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(54) **Titre : SYSTEME D'ADMINISTRATION A COMPOSANT UNIQUE POUR ACIDES NUCLEIQUES**
(54) **Title: ONE-COMPONENT DELIVERY SYSTEM FOR NUCLEIC ACIDS**

(57) **Abrégé/Abstract:**
The invention relates to amphiphilic Janus dendrimers which may form nanoparticles. The invention also relates to methods of inducing an adaptive immune response in a subject comprising administering to the subject an effective amount of a composition comprising at least one nucleoside-modified RNA encoding at least one antigen and at least one amphiphilic Janus dendrimer and to methods of delivering an agent to a subject in need thereof, said method comprising the step of delivering to the subject a composition comprising an agent encapsulated by a nanoparticle.

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Abstract:

The invention relates to amphiphilic Janus dendrimers which may form nanoparticles. The invention also relates to methods of inducing an adaptive immune response in a subject comprising administering to the subject an effective amount of a composition comprising at least one nucleoside-modified RNA encoding at least one antigen and at least one amphiphilic Janus dendrimer and to methods of delivering an agent to a subject in need thereof, said method comprising the step of delivering to the subject a composition comprising an agent encapsulated by a nanoparticle.

TITLE OF THE INVENTION

ONE-COMPONENT DELIVERY SYSTEM FOR NUCLEIC ACIDS

CROSS REFERENCE TO RELATED APPLICATIONS

This application claims priority to and the benefit of U.S. Provisional Application No. 63/192,236, filed on May 24, 2021, U.S. Provisional Application No. 63/253,348, filed on October 07, 2021, and U.S. Provisional Application No. 63/316,794, filed on March 04, 2022, the contents of which are incorporated hereby by reference herein in their entireties.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

This invention was made with government support under 1720530, 1807127, 2104554 awarded by the National Science Foundation and TR002776 awarded by the National Institutes of Health. The government has certain rights in the invention.

BACKGROUND OF THE INVENTION

Delivery of exogenously produced nucleic acids into cells and/or their nucleus to modify protein expression by viral and nonviral vectors represents one of the most fundamental concepts of nanomedicine. Both viral and nonviral delivery systems exhibit advantages and disadvantages. Viral vectors have high transfection efficiency (95%) and higher specificity for cell targeting including for unnatural cells. Some drawbacks of viral gene delivery include immunogenicity, cytotoxicity, difficulty of their assembly, inflammatory responses to repeated administration and the potential for insertional mutagenesis. Nonviral delivery is biosafe, exhibits lower toxicity and lower immunogenicity but it is less transfection efficient (1-2%) and the vectors are less stable than viral vectors. Covalent and supramolecular dendrimers complexed on their cationic periphery groups with the nucleic acid have been employed as nonviral vectors for cell transfection of DNA.

Four component lipid nanoparticles (LNPs) containing ionizable lipids, phospholipids, cholesterol for improved mechanical properties and a poly(ethylene glycol) (PEG)-conjugated lipid that provides stability, represents the current leading nonviral vector for the delivery of mRNA. Shortcomings of LNPs production and stability are demonstrated by a microfluidic device required for their assembly and the need to be stored at extremely low temperatures (–70 °C). Their design, synthesis and assembly was inspired by stealth liposomes developed to deliver low molar mass drugs.

Since RNA is less stable than DNA, it must be protected by encapsulation before being

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released in the cell. At acidic pH (pH 3 to 5), LNPs can encapsulate large quantities of mRNA when the pK_a of the ionizable amine is less than 7. At physiological pH (7.4) LNPs have a nearly neutral surface charge and a high positive charge at the endosomal pH. In endosomal membranes the electrostatic interaction between the catatonically charged LNPs and the naturally occurring anionic lipids has been suggested to be responsible for the release of the RNA. One of the major limitations of the four-component vector is the unknown distribution of its four components in the LNP. The segregation of the neutral ionizable lipid as an oil-phase in the core of the LNPs is considered to be responsible for their very low transfection efficiency (1-2%). The second deficiency of the LNP is provided by the PEG-conjugated lipid and is known as the "PEG dilemma". PEG conjugated to LNP increases the circulation time in blood after intravenous injection. However, the same PEG is known to decrease gene expression with up to four-order of magnitude by decreasing intracellular trafficking of cellular uptake and endosomal escape.

Charge-altering releasable transporters (CARTS) have also been demonstrated for the delivery of mRNA. This delivery concept is unrelated to the viral and nonviral LNP based methodologies discussed above. Artificial and synthetic vesicles, such as liposomes and polymersomes have been elaborated both for drug delivery and also as mimics of natural cells. Dendrimersomes (DSs), which are assembled from amphiphilic Janus dendrimers (JDs), have been shown to exhibit excellent mechanical properties and stability including in serum. Amphiphilic JDs with sugars conjugated on their hydrophilic part, denoted Janus glycodendrimers (JGDs), self-assemble into glycodendrimersomes (GDSs), which mimic the glycans of biological membranes and bind sugar binding proteins. Both JDs and JGDs self-assemble into monodisperse DSs and GDSs with unilamellar or multilamellar structures by simple injection rather than by the microfluidic technology and their dimensions can be predicted. Sequence-defined JGDs self-assemble by injection into GDSs. They demonstrated that a lower sugar density in a defined sequence elicited higher bioactivity to sugar-binding proteins.

Thus, there is a need in the art for compositions and methods for the delivery of mRNA. The present invention satisfies this unmet need.

SUMMARY OF THE INVENTION

In one aspect, the present invention relates, in part, to an ionizable amphiphilic Janus dendrimer having the structure of Formula (I):

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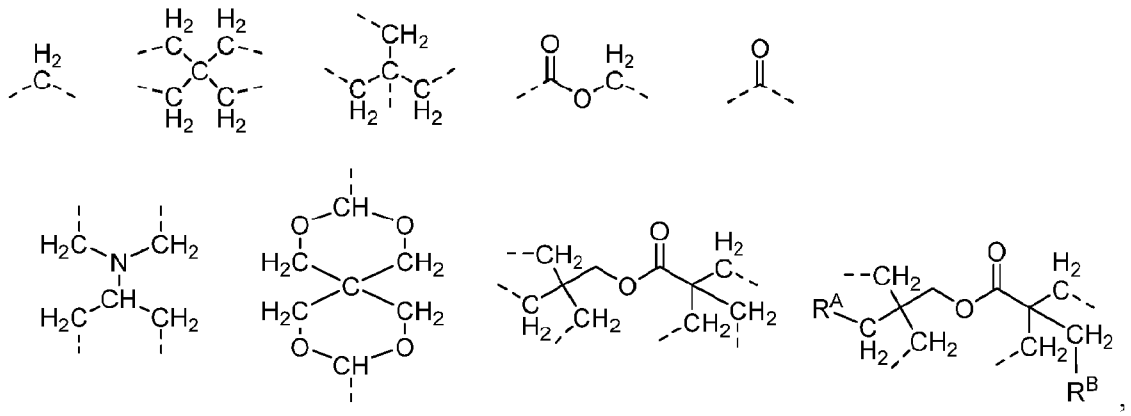
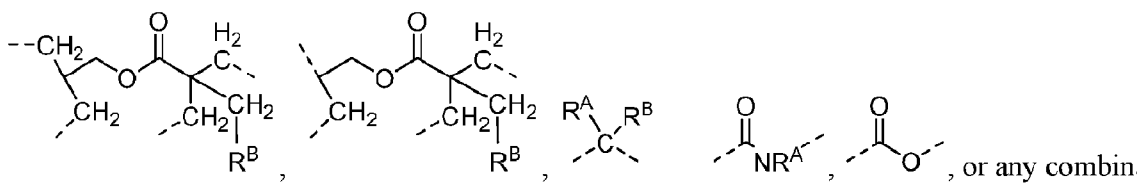
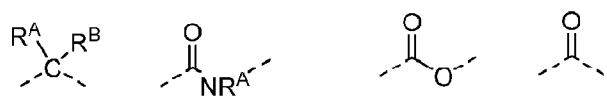
Formula (I).

In some embodiments, A is a polyvalent group comprising at least one selected from





, or any combination thereof. In some embodiments, A is a polyvalent group comprising at least one selected from



, or any combination thereof.

In some embodiments, dashed lines represent a binding site of one of X, Y, or Z.

In some embodiments, X is a hydrophilic group comprising at least one amine.

In some embodiments, Y is a lipophilic group comprising at least one C_1-C_{30}-alkyl chain. In some embodiments, Y is a lipophilic group comprising at least two C_1-C_{30}-alkyl chains having differing numbers of carbon atoms

In some embodiments, Z comprises at least one selected from ethylene glycol, diethylene glycol, triethylene glycol, or polyethylene glycol chain.

In some embodiments, R^A and R^B are independently selected from hydrogen, halide, hydroxy, C_1-C_{30}-alkyl, C_1-C_{30}-alkyl halide, C_1-C_{30}-alkoxy, C_1-C_{30}-alkoxy halide, or any combination thereof.

In some embodiments, s is an integer from 0 to 5. In some embodiments, s is an integer from 1 to 5.

In some embodiments, t is an integer from 0 to 5. In some embodiments, t is an integer from

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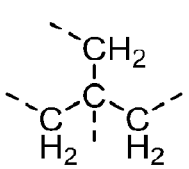
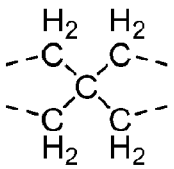
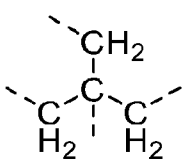
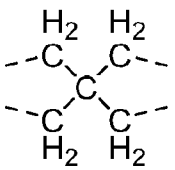
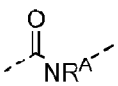
1 to 5.

In some embodiments, u is an integer from 0 to 4.

In some embodiments, the sum of s, t, and u is equal to the valency of A.

For example, in some embodiments, A is represented by  or  and s and t are each 2.

In some embodiments, A is represented by  or  ; t is 2; s is 1; and u is 0 or 1.

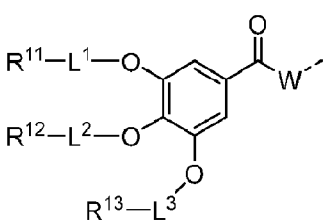
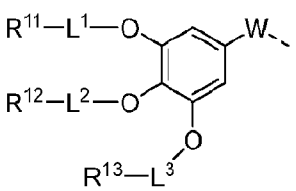
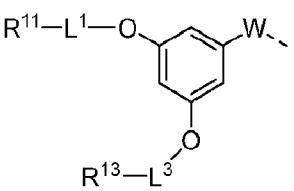
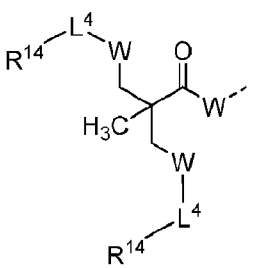
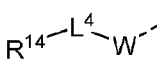
In some embodiments, A is represented by  or and s and t are each 1.

In some embodiments, the ionizable amphiphilic Janus dendrimer having the structure of Formula (I) is an ionizable amphiphilic Janus dendrimer having the structure of Formula (II):

$$(X)_s-A-(Y)_t$$

Formula (II).

In some embodiments, each occurrence of X is independently selected from

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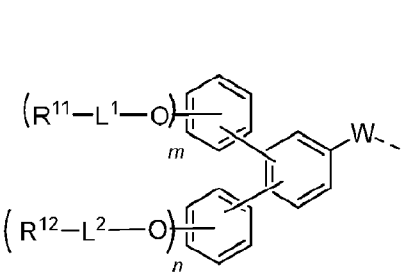
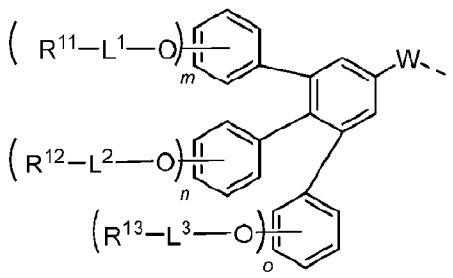


, , or any combination thereof.

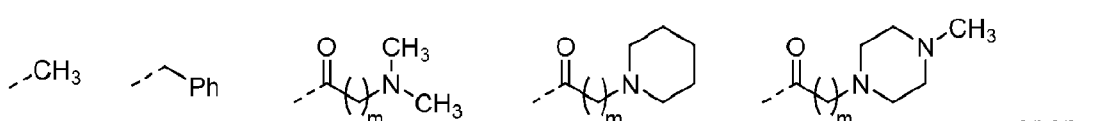
In some embodiments, dashed lines indicate the connection to A.

In some embodiments, each occurrence of m, n, o is independently an integer from 1 to 20. In some embodiments, each occurrence of m, n, o is independently an integer from 1 to 10. In some embodiments, each occurrence of m, n, o is independently an integer from 1 to 5.

In some embodiments, each occurrence of W is independently selected from C=O, $C(R^W)(R^W)$, NR^W, O, or S. In some embodiments, each occurrence of R^W is independently selected from hydrogen, halide, hydroxy, alkyl, alkyl halide, aryl, aryl halide, alkoxy, alkoxy halide, or any combination thereof.

In some embodiments, each occurrence of L^1, L^2, L^3, and L^4 is independently a covalent bond or a divalent linking group selected from alkylene, cycloalkylene, heteroalkylene, heterocycloalkylene, alkenylene, alkynylene, arylene, heteroarylene, silyl, amine, amide, ester, ether, carbonyl, carbamate, thioether, thioester, disulfide, hydrazine, urea, thiourea, phosphate, poly(alkyl ether), heteroatom, or any combinations thereof.

In some embodiments, each occurrence of R^{11}, R^{12}, R^{13}, and R^{14} is independently selected from hydrogen, deuterium, alkyl, aryl, heteroaryl, cycloalkyl, alkoxy, phenoxy, amine, heterocycloalkyl, carbonyl, or any combinations thereof. In some embodiments, at least one occurrence of R^{11}, R^{12}, R^{13}, or R^{14} is $-C(O)(CH_2)_m-N(R^1)(R^2)$. In some embodiments, each occurrence of m is independently an integer from 1 to 10. In some embodiments, each occurrence of R^1 and R^2 is independently selected from hydrogen, deuterium, alkyl, aryl, cycloalkyl, amine, heterocycloalkyl, carbonyl, or any combinations thereof. In some embodiments, R^1 and R^2 may together form a ring. In some embodiments, each occurrence of R^{11}, R^{12}, R^{13}, and R^{14} is independently selected from


, or any combination thereof.

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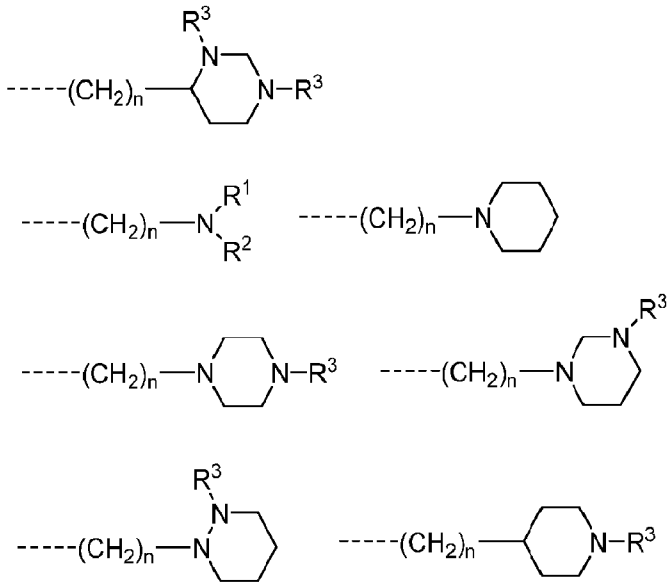
In some embodiments, X comprises at least two tertiary amines.

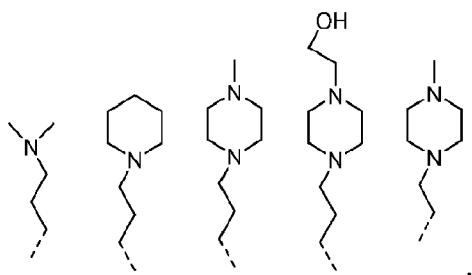
In some embodiments, each occurrence of X is independently selected from



, or any combination thereof.

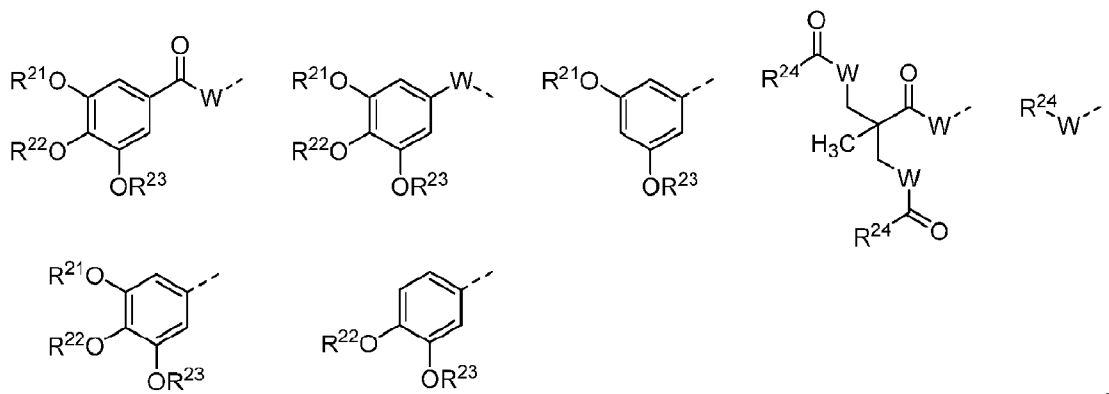
In some embodiments, each occurrence of R^3 is independently selected from hydrogen, $(CH_2)_n$, $(CH_2)_n$-OH, or any combination thereof.

In some embodiments, each occurrence of X is independently selected from



, or any combination thereof.

In some embodiments, each occurrence of Y is independently selected from



, or any

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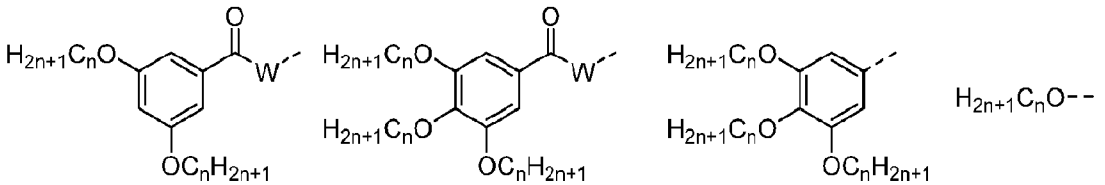
combination thereof.

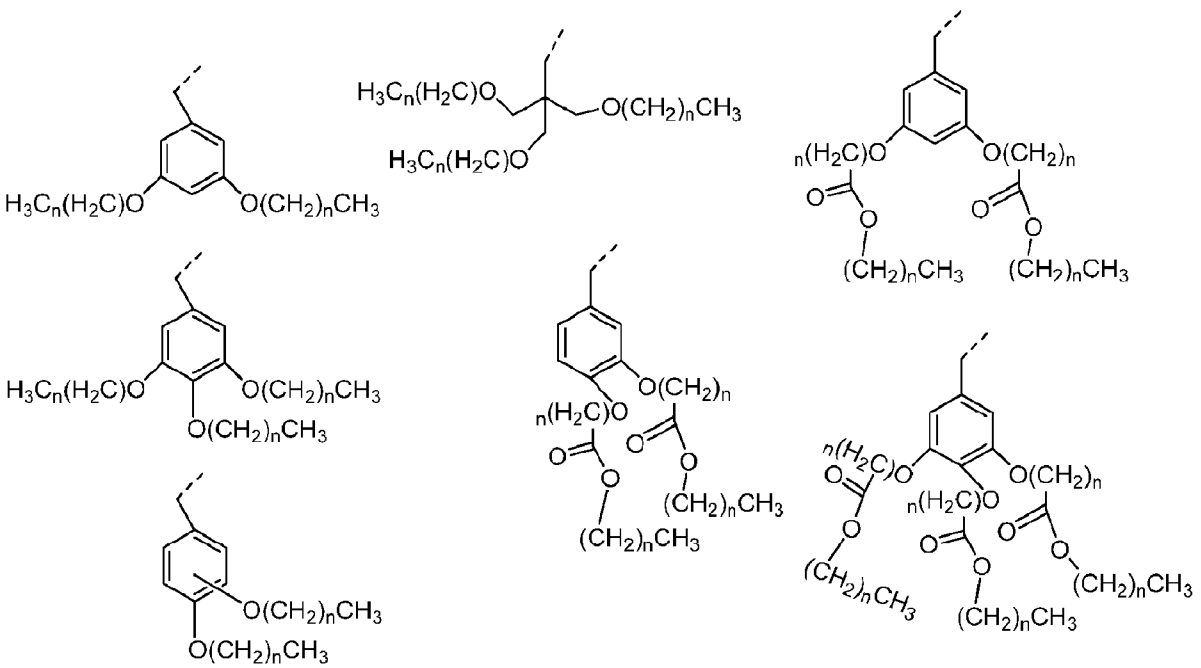
In some embodiments, each occurrence of R^{21}, R^{22}, R^{23}, and R^{24} independently comprises a linear or branched C_1-C_{50}-alkyl. In some embodiments, each occurrence of R^{21}, R^{22}, R^{23}, and R^{24} independently comprises C_1-C_{30}-alkyl.

In some embodiments, each occurrence of Y is independently selected from



, or any combination thereof. In some embodiments, n is an integer from 1 to 30.

In some embodiments, each occurrence of Y is independently selected from



, or any combination thereof.

In some embodiments, each occurrence of $-(CH_2)_nCH_3$, $-O(CH_2)_n-$, and $-O(CH_2)_nCH_3$ is independently linear or branched.

In some embodiments, the ionizable amphiphilic Janus dendrimer comprises a first Y and a second Y. In some embodiments, the first Y comprises an alkyl chain having an even number of carbon atoms and the second Y comprises an alkyl chain having an odd number of carbon atoms. In some embodiments, the ratio between the carbon atoms in the first Y and the carbon atoms in the second Y is greater than or equal to 3 and less than 7.

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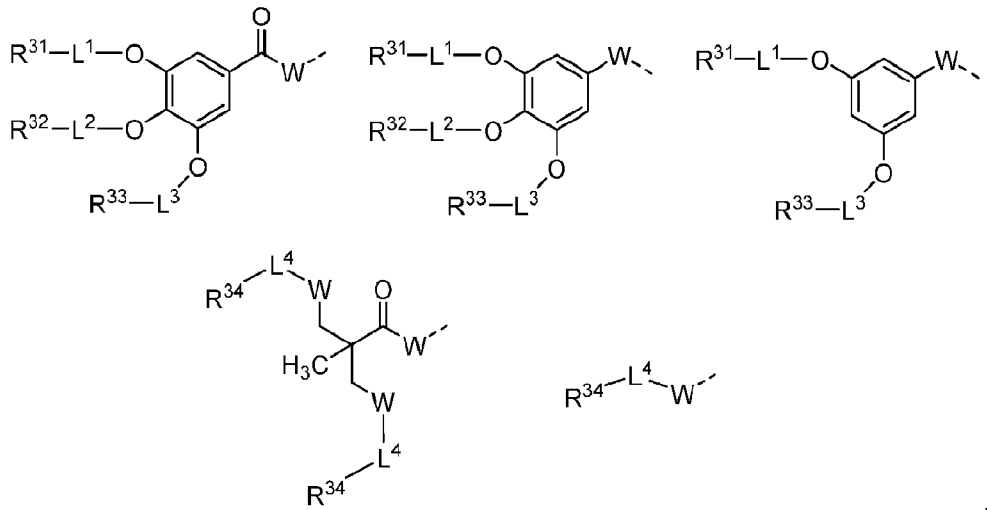
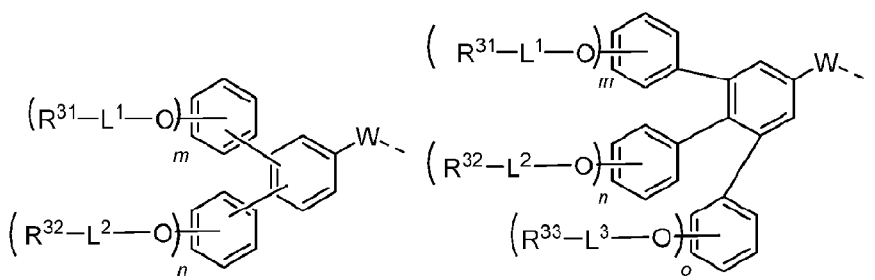
In some embodiments, u is 1 or 2.

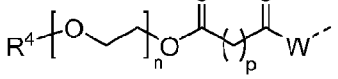
In some embodiments, each occurrence of Z is independently selected from



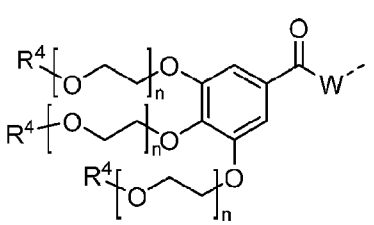


, or any combination thereof.

In some embodiments, each occurrence of R^{31}, R^{32}, R^{33}, and R^{34} is independently selected from hydrogen, deuterium, alkyl, aryl, heteroaryl, cycloalkyl, alkoxy, phenoxy, amine, heterocycloalkyl, carbonyl, or any combinations thereof. In some embodiments, each occurrence of Z is independently selected from



or



. In some embodiments, each occurrence of n is independently an integer from 1 to 100. In some embodiments, each occurrence of p is independently an integer from 1 to 10. In some embodiments, each occurrence of R^4 is independently selected from hydrogen, deuterium, alkyl, aryl, or any combinations thereof.

In some embodiments, the ionizable amphiphilic Janus dendrimer comprises a homochiral, racemic, or achiral branding points.

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In some embodiments, the ionizable amphiphilic Janus dendrimer is a homochiral ionizable amphiphilic Janus dendrimer, racemic ionizable amphiphilic Janus dendrimer, or achiral ionizable amphiphilic Janus dendrimer.

In some embodiments, the ionizable amphiphilic Janus dendrimer is an ionizable amphiphilic Janus dendrimer having a structure selected from the group consisting of at least one structure of Figure 12, at least one structure of Figure 13, at least one structure of Figure 14, at least one structure of Figure 47, at least one structure of Figure 60, at least one structure of Figure 80, and any combination thereof.

In one aspect, the present invention relates, in part, to a nanoparticle comprising at least one ionizable amphiphilic Janus dendrimer of the present invention.

In some embodiments, the nanoparticle comprises a first ionizable amphiphilic Janus dendrimer and a second ionizable amphiphilic Janus dendrimer. In some embodiments, the first ionizable amphiphilic Janus dendrimer has a different structure than the second ionizable amphiphilic Janus dendrimer.

In some embodiments, the nanoparticle comprises a homochiral ionizable amphiphilic Janus dendrimer, achiral ionizable amphiphilic Janus dendrimer, or any combination thereof.

In some embodiments, the nanoparticle is a unilamellar nanoparticle or an onion multilamellar nanoparticle. In some embodiments, the nanoparticle comprises a racemic ionizable amphiphilic Janus dendrimer. In some embodiments, the nanoparticle is a multilamellar nanoparticle.

In some embodiments, the nanoparticle further comprises at least one agent. In some embodiments, the at least one agent comprises a diagnostic agent, detectable agent, therapeutic agent, nucleic acid molecule, or any combination thereof. In some embodiments, the at least one agent is selected from an mRNA, siRNA, microRNA, CRISPR-Cas9, sgRNA, small molecule, protein, antibody, peptide, protein, or any combination thereof.

In some embodiments, the at least one agent comprises a nucleic acid molecule. In some embodiments, the nucleic acid molecule is a DNA molecule or an RNA molecule. In some embodiments, the nucleic acid molecule is selected from cDNA, cRNA, CirRNA, mRNA, miRNA, siRNA, sgRNA, modified RNA, tRNA, antagomir, antisense molecule, targeted nucleic acid, or any combination thereof. In some embodiments, the nucleic acid molecule encodes at least one selected from an antigen, antibody, gene editing molecule, chimeric antigen receptor (CAR), or any combination thereof. In one embodiment, the nucleoside-modified RNA comprises pseudouridine. In one embodiment, the nucleoside-modified RNA comprises pseudouridine plus 5-methyl-

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cytosine. In one embodiment, the nucleoside-modified RNA comprises 5-methyl-uridine. In one embodiment, the nucleoside-modified RNA comprises 1-methyl-pseudouridine.

In one aspect, the invention relates to a composition comprising at least one dendrimersome nanoparticle as described herein.

In one aspect, the present invention relates, in part, a composition comprising at least one ionizable amphiphilic Janus dendrimer of the present invention and/or at least one nanoparticle of the present invention.

In one embodiment, the composition further comprises an adjuvant.

In one embodiment, the composition is a pharmaceutical composition.

In one embodiment, the composition is a vaccine.

In one aspect, the present invention relates, in part, a method of delivering an agent to a subject in need thereof using at least one nanoparticle or a composition of the present invention comprising the same.

In some embodiment, the method further comprises delivering the agent to the liver of the subject.

In some embodiment, the method further comprises delivering the agent to the spleen of the subject.

In some embodiment, the method further comprises delivering the agent to the lungs of the subject.

In some embodiment, the method treats or prevents at least one condition selected from a viral infection, bacterial infection, fungal infection, parasitic infection, cancer, disease or disorder associated with cancer, autoimmune disease or disorder, or any combination thereof.

In some embodiments, the agent is encapsulated within the nanoparticle. In one embodiment, the agent is any agent described herein. For example, in one embodiment, the agent is a composition for protein replacement therapy. In one embodiment, the agent is a composition for gene editing. In one embodiment, the agent is a vaccine.

In one aspect, the present invention relates, in part, a method of preventing or treating a disease or disorder in a subject in need thereof using at least one nanoparticle or a composition of the present invention comprising the same. In some embodiments, the disease or disorder is selected from a viral infection, bacterial infection, fungal infection, parasitic infection, cancer, disease or disorder associated with cancer, autoimmune disease or disorder, or any combination thereof.

In one aspect, the present invention relates, in part, a method of inducing an immune

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response in a subject in need thereof using at least one nanoparticle or a composition of the present invention comprising the same.

In one aspect, the invention relates to a method of delivering an agent to a subject in need thereof, said method comprising administering at least one dendrimersome nanoparticle described herein or a composition comprising the same to a subject.

BRIEF DESCRIPTION OF THE DRAWINGS

The following detailed description of embodiments of the invention will be better understood when read in conjunction with the appended drawings. It should be understood that the invention is not limited to the precise arrangements and instrumentalities of the embodiments shown in the drawings.

Figure 1 depicts a schematic depicting an exemplary one-component nanoparticle (DNP) for mRNA delivery.

Figure 2 depicts a schematic depicting a four-component lipid nanoparticle (LNP) system for mRNA delivery.

Figure 3 depicts a schematic representation of hydrophilic acids, and hydrophobic acids, and linkers employed in the exemplary DNPs.

Figure 4 depicts a schematic representation of the six libraries containing 52 ionizable amphiphilic Janus dendrimers (IAJDs).

Figure 5 depicts representative Luciferase expression in HEK293T cells with DNPs encapsulating Luciferase-mRNA.

Figure 6 depicts representative assay results of *in vivo* transfection results of one-component DNPs. Diameters and polydispersities of DNPs (both in black) and pK_a values of IAJDs (in blue) are shown under the number of the IAJD molecule. All these data are printed on top of each mice image. The luminescence values are also shown.

Figure 7 depicts representative results quantifying the of luciferase signal from *in vivo* images.

Figure 8 depicts representative results comparing concentration and sequence of IAs of IAJDs on activity *in vitro* and *in vivo*. At right are the schematic representations of the IAJDs employed.

Figure 9 depicts representative *in vivo* images of organs.

Figure 10 depicts representative images of mRNA delivery to different organs by one-component DNPs.

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Figure 11 depicts representative results demonstrating examples of excellent stability of DNPs assembled from IAJD9, IAJD22, IAJD33, IAJD34, IAJD32, IAJD33+IAJD32 (2%), IAJD46, and IAJD47.

Figure 12 depicts exemplary compounds from Single-Single IAJD Libraries 1–4. Deep-yellow color means that the molecule shows activity both *in vitro* and *in vivo* except IAJD19, 23 and 45, which only show activity *in vivo* but no activity *in vitro*. Light-yellow color means that the molecule only shows activity *in vitro*. White color means that the molecule shows no activity neither *in vitro* nor *in vivo*.

Figure 13 depicts exemplary compounds from Twin-Twin IAJD Library 5. Deep-yellow color means that the molecule shows activity both *in vitro* and *in vivo*. Light-yellow color means that the molecule only shows activity *in vitro*. White color means that the molecule shows no activity neither *in vitro* nor *in vivo*.

Figure 14 depicts exemplary compounds from Hybrid Twin-Mixed IAJD Library 6. Deep-yellow color means that the molecule shows activity both *in vitro* and *in vivo*. Light-yellow color means that the molecule only shows activity *in vitro*. White color means that the molecule shows no activity neither *in vitro* nor *in vivo*.

Figure 15 depicts representative MALDI-TOF MS spectra of compound 91c ($4/2_{DMBA}{}^{1,3}{}_{Bn}{}^{2}{}_{PEG}{}^{4}$). At right is the zoomed spectrum.

Figure 16 depicts results demonstrating the effect of vortex time on the dimensions of assemblies of exemplary dendrimer IAJD9 (4.0 mg/mL in tris buffer).

Figure 17 depicts representative DLS data of assemblies of IAJD9 (8.00 to 1.00 mg/ml) in tris buffer (pH = 7.4).

Figure 18 depicts representative DLS data of assemblies of IAJD22 (8.0 to 1.0 mg/ml) in tris buffer (pH = 7.4).

Figure 19 depicts representative DLS data of assemblies of IAJD27 (8.0 to 1.0 mg/ml) in tris buffer (pH = 7.4).

Figure 20 depicts representative DLS data of DNPs assembled from IAJD1 and IAJD2.

Figure 21 depicts representative DLS data of DNPs assembled from IAJD8 and IAJD9.

Figure 22 depicts representative DLS data of DNPs assembled from IAJD10 and IAJD17.

Figure 23 depicts representative DLS data of DNPs assembled from IAJD18 and IAJD19.

Figure 24 depicts representative DLS data of DNPs assembled from IAJD20 and IAJD21.

Figure 25 depicts representative DLS data of DNPs assembled from IAJD22 and IAJD23.

Figure 26 depicts representative DLS data of DNPs assembled from IAJD24 and IAJD25.

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Figure 27 depicts representative DLS data of DNPs assembled from IAJD26 and IAJD27.

Figure 28 depicts representative DLS data of DNPs assembled from IAJD28 and IAJD29.

Figure 29 depicts representative DLS data of DNPs assembled from IAJD30 and IAJD31.

Figure 30 depicts representative DLS data of DNPs assembled from IAJD33 and IAJD34.

Figure 31 depicts representative DLS data of DNPs assembled from IAJD37 and IAJD40.

Figure 32 depicts representative DLS data of DNPs assembled from IAJD43 and IAJD44.

Figure 33 depicts representative DLS data of DNPs assembled from IAJD45 and IAJD46.

Figure 34 depicts representative DLS data of DNPs assembled from IAJD47.

Figure 35 depicts representative results showing examples of good stability of DNPs assembled from IAJD27 and not good stability of DNPs assembled from IAJD30 and IAJD31.

Figure 36 depicts representative results showing dimensions of DNPs assembled from IAJD33+IAJD32 (2%) in 1% fetal bovine serum.

Figure 37 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 38 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 39 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 40 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 41 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 42 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 43 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 44 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 45 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 46 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 47 depicts schematic representation of additional compounds of the present

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invention as well as the pKa values determined for these compounds.

Figure 48 depicts representative results demonstrating the distribution of nanoparticles in mice as a function of the IAJD.

Figure 49 depicts representative results demonstrating a comparison of *in vivo* and *in vitro* luminescence for different IAJDs.

Figure 50 depicts representative DLS data of DNPs assembled from IAJD64, IAJD65, IAJD66, IAJD70, IAJD71, IAJD74, IAJD75, IAJD76, and IAJD77.

Figure 51 depicts representative DLS data of DNPs assembled from IAJD78, IAJD79, IAJD81, IAJD82, IAJD83, IAJD84, IAJD85, IAJD86, and IAJD87.

Figure 52 depicts representative DLS data of DNPs assembled from IAJD88, IAJD89, IAJD91, IAJD95, IAJD96, IAJD97, IAJD98, IAJD99, and IAJD103.

Figure 53 depicts representative DLS data of DNPs assembled from IAJD105, IAJD106, IAJD107, and IAJD108.

Figure 54 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 55 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 56 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 57 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 58 depicts representative results demonstrating a comparison of *in vivo* and *in vitro* efficacy.

Figure 59 depicts a schematic representation of a synthesis of nonsymmetric IAJDs.

Figure 60 depicts schematic representations of IAJD 81 through IAJD 159

Figure 61 depicts representative pKa and luminescence of selected IAJDs.

Figure 62 depicts representative results demonstrating the luminescence of selected IAJDs by location in mice.

Figure 63 depicts representative DLS data of DNPs assembled from IAJD113 through IAJD120 and IAJD122.

Figure 64 depicts representative DLS data of DNPs assembled from IAJD124 through IAJD130, IAJD133, IAJD135, and IAJD136.

Figure 65 depicts representative DLS data of DNPs assembled from IAJD138 and IAJD141

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through IAJD148.

Figure 66 depicts representative DLS data of DNPs assembled from IAJD149 through IAJD154, IAJD161, IAJD162, and IAJD171.

Figure 67 depicts representative DLS data of DNPs assembled from IAJD172, IAJD173, IAJD177, IAJD178, IAJD110, IAJD111, and IAJD155 throuhg IAJD159.

Figure 68 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 69 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 70 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 71 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 72 depicts representative titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules.

Figure 73 depicts representative results relating structure and activity of IAJDs.

Figure 74 depicts schematic representations of IAJDs with nonsymmetric alkyl chains and representative results demonstrating selective delivery of Luc-mRNA to spleens and lymph nodes *in vivo* by DNPs and less-selective delivery of Luc-mRNA to organs *in vivo* by DNPs.

Figure 75 depicts representative results demonstrating the ratios between *in vivo* activities of nonsymmetric IAJDs with one C18 alkyl chain and corresponding symmetric IAJDs with two identical C18 alkyl chains.

Figure 76 depicts representative results demonstrating dimensions of DNPs assembled from IAJD125 in 1% fetal bovine serum.

Figure 77 depicts representative results demonstrating dimensions of DNPs assembled from IAJD155 in 1% fetal bovine serum.

Figure 78 depicts representative DLS data of DNP 125 and 178 before and after dialysis in 1X PBS buffer for 3 h.

Figure 79 depicts a schematic representation of glycerol-amphiphillic JDs.

Figure 80 depicts schematic representations of *R*-, *S*-, *rac*- and achiral glycerol with two protected OH and homochiral (1*R*, 1*S*, 2*R*, 2*S*), racemic (1*rac*, 2*rac*), and achiral (3, 4) glycerol-JDs.

Figure 81 depicts a schematic representation of synthesis of Glycerol-JDs from Library 1.

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Reagents and conditions: (*i*) DCC, DPTS, DCM, 23 °C, 12 h; (*ii*) H_2, Pd/C, EtOAc, 23 °C, 8 h; (*iii*) 1 M HCl, MeOH, 23 °C, 1 h.

Figure 82 depicts a schematic representation of synthesis of Glycerol-JDs from Library 2. Reagents and conditions: (*i*) DCC, DPTS, DCM, 23 °C, 12 h; (*ii*) H_2, Pd/C, EtOAc, 23 °C, 8 h; (*iii*) 1 M HCl, 1,4-dioxane, 60 °C, 8 h.

Figure 83 depicts a schematic representation of synthesis of Glycerol-Achiral JDs. Reagents and conditions: (*i*) DCC, DPTS, DCM, 23 °C, 12 h; (*ii*) H_2, Pd/C, EtOAc, 23 °C, 8 h; (*iii*) 1 M HCl, 1,4-dioxane, 60 °C, 8 h.

Figure 84 depicts representative results demonstrating the concentration dependence of diameter (D_h, in nm) and square diameter (D_h^2) of DSs assembled by glycerol-JDs in water. Top two left graphs correspond to glycerol-JDs of library 1. Bottom two right graphs correspond to achiral glycerol-JDs **3** and **4**. The remaining graphs correspond to glycerol-JDs of library 2.

Figure 85 depicts representative cryo-TEM images of DSs assembled by glycerol-JD **2*R***, **2*S***, and **2*rac***.

Figure 86 depicts representative cryo-TEM images of DSs self-assembled by glycerol-based JD **1*R*** (a), **1*S*** (b) and **1*rac*** (c). Scale bar is 100 nm.

Figure 87 depicts representative cryo-TEM images of DSs self-assembled by glycerol-based achiral JD **3**.

Figure 88 depicts representative cryo-TEM images of DSs self-assembled by glycerol-based achiral JD **4**.

Figure 89 depicts representative results demonstrating histograms of normalized frequency of number of vesicle layers of DSs assembled from glycerol-JD **1*R***, **1*S***, **1*rac***, **2*R***, **2*S***, and **2*rac*** by statistical analysis.

Figure 90 depicts representative ^{1}H NMR spectrum of (3,5)-12G1-GC-(*R*)-BMPA-(3,4,5)-3EO-G1 (1*R*). $CDCl_3$, 500 MHz, 298 K. Asterisked signals at δ 7.26 ppm and 2.04 ppm are due to partially nondeuterated residues of $CDCl_3$ and EtOAc respectively.

Figure 91 depicts representative ^{13}C NMR spectrum of (3,5)-12G1-GC-(*R*)-BMPA-(3,4,5)-3EO-G1 (1*R*). $CDCl_3$, 500 MHz, 298 K. The asterisked signal at δ 77.16 ppm is due to $CDCl_3$.

Figure 92 depicts representative ^{1}H NMR spectrum of (3,5)-12G1-GC-(*S*)-BMPA-(3,4,5)-3EO-G1 (1*S*). $CDCl_3$, 500 MHz, 298 K. Asterisked signals at δ 7.26 ppm and 2.04 ppm are due to partially nondeuterated residues of $CDCl_3$ and EA respectively.

Figure 93 depicts representative ^{13}C NMR spectrum of (3,5)-12G1-GC-(*S*)-BMPA-(3,4,5)-3EO-G1 (1*S*). $CDCl_3$, 500 MHz, 298 K. The asterisked signal at δ 77.16 ppm is due to $CDCl_3$.

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Figure 94 depicts representative ^{1}H NMR spectrum of (3,5)-12G1-GC-(*rac*)-BMPA-(3,4,5)-3EO-G1 (1*rac*). $CDCl_3$, 500 MHz, 298 K. Asterisked signals at δ 7.26 ppm, 5.30 ppm and 1.66 ppm are due to partially nondeuterated residues of $CDCl_3$, DCM, and water respectively.

Figure 95 depicts representative ^{13}C NMR spectrum of (3,5)-12G1-GC-(*rac*)-BMPA-(3,4,5)-3EO-G1 (1*rac*). $CDCl_3$, 500 MHz, 298 K. The asterisked signal at δ 77.16 ppm is due to $CDCl_3$.

Figure 96 depicts representative ^{1}H NMR spectrum of (3,5)-12G1-BMPA-GC-(*R*)-(3,4,5)-3EO-G1 (2*R*). $CDCl_3$, 500 MHz, 298 K. Asterisked signals at δ 7.26 ppm and 1.68 ppm are due to partially nondeuterated residues of $CDCl_3$ and water respectively.

Figure 97 depicts representative ^{13}C NMR spectrum of (3,5)-12G1-BMPA-GC-(*R*)-(3,4,5)-3EO-G1 (2*R*). $CDCl_3$, 500 MHz, 298 K. The asterisked signal at δ 77.16 ppm is due to $CDCl_3$.

Figure 98 depicts representative ^{1}H NMR spectrum of (3,5)-12G1-BMPA-GC-(*S*)-(3,4,5)-3EO-G1 (2*S*). $CDCl_3$, 500 MHz, 298 K. Asterisked signals at δ 7.26 ppm and 1.65 ppm are due to partially nondeuterated residues of $CDCl_3$ and water respectively.

Figure 99 depicts representative ^{13}C NMR spectrum of (3,5)-12G1-BMPA-GC-(*S*)-(3,4,5)-3EO-G1 (2*S*). $CDCl_3$, 500 MHz, 298 K. The asterisked signal at δ 77.16 ppm is due to $CDCl_3$.

Figure 100 depicts representative ^{1}H NMR spectrum of (3,5)-12G1-BMPA-GC-(*rac*)-(3,4,5)-3EO-G1 (2*rac*). $CDCl_3$, 500 MHz, 298 K. Asterisked signals at δ 7.26 ppm and 1.60 ppm are due to partially nondeuterated residues of $CDCl_3$ and water respectively.

Figure 101 depicts representative ^{13}C NMR spectrum of (3,5)-12G1-BMPA-GC-(*rac*)-(3,4,5)-3EO-G1 (2*rac*). $CDCl_3$, 500 MHz, 298 K. The asterisked signal at δ 77.16 ppm is due to $CDCl_3$.

Figure 102 depicts representative ^{1}H NMR spectrum of (3,5)-12G1-GC-(achiral)-BMPA-(3,4,5)-3EO-G2 (3). $CDCl_3$, 500 MHz, 298 K. Asterisked signals at δ 7.26 ppm, 2.04 ppm and 1.66 ppm are due to partially nondeuterated residues of $CDCl_3$, EtOAc, and water respectively.

Figure 103 depicts representative ^{13}C NMR spectrum of (3,5)-12G1-GC-(achiral)-BMPA-(3,4,5)-3EO-G2 (3). $CDCl_3$, 500 MHz, 298 K. The asterisked signal at δ 77.16 ppm is due to $CDCl_3$.

Figure 104 depicts representative ^{1}H NMR spectrum of (3,5)-12G2-BMPA-GC-(achiral)-(3,4,5)-3EO-G1 (4). $CDCl_3$, 500 MHz, 298 K. Asterisked signals at δ 7.26 ppm, 5.30 ppm and 1.67 ppm are due to partially nondeuterated residues of $CDCl_3$, DCM, and water respectively.

Figure 105 depicts representative ^{13}C NMR spectrum of (3,5)-12G2-BMPA-GC-(achiral)-(3,4,5)-3EO-G1 (4). $CDCl_3$, 500 MHz, 298 K. The asterisked signal at δ 77.16 ppm is due to

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$CDCl_3$.

Figure 106 depicts representative MALDI-TOF-MS spectra of glycerol-based JDs from library 1.

Figure 107 depicts representative MALDI-TOF-MS spectra of glycerol-based JDs from library 2.

Figure 108 depicts representative MALDI-TOF-MS spectra of glycerol-based achiral JDs **3** (left) and **4** (right).

Figure 109 depicts representative HPLC traces of glycerol-based JDs from library 1.

Figure 110 depicts representative HPLC traces of glycerol-based JDs from library 2.

Figure 111 depicts representative HPLC traces of glycerol-based achiral JDs **3** (left) and **4** (right).

Figure 112 depicts representative results demonstrating normal Q-Q plot of sample **1*rac***. The distribution of **1*rac*** is normal-like.

Figure 113 depicts representative results demonstrating normal Q-Q plot of sample 2rac. The distribution of **2*rac*** is normal-like.

DETAILED DESCRIPTION

The presenet invention is based, in part, on the unexpected results that nanoparticles comprising at least one ionizable amphiphilic Janus dendrimer having the structure of Formula (I) effectively and efficiently delivered an agent to a target of interest. Thus, in one aspect, the present invention relates to an ionizable amphiphilic Janus dendrimer having the structure of Formula (I). In another aspect, the present invention relates to a nanoparticle comprisint at least one ionizable amphiphilic Janus dendrimer of the present invention. In some embodiments, the nanoparticle further comprises at least one agent. In some embodiment, the nanoparticle further comprises at least one agent that is encapsulated by the onizable amphiphilic Janus dendrimer of the present invention. In another aspect, the present invention relates to a composition comprisint at least one ionizable amphiphilic Janus dendrimer of the present invention or a nanoparticle thereof. In some embodiments, the composition is a vaccine.

In one aspect, the present invention relates to methods of delivering an agent to a target of interest using at least one ionizable amphiphilic Janus dendrimer of the present invention or a nanoparticle or a composition thereof. In another aspect, the present invention relates to methods of preventing or treating a disease or disorder in a subject using at least one ionizable amphiphilic Janus dendrimer of the present invention or a nanoparticle or a composition thereof. In another

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aspect, the present invention relates to methods of inducing an adaptive immune response in a subject using at least one ionizable amphiphilic Janus dendrimer of the present invention or a nanoparticle or a composition thereof.

Definitions

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs.

As used herein, each of the following terms has the meaning associated with it in this section.

The articles "a" and "an" are used herein to refer to one or to more than one (i.e., to at least one) of the grammatical object of the article. By way of example, "an element" means one element or more than one element.

"About" as used herein when referring to a measurable value, for example numerical values and/or ranges, such as an amount, a temporal duration, and the like, is meant to encompass variations of ±20%, ±10%, ±5%, ±1%, or ±0.1% from the specified value, as such variations are appropriate to perform the disclosed methods. For example, "about 40 [units]" may mean within ± 25% of 40 (e.g., from 30 to 50), within ± 20%, ± 15%, ± 10%, ± 9%, ± 8%, ± 7%, ± 6%, ± 5%, ± 4%, ± 3%, ± 2%, ± 1%, less than ± 1%, or any other value or range of values therein or therebelow. Furthermore, the phrases "less than about [a value]" or "greater than about [a value]" should be understood in view of the definition of the term "about" provided herein.

The term "compound," as used herein, unless otherwise indicated, refers to any specific chemical compound disclosed herein. In one embodiment, the term also refers to stereoisomers and/or optical isomers (including racemic mixtures) or enantiomerically enriched mixtures of disclosed compounds.

As used herein, the term "analog," "analogue," or "derivative" is meant to refer to a chemical compound or molecule made from a parent compound or molecule by one or more chemical reactions. As such, an analog can be a structure having a structure similar to that of the small molecule therapeutic agents described herein or can be based on a scaffold of a small molecule therapeutic agents described herein, but differing from it in respect to certain components or structural makeup, which may have a similar or opposite action metabolically. An analog or derivative can also be a small molecule that differs in structure from the reference molecule, but retains the essential properties of the reference molecule. An analog or derivative may change its

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interaction with certain other molecules relative to the reference molecule. An analog or derivative molecule may also include a salt, an adduct, tautomer, isomer, prodrug, or other variant of the reference molecule.

As used herein, the term "prodrug" refers to an agent that is converted into the parent drug in vivo. For example, the term "prodrug" refers to a derivative of a known direct acting drug, which derivative has enhanced delivery characteristics and therapeutic value as compared to the drug, and is transformed into the active drug by an enzymatic or chemical process. In some embodiments, "prodrug" refers to an inactive or relatively less active form of an active agent that becomes active by undergoing a chemical conversion through one or more metabolic processes. In one embodiment, upon in vivo administration, a prodrug is chemically converted to the biologically, pharmaceutically, or therapeutically active form of the compound. In another embodiment, a prodrug is enzymatically metabolized by one or more steps or processes to the biologically, pharmaceutically, or therapeutically active form of the compound. For example, the present compounds can be administered to a subject as a prodrug that includes an initiator bound to an active agent, and, by virtue of being degraded by a metabolic process, the active agent is released in its active form.

The term "tautomers" are constitutional isomers of organic compounds that readily interconvert by a chemical process (tautomerization).

The term "isomers" or "stereoisomers" refers to compounds, which have identical chemical constitution, but differ with regard to the arrangement of the atoms or groups in space.

As used herein, the term "alkyl," by itself or as part of another substituent means, unless otherwise stated, a straight or branched chain hydrocarbon having the number of carbon atoms designated (i.e., $C_{1\text{-}50}$ means one to fifty carbon atoms) and including straight, branched chain, or cyclic substituent groups. Examples include methyl, ethyl, propyl, isopropyl, butyl, isobutyl, tert-butyl, pentyl, neopentyl, hexyl, and cyclopropylmethyl.

As used herein, the term "substituted alkyl" means alkyl as defined above, substituted by one, two or three substituents selected from the group consisting of halogen, -OH, alkoxy, $-NH_2$, amino, azido, $-N(CH_3)_2$, -C(=O)OH, trifluoromethyl, -C≡N, $-C(=O)O(C_1\text{-}C_4)$alkyl, $-C(=O)NH_2$, $-SO_2NH_2$, $-C(=NH)NH_2$, and $-NO_2$. Examples of substituted alkyls include, but are not limited to, 2,2-difluoropropyl, 2-carboxycyclopentyl and 3-chloropropyl.

As used herein, the term "heteroalkyl" by itself or in combination with another term means, unless otherwise stated, a stable straight or branched chain alkyl group consisting of the stated number of carbon atoms and one or two heteroatoms selected from the group consisting of O, N,

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and S, and wherein the nitrogen and sulfur atoms may be optionally oxidized and the nitrogen heteroatom may be optionally quaternized. The heteroatom(s) may be placed at any position of the heteroalkyl group, including between the rest of the heteroalkyl group and the fragment to which it is attached, as well as attached to the most distal carbon atom in the heteroalkyl group. Examples include -O-CH_2-CH_2-CH_3, -CH_2-CH_2-CH_2-OH, -CH_2-CH_2-NH-CH_3, -CH_2-S-CH_2-CH_3, and -CH_2CH_2-S(=O)-CH_3. Up to two heteroatoms may be consecutive, such as, for example, -CH_2-NH-OCH_3, or -CH_2-CH_2-S-S-CH_3.

The term "amino" refers to a group of the formula -NRaRa, -NHRa, or -NH2, where each Ra is, independently, an alkyl, alkenyl or alkynyl group as defined above containing 1 to 20 carbon atoms. Unless stated otherwise specifically in the specification, an alkylamino group can be optionally substituted.

The term "hydroxy" or "hydroxyl" refers to a group of the formula OH group.

As used herein, the term "alkoxy" employed alone or in combination with other terms means, unless otherwise stated, an alkyl group having the designated number of carbon atoms, as defined above, connected to the rest of the molecule via an oxygen atom, such as, for example, methoxy, ethoxy, 1-propoxy, 2-propoxy (isopropoxy) and the higher homologs and isomers.

As used herein, the term "halo", "halide", or "halogen" alone or as part of another substituent means, unless otherwise stated, a fluorine, chlorine, bromine, or iodine atom.

"Haloalkyl" or "alkylhalide" refers to an alkyl radical, as defined above, that is substituted by one or more halo radicals, as defined above, e.g., trifluoromethyl, difluoromethyl, trichloromethyl, 2,2,2 trifluoroethyl, 1,2 difluoroethyl, 3 bromo 2 fluoropropyl, 1,2 dibromoethyl, and the like. Unless stated otherwise specifically in the specification, a haloalkyl group can be optionally substituted.

As used herein, the term "cycloalkyl" refers to a mono cyclic or polycyclic non-aromatic radical, wherein each of the atoms forming the ring (i.e. skeletal atoms) is a carbon atom. In one embodiment, the cycloalkyl group is saturated or partially unsaturated. In another embodiment, the cycloalkyl group is fused with an aromatic ring. Cycloalkyl groups include groups having from 3 to 10 ring atoms. Illustrative examples of cycloalkyl groups include, but are not limited to, the following moieties:

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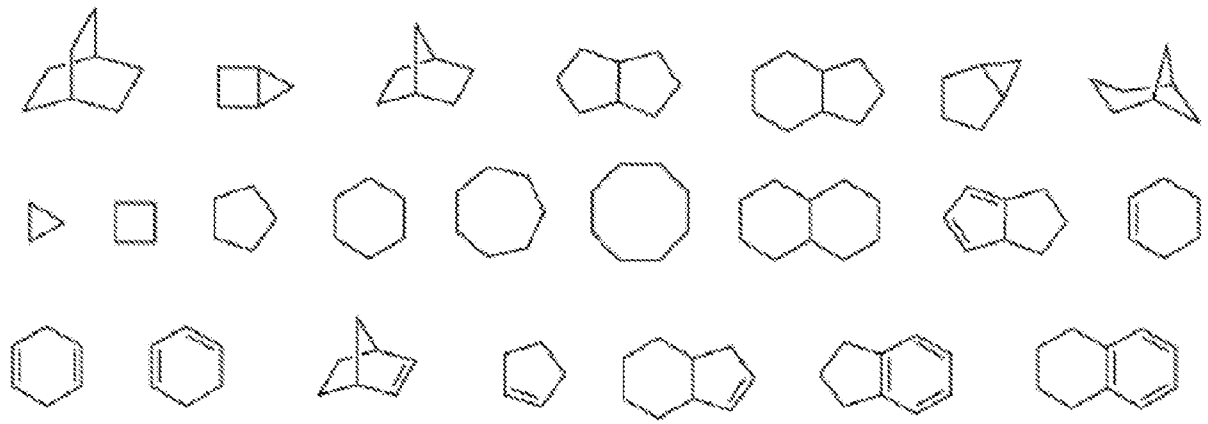

Monocyclic cycloalkyls include, but are not limited to, cyclopropyl, cyclobutyl, cyclopentyl, cyclohexyl, cycloheptyl, and cyclooctyl. Dicyclic cycloalkyls include, but are not limited to, tetrahydronaphthyl, indanyl, and tetrahydropentalene. Polycyclic cycloalkyls include adamantine and norbornane. The term cycloalkyl includes "unsaturated nonaromatic carbocyclyl" or "nonaromatic unsaturated carbocyclyl" groups, both of which refer to a nonaromatic carbocycle as defined herein, which contains at least one carbon double bond or one carbon triple bond.

As used herein, the term "heterocycloalkyl" or "heterocyclyl" refers to a cyclic group containing one to four ring heteroatoms each selected from O, S, and N. In one embodiment, each heterocycloalkyl group has from 4 to 10 atoms in its ring system, with the proviso that the ring of said group does not contain two adjacent O atoms. In another embodiment, the heterocycloalkyl group is fused with an aromatic ring. In one embodiment, the nitrogen and sulfur heteroatoms may be optionally oxidized, and the nitrogen atom may be optionally quaternized. The heterocyclic system may be attached, unless otherwise stated, at any heteroatom or carbon atom that affords a stable structure. A heterocycle may be aromatic or non-aromatic in nature. In one embodiment, the heterocycle is a heteroaryl.

An example of a 3-membered heterocycloalkyl group includes, and is not limited to, aziridine. Examples of 4-membered heterocycloalkyl groups include, and are not limited to, azetidine and a beta lactam. Examples of 5-membered heterocycloalkyl groups include, and are not limited to, pyrrolidine, oxazolidine and thiazolidinedione. Examples of 6-membered heterocycloalkyl groups include, and are not limited to, piperidine, morpholine and piperazine. Other non-limiting examples of heterocycloalkyl groups are:

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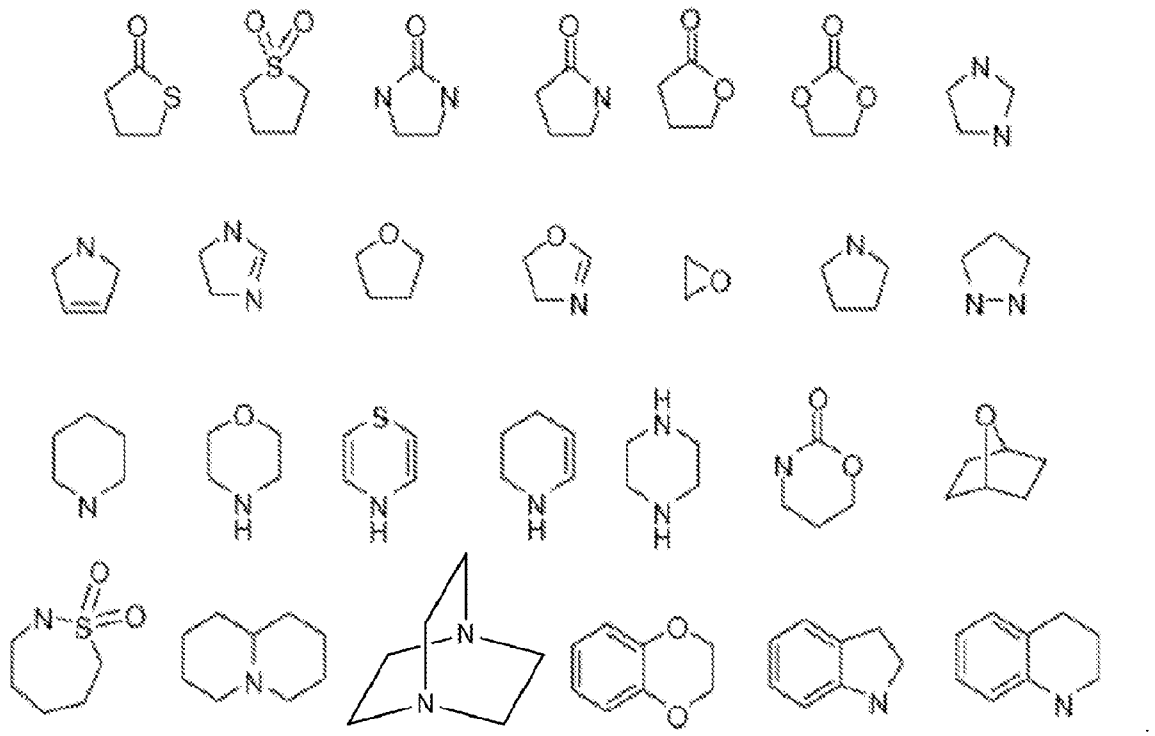

Examples of non-aromatic heterocycles include monocyclic groups such as aziridine, oxirane, thiirane, azetidine, oxetane, thietane, pyrrolidine, pyrroline, pyrazolidine, imidazoline, dioxolane, sulfolane, 2,3-dihydrofuran, 2,5-dihydrofuran, tetrahydrofuran, thiophane, piperidine, 1,2,3,6-tetrahydropyridine, 1,4-dihydropyridine, piperazine, morpholine, thiomorpholine, pyran, 2,3-dihydropyran, tetrahydropyran, 1,4-dioxane, 1,3-dioxane, homopiperazine, homopiperidine, 1,3-dioxepane, 4,7-dihydro-1,3-dioxepin, and hexamethyleneoxide.

As used herein, the term "aromatic" refers to a carbocycle or heterocycle with one or more polyunsaturated rings and having aromatic character, i.e. having $(4n + 2)$ delocalized π (pi) electrons, where n is an integer.

As used herein, the term "aryl," employed alone or in combination with other terms, means, unless otherwise stated, a carbocyclic aromatic system containing one or more rings (typically one, two or three rings), wherein such rings may be attached together in a pendent manner, such as a biphenyl, or may be fused, such as naphthalene. Examples of aryl groups include phenyl, anthracyl, and naphthyl.

"Aralkyl" or "arylalkyl" refers to a radical of the formula $-R_b-R_c$ where R_b is an alkylene group as defined above and R_c is one or more aryl radicals as defined above, for example, benzyl, diphenylmethyl and the like. Unless stated otherwise specifically in the specification, an aralkyl group can be optionally substituted. For example, as used herein, the term "aryl-(C_1-C_3)alkyl"

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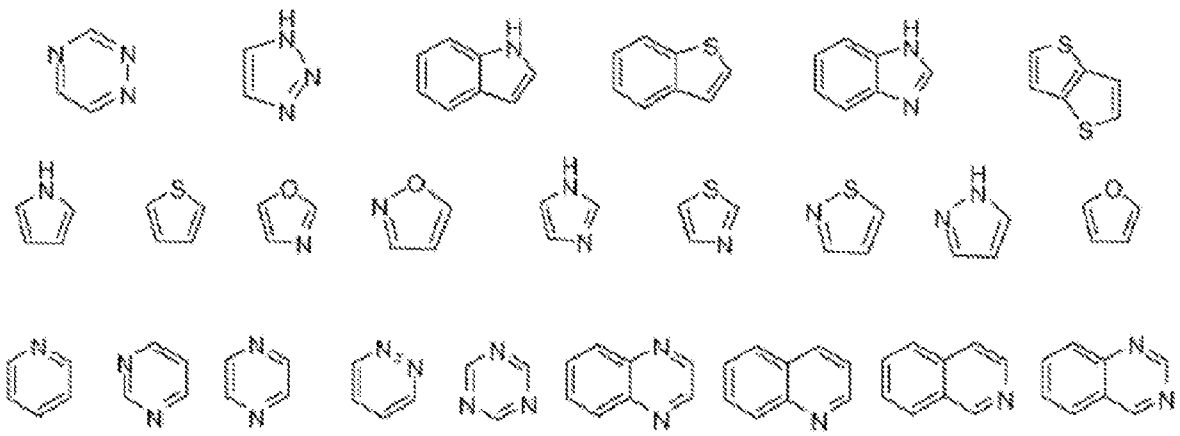
means a functional group wherein a one- to three-carbon alkylene chain is attached to an aryl group, *e.g.*, -CH_2CH_2- phenyl, –CH_2-phenyl (benzyl), aryl-CH_2- and aryl-$CH(CH_3)$-. The term "substituted aryl-(C_1-C_3)alkyl" means an aryl-(C_1-C_3)alkyl functional group in which the aryl group is substituted. Similarly, the term "heteroaryl-(C_1-C_3)alkyl" means a functional group wherein a one to three carbon alkylene chain is attached to a heteroaryl group, *e.g.*, -CH_2CH_2-pyridyl. The term "substituted heteroaryl-(C_1-C_3)alkyl" means a heteroaryl-(C_1-C_3)alkyl functional group in which the heteroaryl group is substituted.

As used herein, the term "heteroaryl" or "heteroaromatic" refers to a heterocycle having aromatic character. A polycyclic heteroaryl may include one or more rings that are partially saturated. Examples include the following moieties:



Examples of heteroaryl groups also include pyridyl, pyrazinyl, pyrimidinyl (particularly 2- and 4-pyrimidinyl), pyridazinyl, thienyl, furyl, pyrrolyl (particularly 2-pyrrolyl), imidazolyl, thiazolyl, oxazolyl, pyrazolyl (particularly 3- and 5-pyrazolyl), isothiazolyl, 1,2,3-triazolyl, 1,2,4-triazolyl, 1,3,4-triazolyl, tetrazolyl, 1,2,3-thiadiazolyl, 1,2,3-oxadiazolyl, 1,3,4-thiadiazolyl and 1,3,4-oxadiazolyl.

Examples of polycyclic heterocycles and heteroaryls include indolyl (particularly 3-, 4-, 5-, 6- and 7-indolyl), indolinyl, quinolyl, tetrahydroquinolyl, isoquinolyl (particularly 1- and 5-isoquinolyl), 1,2,3,4-tetrahydroisoquinolyl, cinnolinyl, quinoxalinyl (particularly 2- and 5-quinoxalinyl), quinazolinyl, phthalazinyl, 1,8-naphthyridinyl, 1,4-benzodioxanyl, coumarin, dihydrocoumarin, 1,5-naphthyridinyl, benzofuryl (particularly 3-, 4-, 5-, 6- and 7-benzofuryl), 2,3-dihydrobenzofuryl, 1,2-benzisoxazolyl, benzothienyl (particularly 3-, 4-, 5-, 6-, and 7-benzothienyl), benzoxazolyl, benzothiazolyl (particularly 2-benzothiazolyl and 5-benzothiazolyl), purinyl, benzimidazolyl (particularly 2-benzimidazolyl), benzotriazolyl, thioxanthinyl, carbazolyl,

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carbolinyl, acridinyl, pyrrolizidinyl, and quinolizidinyl.

As used herein, the term "substituted" means that an atom or group of atoms has replaced hydrogen as the substituent attached to another group. The term "substituted" further refers to any level of substitution, namely mono-, di-, tri-, tetra-, or penta-substitution, where such substitution is permitted. The substituents are independently selected, and substitution may be at any chemically accessible position. In one embodiment, the substituents vary in number between one and four. In another embodiment, the substituents vary in number between one and three. In yet another embodiment, the substituents vary in number between one and two.

As used herein, the term "optionally substituted" means that the referenced group may be substituted or unsubstituted. In one embodiment, the referenced group is optionally substituted with zero substituents, i.e., the referenced group is unsubstituted. In another embodiment, the referenced group is optionally substituted with one or more additional group(s) individually and independently selected from groups described herein.

In one embodiment, the substituents are independently selected from the group consisting of oxo, halogen, -CN, $-NH_2$, -OH, $-NH(CH_3)$, $-N(CH_3)_2$, alkyl (including straight chain, branched and/or unsaturated alkyl), substituted or unsubstituted cycloalkyl, substituted or unsubstituted heterocycloalkyl, fluoro alkyl, substituted or unsubstituted heteroalkyl, substituted or unsubstituted alkoxy, fluoroalkoxy, -S-alkyl, $S(=O)_2$alkyl, $S(=O)_2N$[H, alkyl, or aryl], -C(=O)NH[substituted or unsubstituted alkyl, or substituted or unsubstituted phenyl], -C(=O)N[H or substituted or unsubstituted alkyl or aryl]$_2$, -OC(=O)N[substituted or unsubstituted alkyl]$_2$, -NHC(=O)NH[substituted or unsubstituted alkyl, or substituted or unsubstituted phenyl], -NHC(=O)alkyl, -N[substituted or unsubstituted alkyl]C(=O)[substituted or unsubstituted alkyl], -NHC(=O)[substituted or unsubstituted alkyl], -C(OH)[substituted or unsubstituted alkyl]$_2$, and -$C(NH_2)$[substituted or unsubstituted alkyl]$_2$. In another embodiment, by way of example, an optional substituent is selected from oxo, fluorine, chlorine, bromine, iodine, -CN, $-NH_2$, -OH, $-NH(CH_3)$, $-N(CH_3)_2$, $-CH_3$, $-CH_2CH_3$, $-CH(CH_3)_2$, $-CF_3$, $-CH_2CF_3$, $-OCH_3$, $-OCH_2CH_3$, $-OCH(CH_3)_2$, $-OCF_3$, - OCH_2CF_3, $-S(=O)_2-CH_3$, $-C(=O)NH_2$, -C(=O)-$NHCH_3$, $-NHC(=O)NHCH_3$, -$C(=O)CH_3$, $-ON(O)_2$, and -C(=O)OH. In yet one embodiment, the substituents are independently selected from the group consisting of C_{1-6} alkyl, -OH, C_{1-6} alkoxy, halo, amino, acetamido, oxo and nitro. As used herein, where a substituent is an alkyl or alkoxy group, the carbon chain may be branched, straight or cyclic.

Several references to integers and R, R^1, R^2, R^3, R^4, R^5, R^6, etc. are made in chemical structures and moieties disclosed and described herein. Any description of integers and R, R^1, R^2,

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R^3, R^4, R^5, R^6, etc. in the specification is applicable to any structure or moiety reciting integers and R, R^1, R^2, R^3, R^4, R^5, R^6, etc. respectively.

The term "nanoparticle" refers to a particle having at least one dimension on the order of nanometers (e.g., 1-1,000 nm), which includes one or more amphiphilic Janus dendrimer of Formula (I). In some embodiments, nanoparticles are included in a formulation comprising a nucleoside-modified RNA as described herein. In some embodiments, such nanoparticles an ionizable hydrophilic group and a lipophilic (hydrophobic) group. In one embodiment, the nanoparticles further comprise one or more excipient selected from neutral lipids, charged lipids, steroids and polymer conjugated lipids. In one embodiment, the nanoparticles do not comprise additional excipients. In one embodiment, the nanoparticles do not comprise any of additional lipids, additional cationic polymers, steroids, neutral lipids, charged lipids, or polymer conjugated lipids, besides the at least one compound of Formula (I). In some embodiments, the nucleoside-modified RNA is encapsulated in the lipid portion of the nanoparticle or an aqueous space enveloped by some or all of the lipid portion of the nanoparticle, thereby protecting it from enzymatic degradation or other undesirable effects induced by the mechanisms of the host organism or cells, e.g., an adverse immune response.

The term "antibody," as used herein, refers to an immunoglobulin molecule, which specifically binds with an antigen. Antibodies can be intact immunoglobulins derived from natural sources or from recombinant sources and can be immunoreactive portions of intact immunoglobulins. Antibodies are typically tetramers of immunoglobulin molecules. The antibodies in the invention may exist in a variety of forms including, for example, polyclonal antibodies, monoclonal antibodies, Fv, Fab and $F(ab)_2$, as well as single chain antibodies and humanized antibodies (Harlow et al., 1999, In: Using Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory Press, NY; Harlow et al., 1989, In: Antibodies: A Laboratory Manual, Cold Spring Harbor, New York; Houston et al., 1988, Proc. Natl. Acad. Sci. USA 85:5879-5883; Bird et al., 1988, Science 242:423-426).

The term "antibody fragment" refers to a portion of an intact antibody and refers to the antigenic determining variable regions of an intact antibody. Examples of antibody fragments include, but are not limited to, Fab, Fab', F(ab')2, and Fv fragments, linear antibodies, scFv antibodies, and multispecific antibodies formed from antibody fragments.

An "antibody heavy chain," as used herein, refers to the larger of the two types of polypeptide chains present in all antibody molecules in their naturally occurring conformations.

An "antibody light chain," as used herein, refers to the smaller of the two types of

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polypeptide chains present in all antibody molecules in their naturally occurring conformations. κ and λ light chains refer to the two major antibody light chain isotypes.

By the term "synthetic antibody" as used herein, is meant an antibody, which is generated using recombinant DNA technology, such as, for example, an antibody expressed by a bacteriophage. The term should also be construed to mean an antibody which has been generated by the synthesis of a DNA molecule encoding the antibody and which DNA molecule expresses an antibody protein, or an amino acid sequence specifying the antibody, wherein the DNA or amino acid sequence has been obtained using synthetic DNA or amino acid sequence technology which is available and well known in the art. The term should also be construed to mean an antibody, which has been generated by the synthesis of an RNA molecule encoding the antibody. The RNA molecule expresses an antibody protein, or an amino acid sequence specifying the antibody, wherein the RNA has been obtained by transcribing DNA (synthetic or cloned) or other technology, which is available and well known in the art.

The term "antigen" or "Ag" as used herein is defined as a molecule that provokes an adaptive immune response. This immune response may involve either antibody production, or the activation of specific immunogenically-competent cells, or both. The skilled artisan will understand that any macromolecule, including virtually all proteins or peptides, can serve as an antigen. Furthermore, antigens can be derived from recombinant or genomic DNA or RNA. A skilled artisan will understand that any DNA or RNA, which comprises a nucleotide sequence or a partial nucleotide sequence encoding a protein that elicits an adaptive immune response therefore encodes an "antigen" as that term is used herein. Furthermore, one skilled in the art will understand that an antigen need not be encoded solely by a full length nucleotide sequence of a gene. It is readily apparent that the invention includes, but is not limited to, the use of partial nucleotide sequences of more than one gene and that these nucleotide sequences are arranged in various combinations to elicit the desired immune response. Moreover, a skilled artisan will understand that an antigen need not be encoded by a "gene" at all. It is readily apparent that an antigen can be generated synthesized or can be derived from a biological sample. Such a biological sample can include, but is not limited to a tissue sample, a tumor sample, a cell or a biological fluid.

The term "adjuvant" as used herein is defined as any molecule to enhance an antigen-specific adaptive immune response.

"Encoding" refers to the inherent property of specific sequences of nucleotides in a polynucleotide, such as a gene, a cDNA, or an mRNA, to serve as templates for synthesis of other

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polymers and macromolecules in biological processes having either a defined sequence of nucleotides (i.e., rRNA, tRNA and mRNA) or a defined sequence of amino acids and the biological properties resulting therefrom. Thus, a gene encodes a protein if transcription and translation of mRNA corresponding to that gene produces the protein in a cell or other biological system. Both the coding strand, the nucleotide sequence of which is identical to the mRNA sequence and is usually provided in sequence listings, and the non-coding strand, used as the template for transcription of a gene or cDNA, can be referred to as encoding the protein or other product of that gene or cDNA.

A "vector" is a composition of matter which comprises an isolated nucleic acid and which can be used to deliver the isolated nucleic acid to the interior of a cell. Numerous vectors are known in the art including, but not limited to, linear polynucleotides, polynucleotides associated with ionic or amphiphilic compounds, plasmids, and viruses. Thus, the term "vector" includes an autonomously replicating plasmid or a virus. The term should also be construed to include non-plasmid and non-viral compounds which facilitate transfer of nucleic acid into cells, such as, for example, polylysine compounds, liposomes, and the like. Examples of viral vectors include, but are not limited to, adenoviral vectors, adeno-associated virus vectors, retroviral vectors, and the like.

"Expression vector" refers to a vector comprising a recombinant polynucleotide comprising expression control sequences operatively linked to a nucleotide sequence to be expressed. An expression vector comprises sufficient cis-acting elements for expression; other elements for expression can be supplied by the host cell or in an in vitro expression system. Expression vectors include all those known in the art, such as cosmids, plasmids (e.g., naked or contained in liposomes) RNA, and viruses (e.g., lentiviruses, retroviruses, adenoviruses, and adeno-associated viruses) that incorporate the recombinant polynucleotide.

"Homologous" as used herein, refers to the sequence similarity or sequence identity between two polypeptides or between two nucleic acid molecules. When a position in both of the two compared sequences is occupied by the same base or amino acid monomer subunit, e.g., if a position in each of two DNA molecules is occupied by adenine, then the molecules are homologous at that position. The percent of homology between two sequences is a function of the number of matching or homologous positions shared by the two sequences divided by the number of positions compared X 100. For example, if 6 of 10 of the positions in two sequences are matched or homologous then the two sequences are 60% homologous. By way of example, the DNA sequences ATTGCC and TATGGC share 50% homology. Generally, a comparison is made when two sequences are aligned to give maximum homology.

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"Immunogen" refers to any substance introduced into the body in order to generate an immune response. That substance can a physical molecule, such as a protein, or can be encoded by a vector, such as DNA, mRNA, or a virus.

"Isolated" means altered or removed from the natural state. For example, a nucleic acid or a peptide naturally present in a living animal is not "isolated," but the same nucleic acid or peptide partially or completely separated from the coexisting materials of its natural state is "isolated." An isolated nucleic acid or protein can exist in substantially purified form, or can exist in a non-native environment such as, for example, a host cell.

In the context of the invention, the following abbreviations for the commonly occurring nucleosides (nucleobase bound to ribose or deoxyribose sugar via N-glycosidic linkage) are used. "A" refers to adenosine, "C" refers to cytidine, "G" refers to guanosine, "T" refers to thymidine, and "U" refers to uridine.

By the term "modulating," as used herein, is meant mediating a detectable increase or decrease in the level of a response in a subject compared with the level of a response in the subject in the absence of a treatment or compound, and/or compared with the level of a response in an otherwise identical but untreated subject. The term encompasses perturbing and/or affecting a native signal or response thereby mediating a beneficial therapeutic response in a subject, preferably, a human.

Unless otherwise specified, a "nucleotide sequence encoding an amino acid sequence" includes all nucleotide sequences that are degenerate versions of each other and that encode the same amino acid sequence. Nucleotide sequences that encode proteins and RNA may include introns. In addition, the nucleotide sequence may contain modified nucleosides that are capable of being translation by translational machinery in a cell. For example, an mRNA where all of the uridines have been replaced with pseudouridine, 1-methyl psuedouridine, or another modified nucleoside.

The term "operably linked" refers to functional linkage between a regulatory sequence and a heterologous nucleic acid sequence resulting in expression of the latter. For example, a first nucleic acid sequence is operably linked with a second nucleic acid sequence when the first nucleic acid sequence is placed in a functional relationship with the second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Generally, operably linked DNA or RNA sequences are contiguous and, where necessary to join two protein coding regions, in the same reading frame.

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The term "polynucleotide" as used herein is defined as a chain of nucleotides. Furthermore, nucleic acids are polymers of nucleotides. Thus, nucleic acids and polynucleotides as used herein are interchangeable. One skilled in the art has the general knowledge that nucleic acids are polynucleotides, which can be hydrolyzed into the monomeric "nucleotides." The monomeric nucleotides can be hydrolyzed into nucleosides. As used herein polynucleotides include, but are not limited to, all nucleic acid sequences which are obtained by any means available in the art, including, without limitation, recombinant means, i.e., the cloning of nucleic acid sequences from a recombinant library or a cell genome, using ordinary cloning technology and PCR™, and the like, and by synthetic means.

In certain instances, the polynucleotide or nucleic acid of the invention is a "nucleoside-modified nucleic acid," which refers to a nucleic acid comprising at least one modified nucleoside. A "modified nucleoside" refers to a nucleoside with a modification. For example, over one hundred different nucleoside modifications have been identified in RNA (Rozenski, et al., 1999, The RNA Modification Database: 1999 update. Nucl Acids Res 27: 196-197).

In certain embodiments, "pseudouridine" refers, in another embodiment, to $m^1acp^3\Psi$ (1-methyl-3-(3-amino-3-carboxypropyl) pseudouridine. In another embodiment, the term refers to $m^1\Psi$ (1-methylpseudouridine). In another embodiment, the term refers to Ψm (2'-O-methylpseudouridine). In another embodiment, the term refers to m^5D (5-methyldihydrouridine). In another embodiment, the term refers to $m^3\Psi$ (3-methylpseudouridine). In another embodiment, the term refers to a pseudouridine moiety that is not further modified. In another embodiment, the term refers to a monophosphate, diphosphate, or triphosphate of any of the above pseudouridines. In another embodiment, the term refers to any other pseudouridine known in the art. Each possibility represents a separate embodiment of the invention.

Unless otherwise specified, a "nucleotide sequence encoding an amino acid sequence" includes all nucleotide sequences that are degenerate versions of each other and that encode the same amino acid sequence. The phrase nucleotide sequence that encodes a protein or an RNA may also include introns to the extent that the nucleotide sequence encoding the protein may in some version contain an intron(s).

As used herein, the terms "amino acid", "amino acidic monomer", or "amino acid residue" refer to any of the twenty naturally occurring amino acids including synthetic amino acids with unnatural side chains and including both D and L optical isomers.

As used herein, the terms "peptide," "polypeptide," and "protein" are used interchangeably, and refer to a compound comprised of amino acid residues covalently linked by peptide bonds. A

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protein or peptide must contain at least two amino acids, and no limitation is placed on the maximum number of amino acids that can comprise a protein's or peptide's sequence. Polypeptides include any peptide or protein comprising two or more amino acids joined to each other by peptide bonds. As used herein, the term refers to both short chains, which also commonly are referred to in the art as peptides, oligopeptides and oligomers, for example, and to longer chains, which generally are referred to in the art as proteins, of which there are many types. "Polypeptides" include, for example, biologically active fragments, substantially homologous polypeptides, oligopeptides, homodimers, heterodimers, variants of polypeptides, modified polypeptides, derivatives, analogs, fusion proteins, among others. The polypeptides include natural peptides, recombinant peptides, synthetic peptides, or a combination thereof.

The term "promoter" as used herein is defined as a DNA sequence recognized by the synthetic machinery of the cell, or introduced synthetic machinery, required to initiate the specific transcription of a polynucleotide sequence. For example, the promoter that is recognized by bacteriophage RNA polymerase and is used to generate the mRNA by in vitro transcription.

By the term "specifically binds," as used herein with respect to an antibody, is meant an antibody which recognizes a specific antigen, but does not substantially recognize or bind other molecules in a sample. For example, an antibody that specifically binds to an antigen from one species may also bind to that antigen from one or more other species. But, such cross-species reactivity does not itself alter the classification of an antibody as specific. In another example, an antibody that specifically binds to an antigen may also bind to different allelic forms of the antigen. However, such cross reactivity does not itself alter the classification of an antibody as specific. In some instances, the terms "specific binding" or "specifically binding," can be used in reference to the interaction of an antibody, a protein, or a peptide with a second chemical species, to mean that the interaction is dependent upon the presence of a particular structure (e.g., an antigenic determinant or epitope) on the chemical species; for example, an antibody recognizes and binds to a specific protein structure rather than to proteins generally. If an antibody is specific for epitope "A", the presence of a molecule containing epitope A (or free, unlabeled A), in a reaction containing labeled "A" and the antibody, will reduce the amount of labeled A bound to the antibody.

The term "transfected" or "transformed" or "transduced" as used herein refers to a process by which exogenous nucleic acid is transferred or introduced into the host cell. A "transfected" or "transformed" or "transduced" cell is one which has been transfected, transformed or transduced with exogenous nucleic acid. The cell includes the primary subject cell and its progeny.

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The phrase "under transcriptional control" or "operatively linked" as used herein means that the promoter is in the correct location and orientation in relation to a polynucleotide to control the initiation of transcription by RNA polymerase and expression of the polynucleotide.

The term "therapeutic" as used herein means a treatment and/or prophylaxis. A therapeutic effect is obtained by suppression, diminution, remission, or eradication of at least one sign or symptom of a disease or disorder state.

As used herein, the terms "therapeutic compound", "therapeutic agent", "drug", "active pharmaceutical", and "active pharmaceutical ingredient" are used interchangeably to refer to chemical entities that display certain pharmacological effects in a body and are administered for such purpose. Non-limiting examples of therapeutic agents include, but are not limited to, hydrophilic therapeutic agents, hydrophobic therapeutic agents, antibiotics, antibodies, small molecules, anti-cancer agents, chemotherapeutic agents, immunomodulatory agents, RNA molecules, siRNA molecules, DNA molecules, gene editing agents, gene-silencing agents, CRISPR-associated agents (e.g., guide RNA molecules, endonucleases, and variants thereof), analgesics, vaccines, anticonvulsants; anti-diabetic agents, antifungal agents, antineoplastic agents, anti-parkinsonian agents, anti-rheumatic agents, appetite suppressants, biological response modifiers, cardiovascular agents, central nervous system stimulants, contraceptive agents, dietary supplements, vitamins, minerals, lipids, saccharides, metals, amino acids (and precursors), nucleic acids and precursors, contrast agents, diagnostic agents, dopamine receptor agonists, erectile dysfunction agents, fertility agents, gastrointestinal agents, hormones, immunomodulators, antihypercalcemia agents, mast cell stabilizers, muscle relaxants, nutritional agents, ophthalmic agents, osteoporosis agents, psychotherapeutic agents, parasympathomimetic agents, parasympatholytic agents, respiratory agents, sedative hypnotic agents, skin and mucous membrane agents, smoking cessation agents, steroids, sympatholytic agents, urinary tract agents, uterine relaxants, vaginal agents, vasodilator, anti-hypertensive, hyperthyroids, anti-hyperthyroids, anti-asthmatics and vertigo agents. In certain embodiments, the one or more therapeutic agents are water-soluble, poorly water-soluble drug or a drug with a low, medium or high melting point. The therapeutic agents may be provided with or without a stabilizing salt or salts.

Some examples of active ingredients suitable for use in the pharmaceutical formulations and methods of the present invention include: hydrophilic, lipophilic, amphiphilic or hydrophobic, and that can be solubilized, dispersed, or partially solubilized and dispersed, on or about the nanocluster. The active agent-nanocluster combination may be coated further to encapsulate the agent-nanocluster combination and may be directed to a target by functionalizing the nanocluster

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with, e.g., aptamers and/or antibodies. Alternatively, an active ingredient may also be provided separately from the solid pharmaceutical composition, such as for co-administration. Such active ingredients can be any compound or mixture of compounds having therapeutic or other value when administered to an animal, particularly to a mammal, such as drugs, nutrients, cosmeceuticals, nutraceuticals, diagnostic agents, nutritional agents, and the like. The active agents described herein may be found in their native state, however, they will generally be provided in the form of a salt. The active agents described herein include their isomers, analogs and derivatives.

An "effective amount" as used herein, means an amount which provides a therapeutic or prophylactic benefit.

The term "therapeutically effective amount" refers to the amount of the subject compound that will elicit the biological or medical response of a tissue, system, or subject that is being sought by the researcher, veterinarian, medical doctor or other clinician. The term "therapeutically effective amount" includes that amount of a compound that, when administered, is sufficient to prevent development of, or alleviate to some extent, one or more of the signs or symptoms of the disorder or disease being treated. The therapeutically effective amount will vary depending on the compound, the disease and its severity and the age, weight, etc., of the subject to be treated.

The terms "patient," "subject," "individual," and the like are used interchangeably herein, and refer to any animal, or cells thereof whether in vitro or in situ, amenable to the methods described herein. In certain non-limiting embodiments, the patient, subject or individual is a human.

A "disease" is a state of health of an animal wherein the animal cannot maintain homeostasis, and wherein if the disease is not ameliorated then the animal's health continues to deteriorate.

In contrast, a "disorder" in an animal is a state of health in which the animal is able to maintain homeostasis, but in which the animal's state of health is less favorable than it would be in the absence of the disorder. Left untreated, a disorder does not necessarily cause a further decrease in the animal's state of health.

To "treat" a disease as the term is used herein, means to reduce the frequency or severity of at least one sign or symptom of a disease or disorder experienced by a subject.

As used herein, "treating a disease or disorder" means reducing the frequency with which a symptom of the disease or disorder is experienced by a patient. Disease and disorder are used interchangeably herein.

A disease or disorder is "alleviated" if the severity of a sign or symptom of the disease or disorder, the frequency with which such a sign or symptom is experienced by a patient, or both, is

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reduced.

By the term "modulating," as used herein, is meant mediating a detectable increase or decrease in the level of a response in a subject compared with the level of a response in the subject in the absence of a treatment or compound, and/or compared with the level of a response in an otherwise identical but untreated subject. The term encompasses perturbing and/or affecting a native signal or response thereby mediating a beneficial therapeutic response in a subject, preferably, a human.

"Parenteral" administration of a composition includes, e.g., subcutaneous (s.c.), intravenous (i.v.), intramuscular (i.m.), or intrasternal injection, or infusion techniques.

Ranges: throughout this disclosure, various aspects of the invention can be presented in a range format. It should be understood that the description in range format is merely for convenience and brevity and should not be construed as an inflexible limitation on the scope of the invention. Accordingly, the description of a range should be considered to have specifically disclosed all the possible subranges as well as individual numerical values within that range. For example, description of a range such as from 1 to 6 should be considered to have specifically disclosed subranges such as from 1 to 3, from 1 to 4, from 1 to 5, from 2 to 4, from 2 to 6, from 3 to 6 etc., as well as individual numbers within that range, for example, 1, 2, 2.7, 3, 4, 5, 5.3, and 6. This applies regardless of the breadth of the range.

Description

The presenet invention is based, in part, on the unexpected results that nanoparticles comprising at least one ionizable amphiphilic Janus dendrimer having the structure of Formula (I) effectively and efficiently delivered an agent to a target of interest. Thus, in one aspect, the present invention relates to an ionizable amphiphilic Janus dendrimer having the structure of Formula (I). In another aspect, the present invention relates to a nanoparticle comprisint at least one ionizable amphiphilic Janus dendrimer of the present invention. In some embodiments, the nanoparticle further comprises at least one agent. In some embodiment, the nanoparticle further comprises at least one agent that is encapsulated by the onizable amphiphilic Janus dendrimer of the present invention. In another aspect, the present invention relates to a composition comprisint at least one ionizable amphiphilic Janus dendrimer of the present invention or a nanoparticle thereof. In some embodiments, the composition is a vaccine.

In one aspect, the present invention relates to methods of delivering an agent to a target of interest using at least one ionizable amphiphilic Janus dendrimer of the present invention or a

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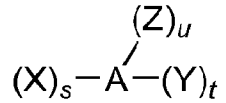
nanoparticle or a composition thereof. In another aspect, the present invention relates to methods of preventing or treating a disease or disorder in a subject using at least one ionizable amphiphilic Janus dendrimer of the present invention or a nanoparticle or a composition thereof. In another aspect, the present invention relates to methods of inducing an adaptive immune response in a subject using at least one ionizable amphiphilic Janus dendrimer of the present invention or a nanoparticle or a composition thereof.

Compounds of the Invention

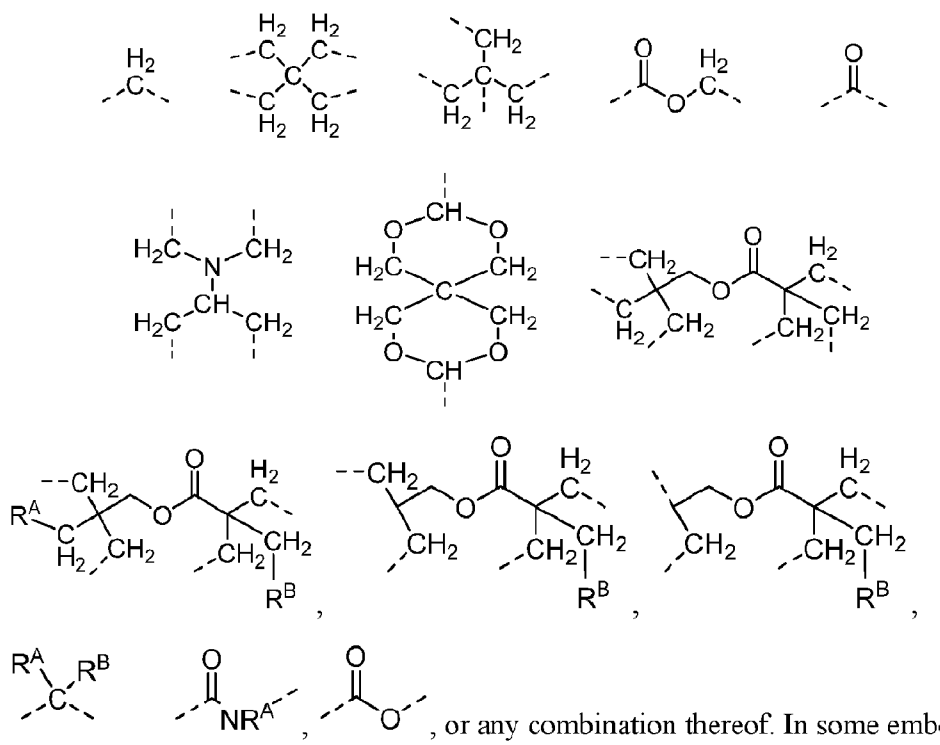
In one aspect, the invention relates to amphiphilic Janus dendrimers of Formula (I):



Formula (I),

or a racemate, an enantiomer, a diastereomer, a pharmaceutically acceptable salt, or a derivative thereof. In various embodiments, the amphiphilic Janus dendrimer is an ionizable amphiphilic Janus dendrimer.

In some embodiments, A is a polyvalent group comprising at least one selected from the following structures:



, or any combination thereof. In some embodiments, A is a

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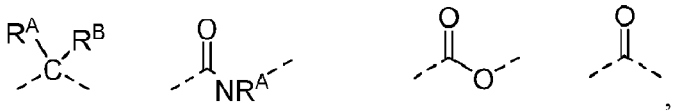
polyvalent group comprising at least one selected from

 , or any combination thereof. In some embodiments, A is optionally substituted.

In some embodiments, dashed lines represent a binding site of one of X, Y, or Z.

In some embodiments, X is a hydrophilic group comprising at least one amine. In some embodiments, the hydrophilic group is optionally substituted. In some embodiments, the amine is optionally substituted.

In some embodiments, Y is a lipophilic group comprising at least one C_1-C_{50} alkyl chain. In some embodiments, Y is a lipophilic group comprising at least one C_1-C_{30} alkyl chain. In some embodiments, the lipophilic group is optionally substituted. In some embodiments, the C_1-C_{50} alkyl chain is optionally substituted.

In some embodiments, Z is a group comprising at least one selected from ethylene glycol, diethylene glycol, triethylene glycol, or polyethylene glycol chain. In some embodiments, Z is optionally substituted. In some embodiments, the ethylene glycol, diethylene glycol, triethylene glycol, and/or polyethylene glycol chain are optionally substituted.

In some embodiments, R^A is selected from hydrogen, halide, hydroxy, C_1-C_{50}-alkyl, C_1-C_{50}-alkyl halide, C_1-C_{50}-alkoxy, C_1-C_{50}-alkoxy halide, or any combination thereof. In some embodiments, R^A is selected from hydrogen, halide, hydroxy, C_1-C_{30}-alkyl, C_1-C_{30}-alkyl halide, C_1-C_{30}-alkoxy, C_1-C_{30}-alkoxy halide, or any combination thereof. In some embodiments, the hydroxy, C_1-C_{50}-alkyl, C_1-C_{50}-alkyl halide, C_1-C_{50}-alkoxy, and/or C_1-C_{50}-alkoxy halide are optionally substituted.

In some embodiments, R^B is selected from hydrogen, halide, hydroxy, C_1-C_{50}-alkyl, C_1-C_{50}-alkyl halide, C_1-C_{50}-alkoxy, C_1-C_{50}-alkoxy halide, or any combination thereof. In some embodiments, R^B is selected from hydrogen, halide, hydroxy, C_1-C_{30}-alkyl, C_1-C_{30}-alkyl halide, C_1-C_{30}-alkoxy, C_1-C_{30}-alkoxy halide, or any combination thereof. In some embodiments, the hydroxy, C_1-C_{50}-alkyl, C_1-C_{50}-alkyl halide, C_1-C_{50}-alkoxy, and/or C_1-C_{50}-alkoxy halide are optionally substituted.

In some embodiments, s is an integer from 0 to 5. In some embodiments, s is an integer from 1 to 5. For example, in one embodiment, s is an integer 5. In one embodiment, s is an integer 4. In one embodiment, s is an integer 3. In one embodiment, s is an integer 2. In one embodiment, s is an integer 1. In one embodiment, s is an integer 0.

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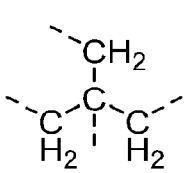
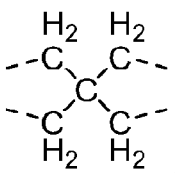
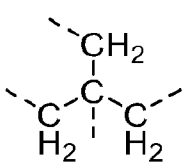
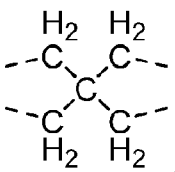
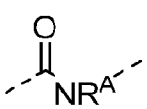
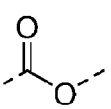
In some embodiments, t is an integer from 0 to 5. In some embodiments, t is an integer from 1 to 5. For example, in one embodiment, t is an integer 5. In one embodiment, t is an integer 4. In one embodiment, t is an integer 3. In one embodiment, t is an integer 2. In one embodiment, t is an integer 1. In one embodiment, t is an integer 0.

In some embodiments, u is an integer from 0 to 4. In some embodiments, u is an integer from 1 to 4. For example, in one embodiment, u is an integer 4. In one embodiment, u is an integer 3. In one embodiment, u is an integer 2. In one embodiment, u is an integer 1. In one embodiment, u is an integer 0.

In some embodiments, the sum of s, t, and u is equal to the valency of A. For example, in one embodiment, the valency of A is 5, and the sum of s, t, and u is 5. In one embodiment, the valency of A is 4, and the sum of s, t, and u is 4. In one embodiment, s and t are each 2. In one embodiment, s is 1 and t is 3. In one embodiment, s is 3 and t is 1. In one embodiment, s is 1, t is 1, and u is 2. In one embodiment, s is 2, t is 1, and u is 1. In one embodiment, s is 1, t is 2, and u is 1. In one embodiment, the valency of A is 2, and the sum of s, t, and u is 2. In one embodiment, s and t are each 1.

For example, in some embodiments, A is represented by  or  ; and s and t are each 2.

In some embodiments, A is represented by  or  ; t is 2; s is 1; and u is 0 or 1.

In some embodiments, A is represented by  or  ; and s and t are each 1.

In one embodiment, u is 0. Thus, in one emdiment, wherein the amphiphilic Janus dendrimer having the structure of Formula (I) is an amphiphilic Janus dendrimer having the structure of Formula (II):

$$(X)_s-A-(Y)_t$$

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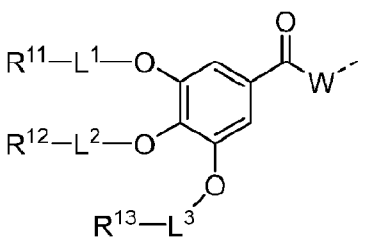
Formula (II),

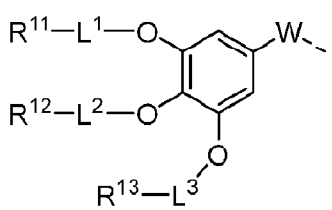
or a racemate, an enantiomer, a diastereomer, a pharmaceutically acceptable salt, or a derivative thereof.

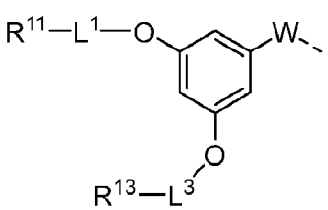
In one embodiment, X is a hydrophilic group comprising one amine. In one embodiment, X is a hydrophilic group comprising two amines. In one embodiment, X is a hydrophilic group comprising three amines. In one embodiment, X comprises at least one primary amine. In one embodiment, X comprises at least one secondary amine. In one embodiment, X comprises at least one tertiary amine. For example, in one embodiment, X comprises at least two tertiary amines.

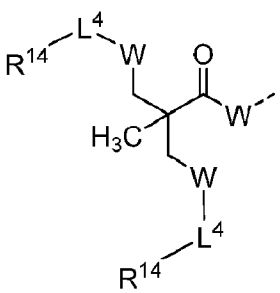
In one embodiment, X comprises at least one amine which is protonated under biological conditions. In one embodiment, X comprises at least one amine which has a formal charge of +1 under biological conditions. In one embodiment, X comprises at least one carbohydrate.

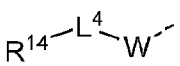
In one embodiment, each occurrence of X is independently selected from the following structures:

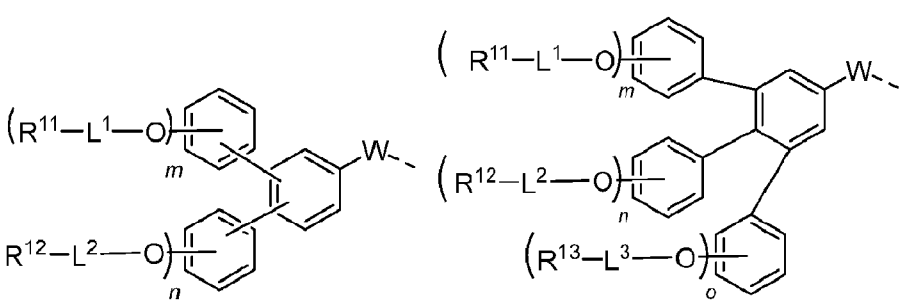












, , or any combination thereof.

In some embodiments, dashed lines indicate the connection to A.

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In some embodiments, each occurrence of m, n, o is independently an integer from 1 to 5.

In some embodiments, each occurrence of W is independently selected from C=O, $C(R^W)(R^W)$, NR^W, O, or S. In some embodiments, each occurrence of R^W is independently selected from hydrogen, halide, hydroxy, alkyl, alkyl halide, aryl, aryl halide, alkoxy, alkoxy halide, or any combination thereof. In one embodiment, each occurrence of W is NH. In one embodiment, each occurrence of W is O. In one embodiment, at least one occurrence of W is NH and at least occurrence of W is O.

In some embodiments, each occurrence of L^1, L^2, L^3, and L^4 is independently a covalent bond or a divalent linking group selected from alkylene, cycloalkylene, heteroalkylene, heterocycloalkylene, alkenylene, alkynylene, arylene, heteroarylene, silyl, amine, amide, ester, ether, carbonyl, carbamate, thioether, thioester, disulfide, hydrazine, urea, thiourea, phosphate, poly(alkyl ether), heteroatom, or any combination thereof. In one embodiment, at least one occurrence of L^1, L^2, L^3, and L^4 is a poly(alkyl ether) or an oligo(alkyl ether). In one embodiment, each occurrence of L^1, L^2, L^3, and L^4 is polyethylene glycol (PEG) or oligoethyleneoxide. In one embodiment, each occurrence of L^1, L^2, L^3, and L^4 independently has the structure $-[CH_2CH_2O]_n-$; wherein n is an integer between 0 and 10.

In some embodiments, each occurrence of R^{11}, R^{12}, R^{13}, and R^{14} is independently selected from hydrogen, deuterium, alkyl, aryl, heteroaryl, cycloalkyl, alkoxy, phenoxy, amine, heterocycloalkyl, carbonyl, or any combination thereof. In some embodiments, at least one of R^{11}, R^{12}, R^{13}, or R^{14} comprises an amine. In one embodiment, one of R^{11}, R^{12}, and R^{13} comprises an amine. In one embodiment, two of R^{11}, R^{12}, and R^{13} comprises an amine. In one embodiment, three of R^{11}, R^{12}, and R^{13} comprises an amine. In one embodiment, at least one of R^{11}, R^{12}, and R^{13} comprises two amines. In one embodiment the amine of one of R^{11}, R^{12}, and R^{13} is protonated under biological conditions. In one embodiment, any of R^{11}, R^{12}, and R^{13} that does not comprise an amine comprises an alkyl group, an aryl group, or a combination thereof.

In one embodiment, at least one occurrence of R^{11}, R^{12}, R^{13}, or R^{14} has the structure

$$-C(O)(CH_2)_m-N(R^1)(R^2).$$

In some embodiments, each occurrence of m is an integer from 1 to 10. In one embodiment, m is 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10.

In some embodiments, each occurrence of R^1 and R^2 is independently selected from the hydrogen, deuterium, alkyl, aryl, cycloalkyl, amine, heterocycloalkyl, carbonyl, or any combination thereof. In some embodiments, R^1 and R^2 may together form a ring.

In one embodiment, R^1 and R^2 are each alkyl. In one embodiment, R^1 and R^2 are each

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methyl. In one embodiment, R^1 and R^2 together form a 6-membered heterocyclic ring. In one embodiment, R^1 and R^2 together form a piperidine ring with the N to which they are bound. In one embodiment, R^1 and R^2 together form an N-alkyl piperzaine ring with the N to which they are bound. In one embodiment, R^1 and R^2 are each hydrogen.

In one embodiment, at least one occurrence of R^{11}, R^{12}, R^{13}, or R^{14} has the structure

$$-C(O)(CR^5R^6)_m-N(R^1)(R^2).$$

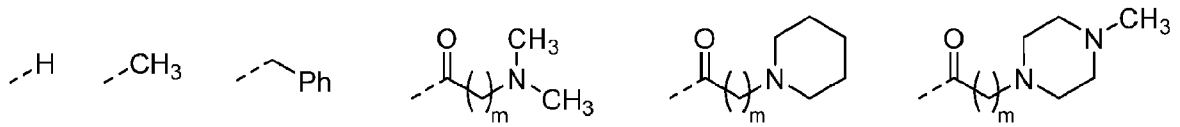
In some embodiments, each occurrence of m is an integer from 1 to 10. In one embodiment, m is 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10.

In some embodiments, each occurrence of R^1 and R^2 is independently selected from hydrogen, deuterium, alkyl, aryl, cycloalkyl, amine, heterocycloalkyl, carbonyl, or any combination thereof.

In some embodiments, each occurrence of R^5 and R^6 is independently selected from hydrogen, deuterium, alkyl, aryl, heteroaryl, cycloalkyl, alkoxy, phenoxy, amine, heterocycloalkyl, carbonyl, or any combination thereof. In some embodiments, any two or more of R^1, R^2, R^5, and R^6 may together form a ring.

In one embodiment, the group having the structure $-C(O)(CR^5R^6)_m-N(R^1)(R^2)$ is derived from an amino acid. In one embodiment, the amino acid is a canonical amino acid. In one embodiment, the amino acid is a non-canonical amino acid. In one embodiment, the amino acid is a beta-amino acid (i.e., m is at least 2, and one geminal pair of R^5 and R^6 is H). In one embodiment, one of R^5 and R^6 is H, and the other of R^5 and R^6 is not H. In one embodiment, the group may be chiral. In one embodiment, the group may be enantiomerically enriched or enantiomerically pure. In one embodiment, the group, the group may be racemic.

In one embodiment, each occurrence of R^{11}, R^{12}, R^{13}, and R^{14} is independently selected from the following structures:



.

In some embodiments, each occurrence of m is an integer from 1 to 10. In one embodiment, m is 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10.

In one embodiment, each occurrence of X is independently selected from the following structures:

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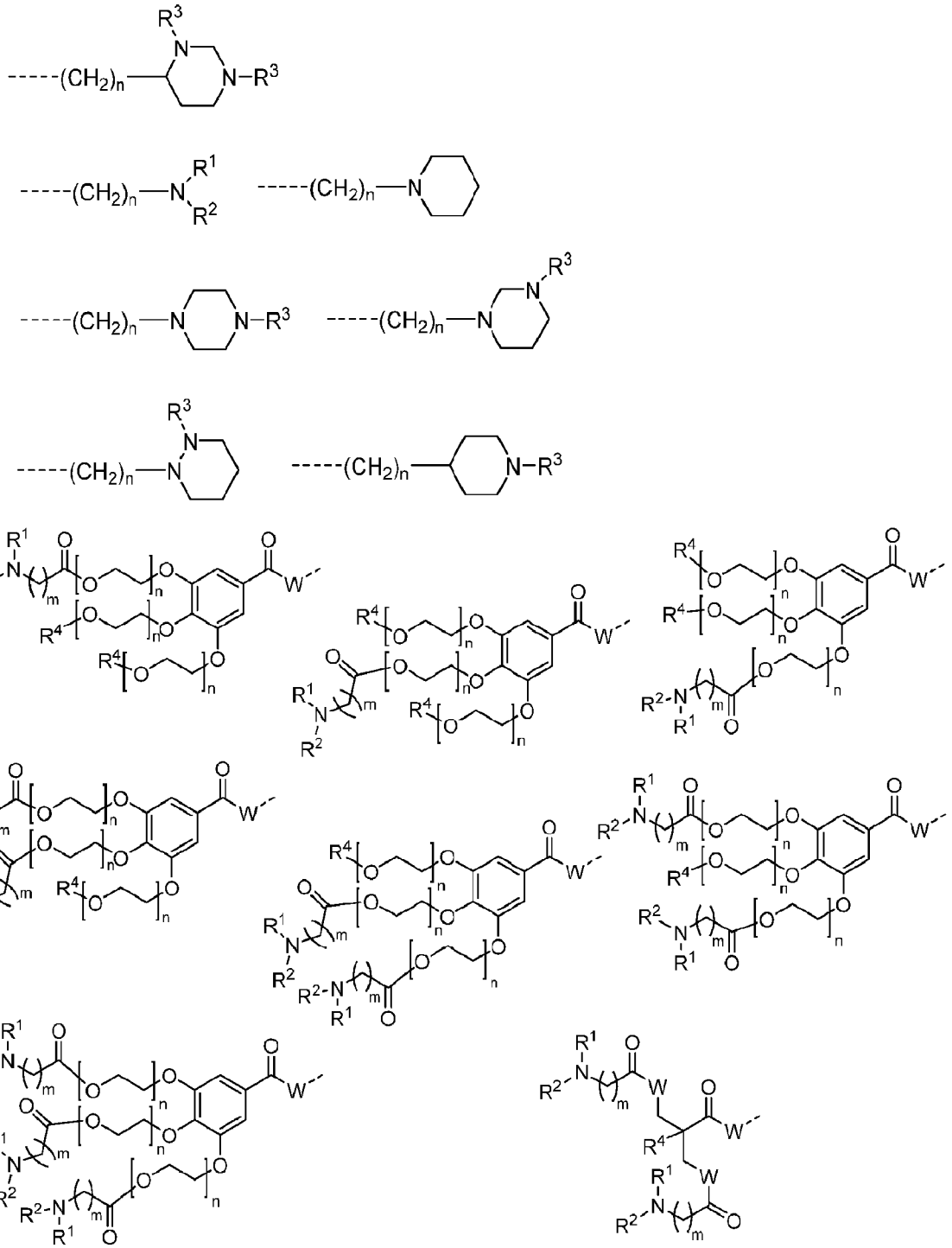

, or any combination thereof.

In some embodiments, each occurrence of n is independently an integer from 1 to 20. In some embodiments, each occurrence of n is independently an integer from 1 to 10.

In some embodiments, each occurrence of m is independently an integer from 1 to 20. In

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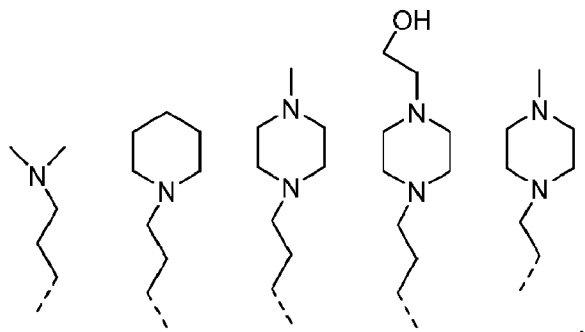
some embodiments, each occurrence of m is independently an integer from 1 to 10.

In some embodiments, each occurrence of R^1 and R^2 is independently selected from hydrogen, deuterium, alkyl, aryl, cycloalkyl, amine, heterocycloalkyl, carbonyl, or any combination thereof. In some embodiments, R^1 and R^2 may together form a ring.

In some embodiments, each occurrence of R^3 is independently selected from hydrogen, $(CH_2)_n$, $(CH_2)_n$-OH, or any combination thereof.

In some embodiments, each occurrence of R^4 is independently selected from hydrogen, deuterium, alkyl, aryl, or any combinations thereof.

For example, in some embodiments, each occurrence of X is independently selected from



, or any combination thereof.

In one embodiment, Y is a lipophilic (hydrophobic) group comprising at least one C_1-C_{50} alkyl chain. In one embodiment, Y is a lipophilic (hydrophobic) group comprising at least one C_1-C_{30} alkyl chain. In one embodiment, Y is a lipophilic (hydrophobic) group comprising at least one C_4-C_{30} alkyl chain. For example, in one embodiment, Y is a lipophilic (hydrophobic) group comprising at least two C_1-C_{30} alkyl chain. In one embodiment, Y is a lipophilic (hydrophobic) group comprising at least two C_1-C_{30} alkyl chain having differing numbers of carbon atoms.

In one embodiment, Y is a lipophilic group further comprising at least one linking group selected from the group consisting of alkylene, cycloalkylene, heteroalkylene, heterocycloalkylene, alkenylene, alkynylene, arylene, heteroarylene, silyl, amine, amide, ester, ether, carbonyl, carbamate, thioether, thioester, disulfide, hydrazine, urea, thiourea, phosphate, poly(alkyl ether), heteroatom, or any combination thereof. In one embodiment, Y comprises alkylene, arylene, alkenylene, alkynylene, disulfide, ether, or any combination thereof. In one embodiment, Y does not comprise an amine. In one embodiment, Y is a lipophilic group comprising at least one C_6 alkyl chain, at least one C_7 alkyl chain, at least one C_8 alkyl chain, at least one C_9 alkyl chain, at least one C_{10} alkyl chain, at least one C_{11} alkyl chain, at least one C_{12} alkyl chain, at least one C_{13} alkyl chain, at least one C_{14} alkyl chain, at least one C_{15} alkyl chain, at least one C_{20} alkyl chain, at least one C_{21} alkyl chain, at least one C_{23} alkyl chain, at least one C_{27} alkyl chain, at least one C_{30} alkyl chain, at

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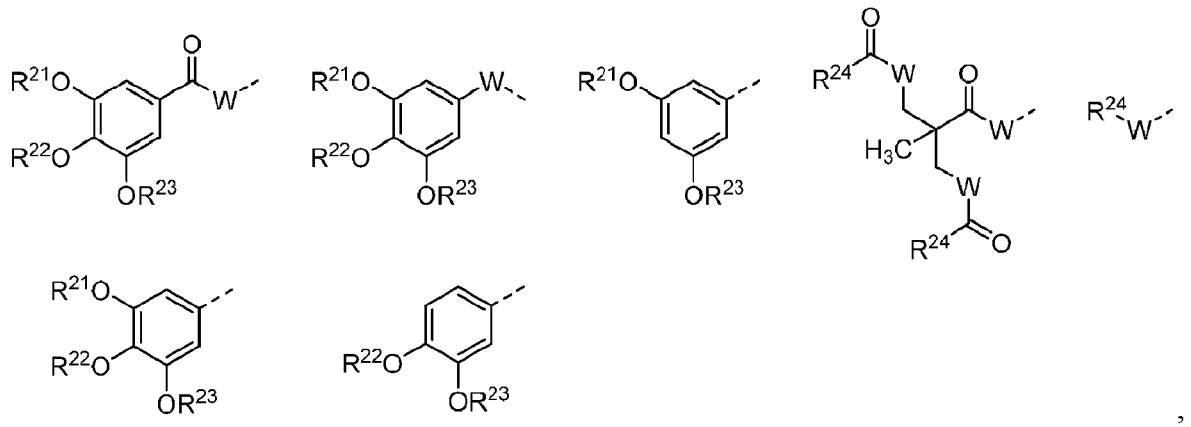
least one C_{35} alkyl chain, at least one C_{40} alkyl chain, or at least one C_{50} alkyl chain. In one embodiment, Y comprises at least one carbohydrate.

In one embodiment, each occurrence of Y is independently selected from the following structures:



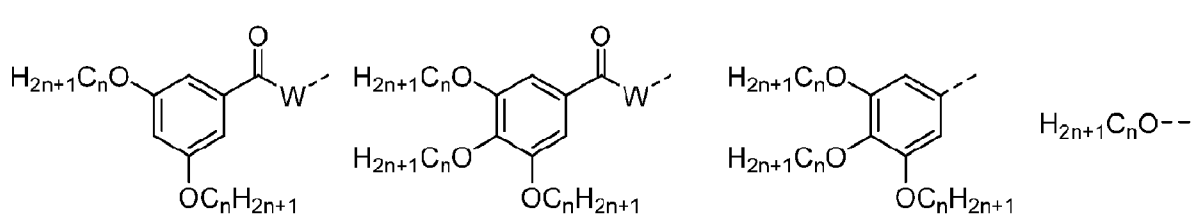
or any combination thereof.

In some embodimetns, dashed lines indicate the connection to A.

In some embodimetns, each occurrence of W is independently selected from C=O, $C(R^W)(R^W)$, NR^W, O, or S. In some embodimetns, each occurrence of R^W is independently selected from hydrogen, halide, hydroxy, alkyl, alkyl halide, aryl, aryl halide, alkoxy, alkoxy halide, or any combination thereof. In one embodiment, each occurrence of W represents O. In one embodiment, each occurrence of W represents NH. In one embodiment, at least one occurrence of W represents O and at least one occurrence of W represents NH.

In some embodments, each occurrence of R^{21}, R^{22}, R^{23}, and R^{24} independently comprises C_1-C_{50}-alkyl. In some embodments, each occurrence of R^{21}, R^{22}, R^{23}, and R^{24} independently comprises a linear or branched C_1-C_{50}-alkyl. In some embodments, each occurrence of R^{21}, R^{22}, R^{23}, and R^{24} independently comprises C_1-C_{30}-alkyl. In some embodments, each occurrence of R^{21}, R^{22}, R^{23}, and R^{24} independently comprises a linear or branched C_1-C_{30} alkyl. In one embodiment, each occurrence of R^{21}, R^{22}, R^{23}, and R^{24} independently represents C_8-C_{12} alkyl.

In one embodiment, each occurrence of Y is independently selected from the following structures:



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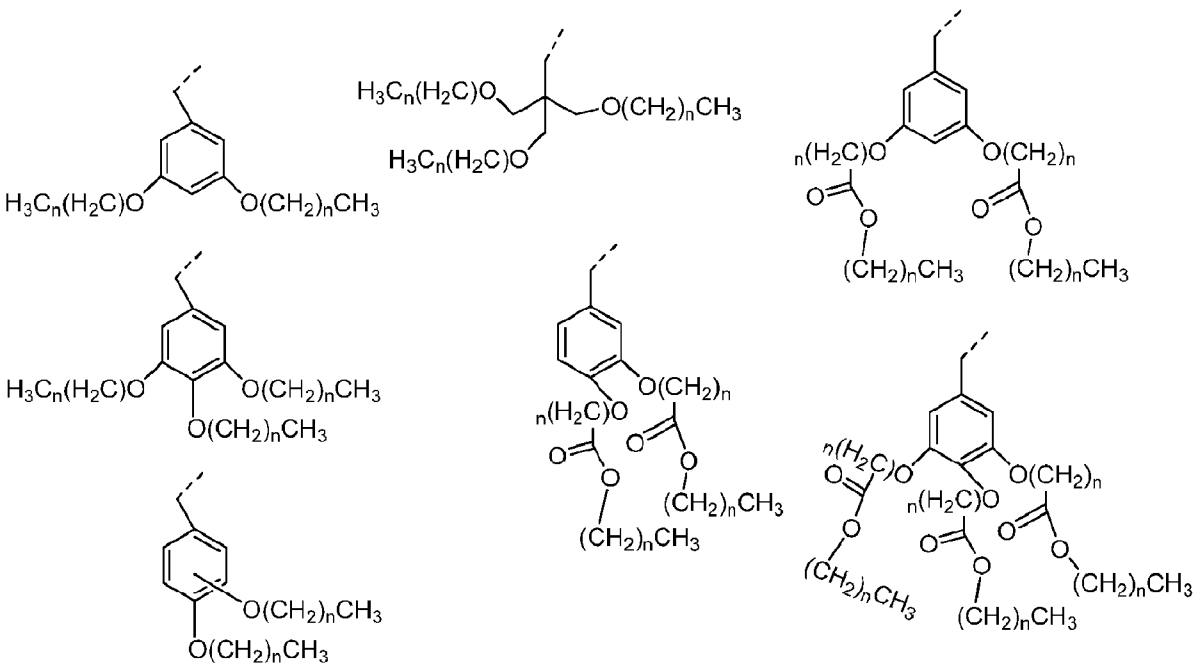
;

In some embodimetns, each occurrence of W is independently selected from C=O, $C(R^W)(R^W)$, NR^W, O, or S. In some embodimetns, each occurrence of R^W is independently selected from hydrogen, halide, hydroxy, alkyl, alkyl halide, aryl, aryl halide, alkoxy, alkoxy halide, or any combination thereof. In one embodiment, each occurrence of W represents O. In one embodiment, each occurrence of W represents NH. In one embodiment, at least one occurrence of W represents O and at least one occurrence of W represents NH.

In some embodiments, n is an integer from 1 to 30. In some embodiments, n is an integer from 6 to 18.

In some embodiments, each occurrence of Y is independently selected from



, or any combination thereof.

. In some embodiments, each occurrence of Y is independently selected from

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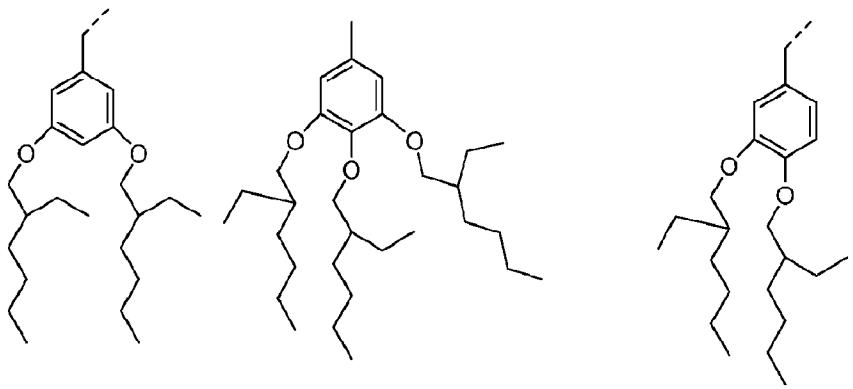

, or any combination thereof.

In some embodiments, dashed lines indicate the connection to A.

In some embodiments, each occurrence of n is independently an integer from 1 to 20.

In some embodiments, each occurrence of $-(CH_2)_nCH_3$, $-O(CH_2)_n-$, and $-O(CH_2)_nCH_3$ is independently linear or branched.

In some embodiments, t is an integer from 2 to 5. Thus, in some embodimetnts, the amphiphilic Janus dendrimer comprises a first Y and a second Y. In some embodiments, the first Y comprises an alkyl chain having a different number of carbon atoms than the second Y comprises an alkyl chain. In some embodiments, the first Y comprises an alkyl chain having an even number of carbon atoms, and the second Y comprises an alkyl chain having an odd number of carbon atoms. In some embodiments, the ratio between the carbon atoms in the first Y and the carbon atoms in the second Y is greater than or equal to 3 and less than 7.

In one embodiment, u is 1 or 2. In one embodiment, u is 1. In one embodiment, u is 2.

In one embodiment, Z does not comprise an amine.

In one embodiment, Z is not a lipophilic group.

In one embodiment, Z comprises at least one carbohydrate.

In one embodiment, each occurrence of Z is independently selected from the following structures:

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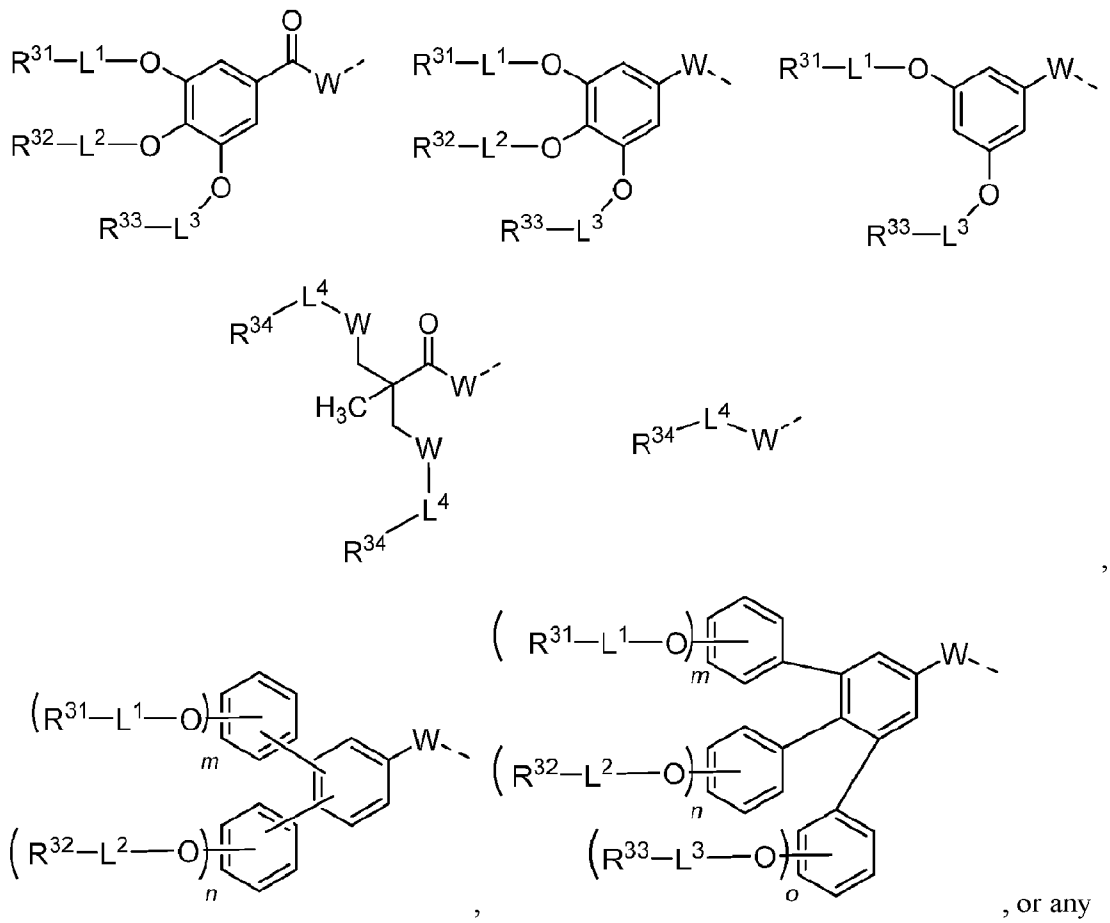

combination thereof.

In some embodiments, dashed lines indicate the connection to A.

In some embodiments, each occurrence of L^1, L^2, L^3, and L^4 is independently a covalent bond or a divalent linking group selected from alkylene, cycloalkylene, heteroalkylene, heterocycloalkylene, alkenylene, alkynylene, arylene, heteroarylene, silyl, amine, amide, ester, ether, carbonyl, carbamate, thioether, thioester, disulfide, hydrazine, urea, thiourea, phosphate, poly(alkyl ether), heteroatom, or any combination thereof. In one embodiment, at least one occurrence of L^1, L^2, L^3, and L^4 in the group Z represents a single bond. In one embodiment, at least one of L^1, L^2, L^3, and L^4 represents a group of formula –W-C(O)$(CH_2)_p$C(O)–W–; wherein p is an integer between 1 and 10 and W represents NH, O, or S. In one embodiment, at least one of L^1, L^2, L^3, and L^4 comprises polyethylene gycol.

In some embodiments, each occurrence of R^{31}, R^{32}, R^{33}, and R^{34} is independently selected from hydrogen, deuterium, alkyl, aryl, heteroaryl, cycloalkyl, alkoxy, phenoxy, amine, heterocycloalkyl, carbonyl, or any combination thereof.

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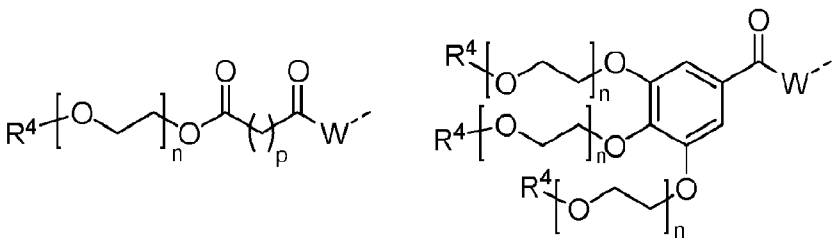
In some embodiments, each occurrence of m, n, o is independently an integer from 1 to 5.

In one embodiment, each occurrence of Z is independently selected from the following structures:



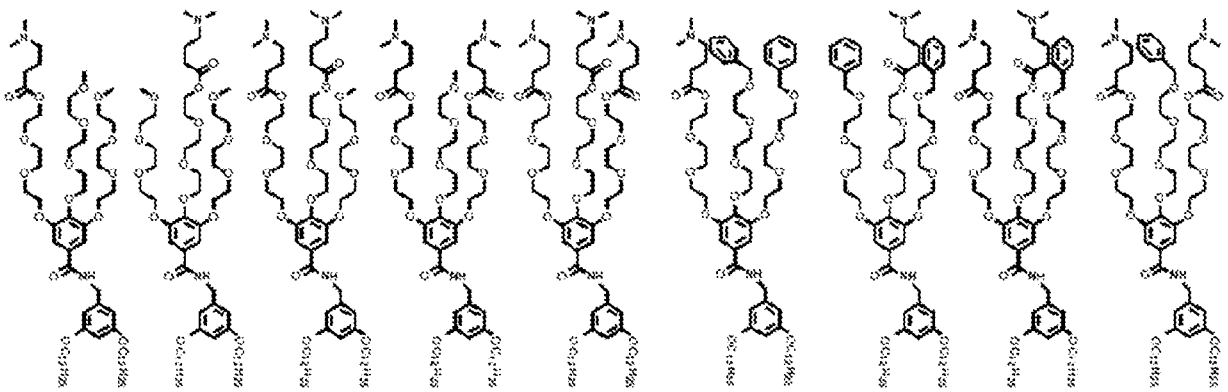
In some embodiments, each occurrence of n is independently an integer from 1 to 100.

In some embodiments, each occurrence of p is independently an integer from 1 to 10.

In some embodiments, each occurrence of R^4 is independently selected from hydrogen, deuterium, alkyl, aryl, or any combination thereof.

In some embodiments, the amphiphilic Janus dendrimer is an amphiphilic Janus dendrimer having a structure selected from at least one structure of Figure 12, at least one structure of Figure 13, at least one structure of Figure 14, at least one structure of Figure 47, at least one structure of Figure 60, at least one structure of Figure 80, or any combination thereof.

In some embodiments, the amphiphilic Janus dendrimer is an amphiphilic Janus dendrimer having a structure selected from:



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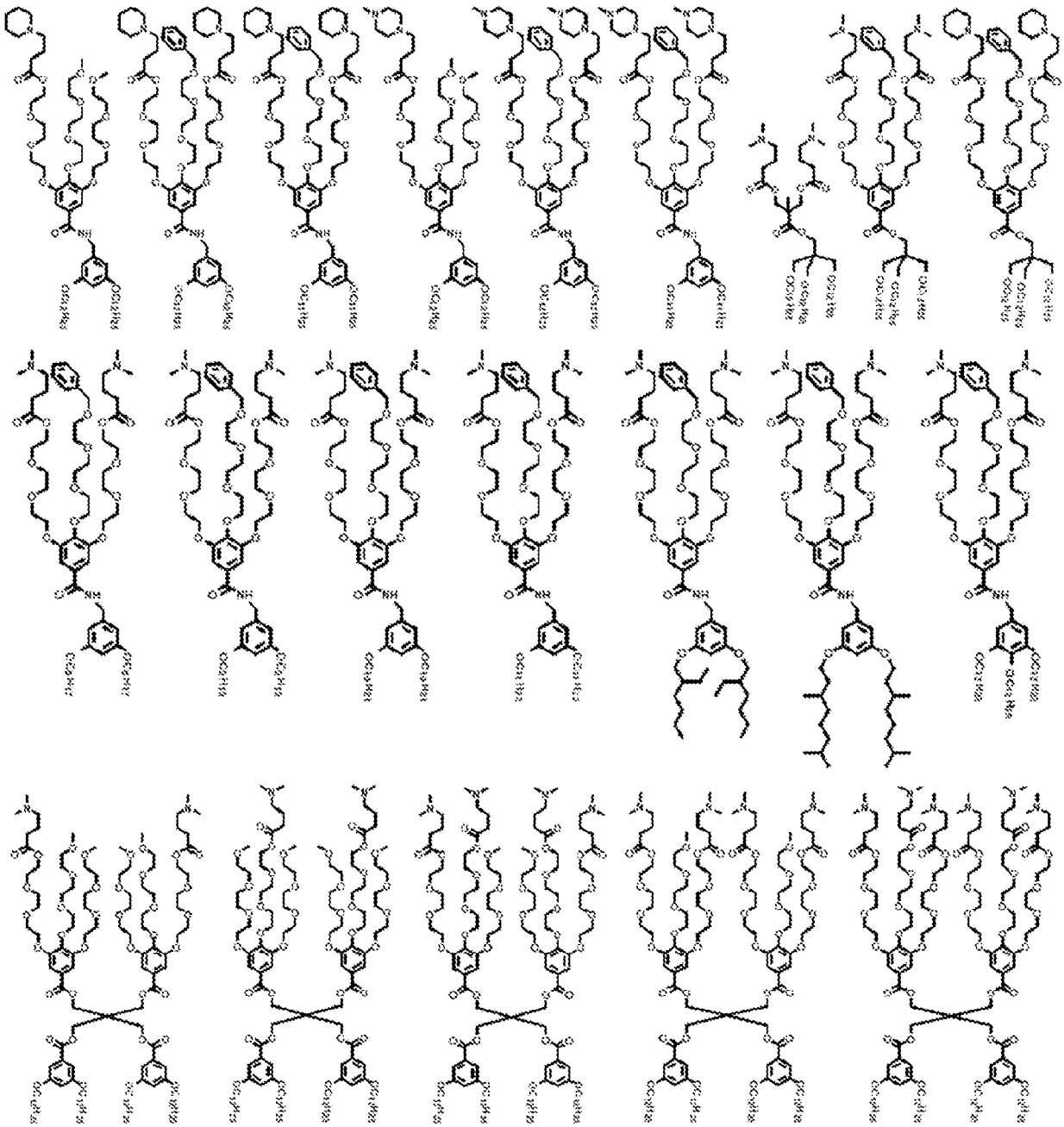

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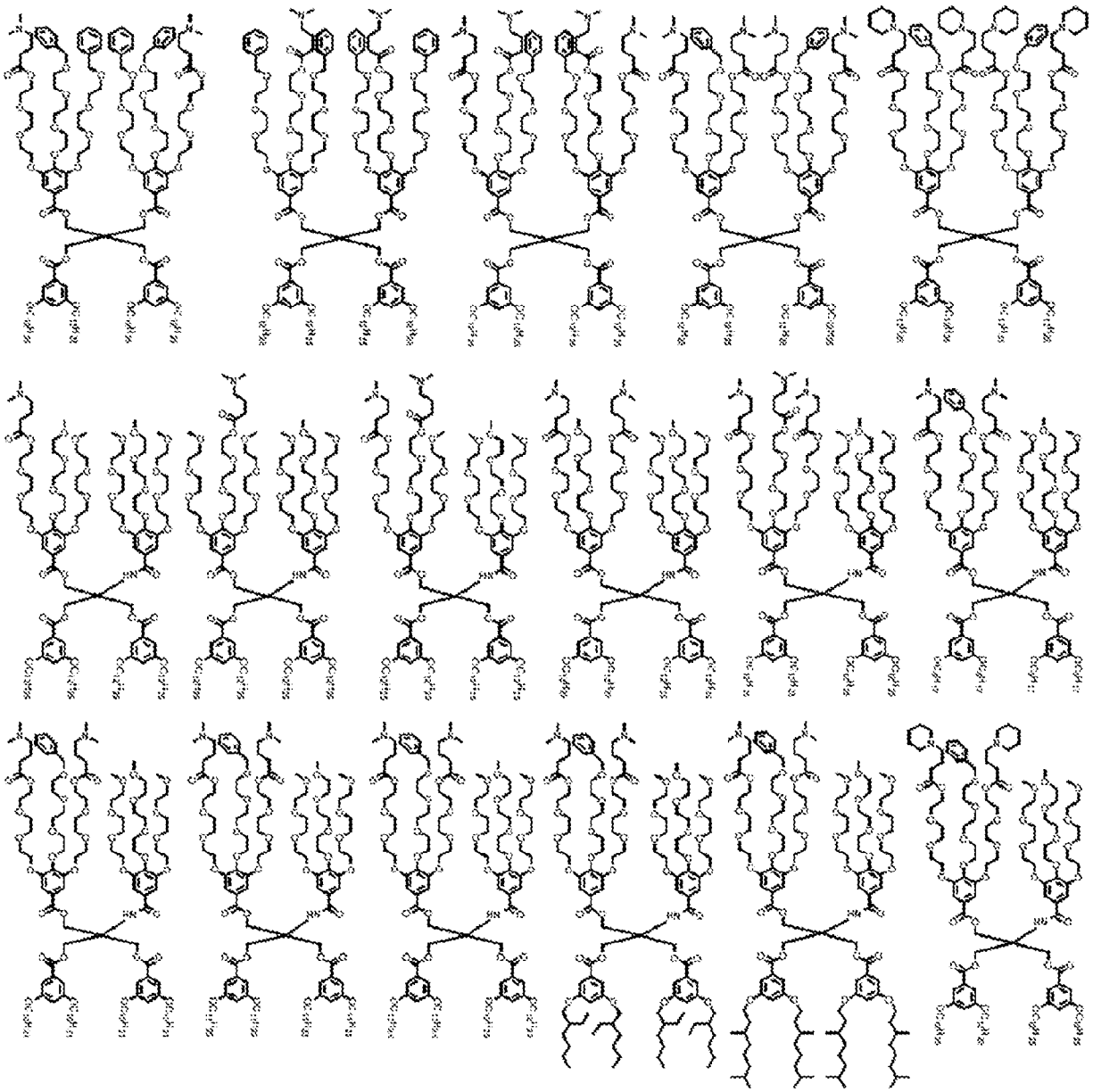

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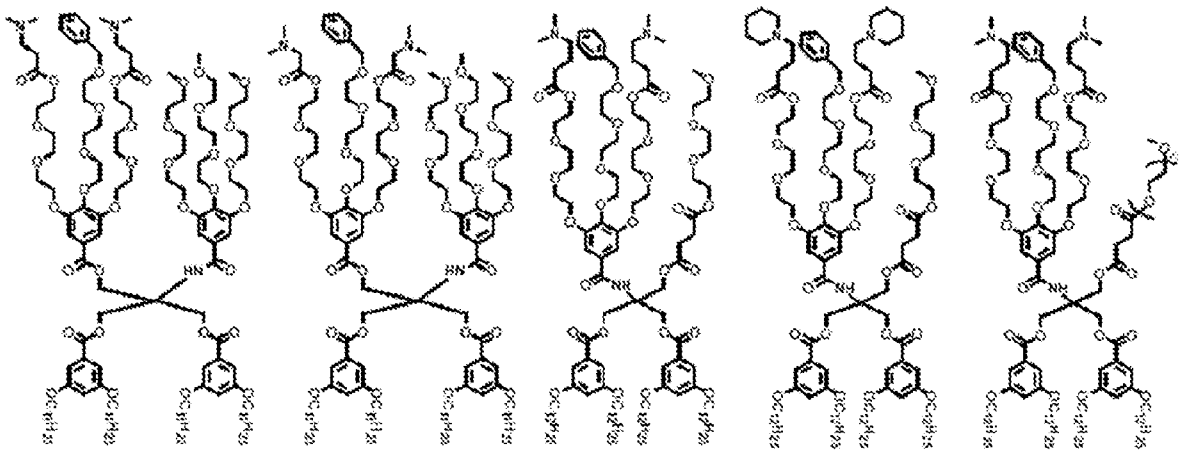

In some embodiments, amphiphilic Janus dendrimer having the structure of Formula (II) comprise an amine (the ionizable amine) in X that is sufficient to provide the dual hydrophilic and binding role in the IAJD. For example, Figure 47 summarizes IAJDs as described herein, synthesized, and investigated *in vitro* and *in vivo*, their schematic representations and the pK_a values. pKa values were determined as described in the examples. Co-assembly of all IAJDs from Figure 47 with Luc-mRNA was performed by the injection of an ethanol solution of IAJD into an acetate buffer of pH = 4. The acetate buffer contained mRNA added to the buffer from neutral water. Further details are found in the Example section below. The co-assembly forms a nanoparticle (DNP). Co-assembly is shown schematically in Figure 1. Figure 48 shows *in vivo* transfection results of the DNPs co-assembled from the IAJDs, shown in Figure 1. The IAJD numbers are shown on the top left, IAJD pK_a values, size in nm, with the polydispersities (PDI) of the resulting DNPs, are shown above each mouse image. The scale of the luminescence values is also shown. Representative images of mRNA delivery to different organs are shown in the bottom part of this figure. Figure 49 shows a comparison of the activity of DNPs assembled from the IAJDs, shown schematically in the top part of the Figure, *in vitro* (in blue) and *in vivo* (in red). Thus, in some embodiments, in dendrimers having the structure of Formula (II), the valency of A may be two, and the sum of s and t may be two.

In some embodimetns, the amphiphilic Janus dendrimer comprises a homochiral, racemic, or achiral branding points. Thus, in some embodiments, the ionizable amphiphilic Janus dendrimer is a homochiral ionizable amphiphilic Janus dendrimer, racemic ionizable amphiphilic Janus dendrimer, or achiral ionizable amphiphilic Janus dendrimer.

In some embodimetns, the amphiphilic Janus dendrimer is a symmetric amphiphilic Janus

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dendrimer.

In some embodimetns, the amphiphilic Janus dendrimer is an asymmetric amphiphilic Janus dendrimer.

In some embodimetns, the amphiphilic Janus dendrimer is a nonsymmetric amphiphilic Janus dendrimer.

For example, in some embodiments, the successful design of the hydrophobic region of the amphiphilic Janus dendrimer is based, in part, on the usage of dissimilar alkyl lengths and discovery of the unexpectedly important role of the primary structure of the hydrophobic part of the IAJDs which increases the activity of targeted delivery of a desired cargo to a target site up to 90.2 fold. In some embodiments, the nonsymmetric one-component amphiphilic Janus dendrimers do not require microfluidic or T-tube technology employed by 4-component LNPs to co-assemble with mRNA. In some embodiments, the one-component systems may co-assemble with mRNA into DNPs with about 97% nucleic acid encapsulation efficiency by simple injection of their ethanol solution into an acidic buffer containing mRNA rather than by microfluidic or T-tube technology required by LNPs.

For example, Figure 2 outlines the structure and co-assembly with mRNA of LNPs. Figure 1 shows the structures of sSS, SS and TM IAJDs with dissimilar and similar alkyl groups in their hydrophobic parts and their co-assembly with mRNA. IAJDs can be described as single-single, (SS, single hydrophilic dendron connected to single lipophilic dendron), twin-twin (TT, two hydrophilic dendrons connected to two lipophilic dendrons) and twin-mixed (TM, two different hydrophilic dendrons connected to two lipophilic dendrons) IAJDs.

For example, Figure 59 shows the synthesis of nonsymmetric IAJDs. In the first step the 3-benzylether of 3,5-dihydroxy methyl benzoate was produced in 39% isolated yield in 5 hours by etherification of **1** with BnCl at 80°C in DMF. Subsequently **2** was alkylated with 1-bromoundecane or 1bromopentadecane in DMF, with K_2CO_3 base at 120°C to produce 80-100% isolated yield of **3**. Hydrogenolysis of **3** (H_2/Pd, DCM/MeOH, 12 hours) produced **4** in 100% isolated yield. Alkylation of **4** with varying alkyl lengths from 1-bromooctane to 1-bromooctadecane in DMF, with K_2CO_3 base at 120°C generated the nonsymmetric compounds **5** in 74-92% isolated yield. Reduction of compounds **5** with $LiAlH_4$ in THF (0-23°C, 1 hour) produced the benzyl alcohols **6** in 93-100% isolated yield. Compounds **6** were reacted with 4-bromobutyric acid either *via* its acid chloride generated with $SOCl_2$ catalyzed by DMF in CH_2Cl_2 at 23°C followed by esterification in the presence of NEt_3/DMAP (0 – 23°C, 2 hours) or by direct esterification with DCC/DPTS in 12 hours to produce compounds **7** in 76 to 98% isolated yield. Compounds **7** were reacted with

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methylpiperazine or hydroxyethyl piperazine (K_2CO_3, MeCN, 95°C, 3 hours) to yield IAJDs **8** (70-90% isolated yield) and **9** (76-98% isolated yield). Their purity was determined by a combination of HPLC, MALDI-TOF and NMR and was higher than 99%. Their structures are shown in Figure 60 (IAJDs 113 to 178). IAJD133 has a similar structure with IAJD105 except that the interconnecting ester group of 105 was replaced with an amide in 133. The benzyl amine precursor of 133 was generated from the corresponding benzyl alcohol *via* its benzyl chloride obtained with $SOCl_2$ followed by reaction with K-phthalimide and subsequently hydrazine as reported.

Single-single (SS) IAJDs reported previously to display very high activity for delivery to lung were synthesized with nonsymmetric alkyl groups in their lipophilic part. They are IAJDs 110 to 159 from Figure 60. Their sequence-defined hydrophilic dendrons were synthesized as reported in Zhang et al., *J. Am. Chem. Soc.* **2021**, *143*, 12315–12327. The hydrophilic dendrons were reacted with selected nonsymmetric lipophilic dendrons **6** or their amine. For convenience, the IAJDs are referred to by their number followed by the ratio between their two alkyl groups forming their nonsymmetric lipophilic part. This nomenclature together with their entire and schematic structures shown in Figure 60 facilitates the discussion of their *in vitro* and *in vivo* activity *vs* molecular structure. For example, 116(11/13) and 117(11/13) both contain a combination of 11 and 13 carbons in their lipophilic part but 116 contains a methyl piperazine while 117 a hydroxyethyl piperazine ionizable amine. The large red dot on the top of the cartoon for 117 refers to hydroxyethyl while the blue thin-line on 116 indicates the methyl group, both attached to piperazine (Figure 60). A combination of 33 IAJDs sSS with 7 IAJDs SS are shown in Figure 60. IAJDs 81, 86, 105, 106 and 107 marked in blue on top left corner of Figure 60 were reported previously (Zhang et al., *J. Am. Chem. Soc.* **2021**, *143*, 17975-17982). Transfection experiments with Luc-mRNA were performed both *in vitro* and *in vivo* by following the methodology reported in Zhang et al., *J. Am. Chem. Soc.* **2021**, *143*, 12315–12327 and Zhang et al., *J. Am. Chem. Soc.* **2021**, *143*, 17975-17982.

The overall transfection activity *in vivo* was analyzed according to its target selectivity and organized in Figure 62. The first important result of the transfection experiments is that 11 IAJDs show approximately 10^8 activity, 2 for lung, 3 for liver and 6 for spleen and lymph (Figure 62 marked in pink, Figure 74). The symmetric IAJDs 110(12/12) and 111(11/11) were previously reported to show the highest activity to lung as IAJDs 33(12/12) and 34(11/11) when they contained an amide interconnecting group. The new IAJDs 110(12/12) and 111(11/11) containing an interconnecting ester rather than amide group show also very high activity to lung. Nonsymmetric IAJDs are stable in serum and PBS buffer and exhibit very high activity in lung by a mechanism

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different from aggregation (Tables shown below and Figure 76 through Figure 78). The highest activity of all IAJDs is for 178(13/18), which displays a total flux luminescence of $4.05x10^8$ p/s, that is 90.2 times higher than of symmetric 99 with the same headgroup (18/18) and only 4.2 times lower than of MC3 (Figure 61). It is also important to mention that the transition from 158(11/17) to 159(11/17), the second an IAJD containing an amide interconnecting group, while the first an ester, increased activity about 6-fold. This demonstrates the significance of amide interconnecting groups for the delivery to lung but simultaneously reveals that the presence of oligooxyethylenes in the hydrophilic part may be important for targeted delivery to lung. These experiments demonstrate the important role of the dissimilar alkyl groups from the hydrophobic part of IAJDs.

Finally, Figure 61 summarizes the activity of all IAJDs made in the examples and compares them with their symmetric (marked in light blue) and nonsymmetric (marked in blue) IAJDs which can be used as control experiments. The results from Figure 61 show that an increase of up to 90.2 times in the activity of the IAJDs was observed by changing their primary structure in the lipophilic part from symmetric to nonsymmetric. Ratios between the two-alkyl length, preferably from odd-even combinations, may be equal or larger than 3 and less than 7 and seem to result in the largest increase in activity. Selected examples of IAJDs supporting this conclusion are provided by IAJDs 119(11/15), 125(11/14), 127(11/17), 128(11/18), 130(11/EH), 153(15/18), 155(11/EH), 159(11/17).

In some embodimetns, the nonsymmetric amphiphilic Janus dendrimer is stable at around 5 °C.

Nanoparticles

In one aspect, the invention relates to nanoparticles comprising at least one amphiphilic Janus dendrimer of the present invention.

In various embodiments, the nanoparticle is a one-component nanoparticle.

In various embodiments, the nanoparticle is a four-component nanoparticle.

In some embodiments, the nanoparticle comprises a homochiral ionizable amphiphilic Janus dendrimer, achiral ionizable amphiphilic Janus dendrimer, or any combination thereof.

In some embodiments, the nanoparticle is a unilamellar nanoparticle.

In some embodiments, the nanoparticle is a multilamellar nanoparticle.

In some embodiments, the nanoparticle is an onion multilamellar nanoparticle.

In some embodiments, the nanoparticle is a racemic ionizable amphiphilic Janus dendrimer.

In some embodiments, the nanoparticle is a dendrimersome nanoparticle (DNP).

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In some embodiments, the nanoparticle comprises at least two amphiphilic Janus dendrimers. Thus, in some embodiments, the the nanoparticle comprises a first ionizable amphiphilic Janus dendrimer and a second ionizable amphiphilic Janus dendrimer. In some embodiments, the first ionizable amphiphilic Janus dendrimer has a different structure than the second ionizable amphiphilic Janus dendrimer.

In some embodiments, the nanoparticles further comprise at least one amphiphilic Janus dendrimer disclosed in Wang, et al., J. Am. Chem. Soc. 2020, 142, 9525−9536; Xiao et al., J. Am. Chem. Soc. 2016, 138, 12655–12663; Torre et al., Proc. Natl. Acad. Sci. U.S.A. 2019, 116, 15378–15385; Percec et al., J. Am. Chem. Soc. 2021, 143, 17724−17743; Wilson et al., J. Polym. Sci. Part A: Polymer Chemistry, 2010, 2498-2508; Xiao et al., Proc. Natl. Acad. Sci. U. S. A. 2017, E7045−E7053; and U.S. Patent Application Publication No. 2012277460; and U.S. Patent No. 8,614,347; each of which is hereby incorporated by reference in their entireties.

In various embodiments, the nanoparticle has a mean diameter of from about 10 nm to about 100,000 nm, about 30 nm to about 1000 nm, about 30 nm to about 500 nm, from about 40 nm to about 150 nm, from about 50 nm to about 150 nm, from about 60 nm to about 130 nm, from about 70 nm to about 110 nm, from about 70 nm to about 100 nm, from about 80 nm to about 100 nm, from about 90 nm to about 100 nm, from about 70 to about 90 nm, from about 80 nm to about 90 nm, from about 70 nm to about 80 nm, or about 30 nm, 35 nm, 40 nm, 45 nm, 50 nm, 55 nm, 60 nm, 65 nm, 70 nm, 75 nm, 80 nm, 85 nm, 90 nm, 95 nm, 100 nm, 105 nm, 110 nm, 115 nm, 120 nm, 125 nm, 130 nm, 135 nm, 140 nm, 145 nm, 150 nm, 160 nm, 170 nm, 200 nm, 250 nm, 300 nm, 310 nm, 375 nm, 400 nm, 500 nm, 800 nm, 1000 nm, 1250 nm, 1400 nm, or 1500 nm. For example, in some embodiments, the the nanoparticle has a mean diameter of from about 10 nm to about 1,000 nm

In various embodiments, the nanoparticle is substantially non-toxic.

In various embodiments, the nanoparticle is biodegradable.

In one aspect of the invention, the nanoparticle comprises at least one cargo. In various aspects, the invention is not limited to any particular cargo or otherwise agent for which the nanoparticle is able to carry or transport. Rather, the invention includes any agent that can be carried by the nanoparticle. For example, agents that can be carried by the nanoparticle of the invention include, but are not limited to, diagnostic agents, detectable agents, and therapeutic agents. Thus, in various embodiments, the nanoparticle comprises at least one agent. In other embodiments, the nanoparticle encapsulates at least one agent.

In some embodiments, the weight ratio of the amphiphilic Janus dendrimer : the at least one

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agent is between about 1 : 1 to about 10,000 : 1. In some embodiments, the nanoparticle comprises, or encapsulates, at least one agent. In some embodiments, the weight ratio of the amphiphilic Janus dendrimer : the at least one agent is between about 2 : 1 to about 1,000 : 1. In some embodiments, the nanoparticle comprises, or encapsulates, at least one agent. In some embodiments, the weight ratio of the amphiphilic Janus dendrimer : the at least one agent is between about 3 : 1 to about 10 : 1. In some embodiments, the nanoparticle comprises, or encapsulates, at least one agent. In some embodiments, the weight ratio of the amphiphilic Janus dendrimer : the at least one agent is between about 4 : 1 to about 10 : 1. In some embodiments, the nanoparticle comprises, or encapsulates, at least one agent. In some embodiments, the weight ratio of the amphiphilic Janus dendrimer : the at least one agent is between about 5 : 1 to about 10 : 1. In some embodiments, the nanoparticle comprises, or encapsulates, at least one agent. In some embodiments, the weight ratio of the amphiphilic Janus dendrimer : the at least one agent is between about 6 : 1 to about 10 : 1. In some embodiments, the nanoparticle comprises, or encapsulates, at least one agent. In some embodiments, the weight ratio of the amphiphilic Janus dendrimer : the at least one agent is between about 7 : 1 to about 10 : 1. In some embodiments, the nanoparticle comprises, or encapsulates, at least one agent. In some embodiments, the weight ratio of the amphiphilic Janus dendrimer : the at least one agent is between about 8 : 1 to about 10 : 1. In some embodiments, the nanoparticle comprises, or encapsulates, at least one agent. In some embodiments, the weight ratio of the amphiphilic Janus dendrimer : the at least one agent is between about 9 : 1 to about 10 : 1. In some embodiments, the nanoparticle comprises, or encapsulates, at least one agent. In some embodiments, the weight ratio of the amphiphilic Janus dendrimer : the at least one agent is between about 9.5 : 1 to about 10 : 1.

In various embodiments, the nanoparticle is suitable for delivering at least one cargo to a cell of interest.

For example, in some embodiments, the cargo is at least one agent comprising a diagnostic agent, detectable agent, therapeutic agent, nucleic acid molecule, gene editing agent, vaccine, composition for protein replacement therapy, or any combination thereof. In some embodiments, the at least one agent is selected from an mRNA, siRNA, microRNA, CRISPR-Cas9, sgRNA, small molecule, protein, antibody, peptide, protein, or any combination thereof. In some embodiments, the at least one agent comprises a nucleic acid molecule. In some embodiments, the nucleic acid molecule encodes at least one selected from an antigen, antibody, gene editing molecule, chimeric antigen receptor (CAR), or any combination thereof. In some embodiments, the nucleic acid molecule is a DNA molecule or an RNA molecule. In some embodiments, the nucleic acid

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molecule is selected from cDNA, cRNA, CirRNA, mRNA, miRNA, siRNA, sgRNA, modified RNA, tRNA, antagomir, antisense molecule, targeted nucleic acid, or any combination thereof. In some embodiments, the modified RNA is a nucleoside-modified RNA. In some embodiments, the nucleoside-modified RNA comprises pseudouridine. In some embodiments, the nucleoside-modified RNA comprises pseudouridine plus 5-methyl-cytosine. In some embodiments, the nucleoside-modified RNA comprises 5-methyl-uridine. In some embodiments, the nucleoside-modified RNA comprises 1-methyl-pseudouridine.

Thus, in one embodiment, the nanoparticles may be used for the delivery of nucleoside-modified RNA to a subject in need thereof. In certain embodiments, delivery of a nucleoside-modified RNA to a subject comprises mixing the nucleoside-modified RNA with at least one dendrimer of Formula (I) prior to the step of contacting. In another embodiment, a method of invention further comprises administering nucleoside-modified RNA together with at least one dendrimer of Formula (I).

In some embodiments, customizable targeting can be achieved based on the identity of the linking group A. Additionally, the identity of the linking group A affects the delivery of the nanoparticle cargo. For an mRNA cargo it was found that an ester linking group resulted in delivery to the liver and/or spleen while an amide group favored delivery to the lungs. This allows the nanoparticle to be tailored to facilitate delivery to the desired target organ. One example of which would be the delivery of anti-inflammatory drugs to the lungs.

In another embodiment, the transfection reagent forms a nanoparticle, which is a liposome. Liposomes, in another embodiment, increase intracellular stability, increase uptake efficiency and improve biological activity. In another embodiment, liposomes are hollow spherical vesicles composed of dendrimers arranged in a similar fashion as those lipids which make up the cell membrane. They have, in another embodiment, an internal aqueous space for entrapping water-soluble compounds and range in size from 0.05 to several microns in diameter. In another embodiment, nanoparticle liposomes can deliver RNA to cells in a biologically active form.

In various embodiments, the nanoparticles have a mean diameter of from about 30 nm to about 150 nm, from about 40 nm to about 150 nm, from about 50 nm to about 150 nm, from about 60 nm to about 130 nm, from about 70 nm to about 110 nm, from about 70 nm to about 100 nm, from about 80 nm to about 100 nm, from about 90 nm to about 100 nm, from about 70 to about 90 nm, from about 80 nm to about 90 nm, from about 70 nm to about 80 nm, or about 30 nm, 35 nm, 40 nm, 45 nm, 50 nm, 55 nm, 60 nm, 65 nm, 70 nm, 75 nm, 80 nm, 85 nm, 90 nm, 95 nm, 100 nm, 105 nm, 110 nm, 115 nm, 120 nm, 125 nm, 130 nm, 135 nm, 140 nm, 145 nm, or 150 nm, and are

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substantially non-toxic. In certain embodiments, the nucleoside-modified RNA, when present in the nanoparticles, is resistant in aqueous solution to degradation with a nuclease.

Small Molecule Therapeutic Agents

In various embodiments, the agent is a therapeutic agent. In various embodiments, the therapeutic agent is a small molecule. When the therapeutic agent is a small molecule, a small molecule may be obtained using standard methods known to the skilled artisan. Such methods include chemical organic synthesis or biological means. Biological means include purification from a biological source, recombinant synthesis and in vitro translation systems, using methods well known in the art. In one embodiment, a small molecule therapeutic agents comprises an organic molecule, inorganic molecule, biomolecule, synthetic molecule, and the like.

Combinatorial libraries of molecularly diverse chemical compounds potentially useful in treating a variety of diseases and conditions are well known in the art, as are method of making the libraries. The method may use a variety of techniques well-known to the skilled artisan including solid phase synthesis, solution methods, parallel synthesis of single compounds, synthesis of chemical mixtures, rigid core structures, flexible linear sequences, deconvolution strategies, tagging techniques, and generating unbiased molecular landscapes for lead discovery vs. biased structures for lead development. In some embodiments of the invention, the therapeutic agent is synthesized and/or identified using combinatorial techniques.

In a general method for small library synthesis, an activated core molecule is condensed with a number of building blocks, resulting in a combinatorial library of covalently linked, core-building block ensembles. The shape and rigidity of the core determines the orientation of the building blocks in shape space. The libraries can be biased by changing the core, linkage, or building blocks to target a characterized biological structure ("focused libraries") or synthesized with less structural bias using flexible cores. In some embodiments of the invention, the therapeutic agent is synthesized via small library synthesis.

The small molecule and small molecule compounds described herein may be present as salts even if salts are not depicted, and it is understood that the invention embraces all salts and solvates of the therapeutic agents depicted here, as well as the non-salt and non-solvate form of the therapeutic agents, as is well understood by the skilled artisan. In some embodiments, the salts of the therapeutic agents of the invention are pharmaceutically acceptable salts.

Where tautomeric forms may be present for any of the therapeutic agents described herein, each and every tautomeric form is intended to be included in the invention, even though only one or

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some of the tautomeric forms may be explicitly depicted. For example, when a 2-hydroxypyridyl moiety is depicted, the corresponding 2-pyridone tautomer is also intended.

The invention also includes any or all of the stereochemical forms, including any enantiomeric or diastereomeric forms of the therapeutic agents described. The recitation of the structure or name herein is intended to embrace all possible stereoisomers of therapeutic agents depicted. All forms of the therapeutic agents are also embraced by the invention, such as crystalline or non-crystalline forms of the therapeutic agent. Compositions comprising a therapeutic agents of the invention are also intended, such as a composition of substantially pure therapeutic agent, including a specific stereochemical form thereof, or a composition comprising mixtures of therapeutic agents of the invention in any ratio, including two or more stereochemical forms, such as in a racemic or non-racemic mixture.

The invention also includes any or all active analog or derivative, such as a prodrug, of any therapeutic agent described herein. In one embodiment, the therapeutic agent is a prodrug. In one embodiment, the small molecules described herein are candidates for derivatization. As such, in certain instances, the analogs of the small molecules described herein that have modulated potency, selectivity, and solubility are included herein and provide useful leads for drug discovery and drug development. Thus, in certain instances, during optimization new analogs are designed considering issues of drug delivery, metabolism, novelty, and safety.

In some instances, small molecule therapeutic agents described herein are derivatives or analogs of known therapeutic agents, as is well known in the art of combinatorial and medicinal chemistry. The analogs or derivatives can be prepared by adding and/or substituting functional groups at various locations. As such, the small molecules described herein can be converted into derivatives/analogs using well known chemical synthesis procedures. For example, all of the hydrogen atoms or substituents can be selectively modified to generate new analogs. Also, the linking atoms or groups can be modified into longer or shorter linkers with carbon backbones or hetero atoms. Also, the ring groups can be changed so as to have a different number of atoms in the ring and/or to include hetero atoms. Moreover, aromatics can be converted to cyclic rings, and vice versa. For example, the rings may be from 5-7 atoms, and may be carbocyclic or heterocyclic.

As used herein, the term "analog," "analogue," or "derivative" is meant to refer to a chemical compound or molecule made from a parent compound or molecule by one or more chemical reactions. As such, an analog can be a structure having a structure similar to that of the small molecule therapeutic agents described herein or can be based on a scaffold of a small molecule therapeutic agents described herein, but differing from it in respect to certain components

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or structural makeup, which may have a similar or opposite action metabolically. An analog or derivative of any of a small molecule inhibitor in accordance with the invention can be used to treat a disease or disorder.

In one embodiment, the small molecule therapeutic agents described herein can independently be derivatized, or analogs prepared therefrom, by modifying hydrogen groups independently from each other into other substituents. That is, each atom on each molecule can be independently modified with respect to the other atoms on the same molecule. Any traditional modification for producing a derivative/analog can be used. For example, the atoms and substituents can be independently comprised of hydrogen, an alkyl, aliphatic, straight chain aliphatic, aliphatic having a chain hetero atom, branched aliphatic, substituted aliphatic, cyclic aliphatic, heterocyclic aliphatic having one or more hetero atoms, aromatic, heteroaromatic, polyaromatic, polyamino acids, peptides, polypeptides, combinations thereof, halogens, halo-substituted aliphatics, and the like. Additionally, any ring group on a compound can be derivatized to increase and/or decrease ring size as well as change the backbone atoms to carbon atoms or hetero atoms.

Nucleic Acid Therapeutic Agents

In other related aspects, the therapeutic agent is an isolated nucleic acid. In certain embodiments, the isolated nucleic acid molecule is one of a DNA molecule or an RNA molecule. In certain embodiments, the isolated nucleic acid molecule is a cDNA, mRNA, siRNA, shRNA or miRNA molecule. In one embodiment, the isolated nucleic acid molecule encodes a therapeutic peptide such a thrombomodulin, endothelial protein C receptor (EPCR), anti-thrombotic proteins including plasminogen activators and their mutants, antioxidant proteins including catalase, superoxide dismutase (SOD) and iron-sequestering proteins. In some embodiments, the therapeutic agent is an siRNA, miRNA, shRNA, or an antisense molecule, which inhibits a targeted nucleic acid including those encoding proteins that are involved in aggravation of the pathological processes.

In one embodiment, the nucleic acid comprises a promoter/regulatory sequence such that the nucleic acid is capable of directing expression of the nucleic acid. Thus, the invention encompasses expression vectors and methods for the introduction of exogenous nucleic acid into cells with concomitant expression of the exogenous nucleic acid in the cells such as those described, for example, in Sambrook et al. (2012, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, New York), and in Ausubel et al. (1997, Current Protocols in Molecular Biology, John Wiley & Sons, New York) and as described elsewhere herein.

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In one embodiment, siRNA is used to decrease the level of a targeted protein. RNA interference (RNAi) is a phenomenon in which the introduction of double-stranded RNA (dsRNA) into a diverse range of organisms and cell types causes degradation of the complementary mRNA. In the cell, long dsRNAs are cleaved into short 21-25 nucleotide small interfering RNAs, or siRNAs, by a ribonuclease known as Dicer. The siRNAs subsequently assemble with protein components into an RNA-induced silencing complex (RISC), unwinding in the process. Activated RISC then binds to complementary transcript by base pairing interactions between the siRNA antisense strand and the mRNA. The bound mRNA is cleaved and sequence specific degradation of mRNA results in gene silencing. See, for example, U.S. Patent No. 6,506,559; Fire et al., 1998, Nature 391(19):306-311; Timmons et al., 1998, Nature 395:854; Montgomery et al., 1998, TIG 14 (7):255-258; David R. Engelke, Ed., RNA Interference (RNAi) Nuts & Bolts of RNAi Technology, DNA Press, Eagleville, PA (2003); and Gregory J. Hannon, Ed., RNAi A Guide to Gene Silencing, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (2003). Soutschek et al. (2004, Nature 432:173-178) describe a chemical modification to siRNAs that aids in intravenous systemic delivery. Optimizing siRNAs involves consideration of overall G/C content, C/T content at the termini, Tm and the nucleotide content of the 3' overhang. See, for instance, Schwartz et al., 2003, Cell, 115:199-208 and Khvorova et al., 2003, Cell 115:209-216. Therefore, the invention also includes methods of decreasing levels of PTPN22 using RNAi technology.

In one aspect, the invention includes a vector comprising an siRNA or an antisense polynucleotide. Preferably, the siRNA or antisense polynucleotide is capable of inhibiting the expression of a target polypeptide. The incorporation of a desired polynucleotide into a vector and the choice of vectors are well-known in the art as described in, for example, Sambrook et al. (2012), and in Ausubel et al. (1997), and elsewhere herein.

In certain embodiments, the expression vectors described herein encode a short hairpin RNA (shRNA) therapeutic agents. shRNA molecules are well known in the art and are directed against the mRNA of a target, thereby decreasing the expression of the target. In certain embodiments, the encoded shRNA is expressed by a cell, and is then processed into siRNA. For example, in certain instances, the cell possesses native enzymes (e.g., dicer) that cleave the shRNA to form siRNA.

In order to assess the expression of the siRNA, shRNA, or antisense polynucleotide, the expression vector to be introduced into a cell can also contain either a selectable marker gene or a reporter gene or both to facilitate identification of expressing cells from the population of cells sought to be transfected or infected using a the delivery vehicle of the invention. In other

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embodiments, the selectable marker may be carried on a separate piece of DNA and also be contained within the delivery vehicle. Both selectable markers and reporter genes may be flanked with appropriate regulatory sequences to enable expression in the host cells. Useful selectable markers are known in the art and include, for example, antibiotic-resistance genes, such as neomycin resistance and the like.

Therefore, in one aspect, the delivery vehicle may contain a vector, comprising the nucleotide sequence or the construct to be delivered. The choice of the vector will depend on the host cell in which it is to be subsequently introduced. In a particular embodiment, the vector of the invention is an expression vector. Suitable host cells include a wide variety of prokaryotic and eukaryotic host cells. In specific embodiments, the expression vector is selected from the group consisting of a viral vector, a bacterial vector and a mammalian cell vector. Prokaryote- and/or eukaryote-vector based systems can be employed for use with the invention to produce polynucleotides, or their cognate polypeptides. Many such systems are commercially and widely available.

By way of illustration, the vector in which the nucleic acid sequence is introduced can be a plasmid, which is or is not integrated in the genome of a host cell when it is introduced in the cell. Illustrative, non-limiting examples of vectors in which the nucleotide sequence of the invention or the gene construct of the invention can be inserted include a tet-on inducible vector for expression in eukaryote cells.

The vector may be obtained by conventional methods known by persons skilled in the art (Sambrook et al., 2012). In a particular embodiment, the vector is a vector useful for transforming animal cells.

In one embodiment, the recombinant expression vectors may also contain nucleic acid molecules, which encode a peptide or peptidomimetic.

A promoter may be one naturally associated with a gene or polynucleotide sequence, as may be obtained by isolating the 5' non-coding sequences located upstream of the coding segment and/or exon. Such a promoter can be referred to as "endogenous." Similarly, an enhancer may be one naturally associated with a polynucleotide sequence, located either downstream or upstream of that sequence. Alternatively, certain advantages will be gained by positioning the coding polynucleotide segment under the control of a recombinant or heterologous promoter, which refers to a promoter that is not normally associated with a polynucleotide sequence in its natural environment. A recombinant or heterologous enhancer refers also to an enhancer not normally associated with a polynucleotide sequence in its natural environment. Such promoters or enhancers

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may include promoters or enhancers of other genes, and promoters or enhancers isolated from any other prokaryotic, viral, or eukaryotic cell, and promoters or enhancers not "naturally occurring," i.e., containing different elements of different transcriptional regulatory regions, and/or mutations that alter expression. In addition to producing nucleic acid sequences of promoters and enhancers synthetically, sequences may be produced using recombinant cloning and/or nucleic acid amplification technology, including PCR™, in connection with the compositions disclosed herein (U.S. Patent 4,683,202, U.S. Patent 5,928,906). Furthermore, it is contemplated the control sequences that direct transcription and/or expression of sequences within non-nuclear organelles such as mitochondria, chloroplasts, and the like, can be employed as well.

Naturally, it will be important to employ a promoter and/or enhancer that effectively directs the expression of the DNA segment in the cell type, organelle, and organism chosen for expression. Those of skill in the art of molecular biology generally know how to use promoters, enhancers, and cell type combinations for protein expression, for example, see Sambrook et al. (2012). The promoters employed may be constitutive, tissue-specific, inducible, and/or useful under the appropriate conditions to direct high level expression of the introduced DNA segment, such as is advantageous in the large-scale production of recombinant proteins and/or peptides. The promoter may be heterologous or endogenous.

The recombinant expression vectors may also contain a selectable marker gene, which facilitates the selection of host cells. Suitable selectable marker genes are genes encoding proteins such as G418 and hygromycin, which confer resistance to certain drugs, β-galactosidase, chloramphenicol acetyltransferase, firefly luciferase, or an immunoglobulin or portion thereof such as the Fc portion of an immunoglobulin preferably IgG. The selectable markers may be introduced on a separate vector from the nucleic acid of interest.

Following the generation of the siRNA polynucleotide, a skilled artisan will understand that the siRNA polynucleotide will have certain characteristics that can be modified to improve the siRNA as a therapeutic compound. Therefore, the siRNA polynucleotide may be further designed to resist degradation by modifying it to include phosphorothioate, or other linkages, methylphosphonate, sulfone, sulfate, ketyl, phosphorodithioate, phosphoramidate, phosphate esters, and the like (see, e.g., Agrawal et al., 1987, Tetrahedron Lett. 28:3539-3542; Stec et al., 1985 Tetrahedron Lett. 26:2191-2194; Moody et al., 1989 Nucleic Acids Res. 12:4769-4782; Eckstein, 1989 Trends Biol. Sci. 14:97-100; Stein, In: Oligodeoxynucleotides. Antisense Inhibitors of Gene Expression, Cohen, ed., Macmillan Press, London, pp. 97-117 (1989)).

Any polynucleotide may be further modified to increase its stability in vivo. Possible

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modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends; the use of phosphorothioate or 2' O-methyl rather than phosphodiester linkages in the backbone; and/or the inclusion of nontraditional bases such as inosine, queuosine, and wybutosine and the like, as well as acetyl- methyl-, thio- and other modified forms of adenine, cytidine, guanine, thymine, and uridine.

In one embodiment of the invention, an antisense nucleic acid sequence, which is expressed by a plasmid vector is used as a therapeutic agent to inhibit the expression of a target protein. The antisense expressing vector is used to transfect a mammalian cell or the mammal itself, thereby causing reduced endogenous expression of the target protein.

Antisense molecules and their use for inhibiting gene expression are well known in the art (see, e.g., Cohen, 1989, In: Oligodeoxyribonucleotides, Antisense Inhibitors of Gene Expression, CRC Press). Antisense nucleic acids are DNA or RNA molecules that are complementary, as that term is defined elsewhere herein, to at least a portion of a specific mRNA molecule (Weintraub, 1990, Scientific American 262:40). In the cell, antisense nucleic acids hybridize to the corresponding mRNA, forming a double-stranded molecule thereby inhibiting the translation of genes.

The use of antisense methods to inhibit the translation of genes is known in the art, and is described, for example, in Marcus-Sakura (1988, Anal. Biochem. 172:289). Such antisense molecules may be provided to the cell via genetic expression using DNA encoding the antisense molecule as taught by Inoue, 1993, U.S. Patent No. 5,190,931.

Alternatively, antisense molecules of the invention may be made synthetically and then provided to the cell. Antisense oligomers of between about 10 to about 30, and more preferably about 15 nucleotides, are preferred, since they are easily synthesized and introduced into a target cell. Synthetic antisense molecules contemplated by the invention include oligonucleotide derivatives known in the art which have improved biological activity compared to unmodified oligonucleotides (see U.S. Patent No. 5,023,243).

In one embodiment of the invention, a ribozyme is used as a therapeutic agent to inhibit expression of a target protein. Ribozymes useful for inhibiting the expression of a target molecule may be designed by incorporating target sequences into the basic ribozyme structure, which are complementary, for example, to the mRNA sequence encoding the target molecule. Ribozymes targeting the target molecule, may be synthesized using commercially available reagents (Applied Biosystems, Inc., Foster City, CA) or they may be genetically expressed from DNA encoding them.

In one embodiment, the therapeutic agent may comprise one or more components of a

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CRISPR-Cas system, where a guide RNA (gRNA) targeted to a gene encoding a target molecule, and a CRISPR-associated (Cas) peptide form a complex to induce mutations within the targeted gene. In one embodiment, the therapeutic agent comprises a gRNA or a nucleic acid molecule encoding a gRNA. In one embodiment, the therapeutic agent comprises a Cas peptide or a nucleic acid molecule encoding a Cas peptide.

In one embodiment, the agent comprises a miRNA or a mimic of a miRNA. In one embodiment, the agent comprises a nucleic acid molecule that encodes a miRNA or mimic of a miRNA.

MiRNAs are small non-coding RNA molecules that are capable of causing post-transcriptional silencing of specific genes in cells by the inhibition of translation or through degradation of the targeted mRNA. A miRNA can be completely complementary or can have a region of noncomplementarity with a target nucleic acid, consequently resulting in a "bulge" at the region of non-complementarity. A miRNA can inhibit gene expression by repressing translation, such as when the miRNA is not completely complementary to the target nucleic acid, or by causing target RNA degradation, which is believed to occur only when the miRNA binds its target with perfect complementarity. The disclosure also can include double-stranded precursors of miRNA. A miRNA or pri-miRNA can be 18- 100 nucleotides in length, or from 18-80 nucleotides in length. Mature miRNAs can have a length of 19-30 nucleotides, or 21-25 nucleotides, particularly 21, 22, 23, 24, or 25 nucleotides. MiRNA precursors typically have a length of about 70-100 nucleotides and have a hairpin conformation. miRNAs are generated in vivo from pre- miRNAs by the enzymes Dicer and Drosha, which specifically process long pre-miRNA into functional miRNA. The hairpin or mature microRNAs, or pri-microRNA agents featured in the disclosure can be synthesized in vivo by a cell-based system or in vitro by chemical synthesis.

In various embodiments, the agent comprises an oligonucleotide that comprises the nucleotide sequence of a disease-associated miRNA. In certain embodiments, the oligonucleotide comprises the nucleotide sequence of a disease-associated miRNA in a pre -microRNA, mature or hairpin form. In other embodiments, a combination of oligonucleotides comprising a sequence of one or more disease-associated miRNAs, any pre -miRNA, any fragment, or any combination thereof is envisioned.

MiRNAs can be synthesized to include a modification that imparts a desired characteristic. For example, the modification can improve stability, hybridization thermodynamics with a target nucleic acid, targeting to a particular tissue or cell -type, or cell permeability, e.g., by an endocytosis-dependent or -independent mechanism.

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Modifications can also increase sequence specificity, and consequently decrease off-site targeting. Methods of synthesis and chemical modifications are described in greater detail below. If desired, miRNA molecules may be modified to stabilize the miRNAs against degradation, to enhance half-life, or to otherwise improve efficacy. Desirable modifications are described, for example, in U.S. Patent Publication Nos. 20070213292, 20060287260, 20060035254. 20060008822. and 2005028824, each of which is hereby incorporated by reference in its entirety. For increased nuclease resistance and/or binding affinity to the target, the single- stranded oligonucleotide agents featured in the disclosure can include 2'-O-methyl, 2'-fluorine, 2'-O-methoxyethyl, 2'-O-aminopropyl, 2'-amino, and/or phosphorothioate linkages. Inclusion of locked nucleic acids (LNA), ethylene nucleic acids (ENA), e.g., 2'-4'-ethylene- bridged nucleic acids, and certain nucleotide modifications can also increase binding affinity to the target. The inclusion of pyranose sugars in the oligonucleotide backbone can also decrease endonucleolytic cleavage. An oligonucleotide can be further modified by including a 3' cationic group, or by inverting the nucleoside at the 3'-terminus with a 3 -3' linkage. In another alternative, the 3 '-terminus can be blocked with an aminoalkyl group. Other 3' conjugates can inhibit 3'-5' exonucleolytic cleavage. While not being bound by theory, a 3' may inhibit exonucleolytic cleavage by sterically blocking the exonuclease from binding to the 3' end of the oligonucleotide. Even small alkyl chains, aryl groups, or heterocyclic conjugates or modified sugars (D-ribose, deoxyribose, glucose etc.) can block 3'-5'-exonucleases.

In one embodiment, the miRNA includes a 2'-modified oligonucleotide containing oligodeoxynucleotide gaps with some or all internucleotide linkages modified to phosphorothioates for nuclease resistance. The presence of methylphosphonate modifications increases the affinity of the oligonucleotide for its target RNA and thus reduces the IC_5Q. This modification also increases the nuclease resistance of the modified oligonucleotide. It is understood that the methods and reagents of the present disclosure may be used in conjunction with any technologies that may be developed to enhance the stability or efficacy of an inhibitory nucleic acid molecule.

miRNA molecules include nucleotide oligomers containing modified backbones or non-natural internucleoside linkages. Oligomers having modified backbones include those that retain a phosphorus atom in the backbone and those that do not have a phosphorus atom in the backbone. For the purposes of this disclosure, modified oligonucleotides that do not have a phosphorus atom in their internucleoside backbone are also considered to be nucleotide oligomers. Nucleotide oligomers that have modified oligonucleotide backbones include, for example, phosphorothioates, chiral phosphorothioates, phosphorodithioates, phosphotriesters, aminoalkyl-phosphotriesters,

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methyl and other alkyl phosphonates including 3'-alkylene phosphonates and chiral phosphonates, phosphinates, phosphoramidates, thionophosphoramidates, thionoalkylphosphonates, thionoalkylphosphotriest- ers, and boranophosphates. Various salts, mixed salts and free acid forms are also included.

A miRNA described herein, which may be in the mature or hairpin form, may be provided as a naked oligonucleotide. In some cases, it may be desirable to utilize a formulation that aids in the delivery of a miRNA or other nucleotide oligomer to cells (see, e.g., U.S. Pat. Nos. 5,656,61 1, 5,753,613, 5,785,992, 6,120,798, 6,221,959, 6,346,613, and 6,353,055, each of which is hereby incorporated by reference).

In some examples, the miRNA composition is at least partially crystalline, uniformly crystalline, and/or anhydrous (e.g., less than 80, 50, 30, 20, or 10% water). In another example, the miRNA composition is in an aqueous phase, e.g., in a solution that includes water. The aqueous phase or the crystalline compositions can be incorporated into a delivery vehicle, e.g., a liposome (particularly for the aqueous phase), or a particle (e.g., a microparticle as can be appropriate for a crystalline composition). Generally, the miRNA composition is formulated in a manner that is compatible with the intended method of administration. A miRNA composition can be formulated in combination with another agent, e.g., another therapeutic agent or an agent that stabilizes an oligonucleotide agent, e.g., a protein that complexes with the oligonucleotide agent. Still other agents include chelators, e.g., EDTA (e.g., to remove divalent cations such as Mg), salts, and RNAse inhibitors (e.g., a broad specificity RNAse inhibitor). In one embodiment, the miRNA composition includes another miRNA, e.g., a second miRNA composition (e.g., a microRNA that is distinct from the first). Still other preparations can include at least three, five, ten, twenty, fifty, or a hundred or more different oligonucleotide species.

In certain embodiments, the composition comprises an oligonucleotide composition that mimics the activity of a miRNA. In certain embodiments, the composition comprises oligonucleotides having nucleobase identity to the nucleobase sequence of a miRNA, and are thus designed to mimic the activity of the miRNA. In certain embodiments, the oligonucleotide composition that mimics miRNA activity comprises a double-stranded RNA molecule which mimics the mature miRNA hairpins or processed miRNA duplexes.

In one embodiment, the oligonucleotide shares identity with endogenous miRNA or miRNA precursor nucleobase sequences. An oligonucleotide selected for inclusion in a composition of the invention may be one of a number of lengths. Such an oligonucleotide can be from 7 to 100 linked nucleosides in length. For example, an oligonucleotide sharing nucleobase

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identity with a miRNA may be from 7 to 30 linked nucleosides in length. An oligonucleotide sharing identity with a miRNA precursor may be up to 100 linked nucleosides in length. In certain embodiments, an oligonucleotide comprises 7 to 30 linked nucleosides. In certain embodiments, an oligonucleotide comprises 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 28, 29, or 30 linked nucleotides. In certain embodiments, an oligonucleotide comprises 19 to 23 linked nucleosides. In certain embodiments, an oligonucleotide is from 40 up to 50, 60, 70, 80, 90, or 100 linked nucleosides in length.

In certain embodiments, an oligonucleotide has a sequence that has a certain identity to a miRNA or a precursor thereof. Nucleobase sequences of mature miRNAs and their corresponding stem-loop sequences described herein are the sequences found in miRBase, an online searchable database of miRNA sequences and annotation. Entries in the miRBase Sequence database represent a predicted hairpin portion of a miRNA transcript (the stem-loop), with information on the location and sequence of the mature miRNA sequence. The miRNA stem-loop sequences in the database are not strictly precursor miRNAs (pre-miRNAs), and may in some instances include the pre-miRNA and some flanking sequence from the presumed primary transcript. The miRNA nucleobase sequences described herein encompass any version of the miRNA, including the sequences described in Release 10.0 of the miRBase sequence database and sequences described in any earlier Release of the miRBase sequence database. A sequence database release may result in the re-naming of certain miRNAs. A sequence database release may result in a variation of a mature miRNA sequence. The compositions of the invention encompass oligomeric compound comprising oligonucleotides having a certain identity to any nucleobase sequence version of a miRNAs described herein.

