ ID NO 176
<211> LENGTH: 30
<212> TYPE: DNA

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<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 176

acatgcatgc ttaaccccc agtttcgatt 30

<210> SEQ ID NO 177
<211> LENGTH: 43
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 177

gctctagagg aggatata tatggtaac tacttcaata cac 43

<210> SEQ ID NO 178
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 178

acatgcatgc ttaaccccc agtttcgatt 30

<210> SEQ ID NO 179
<211> LENGTH: 43
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 179

cccgagctca ggaggatata tatatggata aacagtatcc ggt 43

<210> SEQ ID NO 180
<211> LENGTH: 28
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 180

gctctagatt acagaatttg actcaggt 28

<210> SEQ ID NO 181
<211> LENGTH: 45
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 181

cccgagctca ggaggatata tatatgttga caaaagcaac aaaag 45

<210> SEQ ID NO 182
<211> LENGTH: 25
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

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<400> SEQUENCE: 182
ctctaaatct ctggaaggg taccg 25

<210> SEQ ID NO 183
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 183
gctctagatt agagagcttt cgttttcatg 30

<210> SEQ ID NO 184
<211> LENGTH: 45
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 184
cccagctca ggagatata tatatgttga caaaagcaac aaaag 45

<210> SEQ ID NO 185
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 185
gctctagatt agagagcttt cgttttcatg 30

<210> SEQ ID NO 186
<211> LENGTH: 46
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 186
cgagctcagg aggatata tatgagccag caagtcatta ttttcg 46

<210> SEQ ID NO 187
<211> LENGTH: 35
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 187
aaaactgcag cgtttgatga cgtggacgat agcgg 35

<210> SEQ ID NO 188
<211> LENGTH: 46
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 188
cgagctcagg aggatata tatgagccag caagtcatta ttttcg 46

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<210> SEQ ID NO 189
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 189

agggggtgtaa ggaggatata tatatggcta agacgttata cgaaaaattg 50

<210> SEQ ID NO 190
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 190

cgtcttagcc atatatatat cctcettaca ccccttctgc tacatagcgg 50

<210> SEQ ID NO 191
<211> LENGTH: 35
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 191

aaaactgcag cgtttgatga cgtggacgat agcgg 35

<210> SEQ ID NO 192
<211> LENGTH: 46
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 192

cgagctcagg aggatatata tatgagccag caagtcatta ttttcg 46

<210> SEQ ID NO 193
<211> LENGTH: 35
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 193

aaaactgcag cgtttgatga cgtggacgat agcgg 35

<210> SEQ ID NO 194
<211> LENGTH: 46
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 194

cgagctcagg aggatatata tatgagccag caagtcatta ttttcg 46

<210> SEQ ID NO 195
<211> LENGTH: 50
<212> TYPE: DNA

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<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 195
gaaaccgtgt gaggaggata tatatatgtc gaagaattac catattgccg 50

<210> SEQ ID NO 196
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 196
aggggtgtaa ggaggatata tatatggcta agacgttata cgaaaaattg 50

<210> SEQ ID NO 197
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 197
acattaaata agggagatat atatatggca gagaattta tcaaacacac 50

<210> SEQ ID NO 198
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 198
attcttcgac atatatatat cctcctcaca cggtttcctt gttgttttcg 50

<210> SEQ ID NO 199
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 199
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<210> SEQ ID NO 200
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 200
tttctctgcc atatatatat cctccttatt taatggtgcg aatgtcggcg 50

<210> SEQ ID NO 201
<211> LENGTH: 35
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

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<400> SEQUENCE: 201
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<210> SEQ ID NO 202
<211> LENGTH: 46
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 202
cgagctcagg aggatata tatgagccag caagtcatta ttttcg 46

<210> SEQ ID NO 203
<211> LENGTH: 35
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 203
aaaactgcag cgtttgatga cgtggacgat agcgg 35

<210> SEQ ID NO 204
<211> LENGTH: 40
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 204
aaggaaaaa gcggccgccc ctgaaccgac gaccgggtcg 40

<210> SEQ ID NO 205
<211> LENGTH: 35
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 205
cggggtaccg cgatacata ttgaaatgta ttttag 35

<210> SEQ ID NO 206
<211> LENGTH: 42
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 206
aaggaaaaa gcggccgac ttttcatact cccgccattc ag 42

<210> SEQ ID NO 207
<211> LENGTH: 31
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 207
caaaggccgt ctgcacgcgc cgaaaggcaa a 31

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<210> SEQ ID NO 208
<211> LENGTH: 31
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 208

tttgcctttc ggcgctgca gacggccttt g 31

<210> SEQ ID NO 209
<211> LENGTH: 35
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 209

acatgcatgc cgtttgatga cgtggacgat agcgg 35

<210> SEQ ID NO 210
<211> LENGTH: 42
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 210

aaggaaaaa gcggcgcac ttttcatact cccgccattc ag 42

<210> SEQ ID NO 211
<211> LENGTH: 35
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 211

acatgcatgc cgtttgatga cgtggacgat agcgg 35

<210> SEQ ID NO 212
<211> LENGTH: 48
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 212

cccgagctca ggaggatata tatatgaatt atcagaacga cgatttac 48

<210> SEQ ID NO 213
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 213

gcgtcgcggg taaggaggaa aattttatgt cctcacgtaa agagcttgcc 50

<210> SEQ ID NO 214
<211> LENGTH: 50
<212> TYPE: DNA

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<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 214
gaactgctgt aaggagggtta aaattatgga gaggattgtc gttactctcg 50

<210> SEQ ID NO 215
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 215
caatcagcgt aaggagggtat atataatgaa aaccgtaact gtaaaagatc 50

<210> SEQ ID NO 216
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 216
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<400> SEQUENCE: 218
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<400> SEQUENCE: 220

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<211> LENGTH: 48

<212> TYPE: DNA

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<400> SEQUENCE: 222

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<220> FEATURE:

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<223> OTHER INFORMATION: Primer

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<213> ORGANISM: Artificial Sequence
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<220> FEATURE:

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<220> FEATURE:

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<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 242

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<220> FEATURE:

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<212> TYPE: DNA

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<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 244

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<210> SEQ ID NO 245

<211> LENGTH: 32

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 245

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<210> SEQ ID NO 246
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<212> TYPE: DNA
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<220> FEATURE:
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<400> SEQUENCE: 246

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<210> SEQ ID NO 247
<211> LENGTH: 44
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
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<400> SEQUENCE: 247

aaggaaaaa gcggccgcgc ggatacatat ttgaatgtat ttag 44

<210> SEQ ID NO 248
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
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<400> SEQUENCE: 248

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<210> SEQ ID NO 249
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<212> TYPE: DNA
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<220> FEATURE:
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<400> SEQUENCE: 249

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<210> SEQ ID NO 250
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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 250

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<400> SEQUENCE: 251

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<210> SEQ ID NO 252
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<213> ORGANISM: Artificial Sequence
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<400> SEQUENCE: 252

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<212> TYPE: DNA
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<400> SEQUENCE: 253

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<212> TYPE: DNA
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<400> SEQUENCE: 254

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<210> SEQ ID NO 255
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
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<400> SEQUENCE: 255

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<212> TYPE: DNA
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<210> SEQ ID NO 257
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
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<400> SEQUENCE: 257

gctctagatt atgatttatt ttgttcagca aat 33

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<212> TYPE: DNA
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<400> SEQUENCE: 258

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<212> TYPE: DNA

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<220> FEATURE:

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<220> FEATURE:

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<211> LENGTH: 31

<212> TYPE: DNA

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<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 263

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<212> TYPE: DNA

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<220> FEATURE:

<223> OTHER INFORMATION: Primer

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<212> TYPE: DNA
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<400> SEQUENCE: 265

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<400> SEQUENCE: 266

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<210> SEQ ID NO 268
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<212> TYPE: DNA
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<220> FEATURE:
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<400> SEQUENCE: 268

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<212> TYPE: DNA
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<400> SEQUENCE: 269

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<210> SEQ ID NO 271
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<210> SEQ ID NO 272
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<212> TYPE: DNA
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<400> SEQUENCE: 272
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<210> SEQ ID NO 273
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<400> SEQUENCE: 273
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<210> SEQ ID NO 274
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<212> TYPE: DNA
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<220> FEATURE:
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<400> SEQUENCE: 274
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<210> SEQ ID NO 275
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 275
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<210> SEQ ID NO 276
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<212> TYPE: DNA
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<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 276
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<212> TYPE: DNA
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<220> FEATURE:
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<400> SEQUENCE: 277

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<210> SEQ ID NO 278

<211> LENGTH: 27

<212> TYPE: DNA

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<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 281

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<211> LENGTH: 30

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

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<220> FEATURE:

<223> OTHER INFORMATION: Primer

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<400> SEQUENCE: 286

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<210> SEQ ID NO 287
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<400> SEQUENCE: 287

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<210> SEQ ID NO 288
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<212> TYPE: DNA
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<212> TYPE: DNA
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<210> SEQ ID NO 290
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<213> ORGANISM: Artificial Sequence
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<400> SEQUENCE: 291

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<210> SEQ ID NO 292
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<210> SEQ ID NO 293
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<212> TYPE: DNA
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<400> SEQUENCE: 293

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<210> SEQ ID NO 294
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<212> TYPE: DNA
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<400> SEQUENCE: 294

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<210> SEQ ID NO 295
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
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<400> SEQUENCE: 295

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<210> SEQ ID NO 296
<211> LENGTH: 30
<212> TYPE: DNA
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<400> SEQUENCE: 296

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<210> SEQ ID NO 297

<211> LENGTH: 28

<212> TYPE: DNA

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<223> OTHER INFORMATION: Primer

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cgggatcctt aggatgcgta tgtaacca 28

<210> SEQ ID NO 298

<211> LENGTH: 30

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<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

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<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 300

ggaattccat atgccgatgg cgctcggcca 30

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<220> FEATURE:

<223> OTHER INFORMATION: Primer

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<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 302

ggaattccat atgaaacatt ctcaggacaa 30

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<210> SEQ ID NO 303
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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 303

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<210> SEQ ID NO 304
<211> LENGTH: 27
<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 304

