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— 包括国际检索报告(条约第 21 条(3))。

(54) Title: TARGETED PROTEASE DEGRADATION PLATFORM

(54) 发明名称: 靶向蛋白酶降解平台(TED)

(57) Abstract: Disclosed is a targeted protease degradation platform (TED), which in particular is a conjugate of target molecule-linker-E3 ligase ligand as shown in the structure of A-L1-B (formula I), wherein the A is the monovalent group of the target molecule, the B is the monovalent group of the E3 ligase ligand, the L1 is the linker linking A and B, and L1 is as shown in -X-L2-Y- (formula II).

(57) 摘要: 靶向蛋白酶降解平台 (TED), 具体地作为一种结构为 A-L1-B (式 I) 所示的靶标分子-连接体-E3 连接酶配体的偶联物, 其中, 所述 A 为靶标分子的一价基团, 所述 B 为 E3 连接酶配体的一价基团, 所述 L1 为连接 A 和 B 的连接头, 且 L1 如 -X-L2-Y- (式 II) 所示。



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TARGETED PROTEASE DEGRADATION PLATFORM

TECHNICAL FIELD

The present invention belongs to biomedicine, specifically, relates to a Targeted Protease Degradation platform (TED).

BACKGROUND

Expression level of proteins is regulated on three basic levels according to modern molecular biology. Firstly, at the DNA level, the target protein DNA is inactivated through gene knock-out. Secondly, at the mRNA level, it binds to the mRNA of the target protein through small molecule RNA, thereby inhibiting the translation and expression of mRNA. Thirdly, at the protein level, the amount and activity of the target protein can be regulated by modifying of the target protein after translation, such as methylation, phosphorylation, glycosylation, etc.

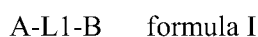
In terms of the overall development of drug research and development, both small molecule and macromolecule drug forms have their own advantages and disadvantages. For example, the development of small molecule drugs has been facing crucial challenges such as how to maintain drug concentration in the body and drug resistance. The shapes of some target sites are adverse to design of small molecule drugs, thus becoming non-drugable targets. For these targets, no effective regulatory methods have yet been found. Although, compared to small molecules, monoclonal antibodies have the advantages of high affinity and high selectivity, and easy to develop highly effective and highly selective drugs, but the biggest drawback thereof is that they cannot penetrate cell membranes and therefore cannot act on intracellular targets. Antibody-drug conjugates (ADC) utilize endocytic antibodies to provide targeting and serve as carriers to deliver super toxin drugs to the targeted site. The bottleneck encountered in the development of ADC drugs is that the treatment window is not wide enough. In addition to the side effects caused by the antibody itself, the super toxins will fall off before reaching the targeting site due to the heterogeneity of coupling, and causing serious side effects. In addition, normal physiological function of ubiquitin- proteasome system is responsible for cleaning up denatured, mutated or harmful proteins in cells.

Summary, there is an urgent need in the art to develop a compound that is able to degrade target proteins more efficiently and re-usably so as to treat related diseases.

SUMMARY OF THE INVENTION

The purpose of the present application is to provide a compound that is able to degrade target proteins more efficiently and re-usably so as to treat related diseases.

In the first aspect of the present invention, a conjugate of formula I is provided:



wherein

A is a monovalent moiety of a targeting molecule;

B is a monovalent moiety of an E3 ligase ligand;

L1 is a linker connecting A and B; and L1 has a formula II:



wherein

when X or Y is NH or NR, X or Y and part of L2 chain to which X or Y is attached are combined together to form a 4 - 8 membered aromatic or non-aromatic heterocyclic ring; wherein, R is substituted or unsubstituted C1-C10 alkyl, $-(C=O)-R'$, $(C=O)NH-R'$, $-NH(C=O)-R'$, $-SO_2-R'$, $-NHSO_2-R'$, $-SO_2NH-R'$,

-SO-R', -NHCO-R', -SONH-R', -PO₃-R', -NHCOO-R', -COO-R', or -NH-CO-NH-R', -NH-CO-O-R', or -X'-L3-Z; wherein L3 is linker, and Z is a polypeptide element or target molecule T; wherein R' is selected from the group consisting of H, substituted or unsubstituted C1-C6 alkyl, substituted or unsubstituted C3-C8 cycloalkyl, amino protecting group, and -X'-L3-Z;

5 or

each X and Y is independently selected from the group consisting of -O-, -S-, -NH-, -NR-, -(C=O)NH-, -NH(C=O)-, -SO₂-, -NHCO₂-, -SO₂NH-, -SO-, -NHCO-, -SONH-, -PO₃-, -NHCOO-, -COO-, -NHCOCH₂O-, and -NH-CO-NH-; wherein R is substituted or unsubstituted C1-C10 alkyl, -(C=O)-R', -(C=O)NH-R', -NH(C=O)-R', -SO₂-R', -NHCO₂-R', -SO₂NH-R', -SO-R', -NHCO-R', -SONH-R', -PO₃-R', -NHCOO-R', -COO-R' or -NH-CO-NH-R', or -X'-L3-Z;

10 wherein L3 is linker, and Z is a polypeptide element or target molecule T; wherein R' is selected from the group consisting of H, substituted or unsubstituted C1-C6 alkyl, substituted or unsubstituted C3-C8 cycloalkyl, protecting group, and -X'-L3-Z;

15 X' is selected from the group consisting of -O-, -S-, -NH-, -(C=O)NH-, -NH(C=O)-, -SO₂-, -NHCO₂-, -SO₂NH-, -SO-, -NHCO-, -SONH-, -PO₃-, -NHCOO-, -COO-, -COOCH₂-, and -NH-CO-NH-;

each L2 and L3 is independently linker selected from the group consisted of

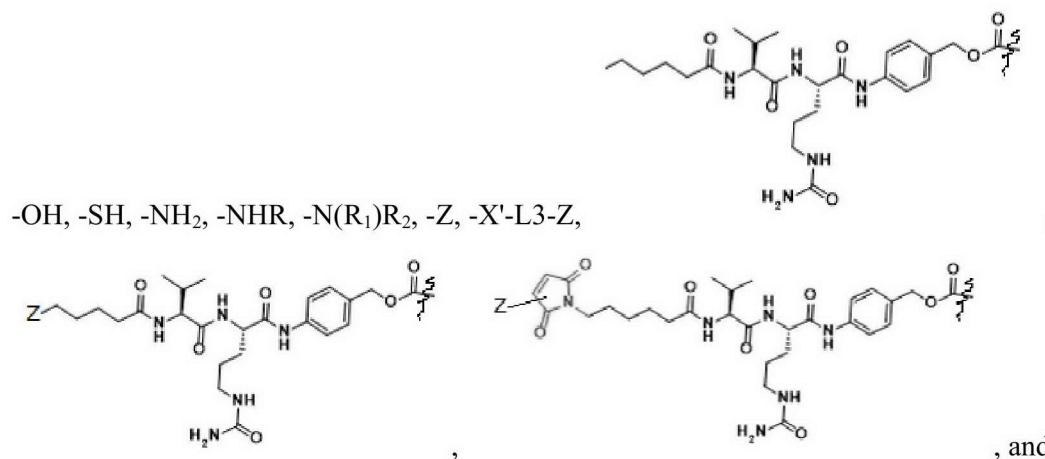
(a) substituted or unsubstituted 5-20 membered carbon chain or 5-50 membered carbon chain;

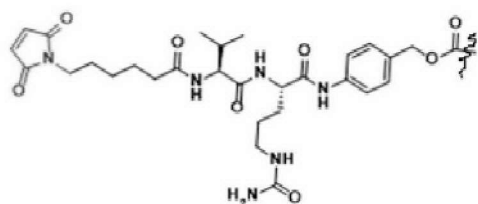
(b) substituted or unsubstituted 5-20 membered heterocarbon chain or 5-50 membered heterocarbon chain containing 1 to 5 heteroatoms selected from N, O and S;

20 (c) substituted or unsubstituted 5-20 membered carbon chain or 5-50 membered carbon chain; and 1 to 4 chain atoms in the chain are replaced with heterocyclic ring selected from the group consisting of 4-10 membered saturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S, and 5-10 membered partially unsaturated or fully unsaturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S;

25 (d) substituted or unsubstituted 5-20 membered heterocarbon chain or 5-50 membered heterocarbon chain containing 1 to 5 heteroatoms selected from N, O and S; and 1 to 4 chain atoms in the chain are replaced with heterocyclic ring selected from the group consisting of 4-10 membered saturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S, and 5-10 membered partially unsaturated or fully unsaturated heterocyclic rings containing 1- 4 heteroatoms selected from N, O and S;

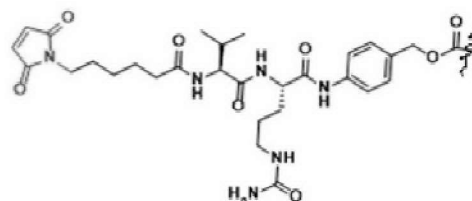
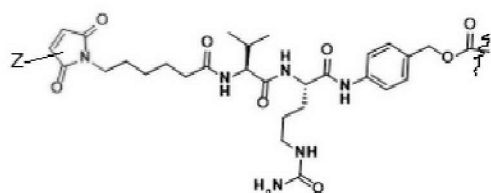
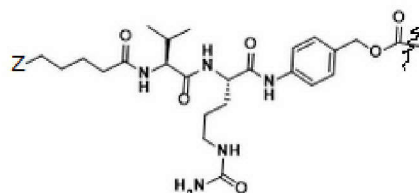
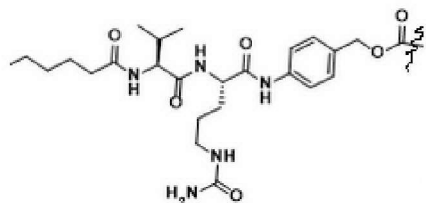
30 wherein the "substituted" means one or more (preferably, 1, 2, 3 or 4) hydrogen atoms in the group is substituted with substituent selected from the group consisting of C1-C10 alkyl, C3-C8 cycloalkyl, -COOH,





; wherein Z is a polypeptide element or target molecule T;

wherein when the substituent is -Z, -Z is located only on N atom; wherein each R₁ and R₂ is indenpendently H, C1-C6 alkyl, C3-C8 cycloalkyl, amino protecting group, -X'-L3-Z,



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, or

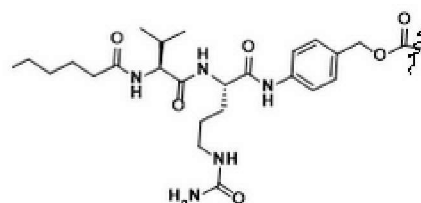
; wherein R is

substituted or unsubstituted C1-C10 alkyl, -(C=O)-R', (C=O)NH-R', -NH(C=O)- R', -SO₂-R', -NHSO₂-R', -SO₂NH-R', -SO-R', -NH-SO-R', -SONH-R', -PO₃-R', -NHCOO-R', -COO-R', or -NH-CO-NH- R',

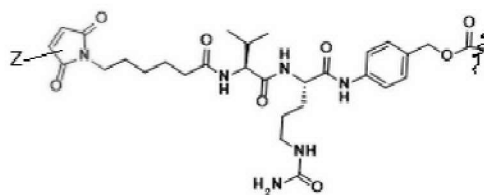
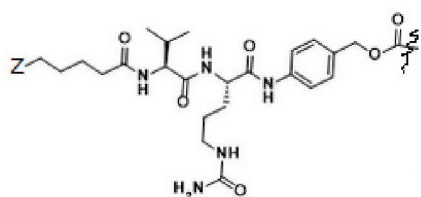
-NH-CO-O-R', or -X'-L3-Z; wherein L3 is linker, and Z is a polypeptide element or target molecule T;

wherein R' is selected from the group consisting of H, substituted or unsubstituted C1-C6 alkyl, substituted or unsubstituted C3-C8 cycloalkyl, protecting group, and X'-L3-Z.

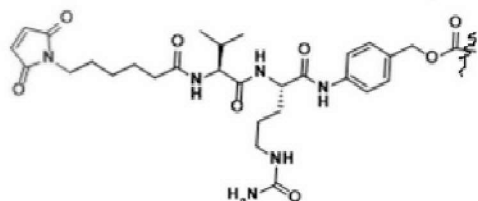
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In another preferred embodiment, R₁ and R₂ are not -X'-L3-Z,

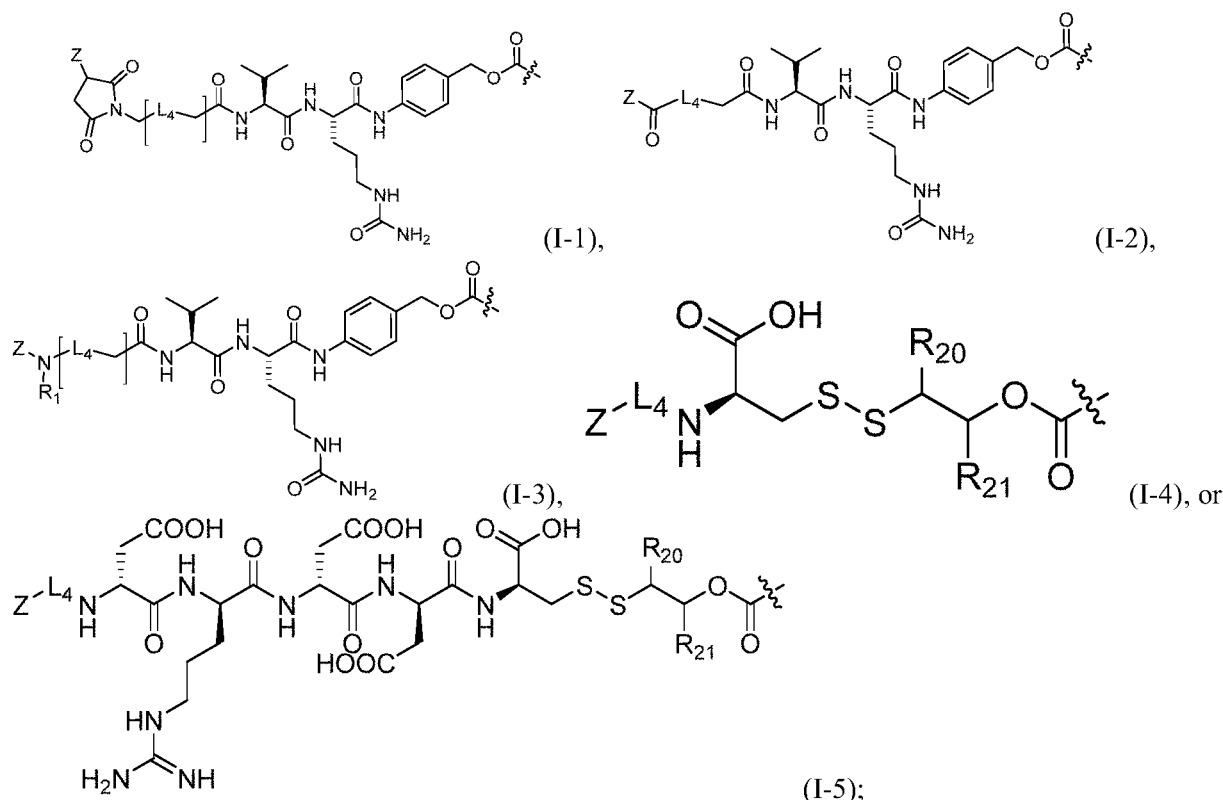


, or



at the same time.

In another preferred embodiment, -X'-L3-Z is a group of fomula I-1, I-2, I-3, I-4, or I-5:



wherein, each R_{20} and R_{21} is independently selected from the group consisting of -H, -Me, -Et, -nPr, iPro, cPro;

L_4 is defined the same as L_3 .

In another preferred embodiment, when $-X'-L_3-Z$ is a group of I-1, I-2, or I-3, Z is a polypeptide element.

In another preferred embodiment, when $-X'-L_3-Z$ is a group of I-4, or I-5, Z is a target molecule T.

In another preferred embodiment, L_4 is a linker selected from the group consisted of

(a) substituted or unsubstituted 5-20 membered carbon chains;

(b) substituted or unsubstituted 5-20 membered heterocarbon chains containing 1 to 5 heteroatoms selected from N, O and S;

(c) substituted or unsubstituted 5-20 membered carbon chains, and 1 to 4 chain atoms in the chain are replaced with heterocyclic rings selected from the group consisting of 4-10 membered saturated heterocyclic rings containing 1- 4 heteroatoms selected from N, O and S, and 5-10 membered partially unsaturated or fully unsaturated heterocyclic rings containing 1- 4 heteroatoms selected from N, O and S;

(d) substituted or unsubstituted 5-20 membered heterocarbon chains containing 1 to 5 heteroatoms selected from N, O and S, and 1 to 4 chain atoms in the chain are replaced with heterocyclic rings selected from the group consisting of 4-10 membered saturated heterocyclic rings containing 1- 4 heteroatoms selected from N, O and S, and 5-10 membered partially unsaturated or fully unsaturated heterocyclic rings containing 1- 4 heteroatoms selected from N, O and S;

wherein the “substituted” means one or more (preferably, 1, 2, 3 or 4) hydrogen atoms in the group is substituted with substituents selected from the group consisting of C1-C10 alkyl, C3-C8 cycloalkyl, -COOH, -OH, -SH, -NH₂, -NHR, -N(R₁)R₂; wherein each R₁ and R₂ is indenpedently H, C1-C6 alkyl, C3-C8 cycloalkyl, amino protecting group; R is substituted or unsubstituted C1-C10 alkyl, -(C=O)-R', (C=O)NH-R', -NH(C=O)- R', -SO₂-R', -NHSO₂-R', - SO₂NH-R', -SO-R', -NHSO-R', -SONH-R', -PO₃-R', -NHCOO-R', -COO-R', or -NH-CO-NH- R', or -NH-CO-O-R'; wherein R' is selected from the group consisting of H, substituted or unsubstituted C1-C6 alkyl, substituted or unsubstituted C3-C8 cycloalkyl,

and protecting group.

In another preferred embodiment, L4 is linker selected from the group consisted of

(a) substituted or unsubstituted 5-20 membered carbon chain;

(b) substituted or unsubstituted 5-20 membered heterocarbon chain containing 1 to 5 heteroatoms
5 selected from N, O and S.

In another preferred embodiment, the polypeptide element is selected from the group consisting of ligand or active fragment thereof, receptor or active fragment thereof, antibody or active fragment thereof, and a combination thereof.

In another preferred embodiment, Z is selected from the group consisting of ligand, soluble receptor,
10 antibody, or antibody-drug conjugate (ADC), and a combination thereof.

In another preferred embodiment, the target molecule T and the target molecule A are the same or different.

In another preferred embodiment, the target molecule T and the target molecule A are small molecules with targeting functions in the drug conjugates.

In another preferred embodiment, the target molecule T and the target molecule A are each
15 independently selected from the group consisting of folic acid, HSP90 inhibitor, and a combination thereof.

In another preferred embodiment, the ligand comprises full-length ligand or active fragment thereof.

In another preferred embodiment, the antibody comprises full-length antibody or active fragment
20 thereof.

In another preferred embodiment, the antibody comprises single-chain antibody or double-chain antibody.

In another preferred embodiment, the 4-10 membered saturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S can be single ring, double ring or triple ring.

In another preferred embodiment, the 4-10 membered saturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S can be fused ring or spiro ring.

In another preferred embodiment, the 4-10 membered saturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S can be a 4 - 6 membered saturated ring containing 1 nitrogen atom or a 4 - 6 membered saturated ring containing 1 nitrogen atom.

In another preferred embodiment, the 5-10 membered partially unsaturated or fully unsaturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S can be single ring, double ring or triple ring.

In another preferred embodiment, the 5-10 membered partially unsaturated or fully unsaturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S can be fused ring or spiro ring.

In another preferred embodiment, the 5-10 membered partially unsaturated or fully unsaturated heterocyclic rings containing 1- 4 heteroatoms selected from N, O and S can be 5 membered partially unsaturated or fully unsaturated heterocyclic rings containing 1- 4 heteroatoms selected from N, O and S.

In another preferred embodiment, L2 and/or L3 are linkers selected from the group consisted of

(a) substituted or unsubstituted $-(CH_2)_s-$;

(b) substituted or unsubstituted $-(CH_2)_s-$, wherein 1-5 of $-CH_2-$ are replaced with O;

(c) substituted or unsubstituted $-(CH_2)_s-$, wherein 1-2 of $-CH_2-$ are replaced with NR and optionally 1 - 2 of $-CH_2-$ are replaced with O; wherein R is H, protecting group or X^1-L3-Z ;

(d) substituted or unsubstituted $-(CH_2)_s-$, wherein 1-4 of $-CH_2-$ are replaced with 5 membered

unsaturated heterocyclic ring containing 1-4 heteroatoms selected from N, O, S and P;

(e) substituted or unsubstituted $-(CH_2)_s-$, wherein 1-4 of $-CH_2-$ are replaced with 4-6 membered saturated heterocyclic ring containing 1-4 heteroatoms selected from N, O, S and P;

(f) substituted or unsubstituted $-(CH_2)_s-$, wherein 1-4 of $-CH_2-$ are replaced with O and 4-6 membered saturated heterocyclic ring containing 1-4 heteroatoms selected from N, O, S and P;

(g) divalent 4-6 membered heterocyclic group containing 1-2 nitrogen atoms;

wherein, s is an integer of 5-20, or an integer of 5-50.

In another preferred embodiment, s is 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20.

In another preferred embodiment, the substituted $-(CH_2)_s-$ means substituted with one or more groups selected from the group consisting of C1-C10 alkyl, C3-C8 cycloalkyl, $-NH_2$, $-NHR$, $-NR_1R_2$, wherein, each R_1 and R_2 is independently H, C1-C6 alkyl, C3-C8 cycloalkyl, protecting group, and R, wherein R is defined as above, and R_1 and R_2 are not the polypeptide element or target molecule T at the same time.

In another preferred embodiment, the L2 and/or L3 are selected from the group consisting of

(d1) substituted or unsubstituted $-(CH_2)_s-$, wherein 1-4 of $-CH_2-$ are replaced with 5 membered unsaturated heterocyclic rings containing 1-3 heteroatoms selected from N, O, S and P;

(e1) substituted or unsubstituted $-(CH_2)_s-$, wherein 1-4 of $-CH_2-$ are replaced with 4-6 membered saturated heterocyclic rings containing 1-3 heteroatoms selected from N, O, S and P;

(f1) substituted or unsubstituted $-(CH_2)_s-$, wherein 1-4 of $-CH_2-$ are replaced with O and 4-6 membered saturated heterocyclic rings containing 1-3 heteroatoms selected from N, O, S and P.

In another preferred embodiment, the L2 and/or L3 are selected from the group consisting of

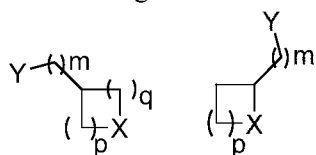
(d2) substituted or unsubstituted $-(CH_2)_s-$, wherein 1-4 of $-CH_2-$ are replaced with 5 membered unsaturated heterocyclic rings containing 1-2 N atoms;

(e2) substituted or unsubstituted $-(CH_2)_s-$, wherein 1-4 of $-CH_2-$ are replaced with 4-6 membered saturated heterocyclic rings containing 1-2 N atoms;

(f2) substituted or unsubstituted $-(CH_2)_s-$, wherein 1-4 of $-CH_2-$ are replaced with O and 4-6 membered saturated heterocyclic rings containing 1-2 N atoms.

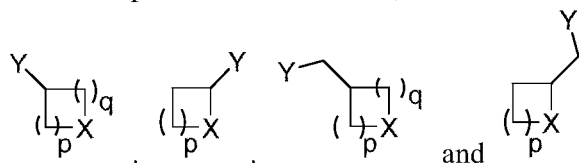
In another preferred embodiment, s is 5-15, preferably 6-10.

In another preferred embodiment, when X is NH or NR, X and part of L2 chain to which X is attached are combined together to form a 4-8 membered heterocyclic ring,



wherein Y and R are defined as above; m is an integer of 1-15; each p and q is independently 0, 1, 2, or 3, and $p + q \geq 2$.

In another preferred embodiment, L1 is selected from the group consisting of

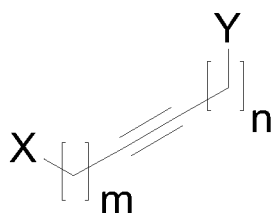


wherein

X and Y are defined as above;

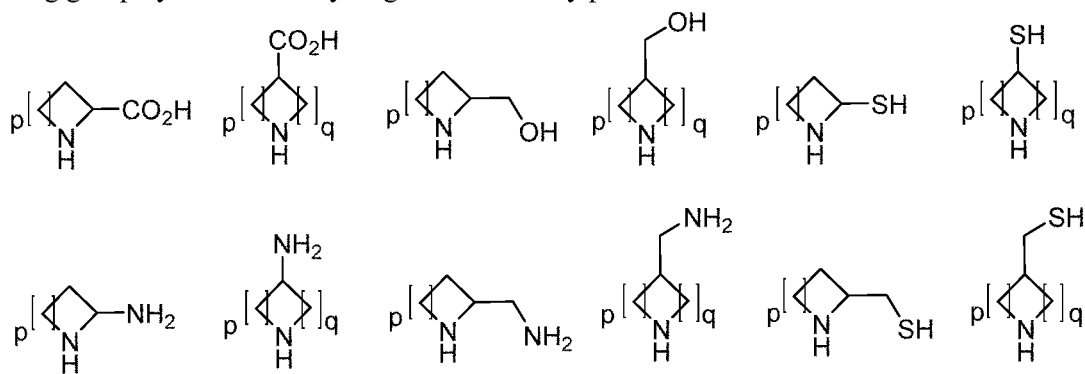
each p and q is independently an integer of 1, 2, or 3.

In another preferred embodiment, L1 is



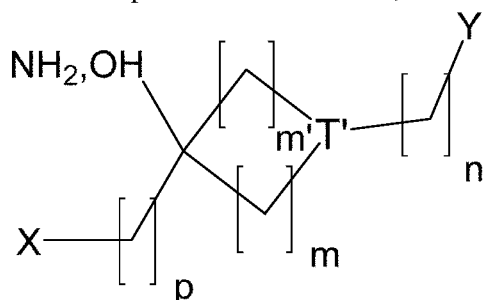
wherein each m and n is independently an integer of 1 - 15.

In another preferred embodiment, L1 is a divalent linker formed from a group selected from the following group by loss of two hydrogen atoms at any position:



in the formula, p and q are defined as above.

In another preferred embodiment, L1 is derived from



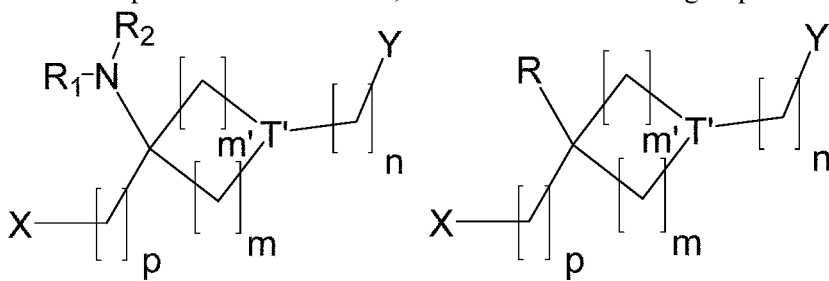
wherein m' and m are each independently 0, 1, 2, 3, 4, or 5 (preferably 0, 1, 2, or 3), and m' and m are not 0 at the same time;

each p and n is independently an integer of 1 - 15;

in the formula, "NH₂,OH" means the terminal is NH₂ or OH;

T' is N or CR_b; wherein R_b is a group selected from the group consisting of H, and substituted or unsubstituted C1-C6 alkyl.

In another preferred embodiment, L1 is selected from the group consisting of

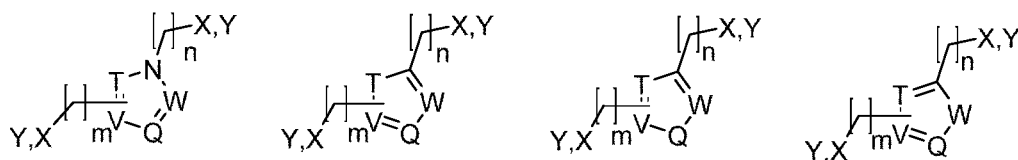


wherein m' and m are each independently 0, 1, 2, 3, 4, or 5 (preferably 0, 1, 2, or 3), and m' and m are not 0 at the same time;

each p and n is independently an integer of 1 - 15;

the definitions of T', R, R₁ and R₂ are as above.

In another preferred embodiment, L1 is selected from the group consisting of



in the formula, “Y, X” means this terminal is X or Y; with the proviso that one end is X, and the other end is Y;

X and Y are defined as above, each m and n is independently an integer of 1 - 15;

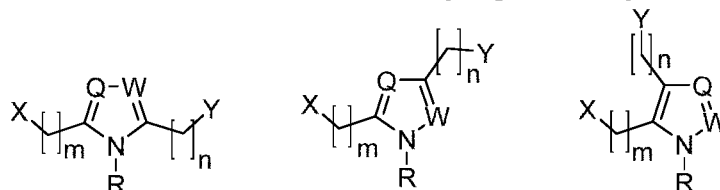
- 5 T, V, Q and W are each independently CH, C=O, S, O, NH or NR; R is C1-C10 alkyl, -(C=O)NH-, -(C=O)NH-R', -NH(C=O)-R', -SO₂-R', -NHSO₂-R', -SO₂NH-R', -SO-R', -NHSO-R', -SONH-R', -PO₃-R', -NHCOO-R', -COO-R', -NH-CO-NH-R', -NH-CO-O-R', or N'-X'-L3-Z (i.e. -X'-L3-Z is a substituent on the N); wherein the definition of L3 is as described above, and Z is a polypeptide element or target molecule T; R' is defined as above.

- 10 In another preferred embodiment, at least one of T, V, Q and W is N'-X'-L3-Z.

In another preferred embodiment, m+n≤20.

In another preferred embodiment, m+n≥2.

In another preferred embodiment, L1 is selected from the group consisting of



- 15 in each formula, X and Y are defined as above, each m and n is independently an integer of 1 - 15;

R is X'-L3-Z, and L3 and Z are defined as above;

Q and W are each independently CH, C=O, S, O, NH or NR; R is -(C=O)-R', (C=O)NH-R', -NH(C=O)-R', -SO₂-R', -NHSO₂-R', -SO₂NH-R', -SO-R', -NHSO-R', -SONH-R', -PO₃-R', -NHCOO-R', -COO-R', -NH-CO-NH-R' or -NH-CO-O-R'; R' is defined as above.

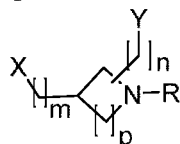
- 20 In another preferred embodiment, Z is the polypeptide element selected from antibody, receptor, ligand, or active fragment thereof.

In another preferred embodiment, Z is ADC.

In another preferred embodiment, m+n≤20.

In another preferred embodiment, m+n≥2.

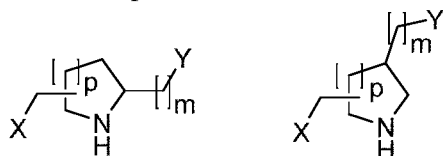
- 25 In another preferred embodiment, L1 is



wherein X, Y and R are defined as above;

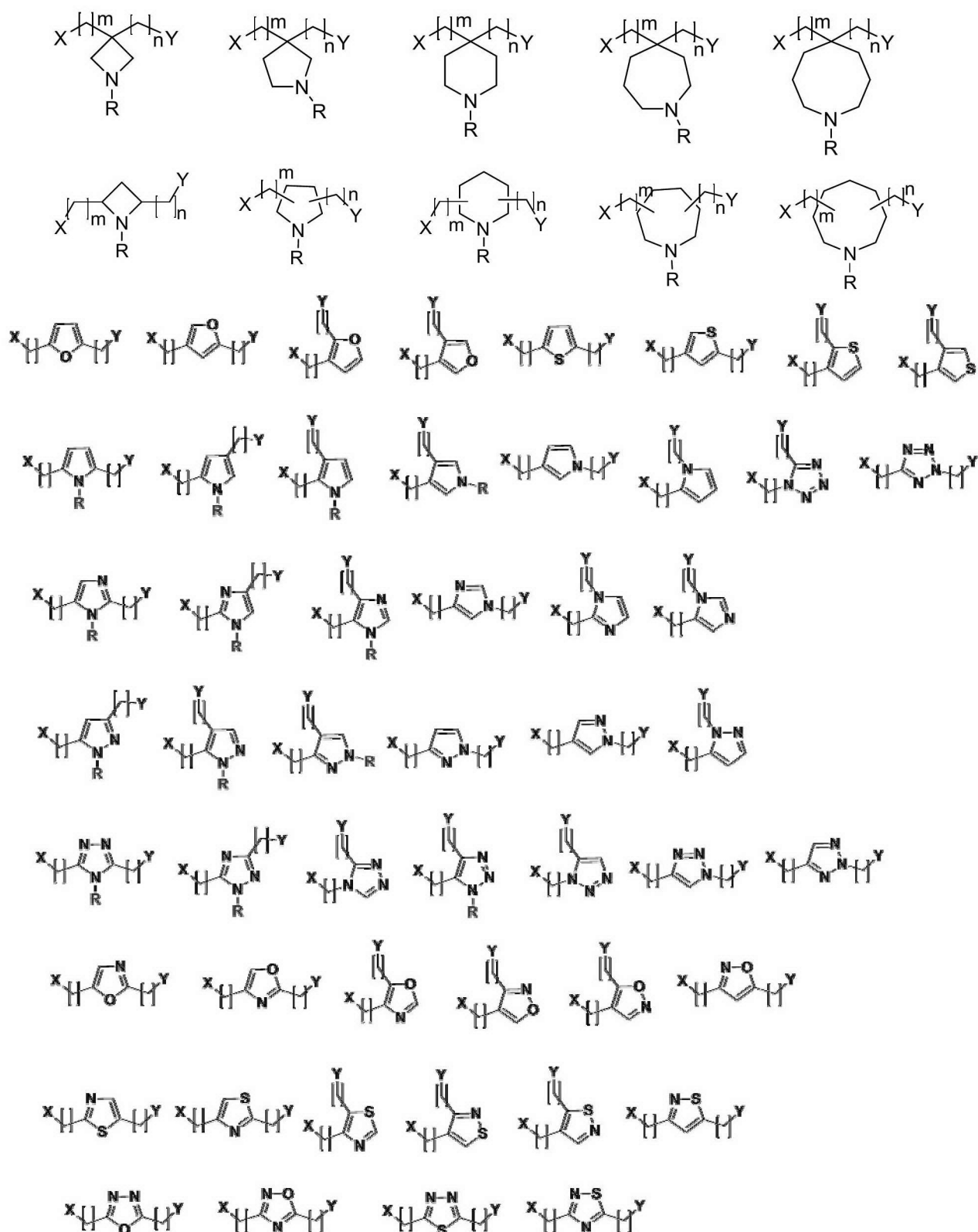
each m and n is independently an integer of 1-15; p is 1, 2, or 3.

In another preferred embodiment, L1 is selected from the group consisting of



- 30 in each formula, X and Y are defined as above, m is an integer of 1 - 15; p is 1, 2 or 3.

In another preferred embodiment, the linker (L1) is selected from the group consisting of



in the formula, X, Y and R are defined as above, each m and n is independently an integer of 1 - 15.

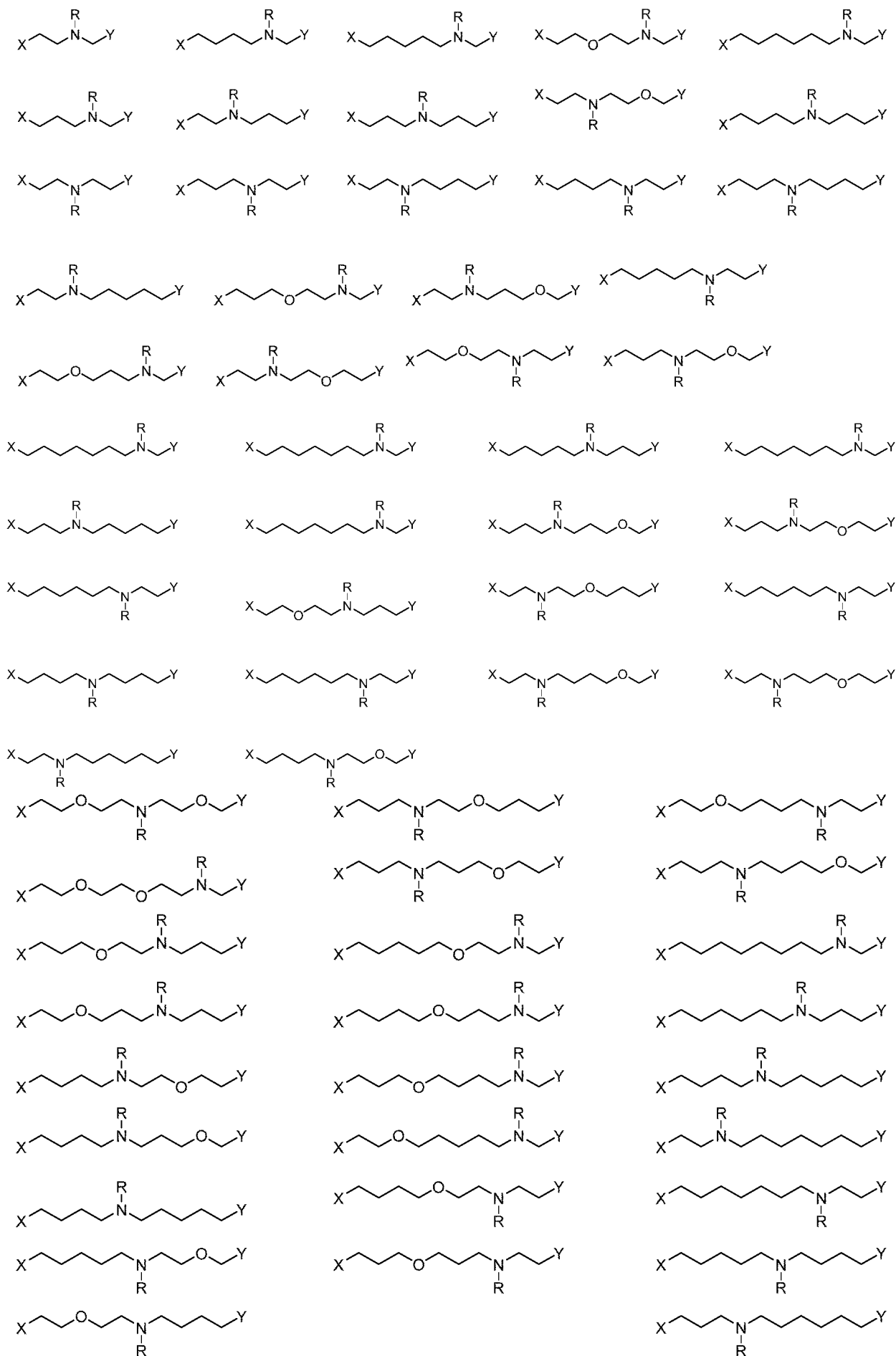
5

In another preferred embodiment, $m+n \leq 20$.

In another preferred embodiment, $m+n \geq 2$.

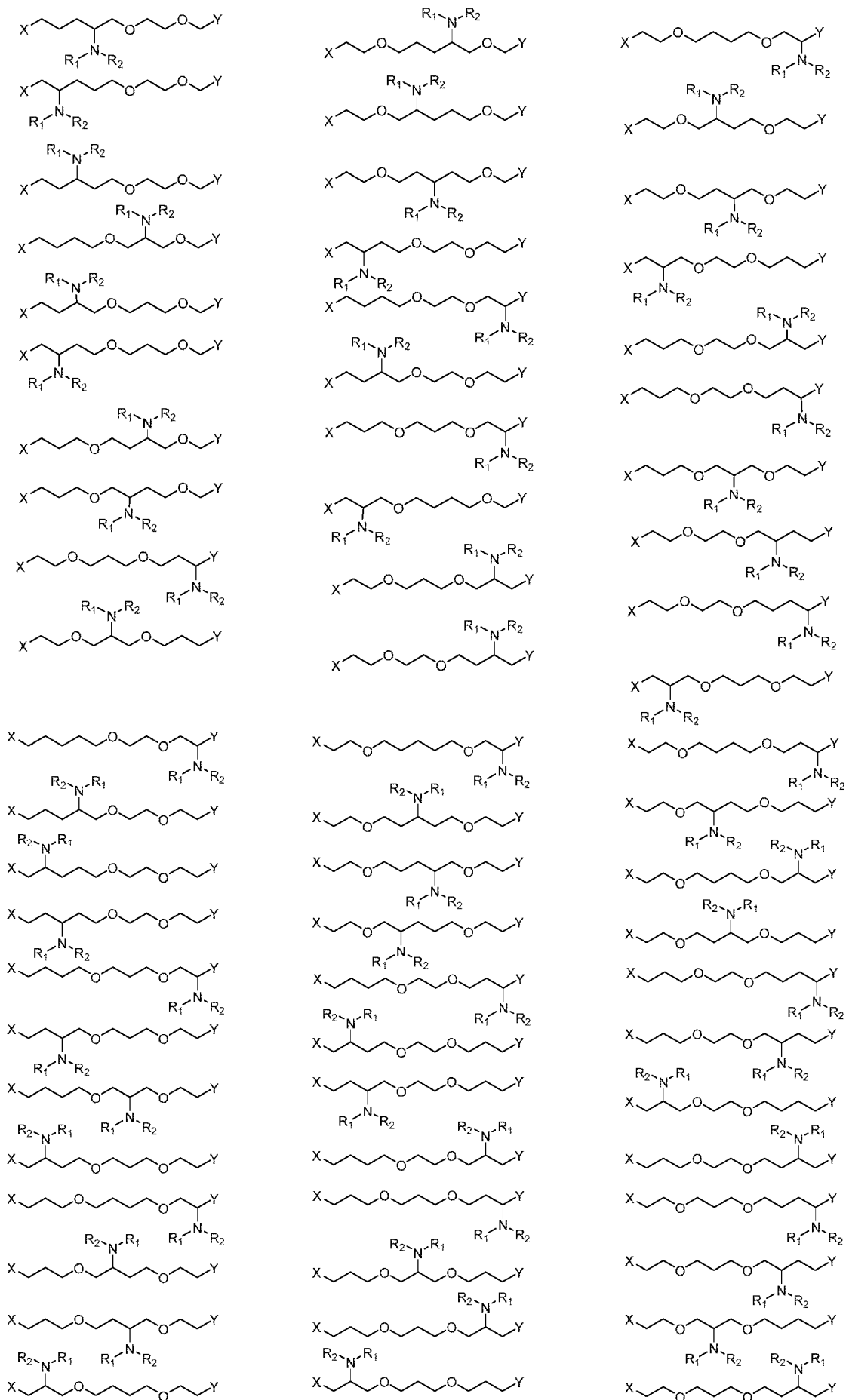
In another preferred embodiment, L1 is selected from the group consisting of





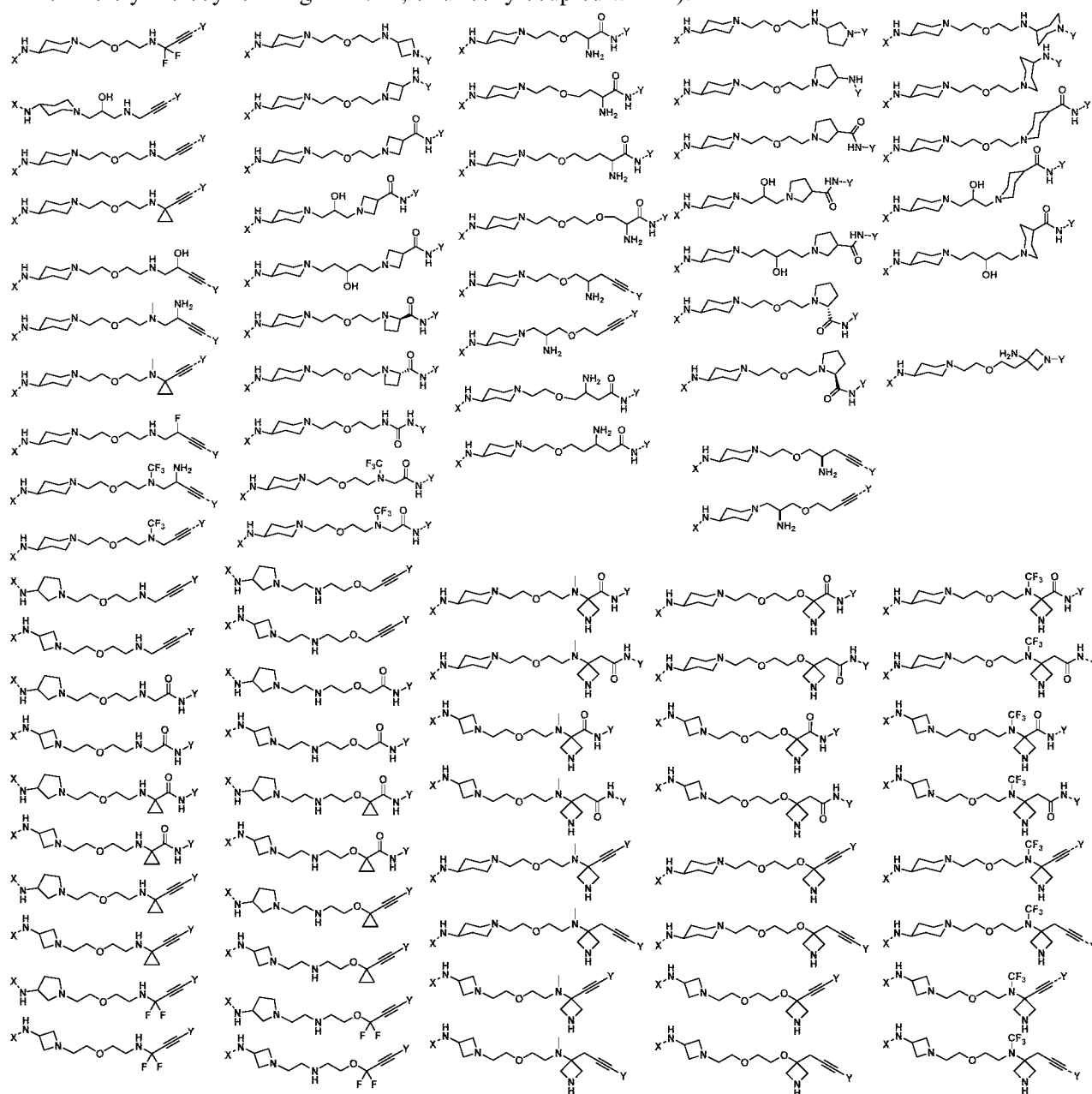


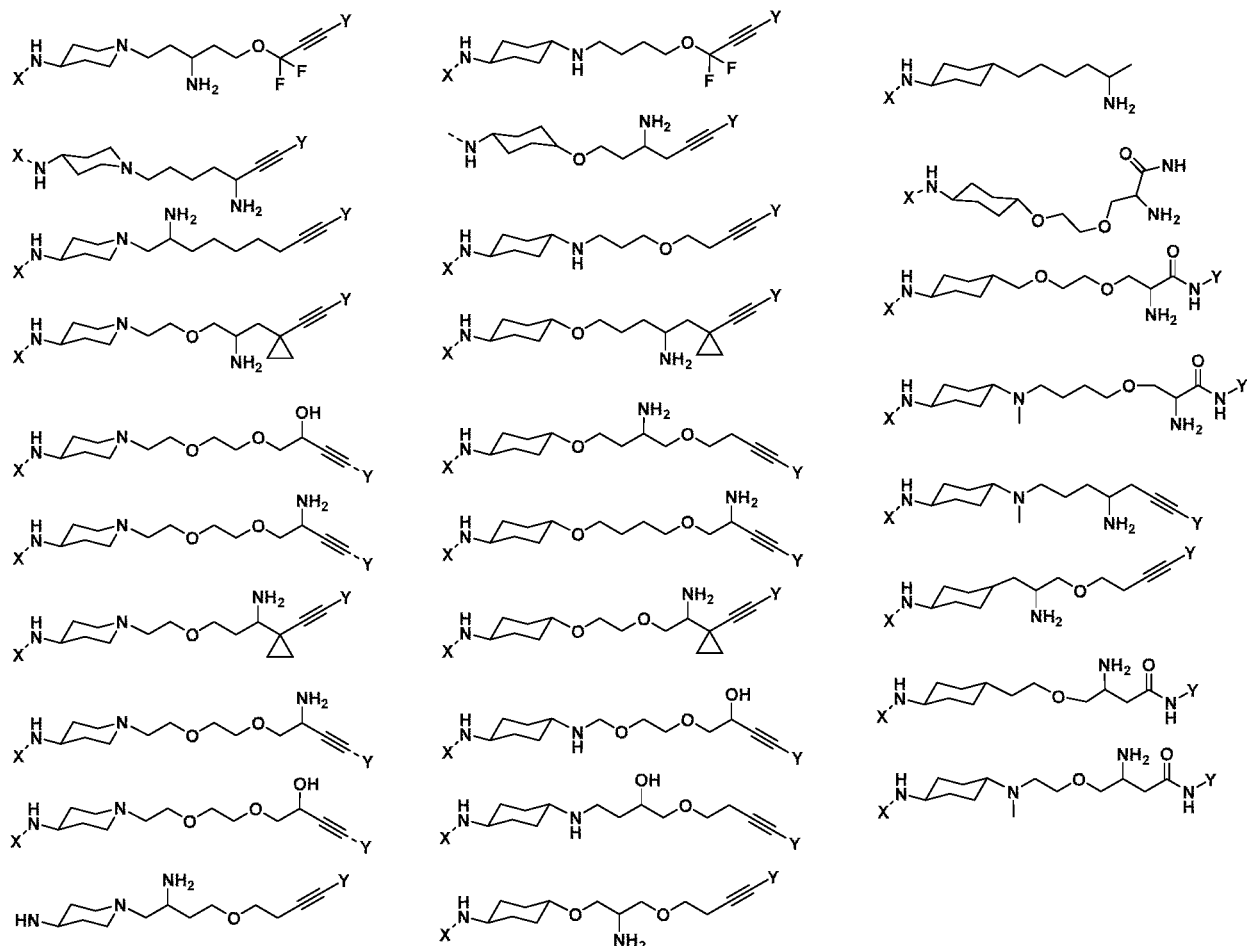




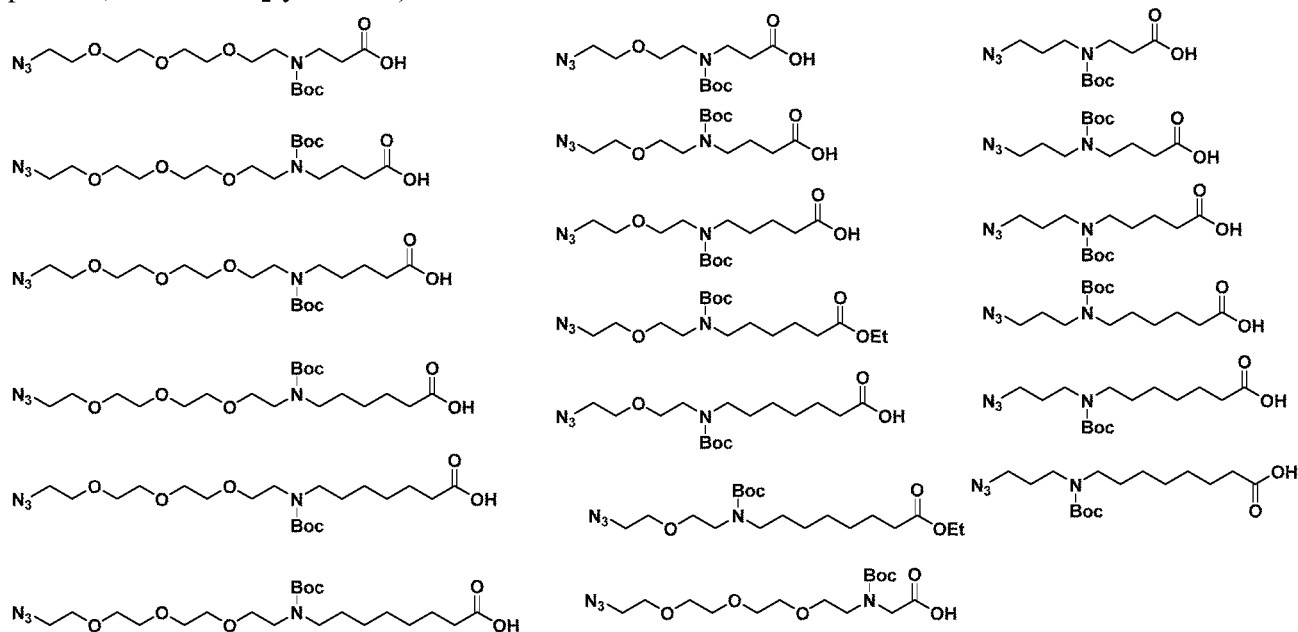


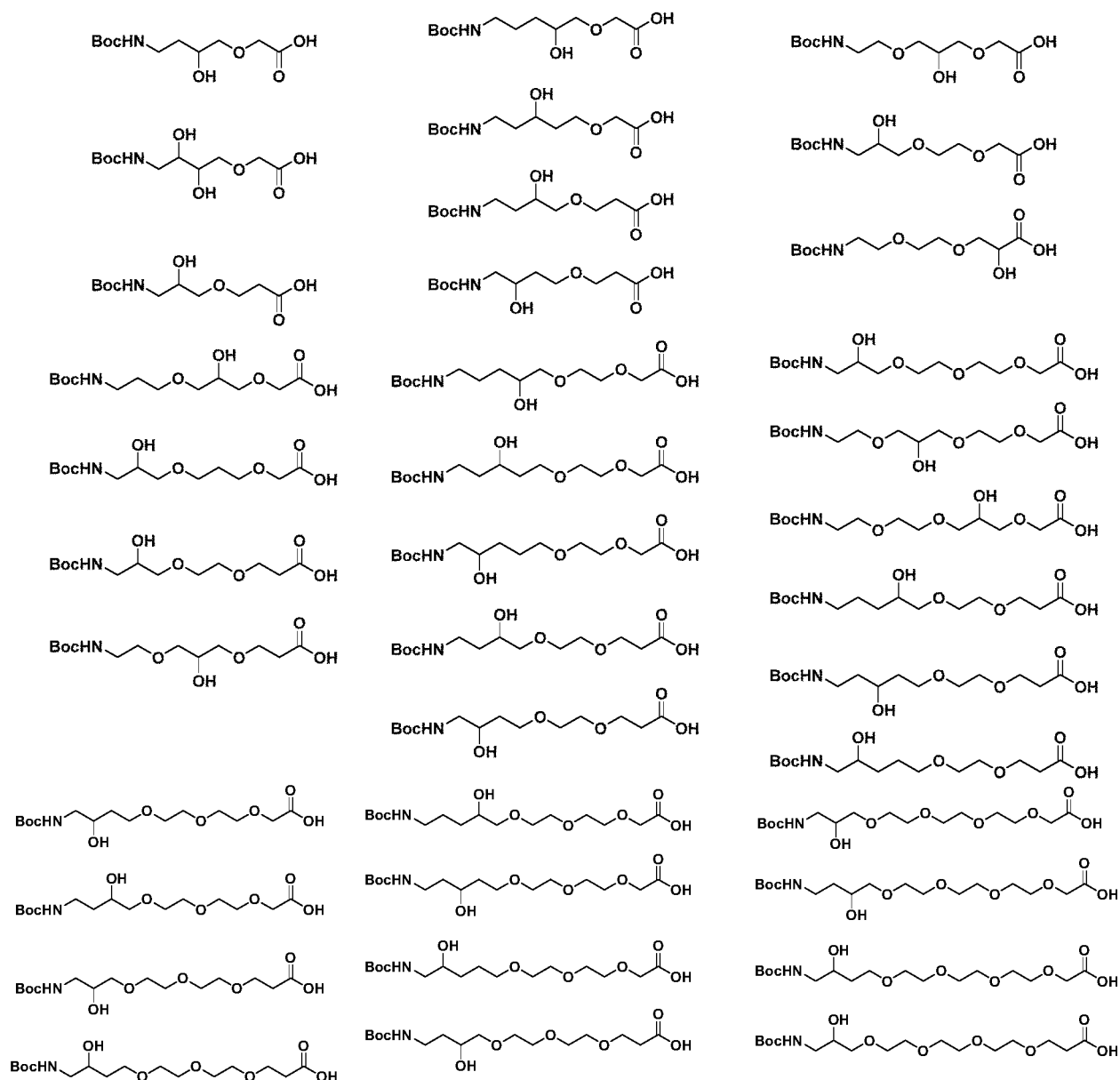
linker moiety thereby forming X'-L3-Z, or directly coupled with Z):





In another preferred embodiment, L_1 is a divalent linker derived from a group selected from the following group of (preferably, by loss two of hydrogen atom, hydroxyl, protecting group (boc) at any position, and/or azide_cyclization):



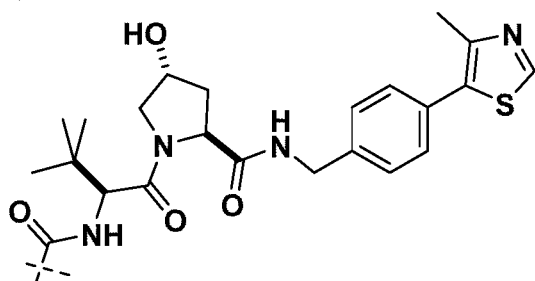


In another preferred embodiment, the target molecule A and T are each independently selected from the group consisted of folic acid, HSP90, TINFRm, TNFR2, NADPH oxidase, BclIBax, C5a receptor, HMG-CoA reductase, PDE I-V, Squalene cyclase inhibitors, CXCR1, CXCR2, Nitric oxide (NO)synthase, cyclo-oxygenase 1-2, 5HT receptors, dopamine receptors, G-proteins, Gq, Histamine receptors, Lipoxigenases, Tryptase serine protease, Thymidylate synthase, Purine nucleotide phosphorylase, GAPDH trypanosomal, Glycogen phosphorylase, Carbonic anhydrase, Chemokine receptors, JAW STAT, RXR and the analogues thereof, HIV 1 protease, HIV 1 integrase, Influenza, hepatitis B reverse transcriptase, neuraminidase, Sodium channel, MDR, protein P-glycoprotein, Tyrosine kinases, CD23, CD124, TK p56 lck, CD4, CD5, IL-1 receptor, IL-2 receptor, TNF- α R, ICAM1, Ca⁺ channels, VCAM, VLA-4 integrin, VLA-4 integrin, Selectins, CD40/40L, Newokinins and receptors, Inosine monophosphate dehydrogenase, p38 MAP kinase, Interleukin-1 converting enzyme, Caspase, HCV NS3 protease, HCV-NS3 RNA helicase, Glycinamide ribonucleotide formyl transferase, rhinovirus 3C protease, HSV-I, CMV, ADP-polymerase, CDK, VEGF, oxytocin receptor, msomalmsomal transfer protein inhibitor, Bile acid transfer protein inhibitor, 5-a reductase, Angiotensin 11, Glycine receptors, noradrenaline reuptake receptor, Endothelin receptors, Neuropeptide Y and receptors, Estrogen receptors, AMP, AMP deaminase, ACC, EGFR, Farnesyltransferase.

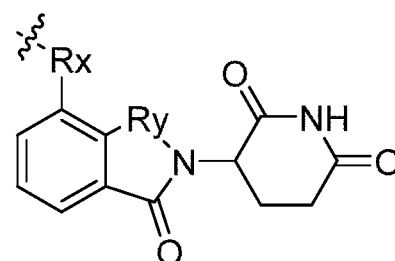
In another preferred embodiment, the polypeptide element is antibody; preferably, the antibody comprises nanobody and/or small molecule antibody (minibody).

In another preferred embodiment, the antibody can bind to an antigen or receptor selected from the group consisting of DLL3, EDAR, CLL1, BMPR1B, E16, STEAP1, 0772P, MPF, 5T4, NaPi2b, Sema 5b, PSCA hlg, ETBR, MSG783, STEAP2, TrpM4, CRIPTO, CD21, CD22, CD79b, CD19, CD37, CD138, FeRH2, B7-H4, HER2, NCA, MDP, IL20R α , Brevican, EphB2R, ASLG659, PSCA, GEDA, BAFF-R, CD79a, CXCR5, HLA-DOB, P2X5, CD72, LY64, FeRH1, IRTA2, TENB2, PMEL17, TMEFF1, GDNF-Ra1, Ly6E, TMEM46, Ly6G6D, LGR5, RET, LY6K, GPR19, GPR54, ASPHD1, Tyrosinase, TMEM118, GPR172A, MUC1, CD70, CD71, MUC16, methothelin, FOLR1, Trop-2, gpNMB, EGFR, ENPP3, PSMA, CA6, GPC-3, PTK7, CD44, CD56, TIM-1, Cadherin-6, ASG-15ME, ASG-22ME, CanAg, AXL, CEACAM5, EphA4, cMet, FGFR2, FGFR3, CD123, Her3, LAMP1, LRRC15, TDGF1, CD66, CD25, BCMA, GCC, Noch3 and CD33.

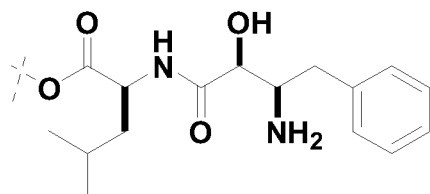
In another preferred embodiment, the E3 ligase ligand is selected from the group consisting of the A¹ groups in WO2017/176957 A1 (such as, A-10, A-11, A-15, A-28, A-48, A-69, A-85, A-93, A-98, A-99 or A-101):



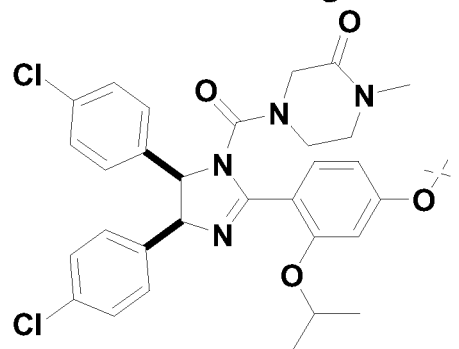
VHL Ligand



CRBN Ligand



cIAP1 Ligand



MDM2 Ligand

in each formula, a dotted line indicates the position connected with other parts (i.e., the position connected with -L1-A);

wherein Rx is selected from the group consisting of none, NH, NH-CO, O, S, SO, SO₂, SO₂(NH₂)NH, C1~C4 alkylene, C2~C5 alkenylene, C2~C5 alkynylene; R_y is C=O, C=S or CH₂.

In another preferred embodiment, the conjugate includes monomer or polymer of the conjugate. Preferably, the polymer of the conjugate shares one or more Z.

In another preferred embodiment, the conjugate of formula I is a conjugate of formula III



wherein, t=1 to 15 (preferably, 1 to 8; more preferably, 2 to 7);

Z is described as above, preferably Z is a polypeptide element, more preferably, is an antibody;

R_a-L3 is the remaining part of the conjugate of formula I after the loss of Z.

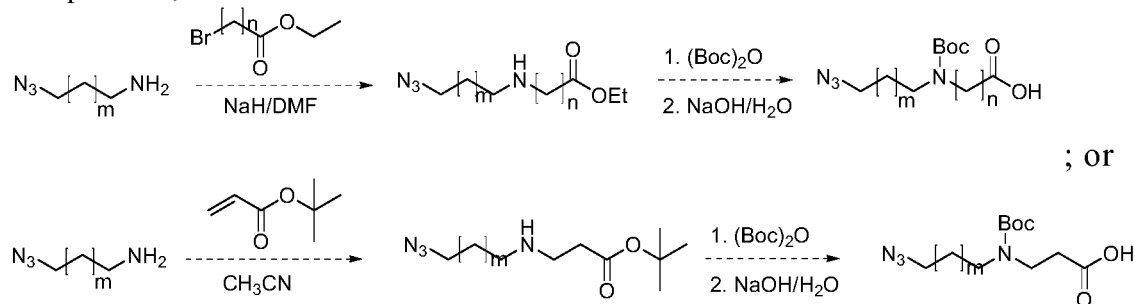
Preferably, R_a is a monovalent group derived from the specific compounds in Tables B1 to B11 (wherein, said derived means a monovalent group formed by the specific compounds shown in Tables B1 to B11 losing a hydrogen from NH or NH₂ on the main chain or the branch chain of the linker group).

In another preferred embodiment, Ab is connected with L3 of formula III (preferably, -NH-, -NHR or -NH₂ group in L3), through amino acid at N-terminal or C-terminal, or a side chain of amino acid (preferably, a side chain of amino acid selected from the group consisting of Lys, and Cys), or a sulfhydryl group formed by reducing and opening disulfide bond.

In another preferred embodiment, the definitions of each group in the conjugate of formula I are the same as the corresponding group of the specific compound in Tables A1 and A2, Table B1 to Table B11, Table C and Table D.

In the second aspect of the present invention, a pharmaceutical composition is provided, wherein the pharmaceutical composition comprises the conjugate described in the first aspect and pharmaceutically acceptable carrier.

In the third aspect of the present invention, a preparation method of the conjugate as described in the first aspect is provided, wherein the reaction route of the method is shown as follows:



wherein m, and n are defined as above.

In the fourth aspect of the present invention, a use of the conjugate as described in the first aspect in the preparation of pharmaceutical compositions for the treatment of diseases associated with excess of the target protein is provided.

In the fifth aspect of the present invention, a method for reducing the content of target proteins in a cell is provided, wherein the cell is contacted with the conjugate as described in the first aspect, thereby reducing the content of the target proteins in the cell.

In another preferred embodiment, the method is a method *in vitro*.

In another preferred embodiment, the method is non-diagnostic and non-therapeutic.

In the sixth aspect of the present invention, a method for reducing the content of target proteins in a subject or treating diseases associated with excess of target proteins is provided, comprising step of administering the conjugate as described in the first aspect to a subject in need thereof.

It should be understood that within the scope of the present invention, the above technical features of the present invention and the technical features specifically described in the following (e.g., examples) can be combined with each other, thereby forming a new or preferred technical solution. Due to space limitations, it will not be repeated herein.

DESCRIPTION OF THE DRAWINGS

Figure 1 is a schematic diagram showing the action principle of degrading the target protein by the Targeted Enzyme Degradation platform (TED).

Figure 2 shows the result of biological test of the conjugate in the examples.

Figure 3 shows the changes of tumor volume of nude mice in animal experiments.

Figure 4 shows the changes of weight of nude mice in animal experiments.

DETAILED DESCRIPTION OF THE INVENTION

After extensively and deeply researching, the inventors developed a TED conjugate with novel structure for the first time, the conjugate of the present invention has a structure of formula I. In addition, the conjugate of the present invention is very suitable for further connected with polypeptide element (especially antibody, protein ligand) or after further connecting with polypeptide element and the like, making the conjugate of the present invention possess excellent dual targeting properties, improve drug selectivity, implement more precise degradation of pathogenic proteins, reduce the possible systemic toxicity induced by non-specific degradation, and is possible to overcome the difficulties encountered in drug absorption and metabolism, and eliminate the possibility for producing drug resistance. The present invention was completed on this basis.

TERMS

As used herein, the term “compound of the present invention”, “conjugate of the present invention”, and “TED conjugate of the present invention” are used interchangeably and refer to the compound or the conjugate of formula I described in the first aspect of the present invention.

As used herein, the term “alkyl”, by itself or as part of another substituent, means, unless otherwise stated, a straight or branched chain hydrocarbon radical, having the number of carbon atoms designated (i.e. C₁₋₈ means 1 - 8 carbons). Examples of alkyl include methyl, ethyl, *n*-propyl, isopropyl, *n*-butyl, *t*-butyl, *iso*-butyl, *sec*-butyl, *n*-pentyl, *n*-hexyl, *n*-heptyl, *n*-octyl, and the like. The term “alkenyl” refers to an unsaturated alkyl group having one or more double bonds. Similarly, the term “alkynyl” refers to an unsaturated alkyl having one or more triple bonds. Examples of such unsaturated alkyl include vinyl, 2-propenyl, crotyl, 2-isopentenyl, 2-(butadienyl), 2,4-pentadienyl, 3-(1,4-pentadienyl), ethynyl, 1- and 3-propynyl, 3-butylnyl, and the higher homologs and isomers. The term “cycloalkyl” refers to hydrocarbon ring having the indicated number of ring atoms (e.g., C₃₋₆ cycloalkyl) and being fully saturated or having no more than one double bond between ring vertices. “Cycloalkyl” is also meant to refer to bicyclic and polycyclic hydrocarbon rings such as, bicyclo[2.2.1]heptane, bicyclo[2.2.2]octane, etc. The term “heterocycloalkyl” refers to a cycloalkyl that contain from one to five heteroatoms selected from N, O, and S, wherein the nitrogen and sulfur atoms are optionally oxidized, and the nitrogen atom(s) are optionally quaternized. The heterocycloalkyl may be a monocyclic, a bicyclic or a polycyclic ring system. Non limiting examples of heterocycloalkyl include pyrrolidine, imidazolidine, pyrazolidine, butyrolactam, valerolactam, imidazolidinone, hydantoin, dioxolane, phthalimide, piperidine, 1,4-dioxane, morpholine, thiomorpholine, thiomorpholine-S-oxide, thiomorpholine-S,S-oxide, piperazine, pyran, pyridone, 3-pyrroline, thiopyran, pyrone, tetrahydrofuran, tetrahydrothiophene, quinuclidine, and the like. The heterocycloalkyl can be attached to the rest of the molecule via a ring carbon or a heteroatom. For terms such as cycloalkylalkyl and heterocycloalkylalkyl, it is meant that a cycloalkyl or a heterocycloalkyl is attached through an alkyl or

alkylene linker to the remainder of the molecule. For example, cyclobutylmethyl – is a cyclobutyl ring that is attached to a methylene linker to the remainder of the molecule.

The term “alkylene” by itself or as part of another substituent means a divalent radical derived from an alkane, for example $-\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2-$. Typically, an alkyl (or alkylene) will have from 1 to 24 carbon atoms, with those groups having 10 or fewer carbon atoms being preferred in the present disclosure. A “lower alkyl” or “lower alkylene” is a shorter chain alkyl or alkylene, generally having four or fewer carbon atoms. Similarly, “alkenylene” and “alkynylene” refer to the unsaturated forms of “alkylene” having double or triple bond, respectively.

Unless otherwise specified, the term “heteroalkyl” by itself or in combination with other terms refers to a stable linear or branched or cyclic hydrocarbon group or a combination thereof, consisting of a specified number of carbon atoms and 1 to 3 heteroatoms selected from O, N, Si and S, and wherein nitrogen and sulfur atoms are optionally oxidized, and nitrogen heteroatoms can be optionally quaternized. The heteroatoms O, N and S can be placed at any internal position of the heteroalkyl. The heteroatom Si can be placed at any position of the heteroalkyl, including the position at which the alkyl is attached to the remainder of the molecule. Examples include $-\text{CH}_2\text{CH}_2\text{OCH}_3$, $-\text{CH}_2\text{CH}_2\text{NHCH}_3$, $-\text{CH}_2\text{CH}_2\text{N}(\text{CH}_3)\text{CH}_3$, $-\text{CH}_2\text{SCH}_2\text{CH}_3$, $-\text{CH}_2\text{CH}_2\text{S}(\text{O})\text{CH}_3$, $-\text{CH}_2\text{CH}_2\text{S}(\text{O})_2\text{CH}_3$, $-\text{CH}=\text{CHOCH}_3$, $-\text{Si}(\text{CH}_3)_3$, $-\text{CH}_2\text{CH}=\text{NOCH}_3$, and $-\text{CH}=\text{CHN}(\text{CH}_3)\text{CH}_3$. Up to two heteroatoms may be consecutive, such as, $-\text{CH}_2\text{NHCH}_3$ and $-\text{CH}_2\text{OSi}(\text{CH}_3)_3$. Similarly, unless otherwise specified, terms “heteroalkenyl” and “heteroalkynyl” by themselves or in combination with another term refer to alkenyl or alkynyl, respectively, that contain the stated number of carbons and 1 to 3 heteroatoms selected from O, N, Si and S, and wherein the nitrogen and sulfur atoms may optionally be oxidized and the nitrogen heteroatom may optionally be quaternized. The heteroatoms O, N and S can be placed at any internal position of the heteroalkyl.

The term “heteroalkylene” by itself or as part of another substituent means a divalent group, saturated or unsaturated or polyunsaturated, derived from heteroalkyl, as exemplified by $-\text{CH}_2\text{CH}_2\text{SCH}_2\text{CH}_2-$ and $-\text{CH}_2\text{SCH}_2\text{CH}_2\text{NHCH}_2-$, $-\text{OCH}_2\text{CH}=\text{CH}-$, $-\text{CH}_2\text{CH}=\text{C}(\text{H})\text{CH}_2\text{OCH}_2-$ and $-\text{SCH}_2\text{C}\equiv\text{C}-$. For heteroalkylene, heteroatoms can also occupy either or both of the chain termini (e.g., alkyleneoxy, alkylenedioxy, alkyleneamino, alkylenediamino, and the like).

The terms “alkoxy”, “alkylamino” and “alkylthio” (or thioalkoxy) are used in their conventional meanings, refer to those alkyl attached to the rest of the molecule via oxygen atom, amino, or sulfur atom, respectively. Additionally, for dialkylamino, the alkyl portions can be the same or different and can also be combined to form a 3-7 membered ring with the nitrogen atom to which each is attached. Accordingly, a group represented as $-\text{NR}^a\text{R}^b$ is meant to include piperidinyl, pyrrolidinyl, morpholinyl, azetidiny and the like.

The terms “halo” or “halogen,” by itself or as part of another substituent, mean, unless otherwise stated, fluorine, chlorine, bromine, or iodine atom. Additionally, terms such as “haloalkyl,” is meant to include monohaloalkyl or polyhaloalkyl. For example, the term “ C_{1-4} haloalkyl” is meant to include trifluoromethyl, 2,2,2-trifluoroethyl, 4-chlorobutyl, 3-bromopropyl, and the like.

The term “aryl” means, unless otherwise stated, polyunsaturated, typically aromatic, hydrocarbon group which can be a single ring or multiple rings (up to three rings) which are fused together or linked covalently. The term “heteroaryl” refers to aryl (or ring) that contain from one to five heteroatoms selected from N, O, and S, wherein the nitrogen and sulfur atoms are optionally oxidized, and the nitrogen atom(s) are optionally quaternized. A heteroaryl can be attached to the remainder of the molecule through a heteroatom. Non-limiting examples of aryl include phenyl, naphthyl and biphenyl, while non-limiting

examples of heteroaryl include pyridyl, pyridazinyl, pyrazinyl, pyrimidinyl, triazinyl, quinolinyl, quinoxaliny, quinazoliny, cinnoliny, phthalazinyl, benzotriazinyl, purinyl, benzimidazolyl, benzopyrazolyl, benzotriazolyl, benzisoxazolyl, isobenzofuryl, isoindolyl, indoliziny, benzotriazinyl, thienopyridinyl, thienopyrimidinyl, pyrazolopyrimidinyl, imidazopyridines, benzothiazolyl, benzofuranyl, benzothieryl, indolyl, quinolyl, isoquinolyl, isothiazolyl, pyrazolyl, indazolyl, pteridinyl, imidazolyl, triazolyl, tetrazolyl, oxazolyl, isoxazolyl, thiadiazolyl, pyrrolyl, thiazolyl, furyl, thienyl and the like. Substituents for above-stated aryl and heteroaryl ring systems are selected from the group of acceptable substituents described below.

For brevity, the term “aryl” when used in combination with other terms (e.g., aryloxy, arylthioxy, arylalkyl) includes both aryl and heteroaryl rings as defined above. Thus, the term “arylalkyl” is meant to include those groups in which an aryl is attached to an alkyl that is attached to the remainder of the molecule (e.g., benzyl, phenethyl, pyridylmethyl and the like).

The above terms (e.g., “alkyl,” “aryl” and “heteroaryl”), in some embodiments, will include both substituted and unsubstituted forms of the indicated group. The preferred substituents for each type of group are provided below. For brevity, the terms aryl and heteroaryl will refer to substituted or unsubstituted versions as provided below, while the term “alkyl” and related aliphatic groups are meant to refer to unsubstituted form, unless indicated to be substituted.

Substituents for the alkyl (including those groups often referred to as alkylene, alkenyl, alkynyl and cycloalkyl) can be a variety of groups selected from: -halogen, -OR', -NR'R'', -SR', -SiR'R''R''', -OC(O)R', -C(O)R', -CO₂R', -CONR'R'', -OC(O)NR'R'', -NR''C(O)R', -NR'-C(O)NR''R''', -NR''C(O)₂R', -NH-C(NH₂)=NH, -NR'C(NH₂)=NH, -NH-C(NH₂)=NR', -S(O)R', -S(O)₂R', -S(O)₂NR'R'', -NR'S(O)₂R'', -CN and -NO₂ in a number ranging from zero to (2m'+1), wherein m' is the total number of carbon atoms in such radical. R', R'' and R''' each independently refers to hydrogen, unsubstituted C₁₋₈ alkyl, unsubstituted heteroalkyl, unsubstituted aryl, aryl substituted with 1-3 halogens, unsubstituted C₁₋₈ alkyl, C₁₋₈ alkoxy or C₁₋₈ thioalkoxy, or unsubstituted aryl-C₁₋₄ alkyl. When R' and R'' are attached to the same nitrogen atom, they can be combined with the nitrogen atom to form a 3-, 4-, 5-, 6-, or 7-membered ring. For example, -NR'R'' is meant to include 1-pyrrolidinyl and 4-morpholinyl. The term “acyl” as used by itself or as part of another group refers to groups wherein two substituents on the carbon that is closest to the point of attachment for the radical is replaced with the substituent =O (e.g., C(O)CH₃, -C(O)CH₂CH₂OR' and the like).

Similarly, substituents for the aryl and heteroaryl are varied and are generally selected from: -halogen, -OR', -OC(O)R', -NR'R'', -SR', -R', -CN, -NO₂, -CO₂R', -CONR'R'', -C(O)R', -OC(O)NR'R'', -NR''C(O)R', -NR''C(O)₂R', -NR'-C(O)NR''R''', -NH-C(NH₂)=NH, -NR'C(NH₂)=NH, -NH-C(NH₂)=NR', -S(O)R', -S(O)₂R', -S(O)₂NR'R'', -NR'S(O)₂R'', -N₃, perfluoro(C₁₋₄)alkoxy, and perfluoro(C₁₋₄)alkyl, in a number ranging from zero to the total number of open valences on the aromatic ring system; and wherein R', R'' and R''' are independently selected from hydrogen, C₁₋₈ alkyl, C₃₋₆ cycloalkyl, C₂₋₈ alkenyl, C₂₋₈ alkynyl, unsubstituted aryl and heteroaryl, (unsubstituted aryl)-C₁₋₄ alkyl, and unsubstituted aryloxy-C₁₋₄ alkyl. Other suitable substituents include each of the above aryl substituents attached to a ring atom by an alkylene tether of from 1-4 carbon atoms.

Two of the substituents on adjacent atoms of the aryl or heteroaryl ring may optionally be replaced with a substituent of the formula -T-C(O)-(CH₂)_q-U-, wherein T and U are independently -NH-, -O-, -CH₂- or a single bond, and q is an integer of from 0 to 2. Alternatively, two of the substituents on adjacent atoms of the aryl or heteroaryl ring may optionally be replaced with formula -A-(CH₂)_r-B-, wherein A and B are independently -CH₂-, -O-, -NH-, -S-, -S(O)-, -S(O)₂-, -S(O)₂NR'- or a single bond, and r is an integer of

from 1 to 3. One of the single bonds of the new ring so formed may optionally be replaced with a double bond. Alternatively, two of the substituents on adjacent atoms of the aryl or heteroaryl ring may optionally be replaced with a substituent of the formula $-(CH_2)_s-X-(CH_2)_t-$, where s and t are independently integers of from 0 to 3, and X is -O-, -NR'-, -S-, -S(O)-, -S(O)₂- or -S(O)₂NR'-. The substituent R' in -NR'- and -S(O)₂NR- is selected from hydrogen or unsubstituted C₁₋₆ alkyl.

In the present invention, when one chain atom (such as a C atom) is replaced with a heterocyclic ring, the heterocyclic ring may lose two hydrogens on the same ring atom (on ring carbon atom) thereby connect with other chain atoms on the chain (forming a structure similar to a spirocyclic ring), or may lose two hydrogens on different ring atoms thereby connect with other chain atoms on the chain (such as -cyclopentylidene-).

As used herein, the term "heteroatom" is meant to include oxygen (O), nitrogen (N), sulfur (S) and silicon (Si).

For the compounds provided herein, a bond that is drawn from a substituent (typically an R group) to the center of an aromatic ring (e.g., benzene, pyridine, and the like) will be understood to refer to a bond providing a connection at any of the available vertices of the aromatic ring. In some embodiments, the depiction will also include connection at a ring which is fused to the aromatic ring. For example, a bond drawn to the center of the benzene portion of an indole, will indicate a bond to any available vertex of the six- or five-membered ring portions of the indole.

The term "pharmaceutically acceptable salts" is meant to include salts of the active compounds which are prepared with relatively nontoxic acids or bases, depending on the particular substituents found on the compounds described herein. When compounds of the present disclosure contain relatively acidic functionalities, base addition salts can be obtained by contacting the neutral form of such compounds with a sufficient amount of the desired base, either neat or in a suitable inert solvent. Examples of salts derived from pharmaceutically acceptable inorganic bases include aluminum, ammonium, calcium, copper, iron, ferrous, lithium, magnesium, manganese, manganous, potassium, sodium, zinc, and the like. Salts derived from pharmaceutically-acceptable organic bases include salts of primary, secondary and tertiary amines, including substituted amines, cyclic amines, naturally-occurring amines and the like, such as arginine, betaine, caffeine, choline, N,N'-dibenzylethylenediamine, diethylamine, 2-diethylaminoethanol, 2-dimethylaminoethanol, ethanolamine, ethylenediamine, N-ethylmorpholine, N-ethylpiperidine, glucamine, glucosamine, histidine, hydrabamine, isopropylamine, lysine, methylglucamine, morpholine, piperazine, piperidine, polyamine resins, procaine, purines, theobromine, triethylamine, trimethylamine, tripropylamine, tromethamine and the like. When compounds of the present disclosure contain relatively basic functionalities, acid addition salts can be obtained by contacting the neutral form of such compounds with a sufficient amount of the desired acid, either neat or in a suitable inert solvent. Examples of pharmaceutically acceptable acid addition salts include those derived from inorganic acids like hydrochloric, hydrobromic, nitric, carbonic, monohydrogencarbonic, phosphoric, monohydrogenphosphoric, dihydrogenphosphoric, sulfuric, monohydrogensulfuric, hydriodic, or phosphorous acids and the like, as well as the salts derived from relatively nontoxic organic acids like acetic, propionic, isobutyric, malonic, benzoic, succinic, suberic, fumaric, mandelic, phthalic, benzenesulfonic, *p*-tolylsulfonic, citric, tartaric, methanesulfonic, and the like. Also included are salts of amino acids such as arginate and the like, and salts of organic acids like glucuronic or galactunoric acids and the like (see, for example, Berge, S.M., et al, "Pharmaceutical Salts", Journal of Pharmaceutical Science, 1977, 66, 1-19). Certain specific compounds of the present disclosure contain both basic and acidic functionalities that allow the compounds to be converted into either base or acid addition salts.

The neutral forms of the compounds may be regenerated by contacting the salt with a base or acid and isolating the parent compound in the conventional manner. The parent form of the compound differs from the various salt forms thereof in certain physical properties, such as solubility in polar solvents, but in addition to the above, those salts are equivalent to the parent form of the compound for the purposes of the present invention.

In addition to salt forms, the present disclosure provides compounds which are in a prodrug form. Prodrugs of the compounds described herein are those compounds that readily undergo chemical changes under physiological conditions to provide the compounds of the present disclosure. Additionally, prodrugs can be converted to the compounds of the present disclosure by chemical or biochemical methods in an *ex vivo* environment. For example, when placed in a transdermal patch reservoir containing suitable enzymes or chemical reagents, the prodrug can be slowly converted to the compound of the invention.

Certain compounds of the present disclosure can exist in unsolvated forms as well as solvated forms, including hydrated forms. The solvated forms are generally equivalent to the non-solvated forms and should be included in the scope of the present invention. Certain compounds of the present disclosure may exist in multiple crystalline or amorphous forms. Generally, as for the application considered in the present invention, all physical forms are equivalent and should be included in the scope of the present invention.

Certain compounds of the present disclosure possess asymmetric carbon atoms (optical centers) or double bonds; the racemates, diastereomers, geometric isomers, regioisomers and individual isomers (e.g., separate enantiomers) are all intended to be encompassed within the scope of the present disclosure. When compounds are provided herein with an identified stereochemistry (indicated as R or S, or with dashed or wedge bond designations), those compounds will be understood by one of skill in the art to be substantially free of other isomers (e.g., at least 80%, 90%, 95%, 98%, 99%, and up to 100% free of the other isomer).

The compounds of the present disclosure may also contain unnatural proportions of isotope atomic isotopes at one or more of isotopic atoms that constitute such compounds. The unnatural proportions of certain isotope can be defined as the amount from the naturally found amount of the atom discussed to 100% of that atom. For example, the compounds may incorporate radioactive isotopes, such as tritium (^3H), iodine-125 (^{125}I) or carbon-14 (^{14}C), or non-radioactive isotopes, such as deuterium (^2H) or carbon-13 (^{13}C). Such isotopic variants may provide additional uses in addition to those described in this application. For instance, isotopic variants of the compounds of the disclosure may find additional utility, including but not limited to, as diagnostic and/or imaging reagents, or as cytotoxic/radiotoxic therapeutic agents. Additionally, isotopic variants of the compounds of the disclosure can have altered pharmacokinetic and pharmacodynamic characteristics which can contribute to enhanced safety, tolerability or efficacy during treatment. All isotopic variations of the compounds of the present disclosure, whether radioactive or not, should be encompassed within the scope of the present disclosure.

Targeted Enzyme Degradation platform TED

The present invention provides a Targeted Enzyme Degradation platform (TED) on basis of the conjugate of the present invention, which utilizes the intracellular “cleaner” - ubiquitin proteasome system.

Typically, according to the TED technology of the present invention, which can utilize cell's intrinsic protein destruction mechanism to remove specific oncogenic and pathogenic proteins from the cell, therefore it is an alternative method of targeted therapy.

Different from action mechanism of conventional protein inhibitors, TED technology of the present invention relates to a bifunctional hybrid compound, one side of which is used to bind target proteins, and another side is used to bind E3 ligases, enabling the target proteins binding the E3 ligases, and the target

proteins being ubiquitinated, thereby being degraded by the proteome. Theoretically, TED technology only provides binding activity without functional activity that directly inhibiting the target protein, and can be reused. Therefore, TED technology has excellent application prospects.

5 **Polypeptide element**

As used herein, the term "polypeptide element" includes peptide fragment (such as oligopeptide comprising 3 - 20 aa) or protein. In addition, this term also includes intact protein or fragment thereof. Preferred polypeptide element includes antibody (such as intact antibody, single-chain antibody, nanobody, Fab), especially those antibodies against tumor cell markers (such as tumor markers located on the surface of tumor cells, such as receptors on the cell surface) or inflammatory factors (such as inflammatory factors associated with autoimmune diseases).

As used herein, term "antibody" or "immunoglobulin" is a heterotetrameric glycoprotein of about 150,000 daltons with the same structural characteristics, which consists of two identical light chains (L) and two identical heavy chains (H). Each light chain is connected to the heavy chain by a covalent disulfide bond, and the number of disulfide bonds between the heavy chains of different immunoglobulin isotypes are different. Each heavy and light chain also has regularly spaced intrachain disulfide bonds. Each heavy chain has a variable region (VH) at one end, followed by multiple constant regions. There are a variable region (VL) at one end of each light chain and a constant region at the other end. The constant region of the light chain is relative to the first constant region of the heavy chain, and the variable region of the light chain is relative to the variable region of the heavy chain. Special amino acid residues form an interface between the variable regions of the light chain and the heavy chain.

As used herein, terms "single-domain antibody" and "nanobody" have the same meaning, and refer to cloneing the variable region of the heavy chain of an antibody, and constructing a single-domain antibody consisting of only one heavy chain variable region, which is the smallest antigen-binding fragment that having complete functions. Usually, after obtaining an antibody naturally missing constant region 1 (CH1) of light chain and heavy chain, the variable region of the antibody heavy chain is cloned to construct a single domain antibody consisting of only one heavy chain variable region.

As used herein, term "variable" means that certain parts of the variable region of the antibody are different in sequence, which forms the binding and specificity to specific antigens of various specific antibodies. However, variabilities are not evenly distributed throughout the variable regions of antibodies. It is concentrated in three fragments that are called complementarity determining regions (CDR) or hypervariable regions in the variable regions of light chain and heavy chain. More conservative parts of the variable region are called the framework region (FR). The variable regions of the natural heavy chain and light chain each contain four FR regions, which are roughly in a β -folded configuration, connected by three CDRs forming a connecting loop, and in some cases can form a partially β -folded structure. The CDRs in each chain are closely placed together through the FR region and form the antigen binding site of the antibody together with the CDRs of the other chain (see Kabat et al., NIH Publ. No. 91-3242, Volume I, pages 647-669 (1991)). Constant regions do not directly participate in the binding of antibodies to antigens, but they exhibit different effector functions, such as participating in antibody-dependent cytotoxicity of antibodies.

The light chains of vertebrate antibodies (immunoglobulins) can be classified into one of two distinct categories (referred to κ and λ) based on the amino acid sequence of their constant regions. According to the amino acid sequence of the constant region of their heavy chains, immunoglobulins can be divided into different types. There are five main classes of immunoglobulins: IgA, IgD, IgE, IgG and IgM, some of which can be further divided into subclasses (isotypes), such as IgG1, IgG2, IgG3, IgG4, IgA and IgA2. The heavy chain constant regions corresponding to different classes of immunoglobulins are called α , δ , ϵ , γ

and μ respectively. The subunit structures and three-dimensional configurations of different classes of immunoglobulins are well known to those skilled in the art.

Generally, the antigen-binding properties of antibodies can be described by 3 specific regions located in the variable regions of the heavy and light chains, called variable regions (CDR), which are divided into 4 framework regions (FR). The amino acid sequence of 4 FR is relatively conservative and does not directly participate in the binding reaction. These CDRs form a circular structure, and the β -folded configuration formed by the FRs in between are close to each other in space structure, and the CDRs on the heavy chain and the corresponding CDRs on the light chain constitute the antigen binding site of the antibody. The amino acid sequences of antibodies of the same type can be compared to determine which amino acids constitute the FR or CDR regions.

In the present invention, the polypeptide element can include not only intact antibody, but also fragment of antibody with immunological activity (such as Fab or (Fab')₂ fragment; heavy chain of antibody; or light chain of antibody) or fusion protein formed by antibody and other sequences. Therefore, the present invention also includes fragment, derivative and analog of the antibody.

Targeting ligand

Targeting ligand (or target protein portion or target protein ligand or ligand) is a small molecule that can bind to a target protein of interest.

Some embodiments of this application relate to target molecules, and representative target molecules include but are not limited to: folic acid, Hsp90 inhibitors, kinase inhibitors, MDM2 inhibitors, compounds targeting proteins containing human BET bromodomain, compounds targeting cytoplasmic signaling protein FKBP12, HDAC inhibitors, human lysine methyltransferase inhibitors, angiogenesis inhibitors, immunosuppressive compounds and compounds targeting aryl hydrocarbon receptor (AHR).