In certain embodiments, an oligonucleotide has a nucleobase sequence at least 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 97%, 98% or 99% identical to the miRNA over a region of 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, or 30 nucleobases. Accordingly, in certain embodiments the nucleobase sequence of an oligonucleotide may have one or more non-identical nucleobases with respect to the miRNA.

In certain embodiments, the composition comprises a nucleic acid molecule encoding a miRNA, precursor, mimic, or fragment thereof. For example, the composition may comprise a viral vector, plasmid, cosmid, or other expression vector suitable for expressing the miRNA, precursor, mimic, or fragment thereof in a desired mammalian cell or tissue.

Nucleic Acids

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In one embodiment, the invention includes a nanoparticle comprising or encapsulating one or more nucleic acid molecule. In one embodiment, the nucleic acid molecule is a nucleoside-modified mRNA molecule. In one embodiment, the nucleoside-modified mRNA molecule encodes an antigen. In one embodiment, the nucleoside-modified mRNA molecule encodes a plurality of antigens. In certain embodiments, the nucleoside-modified mRNA molecule encodes an antigen that induces an adaptive immune response against the antigen. In one embodiment, the invention includes a nucleoside-modified mRNA molecule encoding an adjuvant.

The nucleotide sequences encoding an antigen or adjuvant, as described herein, can alternatively comprise sequence variations with respect to the original nucleotide sequences, for example, substitutions, insertions and/or deletions of one or more nucleotides, with the condition that the resulting polynucleotide encodes a polypeptide according to the invention. Therefore, the scope of the invention includes nucleotide sequences that are substantially homologous to the nucleotide sequences recited herein and encode an antigen or adjuvant of interest.

As used herein, a nucleotide sequence is "substantially homologous" to any of the nucleotide sequences described herein when its nucleotide sequence has a degree of identity with respect to the nucleotide sequence of at least 60%, advantageously of at least 70%, preferably of at least 85%, and more preferably of at least 95%. A nucleotide sequence that is substantially homologous to a nucleotide sequence encoding an antigen can typically be isolated from a producer organism of the antigen based on the information contained in the nucleotide sequence by means of introducing conservative or non-conservative substitutions, for example. Other examples of possible modifications include the insertion of one or more nucleotides in the sequence, the addition of one or more nucleotides in any of the ends of the sequence, or the deletion of one or more nucleotides in any end or inside the sequence. The degree of identity between two polynucleotides is determined using computer algorithms and methods that are widely known for the persons skilled in the art.

Further, the scope of the invention includes nucleotide sequences that encode amino acid sequences that are substantially homologous to the amino acid sequences recited herein and preserve the immunogenic function of the original amino acid sequence.

As used herein, an amino acid sequence is "substantially homologous" to any of the amino acid sequences described herein when its amino acid sequence has a degree of identity with respect to the amino acid sequence of at least 60%, advantageously of at least 70%, preferably of at least 85%, and more preferably of at least 95%. The identity between two amino acid sequences is preferably determined by using the BLASTN algorithm (BLAST Manual, Altschul, S., et al., NCBI

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NLM NIH Bethesda, Md. 20894, Altschul, S., et al., J. Mol. Biol. 215: 403-410 (1990)).

In one embodiment, the invention relates to a construct, comprising a nucleotide sequence encoding an antigen. In one embodiment, the construct comprises a plurality of nucleotide sequences encoding a plurality of antigens. For example, in certain embodiments, the construct encodes 1 or more, 2 or more, 5 or more, 10 or more, 15 or more, or 20 or more antigens. In one embodiment, the invention relates to a construct, comprising a nucleotide sequence encoding an adjuvant. In one embodiment, the construct comprises a first nucleotide sequence encoding an antigen and a second nucleotide sequence encoding an adjuvant.

In one embodiment, the composition comprises a plurality of constructs, each construct encoding one or more antigens. In certain embodiments, the composition comprises 1 or more, 2 or more, 5 or more, 10 or more, 15 or more, or 20 or more constructs. In one embodiment, the composition comprises a first construct, comprising a nucleotide sequence encoding an antigen; and a second construct, comprising a nucleotide sequence encoding an adjuvant.

In another particular embodiment, the construct is operatively bound to a translational control element. The construct can incorporate an operatively bound regulatory sequence for the expression of the nucleotide sequence of the invention, thus forming an expression cassette.

Vectors

The nucleic acid sequences encapsulated in the nanoparticle of the invention can be obtained using recombinant methods known in the art, such as, for example by screening libraries from cells expressing the gene, by deriving the gene from a vector known to include the same, or by isolating directly from cells and tissues containing the same, using standard techniques. Alternatively, the nucleic acid molecule of interest can be produced synthetically.

The nucleic acid can be cloned into a number of types of vectors. For example, the nucleic acid can be cloned into a vector including, but not limited to a plasmid, a phagemid, a phage derivative, an animal virus, and a cosmid. Vectors of particular interest include expression vectors, replication vectors, probe generation vectors, sequencing vectors and vectors optimized for in vitro transcription.

In one embodiment, the composition of the invention comprises in vitro transcribed (IVT) RNA encoding an antigen. In one embodiment, the composition of the invention comprises IVT RNA encoding a plurality of antigens. In one embodiment, the composition of the invention comprises IVT RNA encoding an adjuvant. In one embodiment, the composition of the invention comprises IVT RNA encoding one or more antigens and one or more adjuvants.

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Nucleoside-Modified RNA

In one embodiment, the nucleic acid molecule comprises a nucleoside-modified RNA. Nucleoside-modified mRNA have particular advantages over non-modified mRNA, including for example, increased stability, low or absent innate immunogenicity, and enhanced translation. Nucleoside-modified mRNA useful in the invention is further described in U.S. Patent No. 8,278,036, which is incorporated by reference herein in its entirety.

In certain embodiments, nucleoside-modified mRNA does not activate any pathophysiologic pathways, translates very efficiently and almost immediately following delivery, and serve as templates for continuous protein production in vivo lasting for several days (Karikó et al., 2008, Mol Ther 16:1833-1840; Karikó et al., 2012, Mol Ther 20:948-953). The amount of mRNA required to exert a physiological effect is small and that makes it applicable for human therapy. For example, as described herein, nucleoside-modified mRNA encoding an antigen has demonstrated the ability to induce CD4+ and CD8+ T-cell and antigen-specific antibody production. For example, in certain instances, antigen encoded by nucleoside-modified mRNA induces greater production of antigen-specific antibody production as compared to antigen encoded by non-modified mRNA.

In certain instances, expressing a protein by delivering the encoding mRNA has many benefits over methods that use protein, plasmid DNA or viral vectors. During mRNA transfection, the coding sequence of the desired protein is the only substance delivered to cells, thus avoiding all the side effects associated with plasmid backbones, viral genes, and viral proteins. More importantly, unlike DNA- and viral-based vectors, the mRNA does not carry the risk of being incorporated into the genome and protein production starts immediately after mRNA delivery. For example, high levels of circulating proteins have been measured within 15 to 30 minutes of in vivo injection of the encoding mRNA. In certain embodiments, using mRNA rather than the protein also has many advantages. Half-lives of proteins in the circulation are often short, thus protein treatment would need frequent dosing, while mRNA provides a template for continuous protein production for several days. Purification of proteins is problematic and they can contain aggregates and other impurities that cause adverse effects (Kromminga and Schellekens, 2005, Ann NY Acad Sci 1050:257-265).

In certain embodiments, the nucleoside-modified RNA comprises the naturally occurring modified-nucleoside pseudouridine. In certain embodiments, inclusion of pseudouridine makes the mRNA more stable, non-immunogenic, and highly translatable (Karikó et al., 2008, Mol Ther

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16:1833-1840; Anderson et al., 2010, Nucleic Acids Res 38:5884-5892; Anderson et al., 2011, Nucleic Acids Research 39:9329-9338; Karikó et al., 2011, Nucleic Acids Research 39:e142; Karikó et al., 2012, Mol Ther 20:948-953; Karikó et al., 2005, Immunity 23:165-175).

It has been demonstrated that the presence of modified nucleosides, including pseudouridines in RNA suppress their innate immunogenicity (Karikó et al., 2005, Immunity 23:165-175). Further, protein-encoding, in vitro-transcribed RNA containing pseudouridine can be translated more efficiently than RNA containing no or other modified nucleosides (Karikó et al., 2008, Mol Ther 16:1833-1840). Subsequently, it is shown that the presence of pseudouridine improves the stability of RNA (Anderson et al., 2011, Nucleic Acids Research 39:9329-9338) and abates both activation of PKR and inhibition of translation (Anderson et al., 2010, Nucleic Acids Res 38:5884-5892). A preparative HPLC purification procedure has been established that was critical to obtain pseudouridine-containing RNA that has superior translational potential and no innate immunogenicity (Karikó et al., 2011, Nucleic Acids Research 39:e142). Administering HPLC-purified, pseudourine-containing RNA coding for erythropoietin into mice and macaques resulted in a significant increase of serum EPO levels (Karikó et al., 2012, Mol Ther 20:948-953), thus confirming that pseudouridine-containing mRNA is suitable for in vivo protein therapy.

The invention encompasses RNA, oligoribonucleotide, and polyribonucleotide molecules comprising pseudouridine or a modified nucleoside. In certain embodiments, the composition comprises an isolated nucleic acid encoding an antigen, wherein the nucleic acid comprises a pseudouridine or a modified nucleoside. In certain embodiments, the composition comprises a vector, comprising an isolated nucleic acid encoding an antigen, adjuvant, or combination thereof, wherein the nucleic acid comprises a pseudouridine or a modified nucleoside.

In one embodiment, the nucleoside-modified RNA of the invention is IVT RNA. For example, in certain embodiments, the nucleoside-modified RNA is synthesized by T7 phage RNA polymerase. In another embodiment, the nucleoside-modified mRNA is synthesized by SP6 phage RNA polymerase. In another embodiment, the nucleoside-modified RNA is synthesized by T3 phage RNA polymerase.

In one embodiment, the modified nucleoside is $m^1acp^3\Psi$ (1-methyl-3-(3-amino-3-carboxypropyl) pseudouridine. In another embodiment, the modified nucleoside is $m^1\Psi$ (1-methylpseudouridine). In another embodiment, the modified nucleoside is Ψm (2'-O-methylpseudouridine. In another embodiment, the modified nucleoside is m^5D (5-methyldihydrouridine). In another embodiment, the modified nucleoside is $m^3\Psi$ (3-methylpseudouridine). In another embodiment, the modified nucleoside is a pseudouridine moiety

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that is not further modified. In another embodiment, the modified nucleoside is a monophosphate, diphosphate, or triphosphate of any of the above pseudouridines. In another embodiment, the modified nucleoside is any other pseudouridine-like nucleoside known in the art.

In another embodiment, the nucleoside that is modified in the nucleoside-modified RNA the invention is uridine (U). In another embodiment, the modified nucleoside is cytidine (C). In another embodiment, the modified nucleoside is adenosine (A). In another embodiment the modified nucleoside is guanosine (G).

In another embodiment, the modified nucleoside of the invention is m^5C (5-methylcytidine). In another embodiment, the modified nucleoside is m^5U (5-methyluridine). In another embodiment, the modified nucleoside is m^6A (N^6-methyladenosine). In another embodiment, the modified nucleoside is s^2U (2-thiouridine). In another embodiment, the modified nucleoside is Ψ (pseudouridine). In another embodiment, the modified nucleoside is Um (2'-O-methyluridine).

In other embodiments, the modified nucleoside is m^1A (1-methyladenosine); m^2A (2-methyladenosine); Am (2'-O-methyladenosine); ms^2m^6A (2-methylthio-N^6-methyladenosine); i^6A (N^6-isopentenyladenosine); ms^2i6A (2-methylthio-N^6isopentenyladenosine); io^6A (N^6-(cis-hydroxyisopentenyl)adenosine); ms^2io^6A (2-methylthio-N^6-(cis-hydroxyisopentenyl) adenosine); g^6A (N^6-glycinylcarbamoyladenosine); t^6A (N^6-threonylcarbamoyladenosine); ms^2t^6A (2-methylthio-N^6-threonyl carbamoyladenosine); m^6t^6A (N^6-methyl-N^6-threonylcarbamoyladenosine); hn^6A(N^6-hydroxynorvalylcarbamoyladenosine); ms^2hn^6A (2-methylthio-N^6-hydroxynorvalyl carbamoyladenosine); Ar(p) (2'-O-ribosyladenosine (phosphate)); I (inosine); m^1I (1-methylinosine); m^1Im (1,2'-O-dimethylinosine); m^3C (3-methylcytidine); Cm (2'-O-methylcytidine); s^2C (2-thiocytidine); ac^4C (N^4-acetylcytidine); f^5C (5-formylcytidine); m^5Cm (5,2'-O-dimethylcytidine); ac^4Cm (N^4-acetyl-2'-O-methylcytidine); k^2C (lysidine); m^1G (1-methylguanosine); m^2G (N^2-methylguanosine); m^7G (7-methylguanosine); Gm (2'-O-methylguanosine); $m^2{}_2G$ (N^2,N^2-dimethylguanosine); m^2Gm (N^2,2'-O-dimethylguanosine); $m^2{}_2Gm$ (N^2,N^2,2'-O-trimethylguanosine); Gr(p) (2'-O-ribosylguanosine (phosphate)); yW (wybutosine); o_2yW (peroxywybutosine); OHyW (hydroxywybutosine); OHyW* (undermodified hydroxywybutosine); imG (wyosine); mimG (methylwyosine); Q (queuosine); oQ (epoxyqueuosine); galQ (galactosyl-queuosine); manQ (mannosyl-queuosine); $preQ_0$ (7-cyano-7-deazaguanosine); $preQ_1$ (7-aminomethyl-7-deazaguanosine); G^1 (archaeosine); D (dihydrouridine); m^5Um (5,2'-O-dimethyluridine); s^4U (4-thiouridine); m^5s^2U (5-methyl-2-thiouridine); s^2Um (2-thio-2'-O-methyluridine); acp^3U (3-(3-amino-3-carboxypropyl)uridine); ho^5U (5-hydroxyuridine); mo^5U

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(5-methoxyuridine); cmo^5U (uridine 5-oxyacetic acid); $mcmo^5U$ (uridine 5-oxyacetic acid methyl ester); chm^5U (5-(carboxyhydroxymethyl)uridine)); $mchm^5U$ (5-(carboxyhydroxymethyl)uridine methyl ester); mcm^5U (5-methoxycarbonylmethyluridine); mcm^5Um (5-methoxycarbonylmethyl-2'-O-methyluridine); mcm^5s^2U (5-methoxycarbonylmethyl-2-thiouridine); nm^5s^2U (5-aminomethyl-2-thiouridine); mnm^5U (5-methylaminomethyluridine); mnm^5s^2U (5-methylaminomethyl-2-thiouridine); mnm^5se^2U (5-methylaminomethyl-2-selenouridine); ncm^5U (5-carbamoylmethyluridine); ncm^5Um (5-carbamoylmethyl-2'-O-methyluridine); $cmnm^5U$ (5-carboxymethylaminomethyluridine); $cmnm^5Um$ (5-carboxymethylaminomethyl-2'-O-methyluridine); $cmnm^5s^2U$ (5-carboxymethylaminomethyl-2-thiouridine); $m^6{}_2A$ (N^6,N^6-dimethyladenosine); Im (2'-O-methylinosine); m^4C (N^4-methylcytidine); m^4Cm (N^4,2'-O-dimethylcytidine); hm^5C (5-hydroxymethylcytidine); m^3U (3-methyluridine); cm^5U (5-carboxymethyluridine); m^6Am (N^6,2'-O-dimethyladenosine); $m^6{}_2Am$ (N^6,N^6,O-2'-trimethyladenosine); $m^{2,7}G$ (N^2,7-dimethylguanosine); $m^{2,2,7}G$ (N^2,N^2,7-trimethylguanosine); m^3Um (3,2'-O-dimethyluridine); m^5D (5-methyldihydrouridine); f^5Cm (5-formyl-2'-O-methylcytidine); m^1Gm (1,2'-O-dimethylguanosine); m^1Am (1,2'-O-dimethyladenosine); τm^5U (5-taurinomethyluridine); τm^5s^2U (5-taurinomethyl-2-thiouridine)); imG-14 (4-demethylwyosine); imG2 (isowyosine); or ac^6A (N^6-acetyladenosine).

In another embodiment, a nucleoside-modified RNA of the invention comprises a combination of 2 or more of the above modifications. In another embodiment, the nucleoside-modified RNA comprises a combination of 3 or more of the above modifications. In another embodiment, the nucleoside-modified RNA comprises a combination of more than 3 of the above modifications.

In another embodiment, between 0.1% and 100% of the residues in the nucleoside-modified of the invention are modified (e.g. either by the presence of pseudouridine or a modified nucleoside base). In another embodiment, 0.1% of the residues are modified. In another embodiment, the fraction of modified residues is 0.2%. In another embodiment, the fraction is 0.3%. In another embodiment, the fraction is 0.4%. In another embodiment, the fraction is 0.5%. In another embodiment, the fraction is 0.6%. In another embodiment, the fraction is 0.8%. In another embodiment, the fraction is 1%. In another embodiment, the fraction is 1.5%. In another embodiment, the fraction is 2%. In another embodiment, the fraction is 2.5%. In another embodiment, the fraction is 3%. In another embodiment, the fraction is 4%. In another embodiment, the fraction is 5%. In another embodiment, the fraction is 6%. In another embodiment, the fraction is 8%. In another embodiment, the fraction is 10%. In another

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embodiment, the fraction is 12%. In another embodiment, the fraction is 14%. In another embodiment, the fraction is 16%. In another embodiment, the fraction is 18%. In another embodiment, the fraction is 20%. In another embodiment, the fraction is 25%. In another embodiment, the fraction is 30%. In another embodiment, the fraction is 35%. In another embodiment, the fraction is 40%. In another embodiment, the fraction is 45%. In another embodiment, the fraction is 50%. In another embodiment, the fraction is 60%. In another embodiment, the fraction is 70%. In another embodiment, the fraction is 80%. In another embodiment, the fraction is 90%. In another embodiment, the fraction is 100%.

In another embodiment, the fraction is less than 5%. In another embodiment, the fraction is less than 3%. In another embodiment, the fraction is less than 1%. In another embodiment, the fraction is less than 2%. In another embodiment, the fraction is less than 4%. In another embodiment, the fraction is less than 6%. In another embodiment, the fraction is less than 8%. In another embodiment, the fraction is less than 10%. In another embodiment, the fraction is less than 12%. In another embodiment, the fraction is less than 15%. In another embodiment, the fraction is less than 20%. In another embodiment, the fraction is less than 30%. In another embodiment, the fraction is less than 40%. In another embodiment, the fraction is less than 50%. In another embodiment, the fraction is less than 60%. In another embodiment, the fraction is less than 70%.

In another embodiment, 0.1% of the residues of a given nucleoside (i.e., uridine, cytidine, guanosine, or adenosine) are modified. In another embodiment, the fraction of the given nucleotide that is modified is 0.2%. In another embodiment, the fraction is 0.3%. In another embodiment, the fraction is 0.4%. In another embodiment, the fraction is 0.5%. In another embodiment, the fraction is 0.6%. In another embodiment, the fraction is 0.8%. In another embodiment, the fraction is 1%. In another embodiment, the fraction is 1.5%. In another embodiment, the fraction is 2%. In another embodiment, the fraction is 2.5%. In another embodiment, the fraction is 3%. In another embodiment, the fraction is 4%. In another embodiment, the fraction is 5%. In another embodiment, the fraction is 6%. In another embodiment, the fraction is 8%. In another embodiment, the fraction is 10%. In another embodiment, the fraction is 12%. In another embodiment, the fraction is 14%. In another embodiment, the fraction is 16%. In another embodiment, the fraction is 18%. In another embodiment, the fraction is 20%. In another embodiment, the fraction is 25%. In another embodiment, the fraction is 30%. In another embodiment, the fraction is 35%. In another embodiment, the fraction is 40%. In another embodiment, the fraction is 45%. In another embodiment, the fraction is 50%. In another embodiment, the fraction is 60%. In another embodiment, the fraction is 70%. In another

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embodiment, the fraction is 80%. In another embodiment, the fraction is 90%. In another embodiment, the fraction is 100%.

In another embodiment, the fraction of the given nucleotide that is modified is less than 8%. In another embodiment, the fraction is less than 10%. In another embodiment, the fraction is less than 5%. In another embodiment, the fraction is less than 3%. In another embodiment, the fraction is less than 1%. In another embodiment, the fraction is less than 2%. In another embodiment, the fraction is less than 4%. In another embodiment, the fraction is less than 6%. In another embodiment, the fraction is less than 12%. In another embodiment, the fraction is less than 15%. In another embodiment, the fraction is less than 20%. In another embodiment, the fraction is less than 30%. In another embodiment, the fraction is less than 40%. In another embodiment, the fraction is less than 50%. In another embodiment, the fraction is less than 60%. In another embodiment, the fraction is less than 70%.

In another embodiment, a nucleoside-modified RNA of the invention is translated in the cell more efficiently than an unmodified RNA molecule with the same sequence. In another embodiment, the nucleoside-modified RNA exhibits enhanced ability to be translated by a target cell. In another embodiment, translation is enhanced by a factor of 2-fold relative to its unmodified counterpart. In another embodiment, translation is enhanced by a 3-fold factor. In another embodiment, translation is enhanced by a 5-fold factor. In another embodiment, translation is enhanced by a 7-fold factor. In another embodiment, translation is enhanced by a 10-fold factor. In another embodiment, translation is enhanced by a 15-fold factor. In another embodiment, translation is enhanced by a 20-fold factor. In another embodiment, translation is enhanced by a 50-fold factor. In another embodiment, translation is enhanced by a 100-fold factor. In another embodiment, translation is enhanced by a 200-fold factor. In another embodiment, translation is enhanced by a 500-fold factor. In another embodiment, translation is enhanced by a 1000-fold factor. In another embodiment, translation is enhanced by a 2000-fold factor. In another embodiment, the factor is 10-1000-fold. In another embodiment, the factor is 10-100-fold. In another embodiment, the factor is 10-200-fold. In another embodiment, the factor is 10-300-fold. In another embodiment, the factor is 10-500-fold. In another embodiment, the factor is 20-1000-fold. In another embodiment, the factor is 30-1000-fold. In another embodiment, the factor is 50-1000-fold. In another embodiment, the factor is 100-1000-fold. In another embodiment, the factor is 200-1000-fold. In another embodiment, translation is enhanced by any other significant amount or range of amounts.

In another embodiment, the nucleoside-modified antigen-encoding RNA of the invention

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induces significantly more adaptive immune response than an unmodified in vitro-synthesized RNA molecule with the same sequence. In another embodiment, the modified RNA molecule exhibits an adaptive immune response that is 2-fold greater than its unmodified counterpart. In another embodiment, the adaptive immune response is increased by a 3-fold factor. In another embodiment the adaptive immune response is increased by a 5-fold factor. In another embodiment, the adaptive immune response is increased by a 7-fold factor. In another embodiment, the adaptive immune response is increased by a 10-fold factor. In another embodiment, the adaptive immune response is increased by a 15-fold factor. In another embodiment the adaptive immune response is increased by a 20-fold factor. In another embodiment, the adaptive immune response is increased by a 50-fold factor. In another embodiment, the adaptive immune response is increased by a 100-fold factor. In another embodiment, the adaptive immune response is increased by a 200-fold factor. In another embodiment, the adaptive immune response is increased by a 500-fold factor. In another embodiment, the adaptive immune response is increased by a 1000-fold factor. In another embodiment, the adaptive immune response is increased by a 2000-fold factor. In another embodiment, the adaptive immune response is increased by another fold difference.

In another embodiment, "induces significantly more adaptive immune response" refers to a detectable increase in an adaptive immune response. In another embodiment, the term refers to a fold increase in the adaptive immune response (e.g., 1 of the fold increases enumerated above). In another embodiment, the term refers to an increase such that the nucleoside-modified RNA can be administered at a lower dose or frequency than an unmodified RNA molecule with the same species while still inducing an effective adaptive immune response. In another embodiment, the increase is such that the nucleoside-modified RNA can be administered using a single dose to induce an effective adaptive immune response.

In another embodiment, the nucleoside-modified RNA of the invention exhibits significantly less innate immunogenicity than an unmodified in vitro-synthesized RNA molecule with the same sequence. In another embodiment, the modified RNA molecule exhibits an innate immune response that is 2-fold less than its unmodified counterpart. In another embodiment, innate immunogenicity is reduced by a 3-fold factor. In another embodiment, innate immunogenicity is reduced by a 5-fold factor. In another embodiment, innate immunogenicity is reduced by a 7-fold factor. In another embodiment, innate immunogenicity is reduced by a 10-fold factor. In another embodiment, innate immunogenicity is reduced by a 15-fold factor. In another embodiment, innate immunogenicity is reduced by a 20-fold factor. In another embodiment, innate immunogenicity is reduced by a 50-fold factor. In another embodiment, innate immunogenicity is reduced by a 100-

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fold factor. In another embodiment, innate immunogenicity is reduced by a 200-fold factor. In another embodiment, innate immunogenicity is reduced by a 500-fold factor. In another embodiment, innate immunogenicity is reduced by a 1000-fold factor. In another embodiment, innate immunogenicity is reduced by a 2000-fold factor. In another embodiment, innate immunogenicity is reduced by another fold difference.

In another embodiment, "exhibits significantly less innate immunogenicity" refers to a detectable decrease in innate immunogenicity. In another embodiment, the term refers to a fold decrease in innate immunogenicity (e.g., 1 of the fold decreases enumerated above). In another embodiment, the term refers to a decrease such that an effective amount of the nucleoside-modified RNA can be administered without triggering a detectable innate immune response. In another embodiment, the term refers to a decrease such that the nucleoside-modified RNA can be repeatedly administered without eliciting an innate immune response sufficient to detectably reduce production of the recombinant protein. In another embodiment, the decrease is such that the nucleoside-modified RNA can be repeatedly administered without eliciting an innate immune response sufficient to eliminate detectable production of the recombinant protein.

Polypeptide Therapeutic Agents

In other related aspects, the therapeutic agent includes an isolated peptide that modulates a target. For example, in one embodiment, the peptide of the invention inhibits or activates a target directly by binding to the target thereby modulating the normal functional activity of the target. In one embodiment, the peptide of the invention modulates the target by competing with endogenous proteins. In one embodiment, the peptide of the invention modulates the activity of the target by acting as a transdominant negative mutant.

The variants of the polypeptide therapeutic agents may be (i) one in which one or more of the amino acid residues are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code, (ii) one in which there are one or more modified amino acid residues, e.g., residues that are modified by the attachment of substituent groups, (iii) one in which the polypeptide is an alternative splice variant of the polypeptide of the invention, (iv) fragments of the polypeptides and/or (v) one in which the polypeptide is fused with another polypeptide, such as a leader or secretory sequence or a sequence which is employed for purification (for example, His-tag) or for detection (for example, Sv5 epitope tag). The fragments include polypeptides generated via proteolytic cleavage (including multi-site proteolysis) of an

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original sequence. Variants may be post-translationally, or chemically modified. Such variants are deemed to be within the scope of those skilled in the art from the teaching herein.

The nanoparticles may further comprise any lipid capable of forming a particle to which the one or more nucleic acid molecules are attached, or in which the one or more nucleic acid molecules are encapsulated. The term "lipid" refers to a group of organic compounds that are derivatives of fatty acids (e.g., esters) and are generally characterized by being insoluble in water but soluble in many organic solvents. Lipids are usually divided in at least three classes: (1) "simple lipids" which include fats and oils as well as waxes; (2) "compound lipids" which include phospholipids and glycolipids; and (3) "derived lipids" such as steroids. In one embodiment, the nanoparticles do not further comprise any lipid capable of forming a particle to which the one or more nucleic acid molecules are attached, or in which the one or more nucleic acid molecules are encapsulated. In one embodiment, the nanoparticles do not further comprise any or all of a simple lipid, a compound lipid, or a derived lipid.

In one embodiment, the nanoparticle comprises a cationic lipid. As used herein, the term "cationic lipid" refers to a lipid that is cationic or becomes cationic (protonated) as the pH is lowered below the pK of the ionizable group of the lipid, but is progressively more neutral at higher pH values. At pH values below the pK, the lipid is then able to associate with negatively charged nucleic acids. In certain embodiments, the cationic lipid comprises a zwitterionic lipid that assumes a positive charge on pH decrease. In one embodiment, the nanoparticle does not comprise a cationic lipid.

In certain embodiments, the cationic lipid which is optionally present or not present in the nanoparticles comprises any of a number of lipid species which carry a net positive charge at a selective pH, such as physiological pH. Such lipids include, but are not limited to, N,N-dioleyl-N,N-dimethylammonium chloride (DODAC); N-(2,3-dioleyloxy)propyl)-N,N,N-trimethylammonium chloride (DOTMA); N,N-distearyl-N,N-dimethylammonium bromide (DDAB); N-(2,3-dioleoyloxy)propyl)-N,N,N-trimethylammonium chloride (DOTAP); 3-(N—(N',N'-dimethylaminoethane)-carbamoyl)cholesterol (DC-Chol), N-(1-(2,3-dioleoyloxy)propyl)-N-2-(sperminecarboxamido)ethyl)-N,N-dimethylammonium trifluoracetate (DOSPA), dioctadecylamidoglycyl carboxyspermine (DOGS), 1,2-dioleoyl-3-dimethylammonium propane (DODAP), N,N-dimethyl-2,3-dioleoyloxy)propylamine (DODMA), and N-(1,2-dimyristyloxyprop-3-yl)-N,N-dimethyl-N-hydroxyethyl ammonium bromide (DMRIE). Additionally, a number of commercial preparations of cationic lipids are available which can be used in the invention. These

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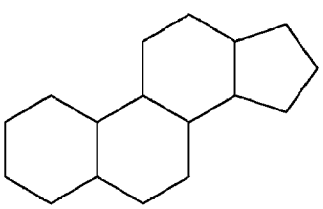
include, for example, LIPOFECTIN® (commercially available cationic liposomes comprising DOTMA and 1,2-dioleoyl-sn-3-phosphoethanolamine (DOPE), from GIBCO/BRL, Grand Island, N.Y.); LIPOFECTAMINE® (commercially available cationic liposomes comprising N-(1-(2,3-dioleyloxy)propyl)-N-(2-(sperminecarboxamido)ethyl)-N,N-dimethylammonium trifluoroacetate (DOSPA) and (DOPE), from GIBCO/BRL); and TRANSFECTAM® (commercially available cationic lipids comprising dioctadecylamidoglycyl carboxyspermine (DOGS) in ethanol from Promega Corp., Madison, Wis.). The following lipids are cationic and have a positive charge at below physiological pH: DODAP, DODMA, DMDMA, 1,2-dilinoleyloxy-N,N-dimethylaminopropane (DLinDMA), 1,2-dilinolenyloxy-N,N-dimethylaminopropane (DLenDMA).

In one embodiment, the cationic lipid is an amino lipid. Such amino lipids include those described in WO 2012/016184, incorporated herein by reference in its entirety. Representative amino lipids include, but are not limited to, 1,2-dilinoleyoxy-3-(dimethylamino)acetoxypropane (DLin-DAC), 1,2-dilinoleyoxy-3-morpholinopropane (DLin-MA), 1,2-dilinoleoyl-3-dimethylaminopropane (DLinDAP), 1,2-dilinoleylthio-3-dimethylaminopropane (DLin-S-DMA), 1-linoleoyl-2-linoleyloxy-3-dimethylaminopropane (DLin-2-DMAP), 1,2-dilinoleyloxy-3-trimethylaminopropane chloride salt (DLin-TMA.Cl), 1,2-dilinoleoyl-3-trimethylaminopropane chloride salt (DLin-TAP.Cl), 1,2-dilinoleyloxy-3-(N-methylpiperazino)propane (DLin-MPZ), 3-(N,N-dilinoleylamino)-1,2-propanediol (DLinAP), 3-(N,N-dioleylamino)-1,2-propanediol (DOAP), 1,2-dilinoleyloxo-3-(2-N,N-dimethylamino)ethoxypropane (DLin-EG-DMA), and 2,2-dilinoleyl-4-dimethylaminomethyl-[1,3]-dioxolane (DLin-K-DMA).

In various embodiments, the nanoparticles further comprise a steroid or steroid analogue. A "steroid" is a compound comprising the following carbon skeleton:



In certain embodiments, the steroid or steroid analogue is cholesterol. In one embodiment, the nanoparticles do not comprise a steroid or steroid analogue. In one embodiment, the nanoparticles do not comprise cholesterol.

In one embodiment, the nanoparticles further comprise a stabilizer. In one embodiment, the stabilizer comprises oligooxyetylenes. In one embodiment, the stabilizer comprises a water soluble macromolecule. In one embodiment the stabilizer comprises a water soluble oligomer. In one

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embodiment, the stabilizer comprises a carbohydrate.

In certain embodiments, the nanoparticle comprises one or more targeting moieties which are capable of targeting the nanoparticle to a cell or cell population. For example, in one embodiment, the targeting moiety is a ligand which directs the nanoparticle to a receptor found on a cell surface.

In certain embodiments, the nanoparticle comprises one or more internalization domains. For example, in one embodiment, the nanoparticle comprises one or more domains which bind to a cell to induce the internalization of the nanoparticle. For example, in one embodiment, the one or more internalization domains bind to a receptor found on a cell surface to induce receptor-mediated uptake of the nanoparticle. In certain embodiments, the nanoparticle is capable of binding a biomolecule in vivo, where the nanoparticle-bound biomolecule can then be recognized by a cell-surface receptor to induce internalization. For example, in one embodiment, the nanoparticle binds systemic ApoE, which leads to the uptake of the nanoparticle and associated cargo.

Compositions

In one aspect, the present invention relates to compositions comprising at least one amphiphilic Janus dendrimer of the presentin invention and/or nanoparticle thereof. In some embodiments, the composition further comprises at least one agent described herein.

The invention also relates to compositions comprising at least one compound of Formula (I) and methods of use thereof for delivering an encapsulated agent to a site of interest. Exemplary agents that can be encapsulated in the compositions of the invention include, but are not limited to, diagnostic agents, detectable agents, and therapeutic agents.

In one embodiment, the composition comprises nanoparticles comprising a compound of Formula (I) and at least one agent encapsulatd by the nanoparticle. In some embodiments, the encapsulated agent comprises an agent for inducing an immune response in a subject. In certain embodiments, the invention provides a composition comprising a nanoparticle encapsulating a nucleic acid molecule encoding an agent for inducing an immune response in a subject. For example, in certain embodiments, the composition comprises a vaccine comprising a nucleic acid molecule encoding an antigen.

In one embodiment, the composition is a vaccine.

In one embodiment, the composition comprises a nanoparticle and one or more nucleic acid molecules described herein. For example, in one embodiment, the composition comprises a

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nanoparticle and one or more nucleoside-modified RNA molecules encoding one or more antigens, adjuvants, or a combination thereof.

In one embodiment, the composition may be prepared by injection of a mixture comprising a compound described herein into a suitable solution, such as a solution comprising the agent to be encapsulated. In one embodiment, microfluidic techniques such as those required for the formation of lipid nanoparticles (LNPs) are not required for the production of the inventive nanoparticles.

In one embodiment, the composition of the invention comprises in vitro transcribed (IVT) RNA molecule. For example, in certain embodiments, the composition of the invention comprises IVT RNA molecule which encodes an agent. In certain embodiments, the IVT RNA molecule of the present composition is a nucleoside-modified mRNA molecule. In certain embodiments, the agent is at least one of a viral antigen, bacterial antigen, fungal antigen, parasitic antigen, tumor-specific antigen, or tumor-associated antigen. However, the invention is not limited to any particular agent or combination of agents. In certain embodiments, the composition comprises an adjuvant. In certain embodiments, the composition comprises a nucleic acid molecule encoding an adjuvant. In one embodiment, the composition comprises a nucleoside-modified RNA encoding an adjuvant.

In one embodiment, the composition comprises at least one nucleoside-modified RNA molecule encoding a combination of at least two agents. In one embodiment, the composition comprises a combination of two or more nucleoside-modified RNA molecules encoding a combination of two or more agents.

In one embodiment, the invention provides a method for inducing an immune response in a subject. For example, the method can be used to provide immunity in the subject against a virus, bacteria, fungus, parasite, cancer, or the like. In some embodiments, the method comprises administering to the subject a composition comprising one or more nanoparticles comprising one or more nucleoside-modified RNA encoding at least one antigen, an adjuvant, or a combination thereof.

In one embodiment, the method comprises the systemic administration of the composition into the subject, including for example intradermal administration. In certain embodiments, the method comprises administering a plurality of doses to the subject. In another embodiment, the method comprises administering a single dose of the composition, where the single dose is effective in inducing a therapeutic response.

Vaccine

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In one embodiment, the invention provides an immunogenic composition for inducing an immune response in a subject. For example, in one embodiment, the immunogenic composition is a vaccine. As used herein, an "immunogenic composition" may comprise an antigen (e.g., a peptide or polypeptide), a nucleic acid encoding an antigen, a cell expressing or presenting an antigen or cellular component, or a combination thereof. In particular embodiments the composition comprises or encodes all or part of any peptide antigen, or an immunogenically functional equivalent thereof. In other embodiments, the composition comprises a mixture of mRNA molecules that encodes one or more additional immunostimulatory agent. Immunostimulatory agents include, but are not limited to, an additional antigen, an immunomodulator, or an adjuvant. In the context of the invention, the term "vaccine" refers to a substance that induces immunity upon inoculation into animals.

A vaccine of the invention may vary in its composition of nucleic acid components. In a non-limiting example, a nucleic acid encoding an antigen might also be formulated with an adjuvant. Of course, it will be understood that various compositions described herein may further comprise additional components. A vaccine of the invention, and its various components, may be prepared and/or administered by any method disclosed herein or as would be known to one of ordinary skill in the art, in light of the present disclosure.

The induction of the immunity by the expression of the antigen can be detected by observing in vivo or in vitro the response of all or any part of the immune system in the host against the antigen.

For example, a method for detecting the induction of cytotoxic T lymphocytes is well known. A foreign substance that enters the living body is presented to T cells and B cells by the action of APCs. T cells that respond to the antigen presented by APC in an antigen specific manner differentiate into cytotoxic T cells (also referred to as cytotoxic T lymphocytes or CTLs) due to stimulation by the antigen. These antigen stimulated cells then proliferate. This process is referred to herein as "activation" of T cells. Therefore, CTL induction by an epitope of a polypeptide or peptide or combinations thereof can be evaluated by presenting an epitope of a polypeptide or peptide or combinations thereof to a T cell by APC, and detecting the induction of CTL. Furthermore, APCs have the effect of activating B cells, CD4+ T cells, CD8+ T cells, macrophages, eosinophils and NK cells.

A method for evaluating the inducing action of CTL using dendritic cells (DCs) as APC is well known in the art. DC is a representative APC having a robust CTL inducing action among APCs. In the methods of the invention, the epitope of a polypeptide or peptide or combinations

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thereof is initially expressed by the DC and then this DC is contacted with T cells. Detection of T cells having cytotoxic effects against the cells of interest after the contact with DC shows that the an epitope of a polypeptide or peptide or combinations thereof has an activity of inducing the cytotoxic T cells. Furthermore, the induced immune response can be also examined by measuring IFN-gamma produced and released by CTL in the presence of antigen-presenting cells that carry immobilized peptide or combination of peptides by visualizing using anti-IFN-gamma antibodies, such as an ELISPOT assay.

Apart from DC, peripheral blood mononuclear cells (PBMCs) may also be used as the APC. The induction of CTL is reported to be enhanced by culturing PBMC in the presence of GM-CSF and IL-4. Similarly, CTL has been shown to be induced by culturing PBMC in the presence of keyhole limpet hemocyanin (KLH) and IL-7.

The antigens confirmed to possess CTL-inducing activity by these methods are antigens having DC activation effect and subsequent CTL-inducing activity. Furthermore, CTLs that have acquired cytotoxicity due to presentation of the antigen by APC can be also used as vaccines against antigen-associated disorders.

The induction of immunity by expression of the antigen can be further confirmed by observing the induction of antibody production against the antigen. For example, when antibodies against an antigen are induced in a laboratory animal immunized with the composition encoding the antigen, and when antigen-associated pathology is suppressed by those antibodies, the composition is determined to induce immunity.

The induction of immunity by expression of the antigen can be further confirmed by observing the induction of CD4+ T cells. CD4+ T cells can also lyse target cells, but mainly supply help in the induction of other types of immune responses, including CTL and antibody generation. The type of CD4+ T cell help can be characterized, as Th1, Th2, Th9, Th17, Tregulatory, or T follicular helper (T_{fh}) cells. Each subtype of CD4+ T cell supplies help to certain types of immune responses. Of particular interest to this invention, the T_{fh} subtype provides help in the generation of high affinity antibodies.

In some embodiments, the therapeutic compounds or compositions of the invention may be administered prophylactically (i.e., to prevent disease or disorder) or therapeutically (i.e., to treat disease or disorder) to subjects suffering from or at risk of (or susceptible to) developing the disease or disorder. Such subjects may be identified using standard clinical methods. In the context of the invention, prophylactic administration occurs prior to the manifestation of overt clinical symptoms of disease, such that a disease or disorder is prevented or alternatively delayed in its progression. In

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the context of the field of medicine, the term "prevent" encompasses any activity which reduces the burden of mortality or morbidity from disease. Prevention can occur at primary, secondary and tertiary prevention levels. While primary prevention avoids the development of a disease, secondary and tertiary levels of prevention encompass activities aimed at preventing the progression of a disease and the emergence of symptoms as well as reducing the negative impact of an already established disease by restoring function and reducing disease-related complications.

Targeting Domain

In one embodiment, the composition comprises a targeting domain that directs the delivery vehicle to a site. In one embodiment, the site is a site in need of the agent comprised within the delivery vehicle. The targeting domain may comprise a nucleic acid, peptide, antibody, small molecule, organic molecule, inorganic molecule, glycan, sugar, hormone, and the like that targets the particle to a site in particular need of the therapeutic agent. In certain embodiments, the particle comprises multivalent targeting, wherein the particle comprises multiple targeting mechanisms described herein. In certain embodiments, the targeting domain of the delivery vehicle specifically binds to a target associated with a site in need of an agent comprised within the delivery vehicle. For example, the targeting domain may be chosen to recognize a ligand that acts as a cell surface marker on target cells associated with a particular disease state. Such a target can be a protein, protein fragment, antigen, or other biomolecule that is associated with the targeted site. In some embodiments, the targeting domain is an affinity ligand which specifically binds to a target. In certain embodiments, the target (e.g. antigen) associated with a site in need of a treatment with an agent. In some embodiments, the targeting domain may be co-polymerized with the composition comprising the delivery vehicle. In some embodiments, the targeting domain may be covalently attached to the composition comprising the delivery vehicle, such as through a chemical reaction between the targeting domain and the composition comprising the delivery vehicle. In some embodiments, the targeting domain is an additive in the delivery vehicle. Targeting domains of the instant invention include, but are not limited to, antibodies, antibody fragments, proteins, peptides, and nucleic acids.

In various embodiments, the targeting domain binds to a cell surface molecule of a cell of interest. For example, in various embodiments, the targeting domain binds to a cell surface molecule of an endothelial cell, a stem cell, or an immune cell.

Peptides

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In one embodiment, the targeting domain of the invention comprises a peptide. In certain embodiments, the peptide targeting domain specifically binds to a target of interest.

The peptide of the invention may be made using chemical methods. For example, peptides can be synthesized by solid phase techniques (Roberge J Y et al (1995) Science 269: 202-204), cleaved from the resin, and purified by preparative high performance liquid chromatography. Automated synthesis may be achieved, for example, using the ABI 431 A Peptide Synthesizer (Perkin Elmer) in accordance with the instructions provided by the manufacturer.

The peptide may alternatively be made by recombinant means or by cleavage from a longer polypeptide. The composition of a peptide may be confirmed by amino acid analysis or sequencing.

The variants of the peptides according to the invention may be (i) one in which one or more of the amino acid residues are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code, (ii) one in which there are one or more modified amino acid residues, e.g., residues that are modified by the attachment of substituent groups, (iii) one in which the peptide is an alternative splice variant of the peptide of the invention, (iv) fragments of the peptides and/or (v) one in which the peptide is fused with another peptide, such as a leader or secretory sequence or a sequence which is employed for purification (for example, His-tag) or for detection (for example, Sv5 epitope tag). The fragments include peptides generated via proteolytic cleavage (including multi-site proteolysis) of an original sequence. Variants may be post-translationally, or chemically modified. Such variants are deemed to be within the scope of those skilled in the art from the teaching herein.

As known in the art the "similarity" between two peptides is determined by comparing the amino acid sequence and its conserved amino acid substitutes of one peptide to a sequence of a second peptide. Variants are defined to include peptide sequences different from the original sequence, preferably different from the original sequence in less than 40% of residues per segment of interest, more preferably different from the original sequence in less than 25% of residues per segment of interest, more preferably different by less than 10% of residues per segment of interest, most preferably different from the original protein sequence in just a few residues per segment of interest and at the same time sufficiently homologous to the original sequence to preserve the functionality of the original sequence. The invention includes amino acid sequences that are at least 60%, 65%, 70%, 72%, 74%, 76%, 78%, 80%, 90%, or 95% similar or identical to the original amino acid sequence. The degree of identity between two peptides is determined using computer algorithms and methods that are widely known for the persons skilled in the art. The identity

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between two amino acid sequences is preferably determined by using the BLASTP algorithm [BLAST Manual, Altschul, S., et al., NCBI NLM NIH Bethesda, Md. 20894, Altschul, S., et al., J. Mol. Biol. 215: 403-410 (1990)].

The peptides of the invention can be post-translationally modified. For example, post-translational modifications that fall within the scope of the invention include signal peptide cleavage, glycosylation, acetylation, isoprenylation, proteolysis, myristoylation, protein folding and proteolytic processing, etc. Some modifications or processing events require introduction of additional biological machinery. For example, processing events, such as signal peptide cleavage and core glycosylation, are examined by adding canine microsomal membranes or Xenopus egg extracts (U.S. Pat. No. 6,103,489) to a standard translation reaction.

The peptides of the invention may include unnatural amino acids formed by post-translational modification or by introducing unnatural amino acids during translation.

Nucleic Acids

In one embodiment, the targeting domain of the invention comprises an isolated nucleic acid, including for example a DNA oligonucleotide and a RNA oligonucleotide. In certain embodiments, the nucleic acid targeting domain specifically binds to a target of interest. For example, in one embodiment, the nucleic acid comprises a nucleotide sequence that specifically binds to a target of interest.

The nucleotide sequences of a nucleic acid targeting domain can alternatively comprise sequence variations with respect to the original nucleotide sequences, for example, substitutions, insertions and/or deletions of one or more nucleotides, with the condition that the resulting nucleic acid functions as the original and specifically binds to the target of interest.

In the sense used in this description, a nucleotide sequence is "substantially homologous" to any of the nucleotide sequences describe herein when its nucleotide sequence has a degree of identity with respect to the nucleotide sequence of at least 60%, advantageously of at least 70%, preferably of at least 85%, and more preferably of at least 95%. Other examples of possible modifications include the insertion of one or more nucleotides in the sequence, the addition of one or more nucleotides in any of the ends of the sequence, or the deletion of one or more nucleotides in any end or inside the sequence. The degree of identity between two polynucleotides is determined using computer algorithms and methods that are widely known for the persons skilled in the art. The identity between two amino acid sequences is preferably determined by using the BLASTN algorithm [BLAST Manual, Altschul, S., et al., NCBI NLM NIH Bethesda, Md. 20894, Altschul,

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S., et al., J. Mol. Biol. 215: 403-410 (1990)].

Antibodies

In one embodiment, the targeting domain of the invention comprises an antibody, or antibody fragment. In certain embodiments, the antibody targeting domain specifically binds to a target of interest. Such antibodies include polyclonal antibodies, monoclonal antibodies, Fab and single chain Fv (scFv) fragments thereof, bispecific antibodies, heteroconjugates, human and humanized antibodies.

The antibodies may be intact monoclonal or polyclonal antibodies, and immunologically active fragments (e.g., a Fab or (Fab)2 fragment), an antibody heavy chain, an antibody light chain, humanized antibodies, a genetically engineered single chain Fv molecule (Ladner et al, U.S. Pat. No. 4,946,778), or a chimeric antibody, for example, an antibody which contains the binding specificity of a murine antibody, but in which the remaining portions are of human origin. Antibodies including monoclonal and polyclonal antibodies, fragments and chimeras, may be prepared using methods known to those skilled in the art.

Such antibodies may be produced in a variety of ways, including hybridoma cultures, recombinant expression in bacteria or mammalian cell cultures, and recombinant expression in transgenic animals. The choice of manufacturing methodology depends on several factors including the antibody structure desired, the importance of carbohydrate moieties on the antibodies, ease of culturing and purification, and cost. Many different antibody structures may be generated using standard expression technology, including full-length antibodies, antibody fragments, such as Fab and Fv fragments, as well as chimeric antibodies comprising components from different species. Antibody fragments of small size, such as Fab and Fv fragments, having no effector functions and limited pharmokinetic activity may be generated in a bacterial expression system. Single chain Fv fragments show low immunogenicity.

Adjuvant

In one embodiment, the composition comprises an adjuvant. In one embodiment, the composition comprises a nucleic acid molecule encoding an adjuvant. In one embodiment, the adjuvant-encoding nucleic acid molecule is IVT RNA. In one embodiment, the adjuvant-encoding nucleic acid molecule is nucleoside-modified mRNA.

Exemplary adjuvants include, but is not limited to, alpha-interferon, gamma-interferon, platelet derived growth factor (PDGF), TNFα, TNFβ, GM-CSF, epidermal growth factor (EGF), cutaneous T cell-attracting chemokine (CTACK), epithelial thymus-expressed chemokine (TECK),

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mucosae-associated epithelial chemokine (MEC), IL-12, IL-15, MHC, CD80, CD86 including IL-15 having the signal sequence deleted and optionally including the signal peptide from IgE. Other genes which may be useful adjuvants include those encoding: MCP-I, MIP-Ia, MIP-Ip, IL-8, RANTES, L-selectin, P-selectin, E-selectin, CD34, GlyCAM-1, MadCAM-1, LFA-I, VLA-I, Mac-1, pl50.95, PECAM, ICAM-I, ICAM-2, ICAM-3, CD2, LFA-3, M-CSF, G-CSF, IL-4, mutant forms of IL-18, CD40, CD40L, vascular growth factor, fibroblast growth factor, IL-7, nerve growth factor, vascular endothelial growth factor, Fas, TNF receptor, Fit, Apo-1, p55, WSL-I, DR3, TRAMP, Apo-3, AIR, LARD, NGRF, DR4, DR5, KILLER, TRAIL-R2, TRICK2, DR6, Caspase ICE, Fos, c-jun, Sp-I, Ap-I, Ap-2, p38, p65Rel, MyD88, IRAK, TRAF6, IkB, Inactive NIK, SAP K, SAP-I, JNK, interferon response genes, NFkB, Bax, TRAIL, TRAILrec, TRAILrecDRC5, TRAIL-R3, TRAIL-R4, RANK, RANK LIGAND, Ox40, Ox40 LIGAND, NKG2D, MICA, MICB, NKG2A, NKG2B, NKG2C, NKG2E, NKG2F, TAP 1, TAP2, anti-CTLA4-sc, anti-LAG3-Ig, anti-TIM3-Ig and functional fragments thereof.

Antibody Therapeutic Agents

The invention also contemplates a delivery vehicle comprising an antibody, or antibody fragment, specific for a target. That is, the antibody can inhibit a target to provide a beneficial effect.

The antibodies may be intact monoclonal or polyclonal antibodies, and immunologically active fragments (e.g., a Fab or (Fab)2 fragment), an antibody heavy chain, an antibody light chain, humanized antibodies, a genetically engineered single chain FV molecule (Ladner et al, U.S. Pat. No. 4,946,778), or a chimeric antibody, for example, an antibody which contains the binding specificity of a murine antibody, but in which the remaining portions are of human origin. Antibodies including monoclonal and polyclonal antibodies, fragments and chimeras, may be prepared using methods known to those skilled in the art.

Antibodies can be prepared using intact polypeptides or fragments containing an immunizing antigen of interest. The polypeptide or oligopeptide used to immunize an animal may be obtained from the translation of RNA or synthesized chemically and can be conjugated to a carrier protein, if desired. Suitable carriers that may be chemically coupled to peptides include bovine serum albumin and thyroglobulin, keyhole limpet hemocyanin. The coupled polypeptide may then be used to immunize the animal (e.g., a mouse, a rat, or a rabbit).

Antigen

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The invention provides a composition that induces a therapeutic response in a subject. In one embodiment, the composition comprises an antigen. In one embodiment, the composition comprises a nucleic acid sequence which encodes an antigen. For example, in certain embodiments, the composition comprises a nucleoside-modified RNA encoding an antigen. The antigen may be any molecule or compound, including but not limited to a polypeptide, peptide or protein that induces a therapeutic response, such as an adaptive immune response, in a subject.

In one embodiment, the antigen comprises a polypeptide or peptide associated with a pathogen, such that the antigen induces an adaptive immune response against the antigen, and therefore the pathogen. In one embodiment, the antigen comprises a fragment of a polypeptide or peptide associated with a pathogen, such that the antigen induces an adaptive immune response against the pathogen.

In certain embodiments, the antigen comprises an amino acid sequence that is substantially homologous to the amino acid sequence of an antigen described herein and retains the immunogenic function of the original amino acid sequence. For example, in certain embodiments, the amino acid sequence of the antigen has a degree of identity with respect to the original amino acid sequence of at least 60%, advantageously of at least 70%, preferably of at least 85%, and more preferably of at least 95%.

In one embodiment, the antigen is encoded by a nucleic acid sequence of a nucleic acid molecule. In certain embodiments, the nucleic acid sequence comprises DNA, RNA, cDNA, a variant thereof, a fragment thereof, or a combination thereof. In one embodiment, the nucleic acid sequence comprises a modified nucleic acid sequence. For example, in one embodiment the antigen-encoding nucleic acid sequence comprises nucleoside-modified RNA, as described in detail elsewhere herein. In certain instances, the nucleic acid sequence comprises include additional sequences that encode linker or tag sequences that are linked to the antigen by a peptide bond.

In certain embodiments, the antigen, encoded by the nucleoside-modified nucleic acid molecule, comprises a protein, peptide, a fragment thereof, or a variant thereof, or a combination thereof from any number of organisms, for example, a virus, a parasite, a bacterium, a fungus, or a mammal. For example, in certain embodiments, the antigen is associated with an autoimmune disease, allergy, or asthma. In other embodiments, the antigen is associated with cancer, herpes, influenza, hepatitis B, hepatitis C, human papilloma virus (HPV), ebola, pneumococcus, Haemophilus influenza, meningococcus, dengue, tuberculosis, malaria, norovirus or human immunodeficiency virus (HIV). In certain embodiments, the antigen comprises a consensus sequence based on the amino acid sequence of two or more different organisms. In certain

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embodiments, the nucleic acid sequence encoding the antigen is optimized for effective translation in the organism in which the composition is delivered.

In one embodiment, the antigen comprises a tumor-specific antigen or tumor-associated antigen, such that the antigen induces an adaptive immune response against the tumor. In one embodiment, the antigen comprises a fragment of a tumor-specific antigen or tumor-associated antigen, such that the antigen induces an adaptive immune response against the tumor. In certain embodiment, the tumor-specific antigen or tumor-associated antigen is a mutation variant of a host protein.

Viral Antigens

In one embodiment, the antigen comprises a viral antigen, or fragment thereof, or variant thereof. In certain embodiments, the viral antigen is from a virus from one of the following families: Adenoviridae, Arenaviridae, Bunyaviridae, Caliciviridae, Coronaviridae, Filoviridae, Hepadnaviridae, Herpesviridae, Orthomyxoviridae, Papovaviridae, Paramyxoviridae, Parvoviridae, Picornaviridae, Poxviridae, Reoviridae, Retroviridae, Rhabdoviridae, or Togaviridae. In certain embodiments, the viral antigen is from papilloma viruses, for example, human papillomoa virus (HPV), human immunodeficiency virus (HIV), polio virus, hepatitis B virus, hepatitis C virus, smallpox virus (Variola major and minor), vaccinia virus, influenza virus, rhinoviruses, dengue fever virus, equine encephalitis viruses, rubella virus, yellow fever virus, Norwalk virus, hepatitis A virus, human T-cell leukemia virus (HTLV-I), hairy cell leukemia virus (HTLV-II), California encephalitis virus, Hanta virus (hemorrhagic fever), rabies virus, Ebola fever virus, Marburg virus, measles virus, mumps virus, respiratory syncytial virus (RSV), herpes simplex 1 (oral herpes), herpes simplex 2 (genital herpes), herpes zoster (varicella-zoster, a.k.a., chickenpox), cytomegalovirus (CMV), for example human CMV, Epstein-Barr virus (EBV), flavivirus, foot and mouth disease virus, chikungunya virus, lassa virus, arenavirus, severe acute respiratory syndrome (SARS) virus, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) or a cancer causing virus.

Parasite Antigens

In certain embodiments, the antigen comprises a parasite antigen or fragment or variant thereof. In certain embodiments, the parasite is a protozoa, helminth, or ectoparasite. In certain embodiments, the helminth (i.e., worm) is a flatworm (e.g., flukes and tapeworms), a thorny-headed worm, or a round worm (e.g., pinworms). In certain embodiments, the ectoparasite is lice, fleas,

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ticks, and mites.

In certain embodiments, the parasite is any parasite causing the following diseases: Acanthamoeba keratitis, Amoebiasis, Ascariasis, Babesiosis, Balantidiasis, Baylisascariasis, Chagas disease, Clonorchiasis, Cochliomyia, Cryptosporidiosis, Diphyllobothriasis, Dracunculiasis, Echinococcosis, Elephantiasis, Enterobiasis, Fascioliasis, Fasciolopsiasis, Filariasis, Giardiasis, Gnathostomiasis, Hymenolepiasis, Isosporiasis, Katayama fever, Leishmaniasis, Lyme disease, Malaria, Metagonimiasis, Myiasis, Onchocerciasis, Pediculosis, Scabies, Schistosomiasis, Sleeping sickness, Strongyloidiasis, Taeniasis, Toxocariasis, Toxoplasmosis, Trichinosis, and Trichuriasis.

In certain embodiments, the parasite is Acanthamoeba, Anisakis, Ascaris lumbricoides, Botfly, Balantidium coli, Bedbug, Cestoda (tapeworm), Chiggers, Cochliomyia hominivorax, Entamoeba histolytica, Fasciola hepatica, Giardia lamblia, Hookworm, Leishmania, Linguatula serrata, Liver fluke, Loa loa, Paragonimus - lung fluke, Pinworm, Plasmodium falciparum, Schistosoma, Strongyloides stercoralis, Mite, Tapeworm, Toxoplasma gondii, Trypanosoma, Whipworm, or Wuchereria bancrofti.

Bacterial Antigens

In one embodiment, the antigen comprises a bacterial antigen or fragment or variant thereof. In certain embodiments, the bacterium is from any one of the following phyla: Acidobacteria, Actinobacteria, Aquificae, Bacteroidetes, Caldiserica, Chlamydiae, Chlorobi, Chloroflexi, Chrysiogenetes, Cyanobacteria, Deferribacteres, Deinococcus-Thermus, Dictyoglomi, Elusimicrobia, Fibrobacteres, Firmicutes, Fusobacteria, Gemmatimonadetes, Lentisphaerae, Nitrospira, Planctomycetes, Proteobacteria, Spirochaetes, Synergistetes, Tenericutes, Thermodesulfobacteria, Thermotogae, and Verrucomicrobia.

In certain embodiments, the bacterium is a gram positive bacterium or a gram negative bacterium. In certain embodiments, the bacterium is an aerobic bacterium or an anaerobic bacterium. In certain embodiments, the bacterium is an autotrophic bacterium or a heterotrophic bacterium. In certain embodiments, the bacterium is a mesophile, a neutrophile, an extremophile, an acidophile, an alkaliphile, a thermophile, psychrophile, halophile, or an osmophile.

In certain embodiments, the bacterium is an anthrax bacterium, an antibiotic resistant bacterium, a disease causing bacterium, a food poisoning bacterium, an infectious bacterium, Salmonella bacterium, Staphylococcus bacterium, Streptococcus bacterium, or tetanus bacterium. In certain embodiments, bacterium is a mycobacteria, Clostridium tetani, Yersinia pestis, Bacillus anthracis, methicillin-resistant Staphylococcus aureus (MRSA), or Clostridium difficile.

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Fungal Antigens

In one embodiment, the antigen comprises a fungal antigen or fragment or variant thereof. In certain embodiments, the fungus is Aspergillus species, Blastomyces dermatitidis, Candida yeasts (e.g., Candida albicans), Coccidioides, Cryptococcus neoformans, Cryptococcus gattii, dermatophyte, Fusarium species, Histoplasma capsulatum, Mucoromycotina, Pneumocystis jirovecii, Sporothrix schenckii, Exserohilum, or Cladosporium.

Tumor Antigens

In certain embodiments, the antigen comprises a tumor antigen, including for example a tumor-associated antigen or a tumor-specific antigen. In the context of the invention, "tumor antigen" or "hyperporoliferative disorder antigen" or "antigen associated with a hyperproliferative disorder" refer to antigens that are common to specific hyperproliferative disorders. In certain aspects, the hyperproliferative disorder antigens of the invention are derived from cancers including, but not limited to, primary or metastatic melanoma, mesothelioma, thymoma, lymphoma, sarcoma, lung cancer, liver cancer, non-Hodgkin's lymphoma, Hodgkins lymphoma, leukemias, uterine cancer, cervical cancer, bladder cancer, kidney cancer and adenocarcinomas such as breast cancer, prostate cancer, ovarian cancer, pancreatic cancer, and the like.

Tumor antigens are proteins that are produced by tumor cells that elicit an immune response, particularly T-cell mediated immune responses. In one embodiment, the tumor antigen of the invention comprises one or more antigenic cancer epitopes immunogenically recognized by tumor infiltrating lymphocytes (TIL) derived from a cancer tumor of a mammal. The selection of the antigen will depend on the particular type of cancer to be treated or prevented by way of the composition of the invention.

Tumor antigens are well known in the art and include, for example, a glioma-associated antigen, carcinoembryonic antigen (CEA), β-human chorionic gonadotropin, alphafetoprotein (AFP), lectin-reactive AFP, thyroglobulin, RAGE-1, MN-CA IX, human telomerase reverse transcriptase, RU1, RU2 (AS), intestinal carboxyl esterase, mut hsp70-2, M-CSF, prostase, prostate-specific antigen (PSA), PAP, NY-ESO-1, LAGE-1a, p53, prostein, PSMA, Her2/neu, survivin and telomerase, prostate-carcinoma tumor antigen-1 (PCTA-1), MAGE, ELF2M, neutrophil elastase, ephrinB2, CD22, insulin growth factor (IGF)-I, IGF-II, IGF-I receptor and mesothelin.

In one embodiment, the tumor antigen comprises one or more antigenic cancer epitopes

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associated with a malignant tumor. Malignant tumors express a number of proteins that can serve as target antigens for an immune attack. These molecules include but are not limited to tissue-specific antigens such as MART-1, tyrosinase and GP 100 in melanoma and prostatic acid phosphatase (PAP) and prostate-specific antigen (PSA) in prostate cancer. Other target molecules belong to the group of transformation-related molecules such as the oncogene HER-2/Neu/ErbB-2. Yet another group of target antigens are onco-fetal antigens such as carcinoembryonic antigen (CEA). In B-cell lymphoma the tumor-specific idiotype immunoglobulin constitutes a truly tumor-specific immunoglobulin antigen that is unique to the individual tumor. B-cell differentiation antigens such as CD19, CD20 and CD37 are other candidates for target antigens in B-cell lymphoma. Some of these antigens (CEA, HER-2, CD19, CD20, idiotype) have been used as targets for passive immunotherapy with monoclonal antibodies with limited success.

The type of tumor antigen referred to in the invention may also be a tumor-specific antigen (TSA) or a tumor-associated antigen (TAA). A TSA is unique to tumor cells and does not occur on other cells in the body. A TAA associated antigen is not unique to a tumor cell and instead is also expressed on a normal cell under conditions that fail to induce a state of immunologic tolerance to the antigen. The expression of the antigen on the tumor may occur under conditions that enable the immune system to respond to the antigen. TAAs may be antigens that are expressed on normal cells during fetal development when the immune system is immature and unable to respond or they may be antigens that are normally present at extremely low levels on normal cells but which are expressed at much higher levels on tumor cells.

Non-limiting examples of TSA or TAA antigens include the following: Differentiation antigens such as MART-1/MelanA (MART-I), gp100 (Pmel 17), tyrosinase, TRP-1, TRP-2 and tumor-specific multilineage antigens such as MAGE-1, MAGE-3, BAGE, GAGE-1, GAGE-2, p15; overexpressed embryonic antigens such as CEA; overexpressed oncogenes and mutated tumor-suppressor genes such as p53, Ras, HER-2/neu; unique tumor antigens resulting from chromosomal translocations; such as BCR-ABL, E2A-PRL, H4-RET, IGH-IGK, MYL-RAR; and viral antigens, such as the Epstein Barr virus antigens EBVA and the human papillomavirus (HPV) antigens E6 and E7. Other large, protein-based antigens include TSP-180, MAGE-4, MAGE-5, MAGE-6, RAGE, NY-ESO, p185erbB2, p180erbB-3, c-met, nm-23H1, PSA, TAG-72, CA 19-9, CA 72-4, CAM 17.1, NuMa, K-ras, beta-Catenin, CDK4, Mum-1, p 15, p 16, 43-9F, 5T4, 791Tgp72, alpha-fetoprotein, beta-HCG, BCA225, BTAA, CA 125, CA 15-3\CA 27.29\BCAA, CA 195, CA 242, CA-50, CAM43, CD68\P1, CO-029, FGF-5, G250, Ga733\EpCAM, HTgp-175, M344, MA-50, MG7-Ag, MOV18, NB/70K, NY-CO-1, RCAS1, SDCCAG16, TA-90\Mac-2 binding

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protein\cyclophilin C-associated protein, TAAL6, TAG72, TLP, and TPS.

In a preferred embodiment, the antigen includes but is not limited to CD19, CD20, CD22, ROR1, Mesothelin, CD33/IL3Ra, c-Met, PSMA, Glycolipid F77, EGFRvIII, GD-2, MY-ESO-1 TCR, MAGE A3 TCR, and the like.

Combinations

In one embodiment, the composition of the invention comprises a combination of agents described herein. In certain embodiments, a composition comprising a combination of agents described herein has an additive effect, wherein the overall effect of the combination is approximately equal to the sum of the effects of each individual agent. In other embodiments, a composition comprising a combination of agents described herein has a synergistic effect, wherein the overall effect of the combination is greater than the sum of the effects of each individual agent.

A composition comprising a combination of agents comprises individual agents in any suitable ratio. For example, in one embodiment, the composition comprises a 1:1 ratio of two individual agents. However, the combination is not limited to any particular ratio. Rather any ratio that is shown to be effective is encompassed.

Conjugation

In various embodiments of the invention, the delivery vehicle is conjugated to a targeting domain. Exemplary methods of conjugation can include, but are not limited to, covalent bonds, electrostatic interactions, and hydrophobic ("van der Waals") interactions. In one embodiment, the conjugation is a reversible conjugation, such that the delivery vehicle can be disassociated from the targeting domain upon exposure to certain conditions or chemical agents. In another embodiment, the conjugation is an irreversible conjugation, such that under normal conditions the delivery vehicle does not dissociate from the targeting domain.

In some embodiments, the conjugation comprises a covalent bond between an activated polymer conjugated lipid and the targeting domain. The term "activated polymer conjugated lipid" refers to a molecule comprising a lipid portion and a polymer portion that has been activated via functionalization of a polymer conjugated lipid with a first coupling group. In one embodiment, the activated polymer conjugated lipid comprises a first coupling group capable of reacting with a second coupling group. In one embodiment, the activated polymer conjugated lipid is an activated pegylated lipid. In one embodiment, the first coupling group is bound to the lipid portion of the pegylated lipid. In another embodiment, the first coupling group is bound to the polyethylene glycol

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portion of the pegylated lipid. In one embodiment, the second functional group is covalently attached to the targeting domain.

The first coupling group and second coupling group can be any functional groups known to those of skill in the art to together form a covalent bond, for example under mild reaction conditions or physiological conditions. In some embodiments, the first coupling group or second coupling group are selected from the group consisting of maleimides, N-hydroxysuccinimide (NHS) esters, carbodiimides, hydrazide, pentafluorophenyl (PFP) esters, phosphines, hydroxymethyl phosphines, psoralen, imidoesters, pyridyl disulfide, isocyanates, vinyl sulfones, alpha-haloacetyls, aryl azides, acyl azides, alkyl azides, diazirines, benzophenone, epoxides, carbonates, anhydrides, sulfonyl chlorides, cyclooctyne, aldehydes, and sulfhydryl groups. In some embodiments, the first coupling group or second coupling group is selected from the group consisiting of free amines ($-NH_2$), free sulfhydryl groups (–SH), free hydroxide groups (–OH), carboxylates, hydrazides, and alkoxyamines. In some embodiments, the first coupling group is a functional group that is reactive toward sulfhydryl groups, such as maleimide, pyridyl disulfide, or a haloacetyl. In one embodiment, the first coupling group is a maleimide.

In one embodiment, the second coupling group is a sulfhydryl group. The sulfhydryl group can be installed on the targeting domain using any method known to those of skill in the art. In one embodiment, the sulfhydryl group is present on a free cysteine residue. In one embodiment, the sulfhydryl group is revealed via reduction of a disulfide on the targeting domain, such as through reaction with 2-mercaptoethylamine. In one embodiment, the sulfhydryl group is installed via a chemical reaction, such as the reaction between a free amine and 2-iminothilane or N-succinimidyl S-acetylthioacetate (SATA).

In some embodiments, the polymer conjugated lipid and targeting domain are functionalized with groups used in "click" chemistry. Bioorthogonal "click" chemistry comprises the reaction between a functional group with a 1,3-dipole, such as an azide, a nitrile oxide, a nitrone, an isocyanide, and the link, with an alkene or an alkyne dipolarophiles. Exemplary dipolarophiles include any strained cycloalkenes and cycloalkynes known to those of skill in the art, including, but not limited to, cyclooctynes, dibenzocyclooctynes, monofluorinated cyclcooctynes, difluorinated cyclooctynes, and biarylazacyclooctynone

Pharmaceutical Compositions

The formulations of the pharmaceutical compositions described herein may be prepared by any method known or hereafter developed in the art of pharmacology. In general, such preparatory

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methods include the step of bringing the active ingredient into association with a carrier or one or more other accessory ingredients, and then, if necessary or desirable, shaping or packaging the product into a desired single- or multi-dose unit.

Although the description of pharmaceutical compositions provided herein are principally directed to pharmaceutical compositions which are suitable for ethical administration to humans, it will be understood by the skilled artisan that such compositions are generally suitable for administration to animals of all sorts. Modification of pharmaceutical compositions suitable for administration to humans in order to render the compositions suitable for administration to various animals is well understood, and the ordinarily skilled veterinary pharmacologist can design and perform such modification with merely ordinary, if any, experimentation. Subjects to which administration of the pharmaceutical compositions of the invention is contemplated include, but are not limited to, humans and other primates, mammals including commercially relevant mammals such as non-human primates, cattle, pigs, horses, sheep, cats, and dogs.

Pharmaceutical compositions that are useful in the methods of the invention may be prepared, packaged, or sold in formulations suitable for ophthalmic, oral, rectal, vaginal, parenteral, topical, pulmonary, intranasal, buccal, intravenous, intracerebroventricular, intradermal, intramuscular, or another route of administration. Other contemplated formulations include projected nanoparticles, liposomal preparations, resealed erythrocytes containing the active ingredient, and immunogenic-based formulations.

A pharmaceutical composition of the invention may be prepared, packaged, or sold in bulk, as a single unit dose, or as a plurality of single unit doses. As used herein, a "unit dose" is discrete amount of the pharmaceutical composition comprising a predetermined amount of the active ingredient. The amount of the active ingredient is generally equal to the dosage of the active ingredient which would be administered to a subject or a convenient fraction of such a dosage such as, for example, one-half or one-third of such a dosage.

The relative amounts of the active ingredient, the pharmaceutically acceptable carrier, and any additional ingredients in a pharmaceutical composition of the invention will vary, depending upon the identity, size, and condition of the subject treated and further depending upon the route by which the composition is to be administered. By way of example, the composition may comprise between 0.1% and 100% (w/w) active ingredient.

In addition to the active ingredient, a pharmaceutical composition of the invention may further comprise one or more additional pharmaceutically active agents.

Controlled- or sustained-release formulations of a pharmaceutical composition of the

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invention may be made using conventional technology.

As used herein, "parenteral administration" of a pharmaceutical composition includes any route of administration characterized by physical breaching of a tissue of a subject and administration of the pharmaceutical composition through the breach in the tissue. Parenteral administration thus includes, but is not limited to, administration of a pharmaceutical composition by injection of the composition, by application of the composition through a surgical incision, by application of the composition through a tissue-penetrating non-surgical wound, and the like. In particular, parenteral administration is contemplated to include, but is not limited to, intraocular, intravitreal, subcutaneous, intraperitoneal, intramuscular, intradermal, intrasternal injection, intratumoral, intravenous, intracerebroventricular and kidney dialytic infusion techniques.

Formulations of a pharmaceutical composition suitable for parenteral administration comprise the active ingredient combined with a pharmaceutically acceptable carrier, such as sterile water or sterile isotonic saline. Such formulations may be prepared, packaged, or sold in a form suitable for bolus administration or for continuous administration. Injectable formulations may be prepared, packaged, or sold in unit dosage form, such as in ampules or in multi-dose containers containing a preservative. Formulations for parenteral administration include, but are not limited to, suspensions, solutions, emulsions in oily or aqueous vehicles, pastes, and implantable sustained-release or biodegradable formulations. Such formulations may further comprise one or more additional ingredients including, but not limited to, suspending, stabilizing, or dispersing agents. In one embodiment of a formulation for parenteral administration, the active ingredient is provided in dry (i.e. powder or granular) form for reconstitution with a suitable vehicle (e.g. sterile pyrogen-free water) prior to parenteral administration of the reconstituted composition.

The pharmaceutical compositions may be prepared, packaged, or sold in the form of a sterile injectable aqueous or oily suspension or solution. This suspension or solution may be formulated according to the known art, and may comprise, in addition to the active ingredient, additional ingredients such as the dispersing agents, wetting agents, or suspending agents described herein. Such sterile injectable formulations may be prepared using a non-toxic parenterally-acceptable diluent or solvent, such as water or 1,3-butane diol, for example. Other acceptable diluents and solvents include, but are not limited to, Ringer's solution, isotonic sodium chloride solution, and fixed oils such as synthetic mono- or di-glycerides. Other parentally-administrable formulations which are useful include those which comprise the active ingredient in microcrystalline form, in a liposomal preparation, or as a component of a biodegradable polymer systems. Compositions for sustained release or implantation may comprise pharmaceutically

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acceptable polymeric or hydrophobic materials such as an emulsion, an ion exchange resin, a sparingly soluble polymer, or a sparingly soluble salt.

A pharmaceutical composition of the invention may be prepared, packaged, or sold in a formulation suitable for pulmonary administration via the buccal cavity. Such a formulation may comprise dry particles which comprise the active ingredient and which have a diameter in the range from about 0.5 to about 7 nanometers, and preferably from about 1 to about 6 nanometers. Such compositions are conveniently in the form of dry powders for administration using a device comprising a dry powder reservoir to which a stream of propellant may be directed to disperse the powder or using a self-propelling solvent/powder-dispensing container such as a device comprising the active ingredient dissolved or suspended in a low-boiling propellant in a sealed container. Preferably, such powders comprise particles wherein at least 98% of the particles by weight have a diameter greater than 0.5 nanometers and at least 95% of the particles by number have a diameter less than 7 nanometers. More preferably, at least 95% of the particles by weight have a diameter greater than 1 nanometer and at least 90% of the particles by number have a diameter less than 6 nanometers. Dry powder compositions preferably include a solid fine powder diluent such as sugar and are conveniently provided in a unit dose form.

Low boiling propellants generally include liquid propellants having a boiling point of below 65°F at atmospheric pressure. Generally the propellant may constitute 50 to 99.9% (w/w) of the composition, and the active ingredient may constitute 0.1 to 20% (w/w) of the composition. The propellant may further comprise additional ingredients such as a liquid non-ionic or solid anionic surfactant or a solid diluent (preferably having a particle size of the same order as particles comprising the active ingredient).

Formulations of a pharmaceutical composition suitable for parenteral administration comprise the active ingredient combined with a pharmaceutically acceptable carrier, such as sterile water or sterile isotonic saline. Such formulations may be prepared, packaged, or sold in a form suitable for bolus administration or for continuous administration. Injectable formulations may be prepared, packaged, or sold in unit dosage form, such as in ampules or in multi-dose containers containing a preservative. Formulations for parenteral administration include, but are not limited to, suspensions, solutions, emulsions in oily or aqueous vehicles, pastes, and implantable sustained-release or biodegradable formulations. Such formulations may further comprise one or more additional ingredients including, but not limited to, suspending, stabilizing, or dispersing agents. In one embodiment of a formulation for parenteral administration, the active ingredient is provided in dry (i.e., powder or granular) form for reconstitution with a suitable vehicle (e.g., sterile

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pyrogen-free water) prior to parenteral administration of the reconstituted composition.

The pharmaceutical compositions may be prepared, packaged, or sold in the form of a sterile injectable aqueous or oily suspension or solution. This suspension or solution may be formulated according to the known art, and may comprise, in addition to the active ingredient, additional ingredients such as the dispersing agents, wetting agents, or suspending agents described herein. Such sterile injectable formulations may be prepared using a non-toxic parenterally-acceptable diluent or solvent, such as water or 1,3-butane diol, for example. Other acceptable diluents and solvents include, but are not limited to, Ringer's solution, isotonic sodium chloride solution, and fixed oils such as synthetic mono- or di-glycerides. Other parentally-administrable formulations that are useful include those that comprise the active ingredient in microcrystalline form, in a liposomal preparation, or as a component of a biodegradable polymer system. Compositions for sustained release or implantation may comprise pharmaceutically acceptable polymeric or hydrophobic materials such as an emulsion, an ion exchange resin, a sparingly soluble polymer, or a sparingly soluble salt.

As used herein, "additional ingredients" include, but are not limited to, one or more of the following: excipients; surface active agents; dispersing agents; inert diluents; granulating and disintegrating agents; binding agents; lubricating agents; sweetening agents; flavoring agents; coloring agents; preservatives; physiologically degradable compositions such as gelatin; aqueous vehicles and solvents; oily vehicles and solvents; suspending agents; dispersing or wetting agents; emulsifying agents, demulcents; buffers; salts; thickening agents; fillers; emulsifying agents; antioxidants; antibiotics; antifungal agents; stabilizing agents; and pharmaceutically acceptable polymeric or hydrophobic materials. Other "additional ingredients" which may be included in the pharmaceutical compositions of the invention are known in the art and described, for example in Remington's Pharmaceutical Sciences (1985, Genaro, ed., Mack Publishing Co., Easton, PA), which is incorporated herein by reference.

Treatment Methods

The invention provides methods of delivering an agent to a cell, tissue, or organ of a subject. In some embodiments, the agent is a diagnostic agent to detect at least one marker associated with a disease or disorder. In some embodiments, the agent is a therapeutic agent for the treatment or prevention of a disease or disorder. Therefore, in some embodiments, the invention provides methods for diagnosing, treating or preventing a disease or disorder comprising

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administering an effective amount of a composition comprising one or more diagnostic or therapeutic agents, one or more adjuvants, or a combination thereof.

In some embodiments, the method provides immunity in the subject to an infection, disease, or disorder associated with an antigen. The invention thus provides a method of treating or preventing the infection, disease, or disorder associated with the antigen. For example, the method may be used to treat or prevent a viral infection, bacterial infection, fungal infection, parasitic infection, or cancer, depending upon the type of antigen of the administered composition. Exemplary antigens and associated infections, diseases, and tumors are described elsewhere herein.

In one embodiment, the composition is administered to a subject having an infection, disease, or cancer associated with the antigen. In one embodiment, the composition is administered to a subject at risk for developing the infection, disease, or cancer associated with the antigen. For example, the composition may be administered to a subject who is at risk for being in contact with a virus, bacteria, fungus, parasite, or the like. In one embodiment, the composition is administered to a subject who has increased likelihood, though genetic factors, environmental factors, or the like, of developing cancer.

In one embodiment, the method comprises administering a composition comprising one or more nucleoside-modified nucleic acid molecules encoding one or more antigens and one or more adjuvant. In one embodiment, the method comprises administering a composition comprising a first nucleoside-modified nucleic acid molecule encoding one or more antigens and a second nucleoside-modified nucleic acid molecule encoding one or more adjuvants. In one embodiment, the method comprises administering a first composition comprising one or more nucleoside-modified nucleic acid molecules encoding one or more antigens and administering a second composition comprising one or more nucleoside-modified nucleic acid molecules encoding one or more adjuvants.

In certain embodiments, the method comprises administering to subject a plurality of nucleoside-modified nucleic acid molecules encoding a plurality of antigens, adjuvants, or a combination thereof.

In certain embodiments, the method of the invention allows for sustained expression of the antigen or adjuvant, described herein, for at least several days following administration. However, the method, in certain embodiments, also provides for transient expression, as in certain embodiments, the nucleic acid is not integrated into the subject genome.

In certain embodiments, the method comprises administering nucleoside-modified RNA which provides stable expression of the antigen or adjuvant described herein. In some embodiments, administration of nucleoside-modified RNA results in little to no innate immune

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response, while inducing an effective adaptive immune response.

Administration of the compositions of the invention in a method of treatment can be achieved in a number of different ways, using methods known in the art. In one embodiment, the method of the invention comprises systemic administration of the subject, including for example enteral or parenteral administration. In certain embodiments, the method comprises intradermal delivery of the composition. In another embodiment, the method comprises intravenous delivery of the composition. In some embodiments, the method comprises intramuscular delivery of the composition. In one embodiment, the method comprises subcutaneous delivery of the composition. In one embodiment, the method comprises inhalation of the composition. In one embodiment, the method comprises intranasal delivery of the composition.

It will be appreciated that the composition of the invention may be administered to a subject either alone, or in conjunction with another agent.

The therapeutic and prophylactic methods of the invention thus encompass the use of pharmaceutical compositions encoding an antigen, adjuvant, or a combination thereof, described herein to practice the methods of the invention. The pharmaceutical compositions useful for practicing the invention may be administered to deliver a dose of from ng/kg/day and 100 mg/kg/day. In one embodiment, the invention envisions administration of a dose which results in a concentration of the compound of the invention from 10nM and 10 μM in a mammal.

Typically, dosages which may be administered in a method of the invention to a mammal, preferably a human, range in amount from 0.01 μg to about 50 mg per kilogram of body weight of the mammal, while the precise dosage administered will vary depending upon any number of factors, including but not limited to, the type of mammal and type of disease state being treated, the age of the mammal and the route of administration. Preferably, the dosage of the compound will vary from about 0.1 μg to about 10 mg per kilogram of body weight of the mammal. More preferably, the dosage will vary from about 1 μg to about 1 mg per kilogram of body weight of the mammal.

The composition may be administered to a mammal as frequently as several times daily, or it may be administered less frequently, such as once a day, once a week, once every two weeks, once a month, or even less frequently, such as once every several months or even once a year or less. The frequency of the dose will be readily apparent to the skilled artisan and will depend upon any number of factors, such as, but not limited to, the type and severity of the disease being treated, the type and age of the mammal, etc.

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In certain embodiments, administration of an immunogenic composition or vaccine of the invention may be performed by single administration or boosted by multiple administrations.

In one embodiment, the invention includes a method comprising administering one or more compositions encoding one or more antigens or adjuvants described herein. In certain embodiments, the method has an additive effect, wherein the overall effect of the administering the combination is approximately equal to the sum of the effects of administering each antigen or adjuvant. In other embodiments, the method has a synergistic effect, wherein the overall effect of administering the combination is greater than the sum of the effects of administering each antigen or adjuvant.

Treatment Methods for SARS-CoV-2

In one embodiment, the invention provides methods of inducing an adaptive immune response against SARS-CoV-2 in a subject comprising administering an effective amount of a composition comprising an amphiphilic Janus dendrimer of Formula (I) and one or more isolated nucleic acids encoding one or more SARS-CoV-2 antigens.

In one embodiment, the method provides immunity in the subject to SARS-CoV-2, SARS-CoV-2 infection, or to a disease or disorder associated with SARS-CoV-2. The invention thus provides a method of treating or preventing the infection, disease, or disorder associated with SARS-CoV-2. In one embodiment, the disease or disorder associated with SARS-CoV-2 is COVID-19 or a comorbidity of COVID-19.

In some embodiments, the invention is a method of administering to a subject a composition comprising at least one nucleoside-modified RNA encoding at least one SARS-CoV-2 antigen.

In one embodiment, the composition is administered to a subject having an infection, disease, or disorder associated with SARS-CoV-2. In one embodiment, the composition is administered to a subject at risk for developing the infection, disease, or disorder associated with SARS-CoV-2. For example, the composition may be administered to a subject who is at risk for being in contact with a SARS-CoV-2. In one embodiment, the composition is administered to a subject who lives in, traveled to, or is expected to travel to a geographic region in which SARS-CoV-2 is prevalent. In one embodiment, the composition is administered to a subject who is in contact with or expected to be in contact with another person who lives in, traveled to, or is expected to travel to a geographic region in which SARS-CoV-2 is prevalent. In one embodiment, the composition is administered to a subject who has knowingly been exposed to SARS-CoV-2

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through their occupation or contact.

In one embodiment, the method comprises administering a composition comprising at least one dendrimer of Formula (I) and one or more nucleoside-modified nucleic acid molecules encoding one or more SARS-CoV-2 antigens and one or more adjuvant. In one embodiment, the method comprises administering a composition comprising a first nucleoside-modified nucleic acid molecule encoding one or more SARS-CoV-2 antigens and a second nucleoside-modified nucleic acid molecule encoding one or more adjuvants. In one embodiment, the method comprises administering a first composition comprising one or more nucleoside-modified nucleic acid molecules encoding one or more SARS-CoV-2 antigens and administering a second composition comprising one or more nucleoside-modified nucleic acid molecules encoding one or more adjuvants.

In certain embodiments, the method comprises administering to subject a plurality of nucleoside-modified nucleic acid molecules encoding a plurality of SARS-CoV-2 antigens, adjuvants, or a combination thereof.

In certain embodiments, the method of the invention allows for sustained expression of the SARS-CoV-2 antigen or adjuvant, described herein, for at least several days following administration. In certain embodiments, the method of the invention allows for sustained expression of the SARS-CoV-2 antigen or adjuvant, described herein, for at least 2 weeks following administration. In certain embodiments, the method of the invention allows for sustained expression of the SARS-CoV-2 antigen or adjuvant, described herein, for at least 1 month following administration. However, the method, in certain embodiments, also provides for transient expression, as in certain embodiments, the nucleic acid is not integrated into the subject genome.

In certain embodiments, the method comprises administering nucleoside-modified RNA, which provides stable expression of the SARS-CoV-2 antigen or adjuvant described herein. In some embodiments, administration of nucleoside-modified RNA results in little to no innate immune response, while inducing an effective adaptive immune response.

In certain embodiments, the method provides sustained protection against SARS-CoV-2. For example, in certain embodiments, the method provides sustained protection against SARS-CoV-2 for more than 2 weeks. In certain embodiments, the method provides sustained protection against SARS-CoV-2 for 1 month or more. In certain embodiments, the method provides sustained protection against SARS-CoV-2 for 2 months or more. In certain embodiments, the method provides sustained protection against SARS-CoV-2 for 3 months or more. In certain embodiments, the method provides sustained protection against SARS-CoV-2 for 4 months or more. In certain

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embodiments, the method provides sustained protection against SARS-CoV-2 for 5 months or more. In certain embodiments, the method provides sustained protection against SARS-CoV-2 for 6 months or more. In certain embodiments, the method provides sustained protection against SARS-CoV-2 for 1 year or more.

In one embodiment, a single immunization of the composition induces a sustained protection against SARS-CoV-2 for 1 month or more, 2 months or more, 3 months or more, 4 months or more, 5 months or more, 6 months or more, or 1 year or more.

Administration of the compositions of the invention in a method of treatment can be achieved in a number of different ways, using methods known in the art. In one embodiment, the method of the invention comprises systemic administration of the subject, including for example enteral or parenteral administration. In certain embodiments, the method comprises intradermal delivery of the composition. In another embodiment, the method comprises intravenous delivery of the composition. In some embodiments, the method comprises intramuscular delivery of the composition. In one embodiment, the method comprises subcutaneous delivery of the composition. In one embodiment, the method comprises inhalation of the composition. In one embodiment, the method comprises intranasal delivery of the composition.

It will be appreciated that the composition of the invention may be administered to a subject either alone, or in conjunction with another agent.

The therapeutic and prophylactic methods of the invention thus encompass the use of pharmaceutical compositions encoding a SARS-CoV-2 antigen, adjuvant, or a combination thereof, described herein to practice the methods of the invention. The pharmaceutical compositions useful for practicing the invention may be administered to deliver a dose of from 1 ng/kg/day and 100 mg/kg/day. In one embodiment, the invention envisions administration of a dose, which results in a concentration of the compound of the invention from 10 nM and 10 μM in a mammal.

Typically, dosages which may be administered in a method of the invention to a mammal, such as a human, range in amount from 0.01 μg to about 50 mg per kilogram of body weight of the mammal, while the precise dosage administered will vary depending upon any number of factors, including but not limited to, the type of mammal and type of disease state being treated, the age of the mammal and the route of administration. In certain embodiments, the dosage of the compound will vary from about 0.1 μg to about 10 mg per kilogram of body weight of the mammal. In certain embodiments, the dosage will vary from about 1 μg to about 1 mg per kilogram of body weight of the mammal.

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The composition may be administered to a mammal as frequently as several times daily, or it may be administered less frequently, such as once a day, once a week, once every two weeks, once a month, or even less frequently, such as once every several months or even once a year or less. The frequency of the dose will be readily apparent to the skilled artisan and will depend upon any number of factors, such as, but not limited to, the type and severity of the disease being treated, the type and age of the mammal, etc.

In certain embodiments, administration of an immunogenic composition or vaccine of the invention may be performed by single administration or boosted by multiple administrations.

In one embodiment, the invention includes a method comprising administering one or more compositions encoding one or more SARS-CoV-2 antigens or adjuvants described herein. In certain embodiments, the method has an additive effect, wherein the overall effect of the administering the combination is approximately equal to the sum of the effects of administering each SARS-CoV-2 antigen or adjuvant. In other embodiments, the method has a synergistic effect, wherein the overall effect of administering the combination is greater than the sum of the effects of administering each SARS-CoV-2 antigen or adjuvant.

EXPERIMENTAL EXAMPLES

The invention is further described in detail by reference to the following experimental examples. These examples are provided for purposes of illustration only and are not intended to be limiting unless otherwise specified. Thus, the invention should in no way be construed as being limited to the following examples, but rather, should be construed to encompass any and all variations which become evident as a result of the teaching provided herein.

Without further description, it is believed that one of ordinary skill in the art can, using the preceding description and the following illustrative examples, make and utilize the invention and practice the claimed methods. The following working examples therefore are not to be construed as limiting in any way the remainder of the disclosure.

Example 1: One-Component Multifunctional Sequence-Defined Ionizable Amphiphilic Janus Dendrimer (IAJD) Delivery Systems of mRNA for Vaccines and Drugs

The present invention relates in part to a one-component multifunctional sequence-defined ionizable amphiphilic Janus dendrimer (IAJD) delivery system that co-assembles with mRNA by simple injection into dendrimersome nanoparticles (DNPs) (Figure 1). Screening experiments with six libraries containing 52 IAJDs was performed both *in-vitro* and *in vivo*. They demonstrated the

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proof of concept of DNPs, their potential applications and utility as a model to elucidate fundamental aspects of nonviral vectors.

One of the major advantages of the synthetic vectors used for the delivery of mRNA consist in their unlimited synthetic capabilities. The transformation of the four-component LNP (Figure 2) into a one-component DNP represents a demonstration of this synthetic capability.

Figure 2 illustrates the process involved in the assembly of four-component LNPs. The four-components composition containing various ratios of the ionizable lipid, phospholipid, PEG-lipid and cholesterol is prepared as a solution in ethanol. This ethanol solution is mixed with a microfluidic device with a pH 3 to 5 buffer solution and with an aqueous solution of mRNA. The mRNA used in the acidic buffer solution is produced and stored in neutral water. The resulting nanoparticles are analyzed by dynamic light scattering (DLS) to determine the diameter (D, nm) and the polydispersity (PDI) of the LNPs, dialyzed to pH 7.4, analyzed by DLS again and stored at -70 °C before the *in vitro* or *in vivo* experiments are performed. Figure 1 illustrates the same process for the one-component DNPs. The IAJD containing an ionizable amine incorporated in a precise sequence is dissolved in ethanol. The ethanol solution is injected into an acidic buffer solution containing mRNA (pH 3 to 5.2). Depending on the original pH of the buffer the resulting DNPs containing mRNA have already a pH between 4.5 and 7.3 and after DLS analysis can be used for *in vitro* and *in vivo* experiment without dialysis. Long time storage of DNPs is at 5 °C. Figure 1 also illustrates the transition from the extracellular to the intracellular process for both LNPs and DNPs. Once injected both LNP and DNP approach the corresponding cells and get encapsulated *via* endocytosis. The extracellular pH is 7.4 and therefore LNPs and DNPs enter the cell with an almost neutral surface. Then the endocytosis of LNPs or DNPs deposits them into endosomes, the pH of which decreases from 6.8 to 4.5 during their morphing into lysosomes due to the ATP-dependent proton pumps on the endosomal membrane (Huotari and Helenius, EMBO J. 2011, 30, 3481–3500). Therefore, both LNPs and DNPs get re-protonated, interact with the naturally occurring anionic lipids and release the mRNA that helps the ribosome to generate new proteins. When the ionizable amines from LNPs are segregated in their center, re-protonation of their periphery cannot occur and therefore, the release of mRNA in the cell has a very low efficiency (1-2%).

Accelerated Modular-Orthogonal Synthesis of Six Libraries of IAJDs: Modular-orthogonal methodologies were employed for the synthesis of sequence-defined amphiphilic Janus glycodendrimers (JGDs). Accelerated modular-orthogonal methodologies (Figure 3 and Figure 4) for the synthesis of single-single (a single hydrophobic combined with a single hydrophilic

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dendrons), twin-twin (two identical hydrophobic and two identical hydrophilic dendrons), and hybrid twin-mix (two identical hydrophobic and two different hydrophilic dendrons), rely on related but improved and accelerated synthetic principles originally employed for the synthesis of sequence-defined JGDs. Two different orthogonal protective groups, 4-methoxybenzyl ether and benzyl ether, were employed in this methodology (Horita, et al., Tetrahedron 1986, 42, 3021−3028). The six libraries synthesized are schematically shown in Figure 4. Four of these libraries are based on single-single IAJDs. Nine IAJDs are available in library 1, seven in library 2, six in library 3 and three in library 4. Library 5 is based on the twin-twin IAJDs generated from IAJD1 to IAJD9 of library 1 and JAJD33 of library 3. Library 6 contains 17 hybrid twin-mix IAJDs selected from all libraries. The selection process was determined by the activity *in vivo* and *in vitro* of their single-single components. The hydrophilic parts of these IAJDs contain sequence-defined compositions based on the dimethylaminobutanoate (DMBA), dimethylaminopropanoate (DMPA), dimethylaminoacetate (DMA), piperidinebutanoate (PIP) and methylpiperazinebutanoate (MPRZ) ionizable amines (Figure 3). They were selected based on the pK_a of their corresponding ionizable lipids available in the literature (Ramishetti, et al., Adv. Mater. 2020, 32, 1906128; Kim, et al., Sci. Adv. 2021, 7, eabf4398). The symbols employed in the schematic representation of these sequence-defined IAJDs are shown in the square from the bottom part of Figure 4. A benzyl ether group, marked in red, was employed to isolate the ionizable amines and construct different sequences in the hydrophilic part of the IAJDs. Aromatic groups such as benzyl ethers are suggested to interact as hydrogen-bond acceptors in molecular recognition with cations including ammonium groups both in biology and in synthetic supramolecular chemistry (Levitt, et al., Philos. Trans. R. Soc., A. 1993, 345, 105 112; Perutz, , et al., J. Am. Chem. Soc. 1986, 108, 1064 1078; Loewenthal, et al., J. Mol. Biol. 1992, 224, 759–770; Zacharias, et al., Trends Pharmacol. Sci. 2002, 23, 281–287; Gallivan, et al., Proc. Natl. Acad. Sci. U.S.A. 1999, 96, 9459–9464; Dougherty, Science 1996, 271, 163−168; Dougherty, Acc. Chem. Res. 2013, 46, 885–893; Ma, et al., Chem. Rev. 1997, 97, 1303–1324; Dougherty, et al., Science 1990, 250, 1558–1560). This cation-π interaction is weaker than traditional H-bonds (about 3 kcal/mol) and therefore, can mediate a dynamic control of the pK_a of the ionizable amines. In the protonated state of the amine, it can increase its pK_a while in the non-protonated state it may facilitate an interaction of the benzyl ether with the nucleic bases of the mRNA to enhance co-assembly, or segregate in the hydrophobic part of the DNPs. This cation-π interaction has not been used previously in delivery vectors for mRNA. The hydrophobic parts of the IAJDs contains both linear and branched alkyl groups of different length (Figure 3 and Figure 4). The hydrophilic acid components of these IAJDs are shown in Figure 3 (Module A); while their

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synthesis is described elsewhere herein. The structures of the hydrophobic benzyl amines are shown in Figure 3 (Module C). Their synthesis is also described elsewhere herein. Combining in an orthogonal way all these modules as illustrated in Figure 4 provides, in an accelerated way, the 6 libraries of IAJDs. The synthesis of the twin-twin part of the hydrophobic IAJDs is described elsewhere herein. The detailed structures of all libraries of IAJDs together with their pK_a values and short names are presented in Figure 12 (libraries 1,2,3,4), Figure 13 (library 5), and Figure 14 (library 6).

In Vitro Transfection Activity of DNPs in Human Embrionic Kidney (HEK) 293T Cells: HEK293T cells were seeded into 96-well plates (20,000 cells/well/200 μL) and cultured for 24 h at 37 °C in 5 % CO_2 complete cell culture media. Screening experiments were performed with nonoptimized DNPs containing naked nucleoside-modified mRNA (Karikó, et al., Immunity 2005, 23, 165–175; Pardi, et al., Methods Mol. Biol. 2013, 969, 29–42), of molar mass 664,341 encoding firefly luciferase (mRNA-Luc). A constant concentration of mRNA-Luc of 125 ng/well was used. The *trans*IT (TransIT-mRNA transfection Kit from Mirus Bio) and MC3-based LNPs (MC3: DLin-MC3-DMA, which is the FDA approved LNP for mRNA delivery) were used as positive controls for cell transfection at a concentration of mRNA-Luc of 125 ng/well, identical to that of the tested DNPs. Subsequently cells were cultured for 24 h, the medium was aspirated under reduced pressure by a glass pipette and cells were lysed with 30 μL /well of cell culture lysis reagent (Promega). The luminescence intensity corresponding to the luciferase protein expressed was characterized and analyzed.

All 52 IAJDs from Figure 4 were co-assembled by injection with mRNA in DNPs and were used in these experiments. Table 1 through Table 5 (vida infra) summarize the conditions employed for the preparation of the DNPs, their diameter (D, in nm) and polydispersity (PDI) as obtained by DLS experiments and the luminescence results. Each experiment was performed at least three times (Figure 5). Out of 52 DNPs, 42 (81 %) showed activity *in vitro* (Figure 5). Without any optimization two of them DNP9 and DNP 22 display higher activity than the positive controls MC3 and *Trans*IT, while four of them DNP8, DNP9, DNP21 and DNP22 show higher activity than the most commonly used positive control MC3.

In vivo mRNA Delivery in Mice with DNPs: Six to eight weeks old female mice were used in these experiments. Four to seven hours after injection with 100 μL solution of DNP encapsulated with 10 μg of mRNA-Luc the mice were imaged 10 min after intraperitoneal injection with D-Luciferin, 15 mg/mL at 10 μL/g of body weight. The exposure time was 1 min. For imaging of the organs, mice were sacrified, the organs were immediately collected and bioluminescence imaging

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was performed. Table 7 (vida infra) summarizes all DNPs injection assembly data including D in nm and PDI determined by DLS together with the resulting results obrained *in vivo*. Figure 6 summarizes all mice experiments including the D in nm and PDI of DNPs (both in black on top of the mice image), pK_a values of the correspoonding IAJDs (in blue also on top) of all compounds used for delivery. The results from Figures 6 and 7 provide a proof of concept for the one-component multifunctional sequence-defined ionizable amphiphilic Janus dendrimers delivery system for mRNA.

The summary of the results from Figure 7 demonstrate that there seem to be no correlation between the activity of the DNPs *in-vivo* and the same experiments *in vitro* (Figure 5) and that the most active IAJDs 33, 34 and 31 do not have the lowest pK_a values. The results from Figure 6 also demonstrate that there is quite a tolerance of the activity of DNPs to their diameters and polydispersity. Larger than 100 nm diameter seem to be as active as lower than 100 nm diameter DNPs (Figure 6). However quantitative correlations for the dependences of activity with pK_a and DNPs dimensions must be studied. The stability of the dimension of 40 DNPs was investigated as a function of time at 5 °C. Unoptimized experiments showed that the dimensions of 19 out of 40 DNPs were very stable after being stored at 5 °C for up to 120 days. Almost all these DNPs were assembled from IAJDs containing a benzyl ether in their hydrophilic part (Table 9, vida infra). A very interesting series of activity trends is observed when comparing the data from Figures 7 and 11. As shown in Figure 11, the stability of the dimensions of the DNPs is excellent for up to 120 days when they are stored at 5 °C. IAJD9, IAJD22, IAJD 33 and IAJD34 single-single compounds form remarkably stable DNPSs that also show high *in-vivo* activity (Figure 7). IAJD46 is the twin-twin of IAJD33. However the activity of the DNP36 is about half that of DNP33 (Figures 6 and 7). The hybrid twin-mix IAJD47 is also based on the structure of IAJD33 or half of the structure of IAJD46. However, the activity of the DNP47 is much lower than both that of DNP33 and DNP46. Even more interesting is the comparison of single-single IAJD9 with the hybrid twin-mix IAJD32 (Figures 6 and 7).

Figure 7 plots the results from Figure 6. Without any optimization, out of the 52 DNPs investigated, 28 (54 %) show activity in vivo. Two of them, DNP33 and DNP34 show very high activity in lung. Single-single IAJD9 derived DNP9 exhibits good activity (Figure 7) and stability (Figure 11). At the same time the corresponding hybrid twin-mix IAJD32 based DNP32 that is based on IAJD9 and a single PEG of degree of polymerization 45 (Figures 3 and 4) exhibits also excellent stability (Figure 11) but it is completely inactive in mice (Figure 7). In order to clarify this result, several co-assembled DNPs were prepared based on very active IAJDs and a small

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concentration of IAJD32. One example is the DNP assembled from IAJD33 and 2% IAJD32. (Figure 11, bottom row, second from left) and Figure 7. The stability of this combined DNP is excellent (Figure 11) and is comparable with that of DNP32 assembled from IAJD32 alone. However, the in vivo activity of DNP co-assembled from IAJD33 with 2% IAJD32 is only a small fraction of the activity of the DNP33 (Figure 7). This confirms the mechanism of the PEG enigma. Therefore, while insertion of a small fraction of PEG-conjugated to an IAJD can increase dramatically the stability of the resulting DNP it also decreases even more dramatically its activity *in vivo*. Incorporation of short oligooxyethylene fragments in the structure of single-single, twin-twin or even hybrid twin-mix IAJDs may serve to address this issue.

The Role of Ionizable Amine Concentration and Sequence on the Activity of the Corresponding DNPs: The binding activity of sugars located on the surface of the glycodendrimersomes (GDSs) assembled from amphiphilic Janus glycodendrimers (JGDs) toward sugar binding proteins increases by decreasing the concentration of the sugar in a sequence-defined process (Percec, et al., J. Am. Chem. Soc. 2013, 135, 9055−9077). This unexpected trend was explained by self-organization on the periphery of the glycodendrimersome of a morphology that facilitated higher binding activity between the sugar and the proteins at lower concentrations of sugar.

Figure 8 plots representative activity data for the sequence-defined IAJDs derived DNPs both *in vitro* and *in vivo* experiments. Low or no activity was observed at high concentrations of ionizable amines in the structure of the IAJD and extremely high activities were observed at lower ionizable amine concentration in very specific sequences. Without discussing in great details, the results from Figure 8, provide a mechanism to engineer activity of NDPs via the sequence and concentration of their ionizable amines. The change in activity observed in Figure 8 is much higher than that observed in the case of sequence-defined dendrimersomes.

Some Comments On Potential Targeted Delivery of mRNA: Figure 9 and Figure 10 summarize representative organ delivery data selected from the experiments reported in Figure 6. They illustrate the luminescence intensity reflecting delivery activity in heart, lung, liver and spleen as a function of the structure of the IAJD employed in the design of the structure of the DNP used in the delivery of mRNA. The highest luminescence is exhibited by single-single IAJD33 and IAJD34 forming DNPs (10^8) in lung. The next higher is again based on the IAJD31, IAJD46 and IAJD27 (10^7) based DNPs and is also in lung.

These lung activities are higher than the activity in lung of the control experiment of MC3 (Figure 9). It is important to realize that organ activities from Figure 10 are much higher than

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overall mice activities from Figures 7 and 9. The next higher activity is in liver (Figures 9 and 10). Without optimization, the luminescence in liver is 10^6 for IAJD31, IAJD34 DNPs and 10^5 for IAJD30, IAJD33 and IAJD46 based DNPs. They are smaller than the values of MC3 that is 10^8. The highest activity in spleen is for IAJD29, IAJD30, IAJD37 based DNPs (10^5) and IAJD27 based DNP (10^4). The control experiment with MC3 is 10^7 in spleen. These results indicate that single component IAJDs could provide a potential strategy to target different organs.

The design and accelerated modular-orthogonal synthesis of six libraries containing 52 multifunctional sequence-defined ionizable amphiphilic Janus dendrimers (IAJDs) are reported. All 52 IAJDs co-assemble with mRNA by simple injection in acidic buffers to generate dendrimersome nanoparticles (DNPs) with predictable dimensions and narrow polydispersity. The dimensions of these DNPs are stable for over 120 days at 5 °C. Among the 52 DNPs encapsulating Luciferase-mRNA, 42 (81%) showed activity *in vitro* and four of them showed higher activity than the four component lipid nanoparticles obtained from the MC3 FDA approved control experiment. 28 (54 %) DNPs displayed activity *in vivo* with two of them exhibiting higher activity than the MC3 control experiment in the lung. These unoptimized preliminary experiments provide proof of concept for the one-component multifunctional sequence-defined amphiphilic Janus dendrimer as an efficient delivery system for mRNA. This one-component delivery platform can be used to elucidate the mechanisms of encapsulation and release of mRNA from supramolecular virus-like assemblies and for the production of vaccines and drugs.

mRNA vaccines rely on a delivery system for the nucleic acid based on four-component ionizable lipid nanoparticles (LNPs), containing phospholipids, cholesterol for mechanical properties, PEG conjugated lipids for stability and ionizable amines.

In this report, the current four-component LNP delivery system is transformed into a simpler and more precise one-component multifunctional JD system, with highest activity mediated by a sequence-defined low concentration of ionizable amines.

A new class of JDs, named ionizable amphiphilic Janus dendrimers (IAJDs), was designed and synthesized. This concept provides the first attempt to elucidate the role of ionizable amine(s) (IA) and their ability to encapsulate and release mRNA as a function of concentration, constitutional isomerism, number of ionizable amines and their sequence by using IAJDs based DNPs as a delivery tool. The assembly of one-component DNPs relies on a simple injection method. This process is much simpler than the microfluidic device required to assemble four-component LNPs.

Six libraries containing **52** sequence-defined IAJDs were synthesized *via* an accelerated

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modular-orthogonal methodology.

Among the **52** DNPs, assembled without any optimization from IAJDs encapsulating Luciferase-mRNA, **45 (87%)** showed activity ***in vitro*** with **two** of them (IAJD9 and 22) displaying higher activity than the positive controls, MC3 and *trans*IT. **28 (54%)** showed activity ***in vivo*** with **two** of them (IAJD33 and 34) exhibited very high activity in the lungs.

The size stability of 40 DNPs was tested. Unoptimized screening experiments showed that sizes of 19 out of 40 DNPs were very stable after storage at 5 °C for up to 120 days. Almost all these DNPs were assembled from IAJDs containing a benzyl ether in the hydrophilic part.

This work provides proof of concept for one-component IAJDs as efficient delivery systems for mRNA. They will be employed to produce vaccines and drugs by mRNA delivery.

The materials and methods employed in these experiments are now described.

Triethylene glycol monomethyl ether (TCI, 98%), triethylene glycol (Alfa Aesar, 99%), benzyl chloride (Alfa Aesar, 99%), 4-methoxybenzyl chloride (TCI, 98%), *p*-toluenesulfonyl chloride (Alfa Aesar, 98%), gallic acid (Chem Impex, anhydrous, ACS grade), triethyl orthoformate (TCI, 98+%), Amberlyst-15(H) (Alfa Aesar), 1-bromooctane (Aldrich, 99%), 1-bromononane (Lancaster, 99%), 1-bromoundecane (Aldrich, 99%), 1-bromododecane (Alfa Aesar, 99%), (*rac*)-3-(bromomethyl)heptane (Aldrich, 95%), (*rac*)-1-bromo-3,7-dimethyloctane (TCI, 93+%), palladium on activated carbon catalyst (Spectrum, 10 wt% loading), pentaerythritol (Aldrich, 98%), 4-(dimethylamino)butyric acid hydrochloride (Alfa Aesar, 98%), 3-(dimethylamino)propionic acid hydrochloride (TCI, 98%), N,N-dimethylglycine hydrochloride (Acros, 99%), piperidine (Beantown Chemical, 99%), 1-methylpiperazine (Alfa Aesar, 98%), citrate buffer (100 mM, pH 3.0, TEKnova), tris buffer (1 M, Thermo Fisher Scientific) and other reagents and solvents were obtained from commercial sources and were used as received. 4-(Dimethylamino)pyridinium 4-toluenesulfonate (DPTS) was prepared according to a literature procedure. 2-Chloro-4,6-dimethoxy-1,3,5-triazine (CDMT) was prepared according to a literature procedure. Citrate buffer (100 mM, pH 3.0, TEKnova) CH_2Cl_2 (DCM) was dried over CaH_2 and freshly distilled before use.

The purity and structural identity of final products and intermediates was determined by a combination of techniques that include thin-layer chromatography (TLC), high-pressure liquid chromatography (HPLC), ^1H and ^{13}C NMR, and matrix assisted laser desorption ionization-time of flight (MALDI-TOF) mass spectrometry.

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^{1}H and ^{13}C NMR spectra were recorded at 400 MHz and 101 MHz, on a Bruker NEO (400 MHz) NMR spectrometer with autosampler, and 500 MHz and 126 MHz respectively, on a Bruker DRX (500 MHz) NMR spectrometer. All NMR spectra were measured at 23 °C in $CDCl_3$. Residual protic solvent of $CDCl_3$ (^{1}H, δ 7.26 ppm; ^{13}C, δ 77.16 ppm), and tetramethylsilane (TMS, δ 0 ppm) were used as the internal reference in the ^{1}H- and ^{13}C-NMR spectra. The absorptions are given in wavenumbers (cm^{-1}). NMR spectra were analyzed and exported by TopSpin 4.07 (Bruker) or MNova 14.

The determination of the purity of molecules by high-pressure liquid chromatography (HPLC) was carried out using Shimadzu LC-20AD high-performance liquid chromatograph pump, a PE Nelson Analytical 900 Series integration data station, a Shimadzu SPD-10A VP (UV-*vis*, λ = 254 nm) and three AM gel columns (a guard column, two 500 Å, 10 μm columns). THF with 5% of NEt_3 was used as solvent at the oven temperature of 23 °C. Detection was done by UV absorbance at 254 nm.

Matrix Assisted Laser Desorption Ionization-Time of Flight (MALDI-TOF) Mass Spectrometry was performed on a PerSeptive Biosystem-Voyager-DE (Framingham, MA) mass spectrometer equipped with nitrogen laser (337 nm) and operating in linear mode. Internal calibration was performed using Angiotensin II and Bombesin as standards. The analytical sample solution was prepared by mixing the THF solution of the sample (5–10 mg/mL) and THF solution of the matrix (2,5-dihydroxybenzoic acid, 10 mg/mL) with a 1/5 (v/v) ratio. The prepared sample solution (0.5 μL) was loaded on the MALDI plate and dried at 23 °C before the plate was inserted into the vacuum chamber of the instrument. The laser intensity and voltages applied were adjusted depending on the molecular weight and the nature of each analyzed compound.

Dynamic Light Scattering (DLS) for the DNPs was performed in buffer with a Malvern Instruments particle sizer (Zetasizer Nano S, Malvern Instruments, UK) equipped with 4 mW He–Ne laser 633 nm and avalanche photodiode positioned at 175° to the beam and temperature-controlled cuvette holder. Instrument parameters were determined automatically along with measurement times.

pK_a Measurements of Individual IAJD Molecules: IAJD Molecules were dissolved in ethanol (Sat. with NaCl) at a concentration of 1.5 mg/mL in a volume of 3 mL. 0.1 M HCl aqueous solution was added in increments of 7.5 μL, with the resulting pH measured using an Hach H170 pH meter. pK_a was calculated using half equivalence point titration.

Formulation of DNPs Containing mRNA-Luc: IAJDs were dissolved in ethanol with

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various initial concentrations (5–160 mg/mL). Nucleoside-modified mRNA encoding firefly luciferase (mRNA-Luc) was dissolved in water with various initial concentrations (1–4 mg/mL). 12.5 μL of mRNA solution was placed into a clean RNAs free eppendorf (1.5 mL) and mixed with 463 μL of citrate buffer (10 mM, pH 3.0)/acetate buffer (10 mM). For encapsulation of mRNA into IAJDs, 25 μL of IAJDs of the ethanol stock solution was rapidly injected into mRNA solution in citrate or acetate buffer followed by vortex for 5 seconds.

Dialysis of DNPs: The 0.5 mL DNP solution was dialyzed against 10 mM tris buffer (pH 7.4) or 1X PBS buffer (pH 7.4) for 2 h in 3,500 – 14,000 molecular weight cut-off dialysis tube (Spectrum Medical Instruments Inc. Spectra/Por molecular porous membrane tubing Flat Width.: 45 mm; Diameter.: 29 mm & Vol/length.: 6.4 mL/cm).

In vitro mRNA Delivery: Human embryonic kidney (HEK) 293T cells (American Type Culture Collection) were cultured in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% inactivated fetal bovine serum (FBS) (Gemini Bio-Products), 2 mM L–glutamine and 100 U/mL penicillin/streptomycin (Life Technologies) (complete medium). For *in vitro* screening experiments, HEK293T cells were seeded into 96-well plates (20,000 cells/well/200 μL) and cultured for 24 h in at 37 °C and 5% CO_2 in complete media. Cell were transfected with IAJDs with encapsulated naked nucleoside-modified mRNA encoding firefly luciferase (mRNA-Luc) with fixed concentration of mRNA-Luc, 125 ng per well. The *trans*IT (TransIT®-mRNA Transfection Kit, Mirus Bio) was used as a positive control for cell transfection, accordingly per manufacturer protocol and added to cells at the concentration of mRNA-Luc, 125 ng per well, the same as for tested particles. Cells were further cultured for 24 h, then medium was aspirated, and cells were lysed with 30 μL/well of cell culture lysis reagent (Promega). For the characterization of luminescence intensity, 2.5 μL of the lysed cells was mixed with 10 μL of luciferase assay substrate and luminescence was analyzed using a MiniLumat LB 9506 luminometer (Berthold/EG&G; Wallac).

In vivo mRNA Delivery: Female BALB/c mice aged 6-8 weeks were purchased from Charles River Laboratories. 100 μL of buffer solution containing IAJDs with encapsulated 10 μg of mRNA-Luc was retro-orbitally injected into mice under Isoflurane-anesthesia. After 4-7 h post injection mice were imaged on a Perkin Elmer IVIS Spectrum CT system 10 minutes after intraperitoneal injection of D-Luciferin, 15 mg/mL (Regis Technologies) at 10 μL/g body weight. The exposure time was set to 1 min using medium binning (binning = 8). For IVIS imaging of the organs, mice were sacrificed, then the organs were immediately collected, and bioluminescence imaging was performed.

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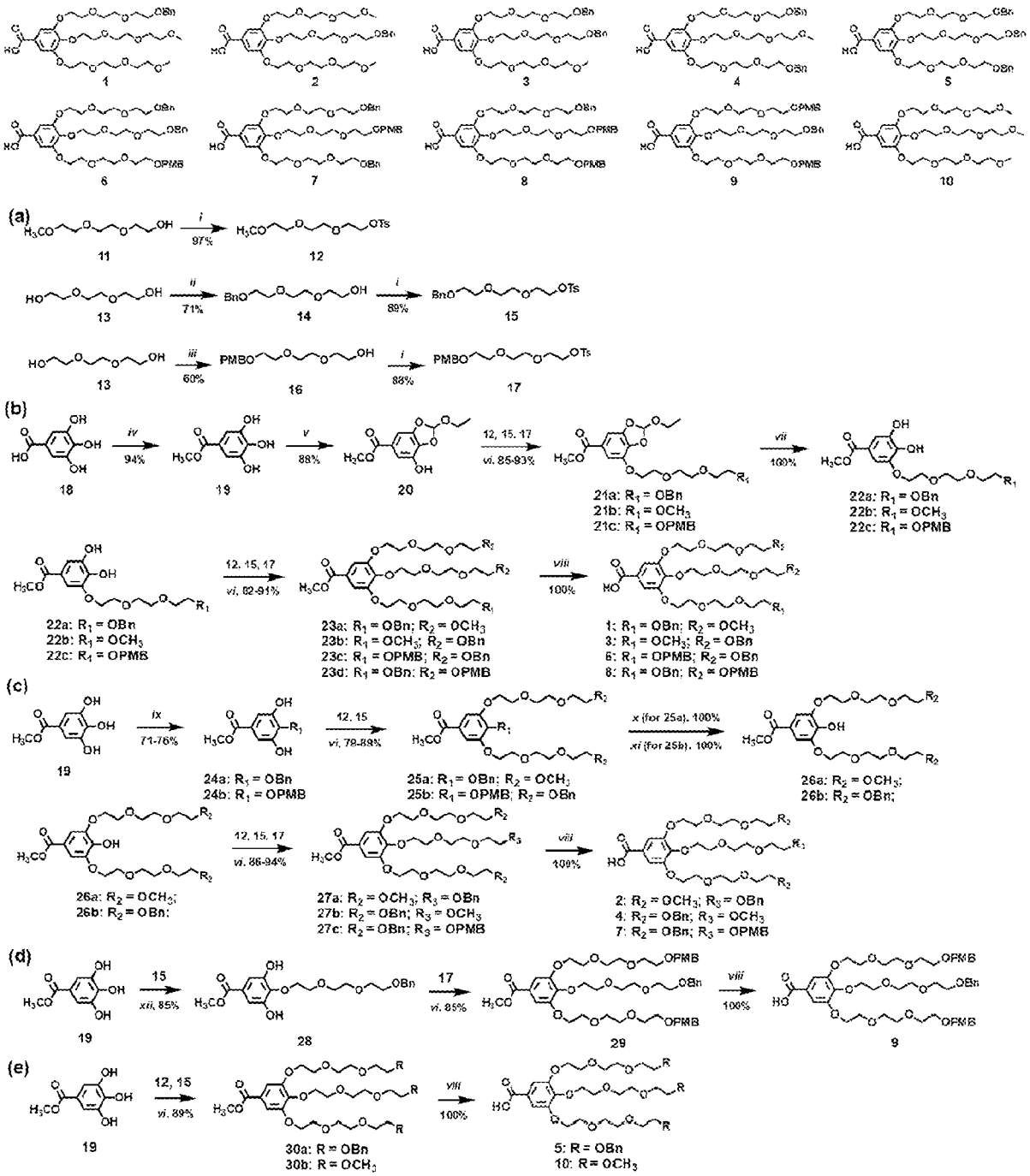
Synthesis of Hydrophilic Acids



Reagents and conditions: (*i*) TsCl, NaOH, THF-water, 0–5 °C, 3 h; (*ii*) Benzyl chloride/, NaOH, water, 100 °C, 12 h; (*iii*) 4-Methoxybenzyl chloride, NaOH, water, 100 °C, 12 h; (*iv*) H_2SO_4 (cat.), MeOH, reflux, 2 h; (*v*) Triethyl orthoformate, Amberlyst-15(H), Toluene, Dean-Stark, reflux, 18 h; (*vi*) K_2CO_3, DMF, 70 °C, 12 h; (*vii*) 2 M HCl, MeOH, 23 °C, 2 h; (*viii*) (a) KOH, EtOH,

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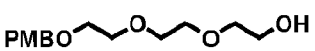
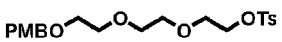
reflux, 2 h; (b) HCl (1 M); (*ix*) $KHCO_3$, KI, 4-Methoxybenzyl chloride, DMF, 60 °C, 24 h; (*x*) H_2, Pd/C, DCM/MeOH, 23 °C, 12 h; (*xi*) TFA, DCM, 23 °C, 2 h. (xii) $KHCO_3$, KI, DMF, 60 °C, 24 h.

The synthesis of 2-(2-(2-methoxyethoxy)ethoxy)ethyl 4-methylbenzenesulfonate (**11**), 2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethan-1-ol (**14**) and 2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethyl 4-methylbenzenesulfonate (**15**) was adapted from literature procedures previously reported. The procedures are optimized to increase the yields of compounds **11**, **14** and **15** to 78.02 g, 20.21 g, and 28.81 g, respectively. The synthesis of methyl 2-ethoxy-7-hydroxybenzo[*d*][1,3]dioxole-5-carboxylate (**20**) and methyl 4-(benzyloxy)-3,5-dihydroxybenzoate (**24a**) was adapted from literature procedures (Wang, et al., J. Am. Chem. Soc. 2020, 142, 9525−9536). Methyl 3,4,5-tris(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy) benzoate (**30a**), methyl 3,4,5-tris(2-(2-(2-methoxyethoxy)ethoxy)ethoxy)benzoate (**30b**), 3,4,5-tris(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)benzoic acid (**5**) and 3,4,5-tris(2-(2-(2-methoxyethoxy)ethoxy)ethoxy)benzoic acid (**10**) were prepared according to literature procedures (Percec, et al., Science 2010, 328, 1009–1014; Percec, et al., J. Am. Chem. Soc. 2013, 135, 4129–4148).



16

2-(2-(2-((4-Methoxybenzyl)oxy)ethoxy)ethoxy)ethan-1-ol (16). Triethylene glycol (**13**, 34.60 g, 229.75 mmol, 4 equiv) was added to 120 mL NaOH aqueous solution (50%). The mixture was stirred at 23 °C for 30 min. Then 4-methoxybenzyl chloride (9.00 g, 57.47 mmol, 1 equiv) was added and the mixture was allowed to stir at 100 °C for 24 h. The reaction mixture was cooled to 23 °C, followed by diluting with water (80 mL) and extracting with diethyl ether (60 mL × 3). The organic phase was dried over anhydrous magnesium sulfate ($MgSO_4$) and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with ethyl acetate (EtOAc)/hexane = 1/1 as the eluent to give the title compound as a brown-red oil (9.82 g, 60%).



17

2-(2-(2-((4-Methoxybenzyl)oxy)ethoxy)ethoxy)ethyl 4-methylbenzenesulfonate **(17).** Compound **16** (8.31 g, 30.74 mmol, 1 equiv) and NaOH (1.84 g, 46.11 mmol, 1 equiv) were dissolved in the mixture of THF and water (1/1, 40 mL) and the mixture was cooled to 0 °C. 4-Toluenesulfonyl chloride (TsCl, 5.86 g, 30.74 mmol, 1 equiv) dissolved in 20 mL THF was added to the mixture over 30 min at 0 °C. The reaction mixture was allowed to stir at 0–5 °C for 2 h. The reaction mixture was poured into the ice-water bath (100 mL), and the mixture was extracted by

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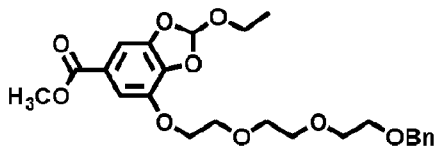
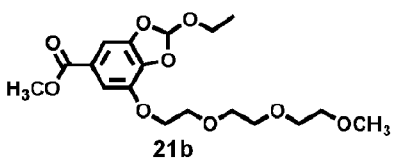
DCM (50 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 1/1 as the eluent to give the title compound as a brown-red oil (11.45 g, 88%).



21a

Methyl 7-(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-2-ethoxybenzo-[*d*][1,3]dioxole-5-carboxylate **(21a).** Methyl 2-ethoxy-7-hydroxybenzo[*d*][1,3]dioxole-5-carboxylate (**20**, 0.80 g, 3.33 mmol, 1 equiv) and K_2CO_3 (1.38 g, 10.00 mmol, 3 equiv) were stirred in dry DMF (50 mL). 2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethyl 4-methylbenzenesulfonate (**15**, 1.58 g, 4.00 mmol, 1.2 equiv) was added and the mixture was stirred at 70 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 1/1 as the eluent to give the title compound as a light-yellow oil (1.43 g, 93%).



21b

Methyl 2-ethoxy-7-(2-(2-(2-methoxyethoxy)ethoxy)ethoxy)benzo-[*d*][1,3]dioxole-5-carboxylate **(21b).** Compound **20** (5.00 g, 20.82 mmol, 1 equiv) and K_2CO_3 (8.63 g, 62.46 mmol, 3 equiv) were stirred in dry DMF (70 mL). Compound **12** (7.95 g, 24.98 mol, 1.2 equiv) was added and the mixture was stirred at 70 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (50 mL) was added, and the mixture was extracted by DCM (50 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 1/2 as the eluent to give the title compound as a light-yellow oil (6.23 g, 86%).

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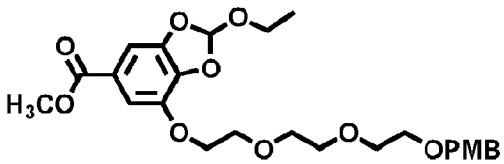
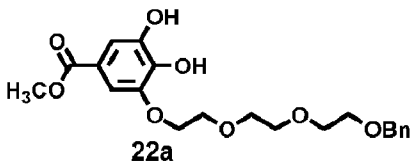


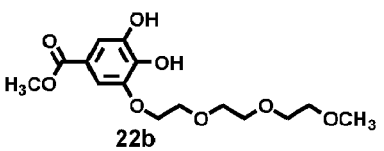
21c

Methyl 2-ethoxy-7-(2-(2-(2-((4-methoxybenzyl)oxy)ethoxy)ethoxy)ethoxy)-benzo[*d*][1,3]dioxole-5-carboxylate **(21c).** Compound **20** (0.70 g, 2.91 mmol, 1 equiv) and K_2CO_3 (1.21 g, 8.73 mmol, 3 equiv) were stirred in dry DMF (30 mL). Compound **17** (1.36 g, 3.20 mmol, 1.1 equiv) was added and the mixture was stirred at 70 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 1/1 as the eluent to give the title compound as a light-yellow oil (1.22 g, 85%).



22a

Methyl 3-(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-4,5-dihydroxybenzoate **(22a).** Compound **21a** (0.97 g, 2.10 mmol) was dissolved in 20 mL MeOH. Then HCl (2 M, 10.0 mL, 20.00 mmol) was added. The mixture was stirred at 23 °C for 2 h. The reaction mixture was extracted by DCM (20 mL × 3) and the organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.85 g, 100%).



22b

Methyl 3,4-dihydroxy-5-(2-(2-(2-methoxyethoxy)ethoxy)ethoxy)benzoate **(22b).** Compound **21b** (5.36 g, 13.88 mmol) was dissolved in 30 mL MeOH. Then HCl (2 M, 30.0 mL, 60.00 mmol) was added. The mixture was stirred at 23 °C for 2 h. The reaction mixture was extracted by DCM (30 mL × 3) and the organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (4.58 g, 100%).

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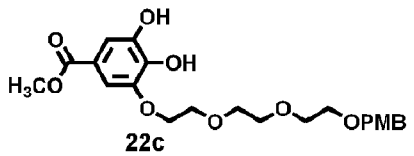
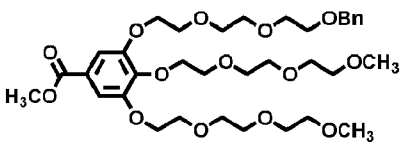


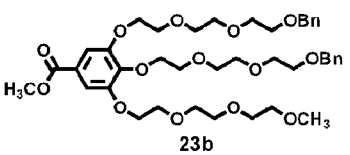
22c

Methyl 3,4-dihydroxy-5-(2-(2-(2-((4-methoxybenzyl)oxy)ethoxy)ethoxy)ethoxy)-benzoate **(22c).** Compound **21c** (0.90 g, 1.83 mmol) was dissolved in 10 mL MeOH. Then HCl (2 M, 4.0 mL, 8.00 mmol) was added. The mixture was stirred at 23 °C for 2 h. The reaction mixture was extracted by DCM (20 mL × 3) and the organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.80 g, 100%).



23a

Methyl 3-(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-4,5-bis(2-(2-(2-methoxyethoxy)ethoxy)ethoxy) benzoate **(23a).** Compound **22a** (0.84 g, 2.07 mmol, 1 equiv) and K_2CO_3 (1.72 g, 12.42 mmol, 6 equiv) were stirred in dry DMF (30 mL). Compound **12** (1.45 g, 4.55 mol, 2.2 equiv) was added and the mixture was stirred at 70 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 30/1 as the eluent to give the title compound as a yellow oil (1.19 g, 82%).



23b

Methyl 3,4-bis(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-5-(2-(2-(2-methoxyethoxy)ethoxy)ethoxy) benzoate **(23b).** Compound **22b** (4.00 g, 12.11 mmol, 1 equiv) and K_2CO_3 (10.04 g, 72.66 mmol, 6 equiv) were stirred in dry DMF (70 mL). Compound **15** (10.98 g, 27.83 mol, 2.3 equiv) was added and the mixture was stirred at 70 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (50 mL) was added, and the mixture was extracted by DCM (50 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 50/1 as the eluent to give the title compound

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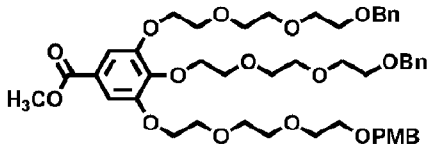
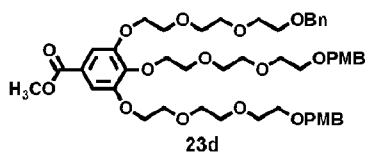
as a light-yellow oil (8.42 g, 90%).



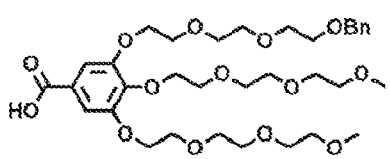
23c

Methyl 3,4-bis(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-5-(2-(2-(2-((4-methoxybenzyl)oxy)ethoxy) ethoxy)ethoxy)benzoate **(23c).** Compound **22c** (0.79 g, 1.81 mmol, 1 equiv) and K_2CO_3 (1.50 g, 10.86 mmol, 6 equiv) were stirred in dry DMF (30 mL). Compound **15** (1.50 g, 3.80 mol, 2.1 equiv) was added and the mixture was stirred at 70 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 1/3 as the eluent to give the title compound as a yellow oil (1.31 g, 82%).



23d

Methyl 3-(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-4,5-bis(2-(2-(2-((4-methoxybenzyl)oxy)ethoxy) ethoxy)ethoxy)benzoate **(23d).** Compound **22a** (1.80 g, 4.43 mmol, 1 equiv) and K_2CO_3 (3.67 g, 26.58 mmol, 6 equiv) were stirred in dry DMF (50 mL). Compound **17** (3.95 g, 9.30 mol, 2.1 equiv) was added and the mixture was stirred at 70 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc as the eluent to give the title compound as a brown-yellow oil (3.65 g, 91%).



1

3-(2-(2-(2-(Benzyloxy)ethoxy)ethoxy)ethoxy)-4,5-bis(2-(2-(2-methoxyethoxy)ethoxy)ethoxy) benzoic acid (**1**). Compound **23a** (1.10 g, 1.57 mmol, 1 equiv) was

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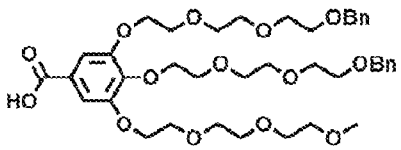
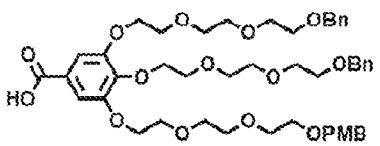
dissolved in ethanol (EtOH, 15 mL). KOH (0.62 g, 10.99 mmol, 7 equiv) was added and the mixture was stirred at reflux for 2 h. The mixture was cooled to 23 °C and acidified with dilute HCl (1 M) to pH ≈ 1. The solution was extracted by DCM (20 mL × 3) and the organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a yellow oil (1.07 g, 100%).



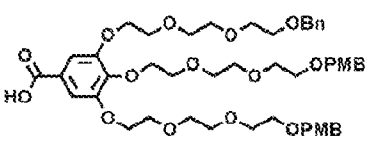
3

3,4-Bis(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-5-(2-(2-(2-methoxyethoxy)ethoxy)ethoxy) benzoic acid **(3).** Compound **23b** (7.88 g, 10.17 mmol, 1 equiv) was dissolved in ethanol (50 mL). KOH (3.99 g, 71.19 mmol, 7 equiv) was added and the mixture was stirred at reflux for 2 h. The mixture was cooled to 23 °C and acidified with dilute HCl (1 M) to pH ≈ 1. The solution was extracted by DCM (40 mL × 3) and the organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a yellow oil (7.73 g, 100%).



6

3,4-Bis(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-5-(2-(2-(2-((4-methoxybenzyl)oxy)ethoxy)ethoxy) ethoxy)benzoic acid **(6).** Compound **23c** (0.98 g, 1.11 mmol, 1 equiv) was dissolved in ethanol (15 mL). KOH (0.44 g, 7.77 mmol, 7 equiv) was added and the mixture was stirred at reflux for 2 h. The mixture was cooled to 23 °C and acidified with dilute HCl (1 M) to pH < 2. The solution was extracted by DCM (30 mL × 3) and the organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a brown-yellow oil (0.96 g, 100%).



8

3-(2-(2-(2-(Benzyloxy)ethoxy)ethoxy)ethoxy)-4,5-bis(2-(2-(2-((4-methoxybenzyl)oxy)ethoxy)ethoxy) ethoxy)benzoic acid **(8).** Compound **23d** (2.80 g, 3.07 mmol, 1

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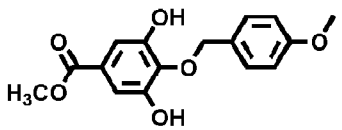
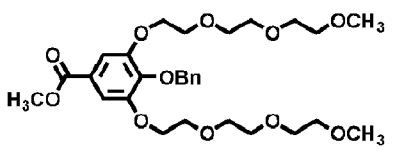
equiv) was dissolved in ethanol (30 mL). KOH (1.20 g, 21.49 mmol, 7 equiv) was added and the mixture was stirred at reflux for 2 h. The mixture was cooled to 23 °C and acidified with dilute HCl (1 M) to pH < 2. The solution was extracted by DCM (30 mL × 3) and the organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a brown-yellow oil (2.73 g, 100%).



24b

Methyl 3,5-dihydroxy-4-((4-methoxybenzyl)oxy)benzoate **(24b).** Methyl 3,4,5-trihydroxybenzoate (**19**, 3.25 g, 17.66 mmol, 1 equiv), 4-methoxybenzyl chloride (2.77 g, 17.66 mmol, 1 equiv), $KHCO_3$ (2.53 g, 35.32 mmol, 3 equiv) and KI (18 mg, 0.11 mmol, 0.006 equiv) were stirred in dry DMF (80 mL) at 60 °C under N_2 atmosphere for 24 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (50 mL) was added, and the mixture was extracted by DCM (50 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 2/1 as the eluent to give the title compound as a white solid (2.30 g, 76%).



25a

Methyl 4-(benzyloxy)-3,5-bis(2-(2-(2-methoxyethoxy)ethoxy)ethoxy)benzoate **(25a).** Methyl 4-(benzyloxy)-3,5-dihydroxybenzoate (**24a**, 4.00 g, 14.59 mmol, 1 equiv) and K_2CO_3 (12.08 g, 87.41 mmol, 6 equiv) were stirred in dry DMF (70 mL). Compound **12** (10.64 g, 33.42 mol, 2.3 equiv) was added and the mixture was stirred at 70 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (50 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc as the eluent to give the title compound as a light-yellow oil (7.27 g, 88%).

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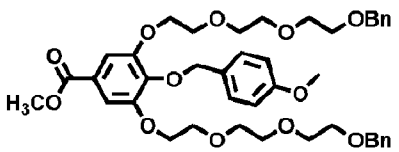
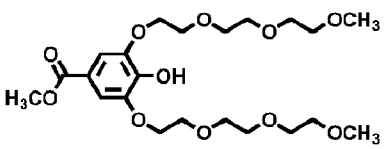


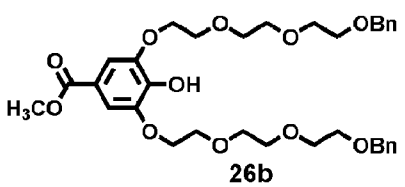
25b

Methyl 3,5-bis(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-4-((4-methoxybenzyl)oxy)benzoate (25b). Compound **24b** (4.00 g, 13.14 mmol, 1 equiv) and K_2CO_3 (10.90 g, 78.86 mmol, 6 equiv) were stirred in dry DMF (60 mL). Compound **15** (11.90 g, 30.17 mol, 2.3 equiv) was added and the mixture was stirred at 70 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (50 mL) was added and the mixture was extracted by DCM (50 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 1/2 as the eluent to give the title compound as a light-yellow oil (7.70 g, 78%).



26a

Methyl 4-hydroxy-3,5-bis(2-(2-(2-methoxyethoxy)ethoxy)ethoxy)benzoate **(26a).** Compound **25a** (7.27 g, 12.83 mmol) was dissolved in 1:2 MeOH:DCM (60 mL). Then Pd/C (0.22 g, 3 wt%) was added and the flask was evacuated and filled with hydrogen for three times. The mixture was stirred at 23 °C for 12 h under hydrogen atmosphere. The reaction mixture was filtered through Celite and the filter cake was washed with DCM. Evaporation of the solvent yielded the title compound as a light-yellow oil (6.11 g, 100%).



26b

Methyl 3,5-bis(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-4-hydroxybenzoate **(26b).** Compound **25b** (3.00 g, 4.01 mmol) was dissolved in 20 mL DCM. Then trifluoroacetic acid (TFA, 1.83 g, 16.04 mmol) was added. The mixture was stirred at 23 °C for 2 h. The reaction was quenched by adding 30 mL saturated $NaHCO_3$ solution and the mixture was extracted by DCM (30 mL × 3). The combined organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (2.52 g, 100%).

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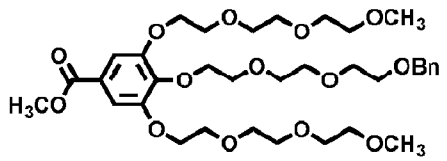
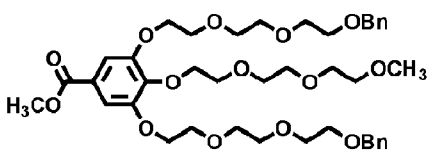


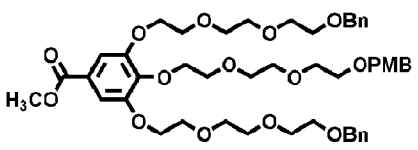
27a

Methyl 4-(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-3,5-bis(2-(2-(2-methoxyethoxy)ethoxy)ethoxy) benzoate **(27a).** Compound **26a** (1.06 g, 2.22 mmol, 1 equiv) and K_2CO_3 (0.92 g, 6.66 mmol, 3 equiv) were stirred in dry DMF (30 mL). Compound **15** (1.00 g, 2.53 mmol, 1.2 equiv) was added and the mixture was stirred at 70 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 30/1 as the eluent to give the title compound as a light-yellow oil (1.46 g, 94%).



27b

Methyl 3,5-bis(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-4-(2-(2-(2-methoxyethoxy)ethoxy)ethoxy) benzoate **(27b).** Compound **26b** (1.70 g, 2.71 mmol, 1 equiv) and K_2CO_3 (1.12 g, 8.13 mmol, 3 equiv) were stirred in dry DMF (70 mL). Compound **12** (1.00 g, 3.25 mol, 1.2 equiv) was added and the mixture was stirred at 70 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 50/1 as the eluent to give the title compound as a light-yellow oil (1.85 g, 89%).



27c

Methyl 3,5-bis(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-4-(2-(2-(2-((4-methoxybenzyl)oxy)ethoxy) ethoxy)ethoxy)benzoate **(27c).** Compound **26b** (1.00 g, 1.59 mmol, 1 equiv) and K_2CO_3 (0.66 g, 4.78 mmol, 3 equiv) were stirred in dry DMF (20 mL). Compound **17**

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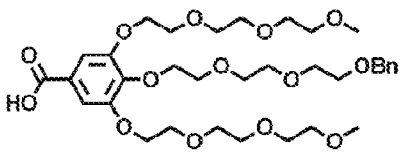
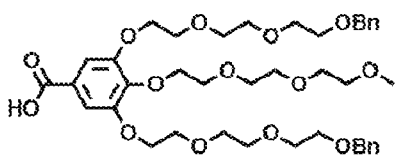
(0.75 g, 1.75 mol, 1.1 equiv) was added and the mixture was stirred at 70 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (20 mL) was added, and the mixture was extracted by DCM (20 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 1/3 as the eluent to give the title compound as a yellow oil (1.21 g, 86%).



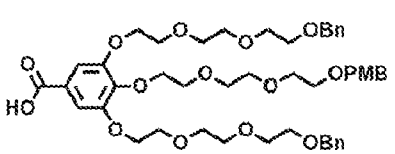
2

4-(2-(2-(2-(Benzyloxy)ethoxy)ethoxy)ethoxy)-3,5-bis(2-(2-(2-methoxyethoxy)ethoxy)ethoxy) benzoic acid **(2).** Compound **27a** (1.46 g, 2.09 mmol, 1 equiv) was dissolved in ethanol (15 mL). KOH (0.82 g, 14.62 mmol, 7 equiv) was added and the mixture was stirred at reflux for 2 h. The mixture was cooled to 23 °C and acidified with dilute HCl (1 M) to pH ≈ 1. The solution was extracted by DCM (20 mL × 3) and the organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (1.42 g, 100%).



4

3,4-Bis(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-5-(2-(2-(2-methoxyethoxy)ethoxy)ethoxy) benzoic acid (**4**). Compound **27b** (1.80 g, 2.32 mmol, 1 equiv) was dissolved in ethanol (20 mL). KOH (0.90 g, 16.30 mmol, 7 equiv) was added and the mixture was stirred at reflux for 2 h. The mixture was cooled to 23 °C and acidified with dilute HCl (1 M) to pH ≈ 1. The solution was extracted by DCM (20 mL × 3) and the organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a yellow oil (1.76 g, 100%).



7

3,5-Bis(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-4-(2-(2-(2-((4-

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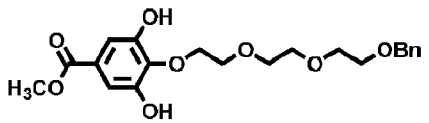
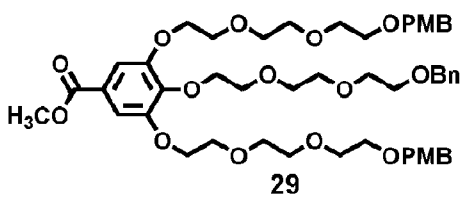
methoxybenzyl)oxy)ethoxy)ethoxy) ethoxy)benzoic acid **(7).** Compound **27c** (1.00 g, 1.14 mmol, 1 equiv) was dissolved in ethanol (15 mL). KOH (0.45 g, 7.98 mmol, 7 equiv) was added and the mixture was stirred at reflux for 2 h. The mixture was cooled to 23 °C and acidified with dilute HCl (1 M) to pH < 2. The solution was extracted by DCM (30 mL × 3) and the organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a brown-yellow oil (0.99 g, 100%).



28

Methyl 4-(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-3,5-dihydroxybenzoate **(28).** Compound **19** (2.73 g, 14.84 mmol, 1 equiv), compound **15** (5.85 g, 14.84 mmol, 1 equiv), $KHCO_3$ (2.97 g, 29.68 mmol, 2 equiv) and KI (15 mg, 0.09 mmol, 0.006 equiv) were stirred in dry DMF (50 mL) at 60 °C under N_2 atmosphere for 24 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 1/1 as the eluent to give the title compound as a yellow oil (5.12 g, 85%).



29

Methyl 4-(2-(2-(2-(benzyloxy)ethoxy)ethoxy)ethoxy)-3,5-bis(2-(2-(2-((4-methoxybenzyl)oxy)ethoxy) ethoxy)ethoxy)benzoate **(29).** Compound **28** (1.80 g, 4.43 mmol, 1 equiv) and K_2CO_3 (3.67 g, 26.58 mmol, 6 equiv) were stirred in dry DMF (50 mL). Compound **17** (3.95 g, 9.30 mol, 2.1 equiv) was added and the mixture was stirred at 70 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc as the eluent to give the title compound as a yellow oil (3.42 g, 85%).

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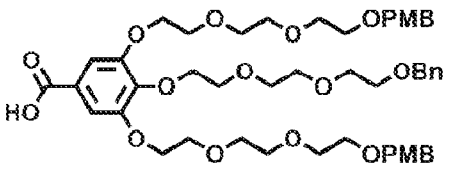
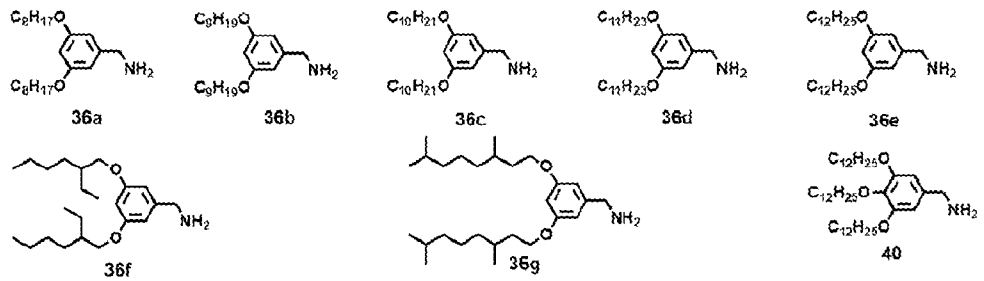


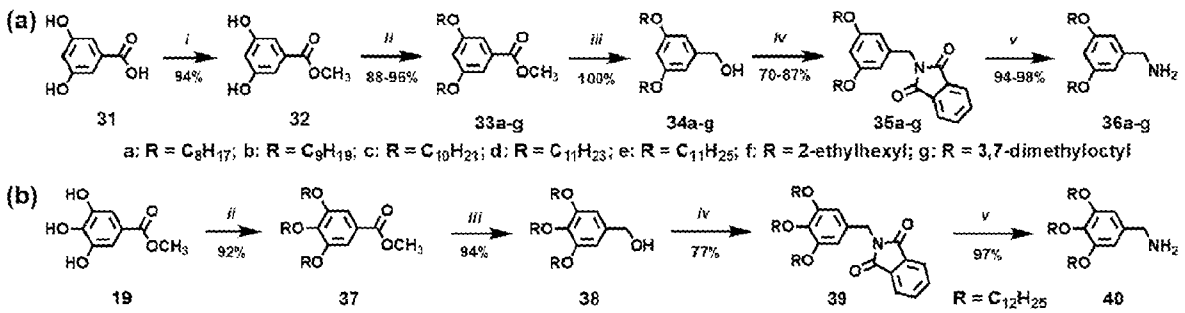
9

4-(2-(2-(2-(Benzyloxy)ethoxy)ethoxy)ethoxy)-3,5-bis(2-(2-(2-((4-methoxybenzyl)oxy)ethoxy)ethoxy) ethoxy)benzoic acid **(9).** Compound **29** (3.40 g, 3.73 mmol, 1 equiv) was dissolved in ethanol (30 mL). KOH (1.46 g, 26.11 mmol, 7 equiv) was added and the mixture was stirred at reflux for 2 h. The mixture was cooled to 23 °C and acidified with dilute HCl (1 M) to pH < 2. The solution was extracted by DCM (30 mL × 3) and the organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a yellow oil (3.35 g, 100%).

Synthesis of Hydrophobic Benzyl Amines



36a 36b 36c 36d 36e 36f 36g 40



a: R = C_8H_{17}; b: R = C_9H_{19}; c: R = $C_{10}H_{21}$; d: R = $C_{11}H_{23}$; e: R = $C_{11}H_{25}$; f: R = 2-ethylhexyl; g: R = 3,7-dimethyloctyl

Reagents and conditions: (*i*) H_2SO_4 (cat.), MeOH, reflux, 2 h; (*ii*) RBr, K_2CO_3, DMF, 120 °C, 2 h; (*iii*) $LiAlH_4$, THF, 0–23 °C, 1 h; (*iv*) (a) $SOCl_2$, DMF (cat.), DCM, 23 °C, 30 min; (b) Potassium phthalimide, DMF, 80 °C, 1 h; (*v*) Hydrazine monohydrate, EtOH, reflux, 2 h.

General Synthetic Procedure for Compounds 33a-g and 37: Compounds **33a-g** and **37** were synthesized according to a literature procedure (Buzzacchera, et al., Biomacromolecules 2019, 20, 712−727). Generally, compound **32/19** (1 equiv) and K_2CO_3 (3 equiv) were stirred in dry DMF.

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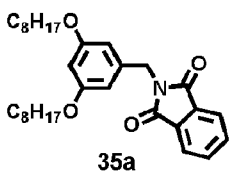
RBr (2.2 equiv for compounds **33a-g**, 3.3 equiv for compound **37**) was added, and the mixture was stirred at 120 °C under N_2 atmosphere for 2 h. The reaction mixture was cooled to 23 °C. For compounds **33a-e and 37**, the reaction mixture was poured into ice/water and the white precipitates were filtered. Then the precipitates were recrystallized from acetone to afford the title compound as a white solid. For compounds **33f, g**, water was added to the reaction mixture and the mixture was extracted by DCM for three times. The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the eluent to give the title compound as a light-yellow oil.

General Synthetic Procedure for Compounds 34a-g and 38. Compounds **34a-g** and **38** were synthesized according to a literature procedure (Li, et al., ACS Nano 2020, 14, 7398–7411). Generally, compound **33a-g** and **38** (1 equiv) dissolved in dry THF was added dropwise to a slurry of $LiAlH_4$ (1.2 equiv) in dry THF at 0 °C under N_2 atmosphere. The mixture was stirred at 23 °C for 1 h. Afterwards, the reaction was quenched by the successive addition of water, 15% NaOH aqueous solution and water. Then the mixture was filtered and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a white solid or light-yellow oil.

General Synthetic Procedure for Compounds 35a-g and 39. Compounds **35a-g** and **39** were synthesized according to a literature procedure (Zhang, et al., ACS Nano 2014, 8, 1554−1565). The characterizations of 2-(3,5-bis(dodecyloxy)benzyl)isoindoline-1,3-dione (**35e**) and 2-(3,4,5-tris(dodecyloxy)benzyl)isoindoline-1,3-dione (**39**) were reported in the literature. A typical example is provided below.



35a

2-(3,5-Bis(octyloxy)benzyl)isoindoline-1,3-dione **(35a).** To a solution of **34a** (3.82 g, 10.48 mmol, 1 equiv) in dry DCM (20 mL) was added 2 drops of DMF, followed by the dropwise addition of $SOCl_2$ (1.87 g, 15.72 mmol, 1.5 equiv). The reaction mixture was stirred at 23 °C for 30 min. Then DCM and excess $SOCl_2$ was removed under vacuum. The obtained intermediate product was dissolved in dry DMF (40 mL) and potassium phthalimide (2.15 g, 11.53 mmol, 1.1 equiv) was added. The mixture was stirred at 80 °C under N_2 atmosphere for 1 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous

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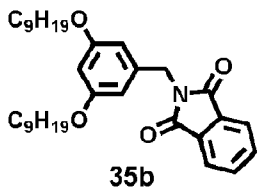
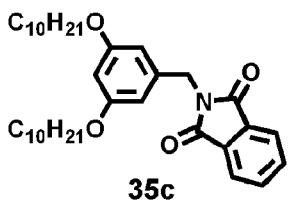
$MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 10/1 as the eluent to give the title compound as a yellow oil (4.64 g, 81%).



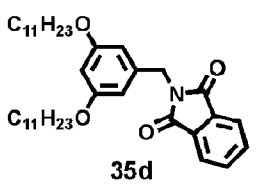
35b

2-(3,5-Bis(nonyloxy)benzyl)isoindoline-1,3-dione **(35b).** From compound **34b** (2.15 g, 5.47 mmol), $SOCl_2$ (0.60 g, 5.51 mmol) and potassium phthalimide (1.21 g, 6.56 mmol), compound **35b** was obtained as a white solid (2.01 g, 70%).



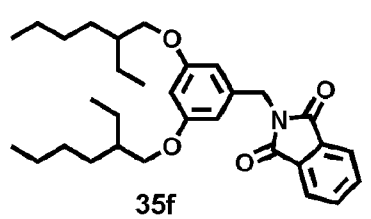
35c

2-(3,5-Bis(decyloxy)benzyl)isoindoline-1,3-dione **(35c).** From compound **34c** (2.94 g, 6.17 mmol), $SOCl_2$ (1.10 g, 9.26 mmol) and potassium phthalimide (1.26 g, 6.79 mmol), compound **35c** was obtained as a red oil (2.84 g, 76%).



35d

2-(3,5-Bis(undecyloxy)benzyl)isoindoline-1,3-dione **(35d).** From compound **34d** (3.10 g, 7.90 mmol), $SOCl_2$ (1.13 g, 9.48 mmol) and potassium phthalimide (1.76 g, 9.48 mmol), compound **35d** was obtained as a yellow solid (3.28 g, 72%).



35f

2-(3,5-Bis((2-ethylhexyl)oxy)benzyl)isoindoline-1,3-dione **(35f).** From compound **34f** (3.20 g, 8.78 mmol), $SOCl_2$ (1.57 g, 13.17 mmol) and potassium phthalimide (1.80 g, 9.66 mmol), compound **35f** was obtained as a yellow oil (3.25 g, 75%).

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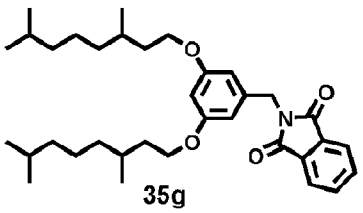
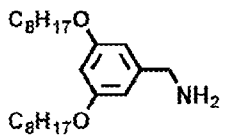


35g

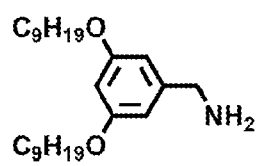
2-(3,5-Bis((3,7-dimethyloctyl)oxy)benzyl)isoindoline-1,3-dione **(35g).** From compound **34g** (3.50 g, 8.33 mmol), $SOCl_2$ (1.19 g, 10.0 mmol) and potassium phthalimide (1.85 g, 10.0 mmol), compound **35g** was obtained as a yellow oil (3.98 g, 87%).

General Synthetic Procedure for Compounds 36a-g and 40. Compounds **36a-g** and **40** were synthesized according to a literature procedure (Zhang, et al., *ACS Nano* **2014**, *8*, 1554−1565). The characterizations of (3,5-bis(dodecyloxy)phenyl)methanamine (**36e**) and (3,4,5-tris(dodecyloxy)phenyl)methanamine (**40**) were reported in the literature. A typical example is provided below.



36a

(3,5-Bis(octyloxy)phenyl)methanamine **(36a).** Compound **35a** (1.52 g, 2.76 mmol, 1 equiv) and $NH_2NH_2{\bullet}H_2O$ (1.38 g, 27.6 mmol, 10 equiv) were dissolved in EtOH (40 mL). The mixture was heated at reflux for 2 h. The reaction mixture was cooled to 23 °C and EtOH was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. Filtration and evaporation of the solvent yielded the title compound as a yellow oil (0.98 g, 98%).



36b

(3,5-Bis(nonyloxy)phenyl)methanamine **(36b).** From compound **35b** (2.01 g, 3.85 mmol) and $NH_2NH_2{\bullet}H_2O$ (1.92 g, 38.5 mmol), compound **36b** was obtained as a yellow oil (1.42 g, 94%).

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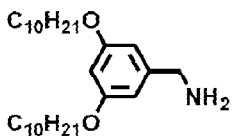
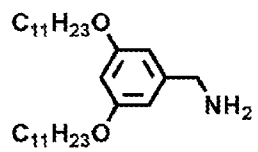


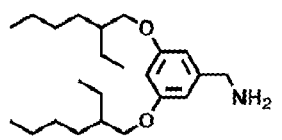
36c

(3,5-Bis(decyloxy)phenyl)methanamine **(36c).** From compound **35c** (1.87 g, 3.09 mmol) and $NH_2NH_2•H_2O$ (1.55 g, 30.9 mmol), compound **36c** was obtained as a brownish-red oil (1.26 g, 97%).



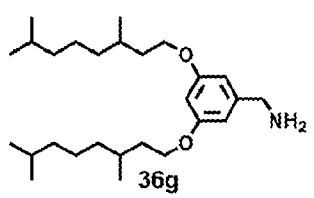
36d

(3,5-Bis(undecyloxy)phenyl)methanamine **(36d).** From compound **35d** (3.22 g, 5.57 mmol) and $NH_2NH_2•H_2O$ (2.79 g, 55.7 mmol), compound **36d** was obtained as a brownish-red oil (2.35 g, 94%).



36f

(3,5-Bis((2-ethylhexyl)oxy)phenyl)methanamine **(36f).** From compound **35f** (2.49 g, 5.04 mmol) and $NH_2NH_2•H_2O$ (2.52 g, 50.4 mmol), compound **36f** was obtained as a brownish-yellow oil (1.80 g, 98%).



36g

(3,5-Bis((3,7-dimethyloctyl)oxy)phenyl)methanamine **(36g).** From compound **35g** (3.50 g, 6.4 mmol) and $NH_2NH_2•H_2O$ (3.19 g, 64 mmol), compound **36g** was obtained as a brownish-yellow oil (2.55 g, 95%).

Synthesis of Hydrophobic Acids

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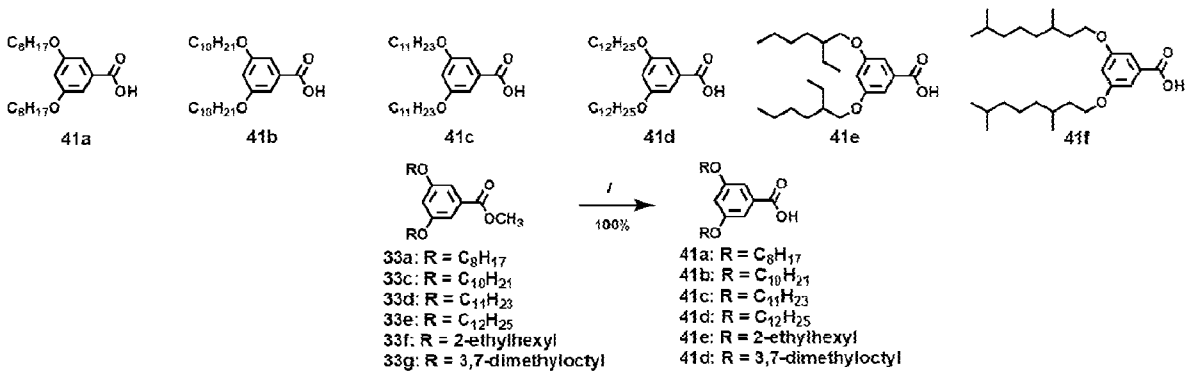
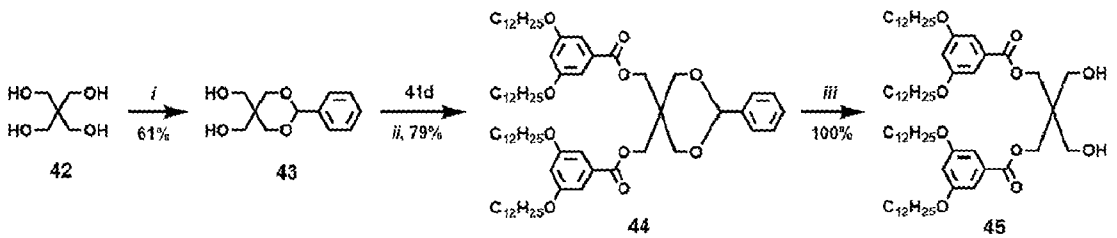


Reagents and conditions: (i) (a) KOH, EtOH, reflux, 2 h; (b) HCl (1 M).

Compounds 41a-f were synthesized according to a literature procedure. The procedures were optimized to increase the yields of compounds 41a-f to around 20 g, respectively.

Synthesis of Twin-Twin Hydrophobic Building Blocks



Reagents and conditions: (*i*) Benzaldehyde, HCl (cat.), water; (*ii*) DCC, DTPS, DCM, 23 °C, 12 h; (*iii*) H_2, Pd/C, MeOH/DCM, 23 °C, 12 h.

Compounds **43-45** were synthesized according to a literature procedure. The procedures were optimized to increase the yields of compound **45** to 30 g.

3.2 Synthesis of Single-Single IAJDs of Library 1

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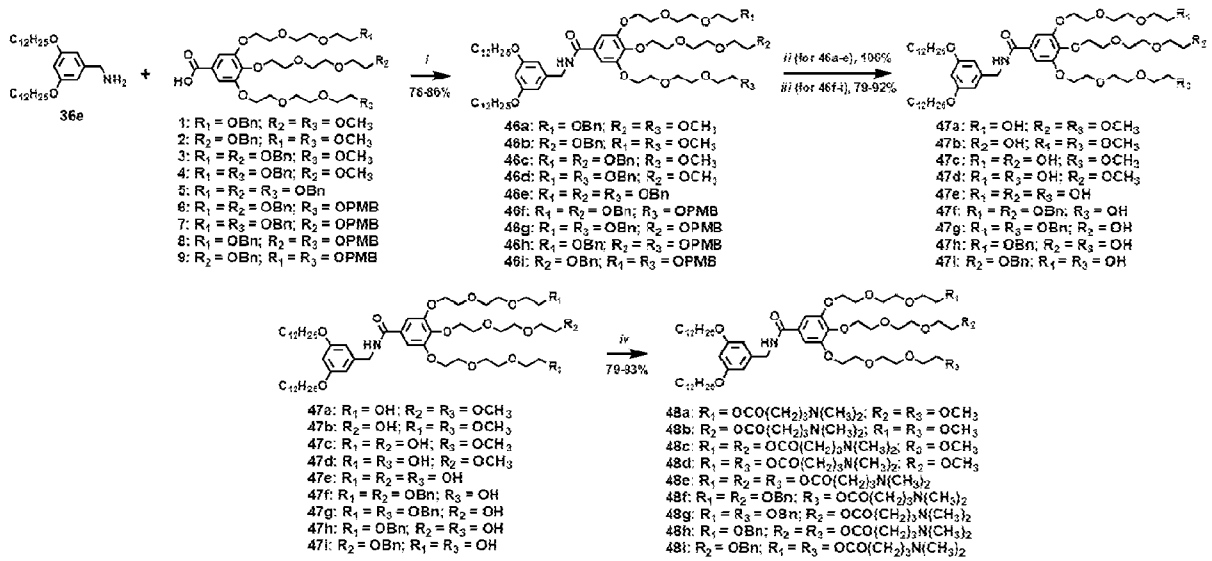
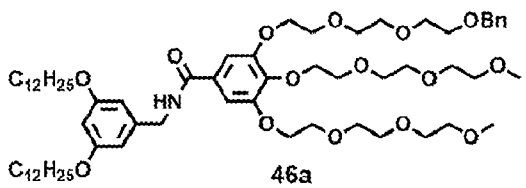
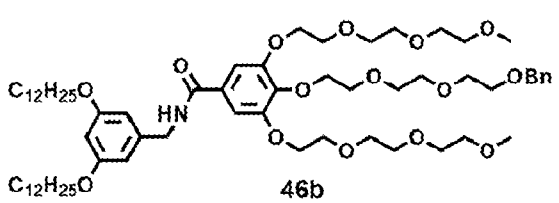


Reagents and conditions: (*i*) EDC.HCl, DMAP, DCM, 23 °C, 12 h; (*ii*) H_2, Pd/C, DCM/MeOH, 23 °C, 12 h; (*iii*) DDQ, DCM/H_2O, 23 °C, 1 h; (*iv*) 4-(Dimethylamino)butyric acid hydrochloride, DCC, DCM, 23 °C, 12 h.



46a

Compound 46a. Compound **1** (0.36 g, 0.53 mmol, 1 equiv), compound **36e** (0.27 g, 0.57 mmol, 1.1 equiv), EDC.HCl (0.11 g, 0.57 mmol, 1.1 equiv) and DMAP (19 mg, 0.16 mmol, 0.3 equiv) were dissolved in dry DCM (6 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 30/1 as the eluent to give the title compound as a light-yellow oil (0.51 g, 85%).



46b

Compound 46b. Compound **2** (0.36 g, 0.53 mmol, 1 equiv), compound **36e** (0.27 g, 0.57 mmol, 1.1 equiv), EDC.HCl (0.11 g, 0.57 mmol, 1.1 equiv) and DMAP (19 mg, 0.16 mmol, 0.3 equiv) were dissolved in dry DCM (6 mL). The reaction mixture was stirred at 23 °C for 12 h.

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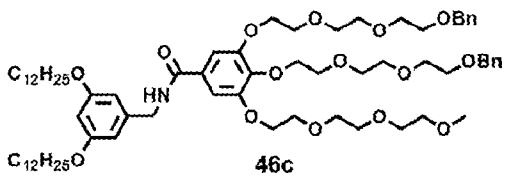
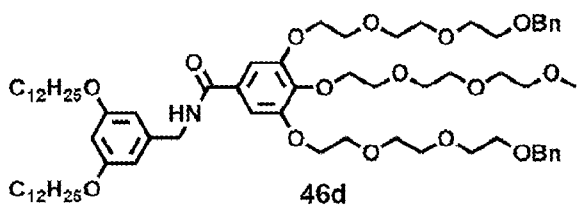
Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 30/1 as the eluent to give the title compound as a light-yellow oil (0.49 g, 82%).



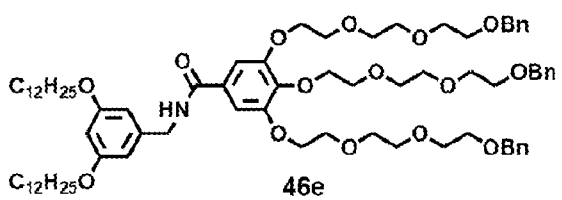
46c

Compound 46c. Compound **3** (1.15 g, 1.51 mmol, 1 equiv), compound **36e** (0.80 g, 1.68 mmol, 1.1 equiv), EDC.HCl (0.32 g, 1.68 mmol, 1.1 equiv) and DMAP (55 mg, 0.45 mmol, 0.3 equiv) were dissolved in dry DCM (10 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 3/1 as the eluent to give the title compound as a light-yellow oil (1.55 g, 84%).



46d

Compound 46d. Compound **4** (1.15 g, 1.51 mmol, 1 equiv), compound **36e** (0.80 g, 1.68 mmol, 1.1 equiv), EDC.HCl (0.32 g, 1.68 mmol, 1.1 equiv) and DMAP (55 mg, 0.45 mmol, 0.3 equiv) were dissolved in dry DCM (10 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 3/1 as the eluent to give the title compound as a light-yellow oil (1.44 g, 78%).



46e

Compound 46e. Compound **5** (0.37 g, 0.44 mmol, 1 equiv), compound **36e** (0.23 g, 0.48 mmol, 1.1 equiv), EDC.HCl (92 mg, 0.48 mmol, 1.1 equiv) and DMAP (27 mg, 0.22 mmol, 0.5

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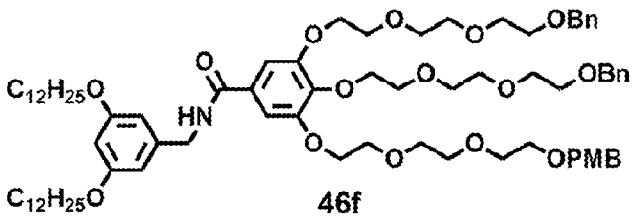
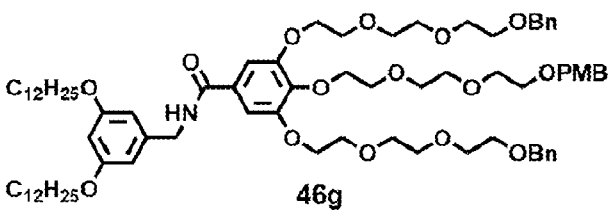
equiv) were dissolved in dry DCM (8 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (20 mL) was added, and the mixture was extracted by DCM (20 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 3/1 as the eluent to give the title compound as a light-yellow oil (0.48 g, 84%).



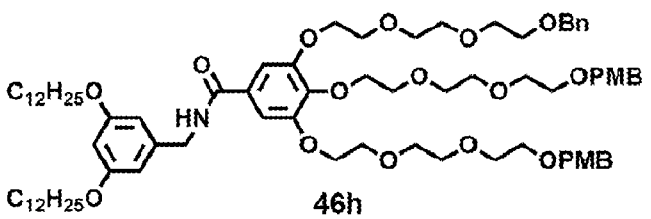
46f

Compound 46f. Compound **6** (0.45 g, 0.52 mmol, 1 equiv), compound **36e** (0.27 g, 0.57 mmol, 1.1 equiv), EDC.HCl (0.11 g, 0.57 mmol, 1.1 equiv) and DMAP (19 mg, 0.16 mmol, 0.3 equiv) were dissolved in dry DCM (6 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 3/1 as the eluent to give the title compound as a light-yellow oil (0.59 g, 86%).



46g

Compound 46g. Compound 7 (0.45 g, 0.52 mmol, 1 equiv), compound **36e** (0.27 g, 0.57 mmol, 1.1 equiv), EDC.HCl (0.11 g, 0.57 mmol, 1.1 equiv) and DMAP (19 mg, 0.16 mmol, 0.3 equiv) were dissolved in dry DCM (6 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 3/1 as the eluent to give the title compound as a light-yellow oil (0.54 g, 78%).



46h

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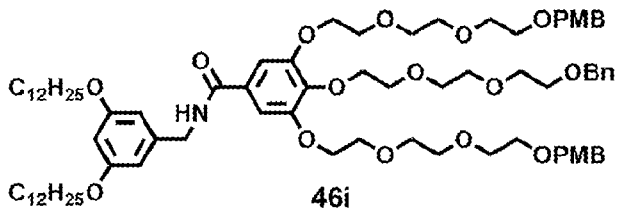
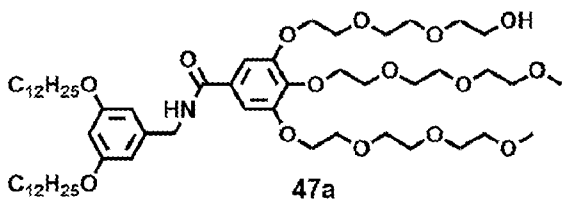
Compound 46h. Compound **8** (0.83 g, 0.92 mmol, 1 equiv), compound **36e** (0.48 g, 1.01 mmol, 1.1 equiv), EDC.HCl (0.19 g, 1.01 mmol, 1.1 equiv) and DMAP (34 mg, 0.28 mmol, 0.3 equiv) were dissolved in dry DCM (8 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 3/1 as the eluent to give the title compound as a light-yellow oil (1.06 g, 85%).



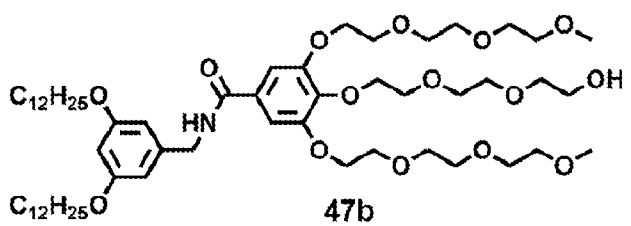
46i

Compound 46i. Compound **9** (0.86 g, 0.96 mmol, 1 equiv), compound **36e** (0.50 g, 1.05 mmol, 1.1 equiv), EDC.HCl (0.20 g, 1.05 mmol, 1.1 equiv) and DMAP (35 mg, 0.29 mmol, 0.3 equiv) were dissolved in dry DCM (8 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 3/1 as the eluent to give the title compound as a light-yellow oil (1.07 g, 83%).



47a

Compound 47a. Compound **46a** (0.46 g, 0.40 mmol) was dissolved in 1:2 MeOH:DCM (15 mL). Then Pd/C (23.0 mg, 5 wt%) was added and the flask was evacuated and filled with hydrogen for three times. The mixture was stirred at 23 °C for 12 h under hydrogen atmosphere. The reaction mixture was filtered through Celite and the filter cake was washed with DCM. Evaporation of the solvent yielded the title compound as a colorless oil (0.42 g, 100%).



47b

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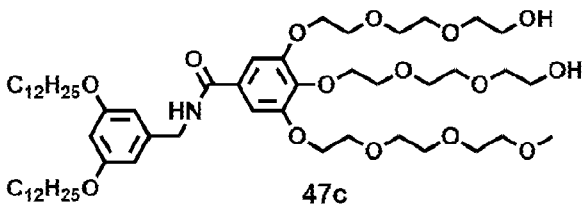
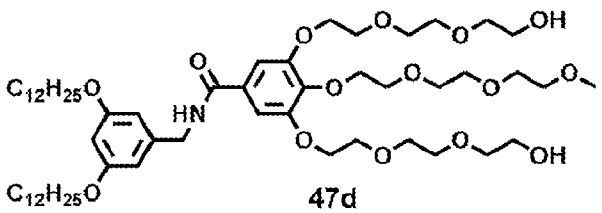
Compound 47b. Compound **47b** was synthesized from compound **46b** (0.45 g, 0.39 mmol) following a procedure similar to that used for the synthesis of compound **47a**. The title compound was obtained as a light-yellow oil (0.41 g, 100%).



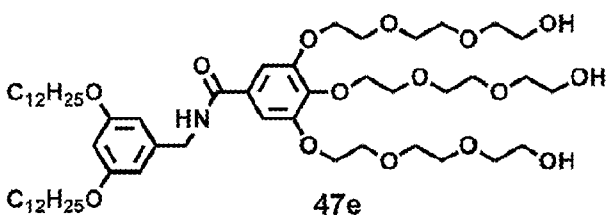
47c

Compound 47c. Compound **47c** was synthesized from compound **46c** (1.55 g, 1.27 mmol) following a procedure similar to that used for the synthesis of compound **47a**. The title compound was obtained as a colorless oil (1.31 g, 100%)..



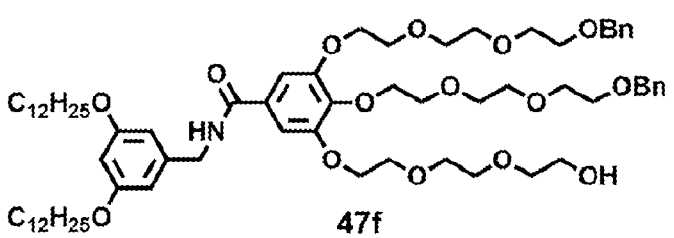
47d

Compound 47d. Compound **47d** was synthesized from compound **46d** (1.40 g, 1.15 mmol) following a procedure similar to that used for the synthesis of compound **47a**. The title compound was obtained as a colorless oil (1.19 g, 100%).



47e

Compound 47e. Compound **47e** was synthesized from compound **46e** (0.40 g, 0.31 mmol) following a procedure similar to that used for the synthesis of compound **47a**. The title compound was obtained as a light-yellow oil (0.31 g, 100%).



47f

Compound 47f. Compound **46f** (0.51 g, 0.38 mmol, 1 equiv) was dissolved in 5 mL DCM and 0.25 mL water (5%) was added. To this solution was added 2,3-dichloro-5,6-dicyano-1,4-

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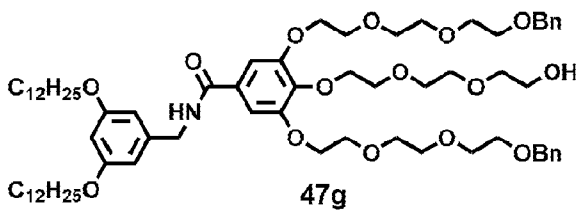
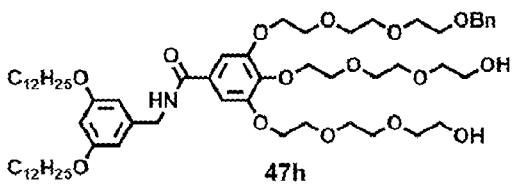
benzoquinone (DDQ, 95 mg, 0.42 mmol, 1.1 equiv). The mixture was allowed to stir at 23 °C for 1 h. The precipitates were filtered out and DCM (20 mL) was added. The mixture was washed by $NaHCO_3$ aqueous solution (saturated), $NaHSO_3$ aqueous solution (2%) and $NaHCO_3$ aqueous solution (saturated) successively. The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 20/1 as the eluent to give the title compound as a yellow oil (0.42 g, 91%).



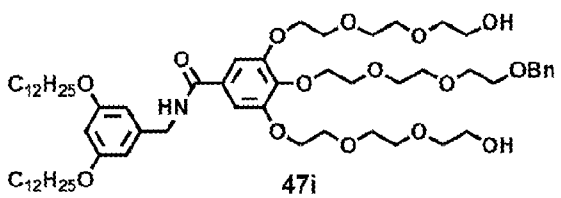
47g

Compound 47g. Compound **47g** was synthesized from compound **46g** (0.44 g, 0.33 mmol) following a procedure similar to that used for the synthesis of compound **47f**. The title compound was obtained as a yellow oil (0.39 g, 82%).



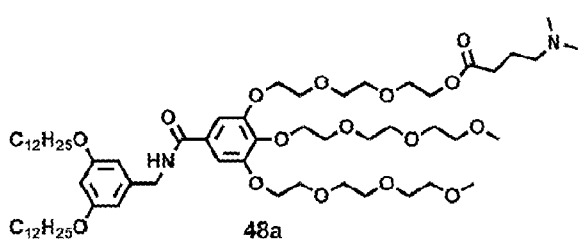
47h

Compound 47h. Compound **47h** was synthesized from compound **46h** (0.74 g, 0.55 mmol) following a procedure similar to that used for the synthesis of compound **47f**. The title compound was obtained as a light-yellow oil (0.56 g, 92%).



47i

Compound 47i. Compound **47i** was synthesized from compound **46i** (0.80 g, 0.59 mmol) following a procedure similar to that used for the synthesis of compound **47f**. The title compound was obtained as a light-yellow oil (0.51 g, 79%).



48a

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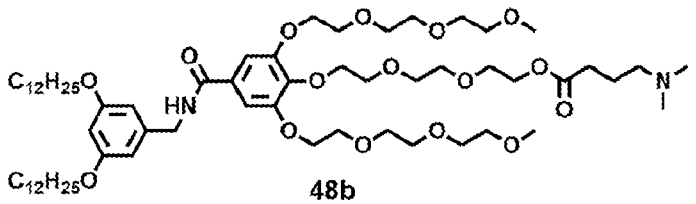
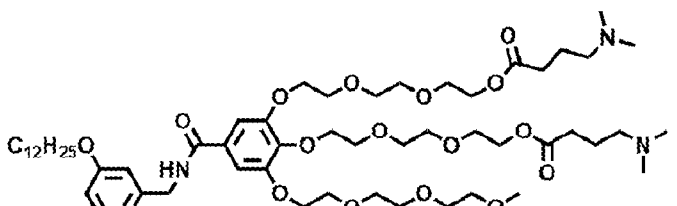
Compound 48a ($3/1_{DMBA}^{1}$). Compound **47a** (0.40 g, 0.38 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (70 mg, 0.42 mmol, 1.1 equiv) were dissolved in 8 mL dry DCM. *N*,*N*'-Dicyclohexylcarbodiimide (DCC, 0.17 g, 0.57 mmol, 1.5 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 8/1 as the eluent to give the title compound as a colorless oil (0.39 g, 89%).



48b

Compound 48b ($3/1_{DMBA}^{2}$). Compound **47b** (0.37 g, 0.35 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (65 mg, 0.39 mmol, 1.1 equiv) were dissolved in 6 mL dry DCM. DCC (0.11 g, 0.53 mmol, 1.5 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 8/1 as the eluent to give the title compound as a light-yellow oil (0.37 g, 89%).



48c

Compound 48c ($3/2_{DMBA}^{1,2}$). Compound **47c** (0.40 g, 0.38 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (0.15 g, 0.89 mmol, 2.3 equiv) were dissolved in 8 mL dry DCM. DCC (0.24 g, 1.16 mmol, 3 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 8/1, 6/1 with 0.1% triethylamine (TEA) as the eluent. Then the crude product was dissolved in sodium bicarbonate ($NaHCO_3$) solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil

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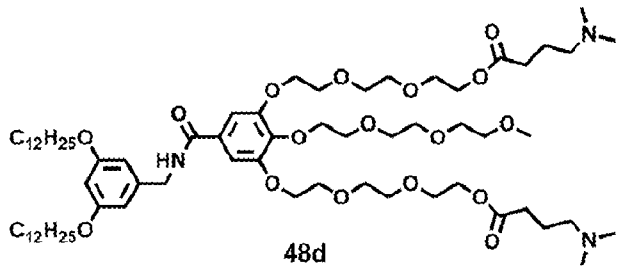
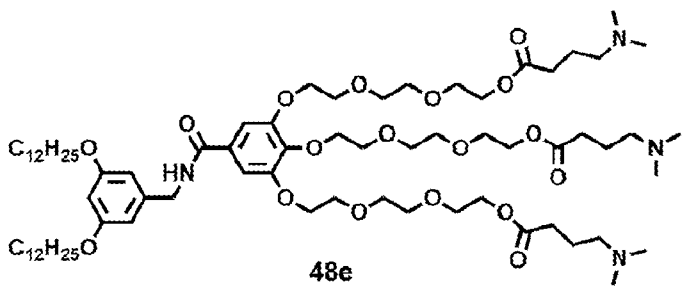
(0.38 g, 79%).



48d

Compound 48d (3/2$_{DMBA}^{1,3}$). Compound **47d** (0.26 g, 0.25 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (97 mg, 0.58 mmol, 2.3 equiv) were dissolved in 6 mL dry DCM. DCC (0.16 g, 0.75 mmol, 3 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 8/1, 6/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.28 g, 88%).



48e

Compound 48e (3/3$_{DMBA}^{1,2,3}$). Compound **47e** (0.27 g, 0.26 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (0.15 g, 0.87 mmol, 3.3 equiv) were dissolved in 6 mL dry DCM. DCC (0.24 g, 1.17 mmol, 4.5 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 8/1, 6/1 and 4/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.31 g, 88%).

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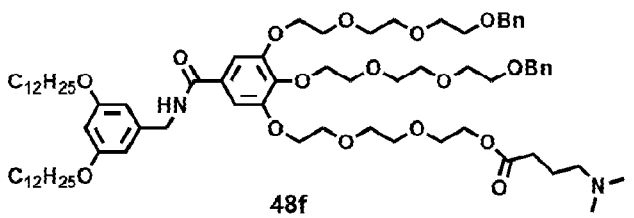
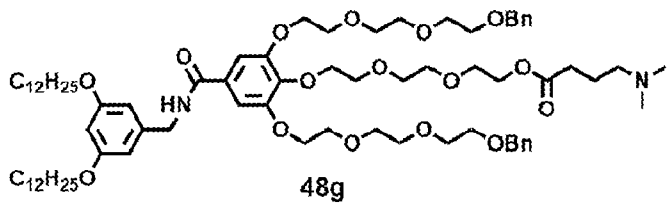


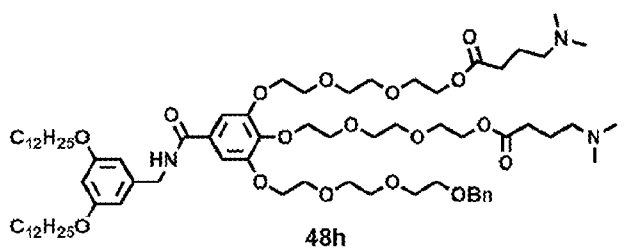
48f

Compound 48f (3/1$_{DMBA}^{1}_{Bn}$^{2,3}). Compound **47f** (0.30 g, 0.25 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (50 mg, 0.30 mmol, 1.2 equiv) were dissolved in 6 mL dry DCM. DCC (0.11 g, 0.50 mmol, 2 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 7/1 as the eluent to give the title compound as a light-yellow oil (0.29 g, 88%).