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<210> SEQ ID NO 305
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<212> TYPE: DNA
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<400> SEQUENCE: 305

cgggatcctt atgccatacg ttccatat          28

<210> SEQ ID NO 306
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 306

ggaattccat atgcagcgtt ttaccaacag          30

<210> SEQ ID NO 307
<211> LENGTH: 28
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 307

cgggatcctt aggaaaacag gacgcgc          28

<210> SEQ ID NO 308
<211> LENGTH: 610
<212> TYPE: PRT
<213> ORGANISM: Klebsiella pneumoniae subsp. pneumoniae MGH 78578

<400> SEQUENCE: 308

Met Arg Tyr Ile Ala Gly Ile Asp Ile Gly Asn Ser Ser Thr Glu Val
1           5           10           15

Ala Leu Ala Thr Val Asp Asp Ala Gly Val Leu Asn Ile Arg His Ser
20          25          30

Ala Leu Ala Glu Thr Thr Gly Ile Lys Gly Thr Leu Arg Asn Val Phe
35          40          45

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Gly Ile Gln Glu Ala Leu Thr Gln Ala Ala Lys Ala Ala Gly Ile Gln
 50 55 60

Leu Ser Asp Ile Ser Leu Ile Arg Ile Asn Glu Ala Thr Pro Val Ile
 65 70 75 80

Gly Asp Val Ala Met Glu Thr Ile Thr Glu Thr Ile Ile Thr Glu Ser
 85 90 95

Thr Met Ile Gly His Asn Pro Lys Thr Pro Gly Gly Val Gly Leu Gly
 100 105 110

Val Gly Ile Thr Ile Thr Pro Glu Ala Leu Leu Ser Cys Ser Ala Asp
 115 120 125

Thr Pro Tyr Ile Leu Val Val Ser Ser Ala Phe Asp Phe Ala Asp Val
 130 135 140

Ala Ala Met Val Asn Ala Ala Thr Ala Ala Gly Tyr Gln Ile Thr Gly
 145 150 155 160

Ile Ile Leu Gln Gln Asp Asp Gly Val Leu Val Asn Asn Arg Leu Gln
 165 170 175

Gln Pro Leu Pro Val Ile Asp Glu Val Gln His Ile Asp Arg Ile Pro
 180 185 190

Leu Gly Met Leu Ala Ala Val Glu Val Ala Leu Pro Gly Lys Ile Ile
 195 200 205

Glu Thr Leu Ser Asn Pro Tyr Gly Ile Ala Thr Val Phe Asp Leu Asn
 210 215 220

Ala Glu Glu Thr Lys Asn Ile Val Pro Met Ala Arg Ala Leu Ile Gly
 225 230 235 240

Asn Arg Ser Ala Val Val Val Lys Thr Pro Ser Gly Asp Val Lys Ala
 245 250 255

Arg Ala Ile Pro Ala Gly Asn Leu Leu Ile Ala Gln Gly Arg Ser
 260 265 270

Val Gln Val Asp Val Ala Ala Gly Ala Glu Ala Ile Met Lys Ala Val
 275 280 285

Asp Gly Cys Gly Lys Leu Asp Asn Val Ala Gly Glu Ala Gly Thr Asn
 290 295 300

Ile Gly Gly Met Leu Glu His Val Arg Gln Thr Met Ala Glu Leu Thr
 305 310 315 320

Asn Lys Pro Ala Gln Glu Ile Arg Ile Gln Asp Leu Leu Ala Val Asp
 325 330 335

Thr Ala Val Pro Val Ser Val Thr Gly Gly Leu Ala Gly Glu Phe Ser
 340 345 350

Leu Glu Gln Ala Val Gly Ile Ala Ser Met Val Lys Ser Asp Arg Leu
 355 360 365

Gln Met Ala Leu Ile Ala Arg Glu Ile Glu His Lys Leu Gln Ile Ala
 370 375 380

Val Gln Val Gly Gly Ala Glu Ala Glu Ala Ala Ile Leu Gly Ala Leu
 385 390 395 400

Thr Thr Pro Gly Thr Thr Arg Pro Leu Ala Ile Leu Asp Leu Gly Ala
 405 410 415

Gly Ser Thr Asp Ala Ser Ile Ile Asn Ala Gln Gly Glu Ile Ser Ala
 420 425 430

Thr His Leu Ala Gly Ala Gly Asp Met Val Thr Met Ile Ile Ala Arg
 435 440 445

-continued

Glu Leu Gly Leu Glu Asp Arg Tyr Leu Ala Glu Glu Ile Lys Lys Tyr
 450 455 460
 Pro Leu Ala Lys Val Glu Ser Leu Phe His Leu Arg His Glu Asp Gly
 465 470 475 480
 Ser Val Gln Phe Phe Pro Ser Ala Leu Pro Pro Ala Val Phe Ala Arg
 485 490 495
 Val Cys Val Val Lys Pro Asp Glu Leu Val Pro Leu Pro Gly Asp Leu
 500 505 510
 Pro Leu Glu Lys Val Arg Ala Ile Arg Arg Ser Ala Lys Ser Arg Val
 515 520 525
 Phe Val Thr Asn Ala Leu Arg Ala Leu Arg Gln Val Ser Pro Thr Gly
 530 535 540
 Asn Ile Arg Asp Ile Pro Phe Val Val Leu Val Gly Gly Ser Ser Leu
 545 550 555 560
 Asp Phe Glu Ile Pro Gln Leu Val Thr Asp Ala Leu Ala His Tyr Arg
 565 570 575
 Leu Val Ala Gly Arg Gly Asn Ile Arg Gly Cys Glu Gly Pro Arg Asn
 580 585 590
 Ala Val Ala Ser Gly Leu Leu Leu Ser Trp Gln Lys Gly Gly Thr His
 595 600 605
 Gly Glu
 610

<210> SEQ ID NO 309
 <211> LENGTH: 116
 <212> TYPE: PRT
 <213> ORGANISM: Klebsiella pneumoniae subsp. pneumoniae MGH78578

<400> SEQUENCE: 309

Met Glu Ser Ser Val Val Ala Pro Ala Ile Val Ile Ala Val Thr Asp
 1 5 10 15
 Glu Cys Ser Glu Gln Trp Arg Asp Val Leu Leu Gly Ile Glu Glu Glu
 20 25 30
 Gly Ile Pro Phe Val Leu Gln Pro Gln Thr Gly Gly Asp Leu Ile His
 35 40 45
 His Ala Trp Gln Ala Ala Gln Arg Ser Pro Leu Gln Val Gly Ile Ala
 50 55 60
 Cys Asp Arg Glu Arg Leu Ile Val His Tyr Lys Asn Leu Pro Ala Ser
 65 70 75 80
 Thr Pro Leu Phe Ser Leu Met Tyr His Gln Asn Arg Leu Ala Arg Arg
 85 90 95
 Asn Thr Gly Asn Asn Ala Ala Arg Leu Val Lys Gly Ile Pro Phe Arg
 100 105 110
 Asp Arg His Ala
 115

<210> SEQ ID NO 310
 <211> LENGTH: 787
 <212> TYPE: PRT
 <213> ORGANISM: Clostridium butyricum

<400> SEQUENCE: 310

Met Ile Ser Lys Gly Phe Ser Thr Gln Thr Glu Arg Ile Asn Ile Leu
 1 5 10 15

-continued

Lys Ala Gln Ile Leu Asn Ala Lys Pro Cys Val Glu Ser Glu Arg Ala
 20 25 30
 Ile Leu Ile Thr Glu Ser Phe Lys Gln Thr Glu Gly Gln Pro Ala Ile
 35 40 45
 Leu Arg Arg Ala Leu Ala Leu Lys His Ile Leu Glu Asn Ile Pro Ile
 50 55 60
 Thr Ile Arg Asp Gln Glu Leu Ile Val Gly Ser Leu Thr Lys Glu Pro
 65 70 75 80
 Arg Ser Ser Gln Val Phe Pro Glu Phe Ser Asn Lys Trp Leu Gln Asp
 85 90
 Glu Leu Asp Arg Leu Asn Lys Arg Thr Gly Asp Ala Phe Gln Ile Ser
 100 105 110
 Glu Glu Ser Lys Glu Lys Leu Lys Asp Val Phe Glu Tyr Trp Asn Gly
 115 120 125
 Lys Thr Thr Ser Glu Leu Ala Thr Ser Tyr Met Thr Glu Glu Thr Arg
 130 135 140
 Glu Ala Val Asn Cys Asp Val Phe Thr Val Gly Asn Tyr Tyr Tyr Asn
 145 150 155 160
 Gly Val Gly His Val Ser Val Asp Tyr Gly Lys Val Leu Arg Val Gly
 165 170 175
 Phe Asn Gly Ile Ile Asn Glu Ala Lys Glu Gln Leu Glu Lys Asn Arg
 180 185 190
 Ser Ile Asp Pro Asp Phe Ile Lys Lys Glu Lys Phe Leu Asn Ser Val
 195 200 205
 Ile Ile Ser Cys Glu Ala Ala Ile Thr Tyr Val Asn Arg Tyr Ala Lys
 210 215 220
 Lys Ala Lys Glu Ile Ala Asp Asn Thr Ser Asp Ala Lys Arg Lys Ala
 225 230 235 240
 Glu Leu Asn Glu Ile Ala Lys Ile Cys Ser Lys Val Ser Gly Glu Gly
 245 250 255
 Ala Lys Ser Phe Tyr Glu Ala Cys Gln Leu Phe Trp Phe Ile His Ala
 260 265 270
 Ile Ile Asn Ile Glu Ser Asn Gly His Ser Ile Ser Pro Ala Arg Phe
 275 280 285
 Asp Gln Tyr Met Tyr Pro Tyr Tyr Glu Asn Asp Lys Asn Ile Thr Asp
 290 295 300
 Lys Phe Ala Gln Glu Leu Ile Asp Cys Ile Trp Ile Lys Leu Asn Asp
 305 310 315 320
 Ile Asn Lys Val Arg Asp Glu Ile Ser Thr Lys His Phe Gly Gly Tyr
 325 330 335
 Pro Met Tyr Gln Asn Leu Ile Val Gly Gly Gln Asn Ser Glu Gly Lys
 340 345 350
 Asp Ala Thr Asn Lys Val Ser Tyr Met Ala Leu Glu Ala Ala Val His
 355 360 365
 Val Lys Leu Pro Gln Pro Ser Leu Ser Val Arg Ile Trp Asn Lys Thr
 370 375 380
 Pro Asp Glu Phe Leu Leu Arg Ala Ala Glu Leu Thr Arg Glu Gly Leu
 385 390 395 400
 Gly Leu Pro Ala Tyr Tyr Asn Asp Glu Val Ile Ile Pro Ala Leu Val
 405 410 415
 Ser Arg Gly Leu Thr Leu Glu Asp Ala Arg Asp Tyr Gly Ile Ile Gly