In certain embodiments, the targeting ligand is capable of binding kinases, BET bromodomain-containing proteins, cytoplasmic signaling proteins (such as FKBP12), nucleoproteins, histone deacetylases, lysine methyltransferase, protein that regulates angiogenesis, protein that regulates immune response, aromatic hydrocarbon receptor (AHR), estrogen receptor, androgen receptor, glucocorticoid receptor, or transcription factor (e.g., SMARCA4, SMARCA2, TRIM24).

In certain embodiments, kinases which targeting ligands are able to bind to, include, but not limited to: tyrosine kinases (for example, AATK, ABL, ABL2, ALK, AXL, BLK, BMX, BTK, CSF1R, CSK, DDR1, DDR2, EGFR, EPHA1, EPHA2, EPHA3, EPHA4, EPHA5, EPHA6, EPHA7, EPHA8, EPHA10, EPHB1, EPHB2, EPHB3, EPHB4, EPHB6, ERBB2, ERBB3, ERBB4, FER, FES, FGFR1, FGFR2, FGFR3, FGFR4, FGR, FLT1, FLT3, FLT4, FRK, FYN, GSG2, HCK, HRAS, HSP90, IGF1R, ILK, INSR, INSR, IRAK4, ITK, JAK1, JAK2, JAK3, KDR, KIT, KRAS, KSP, KSR1, LCK, LMTK2, LMTK3, LTK, LYN, MATK, MERTK, MET, MLTK, MST1R, MUSK, NPR1, NRAS, NTRK1, NTRK2, NTRK3, PDGFRA, PDGFRB, PLK4, PTK2, PTK2B, PTK6, PTK7, RET, ROR1, ROR2, ROS1, RYK, SGK493, SRC, SRMS, STYK1, SYK, TEC, TEK, TEX14, TIE1, TNK1, TNK2, TNKI3K, TXK, TYK2, TYRO3, YES1 or ZAP70), serine/threonine kinase (such as Casein Kinase 2, protein kinase A, protein kinase B, protein kinase C, Raf kinase, CaM kinase, AKT1, AKT2, AKT3, ALK1, ALK2, ALK3, ALK4, Aurora A, Aurora B, Aurora C, CHK1, CHK2, CLK1, CLK2, CLK3, DAPK1, DAPK2, DAPK3, DMPK, ERK1, ERK2, ERK5, GSK, GSK3, HIPK, KHS1, LKB1, LOK, MAPKAPK2, MAPKAPK, MEK, MNK1, MSSK1, MST1, MST2, MST4, NDR, NEK2, NEK3, NEK6, NEK7, NEK9, NEK11, PAK1, PAK2, PAK3, PAK4, PAK5, PAK6, PIM1, PIM2, PLK1, RIP2, RIP5, RSK1, RSK2, SGK2, SGK3, SIK1, STK33, TAO1, TAO2, TGF- β , TLK2, TSSK1, TSSK2, MLK1 or MLK2), cyclin-dependent kinases (such as Cdk1-Cdk11) and Leucine-rich repetitive kinase (such as LRRK2).

Target molecule

The conjugate of formula I in the present application, binds to the target protein through target molecule.

In the present invention, the target molecule can be a target molecule A, a target molecule T, or the combination thereof.

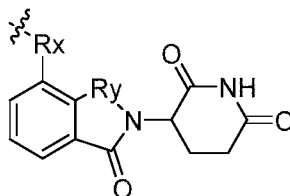
In the present invention, the target molecule can be any inhibitor of the target protein. The target molecule can be a highly effective inhibitor of the target protein, or an inhibitor with relatively poor activity. Specifically, the target molecule of the present invention may be a small molecule inhibitor known in the art against any target protein in the art.

In some embodiments, the target molecule used herein has a radical, such as -O-, -NR_a- (wherein R_a is H, or substituents of C1-C6 alkyl etc., -CO-, -COO-, and the like, that is able to connect to a linker (e.g. L1 in the present invention) monovalently to form an ether, an amine, an amide and the like.

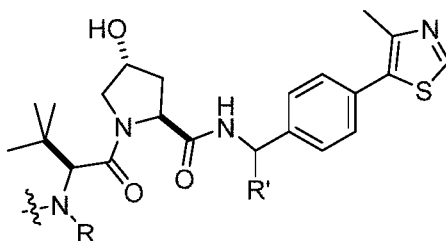
The target protein may be a variety of target proteins known in the art, representative examples include, but are not limited to:MDM2, AKT, BCR-ABL, Tau, BET(BRD2, BRD3, BRD4), ERR α , FKBP12, RIPK2, ERBB3, androgen receptor, MetAP2, TACC3, FRS2 α , PI3K, DHFR, GST, Halo Tag, CRABPI, CRABPII, RAR, aromatic hydrocarbon receptor, strogen receptor. Different target proteins and some corresponding inhibitors can be obtained commercially or prepared by conventional methods. For example, as for MDM2, the inhibitors thereof can be referred to documents such as WO 2017176957, and WO2017176958A1.

E3 Ligase Ligand

In the present invention, the E3 ligase ligand is used for binding E3 ligase. Representative E3 ligase ligands have structures shown as below:



in the above formula, R_X is none, C1-C6 alkyl, C2-C6 alkenyl, C2-C6 alkynyl, O, NH, S, CO or SO_n (n is 1 or 2) and the like; R_Y is CH₂, C=S, CO; and the E3 ligase ligand (B in formula I) is able to connect to L1 of the present invention via R_X group in the E3 ligase ligand, for example -R_X-L1-A (such as -O-L1-A),



in the above formula, R' is H or Me, and R is H, Me or Et.

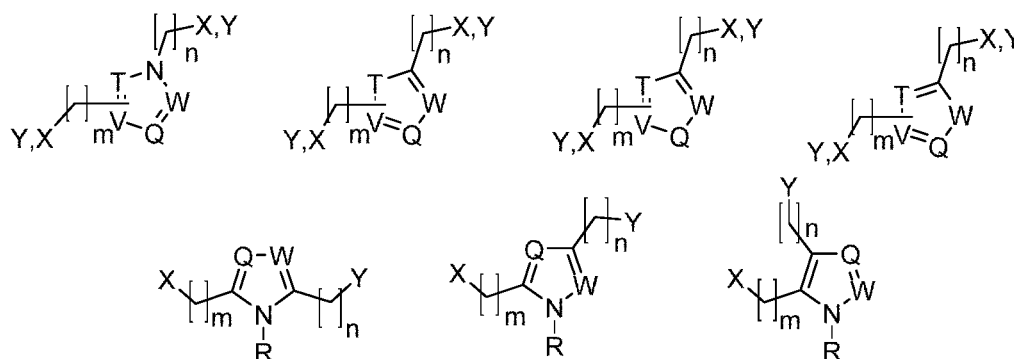
In some embodiments, the E3 ligase ligand used herein has a radical, such as -O-, -NR_a- (wherein R_a is H, or substituents of C1-C6 alkyl etc., -CO-, -COO-, and the like, that is able to connect to a linker (e.g. L1 in the present invention and the like) monovalently to form an ether, an amine, an amide and the like.

Linker molecule (L1 as described herein)

The linker molecule of the present invention is used for connecting the target molecule and the E3

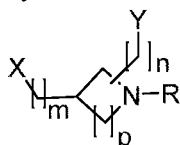
ligase ligand. For example, it can be connected to the target molecule or the E3 ligase ligand through functional groups at both ends (such as -OH, -SH, -NH₂, -NHR, -SOOH or -COOH) ; wherein R is selected from: substituted or unsubstituted C1-C10 alkyl, -(C=O)-R', (C=O)NH-R', -NH(C=O)-R', -SO₂-R', -NH-SO₂-R', -SO₂NH-R', -SO-R', -NH-SO-R', -SONH-R', -PO₃-R', -NHCOO-R', -COO-R' or -NH-CO-NH-R', -NH-CO-O-R' or -X'-L3-Z; where L3 is linker, and Z is polypeptide element (such as ligand, antibody or peptide fragment, etc.) or a targeting molecule such as targeting function small molecule (such as folic acid, HSP90 inhibitor, etc.).

Preferably, when the linker is -(CH₂)_s-; wherein, when 1 to 4 -CH₂- are replaced with five-membered unsaturated ring containing 1 to 4 heteroatoms (such as N, O, S, and P), the linker molecule is show as below:



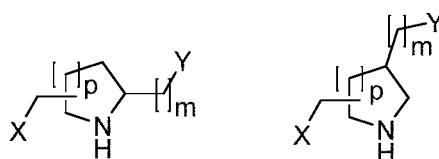
in the formula, X, Y, T, V, Q, W, m, n, s, R are defined as above.

Preferably, when the linker is -(CH₂)_s-; wherein, when 1 to 4 -CH₂- are replaced with O or the 4-6 membered saturated ring containing one nitrogen atom, the linker molecule of the present invention specifically can have the structure selected from the following:



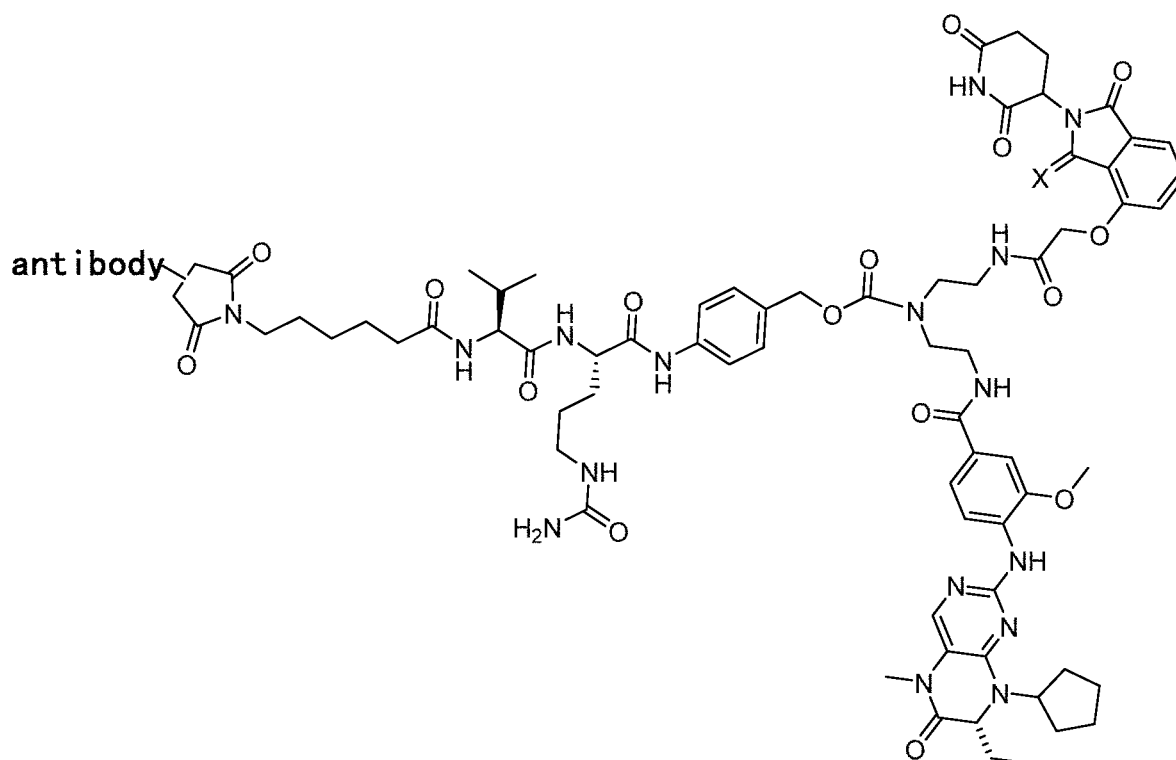
in the formula, X, Y, p, m, n, s, and R are defined as above.

Preferably, the linker molecule of the present invention can have a structure selected from the following:



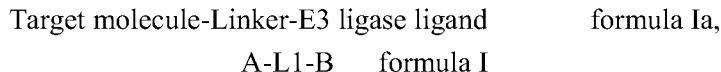
in the formula, X and Y are defined as above, m is an integer of 1-20; p is an integer of 1-3.

Preferably, when the linker is -(CH₂)_s- or the linker is -(CH₂)_s-; wherein 1 to 4 -CH₂- are replaced with O. Preferably, the linker molecule of the present invention can further connect with the target molecule or polypeptide element through the linker structure of -X'-L3-, forming a trifunctional Targeted Enzyme Degradation platform (TED) molecule, for example, the Targeted Enzyme Degradation platform (TED) molecules having the following formula:



Targeted Enzyme Degradation platform (TED) molecule

The structure of the Targeted Enzyme Degradation platform (TED) molecule of the present invention is of formula Ia or formula I:



in the formula, the definitions of A, L1 and B are as described in the first aspect of the present invention.

Linker and coupling method

The linker of the present invention L1 is used for connecting the target molecule and the E3 ligase ligand.

Preferably, the target molecule or the E3 ligase ligand can be connected through -O-, -S-, -NH-, -NR-, -(C=O)-, -(C=O)O-, -SO₂- and other groups.

The linker of the present invention may further contain a variety of other functional groups, such as -OH, -NHR, -SH and the like.

Typically, the linker of the present invention L1, can be represented by the following general formula II:

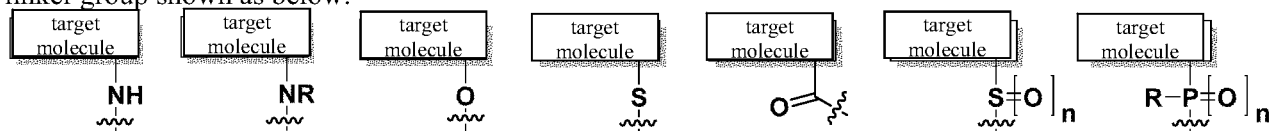


in the formula, the definitions of X, L2, and Y are as described in the first aspect of the present invention.

In another preferred embodiment, X and Y are each independently divalent groups formed by the loss of one hydrogen atom forming bivalence from the following monovalent groups: -OH, -NH₂, -SH, -COOH, -SO₂H.

Specifically, the connection mode of the linker and the target molecule can be connected through the

linker group shown as below:



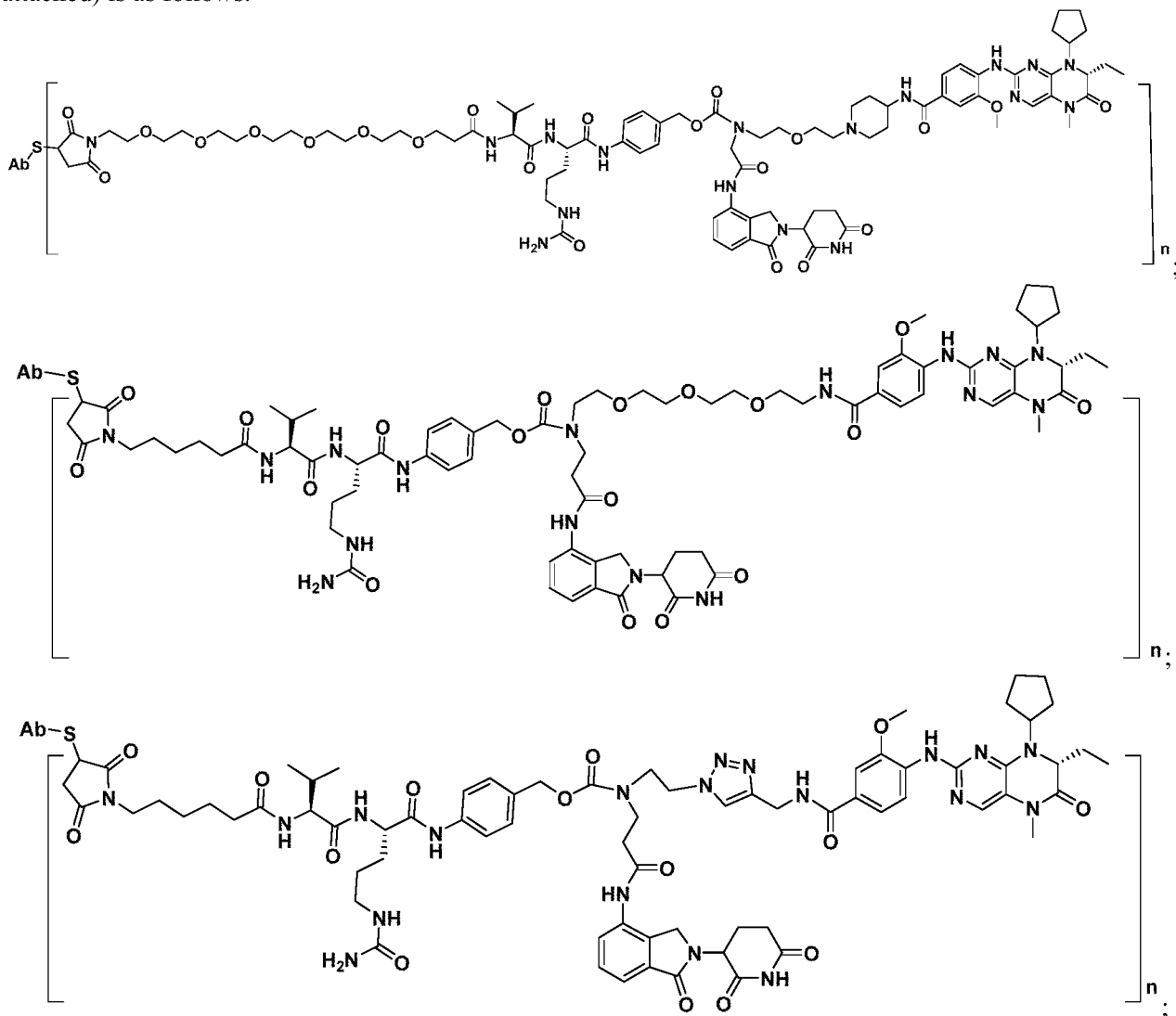
wherein, R in the each of above formulas is defined as above; n is 1 or 2 or 3.

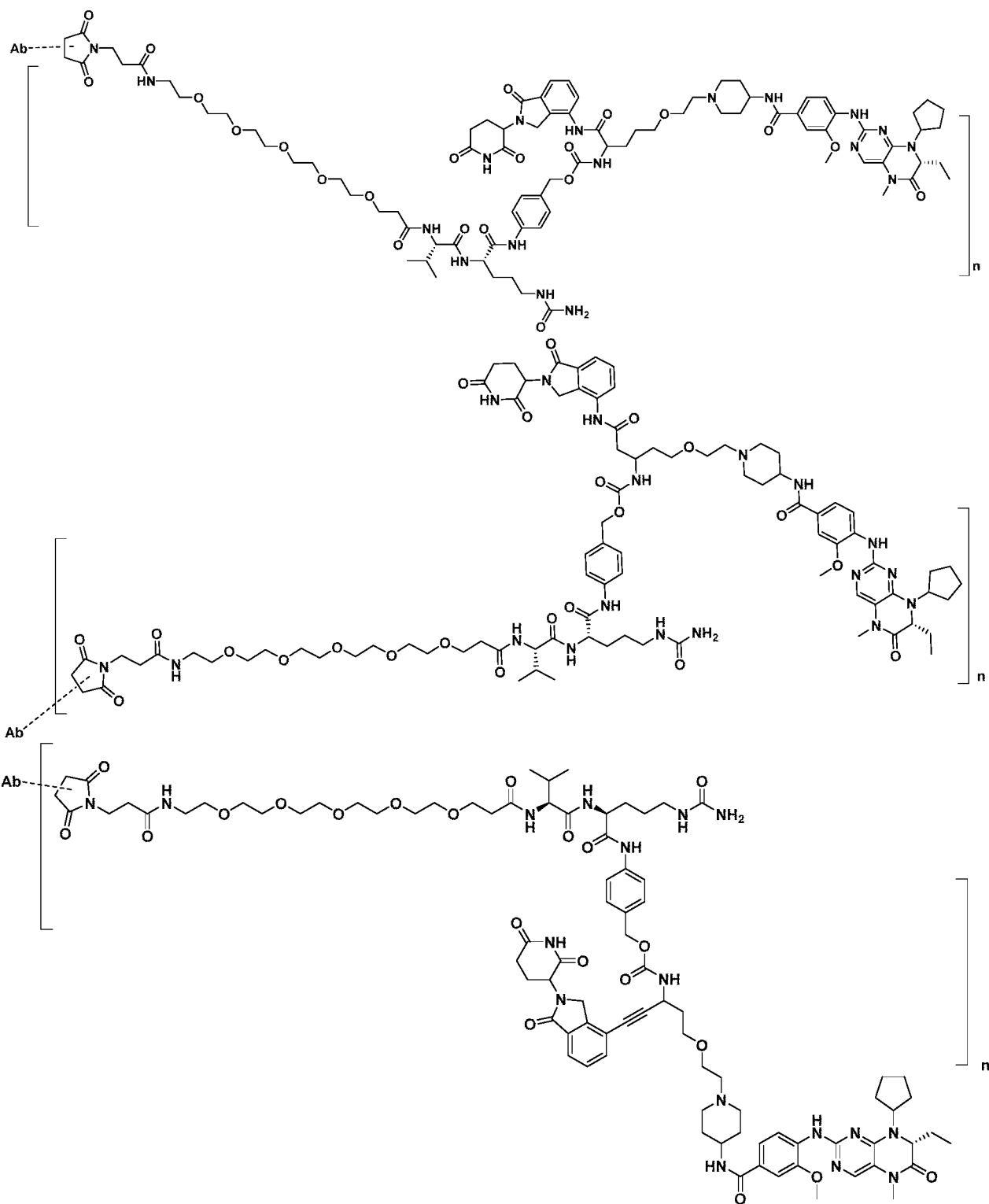
5 ACTED

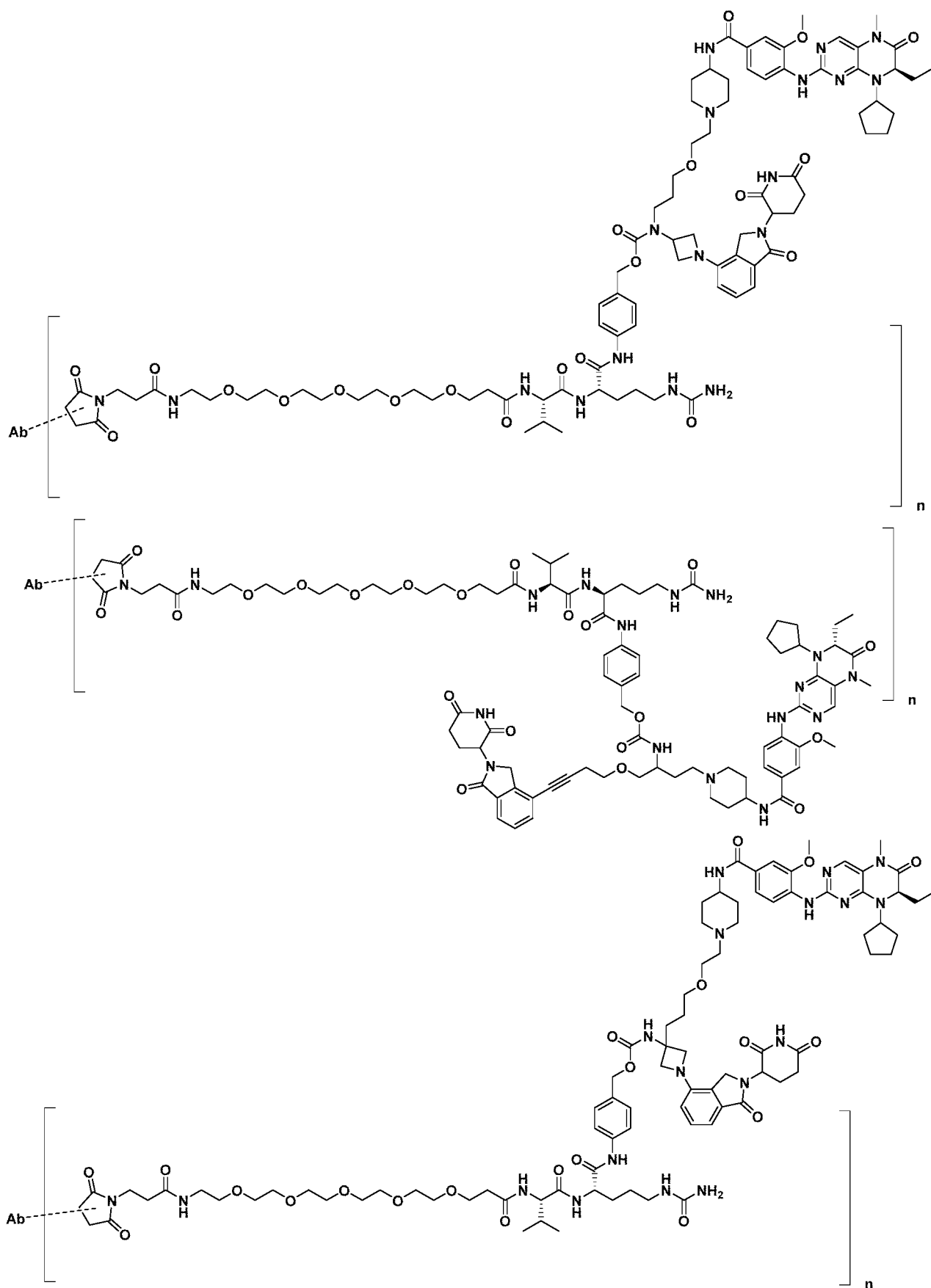
In the present invention, when the target molecule is an antibody, the conjugate of the present invention also can be referred to as ACTED or ACTED molecule or ACTED compound for short.

In the present invention, the structure of exemplary ACTED compounds (i.e, a TED molecule with Ab attached) is as follows:

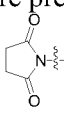
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wherein, $n=1$ to 15 (preferably, $n=1$ to 8, more preferably, $n=2$ to 7), Ab means an antibody; --- means

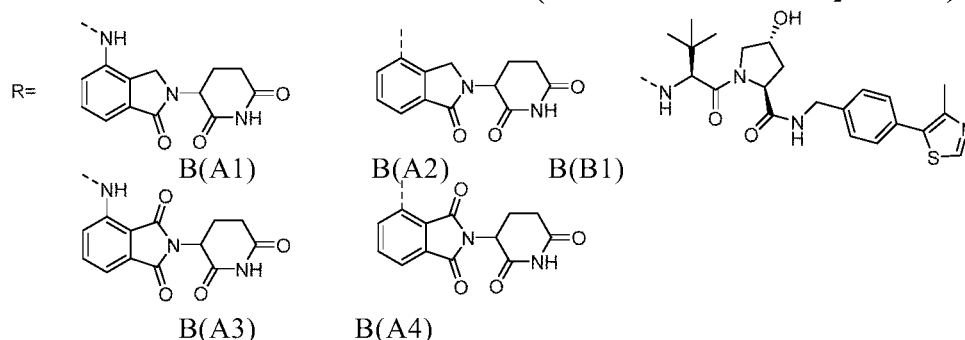
5 Ab is connected with L3 of formula III (such as , -NH-, -NHR or -NH₂ group in L3), through amino

acid at N-terminal or C-terminal, or a side chain of amino acid (such as a side chain of amino acid selected from the group consisting of Lys, and Cys), or a sulfhydryl formed by reducing and opening disulfide bond.

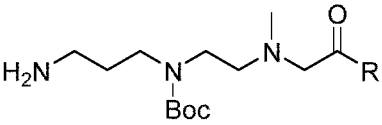
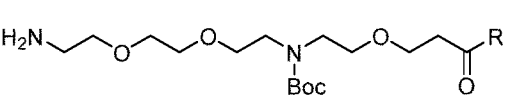
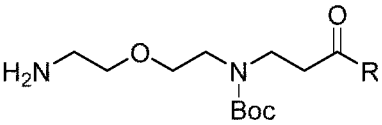
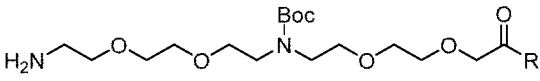
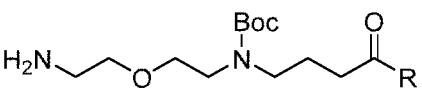
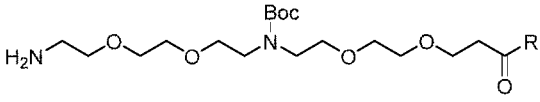
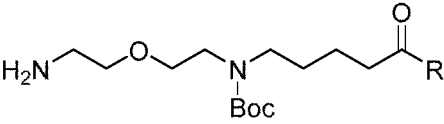
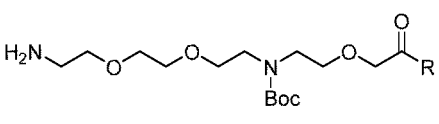
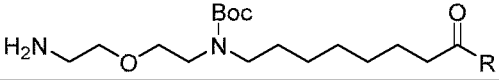
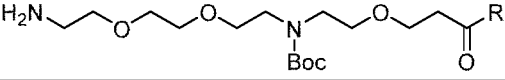
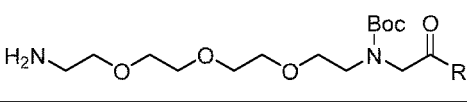
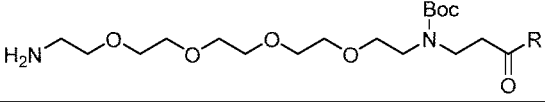
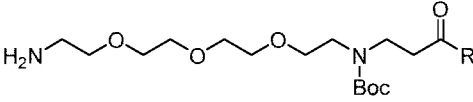
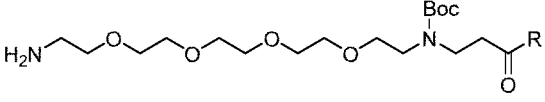
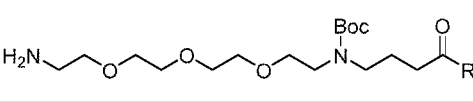
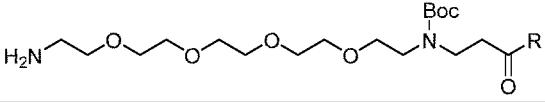
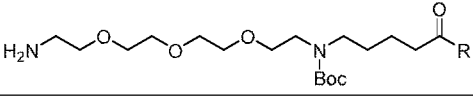
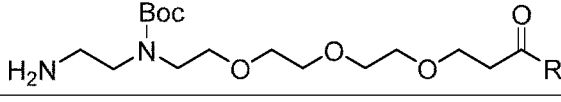
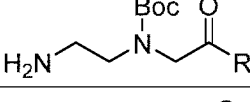
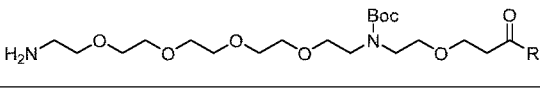
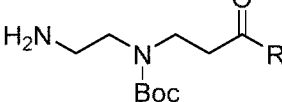
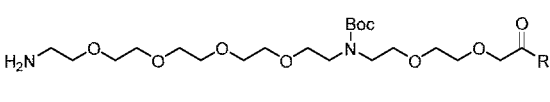
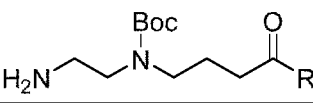
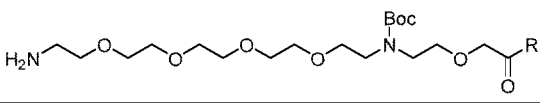
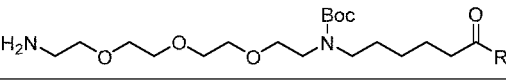
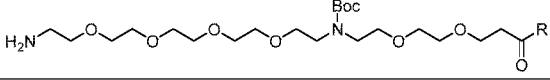
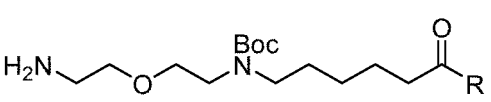
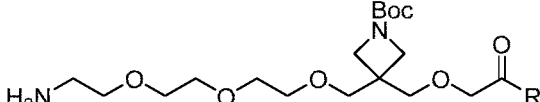
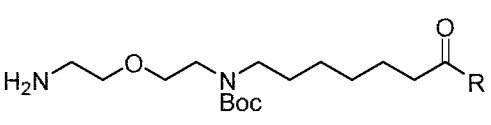
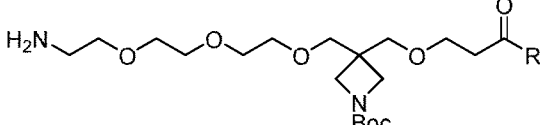
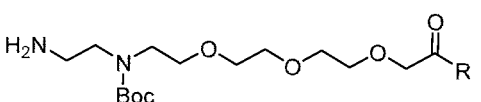
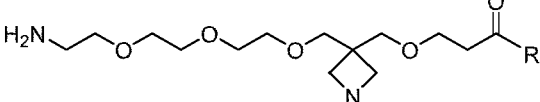
The present invention was further described hereafter in combination with specific embodiments. It should be understood that these examples are only used to illustrate the and not to limit the scope of the invention. The experimental methods without specific conditions in the following examples generally follow the conventional conditions or the conditions suggested by the manufacturer. Unless otherwise stated, percentages and parts are percentages by weight and parts by weight. Unless otherwise specified, the experimental materials and reagents used in the following examples are commercially available.

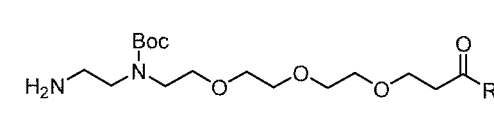
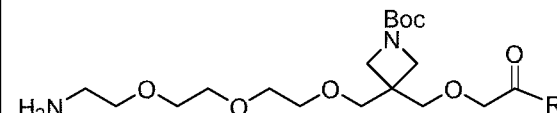
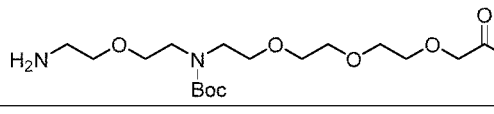
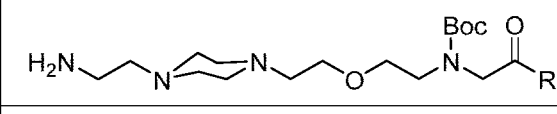
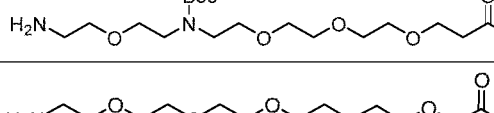
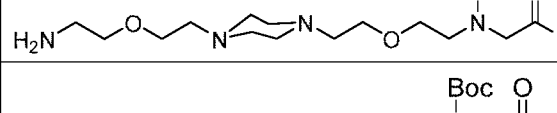
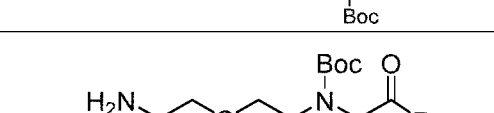
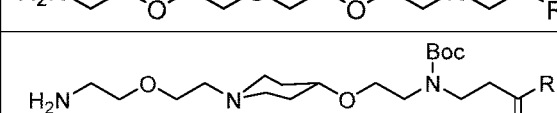
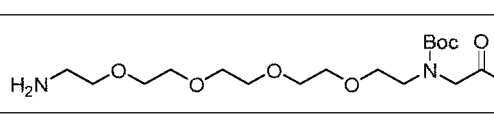
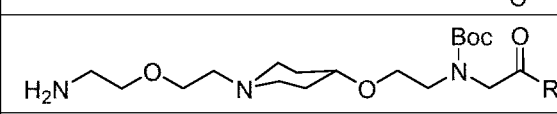
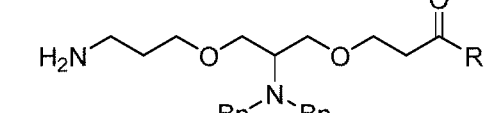
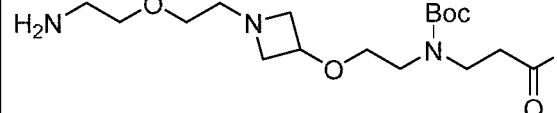
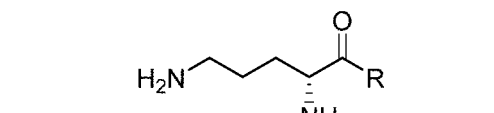
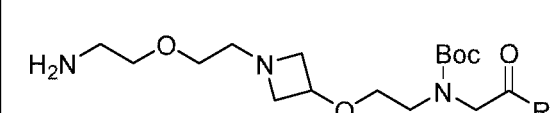
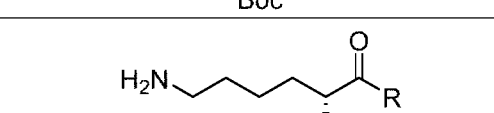
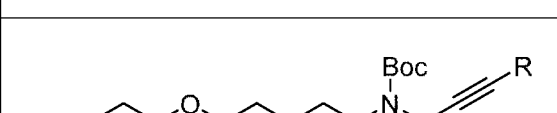
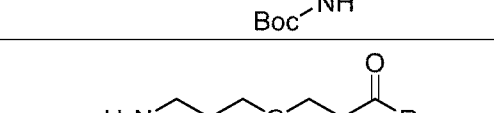
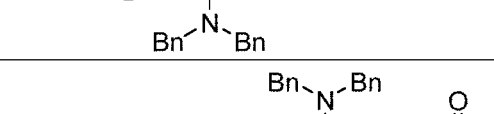
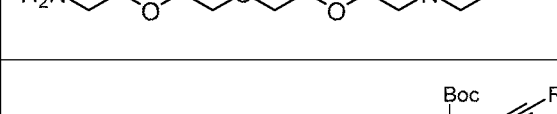
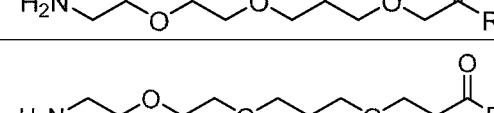
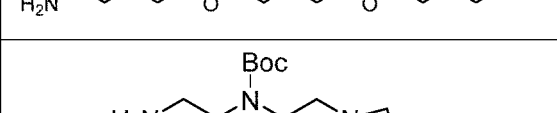
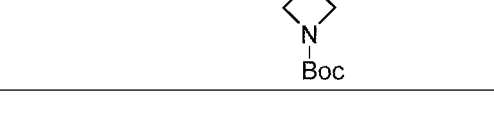
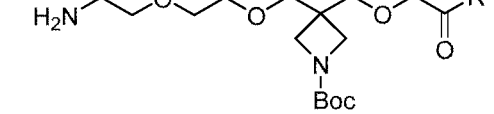
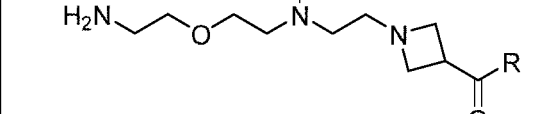
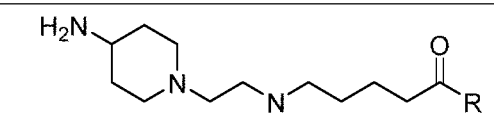
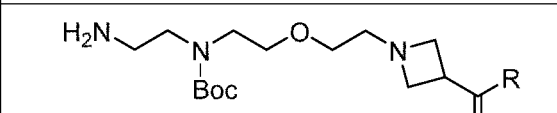
Example 1 Synthesis of L1-B moiety

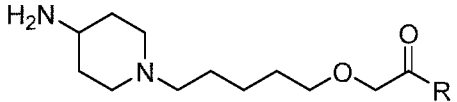
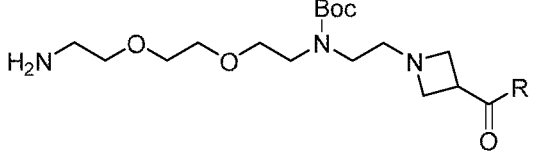
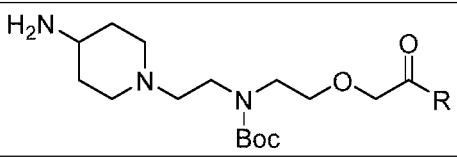
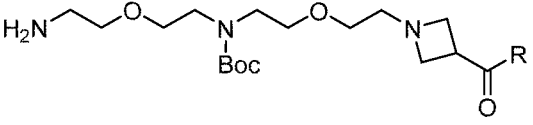
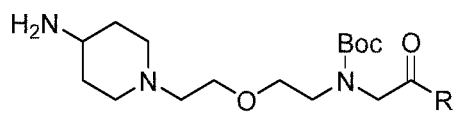
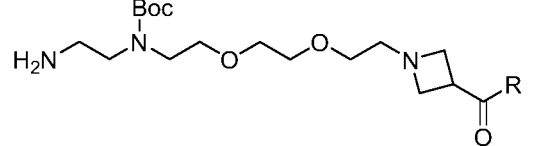
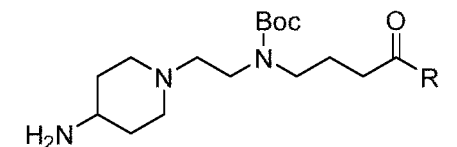
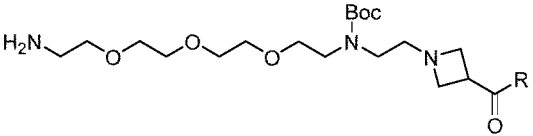
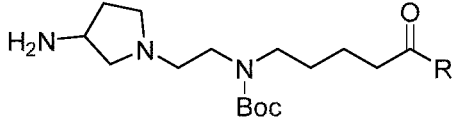
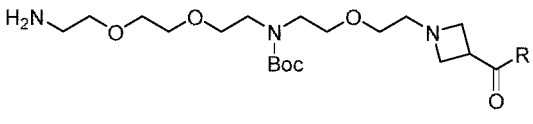
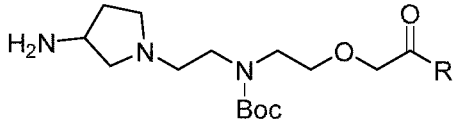
Table A1 the structure of linker in TED (i.e. the structure of NH₂-L2-Y-B)



Cmpd . No.	Structural formula	Cmpd . No.	Structural formula
L1		L44	
L2		L45	
L3		L46	
L4		L47	
L5		L48	
L6		L49	

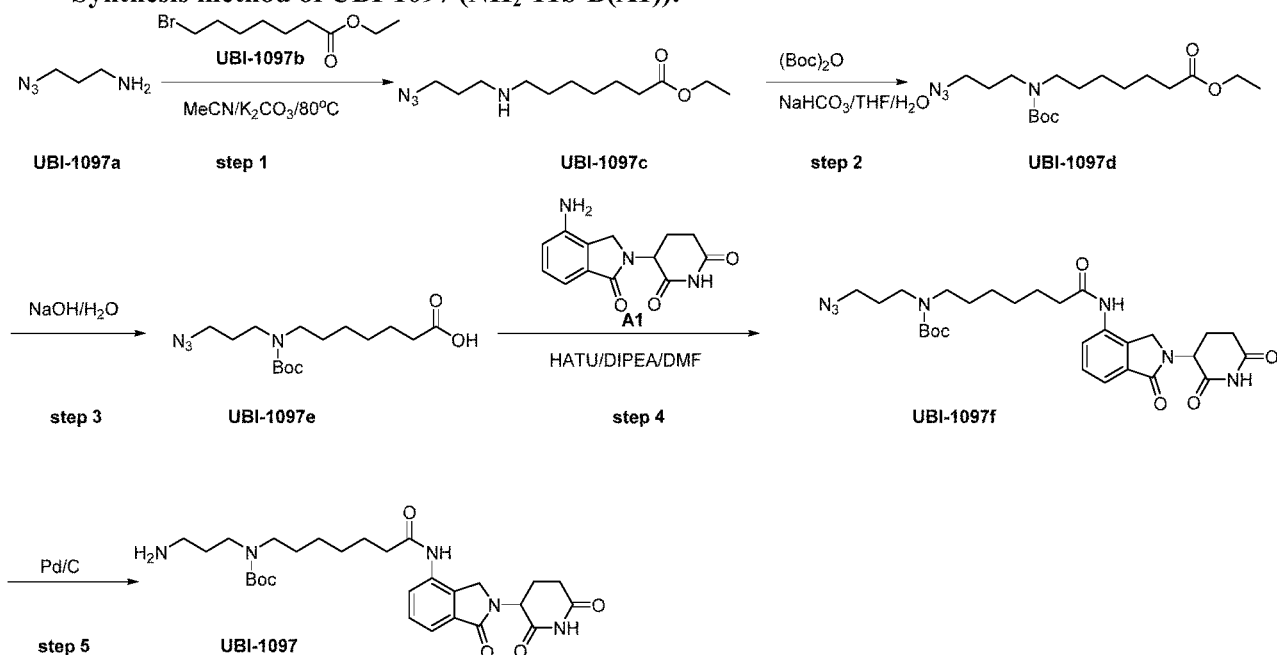
L7		L50	
L8		L51	
L9		L52	
L10		L53	
L11		L54	
L12		L55	
L13		L56	
L14		L57	
L15		L58	
L16		L59	
L17		L60	
L18		L61	
L19		L62	
L20		L63	
L22		L64	
L23		L65	

L24		L66	
L25		L67	
L26		L68	
L27		L69	
L28		L70	
L29		L71	
L30		L72	
L31		L73	
L32		L74	
L33		L75	
L34		L76	
L35		L77	
L36		L78	
L37		L79	

L38		L80	
L39		L81	
L40		L82	
L41		L83	
L42		L84	
L43			

Synthesis method of Compounds Linker b-Ligand B(A)

Synthesis method of UBI-1097 (NH₂-11b-B(A1)):



5

Step 1: UBI-1097c

Compound **UBI-1097a** (1.0 g, 10 mmol), compound **UBI-1097b** (2.36 g, 10 mmol) and K₂CO₃ (2.76 g, 20 mmol) were dissolved in CH₃CN (100 mL), then reacting at room temperature for 18 hours. After the completion of reaction, the reaction was filtrated. The filtrate was concentrated to obtain the crude product.

The crude product was isolated by silica gel column chromatography (petroleum ether/ethyl acetate = 50% to 80%, then dichloromethane/methanol = 0% to 10%) to obtain the target product **UBI-1097c** (450mg, yield 18%) as colorless transparent oil. ¹H NMR (400 MHz, chloroform-*d*) δ 4.12 (q, *J* = 7.1 Hz, 2H), 3.37 (t, *J* = 6.7 Hz, 2H), 2.70 (t, *J* = 7.0 Hz, 2H), 2.60 (t, *J* = 7.2 Hz, 2H), 2.29 (t, *J* = 7.5 Hz, 2H), 1.77 (p, *J* = 6.8 Hz, 2H), 1.67 – 1.60 (m, 2H), 1.53 – 1.44 (m, 2H), 1.38 – 1.30 (m, 4H), 1.25 (t, *J* = 7.1, 1.1 Hz, 3H).

Step 2: UBI-1097d

Compound **UBI-1097c** (450mg, 1.76mmol), di-*tert*-butyl dicarbonate (760mg, 3.52mmol) and NaHCO₃ (295mg, 3.52mmol) were dissolved in THF (20mL) and reacted at room temperature for 2 hours. After the completion of reaction, water (10mL) was added, extracted with dichloromethane (10mL *3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na₂SO₄, and concentrated by rotary evaporation under reduced pressure to obtain the crude product **UBI-1097d** (450mg, yield 71.9%), which was directly used in the next step.

Step 3: UBI-1097e

Compound **UBI-1097d** (450mg, 1.26mmol) and NaOH (200mg, 5.05 mmol) were dissolved in water (0.5mL), MeOH (3mL) and THF (3mL), and reacted at room temperature for 16 hours. After the completion of the reaction, the reaction was concentrated to obtain a crude product. Water (20 mL) was added, then extracted with ether (10 mL*3). The aqueous phase was acidified to pH=5 using dilute hydrochloric acid (1M), and then extracted with dichloromethane (10 mL*3). The organic phases were combined, then dried over anhydrous Na₂SO₄, and concentrated by rotary evaporation under reduced pressure to obtain compound **UBI-1097e** (370 mg, yield 64.4%). ¹H NMR (400 MHz, chloroform-*d*) δ 3.32 (t, *J* = 6.6 Hz, 2H), 3.22 – 3.14 (m, 2H), 3.10 (t, *J* = 7.3 Hz, 2H), 2.18 (t, *J* = 7.3 Hz, 2H), 1.70 (t, *J* = 7.0 Hz, 2H), 1.54 – 1.42 (m, 4H), 1.39 (s, 9H), 1.25 (m, 4H).

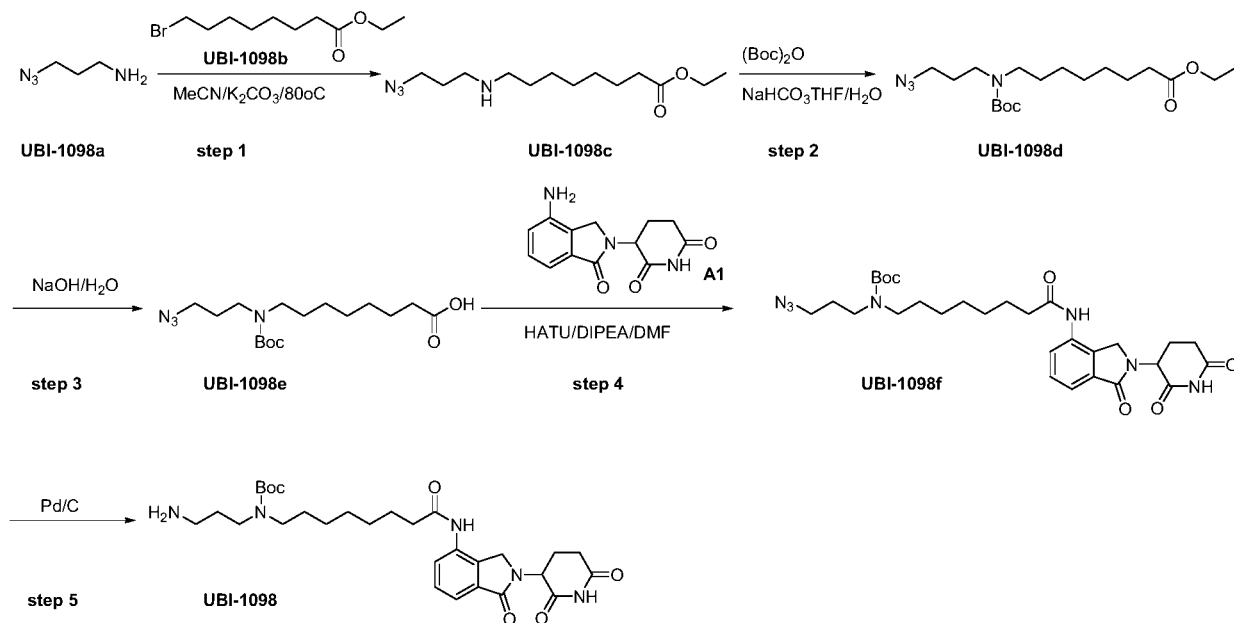
Step 4: UBI-1097f

Compound **UBI-1097e** (370 mg, 1.13 mmol), **B(A1)** (292 mg, 1.13 mmol), and HATU (859 mg, 2.26) were dissolved in DMF (3 mL) and DIPEA (0.3 mL), and reacted at room temperature for 16 hours. The reaction solution was directly isolated by reversed-phase column (MeOH/H₂O=5% to 95%, 45 minutes, collected at 70%) to obtain the target compound **UBI-1097f** (420mg, yield 65.4%) as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.02 (s, 1H), 9.75 (s, 1H), 7.82 (dd, *J* = 7.0, 2.0 Hz, 1H), 7.59 – 7.42 (m, 2H), 5.15 (dd, *J* = 13.3, 5.1 Hz, 1H), 4.51 – 4.27 (m, 2H), 3.23 – 3.08 (m, 4H), 2.92 (m, 1H), 2.61 (m, 1H), 2.41 – 2.29 (m, 3H), 2.02 (m, 1H), 1.71 (t, *J* = 6.9 Hz, 2H), 1.61 (t, *J* = 7.4 Hz, 2H), 1.38 (m, 17H). LC-MS: [M+H]⁺ = 570.2

Step 5: UBI-1097

Compound **UBI-1097f** (420mg, 0.74mmol) and Pd/C (42mg) were dissolved in MeOH (1 mL) and DCM (10mL), and reacted for 2 hours under hydrogen atmosphere. After the completion of reaction, the reaction was filtrated. The filtrate was concentrated to obtain the crude product. The crude product was washed with cold anhydrous ether (10 mL*3), and filtered to obtain target compound **UBI-1097** (340 mg, yield 85%) as white solid. ¹H NMR: (400 MHz, DMSO-*d*₆) δ 9.87 (d, *J* = 7.6 Hz, 1H), 7.82 (dd, *J* = 6.9, 2.1 Hz, 1H), 7.49 (d, *J* = 7.0 Hz, 2H), 5.15 (m, 1H), 4.37 (m, 2H), 3.21 – 3.15 (m, 3H), 3.11 (t, *J* = 7.4 Hz, 2H), 2.73 (t, *J* = 7.6 Hz, 2H), 2.37 (t, *J* = 7.4 Hz, 3H), 2.03 m, 1H), 1.76 (q, *J* = 7.7 Hz, 2H), 1.60 (q, *J* = 7.4 Hz, 2H), 1.39 (m, 15H). LCMS: (M+H)⁺ = 543.3

Synthesis method of Compound UBI-1098 (NH₂-12b-B(A1))



Step 1: UBI-1098c

Compound **UBI-1098a** (1.0 g, 10 mmol), **BUI-1098b** (2.5 g, 10 mmol) and K_2CO_3 (2.76 g, 20 mmol) were dissolved in CH_3CN (100 mL), reacted at 80°C for 18 hours. The reaction solution was filtered, and the filtrate was concentrated to obtain crude product. The crude product was isolated by column chromatography (petroleum ether/ethyl acetate = 50% to 80%, then DCM/MeOH=0% to 10%) to obtain the target compound **UBI-1098c** (600mg, yield 22%) as colorless oil. ^1H NMR (400 MHz, chloroform-*d*) δ 4.12 (q, $J = 7.1$ Hz, 2H), 3.36 (t, $J = 6.8$ Hz, 2H), 2.69 (t, $J = 7.0$ Hz, 2H), 2.59 (t, $J = 7.2$ Hz, 2H), 2.28 (t, $J = 7.5$ Hz, 2H), 1.77 (p, $J = 6.9$ Hz, 2H), 1.66 – 1.58 (m, 2H), 1.52 – 1.44 (m, 2H), 1.33 (m, 6H), 1.25 (t, $J = 7.1$ Hz, 3H).

Step 2: UBI-1098d

Compound **UBI-1098c** (450mg, 2.22mmol), di-tert-butyl dicarbonate (960mg, 4.44mmol) and NaHCO_3 (373mg, 4.44mmol) were dissolved in THF (20mL) and reacted at room temperature for 2 hours. After the completion of reaction, water (10mL) was added, extracted with dichloromethane (10mL *3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na_2SO_4 , and concentrated by rotary evaporation under reduced pressure to obtain the crude product **UBI-1098d** (600mg), which was directly used in the next step.

Step 3: UBI-1098e

Compound **UBI-1098d** (600mg, 1.62mmol), NaOH (260mg, 6.48mmol) were dissolved in water (0.5mL), MeOH (3mL) and THF (3mL), and reacted at room temperature for 16 hours. After the completion of the reaction, water (20 mL) was added, then extracted with ether (10 mL*3). The aqueous phase was acidified to pH=5 using dilute hydrochloric acid (1M), and then extracted with dichloromethane (10 mL*3). The organic phases were combined, then dried over anhydrous Na_2SO_4 , and concentrated by rotary evaporation under reduced pressure to obtain compound **UBI-1098e** (700mg, yield 92%).

^1H NMR (400 MHz, chloroform-*d*) δ 3.31 (t, $J = 6.6$ Hz, 2H), 3.14 (m, $J = 28.8, 7.2$ Hz, 4H), 2.18 (t, $J = 7.4$ Hz, 2H), 1.71 (p, $J = 6.8$ Hz, 2H), 1.39 (s, 13H), 1.31 – 1.12 (m, 6H).

Step 4: UBI-1098f

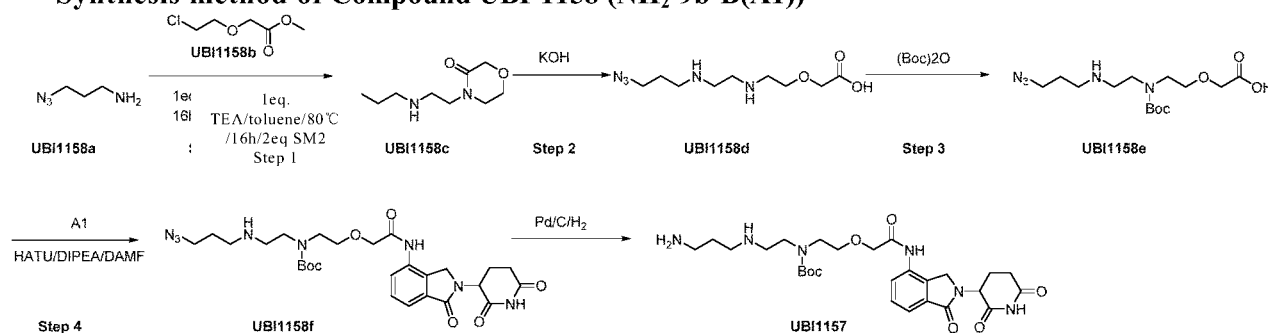
Compound **UBI-1098e** (700mg, 2mmol), **B(A1)** (518mg, 2mmol), and HATU (760mg, 4) were dissolved in DMF (3 mL) and DIPEA (0.3 mL), and reacted at room temperature for 16 hours. The reaction solution was directly isolated by reversed-phase column (MeOH/ H_2O =5% to 95%, 45 minutes, collected at 70%) to obtain the target compound **UBI-1098f** (720mg, yield 62%) as a white solid. ^1H NMR (400 MHz,

DMSO-*d*₆) δ 11.02 (s, 1H), 9.75 (s, 1H), 7.82 (dd, *J* = 7.0, 2.0 Hz, 1H), 7.59 – 7.42 (m, 2H), 5.15 (dd, *J* = 13.3, 5.1 Hz, 1H), 4.51 – 4.27 (m, 2H), 3.23 – 3.08 (m, 4H), 2.92 (m, 1H), 2.61 (m, 1H), 2.41 – 2.29 (m, 3H), 2.02 (m, 1H), 1.71 (t, *J* = 6.9 Hz, 2H), 1.61 (t, *J* = 7.4 Hz, 2H), 1.38 (m, 19H). LC-MS : [M+H]⁺ = 584.2

5 Step 4: UBI-1098

Compound **UBI-1098f** (700mg, 1.2mmol) and Pd/C (70mg) were dissolved in MeOH (1 mL) and DCM (10mL), and reacted for 2 hours under hydrogen atmosphere. After the completion of reaction, the reaction was filtrated. The filtrate was concentrated to obtain the crude product. The crude product was washed with cold anhydrous ether (10 mL*3), and filtered to obtain target compound **UBI-1098** (400mg, yield 60%) as white solid. ¹H NMR: (400 MHz, DMSO-*d*₆) δ 9.84 (s, 1H), 7.82 (dd, *J* = 7.0, 2.0 Hz, 1H), 7.56 – 7.39 (m, 2H), 5.15 (dd, *J* = 13.3, 5.1 Hz, 1H), 4.37 (d, *J* = 12.9 Hz, 2H), 3.33 (s, 2H), 3.17 (q, *J* = 5.4, 4.3 Hz, 2H), 3.10 (t, *J* = 7.4 Hz, 2H), 2.72 (t, *J* = 7.6 Hz, 2H), 2.36 (q, *J* = 8.7, 8.2 Hz, 3H), 2.08 – 1.98 (m, 1H), 1.74 (t, *J* = 7.5 Hz, 2H), 1.60 (t, *J* = 7.2 Hz, 2H), 1.48 – 1.30 (m, 15H), 1.23 (s, 2H).

15 Synthesis method of Compound UBI-1158 (NH₂-9b-B(A1))



Step 1: UBI-1158c

Compound **UBI-1158a** (2g, 20mmol), **UBI-1158b** (3.65g, 24mmol) were dissolved in triethylamine (2.02g, 20mmol) and toluene (100mL), reacted at 80°C for 18 hours. The reaction solution was concentrated and isolated by silica gel column chromatography (petroleum ether/ethyl acetate = 50% to 100%, then MeOH/DCM=0% to 10%) to obtain the target compound **UBI-1158c** (1.4g) as colorless oil. ¹H NMR (400 MHz, chloroform-*d*) δ 4.16 (s, 1H), 4.02 (s, 1H), 3.95 – 3.86 (m, 1H), 3.80 (dd, *J* = 6.0, 4.3 Hz, 1H), 3.70 (dd, *J* = 6.0, 4.3 Hz, 1H), 3.52 – 3.46 (m, 1H), 3.45 – 3.33 (m, 4H), 1.93 – 1.77 (m, 2H).

Step 2: UBI-1158d

Compound **UBI-1158c** (1.3g, 7.06mmol), KOH (1.19g, 21.2mmol) were dissolved in water (5mL), and refluxed at 100°C for 2 hours. The reaction liquid was directly used in the next step after cooling.

Step 3: UBI-1158e

To the aqueous solution of compound **UBI-1158d** (1.3g, 5.4mmol), di-tert-butyl dicarbonate (2.3g, 10.7mmol) and dioxane (30mL) were added, and reacted at room temperature for 2 hours. The reaction solution was concentrated, added water (20 mL), and extracted with ethyl acetate (10 mL*3). The aqueous phase was concentrated to obtain the crude product, which was dissolved in MeOH/DCM=10/1 (30 mL), filtered, and the filtrate was concentrated to obtain the target compound **UBI-1158e** (1.7g).

Step 4: UBI-1158f

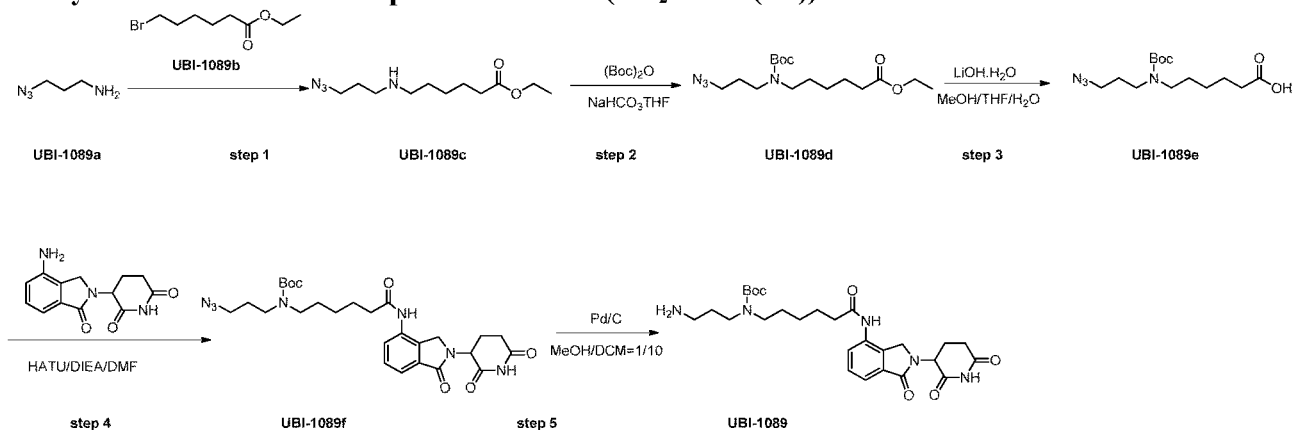
Compound **UBI-1158e** (600mg, 1.99mmol), **B(A1)** (514mg, 1.99mmol), and HATU (1.51g, 3.98mmol) were dissolved in DMF (3 mL) and DIPEA (0.3 mL), and reacted at room temperature for 16 hours. The reaction solution was directly isolated by reversed-phase column (MeOH/H₂O=5% to 95%, 45 minutes, collected at 60%) to obtain the target compound **UBI-1158f** (500mg) as a white solid. LC-MS: [M+H]⁺ = 587.3

Step 5: UBI-1158

Compound **UBI-1157** (100mg, 0.17mmol) and Pd/C (20mg) were dissolved in DCM (10mL), and reacted for 16 hours under hydrogen atmosphere. After the completion of reaction, the reaction was filtrated. The filtrate was concentrated to obtain the target compound which was directly used in the next step.

5 LC-MS: $(M+H)^+ = 561.3$

Synthesis method of Compound UBI-1089 (NH₂-11b-B(A1))



Step 1: UBI-1089c

10 Compound **UBI-1097a** (2.0 g, 20 mmol), **BUI-1097b** (3.57 g, 16 mmol) and K₂CO₃ (5.52 g, 40 mmol) were dissolved in acetonitrile (15 mL), then reacted at 80°C for 18 hours. The reaction solution was filtered and then concentrated to obtain crude product. The crude product was passed through silica gel column chromatography (ethyl acetate/petroleum ether=50% to 100%, 20 minutes, methanol/dichloromethane=0% to 10%, 20 minutes) to obtain product **UBI-1089c** (460mg, yield 9.5%) as light yellow oil.

15 ¹H NMR (400 MHz, chloroform-*d*) δ 4.13 (q, *J* = 7.1 Hz, 2H), 3.38 (dd, *J* = 7.6, 5.9 Hz, 2H), 2.73 (td, *J* = 7.0, 3.1 Hz, 2H), 2.64 (d, *J* = 6.9 Hz, 2H), 2.30 (t, *J* = 7.4 Hz, 2H), 1.88 – 1.76 (m, 2H), 1.65 (p, *J* = 7.5 Hz, 2H), 1.53 (m, 2H), 1.38 (dt, *J* = 10.2, 6.4 Hz, 2H), 1.25 (t, *J* = 7.1 Hz, 3H).

Step 2: UBI-1089d

20 The compound **UBI-1097c** (460 mg, 2 mmol), (Boc)₂O (460 mg, 2 mmol) and NaHCO₃ (320 mg, 4 mmol) were dissolved in tetrahydrofuran (10 mL), and then reacted at room temperature for 2 hours. The reaction solution was added water (10 mL), and extracted with dichloromethane (10 mL*3). Organic phases were combined, dried over Na₂SO₄, and concentrated. After the concentration, the crude product **UBI-1089d** (770mg, 100% yield) as yellow oil was obtained, which was directly used in the next step.

Step 3: UBI-1089e

25 Compound **UBI-1089d** (770mg, 2mmol), LiOH (140mg, 3mmol) were dissolved in water (3mL), methanol (3mL) and tetrahydrofuran (6mL), and then reacted at room temperature for 18 hours. The reaction solution was concentrated and extracted with ethyl acetate (10mL*3), the aqueous phase was acidified to pH=5 using (1M) HCl. Then the aqueous phase was extracted with ethyl acetate (10mL*3). The organic phases were combined, then dried over anhydrous sodium sulfate and concentrated to obtain product **UBI-1089e** (450mg, yield 63.6%) as yellow oil.

Step 4: UBI-1089f

30 Compound **UBI-1089e** (450mg, 1.43mmol), **B(A1)** (371mg, 1.43mmol), and HATU (1.09g, 2.87) were dissolved in DMF (3 mL) and DIPEA (0.3 mL), and reacted at room temperature for 18 hours. The reaction was quenched with water (10 mL), then extracted with ethyl acetate (8 mL*3). The organic phases were combined, then dried over anhydrous sodium sulfate and concentrated to obtain crude product. The crude

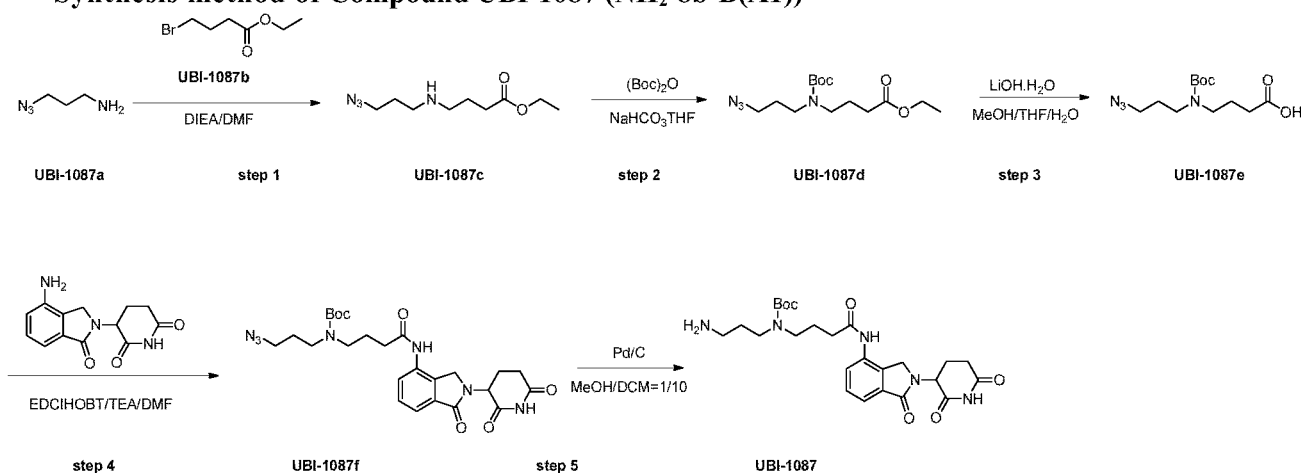
product was purified by reversed-phase column (methanol/water=5% to 95%, collected at 70%) to obtain product **UBI-1089f** (290mg, yield 36.5%) as colorless oil.

¹H NMR (400 MHz, DMSO-*d*₆) δ 11.02 (s, 1H), 9.76 (s, 1H), 7.81 (d, *J* = 6.8 Hz, 1H), 7.49 (d, *J* = 7.2 Hz, 2H), 5.14 (dd, *J* = 13.2, 5.2 Hz, 1H), 4.49 – 4.24 (m, 2H), 3.20 – 3.11 (m, 6H), 2.99 – 2.87 (m, 1H), 2.69 – 2.51 (m, 4H), 2.35 (q, *J* = 5.7, 4.0 Hz, 3H), 1.80 – 1.56 (m, 4H), 1.37 (s, 9H), 1.28 (m, 2H).

Step 5: UBI-1089

Under hydrogen atmosphere, compound **UBI-1089f** (290mg, 0.52mmol), Pd/C (29mg) were dissolved in methanol (1 mL) and dichloromethane (10mL), and the reaction solution was reacted at room temperature for 1 hour. The reaction solution was filtered through Celite. The filtrate was concentrated and purified via reversed-phase column (methanol/water=5% to 95%, collected at 15%) to obtain the product **UBI-1089** (180mg, yield 65%) as yellow oil.

Synthesis method of Compound UBI-1087 (NH₂-8b-B(A1))



Step 1: UBI-1087c

Compound **UBI-1087a** (1.5 g, 15 mmol), **BUI-1087b** (1.46 g, 8 mmol) and DIEA (5.81 g, 45 mmol) were dissolved in DMF (15 mL), and reacted at room temperature for 18 hours. The reaction solution was added water (15 mL), and extracted with ethyl acetate (15 mL*3). The crude product was obtained after concentration of the organic phase. The crude product was passed through silica gel column chromatography (ethyl acetate/petroleum ether=50% to 80%, 20 minutes, methanol/dichloromethane = 0% to 10%, 20 minutes) to obtain product **UBI-1087c** (480mg, yield 15%) as colorless oil.

¹H NMR (400 MHz, chloroform-*d*) δ 4.26 – 3.95 (m, 2H), 3.42 – 3.25 (m, 2H), 2.67 (dtd, *J* = 22.1, 7.0, 1.2 Hz, 4H), 2.36 (tt, *J* = 7.3, 1.8 Hz, 2H), 1.86 – 1.67 (m, 4H), 1.51 (s, 1H), 1.40 – 1.16 (m, 3H).

Step 2: UBI-1087d

The compound **UBI-1087c** (480mg, 2mmol), (Boc)₂O (540mg, 2.2 mmol) and NaHCO₃ (380mg, 4 mmol) were dissolved in tetrahydrofuran (15 mL), and reacted at room temperature for 2 hours. The reaction solution was added water (10 mL), and extracted with ethyl acetate (10 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate and then concentrated to obtain crude product **UBI-1087d** (600 mg, yield 85%) as yellow oil, the crude product was directly used in the next step.

Step 3: UBI-1087e

Compound **UBI-1087d** (600 mg, 1.9 mmol), LiOH water (88 mg, 2.1 mmol) were dissolved in water (2 mL), methanol (2 mL) and tetrahydrofuran (6 mL), reacted at room temperature for 18 hours. The reaction solution was concentrated and extracted with ethyl acetate (10 mL*3). The aqueous phase was acidified to pH=5 using (1M) HCl, and extracted with ethyl acetate (10 mL*3). The organic phases were combined and dried over anhydrous sodium sulfate, then concentrated to obtain product **UBI-1087e** (480mg, yield 77%)

as yellow oil.

^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 12.03 (s, 1H), 3.68 – 3.55 (m, 2H), 3.22 – 3.09 (m, 4H), 2.17 (t, J = 7.2 Hz, 2H), 1.75 – 1.62 (m, 4H), 1.39 (s, 9H).

Step 4: UBI-1087f

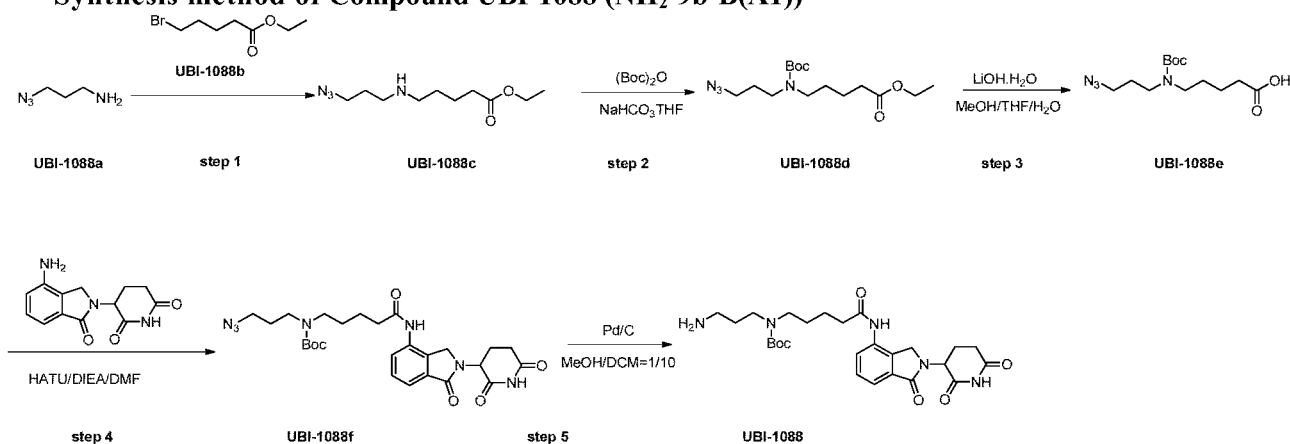
Compound **UBI-1087e** (420 mg, 1.47 mmol), **B(A1)** (380 mg, 1.47 mmol), EDCI (564 mg, 2.94 mmol), HOBT (595 mg, 4.41 mmol) and TEA (445 mg, 4.41 mmol) were dissolved in DMF (12 mL) and reacted at room temperature for 18 hours. The reaction solution was quenched by the addition of water (10 mL), then extracted with ethyl acetate (15 mL*3). The organic phases were combined, then washed with saturated brine, dried over anhydrous sodium sulfate and purified by reversed-phase column to obtain product **UBI-1087f** (280 mg, yield 36%) as yellow solid.

^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 12.06 (s, 1H), 11.02 (s, 1H), 9.81 (s, 1H), 7.80 (d, J = 7.0 Hz, 1H), 7.64 – 7.28 (m, 2H), 5.15 (dd, J = 13.3, 5.1 Hz, 1H), 4.49 – 4.21 (m, 2H), 2.93 (ddd, J = 18.6, 13.5, 5.3 Hz, 1H), 2.61 (d, J = 18.3 Hz, 2H), 2.38 – 2.32 (m, 2H), 2.17 (t, J = 7.3 Hz, 2H), 2.06 – 1.95 (m, 1H), 1.88 – 1.59 (m, 8H), 1.39 (d, J = 1.8 Hz, 9H).

Step 5: UBI-1087

Under hydrogen atmosphere, compound **UBI-1087f** (280mg, 0.53mmol), Pd/C (28mg) were dissolved in methanol (1 mL) and dichloromethane (10mL), and the reaction solution was reacted at room temperature for 1 hour. The reaction solution was filtered through Celite. The filtrate was concentrated and purified via reversed-phase column (methanol/water = 5% to 95%, collected at 20%) to obtain the product **UBI-1087** (80mg, yield 30%) as yellow oil.

Synthesis method of Compound UBI-1088 (NH_2 -9b-B(A1))



Step 1: UBI-1088c

Compound **UBI-1188a** (2g, 20mmol), **BUI-1188b** (3.34g, 16mmol) and K_2CO_3 (5.52g, 40mmol) were dissolved in acetonitrile (15 mL), then the reaction was reacted at room temperature for 18 hours. The reaction solution was filtered. The filtrate was concentrated and passed through silica gel column chromatography (ethyl acetate/petroleum ether = 50% to 100%, 20 minutes, methanol/dichloromethane = 0% to 10%, 30 minutes) to obtain product **UBI-1188c** (700 mg, yield 15%) as light yellow oil.

Step 2: UBI-1088d

Compound **UBI-1088c** (700mg, 3.1mmol), $(\text{Boc})_2\text{O}$ (740mg, 3.4mmol) and NaHCO_3 (520mg, 6.2mmol) were dissolved in tetrahydrofuran (15mL) and reacted at room temperature for 2 hours. The reaction solution was added water (10 mL), then extracted with ethyl acetate (10 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate and then concentrated to obtain crude product **UBI-1088d** (1 g, yield 100%) as yellow oil, the crude product was directly used in the next step.

Step 3: UBI-1088e

Compound **UBI-1088d** (1g, 3 mmol), LiOH.water (190 mg, 5 mmol) were dissolved in water (2 mL), methanol (2 mL) and tetrahydrofuran (6 mL), reacted at room temperature over night. The reaction solution was concentrated and then extracted with ethyl acetate (10 mL*3). The aqueous phase was acidified to pH=5 using (1M) HCl, and extracted with ethyl acetate (10 mL*3). The organic phases were combined and dried over anhydrous sodium sulfate to obtain crude product **UBI-1088e** (600mg, yield 66%) as yellow oil.

Step 4: UBI-1088f

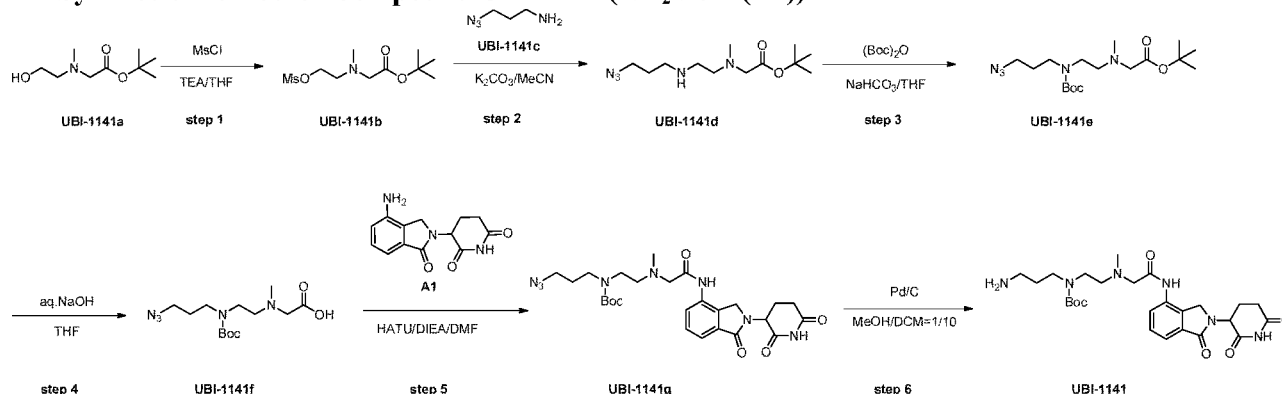
Compound **UBI-1088e** (640mg, 2.13mmol), **B(A1)** (497mg, 1.92mmol), and HATU (1.62g, 4.27) were dissolved in DMF (8 mL) and DIPEA (0.5 mL), and reacted at room temperature for 18 hours. The reaction solution was directly purified via reversed-phase column (methanol/water = 5 % to 95 %, collected at 70%) to obtain product **UBI-1088f** (380mg, yield 33%) as light yellow oil.

¹H NMR (400 MHz, DMSO-*d*₆) δ 11.02 (s, 1H), 9.77 (s, 1H), 7.81 (dd, *J* = 7.0, 2.0 Hz, 1H), 7.57 – 7.43 (m, 2H), 5.15 (dd, *J* = 13.3, 5.1 Hz, 1H), 4.45 – 4.26 (m, 2H), 3.23 – 3.06 (m, 6H), 3.00 – 2.85 (m, 1H), 2.69 – 2.56 (m, 2H), 2.38 (t, *J* = 7.0 Hz, 2H), 2.10 – 1.98 (m, 1H), 1.77 – 1.66 (m, 2H), 1.54 (m, 4H), 1.39 (s, 9H).

Step 5: UBI-1088

Under hydrogen atmosphere, compound **UBI-1188f** (390mg, 0.72mmol), Pd/C (39mg) were dissolved in methanol (1 mL) and dichloromethane (10mL), and reacted at room temperature for 1 hour. The reaction solution was filtered on Celite. The filtrate was concentrated to obtain crude product **UBI-1088** (350 mg, yield 95%) as yellow oil, the crude product was directly used in the next step.

Synthesis method of Compound UBI-1141 (NH₂-9b-B(A1))



Step 1: UBI-1141b

After Compound **UBI-1141a** (2.0 g, 10.6 mmol) was dissolved in tetrahydrofuran (25 mL), TEA (1.6 g, 15.9 mmol) was added, and then MsCl (1.5 g, 12.7 mmol) was added at 0°C. The reaction was reacted at room temperature for 18 hours. The reaction solution was quenched by adding water (30 mL), then extracted with ethyl acetate (20 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate and concentrated to obtain crude product **UBI-1141b** (2.5 g, yield 89%) as yellow oil. The crude product was directly used in the next step.

Step 2: UBI-1141d

Compound **UBI-1141c** (1.3 g, 13mmol), **BUI-1141b** (2.78g, 10mmol) and K₂CO₃ (2.59 g, 26mmol) were dissolved in acetonitrile (15 mL), then reacted at RT for 18 hours. The reaction solution was filtered. The filtrate was concentrated and passed through silica gel column chromatography (ethyl acetate/petroleum ether=50% to 100%, 20 minutes, methanol/dichloromethane = 0% to 10%, 30 minutes) to obtain product **UBI-1141d** (450 mg, yield 16%) as light yellow oil.

Step 3: UBI-1141e

Compound **UBI-1141d** (450mg, 1.7mmol), **(Boc)₂O** (543mg, 2.5mmol) and **NaHCO₃** (279mg, 3.3mmol) were dissolved in tetrahydrofuran (15mL) and reacted at room temperature for 2 hours. The reaction solution was quenched with water (10 mL), and extracted with ethyl acetate (10 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate and concentrated to obtain crude product **UBI-1141e** (550 mg, yield 89%) as light yellow oil. The crude product was directly used in the next step. ¹H NMR (400 MHz, chloroform-*d*) δ 3.30 (q, *J* = 7.1 Hz, 6H), 3.19 (s, 2H), 2.66 (s, 2H), 2.41 (s, 3H), 1.81 (p, *J* = 6.9 Hz, 2H), 1.46 (d, *J* = 2.7 Hz, 18H).

Step 4: UBI-1141f

Compound **UBI-1141e** (550 mg, 1.5 mmol) was dissolved in 50% aq. NaOH (10 mL) and tetrahydrofuran (10 mL), and reacted at room temperature for 18 hours. The reaction solution was concentrated and then extracted with ethyl acetate (10 mL) once. The aqueous phase was acidified to pH=5 using (1M) HCl, and extracted with ethyl acetate (10 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate and concentrated to obtain crude product **UBI-1141f** (400mg, yield 86%) as light yellow oil. ¹H NMR (400 MHz, chloroform-*d*) δ 3.39 – 3.14 (m, 8H), 2.83 (s, 2H), 2.53 (s, 3H), 1.85 – 1.68 (m, 2H), 1.45 (s, 9H).

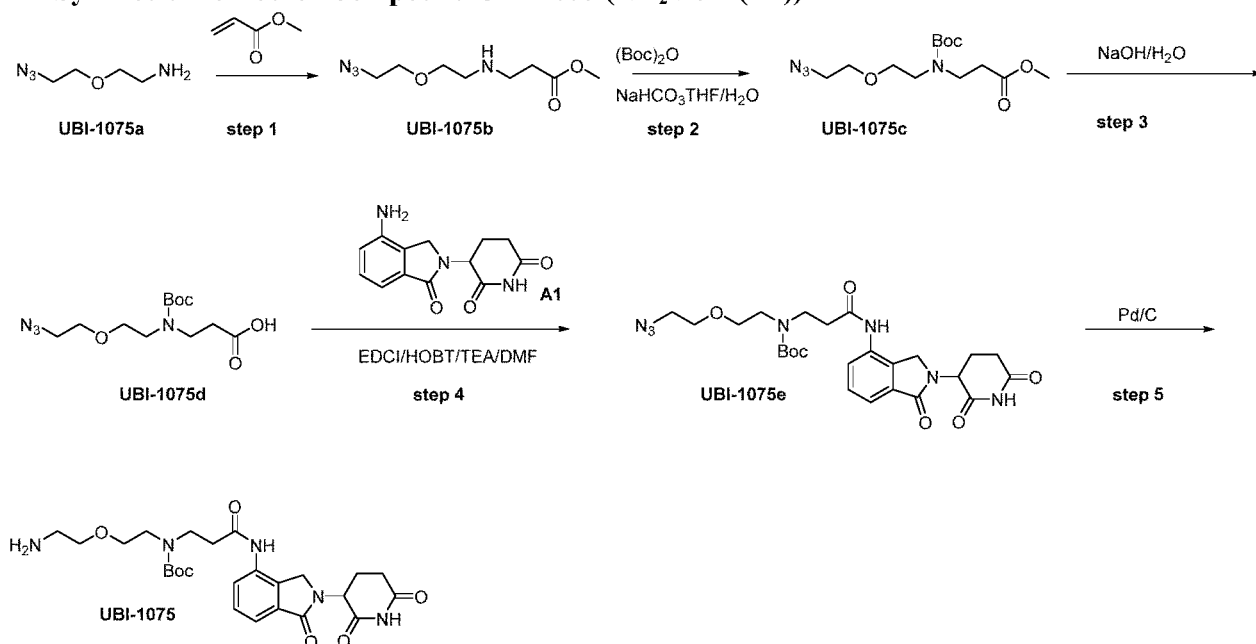
Step 5: UBI-1141g

Compound **UBI-1141f** (200mg, 0.63mmol), **B(A1)** (148mg, 0.57mmol), and HATU (483mg, 4.27) were dissolved in DMF (8 mL) and DIPEA (0.5 mL), and reacted at room temperature for 16 hours. The reaction solution was directly purified via reversed-phase column (methanol/water = 5 % to 95 %, collected at 70%) to obtain product **UBI-1141g** (30mg, yield 9%) as yellow oil.

Step 6: UBI-1141

Under hydrogen atmosphere, compound **UBI-1141g** (50mg, 0.09mmol), Pd/C (5mg) were dissolved in methanol (1 mL) and dichloromethane (10mL), and the reaction was reacted at room temperature for 1 hour. The reaction solution was filtered on Celite. The filtrate was concentrated to obtain crude product **UBI-1141** (50 mg, yield 100%). The crude product was directly used in the next step.

Synthesis method of Compound UBI-1075 (NH₂-9b-B(A1))



Step 1: UBI-1075b

Compound **UBI-1075a** (1.0 g, 7.69 mmol) and methyl acrylate (984 mg, 7.69 mmol) were dissolved in

acetonitrile (20 mL), and reacted at 80 ° C for 18 hours. After the completion of reaction, the reaction solution was poured into 5 mL water and extracted with ethyl acetate (5 mL*3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na₂SO₄, concentrated by rotary evaporation under reduced pressure to obtain crude product. The crude product was isolated by silica gel column chromatography (DCM/MeOH = 10/1) to obtain compound as colorless oil (660 mg, yield 40%).

¹H NMR (400 MHz, d₆-DMSO) δ 3.67 – 3.62 (m, 1H), 3.60 – 3.57 (m, 5H), 3.48– 3.43 (m, 2H), 3.39 – 3.36 (m, 2H), 2.77-2.74 (m, 2H), 2.70-2.63(m, 2H), 2.43-2.40(m, 2H).

Step 2: UBI-1075c

Compound **UBI-1075b** (660 mg, 3.06 mmol), di-tert-butyl dicarbonate (975 mg, 4.58 mmol) and sodium bicarbonate (771 mg, 9.18 mmol) were added to tetrahydrofuran (20mL) and reacted at room temperature for 2 hours. After the completion of reaction, it was poured into 10 mL of water and extracted with dichloromethane (5 mL*3). The organic phases were combined and washed with saturated brine, dried over anhydrous Na₂SO₄, and concentrated by rotary evaporation under reduced pressure to obtain compound **UBI-1075c** (490 mg, yield 64%). ¹H NMR (400 MHz, d₆-DMSO) δ 3.60 – 3.56 (m, 5H), 3.51 – 3.48 (m, 2H), 3.43–3.38 (m, 4H), 1.43 (s, 9H), 1.38 (s, 4H).

Step 3: UBI-1075d

Compound **UBI-1075c** (316 mg, 1.0 mmol) and sodium hydroxide (200 mg, 5.0 mmol) were added to a system of water/methanol/tetrahydrofuran (0.5mL/3mL/3mL), and reacted at room temperature for 16 hours. After the completion of reaction, the organic solvent was removed by concentration, and the aqueous phase was acidified to pH=5 using hydrochloric acid (1M). Then it was extracted with dichloromethane (10ml*3), the combined organic phase was dried over anhydrous Na₂SO₄, and concentrated to obtain the desired compound **UBI-1075d** (170mg, yield 56.3%). ¹H NMR (400 MHz, d₆-DMSO) δ 12.16 (s, 1H), 3.60-3.57 (m, 2H), 3.54-3.49(m,2H), 3.46(dt, *J* = 11.1, 5.1 Hz, 4H), 3.47 – 3.38 (m, 2H), 2.48– 2.40 (m, 2H), 1.39 (s, 9H).