48g

Compound 48g (3/1$_{DMBA}^{2}_{Bn}$^{1,3}). Compound **47g** (0.29 g, 0.24 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (49 mg, 0.29 mmol, 1.2 equiv) were dissolved in 6 mL dry DCM. DCC (0.10 g, 0.48 mmol, 2 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 6/1 as the eluent to give the title compound as a light-yellow oil (0.28 g, 86%).



48h

Compound 48h (3/2$_{DMBA}^{1,2}_{Bn}$^{3}). Compound **47h** (0.31 g, 0.28 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (0.11 g, 0.66 mmol, 2.3 equiv) were dissolved in 6 mL dry DCM. DCC (0.18 g, 0.84 mmol, 3 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea,

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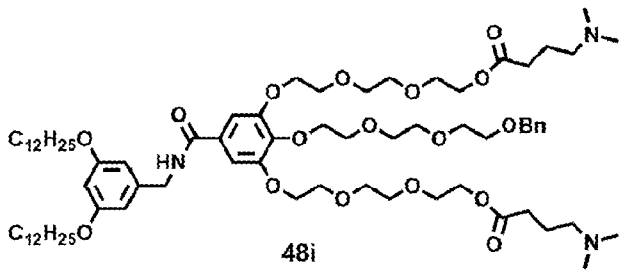
which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 8/1, 6/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.35 g, 93%).



48i

Compound 48i ($3/2_{DMBA}^{1,3}{}_{Bn}^{2}$). Compound **47i** (0.43 g, 0.39 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (0.14 g, 0.85 mmol, 2.3 equiv) were dissolved in 6 mL dry DCM. DCC (0.24 g, 1.16 mmol, 3 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 8/1, 6/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.42 g, 82%).

3.3 Synthesis of Single-Single IAJDs in Library 2

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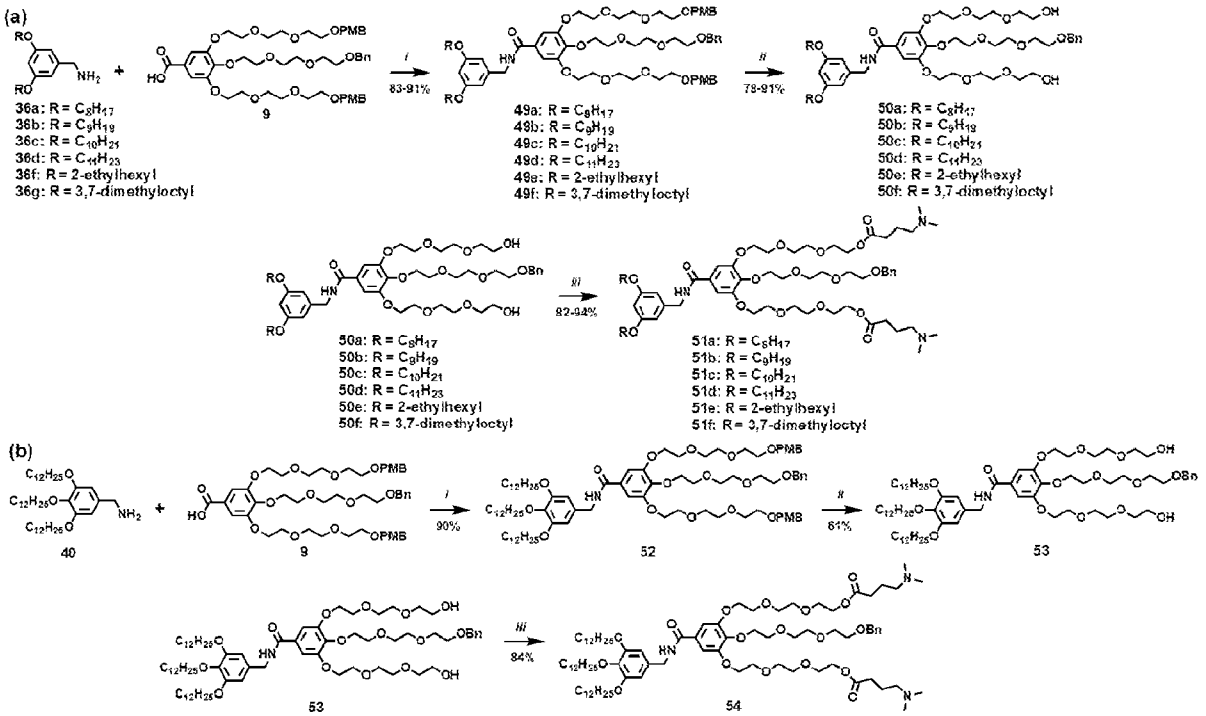
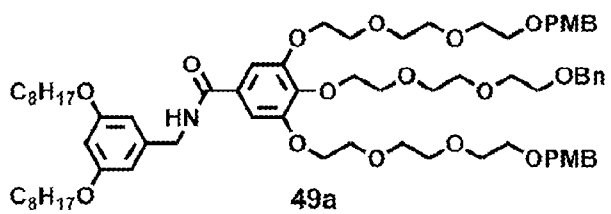


Reagents and conditions: (*i*) EDC.HCl, DMAP, DCM, 23 °C, 12 h; (*ii*) DDQ, DCM/H_2O, 23 °C, 1 h; (*iii*) 4-(Dimethylamino)butyric acid hydrochloride, DCC, DCM, 23 °C, 12 h.



Compound 49a. Compound **9** (0.40 g, 0.45 mmol, 1 equiv), compound **36a** (0.18 g, 0.50 mmol, 1.1 equiv), EDC.HCl (96 mg, 0.50 mmol, 1.1 equiv) and DMAP (17 mg, 0.14 mmol, 0.3 equiv) were dissolved in dry DCM (6 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 4/1 as the eluent to give the title compound as a light-yellow oil (0.51 g, 91%).

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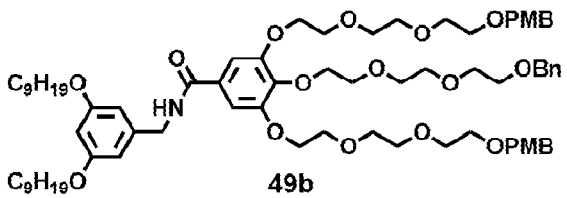
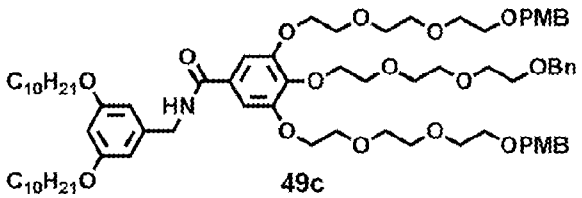


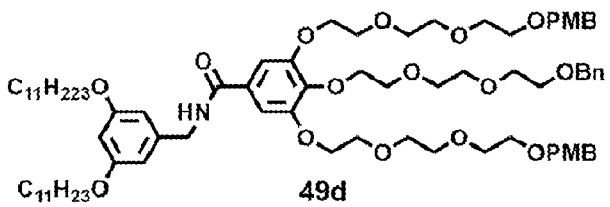
49b

Compound 49b. Compound **49b** was synthesized from compound **9** (0.40 g, 0.45 mmol) and compound **36b** (0.20 g, 0.50 mmol) following a procedure similar to that used for the synthesis of compound **49a**. The title compound was obtained as a yellow oil (0.48 g, 84%).



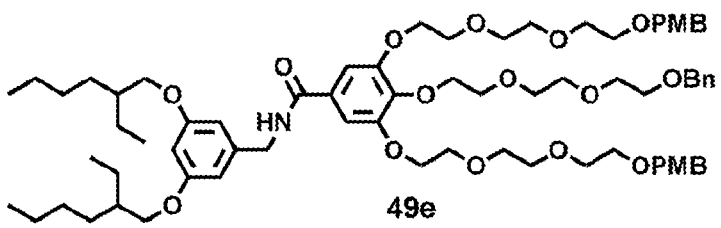
49c

Compound 49c. Compound **49c** was synthesized from compound **9** (0.40 g, 0.45 mmol) and compound **36c** (0.21 g, 0.50 mmol) following a procedure similar to that used for the synthesis of compound **49a**. The title compound was obtained as a yellow oil (0.48 g, 83%).



49d

Compound 49d. Compound **49d** was synthesized from compound **9** (0.35 g, 0.39 mmol) and compound **36d** (0.19 g, 0.43 mmol) following a procedure similar to that used for the synthesis of compound **49a**. The title compound was obtained as a light-yellow oil (0.43 g, 83%).



49e

Compound 49e. Compound **49e** was synthesized from compound **9** (0.40 g, 0.45 mmol) and compound **36f** (0.18 g, 0.50 mmol) following a procedure similar to that used for the synthesis of compound **49a**. The title compound was obtained as a light-yellow oil (0.49 g, 88%).

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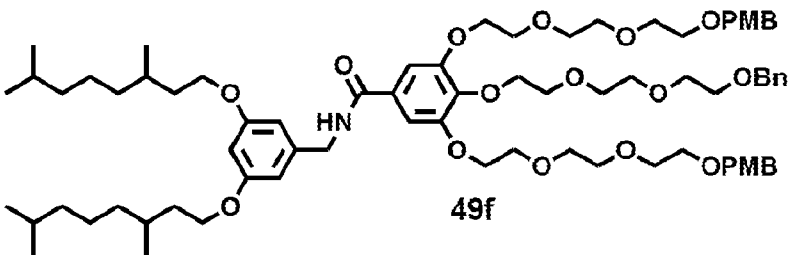
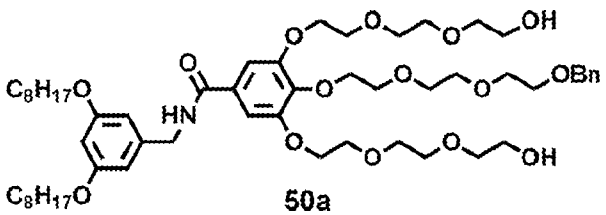


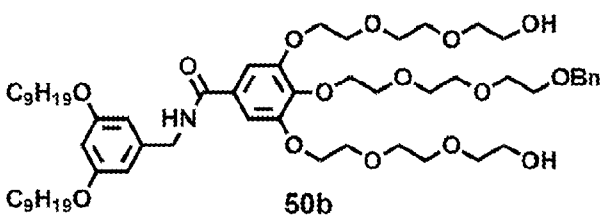
49f

Compound 49f. Compound **49f** was synthesized from compound **9** (0.40 g, 0.45 mmol) and compound **36g** (0.21 g, 0.50 mmol) following a procedure similar to that used for the synthesis of compound **49a**. The title compound was obtained as a light-yellow oil (0.48 g, 85%).



50a

Compound 50a. Compound **49a** (0.49 g, 0.39 mmol, 1 equiv) was dissolved in 6 mL DCM and 0.3 mL water (5%) was added. To this solution was added DDQ (0.20 g, 0.86 mmol, 2.2 equiv). The mixture was allowed to stir at 23 °C for 1 h. The precipitates were filtered out and DCM (20 mL) was added. The mixture was washed by $NaHCO_3$ aqueous solution (saturated), $NaHSO_3$ aqueous solution (2%) and $NaHCO_3$ aqueous solution (saturated) successively. The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 10/1 as the eluent to give the title compound as a light-yellow oil (0.33 g, 85%).



50b

Compound 50b. Compound **50b** was synthesized from compound **49b** (0.48 g, 0.38 mmol) following a procedure similar to that used for the synthesis of compound **50a**. The title compound was obtained as a light-yellow oil (0.32 g, 78%).

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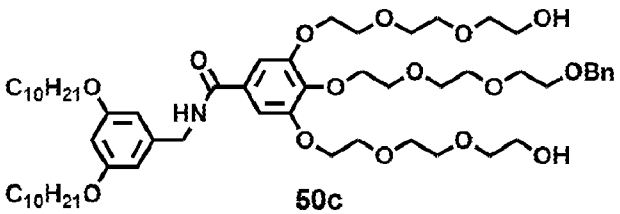
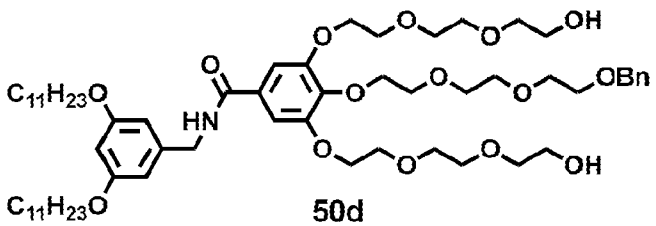


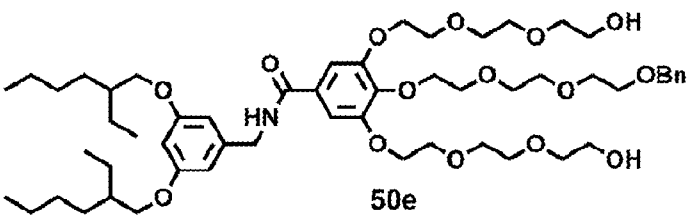
50c

Compound 50c. Compound **50c** was synthesized from compound **49c** (0.43 g, 0.34 mmol) following a procedure similar to that used for the synthesis of compound **50a**. The title compound was obtained as a yellow oil (0.30 g, 86%).



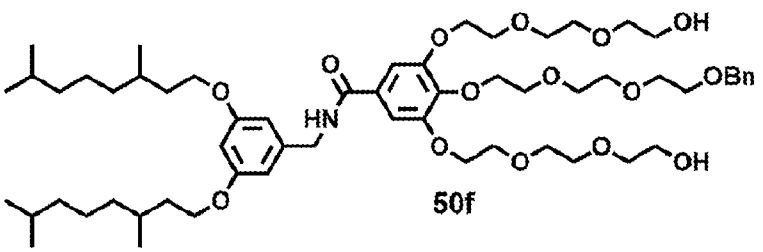
50d

Compound 50d. Compound **50d** was synthesized from compound **49d** (0.41 g, 0.31 mmol) following a procedure similar to that used for the synthesis of compound **50a**. The title compound was obtained as a yellow oil (0.31 g, 91%).



50e

Compound 50e. Compound **50e** was synthesized from compound **49e** (0.44 g, 0.35 mmol) following a procedure similar to that used for the synthesis of compound **50a**. The title compound was obtained as a yellow oil (0.32 g, 91%).



50f

Compound 50f. Compound **50f** was synthesized from compound **49f** (0.48 g, 0.37 mmol) following a procedure similar to that used for the synthesis of compound **50a**. The title compound was obtained as a yellow oil (0.31 g, 78%).

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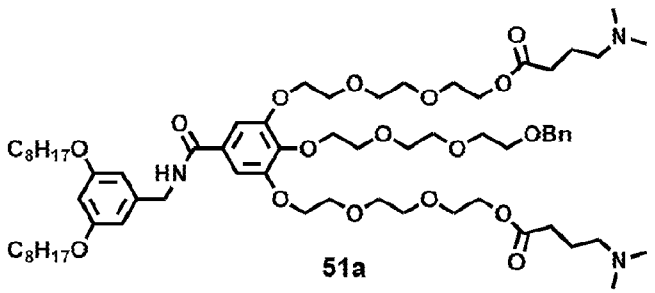
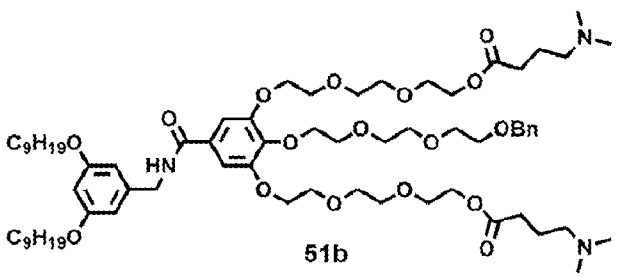
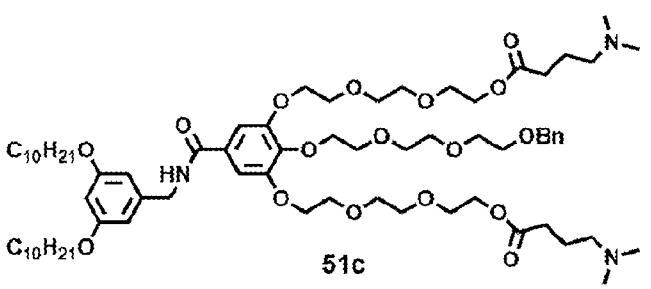


Compound 51a (3/2$_{DMBA}^{1,3}$$_{Bn}^{2}$-C8). Compound **50a** (0.27 g, 0.27 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (0.10 g, 0.59 mmol, 2.2 equiv) were dissolved in 5 mL dry DCM. DCC (0.17 g, 0.81 mmol, 3 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 10/1, 6/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.31 g, 94%).



Compound 51b (3/2$_{DMBA}^{1,3}$$_{Bn}^{2}$-C9). Compound **51b** was synthesized from compound **50b** (0.43 g, 0.42 mmol), 4-(dimethylamino)butyric acid hydrochloride (0.15 g, 0.92 mmol) and DCC (0.26 g, 1.26 mmol) following a procedure similar to that used for the synthesis of compound **51a**. The title compound was obtained as a light-yellow oil (0.40 g, 82%).



Compound 51c (3/2$_{DMBA}^{1,3}$$_{Bn}^{2}$-C10). Compound **51c** was synthesized from compound **50c** (0.25 g, 0.24 mmol), 4-(dimethylamino)butyric acid hydrochloride (87 mg, 0.52 mmol) and DCC

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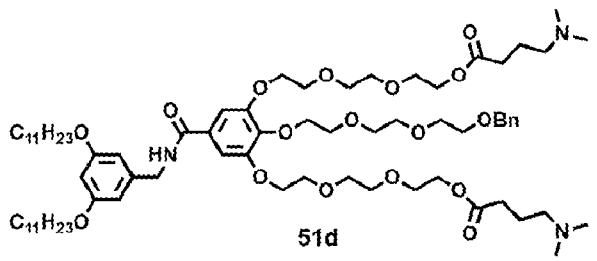
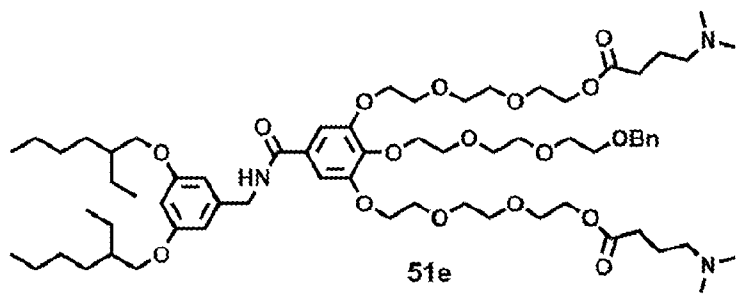
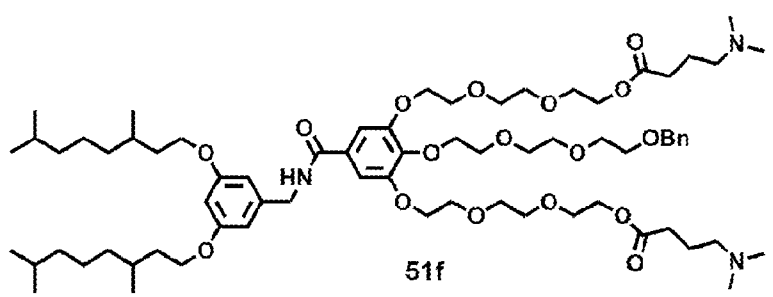
(0.15 g, 0.71 mmol) following a procedure similar to that used for the synthesis of compound **51a**. The title compound was obtained as a light-yellow oil (0.26 g, 84%).



Compound 51d (3/2$_{DMBA}^{1,3}$$_{Bn}^{2}$-C11). Compound **51d** was synthesized from compound **50d** (0.23 g, 0.21 mmol), 4-(dimethylamino)butyric acid hydrochloride (77 mg, 0.46 mmol) and DCC (0.13 g, 0.63 mmol) following a procedure similar to that used for the synthesis of compound **51a**. The title compound was obtained as a light-yellow oil (0.23 g, 84%).



Compound 51e (3/2$_{DMBA}^{1,3}$$_{Bn}^{2}$-EH). Compound **51e** was synthesized from compound **50e** (0.29 g, 0.29 mmol), 4-(dimethylamino)butyric acid hydrochloride (0.11 g, 0.64 mmol) and DCC (0.18 g, 0.87 mmol) following a procedure similar to that used for the synthesis of compound **51a**. The title compound was obtained as a light-yellow oil (0.32 g, 89%).



Compound 51f (3/2$_{DMBA}^{1,3}$$_{Bn}^{2}$-dm8). Compound **51f** was synthesized from compound **50f** (0.30 g, 0.28 mmol), 4-(dimethylamino)butyric acid hydrochloride (104 mg, 0.62 mmol) and DCC (0.17 g, 0.84 mmol) following a procedure similar to that used for the synthesis of compound **51a**. The title compound was obtained as a light-yellow oil (0.31 g, 86%).

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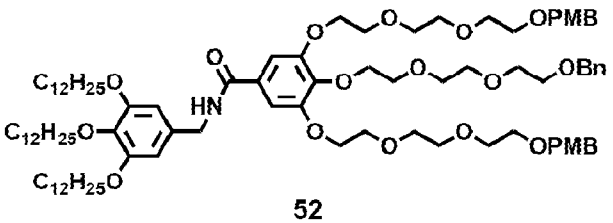
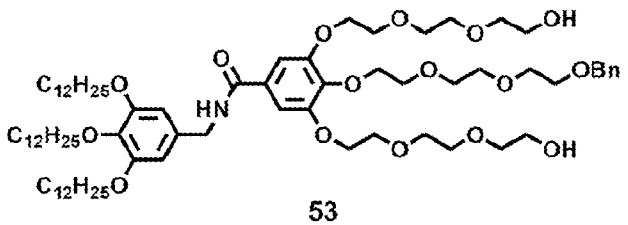


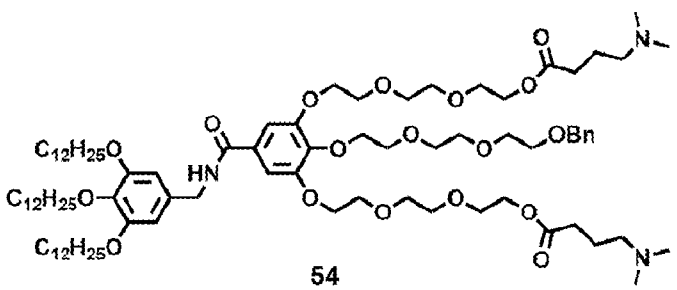
52

Compound 52. Compound **52** was synthesized from compound **9** (0.30 g, 0.33 mmol) and compound **40** (0.24 g, 0.37 mmol) following a procedure similar to that used for the synthesis of compound **49a**. The title compound was obtained as a white viscous solid (0.46 g, 90%).



53

Compound 53. Compound **53** was synthesized from compound **52** (0.45 g, 0.29 mmol) following a procedure similar to that used for the synthesis of compound **50a**. The title compound was obtained as a yellow oil (0.23 g, 61%).



54

Compound 54 ($3/2_{DMBA}^{1,3}{}_{Bn}^{2}$-3,4,5-C12). Compound **54** was synthesized from compound **53** (0.21 g, 0.16 mmol), 4-(dimethylamino)butyric acid hydrochloride (60 mg, 0.36 mmol) and DCC (0.10 g, 0.49 mmol) following a procedure similar to that used for the synthesis of compound **51a**. The title compound was obtained as a light-yellow solid (0.21 g, 84%).

Synthesis of Single-Single IAJDs in Library 3

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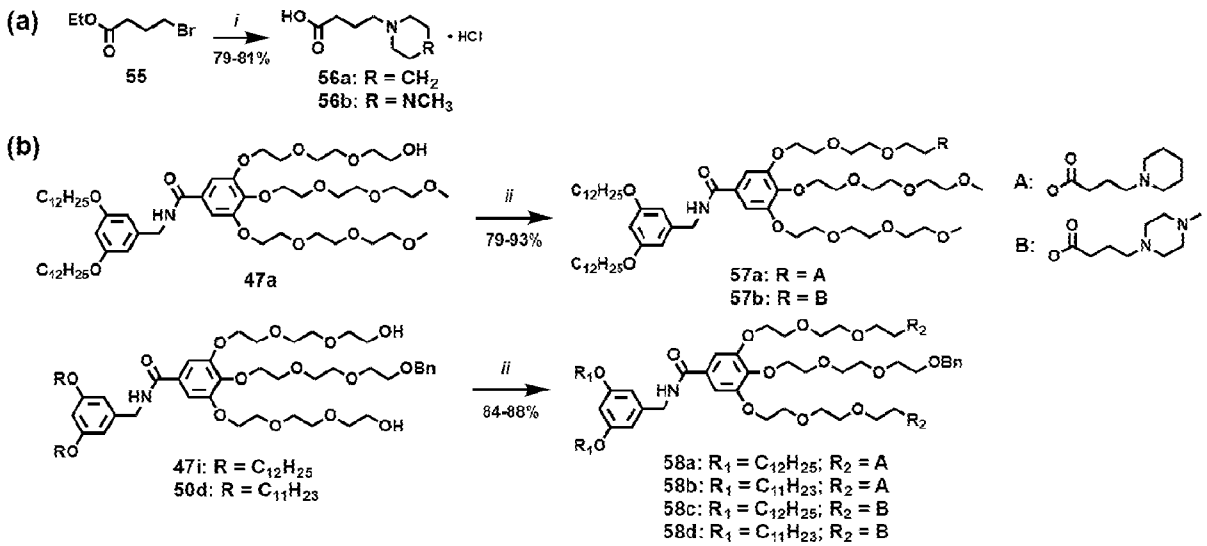
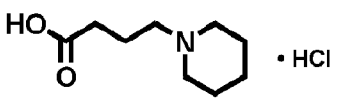


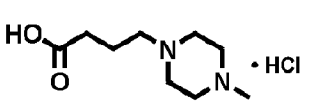
Reagents and conditions: (*i*) (1) piperidine/1-methylpiperazine, K_2CO_3, MeCN, reflux, 3h; (2) conc. HCl/water, reflux, 3 h; (*ii*) 4-(piperidin-1-yl)butanoic acid hydrochloride/4-(4-methylpiperazin-1-yl)butanoic acid hydrochloride, DCC, DCM, 23 °C, 12 h.

The synthesis of compounds **56a** and **56b** was adapted from literature procedure (May, et al., Bioconjugate Chem. 2016, 27, 226−237).



56a

4-(Piperidin-1-yl)butanoic acid hydrochloride (56a). Ethyl 4-bromobutanoate (**55**, 1.00 g, 5.15 mmol, 1 equiv), piperidine (0.46 g, 5.41 mmol, 1.05 equiv) and K_2CO_3 (0.70 g, 5.15 mmol, 1 equiv) were stirred in MeCN (20 mL). The mixture was heated at reflux for 3h. The mixture was cooled and extracted by DCM (30 mL × 3). The organic phase was washed with $NaHCO_3$ aqueous solution (saturated) (20 mL × 2), dried over anhydrous $MgSO_4$ and filtered. The solvent was evaporated to leave an orange precipitate. The precipitate was suspended in conc. HCl/water (1/1, 10 mL) and the mixture was heated at reflux for 3 h. The solvent was evaporated to give the crude product, which was recrystallized in EtOH to give the title compound as a white solid (0.86 g, 81%).



56b

4-(4-Methylpiperazin-1-yl)butanoic acid hydrochloride (56b). Compound **56b** was synthesized from compound **55** (1.50 g, 7.73 mmol) and 1-methylpiperazine (0.81 g, 8.12 mmol)

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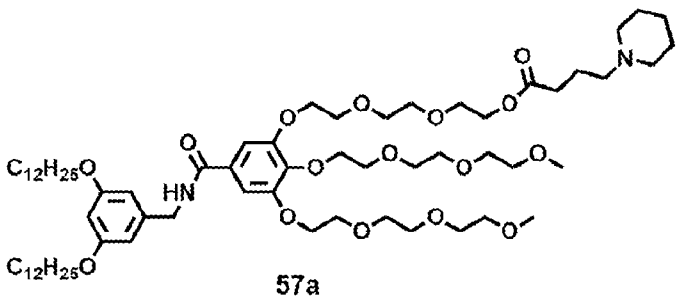
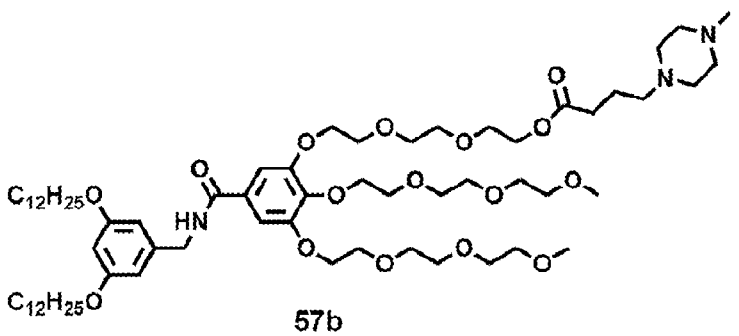
following a procedure similar to that used for the synthesis of compound **56a**. The title compound was obtained as a white solid (1.31 g, 79%).



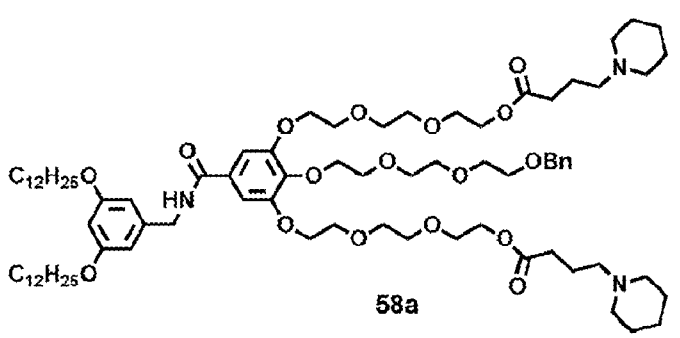
57a

Compound 57a (3/1$_{PIP}^{1}$). Compound **57a** was synthesized from compound **47a** (0.25 g, 0.24 mmol), 4-(piperidin-1-yl)butanoic acid hydrochloride (**56a**, 0.11 g, 0.53 mmol) and DCC (0.15 g, 0.72 mmol) following a procedure similar to that used for the synthesis of compound **51a**. The title compound was obtained as a light-yellow oil (0.23 g, 79%).



57b

Compound 57b (3/1$_{MPRZ}^{1}$). Compound **57b** was synthesized from compound **47a** (0.25 g, 0.24 mmol), 4-(4-methylpiperazin-1-yl)butanoic acid hydrochloride (**56b**, 0.12 g, 0.53 mmol) and DCC (0.15 g, 0.72 mmol) following a procedure similar to that used for the synthesis of compound **51a**. The title compound was obtained as a light-yellow oil (0.27 g, 93%).



58a

Compound 58a (3/2$_{PIP}^{1,3}$$_{Bn}^{2}$). Compound **58a** was synthesized from compound **47i** (0.30 g, 0.27 mmol), compound **56a** (0.12 g, 0.59 mmol) and DCC (0.17 g, 0.81 mmol) following a

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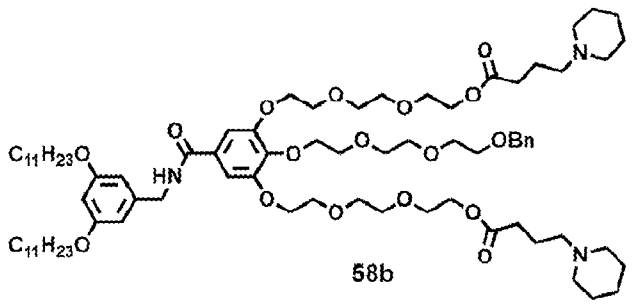
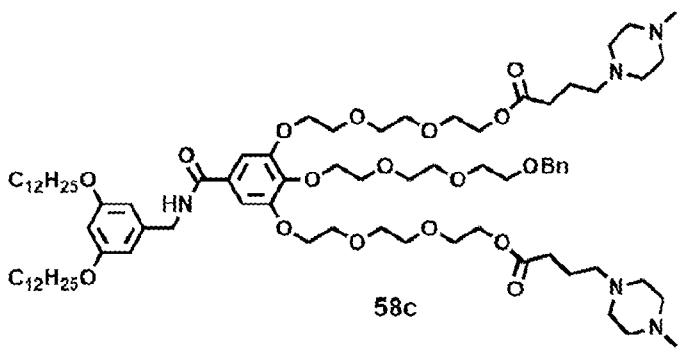
procedure similar to that used for the synthesis of compound **51a**. The title compound was obtained as a light-yellow oil (0.32 g, 84%).



58b

Compound 58b ($3/2_{PIP}^{1,3}{}_{Bn}^{2}$-C11). Compound **58b** was synthesized from compound **50d** (0.28 g, 0.26 mmol), compound **56a** (0.11 g, 0.57 mmol) and DCC (0.16 g, 0.78 mmol) following a procedure similar to that used for the synthesis of compound **51a**. The title compound was obtained as a light-yellow oil (0.31 g, 86%).



58c

Compound 58c ($3/2_{MPRZ}^{1,3}{}_{Bn}^{2}$). Compound **47i** (0.30 g, 0.27 mmol, 1 equiv) and compound **56b** (0.15 g, 0.68 mmol, 2.5 equiv) were dissolved in 6 mL dry DCM. DCC (0.17 g, 0.81 mmol, 3 equiv) was added in one portion into the above mixture. To the reaction mixture was added 3 drops of TEA. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 10/1, 8/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.33 g, 85%).

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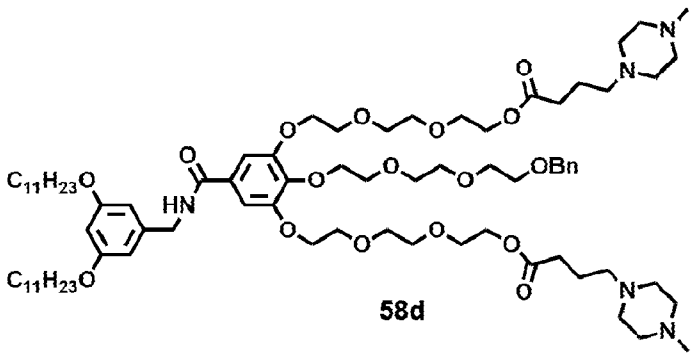
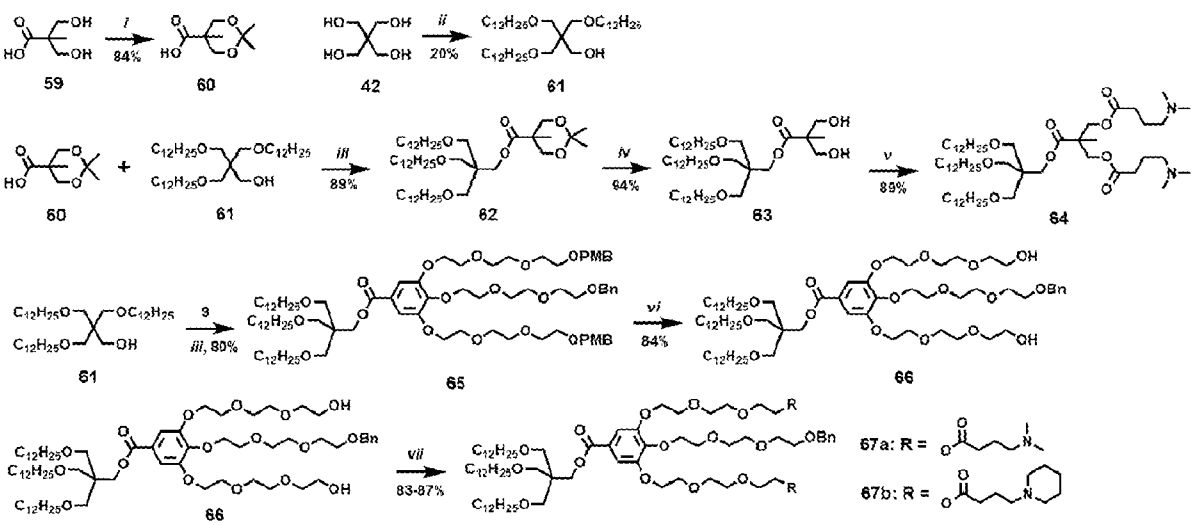


58d

Compound 58d (3/2$_{MPRZ}^{1,3}$$_{Bn}^{2}$-C11). Compound **58d** was synthesized from compound **50d** (0.30 g, 0.28 mmol), compound **56b** (0.14 g, 0.62 mmol) and DCC (0.17 g, 0.84 mmol) following a procedure similar to that used for the synthesis of compound **58c**. The title compound was obtained as a light-yellow oil (0.35 g, 88%).

Synthesis of Single-Single IAJDs in Library 4



Reagents and conditions: (*i*) PTSA·H_2O, 2,2-dimethoxypropane, Acetone, 23 °C, 3 h; (*ii*) $C_{12}H_{25}Br$, 50% NaOH, TBABr (cat.), 80 °C, 5 h; (*iii*) EDC.HCl, DMAP, DCM, 23 °C, 12 h; (*iv*) 2 M HCl, 1,4-dioxane, 60 °C, 5 h; (*v*) 4-(Dimethylamino)butyric acid hydrochloride, DCC, DCM, 23 °C, 12 h; (*vi*) DDQ, DCM/H_2O, 23 °C, 1 h; (*vii*) 4-(Dimethylamino)butyric acid hydrochloride/4-(piperidin-1-yl)butanoic acid hydrochloride, DCC, DCM, 23 °C, 12 h.

Compounds **60** and **61** were synthesized according to literature procedures The procedures were optimized to increase the yields of compounds **60** and **61** to 10 g and 5 g, respectively.

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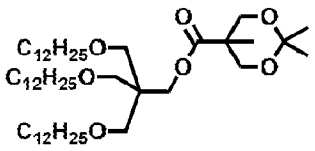
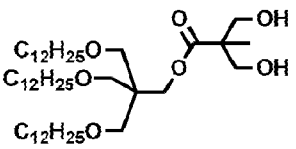


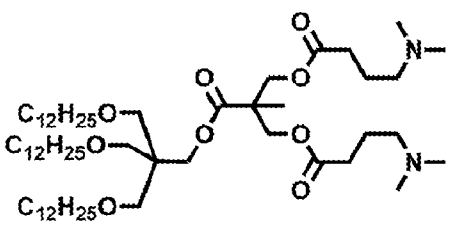
62

Compound 62. Compound **60** (0.22 g, 1.25 mmol, 1 equiv), compound **61** (0.80 g, 1.25 mmol, 1 equiv), EDC.HCl (0.26 g, 1.38 mmol, 1.1 equiv) and DMAP (46 mg, 0.38 mmol, 0.3 equiv) were dissolved in dry DCM (6 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 30/1 as the eluent to give the title compound as a colorless oil (0.89 g, 89%).



63

Compound 63. Compound **62** (0.55 g, 0.69 mmol) was dissolved in 20 mL 1,4-dioxane. Then HCl (2 M, 3.0 mL, 6.0 mmol) was added. The mixture was stirred at 60 °C for 5 h. The reaction mixture was extracted by DCM (3 × 20 mL) and the organic phase was dried over anhydrous $MgSO_4$. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 5/1 as the eluent to give the title compound as a colorless oil (0.49 g, 94%).



64

Compound 64 ($2/2_{DMBA}^{1,2}$). Compound **63** (0.20 g, 0.26 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (96 mg, 0.57 mmol, 2.2 equiv) were dissolved in 5 mL dry DCM. DCC (0.16 g, 0.78 mmol, 3 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 10/1, 6/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM

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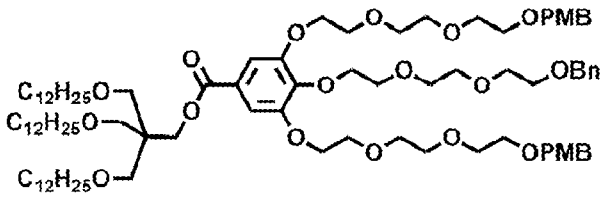
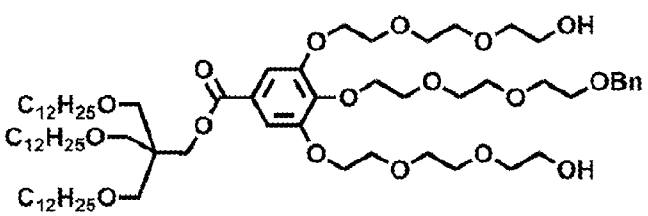
(30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.23 g, 89%).



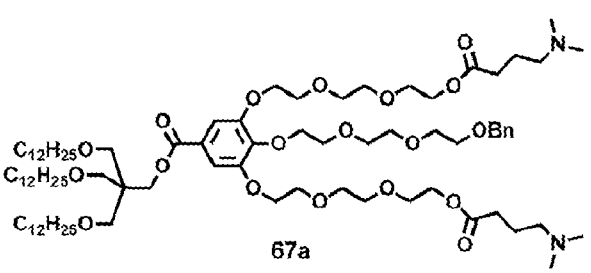
65

Compound 65. Compound **61** (0.30 g, 0.47 mmol, 1.1 equiv), compound **9** (0.38 g, 0.43 mmol, 1 equiv), EDC.HCl (90 mg, 0.47 mmol, 1.1 equiv) and DMAP (16 mg, 0.13 mmol, 0.3 equiv) were dissolved in dry DCM (6 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 3/1 as the eluent to give the title compound as a light-yellow oil (0.52 g, 80%).



66

Compound 66. Compound **65** (0.36 g, 0.24 mmol, 1 equiv) was dissolved in 6 mL DCM and 0.3 mL water (5%) was added. To this solution was added DDQ (0.12 g, 0.53 mmol, 2.2 equiv). The mixture was allowed to stir at 23 °C for 1 h. The precipitates were filtered out and DCM (20 mL) was added. The mixture was washed by $NaHCO_3$ aqueous solution (saturated), $NaHSO_3$ aqueous solution (2%) and $NaHCO_3$ aqueous solution (saturated) successively. The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 10/1 as the eluent to give the title compound as a light-yellow oil (0.26 g, 84%).



67a

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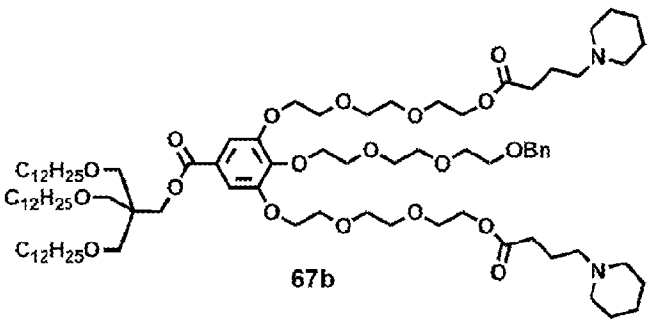
Compound 67a ($3/2_{DMBA}^{1,3}{}_{Bn}^{2}$-PE-3C12). Compound **66** (0.25 g, 0.20 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (74 mg, 0.44 mmol, 2.2 equiv) were dissolved in 5 mL dry DCM. DCC (0.13 g, 0.60 mmol, 3 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 10/1, 5/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.26 g, 87%).



67b

Compound 67b ($3/2_{PIP}^{1,3}{}_{Bn}^{2}$-PE-3C12). Compound **67b** was synthesized from compound **66** (0.24 g, 0.19 mmol), compound **56a** (87 mg, 0.42 mmol) and DCC (0.12 g, 0.57 mmol) following a procedure similar to that used for the synthesis of compound **67a**. The title compound was obtained as a colorless oil (0.25 g, 83%).

Synthesis of Twin-Twin IAJDs of Library 5

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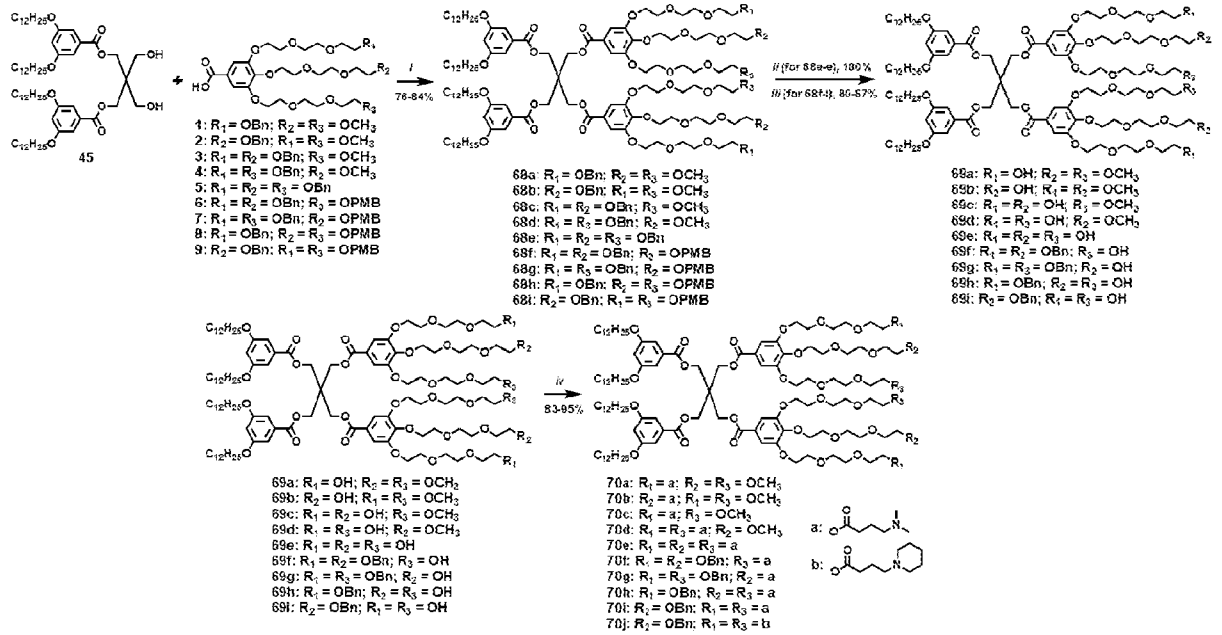
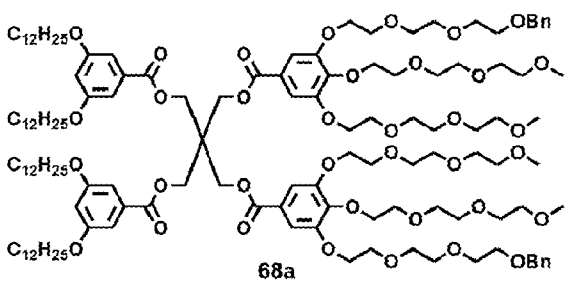


Reagents and conditions: (*i*) EDC.HCl, DMAP, DCM, 23 °C, 12 h; (*ii*) H_2, Pd/C, DCM/MeOH, 23 °C, 12 h; (*iii*) DDQ, DCM/H_2O, 23 °C, 1 h; (*iv*) 4-(Dimethylamino)butyric acid hydrochloride/4-(piperidin-1-yl)butanoic acid hydrochloride, DCC, DCM, 23 °C, 12 h.



Compound 68a. Compound **45** (0.23 g, 0.21 mmol, 1 equiv), compound **1** (0.30 g, 0.44 mmol, 2.1 equiv), EDC.HCl (84 mg, 0.44 mmol, 2.1 equiv) and DMAP (16 mg, 0.13 mmol, 0.6 equiv) were dissolved in dry DCM (5 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 30/1 as the eluent to give the title compound as a light-yellow oil (0.41 g, 81%).

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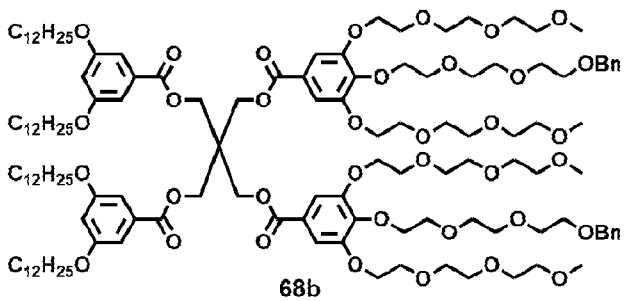
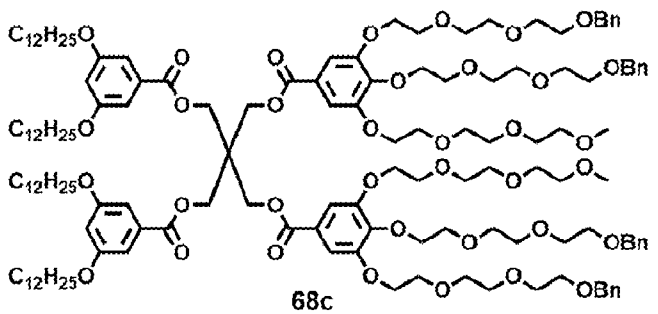


68b

Compound 68b. Compound **45** (0.23 g, 0.21 mmol, 1 equiv), compound **2** (0.30 g, 0.44 mmol, 2.1 equiv), EDC.HCl (84 mg, 0.44 mmol, 2.1 equiv) and DMAP (16 mg, 0.13 mmol, 0.6 equiv) were dissolved in dry DCM (5 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 30/1 as the eluent to give the title compound as a light-yellow oil (0.39 g, 77%).



68c

Compound 68c. Compound **45** (0.21 g, 0.19 mmol, 1 equiv), compound **3** (0.30 g, 0.40 mmol, 2.1 equiv), EDC.HCl (77 mg, 0.40 mmol, 2.1 equiv) and DMAP (14 mg, 0.11 mmol, 0.6 equiv) were dissolved in dry DCM (5 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc as the eluent to give the title compound as a colorless oil (0.41 g, 84%).

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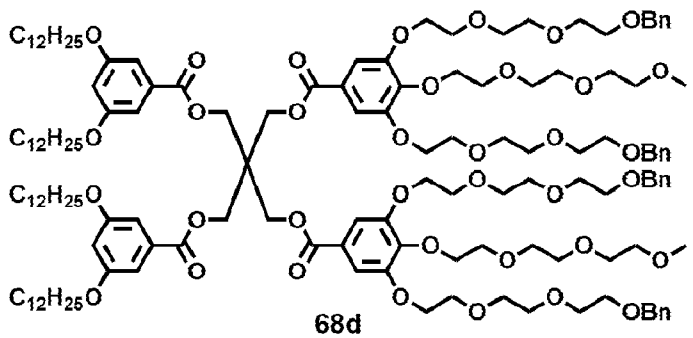
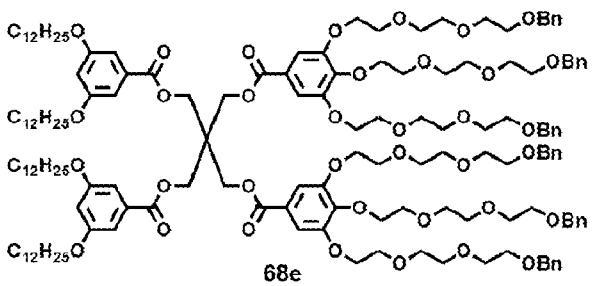


68d

Compound 68d. Compound **45** (0.21 g, 0.19 mmol, 1 equiv), compound **4** (0.30 g, 0.40 mmol, 2.1 equiv), EDC.HCl (77 mg, 0.40 mmol, 2.1 equiv) and DMAP (14 mg, 0.11 mmol, 0.6 equiv) were dissolved in dry DCM (5 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc as the eluent to give the title compound as a colorless oil (0.40 g, 82%).



68e

Compound 68e. Compound **45** (0.18 g, 0.17 mmol, 1 equiv), compound **5** (0.30 g, 0.36 mmol, 2.1 equiv), EDC.HCl (69 mg, 0.36 mmol, 2.1 equiv) and DMAP (12 mg, 0.10 mmol, 0.6 equiv) were dissolved in dry DCM (5 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 3/1 as the eluent to give the title compound as a light-yellow oil (0.36 g, 78%).

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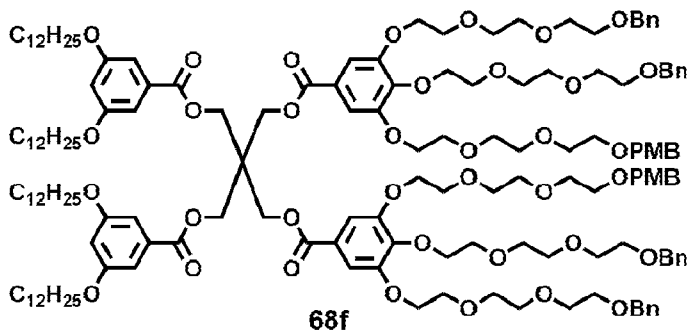
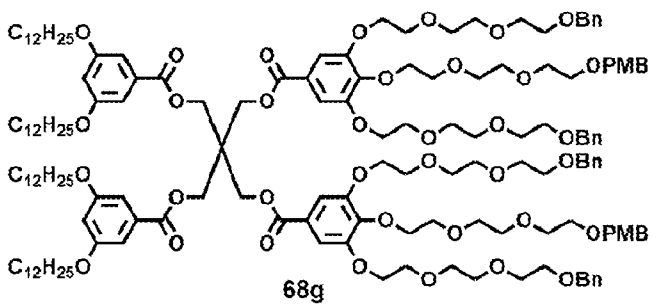


68f

Compound 68f. Compound **45** (0.18 g, 0.17 mmol, 1 equiv), compound **6** (0.30 g, 0.35 mmol, 2.1 equiv), EDC.HCl (67 mg, 0.35 mmol, 2.1 equiv) and DMAP (12 mg, 0.10 mmol, 0.6 equiv) were dissolved in dry DCM (5 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 3/1 as the eluent to give the title compound as a light-yellow oil (0.36 g, 77%).



68g

Compound 68g. Compound **45** (0.18 g, 0.17 mmol, 1 equiv), compound **7** (0.30 g, 0.35 mmol, 2.1 equiv), EDC.HCl (67 mg, 0.35 mmol, 2.1 equiv) and DMAP (12 mg, 0.10 mmol, 0.6 equiv) were dissolved in dry DCM (5 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 3/1 as the eluent to give the title compound as a light-yellow oil (0.39 g, 84%).

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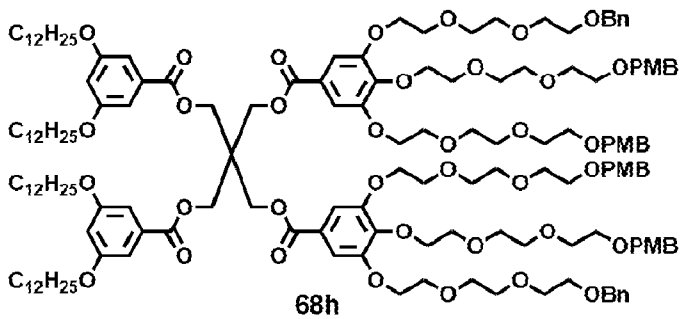
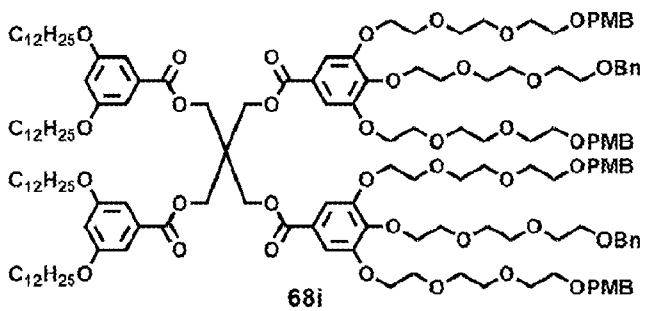


68h

Compound 68h. Compound **45** (0.35 g, 0.32 mmol, 1 equiv), compound **8** (0.60 g, 0.67 mmol, 2.1 equiv), EDC.HCl (0.13 g, 0.67 mmol, 2.1 equiv) and DMAP (24 mg, 0.19 mmol, 0.6 equiv) were dissolved in dry DCM (6 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 4/1 as the eluent to give the title compound as a light-yellow oil (0.68 g, 76%).



68i

Compound 68i. Compound **45** (0.35 g, 0.32 mmol, 1 equiv), compound **9** (0.60 g, 0.67 mmol, 2.1 equiv), EDC.HCl (0.13 g, 0.67 mmol, 2.1 equiv) and DMAP (24 mg, 0.19 mmol, 0.6 equiv) were dissolved in dry DCM (6 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 3/1 as the eluent to give the title compound as a light-yellow oil (0.72 g, 78%).

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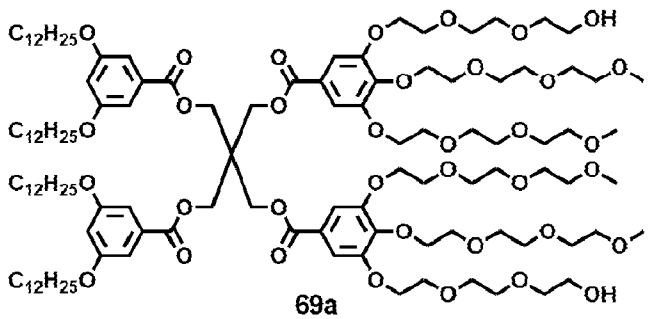
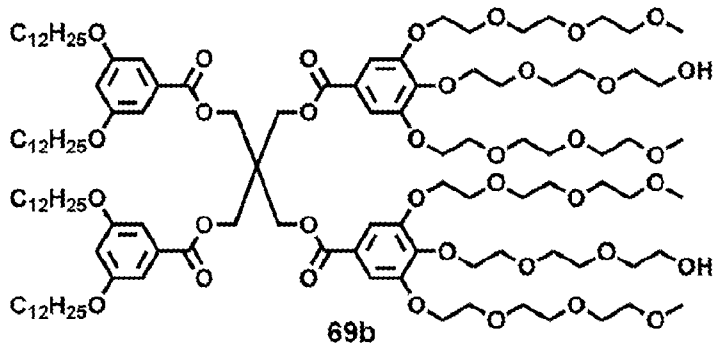


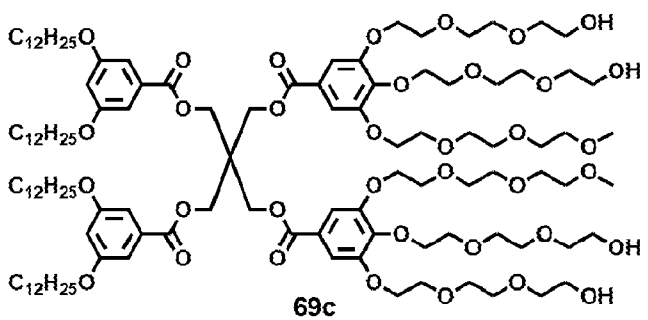
69a

Compound 69a. Compound **68a** (0.39 g, 0.16 mmol) was dissolved in 1:2 MeOH:DCM (15 mL). Then Pd/C (19.5 mg, 5 wt%) was added and the flask was evacuated and filled with hydrogen for three times. The mixture was stirred at 23 °C for 12 h under hydrogen atmosphere. The reaction mixture was filtered through Celite and the filter cake was washed with DCM. Evaporation of the solvent yielded the title compound as a light-yellow oil (0.36 g, 100%).



69b

Compound 69b. Compound **69b** was synthesized from compound **68b** (0.38 g, 0.16 mmol) following a procedure similar to that used for the synthesis of compound **69a**. The title compound was obtained as a light-yellow oil (0.36 g, 100%).



69c

Compound 69c. Compound **69c** was synthesized from compound **68c** (0.39 g, 0.15 mmol) following a procedure similar to that used for the synthesis of compound **69a**. The title compound was obtained as a light-yellow oil (0.33 g, 100%).

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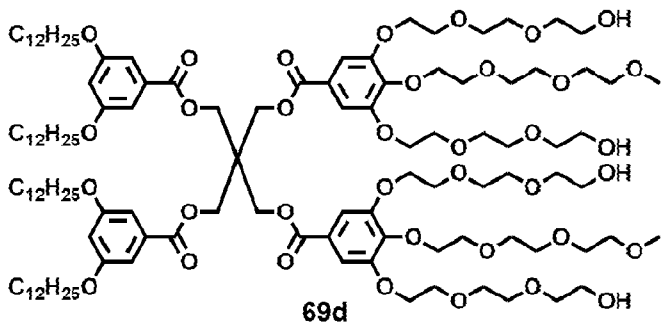
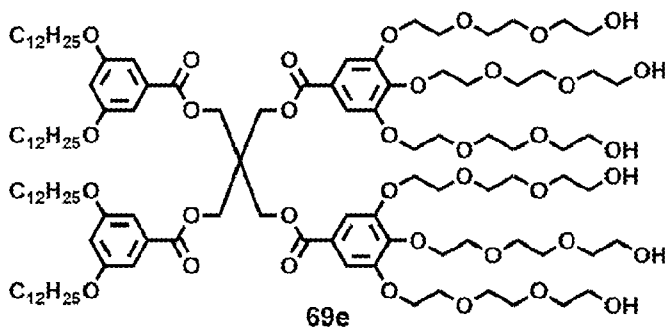


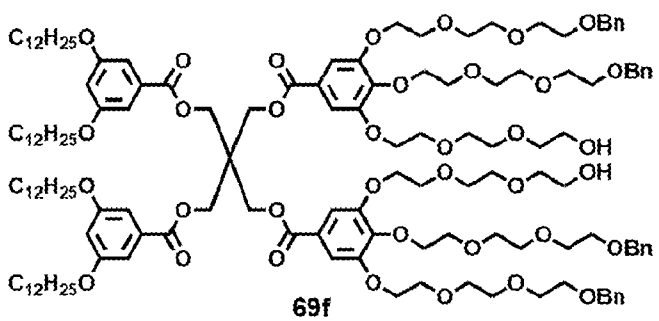
69d

Compound 69d. Compound **69d** was synthesized from compound **68d** (0.39 g, 0.15 mmol) following a procedure similar to that used for the synthesis of compound **69a**. The title compound was obtained as a light-yellow oil (0.33 g, 100%).



69e

Compound 69e. Compound **69e** was synthesized from compound **68e** (0.32 g, 0.12 mmol) following a procedure similar to that used for the synthesis of compound **69a**. The title compound was obtained as a light-yellow oil (0.26 g, 100%).



69f

Compound 69f. Compound **68f** (0.26 g, 0.09 mmol, 1 equiv) was dissolved in 5 mL DCM and 0.25 mL water (5%) was added. To this solution was added DDQ (45 mg, 0.20 mmol, 2.2 equiv). The mixture was allowed to stir at 23 °C for 1 h. The precipitates were filtered out and DCM (20 mL) was added. The mixture was washed by $NaHCO_3$ aqueous solution (saturated), $NaHSO_3$ aqueous solution (2%) and $NaHCO_3$ aqueous solution (saturated) successively. The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and

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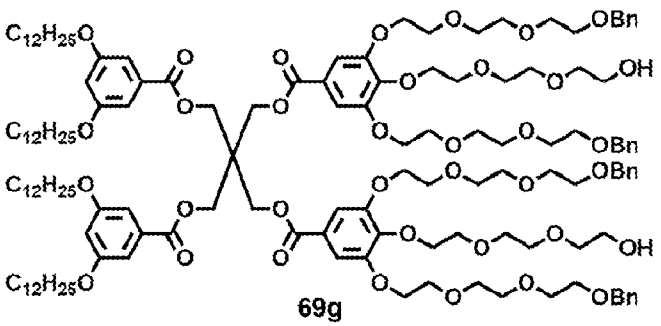
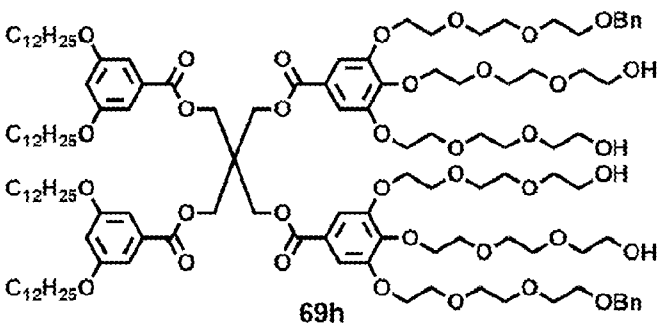
purified by column chromatography (SiO_2) with EtOAc/MeOH = 20/1 as the eluent to give the title compound as a light-yellow oil (0.19 g, 83%).



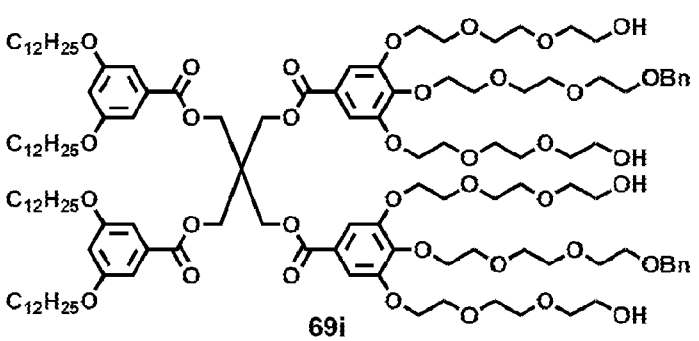
69g

Compound 69g. Compound **69g** was synthesized from compound **68g** (0.18 g, 0.07 mmol) following a procedure similar to that used for the synthesis of compound **69f**. The title compound was obtained as a light-yellow oil (0.13 g, 80%).



69h

Compound 69h. Compound **69h** was synthesized from compound **68h** (0.53 g, 0.19 mmol) following a procedure similar to that used for the synthesis of compound **69f**. The title compound was obtained as a yellow oil (0.39 g, 87%).



69i

Compound 69i. Compound **69i** was synthesized from compound **68i** (0.57 g, 0.20 mmol) following a procedure similar to that used for the synthesis of compound **69f**. The title compound was obtained as a yellow oil (0.40 g, 84%).

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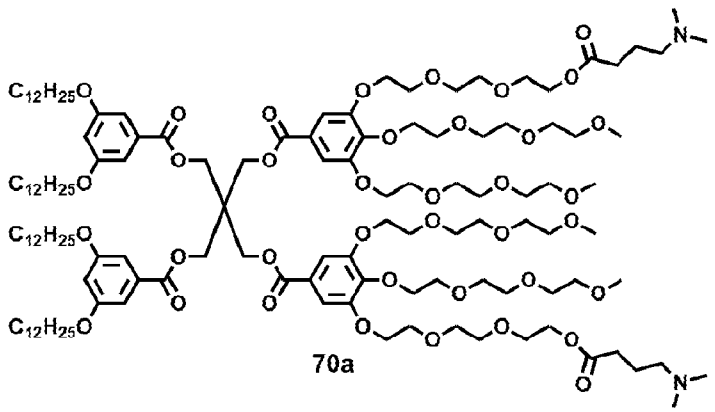
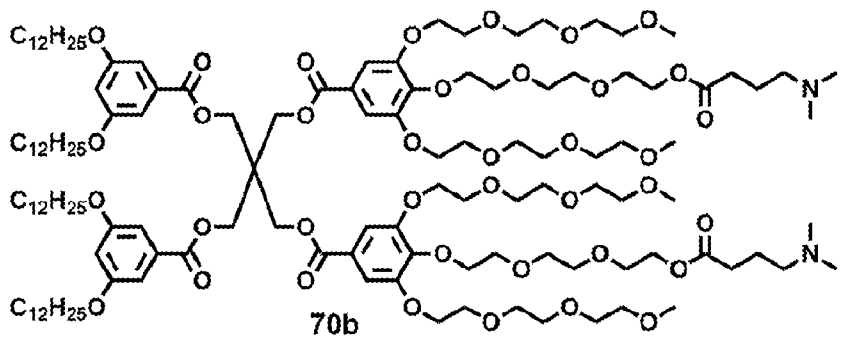


70a

Compound 70a (6/2$_{DMBA}^{1,1'}$). Compound **69a** (0.32 g, 0.14 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (54 mg, 0.32 mmol, 2.3 equiv) were dissolved in 6 mL dry DCM. DCC (87 mg, 0.42 mmol, 3.0 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 8/1, 6/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.33 g, 94%).



70b

Compound 70b (6/2$_{DMBA}^{2,2'}$). Compound **69b** (0.28 g, 0.13 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (48 mg, 0.29 mmol, 2.3 equiv) were dissolved in 6 mL dry DCM. DCC (77 mg, 0.38 mmol, 3.0 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 8/1, 6/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the

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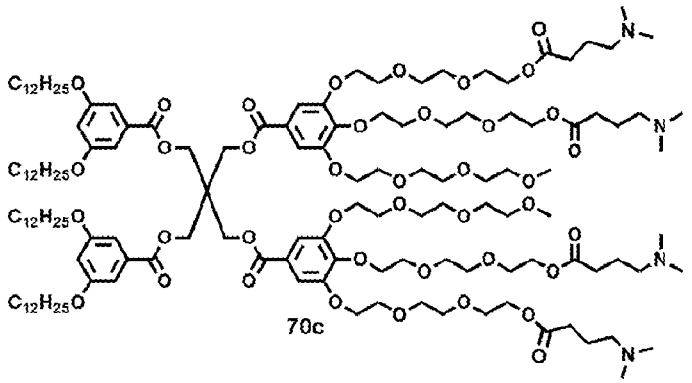
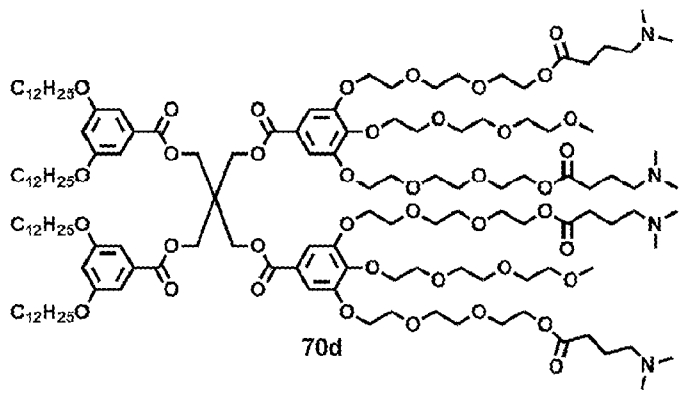
solvent yielded the title compound as a light-yellow oil (0.26 g, 84%).



Compound 70c ($6/4_{DMBA}^{1,1',2,2'}$). Compound **69c** (0.28 g, 0.13 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (0.11 g, 0.65 mmol, 5.0 equiv) were dissolved in 6 mL dry DCM. DCC (0.16 g, 0.78 mmol, 6.0 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 6/1, 3/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.29 g, 83%).



Compound 70d ($6/4_{DMBA}^{1,1',3,3'}$). Compound **69d** (0.18 g, 0.08 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (69 mg, 0.41 mmol, 5.0 equiv) were dissolved in 5 mL dry DCM. DCC (0.10 g, 0.49 mmol, 6.0 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 6/1, 3/1 with 0.1% TEA as the eluent. Then the crude

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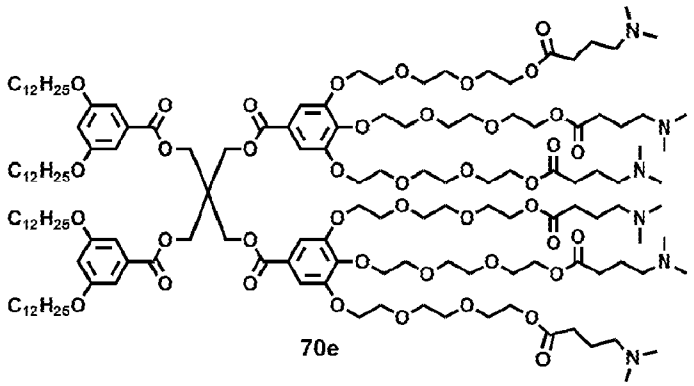
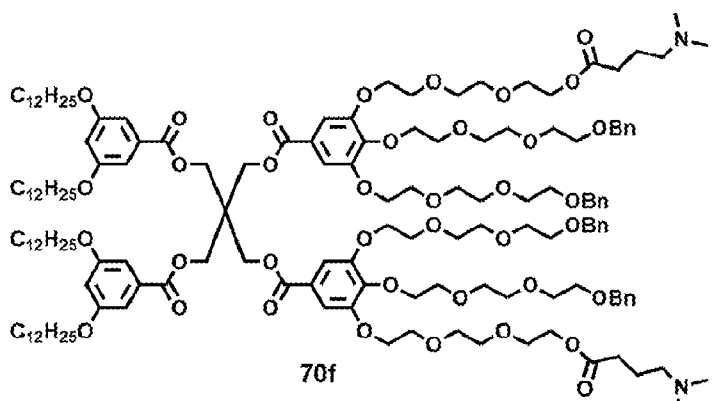
product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.19 g, 88%).



70e

Compound 70e (6/6$_{DMBA}^{1,1',2,2',3,3'}$). Compound **69e** (95 mg, 0.04 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (59 mg, 0.35 mmol, 8.0 equiv) were dissolved in 4 mL dry DCM. DCC (90 mg, 0.44 mmol, 10.0 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH – 6/1, 3/1, 1/1 with 0.2% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 20 mL) and the mixture was extracted by DCM (20 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.11 g, 83%).



70f

Compound 70f (6/2$_{DMBA}^{1,1'}$$_{Bn}^{2,2',3,3'}$). Compound **69f** (0.15 g, 0.06 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (22 mg, 0.13 mmol, 2.2 equiv) were dissolved in 4 mL dry DCM. DCC (37 mg, 0.18 mmol, 3.0 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the

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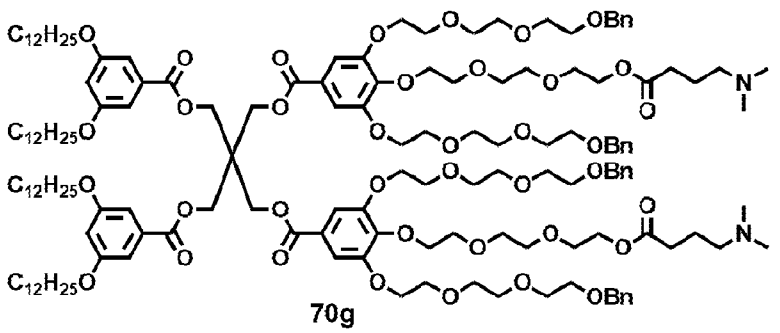
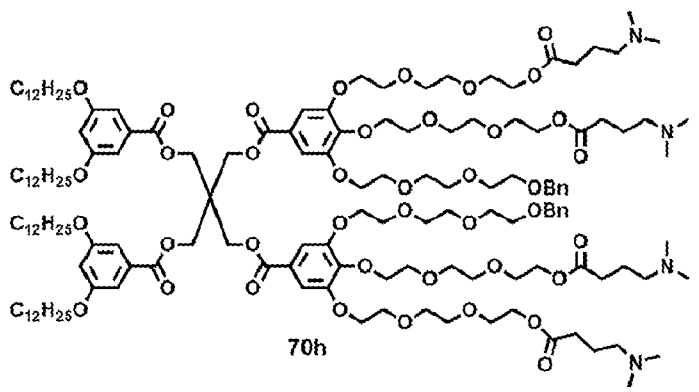
urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 6/1 as the eluent to afford the title compound as a light-yellow oil (0.14 g, 88%).



70g

Compound 70g ($6/2_{DMBA}^{2,2'}{}_{Bn}^{1,1',3,3'}$). Compound **69g** (95 mg, 0.04 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (15 mg, 0.09 mmol, 2.2 equiv) were dissolved in 4 mL dry DCM. DCC (24 mg, 0.12 mmol, 3.0 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 10/1 as the eluent to afford the title compound as a colorless oil (92 mg, 86%).



70h

Compound 70h ($6/4_{DMBA}^{1,1',2,2'}{}_{Bn}^{3,3'}$). Compound **69h** (0.31 g, 0.13 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (97 mg, 0.58 mmol, 4.4 equiv) were dissolved in 6 mL dry DCM. DCC (0.16 g, 0.79 mmol, 6.0 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 6/1, 5/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the

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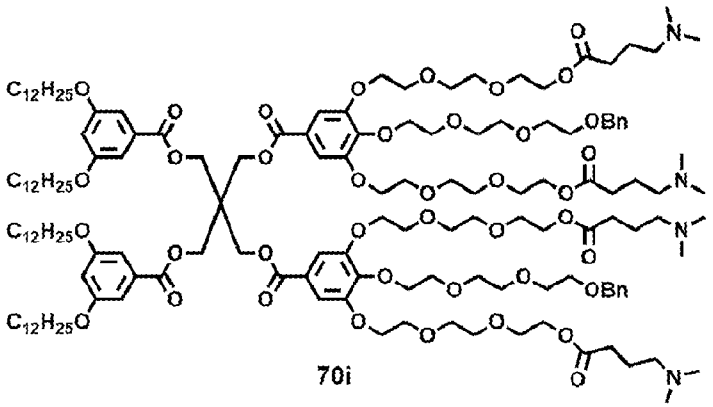
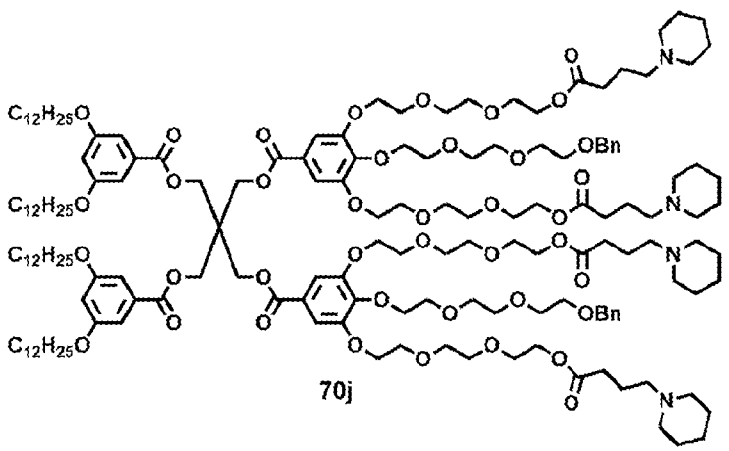
solvent yielded the title compound as a light-yellow oil (0.35 g, 95%).



70i

Compound 70i (6/4$_{DMBA}^{1,1',3,3'}$$_{Bn}^{2,2'}$). Compound **69i** (0.34 g, 0.14 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (0.11 g, 0.63 mmol, 4.4 equiv) were dissolved in 6 mL dry DCM. DCC (0.18 g, 0.86 mmol, 6.0 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 6/1, 5/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.37 g, 89%).



70j

Compound 70j (6/4$_{PIP}^{1,1',3,3'}$$_{Bn}^{2,2'}$). Compound **70j** was synthesized from compound **69i** (0.15 g, 0.064 mmol), compound **56a** (58 mg, 0.28 mmol) and DCC (79 mg, 0.38 mmol) following a procedure similar to that used for the synthesis of compound **70i**. The title compound was obtained as a light-yellow oil (0.17 g, 90%).

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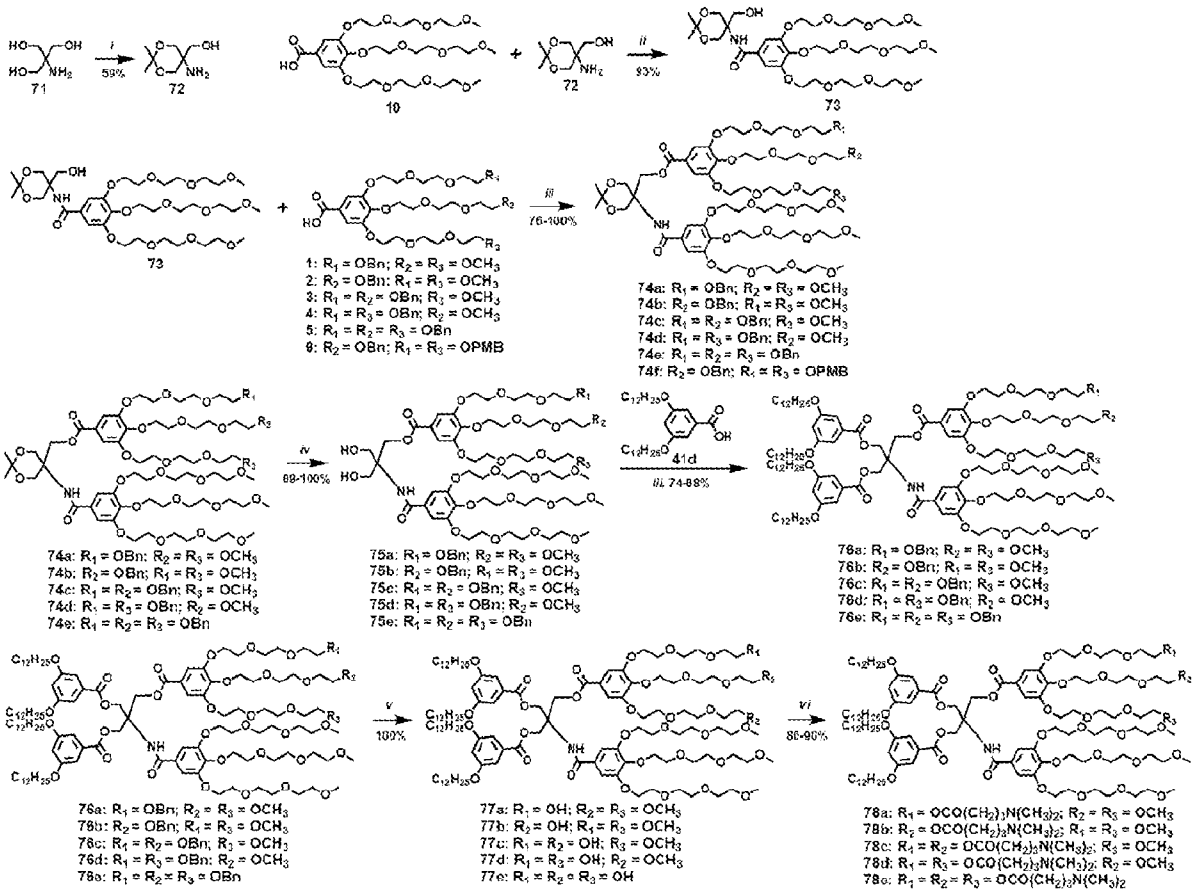
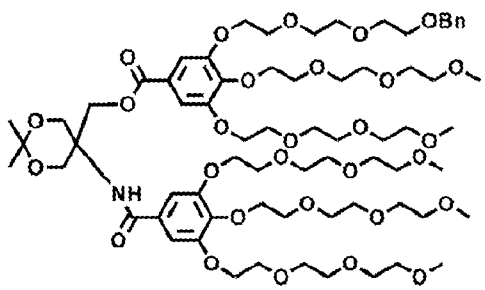
3.7 Synthesis of Hybrid Twin-Mix IAJDs 38-42 of Library 6



Reagents and conditions: (*i*) PTSA·H_2O, 2,2-dimethoxypropane, Acetone, 23 °C, 12 h; (*ii*) CDMT, NMM, THF, 0–23 °C, 12 h; (*iii*) DCC, DPTS, DCM, 23 °C, 12 h; (*iv*) 2 M HCl, MeOH, 23 °C, 2 h; (*v*) H_2, Pd/C, DCM/MeOH, 23 °C, 12 h; (*vi*) 4-(Dimethylamino)butyric acid hydrochloride, DCC, DCM, 23 °C, 12 h.

Compounds **72** and **73** were synthesized according to literature procedures (Zhang, et al., J. Am. Chem. Soc. 2015, 137, 13334–13344; Xiao, et al., Proc. Natl. Acad. Sci. U.S.A. 2020, 117, 11931–11939).



74a

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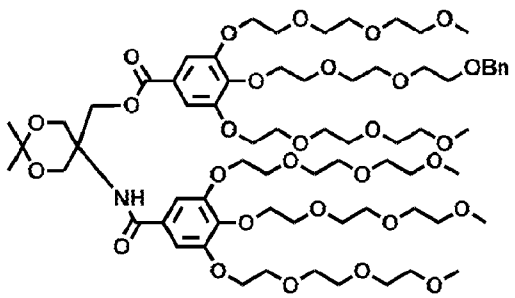
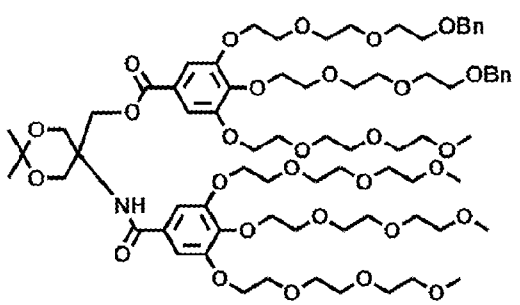
Compound **74a.** To a DCM (5 mL) solution of compound **1** (230 mg, 0.33 mmol), compound **73** (250 mg, 0.33 mmol) and DPTS (100 mg, 0.33 mmol), was added DCC (140 mg, 0.66 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless oil (420 mg, 89%).



74b

Compound **74b.** To a DCM (10 mL) solution of compound **2** (229 mg, 0.304 mmol), compound **73** (229 mg, 0.334 mmol) and DPTS (134 mg, 0.456 mmol), was added DCC (188 mg, 0.912 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless oil (430 mg, 100%).



74c

Compound **74c.** To a DCM (5 mL) solution of compound **3** (250 mg, 0.334 mmol), compound **73** (250 mg, 0.334 mmol) and DPTS (102 mg, 0.334 mmol), was added DCC (142 mg, 0.668 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 60/1 to 20/1 to yield the title compound as a colorless oil (376 mg, 76%).

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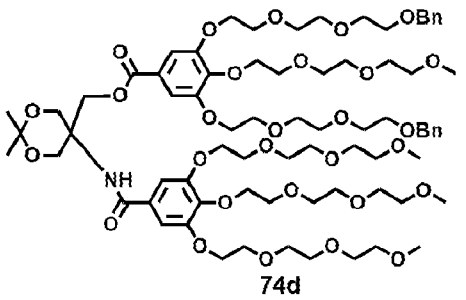
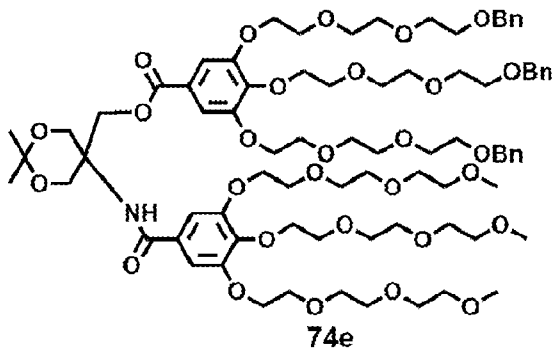


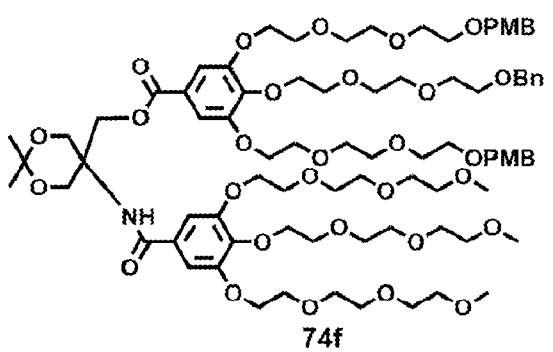
74d

Compound **74d.** To a DCM (5 mL) solution of compound **4** (0.25 mg, 0.33 mmol, 1 equiv), compound **73** (0.25 g, 0.33 mmol, 1 equiv) and DPTS (0.10 g, 0.33 mmol, 1 equiv), was added DCC (0.14 g, 0.66 mmol, 2 equiv). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 30/1 to yield the title compound as a colorless oil (0.43 g, 88%).



74e

Compound **74e.** To a DCM (5 mL) solution of compound **5** (280 mg, 0.330 mmol), compound **73** (250 mg, 0.330 mmol) and DPTS (103 mg, 0.330 mmol), was added DCC (140 mg, 0.660 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless oil (440 mg, 85%).



74f

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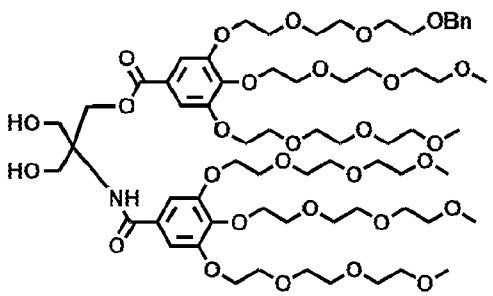
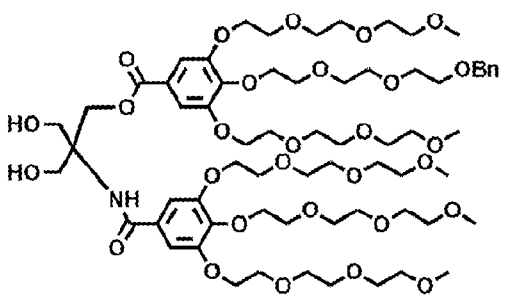
Compound **74f.** To a DCM (10 mL) solution of compound **9** (558 mg, 0.62 mmol), compound **73** (425 mg, 0.57 mmol) and DPTS (252 mg, 0.86 mmol), was added DCC (352 mg, 1.71 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless oil (814 mg, 88%).



75a

Compound **75a.** To a MeOH solution (10 mL) of compound **74a** (400 mg, 0.63 mmol) was added 3 mL of HCl aqueous solution (2 M). The mixture was allowed to stir at 23 °C for 2h and saturated $NaHCO_3$ solution (20 mL) was added. The mixture was extracted with DCM (50 mL) for 3 times. An organic extract was dried over $MgSO_4$ and evaporated to dryness under reduced pressure to yield the title compound as a colorless viscous liquid without further purification (366 mg, 95%).



75b

Compound **75b.** Compound **75b** was synthesized from compound **74b** (430 mg, 0.334 mmol) following a procedure similar to that used for the synthesis of compound **75a**. The title compound was obtained as a colorless viscous liquid without further purification (410 mg, 89%).

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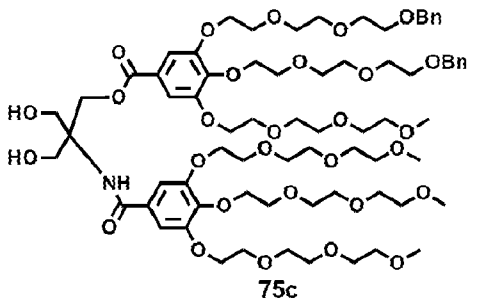
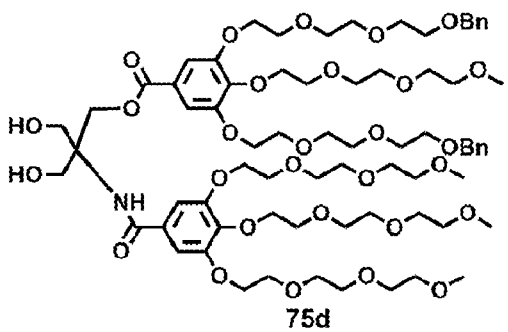


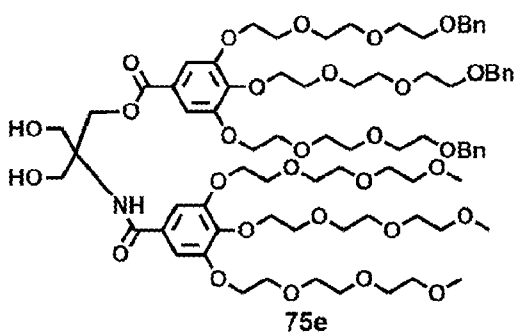
75c

Compound **75c.** Compound **75c** was synthesized from compound **74c** (350 mg, 0.234 mmol) following a procedure similar to that used for the synthesis of compound **75a**. The title compound was obtained as a colorless viscous liquid without further purification (310 mg, 91



75d

Compound **75d.** Compound **75d** was synthesized from compound **74d** (0.36 g, 0.24 mmol) following a procedure similar to that used for the synthesis of compound **75a**. The title compound was obtained as a a colorless oil without further purification (0.35 g, 100%).



75e

Compound **75e.** Compound **75e** was synthesized from compound **74e** (330 mg, 0.210 mmol) following a procedure similar to that used for the synthesis of compound **75a**. The title compound was obtained as a colorless viscous liquid without further purification (320 mg, 100%).

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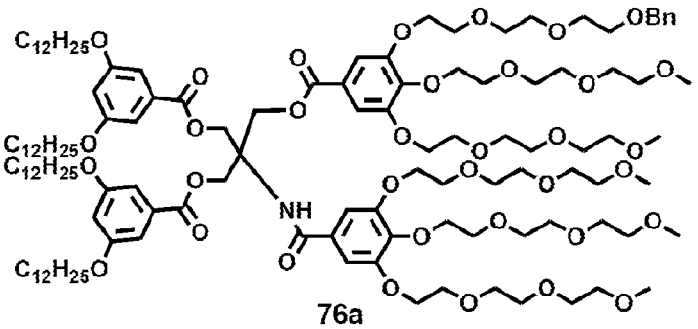
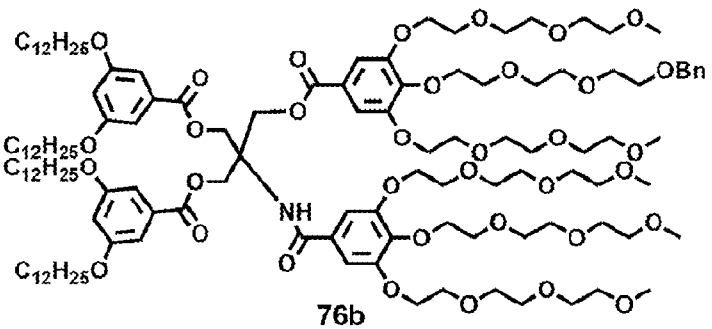


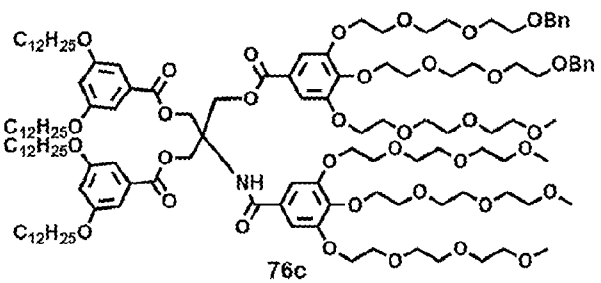
76a

Compound **76a.** To a DCM (5 mL) solution of compound **41d** (307 mg, 0.624 mmol), compound **75a** (410 mg, 0.297 mmol) and DPTS (219 mg, 0.742 mmol), was added DCC (306 mg, 1.49 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless oil (586 mg, 85%).



76b

Compound **76b.** To a DCM (10 mL) solution of compound **41d** (307 mg, 0.624 mmol), compound **75b** (410 mg, 0.297 mmol) and DPTS (219 mg, 0.742 mmol), was added DCC (306 mg, 1.49 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless oil (607 mg, 88%).



76c

Compound **76c.** To a DCM (5 mL) solution of compound **41d** (200 mg, 0.400 mmol),

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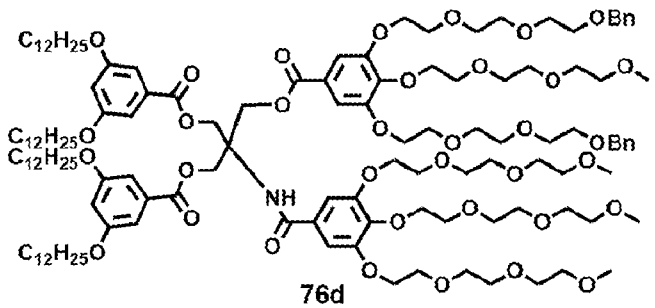
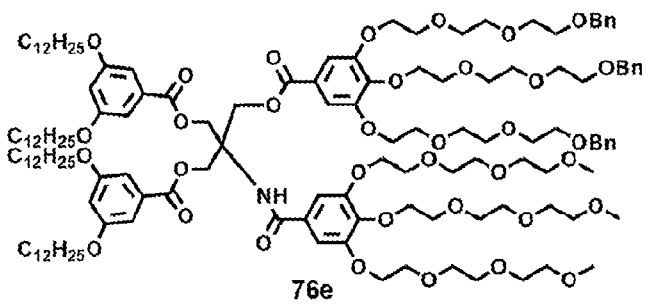
compound **75c** (280 mg, 0.197 mmol) and DPTS (119 mg, 0.380 mmol), was added DCC (118 mg, 0.571 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless oil (375 mg, 83%).



76d

Compound **76d.** To a DCM (10 mL) solution of compound **41d** (0.19 g, 0.38 mmol, 2.1 equiv), compound **75d** (0.26 g, 0.18 mmol, 1 equiv) and DPTS (0.11 g, 0.36 mmol, 2 equiv), was added DCC (0.11 g, 0.54 mmol, 3 equiv). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 30/1 to yield the title compound as a colorless oil (0.32 g, 74%).



76e

Compound **76e.** To a DCM (10 mL) solution of compound **41d** (220 mg, 0.44 mmol), compound **75e** (320 mg, 0.21 mmol) and DPTS (130 mg, 0.42 mmol), was added DCC (130 mg, 0.63 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 30/1 to yield the title compound as a colorless oil (440 mg, 85%).

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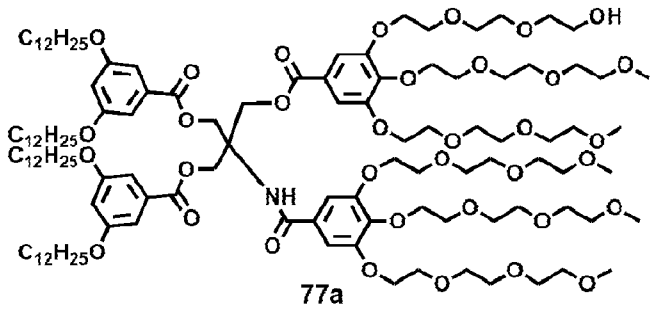
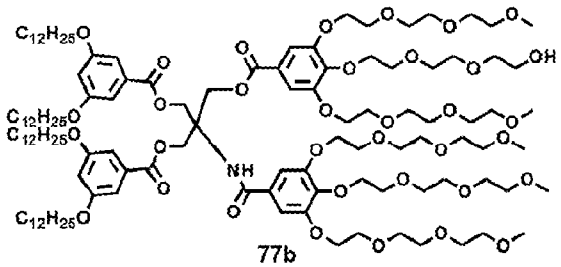


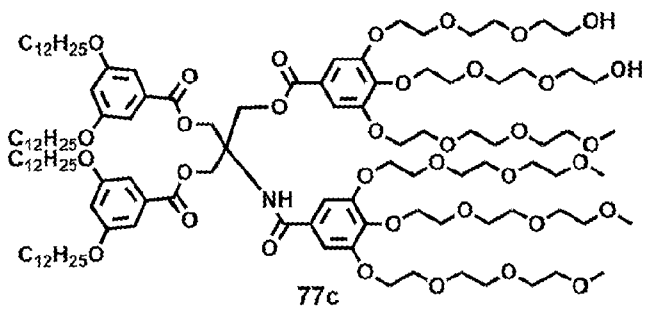
77a

Compound 77**a.** To a mixed DCM (16 mL) and MeOH (8 mL) solution of compound **76a** (250 mg, 0.112 mmol) was added Pd/C (25 mg, 10 wt%). The mixture was bubbling with H_2 for 30 min and then allowed to stir at 23 °C for 12 h under H_2 atmosphere. The mixture was then filtered through Celite®. The filtrate was concentrated to dryness and without further purification to yield the title compound as a colorless oil (240 mg, 100%).



77b

Compound 77**b.** Compound 77**b** was synthesized from compound **76b** (607 mg, 0.261 mmol) following a procedure similar to that used for the synthesis of compound 77**a**. The title compound was obtained as a colorless oil (582 mg, 100%).



77c

Compound 77**c.** Compound 77**c** was synthesized from compound **76c** (360 mg, 0.150 mmol) following a procedure similar to that used for the synthesis of compound 77**a**. The title compound was obtained as a colorless oil (330 mg, 100%).

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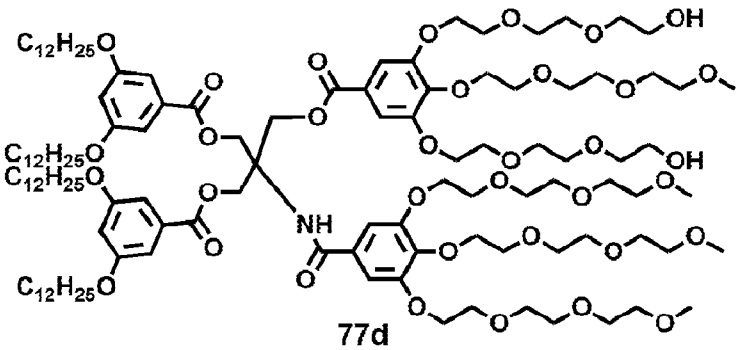
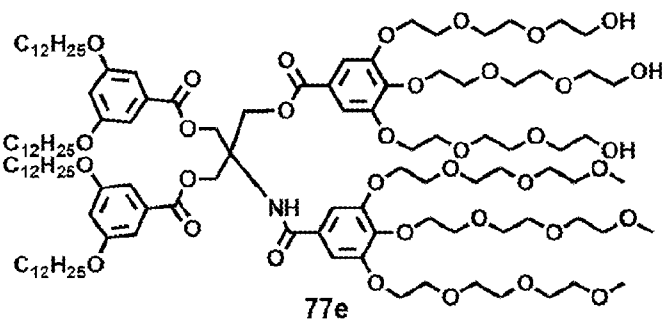


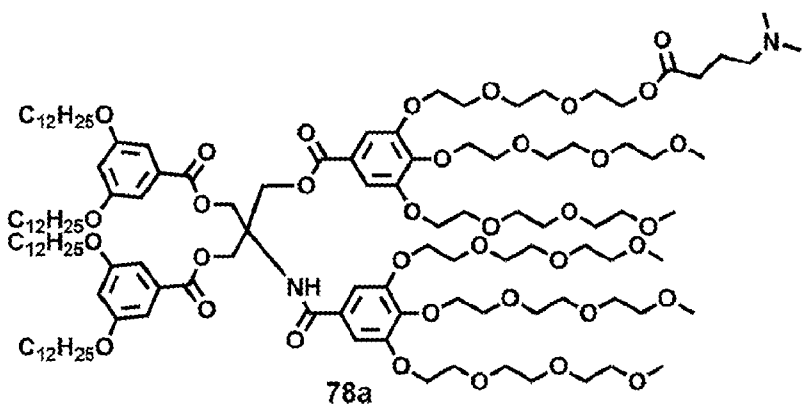
77d

Compound 77**d.** Compound 77**d** was synthesized from compound **76d** (0.23 g, 0.096 mmol) following a procedure similar to that used for the synthesis of compound **77a**. The title compound was obtained as a colorless oil (0.21 g, 100%).



77e

Compound **77e.** Compound **77e** was synthesized from compound **76e** (300 mg, 0.121 mmol) following a procedure similar to that used for the synthesis of compound **77a**. The title compound was obtained as a colorless oil (266 mg, 100%).



78a

Compound 78a (6/1$_{DMBA}$[1]). Compound **77a** (560 mg, 0.25 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (50 mg, 0.30 mmol, 1.2 equiv) were dissolved in 6 mL dry DCM. DCC (155 mg, 0.75 mmol, 3 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the

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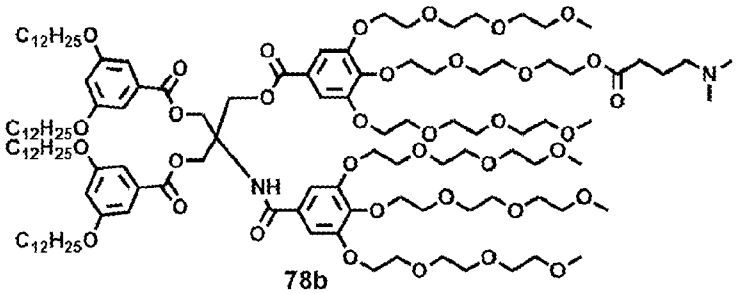
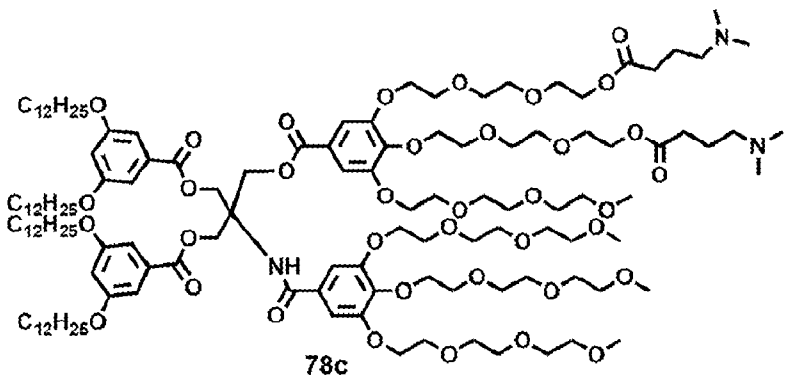
urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 10/1, 6/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (504 mg, 86%).



78b

Compound 78b (6/1$_{DMBA}^{2}$). Compound **78b** was synthesized from compound **77b** (311 mg, 0.139 mmol), 4-(dimethylamino)butyric acid hydrochloride (28 mg, 0.167 mmol) and DCC (57 mg, 0.278 mmol) following a procedure similar to that used for the synthesis of compound **78a**. The title compound was obtained as a light-yellow oil (290 mg, 90%).



78c

Compound 78c (6/2$_{DMBA}^{1,2}$). Compound **78c** was synthesized from compound **77c** (320 mg, 0.144 mmol), 4-(dimethylamino)butyric acid hydrochloride (70 mg, 0.432 mmol) and DCC (119 mg, 0.576 mmol) following a procedure similar to that used for the synthesis of compound **78a**. The title compound was obtained as a light-yellow oil (316 mg, 90%).

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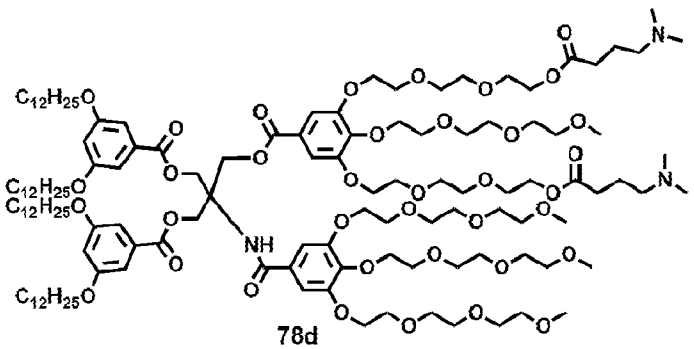
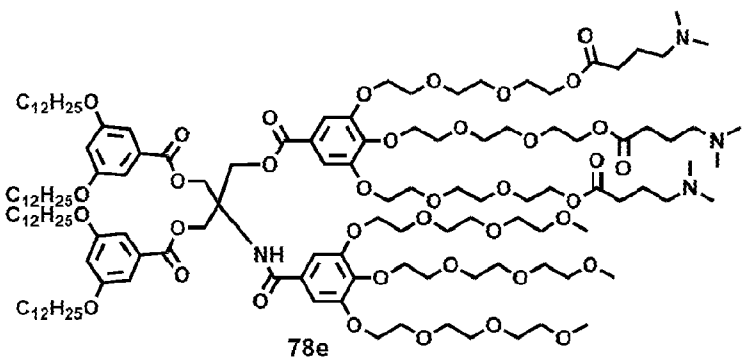


78d

Compound 78d (6/2$_{DMBA}^{1,3}$). Compound **78d** was synthesized from compound **77d** (0.17 g, 0.077 mmol), 4-(dimethylamino)butyric acid hydrochloride (28 mg, 0.170 mmol) and DCC (48 mg, 0.231 mmol) following a procedure similar to that used for the synthesis of compound **78a**. The title compound was obtained as a light-yellow oil (0.17 g, 90%).



78e

Compound 78e (6/3$_{DMBA}^{1,2,3}$). Compound **78e** was synthesized from compound **77e** (260 mg, 0.116 mmol), 4-(dimethylamino)butyric acid hydrochloride (78 mg, 0.467 mmol) and DCC (95 mg, 0.467 mmol) following a procedure similar to that used for the synthesis of compound **78a**. The title compound was obtained as a light-yellow oil (290 mg, 90%).

3.8 Synthesis of Hybrid Twin-Mix IAJDs 26-29, 47, 49-52 in Library 6

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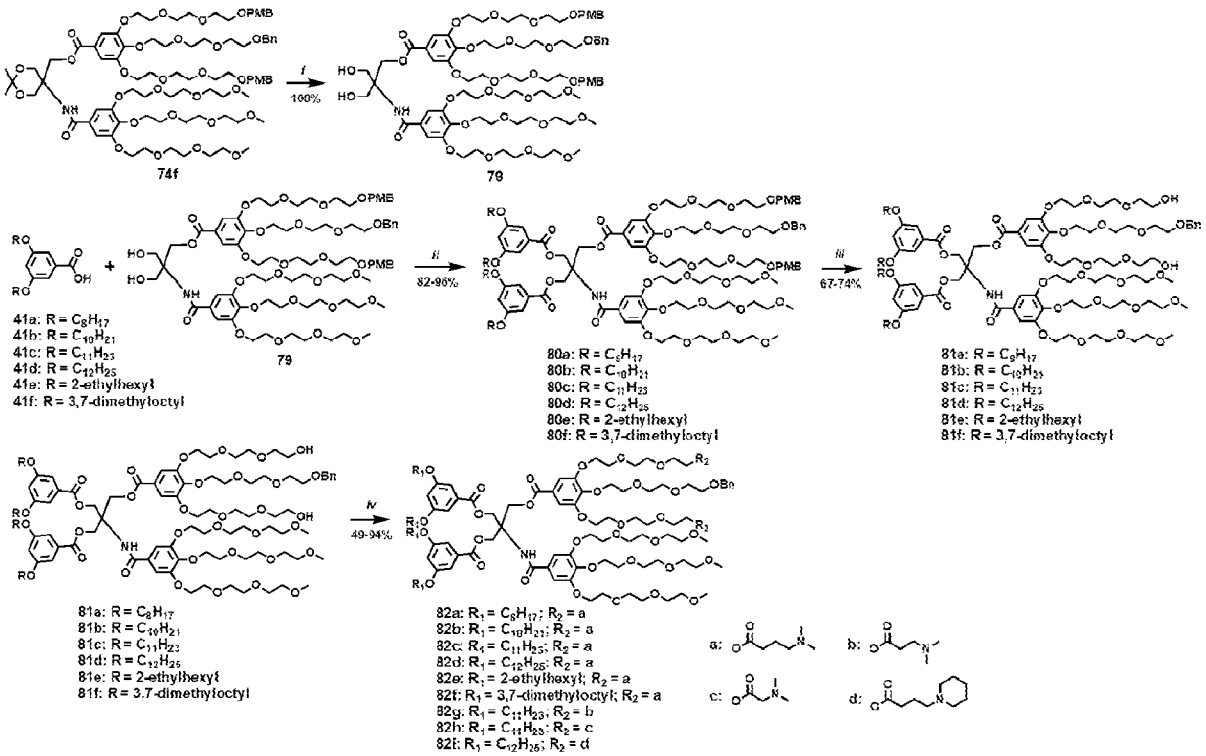
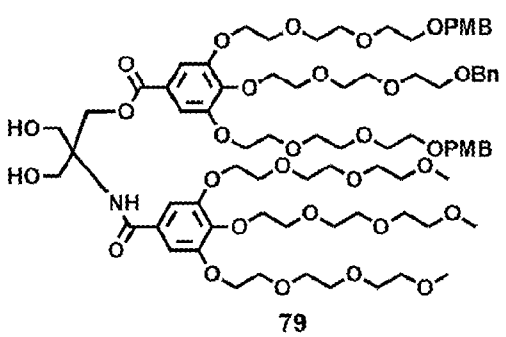


Reagents and conditions: (*i*) 2 M HCl, MeOH, 23 °C, 2 h; (*ii*) DCC, DPTS, DCM, 23 °C, 12 h; (*iii*) DDQ, DCM/H_2O, 23 °C, 1 h; (*iv*) 4-(Dimethylamino)butyric acid hydrochloride, 3-(dimethylamino)propionic acid hydrochloride, N,N-dimethylglycine hydrochloride/4-(piperidin-1-yl)butanoic acid hydrochloride, DCC, DCM, 23 °C, 12 h.



Compound 79. To a MeOH solution (10 mL) of compound **74f** (800 mg, 0.489 mmol) was added 3 mL of HCl aqueous solution (2 M). The mixture was allowed to stir at 23 °C for 2 h and saturated $NaHCO_3$ solution (20 mL) was added. The mixture was extracted with DCM (50 mL) for 3 times. An organic extract was dried over anhydrous $MgSO_4$ and evaporated to dryness under reduced pressure to yield the title compound as a colorless viscous liquid without further purification (778 mg, 100%).

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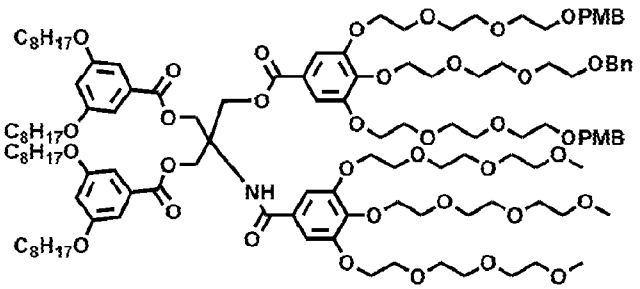
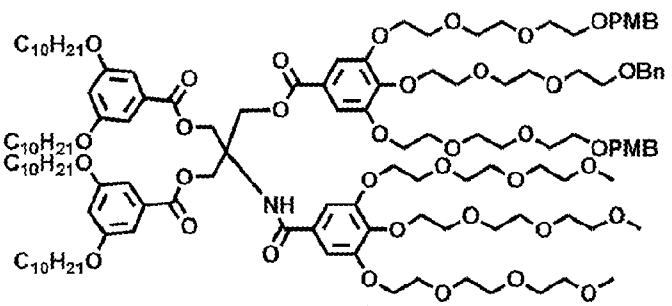


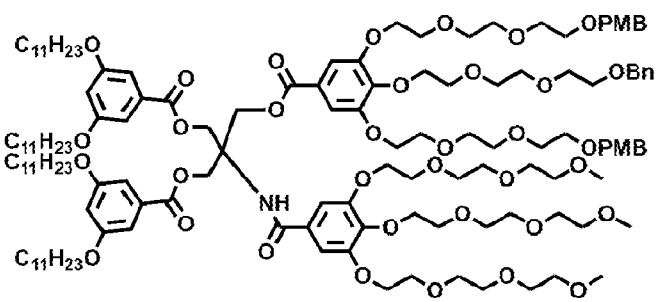
80a

Compound 80a. To a DCM (10 mL) solution of compound **41a** (79 mg, 0.21 mmol), compound **79** (160 mg, 0.10 mmol) and DPTS (62 mg, 0.20 mmol), was added DCC (62 mg, 0.30 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless oil (190 mg, 83%).



80b

Compound 80b. Compound **80b** was synthesized from compound **41b** (174 mg, 0.401 mmol), compound **79** (290 mg, 0.182 mmol), DPTS (134 mg, 0.455 mmol) and DCC (188 mg, 0.910 mmol) following a procedure similar to that used for the synthesis of compound **80a**. The title compound was obtained as a colorless oil (423 mg, 96%).



80c

Compound 80c. Compound **80c** was synthesized from compound **41c** (150 mg, 0.345 mmol), compound **79** (249 mg, 0.157 mmol), DPTS (116 mg, 0.392 mmol) and DCC (161 mg, 0.785 mmol) following a procedure similar to that used for the synthesis of compound **80a**. The

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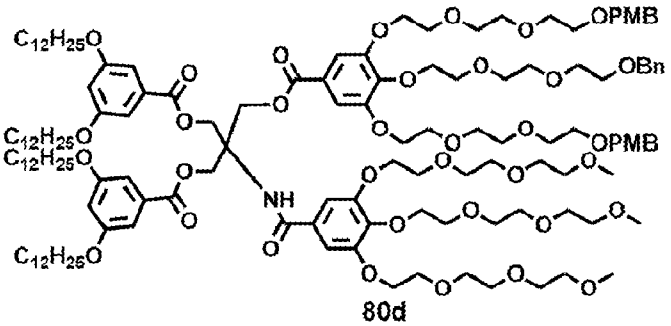
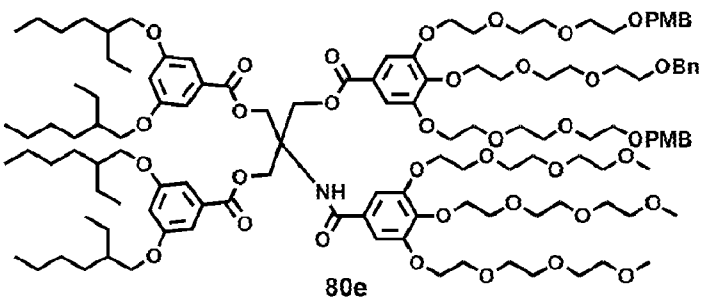
title compound was obtained as a colorless oil (332 mg, 87%).



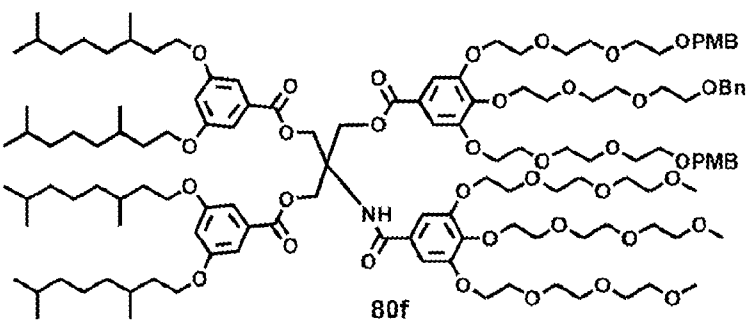
80d

Compound 80d. Compound **80d** was synthesized from compound **41d** (170 mg, 0.347 mmol), compound **79** (251 mg, 0.158 mmol), DPTS (116 mg, 0.371 mmol) and DCC (163 mg, 0.789 mmol) following a procedure similar to that used for the synthesis of compound **80a**. The title compound was obtained as a colorless oil (355 mg, 89%).



80e

Compound 80e. Compound **80e** was synthesized from compound **41e** (79 mg, 0.21 mmol), compound **79** (160 mg, 0.10 mmol), DPTS (62 mg, 0.20 mmol) and DCC (62 mg, 0.30 mmol) following a procedure similar to that used for the synthesis of compound **80a**. The title compound was obtained as a colorless oil (194 mg, 84%).



80f

Compound 80f. Compound **80f** was synthesized from compound **41f** (150 mg, 0.345 mmol), compound **79** (249 mg, 0.157 mmol), DPTS (116 mg, 0.392 mmol) and DCC (161 mg, 0.785 mmol) following a procedure similar to that used for the synthesis of compound **80a**. The title compound was obtained as a colorless oil (314 mg, 82%).

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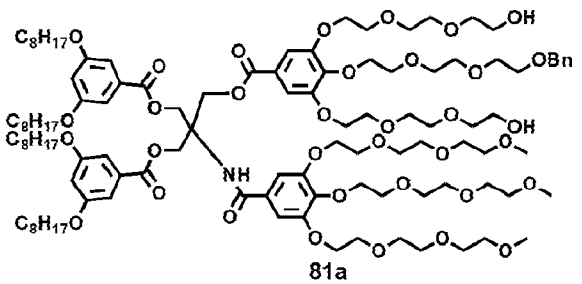
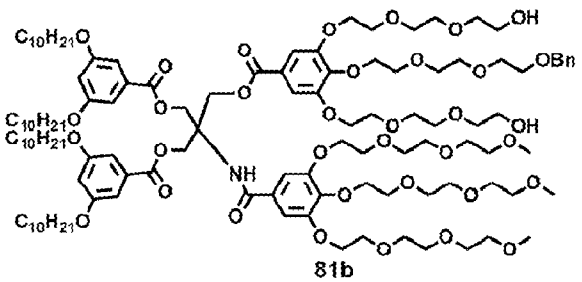


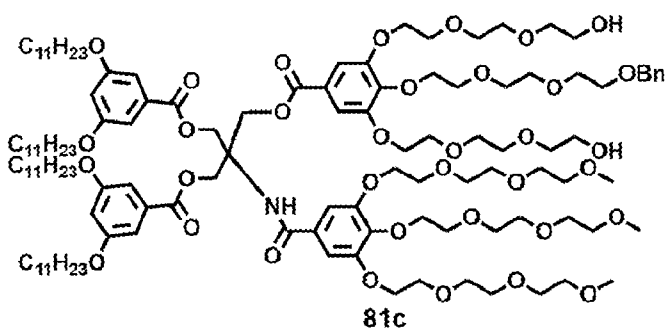
81a

Compound 81a. To a mixed solution of DCM (5 mL) and water (0.25 mL) of compound **80a** (180 mg, 0.078 mmol), was added DDQ (44 mg, 0.195 mmol). The mixture was allowed to stir at 23 °C for 1 h. The precipitates were filtered out and DCM (20 mL) was added. The mixture was washed by $NaHCO_3$ aqueous solution (saturated), $NaHSO_3$ aqueous solution (2%) and $NaHCO_3$ aqueous solution (saturated) successively. The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography with a mobile phase of EtOAc/MeOH = 10/1 to yield the title compound as a light-yellow oil (112 mg, 67%).



81b

Compound 81b. Compound **81b** was synthesized from compound **80b** (420 mg, 0.173 mmol) following a procedure similar to that used for the synthesis of compound **81a**. The title compound was obtained as a colorless oil (278 mg, 73%).



81c

Compound 81c. Compound **81c** was synthesized from compound **80c** (350 mg, 0.141 mmol) following a procedure similar to that used for the synthesis of compound **81a**. The title compound was obtained as a colorless oil (229 mg, 73%).

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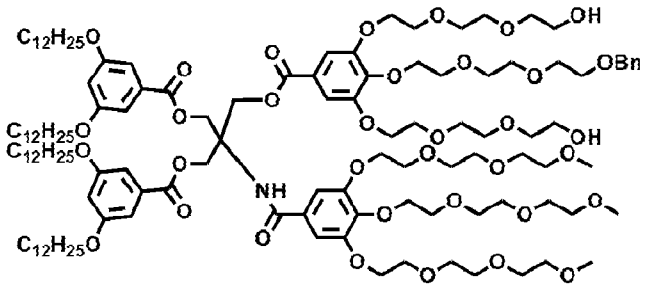
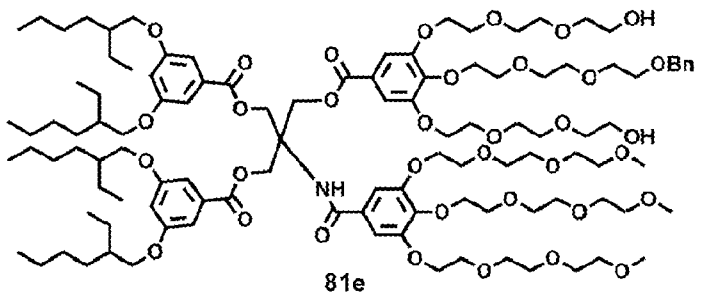


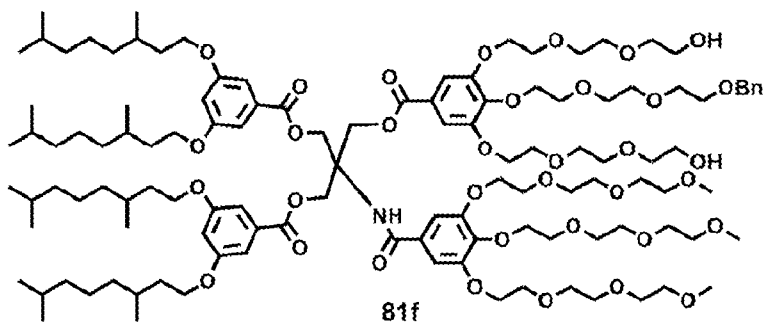
81d

Compound 81d. Compound **81d** was synthesized from compound **80d** (348 mg, 0.137 mmol) following a procedure similar to that used for the synthesis of compound **81a**. The title compound was obtained as a colorless oil (249 mg, 72%).



81e

Compound 81e. Compound **81e** was synthesized from compound **80e** (190 mg, 0.082 mmol) following a procedure similar to that used for the synthesis of compound **81a**. The title compound was obtained as a light-yellow oil (130 mg, 74%).



81f

Compound 81f. Compound **81f** was synthesized from compound **80f** (330 mg, 0.136 mmol) following a procedure similar to that used for the synthesis of compound **81a**. The title compound was obtained as a colorless oil (215 mg, 72%).

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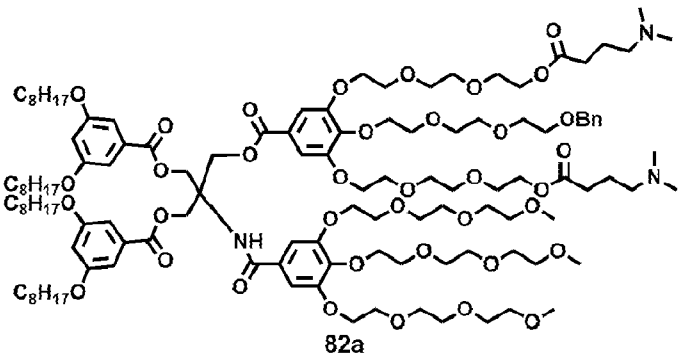
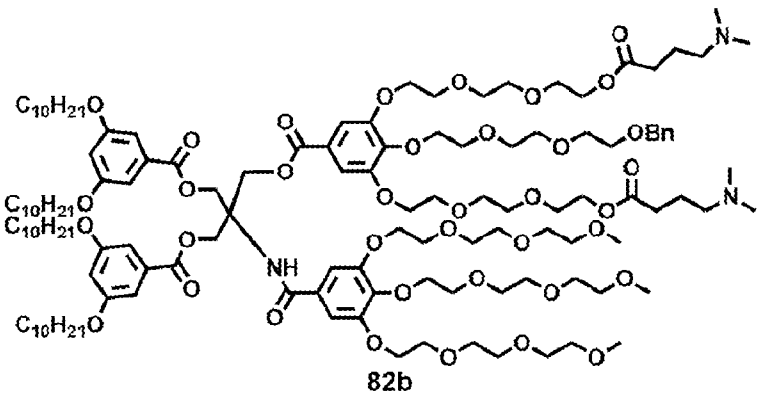


82a

Compound 82a (6/2$_{DMBA}^{1,3}$$_{Bn}^{2}$-C8). Compound **81a** (110 mg, 0.053 mmol) and 4-(dimethylamino)butyric acid hydrochloride (20 mg, 0.117 mmol) were dissolved in 5 mL dry DCM. DCC (33 mg, 0.159 mmol) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 10/1, 6/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (110 mg, 92%).



82b

Compound 82b (6/2$_{DMBA}^{1,3}$$_{Bn}^{2}$-C10). Compound **82b** was synthesized from compound **81b** (270 mg, 0.124 mmol), 4-(dimethylamino)butyric acid hydrochloride (52 mg, 0.310 mmol) and DCC (128 mg, 0.620 mmol) following a procedure similar to that used for the synthesis of compound **82a**. The title compound was obtained as a light-yellow oil (280 mg, 94%).

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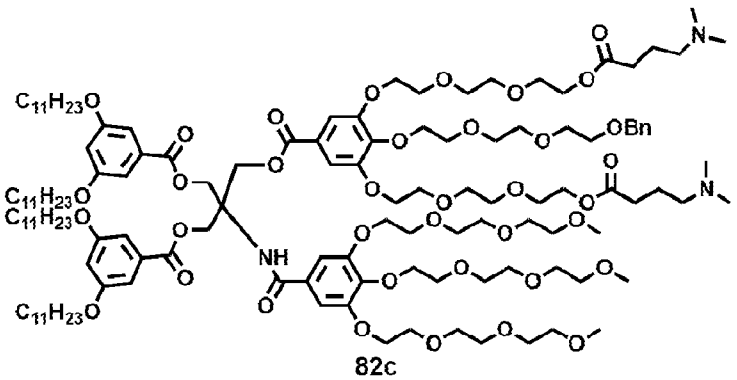
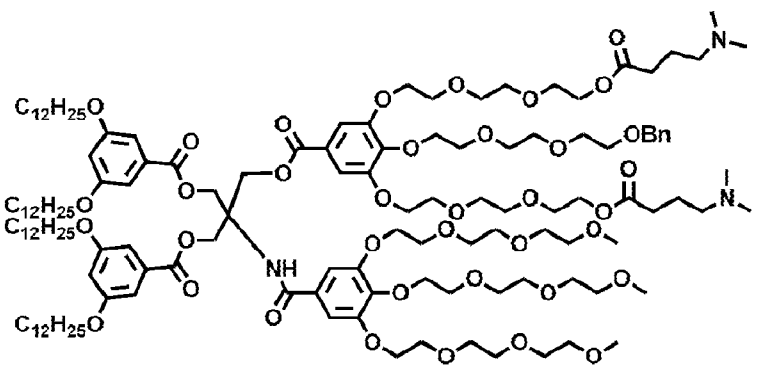


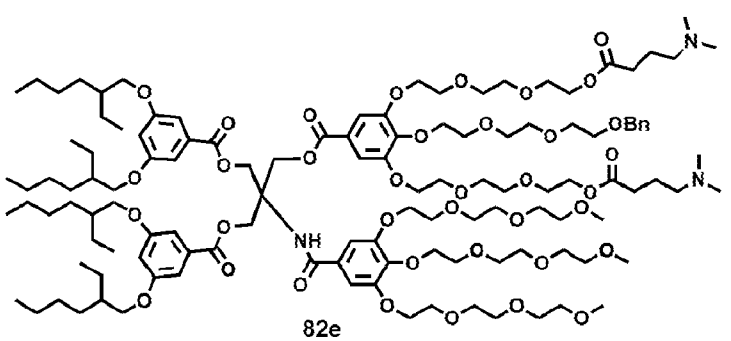
82c

Compound 82c (6/2$_{DMBA}^{1,3}$$_{Bn}^{2}$-C11). Compound **82c** was synthesized from compound **81c** (229 mg, 0.102 mmol), 4-(dimethylamino)butyric acid hydrochloride (42 mg, 0.256 mmol) and DCC (105 mg, 0.510 mmol) following a procedure similar to that used for the synthesis of compound **82a**. The title compound was obtained as a light-yellow oil (227 mg, 90%).



82d

Compound 82d (6/2$_{DMBA}^{1,3}$$_{Bn}^{2}$). Compound **82d** was synthesized from compound **81d** (249 mg, 0.108 mmol), 4-(dimethylamino)butyric acid hydrochloride (46 mg, 0.271 mmol) and DCC (89 mg, 0.432 mmol) following a procedure similar to that used for the synthesis of compound **82a**. The title compound was obtained as a light-yellow oil (245 mg, 90%).



82e

Compound 82e (6/2$_{DMBA}^{1,3}$$_{Bn}^{2}$-EH). Compound **82e** was synthesized from compound **81e** (100 mg, 0.048 mmol), 4-(dimethylamino)butyric acid hydrochloride (18 mg, 0.105 mmol) and

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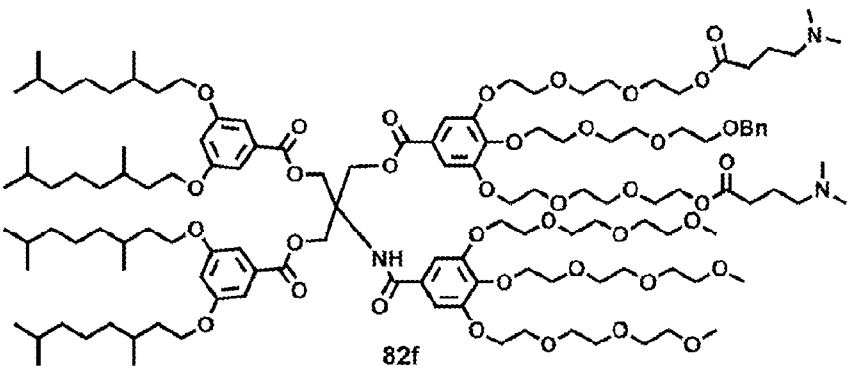
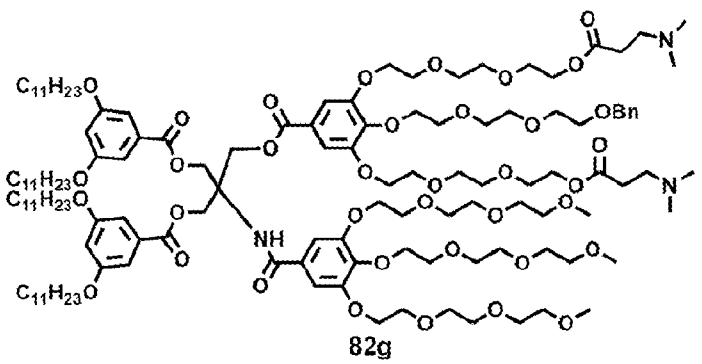
DCC (30 mg, 0.143 mmol) following a procedure similar to that used for the synthesis of compound **82a**. The title compound was obtained as a light-yellow oil (100 mg, 91%).



82f

Compound 82f ($6/2_{DMBA}^{1,3}{}_{Bn}^{2}$-dm8). Compound **82f** was synthesized from compound **81f** (215 mg, 0.098 mmol), 4-(dimethylamino)butyric acid hydrochloride (41 mg, 0.246 mmol) and DCC (88 mg, 0.392 mmol) following a procedure similar to that used for the synthesis of compound **82a**. The title compound was obtained as a light-yellow oil (203 mg, 86%).



82g

Compound 82g ($6/2_{DMPA}^{1,3}{}_{Bn}^{2}$-C11). Compound **82g** was synthesized from compound **81c** (110 mg, 0.049 mmol), 3-(dimethylamino)propionic acid hydrochloride (17 mg, 0.108 mmol) and DCC (30 mg, 0.147 mmol) following a procedure similar to that used for the synthesis of compound **82a**. The title compound was obtained as a light-yellow oil (72 mg, 61%).

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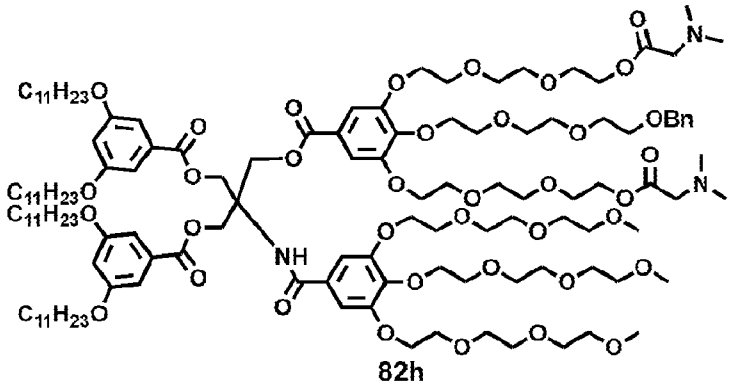
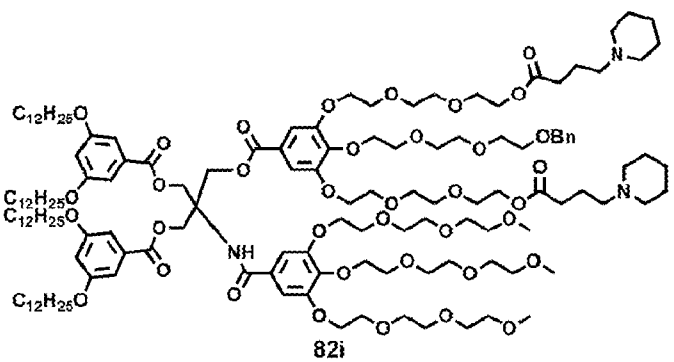


82h

Compound 82h (6/2$_{DMA}^{1,3}$$_{Bn}^{2}$-C11). Compound **81c** (110 mg, 0.049 mmol) and N,N-dimethylglycine hydrochloride (15 mg, 0.108 mmol) were dissolved in 5 mL dry DCM. DCC (30 mg, 0.147 mmol) was added in one portion into the above mixture. Then 3 drops of TEA were added to the mixture. The reaction mixture was stirred at 23 °C for 12 h and more N,N-dimethylglycine hydrochloride (7.5 mg, 0.054 mmol) was added. The reaction mixture was stirred at 23 °C for another 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 30/1, 20/1 and 10/1 as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (58 mg, 49%).



82i

Compound 82i (6/2$_{PIP}^{1,3}$$_{Bn}^{2}$). Compound **82i** was synthesized from compound **81d** (195 mg, 0.085 mmol), compound **56a** (32 mg, 0.187 mmol) and DCC (88 mg, 0.424 mmol) following a procedure similar to that used for the synthesis of compound **82a**. The title compound was obtained as a light-yellow oil (160 mg, 72%).

Synthesis of Hybrid Twin-Mix IAJDs 32, 43, 48 of Library 6

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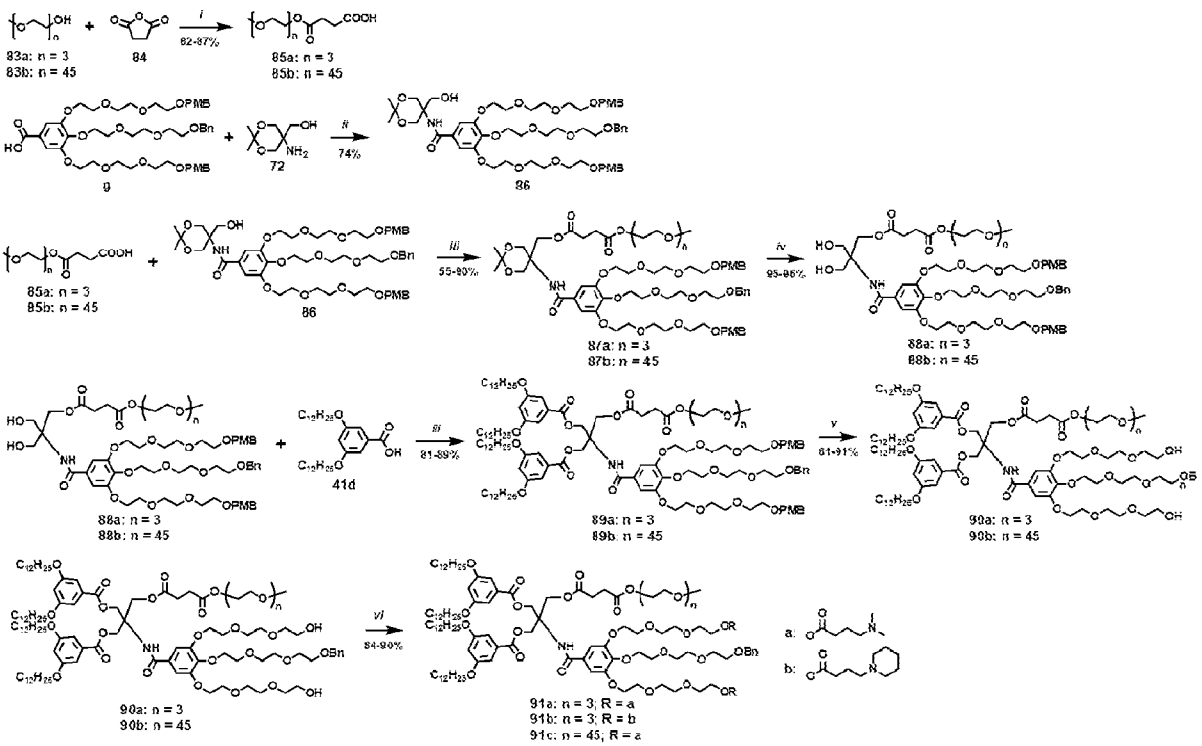
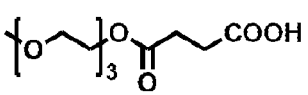
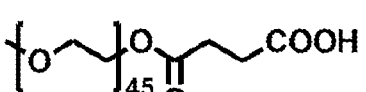


Reagents and conditions: (*i*) DMAP, DCM, 23 °C, 12 h; (*ii*) CDMT, NMM, THF, 0–23 °C, 12 h; (*iii*) DCC, DPTS, DCM, 23 °C, 12 h; (*iv*) 2 M HCl, MeOH, 23 °C, 2 h; (*v*) DDQ, DCM/H_2O, 23 °C, 1 h; (*vi*) 4-(Dimethylamino)butyric acid hydrochloride/4-(piperidin-1-yl)butanoic acid hydrochloride, DCC, DCM, 23 °C, 12 h.



85a

Compound 85a. To a 250 mL flask, DMAP (1.50 g, 12.3 mmol), succinic anhydride (7.32 g, 73.1 mmol), and triethylene glycol monomethyl ether (10.0 g, 60.9 mmol) were dissolved in 50 mL DCM and the reaction mixture was left to react for 12 h at 23 °C. The reaction mixture was diluted with 50 mL of DCM, then 25 mL of water was then added, followed by washing 3 times with aqueous 10% $NaHSO_4$ solution (10 mL). The organic phase was dried over $MgSO_4$, filtered and concentrated to obtain a colorless oil without further purification (14.08 g, 87%).



85b

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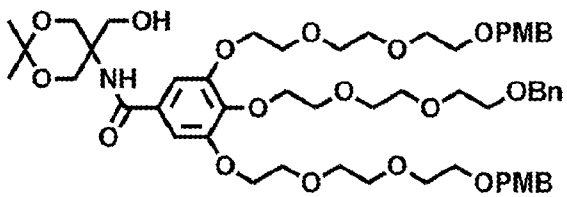
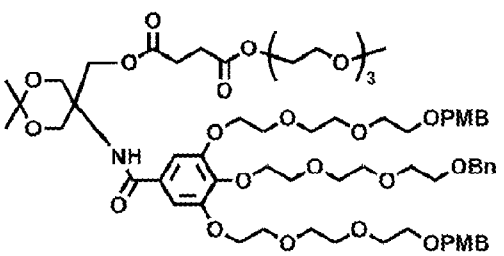
Compound 85b. To a 250 mL flask, DMAP (1.22 g, 10 mmol) and succinic anhydride (1.00 g, 10 mmol) were dissolved in 50 mL DCM. Polyethylene glycol monomethyl ether 2000 (Me-PEG-2000-OH, 2.00 g, 1.0 mmol) was slowly added to the suspension and the reaction mixture was left to react for 12 h at 23 °C. The reaction mixture was diluted with 50 mL of DCM, then 100 mL of water was then added, followed by washing 3 times with aqueous 10% $NaHSO_4$ solution (20 mL). The organic phase was dried over $MgSO_4$, filtered and concentrated to obtain a white solid without further purification (1.72 g, 82%).



86

Compound 86. To a THF (10 mL) solution of compound **9** (675 mg, 0.75 mmol) and compound **72** (136 mg, 0.84 mmol) was added CDMT (148 mg, 0.84 mmol). The mixture was cooled to 0 °C and *N*-methylmorpholine (NMM, 195 mg, 1.92 mmol) was added under N_2. The reaction mixture was allowed to stir at 23 °C for 12 h. The precipitate was then filtered, and the crude product was further purified by column chromatography on silica gel with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless oil (578 mg, 74%).



87a

Compound 87a. To a DCM (10 mL) solution of compound **85a** (179 mg, 0.677 mmol), compound **86** (640 mg, 0.615 mmol) and DPTS (271 mg, 0.922 mmol), was added DCC (380 mg, 1.85 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless viscous liquid (710 mg, 90%).

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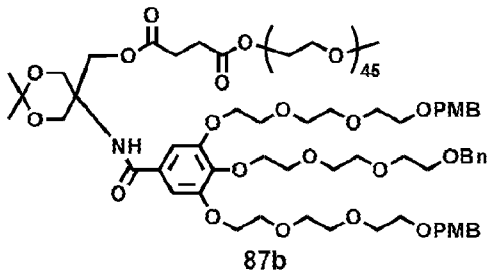
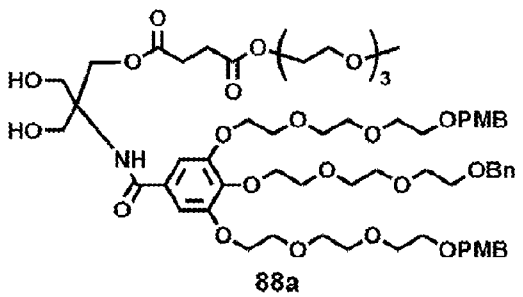


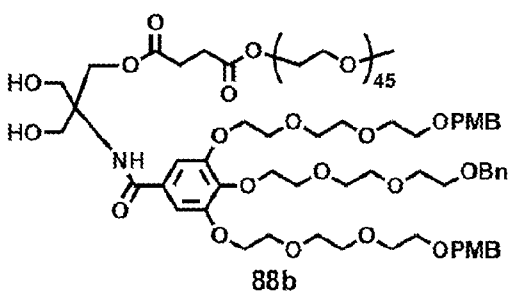
87b

Compound 87b. To a DCM (10 mL) solution of compound **85b** (1.17 g, 0.557 mmol), compound **86** (578 mg, 0.557 mmol) and DPTS (180 mg, 0.613 mmol), was added DCC (287 mg, 1.40 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless viscous liquid (960 mg, 55%).



88a

Compound 88a. To a MeOH solution (10 mL) of compound **87a** (710 mg, 0.552 mmol) was added 3 mL of HCl aqueous solution (2 M). The mixture was allowed to stir at 23 °C for 2 h and saturated $NaHCO_3$ solution (20 mL) was added. The mixture was extracted with DCM (50 mL) for 3 times. An organic extract was dried over anhydrous $MgSO_4$ and evaporated to dryness under reduced pressure to yield the title compound as a colorless viscous liquid without further purification (653 mg, 95%).



88b

Compound 88b. Compound **88b** was synthesized from compound **87b** (960 mg, 0.307 mmol) following a procedure similar to that used for the synthesis of compound **88a**. The title compound was obtained as a colorless viscous liquid without further purification (930 mg, 96%).

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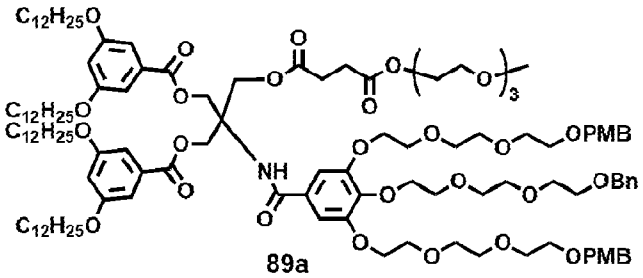
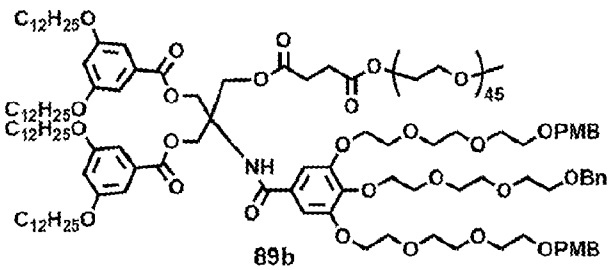


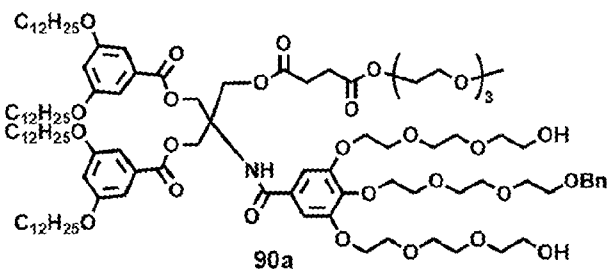
89a

Compound 89a. To a DCM (10 mL) solution of compound **41d** (539 mg, 1.10 mmol), compound **88a** (653 mg, 0.524 mmol) and DPTS (384 mg, 1.31 mmol), was added DCC (541 mg, 2.62 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless viscous liquid (933 mg, 81%).



89b

Compound 89b. To a DCM (10 mL) solution of compound **41d** (106 mg, 0.220 mmol), compound **88b** (302 mg, 0.098 mmol) and DPTS (74 mg, 0.250 mmol), was added DCC (103 mg, 0.500 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless viscous liquid (351 mg, 89%).



90a

Compound 90a. To a mixed solution of DCM (15 mL) and water (0.75 mL) of compound **89a** (933 mg, 0.426 mmol), was added DDQ (241 mg, 1.06 mmol). The mixture was allowed to stir at 23 °C for 3 h. The precipitates were filtered out and DCM (20 mL) was added. The mixture was washed by $NaHCO_3$ aqueous solution (saturated), $NaHSO_3$ aqueous solution (2%) and $NaHCO_3$

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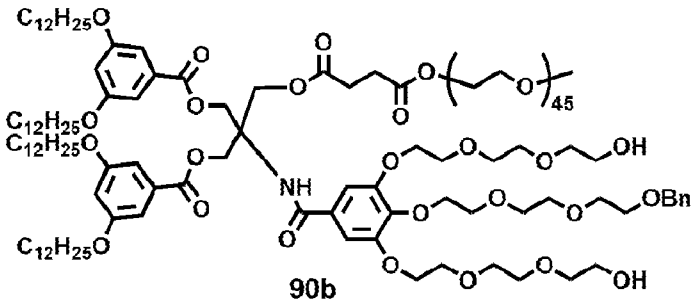
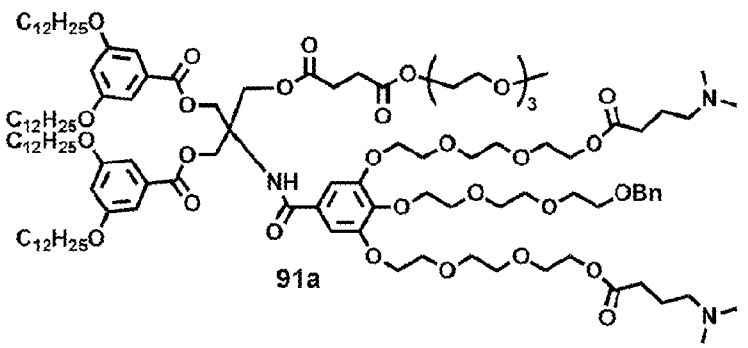
aqueous solution (saturated) successively. The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography with a mobile phase of DCM/MeOH = 20/1 to yield the title compound as a colorless viscous liquid (504 mg, 61%).



90b

Compound 90b. To a mixed solution of DCM (5 mL) and water (0.25 mL) of compound **90a** (270 mg, 0.067 mmol), was added DDQ (51 mg, 0.220 mmol). The mixture was allowed to stir at 23 °C for 2 h. The completion of the reaction was monitored by ^1H NMR when the signal at δ = 4.46 from the p-methoxy benzyl groups was completely disappeared. The precipitates were filtered out and DCM (20 mL) was added. The mixture was washed by $NaHCO_3$ aqueous solution (saturated), $NaHSO_3$ aqueous solution (2%) and $NaHCO_3$ aqueous solution (saturated) successively. The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography with a mobile phase of EtOAc/MeOH = 8/1 to yield the title compound as a colorless viscous liquid (236 mg, 91%).



91a

Compound 91a ($4/2_{DMBA}^{1,3}{}_{Bn}^{2}$). Compound **90a** (205 mg, 0.105 mmol) and 4-(dimethylamino)butyric acid hydrochloride (44 mg, 0.263 mmol) were dissolved in 5 mL dry DCM. DCC (108 mg, 0.525 mmol) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 10/1, 6/1 with 0.1% TEA as the eluent. Then the crude product was dissolved in $NaHCO_3$ solution (2%, 30 mL) and the mixture was extracted by DCM

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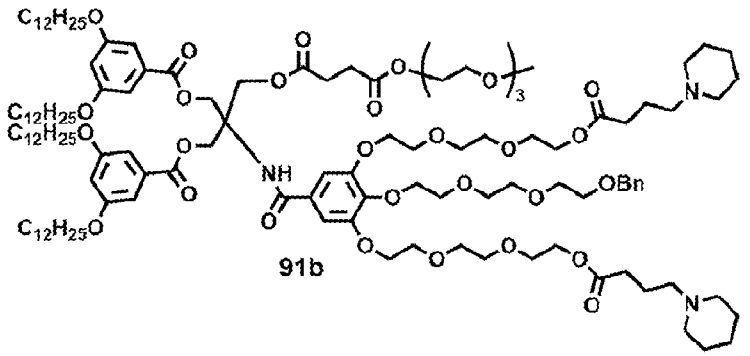
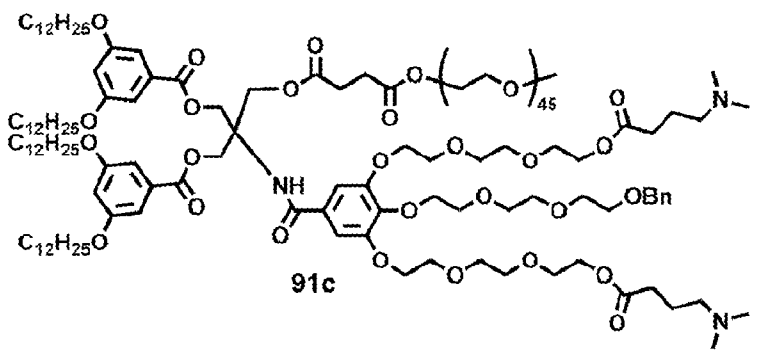
(30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (205 mg, 90%).



Compound 91b ($4/2_{PIP}^{1,3}{}_{Bn}^{2}$). Compound **91b** was synthesized from compound **90a** (154 mg, 0.079 mmol), compound **56a** (44 mg, 0.263 mmol) and DCC (33 mg, 0.193 mmol) following a procedure similar to that used for the synthesis of compound **91a**. The title compound was obtained as a light-yellow oil (160 mg, 90%).



Compound 91c ($4/2_{DMBA}^{1,3}{}_{Bn}^{2}{}_{PEG}^{4}$). Compound **91c** was synthesized from compound **90b** (230 mg, 0.061 mmol), 4-(dimethylamino)butyric acid hydrochloride (26 mg, 0.150 mmol) and DCC (62 mg, 0.300 mmol) following a procedure similar to that used for the synthesis of compound **91a**. The title compound was obtained as a light-yellow solid (209 mg, 84%).

Dimensions of DNPs Containing Luc-mRNA Obtained by Co-assembly of IAJDs from Library 1 with Luc-mRNA by Injection in Citrate Buffer (10 mM, pH 3.0) or Acetate Buffer (10 mM, pH 4.0/5.2) and Luminescence Expression in HEK293T Cells (125 ng/well) are presented in Table 1.

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Table 1

IAJD		Buffer (10 mM)	C_{IAJD} (mg/mL)	$C_{Luc\text{-}mRNA}$ (mg/mL)	Size (d, nm)	PDI	Luminescence	Standard deviation
Library 1 Single-Single	**1**	Citrate (pH 3.0)	4	0.05	73	0.406	552,723	83,190
	1	Acetate (pH 4.0)	4	0.05	85	0.282	1,009,268	180,480
	1	Acetate (pH 5.2)	4	0.10	295	0.501	183,282	55,508
	2	Citrate (pH 3.0)	4	0.05	94	0.530	165,500	15,360
	2	Acetate (pH 4.0)	4	0.05	83	0.254	122,687	38,579
	2	Acetate (pH 5.2)	4	0.10	289	0.521	71,265	43,096
	3	Citrate (pH 3.0)	4	0.05	70	0.311	7,093	1,946
	3	Acetate (pH 4.0)	4	0.05	65	0.272	4,412	199
	3	Acetate (pH 5.2)	4	0.10	95	0.244	3,581	556
	4	Citrate (pH 3.0)	4	0.05	61	0.460	2,063	82
	4	Acetate (pH 4.0)	4	0.05	58	0.330	3,283	1,981
	4	Acetate (pH 5.2)	4	0.10	178	0.284	1,613	156
	5	Citrate (pH 3.0)	4	0.05	77	0.320	18,939	4,425
	5	Acetate (pH 4.0)	4	0.05	60	0.291	17,156	12,967
	5	Acetate (pH 5.2)	4	0.10	156	0.281	1,226	238
	6	Citrate (pH 3.0)	4	0.05	91	0.353	15,207	1,536
	6	Acetate (pH 4.0)	4	0.05	59	0.291	6,845	1,073
	6	Acetate (pH 5.2)	4	0.10	92	0.246	33,562	4,359
	7	Citrate (pH 3.0)	4	0.05	100	0.406	10,119	1,374
	7	Acetate (pH 4.0)	4	0.05	86	0.329	1,810	43
	7	Acetate (pH 5.2)	4	0.10	132	0.338	9,162	801
	8	Citrate (pH 3.0)	4	0.05	73	0.442	1,291,219	204,977
	8	Acetate (pH	4	0.05	70	0.295	1,975,892	540,459

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		4.0)						
	8	Acetate (pH 5.2)	4	0.10	142	0.269	1,473,247	158,493
	9	Citrate (pH 3.0)	4	0.05	87	0.271	4,236,960	428,817
	9	Acetate (pH 4.0)	4	0.05	55	0.306	4,474,957	215,800
	9	Acetate (pH 5.2)	4	0.10	98	0.224	4,390,218	197,649
Negative							81	5
MC3							1,320,533	105,967
***Trans*IT**							3,642,765	484,480

Dimensions of DNPs Containing Luc-mRNA Obtained by Co-assembly of IAJDs from Library 2-4 with Luc-mRNA by Injection in Citrate Buffer (10 mM, pH 3.0) or Acetate Buffer (10 mM, pH 4.0/5.2) and Luminescence Expression in HEK293T Cells (125 ng/well) are presented in Table 2.

Table 2

IAJD		**Buffer (10 mM)**	**C_{IAJD} (mg/mL)**	**$C_{Luc\text{-}mRNA}$ (mg/mL)**	**Size (d, nm)**	**PDI**	**Luminescence**	**Standard deviation**
	19	Citrate (pH 3.0)	4	0.05	77	0.271	39	10
	19	Acetate (pH 4.0)	4	0.05	65	0.274	39	7
	19	Acetate (pH 5.2)	4	0.10	110	0.167	33	7
	20	Citrate (pH 3.0)	4	0.05	72	0.293	249	89
	20	Acetate (pH 4.0)	4	0.05	62	0.386	28	3
Library 2 Single-Single	**20**	Acetate (pH 5.2)	4	0.10	120	0.203	47,808	8,089
	21	Citrate (pH 3.0)	4	0.05	65	0.414	927,496	97,565
	21	Acetate (pH 4.0)	4	0.05	68	0.301	1,098,558	149,158
	21	Acetate (pH 5.2)	4	0.10	102	0.200	2,901,220	237,377
	22	Citrate (pH 3.0)	4	0.05	69	0.296	4,740,529	464,414
	22	Acetate (pH 4.0)	4	0.05	66	0.280	4,369,979	1,133,074
	22	Acetate (pH 5.2)	4	0.10	114	0.232	4,664,358	733,410
	23	Citrate (pH	4	0.05	86	0.213	159	10

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		3.0)						
	23	Acetate (pH 4.0)	4	0.05	68	0.328	38	8
	23	Acetate (pH 5.2)	4	0.10	108	0.230	44	13
	24	Citrate (pH 3.0)	4	0.05	64	0.421	5,578	3,523
	24	Acetate (pH 4.0)	4	0.05	50	0.311	27,766	9,240
	24	Acetate (pH 5.2)	4	0.10	108	0.216	339,002	27,855
	25	Citrate (pH 3.0)	4	0.05	117	0.538	116,292	8,791
	25	Acetate (pH 4.0)	4	0.05	64	0.425	5,658	1,859
	25	Acetate (pH 5.2)	4	0.10	147	0.202	57,464	7,513
	44	Citrate (pH 3.0)	4	0.05	85	0.310	66,956	3,183
	44	Acetate (pH 4.0)	4	0.05	98	0.279	84,746	5,790
	44	Acetate (pH 5.2)	4	0.10	171	0.252	99,996	103,05
	33	Citrate (pH 3.0)	4	0.05	74	0.278	36,871	18,033
	33	Acetate (pH 4.0)	4	0.05	72	0.260	32,404	24,064
	33	Acetate (pH 4.0)	4	0.10	93	0.216	100,259	11,162
	34	Citrate (pH 3.0)	4	0.05	83	0.264	37,381	4,560
Library 3 Single-Single	**34**	Acetate (pH 4.0)	4	0.05	56	0.255	52,554	36,103
	34	Acetate (pH 4.0)	4	0.10	83	0.245	118,838	35,286
	45	Citrate (pH 3.0)	4	0.05	64	0.431	22	12
	45	Acetate (pH 4.0)	4	0.05	102	0.267	32	5
	45	Acetate (pH 5.2)	4	0.10	512	0.536	15	6
	35	Citrate (pH 3.0)	4	0.05	69	0.277	58	11
	35	Acetate (pH 4.0)	4	0.05	53	0.260	76	13
	35	Acetate (pH 4.0)	4	0.10	70	0.245	78	41
	36	Citrate (pH	4	0.05	58	0.269	42	14

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		3.0)						
	36	Acetate (pH 4.0)	4	0.05	53	0.265	46	19
	36	Acetate (pH 5.2)	4	0.10	71	0.224	83	22
Library 4 Single-Single	**30**	Citrate (pH 3.0)	4	0.05	234	0.202	662	142
	30	Acetate (pH 4.0)	4	0.05	73	0.321	5,421	1,812
	30	Acetate (pH 5.2)	4	0.10	165	0.377	674	52
	31	Citrate (pH 3.0)	4	0.05	166	0.881	16,166	4,370
	31	Acetate (pH 4.0)	4	0.05	76	0.489	41,889	5,247
	31	Acetate (pH 5.2)	4	0.10	210	0.261	28,524	3,995
	37	Citrate (pH 3.0)	4	0.05	118	0.526	3,800	265
	37	Acetate (pH 4.0)	4	0.05	79	0.329	30,785	4,803
	37	Acetate (pH 4.0)	4	0.10	131	0.267	2,606	489

Dimensions of DNPs Containing Luc-mRNA Obtained by Co-assembly of IAJDs from Library 5 with Luc-mRNA by Injection in Citrate Buffer (10 mM, pH 3.0) or Acetate Buffer (10 mM, pH 4.0/5.2) and Luminescence Expression in HEK293T Cells (125 ng/well) are presented in Table 3.

Table 3

IAJD		Buffer (10 mM)	C_{IAJD} (mg/mL)	$C_{Luc\text{-}mRNA}$ (mg/mL)	Size (d, nm)	PDI	Luminescence	Standard deviation
Library 5 Twin-Twin	**10**	Citrate (pH 3.0)	4	0.05	110	0.467	52,530	6,718
	10	Acetate (pH 4.0)	4	0.05	92	0.242	64,605	33,615
	10	Acetate (pH 5.2)	4	0.10	347	0.449	131,357	22,629
	11	Citrate (pH 3.0)	4	0.05	108	0.464	26,378	5,030
	11	Acetate (pH 4.0)	4	0.05	90	0.272	26,843	8,193
	11	Acetate (pH 5.2)	4	0.10	369	0.477	24,728	10,049

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12	Citrate (pH 3.0)	4	0.05	66	0.473	87	33
12	Acetate (pH 4.0)	4	0.05	64	0.326	71	11
12	Acetate (pH 5.2)	4	0.10	171	0.467	331	82
13	Citrate (pH 3.0)	4	0.05	66	0.331	83	44
13	Acetate (pH 4.0)	4	0.05	63	0.319	49	8
13	Acetate (pH 5.2)	4	0.10	163	0.299	251	92
14	Citrate (pH 3.0)	4	0.05	75	0.474	187	121
14	Acetate (pH 4.0)	4	0.05	64	0.456	77	41
14	Acetate (pH 5.2)	4	0.10	98	0.206	38	10
15	Citrate (pH 3.0)	4	0.05	76	0.431	18,158	10,971
15	Acetate (pH 4.0)	4	0.05	84	0.265	14,233	1,705
15	Acetate (pH 5.2)	4	0.10	103	0.262	46,433	11,414
16	Citrate (pH 3.0)	4	0.05	77	0.429	12,299	2,475
16	Acetate (pH 4.0)	4	0.05	56	0.270	7,789	2,103
16	Acetate (pH 5.2)	4	0.10	108	0.183	27,883	7,388
17	Citrate (pH 3.0)	4	0.05	82	0.398	32,239	11,612
17	Acetate (pH 4.0)	4	0.05	57	0.472	34,120	5,598
17	Acetate (pH 5.2)	4	0.10	203	0.449	19,080	3,288
18	Citrate (pH 3.0)	4	0.05	69	0.333	368,727	62,081
18	Acetate (pH 4.0)	4	0.05	52	0.325	478,097	29,427
18	Acetate (pH 5.2)	4	0.10	156	0.370	161,286	14,020
46	Citrate (pH 3.0)	4	0.05	71	0.433	186,720	60,386
46	Acetate (pH 4.0)	4	0.05	78	0.293	101,862	45,648
46	Acetate (pH 4.0)	4	0.10	111	0.217	2,819	2,466

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Dimensions of DNPs Containing Luc-mRNA Obtained by Co-assembly of IAJDs from Library 6 with Luc-mRNA by Injection in Citrate Buffer (10 mM, pH 3.0) or Acetate Buffer (10 mM, pH 4.0/5.2) and Luminescence Expression in HEK293T Cells (125 ng/well) are presented in Table 4.

Table 4.

IAJD		Buffer (10 mM)	C_{IAJD} (mg/mL)	$C_{Luc-mRNA}$ (mg/mL)	Size (d, nm)	PDI	Luminescence	Standard deviation
Library 6 Hybrid Twin-Mix	**38**	Citrate (pH 3.0)	4	0.05	96	0.341	32	5
	38	Acetate (pH 4.0)	4	0.05	119	0.353	45	17
	38	Acetate (pH 5.2)	4	0.10	418	0.364	570	206
	39	Citrate (pH 3.0)	4	0.05	117	0.430	59	9
	39	Acetate (pH 4.0)	4	0.05	147	0.478	74	13
	39	Acetate (pH 5.2)	4	0.10	242	0.392	311	88
	40	Citrate (pH 3.0)	4	0.05	98	0.531	9,184	1,028
	40	Acetate (pH 4.0)	4	0.05	73	0.439	12,389	1,296
	40	Acetate (pH 5.2)	4	0.10	423	0.388	11,392	1,209
	41	Citrate (pH 3.0)	4	0.05	107	0.509	10,079	1,494
	41	Acetate (pH 4.0)	4	0.05	104	0.220	38,864	6,688
	41	Acetate (pH 5.2)	4	0.10	254	0.517	45,553	4,585
	42	Citrate (pH 3.0)	4	0.05	59	0.418	637	43
	42	Acetate (pH 4.0)	4	0.05	50	0.323	4,357	90
	42	Acetate (pH 5.2)	4	0.10	307	0.570	5,137	2,297
	49	Citrate (pH 3.0)	4	0.05	70	0.356	7,925	2,909
	49	Acetate (pH 4.0)	4	0.05	72	0.253	7,908	1,832
	49	Acetate (pH 5.2)	4	0.10	242	0.340	4,632	1,337

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26	Citrate (pH 3.0)	4	0.05	74	0.310	330,183	5,311
26	Acetate (pH 4.0)	4	0.05	75	0.296	598,804	98,063
26	Acetate (pH 5.2)	4	0.10	227	0.448	184,041	28,499
27	Citrate (pH 3.0)	4	0.05	72	0.449	246,051	23,408
27	Acetate (pH 4.0)	4	0.05	78	0.266	330,414	133,482
27	Acetate (pH 5.2)	4	0.10	306	0.538	81,351	11,532
28	Citrate (pH 3.0)	4	0.05	87	0.341	87,848	14,593
28	Acetate (pH 4.0)	4	0.05	76	0.284	114,911	15,142
28	Acetate (pH 5.2)	4	0.10	266	0.488	66,366	6,091
50	Citrate (pH 3.0)	4	0.05	86	0.386	6,871	2,245
50	Acetate (pH 4.0)	4	0.05	77	0.376	10,925	3,855
50	Acetate (pH 5.2)	4	0.10	166	0.227	9,872	3,498
29	Citrate (pH 3.0)	4	0.05	65	0.404	418,644	95,190
29	Acetate (pH 4.0)	4	0.05	84	0.285	546,335	32,895
29	Acetate (pH 5.2)	4	0.10	233	0.323	243,618	17,531
47	Citrate (pH 3.0)	4	0.05	82	0.443	279	188
47	Acetate (pH 4.0)	4	0.05	81	0.416	775	276
47	Acetate (pH 4.0)	4	0.10	121	0.236	702	109
51	Citrate (pH 3.0)	4	0.05	89	0.361	1,176	256
51	Acetate (pH 4.0)	4	0.05	99	0.352	734	62
51	Acetate (pH 4.0)	4	0.10	115	0.254	2,498	211
52	Citrate (pH 3.0)	4	0.05	112	0.476	102	35
52	Acetate (pH 4.0)	4	0.05	175	0.754	56	25
52	Acetate (pH 4.0)	4	0.10	323	0.885	38	26

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43	Citrate (pH 3.0)	4	0.05	192	0.283	819	548
43	Acetate (pH 4.0)	4	0.05	106	0.493	3,876	1,046
43	Acetate (pH 5.2)	4	0.10	199	0.311	18,268	1,889
48	Citrate (pH 3.0)	4	0.05	187	0.464	1,831	449
48	Acetate (pH 4.0)	4	0.05	94	0.445	2,056	1,200
48	Acetate (pH 4.0)	4	0.10	110	0.248	1,323	640
32	Citrate (pH 3.0)	4	0.05	39	0.435	51	17
32	Acetate (pH 4.0)	4	0.05	54	0.423	55	14
32	Acetate (pH 5.2)	4	0.10	61	0.245	35	18

Dimensions of DNPs Containing Luc-mRNA Obtained by Co-assembly of IAJD9 or 22 with Different Concentrations and Luc-mRNA in Acetate Buffer (10 mM) and Luminescence Expression in HEK293T Cells (125 ng/well) are presented in Table 5.

Table 5.

IAJD	**pH of acetate buffer (10 mM)**	**C_{IAJD} (mg/mL)**	**$C_{Luc-mRNA}$ (mg/mL)**	**Size (d, nm)**	**PDI**	**Luminescence**	**Standard deviation**
9	4.0	8.00	0.05	66	0.281	2,929,485	200,696
	4.0	6.00	0.05	65	0.295	4,732,744	455,960
	4.0	5.00	0.05	44	0.341	5,357,139	189,684
	4.0	4.00	0.05	59	0.291	5,316,758	140,302
	4.0	3.00	0.05	66	0.282	4,150,126	954,384
	4.0	2.00	0.05	65	0.258	2,533,441	484,087
	4.0	1.00	0.05	71	0.270	1,566,729	370,057
	4.0	0.75	0.05	75	0.241	683,904	155,907
	4.0	0.50	0.05	91	0.270	197,838	47,356
	4.0	0.25	0.05	112	0.243	32,208	8,832
9	4.0	4.00	0.05	68	0.153	4,832,988	612,805
	4.4	4.00	0.05	50	0.280	4,949,850	601,419
	4.8	4.00	0.05	56	0.281	5,278,352	476,945
	5.2	4.00	0.05	74	0.259	5,306,972	421,485
	5.6	4.00	0.05	140	0.209	4,053,924	307,161
22	4.0	8.00	0.05	50	0.302	5,246,616	131,335
	4.0	6.00	0.05	45	0.292	6,493,891	108,856
	4.0	5.00	0.05	52	0.272	7,317,328	192,067
	4.0	4.00	0.05	53	0.288	6,922,629	427,518

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	4.0	3.00	0.05	57	0.291	5,695,413	221,600
	4.0	2.00	0.05	55	0.256	5,269,341	508,483
	4.0	1.00	0.05	65	0.252	2,502,511	388,969
	4.0	0.75	0.05	72	0.232	891,902	191,482
	4.0	0.50	0.05	71	0.195	178,965	53,060
	4.0	0.25	0.05	92	0.204	41,897	5,500
	4.0	4.00	0.05	49	0.307	5,069,642	275,369
	4.4	4.00	0.05	66	0.288	5,409,310	427,776
22	4.8	4.00	0.05	74	0.264	5,106,741	405,158
	5.2	4.00	0.05	71	0.237	5,390,693	402,291
	5.6	4.00	0.05	115	0.225	3,939,728	668,494

Dimensions of DNPs Containing Luc-mRNA Obtained by Co-assembly of IAJD33 or 34 with Different Concentrations and Luc-mRNA in Acetate Buffer (10 mM) and Luminescence Expression in HEK293T Cells (125 ng/well) are presented in Table 6.

Table 6

IAJD	pH of acetate buffer (10 mM)	C_{IAJD} (mg/mL)	$C_{Luc\text{-}mRNA}$ (mg/mL)	Size (d, nm)	PDI	Luminescence	Standard deviation
	4.0	8.00	0.05	94	0.262	4,707	870
	4.0	7.00	0.05	85	0.283	7,539	1,049
	4.0	6.00	0.05	72	0.277	15,845	2,552
	4.0	5.00	0.05	75	0.263	23,237	8,898
	4.0	4.00	0.05	66	0.256	32,404	24,064
33	4.0	3.00	0.05	60	0.274	62,770	14,762
	4.0	2.00	0.05	59	0.263	74,283	11,774
	4.0	1.00	0.05	64	0.207	56,565	18,157
	4.0	0.75	0.05	74	0.221	44,198	23,163
	4.0	0.50	0.05	74	0.209	19,310	1,217
	4.0	0.25	0.05	88	0.198	14,035	3,681
	4.0	8.00	0.05	85	0.245	4,362	2,487
	4.0	7.00	0.05	69	0.289	9,060	3,341
	4.0	6.00	0.05	59	0.238	15,583	7,266
	4.0	5.00	0.05	59	0.256	31,303	7,940
	4.0	4.00	0.05	56	0.255	52,554	36,103
34	4.0	3.00	0.05	50	0.212	51,933	17,038
	4.0	2.00	0.05	49	0.212	90,071	59,289
	4.0	1.00	0.05	64	0.206	95,041	56,801
	4.0	0.75	0.05	73	0.231	46,628	37,069
	4.0	0.50	0.05	65	0.209	5,491	2,269
	4.0	0.25	0.05	88	0.211	61	26

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Dimensions of DNPs Containing Luc-mRNA and Luminescence Expression *in vivo* are presented in Table 7.

Table 7.

IAJD		C_{IAJD} (mg/mL)	$C_{Luc-mRNA}$ (mg/mL)	pH		Size (d, nm)	PDI	T (h)[a]	Organs with luminescence
				Acetate buffer	Sample				
Library 1 Single-Single	**1**	4.0	0.1	5.2	7.10	295	0.501	6	Lung, spleen
	2	4.0	0.1	5.2	7.13	289	0.521	6	Lung, spleen
	3	4.0	0.1	5.2	7.07	95	0.244	7	--
	4	4.0	0.1	5.2	7.45	178	0.284	7	--
	5	4.0	0.1	5.2	7.78	156	0.281	7	--
	6	4.0	0.1	5.2	6.30	92	0.246	--	--
	7	4.0	0.1	5.2	6.08	132	0.338	--	--
	8	4.0	0.1	5.2	7.23	142	0.269	7	Lung
	9a	4.0	0.1	5.2	7.30	98	0.224	6	Lung
	9b	4.0	0.025	3.0[b]	3.71	49	0.464	5	Lung, liver, spleen
Library 2 Single-Single	**19**	4.0	0.1	5.2	7.52	110	0.167	6	Lung, liver, spleen
	20	4.0	0.1	5.2	7.63	120	0.203	6	Lung, spleen
	21	4.0	0.1	5.2	7.23	102	0.200	6	--
	22	4.0	0.1	5.2	7.39	114	0.232	6	Lung
	23	4.0	0.1	5.2	7.74	108	0.230	6	Lung, liver, spleen
	24	4.0	0.1	5.2	7.61	108	0.216	6	Lung, spleen
	25	4.0	0.1	5.2	6.90	147	0.202	6	Lung, spleen
Library 3 Single-Single	**44**	4.0	0.1	5.2	7.15	171	0.252	4	Lung, liver
	33	4.0	0.1	4.0	4.96	73	0.248	6	Lung, liver
	34	4.0	0.1	4.0	5.10	83	0.245	6	Lung, liver

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	45	4.0	0.1	5.2	6.87	512	0.536	4	Lung
	35	4.0	0.1	4.0	4.76	70	0.243	6	--
	36	4.0	0.1	4.0	4.90	71	0.224	6	--
Library 4 Single-Single	**30a**	4.0	0.1	5.2	5.95	165	0.377	7	Lung, liver, spleen
	30b	4.0	0.1	5.2	6.13	134	0.408	7	Lung, liver, spleen
	31	4.0	0.1	5.2	7.02	210	0.261	7	Lung, liver, spleen
	37	4.0	0.1	4.0	4.52	131	0.267	6	Lung, liver, spleen
Library 5 Twin-Twin	**10**	4.0	0.1	5.2	7.76	347	0.449	7	Lung
	11	4.0	0.1	5.2	7.13	369	0.477	7	--
	12	4.0	0.1	5.2	7.35	171	0.467	7	--
	13	4.0	0.1	5.2	7.35	163	0.299	7	--
	14	4.0	0.1	5.2	7.26	98	0.206	6	--
	15	4.0	0.1	5.2	5.78	103	0.262	--	--
	16	4.0	0.1	5.2	5.84	108	0.183	--	--
	17	4.0	0.1	5.2	7.13	203	0.449	6	Lung, spleen
	18	4.0	0.1	5.2	7.17	156	0.370	6	Lung, spleen
	46	4.0	0.1	4.0	4.68	111	0.217	4	Lung, liver, spleen
Library 6 Hybrid Twin-Mix	**38**	4.0	0.1	5.2	6.59	418	0.364	4	--
	39	4.0	0.1	5.2	6.66	242	0.392	4	--
	40	4.0	0.1	5.2	6.87	423	0.388	4	Lung
	41	4.0	0.1	5.2	7.68	254	0.517	4	--
	42	4.0	0.1	5.2	7.26	307	0.570	4	--
	49	4.0	0.1	5.2	7.27	242	0.340	4	--
	26	4.0	0.1	5.2	6.93	227	0.448	6	Lung, liver, spleen

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27	4.0	0.1	5.2	6.83	306	0.538	6	Lung, liver, spleen
28	4.0	0.1	5.2	6.67	266	0.488	7	Lung, liver
50	4.0	0.1	5.2	7.53	166	0.227	**4**	--
29	4.0	0.1	5.2	6.81	233	0.323	7	Lung, liver, spleen
47	4.0	0.1	4.0	4.29	121	0.236	**4**	Lung
51	4.0	0.1	4.0	4.89	115	0.254	6	--
52	4.0	0.1	4.0	4.84	323	0.885	6	--
43	4.0	0.1	5.2	7.15	199	0.311	**4**	Lung, spleen
48	4.0	0.1	4.0	4.34	110	0.248	--	--
32	4.0	0.1	5.2	6.44	61	0.245	6	--

a: Time from injection to the characterization of mice.

b: Sample was prepared in citrate buffer (10 mM, pH = 3.0) and then dialyzed against 1X PBS for 2 h at 4 °C.

The effect of vortex time on the dimensions of assemblies of IAJDS was determined. The results are shown in Table 8 and in Figure 16.

Table 8.

Sample No.	Vortex Time (Sec)	Size (nm)	PDI
1	5	81	0.250
2	10	81	0.251
3	30	85	0.240
4	60	81	0.247
5	120	78	0.250
6	300	77	0.249

Sample preparation: Injected 25 μL of IAJD9 (80 mg/mL) ethanol stock solution into 475 μL tris buffer (pH 7.4), followed by vortex for (5-300) seconds. The final concentration of IAJD9 was 4.0 mg/mL.

The DLS data of assemblies of the dendrimers is presented in Figure 17 through Figure 35.

The stability of the DNPs as assayed by DLS. All the samples are prepared in acetate buffer. The stabilities of the constructs were ranked: Excellent (in bold) – the original sizes and

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distribution remains almost the same after storage; Good – the original relatively big sizes became smaller or broad size distribution became narrower; Not stable – the original relatively big sizes or broad size distribution remained almost the similar level, or the original relatively small sizes became bigger or narrow size distribution became broader. The results are shown in Table 9.

Examples of excellent stability are presented in Figure 11.

Table 9. Stability of DNPs by DLS.

IAJD (storage time)		C_{IAJD} (mg/mL)	$C_{Luc\text{-}mRNA}$ (mg/mL)	pH		Original size (d, nm)	PDI	Size (d, nm)	PDI	Ranking of stability[c]
				Acetate buffer	Sample					
Library 1 Single-Single	1 (60 days)	4.0	0.1	5.2	7.10	295 (M)[b]	0.501	215 (M)	1.000	Not stable
	2 (60 days)	4.0	0.1	5.2	7.13	289 (M)	0.521	164 (M)	1.000	Not stable
	3 (60 days)	4.0	0.1	5.2	7.07	95 (U)	0.244	1,666 (B)	0.733	Not stable
	4 (60 days)	4.0	0.1	5.2	7.45	178 (B)	0.284	4,923 (U)	0.155	Not stable
	5 (60 days)	4.0	0.1	5.2	7.78	156 (U)	0.281	302 (B)	0.509	Not stable
	6 (60 days)	4.0	0.1	5.2	6.30	92 (B)	0.246	251 (U)	0.464	Not stable
	7 (60 days)	4.0	0.1	5.2	6.08	132 (B)	0.338	108 (M)	0.686	Not stable
	8 (60 days)	4.0	0.1	5.2	7.23	142 (U)	0.269	327 (M)	0.526	Not stable
	9 (120 days)	**4.0**	**0.05**	**4.0**	--	**55 (U)**	**0.306**	**59 (U)**	**0.296**	**Excellent**
Library 2 Single-Single	**19 (60 days)**	**4.0**	**0.1**	**5.2**	**7.52**	**110 (U)**	**0.167**	**141 (U)**	**0.118**	**Excellent**
	20 (60 days)	**4.0**	**0.1**	**5.2**	**7.63**	**120 (U)**	**0.203**	**122 (U)**	**0.152**	**Excellent**
	21 (60 days)	4.0	0.1	5.2	7.23	102 (U)	0.200	933 (B)	1.000	Not stable
	22 (120 days)	**4.0**	**0.05**	**4.0**	--	**66 (U)**	**0.280**	**67 (U)**	**0.249**	**Excellent**
	23 (60 days)	**4.0**	**0.1**	**5.2**	**7.74**	**108 (U)**	**0.230**	**117 (U)**	**0.123**	**Excellent**
	24 (60 days)	**4.0**	**0.1**	**5.2**	**7.61**	**108 (U)**	**0.216**	**126 (U)**	**0.217**	**Excellent**
	25 (60 days)	4.0	0.1	5.2	6.90	147 (U)	0.202	107 (B)	0.538	Not stable
Library 3 Single-	**44 (23 days)**	**4.0**	**0.1**	**5.2**	**7.15**	**171 (B)**	**0.252**	**202 (B)**	**0.297**	**Excellent**
	33 (70	**4.0**	**0.1**	**4.0**	**4.96**	**93 (U)**	**0.21**	**96 (U)**	**0.225**	**Excelle**

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Single	**days)**						**6**			**nt**
	34 (70 days)	**4.0**	**0.1**	**4.0**	**5.10**	**83 (U)**	**0.245**	**86 (U)**	**0.205**	**Excellent**
	45 (23 days)	4.0	0.1	5.2	6.87	512 (M)	0.536	283 (B)	0.501	Not stable
Library 4 Single-Single	30 (60 days)	4.0	0.1	5.2	5.95	165 (B)	0.377	657 (U)	0.589	Not stable
	31 (60 days)	4.0	0.1	5.2	7.02	210 (U)	0.261	189 (M)	0.910	Not stable
	37 (50 days)	**4.0**	**0.1**	**4.0**	**4.52**	**131 (U)**	**0.267**	**130 (U)**	**0.215**	**Excellent**
Library 5 Twin-Twin	**10 (60 days)**	**4.0**	**0.05**	**4.0**	**--**	**92 (U)**	**0.242**	**99 (U)**	**0.269**	**Excellent**
	14 (60 days)	4.0	0.1	5.2	7.26	98 (U)	0.206	235 (U)	0.289	Not stable
	15 (60 days)	**4.0**	**0.1**	**5.2**	**5.78**	**103 (U)**	**0.262**	**95 (U)**	**0.287**	**Excellent**
	16 (60 days)	**4.0**	**0.1**	**5.2**	**5.84**	**108 (U)**	**0.183**	**103 (U)**	**0.218**	**Excellent**
	17 (60 days)	4.0	0.1	5.2	7.13	203 (M)	0.449	149 (U)	0.153	Good
	18 (60 days)	4.0	0.1	5.2	7.17	156 (U)	0.370	131 (U)	0.136	Good
	46 (21 days)	**4.0**	**0.1**	**4.0**	**4.68**	**111 (U)**	**0.217**	**102 (U)**	**0.240**	**Excellent**
Library 6 Hybrid Twin-Mix	40 (23 days)	4.0	0.1	5.2	6.87	423 (B)	0.388	302 (U)	0.158	Good
	49 (21 days)	4.0	0.1	5.2	7.27	242 (U)	0.340	193 (U)	0.179	Good
	27 (60 days)	4.0	0.1	5.2	6.83	306 (M)	0.538	115 (U)	0.262	Good
	50 (21 days)	**4.0**	**0.1**	**5.2**	**7.53**	**166 (U)**	**0.227**	**165 (U)**	**0.229**	**Excellent**
	29 (60 days)	4.0	0.1	5.2	6.81	233 (B)	0.323	151 (U)	0.256	Good
	47 (21 days)	**4.0**	**0.1**	**4.0**	**4.29**	**121 (U)**	**0.236**	**114 (U)**	**0.260**	**Excellent**
	51 (2 days)	**4.0**	**0.1**	**4.0**	**4.89**	**115 (U)**	**0.254**	**114 (U)**	**0.271**	**Excellent**
	52 (2 days)	4.0	0.1	4.0	4.84	323 (M)	0.885	322 (M)	0.454	Not stable
	48 (21 days)	**4.0**	**0.1**	**4.0**	**4.34**	**110 (U)**	**0.248**	**108 (U)**	**0.310**	**Excellent**
	32 (60 days)	**4.0**	**0.1**	**5.2**	**6.44**	**61 (U)**	**0.245**	**67 (U)**	**0.251**	**Excellent**

The serum stability of DNPs assembled from IAJD33+IAJD32 (2%) was determined.