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| 420 | | | | | 425 | | | | | 430 | | | | | |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Cys | Val | Glu | Pro | Gln | Lys | Pro | Gly | Lys | Thr | Glu | Gly | Trp | His | Asp | Ser |
| | 435 | | | | | | 440 | | | | | 445 | | | |
| Ala | Phe | Phe | Asn | Leu | Ala | Arg | Ile | Val | Glu | Leu | Thr | Ile | Asn | Ser | Gly |
| | 450 | | | | | 455 | | | | | 460 | | | | |
| Phe | Asp | Lys | Asn | Lys | Gln | Ile | Gly | Pro | Lys | Thr | Gln | Asn | Phe | Glu | Glu |
| 465 | | | | | 470 | | | | | 475 | | | | | 480 |
| Met | Lys | Ser | Phe | Asp | Glu | Phe | Met | Lys | Ala | Tyr | Lys | Ala | Gln | Met | Glu |
| | | | | 485 | | | | | | 490 | | | | | 495 |
| Tyr | Phe | Val | Lys | His | Met | Cys | Cys | Ala | Asp | Asn | Cys | Ile | Asp | Ile | Ala |
| | | | 500 | | | | | 505 | | | | | 510 | | |
| His | Ala | Glu | Arg | Ala | Pro | Leu | Pro | Phe | Leu | Ser | Ser | Met | Val | Asp | Asn |
| | | 515 | | | | | 520 | | | | | 525 | | | |
| Cys | Ile | Gly | Lys | Gly | Lys | Ser | Leu | Gln | Asp | Gly | Gly | Ala | Glu | Tyr | Asn |
| | 530 | | | | | 535 | | | | | 540 | | | | |
| Phe | Ser | Gly | Pro | Gln | Gly | Val | Gly | Val | Ala | Asn | Ile | Gly | Asp | Ser | Leu |
| 545 | | | | | 550 | | | | | 555 | | | | | 560 |
| Val | Ala | Val | Lys | Lys | Ile | Val | Phe | Asp | Glu | Asn | Lys | Ile | Thr | Pro | Ser |
| | | | | 565 | | | | | 570 | | | | | 575 | |
| Glu | Leu | Lys | Lys | Thr | Leu | Asn | Asn | Asp | Phe | Lys | Asn | Ser | Glu | Glu | Ile |
| | | | 580 | | | | | 585 | | | | | 590 | | |
| Gln | Ala | Leu | Leu | Lys | Asn | Ala | Pro | Lys | Phe | Gly | Asn | Asp | Ile | Asp | Glu |
| | | 595 | | | | | 600 | | | | | 605 | | | |
| Val | Asp | Asn | Leu | Ala | Arg | Glu | Gly | Ala | Leu | Val | Tyr | Cys | Arg | Glu | Val |
| | 610 | | | | | 615 | | | | | 620 | | | | |
| Asn | Lys | Tyr | Thr | Asn | Pro | Arg | Gly | Gly | Asn | Phe | Gln | Pro | Gly | Leu | Tyr |
| 625 | | | | | 630 | | | | | 635 | | | | | 640 |
| Pro | Ser | Ser | Ile | Asn | Val | Tyr | Phe | Gly | Ser | Leu | Thr | Gly | Ala | Thr | Pro |
| | | | | 645 | | | | | 650 | | | | | 655 | |
| Asp | Gly | Arg | Lys | Ser | Gly | Gln | Pro | Leu | Ala | Asp | Gly | Val | Ser | Pro | Ser |
| | | | 660 | | | | | 665 | | | | | 670 | | |
| Arg | Gly | Cys | Asp | Val | Ser | Gly | Pro | Thr | Ala | Ala | Cys | Asn | Ser | Val | Ser |
| | | 675 | | | | | 680 | | | | | 685 | | | |
| Lys | Leu | Asp | His | Phe | Ile | Ala | Ser | Asn | Gly | Thr | Leu | Phe | Asn | Gln | Lys |
| | 690 | | | | | 695 | | | | | 700 | | | | |
| Phe | His | Pro | Ser | Ala | Leu | Lys | Gly | Asp | Asn | Gly | Leu | Met | Asn | Leu | Ser |
| 705 | | | | | 710 | | | | | 715 | | | | | 720 |
| Ser | Leu | Ile | Arg | Ser | Tyr | Phe | Asp | Gln | Lys | Gly | Phe | His | Val | Gln | Phe |
| | | | | 725 | | | | | 730 | | | | | 735 | |
| Asn | Val | Ile | Asp | Lys | Lys | Ile | Leu | Leu | Ala | Ala | Gln | Lys | Asn | Pro | Glu |
| | | | 740 | | | | | 745 | | | | | 750 | | |
| Lys | Tyr | Gln | Asp | Leu | Ile | Val | Arg | Val | Ala | Gly | Tyr | Ser | Ala | Gln | Phe |
| | | 755 | | | | | 760 | | | | | 765 | | | |
| Ile | Ser | Leu | Asp | Lys | Ser | Ile | Gln | Asn | Asp | Ile | Ile | Ala | Arg | Thr | Glu |
| | 770 | | | | | 775 | | | | | 780 | | | | |
| His | Val | Met | | | | | | | | | | | | | |
| 785 | | | | | | | | | | | | | | | |

<210> SEQ ID NO 311

<211> LENGTH: 304

<212> TYPE: PRT

<213> ORGANISM: Clostridium buyricum

-continued

<400> SEQUENCE: 311

Met Ser Lys Glu Ile Lys Gly Val Leu Phe Asn Ile Gln Lys Phe Ser
 1 5 10 15
 Leu His Asp Gly Pro Gly Ile Arg Thr Ile Val Phe Phe Lys Gly Cys
 20 25 30
 Ser Met Ser Cys Leu Trp Cys Ser Asn Pro Glu Ser Gln Asp Ile Lys
 35 40 45
 Pro Gln Val Met Phe Asn Lys Asn Leu Cys Thr Lys Cys Gly Arg Cys
 50 55 60
 Lys Ser Gln Cys Lys Ser Ala Ala Ile Asp Met Asn Ser Glu Tyr Arg
 65 70 75 80
 Ile Asp Lys Ser Lys Cys Thr Glu Cys Thr Lys Cys Val Asp Asn Cys
 85 90 95
 Leu Ser Gly Ala Leu Val Ile Glu Gly Arg Asn Tyr Ser Val Glu Asp
 100 105 110
 Val Ile Lys Glu Leu Lys Lys Asp Ser Val Gln Tyr Arg Arg Ser Asn
 115 120 125
 Gly Gly Ile Thr Leu Ser Gly Gly Glu Val Leu Leu Gln Pro Asp Phe
 130 135 140
 Ala Val Glu Leu Leu Lys Glu Cys Lys Ser Tyr Gly Trp His Thr Ala
 145 150 155 160
 Ile Glu Thr Ala Met Tyr Val Asn Ser Glu Ser Val Lys Lys Val Ile
 165 170 175
 Pro Tyr Ile Asp Leu Ala Met Ile Asp Ile Lys Ser Met Asn Asp Glu
 180 185 190
 Ile His Arg Lys Phe Thr Gly Val Ser Asn Glu Ile Ile Leu Gln Asn
 195 200 205
 Ile Lys Leu Ser Asp Glu Leu Ala Lys Glu Ile Ile Ile Arg Ile Pro
 210 215 220
 Val Ile Glu Gly Phe Asn Ala Asp Leu Gln Ser Ile Gly Ala Ile Ala
 225 230 235 240
 Gln Phe Ser Lys Ser Leu Thr Asn Leu Lys Arg Ile Asp Leu Leu Pro
 245 250 255
 Tyr His Asn Tyr Gly Glu Asn Lys Tyr Gln Ala Ile Gly Arg Glu Tyr
 260 265 270
 Ser Leu Lys Glu Leu Lys Ser Pro Ser Lys Asp Lys Met Glu Arg Leu
 275 280 285
 Lys Ala Leu Val Glu Ile Met Gly Ile Pro Cys Thr Ile Gly Ala Glu
 290 295 300

<210> SEQ ID NO 312

<211> LENGTH: 545

<212> TYPE: PRT

<213> ORGANISM: Azospirillum brasilense

<400> SEQUENCE: 312

Met Lys Leu Ala Glu Ala Leu Leu Arg Ala Leu Lys Asp Arg Gly Ala
 1 5 10 15
 Gln Ala Met Phe Gly Ile Pro Gly Asp Phe Ala Leu Pro Phe Phe Lys
 20 25 30
 Val Ala Glu Glu Thr Gln Ile Leu Pro Leu His Thr Leu Ser His Glu
 35 40 45