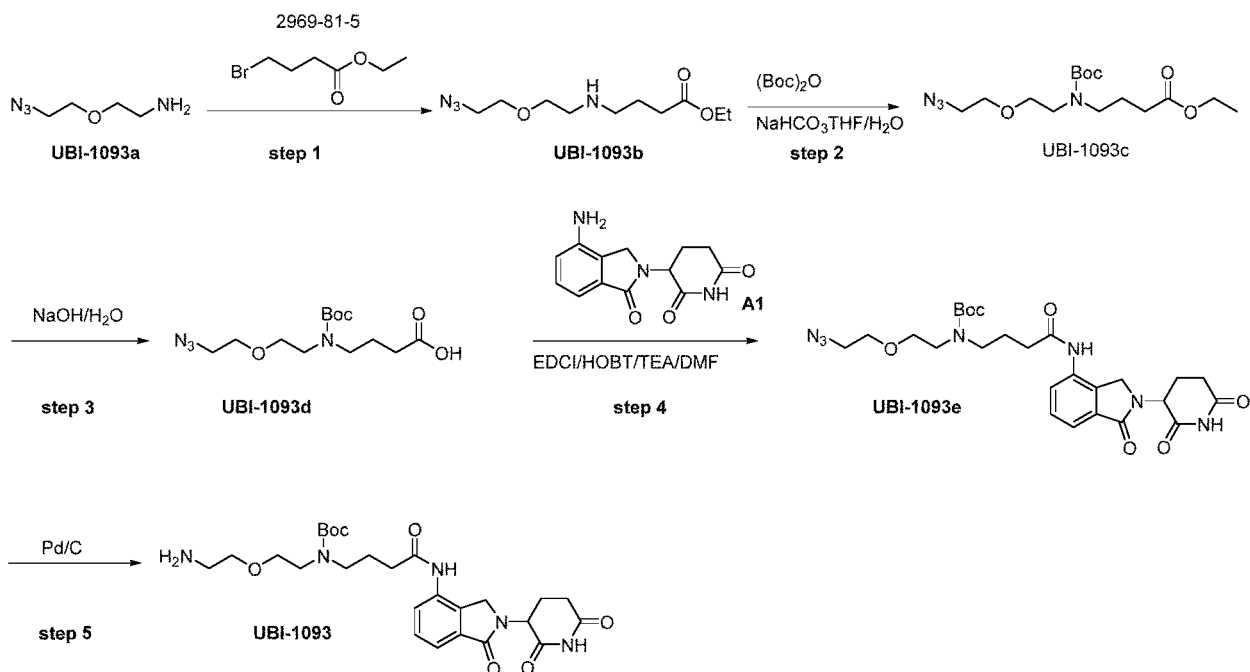
Step 4: UBI-1075e

Compound **UBI-1075d** (268 mg, 0.88 mmol), **B(A1)** (230 mg, 0.88mmol), HATU (337mg, 0.97) and DIPEA (0.3mL) were added to anhydrous DMF (3mL). The mixture was reacted at room temperature for 16 hours. After the completion of reaction, the organic solvent was removed by concentration. The crude product was isolated by reverse chromatography on silica gel column chromatography (MeOH/H₂O = 5% to 95%, 45mins, collected at 60%) to obtain compound **UBI-1075e** (350 mg, yield 72.7%) as colorless oil.

Step 5: UBI-1075

Compound **UBI-1075e** (200 mg, 0.37 mmol), 10% palladium on carbon (42 mg) was added to a mixed solvent of methanol/dichloromethane (1 mL/10 mL) , and reacted at room temperature for 2 hours under hydrogen atmosphere. After filtration, the filtrate was concentrated to obtain the crude product. The crude product was washed with cold ether (10 mL*3), and dried to obtain target compound **UBI-1075** (169 mg, yield 88.9%) as colorless oil.

Synthesis method of Compound UBI-1093 (NH₂-10b-B(A1))



Step 1: UBI-1093b

Compound **UBI-1093a** (970 mg, 7.46 mmol), ethyl 4-bromobutyrate (1.45 g, 7.46 mmol) and K_2CO_3 (2.06 g, 14.9 mmol) were dissolved in acetonitrile (20 mL), and reacted at 80° C for 18 hours, filtered after the completion of reaction, the filtrate was concentrated by rotary evaporation under reduced pressure, and isolated by silica gel column chromatography (DCM/MeOH = 10/1) to obtain compound **UBI-1093b** (710 mg, yield 22 %) as colorless oil.

^1H NMR (400 MHz, d_6 -DMSO) δ 4.09 – 4.02 (m, 2H), 3.62 – 3.58 (m, 2H), 3.52 – 3.48 (m, 2H), 3.41 – 3.37 (m, 2H), 2.68 – 2.64 (m, 2H), 2.58 – 2.54 (m, 2H), 2.37 – 2.35 (m, 2H), 1.63 (ddd, J = 13.8, 10.9, 7.0 Hz, 2H), 1.19 – 1.16 (m, 3H).

Step 2: UBI-1093c

Compound **UBI-1093b** (350 mg, 1.43 mmol), di-tert-butyl dicarbonate (469 mg, 2.15 mmol) and sodium bicarbonate (360 mg, 4.29 mmol) were added to tetrahydrofuran (20 mL) successively and reacted at room temperature for 2 hours. After the completion of reaction, the reaction was poured into 10 mL of water and extracted with dichloromethane (5 mL*3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na_2SO_4 , and concentrated by rotary evaporation under reduced pressure to obtain compound **UBI-1093c** (320 mg, yield 71%) as colorless oil.

Step 3: UBI-1093d

Compound **UBI-1093c** (300 mg, 0.87 mmol) and sodium hydroxide (69.6 mg, 1.74 mmol) were added to a system of water/methanol/tetrahydrofuran (0.5 mL/3 mL/3 mL), and reacted at room temperature for 16 hours. After the completion of reaction, the organic solvent was removed by concentration, and the aqueous phase was acidified to pH=5 using hydrochloric acid (1M). Then it was extracted with dichloromethane (10 mL*3), the combined organic layers were dried over anhydrous Na_2SO_4 , and concentrated to obtain desired compound **UBI-1093d** (250 mg, yield 91%) as colorless oil.

^1H NMR (400 MHz, d_6 -DMSO) δ 12.03 (s, 1H), 3.67 – 3.51 (m, 4H), 3.51 – 3.30 (m, 4H), 3.24 (dt, J = 12.6, 6.0 Hz, 2H), 2.16 (t, J = 7.2 Hz, 2H), 1.73 – 1.69 (m, 2H), 1.39 (s, 9H).

Step 4: UBI-1093e

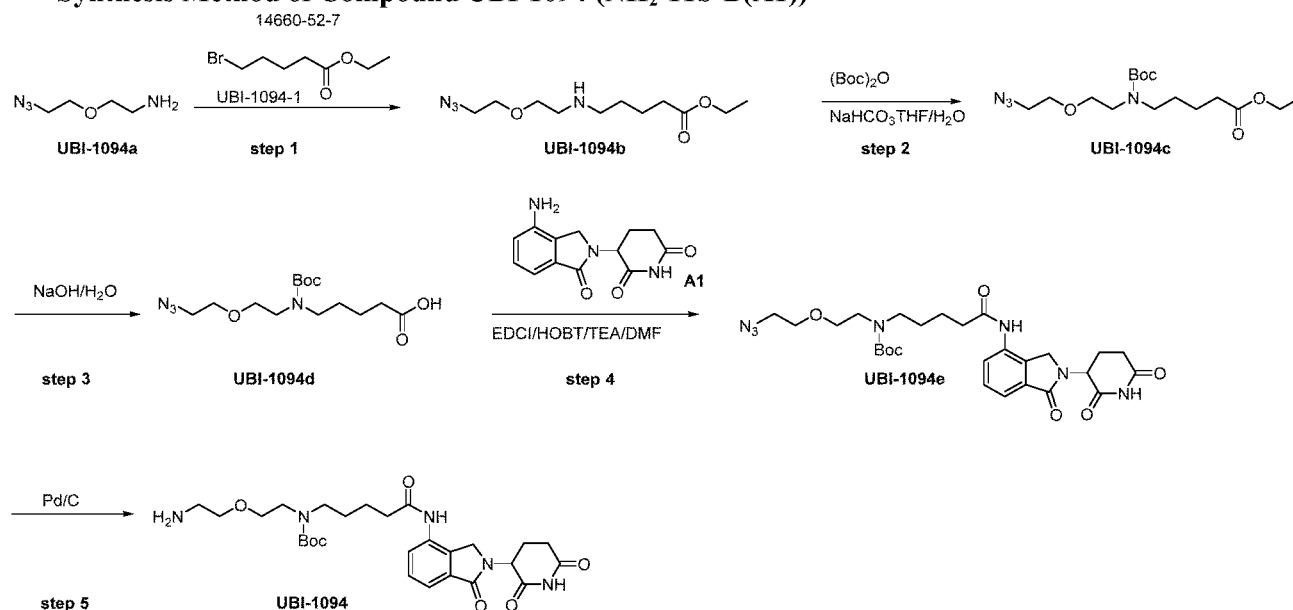
Compound **UBI-1093d** (254 mg, 0.80 mmol), **B(A1)** (230 mg, 0.88 mmol), HATU (337 mg, 0.97) and DIPEA (0.3 mL) were added to anhydrous DMF (3 mL), reacted at room temperature for 16 hours. After the completion of reaction, the reaction was concentrated. The crude was isolated by silica gel column

chromatography (MeOH/DCM = 1% to 10%) to obtain compound **UBI-1158c** (220mg, yield 49%) as colorless oil.

Step 4: UBI-1093

Compound **UBI-1093e** (200 mg, 0.35 mmol), 10% palladium on carbon (20 mg) was added to a mixed solvent of methanol/dichloromethane (1 mL/10 mL), and reacted at room temperature for 2 hours under hydrogen atmosphere. After filtration, the filtrate was concentrated to obtain the crude product. The crude product was washed with cold ether (10 mL*3), and dried to obtain the compound **UBI-1093** (170 mg, yield 89.4%) as colorless oil.

Synthesis Method of Compound UBI-1094 (NH₂-11b-B(A1))



Step 1: UBI-1094b

Compound **UBI-1094a** (1.0g, 7.69 mmol), **UBI-1094-1** (1.6g, 7.69 mmol) and triethylamine (2.02g, 20mmol) were added to anhydrous toluene (10 mL). The mixture was reacted at 80 °C for 18 hours. After the completion of reaction, the reaction solution was concentrated. The crude was isolated by silica gel column chromatography (DCM/MeOH = 10/1) to obtain compound **UBI-1094b** (740 mg, yield 37 %) as colorless oil.

Step 2: UBI-1094c

Compound **UBI-1094b** (350 mg, 1.35 mmol), di-tert-butyl dicarbonate (441 mg, 2.03 mmol) and sodium bicarbonate (360 mg, 4.29 mmol) were added to dioxane (20mL) successively, reacted at room temperature for 2 hours. After the completion of reaction, the reaction solution was poured into 10 mL of water, extracted with ethyl acetate (10 mL*3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na₂SO₄, and concentrated by rotary evaporation under reduced pressure to obtain compound **UBI-1094c** (390 mg, yield 87%) as colorless oil.

Step 3: UBI-1094d

Compound **UBI-1094c** (390 mg, 1.09 mmol), sodium hydroxide (1.19g, 21.2mmol) were added to water (25 mL) successively, reacted at 30°C for 12 hours. After the completion of reaction, it was concentrated and the aqueous phase was acidified to pH=5 using hydrochloric acid (1M). Then it was extracted with dichloromethane (10ml*3), the combined organic layers was dried over anhydrous Na₂SO₄, and concentrated to obtain target compound **UBI-1094d** (250 mg, yield 91%).

¹H NMR (400 MHz, d₆-DMSO) δ 11.99 (s, 1H), 3.69 – 3.48 (m, 4H), 3.39 – 3.27 (m, 6H), 2.22 (t, *J* = 6.9 Hz, 2H), 1.47 (s, 4H), 1.36 (s, 9H).

Step 4: UBI-1094e

Compound **UBI-1093d** (210 mg, 0.665 mmol), **B(A1)** (160 mg, 0.618 mmol), HATU (305 mg, 0.803mmol) and DIPEA (0.5mL) were added to anhydrous DMF (5mL), and reacted at room temperature for 16 hours, concentrated after the completion of reaction. The crude product was isolated by reverse chromatography on silica gel column chromatography (DCM/MeOH=10/1) to obtain compound **UBI-1093e** (157 mg, 43% yield) as white solid.

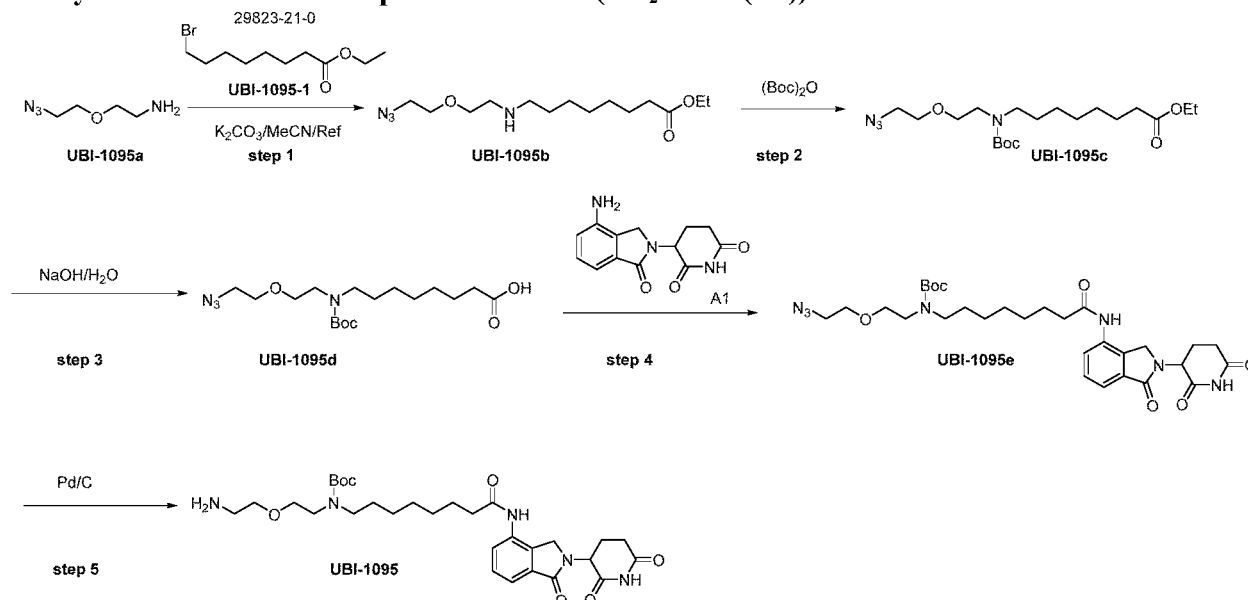
LC-MS: $[M-100+H]^+ = 472.3$

Step 5: UBI-1094

Compound **UBI-1094e** (100 mg, 0.17mmol), 10% palladium on carbon (20 mg) was added to a mixed solvent of methanol/dichloromethane (1 mL/10 mL), and reacted at room temperature for 2 hours under hydrogen atmosphere. The reaction solution was filtrated, and the filtrate was concentrated to obtain the crude product. The crude product was washed with cold ether (10 mL*3), and dried to obtain target compound **UBI-1094** (81 mg, yield 85%) as colorless oil.

LCMS: $(M-100+H)^+ = 446.3$

Synthesis Method of Compound UBI-1095 (NH₂-14b-B(A1))



Step 1: UBI-1095b

Compound **UBI-1095a** (2g, 15.3mmol), **UBI-1095-1** (3.80g, 15.3mmol) and triethylamine (2.02g, 20mmol) were added to anhydrous toluene (100 mL), and reacted at 80 °C for 18 hours, concentrated after the completion of reaction. The crude product was isolated by silica gel column chromatography (DCM/MeOH = 10/1) to obtain compound **UBI-1095b** (740 mg, yield 16 %) as colorless oil.

Step 2: UBI-1095c

Compound **UBI-1095b** (1.3g, 3.7 mmol), di-tert-butyl dicarbonate (2.3g, 10.7mmol) and sodium bicarbonate (360 mg, 4.29mmol) were added to dioxane (30mL) successively and reacted at room temperature for 2 hours. After the completion of the reaction, the reaction solution was poured into 10 mL of water and extracted with ethyl acetate (10 mL*3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na₂SO₄, and concentrated by rotary evaporation under reduced pressure to obtain compound **UBI-1095c** (1.4g, yield 83%).

¹H NMR (400 MHz, d₆-DMSO) δ 12.03 (s, 1H), 3.63 – 3.51 (m, 4H), 3.45 – 3.41 (m, 2H), 3.32-3.29 (m,

2H), 3.24 (dt, $J = 12.6, 6.0$ Hz, 2H), 2.16 (t, $J = 7.2$ Hz, 2H), 1.54 (d, $J = 12.2$ Hz, 4H), 1.39 (s, 9H), 1.35 – 1.28 (m, 6H).

Step 3: UBI-1095d

Compound **UBI-1095c** (1.5g, 6.0 mmol), sodium hydroxide (1.19g, 21.2mmol) were added to water (50 mL) successively, reacted at 30°C for 12 hours. After the completion of reaction, it was concentrated and the aqueous phase was acidified to pH=5 using hydrochloric acid (1M). Then it was extracted with dichloromethane (10ml*3), the combined organic layer was dried over anhydrous Na_2SO_4 , and concentrated to obtain the desired compound **UBI-1095d** (1.2 g, yield 83%).

Step 4: UBI-1095e

Compound **UBI-1095d** (600 mg, 1.57 mmol), **B(A1)** (406 mg, 1.57 mmol), HATU (1.51g, 3.98 mmol) and DIPEA (0.5mL) were added to anhydrous DMF (5 mL), and reacted at room temperature for 16 hours, concentrated after the completion of reaction. The crude product was isolated by reverse chromatography on silica gel column chromatography (DCM/MeOH=10/1) to obtain compound **UBI-1095e** (500 mg, yield 51%) as colorless oil.

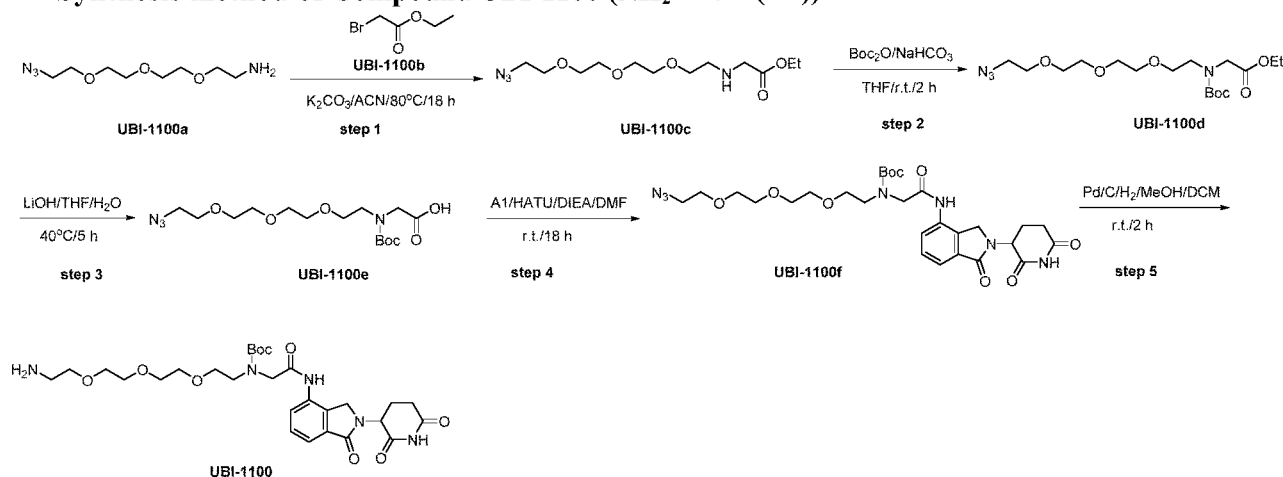
LC-MS: $[\text{M}-100+\text{H}]^+ = 515.3$

Step 5: UBI-1095

Compound **UBI-1095e** (100 mg, 0.17mmol), 10% palladium on carbon (20 mg) was added to a mixed solvent of methanol/dichloromethane (1 mL/10 mL), and reacted at room temperature for 16 hours under hydrogen atmosphere. After filtration, the filtrate was concentrated to obtain the crude product. The crude product was washed with cold ether (10 mL*3), and dried to obtain target compound **UBI-1095** (83 mg, yield 86%) as white solid.

LCMS: $(\text{M}-100+\text{H})^+ = 489.3$

Synthesis method of Compound UBI-1100 (NH_2 -14b-B(A1))



Step 1: UBI-1100c

Compound **UBI-1100a** (1.0 g, 4.58 mmol) was dissolved in acetonitrile (30 mL), added anhydrous potassium carbonate (949.8 mg, 6.87 mmol) and **UBI-1100b** (765.2 mg, 4.58 mmol), and then heated to 80 °C reacted for 18 hours. The reaction was cooled to room temperature and filtered. The filtrate was concentrated and subjected to silica gel column chromatography (petroleum ether/ethyl acetate = 50% to 100%, then dichloromethane /methanol = 0% to 10%) to obtain product **UBI-1100c** (0.6 g, yield 43%) as white solid.

Step 2: UBI-1100d

Compound **UBI-1100c** (0.6 g, 1.97 mmol) was dissolved in tetrahydrofuran (20 mL), added sodium

bicarbonate (331.2 mg, 3.94 mmol) and Boc_2O (0.64 g, 2.92 mmol). The reaction was reacted at room temperature for 2 hours. The reaction was quenched with water (20 mL), then extracted with ethyl acetate (20 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate and concentrated to obtain crude product **UBI-1100d** (**V1084-016**, 400 mg, yield 50.2%) as yellow oil. LCMS $[\text{M}-100]^+ = 305.1$

Step 3: UBI-1100e

Compound **UBI-1100d** (400 mg, 1 mmol), lithium hydroxide (47 mg, 1.98 mmol) were dissolved in tetrahydrofuran (20 mL) and water (10 mL). The system was reacted at 40°C for 5 hours. After complete hydrolysis, the reaction was added water (15 mL), extracted with ethyl acetate (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain product **UBI-1100e** (**V1084-019**, 320 mg, yield 86%) as colorless oil. ^1H NMR (400 MHz, chloroform-*d*) δ 4.00 (s, 2H), 3.72 – 3.63 (m, 10H), 3.52 (s, 2H), 3.42 (d, $J = 5.2$ Hz, 2H), 1.46 (d, $J = 10.2$ Hz, 11H). LCMS $[\text{M}-100]^+ = 277.2$

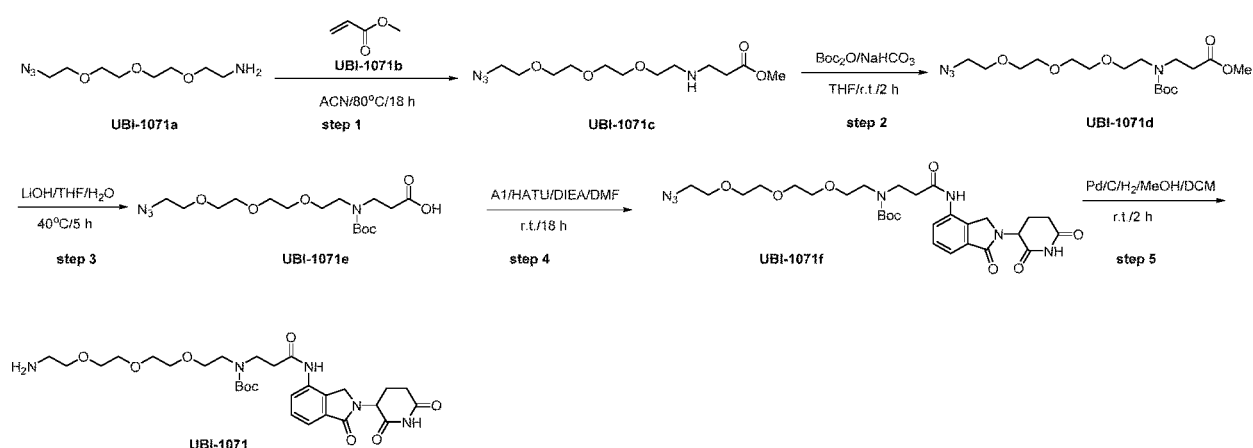
Step 4: UBI-1100f

Compound **UBI-1100e** (200 mg, 0.53 mmol), B(A1) (137.7 mg, 0.53 mmol), HATU (303 mg, 0.8 mmol), DIPEA (0.5 mL) were dissolved in DMF (5 mL) and reacted at room temperature overnight. After the completion of reaction, it was purified by reversed-phase column (MeOH/H₂O = 5 % to 95 %, 45 minutes, collected at 70%). After purification, product **UBI-1100f** (**V1084-035**, 180 mg, yield 54.9%) was obtained as yellow oil. ^1H NMR (400 MHz, chloroform-*d*) δ 9.09 (s, 1H), 7.99 (s, 1H), 7.72 (s, 2H), 7.48 (t, $J = 7.7$ Hz, 1H), 5.22 (dd, $J = 13.2, 5.2$ Hz, 1H), 4.41 (s, 2H), 3.62 (d, $J = 35.7$ Hz, 12H), 3.33 (s, 2H), 2.95 – 2.80 (m, 2H), 2.42 – 2.31 (m, 1H), 2.27 – 2.21 (m, 1H), 1.47 (s, 9H). LCMS $[\text{M}-100]^+ = 518.1$

Step 5: UBI-1100

Compound **UBI-1100e** (100 mg, 0.16 mmol) was dissolved in dichloromethane (10 mL) and methanol (1 mL), added Pd/C (20 mg), and reacted out at room temperature for 2 hours under hydrogen atmosphere. The reaction solution was filtered on Celite. The filtrate was concentrated to obtain product **UBI-1100f** (130 mg, yield 100%) as yellow oil. LCMS $[\text{M}+\text{H}]^+ = 592.2$

Synthesis method of Compound UBI-1071 (NH₂-15b-B(A1))



Step 1: UBI-1071c

Compound **UBI-1071a** (1.0 g, 4.58 mmol) and **UBI-1071b** (0.39 g, 4.58 mmol) were dissolved in acetonitrile (30 mL), and then heated to 80 °C and reacted for 18 hours. The reaction solution was concentrated and subjected to silica gel column chromatography (petroleum ether/ethyl acetate = 50% to 100%, then dichloromethane/methanol = 0% to 10%) to obtain product **UBI-1071c** (0.4 g, yield 28.7%) as

yellow oil.

^1H NMR (400 MHz, DMSO-*d*₆) δ 3.63 - 3.47 (m, 13H), 3.44 (t, J = 5.6 Hz, 2H), 3.41 - 3.36 (m, 2H), 2.76 (t, J = 6.8 Hz, 2H), 2.64 (t, J = 5.6 Hz, 2H), 2.44 (t, J = 6.8 Hz, 2H).

Step 2: UBI-1071d

Compound **UBI-1071c** (0.4 g, 1.31 mmol) was dissolved in tetrahydrofuran (20 mL), added sodium bicarbonate (220.8 mg, 2.63 mmol) and Boc₂O (430 mg, 1.97 mmol) and reacted at room temperature for 2 hours. The reaction was quenched with water (20 mL), extracted with ethyl acetate (20 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate and concentrated to obtain crude product **UBI-1071d** (0.42 g, yield 79%) as yellow oil. LCMS $[\text{M}-100]^+ = 305.1$

Step 3: UBI-1071e

Compound **UBI-1071d** (0.4 g, 0.99 mmol), lithium hydroxide (47.8 mg, 1.98 mmol) were dissolved in tetrahydrofuran (20 mL) and water (10 mL). The system was reacted at 40°C for 5 hours. After complete hydrolysis, the reaction was added water (30 mL), extracted impurities using ethyl acetate (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain product **UBI-1071e** (**V1084-010**, 320 mg, yield 82.9%) as colorless oil. ^1H NMR (400 MHz, DMSO-*d*₆) δ 12.15 (s, 1H), 3.63 - 3.58 (m, 2H), 3.58 - 3.49 (m, 8H), 3.46 (s, 2H), 3.42 - 3.34 (m, 4H), 3.32 - 3.27 (m, 2H), 2.45 (t, J = 7.2 Hz, 2H), 1.39 (s, 9H). LCMS $[\text{M}-100]^+ = 291.2$

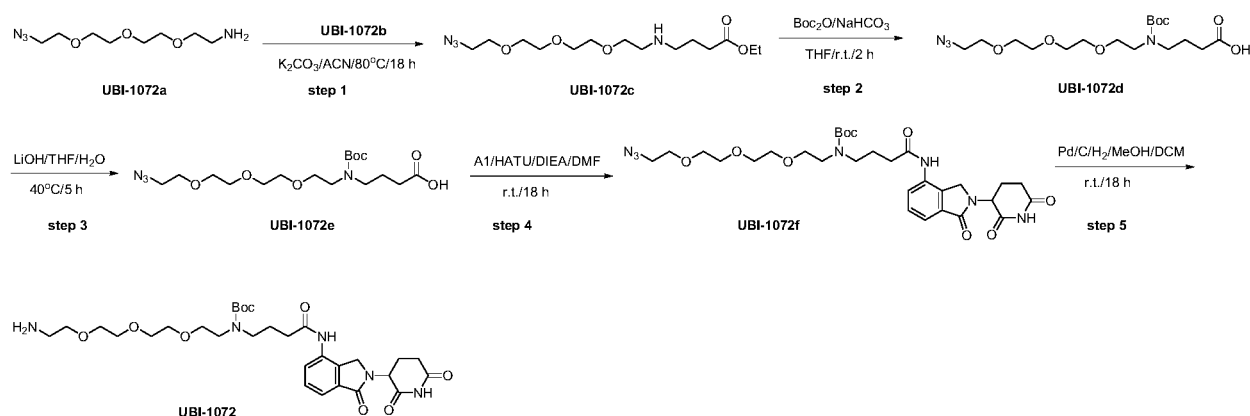
Step 4: UBI-1071f

Compound **UBI-1071e** (200 mg, 0.51 mmol), B(A1) (132.8 mg, 0.51 mmol), HATU (292.2 mg, 0.77 mmol), DIPEA (0.5 mL) were dissolved in DMF (5 mL) and reacted at room temperature overnight. After the completion of reaction, it was purified by reversed-phase column (MeOH/H₂O = 5 % to 95 %, 45 minutes, collected at 70%). After purification, product **UBI-1071f** (150 mg, yield 46.7%) was obtained as yellow oil. ^1H NMR (400 MHz, DMSO-*d*₆) δ 11.02 (s, 1H), 9.83 (d, J = 14.0 Hz, 1H), 7.90 - 7.75 (m, 1H), 7.57 - 7.37 (m, 2H), 5.21 - 5.10 (m, 1H), 4.47 - 4.26 (m, 2H), 3.55 - 3.47 (m, 16H), 3.31 - 3.26 (m, 2H), 3.00 - 2.85 (m, 1H), 2.66 - 2.56 (m, 3H), 2.38 - 2.27 (m, 1H), 2.08 - 1.98 (m, 1H), 1.38 (s, 9H). LCMS $[\text{M}-100]^+ = 532.2$

Step 5: UBI-1071

Compound **UBI-1071e** (100 mg, 0.74 mmol) was dissolved in a mixed solvent of dichloromethane (10 mL) and methanol (1 mL), added Pd/C (20 mg), and reacted out at room temperature for 2 hours under hydrogen atmosphere. The reaction solution was filtered on Celite. The filtrate was concentrated to obtain product **UBI-1071f** (130 mg, yield 86.5%) as yellow oil. LCMS $[\text{M}+\text{H}]^+ = 606.2$.

Synthesis method of Compound UBI-1072 (NH₂-16b-B(A1))



Step 1: UBI-1072c

Compound **UBI-1072a** (1.0 g, 4.58 mmol) was dissolved in acetonitrile (30 mL), added anhydrous potassium carbonate (0.95 g, 6.87 mmol) and **UBI-1160b** (0.89 g, 4.58 mmol), and then heated to 80 °C reacted for 18 hours. The reaction was cooled to room temperature and filtered. The filtrate was concentrated and subjected to silica gel column chromatography (petroleum ether/ethyl acetate = 50% to 100%, then dichloromethane /methanol = 0% to 10%) to obtain product **UBI-1072c** (530 mg, yield 34.8%) as yellow oil.

¹H NMR (400 MHz, chloroform-*d*) δ 5.83 (s, 1H), 4.14 (d, *J* = 7.1 Hz, 2H), 3.89 – 3.81 (m, 2H), 3.69 (d, *J* = 3.1 Hz, 10H), 3.43 (dd, *J* = 5.6, 4.4Hz, 2H), 3.16 – 3.09 (m, 2H), 3.03 (s, 2H), 2.47 (t, *J* = 7.0 Hz, 2H), 2.11 (s, 2H), 1.26 (t, *J* = 7.1 Hz, 3H).

Step 2: UBI-1072d

Compound **UBI-1072c** (0.5 g, 1.5 mmol) was dissolved in tetrahydrofuran (20 mL), added sodium bicarbonate (252.7 mg, 3.01 mmol) and Boc₂O (492.4 mg, 2.26 mmol). The reaction was reacted at room temperature for 2 hours. The reaction was quenched with water (20 mL), extracted with ethyl acetate (20 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate and concentrated to obtain crude product **UBI-1072d** (0.52 g, yield 80%) as yellow oil. LCMS [M-100]⁺ = 233.1

Step 3: UBI-1072e

Compound **UBI-1072d** (400 mg, 0.93 mmol), lithium hydroxide (44.3 mg, 1.85 mmol) were dissolved in tetrahydrofuran (20 mL) and water (10 mL). The system was reacted at 40°C for 5 hours. After complete hydrolysis, the reaction was added water (15 mL), extracted impurities using ethyl acetate (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain product **UBI-1072e** (300 mg, yield 80.2%) as colorless oil. ¹H NMR (400 MHz, chloroform-*d*) δ 3.67 (d, *J* = 5.0 Hz, 8H), 3.64 – 3.60 (m, 4H), 3.40 (d, *J* = 5.2 Hz, 2H), 3.35 – 3.30 (m, 2H), 2.34 (t, *J* = 7.2 Hz, 2H), 1.85 (q, *J* = 7.2 Hz, 2H), 1.45 (s, 11H). LCMS [M-100]⁺ = 305.2

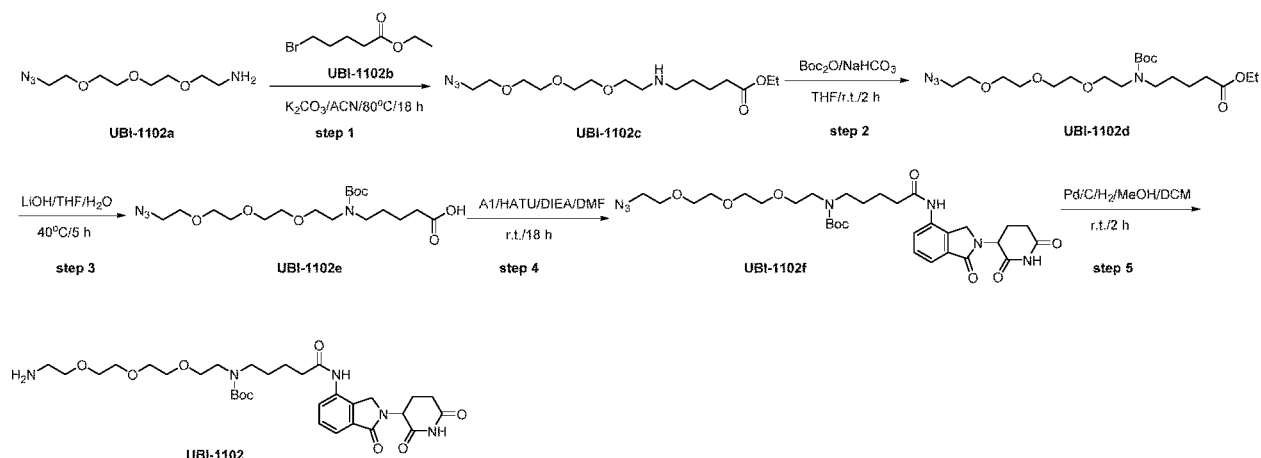
Step 4: UBI-1072f

Compound **UBI-1072e** (200 mg, 0.49 mmol), **B(A1)** (128.2 mg, 0.49 mmol), HATU (282 mg, 0.74 mmol), and DIPEA (0.5 mL) were dissolved in DMF (5 mL), reacted at room temperature overnight. After the completion of reaction, it was purified by reversed-phase column (MeOH/H₂O = 5 % to 95 %, 45 minutes, collected at 70%). After purification, product **UBI-1072f** (150 mg, yield 47%) was obtained as yellow oil. LCMS [M-100]⁺ = 546.1

Step5: UBI-1072

Compound **UBI-1072e** (100 mg, 0.15 mmol) was dissolved in a mixed solvent of dichloromethane (10 mL) and methanol (1 mL), added Pd/C (20 mg), and reacted at room temperature for 2 hours under hydrogen atmosphere. The reaction solution was filtered on Celite. The filtrate was concentrated to obtain product **UBI-1072** (80 mg, yield 55.6%) as yellow oil. LCMS [M+H]⁺ = 620.2

Synthesis method of Compound UBI-1102 (NH₂-17b-B(A1))



Step 1: UBI-1102c

Compound **UBI-1102a** (1.0 g, 4.58 mmol) was dissolved in acetonitrile (30 mL), added anhydrous potassium carbonate (950 mg, 6.87 mmol) and **UBI-1102b** (0.88 g, 1.92 mmol), and then heated to 80 °C reacted for 18 hours. The reaction was cooled to room temperature and filtered. The filtrate was concentrated and subjected to silica gel column chromatography (petroleum ether/ethyl acetate = 50% to 100%, then dichloromethane /methanol = 0% to 10%) to obtain product **UBI-1102c** (0.5 g, yield 25%) as yellow oil.

¹H NMR (400 MHz, chloroform-*d*) δ 4.12 (q, *J* = 7.1 Hz, 2H), 3.71 – 3.60 (m, 11H), 3.40 (t, *J* = 5.1 Hz, 2H), 2.86 – 2.77 (m, 2H), 2.70 – 2.62 (m, 2H), 2.32 (t, *J* = 7.4 Hz, 2H), 2.17 (s, 2H), 1.73 – 1.50 (m, 4H), 1.25 (t, *J* = 7.1 Hz, 3H).

Step 2: UBI-1102d

Compound **UBI-1102c** (0.5 g, 1.44 mmol) was dissolved in tetrahydrofuran (30 mL) and added sodium bicarbonate (121 mg, 2.16 mmol) and Boc₂O (0.47 g, 2.17 mmol). The reaction was reacted at room temperature for 2 hours. The reaction was quenched with water (20 mL), extracted with ethyl acetate (20 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate and concentrated to obtain crude product **UBI-1102d** (0.45 g, yield 60.7%) as yellow oil.

Step 3: UBI-1102e

Compound **UBI-1102d** (0.4 g, 0.9 mmol), lithium hydroxide (40 mg, 1.79 mmol) were dissolved in tetrahydrofuran (20 mL) and water (10 mL). The system was reacted at 40°C for 5 hours. After complete hydrolysis, the reaction was added water (15 mL), extracted impurities using ethyl acetate (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain product **UBI-1102e** (300 mg, yield 83.6%) as colorless oil.

¹H NMR (400 MHz, chloroform-*d*) δ 10.30 (s, 1H), 3.71 – 3.55 (m, 12H), 3.38 (q, *J* = 11.2, 8.1 Hz, 4H), 3.26 (s, 2H), 2.41 – 2.32 (m, 2H), 1.65 – 1.56 (m, 4H), 1.45 (d, *J* = 1.7 Hz, 9H).

Step 4: UBI-1102f

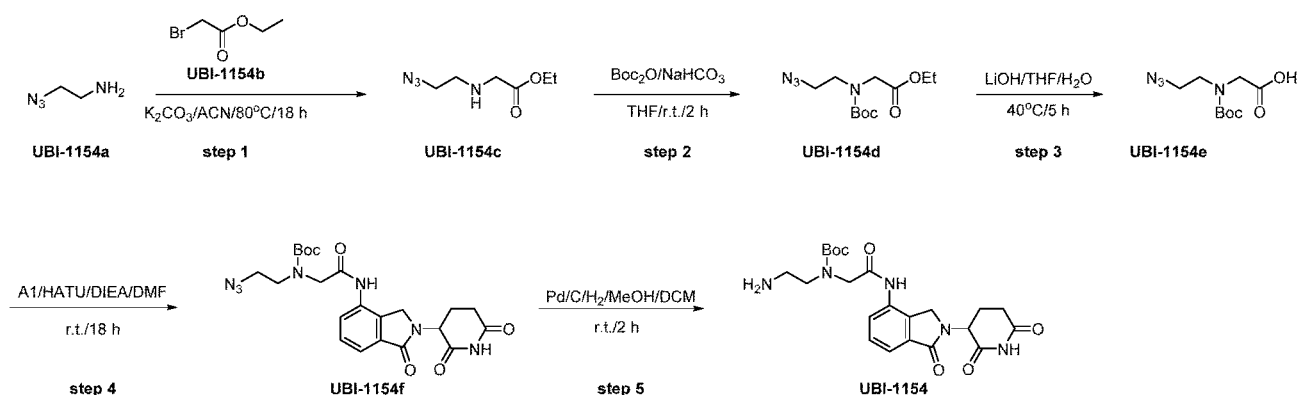
Compound **UBI-1102e** (300 mg, 0.72 mmol), **B(A1)** (150 mg, 0.57 mmol), HATU (408.9 mg, 1.08 mmol), and DIPEA (0.5 mL) were dissolved in DMF (5 mL), reacted at room temperature overnight. After the completion of reaction, it was purified by reversed-phase column (MeOH/H₂O = 5 % to 95 %, 45 minutes, collected at 70%). After purification, product **UBI-1102f** (135 mg, yield 35.2%) was obtained as yellow oil. LCMS [M-100]⁺ = 574

Step 5: UBI-1102

Dissolving in a mixed solvent of dichloromethane (30 mL) and methanol (3 mL), adding Pd/C (50 mg), and reacting at room temperature for 2 hours under hydrogen atmosphere. The reaction solution was filtered

on Celite. The filtrate was concentrated to obtain product **UBI-1102** (130 mg, yield 89.8%) as yellow oil. LCMS $[M+H]^+ = 648$

Synthesis method of Compound **UBI-1154** (**NH₂-5b-B(A1)**)



Step 1: **UBI-1154c**

Compound **UBI-1154a** (1.0 g, 11.63 mmol) was dissolved in acetonitrile (30 mL), added anhydrous potassium carbonate (2.41 g, 17.44 mmol) and **UBI-1154b** (1.93 g, 11.63 mmol), and then heated to 80 °C reacted for 18 hours. The reaction was cooled to room temperature and filtered. The filtrate was concentrated and subjected to silica gel column chromatography (petroleum ether/ethyl acetate = 50% to 100%, then dichloromethane /methanol = 0% to 10%) to obtain product **UBI-1154c** (500 mg, yield 25%) as yellow oil.

Step 2: **UBI-1154d**

Compound **UBI-1154c** (500 mg, 2.91 mmol) was dissolved in tetrahydrofuran (20 mL) and added sodium bicarbonate (731.8mg, 8.72 mmol) and Boc_2O (1.27 g, 5.81 mmol). The reaction was reacted at room temperature for 2 hours. The reaction was quenched with water (20 mL), extracted with ethyl acetate (20 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate and concentrated to obtain crude product **UBI-1154d** (0.48 g, yield 60.7%) as yellow oil. LCMS $[M-100]^+ = 173.1$

Step 3: **UBI-1154e**

Compound **UBI-1154d** (0.4 g, 1.47 mmol), lithium hydroxide (70 mg, 2.94 mmol) were dissolved in tetrahydrofuran (20 mL) and water (10 mL). The system was reacted at 40 °C for 5 hours. After complete hydrolysis, the reaction was added water (30 mL), extracted impurities using ethyl acetate (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain product **UBI-1154e** (230 mg, yield 83.6%) as colorless oil. ^1H NMR (400 MHz, chloroform-*d*) δ 4.03 (s, 2H), 3.45 (d, $J = 4.1$ Hz, 4H), 1.44 (s, 9H).

Step 4: **UBI-1154f**

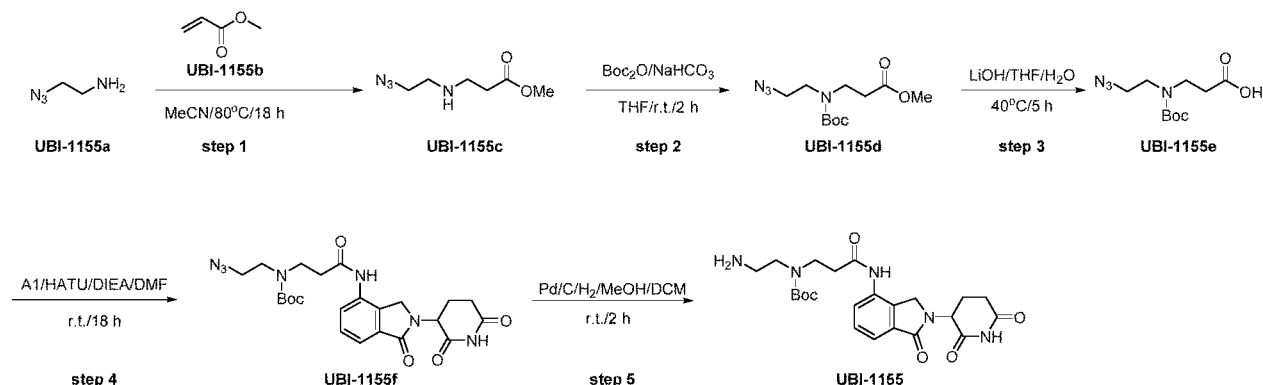
Compound **UBI-1154e** (200 mg, 0.82 mmol), **B(A1)** (212.3 mg, 0.82 mmol), HATU (467 mg, 0.13 mmol), and DIPEA (0.5 mL) were dissolved in DMF (5 mL), reacted at room temperature overnight. After the completion of reaction, it was purified by reversed-phase column (MeOH/H₂O = 5 % to 95 %, 45 minutes, collected at 70%). After purification, product **UBI-1154f** (140 mg, yield 35.2%) was obtained as yellow oil. LCMS $[M+H]^+ = 486.1$

Step 5: **UBI-1154**

Compound **UBI-1154e** (100 mg, 0.21 mmol) was dissolved in a mixed solvent of dichloromethane (10 mL) and methanol (1 mL), added Pd/C (20 mg), and reacted at room temperature for 2 hours under hydrogen atmosphere. The reaction solution was filtered on Celite. The filtrate was concentrated to obtain

crude product **UBI-1154f** (85 mg, yield 89.8%) as yellow oil. LCMS $[M+H]^+ = 460.2$

Synthesis method of Compound **UBI-1155** (**NH₂-6b-B(A1)**)



5 **Step 1: UBI-1155c**

Compound **UBI-1155a** (1.0 g, 11.63 mmol) and **UBI-1155b** (1.0 g, 11.63 mmol) were dissolved in acetonitrile (30 mL), and then heated to 80 °C reacted for 18 hours. The reaction solution was concentrated and subjected to silica gel column chromatography (petroleum ether/ethyl acetate = 50% to 100%, then dichloromethane/methanol = 0% to 10%) to obtain product **UBI-1071c** (0.4 g, yield 30%) as yellow oil.

10 **Step 2: UBI-1155d**

Compound **UBI-1155c** (500 mg, 2.90 mmol) was dissolved in tetrahydrofuran (20 mL), added sodium bicarbonate (732 mg, 8.71 mmol) and Boc_2O (1.27 g, 5.81 mmol), and reacted at room temperature for 2 hours. The reaction was quenched with water (20 mL), extracted with ethyl acetate (20 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate and concentrated to obtain crude product **UBI-1155d** (500 mg, yield 44.3%) as yellow oil. ^1H NMR (400 MHz, chloroform-*d*) δ 3.69 (s, 3H), 3.54 (t, $J = 6.8$ Hz, 2H), 3.48 - 3.35 (m, 4H), 2.68 - 2.53 (m, 2H), 1.47 (s, 9H). LCMS $[M-100]^+ = 173.1$

15 **Step 3: UBI-1155e**

Compound **UBI-1155d** (500 mg, 1.84 mmol), lithium hydroxide (88.0 mg, 3.67 mmol) were dissolved in tetrahydrofuran (20 mL) and water (10 mL). The system was reacted at 40°C for 5 hours. After complete hydrolysis, the reaction was added water (30 mL), extracted impurities using ethyl acetate (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain product **UBI-1155e** (350 mg, yield 73.8%) as colorless oil. ^1H NMR (400 MHz, chloroform-*d*) δ 7.68 (s, 1H), 3.54 (t, $J = 6.9$ Hz, 2H), 3.49 - 3.30 (m, 4H), 2.65 (s, 2H), 1.47 (s, 9H). LCMS $[M-100]^+ = 159.1$

20 **Step 4: UBI-1155f**

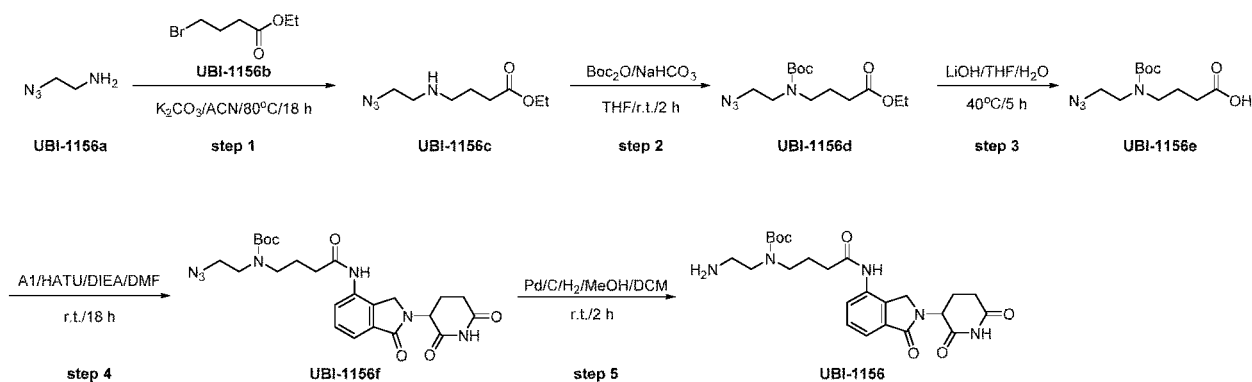
Compound **UBI-1155e** (200 mg, 0.74 mmol), **B(A1)** (183.9 mg, 0.74 mmol), HATU (419 mg, 1.10 mmol), and DIPEA (0.5 mL) were dissolved in DMF (5 mL), reacted at room temperature overnight. After the completion of reaction, it was purified by reversed-phase column (MeOH/H₂O = 5 % to 95 %, 45 minutes, collected at 70%). After purification, product **UBI-1155f** (**V1084-066**, 200 mg, yield 51.7%) was obtained as yellow oil. ^1H NMR (400 MHz, DMSO-*d*₆) δ 11.03 (s, 1H), 9.88 (s, 1H), 7.83 (s, 1H), 7.56 - 7.43 (m, 2H), 5.15 (dd, $J = 13.3, 5.1$ Hz, 1H), 4.45 - 4.28 (m, 2H), 3.50 (t, $J = 7.0$ Hz, 2H), 3.46 - 3.37 (m, 4H), 3.27 - 2.87 (m, 2H), 2.64 (t, $J = 6.9$ Hz, 2H), 2.39 - 2.00 (m, 2H), 1.40 (s, 9H). LCMS $[M+H]^+ = 400.2$

25 **Step 5: UBI-1155**

Compound **UBI-1155e** (200 mg, 0.74 mmol) was dissolved in a mixed solvent of dichloromethane (30 mL) and methanol (3 mL), added Pd/C (40 mg), and reacted at room temperature for 2 hours under

hydrogen atmosphere. The reaction solution was filtered on Celite. The filtrate was concentrated to obtain crude product **UBI-1155f** (130 mg, yield 68.6%) as yellow oil. LCMS $[M+H]^+ = 474.2$

Synthesis Method of Compound: **UBI-1156 (NH₂-7b-B(A1))**



Step 1: **UBI-1156c**

Compound **UBI-1156a** (1.0 g, 11.63 mmol) was dissolved in acetonitrile (30 mL), added anhydrous potassium carbonate (2.41 g, 17.44 mmol) and **UBI-1156b** (2.26 g, 11.63 mmol), and then heated to 80 °C reacted for 18 hours. The reaction was cooled to room temperature and filtered. The filtrate was concentrated and subjected to silica gel column chromatography (petroleum ether/ethyl acetate = 50% to 100%, then dichloromethane /methanol = 0% to 10%) to obtain product **UBI-1156c** (650 mg, yield 27%) as yellow oil. LCMS $[M+H]^+ = 201.1$

Step 2: **UBI-1156d**

Compound **UBI-1156c** (500 mg, 2.5 mmol) was dissolved in tetrahydrofuran (10 mL) and added sodium bicarbonate (731.8 mg, 7.50 mmol) and Boc_2O (0.47 g, 5.0 mmol). The reaction was reacted at room temperature for 2 hours. The reaction was quenched with water (40 mL), extracted with ethyl acetate (20 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate and concentrated to obtain crude product **UBI-1156d** (500 mg, yield 66.7%) as yellow oil.

^1H NMR (400 MHz, chloroform-*d*) δ 4.14 (q, $J = 7.1$ Hz, 2H), 3.48 – 3.32 (m, 4H), 3.29 (t, $J = 7.3$ Hz, 2H), 2.31 (t, $J = 7.3$ Hz, 2H), 1.86 (p, $J = 7.3$ Hz, 2H), 1.47 (s, 9H), 1.26 (t, $J = 7.1$ Hz, 3H).

Step 3: **UBI-1156e**

Compound **UBI-1156d** (500 mg, 1.66 mmol), lithium hydroxide (79.73 mg, 3.33 mmol) were dissolved in tetrahydrofuran (20 mL) and water (10 mL). The system was reacted at 40 °C for 5 hours. After complete hydrolysis, the reaction was added water (30 mL), extracted impurities using ethyl acetate (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain product **UBI-1156e** (400 mg, yield 88.2%) as colorless oil. LCMS $[M+H]^+ = 173.2$

^1H NMR (400 MHz, DMSO-*d*₆) δ 3.40 (dd, $J = 6.3, 4.7$ Hz, 2H), 3.33 (dd, $J = 7.1, 5.5$ Hz, 2H), 3.18 (t, $J = 7.3$ Hz, 2H), 2.16 (t, $J = 7.3$ Hz, 2H), 1.68 (p, $J = 7.3$ Hz, 2H), 1.40 (s, 9H).

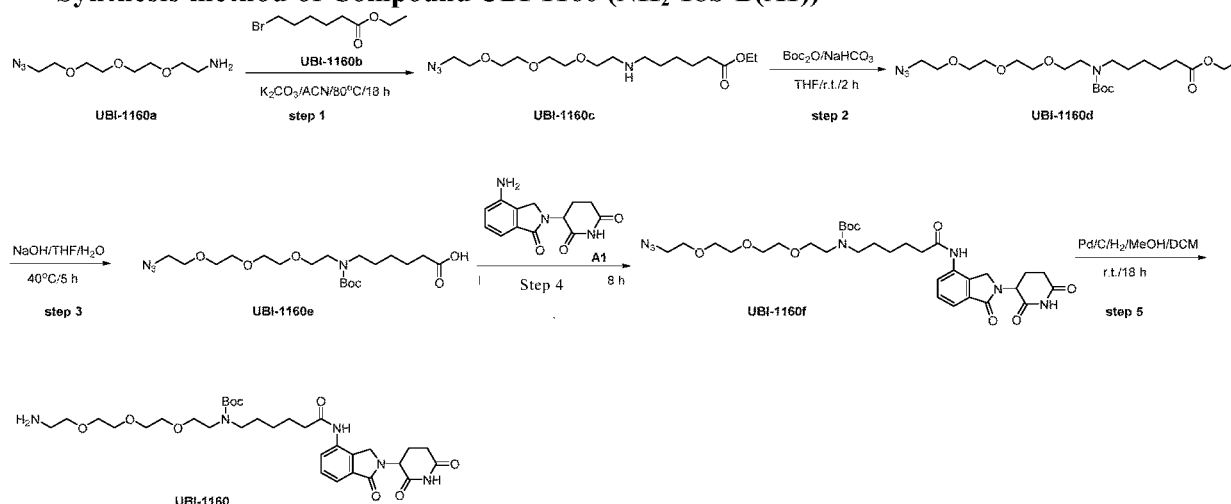
Step 4: **UBI-1156f**

Compound **UBI-1156e** (200 mg, 0.735 mmol), **B(A1)** (191.2 mg, 0.735 mmol), HATU (419.12 mg, 1.1 mmol), and DIPEA (0.5 mL) were dissolved in DMF (5 mL), reacted at room temperature overnight. After the completion of reaction, it was purified by reversed-phase column (MeOH/H₂O = 5 % to 95 %, 45 minutes, collected at 70%). After purification, product **UBI-1156f** (100 mg, yield 26.5%) was obtained as yellow oil. LCMS $[M-100]^+ = 414.1$

Step 5: **UBI-1156**

Compound **UBI-1156e** (100 mg, 0.20 mmol) was dissolved in a mixed solvent of dichloromethane (20 mL) and methanol (2 mL), added Pd/C (20 mg), and reacted at room temperature for 2 hours under hydrogen atmosphere. The reaction solution was filtered on Celite. The filtrate was concentrated to obtain product **UBI-1156f** (100 mg, yield 80.3%) as yellow oil. LCMS $[M+H]^+ = 488.1$

Synthesis method of Compound **UBI-1160** (**NH₂-18b-B(A1)**)



Step 1: **UBI-1160c**

Compound **UBI-1160a** (1.5 g, 6.87 mmol) was dissolved in acetonitrile (30 mL), added anhydrous potassium carbonate (1.9 g, 6.87 mmol) and **UBI-1160b** (1.53 g, 6.87 mmol), reacted at 80 °C overnight. The reaction was cooled to room temperature and filtered. The filtrate was concentrated and isolated by silica gel column chromatography (dichloromethane /methanol = 95/5) to obtain product **UBI-1160c** (1 g, yield 40%) as white solid.

Step 2: **UBI-1160d**

Compound **UBI-1160c** (1 g, 2.77 mmol) was dissolved in tetrahydrofuran (30 mL), added NaHCO₃ (700 mg, 8.31 mmol) and Boc₂O (0.72 g, 3.33 mmol), and reacted at room temperature for 2 hours. The reaction was quenched with water (20 mL), extracted with ethyl acetate (20 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 90/10) to obtain product **UBI-1160d** (2 g, yield 100%) as yellow oil.

Step 3: **UBI-1160e**

Compound **UBI-1160d** (1.27 g, 2.77 mmol) was dissolved in tetrahydrofuran (20 mL) and water (10 mL), added NaOH (332 mg, 8.31 mmol), and reacted at 40°C for 5 hours. The reaction solution was concentrated, and added saturated brine (15 mL) and extracted with ether (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain crude product **UBI-1160e** (500 mg, yield 42%) as colorless oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.97 (s, 1H), 3.60 (m, 2H), 3.51-3.49 (m, 8H), 3.46 (m, 2H), 3.40 (m, 2H), 3.25 (m, 2H), 3.16-3.12 (m, 2H), 2.17-2.19 (m, 2H), 1.54-1.35 (m, 13H), 1.16-1.21 (m, 2H).

Step 4: **UBI-1160f**

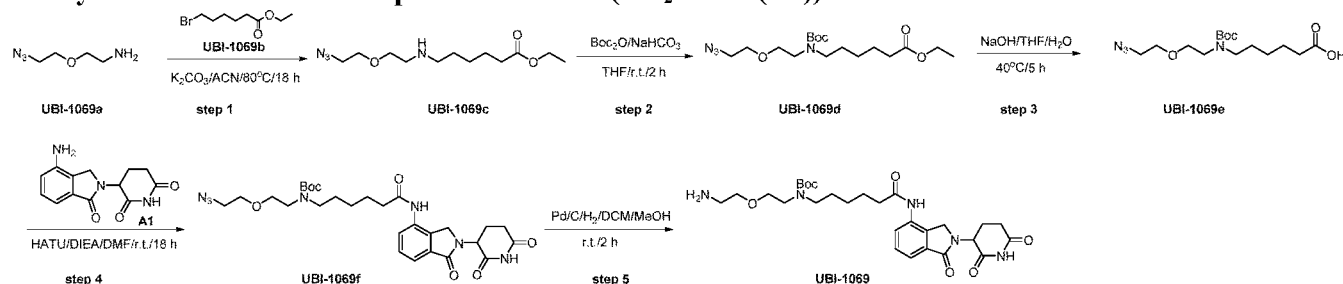
Compound **UBI-1160e** (70 mg, 0.22 mmol), **B(A1)** (50 mg, 0.11 mmol), EDCI (32 mg, 0.165 mmol), HOBT (18 mg, 0.13 mmol), DIEA (0.5 mL) were dissolved in DMF (5 mL) and reacted at room temperature overnight. Further adding HATU (870 mg, 2.3 mmol) and reacting at room temperature overnight. The reaction was quenched with water (50 mL), extracted with ethyl acetate (50 mL*3). The

organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 97/3) to obtain crude product **UBI-1160f** (500 mg, yield 76%) as yellow oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.02 (s, 1H), 9.75 (s, 1H), 7.82 (m, 1H), 7.55 – 7.43 (m, 2H), 5.15 (m, 1H), 4.43 – 4.28 (m, 2H), 3.66 – 3.57 (m, 8H), 3.55 – 3.50 (m, 8H), 3.15 (m, 8H), 2.06 – 1.95 (m, 2H), 1.62 (m, 2H), 1.49 (m, 4H), 1.38 (d, 9H). LCMS: [M-100]⁺ = 574

Step 5: UBI-1160

Compound **UBI-1160e** (500 mg, 0.74 mmol) was dissolved in a mixed solvent of dichloromethane (30 mL) and methanol (3 mL), added Pd/C (500 mg), and reacted for 2 hours under hydrogen atmosphere. It was filtered on Celite. The filtrate was concentrated to obtain product **UBI-1160f** (130 mg, yield 27%) as yellow oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.02 (s, 1H), 9.76 (s, 1H), 7.54 – 7.46 (m, 1H), 7.25 (t, 1H), 7.20 – 7.11 (m, 1H), 5.15 (dd, 1H), 4.45 – 4.24 (m, 2H), 3.68 – 3.38 (m, 15H), 3.27 (m, 2H), 3.14 (m, 5H), 3.01 – 2.93 (m, 3H), 2.68 – 2.59 (m, 1H), 2.39 – 2.31 (m, 2H), 2.08 – 1.99 (m, 1H), 1.61 (m, 2H), 1.48 (m, 2H), 1.38 (d, 9H). LCMS: [M+H]⁺ = 648

Synthesis method of Compound UBI-1069 (NH₂-12b-B(A1))



Step 1: UBI-1169c

Compound **UBI-1169a** (1.5 g, 11.5 mmol) was dissolved in acetonitrile (30 mL), added anhydrous potassium carbonate (3.18 g, 23 mmol) and **UBI-1169b** (2.56 g, 11.5 mmol), reacted at 80 °C overnight. The reaction was cooled to room temperature and filtered. The filtrate was concentrated and isolated by silica gel column chromatography (dichloromethane /methanol = 95/5) to obtain product **UBI-1169c** (1.3 g, yield 41%) as white solid. ¹H NMR (400 MHz, chloroform-*d*) δ 4.12 (q, *J* = 7.1 Hz, 2H), 3.70 – 3.59 (m, 4H), 3.38 (t, *J* = 5.0 Hz, 2H), 2.86 – 2.79 (m, 2H), 2.70 – 2.60 (m, 2H), 2.30 (t, *J* = 7.5 Hz, 2H), 1.69 – 1.60 (m, 2H), 1.53 (p, *J* = 7.4 Hz, 2H), 1.42 – 1.34 (m, 2H), 1.25 (t, *J* = 7.1 Hz, 3H).

Step 2: UBI-1169d

Compound **UBI-1169c** (1.3 g, 4.77 mmol) was dissolved in tetrahydrofuran (30 mL), added NaHCO₃ (1.2 g, 14.33 mmol) and Boc₂O (1.24 g, 5.72 mmol), and reacted at room temperature for 2 hours. The reaction was quenched by adding water (20 mL) and extracted with ethyl acetate (20 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated to obtain crude product **UBI-1169d** (2.5 g, yield 100%) as yellow oil.

Step 3: UBI-1169e

Compound **UBI-1169d** (1.7 g, 4.77 mmol) was dissolved in tetrahydrofuran (20 mL) and water (10 mL), added sodium hydroxide (572 mg, 14.31 mmol), and reacted at 40 °C for 5 hours under string. The reaction solution was concentrated, and added saturated brine (15 mL) and extracted with ether (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain crude product **UBI-1169e** (300 mg, yield 18%) as colorless oil.

¹H NMR (400 MHz, DMSO-*d*₆) δ 11.96 (s, 1H), 3.62 – 3.56 (m, 2H), 3.51 (t, *J* = 5.9 Hz, 2H), 3.38 (d, *J* = 9.5 Hz, 2H), 3.28 (t, *J* = 5.9 Hz, 2H), 3.16 (q, *J* = 7.4, 6.3 Hz, 2H), 2.19 (t, *J* = 7.3 Hz, 2H), 1.50 (dt, *J* = 15.0, 7.5 Hz, 4H), 1.39 (s, 9H), 1.25 – 1.17 (m, 2H).

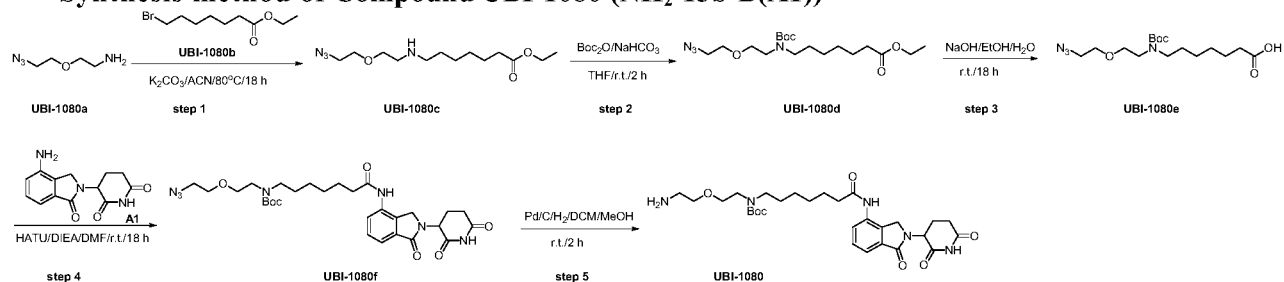
Step 4: UBI-1069f

5 Compound **UBI-1169e** (300 mg, 0.87 mmol), **B(A1)** (226 mg, 0.87 mmol), HATU (660 mg, 1.7 mmol), and DIPEA (561 mg, 4.35 mmol) were dissolved in DMF (5 mL), reacted at room temperature overnight. The reaction was quenched with water (20 mL), extracted with ethyl acetate (50 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 97/3) to obtain crude product **UBI-1069f**
 10 (300 mg, yield 71%) as yellow oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.02 (s, 1H), 9.76 (s, 1H), 7.81 (m, 1H), 7.53 – 7.44 (m, 2H), 5.15 (m, 1H), 4.35 (m, 2H), 3.65 – 3.57 (m, 6H), 3.38 (d, *J* = 4.8 Hz, 2H), 3.16 (m, 6H), 2.02 (m, 2H), 1.62 (m, 2H), 1.49 (m, 4H), 1.38 (d, 9H). LCMS: [M-100]⁺ = 486

Step 5: UBI-1069

15 Compound **UBI-1069f** (300 mg, 0.51 mmol) was dissolved in a mixed solvent of dichloromethane (30 mL) and methanol (3 mL), added Pd/C (300 mg), and reacted for 2 hours under hydrogen atmosphere. It was filtered on Celite. The filtrate was concentrated to obtain product **UBI-1069** (200 mg, yield 70%) as yellow oil. LCMS: [M+H]⁺ = 560

Synthesis method of Compound UBI-1080 (NH₂-13b-B(A1))



Step 1: UBI-1180c

20 Compound **UBI-1180a** (1.5 g, 11.5 mmol) was dissolved in acetonitrile (30 mL), added anhydrous potassium carbonate (3.17 g, 23 mmol) and **UBI-1180b** (2.73 g, 11.5 mmol), reacted at 80 °C overnight. The reaction was cooled to room temperature and filtered. The filtrate was concentrated and isolated by silica gel column chromatography (dichloromethane /methanol = 95/5) to obtain crude product **UBI-1180c**
 25 (1.4 g, yield 42%) as a yellow oil. ¹H NMR (400 MHz, chloroform-*d*) δ 4.12 (q, 2H), 3.72 – 3.60 (m, 4H), 3.47 (s, 1H), 3.38 (t, 2H), 2.86 – 2.76 (m, 2H), 2.65 – 2.59 (m, 2H), 2.29 (t, 2H), 1.69 – 1.58 (m, 2H), 1.56 – 1.47 (m, 2H), 1.34 (m, 4H), 1.25 (t, 3H).

Step 2: UBI-1180d

30 Compound **UBI-1180c** (1.4 g, 4.8 mmol) was dissolved in tetrahydrofuran (30 mL), added NaHCO₃ (1.2 g, 14.4 mmol) and Boc₂O (1.15 g, 5.28 mmol), and reacted at room temperature for 2 hours. The reaction was quenched with water (20 mL), extracted with ethyl acetate (20 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated to obtain crude product **UBI-1180d** (2 g, yield 100%) as yellow oil.

Step 3: UBI-1180

35 Compound **UBI-1180d** (1.85 g, 4.8 mmol) was dissolved in EtOH (20 mL) and water (10 mL), added sodium hydroxide (1.15 g, 28.8 mmol), and reacted at room temperature overnight. The reaction solution was concentrated, and added saturated brine (15 mL) and extracted with ether (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, then extracted with ethyl acetate (40 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by

silica gel column chromatography (dichloromethane/methanol = 94/6) to obtain product **UBI-1180** (1.4 g, yield 82%) as colorless oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.96 (s, 1H), 3.58 (t, *J* = 4.8 Hz, 2H), 3.51 (t, *J* = 5.9 Hz, 2H), 3.38 (t, *J* = 4.9 Hz, 2H), 3.28 (t, *J* = 5.9 Hz, 2H), 3.15 (t, *J* = 7.4 Hz, 2H), 2.18 (t, *J* = 7.4 Hz, 2H), 1.53 – 1.41 (m, 4H), 1.39 (s, 9H), 1.24 (m, 4H).

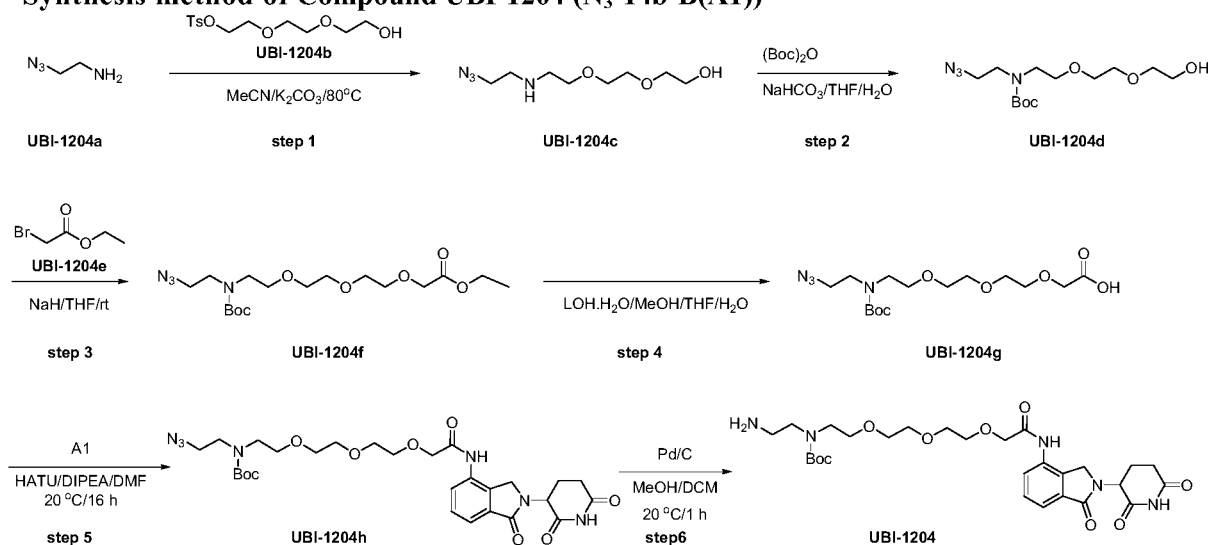
Step 4: UBI-1180f

Compound **UBI-1180e** (500 mg, 1.39 mmol), **B(A1)** (362 mg, 1.39 mmol), HATU (633 mg, 1.67 mmol), and DIPEA (3 mL) were dissolved in DMF (8 mL), reacted at room temperature overnight. The reaction was quenched with water (20 mL), extracted with ethyl acetate (50 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane /methanol = 93/7) to obtain product **UBI-1180f** (500 mg, yield 60%) as yellow oil. LCMS: [M+H]⁺ = 600

Step 5: UBI-1180

Compound **UBI-1180f** (500 mg, 0.83 mmol) was dissolved in a mixed solvent of dichloromethane (30 mL) and methanol (3 mL), added Pd/C (300 mg), and reacted for 2 hours under hydrogen atmosphere. It was filtered on Celite. The filtrate was concentrated to obtain product as yellow oil. The product was purified by a reversed-phase column (water: methanol = 97:3) to obtain product **UBI-1180** (270 mg, yield 57%) as yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.77 (s, 1H), 7.82 (m, 1H), 7.53 – 7.46 (m, 2H), 5.15 (m, 1H), 4.42 – 4.27 (m, 2H), 3.56 (m, 3H), 3.50 (m, 3H), 3.19 – 3.11 (m, 4H), 2.95 (m, 4H), 2.37 (m, 2H), 1.61 (m, 2H), 1.46 (m, 4H), 1.38 (m, 13H), 1.26 (m, 5H), 1.14 – 1.09 (m, 3H), 1.07 – 0.98 (m, 2H). LCMS: [M+H]⁺ = 574

Synthesis method of Compound UBI-1204 (N₃-14b-B(A1))



Step 1: UBI-1204c

Compound **UBI-1204a** (5.986g, 69.52mmol), **BUI-1204b** (19.05g, 66.05mmol) and K₂CO₃ (11.53 g, 83.43 mmol) were dissolved in acetonitrile (200 mL), then the reaction was reacted at room temperature for 3 hours. The reaction solution was filtered. The filtrate was concentrated and isolated by silica gel column chromatography (methanol/dichloromethane = 0% to 10%) to obtain product **UBI-1204c** (4.10g, yield 26%) as colorless transparent oil. ¹H NMR (400 MHz, chloroform-*d*) δ 3.76 - 3.71 (m, 2H), 3.70 - 3.59 (m, 8H), 3.49 - 3.41 (m, 2H), 2.86 - 2.79 (m, 4H), 2.14 - 1.97 (m, 2H).

Step 2: UBI-1204d

Compound **UBI-1204c** (4.100g, 18.76mmol), (Boc)₂O (8.200g, 37.57mmol) and NaHCO₃ (3.156g, 37.57mmol) were dissolved in tetrahydrofuran (50 mL) and reacted at room temperature for 16 hours. The

reaction was filtered. The filtrate was concentrated and isolated by column chromatography (ethyl acetate/petroleum ether = 0% to 95%) to obtain product **UBI-1204d** (4.85g, yield 71%) as colorless transparent oil. ¹H NMR (400 MHz, chloroform-*d*) δ 3.76 - 3.72 (m, 2H), 3.69 - 3.64 (m, 2H), 3.64 - 3.56 (m, 6H), 3.49 - 3.39 (m, 6H), 1.93 (s, 1H), 1.47 (s, 9H).

Step 3: UBI-1204e

Compound **UBI-1097d** (1.00g, 3.14mmol) was dissolved in tetrahydrofuran (10 mL) and cooled to 0°C, added NaH (0.251g, 6.28mmol), and reacted for ten minutes. Then **UBI-1204e** (0.787g, 4.71mmol) was added, and the reaction was carried out at room temperature for 16 hours. The reaction was filtered. The filtrate was concentrated, added water (50 mL), and extracted with ethyl acetate (50 mL*3). The organic phases were combined, washed with water, saturated brine, dried over anhydrous sodium sulfate, and concentrated under reduced pressure, then isolated by silica gel column chromatography (methanol/dichloromethane = 0% to 3%) to obtain product **UBI-1204f** (705mg, 53%) as colorless transparent oil. ¹H NMR (400 MHz, chloroform-*d*) δ 4.22 (q, *J* = 7.2 Hz, 2H), 4.15 (s, 2H), 3.77 - 3.72 (m, 2H), 3.72 - 3.67 (m, 2H), 3.67 - 3.63 (m, 2H), 3.63 - 3.55 (m, 4H), 3.48 - 3.39 (m, 6H), 1.47 (s, 9H), 1.29 (t, *J* = 7.2 Hz, 3H).

Step 4: UBI-1204g

Compound **UBI-1204f** (698mg, 1.21mmol), LiOH.H₂O (253mg, 6.04mmol) were dissolved in a mixed solvent consisting of methanol (3 mL), tetrahydrofuran (9 mL) and water (3 mL), stirred at room temperature overnight. The reaction solution was added water, concentrated and extracted with ethyl acetate. The aqueous phase was acidified to pH=5 using hydrochloric acid (1M), and then lyophilized. The solid was added to a mixed solvent consisting of methanol (2 mL) and dichloromethane (20 mL), filtered after stirring for 10 minutes. The filtrate was concentrated to obtain product **UBI-1204g** (650 mg, yield 90%) as colorless oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 3.86 (s, 2H), 3.58 - 3.55 (m, 2H), 3.54 - 3.46 (m, 8H), 3.43 - 3.38 (m, 4H), 3.36 - 3.32 (m, 2H), 1.40 (s, 9H). LCMS: [M+Na]⁺ = 399.3.

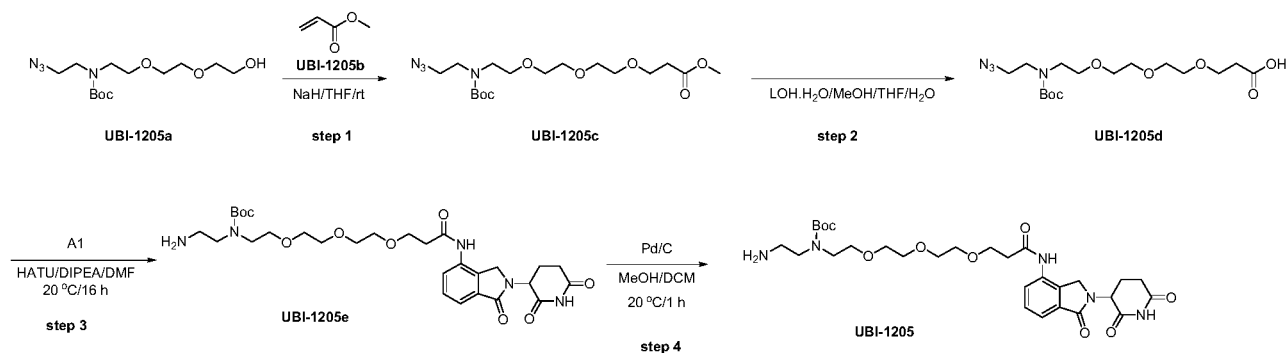
Step 5: UBI-1204h

Compound **UBI-1204g** (120mg, 0.32mmol), **B(A1)** (82.6mg, 0.32mmol), and HATU (181.8 mg, 0.478 mmol) were dissolved in DMF (3 mL) and DIPEA (0.3 mL), and reacted at room temperature for 16 hours. The reaction was quenched with water (10 mL), then extracted with ethyl acetate (8 mL*3). The organic phases were combined, then dried over anhydrous sodium sulfate and concentrated to obtain crude product. The crude product was purified by reversed-phase column (methanol/water = 5% to 95%, collected at 70%) to obtain product **UBI-1204h** (90mg, yield 45.7%) as colorless oil. LCMS: [M+H-100]⁺ = 518.3.

Step 6: UBI-1204

Under hydrogen atmosphere, compound **UBI-1204h** (90mg, 0.15mmol), Pd/C (30mg) were dissolved in methanol (1 mL) and dichloromethane (10mL), and the reaction solution was reacted at room temperature for 1 hour. The reaction solution was filtered through Celite. The filtrate was concentrated and purified via reversed-phase column (methanol/water = 5% to 95%, collected at 15%) to obtain the product **UBI-1204** (80mg, yield 93%) as yellow oil. LCMS: [M+H]⁺ = 592.7.

Synthesis method of Compound UBI-1205 (N₃-15b-B(A1))



Step 1: UBI-1205c

Compound **UBI-1205a** (1.050g, 3.298mmol) was dissolved in tetrahydrofuran (10 mL), added NaH (26.4mg, 0.660mmol) and reacted at room temperature for 10 minutes. Then **UBI-1205b** (0.426 g, 4.95 mmol) was added, and the reaction was carried out at room temperature overnight. The reaction solution was added methanol, then concentrated, and isolated by silica gel column chromatography (ethyl acetate/petroleum ether = 0% to 95%) to obtain product **UBI-1205c** (710mg, yield 51%) as colorless and transparent oil. ^1H NMR (400 MHz, chloroform- d) δ 3.76 (t, J = 6.4 Hz, 2H), 3.69 (s, 3H), 3.66 - 3.53 (m, 10H), 3.48 - 3.37 (m, 6H), 2.61 (t, J = 6.4 Hz, 2H), 1.47 (s, 9H).

Step 2: UBI-1205d

Compound **UBI-1205c** (700mg, 1.21mmol) and LiOH.H₂O (254mg, 6.06mmol) were dissolved in a mixed solvent consisting of methanol (3 mL), tetrahydrofuran (9 mL) and water (3 mL). The reaction was stirred at room temperature overnight. The reaction solution was added water, concentrated and then extracted with ethyl acetate. The aqueous phase was acidified to pH=5 using hydrochloric acid (1M), and then lyophilized. The solid was poured into a mixed solvent of methanol (2 mL) and dichloromethane (20 mL), stirred for 10 minutes, filtered. The filtrate was concentrated to obtain product **UBI-1205d** (650mg, yield 87%) as colorless transparent oil.

^1H NMR (400 MHz, DMSO- d_6) δ 11.98 (s, 1H), 3.59 (t, J = 6.4 Hz, 2H), 3.55 - 3.44 (m, 10H), 3.43 - 3.38 (m, 4H), 3.35 - 3.32 (m, 2H), 2.43 (t, J = 6.4 Hz, 2H), 1.40 (s, 9H). LCMS: $[\text{M}+\text{Na}]^+ = 413.3$.

Step 3: UBI-1205e

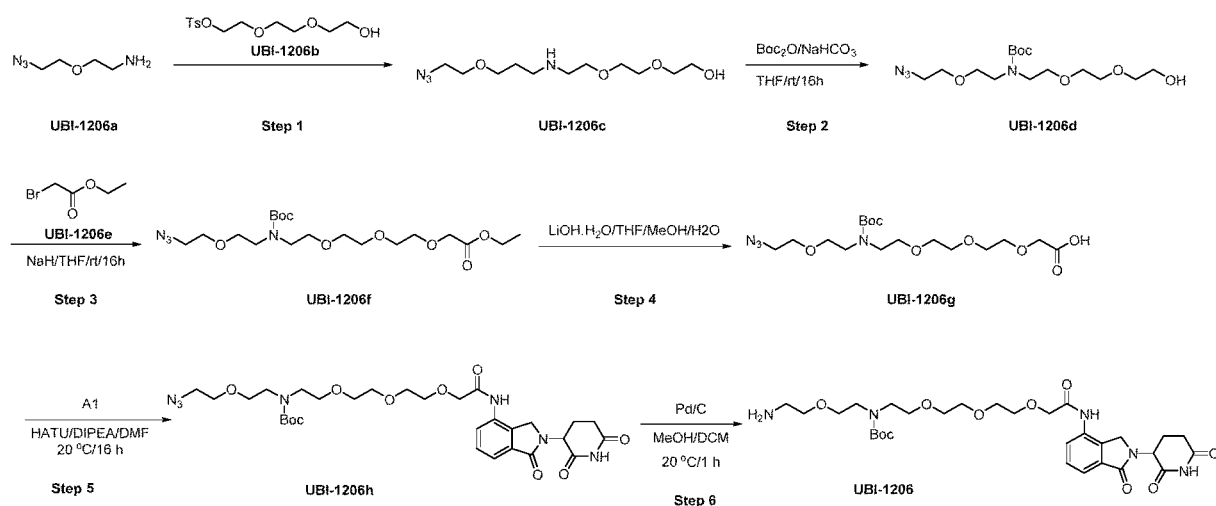
Compound **UBI-1205d** (150mg, 0.38mmol), **B(A1)** (100mg, 0.38mmol), and HATU (0.22g, 0.58 mmol) were dissolved in DMF (3 mL) and DIPEA (0.3 mL), and reacted at room temperature for 16 hours. The reaction was quenched with water (10 mL), then extracted with ethyl acetate (8 mL*3). The organic phases were combined, then dried over anhydrous sodium sulfate and concentrated to obtain crude product. The crude product was purified by reversed-phase column (methanol/water = 5% to 95%, collected at 70%) to obtain product **UBI-1205e** (95mg, yield 40%) as colorless oil.

LCMS: $[\text{M}+\text{H}-100]^+ = 532.3$.

Step 4: UBI-1205

Under hydrogen atmosphere, compound **UBI-1205e** (95mg, 0.15mmol), Pd/C (29mg) were dissolved in methanol (1 mL) and dichloromethane (10mL), and the reaction solution was reacted at room temperature for 1 hour. The reaction solution was filtered through Celite. The filtrate was concentrated and purified via reversed-phase column (methanol/water = 5% to 95%, collected at 15%) to obtain the product **UBI-1205** (70mg, yield 77%) as yellow oil. LCMS: $[\text{M}+\text{H}]^+ = 606.3$.

Synthesis method of Compound UBI-1206 (N₃-17b-B(A1))



Step 1: UBI-1206c

Compound **UBI-1206a** (5.000g, 38.42mmol), **UBI-1206b** (11.69g, 38.42mmol), K_2CO_3 (6.371g, 46.10mmol) were added to acetonitrile (200 mL) and reacted at room temperature overnight under stirring. The reaction was filtered. The filtrate was concentrated and isolated by silica gel column chromatography (methanol/dichloromethane = 0% to 6%) to obtain product **UBI-1206c** (4.54g, yield 43%) as colorless transparent oil. ^1H NMR (400 MHz, chloroform-*d*) δ 3.76 - 3.70 (m, 2H), 3.68 - 3.60 (m, 12H), 3.40 - 3.37 (m, 2H), 2.87 - 2.80 (m, 4H), 2.18 (s, 2H). LCMS: $[M+H]^+ = 263.2$.

Step 2: UBI-1206d

Compound **UBI-1206c** (4.524g, 17.25mmol), $(Boc)_2O$ (7.528g, 34.49mmol), $NaHCO_3$ (2.898g, 34.49mmol) were added into tetrahydrofuran (50 mL), and reacted at room temperature overnight under stirring. The reaction solution was filtered. The filtrate was concentrated and isolated by silica gel column chromatography (ethyl acetate/petroleum ether = 0% to 95%) to obtain product **UBI-1206d** (6.04g, yield 90%) as colorless transparent oil. ^1H NMR (400 MHz, chloroform-*d*) δ 3.77 - 3.70 (m, 2H), 3.70 - 3.54 (m, 12H), 3.47 (s, 4H), 3.37 (t, $J = 5.2$ Hz, 2H), 2.10 (s, 1H), 1.46 (s, 9H).

Step 3: UBI-1206e

Compound **UBI-1206d** (2.90 g, 8.00 mmol) was dissolved in tetrahydrofuran (20 mL), added NaH (640 mg, 16.0 mmol) under ice bath, and reacted for ten minutes. Then **UBI-1206e** (2.004 g, 12.00 mol) was added, and reacted at room temperature for 16 hours. the reaction was filtered. The filtrate was concentrated, added (50 mL) and extracted with ethyl acetate (50 mL*3). The organic phases were combined, washed with water (50 mL*3), saturated brine (50 mL*3) successively, dried over anhydrous sodium sulfate, and concentrated, then isolated by silica gel column chromatography (methanol/dichloromethane = 0% to 10%) to obtain product **UBI-1206f** (910 mg, 25%) as yellow oil. LCMS: $[M+H-100]^+ = 349.3$.

Step 4: UBI-1206g

Compound **UBI-1206f** (900mg, 1.41mmol) and $LiOH.H_2O$ (295mg, 7.02mmol) were dissolved in a mixed solvent consisting of methanol (3 mL), water (3 mL) and tetrahydrofuran (9 mL), and reacted at room temperature overnight under stirring. The reaction solution was added water, concentrated and then extracted with ethyl acetate. The aqueous phase was acidified to pH=5 using hydrochloric acid (1M), then extracted with ethyl acetate. The organic phase was combined, washed with water and saturated brine, then dried over anhydrous sodium sulfate, and concentrated to obtain product **UBI-1206g** (740mg, yield 83%) as colorless oil.

^1H NMR (400 MHz, DMSO-*d*₆) δ 12.42 (s, 1H), 4.01 (s, 2H), 3.64 - 3.56 (m, 4H), 3.55 - 3.43 (m, 10H), 3.39 (d, $J = 4.2$ Hz, 2H), 3.37 - 3.33 (m, 4H), 1.39 (s, 9H). LCMS: $[M+H-100]^+ = 321.2$.

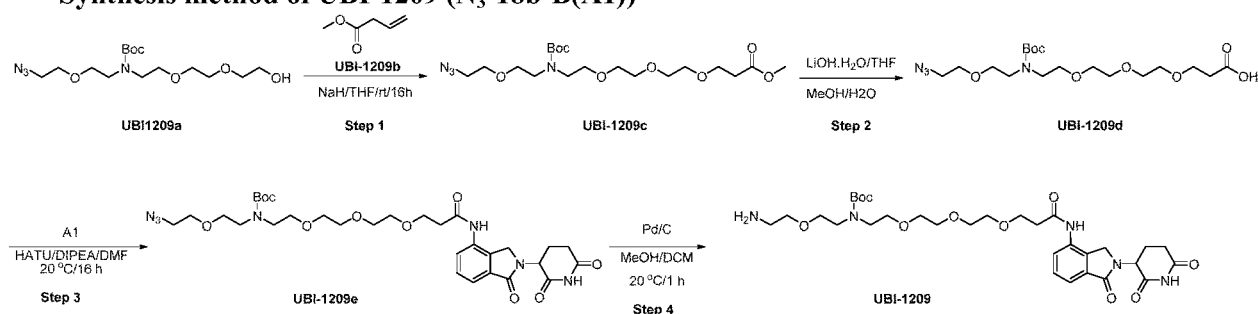
Step 5: UBI-1206h

Compound **UBI-1206g** (120mg, 0.28 mmol), **B(A1)** (74mg, 0.28mmol), and HATU (163 mg, 0.43 mmol) were dissolved in DMF (3 mL) and DIPEA (0.3 mL), and reacted at room temperature for 16 hours. The reaction was quenched with water (10 mL), then extracted with ethyl acetate (8 mL*3). The organic phases were combined, then dried over anhydrous sodium sulfate and concentrated to obtain crude product. The crude product was purified by reversed-phase column (methanol/water = 5% to 95%, collected at 70%) to obtain product **UBI-1206h** (90mg, yield 51%) as colorless oil. LCMS: $[M+H]^+ = 562.3$.

Step 6: UBI-1206

Under hydrogen atmosphere, compound **UBI-1206g** (90mg, 0.15mmol), Pd/C (29mg) were dissolved in methanol (1 mL) and dichloromethane (10mL), and the reaction solution was reacted at room temperature for 1 hour. The reaction solution was filtered through Celite. The filtrate was concentrated and purified via reversed-phase column (methanol/water=5% to 95%, collected at 15%) to obtain the product **UBI-1206** (65mg, yield 75%) as yellow oil. LCMS: $[M+H]^+ = 636.5$.