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Sample preparation: injected 25 μL ethanol stock solution of IAJD33+IAJD32 (2%) (80 mg/mL of IAJDs) into the mixture of 462.5 μL acetate buffer (10 mM, pH 4.0) and 12.5 μL mRNA (4.0 mg/mL in water), followed by vortex for 5 seconds.

Final composition of DNP solution: IAJD33+IAJD32 (2%) (4.0 mg/mL) + mRNA (0.1 mg/mL) in 500 μL acetate buffer. The dimension of assemblies was checked by DLS.

The 500 μL DNP solution was dialyzed against 10 mM tris buffer (10 mM, pH 7.4) for 120 minutes in 3,500 – 14,000 molecular weight cut-off dialysis tube (Spectrum Medical Instruments Inc. Spectra/Por molecular porous membrane tubing Flat Width.: 45 mm; Diameter.: 29 mm & Vol/length.: 6.4 mL/cm).

DNPs in 1% Serum: Serum (5.5 μL) + [IAJDs (4.0 mg/mL) + mRNA (0.1 mg/mL) in 500 μL solution after dialysis in tris buffer]. The dimensions of the resulting DNPs are presented in Table 10 and Figure 36.

Table 10.

Samples	Size (nm)	PDI
DNPs of IAJD33+32 (2%) in acetate buffer (pH 4.0)	92	0.352
DNPs of IAJD33+32 (2%) after dialysis	105	0.170
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 0 min)	146	0.150
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 10 min)	147	0.183
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 20 min)	145	0.180
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 30 min)	148	0.171
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 40 min)	144	0.161
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 50 min)	145	0.154
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 60 min)	143	0.141

The pK_a values of the individual IAJD molecules was assayed. The results for each library are presented in Table 11. Titration curves for each compound are presented in Figure 37 through Figure 46.

Table 11. pK_a of Individual IAJDs Molecules

Library 1					
IAJD No.	pK_a 1	pK_a 2	pK_a 3	Avg pK_a	Std Div
IAJD 1	6.27			6.27	
IAJD 2	6.20			6.20	
IAJD 3	6.41			6.41	
IAJD 4	6.43			6.43	
IAJD 5	6.40			6.40	
IAJD 6	6.32	6.28		6.30	0.03
IAJD 7	6.28	6.08		6.18	0.14
IAJD 8	6.53			6.53	
IAJD 9	6.52	6.52	6.64	6.56	0.07

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Library 2					
IAJD No.	pK_a 1	pK_a 2	pK_a 3	Avg pK_a	Std Div
IAJD 19	6.38			6.38	
IAJD 20	6.34			6.34	
IAJD 21	6.28			6.28	
IAJD 22	6.32	6.42	6.41	6.38	0.06
IAJD 23	6.38			6.38	
IAJD 24	6.40			6.40	
IAJD 25	6.15			6.15	
Library 3					
IAJD No.	pK_a 1	pK_a 2	pK_a 3	Avg pK_a	Std Div
IAJD 44	6.75			6.75	
IAJD 33	6.61	6.66	6.72	6.66	0.06
IAJD 34	6.68	6.77	6.77	6.74	0.05
IAJD 45	5.88	5.98		5.93	0.07
IAJD 35	6.16			6.16	
IAJD 36	5.89			5.89	
Library 4					
IAJD No.	pK_a 1	pK_a 2	pK_a 3	Avg pK_a	Std Div
IAJD 30	6.42			6.42	
IAJD 31	6.70			6.70	
IAJD 37	6.83			6.83	
Library 5					
IAJD No.	pK_a 1	pK_a 2	pK_a 3	Avg pK_a	Std Div
IAJD 10	6.34			6.34	
IAJD 11	6.27	6.33		6.30	0.04
IAJD 12	6.25			6.25	
IAJD 13	6.20			6.20	
IAJD 14	6.02			6.02	
IAJD 15	5.71	5.96		5.84	0.18
IAJD 16	5.52			5.52	
IAJD 17	6.16			6.16	
IAJD 18	6.14	6.41	6.40	6.32	0.15
IAJD 46	6.98			6.98	
Library 6					
IAJD No.	pK_a 1	pK_a 2	pK_a 3	Avg pK_a	Std Div
IAJD 38	6.43	6.32		6.38	0.07
IAJD 39	6.20	6.21	6.30	6.24	0.06
IAJD 40	6.48	6.23		6.36	0.18
IAJD 41	6.38	6.34		6.36	0.03
IAJD 42	6.06			6.06	
IAJD 49	6.39			6.39	
IAJD 26	6.24			6.24	
IAJD 27	6.14	6.38	6.26	6.26	0.12
IAJD 28	6.09	6.30	6.37	6.25	0.15
IAJD 50	6.47			6.47	
IAJD 29	6.10			6.10	
IAJD 47	6.63	6.54		6.59	0.06

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IAJD 51	5.90		5.90	
IAJD 52	4.91		4.91	
IAJD 43	6.48	6.42	6.45	0.04
IAJD 48	6.68		6.68	
IAJD 32	6.11	6.32	6.22	0.15

Example 2: Targeted Delivery of mRNA with One-Component IAJDs

One aspect of the invention relates in part to a one-component multifunctional sequence-defined IAJD delivery system that co-assembles with mRNA by simple injection into DNPs (Figure 1). IAJDs described in the preceding examples require a 14-step synthesis for their preparation. Earlier IAJDs use a combination of a hydrophobic dendron conjugated to a hydrophilic dendron based on methoxytriethylene glycol fragments (Z) enclosing an ionizable amine incorporated in a sequence-defined arrangement. In order to reduce the number of synthetic steps employed in the construction of IAJDs, the methoxytriethylene glycol fragments (Z) attached to the branching point of the hydrophilic fragment of the JD were eliminated since these fragments required multiple protection-deprotection steps. In the IAJDs described in the following examples only its ionizable amine is maintained, which when protonated, may provide the dual hydrophilic and binding component of the IAJD to mRNA (Figure 47 through Figure 49). This transformation reduced the number of reaction steps significantly. The synthesis is shown in Scheme 1.

Scheme 1. Synthesis of IAJD with Benzyl Esters and Aliphatic Chains. Reagents and Conditions: i) $C_nH_{2n+1}Br$, K_2CO_3, DMF, 80 °C, 5 h; ii) $LiAlH_4$, THF, 0–23 °C, 1 h; iii) 4-Bromobutyric acid, $SOCl_2$, DMF (cat.), DCM, 23 °C, 1 h, then DMAP, NEt_3, DCM, 0–23 °C, 2 h; iv) K_2CO_3, MeCN, 95 °C, 3 h; v) $C_{12}H_{25}Br$, NaOH, water, 80 °C, 5 h.

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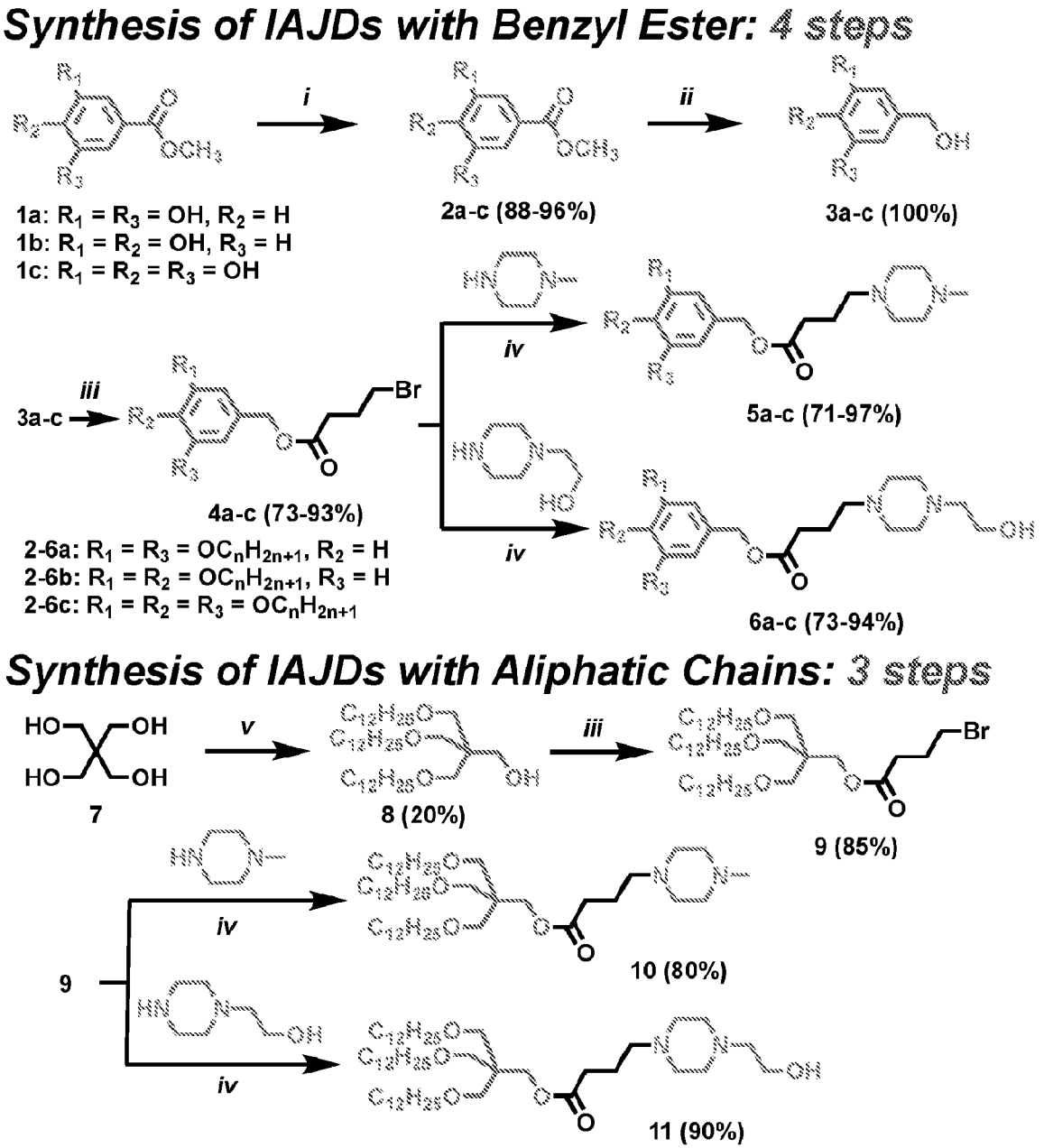

The materials and methods employed in these experiments are now described.

Materials

3,5-Dihydroxybenzoic acid (Acros, 97%), 3,4-dihydroxybenzoic acid (Acros, 97%), 3,4,5-trihydroxybenzoic acid (gallic acid, Chem Impex, anhydrous, ACS grade), 1-bromooctane (Aldrich, 99%), 1-bromononane (Lancaster, 99%), 1-bromodecane (Acros, 98%), 1-bromoundecane (Aldrich, 99%), 1-bromododecane (Alfa Aesar, 99%), 1-bromotetradecane (Acros, 98%), 1-bromohexadecane (TCI, 96%), 1-bromooctadecane (Acros, 96%), (*rac*)-3-(bromomethyl)heptane

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(2-ethylhexyl bromide, Aldrich, 95%), (*rac*)-1-bromo-3,7-dimethyloctane (TCI, 93+%), $LiAlH_4$ (TCI, 95%), palladium on activated carbon catalyst (Spectrum, 10 wt% loading), pentaerythritol (Aldrich, 98%), 4-(dimethylamino)butyric acid hydrochloride (Alfa Aesar, 98%), 3-bromopropanoic acid (Aldrich, 97%), 4-bromobutyric acid (Acros, 98%), 5-bromovaleric acid (Aldrich, 97%), 1-methylpiperazine (Alfa Aesar, 98%), 1-(2-hydroxyethyl)piperazine (Acros, 99%), thionyl chloride (Alfa Aesar, 99+%), triethylamine (TCI, 99%), other reagents and solvents for chemical synthesis were obtained from commercial sources and were used as received. 4-(Dimethylamino)pyridinium 4-toluenesulfonate (DPTS) was prepared according to a literature procedure.[1] CH_2Cl_2 (DCM) was dried over CaH_2 and freshly distilled before use. Acetate buffer (10 mM) was prepared with the following composition: sodium acetate (0.0023 M) and acetic acid (0.0077 M) in ultra-pure water. Final pH was adjusted with 0.1 M NaOH or 0.1 M HCl solution. Nucleoside-modified mRNA encoding firefly luciferase (Luc-mRNA) was produced as previously described. Human embryonic kidney (HEK) 293T cells (American Type Culture Collection) were cultured in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% inactivated fetal bovine serum (FBS) (Gemini Bio-Products), 2 mM L–glutamine and 100 U/mL penicillin/streptomycin (Life Technologies). OptiMEM (Gibco), DPBS (Corning), UltraPure DNase/RNase-Free Distilled Water (Invitrogen), Trypan Blue (Sigma-Aldrich), Trypsin-EDTA (0.25%), (Gibco), Cell Culture Lysis 5X Reagent (Promega), Luciferase Assay System (Promega) and D-luciferin sodium salt (Regis Technologies) were used as received.

Methods and Techniques

The purity and structural identity of intermediate compounds and final products and was determined by a combination of techniques including thin-layer chromatography (TLC), ^1H and ^{13}C NMR, high-pressure liquid chromatography (HPLC), and matrix assisted laser desorption ionization-time of flight (MALDI-TOF) mass spectrometry as described above.

Evolution of the reaction was monitored by TLC using silica gel 60 F_{254} precoated plates (E. Merck) and individual compounds were visualized by UV light with a wavelength of 254 nm or by staining of the TLC plate with iodine vapor. Purification by flash column chromatography was performed using flash silica gel from Silicycle (60 Å, 40–63 μm) with the indicated eluent.

Dynamic Light Scattering (DLS) for the sizes and polydispersities of DNPs was performed on a Malvern Instruments particle sizer (Zetasizer Nano S, Malvern Instruments, UK) equipped with 4 mW He–Ne laser 633 nm and avalanche photodiode positioned at 175° to the beam and temperature-controlled cuvette holder. Instrument parameters were determined automatically along

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with measurement times. Sample solution (0.4 mL) was placed in a semi-micro cuvette (1.6 mL, polystyrene, 10 × 10 × 45 mm, manufactured by Greiner Bio-One) and the measurements were performed at 23 °C.

pK_a measurements of individual IAJD Molecules was performed as follows. IAJD Molecules were first dissolved in ethanol (Sat. with NaCl) at a concentration of 1.5 mg/mL and the solution volume was 3 mL. Then 0.1 M HCl aqueous solution was added to the above ethanol solution in increments of 7.5 μL, with the resulting pH measured using an Thermo Scientific Orion Star A121 meter with Thermo Scientific Orion 8220BNWP pH probe. pK_a was calculated using the half equivalence point titration.

Formulation of DNPs obtained by co-assembly of IAJDs and Luc-mRNA was performed as follows. IAJDs were dissolved in ethanol with an initial concentration of 80 mg/mL. Nucleoside-modified mRNA encoding firefly luciferase (Luc-mRNA) was dissolved in UltraPure DNase/RNase-free distilled water with an initial concentration of 4.0 mg/mL. 12.5 μL of Luc-mRNA solution was placed into a clean RNAs free eppendorf (1.5 mL) and 463 μL acetate buffer (10 mM, pH 4.0) was added. Then, 25 μL of IAJD in the ethanol stock solution was rapidly injected into the Luc-mRNA solution in acetate buffer followed by vortex for 5 seconds.

Luminescence characterization for *in vitro* transfection experiments was performed by determining the luminescence intensity for *in vitro* Luc-mRNA transfection experiments using a MiniLumat LB 9506 luminometer (Berthold/EG&G; Wallac).

Luminescence characterization for *in vivo* transfection experiments employed bioluminescence imaging with an IVIS Spectrum imaging system (PerkinElmer, Waltham, MA). Mice were anesthetized with 3% of isoflurane (Piramal Healthcare Limited) and intraperitoneally (i.p.) administered with D-luciferin (Regis Technologies) at a dose of 150 mg/kg of body weight. Ten minutes post administration of D-luciferin, mice were placed on the imaging platform while being maintained on isoflurane via a nose cone and imaged using an exposure time of 60 seconds. Bioluminescence values were quantified by measuring photon flux (photons/second, p/s) in the region of interest (ROI) on mice where bioluminescence signal emanated using the Living Image Software (PerkinElmer). To quantify luminescent flux, an oval ROI was placed over each organ of interest and analyzed.

For *in vitro* Luc-mRNA transfection screening experiments, human embryonic kidney 293 cells (HEK 293) were seeded into 96-well plates (20,000 cells/well/200 μL) and cultured for 18-20 hours at 37 °C, 5% CO_2 in complete Dulbecco's modified Eagle's medium (DMEM) supplemented

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with 2 mM L–glutamine (Life Technologies), 100 U/mL penicillin/streptomycin (Invitrogen) and 10% fetal bovine serum (FBS, Gemini Bio-Products). Cells were transfected with DNPs with encapsulated naked nucleoside-modified mRNA encoding firefly luciferase (Luc-mRNA) at constant concentration of Luc-mRNA, 125 ng per well. Luc-mRNA complexed with *Trans*IT (TransIT®-mRNA Transfection Kit, Mirus Bio) or encapsulated in FDA approved MC3-based LNPs were used as positive controls for cell transfection, at the concentration of Luc-mRNA of 125 ng per well, the same as for tested DNPs. After transfection cells were further cultured at 37 °C, 5% CO_2 for 18-20 hours, then medium was aspirated and cells were lysed with luciferase cell culture lysis reagent (30 μL/well) (Promega, Madison, WI). For the determination of the luciferase enzymatic activity as luminescence, 2.5 μL of the lysed cells was mixed with 10 μL of firefly luciferase assay substrate ((Luciferase assay system, Promega) and luminescence was analyzed by MiniLumat LB9506 luminometer. Transfections were performed in triplicate.

In vivo mRNA Delivery in Mice with DNPs employed female or male BALB/c mice (6-8 weeks old, from Charles River Laboratories) anesthetized with isoflurane (Piramal Healthcare Limited) and injected *via* retro-orbital sinus with 100 μL of buffer solution containing DNPs with encapsulated 10 μg of Luc-mRNA. At 4-7 hours post injection, mice were i.p. injected with D-Luciferin (150 mg/kg of body weight, Regis Technologies) and imaged on a PerkinElmer IVIS Spectrum CT system (PerkinElmer, Waltham, MA). Tissue luminescence signal was measured on the IVIS imaging system using an exposure time of 60 seconds using medium binning (binning = 8) to ensure that the signal obtained was within operative detection range. For IVIS imaging of the organs, mice were sacrificed and heart, lungs, liver, and spleen were immediately collected. Bioluminescence imaging was performed as described above. Image analysis was conducted with the Living Image software (PerkinElmer). Bioluminescence values were quantified by measuring photon flux (photons/s) in the region of interest using Living Image software.

Synthesis of IAJDs with ester bonds in hydrophobic alkyl chains was performed as shown in Scheme 2.

Scheme 2.

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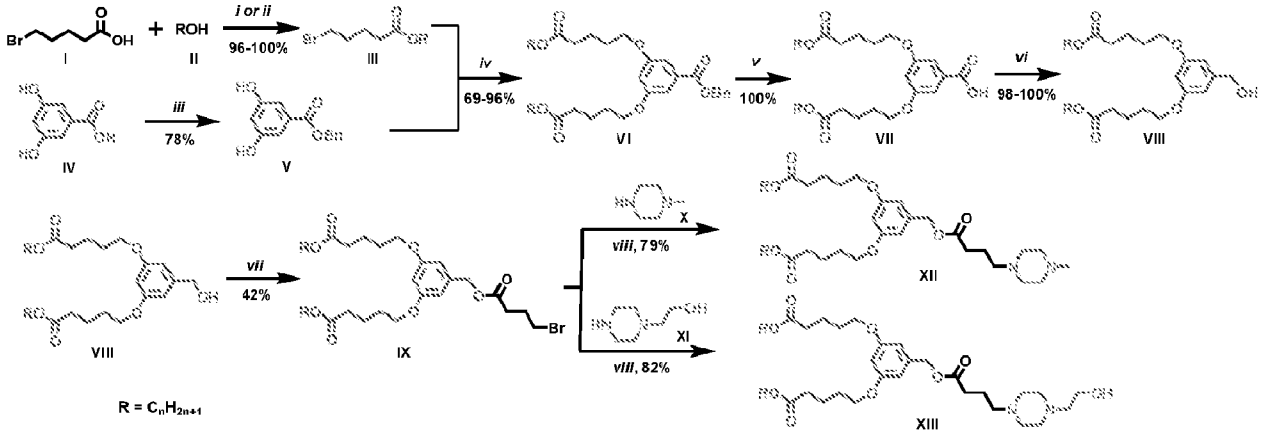
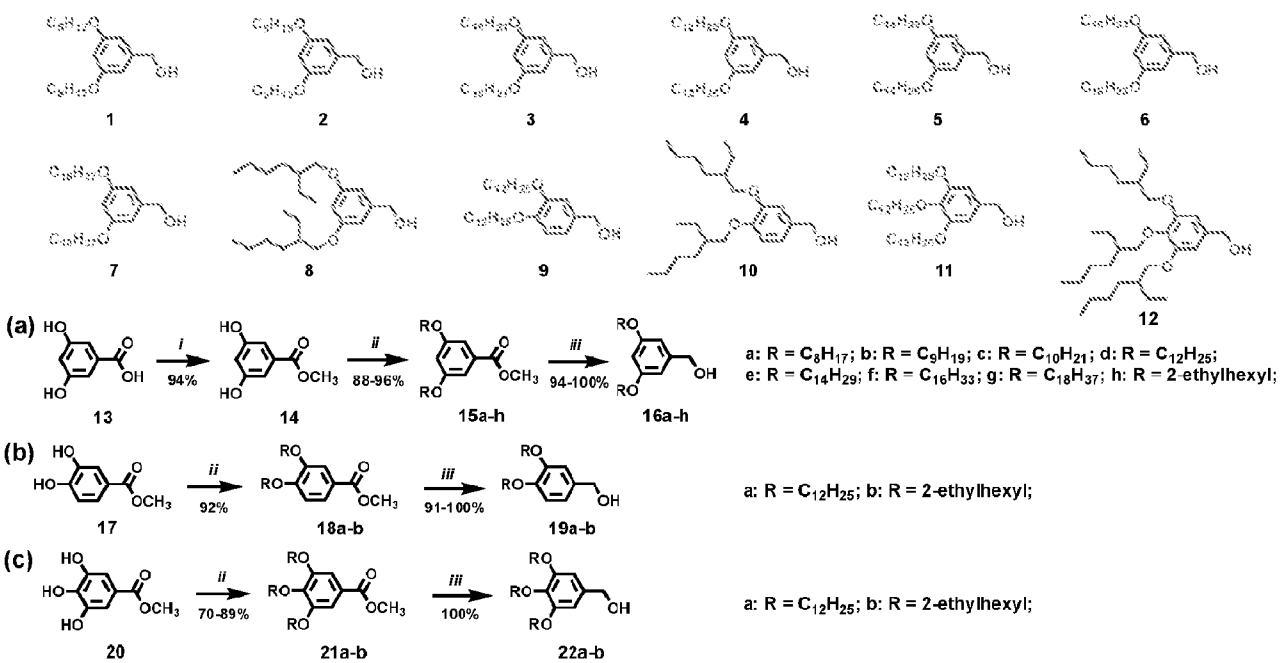
General Synthesis of IAJDs Containing Alkyl Chains with Ester Bonds: 7 steps



Reagents and Conditions: (*i*) $SOCl_2$, DMF (cat.), DCM, 23 °C, 1 h, then DMAP, NEt_3, DCM, 0–23 °C, 2 h; (*ii*) DCC, DPTS, DCM, 23 °C, 12 h; (*iii*) BnCl, $NaHCO_3$, DMF, 40 °C, 12 h; (*iv*) K_2CO_3, DMF, 80 °C, 5 h; (*v*) H_2, Pd/C, DCM, MeOH, 12 h; (*vi*) $NaBH_4$, I_2, THF, 0–23 °C, 2 h; (*vii*) 4-Bromobutyric acid, $SOCl_2$, DMF (cat.), DCM, 23 °C, 1 h, then DMAP, NEt_3, DCM, 0–23 °C, 2 h; (*viii*) K_2CO_3, MeCN, 95 °C, 3 h.

Synthesis of Hydrophobic Building Blocks of IAJDs was performed as shown in Scheme 3.

Scheme 3.



Reagents and conditions: (*i*) H_2SO_4 (cat.), MeOH, reflux, 2 h; (*ii*) RBr, K_2CO_3, DMF, 120 °C, 2 h; (*iii*) $LiAlH_4$, THF, 0–23 °C, 1 h.

General Synthetic Procedure for Compounds 15a-h, 18a-b, and 21a-b: Compounds 15a-h,

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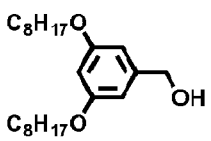
18a-b, and 21a-b were synthesized according to a procedure elaborated and optimized. Generally, compound 14/17/20 (1 equiv) and K_2CO_3 (3 equiv for compound 14/17, 4.5 equiv for compound 20) were stirred in dry DMF. RBr (2.2 equiv for compound 14/17, 3.3 equiv for compound 20) was added, and the mixture was stirred at 120 °C under N_2 atmosphere for 2 h. The reaction mixture was cooled to 23 °C. For compounds 15a-g, 18a and 21a, the reaction mixture was poured into ice/water and the white precipitates were filtered and collected. Then the precipitates were purified by recrystallization from acetone to give the title compound as a white solid. For compounds 15h, 18b and 21b, water was added to the reaction mixture, which was then extracted by DCM for three times. The organic phase was collected and dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and further purified by column chromatography on silica gel with a mobile phase of hexane/EtOAc = 20/1 (v/v) to afford the title compound as a light-yellow oil.

General Synthetic Procedure for Compounds 16a-h (1-8), 19a-b (9,10), and 22a-b (11,12): Compounds 16a-h, 19a-b, and 22a-b were synthesized according to a procedure previously reported by us. Generally, compounds 15a-h, 18a-b, and 21a-b (1 equiv) were dissolved in dry THF, which was added dropwise to a slurry of $LiAlH_4$ (1.2 equiv) in dry THF at 0 °C under N_2 atmosphere. The resulted mixture was stirred at 23 °C for 1 h. The reaction was quenched by the successive addition of water, 15% NaOH aqueous solution and water. Then the mixture was filtered and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a white solid or light-yellow oil. The synthesis and characterizations of compounds 16d (4), 19a (9) and 22a (11) were reported in the literature.

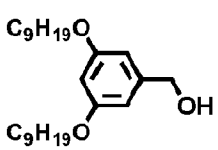
(3,5-Bis(octyloxy)phenyl)methanol (1).



1

From compound 15a (4.50 g, 11.46 mmol) and $LiAlH_4$ (0.52 g, 13.75 mmol), compound 1 was obtained as a colorless oil (4.10 g, 98%).

(3,5-Bis(nonyloxy)phenyl)methanol (2).



2

From compound 15b (1.16 g, 2.75 mmol) and $LiAlH_4$ (0.13 g, 3.30 mmol), compound 2 was obtained

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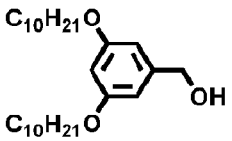
as a colorless oil (1.08 g, 100%).

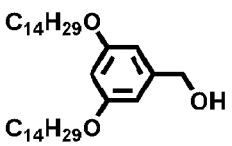
(3,5-bis(decyloxy)phenyl)methanol (3).



3

From compound 15c (3.40 g, 8.08 mmol) and $LiAlH_4$ (0.37 g, 9.70 mmol), compound 3 was obtained as a white solid (3.21 g, 94%).

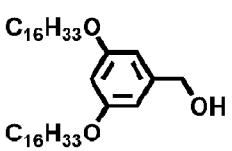
(3,5-Bis(tetradecyloxy)phenyl)methanol (5).



5

From compound 15e (0.50 g, 0.89 mmol) and $LiAlH_4$ (51 mg, 1.34 mmol), compound 5 was obtained as a white solid (0.48 g, 100%).

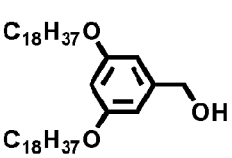
(3,5-Bis(hexadecyloxy)phenyl)methanol (6).



6

From compound 15f (0.50 g, 0.81 mmol) and $LiAlH_4$ (46 mg, 1.22 mmol), compound 6 was obtained as a white solid (0.48 g, 100%).

(3,5-Bis(octadecyloxy)phenyl)methanol (7).



7

From compound 15g (0.60 g, 0.89 mmol) and $LiAlH_4$ (68 mg, 1.78 mmol), compound 7 was obtained as a white solid (0.55 g, 96%).

(3,5-Bis((2-ethylhexyl)oxy)phenyl)methanol (8).

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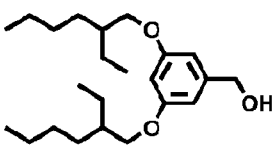
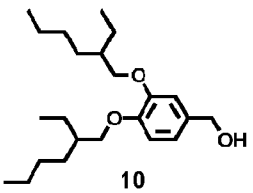


8

From compound 15h (4.10 g, 10.44 mmol) and $LiAlH_4$ (0.48 g, 12.53 mmol), compound 8 was obtained as a light-yellow oil (3.64 g, 96%).

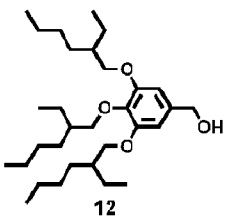
(3,4-Bis((2-ethylhexyl)oxy)phenyl)methanol (10).



10

From compound 18b (2.77 g, 7.10 mmol) and $LiAlH_4$ (0.32 g, 8.50 mmol), compound 10 was obtained as a colorless oil (2.36 g, 91%).

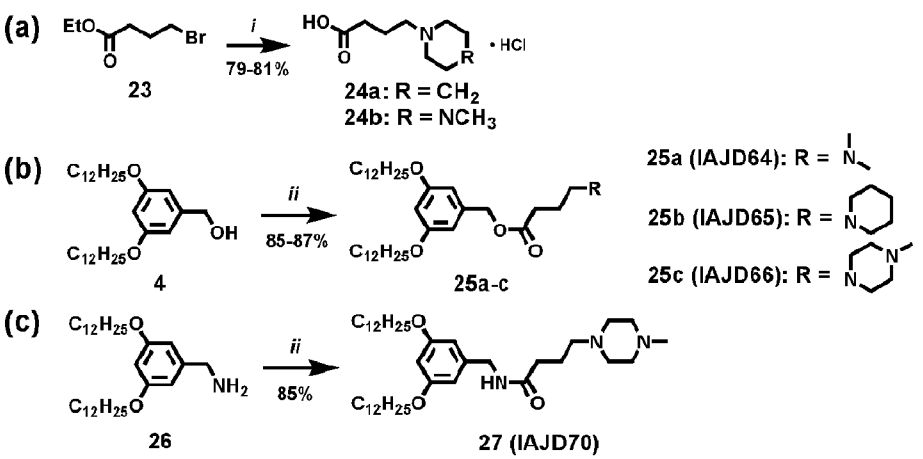
(3,4,5-Tris((2-ethylhexyl)oxy)phenyl)methanol (12).



12

From compound 21b (0.60 g, 1.15 mmol) and $LiAlH_4$ (0.12 g, 3.16 mmol), compound 12 was obtained as a colorless oil (0.57 g, 100%).

Synthesis of IAJDs 64-66 and 70 is shown in Scheme 4.



Reagents and conditions: (*i*) (1) piperidine/1-methylpiperazine, K_2CO_3, MeCN, reflux, 3 h; (2) conc.

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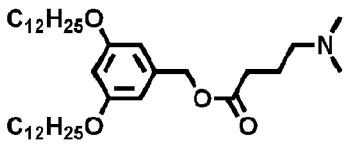
HCl/water, reflux, 3 h; (*ii*) 4-(Dimethylamino)butyric acid hydrochloride/4-(piperidin-1-yl)butanoic acid hydrochloride/4-(4-methylpiperazin-1-yl)butanoic acid hydrochloride, DCC, DCM, 23 °C, 12 h.

The synthesis and characterizations of compounds 24a and 24b are available in the literature.

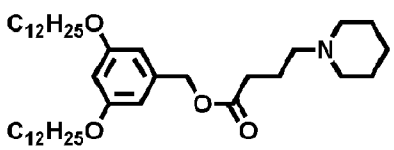
3,5-Bis(dodecyloxy)benzyl 4-(dimethylamino)butanoate (25a, IAJD64).



25a

Compound 4 (0.50 g, 1.05 mmol, 1 equiv) and 4-(dimethylamino)butyric acid hydrochloride (0.18 g, 1.05 mmol, 1 equiv) were dissolved in 6 mL dry DCM. *N,N'*-Dicyclohexylcarbodiimide (DCC, 0.43 g, 2.10 mmol, 2 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. Afterwards, urea was removed by filtration, and washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 40/1, 20/1 and 15/1 as the eluent. Then the product was dissolved in DCM (30 mL), which was washed by $NaHCO_3$ solution (2%, 30 mL). The aqueous phase was extracted by DCM (30 mL) for another two times. The organic phase was combined and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.54 g, 87%).

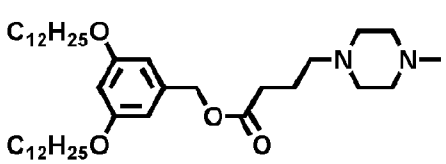
3,5-Bis(dodecyloxy)benzyl 4-(piperidin-1-yl)butanoate (25b, IAJD65).



25b

Compound 25b was synthesized from compound 4 (0.50 g, 1.05 mmol, 1 equiv), 4-(piperidin-1-yl)butanoic acid hydrochloride (24a, 0.22 g, 1.05 mmol, 1 equiv) and DCC (0.43 g, 2.10 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound 25a. The title compound was obtained as a colorless oil (0.57 g, 86%).

3,5-Bis(dodecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (25c, IAJD66).



25c

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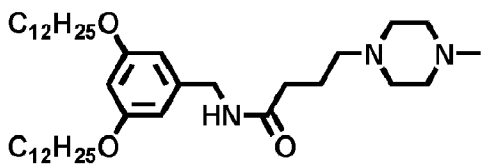
Compound 25c was synthesized from compound 4 (0.20 g, 0.42 mmol, 1 equiv), 4-(4-methylpiperazin-1-yl)butanoic acid hydrochloride (24b, 94 mg, 0.42 mmol, 1 equiv) and DCC (0.17 g, 0.84 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound 25a. The title compound was obtained as a colorless oil (0.23 g, 85%).

N-(3,5-bis(dodecyloxy)benzyl)-4-(4-methylpiperazin-1-yl)butanamide (27, IAJD70).



27

The synthesis and characterizations of (3,5-bis(dodecyloxy)phenyl)methanamine (26) were reported in the literature.[7] Compound 26 (0.20 g, 0.42 mmol, 1 equiv) and 4-(4-methylpiperazin-1-yl)butanoic acid hydrochloride (24b, 94 mg, 0.42 mmol, 1 equiv) were dissolved in 6 mL dry DCM. NEt_3 (3 drops) was added. DCC (0.17 g, 0.84 mmol, 2 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. Afterwards, urea was removed by filtration, and washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 30/1, 10/1, 8/1 and 8/1 with 0.1% TEA as the eluent. Then the product was dissolved in DCM (30 mL), which was washed by $NaHCO_3$ solution (2%, 30 mL). The aqueous phase was extracted by DCM (30 mL) for another two times. The organic phase was combined and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a white solid (0.23 g, 85%).

Synthesis of IAJDs 71 and 74-79 is shown in Scheme 5.

Scheme 5.

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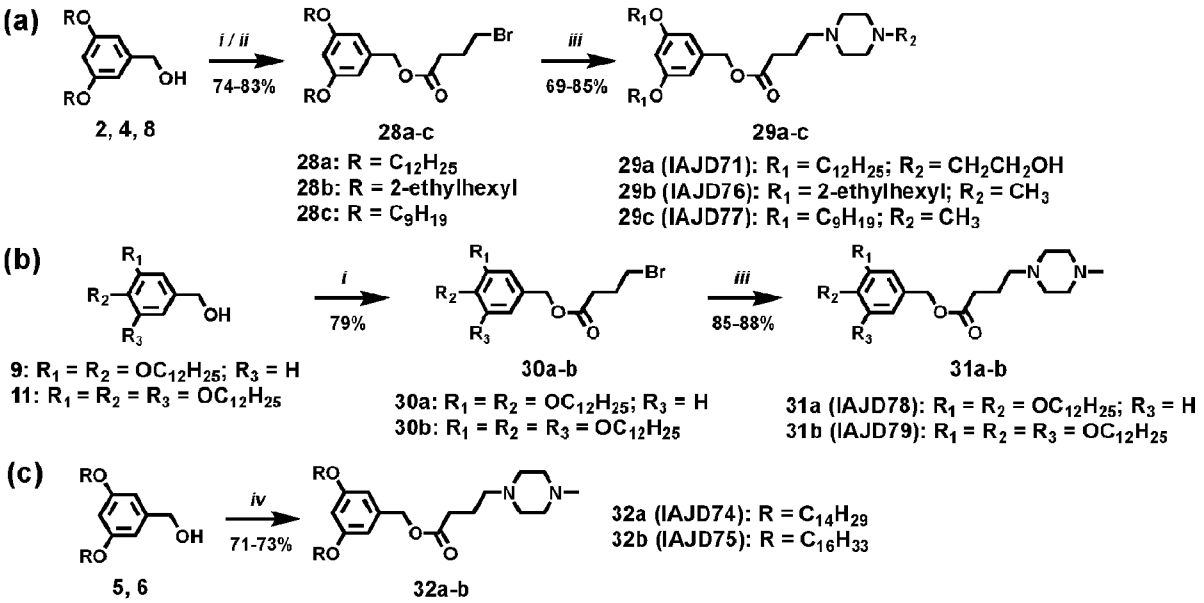
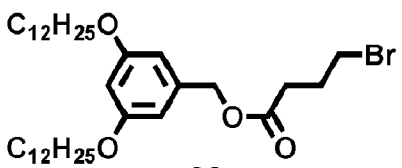


Reagents and conditions: (*i*) EDC.HCl, DMAP, DCM, 23 °C, 12 h; (*ii*) 4-bromobutyric acid, $SOCl_2$, DMF (cat.), DCM, 23 °C, 1 h, then DMAP, NEt_3, DCM, 0–23 °C, 2 h; (*iii*) 1-methylpiperazine/1-(2-hydroxyethyl)piperazine, K_2CO_3, MeCN, 95 °C, 3 h. (*iv*) 4-(4-methylpiperazin-1-yl)butanoic acid hydrochloride, DCC, DCM, 23 °C, 12 h.

3,5-Bis(dodecyloxy)benzyl 4-bromobutanoate (28a).



28a

Compound 4 (0.60 g, 1.26 mmol, 1 equiv), 4-bromobutyric acid (0.21 g, 1.26 mmol, 1 equiv), EDC.HCl (0.24 g, 1.26 mmol, 1 equiv) and DMAP (46 mg, 0.38 mmol, 0.3 equiv) were dissolved in dry DCM (6 mL). The reaction mixture was stirred at 23 °C for 12 h. Afterwards, brine (30 mL) was added, and the mixture was extracted by DCM (30 mL) for three times. The organic phase was collected, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the eluent to give the title compound as a colorless oil (0.62 g, 79%).

3,5-Bis((2-ethylhexyl)oxy)benzyl 4-bromobutanoate (28b).

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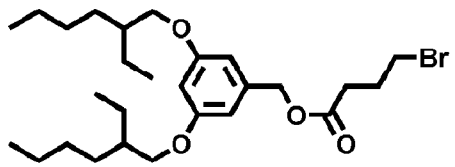
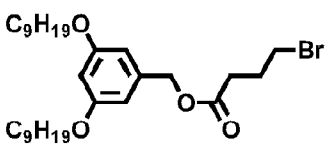


28b

4-Bromobutyric acid (0.11 g, 0.63 mmol, 1.05 equiv) was dissolved in 5 mL anhydrous DCM and 1 drop of DMF was added. Thionyl chloride ($SOCl_2$, 0.14 g, 1.20 mmol, 2 equiv) was added and the mixture was stirred at 23 °C for 1 h. Afterwards, DCM and excess $SOCl_2$ was removed under reduced pressure to give the 4-bromobutanoyl chloride. Compound 8 (0.22 g, 0.60 mmol, 1.0 equiv), NEt_3 (67 mg, 0.66 mmol, 1.1 equiv) and DMAP (14 mg, 0.12 mmol, 0.2 equiv) were dissolved in 6 mL anhydrous DCM. To this mixture was added the DCM solution (3 mL) of above 4-bromobutanoyl chloride dropwise at 0 °C. The mixture was allowed to warm to 23 °C with stirring during a 2 h period. Afterwards, water (30 mL) was added, and the mixture was extracted by DCM (30 mL) for three times. The organic phase was collected, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the eluent to give the title compound as a colorless oil (0.26 g, 83%).

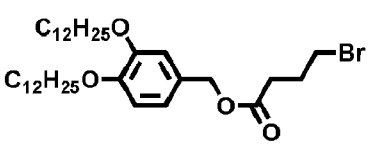
3,5-Bis(nonyloxy)benzyl 4-bromobutanoate (28c).



28c

Compound 28c was synthesized from compound 2 (0.50 g, 1.30 mmol), 4-bromobutyric acid (0.22 g, 1.30 mmol) and NEt_3 (0.14 g, 1.43 mmol) following a procedure similar to that used for the synthesis of compound 28b. The title compound was obtained as a light-yellow oil (0.51 g, 74%).

3,4-Bis(dodecyloxy)benzyl 4-bromobutanoate (30a).



30a

Compound 30a was synthesized from compound 9 (0.50 g, 1.05 mmol), 4-bromobutyric acid (0.18 g, 1.05 mmol) and NEt_3 (0.12 g, 1.16 mmol) following a procedure similar to that used for the synthesis of compound 28b. The title compound was obtained as a white solid (0.52 g, 79%).

3,4,5-Tris(dodecyloxy)benzyl 4-bromobutanoate (30b).

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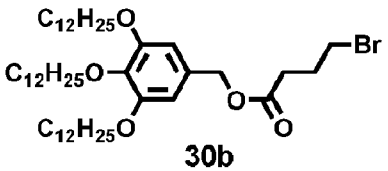
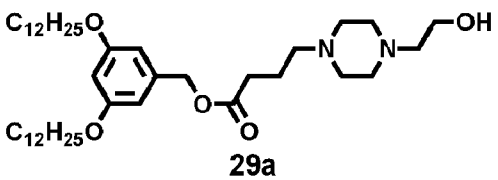


30b

Compound 30b was synthesized from compound 11 (0.50 g, 0.76 mmol), 4-bromobutyric acid (0.13 g, 0.76 mmol) and NEt_3 (0.12 g, 0.84 mmol) following a procedure similar to that used for the synthesis of compound 28b. The title compound was obtained as a colorless oil (0.48 g, 79%).

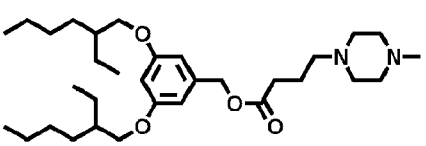
3,5-Bis(dodecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (29a, IAJD71).



29a

Compound 28a (0.24 g, 0.38 mmol, 1.0 equiv), 1-(2-hydroxyethyl)piperazine (52 mg, 0.40 mmol, 1.05 equiv), K_2CO_3 (55 mg, 0.40 mmol, 1.05 equiv) were stirred in MeCN (20 mL). The mixture was heated at reflux (95 °C) for 3 h. The reaction mixture was cooled to 23 °C and MeCN was removed under reduced pressure. Then water (20 mL) was added, and the mixture was extracted by DCM (20 mL × 3). The organic phase was collected, dried over anhydrous $MgSO_4$, filtered and dried to give the crude product. The crude product was purified by column chromatography (SiO_2) with DCM/MeOH = 30/1 and 15/1 as the eluent. Then the obtained product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was combined and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.22 g, 85%).

3,5-Bis((2-ethylhexyl)oxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (29b, IAJD76).



29b

Compound 29b was synthesized from compound 28b (0.26 g, 0.50 mmol), 1-methylpiperazine (56 mg, 0.56 mmol) and K_2CO_3 (0.10 g, 0.75 mmol) following a procedure similar to that used for the synthesis of compound 29a. The title compound was obtained as a colorless oil (0.19 g, 69%).

3,5-Bis(nonyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (29c, IAJD77).

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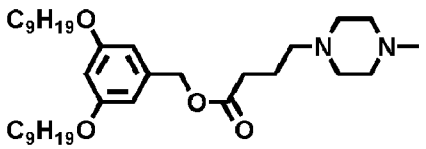
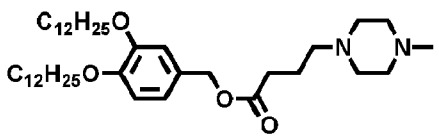


29c

Compound 29c was synthesized from compound 28c (0.35 g, 0.64 mmol), 1-methylpiperazine (70 mg, 0.70 mmol) and K_2CO_3 (98 mg, 0.70 mmol) following a procedure similar to that used for the synthesis of compound 29a. The title compound was obtained as a light-yellow oil (0.26 g, 72%).

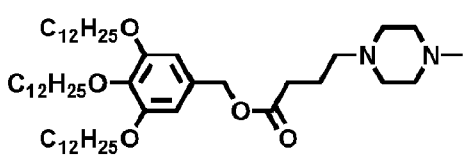
3,4-Bis(dodecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (31a, IAJD78).



31a

Compound 31a was synthesized from compound 30a (0.40 g, 0.64 mmol), 1-methylpiperazine (67 mg, 0.67 mmol) and K_2CO_3 (93 mg, 0.67 mmol) following a procedure similar to that used for the synthesis of compound 29a. The title compound was obtained as a viscous white solid (0.36 g, 88%)..

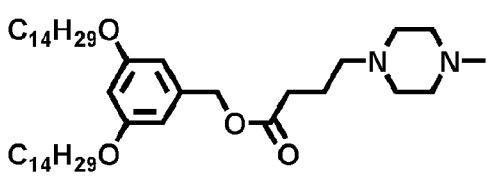
3,4,5-Tris(dodecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (31b, IAJD79).



31b

Compound 31b was synthesized from compound 30b (0.47 g, 0.58 mmol), 1-methylpiperazine (61 mg, 0.61 mmol) and K_2CO_3 (84 mg, 0.61 mmol) following a procedure similar to that used for the synthesis of compound 29a. The title compound was obtained as a light-yellow oil (0.41 g, 85%).

3,5-Bis(tetradecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (32a, IAJD74).



32a

Compound 5 (0.24 g, 0.45 mmol, 1 equiv) and 4-(4-methylpiperazin-1-yl)butanoic acid hydrochloride (24b, 0.10 g, 0.45 mmol, 1 equiv) were dissolved in 6 mL dry DCM. NEt_3 (3 drops)

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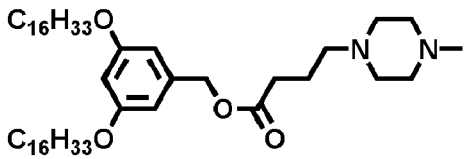
was added. DCC (0.19 g, 0.90 mmol, 2 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. Afterwards, urea was removed by filtration, and washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 40/1, 20/1 and 15/1 as the eluent. Then the product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was combined and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.22 g, 71%).

3,5-Bis(hexadecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (32b, IAJD75).

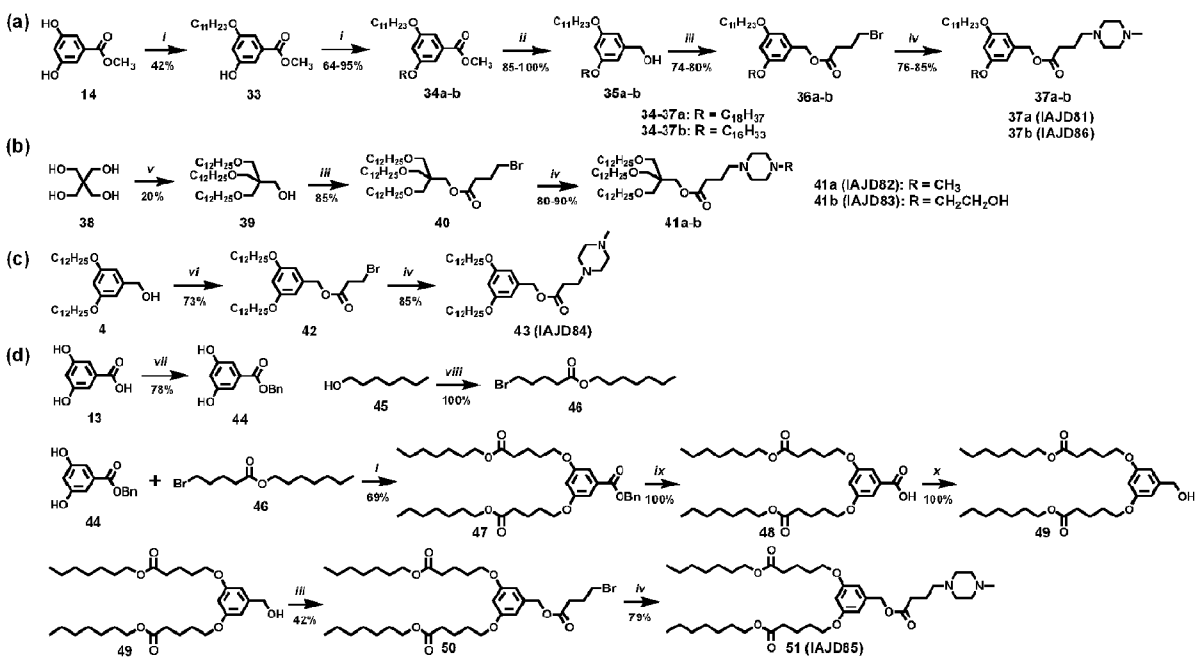


32b

Compound 32b was synthesized from compound 6 (0.27 g, 0.45 mmol, 1 equiv), 4-(4-methylpiperazin-1-yl)butanoic acid hydrochloride (24b, 0.10 g, 0.45 mmol, 1 equiv) and DCC (0.19 g, 0.90 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound 32a. The title compound was obtained as a light-yellow oil (0.25 g, 73%).

Synthesis of IAJDs 81-86 was performed according to Scheme 6.

Scheme 6.



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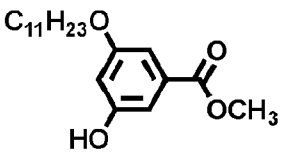
Reagents and conditions: (*i*) RBr, K_2CO_3, DMF, 60 °C, 6 h; (*ii*) $LiAlH_4$, THF, 0–23 °C, 1 h; (*iii*) 4-bromobutyric acid, $SOCl_2$, DMF (cat.), DCM, 23 °C, 1 h, then DMAP, NEt_3, DCM, 0–23 °C, 2 h; (*iv*) 1-methylpiperazine/1-(2-hydroxyethyl)piperazine, K_2CO_3, MeCN, 95 °C, 3 h; (*v*) $C_{12}H_{25}Br$, 50% NaOH, TBABr (cat.), 80 °C, 5 h; (*vi*) 3-bromopropanoic acid, $SOCl_2$, DMF (cat.), DCM, 23 °C, 1 h, then NEt_3, DCM, 0–23 °C, 2 h; (*vii*) BnCl, $NaHCO_3$, DMF, 40 °C, 12 h; (*viii*) 5-bromovaleric acid, $SOCl_2$, DMF (cat.), DCM, 23 °C, 1 h, then DMAP, NEt_3, DCM, 0–23 °C, 2 h; (*ix*) H_2, Pd/C, DCM, MeOH, 12 h; (*x*) $NaBH_4$, I_2, THF, 0–23 °C, 2 h.

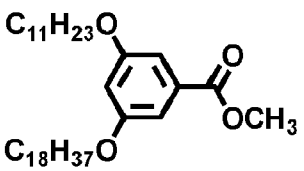
Methyl 3-hydroxy-5-(undecyloxy)benzoate (33).



33

Methyl 3,5-dihydroxybenzoate (14, 5.00 g, 29.7 mmol, 1.2 equiv), 1-bromoundecane (5.83 g, 24.8 mmol, 1 equiv), and K_2CO_3 (3.42 g, 24.8 mmol, 1 equiv) were stirred in DMF (50 mL). The mixture was stirred at 60 °C under N_2 atmosphere for 6 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 10/1 as the eluent to give the title compound as a white solid (3.31g, 42%).

Methyl 3-(octadecyloxy)-5-(undecyloxy)benzoate (34a).



34a

Compound 34a was synthesized from compound 33 (1.0 g, 3.10 mmol, 1 equiv), 1-bromooctadecane (1.24 g, 3.72 mmol, 1.2 equiv), and K_2CO_3 (1.28 g, 9.30 mmol, 3 equiv) following a procedure similar to that used for the synthesis of compound 33. The title compound was obtained as a white solid (1.69 g, 95%).

Methyl 3-(hexadecyloxy)-5-(undecyloxy)benzoate (34b).

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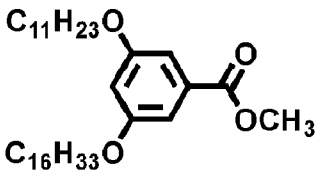
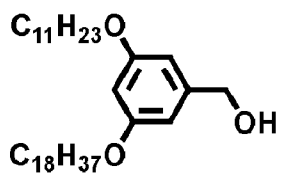


34b

Compound 34b was synthesized from compound 33 (0.50 g, 1.55 mmol, 1 equiv), 1-bromohexadecane (0.52 g, 1.71 mmol, 1.1 equiv), and K_2CO_3 (0.64 g, 4.65 mmol, 3 equiv) following a procedure similar to that used for the synthesis of compound 33. The title compound was obtained as a white solid (0.54 g, 64%).

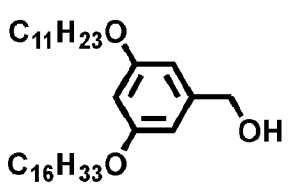
(3-(Octadecyloxy)-5-(undecyloxy)phenyl)methanol (35a).



35a

Compound 34a (1.0 g, 1.74 mmol, 1 equiv) was dissolved in 5 mL dry THF, which was added dropwise to a slurry of $LiAlH_4$ (0.10 g, 2.61 mmol, 1.5 equiv) in dry THF (5 mL) at 0 °C under N_2 atmosphere. The resulted mixture was stirred at 23 °C for 1 h. The reaction was quenched by the successive addition of water (0.3 mL), 15% NaOH aqueous solution (0.3 mL) and water (1.5 mL). Then the mixture was filtered to remove white precipitates and the filtrate was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a white solid (0.95 g, 100%).

(3-(Hexadecyloxy)-5-(undecyloxy)phenyl)methanol (35b).



35b

Compound 35b was synthesized from compound 34b (0.50 g, 0.91 mmol, 1 equiv) and $LiAlH_4$ (52 mg, 1.37 mmol, 1.5 equiv) following a procedure similar to that used for the synthesis of compound 35a. The title compound was obtained as a white solid (0.40 g, 85%).

3-(Octadecyloxy)-5-(undecyloxy)benzyl 4-bromobutanoate (36a).

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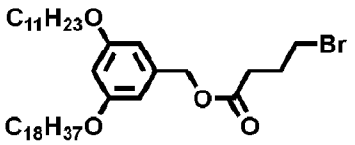
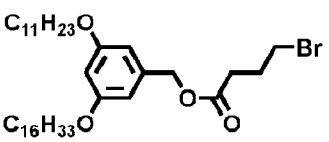


36a

4-Bromobutyric acid (0.10 g, 0.60 mmol, 1.0 equiv) was dissolved in 5 mL anhydrous DCM and 1 drop of DMF was added. $SOCl_2$ (0.14 g, 1.20 mmol, 2 equiv) was added and the mixture was stirred at 23 °C for 1 h. Afterwards, DCM and excess $SOCl_2$ was removed under reduced pressure to give the 4-bromobutanoyl chloride. Compound 35a (0.35 g, 0.60 mmol, 1 equiv) and NEt_3 (0.12 g, 1.20 mmol, 2 equiv) were dissolved in 6 mL anhydrous DCM. Then DMAP (15 mg, 0.12 mmol, 0.2 equiv) was added. To this mixture was added the DCM solution (3 mL) of above 4-bromobutanoyl chloride dropwise at 0 °C. The mixture was allowed to warm to 23 °C with stirring during a 2 h period. Afterwards, water (30 mL) was added, and the mixture was extracted by DCM (30 mL) for three times. The organic phase was collected, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the eluent to give the title compound as a colorless oil (0.31 g, 74%).

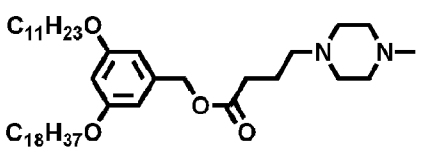
3-(Hexadecyloxy)-5-(undecyloxy)benzyl 4-bromobutanoate (36b).



36b

Compound 36b was synthesized from compound 35b (0.37 g, 0.60 mmol, 1 equiv), 4-bromobutyric acid (0.10 g, 0.60 mmol, 1.0 equiv), NEt_3 (0.12 g, 1.20 mmol, 2 equiv) and DMAP (15 mg, 0.12 mmol, 0.2 equiv) following a procedure similar to that used for the synthesis of compound 36a. The title compound was obtained as a colorless oil (0.33 g, 80%).

3-(Octadecyloxy)-5-(undecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (37a, IAJD81).



37a

Compound 36a (0.24 g, 0.35 mmol, 1 equiv), 1-methylpiperazine (37 mg, 0.36 mmol, 1.05 equiv), K_2CO_3 (53 mg, 0.38 mmol, 1.1 equiv) were stirred in MeCN (20 mL). The mixture was heated at reflux (95 °C) for 3 h. The reaction mixture was cooled to 23 °C and MeCN was removed under reduced pressure. Then water (20 mL) was added, and the mixture was extracted by DCM (20 mL

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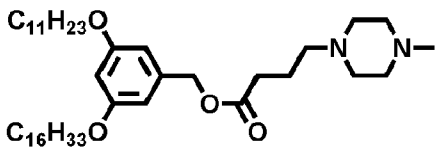
× 3). The organic phase was collected, dried over anhydrous $MgSO_4$, filtered and dried to give the crude product. The crude product was purified by column chromatography (SiO_2) with DCM/MeOH = 30/1 and 15/1 as the eluent. Then the obtained product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was combined and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.21 g, 85%).

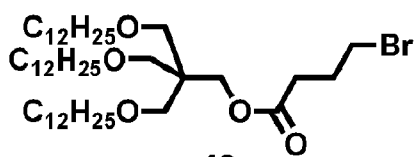
3-(Hexadecyloxy)-5-(undecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (37b, IAJD86).



37b

Compound 37b was synthesized from compound 36b (0.33 g, 0.49 mmol, 1 equiv), 1-methylpiperazine (52 mg, 0.52 mmol, 1.05 equiv) and K_2CO_3 (74 mg, 0.53 mmol, 1.1 equiv) following a procedure similar to that used for the synthesis of compound 37a. The title compound was obtained as a light-yellow oil (0.31 g, 76%).

3-(Dodecyloxy)-2,2-bis((dodecyloxy)methyl)propyl 4-bromobutanoate (40).



40

The synthesis and characterizations of 3-(dodecyloxy)-2,2-bis((dodecyloxy)methyl)propan-1-ol (39) were reported in the literature.[5] 4-Bromobutyric acid (0.15 g, 0.86 mmol, 1.1 equiv) was dissolved in 5 mL anhydrous DCM and 1 drop of DMF was added. $SOCl_2$ (0.19 g, 1.56 mmol, 2 equiv) was added and the mixture was stirred at 23 °C for 1 h. Afterwards, DCM and excess $SOCl_2$ was removed under reduced pressure to give the 4-bromobutanoyl chloride. Compound 39 (0.50 g, 0.78 mmol, 1.0 equiv) and NEt_3 (87 mg, 0.86 mmol, 1.1 equiv) were dissolved in 6 mL anhydrous DCM. To this mixture was added the DCM solution (3 mL) of above 4-bromobutanoyl chloride dropwise at 0 °C. The mixture was allowed to warm to 23 °C with stirring during a 2 h period. Afterwards, water (20 mL) was added, and the mixture was extracted by DCM (20 mL) for three times. The organic phase was collected, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 40/1 as the

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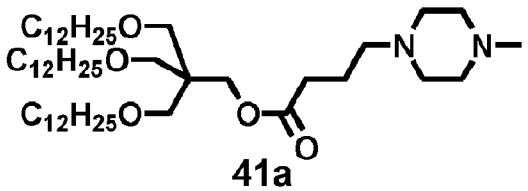
eluent to give the title compound as a colorless oil (0.48 g, 85%).

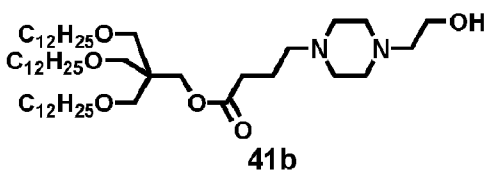
3-(Dodecyloxy)-2,2-bis((dodecyloxy)methyl)propyl 4-(4-methylpiperazin-1-yl)butanoate (41a, IAJD82).



41a

Compound 40 (0.15 g, 0.19 mmol, 1 equiv), 1-methylpiperazine (26 mg, 0.20 mmol, 1.05 equiv), K_2CO_3 (26 mg, 0.20 mmol, 1.05 equiv) were stirred in MeCN (15 mL). The mixture was heated at reflux (95 °C) for 3 h. The reaction mixture was cooled to 23 °C and MeCN was removed under reduced pressure. Then water (20 mL) was added, and the mixture was extracted by DCM (20 mL × 3). The organic phase was collected, dried over anhydrous $MgSO_4$, filtered and dried to give the crude product. The crude product was purified by column chromatography (SiO_2) with DCM/MeOH = 30/1 and 20/1 as the eluent. Then the obtained product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was combined and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.13 g, 80%).

3-(Dodecyloxy)-2,2-bis((dodecyloxy)methyl)propyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (41b, IAJD83).



41b

Compound 41b was synthesized from compound 40 (0.19 g, 0.24 mmol, 1 equiv), 1-(2-hydroxyethyl)piperazine (33 mg, 0.25 mmol, 1.05 equiv) and K_2CO_3 (33 mg, 0.25 mmol, 1.05 equiv) following a procedure similar to that used for the synthesis of compound 41a. The title compound was obtained as a light-yellow oil (0.18 g, 90%).

3,5-Bis(dodecyloxy)benzyl 3-bromopropanoate (42).

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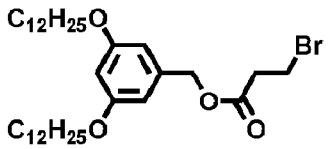
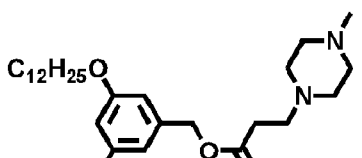


42

3-Bromopropanoic acid (0.16 g, 1.05 mmol, 1.0 equiv) was dissolved in 5 mL anhydrous DCM and 1 drop of DMF was added. $SOCl_2$ (0.25 g, 2.10 mmol, 2.0 equiv) was added and the mixture was stirred at 23 °C for 1 h. Afterwards, DCM and excess $SOCl_2$ was removed under reduced pressure to give the 3-bromopropanoyl chloride. Compound 4 (0.50 g, 1.05 mmol, 1.0 equiv) and NEt_3 (0.12 g, 1.16 mmol, 1.1 equiv) were dissolved in 6 mL anhydrous DCM. To this mixture was added the DCM solution (3 mL) of above 3-bromopropanoyl chloride dropwise at 0 °C. The mixture was allowed to warm to 23 °C with stirring during a 2 h period. Afterwards, water (30 mL) was added, and the mixture was extracted by DCM (30 mL) for three times. The organic phase was collected, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the eluent to give the title compound as a colorless oil (0.47 g, 73%).

3,5-Bis(dodecyloxy)benzyl 3-(4-methylpiperazin-1-yl)propanoate (43, IAJD84).



43

Compound 42 (0.26 g, 0.43 mmol, 1 equiv), 1-methylpiperazine (45 mg, 0.45 mmol, 1.05 equiv), K_2CO_3 (62 mg, 0.45 mmol, 1.05 equiv) were stirred in MeCN (15 mL). The mixture was heated at reflux (95 °C) for 3 h. The reaction mixture was cooled to 23 °C and MeCN was removed under reduced pressure. Then water (20 mL) was added, and the mixture was extracted by DCM (20 mL × 3). The organic phase was collected, dried over anhydrous $MgSO_4$, filtered and dried to give the crude product. The crude product was purified by column chromatography (SiO_2) with DCM/MeOH = 40/1, 20/1 and 15/1 as the eluent. Then the obtained product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was combined and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.23 g, 85%).

Benzyl 3,5-dihydroxybenzoate (44).

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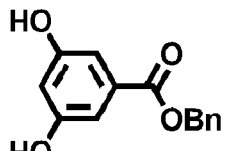
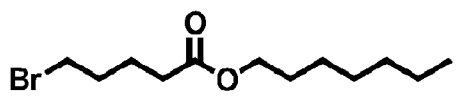


44

The synthesis of compound 44 was adapted from literature procedure.[8] 3,5-Dihydroxyl benzoic acid (13, 5.0 g, 32.4 mmol), benzyl chloride (6.1 g, 48.5 mmol) and $NaHCO_3$ (3.3 g, 38.9 mmol) were mixed with DMF (60 mL), and allowed to stir at 40 °C for 12 h. After the reaction mixture was cooled to 23 °C, DCM (350 mL) and saturated NH_4Cl aqueous solution (350 mL) was added. The mixture was extracted with DCM (100 mL) for 3 times. The organic extract was dried over anhydrous $MgSO_4$, filtered and dried to give the crude product. The crude product was further purified by column chromatography (SiO_2) with a mobile phase of hexane/EtOAc= 3/2 (v/v) to yield the title compound as a white solid (6.2 g, 78%).

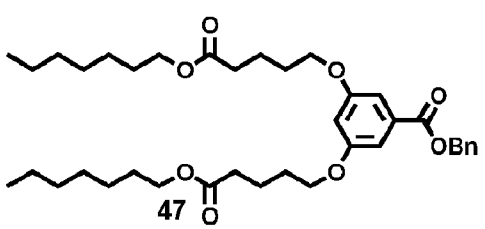
Heptyl 5-bromopentanoate (46).



46

To a solution of 5-bromovaleric acid (5.00 g, 27.6 mmol) in dry DCM (20 mL) was added 1 drop of DMF, followed by the dropwise addition of $SOCl_2$ (6.57 g, 55.2 mmol). The reaction mixture was stirred at 23 °C for 30 min, then DCM and excess $SOCl_2$ was removed under reduced pressure. The obtained acyl chloride was dissolved in dry DCM (10 mL) and added dropwise into a solution of 1-heptanol (3.55 g, 30.4 mmol) and NEt_3 (3.07 g, 30.4 mmol) in dry DCM (20 mL) at 0 °C. After the mixture was stirred at 23 °C for 12 h, water (50 mL) was added, and the mixture was extracted by DCM (50 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 10/1 as the eluent to give the title compound as a colorless oil (7.70 g, 100%).

Diheptyl 5,5'-((5-((benzyloxy)carbonyl)-1,3-phenylene)bis(oxy))dipentanoate (47).



47

Compound 44 (0.39 g, 1.58 mmol), compound 46 (1.10 g, 4.00 mmol), and K_2CO_3 (1.30 g, 9.50

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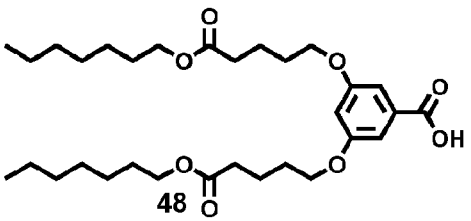
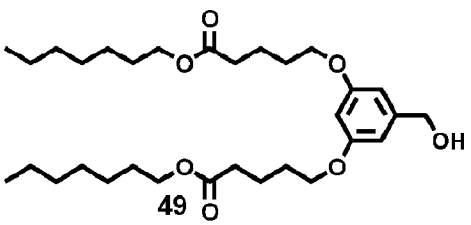
mmol) were stirred in DMF (25 mL). The mixture was stirred at 60 °C under N_2 atmosphere for 6 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the eluent to give the title compound as a colorless oil (0.70 g, 69%).

3,5-Bis((5-(heptyloxy)-5-oxopentyl)oxy)benzoic acid (48).



Compound 47 (0.70 g, 1.09 mmol) was dissolved in 1:2 MeOH:DCM (30 mL). Then Pd/C (50 mg, 7 wt%) was added and the flask was evacuated and filled with hydrogen for three times. The mixture was stirred at 23 °C for 12 h under hydrogen atmosphere. The reaction mixture was filtered through Celite and the filter cake was washed with DCM. Evaporation of the solvent yielded the title compound as a colorless oil (0.60 g, 100%).

Diheptyl 5,5'-((5-(hydroxymethyl)-1,3-phenylene)bis(oxy))dipentanoate (49).



The synthesis of compound 49 was adapted from literature procedure.[9] To a solution of compound 48 (0.60 g, 1.09 mmol) in 30 mL THF was added $NaBH_4$ (0.10 g, 2.75 mmol). The reaction mixture was stirred at 23 °C for 30 min under N_2 until no more gas was generated. A solution of iodine (0.34 g, 1.32 mmol) in THF (10 mL) was added dropwise into the reaction mixture under N_2 in 20 min at 0 °C. After stirring at 0 °C for 30 min, the reaction was returned to 23 °C and stirred for 1.5 h. The reaction was quenched with HCl (2 M) solution (10 mL) until no more gas was generated, and then saturated $NaHCO_3$ aqueous solution was added until pH = 7. The mixture was extracted by DCM (50 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The alcohol as the product (0.59 g, 100%) was obtained by concentrating the filtrate as colorless oil without further purification.

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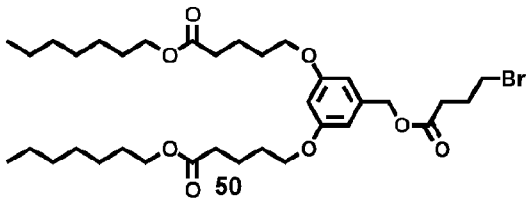
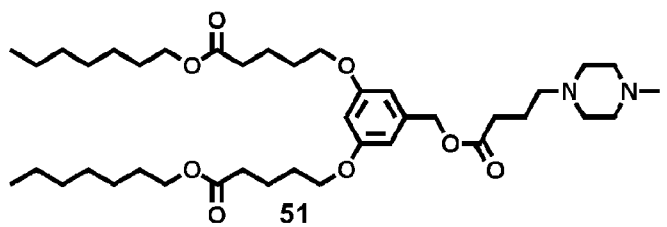
Diheptyl 5,5'-((5-(((4-bromobutanoyl)oxy)methyl)-1,3-phenylene)bis(oxy))dipentanoate (50).



4-Bromobutyric acid (0.22 g, 1.30 mmol, 1.2 equiv) was dissolved in 5 mL anhydrous DCM and 1 drop of DMF was added. $SOCl_2$ (0.26 g, 2.20 mmol, 2.0 equiv) was added and the mixture was stirred at 23 °C for 1 h. Afterwards, DCM and excess $SOCl_2$ was removed under reduced pressure to give the 4-bromobutanoyl chloride. Compound 49 (0.59 g, 1.10 mmol, 1.0 equiv) and NEt_3 (0.13 g, 1.30 mmol, 1.2 equiv) were dissolved in 6 mL anhydrous DCM. Then DMAP (24 mg, 0.20 mmol, 0.2 equiv) was added. To this mixture was added the DCM solution (3 mL) of above 4-bromobutanoyl chloride dropwise at 0 °C. The mixture was allowed to warm to 23 °C with stirring during a 2 h period. Afterwards, water (20 mL) was added, and the mixture was extracted by DCM (20 mL) for three times. The organic phase was collected, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 10/1 as the eluent to give the title compound as a colorless oil (0.32 g, 42%).

Diheptyl 5,5'-((5-(((4-(4-methylpiperazin-1-yl)butanoyl)oxy)methyl)-1,3-phenylene)bis(oxy)) dipentanoate (51, IAJD85).



Compound 50 (0.32 g, 0.47 mmol, 1 equiv), 1-methylpiperazine (50 mg, 0.50 mmol, 1.05 equiv), K_2CO_3 (69 mg, 0.50 mmol, 1.05 equiv) were stirred in MeCN (15 mL). The mixture was heated at reflux (95 °C) for 3 h. The reaction mixture was cooled to 23 °C and MeCN was removed under reduced pressure. Then water (20 mL) was added, and the mixture was extracted by DCM (20 mL × 3). The organic phase was collected, dried over anhydrous $MgSO_4$, filtered and dried to give the crude product. The crude product was purified by column chromatography (SiO_2) with DCM/MeOH = 40/1, 20/1 and 10/1 as the eluent. Then the obtained product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was combined and dried over

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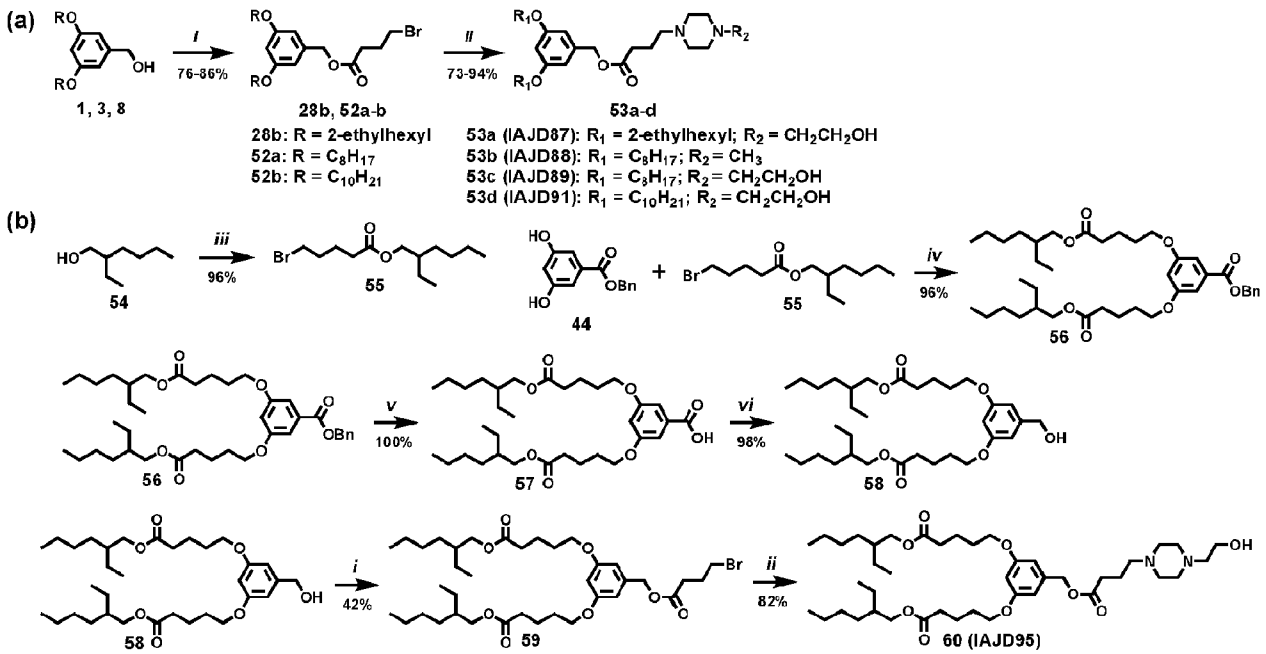
anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.28 g, 79%).

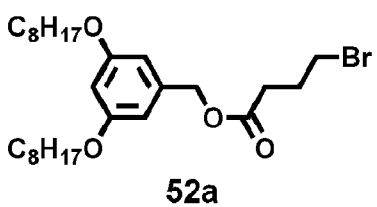
4.6 Synthesis of IAJDs 87-89, 91 and 95 was performed according to Scheme 7

Scheme 7.



Reagents and conditions: (*i*) 4-Bromobutyric acid, $SOCl_2$, DMF (cat.), DCM, 23 °C, 1 h, then DMAP, NEt_3, DCM, 0–23 °C, 2 h; (*ii*) 1-methylpiperazine/1-(2-hydroxyethyl)piperazine, K_2CO_3, MeCN, 95 °C, 3 h; (*iii*) 5-bromovaleric acid, DCC, DPTS, DCM, 23 °C, 12 h; (*iv*) K_2CO_3, DMF, 60 °C, 6 h; (*v*) H_2, Pd/C, DCM, MeOH, 12 h; (*vi*) $NaBH_4$, I_2, THF, 0–23 °C, 2 h.

3,5-Bis(octyloxy)benzyl 4-bromobutanoate (52a).



52a

4-Bromobutyric acid (0.45 g, 2.70 mmol, 1.0 equiv) was dissolved in 6 mL anhydrous DCM and 1 drop of DMF was added. $SOCl_2$ (0.64 g, 5.40 mmol, 2.0 equiv) was added and the mixture was stirred at 23 °C for 1 h. Afterwards, DCM and excess $SOCl_2$ was removed under reduced pressure to give the 4-bromobutanoyl chloride. Compound 1 (1.00 g, 2.70 mmol, 1.0 equiv) and NEt_3 (0.30 g, 3.00 mmol, 1.1 equiv) were dissolved in 10 mL anhydrous DCM. To this mixture was added the DCM solution (5 mL) of above 4-bromobutanoyl chloride dropwise at 0 °C. The mixture was

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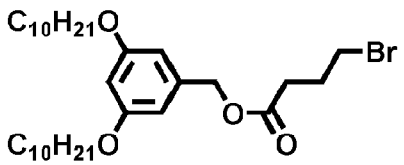
allowed to warm to 23 °C with stirring during a 2 h period. Afterwards, water (30 mL) was added, and the mixture was extracted by DCM (30 mL) for three times. The organic phase was collected, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the eluent to give the title compound as a light-yellow oil (1.22 g, 86%).

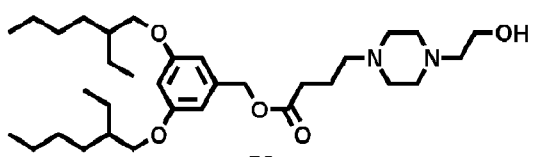
3,5-Bis(decyloxy)benzyl 4-bromobutanoate (52b).



52b

Compound 52b was synthesized from compound 3 (1.00 g, 2.38 mmol, 1.0 equiv), 4-bromobutyric acid (0.40 g, 2.38 mmol, 1.0 equiv) and NEt_3 (0.27 g, 2.62 mmol, 1.1 equiv) following a procedure similar to that used for the synthesis of compound 52a. The title compound was obtained as a light-yellow oil (1.02 g, 76%).

3,5-Bis((2-ethylhexyl)oxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (53a, IAJD87).



53a

Compound 28b (0.30 g, 0.58 mmol, 1.0 equiv), 1-(2-hydroxyethyl)piperazine (79 mg, 0.61 mmol, 1.05 equiv), K_2CO_3 (84 mg, 0.61 mmol, 1.05 equiv) were stirred in MeCN (20 mL). The mixture was heated at reflux (95 °C) for 3 h. The reaction mixture was cooled to 23 °C and MeCN was removed under reduced pressure. Then water (20 mL) was added, and the mixture was extracted by DCM (20 mL × 3). The organic phase was collected, dried over anhydrous $MgSO_4$, filtered and dried to give the crude product. The crude product was purified by column chromatography (SiO_2) with DCM/MeOH = 40/1 and 15/1 as the eluent. Then the obtained product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was combined and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.31 g, 94%).

3,5-Bis(octyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (53b, IAJD88).

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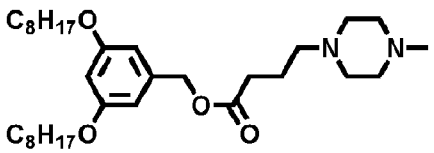
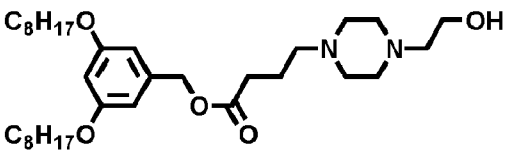


53b

Compound 53b was synthesized from compound 52a (0.30 g, 0.58 mmol, 1 equiv), 1-methylpiperazine (61 mg, 0.61 mmol, 1.05 equiv) and K_2CO_3 (85 mg, 0.61 mmol, 1.05 equiv) following a procedure similar to that used for the synthesis of compound 53a. The title compound was obtained as a light-yellow oil (0.23 g, 74%).

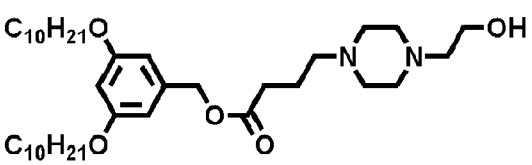
3,5-Bis(octyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (53c, IAJD89).



53c

Compound 53c was synthesized from compound 52a (0.30 g, 0.58 mmol, 1 equiv), 1-(2-hydroxyethyl)piperazine (82 mg, 0.61 mmol, 1.05 equiv) and K_2CO_3 (85 mg, 0.61 mmol, 1.05 equiv) following a procedure similar to that used for the synthesis of compound 53a. The title compound was obtained as a light-yellow oil (0.24 g, 73%).

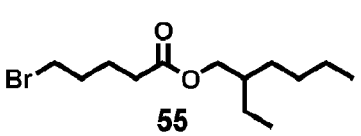
3,5-Bis(decyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (53d, IAJD91).



53d

Compound 53d was synthesized from compound 52b (0.30 g, 0.53 mmol, 1 equiv), 1-(2-hydroxyethyl)piperazine (73 mg, 0.56 mmol, 1.05 equiv) and K_2CO_3 (77 mg, 0.56 mmol, 1.05 equiv) following a procedure similar to that used for the synthesis of compound 53a. The title compound was obtained as a colorless oil (0.31 g, 94%).

2-Ethylhexyl 5-bromopentanoate (55).



55

To a DCM (10 mL) solution of 2-ethylhexanol (54, 2.00 g, 11.1 mmol), 5-bromovaleric acid (1.44 g, 11.1 mmol) and DPTS (3.60 g, 12.2 mmol), was added DCC (4.58 g, 22.2 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated

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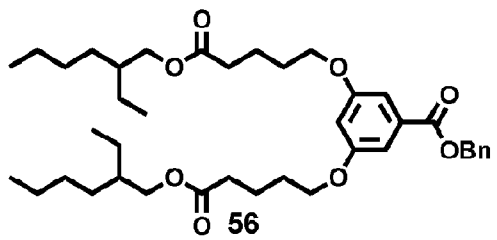
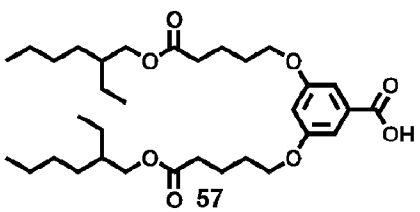
to dryness. The crude product was further purified by column chromatography with a mobile phase of hexane/DCM = 1/1 to yield the title compound as a colorless oil (3.13 g, 96%).

Bis(2-ethylhexyl) 5,5'-((5-((benzyloxy)carbonyl)-1,3-phenylene)bis(oxy))dipentanoate (56).



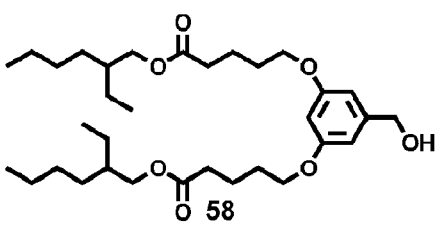
Benzyl 3,5-hydroxyl benzoate (44, 0.78 g, 3.21 mmol), compound 55 (2.00 g, 6.75 mmol) and K_2CO_3 (2.66 g, 19.3 mmol) were stirred in DMF (20 mL). The mixture was stirred at 60 °C under N_2 atmosphere for 6 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM as the eluent to give the title compound as a colorless oil (2.95 g, 96%).

3,5-Bis((5-((2-ethylhexyl)oxy)-5-oxopentyl)oxy)benzoic acid (57).



Compound 56 (2.13 g, 3.18 mmol) was dissolved in 1:2 MeOH:DCM (45 mL). Then Pd/C (0.21 g, 10 wt%) was added and the flask was evacuated and filled with hydrogen for three times. The mixture was stirred at 23 °C for 12 h under hydrogen atmosphere. The reaction mixture was filtered through Celite and the filter cake was washed with DCM. Evaporation of the solvent yielded the title compound as a colorless oil (1.84 g, 100%).

Bis(2-ethylhexyl) 5,5'-((5-(hydroxymethyl)-1,3-phenylene)bis(oxy))dipentanoate (58).



To a solution of compound 57 (1.78 g, 3.08 mmol) in 50 mL THF was added $NaBH_4$ (0.35 g, 9.23

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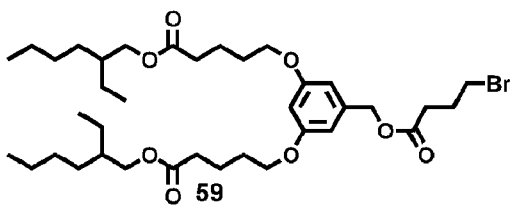
mmol). The reaction mixture was stirred at 23 °C for 30 min under N_2 until no more gas was generated. A solution of iodine (1.02 g, 4.00 mmol) in THF (20 mL) was added dropwise into the reaction mixture under N_2 in 20 min at 0 °C. After stirring at 0 °C for 30 min, the reaction was returned to 23 °C and stirred for 1.5 h. The reaction was quenched with HCl (2 M) solution (10 mL) until no more gas was generated, and then saturated $NaHCO_3$ aqueous solution was added until pH = 7. The mixture was extracted by DCM (50 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The alcohol as the product (1.70 g, 98%) was obtained by concentrating the filtrate as colorless oil without further purification.

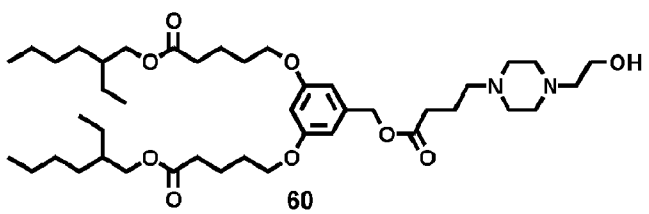
Compound 59.



59

Compound 59 was synthesized from compound 58 (1.70 g, 3.0 mmol), 4-bromobutyric acid (0.60 g, 3.6 mmol) and NEt_3 (0.36 g, 3.6 mmol) following a procedure similar to that used for the synthesis of compound 52a. Besides, DMAP (24 mg, 0.2 mmol) was added together with NEt_3 to perform the esterification. The title compound was obtained as a colorless oil (0.90 g, 42%).

Compound 60 (IAJD95).



60

Compound 60 was synthesized from compound 59 (0.45 g, 0.63 mmol, 1.0 equiv), 1-(2-hydroxyethyl)piperazine (90 mg, 0.69 mmol, 1.1 equiv) and K_2CO_3 (95 mg, 0.69 mmol, 1.1 equiv) following a procedure similar to that used for the synthesis of compound 53a. The title compound was obtained as a colorless oil (0.38 g, 82%).

Synthesis of IAJDs 96-99 and 103 was performed according to Scheme 8.

Scheme 8.

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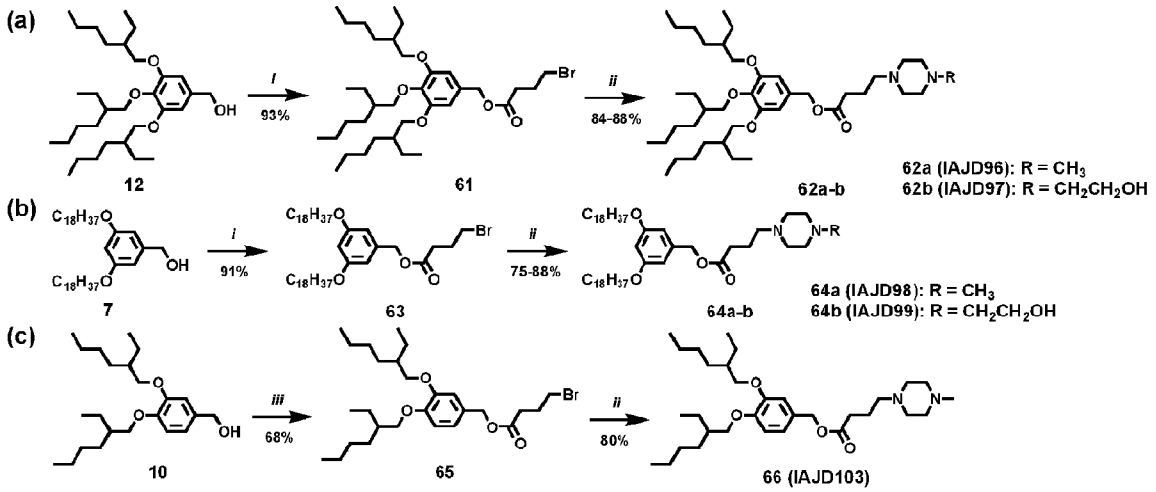
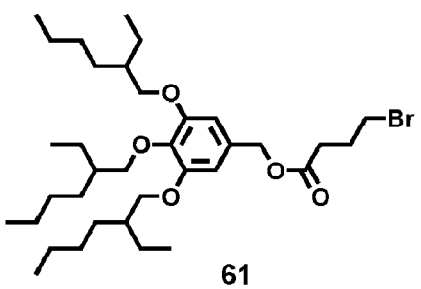


Reagents and conditions: (*i*) 4-Bromobutyric acid, $SOCl_2$, DMF (cat.), DCM, 23 °C, 1 h, then DMAP, NEt_3, DCM, 0–23 °C, 2 h; (*ii*) 1-methylpiperazine/1-(2-hydroxyethyl)piperazine, K_2CO_3, MeCN, 95 °C, 3 h; (*iii*) DCC, DPTS, DCM, 23 °C, 12 h.

3,4,5-Tris((2-ethylhexyl)oxy)benzyl 4-bromobutanoate (61).



61

4-Bromobutyric acid (0.33 g, 1.95 mmol, 1.1 equiv) was dissolved in 6 mL anhydrous DCM and 1 drop of DMF was added. $SOCl_2$ (0.42 g, 3.54 mmol, 2.0 equiv) was added and the mixture was stirred at 23 °C for 1 h. Afterwards, DCM and excess $SOCl_2$ was removed under reduced pressure to give the 4-bromobutanoyl chloride. Compound 12 (0.87 g, 1.77 mmol, 1.0 equiv) and NEt_3 (0.27 g, 2.66 mmol, 1.5 equiv) were dissolved in 6 mL anhydrous DCM. To this mixture was added the DCM solution (4 mL) of above 4-bromobutanoyl chloride dropwise at 0 °C. The mixture was allowed to warm to 23 °C with stirring during a 2 h period. Afterwards, water (20 mL) was added, and the mixture was extracted by DCM (20 mL) for three times. The organic phase was collected, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the eluent to give the title compound as a colorless oil (1.06 g, 93%).

3,4,5-Tris((2-ethylhexyl)oxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (62a, IAJD96).

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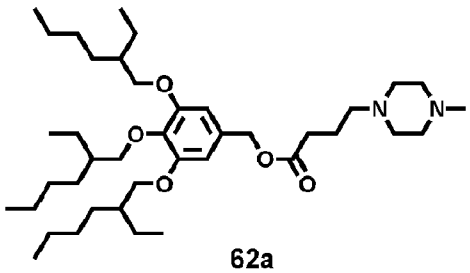
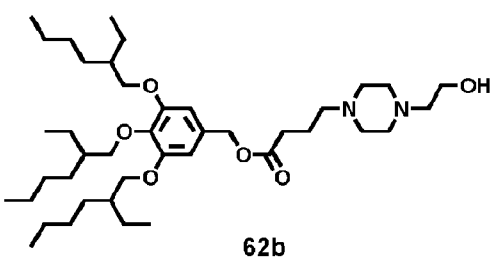


62a

Compound 61 (0.30 g, 0.47 mmol, 1.0 equiv), 1-methylpiperazine (49 mg, 0.49 mmol, 1.05 equiv), K_2CO_3 (68 mg, 0.49 mmol, 1.05 equiv) were stirred in MeCN (20 mL). The mixture was heated at reflux (95 °C) for 3 h. The reaction mixture was cooled to 23 °C and MeCN was removed under reduced pressure. Then water (20 mL) was added, and the mixture was extracted by DCM (20 mL × 3). The organic phase was collected, dried over anhydrous $MgSO_4$, filtered and dried to give the crude product. The crude product was purified by column chromatography (SiO_2) with DCM/MeOH = 40/1 and 15/1 as the eluent. Then the obtained product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was combined and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.26 g, 84%).

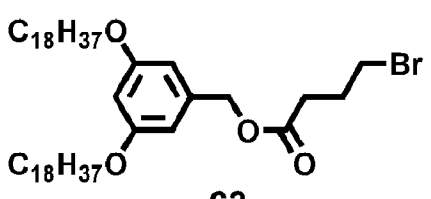
3,4,5-Tris((2-ethylhexyl)oxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (62b, IAJD97).



62b

Compound 62b was synthesized from compound 61 (0.30 g, 0.47 mmol, 1.0 equiv), 1-(2-hydroxyethyl)piperazine (64 mg, 0.49 mmol, 1.05 equiv) and K_2CO_3 (68 mg, 0.49 mmol, 1.05 equiv) following a procedure similar to that used for the synthesis of compound 62a. The title compound was obtained as a light-yellow oil (0.28 g, 88%).

3,5-Bis(octadecyloxy)benzyl 4-bromobutanoate (63).



63

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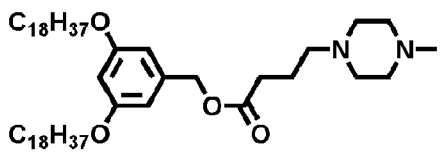
Compound 63 was synthesized from compound 7 (0.85 g, 1.32 mmol, 1.0 equiv), 4-bromobutyric acid (0.24 g, 1.45 mmol, 1.1 equiv) and NEt_3 (0.15 g, 1.45 mmol, 1.1 equiv) following a procedure similar to that used for the synthesis of compound 61. Besides, DMAP (32 mg, 0.26 mmol, 0.2 equiv) was added together with NEt_3 to perform the esterification. The title compound was obtained as a colorless oil (0.94 g, 91%).

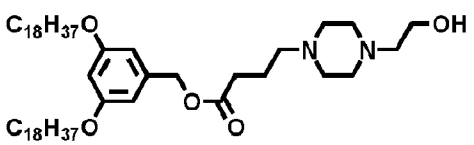
3,5-Bis(octadecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (64a, IAJD98).



64a

Compound 64a was synthesized from compound 63 (0.30 g, 0.38 mmol, 1.0 equiv), 1-methylpiperazine (41 mg, 0.40 mmol, 1.05 equiv) and K_2CO_3 (58 mg, 0.42 mmol, 1.1 equiv) following a procedure similar to that used for the synthesis of compound 62a. The title compound was obtained as a light-yellow oil (0.27 g, 88%).

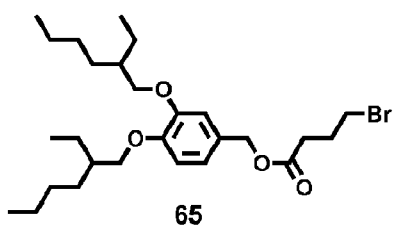
3,5-Bis(octadecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (64b, IAJD99).



64b

Compound 64b was synthesized from compound 63 (0.30 g, 0.38 mmol, 1.0 equiv), 1-(2-hydroxyethyl)piperazine (53 mg, 0.40 mmol, 1.05 equiv) and K_2CO_3 (58 mg, 0.42 mmol, 1.1 equiv) following a procedure similar to that used for the synthesis of compound 62a. The title compound was obtained as a light-yellow oil (0.24 g, 75%).

3,4-Bis((2-ethylhexyl)oxy)benzyl 4-bromobutanoate (65).



65

To a DCM (10 mL) solution of compound 10 (0.60 g, 1.65 mmol), 4-bromobutyric acid (0.28 g, 1.65 mmol) and DPTS (0.53 g, 1.81 mmol), was added DCC (0.68 g, 3.30 mmol). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of

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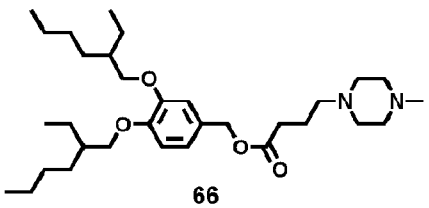
hexane/DCM = 1/1 to yield the title compound as a colorless oil (0.58 g, 68%).

3,4-Bis((2-ethylhexyl)oxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (66, IAJD103).

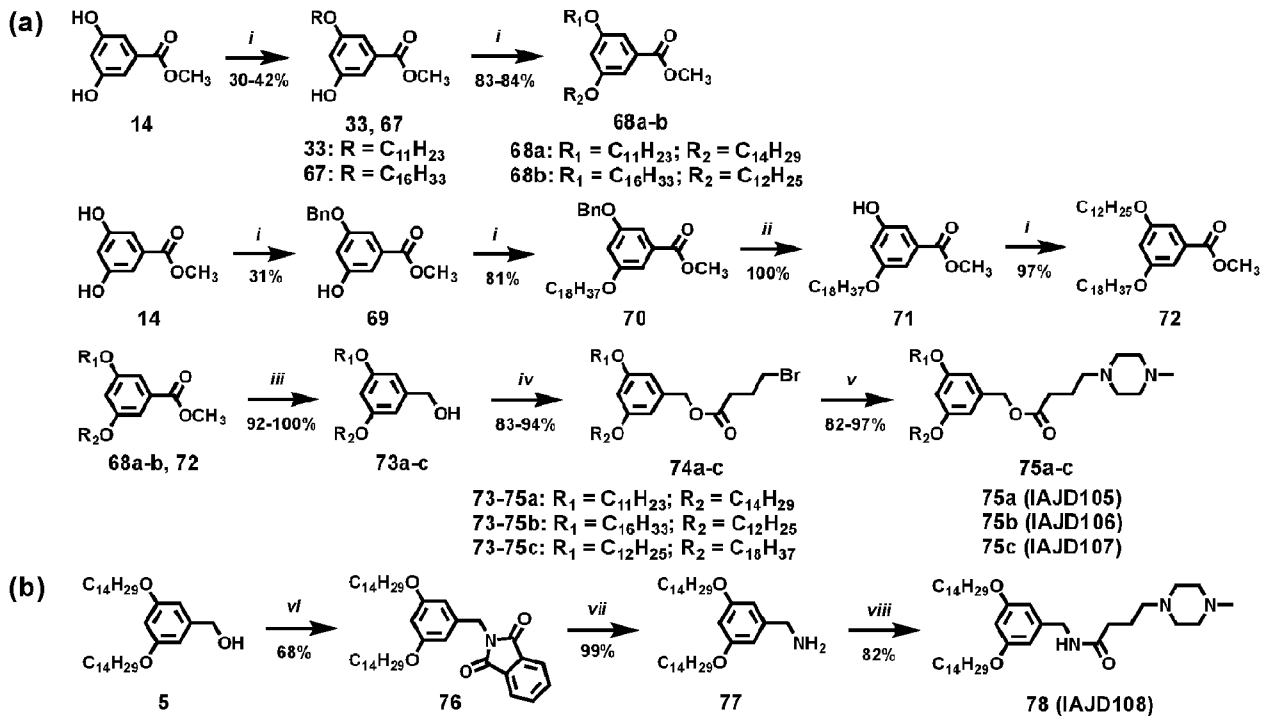


66

Compound 66 was synthesized from compound 65 (0.54 g, 1.05 mmol, 1.0 equiv), 1-methylpiperazine (0.12 g, 1.16 mmol, 1.1 equiv) and K_2CO_3 (0.16 g, 1.16 mmol, 1.1 equiv) following a procedure similar to that used for the synthesis of compound 62a. The title compound was obtained as a colorless oil (0.45 g, 80%).

Synthesis of IAJDs 105-108 was according to Scheme 9.

Scheme 9.



Reagents and conditions: (*i*) RBr, K_2CO_3, DMF, 60 °C, 6 h; (*ii*) H_2, Pd/C, DCM, MeOH, 12 h; (*iii*) $LiAlH_4$, THF, 0–23 °C, 1 h; (*iv*) DCC, DPTS, DCM, 23 °C, 12 h; (*v*) 1-methylpiperazine, K_2CO_3, MeCN, 95 °C, 3 h; (*vi*) $SOCl_2$, DMF (cat.), DCM, 23 °C, 0.5 h, then potassium phthalimide, DMF, 80 °C, 1 h; (*vii*) Hydrazine monohydrate, EtOH, reflux, 2 h; (*viii*) DCC, DCM, 23 °C, 12 h.

Methyl 3-(hexadecyloxy)-5-hydroxybenzoate (67).

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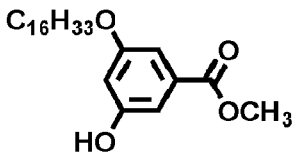
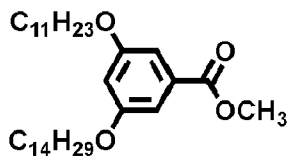


67

Methyl 3,5-dihydroxybenzoate (14, 2.62 g, 15.6 mmol, 1 equiv), 1-bromohexadecane (10.0 g, 32.7 mmol, 2.1 equiv), and K_2CO_3 (8.61 g, 62.4 mmol, 4 equiv) were stirred in DMF (50 mL). The mixture was stirred at 60 °C under N_2 atmosphere for 6 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 10/1 as the eluent to give the title compound as a white solid (1.86 g, 30%).

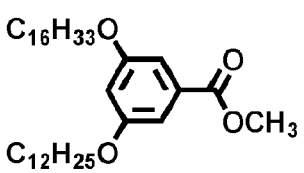
Methyl 3-(tetradecyloxy)-5-(undecyloxy)benzoate (68a).



68a

Compound 68a was synthesized from compound 33 (0.45 g, 1.39 mmol, 1.0 equiv), 1-bromotetradecane (0.41 g, 1.46 mmol, 1.05 equiv), and K_2CO_3 (0.58 g, 4.17 mmol, 3 equiv) following a procedure similar to that used for the synthesis of compound 67. The title compound was obtained as a white solid (0.60 g, 83%).

Methyl 3-(dodecyloxy)-5-(hexadecyloxy)benzoate (68b).



68b

Compound 68b was synthesized from compound 67 (0.30 g, 0.76 mmol, 1.0 equiv), 1-bromododecane (0.20 g, 0.80 mmol, 1.05 equiv), and K_2CO_3 (0.21 g, 1.53 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound 67. The title compound was obtained as a white solid (0.36 g, 84%).

Methyl 3-(benzyloxy)-5-hydroxybenzoate (69).

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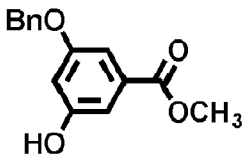
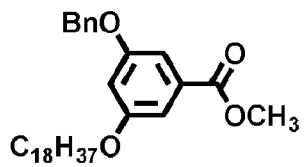


69

Compound 69 was synthesized from compound 14 (10.00 g, 59.5 mmol, 1.0 equiv), benzyl chloride (8.30 g, 65.4 mmol, 1.1 equiv), and K_2CO_3 (9.06 g, 65.4 mmol, 1.1 equiv) following a procedure similar to that used for the synthesis of compound 67. The title compound was obtained as a white solid (4.80 g, 31%).

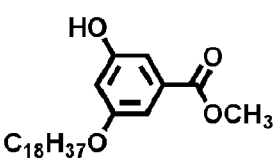
Methyl 3-(benzyloxy)-5-(octadecyloxy)benzoate (70).



70

Compound 70 was synthesized from compound 69 (2.00 g, 7.74 mmol, 1.0 equiv), 1-bromooctadecane (2.70 g, 8.13 mmol, 1.05 equiv), and K_2CO_3 (2.14 g, 15.50 mmol, 2.0 equiv) following a procedure similar to that used for the synthesis of compound 67. The title compound was obtained as a white solid (3.20 g, 81%).

Methyl 3-hydroxy-5-(octadecyloxy)benzoate (71).



71

Compound 70 (3.20 g, 6.26 mmol) was dissolved in the mixture of DCM (50 mL) and methanol (10 mL). Then Pd/C (0.20 g, 6 wt%) was added and the flask was evacuated and filled with hydrogen for three times. The mixture was stirred at 23 °C for 12 h under hydrogen atmosphere. The reaction mixture was filtered through Celite and the filter cake was washed with DCM. Evaporation of the solvent yielded the title compound as a white solid (2.63 g, 100%).

Methyl 3-(dodecyloxy)-5-(octadecyloxy)benzoate (72).

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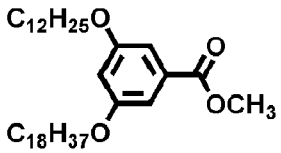
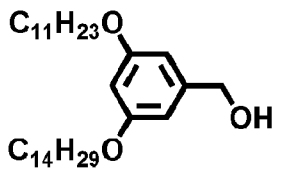


72

Compound 72 was synthesized from compound 71 (0.53 g, 1.26 mmol, 1.0 equiv), 1-bromododecane (0.33 g, 1.32 mmol, 1.05 equiv), and K_2CO_3 (0.35 g, 2.52 mmol, 2.0 equiv) following a procedure similar to that used for the synthesis of compound 67. The title compound was obtained as a white solid (0.72 g, 97%).

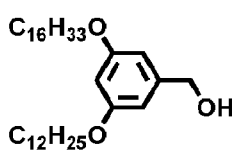
(3-(Tetradecyloxy)-5-(undecyloxy)phenyl)methanol (73a).



73a

Compound 68a (0.59 g, 1.14 mmol, 1.0 equiv) was dissolved in 5 mL dry THF, which was added dropwise to a slurry of $LiAlH_4$ (44 mg, 1.14 mmol, 1.0 equiv) in dry THF (5 mL) at 0 °C under N_2 atmosphere. The resulted mixture was stirred at 23 °C for 1 h. The reaction was quenched by the successive addition of water (0.3 mL), 15% NaOH aqueous solution (0.3 mL) and water (1.5 mL). Then the mixture was filtered to remove white precipitates and the filtrate was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.55 g, 98%).

(3-(Dodecyloxy)-5-(hexadecyloxy)phenyl)methanol (73b).



73b

Compound 73b was synthesized from compound 68b (0.32 g, 0.57 mmol, 1.0 equiv) and $LiAlH_4$ (22 mg, 0.57 mmol, 1.0 equiv) following a procedure similar to that used for the synthesis of compound 73a. The title compound was obtained as a white solid (0.30 g, 100%).

(3-(Dodecyloxy)-5-(octadecyloxy)phenyl)methanol (73c).

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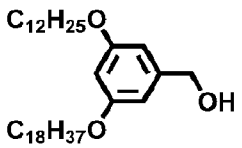
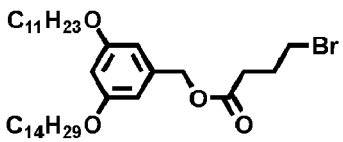


73c

Compound 73c was synthesized from compound 72 (0.72 g, 1.22 mmol, 1.0 equiv) and $LiAlH_4$ (47 mg, 1.22 mmol, 1.0 equiv) following a procedure similar to that used for the synthesis of compound 73a. The title compound was obtained as a colorless oil (0.63 g, 92%).

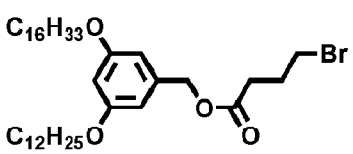
3-(Tetradecyloxy)-5-(undecyloxy)benzyl 4-bromobutanoate (74a).



74a

To a DCM (10 mL) solution of compound 73a (0.53 g, 1.08 mmol, 1.0 equiv), 4-bromobutyric acid (0.20 g, 1.19 mmol, 1.1 equiv) and DPTS (0.35 g, 1.19 mmol, 1.1 equiv), was added DCC (0.45 g, 2.16 mmol, 2.0 equiv). The mixture was allowed to stir at 23 °C for 12 h. The precipitate was filtered, and the filtrate was concentrated to dryness. The crude product was further purified by column chromatography with a mobile phase of hexane/DCM = 1/1 to yield the title compound as a colorless oil (0.61 g, 88%).

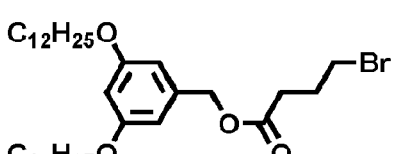
3-(Dodecyloxy)-5-(hexadecyloxy)benzyl 4-bromobutanoate (74b).



74b

Compound 74b was synthesized from compound 73b (0.30 g, 0.56 mmol, 1.0 equiv), 4-bromobutyric acid (94 mg, 0.56 mmol, 1.0 equiv), DPTS (0.18 g, 0.62 mmol, 1.1 equiv) and DCC (0.23 g, 1.13 mmol, 2.0 equiv) following a procedure similar to that used for the synthesis of compound 74a. The title compound was obtained as a white solid (0.36 g, 94%).

3-(Dodecyloxy)-5-(octadecyloxy)benzyl 4-bromobutanoate (74c).



74c

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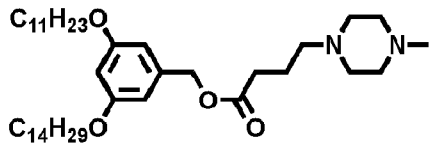
Compound 74c was synthesized from compound 73c (0.63 g, 1.12 mmol, 1.0 equiv), 4-bromobutyric acid (0.19 g, 1.12 mmol, 1.0 equiv), DPTS (0.33 g, 1.23 mmol, 1.1 equiv) and DCC (0.46 g, 2.24 mmol, 2.0 equiv) following a procedure similar to that used for the synthesis of compound 74a. The title compound was obtained as a colorless oil (0.66 g, 83%).

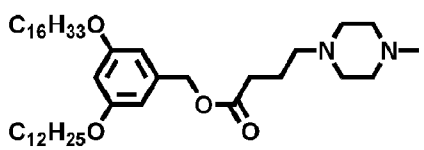
3-(Tetradecyloxy)-5-(undecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (75a, IAJD105).



75a

Compound 74a (0.61 g, 0.95 mmol, 1.0 equiv), 1-methylpiperazine (0.11 g, 1.10 mmol, 1.1 equiv), K_2CO_3 (0.20 g, 1.42 mmol, 1.5 equiv) were stirred in MeCN (20 mL). The mixture was heated at reflux (95 °C) for 3 h. The reaction mixture was cooled to 23 °C and MeCN was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was collected, dried over anhydrous $MgSO_4$, filtered and dried to give the crude product. The crude product was purified by column chromatography (SiO_2) with DCM/MeOH = 20/1 as the eluent. Then the obtained product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was combined and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.56 g, 90%).

3-(Dodecyloxy)-5-(hexadecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (75b, IAJD106).



75b

Compound 75b was synthesized from compound 74b (0.36 g, 0.53 mmol, 1.0 equiv), 1-methylpiperazine (79 mg, 0.79 mmol, 1.5 equiv) and K_2CO_3 (0.11 g, 0.79 mmol, 1.5 equiv) following a procedure similar to that used for the synthesis of compound 75a. The title compound was obtained as a light-yellow oil (0.36 g, 97%).

3-(Dodecyloxy)-5-(octadecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (75c, IAJD107).

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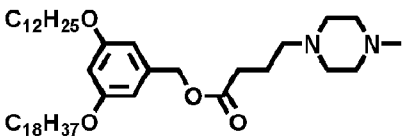
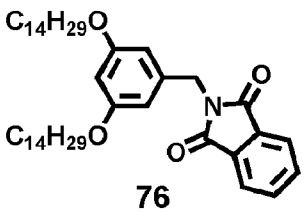


75c

Compound 75c was synthesized from compound 74c (0.64 g, 0.90 mmol, 1.0 equiv), 1-methylpiperazine (99 mg, 0.99 mmol, 1.1 equiv) and K_2CO_3 (0.15 g, 1.10 mmol, 1.2 equiv) following a procedure similar to that used for the synthesis of compound 75a. The title compound was obtained as a light-yellow oil (0.54 g, 82%).

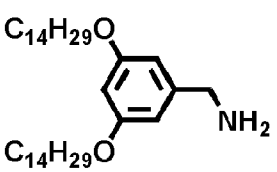
2-(3,5-Bis(tetradecyloxy)benzyl)isoindoline-1,3-dione (76).



76

To a DCM solution (20 mL) of compound 5 (4.14 g, 7.77 mmol, 1.0 equiv) was added 2 drops of DMF, followed by the dropwise addition of $SOCl_2$ (1.12 g, 9.37 mmol, 1.2 equiv). The reaction mixture was stirred at 23 °C for 0.5 h. Then DCM and excess $SOCl_2$ was removed under reduced pressure. The obtained intermediate product was dissolved in dry DMF (40 mL) and potassium phthalimide (1.73 g, 9.32 mmol, 1.2 equiv) was added. The mixture was stirred at 80 °C under N_2 atmosphere for 1 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL × 3). The organic phase was dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the eluent to give the title compound as a white solid (3.48 g, 68%).

(3,5-Bis(tetradecyloxy)phenyl)methanamine (77).



77

Compound 76 (1.00 g, 1.50 mmol, 1 equiv) and hydrazine monohydrate ($NH_2NH_2•H_2O$, 0.75 g, 15.0 mmol, 10 equiv) were dissolved in EtOH (20 mL). The mixture was heated at reflux for 2 h. The reaction mixture was cooled to 23 °C and EtOH was removed under reduced pressure. Then water (10 mL) was added, and the mixture was extracted by DCM (10 mL × 3). The organic phase

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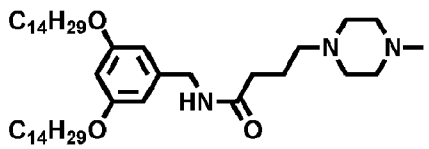
was dried over anhydrous $MgSO_4$ and filtered. Filtration and evaporation of the solvent yielded the title compound as a white solid (0.85 g, 99%).

N-(3,5-bis(tetradecyloxy)benzyl)-4-(4-methylpiperazin-1-yl)butanamide (78, IAJD108).



78

Compound 77 (0.25 g, 0.47 mmol, 1 equiv) and 4-(4-methylpiperazin-1-yl)butanoic acid hydrochloride (24b, 0.10 g, 0.47 mmol, 1 equiv) were dissolved in 6 mL dry DCM. NEt_3 (3 drops) was added. DCC (0.19 g, 0.94 mmol, 2 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. Afterwards, urea was removed by filtration, and washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 30/1, 10/1, 8/1 and 8/1 with 0.1% TEA as the eluent. Then the product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was combined and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a white solid (0.27 g, 82%).

Dimensions of DNPs Containing Luc-mRNA Obtained by Co-assembly of IAJDs with Luc-mRNA by Injection in Acetate Buffer (10 mM, pH 4.0) and Luminescence Expression in HEK293T Cells (125 ng/well, $n = 3$) are shown in Table 12.

Table 12.

IAJD	Buffer (10 mM)	C_{IAJD} (mg/mL)	$C_{Luc\text{-}mRNA}$ (mg/mL)	Size (d, nm)	PDI	Luminescence	Standard deviation
64	Acetate (pH 4.0)	4	0.10	135	0.271	125	7
65	Acetate (pH 4.0)	4	0.10	241	0.249	18	2
66	Acetate (pH 4.0)	4	0.10	165	0.320	234	43
70	Acetate (pH 4.0)	4	0.10	205	0.438	455,361	66,604
71	Acetate (pH 4.0)	4	0.10	237	0.451	473,548	123,589
74	Acetate (pH 4.0)	4	0.10	180	0.317	3,926	1,559
75	Acetate (pH	4	0.10	235	0.32	68,572	6,794

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	4.0)				8		
76	Acetate (pH 4.0)	4	0.10	78	0.287	17	3
77	Acetate (pH 4.0)	4	0.10	110	0.267	129	6
78	Acetate (pH 4.0)	4	0.10	629	1.000	8,688	1,914
79	Acetate (pH 4.0)	4	0.10	352	0.763	27,615	6,704
81	Acetate (pH 4.0)	4	0.10	111	0.287	206,161	21,200
82	Acetate (pH 4.0)	4	0.10	162	0.144	17	2
83	Acetate (pH 4.0)	4	0.10	143	0.430	68,691	2,498
84	Acetate (pH 4.0)	4	0.10	171	0.303	205	152
85	Acetate (pH 4.0)	4	0.10	102	0.250	47,369	18,262
86	Acetate (pH 4.0)	4	0.10	91	0.234	47,066	21,163
87	Acetate (pH 4.0)	4	0.10	146	0.486	33	7
88	Acetate (pH 4.0)	4	0.10	100	0.342	17	2
89	Acetate (pH 4.0)	4	0.10	149	0.467	55	12
91	Acetate (pH 4.0)	4	0.10	188	0.498	65,255	5,573
95	Acetate (pH 4.0)	4	0.10	105	0.352	153,564	25,782
96	Acetate (pH 4.0)	4	0.10	139	0.402	22,799	4,244
97	Acetate (pH 4.0)	4	0.10	204	0.482	342,016	70,616
98	Acetate (pH 4.0)	4	0.10	187	0.221	22,209	10,213
99	Acetate (pH 4.0)	4	0.10	277	0.571	87,433	7,107
103	Acetate (pH 4.0)	4	0.10	88	0.275	19	8
105	Acetate (pH 4.0)	4	0.10	94	0.294	85	21
106	Acetate (pH 4.0)	4	0.10	105	0.275	309	116
107	Acetate (pH 4.0)	4	0.10	146	0.279	93,745	12,116
108	Acetate (pH	4	0.10	214	0.33	136,424	3,167

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	4.0)	4		
Negative Control (untreated cells)			81	5
MC3-Luc-mRNA			1,320,533	105,967

Quantification of luminescence expression *in vivo* is shown in Table 13.

Table 13

IAJD	C_{IAJD} (mg/mL)	$C_{Luc-mRNA}$ (mg/mL)	pH		Size (d, nm)	PDI	T (h)[a]	Total flux (p/s)	Organs with luminescence
			Acetate buffer	Sample					
64	4	0.10	4.0	4.53	135	0.271	4	0	--
65	4	0.10	4.0	4.22	241	0.249	4	0	--
66	4	0.10	4.0	4.50	165	0.320	4	2.58×10^7	Lung, liver, spleen
70	4	0.10	4.0	4.50	205	0.438	5	6.62×10^6	Lung, spleen
71	4	0.10	4.0	4.39	237	0.451	5	3.79×10^7	Lung, liver, spleen
74	4	0.10	4.0	4.46	180	0.317	5	1.71×10^7	Lung, liver, spleen
75	4	0.10	4.0	4.62	235	0.328	5	3.37×10^7	Lung, liver, spleen
76	4	0.10	4.0	4.47	78	0.287	5	7.81×10^7	Lung, liver, spleen
77	4	0.10	4.0	4.50	110	0.267	5	2.44×10^6	Lung, liver, spleen
78	4	0.10	4.0	4.46	629	1.000	5	1.12×10^5	Lung, spleen
79	4	0.10	4.0	4.43	352	0.763	6	1.34×10^7	Lung, liver, spleen
81	4	0.10	4.0	4.38	111	0.287	6	2.32×10^7	Lung, liver, spleen
82	4	0.10	4.0	4.26	162	0.144	5	1.62×10^6	spleen
83	4	0.10	4.0	4.31	143	0.430	5	7.11×10^7	Lung, liver, spleen
84	4	0.10	4.0	4.44	171	0.303	5	6.89×10^5	Lung, liver, spleen
85	4	0.10	4.0	4.44	102	0.250	5	2.87×10^6	Lung, liver, spleen
86	4	0.10	4.0	4.39	91	0.234	5	4.73×10^7	Lung, liver, spleen
87	4	0.10	4.0	4.47	146	0.486	6	3.24×10^7	Lung, liver, spleen

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88	4	0.10	4.0	4.55	100	0.342	6	2.34×10^6	Lung, liver, spleen
89	4	0.10	4.0	4.45	149	0.467	6	5.92×10^7	Lung, liver, spleen
91	4	0.10	4.0	4.44	188	0.498	6	9.94×10^6	Lung, liver, spleen
95	4	0.10	4.0	4.38	105	0.352	6	6.61×10^7	Lung, liver, spleen
96	4	0.10	4.0	4.52	139	0.402	5	4.79×10^7	Lung, liver, spleen
97	4	0.10	4.0	4.42	204	0.482	5	5.89×10^7	Lung, liver, spleen
98	4	0.10	4.0	4.35	187	0.221	6	2.06×10^6	Lung, liver, spleen
99	4	0.10	4.0	4.32	277	0.571	6	4.49×10^6	Lung, liver, spleen
103	4	0.10	4.0	4.58	88	0.275	6	1.70×10^6	Lung, spleen
105	4	0.10	4.0	4.42	94	0.294	6	4.74×10^7	Lung, liver, spleen
106	4	0.10	4.0	4.41	105	0.275	6	3.48×10^7	Lung, liver, spleen
107	4	0.10	4.0	4.42	146	0.279	6	2.19×10^7	Lung, liver, spleen
108	4	0.10	4.0	4.54	214	0.334	6	9.66×10^6	Lung, spleen

a. Time from Luc-mRNA-IAJD injection to the imaging of mice on the IVIS imaging system.

DLS data of DNPs assembled from IAJDs and Luc-mRNA is shown in Figure 50 through Figure 53. Stability of DNPs by determined by DLS[a] at 5 °C after different periods of time is shown in Table 14.

Table 14. Stability of DNPs by determined by DLS[a] at 5 °C after different periods of time. *a*. All the samples are prepared in acetate buffer. *b*. U: Unimodal size distribution of DLS curve; B: Bimodal size distribution of DLS curve; M: Multimodal size distribution of DLS curve. *c*. Ranking of stability: Excellent (in green color) – the original sizes and distribution remained almost the same after storage; Good – the original sizes and distribution changed slightly (size difference was less than 100 nm) after storage or the original size remained almost the same but with multimodal distributions after storage; Not stable – the original sizes and distribution changed obviously (size difference was more than 100 nm and distribution type of DLS curve changed) after storage.

IAJD	**C_{IAJD} (mg/mL)**	**$C_{Luc\text{-}mRNA}$ (mg/mL)**	**pH**		**Size (d, nm)**	**PDI**	**T (h)** [a]	**Total flux (p/s)**	**Organs with luminescence**[b]
			Acetate buffer	**Sample**					
64	4	0.10	4.0	4.53	135	0.271	4	0	--

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65	4	0.10	4.0	4.22	241	0.249	4	0	--
66	4	0.10	4.0	4.50	165	0.320	4	2.58×10^7	Lung (weak), liver (strong), spleen (strong)
70	4	0.10	4.0	4.50	205	0.438	5	6.62×10^6	Lung (strong), spleen (weak)
71	4	0.10	4.0	4.39	237	0.451	5	3.79×10^7	Lung (weak), liver (weak), spleen (strong)
74	4	0.10	4.0	4.46	180	0.317	5	1.71×10^7	Lung (weak), liver (weak), spleen (strong)
75	4	0.10	4.0	4.62	235	0.328	5	3.37×10^7	Lung (weak), liver (weak), spleen (strong)
76	4	0.10	4.0	4.47	78	0.287	5	7.81×10^7	Lung (weak), liver (weak), spleen (strong)
77	4	0.10	4.0	4.50	110	0.267	5	2.44×10^6	Lung (weak), liver (weak), spleen (strong)
78	4	0.10	4.0	4.46	629	1.000	5	1.12×10^5	Lung (strong), spleen (weak)
79	4	0.10	4.0	4.43	352	0.763	6	1.34×10^7	Lung (weak), liver (weak), spleen (strong)
81	4	0.10	4.0	4.38	111	0.287	6	2.32×10^7	Lung (weak), liver (strong), spleen (weak)
82	4	0.10	4.0	4.26	162	0.144	5	1.62×10^6	Spleen (strong)
83	4	0.10	4.0	4.31	143	0.430	5	7.11×10^7	Lung (weak), liver (weak), spleen (strong)
84	4	0.10	4.0	4.44	171	0.303	5	6.89×10^5	Lung (weak), liver (weak), spleen (strong)
85	4	0.10	4.0	4.44	102	0.250	5	2.87×10^6	Lung (weak), liver (weak), spleen (strong)
86	4	0.10	4.0	4.39	91	0.234	5	4.73×10^7	Lung (weak), liver (strong), spleen (weak)
87	4	0.10	4.0	4.47	146	0.486	6	3.24×10^7	Lung (weak), liver (weak), spleen (strong)
88	4	0.10	4.0	4.55	100	0.34	6	$2.34 \times$	Lung (weak),

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						2		10^6	liver (weak), spleen (strong)
89	4	0.10	4.0	4.45	149	0.467	6	5.92×10^7	Lung (weak), liver (weak), spleen (strong)
91	4	0.10	4.0	4.44	188	0.498	6	9.94×10^6	Lung (weak), liver (weak), spleen (strong)
95	4	0.10	4.0	4.38	105	0.352	6	6.61×10^7	Lung (weak), liver (weak), spleen (strong)
96	4	0.10	4.0	4.52	139	0.402	5	4.79×10^7	Lung (weak), liver (weak), spleen (strong)
97	4	0.10	4.0	4.42	204	0.482	5	5.89×10^7	Lung (weak), liver (weak), spleen (strong)
98	4	0.10	4.0	4.35	187	0.221	6	2.06×10^6	Lung (weak), liver (weak), spleen (strong)
99	4	0.10	4.0	4.32	277	0.571	6	4.49×10^6	Lung (weak), liver (weak), spleen (strong)
103	4	0.10	4.0	4.58	88	0.275	6	1.70×10^6	Lung (strong), spleen (weak)
105	4	0.10	4.0	4.42	94	0.294	6	4.74×10^7	Lung (weak), liver (strong), spleen (weak)
106	4	0.10	4.0	4.41	105	0.275	6	3.48×10^7	Lung (weak), liver (weak), spleen (strong)
107	4	0.10	4.0	4.42	146	0.279	6	2.19×10^7	Lung (weak), liver (weak), spleen (strong)
108	4	0.10	4.0	4.54	214	0.334	6	9.66×10^6	Lung (strong), spleen (weak)

pK_a Measurements of individual IAJD molecules is shown in Table 15.

Table 15.

IAJD No.	pK_a
IAJD 64	7.06
IAJD 65	7.40
IAJD 66	6.37
IAJD 70	6.44
IAJD 71	6.38
IAJD 74	6.32
IAJD 75	6.22

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IAJD 76	6.28
IAJD 77	6.40
IAJD 78	6.46
IAJD 79	6.25
IAJD 81	6.19
IAJD 82	6.18
IAJD 83	6.30
IAJD 84	6.19
IAJD 85	6.21
IAJD 86	6.24
IAJD 87	6.49
IAJD 88	6.28
IAJD 89	6.45
IAJD 91	6.48
IAJD 95	6.33
IAJD 96	6.32
IAJD 97	6.46
IAJD 98	6.25
IAJD 99	6.31
IAJD 103	6.23
IAJD 105	6.50
IAJD 106	6.38
IAJD 107	6.42
IAJD 108	6.59

Titration curves showing changes in solution pH in response to addition of a strong acid for IAJD molecules is shown in Figure 54 through Figure 57.

A comparison of i*n vitro vs* i*n vivo* efficacy is shown in Figure 58. No correlations were found between *in vitro* and *in vivo* activities when all data points were considered; positive and negative correlations between *in vitro* and *in vivo* activities were found for two different groups of bias selective data points shown by the lines in Figure 58 (right) (the selection of data points has no scientific base).

Example 3: The Unexpected Importance of the Primary Structure of the Hydrophobic Part of One-Component IAJDs in Targeted mRNA Delivery Activity

The invention relates in part to a one-component multifunctional sequence-defined IAJD delivery system that co-assembles with mRNA by simple injection into DNPs (Figure 1, Figure 59 through Figure 62, and Figure 73 through Figure 75).

The materials and methods employed in these experiments are now described.

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Materials

3,5-Dihydroxybenzoic acid (Acros, 97%), (*rac*)-3-(bromomethyl)heptane, 2-ethylhexyl bromide, (Aldrich, 95%), 1-bromooctane (Aldrich, 99%), 1-bromononane (Lancaster, 99%), 1-bromodecane (Acros, 98%), 1-bromoundecane (Aldrich, 99%), 1-bromododecane (Alfa Aesar, 99%), 1bromotetradecane (Acros, 98%), 1-bromopentadecane (Aldrich, 98%), 1-bromohexadecane (TCI, 96%), 1-bromooctadecane (Acros, 96%), 1-heptadecanol (TCI, 97%), benzyl chloride (Alfa Aesar, 99%), 4toluenesulfonyl chloride (Alfa Aesar, 98%), palladium on activated carbon catalyst (Spectrum, 10 wt% loading), lithium aluminium hydride (LiAlH4, TCI, 95%), 4-bromobutyric acid (Acros, 98%), thionyl chloride (Alfa Aesar, 99+%), potassium phthalimide (Chem Impex, 99.8%), 1-methylpiperazine (Alfa Aesar, 98%), 1-(2-hydroxyethyl)piperazine (Acros, 99%), triethylamine (TCI, 99%), other reagents and solvents for chemical synthesis were obtained from commercial sources and were used as received. CH2Cl2 (DCM) was dried over CaH2 and distilled freshly before use. 4-(Dimethylamino)pyridinium 4toluenesulfonate (DPTS) was prepared according to a literature procedure (Moore and Stupp, *Macromolecules* **1990**, *23*, 65-70). Acetate buffer (10 mM) was prepared by dissolving sodium acetate (2.3 mM) and acetic acid (7.7 mM) in ultra-pure water. Final pH of the buffer was adjusted with 0.1 M HCl or 0.1 M NaOH solution. Nucleoside-modified messenger RNA encoding firefly luciferase (Luc-mRNA) was produced as previously described. Human embryonic kidney (HEK) 293T cells (American Type Culture Collection) were cultured in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% inactivated fetal bovine serum (FBS) (Gemini Bio-Products), 2 mM L–glutamine and 100 U/mL penicillin/streptomycin (Life Technologies). DPBS (Corning), OptiMEM (Gibco), UltraPure DNase/RNase-Free Distilled Water (Invitrogen), Trypsin-EDTA (0.25%, Gibco), Trypan Blue (Sigma-Aldrich), Cell Culture Lysis 5X Reagent (Promega), Luciferase Assay System (Promega) and D-luciferin sodium salt (Regis Technologies) were used as received.

2. Methods and Techniques

The purity and structural identity of intermediate compounds and final products were determined by a combination of characterization techniques including thin-layer chromatography (TLC), ^{1}H and ^{13}C NMR, high-pressure liquid chromatography (HPLC) and matrix assisted laser desorption ionization-time of flight (MALDI-TOF) mass spectrometry. ^{1}H and ^{13}C NMR spectra were recorded at 400 MHz and 101 MHz respectively, on a Bruker NEO (400 MHz) NMR spectrometer with autosampler, or 500 MHz and 126 MHz respectively, on a Bruker DRX (500

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MHz) NMR spectrometer. All NMR spectra were measured at 23°C in $CDCl_3$. Residual protic solvent of $CDCl_3$ (^1H, δ 7.26 ppm; ^{13}C, δ 77.16 ppm) and tetramethylsilane (TMS, δ 0 ppm) are used as the internal reference in the ^1H and ^{13}C NMR spectra. NMR spectra were analyzed by MNova 14 or TopSpin 4.07 (Bruker). TLC was used to monitor the evolution of the reaction by using silica gel 60 F254 precoated plates (E. Merck). Individual compounds with aromatic groups were visualized by UV light (λ = 254 nm). For compounds without aromatic groups, the TLC plate was stained with iodine vapor to help visualization. Purification by flash column chromatography (SiO_2) was performed using silica gel from Silicycle (60 Å, 40–63 μm) with the indicated eluent. The determination of the purity of individual compounds by high-pressure liquid chromatography (HPLC) was performed by using Shimadzu LC20AD high-performance liquid chromatograph pump, a PE Nelson Analytical 900 Series integration data station, a Shimadzu SPD-10A VP (UV-*vis*, λ = 254 nm) and three AM gel columns (a guard column, two 500 Å, 10 μm columns). THF with 5% of NEt_3 was used as the solvent and the characterization was carried out at 23°C. Detection of compounds was determined by UV absorbance (λ = 254 nm) or RI (refractive index) detector. The molar mass of molecules was determined by MALDI-TOF mass spectrometry on a PerSeptive Biosystem-Voyager-DE (Framingham, MA) mass spectrometer equipped with nitrogen laser (337 nm) and operating in linear mode. Angiotensin II and Bombesin were used as standards for internal calibration. For the preparation of sample solution, compound for analysis was dissolved in THF (5–10 mg/mL) firstly. Then the matrix (2,5-dihydroxybenzoic acid) was dissolved in THF 10 mg/mL. Finally, the above two solutions were mixed with a 1/5 (v/v, compound solution/matrix solution) ratio. Then one drop of sample solution was placed on the MALDI plate and dried at 23 °C. Afterwards, the plate was inserted into the vacuum chamber of the instrument for analysis. The laser intensity and voltages applied for the analysis were adjusted based on the molar mass and nature of each analyzed compound.

Dynamic Light Scattering (DLS) for the dimensions (sizes and polydispersities) of DNPs was performed on a Malvern Instruments particle sizer (Zetasizer Nano S, Malvern Instruments, UK) equipped with 4 mW He–Ne laser 633 nm and avalanche photodiode positioned at 175° to the beam and temperature-controlled cuvette holder. Instrument parameters were set up automatically along with measurement times. Sample solution (c.a. 0.4 mL) was placed in a semi-micro cuvette (1.6 mL, polystyrene, 10 × 10 × 45 mm, Greiner Bio-One) and the measurements were performed at 23 °C.

For pKa measurements of individual IAJD molecules, the IAJD molecules were dissolved in ethanol (saturated with NaCl) at a concentration of 1.5 mg/mL and in a volume of 3 mL. Then

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0.1 M HCl solution was added to the above ethanol solution with an increment of 7.5 μL. The resulting pH after each addition of HCl solution was measured by an Thermo Scientific Orion Star A121 meter with Thermo Scientific Orion 8220BNWP pH probe. pKa was determined using the half equivalence point titration.

Formulation of DNPs Co-assembled from IAJDs and Luc-mRNA: nucleoside-modified mRNA encoding firefly luciferase (Luc-mRNA) was dissolved in UltraPure DNase/RNase-free distilled water with an initial concentration of 4.0 mg/mL. IAJD molecules were dissolved in ethanol with an initial concentration of 80 mg/mL. 12.5 μL of Luc-mRNA solution was taken and placed into a clean RNAs free eppendorf (1.5 mL). Then 463 μL of acetate buffer (10 mM, pH 4.0) was added. Afterwards, 25 μL of IAJD in the ethanol stock solution was taken and rapidly injected into the above Luc-mRNA solution in acetate buffer followed by vortex for 5 seconds.

Luminescence Characterization for *In Vitro* Transfection Experiments: the determination of luminescence intensity for *in vitro* Luc-mRNA transfection experiments with HEK 293T Cells was performed using a MiniLumat LB 9506 luminometer (Berthold/EG&G; Wallac).

Luminescence Characterization for *In Vivo* Transfection Experiments: bioluminescence imaging was performed with an IVIS Spectrum imaging system (PerkinElmer, Waltham, MA). Mice were anesthetized with 3% of isoflurane (Piramal Healthcare Limited) and intraperitoneally (i.p.) administered with D-luciferin (Regis Technologies) at a dose of 150 mg/kg of body weight. Ten minutes post administration of D-luciferin, mice were placed on the imaging platform while being maintained on isoflurane *via* a nose cone and imaged using a certain exposure time (60, 30, or 15 seconds).

Bioluminescence values were quantified by measuring photon flux (photons/second, p/s) in the region of interest (ROI) on mice where bioluminescence signal emanated using the Living Image Software (PerkinElmer). To quantify luminescent flux, an oval ROI was placed over each organ of interest and analyzed.

In Vitro mRNA Delivery: for *in vitro* Luc-mRNA transfection screening experiments, human embryonic kidney 293T cells (HEK 293T) were seeded into 96-well plates (20,000 cells/well/200 μL) and cultured for 18-20 hours at 37 °C, 5% CO2 in complete Dulbecco's modified Eagle's medium (DMEM) supplemented with 2 mM L–glutamine (Life Technologies), 100 U/mL penicillin/streptomycin (Invitrogen) and 10% fetal bovine serum (FBS, Gemini Bio-Products). Cells were transfected with DNPs with encapsulated Luc-mRNA at a constant concentration of Luc-mRNA, 125 ng per well. Luc-mRNA encapsulated in FDA approved MC3-based LNPs were used as positive controls for cell transfection, at the concentration of Luc-mRNA

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of 125 ng per well, the same as for tested DNPs. After transfection cells were further cultured at 37 °C, 5% CO2 for 18-20 hours, then medium was aspirated and cells were lysed with luciferase cell culture lysis reagent (30 µL/well) (Promega, Madison, WI). For the determination of the luciferase enzymatic activity as luminescence, 2.5 µL of the lysed cells was mixed with 10 µL of firefly luciferase assay substrate (Luciferase assay system, Promega) and luminescence was analyzed by MiniLumat LB9506 luminometer. Transfections were performed in triplicate.

In Vivo mRNA Delivery in Mice with DNPs: All animals used were in accordance with the guidelines and approval from the Pennsylvania University Institution of Animal Care and Use Committee. Female or male BALB/c mice (6-8 weeks old, from Charles River Laboratories) were anesthetized with isoflurane (Piramal Healthcare Limited) and injected *via* retro-orbital sinus with 100 µL of DNP solution containing 10 µg of Luc-mRNA. At 4-7 hours post injection, mice were i.p. injected with D-Luciferin (150 mg/kg of body weight, Regis Technologies) and imaged on a PerkinElmer IVIS Spectrum CT system (PerkinElmer, Waltham, MA). Tissue luminescence signal was measured on the IVIS imaging system using a certain exposure time (60, 30 or 15 seconds) and medium binning (binning = 8) to ensure that the signal obtained was within operative detection range. For IVIS imaging of the organs, mice were sacrificed, and heart, lungs, liver, and spleen were immediately collected, and bioluminescence imaging was performed as described above. Image analysis was conducted with the Living Image software (PerkinElmer). Bioluminescence values were quantified by measuring photon flux (photons/second) in the region of interest (ROI) using Living Image software.

Scheme 10 (below) shows the synthesis of undecane-based IAJDs with nonsymmetric alkyl chains.

Scheme 10. Reagents and conditions: (*i*) K_2CO_3, KI, DMF, 80 °C, 5 h; (*ii*) RBr, K_2CO_3, DMF, 120 °C, 2 h; (*iii*) H_2, Pd/C, DCM, MeOH, 12 h; (*iv*) $LiAlH_4$, THF, 0–23 °C, 1 h; (*v*) DCC, DPTS, DCM, 12 h; (*vi*) K_2CO_3, MeCN, 95 °C, 3 h; (*vii*) Mg, iodine, THF, 80 °C, 1 h, then CO_2, HCl (aq); (*viii*) $LiAlH_4$, THF, 80 °C, 1 h; (*ix*) 4-toluenesulfonyl chloride, pyridine, DCM, 24 h; (*x*) K_2CO_3, DMF, 80 °C, 12 h; (*xi*) $SOCl_2$, DMF (cat.), DCM, 23 °C, 0.5 h, then potassium phthalimide, DMF, 80 °C, 1 h; (*xii*) Hydrazine monohydrate, EtOH, reflux, 2 h; (*xiii*) 4-(4-methylpiperazin-1-yl)butanoic acid hydrochloride, DCC, DCM, 23 °C, 12 h.

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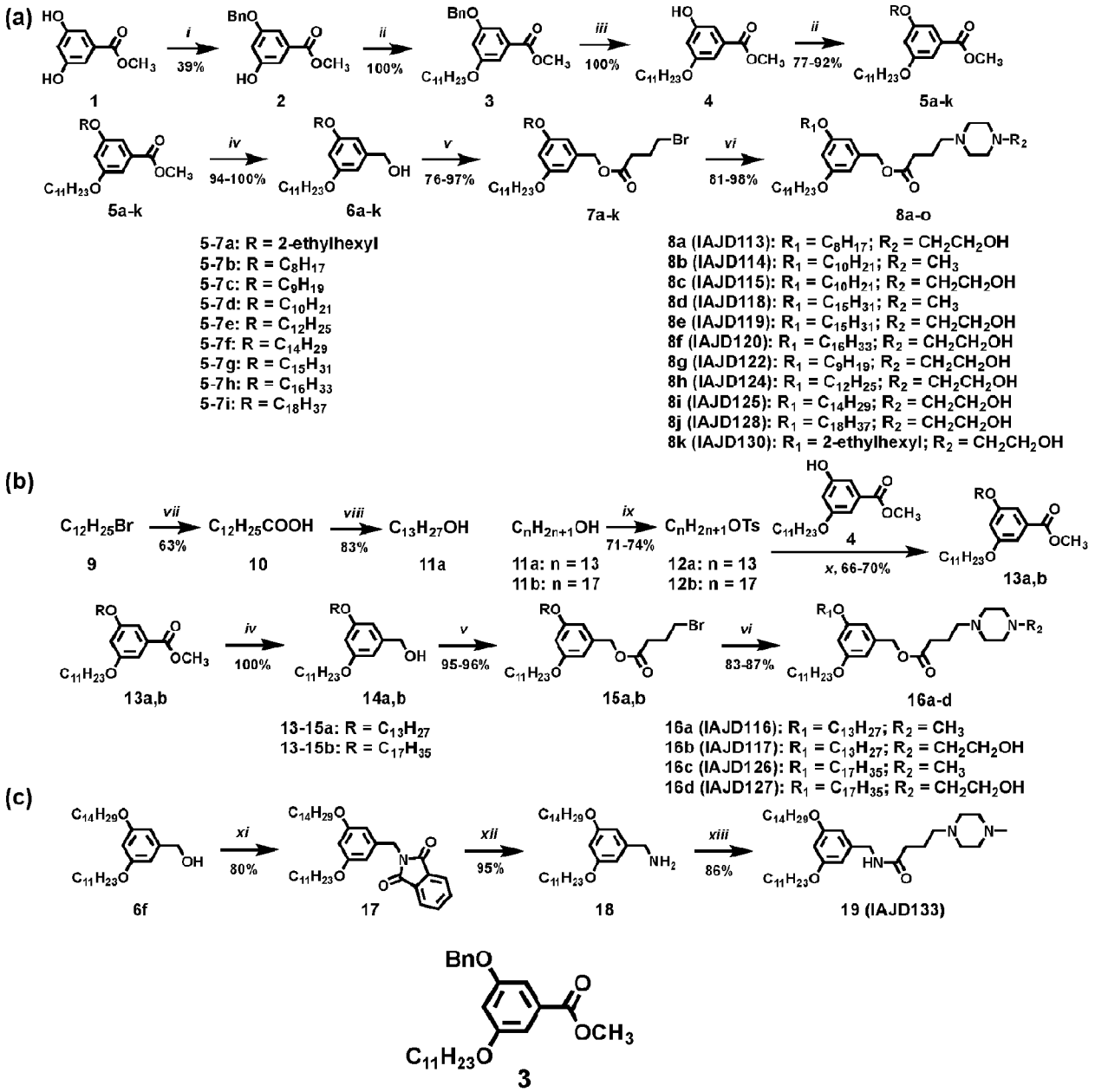

Methyl 3-(benzyloxy)-5-(undecyloxy)benzoate (3). The synthesis of methyl 3-(benzyloxy)-5-hydroxybenzoate (**2**) was adapted from literature procedures. In this work, the procedures were optimized and increased the scale of compound **2** to 23 g. Compound **2** (4.00 g, 15.49 mmol, 1 equiv), 1-bromoundecane (4.01 g, 17.04 mmol, 1.1 equiv) and K_2CO_3 (4.28 g, 30.98 mmol, 2 equiv) were stirred in DMF (50 mL). The mixture was stirred at 120 °C under N_2 atmosphere for 2 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL), three times. The organic phasees were combined, dried over anhydrous $MgSO_4$ and filtered. The filtrate was

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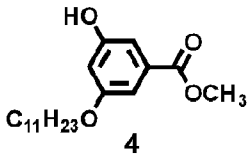
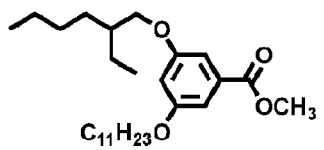
concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the mobile phase to give the title compound as a colorless oil (6.39 g, 100%).



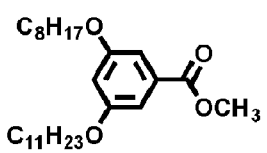
4

Methyl 3-hydroxy-5-(undecyloxy)benzoate (4). Compound **3** (6.39 g, 15.49 mmol) was dissolved in the mixture of DCM (40 mL) and methanol (20 mL). Then Pd/C (0.32 g, 5 wt%) was added and the flask was evacuated and filled with hydrogen, three times. The mixture was stirred at 23 °C for 12 h under hydrogen atmosphere. The reaction mixture was filtered through Celite, and the filter cake was washed with DCM carefully. Evaporation of the solvent yielded the title compound as a white solid (5.02 g, 100%).



5a

Methyl 3-((2-ethylhexyl)oxy)-5-(undecyloxy)benzoate (5a). Compound **5a** was synthesized from compound **4** (1.00 g, 3.10 mmol, 1 equiv), 3-(bromomethyl)heptane (0.72 g, 3.72 mmol, 1.2 equiv) and K_2CO_3 (1.03 g, 7.45 mmol, 2.4 equiv) following a procedure similar to that used for the synthesis of compound **3**. The title compound was obtained as a yellow oil (1.16 g, 92%).



5b

Methyl 3-(octyloxy)-5-(undecyloxy)benzoate (5b). Compound **4** (1.00 g, 3.10 mmol, 1 equiv), 1-bromooctane (0.63 g, 3.26 mmol, 1.05 equiv) and K_2CO_3 (0.87 g, 6.20 mmol, 2 equiv) were stirred in DMF (15 mL). The mixture was stirred at 120 °C under N_2 atmosphere for 2 h. After cooled to 23 °C, the reaction mixture was poured into ice/water (100 mL) and the resulted white precipitates were filtered and collected. Then the precipitates were purified by recrystallization from a minimum amount of acetone to give the title compound as a white solid (1.10 g, 82%).

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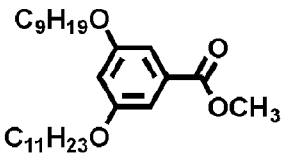
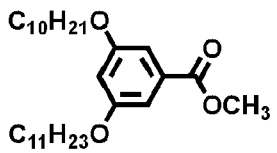


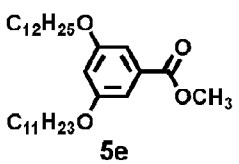
5c

Methyl 3-(nonyloxy)-5-(undecyloxy)benzoate (5c). Compound **5c** was synthesized from compound **4** (1.00 g, 3.10 mmol, 1 equiv), 1-bromononane (0.69 g, 3.31 mmol, 1.1 equiv) and K_2CO_3 (0.92 g, 6.63 mmol, 2.2 equiv) following a procedure similar to that used for the synthesis of compound **5b**. The title compound was obtained as a white solid (1.11 g, 80%).



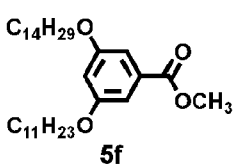
5d

Methyl 3-(decyloxy)-5-(undecyloxy)benzoate (5d). Compound **5d** was synthesized from compound **4** (1.00 g, 3.10 mmol, 1 equiv), 1-bromodecane (0.72 g, 3.26 mmol, 1.05 equiv) and K_2CO_3 (0.87 g, 6.20 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **5b**. The title compound was obtained as a white solid (1.25 g, 87%).



5e

Methyl 3-(dodecyloxy)-5-(undecyloxy)benzoate (5e). Compound **5e** was synthesized from compound **4** (1.00 g, 3.10 mmol, 1 equiv), 1-bromododecane (0.82 g, 3.30 mmol, 1.1 equiv) and K_2CO_3 (0.92 g, 6.63 mmol, 2.2 equiv) following a procedure similar to that used for the synthesis of compound **5b**. The title compound was obtained as a white solid (1.25 g, 82%).



5f

Methyl 3-(tetradecyloxy)-5-(undecyloxy)benzoate (5f). Compound **5f** was synthesized from compound **4** (1.00 g, 3.10 mmol, 1 equiv), 1-bromotetradecane (0.90 g, 3.26 mmol, 1.05 equiv) and K_2CO_3 (0.87 g, 6.20 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **5b**. The title compound was obtained as a white solid (1.32 g, 82%).

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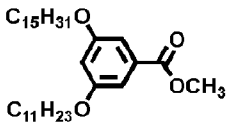
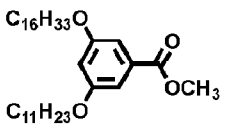


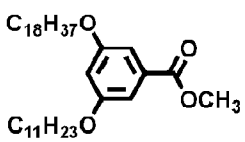
5g

Methyl 3-(pentadecyloxy)-5-(undecyloxy)benzoate (5g). Compound **5g** was synthesized from compound **4** (1.00 g, 3.10 mmol, 1 equiv), 1-bromopentadecane (0.95 g, 3.26 mmol, 1.05 equiv) and K_2CO_3 (0.87 g, 6.20 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **5b**. The title compound was obtained as a white solid (1.37 g, 83%).



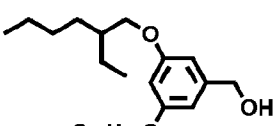
5h

Methyl 3-(hexadecyloxy)-5-(undecyloxy)benzoate (5h). Compound **5h** was synthesized from compound **4** (1.00 g, 3.10 mmol, 1 equiv), 1-bromohexadecane (1.01 g, 3.31 mmol, 1.1 equiv) and K_2CO_3 (0.92 g, 6.63 mmol, 2.2 equiv) following a procedure similar to that used for the synthesis of compound **5b**. The title compound was obtained as a white solid (1.30 g, 77%).



5i

Methyl 3-(octadecyloxy)-5-(undecyloxy)benzoate (5i). Compound **5i** was synthesized from compound **4** (1.00 g, 3.10 mmol, 1 equiv), 1-bromoctadecanene (1.09 g, 3.26 mmol, 1.05 equiv) and K_2CO_3 (0.87 g, 6.20 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **5b**. The title compound was obtained as a white solid (1.63 g, 92%).



6a

(3-((2-Ethylhexyl)oxy)-5-(undecyloxy)phenyl)methanol (6a). Compound **5a** (1.14 g, 2.62 mmol, 1 equiv) was dissolved in 5 mL dry THF, which was added dropwise to a slurry of $LiAlH_4$ (78 mg, 2.06 mmol, 0.8 equiv) in dry THF (5 mL) at 0 °C under N_2 atmosphere. The resulted mixture was stirred at 23 °C for 1 h. The reaction was quenched by the successive addition of water (0.3 mL), 15% NaOH aqueous solution (0.3 mL) and water (1.5 mL). Then the white precipitates in the

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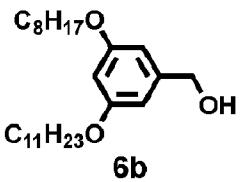
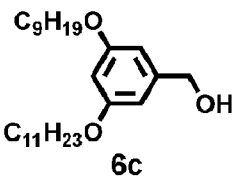
mixture were filtered out and the filtrate was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (1.02 g, 96%).



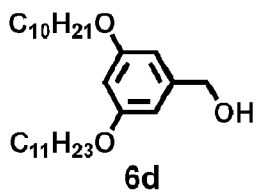
6b

(3-(Octyloxy)-5-(undecyloxy)phenyl)methanol (6b). Compound **6b** was synthesized from compound **5b** (0.92 g, 2.12 mmol, 1 equiv) and $LiAlH_4$ (80 mg, 2.12 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **6a**. The title compound was obtained as a colorless oil (0.82 g, 95%).



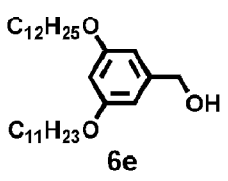
6c

(3-(Nonyloxy)-5-(undecyloxy)phenyl)methanol (6c). Compound **6c** was synthesized from compound **5c** (1.00 g, 2.23 mmol, 1 equiv) and $LiAlH_4$ (85 mg, 2.23 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **6a**. The title compound was obtained as a colorless oil (0.88 g, 94%).



6d

(3-(Decyloxy)-5-(undecyloxy)phenyl)methanol (6d). Compound **6d** was synthesized from compound **5d** (1.05 g, 2.27 mmol, 1 equiv) and $LiAlH_4$ (86 mg, 2.25 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **6a**. The title compound was obtained as a colorless oil (0.98 g, 98%).



6e

(3-(Dodecyloxy)-5-(undecyloxy)phenyl)methanol (6e). Compound **6e** was synthesized from compound **5e** (1.15 g, 2.34 mmol, 1 equiv) and $LiAlH_4$ (89 mg, 2.34 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **6a**. The title compound was obtained

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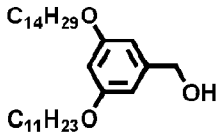
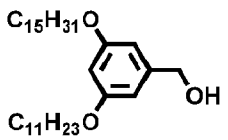
as a colorless oil (1.08 g, 99%).



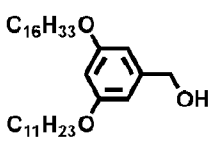
6f

(3-(Tetradecyloxy)-5-(undecyloxy)phenyl)methanol (6f). Compound **6f** was synthesized from compound **5f** (1.15 g, 2.21 mmol, 1 equiv) and $LiAlH_4$ (84 mg, 2.21 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **6a**. The title compound was obtained as a colorless oil (1.08 g, 99%).



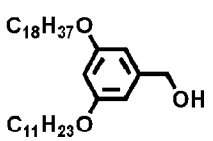
6g

(3-(Pentadecyloxy)-5-(undecyloxy)phenyl)methanol (6g). Compound **6g** was synthesized from compound **5g** (1.12 g, 2.10 mmol, 1 equiv) and $LiAlH_4$ (79 mg, 2.10 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **6a**. The title compound was obtained as a white solid (1.06 g, 100%).



6h

(3-(Hexadecyloxy)-5-(undecyloxy)phenyl)methanol (6h). Compound **6h** was synthesized from compound **5h** (1.10 g, 2.01 mmol, 1 equiv) and $LiAlH_4$ (76 mg, 2.01 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **6a**. The title compound was obtained as a white solid (1.03 g, 99%).



6i

(3-(Octadecyloxy)-5-(undecyloxy)phenyl)methanol (6i). Compound **6i** was synthesized from compound **5i** (1.15 g, 2.05 mmol, 1 equiv) and $LiAlH_4$ (78 mg, 2.05 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **6a**. The title compound was obtained as a white solid (1.32 g, 94%).

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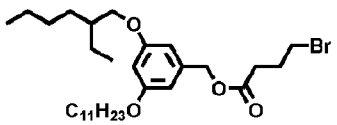
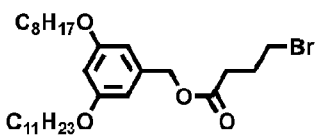


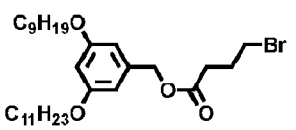
7a

3-((2-Ethylhexyl)oxy)-5-(undecyloxy)benzyl 4-bromobutanoate (7a). Compound **6a** (0.50 g, 1.23 mmol, 1 equiv), 4-bromobutyric acid (0.23 g, 1.35 mmol, 1.1 equiv) and DPTS (0.40 g, 1.35 mmol, 1.1 equiv) were dissolved in 8 mL DCM. *N,N'*-Dicyclohexylcarbodiimide (DCC, 0.51 g, 2.46 mmol, 2 equiv) was added in one portion into the above mixture. The reaction mixture was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/DCM = 1/1 as the eluent to give the title compound as a colorless oil (0.61 g, 90%).



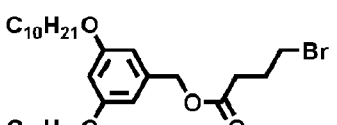
7b

3-(Octyloxy)-5-(undecyloxy)benzyl 4-bromobutanoate (7b). Compound **7b** was synthesized from compound **6b** (0.40 g, 0.98 mmol, 1 equiv), 4-bromobutyric acid (0.18 g, 1.08 mmol, 1.1 equiv), DPTS (0.32 g, 1.08 mmol, 1.1 equiv) and DCC (0.41 g, 1.97 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **7a**. The title compound was obtained as a colorless oil (0.53 g, 97%).



7c

3-(Nonyloxy)-5-(undecyloxy)benzyl 4-bromobutanoate (7c). Compound **7c** was synthesized from compound **6c** (0.40 g, 0.95 mmol, 1 equiv), 4-bromobutyric acid (0.16 g, 0.95 mmol, 1 equiv), DPTS (0.28 g, 0.95 mmol, 1 equiv) and DCC (0.39 g, 1.90 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **7a**. The title compound was obtained as a colorless oil (0.47 g, 87%).



7d

3-(Decyloxy)-5-(undecyloxy)benzyl 4-bromobutanoate (7d). Compound **7d** was synthesized

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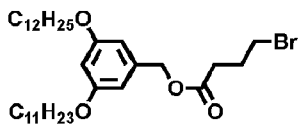
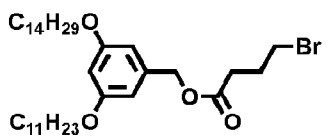
from compound **6d** (0.40 g, 0.92 mmol, 1 equiv), 4-bromobutyric acid (0.17 g, 1.01 mmol, 1.1 equiv), DPTS (0.30 g, 1.01 mmol, 1.1 equiv) and DCC (0.38 g, 1.84 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **7a**. The title compound was obtained as a colorless oil (0.51 g, 95%).



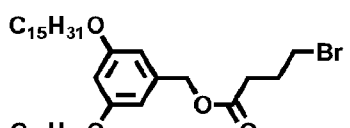
7e

3-(Dodecyloxy)-5-(undecyloxy)benzyl 4-bromobutanoate (7e). Compound **7e** was synthesized from compound **6e** (0.40 g, 0.86 mmol, 1 equiv), 4-bromobutyric acid (0.16 g, 0.95 mmol, 1.1 equiv), DPTS (0.28 g, 0.95 mmol, 1.1 equiv) and DCC (0.36 g, 1.73 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **7a**. The title compound was obtained as a colorless oil (0.50 g, 95%).



7f

3-(Tetradecyloxy)-5-(undecyloxy)benzyl 4-bromobutanoate (7f). Compound **7f** was synthesized from compound **6f** (0.40 g, 0.82 mmol, 1 equiv), 4-bromobutyric acid (0.15 g, 0.90 mmol, 1.1 equiv), DPTS (0.26 g, 0.90 mmol, 1.1 equiv) and DCC (0.34 g, 1.63 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **7a**. The title compound was obtained as a colorless oil (0.45 g, 86%).



7g

3-(Pentadecyloxy)-5-(undecyloxy)benzyl 4-bromobutanoate (7g). Compound **7g** was synthesized from compound **6g** (0.40 g, 0.79 mmol, 1 equiv), 4-bromobutyric acid (0.145 g, 0.87 mmol, 1.1 equiv), DPTS (0.256 g, 0.87 mmol, 1.1 equiv) and DCC (0.33 g, 1.58 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **7a**. The title compound was obtained as a colorless oil (0.48 g, 93%).

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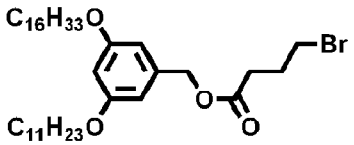
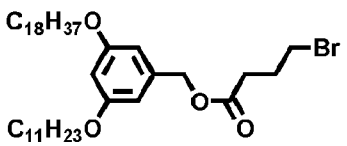


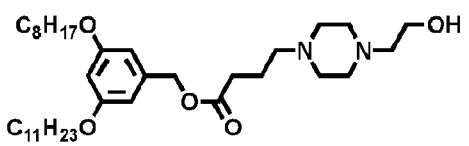
7h

3-(Hexadecyloxy)-5-(undecyloxy)benzyl 4-bromobutanoate (7h). Compound **7h** was synthesized from compound **6h** (0.40 g, 0.77 mmol, 1 equiv), 4-bromobutyric acid (0.13 g, 0.77 mmol, 1 equiv), DPTS (0.23 g, 0.77 mmol, 1 equiv) and DCC (0.32 g, 1.54 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **7a**. The title compound was obtained as a colorless oil (0.39 g, 76%).



7i

3-(Octadecyloxy)-5-(undecyloxy)benzyl 4-bromobutanoate (7i). Compound **7i** was synthesized from compound **6i** (0.40 g, 0.73 mmol, 1 equiv), 4-bromobutyric acid (0.14 g, 0.81 mmol, 1.1 equiv), DPTS (0.24 g, 0.81 mmol, 1.1 equiv) and DCC (0.30 g, 1.43 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **7a**. The title compound was obtained as a white solid (0.49 g, 97%).



8a (IAJD113)

3-(Octyloxy)-5-(undecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (8a, IAJD113). Compound **7b** (0.25 g, 0.45 mmol, 1.0 equiv), 1-(2-hydroxyethyl)piperazine (61 mg, 0.47 mmol, 1.05 equiv), K_2CO_3 (65 mg, 0.47 mmol, 1.05 equiv) were stirred in MeCN (15 mL). The mixture was heated at 95 °C for 3 h. The reaction mixture was cooled to 23 °C and solvent was removed under reduced pressure. Then water (20 mL) was added, and the resulted mixture was extracted by DCM (20 mL) for three times. The organic phase was combined, dried over anhydrous $MgSO_4$, filtered and dried to give the crude product. The crude product was purified by column chromatography (SiO_2) with DCM/MeOH = 30/1 and 15/1 as the mobile phase. Then the obtained product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was collected and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the

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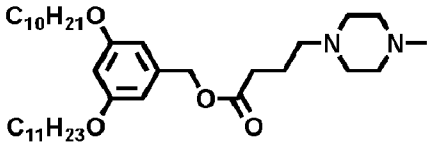
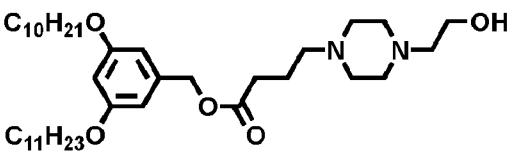
title compound as a light-yellow oil (0.25 g, 93%).



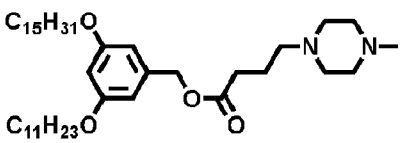
8b (IAJD114)

3-(Decyloxy)-5-(undecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (8b, IAJD114). Compound **8b** was synthesized from compound **7d** (0.24 g, 0.41 mmol), 1-methylpiperazine (49 mg, 0.49 mmol) and K_2CO_3 (68 mg, 0.49 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a light-yellow oil (0.24 g, 88%).



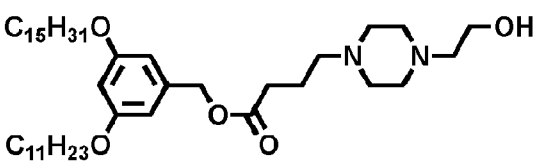
8c (IAJD115)

3-(Decyloxy)-5-(undecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (8c, IAJD115). Compound **8c** was synthesized from compound **7d** (0.24 g, 0.41 mmol), 1-(2-hydroxyethyl)piperazine (64 mg, 0.49 mmol) and K_2CO_3 (68 mg, 0.49 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a light-yellow oil (0.24 g, 92%).



8d (IAJD118)

3-(Pentadecyloxy)-5-(undecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (8d, IAJD118). Compound **8d** was synthesized from compound **7g** (0.22 g, 0.34 mmol), 1-methylpiperazine (38 mg, 0.37 mmol) and K_2CO_3 (70 mg, 0.51 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a colorless oil (0.20 g, 87%).



8e (IAJD119)

3-(Pentadecyloxy)-5-(undecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (8e, IAJD119). Compound **8e** was synthesized from compound **7g** (0.20 g, 0.30 mmol), 1-(2-

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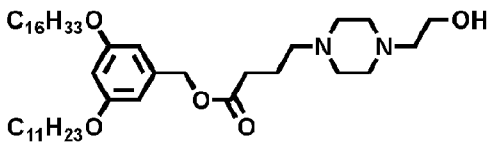
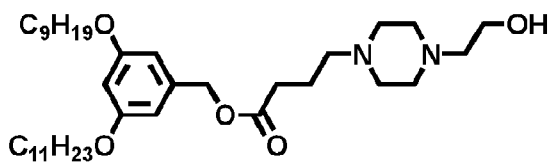
hydroxyethyl)piperazine (43 mg, 0.33 mmol) and K_2CO_3 (62 mg, 0.45 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a colorless oil (0.16 g, 81%).



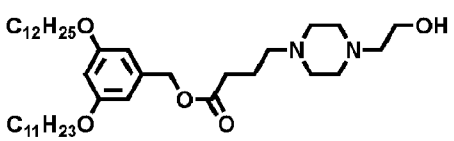
8f (IAJD120)

3-(Hexadecyloxy)-5-(undecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (8f, IAJD120). Compound **8f** was synthesized from compound **7h** (0.20 g, 0.30 mmol), 1-(2-hydroxyethyl)piperazine (51 mg, 0.39 mmol) and K_2CO_3 (57 mg, 0.41 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a light-yellow oil (0.21 g, 98%).



8g (IAJD122)

3-(Nonyloxy)-5-(undecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (8g, IAJD122). Compound **8g** was synthesized from compound **7c** (0.13 g, 0.23 mmol), 1-(2-hydroxyethyl)piperazine (31 mg, 0.24 mmol) and K_2CO_3 (33 mg, 0.24 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a light-yellow oil (0.12 g, 86%).



8h (IAJD124)

3-(Dodecyloxy)-5-(undecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (8h, IAJD124). Compound **8h** was synthesized from compound **7e** (0.20 g, 0.33 mmol), 1-(2-hydroxyethyl)piperazine (64 mg, 0.49 mmol) and K_2CO_3 (68 mg, 0.49 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a light-yellow oil (0.20 g, 93%).

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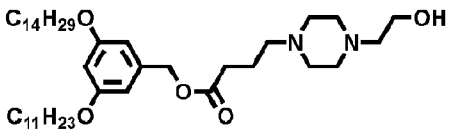
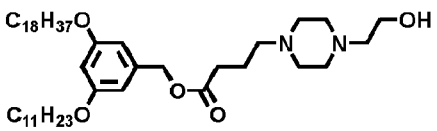


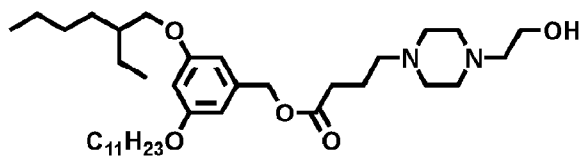
8i (IAJD125)

3-(Tetradecyloxy)-5-(undecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (8i, IAJD125). Compound **8i** was synthesized from compound **7f** (0.20 g, 0.31 mmol), 1-(2-hydroxyethyl)piperazine (45 mg, 0.34 mmol) and K_2CO_3 (47 mg, 0.34 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a light-yellow oil (0.19 g, 88%).



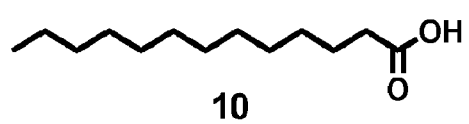
8j (IAJD128)

3-(Octadecyloxy)-5-(undecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (8j, IAJD128). Compound **8j** was synthesized from compound **7i** (0.24 g, 0.41 mmol), 1-(2-hydroxyethyl)piperazine (64 mg, 0.49 mmol) and K_2CO_3 (68 mg, 0.49 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a light-yellow oil (0.23 g, 90%).



8k (IAJD130)

3-((2-Ethylhexyl)oxy)-5-(undecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (8k, IAJD130). Compound **8k** was synthesized from compound **7a** (0.24 g, 0.43 mmol), 1-(2-hydroxyethyl)piperazine (68 mg, 0.52 mmol) and K_2CO_3 (72 mg, 0.52 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a light-yellow oil (0.24 g, 92%).



10

Tridecanoic acid (10, $C_{12}H_{25}COOH$). To a 250 mL two-necked round bottom flask, magnesium (1.00 g, 40 mmol) and catalytic amount of iodine (1 mg) were added. 1-Bromododecane (10.00 g, 40 mmol) in 50 mL of dry THF was mixed in an additional funnel and installed to one of the necks

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of the flask. The other neck of flask was connected to a condenser under N_2. Part solution of 1-bromododecane in THF (10 mL) was added to the flask, and the reaction mixture was heated at 80 °C. Once the color of iodine disappeared and reaction mixture became cloudy, the rest of THF solution of 1-bromododecane was added dropwise. The reaction mixture was maintained at 80 °C for 1 hour until most of the magnesium was consumed. The reaction mixture was cooled to 23°C and poured onto crushed dry ice (100 g). Once all the dry ice disappeared, hydrochloride acid (2M) was added to the reaction mixture until pH = 1. The mixture was extracted with EtOAc (50 mL × 3), dried with anhydrous $MgSO_4$, filtered and concentrated, and then purified by column chromatography on silica gel with a mobile phase of hexane/EtOAc = 10/1 (v/v) to afford the title compound as a colorless oil (5.40 g, 63%).

11a

Tridecan-1-ol (11a, $C_{13}H_{27}OH$). Compound **10** (13.00 g, 61 mmol) were dissolved in dry THF (50 mL), which was added dropwise to a slurry of $LiAlH_4$ (3.60 g, 91 mmol) in dry THF under N_2 atmosphere. The resulted mixture was reflux at 80 °C for 1 h. The reaction was quenched by the successive addition of water, 15% NaOH aqueous solution and water. Then the mixture was filtered and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (10.10 g, 83%) without further purification.

12a

Tridecyl 4-methylbenzenesulfonate (12a, $C_{13}H_{27}OTs$). Compound **11a** (10.10 g, 50 mmol) and pyridine (6.00 g, 75.60 mmol) were dissolved in dry DCM (50 mL), which was added 4-toluenesulfonyl chloride (10.50 g, 55.10 mmol) in dry DCM at 23 °C. The resulted mixture was stirred at 23 °C for 24 h. Ethyl acetate (50 mL) was added, and the precipitates were removed by filtration. The filtrate was washed with saturated $NaHCO_3$ solution, hydrochloride acid (2M), and brine, dried with anhydrous $MgSO_4$, filtered and concentrated. Cold methanol (100 mL) was added to precipitate the title compound as a white solid without further purification (12.60 g, 71%).

12b

Heptadecyl 4-methylbenzenesulfonate (12b, $C_{17}H_{35}OTs$). Compound **12b** was synthesized from 1-heptadecanol (7.00 g, 27.30 mmol), 4-toluenesulfonyl chloride (6.76 g, 35.50 mmol) and pyridine

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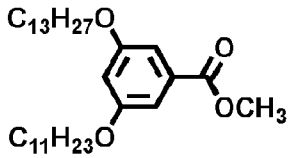
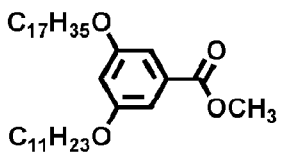
(3.00 g, 37.80 mmol) following a procedure similar to that used for the synthesis of compound **12a**. The title compound was obtained as a white solid (8.30 g, 74%).



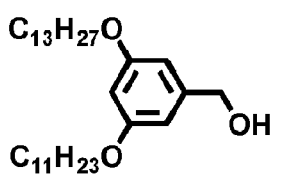
13a

Methyl 3-(tridecyloxy)-5-(undecyloxy)benzoate (13a). Compound **4** (1.00 g, 3.10 mmol, 1 equiv), $C_{13}H_{27}OTs$ (**12a**, 1.16 g, 3.26 mmol, 1.05 equiv) and K_2CO_3 (0.87 g, 6.20 mmol, 2 equiv) were stirred in DMF (10 mL). The mixture was stirred at 80 °C under N_2 atmosphere for 12 h. After cooled to 23 °C, the reaction mixture was poured into ice/water (100 mL) and the resulted white precipitates were filtered and collected. Then the precipitates were purified by recrystallization from minimum acetone to give the title compound as a white solid (1.03 g, 66%).



13b

Methyl 3-(heptadecyloxy)-5-(undecyloxy)benzoate (13b). Compound **13b** was synthesized from compound **4** (1.00 g, 3.10 mmol), $C_{17}H_{35}OTs$ (**12b**, 1.34 g, 3.26 mmol) and K_2CO_3 (0.87 g, 6.20 mmol) following a procedure similar to that used for the synthesis of compound **13a**. The title compound was obtained as a white solid (1.20 g, 70%).



14a

(3-(Tridecyloxy)-5-(undecyloxy)phenyl)methanol (14a). Compound **13a** (0.90 g, 1.78 mmol, 1 equiv) was dissolved in 5 mL dry THF, which was added dropwise to a slurry of $LiAlH_4$ (67 mg, 1.78 mmol, 1 equiv) in dry THF (5 mL) at 0 °C under N_2 atmosphere. The resulted mixture was stirred at 23 °C for 1 h. The reaction was quenched by the successive addition of water (0.3 mL), 15% NaOH aqueous solution (0.3 mL) and water (1.5 mL). Then the white precipitates in the mixture were filtered out and the filtrate was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.85 g, 100%).

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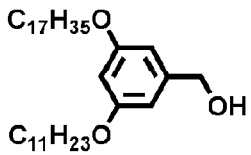
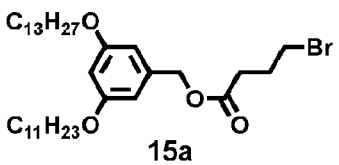


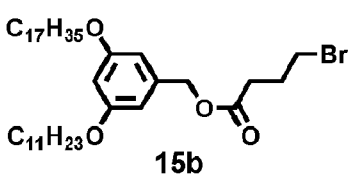
14b

(3-(Heptadecyloxy)-5-(undecyloxy)phenyl)methanol (14b). Compound **14b** was synthesized from compound **13b** (1.15 g, 2.05 mmol, 1 equiv) and $LiAlH_4$ (78 mg, 2.05 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **14a**. The title compound was obtained as a white solid (1.09 g, 100%).



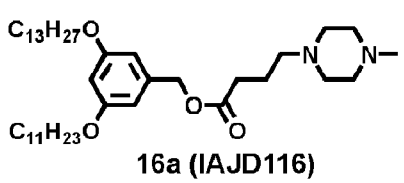
15a

3-(Tridecyloxy)-5-(undecyloxy)benzyl 4-bromobutanoate (15a). Compound **14a** (0.40 g, 0.84 mmol, 1 equiv), 4-bromobutyric acid (0.15 g, 0.92 mmol, 1.1 equiv) and DPTS (0.27 g, 0.92 mmol, 1.1 equiv) were dissolved in 8 mL DCM. DCC (0.35 g, 1.68 mmol, 2 equiv) was added in one portion into the above mixture. The reaction mixture was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with Hexane/DCM = 1/1 as the mobile phase to give the title compound as a colorless oil (0.50 g, 95%).



15b

3-(Heptadecyloxy)-5-(undecyloxy)benzyl 4-bromobutanoate (15b). Compound **15b** was synthesized from compound **14b** (0.40 g, 0.75 mmol, 1 equiv), 4-bromobutyric acid (0.14 g, 0.83 mmol, 1.1 equiv), DPTS (0.26 g, 0.83 mmol, 1.1 equiv) and DCC (0.31 g, 1.50 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **15a**. The title compound was obtained as a white solid (0.49 g, 96%).



16a (IAJD116)

3-(Tridecyloxy)-5-(undecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (16a, IAJD116).

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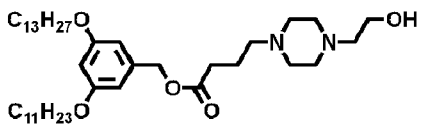
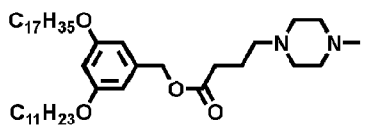
Compound **16a** was synthesized from compound **15a** (0.26 g, 0.36 mmol), 1-methylpiperazine (43 mg, 0.43 mmol) and K_2CO_3 (60 mg, 0.43 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a light-yellow oil (0.19 g, 83%).



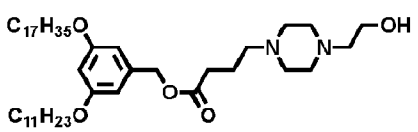
16b (IAJD117)

3-(Tridecyloxy)-5-(undecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (16b, IAJD117). Compound **16b** was synthesized from compound **15a** (0.20 g, 0.31 mmol), 1-(2-hydroxyethyl)piperazine (56 mg, 0.43 mmol) and K_2CO_3 (60 mg, 0.43 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a light-yellow oil (0.18 g, 86%).



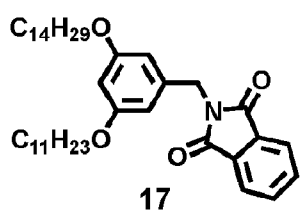
16c (IAJD126)

3-(Heptadecyloxy)-5-(undecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (16c, IAJD126). Compound **16c** was synthesized from compound **15b** (0.22 g, 0.33 mmol), 1-methylpiperazine (36 mg, 0.36 mmol) and K_2CO_3 (68 mg, 0.50 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a colorless oil (0.20 g, 87%).



16d (IAJD127)

3-(Heptadecyloxy)-5-(undecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (16d, IAJD127). Compound **16d** was synthesized from compound **15b** (0.21 g, 0.30 mmol), 1-(2-hydroxyethyl)piperazine (40 mg, 0.33 mmol) and K_2CO_3 (62 mg, 0.45 mmol) following a procedure similar to that used for the synthesis of compound **8a**. The title compound was obtained as a colorless oil (0.20 g, 87%).



17

2-(3-(Tetradecyloxy)-5-(undecyloxy)benzyl)isoindoline-1,3-dione (17). To a DCM solution (10

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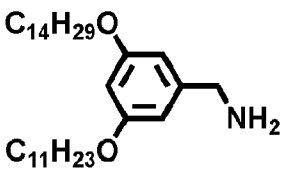
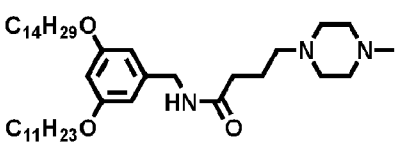
mL) of compound **6f** (0.40 g, 0.82 mmol, 1.0 equiv) was added 1 drop of DMF, followed by the dropwise addition of $SOCl_2$ (0.15 g, 1.22 mmol, 1.5 equiv). The reaction mixture was stirred at 23 °C for 0.5 h. Then DCM and excess $SOCl_2$ was removed under reduced pressure. The obtained intermediate product was dissolved in dry DMF (10 mL) and potassium phthalimide (0.18 g, 0.98 mmol, 1.2 equiv) was added. The reaction mixture was stirred at 80 °C under N_2 atmosphere for 1 h. The mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (20 mL) was added, and the mixture was extracted by DCM (20 mL) for three times. The organic phase was combined, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the mobile phase to give the title compound as a light-yellow oil (0.40 g, 80%).



18

(3-(Tetradecyloxy)-5-(undecyloxy)phenyl)methanamine (18). Compound **17** (0.38 g, 0.61 mmol, 1 equiv) and hydrazine monohydrate ($NH_2NH_2•H_2O$, 0.31 g, 6.10 mmol, 10 equiv) were dissolved in EtOH (10 mL). The reaction mixture was heated at reflux for 2 h. The mixture was cooled to 23 °C and the solvent was removed under reduced pressure. Then water (10 mL) was added, and the mixture was extracted by DCM (10 mL) for three times. The organic phase was combined, dried over anhydrous $MgSO_4$ and filtered. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.28 g, 95%).



19 (IAJD133)

4-(4-Methylpiperazin-1-yl)-N-(3-(tetradecyloxy)-5-(undecyloxy)benzyl)butanamide (19, IAJD133). The synthesis and characterizations of 4-(4-methylpiperazin-1-yl)butanoic acid hydrochloride were available in the literature.[4] Compound **18** (0.27 g, 0.55 mmol, 1 equiv) and 4-(4-methylpiperazin-1-yl)butanoic acid hydrochloride (0.12 g, 0.55 mmol, 1 equiv) were dissolved in 6 mL dry DCM. NEt_3 (3 drops) was added. DCC (0.23 g, 1.10 mmol, 2 equiv) was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. Afterwards, the resulted white precipitates (urea) were removed by filtration and washed with DCM carefully. The

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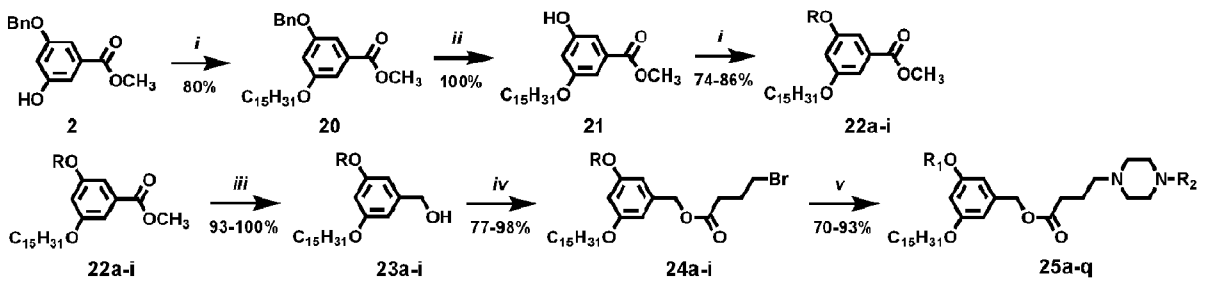
filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 30/1, 10/1, 8/1 and 8/1 with 0.1% NEt_3 as the mobile phase. Then the product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was combined and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a white solid (0.31 g, 86%).

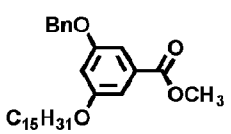
Synthesis of Pentadecane-Based IAJDs with Nonsymmetric Alkyl Chains was performed according to Scheme 11 shown below:

Scheme 11. Reagents and conditions: (*i*) RBr/ROTs, K_2CO_3, DMF, 120 °C, 2 h; (*ii*) H_2, Pd/C, DCM, MeOH, 12 h; (*iii*) $LiAlH_4$, THF, 0–23 °C, 1 h; (*iv*) 4-Bromobutyric acid, $SOCl_2$, DMF (cat.), DCM, 23 °C, 1 h, then DMAP, NEt_3, DCM, 0–23 °C, 2 h, or DCC, DPTS, 12 h; (*v*) K_2CO_3, MeCN, 95 °C, 3 h.