-continued

Pro Ala Val Gly Phe Ala Ala Asp Ala Ala Ala Arg Tyr Ser Ser Thr
 50 55 60

Leu Gly Val Ala Ala Val Thr Tyr Gly Ala Gly Ala Phe Asn Met Val
 65 70 75 80

Asn Ala Val Ala Gly Ala Tyr Ala Glu Lys Ser Pro Val Val Val Ile
 85 90 95

Ser Gly Ala Pro Gly Thr Thr Glu Gly Asn Ala Gly Leu Leu Leu His
 100 105 110

His Gln Gly Arg Thr Leu Asp Thr Gln Phe Gln Val Phe Lys Glu Ile
 115 120 125

Thr Val Ala Gln Ala Arg Leu Asp Asp Pro Ala Lys Ala Pro Ala Glu
 130 135 140

Ile Ala Arg Val Leu Gly Ala Ala Arg Ala Gln Ser Arg Pro Val Tyr
 145 150 155 160

Leu Glu Ile Pro Arg Asn Met Val Asn Ala Glu Val Glu Pro Val Gly
 165 170 175

Asp Asp Pro Ala Trp Pro Val Asp Arg Asp Ala Leu Ala Ala Cys Ala
 180 185 190

Asp Glu Val Leu Ala Ala Met Arg Ser Ala Thr Ser Pro Val Leu Met
 195 200 205

Val Cys Val Glu Val Arg Arg Tyr Gly Leu Glu Ala Lys Val Ala Glu
 210 215 220

Leu Ala Gln Arg Leu Gly Val Pro Val Val Thr Thr Phe Met Gly Arg
 225 230 235 240

Gly Leu Leu Ala Asp Ala Pro Thr Pro Pro Leu Gly Thr Tyr Ile Gly
 245 250 255

Val Ala Gly Asp Ala Glu Ile Thr Arg Leu Val Glu Glu Ser Asp Gly
 260 265 270

Leu Phe Leu Leu Gly Ala Ile Leu Ser Asp Thr Asn Phe Ala Val Ser
 275 280 285

Gln Arg Lys Ile Asp Leu Arg Lys Thr Ile His Ala Phe Asp Arg Ala
 290 295 300

Val Thr Leu Gly Tyr His Thr Tyr Ala Asp Ile Pro Leu Ala Gly Leu
 305 310 315 320

Val Asp Ala Leu Leu Glu Arg Leu Pro Pro Ser Asp Arg Thr Thr Arg
 325 330 335

Gly Lys Glu Pro His Ala Tyr Pro Thr Gly Leu Gln Ala Asp Gly Glu
 340 345 350

Pro Ile Ala Pro Met Asp Ile Ala Arg Ala Val Asn Asp Arg Val Arg
 355 360 365

Ala Gly Gln Glu Pro Leu Leu Ile Ala Ala Asp Met Gly Asp Cys Leu
 370 375 380

Phe Thr Ala Met Asp Met Ile Asp Ala Gly Leu Met Ala Pro Gly Tyr
 385 390 395 400

Tyr Ala Gly Met Gly Phe Gly Val Pro Ala Gly Ile Gly Ala Gln Cys
 405 410 415

Val Ser Gly Gly Lys Arg Ile Leu Thr Val Val Gly Asp Gly Ala Phe
 420 425 430

Gln Met Thr Gly Trp Glu Leu Gly Asn Cys Arg Arg Leu Gly Ile Asp
 435 440 445

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Pro Ile Val Ile Leu Phe Asn Asn Ala Ser Trp Glu Met Leu Arg Thr
 450 455 460

Phe Gln Pro Glu Ser Ala Phe Asn Asp Leu Asp Asp Trp Arg Phe Ala
 465 470 475 480

Asp Met Ala Ala Gly Met Gly Gly Asp Gly Val Arg Val Arg Thr Arg
 485 490 495

Ala Glu Leu Lys Ala Ala Leu Asp Lys Ala Phe Ala Thr Arg Gly Arg
 500 505 510

Phe Gln Leu Ile Glu Ala Met Ile Pro Arg Gly Val Leu Ser Asp Thr
 515 520 525

Leu Ala Arg Phe Val Gln Gly Gln Lys Arg Leu His Ala Ala Pro Arg
 530 535 540

Glu
 545

<210> SEQ ID NO 313
 <211> LENGTH: 348
 <212> TYPE: PRT
 <213> ORGANISM: Rhodococcus sp. ST-10

<400> SEQUENCE: 313

Met Lys Ala Ile Gln Tyr Thr Arg Ile Gly Ala Glu Pro Glu Leu Thr
 1 5 10 15

Glu Ile Pro Lys Pro Glu Pro Gly Pro Gly Glu Val Leu Leu Glu Val
 20 25 30

Thr Ala Ala Gly Val Cys His Ser Asp Asp Phe Ile Met Ser Leu Pro
 35 40 45

Glu Glu Gln Tyr Thr Tyr Gly Leu Pro Leu Thr Leu Gly His Glu Gly
 50 55 60

Ala Gly Lys Val Ala Ala Val Gly Glu Gly Val Glu Gly Leu Asp Ile
 65 70 75 80

Gly Thr Asn Val Val Val Tyr Gly Pro Trp Gly Cys Gly Asn Cys Trp
 85 90 95

His Cys Ser Gln Gly Leu Glu Asn Tyr Cys Ser Arg Ala Gln Glu Leu
 100 105 110

Gly Ile Asn Pro Pro Gly Leu Gly Ala Pro Gly Ala Leu Ala Glu Phe
 115 120 125

Met Ile Val Asp Ser Pro Arg His Leu Val Pro Ile Gly Asp Leu Asp
 130 135 140

Pro Val Lys Thr Val Pro Leu Thr Asp Ala Gly Leu Thr Pro Tyr His
 145 150 155 160

Ala Ile Lys Arg Ser Leu Pro Lys Leu Arg Gly Gly Ser Tyr Ala Val
 165 170 175

Val Ile Gly Thr Gly Gly Leu Gly His Val Ala Ile Gln Leu Leu Arg
 180 185 190

His Leu Ser Ala Ala Thr Val Ile Ala Leu Asp Val Ser Ala Asp Lys
 195 200 205

Leu Glu Leu Ala Thr Lys Val Gly Ala His Glu Val Val Leu Ser Asp
 210 215 220

Lys Asp Ala Ala Glu Asn Val Arg Lys Ile Thr Gly Ser Gln Gly Ala
 225 230 235 240

Ala Leu Val Leu Asp Phe Val Gly Tyr Gln Pro Thr Ile Asp Thr Ala
 245 250 255

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ccgctcgagg aggatatata tatgatttct aaaggcttta gcaccc 46

<210> SEQ ID NO 319
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 319

acgtgatgta atctagagga ggatatatat atgagcaaag aaattaaagg 50

<210> SEQ ID NO 320
<211> LENGTH: 50
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 320

tctttgctca tatatatatc ctctctaga ttacatcacg tgttcagtac 50

<210> SEQ ID NO 321
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 321

cgagctctta ttcggcgcca atggtgcacg gg 32

<210> SEQ ID NO 322
<211> LENGTH: 46
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 322

ccgctcgagg aggatatata tatgatttct aaaggcttta gcaccc 46

<210> SEQ ID NO 323
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 323

cgagctctta ttcggcgcca atggtgcacg gg 32

<210> SEQ ID NO 324
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 324

cacccaagcg atagtttata tagcgt 26

-continued

<210> SEQ ID NO 325
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 325

gaaatgaacg gatattacgt 20

<210> SEQ ID NO 326
<211> LENGTH: 19
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 326

cggaacaggt gattgtggt 19

<210> SEQ ID NO 327
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 327

caccgcccac ttcaagatga agctgt 26

<210> SEQ ID NO 328
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 328

cacccaagcg atagtttata tagcgt 26

<210> SEQ ID NO 329
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 329

gtggctaagt acatgccggt 20

<210> SEQ ID NO 330
<211> LENGTH: 35
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 330

ggaattccat atgacaaaga atatgacgac taaac 35

<210> SEQ ID NO 331
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence

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<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 331

cgggatcctt attatttccc ctgccctgca gt                               32

<210> SEQ ID NO 332
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 332

ggaattccat atgagctatc aaccactttt ac                               32

<210> SEQ ID NO 333
<211> LENGTH: 29
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 333

cgggatcctt acagttgagc aaatgatcc                                   29

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1. A method for converting a polysaccharide to a commodity chemical, comprising:

(a) contacting the polysaccharide, wherein the polysaccharide is optionally derived from biomass, with a polysaccharide degrading or depolymerizing metabolic system, wherein the metabolic system is selected from;

- (i) enzymatic or chemical catalysis, and
- (ii) a microbial system, wherein the microbial system comprises a recombinant microorganism, wherein the recombinant microorganism comprises one or exogenous genes that allow it to grow on the polysaccharide as a sole source of carbon,

thereby converting the polysaccharide to a suitable monosaccharide or oligosaccharide; and

(b) contacting the suitable monosaccharide or oligosaccharide with commodity chemical biosynthesis pathway, wherein the commodity chemical biosynthesis pathway comprises an aldehyde or ketone biosynthesis pathway, thereby converting the polysaccharide to the commodity chemical

2. The method of claim 1, wherein the biomass is selected from marine biomass and vegetable/fruit/plant biomass.

3. The method of claim 2, wherein the marine biomass is selected from kelp, giant kelp, sargasso, seaweed, algae, marine microflora, microalgae, and sea grass.

4. The method of claim 2, wherein the vegetable/fruit/plant biomass comprises plant peel or pomace.

5. The method of claim 2, wherein the vegetable/fruit/plant biomass is selected from citrus, potato, tomato, grape, gooseberry, carrot, mango, sugar-beet, apple, switchgrass, wood, and stover.

6. The method of claim 1, wherein the polysaccharide is selected from alginate, agar, carrageenan, fucoidan, pectin, polygalacturonate, cellulose, hemicellulose, xylan, arabinan, and mannan.

7. The method of claim 1, wherein the suitable monosaccharide or oligosaccharide is selected from 2-keto-3-deoxy D-gluconate (KDG), D-mannitol, guluronate, mannuronate, mannitol, lyxose, glycerol, xylitol, glucose, mannose, galactose, xylose, arabinose, glucuronate, galacturonates, and rhamnose.