Synthesis method of UBI-1209 (N₃-18b-B(A1))



Step 1: UBI-1209c

Compound **UBI-1209a** (2.10g, 5.79mmol) was dissolved in tetrahydrofuran THF (10 mL), added NaH (46mg, 1.2mmol) and reacted at room temperature for 10 minutes. Then **UBI-1209b** (0.748 g, 8.69 mmol) was added, and the reaction was stirred at room temperature overnight. The reaction was quenched, concentrated and isolated by silica gel column chromatography (ethyl acetate/petroleum ether = 5% to 95%) to obtain product **UBI-1209c** (645mg, yield 25%) as yellow oil.

¹H NMR (400 MHz, chloroform-*d*) δ 3.81 - 3.72 (m, 2H), 3.71 - 3.66 (m, 3H), 3.65 - 3.54 (m, 12H), 3.46 (d, $J = 5.2$ Hz, 4H), 3.38 - 3.34 (m, 2H), 2.68 - 2.53 (m, 2H), 1.58 - 1.57 (m, 2H), 1.45 (s, 9H).

Step 2: UBI-1209d

Compound **UBI-1209c** (600mg, 0.936mmol) and LiOH.H₂O (0.196g, 4.68mmol) were dissolved in methanol (3 mL), tetrahydrofuran (3 mL) and water (9 mL), and reacted at room temperature overnight under stirring. The reaction solution was added water, concentrated and then extracted with ethyl acetate. The aqueous phase was acidified to pH=5 using hydrochloric acid (1M), then extracted with ethyl acetate. The organic phases were combined, then washed with water, saturated brine, and dried over anhydrous sodium sulfate. After concentration, product **UBI-1209d** (305 mg, yield 47%) as yellow oil was obtained.

¹H NMR (400 MHz, DMSO-*d*₆) δ 12.19 (s, 1H), 3.65 - 3.55 (m, 2.9 Hz, 4H), 3.55 - 3.43 (m, 12H), 3.42 - 3.37 (m, 2H), 3.36 - 3.32 (m, 4H), 2.43 (t, $J = 6.4$ Hz, 2H), 1.39 (s, 9H). LCMS: $[M+H-100]^+ = 335.3$.

Step3: UBI-1209e

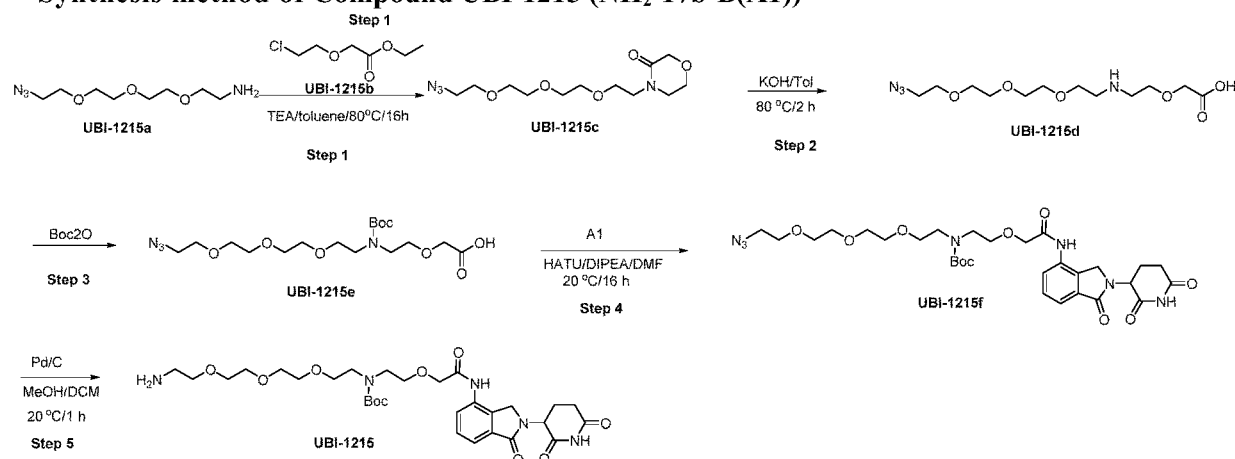
Compound **UBI-1209d** (120mg, 0.27mmol), **B(A1)** (71.6mg, 0.27mmol), and HATU (157 mg, 0.41 mmol) were dissolved in DMF (3 mL) and DIPEA (0.3 mL), and reacted at room temperature for 16 hours. The reaction was quenched with water (10 mL), then extracted with ethyl acetate (8 mL*3). The organic phases were combined, then dried over anhydrous sodium sulfate and concentrated to obtain crude product.

The crude product was purified by reversed-phase column (methanol/water = 5% to 95%, collected at 70%) to obtain product **UBI-1209e** (90mg, yield 52.7%) as colorless oil. LCMS: $[M+H-100]^+ = 676.3$

Step 4: UBI-1209

Under hydrogen atmosphere, compound **UBI-1209e** (90mg, 0.15mmol), Pd/C (29mg) were dissolved in methanol (1 mL) and dichloromethane (10mL), and the reaction solution was reacted at room temperature for 1 hour. The reaction solution was filtered through Celite. The filtrate was concentrated and purified via reversed-phase column (methanol/water=5% to 95%, collected at 15%) to obtain the product **UBI-1209** (60mg, yield 70%) as yellow oil. LCMS: $[M+H]^+ = 650.3$

Synthesis method of Compound UBI-1215 (NH₂-17b-B(A1))



Step 1: UBI-1215c

Compound **UBI-1215a** (2.0 g, 9.17mmol), **UBI-1215b** (1.39g, 9.17mmol) were dissolved in TEA (1.39 g, 13.76mmol) and toluene (25mL), reacted at 80°C for 18 hours. The reaction solution was concentrated and isolated by silica gel column chromatography (petroleum ether/ethyl acetate = 50% to 100%, then MeOH/DCM=0% to 10%) to obtain target compound **UBI-1215** (500 mg, yield 18%) as colorless oil. ¹H NMR (400 MHz, chloroform-*d*) δ 4.17 (s, 2H), 3.89 – 3.83 (m, 2H), 3.69 (ddd, $J = 4.7, 1.9, 0.7$ Hz, 4H), 3.66 (d, $J = 1.3$ Hz, 4H), 3.63 (dt, $J = 4.6, 2.9, 1.4$ Hz, 4H), 3.61 – 3.58 (m, 2H), 3.53 (dd, $J = 6.0, 4.2$ Hz, 2H), 3.39 (t, $J = 5.0$ Hz, 2H).

Step 2: UBI-1215d

Compound **UBI-1215c** (400mg, 1.32mmol), KOH (371mg, 6.62mmol) were dissolved in water (10mL), and refluxed at 100°C for 2 hours. The reaction liquid was directly used in the next step after cooling. LCMS: $[M+H]^+ = 321.1$.

Step 3: UBI-1215e

To the aqueous solution of compound **UBI-1215d** (400mg, 1.25mmol), di-tert-butyl dicarbonate (545mg, 2.5 mmol) and dioxane (4mL) were added, and reacted at room temperature for 2 hours. The reaction solution was concentrated, added water (20 mL), and extracted with ethyl acetate (20 mL*3). The aqueous phase was concentrated to obtain the crude product, which was dissolved in MeOH/DCM=10/1 (30 mL), filtered, and the filtrate was concentrated to obtain the target compound **UBI-1215e** (300 mg). LCMS: $[M+H-100]^+ = 335.3$.

Step 4: UBI-1215f

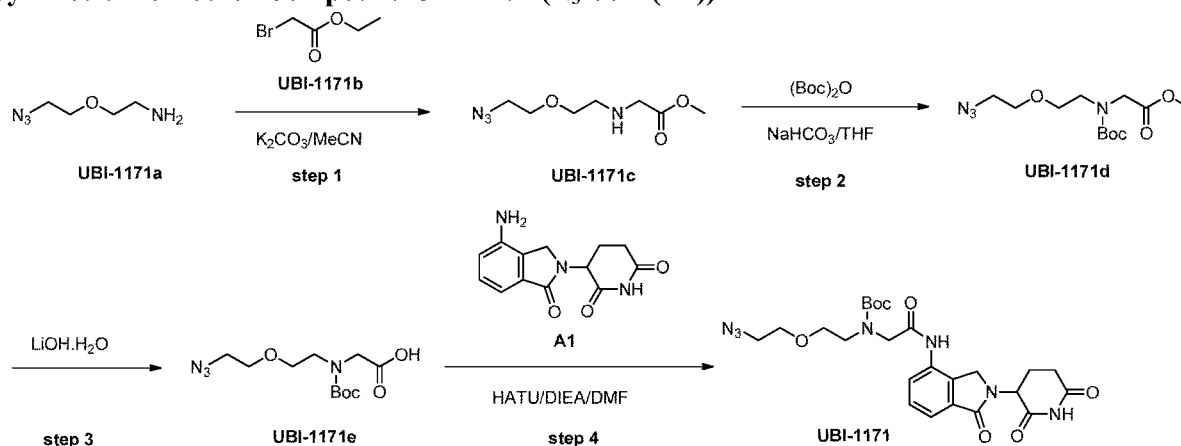
Compound **UBI-1215e** (300mg, 0.71mmol), **B(A1)** (184mg, 0.71mmol), and HATU (406.9 mg, 1.07 mmol) were dissolved in DMF (10 mL) and DIPEA (0.3 mL), and reacted at room temperature for 16 hours. The reaction solution was directly isolated by reversed-phase column (MeOH/H₂O=5% to 95%, 45 minutes, collected at 60%) to obtain the target compound **UBI-1215f** (150mg, yield 32%) as white solid. LCMS:

$[M+H-100]^+ = 562.3$.

Step 5: UBI-1215

Compound **UBI-1215f** (150mg, 0.08mmol) and Pd/C (20mg) were dissolved in DCM (5mL), and reacted under hydrogen atmosphere for 1 hours. After the completion of reaction, the reaction was filtrated. The filtrate was concentrated to obtain the target compound **UBI-1215** (100mg, yield 69%), which was directly used in the next step. LCMS: $[M+H]^+ = 636.3$.

Synthesis method of Compound UBI-1171 (**N₃-8b-B(A1)**)



Step 1: UBI-1171c

Compound **UBI-1171a** (3 g, 23 mmol) was dissolved in acetonitrile (50 mL), added **UBI-1171b** (3.9 g, 23 mmol) and K_2CO_3 (4.8 g, 35 mmol), and reacted at room temperature for 16 hours. The reaction solution was filtered. Then the filtrate was concentrated and isolated by silica gel column chromatography (ethyl acetate/petroleum ether = 1/1 to methanol/dichloromethane = 1/10) to obtain target product **UBI-1171c** (2.6 g, yield 57 %) as colorless transparent oil.

^1H NMR (400 MHz, chloroform-*d*) δ 3.73 (s, 3H), 3.70 – 3.57 (m, 4H), 3.48 (dd, $J = 5.6, 3.0$ Hz, 2H), 3.39 (td, $J = 5.1, 1.7$ Hz, 2H), 2.83 (td, $J = 5.0, 2.6$ Hz, 2H), 1.79 (s, 1H).

Step 2: UBI-1171d

Compound **UBI-1171c** (2.6 g, 13 mmol) was dissolved in tetrahydrofuran (50 mL), then added (Boc) $_2$ O (4.2 g, 19 mmol) and $NaHCO_3$ (2.2 g, 26 mmol), and then reacted at room temperature for 16 hours. The reaction solution was added with water (10 mL) and extracted with ethyl acetate (15 mL*2). The organic phase was dried over Na_2SO_4 and then concentrated to obtain crude product **UBI-1171d** (3.6 g, yield 93%) as light yellow oil.

Step 3: UBI-1171e

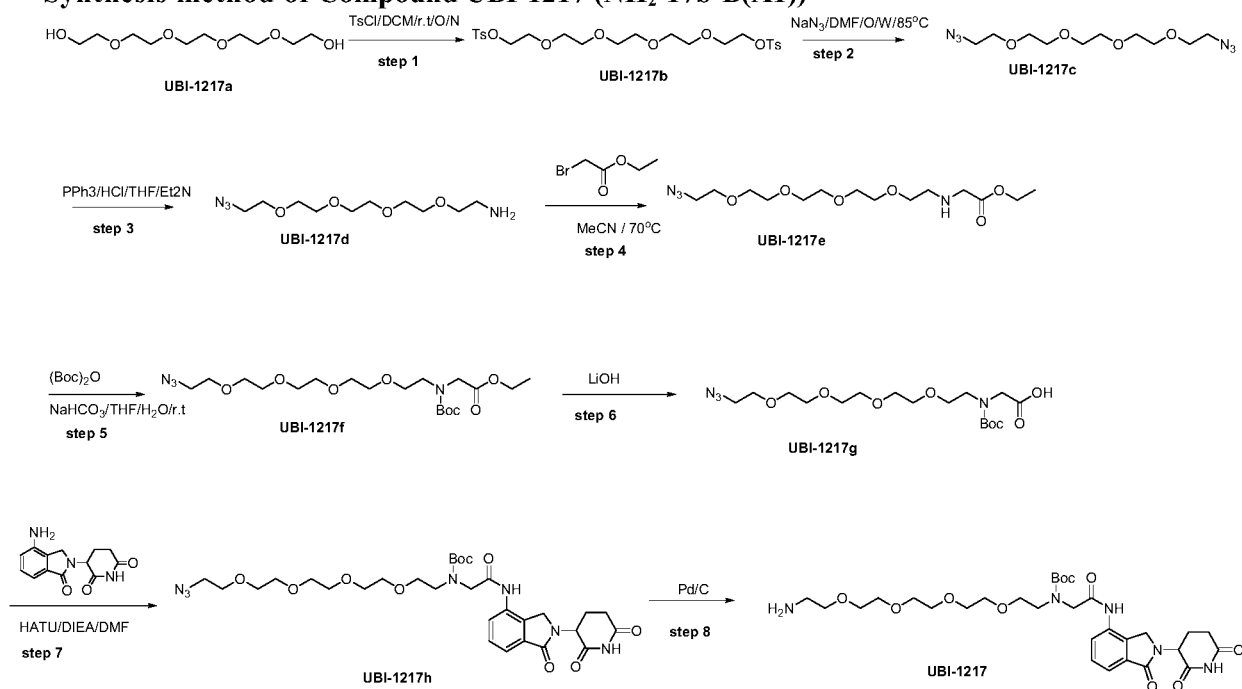
Compound **UBI-1171d** (3.6 g, 12 mmol) was dissolved in methanol (10 mL), tetrahydrofuran (30 mL) and water (10 mL), and then added $LiOH \cdot H_2O$ (751 mg, 18 mmol). The reaction was allowed to react at room temperature for 16 hours. The reaction solution was washed with ethyl acetate (20 mL) once, the aqueous phase was adjusted to pH=6 with 3N HCl, and then extracted with ethyl acetate (25 mL*2). After concentration of the organic phase, crude product **UBI-1171e** (2.9g, 85% yield), as colorless oil, was obtained. The crude product was directly used in the next step. ^1H NMR (400 MHz, chloroform-*d*) δ 10.52 (s, 1H), 4.06 (d, $J = 14.6$ Hz, 2H), 3.62 (dt, $J = 15.8, 5.1$ Hz, 4H), 3.55 – 3.42 (m, 2H), 3.35 (q, $J = 4.5$ Hz, 2H), 1.45 (d, $J = 15.0$ Hz, 9H).

Step 4: UBI-1171

At 0°C, compound **UBI-1171e** (350 mg, 1.2 mmol) was dissolved in N,N-dimethylformamide (8 mL) and then added HATU (924 mg, 2.4 mmol) and DIEA (313 mg, 2.4 mmol), followed by **B(A1)** (315 mg,

1.2 mmol). The system was reacted at room temperature for 16 h. The reaction solution was quenched with water (10 mL), extracted with ethyl acetate (10 mL*2). The organic phase was dried over Na₂SO₄ and then concentrated to obtain crude product. The crude product was purified by a reversed-phase column (methanol/water=5% to 95%, 45 minutes, collected at 80%) to obtain target product **UBI-1171** (350 mg, yield 54%) as yellow solid. LCMS [M+H]⁺=530.0

Synthesis method of Compound **UBI-1217** (**NH₂-17b-B(A1)**)



Step 1: **UBI-1217b**

UBI-1217a (8.0 g, 34 mmol) was dissolved in dichloromethane (100 ml), added *p*-toluenesulfonyl chloride (14.7 g, 7.7 mmol), added triethylamine (10.2 g, 101 mmol) at 25°C, and stirred overnight. After completion of reaction, to the reaction solution was added water (10ml) and extracted with dichloromethane (10ml*3). The organic phase was dried over Na₂SO₄ and concentrated to obtain crude product, which was purified by silica gel column chromatography (dichloromethane/methanol=30/1) to obtain colorless oil (18 g, yield 99%). LC-MS : (M+H)⁺=547.1

Step 2: **UBI-1217c**

UBI-1217b (18 g, 33 mmol) was dissolved in DMF (100 ml), added NaN₃ (6.43 g, 99 mmol), and stirred overnight at 85°C. After the completion of the reaction, the reaction solution was filtered and concentrated *in vacuo* to obtain crude product, which was purified with FFC (dichloromethane/methanol=30/1) to obtain colorless oil (8.7g, yield 92%).

Step 3: **UBI-1217d**

Compound **UBI-1217c** (8.7 g, 30 mmol) and triphenylphosphine (7.9 g, 30 mmol) were mixed and then dissolved in Et₂O/THF/1M HCl (100 ml), and stirred at 25°C for 12 h. After the completion of the reaction, the reaction was added water, and extracted with ethyl acetate (50 ml*2), followed by the organic layer was dried over anhydrous magnesium sulfate and concentrated, the crude product was purified by silica gel column chromatography (EtOAc/PE=1/1) to obtain oil (3.9 g, yield 45%). ¹H NMR (400 MHz, CDCl₃) δ 3.69 – 3.62 (m, 14H), 3.52 – 3.40 (m, 2H), 3.39 – 3.37 (m, 2H), 2.87 – 2.80 (m, 2H), 1.40 (s, 2H).

Step 4: **UBI-1217e**

Compound **UBI-1217d** (2 g, 7.6 mmol), ethyl bromoacetate (1.27 g, 7.6 mmol) and potassium

carbonate (2.06 g, 14.9 mmol) were mixed and dissolved in acetonitrile (20 mL), reacted at 80 °C for 18 hours. After the completion of reaction, the filtrate was concentrated by rotary evaporation under reduced pressure, and isolated by silica gel column chromatography (dichloromethane/methanol = 10/1) to obtain **UBI-1217e** (1.4g, yield 30%) as colorless oil.

Step 5: **UBI-1217f**

Compound **UBI-1217e** (1.3g, 3.7 mmol), di-tert-butyl dicarbonate (2.3g, 10.7mmol) and sodium bicarbonate (318 mg, 3.79 mmol) were added to dioxane (30mL) successively and reacted at room temperature for 2 hours. After the completion of the reaction, the reaction solution was poured into 10 mL of water and extracted with ethyl acetate (10 mL*3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure to obtain compound **UBI-1217f** (1.4g, yield 83%).

Step 6: **UBI-1217g**

Compound **UBI-1217f** (150 mg, 0.335 mmol), sodium hydroxide (26.8 mg, 0.669 mmol) were added to water (5 mL) successively, reacted at RT for 12 hours. After the completion of reaction, it was concentrated and the aqueous phase was acidified to pH=5 using hydrochloric acid (1M). Then it was extracted with dichloromethane (10ml*3), the combined organic layer was dried over anhydrous Na₂SO₄, and concentrated to obtain the desired compound **UBI-1217g** (131 mg, yield 93%).

Step 7: **UBI-1217h**

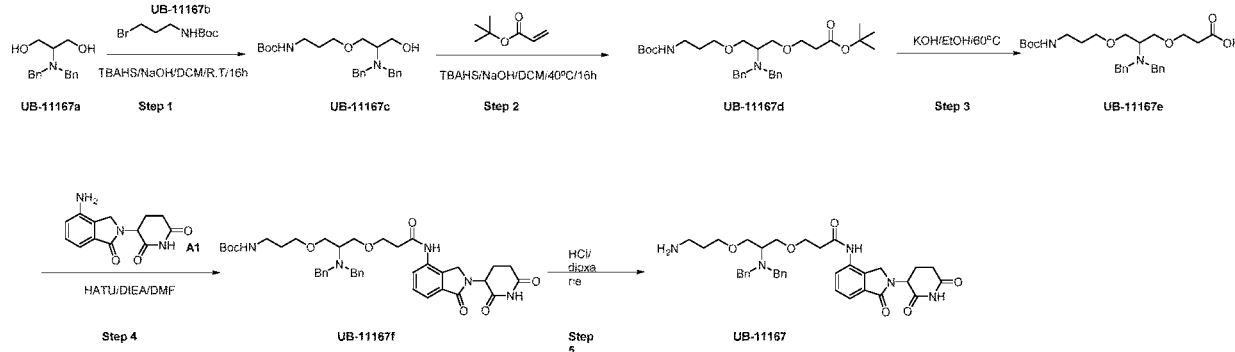
Compound **UBI-1217d** (101 mg, 0.24 mmol), **B(A1)** (75 mg, 0.29 mmol), HATU (137 mg, 0.36 mmol) and DIPEA (0.5mL) were added to anhydrous DMF (5 mL). The mixture was reacted at room temperature for 16 hours, concentrated after the completion of reaction. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 10/1) to obtain compound **UBI-1217h** (81 mg, 51% yield) as white solid. LC-MS: [M-100+H]⁺ = 562.3

Step 8: **UBI-1217**

Compound **UBI-1217h** (81 mg, 0.12 mmol), 10% palladium on carbon (20 mg) was added to a mixed solvent of methanol/dichloromethane (1 mL/10 mL), and reacted at room temperature for 16 hours under hydrogen atmosphere. After filtration, the filtrate was concentrated to obtain the crude product. The crude product was washed with cold ether (10 mL*3), and dried to obtain target compound **UBI-1217** (63 mg, yield 82%) as white solid. LCMS: (M-100+H)⁺ = 536.3

Synthesis method of Compound Linker c- Ligand B(A1)

Synthesis method of Compound **UBI-1167** (NH₂-12c-B(A1))



Step 1: **UBI-1167c**

A solution of Compound **UBI-1167a** (5g, 4.8mmol) in DCM (20mL) was cooled to 0°C, added NaOH (50% aq. 20mL), followed by tetrabutylammonium bisulfate (12.5g, 16.9mmol) and compound **UBI-1167b** (4.8g, 20.3mmol), reacted at 40°C for 16 hours. After cooling of reaction solution, the upper organic phase

was collected, and the aqueous phase was extracted with dichloromethane (10 mL * 3). All organic phases were combined, then dried over anhydrous Na₂SO₄, concentrated by rotary evaporation under reduced pressure, and isolated by silica gel column chromatography (petroleum ether/ethyl acetate = 10%) to obtain target compound **UBI-1167c** (2.25g) as colorless oil. ¹H NMR (400 MHz, chloroform-*d*) δ 7.39 – 7.15 (m, 11H), 4.80 (s, 1H), 3.88 (d, *J* = 13.3 Hz, 2H), 3.71 – 3.43 (m, 8H), 3.31 – 3.20 (m, 2H), 3.17 – 3.02 (m, 1H), 1.77 (p, *J* = 12.8, 9.5 Hz, 2H), 1.43 (s, 9H). LC-MS: [M+H]⁺ = 429.1

Step 2: UBI-1167d

A solution of Compound **UBI-1167c** (1.44g, 3.36mmol) in DCM (20mL) was cooled to 0°C, added NaOH (50% aq. 20mL), followed by tetrabutylammonium bisulfate (12.28g, 6.72mmol) and tert-butyl acrylate (861mg, 6.72mmol), then reacted at 40°C for 16 hours. The reaction solution was extracted with dichloromethane (10 mL*3). Organic phases were combined, then dried over anhydrous Na₂SO₄, concentrated by rotary evaporation under reduced pressure, and isolated by silica gel column chromatography (petroleum ether/ethyl acetate = 10%) to obtain target compound **UBI-1167d** (480mg) as colorless oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.40 – 7.25 (m, 8H), 7.22 – 7.14 (m, 2H), 6.75 (t, *J* = 5.8 Hz, 1H), 3.70 (s, 4H), 3.61 – 3.41 (m, 6H), 3.30 (d, *J* = 1.4 Hz, 2H), 3.03 – 2.95 (m, 2H), 2.89 (q, *J* = 6.0 Hz, 1H), 2.42 (t, *J* = 6.0 Hz, 2H), 1.61 (p, *J* = 6.5 Hz, 2H), 1.38 (d, *J* = 6.5 Hz, 18H).

Step 3: UBI-1167e

Compound **UBI-1167d** (2.45g, 4.4mmol) was dissolved in EtOH (20mL) and H₂O (5mL), then added KOH (2.46g, 44mmol), and reacted at 60°C for 3 hours. After the completion of reaction, it was concentrated and the aqueous phase was acidified to pH=6 using diluted hydrochloric acid (1M). Then it was extracted with dichloromethane (20 mL*3). Organic phases were combined, then dried over anhydrous Na₂SO₄, concentrated by rotary evaporation under reduced pressure to obtain target compound **UBI-1167e** (2g). LC-MS: [M+H]⁺ = 500.7

Step 4: UBI-1167f

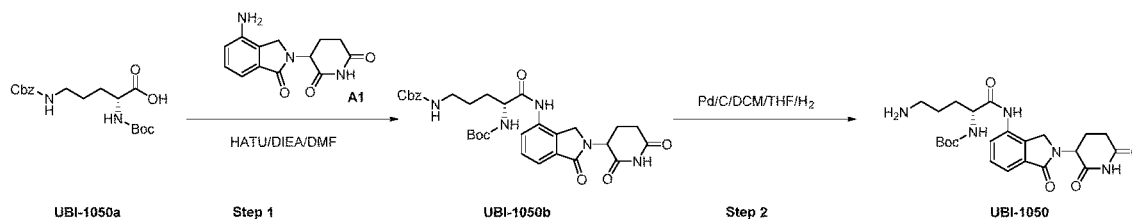
Compound **UBI-1167e** (1g, 2mmol), B(A1) (518mg, 2mmol), HATU (1.52g, 4mmol) were dissolved in DIPEA (1mL) and DMF (10mL), and reacted at room temperature for 16 hours. The reaction solution was directly isolated by reversed-phase column (MeOH/H₂O = 5% to 95%, collected at 48%) to obtain target compound **UBI-1167f** (440mg) as white solid.

¹H NMR (400 MHz, DMSO-*d*₆) δ 11.01 (d, *J* = 4.4 Hz, 1H), 9.87 (d, *J* = 8.4 Hz, 1H), 7.94 – 7.79 (m, 1H), 7.53 – 7.43 (m, 2H), 7.40 – 7.11 (m, 10H), 6.75 (t, *J* = 5.5 Hz, 1H), 5.20 – 5.01 (m, 1H), 4.41 – 4.19 (m, 2H), 3.74 – 3.62 (m, 7H), 3.56 – 3.51 (m, 2H), 3.47 – 3.41 (m, 1H), 3.32 – 3.27 (m, 1H), 3.02 – 2.87 (m, 4H), 2.70 – 2.54 (m, 4H), 2.27 – 2.12 (m, 1H), 1.98 – 1.83 (m, 1H), 1.64 – 1.54 (m, 2H), 1.37 (s, 9H). LC-MS: [M+H]⁺ = 742.6.

Step 5: UBI-1167

Compound **UBI-1167f** (100mg, 0.135mmol) was dissolved in HCl/dioxane (10 mL), reacted at room temperature for 16 hours. The reaction solution was concentrated to obtain the target compound crude **UBI-1167g** (100mg), which was directly used in the next step. LC-MS: (M+H)⁺ = 641.7

Synthesis method of Compound UBI-1050 (NH₂-5c-B(A1))



Step 1: UBI-1050b

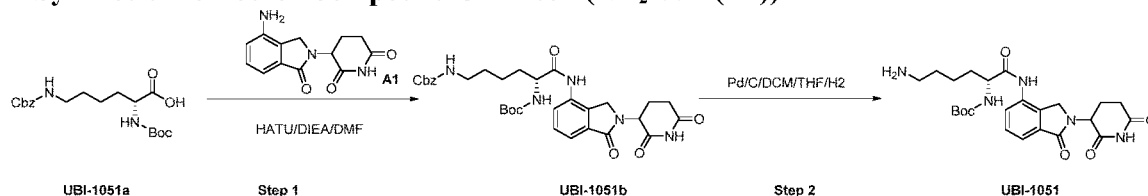
Compound **UBI-1050a** (2g, 5.4mmol), **B(A1)** (1.4g, 5.4mmol), and HATU (4.1g, 10.8mmol) were dissolved in DMF (3 mL) and DIPEA (0.3 mL), and reacted at room temperature for 16 hours. The reaction solution was directly isolated by reversed-phase column (MeOH/H₂O = 5% to 95%, 40 minutes, collected at 65%) to obtain target compound **UBI-1050b** (2g) as white solid. LCMS: (M+H)⁺ = 608.3.

Step 2: UBI-1050

Compound **UBI-1050b** (1g, 1.64mmol) and Pd/C (200 mg) were dissolved in DCM (200mL) and THF (100mL), and reacted under hydrogen atmosphere for 48 hours. After completion of the reaction, the reaction solution was concentrated, added 300 mL of MeOH/DCM=10:1 and stirred at room temperature for 3 hours, then filtered. The filtrate was concentrated to obtain the crude of target compound. The crude product was isolated and purified by reversed-phase column chromatography (MeOH/H₂O = 5% to 95%, collected at 45%) to obtain target compound **UBI-1050** (200mg) as white solid.

¹H NMR (400 MHz, DMSO-*d*₆) δ 10.06 (s, 1H), 7.81 – 7.73 (m, 1H), 7.57 – 7.44 (m, 3H), 7.21 – 7.10 (m, 1H), 5.26 – 5.12 (m, 1H), 4.44 – 4.27 (m, 2H), 4.13 (d, *J* = 29.1 Hz, 1H), 2.97 – 2.88 (m, 1H), 2.83 – 2.72 (m, 2H), 2.65 – 2.56 (m, 1H), 2.36 – 2.27 (m, 1H), 2.10 – 2.01 (m, 1H), 1.64 (s, 4H), 1.39 (s, 9H). LCMS: (M+H)⁺ = 474.2.

Synthesis method of Compound UBI-1051 (NH₂-6c-B(A1))



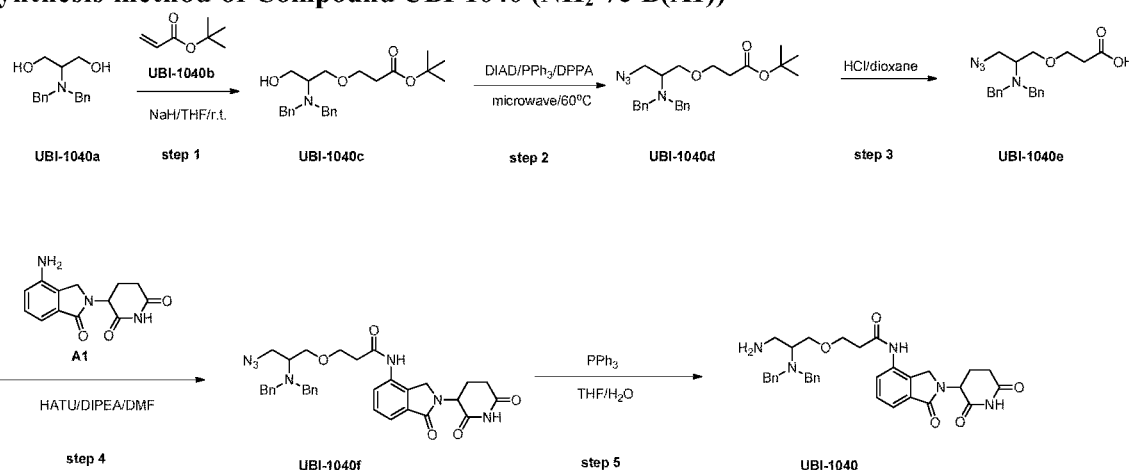
Step 1: UBI-1051b

Compound **UBI-1051a** (2g, 5.26mmol), **B(A1)** (1.36g, 5.26mmol), and HATU (4g, 10.52mmol) were dissolved in DMF (3 mL) and DIPEA (0.3 mL), and reacted at room temperature for 16 hours. The reaction solution was directly isolated by reversed-phase column (MeOH/H₂O = 5% to 95%, 40 minutes, collected at 65%) to obtain target compound **UBI-1051b** (1.8g) as white solid. LCMS: (M+H)⁺ = 622.3.

Step 2: UBI-1051

Compound **UBI-1051b** (1g, 1.64mmol) and Pd/C (200 mg) were dissolved in DCM (200mL) and THF (100mL), and reacted under hydrogen atmosphere for 48 hours. After completion of the reaction, the reaction solution was concentrated, added 300 mL solvent of MeOH/DCM=10:1 and stirred at room temperature for 3 hours, then filtered. The filtrate was concentrated to obtain the crude of target compound. The crude product was washed with methanol (3mL*3) and filtered to obtain target compound **UBI-1051** (800mg) as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.03 (s, 1H), 10.03 (s, 1H), 7.96 – 7.71 (m, 3H), 7.60 – 7.43 (m, 2H), 7.08 (dd, *J* = 7.7, 4.1 Hz, 1H), 5.16 (dt, *J* = 13.3, 5.0 Hz, 1H), 4.34 (q, *J* = 18.4, 17.9 Hz, 2H), 4.15 – 4.05 (m, 1H), 3.03 – 2.88 (m, 1H), 2.78 (t, *J* = 7.5 Hz, 2H), 2.67 – 2.56 (m, 1H), 2.37 – 2.21 (m, 1H), 2.09 – 1.99 (m, 1H), 1.79 – 1.54 (m, 4H), 1.39 (s, 11H). LCMS: (M+H)⁺ = 488.4.

Synthesis method of Compound UBI-1040 (NH₂-7c-B(A1))



Step 1: UBI-1040c

5 Compound **UBI-1040a** (5g, 18.5 mmol), **UBI-1040b** (2.4g, 18.5 mmol) were dissolved in tetrahydrofuran (100 mL), then cooled to 0°C under ice bath, and then added catalytic amount of NaH. The reaction was carried out at room temperature for 18 hours. The reaction solution was added water (30 mL), extracted with ethyl acetate (40 mL*3). The organic phases were combined, concentrated and subjected to silica gel column chromatography (ethyl acetate/petroleum ether = 1/5) to obtain product. **UBI-1040c** (3.6 g, yield 48%) as colorless oil.

Step 2: UBI-1040d

15 Compound **UBI-1040c** (800mg, 2.0 mmol), PPh₃ (1.3 g, 5.0 mmol) and DIAD (1.0 g, 5.0 mmol) were dissolved in dry tetrahydrofuran (15 mL) and cooled to 0 °C under ice bath, then added DPPA (1.4 g, 5.0 mmol) slowly. The reaction solution was heated to 60 °C by microwave and reacted for 2 hours. The reaction solution was concentrated and subjected to silica gel column chromatography (dichloromethane/petroleum ether = 1/5) to obtain product **UBI-1040d** (300 mg, yield 36%) as colorless oil.

Step 3: UBI-1040e

20 Compound **UBI-1040e** (860 mg, 2.0 mmol) was dissolved in a solution of HCl in dioxane (15 mL), reacted at room temperature for 2 hours. The reaction solution was concentrated to obtain crude product **UBI-1040e** (750 mg, yield 100%) as light pink oil. The crude product was directly used in the next step.

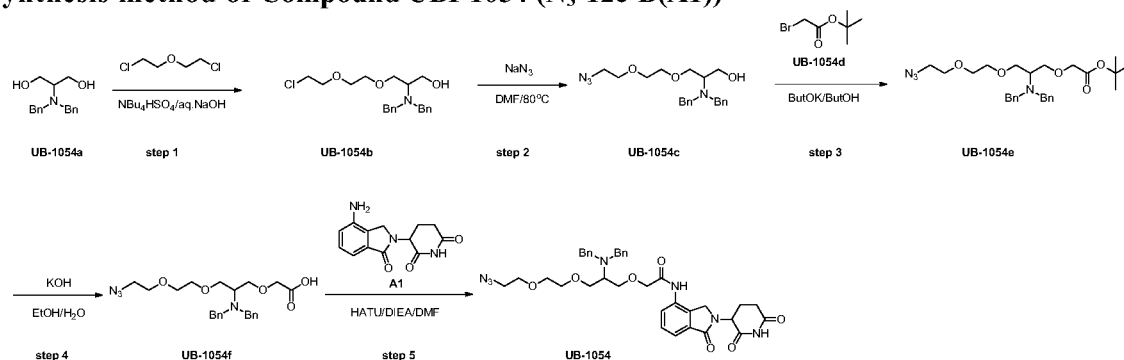
Step 4: UBI-1040f

25 Compound **UBI-1140e** (750mg, 2.0mmol), **B(A1)** (528mg, 2.0mmol), and HATU (929mg, 2.4mmol) were dissolved in DMF (8 mL) and DIPEA (0.5 mL), and the reaction was reacted at room temperature for 18 hours. The reaction solution was directly subjected to silica gel column chromatography (methanol/dichloromethane = 1/3) to obtain product **UBI-1040f** (750mg, yield 60%) as colorless oil.

Step 5: UBI-1040

30 Compound **UBI-1040f** (750 mg, 1.2 mmol) and PPh₃ (387 mg, 1.5 mmol) were dissolved in tetrahydrofuran (30 mL), reacted at room temperature for 18 hours, then added water (10 mL), and continued to react for 1 hour. The reaction solution was added water (10 mL), extracted with ethyl acetate (20 mL*3). The organic phases were combined and concentrated, purified by a reversed-phase column (methanol/water = 5% to 95%, 45 minutes, collected at 40%) to obtain product **UBI-1040** (54mg, yield 8%) as white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.90 (s, 1H), 7.87 (dd, *J* = 7.7, 4.5 Hz, 2H), 7.63 – 7.06 (m, 11H), 5.13 (dt, *J* = 13.4, 4.3 Hz, 1H), 4.34 (d, *J* = 5.9 Hz, 2H), 3.85 – 3.48 (m, 6H), 3.30 (s, 2H), 3.05 – 2.78 (m, 3H), 2.67 (d, *J* = 4.6 Hz, 2H), 2.29 – 1.81 (m, 6H).

Synthesis method of Compound UBI-1054 (N₃-12c-B(A1))



Step 1: UBI-1054b

Compound **UBI-1054a** (10 g, 37 mmol) was dissolved in dichloroether (50 mL), and then added **N(Bu)₄HSO₄** (12.5 g, 37 mmol). After the reaction was cooled to 0°C under ice bath, 50% NaOH (100 mL) was slowly added dropwise to the reaction, with dropping time of over 15 minutes. At the final reaction was carried out at 40°C for 18 hours. The reaction solution was added water (50 mL), extracted with dichloromethane (80 mL*3). The organic phases were combined and then concentrated and subjected to silica gel column chromatography (ethyl acetate/petroleum ether = 1/5) to obtain light yellow product **UBI-1054b** (8.8 g, yield 63%).

¹H NMR (400 MHz, chloroform-*d*) δ 7.43 – 7.14 (m, 10H), 3.86 (d, *J* = 13.4 Hz, 2H), 3.81 – 3.75 (m, 3H), 3.72 – 3.52 (m, 11H), 3.29 – 3.00 (m, 1H), 2.89 (s, 1H).

Step 2: UBI-1054c

Compound **UBI-1054b** (7.5 g, 20 mmol) was dissolved in DMF (50 mL), then added NaN₃ (1.6 g, 24 mmol). The reaction was heated to 80°C reacted for 4 hours. The reaction solution was added iced water (30 mL), then extracted with dichloromethane (50 mL*3). The organic phases were combined, then dried over anhydrous sodium sulfate, and then concentrated to obtain crude product as yellow oil. The crude product was subjected to silica gel column chromatography (ethyl acetate/petroleum ether = 1/4) to obtain product **UBI-1054c** (10 g, yield 100%) as light yellow oil. LCMS [*M*+1]⁺ = 384.0

Step 3: UBI-1054e

Compound **UBI-1054c** (4.0 g, 10.42 mmol) was dissolved in ButOH (150 mL), then added ButOK (1.4 g, 12.50 mmol) and **UBI-1054d** (2.0 g, 10.42 mmol) at 0 °C. The reaction was reacted at room temperature for 18 hours. The reaction solution was concentrated, then added water (50 mL), and extracted with ethyl acetate (80 mL*3). The organic phases were combined, then dried over anhydrous sodium sulfate, and concentrated to obtain crude product. The crude product was subjected to silica gel column chromatography (ethyl acetate/petroleum ether = 1/4) to obtain product **UBI-1054e** (520mg, yield 10%) as white solid.

¹H NMR (400 MHz, chloroform-*d*) δ 7.43 – 7.15 (m, 10H), 3.96 – 3.89 (m, 2H), 3.81 (s, 4H), 3.77 – 3.54 (m, 10H), 3.35 (t, *J* = 5.1 Hz, 2H), 3.13 (p, *J* = 6.0 Hz, 1H), 1.48 (s, 9H).

Step 4: UBI-1054f

Compound **UBI-1054e** (520 mg, 1.04 mmol) was dissolved in EtOH (10 mL) and water (1 mL), and added KOH (234 mg, 4.18 mmol) at 0 °C, and reacted at room temperature for 1 hour. The reaction solution was concentrated to obtain crude product. The crude product was added water (10 mL), acidified to pH=3 with 3N HCl, and then lyophilized to obtain 800 mg crude product. The crude product was eluted with methanol/dichloromethane=1/10 (20 mL) and concentrated to obtain product **UBI-1054f** (500 mg, yield 100%) as white solid.

Step 5: UBI-1054

Compound **UBI-1054f** (250mg, 0.57mmol), **B(A1)** (146 mg, 0.57mmol), and HATU (258mg, 4.27)

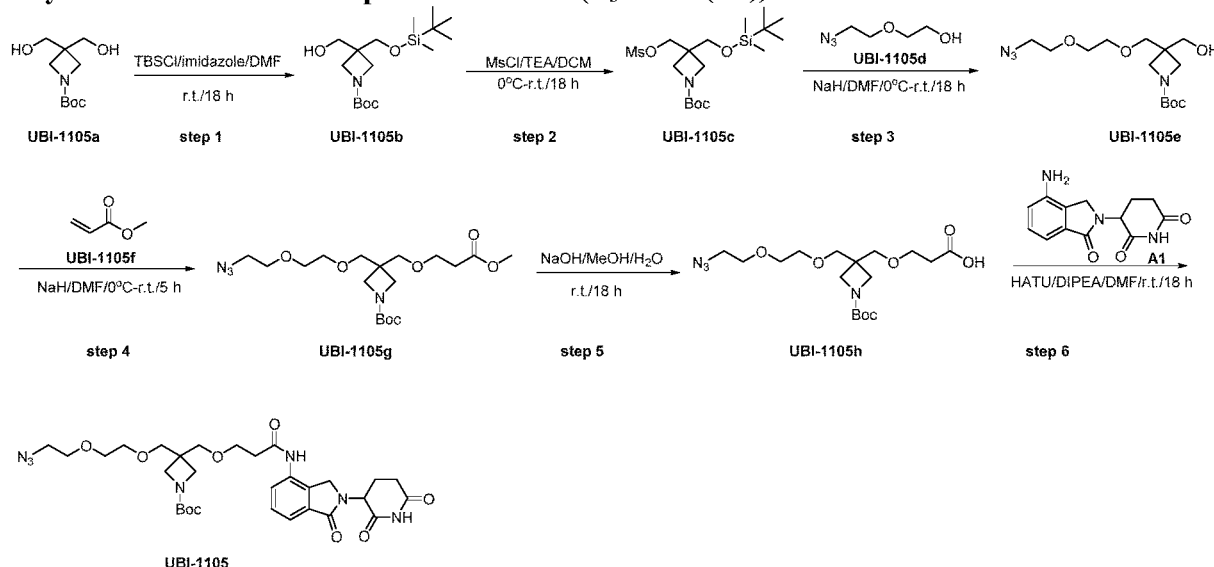
were dissolved in DMF (8 mL) and DIPEA (0.5 mL), and reacted at room temperature for 18 hours. The reaction solution was directly purified via reversed-phase column (methanol/water = 5 % to 95 %, collected at 80%) to obtain product **UBI-1054** (50mg, yield 13%) as yellow solid.

¹H NMR (400 MHz, DMSO-*d*₆) δ 11.00 (s, 1H), 9.63 (d, *J* = 5.2 Hz, 1H), 7.77 (ddd, *J* = 7.6, 3.8, 1.3 Hz, 1H), 7.62 – 7.46 (m, 2H), 7.38 (d, *J* = 8.1 Hz, 4H), 7.28 (t, *J* = 7.4 Hz, 4H), 7.23 – 7.12 (m, 2H), 5.12 (dd, *J* = 13.2, 5.2 Hz, 1H), 4.41 – 4.24 (m, 2H), 4.09 (d, *J* = 1.5 Hz, 2H), 3.89 – 3.45 (m, 16H), 3.08 – 2.82 (m, 2H), 2.58 (s, 1H), 2.31 – 2.11 (m, 1H), 2.03 – 1.90 (m, 1H).

Step 6: compound **UBI-1054** was reduced by the method in accordance with the preparation of compound **UBI-1040**.

Synthesis method of Compound Linker d-Ligand B(A1)

Synthesis method of Compound **UBI-1105** (**N**₃-13d-B(A1)):



Step 1: **UBI-1105b**

Compound **UBI-1105a** (1 g, 4.6 mmol) was dissolved in anhydrous DMF (20 mL), added imidazole (1.56 g, 23 mmol) and TBSCl (829 mg, 5.5 mmol) after cooling to 0°C, and reacted at 0°C for 1 hour, then reacted at room temperature overnight. The reaction solution was concentrated and isolated by silica gel column chromatography (dichloromethane /methanol = 20/1, *R*_f=0.5) to obtain product **UBI-1105b** (500 mg, yield 32%) as white solid. ¹H NMR (400 MHz, chloroform-*d*) δ 3.73 (d, 4H), 3.58 – 3.60 (d, 2H), 3.51 – 3.53 (d, 2H), 1.35 (s, 9H), 0.82 (s, 9H), 0.0 (s, 6H). LCMS: [*M*+*H*]⁺ = 332

Step 2: **UBI-1105c**

Compound **UBI-1105b** (800 mg, 2.4 mmol) was dissolved in anhydrous dichloromethane (30 mL), and then added triethylamine (48 mg, 4.8 mmol) and MsCl (304 mg, 2.64 mmol) at 0°C, and reacted at 0°C for 1 hour, then reacted at room temperature overnight. The reaction was quenched with water (50 mL), then extracted with dichloromethane (50 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 20/1) to obtain product **UBI-1105c** (1 g, yield 100%) as white solid. ¹H NMR (400 MHz, chloroform-*d*) δ 4.27(s, 2H), 3.65 (s, 2H), 3.62 (s, 4H), 2.97 (s, 3H), 1.38 (s, 9H), 0.82 (s, 9H), 0.0 (s, 6H). LCMS: [*M*+*H*]⁺ = 310

Step 3: **UBI-1105e**

Compound **UBI-1105d** (445 mg 3.4 mmol) was dissolved in anhydrous DMF (10 mL), added 60% NaH (272 mg, 6.8 mmol) in batches at 0°C, reacted for 1 hour at 0°C, then added **UBI-1105c** (1 g crude, 1.7

mmol) to dissolve, and reacted overnight. The reaction was quenched with water (20 mL), then extracted with dichloromethane (50 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane /methanol = 94/6, R_f = 0.7) to obtain product **UBI-1105e** (450 mg, yield 32%) as yellow oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 4.62 (s, 2H), 3.98 (s, 2H), 3.61 (m, 2H), 3.58 (m, 2H), 3.54 – 3.44 (m, 6H), 3.39 (m, 2H), 1.36 (s, 9H).

Step 4: UBI-1105g

Compound **UBI-1105e** (400 mg, 1.21 mmol), **UBI-1105f** (104 mg, 1.21 mmol) were dissolved in anhydrous DMF (5 mL), added 60% NaH (5 mg, 0.12 mmol) in batches at 0 °C. After that, reacted for 4 hours, the reaction was quenched with water (20 mL), then extracted with ethyl acetate (20 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane /methanol = 94/6, R_f = 0.7) to obtain product **UBI-1105g** (300 mg, yield 59%) as yellow oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 4.62 (s, 2H), 3.98 (s, 2H), 3.64 (m, 3H), 3.62 – 3.56 (m, 5H), 3.56 – 3.43 (m, 6H), 3.39 (d, J = 5.0 Hz, 2H), 3.24 – 3.20 (m, 1H), 2.54 (t, J = 6.2 Hz, 2H), 1.36 (d, J = 1.1 Hz, 9H).

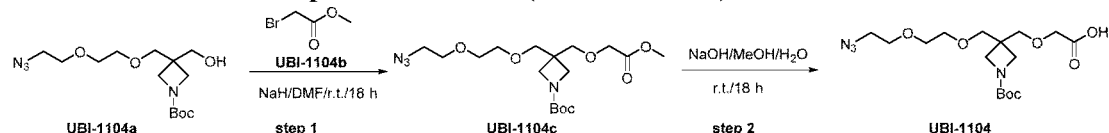
Step 5: UBI-1105h

Compound **UBI-1105g** (260 mg, 0.62 mmol) was dissolved in methanol (5 mL) and added 2M NaOH (1.6 mL, 3.1 mmol), reacted at room temperature overnight. The reaction solution was concentrated and added saturated brine (15 mL), extracted with ether (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain crude product **UBI-1105h** (120 mg, yield 46%) as yellow oil, which is directly used in the next step.

Step 6: UBI-1105

Compound **UBI-1105h** (120 mg, 0.3 mmol), **B(A1)** (93 mg, 0.36 mmol), HATU (137 mg, 0.36 mmol), DIPEA (0.3 mL) were dissolved in DMF (3 mL), reacted at room temperature overnight. The reaction was quenched with water (20 mL) and extracted with ethyl acetate (50 mL*3). The organic layer was combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol=90/10) and reversed phase (water/methanol=20/80) to obtain crude product **UBI-1105** (12 mg, yield 6%) as white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.01 (s, 1H), 9.86 (s, 1H), 7.80 (m, 1H), 7.54 – 7.45 (m, 2H), 5.15 (dd, J = 13.3, 5.1 Hz, 1H), 4.39 – 4.29 (m, 2H), 3.75 (m, 2H), 3.65 – 3.56 (m, 8H), 3.55 – 3.47 (m, 8H), 3.37 (m, 2H), 2.97 – 2.88 (m, 1H), 2.63 (m, 2H), 2.36 – 2.27 (m, 2H), 2.02 (m, 1H), 1.34 (d, J = 15.4 Hz, 9H). LCMS: [M+H]⁺ = 644

Synthesis method of Compound UBI-1104 (N₃-12d-COOH):



Step 1: UBI-1104c

Compound **UBI-1104a** (700 mg 2.1 mmol) was dissolved in anhydrous DMF (5 mL), added 60% NaH (252 mg, 6.3 mmol) in batches at 0 °C, reacted for 1 hour at 0 °C. Compound **UBI-1104b** (324 mg, 2.1 mmol) was added at 0 °C, and then reacted overnight. The reaction was quenched by adding water (20 mL), then extracted with dichloromethane (50 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 90/10) to obtain crude product **UBI-1104c** (300 mg, yield

35%) as yellow oil. ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 4.63 (s, 2H), 3.98 (s, 2H), 3.72 – 3.46 (m, 11H), 3.39 (m, 1H), 2.90 (m, 3H), 2.59 – 2.53 (m, 2H), 1.36 (s, 9H). LCMS: $[\text{M}+\text{H}]^+ = 403$.

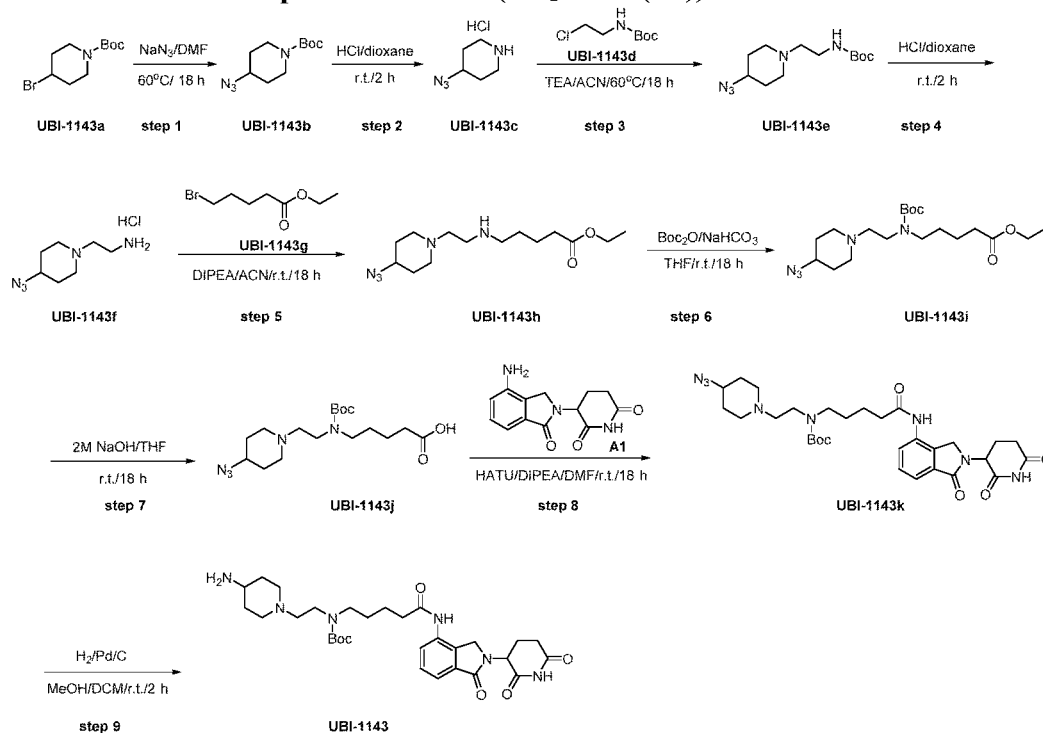
Step 2: UBI-1104

Compound **UBI-1104c** (200 mg, 0.49 mmol) was dissolved in methanol (5 mL) and added 2M NaOH (1.24 mL, 2.48 mmol). Reacted at room temperature overnight. The reaction solution was concentrated, added saturated brine (15 mL), extracted with ether twice (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, and then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain crude product **UBI-1104** (80 mg, yield 42%) as yellow oil.

Step 3: compound UBI-1104 was reduced by the method in accordance with the preparation of compound UBI-1040.

Synthesis method of Compound Linkere- Ligand B(A1)/B(B1)

Synthesis method of Compound UBI-1143 (NH_2 -12e-B(A1))



Step 1: UBI-1143b

Compound **UBI-1143a** (10 g, 37.8 mmol) was dissolved in anhydrous DMF (50 mL), added sodium azide (2.95 g, 45.4 mmol), reacted 60°C overnight. The reaction was quenched by adding water (20 mL) and extracted with ethyl acetate (80 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (petroleum ether/ethyl acetate = 9/1) to obtain product **UBI-1143b** (8.6 g, yield 100%) as colorless oil. ^1H NMR (400 MHz, $\text{chloroform}-d$) δ 3.82 (m, 2H), 3.61 – 3.52 (m, 1H), 3.09 (m, 2H), 1.87 (m, 2H), 1.62 – 1.53 (m, 2H), 1.46 (s, 9H). LCMS: $[\text{M}-56]^+ = 171$

Step 2: UBI-1143c

Compound **UBI-1143b** (8.6 g, 38 mmol) was added to a solution of 4M hydrochloric acid in dioxane (47.5 mL) in ice bath, and reacted at room temperature for 2 hours. The reaction solution was concentrated to obtain crude product **UBI-1143c** (6.3 g, yield 100%) as yellow solid, which was directly used in the next step.

Step 3: UBI-1143e

Compound **UBI-1143c** (5.6 g, 34.4 mmol), **UBI-1143d** (8.8 g, 49 mmol), triethylamine (11 mL, 78 mmol) were dissolved in acetonitrile (100 mL), then reacted 60 °C overnight. The reaction was quenched with water (50 mL), extracted with ethyl acetate (50 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (petroleum ether/ethyl acetate = 70/30, R_f = 0.3) to obtain product **UBI-1143e** (3.1 g, yield 33%) as yellow oil.

Step 4: UBI-1143f

Compound **UBI-1143e** (3.1 g, 11.5 mmol) was added to a solution of 4M hydrochloric acid in dioxane (35 mL) under ice bath, and reacted at room temperature for 2 hours. The reaction solution was concentrated to obtain product **UBI-1143f** (2.36 g, yield 100%) as yellow oil, which was directly used in the next step.

Step 5: UBI-1143h

Compound **UBI-1143f** (1.52 g, 7.3 mmol) was dissolved in acetonitrile (60 mL), added DIPEA (6 mL, 36.8 mmol), **UBI-1143g** (1.54 g, 7.3 mmol), and reacted at room temperature overnight. The reaction was quenched with water (50 mL), extracted with ethyl acetate (50 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane /methanol = 40/60, R_f = 0.7) to obtain **UBI-1143g** (400 mg, yield 18%) as yellow solid.

¹H NMR (400 MHz, DMSO-*d*₆) δ 4.06 (q, J = 7.1 Hz, 2H), 3.56 (m, 1H), 3.28 (m, 1H), 2.98 (m, 2H), 2.89 (t, J = 7.5 Hz, 2H), 2.73 (m, 2H), 2.55 (m, 2H), 2.34 (t, J = 7.1 Hz, 2H), 2.18 (m, 2H), 1.85 (m, 2H), 1.68 – 1.49 (m, 6H), 1.31 – 1.23 (m, 3H), 1.19 (t, J = 7.1 Hz, 3H). LCMS: [M+H]⁺ = 298

Step 6: UBI-1143i

Compound **UBI-1143h** (400 mg, 1.34 mmol) was dissolved in tetrahydrofuran (10 mL), added NaHCO₃ (338 mg, 1.61 mmol) and Boc₂O (352 mg, 1.61 mmol), and reacted at room temperature overnight. After being quenched by adding water (20 mL), the reaction solution was extracted with ethyl acetate (20 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 90/10) to obtain product **UBI-1143i** (400 mg, yield 75%) as yellow oil. LCMS: [M+H]⁺ = 398

Step 7: UBI-1143j

Compound **UBI-1143i** (400 mg, 1 mmol) was dissolved in tetrahydrofuran (10 mL), added 2M NaOH (5 mL, 10 mmol), and reacted at room temperature overnight. The reaction was added saturated brine (15 mL) and extracted with ether twice (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain crude product **UBI-1143j** (380 mg, yield 100%) as colorless oil. LCMS: [MS+H]⁺=370

Step 8: UBI-1143k

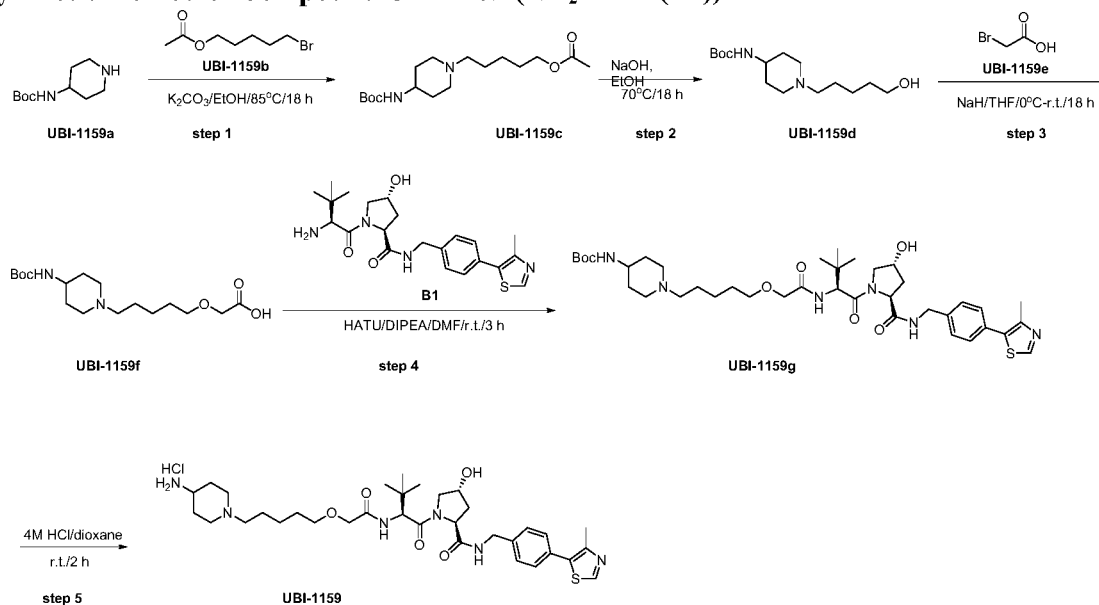
Compound **UBI-1143j** (370 mg, 1 mmol), **B(A1)** (260 mg, 1 mmol), HATU (570 mg, 1.5 mmol), and DIPEA (1 mL) were dissolved in DMF (5 mL), reacted at room temperature overnight. The reaction was quenched with water (50 mL), extracted with ethyl acetate (50 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 93/7) to obtain crude product **UBI-1143k** (360 mg, yield 59%) as colorless oil. LCMS: [M+H]⁺ = 611 ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.03 (s, 1H), 9.78 (s, 1H), 7.82 (dd, J = 7.1, 1.9 Hz, 1H), 7.50 (m, 2H), 5.16 (m, 1H), 4.44 – 4.27 (m, 2H), 3.61 (m, 1H), 3.48 (m,

1H), 3.17 (m, 4H), 2.98 – 2.87 (m, 1H), 2.78 – 2.57 (m, 2H), 2.43 – 2.22 (m, 4H), 2.21 – 2.00 (m, 3H), 1.81 (m, 2H), 1.47 (m, 13H), 1.28 – 1.22 (m, 2H).

Step 9: UBI-1143

Compound **UBI-1143k** (360 mg, 0.58 mmol) was dissolved in dichloromethane (20 mL) and methanol (2 mL), added Pd/C (250 mg), and reacted for 2 hours under hydrogen atmosphere. Filtered through Celite, the filtrate was concentrated to obtain product **UBI-1143** (300 mg, yield 88%) as yellow solid, which was directly used in the next step. LCMS: $[M+H]^+ = 585$

Synthesis method of Compound UBI-1159 (NH₂-12e-B(B1)):



Step 1: UBI-1159c

Compound **UBI-1159a** (2 g, 10 mmol), **UBI-1159b** (2 mL, 12 mmol), anhydrous potassium carbonate (11 g, 80 mmol) were dissolved in acetonitrile (40 mL), reacted at 85 °C overnight. The reaction was cooled to room temperature, poured into water and then extracted with ethyl acetate (40 mL*2). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 10/1) to obtain product **UBI-1159c** (1.9 g, yield 58%) as white solid. LCMS: $[M+H]^+ = 329$

Step 2: UBI-1159d

Compound **UBI-1159c** (1.9 g, 5.8 mmol), 2N NaOH (8.7 mL, 17.4 mmol) were dissolved in ethanol (30 mL), and reacted at 70°C overnight. The reaction was cooled to room temperature, poured into water and then extracted with ethyl acetate (40 mL*2). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 10/1) to obtain product **UBI-1159c** (1.3 g, yield 78%) as white solid. LCMS: $[M+H]^+ = 287$

Step 3: UBI-1159

Compound **UBI-1159d** (1 g, 3.5 mmol) was dissolved in anhydrous tetrahydrofuran (10 mL) and cooled to 0°C, added 60% NaH (276 mg, 6.9 mmol) in batches, reacted at 0 °C for half an hour. Then the compound **UBI-1159e** (324 mg, 2.3 mmol) was dissolved in anhydrous tetrahydrofuran (2 mL) and added to the above reaction solution, and reacted at room temperature overnight. The reaction was poured into water and then extracted with ethyl acetate (40 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl. The crude product was concentrated and isolated by silica gel column chromatography

(dichloromethane /methanol = 90/10) to obtain product as white solid (290 mg, yield 36%) . ¹H NMR (400 MHz, DMSO-*d*₆) δ 6.93 (d, 1H), 3.89 (s, 2H), 3.44 (m, 3H), 3.36 (m, 2H), 3.10 (m, 2H), 2.64 (m, 2H), 1.85 – 1.75 (m, 2H), 1.53 (m, 6H), 1.38 (s, 9H), 1.29 (m, 2H). LCMS: [M+H]⁺ = 345

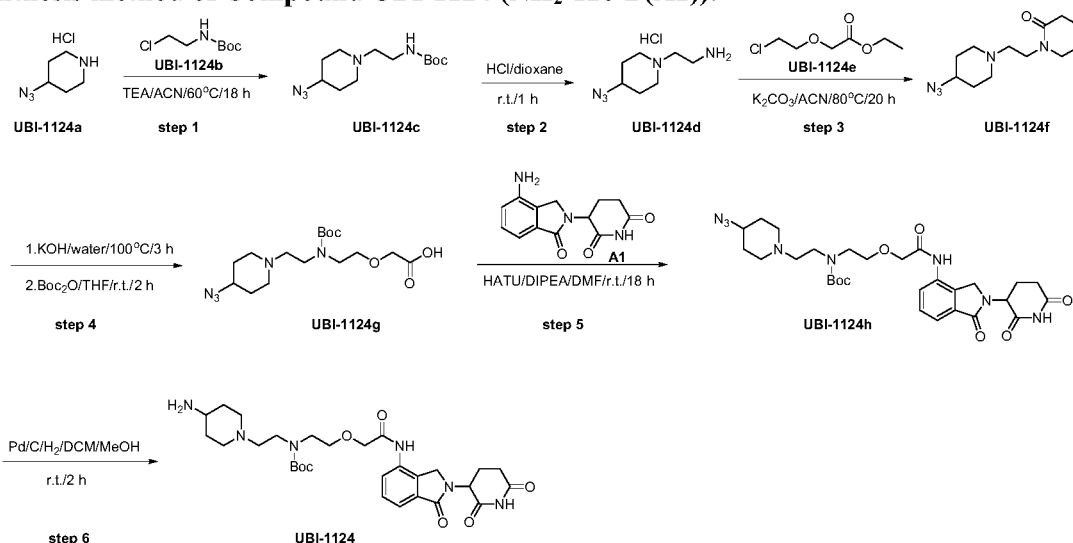
Step 4: UBI-1159g

5 Compound **UBI-1159f** (190 mg, 0.55 mmol), **B(B1)** (237 mg, 0.55 mmol), HATU (250 mg, 0.66 mmol), and DIPEA (1 mL) were dissolved in DMF (5 mL), reacted at room temperature for 3 hours. The reaction was poured into water (30 ml) and then extracted with ethyl acetate (40 mL*3). The combined organic phases were with 1M HCl, saturated NaHCO₃. The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane /methanol = 10/1, R_f = 0.3) to obtain **UBI-1159g** (180 mg, yield 43%) as white solid. ¹H NMR (400 MHz, chloroform-*d*) δ 8.68 (s, 1H), 7.36 (s, 4H), 5.35 (t, 1H), 4.73 (s, 1H), 4.67 – 4.58 (m, 1H), 4.53 (s, 1H), 4.45 (d, 1H), 4.00 (s, 1H), 3.65 (m, 5H), 3.03 (m, 3H), 2.52 (m, 2H), 2.30 (s, 3H), 2.27 – 2.20 (m, 1H), 2.09 (m, 1H), 2.01 (m, 5H), 1.86 (m, 3H), 1.59 (m, 3H), 1.41 (m, 5H), 1.25 (s, 9H), 1.01 (s, 9H), 0.97 – 0.84 (m, 5H). LCMS: [M+H]⁺ = 757

15 Step 5: UBI-1159

Compound **UBI-1159g** (50 mg, 0.066 mmol) was added to 4M hydrochloride in dioxane (1 mL) under ice bath, and reacted at room temperature for 2 hours. The reaction solution was concentrated to obtain crude product **UBI-1159h** (50 mg, 100% yield) as yellow oil. LCMS: [M+H]⁺ = 657

20 Synthesis method of Compound UBI-1124 (NH₂-12e-B(A1)):



Step 1: UBI-1124c

25 Compound **UBI-1124a** (5.6 g, 34.4 mmol), **UBI-1124b** (8.8 g, 49 mmol), and triethylamine (11 mL, 78 mmol) were dissolved in acetonitrile (100 mL), reacted at 60°C overnight. The reaction was cooled to room temperature, poured into water and then extracted with ethyl acetate (50 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (petroleum ether/ethyl acetate = 70/30, R_f = 0.3) to obtain product **UBI-1143e** (3.1 g, yield 33%) as yellow oil.

Step 2: UBI-1124d

30 Compound **UBI-1124c** (3.1 g, 11.5 mmol) was added to a solution of 4M hydrochloric acid in dioxane (35 mL) under ice bath, and reacted at room temperature for 2 hours. The reaction solution was concentrated to obtain product **UBI-1124d** (2.36 g, yield 100%) as yellow oil.

Step 3: UBI-1124f

Compound **UBI-1124d** (1.5g, 8.8 mmol) was dissolved in acetonitrile (150 mL), added anhydrous potassium carbonate (3.64 g, 26.4 mmol) and **UBI-1124e** (1.7 g, 10.2 mmol), reacted at 80 °C overnight. The reaction was cooled to room temperature and filtered. The filtrate was concentrated and isolated by silica gel column chromatography (dichloromethane /methanol = 97/3, R_f = 0.2) to obtain product **UBI-1124f** (260 mg, yield 11%) as a yellow oil. LCMS: [M+H]⁺ = 254

Step 4: UBI-1124g

Compound **UBI-1124f** (800 mg, 3.15 mmol) was dissolved in water (5 mL), added potassium hydroxide (530 mg, 9.47 mmol), and reacted at 100°C for 3 hours. After completion of reaction, the reaction was cooled to room temperature, added Boc₂O (1.37 g, 6.3 mmol) and tetrahydrofuran (5 mL), and reacted at room temperature for 2 hours. The reaction added saturated brine (15 mL), extracted with ether twice (20 mL*2). The aqueous phase was acidified to pH=5 using 1M HCl, and then extracted with a mixed solvent of ethyl acetate and acetone (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain product **UBI-1124g** (1.1 g, yield 100%) as colorless oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 12.58 (s, 1H), 3.61 – 3.52 (m, 6H), 3.35 (m, 4H), 3.24 (m, 2H), 3.06 (m, 2H), 2.11 (m, 2H), 1.77 (m, 2H), 1.41 (d, *J* = 5.0 Hz, 9H). LCMS: [MS+H]⁺=372.2

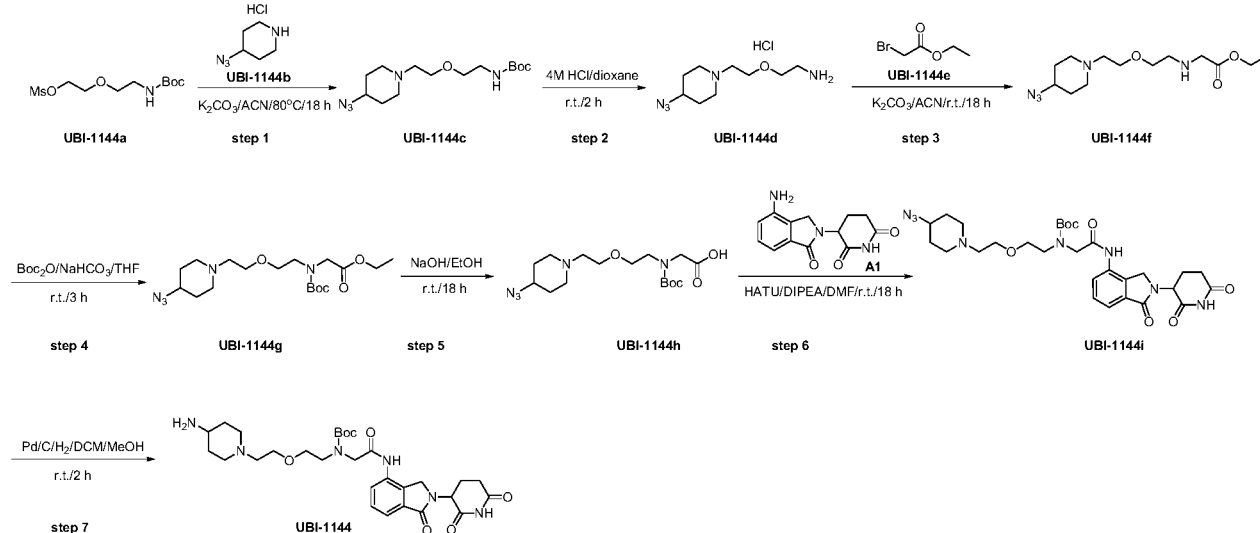
Step 5: UBI-1124h

Compound **UBI-1124g** (500 mg, 1.34 mmol), **B(A1)** (347 mg, 1.34 mmol), HATU (763 mg, 2 mmol), and DIPEA (1 mL) were dissolved in DMF (5 mL), reacted at room temperature overnight. The reaction was quenched with water (50 mL) and extracted with ethyl acetate (50 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 93/7) to obtain product **UBI-1124h** (370 mg, yield 45%) as yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.03 (s, 1H), 9.85 (m, 1H), 7.78 (m, 1H), 7.62-7.45 (m, 2H), 5.16 (m, 1H), 4.45-4.30 (m, 2H), 4.16 (m, 3H), 3.69-3.56 (m, 4H), 3.41 (m, 2H), 3.30 (m, 2H), 3.15 (m, 2H), 3.04-2.84 (m, 2H), 2.69 – 2.55 (m, 2H), 2.33 (m, 2H), 2.16 (m, 1H), 2.08 – 1.89 (m, 3H), 1.39 (s, 9H). LCMS: [M+H]⁺ = 613

Step 6: UBI-1124

Compound **UBI-1124h** (370 mg, 0.6 mmol) was dissolved in a mixed solvent of dichloromethane (10 mL) and methanol (2 mL), added Pd/C (300 mg), and reacted for 2 hours under hydrogen atmosphere. Filtered through Celite, the filtrate was concentrated to obtain product **UBI-1124** (370 mg, yield 100%) as yellow solid, which was directly used in the next step. LCMS: [M+H]⁺ = 587.1

Synthesis method of Compound UBI-1144 (NH₂-12e-B(A1)):



Step 1: UBI-1144c

Compound **UBI-1144a** (1.5 g, 6.8 mmol) was dissolved in acetonitrile (30 mL), added anhydrous potassium carbonate (2.82 g, 20.4 mmol) and **UBI-1144b** (1.14 g, 6.8 mmol), reacted at 80 °C overnight. The reaction was cooled to room temperature and filtered. The filtrate was concentrated and isolated by silica gel column chromatography (dichloromethane /methanol = 95/5) to obtain product **UBI-1144c** (1.1 g, yield 51%) as a yellow oil. ¹H NMR (400 MHz, chloroform-*d*) δ 5.22 (s, 1H), 3.54 (m, 4H), 3.42 (m, 1H), 3.30 (m, 2H), 2.81 (m, 2H), 2.57 (m, 2H), 2.23 (m, 2H), 1.91 (m, 2H), 1.71 (m, 2H), 1.45 (s, 9H). LCMS: [M+H]⁺ = 314

Step 2: UBI-1144d

Compound **UBI-1144c** (1.1 g, 3.5 mmol) was added to a solution of 4M hydrochloric acid in dioxane (5 mL) under ice bath, and reacted at room temperature for 2 hours. The reaction solution was concentrated to obtain product **UBI-1144d** (874 mg, yield 100%) as yellow oil, which was directly used in the next step.

Step 3: UBI-1144f

Compound **UBI-1144d** (1.5 g, 6.8 mmol) was dissolved in acetonitrile (30 mL), added anhydrous potassium carbonate (2.82 g, 20.4 mmol) and **UBI-1144e** (1.14 g, 6.8 mmol), reacted at RT overnight. The reaction was filtered. The filtrate was concentrated and isolated by silica gel column chromatography (dichloromethane /methanol = 95/5) to obtain product **UBI-1144f** (670 mg, yield 33%) as a yellow oil. LCMS: [M+H]⁺ = 300

Step 4: UBI-1144g

Compound **UBI-1144c** (670 mg, 2.2 mmol) was dissolved in tetrahydrofuran (10 mL) and added sodium bicarbonate (554 mg, 6.6 mmol) and Boc₂O (590 mg, 2.7 mmol). Reacted at room temperature for 3 hours, the reaction was quenched with water (20 mL), extracted with ethyl acetate (20 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 90/10) to obtain product **UBI-1144g** (800 mg, yield 91%) as yellow oil. LCMS: [M+H]⁺ = 400

Step 5: UBI-1144h

Compound **UBI-1144g** (800 mg, 2 mmol) was dissolved in ethanol(10 mL), added 2M NaOH (4 mL, 8 mmol), and reacted at room temperature overnight. The reaction solution was concentrated, and added saturated brine (15 mL) and extracted with ether (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, and then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain crude product **UBI-1144h** (650 mg, yield 87%) as colorless oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.62 (s, 1H), 3.87 (m, 3H), 3.71 – 3.64 (m, 2H), 3.50 (m, 2H), 3.36 (m, 2H), 3.17 (m, 2H), 3.04 (m, 2H), 2.89 (m, 2H), 2.01 (m, 2H), 1.74 (m, 2H), 1.37 (d, J = 19.6 Hz, 9H). LCMS: [MS+H]⁺=372.2

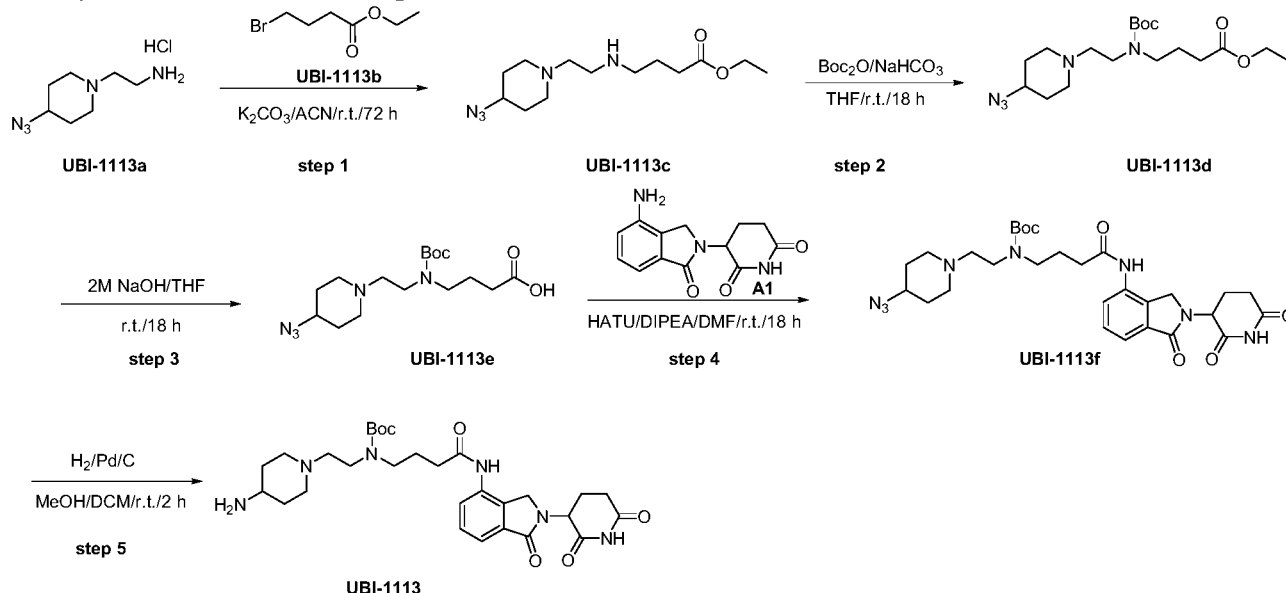
Step 6: UBI-1144i

Compound **UBI-1144h** (300 mg, 0.8 mmol), **B(A1)** (251 mg, 0.96 mmol), HATU (456 mg, 1.2 mmol), and DIPEA (1 mL) were dissolved in DMF (5 mL), reacted at room temperature overnight. The reaction was quenched with water (50 mL) and extracted with ethyl acetate (50 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 93/7) to obtain product **UBI-1144i** (250 mg, yield 51%) as yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.04 (s, 1H), 9.88 (s, 1H), 9.26 (s, 1H), 7.88-7.72 (m, 1H), 7.59-7.46 (m, 2H), 5.23-5.13 (m, 1H), 4.43-4.27 (m, 2H), 4.08-3.98 (m, 3H), 3.80-3.39 (m, 8H), 3.15-2.88 (m, 3H), 2.70-2.58 (m, 2H), 2.35-2.18 (m, 2H), 2.08 (m, 2H), 1.90 (m, 1H), 1.68 (m, 1H), 1.35 (s, 9H). LCMS: [M+H]⁺ = 613

Step 7: UBI-1144

Compound **UBI-1144i** (250 mg, 0.4 mmol) was dissolved in a mixed solvent of dichloromethane (10 mL) and methanol (2 mL), added Pd/C (150 mg), and reacted for 2 hours under hydrogen atmosphere. Filtered through Celite, the filtrate was concentrated to obtain product **UBI-1144** (210 mg, yield 89%) as yellow solid, which was directly used in the next step. LCMS: $[M+H]^+ = 587.1$

Synthesis method of Compound UBI-1113 (NH₂-11e-B(A1))



Step 1: UBI-1113c

Compound **UBI-1113a** (1 g, 3.7 mmol) was dissolved in acetonitrile (30 mL), anhydrous potassium carbonate (1.5 g, 11.1 mmol) and **UBI-1113b** (770 mg, 3.7 mmol) were added, and reacted at room temperature for 72 hours. The reaction was filtered at room temperature, the filtrate was concentrated and isolated by silica gel column chromatography (dichloromethane/methanol = 95/5) to obtain product **UBI-1113c** (600 mg, yield 57%) as yellow oil. LCMS: $[M+H]^+ = 284$

Step2: UBI-1113d

Compound **UBI-1113c** (600 mg, 2.1 mmol) was dissolved in tetrahydrofuran (10 mL), added sodium bicarbonate (530 mg, 6.3 mmol) and Boc_2O (545 mg, 2.5 mmol), and reacted at room temperature overnight. The reaction was quenched with water (20 mL), extracted with ethyl acetate (20 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 90/10) to obtain product **UBI-1113d** (600 mg, yield 74%) as yellow oil. LCMS: $[M+H]^+ = 384$

Step 3: UBI-1113e

Compound **UBI-1113d** (400 mg, 1 mmol) was dissolved in tetrahydrofuran (10 mL), added 2M NaOH (5 mL, 10 mmol), and reacted at room temperature overnight. The reaction solution was concentrated, added saturated brine (15 mL), extracted with ether twice (20 mL*2). The aqueous phase was acidified to pH=6 using 1M HCl, then extracted with ethyl acetate (40 mL*3). The organic phases were combined, dried over anhydrous sodium sulfate, filtered and concentrated to obtain crude product **UBI-1113e** (530 mg, yield 100%) as colorless oil, which was directly used in the next step. ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 12.00 (s, 1H), 3.54 (m, 2H), 3.17 (m, 3H), 2.49 – 2.41 (m, 2H), 2.18 (m, 2H), 1.91 (m, 5H), 1.76 – 1.63 (m, 3H), 1.40 (s, 9H). LCMS: $[M+H]^+ = 356.2$

Step 4: UBI-1113f

Compound **UBI-1113e** (530 mg, 1.5 mmol), **B(A1)** (460 mg, 1.8 mmol), HATU (855 mg, 2.2 mmol),

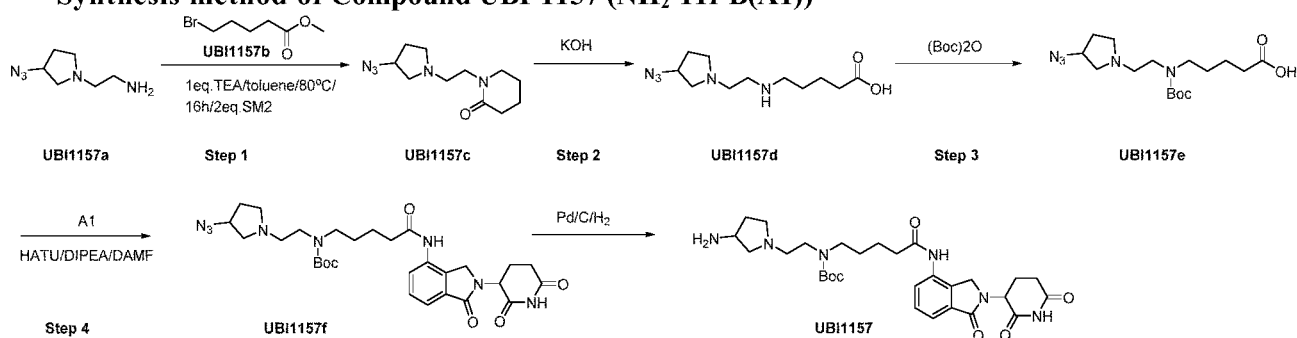
and DIPEA (1.2 mL) were dissolved in DMF (5 mL), reacted at room temperature overnight. The reaction was quenched with water (50 mL), extracted with ethyl acetate (50 mL*3). The organic layers were combined, dried over anhydrous sodium sulfate and concentrated. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 93/7) to obtain product **UBI-1113f** (350 mg, yield 39%) as white solid. LCMS: $[M+H]^+ = 597$

Step 5: UBI-1113

Compound **UBI-1113f** (350 mg, 0.58 mmol) was dissolved in a mixed solvent of dichloromethane (10 mL) and methanol (2 mL), added Pd/C (250 mg), and reacted for 2 hours under hydrogen atmosphere. Filtered through Celite, the filtrate was concentrated to obtain product **UBI-1113** (300 mg, yield 91%) as white solid, which was directly used in the next step. LCMS: $[M+H]^+ = 571$

Synthesis method of Compound Linker f - Ligand B(A1)

Synthesis method of Compound UBI-1157 (NH₂-11f-B(A1))



Step 1: UBI-1157c

Compound **UBI-1157a** (700mg, 4.48mmol), and **UBI-1157b** (1.74 g, 8.98mmol) were dissolved in triethylamine (452mg, 4.48mmol) and toluene (10mL), reacted at 80°C for 18 hours. The reaction solution was concentrated and isolated by silica gel column chromatography (petroleum ether/ethyl acetate = 50% to 100%, then MeOH/DCM=0% to 10%) to obtain the target compound **UBI-1158c** (1g) as colorless oil.

LC-MS: $[M+H]^+ = 238.5$

Step 2: UBI-1157d

Compound **UBI-11157c** (600mg, 2.5mmol), KOH (708mg, 12.5mmol) were dissolved in water (3mL), and refluxed at 100°C for 1 hours. After cooling, the reaction solution was extracted with ethyl acetate (10 mL * 3), and the aqueous phase was directly used in the next step. LC-MS: $[M+H]^+ = 257.5$

Step 3: UBI-1157e

To the aqueous solution of compound UBI-1157d (600mg, 2.35mmol), di-tert-butyl dicarbonate (1.01g, 4.7mmol) and dioxane (20mL) were added, and reacted at room temperature for 2 hours. The reaction solution was concentrated, added water (20 mL), and extracted with ethyl acetate (10 mL*3). The aqueous phase was concentrated to obtain the crude product, which was dissolved in MeOH/DCM=10/1 (30 mL), filtered, and the filtrate was concentrated to obtain the target compound **UBI-1157e** (600mg). LC-MS: $[M+H]^+ = 357.2$

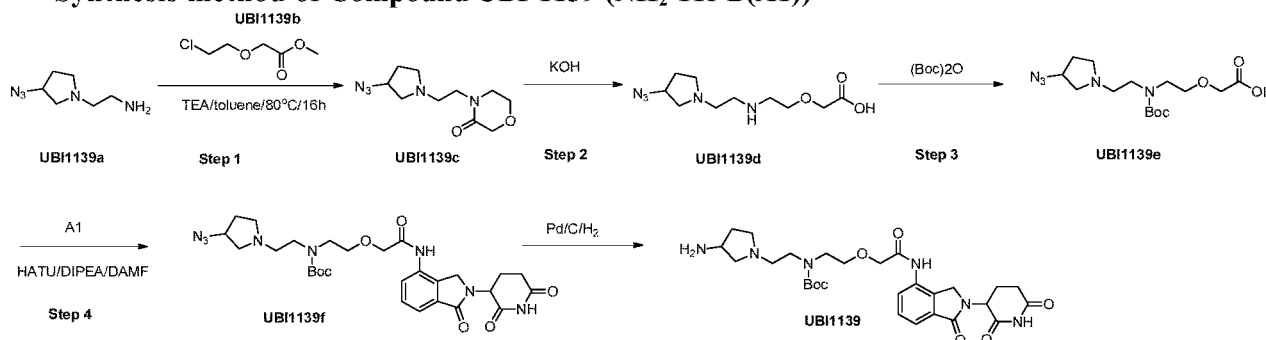
Step 4: UBI-1157f

Compound **UBI-1157e** (600mg, 1.69mmol), **B(A1)** (437mg, 1.69mmol), HATU (1.28g, 3.38mmol) were dissolved in DMF (3 mL) and DIPEA (0.3 mL), and reacted at room temperature for 16 hours. The reaction solution was directly isolated by reversed-phase column (MeOH/H₂O = 5% to 95%, 45 minutes, collected at 40%) to obtain target compound **UBI-11157f** (400mg) as white solid. LC-MS: $[M+H]^+ = 597.5$

Step 5: UBI-1157

Compound **UBI-1157f** (100mg, 0.17mmol) and Pd/C (20mg) were dissolved in MeOH (1 mL) and DCM (10mL), and reacted for 16 hours under hydrogen atmosphere. The reaction was filtered after completion, and the filtrate was concentrated to obtain crude product. The crude product was washed with anhydrous ether (3mL*3), and filtered to obtain target compound **UBI-1157** (50mg) as a white solid. LCMS: (M+H)⁺ = 543.3

Synthesis method of Compound **UBI-1139** (NH₂-11f-B(A1))



Step 1: **UBI-1139c**

Compound **UBI-1157a** (1.5 g, 10mmol), **UBI-1139b** (4.4 g, 29mmol) and TEA (980mg, 10mmol) were dissolved in toluene (25mL) and reacted at 80°C for 18 hours. The reaction solution was concentrated and subjected to silica gel column chromatography (petroleum ether/ethyl acetate = 50% to 100%, 20 minutes, methanol/dichloromethane = 0% to 10%, 30 minutes) to obtain crude product **UBI-1139c** (500mg, yield 22%) as light yellow oil.

¹H NMR (400 MHz, chloroform-*d*) δ 4.03 (s, 2H), 3.92 (s, 1H), 3.79 (dd, *J* = 5.8, 4.2 Hz, 2H), 3.68 (dd, *J* = 6.2, 4.5 Hz, 2H), 3.43 (d, *J* = 6.4 Hz, 2H), 2.87 (s, 2H), 2.76 (d, *J* = 9.7 Hz, 2H), 2.65 (s, 2H), 2.45 (s, 1H), 2.32 – 2.15 (m, 1H), 1.92 (s, 1H).

Step 2: **UBI-1139d**

Compound **UBI-1139c** (200mg, 0.8mmol), KOH (141mg, 2.5mmol) was dissolved in water (2mL), and the reaction was reacted at 100°C for 3 hours. The reaction liquid was directly used in the next step.

Step 3: **UBI-1139e**

Compound **UBI-1139d** (200mg, 0.8mmol) and (Boc)₂O (340mg, 1.6mmol) were dissolved in dioxane (3mL) and water (2 mL), and reacted at room temperature for 2 hours. The reaction solution was concentrated and washed with ethyl acetate(10mL*3). The aqueous phase was acidified to pH=6 using (1M) HCl, and lyophilized to obtain crude product. The crude product was soaked in methanol/dichloromethane=1/10 (30mL) and filtered. The mother solution was dried by rotary evaporation to obtain crude product **UBI-1139e** (300mg, yield 100%) as colorless oil.