22-24a: R = 2-ethylhexyl
22-24b: R = C_8H_{17}
22-24c: R = $C_{10}H_{21}$
22-24d: R = $C_{12}H_{25}$
22-24e: R = $C_{13}H_{27}$
22-24f: R = $C_{14}H_{29}$
22-24g: R = $C_{16}H_{33}$
22-24h: R = $C_{17}H_{35}$
22-24i: R = $C_{18}H_{37}$

25a (IAJD135): R_1 = 2-ethylhexyl; R_2 = CH_3
25b (IAJD136): R_1 = 2-ethylhexyl; R_2 = CH_2CH_2OH
25c (IAJD138): R_1 = C_8H_{17}; R_2 = CH_2CH_2OH
25d (IAJD141): R_1 = $C_{10}H_{21}$; R_2 = CH_3
25e (IAJD142): R_1 = $C_{10}H_{21}$; R_2 = CH_2CH_2OH
25f (IAJD143): R_1 = $C_{12}H_{25}$; R_2 = CH_3
25g (IAJD144): R_1 = $C_{12}H_{25}$; R_2 = CH_2CH_2OH
25h (IAJD145): R_1 = $C_{13}H_{27}$; R_2 = CH_3
25i (IAJD146): R_1 = $C_{13}H_{27}$; R_2 = CH_2CH_2OH
25j (IAJD147): R_1 = $C_{14}H_{29}$; R_2 = CH_3
25k (IAJD148): R_1 = $C_{14}H_{29}$; R_2 = CH_2CH_2OH
25l (IAJD149): R_1 = $C_{16}H_{33}$; R_2 = CH_3
25m (IAJD150): R_1 = $C_{16}H_{33}$; R_2 = CH_2CH_2OH
25n (IAJD151): R_1 = $C_{17}H_{35}$; R_2 = CH_3
25o (IAJD152): R_1 = $C_{17}H_{35}$; R_2 = CH_2CH_2OH
25p (IAJD153): R_1 = $C_{18}H_{37}$; R_2 = CH_3
25q (IAJD154): R_1 = $C_{18}H_{37}$; R_2 = CH_2CH_2OH



20

Methyl 3-(benzyloxy)-5-(pentadecyloxy)benzoate (20). Compound **2** (5.00 g, 19.36 mmol, 1 equiv), 1-bromopentadecane (6.20 g, 21.30 mmol, 1.1 equiv) and K_2CO_3 (5.35 g, 38.72 mmol, 2

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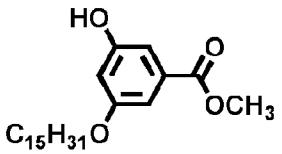
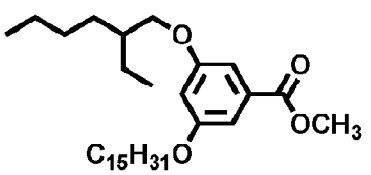
equiv) were stirred in DMF (40 mL). The mixture was stirred at 120 °C under N_2 atmosphere for 2 h. After cooled to 23 °C, the reaction mixture was poured into ice/water (150 mL) and the resulted white precipitates were filtered and collected. Then the precipitates were purified by recrystallization from minimum acetone to give the title compound as a white solid (7.20 g, 80%).



21

Methyl 3-hydroxy-5-(pentadecyloxy)benzoate (21). Compound **20** (7.20 g, 15.36 mmol) was dissolved in a mixture of DCM (40 mL) and methanol (20 mL). Then Pd/C (0.36 g, 5 wt%) was added and the flask was evacuated and filled with hydrogen for three times. The mixture was stirred at 23 °C for 12 h under hydrogen atmosphere. The reaction mixture was filtered through Celite, and the filter cake was washed with DCM carefully. Evaporation of the solvent yielded the title compound as a white solid (5.81 g, 100%).



22a

Methyl 3-((2-ethylhexyl)oxy)-5-(pentadecyloxy)benzoate (22a). Compound **21** (0.50 g, 1.32 mmol, 1 equiv), 3-(bromomethyl)heptane (0.27 g, 1.39 mmol, 1.05 equiv) and K_2CO_3 (0.37 g, 2.64 mmol, 2 equiv) were stirred in DMF (15 mL). The mixture was stirred at 80 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (20 mL) was added, and the mixture was extracted with DCM (20 mL) for three times. The organic phases were combined, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the mobile phase to give the title compound as a light-yellow oil (0.48 g, 74%).

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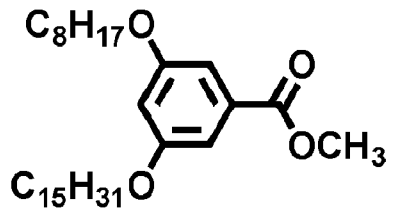
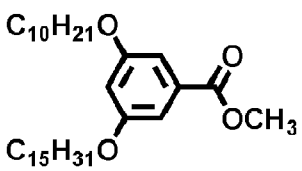


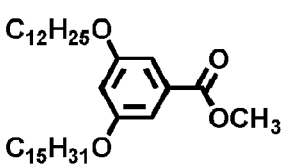
22b

Methyl 3-(octyloxy)-5-(pentadecyloxy)benzoate (22b). Compound **21** (0.50 g, 1.32 mmol, 1 equiv), 1-bromooctane (0.27 g, 1.39 mmol, 1.05 equiv) and K_2CO_3 (0.37 g, 2.64 mmol, 2 equiv) were stirred in DMF (15 mL). The mixture was stirred at 120 °C under N_2 atmosphere for 2 h. After cooled to 23 °C, the reaction mixture was poured into ice/water (100 mL) and the resulted white precipitates were filtered and collected. Then the precipitates were purified by recrystallization from minimum acetone to give the title compound as a white solid (0.48 g, 84%).



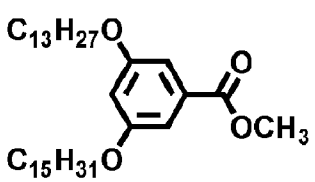
22c

Methyl 3-(decyloxy)-5-(pentadecyloxy)benzoate (22c). Compound **22c** was synthesized from compound **21** (0.50 g, 1.32 mmol, 1 equiv), 1-bromodecane (0.31 g, 1.39 mmol, 1.05 equiv) and K_2CO_3 (0.37 g, 2.64 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **22b**. The title compound was obtained as a white solid (0.58 g, 86%).



22d

Methyl 3-(dodecyloxy)-5-(pentadecyloxy)benzoate (22d). Compound **22d** was synthesized from compound **21** (0.50 g, 1.32 mmol, 1 equiv), 1-bromododecane (0.35 g, 1.39 mmol, 1.05 equiv) and K_2CO_3 (0.37 g, 2.64 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **22b**. The title compound was obtained as a white solid (0.60 g, 83%).



22e

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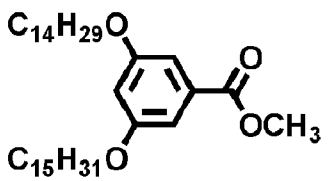
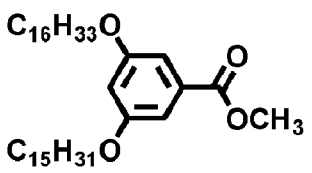
Methyl 3-(pentadecyloxy)-5-(tridecyloxy)benzoate (22e). Compound **22e** was synthesized from compound **21** (0.50 g, 1.32 mmol, 1 equiv), $C_{13}H_{27}OTs$ (**12a**, 0.51 g, 1.39 mmol, 1.05 equiv) and K_2CO_3 (0.37 g, 2.64 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **22b**. The title compound was obtained as a white solid (0.61 g, 82%).



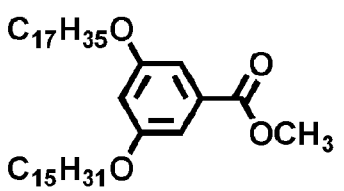
22f

Methyl 3-(pentadecyloxy)-5-(tetradecyloxy)benzoate (22f). Compound **22f** was synthesized from compound **21** (0.50 g, 1.32 mmol, 1 equiv), 1-bromotetradecane (0.39 g, 1.39 mmol, 1.05 equiv) and K_2CO_3 (0.37 g, 2.64 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **22b**. The title compound was obtained as a white solid (0.56 g, 74%).



22g

Methyl 3-(hexadecyloxy)-5-(pentadecyloxy)benzoate (22g). Compound **22g** was synthesized from compound **21** (0.50 g, 1.32 mmol, 1 equiv), 1-bromohexadecane (0.43 g, 1.39 mmol, 1.05 equiv) and K_2CO_3 (0.37 g, 2.64 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **22b**. The title compound was obtained as a white solid (0.60 g, 75%).



22h

Methyl 3-(heptadecyloxy)-5-(pentadecyloxy)benzoate (22h). Compound **22h** was synthesized from compound **21** (0.50 g, 1.32 mmol, 1 equiv), $C_{17}H_{35}OTs$ (**12b**, 0.43 g, 1.39 mmol, 1.05 equiv) and K_2CO_3 (0.37 g, 2.64 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **22b**. The title compound was obtained as a white solid (0.67 g, 83%).

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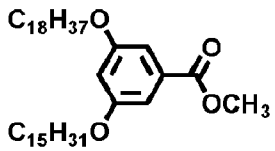
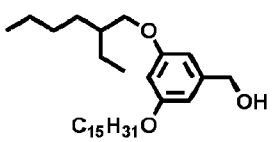


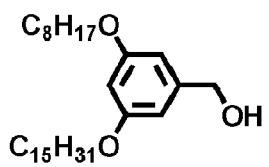
22i

Methyl 3-(octadecyloxy)-5-(pentadecyloxy)benzoate (22i). Compound **22i** was synthesized from compound **21** (0.50 g, 1.32 mmol, 1 equiv), 1-bromoctadecanene (0.46 g, 1.39 mmol, 1.05 equiv) and K_2CO_3 (0.37 g, 2.64 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **22b**. The title compound was obtained as a white solid (0.71 g, 85%).



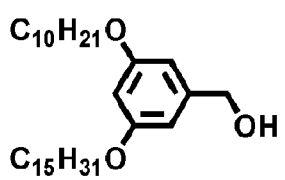
23a

(3-((2-Ethylhexyl)oxy)-5-(pentadecyloxy)phenyl)methanol (23a). Compound **22a** (0.32 g, 0.65 mmol, 1 equiv) was dissolved in 5 mL dry THF, which was added dropwise to a slurry of $LiAlH_4$ (25 mg, 0.65 mmol, 1 equiv) in dry THF (5 mL) at 0 °C under N_2 atmosphere. The resulted mixture was stirred at 23 °C for 1 h. The reaction was quenched by the successive addition of water (0.3 mL), 15% NaOH aqueous solution (0.3 mL) and water (1.5 mL). Then the white precipitates in the mixture were filtered out and the filtrate was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.29 g, 97%).



23b

(3-(Octyloxy)-5-(pentadecyloxy)phenyl)methanol (23b). Compound **23b** was synthesized from compound **22b** (0.45 g, 0.91 mmol, 1 equiv) and $LiAlH_4$ (35 mg, 0.91 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **23a**. The title compound was obtained as a colorless oil (0.42 g, 100%).



23c

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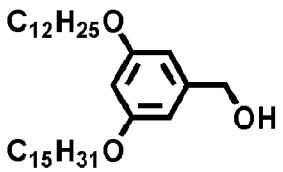
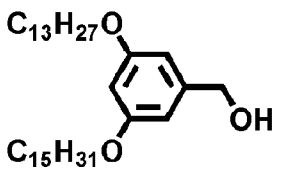
(3-(Decyloxy)-5-(pentadecyloxy)phenyl)methanol (23c). Compound **23c** was synthesized from compound **22c** (0.43 g, 0.83 mmol, 1 equiv) and $LiAlH_4$ (32 mg, 0.83 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **23a**. The title compound was obtained as a colorless oil (0.41 g, 100%).



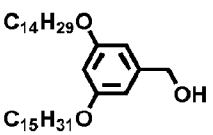
23d

(3-(Dodecyloxy)-5-(pentadecyloxy)phenyl)methanol (23d). Compound **23d** was synthesized from compound **22d** (0.28 g, 0.51 mmol, 1 equiv) and $LiAlH_4$ (20 mg, 0.51 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **23a**. The title compound was obtained as a colorless oil (0.26 g, 100%).



23e

(3-(Pentadecyloxy)-5-(tridecyloxy)phenyl)methanol (23e). Compound **23e** was synthesized from compound **22e** (0.28 g, 0.49 mmol, 1 equiv) and $LiAlH_4$ (19 mg, 0.49 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **23a**. The title compound was obtained as a colorless oil (0.25 g, 96%).



23f

(3-(Pentadecyloxy)-5-(tetradecyloxy)phenyl)methanol (23f). Compound **23f** was synthesized from compound **22f** (0.53 g, 0.91 mmol, 1 equiv) and $LiAlH_4$ (35 mg, 0.91 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **23a**. The title compound was obtained as a colorless oil (0.50 g, 100%).

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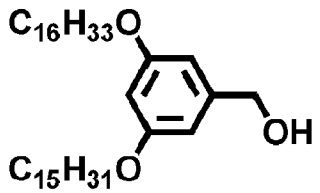
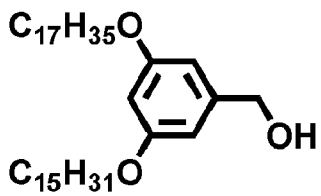


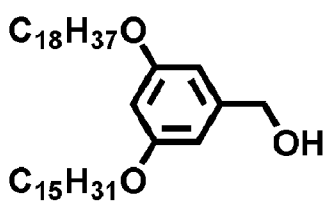
23g

(3-(Hexadecyloxy)-5-(pentadecyloxy)phenyl)methanol (23g). Compound **23g** was synthesized from compound **22g** (0.50 g, 0.85 mmol, 1 equiv) and $LiAlH_4$ (32 mg, 0.85 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **23a**. The title compound was obtained as a white solid (0.44 g, 93%).



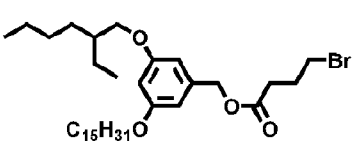
23h

(3-(Heptadecyloxy)-5-(pentadecyloxy)phenyl)methanol (23h). Compound **23h** was synthesized from compound **22h** (0.55 g, 0.89 mmol, 1 equiv) and $LiAlH_4$ (34 mg, 0.89 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **23a**. The title compound was obtained as a white solid (0.49 g, 94%).



23i

(3-(Octadecyloxy)-5-(pentadecyloxy)phenyl)methanol (23i). Compound **23i** was synthesized from compound **22i** (0.57 g, 0.90 mmol, 1 equiv) and $LiAlH_4$ (35 mg, 0.90 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **23a**. The title compound was obtained as a white solid (0.55 g, 100%).



24a

3-((2-Ethylhexyl)oxy)-5-(pentadecyloxy)benzyl 4-bromobutanoate (24a). Compound **23a** (0.25

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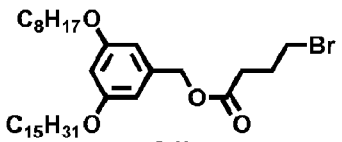
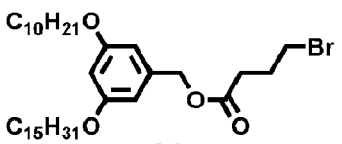
g, 0.54 mmol, 1 equiv), 4-bromobutyric acid (99 mg, 0.59 mmol, 1.1 equiv) and DPTS (0.17 g, 0.59 mmol, 1.1 equiv) were dissolved in 6 mL DCM. DCC (0.22 g, 1.08 mmol, 2 equiv) was added in one portion into the above mixture. The reaction mixture was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/DCM = 1/1 as the mobile phase to give the title compound as a colorless oil (0.31 g, 94%).



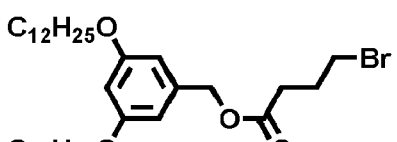
24b

3-(Octyloxy)-5-(pentadecyloxy)benzyl 4-bromobutanoate (24b). Compound **24b** was synthesized from compound **23b** (0.40 g, 0.91 mmol, 1 equiv), 4-bromobutyric acid (0.17 g, 1.00 mmol, 1.1 equiv), DPTS (0.29 g, 1.00 mmol, 1.1 equiv) and DCC (0.38 g, 1.82 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **24a**. The title compound was obtained as a colorless oil (0.53 g, 95%).



24c

3-(Decyloxy)-5-(pentadecyloxy)benzyl 4-bromobutanoate (24c). Compound **24c** was synthesized from compound **23c** (0.40 g, 0.81 mmol, 1 equiv), 4-bromobutyric acid (0.15 g, 0.89 mmol, 1.1 equiv), DPTS (0.26 g, 0.89 mmol, 1.1 equiv) and DCC (0.33 g, 1.62 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **24a**. The title compound was obtained as a colorless oil (0.43 g, 83%).



24d

3-(Dodecyloxy)-5-(pentadecyloxy)benzyl 4-bromobutanoate (24d). Compound **24d** was synthesized from compound **23d** (0.26 g, 0.51 mmol, 1 equiv), 4-bromobutyric acid (94 mg, 0.56 mmol, 1.1 equiv), DPTS (0.16 g, 0.56 mmol, 1.1 equiv) and DCC (0.21 g, 1.02 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **24a**. The title compound was obtained as a colorless oil (0.30 g, 88%).

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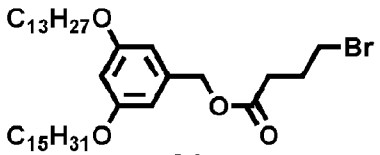
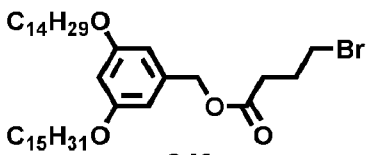


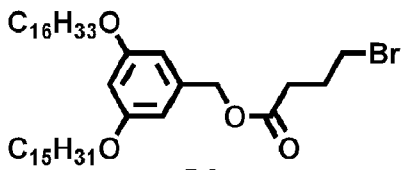
24e

3-(Pentadecyloxy)-5-(tridecyloxy)benzyl 4-bromobutanoate (24e). Compound **24e** was synthesized from compound **23e** (0.23 g, 0.43 mmol, 1 equiv), 4-bromobutyric acid (79 mg, 0.47 mmol, 1.1 equiv), DPTS (0.14 g, 0.47 mmol, 1.1 equiv) and DCC (0.18 g, 0.86 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **24a**. The title compound was obtained as a colorless oil (0.29 g, 98%).



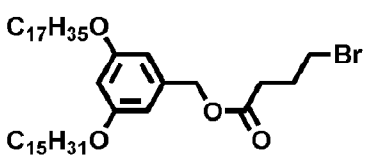
24f

3-(Pentadecyloxy)-5-(tetradecyloxy)benzyl 4-bromobutanoate (24f). Compound **24f** was synthesized from compound **23f** (0.50 g, 0.91 mmol, 1 equiv), 4-bromobutyric acid (0.17 g, 1.00 mmol, 1.1 equiv), DPTS (0.29 g, 1.00 mmol, 1.1 equiv) and DCC (0.38 g, 1.82 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **24a**. The title compound was obtained as a colorless oil (0.60 g, 95%).



24g

3-(Hexadecyloxy)-5-(pentadecyloxy)benzyl 4-bromobutanoate (24 g). Compound **24g** was synthesized from compound **23g** (0.42 g, 0.73 mmol, 1 equiv), 4-bromobutyric acid (0.13 g, 0.80 mmol, 1.1 equiv), DPTS (0.24 g, 0.80 mmol, 1.1 equiv) and DCC (0.30 g, 1.46 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **24a**. The title compound was obtained as a colorless oil (0.50 g, 96%).



24h

3-(Heptadecyloxy)-5-(pentadecyloxy)benzyl 4-bromobutanoate (24h). 4-Bromobutyric acid

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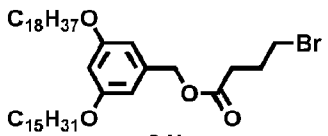
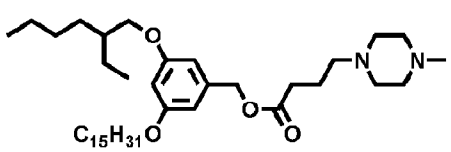
(0.15 g, 0.87 mmol, 1.1 equiv) was dissolved in 6 mL anhydrous DCM and 1 drop of DMF was added. $SOCl_2$ (0.21 g, 1.74 mmol, 2.2 equiv) was added and the mixture was stirred at 23 °C for 1 h. Afterwards, DCM and excess $SOCl_2$ was removed under reduced pressure to give the 4-bromobutanoyl chloride. Compound **23h** (0.45 g, 0.76 mmol, 1.0 equiv) and NEt_3 (97 mg, 0.96 mmol, 1.3 equiv) were dissolved in 3 mL anhydrous DCM. To this mixture was added the DCM solution (3 mL) of above 4-bromobutanoyl chloride dropwise at 0 °C. The mixture was allowed to warm to 23 °C with stirring during a 2 h period. Afterwards, water (30 mL) was added, and the mixture was extracted by DCM (30 mL) for three times. The organic phase was combined, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 30/1 as the mobile phase to give the title compound as a colorless oil (0.43 g, 77%).



24i

3-(Octadecyloxy)-5-(pentadecyloxy)benzyl 4-bromobutanoate (24i). Compound **24i** was synthesized from compound **23i** (0.56 g, 0.93 mmol, 1.0 equiv), 4-bromobutyric acid (0.17 g, 1.02 mmol, 1.1 equiv) and NEt_3 (0.12 g, 1.21 mmol, 1.3 equiv) following a procedure similar to that used for the synthesis of compound **24h**. The title compound was obtained as a light-yellow oil (0.61 g, 88%).



25a (IAJD135)

3-((2-Ethylhexyl)oxy)-5-(pentadecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (25a, IAJD135). Compound **24a** (0.15 g, 0.25 mmol, 1.0 equiv), 1-methylpiperazine (28 mg, 0.28 mmol, 1.1 equiv), K_2CO_3 (39 mg, 0.28 mmol, 1.1 equiv) were stirred in MeCN (15 mL). The mixture was heated at 95 °C for 3 h. The reaction mixture was cooled to 23 °C and the solvent was removed under reduced pressure. Then water (20 mL) was added, and the resulted mixture was extracted by DCM (20 mL) for three times. The organic phase was combined, dried over anhydrous $MgSO_4$, filtered and dried to give the crude product. The crude product was purified by column chromatography (SiO_2) with DCM/MeOH = 30/1 and 15/1 as the mobile phase. Then the obtained product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The

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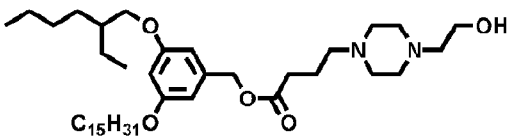
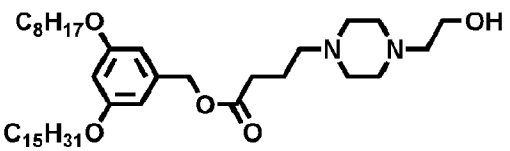
aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was collected and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.13 g, 82%).



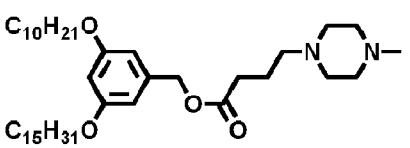
25b (IAJD136)

3-((2-Ethylhexyl)oxy)-5-(pentadecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (25b, IAJD136). Compound **25b** was synthesized from compound **24a** (0.16 g, 0.26 mmol), 1-(2-hydroxyethyl)piperazine (37 mg, 0.28 mmol) and K_2CO_3 (53 mg, 0.39 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a colorless oil (0.13 g, 76%).



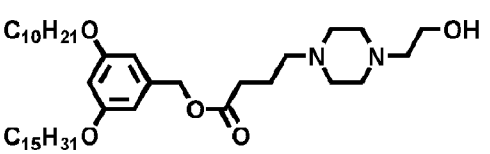
25c (IAJD138)

3-(Octyloxy)-5-(pentadecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (25c, IAJD138). Compound **25c** was synthesized from compound **24b** (0.24 g, 0.41 mmol), 1-(2-hydroxyethyl)piperazine (64 mg, 0.49 mmol) and K_2CO_3 (68 mg, 0.49 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a light-yellow oil (0.25 g, 93%).



25d (IAJD141)

3-(Decyloxy)-5-(pentadecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (25d, IAJD141). Compound **25d** was synthesized from compound **24c** (0.19 g, 0.30 mmol), 1-methylpiperazine (32 mg, 0.32 mmol) and K_2CO_3 (44 mg, 0.32 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a colorless oil (0.17 g, 90%).



25e (IAJD142)

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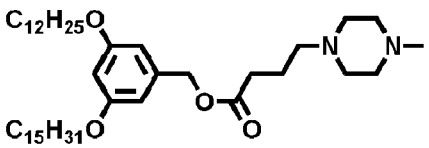
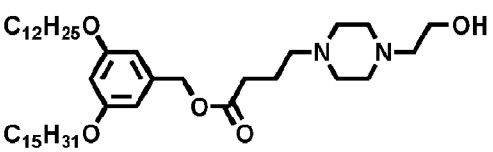
3-(Decyloxy)-5-(pentadecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (25e, IAJD142). Compound **25e** was synthesized from compound **24c** (0.19 g, 0.30 mmol), 1-(2-hydroxyethyl)piperazine (42 mg, 0.32 mmol) and K_2CO_3 (44 mg, 0.32 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a light-yellow oil (0.18 g, 90%).



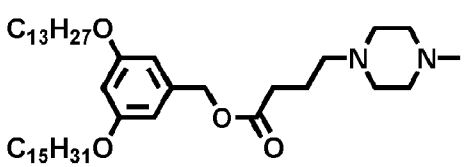
25f (IAJD143)

3-(Dodecyloxy)-5-(pentadecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (25f, IAJD143). Compound **25f** was synthesized from compound **24d** (0.15 g, 0.22 mmol), 1-methylpiperazine (23 mg, 0.23 mmol) and K_2CO_3 (32 mg, 0.23 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a colorless oil (0.13 g, 87%).



25g (IAJD144)

3-(Dodecyloxy)-5-(pentadecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (25g, IAJD144). Compound **25g** was synthesized from compound **24d** (0.15 g, 0.22 mmol), 1-(2-hydroxyethyl)piperazine (30 mg, 0.23 mmol) and K_2CO_3 (32 mg, 0.23 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a colorless oil (0.14 g, 88%).



25h (IAJD145)

3-(pentadecyloxy)-5-(tridecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (25h, IAJD145). Compound **25h** was synthesized from compound **24e** (0.15 g, 0.22 mmol), 1-methylpiperazine (23 mg, 0.23 mmol) and K_2CO_3 (32 mg, 0.23 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a colorless oil (0.13 g, 87%).

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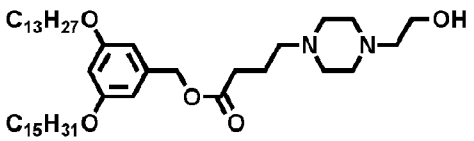
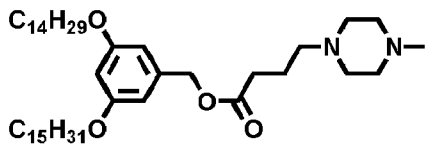


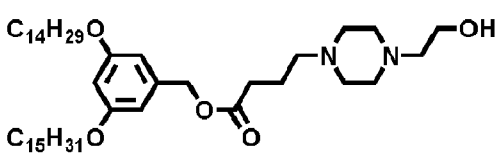
25i (IAJD146)

3-(Pentadecyloxy)-5-(tridecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (25i, IAJD146). Compound **25i** was synthesized from compound **24e** (0.15 g, 0.22 mmol), 1-(2-hydroxyethyl)piperazine (30 mg, 0.23 mmol) and K_2CO_3 (32 mg, 0.23 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a light-yellow oil (0.14 g, 89%).



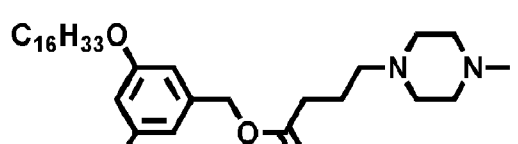
25j (IAJD147)

3-(Pentadecyloxy)-5-(tetradecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (25j, IAJD147). Compound **25j** was synthesized from compound **24f** (0.28 g, 0.40 mmol), 1-methylpiperazine (44 mg, 0.44 mmol) and K_2CO_3 (67 mg, 0.48 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a light-yellow oil (0.24 g, 88%).



25k (IAJD148)

3-(Pentadecyloxy)-5-(tetradecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (25k, IAJD148). Compound **25k** was synthesized from compound **24f** (0.28 g, 0.40 mmol), 1-(2-hydroxyethyl)piperazine (57 mg, 0.44 mmol) and K_2CO_3 (67 mg, 0.48 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a light-yellow oil (0.25 g, 92%).



25l (IAJD149)

3-(Hexadecyloxy)-5-(pentadecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (25l,

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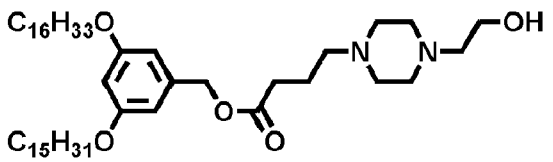
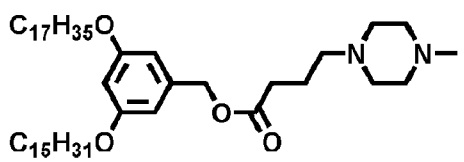
IAJD149). Compound **25l** was synthesized from compound **24g** (0.25 g, 0.35 mmol), 1-methylpiperazine (38 mg, 0.38 mmol) and K_2CO_3 (72 mg, 0.52 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a colorless oil (0.21 g, 81%).



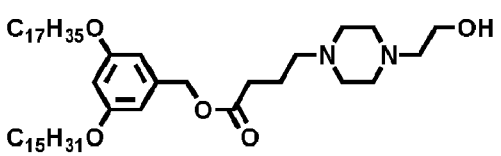
25m (IAJD150)

3-(Hexadecyloxy)-5-(pentadecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (25m, IAJD150). Compound **25m** was synthesized from compound **24g** (0.25 g, 0.35 mmol), 1-(2-hydroxyethyl)piperazine (50 mg, 0.38 mmol) and K_2CO_3 (72 mg, 0.52 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a colorless oil (0.22 g, 81%).



25n (IAJD151)

3-(Heptadecyloxy)-5-(pentadecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (25n, IAJD151). Compound **25n** was synthesized from compound **24h** (0.16 g, 0.22 mmol), 1-methylpiperazine (23 mg, 0.23 mmol) and K_2CO_3 (32 mg, 0.23 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a light-yellow oil (0.15 g, 88%).



25o (IAJD152)

3-(Heptadecyloxy)-5-(pentadecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (25o, IAJD152). Compound **25o** was synthesized from compound **24h** (0.15 g, 0.20 mmol), 1-(2-hydroxyethyl)piperazine (27 mg, 0.21 mmol) and K_2CO_3 (29 mg, 0.21 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a colorless oil (0.14 g, 88%).

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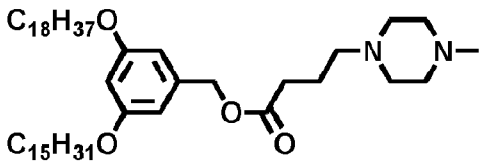
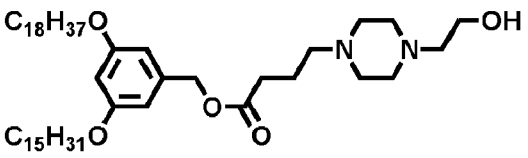


25p (IAJD153)

3-(Octadecyloxy)-5-(pentadecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (25p, IAJD153). Compound **25p** was synthesized from compound **24i** (0.31 g, 0.41 mmol), 1-methylpiperazine (45 mg, 0.45 mmol) and K_2CO_3 (85 mg, 0.62 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a colorless oil (0.21 g, 70%).



25q (IAJD154)

3-(Octadecyloxy)-5-(pentadecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (25q, IAJD154). Compound **25q** was synthesized from compound **24i** (0.31 g, 0.41 mmol), 1-(2-hydroxyethyl)piperazine (60 mg, 0.44 mmol) and K_2CO_3 (85 mg, 0.62 mmol) following a procedure similar to that used for the synthesis of compound **25a**. The title compound was obtained as a colorless oil (0.25 g, 76%).

Synthesis of tridecane-based IAJDs with nonsymmetric alkyl chains was performed according to Scheme 12 shown below:

Scheme 12. Reagents and conditions: (*i*) RBr/ROTs, K_2CO_3, DMF, 120 °C, 2 h; (*ii*) H_2, Pd/C, DCM, MeOH, 12 h; (*iii*) $LiAlH_4$, THF, 0–23 °C, 1 h; (*iv*) DCC, DPTS, 12 h; (*v*) K_2CO_3, MeCN, 95 °C, 3 h.

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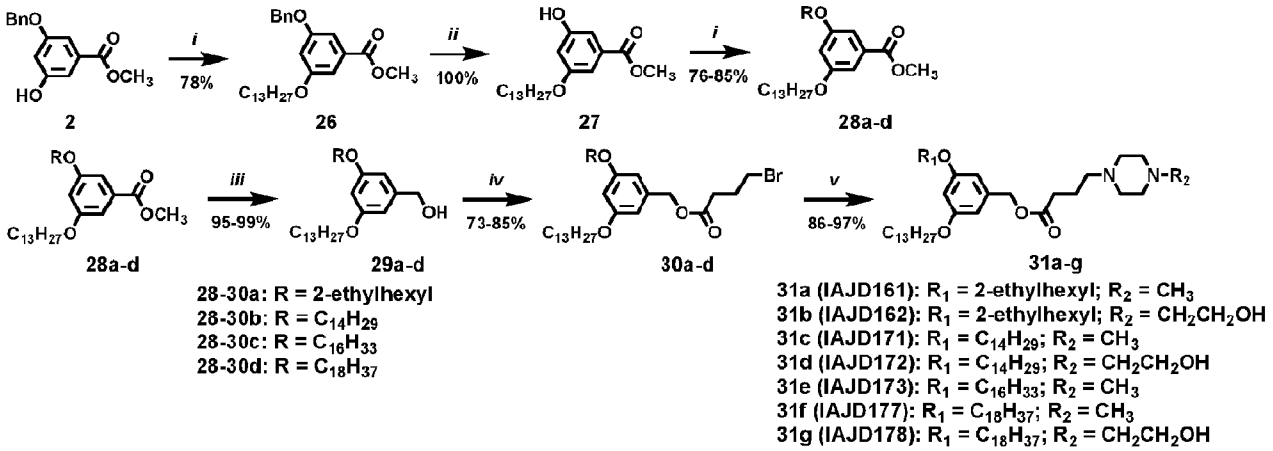
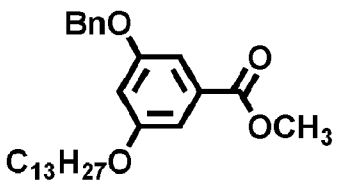
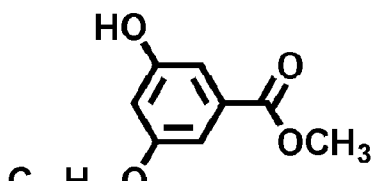




26

Methyl 3-(benzyloxy)-5-(tridecyloxy)benzoate (26). Compound **2** (5.50 g, 21.30 mmol, 1 equiv), $C_{13}H_{27}OTs$ (**12a**, 8.30 g, 23.43 mmol, 1.1 equiv) and K_2CO_3 (5.89 g, 42.60 mmol, 2 equiv) were stirred in DMF (40 mL). The mixture was stirred at 120 °C under N_2 atmosphere for 2 h. After cooled to 23 °C, the reaction mixture was poured into ice/water (150 mL) and the resulted white precipitates were filtered and collected. Then the precipitates were purified by recrystallization from minimum acetone to give the title compound as a white solid (7.30 g, 78%).



27

Methyl 3-hydroxy-5-(tridecyloxy)benzoate (27). Compound **26** (6.86 g, 15.57 mmol) was dissolved in a mixture of DCM (50 mL) and methanol (25 mL). Then Pd/C (0.41 g, 6 wt%) was added and the flask was evacuated and filled with hydrogen for three times. The mixture was stirred at 23 °C for 12 h under hydrogen atmosphere. The reaction mixture was filtered through Celite, and the filter cake was washed with DCM carefully. Evaporation of the solvent yielded the title compound as a white solid (5.46 g, 100%).

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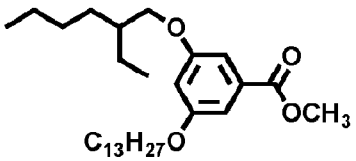
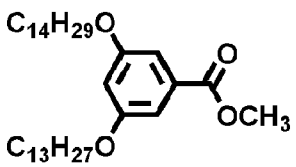


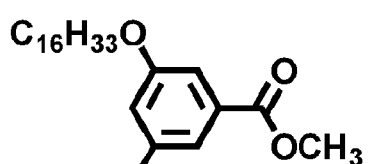
28a

Methyl 3-((2-ethylhexyl)oxy)-5-(tridecyloxy)benzoate (28a). Compound **27** (0.60 g, 1.71 mmol, 1 equiv), 3-(bromomethyl)heptane (0.36 g, 1.88 mmol, 1.1 equiv) and K_2CO_3 (0.47 g, 3.42 mmol, 2 equiv) were stirred in DMF (15 mL). The mixture was stirred at 80 °C under N_2 atmosphere for 12 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (20 mL) was added, and the mixture was extracted with DCM (20 mL) for three times. The organic phases were combined, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the mobile phase to give the title compound as a light-yellow oil (0.60 g, 76%).



28b

Methyl 3-(tetradecyloxy)-5-(tridecyloxy)benzoate (28b). Compound **27** (0.60 g, 1.71 mmol, 1 equiv), 1-bromotetradecane (0.52 g, 1.88 mmol, 1.1 equiv) and K_2CO_3 (0.47 g, 3.42 mmol, 2 equiv) were stirred in DMF (15 mL). The mixture was stirred at 120 °C under N_2 atmosphere for 2 h. After cooled to 23 °C, the reaction mixture was poured into ice/water (100 mL) and the resulted white precipitates were filtered and collected. Then the precipitates were purified by recrystallization from minimum acetone to give the title compound as a white solid (0.76 g, 81%).



28c

Methyl 3-(hexadecyloxy)-5-(tridecyloxy)benzoate (28c). Compound **28c** was synthesized from compound **27** (0.60 g, 1.71 mmol, 1 equiv), 1-bromohexadecane (0.57 g, 1.88 mmol, 1.1 equiv) and K_2CO_3 (0.47 g, 3.42 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **28b**. The title compound was obtained as a white solid (0.84 g, 85%).

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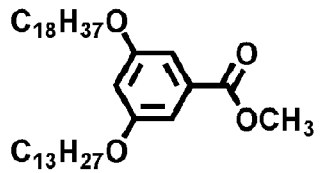
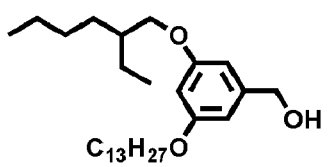


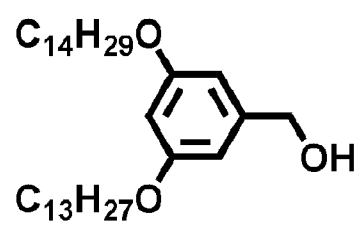
28d

Methyl 3-(octadecyloxy)-5-(tridecyloxy)benzoate (28d). Compound **28d** was synthesized from compound **27** (0.42 g, 1.20 mmol, 1 equiv), 1-bromoctadecanene (0.44 g, 1.32 mmol, 1.1 equiv) and K_2CO_3 (0.33 g, 2.40 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **28b**. The title compound was obtained as a white solid (0.56 g, 78%).



29a

(3-((2-Ethylhexyl)oxy)-5-(tridecyloxy)phenyl)methanol (29a). Compound **28a** (0.60 g, 1.30 mmol, 1 equiv) was dissolved in 5 mL dry THF, which was added dropwise to a slurry of $LiAlH_4$ (49 mg, 1.30 mmol, 1 equiv) in dry THF (5 mL) at 0 °C under N_2 atmosphere. The resulted mixture was stirred at 23 °C for 1 h. The reaction was quenched by the successive addition of water (0.3 mL), 15% NaOH aqueous solution (0.3 mL) and water (1.5 mL). Then the white precipitates in the mixture were filtered out and the filtrate was dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.54 g, 95%).



29b

(3-(Tetradecyloxy)-5-(tridecyloxy)phenyl)methanol (29b). Compound **29b** was synthesized from compound **28b** (0.75 g, 1.37 mmol, 1 equiv) and $LiAlH_4$ (52 mg, 1.37 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **29a**. The title compound was obtained as a colorless oil (0.68 g, 96%).

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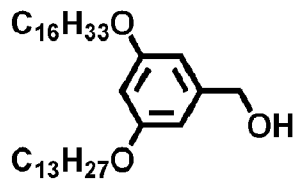
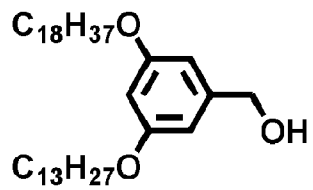


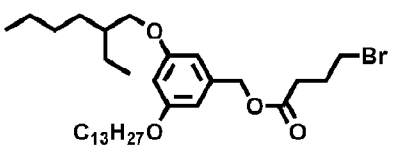
29c

(3-(Hexadecyloxy)-5-(tridecyloxy)phenyl)methanol (29c). Compound **29c** was synthesized from compound **28c** (0.86 g, 1.50 mmol, 1 equiv) and $LiAlH_4$ (58 mg, 1.50 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **29a**. The title compound was obtained as a white solid (0.80 g, 98%).



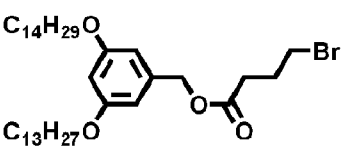
29d

(3-(Octadecyloxy)-5-(tridecyloxy)phenyl)methanol (29d). Compound **29d** was synthesized from compound **28d** (0.53 g, 0.88 mmol, 1 equiv) and $LiAlH_4$ (34 mg, 0.88 mmol, 1 equiv) following a procedure similar to that used for the synthesis of compound **29a**. The title compound was obtained as a white solid (0.50 g, 99%).



30a

3-((2-Ethylhexyl)oxy)-5-(tridecyloxy)benzyl 4-bromobutanoate (30a). Compound **29a** (0.46 g, 1.06 mmol, 1 equiv), 4-bromobutyric acid (0.20 g, 1.17 mmol, 1.1 equiv) and DPTS (0.34 g, 1.17 mmol, 1.1 equiv) were dissolved in 6 mL DCM. DCC (0.44 g, 2.12 mmol, 2 equiv) was added in one portion into the above mixture. The reaction mixture was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/DCM = 1/1 as the mobile phase to give the title compound as a colorless oil (0.46 g, 74%).



30b

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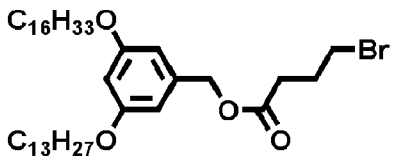
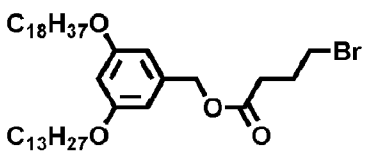
3-(Tetradecyloxy)-5-(tridecyloxy)benzyl 4-bromobutanoate (30b). Compound **30b** was synthesized from compound **29b** (0.30 g, 0.58 mmol, 1 equiv), 4-bromobutyric acid (0.11 g, 0.64 mmol, 1.1 equiv), DPTS (0.19 g, 0.64 mmol, 1.1 equiv) and DCC (0.24 g, 1.16 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **30a**. The title compound was obtained as a colorless oil (0.31 g, 79%).



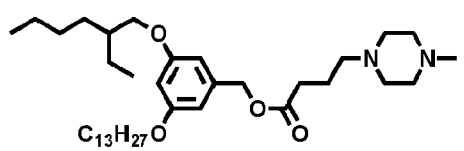
30c

3-(Hexadecyloxy)-5-(tridecyloxy)benzyl 4-bromobutanoate (30c). Compound **30c** was synthesized from compound **29c** (0.30 g, 0.55 mmol, 1 equiv), 4-bromobutyric acid (0.10 g, 0.61 mmol, 1.1 equiv), DPTS (0.18 g, 0.61 mmol, 1.1 equiv) and DCC (0.23 g, 1.10 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **30a**. The title compound was obtained as a colorless oil (0.28 g, 73%).



30d

3-(Octadecyloxy)-5-(tridecyloxy)benzyl 4-bromobutanoate (30d). Compound **30d** was synthesized from compound **29d** (0.44 g, 0.77 mmol, 1 equiv), 4-bromobutyric acid (0.14 g, 0.85 mmol, 1.1 equiv), DPTS (0.25 g, 0.85 mmol, 1.1 equiv) and DCC (0.32 g, 1.54 mmol, 2 equiv) following a procedure similar to that used for the synthesis of compound **30a**. The title compound was obtained as a colorless oil (0.47 g, 85%).



31a (IAJD161)

3-((2-Ethylhexyl)oxy)-5-(tridecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (31a, IAJD161). Compound **30a** (0.17 g, 0.29 mmol, 1.0 equiv), 1-methylpiperazine (30 mg, 0.30 mmol, 1.05 equiv), K_2CO_3 (42 mg, 0.30 mmol, 1.05 equiv) were stirred in MeCN (15 mL). The mixture was heated at 95 °C for 3 h. The reaction mixture was cooled to 23 °C and the solvent was removed under reduced pressure. Then water (20 mL) was added, and the resulted mixture was extracted by

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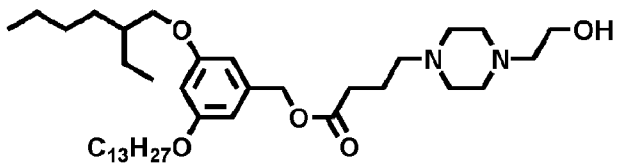
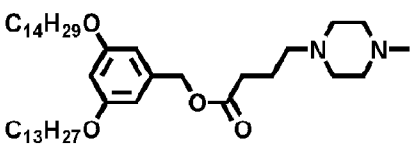
DCM (20 mL) for three times. The organic phase was combined, dried over anhydrous $MgSO_4$, filtered and dried to give the crude product. The crude product was purified by column chromatography (SiO_2) with DCM/MeOH = 30/1 and 15/1 as the mobile phase. Then the obtained product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was collected and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.15 g, 88%).



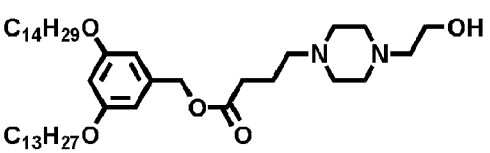
31b (IAJD162)

3-((2-Ethylhexyl)oxy)-5-(tridecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (31b, IAJD162). Compound **31b** was synthesized from compound **30a** (0.17 g, 0.29 mmol), 1-(2-hydroxyethyl)piperazine (39 mg, 0.30 mmol) and K_2CO_3 (42 mg, 0.30 mmol) following a procedure similar to that used for the synthesis of compound **31a**. The title compound was obtained as a colorless oil (0.16 g, 89%).



31c (IAJD171)

3-(Tetradecyloxy)-5-(tridecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (31c, IAJD171). Compound **31c** was synthesized from compound **30b** (0.14 g, 0.21 mmol), 1-methylpiperazine (22 mg, 0.22 mmol) and K_2CO_3 (31 mg, 0.22 mmol) following a procedure similar to that used for the synthesis of compound **31a**. The title compound was obtained as a colorless oil (0.12 g, 86%).



31d (IAJD172)

3-(Tetradecyloxy)-5-(tridecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (31d, IAJD172). Compound **31d** was synthesized from compound **30b** (0.14 g, 0.21 mmol), 1-(2-hydroxyethyl)piperazine (29 mg, 0.22 mmol) and K_2CO_3 (31 mg, 0.22 mmol) following a procedure similar to that used for the synthesis of compound **31a**. The title compound was obtained

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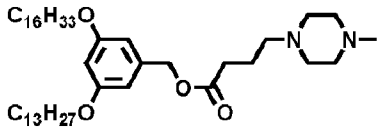
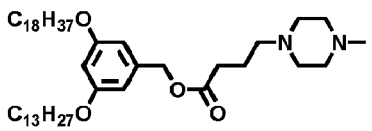
as a colorless oil (0.13 g, 87%).



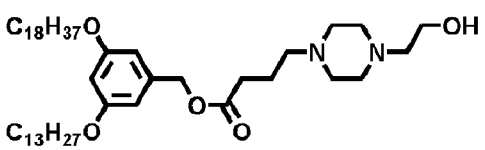
31e (IAJD173)

3-(Hexadecyloxy)-5-(tridecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (31e, IAJD173). Compound **31e** was synthesized from compound **30c** (0.14 g, 0.20 mmol), 1-methylpiperazine (22 mg, 0.22 mmol) and K_2CO_3 (31 mg, 0.22 mmol) following a procedure similar to that used for the synthesis of compound **31a**. The title compound was obtained as a light-yellow oil (0.12 g, 86%).



31f (IAJD177)

3-(Octadecyloxy)-5-(tridecyloxy)benzyl 4-(4-methylpiperazin-1-yl)butanoate (31f, IAJD177). Compound **31f** was synthesized from compound **30d** (0.20 g, 0.27 mmol), 1-methylpiperazine (30 mg, 0.30 mmol) and K_2CO_3 (41 mg, 0.30 mmol) following a procedure similar to that used for the synthesis of compound **31a**. The title compound was obtained as a light-yellow oil (0.20 g, 97%).



31g (IAJD178)

3-(Octadecyloxy)-5-(tridecyloxy)benzyl 4-(4-(2-hydroxyethyl)piperazin-1-yl)butanoate (31g, IAJD178). Compound **31g** was synthesized from compound **30d** (0.26 g, 0.36 mmol), 1-(2-hydroxyethyl)piperazine (48 mg, 0.37 mmol) and K_2CO_3 (51 mg, 0.37 mmol) following a procedure similar to that used for the synthesis of compound **31a**. The title compound was obtained as a light-yellow oil (0.27 g, 96%).

Synthesis of Triethylene Glycol-Based IAJDs with Nonsymmetric Alkyl Chains was performed according to Scheme 13 shown below:

Scheme 13. Reagents and conditions: (*i*) EDC.HCl, DMAP, DCM, 23 °C, 12 h; (*ii*) DDQ, DCM/H_2O, 23 °C, 1 h; (*iii*) 4-(piperidin-1-yl)butanoic acid hydrochloride, DCC, DCM, 23 °C, 12 h; (*iv*) $SOCl_2$, DMF (cat.), DCM, 23 °C, 0.5 h, then potassium phthalimide, DMF, 80 °C, 1 h; (*v*)

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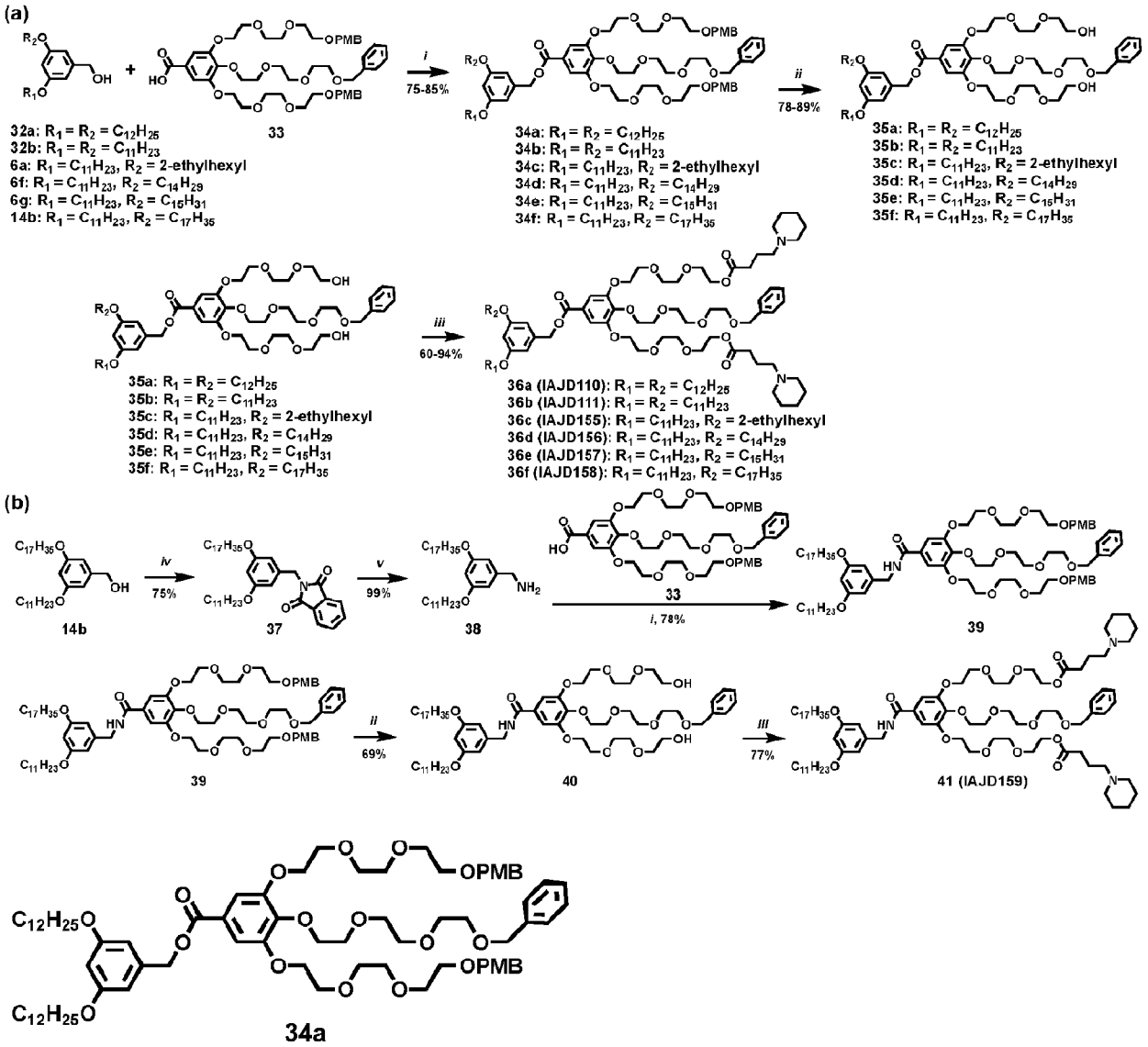
Hydrazine monohydrate, EtOH, reflux, 2 h.



Compound 34a. Compound **32a** was synthesized according to a previously reported procedure. Compound **33** was synthesized according to a procedure that was elaborated and optimized. Compound **32a** (0.41 g, 0.86 mmol, 1.1 equiv), compound **33** (0.70 g, 0.78 mmol, 1.0 equiv), EDC.HCl (0.16 g, 0.86 mmol, 1.1 equiv) and DMAP (28 mg, 0.23 mmol, 0.3 equiv) were dissolved in dry DCM (8 mL). The reaction mixture was stirred at 23 °C for 12 h. Brine (30 mL) was added, and the mixture was extracted by DCM (30 mL) for three times. The organic phase was collected, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/hexane = 1/1 as the mobile phase to give the title compound as a colorless oil (0.83 g, 78%).

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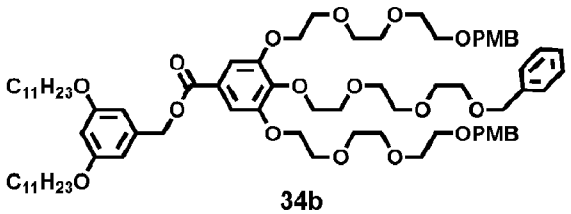
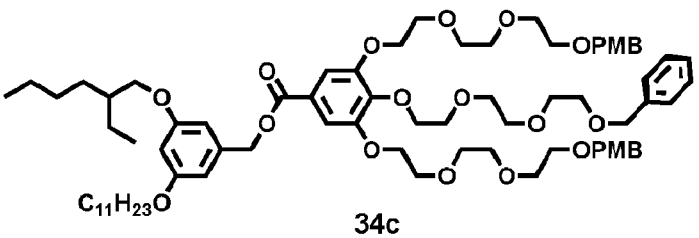


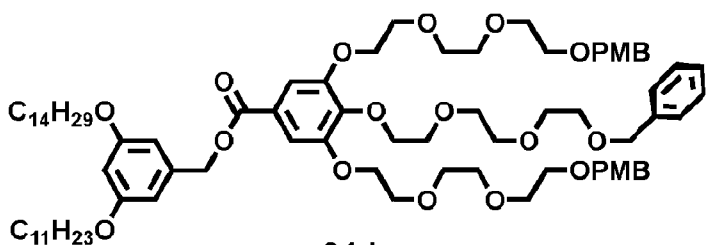
34b

Compound 34b. Compound **32b** was synthesized according to a previously reported procedure. Compound **34b** was synthesized from compound **32b** (0.39 g, 0.86 mmol) and compound **33** (0.70 g, 0.78 mmol) following a procedure similar to that used for the synthesis of compound **34a**. The title compound was obtained as a colorless oil (0.78 g, 75%).



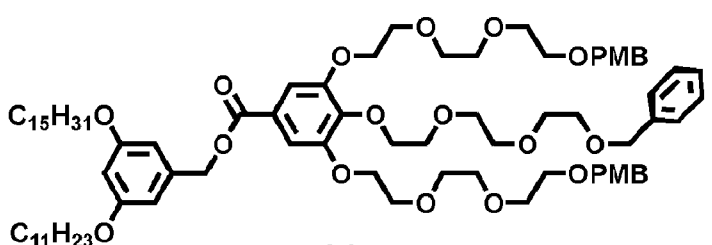
34c

Compound 34c. Compound **34c** was synthesized from compound **6a** (0.10 g, 0.25 mmol) and compound **33** (0.22 g, 0.25 mmol) following a procedure similar to that used for the synthesis of compound **34a**. The title compound was obtained as a colorless oil (0.26 g, 81%).



34d

Compound 34d. Compound **34d** was synthesized from compound **6f** (0.12 g, 0.24 mmol) and compound **33** (0.22 g, 0.24 mmol) following a procedure similar to that used for the synthesis of compound **34a**. The title compound was obtained as a colorless oil (0.28 g, 85%).



34e

Compound 34e. Compound **34e** was synthesized from compound **6g** (0.12 g, 0.24 mmol) and

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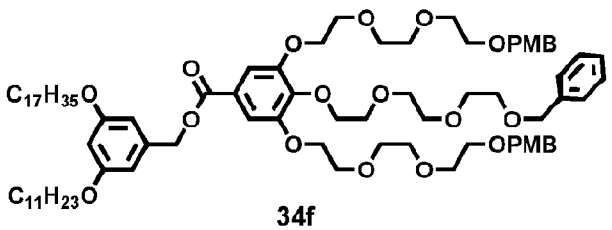
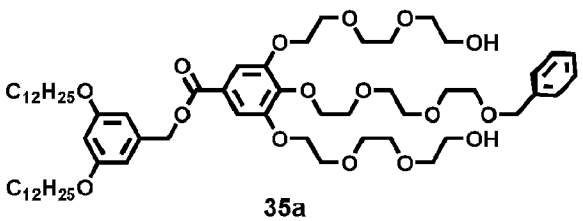
compound **33** (0.22 g, 0.24 mmol) following a procedure similar to that used for the synthesis of compound **34a**. The title compound was obtained as a colorless oil (0.27 g, 82%).



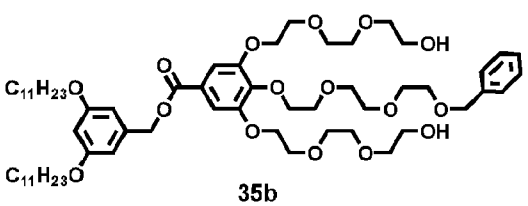
34f

Compound 34f. Compound **34f** was synthesized from compound **14b** (0.17 g, 0.32 mmol) and compound **33** (0.29 g, 0.32 mmol) following a procedure similar to that used for the synthesis of compound **34a**. The title compound was obtained as a colorless oil (0.35 g, 78%).



35a

Compound 35a. Compound **34a** (0.68 g, 0.50 mmol, 1 equiv) was dissolved in 6 mL DCM and 0.42 mL water (7 v%) was added. To this solution was added 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ, 0.25 g, 1.10 mmol, 2.2 equiv). The reaction mixture was stirred at 23 °C for 1 h. Then the precipitates were filtered out and DCM (20 mL) was added. The mixture was washed by $NaHCO_3$ aqueous solution (saturated), $NaHSO_3$ aqueous solution (2%) and $NaHCO_3$ aqueous solution (saturated) successively. The organic phase was collected, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 20/1 as the mobile phase to give the title compound as a light-yellow oil (0.45 g, 80%).



35b

Compound 35b. Compound **35b** was synthesized from compound **34b** (0.66 g, 0.50 mmol) and DDQ (0.25 g, 1.10 mmol) following a procedure similar to that used for the synthesis of compound **35a**. The title compound was obtained as a light-yellow oil (0.42 g, 78%).

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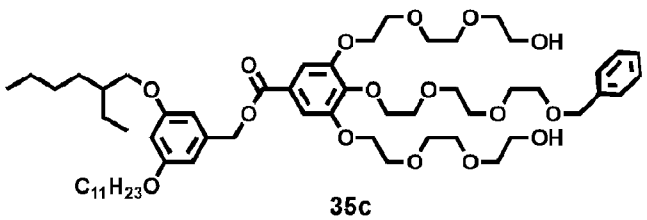
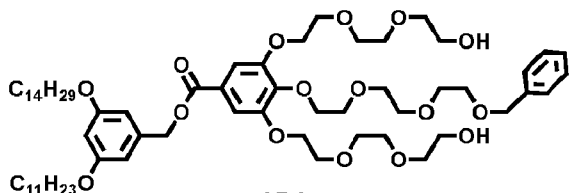


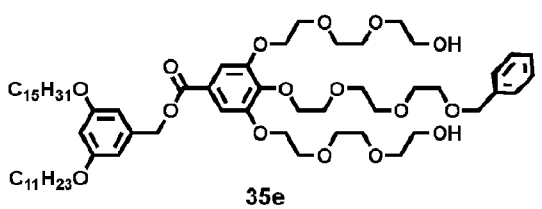
35c

Compound 35c. Compound **35c** was synthesized from compound **34c** (0.25 g, 0.19 mmol) and DDQ (95 mg, 0.42 mmol) following a procedure similar to that used for the synthesis of compound **35a**. The title compound was obtained as a light-yellow oil (0.17 g, 85%).



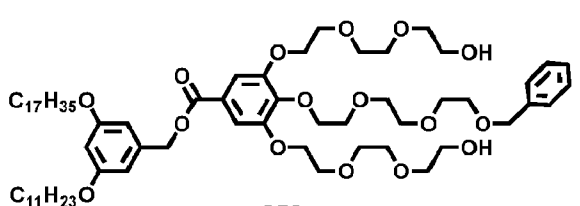
35d

Compound 35d. Compound **35d** was synthesized from compound **34d** (0.26 g, 0.19 mmol) and DDQ (95 mg, 0.42 mmol) following a procedure similar to that used for the synthesis of compound **35a**. The title compound was obtained as a light-yellow oil (0.17 g, 81%).



35e

Compound 35e. Compound **35e** was synthesized from compound **34e** (0.25 g, 0.18 mmol) and DDQ (0.10 g, 0.44 mmol) following a procedure similar to that used for the synthesis of compound **35a**. The title compound was obtained as a colorless oil (0.18 g, 89%).



35f

Compound 35f. Compound **35f** was synthesized from compound **34f** (0.34 g, 0.24 mmol) and DDQ (0.14 g, 0.61 mmol) following a procedure similar to that used for the synthesis of compound **35a**. The title compound was obtained as a colorless oil (0.23 g, 81%).

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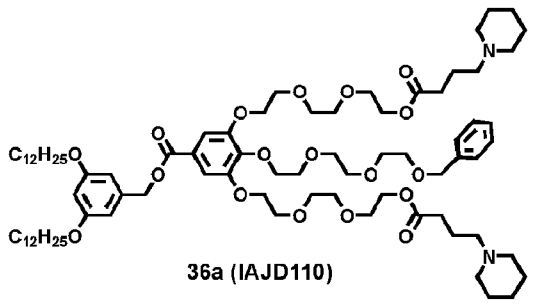
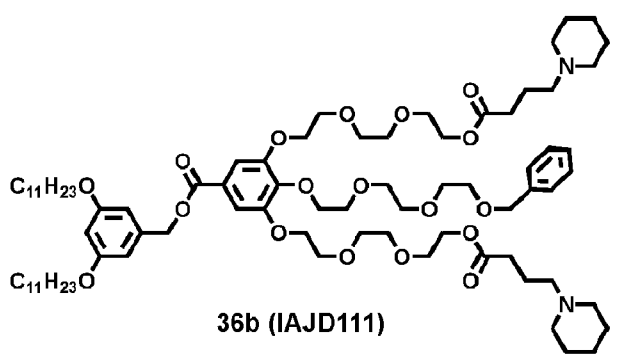


36a (IAJD110)

Compound 36a (IAJD110). Compound **35a** (0.20 g, 0.18 mmol, 1 equiv) and 4-(piperidin-1-yl)butanoic acid hydrochloride (83 mg, 0.40 mmol, 2.2 equiv) were dissolved in 5 mL dry DCM. DCC (0.11 g, 0.54 mmol, 3 equiv) was added in one portion into the above mixture. The reaction mixture was stirred at 23 °C for 12 h. The mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 30/1, 10/1, 8/1 and 8/1 with 0.1% NEt_3 as the mobile phase. Then the product was dissolved in DCM (20 mL), which was washed by $NaHCO_3$ aqueous solution (2%, 20 mL). The aqueous phase was extracted by DCM (20 mL) for another two times. The organic phase was combined and dried over anhydrous $MgSO_4$. Filtration and evaporation of the solvent yielded the title compound as a light-yellow oil (0.23 g, 89%).



36b (IAJD111)

Compound 36b (IAJD111). Compound **36b** was synthesized from compound **35b** (0.20 g, 0.18 mmol), 4-(piperidin-1-yl)butanoic acid hydrochloride (83 mg, 0.40 mmol) and DCC (0.11 g, 0.54 mmol) following a procedure similar to that used for the synthesis of compound **36a**. The title compound was obtained as a light-yellow oil (0.23 g, 92%).

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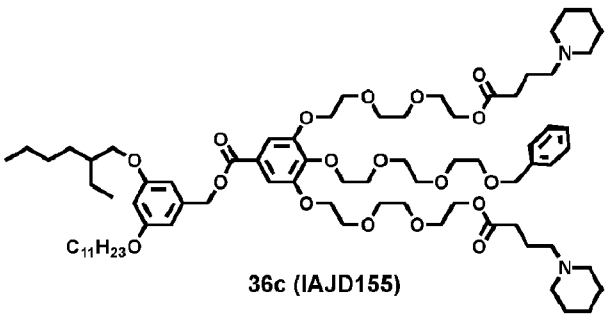
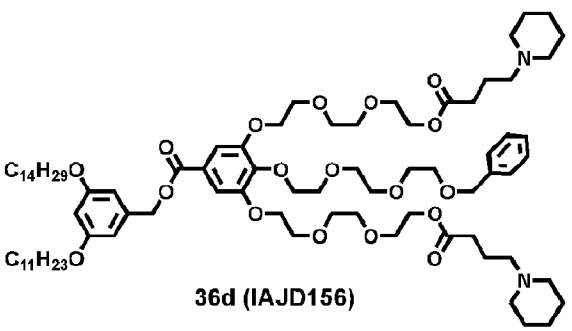


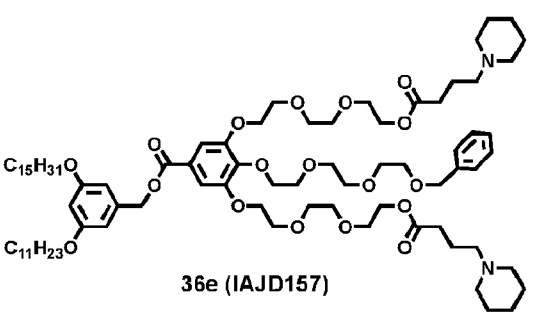
36c (IAJD155)

Compound 36c (IAJD155). Compound **36c** was synthesized from compound **35c** (0.12 g, 0.11 mmol), 4-(piperidin-1-yl)butanoic acid hydrochloride (50 mg, 0.24 mmol) and DCC (68 mg, 0.33 mmol) following a procedure similar to that used for the synthesis of compound **36a**. The title compound was obtained as a light-yellow oil (0.13 g, 87%).



36d (IAJD156)

Compound 36d (IAJD156). Compound **36d** was synthesized from compound **35d** (0.14 g, 0.12 mmol), 4-(piperidin-1-yl)butanoic acid hydrochloride (54 mg, 0.26 mmol) and DCC (74 mg, 0.36 mmol) following a procedure similar to that used for the synthesis of compound **36a**. The title compound was obtained as a light-yellow oil (0.14 g, 82%).



36e (IAJD157)

Compound 36e (IAJD157). Compound **36e** was synthesized from compound **35e** (0.18 g, 0.16 mmol), 4-(piperidin-1-yl)butanoic acid hydrochloride (84 mg, 0.40 mmol) and DCC (0.17 g, 0.80 mmol) following a procedure similar to that used for the synthesis of compound **36a**. The title compound was obtained as a colorless oil (0.22 g, 94%).

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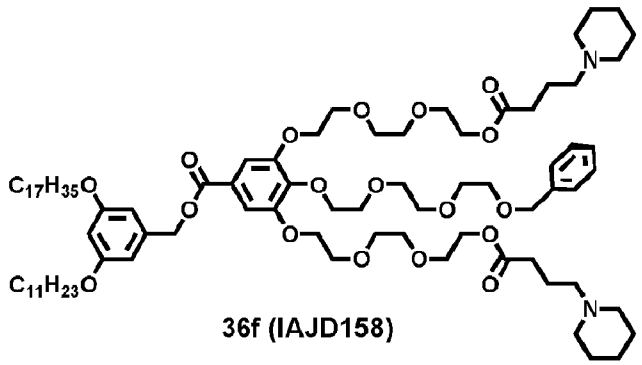
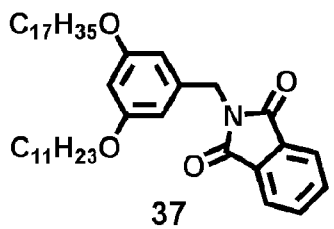


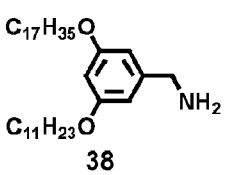
36f (IAJD158)

Compound 36f (IAJD158). Compound **36f** was synthesized from compound **35f** (0.23 g, 0.20 mmol), 4-(piperidin-1-yl)butanoic acid hydrochloride (0.10 g, 0.49 mmol) and DCC (0.20 g, 0.98 mmol) following a procedure similar to that used for the synthesis of compound **36a**. The title compound was obtained as a colorless oil (0.17 g, 60%).



37

Compound 37. Compound **14b** (0.35 g, 0.66 mmol, 1 equiv) was dissolved in dry DCM (10 mL) and 1 drop of DMF was added, followed by the dropwise addition of $SOCl_2$ (0.18 g, 0.99 mmol, 1.5 equiv). The reaction mixture was stirred at 23 °C for 0.5 h. Then DCM and excess $SOCl_2$ were removed under reduced pressure. The obtained intermediate product was dissolved in dry DMF (10 mL) and potassium phthalimide (0.15 g, 0.79 mmol, 1.2 equiv) was added. The mixture was stirred at 80 °C for 1 h. The reaction mixture was cooled to 23 °C and DMF was removed under reduced pressure. Then water (30 mL) was added, and the mixture was extracted by DCM (30 mL) for three times. The organic phase was combined, dried over anhydrous $MgSO_4$ and filtered. The filtrate was concentrated and purified by column chromatography (SiO_2) with hexane/EtOAc = 20/1 as the mobile phase to give the title compound as a yellow solid (0.30 g, 75%).



38

Compound 38. Compound **37** (0.21 g, 0.32 mmol, 1 equiv) and hydrazine monohydrate ($NH_2NH_2 \cdot H_2O$, 0.16 g, 3.17 mmol, 10 equiv) were dissolved in EtOH (10 mL). The reaction

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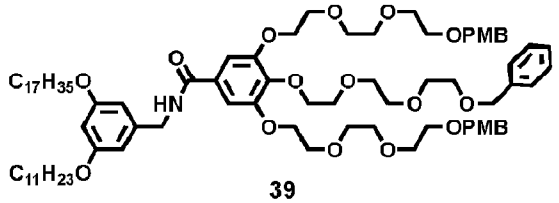
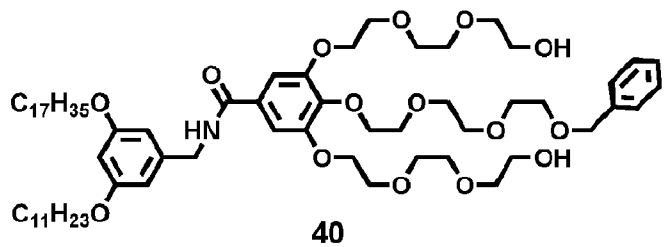
mixture was heated at reflux for 2 h. The mixture was cooled to 23 °C and the solvent was removed under reduced pressure. Then water (10 mL) was added, and the mixture was extracted by DCM (10 mL) for three times. The organic phases were combined, dried over anhydrous $MgSO_4$ and filtered. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.20 g, 99%).



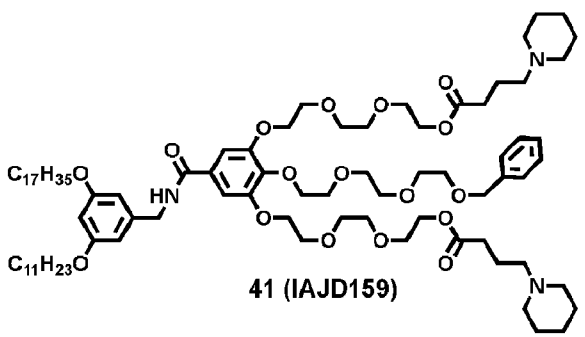
39

Compound 39. Compound **39** was synthesized from compound **38** (70 mg, 0.13 mmol) and compound **33** (0.12 g, 0.13 mmol) following a procedure similar to that used for the synthesis of compound **34a**. The title compound was obtained as a colorless oil (0.14 g, 78%).



40

Compound 40. Compound **40** was synthesized from compound **39** (0.14 g, 0.098 mmol) and DDQ (56 mg, 0.25 mmol) following a procedure similar to that used for the synthesis of compound **35a**. The title compound was obtained as a colorless oil (80 mg, 69%).



41 (IAJD159)

Compound 41 (IAJD159). Compound **41** was synthesized from compound **40** (80 mg, 0.068 mmol), 4-(piperidin-1-yl)butanoic acid hydrochloride (36 mg, 0.17 mmol) and DCC (70 mg, 0.34 mmol) following a procedure similar to that used for the synthesis of compound **36a**. The title compound was obtained as a colorless oil (77 mg, 77%).

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The dimensions of DNPs co-assembled from IAJDs and Luc-mRNA by injection in acetate buffer (10 mM, pH 4.0) and luminescence expression results in HEK 293T cells (125 ng/well, $n=3$) is shown in the Table 16 below.

Table 16.

IAJD	Buffer (10 mM)	C_{IAJD} (mg/mL)	$C_{Luc\text{-}mRNA}$ (mg/mL)	Size (d, nm)	PDI	Luminescence	Standard deviation
66[a]	Acetate (pH 4.0)	4	0.10	165	0.320	234	43
71[a]	Acetate (pH 4.0)	4	0.10	237	0.451	473,548	123,589
81[a]	Acetate (pH 4.0)	4	0.10	111	0.287	206,161	21,200
86[a]	Acetate (pH 4.0)	4	0.10	91	0.234	47,066	21,163
105[a]	Acetate (pH 4.0)	4	0.10	94	0.294	85	21
106[a]	Acetate (pH 4.0)	4	0.10	105	0.275	309	116
107[a]	Acetate (pH 4.0)	4	0.10	146	0.279	93,745	12,116
113	Acetate (pH 4.0)	4	0.10	181	0.511	78,118	13,787
114	Acetate (pH 4.0)	4	0.10	105	0.351	1,580	1,541
115	Acetate (pH 4.0)	4	0.10	123	0.367	390,021	17,022
116	Acetate (pH 4.0)	4	0.10	150	0.264	192,213	35,528
117	Acetate (pH 4.0)	4	0.10	255	0.367	516,522	54,783
118	Acetate (pH 4.0)	4	0.10	104	0.278	568,354	15,820
119	Acetate (pH 4.0)	4	0.10	163	0.459	565,906	39,932
120	Acetate (pH 4.0)	4	0.10	212	0.474	443,836	64,704
122	Acetate (pH 4.0)	4	0.10	215	0.337	55,101	3,169
124	Acetate (pH 4.0)	4	0.10	276	0.280	145,115	5,020
125	Acetate (pH 4.0)	4	0.10	255	0.300	119,405	21,570
126	Acetate (pH 4.0)	4	0.10	158	0.247	675	137

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127	Acetate (pH 4.0)	4	0.10	276	0.270	147,731	24,320
128	Acetate (pH 4.0)	4	0.10	291	0.381	449,311	104,462
130	Acetate (pH 4.0)	4	0.10	172	0.271	76,345	5,712
133	Acetate (pH 4.0)	4	0.10	228	0.414	107,307	17,263
135	Acetate (pH 4.0)	4	0.10	92	0.267	432	236
136	Acetate (pH 4.0)	4	0.10	149	0.307	283,656	146,748
138	Acetate (pH 4.0)	4	0.10	99	0.305	102,620	5,680
141	Acetate (pH 4.0)	4	0.10	158	0.313	140,589	107,411
142	Acetate (pH 4.0)	4	0.10	230	0.293	463,750	213,429
143	Acetate (pH 4.0)	4	0.10	169	0.270	71,851	10,736
144	Acetate (pH 4.0)	4	0.10	236	0.432	253,672	25,719
145	Acetate (pH 4.0)	4	0.10	194	0.306	296	353
146	Acetate (pH 4.0)	4	0.10	231	0.276	50,625	6,496
147	Acetate (pH 4.0)	4	0.10	144	0.287	5,844	1,645
148	Acetate (pH 4.0)	4	0.10	363	0.339	139,420	10,045
149	Acetate (pH 4.0)	4	0.10	193	0.304	149,935	22,183
150	Acetate (pH 4.0)	4	0.10	285	0.454	134,225	13,323
151	Acetate (pH 4.0)	4	0.10	287	0.216	9,310	1,977
152	Acetate (pH 4.0)	4	0.10	281	0.370	88,202	24,833
153	Acetate (pH 4.0)	4	0.10	232	0.238	8,860	2,558
154	Acetate (pH 4.0)	4	0.10	429	0.407	17,871	6,419
161	Acetate (pH 4.0)	4	0.10	129	0.235	128	49
162	Acetate (pH 4.0)	4	0.10	191	0.301	222,235	32,067
171	Acetate (pH 4.0)	4	0.10	174	0.268	1,436	832

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172	Acetate (pH 4.0)	4	0.10	235	0.203	166,377	22,769
173	Acetate (pH 4.0)	4	0.10	139	0.301	2,302	410
177	Acetate (pH 4.0)	4	0.10	159	0.275	4,343	755
178	Acetate (pH 4.0)	4	0.10	344	0.223	235,419	43,906
110	Acetate (pH 4.0)	4	0.10	111	0.274	47	30
111	Acetate (pH 4.0)	4	0.10	99	0.232	41	35
155	Acetate (pH 4.0)	4	0.10	83	0.279	74	49
156	Acetate (pH 4.0)	4	0.10	75	0.236	59	23
157	Acetate (pH 4.0)	4	0.10	68	0.238	47	7
158	Acetate (pH 4.0)	4	0.10	75	0.246	83	36
159	Acetate (pH 4.0)	4	0.10	91	0.218	24,055	15,173
Negative Control (untreated cells)						81	5
MC3-Luc-mRNA						1,320,533	105,967

a. The synthesis and characterizations of IAJDs 66, 71, 81, 86, 105, 106, and 107 as well as the dimensions of DNPs and the luminescence expression results in HEK 293T cells are available in literature recently reported (Zhang et al., *J. Am. Chem. Soc.* **2021**, *143*, 17975−17982).

5. Luminescence Expression *In Vivo*

The Table 17 below shows the quantification of luminescence expression *in vivo*.

Table 17.

IAJD	**C_{IAJD} (mg/mL)**	**$C_{Luc\text{-}mRNA}$ (mg/mL)**	**pH**		**Size (d, nm)**	**PDI**	**T (h)[a]**	**Total flux (p/s)**	**Organs with luminescence[b]**
			Acetate buffer	**Sample**					
66	4	0.10	4.0	4.50	165	0.320	4	2.58×10^7	Lung (weak), liver (strong), spleen (strong)
71	4	0.10	4.0	4.39	237	0.451	5	3.79×10^7	Lung (weak), liver (weak),

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									spleen (strong)
81	4	0.10	4.0	4.38	111	0.287	6	2.32×10^7	Lung (weak), liver (strong), spleen (weak)
86	4	0.10	4.0	4.39	91	0.234	5	4.73×10^7	Lung (weak), liver (strong), spleen (weak)
105	4	0.10	4.0	4.42	94	0.294	6	4.74×10^7	Lung (weak), liver (strong), spleen (weak)
106	4	0.10	4.0	4.41	105	0.275	6	3.48×10^7	Lung (weak), liver (weak), spleen (strong)
107	4	0.10	4.0	4.42	146	0.279	6	2.19×10^7	Lung (weak), liver (weak), spleen (strong)
113	4	0.10	4.0	4.48	181	0.511	5	1.70×10^7	Spleen (strong)
114	4	0.10	4.0	4.46	105	0.351	5	3.56×10^6	Lung (weak), liver (weak), spleen (strong)
115	4	0.10	4.0	4.44	123	0.367	5	4.59×10^6	Lung (weak), spleen (strong)
116	4	0.10	4.0	4.47	150	0.264	5	2.38×10^7	Lung (weak), liver (weak), spleen (strong)
117	4	0.10	4.0	4.40	255	0.367	5	8.64×10^6	Lung (weak), liver (weak), spleen (strong)
118	4	0.10	4.0	4.43	104	0.278	6	1.46×10^7	Lung (weak), liver (weak), spleen (strong)
119	4	0.10	4.0	4.36	163	0.459	5	8.22×10^7	Lung (weak), liver (weak), spleen (strong)
120	4	0.10	4.0	4.40	212	0.474	5	2.48×10^7	Spleen (strong)
122	4	0.10	4.0	4.49	215	0.337	5	2.50×10^7	Lung (weak), spleen (strong)
124	4	0.10	4.0	4.51	276	0.280	5	3.77×10^7	Lung (weak), spleen (strong)
125	4	0.10	4.0	4.44	255	0.300	5	2.15×10^8	Lung (weak), liver (weak), spleen (strong)
126	4	0.10	4.0	4.48	158	0.247	5	3.21×10^7	Lung (weak), liver (weak), spleen (strong)
127	4	0.10	4.0	4.46	276	0.27	5	$1.07 \times$	Lung (weak),

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						0		10^8	liver (weak), spleen (strong)
128	4	0.10	4.0	4.40	291	0.381	5	5.53×10^7	Lung (weak), liver (weak), spleen (strong)
130	4	0.10	4.0	4.50	172	0.271	5	1.18×10^8	Lung (weak), liver (weak), spleen (strong)
133	4	0.10	4.0	4.63	228	0.414	6	1.11×10^7	Lung (strong), spleen (weak)
135	4	0.10	4.0	4.95	92	0.267	5	1.22×10^7	Lung (weak), liver (strong), spleen (weak)
136	4	0.10	4.0	4.57	149	0.307	5	1.04×10^7	Lung (weak), liver (weak), spleen (strong)
138	4	0.10	4.0	4.52	99	0.305	5	1.49×10^7	Lung (weak), spleen (strong)
141	4	0.10	4.0	4.62	158	0.313	5	2.06×10^7	Lung (weak), liver (strong), spleen (strong)
142	4	0.10	4.0	4.52	230	0.293	5	2.51×10^7	Lung (weak), liver (strong), spleen (strong)
143	4	0.10	4.0	4.48	169	0.270	5	1.28×10^7	Lung (strong), liver (strong), spleen (strong)
144	4	0.10	4.0	4.40	236	0.432	5	1.39×10^7	Lung (weak), liver (weak), spleen (strong)
145	4	0.10	4.0	4.51	194	0.306	5	9.25×10^6	Lung (strong), liver (weak), spleen (strong)
146	4	0.10	4.0	4.40	231	0.276	5	4.70×10^7	Lung (strong), liver (strong), spleen (strong)
147	4	0.10	4.0	4.50	144	0.287	5	9.24×10^6	Lung (weak), liver (weak), spleen (strong)
148	4	0.10	4.0	4.46	363	0.339	5	6.33×10^7	Lung (weak), liver (weak), spleen (strong)
149	4	0.10	4.0	4.50	193	0.304	5	2.26×10^7	Lung (weak), liver (weak), spleen (strong)
150	4	0.10	4.0	4.51	285	0.454	5	3.47×10^7	Lung (weak), liver (weak), spleen (strong)

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151	4	0.10	4.0	4.57	287	0.216	5	3.12×10^7	Lung (weak), liver (weak), spleen (strong)
152	4	0.10	4.0	4.45	281	0.370	5	1.97×10^7	Liver (weak), spleen (strong)
153	4	0.10	4.0	4.52	232	0.238	6	7.33×10^7	Lung (weak), liver (strong), spleen (strong)
154	4	0.10	4.0	4.47	429	0.407	6	2.39×10^7	Lung (weak), liver (weak), spleen (strong)
161	4	0.10	4.0	4.97	129	0.235	5	8.48×10^7	Lung (weak), liver (strong), spleen (strong)
162	4	0.10	4.0	4.66	191	0.301	5	2.03×10^8	Lung (weak), liver (weak), spleen (strong)
171	4	0.10	4.0	4.88	174	0.268	5	7.68×10^7	Lung (weak), liver (strong), spleen (strong)
172	4	0.10	4.0	4.84	235	0.203	5	6.82×10^7	Lung (weak), liver (weak), spleen (strong)
173	4	0.10	4.0	4.79	139	0.301	5	1.23×10^8	Lung (weak), liver (strong), spleen (weak)
177	4	0.10	4.0	4.76	159	0.275	6	9.18×10^7	Lung (weak), liver (strong), spleen (strong)
178	4	0.10	4.0	4.58	344	0.223	6	4.05×10^8	Lung (weak), liver (weak), spleen (strong)
110	4	0.10	4.0	4.91	111	0.274	5	8.85×10^7	Lung (strong)
111	4	0.10	4.0	4.84	99	0.232	5	7.21×10^7	Lung (strong), spleen (weak)
155	4	0.10	4.0	4.93	83	0.279	5	1.01×10^8	Lung (strong), spleen (weak)
156	4	0.10	4.0	4.82	75	0.236	5	5.83×10^6	Lung (strong)
157	4	0.10	4.0	4.84	68	0.238	5	3.66×10^7	Lung (strong)
158	4	0.10	4.0	4.83	75	0.246	5	1.23×10^7	Lung (strong)
159	4	0.10	4.0	4.72	91	0.218	6	6.86×10^7	Lung (strong)

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a. Time from injection of DNP (co-assembled from IAJDs and Luc-mRNA) solution to the imaging of mice on the IVIS imaging system.

b. Organs showing luminescence indicated the expression of luciferase enzyme in the corresponding organs which interacted with D-Luciferin to generate oxy-Luciferin emitting light. Strong meant the luminescence intensity in that organ was relatively high, compared to other organs. The organ highlighted with green color meant it showed strongest luminescence intensity. Weak meant the luminescence intensity in that organ was relatively low, compared to other organs.

6. Comparison of Luminescence Expression *In Vivo* by Symmetric IAJDs and Nonsymmetric IAJDs

The Table 18 below shows in vivo activity of symmetric IAJDs with identical number of carbons on each alkyl chain

Table 18.

IAJD[a]	**Number of carbons on each alkyl chain**	**Average number of carbons per alkyl chain**	**Ionizable amine**[b]	***In vivo* activity (total flux, p/s)**
76	8+8 (EH+EH)	8	MP	7.81×10^7
87	8+8 (EH+EH)	8	HP	3.24×10^7
88	8+8	8	MP	2.34×10^6
89	8+8	8	HP	5.92×10^7
77	9+9	9	MP	2.44×10^6
91	10+10	10	HP	9.94×10^6
66	12+12	12	MP	2.58×10^7
71	12+12	12	HP	3.79×10^7
74	14+14	14	MP	1.71×10^7
75	16+16	16	MP	3.37×10^7
98	18+18	18	MP	2.06×10^6
99	18+18	18	HP	4.49×10^6

a. The synthesis and characterizations of IAJDs 66, 71, 74, 75, 76, 77, 87, 88, 89, 91, 98 and 99 as well as their luminescence expression results *in vivo* were available in the literature as recently reported.

b. MP: methyl piperazine; HP: hydroxy piperazine.

The Table 19 below shows in vivo activity of nonsymmetric IAJDs with different number of carbons on each alkyl chain.

Table 19.

IAJD	**Number of carbons on each alkyl chain**	**Average number of carbons per alkyl chain**	**Ionizable amine**[a]	***In vivo* activity (total flux, p/s)**

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81	11+18	14.5	MP	2.32×10^7
86	11+16	13.5	MP	4.73×10^7
105	11+14	12.5	MP	4.74×10^7
106	12+16	14	MP	3.48×10^7
107	12+18	15	MP	2.19×10^7
113	11+8	9.5	HP	1.70×10^7
114	11+10	10.5	MP	3.56×10^6
115	11+10	10.5	HP	4.59×10^6
116	11+13	12	MP	2.38×10^7
117	11+13	12	HP	8.64×10^6
118	11+15	13	MP	1.46×10^7
119	11+15	13	HP	8.22×10^7
120	11+16	13.5	HP	2.48×10^7
122	11+9	10	HP	2.50×10^7
124	11+12	11.5	HP	3.77×10^7
125	11+14	12.5	HP	2.15×10^8
126	11+17	14	MP	3.21×10^7
127	11+17	14	HP	1.07×10^8
128	11+18	14.5	HP	5.53×10^7
130	11+8 (11+EH)	9.5	HP	1.18×10^8
135	15+8 (15+EH)	11.5	MP	1.22×10^7
136	15+8 (15+EH)	11.5	HP	1.04×10^7
138	15+8	11.5	HP	1.49×10^7
141	15+10	12.5	MP	2.06×10^7
142	15+10	12.5	HP	2.51×10^7
143	15+12	13.5	MP	1.28×10^7
144	15+12	13.5	HP	1.39×10^7
145	15+13	14	MP	9.25×10^6
146	15+13	14	HP	4.70×10^7
147	15+14	14.5	MP	9.24×10^6
148	15+14	14.5	HP	6.33×10^7
149	15+16	15.5	MP	2.26×10^7
150	15+16	15.5	HP	3.47×10^7
151	15+17	16	MP	3.12×10^7
152	15+17	16	HP	1.97×10^7
153	15+18	16.5	MP	7.33×10^7
154	15+18	16.5	HP	2.39×10^7
161	13+8 (13+EH)	10.5	MP	8.48×10^7
162	13+8 (13+EH)	10.5	HP	2.03×10^8
171	13+14	13.5	MP	7.68×10^7
172	13+14	13.5	HP	6.82×10^7
173	13+16	14.5	MP	1.23×10^8
177	13+18	15.5	MP	9.18×10^7
178	13+18	15.5	HP	4.05×10^8

The Table 20 below shows representative ratios between in vivo activities of nonsymmetric IAJDs and corresponding symmetric IAJDs.

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Table 20.

Nonsymmetric IAJDs	*In vivo* activity (total flux, p/s)	Corresponding symmetric IAJDs	*In vivo* activity (total flux, p/s)	Ratio between *in vivo* activities (nonsymmetric/ symmetric IAJDs)
125 (11/14-HP)	2.15×10^8	**71 (12/12-HP)**	3.79×10^7	IAJD125/IAJD71 = **5.7**
130 (11/EH-HP)	1.18×10^8	**87 (EH/EH-HP)**	3.24×10^7	IAJD130/IAJD87 = **3.6**
162 (13/EH-HP)	2.03×10^8	**87 (EH/EH-HP)**	3.24×10^7	IAJD162/IAJD87 = **6.3**
171 (13/14-MP)	7.68×10^7	**74 (14/14-MP)**	1.71×10^7	IAJD171/IAJD74 = **4.5**
173 (13/16-MP)	1.23×10^8	**75 (16/16-MP)**	3.37×10^7	IAJD173/IAJD75 = **3.6**
81 (11/18-MP)	2.32×10^7	**98 (18/18-MP)**	2.06×10^6	IAJD81/IAJD98 = **11.3**
128 (11/18-HP)	5.53×10^7	**99 (18/18-HP)**	4.49×10^6	IAJD128/IAJD99 = **12.3**
177 (13/18-MP)	9.18×10^7	**98 (18/18-MP)**	2.06×10^6	IAJD177/IAJD98 = **44.6**
178 (13/18-HP)	4.05×10^8	**99 (18/18-HP)**	4.49×10^6	IAJD178/IAJD99 = **90.2**
153 (15/18-MP)	7.33×10^7	**98 (18/18-MP)**	2.06×10^6	IAJD153/IAJD98 = **35.6**
154 (15/18-HP)	2.39×10^7	**99 (18/18-HP)**	4.49×10^6	IAJD154/IAJD99 = **5.3**
MC3-Luc-mRNA	17.0×10^8	**178 (13/18-HP)**	4.05×10^8	MC3/178 = **4.2**

The DLS data of DNPs co-assembled from IAJDs and Luc-mRNA is shown in Figure 63 through Figure 67. The stability of DNPs by DLS is shown in the Table 21 below.

Table 21. Stability of DNPs Determined by DLS After Different Periods of Time at 5 °C

IAJD (storage time)	C_{IAJD} (mg/mL)	$C_{Luc\text{-}mRNA}$ (mg/mL)	pH		Original size (d, nm)	PDI	Size (d, nm)	PDI	Ranking of stability[c]
			Acetate buffer	Sample					
113 (70 days)	4.0	0.1	4.00	4.48	181 (B)[b]	0.511	184 (B)	0.365	Excellent
114 (75 days)	4.0	0.1	4.00	4.46	105 (U)	0.351	107 (B)	0.369	Good
115 (70 days)	4.0	0.1	4.00	4.44	123 (U)	0.367	134 (U)	0.383	Excellent
116 (62 days)	4.0	0.1	4.00	4.47	150 (U)	0.264	168 (U)	0.272	Excellent

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117 (70 days)	4.0	0.1	4.00	4.40	255 (B)	0.367	323 (B)	0.312	Excellent
118 (75 days)	4.0	0.1	4.00	4.43	104 (B)	0.278	105 (B)	0.262	Excellent
119 (62 days)	4.0	0.1	4.00	4.36	163 (M)	0.459	254 (B)	0.489	Good
120 (70 days)	4.0	0.1	4.00	4.40	212 (B)	0.474	177 (B)	0.289	Excellent
122 (62 days)	4.0	0.1	4.00	4.49	215 (B)	0.337	192 (B)	0.346	Excellent
124 (62 days)	4.0	0.1	4.00	4.51	276 (U)	0.280	284 (B)	0.240	Good
125 (62 days)	4.0	0.1	4.00	4.44	255 (U)	0.300	276 (U)	0.283	Excellent
126 (70 days)	4.0	0.1	4.00	4.48	158 (U)	0.247	154 (U)	0.239	Excellent
127 (62 days)	4.0	0.1	4.00	4.46	276 (U)	0.270	263 (B)	0.248	Good
128 (70 days)	4.0	0.1	4.00	4.40	291 (B)	0.381	210 (B)	0.459	Excellent
130 (62 days)	4.0	0.1	4.00	4.50	172 (U)	0.271	149 (U)	0.282	Excellent
133 (42 days)	4.0	0.1	4.00	4.63	228 (B)	0.414	216 (B)	0.454	Excellent
135 (27 days)	4.0	0.1	4.00	4.95	92 (U)	0.267	92 (U)	0.278	Excellent
136 (27 days)	4.0	0.1	4.00	4.57	149 (U)	0.307	153 (U)	0.296	Excellent
138 (35 days)	4.0	0.1	4.00	4.52	99 (U)	0.305	112 (B)	0.261	Good
141 (27 days)	4.0	0.1	4.00	4.62	158 (U)	0.313	156 (U)	0.286	Excellent
142 (27 days)	4.0	0.1	4.00	4.52	230 (B)	0.293	214 (U)	0.266	Excellent
143 (27 days)	4.0	0.1	4.00	4.48	169 (U)	0.270	166 (U)	0.243	Excellent
144 (27 days)	4.0	0.1	4.00	4.40	236 (B)	0.432	216 (U)	0.276	Excellent
145 (27 days)	4.0	0.1	4.00	4.51	194 (U)	0.306	189 (U)	0.330	Excellent
146 (27 days)	4.0	0.1	4.00	4.40	231 (U)	0.276	227 (U)	0.285	Excellent
147 (19 days)	4.0	0.1	4.00	4.50	144 (U)	0.287	145 (U)	0.283	Excellent
148 (19 days)	4.0	0.1	4.00	4.46	363 (U)	0.339	405 (B)	0.374	Good
149 (19 days)	4.0	0.1	4.00	4.50	193 (B)	0.304	193 (B)	0.291	Excellent

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150 (19 days)	4.0	0.1	4.00	4.51	285 (B)	0.454	308 (U)	0.314	Excellent
151 (35 days)	4.0	0.1	4.00	4.57	287 (U)	0.216	271 (U)	0.240	Excellent
152 (35 days)	4.0	0.1	4.00	4.45	281 (B)	0.370	253 (U)	0.286	Excellent
153 (35 days)	4.0	0.1	4.00	4.52	232 (U)	0.238	222 (U)	0.232	Excellent
154 (35 days)	4.0	0.1	4.00	4.47	429 (B)	0.407	372 (B)	0.471	Excellent
161 (21 days)	4.0	0.1	4.00	4.97	129 (U)	0.235	125 (U)	0.226	Excellent
162 (21 days)	4.0	0.1	4.00	4.66	191 (B)	0.301	182 (B)	0.291	Excellent
171 (21 days)	4.0	0.1	4.00	4.88	174 (U)	0.268	175 (U)	0.277	Excellent
172 (14 days)	4.0	0.1	4.00	4.84	235 (U)	0.203	225 (U)	0.219	Excellent
173 (21 days)	4.0	0.1	4.00	4.79	139 (U)	0.301	132 (B)	0.376	Good
177 (21 days)	4.0	0.1	4.00	4.76	159 (B)	0.275	153 (B)	0.218	Excellent
178 (21 days)	4.0	0.1	4.00	4.58	344 (U)	0.223	261 (U)	0.261	Excellent
110 (75 days)	4.0	0.1	4.00	4.91	111 (U)	0.274	103 (U)	0.222	Excellent
111 (75 days)	4.0	0.1	4.00	4.84	99 (U)	0.232	96 (U)	0.227	Excellent
155 (19 days)	4.0	0.1	4.00	4.93	83 (U)	0.279	86 (U)	0.259	Excellent
156 (19 days)	4.0	0.1	4.00	4.82	75 (U)	0.236	78 (U)	0.242	Excellent
157 (19 days)	4.0	0.1	4.00	4.84	68 (U)	0.238	71 (U)	0.255	Excellent
158 (19 days)	4.0	0.1	4.00	4.83	75 (U)	0.246	77 (U)	0.258	Excellent
159 (11 days)	4.0	0.1	4.00	4.72	91 (U)	0.218	93 (U)	0.220	Excellent

a. All samples were prepared by injection in acetate buffer.

b. U: DLS curve showed unimodal size distribution; B: DLS curve showed bimodal size distribution; M: DLS curve showed multimodal size distribution.

c. Ranking of stability: Excellent (highlighted in green color) – the original sizes and distribution types remained almost the same after storage; Good – the original sizes and distribution types changed slightly (size difference was less than 100 nm) or the original

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size remained almost the same but the DLS curve showed more size distributions after storage.

9. Serum Stability of DNPs

(1) DNPs assembled from IAJD125 and IAJD155

Sample preparation: injected 25 μL ethanol stock solution of IAJD125/155 (80 mg/mL) into the mixture of 462.5 μL acetate buffer (10 mM, pH 4.0) and 12.5 μL Luc-mRNA (4.0 mg/mL in water), followed by vortex for 5 seconds.

Final composition of DNP solution: IAJD125/155 (4.0 mg/mL) + Luc-mRNA (0.1 mg/mL) in 500 μL acetate buffer. The dimension of assemblies in solution was checked by DLS.

(2) Mixture of DNPs and fetal bovine serum

DNPs in 1% Serum: Serum (5.5 μL) + [500 μL solution of IAJD (4.0 mg/mL) and Luc-mRNA (0.1 mg/mL].

The Table 22 below shows the dimensions of DNPs assembled from IAJD125 in 1% fetal bovine serum.

Table 22.

Samples	Size (nm)	PDI
DNPs of IAJD125 in acetate buffer (pH 4.0)	196	0.273
DNPs of IAJD125 in 1% Serum (t = 0 min)	207	0.234
DNPs of IAJD125 in 1% Serum (t = 10 min)	206	0.290
DNPs of IAJD125 in 1% Serum (t = 20 min)	199	0.303
DNPs of IAJD125 in 1% Serum (t = 30 min)	197	0.256
DNPs of IAJD125 in 1% Serum (t = 40 min)	200	0.284
DNPs of IAJD125 in 1% Serum (t = 50 min)	190	0.339
DNPs of IAJD125 in 1% Serum (t = 60 min)	194	0.258

Figure 76 shows dimensions of DNPs assembled from IAJD125 in 1% fetal bovine serum.

The Table 23 below shows the dimensions of DNPs assembled from IAJD155 in 1% fetal bovine serum.

Table 23.

Samples	Size (nm)	PDI
DNPs of IAJD155 in acetate buffer (pH 4.0)	83	0.256
DNPs of IAJD155 in 1% Serum (t = 0 min)	93	0.186
DNPs of IAJD155 in 1% Serum (t = 10 min)	101	0.171

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DNPs of IAJD155 in 1% Serum (t = 20 min)	105	0.186
DNPs of IAJD155 in 1% Serum (t = 30 min)	107	0.173
DNPs of IAJD155 in 1% Serum (t = 40 min)	108	0.161
DNPs of IAJD155 in 1% Serum (t = 50 min)	111	0.174
DNPs of IAJD155 in 1% Serum (t = 60 min)	108	0.149

Figure 77 shows dimensions of DNPs assembled from IAJD155 in 1% fetal bovine serum.

Table 24. pKa Measurements of Individual IAJD Molecules

IAJD No.	**pK_a**
IAJD 113	6.52
IAJD 114	6.43
IAJD 115	6.32
IAJD 116	6.51
IAJD 117	6.49
IAJD 118	6.33
IAJD 119	6.44
IAJD 120	6.37
IAJD 122	6.66
IAJD 124	6.65
IAJD 125	6.64
IAJD 126	6.52
IAJD 127	6.52
IAJD 128	6.68
IAJD 130	6.62
IAJD 133	6.57
IAJD 135	6.32
IAJD 136	6.50
IAJD 138	6.62
IAJD 141	6.56
IAJD 142	6.69
IAJD 143	6.33
IAJD 144	6.51
IAJD 145	6.28
IAJD 146	6.59
IAJD 147	6.32
IAJD 148	6.65
IAJD 149	6.50
IAJD 150	6.65
IAJD 151	6.39
IAJD 152	6.65
IAJD 153	6.42
IAJD 154	6.48
IAJD 161	6.38
IAJD 162	6.33
IAJD 171	6.16
IAJD 172	6.03

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IAJD 173	5.91
IAJD 177	5.90
IAJD 178	5.97
IAJD 110	7.64
IAJD 111	7.58
IAJD 155	7.40
IAJD 156	7.43
IAJD 157	7.42
IAJD 158	7.38
IAJD 159	7.40

The titration curves and calculated pKa for IAJDs 110, 111, 113-120, 122, 124-128, 130, 133, 135, 136, 138, 141-159, are shown in Figure 68 through Figure 72.

DLS Data of DNPs After Dialysis in PBS

The 0.5 mL DNP solution (in acetate buffer) was dialyzed against 1000 mL 1X PBS buffer (pH 6.2) at 23 °C for 3 h in a dialysis tube with molecular weight cut-off of 3,500 – 14,000 (Spectrum Medical Instruments Inc. Spectra/Por molecular porous membrane tubing; Flat Width.: 45 mm; Diameter.: 29 mm & Vol/length.: 6.4 mL/cm). Figure 78 shows the DLS data of DNP 125 (a) and 178 (b) before and after dialysis in 1X PBS buffer for 3 h.

Example 4: Self-Assembly of Glycerol-Amphiphilic Janus Dendrimers Indicates Principles for the Selection of Stereochemistry by Biological Membranes

Homochirality is an essential signature of living matter. During the evolution of life, nature selected homochirality for most biological molecules and macromolecules (Franck et al., On the One Hand but Not the Other: The Challenge of the Origin and Survival of Homochirality in Prebiotic Chemistry. In *Chemistry for the 21th Century*; Wiley-VCH: Weinheim, 2000; pp 175–208; Lehn et al., *Chem. Soc. Rev.* **2007**, *36*, 151–160; Hein et al., *Top. Curr. Chem.* **2013**, *333*, 83–108; Rowan et al., *Angew. Chem. Int. Ed.* **1998**, *37*, 63–68; Weissbuch et al., *Chem. Rev.* **2011**, *111*, 3236–3267). L-amino acids and D-sugars have been selected as the main components of biological systems (Kumar et al., *Proc. Natl. Acad. Sci. U. S. A.* **2017**, *114*, 2474–2478; Yeom et al., *Adv. Mater.* **2020**, *32*, 1903878). However, phospholipids, the backbone of cell and endosome membranes, display different chirality in different living systems (Sojo et al., *Orig. Life. Evol. Biosph.* **2015**, *45*, 219–224; Liu et al., *Nat. Chem.* **2020**, *12*, 1029–1034). All natural phospholipids are homochiral. Nevertheless, a case of dual homochirality is also known for phospholipids. Archaea and bacteria, which are the two basal domains of life, display

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different homochirality in their phospholipids (Sojo et al., *Orig. Life. Evol. Biosph.* **2015**, *45*, 219–224; Liu et al., *Nat. Chem.* **2020**, *12*, 1029–1034; Coleman et al., *Genome Biol. Evol.* **2011**, *3*, 883–898.). Archaea are based on *sn*-glycerol-1-phosphate (G1P) enantiomer, while bacteria employ exclusively *sn*-glycerol-3-phosphate (G3P) enantiomer (Koga et al., *J. Mol. Evol.* **1998**, *46*, 54–63.). G1P and G3P exhibit opposite handedness, but both of them provide efficient membranes. A possible explanation is that the enzyme producing G1P existed simultaneously with that producing G3P but they evolved independently after the divergence of archaea and bacteria (Sojo et al., *Orig. Life. Evol. Biosph.* **2015**, *45*, 219–224; Liu et al., *Nat. Chem.* **2020**, *12*, 1029–1034).

Vesicles, an important class of biological assemblies, represent the basis of cell membranes. Synthetic variants, including liposomes (Bangham et al., *J. Mol. Biol.* **1964**, *8*, 660−668; Bangham et al., *J. Mol. Biol.* **1965**, *13*, 238–252; Seoane et al., *J. Am. Chem. Soc.* **2018**, *140*, 8388–8391; Ringsdorf et al., *Angew. Chem. Int. Ed. Engl.* **1988**, *27*, 113–158; Kunitake et al., *Angew. Chem. Int. Ed. Engl.* **1992**, *31*, 709–726; Brea et al., *Chem. Eur. J.* **2015**, *21*, 12564–12570; Flores et al., *Chem. Soc. Rev.* **2020**, *49*, 4602–4614; Bangham et al., *J. Mol. Biol.* **1964**, *8*, 660−668; Bangham et al., *J. Mol. Biol.* **1965**, *13*, 238–252; Seoane et al., *J. Am. Chem. Soc.* **2018**, *140*, 8388–8391; Ringsdorf et al., *Angew. Chem. Int. Ed. Engl.* **1988**, *27*, 113–158; Kunitake et al., *Angew. Chem. Int. Ed. Engl.* **1992**, *31*, 709–726; Brea et al., *Chem. Eur. J.* **2015**, *21*, 12564–12570; Flores et al., *Chem. Soc. Rev.* **2020**, *49*, 4602–4614), stealth liposomes (Lasic et al., *Chem. Rev.* **1995**, *95*, 2601–2628: Immordino et al., *Int. J. Nanomedicine* **2006**, *1*, 297−315), and polymersomes (Discher et al., *Science* **1999**, *284*, 1143−1146; Lee et al., *J. Controlled Release* **2012**, *161*, 473−483; Deming et al., *WIREs Nanomed. Nanobiotechnol.* **2014**, *6*, 283−297) have been elaborated to mimic natural cells. In addition to being models for biological membranes synthetic vesicles have been used to deliver active agents such as drugs and genes in *vitro* and in *vivo* (Allen et al., *Science* **2004**, *303*, 1818–1822; Torchilin et al., *Nat. Rev. Drug Discovery* **2005**, *4*, 145–160; de Jong et al., *Acc. Chem. Res.* **2019**, *52*, 1761−1770; EL Andaloussi et al., *Nat. Rev. Drug. Discov.* **2013**, *12*, 347–357; Dong et al., *J. Am. Chem. Soc.* **2018**, *140*, 16264–16274; Lee et al., *Nat. Biotechnol.* **2005**, *23*, 1517−1526; Tu et al., *Chem. Rev.* **2016**, *116*, 2023–2078; Thota et al., *Chem. Rev.* **2016**, *116*, 2079–2102; Weiss et al., *Nat. Mater.* **2018**, *17*, 89–96; Gillies et al., *Drug Discov. Today* **2005**, *10*, 35–43; Dendrimers and Other Dendritic Polymers; Tomalia, D. A.; Fréchet, J. M. J., Eds.; Wiley: Amsterdam, The

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Netherlands, 2001). Liposomes assembled from phospholipids are unstable because the alkene units in the hydrophobic tails are readily oxidized and exhibit poor mechanical properties. Stealth liposomes co-assembled from phospholipids, phospholipid conjugated with poly(ethylene glycol) (PEG) and cholesterol are more stable, but are polydisperse requiring time-consuming fractionation to desirable dimensions and narrow polydispersity. Polymersomes assembled from amphiphilic block copolymers exhibit excellent stability but are not always biocompatible and their membrane thickness is wider than that of cell membranes (Discher et al., *Science* **1999**, *284*, 1143−1146; Lee et al., *J. Controlled Release* **2012**, *161*, 473−483; Deming et al., *WIREs Nanomed. Nanobiotechnol.* **2014**, *6*, 283−297).

The previous studies have developed a class of vesicles named dendrimersomes (DSs), which are assembled from amphiphilic JDs (Percec et al., *Science* **2010**, *328*, 1009–1014; Percec et al., *J. Am. Chem. Soc.* **2013**, *135*, 9055–9077; Xiao et al., *J. Am. Chem. Soc.* **2016**, *138*, 12655–12663; Rodriguez-Emmenegger et al., *Proc. Natl. Acad. Sci. U. S. A.* **2019**, *116*, 5376–5382; Sherman et al., *Chem. Rev.* **2017**, *117*, 6538–6631; Peterca et al., *J. Am. Chem. Soc.* **2011**, *133*, 20507–20520). DSs exhibit better stability than liposomes and can be functionalized on their surface through their multivalency both in the hydrophilic and hydrophobic parts of JDs (Torre et al., *Proc. Natl. Acad. Sci. U.S.A.* **2019**, *116*, 15378–15385; Xiao et al., *Proc. Natl. Acad. Sci. U.S.A.* **2020**, *117*, 11931–11939; Li et al., *ACS Nano* **2020**, *14*, 7398–7411). Amphiphilic JDs with sugars conjugated on their hydrophilic part denoted Janus glycodendrimers (JGDs) self-assemble into glycodendrimersomes (GDSs), which mimic the glycan of biological membranes and bind sugar-binding proteins (Xiao et al., *Proc. Natl. Acad. Sci. U. S. A.* **2018**, *115*, E2509–E2518). DSs are assembled by injection of their JD in water-miscible solvent solution such as ethanol or THF into water or buffer. This simple assembling procedure provides monodisperse, impermeable and stable vesicles with excellent mechanical properties.

The properties of DSs rely on the architecture of the assembling amphiphilic JDs. JDs assembling DSs were classified into "Twin-Twin" and "Single-Single". "Twin-Twin" JDs are constructed by combinations of twin-hydrophobic and twin-hydrophilic dendrons with an achiral pentaerythritol (PE) or tris(hydroxymethyl)aminomethane (Tris) core, while "Single-Single" JDs are made from single-hydrophobic and single-hydrophilic dendrons connected by an ester or amide linker. Unilamellar and onion-like DSs have been discovered, designed and predicted (Peterca et al., *J. Am. Chem. Soc.* **2011**, *133*, 20507–20520) by self-assembly of "Twin-Twin" and "Single-Single" JDs (Zhang et al., *Proc. Natl. Acad. Sci. U. S. A.* **2014**, *111*, 9058–9063; Zhang et al.,

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ACS Nano **2014**, *8*, 1554−1565; Zhang et al., *J. Am. Chem. Soc.* **2015**, *137*, 13334−13344; Xiao et al., *Proc. Natl. Acad. Sci. U. S. A.* **2016**, *113*, 1162−1167; Kostina et al., *Soft Matter* **2020**, DOI: 10.1039/D0SM01097A). However, there is no report on amphiphilic JDs with a chiral or achiral glycerol core connecting hydrophilic and hydrophobic parts (Figure 79). Nature selects homochiral rather than racemic but not achiral glycerol (Figure 80) phospholipids to construct biological membranes.

The present studies investigated the self-assembly of all constitutional isomeric glycerol-JDs that facilitate the construction of homochiral, racemic and achiral structures *via* the stereochemistry of the branching point. Eight JDs including four homochiral, two racemic and two achiral were synthesized (Figure 80). Injection of their ethanol solution into water produced DSs with different dimensions and morphologies. They were characterized by dynamic light scattering (DLS) and cryogenic-transmission electron microscopy (cryo-TEM). Statistical analysis of the number of bilayers showed that homochiral JDs self-assemble predominantly unilamellar, achiral only unilamellar, while racemic predominantly multilamellar, onion-like DSs. This result provides the rationale for the selection of homochirality of phospholipids by nature.

Three libraries of glycerol-based JDs were synthesized by an orthogonal-modular methodology. For library 1 and 2, each library has two JDs with homochiral glycerol branching point and one JD with racemic branching point (Figure 80). Library 3 contains two constitutional isomeric glycerol-achiral JDs (Figure 80). The main building blocks employed in the orthogonal-modular synthesis include the hydrophilic acid (**5**), hydrophilic second-generation acid (**7**), hydrophobic acid (**10**), hydrophobic second-generation acid (**12**), *R/S/rac*- 2,2-dimethyl-1,3-dioxolane-4-methanol (isopropylidene glycerol) which contains the stereocenter and 5-hydroxy-2-phenyl-1,3-dioxane (Figure 81 through Figure 83).

The hydrophilic acid (**5**) was synthesized by introducing triethylene glycol monomethyl ether in the 3, 4, 5 positions of gallic acid. Two hydrophilic acids (**5**) were connected by a 2,2-bis(hydroxylmethyl)propionate linker to afford the hydrophilic second-generation acid (**7**). Similarly, hydrophobic acid (**10**) was synthesized by attaching dodecyl groups on the 3, 5 positions of 3,5-dihydroxybenzoic acid. The hydrophobic second-generation acid (**12**) was prepared by connecting two hydrophobic acids (**10**) with a 2,2-bis(hydroxylmethyl)propionate linker. For library 1, hydrophilic second-generation acid (**7**) reacted with *rac/R/S*- 2,2-dimethyl-1,3-dioxolane-4-methanol, followed by deprotection to afford the corresponding diol (**9**). The esterification of this hydrophilic diol and two hydrophobic first-generation acids produced **1*R***, **1*S*** and **1*rac*** in 75-86 %

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yield.

The synthesis of the three molecules of library 2 (**2*R***, **2*S*** and **2*rac***) (Figure 82) was accomplished with the first esterification performed with hydrophobic second-generation acid and the second esterification with the hydrophobic diol (**14**) and two hydrophilic acids (**5**) in 73-78 % yield. Detailed synthesis and characterization are in the supporting information. Figure 83 and Scheme 21 outline the synthesis of the achiral JDs (**3**) and (**4**).

Assembly of all eight glycerol-JDs was performed by injection of their ethanol solution (50 μL) into 1 mL of Milli-Q water. The size distribution and structure of resulting DSs were analyzed by DLS (Figure 84) and cryo-TEM. Previously, it was found that unilamellar and onion-like DSs assembled from "Twin-Twin" and "Single-Single" JDs displayed size-concentration dependence (Peterca et al., *J. Am. Chem. Soc.* **2011**, *133*, 20507–20520; Zhang et al., *ACS Nano* **2014**, *8*, 1554−1565). In a certain concentration range the size of DSs increased with the concentration of JDs injected in water or buffer. A similar size-concentration dependence was observed for glycerol-JDs. Figure 84 shows how the size of the resulting DSs increased with the concentration of JDs in water for **1*R***, **1*S*** and **1*rac*** for concentration range from 0.025 to 0.5 $mg \cdot mL^{-1}$. The dimensions of the assemblies were identical when the concentrations of the JDs in water were identical. Sizes of assemblies from mixtures of **1*R*** and **1*S*** in different ratios (1:3, 1:1 and 3:1), were also identical (Figure 84). This indicated that sizes of assemblies from JD enantiomers (**1*R*** and **1*S***) and racemic mixtures with different ratios of enantiomers were identical at the same concentrations of JDs in water. With the exception of the rotation of polarized light, the physical properties of enantiomers and their racemic mixtures must be identical. In addition, the square of the diameter of DSs displayed a linear dependence on concentration in the range from 0.025 to 0.4, for homochiral and racemic, and to 1.0 $mg \cdot mL^{-1}$ for achiral (Figure 84). This result is consistent with the size-concentration relationship of JDs already reported (Peterca et al., *J. Am. Chem. Soc.* **2011**, *133*, 20507–20520; Zhang et al., *ACS Nano* **2014**, *8*, 1554−1565) and helps predict the dimensions of DSs with the aid of JDs concentration.

2*R*, **2*S*** and **2*rac*** as well as mixtures of **2*R*** and **2*S*** with different molar ratios (1:3, 1:1 and 3:1) showed similar size-concentration dependence (Figure 84). The diameter rather than the square of diameter of DSs displayed a linear relationship versus concentration of glycerol-JD in the concentration range from 0.025 to 0.4 for homochiral and racemic and to 1.0 $mg \cdot mL^{-1}$ for achiral (Figure 84).

Although dimensions do not show differences when comparing homochiral, achiral and

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racemic JDs, the structure of the resulting DSs was investigated by cryo-TEM. Substantial differences were observed. cryo-TEM of enantiomers **2*R*** and **2*S*** (Figure 85) showed unilamellar DSs predominantly, being accompanied by a small amount of multilamellar. cryo-TEM images of enantiomers **2*rac*** (Figure 85) showed the opposite result, in which multilamellar DSs were predominant while unilamellar were observed in small amount. The red numbers in Figure 85 stand for the unilamellar while yellow numbers for multilamellar DSs. The calculated percentages of unilamellar DSs in the selected images for **2*R***, **2*S*** and **2*rac*** are 91%, 70% and 13%, respectively. Same trend was found for cryo-TEM of **1*R***, **1*S*** and **1*rac*** (Figure 86 through Figure 88). This indicates that homochiral JDs tend to self-assemble predominantly into unilamellar while racemic favor the formation of multilamellar DSs. Achiral DSs are all unilamellar.

Statistical analysis was used to organize the number of bilayers from cryo-TEM. The distribution of the number of lamellae in DSs was analyzed for each sample to construct the respective histograms (Figure 89). The histograms represent the distribution function of probability. The number of bilayers of racemic JDs (**1*rac*** and **2*rac***) exhibit a near normal distribution (Gaussian) as demonstrated by various statistic tests. However, the enantiomerically pure JDs (**1*R***, **1*S***, **2*R*** and **2*S***) exhibit chi-squared distribution (χ^2 distribution) with a shape parameter of 1 or 2. The difference in the distributions of number of bilayers for homochiral vs racemic glycerol-based JDs emphasizes the difference between the self-assembly structures, unilamellar vs multilamellar.

To investigate the normality of the distribution of racemic JDs, SPSS statistics software, which utilized normality tests of Kolmogorov-Smirnov and Shapiro-Wilk, was used. Both methods demonstrated that the distribution of racemic JDs (**1*rac*** and **2*rac***) were normal-like. For a more detailed study of distribution functions, the skewness (third standardized moment and a measure of the asymmetry) and kurtosis (fourth standardized moment and measure the extreme values in either tail) of the data were calculated (number of DSs layers). The skewness of the distributions of the number of bilayers for **1*rac*** and **2*rac*** is slightly positive (0.70 and 0.58, respectively). Positive skewness means the tails are extending to the right. These relatively low values are in line with the previous conclusion that their distribution is normal-like. Comparing the skewness of the distribution of racemic JDs with homochiral JDs shows that **1*R***, **1*S***, **2*R*** and **2*S*** are very positively skewed (1.977, 1.687, 3.313 and 3.029, respectively) and non-symmetric. This difference in distribution parameter (skewness) strongly supports the difference between homochiral (unilamellar) and racemic (multilamellar) glycerol-based JDs.

The second distribution parameter, which was investigated, is the kurtosis and excess kurtosis (kurtosis – 3). For normal distribution the kurtosis is 3 and the excess of kurtosis is zero.

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The distributions of racemic JDs have negative Kurtosis (-1.623 for **1*rac*** and -0.423 for **2*rac***), indicating that the peak is lower than the peak of a normal distribution. While homochiral JDs show distributions with positive kurtosis (3.932 for **1*R***, 2.797 for **1*S***, 10.980 for **2*R*** and 9.502 for **2*S***), indicating that corresponding peaks are higher than those of the normal distribution. This result highlights the difference between the number of bilayers of DSs assembled from homochiral and racemic glycerol-JDs.

Glycerol-JDs with homochiral, achiral and racemic branching points were synthesized. They self-assemble into monodisperse DSs by simple injection of their ethanol solution into water. DLS showed that the resulting DSs are concentration-size dependent with an identical trend regardless of achiral, homochiral or racemic branching point. Cryo-TEM and statistical analysis showed that homochiral JDs form predominantly unilamellar while racemic JDs favor multilamellar DSs. Achiral produce only unilamellar DSs. This study explains the selection of homochiral lipids but does not why achiral were eliminated, since single-bilayer membranes are required for the construction of biological cell membranes.

In conclusion, three libraries of constitutional isomeric glycerol-amphiphilic Janus dendrimers (JDs) with homochiral, racemic and achiral branching points were synthesized by an orthogonal-modular methodology. Monodisperse vesicles known as dendrimersomes (DSs) with predictable dimensions programmed by JD concentration were assembled by injection of their ethanol solution into water. DSs of JD enantiomers, racemic including mixtures of different enantiomers, and achiral exhibited identical concentration-size dependence. However, the number of bilayers of DSs from homochiral, achiral and racemic JDs determined by cryo-TEM were different. Statistical analysis of the number of bilayers demonstrated that homochiral and achiral (JDs) formed predominantly or only unilamellar while racemic (JDs) favored multilamellar DSs. These results provide a rationale for homochiral *vs* racemic selection.

The materials and methods employed in these experiments are now described.

Materials

(*R*)-2,2-dimethyl-1,3-dioxolane-4-methanol (TCI, 98%) and (*S*)-2,2-dimethyl-1,3-dioxolane-4-methanol (Alfa Aesar, 98%) and concentrated hydrochloric acid (Fisher, ACS grade) were used as received. 4-(Dimethylamino)pyridinium 4-toluenesulfonate (DPTS) was prepared according to a literature procedure (Moore et al., *Macromolecules* **1990**, *23*, 65–70). Benzyl 2,2-bis(hydroxymethyl)propionate was prepared according to a literature procedure (Córdova et al., *J.*

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Am. Chem. Soc. **2001**, *123*, 8248–8259). CH_2Cl_2 (DCM) was dried over CaH_2 and freshly distilled before use. All other reagents and solvents were obtained from commercial sources and were used without purification unless otherwise stated.

Methods

NMR. ^1H NMR and ^{13}C NMR spectra were recorded on Bruker DRX (500 MHz) spectrometer at 298 K. Deuterated chloroform ($CDCl_3$) was used as solvent. The resonance multiplicities in the ^1H NMR spectra are described as "s" (singlet), "d" (doublet), "t" (triplet), and "m" (multiplet) and broad resonances are indicated by "br". NMR chemical shifts were calculated with the residual protic solvent of $CDCl_3$ (^1H, δ 7.26 ppm; ^{13}C, δ 77.16 ppm) as internal reference (Figure 90 through Figure 105).

HPLC. The purity of the products was determined by high-performance liquid chromatography (HPLC) on a Shimadzu LC-10AT high-performance liquid chromatograph equipped with a Perkin Elmer LC- 100 oven, containing two Perkin-Elmer PL gel columns of 5×10^2 and 1×10^4 Å, a Shimadzu SPD- 10A UV detector ($\lambda = 254$ nm), a Shimadzu RID-10A RI-detector, and a PE Nelson Analytical 900 Series integrator data station. All measurements were performed with THF as eluent at a flow rate of 1.0 mL/min at 23 °C (Figure 109 through Figure 111).

Matrix-assisted laser desorption/ionization time of flight (MALDI-TOF) mass spectrometry. MALDI-TOF mass spectrometry was performed on a PerSeptive Biosystems-Voyager-DE (Framingham, MA) mass spectrometer equipped with a nitrogen laser (337 μm) and operated in linear mode. Internal calibration was performed using Angiotensin II and Bombesin as standards. The analytical samples were prepared by mixing the THF solutions of dendrimer molecules (2–5 mg/mL) and the matrix (2,5-dihydroxybenzoic acid) (10 mg/mL in THF) with a 1:1 to 1:5 v/v ratio. The prepared solution (0.5 μL) was then loaded on a MALDI plate and dried at room temperature before the plate was inserted into the vacuum chamber. The laser intensity and voltage were adjusted depending on the molecular weight and the nature of each analyzed molecule (Figure 106 through Figure 108).

Dynamic light scattering (DLS). Hydrodynamic diameters (D_h) of the self-assemblies of dendrimer molecules and their size distributions were measured on a Malvern Instruments particle sizer (Zetasizer® Nano S, Malvern Instruments, UK) equipped with 4 mW He-Ne laser (633 nm) and avalanche photodiode positioned at 175° to the beam and temperature controlled cuvette holder. Experiments were performed at 25 °C in triplicate. The distribution of radius was intensity.

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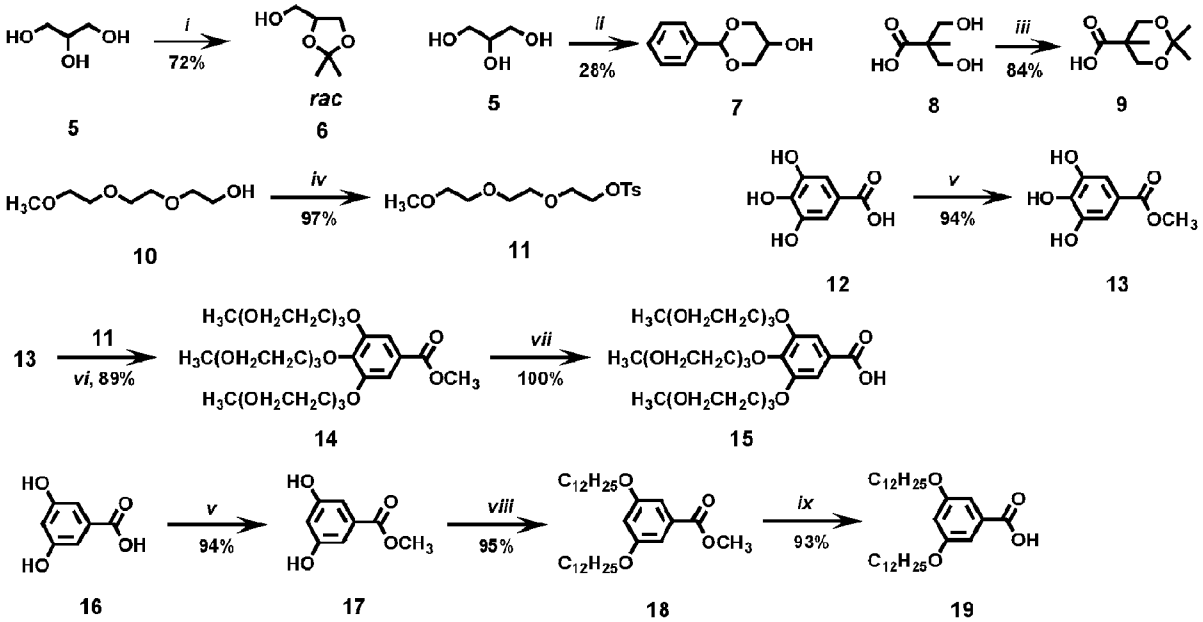
Cryogenic Transmission Electron Microscopy (Cryo-TEM). Morphologies of the Janus Dendrimer assemblies were characterized by cryo-TEM. Images were acquired on a Carl Zeiss Libra 120 microscope. Samples were prepared by plunge freezing of aqueous dispersion on plasma-treated lacey grids. The vitrified specimens were then transferred to a Gatan-910 cryoholder. The images were recorded at −170 °C with an acceleration voltage of 120 kV. The concentration of JD in the aqueous solution of DSs was 1 mg·mL^{-1}.

Synthesis

Synthesis of (*rac*)-2,2-Dimethyl-1,3-dioxolane-4-methanol, 5-Hydroxy-2-phenyl-1,3-dioxane, Isopropylidene-2,2-bis(methoxy)propionic acid, Hydrophilic First-Generation Acid and Hydrophobic First-Generation Acid

Scheme 14. Synthesis of (*rac*)-2,2-Dimethyl-1,3-dioxolane-4-methanol, 5-Hydroxy-2-phenyl-1,3-dioxane, Isopropylidene-2,2-bis(methoxy)propionic acid, Hydrophilic First-Generation Acid and Hydrophobic First-Generation Acid. Reagents and conditions: (*i*) PTSA·H_2O, 4Å MS, Hexane/Acetone, Dean-Stark, reflux, 12 h; (*ii*) Benzaldehyde, H_2SO_4, Toluene, Dean-Stark, reflux, 6 h; (*iii*) PTSA·H_2O, 2,2-dimethoxypropane, Acetone, 23 °C, 3 h; (*iv*) TsCl, NaOH, THF-water, 0–5 °C, 3 h; (*v*) H_2SO_4 (cat.), MeOH, reflux, 2 h; (*vi*) K_2CO_3, DMF, 70 °C, 15 h; (*vii*) KOH, water, 100 °C, 2 h; (*viii*) 1-Bromododecane, K_2CO_3, DMF, 70 °C, 15 h; (*ix*) KOH, EtOH, reflux, 2 h.



(*rac*)-2,2-Dimethyl-1,3-dioxolane-4-methanol (**6**) was synthesized according to a literature

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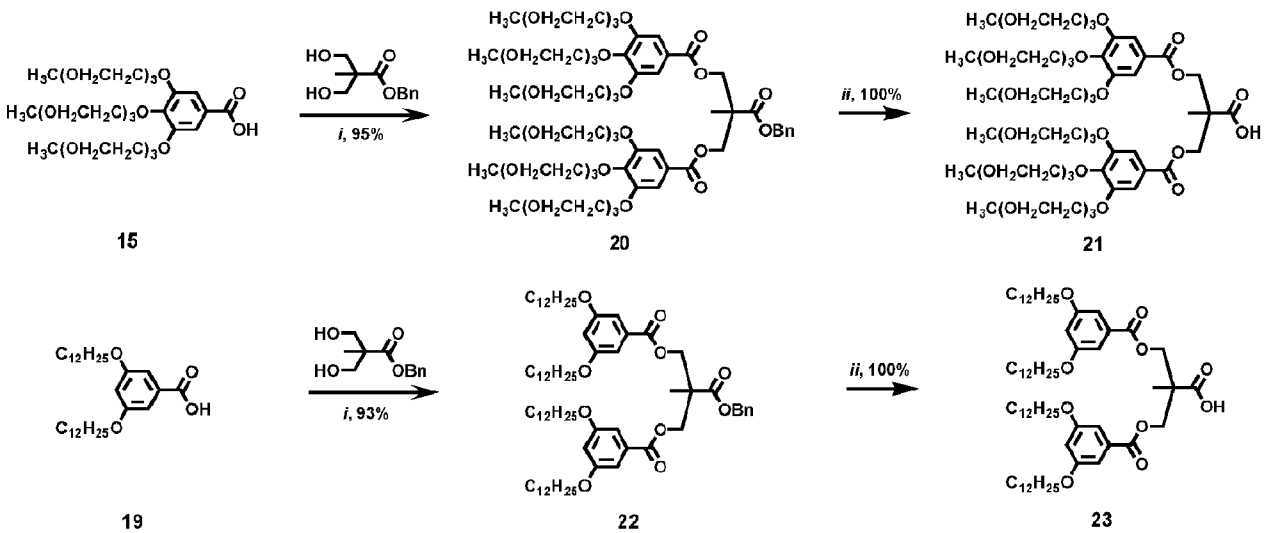
procedure (Xu et al., *Org. Lett.* **2017**, *19*, 3703–3706). 5-Hydroxy-2-phenyl-1,3-dioxane (**7**) was synthesized according to a literature procedure (Crich et al., *J. Am. Chem. Soc.* **1995**, *117*, 8757–8768). Isopropylidene-2,2-bis(methoxy)propionic acid (**9**) was synthesized according to a literature procedure (Ihre et al., *Macromolecules* **1998**, *31*, 4061–4068; Percec et al., *Science* **2010**, *328*, 1009–1014). Hydrophilic first-generation acid (3,4,5)-tris(methyl triethylene glycol)benzoic acid (**(3,4,5)-3EO-G1-COOH**, **15**) and hydrophobic first-generation acid 3,5-bis(dodecyloxy)benzoic acid (**(3,5)-12G1-COOH**, **19**) were prepared according to literature procedures (Percec et al., *Science* **2010**, *328*, 1009–1014; Percec et al., *J. Am. Chem. Soc.* **2013**, *135*, 4129–4148).

Synthesis of Hydrophilic and Hydrophobic Second-Generation Acids

Scheme 15. Synthesis of Hydrophilic and Hydrophobic Second-Generation Acids. Reagents and conditions: (*i*) DCC, DPTS, DCM, 23 °C, 12 h; (*ii*) H_2, Pd/C, EtOAc, 23 °C, 8 h.



The synthesis of compounds **20**, **21**, **22** and **23** was adapted from literature procedures (Zhang et al., *J. Am. Chem. Soc.* **2015**, *137*, 13334–13344).

Benzyl 2,2-bis(3,4,5-tris{2-[2-(2-methoxyethoxy)ethoxy]ethoxy}benzonate)propionate (20). (3,4,5)-3EO-G1-COOH (**15**, 3.00 g, 4.93 mmol, 2.1 equiv), benzyl 2,2-bis(hydroxymethyl)propionate (0.53 g, 2.35 mmol, 1.0 equiv) and DPTS (1.54 g, 4.93 mmol, 2.1 equiv) were dissolved in 10 mL dry DCM. N,N'-Dicyclohexylcarbodiimide (DCC, 1.94 g, 9.40 mmol, 4.0 equiv) dissolved in 2 mL dry DCM was added in one portion to the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea,

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which was washed with DCM carefully. The solvent was then removed under vacuum and the crude product was dissolved in 10 mL ethyl acetate (EtOAc). The EtOAc solution was filtered through Celite and the filter cake was washed with EtOAc. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc, then EtOAc/MeOH = 20/1, 10/1 as the eluent to give the title compound as a light-yellow oil (3.15 g, 95%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.24–7.20 (m, 9 H, Ph*H*), 5.17 (s, 2 H, PhC*H*2OCO-), 4.53 (d, 4 H, -C((C*H*2O-)2)CH3), 4.21 (t, 4 H, PhOC*H*2CH2O-), 4.13 (t, 8 H, PhOC*H*2CH2O-), 3.84 (t, 8 H, PhOCH2C*H*2O-), 3.78 (t, 4 H, PhOCH2C*H*2O-), 3.73–3.71 (m, 12 H, PhOCH2CH2OC*H*2CH2-), 3.70–3.61 (m, 24 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2-), 3.52 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.35 (ds, 18 H, -OC*H*3), 1.42 (s, 3 H, C(CH2O-)2C*H*3). Purity by HPLC: 99+%. MALDI-TOF MS *m/z* of $[M + Na]^+$ calculated for $C_{68}H_{108}O_{30}Na$: 1428.6; Found: 1428.9.

2,2-Bis(3,4,5-tris{2-[2-(2-methoxyethoxy)ethoxy]ethoxy}benzonate)propionic acid ((3,4,5)-3EO-G1-BMPA-COOH, **21**). Compound **20** (1.40 g, 1.00 mmol) was dissolved in 30 mL EtOAc. Then Pd/C (70 mg, 5 wt%) was added and the flask that was evacuated and filled with hydrogen for three times. The mixture was stirred at 23 °C for 8 h under hydrogen atmosphere. The reaction mixture was filtered through Celite and the filter cake was washed with EtOAc. Evaporation of the solvent yielded the title compound as a light-yellow oil (1.31 g, 100%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.30 (s, 4 H, Ph*H*), 4.56 (d, 2 H, 0.5 × -C((C*H*2O-)2)CH3), 4.46 (d, 2 H, 0.5 × -C((C*H*2O-)2)CH3), 4.21 (m, 12 H, PhOC*H*2CH2O-), 3.84 (t, 8 H, PhOCH2C*H*2O-), 3.78 (t, 4 H, PhOCH2C*H*2O-), 3.75–3.68 (m, 12 H, PhOCH2CH2OC*H*2CH2-), 3.68–3.60 (m, 24 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2-), 3.54 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.36 (ds, 18 H, -OC*H*3), 1.40 (s, 3 H, C(CH2O-)2C*H*3). Purity by HPLC: 99+%. MALDI-TOF MS *m/z* of $[M + Na]^+$ calculated for $C_{61}H_{102}O_{30}Na$: 1338.5; Found: 1338.9.

Benzyl 2,2-bis(3,5-didodecyloxybenzonate)propionate (22). (3,5)-12G1-COOH (**19**, 1.50 g, 3.06 mmol, 2.1 equiv), benzyl 2,2-bis(hydroxymethyl)propionate (0.33 g, 1.46 mmol, 1.0 equiv) and DPTS (0.96 g, 3.06 mmol, 2.1 equiv) were dissolved in 10 mL dry DCM. DCC (1.20 g, 5.84 mmol, 4.0 equiv) dissolved in 2 mL dry DCM was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The solvent was then removed under vacuum and the crude product was dissolved in 10 mL EtOAc. The EtOAc solution was filtered through Celite and

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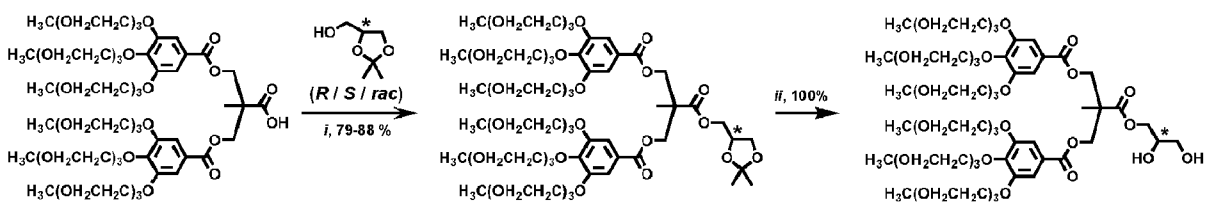
the filter cake was washed with EtOAc. The filtrate was concentrated and purified by column chromatography (SiO_2) with Hexane/EtOAc = 20/1 as the eluent to give the title compound as a light-yellow oil (1.59 g, 93%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.27–7.23 (m, 5 H, Ph*H*), 7.06 (d, 4 H, Ph*H*), 6.62 (t, 2 H, Ph*H*), 5.18 (s, 2 H, PhC*H*2OCO-), 4.56 (d, 4 H, -C(C*H*2O-)2CH3), 3.92 (t, 8 H, PhOC*H*$_2$(CH_2)$_{10}$$CH_3$), 1.77 (m, 8 H, PhOCH$_2$C*H*$_2$($CH_2$)$_9$$CH_3$), 1.44 (m, 8 H, PhOCH$_2CH_2$C*H*$_2$($CH_2$)$_8$$CH_3$), 1.44 (s, 3 H, -C(CH2O-)$_2$C*H*$_3$), 1.26 (br, 64 H, PhOCH$_2$CH$_2$CH$_2$(C*H*$_2$)$_8$$CH_3$), 0.87 (t, 12 H, PhO($CH_2$)$_{11}$C*H*$_3$). Purity by HPLC: 99+%. MALDI-TOF MS *m/z* of $[M + Na]^+$ calculated for $C_{74}H_{120}O_{10}Na$: 1192.8; Found: 1193.3.

2,2-Bis(3,5-didodecyloxybenzonate)propionic acid ((3,5)-12G1-BMPA-COOH, **23**). Compound **22** (1.50 g, 1.30 mmol) was dissolved in 30 mL EtOAc. Then Pd/C (75 mg, 5 wt%) was added and the flask was evacuated and filled three times with hydrogen. The mixture was stirred at 23 °C for 8 h under hydrogen atmosphere. The reaction mixture was filtered through Celite and the filter cake was washed with EtOAc. Evaporation of the solvent yielded the title compound as a colorless oil (1.40 g, 100%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.09 (d, 4 H, Ph*H*), 6.61 (t, 2 H, Ph*H*), 4.56 (m, 4 H, -C((C*H*2O-)2)CH3), 3.93 (t, 8 H, PhOC*H*$_2$(CH_2)$_{10}$$CH_3$), 1.76 (m, 8 H, PhOCH$_2$C*H*$_2$($CH_2$)$_9$$CH_3$), 1.45 (s, 3 H, -C(CH2O-)$_2$C*H*$_3$), 1.43 (m, 8 H, PhOCH$_2$CH$_2$C*H*$_2$(CH_2)$_8$$CH_3$), 1.26 (br, 64 H, PhOCH$_2CH_2CH_2$(C*H*$_2$)$_8$$CH_3$), 0.88 (t, 12 H, PhO($CH_2$)$_{11}$C*H*$_3$). Purity by HPLC: 99+%. MALDI-TOF MS *m/z* of $[M + Na]^+$ calculated for $C_{67}H_{114}O_{10}Na$: 1102.6; Found: 1102.9.

Synthesis of Hydrophilic Homochiral/Racemic Minidendron

Scheme 16. Synthesis of Hydrophilic Homochiral/Racemic Minidendron. Reagents and conditions: (*i*) DCC, DPTS, DCM, 23 °C, 12 h; (*ii*) 1 M HCl, MeOH, 23 °C, 1 h.



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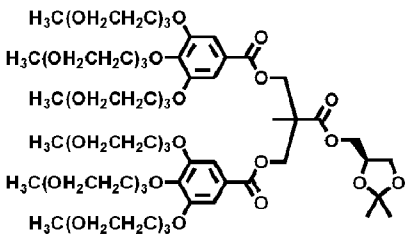
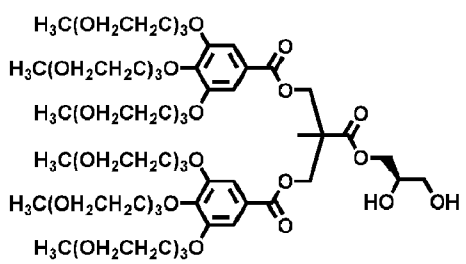


24*R*

(3,4,5)-3EO-G1-BMPA-GC-(*R*)-(1,2)acetonide (24*R*). (3,4,5)-3EO-G1-BMPA-COOH (**21**, 0.40 g, 0.30 mmol, 1.0 equiv), (*S*)-2,2-dimethyl-1,3-dioxolane-4-methanol (0.044 g, 0.33 mmol, 1.1 equiv) and DPTS (0.103 g, 0.33 mmol, 1.1 equiv) were dissolved in 5 mL dry DCM. DCC (0.093 g, 0.45 mmol, 1.5 equiv) dissolved in 2 mL dry DCM was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc, then DCM/MeOH = 20/1 as the eluent to give the title compound as a colorless oil (0.38 g, 88%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.23 (s, 4 H, Ph*H*), 4.53 (m, 4 H, -C((C*H*2O-)2)CH3), 4.26 (m, 1 H, C*H*(O-)(CH2O-)2), 4.23 (q, 1 H, 0.5 × (-COO-C*H*2O-CH(O-)-)), 4.21 (t, 4 H, PhOC*H*2CH2O-), 4.19 (t, 8 H, PhOC*H*2CH2O-), 4.15 (q, 1 H, 0.5 × (-COO-C*H*2O-CH(O-)-)), 3.99 (q, 1 H, 0.5 × (-CH(O-)-C*H*2O-)), 3.86 (t, 8 H, PhOCH2C*H*2O-), 3.78 (t, 4 H, PhOCH2C*H*2O-), 3.75–3.70 (m, 12 H, PhOCH2CH2OC*H*2CH2-), 3.70 (q, 1 H, 0.5 × (-CH(O-)-C*H*2O-)), 3.67–3.61 (m, 24 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2-), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.36 (ds, 18 H, -OC*H*3), 1.42 (s, 3 H, -C(CH2O-)2C*H*3), 1.36 (s, 3 H, C(O-)2(CH3)C*H*3), 1.26 (s, 3 H, C(O-)2(C*H*3)CH3). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.2, 165.1, 152.2, 142.8, 124.1, 109.5, 108.9, 73.0, 72.2, 71.7, 70.6, 70.5, 70.3, 70.3, 69.4, 68.7, 65.9, 65.6, 65.0, 58.8, 46.8, 26.5, 24.9, 17.8.



25*R*

(3,4,5)-3EO-G1-BMPA-GC-(*R*)-(1,2)(OH)2 (25*R*). (3,4,5)-3EO-G1-BMPA-GC-(*R*)-(1,2)acetonide (**24*R***, 0.22 g, 0.15 mmol) was dissolved in 10 mL MeOH. Then HCl (1 M, 2.0 mL, 2.0 mmol) was added. The mixture was stirred at 23 °C for 1 h. The reaction mixture was extracted by DCM (3 × 20 mL) and the organic phase was dried over

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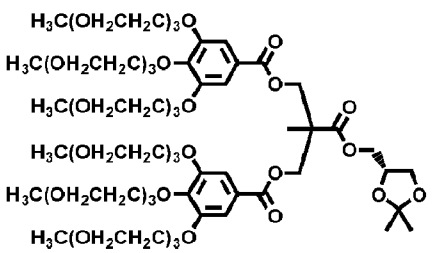
anhydrous magnesium sulfate. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.21 g, 100%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.27 (d, 4 H, Ph*H*), 4.58 (q, 2 H, -C((C*H*2O-)2)CH3), 4.48 (q, 2 H, -C((C*H*2O-)2)CH3), 4.29 (q, 1 H, 0.5 × (-COO-C*H*2O-CH(O-)-)), 4.20 (m, 13 H, 0.5 × (-COO-C*H*2O-CH(O-)-) and PhOC*H*2CH2O-), 3.91 (m, 1 H, C*H*(O-)(CH2O-)2), 3.86 (t, 8 H, PhOCH2C*H*2O-), 3.78 (t, 4 H, PhOCH2C*H*2O-), 3.75–3.69 (m, 12 H, PhOCH2CH2OC*H*2CH2-), 3.70 (q, 1 H, 0.5 × (-CH(O-)-C*H*2O-)), 3.68–3.60 (m, 25 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2- and 0.5 × (-CH(O-)-C*H*2O-)), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.36 (ds, 18 H, -OC*H*3), 1.41 (s, 3 H, -C(CH2O-)2C*H*3). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.9, 165.4, 152.3, 143.0, 124.2, 109.3, 72.4, 71.8, 71.8, 70.7, 70.7, 70.6, 70.6, 70.5, 70.4, 69.9, 69.5, 68.8, 66.1, 63.3, 58.9, 47.1, 18.0.



24*S*

(3,4,5)-3EO-G1-BMPA-GC-(*S*)-(1,2)acetonide (24*S*).

Compound 24*S* was synthesized from (3,4,5)-3EO-G1-BMPA-COOH (**21**, 0.40 g, 0.30 mmol, 1.0 equiv) and (*R*)-2,2-dimethyl-1,3-dioxolane-4-methanol (0.044 g, 0.33 mmol, 1.1 equiv) following a procedure similar to that used for the synthesis of **24*R***. The title compound was obtained as a colorless oil (0.37 g, 86%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.23 (s, 4 H, Ph*H*), 4.53 (m, 4 H, -C((C*H*2O-)2)CH3), 4.26 (m, 1 H, C*H*(O-)(CH2O-)2), 4.24 (q, 1 H, 0.5 × (-COO-C*H*2O-CH(O-)-)), 4.21 (t, 4 H, PhOC*H*2CH2O-), 4.17 (t, 8 H, PhOC*H*2CH2O-), 4.15 (q, 1 H, 0.5 × (-COO-C*H*2O-CH(O-)-)), 3.99 (q, 1 H, 0.5 × (-CH(O-)-C*H*2O-)), 3.86 (t, 8 H, PhOCH2C*H*2O-), 3.78 (t, 4 H, PhOCH2C*H*2O-), 3.74–3.70 (m, 12 H, PhOCH2CH2OC*H*2CH2-), 3.70 (q, 1 H, 0.5 × (-CH(O-)-C*H*2O-)), 3.67–3.61 (m, 24 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2-), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.36 (ds, 18 H, -OC*H*3), 1.42 (s, 3 H, -C(CH2O-)2C*H*3), 1.36 (s, 3 H, C(O-)2(CH3)C*H*3), 1.27 (s, 3 H, C(O-)2(C*H*3)CH3). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.3, 165.2, 152.2, 142.8, 124.1, 109.5, 108.9, 73.0, 72.2, 71.7, 70.6, 70.5, 70.3, 70.3, 69.4, 68.7, 65.9, 65.6, 65.0, 58.8, 46.8, 26.5, 24.9, 17.8.

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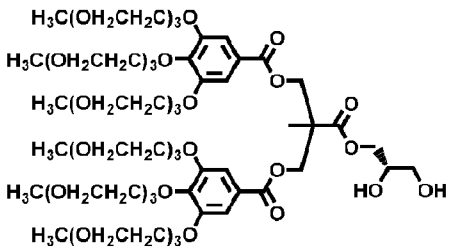
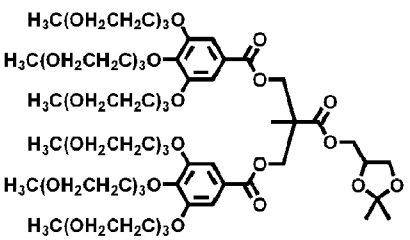


25S

(3,4,5)-3EO-G1-BMPA-GC-(*S*)-(1,2)(OH)$_2$ (25*S*).

Compound 25*S* was synthesized from (3,4,5)-3EO-G1-BMPA-GC-(*S*)-(1,2)acetonide (**24*S***, 0.36 g, 0.25 mmol) following a procedure similar to that used for the synthesis of **25*R***. The title compound was obtained as a colorless oil (0.35 g, 100%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.26 (d, 4 H, Ph*H*), 4.56 (q, 2 H, -C((C*H*$_2$O-)$_2$)CH3), 4.48 (q, 2 H, -C((C*H*$_2$O-)$_2$)CH3), 4.27 (q, 1 H, 0.5 × (-COO-C*H*$_2$O-CH(O-)-)), 4.20 (m, 13 H, 0.5 × (-COO-C*H*$_2$O-CH(O-)- and PhOC*H*$_2$CH$_2$O-), 3.90 (m, 1 H, C*H*(O-)(CH$_2$O-)$_2$), 3.85 (t, 8 H, PhOCH$_2$C*H*$_2$O-), 3.78 (t, 4 H, PhOCH$_2$C*H*$_2$O-), 3.75–3.69 (m, 12 H, PhOCH$_2$CH$_2$OC*H*$_2$CH$_2$-), 3.69 (q, 1 H, 0.5 × (-CH(O-)-C*H*$_2$O-)), 3.67–3.60 (m, 25 H, PhOCH$_2$CH$_2$OCH$_2$C*H*$_2$OC*H*$_2$CH$_2$- and 0.5 × (-CH(O-)-C*H*$_2$O-)), 3.53 (m, 12 H, PhOCH$_2$CH$_2$OCH$_2$CH$_2$OCH$_2$C*H*$_2$-), 3.36 (ds, 18 H, -OC*H*3), 1.40 (s, 3 H, -C(CH$_2$O-)$_2$C*H*$_3$). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.8, 165.3, 152.2, 142.9, 142.9, 124.2, 109.2, 109.1, 72.3, 71.7, 71.7, 70.6, 70.6, 70.5, 70.5, 70.4, 70.3, 70.3, 69.7, 69.5, 68.7, 66.0, 65.9, 63.2, 58.8, 47.0, 17.9.



24*rac*

(3,4,5)-3EO-G1-BMPA-GC-(*rac*)-(1,2)acetonide (24*rac*).

Compound 24*rac* was synthesized from (3,4,5)-3EO-G1-BMPA-COOH (**21**, 0.45 g, 0.34 mmol, 1.0 equiv) and (*rac*)-2,2-dimethyl-1,3-dioxolane-4-methanol (0.049 g, 0.37 mmol, 1.1 equiv) following a procedure similar to that used for the synthesis of **24*R***. The title compound was obtained as a colorless oil (0.38 g, 79%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.23 (s, 4 H, Ph*H*), 4.53 (m, 4 H, -C((C*H*$_2$O-)$_2$)CH3), 4.26 (m, 1 H, C*H*(O-)(CH$_2$O-)$_2$), 4.23 (q, 1 H, 0.5 × (-COO-C*H*$_2$O-CH(O-)-)), 4.21 (t, 4 H, PhOC*H*$_2$CH$_2$O-), 4.17 (t, 8 H, PhOC*H*$_2$CH$_2$O-), 4.15 (q, 1 H, 00.5 × (-COO-C*H*$_2$O-CH(O-)-)), 3.99 (q, 1 H, 0.5 × (-CH(O-)-C*H*$_2$O-)), 3.86 (t, 8 H, PhOCH$_2$C*H*$_2$O-), 3.78 (t, 4 H, PhOCH$_2$C*H*$_2$O-), 3.74–3.70 (m, 12 H, PhOCH$_2$CH$_2$OC*H*$_2$CH$_2$-), 3.70 (q, 1 H, 0.5 ×

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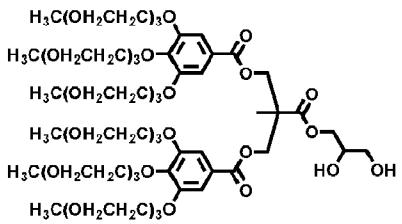
(-CH(O-)-C*H*2O-)), 3.67–3.61 (m, 24 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2-), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.36 (ds, 18 H, -OC*H*3), 1.42 (s, 3 H, -C(CH2O-)2C*H*3), 1.36 (s, 3 H, C(O-)2(CH3)C*H*3), 1.26 (s, 3 H, C(O-)2(C*H*3)CH3). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.3, 165.2, 152.3, 142.9, 124.2, 109.6, 109.0, 73.1, 72.3, 71.8, 70.7, 70.5, 70.4, 69.5, 68.8, 66.0, 65.7, 65.1, 58.9, 46.8, 26.6, 25.0, 17.9.



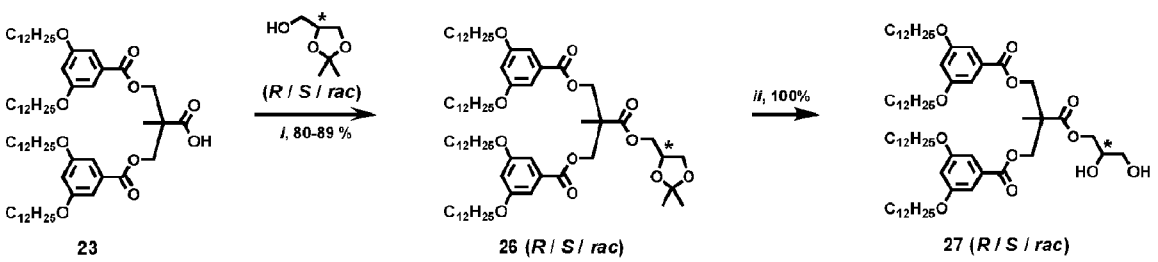
25*rac*

(3,4,5)-3EO-G1-BMPA-GC-(*rac*)-(1,2)(OH)2 (**25*rac***).

Compound **25*rac*** was synthesized **from (3,4,5)-3EO-G1-BMPA-GC-(*rac*)-(1,2)acetonide** (**24*rac***, 0.31 g, 0.22 mmol) following a procedure similar to that used for the synthesis of **25*R***. The title compound was obtained as a colorless oil (0.30 g, 100%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.27 (d, 4 H, Ph*H*), 4.57 (q, 2 H, -C((C*H*2O-)2)CH3), 4.48 (q, 2 H, -C((C*H*2O-)2)CH3), 4.27 (q, 1 H, 0.5 × (-COO-C*H*2O-CH(O-)-)), 4.20 (m, 13 H, 0.5 × (-COO-C*H*2O-CH(O-)-)) and PhOC*H*2CH2O-), 3.91 (m, 1 H, C*H*(O-)(CH2O-)2), 3.86 (t, 8 H, PhOCH2C*H*2O-), 3.78 (t, 4 H, PhOCH2C*H*2O-), 3.75–3.69 (m, 12 H, PhOCH2CH2OC*H*2CH2-), 3.70 (q, 1 H, 0.5 × (-CH(O-)-C*H*2O-)), 3.68–3.60 (m, 25 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2- and 0.5 × (-CH(O-)-C*H*2O-)), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.36 (ds, 18 H, -OC*H*3), 1.41 (s, 3 H, -C(CH2O-)2C*H*3). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.9, 165.4, 152.3, 142.9, 124.2, 109.3, 109.2, 72.3, 71.8, 71.8, 70.7, 70.7, 70.6, 70.5, 70.5, 70.4, 70.4, 70.4, 69.8, 69.5, 68.8, 66.1, 66.0, 63.2, 58.9, 47.0, 17.9.

Synthesis of Hydrophobic Homochiral/Racemic Minidendron

Scheme 17. Synthesis of Hydrophobic Homochiral/Racemic Minidendron. Reagents and conditions: (*i*) DCC, DPTS, DCM, 23 °C, 12 h; (*ii*) 1 M HCl, 1,4-dioxane, 60 °C, 8 h.



23 **26 (*R* / *S* / *rac*)** **27 (*R* / *S* / *rac*)**

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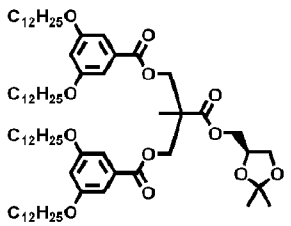
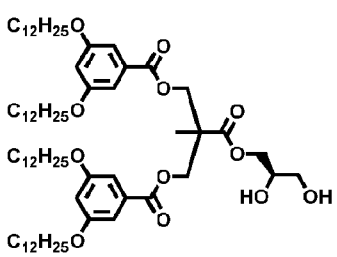


26*R*

(3,5)-12G1-BMPA-GC-(*R*)-(1,2)acetonide (26*R*). (3,5)-12G1-BMPA-COOH (**23**, 0.40 g, 0.37 mmol, 1.0 equiv), (*S*)-2,2-dimethyl-1,3-dioxolane-4-methanol (0.054 g, 0.41 mmol, 1.1 equiv) and DPTS (0.13 g, 0.41 mmol, 1.1 equiv) were dissolved in 5 mL dry DCM. DCC (0.12 g, 0.56 mmol, 1.5 equiv) dissolved in 2 mL of dry DCM was added in one portion to the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with Hexane/EtOAc = 20/1 as the eluent to give the title compound as a colorless oil (0.38 g, 86%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.10 (d, 4 H, Ph*H*), 6.62 (t, 2 H, Ph*H*), 4.56 (m, 4 H, -C((C*H*2O-)2)CH3), 4.27 (m, 1 H, C*H*(O-)(CH2O-)2), 4.23 (q, 1 H, 0.5 × (-COO-C*H*2O-CH(O-)-)), 4.16 (q, 1 H, 0.5 × (-COO-C*H*2O-CH(O-)-)), 3.99 (q, 1 H, 0.5 × (-CH(O-)-C*H*2O-)), 3.94 (t, 8 H, PhOC*H*$_2$(CH$_2$)$_{10}$CH$_3$), 3.71 (q, 1 H, 0.5 × (-CH(O-)-C*H*2O-)), 1.77 (m, 8 H, PhOCH$_2$C*H*$_2$(CH$_2$)$_9$CH$_3$), 1.44 (m, 8 H, PhOCH$_2$CH$_2$C*H*$_2$(CH$_2$)$_8$CH$_3$), 1.43 (s, 3 H, -C(CH2O-)$_2$C*H*$_3$), 1.38 (s, 3 H, C(O-)$_2$(CH$_3$)C*H*$_3$), 1.27 (s, 3 H, C(O-)$_2$(C*H*$_3$)CH$_3$), 1.26 (br, 64 H, PhOCH$_2$CH$_2$CH$_2$(C*H*$_2$)$_8$CH$_3$), 0.88 (t, 12 H, PhO(CH$_2$)$_{11}$C*H*$_3$). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.6, 165.9, 160.3, 131.3, 131.3, 109.8, 107.8, 106.6, 73.3, 68.4, 66.4, 66.1, 66.1, 65.4, 47.0, 32.0, 29.8, 29.7, 29.7, 29.7, 29.5, 29.5, 29.3, 26.8, 26.1, 25.2, 22.8, 18.1, 14.2.



27*R*

(3,5)-12G1-BMPA-GC-(*R*)-(1,2)(OH)$_2$ (27*R*). (3,5)-12G1-BMPA-GC-(*R*)-(1,2)acetonide (**26*R***, 0.29 g, 0.24 mmol) was dissolved in 10 mL 1,4-dioxane. Then HCl (2 M, 2.0 mL, 4.0 mmol) was added. The mixture was stirred at 60 °C for 8 h. The reaction mixture was extracted by DCM (3 × 20 mL) and the organic phase was dried over anhydrous magnesium sulfate. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.28 g, 100%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.11 (d, 4 H, Ph*H*), 6.63 (t, 2 H, Ph*H*), 4.58 (m, 4 H, -

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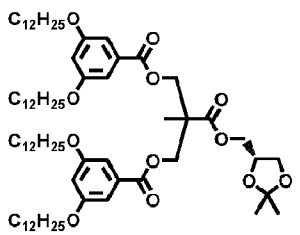
C((C*H*2O-)2)CH3), 4.27 (q, 1 H, 0.5 × (-COO-C*H*2O-CH(O-)-)), 4.21 (q, 1 H, 0.5 × (-COO-C*H*2O-CH(O-)-)), 3.94 (t, 8 H, PhOC*H*2(CH2)10CH3), 3.92 (m, 1 H, C*H*(O-)(CH2O-)2), 3.65 (q, 1 H, 0.5 × (-CH(O-)-C*H*2O-)), 3.60 (q, 1 H, 0.5 × (-CH(O-)-C*H*2O-)), 2.74 (s, 1 H, -CH(O*H*)(CH2O-)2), 2.17 (s, 1 H, -CH2O*H*), 1.77 (m, 8 H, PhOCH2C*H*2(CH2)9CH3), 1.44 (m, 8 H, PhOCH2CH2C*H*2(CH2)8CH3), 1.44 (s, 3 H, -C(CH2O-)2C*H*3), 1.26 (br, 64 H, PhOCH2CH2CH2(C*H*2)8CH3), 0.88 (t, 12 H, PhO(CH2)11C*H*3). ^{13}C NMR (500 MHz, CDCl3) δ 173.0, 166.0, 166.0, 160.2, 131.2, 131.2, 107.8, 106.6, 69.9, 68.4, 66.1, 63.2, 47.1, 32.0, 29.7, 29.7, 29.7, 29.7, 29.5, 29.4, 29.2, 26.1, 22.8, 18.1, 14.2.



26S

(3,5)-12G1-BMPA-GC-(*S*)-(1,2)acetonide (**26*S***). Compound **26*S*** was synthesized from (3,5)-12G1-BMPA-COOH (**23**, 0.40 g, 0.37 mmol, 1.0 equiv) and (*R*)-2,2-dimethyl-1,3-dioxolane-4-methanol (0.054 g, 0.41 mmol, 1.1 equiv) following a procedure similar to that used for the synthesis of **26*R***. The title compound was obtained as a colorless oil (0.39 g, 89%). ^{1}H NMR (500 MHz, CDCl3) δ 7.10 (d, 4 H, Ph*H*), 6.62 (t, 2 H, Ph*H*), 4.56 (m, 4 H, -C((C*H*2O-)2)CH3), 4.27 (m, 1 H, C*H*(O-)(CH2O-)2), 4.23 (q, 1 H, 0.5 × (-COO-C*H*2O-CH(O-)-)), 4.16 (q, 1 H, 0.5 × (-COO-C*H*2O-CH(O-)-)), 3.99 (q, 1 H, 0.5 × (-CH(O-)-C*H*2O-)), 3.94 (t, 8 H, PhOC*H*2(CH2)10CH3), 3.72 (q, 1 H, 0.5 × (-CH(O-)-C*H*2O-)), 1.77 (m, 8 H, PhOCH2C*H*2(CH2)9CH3), 1.44 (m, 8 H, PhOCH2CH2C*H*2(CH2)8CH3), 1.43 (s, 3 H, -C(CH2O-)2C*H*3), 1.38 (s, 3 H, C(O-)2(CH3)C*H*3), 1.27 (s, 3 H, C(O-)2(C*H*3)CH3), 1.26 (br, 64 H, PhOCH2CH2CH2(C*H*2)8CH3), 0.88 (t, 12 H, PhO(CH2)11C*H*3). ^{13}C NMR (500 MHz, CDCl3) δ 172.6, 165.9, 160.3, 131.4, 131.3, 109.8, 107.8, 106.6, 73.3, 68.4, 66.4, 66.2, 66.1, 65.4, 47.0, 32.0, 29.8, 29.8, 29.7, 29.7, 29.5, 29.5, 29.3, 26.8, 26.1, 25.2, 22.8, 18.1, 14.2.

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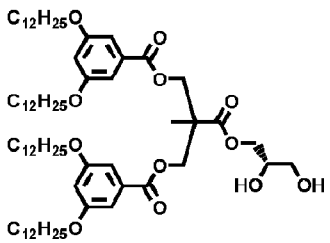
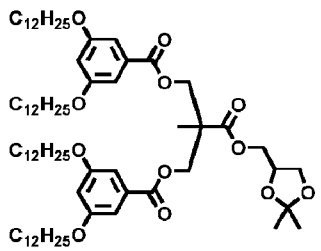


27*S*

(3,5)-12G1-BMPA-GC-(*S*)-(1,2)(OH)$_2$ (**27*S***). Compound **27*S*** was synthesized from (3,5)-12G1-BMPA-GC-(*S*)-(1,2)acetonide (**26*S***, 0.31 g, 0.26 mmol) following a procedure similar to that used for the synthesis of **27*R***. The title compound was obtained as a colorless oil (0.30 g, 100%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.11 (d, 4 H, Ph*H*), 6.64 (t, 2 H, Ph*H*), 4.58 (m, 4 H, -C((C*H*$_2$O-)$_2$)CH3), 4.27 (q, 1 H, 0.5 × (-COO-C*H*$_2$O-CH(O-)-)), 4.21 (q, 1 H, 0.5 × (-COO-C*H*$_2$O-CH(O-)-)), 3.94 (t, 8 H, PhOC*H*$_2$(CH$_2$)$_{10}$CH$_3$), 3.91 (m, 1 H, C*H*(O-)(CH$_2$O-)$_2$), 3.65 (q, 1 H, 0.5 × (-CH(O-)-C*H*$_2$O-)), 3.59 (q, 1 H, 0.5 × (-CH(O-)-C*H*$_2$O-)), 2.73 (s, 1 H, -CH(O*H*)(CH$_2$O-)$_2$), 2.16 (s, 1 H, -CH$_2$O*H*), 1.77 (m, 8 H, PhOCH$_2$C*H*$_2$(CH$_2$)$_9$CH$_3$), 1.44 (m, 8 H, PhOCH$_2$CH$_2$C*H*$_2$(CH$_2$)$_8$CH$_3$), 1.44 (s, 3 H, -C(CH$_2$O-)$_2$C*H*$_3$), 1.26 (br, 64 H, PhOCH$_2$CH$_2$CH$_2$(C*H*$_2$)$_8$CH$_3$), 0.88 (t, 12 H, PhO(CH$_2$)$_{11}$C*H*$_3$). ^{13}C NMR (500 MHz, $CDCl_3$) δ 173.0, 166.1, 166.1, 160.3, 131.2, 131.2, 107.8, 106.6, 70.0, 68.4, 66.2, 63.2, 47.2, 32.0, 29.8, 29.7, 29.7, 29.7, 29.5, 29.5, 29.3, 26.1, 22.8, 18.1, 14.2.



26*rac*

(3,5)-12G1-BMPA-GC-(*rac*)-(1,2)acetonide (**26*rac***). Compound **26*rac*** was synthesized from (3,5)-12G1-BMPA-COOH (**23**, 0.53 g, 0.49 mmol, 1.0 equiv) and (*rac*)-2,2-dimethyl-1,3-dioxolane-4-methanol (0.071 g, 0.54 mmol, 1.1 equiv) following a procedure similar to that used for the synthesis of **26*R***. The title compound was obtained as a colorless oil (0.46 g, 80%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.10 (d, 4 H, Ph*H*), 6.62 (t, 2 H, Ph*H*), 4.56 (m, 4 H, -C((C*H*$_2$O-)$_2$)CH3), 4.27 (m, 1 H, C*H*(O-)(CH$_2$O-)$_2$), 4.23 (q, 1 H, 0.5 × (-COO-C*H*$_2$O-CH(O-)-)), 4.16 (q, 1 H, 0.5 × (-COO-C*H*$_2$O-CH(O-)-)), 3.99 (q, 1 H, 0.5 × (-CH(O-)-C*H*$_2$O-)), 3.94 (t, 8 H, PhOC*H*$_2$(CH$_2$)$_{10}$CH$_3$), 3.71 (q, 1 H, 0.5 × (-CH(O-)-C*H*$_2$O-)), 1.77 (m, 8 H, PhOCH$_2$C*H*$_2$(CH$_2$)$_9$CH$_3$), 1.44 (m, 8 H, PhOCH$_2$CH$_2$C*H*$_2$(CH$_2$)$_8$CH$_3$), 1.43 (s, 3 H, -C(CH$_2$O-

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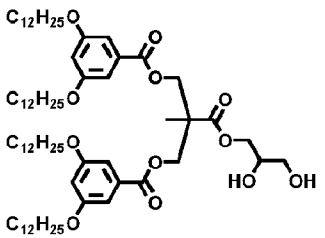
)$_2$C*H*$_3$), 1.38 (s, 3 H, C(O-)$_2$(CH$_3$)C*H*$_3$), 1.27 (s, 3 H, C(O-)$_2$(C*H*$_3$)CH$_3$), 1.26 (br, 64 H, PhOCH$_2$CH$_2$CH$_2$(C*H*$_2$)$_8$CH$_3$), 0.88 (t, 12 H, PhO(CH$_2$)$_{11}$C*H*$_3$). ^{13}C NMR (500 MHz, CDCl$_3$) δ 172.6, 165.9, 160.3, 131.4, 131.3, 109.8, 107.8, 106.6, 73.3, 68.4, 66.4, 66.2, 66.1, 65.4, 47.0, 32.0, 29.8, 29.8, 29.7, 29.7, 29.5, 29.5, 29.3, 26.8, 26.1, 25.2, 22.8, 18.1, 14.2.

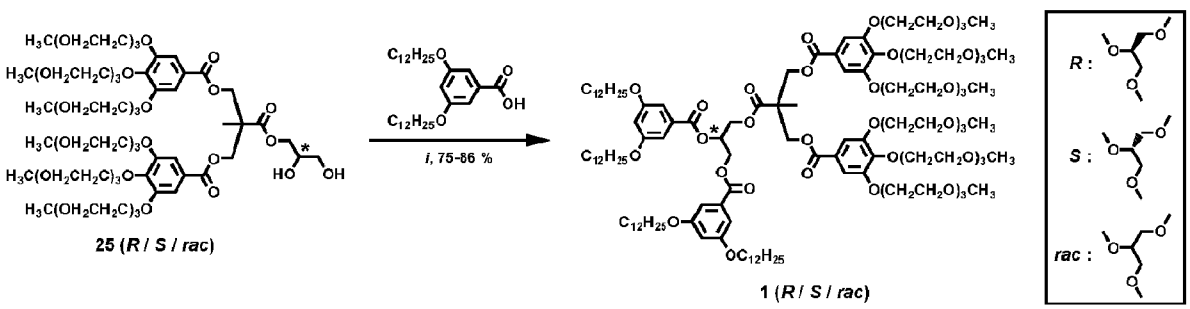


27*rac*

(3,5)-12G1-BMPA-GC-(*rac*)-(1,2)(OH)$_2$ (**27*rac***). Compound **27*rac*** was synthesized from (3,5)-12G1-BMPA-GC-(*rac*)-(1,2)acetonide (**26*rac***, 0.30 g, 0.25 mmol) following a procedure similar to that used for the synthesis of **27*R***. The title compound was obtained as a colorless oil (0.29 g, 100%). ^{1}H NMR (500 MHz, CDCl$_3$) δ 7.11 (d, 4 H, Ph*H*), 6.64 (t, 2 H, Ph*H*), 4.58 (m, 4 H, -C((C*H*$_2$O-)2)CH3), 4.27 (q, 1 H, 0.5 × (-COO-C*H*$_2$O-CH(O-)-)), 4.21 (q, 1 H, 0.5 × (-COO-C*H*$_2$O-CH(O-)-)), 3.95 (t, 8 H, PhOC*H*$_2$(CH$_2$)$_{10}$CH$_3$), 3.91 (m, 1 H, C*H*(O-)(CH2O-)2), 3.65 (q, 1 H, 0.5 × (-CH(O-)-C*H*$_2$O-)), 3.59 (q, 1 H, 0.5 × (-CH(O-)-C*H*$_2$O-)), 2.72 (s, 1 H, -CH(O*H*)(CH2O-)2), 2.15 (s, 1 H, -CH$_2$O*H*), 1.77 (m, 8 H, PhOCH$_2$C*H*$_2$(CH$_2$)$_9$CH$_3$), 1.44 (m, 8 H, PhOCH$_2$CH$_2$C*H*$_2$(CH$_2$)$_8$CH$_3$), 1.44 (s, 3 H, -C(CH2O-)$_2$C*H*$_3$), 1.26 (br, 64 H, PhOCH$_2$CH$_2$CH$_2$(C*H*$_2$)$_8$CH$_3$), 0.88 (t, 12 H, PhO(CH$_2$)$_{11}$C*H*$_3$). ^{13}C NMR (500 MHz, CDCl$_3$) δ 173.0, 166.1, 166.1, 160.3, 131.2, 131.2, 107.9, 106.6, 70.0, 68.4, 66.2, 63.2, 47.2, 32.0, 29.8, 29.7, 29.7, 29.7, 29.5, 29.5, 29.3, 26.1, 22.8, 18.1, 14.2.

Synthesis of Amphiphilic Glycerol-Based Janus Dendrimers from Library 1

Scheme 18. Synthesis of Amphiphilic Glycerol-Based Janus Dendrimers from Library 1. Reagents and conditions: (*i*) DCC, DPTS, DCM, 23 °C, 12 h.



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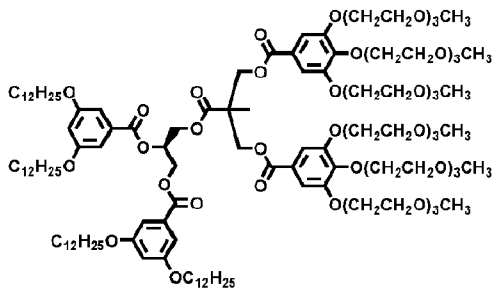

1*R*

(3,5)-12G1-GC-(*R*)-BMPA-(3,4,5)-3EO-G1 (1*R*). (3,4,5)-3EO-G1-BMPA-GC-(*R*)-(1,2)(OH)$_2$ (25*R*, 0.18 g, 0.13 mmol, 1.0 equiv), (3,5)-12G1-COOH (0.15 g, 0.30 mmol, 2.3 equiv) and DPTS (0.094 g, 0.30 mmol, 2.3 equiv) were dissolved in 5 mL dry DCM. DCC (0.080 g, 0.39 mmol, 3.0 equiv) dissolved in 2 mL dry DCM was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 20/1 as the eluent to give the title compound as a light-yellow oil (0.23 g, 86%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.20 (s, 4 H, PhH), 7.07 (t, 4 H, PhH), 6.60 (t, 1 H, PhH), 6.58 (t, 1 H, PhH), 5.66 (m, 1 H, CH(O-)(CH2O-)2), 4.63 (q, 1 H, 0.25 × -C((CH2O-)2)CH3), 4.58–4.42 (m, 7 H, 0.75 × -C((CH2O-)2)CH3, -COO-CH2O-CH(O-)- and -CH(O-)-CH2O-), 4.20 (t, 4 H, PhOCH2CH2O-), 4.15 (t, 8 H, PhOCH2CH2O-), 3.89 (m, 8 H, PhOCH2CH2O-), 3.84 (t, 8 H, PhOCH$_2$(CH$_2$)$_{10}$CH$_3$), 3.78 (t, 4 H, PhOCH2CH2O-), 3.74–3.69 (m, 12 H, PhOCH2CH2OCH2CH2-), 3.67–3.60 (m, 24 H, PhOCH2CH2OCH2CH2OCH2CH2-), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2CH2-), 3.36 (ds, 18 H, -OCH3), 1.73 (m, 8 H, PhOCH$_2$CH$_2$(CH$_2$)$_9$CH$_3$), 1.41 (m, 8 H, PhOCH$_2$CH$_2$CH$_2$(CH$_2$)$_8$CH$_3$), 1.38 (s, 3 H, -C(CH2O-)$_2$CH$_3$), 1.25 (br, 64 H, PhOCH$_2$CH$_2$CH$_2$(CH$_2$)$_8$CH$_3$), 0.87 (t, 12 H, PhO(CH$_2$)$_{11}$CH$_3$). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.3, 165.9, 165.5, 165.4, 160.2, 152.4, 143.0, 131.1, 131.0, 124.4, 124.3, 109.1, 107.8, 107.7, 106.9, 72.5, 72.0, 70.9, 70.7, 70.6, 70.6, 69.7, 69.0, 68.4, 65.6, 63.2, 62.8, 59.1, 47.0, 32.0, 29.8, 29.7, 29.7, 29.7, 29.5, 29.5, 29.4, 29.3, 29.3, 26.1, 26.1, 22.8, 18.0, 14.2. Purity by HPLC: 99+%. MALDI-TOF MS *m/z* of $[M + Na]^+$ calculated for $C_{126}H_{212}O_{38}Na$: 2358.0; Found: 2358.3. The ^{1}H NMR and ^{13}C NMR spectra of compound **1*R*** are shown in Figure 90 and Figure 91, respectively.

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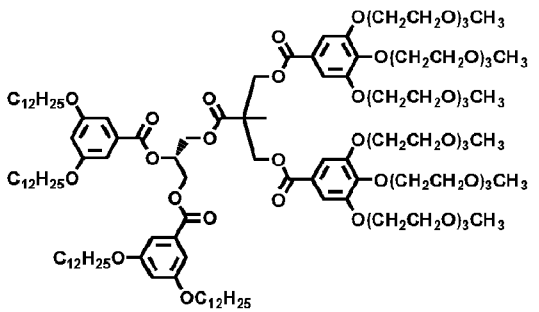

1S

(3,5)-12G1-GC-(*S*)-BMPA-(3,4,5)-3EO-G1 (1*S*).

Compound 1*S* was synthesized from (3,4,5)-3EO-G1-BMPA-GC-(*S*)-(1,2)(OH)$_2$ (25*S*, 0.20 g, 0.14 mmol, 1.0 equiv) and (3,5)-12G1-COOH (0.16 g, 0.32 mmol, 2.3 equiv) following a procedure similar to that used for the synthesis of **1*R***. The title compound was obtained as a light-yellow oil (0.26 g, 80%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.20 (s, 4 H, Ph*H*), 7.07 (t, 4 H, Ph*H*), 6.60 (t, 1 H, Ph*H*), 6.58 (t, 1 H, Ph*H*), 5.66 (m, 1 H, C*H*(O-)(CH2O-)2), 4.63 (q, 1 H, 0.25 × -C((C*H*2O-)2)CH3), 4.58–4.42 (m, 7 H, 0.75 × -C((C*H*2O-)2)CH3, -COO-C*H*2O-CH(O-)- and -CH(O-)-C*H*2O-), 4.20 (t, 4 H, PhOC*H*2CH2O-), 4.16 (t, 8 H, PhOC*H*2CH2O-), 3.89 (m, 8 H, PhOCH2C*H*2O-), 3.85 (t, 8 H, PhOC*H*$_2$(CH$_2$)$_{10}$CH$_3$), 3.78 (t, 4 H, PhOCH2C*H*2O-), 3.74–3.69 (m, 12 H, PhOCH2CH2OC*H*2CH2-), 3.67–3.60 (m, 24 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2-), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.36 (ds, 18 H, -OC*H*3), 1.73 (m, 8 H, PhOCH$_2$C*H*$_2$(CH$_2$)$_9$CH$_3$), 1.41 (m, 8 H, PhOCH$_2$CH$_2$C*H*$_2$(CH$_2$)$_8$CH$_3$), 1.38 (s, 3 H, -C(CH2O-)$_2$C*H*$_3$), 1.25 (br, 64 H, PhOCH$_2$CH$_2$CH$_2$(C*H*$_2$)$_8$CH$_3$), 0.87 (t, 12 H, PhO(CH$_2$)$_{11}$C*H*$_3$). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.2, 165.8, 165.5, 165.3, 160.2, 152.4, 142.9, 131.0, 130.9, 124.3, 124.3, 109.0, 107.8, 107.6, 106.8, 72.4, 71.9, 70.8, 70.7, 70.6, 69.6, 68.9, 68.3, 65.6, 63.1, 62.7, 59.0, 47.0, 31.9, 29.7, 29.7, 29.6, 29.6, 29.5, 29.4, 29.4, 29.2, 29.2, 26.1, 26.0, 22.7, 17.9, 14.1. Purity by HPLC: 99+%. MALDI-TOF MS *m/z* of $[M + Na]^+$ calculated for $C_{126}H_{212}O_{38}Na$: 2358.0; Found: 2358.4. The ^{1}H NMR and ^{13}C NMR spectra of compound **1*S*** are shown in Figure 92 and Figure 93, respectively.

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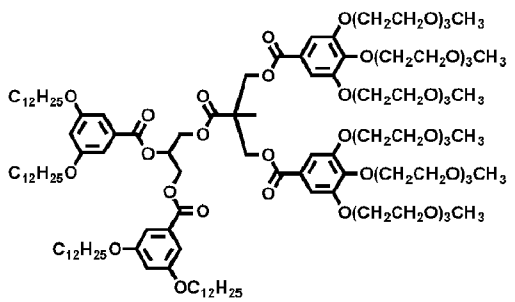

1*rac*

(3,5)-12G1-GC-(*rac*)-BMPA-(3,4,5)-3EO-G1 (1*rac*).