8. The method of claim 1, wherein the commodity chemical is selected from methane, methanol, ethane, ethene, ethanol, n-propane, 1-propene, 1-propanol, propanal, acetone, propionate, n-butane, 1-butene, 1-butanol, butanal, butanoate, isobutanol, 2-methylbutanal, 2-methylbutanol, 3-methylbutanal, 3-methylbutanol, 2-butene, 2-butanol, 2-butanone, 2,3-butanediol, 3-hydroxy-2-butanone, 2,3-butanedione, ethylbenzene, ethenylbenzene, 2-phenylethanol, phenylacetaldehyde, 1-phenylbutane, 4-phenyl-1-butene, 4-phenyl-2-butene, 1-phenyl-2-butene, 1-phenyl-2-butanol, 4-phenyl-2-butanol, 1-phenyl-2-butanone, 4-phenyl-2-butanone, 1-phenyl-2,3-butanediol, 1-phenyl-3-hydroxy-2-butanone, 4-phenyl-3-hydroxy-2-butanone, 1-phenyl-2,3-butanedione, n-pentane, ethylphenol, ethenylphenol, 2-(4-hydroxyphenyl)ethanol, 4-hydroxyphenylacetaldehyde, 1-(4-hydroxyphenyl) butane, 4-(4-hydroxyphenyl)-1-butene, 4-(4-hydroxyphenyl)-2-butene, 1-(4-hydroxyphenyl)-1-butene, 1-(4-hydroxyphenyl)-2-butanol, 4-(4-hydroxyphenyl)-2-butanol, 1-(4-hydroxyphenyl)-2-butanone, 4-(4-hydroxyphenyl)-2-butanone, 1-(4-hydroxyphenyl)-2,3-butanediol, 1-(4-hydroxyphenyl)-3-hydroxy-2-butanone, 4-(4-hydroxyphenyl)-3-hydroxy-2-butanone, 1-(4-hydroxyphenyl)-2,3-butanedione, indolylethane, indolylethene, 2-(indole-3-yl)ethanol, n-pentane, 1-pentene, 1-pentanol, pentanal, pentanoate, 2-pentene, 2-pentanol, 3-pentanol, 2-pentanone, 3-pentanone, 4-methylpentanal, 4-methylpentanol, 2,3-pentanediol, 2-hydroxy-3-pentanone, 3-hydroxy-2-pentanone, 2,3-pentanedione, 2-methylpentane, 4-methyl-1-pentene, 4-methyl-2-pentene, 4-methyl-3-

pentene, 4-methyl-2-pentanol, 2-methyl-3-pentanol, 4-methyl-2-pentanone, 2-methyl-3-pentanone, 4-methyl-2,3-pentanediol, 4-methyl-2-hydroxy-3-pentanone, 4-methyl-3-hydroxy-2-pentanone, 4-methyl-2,3-pentanedione, 1-phenylpentane, 1-phenyl-1-pentene, 1-phenyl-2-pentene, 1-phenyl-3-pentene, 1-phenyl-2-pentanol, 1-phenyl-3-pentanol, 1-phenyl-2-pentanone, 1-phenyl-3-pentanone, 1-phenyl-2,3-pentanediol, 1-phenyl-2-hydroxy-3-pentanone, 1-phenyl-3-hydroxy-2-pentanone, 1-phenyl-2,3-pentanedione, 4-methyl-1-phenylpentane, 4-methyl-1-phenyl-1-pentene, 4-methyl-1-phenyl-2-pentene, 4-methyl-1-phenyl-3-pentene, 4-methyl-1-phenyl-3-pentanol, 4-methyl-1-phenyl-2-pentanol, 4-methyl-1-phenyl-3-pentaneone, 4-methyl-1-phenyl-2-pentanone, 4-methyl-1-phenyl-2,3-pentanediol, 4-methyl-1-phenyl-2,3-pentanedione, 4-methyl-1-phenyl-3-hydroxy-2-pentanone, 4-methyl-1-phenyl-2-hydroxy-3-pentanone, 1-(4-hydroxyphenyl) pentane, 1-(4-hydroxyphenyl)-1-pentene, 1-(4-hydroxyphenyl)-2-pentene, 1-(4-hydroxyphenyl)-3-pentene, 1-(4-hydroxyphenyl)-2-pentanol, 1-(4-hydroxyphenyl)-3-pentanol, 1-(4-hydroxyphenyl)-2-pentanone, 1-(4-hydroxyphenyl)-3-pentanone, 1-(4-hydroxyphenyl)-2,3-pentanediol, 1-(4-hydroxyphenyl)-2-hydroxy-3-pentanone, 1-(4-hydroxyphenyl)-3-hydroxy-2-pentanone, 1-(4-hydroxyphenyl)-2,3-pentanedione, 4-methyl-1-(4-hydroxyphenyl) pentane, 4-methyl-1-(4-hydroxyphenyl)-2-pentene, 4-methyl-1-(4-hydroxyphenyl)-3-pentene, 4-methyl-1-(4-hydroxyphenyl)-1-pentene, 4-methyl-1-(4-hydroxyphenyl)-3-pentanol, 4-methyl-1-(4-hydroxyphenyl)-2-pentanol, 4-methyl-1-(4-hydroxyphenyl)-3-pentanone, 4-methyl-1-(4-hydroxyphenyl)-2-pentanone, 4-methyl-1-(4-hydroxyphenyl)-2,3-pentanediol, 4-methyl-1-(4-hydroxyphenyl)-2,3-pentanedione, 4-methyl-1-(4-hydroxyphenyl)-3-hydroxy-2-pentanone, 1-indole-3-pentane, 1-(indole-3)-1-pentene, 1-(indole-3)-2-pentene, 1-(indole-3)-3-pentene, 1-(indole-3)-2-pentanol, 1-(indole-3)-3-pentanol, 1-(indole-3)-2-pentanone, 1-(indole-3)-3-pentanone, 1-(indole-3)-2,3-pentanediol, 1-(indole-3)-2-hydroxy-3-pentanone, 1-(indole-3)-3-hydroxy-2-pentanone, 1-(indole-3)-2,3-pentanedione, 4-methyl-1-(indole-3)pentane, 4-methyl-1-(indole-3)-2-pentene, 4-methyl-1-(indole-3)-3-pentene, 4-methyl-1-(indole-3)-1-pentene, 4-methyl-2-(indole-3)-3-pentanol, 4-methyl-1-(indole-3)-2-pentanol, 4-methyl-1-(indole-3)-3-pentanone, 4-methyl-1-(indole-3)-2-pentanone, 4-methyl-1-(indole-3)-2,3-pentanediol, 4-methyl-1-(indole-3)-2,3-pentanedione, 4-methyl-1-(indole-3)-3-hydroxy-2-pentanone, 4-methyl-1-(indole-3)-2-hydroxy-3-pentanone, n-hexane, 1-hexene, 1-hexanol, hexanal, hexanoate, 2-hexene, 3-hexene, 2-hexanol, 3-hexanol, 2-hexanone, 3-hexanone, 2,3-hexanediol, 2,3-hexanedione, 3,4-hexanediol, 3,4-hexanedione, 2-hydroxy-3-hexanone, 3-hydroxy-2-hexanone, 3-hydroxy-4-hexanone, 4-hydroxy-3-hexanone, 2-methylhexane, 3-methylhexane, 2-methyl-2-hexene, 2-methyl-3-hexene, 5-methyl-1-hexene, 5-methyl-2-hexene, 4-methyl-1-hexene, 4-methyl-2-hexene, 3-methyl-3-hexene, 3-methyl-2-hexene, 3-methyl-1-hexene, 2-methyl-3-hexanol, 5-methyl-2-hexanol, 5-methyl-3-hexanol, 2-methyl-3-hexanone, 5-methyl-2-hexanone, 5-methyl-3-hexanone, 2-methyl-3,4-hexanediol, 2-methyl-3,4-hexanedione, 5-methyl-2,3-hexanediol, 5-methyl-2,3-hexanedione, 4-methyl-2,3-hexanediol, 4-methyl-2,3-hexanedione, 2-methyl-3-hydroxy-4-hexanone, 2-methyl-4-hydroxy-3-hexanone, 5-methyl-2-hydroxy-3-hexanone, 5-methyl-3-hy-

droxy-2-hexanone, 4-methyl-2-hydroxy-3-hexanone, 4-methyl-3-hydroxy-2-hexanone, 2,5-dimethylhexane, 2,5-dimethyl-2-hexene, 2,5-dimethyl-3-hexene, 2,5-dimethyl-3-hexanol, 2,5-dimethyl-3-hexanone, 2,5-dimethyl-3,4-hexanediol, 2,5-dimethyl-3,4-hexanedione, 2,5-dimethyl-3-hydroxy-4-hexanone, 5-methyl-1-phenylhexane, 4-methyl-1-phenylhexane, 5-methyl-1-phenyl-1-hexene, 5-methyl-1-phenyl-2-hexene, 5-methyl-1-phenyl-3-hexene, 4-methyl-1-phenyl-1-hexene, 4-methyl-1-phenyl-2-hexene, 4-methyl-1-phenyl-3-hexene, 5-methyl-1-phenyl-2-hexanol, 5-methyl-1-phenyl-3-hexanol, 4-methyl-1-phenyl-2-hexanol, 4-methyl-1-phenyl-3-hexanol, 5-methyl-1-phenyl-2-hexanone, 5-methyl-1-phenyl-3-hexanone, 4-methyl-1-phenyl-2-hexanone, 4-methyl-1-phenyl-3-hexanone, 5-methyl-1-phenyl-2,3-hexanediol, 4-methyl-1-phenyl-2,3-hexanediol, 5-methyl-1-phenyl-3-hydroxy-2-hexanone, 5-methyl-1-phenyl-2-hydroxy-3-hexanone, 4-methyl-1-phenyl-3-hydroxy-2-hexanone, 4-methyl-1-phenyl-2-hydroxy-3-hexanone, 5-methyl-1-phenyl-2,3-hexanedione, 4-methyl-1-phenyl-2,3-hexanedione, 4-methyl-1-(4-hydroxyphenyl)hexane, 5-methyl-1-(4-hydroxyphenyl)-1-hexene, 5-methyl-1-(4-hydroxyphenyl)-2-hexene, 5-methyl-1-(4-hydroxyphenyl)-3-hexene, 4-methyl-1-(4-hydroxyphenyl)-1-hexene, 4-methyl-1-(4-hydroxyphenyl)-2-hexene, 4-methyl-1-(4-hydroxyphenyl)-3-hexene, 5-methyl-1-(4-hydroxyphenyl)-2-hexanol, 5-methyl-1-(4-hydroxyphenyl)-3-hexanol, 4-methyl-1-(4-hydroxyphenyl)-2-hexanol, 4-methyl-1-(4-hydroxyphenyl)-3-hexanol, 5-methyl-1-(4-hydroxyphenyl)-2-hexanone, 5-methyl-1-(4-hydroxyphenyl)-3-hexanone, 4-methyl-1-(4-hydroxyphenyl)-2-hexanone, 4-methyl-1-(4-hydroxyphenyl)-3-hexanone, 5-methyl-1-(4-hydroxyphenyl)-2,3-hexanediol, 4-methyl-1-(4-hydroxyphenyl)-2,3-hexanediol, 5-methyl-1-(4-hydroxyphenyl)-3-hydroxy-2-hexanone, 5-methyl-1-(4-hydroxyphenyl)-2-hydroxy-3-hexanone, 4-methyl-1-(4-hydroxyphenyl)-3-hydroxy-2-hexanone, 4-methyl-1-(4-hydroxyphenyl)-2-hydroxy-3-hexanone, 5-methyl-1-(4-hydroxyphenyl)-2,3-hexanedione, 4-methyl-1-(4-hydroxyphenyl)-2,3-hexanedione, 4-methyl-1-(indole-3)hexane, 5-methyl-1-(indole-3)-1-hexene, 5-methyl-1-(indole-3)-2-hexene, 5-methyl-1-(indole-3)-3-hexene, 4-methyl-1-(indole-3)-2-hexanol, 5-methyl-1-(indole-3)-3-hexanol, 4-methyl-1-(indole-3)-2-hexanol, 4-methyl-1-(indole-3)-3-hexanol, 5-methyl-1-(indole-3)-2-hexanone, 5-methyl-1-(indole-3)-3-hexanone, 4-methyl-1-(indole-3)-2-hexanone, 4-methyl-1-(indole-3)-3-hexanone, 5-methyl-1-(indole-3)-2,3-hexanediol, 4-methyl-1-(indole-3)-2,3-hexanediol, 5-methyl-1-(indole-3)-3-hydroxy-2-hexanone, 5-methyl-1-(indole-3)-2-hydroxy-3-hexanone, 4-methyl-1-(indole-3)-2-hydroxy-3-hexanone, 5-methyl-1-(indole-3)-2,3-hexanedione, 4-methyl-1-(indole-3)-2,3-hexanedione, n-heptane, 1-heptene, 1-heptanol, heptanal, heptanoate, 2-heptene, 3-heptene, 2-heptanol, 3-heptanol, 4-heptanol, 2-heptanone, 3-heptanone, 4-heptanone, 2,3-heptanediol, 2,3-heptanedione, 3,4-heptanediol, 3,4-heptanedione, 2-hydroxy-3-heptanone, 3-hydroxy-2-heptanone, 3-hydroxy-4-heptanone, 4-hydroxy-3-heptanone, 2-methylheptane, 3-methylheptane, 6-methyl-2-heptene, 6-methyl-3-heptene, 2-methyl-3-heptene, 2-methyl-2-heptene, 5-methyl-2-heptene, 5-methyl-3-heptene, 3-methyl-3-heptene, 2-methyl-3-heptanol, 2-methyl-4-heptanol, 6-methyl-3-heptanol, 5-methyl-3-heptanol, 3-methyl-4-heptanol, 2-methyl-3-heptanone, 2-methyl-4-