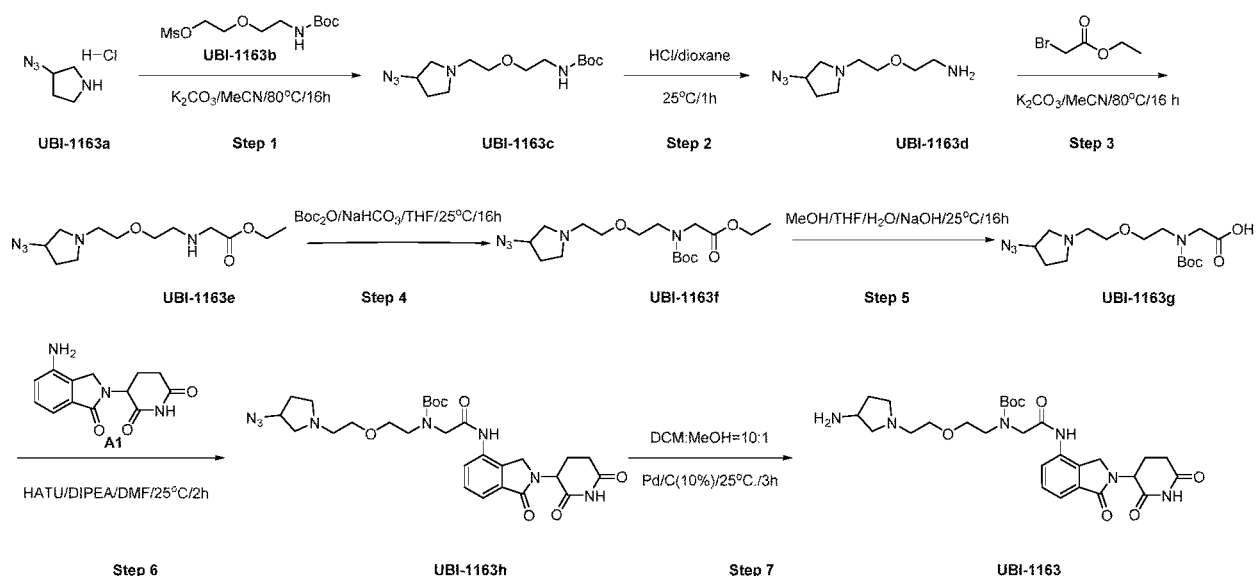
Step 4: **UBI-1139f**

Compound **UBI-1139e** (300mg, 0.84mmol), **B(A1)** (196mg, 0.76mmol), and HATU (639mg, 1.68mmol) were dissolved in DMF (10 mL) and DIPEA (0.3 mL), and reacted at room temperature for 16 hours. The reaction solution was directly purified via reversed-phase column (methanol/water = 5 % to 95 %, 45 minutes, collected at 40%) to obtain product **UBI-1139f** (100mg, yield 20%) as light yellow oil.

Step 5: **UBI-1039**

Under hydrogen atmosphere, compound **UBI-1139f** (50mg, 0.08 mmol), Pd/C (5 mg) were dissolved in methanol (0.5 mL) and dichloromethane (5mL), and reacted at room temperature for 1 hour. The reaction solution was filtered on Celite. The filtrate was concentrated to obtain crude product **UBI-1139** (50mg, yield 100%) as yellow oil.

Synthesis method of Compound **UB-1163** (NH₂-11f-B(A1))



Step 1: UBI-1163c

Compound **UBI-1163a** (1.40 g, 8.92 mmol), **BUI-1163b** (3.032 g, 10.70 mmol) and K_2CO_3 (2.456 g, 17.83 mmol) were dissolved in CH_3CN (40 mL), then heated to 80°C and reacted for 16 hours. After completion of reaction, the reaction was concentrated under reduced pressure, added water (50 mL), and then extracted with ethyl acetate (50 mL*3). The organic phases were combined, washed with water (50 mL*3), saturated brine (50 mL*3) successively, then dried over anhydrous sodium sulfate, and concentrated under reduced pressure to obtain crude product. The crude product was isolated by silica gel column chromatography (methanol/ dichloromethane = 0% to 10%) to obtain target compound **UBI-1163c** (1.32g, yield 47%) as yellow solid. ^1H NMR (400 MHz, chloroform- d) δ 5.29 - 4.94 (m, 1H), 4.15 - 3.89 (m, 1H), 3.58 (t, J = 5.6 Hz, 2H), 3.52 (t, J = 5.2 Hz, 2H), 3.36 - 3.25 (m, 2H), 2.93 - 2.79 (m, 1H), 2.79 - 2.67 (m, 2H), 2.67 - 2.56 (m, 2H), 2.30 - 2.13 (m, 1H), 1.96 - 1.79 (m, 1H), 1.45 (s, 9H).

Step 2: UBI-1163d

Compound **UBI-1163c** (1.300g, 4.342mmol) was dissolved in a solution of HCl in dioxane (40 mL), reacted at 25°C for 1 hour. The reaction solution was concentrated to obtain product **UBI-1163d** (1.05g, yield 100%) as yellow oil. The crude product was directly used in the next step.

Step 3: UBI-1163e

Compound **UBI-1163d** (1.05 g, 4.34 mmol), ethyl 2-bromoacetate (1.09 g, 6.51 mmol) and K_2CO_3 (1.8 g, 13.03 mmol) were dissolved in CH_3CN (50 mL) and heated to 80°C and reacted for 16 hours. After completion of reaction, the reaction was concentrated, added water (50 mL), and then extracted with ethyl acetate (50 mL*3). The organic phases were combined, washed with water (50 mL*3), saturated brine (50 mL*3) successively, then dried over anhydrous sodium sulfate, and concentrated under reduced pressure to obtain crude product. The crude product was isolated by silica gel column chromatography (methanol/ dichloromethane = 0% to 10%) to obtain target compound **UBI-1163e** (810 mg, yield 62%) as yellow oil. ^1H NMR (400 MHz, chloroform- d) δ 4.19 (q, J = 7.2 Hz, 2H), 4.10 = 3.96 (m, 1H), 3.67 - 3.52 (m, 4H), 3.51 - 3.41 (m, 2H), 3.01 - 2.54 (m, 8H), 2.29 - 2.14 (m, 2H), 1.95 - 1.81 (m, 1H), 1.28 (t, J = 7.2 Hz, 3H).

Step 4: UBI-1163f

Compound **UBI-1163e** (790mg, 2.22mmol), di-*tert*-butyl dicarbonate (1.208g, 5.537mmol) and $NaHCO_3$ (465mg, 5.54mmol) were dissolved in THF (30mL) and reacted at 25 °C for 1 hours. After the completion of reaction, it was concentrated, added water (50mL), then extracted with ethyl acetate (50 mL * 3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na_2SO_4 , concentrated by rotary evaporation under reduced pressure to obtain crude product. The crude product was

isolated by silica gel column chromatography (ethyl acetate/petroleum ether = 0% to 20%) to obtain target product **UBI-1163f** (900mg, yield 83%) as colorless transparent oil. ¹H NMR (400 MHz, chloroform-*d*) δ 4.25 - 4.13 (m, 2H), 4.10 - 3.91 (m, 3H), 3.64 - 3.40 (m, 6H), 2.97 - 2.41 (m, 6H), 2.27 - 2.12 (m, 1H), 1.95 - 1.79 (m, 1H), 1.47 (s, 4H), 1.42 (s, 5H), 1.28 (t, *J* = 7.2 Hz, 3H).

5 Step 5: UBI-1163g

Compound **UBI-11163f** (900mg, 2.34mmol), NaOH (280mg, 7.00mmol) were dissolved in water (3mL), THF(9mL) and MeOH(3mL), and reacted at 25°C for 16 hours. After completion of reaction, the reaction was concentrated, added water (20 mL), and then extracted with ethyl acetate (50 mL*3). The aqueous phase was acidified to pH=5 using diluted hydrochloric acid (1M), and then extracted with dichloromethane (50 mL*3). The organic phases were combined, then washed with water (50mL*3), saturated brine (50mL*3) successively, further dried over anhydrous Na₂SO₄, and concentrated by rotary evaporation under reduced pressure to obtain colorless transparent oily liquid, which was the target compound **UBI-1163g** (490mg, yield 59%). ¹H NMR (400 MHz, DMSO-*d*₆) δ 4.09 - 3.97 (m, 1H), 3.85 (d, *J* = 8.8 Hz, 2H), 3.50 - 3.42 (m, 4H), 3.36 - 3.31 (m, 2H), 2.80 - 2.69 (m, 1H), 2.69 - 2.61 (m, 2H), 2.58 - 2.54 (m, 2H), 2.37 - 2.31 (m, 1H), 2.19 - 2.07 (m, 1H), 1.78 - 1.62 (m, 1H), 1.49 - 1.25 (m, 9H).

15 Step 6: UBI-1163h

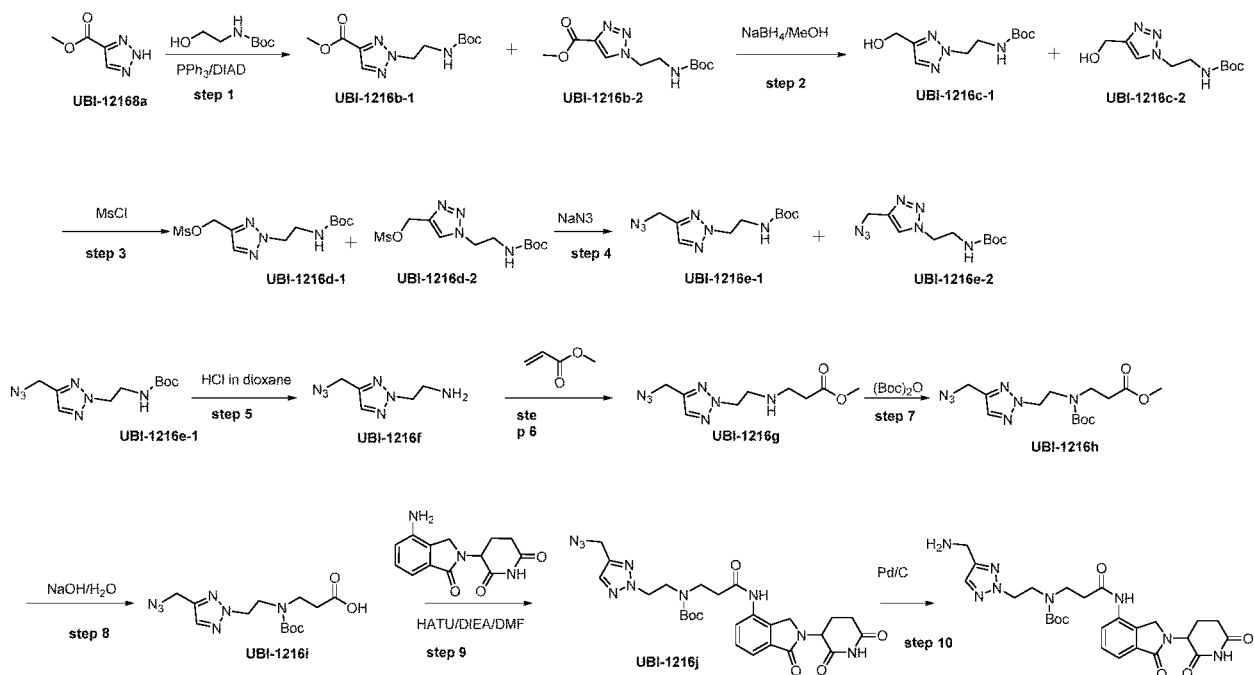
Compound **UBI-1098e** (455mg, 1.27mmol), **B(A1)** (330mg, 1.27mmol), HATU (968mg, 2.55mmol), DIPEA (823mg, 6.36mmol) were dissolved in DMF (4 mL) and reacted at 25°C for 2 hours. After completion of reaction, the reaction was added water (30 mL), and then extracted with ethyl acetate (50 mL*3). The organic phases were combined, washed with water (50 mL*3), saturated brine (50 mL*3) successively, then dried over anhydrous sodium sulfate, and concentrated by rotary evaporation under reduced pressure to obtain crude product. Then the crude product was isolated by silica gel column chromatography (methanol/ dichloromethane = 0% to 20%) to obtain yellow solid, which was the target compound **UBI-1163h** (384mg, yield 50%). ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.04 (s, 1H), 9.87 (s, 1H), 7.80 (s, 1H), 7.62 - 7.44 (m, 2H), 5.27 - 5.09 (m, 1H), 4.45 - 4.26 (m, 2H), 4.05 (d, *J* = 11.6 Hz, 2H), 3.71 - 3.52 (m, 4H), 3.44 (d, *J* = 5.4 Hz, 2H), 3.32 (s, 7H), 3.17 (d, *J* = 3.6 Hz, 1H), 2.99 - 2.89 (m, 1H), 2.66 - 2.58 (m, 1H), 2.27 (d, *J* = 13.2 Hz, 1H), 2.09 - 2.00 (m, 1H), 1.91 (s, 1H), 1.38 (d, *J* = 26.4 Hz, 9H). LCMS [M+H]⁺: 599.1.

25 Step 7: UBI-1163

Compound **UBI-1163h** (173mg, 0.290mmol) and Pd/C (156mg) were dissolved in MeOH (3 mL) and DCM (30mL), and reacted for 3 hours under hydrogen atmosphere. After completion of reaction, the reaction was filtered and washed with methanol (4 mL) and dichloromethane (40 mL). The filtrate was concentrated by rotary evaporation under reduced pressure to obtain the target compound **UBI-11163** (140 mg, yield 82%) as yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.03 (s, 1H), 9.94 (s, 1H), 8.34 (s, 2H), 7.89 - 7.74 (m, 1H), 7.65 - 7.40 (m, 2H), 5.26 - 5.08 (m, 1H), 4.51 - 4.24 (m, 2H), 4.14 - 4.03 (m, 2H), 3.98 - 3.80 (m, 1H), 3.80 - 3.50 (m, 5H), 3.50 - 3.36 (m, 4H), 3.17 (s, 2H), 2.99 - 2.89 (m, 1H), 2.67 - 2.56 (m, 3H), 2.49 - 2.43 (m, 2H), 2.31 - 2.22 (m, 1H), 2.09 - 2.02 (m, 1H), 1.38 (d, *J* = 27.6 Hz, 9H). LCMS [M+H]⁺: 573.1.

35 Synthesis method of Compound Linker g-Ligand B(A1)

40 Synthesis method of Compound UBI-1216 (NH₂-9g-B(A1))



Step 1: UBI-1216b

Methyl -1H-1,2,4-triazole-3-carboxylate (200 mg, 1.57 mmol) and N-Boc-ethanolamine (304 mg, 1.89 mmol) were added to THF (10 ml). Triphenylphosphine (495 mg, 1.89 mmol) and diisopropyl azodicarboxylate (381 mg, 1.89 mmol) were added to the above solution, and stirred overnight at room temperature. The reaction solution was purified by a reversed-phase column (methanol/H₂O = 5% to 95%, 45mins) and was collected at 60% to obtain white solid (230 mg, yield 54%). LC-MS: (M+H)⁺=271.1

Step 2: UBI-1216c

At 0°C, to a solution of UBI-1216b (1 g, 33.3 mmol) in THF (10 ml) was added lithium aluminum hydride (1.9 g, 50 mmol) for 30 min. The reaction was stirred for 3 hours, added water (1 ml) and 10% NaOH solution (2 ml), filtered, and then the filtrate was dried over Na₂SO₄, concentrated under reduced pressure to obtain the product (8.01 g, yield 51%). LC-MS: (M+H)⁺=243.1

Step 3: UBI-1216d

To a solution of UBI-1216c (1.08 g, 4 mmol) and triethylamine (0.68 g, 7 mmol) in dichloromethane (10 ml) was added methanesulfonyl chloride (0.76 g, 7 mmol) dropwise, and stirred overnight at room temperature. After completion of reaction, to the mixture was added water (10ml) and extracted with dichloromethane (10ml*3). The organic layer was dried over Na₂SO₄ and concentrated to obtain white solid (1.15 g, crude product). LC-MS: (M+H)⁺=321.1

Step 4: UBI-1216e

UBI-1216d (1.7 g, 5 mmol) and NaN₃ (520 mg, 8 mmol) were mixed and dissolved in DMF (10 ml), stirred overnight at room temperature. After completion of reaction, the reaction was diluted with H₂O (300 ml) and extracted with ether (2x150 ml). The organic phase was washed with H₂O (3x100 ml) and brine (1x100 ml), dried over MgSO₄, filtered, and the solvent was removed under low pressure to obtain light yellow oil (1.1 g, yield 79%).

¹H NMR (400 MHz, CDCl₃) δ 7.61 (d, *J* = 8.5 Hz, 1H), 4.94 (s, 1H), 4.51 (dd, *J* = 17.7, 12.2 Hz, 2H), 4.43 (s, 2H), 3.70 (dd, *J* = 10.7, 5.6 Hz, 2H), 1.44 (s, 9H). LC-MS: (M+H)⁺=268.3

Step 5: UBI-1216f

UBI-1216e (500 mg, 1.416 mmol) was dissolved in a solution of hydrochloric acid/dioxane (4N) (10 ml), and stirred overnight at room temperature. After the completion of reaction, the solvent was removed and used directly in the next step. (430mg, crude product). LC-MS: (M+H)⁺=168.3

Step 6: UBI-1216g

Compound **UBI-1216f** (500 mg, 2.99 mmol) and methyl acrylate (257 mg, 2.99 mmol) were dissolved in acetonitrile (20 mL), and reacted at 80 °C for 18 hours. After completion of reaction, the reaction solution was poured into 5 mL water and extracted with ethyl acetate (5 mL*3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na₂SO₄, concentrated under reduced pressure. Obtained crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 10/1) to obtain compound as colorless oil (170 mg, yield 23%). LC-MS: (M+H)⁺=254.1

Step 7: UBI-1216h

Compound **UBI-1216g** (650 mg, 2.57 mmol), di-tert-butyl dicarbonate (840 mg, 3.85 mmol) and sodium bicarbonate (318 mg, 3.79 mmol) were added to dioxane (30mL) successively and reacted at room temperature for 2 hours. After the completion of the reaction, the reaction solution was poured into 10 mL of water and extracted with ethyl acetate (10 mL*3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure to obtain compound **UBI-1216h** (625 mg, yield 69%). LC-MS: (M+H)⁺=354.1

Step 8: UBI-1216i

Compound **UBI-1216h** (500 mg, 1.416 mmol), aqueous solution of sodium hydroxide (4N, 10 mL) were sequentially added into water (50 mL), and reacted at 30 °C for 12 hours. After the completion of reaction, it was concentrated and the aqueous phase was acidified to pH=5 using hydrochloric acid (1M). Then it was extracted with dichloromethane (10mL*3), the combined organic layers was dried over anhydrous Na₂SO₄, and concentrated to obtain compound **UBI-1216i** (1.2 g, yield 83%). LC-MS:(M+H)⁺=340.1

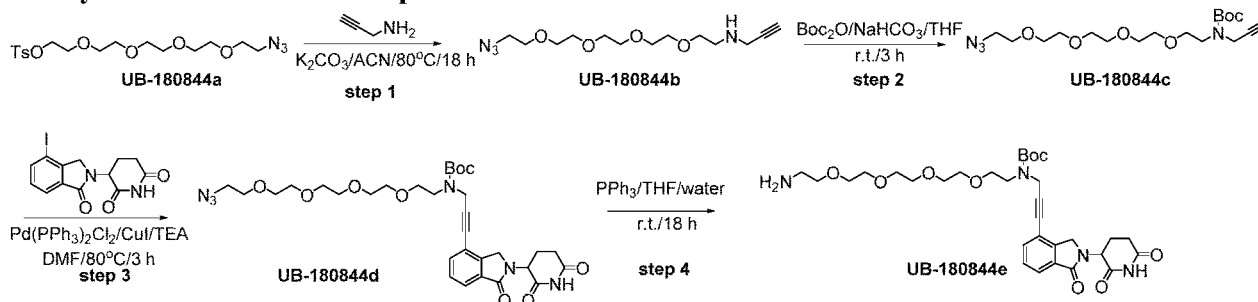
Step 9: UBI-1216j

Compound **UBI-1216i** (20 mg, 0.059 mmol), **B(A1)** (15 mg, 0.059 mmol), HATU (33 mg, 0.088 mmol) and DIPEA (0.5mL) were added to anhydrous DMF (5 mL). The mixture was reacted at room temperature for 16 hours, concentrated after the completion of reaction. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 10/1) to obtain compound **UBI-1216j** (16 mg, 49% yield) as white solid. LC-MS (M+H)⁺=581.1

Step 10: UBI-1216

Compound **UBI-1216j** (100 mg, 0.17mmol), 10% palladium on carbon (20 mg) were added to a mixed solvent of methanol/dichloromethane (1 mL/10 mL), and reacted at room temperature for 16 hours under hydrogen atmosphere. After filtration, the filtrate was concentrated to obtain the crude product. The crude product was washed with cold ether (10 mL*3), and dried to obtain target compound **UBI-1095** (83 mg, yield 88%) as white solid. LCMS: (M-100+H)⁺=455.3

Synthesis method of Compound UB-180844e



Step 1: UB-180844b

Compound **UB-180844a** (600 mg, 1.4 mmol) was dissolved in acetonitrile (20 mL), added

propargylamine (400 mg, 7.2 mmol), potassium carbonate (386 mg, 2.8 mmol). reacted at 80°C for 18 hours. The reaction solution was cooled, filtered, concentrated and was directly used in the next reaction. LCMS $[M+H]^+ = 301$

Step 2: UB-180844c

Compound **UB-180844b** (459 mg, 1.53 mmol) was dissolved in tetrahydrofuran (10 mL). Di-*tert*-butyl dicarbonate (1.5 g, 6.88 mmol) and sodium bicarbonate (257 mg, 3.06 mmol) were added. Reacted at room temperature for 3 hours. After completion of the reaction, the filtrate was concentrated to obtain the crude product, which was isolated by silica gel column chromatography (petroleum ether/ethyl acetate = 0% to 30%) to obtain target product **UB-180844c** (340 mg, yield 60%) as a yellow transparent oil. LCMS $[M+H]^+ = 401$

Step 3: UB-180844d

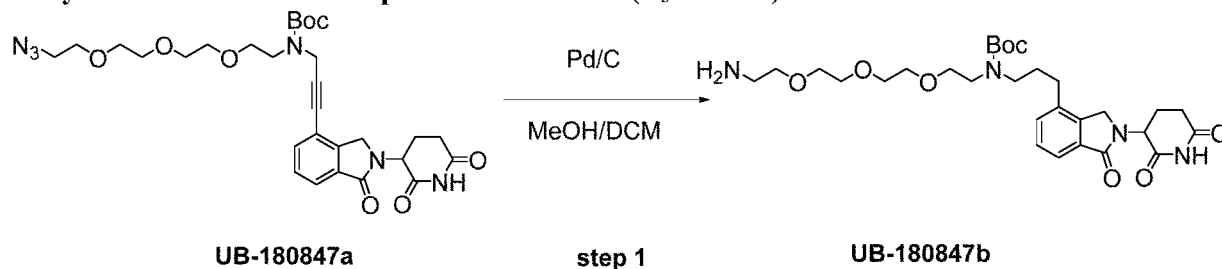
Compound **UB-180844c** (346 mg, 0.86 mmol), iodine-substituted lenalidomide (150 mg, 0.55 mmol), $Pd(PPh_3)_2Cl_2$ (35 mg, 0.05 mmol), CuI (21 mg, 0.11 mmol), triethylamine (55 mg, 0.55 mmol) were dissolved in anhydrous DMF (6 mL), reacted at 80°C for 3 hours under nitrogen protection. After completion of reaction, the filtrate was concentrated to obtain the crude product, which was isolated by silica gel column chromatography (ethyl acetate-dichloromethane/methanol=100% - 0% to 10%) to obtain target product **UB-180844d** (200 mg, yield 42%) as brown oil. LCMS $[M-100]^+ = 543$

^1H NMR (400 MHz, chloroform-*d*) δ 7.84 (d, $J = 7.6$ Hz, 1H), 7.71 – 7.64 (m, 1H), 7.63 – 7.51 (m, 1H), 7.46 (t, $J = 7.8$ Hz, 1H), 5.24 (dd, $J = 13.3, 5.2$ Hz, 1H), 4.50 (d, $J = 16.8$ Hz, 1H), 4.37 (m, 2H), 3.70 – 3.60 (m, 14H), 3.55 (m, 2H), 3.42 – 3.36 (m, 2H), 2.91 (m, 1H), 2.30 – 2.20 (m, 1H), 2.03 (d, $J = 14.4$ Hz, 1H), 1.49 (s, 9H), 1.31 (m, 2H), 0.88 (t, $J = 6.2$ Hz, 2H).

Step 4: UB-180844e

Compound **UB-180844d** (240 mg, 0.37 mmol) was dissolved in tetrahydrofuran (20 mL) and added triphenylphosphine (262 mg, 1 mmol), reacted at room temperature for 3 hours. The reaction was added water (0.3 mL) and reacted at room temperature for 18 hours. After the completion of reaction, the crude product was obtained by concentration, and isolated by silica gel column chromatography (dichloromethane/methanol=0% to 10%) to obtain crude product **UB-180844e** (60 mg) as yellow oil. LCMS $[M+H]^+ = 617$

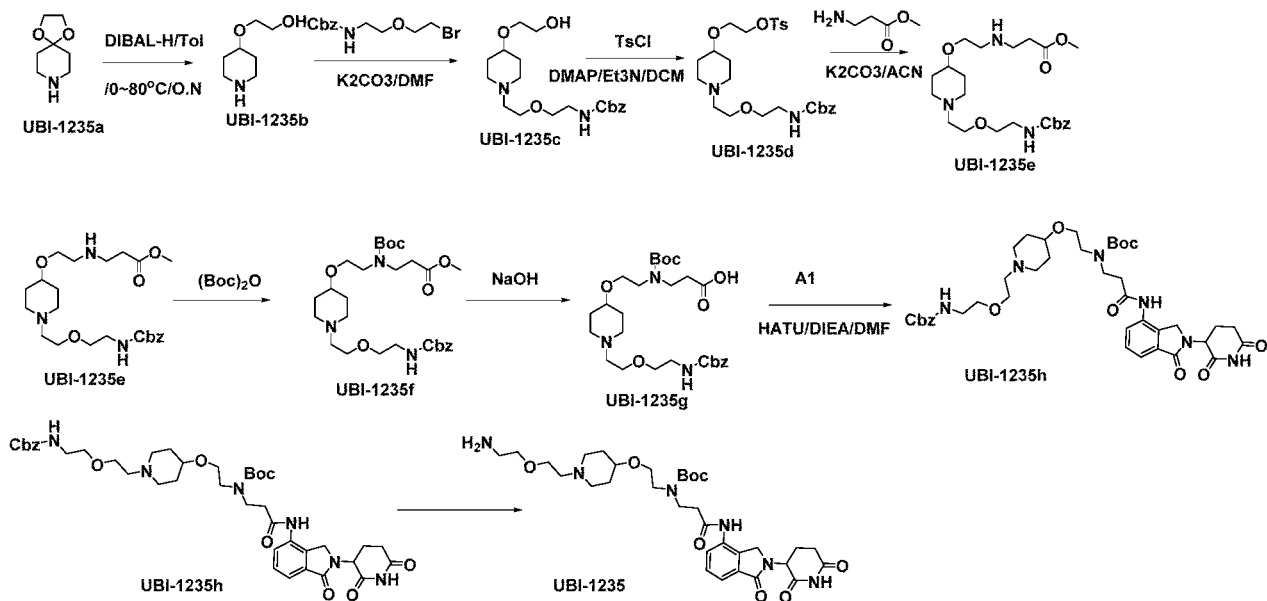
Synthetic method of Compound UB-180847b (N₃-13b-A1)



Step 1: UB-180847b

Compound **UB-180847a** (100 mg, 2.3 mmol) and Pd/C (10 mg) were dissolved in methanol (1 mL) and dichloromethane (10 mL), and then reacted at room temperature for 1 hour under hydrogen. After the reaction solution was filtered, the filtrate was concentrated to obtain crude product. The crude product was purified by a reversed-phase column (methanol/water=5% to 95%, collected at 20%) to obtain target product **UB180847b** (80mg, yield 84 %) as yellow oil.

Synthesis method of Compound UBI-1235 (NH₂-16e-A1)



Step 1: UBI-1235b

UBI-1235a (1.0 g, 7.0 mmol, 1.0 equiv) was dissolved in toluene (20 mL), added 1.0 M diisobutylaluminum hydride in hexane (20 mL, 20 mmol, 2.9 equiv) at 0° C. The solution was heated to 80°C and reacted for 12 hours. added methanol (20 mL), sodium tartrate (20 mL) and chloroform (100 mL). The solution was stirred for 30 minutes. The chloroform was separated, and the aqueous solution was extracted 5 times. The mixture was dried (Na₂SO₄), filtered and concentrated to obtain 980 mg of crude UBI-1235b as white solid. LC-MS: [M+H]⁺ = 146.3

Step 2: UBI-1235c

UBI-1235b (0.28 g, 1.93 mmol) was added to DMF (10 ml), and added UBI-1235b (581 mg, 1.93 mmol) and K₂CO₃ (748 mg, 5.42 mmol). The above solution was stirred at 80°C overnight. TLC showed that the reaction was completed. The reaction mixture was added to a mixed solvent of DCM/MeOH (V/V=20 ml), stirred for 10 minutes, filtered, and concentrated *in vacuo* to obtain the crude product, which was isolated by column chromatography (DCM/MeOH=10/ 1) and purified to obtain product UBI-1235C (625 mg, yield: 89%) as a yellow oil. LC-MS: [M+H]⁺ = 367.5

Step 3: UBI-1235d

Compound UBI-1235C (500 mg, 1.366 mmol) was dissolved in DCM (10 ml), added TsCl (313 mg, 1.639 mmol) and triethylamine (275 mg, 2.732 mmol), and stirred overnight at room temperature. After completion of reaction, the reaction solution was filtered, concentrated, and purified by column chromatography (DCM/MeOH=10/1) to obtain the product UBI-1235d (510 mg, yield: 72%) as a colorless oil. LC-MS: [M+H]⁺ = 521.6

Step 4: UBI-1235e

UBI-1235d (450 mg, 0.865 mmol) and methyl 3-aminopropionate (200 mg, 1.30 mmol) were added to MeCN (30 ml), potassium carbonate (239 mg, 1.731 mmol) was added. The mixture was stirred overnight at room temperature. After completion of reaction, the reaction was filtered and concentrated, and isolated by column chromatography (DCM/MeOH=10/1) and purified to obtain the product UBI-1235e (190 mg, yield 49%) as a colorless oil. LC-MS: [M+H]⁺ = 452.5

Step 5: UBI-1235f

UBI-1235e (100mg, 0.323mmol) was added to tetrahydrofuran/water (30ml), then di-*tert*-butyl dicarbonate (105mg, 0.484mmol) and sodium bicarbonate (54mg, 0.643mmol) were added, and stirred overnight at room temperature. After completion of reaction, the reaction was filtered and the filtrate was concentrated to obtain crude product, which was isolated by column chromatography (DCM/MeOH=10/1) and purified to obtain the product UBI-1235f (99 mg, yield 81%) as a yellow oil. LC-MS: $[M+H]^+ = 552.3$

Step 6: UBI-1235g

UBI-1235f (100 mg, 0.181 mmol) was dissolved in 4N NaOH (10 ml) and stirred overnight at room temperature. After completion of reaction, the mixture was adjusted to pH = 5, and added brine and ethyl acetate for stratification. The aqueous layer was extracted with ethyl acetate three times. The combined extract was dried over sodium sulfate, filtered and concentrated to obtain product UBI-1235g (56 mg).

Step 7: UBI-1235h

UBI-1235g (50 mg, 0.093 mmol) and lenalidomide (24 mg, 0.093 mmol) were added to DMF (10 ml), HATU (53 mg, 0.139 mmol) and DIEA (36 mg, 0.279 mmol) were added, and stirred at room temperature overnight. After completion of reaction, the crude product was concentrated to obtain crude product. The crude product was purified by Pre-TLC (DCM/MeOH=10/1) to obtain the product UBI-1235h (65 mg, yield 90%) as a white solid. LC-MS: $[M+H]^+ = 779.1$

Step 8: UBI-1235i

UBI-1235h (85mg, 0.109mmol) was added to THF (10ml), and palladium on carbon (10%, 10mg) was added, and the mixture was stirred overnight at room temperature under H₂ atmosphere. After completion of reaction, the reaction solution was added to a mixture of DCM/MeOH (v/v=20 ml), stirred for 10 min, filtered, and concentrated to obtain crude product, which was purified by Pre-TLC (DCM/MeOH=10/1) to obtain UBI-1235i (56 mg, 80% yield) as white solid. LC-MS: $[M+H]^+ = 645.3$

L1-B conjugates, shown in Preparation Table A, in which H in structural formula of the conjugate being replaced with structures of E3 ligase ligand B, i.e. the corresponding linker L1 molecule.

Table A

Compound No.	Structural formula of L1-B
	L1-B(A1)
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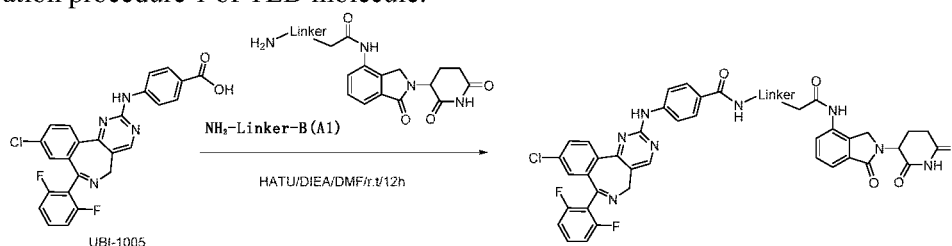
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L1-B(B1)	
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Example Preparation of 2TED

(2.1) Preparation procedure 1 of TED molecule:



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in the formula, refers to L2.

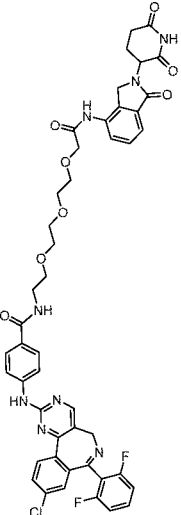
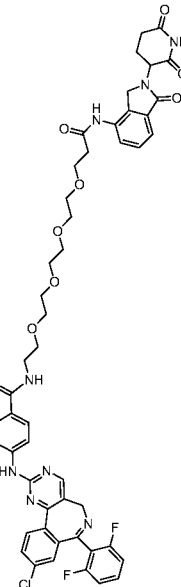
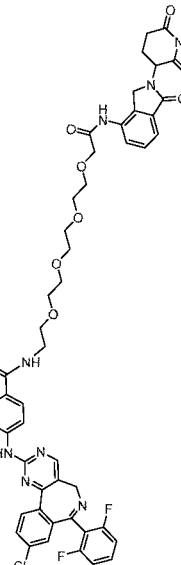
Compound **UBI-1005** (20 mg, 0.042 mmol) was weighed and dissolved in DMF (2 mL), added NH₂-linker-B(A1) (1 eq.), HATU (2eq. 32mg, 0.084mmol) and DIEA (3eq. 16.2mg, 0.126mmol) under ice-water bath successively. After completion of addition, the material system was stirred and reacted at room temperature under the protection of nitrogen for 18 hours. After completion of reaction, the reaction solution was poured into 5 mL of water and extracted with ethyl acetate (5 mL*3) three times. The organic phases were combined and washed with saturated brine, dried over anhydrous Na₂SO₄ and concentrated by rotary evaporation under reduced pressure to obtain the crude product. After that, the crude product was isolated by thin-layer chromatography on silica gel plate with the polarity of the developing solvent (DCM/MeOH = 10/1) to prepare the corresponding TED compound.

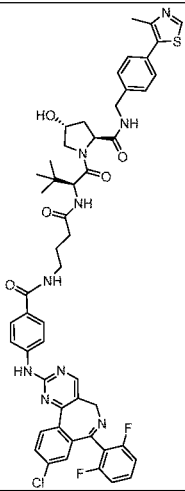
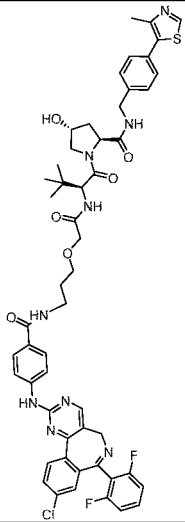
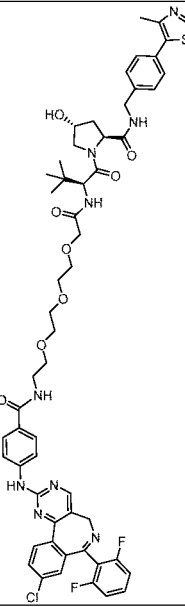
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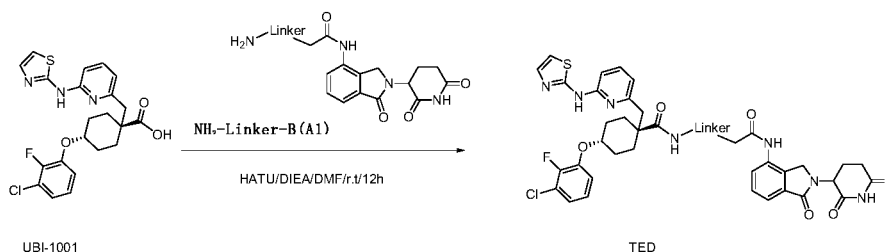
The conjugate of A-L1-B as shown in Table B1 was prepared through similar method.

Table B1

Examples	Compound structure	Analyze data
UB-180501		¹ H NMR (400 MHz, DMSO) δ 11.01 (s, 1H), 10.16 (s, 1H), 9.84 (s, 1H), 8.73 (s, 1H), 8.49 – 8.26 (m, 2H), 7.98 – 7.76 (m, 6H), 7.76 – 7.39 (m, 3H), 7.34 (s, 1H), 7.31-7.18 (m, 2H), 5.14 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.52 – 4.26 (m, 2H), 3.53 – 3.39 (m, 2H), 3.13 – 2.74 (m, 1H), 2.95 – 2.74 (m, 1H), 2.95 – 2.56 (m, 3H), 2.45 – 2.30 (m, 2H), 2.00-581.98 (m, 1H), 1.88 (dd, <i>J</i> = 13.9, 6.9 Hz, 2H), 1.39 – 1.15 (m, 2H). LCMS [M+ H] ⁺ = 803.2

UB-180520		^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 11.01 (s, 1H), 10.16 (s, 1H), 9.70 (s, 1H), 8.72 (s, 1H), 8.40 – 8.24 (m, 2H), 7.97 – 7.80 (m, 5H), 7.74 (d, J = 7.7 Hz, 1H), 7.52 (m, 3H), 7.34 (d, J = 2.2 Hz, 1H), 7.17 (t, J = 8.4 Hz, 2H), 5.14 (m, 1H), 4.46 – 4.28 (m, 2H), 4.14 (s, 2H), 3.76 – 3.36 (m, 14H), 2.98 – 2.80 (m, 1H), 2.59 (d, J = 18.3 Hz, 1H), 2.37 (dd, J = 13.3, 4.3 Hz, 1H), 2.05 – 1.96 (m, 1H). LCMS $[\text{M}+\text{H}]^+ = 1362.1$
UB-180530		^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 11.02 (s, 1H), 10.16 (s, 1H), 9.85 (s, 1H), 8.73 (s, 1H), 8.42 – 8.18 (m, 2H), 7.97 – 7.74 (m, 6H), 7.60 – 7.45 (m, 3H), 7.34 (d, J = 2.1 Hz, 1H), 7.17 (t, J = 8.7 Hz, 3H), 5.14 (m, 1H), 4.47 – 4.24 (m, 2H), 3.70 (t, J = 6.3 Hz, 2H), 3.58 – 3.45 (m, 15H), 3.39 (m, 2H), 3.01 – 2.84 (m, 1H), 2.71 – 2.51 (m, 5H), 2.33 (m, 1H), 2.10 – 1.96 (m, 1H). LCMS $[\text{M}+\text{H}]^+ = 1449.3$
UB-180534		^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 10.16 (s, 1H), 9.69 (s, 1H), 8.72 (s, 1H), 8.40 – 8.24 (m, 2H), 7.86 (m, 6H), 7.74 (d, J = 7.7 Hz, 1H), 7.53 (m, 3H), 7.34 (d, J = 2.1 Hz, 1H), 7.17 (t, J = 8.7 Hz, 2H), 5.14 (dd, J = 13.3, 5.1 Hz, 1H), 4.49 – 4.30 (m, 2H), 4.13 (s, 2H), 3.64 (m, 4H), 3.57 – 3.45 (m, 10H), 3.40 (m, 2H), 2.91 (m, 1H), 2.59 (m, 2H), 2.36 (m, 2H), 2.08 – 1.92 (m, 1H). LCMS $[\text{M}+\text{H}]^+ = 1428.2$

UB-180505		¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 10.15 (s, 1H), 8.97 (s, 1H), 8.73 (s, 1H), 8.57 (t, <i>J</i> = 6.1 Hz, 1H), 8.30 (d, <i>J</i> = 8.4 Hz, 2H), 8.00 – 7.78 (m, 6H), 7.59 – 7.51 (m, 1H), 7.40 (q, <i>J</i> = 8.3 Hz, 4H), 7.17 (t, <i>J</i> = 8.5 Hz, 3H), 5.13 (d, <i>J</i> = 3.6 Hz, 1H), 4.56 (d, <i>J</i> = 9.3 Hz, 1H), 4.50 – 4.32 (m, 3H), 4.22 (m, 1H), 3.75 – 3.62 (m, 2H), 3.26 (p, <i>J</i> = 6.6, 6.0 Hz, 2H), 2.44 (s, 3H), 2.27 (m, 2H), 2.09 – 1.87 (m, 2H), 1.75 (s, 2H), 1.24 (d, <i>J</i> = 5.8 Hz, 2H), 0.95 (s, 9H). LCMS [M+H] ⁺ = 974.3
UB-180506		¹ H NMR (400 MHz, DMSO) δ 10.16 (s, 1H), 8.96 (d, <i>J</i> = 4.7 Hz, 1H), 8.73 (s, 1H), 8.61 (t, <i>J</i> = 5.9 Hz, 1H), 8.50 – 8.25 (m, 2H), 7.86 (dt, <i>J</i> = 19.7, 8.9 Hz, 6H), 7.65 – 7.34 (m, 7H), 7.34 – 7.00 (m, 3H), 4.58 (d, <i>J</i> = 9.6 Hz, 4H), 5.23-5.18 (m, 1H), 4.55 – 4.32 (m, 6H), 3.91 (s, 2H), 4.32 – 3.51 (m, 3H), 2.46 (s, 3H), 2.08 (d, <i>J</i> = 8.2 Hz, 2H), 2.07 – 1.85 (m, 1H), 1.85 – 1.59 (m, 2H), 0.93 (d, <i>J</i> = 6.2 Hz, 9H). LCMS [M+H] ⁺ = 1003.8
UB-180514		¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 10.16 (s, 1H), 8.97 (s, 1H), 8.72 (s, 1H), 8.60 (s, 1H), 8.44 – 8.22 (m, 2H), 7.96 – 7.73 (m, 5H), 7.54 (s, 1H), 7.40 (d, <i>J</i> = 12.1 Hz, 3H), 7.32 (d, <i>J</i> = 13.5 Hz, 2H), 7.17 (m, 2H), 7.05 (m, 2H), 5.16 (s, 1H), 4.56 (d, <i>J</i> = 9.4 Hz, 1H), 4.49 – 4.18 (m, 3H), 3.97 (s, 2H), 3.74 – 3.45 (m, 12H), 3.40 (d, <i>J</i> = 5.5 Hz, 2H), 2.43 (s, 3H), 1.98 (m, 2H), 0.94 (s, 9H). LCMS [M+H] ⁺ = 1078.4

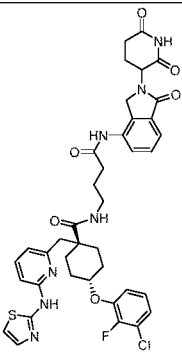
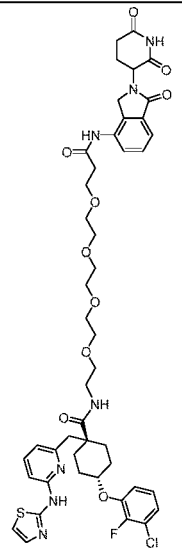
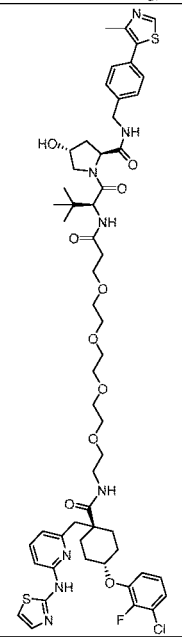
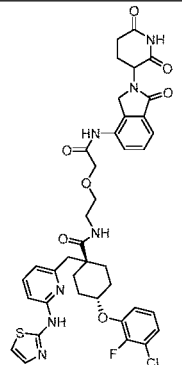


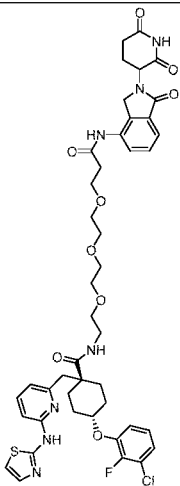
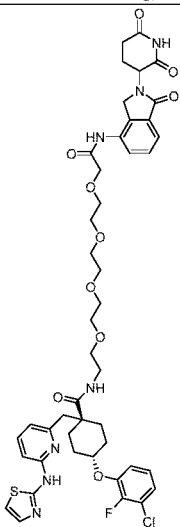
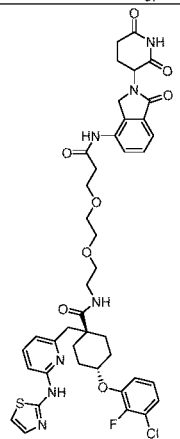
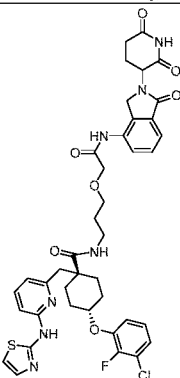
Compound **UBI-1001** (20 mg, 0.043 mmol) was weighed and dissolved in DMF (2 mL), added **NH₂-linker-B(A1)** (1 eq.), **HOBT** (2eq. 23mg, 0.086mmol), **EDCI** (2eq. 16.4mg, 0.086mmol) and **TEA** (3eq. 13mg, 0.129mmol) under ice-water bath successively. After completion of addition, the material system was stirred and reacted at room temperature under the protection of nitrogen for 18 hours. After completion of reaction, the reaction solution was poured into 5 mL of water and extracted with ethyl acetate (5 mL*3) three times. The organic phases were combined and washed with saturated brine, dried over anhydrous Na₂SO₄ and concentrated by rotary evaporation under reduced pressure to obtain the crude product. After that, the crude product was isolated by thin-layer chromatography on silica gel plate with the polarity of the developing solvent (DCM/MeOH = 10/1) to prepare the target compound TED.

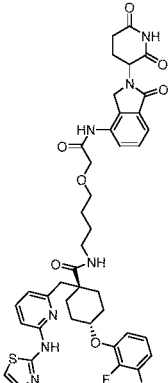
The conjugate of A-L1-B as shown in Table B3 was prepared through similar method.

Table B3

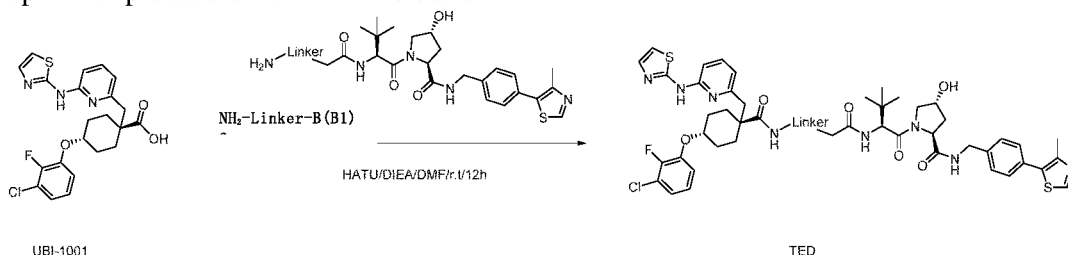
Examples	Compound structure	Analyze data
UB-180572		¹ H NMR (400 MHz, DMSO-d ₆) δ 11.12 (s, 1H), 11.01 (s, 1H), 9.71 (s, 1H), 7.73 (d, J = 7.7 Hz, 1H), 7.63 (d, J = 6.0 Hz, 1H), 7.52 (dt, J = 19.2, 7.5 Hz, 3H), 7.35 (d, J = 3.6 Hz, 1H), 7.16 – 7.03 (m, 3H), 6.95 (d, J = 3.7 Hz, 1H), 6.86 (d, J = 8.2 Hz, 1H), 6.62 (d, J = 7.3 Hz, 1H), 5.13 (dd, J = 13.2, 5.1 Hz, 1H), 4.52 (s, 1H), 4.45 – 4.26 (m, 2H), 4.14 (s, 2H), 3.68 (t, J = 4.6 Hz, 2H), 3.59 (t, J = 4.7 Hz, 2H), 3.41 (m, 3H), 3.21 (d, J = 6.1 Hz, 1H), 2.90 (s, 2H), 2.59 (m, 2H), 2.35 (m, 1H), 2.12 – 1.55 (m, 9H). LCMS [M+1] ⁺ = 849.3
UB-180573		¹ H NMR (400 MHz, DMSO-d ₆) δ 11.12 (s, 1H), 11.01 (s, 1H), 9.72 (s, 1H), 7.73 (d, J = 7.7 Hz, 1H), 7.63 (s, 1H), 7.58 – 7.44 (m, 3H), 7.35 (d, J = 3.6 Hz, 1H), 7.23 – 7.01 (m, 3H), 6.95 (d, J = 3.6 Hz, 1H), 6.86 (d, J = 8.2 Hz, 1H), 6.62 (d, J = 7.4 Hz, 1H), 5.76 (s, 1H), 5.32 (t, J = 4.9 Hz, 1H), 5.14 (dd, J = 13.3, 5.2 Hz, 1H), 4.54 (s, 1H), 4.44 – 4.29 (m, 2H), 4.13 (s, 2H), 3.70 – 3.46 (m, 8H), 3.19 – 3.12 (m, 3H), 2.90 (m, 3H), 2.64 (m, 1H), 2.39 – 2.26 (m, 1H), 1.63 (m, 10H). LCMS [M+1] ⁺ = 892.5

UB-180574		¹ H NMR (400 MHz, DMSO-d ₆) δ 11.12 (s, 1H), 11.02 (s, 1H), 9.83 (s, 1H), 7.82 (dd, <i>J</i> = 6.9, 2.0 Hz, 1H), 7.68 (t, <i>J</i> = 5.5 Hz, 1H), 7.57 – 7.44 (m, 3H), 7.35 (d, <i>J</i> = 3.6 Hz, 1H), 7.17 – 7.04 (m, 3H), 6.95 (d, <i>J</i> = 3.6 Hz, 1H), 6.86 (d, <i>J</i> = 8.2 Hz, 1H), 6.60 (d, <i>J</i> = 7.3 Hz, 1H), 5.15 (dd, <i>J</i> = 13.3, 5.2 Hz, 1H), 4.52 (s, 1H), 4.47 – 4.18 (m, 2H), 3.10 (q, <i>J</i> = 6.7 Hz, 2H), 2.99 – 2.79 (m, 3H), 2.58 (d, <i>J</i> = 18.0 Hz, 1H), 2.33 (p, <i>J</i> = 6.7, 5.9 Hz, 3H), 2.07 – 1.96 (m, 1H), 1.74 (m, 10H). LCMS [M+1] ⁺ = 789.2
UB-180575		¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.07 (d, <i>J</i> = 38.3 Hz, 2H), 9.85 (s, 1H), 7.82 (m, 1H), 7.64 (t, <i>J</i> = 5.7 Hz, 1H), 7.59 – 7.43 (m, 3H), 7.36 (d, <i>J</i> = 3.6 Hz, 1H), 7.21 – 7.07 (m, 3H), 6.95 (d, <i>J</i> = 3.6 Hz, 1H), 6.86 (d, <i>J</i> = 8.2 Hz, 1H), 6.63 (d, <i>J</i> = 7.4 Hz, 1H), 5.15 (m, 1H), 4.54 (s, 1H), 4.47 – 4.26 (m, 2H), 3.70 (t, <i>J</i> = 6.3 Hz, 2H), 3.56 – 3.42 (m, 12H), 3.37 (m, 2H), 3.18 (q, <i>J</i> = 5.9 Hz, 2H), 2.90 (m, 3H), 2.68 – 2.57 (m, 4H), 2.40 – 2.27 (m, 1H), 2.03 (m, 1H), 1.84 (m, 6H), 1.65 (m, 2H), 1.23 (d, <i>J</i> = 4.1 Hz, 2H). LCMS [M+H] ⁺ = 950.6
UB-180576		¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.12 (s, 1H), 8.98 (s, 1H), 8.57 (t, <i>J</i> = 6.1 Hz, 1H), 7.92 (d, <i>J</i> = 9.3 Hz, 1H), 7.64 (t, <i>J</i> = 5.6 Hz, 1H), 7.54 (t, <i>J</i> = 7.8 Hz, 1H), 7.48 – 7.31 (m, 5H), 7.18 – 7.05 (m, 3H), 6.95 (d, <i>J</i> = 3.6 Hz, 1H), 6.86 (d, <i>J</i> = 8.2 Hz, 1H), 6.63 (d, <i>J</i> = 7.3 Hz, 1H), 5.13 (d, <i>J</i> = 3.5 Hz, 1H), 4.55 (d, <i>J</i> = 9.1 Hz, 2H), 4.43 (m, 2H), 4.35 (s, 1H), 4.22 (m, 1H), 3.71 – 3.55 (m, 5H), 3.47 (d, <i>J</i> = 7.6 Hz, 12H), 3.39 – 3.33 (m, 2H), 3.19 (q, <i>J</i> = 6.2 Hz, 2H), 2.91 (s, 2H), 2.44 (s, 3H), 2.35 (m, 1H), 2.03 (m, 1H), 1.90 (m, 2H), 1.78 (m, 4H), 1.66 (m, 2H), 0.93 (s, 9H). LCMS [M+H] ⁺ = 1121.7
UB-180577		¹ H NMR (400 MHz, DMSO-d ₆) δ 11.11 (s, 1H), 11.00 (s, 1H), 9.74 (s, 1H), 7.76 (t, <i>J</i> = 5.7 Hz, 1H), 7.71 (dd, <i>J</i> = 7.9, 1.2 Hz, 1H), 7.57 – 7.46 (m, 3H), 7.35 (d, <i>J</i> = 3.6 Hz, 1H), 7.14 – 7.04 (m, 3H), 6.95 (d, <i>J</i> = 3.6 Hz, 1H), 6.84 (d, <i>J</i> = 8.2 Hz, 1H), 6.62 (d, <i>J</i> = 7.3 Hz, 1H), 5.13 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.49 (s, 1H), 4.43 – 4.28 (m, 2H), 4.11 (s, 2H), 3.53 (t, <i>J</i> = 5.9 Hz, 2H), 2.91 (m, 3H), 2.33 (dd, <i>J</i> = 13.0, 4.8 Hz, 1H), 2.01 – 1.53 (m, 10H). LCMS [M+1] ⁺ = 804.4

UB-180581		¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.12 (s, 1H), 11.02 (s, 1H), 9.85 (s, 1H), 7.82 (d, <i>J</i> = 8.4 Hz, 1H), 7.63 (t, <i>J</i> = 5.6 Hz, 1H), 7.59 – 7.43 (m, 3H), 7.35 (d, <i>J</i> = 3.6 Hz, 1H), 7.21 – 7.05 (m, 3H), 6.95 (d, <i>J</i> = 3.6 Hz, 1H), 6.86 (d, <i>J</i> = 8.0 Hz, 1H), 6.63 (d, <i>J</i> = 7.2 Hz, 1H), 5.15 (dd, <i>J</i> = 13.2, 5.2 Hz, 1H), 4.54 (s, 1H), 4.45 – 4.26 (m, 2H), 3.70 (t, <i>J</i> = 6.4 Hz, 2H), 3.55 – 3.41 (m, 10H), 3.17 (q, <i>J</i> = 5.6 Hz, 2H), 2.90 (s, 2H), 2.65 – 2.55 (m, 4H), 2.33 (d, <i>J</i> = 17.5 Hz, 1H), 2.08 – 1.94 (m, 2H), 1.91 – 1.74 (m, 5H), 1.70 – 1.59 (m, 2H). LCMS [M+ H] ⁺ = 906.3.
UB-180582		¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.12 (s, 1H), 11.01 (s, 1H), 9.70 (s, 1H), 7.74 (d, <i>J</i> = 7.6 Hz, 1H), 7.63 (t, <i>J</i> = 5.6 Hz, 1H), 7.59 – 7.43 (m, 3H), 7.36 (d, <i>J</i> = 3.6 Hz, 1H), 7.23 – 7.04 (m, 3H), 6.95 (d, <i>J</i> = 3.6 Hz, 1H), 6.86 (d, <i>J</i> = 8.0 Hz, 1H), 6.63 (d, <i>J</i> = 7.2 Hz, 1H), 5.14 (dd, <i>J</i> = 13.2, 5.2 Hz, 1H), 4.58 – 4.49 (m, 1H), 4.46 – 4.30 (m, 2H), 4.13 (s, 2H), 3.73 – 3.64 (m, 2H), 3.64 – 3.56 (m, 2H), 3.56 – 3.41 (m, 8H), 3.31 – 3.25 (m, 2H), 3.24 – 3.06 (m, 3H), 2.98 – 2.82 (m, 3H), 2.65 – 2.55 (m, 1H), 2.43 – 2.33 (m, 1H), 2.07 – 1.93 (m, 2H), 1.90 – 1.74 (m, 5H), 1.70 – 1.58 (m, 2H). LCMS [M+ H] ⁺ = 936.6.
UB-180590		¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.11 (s, 1H), 11.02 (s, 1H), 9.85 (s, 1H), 7.81 (dd, <i>J</i> = 7.2, 1.6 Hz, 1H), 7.63 (t, <i>J</i> = 5.6 Hz, 1H), 7.58 – 7.39 (m, 3H), 7.35 (d, <i>J</i> = 3.6 Hz, 1H), 7.22 – 7.04 (m, 3H), 6.95 (d, <i>J</i> = 3.6 Hz, 1H), 6.86 (d, <i>J</i> = 8.4 Hz, 1H), 6.62 (d, <i>J</i> = 7.2 Hz, 1H), 5.32 (t, <i>J</i> = 4.8 Hz, 1H), 5.14 (dd, <i>J</i> = 13.2, 5.2 Hz, 1H), 4.54 (s, 1H), 4.42 – 4.28 (m, 2H), 3.71 (t, <i>J</i> = 6.3 Hz, 2H), 3.51 (q, <i>J</i> = 5.2 Hz, 3H), 3.17 (d, <i>J</i> = 5.9 Hz, 2H), 2.99 – 2.81 (m, 3H), 2.65 – 2.56 (m, 2H), 2.39 – 2.26 (m, 2H), 2.07 – 1.94 (m, 3H), 1.93 – 1.71 (m, 5H), 1.65 (d, <i>J</i> = 12.8 Hz, 2H), 1.46 (d, <i>J</i> = 7.6 Hz, 1H). LCMS [M+ H]: 862.5.
UB-180588		¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.12 (s, 1H), 11.00 (s, 1H), 9.74 (s, 1H), 7.75 (dd, <i>J</i> = 7.8, 1.1 Hz, 1H), 7.63 (t, <i>J</i> = 5.6 Hz, 1H), 7.57 – 7.44 (m, 3H), 7.35 (d, <i>J</i> = 3.6 Hz, 1H), 7.10 (d, <i>J</i> = 4.0 Hz, 3H), 6.94 (d, <i>J</i> = 3.6 Hz, 1H), 6.86 (d, <i>J</i> = 8.2 Hz, 1H), 6.58 (d, <i>J</i> = 7.2 Hz, 1H), 5.13 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.50 (s, 1H), 4.48 – 4.25 (m, 2H), 4.08 (s, 2H), 3.48 (t, <i>J</i> = 6.2 Hz, 2H), 3.17 (d, <i>J</i> = 6.3 Hz, 2H), 2.89 (s, 3H), 2.59 (s, 1H), 2.36 (dd, <i>J</i> = 13.2, 4.6 Hz, 1H), 2.02 – 1.95 (m, 1H), 1.92 – 1.51 (m, 10H). LCMS [M+H] ⁺ = 818.5

UB-180586		¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.12 (s, 1H), 11.00 (s, 1H), 9.75 (s, 1H), 7.74 (dd, <i>J</i> = 7.8, 1.2 Hz, 1H), 7.64 – 7.57 (m, 1H), 7.59 – 7.46 (m, 3H), 7.35 (d, <i>J</i> = 3.6 Hz, 1H), 7.23 – 7.02 (m, 3H), 6.95 (d, <i>J</i> = 3.6 Hz, 1H), 6.86 (d, <i>J</i> = 8.2 Hz, 1H), 6.59 (d, <i>J</i> = 7.3 Hz, 1H), 5.13 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.53 (s, 1H), 4.47 – 4.21 (m, 2H), 4.09 (s, 2H), 3.52 (t, <i>J</i> = 6.2 Hz, 2H), 3.06 (d, <i>J</i> = 6.2 Hz, 2H), 2.90 (s, 3H), 2.59 (d, <i>J</i> = 17.4 Hz, 1H), 2.37 (dd, <i>J</i> = 13.1, 4.5 Hz, 1H), 2.07 – 1.93 (m, 1H), 1.93 – 1.40 (m, 12H). LCMS [M+H] ⁺ = 832.3
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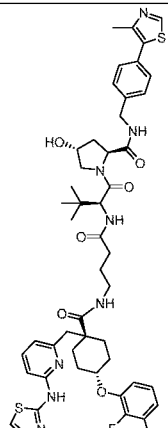
(2.4) Preparation procedure 4 of TED molecule:

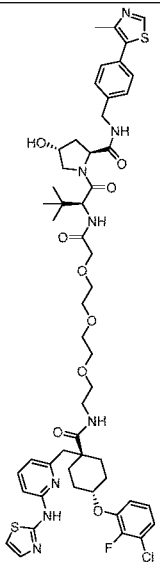
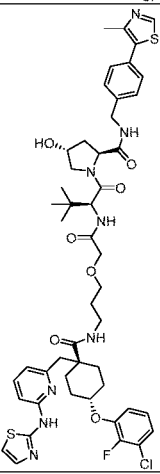
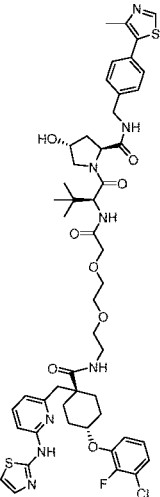


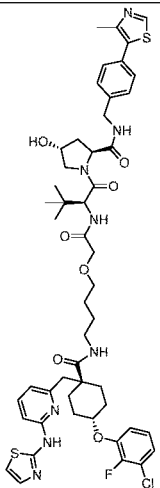
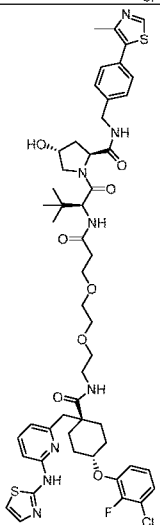
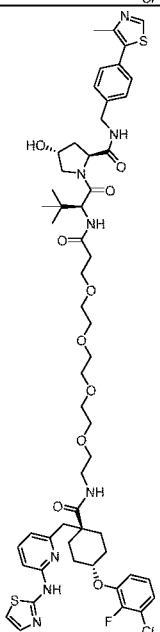
- Compound **UBI-1001** (20 mg, 0.043 mmol) was weighed and dissolved in DMF (2 mL), added
- 5 **NH₂-linker-B(B1)** (1 eq.), **HOBT** (2eq. 23mg, 0.086mmol), **EDCI** (2eq. 16.4mg, 0.086mmol) and **TEA** (3eq. 13mg, 0.129mmol) in an ice-water bath successively. After completion of addition, the material system was stirred and reacted at room temperature under the protection of nitrogen for 18 hours. After completion of reaction, the reaction solution was poured into 5 mL of water and extracted with ethyl acetate (5 mL*3) three times. The organic phases were combined and washed with saturated brine, dried over
- 10 anhydrous Na₂SO₄ and concentrated by rotary evaporation under reduced pressure to obtain the crude product. After that, the crude product was isolated by thin-layer chromatography on silica gel plate with the polarity of the developing solvent (DCM/MeOH = 10/1) to prepare the target compound TED.

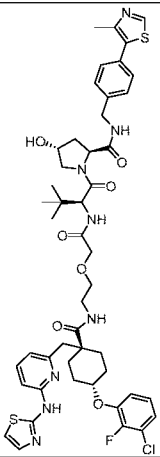
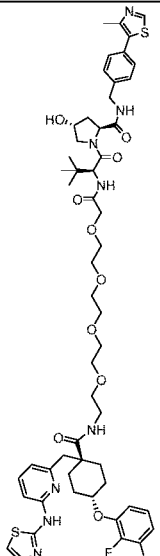
The conjugate of A-L1-B as shown in Table B4 was prepared through similar method.

Table B4

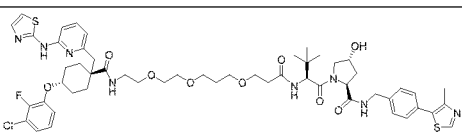
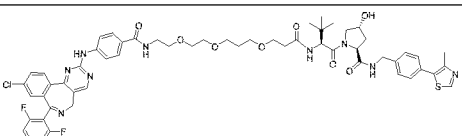
Examples	Compound structure	Analyze data
UB-180583		¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.11 (s, 1H), 8.98 (s, 1H), 8.57 (t, <i>J</i> = 6.1 Hz, 1H), 7.91 (d, <i>J</i> = 9.3 Hz, 1H), 7.63 (t, <i>J</i> = 5.6 Hz, 1H), 7.53 (dd, <i>J</i> = 8.2, 7.4 Hz, 1H), 7.45 – 7.32 (m, 5H), 7.21 – 7.03 (m, 3H), 6.96 (d, <i>J</i> = 3.6 Hz, 1H), 6.86 (d, <i>J</i> = 8.1 Hz, 1H), 6.59 (d, <i>J</i> = 7.3 Hz, 1H), 5.14 (d, <i>J</i> = 3.6 Hz, 1H), 4.55 (d, <i>J</i> = 8.9 Hz, 2H), 4.50 – 4.40 (m, 2H), 4.36 (s, 1H), 4.22 (dd, <i>J</i> = 15.8, 5.4 Hz, 1H), 3.73 – 3.58 (m, 2H), 3.02 (q, <i>J</i> = 6.8 Hz, 2H), 2.91 (s, 2H), 2.44 (s, 3H), 2.26 – 2.18 (m, 1H), 2.20 – 1.92 (m, 2H), 1.98 – 1.46 (m, 10H), 0.94 (s, 9H). LCMS [M/2+H] ⁺ = 480.3

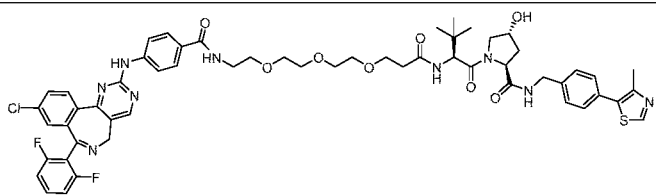
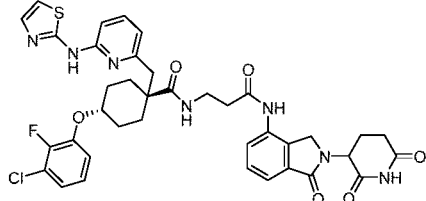
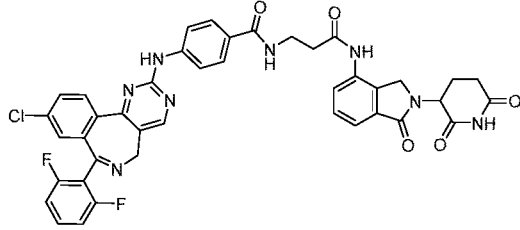
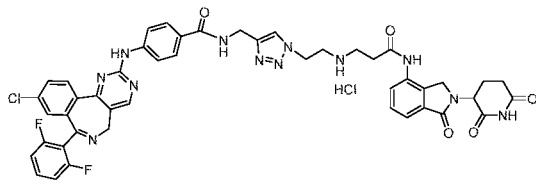
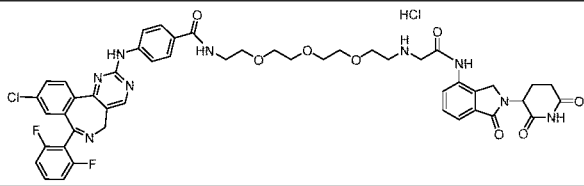
UB-180584	 The chemical structure of UB-180584 features a central core consisting of a 1,2,3,4-tetrahydro-1H-benzopyrrolidine ring system. This core is substituted with a 4-(4-methyl-1,3,4-thiazol-5-yl)phenyl group at the 1-position, a 4-chloro-2-fluorophenyl group at the 3-position, and a 4-(4-methyl-1,3,4-thiazol-5-yl)phenyl group at the 4-position. The 1-position is also substituted with a 4-(4-methyl-1,3,4-thiazol-5-yl)phenyl group. The 3-position is substituted with a 4-chloro-2-fluorophenyl group. The 4-position is substituted with a 4-(4-methyl-1,3,4-thiazol-5-yl)phenyl group. The 1-position is also substituted with a 4-(4-methyl-1,3,4-thiazol-5-yl)phenyl group.	^1H NMR (400 MHz, DMSO-d_6) δ 11.12 (s, 1H), 8.98 (s, 1H), 8.60 (t, J = 6.0 Hz, 1H), 7.64 (t, J = 5.6 Hz, 1H), 7.54 (t, J = 7.8 Hz, 1H), 7.45 – 7.35 (m, 6H), 7.16 – 7.08 (m, 3H), 6.95 (d, J = 3.7 Hz, 1H), 6.86 (d, J = 8.2 Hz, 1H), 6.63 (d, J = 7.3 Hz, 1H), 5.17 (d, J = 3.5 Hz, 1H), 4.58 – 4.52 (m, 2H), 4.46 – 4.35 (m, 3H), 4.30 – 4.23 (m, 1H), 3.96 (s, 2H), 3.76 – 3.41 (m, 11H), 3.18 (q, J = 6.0 Hz, 2H), 2.91 (s, 2H), 2.44 (s, 3H), 2.11 – 2.03 (m, 1H), 1.95 – 1.58 (m, 9H), 0.94 (s, 9H). LCMS $[\text{M}/2+\text{H}]^+ = 532.3$
UB-180585	 The chemical structure of UB-180585 is similar to UB-180584, but with a different substitution pattern on the central core. It features a 1,2,3,4-tetrahydro-1H-benzopyrrolidine ring system. The 1-position is substituted with a 4-(4-methyl-1,3,4-thiazol-5-yl)phenyl group. The 3-position is substituted with a 4-chloro-2-fluorophenyl group. The 4-position is substituted with a 4-(4-methyl-1,3,4-thiazol-5-yl)phenyl group. The 1-position is also substituted with a 4-(4-methyl-1,3,4-thiazol-5-yl)phenyl group.	^1H NMR (400 MHz, DMSO-d_6) δ 11.11 (s, 1H), 8.96 (s, 1H), 8.60 (t, J = 6.0 Hz, 1H), 7.62 (d, J = 6.2 Hz, 1H), 7.54 (t, J = 7.8 Hz, 1H), 7.47 – 7.27 (m, 6H), 7.21 – 7.04 (m, 3H), 6.96 (d, J = 3.6 Hz, 1H), 6.86 (d, J = 8.2 Hz, 1H), 6.58 (d, J = 7.3 Hz, 1H), 5.17 (d, J = 3.5 Hz, 1H), 4.56 (m, 2H), 4.45 (t, J = 8.3 Hz, 1H), 4.32 (dd, J = 26.7, 5.6 Hz, 3H), 3.91 (m, 2H), 3.71 – 3.53 (m, 2H), 3.40 (m, 2H), 3.11 (m, 2H), 2.90 (s, 2H), 2.43 (s, 3H), 2.06 (s, 1H), 1.96 – 1.53 (m, 10H), 0.94 (s, 9H). LCMS $[\text{M}/2+1]^+ = 495.3$
UB-180587	 The chemical structure of UB-180587 is similar to UB-180584, but with a different substitution pattern on the central core. It features a 1,2,3,4-tetrahydro-1H-benzopyrrolidine ring system. The 1-position is substituted with a 4-(4-methyl-1,3,4-thiazol-5-yl)phenyl group. The 3-position is substituted with a 4-chloro-2-fluorophenyl group. The 4-position is substituted with a 4-(4-methyl-1,3,4-thiazol-5-yl)phenyl group. The 1-position is also substituted with a 4-(4-methyl-1,3,4-thiazol-5-yl)phenyl group.	^1H NMR (400 MHz, DMSO-d_6) δ 11.11 (s, 1H), 8.97 (s, 1H), 8.58 (t, J = 5.9 Hz, 1H), 7.65 (t, J = 5.6 Hz, 1H), 7.54 (dd, J = 8.2, 7.4 Hz, 1H), 7.46 – 7.32 (m, 6H), 7.18 – 7.05 (m, 3H), 6.95 (d, J = 3.6 Hz, 1H), 6.86 (d, J = 8.2 Hz, 1H), 6.62 (d, J = 7.4 Hz, 1H), 5.17 (d, J = 3.5 Hz, 1H), 4.58 (d, J = 9.6 Hz, 1H), 4.53 (s, 1H), 4.45 (t, J = 8.1 Hz, 1H), 4.39 – 4.22 (m, 3H), 3.97 (s, 2H), 3.67 – 3.50 (m, 5H), 3.40 (t, J = 6.4 Hz, 2H), 3.22 (d, J = 6.1 Hz, 2H), 2.91 (s, 2H), 2.43 (s, 3H), 2.07 (t, J = 10.3 Hz, 1H), 1.98 – 1.57 (m, 10H), 0.94 (s, 9H). LCMS $[\text{M}+\text{H}]^+ = 1019.6$

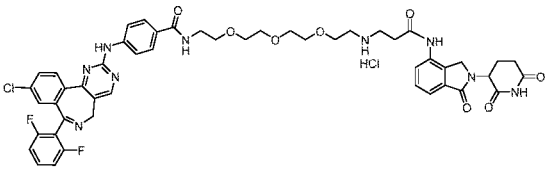
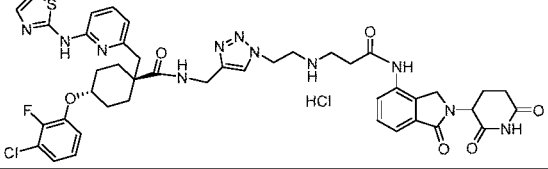
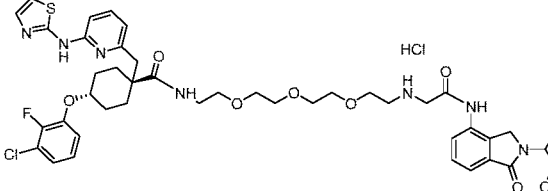
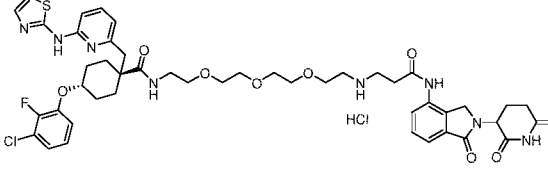
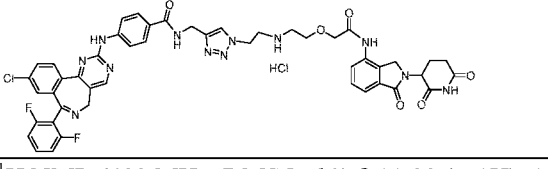
UB-180589	 The chemical structure of UB-180589 features a central core consisting of a 4-chloro-2-fluorophenyl group linked via an ether to a cyclohexane ring. This cyclohexane is also linked to a pyridine ring, which is further connected to a thiazole ring. A side chain on the cyclohexane includes a carbonyl group and a hydroxyl group. The side chain is further extended with a long aliphatic chain containing multiple ether linkages, ending in a thiazole ring.	^1H NMR (400 MHz, DMSO-d_6) δ 11.11 (s, 1H), 8.97 (s, 1H), 8.61 (t, J = 6.0 Hz, 1H), 7.63 – 7.50 (m, 2H), 7.37 (m, 6H), 7.21 – 7.05 (m, 3H), 6.95 (d, J = 3.6 Hz, 1H), 6.86 (d, J = 8.2 Hz, 1H), 6.59 (d, J = 7.4 Hz, 1H), 5.17 (d, J = 3.5 Hz, 1H), 4.57 (m, 2H), 4.51 – 4.19 (m, 4H), 3.92 (d, J = 1.4 Hz, 2H), 3.75 – 3.54 (m, 2H), 3.45 (t, J = 5.2 Hz, 2H), 3.05 (d, J = 6.0 Hz, 2H), 2.90 (s, 2H), 2.44 (s, 3H), 2.18 – 2.00 (m, 1H), 1.95 – 1.34 (m, 13H), 0.94 (s, 9H). LCMS $[\text{M}+2+\text{H}]^+ = 502.9$
UB-180599 (199)	 The chemical structure of UB-180599 is similar to UB-180589, but with a different side chain configuration. It features the same central core, but the side chain on the cyclohexane ring is modified, leading to a different overall molecular weight and NMR profile.	^1H NMR (400 MHz, DMSO-d_6) δ 11.12 (s, 1H), 8.98 (s, 1H), 8.58 (t, J = 6.0 Hz, 1H), 7.93 (d, J = 9.3 Hz, 1H), 7.66 (t, J = 5.6 Hz, 1H), 7.54 (t, J = 7.8 Hz, 1H), 7.47 – 7.30 (m, 4H), 7.22 – 7.05 (m, 3H), 6.95 (d, J = 3.6 Hz, 1H), 6.86 (d, J = 8.1 Hz, 1H), 6.63 (d, J = 7.2 Hz, 1H), 5.39 – 5.27 (m, 1H), 5.14 (d, J = 3.5 Hz, 1H), 4.64 – 4.51 (m, 2H), 4.50 – 4.38 (m, 2H), 4.38 – 4.31 (m, 1H), 4.22 (dd, J = 15.9, 5.4 Hz, 1H), 3.68 – 3.56 (m, 3H), 3.52 – 3.43 (m, 4H), 3.23 – 3.13 (m, 2H), 2.91 (s, 2H), 2.44 (s, 3H), 2.39 – 2.34 (m, 1H), 2.08 – 1.95 (m, 3H), 1.94 – 1.71 (m, 6H), 1.70 – 1.59 (m, 2H), 1.54 – 1.42 (m, 2H), 1.24 (s, 9H). $[\text{M}+\text{H}]^+ = 1033.6$.
UB-180576	 The chemical structure of UB-180576 is similar to the others, but with a different side chain configuration. It features the same central core, but the side chain on the cyclohexane ring is modified, leading to a different overall molecular weight and NMR profile.	^1H NMR (400 MHz, DMSO-d_6) δ 11.12 (s, 1H), 8.98 (s, 1H), 8.57 (t, J = 6.1 Hz, 1H), 7.92 (d, J = 9.3 Hz, 1H), 7.64 (t, J = 5.6 Hz, 1H), 7.54 (t, J = 7.8 Hz, 1H), 7.48 – 7.31 (m, 5H), 7.18 – 7.05 (m, 3H), 6.95 (d, J = 3.6 Hz, 1H), 6.86 (d, J = 8.2 Hz, 1H), 6.63 (d, J = 7.3 Hz, 1H), 5.13 (d, J = 3.5 Hz, 1H), 4.55 (d, J = 9.1 Hz, 2H), 4.43 (m, 2H), 4.35 (s, 1H), 4.22 (m, 1H), 3.71 – 3.55 (m, 5H), 3.47 (d, J = 7.6 Hz, 12H), 3.39 – 3.33 (m, 2H), 3.19 (q, J = 6.2 Hz, 2H), 2.91 (s, 2H), 2.44 (s, 3H), 2.35 (m, 1H), 2.03 (m, 1H), 1.90 (m, 2H), 1.78 (m, 4H), 1.66 (m, 2H), 0.93 (s, 9H). LCMS $[\text{M}+\text{H}]^+ = 1121.7$

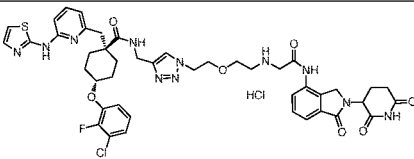
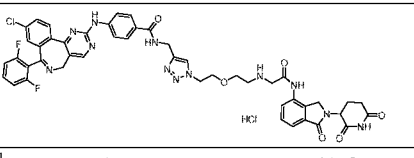
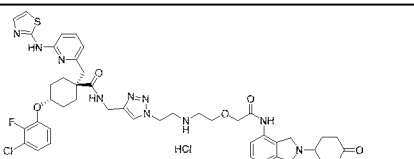
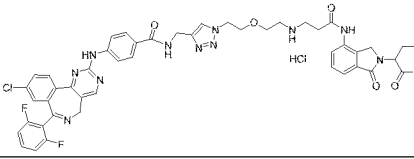
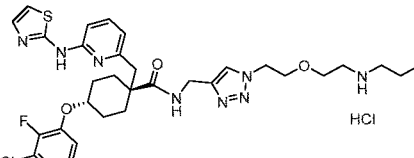
UB-180578		^1H NMR (400 MHz, DMSO-d_6) δ 11.11 (s, 1H), 8.96 (d, J = 2.8 Hz, 1H), 8.59 (d, J = 7.1 Hz, 1H), 7.76 (s, 1H), 7.55 (t, J = 7.9 Hz, 1H), 7.47 (d, J = 9.6 Hz, 1H), 7.37 (m, 5H), 7.11 (m, 3H), 6.95 (t, J = 3.2 Hz, 1H), 6.86 (d, J = 8.3 Hz, 1H), 6.63 (d, J = 7.3 Hz, 1H), 5.16 (d, J = 3.5 Hz, 1H), 4.56 (t, J = 9.7 Hz, 2H), 4.48 – 4.23 (m, 4H), 3.94 (t, J = 10.2 Hz, 2H), 3.71 – 3.58 (m, 2H), 3.48 (m, 2H), 3.24 (m, 2H), 2.92 (m, 2H), 2.43 (d, J = 2.9 Hz, 3H), 2.13 – 1.57 (m, 10H), 0.93 (s, 9H). LCMS $[\text{M}+\text{H}]^+$ = 975.6
UB-180580		^1H NMR (400 MHz, DMSO-d_6) δ 11.12 (s, 1H), 8.98 (s, 1H), 8.60 (t, J = 6.0 Hz, 1H), 7.64 (t, J = 5.6 Hz, 1H), 7.54 (t, J = 7.6 Hz, 1H), 7.47 – 7.32 (m, 6H), 7.19 – 7.07 (m, 3H), 6.95 (d, J = 3.6 Hz, 1H), 6.86 (d, J = 8.0 Hz, 1H), 6.63 (d, J = 7.2 Hz, 1H), 5.15 (d, J = 3.6 Hz, 1H), 4.60 – 4.49 (m, 2H), 4.47 – 4.33 (m, 3H), 4.25 (dd, J = 15.6, 5.6 Hz, 1H), 3.96 (s, 2H), 3.70 – 3.64 (m, 1H), 3.63 – 3.57 (m, 3H), 3.57 – 3.50 (m, 6H), 3.49 – 3.44 (m, 4H), 3.22 – 3.13 (m, 2H), 2.90 (s, 1H), 2.89 (s, 1H), 2.44 (s, 3H), 2.09 – 2.02 (m, 1H), 1.94 – 1.84 (m, 3H), 1.84 – 1.73 (m, 4H), 1.69 – 1.60 (m, 2H), 0.94 (s, 9H). LCMS $[\text{M}+\text{H}]^+$ = 1107.7.