Compound 1*rac* was synthesized from (3,4,5)-3EO-G1-BMPA-GC-(*rac*)-(1,2)$(OH)_2$ (25*rac*, 0.28 g, 0.20 mmol, 1.0 equiv) and (3,5)-12G1-COOH (0.23 g, 0.46 mmol, 2.3 equiv) following a procedure similar to that used for the synthesis of **1*R***. The title compound was obtained as a light-yellow oil (0.35 g, 75%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.20 (s, 4 H, Ph*H*), 7.07 (t, 4 H, Ph*H*), 6.60 (t, 1 H, Ph*H*), 6.58 (t, 1 H, Ph*H*), 5.66 (m, 1 H, C*H*(O-)(CH2O-)2), 4.63 (q, 1 H, 0.25 × -C((C*H*2O-)2)CH3), 4.58–4.42 (m, 7 H, 0.75 × -C((C*H*2O-)2)CH3, -COO-C*H*2O-CH(O-)- and -CH(O-)-C*H*2O-), 4.20 (t, 4 H, PhOC*H*2CH2O-), 4.16 (t, 8 H, PhOC*H*2CH2O-), 3.89 (m, 8 H, PhOCH2C*H*2O-), 3.85 (t, 8 H, PhOC*H*2$(CH_2)_{10}CH_3$), 3.78 (t, 4 H, PhOCH2C*H*2O-), 3.74–3.69 (m, 12 H, PhOCH2CH2OC*H*2CH2-), 3.67–3.60 (m, 24 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2-), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.36 (ds, 18 H, -OC*H*3), 1.73 (m, 8 H, PhOCH2C*H*2$(CH_2)_9CH_3$), 1.41 (m, 8 H, PhOCH2CH2C*H*2$(CH_2)_8CH_3$), 1.38 (s, 3 H, -C(CH2O-)2C*H*3), 1.25 (br, 64 H, PhOCH2CH2CH2(C*H*2)8CH3), 0.87 (t, 12 H, PhO$(CH_2)_{11}CH_3$). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.2, 165.9, 165.5, 165.4, 160.2, 152.4, 143.0, 131.1, 131.0, 124.3, 124.3, 109.1, 107.8, 107.7, 106.9, 72.5, 72.0, 70.9, 70.7, 70.6, 70.6, 69.7, 68.9, 68.3, 65.6, 63.2, 62.8, 59.1, 47.0, 32.0, 29.8, 29.7, 29.7, 29.7, 29.5, 29.5, 29.4, 29.3, 29.3, 26.1, 26.1, 22.8, 18.0, 14.2. Purity by HPLC: 99+%. MALDI-TOF MS *m/z* of $[M + Na]^+$ calculated for $C_{126}H_{212}O_{38}Na$: 2358.0; Found: 2358.1. The ^{1}H NMR and ^{13}C NMR spectra of compound ***1rac*** are shown in Figure 94 and Figure 95, respectively.

Synthesis of Amphiphilic Glycerol-Based Janus Dendrimer from Library 2

Scheme 19. Synthesis of Amphiphilic Glycerol-Based Janus Dendrimers from Library 2. Reagents and conditions: (*i*) DCC, DPTS, DCM, 23 °C, 12 h.

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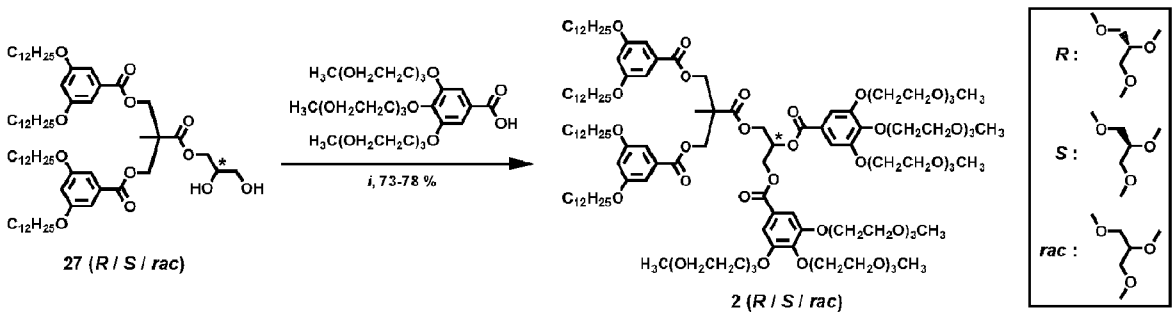

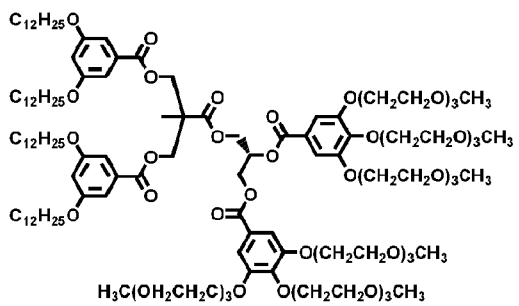

(3,5)-12G1-BMPA-GC-(*R*)-(3,4,5)-3EO-G1 (2*R*). (3,5)-12G1-BMPA-GC-(*R*)-(1,2)(OH)$_2$ (27*R*, 0.28 g, 0.24 mmol, 1.0 equiv), (3,4,5)-3EO-G1-COOH (0.33 g, 0.55 mmol, 2.3 equiv) and DPTS (0.17 g, 0.55 mmol, 2.3 equiv) were dissolved in 6 mL dry DCM. DCC (0.15 g, 0.72 mmol, 3.0 equiv) dissolved in 2 mL dry DCM was added in one portion into the above mixture. The reaction was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 20/1 as the eluent to give the title compound as a light-yellow oil (0.41 g, 73%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.22 (d, 4 H, Ph*H*), 7.05 (d, 2 H, Ph*H*), 7.05 (d, 2 H, Ph*H*), 6.58 (t, 2 H, Ph*H*), 5.63 (m, 1 H, C*H*(O-)(CH2O-)2), 4.62–4.42 (m, 8 H, -C((C*H*2O-)2)CH3, -COO-C*H*2O-CH(O-)- and -CH(O-)-C*H*2O-), 4.20 (m, 4 H, PhOC*H*2CH2O-), 4.13 (m, 8 H, PhOC*H*2CH2O-), 3.91 (t, 8 H, PhOCH2C*H*2O-), 3.83 (m, 8 H, PhOC*H*$_2$(CH$_2$)$_{10}$CH$_3$), 3.78 (t, 4 H, PhOCH2C*H*2O-), 3.74–3.68 (m, 12 H, PhOCH2CH2OC*H*2CH2-), 3.67–3.60 (m, 24 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2-), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.38-3.33 (m, 18 H, -OC*H*3), 1.74 (m, 8 H, PhOCH$_2$C*H*$_2$(CH$_2$)$_9$CH$_3$), 1.42 (m, 8 H, PhOCH$_2$CH$_2$C*H*$_2$(CH$_2$)$_8$CH$_3$), 1.39 (s, 3 H, -C(CH2O-)$_2$C*H*$_3$), 1.25 (br, 64 H, PhOCH$_2$CH$_2$CH$_2$(C*H*$_2$)$_8$CH$_3$), 0.87 (t, 12 H, PhO(CH$_2$)$_{11}$C*H*$_3$). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.2, 165.7, 165.4, 165.0, 160.1, 152.3, 142.9, 142.9, 131.1, 131.1, 124.1, 123.9, 109.0, 109.0, 107.6, 106.4, 72.4, 71.9, 71.9, 70.8, 70.8, 70.7, 70.6, 70.5, 70.5, 70.5, 69.7, 69.6, 68.8, 68.3, 67.0, 65.7, 63.1, 62.4, 59.0, 46.9, 31.9, 29.7, 29.6, 29.6, 29.6, 29.4, 29.3, 29.2,

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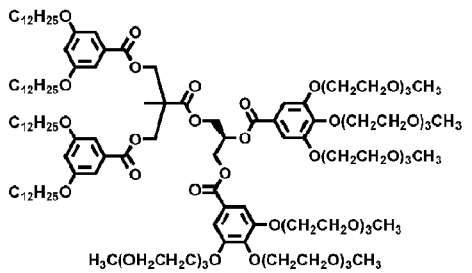
26.0, 22.7, 18.1, 14.1. Purity by HPLC: 99+%. MALDI-TOF MS *m/z* of $[M + Na]^+$ calculated for $C_{126}H_{212}O_{38}Na$: 2358.0; Found: 2358.4. The ^1H NMR and ^{13}C NMR spectra of compound **2*R*** are shown in Figure 96 and Figure 97, respectively.



2S

(3,5)-12G1-BMPA-GC-(*S*)-(3,4,5)-3EO-G1 (2*S*). Compound 2*S* was synthesized from (3,5)-12G1-BMPA-GC-(*S*)-(1,2)$(OH)_2$ (27*S*, 0.24 g, 0.21 mmol, 1.0 equiv) and (3,4,5)-3EO-G1-COOH (0.29 g, 0.48 mmol, 2.3 equiv) following a procedure similar to that used for the synthesis of **2*R***. The title compound was obtained as a light-yellow oil (0.37 g, 76%). ^1H NMR (500 MHz, $CDCl_3$) δ 7.22 (d, 4 H, Ph*H*), 7.05 (d, 2 H, Ph*H*), 7.05 (d, 2 H, Ph*H*), 6.58 (t, 2 H, Ph*H*), 5.63 (m, 1 H, C*H*(O-)$(CH_2O-)_2$), 4.63–4.42 (m, 8 H, -C((C*H*2O-)2)CH3, -COO-C*H*2O-CH(O-)- and -CH(O-)-C*H*2O-), 4.20 (m, 4 H, PhOC*H*2CH2O-), 4.13 (m, 8 H, PhOC*H*2CH2O-), 3.91 (t, 8 H, PhOCH2C*H*2O-), 3.83 (m, 8 H, PhOC*H*$_2(CH_2)_{10}CH_3$), 3.78 (t, 4 H, PhOCH2C*H*2O-), 3.74–3.68 (m, 12 H, PhOCH2CH2OC*H*2CH2-), 3.67–3.60 (m, 24 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2-), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.37-3.33 (m, 18 H, -OC*H*3), 1.74 (m, 8 H, PhOCH$_2$C*H*$_2(CH_2)_9CH_3$), 1.42 (m, 8 H, PhOCH$_2$CH$_2$C*H*$_2(CH_2)_8CH_3$), 1.39 (s, 3 H, -C(CH2O-)$_2$C*H*$_3$), 1.25 (br, 64 H, PhOCH$_2$CH$_2$CH$_2$(C*H*$_2$)$_8$CH$_3$), 0.87 (t, 12 H, PhO$(CH_2)_{11}$C*H*$_3$). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.3, 165.8, 165.5, 165.1, 160.2, 152.4, 143.0, 142.9, 131.2, 131.2, 124.2, 124.0, 109.1, 109.0, 107.7, 106.5, 72.5, 72.0, 72.0, 70.8, 70.7, 70.7, 70.6, 70.6, 70.6, 69.8, 69.6, 68.9, 68.3, 67.1, 65.7, 63.2, 62.5, 59.1, 46.9, 32.0, 29.7, 29.7, 29.7, 29.6, 29.5, 29.4, 29.2, 26.1, 22.7, 18.1, 14.2. Purity by HPLC: 99+%. MALDI-TOF MS *m/z* of $[M + Na]^+$ calculated for $C_{126}H_{212}O_{38}Na$: 2358.0; Found: 2358.3. The ^1H NMR and ^{13}C NMR spectra of compound **2*S*** are shown in Figure 98 and Figure 99, respectively.

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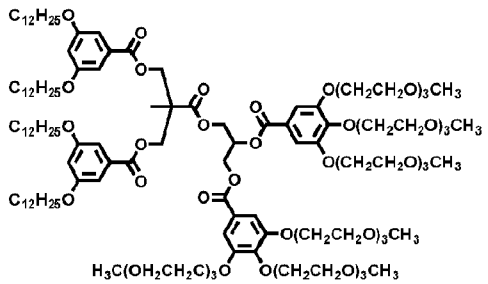

2*rac*

(3,5)-12G1-BMPA-GC-(*rac*)-(3,4,5)-3EO-G1 (2*rac*).

Compound 2*rac* was synthesized from (3,5)-12G1-BMPA-GC-(*rac*)-(1,2)(OH)$_2$ (27*rac*, 0.24 g, 0.21 mmol, 1.0 equiv) and (3,4,5)-3EO-G1-COOH (0.29 g, 0.48 mmol, 2.3 equiv) following a procedure similar to that used for the synthesis of **2*R***. The title compound was obtained as a light-yellow oil (0.38 g, 78%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.22 (d, 4 H, PhH), 7.05 (d, 2 H, PhH), 7.05 (d, 2 H, PhH), 6.58 (t, 2 H, PhH), 5.64 (m, 1 H, CH(O-)(CH2O-)2), 4.63–4.43 (m, 8 H, -C((CH2O-)2)CH3, -COO-CH2O-CH(O-)- and -CH(O-)-CH2O-), 4.20 (m, 4 H, PhOCH2CH2O-), 4.13 (m, 8 H, PhOCH2CH2O-), 3.91 (t, 8 H, PhOCH2CH2O-), 3.84 (m, 8 H, PhOCH$_2$(CH$_2$)$_{10}$CH$_3$), 3.78 (t, 4 H, PhOCH2CH2O-), 3.74–3.69 (m, 12 H, PhOCH2CH2OCH2CH2-), 3.67–3.60 (m, 24 H, PhOCH2CH2OCH2CH2OCH2CH2-), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2CH2-), 3.38-3.33 (m, 18 H, -OCH3), 1.74 (m, 8 H, PhOCH$_2$CH$_2$(CH$_2$)$_9$CH$_3$), 1.42 (m, 8 H, PhOCH$_2$CH$_2$CH$_2$(CH$_2$)$_8$CH$_3$), 1.39 (s, 3 H, -C(CH2O-)2CH3), 1.25 (br, 64 H, PhOCH$_2$CH$_2$CH$_2$(CH$_2$)$_8$CH$_3$), 0.87 (t, 12 H, PhO(CH$_2$)$_{11}$CH$_3$). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.3, 165.8, 165.5, 165.1, 160.2, 152.4, 143.0, 143.0, 131.2, 131.2, 124.2, 124.0, 109.1, 109.1, 107.7, 106.5, 72.5, 72.0, 70.9, 70.8, 70.7, 70.6, 70.6, 69.8, 69.7, 68.9, 68.4, 65.7, 63.2, 62.5, 59.1, 46.9, 32.0, 29.7, 29.7, 29.7, 29.7, 29.5, 29.4, 29.3, 26.1, 22.8, 18.2, 14.2. Purity by HPLC: 99+%. MALDI-TOF MS *m/z* of $[M + Na]^+$ calculated for $C_{126}H_{212}O_{38}Na$: 2358.0; Found: 2358.5. The ^{1}H NMR and ^{13}C NMR spectra of compound **2*rac*** are shown in Figure 100 and Figure 101, respectively.

Synthesis of Amphiphilic Glycerol-Based Achiral Janus Dendrimer 3

Scheme 20. Synthesis of Amphiphilic Glycerol-Based Achiral Janus Dendrimer 3. Reagents and conditions: (*i*) DCC, DPTS, DCM, 23 °C, 12 h; (*ii*) H_2, Pd/C, EtOAc, 23 °C, 8 h; (*iii*) 1 M HCl, 1,4-dioxane, 60 °C, 8 h.

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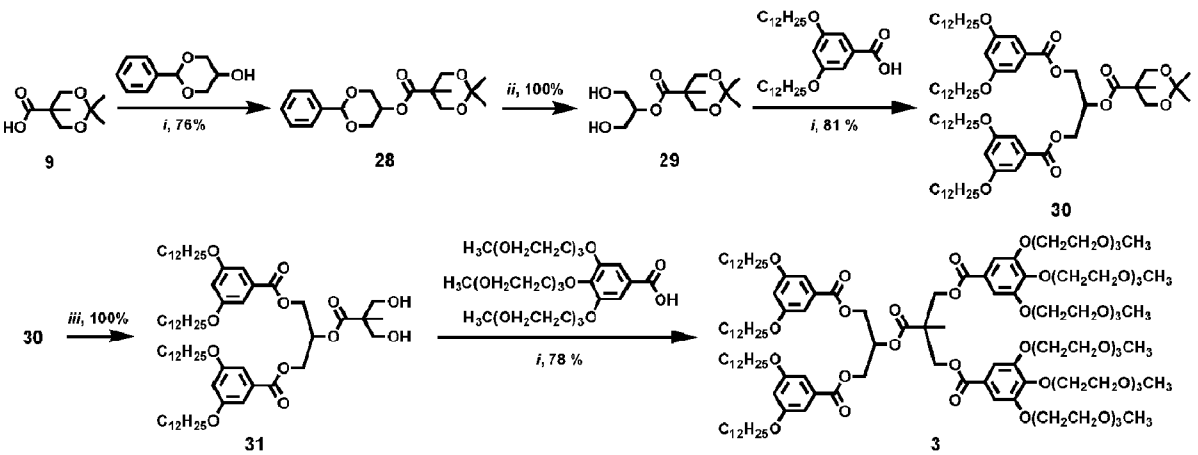
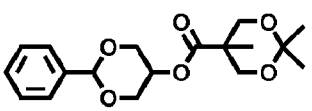
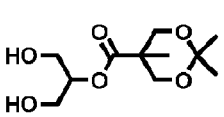




28 2-Phenyl-1,3-dioxan-5-yl-2,2,5-trimethyl-1,3-dioxane-5-carboxylate (28). 2,2,5-Trimethyl-1,3-dioxane-5-carboxylic acid (**9**, 1.00 g, 5.74 mmol, 1.0 equiv), 5-hydroxy-2-phenyl-1,3-dioxane (1.04 g, 5.74 mmol, 1.0 equiv) and DPTS (1.79 g, 5.74 mmol, 1.0 equiv) were dissolved in 10 mL dry DCM. DCC (1.78 g, 8.61 mmol, 1.5 equiv) dissolved in 2 mL dry DCM was added in one portion into the above mixture. The reaction mixture was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The solvent was then removed under vacuum and the crude product was dissolved in EtOAc. The EtOAc solution was filtered through Celite and the filter cake was washed with EtOAc. The filtrate was collected, concentrated and purified by column chromatography (SiO_2) with Hexane/EtOAc = 3/1 to give the title compound as a white solid (1.46 g, 76%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.49 (m, 2 H, Ph*H*), 7.38 (m, 3 H, Ph*H*), 5.55 (s, 1 H, PhC*H*$(OCH_2)_2$-), 4.76 (m, 1 H, -$(OCH_2)_2$-C*H*OCO-), 4.27 (d, 4 H, 2 × -OC*H*$_2$-), 4.18 (q, 2 H, -OC*H*$_2$-), 3.73 (d, 2 H, -OC*H*$_2$-), 1.45 (s, 3 H, C(O-)$_2$(CH_3)C*H*$_3$), 1.41 (s, 3 H, C(O-)$_2$(C*H*$_3$)CH_3), 1.30 (s, 3 H, C(CH2O-)$_2$C*H*$_3$). ^{13}C NMR (500 MHz, $CDCl_3$) δ 174.2, 138.1, 129.3, 128.4, 126.3, 101.5, 98.2, 68.9, 66.3, 66.0, 42.1, 24.4, 23.1, 18.8. Mp = 93 °C.



29 1,3-Dihydroxypropan-2-yl-2,2,5-trimethyl-1,3-dioxane-5-carboxylate (29).

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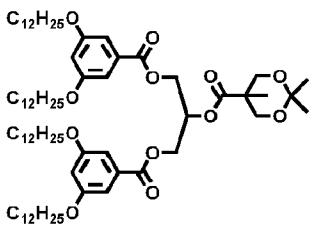
Compound 28 (0.78 g, 2.32 mmol) was dissolved in EtOAc (20 mL). Then Pd/C (39 mg, 5 wt%) was added and the flask was evacuated and filled with hydrogen for three times. The mixture was stirred at 23 °C for 8 h under hydrogen atmosphere. The reaction mixture was filtered through Celite and the filter cake was washed with EtOAc. Evaporation of the solvent yielded the title compound as a colorless oil (0.58 g, 100%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 5.02 (m, 1 H, -$(OCH_2)_2$-C*H*OCO-), 4.24 (d, 2 H, -OC*H*$_2$-), 3.84 (m, 4 H, 2 × -OC*H*$_2$-), 3.70 (d, 2 H, OC*H*$_2$-), 2.84 (br, 2 H, 2 × -CH_2OH), 1.45 (s, 3 H, C(O-)$_2$(CH_3)C*H*$_3$), 1.36 (s, 3 H, C(O-)$_2$(C*H*$_3$)CH_3), 1.09 (s, 3 H, C(CH_2O-)$_2$C*H*$_3$). ^{13}C NMR (500 MHz, $CDCl_3$) δ 174.4, 98.6, 75.9, 66.6, 61.8, 42.7, 27.4, 20.0, 17.8.



30

1,3-bis[3,5-bis(dodecyloxy)benzoate]propan-2-yl-2,2,5-trimethyl-1,3-dioxane-5-carboxylate (30). Compound 29 (0.16 g, 0.64 mmol, 1 equiv), (3,5)-12G1-COOH (0.73 g, 1.47 mmol, 2.3 equiv) and DPTS (0.46 g, 1.47 mmol, 2.3 equiv) were dissolved in 8 mL dry DCM. DCC (0.40 g, 1.92 mmol, 3.0 equiv) dissolved in 2 mL dry DCM was added in one portion into the above mixture. The reaction mixture was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with Hexane/EtOAc = 20/1 to give the title compound as a colorless oil (0.58 g, 81%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.13 (d, 4 H, Ph*H*), 6.64 (t, 2 H, Ph*H*), 5.59 (m, 1 H, -$(OCH_2)_2$-C*H*OCO-), 4.61 (q, 2 H, -OC*H*$_2$-), 4.51 (q, 2 H, -OC*H*$_2$-), 4.14 (d, 2 H, -OC*H*$_2$-), 3.95 (t, 8 H, PhOC*H*$_2$(CH_2)$_{10}$$CH_3$), 3.61 (d, 2 H, -OC*H*$_2$-), 1.77 (m, 8 H, PhOC$H_2$C*H*$_2$($CH_2$)$_9$$CH_3$), 1.44 (m, 8 H, PhOC$H_2CH_2$C*H*$_2$($CH_2$)$_8$$CH_3$), 1.38 (s, 3 H, C(O-)$_2$($CH_3$)C*H*$_3$), 1.28 (s, 3 H, C(O-)$_2$(C*H*$_3$)$CH_3$), 1.26 (br, 64 H, PhOC$H_2CH_2CH_2$(C*H*$_2$)$_8$$CH_3$), 1.20 (s, 3 H, C($CH_2O$-)$_2$C*H*$_3$), 0.88 (t, 12 H, PhO($CH_2$)$_{11}$C*H*$_3$). ^{13}C NMR (500 MHz, $CDCl_3$) δ 173.5, 166.0, 160.4, 131.2, 107.9, 107.0, 98.2, 69.7, 68.4, 65.9, 62.8, 42.0, 32.0, 29.8, 29.8, 29.7, 29.7, 29.5, 29.5, 29.3, 26.2, 23.7, 23.6, 22.8, 18.7, 14.2.

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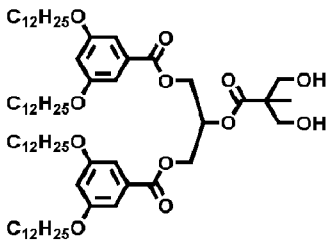
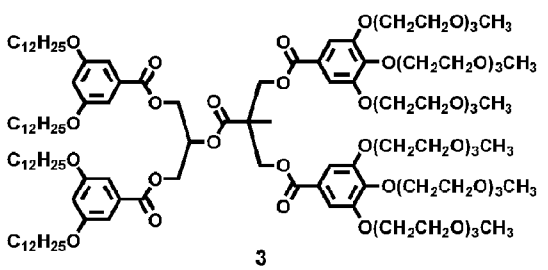


31

1,3-bis[3,5-bis(dodecyloxy)benzoate]propan-2-yl-3-hydroxy-2-(hydroxymethyl)-2-methylpropanoate (31). Compound 30 (0.43 g, 0.36 mmol) was dissolved in 10 mL 1,4-dioxane. Then HCl (2 M, 2.0 mL, 4.0 mmol) was added. The mixture was stirred at 60 °C for 8 h. The reaction mixture was extracted by DCM (3 × 20 mL) and the organic phase was dried over anhydrous magnesium sulfate. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.42 g, 100%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.13 (d, 4 H, Ph*H*), 6.64 (t, 2 H, Ph*H*), 5.60 (m, 1 H, -$(OCH_2)_2$-C*H*OCO-), 4.71 (q, 2 H, -OC*H*$_2$-), 4.51 (q, 2 H, -OC*H*$_2$-), 3.95 (t, 8 H, PhOC*H*$_2(CH_2)_{10}CH_3$), 3.82 (q, 2 H, -OC*H*$_2$-), 3.70 (q, 2 H, -OC*H*$_2$-), 2.75 (t, 2 H, 2 × -CH_2OH), 1.77 (m, 8 H, PhOCH$_2$C*H*$_2(CH_2)_9CH_3$), 1.44 (m, 8 H, PhOCH$_2$CH$_2$C*H*$_2(CH_2)_8CH_3$), 1.26 (br, 64 H, PhOCH$_2$CH$_2$CH$_2$(C*H*$_2)_8CH_3$), 1.06 (s, 3 H, C(CH_2O-)$_2$C*H*$_3$), 0.88 (t, 12 H, PhO$(CH_2)_{11}$C*H*$_3$). ^{13}C NMR (500 MHz, $CDCl_3$) δ 175.0, 166.3, 160.4, 131.0, 107.9, 107.0, 70.1, 68.5, 68.0, 67.6, 62.9, 49.9, 32.0, 29.8, 29.7, 29.7, 29.7, 29.5, 29.5, 29.3, 26.1, 22.8, 17.2, 14.2.



3

(3,5)-12G1-GC-(achiral)-BMPA-(3,4,5)-3EO-G2 (3). Compound 31 (0.21 g, 0.18 mmol, 1 equiv), (3,4,5)-3EO-G1-COOH (0.25 g, 0.41 mmol, 2.3 equiv) and DPTS (0.13 g, 0.41 mmol, 2.3 equiv) were dissolved in 6 mL dry DCM. DCC (0.11 g, 0.54 mmol, 3.0 equiv) dissolved in 2 mL dry DCM was added in one portion into the above mixture. The reaction mixture was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with EtOAc/MeOH = 20/1 as the eluent to give the title compound as a light-yellow oil (0.31 g, 78%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.16 (s, 4 H, Ph*H*), 7.05 (d, 4 H, Ph*H*), 6.58 (t, 2 H, Ph*H*), 5.55 (m, 1 H, C*H*(O-)(CH_2O-)2), 4.61–4.45 (m, 8 H, -C((C*H*2O-)2)CH3 and CH(O-)(C*H*2O-)2), 4.19 (m, 4 H, PhOC*H*2CH2O-), 4.13 (m, 8 H,

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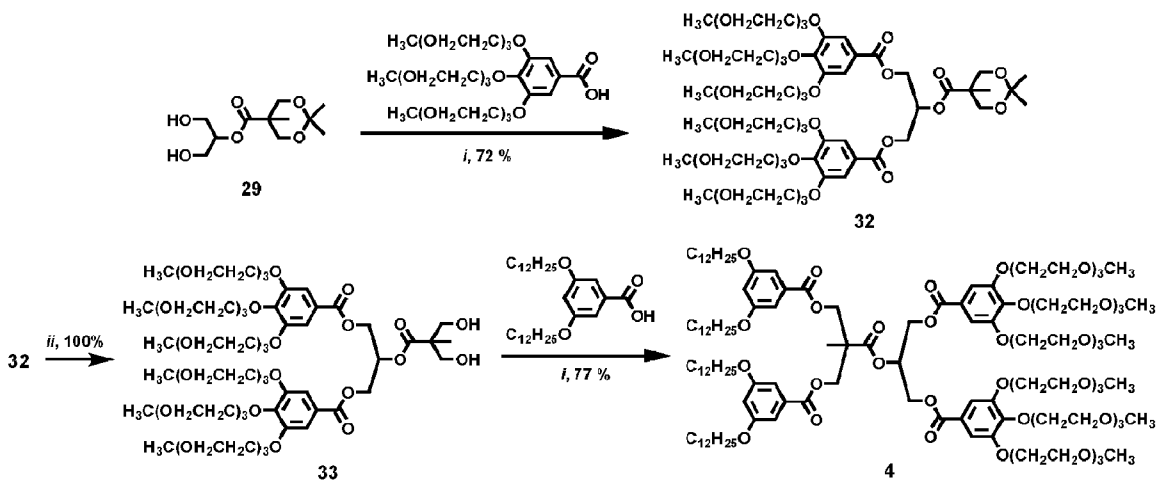
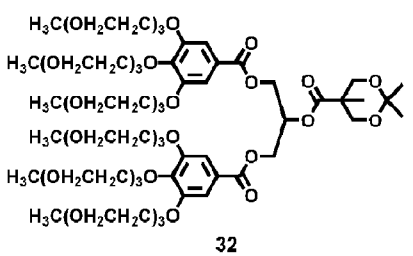
PhOC*H*2CH2O-), 3.91 (t, 8 H, PhOCH2C*H*2O-), 3.84 (m, 8 H, PhOC*H*2(CH2)10CH3), 3.78 (t, 4 H, PhOCH2C*H*2O-), 3.74 3.69 (m, 12 H, PhOCH2CH2OC*H*2CH2-), 3.67 3.61 (m, 24 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2-), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.37-3.34 (m, 18 H, -OC*H*3), 1.75 (m, 8 H, PhOCH2C*H*2(CH2)9CH3), 1.43 (m, 8 H, PhOCH2CH2C*H*2(CH2)8CH3), 1.37 (s, 3 H, -C(CH2O-)2C*H*3), 1.25 (br, 64 H, PhOCH2CH2CH2(C*H*2)8CH3), 0.87 (t, 12 H, PhO(CH2)11C*H*3). ^{13}C NMR (500 MHz, $CDCl_3$) δ 171.9, 165.7, 165.2, 160.2, 152.3, 142.9, 130.8, 124.2, 108.9, 107.6, 106.8, 72.4, 72.0, 71.9, 70.8, 70.7, 70.6, 70.2, 69.6, 68.8, 68.3, 65.6, 62.5, 59.0, 47.1, 31.9, 29.7, 29.7, 29.6, 29.6, 29.4, 29.4, 29.2, 26.1, 22.7, 18.0, 14.1. Purity by HPLC: 99+%. MALDI-TOF MS *m/z* of $[M + Na]^+$ calculated for $C_{126}H_{212}O_{38}Na$: 2358.0; Found: 2358.1. The ^{1}H NMR and ^{13}C NMR spectra of compound **3** are shown in Figure 102 and Figure 103, respectively.

Synthesis of Amphiphilic Glycerol-Based Achiral Janus Dendrimer 4

Scheme 21. Synthesis of Amphiphilic Glycerol-Based Achiral Janus Dendrimer 4. Reagents and conditions: (*i*) DCC, DPTS, DCM, 23 °C, 12 h; (*ii*) 1 M HCl, MeOH, 23 °C, 1 h.





1,3-bis[3,4,5-tris(methyl triethylene glycol)benzoate]propan-2-

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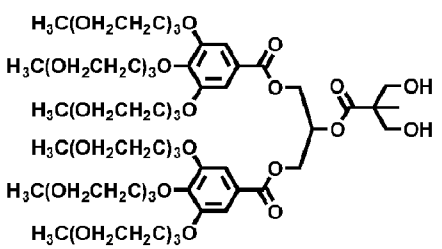
yl-2,2,5-trimethyl-1,3-dioxane-5-carboxylate (32). Compound 29 (0.15 g, 0.60 mmol, 1 equiv), (3,4,5)-3EO-G1-COOH (0.84 g, 1.38 mmol, 2.3 equiv) and DPTS (0.43 g, 1.38 mmol, 2.3 equiv) were dissolved in 8 mL dry DCM. DCC (0.37 g, 1.80 mmol, 3.0 equiv) dissolved in 2 mL dry DCM was added in one portion into the above mixture. The reaction mixture was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 20/1 to give the title compound as a colorless oil (0.61 g, 72%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.26 (s, 4 H, Ph*H*), 5.59 (m, 1 H, C*H*(O-)(CH2O-)2), 4.57 (q, 2 H, -OC*H*2-), 4.49 (q, 2 H, -OC*H*2-), 4.21 (t, 4 H, PhOC*H*2CH2O-), 4.17 (t, 8 H, PhOC*H*2CH2O-), 4.14 (d, 2 H, -OC*H*2-), 3.86 (t, 8 H, PhOCH2C*H*2O-), 3.79 (t, 4 H, PhOCH2C*H*2O-), 3.75–3.69 (m, 12 H, PhOCH2CH2OC*H*2CH2-), 3.67–3.61 (m, 24 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2-), 3.60 (d, 2 H, -OC*H*2-), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.36 (ds, 18 H, -OC*H*3), 1.37 (s, 3 H, C(O-)2(CH3)C*H*3), 1.21 (s, 3 H, C(O-)2(C*H*3)CH3), 1.12 (s, 3 H, -C(CH2O-)2C*H*3). ^{13}C NMR (500 MHz, $CDCl_3$) δ 173.3, 165.4, 152.3, 142.8, 124.0, 109.0, 97.9, 72.3, 71.8, 70.7, 70.6, 70.4, 70.4, 69.6, 69.5, 68.8, 65.7, 62.6, 58.9, 41.9, 24.6, 22.3, 18.4.



33

1,3-bis[3,4,5-tris(methyl triethylene glycol)benzoate]propan-2-yl-3-hydroxy-2-(hydroxymethyl)-2-methylpropanoate (33). Compound 32 (0.20 g, 0.14 mmol) was dissolved in 10 mL MeOH. Then HCl (1 M, 2.0 mL, 2.0 mmol) was added. The mixture was stirred at 23 °C for 1 h. The reaction mixture was extracted by DCM (3 × 20 mL) and the organic phase was dried over anhydrous magnesium sulfate. Filtration and evaporation of the solvent yielded the title compound as a colorless oil (0.19 g, 100%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.29 (s, 4 H, Ph*H*), 5.60 (m, 1 H, C*H*(O-)(CH2O-)2), 4.66 (q, 2 H, -OC*H*2-), 4.44 (q, 2 H, -OC*H*2-), 4.22 (t, 4 H, PhOC*H*2CH2O-), 4.19 (t, 8 H, PhOC*H*2CH2O-), 3.86 (t, 8 H, PhOCH2C*H*2O-), 3.81 (q, 2 H, -OC*H*2-), 3.79 (t, 4 H, PhOCH2C*H*2O-), 3.75–3.68 (m, 14 H, PhOCH2CH2OC*H*2CH2- and -OC*H*2-), 3.67–3.60 (m, 24 H, PhOCH2CH2OCH2C*H*2OC*H*2CH2-), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2C*H*2-), 3.36 (ds, 18 H, -OC*H*3), 3.06 (t, 2 H, 2 × -CH2O*H*), 1.07 (s,

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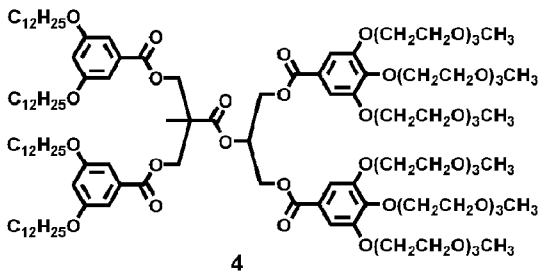
3 H, -C(CH2O-)2CH3). ^{13}C NMR (500 MHz, $CDCl_3$) δ 174.6, 165.6, 152.3, 143.0, 124.0, 109.2, 72.3, 71.8, 71.8, 70.7, 70.6, 70.5, 70.5, 70.4, 69.7, 69.5, 68.8, 62.7, 58.9, 49.7, 17.2.



4

(3,5)-12G2-BMPA-GC-(achiral)-(3,4,5)-3EO-G1 (4). Compound 33 (0.18 g, 0.13 mmol, 1 equiv), (3,5)-12G1-COOH (0.15 g, 0.30 mmol, 2.3 equiv) and DPTS (94 mg, 0.30 mmol, 2.3 equiv) were dissolved in 4 mL dry DCM. DCC (80 mg, 0.39 mmol, 3.0 equiv) dissolved in 2 mL dry DCM was added in one portion into the above mixture. The reaction mixture was allowed to stir at 23 °C for 12 h. The reaction mixture was filtered to remove the urea, which was washed with DCM carefully. The filtrate was concentrated and purified by column chromatography (SiO_2) with DCM/MeOH = 20/1 to give the title compound as a colorless oil (0.23 g, 77%). ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.19 (s, 4 H, PhH), 7.00 (d, 4 H, PhH), 6.55 (t, 2 H, PhH), 5.58 (m, 1 H, CH(O-)(CH2O-)2), 4.63–4.43 (m, 8 H, -C((CH2O-)2)CH3 and CH(O-)(CH2O-)2), 4.20 (t, 4 H, PhOCH2CH2O-), 4.15 (m, 8 H, PhOCH2CH2O-), 3.89 (t, 8 H, PhOCH2CH2O-), 3.85 (t, 8 H, PhOCH2(CH2)10CH3), 3.79 (t, 4 H, PhOCH2CH2O-), 3.74–3.70 (m, 12 H, PhOCH2CH2OCH2CH2-), 3.68–3.60 (m, 24 H, PhOCH2CH2OCH2CH2OCH2CH2-), 3.53 (m, 12 H, PhOCH2CH2OCH2CH2OCH2CH2-), 3.37-3.34 (m, 18 H, -OCH3), 1.74 (m, 8 H, PhOCH2CH2(CH2)9CH3), 1.42 (m, 8 H, PhOCH2CH2CH2(CH2)8CH3), 1.38 (s, 3 H, -C(CH2O-)2CH3), 1.25 (br, 64 H, PhOCH2CH2CH2(CH2)8CH3), 0.87 (t, 12 H, PhO(CH2)11CH3). ^{13}C NMR (500 MHz, $CDCl_3$) δ 172.0, 165.7, 165.4, 160.2, 152.3, 142.8, 131.1, 124.0, 108.9, 107.6, 106.5, 72.5, 72.0, 72.0, 70.8, 70.7, 70.6, 69.7, 68.8, 68.3, 65.7, 62.7, 59.1, 47.1, 32.0, 29.7, 29.7, 29.7, 29.5, 29.4, 29.2, 26.1, 22.7, 18.2, 14.2. Purity by HPLC: 99+%. MALDI-TOF MS *m/z* of $[M + Na]^+$ calculated for $C_{126}H_{212}O_{38}Na$: 2358.0; Found: 2358.2. The ^{1}H NMR and ^{13}C NMR spectra of compound **4** are shown in Figure 104 and Figure 105, respectively.

DLS Data of the Assemblies from Glycerol-Based JDs

Table 25. Size of assemblies from glycerol-based JDs in library 1 at different concentrations of JDs in water (Size distribution is recorded with intensity).

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Concentration of JD (mg·mL^{-1})	1*R*		1*S*		1*rac*		1*R*:1*S* – 1:3		1*R*:1*S* – 1:1		1*R*:1*S* – 3:1	
	D_h (nm)	PDI	D_h (nm)	PDI	D_h (nm)	PDI	D_h (nm)	PDI	D_h (nm)	PDI	D_h (nm)	PDI
0.5	233	0.326	229	0.438	281	0.240	179	0.421	184	0.271	266	0.376
0.4	178	0.228	184	0.258	174	0.275	181	0.268	171	0.274	164	0.246
0.3	148	0.190	147	0.186	153	0.203	144	0.226	147	0.236	153	0.180
0.25	140	0.202	142	0.203	142	0.238	138	0.207	143	0.214	134	0.186
0.2	135	0.175	135	0.195	136	0.216	139	0.197	126	0.201	127	0.234
0.15	124	0.195	124	0.190	128	0.232	116	0.206	119	0.247	117	0.210
0.1	110	0.197	100	0.223	123	0.268	94	0.243	96	0.269	110	0.257
0.05	87	0.336	86	0.359	109	0.277	89	0.401	78	0.368	83	0.334
0.025	82	0.455	80	0.446	86	0.398	84	0.502	76	0.410	74	0.451

Table 26. Size of assemblies from glycerol-based JDs in library 2 at different concentrations of JDs in water (Size distribution is recorded with intensity).

Concentration of JD (mg·mL^{-1})	2*R*		2*S*		2*rac*		2*R*:2*S* – 1:3		2*R*:2*S* – 1:1		2*R*:2*S* – 3:1	
	D_h (nm)	PDI	D_h (nm)	PDI	D_h (nm)	PDI	D_h (nm)	PDI	D_h (nm)	PDI	D_h (nm)	PDI
0.5	319	0.497	288	0.295	412	0.449	304	0.511	324	0.271	390	0.376
0.4	180	0.263	165	0.205	181	0.283	156	0.170	165	0.274	166	0.246
0.3	152	0.227	153	0.235	155	0.225	144	0.295	144	0.236	147	0.180
0.25	139	0.186	132	0.223	147	0.211	134	0.192	138	0.214	133	0.186
0.2	134	0.242	128	0.203	139	0.260	125	0.189	122	0.201	117	0.234
0.15	125	0.257	119	0.212	129	0.236	116	0.223	117	0.247	111	0.210
0.1	111	0.258	112	0.291	122	0.277	113	0.359	110	0.269	103	0.213
0.05	108	0.368	107	0.422	119	0.365	98	0.359	92	0.280	89	0.361
0.025	107	0.416	106	0.465	118	0.361	81	0.419	87	0.309	89	0.439

Table 27. Size of assemblies from glycerol-based achiral JDs **3** and **4** at different concentrations of JDs in water (Size distribution is recorded with intensity).

Concentration	3	4

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of JD (mg·mL^{-1})	D_h (nm)	PDI	D_h (nm)	PDI
1.0	266	0.233	284	0.291
0.75	224	0.236	210	0.233
0.5	162	0.152	166	0.183
0.25	144	0.153	132	0.180
0.1	123	0.183	119	0.136
0.05	97	0.241	102	0.266
0.025	72	0.272	100	0.391

Statistical Analysis of Number of Vesicle Layers of DSs Self-Assembled from Glycerol-Based JDs

Table 28. Number of vesicle layers for each molecule of library 1.

Glycerol JDs	Sample quantity	Number of vesicle layers					
		1	2	3	4	5	6
1*rac*	20	7	6	2	1	1	3
1*R*	82	61	15	4	1	1	0
1*S*	40	23	9	3	4	1	0

Table 29. Number of vesicle layers for each molecule of library **2**.

Glycerol JDs	Sample quantity	Number of vesicle layers											
		1	2	3	4	5	6	7	8	9	10	11	12
2*rac*	306	29	31	40	62	54	27	25	14	14	4	4	2
2*R*	216	202	2	5	1	3	2	0	0	0	0	1	0
2*S*	92	63	13	9	1	0	0	1	1	0	3	1	0

Table 30. Comparison of the distribution parameters, skewness and excess kurtosis.

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Glycerol JDs	Sample quantity	Skewness	Excess Kurtosis
1*rac*	20	0.705	-1.623
1*R*	82	1.977	3.932
1*S*	40	1.687	2.797
2*rac*	306	0.585	-0.423
2*R*	216	3.313	10.980
2*S*	92	3.029	9.502

Table 31. Distribution parameters of **1*rac***.

1*rac*		Statistic	Std. Error
Mean		3.333	1.054
95% Confidence Interval for mean	Lower Bound	0.624	
	Upper Bound	6.043	
5% Trimmed Mean		3.259	
Median		2.500	
Mode		1	
Variance		6.667	
Std. Deviation		2.582	
Minimum		1.00	
Maximum		7.00	
Range		6.00	
Interquartile Range		5.25	
Skewness		0.705	0.845
Excess Kurtosis		-1.623	1.741

Table 32. Test of normality for **1*rac***. *This is a lower bound of the true significance.

	Kolmogorov-Smirnov			Shapiro-Wilk		
	Statistic	df	Sig.	Statistic	df	Sig.
1*rac*	0.218	6	0.200*	0.859	6	0.184

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Table 33. Distribution parameters of ***2rac***.

2rac		Statistic	Std. Error
Mean		25.5	5.601
95% Confidence Interval for mean	Lower Bound	13.172	
	Upper Bound	37.828	
5% Trimmed Mean		24.778	
Median		26.000	
Mode		4	
Variance		376.455	
Std. Deviation		19.402	
Minimum		2.00	
Maximum		62.00	
Range		60.00	
Interquartile Range		31.25	
Skewness		0.585	0.637
Excess Kurtosis		-0.423	1.232

Table 34. Test of normality for ***2rac***. *This is a lower bound of the true significance.

	Kolmogorov-Smirnov			Shapiro-Wilk		
	Statistic	df	Sig.	Statistic	df	Sig.
2rac	0.140	12	0.200*	0.930	12	0.383

The disclosures of each and every patent, patent application, and publication cited herein are hereby incorporated herein by reference in their entirety. While this invention has been disclosed with reference to specific embodiments, it is apparent that other embodiments and variations of this invention may be devised by others skilled in the art without departing from the true spirit and scope of the invention. The appended claims are intended to be construed to include all such embodiments and equivalent variations.

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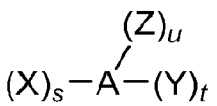
CLAIMS

What is claimed is:

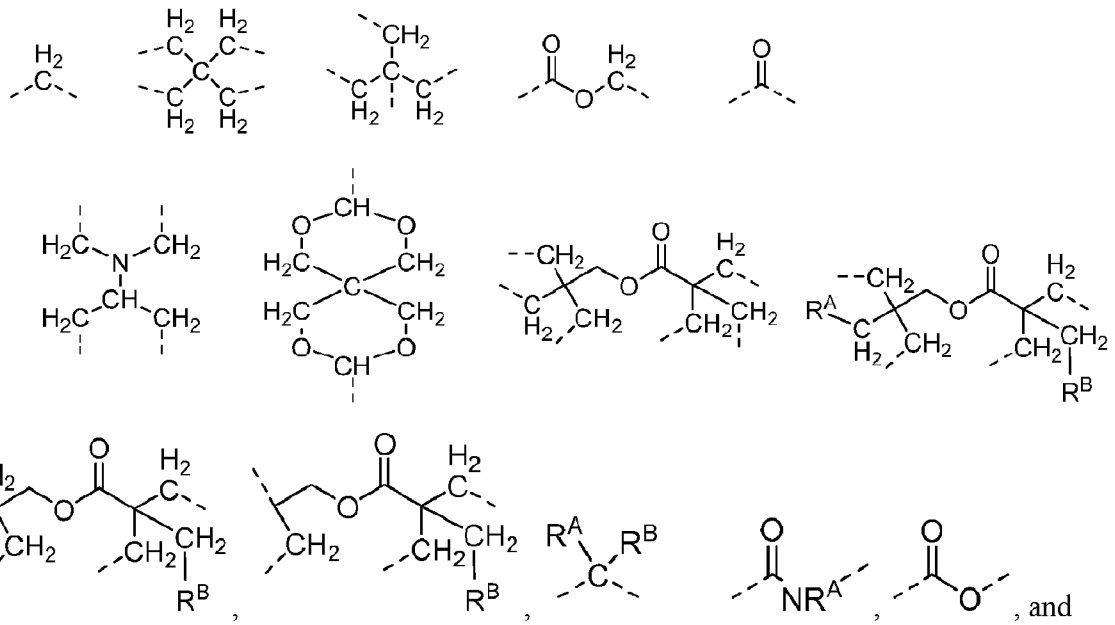
1. An ionizable amphiphilic Janus dendrimer having the structure of Formula (I):



Formula (I)

wherein:

A is a polyvalent group comprising at least one selected from the group consisting of:



, and any combination thereof;

dashed lines represent a binding site of one of X, Y, or Z;

X is a hydrophilic group comprising at least one amine;

Y is a lipophilic group comprising at least one C_1-C_{30}-alkyl chain;

Z comprises at least one selected from the group consisting of ethylene glycol, diethylene glycol, triethylene glycol, and polyethylene glycol chain;

R^A and R^B are independently selected from the group consisting of hydrogen, halide, hydroxy, C_1-C_{30}-alkyl, C_1-C_{30}-alkyl halide, C_1-C_{30}-alkoxy, C_1-C_{30}-alkoxy halide, and any combination thereof;

s is an integer from 0 to 5;

t is an integer from 0 to 5;

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u is an integer from 0 to 4; and

the sum of s, t, and u is equal to the valency of A.

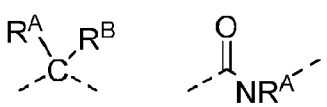
2. The ionizable amphiphilic Janus dendrimer of claim 1, wherein the ionizable amphiphilic Janus dendrimer having the structure of Formula (I) is an ionizable amphiphilic Janus dendrimer having the structure of Formula (II):

$$(X)_s-A-(Y)_t$$

Formula (II)

wherein:

A is a polyvalent group comprising at least one selected from the group consisting of



, and any combination thereof;

dashed lines represent a binding site of one of X and Y;

R^A and R^B are independently selected from the group consisting of hydrogen, halide, hydroxy, C_1-C_{30}-alkyl, C_1-C_{30}-alkyl halide, C_1-C_{30}-alkoxy, C_1-C_{30}-alkoxy halide, and any combination thereof;

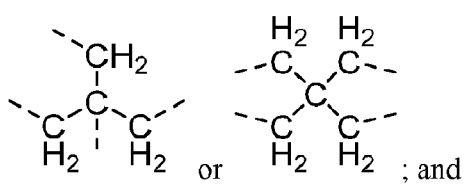
s is an integer from 1 to 5;

t is an integer from 1 to 5; and

the sum of s and t is equal to the valency of A.

3. The ionizable amphiphilic Janus dendrimer of claim 1 or 2, wherein Y is a lipophilic group comprising at least two C_1-C_{30}-alkyl chains having differing numbers of carbon atoms.

4. The ionizable amphiphilic Janus dendrimer of any one of claims 1-3, wherein A is represented by



; and

s and t are each 2.

5. The ionizable amphiphilic Janus dendrimer of any one of claims 1-3, wherein A is

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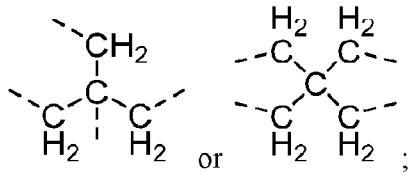
represented by

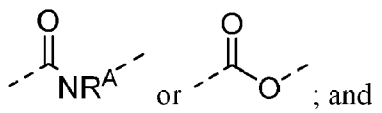


t is 2;

s is 1; and

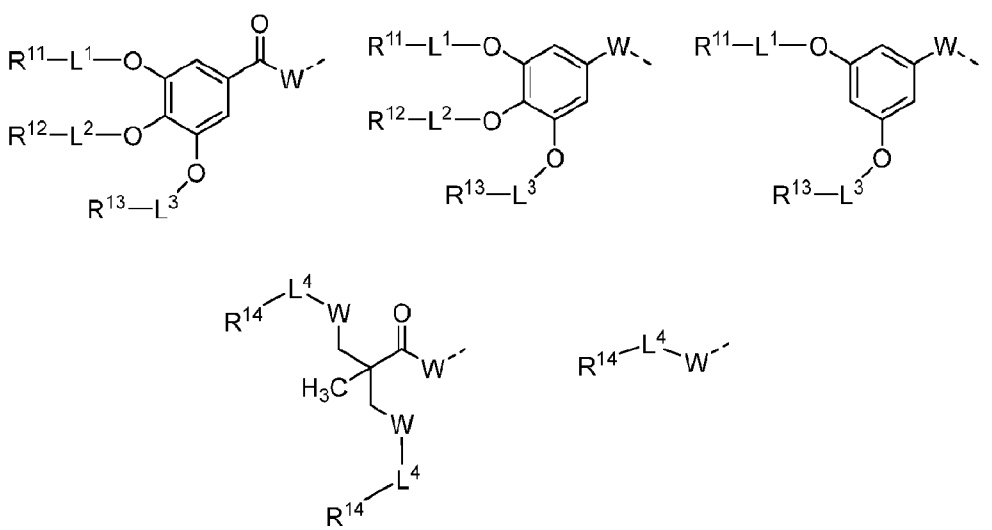
u is 0 or 1.

6. The ionizable amphiphilic Janus dendrimer of any one of claims 1-3, wherein A is represented by



s and t are each 1.

7. The ionizable amphiphilic Janus dendrimer of any one of claims 1-6, wherein each occurrence of X is independently selected from the group consisting of:



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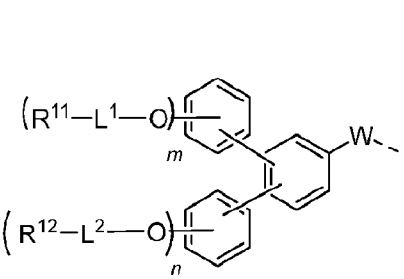
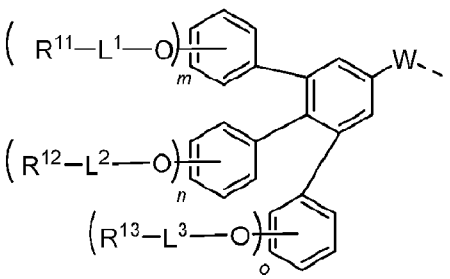
, and any combination thereof;

wherein dashed lines indicate the connection to A;

each occurrence of W is independently selected from the group consisting of C=O, $C(R^W)(R^W)$, NR^W, O, and S;

wherein each occurrence of R^W is independently selected from the group consisting of hydrogen, halide, hydroxy, alkyl, alkyl halide, aryl, aryl halide, alkoxy, alkoxy halide, and any combination thereof; and

each occurrence of L^1, L^2, L^3, and L^4 is independently a covalent bond or a divalent linking group selected from the group consisting of alkylene, cycloalkylene, heteroalkylene, heterocycloalkylene, alkenylene, alkynylene, arylene, heteroarylene, silyl, amine, amide, ester, ether, carbonyl, carbamate, thioether, thioester, disulfide, hydrazine, urea, thiourea, phosphate, poly(alkyl ether), heteroatom, and any combinations thereof;

each occurrence of R^{11}, R^{12}, R^{13}, and R^{14} is independently selected from the group consisting of hydrogen, deuterium, alkyl, aryl, heteroaryl, cycloalkyl, alkoxy, phenoxy, amine, heterocycloalkyl, carbonyl, and any combinations thereof; and

each occurrence of m, n, o is independently an integer from 1 to 5.

8. The ionizable amphiphilic Janus dendrimer of claim 7, wherein at least one occurrence of R^{11}, R^{12}, R^{13}, or R^{14} is $-C(O)(CH_2)_m-N(R^1)(R^2)$;

each occurrence of m is independently an integer from 1 to 10; and

each occurrence of R^1 and R^2 is independently selected from the group consisting of hydrogen, deuterium, alkyl, aryl, cycloalkyl, amine, heterocycloalkyl, carbonyl, and any combinations thereof; wherein R^1 and R^2 may together form a ring.

9. The ionizable amphiphilic Janus dendrimer of claim 7, wherein each occurrence of R^{11}, R^{12}, R^{13}, and R^{14} is independently selected from the group consisting of:

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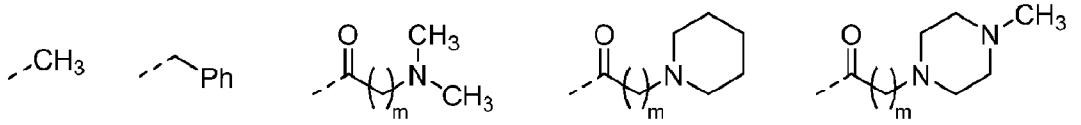
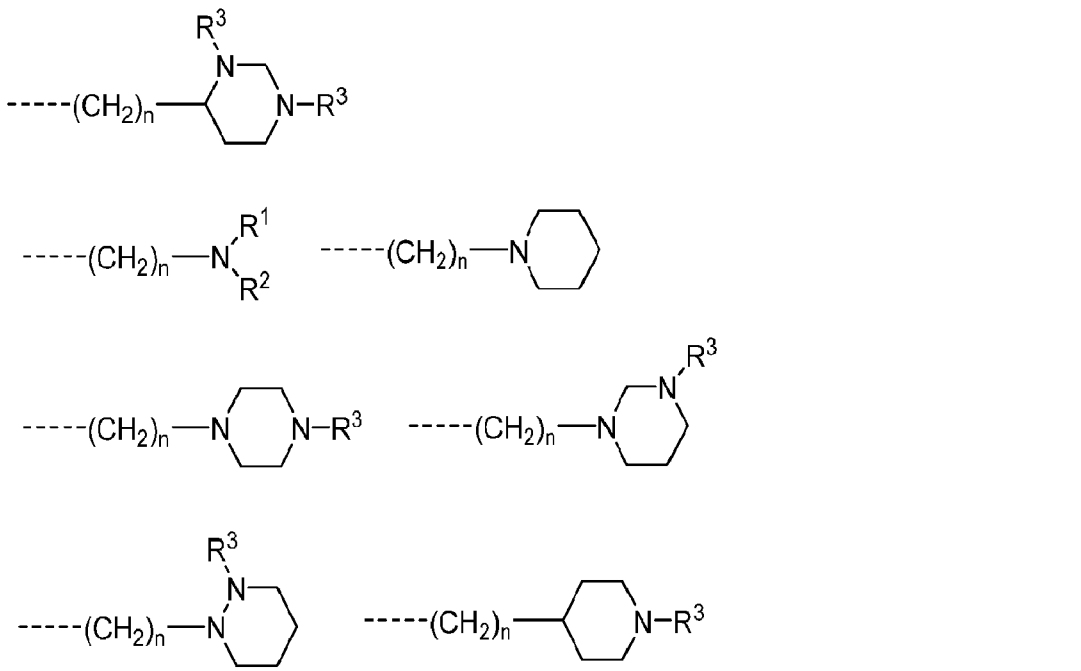


and any combination thereof;

wherein each occurrence of m is independently an integer from 1 to 10.

10. The ionizable amphiphilic Janus dendrimer of any one of the preceding claims, wherein X comprises at least two tertiary amines.

11. The ionizable amphiphilic Janus dendrimer of any one of the preceding claims, wherein each occurrence of X is independently selected from the group consisting of:



and any combination thereof;

wherein dashed lines indicate the connection to A;

each occurrence of R^1 and R^2 is independently selected from the group consisting of hydrogen, deuterium, alkyl, aryl, cycloalkyl, amine, heterocycloalkyl, carbonyl, and any combinations thereof; wherein R^1 and R^2 may together form a ring;

each occurrence of R^3 is independently selected from the group consisting of hydrogen, $(CH_2)_n$, $(CH_2)_n$-OH, and any combination thereof; and

each occurrence of n is independently an integer from 1 to 20.

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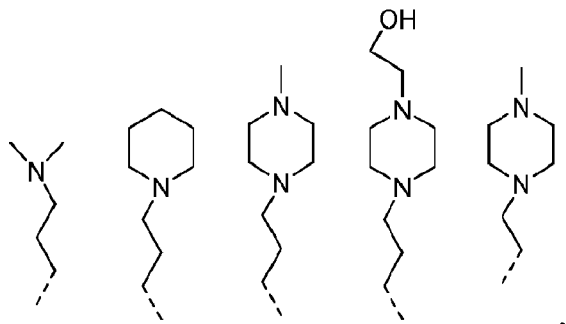
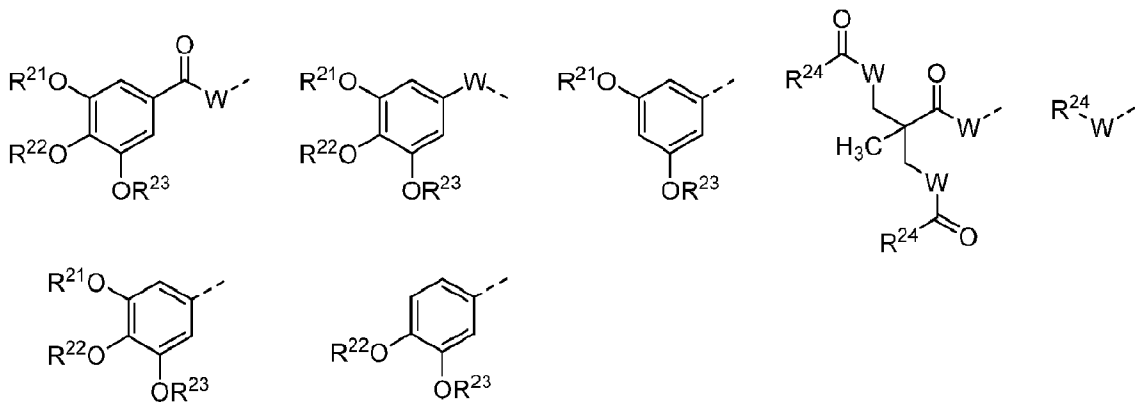
12. The ionizable amphiphilic Janus dendrimer of any one of the preceding claims, wherein each occurrence of X is independently selected from the group consisting of:



, and any combination thereof.

13. The ionizable amphiphilic Janus dendrimer of any one of the preceding claims, wherein each occurrence of Y is independently selected from the group consisting of:



, and any combination thereof;

wherein dashed lines indicate the connection to A;

each occurrence of W is independently selected from the group consisting of C=O, $C(R^W)(R^W)$, NR^W, O, and S;

wherein each occurrence of R^W is independently selected from the group consisting of hydrogen, halide, hydroxy, alkyl, alkyl halide, aryl, aryl halide, alkoxy, alkoxy halide, and any combination thereof; and

each occurrence of R^{21}, R^{22}, R^{23}, and R^{24} independently comprises a linear or branched C_1-C_{50}-alkyl.

14. The ionizable amphiphilic Janus dendrimer of claim 13, wherein each occurrence of R^{21}, R^{22}, R^{23}, and R^{24} independently comprises C_1-C_{30}-alkyl.

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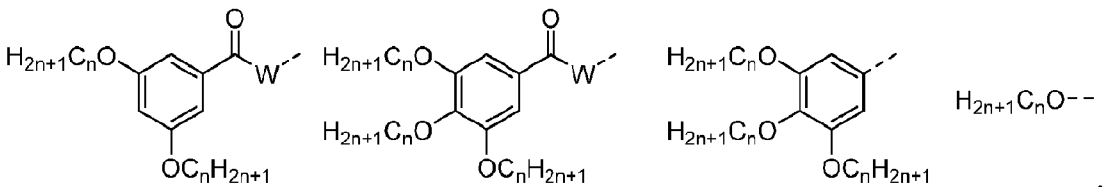
15. The ionizable amphiphilic Janus dendrimer of claim 13, wherein each occurrence of Y is independently selected from the group consisting of:



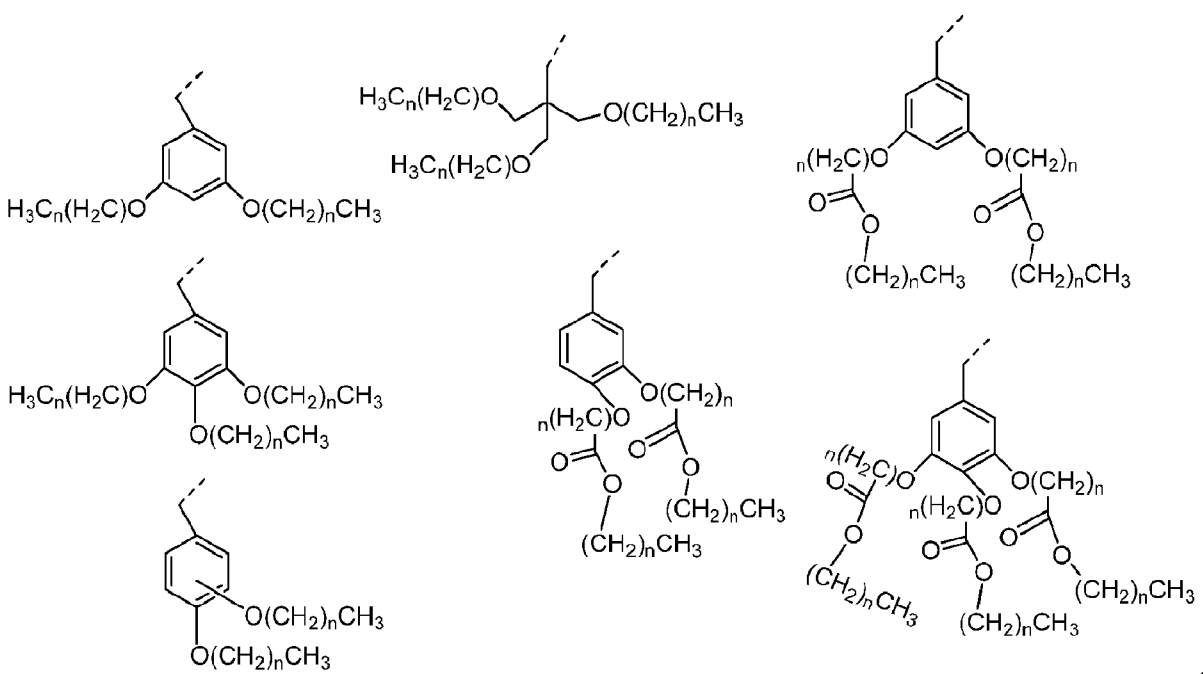
and any combination thereof;

wherein each occurrence of W is independently selected from the group consisting of C=O, $C(R^W)(R^W)$, NR^W, O, and S;

wherein each occurrence of R^W is independently selected from the group consisting of hydrogen, halide, hydroxy, alkyl, alkyl halide, aryl, aryl halide, alkoxy, alkoxy halide, and any combination thereof; and

n is an integer from 1 to 30.

16. The ionizable amphiphilic Janus dendrimer of any one of the preceding claims, wherein each occurrence of Y is independently selected from the group consisting of:



and any combination thereof;

wherein dashed lines indicate the connection to A;

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each occurrence of n is independently an integer from 1 to 20; and

each occurrence of $-(CH_2)_nCH_3$, $-O(CH_2)_n-$, and $-O(CH_2)_nCH_3$ is independently linear or branched.

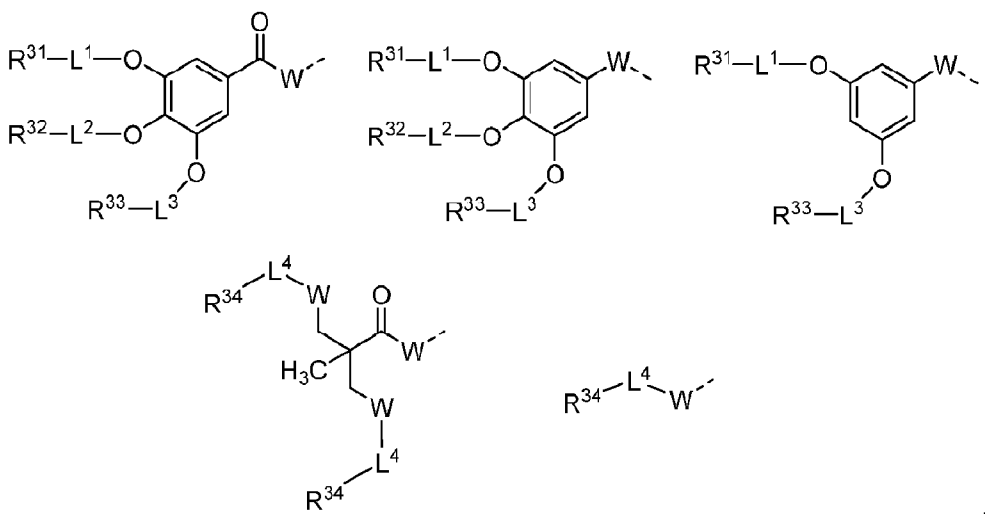
17. The ionizable amphiphilic Janus dendrimer of any one of the preceding claims, wherein the ionizable amphiphilic Janus dendrimer comprises a first Y and a second Y,

wherein the first Y comprises an alkyl chain having an even number of carbon atoms, and the second Y comprises an alkyl chain having an odd number of carbon atoms.

18. The ionizable amphiphilic Janus dendrimer of claim 17, wherein the ratio between the carbon atoms in the first Y and the carbon atoms in the second Y is greater than or equal to 3 and less than 7.

19. The ionizable amphiphilic Janus dendrimer of claim 1, wherein u is 1 or 2;

each occurrence of Z is independently selected from the group consisting of:



, and any combination thereof;

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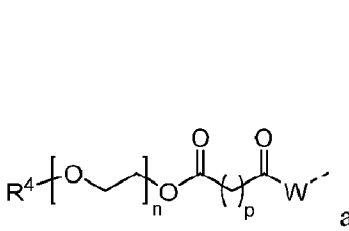
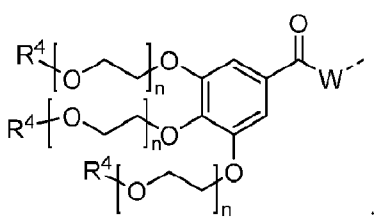
wherein dashed lines indicate the connection to A;

each occurrence of L^1, L^2, L^3, and L^4 is independently a covalent bond or a divalent linking group selected from the group consisting of alkylene, cycloalkylene, heteroalkylene, heterocycloalkylene, alkenylene, alkynylene, arylene, heteroarylene, silyl, amine, amide, ester, ether, carbonyl, carbamate, thioether, thioester, disulfide, hydrazine, urea, thiourea, phosphate, poly(alkyl ether), heteroatom, and any combinations thereof;

each occurrence of R^{31}, R^{32}, R^{33}, and R^{34} is independently selected from the group consisting of hydrogen, deuterium, alkyl, aryl, heteroaryl, cycloalkyl, alkoxy, phenoxy, amine, heterocycloalkyl, carbonyl, and any combinations thereof; and

each occurrence of m, n, o is independently an integer from 1 to 5.

20. The ionizable amphiphilic Janus dendrimer of claim 19, wherein each occurrence of Z is independently selected from the group consisting of:


and

;

wherein each occurrence of n is independently an integer from 1 to 100;

each occurrence of p is independently an integer from 1 to 10; and

each occurrence of R^4 is independently selected from the group consisting of hydrogen, deuterium, alkyl, aryl, and any combinations thereof.

21. The ionizable amphiphilic Janus dendrimer of any one of the preceding claims, wherein the ionizable amphiphilic Janus dendrimer comprises a homochiral, racemic, or achiral branding points.

22. The ionizable amphiphilic Janus dendrimer of any one of the preceding claims, wherein the ionizable amphiphilic Janus dendrimer is a homochiral ionizable amphiphilic Janus dendrimer, racemic ionizable amphiphilic Janus dendrimer, or achiral ionizable amphiphilic Janus dendrimer.

23. The ionizable amphiphilic Janus dendrimer of any one of the preceding claims,

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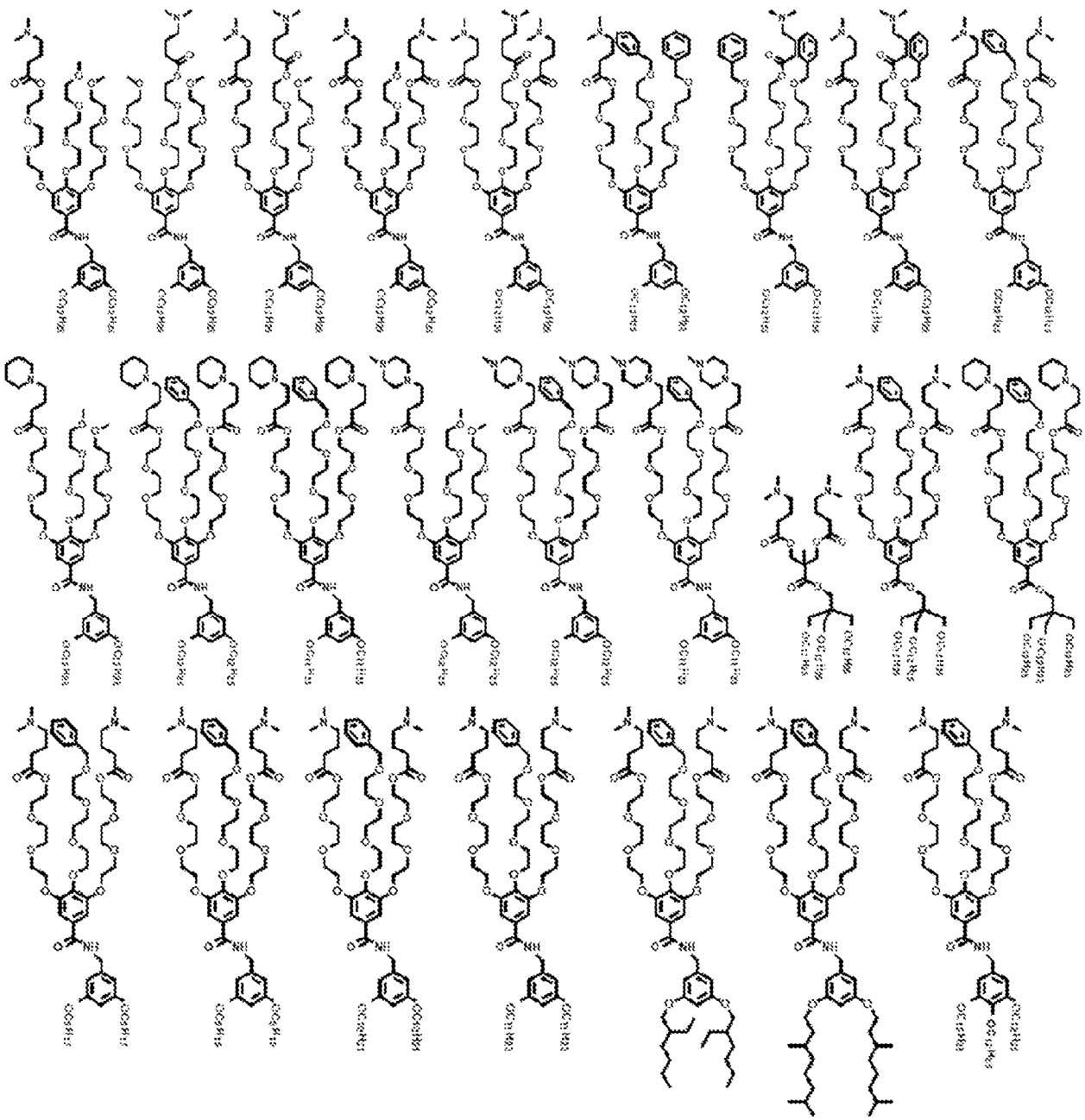
wherein the ionizable amphiphilic Janus dendrimer is an ionizable amphiphilic Janus dendrimer having a structure selected from the group consisting of at least one structure of Figure 12, at least one structure of Figure 13, at least one structure of Figure 14, at least one structure of Figure 47, at least one structure of Figure 60, at least one structure of Figure 80, and any combination thereof.

24. The ionizable amphiphilic Janus dendrimer of any one of the preceding claims, wherein the ionizable amphiphilic Janus dendrimer is an ionizable amphiphilic Janus dendrimer having a structure selected from the group consisting of



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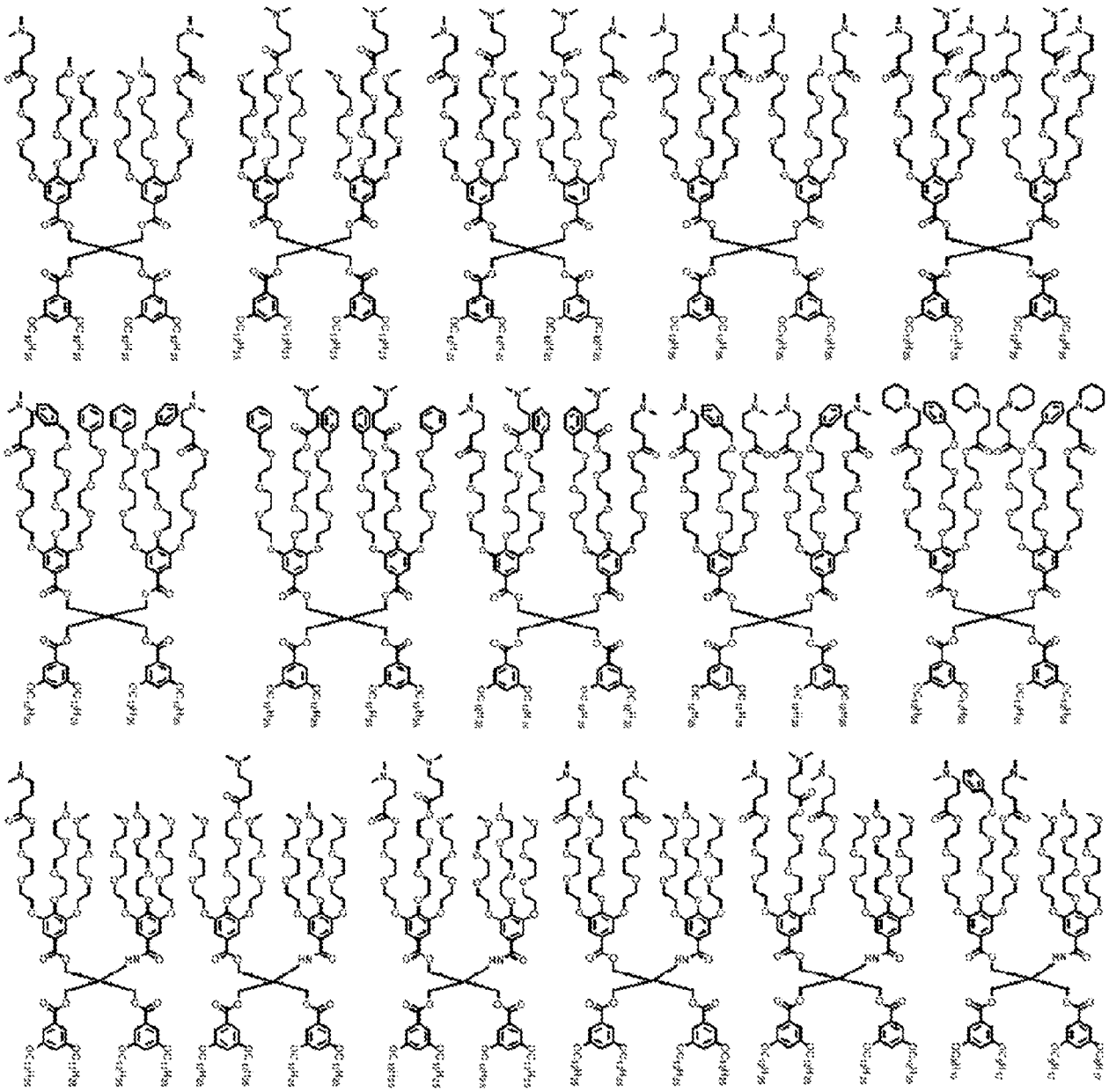

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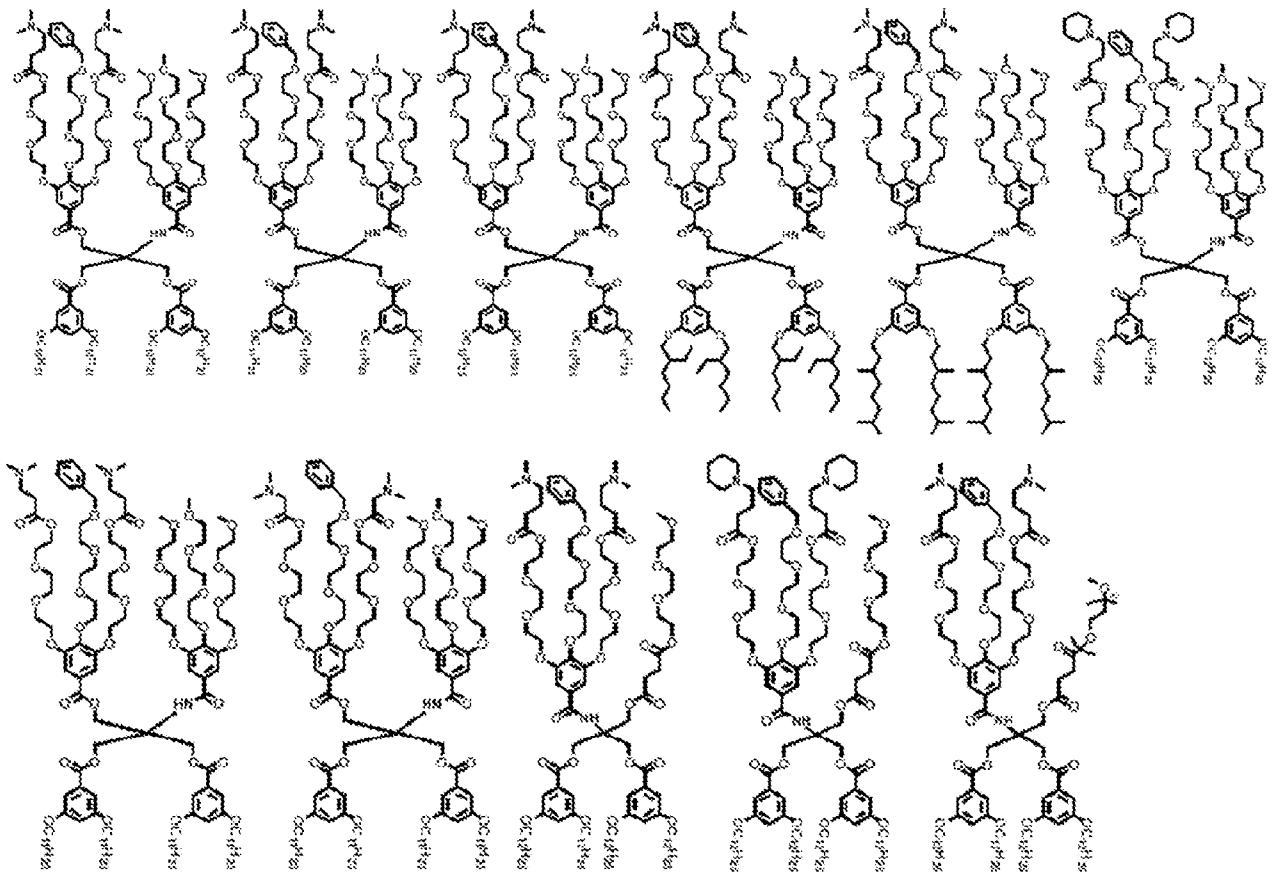

25. A nanoparticle comprising at least one ionizable amphiphilic Janus dendrimer of any one of the preceding claims.

26. The nanoparticle of claim 25, wherein the nanoparticle comprises a first ionizable amphiphilic Janus dendrimer and a second ionizable amphiphilic Janus dendrimer,

wherein the first ionizable amphiphilic Janus dendrimer has a different structure than the second ionizable amphiphilic Janus dendrimer.

27. The nanoparticle of claim 25, wherein the nanoparticle comprises a homochiral ionizable amphiphilic Janus dendrimer, achiral ionizable amphiphilic Janus dendrimer, or any combination thereof.

28. The nanoparticle of claim 27, wherein the nanoparticle is a unilamellar nanoparticle or an onion multilamellar nanoparticle.

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29. The nanoparticle of claim 25, wherein the nanoparticle comprises a racemic ionizable amphiphilic Janus dendrimer.

30. The nanoparticle of claim 29, wherein the nanoparticle is a multilamellar nanoparticle.

31. The nanoparticle of any one of claims 25-30, wherein the nanoparticle further comprises at least one agent.

32. The nanoparticle of claim 31, wherein the at least one agent comprises a diagnostic agent, detectable agent, therapeutic agent, nucleic acid molecule, or any combination thereof.

33. The nanoparticle of claim 31, wherein the at least one agent is selected from the group consisting of an mRNA, siRNA, microRNA, CRISPR-Cas9, sgRNA, small molecule, protein, antibody, peptide, protein, and any combination thereof.

34. The nanoparticle of claim 31, wherein the at least one agent comprises a nucleic acid molecule.

35. The nanoparticle of claim 34, wherein the nucleic acid molecule is a DNA molecule or an RNA molecule.

36. The nanoparticle of claim 34, wherein the nucleic acid molecule is selected from the group consisting of cDNA, cRNA, CirRNA, mRNA, miRNA, siRNA, sgRNA, modified RNA, tRNA, antagomir, antisense molecule, targeted nucleic acid, and any combination thereof.

37. The nanoparticle of claim 34, wherein the nucleic acid molecule encodes at least one selected from the group consisting of an antigen, antibody, gene editing molecule, chimeric antigen receptor (CAR), and any combination thereof.

38. A composition comprising at least one selected from the group consisting of at least one ionizable amphiphilic Janus dendrimer of any one of claims 1-24, at least one nanoparticle of

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any one of claims 25-37, and any combination thereof.

39. The composition of claim 38, wherein the composition further comprises an adjuvant.

40. The composition of claim 38, wherein the composition is a pharmaceutical composition.

41. The composition of claim 38, wherein the composition is a vaccine.

42. A method of delivering an agent to a subject in need thereof, wherein the method comprises administering at least one nanoparticle of any one of claims 31-37 or a composition comprising the same to the subject.

43. The method of claim 42, wherein the composition further comprises an adjuvant.

44. The method of claim 42, wherein the agent is encapsulated within the nanoparticle.

45. The method of claim 42, wherein the method treats or prevents at least one condition selected from the group consisting of a viral infection, bacterial infection, fungal infection, parasitic infection, cancer, disease or disorder associated with cancer, autoimmune disease or disorder, and any combination thereof.

46. The method of claim 42, wherein the agent is a composition for protein replacement therapy.

47. The method of claim 42, wherein the agent is a composition for gene editing.

48. The method of claim 42, wherein the agent is a vaccine.

49. The method of claim 42, wherein the method further comprises delivering the agent to the liver of the subject.

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50. The method of claim 42, wherein the method further comprises delivering the agent to the spleen of the subject.

51. The method of claim 42, wherein the method further comprises delivering the agent to the lungs of the subject.

52. The method of claim 42, wherein the agent comprises at least one selected from the group consisting of cDNA, cRNA, CirRNA, mRNA, miRNA, siRNA, sgRNA, modified RNA, tRNA, antagomir, antisense molecule, targeted nucleic acid, and any combination thereof.

53. The method of claim 52, wherein the modified RNA is a nucleoside-modified RNA.

54. The method of claim 53, wherein the nucleoside-modified RNA comprises pseudouridine.

55. The method of claim 53, wherein the nucleoside-modified RNA comprises pseudouridine plus 5-methyl-cytosine.

56. The method of claim 53, wherein the nucleoside-modified RNA comprises 5-methyl-uridine.

57. The method of claim 53, wherein the nucleoside-modified RNA comprises 1-methyl-pseudouridine.

58. A method of preventing or treating a disease or disorder in a subject in need thereof, wherein the method comprises administering at least one nanoparticle of any one of claims 25-37 or a composition comprising the same to the subject.

59. The method of claim 58, wherein the disease or disorder is selected from the group consisting of a viral infection, bacterial infection, fungal infection, parasitic infection, cancer, disease or disorder associated with cancer, autoimmune disease or disorder, and any combination thereof.

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60. A method of inducing an immune response in a subject in need thereof, wherein the method comprises administering at least one nanoparticle of any one of claims 25-37 or a composition comprising the same to the subject.

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One-Component IAJD Dendrimersome Nanoparticle (DNP)
One-Component
Ionizable Amphiphilic
Janus Dendrimer
(IAJD)
SS
SS
TM
Ethanol
Injection
Buffer
(pH = 3 ~ 4)
(e.g. Luc-mRNA in Acidic Buffer)
No Dialysis
-or-
Dialysis
(pH = 7)
-or-
pH ≅ 7.4)
DNPs Encapsulating
mRNA
Extracellular
(pH = 7.4)
Endocytosis
Cytoplasm
(pH = 7.2)
Endosomal
Escape
mRNA
Translation
Ribosome
Peptides
Nucleus
DNP Encapsulating mRNA

Figure 1

Four-Component Ionizable Lipid Nanoparticle (LNP)
Four-Component
Ionizable lipid
Phospholipid
PEGylated lipid
Cholesterol
mRNA
LNP Encapsulating mRNA
Ethanol
Buffer
(pH = 3 ~ 4)
1. Microfluidic mix
2. Dialysis (pH = 7)
LNPs Encapsulating mRNA
Four-Component
Ionizable lipid
Phospholipid
PEGylated lipid
Cholesterol
mRNA/Luc-mRNA
1. Mix via Microfluidic Device
2. Dialysis (pH = 7.4)
LNPs

Figure 2

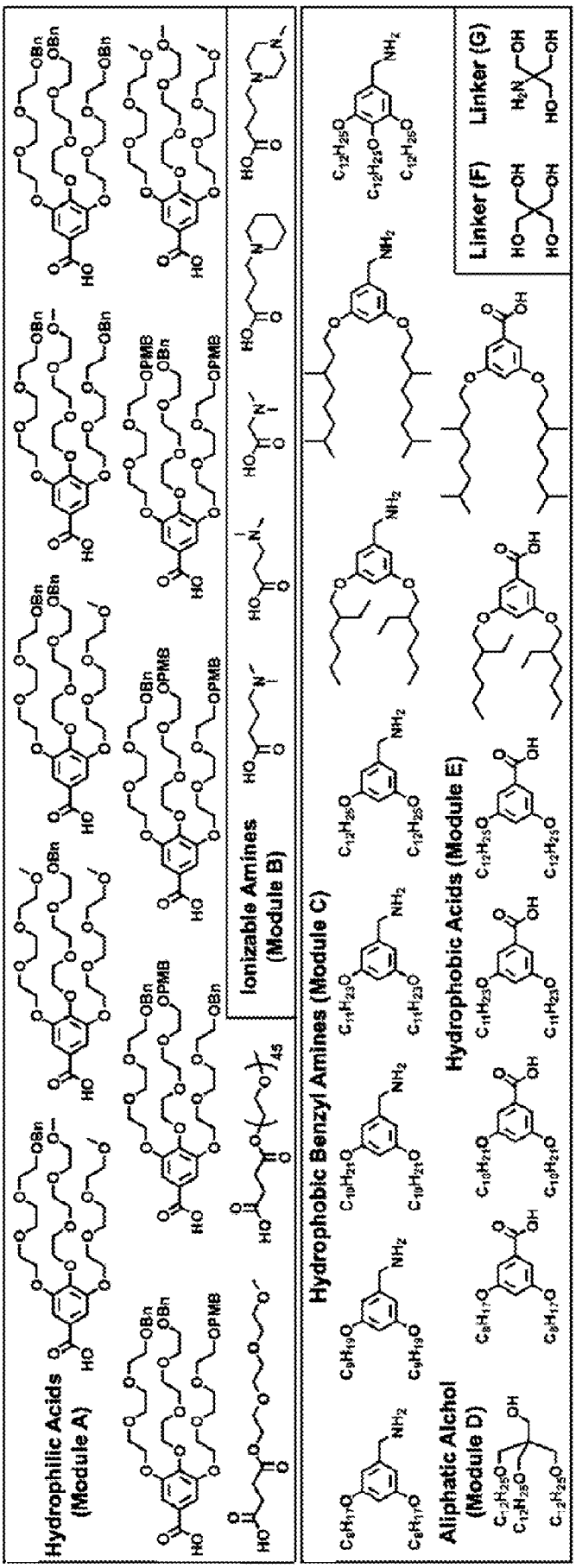
Hydrophilic Acids (Module A)
Ionizable Amines (Module B)
Hydrophobic Benzyl Amines (Module C)
Hydrophobic Acids (Module E)
Aliphatic Alchol (Module D)
Linker (F)
Linker (G)

Figure 3

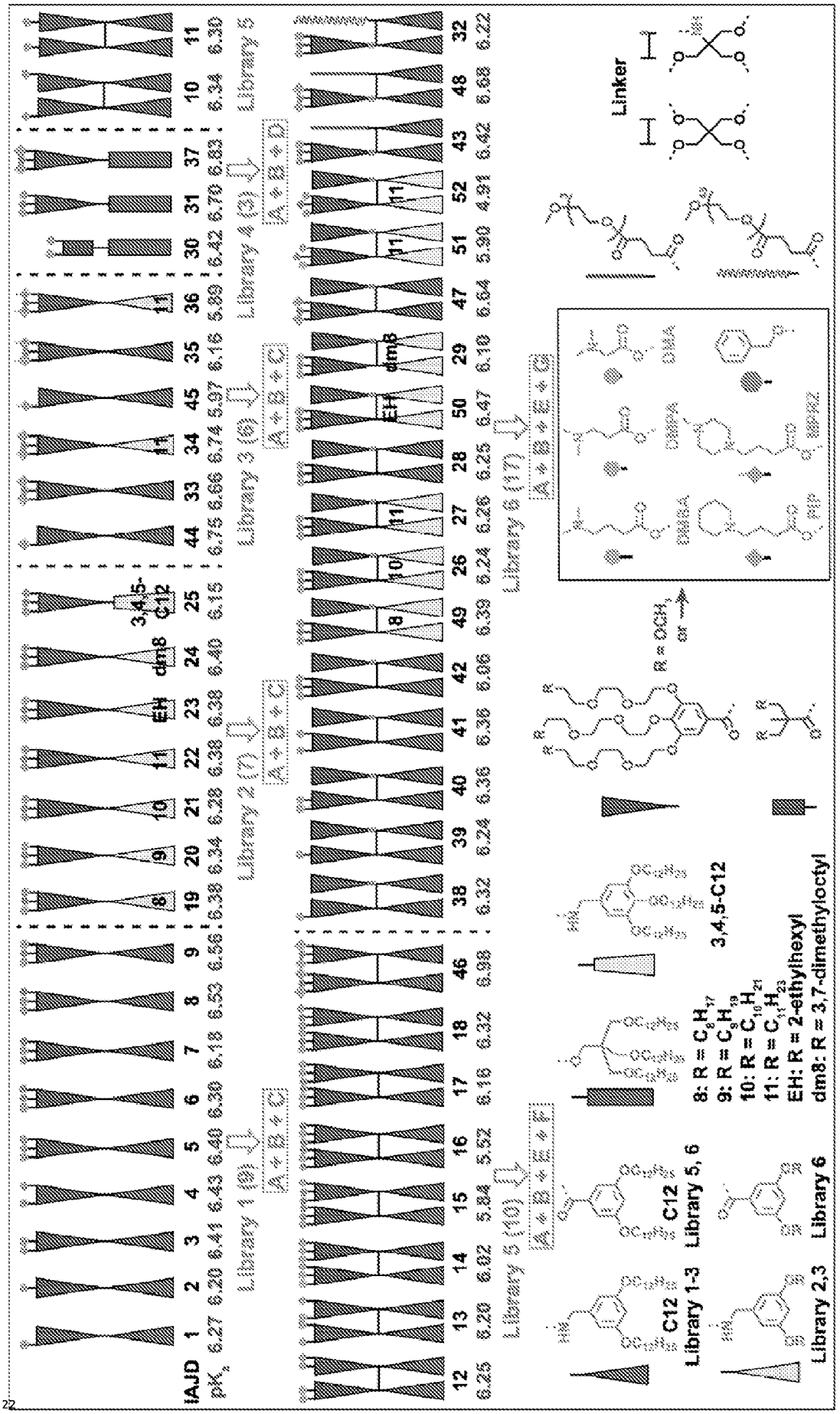
IAJD
pK$_a$
Library 1 (8)
A + B + C
Library 2 (7)
A + B + C
Library 3 (6)
A + B + C
Library 4 (3)
A + B + D
Library 5
Library 5 (10)
A + B + E + F
Library 6 (17)
A + B + E + G
Linker
C12
Library 1-3
Library 5, 6
Library 2,3
Library 6
8: R = C_8H_{17}
9: R = C_9H_{19}
10: R = $C_{10}H_{21}$
11: R = $C_{11}H_{23}$
EH: R = 2-ethylhexyl
dm8: R = 3,7-dimethyloctyl
3,4,5-C12

Figure 4

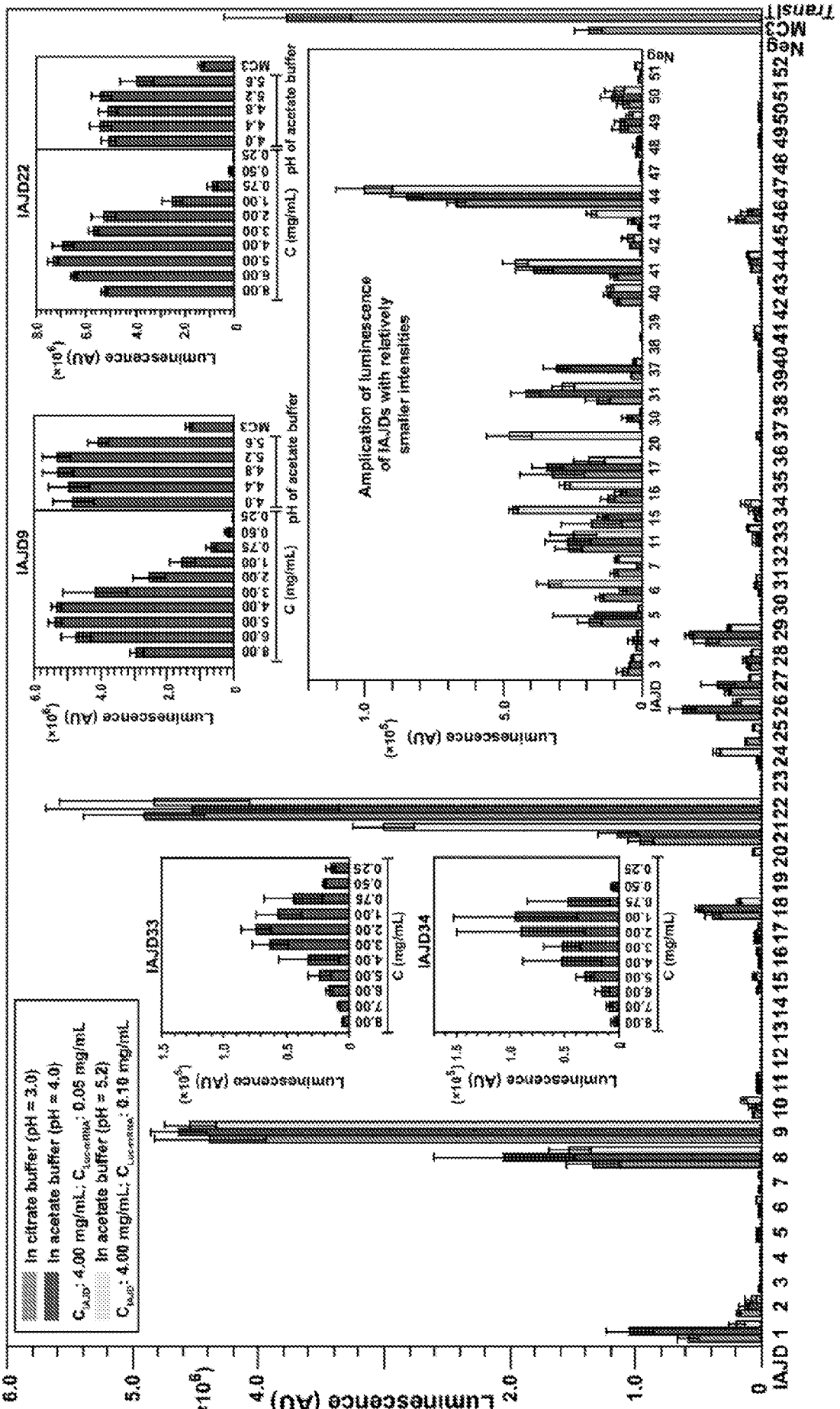
In citrate buffer (pH = 3.0)
In acetate buffer (pH = 4.0)
C_{IAJD}: 4.00 mg/mL; $C_{succinic}$: 0.05 mg/mL
In acetate buffer (pH = 5.2)
C_{IAJD}: 4.00 mg/mL; $C_{succinic}$: 0.10 mg/mL
Luminescence (AU)
(×10^6)
IAJD 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52
Neg
MC1
MC2
Transit
IAJD33
IAJD34
IAJD9
IAJD22
C (mg/mL)
pH of acetate buffer
MC3
Amplification of luminescence of IAJDs with relatively smaller intensities
Neg

Figure 5

Figure 6

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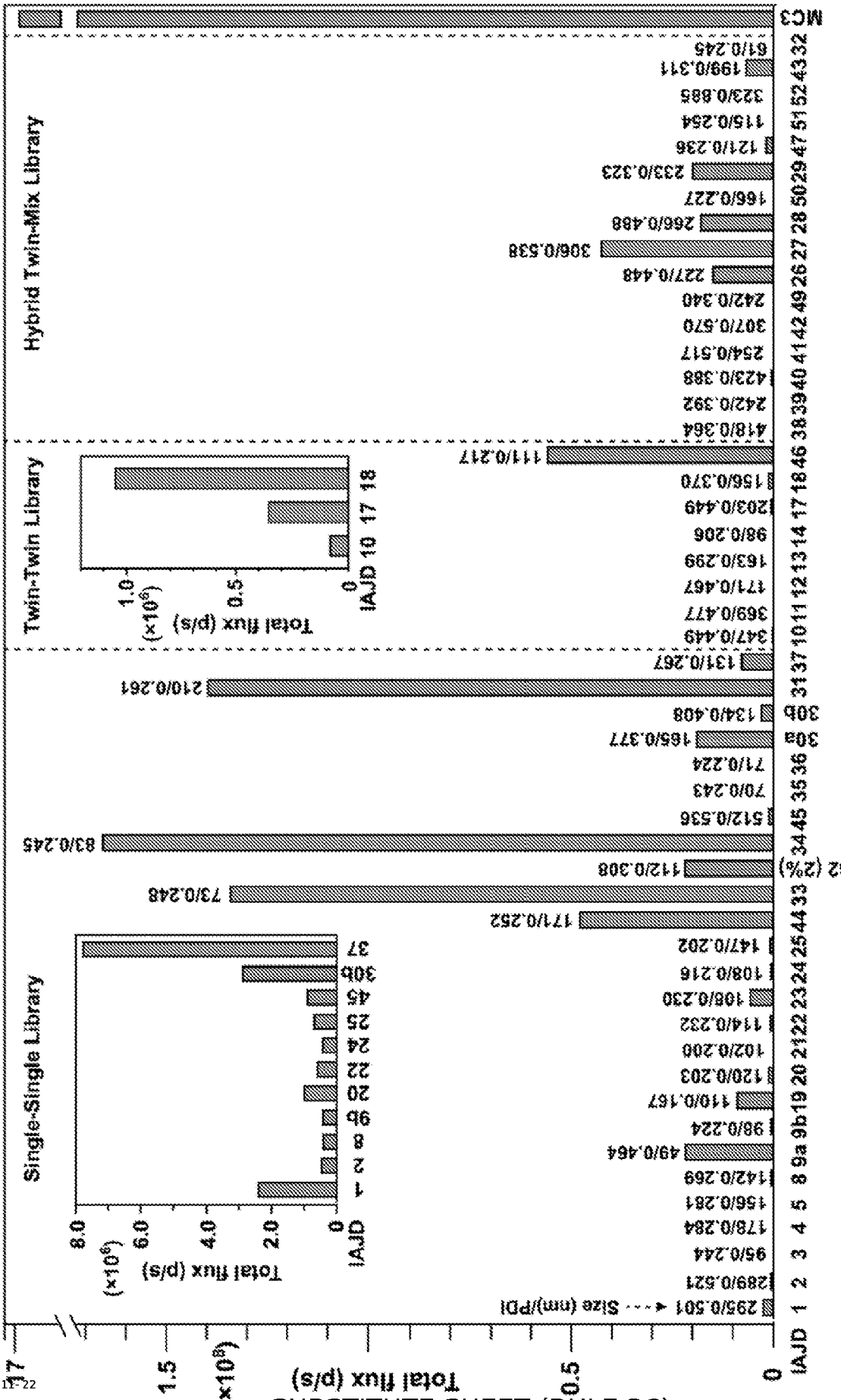

Hybrid Twin-Mix Library
Twin-Twin Library
Single-Single Library
IAJD
Size (nm)/PDI
Total flux (p/s)
(×10^8)
(×10^6)
33+32 (%)
MC3

Figure 7

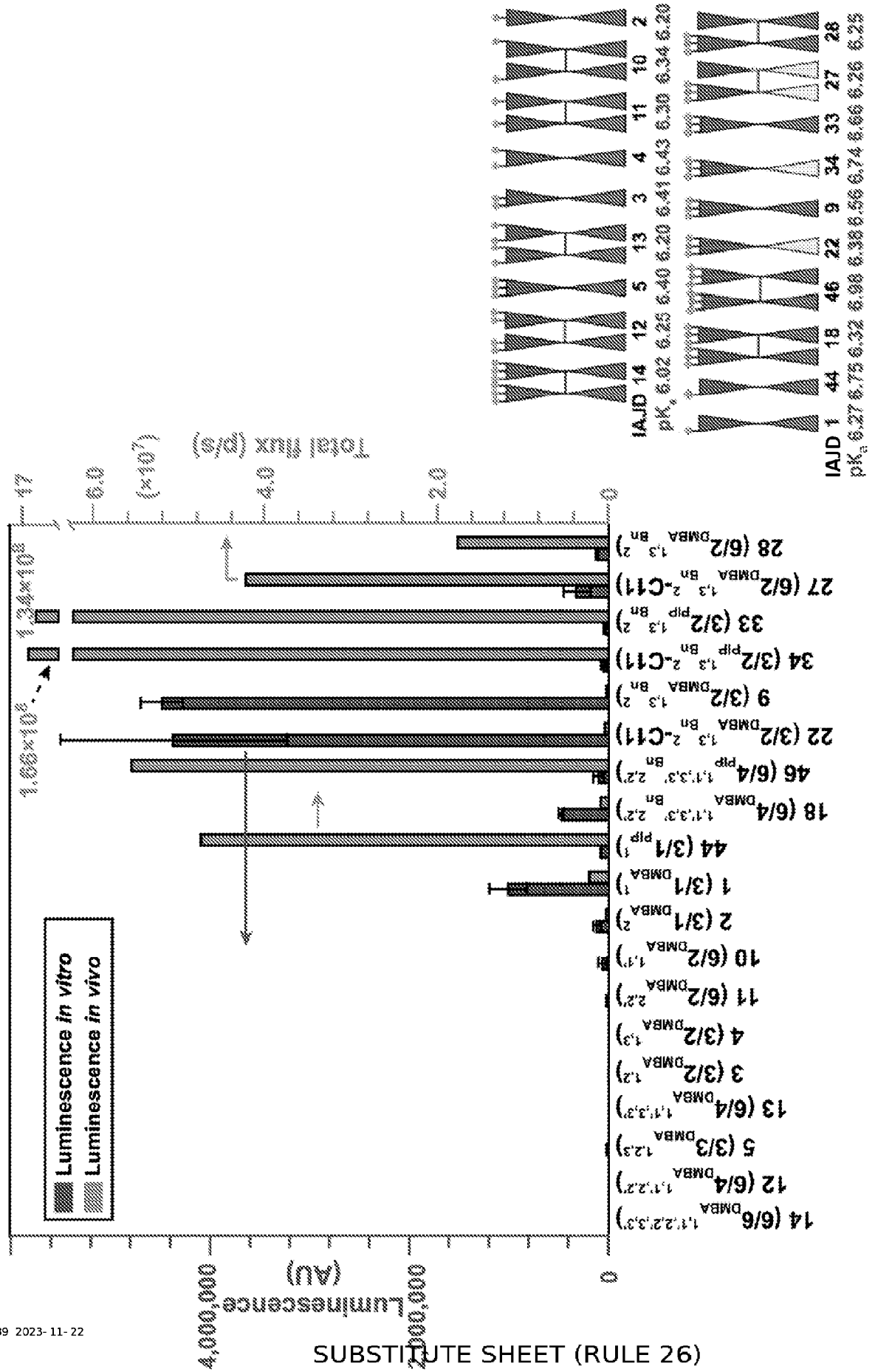

IAJD 1 44 18 46 22 9 34 33 27 28
pKa 6.27 6.75 6.32 6.98 6.38 6.56 6.74 6.66 6.26 6.25
IAJD 14 12 5 13 3 4 11 10 2
pKa 6.02 6.25 6.40 6.20 6.41 6.43 6.30 6.34 6.20
Total flux (p/s) (×10^7)
17
6.0
4.0
2.0
0
1.66×10^8
1.24×10^8
Luminescence in vitro
Luminescence in vivo
Luminescence (AU)
4,000,000
2,000,000
0
14 (6/6DMBA 1,1',2,2',3,3')
12 (6/4DMBA 1,1',2,2')
5 (3/3DMBA 1,2,3)
13 (6/4DMBA 1,1',3,3')
3 (3/2DMBA 1,2)
4 (3/2DMBA 1,3)
11 (6/2DMBA 2,2')
10 (6/2DMBA 1,1')
2 (3/1DMBA 2)
1 (3/1DMBA 1)
44 (3/1PIP 1)
18 (6/4DMBA 1,1',3,3' Bn 2,2')
46 (6/4PIP 1,1',3,3' Bn 2,2')
22 (3/2DMBA 1,3 Bn 2-C11)
9 (3/2DMBA 1,3 Bn 2)
34 (3/2PIP 1,3 Bn 2-C11)
33 (3/2PIP 1,3 Bn 2)
27 (6/2DMBA 1,3 Bn 2-C11)
28 (6/2DMBA 1,3 Bn 2)

Figure 8

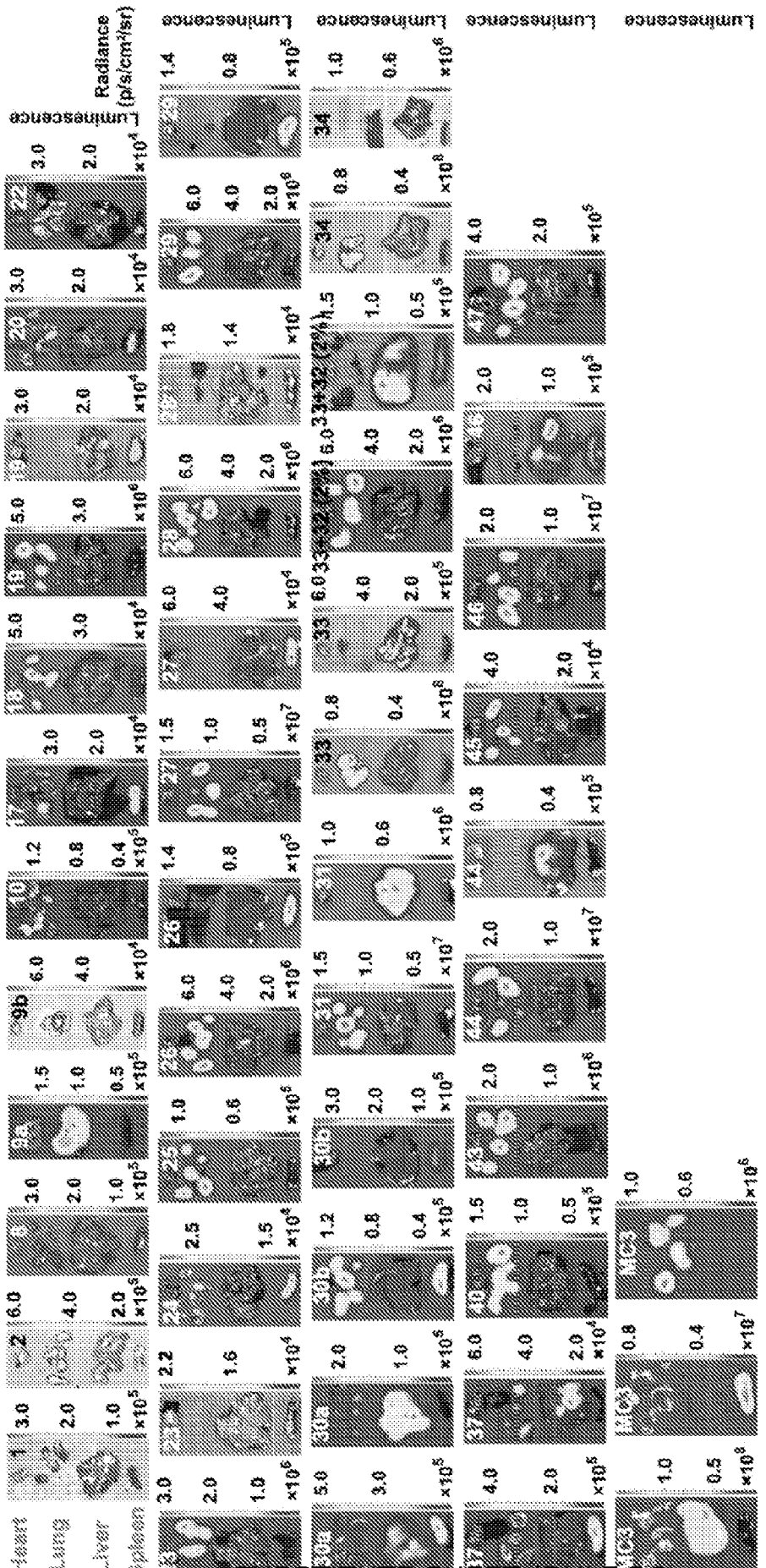


Figure 9

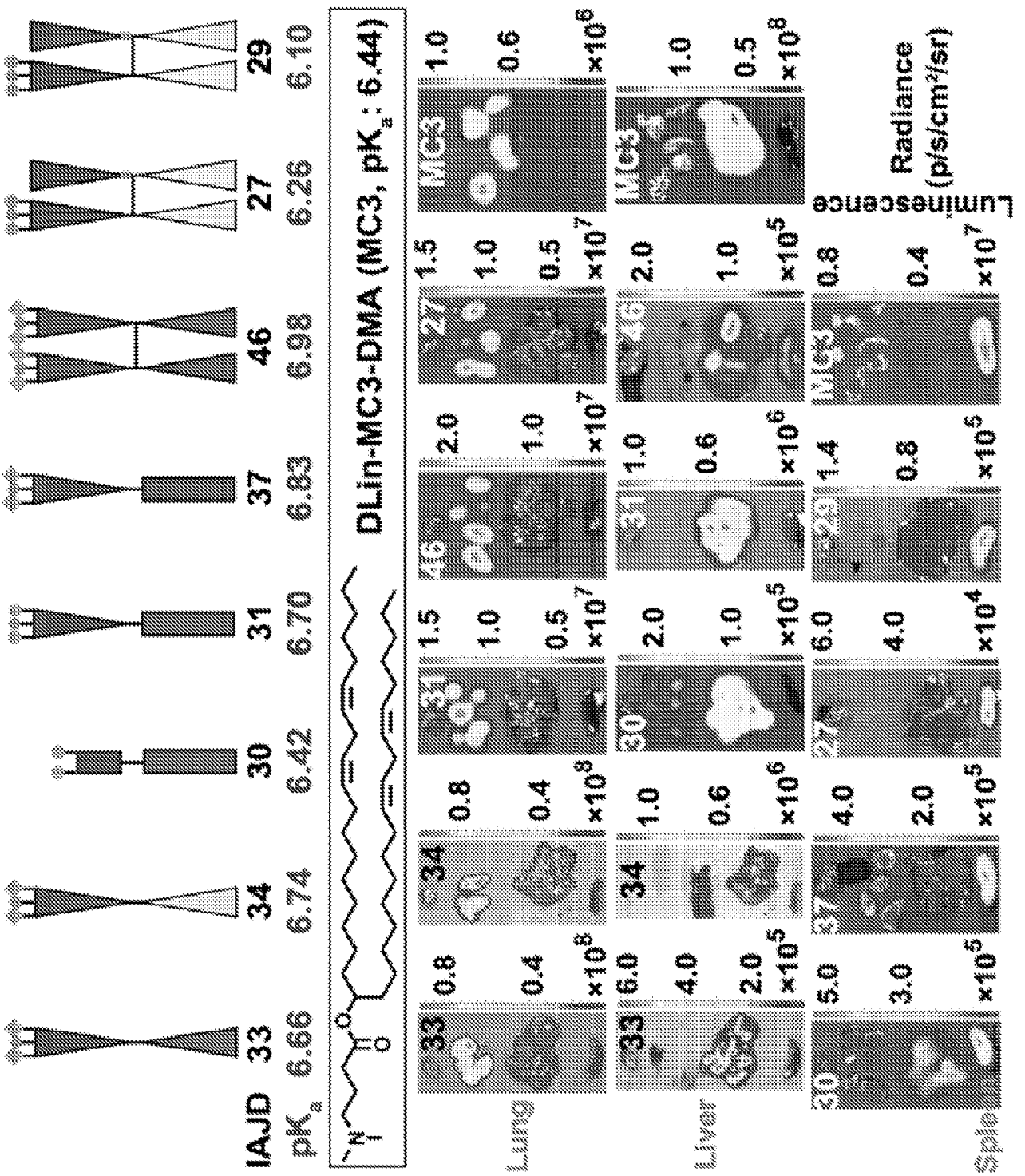

IAJD 33 34 30 31 37 46 27 29
pKa 6.66 6.74 6.42 6.70 6.83 6.98 6.26 6.10
DLin-MC3-DMA (MC3, pKa: 6.44)
Lung
Liver
Spleen
MC3
Luminescence
Radiance (p/s/cm²/sr)

Figure 10

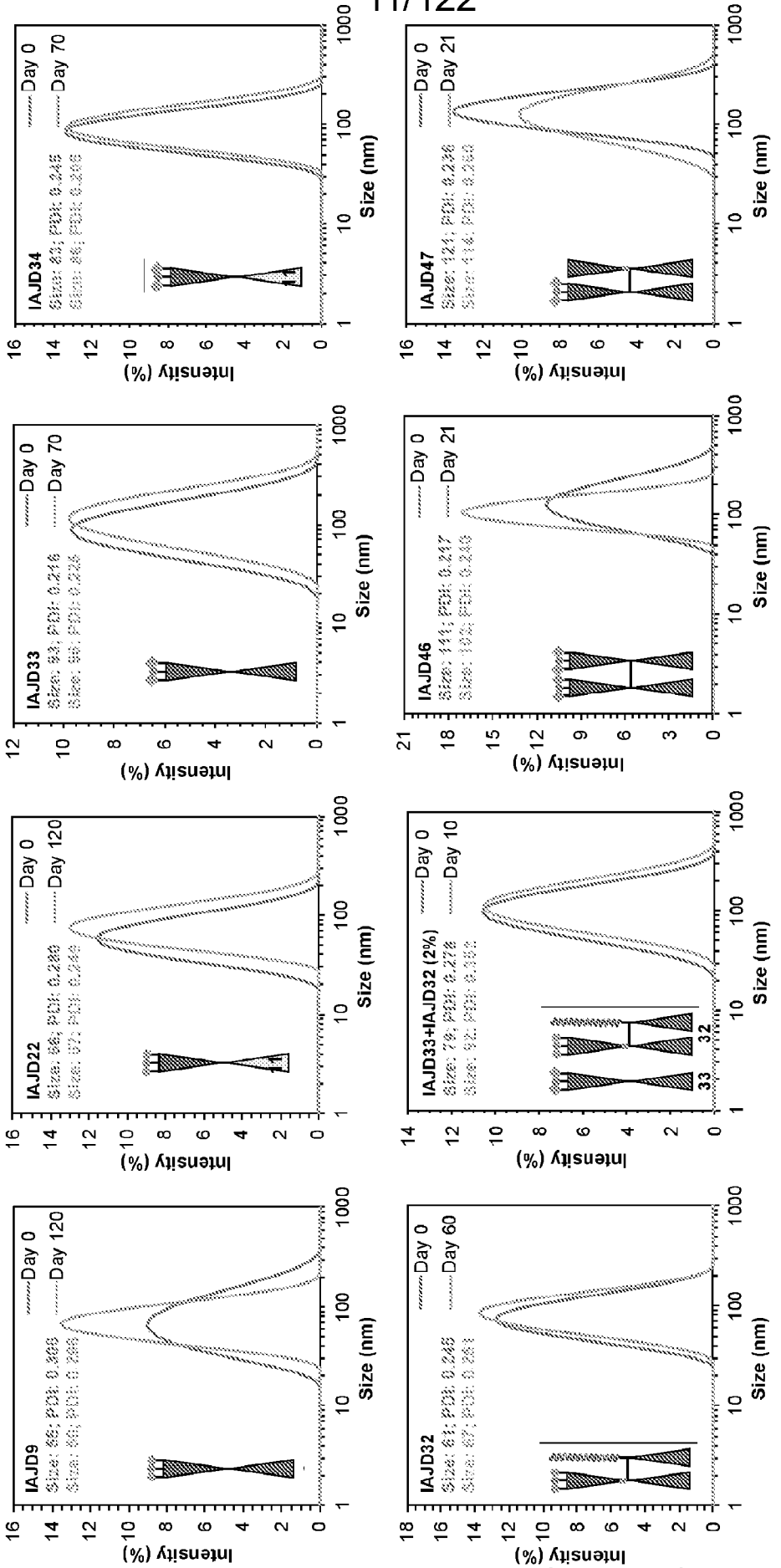

IAJD9
Day 0
Day 120
IAJD32
Day 0
Day 60
IAJD22
Day 0
Day 120
IAJD33+IAJD32 (2%)
Day 0
Day 10
IAJD33
Day 0
Day 70
IAJD46
Day 0
Day 21
IAJD34
Day 0
Day 70
IAJD47
Day 0
Day 21
Intensity (%)
Size (nm)

Figure 11

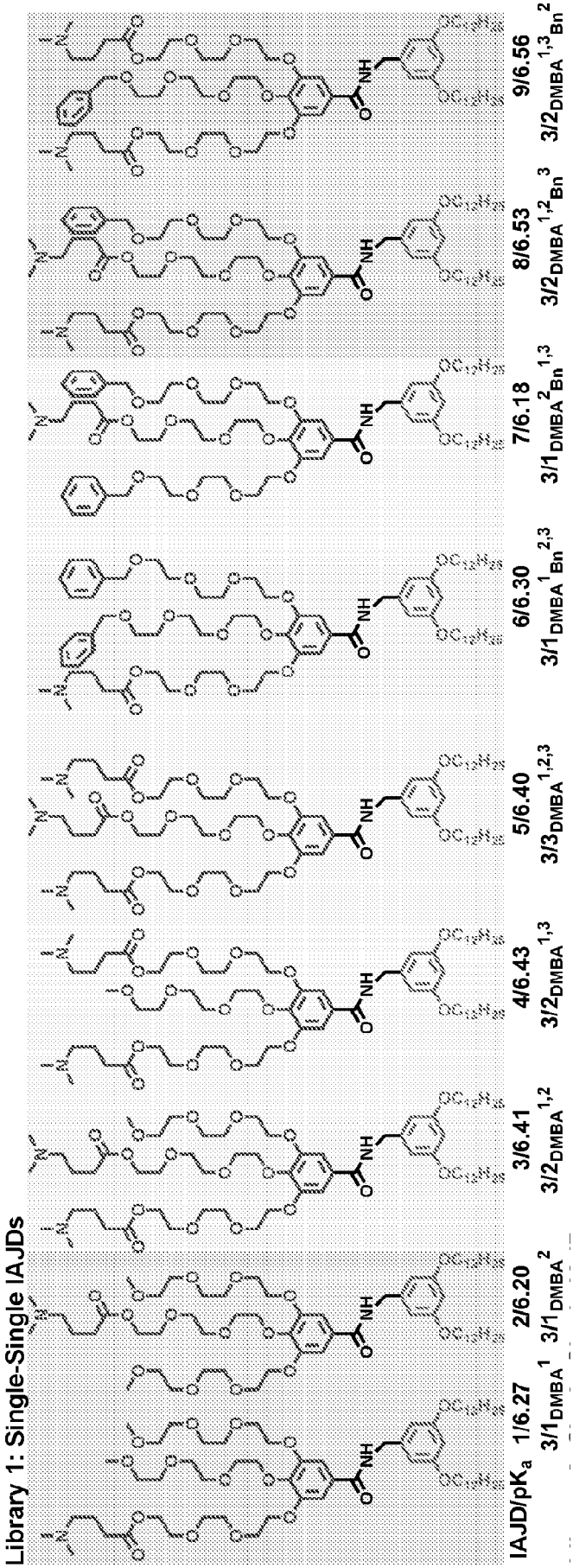

Library 1: Single-Single IAJDs
IAJD/pKa
1/6.27
3/1DMBA1
2/6.20
3/1DMBA2
3/6.41
3/2DMBA1,2
4/6.43
3/2DMBA1,3
5/6.40
3/3DMBA1,2,3
6/6.30
3/1DMBA1Bn2,3
7/6.18
3/1DMBA2Bn1,3
8/6.53
3/2DMBA1,2Bn3
9/6.56
3/2DMBA1,3Bn2

Figure 12

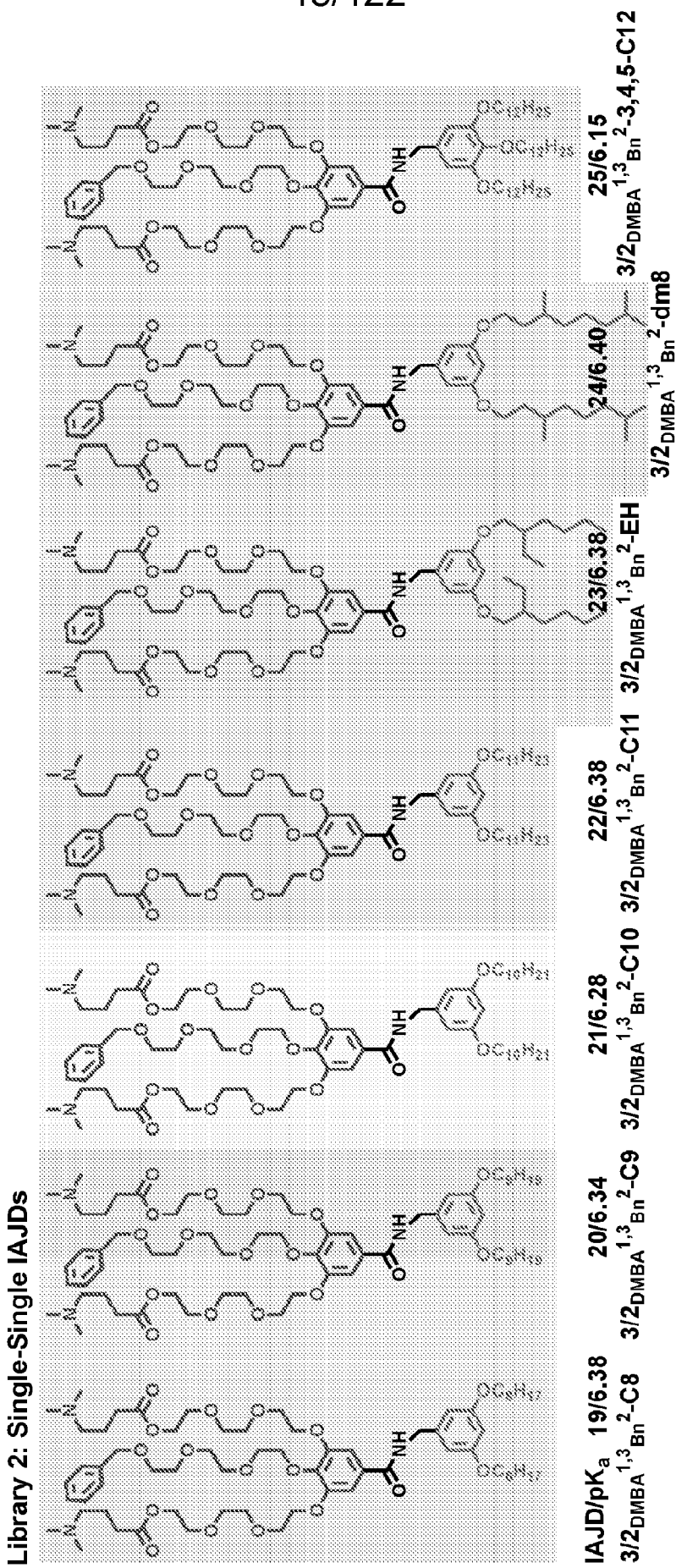


Figure 12 (cont.)

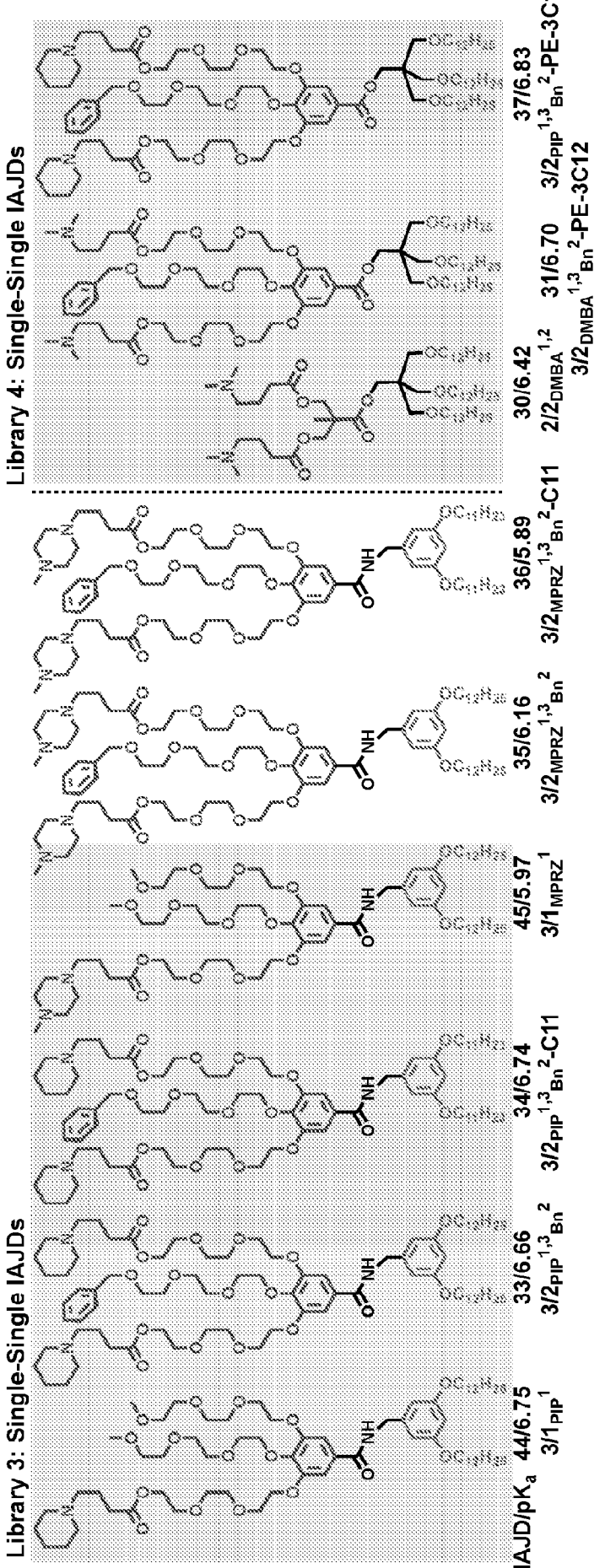


Figure 12 (cont.)

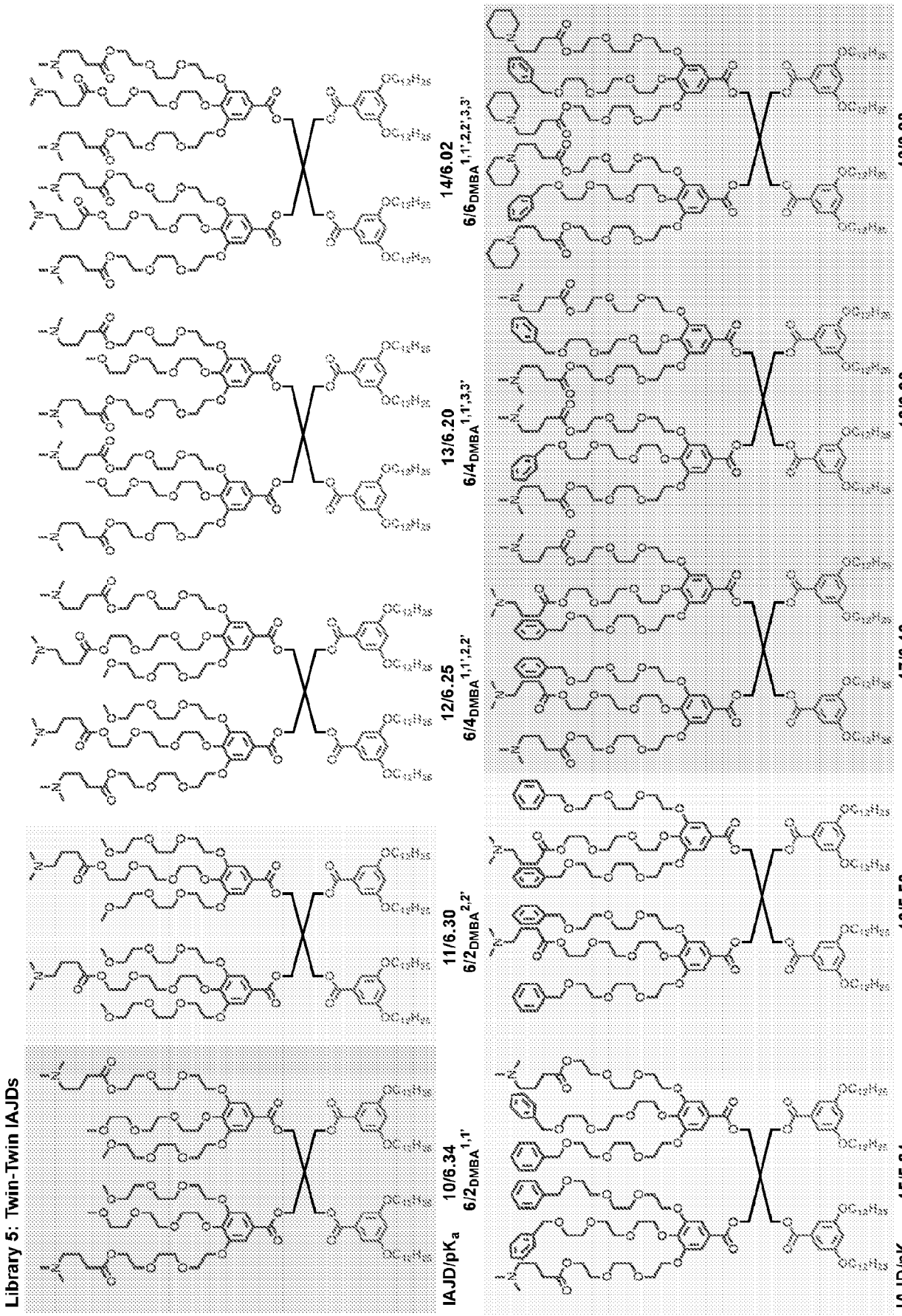

Library 5: Twin-Twin IAJDs
IAJD/pKa
10/6.34
6/2DMBA1,1'
11/6.30
6/2DMBA2,2'
12/6.25
6/4DMBA1,1',2,2'
13/6.20
6/4DMBA1,1',3,3'
14/6.02
6/6DMBA1,1',2,2',3,3'
IAJD/pKa
15/5.84
6/2DMBA1,1' Bn2,2',3,3'
16/5.52
6/2DMBA2,2' Bn1,1',3,3'
17/6.16
6/4DMBA1,1',2,2' Bn3,3'
18/6.32
6/4DMBA1,1',3,3' Bn2,2'
46/6.98
6/4PIP1,1',3,3' Bn2,2'

Figure 13

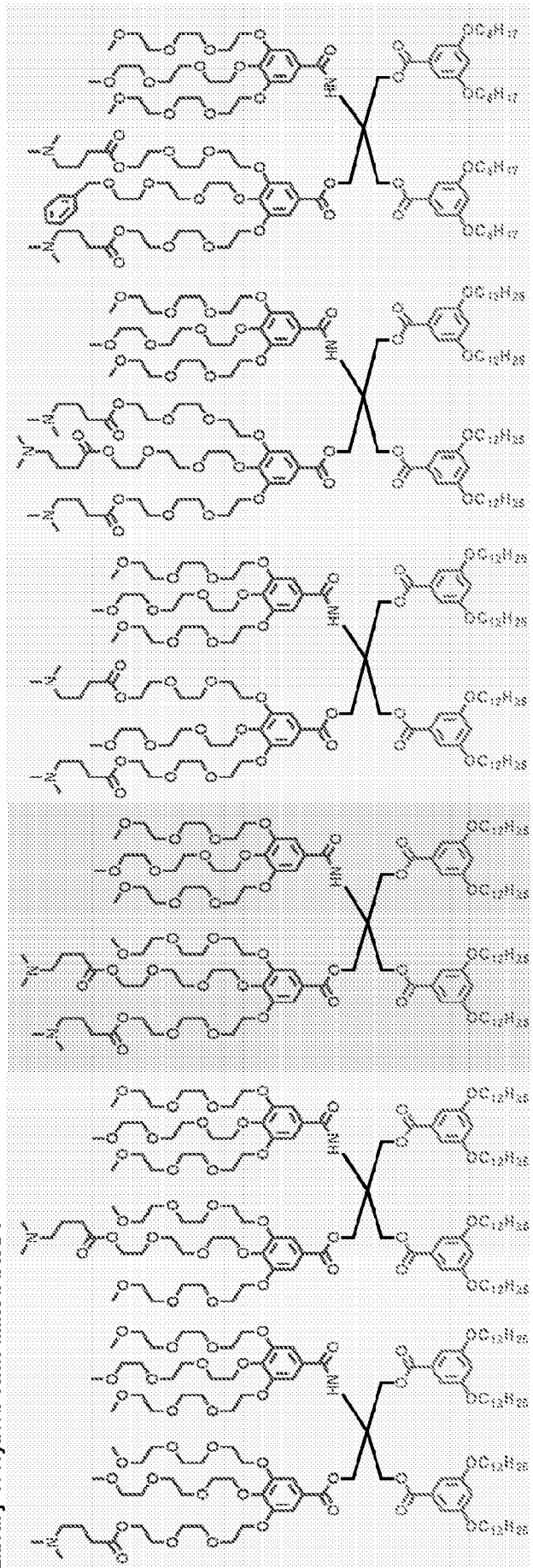

Library 6: Hybrid Twin-Mixed IAJDs
IAJD/pKa
38/6.32
6/1DMBA1
39/6.24
6/1DMBA2
40/6.36
6/2DMBA1,2
41/6.36
6/2DMBA1,3
42/6.06
6/3DMBA1,2,3
49/6.39
6/2DMBA1,3Bn2-C8

Figure 14

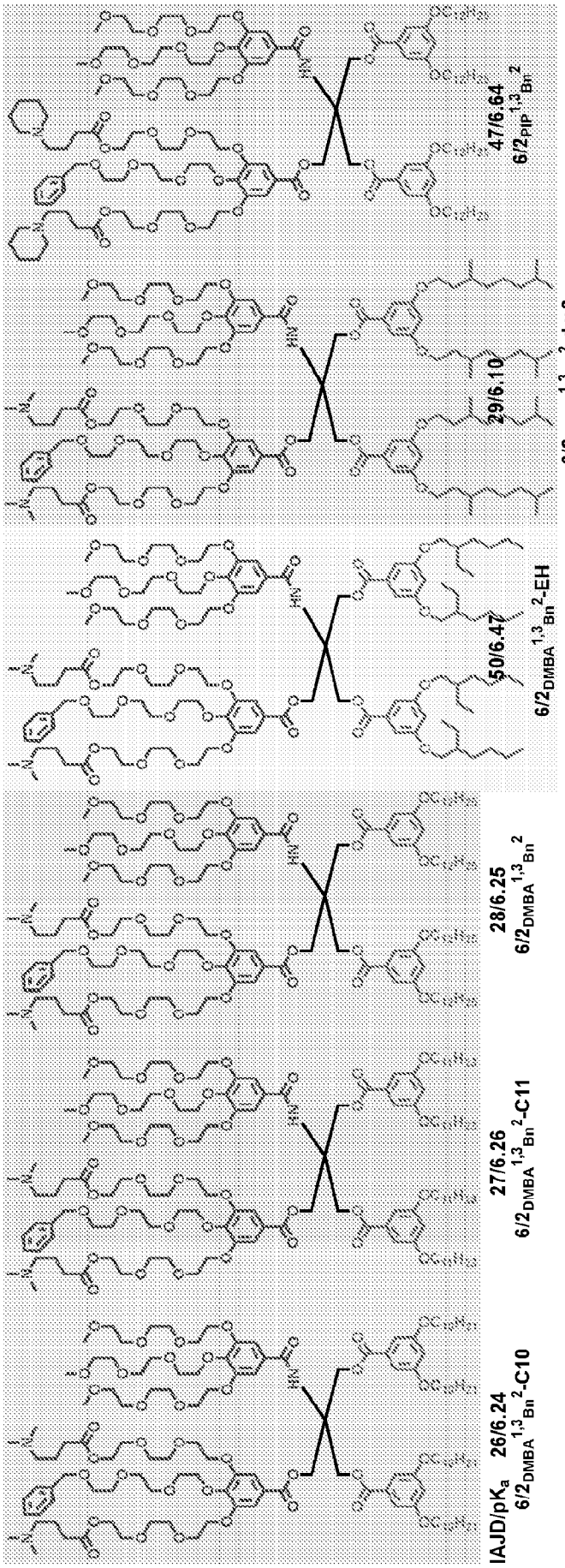


Figure 14 (cont.)

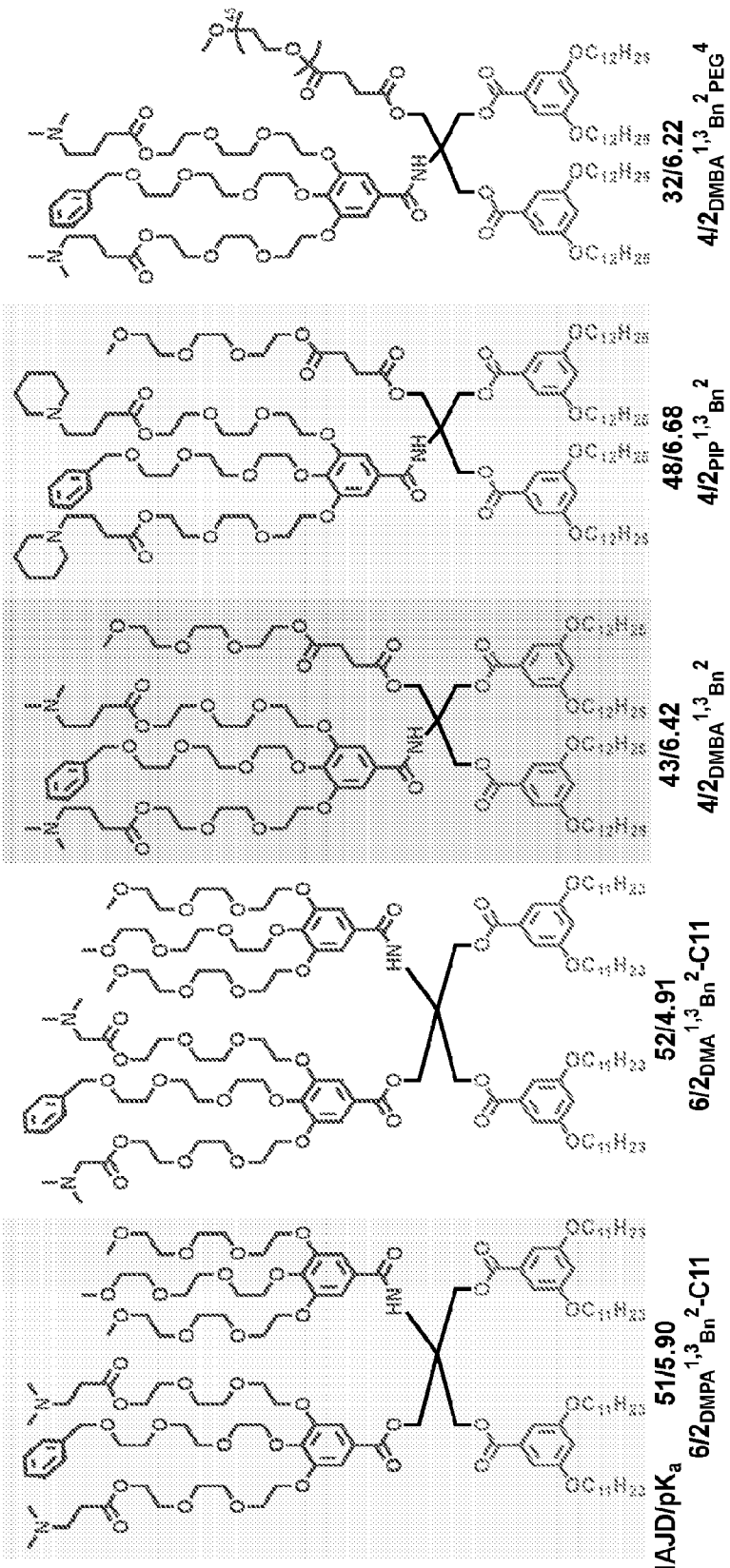


Figure 14 (cont.)

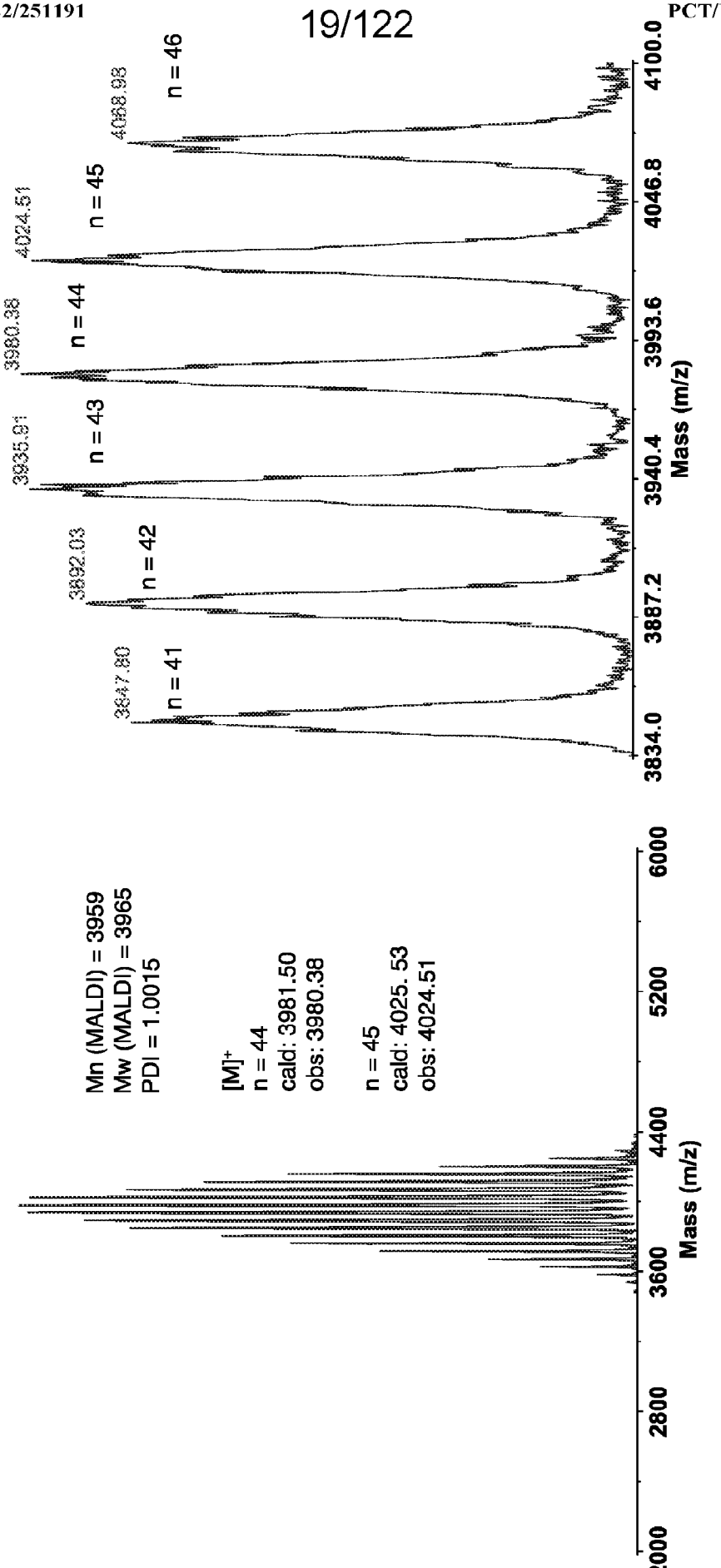

Mn (MALDI) = 3959
Mw (MALDI) = 3965
PDI = 1.0015
[M]+
n = 44
cald: 3981.50
obs: 3980.38
n = 45
cald: 4025. 53
obs: 4024.51
Mass (m/z)
2000
2800
3600
4400
5200
6000
3847.80
n = 41
3892.03
n = 42
3935.91
n = 43
3980.38
n = 44
4024.51
n = 45
4068.98
n = 46
3834.0
3887.2
3940.4
3993.6
4046.8
4100.0
Mass (m/z)

Figure 15

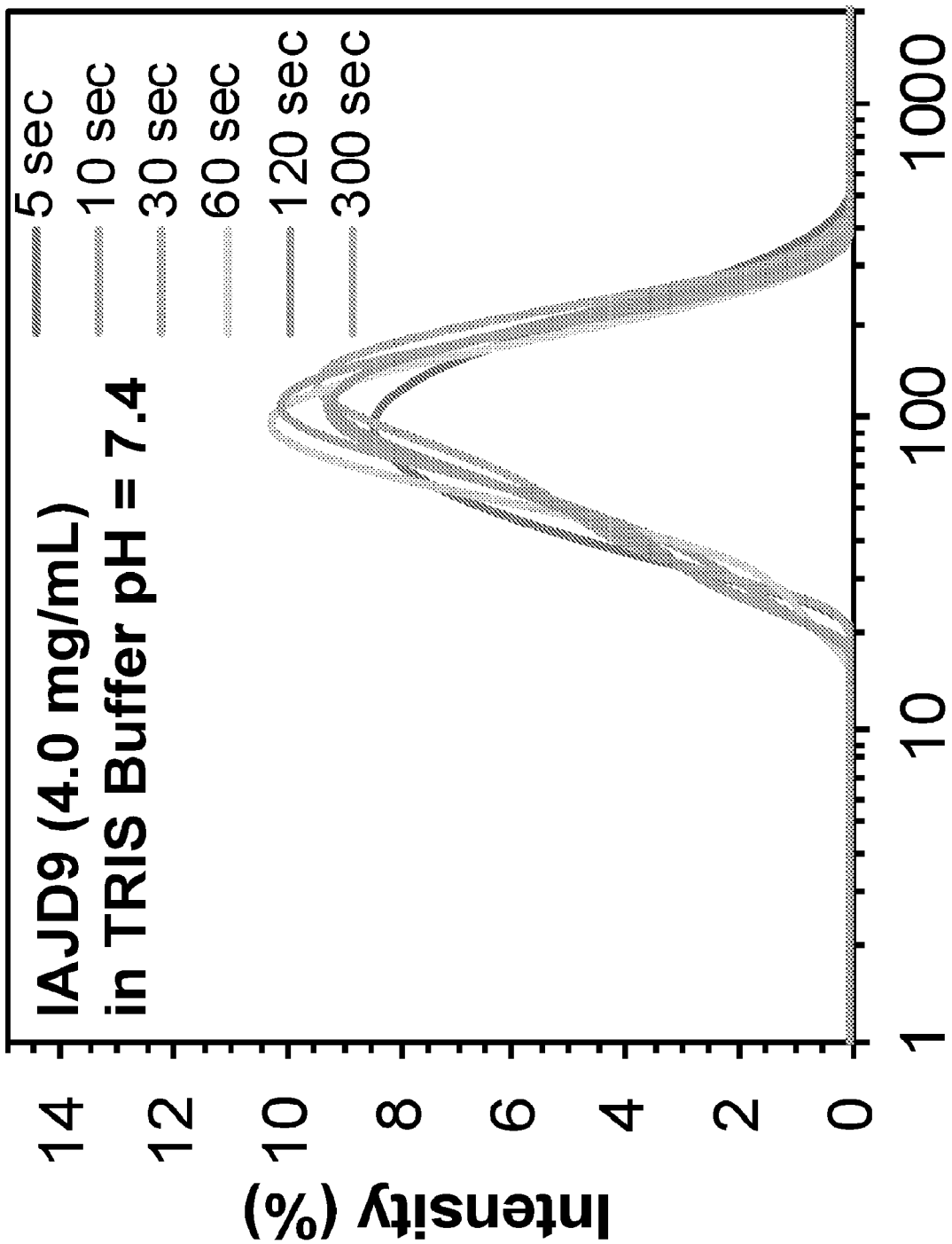

IAJD9 (4.0 mg/mL)
in TRIS Buffer pH = 7.4
5 sec
10 sec
30 sec
60 sec
120 sec
300 sec
Intensity (%)
0
2
4
6
8
10
12
14
1
10
100
1000
Size (nm)

Figure 16

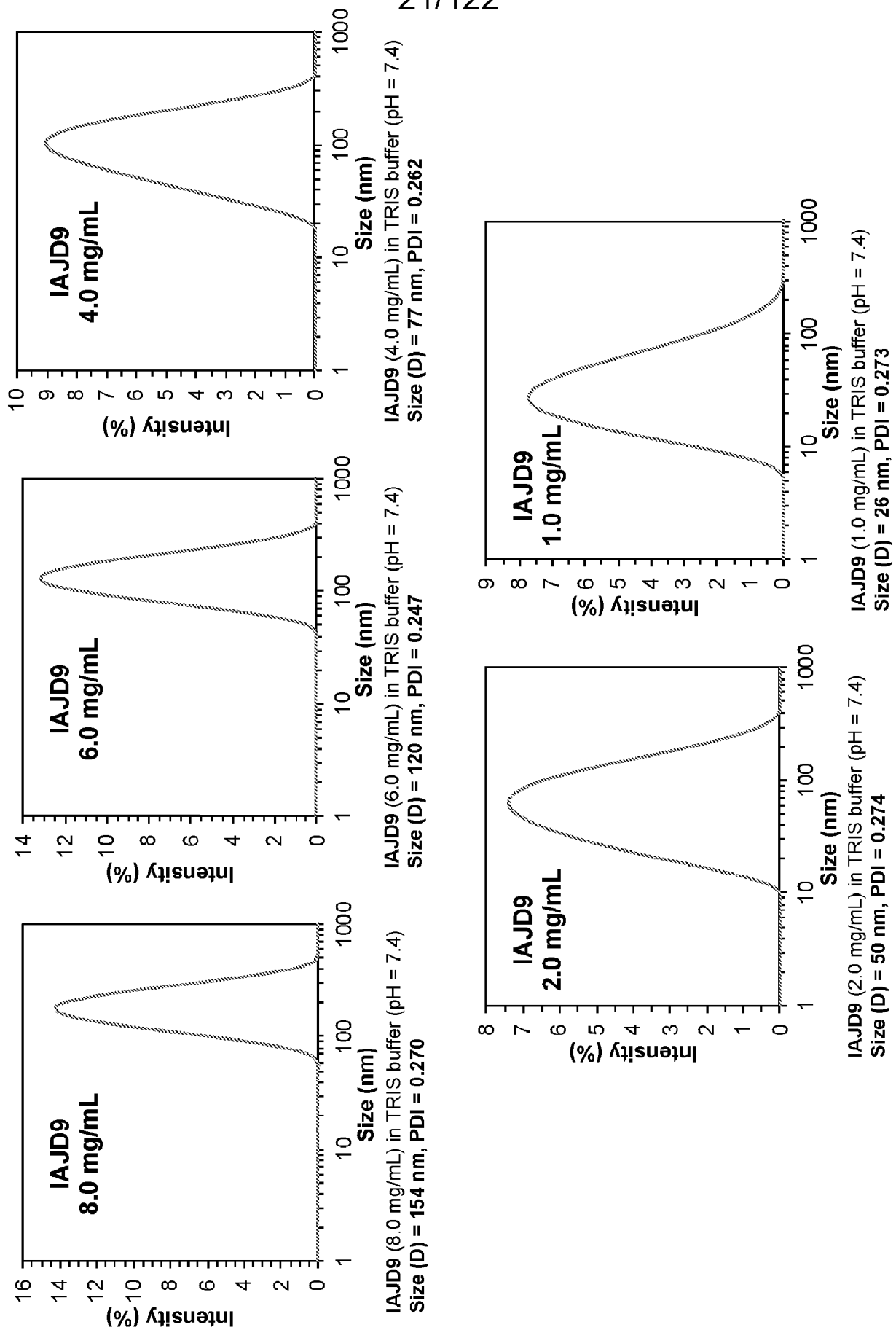
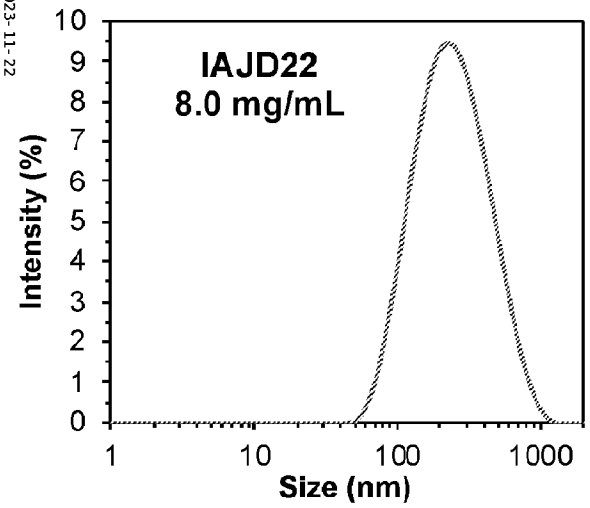
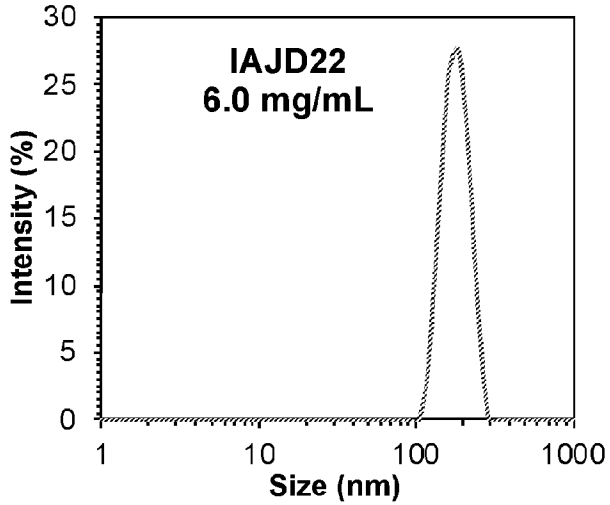

IAJD9
8.0 mg/mL
Intensity (%)
Size (nm)
IAJD9 (8.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 154 nm, PDI = 0.270
IAJD9
6.0 mg/mL
Intensity (%)
Size (nm)
IAJD9 (6.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 120 nm, PDI = 0.247
IAJD9
4.0 mg/mL
Intensity (%)
Size (nm)
IAJD9 (4.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 77 nm, PDI = 0.262
IAJD9
2.0 mg/mL
Intensity (%)
Size (nm)
IAJD9 (2.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 50 nm, PDI = 0.274
IAJD9
1.0 mg/mL
Intensity (%)
Size (nm)
IAJD9 (1.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 26 nm, PDI = 0.273

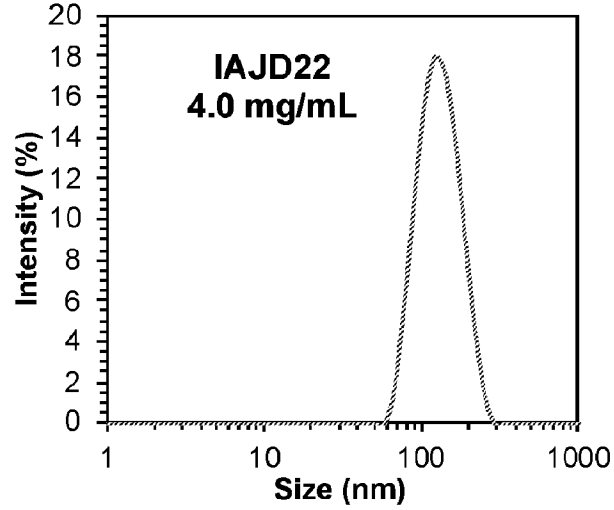
Figure 17



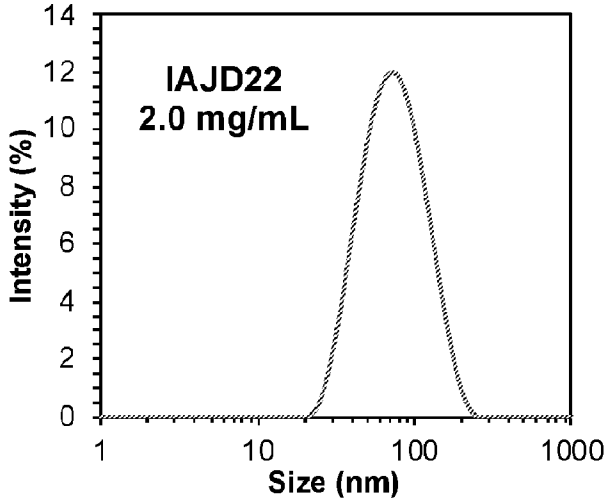
IAJD22 (8.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 202 nm, PDI = 0.22



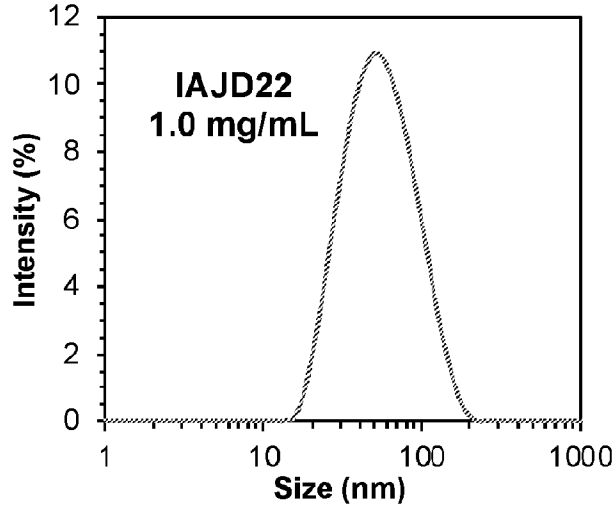
IAJD22 (6.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 183 nm, PDI = 0.103



IAJD22 (4.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 123 nm, PDI = 0.081

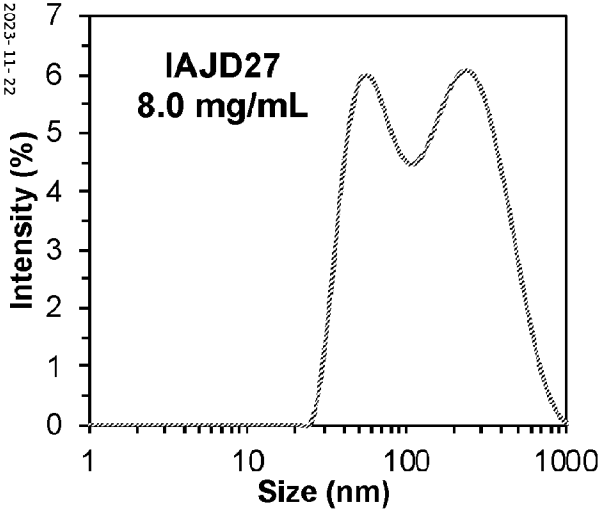


IAJD22 (2.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 67 nm, PDI = 0.157

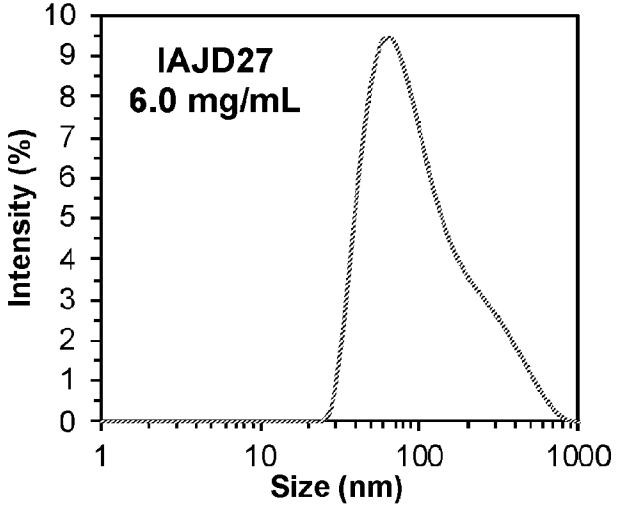


IAJD22 (1.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 49 nm, PDI = 0.204

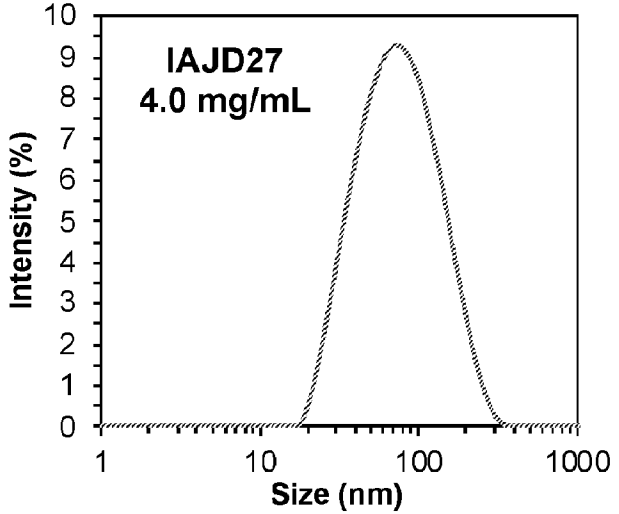
Figure 18



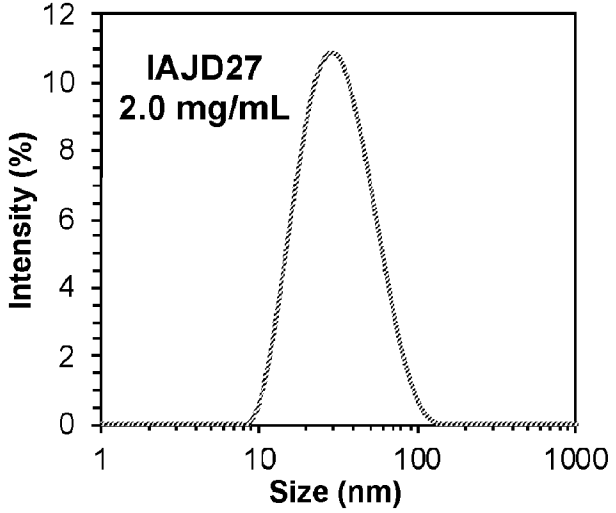
IAJD27 (8.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 103 nm, PDI = 0.401



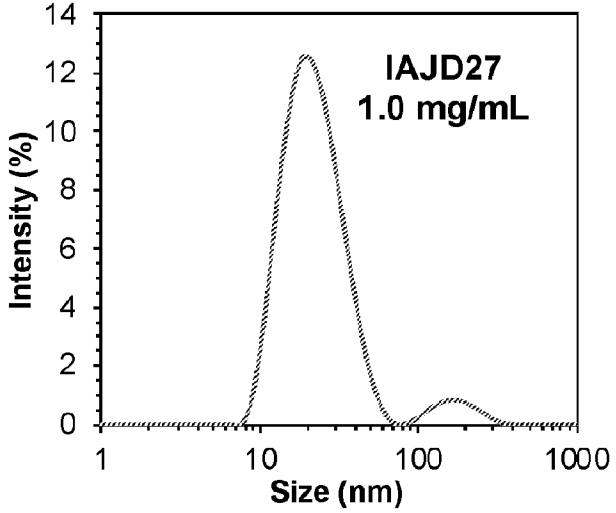
IAJD27 (6.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 81 nm, PDI = 0.252



IAJD27 (4.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 64 nm, PDI = 0.223



IAJD27 (2.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 28 nm, PDI = 0.187



IAJD27 (1.0 mg/mL) in TRIS buffer (pH = 7.4)
Size (D) = 23 nm, PDI = 0.335

Figure 19

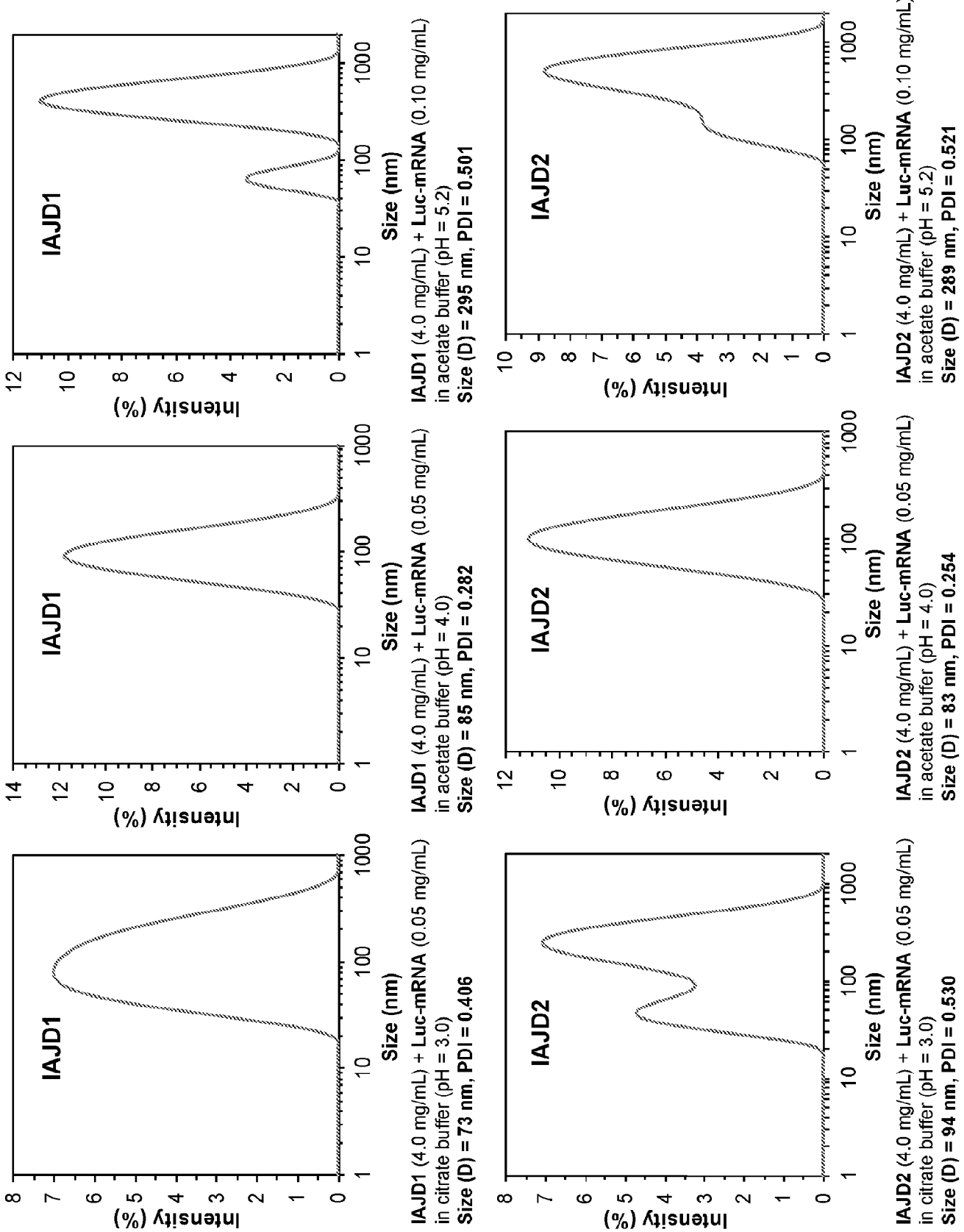

IAJD1
Intensity (%)
Size (nm)
IAJD1 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 73 nm, PDI = 0.406
IAJD1 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 85 nm, PDI = 0.282
IAJD1 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 295 nm, PDI = 0.501
IAJD2
Intensity (%)
Size (nm)
IAJD2 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 94 nm, PDI = 0.530
IAJD2 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 83 nm, PDI = 0.254
IAJD2 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 289 nm, PDI = 0.521

Figure 20

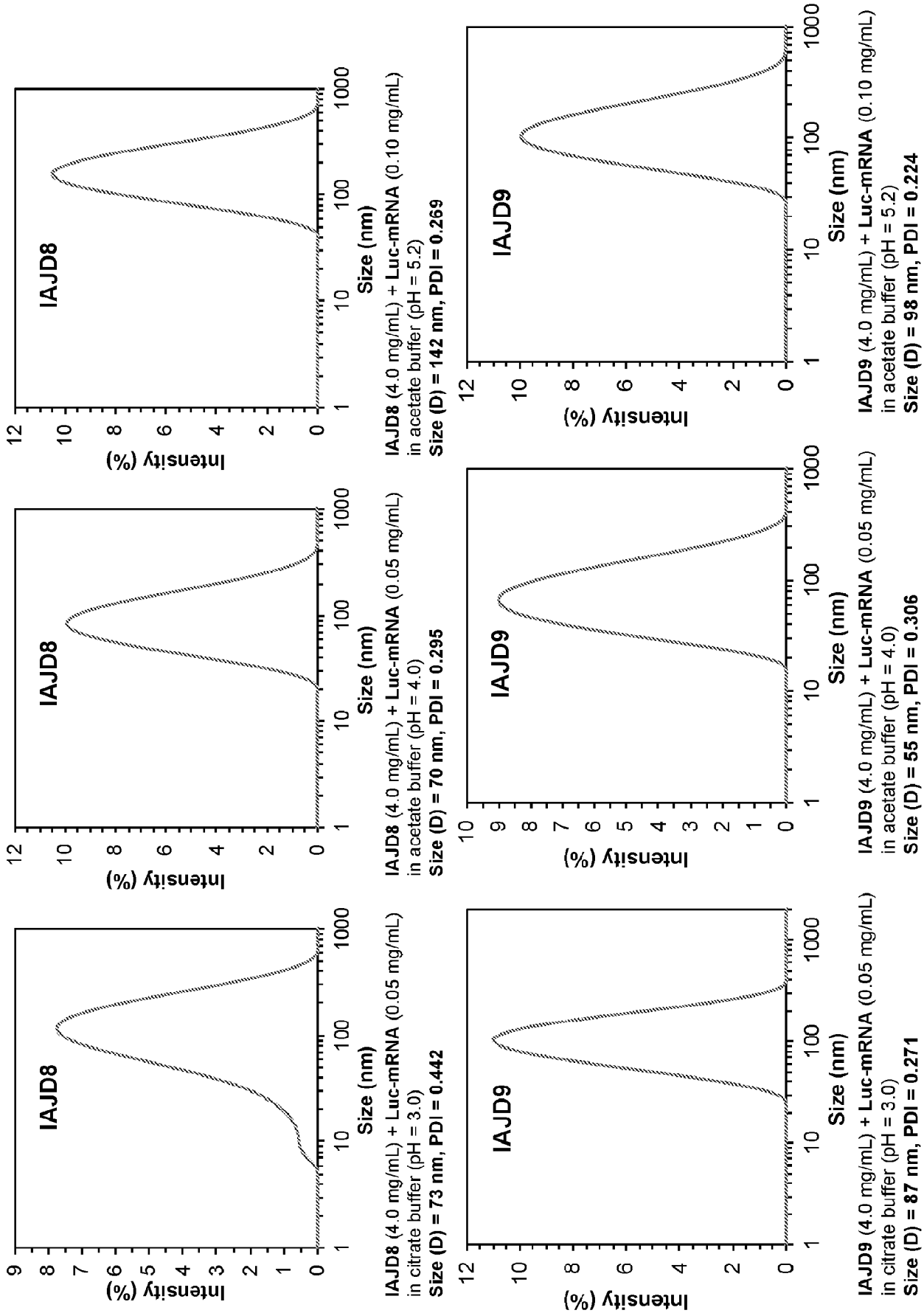
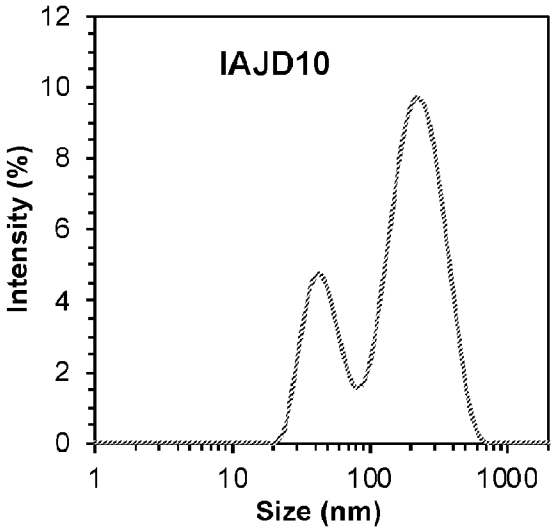
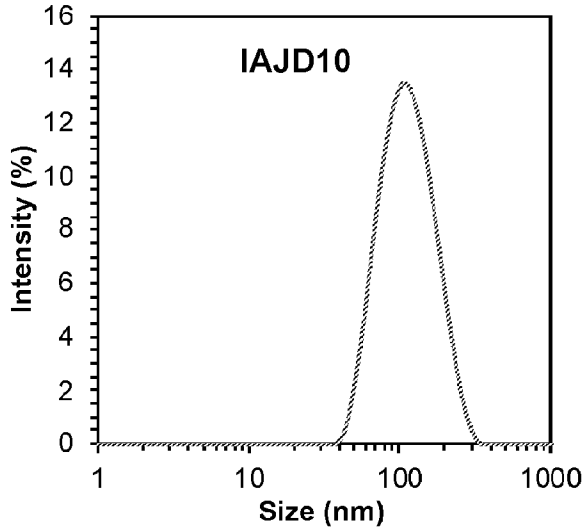

IAJD8
Intensity (%)
Size (nm)
IAJD8 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 73 nm, PDI = 0.442
IAJD8 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 70 nm, PDI = 0.295
IAJD8 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 142 nm, PDI = 0.269
IAJD9
IAJD9 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 87 nm, PDI = 0.271
IAJD9 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 55 nm, PDI = 0.306
IAJD9 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 98 nm, PDI = 0.224

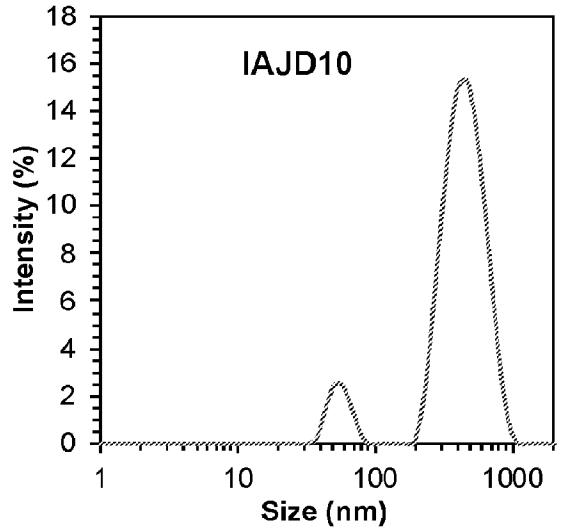
Figure 21



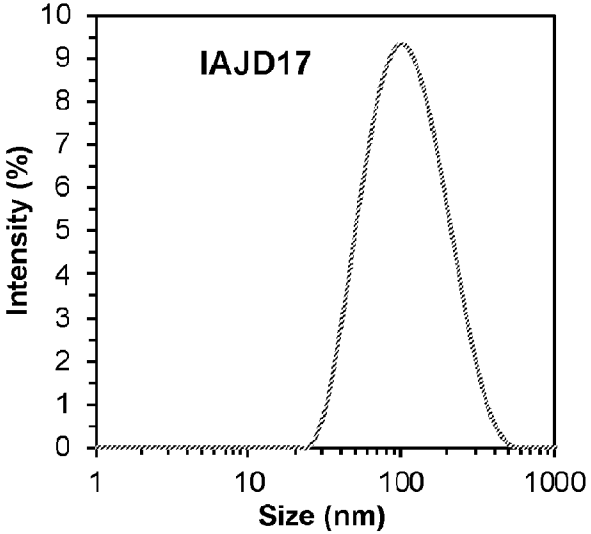
IAJD10 (4.0 mg/mL) + **Luc-mRNA** (0.05 mg/mL)
in citrate buffer (pH = 3.0)
Size (D) = 110 nm, PDI = 0.467



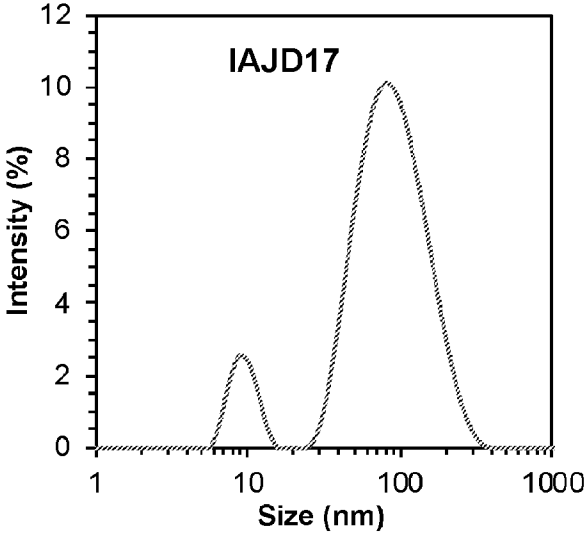
IAJD10 (4.0 mg/mL) + **Luc-mRNA** (0.05 mg/mL)
in acetate buffer (pH = 4.0)
Size (D) = 92 nm, PDI = 0.242



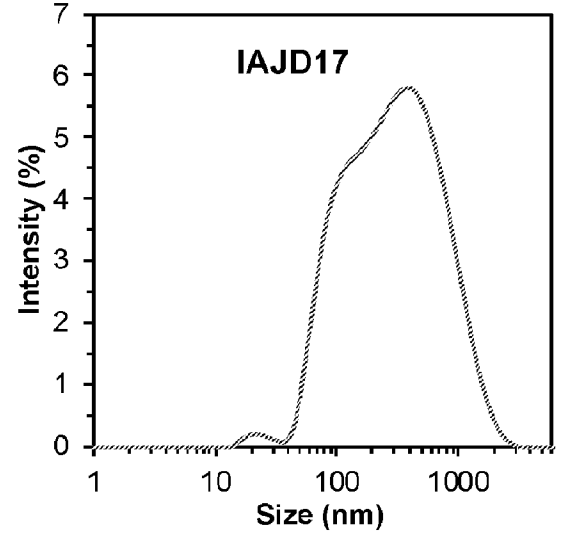
IAJD10 (4.0 mg/mL) + **Luc-mRNA** (0.10 mg/mL)
in acetate buffer (pH = 5.2)
Size (D) = 347 nm, PDI = 0.449



IAJD17 (4.0 mg/mL) + **Luc-mRNA** (0.05 mg/mL)
in citrate buffer (pH = 3.0)
Size (D) = 82 nm, PDI = 0.398



IAJD17 (4.0 mg/mL) + **Luc-mRNA** (0.05 mg/mL)
in acetate buffer (pH = 4.0)
Size (D) = 57 nm, PDI = 0.472



IAJD17 (4.0 mg/mL) + **Luc-mRNA** (0.10 mg/mL)
in acetate buffer (pH = 5.2)
Size (D) = 203 nm, PDI = 0.449

Figure 22

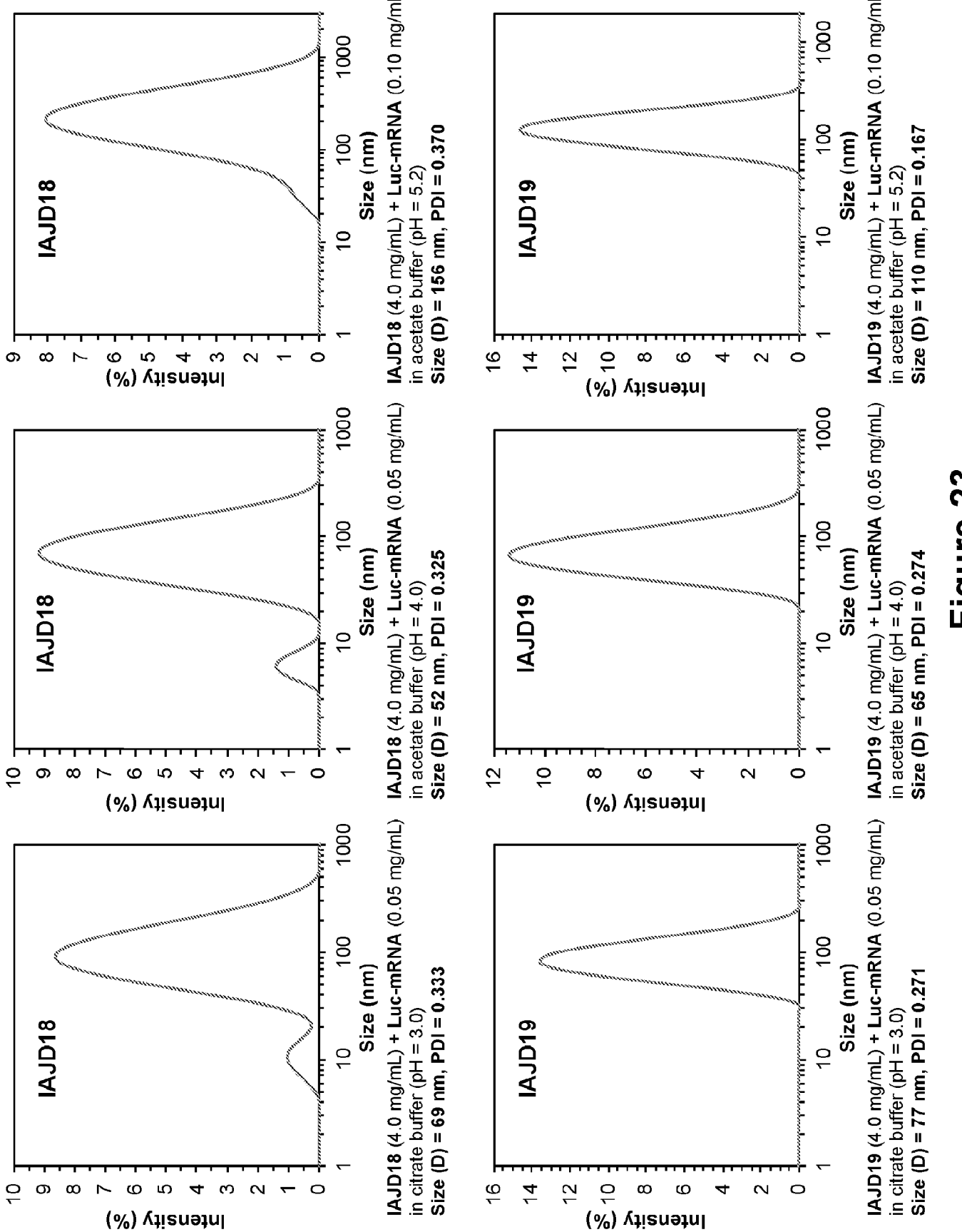

IAJD18
Intensity (%)
Size (nm)
IAJD18 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 69 nm, PDI = 0.333
IAJD18 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 52 nm, PDI = 0.325
IAJD18 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 156 nm, PDI = 0.370
IAJD19
IAJD19 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 77 nm, PDI = 0.271
IAJD19 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 65 nm, PDI = 0.274
IAJD19 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 110 nm, PDI = 0.167