heptanone, 6-methyl-3-heptanone, 5-methyl-3-heptanone, 3-methyl-4-heptanone, 2-methyl-3,4-heptanediol, 2-methyl-3,4-heptanedione, 6-methyl-3,4-heptanediol, 6-methyl-3,4-heptanedione, 5-methyl-3,4-heptanediol, 5-methyl-3,4-heptanedione, 2-methyl-3-hydroxy-4-heptanone, 2-methyl-4-hydroxy-3-heptanone, 6-methyl-3-hydroxy-4-heptanone, 6-methyl-4-hydroxy-3-heptanone, 5-methyl-3-hydroxy-4-heptanone, 5-methyl-4-hydroxy-3-heptanone, 2,6-dimethylheptane, 2,5-dimethylheptane, 2,6-dimethyl-2-heptene, 2,6-dimethyl-3-heptene, 2,5-dimethyl-2-heptene, 2,5-dimethyl-3-heptene, 3,6-dimethyl-3-heptene, 2,6-dimethyl-3-heptanol, 2,6-dimethyl-4-heptanol, 2,5-dimethyl-3-heptanol, 2,5-dimethyl-4-heptanol, 2,6-dimethyl-3,4-heptanediol, 2,6-dimethyl-3,4-heptanedione, 2,5-dimethyl-3,4-heptanediol, 2,5-dimethyl-3,4-heptanedione, 2,6-dimethyl-3-hydroxy-4-heptanone, 2,6-dimethyl-4-hydroxy-3-heptanone, 2,5-dimethyl-3-hydroxy-4-heptanone, 2,5-dimethyl-4-hydroxy-3-heptanone, n-octane, 1-octene, 2-octene, 1-octanol, octanal, octanoate, 3-octene, 4-octene, 4-octanol, 4-octanone, 4,5-octanediol, 4,5-octanedione, 4-hydroxy-5-octanone, 2-methyloctane, 2-methyl-3-octene, 2-methyl-4-octene, 7-methyl-3-octene, 3-methyl-3-octene, 3-methyl-4-octene, 6-methyl-3-octene, 2-methyl-4-octanol, 7-methyl-4-octanol, 3-methyl-4-octanol, 6-methyl-4-octanol, 2-methyl-4-octanone, 7-methyl-4-octanone, 3-methyl-4-octanone, 6-methyl-4-octanone, 2-methyl-4,5-octanediol, 2-methyl-4,5-octanedione, 3-methyl-4,5-octanediol, 3-methyl-4,5-octanedione, 2-methyl-4-hydroxy-5-octanone, 2-methyl-5-hydroxy-4-octanone, 3-methyl-4-hydroxy-5-octanone, 3-methyl-5-hydroxy-4-octanone, 2,7-dimethyloctane, 2,7-dimethyl-3-octene, 2,7-dimethyl-4-octene, 2,7-dimethyl-4-octanol, 2,7-dimethyl-4-octanone, 2,7-dimethyl-4,5-octanediol, 2,7-dimethyl-4,5-octanedione, 2,7-dimethyl-4-hydroxy-5-octanone, 2,6-dimethyloctane, 2,6-dimethyl-3-octene, 2,6-dimethyl-4-octene, 3,7-dimethyl-3-octene, 2,6-dimethyl-4-octanol, 3,7-dimethyl-4-octanol, 2,6-dimethyl-4-octanone, 3,7-dimethyl-4-octanone, 2,6-dimethyl-4,5-octanediol, 2,6-dimethyl-4,5-octanedione, 2,6-dimethyl-4-hydroxy-5-octanone, 2,6-dimethyl-5-hydroxy-4-octanone, 3,6-dimethyloctane, 3,6-dimethyl-3-octene, 3,6-dimethyl-4-octene, 3,6-dimethyl-4-octanol, 3,6-dimethyl-4-octanone, 3,6-dimethyl-4,5-octanediol, 3,6-dimethyl-4,5-octanedione, 3,6-dimethyl-4-hydroxy-5-octanone, n-nonane, 1-nonene, 1-nonanol, nonanal, nonanoate, 2-methylnonane, 2-methyl-4-nonene, 2-methyl-5-nonene, 8-methyl-4-nonene, 2-methyl-5-nonanol, 8-methyl-4-nonanol, 2-methyl-5-nonanone, 8-methyl-4-nonanone, 8-methyl-4,5-nonanediol, 8-methyl-4,5-nonanedione, 8-methyl-4-hydroxy-5-nonanone, 8-methyl-5-hydroxy-4-nonanone, 2,8-dimethylnonane, 2,8-dimethyl-3-nonene, 2,8-dimethyl-4-nonene, 2,8-dimethyl-5-nonene, 2,8-dimethyl-4-nonanol, 2,8-dimethyl-5-nonanol, 2,8-dimethyl-4-nonanone, 2,8-dimethyl-5-nonanone, 2,8-dimethyl-4,5-nonanediol, 2,8-dimethyl-4,5-nonanedione, 2,8-dimethyl-4-hydroxy-5-nonanone, 2,8-dimethyl-5-hydroxy-4-nonanone, 2,7-dimethylnonane, 3,8-dimethyl-3-nonene, 3,8-dimethyl-4-nonene, 3,8-dimethyl-5-nonene, 3,8-dimethyl-4-nonanol, 3,8-dimethyl-5-nonanol, 3,8-dimethyl-4-nonanone, 3,8-dimethyl-5-nonanone, 3,8-dimethyl-4,5-nonanediol, 3,8-dimethyl-4,5-nonanedione, 3,8-dimethyl-4-hydroxy-5-nonanone, 3,8-dimethyl-5-hydroxy-4-nonanone, n-decane, 1-decene, 1-decanol, decanoate, 2,9-dimethyldecane, 2,9-dimethyl-3-decene, 2,9-dimethyl-4-decene, 2,9-dimethyl-5-decanol, 2,9-dimethyl-5-decanone, 2,9-dimethyl-5,6-decanediol, 2,9-dimethyl-6-hydroxy-5-de-