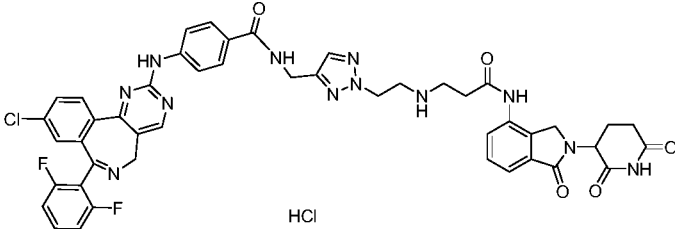
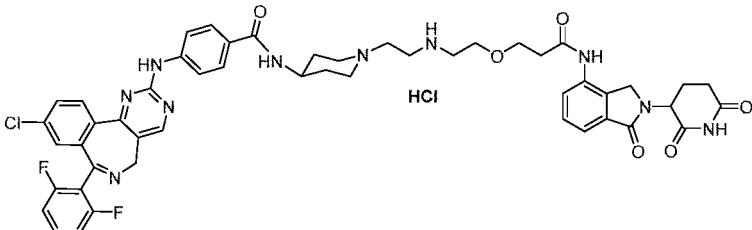
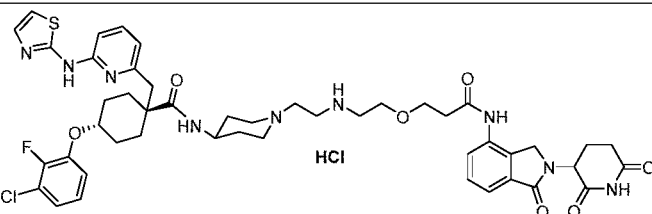
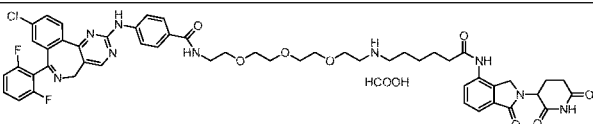
Compounds in Table B9 were prepared by preparation procedures similar to (2.1), (2.2), (2.3) or (2.4)
Table B9

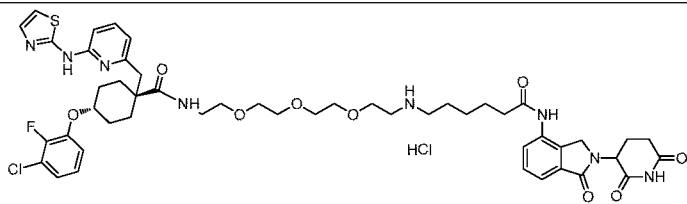
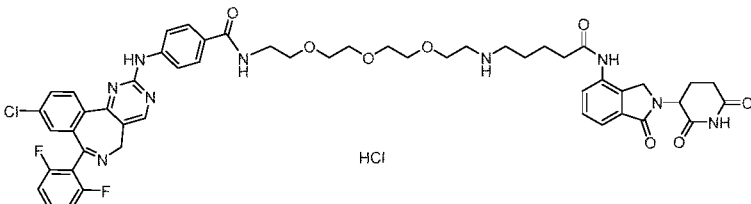
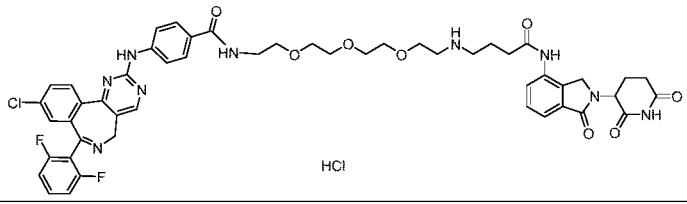
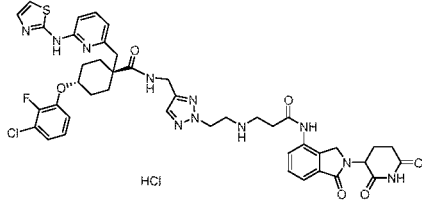
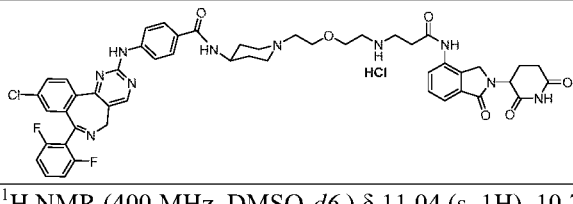
Examples	Structural data analysis
UB-180600 (100)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.12 (s, 1H), 8.98 (s, 1H), 8.57 (t, J = 6.1 Hz, 1H), 7.90 (d, J = 9.4 Hz, 1H), 7.65 (t, J = 5.5 Hz, 1H), 7.54 (dd, J = 8.2, 7.3 Hz, 1H), 7.45 – 7.32 (m, 5H), 7.22 – 7.05 (m, 3H), 6.95 (d, J = 3.6 Hz, 1H), 6.87 (d, J = 8.2 Hz, 1H), 6.64 (d, J = 7.3 Hz, 1H), 5.14 (d, J = 3.6 Hz, 1H), 4.56 (d, J = 9.8 Hz, 2H), 4.49 – 4.38 (m, 2H), 4.35 (s, 1H), 4.22 (dd, J = 15.9, 5.5 Hz, 1H), 3.74 – 3.42 (m, 9H), 3.42 – 3.36 (m, 6H), 3.19 (q, J = 5.9 Hz, 2H), 2.91 (s, 2H), 2.44 (s, 3H), 2.31 (dt, J = 14.5, 5.8 Hz, 1H), 2.02 (d, J = 9.1 Hz, 1H), 1.95 – 1.55 (m, 11H), 0.93 (s, 9H). LCMS $[\text{M}/2+1]^+$ = 546.9
UB-180609 (109)	 ^1H NMR (400 MHz, DMSO-d_6) δ 10.16 (s, 1H), 8.98 (s, 1H), 8.72 (s, 1H), 8.56 (t, J = 6.1 Hz, 1H), 8.34 (t, J = 5.6 Hz, 1H), 8.30 (d, J = 8.5 Hz, 1H), 7.92 – 7.83 (m, 6H), 7.55 (ddd, J = 15.0, 8.4, 6.6 Hz, 1H), 7.44 – 7.31 (m, 5H), 7.17 (t, J = 8.8 Hz, 2H), 5.13 (d, J = 3.5 Hz, 1H), 4.55 (d, J = 9.4 Hz, 1H), 4.42 (td, J = 7.3, 6.5, 3.9 Hz, 2H), 4.35 (s, 1H), 4.22 (dd, J = 15.8, 5.4 Hz, 1H), 3.74 – 3.44 (m, 11H), 3.44 – 3.36 (m, 7H), 3.14 (s, 1H), 2.44 (s, 3H), 2.32 (dt, J = 14.9, 5.9 Hz, 1H), 2.03 (t, J = 10.3 Hz, 1H), 1.90 (ddd, J = 12.9, 8.6, 4.6 Hz, 1H), 1.69 (p, J = 6.5 Hz, 2H), 0.93 (s, 9H). LCMS $[\text{M}+1]^+$ = 1106.3

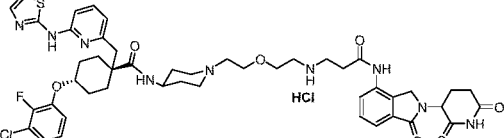
UB-180615 (115)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 10.16 (s, 1H), 8.97 (s, 1H), 8.72 (s, 1H), 8.56 (t, <i>J</i> = 6.0 Hz, 1H), 8.34 (t, <i>J</i> = 5.6 Hz, 1H), 8.30 (d, <i>J</i> = 8.4 Hz, 1H), 7.97 - 7.79 (m, 6H), 7.58 - 7.51 (m, 1H), 7.44 - 7.36 (m, 4H), 7.34 (d, <i>J</i> = 2.0 Hz, 1H), 7.21 - 7.13 (m, 2H), 5.13 (d, <i>J</i> = 3.6 Hz, 1H), 4.55 (d, <i>J</i> = 9.2 Hz, 1H), 4.47 - 4.39 (m, 2H), 4.37 - 4.32 (m, 1H), 4.21 (dd, <i>J</i> = 15.6, 5.6 Hz, 1H), 3.69 - 3.57 (m, 4H), 3.55 - 3.45 (m, 10H), 3.43 - 3.38 (m, 3H), 2.48 - 2.46 (m, 1H), 2.44 (s, 3H), 2.39 - 2.34 (m, 1H), 2.08 - 1.95 (m, 2H), 1.93 - 1.87 (m, 1H), 0.93 (s, 9H). LCMS [M+H]⁺: 1092.3.
UB-180616 (116)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.12 (s, 1H), 11.02 (s, 1H), 9.86 (s, 1H), 7.85 - 7.75 (m, 2H), 7.57 - 7.44 (m, 3H), 7.36 (d, <i>J</i> = 3.6 Hz, 1H), 7.09 (d, <i>J</i> = 4.3 Hz, 3H), 6.95 (d, <i>J</i> = 3.6 Hz, 1H), 6.85 (d, <i>J</i> = 8.2 Hz, 1H), 6.61 (d, <i>J</i> = 7.4 Hz, 1H), 5.14 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.47 (s, 1H), 4.35 (s, 2H), 2.91 (m, 3H), 2.61 - 2.54 (m, 3H), 2.37 - 2.23 (m, 2H), 2.08 - 1.57 (m, 10H). LCMS [M+H]⁺ = 774.2
UB-180617 (117)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.01 (s, 1H), 10.16 (s, 1H), 9.93 (s, 1H), 8.73 (s, 1H), 8.48 (t, <i>J</i> = 5.7 Hz, 1H), 8.30 (d, <i>J</i> = 8.5 Hz, 1H), 7.91 - 7.79 (m, 6H), 7.62 - 7.43 (m, 3H), 7.34 (d, <i>J</i> = 2.2 Hz, 1H), 7.17 (t, <i>J</i> = 8.7 Hz, 2H), 5.11 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.33 (s, 2H), 4.06 - 3.94 (m, 1H), 3.67 - 3.52 (m, 2H), 2.98 - 2.82 (m, 1H), 2.73 - 2.63 (m, 2H), 2.36 - 2.19 (m, 2H), 1.98 (d, <i>J</i> = 5.4 Hz, 2H). LCMS [M+H]⁺ = 789.2
UB-180726 (226)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 10.21 (d, <i>J</i> = 12.0 Hz, 2H), 9.17 (s, 2H), 8.92 (t, <i>J</i> = 5.8 Hz, 1H), 8.73 (s, 1H), 8.33 - 8.25 (m, 1H), 8.09 (s, 1H), 7.94 - 7.80 (m, 7H), 7.59 - 7.47 (m, 3H), 7.34 (d, <i>J</i> = 2.1 Hz, 1H), 7.17 (t, <i>J</i> = 8.6 Hz, 2H), 5.16 (dd, <i>J</i> = 13.3, 5.2 Hz, 1H), 4.73 (t, <i>J</i> = 6.4 Hz, 2H), 4.53 (d, <i>J</i> = 5.6 Hz, 2H), 4.49 - 4.33 (m, 2H), 3.52 (d, <i>J</i> = 6.2 Hz, 2H), 3.32 - 3.21 (m, 2H), 2.98 - 2.86 (m, 3H), 2.67 - 2.58 (m, 1H), 2.32 (dt, <i>J</i> = 13.6, 6.7 Hz, 1H), 2.09 - 1.98 (m, 1H). LCMS [M+H]⁺ = 913.3
UB-180727 (227)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.05 (s, 1H), 10.73 (s, 1H), 10.18 (s, 1H), 9.15 (s, 2H), 8.73 (s, 1H), 8.40 (t, <i>J</i> = 5.7 Hz, 1H), 8.30 (d, <i>J</i> = 8.5 Hz, 1H), 7.97 - 7.78 (m, 7H), 7.64 - 7.49 (m, 3H), 7.35 (d, <i>J</i> = 2.3 Hz, 1H), 7.17 (t, <i>J</i> = 8.8 Hz, 2H), 4.52 - 4.31 (m, 2H), 4.05 (t, <i>J</i> = 5.5 Hz, 2H), 3.73 (t, <i>J</i> = 5.1 Hz, 2H), 3.55 (t, <i>J</i> = 8.9 Hz, 1H), 3.41 (q, <i>J</i> = 5.8 Hz, 2H), 3.26 - 3.17 (m, 2H), 2.93 (ddd, <i>J</i> = 18.8, 11.2, 4.5 Hz, 1H), 2.65 (s, 1H), 2.37 - 2.24 (m, 1H), 2.11 - 2.00 (m, 1H). LCMS [M+H]⁺ = 950.2

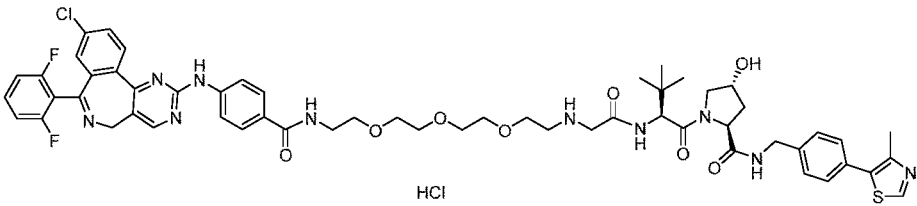
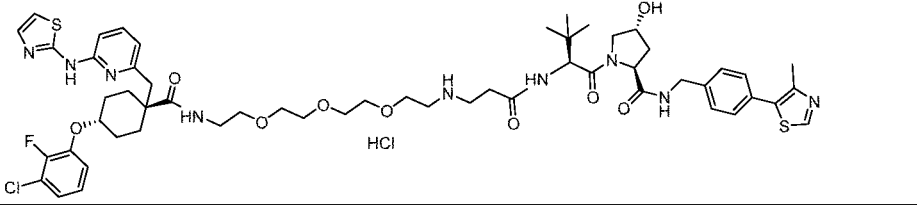
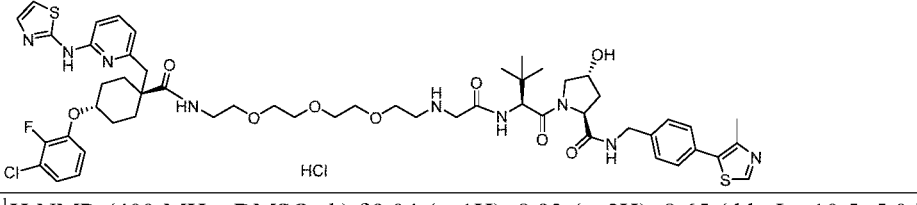
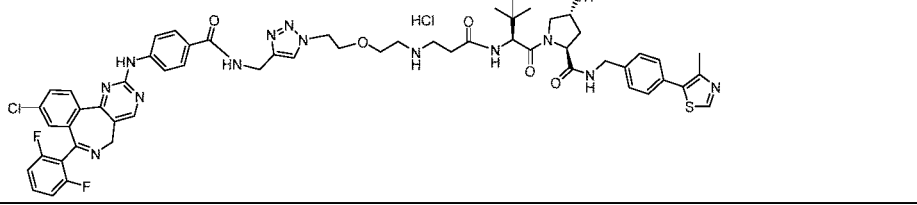
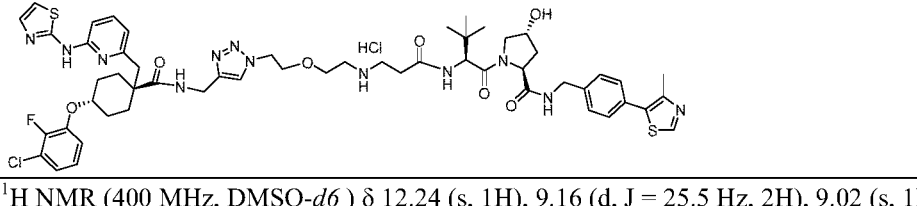
UB-180728 (228)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 10.20 (d, J = 16.5 Hz, 2H), 8.87 (s, 2H), 8.73 (s, 1H), 8.39 (t, J = 5.7 Hz, 1H), 8.30 (d, J = 8.5 Hz, 1H), 7.96 – 7.79 (m, 6H), 7.62 – 7.46 (m, 3H), 7.34 (d, J = 2.2 Hz, 1H), 7.17 (t, J = 8.8 Hz, 2H), 4.51 – 4.32 (m, 2H), 3.70 (t, J = 5.1 Hz, 2H), 3.63 – 3.49 (m, 11H), 3.41 (q, J = 6.0 Hz, 2H), 3.24 (d, J = 7.2 Hz, 2H), 3.14 (s, 2H), 2.90 (dt, J = 9.4, 6.4 Hz, 3H), 2.63 (s, 1H), 2.38 – 2.26 (m, 1H), 2.09 – 1.99 (m, 1H). LCMS [M+H]⁺ = 964.2
UB-180729 (229)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.77 (s, 1H), 11.03 (s, 1H), 10.25 (s, 1H), 9.29 (s, 2H), 8.29 (t, J = 5.6 Hz, 1H), 7.95 (s, 1H), 7.84 (dd, J = 7.3, 1.7 Hz, 1H), 7.59 – 7.42 (m, 4H), 7.18 – 7.04 (m, 4H), 6.96 (d, J = 8.2 Hz, 1H), 6.62 (d, J = 7.4 Hz, 1H), 5.16 (dd, J = 13.2, 5.2 Hz, 1H), 4.76 (t, J = 6.4 Hz, 2H), 4.55 (s, 1H), 4.50 – 4.33 (m, 2H), 4.29 (d, J = 5.4 Hz, 2H), 3.50 (s, 2H), 3.27 (s, 2H), 3.00 – 2.88 (m, 5H), 2.62 (d, J = 17.4 Hz, 1H), 2.33 (d, J = 17.6 Hz, 1H), 2.09 – 1.99 (m, 1H), 1.92 (d, J = 10.9 Hz, 2H), 1.73 (dd, J = 46.8, 12.0 Hz, 6H). LCMS [M+H]⁺ = 898.2
UB-180730 (230)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.75 (s, 1H), 11.05 (s, 1H), 10.66 (s, 1H), 9.11 (s, 2H), 7.87 (dd, J = 7.5, 1.6 Hz, 1H), 7.71 (d, J = 5.7 Hz, 1H), 7.65 – 7.41 (m, 4H), 7.20 – 7.05 (m, 4H), 6.97 (d, J = 8.2 Hz, 1H), 6.73 (d, J = 7.4 Hz, 1H), 5.18 (dd, J = 13.2, 5.1 Hz, 1H), 4.54 (s, 1H), 4.50 – 4.33 (m, 2H), 4.05 (s, 2H), 3.71 (d, J = 5.1 Hz, 2H), 3.51 (dq, J = 13.1, 5.2, 4.4 Hz, 5H), 3.35 (t, J = 6.2 Hz, 2H), 3.19 (d, J = 12.4 Hz, 4H), 3.00 – 2.89 (m, 3H), 2.64 (d, J = 3.7 Hz, 1H), 2.34 – 2.25 (m, 1H), 2.13 – 2.00 (m, 1H), 1.96 – 1.56 (m, 8H). LCMS [M+H]⁺ = 935.2
UB-180731 (231)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.95 (s, 1H), 11.03 (s, 1H), 10.25 (s, 1H), 8.95 (s, 2H), 7.85 (dd, J = 7.3, 1.7 Hz, 1H), 7.72 (t, J = 5.7 Hz, 1H), 7.65 (t, J = 7.8 Hz, 1H), 7.55 – 7.45 (m, 3H), 7.20 – 7.06 (m, 4H), 7.00 (d, J = 8.2 Hz, 1H), 6.76 (d, J = 7.4 Hz, 1H), 5.16 (dd, J = 13.2, 5.1 Hz, 1H), 4.55 (s, 1H), 4.51 – 4.36 (m, 2H), 3.70 (t, J = 5.2 Hz, 2H), 3.54 – 3.43 (m, 5H), 3.35 (t, J = 6.2 Hz, 2H), 3.28 – 3.10 (m, 6H), 2.93 (dt, J = 19.0, 6.8 Hz, 5H), 2.63 (d, J = 4.0 Hz, 1H), 2.36 – 2.27 (m, 1H), 2.09 – 1.98 (m, 1H), 1.90 (d, J = 12.5 Hz, 2H), 1.85 – 1.57 (m, 6H). LCMS [M+H]⁺ = 949.3
UB-180732 (232)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.00 (s, 1H), 10.18 (d, J = 14.5 Hz, 2H), 9.52 (s, 2H), 8.91 (t, J = 5.8 Hz, 1H), 8.74 (d, J = 2.4 Hz, 1H), 8.30 (d, J = 8.5 Hz, 1H), 8.10 (s, 1H), 7.94 – 7.85 (m, 5H), 7.75 (dd, J = 7.8, 1.2 Hz, 1H), 7.60 – 7.49 (m, 3H), 7.35 (d, J = 2.1 Hz, 1H), 7.17 (t, J = 8.7 Hz, 2H), 5.14 (dd, J = 13.3, 5.1 Hz, 1H), 4.80 (t, J = 6.6 Hz, 2H), 4.55 – 4.33 (m, 4H), 4.20 (s, 2H), 3.82 (t, J = 4.9 Hz, 2H), 3.59 – 3.49 (m, 2H), 3.22 (s, 2H), 2.91 (ddd, J = 17.9, 13.5, 5.4 Hz, 1H), 2.62 (s, 1H), 2.37 (dd, J = 13.4, 4.7 Hz, 1H), 2.07 – 1.94 (m, 1H). LCMS [M+H]⁺ = 943.4

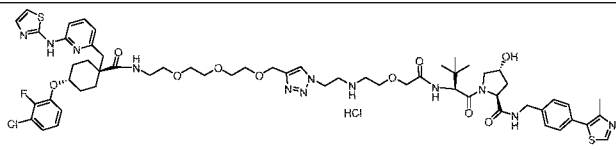
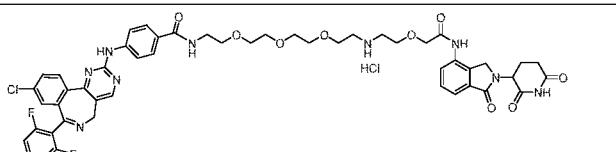
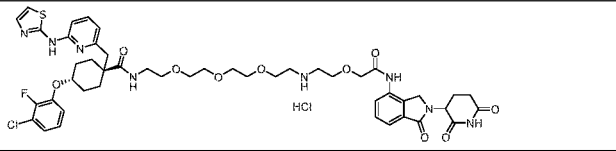
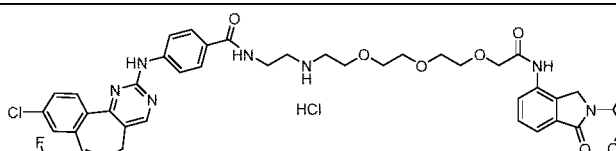
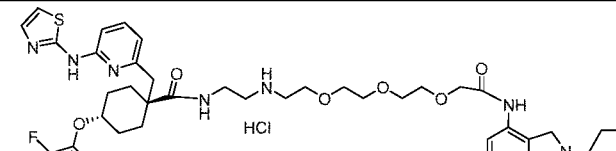
	^1H NMR (400 MHz, DMSO-d_6) δ 11.47 (s, 1H), 11.01 (s, 1H), 10.74 (s, 1H), 10.12 (s, 1H), 9.76 (s, 2H), 7.73 (t, J = 8.8 Hz, 2H), 7.55 (dt, J = 24.3, 7.6 Hz, 3H), 7.40 (d, J = 3.8 Hz, 1H), 7.20 – 7.08 (m, 3H), 7.02 (d, J = 3.7 Hz, 1H), 6.93 (d, J = 8.2 Hz, 1H), 6.65 (d, J = 7.3 Hz, 1H), 5.15 (dd, J = 13.2, 5.2 Hz, 1H), 4.58 – 4.36 (m, 3H), 4.21 (s, 2H), 3.98 – 3.79 (m, 7H), 3.58 (d, J = 11.7 Hz, 3H), 3.27 (s, 2H), 3.08 (t, J = 12.4 Hz, 2H), 2.91 (d, J = 24.1 Hz, 3H), 2.62 (s, 1H), 2.38 (d, J = 4.5 Hz, 1H), 2.06 – 1.72 (m, 10H), 1.65 (d, J = 13.2 Hz, 2H). LCMS $[\text{M}+\text{H}]^+ = 930.1$ 
UB-180742 (242)	^1H NMR (400 MHz, DMSO-d_6) δ 11.82 (s, 1H), 11.04 (s, 1H), 10.73 (d, J = 12.4 Hz, 1H), 9.19 (s, 2H), 8.29 (s, 1H), 7.92 (d, J = 1.5 Hz, 1H), 7.87 (dd, J = 7.7, 1.4 Hz, 1H), 7.63 – 7.35 (m, 4H), 7.11 (dtd, J = 13.4, 6.5, 3.0 Hz, 4H), 6.95 (d, J = 8.6 Hz, 1H), 6.58 (d, J = 8.3 Hz, 1H), 5.17 (dd, J = 13.2, 5.1 Hz, 1H), 4.58 (t, J = 5.3 Hz, 2H), 4.52 – 4.33 (m, 2H), 4.27 (d, J = 5.5 Hz, 2H), 4.04 (s, 2H), 3.85 (t, J = 5.3 Hz, 2H), 3.72 (t, J = 5.0 Hz, 2H), 3.19 (s, 2H), 3.01 – 2.83 (m, 3H), 2.64 (dd, J = 16.8, 12.8 Hz, 1H), 2.36 – 2.16 (m, 2H), 2.05 (dd, J = 9.7, 4.4 Hz, 1H), 1.95 – 1.90 (m, 2H), 1.76 (t, J = 11.8 Hz, 4H), 1.65 (d, J = 13.3 Hz, 2H). LCMS $[\text{M}+\text{H}]^+ = 928.1$ 
UB-180743 (243)	^1H NMR (400 MHz, DMSO-d_6) δ 11.04 (s, 1H), 10.66 (s, 1H), 10.18 (s, 1H), 9.14 (s, 2H), 8.92 (t, J = 5.8 Hz, 1H), 8.72 (s, 1H), 8.28 (d, J = 8.5 Hz, 1H), 8.06 (s, 1H), 7.88 (dd, J = 8.5, 2.2 Hz, 6H), 7.67 – 7.45 (m, 3H), 7.34 (d, J = 2.2 Hz, 1H), 7.17 (t, J = 8.9 Hz, 2H), 5.17 (dd, J = 13.3, 5.2 Hz, 1H), 4.57 (t, J = 5.2 Hz, 2H), 4.49 (t, J = 4.8 Hz, 2H), 4.46 – 4.32 (m, 2H), 4.03 (t, J = 5.6 Hz, 2H), 3.86 (t, J = 5.2 Hz, 2H), 3.72 (t, J = 5.0 Hz, 2H), 3.20 (d, J = 7.6 Hz, 2H), 3.04 – 2.83 (m, 1H), 2.75 – 2.55 (m, 2H), 2.36 – 2.17 (m, 2H), 2.11 – 1.95 (m, 1H). LCMS $[\text{M}+\text{H}]^+ = 943.1$ 
UB-180744 (244)	^1H NMR (400 MHz, DMSO-d_6) δ 11.84 (s, 1H), 11.00 (s, 1H), 10.13 (s, 1H), 9.48 (s, 2H), 8.27 (t, J = 5.7 Hz, 1H), 7.95 (s, 1H), 7.75 (dd, J = 7.8, 1.2 Hz, 1H), 7.62 – 7.43 (m, 4H), 7.18 – 7.05 (m, 4H), 6.97 (d, J = 8.2 Hz, 1H), 6.63 (d, J = 7.4 Hz, 1H), 5.14 (dd, J = 13.3, 5.1 Hz, 1H), 4.80 (t, J = 6.6 Hz, 2H), 4.54 (s, 1H), 4.52 – 4.35 (m, 2H), 4.28 (d, J = 5.4 Hz, 2H), 4.20 (s, 3H), 3.82 (t, J = 4.9 Hz, 2H), 3.53 (s, 2H), 3.23 (s, 2H), 3.03 – 2.83 (m, 3H), 2.61 (d, J = 3.8 Hz, 1H), 2.38 – 2.28 (m, 1H), 2.06 – 1.58 (m, 9H). LCMS $[\text{M}+\text{H}]^+ = 928.1$ 
UB-180748 (248)	^1H NMR (400 MHz, DMSO-d_6) δ 11.02 (s, 1H), 10.19 (s, 1H), 10.05 (s, 1H), 9.11 (s, 2H), 9.01 (s, 1H), 8.92 (t, J = 6.1 Hz, 1H), 8.73 (d, J = 1.1 Hz, 1H), 8.29 (d, J = 8.5 Hz, 1H), 8.06 (d, J = 2.6 Hz, 1H), 7.94 – 7.85 (m, 7H), 7.62 – 7.47 (m, 3H), 7.34 (d, J = 2.2 Hz, 1H), 7.17 (t, J = 8.8 Hz, 3H), 5.15 (dd, J = 13.3, 5.1 Hz, 1H), 4.70 (td, J = 6.6, 2.6 Hz, 2H), 4.52 (dd, J = 5.8, 3.4 Hz, 2H), 4.40 (q, J = 17.5 Hz, 2H), 3.77 (t, J = 6.2 Hz, 2H), 3.70 (t, J = 5.2 Hz, 2H), 3.67 – 3.63 (m, 2H), 3.50 – 3.46 (m, 2H), 3.15 (s, 2H), 2.92 (ddd, J = 17.2, 13.6, 5.3 Hz, 1H), 2.70 (t, J = 6.3 Hz, 2H), 2.61 (d, J = 17.7 Hz, 1H), 2.36 – 2.29 (m, 1H), 2.11 – 1.98 (m, 1H). LCMS $[\text{M}+\text{H}]^+ = 957.1$ 
UB-180749 (249)	

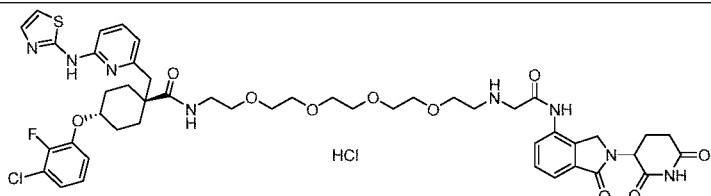
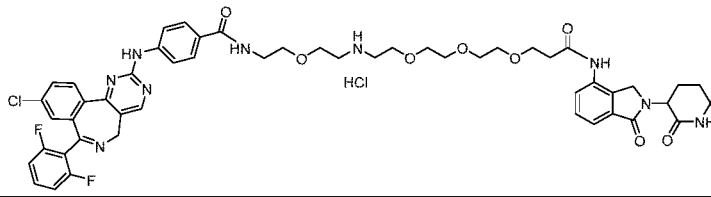
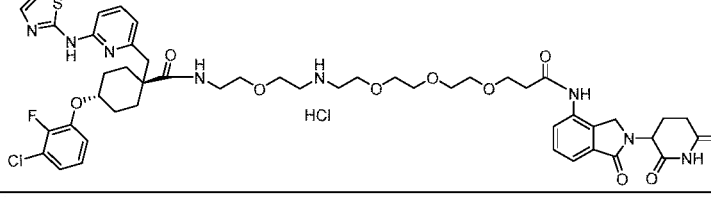
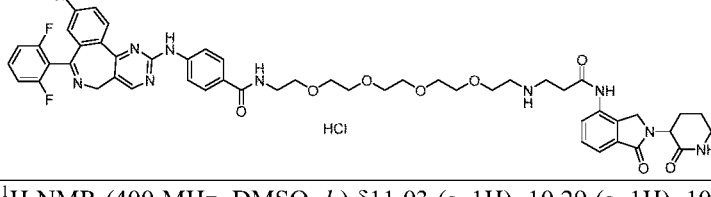
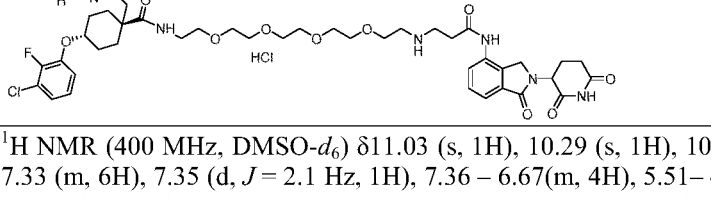
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 12.30 (s, 1H), 11.02 (s, 1H), 10.12 (s, 1H), 9.27 (d, <i>J</i> = 36.0 Hz, 3H), 8.30 (dd, <i>J</i> = 8.3, 3.4 Hz, 1H), 7.93 (d, <i>J</i> = 1.9 Hz, 1H), 7.85 (dd, <i>J</i> = 7.2, 1.7 Hz, 1H), 7.60 (t, <i>J</i> = 7.8 Hz, 1H), 7.56 – 7.47 (m, 3H), 7.16 (td, <i>J</i> = 4.1, 2.4 Hz, 2H), 7.13 – 7.04 (m, 4H), 6.71 (d, <i>J</i> = 7.4 Hz, 1H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.76 – 4.73 (m, 2H), 4.56 (d, <i>J</i> = 4.1 Hz, 2H), 4.41 (d, <i>J</i> = 21.4 Hz, 2H), 4.27 (d, <i>J</i> = 5.1 Hz, 2H), 3.77 (t, <i>J</i> = 6.2 Hz, 2H), 3.73 (t, <i>J</i> = 5.2 Hz, 2H), 3.15 (s, 2H), 2.61 (d, <i>J</i> = 17.6 Hz, 1H), 2.33 (dd, <i>J</i> = 19.6, 6.2 Hz, 1H), 2.07 – 1.64 (m, 12H). LCMS [M+H] ⁺ = 942.1
UB-180761 (261)	 HCl ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.03 (s, 1H), 10.30 (s, 1H), 10.20 (s, 1H), 9.41 (s, 2H), 8.94 (t, <i>J</i> = 5.5 Hz, 1H), 8.74 (d, <i>J</i> = 9.2 Hz, 1H), 8.31 (t, <i>J</i> = 8.4 Hz, 1H), 8.13 – 7.77 (m, 6H), 7.74 (s, 1H), 7.67 – 7.37 (m, 3H), 7.36 – 7.02 (m, 3H), 5.17 – 5.08 (m, 1H), 4.79 (t, <i>J</i> = 6.3 Hz, 2H), 4.55 – 4.29 (m, 4H), 3.53 (dd, <i>J</i> = 6.1, 1.6 Hz, 2H), 3.27 (s, 2H), 2.87 (dt, <i>J</i> = 8.8, 4.5 Hz, 3H), 2.61 (d, <i>J</i> = 16.9 Hz, 1H), 2.56 – 2.47 (m, 1H), 2.47 – 2.14 (m, 1H), 2.14 – 1.88 (m, 1H). LCMS [M+H] ⁺ = 913.2
UB-180768 (268)	 HCl ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.04 (s, 1H), 10.83 (s, 1H), 10.53 (s, 1H), 9.42 (s, 2H), 8.49 (d, <i>J</i> = 7.1 Hz, 1H), 8.19 (s, 1H), 7.98 – 7.82 (m, 2H), 7.62 – 7.46 (m, 5H), 7.22 (t, <i>J</i> = 2.9 Hz, 1H), 5.27 – 5.12 (m, 2H), 4.79 (d, <i>J</i> = 15.4 Hz, 1H), 4.56 – 4.33 (m, 3H), 4.10 (s, 2H), 3.95 (d, <i>J</i> = 5.5 Hz, 3H), 3.83 (d, <i>J</i> = 5.3 Hz, 2H), 3.77 (d, <i>J</i> = 6.4 Hz, 2H), 3.61 (d, <i>J</i> = 11.7 Hz, 2H), 3.30 (s, 2H), 3.24 (s, 3H), 3.14 (d, <i>J</i> = 12.0 Hz, 2H), 2.98 – 2.88 (m, 1H), 2.64 (s, 1H), 2.35 – 2.27 (m, 1H), 2.05 (q, <i>J</i> = 20.1, 13.6 Hz, 5H), 1.88 (t, <i>J</i> = 6.3 Hz, 2H), 0.70 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 959.4
UB-180769 (269)	 HCl ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.49 (s, 1H), 11.03 (s, 1H), 10.38 (s, 1H), 10.25 (d, <i>J</i> = 18.6 Hz, 1H), 9.30 (s, 2H), 7.89 – 7.75 (m, 2H), 7.62 – 7.49 (m, 3H), 7.43 – 7.38 (m, 1H), 7.18 – 7.09 (m, 4H), 7.02 (d, <i>J</i> = 3.6 Hz, 1H), 6.93 (d, <i>J</i> = 8.4 Hz, 1H), 6.66 (d, <i>J</i> = 7.3 Hz, 1H), 5.15 (dd, <i>J</i> = 13.3, 5.2 Hz, 1H), 4.54 (s, 1H), 4.50 – 4.35 (m, 2H), 3.81 (s, 5H), 3.55 (d, <i>J</i> = 4.4 Hz, 2H), 3.22 (d, <i>J</i> = 30.1 Hz, 7H), 3.00 (d, <i>J</i> = 6.1 Hz, 5H), 2.63 (s, 1H), 2.31 (s, 1H), 1.98 (d, <i>J</i> = 21.8 Hz, 5H), 1.80 (dd, <i>J</i> = 11.9, 5.5 Hz, 6H), 1.64 (d, <i>J</i> = 12.9 Hz, 2H). LCMS [M+H] ⁺ = 944.4
UB-180770 (270)	 HCOOH ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 10.17 (s, 1H), 9.79 (s, 1H), 8.73 (d, <i>J</i> = 1.6 Hz, 1H), 8.46 – 8.19 (m, 4H), 7.96 – 7.76 (m, 8H), 7.58 – 7.45 (m, 3H), 7.34 (d, <i>J</i> = 2.1 Hz, 1H), 7.17 (t, <i>J</i> = 8.8 Hz, 2H), 5.14 (dd, <i>J</i> = 13.2, 5.1 Hz, 1H), 4.49 – 4.23 (m, 2H), 3.05 – 2.95 (m, 2H), 2.96 – 2.83 (m, 2H), 2.82 – 2.70 (m, 4H), 2.60 (tt, <i>J</i> = 6.8, 3.6 Hz, 5H), 2.40 – 2.31 (m, 2H), 2.27 – 2.18 (m, 1H), 2.08 – 1.94 (m, 2H), 1.67 – 1.54 (m, 2H), 1.52 – 1.38 (m, 5H), 1.37 – 1.20 (m, 5H), 1.04 – 0.90 (m, 2H). LCMS [M+H] ⁺ = 1006.5

UB-180771 (271)	
	^1H NMR (400 MHz, DMSO- d_6) δ 11.02 (s, 1H), 9.95 (s, 1H), 8.86 (s, 2H), 7.83 (dd, J = 7.2, 1.9 Hz, 1H), 7.79 – 7.67 (m, 2H), 7.61 – 7.41 (m, 3H), 7.28 – 7.03 (m, 6H), 6.84 (d, J = 7.5 Hz, 1H), 5.15 (dd, J = 13.3, 5.1 Hz, 1H), 4.55 (s, 1H), 4.39 (q, J = 17.6 Hz, 2H), 3.68 (t, J = 5.3 Hz, 3H), 3.55 (s, 4H), 3.49 (dt, J = 8.8, 3.7 Hz, 4H), 3.36 (t, J = 6.2 Hz, 2H), 3.18 (d, J = 5.8 Hz, 2H), 3.06 (t, J = 5.7 Hz, 2H), 2.97 (s, 2H), 2.90 (dd, J = 11.5, 4.6 Hz, 3H), 2.69 – 2.58 (m, 1H), 2.40 (t, J = 7.4 Hz, 2H), 1.99 – 1.87 (m, 2H), 1.77 (q, J = 13.6, 12.5 Hz, 4H), 1.70 – 1.56 (m, 6H), 1.35 (t, J = 7.5 Hz, 3H). LCMS $[\text{M}+\text{H}]^+$ = 991.5
UB-180772 (272)	
	^1H NMR (400 MHz, DMSO- d_6) δ 11.03 (s, H), 10.30 (s, 1H), 10.20 (s, 1H), 9.41 (s, 2H), 8.94 (t, J = 5.5 Hz, 1H), 8.74 (d, J = 9.2 Hz, 1H), 8.31 (t, J = 8.4 Hz, 1H), 8.13 – 7.77 (m, 6H), 7.67 – 7.37 (m, 3H), 7.36 – 7.02 (m, 3H), 5.17 – 5.08 (m, 1H), 4.45 – 4.29 (m, 2H), 3.75 – 3.44 (m, 15H), 3.43–3.27 (m, 3H), 2.87 (dt, J = 8.8, 4.5 Hz, 4H), 2.61 (d, J = 16.9 Hz, 1H), 2.47 – 2.14 (m, 2H), 2.14 – 1.88 (m, 2H), 1.67 (s, 4H). LCMS $[\text{M}+\text{H}]^+$ = 992.5
UB-180773 (273)	
	^1H NMR (400 MHz, DMSO- d_6) δ 11.03 (s, H), 10.30 (s, 1H), 10.20 (s, 1H), 8.73 (s, 1H), 8.66 (s, 2H), 8.33 (dd, J = 2.6, 7.0 Hz, 2H), 7.98 – 7.77 (m, 6H), 7.63 – 7.43 (m, 3H), 7.17 (s, 1H), 5.15 (dd, J = 13.3, 5.2 Hz, 1H), 4.38 (dd, J = 3.2, 1.8 Hz, 2H), 3.94 (s, 3H), 3.94 – 3.29 (m, 20H), 3.29 – 2.87 (m, 5H), 2.89 (s, 2H), 2.77 – 2.61 (m, 1H), 2.32 (dd, J = 7.5, 5.7 Hz, 1H), 1.95 (d, J = 7.9 Hz, 1H), 1.24 (ddd, J = 3.3, 1.9, 7.0 Hz, 2H). LCMS $[\text{M}+\text{H}]^+$ = 978.4
UB-180774 (274)	
	^1H NMR (400 MHz, DMSO- d_6) δ 11.03 (s, 1H), 10.25 (s, 1H), 9.29 (s, 2H), 8.29 (s, 1H), 7.96 (d, J = 10.8 Hz, 1H), 7.92 – 7.79 (m, 1H), 7.67 – 7.24 (m, 5H), 7.28 – 6.96 (m, 5H), 6.95 (s, 1H), 6.62 (d, J = 7.4 Hz, 1H), 5.16 (dd, J = 13.3, 5.1 Hz, 1H), 4.76 (t, J = 6.4 Hz, 2H), 4.67 – 4.40 (m, 5H), 3.43 (dd, J = 8.7, 2.7 Hz, 2H), 3.28 (s, 2H), 3.10 – 2.77 (m, 5H), 2.68 – 2.57 (m, 1H), 2.20 – 2.00 (m, 2H), 2.00 – 1.51 (m, 9H). LCMS $[\text{M}+\text{H}]^+$ = 990.2.
UB-180775 (275)	
	^1H NMR (400 MHz, DMSO- d_6) δ 11.04 (s, 1H), 10.75 (s, 1H), 10.41 (s, 1H), 10.18 (d, J = 4.7 Hz, 1H), 9.40 (s, 2H), 8.73 (s, 1H), 8.41 (d, J = 7.2 Hz, 1H), 8.30 (dd, J = 8.5, 2.8 Hz, 1H), 7.96 – 7.81 (m, 6H), 7.62 – 7.51 (m, 3H), 7.35 (d, J = 2.1 Hz, 1H), 7.17 (t, J = 8.8 Hz, 2H), 5.17 (dd, J = 13.3, 5.2 Hz, 4H), 4.55 – 4.34 (m, 3H), 4.06 (d, J = 30.6 Hz, 4H), 3.89 – 3.73 (m, 5H), 3.66 (d, J = 11.4 Hz, 1H), 3.58 (dd, J = 14.2, 8.7 Hz, 2H), 3.39 (s, 1H), 3.29 (d, J = 14.3 Hz, 4H), 3.19 – 3.07 (m, 2H), 2.99 – 2.87 (m, 1H), 2.64 (s, 1H), 2.30 (d, J = 4.1 Hz, 2H), 2.05 (q, J = 13.7 Hz, 5H). LCMS $[\text{M}+\text{H}]^+$ = 959.4

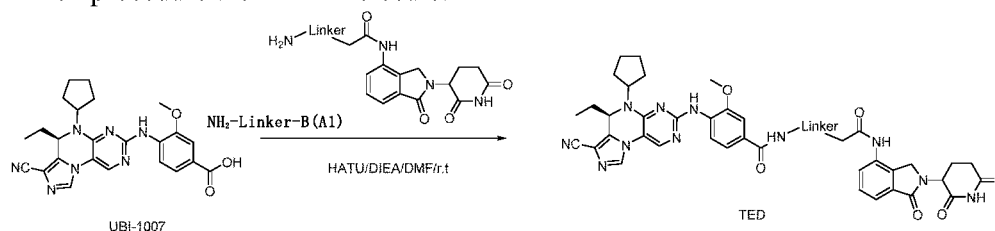
UB-180776 (276)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.01 (s, 1H), 10.77 (s, 1H), 10.16 (dd, <i>J</i> = 17.7, 7.1 Hz, 2H), 9.75 (s, 2H), 8.74 (s, 1H), 8.43 (d, <i>J</i> = 7.5 Hz, 1H), 8.30 (d, <i>J</i> = 8.5 Hz, 1H), 7.98 – 7.83 (m, 5H), 7.75 (dd, <i>J</i> = 7.9, 1.2 Hz, 1H), 7.62 – 7.48 (m, 3H), 7.34 (d, <i>J</i> = 2.1 Hz, 1H), 7.17 (t, <i>J</i> = 8.8 Hz, 2H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.61 – 4.32 (m, 3H), 4.22 (s, 2H), 4.07 (s, 1H), 3.87 (t, <i>J</i> = 4.8 Hz, 3H), 3.29 (s, 3H), 3.16 (d, <i>J</i> = 11.0 Hz, 2H), 2.92 (ddd, <i>J</i> = 17.9, 13.5, 5.4 Hz, 1H), 2.62 (d, <i>J</i> = 3.8 Hz, 1H), 2.42 – 2.33 (m, 1H), 2.05 (d, <i>J</i> = 25.8 Hz, 5H), 1.45 – 1.18 (m, 1H). LCMS [M+H]⁺ = 944.5
UB-180777 (277)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 10.25 (s, 1H), 10.17 (s, 1H), 8.72 (s, 1H), 8.40 (t, <i>J</i> = 5.6 Hz, 1H), 8.29 (d, <i>J</i> = 8.5 Hz, 1H), 7.96 – 7.81 (m, 5H), 7.63 – 7.44 (m, 3H), 7.34 (d, <i>J</i> = 2.2 Hz, 1H), 7.17 (t, <i>J</i> = 9.0 Hz, 2H), 5.32 (t, <i>J</i> = 4.8 Hz, 1H), 5.15 (dd, <i>J</i> = 13.3, 5.2 Hz, 1H), 4.47 – 4.30 (m, 3H), 3.66 (s, 2H), 3.54 (t, <i>J</i> = 6.2 Hz, 5H), 3.42 (d, <i>J</i> = 5.9 Hz, 3H), 3.16 (s, 2H), 3.04 (s, 2H), 2.81 (s, 2H), 2.67 (p, <i>J</i> = 1.9 Hz, 2H), 2.35 – 2.27 (m, 2H), 2.00 (dq, <i>J</i> = 13.3, 5.9 Hz, 3H). LCMS [M+H]⁺ = 920
UB-180778 (278)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 12.67 (s, 1H), 11.02 (s, 1H), 10.03 (s, 7H), 8.93 (s, 2H), 7.90 – 7.63 (m, 3H), 7.58 – 7.28 (m, 3H), 7.09 (dddd, <i>J</i> = 11.5, 10.4, 1.8, 9.0 Hz, 6H), 5.35 (s, 1H), 4.63 – 4.34 (m, 5H), 3.68 (dd, <i>J</i> = 13.5, 8.3 Hz, 2H), 3.59 – 3.09 (m, 10H), 3.05 – 2.85 (m, 5H), 2.64 (dd, <i>J</i> = 2.5, 9.3 Hz, 2H), 2.46 – 2.31 (m, 4H), 2.31 – 1.99 (m, 2H), 1.93–1.49 (m, 13H). LCMS [M+H]⁺ = 977.6
UB-180779 (279)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 10.04 (s, 1H), 8.85 (s, 1H), 7.84 (dd, <i>J</i> = 7.2, 1.5 Hz, 1H), 7.80 – 7.59 (m, 2H), 7.49 (ddd, <i>J</i> = 22.8, 11.4, 2.7 Hz, 3H), 7.29 – 7.05 (m, 4H), 6.96 (d, <i>J</i> = 8.2 Hz, 1H), 6.73 (d, <i>J</i> = 7.3 Hz, 1H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.66 – 4.29 (m, 5H), 3.76 – 3.42 (m, 12H), 2.89 (d, <i>J</i> = 5.2 Hz, 2H), 2.72 – 2.57 (m, 3H), 2.40 – 2.25 (m, 2H), 2.16 – 1.71 (m, 12H), 1.60 (t, <i>J</i> = 3.6 Hz, 2H). LCMS [M+H]⁺ = 963.6
UB-180780 (280)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 10.18 (s, 1H), 9.16 – 8.66 (m, 5H), 8.43 – 8.27 (m, 3H), 7.91 – 7.79 (m, 5H), 7.47 – 7.33 (m, 6H), 7.12 (dt, <i>J</i> = 42.1, 8.5 Hz, 2H), 4.99 – 3.95 (m, 6H), 3.85 – 3.36 (m, 15H), 3.23 – 2.91 (m, 4H), 2.70 (dd, <i>J</i> = 11.4, 7.3 Hz, 3H), 2.55 – 2.42 (m, 3H), 2.13 – 2.01 (m, 2H), 0.93 (d, <i>J</i> = 10.1 Hz, 9H). LCMS [M+H]⁺ = 1135.7

UB-180781 (281)	 HCl
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 10.19 (s, 1H), 9.01 (d, <i>J</i> = 6.0 Hz, 3H), 8.67 (dd, <i>J</i> = 2.2, 1.7 Hz, 3H), 8.36 (dd, <i>J</i> = 4.7, 7.0 Hz, 2H), 7.98 – 7.75 (m, 5H), 7.75 – 7.31 (m, 6H), 7.17 (t, <i>J</i> = 8.2 Hz, 2H), 4.53 (dd, <i>J</i> = 4.0, 8.1 Hz, 1H), 4.40 (dt, <i>J</i> = 6.8, 2.7 Hz, 3H), 4.25 (s, 3H), 4.28 – 3.76 (m, 5H), 3.75 – 3.19 (m, 13H), 2.45 (s, 3H), 2.14 – 2.02 (m, 2H), 0.95 (d, <i>J</i> = 9.1 Hz, 9H). LCMS [M+H] ⁺ = 1121.5
UB-180782 (282)	 HCl
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 9.07 (s, 1H), 8.97 (s, 2H), 8.76–8.71 (m, 1H), 8.46–8.31 (m, 1H), 7.75 (dd, <i>J</i> = 24.2, 16.5 Hz, 2H), 7.49 (dt, <i>J</i> = 16.3, 6.1 Hz, 5H), 7.38 – 7.09 (m, 5H), 6.85–6.77 (m, 1H), 4.55 – 4.22 (m, 2H), 4.16 – 3.90 (m, 10H), 4.16 – 3.38 (m, 10H), 3.09 – 3.04 (m, 2H), 3.09 – 2.72 (m, 3H), 3.59 – 2.14 (m, 3H), 2.31 – 1.86 (m, 4H), 1.72 (d, <i>J</i> = 12.7 Hz, 6H), 1.72 (d, <i>J</i> = 12.7 Hz, 6H), 1.06 – 0.72 (m, 9H). LCMS [M+H] ⁺ = 1120.6
UB-180783 (283)	 HCl
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 9.04 (s, 1H), 8.93 (s, 2H), 8.65 (dd, <i>J</i> = 10.5, 5.0 Hz, 2H), 7.90 – 7.64 (m, 2H), 7.54 (d, <i>J</i> = 4.0 Hz, 1H), 7.42 (dq, <i>J</i> = 16.5, 8.3 Hz, 4H), 7.33 – 7.03 (m, 5H), 6.86 (d, <i>J</i> = 7.4 Hz, 1H), 5.62 (s, 1H), 4.76 – 4.44 (m, 4H), 4.44 – 4.10 (m, 2H), 4.02 – 3.56 (m, 15H), 3.56 – 3.12 (m, 6H), 3.02 (s, 2H), 2.45 (s, 3H), 2.07 (dd, <i>J</i> = 13.2, 6.1 Hz, 3H), 1.97 – 1.25 (m, 9H), 0.95 (d, <i>J</i> = 9.6 Hz, 9H). LCMS [M+H] ⁺ = 1106.6
UB-180784 (284)	 HCl
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 10.19 (s, 1H), 9.01 (s, 2H), 8.94 (t, <i>J</i> = 5.7 Hz, 1H), 8.73 (s, 1H), 8.62 (t, <i>J</i> = 6.1 Hz, 1H), 8.30 (t, <i>J</i> = 8.3 Hz, 2H), 8.05 (s, 1H), 7.94 – 7.85 (m, 5H), 7.55 (tt, <i>J</i> = 8.3, 6.5 Hz, 1H), 7.43 – 7.33 (m, 5H), 7.17 (t, <i>J</i> = 8.8 Hz, 2H), 4.94 (s, 1H), 4.59 – 4.49 (m, 5H), 4.43 (q, <i>J</i> = 7.4 Hz, 3H), 4.35 (s, 1H), 4.21 (dd, <i>J</i> = 15.9, 5.4 Hz, 2H), 3.84 (s, 2H), 3.68 (dt, <i>J</i> = 10.5, 4.7 Hz, 4H), 3.07 (d, <i>J</i> = 6.6 Hz, 4H), 2.72 (t, <i>J</i> = 7.4 Hz, 2H), 2.44 (s, 3H), 2.05 (t, <i>J</i> = 10.4 Hz, 1H), 1.90 (ddd, <i>J</i> = 13.0, 8.6, 4.7 Hz, 1H), 0.94 (s, 9H). LCMS [M+H] ⁺ = 1128.6
UB-180785 (285)	 HCl
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 12.24 (s, 1H), 9.16 (d, <i>J</i> = 25.5 Hz, 2H), 9.02 (s, 1H), 8.63 (t, <i>J</i> = 6.1 Hz, 1H), 8.40 – 8.25 (m, 2H), 7.92 (s, 1H), 7.56 (t, <i>J</i> = 7.8 Hz, 1H), 7.48 (s, 1H), 7.40 (q, <i>J</i> = 8.4 Hz, 4H), 7.21 – 7.08 (m, 4H), 7.03 (d, <i>J</i> = 8.2 Hz, 1H), 6.67 (d, <i>J</i> = 7.4 Hz, 1H), 4.61 – 4.53 (m, 4H), 4.45 (t, <i>J</i> = 7.8 Hz, 3H), 4.28 (d, <i>J</i> = 5.6 Hz, 4H), 4.23 (d, <i>J</i> = 5.5 Hz, 2H), 4.19 (d, <i>J</i> = 5.4 Hz, 2H), 3.83 (t, <i>J</i> = 5.2 Hz, 2H), 3.68 (dt, <i>J</i> = 19.9, 6.8 Hz, 4H), 3.14 – 3.01 (m, 4H), 2.73 (t, <i>J</i> = 7.5 Hz, 2H), 2.15 – 1.58 (m, 11H), 0.94 (s, 9H). LCMS [M+H] ⁺ = 1113.6

	
UB-180791 (291)	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.45 (s, 1H), 9.21 (s, 2H), 9.00 (s, 1H), 8.60 (d, <i>J</i> = 6.0 Hz, 1H), 8.17 (s, 1H), 7.83 (d, <i>J</i> = 9.4 Hz, 1H), 7.68 (s, 1H), 7.58 (t, <i>J</i> = 7.8 Hz, 1H), 7.45 – 7.35 (m, 5H), 7.19 – 7.06 (m, 3H), 7.02 (s, 1H), 6.92 (d, <i>J</i> = 8.3 Hz, 1H), 6.68 (d, <i>J</i> = 7.4 Hz, 1H), 4.74 (t, <i>J</i> = 6.3 Hz, 2H), 4.56 (d, <i>J</i> = 27.7 Hz, 4H), 4.41 (dt, <i>J</i> = 22.4, 7.5 Hz, 3H), 4.24 (dd, <i>J</i> = 16.2, 5.6 Hz, 1H), 4.08 (s, 2H), 3.77 (s, 2H), 3.35 (d, <i>J</i> = 6.0 Hz, 6H), 3.18 (d, <i>J</i> = 5.7 Hz, 4H), 2.92 (s, 1H), 2.44 (s, 4H), 2.21 (s, 1H), 2.07 (t, <i>J</i> = 10.0 Hz, 1H), 1.77 (m, 10H), 0.95 (s, 9H). LCMS [M+H] ⁺ = 1231.7
UB-180792 (292)	
UB-180792 (292)	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.01 (s, 1H), 10.17 (s, 2H), 9.15 (s, 2H), 8.73 (s, 1H), 8.40 (t, <i>J</i> = 5.6 Hz, 1H), 8.30 (d, <i>J</i> = 8.5 Hz, 1H), 7.93 – 7.81 (m, 5H), 7.74 (dd, <i>J</i> = 7.9, 1.2 Hz, 1H), 7.61 – 7.47 (m, 3H), 7.34 (d, <i>J</i> = 2.1 Hz, 1H), 7.17 (t, <i>J</i> = 8.7 Hz, 2H), 5.14 (dd, <i>J</i> = 13.2, 5.1 Hz, 1H), 4.52 – 4.36 (m, 2H), 4.19 (s, 3H), 3.82 (t, <i>J</i> = 5.0 Hz, 2H), 3.75 (t, <i>J</i> = 5.2 Hz, 2H), 3.55 (s, 4H), 3.45 – 3.34 (m, 3H), 3.27 – 3.09 (m, 5H), 2.92 (td, <i>J</i> = 13.0, 12.4, 6.7 Hz, 1H), 2.60 (d, <i>J</i> = 17.4 Hz, 1H), 2.36 (d, <i>J</i> = 4.5 Hz, 1H), 2.06 – 1.96 (m, 1H), 1.35 – 1.24 (m, 4H). LCMS [M+H] ⁺ = 994.45
	
UB-180793 (293)	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.72 (s, 1H), 11.00 (s, 1H), 10.30 (s, 1H), 9.35 (s, 2H), 7.74 (dd, <i>J</i> = 7.1, 4.3 Hz, 2H), 7.65 – 7.47 (m, 3H), 7.42 (d, <i>J</i> = 3.8 Hz, 1H), 7.19 – 7.03 (m, 4H), 6.97 (d, <i>J</i> = 8.2 Hz, 1H), 6.72 (d, <i>J</i> = 7.3 Hz, 1H), 5.14 (dd, <i>J</i> = 13.4, 5.1 Hz, 1H), 4.44 (dd, <i>J</i> = 64.8, 17.4 Hz, 3H), 4.19 (s, 2H), 3.82 (s, 2H), 3.76 (d, <i>J</i> = 5.4 Hz, 2H), 3.48 (q, <i>J</i> = 2.2 Hz, 4H), 3.35 (t, <i>J</i> = 6.3 Hz, 2H), 3.26 – 3.09 (m, 7H), 2.98 – 2.85 (m, 3H), 2.62 (s, 1H), 2.38 – 2.30 (m, 1H), 2.10 – 1.58 (m, 10H). LCMS [M+H] ⁺ = 979.5
	
UB-180794 (294)	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.01 (s, 1H), 10.21 (s, 1H), 9.87 (s, 1H), 9.16 (s, 2H), 8.76 (d, <i>J</i> = 18.6 Hz, 2H), 8.29 (d, <i>J</i> = 8.5 Hz, 1H), 7.96 – 7.84 (m, 5H), 7.75 (dd, <i>J</i> = 7.6, 1.3 Hz, 1H), 7.59 – 7.47 (m, 3H), 7.34 (d, <i>J</i> = 2.2 Hz, 1H), 7.17 (t, <i>J</i> = 8.6 Hz, 2H), 5.14 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.39 (q, <i>J</i> = 17.5 Hz, 3H), 4.17 (s, 2H), 3.77 – 3.67 (m, 8H), 3.64 (s, 3H), 3.14 (t, <i>J</i> = 5.8 Hz, 4H), 2.91 (ddd, <i>J</i> = 18.1, 13.3, 5.4 Hz, 1H), 2.60 (d, <i>J</i> = 17.1 Hz, 1H), 2.39 – 2.31 (m, 1H), 2.08 – 1.95 (m, 1H), 1.29 (ddd, <i>J</i> = 16.8, 8.0, 5.7 Hz, 2H). LCMS [M+H] ⁺ = 950.5
	
UB-180795 (295)	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.57 (s, 1H), 11.01 (s, 1H), 9.87 (s, 1H), 9.23 (s, 2H), 8.13 (s, 1H), 7.76 (d, <i>J</i> = 7.5 Hz, 1H), 7.56 (ddt, <i>J</i> = 22.0, 14.9, 7.5 Hz, 3H), 7.41 (d, <i>J</i> = 3.6 Hz, 1H), 7.12 (td, <i>J</i> = 9.4, 7.4, 4.2 Hz, 3H), 7.03 (d, <i>J</i> = 3.6 Hz, 1H), 6.95 (d, <i>J</i> = 8.1 Hz, 1H), 6.69 (d, <i>J</i> = 7.3 Hz, 1H), 5.14 (dd, <i>J</i> = 13.4, 5.0 Hz, 1H), 4.53 (s, 1H), 4.39 (q, <i>J</i> = 17.5 Hz, 2H), 4.17 (s, 2H), 3.62 (d, <i>J</i> = 5.6 Hz, 10H), 3.38 (d, <i>J</i> = 5.9 Hz, 2H), 3.06 (s, 2H), 3.02 – 2.87 (m, 5H), 2.62 (s, 1H), 2.38 (s, 1H), 1.81 (td, <i>J</i> = 61.1, 59.4, 14.2 Hz, 10H). LCMS [M+H] ⁺ = 935.5

UB-180802 (302)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.86 (s, 1H), 11.05 (s, 1H), 10.67 (s, 1H), 9.12 (s, 2H), 7.87 (d, J = 6.1 Hz, 1H), 7.81 – 7.38 (m, 5H), 7.45 (s, 1H), 7.45 – 7.05 (m, 4H), 6.98 (d, J = 8.2 Hz, 1H), 6.75 (d, J = 7.4 Hz, 1H), 5.18 (dd, J = 13.3, 5.1 Hz, 1H), 4.55 (s, 1H), 4.46 (d, J = 17.6 Hz, 2H), 4.05 (s, 2H), 3.92 – 3.40 (m, 14H), 3.40 – 2.96 (m, 6H), 2.96 – 2.67 (m, 2H), 2.63 (d, J = 18.8 Hz, 2H), 2.29 (dd, J = 15.7, 11.0 Hz, 1H), 2.25 – 2.02 (m, 2H), 2.13 – 2.02 (m, 1H), 1.91-1.82 (m, 2H), 1.71-1.58 (m, 2H). LCMS $[\text{M}+\text{H}]^+ = 979.7$
UB-180803 (303)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.03 (s, 1H), 10.18 (d, J = 9.5 Hz, 1H), 9.99 (s, 1H), 9.33 (s, 2H), 8.88 (s, 1H), 8.52 (t, J = 5.6 Hz, 1H), 8.31 (d, J = 2.1 Hz, 1H), 7.91 – 7.79 (m, 6H), 7.57 – 7.49 (m, 3H), 7.34 (d, J = 2.1 Hz, 1H), 7.17 (s, 2H), 5.15 (dd, J = 13.3, 5.1 Hz, 1H), 4.44 – 4.29 (m, 2H), 3.72 (d, J = 4.8 Hz, 4H), 3.57 (d, J = 2.9 Hz, 4H), 3.47 (d, J = 5.1 Hz, 3H), 3.12 (qd, J = 7.4, 4.2 Hz, 10H), 2.96 – 2.87 (m, 2H), 2.78 (d, J = 4.5 Hz, 1H), 2.63 (t, J = 6.2 Hz, 3H), 2.30 (d, J = 4.4 Hz, 1H), 2.03 (dd, J = 9.9, 4.7 Hz, 1H). LCMS $[\text{M}+\text{H}]^+ = 1088.4$
UB-180804 (304)	 ^1H NMR (400 MHz, DMSO-d_6) δ 12.29 (s, 1H), 11.03 (s, 1H), 10.02 (s, 1H), 8.97 (s, 2H), 7.92 (d, J = 7.0 Hz, 1H), 7.83 (dd, J = 7.1, 1.9 Hz, 1H), 7.71 (td, J = 7.8, 3.7 Hz, 2H), 7.53 – 7.47 (m, 3H), 7.20 – 7.14 (m, 4H), 6.85 (dd, J = 15.7, 7.4 Hz, 2H), 5.15 (dd, J = 13.2, 5.1 Hz, 1H), 4.60 (s, 2H), 4.38 (d, J = 19.7 Hz, 3H), 3.73 – 3.67 (m, 4H), 3.58 (s, 2H), 3.52 (d, J = 1.8 Hz, 5H), 3.42 (t, J = 6.0 Hz, 2H), 3.24 (t, J = 5.6 Hz, 2H), 3.13 – 3.06 (m, 4H), 3.01 (s, 3H), 2.67 – 2.61 (m, 2H), 2.41 – 2.29 (m, 2H), 1.99 (dd, J = 20.4, 7.6 Hz, 2H), 1.89 – 1.81 (m, 4H), 1.66 (q, J = 12.6, 10.2 Hz, 4H). LCMS $[\text{M}+\text{H}]^+ = 993.4$
UB-180807 (307)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.03 (s, 1H), 10.29 (s, 1H), 10.18 (s, 1H), 9.49 – 8.56 (m, 2H), 8.79 – 8.56 (m, 1H), 8.40 (t, J = 5.4 Hz, 1H), 8.31 (t, J = 8.1 Hz, 1H), 8.37 – 7.77 (m, 6H), 7.62 – 7.33 (m, 3H), 7.35 (d, J = 2.1 Hz, 1H), 7.36 – 6.99 (m, 2H), 5.17-5.21 (m, 1H), 4.51-4.22 (m, 2H), 3.95 – 2.89 (m, 23H), 2.99 – 2.78 (m, 3H), 2.63 (t, J = 16.3 Hz, 2H), 2.40 – 2.14 (m, 1H), 2.04 (dd, J = 14.3, 9.0 Hz, 1H). LCMS $[\text{M}+\text{H}]^+ = 1008.4$
UB-180808 (308)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.03 (s, 1H), 10.29 (s, 1H), 10.18 (s, 1H), 8.79 – 8.56 (m, 2H), 7.92 – 7.33 (m, 6H), 7.35 (d, J = 2.1 Hz, 1H), 7.36 – 6.67 (m, 4H), 5.51 – 4.98 (m, 1H), 4.74 – 4.26 (m, 4H), 3.95 – 3.12 (m, 28H), 2.99 – 2.78 (m, 2H), 2.63 (t, J = 16.3 Hz, 2H), 2.40 – 2.14 (m, 2H), 2.04 (dd, J = 14.3, 9.0 Hz, 1H), 1.93 – 1.51 (m, 11H). LCMS $[\text{M}+\text{H}]^+ = 993.4$

(2.5) Preparation procedure 5 of TED molecule:

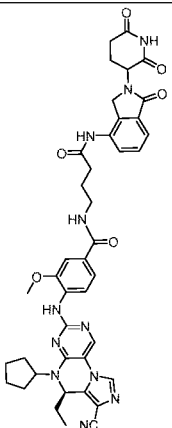
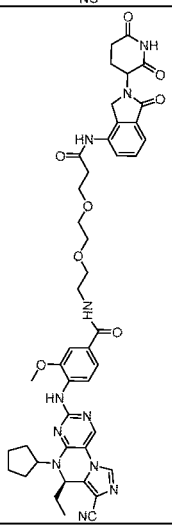
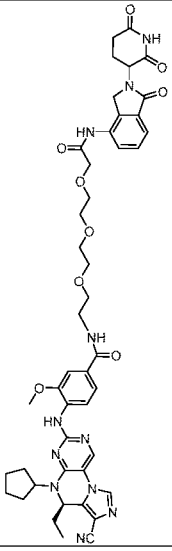


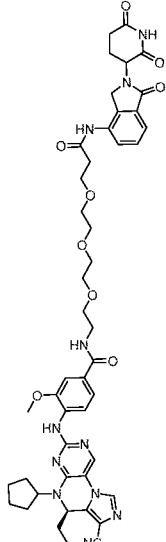
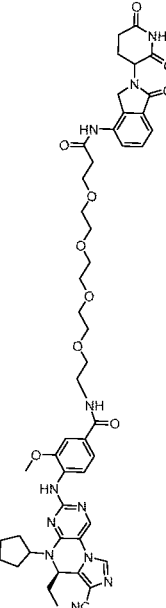
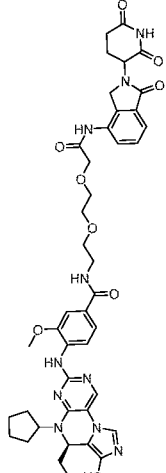
Compound UBI-1007 (20 mg, 0.0436 mmol) was weighed and dissolved in anhydrous DMF (1 mL), added NH₂-linker-B(A1) (0.0436 mmol), HATU (33 mg, 0.0872 mmol) and DIEA (0.072 mL, 0.436 mmol) under ice-water bath successively. After completion of addition, the material system was stirred and reacted at room temperature under the protection of nitrogen for 18 hours. After completion of reaction, the reaction solution was poured into 5 mL of water and extracted with ethyl acetate (5 mL*3) three times. The organic phases were combined and washed with saturated brine, dried over anhydrous Na₂SO₄ and concentrated by rotary evaporation under reduced pressure to obtain the crude product. After that, the crude product was isolated by thin-layer chromatography on silica gel plate with the polarity of the developing solvent (DCM/MeOH = 10/1) to prepare the target compound.

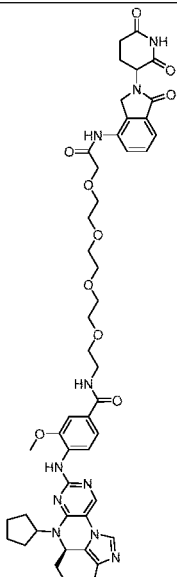
The conjugate of A-L1-B as shown in Table B5 was prepared through similar method.

Table B5

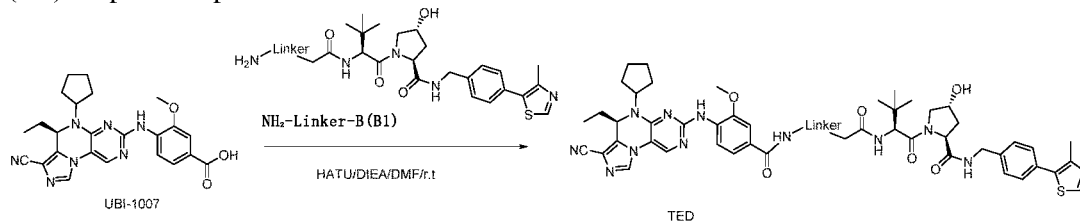
Examples	Compound structure	Analyze data
UB-180512		¹ H NMR (400 MHz, <i>d</i> ₆ -DMSO) δ 11.00 (s, 1H), 9.74 (s, 1H), 8.70 (s, 1H), 8.64 (s, 1H), 8.40 (t, <i>J</i> = 5.6 Hz, 1H), 8.23 (d, <i>J</i> = 8.2 Hz, 1H), 8.00 (s, 1H), 7.74 (d, <i>J</i> = 7.7 Hz, 1H), 7.60 – 7.41 (m, 4H), 5.24 – 5.05 (m, 2H), 4.60 – 4.22 (m, 3H), 4.10 (s, 2H), 3.91 (s, 3H), 3.71 – 3.37 (m, 4H), 2.97 – 2.83 (m, 1H), 2.45 – 2.13 (m, 2H), 2.06 – 1.87 (m, 11H), 0.78 (dt, <i>J</i> = 14.7, 7.3 Hz, 3H). LCMS [M+H] ⁺ = 830.3
UB-180518		¹ H NMR (400 MHz, <i>d</i> ₆ -DMSO) δ 10.99 (s, 1H), 9.79 (s, 1H), 8.70 (s, 1H), 8.64 (s, 1H), 8.55 (t, <i>J</i> = 5.4 Hz, 1H), 8.24 (d, <i>J</i> = 8.9 Hz, 1H), 7.99 (s, 1H), 7.71 (d, <i>J</i> = 7.7 Hz, 1H), 7.52 (ddd, <i>J</i> = 12.7, 8.6, 4.3 Hz, 3H), 5.24 – 5.05 (m, 2H), 4.58 – 4.28 (m, 3H), 4.18 (s, 2H), 3.87 (s, 3H), 3.70 (t, <i>J</i> = 5.7 Hz, 2H), 3.63 – 3.34 (m, 2H), 2.94-2.85 (m, 1H), 2.58-2.52 (m, 1H), 2.43 – 2.07 (m, 1H), 2.07 – 1.57 (m, 11H), 0.74 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [M+H] ⁺ = 802.8

UB-180537		¹ H NMR (400 MHz, <i>d</i> ₆ -DMSO) δ 11.01 (s, 1H), 9.86 (s, 1H), 8.70 (s, 1H), 8.64 (s, 1H), 8.45 (t, <i>J</i> = 5.7 Hz, 1H), 8.25 (d, <i>J</i> = 8.2 Hz, 1H), 8.00 (s, 1H), 7.81 (dd, <i>J</i> = 7.1, 1.7 Hz, 1H), 7.49 (dd, <i>J</i> = 15.4, 8.2 Hz, 4H), 5.27 – 5.07 (m, 2H), 4.55 – 4.28 (m, 3H), 3.92 (s, 3H), 3.30 (s, 2H), 3.12 – 2.86 (m, 1H), 2.41-2.49 (m, 4H), 2.07 – 1.76 (m, 13H), 0.74 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [<i>M</i> + <i>H</i>] ⁺ = 786.3
UB-180538		¹ H NMR (400 MHz, <i>d</i> ₆ -DMSO) δ 11.01 (s, 1H), 9.86 (s, 1H), 8.70 (s, 1H), 8.64 (s, 1H), 8.45 (t, <i>J</i> = 5.7 Hz, 1H), 8.25 (d, <i>J</i> = 8.2 Hz, 1H), 8.00 (s, 1H), 7.81 (dd, <i>J</i> = 7.1, 1.7 Hz, 1H), 7.49 (dd, <i>J</i> = 15.4, 8.2 Hz, 4H), 5.27 – 5.07 (m, 2H), 4.55 – 4.28 (m, 3H), 3.92 (s, 3H), 3.53 – 3.32 (m, 8H), 3.12 – 2.86 (m, 1H), 2.61-2.55 (m, 1H), 2.37 – 1.76 (m, 16H), 0.74 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [<i>M</i> + <i>H</i>] ⁺ = 860.9
UB-180545		¹ H NMR (400 MHz, <i>d</i> ₆ -DMSO) δ 11.01 (s, 1H), 9.70 (s, 1H), 8.70 (s, 1H), 8.64 (s, 1H), 8.45 (t, <i>J</i> = 5.6 Hz, 1H), 8.24 (d, <i>J</i> = 8.3 Hz, 1H), 7.99 (s, 1H), 7.77 – 7.70 (m, 1H), 7.52 (ddd, <i>J</i> = 11.4, 10.5, 4.6 Hz, 4H), 5.24 – 5.07 (m, 2H), 4.38 (dt, <i>J</i> = 27.7, 14.0 Hz, 3H), 4.13 (s, 2H), 3.91 (s, 3H), 3.75 – 3.52 (m, 9H), 3.42 – 3.36 (m, 2H), 3.02 – 2.79 (m, 1H), 2.10 – 1.85 (m, 11H), 1.35 – 1.14 (m, 2H), 0.74 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [<i>M</i> + <i>H</i>] ⁺ = 890.5

UB-180548		¹ H NMR (400 MHz, <i>d</i> ₆ -DMSO) δ 11.02 (s, 1H), 9.83 (s, 1H), 8.70 (s, 1H), 8.63 (s, 1H), 8.44 (t, <i>J</i> = 5.4 Hz, 1H), 8.25 (d, <i>J</i> = 8.3 Hz, 1H), 8.03 – 7.82 (m, 2H), 7.53 – 7.29 (m, 4H), 5.25 – 5.07 (m, 2H), 4.43-4.39 (m, 3H), 3.92 (s, 3H), 3.78 – 3.61 (m, 5H), 3.46 (dt, <i>J</i> = 11.0, 4.2 Hz, 10H), 3.41 (d, <i>J</i> = 9.6 Hz, 2H), 2.75 – 2.10 (m, 5H), 2.49 – 2.10 (m, 15H), 2.10 – 1.26 (m, 11H), 0.74 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 904.4
UB-180551		¹ H NMR (400 MHz, <i>d</i> ₆ -DMSO) δ 11.01 (s, 1H), 9.86 (s, 1H), 8.70 (s, 1H), 8.64 (s, 1H), 8.45 (t, <i>J</i> = 5.7 Hz, 1H), 8.25 (d, <i>J</i> = 8.2 Hz, 1H), 8.00 (s, 1H), 7.81 (dd, <i>J</i> = 7.1, 1.7 Hz, 1H), 7.49 (dd, <i>J</i> = 15.4, 8.2 Hz, 4H), 5.27 – 5.07 (m, 2H), 4.55 – 4.28 (m, 3H), 3.92 (s, 3H), 3.53 – 3.32 (m, 7H), 3.12 – 2.86 (m, 3H), 2.41-2.32 (m, 3H), 2.07 – 1.76 (m, 18H), 0.86 -0.79(m,3H), 0.74 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [M+H] ⁺ =948.7
UB-180557		¹ H NMR (400 MHz, <i>d</i> ₆ -DMSO) δ 11.00 (s, 1H), 9.70 (s, 1H), 8.70 (s, 1H), 8.64 (s, 1H), 8.45 (t, <i>J</i> = 5.4 Hz, 1H), 8.24 (d, <i>J</i> = 8.2 Hz, 1H), 7.99 (s, 1H), 7.73 (d, <i>J</i> = 7.6 Hz, 1H), 7.62 – 7.40 (m, 4H), 5.27 – 5.04 (m, 2H), 4.65 – 4.23 (m, 3H), 4.14 (s, 2H), 3.91 (s, 3H), 3.80 – 3.60 (m, 5H), 3.58 (d, <i>J</i> = 5.8 Hz, 2H), 3.04 – 2.76 (m, 1H), 2.71 – 2.57 (m, 1H), 2.07 – 1.59 (m, 11H), 0.74 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [M+H] ⁺ =846.4

UB-180560		¹ H NMR (400 MHz, <i>d</i> ₆ -DMSO) δ 11.00 (s, 1H), 9.68 (s, 1H), 8.70 (s, 1H), 8.63 (s, 1H), 8.44 (s, 1H), 8.25 (d, <i>J</i> = 8.3 Hz, 1H), 8.00 (s, 1H), 7.73 (d, <i>J</i> = 7.7 Hz, 1H), 7.52 (dt, <i>J</i> = 8.5, 5.3 Hz, 4H), 5.23 – 5.04 (m, 2H), 4.49 – 4.22 (m, 3H), 4.13 (s, 2H), 3.92 (s, 3H), 3.68 (dd, <i>J</i> = 5.8, 3.2 Hz, 2H), 3.65 – 3.42 (m, 13H), 3.03 – 2.72 (m, 1H), 2.64 (d, <i>J</i> = 22.0 Hz, 1H), 2.41 – 2.08 (m, 2H), 2.08 – 1.88 (m, 5H), 1.79 – 1.71 (m, 3H), 1.61 – 1.26 (m, 3H), 1.46 – 1.26 (m, 1H), 1.46 – 1.05 (m, 2H), 0.74 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 934.4
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(2.6) Preparation procedure 6 of TED molecule:

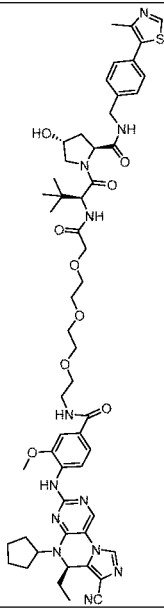
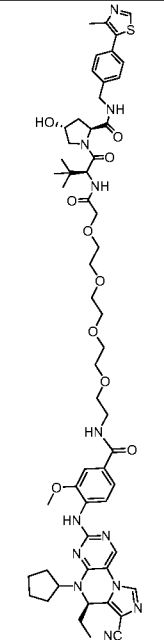
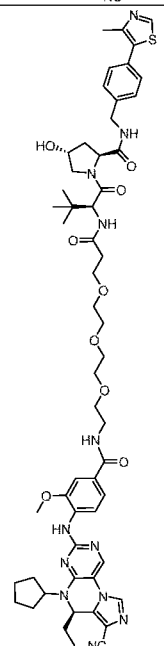


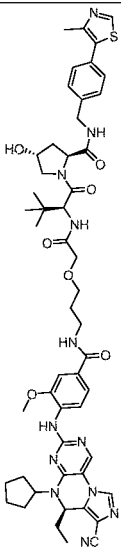
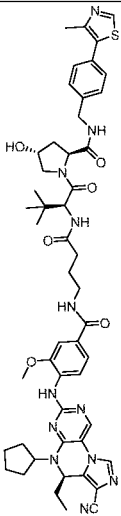
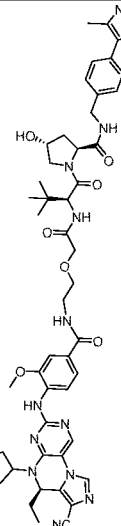
Compound UBI-1007 (20 mg, 0.0436 mmol) was weighed and dissolved in anhydrous DMF (1 mL), added NH₂-linker-B(B1) (0.0436 mmol), HATU(33 mg, 0.0872 mmol) and DIEA (0.072mL, 0.436 mmol) under ice-water bath successively. After completion of addition, the material system was stirred and reacted at room temperature under the protection of nitrogen for 18 hours. After completion of reaction, the reaction solution was poured into 5 mL of water and extracted with ethyl acetate (5 mL*3) three times. The organic phases were combined and washed with saturated brine, dried over anhydrous Na₂SO₄ and concentrated by rotary evaporation under reduced pressure to obtain the crude product. After that, the crude product was isolated by thin-layer chromatography on silica gel plate with the polarity of the developing solvent (DCM/MeOH = 10/1) to prepare the target compound.

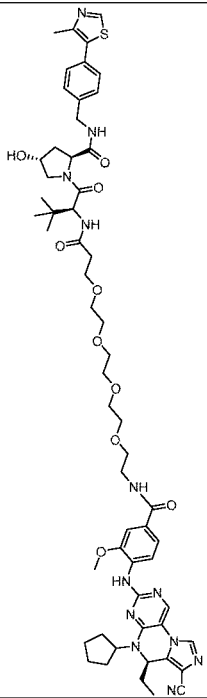
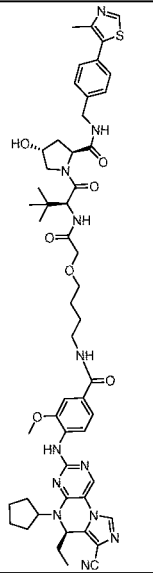
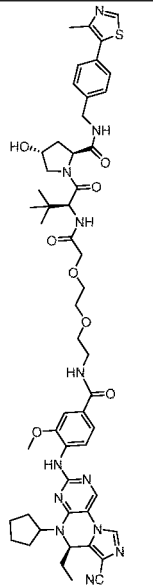
The conjugate of A-L1-B as shown in Table B6 was prepared through similar method.

Table B6

Examples	Compound structure	Analyze data
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UB-180519		¹ H NMR (400 MHz, <i>d6</i> -DMSO) δ 8.97 (d, <i>J</i> = 3.9 Hz, 1H), 8.70 (s, 1H), 8.66 – 8.53 (m, 2H), 8.46 (t, <i>J</i> = 5.4 Hz, 1H), 8.25 (d, <i>J</i> = 8.3 Hz, 1H), 7.99 (s, 1H), 7.52 (s, 1H), 7.52 – 7.36 (m, 5H), 5.25 – 5.10 (m, 2H), 4.56 (d, <i>J</i> = 9.6 Hz, 1H), 4.39-4.31 (m, 5H), 4.25 (dd, <i>J</i> = 15.8, 5.7 Hz, 1H), 3.70 – 3.48 (m, 13H), 3.24 (dd, <i>J</i> = 7.9, 8.5 Hz, 2H), 2.33 (s, 3H), 2.01 – 1.59 (m, 12H), 1.00 – 0.73 (m, 9H), 0.74 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 1061.3
UB-180525		¹ H NMR (400 MHz, <i>d6</i> -DMSO) δ 8.97 (s, 1H), 8.74 – 8.57 (m, 3H), 8.46 (s, 1H), 8.25 (d, <i>J</i> = 8.2 Hz, 1H), 8.01 (s, 1H), 7.58 – 7.35 (m, 2H), 7.35 – 7.32 (m, 5H), 5.27 – 4.84 (m, 2H), 4.62 – 4.38 (m, 3H), 4.38 – 4.17 (m, 3H), 4.06 – 4.00 (m, 3H), 3.94 (d, <i>J</i> = 15.3 Hz, 3H), 3.70 – 3.39 (m, 11H), 3.41 – 3.13 (m, 2H), 2.44 (s, 3H), 2.04-1.55 (m, 10H), 0.94 (s, 9H), 0.74 (t, <i>J</i> = 7.2 Hz, 3H). LCMS [M+H] ⁺ = 1105.6
UB-180529		¹ H NMR (400 MHz, <i>d6</i> -DMSO) δ 8.98 (s, 1H), 8.57 (t, <i>J</i> = 5.9 Hz, 1H), 8.46 – 8.34 (m, 2H), 8.02 – 7.83 (m, 2H), 7.60 (s, 1H), 7.55 – 7.34 (m, 6H), 5.14 (s, 1H), 4.55 – 4.18 (m, 9H), 3.87 (s, 3H), 3.71 – 3.38 (m, 18H), 2.43 (s, 3H), 2.02 – 1.55 (m, 10H), 0.93 (d, <i>J</i> = 5.7 Hz, 9H), 0.87 – 0.48 (m, 3H). LCMS [M+H] ⁺ = 1075.6

UB-180542	 The chemical structure of UB-180542 features a central purine core. At the 2-position, there is a 4-cyclopentyl-6-methyl-1H-pyrimidin-2-yl group. At the 6-position, there is a 4-methoxyphenyl group. The purine core is substituted with a 4-methoxyphenyl group at the 2-position and a 4-cyclopentyl-6-methyl-1H-pyrimidin-2-yl group at the 6-position. The purine core is also substituted with a 4-methoxyphenyl group at the 2-position and a 4-cyclopentyl-6-methyl-1H-pyrimidin-2-yl group at the 6-position.	^1H NMR (400 MHz, <i>d6</i>-DMSO) δ 8.70 (s, 1H), 8.66 – 8.49 (m, 2H), 8.40 (t, <i>J</i> = 5.6 Hz, 1H), 8.23 (d, <i>J</i> = 8.3 Hz, 1H), 8.00 (s, 1H), 7.58 – 6.95 (m, 7H), 5.27 – 5.07 (m, 2H), 4.57 (d, <i>J</i> = 9.6 Hz, 1H), 4.54 – 4.05 (m, 2H), 4.07 – 3.85 (m, 11H), 2.38 (s, 3H), 2.06 – 1.60 (m, 13H), 0.94 (d, <i>J</i> = 6.9 Hz, 9H), 0.74 (t, <i>J</i> = 7.3 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 987.6$
UB-180543	 The chemical structure of UB-180543 is similar to UB-180542, but with a different substitution pattern on the purine core. It features a 4-cyclopentyl-6-methyl-1H-pyrimidin-2-yl group at the 2-position and a 4-methoxyphenyl group at the 6-position. The purine core is also substituted with a 4-methoxyphenyl group at the 2-position and a 4-cyclopentyl-6-methyl-1H-pyrimidin-2-yl group at the 6-position.	^1H NMR (400 MHz, <i>d6</i>-DMSO) δ 8.96 (d, <i>J</i> = 12.3 Hz, 1H), 8.70 (s, 1H), 8.64 (s, 1H), 8.57 (t, <i>J</i> = 6.0 Hz, 1H), 8.41 (t, <i>J</i> = 5.2 Hz, 1H), 8.24 (d, <i>J</i> = 8.3 Hz, 1H), 8.03 – 7.87 (m, 2H), 7.50 (dd, <i>J</i> = 10.8, 2.3 Hz, 2H), 7.40 (q, <i>J</i> = 8.4 Hz, 4H), 5.23 – 5.10 (m, 2H), 4.59 – 4.27 (m, 5H), 4.21 (dd, <i>J</i> = 15.8, 5.4 Hz, 1H), 3.92 (d, <i>J</i> = 5.5 Hz, 3H), 3.78 – 3.41 (m, 2H), 3.29 – 3.10 (m, 3H), 2.36 (s, 3H), 2.37 – 1.63 (m, 15H), 0.94 (d, <i>J</i> = 8.3 Hz, 9H), 0.74 (t, <i>J</i> = 7.4 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 957.5$
UB-180544	 The chemical structure of UB-180544 is similar to UB-180542, but with a different substitution pattern on the purine core. It features a 4-cyclopentyl-6-methyl-1H-pyrimidin-2-yl group at the 2-position and a 4-methoxyphenyl group at the 6-position. The purine core is also substituted with a 4-methoxyphenyl group at the 2-position and a 4-cyclopentyl-6-methyl-1H-pyrimidin-2-yl group at the 6-position.	^1H NMR (400 MHz, <i>d6</i>-DMSO) δ 8.96 (s, 1H), 8.70 (s, 1H), 8.68 – 8.39 (m, 4H), 8.25 (d, <i>J</i> = 8.4 Hz, 1H), 7.99 (s, 1H), 7.60 – 7.51 (m, 3H), 7.51 – 7.36 (m, 4H), 5.17 (dd, <i>J</i> = 7.9, 3.3 Hz, 2H), 4.56 (d, <i>J</i> = 9.6 Hz, 1H), 4.50 – 4.21 (m, 5H), 4.09 – 3.89 (m, 5H), 3.74 – 3.39 (m, 4H), 3.13 – 3.06 (m, 1H), 2.43 (s, 3H), 1.97 – 1.47 (m, 12H), 0.91 (s, 9H), 0.73 (d, <i>J</i> = 7.4 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 973.6$

UB-180552		¹ H NMR (400 MHz, <i>d6</i> -DMSO) δ 11.01 (s, 1H), 9.70 (s, 1H), 8.70 (s, 1H), 8.64 (s, 1H), 8.45 (t, <i>J</i> = 5.6 Hz, 1H), 8.24 (d, <i>J</i> = 8.3 Hz, 1H), 7.99 (s, 1H), 7.77 – 7.70 (m, 1H), 7.52 (ddd, <i>J</i> = 11.4, 10.5, 4.6 Hz, 4H), 5.24 – 5.07 (m, 2H), 4.28–4.59 (m, 6H), 3.91 (s, 3H), 3.75 – 3.52 (m, 20H), 2.41 – 2.10 (m, 4H), 2.10 – 1.55 (m, 12H), 0.84 (s, 9H), 0.74 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [<i>M</i> + <i>H</i>] ⁺ = 1119.8
UB-180554		¹ H NMR (400 MHz, <i>d6</i> -DMSO) δ 8.70 (s, 1H), 8.66 – 8.49 (m, 2H), 8.40 (t, <i>J</i> = 5.6 Hz, 1H), 8.23 (d, <i>J</i> = 8.3 Hz, 1H), 8.00 (s, 1H), 7.58 – 6.95 (m, 6H), 5.27 – 5.07 (m, 2H), 4.28–4.59 (m, 6H), 3.96 – 3.45 (m, 14H), 3.36 (s, 6H), 2.46 – 2.26 (m, 3H), 2.26 – 1.60 (m, 16H), 0.94 (d, <i>J</i> = 6.9 Hz, 9H), 0.74 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [<i>M</i> + <i>H</i>] ⁺ = 1119.8
UB-180563		¹ H NMR (400 MHz, <i>d6</i> -DMSO) δ 8.97 (d, <i>J</i> = 9.0 Hz, 1H), 8.73 (s, 1H), 8.70 – 8.53 (m, 3H), 8.24 (d, <i>J</i> = 8.4 Hz, 1H), 7.99 (s, 1H), 7.59 – 7.33 (m, 7H), 5.27 – 5.10 (m, 2H), 4.57 (d, <i>J</i> = 9.6 Hz, 1H), 4.51 – 4.30 (m, 4H), 4.25 (dd, <i>J</i> = 15.8, 5.7 Hz, 1H), 3.98 (s, 2H), 3.88 (s, 3H), 3.43 – 2.65 (m, 9H), 2.44 (d, <i>J</i> = 5.2 Hz, 3H), 2.06–1.55 (m, 12H), , 0.93 (d, <i>J</i> = 5.7 Hz, 9H), 0.87 – 0.48 (m, 3H). LCMS [<i>M</i> + <i>H</i>] ⁺ = 1017.2

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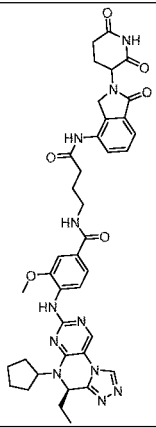
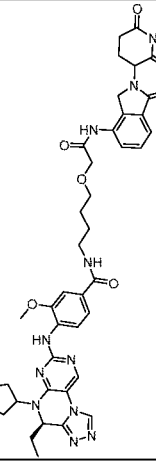
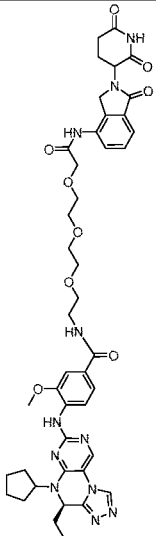


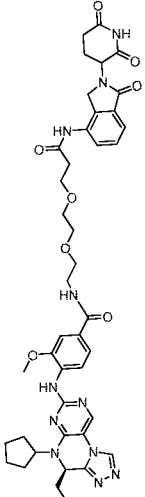
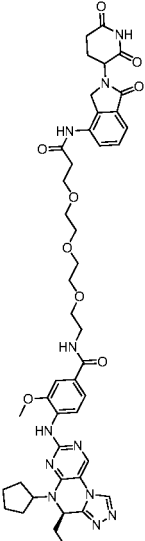
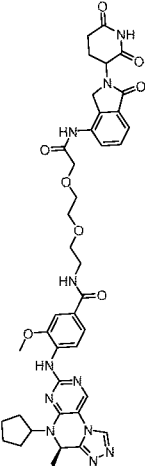
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UB-180521		¹ H NMR (400 MHz, <i>d6</i> -DMSO) δ 11.01 (s, 1H), 9.86 (s, 1H), 9.25 (s, 1H), 8.59 (s, 1H), 8.45 (s, 1H), 8.28 (d, <i>J</i> = 8.4 Hz, 1H), 7.94 (s, 1H), 7.81 (d, <i>J</i> = 7.0 Hz, 1H), 7.49 (dd, <i>J</i> = 15.3, 8.1 Hz, 4H), 5.29 – 4.98 (m, 2H), 4.72 – 3.90 (m, 1H), 3.90 – 3.85 (m, 3H), 2.98 – 2.86 (m, 2H), 2.75 (s, 1H), 2.60 (d, <i>J</i> = 17.4 Hz, 2H), 2.57 – 2.31 (m, 3H), 2.05-1.54 (m, 16H), 0.71 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [M+H] ⁺ = 762.3
UB-180524		¹ H NMR (400 MHz, <i>d6</i> -DMSO) δ 11.00 (s, 1H), 9.75 (s, 1H), 9.25 (s, 1H), 8.58 (s, 1H), 8.40 (t, <i>J</i> = 5.6 Hz, 1H), 8.26 (d, <i>J</i> = 8.2 Hz, 1H), 7.94 (s, 1H), 7.78 – 7.71 (m, 1H), 7.52 (qd, <i>J</i> = 7.8, 4.8 Hz, 4H), 5.17-5.08 (m, 2H), 4.69–4.29 (m, 3H), 4.10 (s, 2H), 3.95 (s, 3H), 3.57 (d, <i>J</i> = 5.5 Hz, 2H), 3.19 – 2.85 (m, 1H), 2.67 (d, <i>J</i> = 1.8 Hz, 1H), 2.06– 1.59 (m, 16H), 0.71 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 807.2.
UB-180546		¹ H NMR (400 MHz, <i>d6</i> -DMSO) δ 11.01 (s, 1H), 9.68 (s, 1H), 9.25 (s, 1H), 8.59 (s, 1H), 8.58 (s, 1H), 8.44 (t, <i>J</i> = 5.6 Hz, 1H), 8.28 (d, <i>J</i> = 8.3 Hz, 1H), 7.94 (s, 1H), 7.61 – 7.43 (m, 4H), 5.17-5.11 (m, 2H), 4.50 – 4.29 (m, 3H), 4.29 (s, 2H), 3.97 – 3.48 (m, 13H), 2.41 – 2.27 (m, 2H), 2.07 – 1.56 (m, 12H), 0.70 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 866.4

UB-180547		^1H NMR (400 MHz, <i>d6</i>-DMSO) δ 8.97 (d, J = 9.0 Hz, 1H), 8.73 (s, 1H), 8.70 – 8.53 (m, 3H), 8.24 (d, J = 8.4 Hz, 1H), 7.99 (s, 1H), 7.59 – 7.33 (m, 4H), 5.27 – 5.10 (m, 2H), 4.57 (d, J = 9.6 Hz, 1H), 4.51 – 4.30 (m, 3H), 4.25 (dd, J = 15.8, 5.7 Hz, 1H), 3.98 (s, 3H), 3.95 – 3.28 (m, 8H), 2.83 – 2.65 (m, 5H), 2.44 (d, J = 5.2 Hz, 2H), 2.06-1.65 (m, 10H), 0.73 (t, J = 7.2 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 836.5$
UB-180549		^1H NMR (400 MHz, <i>d6</i>-DMSO) δ 11.01 (s, 1H), 9.68 (s, 1H), 9.25 (s, 1H), 8.59 (s, 1H), 8.58 (s, 1H), 8.44 (t, J = 5.6 Hz, 1H), 8.28 (d, J = 8.3 Hz, 1H), 7.94 (s, 1H), 7.57 – 7.33 (m, 4H), 5.17-5.11 (m, 2H), 4.52 – 4.25 (m, 3H), 3.92 (s, 3H), 3.70 (t, J = 6.3 Hz, 3H), 3.61 – 3.35 (m, 14H), 2.53 – 2.44 (m, 1H), 2.06-1.60 (m, 11H), 0.92 (s, 9H), 0.70 (t, J = 7.4 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 880.4$
UB-180556		^1H NMR (400 MHz, <i>d6</i>-DMSO) δ 11.01 (s, 1H), 9.68 (s, 1H), 9.25 (s, 1H), 8.58 (s, 1H), 8.44 (t, J = 5.6 Hz, 1H), 8.28 (d, J = 8.3 Hz, 1H), 7.95 (s, 1H), 7.74 (d, J = 6.9 Hz, 1H), 7.61 – 7.41 (m, 4H), 5.17-5.11 (m, 2H), 4.59 – 4.33 (m, 3H), 4.33 – 4.09 (m, 2H), 3.92 (s, 3H), 3.74 – 3.55 (m, 5H), 2.35 – 1.94 (m, 1H), 2.35 – 1.58 (m, 14H), 0.70 (t, J = 7.3 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 822.6$

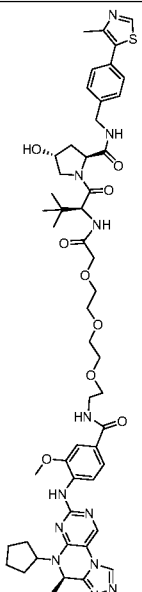
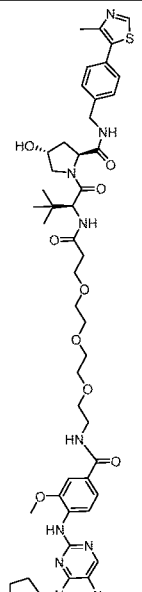
UB-180561		^1H NMR (400 MHz, <i>d6-DMSO</i>) δ 11.01 (s, 1H), 9.68 (s, 1H), 9.25 (s, 1H), 8.58 (s, 1H), 8.44 (t, $J = 5.6$ Hz, 1H), 8.28 (d, $J = 8.3$ Hz, 1H), 7.94 (s, 1H), 7.57 – 7.33 (m, 4H), 5.17–5.11 (m, 2H), 4.52 – 4.25 (m, 3H), 3.92 (s, 3H), 3.77 – 3.45 (m, 15H), 3.45 – 3.21 (m, 3H), 3.01 – 2.78 (m, 1H), 2.07–1.46 (m, 9H), 0.70 (t, $J = 7.3$ Hz, 3H). LCMS $[M+H]^+ = 910.6$
UB-180562		^1H NMR (400 MHz, DMSO) δ 11.02 (s, 1H), 9.84 (s, 1H), 9.25 (s, 1H), 8.59 (s, 1H), 8.45 (t, $J = 5.5$ Hz, 1H), 8.28 (d, $J = 8.3$ Hz, 1H), 7.95 (s, 1H), 7.82 (dd, $J = 7.1, 1.7$ Hz, 1H), 7.56 – 7.31 (m, 4H), 5.18–5.11 (m, 2H), 4.55 – 4.25 (m, 3H), 3.92 (s, 3H), 3.70 (t, $J = 6.3$ Hz, 2H), 3.64 – 2.89 (m, 16H), 3.13 (dd, $J = 7.3, 4.2$ Hz, 1H), 2.92 – 2.85 (m, 1H), 2.60 (t, $J = 6.3$ Hz, 3H), 2.45 – 2.39 (m, 1H), 2.19 – 1.55 (m, 11H), 0.70 (t, $J = 7.4$ Hz, 3H). LCMS $[M+H]^+ = 924.7$.