Figure 23

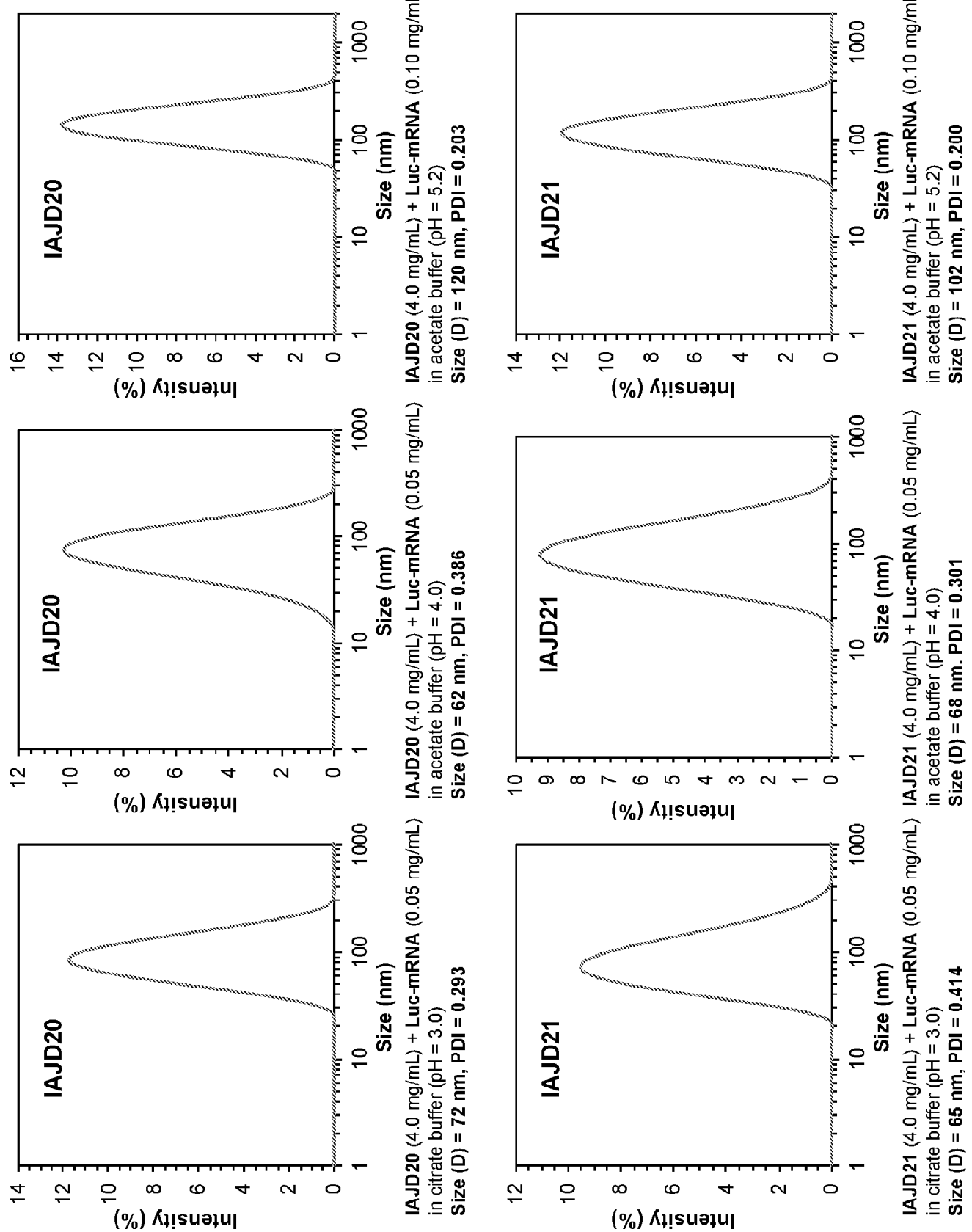

IAJD20
Intensity (%)
Size (nm)
IAJD20 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 120 nm, PDI = 0.203
IAJD20
Intensity (%)
Size (nm)
IAJD20 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 62 nm, PDI = 0.386
IAJD20
Intensity (%)
Size (nm)
IAJD20 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 72 nm, PDI = 0.293
IAJD21
Intensity (%)
Size (nm)
IAJD21 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 102 nm, PDI = 0.200
IAJD21
Intensity (%)
Size (nm)
IAJD21 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 68 nm. PDI = 0.301
IAJD21
Intensity (%)
Size (nm)
IAJD21 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 65 nm, PDI = 0.414

Figure 24

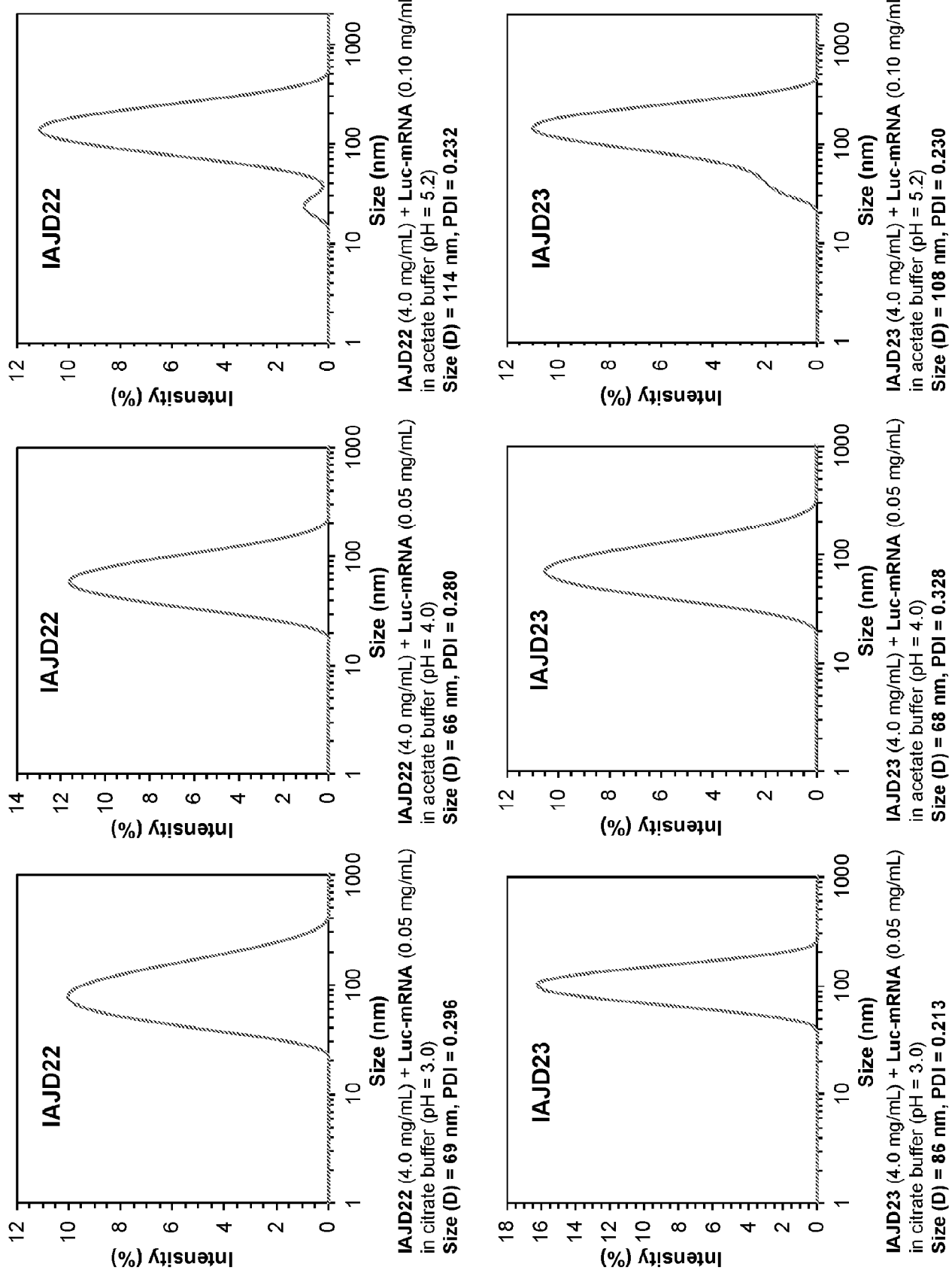

IAJD22
Intensity (%)
Size (nm)
IAJD22 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 69 nm, PDI = 0.296
IAJD22 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 66 nm, PDI = 0.280
IAJD22 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 114 nm, PDI = 0.232
IAJD23
IAJD23 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 86 nm, PDI = 0.213
IAJD23 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 68 nm, PDI = 0.328
IAJD23 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 108 nm, PDI = 0.230

Figure 25

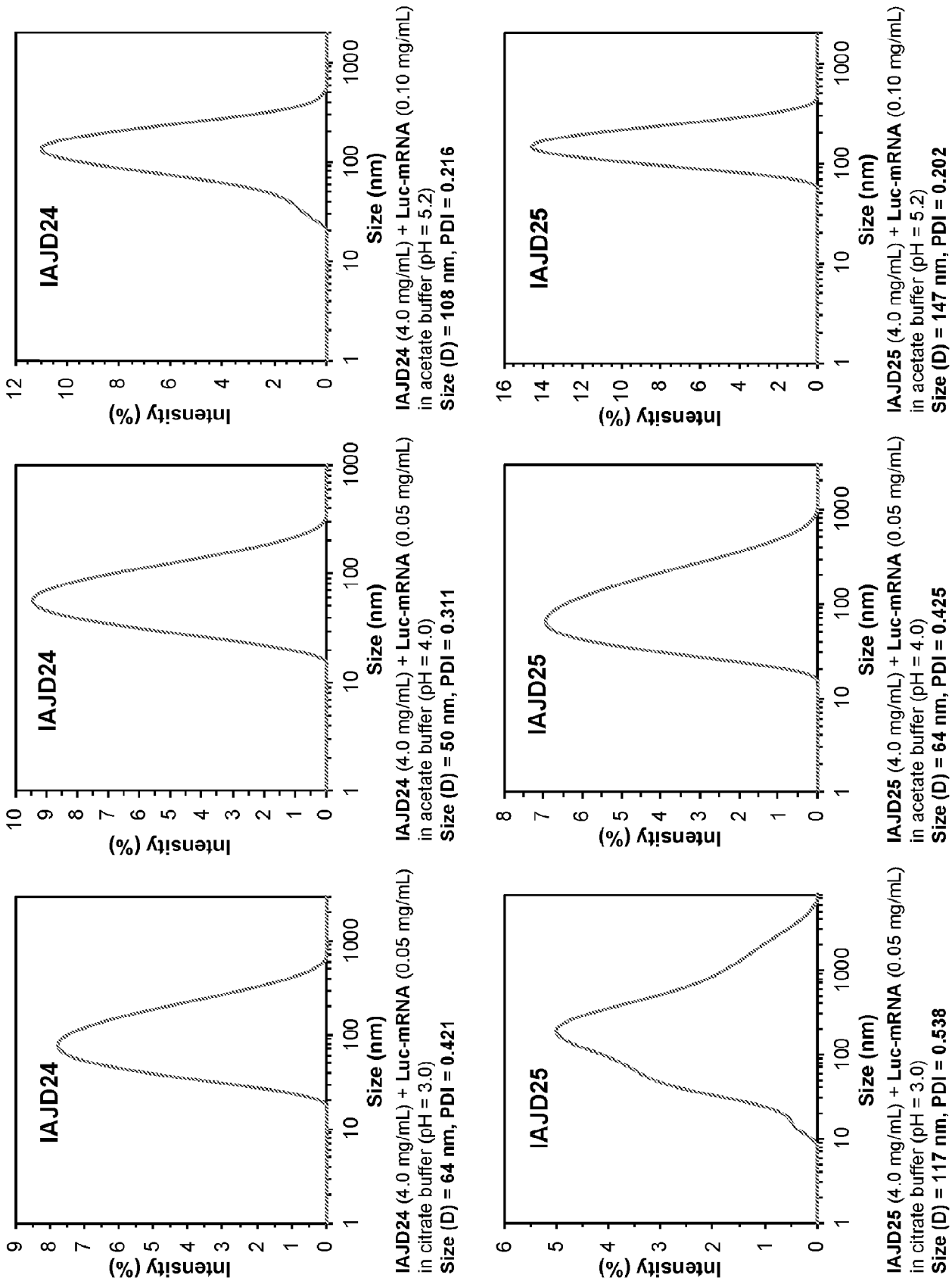

IAJD24
Intensity (%)
Size (nm)
IAJD24 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 64 nm, PDI = 0.421
IAJD24 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 50 nm, PDI = 0.311
IAJD24 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 108 nm, PDI = 0.216
IAJD25
IAJD25 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 117 nm, PDI = 0.538
IAJD25 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 64 nm, PDI = 0.425
IAJD25 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 147 nm, PDI = 0.202

Figure 26

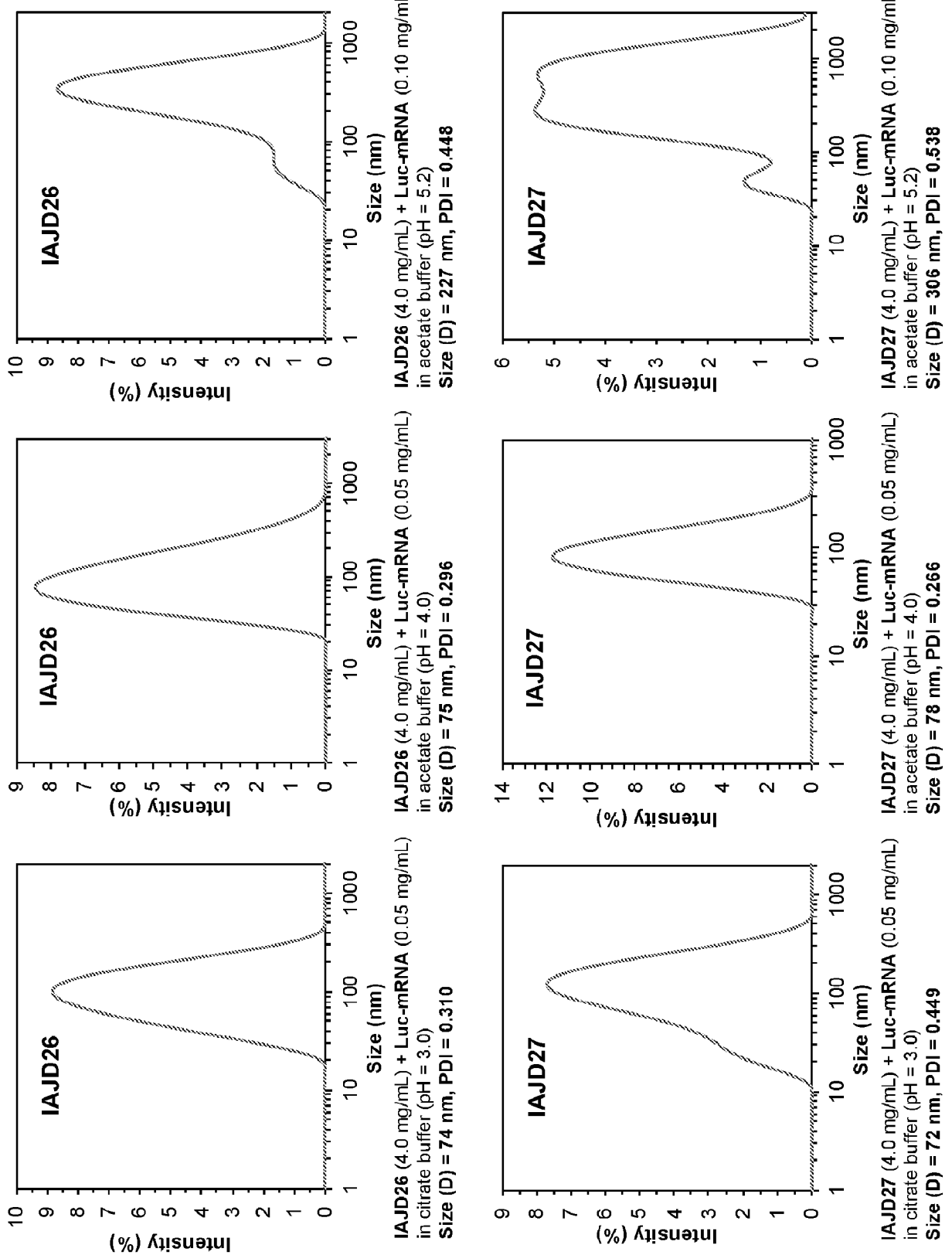

IAJD26
Intensity (%)
Size (nm)
IAJD26 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 74 nm, PDI = 0.310
IAJD26 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 75 nm, PDI = 0.296
IAJD26 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 227 nm, PDI = 0.448
IAJD27
Intensity (%)
Size (nm)
IAJD27 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 72 nm, PDI = 0.449
IAJD27 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 78 nm, PDI = 0.266
IAJD27 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 306 nm, PDI = 0.538

Figure 27

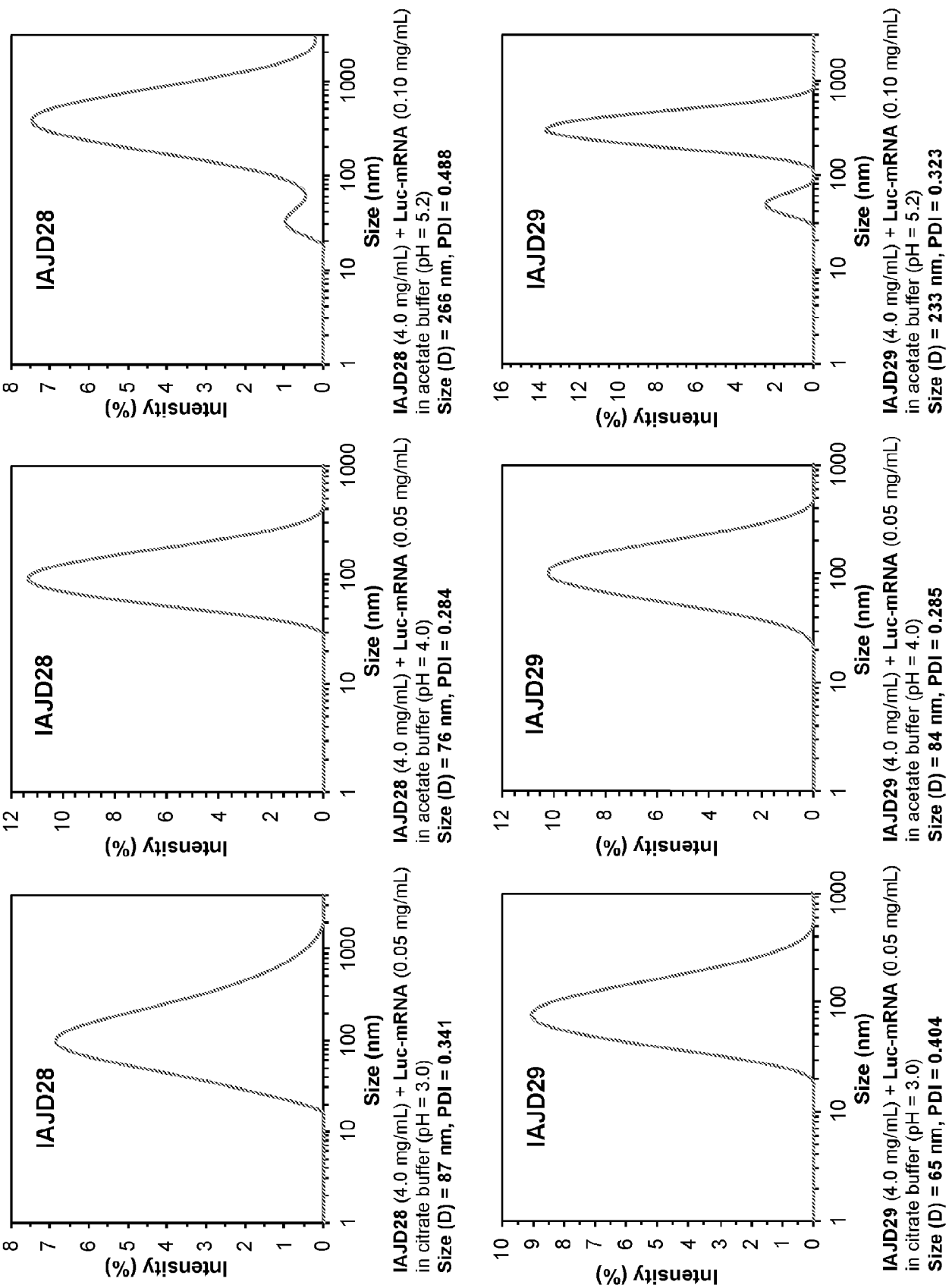

IAJD28
Intensity (%)
Size (nm)
IAJD28 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 87 nm, PDI = 0.341
IAJD28 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 76 nm, PDI = 0.284
IAJD28 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 266 nm, PDI = 0.488
IAJD29
IAJD29 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 65 nm, PDI = 0.404
IAJD29 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 84 nm, PDI = 0.285
IAJD29 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 233 nm, PDI = 0.323

Figure 28

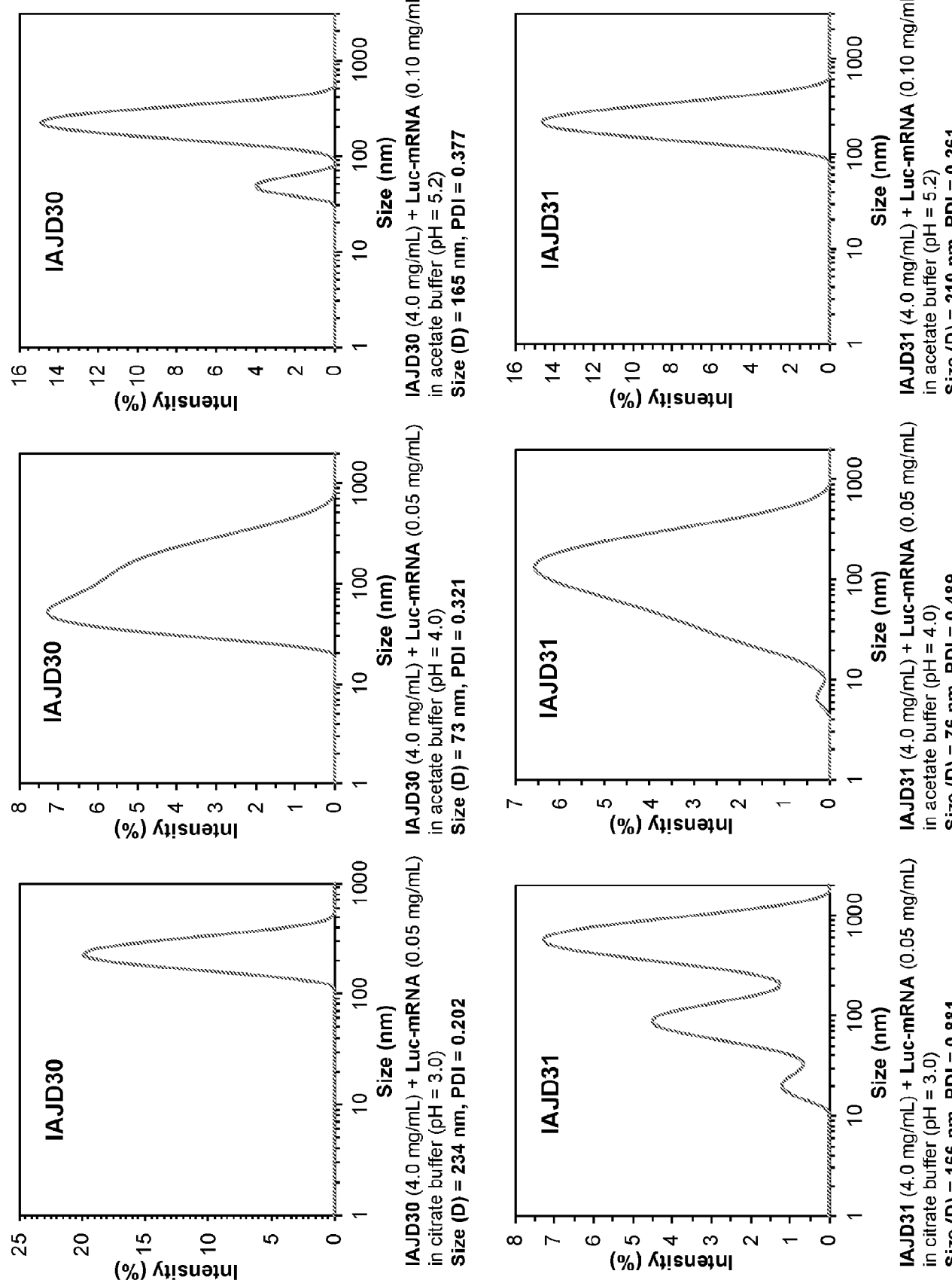

IAJD30
Intensity (%)
Size (nm)
IAJD30 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 234 nm, PDI = 0.202
IAJD30 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 73 nm, PDI = 0.321
IAJD30 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 165 nm, PDI = 0.377
IAJD31
IAJD31 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 166 nm, PDI = 0.881
IAJD31 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 76 nm, PDI = 0.489
IAJD31 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 210 nm, PDI = 0.261

Figure 29

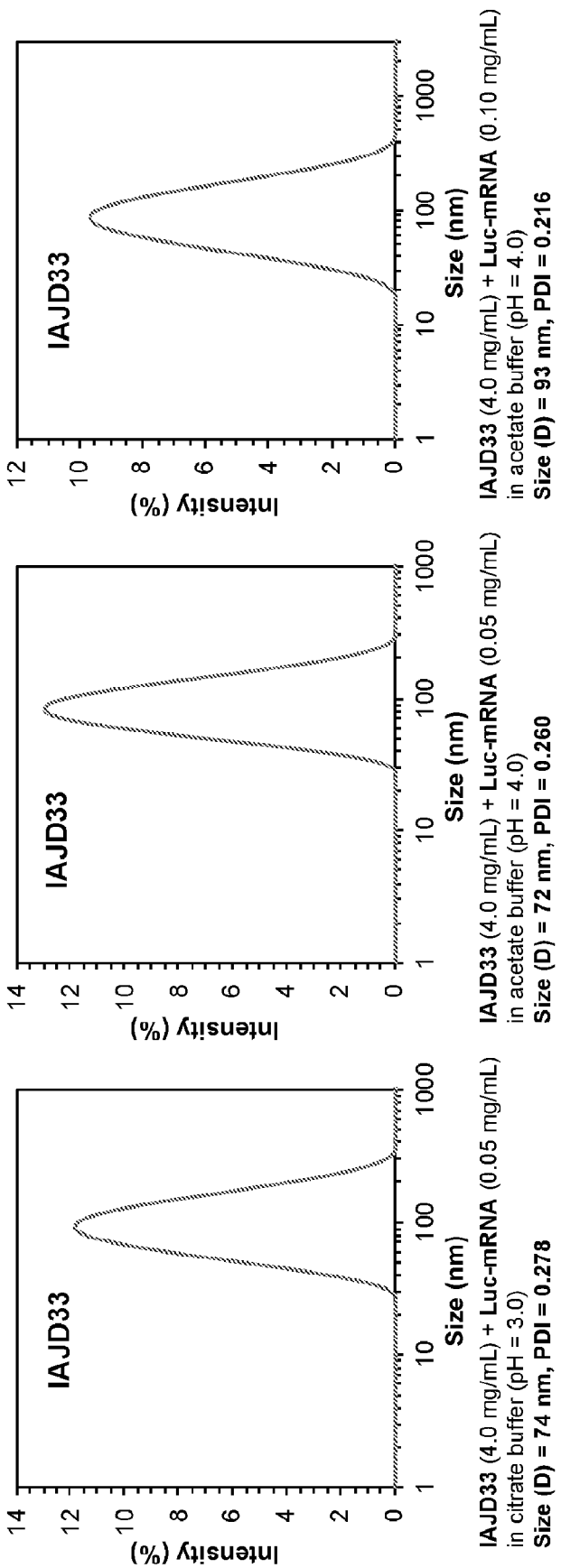

IAJD33
Intensity (%)
Size (nm)
IAJD33 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 74 nm, PDI = 0.278
IAJD33
Intensity (%)
Size (nm)
IAJD33 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 72 nm, PDI = 0.260
IAJD33
Intensity (%)
Size (nm)
IAJD33 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 93 nm, PDI = 0.216
IAJD34
Intensity (%)
Size (nm)
IAJD34 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 83 nm, PDI = 0.264
IAJD34
Intensity (%)
Size (nm)
IAJD34 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 56 nm, PDI = 0.255
IAJD34
Intensity (%)
Size (nm)
IAJD34 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 83 nm, PDI = 0.245

Figure 30

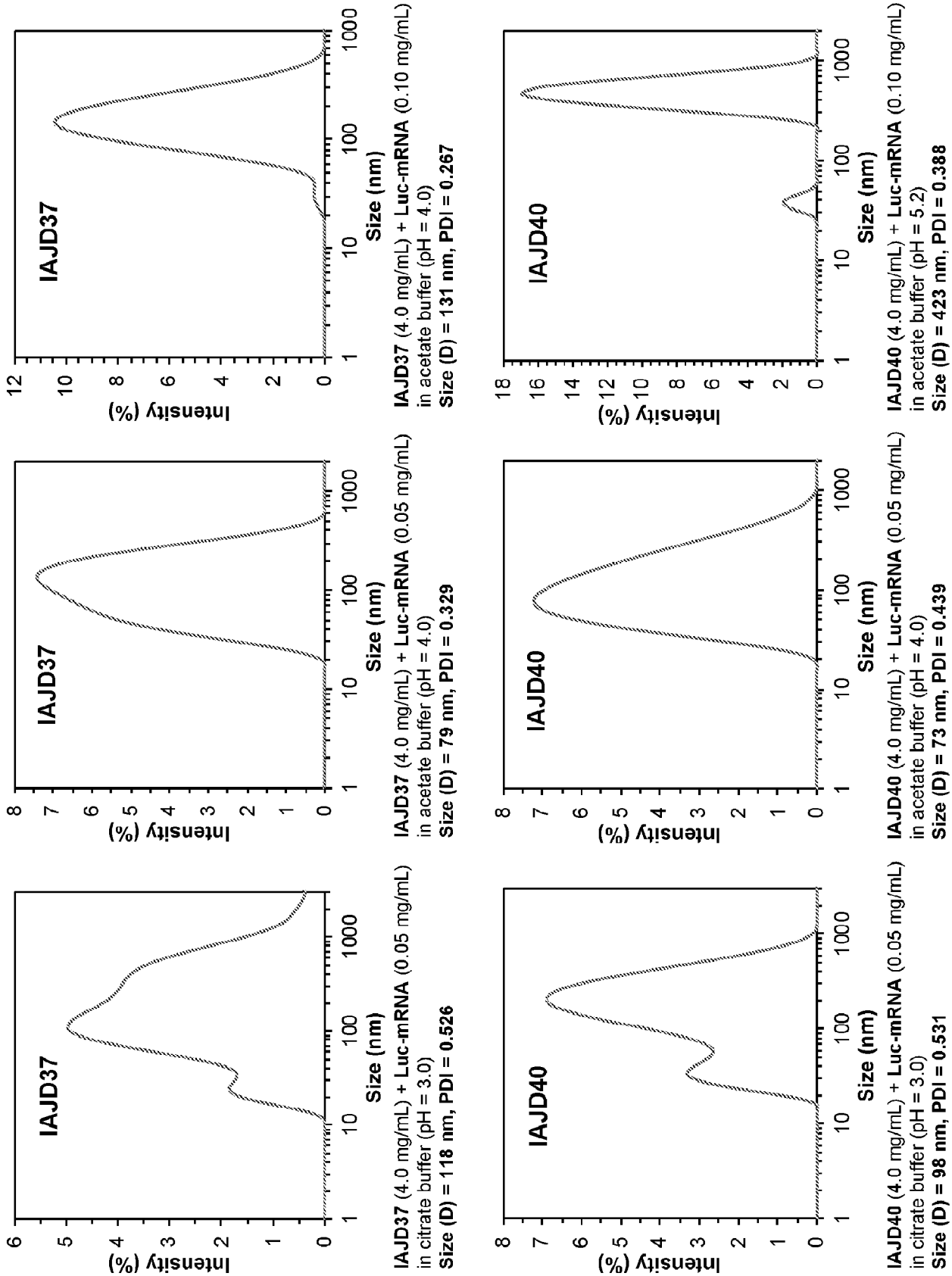

IAJD37
Intensity (%)
Size (nm)
IAJD37 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 118 nm, PDI = 0.526
IAJD37
Intensity (%)
Size (nm)
IAJD37 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 79 nm, PDI = 0.329
IAJD37
Intensity (%)
Size (nm)
IAJD37 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 131 nm, PDI = 0.267
IAJD40
Intensity (%)
Size (nm)
IAJD40 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 98 nm, PDI = 0.531
IAJD40
Intensity (%)
Size (nm)
IAJD40 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 73 nm, PDI = 0.439
IAJD40
Intensity (%)
Size (nm)
IAJD40 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 423 nm, PDI = 0.388

Figure 31

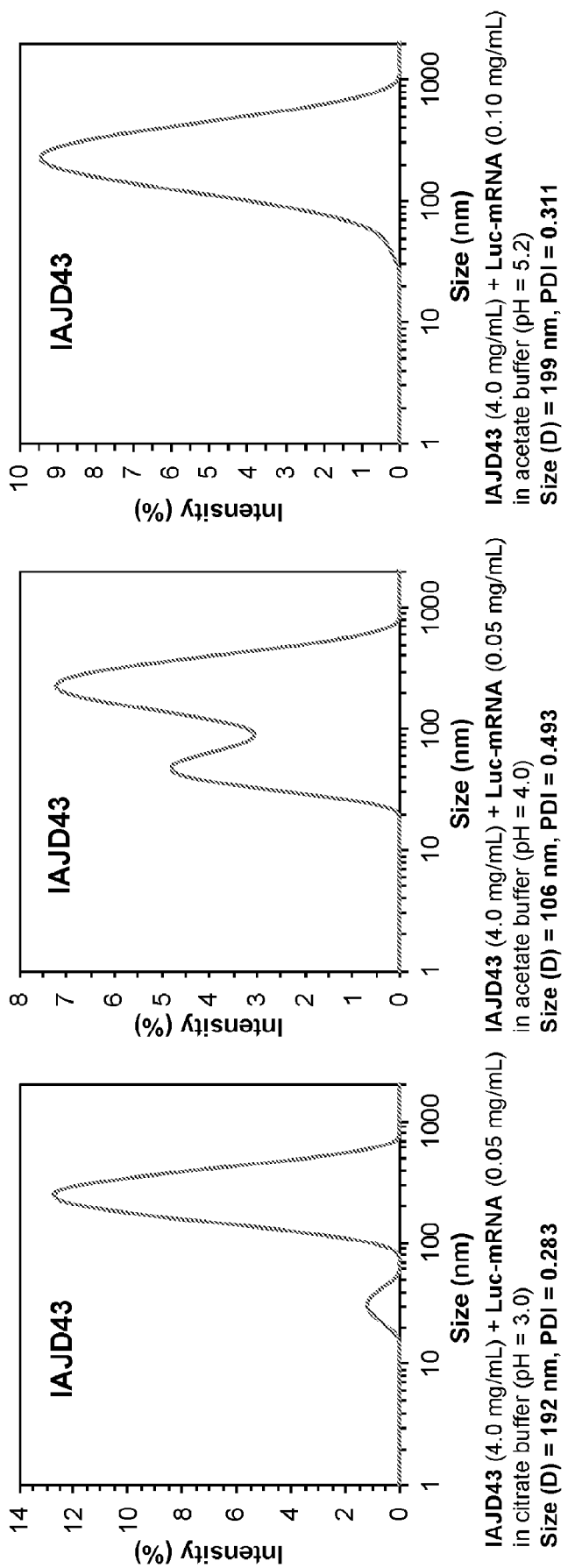

IAJD43
Intensity (%)
Size (nm)
IAJD43 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0) Size (D) = 192 nm, PDI = 0.283
IAJD43 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0) Size (D) = 106 nm, PDI = 0.493
IAJD43 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2) Size (D) = 199 nm, PDI = 0.311
IAJD44
Intensity (%)
Size (nm)
IAJD44 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0) Size (D) = 85 nm, PDI = 0.310
IAJD44 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0) Size (D) = 98 nm. PDI = 0.279
IAJD44 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2) Size (D) = 171 nm, PDI = 0.252

Figure 32

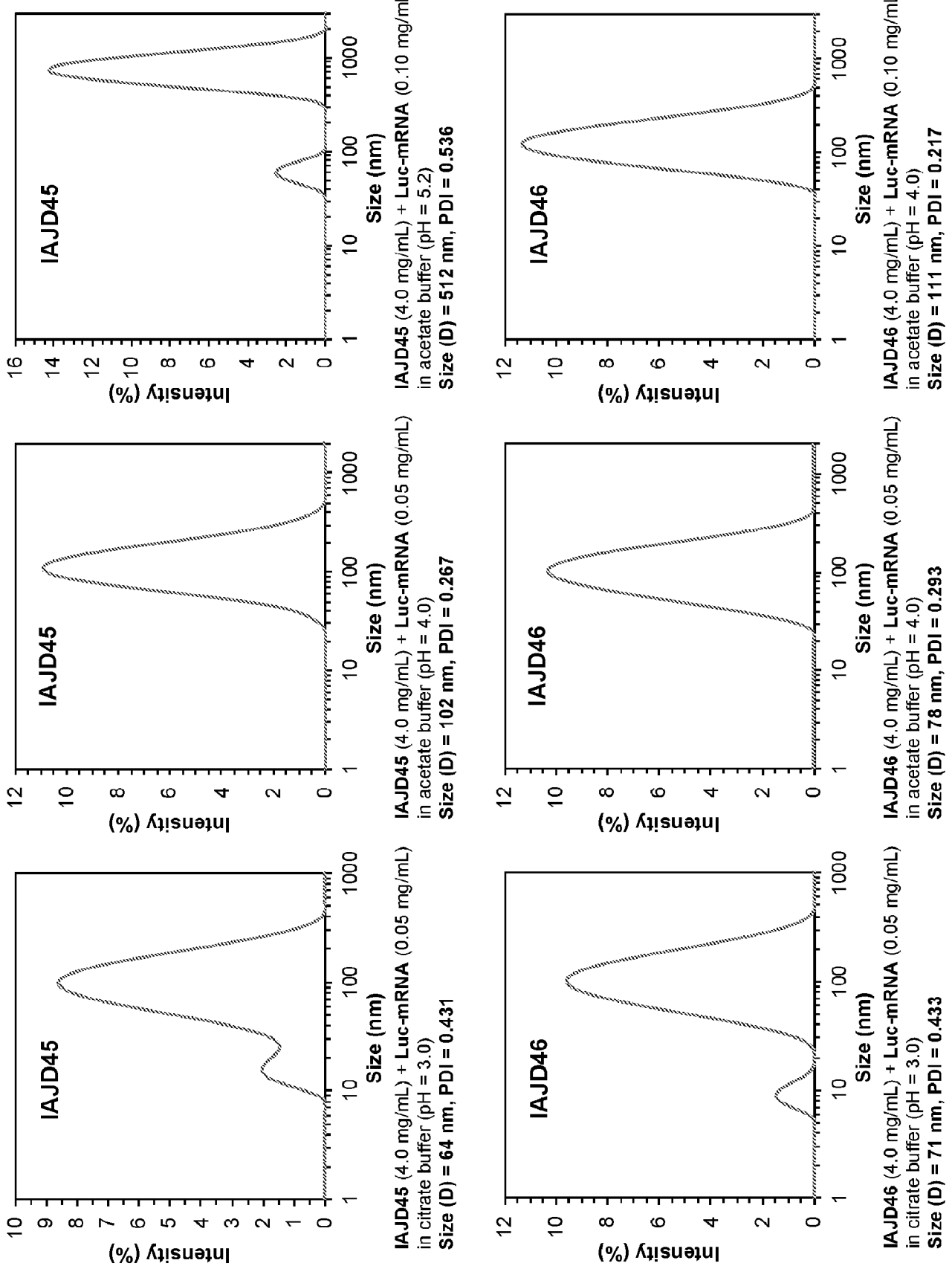

IAJD45
Intensity (%)
Size (nm)
IAJD45 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 64 nm, PDI = 0.431
IAJD45 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 102 nm, PDI = 0.267
IAJD45 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 5.2)
Size (D) = 512 nm, PDI = 0.536
IAJD46
IAJD46 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 71 nm, PDI = 0.433
IAJD46 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 78 nm, PDI = 0.293
IAJD46 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 111 nm, PDI = 0.217

Figure 33

IAJD47
Intensity (%)
Size (nm)
IAJD47 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in citrate buffer (pH = 3.0)
Size (D) = 82 nm, PDI = 0.443
IAJD47
Intensity (%)
Size (nm)
IAJD47 (4.0 mg/mL) + Luc-mRNA (0.05 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 81 nm, PDI = 0.416
IAJD47
Intensity (%)
Size (nm)
IAJD47 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 121 nm, PDI = 0.236

Figure 34

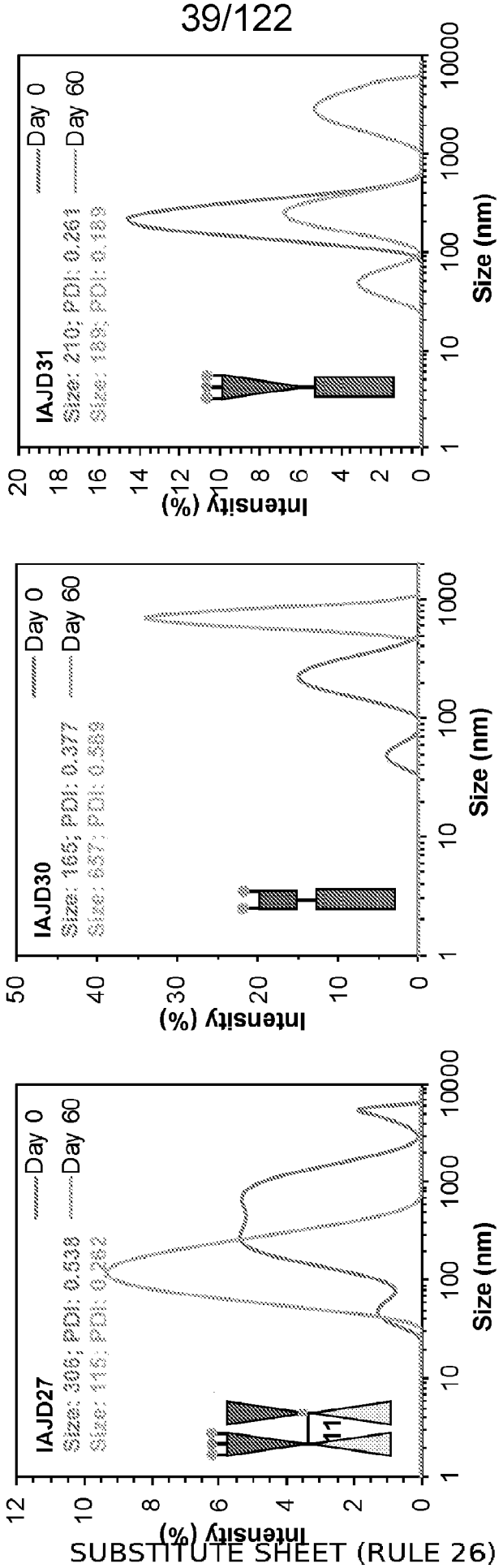

IAJD27
Size: 306; PDI: 0.338
Size: 115; PDI: 0.782
Day 0
Day 60
Intensity (%)
Size (nm)
IAJD30
Size: 165; PDI: 0.377
Size: 687; PDI: 0.589
Day 0
Day 60
Intensity (%)
Size (nm)
IAJD31
Size: 210; PDI: 0.261
Size: 185; PDI: 0.185
Day 0
Day 60
Intensity (%)
Size (nm)

Figure 35

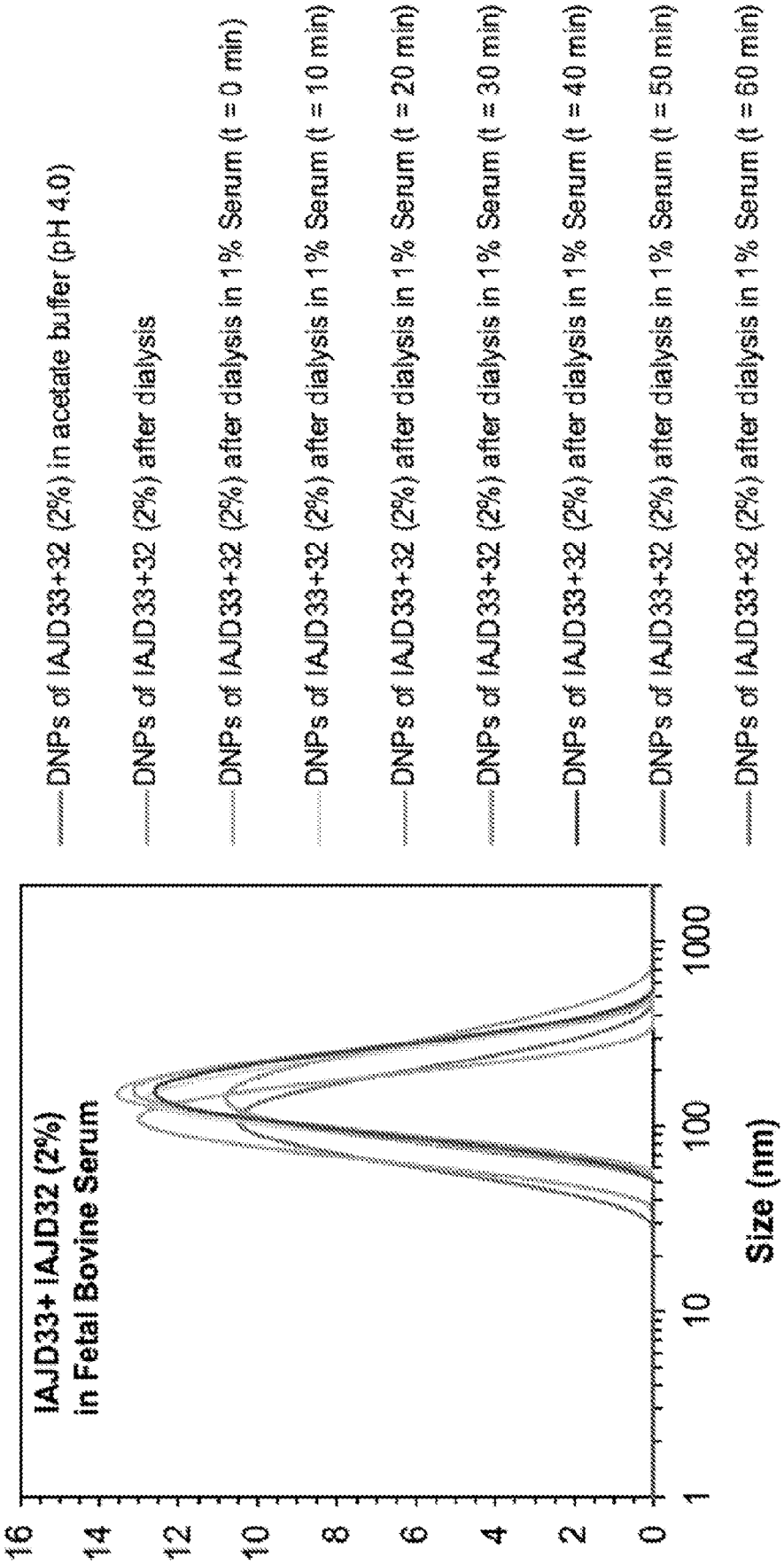

IAJD33+ IAJD32 (2%)
in Fetal Bovine Serum
Intensity (%)
Size (nm)
0
2
4
6
8
10
12
14
16
1
10
100
1000
DNPs of IAJD33+32 (2%) in acetate buffer (pH 4.0)
DNPs of IAJD33+32 (2%) after dialysis
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 0 min)
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 10 min)
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 20 min)
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 30 min)
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 40 min)
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 50 min)
DNPs of IAJD33+32 (2%) after dialysis in 1% Serum (t = 60 min)

Figure 36

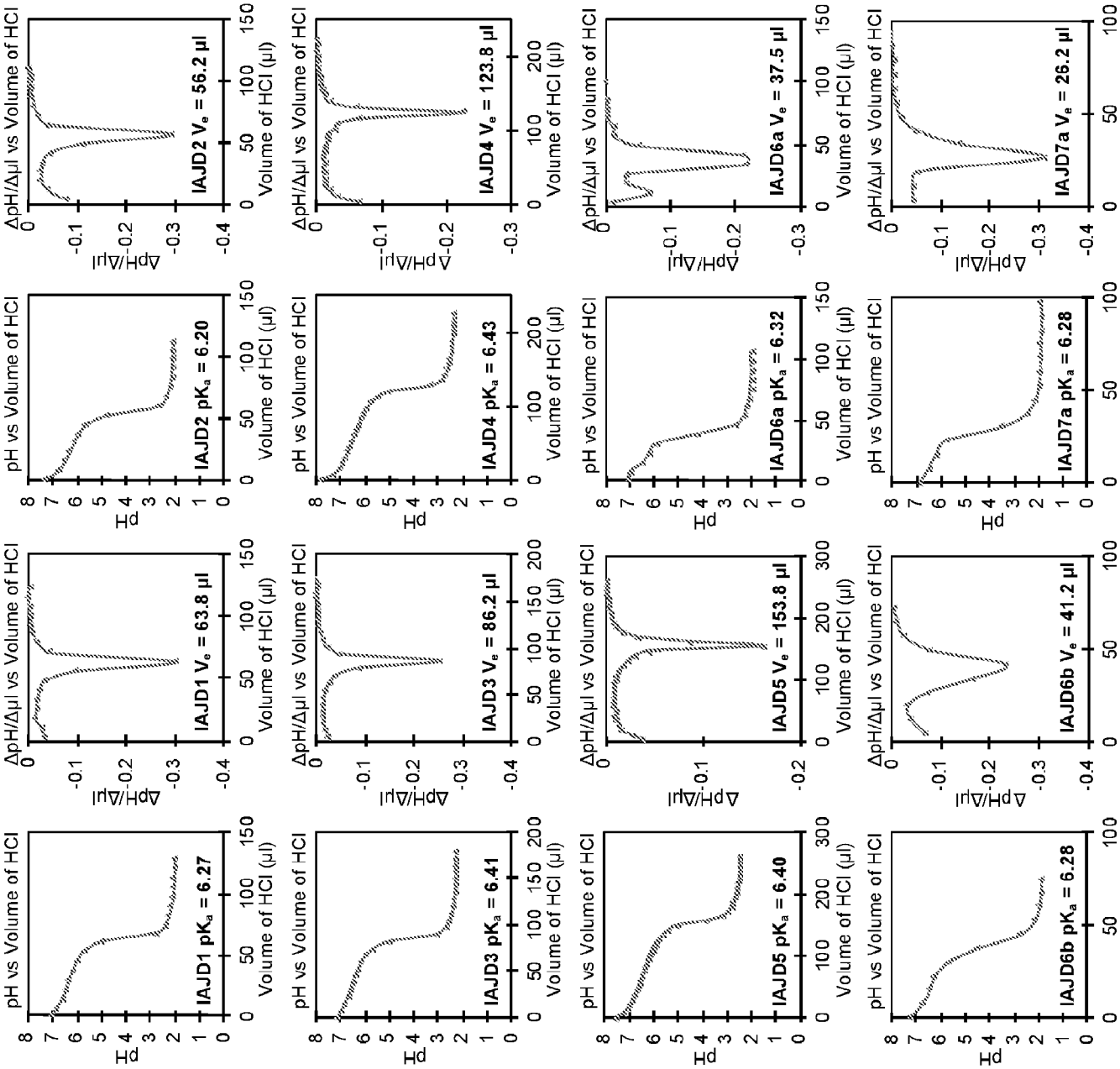

pH vs Volume of HCl
pH
Volume of HCl (µl)
IAJD1 pKa = 6.27
ΔpH/Δµl vs Volume of HCl
ΔpH/Δµl
IAJD1 Ve = 63.8 µl
IAJD2 pKa = 6.20
IAJD2 Ve = 56.2 µl
IAJD3 pKa = 6.41
IAJD3 Ve = 86.2 µl
IAJD4 pKa = 6.43
IAJD4 Ve = 123.8 µl
IAJD5 pKa = 6.40
IAJD5 Ve = 153.8 µl
IAJD6a pKa = 6.32
IAJD6a Ve = 37.5 µl
IAJD6b pKa = 6.28
IAJD6b Ve = 41.2 µl
IAJD7a pKa = 6.28
IAJD7a Ve = 26.2 µl

Figure 37

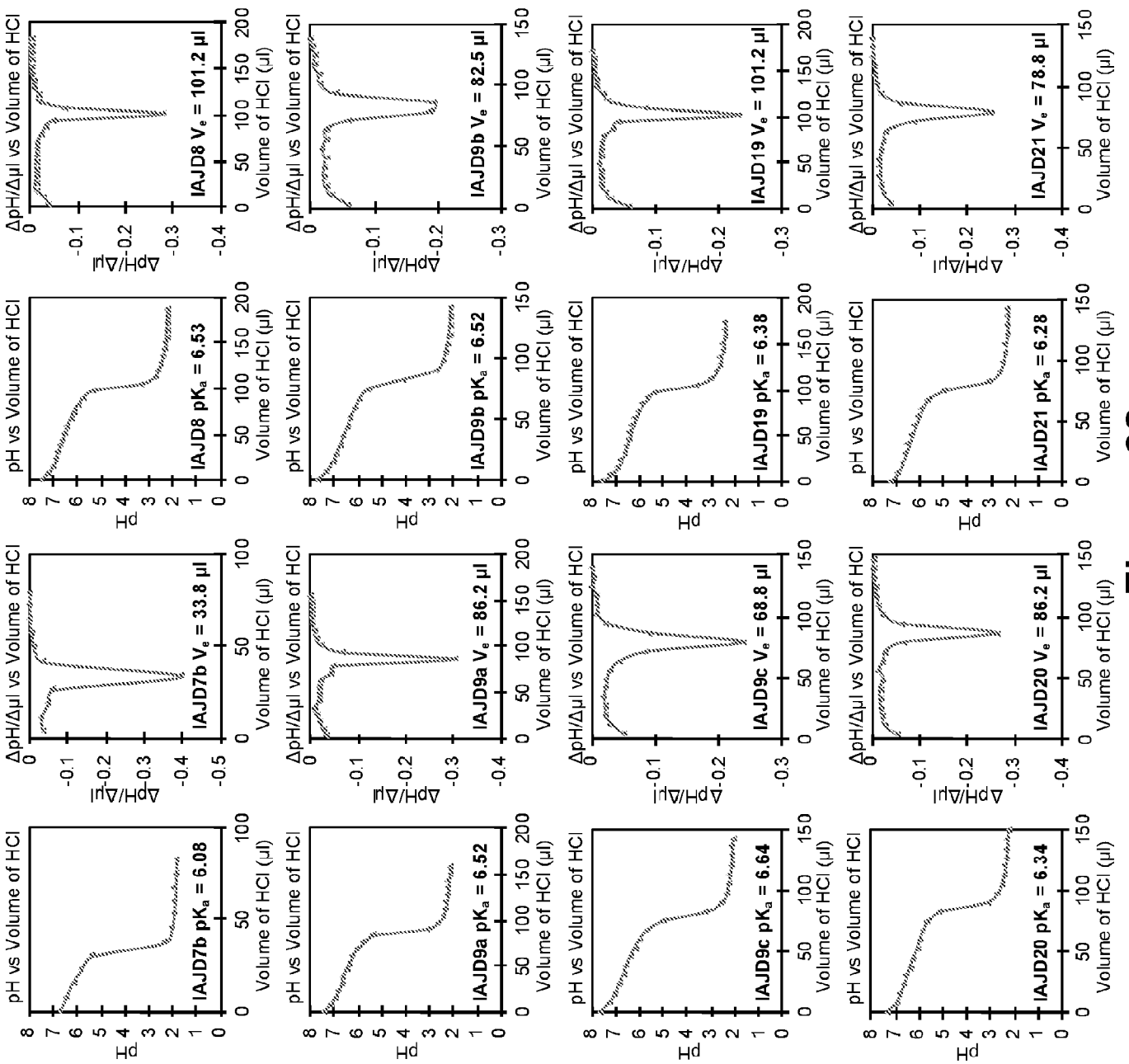

pH vs Volume of HCl
ΔpH/Δμl vs Volume of HCl
Volume of HCl (μl)
pH
ΔpH/Δμl
IAJD7b pKa = 6.08
IAJD7b Ve = 33.8 μl
IAJD8 pKa = 6.53
IAJD8 Ve = 101.2 μl
IAJD9a pKa = 6.52
IAJD9a Ve = 86.2 μl
IAJD9b pKa = 6.52
IAJD9b Ve = 82.5 μl
IAJD9c pKa = 6.64
IAJD9c Ve = 68.8 μl
IAJD19 pKa = 6.38
IAJD19 Ve = 101.2 μl
IAJD20 pKa = 6.34
IAJD20 Ve = 86.2 μl
IAJD21 pKa = 6.28
IAJD21 Ve = 78.8 μl

Figure 38

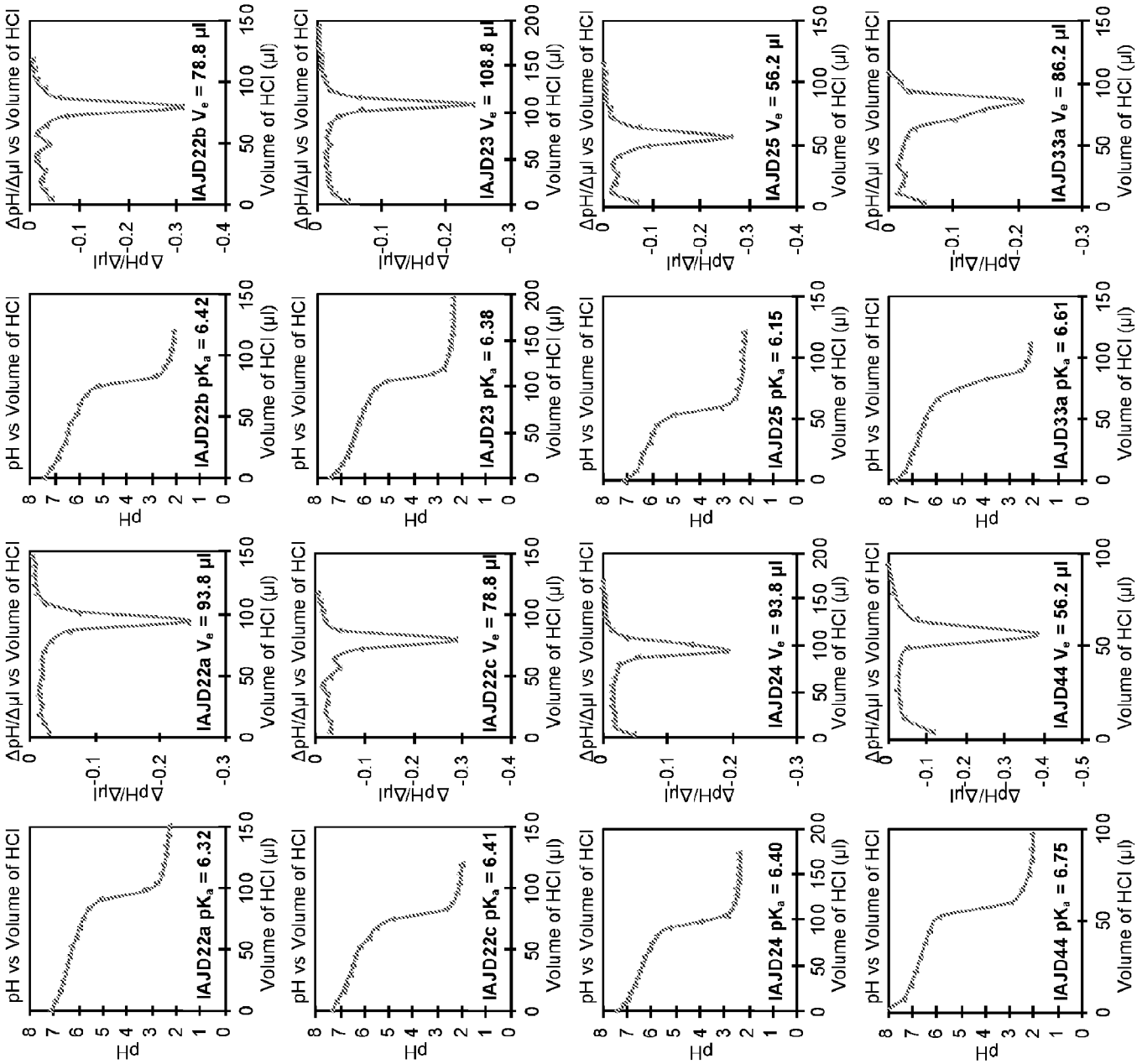

pH vs Volume of HCl
ΔpH/Δμl vs Volume of HCl
Volume of HCl (μl)
IAJD22a pKa = 6.32
IAJD22a Ve = 93.8 μl
IAJD22b pKa = 6.42
IAJD22b Ve = 78.8 μl
IAJD22c pKa = 6.41
IAJD22c Ve = 78.8 μl
IAJD23 pKa = 6.38
IAJD23 Ve = 108.8 μl
IAJD24 pKa = 6.40
IAJD24 Ve = 93.8 μl
IAJD25 pKa = 6.15
IAJD25 Ve = 56.2 μl
IAJD33a pKa = 6.61
IAJD33a Ve = 86.2 μl
IAJD44 pKa = 6.75
IAJD44 Ve = 56.2 μl

Figure 39

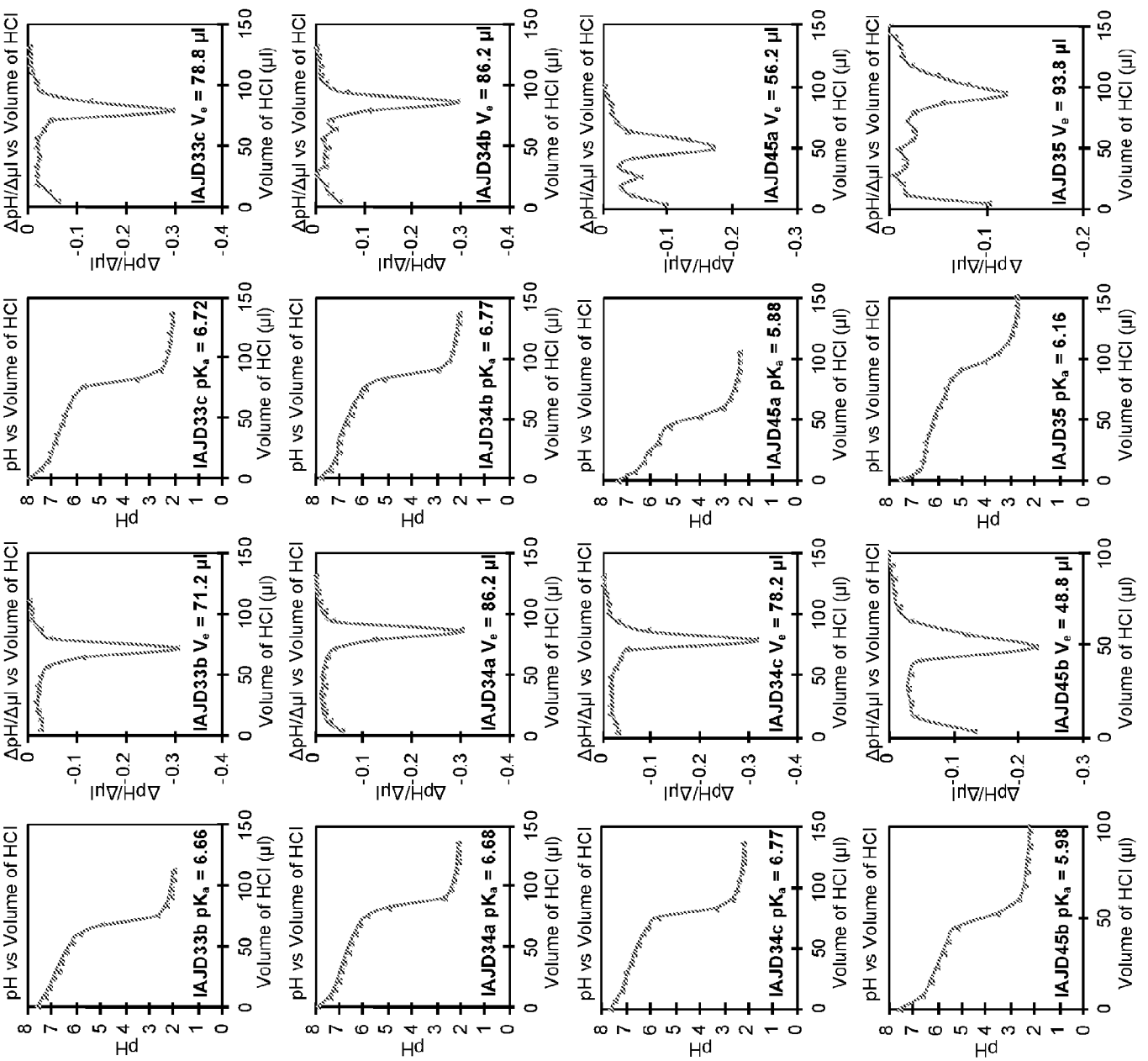

pH vs Volume of HCl
ΔpH/Δμl vs Volume of HCl
pH
ΔpH/Δμl
Volume of HCl (μl)
IAJD33b pKa = 6.66
IAJD33b Ve = 71.2 μl
IAJD33c pKa = 6.72
IAJD33c Ve = 78.8 μl
IAJD34a pKa = 6.68
IAJD34a Ve = 86.2 μl
IAJD34b pKa = 6.77
IAJD34b Ve = 86.2 μl
IAJD34c pKa = 6.77
IAJD34c Ve = 78.2 μl
IAJD45a pKa = 5.88
IAJD45a Ve = 56.2 μl
IAJD45b pKa = 5.98
IAJD45b Ve = 48.8 μl
IAJD35 pKa = 6.16
IAJD35 Ve = 93.8 μl

Figure 40

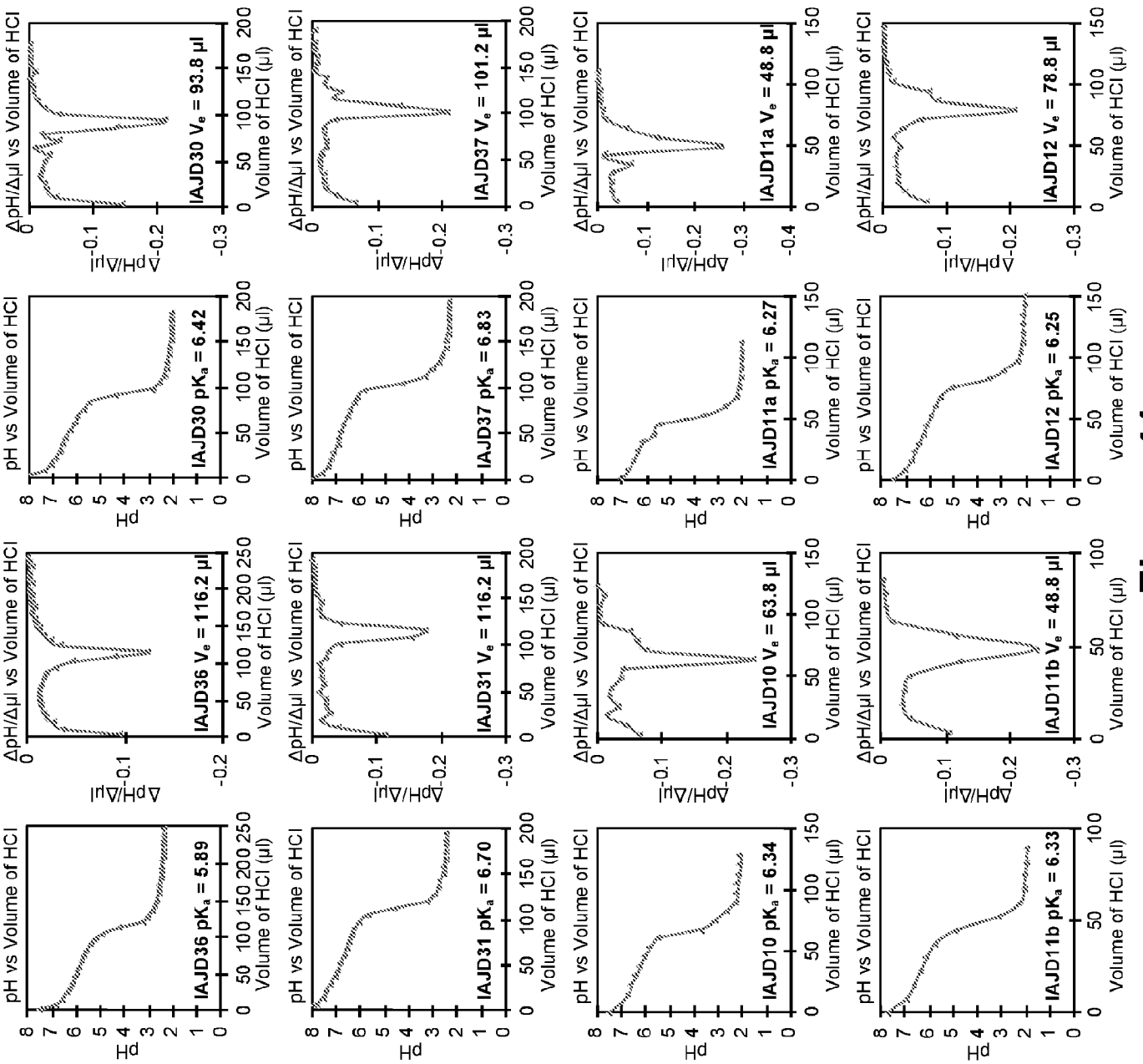

pH vs Volume of HCl
ΔpH/Δµl vs Volume of HCl
Volume of HCl (µl)
IAJD36 pKa = 5.89
IAJD36 Ve = 116.2 µl
IAJD31 pKa = 6.70
IAJD31 Ve = 116.2 µl
IAJD10 pKa = 6.34
IAJD10 Ve = 63.8 µl
IAJD11b pKa = 6.33
IAJD11b Ve = 48.8 µl
IAJD30 pKa = 6.42
IAJD30 Ve = 93.8 µl
IAJD37 pKa = 6.83
IAJD37 Ve = 101.2 µl
IAJD11a pKa = 6.27
IAJD11a Ve = 48.8 µl
IAJD12 pKa = 6.25
IAJD12 Ve = 78.8 µl

Figure 41

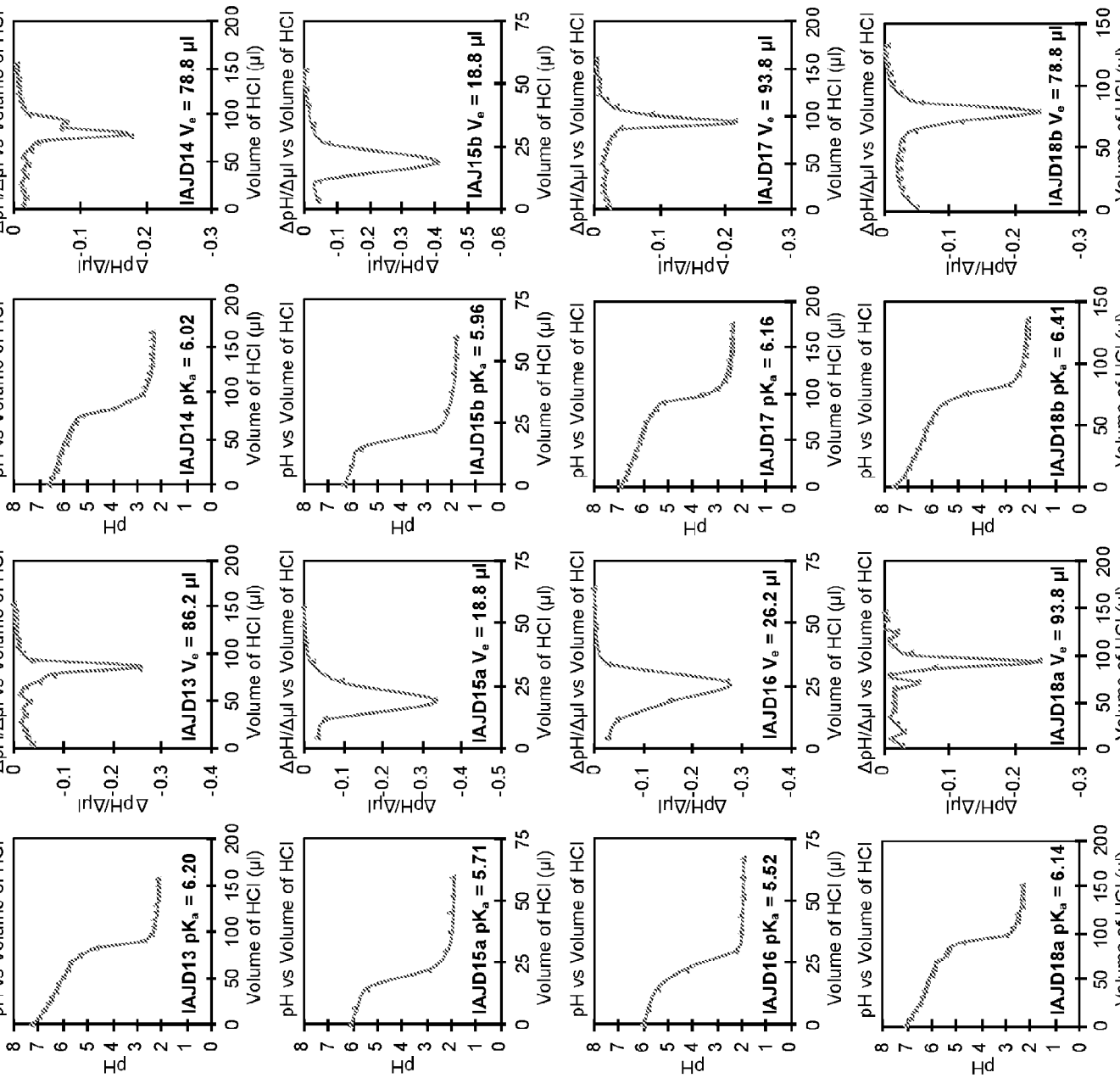

pH vs Volume of HCl
pH
Volume of HCl (µl)
IAJD13 pKa = 6.20
ΔpH/Δµl vs Volume of HCl
ΔpH/Δµl
IAJD13 Ve = 86.2 µl
IAJD14 pKa = 6.02
IAJD14 Ve = 78.8 µl
IAJD15a pKa = 5.71
IAJD15a Ve = 18.8 µl
IAJD15b pKa = 5.96
IAJD15b Ve = 18.8 µl
IAJD16 pKa = 5.52
IAJD16 Ve = 26.2 µl
IAJD17 pKa = 6.16
IAJD17 Ve = 93.8 µl
IAJD18a pKa = 6.14
IAJD18a Ve = 93.8 µl
IAJD18b pKa = 6.41
IAJD18b Ve = 78.8 µl

Figure 42

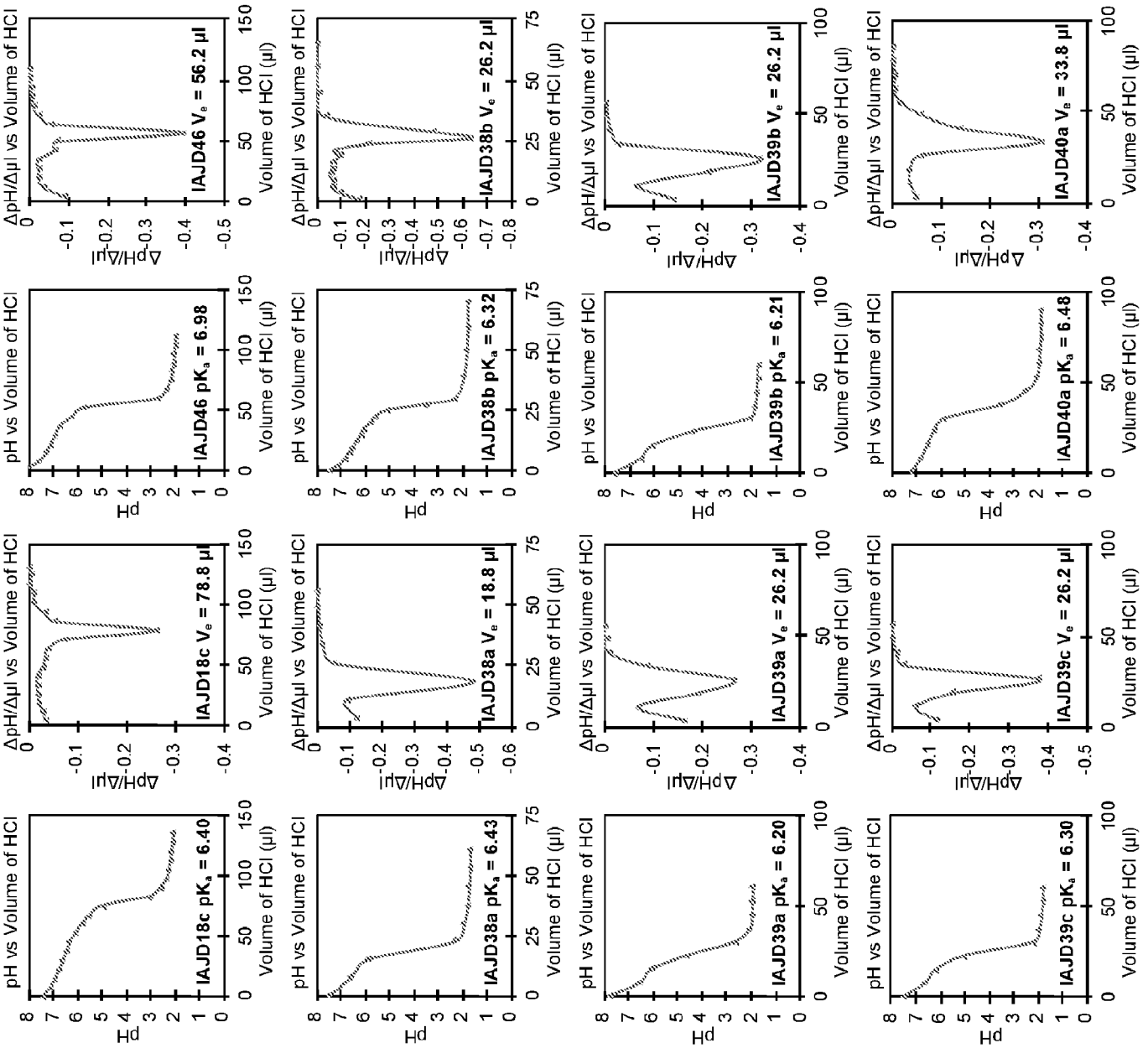

pH vs Volume of HCl
pH
Volume of HCl (µl)
IAJD18c pKa = 6.40
ΔpH/Δµl vs Volume of HCl
ΔpH/Δµl
IAJD18c Ve = 78.8 µl
pH vs Volume of HCl
IAJD46 pKa = 6.98
ΔpH/Δµl vs Volume of HCl
IAJD46 Ve = 56.2 µl
pH vs Volume of HCl
IAJD38a pKa = 6.43
ΔpH/Δµl vs Volume of HCl
IAJD38a Ve = 18.8 µl
pH vs Volume of HCl
IAJD38b pKa = 6.32
ΔpH/Δµl vs Volume of HCl
IAJD38b Ve = 26.2 µl
pH vs Volume of HCl
IAJD39a pKa = 6.20
ΔpH/Δµl vs Volume of HCl
IAJD39a Ve = 26.2 µl
pH vs Volume of HCl
IAJD39b pKa = 6.21
ΔpH/Δµl vs Volume of HCl
IAJD39b Ve = 26.2 µl
pH vs Volume of HCl
IAJD39c pKa = 6.30
ΔpH/Δµl vs Volume of HCl
IAJD39c Ve = 26.2 µl
pH vs Volume of HCl
IAJD40a pKa = 6.48
ΔpH/Δµl vs Volume of HCl
IAJD40a Ve = 33.8 µl

Figure 43

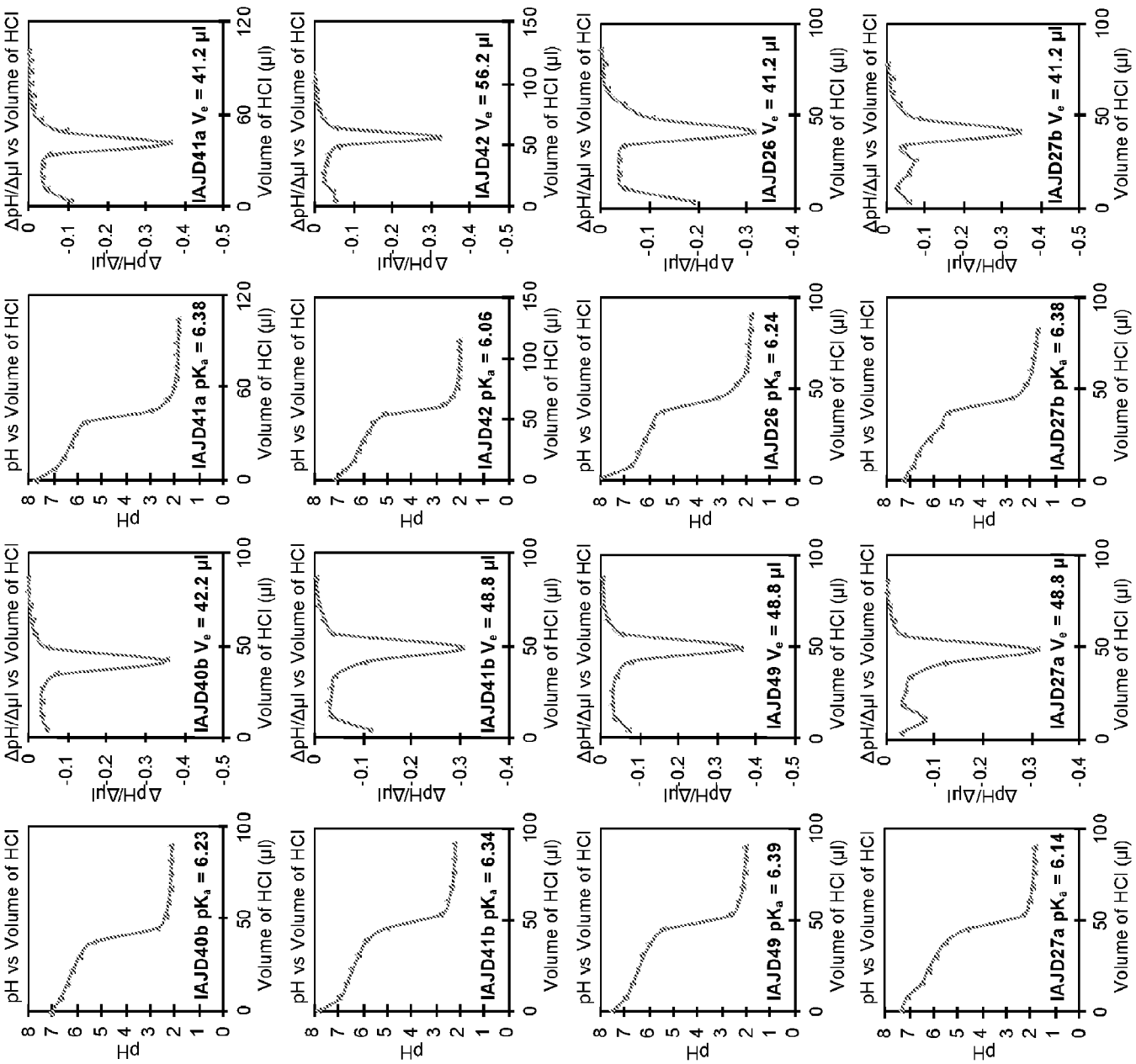

pH vs Volume of HCl
ΔpH/Δμl vs Volume of HCl
pH
ΔpH/Δμl
Volume of HCl (μl)
IAJD40b pKa = 6.23
IAJD40b Ve = 42.2 μl
IAJD41a pKa = 6.38
IAJD41a Ve = 41.2 μl
IAJD41b pKa = 6.34
IAJD41b Ve = 48.8 μl
IAJD42 pKa = 6.06
IAJD42 Ve = 56.2 μl
IAJD49 pKa = 6.39
IAJD49 Ve = 48.8 μl
IAJD26 pKa = 6.24
IAJD26 Ve = 41.2 μl
IAJD27a pKa = 6.14
IAJD27a Ve = 48.8 μl
IAJD27b pKa = 6.38
IAJD27b Ve = 41.2 μl

Figure 44

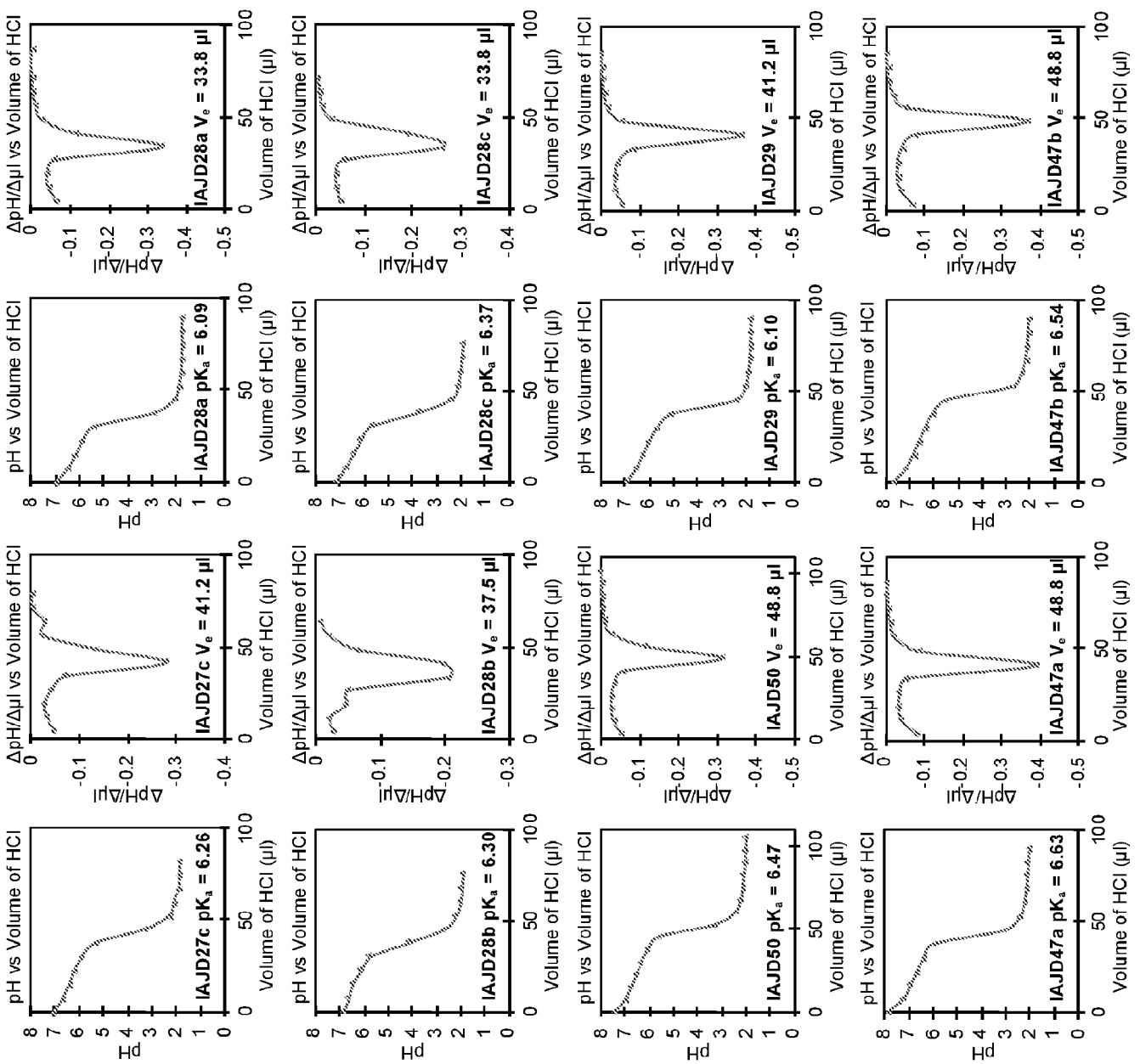

pH vs Volume of HCl
ΔpH/Δμl vs Volume of HCl
Volume of HCl (μl)
IAJD27c pKa = 6.26
IAJD27c Ve = 41.2 μl
IAJD28b pKa = 6.30
IAJD28b Ve = 37.5 μl
IAJD50 pKa = 6.47
IAJD50 Ve = 48.8 μl
IAJD47a pKa = 6.63
IAJD47a Ve = 48.8 μl
IAJD28a pKa = 6.09
IAJD28a Ve = 33.8 μl
IAJD28c pKa = 6.37
IAJD28c Ve = 33.8 μl
IAJD29 pKa = 6.10
IAJD29 Ve = 41.2 μl
IAJD47b pKa = 6.54
IAJD47b Ve = 48.8 μl

Figure 45

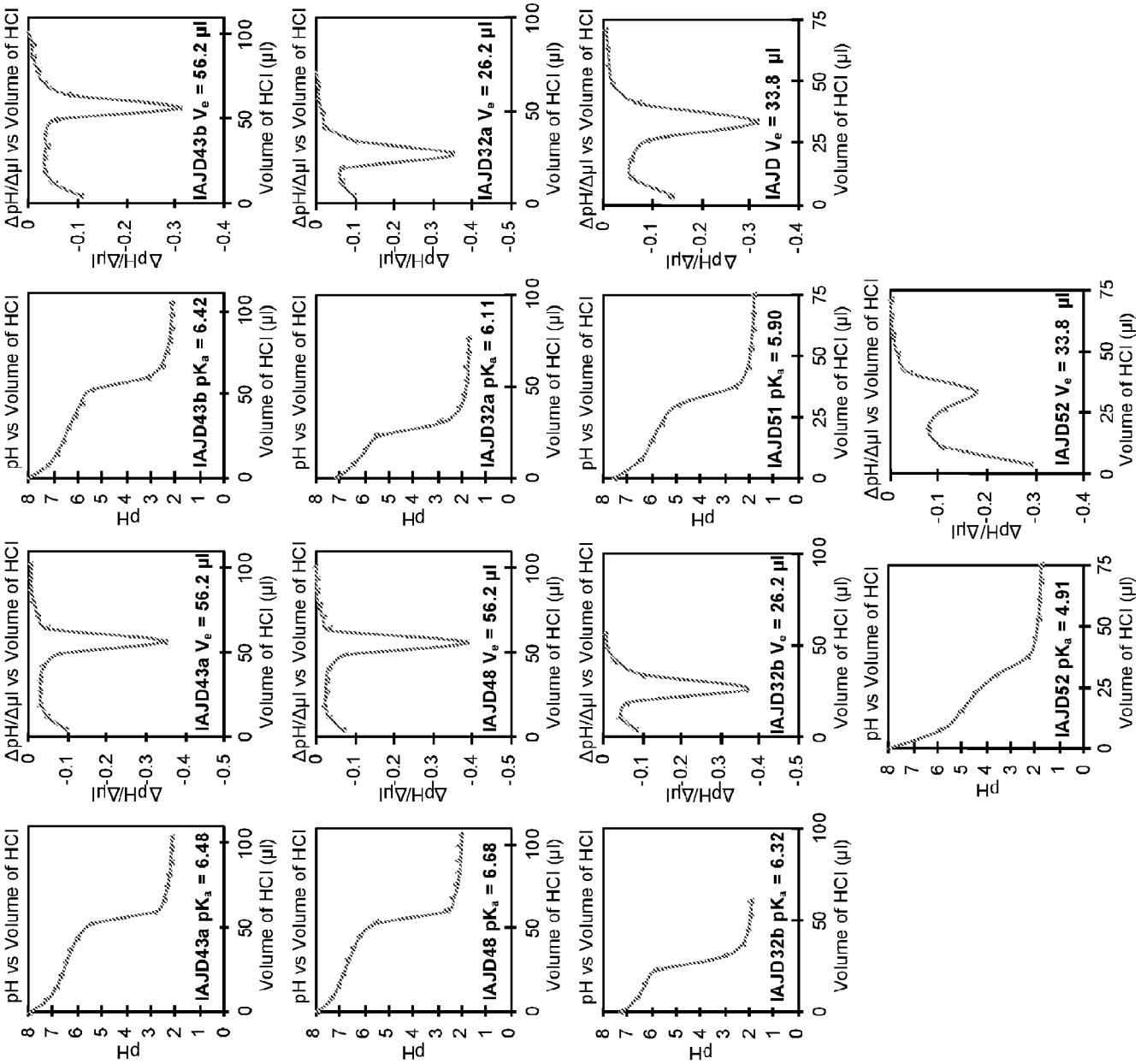

pH vs Volume of HCl
ΔpH/Δμl vs Volume of HCl
Volume of HCl (μl)
IAJD43a pKa = 6.48
IAJD43a Ve = 56.2 μl
IAJD43b pKa = 6.42
IAJD43b Ve = 56.2 μl
IAJD48 pKa = 6.68
IAJD48 Ve = 56.2 μl
IAJD32a pKa = 6.11
IAJD32a Ve = 26.2 μl
IAJD32b pKa = 6.32
IAJD32b Ve = 26.2 μl
IAJD51 pKa = 5.90
IAJD pKa = 33.8 μl
IAJD52 pKa = 4.91
IAJD52 Ve = 33.8 μl

Figure 46

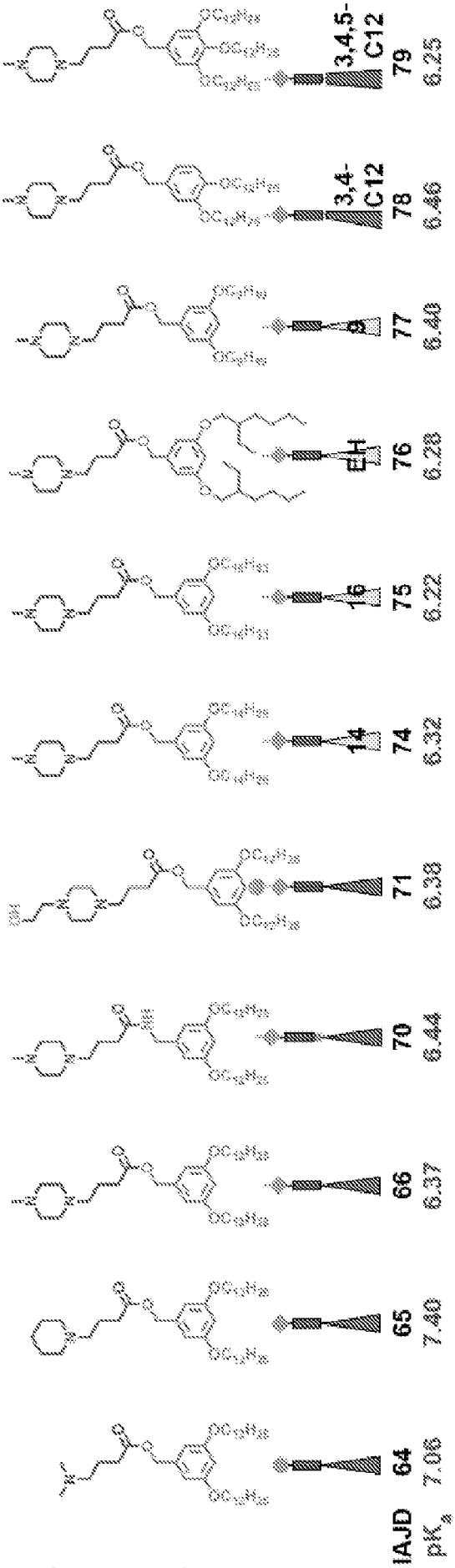

IAJD 64 7.06
65 7.40
66 6.37
70 6.44
71 6.38
74 6.32
75 6.22
76 6.28
77 6.48
78 6.46
79 6.25
3,4,5-C12
3,4-C12
9
16
14
EH

Figure 47

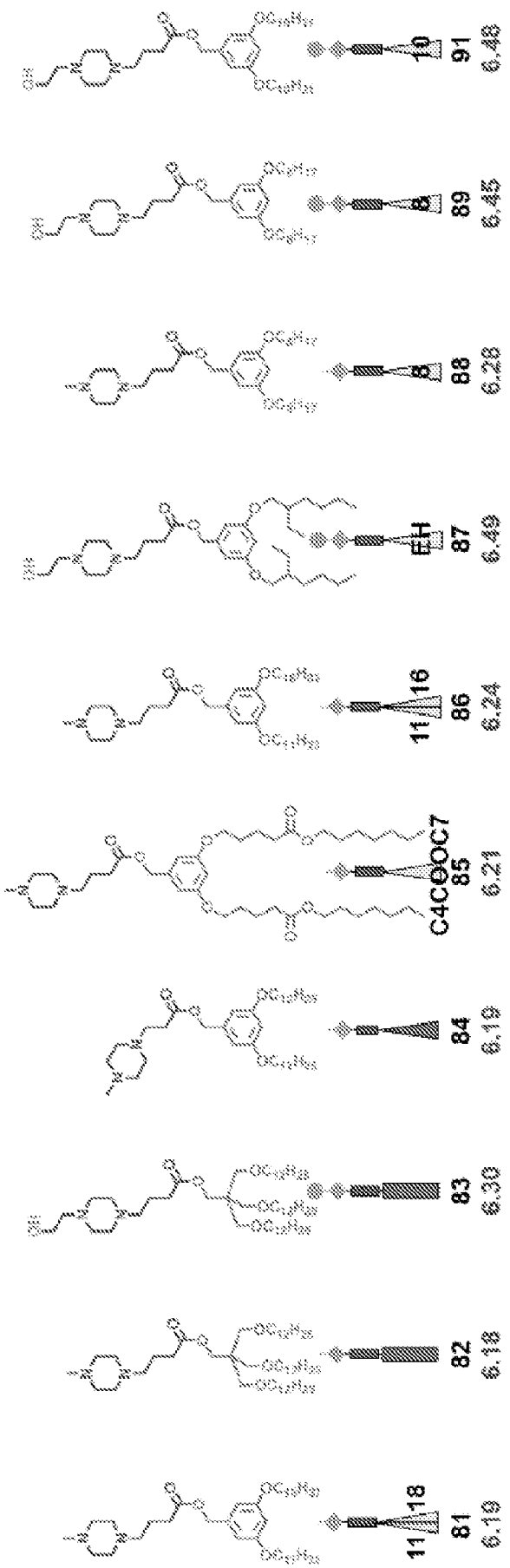


Figure 47 (cont.)

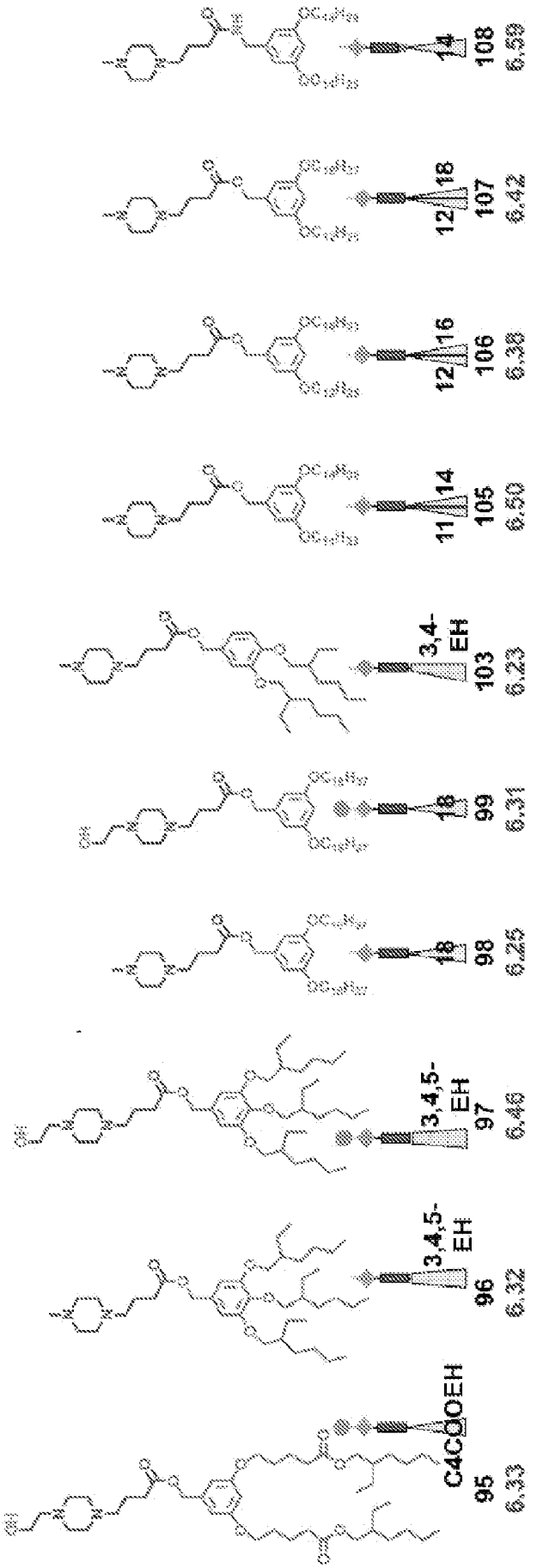


Figure 47 (cont.)

Figure 48

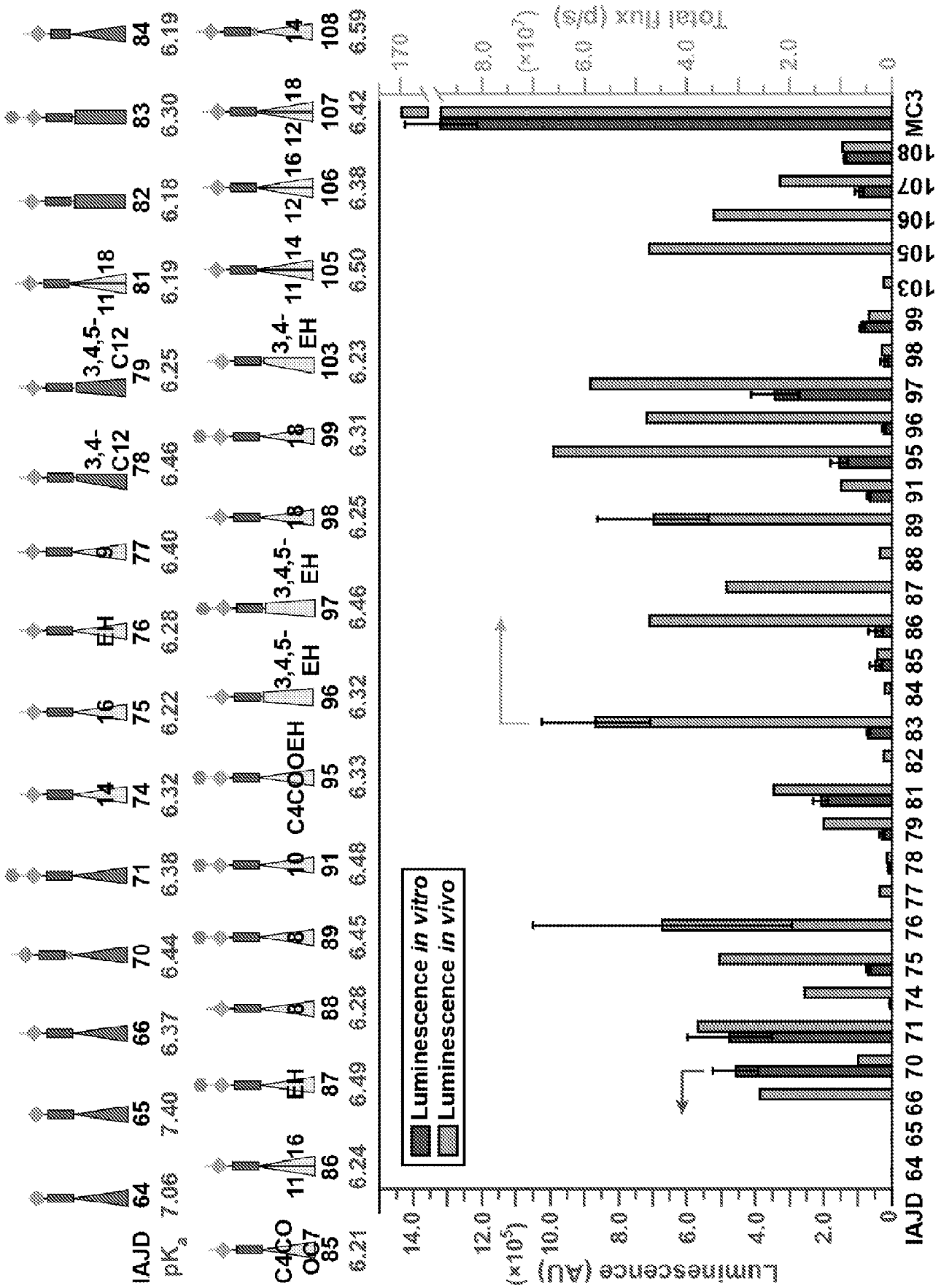

Luminescence in vitro
Luminescence in vivo
Luminescence (AU) (×10^3)
Total flux (p/s) (×10^7)
IAJD
MC3
pKa

Figure 49

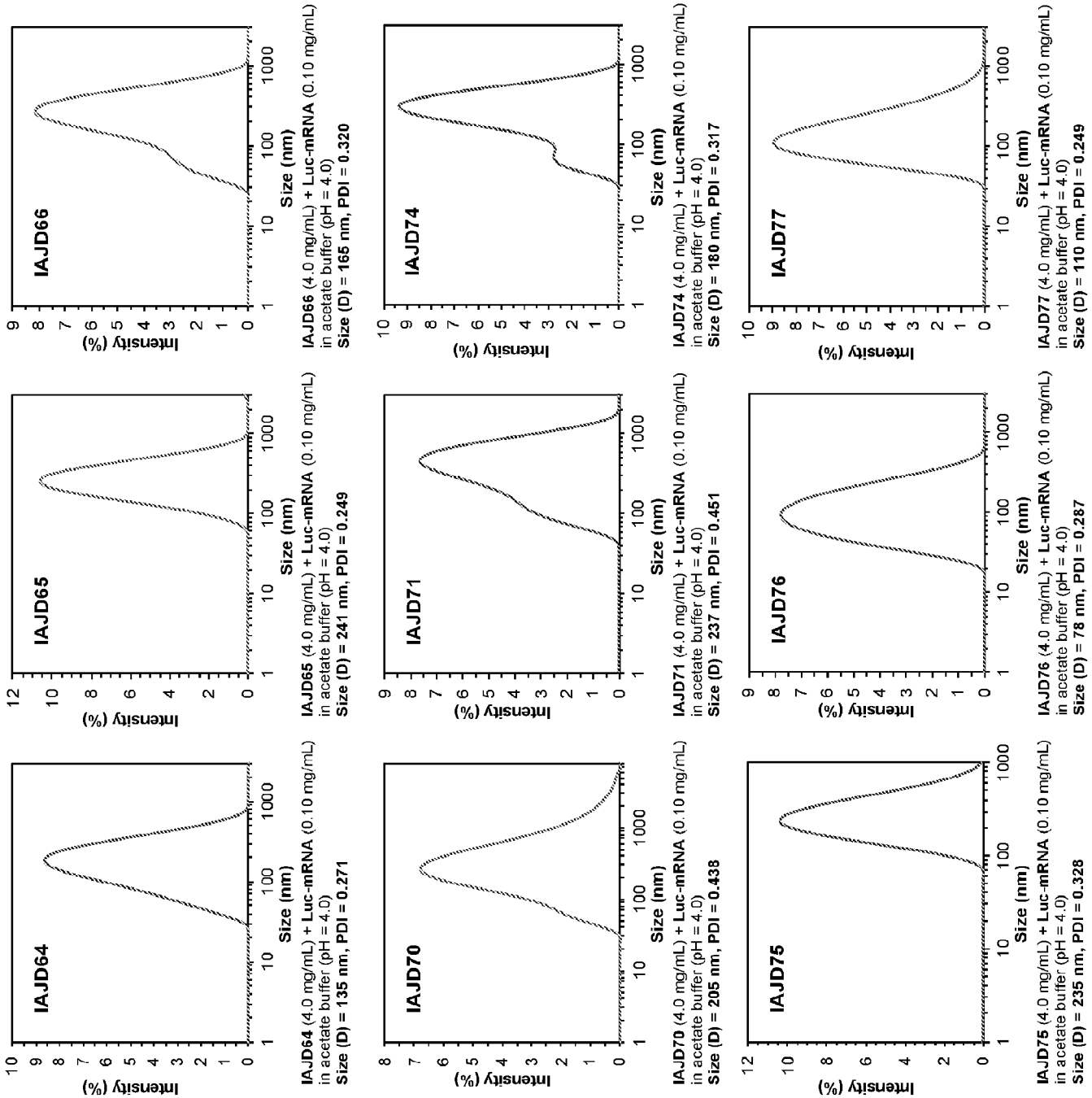

IAJD64
Intensity (%)
Size (nm)
IAJD64 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 135 nm, PDI = 0.271
IAJD65
Intensity (%)
Size (nm)
IAJD65 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 241 nm, PDI = 0.249
IAJD66
Intensity (%)
Size (nm)
IAJD66 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 165 nm, PDI = 0.320
IAJD70
Intensity (%)
Size (nm)
IAJD70 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 205 nm, PDI = 0.438
IAJD71
Intensity (%)
Size (nm)
IAJD71 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 237 nm, PDI = 0.451
IAJD74
Intensity (%)
Size (nm)
IAJD74 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 180 nm, PDI = 0.317
IAJD75
Intensity (%)
Size (nm)
IAJD75 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 235 nm, PDI = 0.328
IAJD76
Intensity (%)
Size (nm)
IAJD76 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 78 nm, PDI = 0.287
IAJD77
Intensity (%)
Size (nm)
IAJD77 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 110 nm, PDI = 0.249

Figure 50

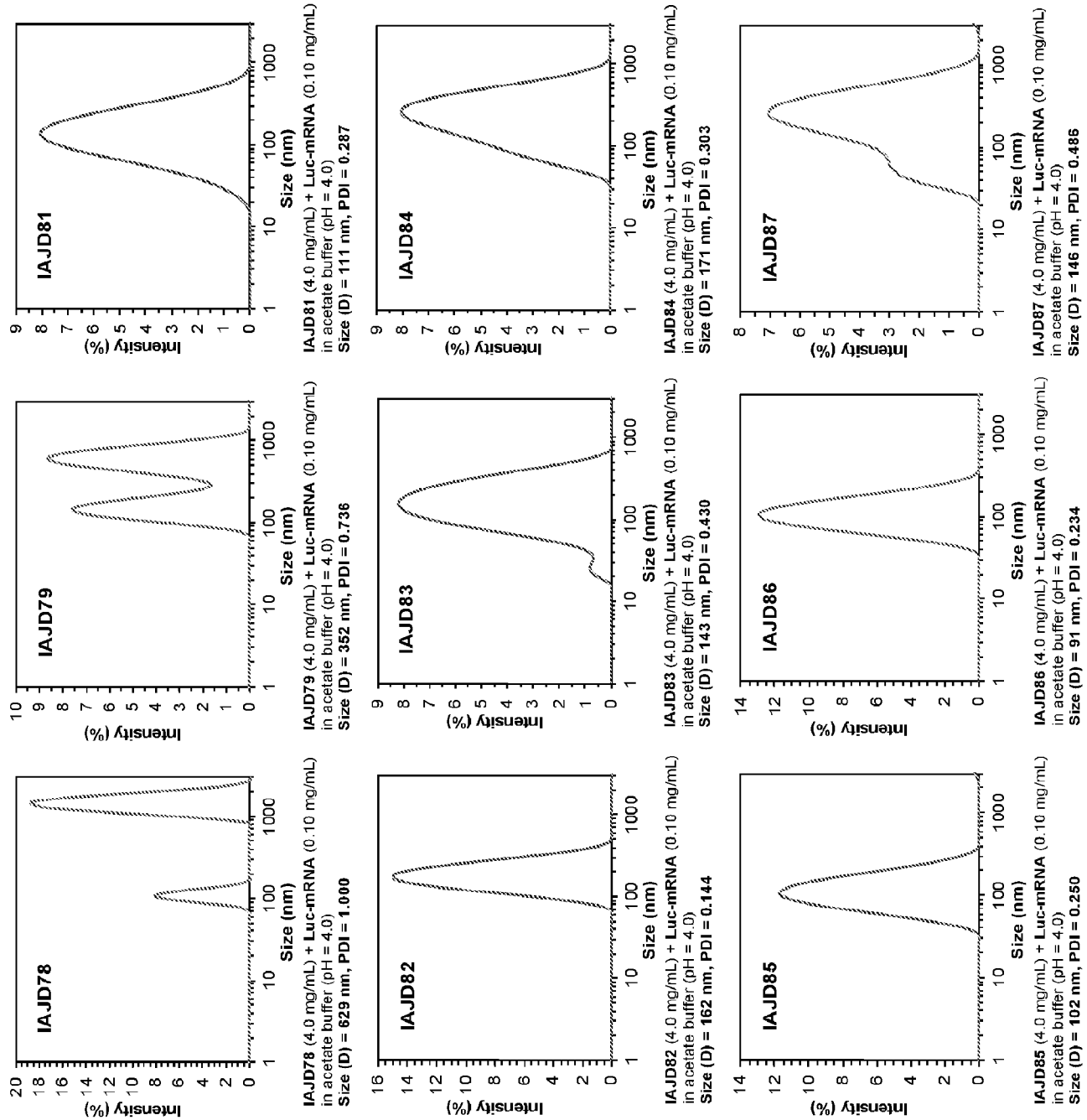

IAJD78
Intensity (%)
Size (nm)
IAJD78 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 629 nm, PDI = 1.000
IAJD79
Intensity (%)
Size (nm)
IAJD79 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 352 nm, PDI = 0.736
IAJD81
Intensity (%)
Size (nm)
IAJD81 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 111 nm, PDI = 0.287
IAJD82
Intensity (%)
Size (nm)
IAJD82 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 162 nm, PDI = 0.144
IAJD83
Intensity (%)
Size (nm)
IAJD83 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 143 nm, PDI = 0.430
IAJD84
Intensity (%)
Size (nm)
IAJD84 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 171 nm, PDI = 0.303
IAJD85
Intensity (%)
Size (nm)
IAJD85 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 102 nm, PDI = 0.250
IAJD86
Intensity (%)
Size (nm)
IAJD86 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 91 nm, PDI = 0.234
IAJD87
Intensity (%)
Size (nm)
IAJD87 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 146 nm, PDI = 0.486

Figure 51

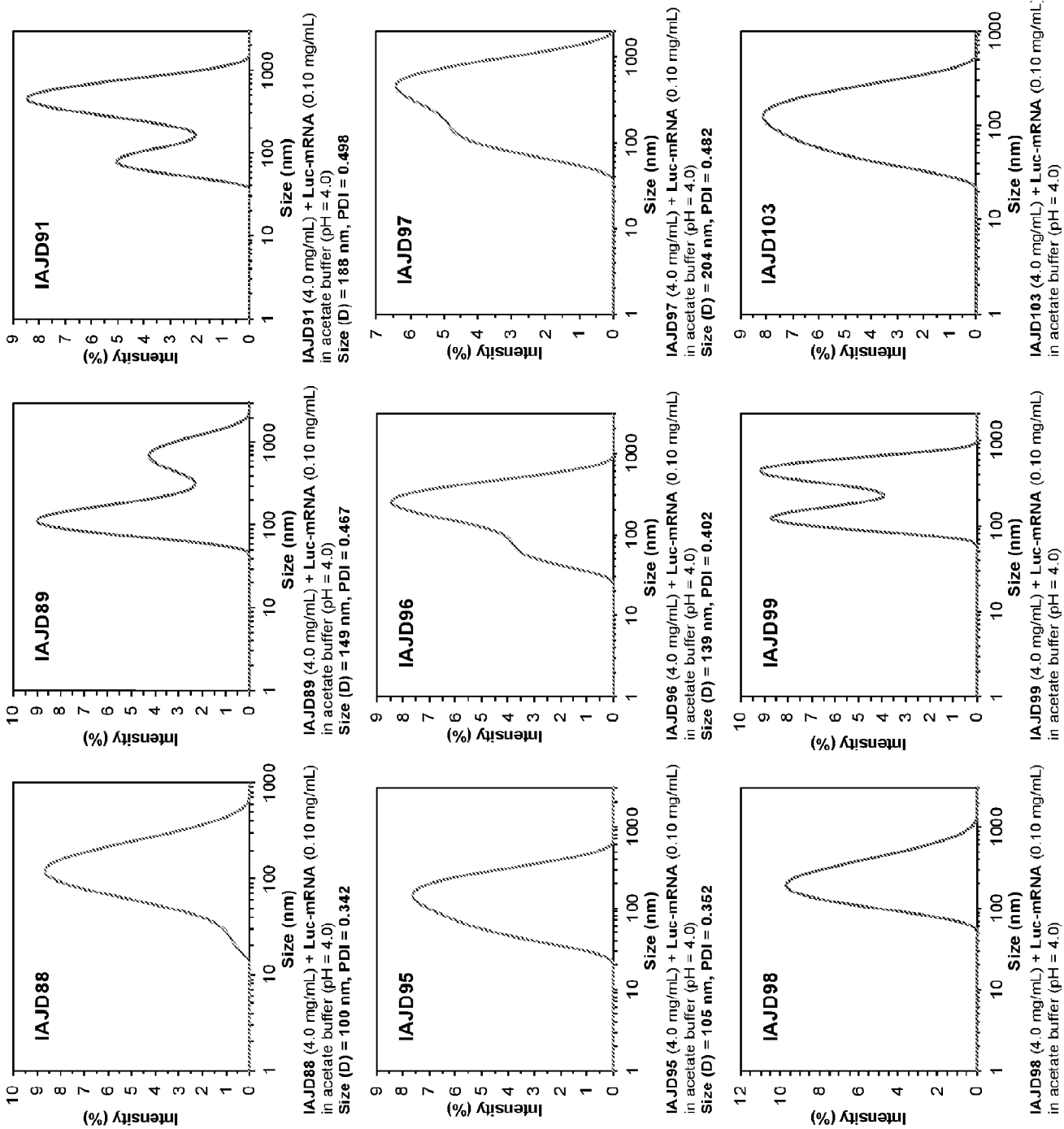
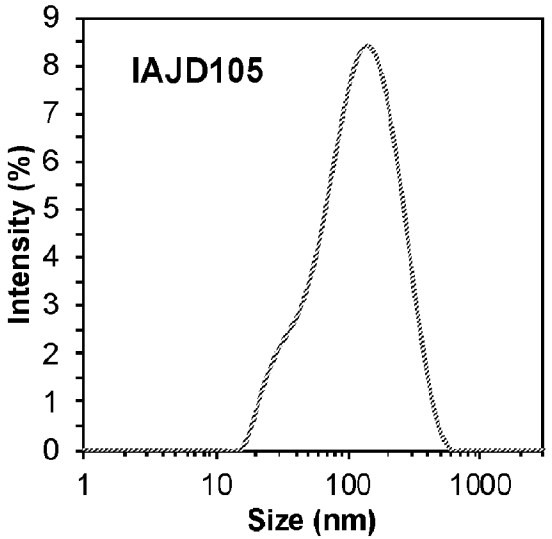
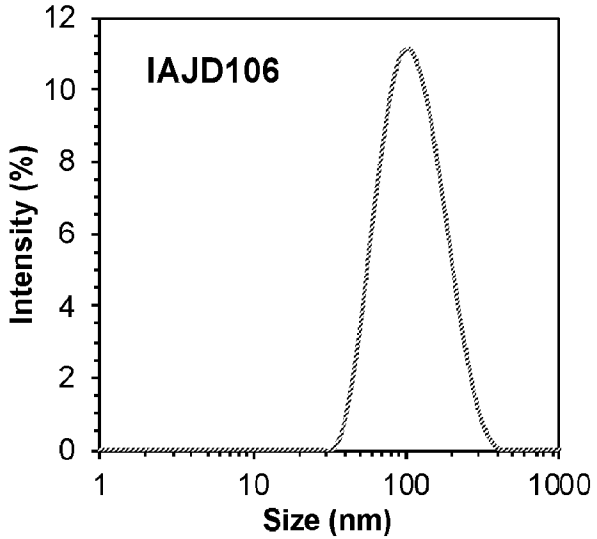

IAJD88
Intensity (%)
Size (nm)
IAJD88 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 100 nm, PDI = 0.342
IAJD89
Intensity (%)
Size (nm)
IAJD89 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 149 nm, PDI = 0.467
IAJD91
Intensity (%)
Size (nm)
IAJD91 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 188 nm, PDI = 0.498
IAJD95
Intensity (%)
Size (nm)
IAJD95 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 105 nm, PDI = 0.352
IAJD96
Intensity (%)
Size (nm)
IAJD96 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 139 nm, PDI = 0.402
IAJD97
Intensity (%)
Size (nm)
IAJD97 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 204 nm, PDI = 0.482
IAJD98
Intensity (%)
Size (nm)
IAJD98 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 187 nm, PDI = 0.221
IAJD99
Intensity (%)
Size (nm)
IAJD99 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 277 nm, PDI = 0.571
IAJD103
Intensity (%)
Size (nm)
IAJD103 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 88 nm, PDI = 0.275

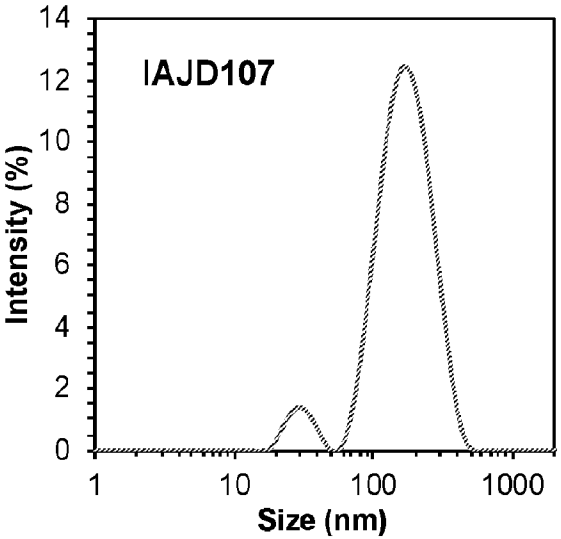
Figure 52



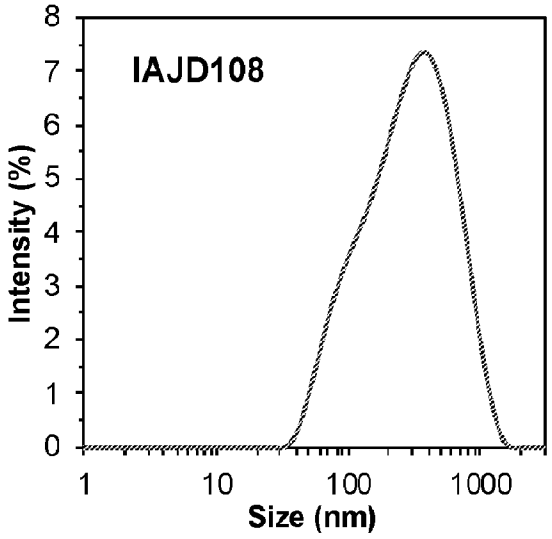
IAJD105 (4.0 mg/mL) + **Luc-mRNA** (0.10 mg/mL)
in acetate buffer (pH = 4.0)
Size (D) = 94 nm, PDI = 0.294



IAJD106 (4.0 mg/mL) + **Luc-mRNA** (0.10 mg/mL)
in acetate buffer (pH = 4.0)
Size (D) = 105 nm, PDI = 0.275



IAJD107 (4.0 mg/mL) + **Luc-mRNA** (0.10 mg/mL)
in acetate buffer (pH = 4.0)
Size (D) = 146 nm, PDI = 0.279



IAJD108 (4.0 mg/mL) + **Luc-mRNA** (0.10 mg/mL)
in acetate buffer (pH = 4.0)
Size (D) = 214 nm, PDI = 0.334

Figure 53

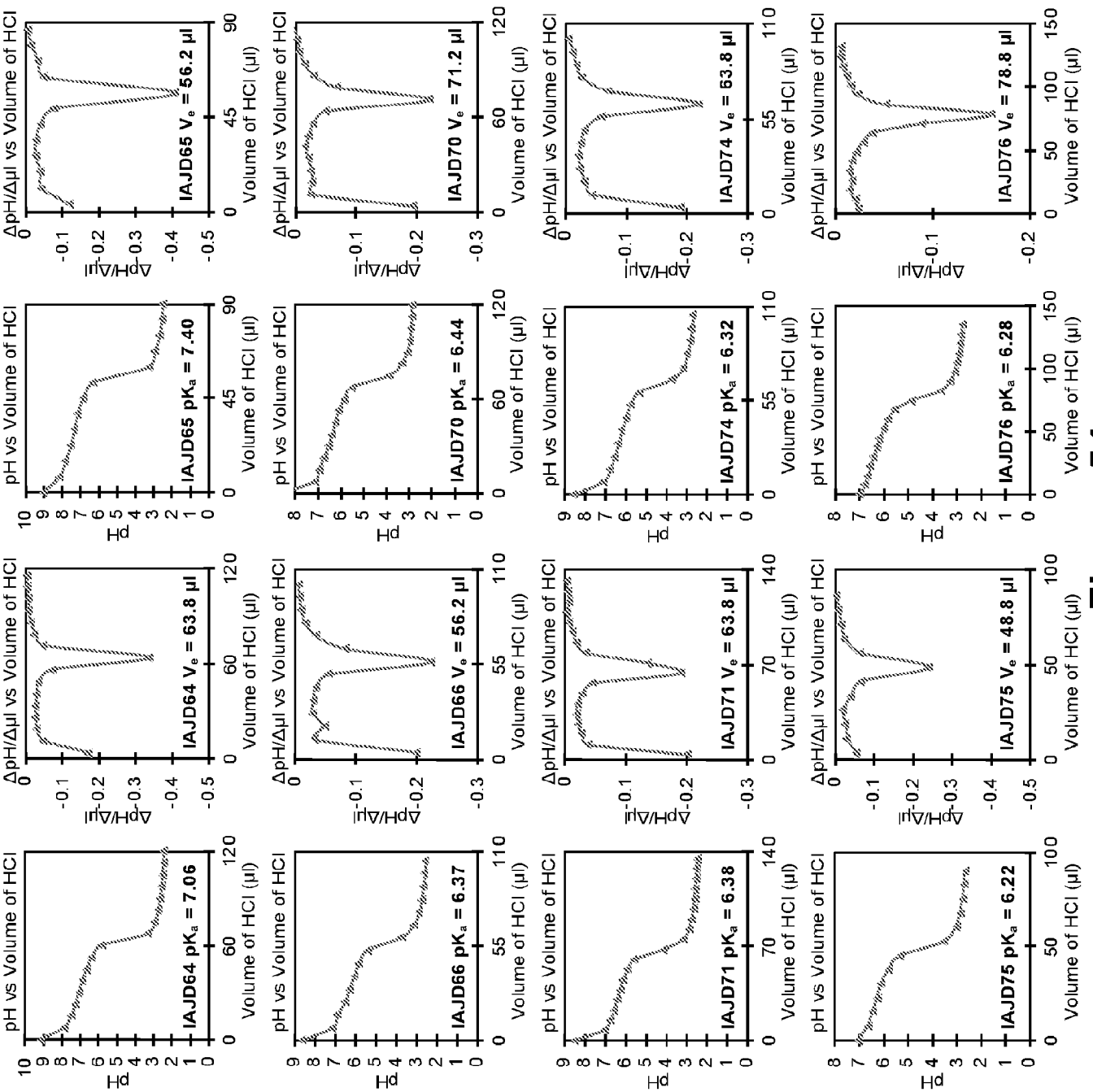

pH vs Volume of HCl
pH
Volume of HCl (µl)
IAJD64 pKa = 7.06
ΔpH/Δµl vs Volume of HCl
ΔpH/Δµl
IAJD64 Ve = 63.8 µl
IAJD65 pKa = 7.40
IAJD65 Ve = 56.2 µl
IAJD66 pKa = 6.37
IAJD66 Ve = 56.2 µl
IAJD70 pKa = 6.44
IAJD70 Ve = 71.2 µl
IAJD71 pKa = 6.38
IAJD71 Ve = 63.8 µl
IAJD74 pKa = 6.32
IAJD74 Ve = 63.8 µl
IAJD75 pKa = 6.22
IAJD75 Ve = 48.8 µl
IAJD76 pKa = 6.28
IAJD76 Ve = 78.8 µl

Figure 54

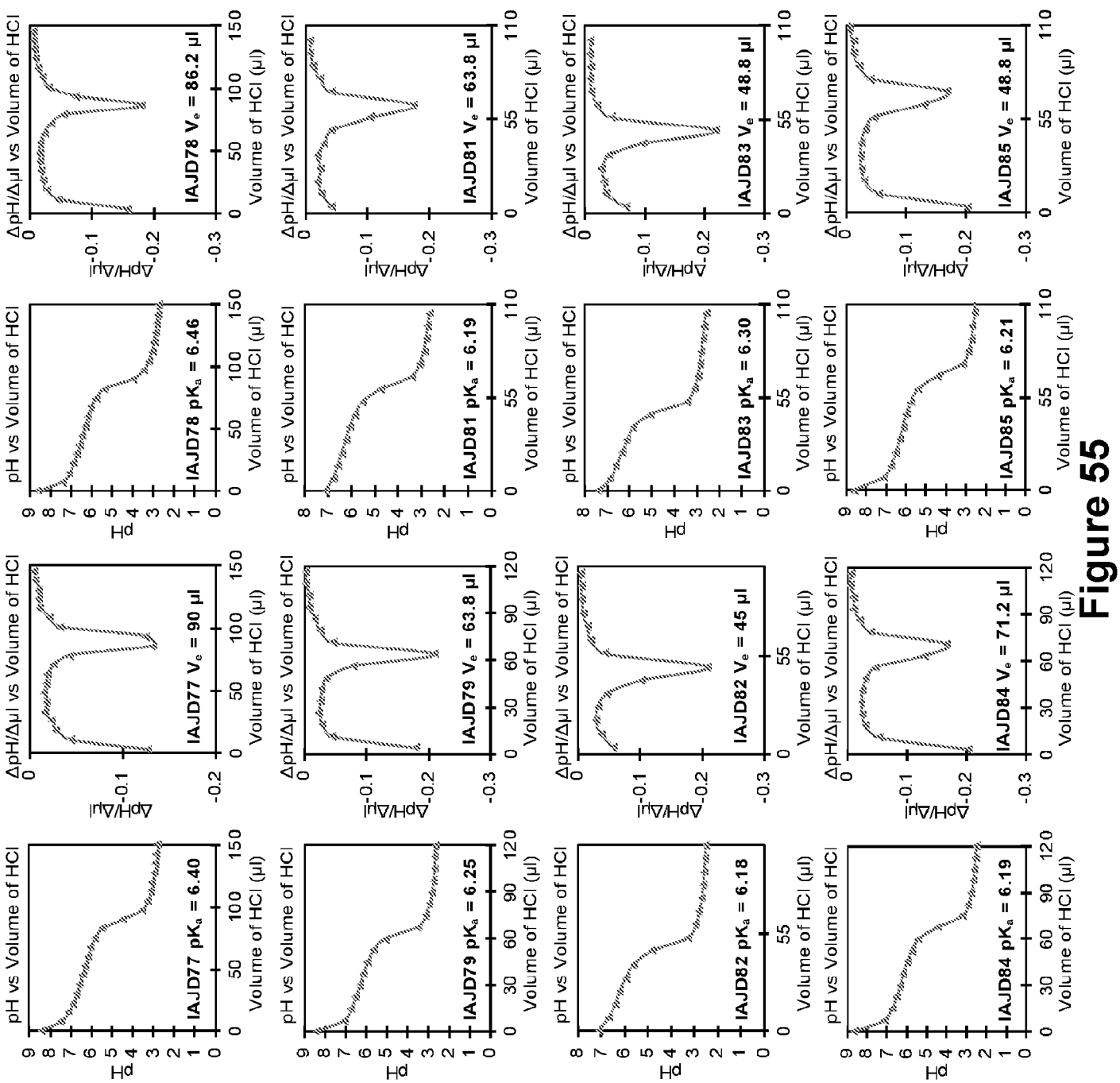

pH vs Volume of HCl
ΔpH/Δμl vs Volume of HCl
Volume of HCl (μl)
IAJD77 pKa = 6.40
IAJD77 Ve = 90 μl
IAJD78 pKa = 6.46
IAJD78 Ve = 86.2 μl
IAJD79 pKa = 6.25
IAJD79 Ve = 63.8 μl
IAJD81 pKa = 6.19
IAJD81 Ve = 63.8 μl
IAJD82 pKa = 6.18
IAJD82 Ve = 45 μl
IAJD83 pKa = 6.30
IAJD83 Ve = 48.8 μl
IAJD84 pKa = 6.19
IAJD84 Ve = 71.2 μl
IAJD85 pKa = 6.21
IAJD85 Ve = 48.8 μl

Figure 55

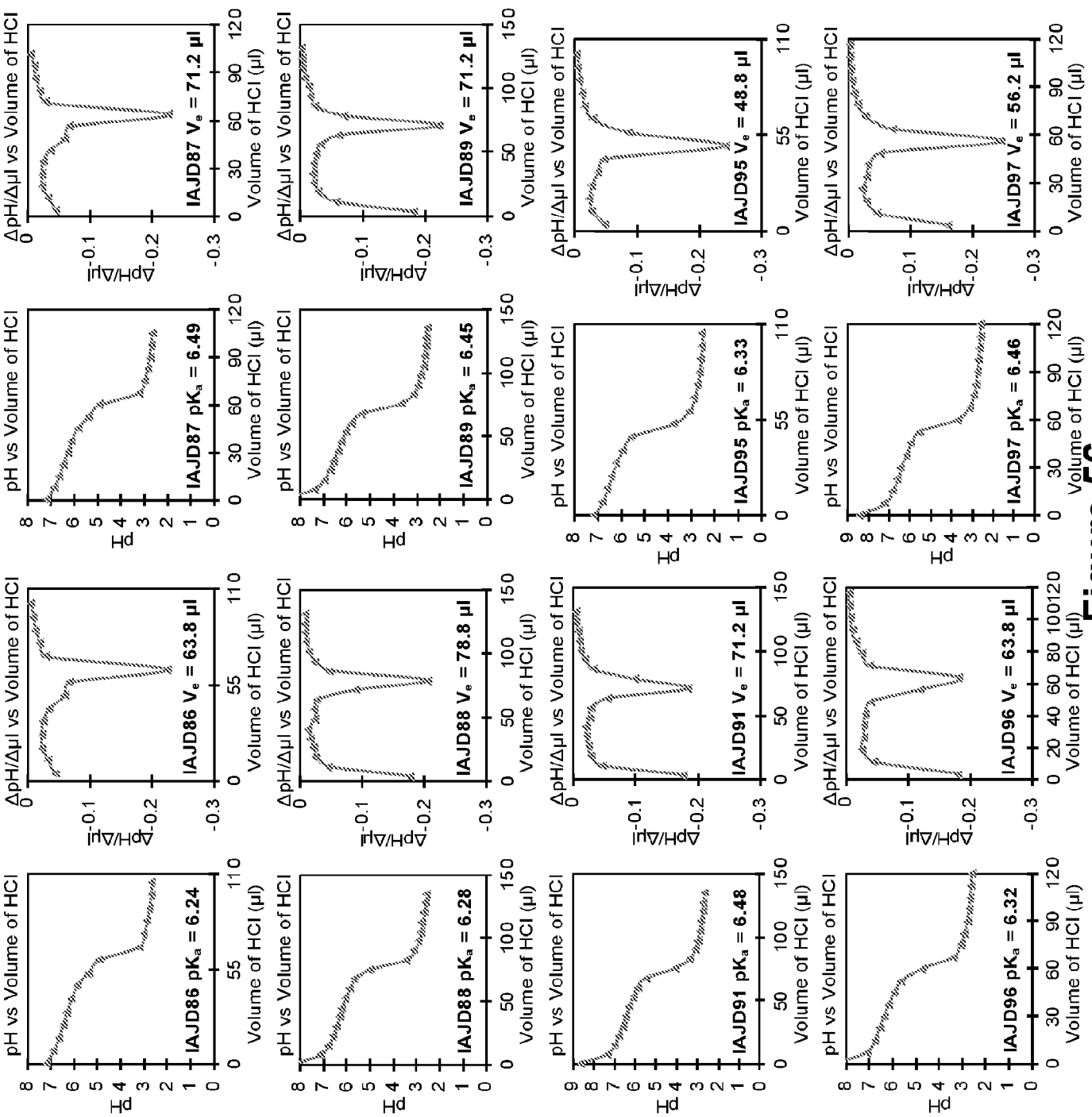

pH vs Volume of HCl
pH
Volume of HCl (µl)
IAJD86 pKa = 6.24
ΔpH/Δµl vs Volume of HCl
ΔpH/Δµl
IAJD86 Ve = 63.8 µl
IAJD87 pKa = 6.49
IAJD87 Ve = 71.2 µl
IAJD88 pKa = 6.28
IAJD88 Ve = 78.8 µl
IAJD89 pKa = 6.45
IAJD89 Ve = 71.2 µl
IAJD91 pKa = 6.48
IAJD91 Ve = 71.2 µl
IAJD95 pKa = 6.33
IAJD95 Ve = 48.8 µl
IAJD96 pKa = 6.32
IAJD96 Ve = 63.8 µl
IAJD97 pKa = 6.46
IAJD97 Ve = 56.2 µl

Figure 56

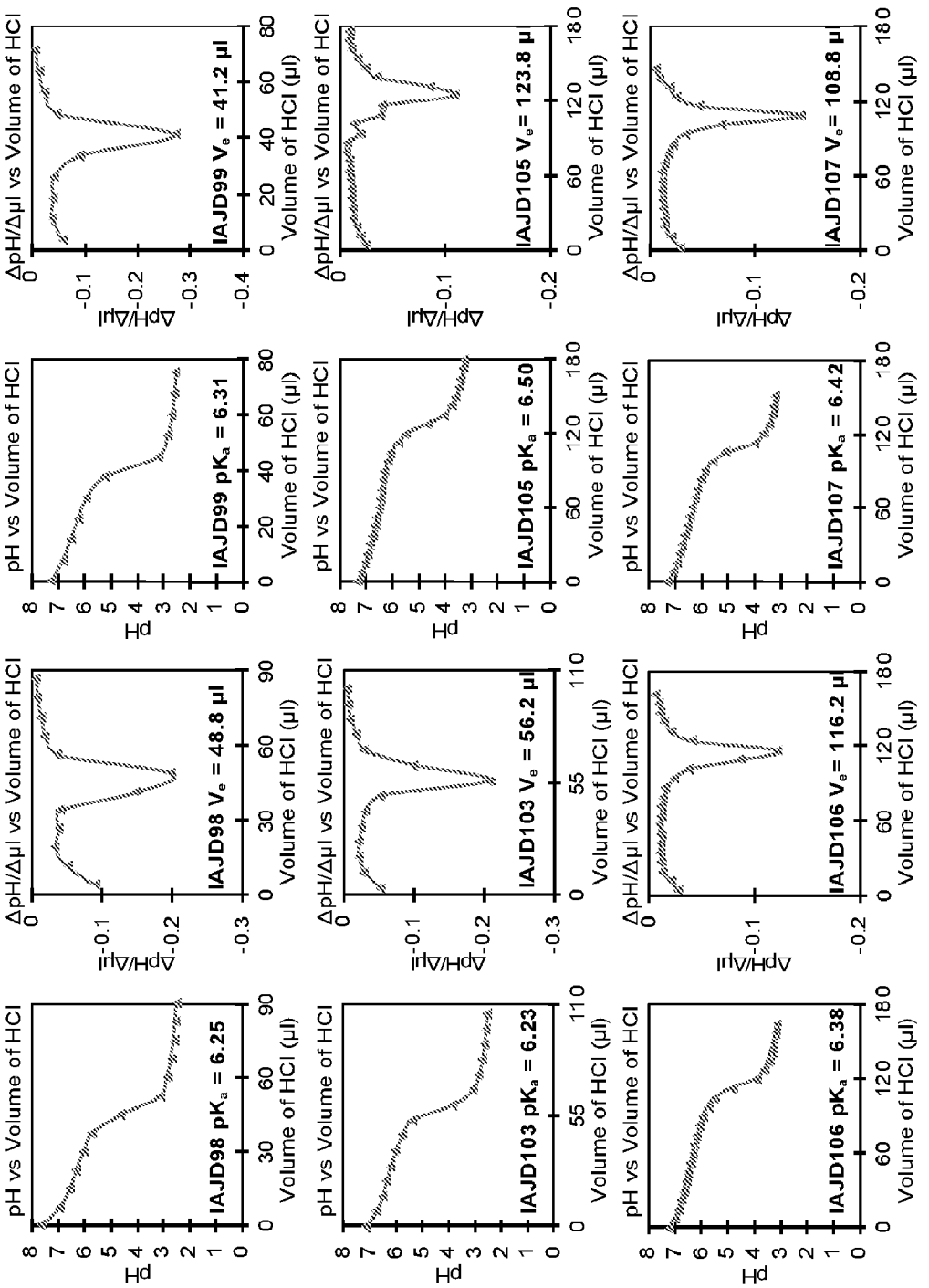

pH vs Volume of HCl
pH
Volume of HCl (µl)
IAJD98 pKa = 6.25
ΔpH/Δµl vs Volume of HCl
ΔpH/Δµl
IAJD98 Ve = 48.8 µl
IAJD99 pKa = 6.31
IAJD99 Ve = 41.2 µl
IAJD103 pKa = 6.23
IAJD103 Ve = 56.2 µl
IAJD105 pKa = 6.50
IAJD105 Ve = 123.8 µ
IAJD106 pKa = 6.38
IAJD106 Ve = 116.2 µl
IAJD107 pKa = 6.42
IAJD107 Ve = 108.8 µl
IAJD108 pKa = 6.59
IAJD108 Ve = 123.8 µl

Figure 57

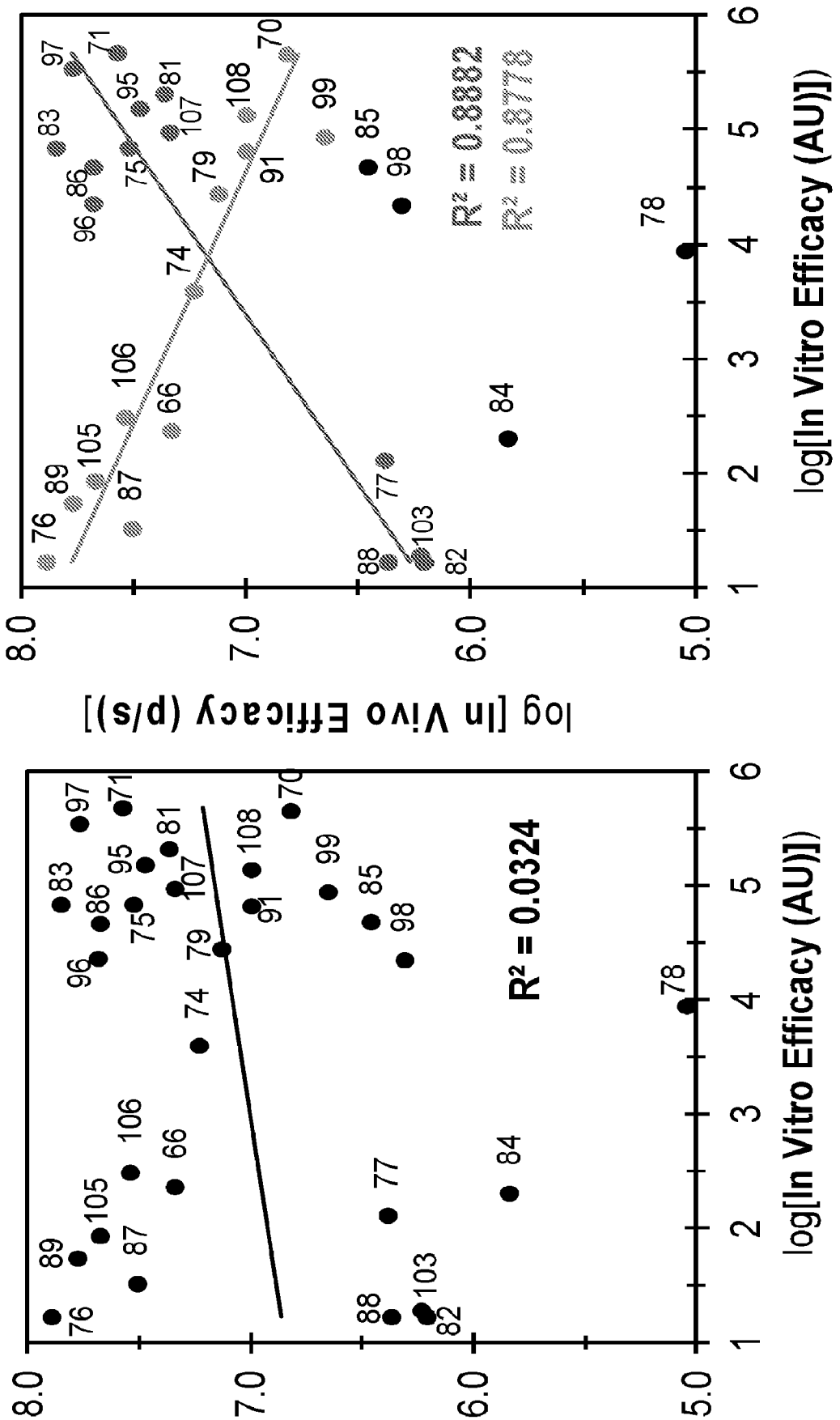

log [In Vivo Efficacy (p/s)]
log[In Vitro Efficacy (AU)]
R² = 0.0324
R² = 0.8882
R² = 0.8778

Figure 58

Synthesis of IAJDs with 3,5-Nonsymmetric Alkyl Chains:7 steps
1
i
2 (39%)
ii
3 (80-100%)
3
iii
4 (100%)
ii
5 (74-92%)
iv
6 (93-100%)
6
v/vi
7 (76-98%)
vii
8 (70-90%)
vii
9 (76-98%)
3-8: R1 = CnH2n+1 (n = 11,15)
R2 = CnH2n+1 (n = 8-18)
Reagents and Conditions: i) BnCl, K2CO3, KI, DMF, 80 °C, 5 h; ii) RBr, K2CO3, DMF, 120 °C, 2 h; iii) H2, Pd/C, DCM, MeOH, 12 h; iv) LiAlH4, THF, 0–23 °C, 1 h; v) 4-Bromobutyric acid, SOCl2, DMF (cat.), DCM, 23 °C, 1 h, then DMAP, NEt3, DCM, 0–23 °C, 2 h; vi) DCC, DPTS, 12 h; vii) K2CO3, MeCN, 95 °C, 3 h.

Figure 59

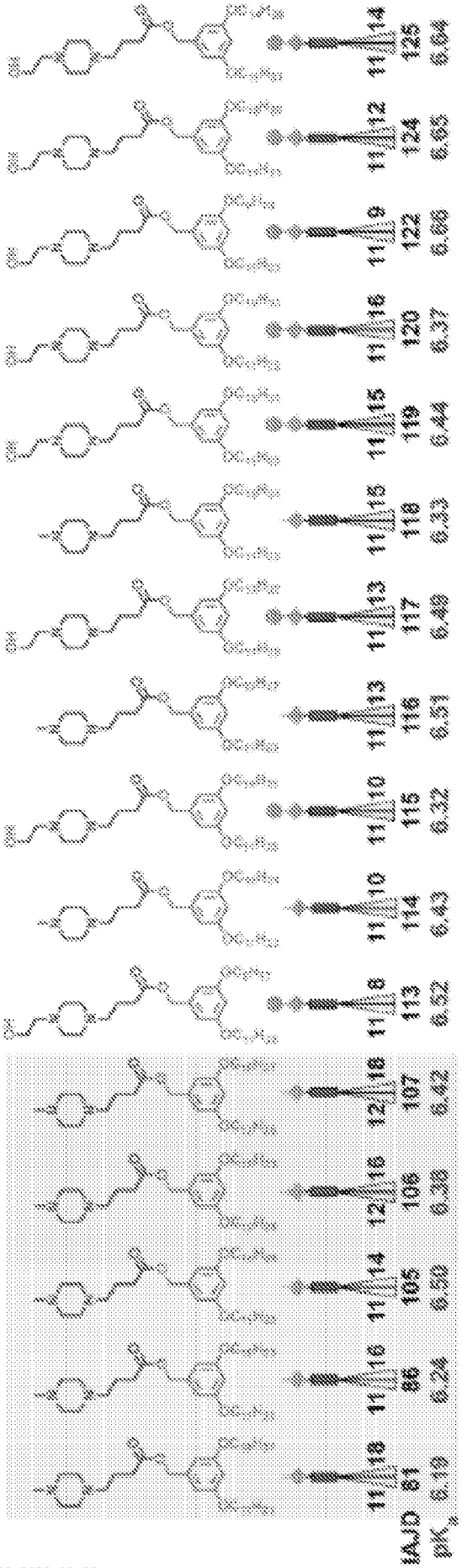

IAJD 81 86 105 106 107 113 114 115 116 117 118 119 120 122 124 125
pK$_a$ 6.19 6.24 6.50 6.38 6.42 6.52 6.43 6.32 6.51 6.49 6.33 6.44 6.37 6.66 6.65 6.64

Figure 60

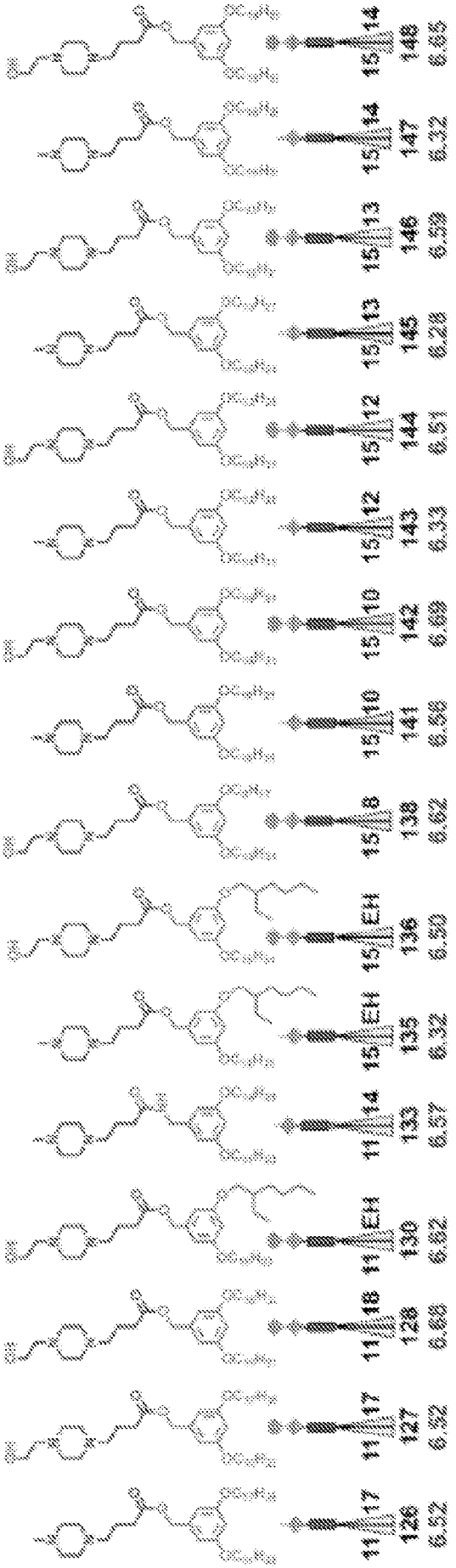


Figure 60 (cont.)

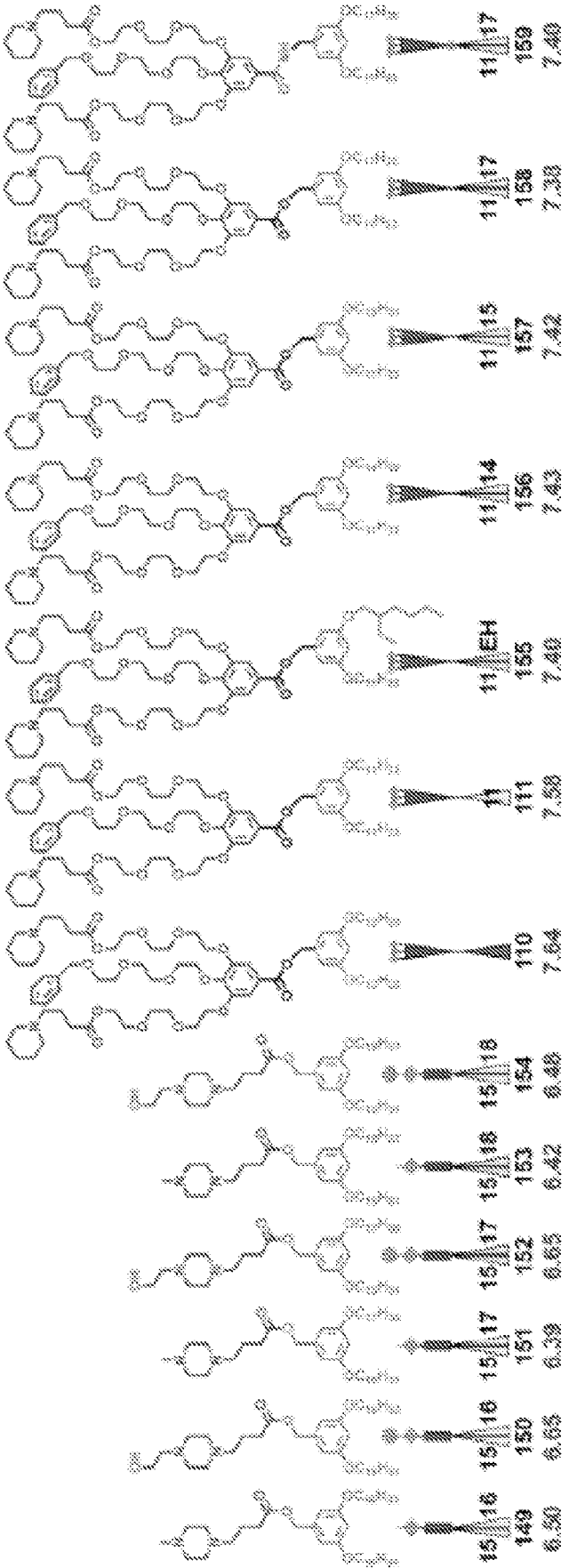


Figure 60 (cont.)

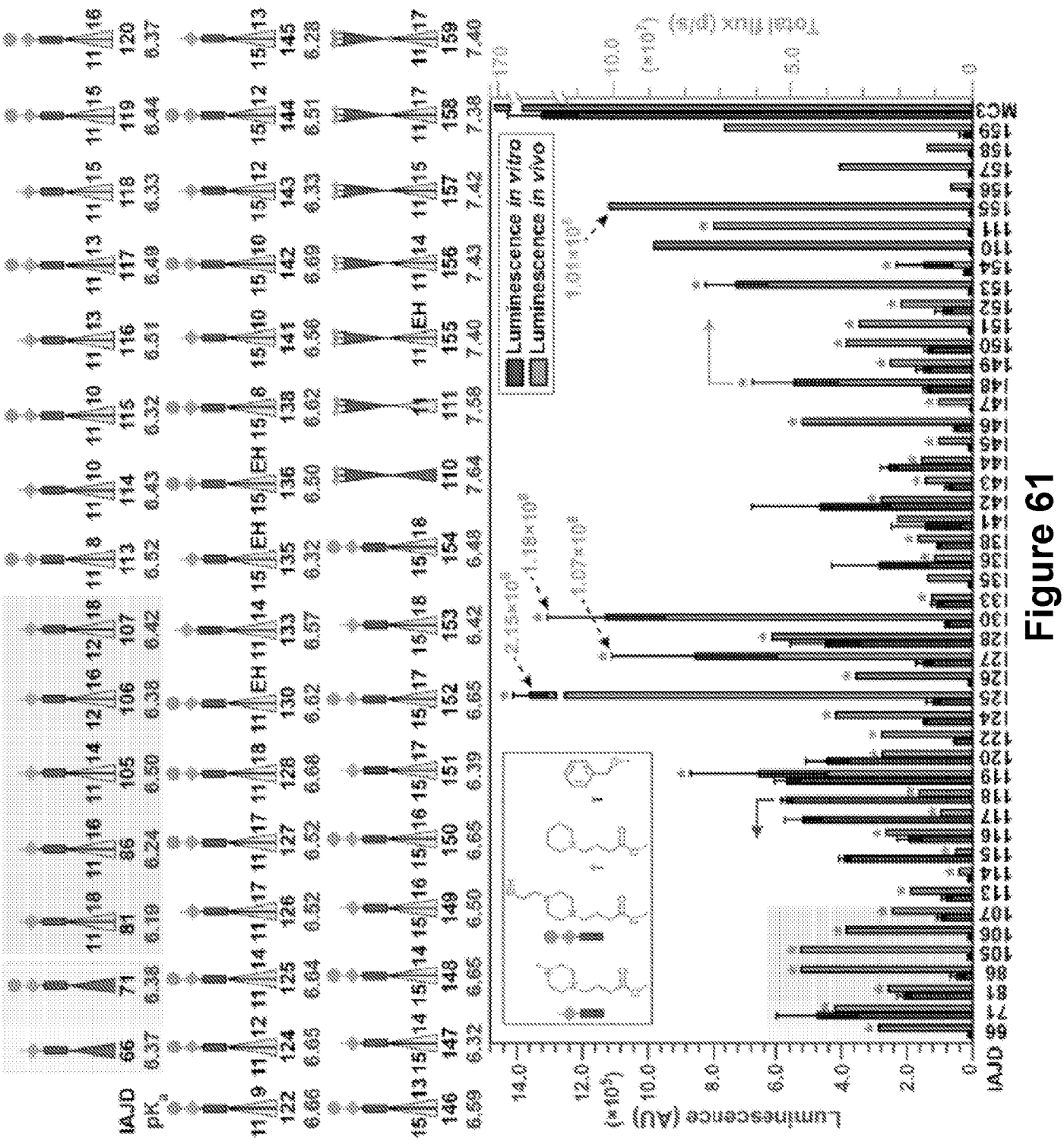

Luminescence in vitro
Luminescence in vivo
Luminescence (AU) (*10^5)
Total flux (p/s) (*10^7)
IAJD
pKa
2.15×10^8
1.18×10^8
1.07×10^8
1.08×10^9

Figure 61

a Selective delivery to lung: 8 hA3Bs
b Selective delivery to liver: 11 hA3Bs
c Selective delivery to spleen and lymph nodes: 26 hA3Bs
Luminescence
High
Low

Figure 62

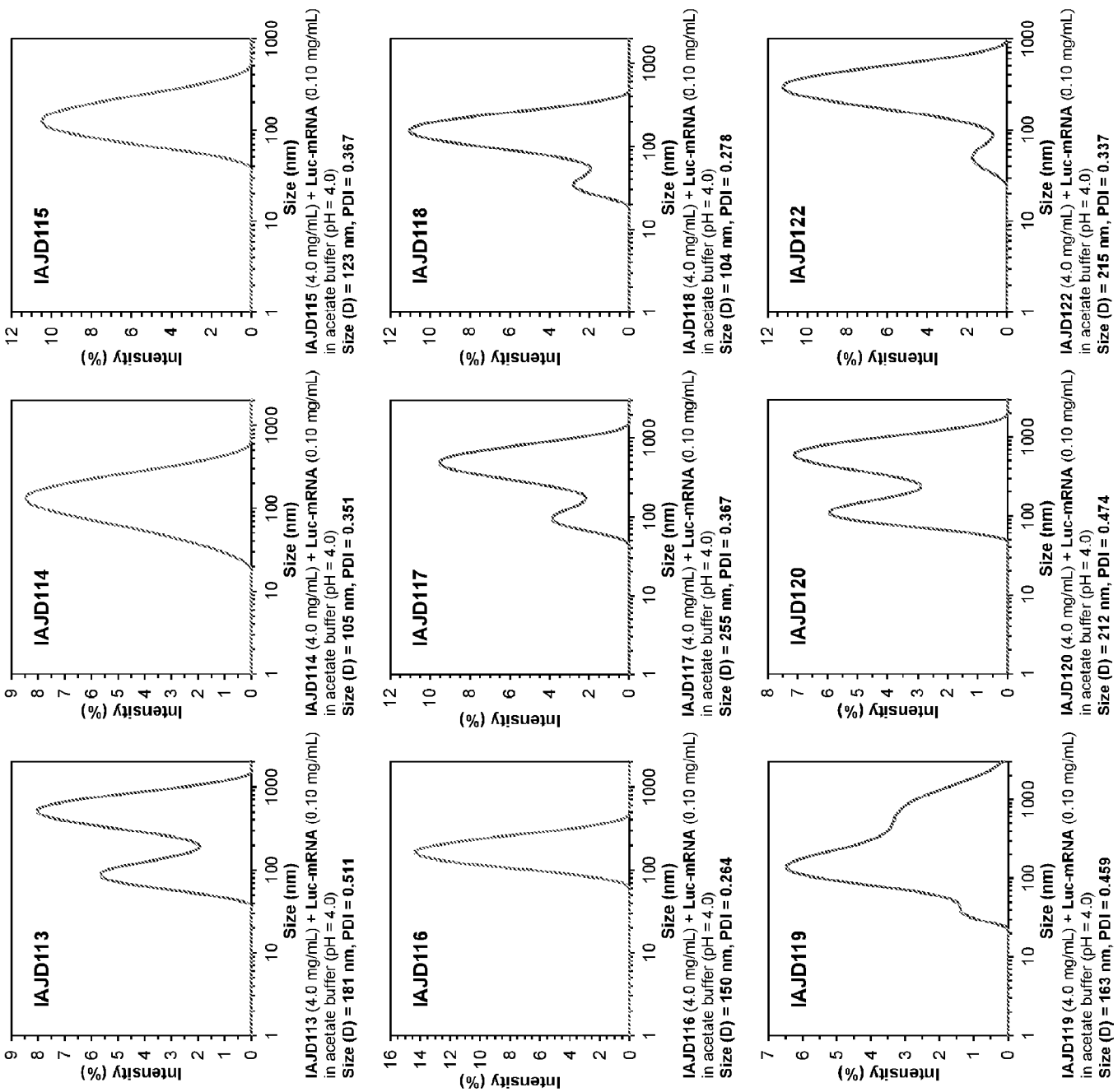

IAJD113
Intensity (%)
Size (nm)
IAJD113 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 181 nm, PDI = 0.511
IAJD114
Intensity (%)
Size (nm)
IAJD114 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 105 nm, PDI = 0.351
IAJD115
Intensity (%)
Size (nm)
IAJD115 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 123 nm, PDI = 0.367
IAJD116
Intensity (%)
Size (nm)
IAJD116 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 150 nm, PDI = 0.264
IAJD117
Intensity (%)
Size (nm)
IAJD117 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 255 nm, PDI = 0.367
IAJD118
Intensity (%)
Size (nm)
IAJD118 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 104 nm, PDI = 0.278
IAJD119
Intensity (%)
Size (nm)
IAJD119 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 163 nm, PDI = 0.459
IAJD120
Intensity (%)
Size (nm)
IAJD120 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 212 nm, PDI = 0.474
IAJD122
Intensity (%)
Size (nm)
IAJD122 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 215 nm, PDI = 0.337

Figure 63

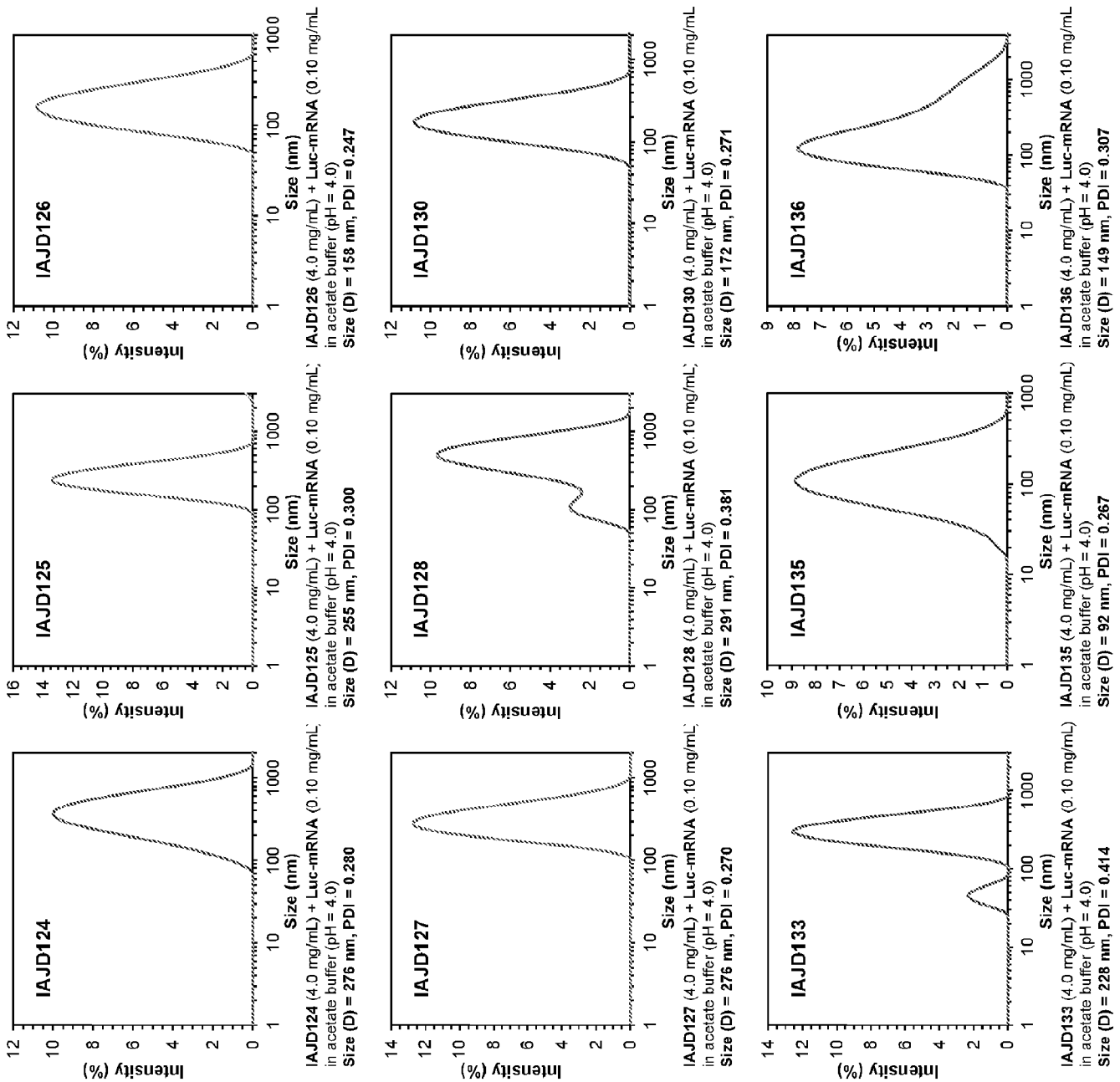

IAJD124
Intensity (%)
Size (nm)
IAJD124 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 276 nm, PDI = 0.280
IAJD125
Intensity (%)
Size (nm)
IAJD125 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 255 nm, PDI = 0.300
IAJD126
Intensity (%)
Size (nm)
IAJD126 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 158 nm, PDI = 0.247
IAJD127
Intensity (%)
Size (nm)
IAJD127 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 276 nm, PDI = 0.270
IAJD128
Intensity (%)
Size (nm)
IAJD128 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 291 nm, PDI = 0.381
IAJD130
Intensity (%)
Size (nm)
IAJD130 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 172 nm, PDI = 0.271
IAJD133
Intensity (%)
Size (nm)
IAJD133 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 228 nm, PDI = 0.414
IAJD135
Intensity (%)
Size (nm)
IAJD135 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 92 nm, PDI = 0.267
IAJD136
Intensity (%)
Size (nm)
IAJD136 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 149 nm, PDI = 0.307

Figure 64

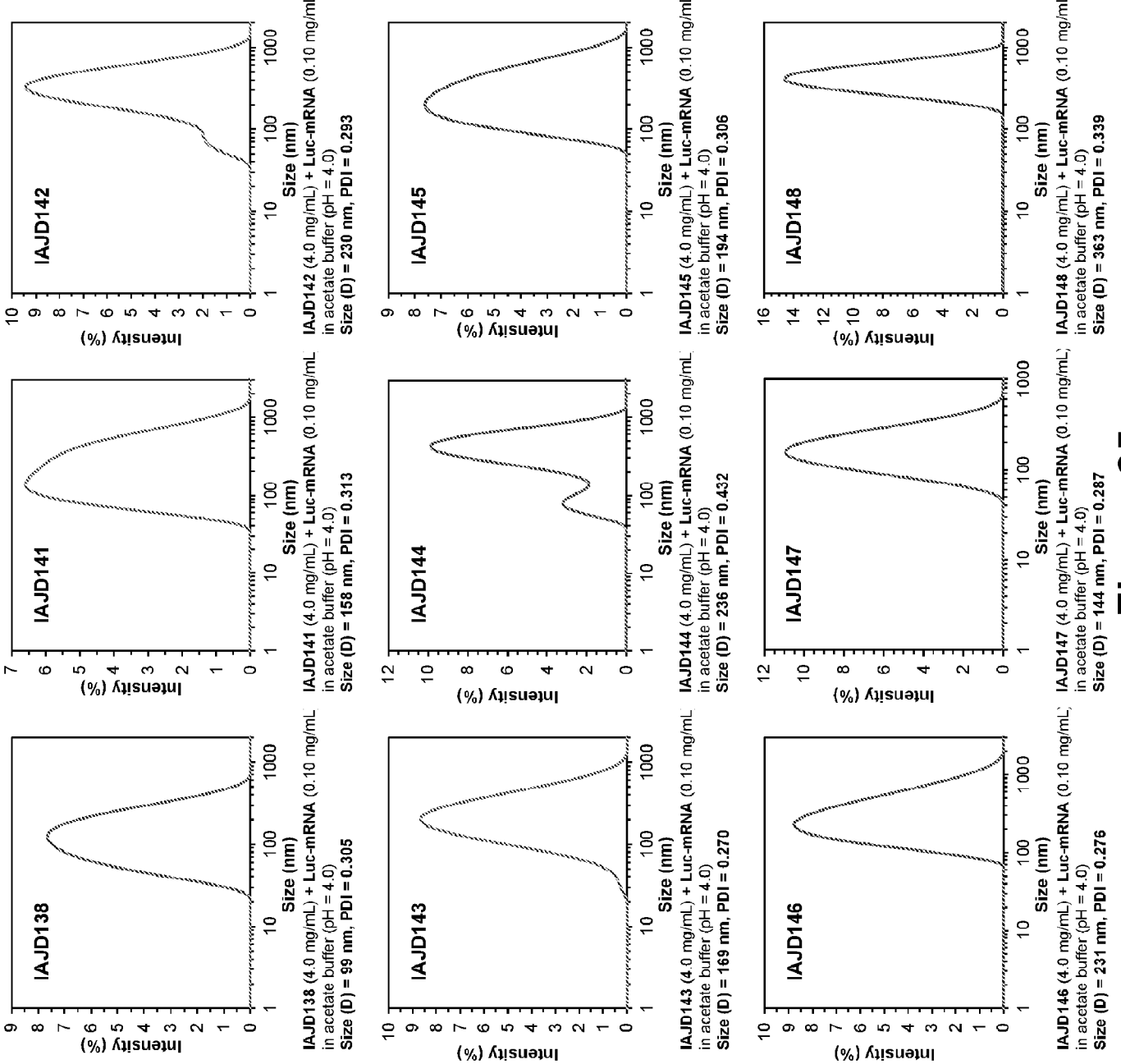

IAJD138
Intensity (%)
Size (nm)
IAJD138 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL in acetate buffer (pH = 4.0)
Size (D) = 99 nm, PDI = 0.305
IAJD141
Intensity (%)
Size (nm)
IAJD141 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL in acetate buffer (pH = 4.0)
Size (D) = 158 nm, PDI = 0.313
IAJD142
Intensity (%)
Size (nm)
IAJD142 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL in acetate buffer (pH = 4.0)
Size (D) = 230 nm, PDI = 0.293
IAJD143
Intensity (%)
Size (nm)
IAJD143 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL in acetate buffer (pH = 4.0)
Size (D) = 169 nm, PDI = 0.270
IAJD144
Intensity (%)
Size (nm)
IAJD144 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL in acetate buffer (pH = 4.0)
Size (D) = 236 nm, PDI = 0.432
IAJD145
Intensity (%)
Size (nm)
IAJD145 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL in acetate buffer (pH = 4.0)
Size (D) = 194 nm, PDI = 0.306
IAJD146
Intensity (%)
Size (nm)
IAJD146 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL in acetate buffer (pH = 4.0)
Size (D) = 231 nm, PDI = 0.276
IAJD147
Intensity (%)
Size (nm)
IAJD147 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL in acetate buffer (pH = 4.0)
Size (D) = 144 nm, PDI = 0.287
IAJD148
Intensity (%)
Size (nm)
IAJD148 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL in acetate buffer (pH = 4.0)
Size (D) = 363 nm, PDI = 0.339

Figure 65

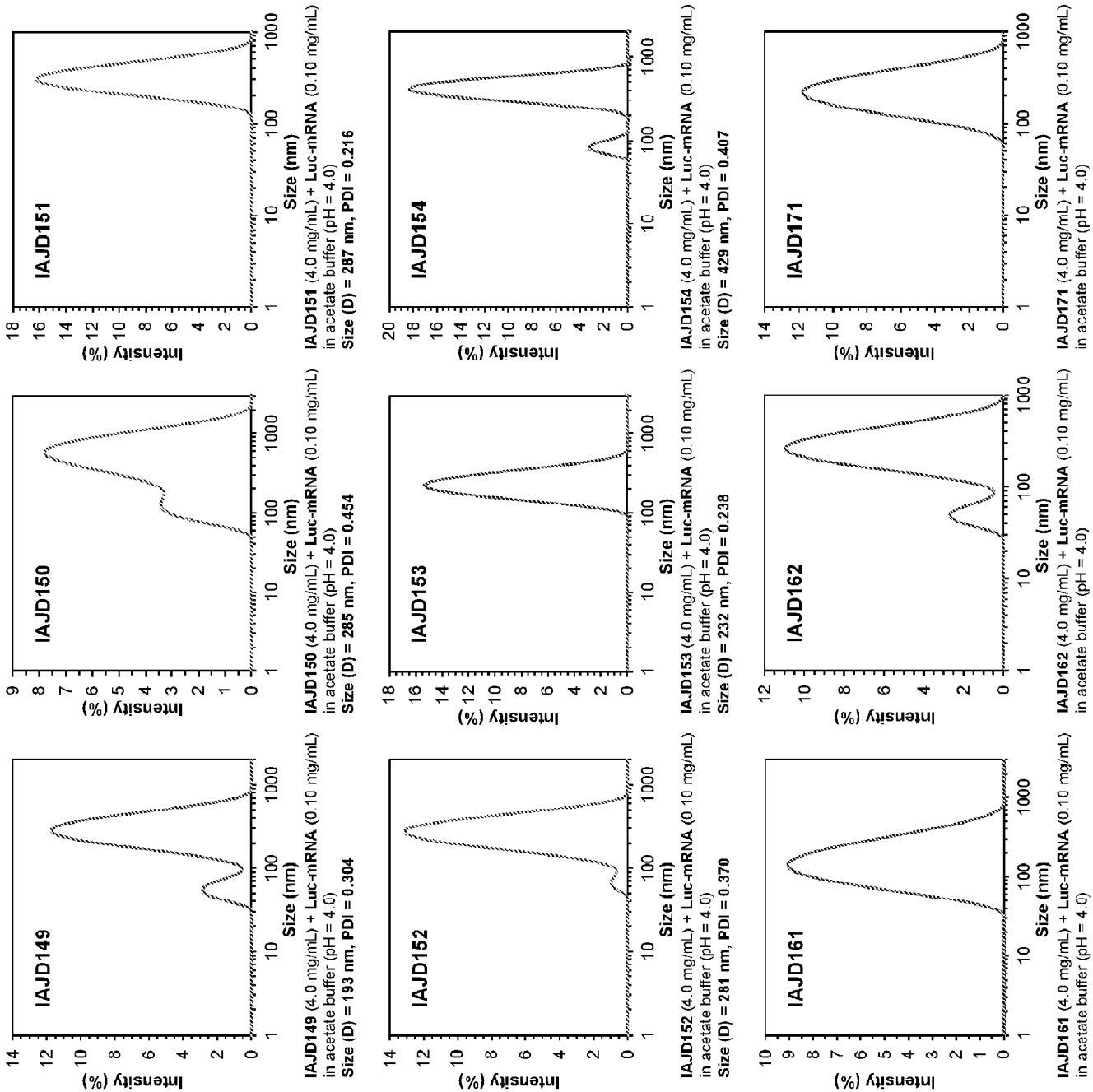

IAJD149
Intensity (%)
Size (nm)
IAJD149 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 193 nm, PDI = 0.304
IAJD150
Intensity (%)
Size (nm)
IAJD150 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 285 nm, PDI = 0.454
IAJD151
Intensity (%)
Size (nm)
IAJD151 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 287 nm, PDI = 0.216
IAJD152
Intensity (%)
Size (nm)
IAJD152 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 281 nm, PDI = 0.370
IAJD153
Intensity (%)
Size (nm)
IAJD153 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 232 nm, PDI = 0.238
IAJD154
Intensity (%)
Size (nm)
IAJD154 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 429 nm, PDI = 0.407
IAJD161
Intensity (%)
Size (nm)
IAJD161 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 129 nm, PDI = 0.235
IAJD162
Intensity (%)
Size (nm)
IAJD162 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 191 nm, PDI = 0.301
IAJD171
Intensity (%)
Size (nm)
IAJD171 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 174 nm, PDI = 0.268

Figure 66

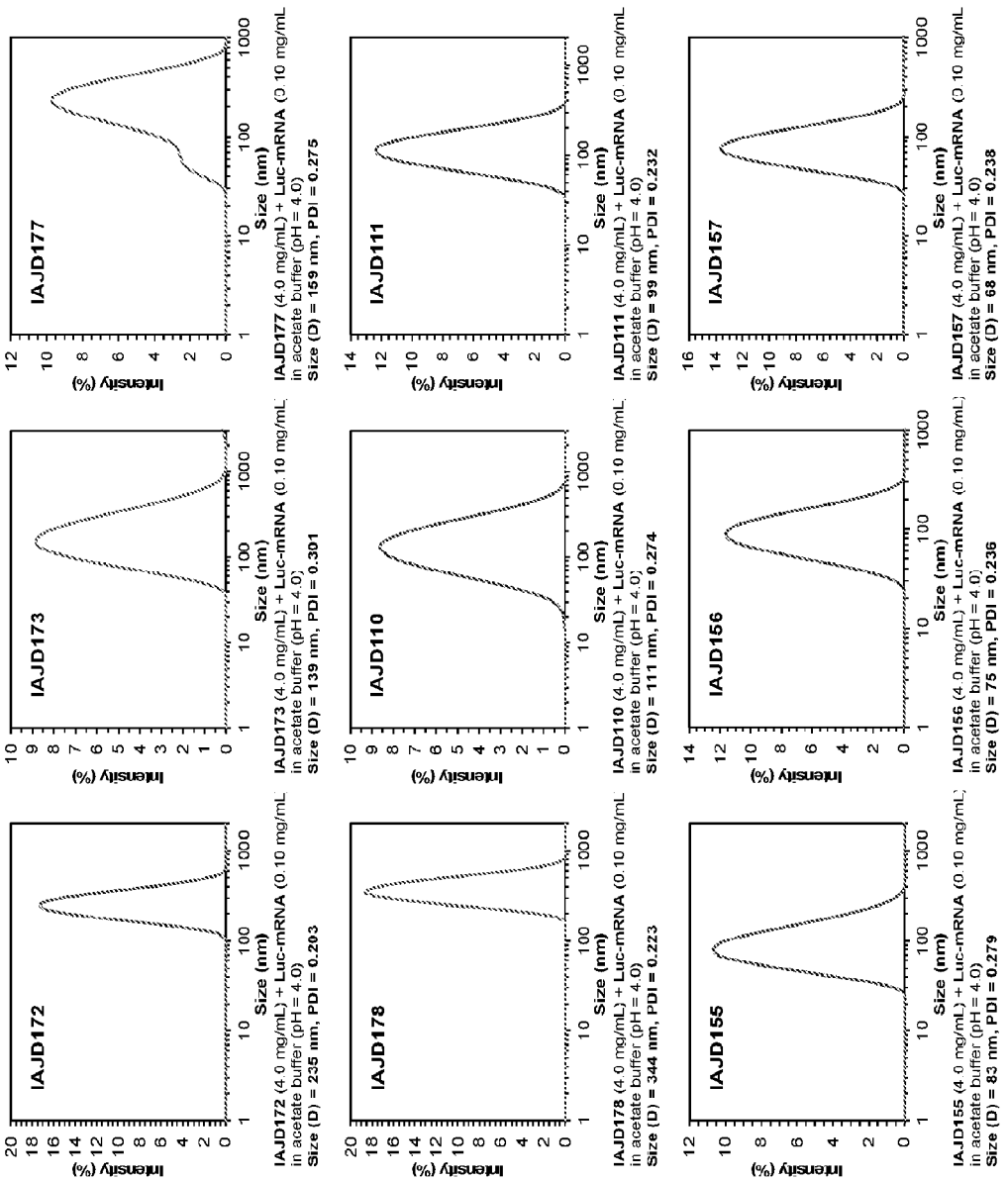

IAJD172
Intensity (%)
Size (nm)
IAJD172 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 235 nm, PDI = 0.203
IAJD173
Intensity (%)
Size (nm)
IAJD173 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 139 nm, PDI = 0.301
IAJD177
Intensity (%)
Size (nm)
IAJD177 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 159 nm, PDI = 0.275
IAJD178
Intensity (%)
Size (nm)
IAJD178 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 344 nm, PDI = 0.223
IAJD110
Intensity (%)
Size (nm)
IAJD110 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 111 nm, PDI = 0.274
IAJD111
Intensity (%)
Size (nm)
IAJD111 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 99 nm, PDI = 0.232
IAJD155
Intensity (%)
Size (nm)
IAJD155 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 83 nm, PDI = 0.279
IAJD156
Intensity (%)
Size (nm)
IAJD156 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 75 nm, PDI = 0.236
IAJD157
Intensity (%)
Size (nm)
IAJD157 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 68 nm, PDI = 0.238
IAJD158
Intensity (%)
Size (nm)
IAJD158 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 75 nm, PDI = 0.246
IAJD159
Intensity (%)
Size (nm)
IAJD159 (4.0 mg/mL) + Luc-mRNA (0.10 mg/mL) in acetate buffer (pH = 4.0)
Size (D) = 91 nm, PDI = 0.218

Figure 67

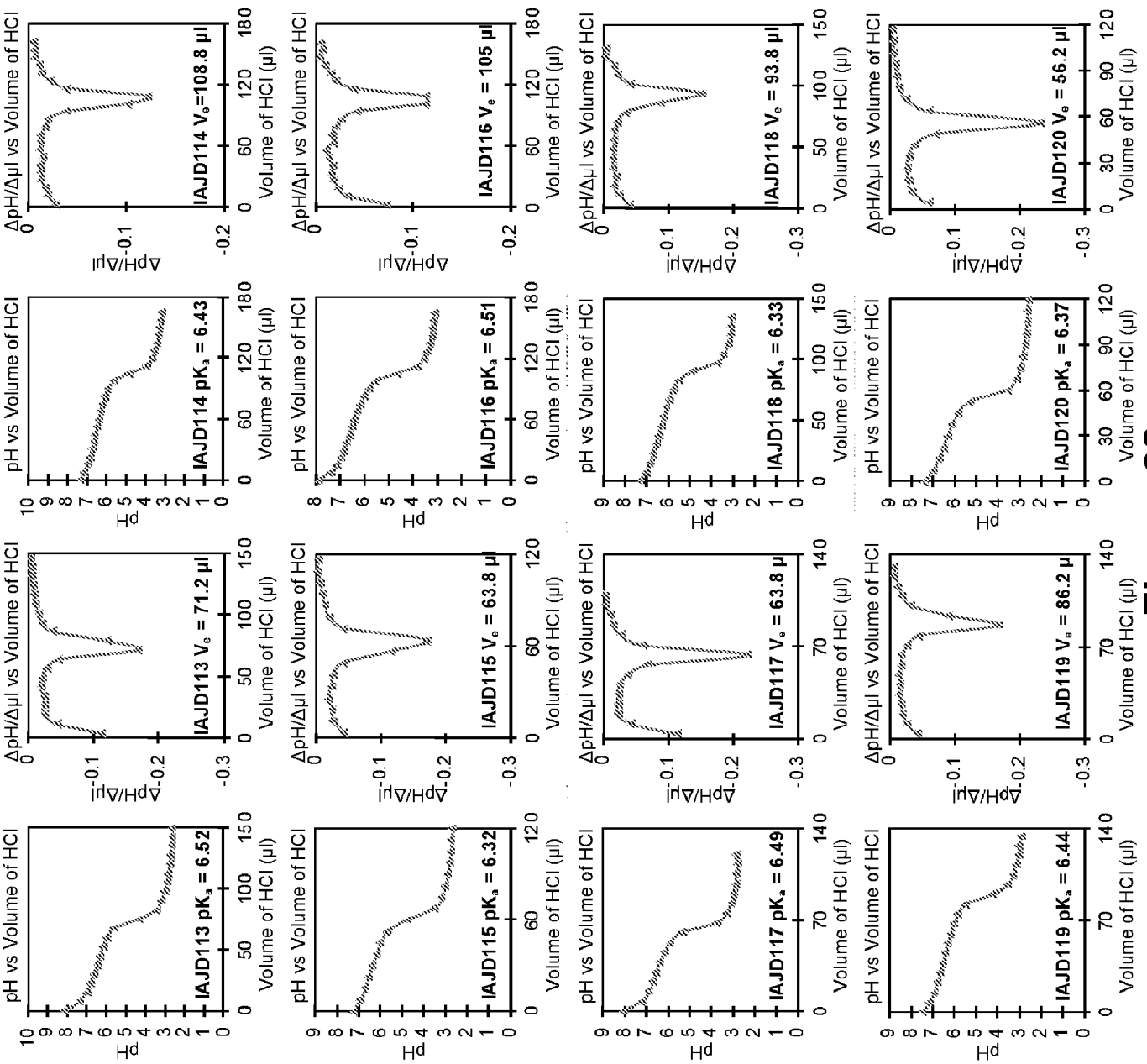

pH vs Volume of HCl
pH
Volume of HCl (µl)
IAJD113 pKa = 6.52
ΔpH/Δµl vs Volume of HCl
ΔpH/Δµl
IAJD113 Ve = 71.2 µl
IAJD114 pKa = 6.43
IAJD114 Ve=108.8 µl
IAJD115 pKa = 6.32
IAJD115 Ve = 63.8 µl
IAJD116 pKa = 6.51
IAJD116 Ve = 105 µl
IAJD117 pKa = 6.49
IAJD117 Ve = 63.8 µl
IAJD118 pKa = 6.33
IAJD118 Ve = 93.8 µl
IAJD119 pKa = 6.44
IAJD119 Ve = 86.2 µl
IAJD120 pKa = 6.37
IAJD120 Ve = 56.2 µl

Figure 68

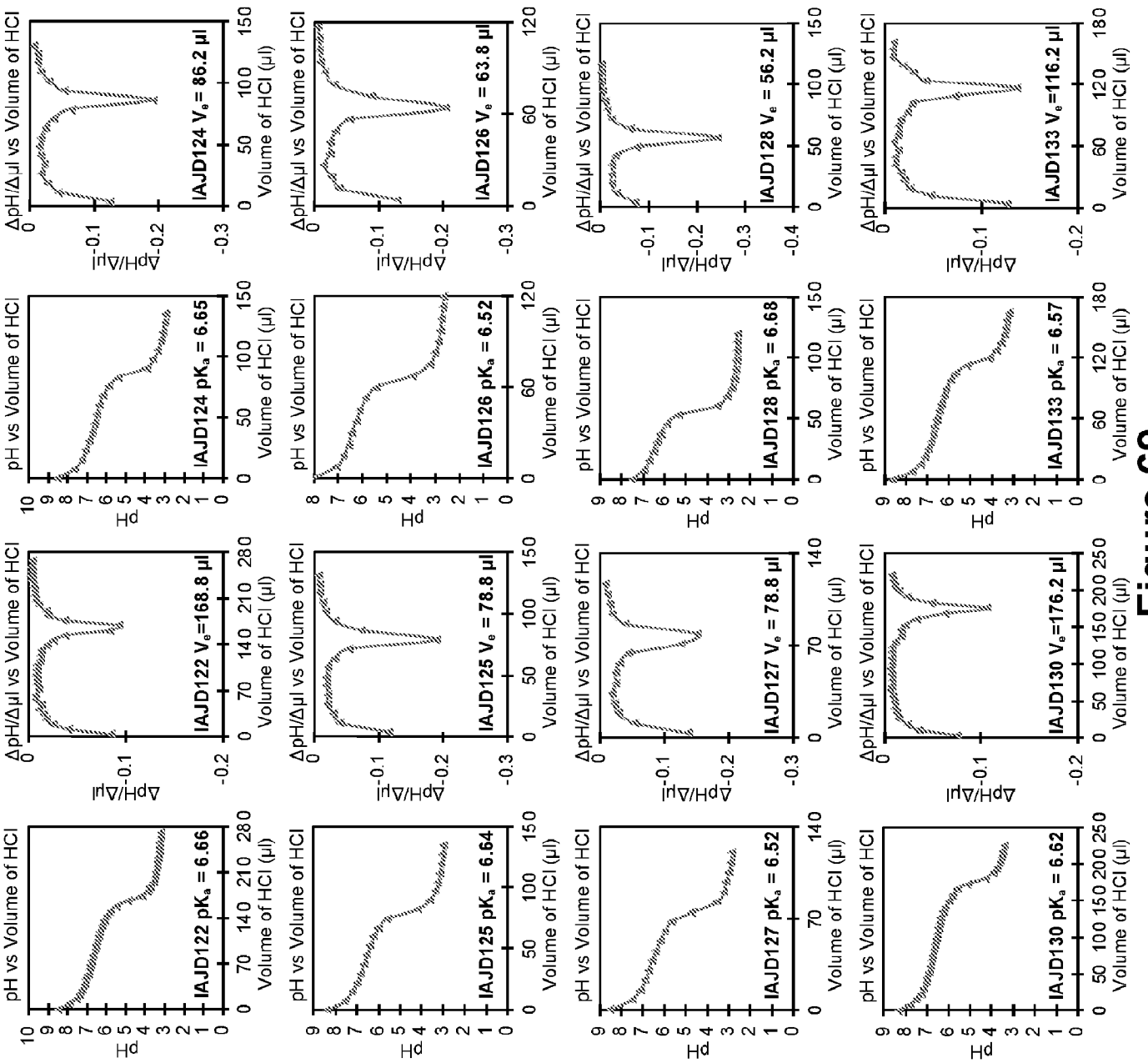

pH vs Volume of HCl
pH
Volume of HCl (µl)
IAJD122 pKa = 6.66
ΔpH/Δµl vs Volume of HCl
ΔpH/Δµl
IAJD122 Ve=168.8 µl
IAJD124 pKa = 6.65
IAJD124 Ve= 86.2 µl
IAJD125 pKa = 6.64
IAJD125 Ve = 78.8 µl
IAJD126 pKa = 6.52
IAJD126 Ve = 63.8 µl
IAJD127 pKa = 6.52
IAJD127 Ve = 78.8 µl
IAJD128 pKa = 6.68
IAJD128 Ve = 56.2 µl
IAJD130 pKa = 6.62
IAJD130 Ve=176.2 µl
IAJD133 pKa = 6.57
IAJD133 Ve=116.2 µl

Figure 69

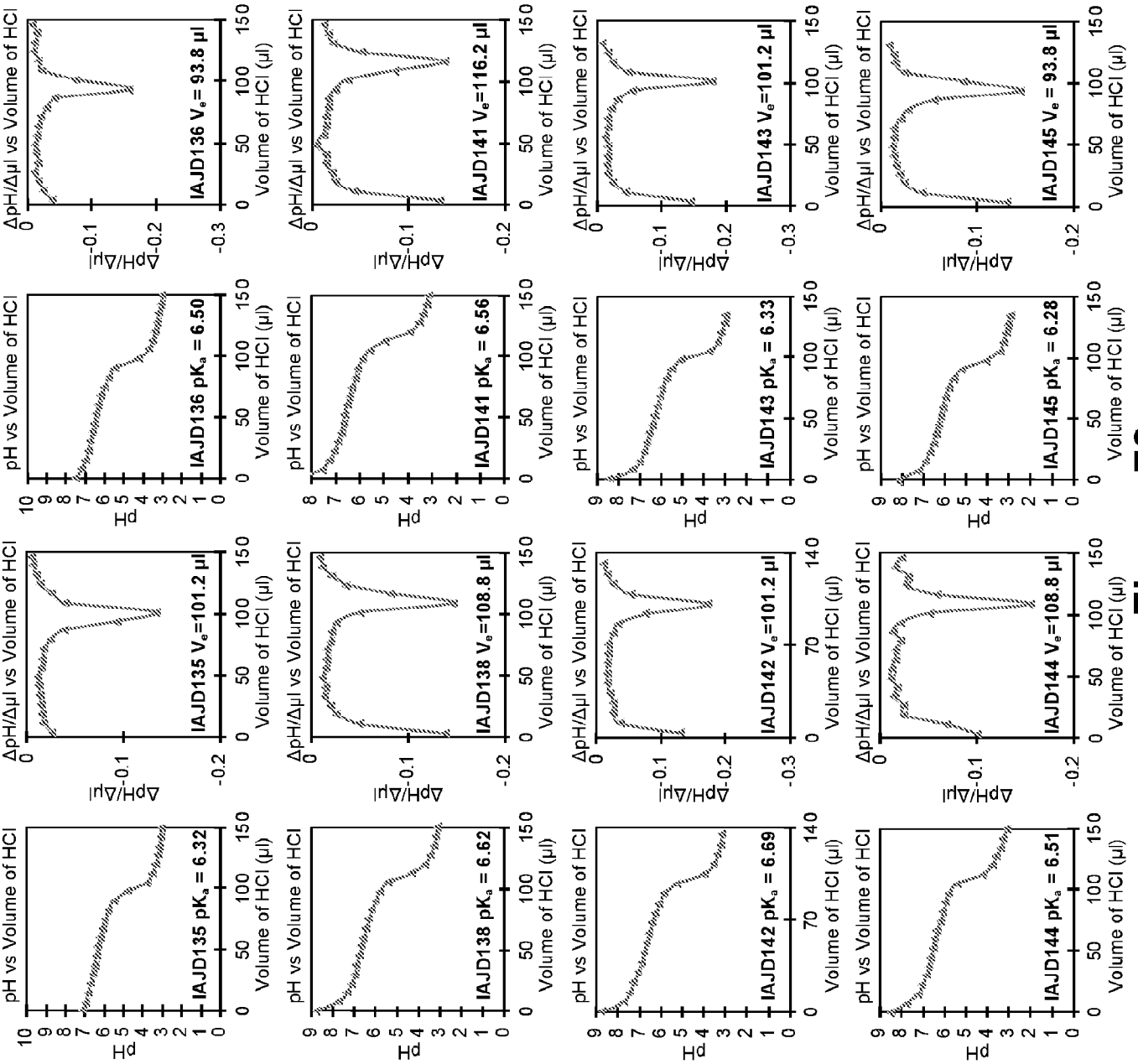

pH vs Volume of HCl
pH
Volume of HCl (µl)
IAJD135 pKa = 6.32
ΔpH/Δµl vs Volume of HCl
ΔpH/Δµl
IAJD135 Ve=101.2 µl
IAJD136 pKa = 6.50
IAJD136 Ve= 93.8 µl
IAJD138 pKa = 6.62
IAJD138 Ve=108.8 µl
IAJD141 pKa = 6.56
IAJD141 Ve=116.2 µl
IAJD142 pKa = 6.69
IAJD142 Ve=101.2 µl
IAJD143 pKa = 6.33
IAJD143 Ve=101.2 µl
IAJD144 pKa = 6.51
IAJD144 Ve=108.8 µl
IAJD145 pKa = 6.28
IAJD145 Ve = 93.8 µl

Figure 70

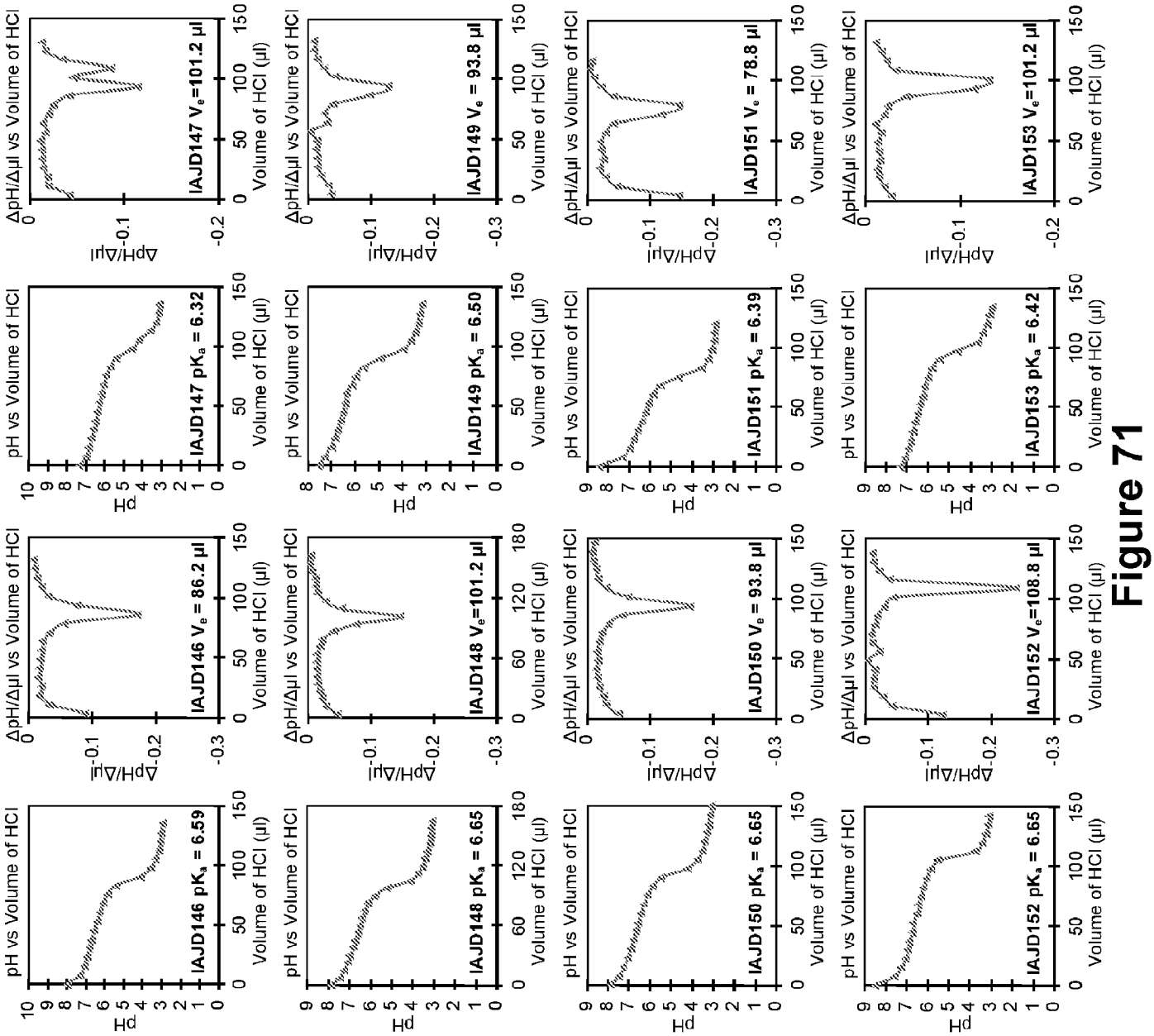

pH vs Volume of HCl
ΔpH/Δμl vs Volume of HCl
Volume of HCl (μl)
pH
ΔpH/Δμl
IAJD146 pKa = 6.59
IAJD146 Ve = 86.2 μl
IAJD147 pKa = 6.32
IAJD147 Ve=101.2 μl
IAJD148 pKa = 6.65
IAJD148 Ve=101.2 μl
IAJD149 pKa = 6.50
IAJD149 Ve = 93.8 μl
IAJD150 pKa = 6.65
IAJD150 Ve = 93.8 μl
IAJD151 pKa = 6.39
IAJD151 Ve = 78.8 μl
IAJD152 pKa = 6.65
IAJD152 Ve=108.8 μl
IAJD153 pKa = 6.42
IAJD153 Ve=101.2 μl

Figure 71

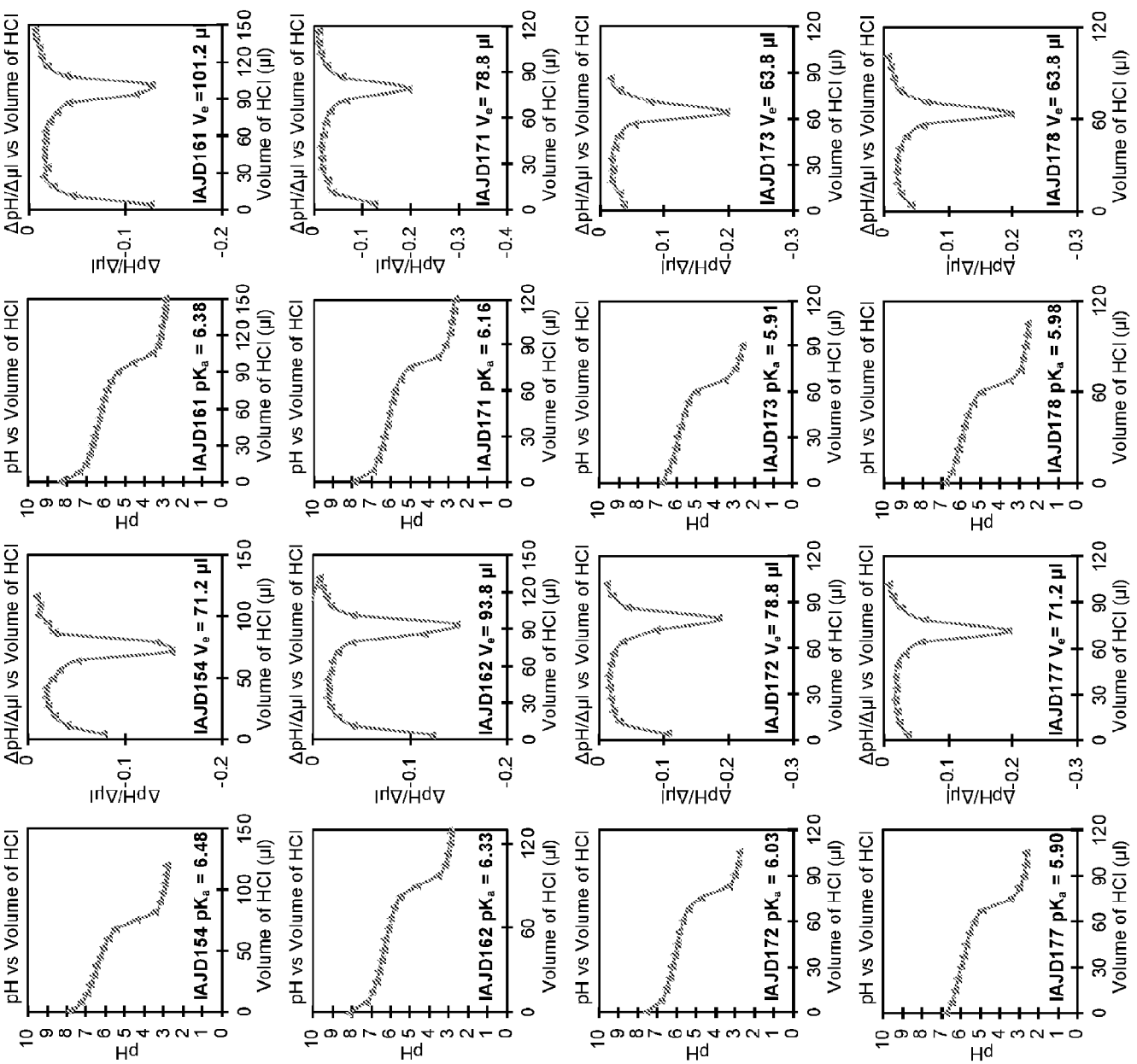

pH vs Volume of HCl
pH
Volume of HCl (µl)
IAJD154 pKa = 6.48
ΔpH/Δµl vs Volume of HCl
ΔpH/Δµl
IAJD154 Ve = 71.2 µl
IAJD161 pKa = 6.38
IAJD161 Ve =101.2 µl
IAJD162 pKa = 6.33
IAJD162 Ve= 93.8 µl
IAJD171 pKa = 6.16
IAJD171 Ve= 78.8 µl
IAJD172 pKa = 6.03
IAJD172 Ve= 78.8 µl
IAJD173 pKa = 5.91
IAJD173 Ve= 63.8 µl
IAJD177 pKa = 5.90
IAJD177 Ve= 71.2 µl
IAJD178 pKa = 5.98
IAJD178 Ve= 63.8 µl

Figure 72

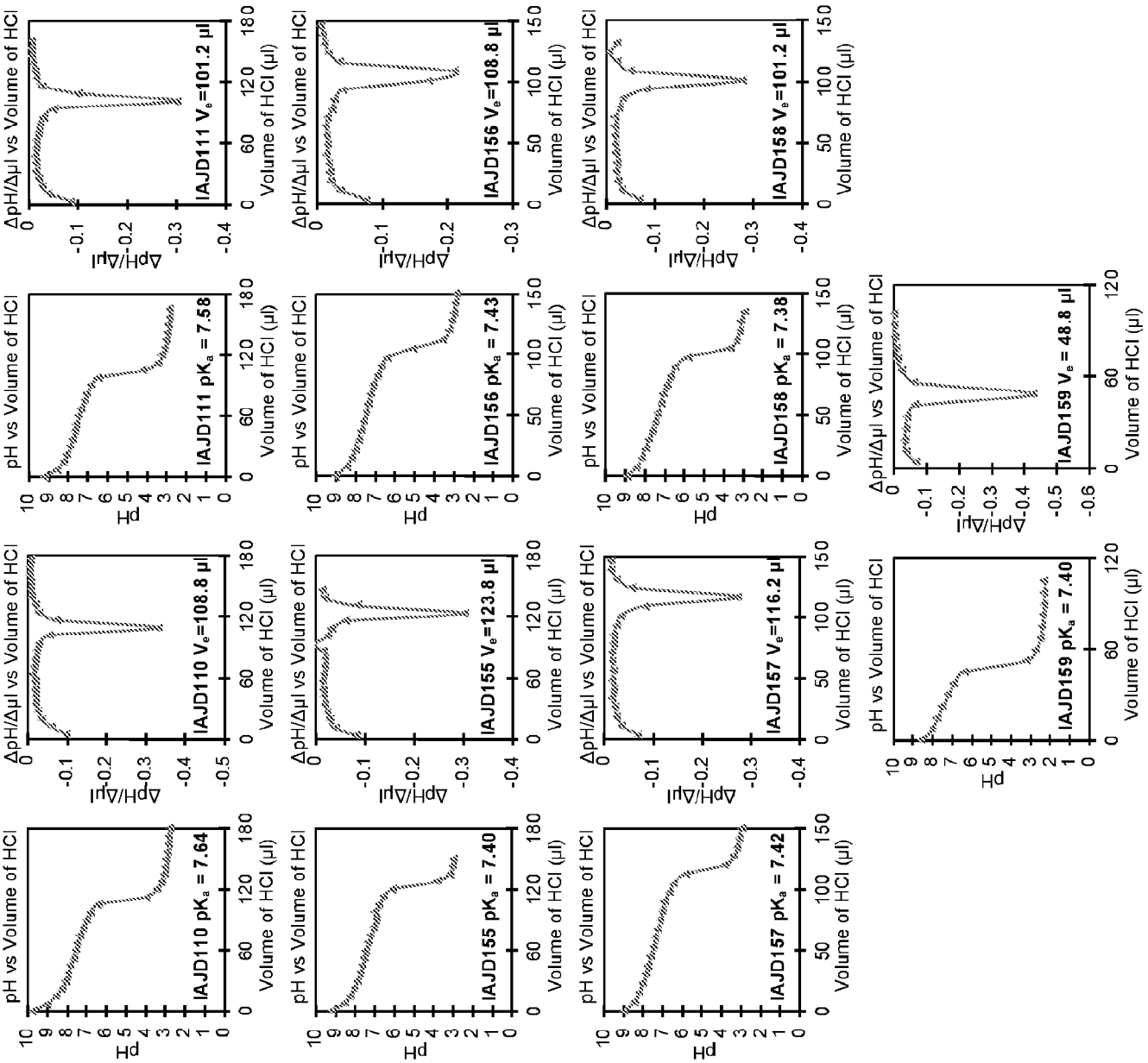


Figure 72 (cont.)