canone, 2,9-dimethyl-5,6-decanedione, undecane, 1-undecene, 1-undecanol, undecanal, undecanoate, n-dodecane, 1-dodecene, 1-dodecanol, dodecanal, dodecanoate, n-dodecane, 1-decdecene, 1-dodecanol, ddodecanal, dodecanoate, n-tridecane, 1-tridecene, 1-tridecanol, tridecanal, tridecanoate, n-tetradecane, 1-tetradecene, 1-tetradecanol, tetradecanal, tetradecanoate, n-pentadecane, 1-pentadecene, 1-pentadecanol, pentadecanal, pentadecanoate, n-hexadecane, 1-hexadecene, 1-hexadecanol, hexadecanal, hexadecanoate, n-heptadecane, 1-heptadecene, 1-heptadecanol, heptadecanal, heptadecanoate, n-octadecane, 1-octadecene, 1-octadecanol, octadecanal, octadecanoate, n-nonadecane, 1-nonadecene, 1-nonadecanol, nonadecanal, nonadecanoate, eicosane, 1-eicosene, 1-eicosanol, eicosanal, eicosanoate, 3-hydroxy propanal, 1,3-propanediol, 4-hydroxybutanal, 1,4-butanediol, 3-hydroxy-2-butanone, 2,3-butanediol, 1,5-pentane diol, homocitrate, homoisocitrate, b-hydroxy adipate, glutarate, glutarsemialdehyde, glutaraldehyde, 2-hydroxy-1-cyclopentanone, 1,2-cyclopentanediol, cyclopentanone, cyclopentanol, (S)-2-acetolactate, (R)-2,3-Dihydroxy-isovalerate, 2-oxoisovalerate, isobutyryl-CoA, isobutyrate, isobutyraldehyde, 5-amino pentaldehyde, 1,10-diaminodecane, 1,10-diamino-5-decene, 1,10-diamino-5-hydroxydecane, 1,10-diamino-5-decanone, 1,10-diamino-5,6-decanediol, 1,10-diamino-6-hydroxy-5-decanone, phenylacetaldehyde, 1,4-diphenylbutane, 1,4-diphenyl-1-butene, 1,4-diphenyl-2-butene, 1,4-diphenyl-2-butanol, 1,4-diphenyl-2-butanone, 1,4-diphenyl-2,3-butanediol, 1,4-diphenyl-3-hydroxy-2-butanone, 1-(4-hydroxyphenyl)-4-phenylbutane, 1-(4-hydroxyphenyl)-4-phenyl-1-butene, 1-(4-hydroxyphenyl)-4-phenyl-2-butene, 1-(4-hydroxyphenyl)-4-phenyl-2-butanol, 1-(4-hydroxyphenyl)-4-phenyl-2-butanone, 1-(4-hydroxyphenyl)-4-phenyl-2,3-butanediol, 1-(4-hydroxyphenyl)-4-phenyl-3-hydroxy-2-butanone, 1-(indole-3)-4-phenylbutane, 1-(indole-3)-4-phenyl-1-butene, 1-(indole-3)-4-phenyl-2-butene, 1-(indole-3)-4-phenyl-2-butanol, 1-(indole-3)-4-phenyl-2-butanone, 1-(indole-3)-4-phenyl-2,3-butanediol, 1-(indole-3)-4-phenyl-3-hydroxy-2-butanone, 4-hydroxyphenylacetaldehyde, 1,4-di(4-hydroxyphenyl)butane, 1,4-di(4-hydroxyphenyl)-1-butene, 1,4-di(4-hydroxyphenyl)-2-butene, 1,4-di(4-hydroxyphenyl)-2-butanol, 1,4-di(4-hydroxyphenyl)-2-butanone, 1,4-di(4-hydroxyphenyl)-2,3-butanediol, 1,4-di(4-hydroxyphenyl)-3-hydroxy-2-butanone, 1-(4-hydroxyphenyl)-4-(indole-3)-butane, 1-(4-hydroxyphenyl)-4-(indole-3)-1-butene, 1-di(4-hydroxyphenyl)-4-(indole-3)-2-butene, 1-(4-hydroxyphenyl)-4-(indole-3)-2-butanol, 1-(4-hydroxyphenyl)-4-(indole-3)-2-butanone, 1-(4-hydroxyphenyl)-4-(indole-3)-2,3-butanediol, 1-(4-hydroxyphenyl)-4-(indole-3)-3-hydroxy-2-butanone, indole-3-acetaldehyde, 1,4-di(indole-3)-butane, 1,4-di(indole-3)-1-butene, 1,4-di(indole-3)-2-butene, 1,4-di(indole-3)-2-butanol, 1,4-di(indole-3)-2-butanone, 1,4-di(indole-3)-2,3-butanediol, 1,4-di(indole-3)-3-hydroxy-2-butanone, succinate semialdehyde, hexane-1,8-dicarboxylic acid, 3-hexene-1,8-dicarboxylic acid, 3-hydroxy-hexane-1,8-dicarboxylic acid, 3-hexanone-1,8-dicarboxylic acid, 3,4-hexanediol-1,8-dicarboxylic acid, 4-hydroxy-3-hexanone-1,8-dicarboxylic acid, fucoidan, iodine, chlorophyll, carotenoid, calcium, magnesium, iron, sodium, potassium, and phosphate.

9. A method for converting a polysaccharide to a suitable monosaccharide or oligosaccharide, comprising:

- (a) contacting the polysaccharide, wherein the polysaccharide is optionally obtained from biomass, with a microbial system for a time sufficient to convert the polysaccharide to a suitable monosaccharide or oligosaccharide, wherein the microbial system comprises,
- (i) at least one gene encoding and expressing an enzyme selected from a lyase and a hydrolase, wherein the lyase and/or hydrolase optionally comprises at least one signal peptide or at least one autotransporter domain;
- (ii) at least one gene encoding and expressing an enzyme selected from a monosaccharide transporter, a disaccharide transporter, a trisaccharide transporter, an oligosaccharide transporter, a polysaccharide transporter, and a superchannel; and
- (iii) at least one gene encoding and expressing an enzyme selected from a monosaccharide dehydrogenase, an isomerase, a dehydratase, a kinase, and an aldolase,

thereby converting the polysaccharide to a suitable monosaccharide or oligosaccharide.

10. A method for converting a polysaccharide to a suitable monosaccharide or oligosaccharide, comprising:

- (a) contacting the polysaccharide, wherein the polysaccharide is optionally obtained from biomass, with a chemical or enzymatic catalysis pathway for a time sufficient to convert the polysaccharide to a first monosaccharide or oligosaccharide; and
- (b) contacting the first monosaccharide or oligosaccharide with a microbial system for a time sufficient to convert the first monosaccharide or oligosaccharide to the suitable monosaccharide or oligosaccharide, wherein the microbial system comprises,
- (i) at least one gene encoding and expressing an enzyme selected from a lyase and a hydrolase
- (ii) at least one gene encoding and expressing an enzyme selected from a monosaccharide transporter, a disaccharide transporter, a trisaccharide transporter, an oligosaccharide transporter, a polysaccharide transporter, and a superchannel; and
- (ii) at least one gene encoding and expressing an enzyme selected from a monosaccharide dehydrogenase, an isomerase, a dehydratase, a kinase, and an aldolase,
- thereby converting the polysaccharide to the suitable monosaccharide or oligosaccharide.

11. The method of claim **9**, wherein the lyase is selected from an alginate lyase, a pectate lyase, a polymannuronate lyase, a polygluronate lyase, a polygalacturonate lyase and a rhamnogalacturonate lyase.

12. The method of claim **9**, wherein the hydrolase is selected from an alginate hydrolase, a rhamnogalacturonate hydrolase, a polymannuronate hydrolase, a pectin hydrolase, and a polygalacturonate hydrolase.

13. The method of claim **9**, wherein the transporter is selected from an ABC transporter, a symporter, and an outer membrane porin.

14. The method of claim **13**, wherein the ABC transporter is selected from Atu3021, Atu3022, Atu3023, Atu3024, algM1, algM2, AlgQ1, AlgQ2, AlgS, OG2516_05558, OG2516_05563, OG2516_05568, OG2516_05573, TogM, TogN, TogA, TogB, and functional variants thereof.

15. The method of claim **13**, wherein the symporter is selected from V12B01_24239 (SEQ ID NO:26), V12B01_24194 (SEQ ID NO:8), and TogT, and functional variants thereof.

16. The method of claim **13**, wherein the outer membrane porin comprises a porin selected from V12B01_24269, KdgM, and KdgN, and functional variants thereof.

17. A recombinant microorganism that is capable of growing on a polysaccharide as a sole source of carbon, wherein the polysaccharide is selected from alginate, pectin, tri-galacturonate, di-galacturonate, cellulose, and hemi-cellulose.

18. The recombinant microorganism of claim **17**, wherein the polysaccharide is alginate.

19. The recombinant microorganism of claim **17**, wherein the polysaccharide is pectin.

20. The recombinant microorganism of claim **17**, wherein the polysaccharide is tri-galacturonate.

21. A recombinant microorganism, comprising (i) at least one gene encoding and expressing an enzyme selected from a lyase and a hydrolase, wherein the lyase or hydrolase optionally comprises at least one signal peptide or at least one autotransporter domain; (ii) at least one gene encoding and expressing an enzyme selected from a monosaccharide transporter, a disaccharide transporter, a trisaccharide transporter, an oligosaccharide transporter, a polysaccharide transporter, and a superchannel; and (iii) at least one gene encoding and expressing an enzyme selected from a monosaccharide dehydrogenase, an isomerase, a dehydratase, a kinase, and an aldolase.

22. The recombinant microorganism of claim **21**, wherein the microorganism is capable of growing on a polysaccharide as a sole source of carbon.

23. The recombinant microorganism of claim **22**, wherein the polysaccharide is selected from alginate, pectin, and tri-galacturonate.

24. A method for converting a suitable monosaccharide or oligosaccharide to a first commodity chemical comprising,

- (a) contacting the suitable monosaccharide or oligosaccharide with a microbial system for a time sufficient to convert to the suitable monosaccharide or oligosaccharide to the commodity chemical, wherein the microbial system comprises a recombinant microorganism, wherein the microorganism comprises a commodity chemical biosynthesis pathway,

thereby converting the suitable monosaccharide or oligosaccharide to the first commodity chemical.

25. The method of claim **24**, wherein the commodity chemical pathway comprises one or more genes encoding an aldehyde or ketone biosynthesis pathway.

26. The method of claim **25**, wherein the aldehyde or ketone biosynthesis pathway is selected from one or more of an acetaldehyde, a propionaldehyde, a butyraldehyde, an isobutyraldehyde, a 2-methyl-butyraldehyde, a 3-methyl-butyraldehyde, a 2-phenyl acetaldehyde, a 2-(4-hydroxyphenyl) acetaldehyde, a 2-Indole-3-acetaldehyde, a glutaraldehyde, a 5-amino-pentaldehyde, a succinate semialdehyde, and a succinate 4-hydroxyphenyl acetaldehyde biosynthesis pathway.

27. The method of claim **26**, wherein the aldehyde or ketone biosynthesis pathway comprises an acetaldehyde biosynthesis pathway and a biosynthesis pathway selected from a propionaldehyde, butyraldehyde, isobutyraldehyde, 2-methyl-butyraldehyde, 3-methyl-butyraldehyde, a 2-phe-

nyl acetoaldehyde, a 2-(4-hydroxyphenyl) acetaldehyde, and a 2-Indole-3-acetoaldehyde biosynthesis pathway.

28. The method of claim **26**, wherein the aldehyde or ketone biosynthesis pathway comprises a propionaldehyde biosynthesis pathway and a biosynthesis pathway selected from a butyraldehyde, isobutyraldehyde, 2-methyl-butyraldehyde, 3-methyl-butyraldehyde, and phenylacetoaldehyde biosynthesis pathway.

29. The method of claim **26**, wherein the aldehyde or ketone biosynthesis pathway comprises a butyraldehyde biosynthesis pathway and a biosynthesis pathway selected from an isobutyraldehyde, 2-methyl-butyraldehyde, 3-methyl-butyraldehyde, a 2-phenyl acetoaldehyde, a 2-(4-hydroxyphenyl)acetaldehyde, and a 2-Indole-3-acetoaldehyde biosynthesis pathway.

30. The method of claim **26**, wherein the aldehyde or ketone biosynthesis pathway comprises an isobutyraldehyde biosynthesis pathway and a biosynthesis pathway selected from a 2-methyl-butyraldehyde, 3-methyl-butyraldehyde, a 2-phenyl acetoaldehyde, a 2-(4-hydroxyphenyl)acetaldehyde, and a 2-Indole-3-acetoaldehyde biosynthesis pathway.