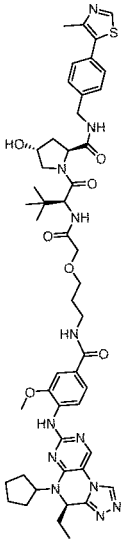
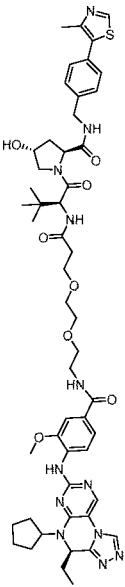
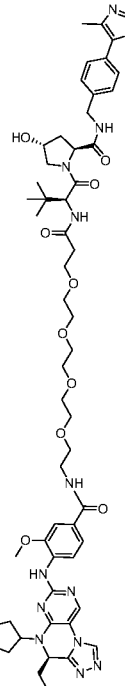
(2.8) Preparation procedure 8 of TED molecule:

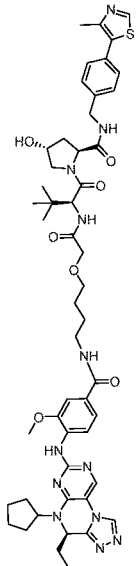
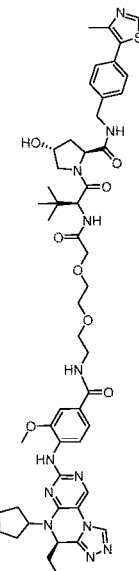
Compound UBI-1008 (20 mg, 0.0436 mmol) was weighed and dissolved in anhydrous DMF (1 mL), added NH₂-linker-B(B1) (0.0436 mmol), HATU(33 mg, 0.0872 mmol) and DIEA (0.072mL, 0.436 mmol) under ice-water bath successively. After completion of addition, the material system was stirred and reacted at room temperature under the protection of nitrogen for 18 hours. After completion of reaction, the reaction solution was poured into 5 mL of water and extracted with ethyl acetate (5 mL*3) three times. The organic phases were combined and washed with saturated brine, dried over anhydrous Na₂SO₄ and concentrated by rotary evaporation under reduced pressure to obtain the crude product. After that, the crude product was isolated by thin-layer chromatography on silica gel plate with the polarity of the developing solvent (DCM/MeOH = 10/1) to prepare the target compound.

The conjugate of A-L1-B as shown in Table B8 was prepared through similar method.

Table B8

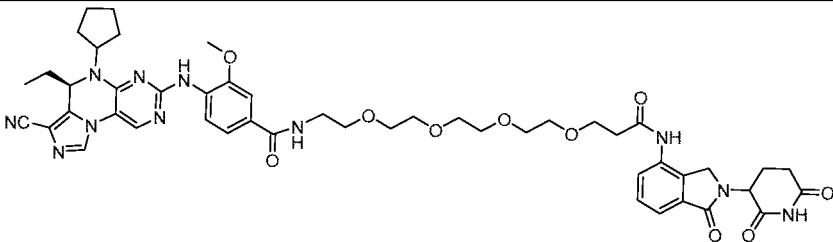
Examples	Compound structure	Analyze data
UB-180516		^1H NMR (400 MHz, <i>d6</i> -DMSO) δ 9.25 (s, 1H), 8.97 (s, 1H), 8.74 – 8.57 (m, 2H), 8.46 (s, 1H), 8.25 (d, <i>J</i> = 8.2 Hz, 1H), 8.01 (s, 1H), 7.58 – 7.35 (m, 5H), 7.35 – 7.32 (m, 2H), 5.27 – 5.14 (m, 2H), 4.62 – 4.17 (m, 6H), 3.94-3.89 (m, 5H), 3.81 – 3.43 (m, 11H), 2.52 (s, 3H), 2.04 – 1.55 (m, 12H), 0.94 (s, 9H), 0.74 (t, <i>J</i> = 7.2 Hz, 3H). LCMS [<i>M</i> + <i>H</i>] ⁺ = 1037.5
UB-180522		^1H NMR (400 MHz, <i>d6</i> -DMSO) δ 9.25 (s, 1H), 8.98 (s, 1H), 8.61–8.46 (m, 2H), 8.45 (s, 1H), 8.28 (d, <i>J</i> = 8.2 Hz, 1H), 7.98 – 7.82 (m, 2H), 7.51 (d, <i>J</i> = 10.1 Hz, 2H), 7.40 (q, <i>J</i> = 8.3 Hz, 4H), 5.28 – 5.16 (m, 1H), 5.13 (d, <i>J</i> = 3.5 Hz, 1H), 4.52 (t, <i>J</i> = 19.4 Hz, 2H), 4.42 (dd, <i>J</i> = 9.4, 6.2 Hz, 3H), 4.29 (d, <i>J</i> = 42.2 Hz, 2H), 4.24 – 4.01 (m, 2H), 3.92 (s, 3H), 3.67 – 2.65 (m, 9H), 2.48 (s, 3H), 2.02-1.57 (m, 11H), 0.92 (s, 9H), 0.70 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [<i>M</i> + <i>H</i>] ⁺ = 1051.6.

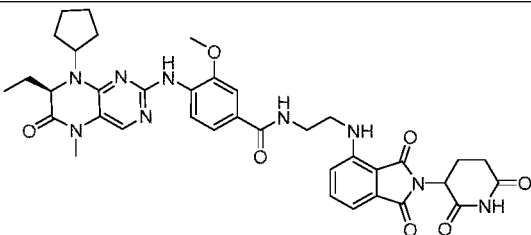
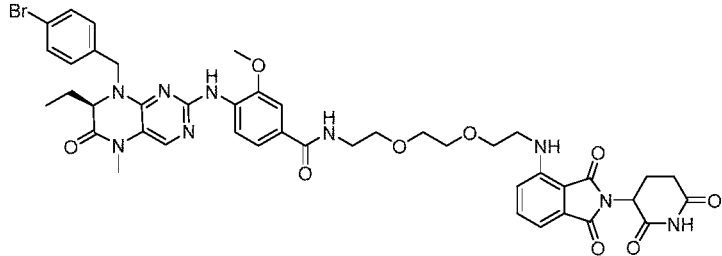
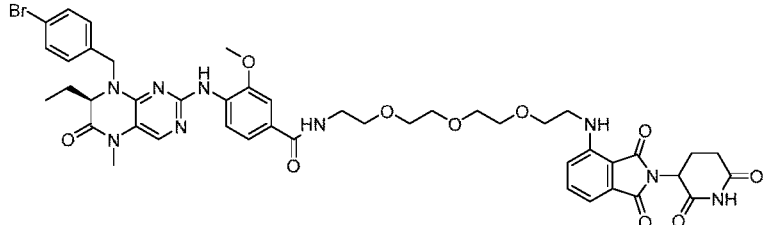
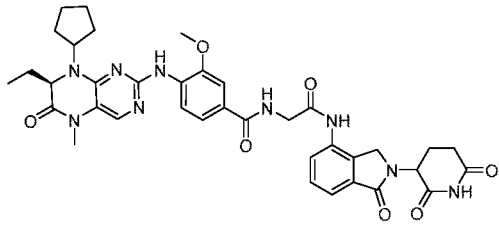
UB-180541	 The chemical structure of UB-180541 features a 1,2,4-triazole core substituted with a cyclopentyl group and a 4-methoxyphenyl group. This core is linked via an amide bond to a 4-methoxyphenyl ring, which is further connected through a long, flexible ether chain to a chiral center. This chiral center is part of a complex side chain that includes a hydroxyl group, a carbamate, and a 4-methyl-1,3,4-thiadiazole moiety.	^1H NMR (400 MHz, <i>d6</i>-DMSO) δ 9.25 (s, 1H), 8.96 (d, J = 6.3 Hz, 1H), 8.59 (d, J = 5.5 Hz, 2H), 8.44 (s, 1H), 8.27 (d, J = 8.3 Hz, 1H), 7.94 (s, 1H), 7.56 – 7.37 (m, 7H), 5.26 – 5.12 (m, 2H), 4.57 (d, J = 9.6 Hz, 1H), 4.51 – 4.27 (m, 5H), 4.24 (d, J = 5.5 Hz, 1H), 3.94–3.92 (m, 5H), 3.67 – 3.52 (m, 4H), 2.09 – 1.60 (m, 13H), 1.34 – 1.11 (m, 2H), 0.93 (d, J = 5.9 Hz, 9H), 0.70 (t, J = 7.3 Hz, 3H). LCMS $[\text{M}+\text{H}]^+$ = 963.4
UB-180550	 The chemical structure of UB-180550 is similar to UB-180541 but features a different ether chain linker, resulting in a distinct mass and NMR profile.	^1H NMR (400 MHz, <i>d6</i>-DMSO) δ 9.25 (s, 1H), 8.85 (d, J = 10.9 Hz, 1H), 8.61 – 8.51 (m, 2H), 8.46 (t, J = 5.5 Hz, 1H), 8.27 (d, J = 8.3 Hz, 1H), 8.11 – 7.86 (m, 2H), 7.52 (d, J = 2.7 Hz, 2H), 7.43 (dt, J = 15.7, 5.1 Hz, 4H), 5.20–5.15 (m, 1H), 5.14 (s, 1H), 4.55 (d, J = 9.4 Hz, 1H), 4.50 – 4.31 (m, 4H), 4.31 – 4.13 (m, 1H), 3.92 (s, 3H), 3.85 – 3.41 (m, 13H), 3.14 (td, J = 7.4, 4.3 Hz, 2H), 2.47 (s, 3H), 2.09 (s, 4H), 2.09 – 1.51 (m, 11H), 0.92 (d, J = 7.1 Hz, 9H), 0.70 (t, J = 7.4 Hz, 3H). LCMS $[\text{M}+\text{H}]^+$ = 1007.4
UB-180553	 The chemical structure of UB-180553 is similar to the others but has a different ether chain linker, leading to a unique mass and NMR profile.	^1H NMR (400 MHz, <i>d6</i>-DMSO) δ 9.25 (s, 1H), 8.98 (s, 1H), 8.61 – 8.51 (m, 2H), 8.46 (t, J = 5.6 Hz, 1H), 8.28 (d, J = 8.3 Hz, 1H), 7.98 – 7.84 (m, 2H), 7.51 (dd, J = 10.2, 1.8 Hz, 2H), 7.46 – 7.32 (m, 4H), 5.21 (dd, J = 7.8, 3.3 Hz, 1H), 5.13 (d, J = 3.5 Hz, 1H), 4.60 – 4.32 (m, 5H), 4.21 (dd, J = 15.8, 5.3 Hz, 1H), 3.93 (s, 3H), 3.80 – 3.46 (m, 20H), 2.42 – 2.19 (m, 4H), 2.02–1.60 (m, 12H), 0.92 (d, J = 8.1 Hz, 9H), 0.70 (t, J = 7.4 Hz, 3H). LCMS $[\text{M}+\text{H}]^+$ = 1095.8

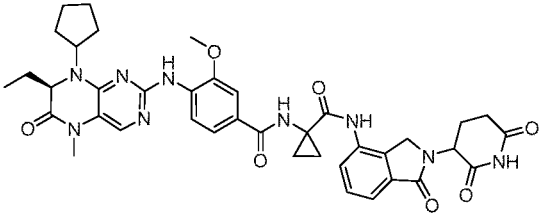
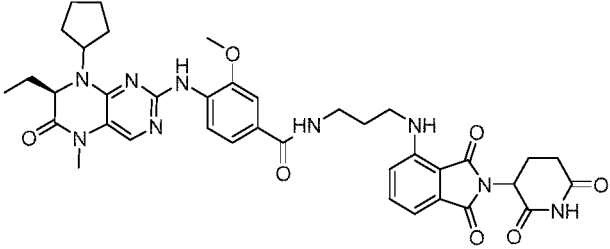
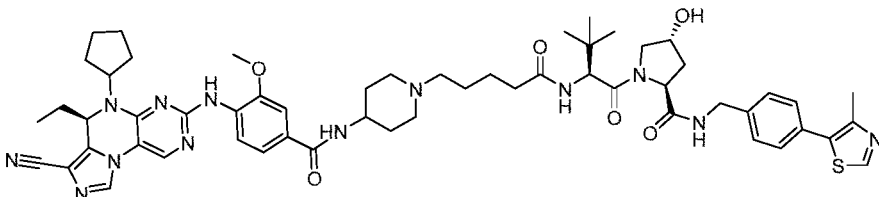
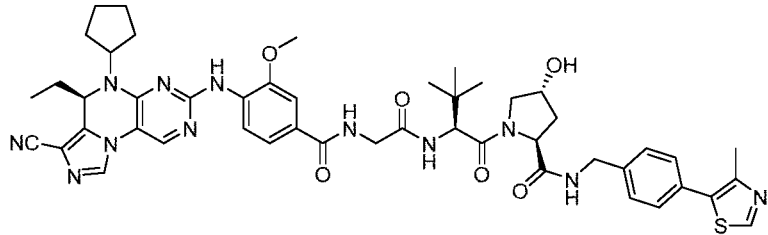
UB-180555		¹ H NMR (400 MHz, <i>d6</i> -DMSO) δ 9.25 (s, 1H), 8.98 (s, 1H), 8.61 – 8.51 (m, 2H), 8.43 – 8.30 (m, 1H), 8.26 (d, <i>J</i> = 8.2 Hz, 1H), 7.94 (s, 1H), 7.57 – 7.38 (m, 7H), 5.29 – 5.12 (m, 4H), 4.58 (t, <i>J</i> = 14.8 Hz, 1H), 4.53 – 4.25 (m, 5H), 3.98 – 3.85 (m, 5H), 3.71 – 3.57 (m, 8H), 3.17 (s, 1H), 2.12–1.76 (m, 12H), 0.93 (d, <i>J</i> = 7.1 Hz, 9H), 0.70 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [<i>M</i> + <i>H</i>] ⁺ = 977.8
UB-180564		¹ H NMR (400 MHz, <i>d6</i> -DMSO) δ 9.25 (s, 1H), 8.97 (s, 1H), 8.65 (s, 2H), 8.44 (s, 1H), 8.27 (d, <i>J</i> = 8.3 Hz, 1H), 7.94 (s, 1H), 7.46 – 7.13 (m, 7H), 5.28 – 4.59 (m, 2H), 4.57 (d, <i>J</i> = 9.6 Hz, 1H), 4.46–4.39 (m, 4H), 4.25 (dd, <i>J</i> = 15.7, 5.6 Hz, 1H), 3.98 (s, 2H), 3.91 (s, 3H), 3.86 – 3.69 (m, 10H), 2.93 (s, 3H), 2.01–1.59 (m, 11H), 0.93 (d, <i>J</i> = 6.5 Hz, 9H), 0.70 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [<i>M</i> + <i>H</i>] ⁺ = 994.5

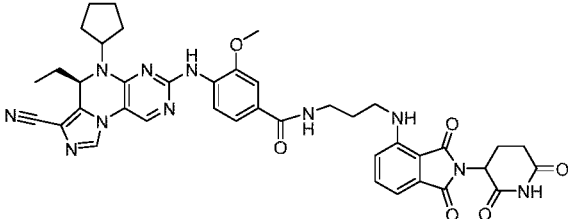
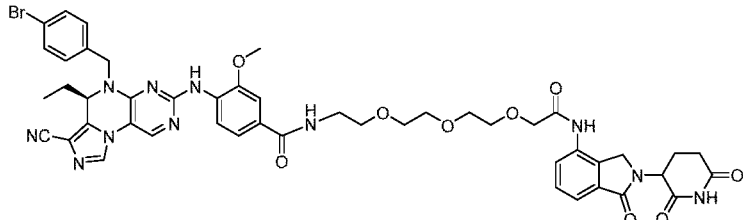
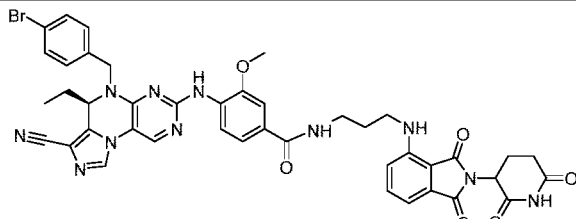
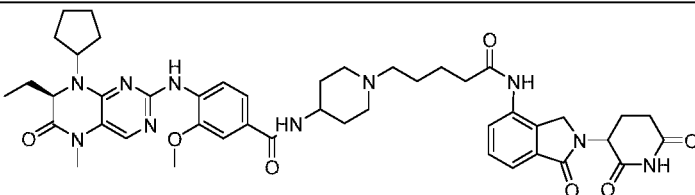
Compounds in Table B10 were prepared by preparation procedures similar to (2.5), (2.6), (2.7) or (2.8)

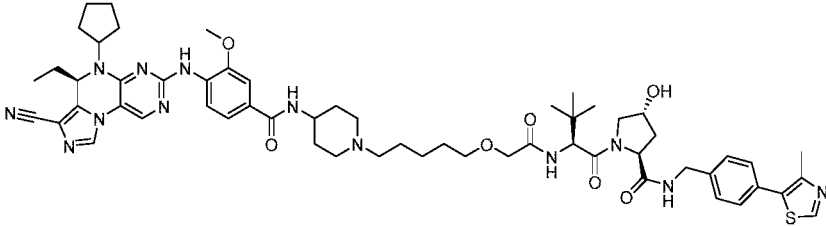
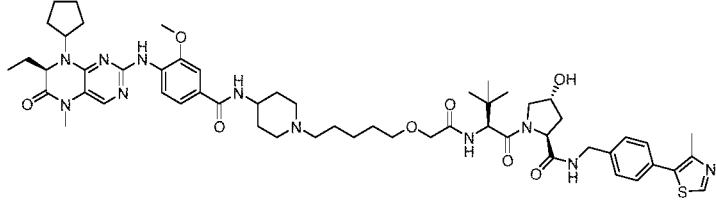
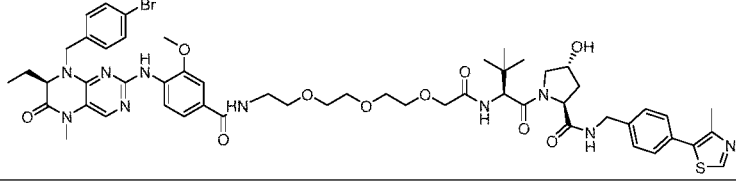
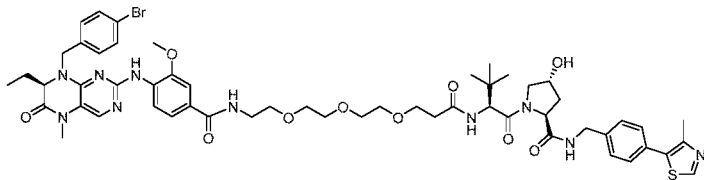
Table B10

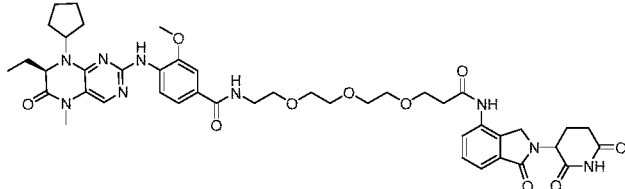
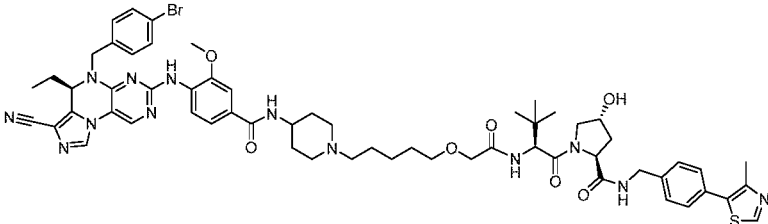
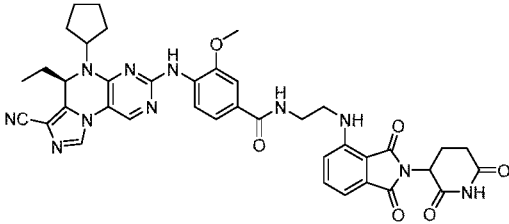
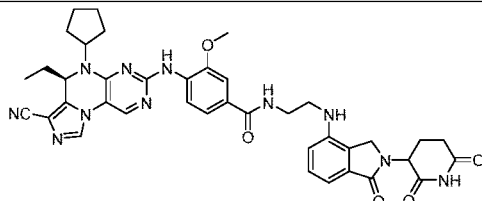
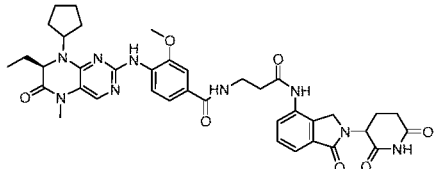
Examples	Structure and data analysis
UB-180551 (51)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.01 (s, 1H), 9.86 (s, 1H), 8.70 (s, 1H), 8.64 (s, 1H), 8.45 (t, <i>J</i> = 5.7 Hz, 1H), 8.25 (d, <i>J</i> = 8.2 Hz, 1H), 8.00 (s, 1H), 7.81 (dd, <i>J</i> = 7.1, 1.7 Hz, 1H), 7.49 (dd, <i>J</i> = 15.4, 8.2 Hz, 4H), 5.27 – 5.07 (m, 2H), 4.55 – 4.28 (m, 3H), 3.92 (s, 3H), 3.53 – 3.32 (m, 7H), 3.12 – 2.86 (m, 3H), 2.41–2.32 (m, 3H), 2.07 – 1.76 (m, 18H), 0.86–0.79 (m, 3H), 0.74 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [<i>M</i>+<i>H</i>]⁺ = 948.7

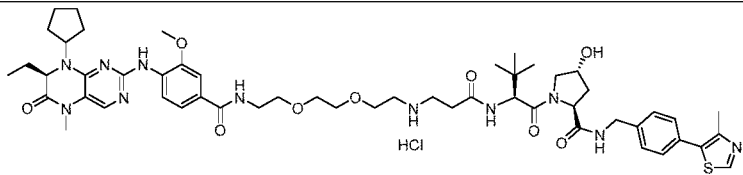
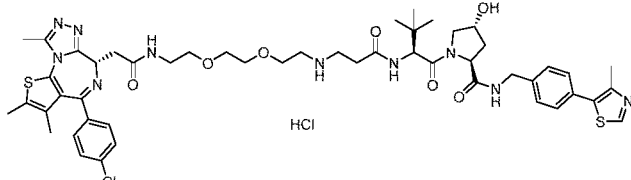
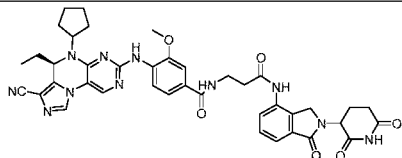
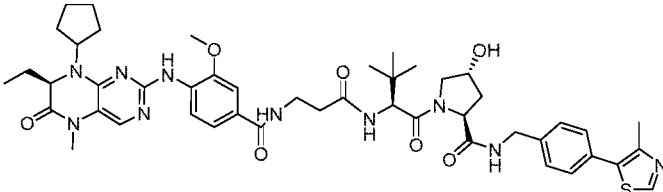
	¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.01 (s, 1H), 9.71(s, 1H), 8.37-8.32(m, 1H), 8.25 (d, <i>J</i> = 8.2 Hz, 1H), 7.85 (s, 1H), 7.62 – 7.41(m, 5H), 7.38 (dt, <i>J</i> = 28.2, 20.4 Hz, 1H), 5.25 (dd, <i>J</i> = 12.8, 5.4 Hz, 1H), 5.21-5.14 (m, 1H), 4.41 – 4.26 (m, 3H), 4.23 – 4.00 (m, 3H), 3.84 (m, 1H), 3.77 – 3.59 (m, 11H), 2.89-2.67 (m, 1H), 2.06 – 1.97 (m, 2H), 1.75 – 1.66 (m, 2H), 0.82 – 0.78 (m, 2H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 956.3
UB-180567 (67)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.09 (s, 1H), 8.59 (s, 1H), 8.42 (d, <i>J</i> = 8.2 Hz, 1H), 7.85 (s, 1H), 7.64 – 7.53 (m, 2H), 7.47 (d, <i>J</i> = 8.9 Hz, 2H), 7.31 (t, <i>J</i> = 11.1 Hz, 1H), 7.03 (d, <i>J</i> = 7.0 Hz, 1H), 6.85 (d, <i>J</i> = 6.1 Hz, 1H), 5.05 (dd, <i>J</i> = 12.8, 5.4 Hz, 1H), 4.59 – 4.19 (m, 2H), 3.93 (s, 3H), 3.70 – 3.34 (m, 4H), 3.31 – 3.21 (m, 5H), 2.89-2.75 (m, 1H), 2.64 – 2.54 (m, 1H), 2.05 – 1.51 (m, 11H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 725.3
UB-180568 (68)	 ¹H NMR (400 MHz, chloroform-<i>d</i>) δ 8.55 (s, 1H), 8.29 (s, 1H), 7.71 (s, 1H), 7.45 (m, 3H), 7.20 (m, 2H), 7.07 (d, 1H), 6.83 (d, 1H), 6.52 (m, 1H), 5.50 (d, 1H), 4.79 (m, 1H), 4.10 (m, 2H), 3.93 (s, 3H), 3.70 (m, 8H), 3.33 (m, 4H), 2.70 (m, 3H), 2.03 (d, 2H), 1.92 (m, 1H), 1.80 (m, 1H), 1.25 (m, 7H), 0.80 (m, 3H). LCMS [M+H]⁺ = 912.6
UB-180569 (69)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 8.68 (d, <i>J</i> = 11.3 Hz, 1H), 8.38 (d, <i>J</i> = 8.4 Hz, 1H), 7.70 (d, <i>J</i> = 21.3 Hz, 2H), 7.40-7.53 (m, 3H), 7.19 (s, 2H), 7.08 (d, <i>J</i> = 7.1 Hz, 1H), 6.86 (d, <i>J</i> = 8.5 Hz, 1H), 6.73 (s, 1H), 6.44 (d, <i>J</i> = 5.0 Hz, 1H), 5.50 (d, <i>J</i> = 15.3 Hz, 1H), 4.90 (m, 1H), 4.10-4.22 (m, 2H), 3.93 (d, <i>J</i> = 1.5 Hz, 2H), 3.59-3.78 (m, 12H), 3.26-3.46 (m, 5H), 2.61-2.98 (m, 3H), 2.03-2.19 (m, 1H), 1.88-1.97 (m, 1H), 1.74-1.86 (m, 1H), 0.84 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 956.5.
UB-180570 (70)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 9.97 (s, 1H), 8.76 (t, <i>J</i> = 5.8 Hz, 1H), 8.47 (d, <i>J</i> = 8.9 Hz, 1H), 7.91 – 7.79 (m, 3H), 7.63 (s, 1H), 7.62 – 7.20 (m, 6H), 5.16 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.51 – 4.29 (m, 4H), 4.29 – 4.14 (m, 2H), 3.95 (s, 3H), 3.25 (s, 3H), 3.00 – 2.86 (m, 1H), 2.04 – 1.58 (m, 11H), 0.77 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 725.3

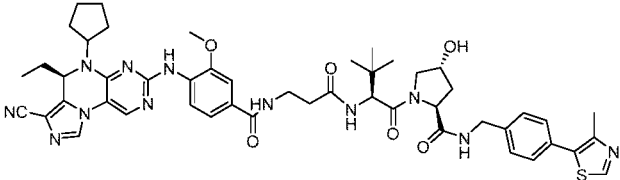
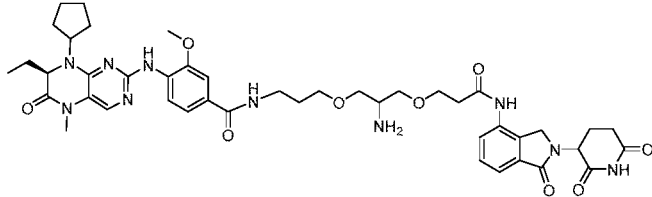
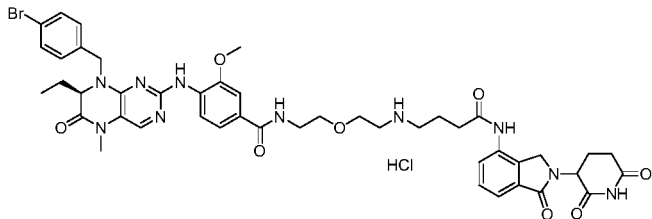
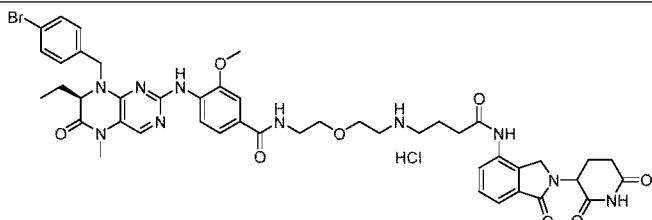
UB-180571 (71)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.09 (s, 1H), , 9.81 (s, 1H), 8.96 (s, 1H), 8.47 (d, J = 8.9 Hz, 1H), 8.42 (d, J = 8.2 Hz, 1H), 7.85 (s, 1H), 7.64 – 7.53 (m, 6H), 5.05 (dd, J = 12.8, 5.4 Hz, 1H), 4.59 – 4.19 (m, 4H), 3.93 (s, 3H), 3.24 (s, 3H), 2.89-2.75 (m, 1H), 2.64 – 2.54 (m, 1H), 2.54 – 2.46 (m, 52H), 2.33 (s, 2H), 2.05 – 1.51 (m, 13H), 1.26 (dd, J = 14.4, 10.8 Hz, 3H), 0.76 (t, J = 7.4 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 750.58$
UB-180579 (79)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.09 (s, 1H), 8.49-8.40 (m, 2H), 7.85 (s, 1H), 7.64 – 7.53 (m, 2H), 7.47 (d, J = 8.9 Hz, 2H), 7.31 (t, J = 11.1 Hz, 1H), 7.03 (d, J = 7.0 Hz, 1H), 6.85 (d, J = 6.1 Hz, 1H), 5.05 (dd, J = 12.8, 5.4 Hz, 1H), 4.59 – 4.19 (m, 2H), 3.93 (s, 3H), 3.47 – 3.44 (m, 3H), 3.27 – 3.24 (m, 3H), 3.21 (s, 3H), 2.89-2.81 (m, 1H), 2.05 – 1.51 (m, 13H), 0.76 (t, J = 7.4 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 739.5$
UB-180591 (91)	 ^1H NMR (400 MHz, DMSO-d_6) δ 8.98 (s, 1H), 8.70 (s, 1H), 8.64 (s, 1H), 8.57 (m, 1H), 8.24 (d, J = 8.0 Hz, 1H), 8.14 (m, 1H), 7.99 (m, 1H), 7.86 (m, 1H), 7.50 (m, 1H), 7.40 (m, 3H), 7.21 (m, 2H), 6.66 (m, 1H), 5.32 (m, 2H), 5.15 (m, 1H), 4.55 (m, 1H), 4.47 (m, 1H), 4.47 (m, 2H), 4.36 (m, 1H), 3.93 (s, 3H), 2.89 (m, 2H), 2.44 (s, 3H), 2.31 (m, 2H), 2.08-1.87 (m, 8H), 1.77 (m, 4H), 1.46 (m, 10H), 0.94 (s, 9H), 0.85 (t, J = 8.0 Hz, 3H), 0.74 (t, J = 8.0 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 1054.7$
UB-180592 (92)	 ^1H NMR (400 MHz, DMSO-d_6) δ 8.96 (d, J = 6.3 Hz, 1H), 8.71 (d, J = 6.2 Hz, 2H), 8.67 – 8.44 (m, 2H), 8.27 (d, J = 8.1 Hz, 1H), 8.02 (s, 1H), 7.86 (d, J = 9.3 Hz, 1H), 7.60 – 7.37 (m, 6H), 5.29 – 5.10 (m, 2H), 4.56 (d, J = 9.3 Hz, 1H), 4.56-4.09 (m, 6H), 4.09 – 3.97 (m, 2H), 3.93 (s, 3H), 3.73 – 3.60 (m, 2H), 2.43 (s, 3H), 2.18 – 1.67 (m, 12H), 0.94 (s, 9H), 0.74 (t, J = 7.3 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 950.3$

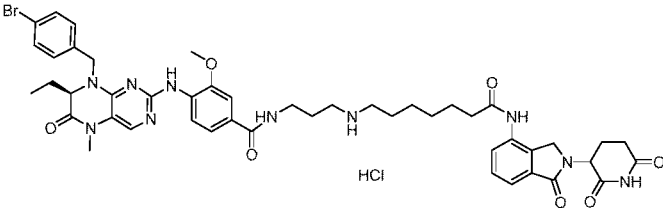
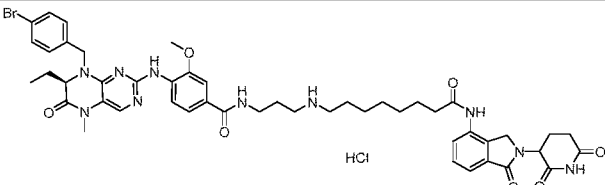
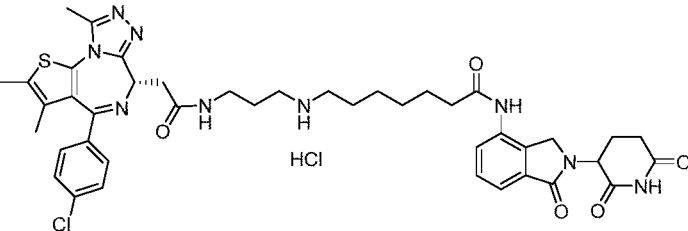
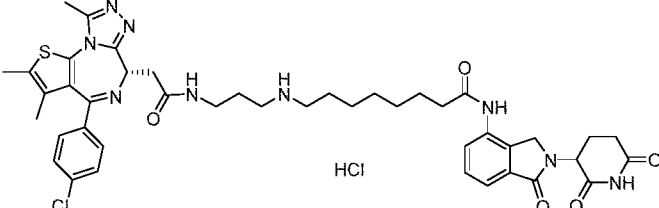
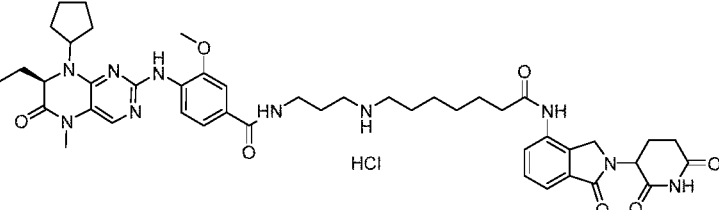
UB-180593 (93)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.10 (s, 1H), 8.70 (s, 1H), 8.64 (s, 1H), 8.49 (s, 1H), 8.24 (d, <i>J</i> = 8.1 Hz, 1H), 8.01 (s, 1H), 7.71 – 7.44 (m, 3H), 7.12 (d, <i>J</i> = 8.7 Hz, 1H), 7.03 (d, <i>J</i> = 7.1 Hz, 1H), 6.77 (s, 1H), 5.16 (s, 2H), 5.05 (dd, <i>J</i> = 12.1, 4.9 Hz, 1H), 4.45 – 4.36 (m, 1H), 3.93(s,3H), 2.87 (d, <i>J</i> = 14.0 Hz, 2H), 2.44 – 2.36 (m, 1H), 2.03– 1.55 (m, 15H), 0.74 (t, <i>J</i> = 6.8 Hz, 3H). LCMS [M+H]⁺ =773.3
UB-180594 (94)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.01 (s, 1H), 9.70 (s, 1H), 8.96 (d, <i>J</i> = 6.3 Hz, 1H), 8.71 (d, <i>J</i> = 6.2 Hz, 1H), 8.67 – 8.44 (m, 2H), 8.27 (d, <i>J</i> = 8.1 Hz, 1H), 8.02 (s, 1H), 7.86 (d, <i>J</i> = 9.3 Hz, 1H), 7.60 – 7.37 (m, 5H), 7.36 (s, 1H), 5.29 – 5.10 (m, 3H), 4.56 (d, <i>J</i> = 9.3 Hz, 1H), 4.41 – 4.29 (m, 2H), 4.21(s,3H), 3.93(s, 3H), 3.73 – 3.40 (m, 13H), 2.87-2.81 (m, 1H), 2.43-2.33 (s, 2H), 2.05-1.95 (m, 1H), 1.92 (d, <i>J</i> = 8.5 Hz, 2H), 0.74 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [M+H]⁺ =992.5
UB-180595 (95)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.09 (s, 1H), 8.75 (s, 1H), 8.70 (s, 1H), 8.46 (t, <i>J</i> = 5.6 Hz, 1H), 8.17 – 7.99 (m, 2H), 7.68 – 7.50 (m, 4H), 7.50 – 7.40 (m, 1H), 7.36 (t, <i>J</i> = 14.5 Hz, 2H), 7.12 (d, <i>J</i> = 8.7 Hz, 1H), 7.03 (d, <i>J</i> = 7.1 Hz, 1H), 6.76 (t, <i>J</i> = 6.0 Hz, 1H), 5.32 – 5.11 (m, 3H), 5.05 (dd, <i>J</i> = 12.9, 5.3 Hz, 1H), 3.91 (s, 3H), 2.89 – 2.76 (m, 1H), 2.29 – 1.96 (m, 1H), 1.83 (dt, <i>J</i> = 13.8, 6.9 Hz, 5H), 0.72 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [M+H]⁺ =872.2
UB-180596 (96)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 9.88 (s, 1H), 8.43-7.40 (d, <i>J</i> = 12 Hz, 1H), 8.16 (m, 1H), 7.85-7.81 (m, 2H), 7.60 (s, 1H), 7.51-7.48 (m, 2H), 5.17-5.13 (m, 1H), 4.40-4.32 (m, 2H), 4.25-4.22 (m, 1H), 3.94 (s, 3H), 3.83 (m, 2H), 3.25 (s, 3H), 3.02-2.96 (m, 5H), 2.45-2.32 (m, 6H), 2.01-1.73 (m, 11H), 1.67-1.62 (m, 8H), 0.78 (t, <i>J</i> = 14.7, 8.0 Hz, 3H). LCMS [M+H]⁺ =850.4

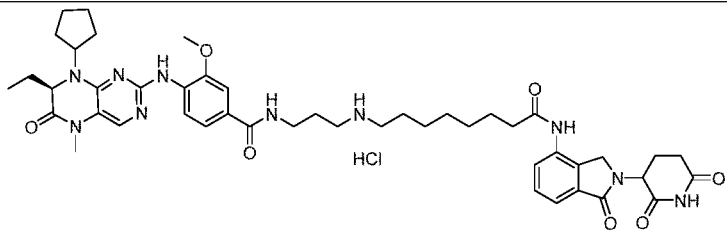
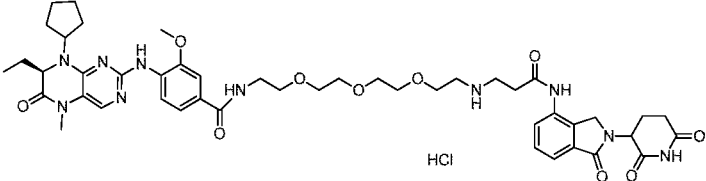
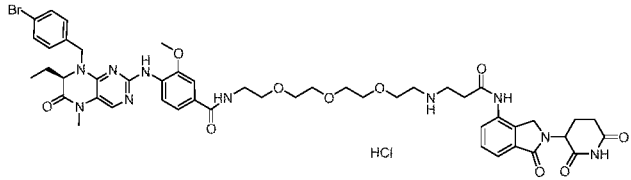
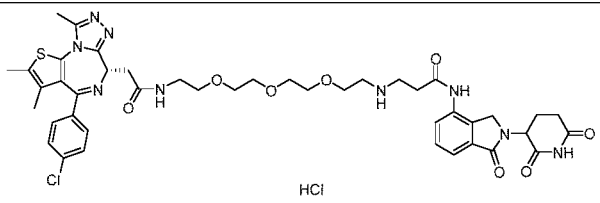
UB-180597 (97)	 ^1H NMR (400 MHz, DMSO-d_6) δ 8.98 (s, 1H), 8.70 (s, 1H), 8.64 (m, 2H), 8.25 (m, 2H), 8.00 (s, 1H), 7.56-7.46 (m, 2H), 7.40 (m, 4H), 7.21 (m, 1H), 6.66 (m, 1H), 6.53 (m, 1H), 5.32 (m, 3H), 5.18 (m, 1H), 4.56 (m, 1H), 4.47-4.41 (m, 2H), 4.36 (m, 2H), 4.29 (m, 1H), 3.93 (s, 3H), 3.64 (m, 3H), 2.44 (s, 3H), 1.99 (m, 10H), 1.76 (m, 4H), 1.64-1.50 (m, 4H), 1.45 (m, 4H), 0.95 (s, 9H), 0.90-0.83 (m, 4H), 0.74 (t, J = 8.0 Hz, 3H). LCMS $[\text{M}+\text{H}]^+$ = 1098.8
UB-180598 (98)	 ^1H NMR (400 MHz, DMSO-d_6) δ 8.98 (s, 1H), 8.65 (m, 1H), 8.42 (m, 1H), 8.30 (m, 1H), 7.92 (m, 1H), 7.85 (s, 1H), 7.65 (m, 1H), 7.61 (s, 1H), 7.52 (m, 1H), 7.39 (m, 4H), 5.32 (m, 1H), 5.19 (m, 1H), 4.56 (m, 1H), 4.47-4.22 (m, 7H), 3.94 (m, 4H), 3.64 (m, 2H), 3.48 (m, 4H), 3.25 (s, 3H), 2.44 (s, 3H), 2.09-1.95 (m, 4H), 1.93-1.86 (m, 6H), 1.78 (m, 5H), 1.61 (m, 8H), 0.95 (s, 9H), 0.84 (m, 3H), 0.76 (t, J = 8.0 Hz, 3H). LCMS $[\text{M}+\text{H}]^+$ = 1064.8
UB-180601 (101)	 ^1H NMR (400 MHz, DMSO-d_6) δ 8.97 (s, 1H), 8.61 (t, J = 6.0 Hz, 1H), 8.39 (t, J = 5.6 Hz, 1H), 8.27 (d, J = 8.5 Hz, 1H), 7.87 (s, 1H), 7.65 (s, 1H), 7.54 (d, J = 8.3 Hz, 2H), 7.48 (d, J = 1.9 Hz, 1H), 7.45 – 7.37 (m, 6H), 7.34 (d, J = 8.2 Hz, 2H), 5.25 (d, J = 15.6 Hz, 1H), 5.17 (d, J = 3.6 Hz, 1H), 4.56 (d, J = 9.6 Hz, 1H), 4.48 – 4.33 (m, 4H), 4.30 – 4.15 (m, 2H), 3.96 (s, 2H), 3.90 (s, 3H), 3.69 – 3.48 (m, 12H), 3.40 (t, J = 5.5 Hz, 2H), 3.27 (s, 3H), 2.44 (s, 3H), 2.06 (t, J = 10.4 Hz, 1H), 1.95 – 1.86 (m, 1H), 1.76 (dq, J = 14.2, 6.8 Hz, 2H), 0.94 (s, 10H), 0.73 (t, J = 7.4 Hz, 3H). LC-MS : V879-129-F LCMS $[\text{M}+\text{H}]^+$ = 1127.1
UB-180602 (102)	 ^1H NMR (400 MHz, DMSO-d_6) δ 8.98 (s, 1H), 8.57 (t, J = 6.1 Hz, 1H), 8.38 (t, J = 5.7 Hz, 1H), 8.27 (d, J = 8.5 Hz, 1H), 7.89 (q, J = 9.6 Hz, 2H), 7.65 (s, 1H), 7.56 – 7.51 (m, 2H), 7.48 (d, J = 1.9 Hz, 1H), 7.44 – 7.33 (m, 7H), 5.25 (d, J = 15.6 Hz, 1H), 5.13 (d, J = 3.5 Hz, 1H), 4.55 (d, J = 9.4 Hz, 1H), 4.49 – 4.39 (m, 3H), 4.35 (s, 1H), 4.28 – 4.13 (m, 2H), 3.91 (s, 3H), 3.66-3.50 (m, 18H), 3.27 (s, 3H), 2.44 (s, 3H), 2.08 – 1.99 (m, 1H), 1.90 (M, 1H), 1.77 (dt, J = 14.4, 7.0 Hz, 2H), 0.93 (s, 9H), 0.73 (t, J = 7.4 Hz, 3H). LC-MS : V879-130-F LCMS $[\text{M}+\text{H}]^+$ = 1143.4

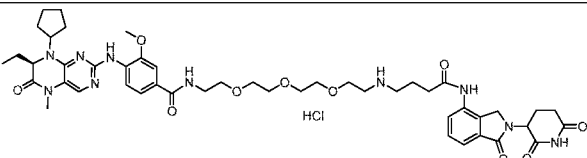
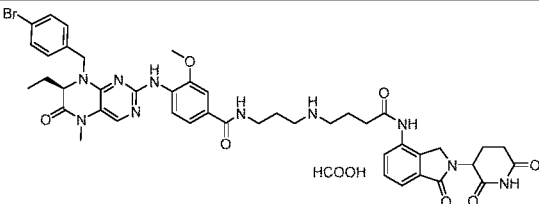
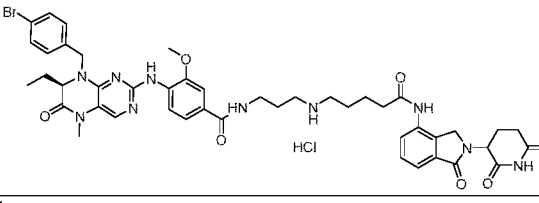
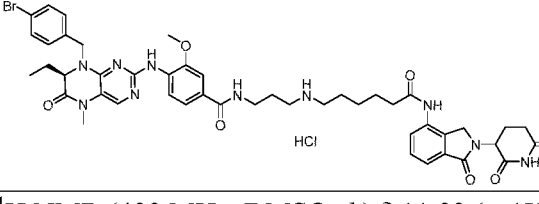
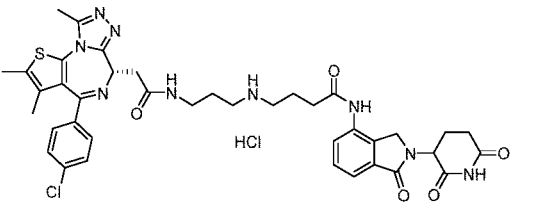
UB-180605 (105)	
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 9.83 (s, 1H), 8.50 - 8.32 (m, 2H), 7.92 - 7.73 (m, 2H), 7.60 (s, 1H), 7.55 - 7.36 (m, 4H), 5.14 (dd, <i>J</i> = 13.2, 5.2 Hz, 1H), 4.47 - 4.28 (m, 3H), 4.27 - 4.19 (m, 1H), 3.93 (s, 3H), 3.70 (t, <i>J</i> = 6.4 Hz, 2H), 3.63 - 3.44 (m, 9H), 3.44 - 3.37 (m, 2H), 3.24 (s, 3H), 2.97 - 2.85 (m, 1H), 2.62 - 2.57 (m, 2H), 2.46 - 2.42 (m, 1H), 2.38 - 2.29 (m, 1H), 2.09 - 1.96 (m, 2H), 1.96 - 1.70 (m, 6H), 1.69 - 1.52 (m, 3H), 0.94 (s, 1H), 0.76 (t, <i>J</i> = 7.6 Hz, 3H). LCMS [M+H] ⁺ : 870.5.
UB-180606 (106)	
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 8.98 (s, 1H), 8.75 (d, 1H), 8.70 (s, 1H), 8.62 (m, 1H), 8.09 (m, 1H), 8.04 (m, 1H), 7.63-7.48 (m, 4H), 7.36-7.31 (m, 4H), 7.35 (m, 3H), 7.20 (m, 1H), 6.65 (s, 1H), 5.32 (t, 1H), 5.27-5.06 (m, 4H), 4.69-4.53 (m, 2H), 4.45 (t, 1H), 4.35 (m, 1H), 4.28 (m, 1H), 3.96-3.87 (m, 5H), 3.66 (m, 2H), 3.50 (m, 4H), 2.44 (s, 3H), 2.09-1.93 (m, 6H), 1.91-1.82 (m, 6H), 1.58 (m, 2H), 1.45 (m, 2H), 0.95 (s, 9H), 0.88-0.83 (m, 3H), 0.72 (t, <i>J</i> = 8.0 Hz, 3H). LCMS [M+H] ⁺ =1198.3
UB-180607 (107)	
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.09 (s, 1H), 8.70 (s, 1H), 8.71 (d, <i>J</i> = 6.2 Hz, 2H), 8.27 - 8.24 (m, 1H), 8.02 (s, 1H), 7.86 (d, <i>J</i> = 9.3 Hz, 3H), 7.60 - 7.37 (m, 17H), 7.36 (s, 2H), 5.29 - 5.10 (m, 2H), 4.56 (d, <i>J</i> = 9.3 Hz, 1H), 4.09 - 3.73 (m, 16H), 3.91 (s, 3H), 3.73 - 3.40 (m, 4H), 2.67 - 2.43 (m, 4H), 2.03 - 1.52 (m, 13H), 0.74 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [M+H] ⁺ =758.3
UB-180608 (108)	
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.01 (s, 1H), 8.72 (d, <i>J</i> = 8.6 Hz, 1H), 8.69 - 8.46 (m, 2H), 8.27 (d, <i>J</i> = 8.3 Hz, 1H), 8.01 (s, 1H), 7.57 - 7.45 (m, 2H), 7.32 (t, <i>J</i> = 7.7 Hz, 1H), 6.94 (t, <i>J</i> = 8.0 Hz, 2H), 5.83 (dd, <i>J</i> = 24.0, 18.3 Hz, 1H), 5.15 (ddd, <i>J</i> = 18.3, 10.6, 4.3 Hz, 2H), 4.48 - 4.23 (m, 3H), 3.93 (s, 3H), 3.46 - 3.26 (m, 2H), 3.05 - 2.83 (m, 1H), 2.38 - 2.21 (m, 1H), 2.02 - 1.50 (m, 11H), 1.34 (s, 2H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ =744.2
UB-180618 (118)	

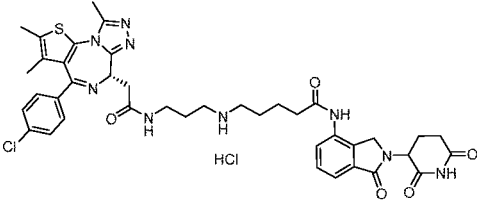
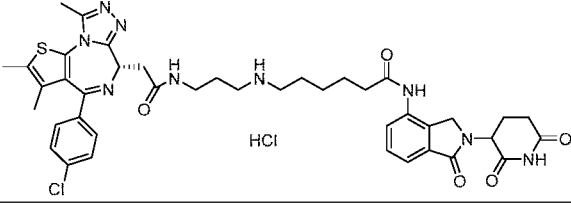
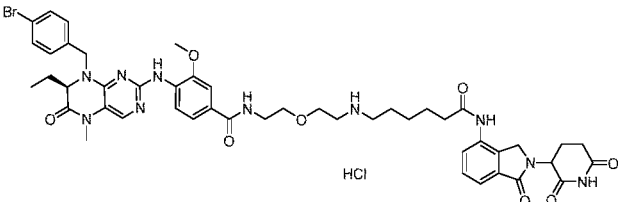
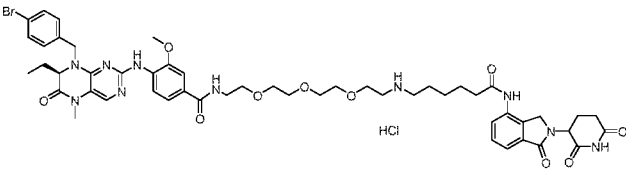
	¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.00 (s, 1H), 9.90 (s, 1H), 8.53 (t, <i>J</i> = 5.7 Hz, 1H), 8.39 (d, <i>J</i> = 8.3 Hz, 1H), 7.84 (s, 1H), 7.81 (dd, <i>J</i> = 7.0, 2.1 Hz, 1H), 7.64 (s, 1H), 7.54 – 7.45 (m, 4H), 5.09 (dd, <i>J</i> = 13.3, 5.2 Hz, 1H), 4.42 – 4.18 (m, 4H), 3.91 (s, 3H), 3.58 (q, <i>J</i> = 6.5 Hz, 2H), 3.25 (s, 3H), 2.97 – 2.81 (m, 1H), 2.67 (t, <i>J</i> = 6.7 Hz, 2H), 2.05 – 1.51 (m, 12H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 738.3
UB-180619 (119)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 8.98 (s, 1H), 8.60 (s, 1H), 8.49 (s, 1H), 8.42 (d, <i>J</i> = 8.3 Hz, 1H), 8.33 (d, <i>J</i> = 9.2 Hz, 1H), 7.84 (s, 1H), 7.60 (s, 1H), 7.53 (t, <i>J</i> = 2.7 Hz, 2H), 7.40 (q, <i>J</i> = 8.4 Hz, 4H), 7.21 (s, 1H), 6.65 (s, 1H), 5.32 (dd, <i>J</i> = 5.6, 4.1 Hz, 1H), 5.18 (d, <i>J</i> = 3.4 Hz, 1H), 4.54 (d, <i>J</i> = 9.3 Hz, 1H), 4.43 (s, 2H), 4.35 (d, <i>J</i> = 6.8 Hz, 2H), 4.27 – 4.18 (m, 2H), 3.94 (s, 3H), 3.70 – 3.60 (m, 4H), 3.58 – 3.56 (m, 3H), 3.53 – 3.49 (m, 2H), 3.45 – 3.40 (m, 2H), 3.24 (s, 3H), 3.07 – 2.91 (m, 4H), 2.59 (d, <i>J</i> = 7.3 Hz, 1H), 2.44 (s, 3H), 1.99 (dd, <i>J</i> = 8.6, 6.5 Hz, 4H), 1.89 (q, <i>J</i> = 4.3 Hz, 2H), 1.80 – 1.76 (m, 2H), 1.65 – 1.59 (m, 2H), 1.46 (d, <i>J</i> = 7.3 Hz, 2H), 0.94 (s, 9H), 0.76 (t, 3H). LCMS [M+H]⁺ = 1040.2
UB-180620 (120)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 8.96 (s, 1H), 8.60 (t, <i>J</i> = 6.1 Hz, 1H), 8.34 – 8.25 (m, 2H), 7.48 – 7.36 (m, 8H), 7.29 (s, 1H), 6.70 – 6.63 (m, 1H), 5.32 – 5.30 (m, 1H), 5.26 (d, <i>J</i> = 3.5 Hz, 1H), 4.54 – 4.47 (m, 2H), 4.45 – 4.39 (m, 2H), 4.36 (s, 1H), 4.26 (q, <i>J</i> = 5.3 Hz, 8H), 4.19 (d, <i>J</i> = 5.4 Hz, 1H), 3.26 (d, <i>J</i> = 4.7 Hz, 2H), 3.08 (d, <i>J</i> = 7.2 Hz, 2H), 2.70 – 2.62 (m, 2H), 2.59 (s, 3H), 2.43 (s, 3H), 2.39 (s, 3H), 2.00 – 1.93 (m, 4H), 1.61 (s, 3H), 1.45 (t, <i>J</i> = 7.1 Hz, 2H), 0.93 (s, 9H). LCMS [M+H]⁺ = 1015.2
UB-180621 (121)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.00 (s, 1H), 9.90 (s, 1H), 8.70 (s, 1H), 8.64 (d, <i>J</i> = 1.0 Hz, 1H), 8.58 (d, <i>J</i> = 5.8 Hz, 1H), 8.23 (dd, <i>J</i> = 8.3, 2.0 Hz, 1H), 7.99 (s, 1H), 7.81 (dd, <i>J</i> = 6.9, 2.0 Hz, 1H), 7.53 – 7.45 (m, 4H), 5.17 (dd, <i>J</i> = 8.1, 3.7 Hz, 1H), 5.10 (dd, <i>J</i> = 13.2, 5.2 Hz, 1H), 4.39 (dd, <i>J</i> = 10.7, 6.7 Hz, 1H), 4.31 (s, 1H), 3.90 (s, 3H), 3.59 (q, <i>J</i> = 6.7 Hz, 2H), 2.71 – 2.62 (m, 2H), 2.23 (tt, <i>J</i> = 15.7, 7.0 Hz, 2H), 2.01 – 1.51 (m, 12H), 0.74 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [M+H]⁺ = 772.3
UB-180622 (122)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 8.98 (s, 1H), 8.56 (t, <i>J</i> = 6.1 Hz, 1H), 8.42 – 8.29 (m, 2H), 7.99 (d, <i>J</i> = 9.2 Hz, 1H), 7.84 (s, 1H), 7.53 – 7.43 (m, 2H), 7.40 (q, <i>J</i> = 8.4 Hz, 4H), 5.15 (s, 1H), 4.56 (d, <i>J</i> = 9.3 Hz, 1H), 4.50 – 4.38 (m, 2H), 4.34 (d, <i>J</i> = 17.3 Hz, 2H), 4.29 – 4.15 (m, 2H), 3.92 (s, 3H), 3.72 – 3.55 (m, 3H), 3.51 – 3.36 (m, 2H), 3.24 (s, 3H), 3.14 (qd, <i>J</i> = 7.3, 4.2 Hz, 1H), 2.44 (s, 3H), 2.10 – 1.52 (m, 12H), 0.93 (s, 9H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 909.5

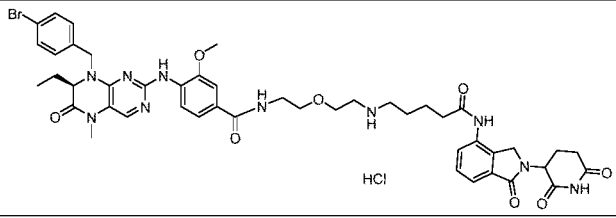
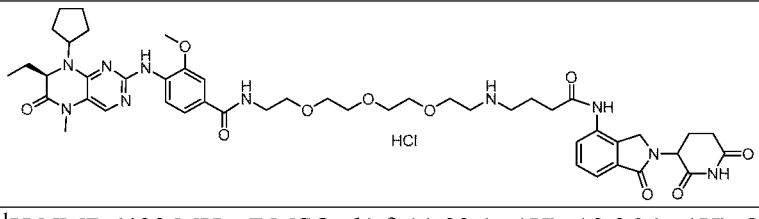
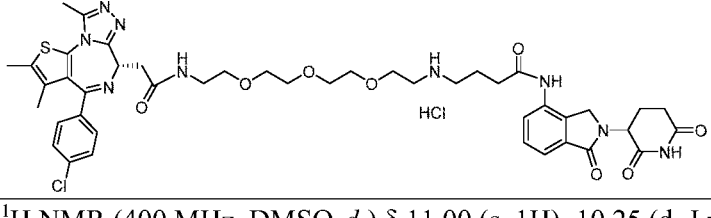
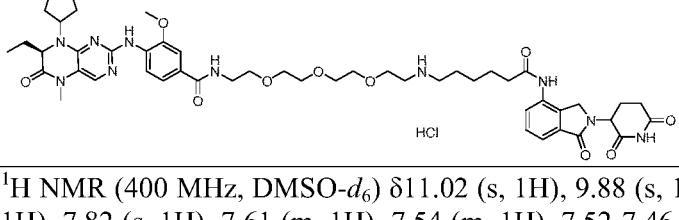
UB-180623 (123)	 ^1H NMR (400 MHz, DMSO-d_6) δ 8.97 (s, 1H), 8.70 (s, 1H), 8.64 (s, 1H), 8.57 (t, J = 6.1 Hz, 1H), 8.37 (t, J = 5.5 Hz, 1H), 8.24 (d, J = 8.3 Hz, 1H), 8.05 – 7.96 (m, 2H), 7.60 – 7.45 (m, 2H), 7.40 (q, J = 8.3 Hz, 4H), 5.26 – 5.08 (m, 2H), 4.56 (d, J = 9.3 Hz, 1H), 4.49 – 4.32 (m, 4H), 4.22 (dd, J = 15.8, 5.5 Hz, 1H), 3.92 (s, 3H), 3.68 (d, J = 5.0 Hz, 2H), 3.52 – 3.36 (m, 2H), 2.44 (s, 3H), 2.08 – 1.53 (m, 13H), 0.93 (s, 9H), 0.74 (t, J = 7.4 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 943.4$
UB-180624 (124)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.02 (s, 1H), 10.21 (s, 1H), 8.58 (t, J = 5.8 Hz, 1H), 8.41 (d, J = 8.4 Hz, 1H), 7.88 – 7.78 (m, 2H), 7.67 – 7.42 (m, 5H), 5.14 (dd, J = 13.3, 5.1 Hz, 1H), 4.52 – 4.30 (m, 3H), 4.24 (dd, J = 7.6, 3.6 Hz, 1H), 3.94 (s, 3H), 3.77 (q, J = 6.0 Hz, 2H), 3.61 (m, 4H), 3.47 (m, 4H), 3.25 (s, 3H), 2.98 – 2.85 (m, 2H), 2.71 (m, 1H), 2.66 – 2.56 (m, 2H), 2.09 – 1.99 (m, 2H), 1.88 (m, 2H), 1.80 – 1.75 (m, 7H), 1.64 (dt, J = 14.4, 7.4 Hz, 2H), 1.28 – 1.20 (m, 2H), 0.76 (t, J = 7.4 Hz, 3H). LCMS: $(\text{M}+\text{H})^+ = 869.4$
UB-180625 (125)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.09 (s, 1H), 8.75 (s, 1H), 8.70 (s, 1H), 8.46 (t, J = 5.6 Hz, 1H), 8.17 – 7.99 (m, 2H), 7.68 – 7.50 (m, 3H), 7.50 – 7.40 (m, 2H), 7.36 (t, J = 14.5 Hz, 3H), 7.12 (d, J = 8.7 Hz, 2H), 7.03 (d, J = 7.1 Hz, 1H), 6.76 (t, J = 6.0 Hz, 1H), 5.32 – 5.11 (m, 2H), 5.05 (dd, J = 12.9, 5.3 Hz, 1H), 4.60 (d, J = 15.7 Hz, 6H), 3.91 (s, 3H), 3.87 – 3.39 (m, 12H), 3.19 – 2.94 (m, 4H), 3.19 – 2.85 (m, 4H), 2.09 – 1.96 (m, 2H), 0.72 (t, J = 7.3 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 927.3$
UB-180626 (126)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.02 (s, 1H), 10.01 (s, 7H), 9.03-8.69 (m, 3H), 7.93 (s, 1H), 7.81 (t, J = 16.4 Hz, 2H), 7.67 (s, 1H), 7.58 – 7.40 (m, 6H), 7.34 (d, J = 8.4 Hz, 2H), 5.15 (dd, J = 13.0, 5.2 Hz, 2H), 4.77 – 4.46 (m, 1H), 4.37 (t, J = 17.5 Hz, 3H), 3.93 (s, 3H), 3.93 (s, 46H), 3.98 – 3.62 (m, 8H), 3.24 (s, 3H), 3.22 – 2.86 (m, 2H), 2.70 – 2.06 (m, 4H), 2.28 – 2.16 (m, 1H), 2.06-1.89 (m, 6H), 0.74 (t, J = 7.3 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 939.3$

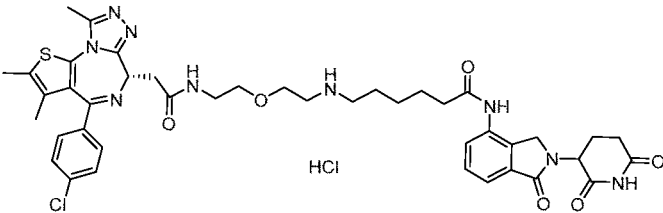
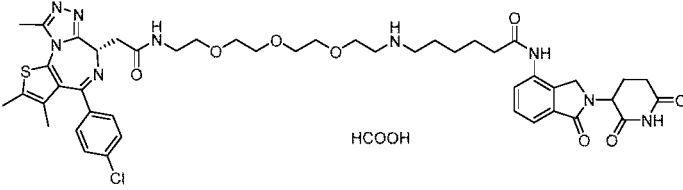
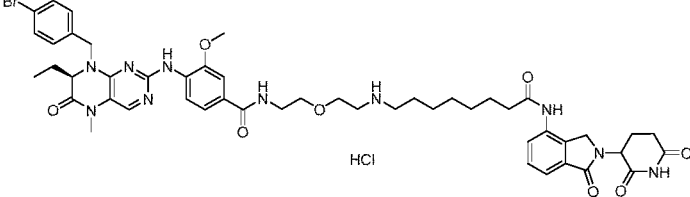
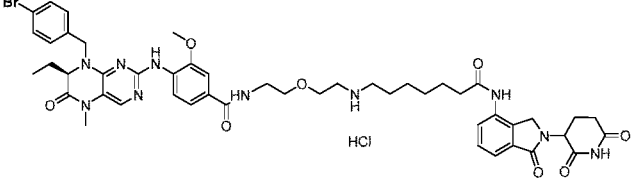
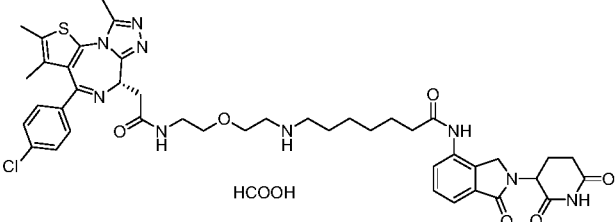
UB-180627 (127)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.02 (s, 1H), 9.86 (s, 1H), 8.77 (s, 4H), 7.93 (s, 1H), 7.88 – 7.78 (m, 2H), 7.60 – 7.44 (m, 5H), 7.42 – 7.31 (m, 3H), 5.15 (dd, J = 14.3, 4.1 Hz, 2H), 4.61 (d, J = 15.6 Hz, 1H), 4.46 – 4.30 (m, 4H), 3.92 (s, 3H), 3.56 (d, J = 2.6 Hz, 3H), 3.24 (s, 3H), 2.90 (d, J = 18.4 Hz, 6H), 2.61 (d, J = 18.1 Hz, 2H), 2.41 – 2.28 (m, 3H), 2.03 (d, J = 6.1 Hz, 1H), 1.88 (s, 4H), 1.62 (s, 4H), 1.35 (s, 4H), 0.74 (d, J = 7.5 Hz, 3H). LC-MS : $[\text{M}+\text{H}]^+ = 953.3$
UB-180628 (128)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.02 (s, 1H), 9.85 (s, 1H), 8.76 (s, 3H), 8.08 – 7.74 (m, 3H), 7.61 – 7.45 (m, 5H), 7.46 – 7.31 (m, 3H), 5.22 – 5.10 (m, 2H), 4.59 (d, J = 15.5 Hz, 1H), 4.46 – 4.27 (m, 3H), 3.92 (s, 3H), 3.25 (s, 3H), 2.99 – 2.79 (m, 6H), 2.70 – 2.57 (m, 2H), 2.43 – 2.24 (m, 3H), 2.08 – 1.80 (m, 6H), 1.61 (s, 4H), 1.32 (s, 6H), 0.75 (t, J = 7.4 Hz, 3H). LC-MS : $[\text{M}+\text{H}]^+ = 967.3$
UB-180629 (129)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.02 (s, 1H), 9.89 (s, 1H), 8.77 (s, 2H), 8.47 (t, J = 5.9 Hz, 1H), 7.82 (dd, J = 7.0, 2.0 Hz, 1H), 7.56 – 7.38 (m, 5H), 5.15 (dd, J = 13.3, 5.1 Hz, 1H), 4.56 – 4.50 (m, 1H), 4.43 – 4.30 (m, 2H), 3.30 – 3.14 (m, 4H), 2.95 – 2.77 (m, 5H), 2.61 (s, 4H), 2.44 – 2.32 (m, 6H), 2.07 – 1.99 (m, 1H), 1.84 – 1.78 (m, 2H), 1.64 – 1.56 (m, 7H), 1.37 – 1.30 (m, 4H). LC-MS : $[\text{M}+\text{H}]^+ = 826.3$
UB-180630 (130)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.03 (s, 1H), 9.88 (s, 1H), 8.76 (s, 2H), 8.48 (t, J = 5.9 Hz, 1H), 7.82 (d, J = 6.9 Hz, 1H), 7.57 – 7.39 (m, 6H), 5.14 (dd, J = 13.3, 5.2 Hz, 1H), 4.56 (t, J = 7.2 Hz, 2H), 4.38 (q, J = 17.5 Hz, 2H), 3.24 (m, 4H), 2.97 – 2.76 (m, 6H), 2.63 (s, 5H), 2.39 (m, 6H), 2.12 – 1.99 (m, 1H), 1.83 (d, J = 8.6 Hz, 2H), 1.61 (mz, 6H), 1.30 (s, 6H). LC-MS : $[\text{M}+\text{H}]^+ = 840.3$
UB-180631 (131)	

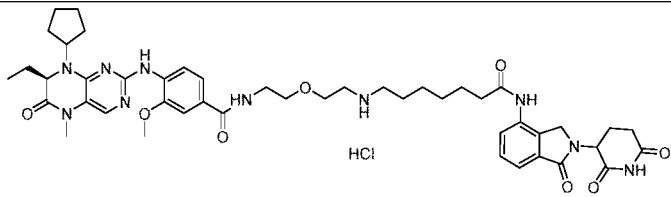
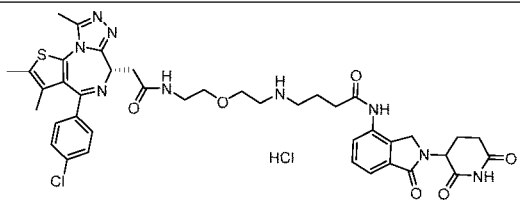
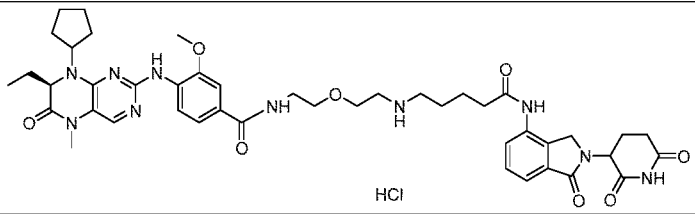
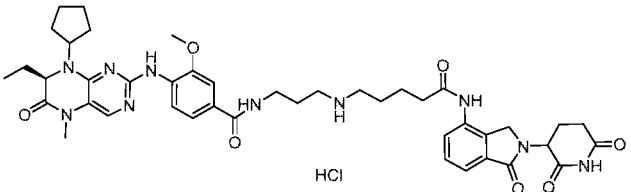
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 9.88 (s, 1H), 8.90 – 8.79 (m, 3H), 7.97 – 7.77 (m, 3H), 7.65 (d, <i>J</i> = 1.9 Hz, 1H), 7.61 – 7.41 (m, 3H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.54 – 4.28 (m, 3H), 4.17 (t, <i>J</i> = 8.8 Hz, 1H), 3.92 (s, 3H), 3.22 (s, 3H), 3.03 – 2.82 (m, 6H), 2.68 – 2.58 (m, 2H), 2.45 – 2.27 (m, 3H), 2.09 – 1.76 (m, 9H), 1.69 – 1.40 (m, 8H), 1.35 (d, <i>J</i> = 5.9 Hz, 4H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LC-MS : [M+H] ⁺ = 851.4
UB-180632 (132)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 9.86 (s, 1H), 9.65 (s, 1H), 8.84 (d, <i>J</i> = 18.1 Hz, 3H), 7.93 – 7.80 (m, 3H), 7.73 – 7.42 (m, 4H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.60 – 4.26 (m, 3H), 4.16 (t, <i>J</i> = 8.8 Hz, 1H), 3.92 (s, 3H), 3.33–3.36 (m, 2H), 3.22 (s, 3H), 2.92 (m, 5H), 2.63 (m, 2H), 2.43 – 2.25 (m, 3H), 2.09 – 1.75 (m, 8H), 1.69 – 1.44 (m, 8H), 1.32 (s, 6H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LC-MS : [M+H] ⁺ = 865.3
UB-180633 (133)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 10.12 (s, 1H), 9.77 (s, 1H), 9.00 (s, 2H), 8.69 (s, 1H), 7.85 (d, <i>J</i> = 4.8 Hz, 3H), 7.64 (d, <i>J</i> = 1.8 Hz, 1H), 7.51 (d, <i>J</i> = 1.8 Hz, 3H), 5.15 (dd, <i>J</i> = 13.3, 5.2 Hz, 1H), 4.53 – 4.33 (m, 3H), 4.15 (p, <i>J</i> = 8.8 Hz, 1H), 3.90 (s, 3H), 3.58 – 3.50 (m, 12H), 3.44 (t, <i>J</i> = 5.8 Hz, 2H), 3.21 (s, 3H), 3.10 (d, <i>J</i> = 5.6 Hz, 2H), 3.02 – 2.88 (m, 4H), 2.09 – 1.75 (m, 10H), 1.57 – 1.39 (m, 4H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 913.58
UB-180634 (134)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.03 (s, 1H), 10.28 (s, 1H), 9.60 (s, 1H), 9.00 (s, 2H), 8.62 (s, 1H), 7.96 (s, 1H), 7.85 (dd, <i>J</i> = 7.3, 1.7 Hz, 1H), 7.78 (d, <i>J</i> = 8.4 Hz, 1H), 7.59 (d, <i>J</i> = 1.9 Hz, 1H), 7.53 (dd, <i>J</i> = 7.0, 5.1 Hz, 4H), 7.34 (d, <i>J</i> = 8.5 Hz, 3H), 5.21 – 5.07 (m, 2H), 4.63 (d, <i>J</i> = 15.5 Hz, 1H), 4.51 – 4.32 (m, 3H), 3.91 (s, 3H), 3.72 – 3.69 (m, 2H), 3.56 (s, 10H), 3.24 (s, 3H), 3.13 (t, <i>J</i> = 5.5 Hz, 3H), 2.93 (hept, <i>J</i> = 5.3, 4.6 Hz, 3H), 2.65 (d, <i>J</i> = 15.0 Hz, 2H), 2.32 (qd, <i>J</i> = 13.4, 5.0 Hz, 2H), 2.16 – 1.97 (m, 2H), 1.96 – 1.85 (m, 2H), 0.75 (t, 3H). LCMS [M-100] ⁺ = 913.6
UB-180635 (135)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.03 (s, 1H), 10.28 (d, <i>J</i> = 3.7 Hz, 1H), 8.99 (s, 2H), 8.33 (t, <i>J</i> = 5.7 Hz, 1H), 7.85 (dd, <i>J</i> = 7.2, 1.8 Hz, 1H), 7.55 – 7.47 (m, 4H), 7.43 (d, <i>J</i> = 8.4 Hz, 2H), 5.15 (dd, <i>J</i> = 13.3, 5.2 Hz, 1H), 4.55 (t, <i>J</i> = 7.0 Hz, 1H), 4.52 – 4.32 (m, 2H), 3.57 (d, <i>J</i> = 14.9 Hz, 11H), 3.46 (t, <i>J</i> = 5.9 Hz, 3H), 3.34 – 3.19 (m, 7H), 3.14 (p, <i>J</i> = 5.9, 5.4 Hz, 3H), 2.93 (qd, <i>J</i> = 11.6, 10.9, 5.1 Hz, 3H), 2.63 (s, 3H), 2.42 (s, 3H), 2.38 – 2.13 (m, 2H), 2.02 (dtd, <i>J</i> = 16.8, 9.0, 8.1, 5.2 Hz, 2H). [M-100] ⁺ = 888.4

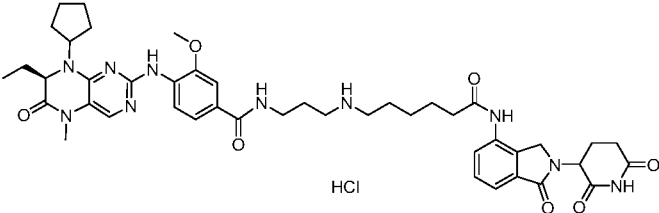
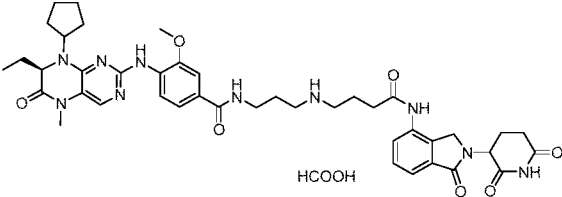
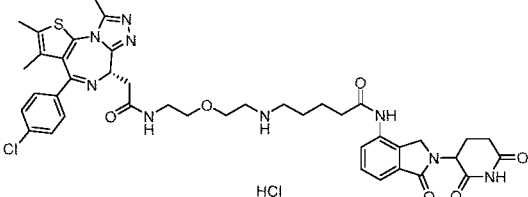
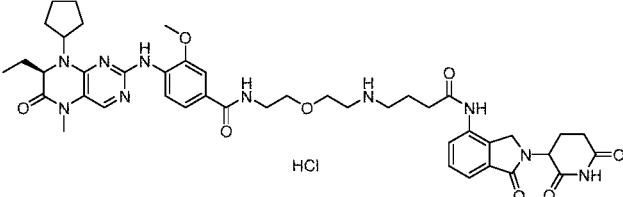
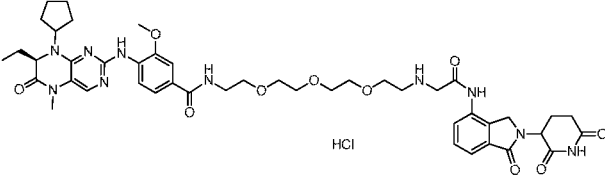
UB-180636 (136)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 10.23 (s, 1H), 8.92 (s, 2H), 8.64 (s, 1H), 7.84 (d, <i>J</i> = 4.5 Hz, 3H), 7.61 (d, <i>J</i> = 1.8 Hz, 1H), 7.57 – 7.48 (m, 3H), 5.16 (dd, <i>J</i> = 13.2, 5.1 Hz, 1H), 4.53 – 4.45 (m, 2H), 4.40 (d, <i>J</i> = 20.7 Hz, 2H), 4.25 – 4.12 (m, 1H), 3.91 (s, 3H), 3.71 (td, <i>J</i> = 6.4, 5.5, 2.9 Hz, 4H), 3.57 (s, 8H), 3.22 (s, 3H), 3.14 (s, 3H), 2.91 (q, <i>J</i> = 6.9 Hz, 3H), 2.38 – 2.25 (m, 2H), 2.10 – 1.75 (m, 8H), 1.54 (s, 4H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [<i>M</i>+<i>H</i>]⁺ = 888.43
UB-180637 (137)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 10.08 (s, 1H), 9.23 (s, 2H), 9.01 (s, 2H), 8.81 (t, <i>J</i> = 5.8 Hz, 1H), 7.95 (s, 1H), 7.84 (dd, <i>J</i> = 7.3, 1.8 Hz, 2H), 7.61 (d, <i>J</i> = 1.8 Hz, 1H), 7.58 – 7.46 (m, 4H), 7.42 (dd, <i>J</i> = 8.5, 1.8 Hz, 1H), 7.34 (d, <i>J</i> = 8.3 Hz, 2H), 5.26 – 5.05 (m, 2H), 4.60 (d, <i>J</i> = 15.6 Hz, 1H), 4.46 – 4.36 (m, 2H), 3.92 (s, 3H), 3.61 (m, 2H), 3.25 (s, 3H), 3.12 (qd, <i>J</i> = 7.3, 4.2 Hz, 4H), 3.00 – 2.91 (m, 4H), 1.99 (m, 4H), 1.95 – 1.81 (m, 4H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [<i>M</i>+<i>H</i>]⁺ = 909.2
UB-180638 (138)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 9.94 (s, 1H), 8.73 (m, 4H), 8.02 (s, 1H), 7.91 (d, <i>J</i> = 2.8 Hz, 1H), 7.83 (dd, <i>J</i> = 7.1, 1.9 Hz, 1H), 7.59 – 7.39 (m, 6H), 7.38 – 7.29 (m, 2H), 5.28 – 5.04 (m, 2H), 4.65 – 4.22 (m, 4H), 3.92 (s, 3H), 3.64 – 3.55 (m, 2H), 3.26 (s, 3H), 3.13 (qt, <i>J</i> = 7.3, 3.8 Hz, 2H), 3.01 – 2.90 (m, 5H), 2.43 (m, 2H), 2.09 – 1.98 (m, 1H), 1.88 (m, 4H), 1.68 (s, 4H), 0.74 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [<i>M</i>+<i>H</i>]⁺ = 926.2
UB-180639 (139)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 9.86 (s, 1H), 8.70 (d, <i>J</i> = 28.6 Hz, 4H), 7.91 (s, 1H), 7.82 (dd, <i>J</i> = 7.2, 1.8 Hz, 2H), 7.59 (s, 1H), 7.58 – 7.51 (m, 2H), 7.53 – 7.43 (m, 2H), 7.41 (dd, <i>J</i> = 8.6, 1.8 Hz, 1H), 7.34 (d, <i>J</i> = 8.5 Hz, 2H), 5.25 – 5.07 (m, 2H), 4.61 (d, <i>J</i> = 15.6 Hz, 1H), 4.48 – 4.25 (m, 3H), 3.92 (s, 3H), 3.40 – 3.30 (m, 2H), 3.25 (s, 3H), 2.90 (m, 5H), 2.42 – 2.24 (m, 4H), 2.03 (m, 1H), 1.88 (m, 4H), 1.73 – 1.55 (m, 4H), 1.45 – 1.26 (m, 2H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [<i>M</i>+<i>H</i>]⁺ = 938.4
UB-180640 (140)	

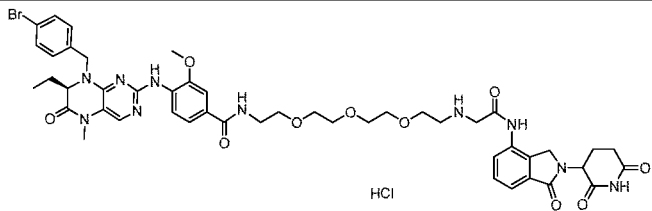
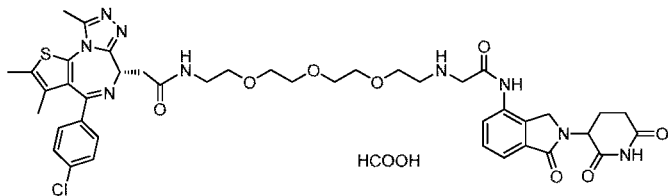
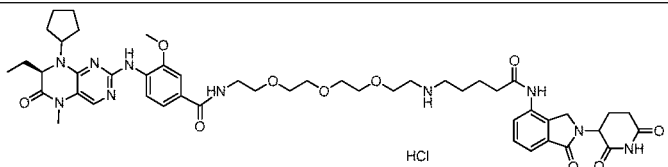
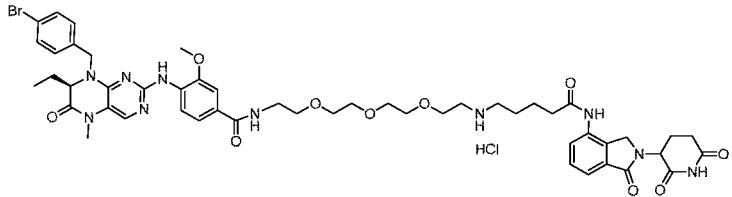
	¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 10.07 (s, 1H), 8.93 (s, 2H), 8.46 (t, <i>J</i> = 5.8 Hz, 1H), 7.85 (dd, <i>J</i> = 7.2, 1.8 Hz, 1H), 7.64 – 7.32 (m, 6H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.55 (t, <i>J</i> = 7.2 Hz, 1H), 4.50 – 4.28 (m, 2H), 3.67 – 3.57 (m, 2H), 3.27 (d, <i>J</i> = 7.3 Hz, 2H), 3.20 (q, <i>J</i> = 6.6 Hz, 2H), 3.11 (td, <i>J</i> = 7.4, 4.3 Hz, 1H), 2.90 (dd, <i>J</i> = 18.0, 6.2 Hz, 5H), 2.62 (s, 3H), 2.40 (s, 3H), 2.32 (t, <i>J</i> = 13.9 Hz, 1H), 1.99 (dt, <i>J</i> = 32.7, 7.1 Hz, 3H), 1.83 (t, <i>J</i> = 8.2 Hz, 2H), 1.62 (s, 3H). LCMS [M+H]⁺ = 784.5
UB-180641 (141)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 9.95 (s, 1H), 8.76 (s, 2H), 8.46 (t, <i>J</i> = 5.8 Hz, 1H), 7.84 (dd, <i>J</i> = 7.0, 2.0 Hz, 1H), 7.61 – 7.31 (m, 6H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.54 (t, <i>J</i> = 7.3 Hz, 1H), 4.40 (q, <i>J</i> = 17.5 Hz, 2H), 3.63 – 3.57 (m, 1H), 3.35 – 3.05 (m, 5H), 2.98 – 2.78 (m, 5H), 2.61 (s, 3H), 2.43 (m, 2H), 2.40 (s, 3H), 2.35 (dd, <i>J</i> = 13.3, 4.2 Hz, 1H), 2.10 – 1.96 (m, 1H), 1.82 (d, <i>J</i> = 10.3 Hz, 2H), 1.72 – 1.62 (m, 3H), 1.62 (s, 3H). LCMS [M+H]⁺ = 798.5
UB-180642 (142)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 9.89 (s, 1H), 8.72 (s, 2H), 8.46 (t, <i>J</i> = 5.9 Hz, 1H), 7.83 (dd, <i>J</i> = 7.0, 2.0 Hz, 1H), 7.56 – 7.36 (m, 6H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.53 (t, <i>J</i> = 7.2 Hz, 1H), 4.43 – 4.30 (m, 2H), 3.30 – 3.24 (m, 2H), 3.25 – 3.17 (m, 1H), 3.17 (s, 2H), 3.12 (dd, <i>J</i> = 7.4, 4.2 Hz, 1H), 2.90 (qd, <i>J</i> = 15.3, 14.0, 8.9 Hz, 4H), 2.61 (d, <i>J</i> = 1.1 Hz, 3H), 2.40 (s, 3H), 2.38 (d, <i>J</i> = 7.6 Hz, 2H), 2.09 – 1.94 (m, 1H), 1.89 – 1.73 (m, 2H), 1.62 (m, 6H), 1.35 (d, <i>J</i> = 7.2 Hz, 2H). LCMS [M+H]⁺ = 812.0
UB-180643 (143)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 9.85 (s, 1H), 8.76 (m, 3H), 7.92 (m, 1H), 7.84-7.80 (m, 1H), 7.65 (d, <i>J</i> = 1.8 Hz, 1H), 7.57-7.51 (m, 2H), 7.50-7.43 (m, 2H), 7.35 (m, 2H), 5.15 (m, 2H), 4.62 (m, 1H), 4.44-4.34 (m, 2H), 3.93 (s, 3H), 3.72 (d, <i>J</i> = 5.7 Hz, 2H), 3.61-3.58 (m, 3H), 3.49 (m, 2H), 3.24 (d, <i>J</i> = 4.6 Hz, 3H), 3.10 (m, 2H), 2.97 (m, 1H), 2.89 (s, 3H), 2.68-2.59 (m, 1H), 2.35 (m, 2H), 2.22 (m, 1H), 1.89 (m, 2H), 1.62 (m, 2H), 1.60 (m, 2H), 1.57 (m, 1H), 1.46 (m, 1H), 1.31 (t, <i>J</i> = 6.4 Hz, 1H), 1.24 (m, 1H), 0.96 (t, <i>J</i> = 7.1 Hz, 1H), 0.75 (q, <i>J</i> = 7.0 Hz, 3H). LCMS [M+H]⁺ = 967.3
UB-180644 (144)	