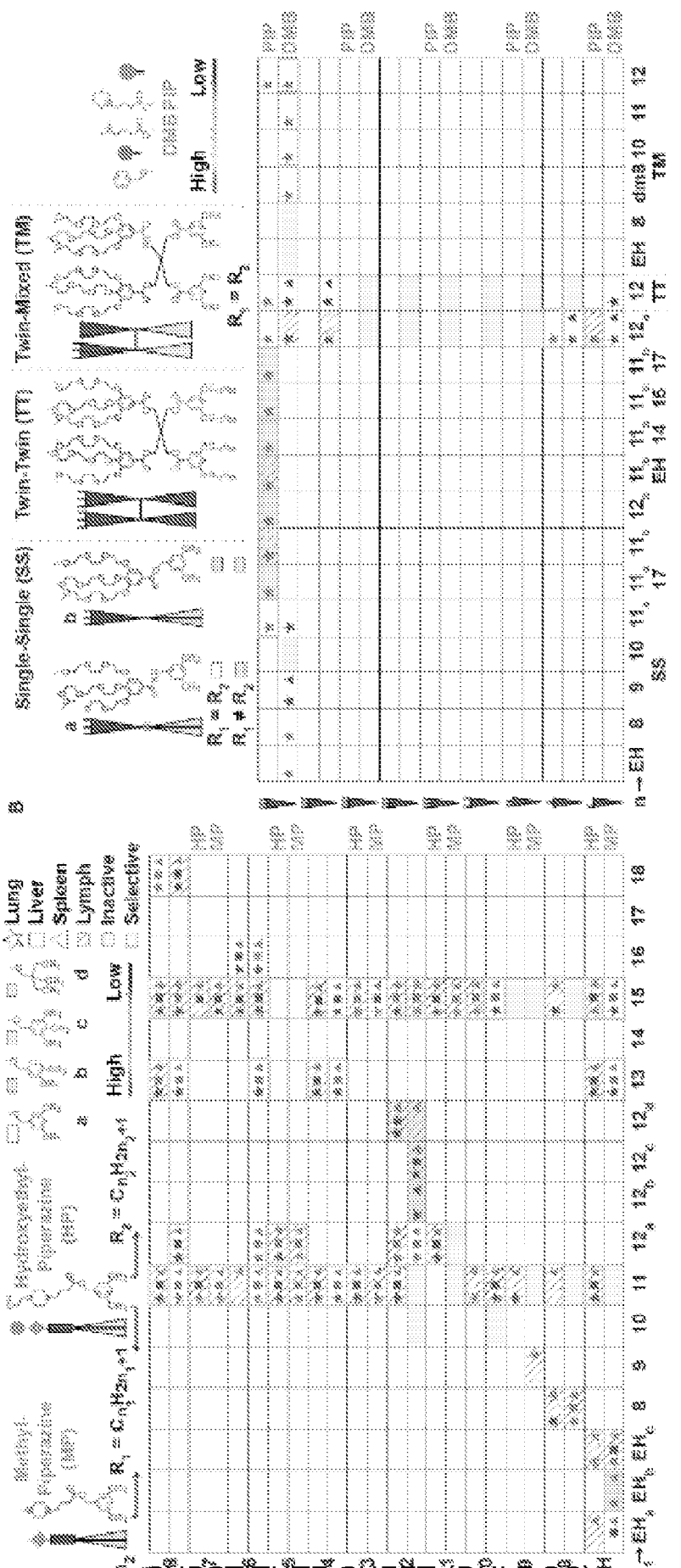


Figure 73

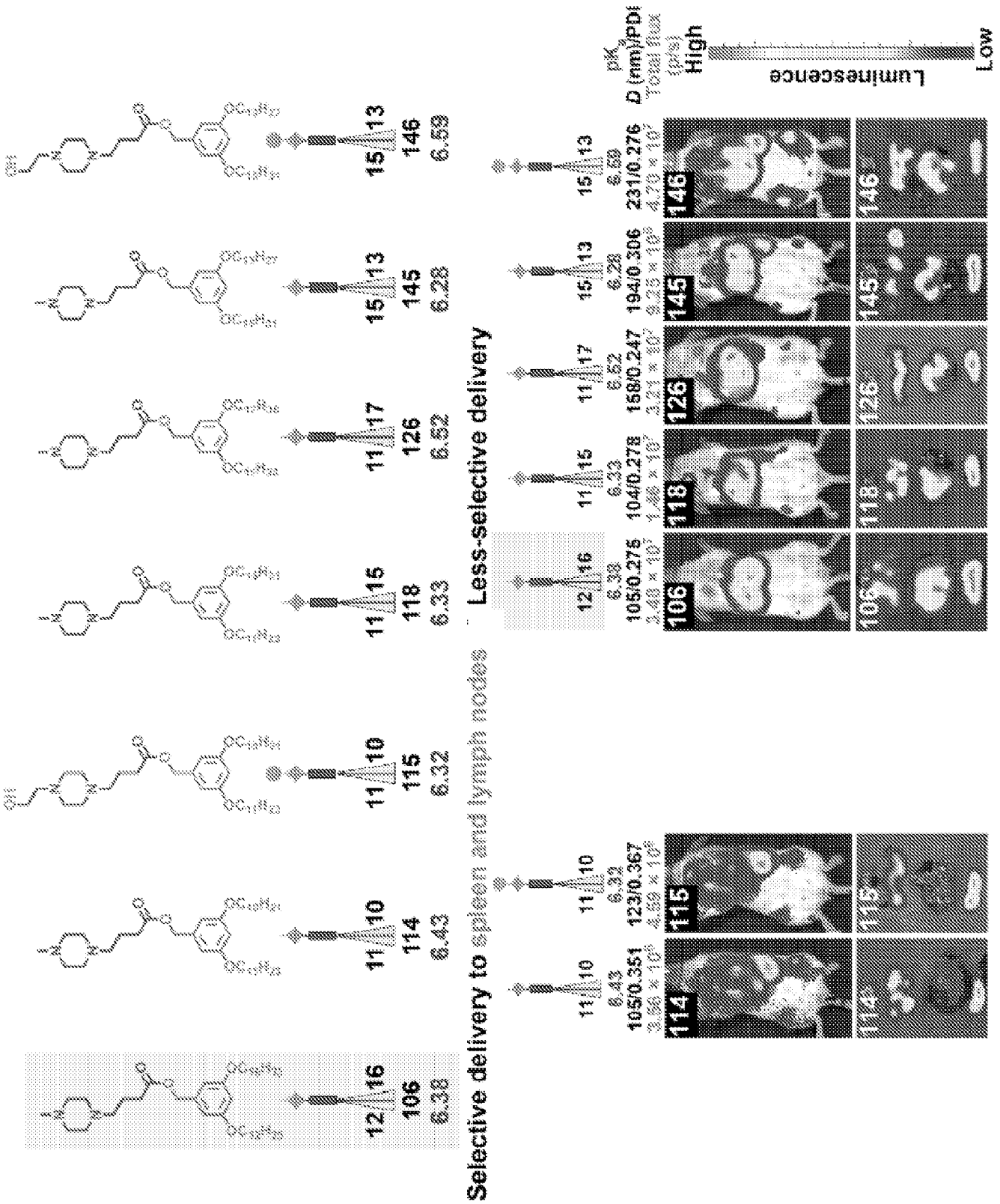

Selective delivery to spleen and lymph nodes
Less-selective delivery
12/16 106 6.38
11/10 114 6.43 105/0.351 3.66 × 10⁵
11/10 115 6.32 123/0.367 4.69 × 10⁵
11/15 118 6.33 104/0.278 1.48 × 10⁷
11/17 126 6.52 158/0.247 3.21 × 10⁷
15/13 145 6.28 194/0.306 9.18 × 10⁵
15/13 146 6.59 231/0.276 4.70 × 10⁷
pK
D (nm)/PDI
Total flux (p/s)
Luminescence
High
Low

Figure 74

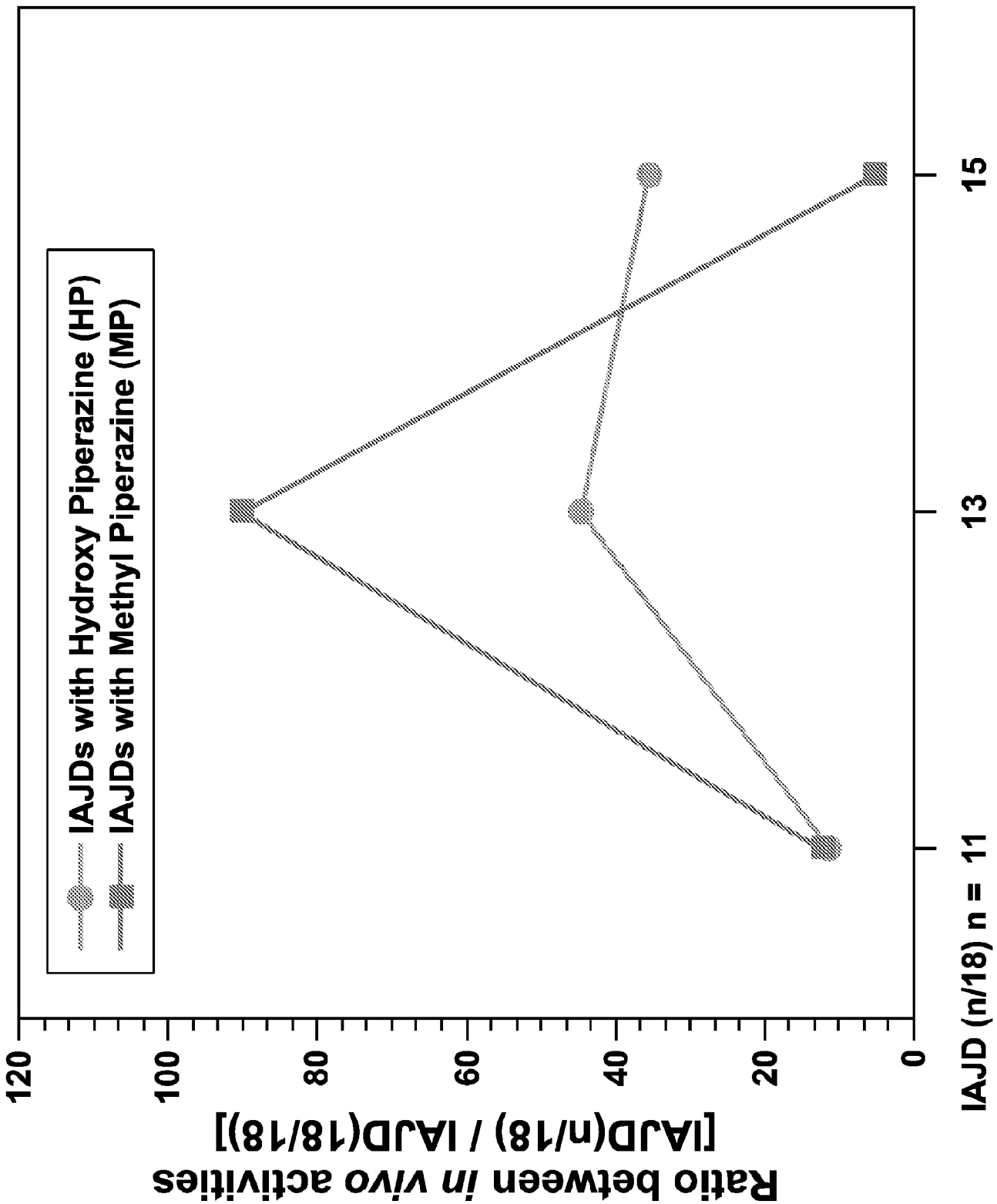

IAJDs with Hydroxy Piperazine (HP)
IAJDs with Methyl Piperazine (MP)
Ratio between in vivo activities
[IAJD(n/18) / IAJD(18/18)]
120
100
80
60
40
20
0
IAJD (n/18) n = 11
13
15

Figure 75

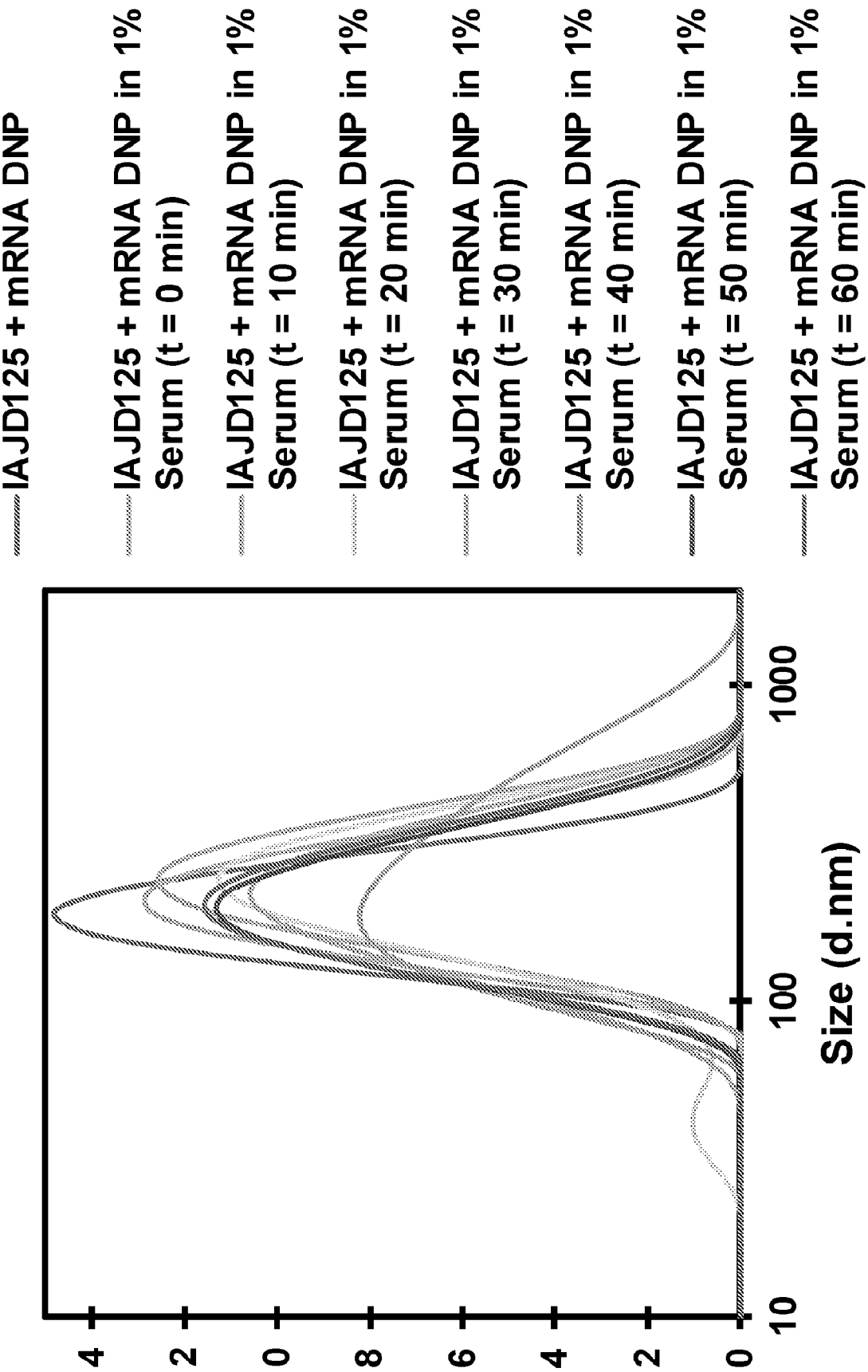

IAJD125 + mRNA DNP
IAJD125 + mRNA DNP in 1% Serum (t = 0 min)
IAJD125 + mRNA DNP in 1% Serum (t = 10 min)
IAJD125 + mRNA DNP in 1% Serum (t = 20 min)
IAJD125 + mRNA DNP in 1% Serum (t = 30 min)
IAJD125 + mRNA DNP in 1% Serum (t = 40 min)
IAJD125 + mRNA DNP in 1% Serum (t = 50 min)
IAJD125 + mRNA DNP in 1% Serum (t = 60 min)
Intensity (Percent)
0
2
4
6
8
10
12
14
10
100
1000
Size (d.nm)

Figure 76

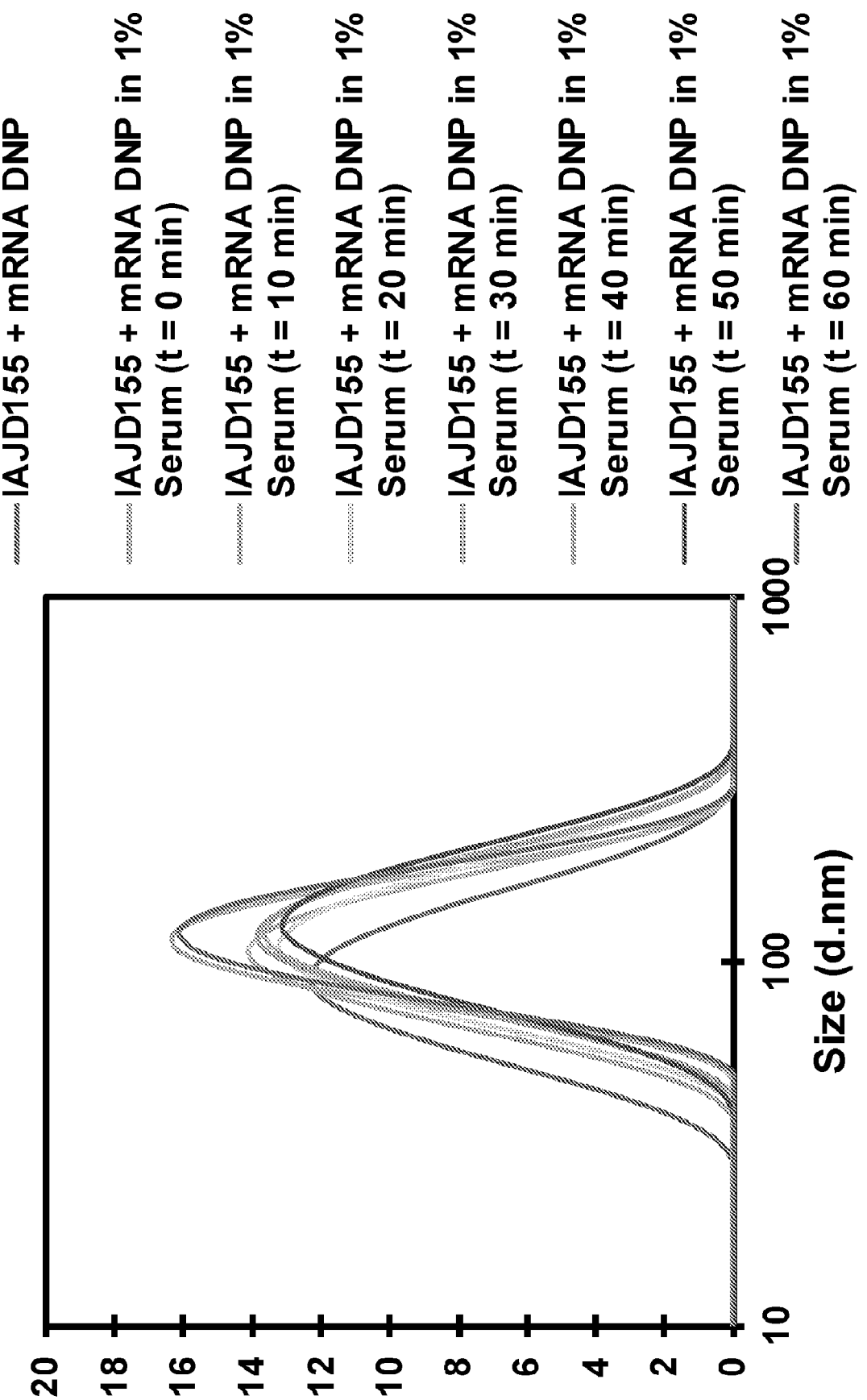

IAJD155 + mRNA DNP
IAJD155 + mRNA DNP in 1% Serum (t = 0 min)
IAJD155 + mRNA DNP in 1% Serum (t = 10 min)
IAJD155 + mRNA DNP in 1% Serum (t = 20 min)
IAJD155 + mRNA DNP in 1% Serum (t = 30 min)
IAJD155 + mRNA DNP in 1% Serum (t = 40 min)
IAJD155 + mRNA DNP in 1% Serum (t = 50 min)
IAJD155 + mRNA DNP in 1% Serum (t = 60 min)
Intensity (Percent)
0
2
4
6
8
10
12
14
16
18
20
10
100
1000
Size (d.nm)

Figure 77

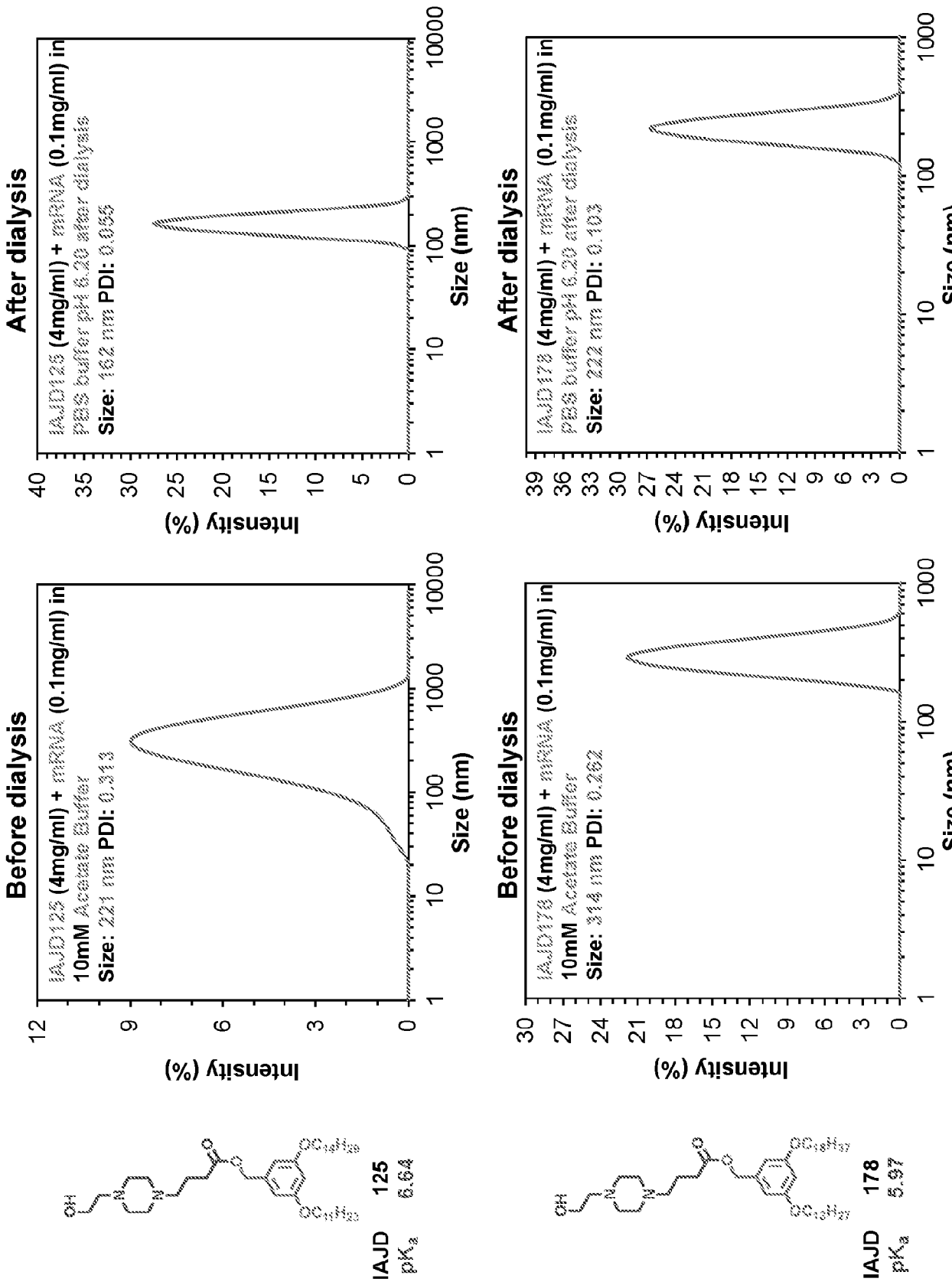

Before dialysis
IAJD123 (4mg/ml) + mRNA (0.1mg/ml) in 10mM Acetate Buffer
Size: 221 nm PDI: 0.313
Intensity (%)
Size (nm)
After dialysis
IAJD123 (4mg/ml) + mRNA (0.1mg/ml) in PBS buffer pH 6.20 after dialysis
Size: 162 nm PDI: 0.055
Intensity (%)
Size (nm)
IAJD 125
pKa 6.64
Before dialysis
IAJD178 (4mg/ml) + mRNA (0.1mg/ml) in 10mM Acetate Buffer
Size: 314 nm PDI: 0.262
Intensity (%)
Size (nm)
After dialysis
IAJD178 (4mg/ml) + mRNA (0.1mg/ml) in PBS buffer pH 6.20 after dialysis
Size: 222 nm PDI: 0.103
Intensity (%)
Size (nm)
IAJD 178
pKa 5.97

Figure 78

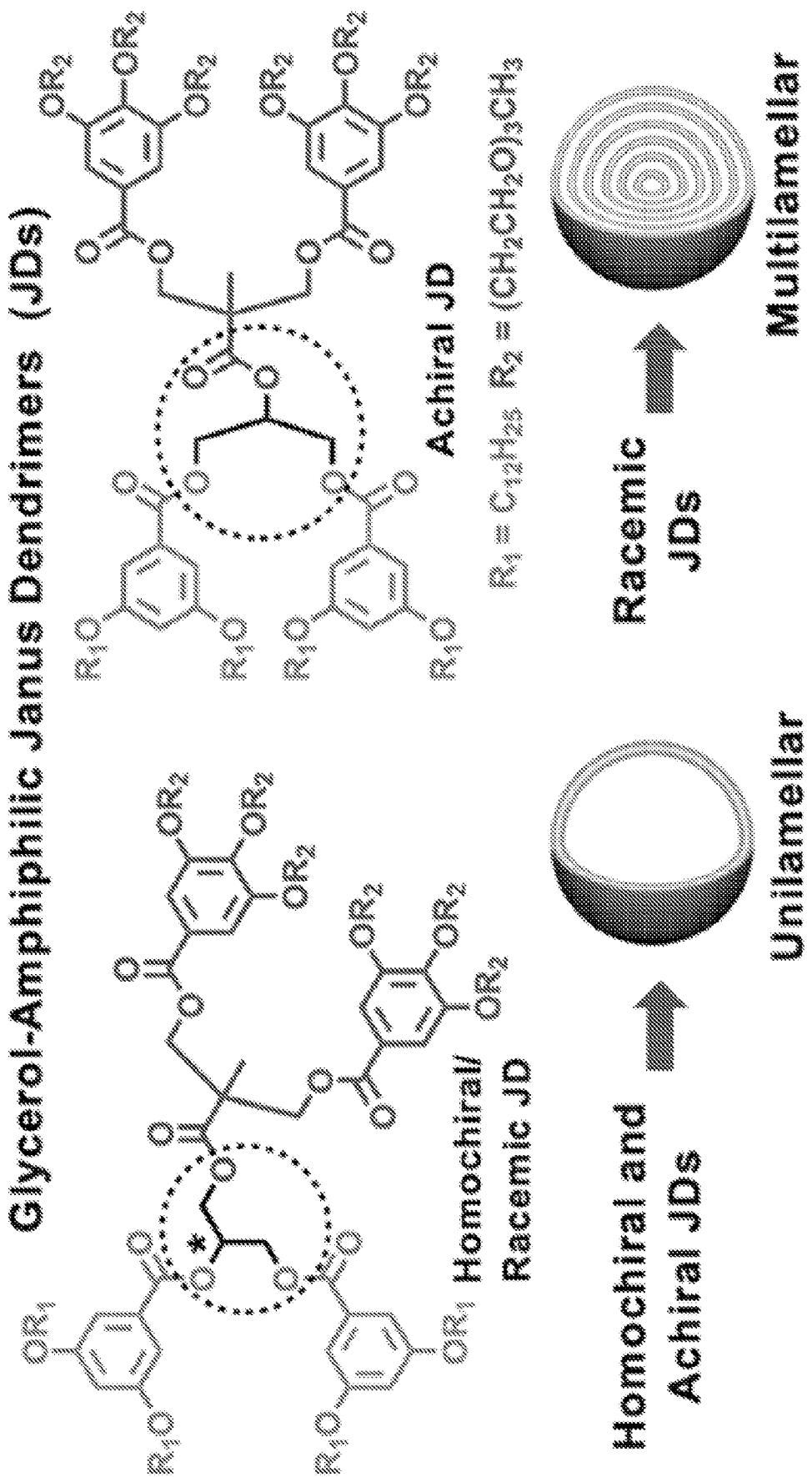

Glycerol-Amphiphilic Janus Dendrimers (JDs)
Achiral JD
$R_1 = C_{12}H_{25}$ $R_2 = (CH_2CH_2O)_3CH_3$
Homochiral/ Racemic JD
Homochiral and Achiral JDs
Racemic JDs
Unilamellar
Multilamellar

Figure 79

R
S
rac
achiral
1R
(3,5)-12G1-GC-(R)-BMPA-(3,4,5)-3EO-G2
1S
(3,5)-12G1-GC-(S)-BMPA-(3,4,5)-3EO-G2
1rac
(3,5)-12G1-GC-(rac)-BMPA-(3,4,5)-3EO-G2
2R
(3,5)-12G2-BMPA-GC-(R)-(3,4,5)-3EO-G1
2S
(3,5)-12G2-BMPA-GC-(S)-(3,4,5)-3EO-G1
2rac
(3,5)-12G2-BMPA-GC-(rac)-(3,4,5)-3EO-G1
3
(3,5)-12G1-GC-(achiral)-BMPA-(3,4,5)-3EO-G2
4
(3,5)-12G2-BMPA-GC-(achiral)-(3,4,5)-3EO-G1

Figure 80

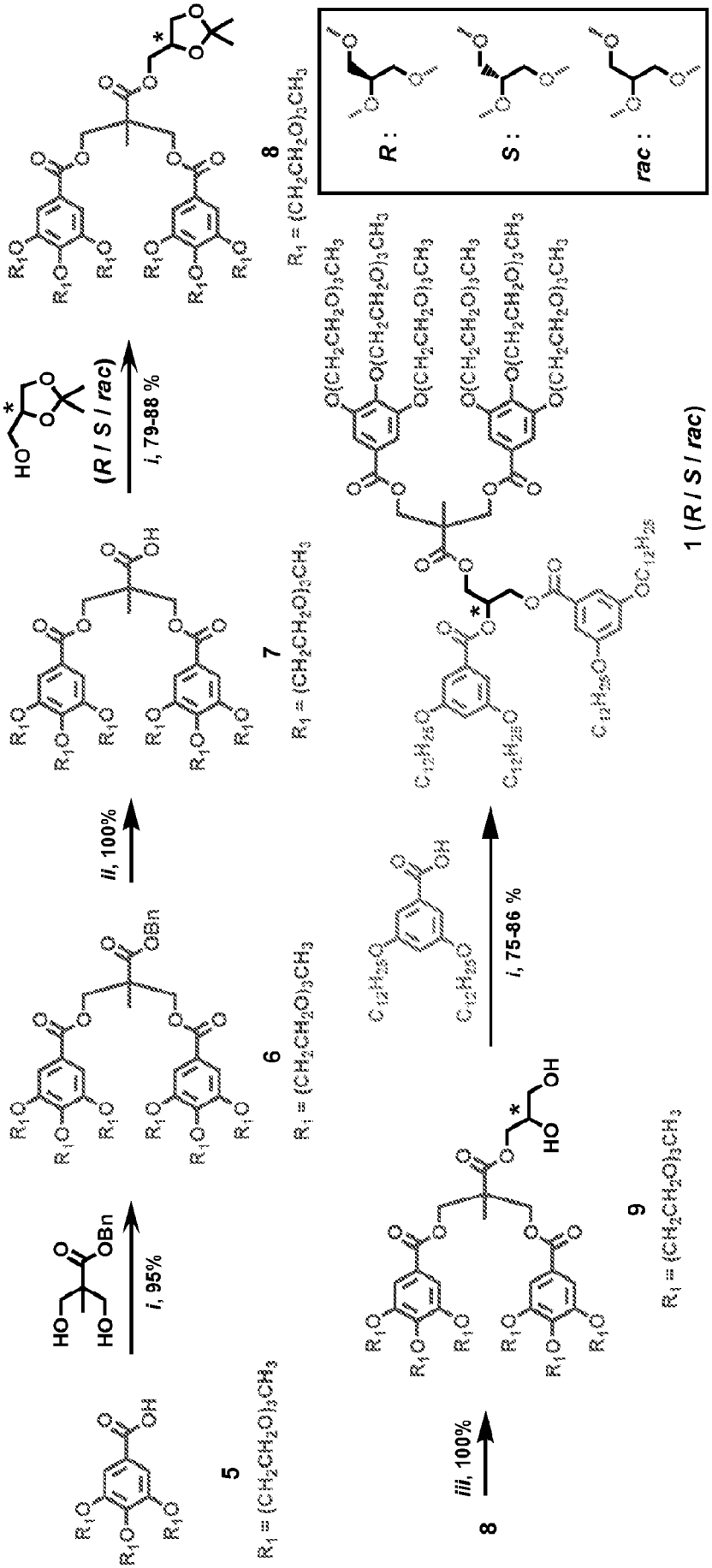

5
R1 = (CH2CH2O)3CH3
i, 95%
6
R1 = (CH2CH2O)3CH3
ii, 100%
7
R1 = (CH2CH2O)3CH3
(R / S / rac)
i, 79-88 %
8
R1 = (CH2CH2O)3CH3
R :
S :
rac :
8
iii, 100%
9
R1 = (CH2CH2O)3CH3
i, 75-86 %
1 (R / S / rac)

Figure 81

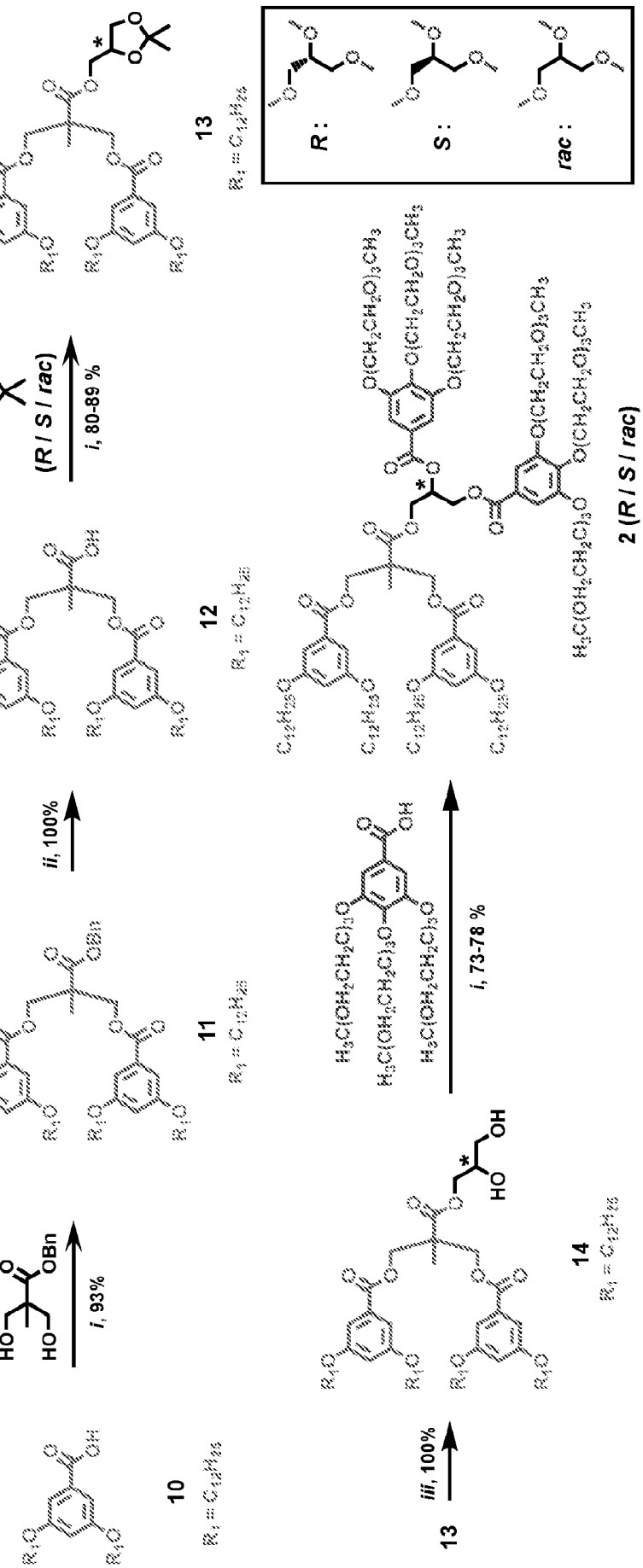

i, 93%
ii, 100%
iii, 100%
i, 80-89 %
i, 73-78 %
(R / S / rac)
10
11
12
13
14
2 (R / S / rac)
R:
S:
rac :

Figure 82

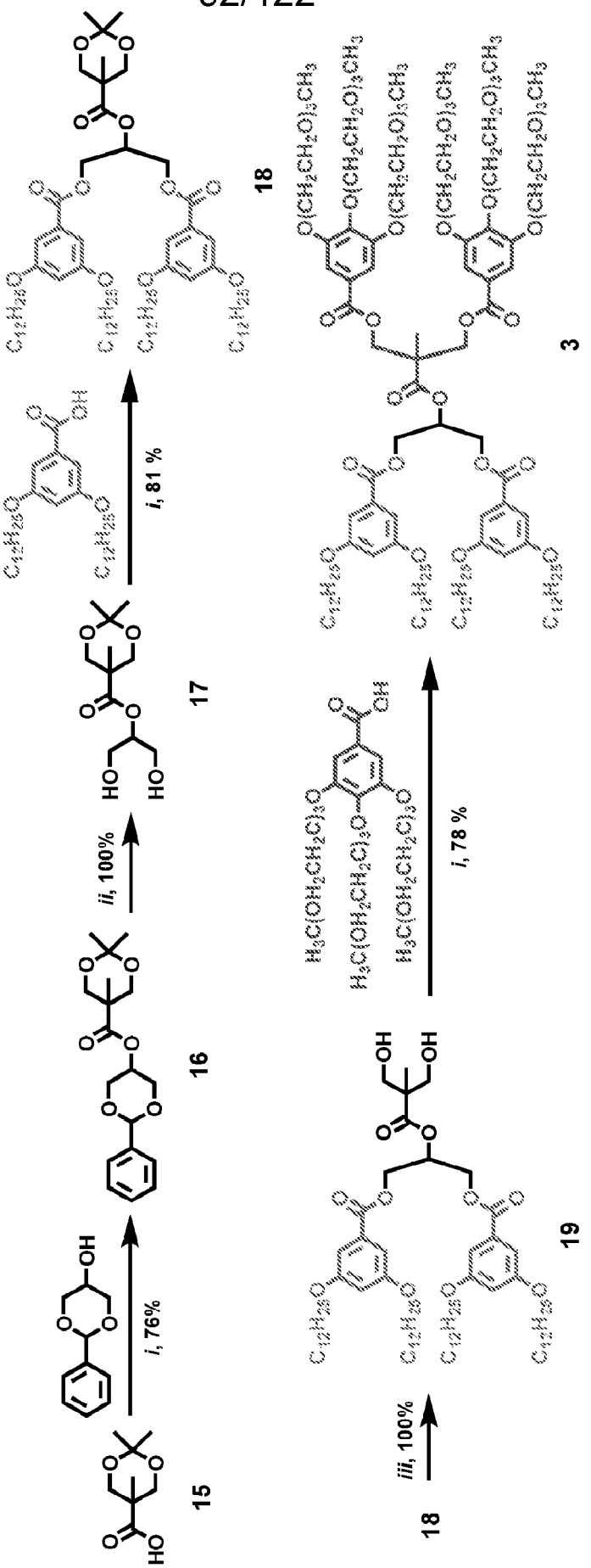


Figure 83

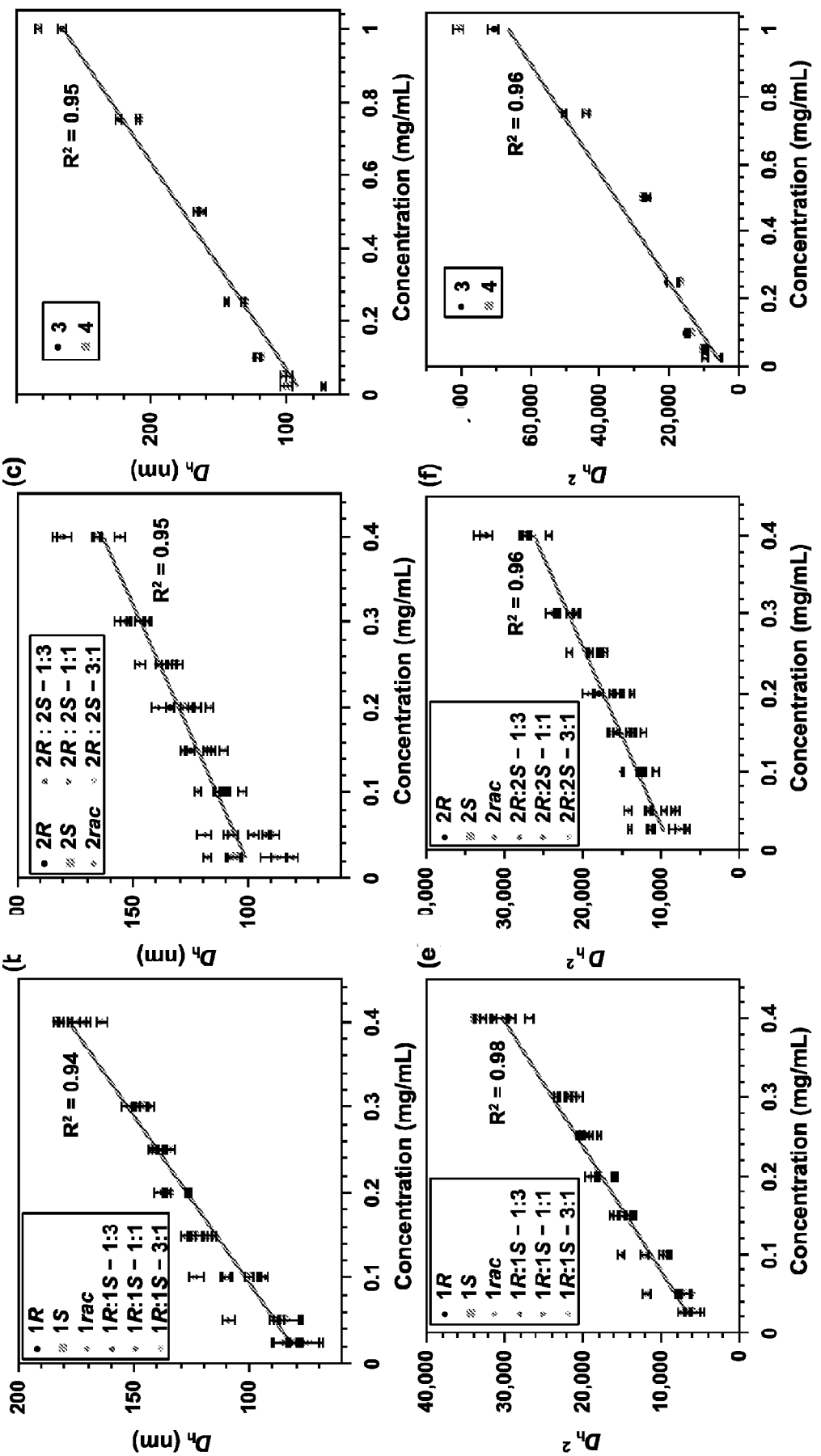

(a)
(b)
(c)
(d)
(e)
(f)
Dh (nm)
Dh ²
Concentration (mg/mL)
R² = 0.94
R² = 0.95
R² = 0.98
R² = 0.96
• 1R
1S
1rac
1R:1S – 1:3
1R:1S – 1:1
1R:1S – 3:1
• 2R
2S
2rac
2R : 2S – 1:3
2R : 2S – 1:1
2R : 2S – 3:1
• 3
4

Figure 84

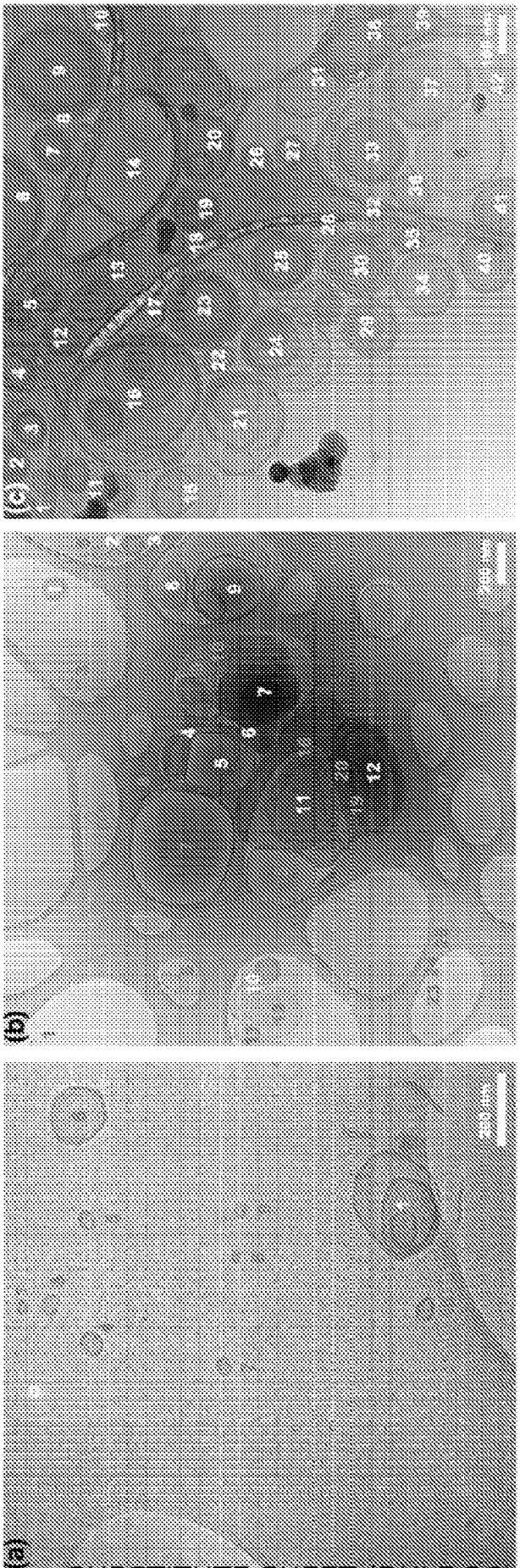


Figure 85

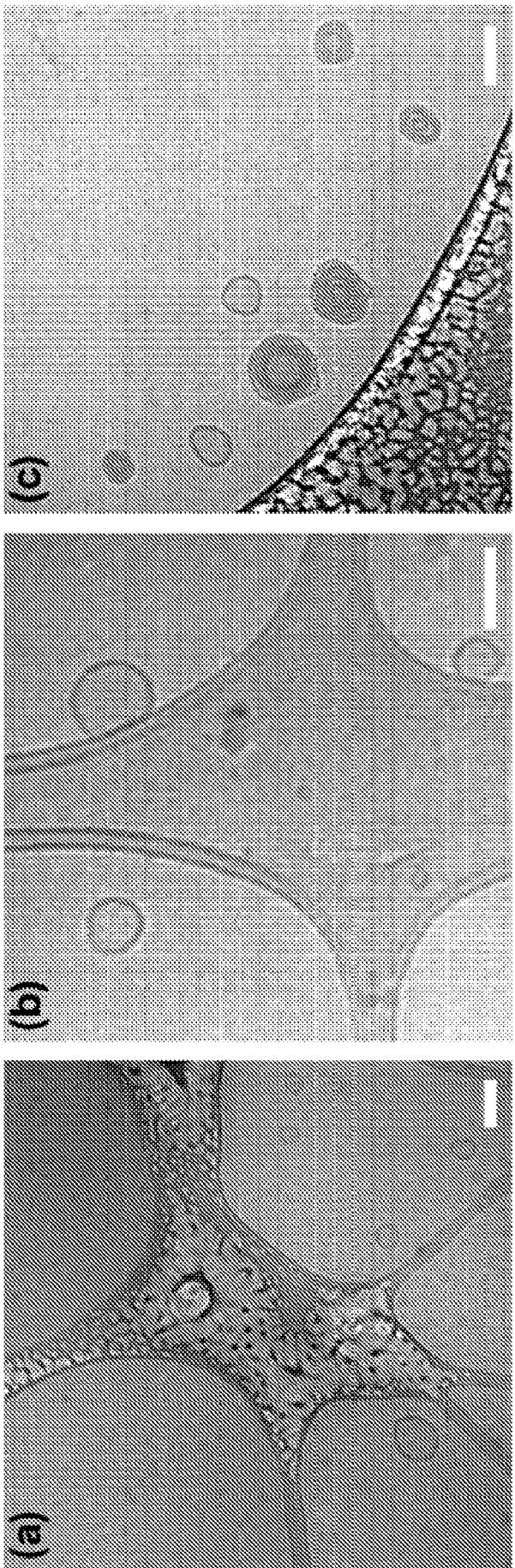


Figure 86

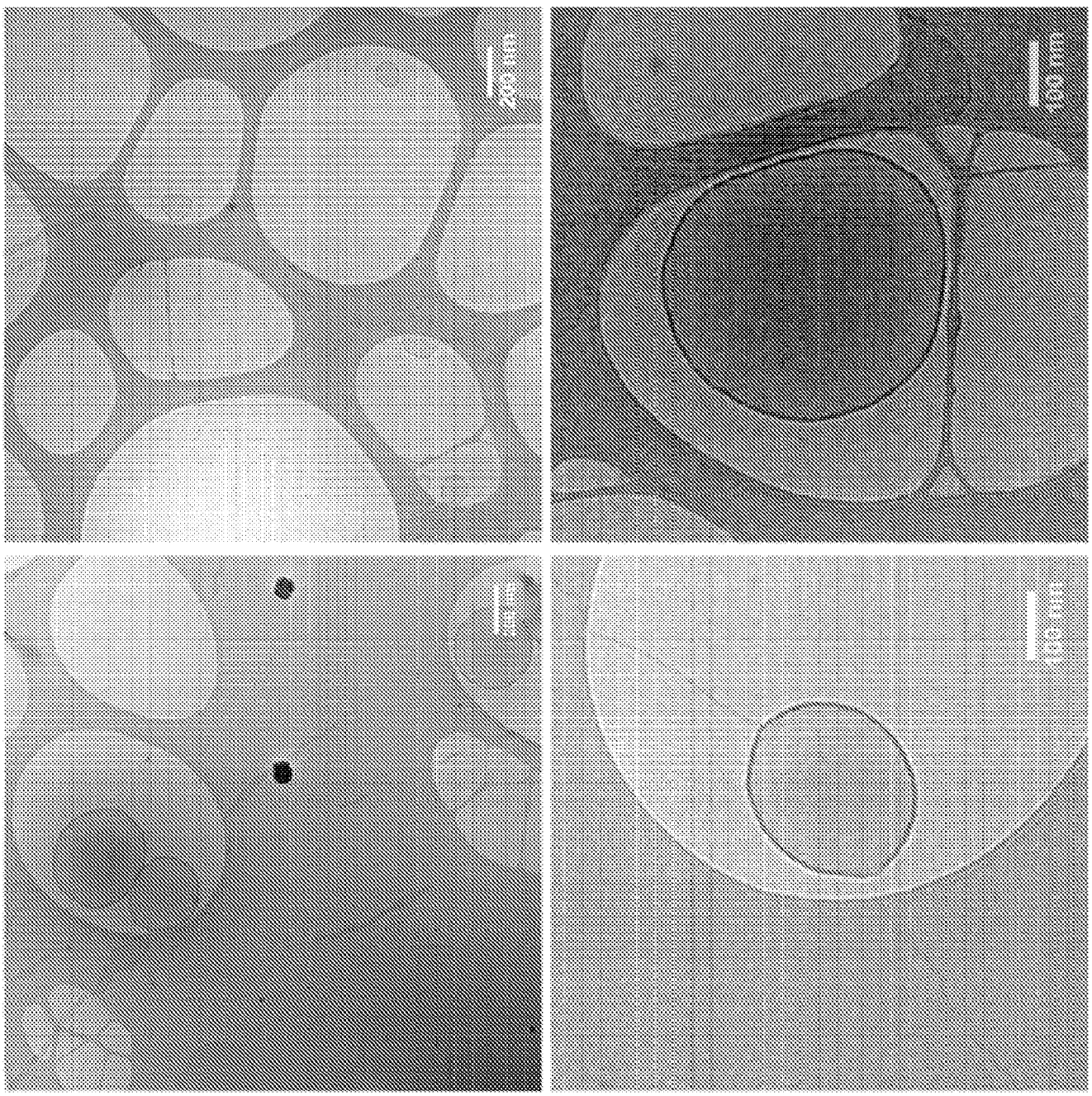
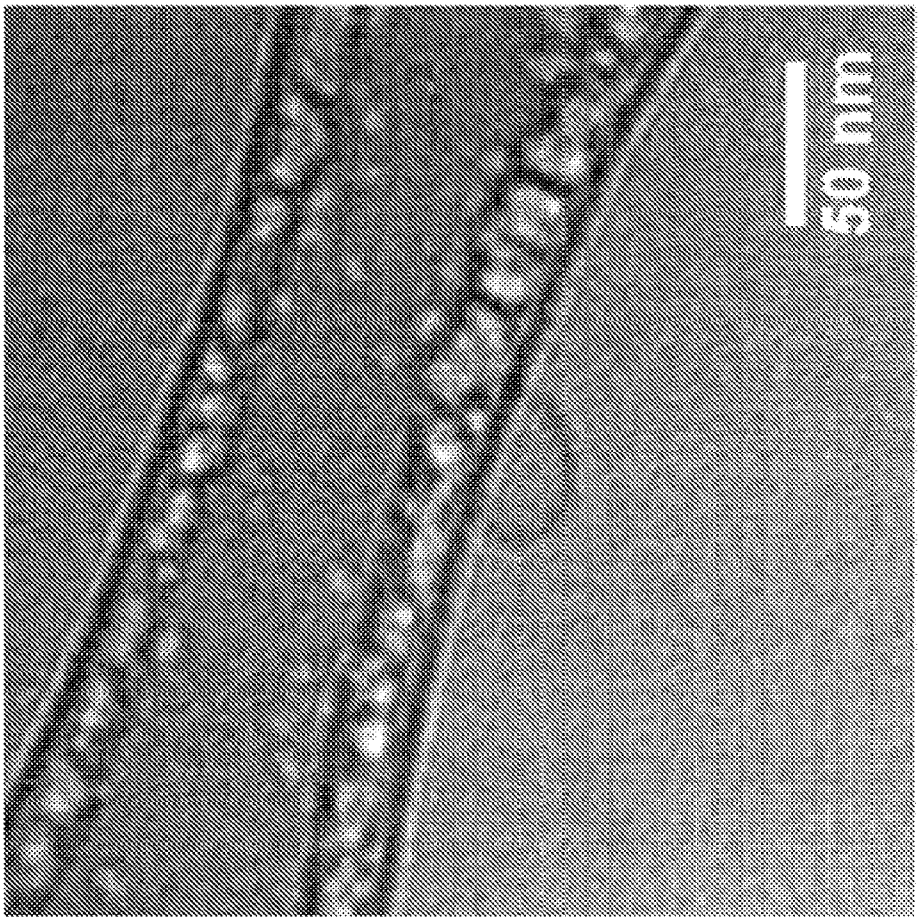


Figure 87



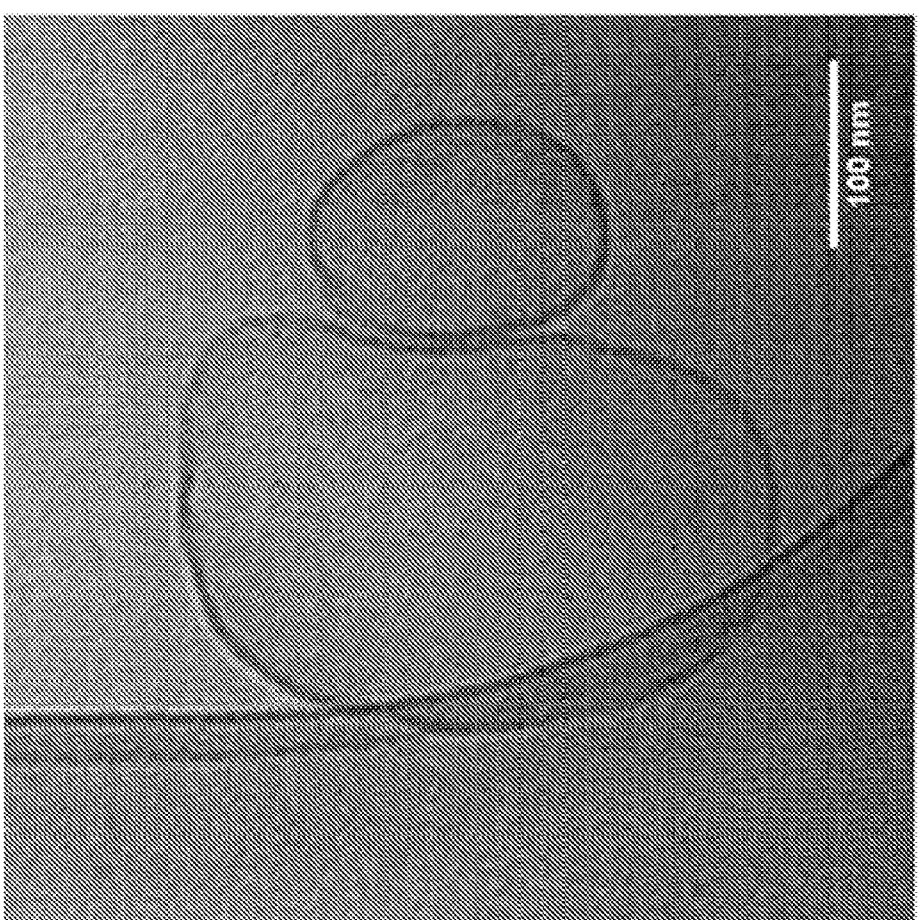


Figure 88

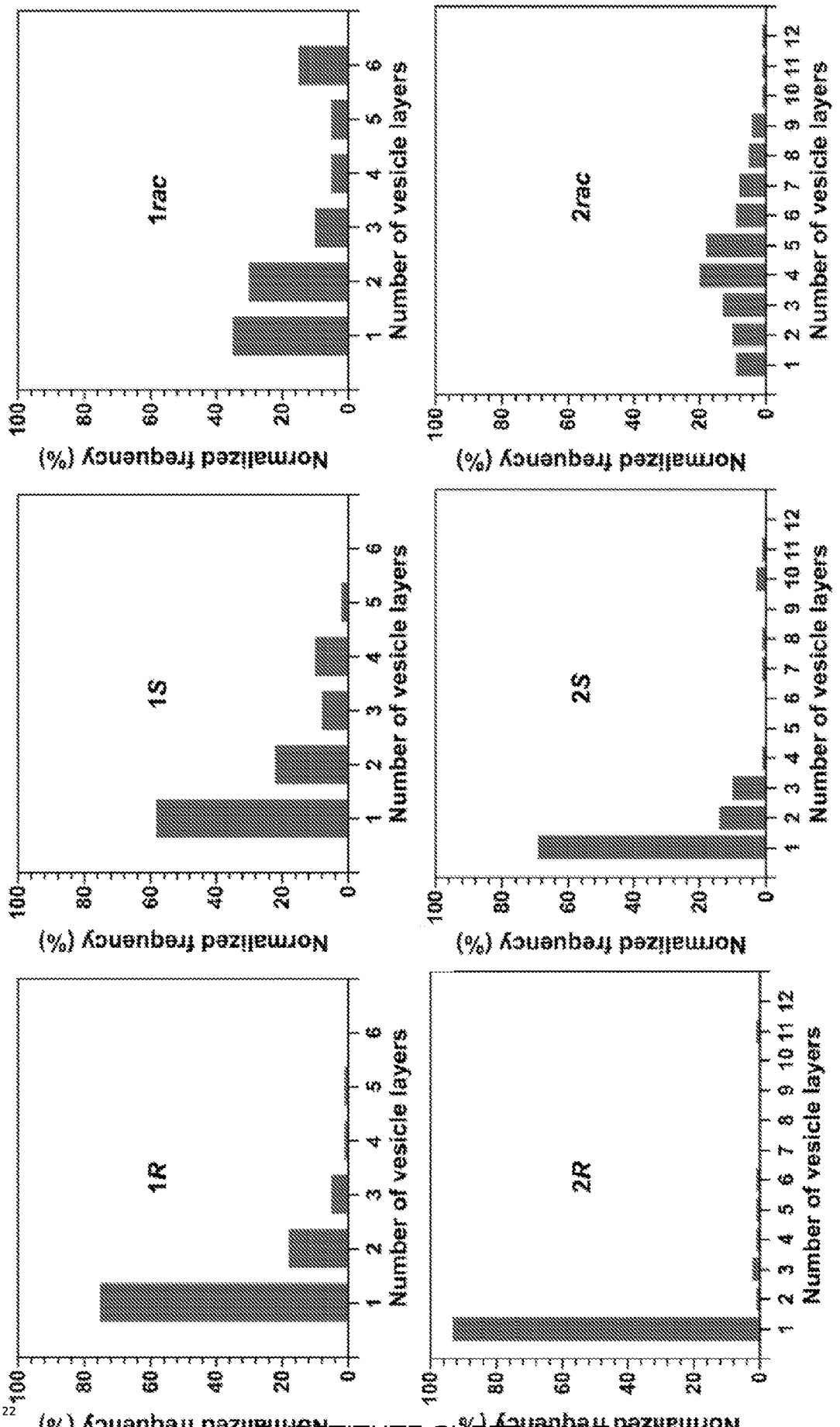

1rac
2rac
1S
2S
1R
2R
Normalized frequency (%)
Number of vesicle layers
0
20
40
60
80
100
1 2 3 4 5 6
1 2 3 4 5 6 7 8 9 10 11 12

Figure 89

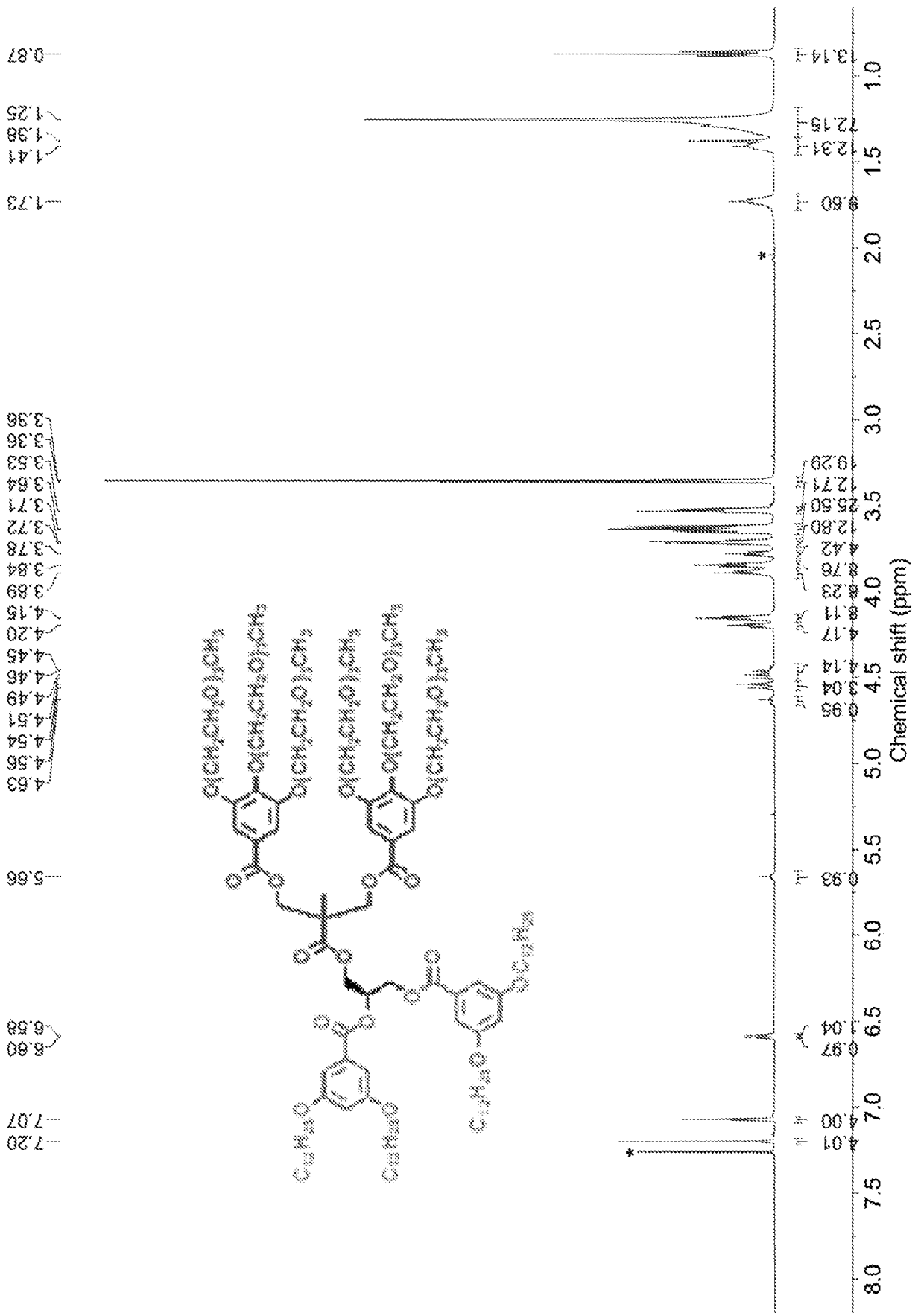


Figure 90

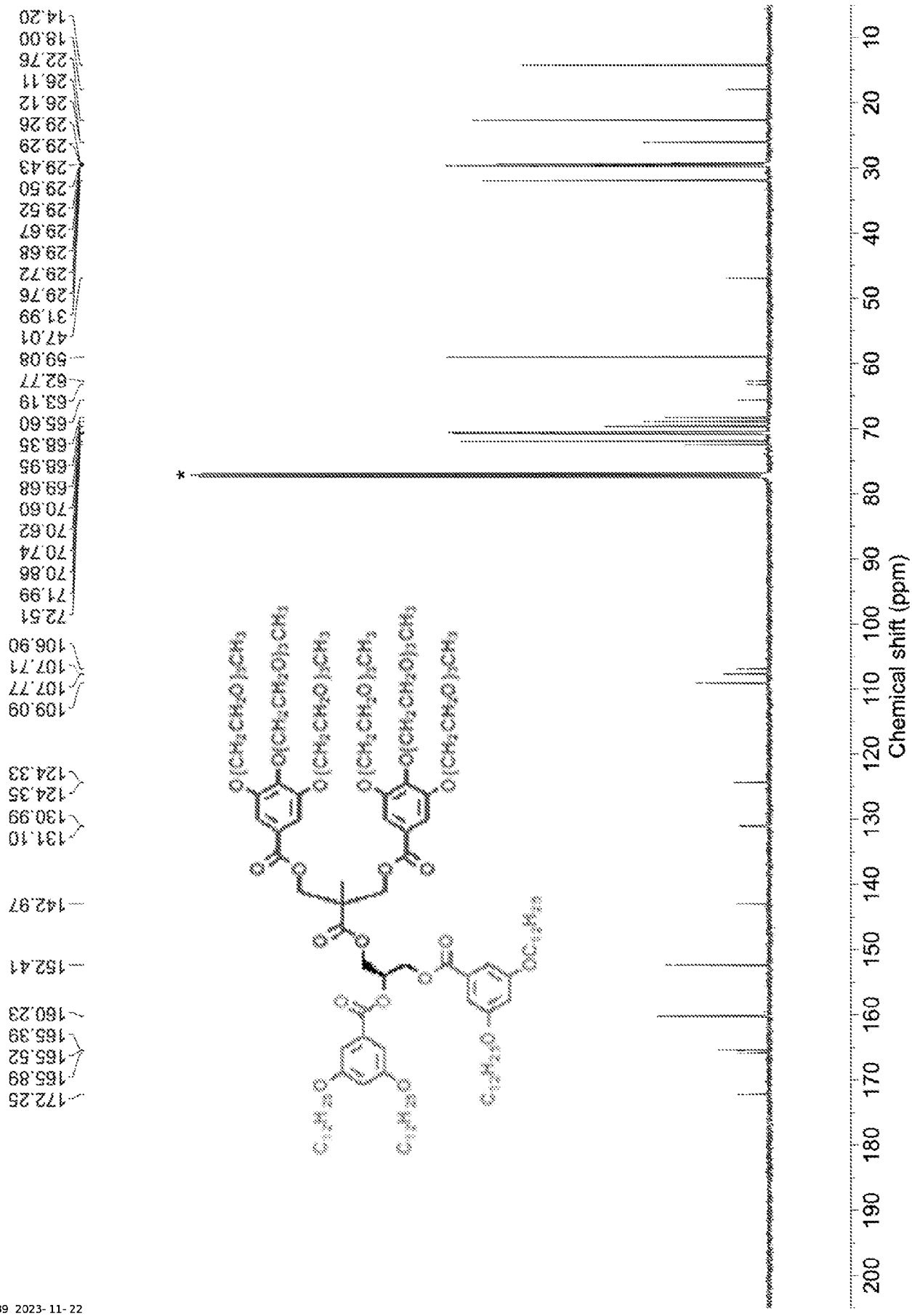

14.20
18.00
22.76
26.11
26.12
29.26
29.29
29.43
29.50
29.52
29.67
29.68
29.72
29.76
31.99
47.01
59.08
62.77
63.19
65.60
68.35
68.95
69.68
70.60
70.62
70.74
70.86
71.99
72.51
106.90
107.71
107.77
109.09
124.33
124.35
130.99
131.10
142.97
152.41
160.23
165.39
165.52
165.89
172.25
*
Chemical shift (ppm)

Figure 91

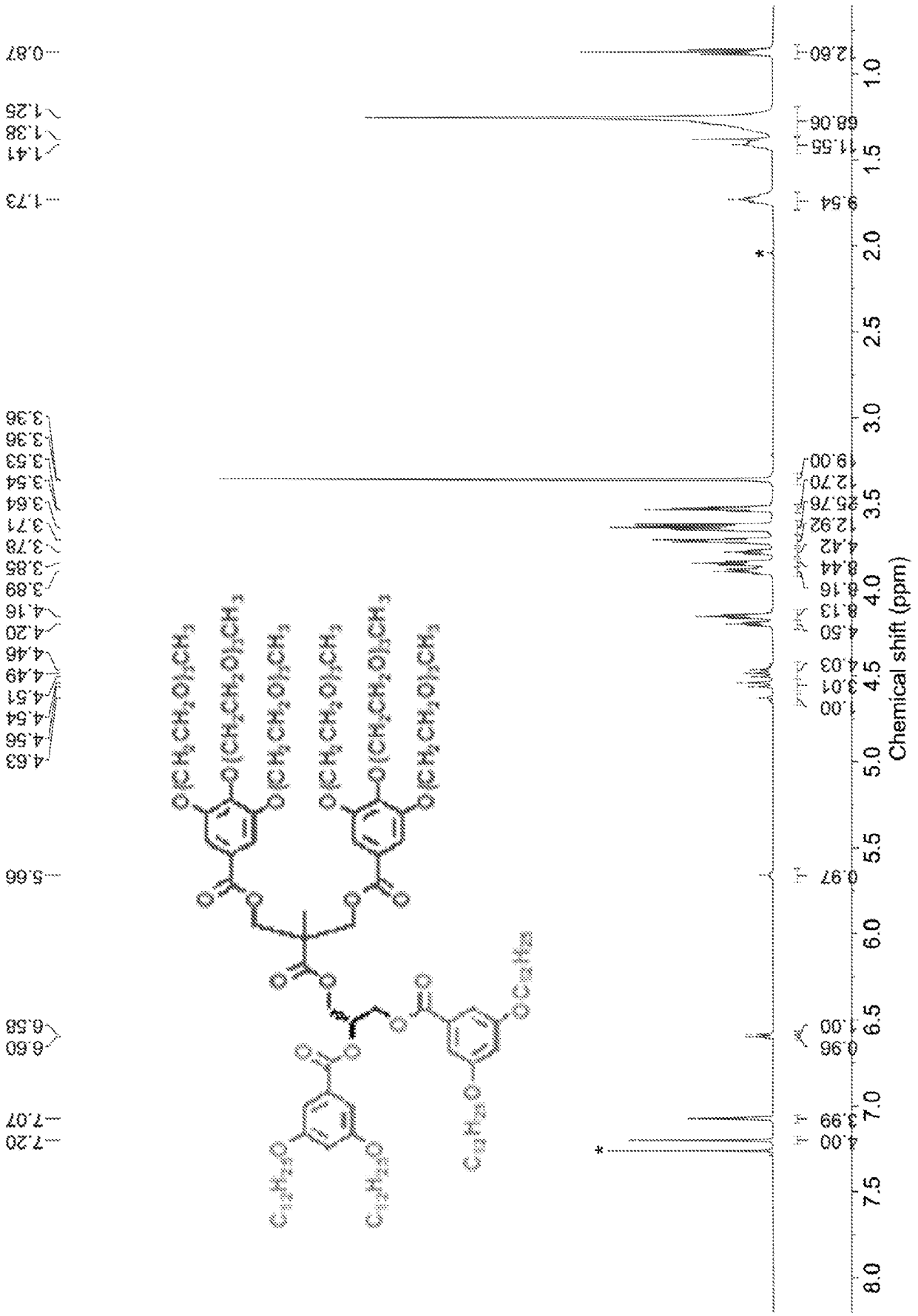

0.87
1.25
1.38
1.41
1.73
3.36
3.36
3.53
3.54
3.64
3.71
3.78
3.85
3.89
4.16
4.20
4.46
4.49
4.51
4.54
4.56
4.63
5.66
6.58
6.60
7.07
7.20
2.60
68.06
11.55
9.54
9.00
2.70
5.76
2.92
4.42
8.44
4.16
4.13
4.50
4.03
4.03
4.00
0.97
1.00
0.96
1.00
1.99
1.00
8.0
7.5
7.0
6.5
6.0
5.5
5.0
4.5
4.0
3.5
3.0
2.5
2.0
1.5
1.0
Chemical shift (ppm)

Figure 92

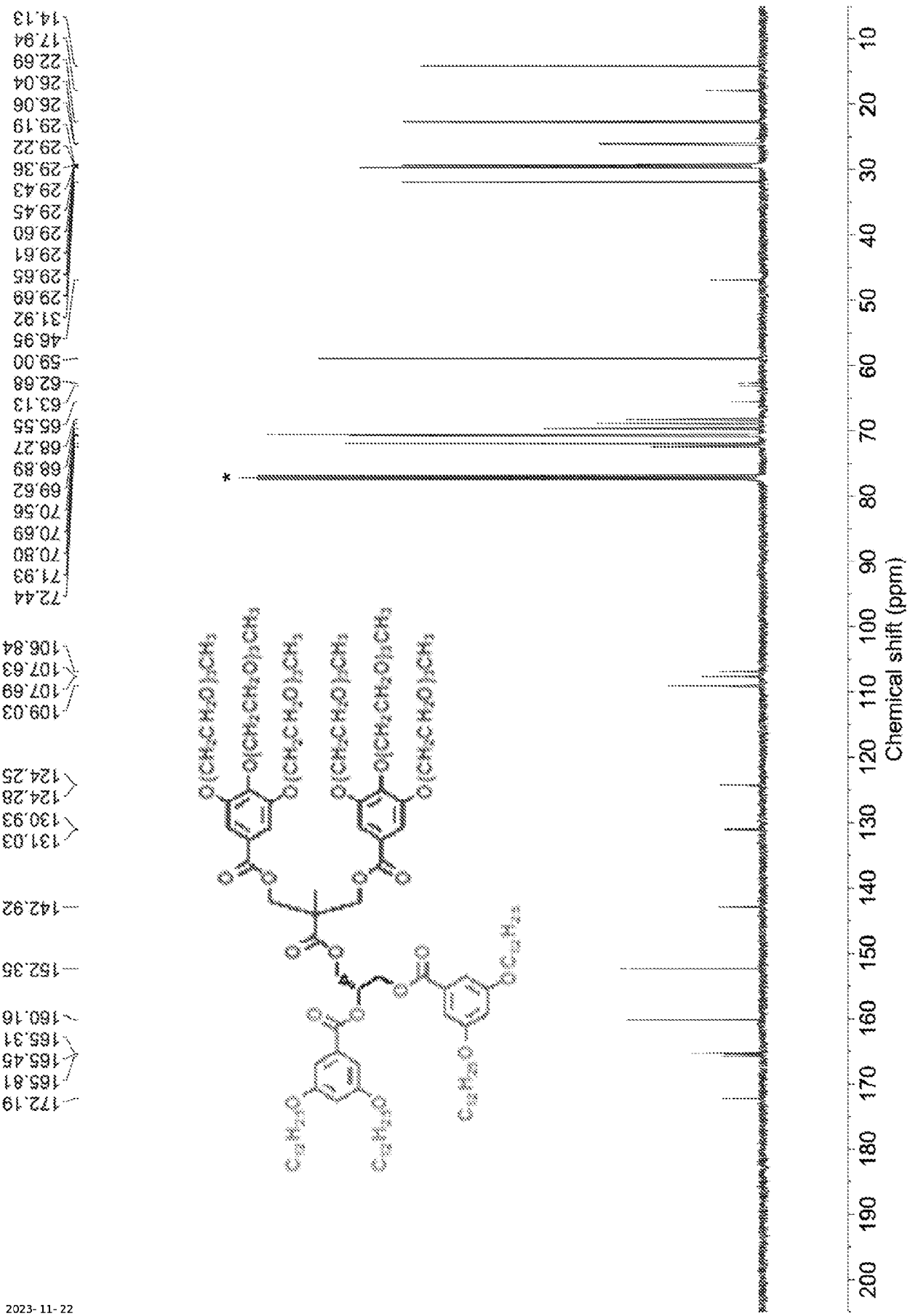

14.13
17.94
22.69
26.04
26.06
29.19
29.22
29.36
29.43
29.45
29.60
29.61
29.65
29.69
31.92
46.95
59.00
62.68
63.13
65.55
68.27
68.89
69.62
70.56
70.69
70.80
71.93
72.44
106.84
107.63
107.69
109.03
124.25
124.28
130.93
131.03
142.92
152.35
160.16
165.31
165.45
165.81
172.19
*
Chemical shift (ppm)

Figure 93

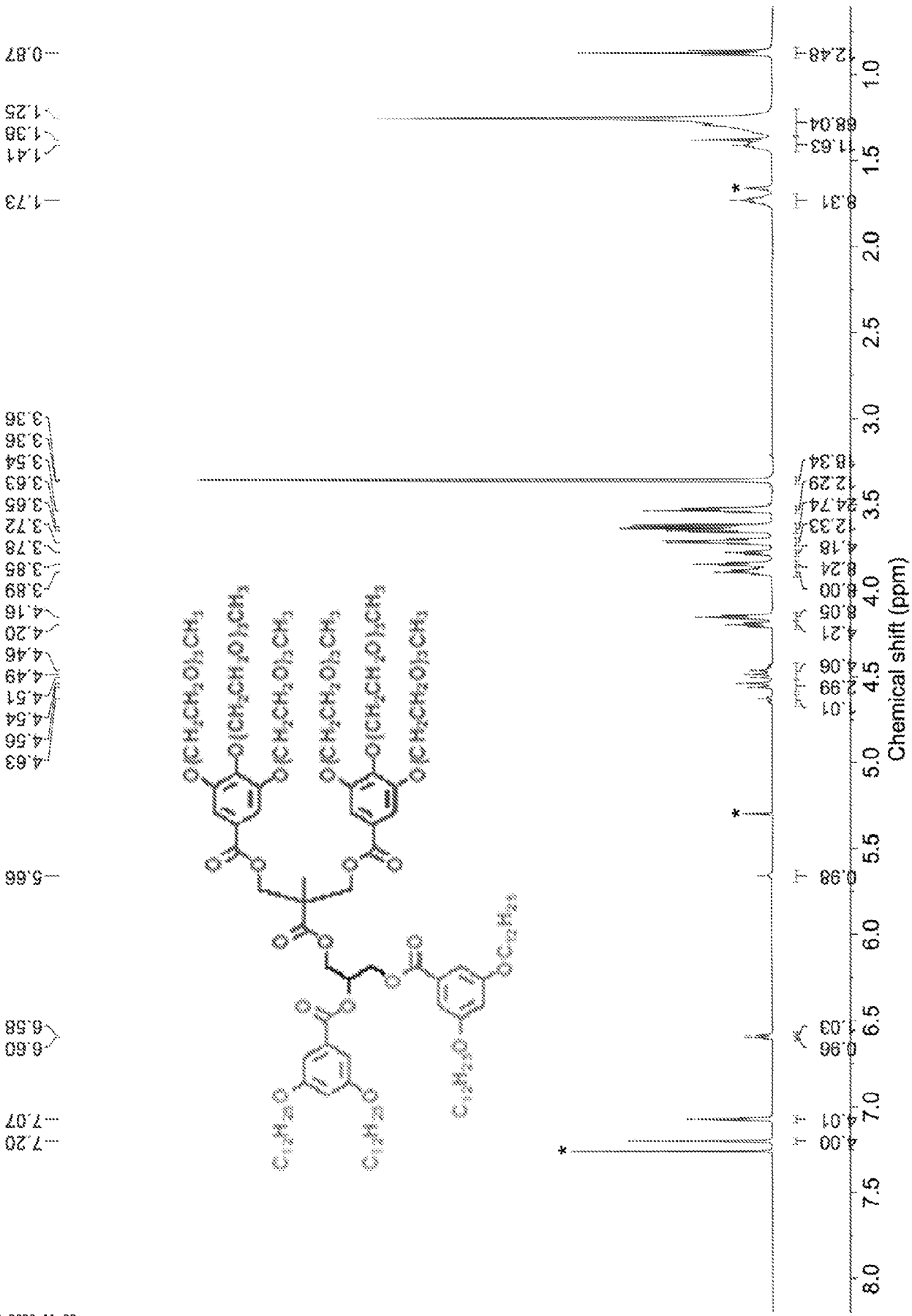

0.87
1.25
1.38
1.41
1.73
3.36
3.36
3.54
3.63
3.65
3.72
3.78
3.85
3.89
4.16
4.20
4.46
4.49
4.51
4.54
4.56
4.63
5.66
6.58
6.60
7.07
7.20
2.48
68.04
11.63
8.31
18.34
2.29
24.74
2.33
4.18
8.24
8.00
8.05
4.21
4.06
2.99
1.01
8.98
1.03
0.96
4.01
4.00
1.0
1.5
2.0
2.5
3.0
3.5
4.0
4.5
5.0
5.5
6.0
6.5
7.0
7.5
8.0
Chemical shift (ppm)

Figure 94

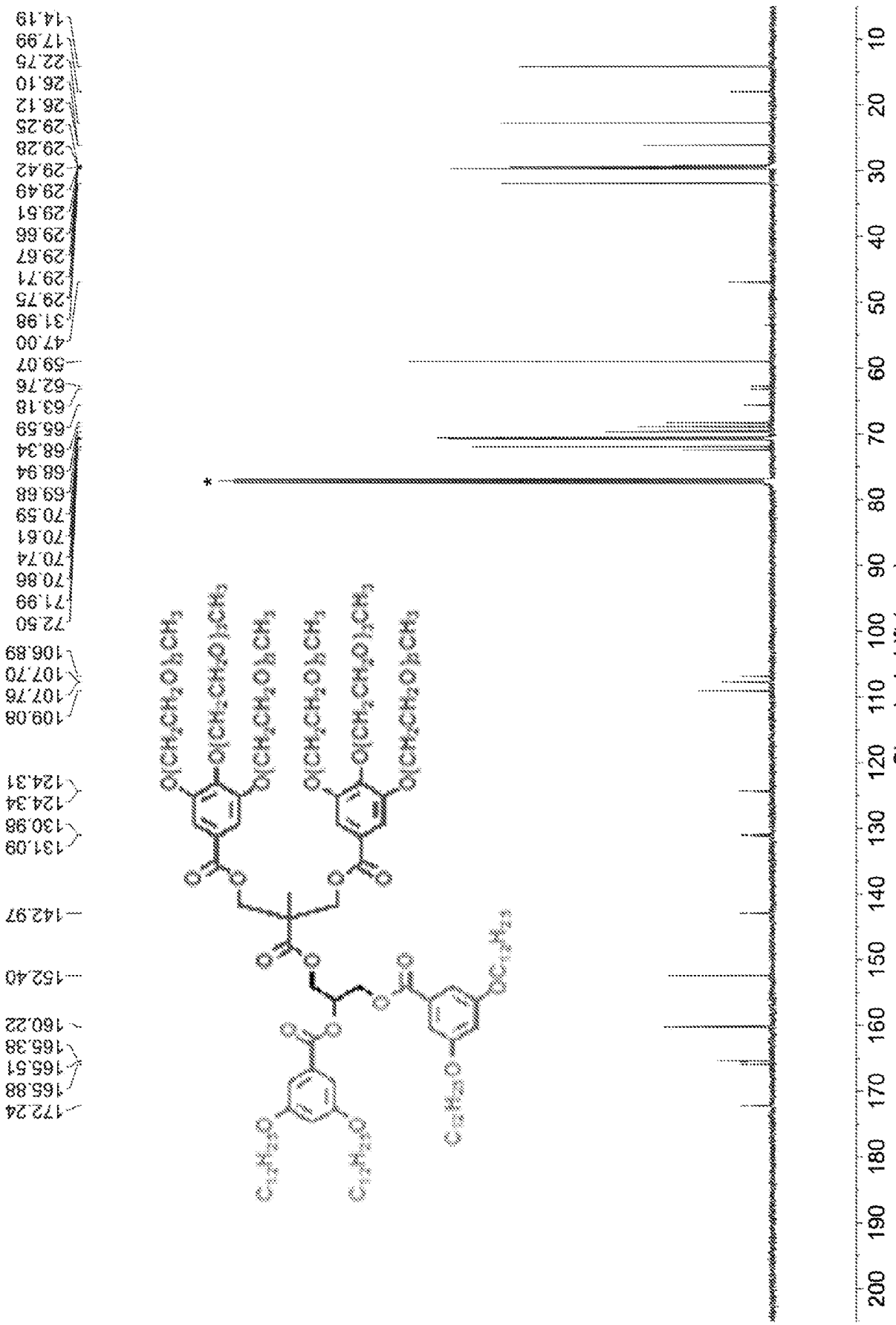

14.19
17.99
22.75
26.10
26.12
29.25
29.28
29.42
29.49
29.61
29.66
29.67
29.71
29.75
31.98
47.00
59.07
62.76
63.18
65.59
68.34
68.94
69.68
70.59
70.61
70.74
70.86
71.99
72.50
106.89
107.70
107.76
109.08
124.31
124.34
130.98
131.09
142.97
152.40
160.22
165.38
165.51
165.88
172.24
Chemical shift (ppm)

Figure 95

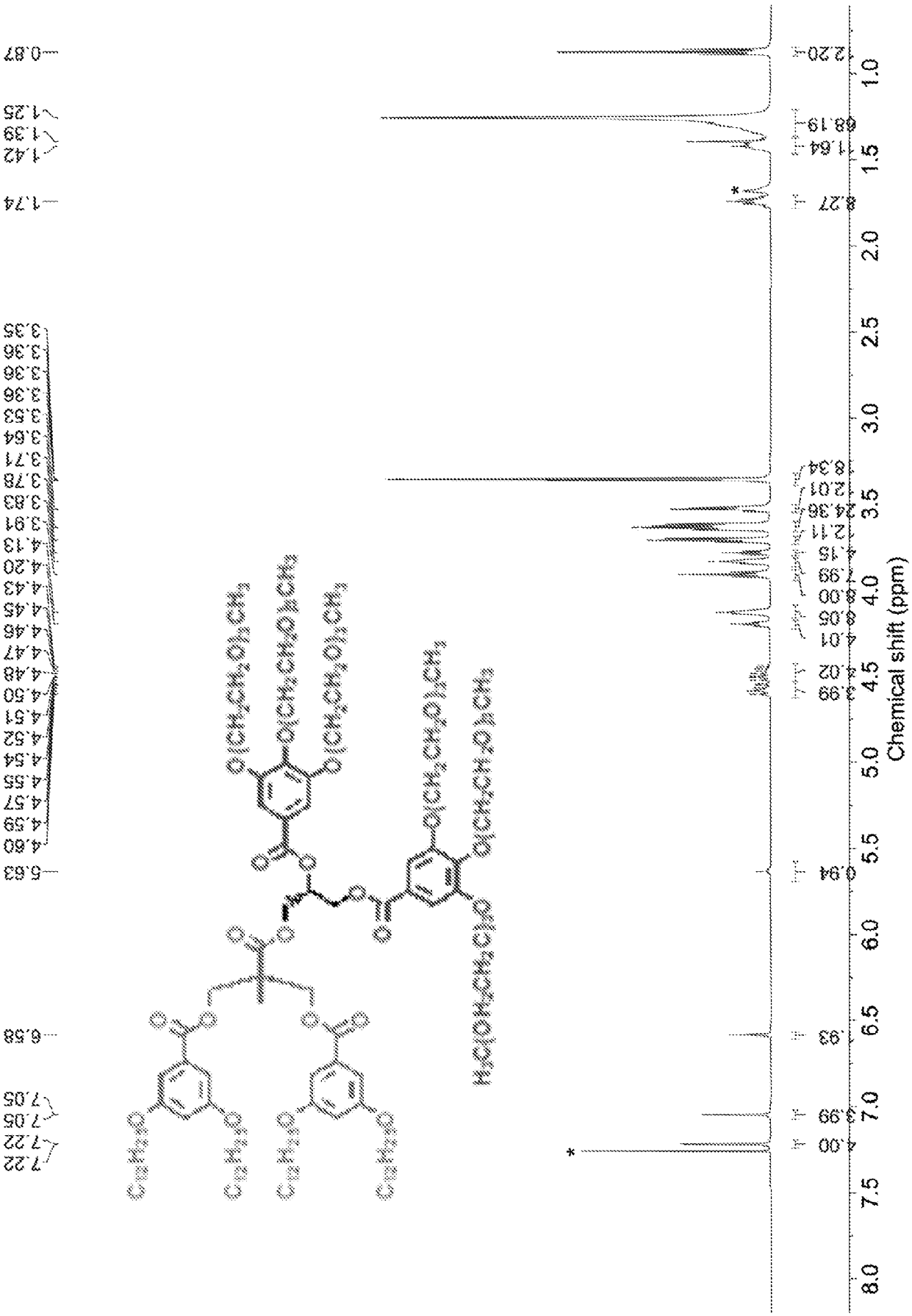


Figure 96

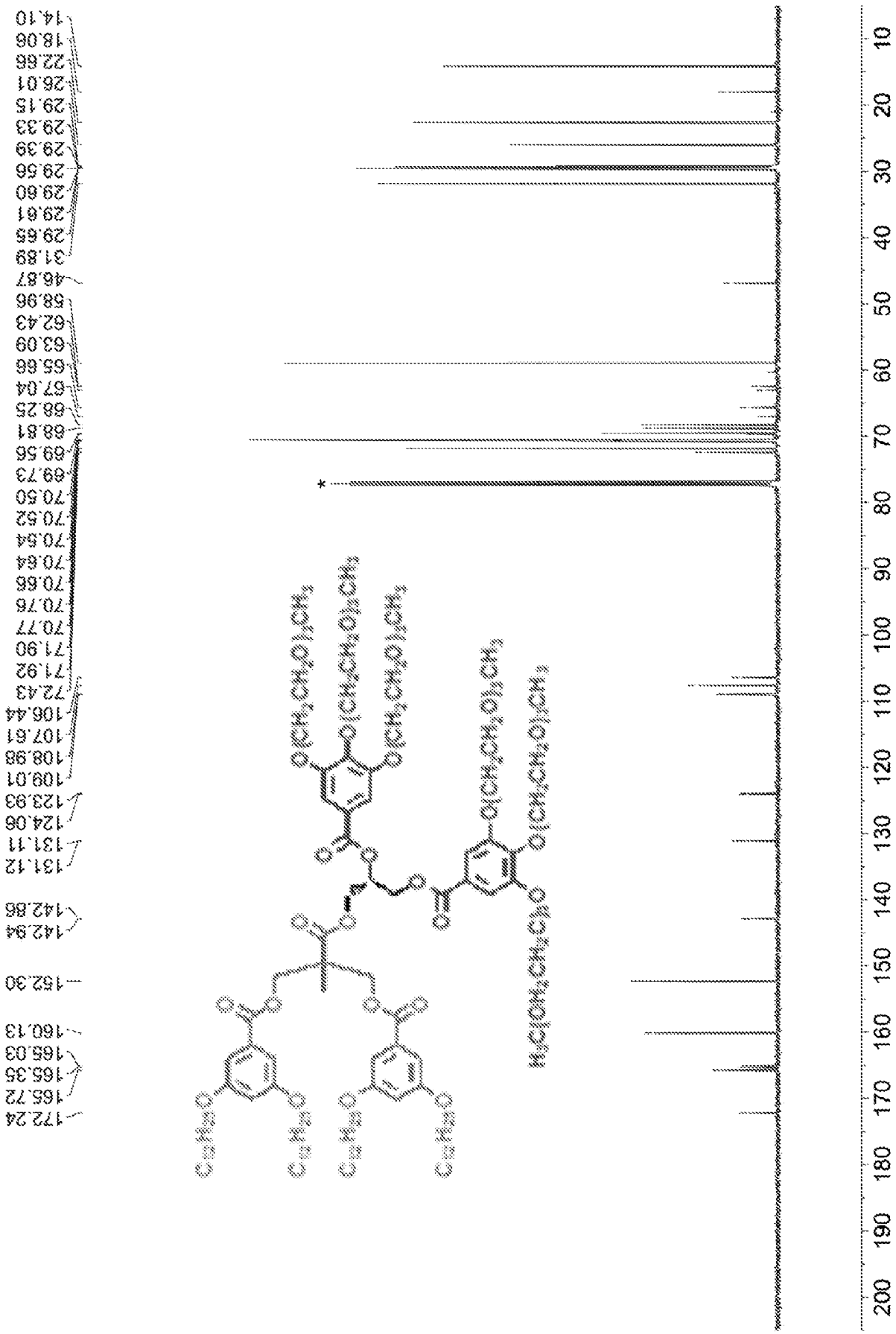

14.10
18.06
22.66
26.01
29.15
29.33
29.39
29.56
29.60
29.61
29.65
31.89
46.87
58.96
62.43
63.09
65.66
67.04
68.25
68.81
69.56
69.73
70.50
70.52
70.54
70.64
70.66
70.76
70.77
71.90
71.92
72.43
106.44
107.61
108.98
109.01
123.93
124.06
131.11
131.12
142.86
142.94
152.30
160.13
165.03
165.35
165.72
172.24
*
Chemical shift (ppm)
10
20
30
40
50
60
70
80
90
100
110
120
130
140
150
160
170
180
190
200

Figure 97

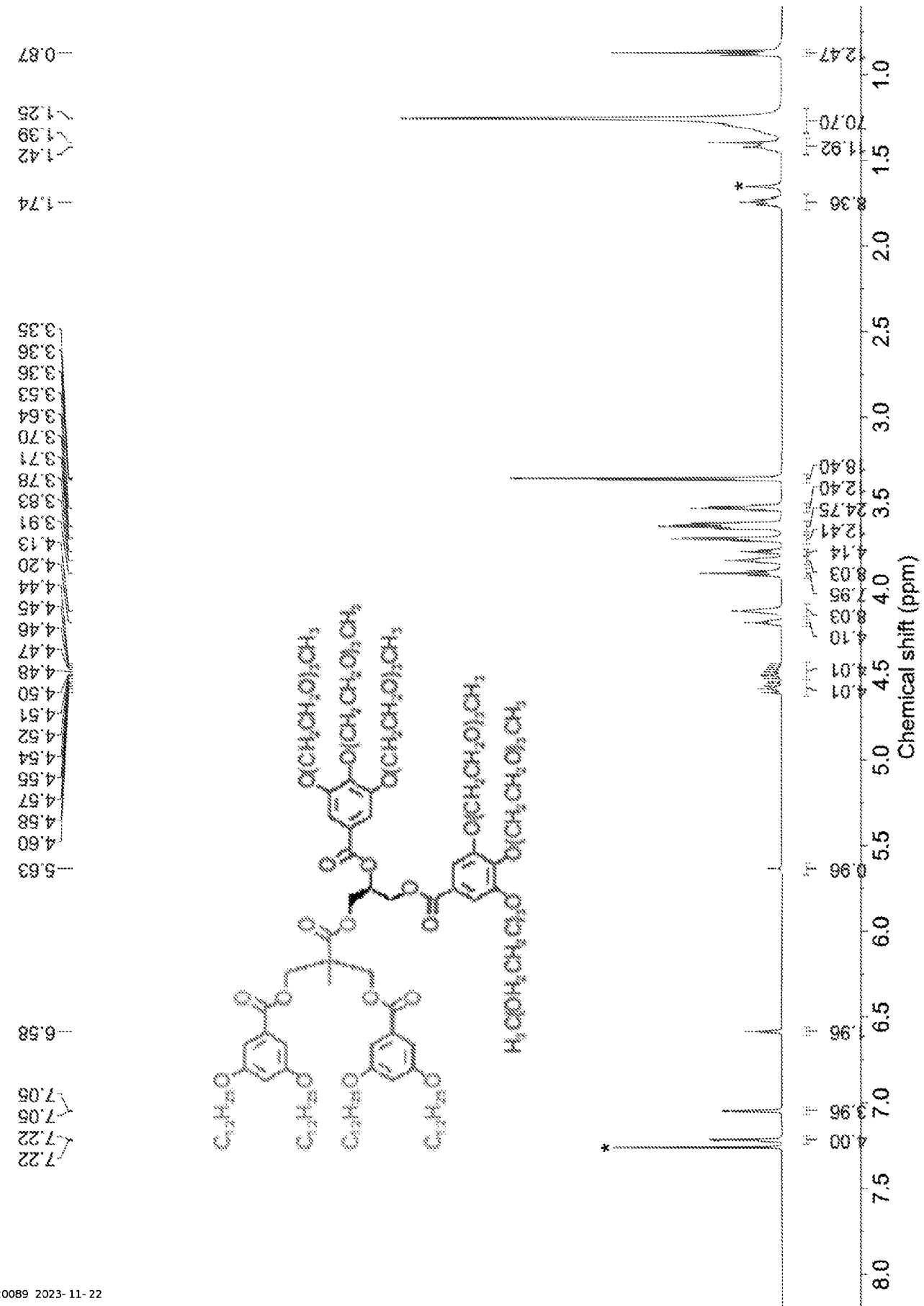


Figure 98

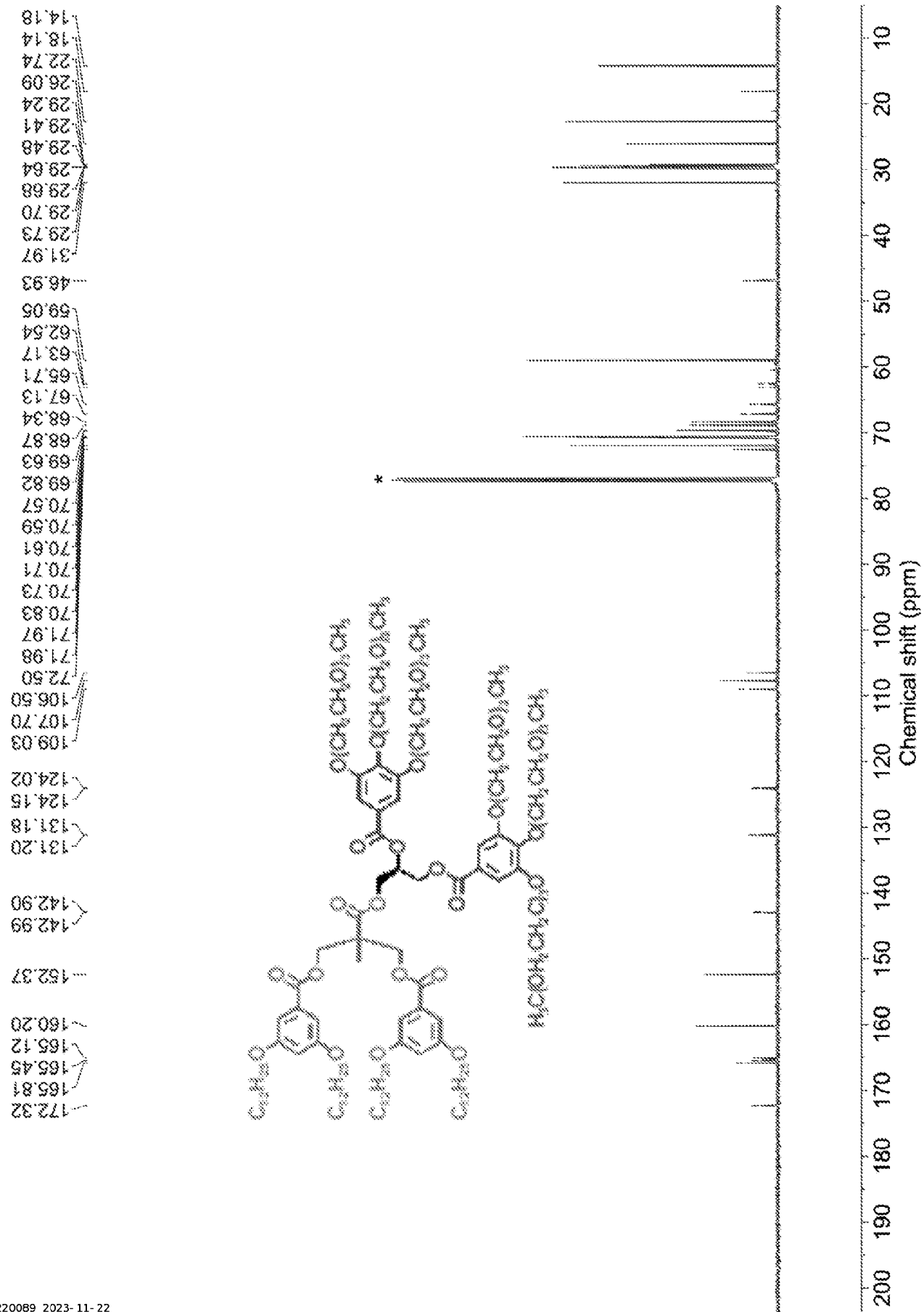

14.18
18.14
22.74
26.09
29.24
29.41
29.48
29.64
29.68
29.70
29.73
31.97
46.93
59.05
62.54
63.17
65.71
67.13
68.34
68.87
69.63
69.82
70.57
70.59
70.61
70.71
70.73
70.83
71.97
71.98
72.50
106.50
107.70
109.03
124.02
124.15
131.18
131.20
142.90
142.99
152.37
160.20
165.12
165.45
165.81
172.32
*
200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10
Chemical shift (ppm)

Figure 99

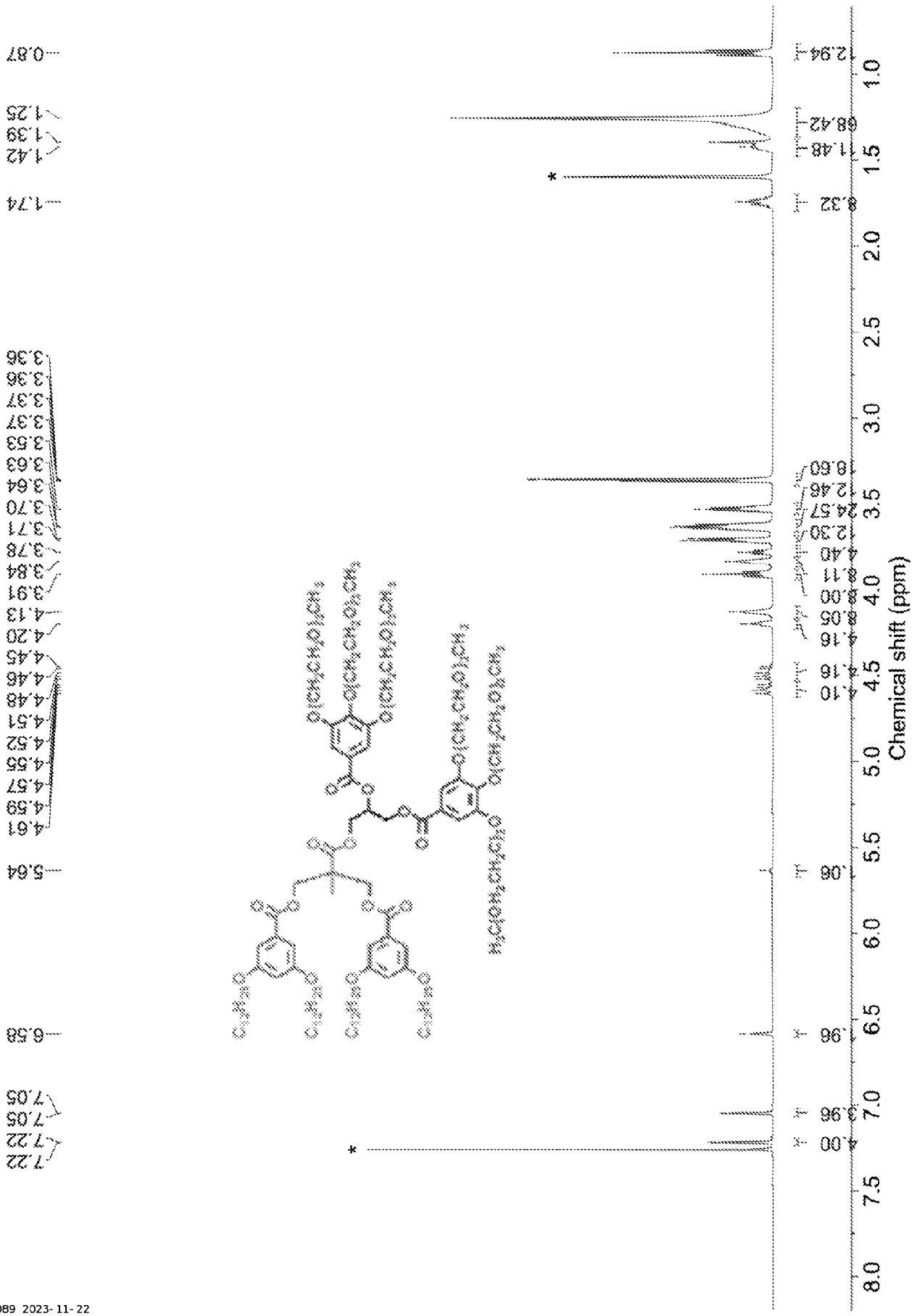


Figure 100

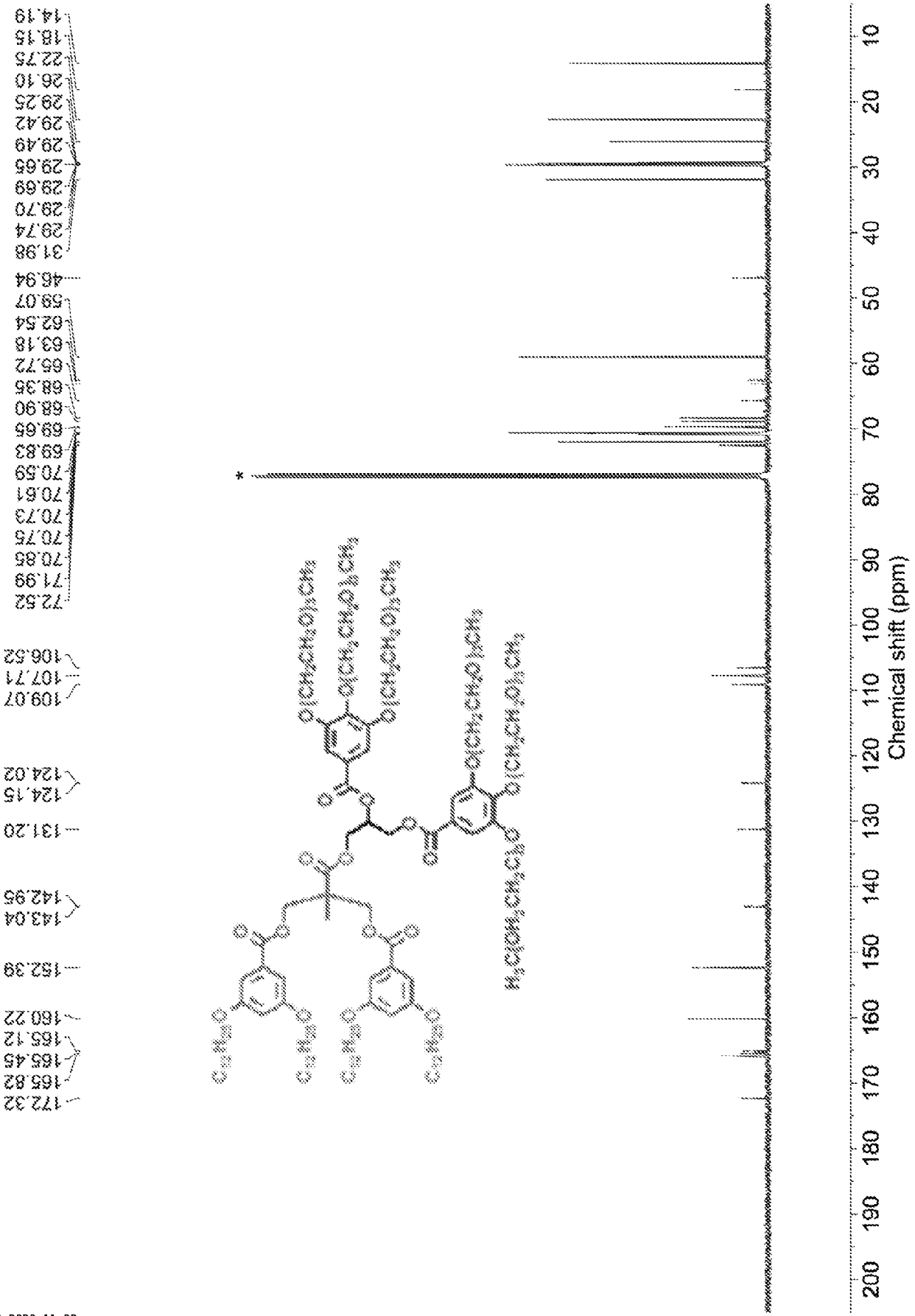

14.19
18.15
22.75
26.10
29.25
29.42
29.49
29.65
29.69
29.70
29.74
31.98
46.94
59.07
62.54
63.18
66.72
68.35
68.90
69.65
69.83
70.59
70.61
70.73
70.75
70.85
71.99
72.52
106.52
107.71
109.07
124.02
124.15
131.20
142.95
143.04
152.39
160.22
165.12
165.45
165.82
172.32
*
200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10
Chemical shift (ppm)

Figure 101

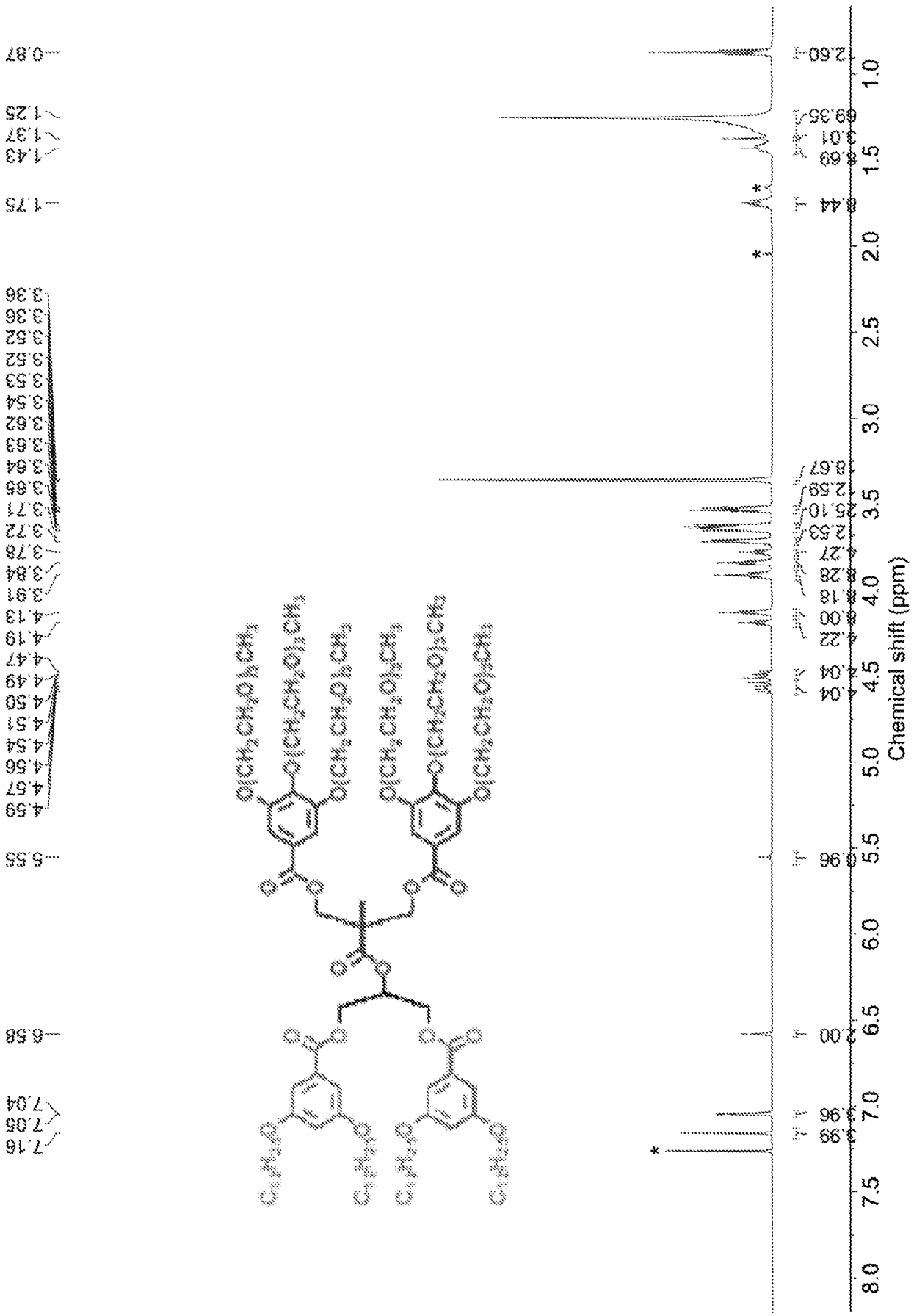


Figure 102

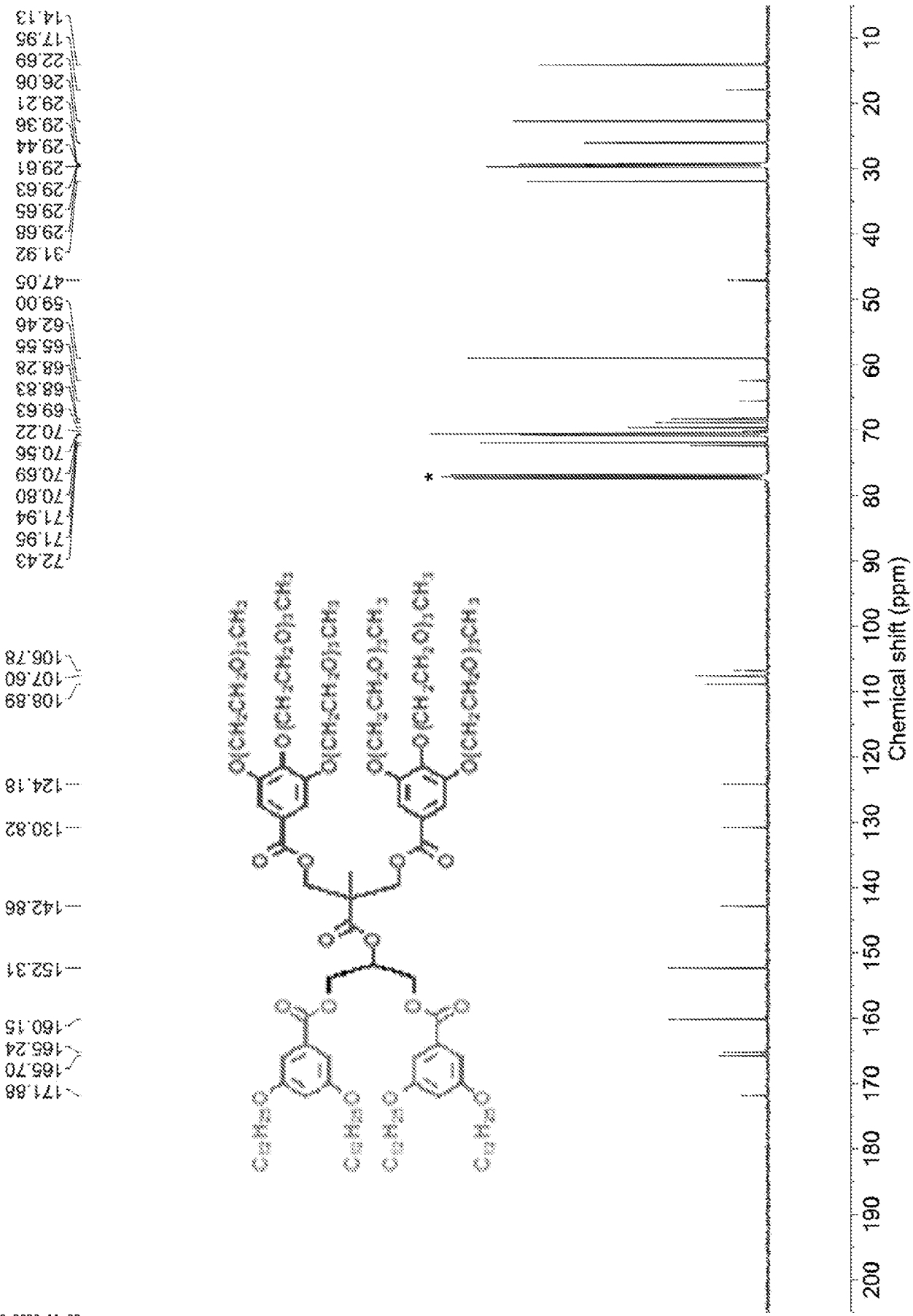

14.13
17.95
22.69
26.06
29.21
29.36
29.44
29.61
29.63
29.65
29.68
31.92
47.05
59.00
62.46
65.55
68.28
68.83
69.63
70.22
70.56
70.69
70.80
71.94
71.95
72.43
106.78
107.60
108.89
124.18
130.82
142.86
152.31
160.15
165.24
165.70
171.88
*
Chemical shift (ppm)

Figure 103

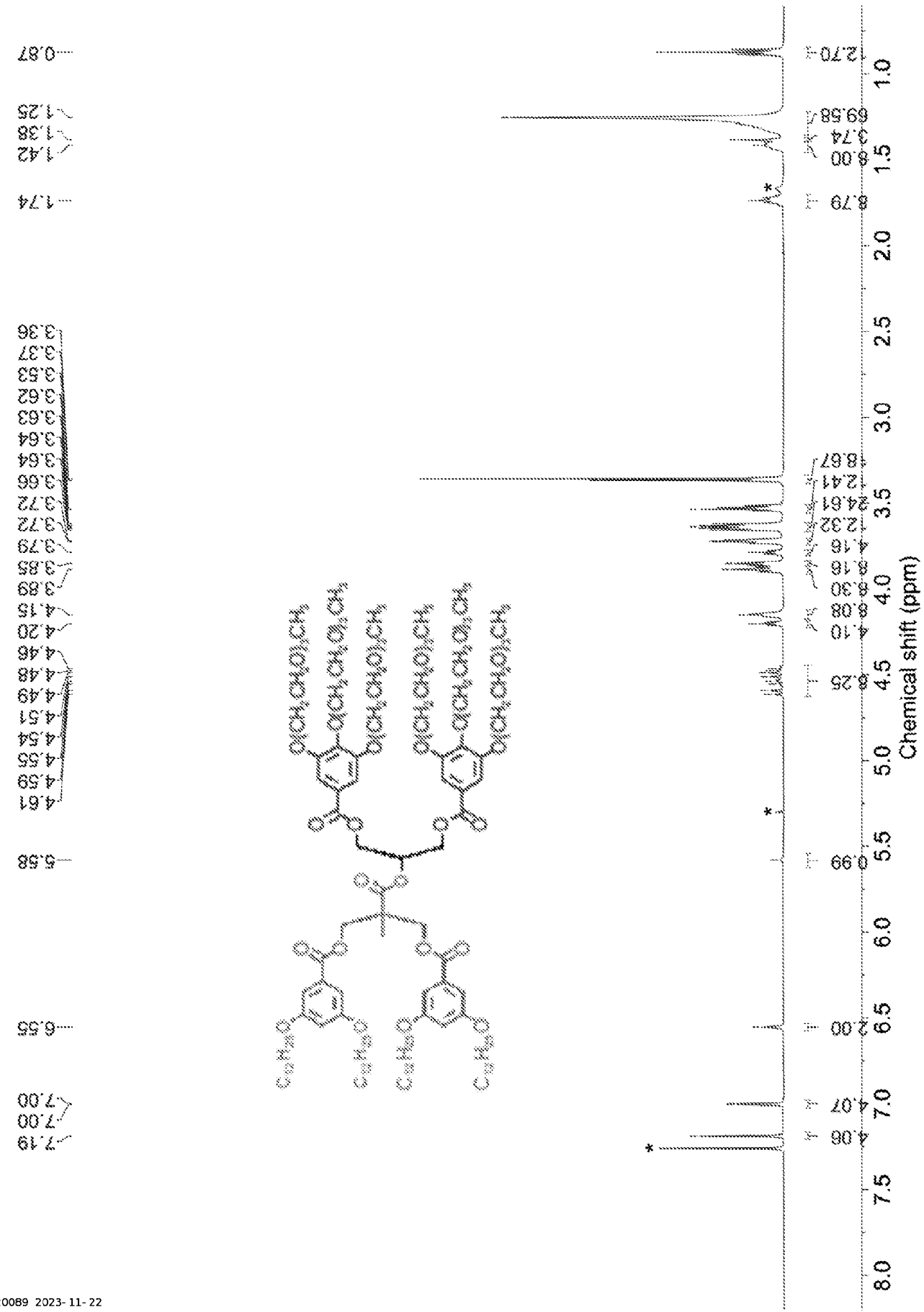


Figure 104

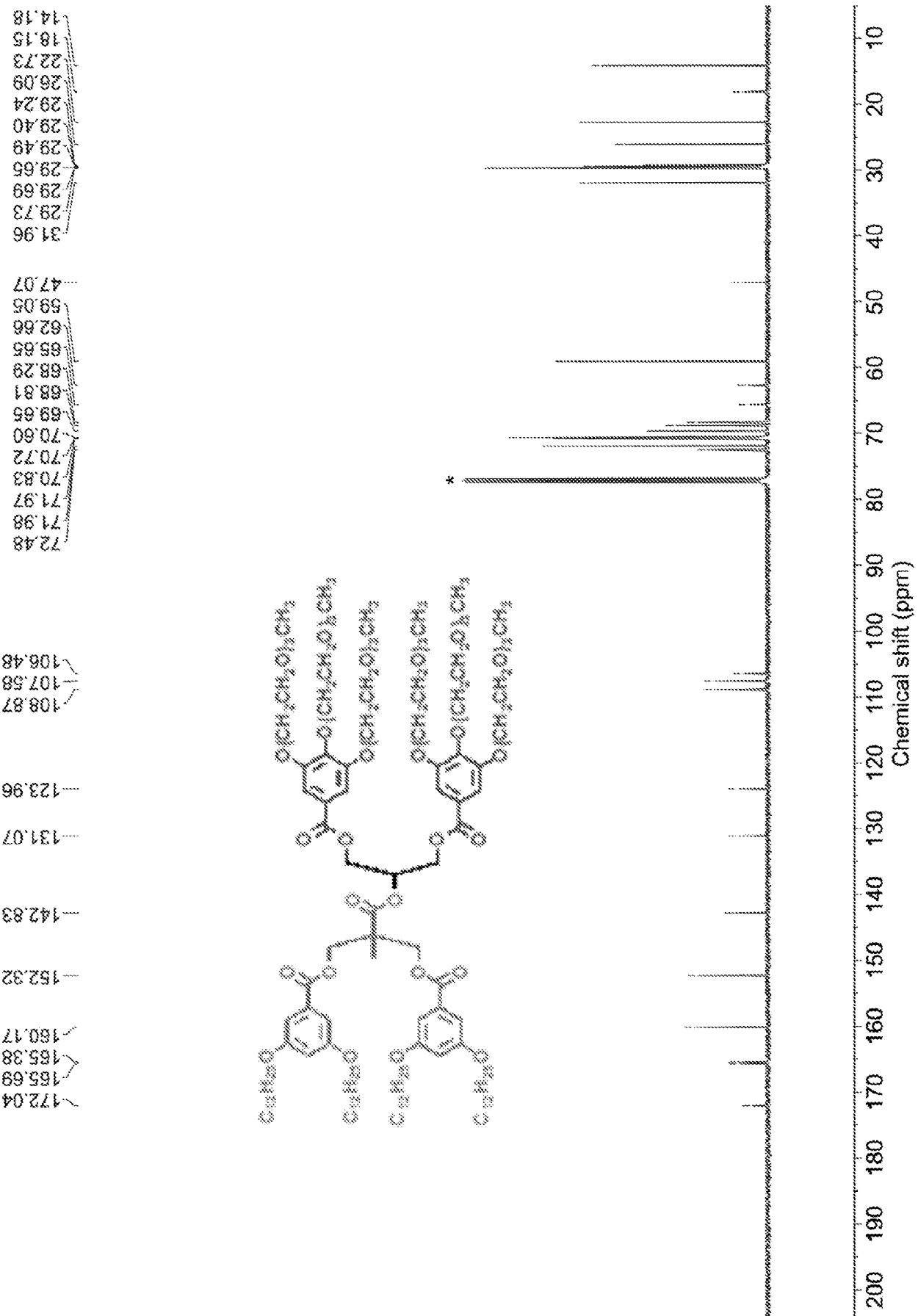

14.18
18.15
22.73
26.09
29.24
29.40
29.49
29.65
29.69
29.73
31.96
47.07
59.05
62.66
65.65
68.29
68.81
69.65
70.60
70.72
70.83
71.97
71.98
72.48
106.48
107.58
108.87
123.96
131.07
142.83
152.32
160.17
165.38
165.69
172.04
*
Chemical shift (ppm)

Figure 105

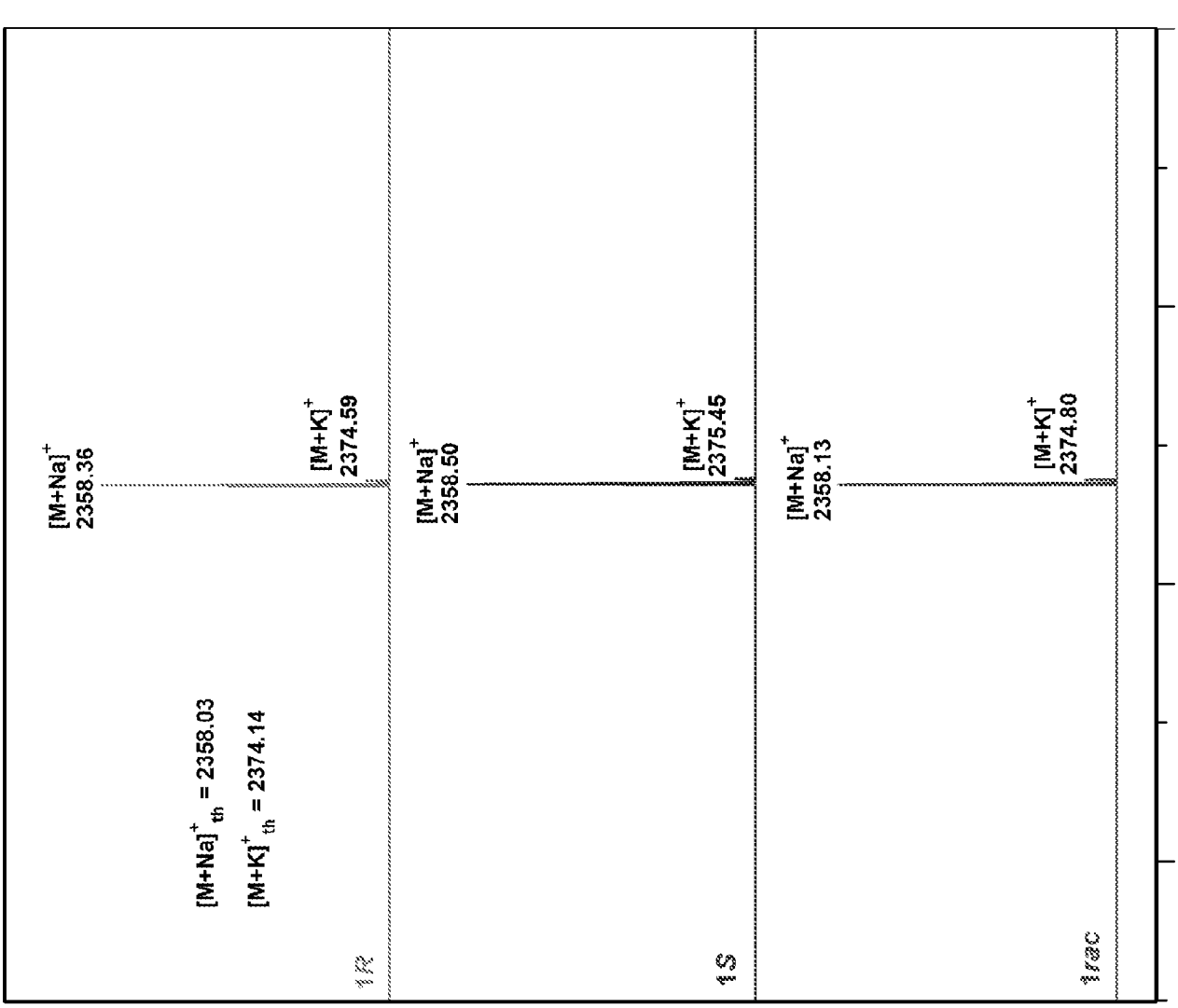

[M+Na]+
2358.36
[M+K]+
2374.59
[M+Na]+th = 2358.03
[M+K]+th = 2374.14
1R
[M+Na]+
2358.50
[M+K]+
2375.45
1S
[M+Na]+
2358.13
[M+K]+
2374.80
1sc
1000
2000
3000
4000
Mass (m/z)

Figure 106

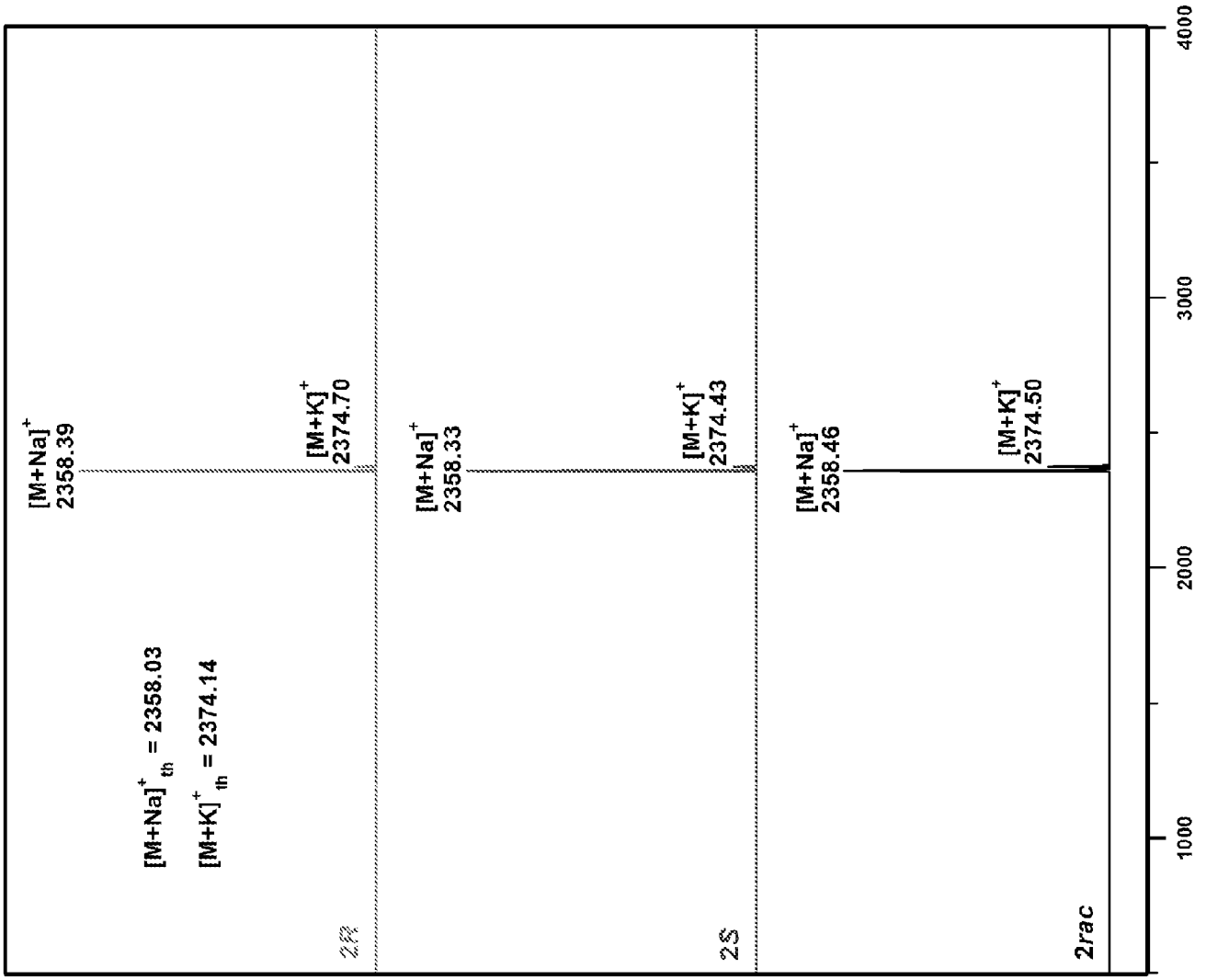
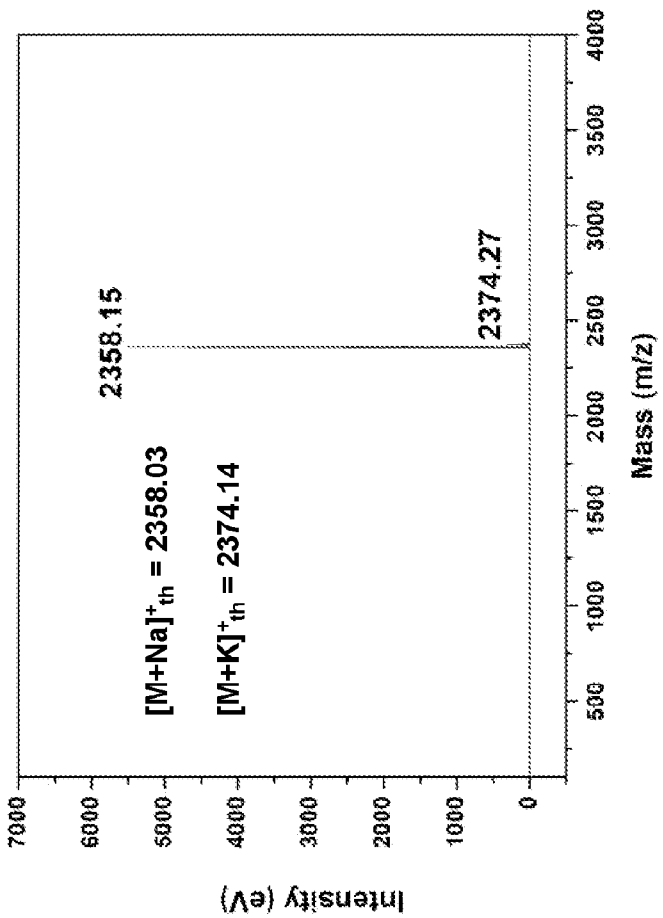

[M+Na]+
2358.39
[M+K]+
2374.70
[M+Na]+th = 2358.03
[M+K]+th = 2374.14
2R
[M+Na]+
2358.33
[M+K]+
2374.43
2S
[M+Na]+
2358.46
[M+K]+
2374.50
2rac
1000
2000
3000
4000
Mass (m/z)

Figure 107


2358.15
2374.27
[M+Na]+th = 2358.03
[M+K]+th = 2374.14
Intensity (eV)
Mass (m/z)

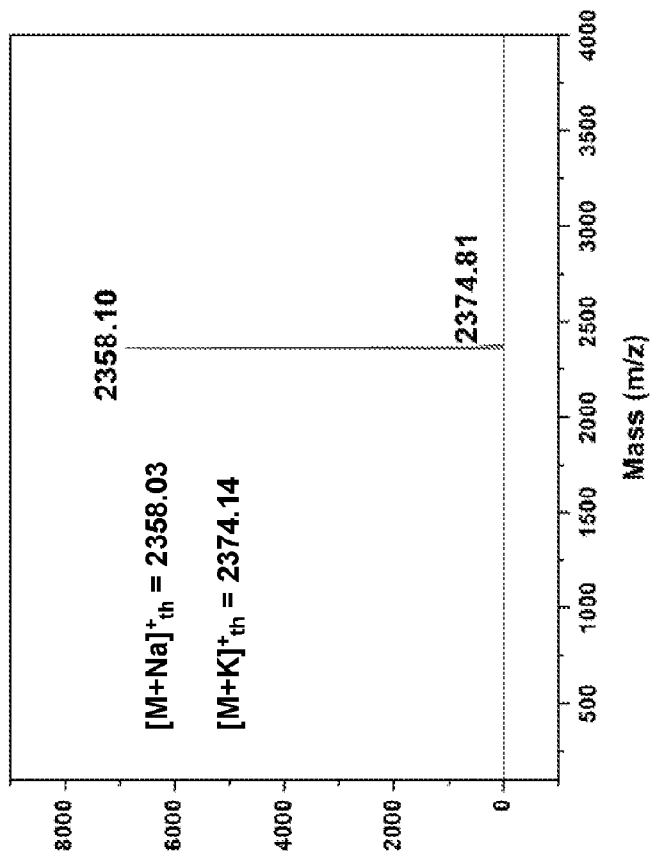

2358.10
2374.81
[M+Na]+th = 2358.03
[M+K]+th = 2374.14
Intensity (eV)
Mass (m/z)

Figure 108

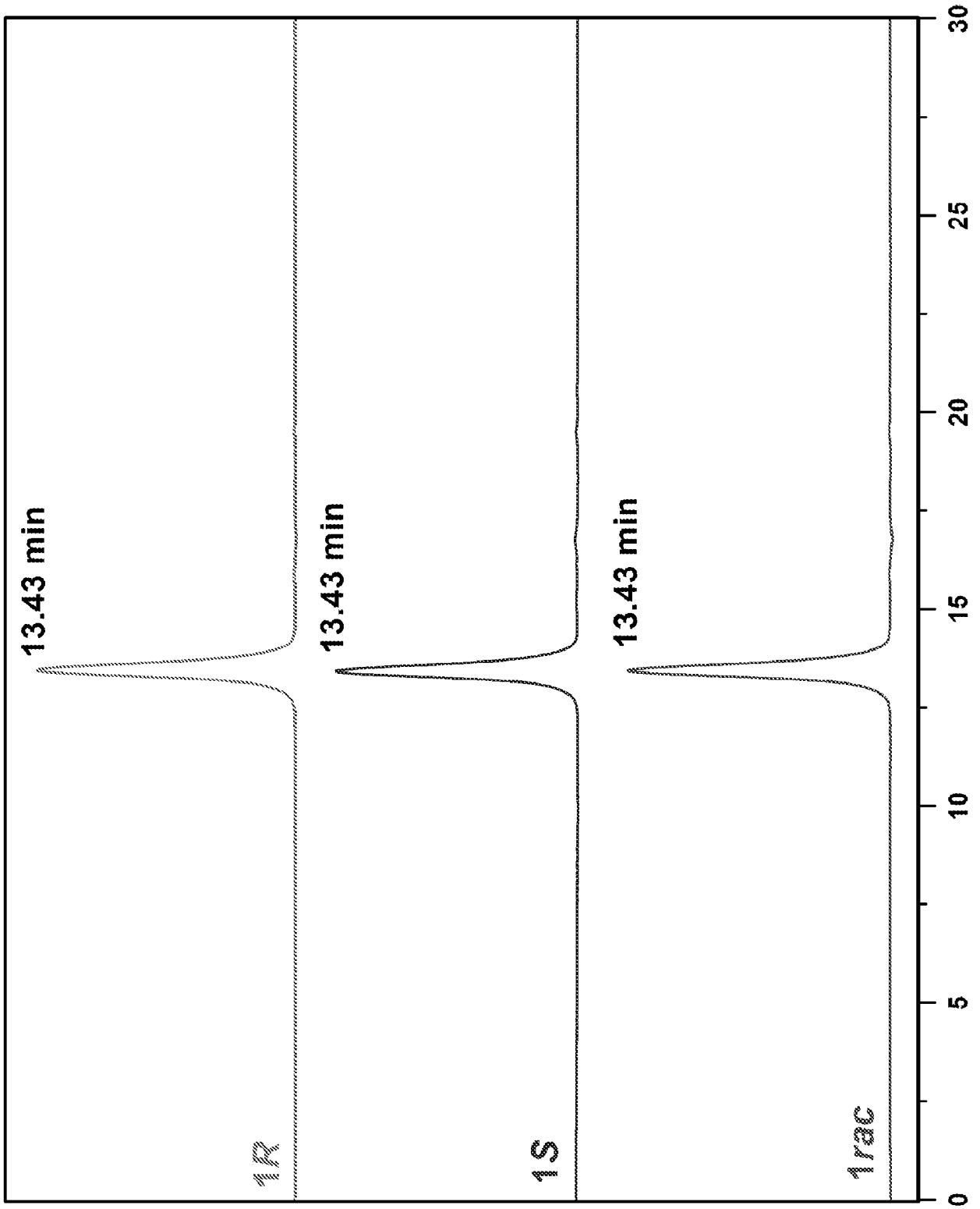

13.43 min
13.43 min
13.43 min
1R
1S
1rac
0
5
10
15
20
25
30
Retention time (min)

Figure 109

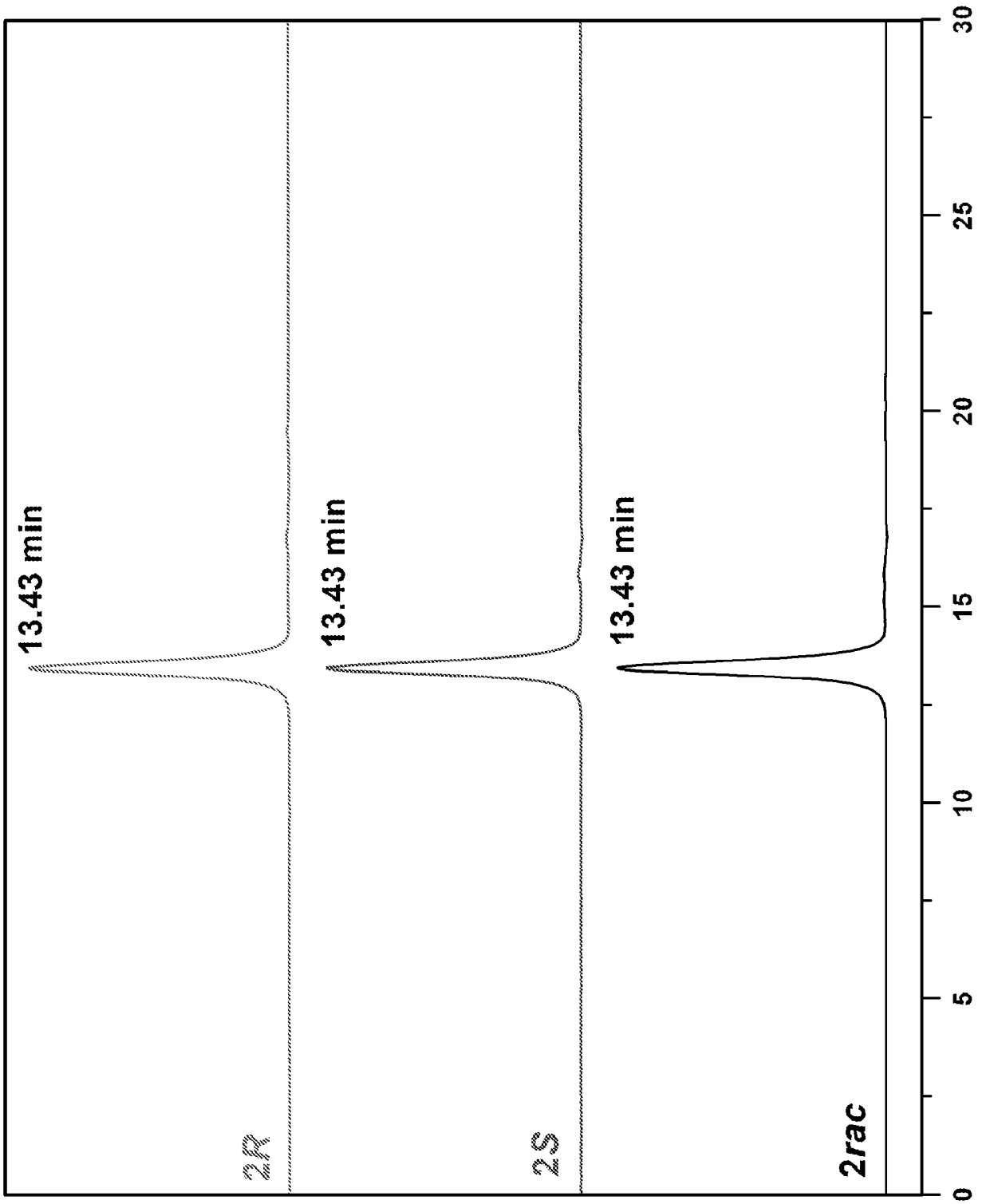
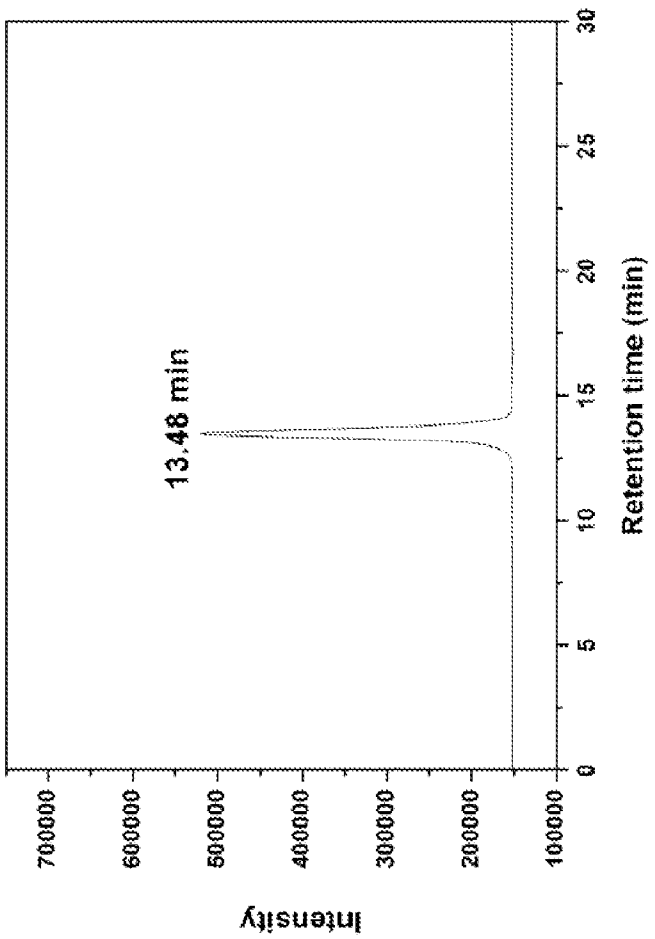

13.43 min
13.43 min
13.43 min
2R
2S
2rac
0
5
10
15
20
25
30
Retention time (min)

Figure 110


13.48 min
Intensity
Retention time (min)
700000
600000
500000
400000
300000
200000
100000
0
5
10
15
20
25
30

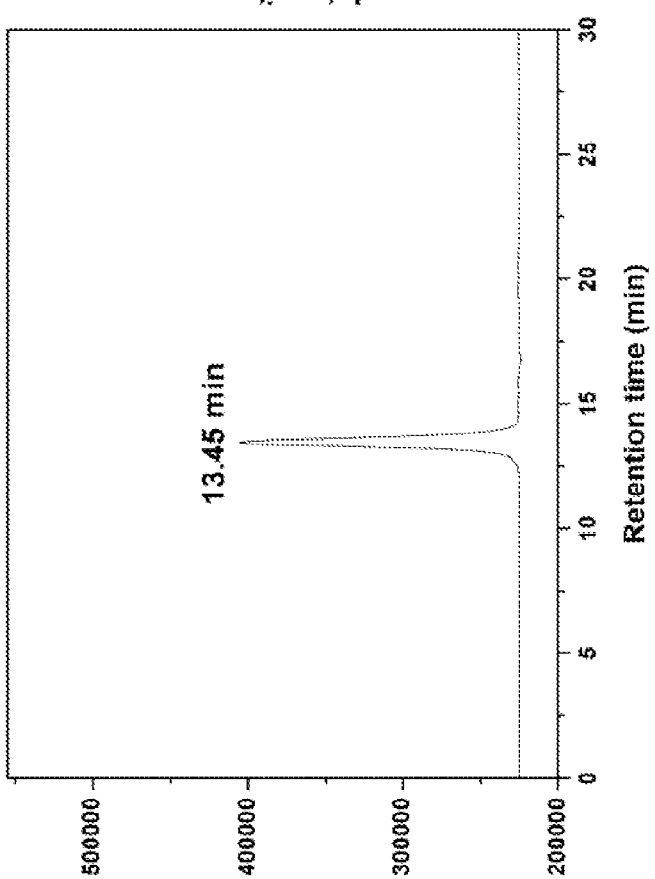

13.45 min
Intensity
Retention time (min)
500000
400000
300000
200000
0
5
10
15
20
25
30

Figure 111

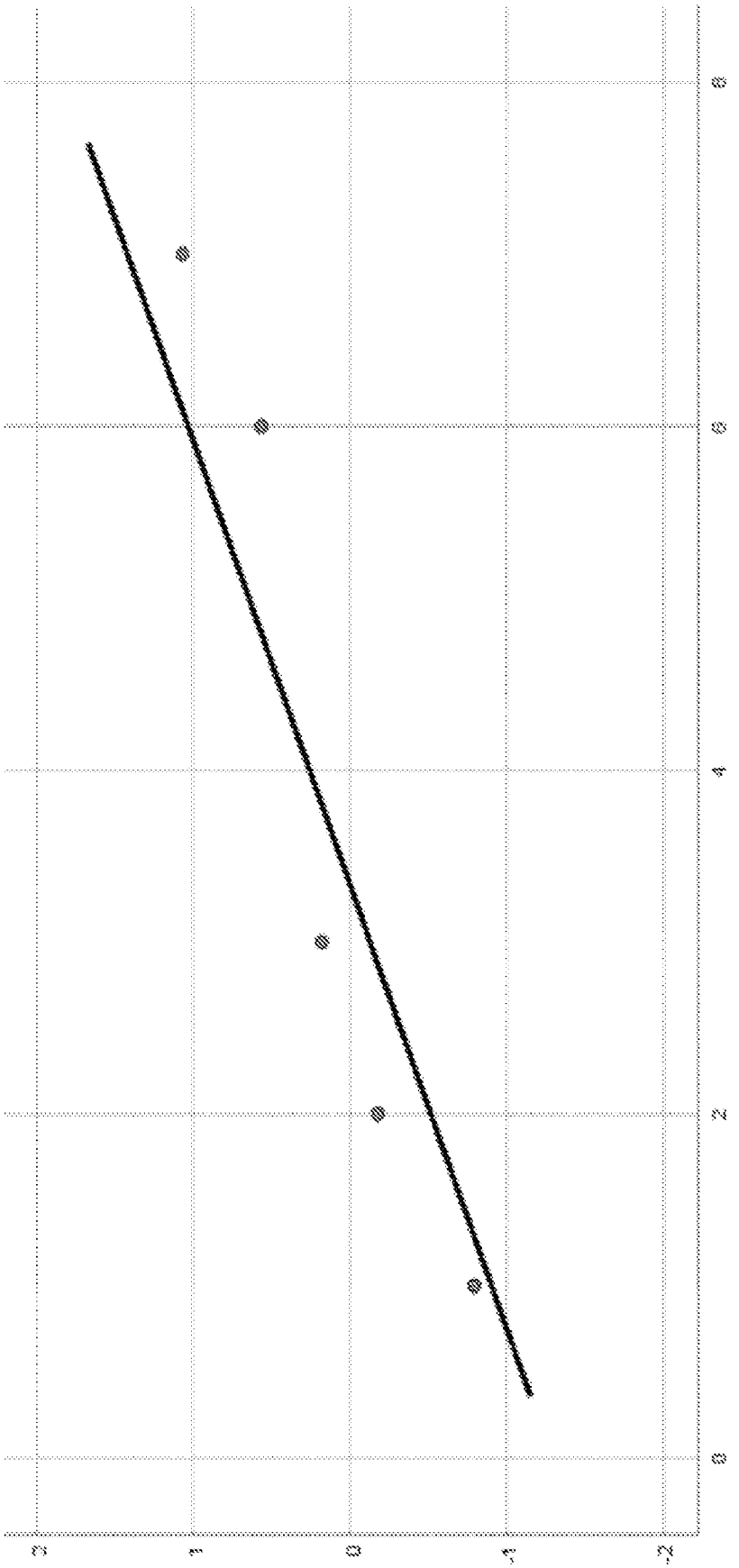


Figure 112

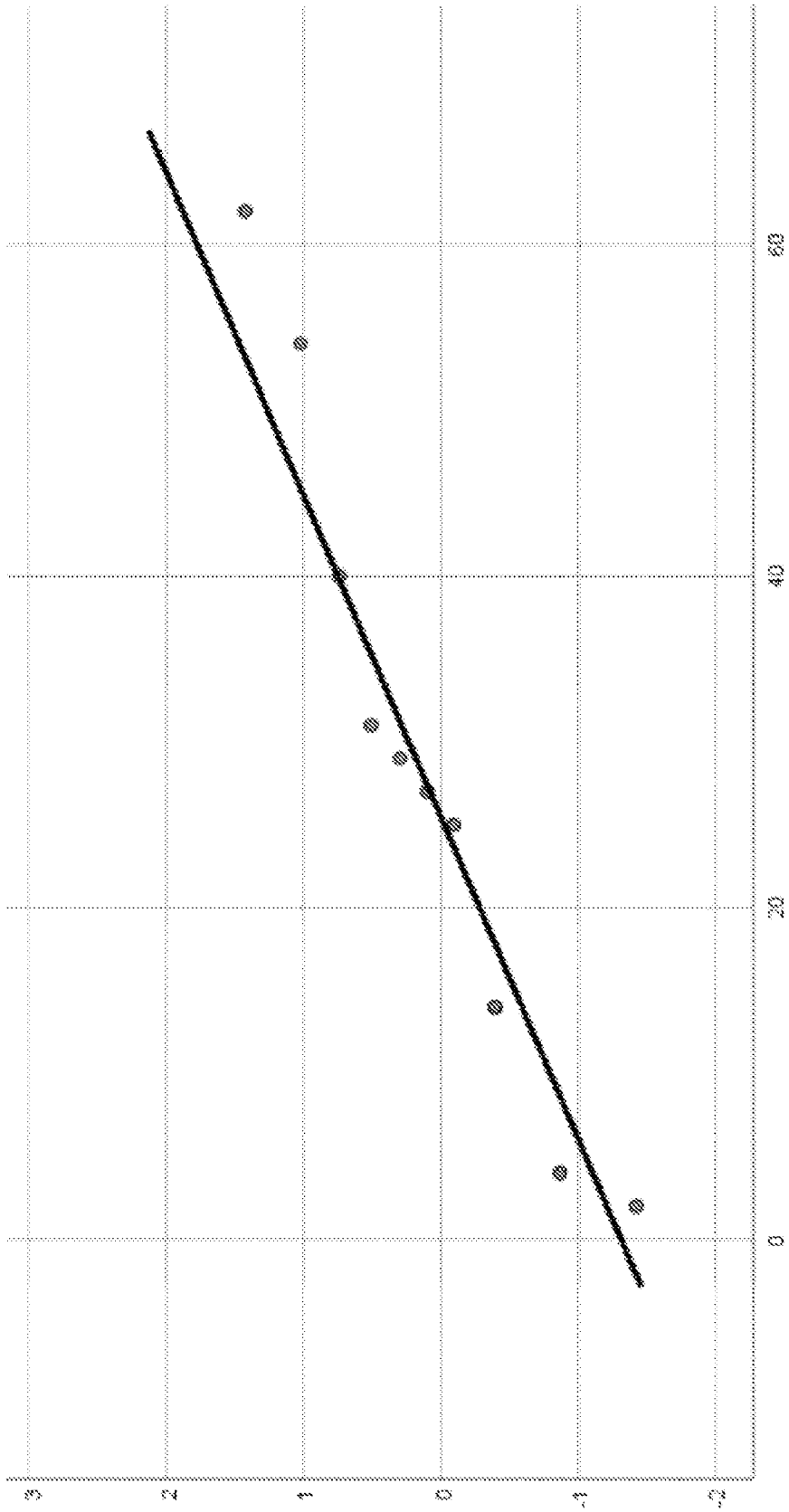


Figure 113