31. The method of claim **26**, wherein the aldehyde or ketone biosynthesis pathway comprises a 2-methyl-butyraldehyde biosynthesis pathway and a biosynthesis pathway selected from a 3-methyl-butyraldehyde, a 2-phenyl acetoaldehyde, a 2-(4-hydroxyphenyl)acetaldehyde, and a 2-Indole-3-acetoaldehyde biosynthesis pathway.

32. The method of claim **26**, wherein the aldehyde or ketone biosynthesis pathway comprises a 3-methyl-butyraldehyde biosynthesis pathway and a biosynthesis pathway selected from a 2-phenyl acetoaldehyde, a 2-(4-hydroxyphenyl) acetaldehyde, and a 2-Indole-3-acetoaldehyde biosynthesis pathway.

33. The method of claim **26**, wherein the aldehyde or ketone biosynthesis pathway comprises a 2-phenyl acetoaldehyde biosynthesis pathway and a biosynthesis pathway selected from a 2-(4-hydroxyphenyl)acetaldehyde and a 2-Indole-3-acetoaldehyde biosynthesis pathway.

34. The method of claim **26**, wherein the aldehyde or ketone biosynthesis pathway comprises a 2-(4-hydroxyphenyl)acetaldehyde biosynthesis pathway and a 2-Indole-3-acetoaldehyde biosynthesis pathway.

35. The method of claim **24**, wherein the first commodity chemical is further enzymatically and/or chemically reduced and dehydrated to a second commodity chemical.

36. A method for converting a suitable monosaccharide or oligosaccharide to a commodity chemical comprising,

(a) contacting the suitable monosaccharide or oligosaccharide with a microbial system for a time sufficient to convert to the suitable monosaccharide or oligosaccharide to the commodity chemical, wherein the microbial system comprises;

(i) one or more genes encoding and expressing an aldehyde biosynthesis pathway, wherein the aldehyde biosynthesis pathway comprises one or more genes encoding and expressing a decarboxylase enzyme; and

(ii) one or more genes encoding and expressing an aldehyde reductase,

thereby converting the suitable monosaccharide or oligosaccharide to the commodity chemical.

37. The method of claim **36**, wherein the decarboxylase enzyme is an indole-3-pyruvate decarboxylase (IPDC).

38. The method of claim **37**, wherein the IPDC comprises an amino acid sequence that is at least 80%, 90%, 95%, 98%, or 99% identical to the amino acid sequence set forth in SEQ ID NO: 312

39. The method of claim **36**, wherein the aldehyde reductase enzyme is a phenylacetaldehyde reductase (PAR).

40. The method of claim **39**, wherein the PAR comprises an amino acid sequence that is at least 80%, 90%, 95%, 98%, or 99% identical to the amino acid sequence set forth in SEQ ID NO: 313.

41. The method of claim **36**, wherein the commodity chemical is selected from 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol, and indole-3-ethanol.

42. A recombinant microorganism, comprising (i) one or more genes encoding and expressing an aldehyde biosynthesis pathway, wherein the aldehyde biosynthesis pathway comprises one or more genes encoding and expressing a decarboxylase enzyme; and (ii) one or more genes encoding and expressing an aldehyde reductase.

43. The recombinant microorganism of claim **42**, wherein the aldehyde biosynthesis pathway further comprises one or more genes encoding and expressing an enzyme selected from a CoA-linked aldehyde dehydrogenase, an aldehyde dehydrogenase, and an alcohol dehydrogenase.

44. The recombinant microorganism of claim **42**, wherein the decarboxylase enzyme is an indole-3-pyruvate decarboxylase (IPDC).

45. The recombinant microorganism of claim **42**, wherein the aldehyde reductase enzyme is a phenylacetoaldehyde reductase (PAR).

46. The recombinant microorganism of claim **42**, wherein the microorganism is capable of converting a suitable monosaccharide or oligosaccharide to a commodity chemical.

47. The recombinant microorganism of claim **46**, wherein the commodity chemical is selected from 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol, and indole-3-ethanol.

48. The recombinant microorganism of claim **17**, wherein the microorganism comprises reduced ethanol production capability compared to a wild-type microorganism.

49. The recombinant microorganism of claim **48**, wherein the microorganism comprises a reduction or inhibition in the conversion of acetyl-coA to ethanol.

50. The recombinant microorganism of claim **48**, wherein the recombinant microorganism comprises a reduction of an ethanol dehydrogenase, thereby providing a reduced ethanol production capability.

51. The recombinant microorganism of claim **50**, wherein the ethanol dehydrogenase is an adhE, homolog or variant thereof.

52. The recombinant microorganism of claim **50**, wherein the microorganism comprises a deletion or knockout of an adhE, homolog or variant thereof.

53. The recombinant microorganism of claim **17** or **42**, wherein the recombinant microorganism comprises one or more deletions or knockouts in a gene encoding an enzyme selected from an enzyme that catalyzes the conversion of acetyl-coA to ethanol, an enzyme that catalyzes the conversion of pyruvate to lactate, an enzyme that catalyzes the conversion of fumarate to succinate, an enzyme that catalyzes the conversion of acetyl-coA and phosphate to coA and acetyl phosphate, an enzyme that catalyzes the conversion of acetyl-

coA and formate to coA and pyruvate, and an enzyme that catalyzes the conversion of alpha-keto acid to branched chain amino acids.

54. The microbial system or recombinant microorganism of claim 1, wherein the recombinant microorganism or microbial system comprises a microorganism selected from *Acetobacter aceti*, *Achromobacter*, *Acidiphilium*, *Acinetobacter*, *Actinomadura*, *Actinoplanes*, *Aeropyrum pernix*, *Agrobacterium*, *Alcaligenes*, *Ananas comosus* (M), *Arthrobacter*, *Aspergillus niger*, *Aspergillus oryzae*, *Aspergillus melleus*, *Aspergillus pulverulentus*, *Aspergillus saitoi*, *Aspergillus sojae*, *Aspergillus usamii*, *Bacillus alcalophilus*, *Bacillus amyloliquefaciens*, *Bacillus brevis*, *Bacillus circulans*, *Bacillus clausii*, *Bacillus lentus*, *Bacillus licheniformis*, *Bacillus macerans*, *Bacillus stearothermophilus*, *Bacillus subtilis*, *Bifidobacterium*, *Brevibacillus brevis*, *Burkholderia cepacia*, *Candida cylindracea*, *Candida rugosa*, *Carica papaya* (L), *Cellulosimicrobium*, *Cephalosporium*, *Chaetomium erraticum*, *Chaetomium gracile*, *Clostridium*, *Clostridium butyricum*, *Clostridium acetobutylicum*, *Clostridium thermocellum*, *Corynebacterium* (glutamicum), *Corynebacterium efficiens*, *Escherichia coli*, *Enterococcus*, *Erwina chrysanthemi*, *Gliconobacter*, *Gluconacetobacter*, *Haloarcula*, *Humicola insolens*, *Humicola insolens*, *Kitasatospora setae*, *Klebsiella*, *Klebsiella oxytoca*, *Kluyveromyces*, *Kluyveromyces fragilis*, *Kluyveromyces lactis*, *Kocuria*, *Lactolactis*, *Lactobacillus*, *Lactobacillus fermentum*, *Lactobacillus sake*, *Lactococcus*, *Lactococcus lactis*, *Leuconostoc*, *Methylocystis*, *Methanobolus siciliae*, *Methanogenium organophilum*, *Methanobacterium bryantii*, *Microbacterium imperiale*, *Micrococcus lysodeikticus*, *Microthumatus*, *Mucor javanicus*, *Mycobacterium*, *Myrothecium*, *Nitrobacter*, *Nitrosomonas*, *Nocardia*, *Papaya carica*, *Pediococcus*, *Pediococcus halophilus*, *Penicillium*, *Penicillium camemberti*, *Penicillium citrinum*, *Penicillium emersonii*, *Penicillium roqueforti*, *Penicillium lilactinum*, *Penicillium multicolor*, *Paracoccus pantotrophus*, *Propionibacterium*, *Pseudomonas*, *Pseudomonas fluorescens*, *Pseudomonas denitrificans*, *Pyro-*

coccus, *Pyrococcus furiosus*, *Pyrococcus horikoshii*, *Rhizobium*, *Rhizomucor miehei*, *Rhizomucor pusillus* Lindt, *Rhizopus*, *Rhizopus delemar*, *Rhizopus japonicus*, *Rhizopus niveus*, *Rhizopus oryzae*, *Rhizopus oligosporus*, *Rhodococcus*, *Sccharomyces cerevisiae*, *Sclerotinia libertina*, *Sphingobacterium multivorum*, *Sphingobium*, *Sphingomonas*, *Streptococcus*, *Streptococcus thermophilus* Y-1, *Streptomyces*, *Streptomyces griseus*, *Streptomyces lividans*, *Streptomyces murinus*, *Streptomyces rubiginosus*, *Streptomyces violaceoruber*, *Streptoverticillium mobaraense*, *Tetragenococcus*, *Thermus*, *Thiosphaera pantotropha*, *Trametes*, *Trichoderma*, *Trichoderma longibrachiatum*, *Trichoderma reesei*, *Trichoderma viride*, *Trichosporon penicillatum*, *Vibrio alginolyticus*, *Xanthomonas*, yeast, *Zygosaccharomyces rouxii*, *Zymomonas*, and *Zymomonas mobilis*.

55. A commodity chemical produced by the method of claim 1.

56. A blended commodity chemical comprising the commodity chemical of claim 55 and a refinery-produced petroleum product.

57. The blended commodity chemical of claim 56, wherein the commodity chemical is selected from a C10-C12 hydrocarbon, 2-phenylethanol, 2-(4-hydroxyphenyl)ethanol, and indole-3-ethanol.

58. The blended commodity chemical of claim 57, wherein the C10-C12 hydrocarbon is selected from 2,7-dimethyloctane and 2,9-dimethyldecane.

59. The blended commodity chemical of claim 56, wherein the refinery-produced petroleum product is selected from jet fuel and diesel fuel.

60. A method of producing a commodity chemical enriched refinery-produced petroleum product, comprising

(a) blending the refinery-produced petroleum product with the commodity chemical produced by the method of claim 1,

thereby producing the commodity chemical enriched refinery-produced petroleum product.

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