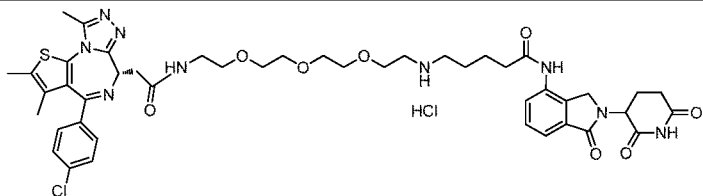
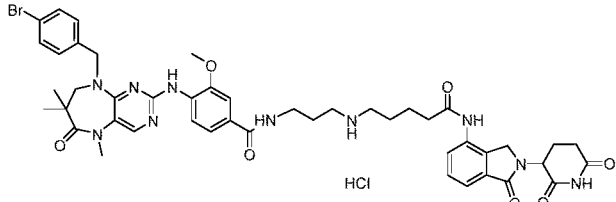
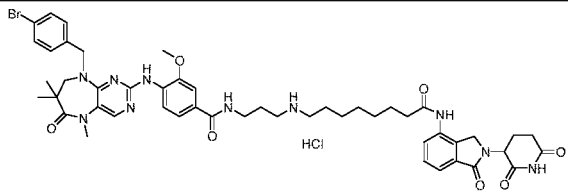
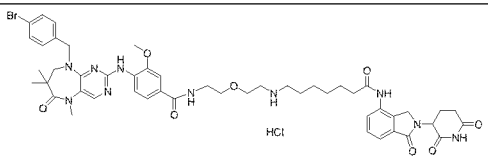
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 9.93 (s, 1H), 9.69 (s, 1H), 8.82 (s, 2H), 8.62 (s, 1H), 7.96 (s, 1H), 7.83 (m, 1H), 7.75 (d, <i>J</i> = 8.4 Hz, 1H), 7.59 (d, <i>J</i> = 1.8 Hz, 1H), 7.56-7.52 (m, 2H), 7.49 (s, 1H), 7.40 -7.32 (m, 3H), 5.20-5.10 (m, 2H), 4.65 (m, 1H), 4.47-4.32 (m, 3H), 3.91 (m, 3H), 3.55 (m, 5H), 3.43 (m, 2H), 3.24 (s, 3H), 3.06 (t, <i>J</i> = 5.6 Hz, 4H), 3.01-2.86 (m, 5H), 2.68-2.58 (m, 2H), 2.39 (m, 2H), 2.33-2.24 (m, 1H), 2.08-1.87 (m, 4H), 1.71-1.58 (m, 4H), 1.49 (m, 1H), 1.36 (m, 2H), 0.97 (m, 1H), 0.75 (t, <i>J</i> = 7.2 Hz, 3H). LCMS [M+H] ⁺ = 1057.1
UB-180645 (145)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 9.95 (s, 1H), 8.90-8.71 (m, 3H), 7.93 (s, 1H), 7.91 - 7.67 (m, 2H), 7.67 (d, <i>J</i> = 1.5 Hz, 1H), 7.49 (ddd, <i>J</i> = 25.5, 18.0, 8.5 Hz, 5H), 7.35 (d, <i>J</i> = 8.4 Hz, 2H), 5.27 - 5.06 (m, 2H), 4.63 (d, <i>J</i> = 15.7 Hz, 1H), 4.41 (dt, <i>J</i> = 35.1, 12.2 Hz, 3H), 3.97 (s, 1H), 4.12 - 3.48 (m, 9H), 3.24 (s, 3H), 2.90 (dd, <i>J</i> = 17.1, 5.4 Hz, 2H), 2.77 - 2.57 (m, 1H), 2.36 (dt, <i>J</i> = 22.0, 10.0 Hz, 3H), 2.03 (d, <i>J</i> = 5.4 Hz, 1H), 1.87 - 1.62 (m, 6H), 1.35 - 1.17 (m, 2H), 0.88 - 0.55 (m, 3H). LCMS [M+H] ⁺ = 954.3
UB-180647 (147)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 10.06 (s, 1H), 8.89 (s, 2H), 8.59 (t, <i>J</i> = 5.6 Hz, 1H), 7.94 (s, 1H), 7.84 (dd, <i>J</i> = 7.3, 1.8 Hz, 2H), 7.59 - 7.45 (m, 5H), 7.39 (dd, <i>J</i> = 8.5, 1.9 Hz, 1H), 7.34 (d, <i>J</i> = 8.2 Hz, 2H), 5.20 - 5.10 (m, 2H), 4.61 (d, <i>J</i> = 15.6 Hz, 1H), 4.49 - 4.32 (m, 3H), 3.91 (s, 3H), 3.69 (t, <i>J</i> = 5.2 Hz, 3H), 3.57 (s, 11H), 3.24 (s, 3H), 3.15 - 3.05 (m, 3H), 3.01 - 2.88 (m, 4H), 2.09 - 1.79 (m, 6H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 927.5
UB-180648 (148)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.00 (s, 1H), 10.25 (d, <i>J</i> = 3.6 Hz, 1H), 8.97 (s, 2H), 8.30 (s, 1H), 7.82 (dd, <i>J</i> = 7.2, 1.8 Hz, 1H), 7.52 - 7.45 (m, 4H), 7.41 (d, <i>J</i> = 8.4 Hz, 2H), 5.13 (d, <i>J</i> = 8.1 Hz, 1H), 4.53 (d, <i>J</i> = 6.9 Hz, 1H), 4.49 - 4.30 (m, 2H), 3.54 (d, <i>J</i> = 14.8 Hz, 12H), 3.43 (t, <i>J</i> = 5.9 Hz, 3H), 3.31 - 3.16 (m, 7H), 3.12 (q, <i>J</i> = 5.4 Hz, 3H), 2.90 (q, <i>J</i> = 7.4, 6.8 Hz, 3H), 2.61 (s, 3H), 2.39 (s, 3H), 2.36 - 2.10 (m, 2H), 1.99 (t, <i>J</i> = 16.8, 9.0, 8.1, 5.2 Hz, 2H). LCMS [M+H] ⁺ = 902.3
UB-180649 (149)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 9.88 (s, 1H), 8.76-8.57 (m, 3H), 7.90 (m, 1H), 7.82 (s, 1H), 7.61 (m, 1H), 7.54 (m, 1H), 7.52-7.46 (m, 1H), 5.15 (m, 1H), 4.47 (m, 1H), 4.44-4.30 (m, 2H), 4.19 (m, 1H), 3.91 (m, 3H), 3.68 (m, 4H), 3.55 (m, 10H), 3.43 (m, 4H), 3.22 (s, 3H), 3.08 (m, 2H), 2.91 (m, 3H), 2.39 (m, 2H), 1.93 (m, 4H), 1.87-1.78 (m, 3H), 1.62 (m, 4H), 1.56-1.44 (m, 4H), 1.36 (m, 2H), 0.97 (m, 1H), 0.75 (t, <i>J</i> = 8.0 Hz, 3H). LCMS [M+H] ⁺ = 955.7

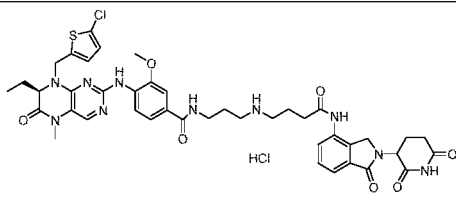
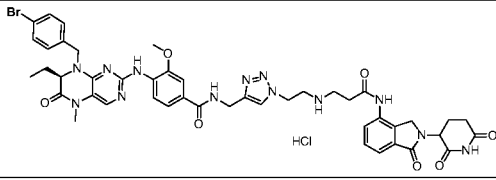
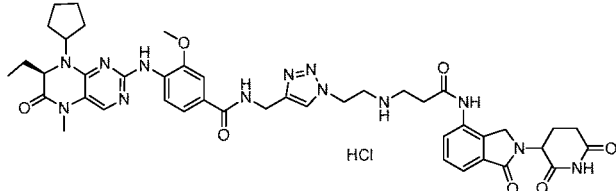
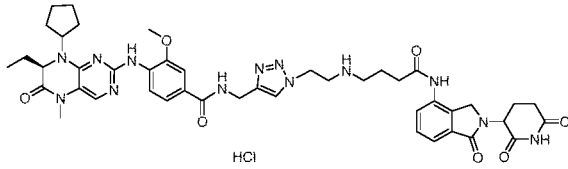
UB-180650 (150)	 HCl
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 9.80 (s, 1H), 8.32 (d, 2H), 7.81 (m, 1H), 7.48 (m, 3H), 7.42 (m, 2H), 5.14 (m, 1H), 4.51 (m, 1H), 4.46-4.27 (m, 2H), 3.56-3.54 (m, 2H), 3.45 (m, 4H), 3.27 (m, 4H), 2.97 (m, 3H), 2.82 (m, 3H), 2.67 (m, 3H), 2.59 (m, 3H), 2.41 (m, 3H), 2.34 (m, 2H), 2.24 (m, 1H), 2.07- 1.96 (m, 1H), 1.62 (m, 4H), 1.54-1.41 (m, 3H), 1.32 (m, 2H), 0.96 (m, 1H). LCMS [M+H] ⁺ =842.3
UB-180651 (151)	 HCOOH
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 9.80 (s, 1H), 8.30 (m, 2H), 7.81 (m, 1H), 7.45 (m, 6H), 5.14 (m, 1H), 4.50 (m, 1H), 4.43-4.27 (m, 2H), 3.51 (m, 4H), 3.46 (m, 4H), 3.25 (m, 2H), 2.92 (m, 2H), 2.78 (d, <i>J</i> = 5.8 Hz, 2H), 2.66 – 2.61 (m, 4H), 2.59 (s, 3H), 2.41 (s, 3H), 2.38-2.31 (m, 3H), 2.06-1.93 (m, 2H), 1.62 (m, 5H), 1.49 (m, 2H), 1.37-1.30 (m, 2H).LCMS [M+H] ⁺ =930.4
UB-180652 (152)	 HCl
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 9.89 (d, <i>J</i> = 10.9 Hz, 1H), 9.21 – 8.73 (m, 5H), 8.15 – 7.84 (m, 3H), 7.82 (d, <i>J</i> = 6.9 Hz, 1H), 7.66 (s, 1H), 7.61 – 7.41 (m, 5H), 7.35 (d, <i>J</i> = 8.2 Hz, 2H), 5.28 – 5.05 (m, 2H), 4.54- 4.10 (m, 4H), 3.93 (s, 3H), 3.93 – 3.37 (m, 10H), 3.24 (s, 3H), 3.18 – 2.79 (m, 3H), 2.78 – 2.56 (m, 4H), 2.38 – 2.29 (m, 38H), 2.11 (s, 1H), 2.29 – 1.50 (m, 3H), 1.54 – 1.50 (m, 1H), 1.42 (d, <i>J</i> = 7.1 Hz, 2H), 0.88 – 0.19 (m, 3H). LCMS [M+H] ⁺ =996.3
UB-180653 (153)	 HCl
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 9.84 (s, 1H), 8.73 (m, 3H), 7.91 (m, 1H), 7.82 (m, 2H), 7.64 (m, 1H), 7.54 (m, 2H), 7.52-7.42 (m, 3H), 7.35 (m, 2H), 5.18-5.11 (m, 2H), 4.62 (m, 1H), 4.44-4.31 (m, 3H), 3.93 (s, 3H), 3.71 (m, 2H), 3.62-3.58 (m, 3H), 3.48 (m, 4H), 3.23 (m, 3H), 3.11 (m, 3H), 2.88 (m, 3H), 2.63 (m, 2H), 2.35 (t, 3H), 2.05-2.00 (m, 1H), 1.88 (m, 2H), 1.60 (m, 4H), 0.74 (t, 3H).LCMS [M+H] ⁺ =980.8
UB-180654 (154)	 HCOOH

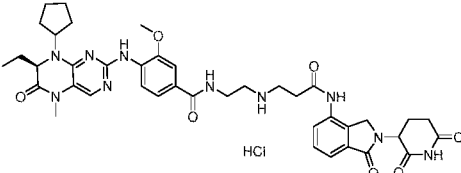
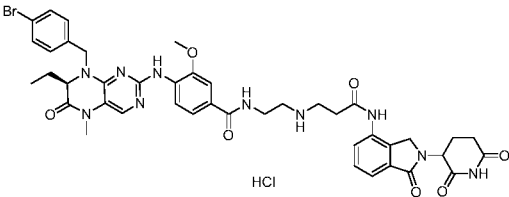
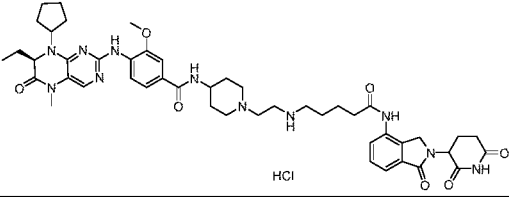
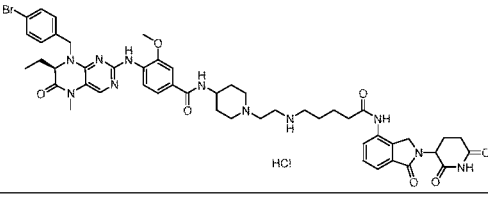
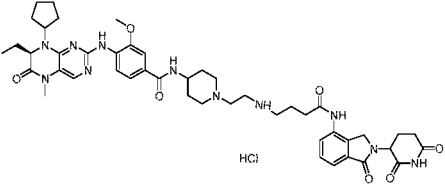
	^1H NMR (400 MHz, DMSO-d_6) δ 11.02 (s, 1H), 9.86 (s, 1H), 8.80 (m, 2H), 8.44 (m, 1H), 7.82 (m, 1H), 7.54-7.41 (m, 6H), 5.15 (m, 1H), 4.55 (m, 1H), 4.45-4.31 (m, 2H), 3.70 (m, 2H), 3.51 (m, 2H), 3.36- 3.28 (m, 4H), 3.16-3.10 (m, 2H), 2.97-2.86 (m, 3H), 2.62 (m, 4H), 2.41 (s, 3H), 2.34 (m, 3H), 2.02 (m, 1H), 1.62 (m, 5H), 1.59 (m, 2H), 1.30 (m, 4H). LCMS $[\text{M}+\text{H}]^+ = 856.4$
UB-180655 (155)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.02 (s, 1H), 9.81 (s, 1H), 8.69 (m, 2H), 7.97 (m, 1H), 7.85-7.78 (m, 2H), 7.65 (m, 1H), 7.58 (m, 1H), 7.52-7.43 (m, 2H), 5.19-5.12 (m, 1H), 4.44 (m, 1H), 4.41-4.29 (m, 2H), 4.20 (m, 1H), 3.93 (s, 3H), 3.71 (m, 2H), 3.61 (m, 2H), 3.50 (m, 3H), 3.21 (s, 3H), 3.12 (m, 3H), 2.90 (m, 3H), 2.68-2.59 (m, 2H), 2.36 (m, 3H), 2.00 (m, 4H), 1.82 (m, 3H), 1.65-1.57 (m, 5H), 1.49 (m, 2H), 1.29 (m, 4H), 0.75 (t, $J = 7.4$ Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 881.6$.
UB-180656 (156)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.02 (s, 1H), 10.08 (s, 1H), 9.06 (s, 2H), 8.49 (t, $J = 5.4$ Hz, 1H), 7.80 (t, $J = 29.2$ Hz, 1H), 7.61 – 7.36 (m, 6H), 5.25-5.10 (m, 1H), 4.41 (d, $J = 21.5$ Hz, 3H), 3.83 – 3.57 (m, 4H), 3.57 – 3.29 (m, 4H), 3.29 – 3.22 (m, 2H), 3.07– 2.98 (m, 3H), 3.01 – 2.69 (m, 11H), 3.01 – 2.26 (m, 137H), 2.04 -1.96 (m, 3H), 1.62 (s, 3H), 1.37 – 1.17 (m, 4H). LCMS $[\text{M}+\text{H}]^+ = 814.3$
UB-180657 (157)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.02 (s, 1H), 9.93 (s, 1H), 8.90 (s, 1H), 8.77 (s, 1H), 7.97 – 7.82 (m, 3H), 7.69 (s, 2H), 7.69 (s, 1H), 7.69 – 7.39 (m, 4H), 5.15 (dd, $J = 13.2, 5.1$ Hz, 1H), 4.39 -4.16 (m, 4H), 3.92 (m, 3H), 3.72 – 3.57 (m, 4H), 3.24 (s, 3H), 3.13– 2.76 (m, 5H), 2.09 – 1.49 (m, 16H), 0.88 – 0.09 (m, 3H). LCMS $[\text{M}+\text{H}]^+ = 854.4$
UB-180658 (158)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.02 (s, 1H), 9.94 (s, 1H), 9.39 (s, 1H), 8.80 (d, $J = 8.0$ Hz, 3H), 7.92 (d, $J = 8.1$ Hz, 1H), 7.90 – 7.76 (m, 2H), 7.64 (d, $J = 1.9$ Hz, 1H), 7.60 – 7.44 (m, 3H), 5.15 (dd, $J = 13.2, 5.2$ Hz, 1H), 4.56 – 4.29 (m, 3H), 3.92 (s, 3H), 3.22 (s, 3H), 3.06 – 2.78 (m, 5H), 2.70 – 2.56 (m, 2H), 2.44 (d, $J = 6.7$ Hz, 2H), 2.39 – 2.26 (m, 2H), 1.91 (m, 10H), 1.75 – 1.63 (m, 4H), 1.58 – 1.39 (m, 4H), 0.75 (t, $J = 7.4$ Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 823.4$

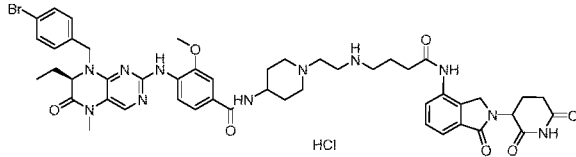
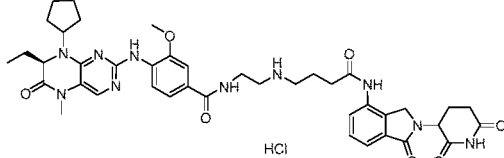
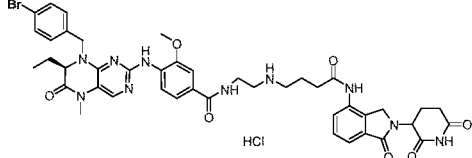
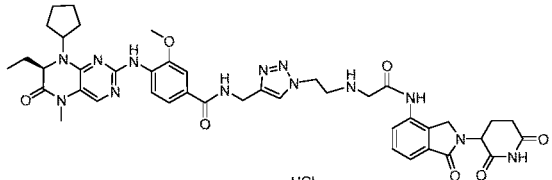
UB-180659 (159)	
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 9.83 (s, 1H), 8.52 (s, 1H), 8.42 (d, <i>J</i> = 8.3 Hz, 1H), 8.34 (s, 1H), 7.95 (t, <i>J</i> = 4.5 Hz, 1H), 7.84 (s, 1H), 7.60 (s, 1H), 7.54 – 7.42 (m, 4H), 7.22 (s, 1H), 5.14 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.46 – 4.27 (m, 3H), 4.24 (dd, <i>J</i> = 7.6, 3.5 Hz, 1H), 3.93 (s, 3H), 3.24 (s, 3H), 3.05 – 2.83 (m, 2H), 2.72 (td, <i>J</i> = 15.4, 7.6 Hz, 5H), 2.44 – 2.29 (m, 3H), 2.08 – 1.48 (m, 20H), 1.36 (d, <i>J</i> = 7.3 Hz, 2H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 837.4
UB-180660 (160)	
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 9.86 (s, 1H), 8.53 – 8.35 (m, 2H), 8.29 (s, 1H), 7.83 (d, <i>J</i> = 8.0 Hz, 2H), 7.60 (s, 1H), 7.48 (td, <i>J</i> = 7.7, 7.3, 1.8 Hz, 4H), 5.32 (t, <i>J</i> = 4.8 Hz, 1H), 5.14 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.45 – 4.28 (m, 3H), 4.24 (dd, <i>J</i> = 7.6, 3.5 Hz, 1H), 3.93 (s, 3H), 3.25 (s, 3H), 2.79 – 2.64 (m, 5H), 2.33 (dd, <i>J</i> = 3.7, 1.8 Hz, 1H), 2.00 (q, <i>J</i> = 6.9, 6.4 Hz, 4H), 1.91 – 1.55 (m, 15H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 809.5
UB-180661 (161)	
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 9.98 (s, 1H), 8.96 (s, 2H), 8.49 (s, 1H), 7.83 (d, <i>J</i> = 6.7 Hz, 2H), 7.59 – 7.28 (m, 4H), 5.63 – 4.52 (m, 1H), 4.07-4.21 (m, 4H), 3.93 (s, 3H), 3.87 – 3.57 (m, 4H), 3.63 – 3.24 (m, 4H), 3.21 – 2.88 (m, 9H), 2.86 – 2.74 (m, 4H), 2.03 – 1.78 (m, 9H), 1.54 – 1.15 (m, 5H), 1.38 – 0.99 (m, 3H). LCMS [M+H] ⁺ = 828.3
UB-180662 (162)	
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 10.03 (d, <i>J</i> = 3.1 Hz, 1H), 9.65 (s, 1H), 9.05 (s, 2H), 8.84 (t, <i>J</i> = 5.3 Hz, 1H), 7.85 (dd, <i>J</i> = 8.6, 4.0 Hz, 3H), 7.72 (d, <i>J</i> = 1.4 Hz, 1H), 7.62 (d, <i>J</i> = 8.3 Hz, 1H), 7.61 – 7.28 (m, 2H), 5.14 (dd, <i>J</i> = 13.2, 5.1 Hz, 1H), 4.64 – 4.30 (m, 3H), 4.37 (t, <i>J</i> = 16.4 Hz, 1H), 3.93 (s, 3H), 3.89-3.41 (m, 7H), 3.16 – 2.92 (m, 4H), 2.27 – 1.48 (m, 16H), 0.78 (t, <i>J</i> = 7.2 Hz, 3H). LCMS [M+H] ⁺ = 839.4
UB-180663 (163)	

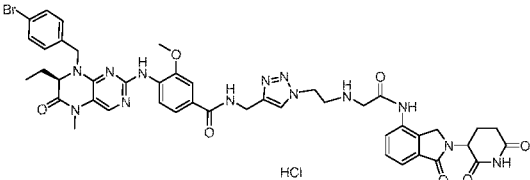
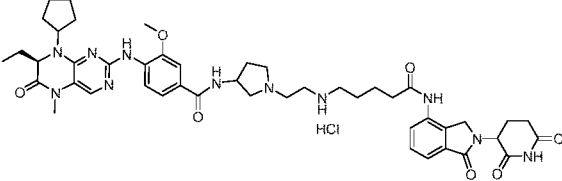
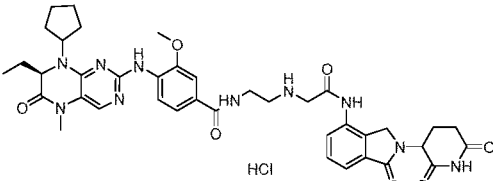
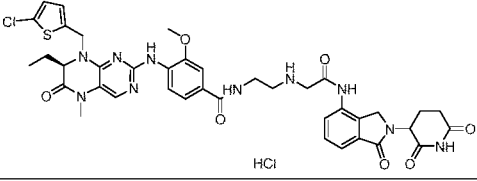
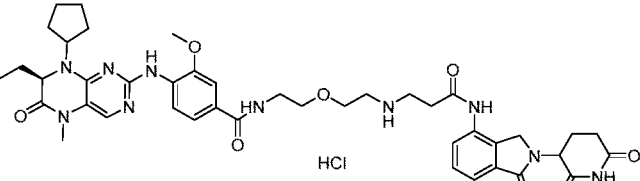
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 10.04 (d, J = 15.1 Hz, 1H), 9.76 (s, 1H), 8.91 (s, 3H), 8.69 (t, J = 5.6 Hz, 1H), 7.85 (dt, J = 8.0, 4.6 Hz, 3H), 7.64 (d, J = 1.9 Hz, 1H), 7.57 – 7.47 (m, 3H), 5.15 (dd, J = 13.3, 5.1 Hz, 1H), 4.50 (dd, J = 6.4, 3.3 Hz, 1H), 4.48 – 4.38 (m, 2H), 3.90 (s, 3H), 3.70 (h, J = 4.7 Hz, 4H), 3.43 (q, J = 5.9 Hz, 2H), 3.21 (s, 3H), 3.07 (d, J = 7.6 Hz, 2H), 2.97 – 2.89 (m, 3H), 2.61 (d, J = 18.2 Hz, 3H), 2.46 – 2.40 (m, 2H), 2.05 – 2.00 (m, 1H), 1.97 – 1.90 (m, 2H), 1.82 (q, J = 7.1 Hz, 3H), 1.71 – 1.63 (m, 4H), 1.49 (d, J = 16.1 Hz, 3H), 0.75 (t, J = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 899.4
UB-180664 (164)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 10.00 (d, J = 10.0 Hz, 1H), 9.61 (s, 1H), 8.84 (s, 2H), 8.61 (t, J = 5.5 Hz, 1H), 7.95 (d, J = 3.6 Hz, 1H), 7.89 – 7.74 (m, 2H), 7.58 (d, J = 1.9 Hz, 1H), 7.55 – 7.44 (m, 4H), 7.40 – 7.31 (m, 3H), 5.22 – 5.10 (m, 2H), 4.63 (d, J = 15.4 Hz, 1H), 4.51 – 4.31 (m, 3H), 3.90 (s, 3H), 3.70 (d, J = 6.3 Hz, 2H), 3.24 (s, 4H), 3.13 – 3.00 (m, 3H), 2.99 – 2.83 (m, 4H), 2.63 (s, 1H), 2.38 (d, J = 15.0 Hz, 1H), 2.14 – 1.97 (m, 2H), 1.95 – 1.85 (m, 2H), 1.67 (s, 5H), 0.74 (t, J = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 999.31
UB-180665 (165)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 8.26 (t, J = 5.7 Hz, 1H), 7.87 (dd, J = 7.1, 1.9 Hz, 1H), 7.56 – 7.40 (m, 7H), 5.15 (dd, J = 13.3, 5.1 Hz, 1H), 4.50 (dd, J = 7.9, 6.2 Hz, 1H), 4.45 – 4.31 (m, 2H), 3.53 (d, J = 9.7 Hz, 10H), 3.44 (d, J = 11.8 Hz, 2H), 3.27 – 3.17 (m, 4H), 3.03 – 2.81 (m, 4H), 2.59 (s, 4H), 2.46 (dd, J = 4.6, 2.8 Hz, 1H), 2.41 (s, 3H), 2.37 – 2.27 (m, 2H), 2.03 (d, J = 12.6 Hz, 1H), 1.62 (s, 3H). LCMS [M+H] ⁺ = 874.20
UB-180666 (166)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 10.04 (d, J = 15.1 Hz, 1H), 9.76 (s, 1H), 8.91 (s, 3H), 8.69 (t, J = 5.6 Hz, 1H), 7.85 (dt, J = 8.0, 4.6 Hz, 3H), 7.64 (d, J = 1.9 Hz, 1H), 7.58 – 7.43 (m, 3H), 5.15 (dd, J = 13.3, 5.1 Hz, 1H), 4.52 – 4.47 (m, 1H), 4.44 – 4.32 (m, 2H), 3.90 (s, 3H), 3.70 (dt, J = 10.6, 5.0 Hz, 3H), 3.48 – 3.38 (m, 2H), 3.21 (s, 3H), 3.07 (d, J = 7.6 Hz, 3H), 3.00 – 2.89 (m, 3H), 2.70 – 2.55 (m, 4H), 2.45 – 2.36 (m, 2H), 2.14 – 1.98 (m, 2H), 1.98 – 1.75 (m, 6H), 1.72 – 1.60 (m, 4H), 1.48 (d, J = 22.8 Hz, 4H), 0.75 (t, J = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 941.30
UB-180667 (167)	

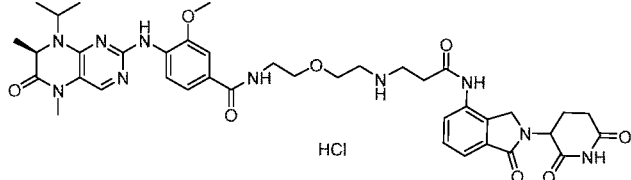
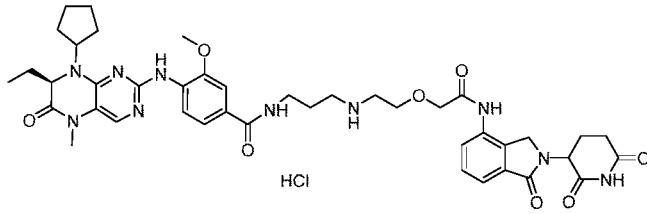
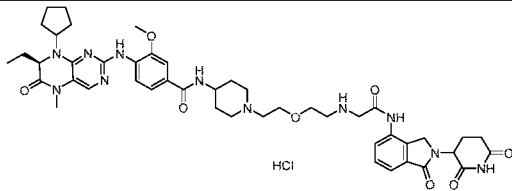
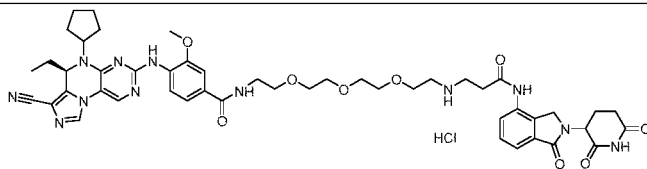
	¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 10.00 (d, <i>J</i> = 10.0 Hz, 1H), 9.61 (s, 1H), 8.84 (s, 2H), 8.61 (t, <i>J</i> = 5.5 Hz, 1H), 7.96 (d, <i>J</i> = 3.6 Hz, 1H), 7.87–7.74 (m, 2H), 7.58 (d, <i>J</i> = 1.9 Hz, 1H), 7.56–7.46 (m, 4H), 7.40–7.32 (m, 3H), 5.23–5.06 (m, 2H), 4.64 (d, <i>J</i> = 15.4 Hz, 1H), 4.51–4.31 (m, 3H), 3.91 (s, 3H), 3.68 (t, <i>J</i> = 5.3 Hz, 2H), 3.57 (d, <i>J</i> = 6.8 Hz, 10H), 3.24 (s, 3H), 3.12–3.01 (m, 3H), 3.00–2.85 (m, 4H), 2.61 (d, <i>J</i> = 16.8 Hz, 2H), 2.44–2.35 (m, 2H), 2.16–1.97 (m, 2H), 1.96–1.85 (m, 2H), 1.67 (s, 4H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 1040.96
UB-180668 (168)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 10.03 (s, 1H), 8.88 (d, <i>J</i> = 33.7 Hz, 2H), 8.34 (t, <i>J</i> = 5.7 Hz, 1H), 7.83 (dd, <i>J</i> = 7.0, 2.0 Hz, 1H), 7.59–7.37 (m, 7H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.56 (dd, <i>J</i> = 7.8, 6.3 Hz, 1H), 4.51–4.31 (m, 2H), 3.70 (q, <i>J</i> = 5.3 Hz, 3H), 3.60–3.52 (m, 10H), 3.46 (t, <i>J</i> = 5.9 Hz, 2H), 3.31–3.20 (m, 4H), 3.10–3.05 (m, 2H), 2.98–2.88 (m, 3H), 2.65 (d, <i>J</i> = 1.4 Hz, 3H), 2.42 (d, <i>J</i> = 1.9 Hz, 5H), 2.07–1.98 (m, 1H), 1.62 (s, 3H). LCMS [M+H]⁺ = 915.99
UB-180670 (170)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 9.98 (s, 1H), 8.66–8.40 (m, 4H), 8.10 (s, 1H), 7.83 (d, <i>J</i> = 7.0 Hz, 1H), 7.64–7.48 (m, 5H), 7.46–7.35 (m, 3H), 5.15 (dd, <i>J</i> = 13.3, 5.2 Hz, 1H), 4.95 (s, 2H), 4.39 (dd, <i>J</i> = 15.8, 8.7 Hz, 2H), 3.92 (s, 3H), 3.67–3.56 (m, 4H), 3.25–3.19 (m, 4H), 2.93–2.78 (m, 4H), 2.56–2.45 (m, 1H), 1.88 (s, 1H), 1.05 (s, 6H). LCMS [M+H]⁺ = 937.3
UB-180671 (171)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 9.98 (s, 1H), 9.65 (s, 1H), 9.05 (s, 2H), 8.84 (d, <i>J</i> = 5.3 Hz, 2H), 8.15 (s, 1H), 7.85 (dd, <i>J</i> = 8.6, 4.0 Hz, 1H), 7.72 (d, <i>J</i> = 1.4 Hz, 1H), 7.62 (d, <i>J</i> = 8.3 Hz, 1H), 7.61–7.28 (m, 2H), 5.14 (dd, <i>J</i> = 13.2, 5.1 Hz, 1H), 4.98 (s, 2H), 4.45–4.30 (m, 2H), 3.98 (s, 3H), 3.63–3.47 (m, 2H), 3.39 (s, 2H), 3.21 (s, 3H), 3.16–2.92 (m, 2H), 2.92–2.66 (m, 6H), 2.57–2.46 (m, 4H), 2.05–1.58 (m, 7H), 0.78 (s, 6H). LCMS [M+H]⁺ = 982.3
UB-180672 (172)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 10.03 (d, <i>J</i> = 3.1 Hz, 1H), 9.65 (s, 9H), 9.05 (s, 2H), 8.84 (t, <i>J</i> = 5.3 Hz, 1H), 8.15 (s, 2H), 7.85 (dd, <i>J</i> = 8.6, 4.0 Hz, 3H), 7.72 (d, <i>J</i> = 1.4 Hz, 1H), 7.62 (d, <i>J</i> = 8.3 Hz, 1H), 7.61–7.28 (m, 2H), 5.14 (dd, <i>J</i> = 13.2, 5.1 Hz, 1H), 4.94–4.30 (m, 3H), 3.98 (m, 3H), 3.79–3.39 (m, 10H), 3.21 (s, 3H), 3.16–2.92 (m, 5H), 2.62–2.56 (m, 1H), 2.47–2.36 (m, 4H), 2.05–1.98 (m, 1H), 1.65–1.56 (m, 4H), 1.23 (s, 4H), 1.08 (s, 6H). LCMS [M+H]⁺ = 996.3

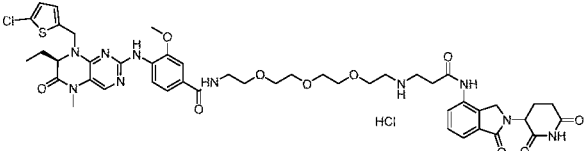
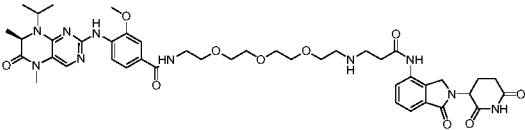
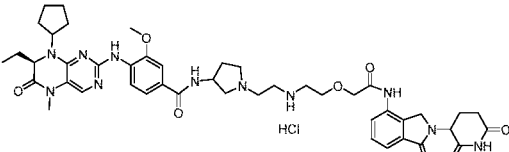
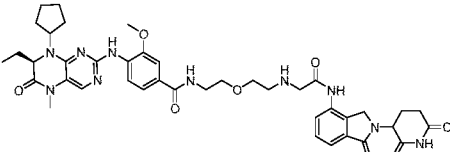
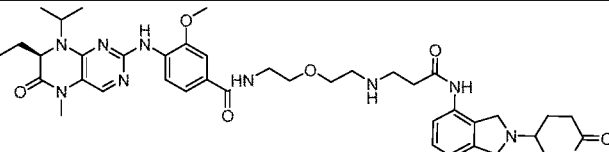
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 10.03 (d, <i>J</i> = 3.1 Hz, 1H), 8.65 (s, 3H), 8.11 (s, 1H), 7.93 (t, <i>J</i> = 5.3 Hz, 1H), 7.85 (dd, <i>J</i> = 8.6, 4.0 Hz, 1H), 7.62 - 7.49 (m, 6H), 7.42 (d, <i>J</i> = 8.3 Hz, 4H), 5.14 (dd, <i>J</i> = 13.2, 5.1 Hz, 1H), 4.94 (s, 2H), 4.43 - 4.10 (m, 2H), 3.98 (s, 3H), 3.63 - 3.51 (m, 4H), 3.21 (s, 3H), 2.92 - 2.86 (m, 4H), 2.66 (s, 1H), 2.47 - 2.36 (m, 2H), 2.06 - 1.80 (m, 7H), 1.18 (s, 6H). LCMS [M+H] ⁺ = 925.3
UB-180678 (178)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 10.02 (s, 1H), 8.11 (d, <i>J</i> = 8.2 Hz, 1H), 7.90 (s, 1H), 7.84 (d, <i>J</i> = 7.2 Hz, 1H), 7.63 (d, <i>J</i> = 1.9 Hz, 1H), 7.51 (td, <i>J</i> = 7.4, 6.3, 3.2 Hz, 3H), 7.11 (d, <i>J</i> = 3.9 Hz, 1H), 6.98 (d, <i>J</i> = 3.8 Hz, 1H), 5.14 (dd, <i>J</i> = 14.0, 7.7 Hz, 2H), 4.79 (d, <i>J</i> = 15.4 Hz, 1H), 4.50 (t, <i>J</i> = 4.7 Hz, 1H), 4.40 (q, <i>J</i> = 17.5 Hz, 2H), 3.95 (s, 3H), 3.22 (s, 3H), 3.04 - 2.93 (m, 5H), 2.69 - 2.52 (m, 2H), 2.33 (h, <i>J</i> = 10.6, 10.1 Hz, 2H), 2.10 - 1.80 (m, 8H), 1.34 - 1.26 (m, 1H), 0.67 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 871.2
UB-180679 (179)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.03 (s, 1H), 10.21 (s, 1H), 9.10 (d, <i>J</i> = 33.5 Hz, 4H), 8.09 (d, <i>J</i> = 7.4 Hz, 1H), 8.00 - 7.82 (m, 3H), 7.60 (d, <i>J</i> = 1.9 Hz, 1H), 7.53 (h, <i>J</i> = 7.3, 6.9 Hz, 4H), 7.43 (dd, <i>J</i> = 8.5, 1.8 Hz, 1H), 7.34 (d, <i>J</i> = 8.2 Hz, 2H), 5.21 - 5.11 (m, 2H), 4.71 (dt, <i>J</i> = 14.4, 6.3 Hz, 2H), 4.63 - 4.37 (m, 6H), 3.91 (s, 3H), 3.25 (s, 3H), 2.90 (d, <i>J</i> = 6.5 Hz, 2H), 2.67 (d, <i>J</i> = 20.1 Hz, 2H), 2.31 (s, 1H), 2.05 (d, <i>J</i> = 5.8 Hz, 1H), 1.89 (d, <i>J</i> = 7.6 Hz, 2H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 963.9
UB-180680 (180)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 10.30 (s, 1H), 9.76 (s, 1H), 9.41 (s, 2H), 9.17 (t, <i>J</i> = 5.7 Hz, 1H), 8.13 (s, 1H), 7.90 - 7.81 (m, 3H), 7.67 (d, <i>J</i> = 1.9 Hz, 1H), 7.60 (dd, <i>J</i> = 8.3, 1.8 Hz, 1H), 7.57 - 7.46 (m, 2H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.76 (t, <i>J</i> = 6.5 Hz, 2H), 4.54 (d, <i>J</i> = 5.5 Hz, 2H), 4.52 - 4.39 (m, 4H), 4.22 - 4.14 (m, 2H), 3.90 (s, 3H), 3.51 (q, <i>J</i> = 6.7, 6.0 Hz, 2H), 3.21 (s, 5H), 3.00 - 2.86 (m, 3H), 2.62 (d, <i>J</i> = 17.5 Hz, 1H), 2.40 - 2.26 (m, 1H), 2.08 - 1.75 (m, 7H), 1.58 - 1.41 (m, 4H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 862.48
UB-180681 (181)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 10.08 (s, 1H), 9.74 (s, 1H), 9.33 (s, 2H), 9.16 (t, <i>J</i> = 5.6 Hz, 1H), 8.12 (s, 1H), 7.93 - 7.80 (m, 3H), 7.67 (d, <i>J</i> = 1.8 Hz, 1H), 7.59 (dd, <i>J</i> = 8.4, 1.8 Hz, 1H), 7.54 - 7.40 (m, 1H), 5.15 (dd, <i>J</i> = 13.2, 5.2 Hz, 1H), 4.75 (t, <i>J</i> = 6.5 Hz, 2H), 4.62 - 4.42 (m, 6H), 4.24 - 4.15 (m, 2H), 3.90 (s, 3H), 3.53 - 3.41 (m, 2H), 3.22 (s, 3H), 3.07 - 2.83 (m, 3H), 2.40 - 2.27 (m, 1H), 2.08 - 1.76 (m, 9H), 1.63 - 1.40 (m, 4H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 876.54

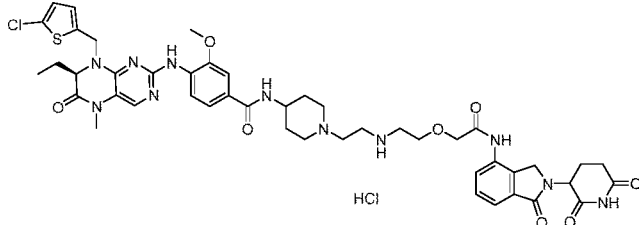
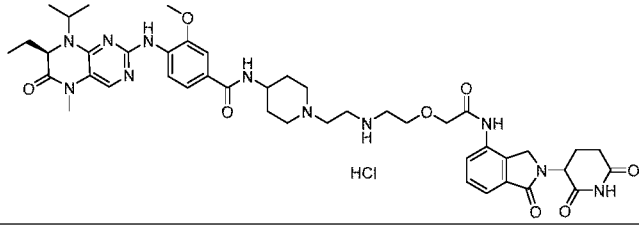
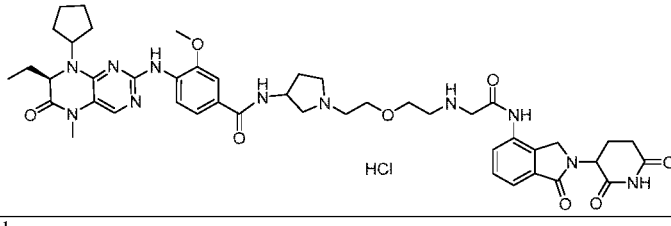
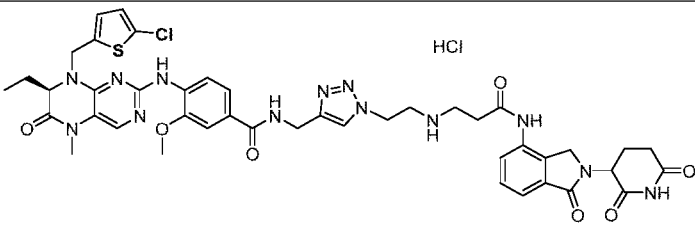
UB-180682 (182)	 ^1H NMR (400 MHz, <i>DMSO-d</i>₆) δ 11.02 (s, 1H), 10.25 (s, 1H), 9.71 (s, 1H), 9.26 (s, 2H), 9.00 (t, <i>J</i> = 5.5 Hz, 1H), 7.92 – 7.80 (m, 3H), 7.75 (d, <i>J</i> = 1.8 Hz, 1H), 7.63 (dd, <i>J</i> = 8.3, 1.8 Hz, 1H), 7.58 – 7.45 (m, 2H), 5.16 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.57 – 4.47 (m, 2H), 4.47 – 4.32 (m, 4H), 4.16 (s, 2H), 3.65 (q, <i>J</i> = 5.7 Hz, 2H), 3.22 (s, 7H), 3.00 – 2.89 (m, 3H), 2.61 (d, <i>J</i> = 17.6 Hz, 1H), 2.40 – 2.25 (m, 1H), 2.10 – 1.72 (m, 7H), 1.61 – 1.38 (m, 4H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 781.48$
UB-180683 (183)	 ^1H NMR (400 MHz, <i>DMSO-d</i>₆) δ 11.02 (s, 1H), 10.24 (s, 1H), 9.64 (s, 1H), 9.19 (s, 2H), 8.94 (t, <i>J</i> = 5.6 Hz, 1H), 7.97 (s, 1H), 7.86 – 7.74 (m, 2H), 7.70 (d, <i>J</i> = 1.9 Hz, 1H), 7.59 – 7.41 (m, 5H), 7.35 (d, <i>J</i> = 8.4 Hz, 2H), 5.22 – 5.08 (m, 2H), 4.65 (d, <i>J</i> = 15.7 Hz, 1H), 4.51 – 4.32 (m, 4H), 3.93 (s, 3H), 3.63 (q, <i>J</i> = 6.0 Hz, 2H), 3.24 (s, 5H), 3.20 – 3.10 (m, 2H), 2.94 (q, <i>J</i> = 6.8, 6.0 Hz, 3H), 2.61 (d, <i>J</i> = 18.1 Hz, 2H), 2.31 (t, <i>J</i> = 13.4 Hz, 1H), 2.07 – 1.85 (m, 3H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 883.38$
UB-180684 (184)	 ^1H NMR (400 MHz, <i>d</i>₆-DMSO) δ 11.02 (s, 1H), 10.81 (s, 1H), 9.97 (s, 1H), 9.28 (s, 2H), 8.56 (m, 1H), 7.92 (s, 1H), 7.82 (m, 2H), 7.60 (m, 2H), 7.55-7.47 (m, 2H), 5.16 (m, 1H), 4.50-4.34 (m, 3H), 4.21 (m, 1H), 4.10 (m, 1H), 3.92 (s, 3H), 3.67 (m, 2H), 3.22 (s, 3H), 3.16 (m, 2H), 2.97 (m, 3H), 2.65 (m, 3H), 2.43-2.31 (m, 2H), 2.12-1.64 (m, 17H), 1.51 (m, 4H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 892.4$
UB-180685 (185)	 ^1H NMR (400 MHz, <i>DMSO-d</i>₆) δ 11.02 (s, 1H), 10.74 (s, 1H), 9.94 (s, 1H), 9.21 (m, 2H), 8.49 (m, 1H), 7.92 (s, 1H), 7.82 (m, 1H), 7.58 – 7.49 (m, 4H), 7.47 – 7.40 (m, 1H), 7.35 (m, 2H), 5.16 (m, 2H), 4.58 (m, 1H), 4.47 – 4.33 (m, 3H), 4.08 (m, 1H), 3.93 (s, 3H), 3.66 (m, 2H), 3.25 (s, 3H), 3.16 (m, 2H), 3.05 – 2.89 (m, 4H), 2.69 – 2.58 (m, 2H), 2.47 – 2.41 (m, 2H), 2.33 (m, 1H), 2.10 – 1.94 (m, 6H), 1.88 (m, 2H), 1.71 (m, 4H), 1.46 (m, 1H), 1.27 (m, 2H), 0.86 (m, 1H), 0.74 (t, <i>J</i> = 7.4 Hz, 3H). LCMS $[\text{M}+\text{H}]^+ = 992.4$
UB-180687 (187)	

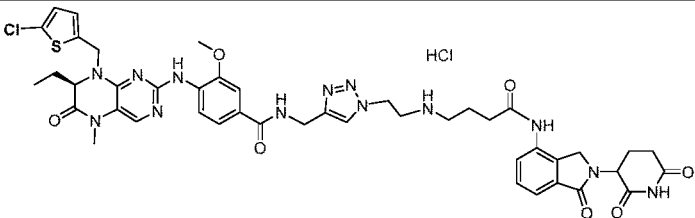
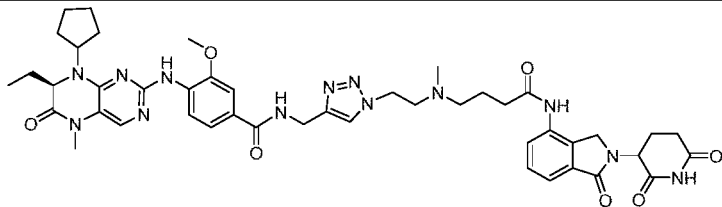
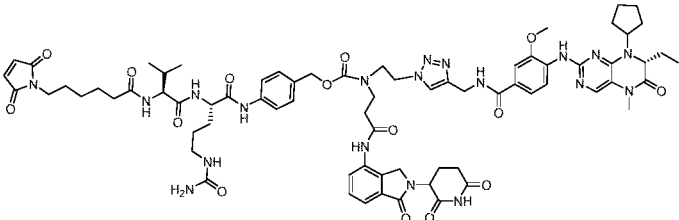
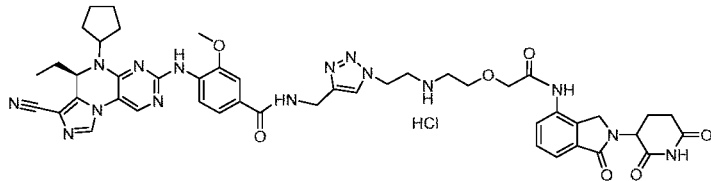
	¹ H NMR (400 MHz, <i>d</i> 6-DMSO) δ 11.03 (s, 1H), 10.73 (s, 1H), 9.99 (s, 1H), 9.23 (m, 2H), 8.53 (s, 1H), 7.88-7.76 (m, 2H), 7.65-7.44 (m, 4H), 5.16 (m, 1H), 4.40 (m, 3H), 4.23 (m, 1H), 4.09 (m, 1H), 3.93 (s, 2H), 3.68 (m, 2H), 3.57 (s, 4H), 3.23 (s, 3H), 3.17 (m, 2H), 3.05 (m, 2H), 2.96-2.87 (m, 1H), 2.70-2.58 (m, 2H), 2.35-2.28 (m, 2H), 2.00 (m, 7H), 1.83 (m, 3H), 1.60 (m, 2H), 1.50 (m, 2H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 879.4
UB-180688 (188)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.03 (s, 1H), 10.71 (s, 1H), 10.00 (s, 1H), 9.24 (m, 2H), 8.49 (d, <i>J</i> = 7.5 Hz, 1H), 7.98 (s, 1H), 7.91 (s, 1H), 7.86-7.80 (m, 1H), 7.59-7.48 (m, 4H), 7.43 (d, <i>J</i> = 8.5 Hz, 1H), 7.35 (m, 2H), 5.24-5.12 (m, 2H), 4.54 (m, 1H), 4.40 (m, 3H), 4.07 (m, 1H), 3.93 (s, 2H), 3.65 (m, 4H), 3.57 (s, 3H), 3.25 (s, 3H), 3.15 (m, 2H), 3.05 (m, 2H), 2.99-2.89 (m, 1H), 2.68-2.62 (m, 2H), 2.38-2.30 (m, 2H), 2.11-1.81 (m, 9H), 1.30-1.24 (m, 1H), 0.74 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 978.3
189 (189)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 10.04 (d, <i>J</i> = 6.0 Hz, 1H), 9.52 (s, 1H), 9.16 (s, 2H), 8.98 (s, 1H), 7.93 (d, <i>J</i> = 7.8 Hz, 1H), 7.87 – 7.80 (m, 2H), 7.74 (d, <i>J</i> = 2.2 Hz, 1H), 7.62 (dd, <i>J</i> = 8.3, 1.9 Hz, 1H), 7.54 – 7.45 (m, 2H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.56 – 4.32 (m, 3H), 4.20 (p, <i>J</i> = 8.9 Hz, 1H), 3.93 (s, 3H), 3.65 (dd, <i>J</i> = 10.5, 4.3 Hz, 2H), 3.22 (s, 3H), 3.15 (s, 2H), 3.02 (s, 2H), 2.98 – 2.89 (m, 1H), 2.67 – 2.57 (m, 1H), 2.40 – 2.29 (m, 1H), 2.06 – 1.71 (m, 10H), 1.52 (d, <i>J</i> = 32.6 Hz, 4H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 795.20
UB-180690 (190)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.02 (s, 1H), 10.03 (s, 1H), 9.07 (s, 2H), 8.89 (s, 1H), 7.94 (d, <i>J</i> = 5.5 Hz, 1H), 7.83 (dd, <i>J</i> = 7.5, 1.6 Hz, 1H), 7.67 (s, 1H), 7.57 – 7.44 (m, 5H), 7.39 – 7.32 (m, 2H), 5.15 (dd, <i>J</i> = 13.8, 5.8 Hz, 2H), 4.61 (d, <i>J</i> = 16.1 Hz, 1H), 4.48 – 4.31 (m, 3H), 3.93 (s, 3H), 3.66 (s, 4H), 3.25 (s, 3H), 3.13 (s, 2H), 3.02 (s, 2H), 2.94 (dd, <i>J</i> = 8.8, 3.9 Hz, 1H), 2.63 (s, 1H), 2.32 (d, <i>J</i> = 30.5 Hz, 1H), 2.10 – 1.81 (m, 6H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 896.49
UB-180691 (191)	 ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.04 (s, 1H), 10.81 (d, <i>J</i> = 8.5 Hz, 1H), 9.79 (s, 1H), 9.56 (s, 2H), 9.20 (t, <i>J</i> = 5.5 Hz, 1H), 8.12 (s, 1H), 7.91 – 7.80 (m, 3H), 7.68 (d, <i>J</i> = 1.8 Hz, 1H), 7.62 – 7.50 (m, 3H), 5.17 (dd, <i>J</i> = 13.2, 5.1 Hz, 1H), 4.77 (t, <i>J</i> = 6.3 Hz, 2H), 4.57 – 4.48 (m, 4H), 4.46 (s, 1H), 4.06 (s, 2H), 3.90 (s, 3H), 3.57 (s, 2H), 3.22 (s, 3H), 2.94 (ddd, <i>J</i> = 17.9, 13.5, 5.4 Hz, 1H), 2.63 (d, <i>J</i> = 17.0 Hz, 1H), 2.28 (dd, <i>J</i> = 13.1, 4.5 Hz, 1H), 2.06 (dd, <i>J</i> = 9.3, 4.0 Hz, 1H), 2.01 – 1.74 (m, 6H), 1.59 – 1.40 (m, 4H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 848.35

UB-180692 (192)	 HCl
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.04 (s, 1H), 10.70 (d, <i>J</i> = 13.8 Hz, 1H), 9.46 (s, 2H), 9.14 – 9.05 (m, 1H), 8.10 (s, 1H), 7.95 (d, <i>J</i> = 3.7 Hz, 1H), 7.86 (dd, <i>J</i> = 7.5, 1.5 Hz, 2H), 7.63 – 7.56 (m, 2H), 7.54 (d, <i>J</i> = 8.3 Hz, 2H), 7.43 (dd, <i>J</i> = 8.4, 1.9 Hz, 1H), 7.34 (d, <i>J</i> = 8.3 Hz, 2H), 5.16 (dd, <i>J</i> = 13.9, 7.6 Hz, 2H), 4.75 (t, <i>J</i> = 6.4 Hz, 2H), 4.60 (d, <i>J</i> = 15.6 Hz, 1H), 4.53 (d, <i>J</i> = 5.6 Hz, 2H), 4.49 – 4.32 (m, 3H), 4.05 (s, 2H), 3.90 (s, 3H), 3.57 (s, 2H), 3.25 (s, 3H), 2.94 (ddd, <i>J</i> = 17.3, 13.3, 5.3 Hz, 1H), 2.69 – 2.58 (m, 1H), 2.26 (dt, <i>J</i> = 14.2, 7.2 Hz, 2H), 2.10 – 2.00 (m, 1H), 1.89 (d, <i>J</i> = 9.6 Hz, 2H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 950.32
UB-180693 (193)	 HCl
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.43 – 11.24 (m, 1H), 11.02 (s, 1H), 9.95 (s, 1H), 9.25 (s, 2H), 8.04 – 7.93 (m, 1H), 7.88 – 7.79 (m, 2H), 7.72 – 7.56 (m, 2H), 7.53 – 7.43 (m, 2H), 5.16 (m, 1H), 4.80 – 4.69 (m, 1H), 4.52 – 4.30 (m, 3H), 4.29 – 4.16 (m, 1H), 4.00 – 3.90 (m, 3H), 3.72 – 3.43 (m, 6H), 3.22 (s, 3H), 3.07 – 2.81 (m, 4H), 2.70 – 2.54 (m, 4H), 2.11 – 1.39 (m, 14H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LC-MS: (M+H) ⁺ = 878.43
UB-180694 (194)	 HCl
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.04 (s, 1H), 10.65 (s, 1H), 9.31 (s, 2H), 8.91 (s, 1H), 7.98 (s, 1H), 7.89 – 7.83 (m, 2H), 7.68 (s, 1H), 7.61 – 7.53 (m, 3H), 5.18 (dd, <i>J</i> = 13.3, 5.2 Hz, 1H), 4.56 – 4.32 (m, 3H), 4.28 – 4.03 (m, 3H), 3.93 (s, 3H), 3.65 (d, <i>J</i> = 5.8 Hz, 2H), 2.95 (td, <i>J</i> = 16.6, 15.1, 5.4 Hz, 1H), 2.62 (d, <i>J</i> = 17.2 Hz, 1H), 2.27 (dd, <i>J</i> = 13.1, 4.5 Hz, 1H), 2.13 – 2.02 (m, 1H), 2.02 – 1.70 (m, 7H), 1.53 (d, <i>J</i> = 38.8 Hz, 4H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 767.41
UB-180695 (195)	 HCl
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.04 (s, 1H), 10.66 (s, 1H), 9.28 (s, 2H), 8.85 (s, 1H), 8.16 (s, 1H), 7.93 (s, 1H), 7.87 (dd, <i>J</i> = 7.5, 1.6 Hz, 1H), 7.68 (d, <i>J</i> = 1.9 Hz, 1H), 7.63 – 7.50 (m, 3H), 7.12 (d, <i>J</i> = 3.8 Hz, 1H), 6.99 (d, <i>J</i> = 3.8 Hz, 1H), 5.25 – 5.09 (m, 2H), 4.80 (d, <i>J</i> = 15.4 Hz, 1H), 4.54 – 4.32 (m, 3H), 4.08 (s, 2H), 3.96 (s, 3H), 3.22 (s, 5H), 3.00 – 2.88 (m, 1H), 2.62 (d, <i>J</i> = 17.1 Hz, 1H), 2.29 (d, <i>J</i> = 4.4 Hz, 1H), 2.10 – 2.01 (m, 1H), 1.96 – 1.82 (m, 2H), 0.68 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 829.04
UB-180696 (196)	 HCl

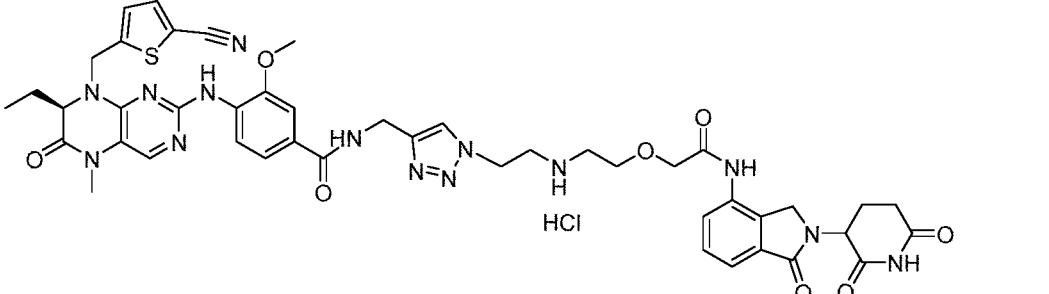
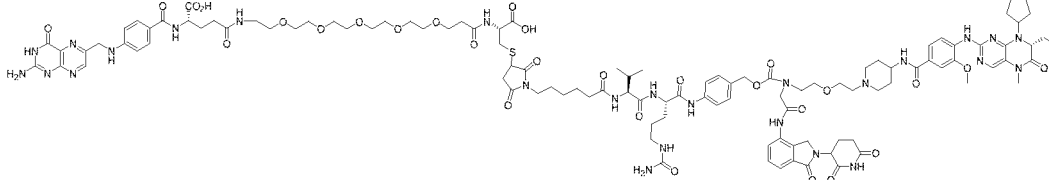
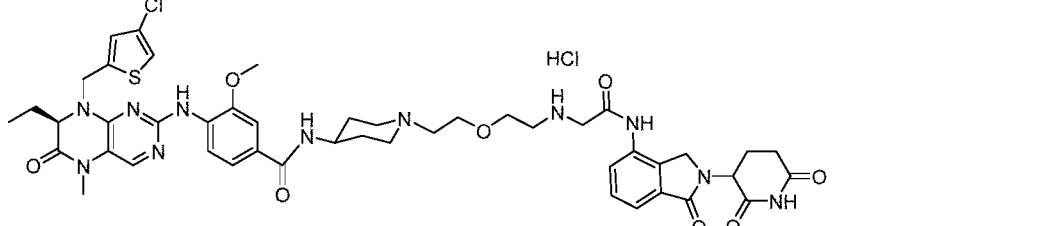
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.03 (s, 1H), 10.15 (d, <i>J</i> = 2.7 Hz, 1H), 8.87 (s, 2H), 8.70 (s, 1H), 7.93 (s, 1H), 7.87 – 7.76 (m, 2H), 7.65 (s, 1H), 7.61 – 7.43 (m, 3H), 5.16 (dd, <i>J</i> = 13.4, 5.2 Hz, 1H), 4.56 – 4.27 (m, 3H), 4.19 (t, <i>J</i> = 8.8 Hz, 1H), 3.92 (s, 3H), 3.74 (t, <i>J</i> = 5.2 Hz, 2H), 3.61 (d, <i>J</i> = 5.8 Hz, 2H), 3.27 (d, <i>J</i> = 7.0 Hz, 2H), 3.22 (s, 3H), 3.18 (m, 2H), 3.00 – 2.82 (m, 3H), 2.71 – 2.59 (m, 1H), 2.37 – 2.22 (m, 2H), 2.07 – 1.72 (m, 8H), 1.53 (d, <i>J</i> = 34.6 Hz, 4H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M/2+1] ⁺ = 413.2
UB-180697 (197)	 HCl ¹ H NMR (400 MHz, <i>d</i> ₆ -DMSO) δ 11.02(s, 1H), 10.24 (s, 1H), 9.13(s,2H), 8.79 (d, <i>J</i> = 13.3 Hz, 1H), 7.99 (t, <i>J</i> = 2.4 Hz, 1H), 7.90 (s, 1H), 7.88 – 7.78 (m, 1H), 7.71(s 1H), 7.68 – 7.64 (m, 1H), 7.52-7.17 (m, 2H), 5.15 (dd, <i>J</i> = 13.2, 5.1 Hz, 1H), 4.58 – 4.36 (m, 4H), 3.92 (s, 3H), 3.77 – 3.49 (m, 6H), 3.39 – 3.30 (m, 7H), 2.96 -2.87(m, 3H), 2.67-2.53(m, 2H), 2.37 – 2.32 (m, 1H), 2.04-1.99 (m, 1H), 1.46 – 1.21 (m, 9H).LCMS [M+H] ⁺ = 785.4
UB-180698 (198)	 HCl ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.00 (s, 1H), 10.12 (s, 1H), 9.18 (s, 2H), 8.84 (s, 1H), 7.90 (s, 1H), 7.84 (s, 1H), 7.74 (d, <i>J</i> = 7.8 Hz, 1H), 7.64 (s, 1H), 7.60 – 7.44 (m, 3H), 5.14 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.58 – 4.32 (m, 3H), 4.26 – 4.10 (m, 3H), 3.90 (s, 3H), 3.83 (t, <i>J</i> = 5.0 Hz, 2H), 3.41 – 3.37 (m, 3H), 3.22 (s, 3H), 3.02 (s, 2H), 2.97 – 2.83 (m, 1H), 2.69 – 2.62 (m, 1H), 2.39 – 2.33 (m, 1H), 2.04 – 1.73 (m, 10H), 1.51 (m, 4H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LC-MS: (M+H) ⁺ = 824.31
UB-180699 (199)	 HCl ¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.04 (s, 1H), 10.69 (s, 1H), 10.44 (s, 1H), 9.35 (s, 2H), 8.53 (d, <i>J</i> = 7.1 Hz, 1H), 7.87 (t, <i>J</i> = 5.5 Hz, 2H), 7.82 (s, 2H), 7.61 – 7.50 (m, 4H), 5.32 (t, <i>J</i> = 4.8 Hz, 1H), 5.20 – 5.15 (m, 1H), 4.51 – 4.34 (m, 4H), 4.22 (m, 2H), 3.92 (s, 3H), 3.82 (m, 2H), 3.75 (m, 2H), 3.59 (m, 2H), 3.24 (s, 3H), 2.92 (m,2H), 2.65 (m, 2H), 2.33-1.83 (m, 11H), 1.59-1.48 (m, 6H), 0.87-0.84 (t, <i>J</i> = 6.6 Hz, 2H), 0.74-0.78 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H] ⁺ = 956
UB-180700 (200)	 HCl ¹ H NMR (400 MHz, <i>DMSO-d</i> ₆) δ 11.03 (s, 1H), 10.23 (s, 1H), 8.90 (s, 2H), 8.77 (d, <i>J</i> = 1.7 Hz, 1H), 8.68 (s, 1H), 8.59 (t, <i>J</i> = 5.5 Hz, 1H), 7.98 (dd, <i>J</i> = 19.3, 6.6Hz, 1H), 7.85 (dd, <i>J</i> = 7.2, 1.8 Hz, 1H), 7.61 – 7.43 (m, 4H), 5.31 (t, <i>J</i> = 5.8 Hz, 1H), 5.16 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.53 – 4.39 (m, 1H), 4.39 – 4.28 (m, 2H), 3.91(d, <i>J</i> = 1.9 Hz, 3H), 3.70 (t, <i>J</i> = 5.2 Hz, 4H), 3.43 (q, <i>J</i> = 5.9 Hz, 6H), 3.24 (d, <i>J</i> = 7.3 Hz, 2H), 3.14 (s, 3H), 2.90 (q, <i>J</i> = 7.3, 6.2 Hz, 2H), 2.62 (d, <i>J</i> = 17.1 Hz, 1H),2.28 (dd, <i>J</i> = 11.7, 5.3 Hz, 1H), 2.05 – 1.78 (m, 8H), 1.56 (d, <i>J</i> = 45.4 Hz, 4H), 1.27 (d, <i>J</i> = 20.9 Hz, 2H), 0.73 (t, <i>J</i> = 7.3 Hz, 3H).LCMS [M+H] ⁺ = 947.62

UB-180701 (201)	 ^1H NMR (400 MHz, <i>DMSO-d</i>₆) δ 11.03 (s, 1H), 10.23 (s, 1H), 9.34 (s, 1H), 8.89 (s, 2H), 8.58 (t, <i>J</i> = 5.7 Hz, 1H), 8.09 (d, <i>J</i> = 8.4 Hz, 1H), 7.91 (s, 1H), 7.85 (dd, <i>J</i> = 7.2, 1.8 Hz, 1H), 7.60 (d, <i>J</i> = 1.9 Hz, 1H), 7.50 (dddd, <i>J</i> = 11.5, 9.7, 6.7, 2.1 Hz, 3H), 7.11 (d, <i>J</i> = 3.8 Hz, 1H), 6.98 (d, <i>J</i> = 3.8 Hz, 1H), 5.23 – 5.08 (m, 2H), 4.80 (d, <i>J</i> = 15.5 Hz, 1H), 4.50 (t, <i>J</i> = 4.6 Hz, 1H), 4.48 – 4.33 (m, 2H), 3.94 (s, 3H), 3.70 (t, <i>J</i> = 5.1 Hz, 2H), 3.56 (s, 5H), 3.22 (s, 5H), 3.17 – 3.12 (m, 3H), 2.91 (dt, <i>J</i> = 14.4, 6.2 Hz, 3H), 2.62 (d, <i>J</i> = 17.2 Hz, 2H), 2.28 (dd, <i>J</i> = 11.9, 5.3 Hz, 2H), 2.10 – 1.83 (m, 4H), 1.30 – 1.18 (m, 2H), 0.67 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [<i>M</i>+<i>H</i>]⁺ = 975.47
UB-180702 (202)	 ^1H NMR (400 MHz, <i>DMSO-d</i>₆) δ 11.03 (s, 1H), 10.32 – 10.18 (m, 1H), 9.57 (s, 1H), 8.98 (s, 2H), 8.63 (s, 1H), 8.04 (d, <i>J</i> = 8.3 Hz, 1H), 7.91 (d, <i>J</i> = 3.6 Hz, 1H), 7.85 (dd, <i>J</i> = 7.2, 1.9 Hz, 1H), 7.62 (d, <i>J</i> = 2.0 Hz, 1H), 7.56 (dd, <i>J</i> = 8.4, 1.8 Hz, 1H), 7.53 – 7.46 (m, 2H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.58 – 4.31 (m, 5H), 3.93 (s, 3H), 3.72 (d, <i>J</i> = 5.1 Hz, 2H), 3.56 (d, <i>J</i> = 8.6 Hz, 7H), 3.20 (s, 5H), 3.13 (s, 3H), 2.93 (dp, <i>J</i> = 13.6, 5.5 Hz, 4H), 2.62 (d, <i>J</i> = 17.6 Hz, 1H), 2.31 (dd, <i>J</i> = 14.8, 10.5 Hz, 1H), 2.09 – 1.94 (m, 2H), 1.41 (d, <i>J</i> = 6.8 Hz, 3H), 1.35 (t, <i>J</i> = 7.2 Hz, 6H). LCMS [<i>M</i>+<i>H</i>]⁺ = 873.54
UB-180703 (203)	 ^1H NMR (400 MHz, <i>DMSO-d</i>₆) δ 11.44 (s, 1H), 11.00 (s, 1H), 10.13 (d, <i>J</i> = 10.0 Hz, 1H), 9.68 (s, 2H), 8.87 (d, <i>J</i> = 6.8 Hz, 1H), 7.95 – 7.83 (m, 2H), 7.79 – 7.67 (m, 2H), 7.62 (d, <i>J</i> = 8.3 Hz, 1H), 7.57 (d, <i>J</i> = 7.4 Hz, 1H), 7.51 (d, <i>J</i> = 7.6 Hz, 1H), 5.14 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.59 – 4.31 (m, 3H), 4.22 (s, 2H), 4.18 (d, <i>J</i> = 8.9 Hz, 1H), 3.93 (d, <i>J</i> = 4.3 Hz, 3H), 3.86 (t, <i>J</i> = 4.8 Hz, 3H), 3.74 (s, 2H), 3.48 (s, 6H), 3.29 (s, 3H), 3.22 (s, 3H), 2.91 (ddd, <i>J</i> = 18.0, 13.5, 5.3 Hz, 1H), 2.67 – 2.56 (m, 1H), 2.43 – 2.29 (m, 1H), 2.07 – 1.70 (m, 7H), 1.59 – 1.40 (m, 4H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [<i>M</i>+<i>H</i>]⁺ = 879.99
UB-180705 (205)	 ^1H NMR (400 MHz, <i>DMSO-d</i>₆) δ 11.01 (s, 1H), 8.44 (t, <i>J</i> = 5.7 Hz, 1H), 8.40 (d, <i>J</i> = 8.3 Hz, 1H), 7.87 (dd, <i>J</i> = 7.1, 1.9 Hz, 1H), 7.83 (s, 1H), 7.58 (s, 1H), 7.54 – 7.46 (m, 4H), 5.14 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.48 – 4.28 (m, 3H), 4.24 (dd, <i>J</i> = 7.6, 3.6 Hz, 1H), 3.92 (s, 3H), 3.57 (dt, <i>J</i> = 12.5, 5.5 Hz, 4H), 3.51 (s, 2H), 3.44 (q, <i>J</i> = 5.8 Hz, 2H), 3.25 (s, 3H), 2.97 – 2.81 (m, 3H), 2.66 – 2.57 (m, 1H), 2.01 (dd, <i>J</i> = 7.2, 4.7 Hz, 2H), 1.93 – 1.52 (m, 10H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [<i>M</i>+<i>H</i>]⁺ = 811.0
UB-180706 (206)	

	¹H NMR (400 MHz, <i>d6</i>-DMSO) δ 11.02 (s, 1H), 10.37 (s, 1H), 9.85(s,1H),9.19(s,2H),8.72 (d, <i>J</i> = 13.3 Hz, 1H), 7.99 (t, <i>J</i> = 2.4 Hz, 1H), 7.90 (s, 1H), 7.88 – 7.78 (m, 1H), 7.66 (d, <i>J</i> = 3.3 Hz, 2H), 7.61 – 7.47 (m, 4H), 5.15 (dd, <i>J</i> = 13.2, 5.1 Hz, 1H), 4.68 – 4.36 (m, 4H), 3.93 (s, 3H), 3.92-3.73(m,2H), 3.66 – 3.51 (m, 9H), 3.49 – 3.40 (m, 2H), 3.30 – 3.11 (m, 7H), 2.96 – 2.81 (m, 43H), 2.53-2.51(m,1H)2.47 – 2.36 (m, 1H), 2.19 – 1.71(m, 3H), 1.46 – 1.21 (m, 6H), 0.77-0.74(m,3H). LCMS [M+H]⁺=887.3
UB-180711 (211)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.01 (s, 1H), 10.66 (s, 1H), 10.01 (s, 1H), 9.49 (m, 2H), 8.51 (m, 1H), 8.16 (m, 1H), 7.90 (s, 1H), 7.74 (d, <i>J</i> = 7.7 Hz, 1H), 7.61 – 7.50 (m, 4H), 7.12 (d, <i>J</i> = 3.9 Hz, 1H), 6.99 (d, <i>J</i> = 3.8 Hz, 1H), 5.19 – 5.10 (m, 2H), 4.79 (m, 1H), 4.53 – 4.33 (m, 4H), 4.23 (s, 2H), 4.07 (m, 1H), 3.96 (s, 3H), 3.86 (m, 2H), 3.66 (m, 3H), 3.23 (m, 4H), 3.15 (m, 3H), 2.98 – 2.87 (m, 2H), 2.10 – 1.83 (m, 10H), 0.67 (t, <i>J</i> = 7.4 Hz, 3H).LCMS [M+H]⁺=956.3
UB-180712 (212)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.01 (s, 1H), 10.74 (s, 1H), 10.04 (s, 1H), 9.56 (m, 2H), 8.55 (d, <i>J</i> = 7.5 Hz, 1H), 8.08 (d, <i>J</i> = 8.4 Hz, 1H), 7.87 (s, 1H), 7.75 (dd, <i>J</i> = 7.9, 1.2 Hz, 1H), 7.63 – 7.49 (m, 4H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.55 – 4.48 (m, 2H), 4.46 – 4.34 (m, 2H), 4.23 (s, 2H), 4.09 (s, 1H), 3.94 (s, 3H), 3.87 (m, 2H), 3.67 (m, 2H), 3.51 (m, 6H), 3.22 (s, 3H), 3.15 (s, 2H), 2.68 – 2.61 (m, 1H), 2.39 – 2.32 (m, 1H), 2.01 (m, 7H), 1.81 (m, 1H), 1.37 (t, <i>J</i> = 7.2 Hz, 6H), 0.77 (t, <i>J</i> = 7.4 Hz, 3H).LCMS [M+H]⁺=868.3
UB-180713 (213)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.04 (s, 1H), 10.83 (d, <i>J</i> = 12.4 Hz, 1H), 10.77 - 10.58 (m, 1H), 9.54 - 9.27 (m, 2H), 9.21 - 8.86 (m, 1H), 8.22 - 8.00 (m, 1H), 7.96 - 7.78 (m, 2H), 7.76 - 7.47 (m, 4H), 5.24 - 5.09 (m, 1H), 4.83 - 4.63 (m, 1H), 4.55 - 4.46 (m, 1H), 4.44 - 4.34 (m, 2H), 4.28 - 4.19 (m, 1H), 4.16 - 4.07 (m, 2H), 3.97 - 3.89 (m, 3H), 3.84 - 3.76 (m, 4H), 3.62 - 3.54 (m, 6H), 3.53 - 3.44 (m, 6H), 3.23 (s, 3H), 2.99 - 2.90 (m, 1H), 2.66 - 2.60 (m, 1H), 2.32 - 2.20 (m, 1H), 2.10 - 2.02 (m, 1H), 1.98 - 1.92 (m, 1H), 1.90 - 1.77 (m, 3H), 1.68 - 1.58 (m, 2H), 1.57 - 1.45 (m, 2H), 0.82 - 0.69 (m, 3H). LCMS [M+H]⁺=880.35.
UB-180714 (214)	

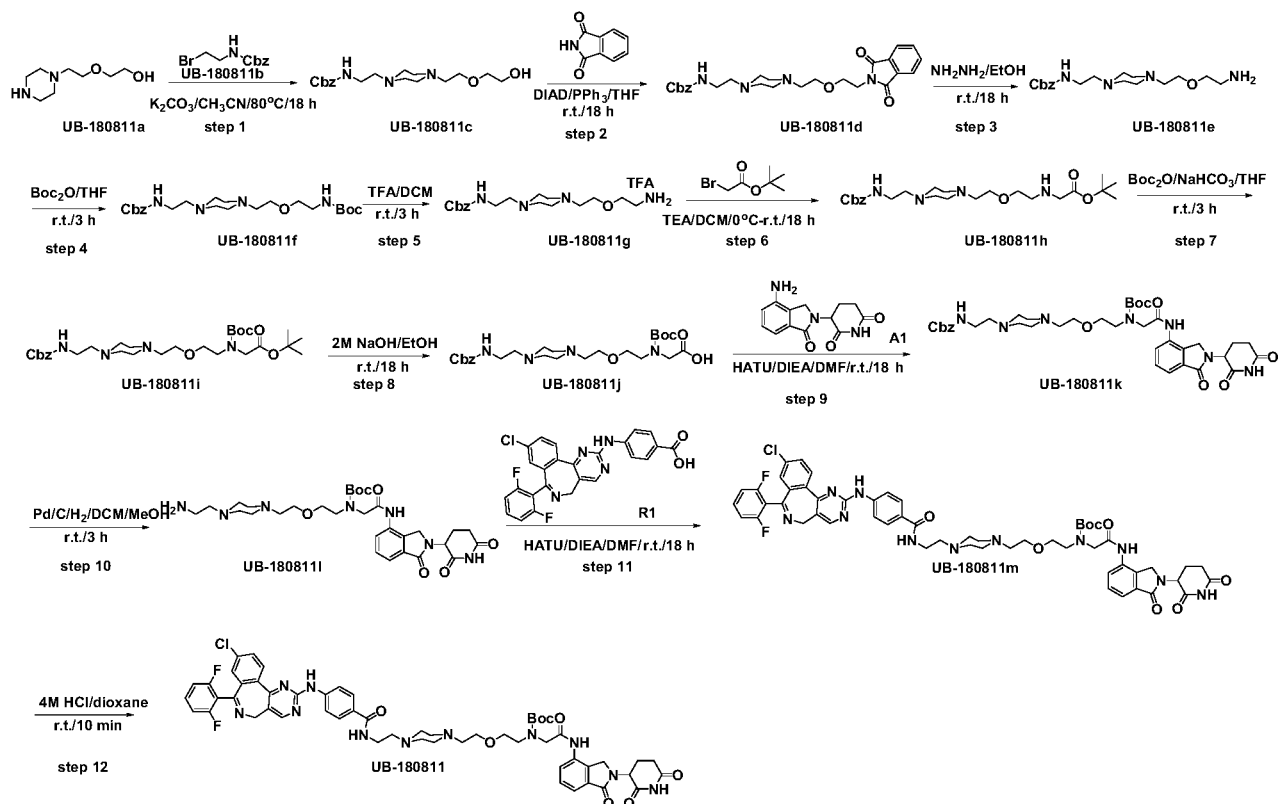
	¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 10.25 (s, 1H), 9.25 (s, 2H), 9.09 (t, <i>J</i> = 5.7 Hz, 1H), 8.09 (d, <i>J</i> = 9.3 Hz, 2H), 7.92 (s, 1H), 7.84 (dd, <i>J</i> = 7.3, 1.7 Hz, 1H), 7.65 (d, <i>J</i> = 1.9 Hz, 1H), 7.59 – 7.47 (m, 3H), 7.11 (d, <i>J</i> = 3.8 Hz, 1H), 6.98 (d, <i>J</i> = 3.8 Hz, 1H), 5.22 – 5.08 (m, 2H), 4.88 – 4.67 (m, 3H), 4.55 – 4.32 (m, 5H), 3.94 (s, 3H), 3.52 – 3.48 (m, 2H), 3.22 (s, 5H), 2.93 – 2.86 (m, 2H), 2.62 (d, <i>J</i> = 14.8 Hz, 1H), 2.39 – 2.27 (m, 1H), 2.04 (dd, <i>J</i> = 9.1, 4.1 Hz, 1H), 1.98 – 1.83 (m, 2H), 0.67 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 924.2
UB-180715 (215)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 10.02 (s, 1H), 9.07 (d, <i>J</i> = 53.3 Hz, 3H), 8.55 (d, <i>J</i> = 8.3 Hz, 1H), 8.34 (s, 1H), 8.13 (s, 1H), 7.84 (d, <i>J</i> = 6.7 Hz, 1H), 7.63 – 7.44 (m, 4H), 7.10 (d, <i>J</i> = 3.6 Hz, 1H), 6.98 (d, <i>J</i> = 3.5 Hz, 1H), 5.22 – 5.06 (m, 2H), 4.80 – 4.66 (m, 3H), 4.61 – 4.32 (m, 5H), 3.96 (s, 3H), 3.59 (d, <i>J</i> = 7.8 Hz, 2H), 3.25 (s, 3H), 2.96 (d, <i>J</i> = 26.1 Hz, 4H), 2.66 (d, <i>J</i> = 18.4 Hz, 1H), 2.34 (s, 1H), 1.95 (s, 2H), 1.83 (s, 2H), 0.66 (t, <i>J</i> = 7.2 Hz, 3H). LCMS [M+H]⁺ = 938.2
UB-180716 (216)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 9.93 (s, 1H), 9.73 (s, 1H), 9.07 (t, <i>J</i> = 5.8 Hz, 1H), 8.83 (s, 1H), 8.08 (d, <i>J</i> = 17.8 Hz, 2H), 7.88 – 7.76 (m, 2H), 7.64 – 7.44 (m, 4H), 5.16 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.80 (t, <i>J</i> = 6.7 Hz, 2H), 4.53 (d, <i>J</i> = 5.7 Hz, 2H), 4.46 – 4.29 (m, 4H), 4.22 (p, <i>J</i> = 8.9 Hz, 2H), 3.91 (s, 4H), 3.72 (s, 4H), 3.23 (s, 5H), 2.93 (ddd, <i>J</i> = 17.8, 13.5, 5.4 Hz, 1H), 2.84 (s, 3H), 2.61 (d, <i>J</i> = 17.6 Hz, 1H), 2.37 – 2.25 (m, 1H), 2.11 – 1.68 (m, 9H), 1.68 – 1.42 (m, 4H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 890.4
UB-180717 (217)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 9.98 (s, 1H), 9.87 (s, 1H), 8.53 (d, <i>J</i> = 6.0 Hz, 1H), 8.07 (d, <i>J</i> = 7.4 Hz, 1H), 7.90 (d, <i>J</i> = 8.5 Hz, 1H), 7.83 – 7.71 (m, 3H), 7.64 – 7.44 (m, 6H), 7.28 (d, <i>J</i> = 8.3 Hz, 2H), 7.00 (s, 2H), 6.05 – 5.92 (m, 1H), 5.14 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.99 (s, 2H), 4.51 – 4.41 (m, 1H), 4.40 – 4.30 (m, 4H), 4.23 – 4.11 (m, 4H), 3.90 (s, 3H), 3.51 – 3.41 (m, 9H), 3.41 – 3.30 (m, 6H), 3.08 – 2.85 (m, 3H), 2.72 – 2.58 (m, 4H), 2.38 – 2.28 (m, 4H), 2.21 – 2.07 (m, 3H), 1.95 (dt, <i>J</i> = 13.2, 6.8 Hz, 4H), 1.79 (dd, <i>J</i> = 14.8, 7.4 Hz, 4H), 1.57 – 1.39 (m, 10H), 1.19 (q, <i>J</i> = 7.6 Hz, 2H), 0.87 – 0.79 (m, 6H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 1512
UB-180718 (218)	

	¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.05 (s, 1H), 10.59 (s, 1H), 10.25 (s, 1H), 9.29 (s, 2H), 8.45 (d, <i>J</i> = 7.1 Hz, 1H), 8.27 (s, 1H), 7.87 (d, <i>J</i> = 5.5 Hz, 2H), 7.61 – 7.50 (m, 4H), 7.11 (d, <i>J</i> = 3.7 Hz, 1H), 6.99 (t, <i>J</i> = 3.6 Hz, 1H), 5.32 (t, <i>J</i> = 4.8 Hz, 1H), 5.23 – 5.09 (m, 3H), 4.75 (m, 1H), 4.51 – 4.34 (m, 4H), 4.07 (m, 3H), 3.96 (s, 3H), 3.82 (m, 2H), 3.75 (m, 2H), 3.59 (m, 2H), 3.24 (s, 3H), 2.10 – 1.95 (m, 8H), 1.86 (m, 2H), 1.46 (m, 2H), 0.85 (t, <i>J</i> = 6.6 Hz, 2H), 0.67 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 956
UB-180733 (233)	HCl
	¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 10.02 (s, 1H), 9.79 (s, 1H), 8.89 (m, 2H), 8.64 (t, <i>J</i> = 5.6 Hz, 1H), 8.00 (d, <i>J</i> = 8.4 Hz, 1H), 7.94 (s, 1H), 7.83 (dd, <i>J</i> = 7.1, 2.0 Hz, 1H), 7.63 (d, <i>J</i> = 1.9 Hz, 1H), 7.53 – 7.46 (m, 3H), 7.11 (d, <i>J</i> = 3.8 Hz, 1H), 6.98 (d, <i>J</i> = 3.8 Hz, 1H), 5.20 – 5.10 (m, 2H), 4.83 (m, 1H), 4.54 (m, 1H), 4.48 – 4.33 (m, 2H), 3.94 (s, 3H), 3.69 (m, 3H), 3.54 (m, 10H), 3.42 (m, 3H), 3.21 (s, 3H), 3.07 (m, 2H), 2.93 (m, 3H), 2.42 (t, <i>J</i> = 6.8 Hz, 2H), 2.02 (m, 1H), 1.98 – 1.88 (m, 2H), 1.72 – 1.62 (m, 4H), 0.67 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 1003
UB-180734 (234)	HCl ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.05 (s, 1H), 10.81 (s, 1H), 9.89 (s, 1H), 9.21 (s, 2H), 8.65 (t, <i>J</i> = 5.6 Hz, 1H), 8.05 – 7.93 (m, 2H), 7.87 (dd, <i>J</i> = 7.3, 1.7 Hz, 1H), 7.64 (d, <i>J</i> = 1.9 Hz, 1H), 7.61 – 7.44 (m, 3H), 7.11 (d, <i>J</i> = 3.8 Hz, 1H), 6.98 (d, <i>J</i> = 3.8 Hz, 1H), 5.24 – 5.10 (m, 3H), 4.83 (d, <i>J</i> = 15.5 Hz, 1H), 4.61 – 4.24 (m, 4H), 4.06 (t, <i>J</i> = 5.5 Hz, 2H), 3.94 (s, 3H), 3.72 (t, <i>J</i> = 5.2 Hz, 2H), 3.56 (s, 3H), 3.53 (m, <i>J</i> = 4.4 Hz, 5H), 3.41 (q, <i>J</i> = 6.4, 6.0 Hz, 2H), 3.21 (m, 5H), 2.99 – 2.85 (m, 1H), 2.70 – 2.57 (m, 1H), 2.35 – 2.23 (m, 1H), 2.14 – 1.98 (m, 1H), 1.97 – 1.85 (m, 2H), 0.67 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 961.1
UB-180745 (245)	HCl ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.04 (d, <i>J</i> = 3.2 Hz, 1H), 10.97 (d, <i>J</i> = 5.5 Hz, 1H), 10.63 (s, 1H), 9.83 (s, 1H), 9.48 (s, 2H), 8.60 (dd, <i>J</i> = 18.3, 7.0 Hz, 1H), 8.05 (dd, <i>J</i> = 8.5, 4.9 Hz, 1H), 7.98 (s, 1H), 7.88 (dd, <i>J</i> = 7.4, 1.6 Hz, 1H), 7.77 – 7.70 (m, 1H), 7.63 (d, <i>J</i> = 1.8 Hz, 1H), 7.60 – 7.50 (m, 3H), 5.28 – 5.12 (m, 3H), 4.85 (d, <i>J</i> = 15.5 Hz, 2H), 4.58 – 4.36 (m, 4H), 4.11 (d, <i>J</i> = 7.1 Hz, 2H), 3.96 (d, <i>J</i> = 8.1 Hz, 3H), 3.84 (s, 2H), 3.78 (t, <i>J</i> = 5.1 Hz, 2H), 3.61 (d, <i>J</i> = 11.5 Hz, 2H), 3.14 (d, <i>J</i> = 11.6 Hz, 2H), 3.01 – 2.85 (m, 1H), 2.64 (s, 1H), 2.36 – 2.27 (m, 1H), 2.18 – 1.92 (m, 7H), 0.69 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 937.2
UB-180746 (246)	HCl

UB-180765 (265)	
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.00 (s, 1H), 10.11 (s, 1H), 9.43 (s, 2H), 9.07 (s, 1H), 8.10 (s, 1H), 8.00 (s, 1H), 7.93 (s, 1H), 7.84 (d, J = 3.8 Hz, 1H), 7.75 (dd, J = 7.8, 1.2 Hz, 1H), 7.65 – 7.60 (m, 1H), 7.59 – 7.47 (m, 3H), 7.37 (d, J = 3.9 Hz, 1H), 5.24 (d, J = 15.8 Hz, 1H), 5.14 (dd, J = 13.2, 5.1 Hz, 1H), 4.95 (d, J = 15.8 Hz, 1H), 4.79 (t, J = 6.6 Hz, 2H), 4.56 (t, J = 4.2 Hz, 1H), 4.54 – 4.49 (m, 2H), 4.49 – 4.34 (m, 2H), 4.20 (s, 2H), 3.93 (d, J = 2.9 Hz, 3H), 3.82 (d, J = 4.9 Hz, 2H), 3.23 (s, 5H), 2.91 (ddd, J = 18.3, 13.7, 5.5 Hz, 1H), 2.60 (d, J = 17.5 Hz, 1H), 2.39 – 2.32 (m, 1H), 2.02 (dd, J = 11.0, 5.5 Hz, 1H), 1.91 (d, J = 6.3 Hz, 2H), 0.67 (t, J = 7.3 Hz, 3H). LCMS [M+H] ⁺ = 945.2
UB-180766 (266)	
	LCMS [M/2+H] ⁺ = 1165
UB-180767 (267)	
	¹ H NMR (400 MHz, DMSO- <i>d</i> ₆) δ 11.03 (d, J = 3.2 Hz, 1H), 10.39 (d, J = 17.5 Hz, 1H), 10.17 (s, 1H), 9.30 (s, 2H), 8.73 (s, 1H), 8.43 (d, J = 7.1 Hz, 1H), 8.30 (d, J = 8.5 Hz, 1H), 7.96 – 7.84 (m, 8H), 7.55 – 7.44 (m, 3H), 7.34 (d, J = 2.2 Hz, 2H), 7.18 (d, J = 8.8 Hz, 2H), 5.15 (dt, J = 13.3, 4.8 Hz, 1H), 4.49 – 4.32 (m, 2H), 4.09 (d, J = 51.8 Hz, 2H), 3.61 (q, J = 10.7, 9.2 Hz, 2H), 3.38 (s, 2H), 3.28 (s, 5H), 3.15 (d, J = 39.6 Hz, 5H), 3.05 – 2.85 (m, 4H), 2.60 (d, J = 17.1 Hz, 1H), 2.37 – 2.15 (m, 3H), 2.01 (d, J = 11.9 Hz, 4H). LCMS [M+H] ⁺ = 956.4

(2.9) Preparation procedure 9 of TED molecule:

Synthesis method of Compound UB-180811



Step 1: UB-180811c

Compound **UB-180811a** (3.2 g, 18.2 mmol) was dissolved in acetonitrile (100 mL). Compound **UB-180811b** (4.7 g, 18.2 mmol) was added slowly dropwise, followed by the addition of potassium carbonate (5 g, 36.4 mmol), then reacted at $80^\circ C$ for 18 hours. After the completion of reaction, the reaction was filtrated. The filtrate was concentrated to obtain the crude product. The crude product was isolated by silica gel column chromatography (dichloromethane/methanol = 0% to 30%) to obtain target product **UB-180811c** (5.8 g, yield 90%) as colorless transparent oil. LCMS $[M+H]^+ = 352$

Step 2: UB-180811d

Compound **UB-180811c** (1.9 g, 5.4 mmol) was dissolved in tetrahydrofuran (40 mL) and diisopropyl azodicarboxylate (1.6 g, 8.1 mmol) and triphenylphosphine (2.1 g, 8.1 mmol) dropwise, reacted at room temperature for 2 hours, added the compound phthalimide (1.2 g, 8.1 mmol) and reacted at room temperature over the weekend. After completion of reaction, the reaction solution was concentrated to obtain a crude product, which was isolated by silica gel column chromatography (dichloromethane/methanol=0% to 10%) to obtain the target product **UB-180811d** (3.2 g) as a yellow transparent oil. LCMS $[M+H]^+ = 481$

^1H NMR (400 MHz, $DMSO-d_6$) δ 7.57 – 7.51 (m, 4H), 7.43 – 7.23 (m, 5H), 7.11 (m, 1H), 5.01 (d, $J = 1.9$ Hz, 2H), 3.75 (t, $J = 5.7$ Hz, 1H), 3.58 (q, $J = 7.0, 6.4$ Hz, 1H), 3.47 (d, $J = 5.8$ Hz, 4H), 3.14 – 2.97 (m, 2H), 2.47 – 1.92 (m, 12H).

Step 3: UB-180811e

Compound **UB-180811d** (1.5 g, 3.1 mmol) was dissolved in ethanol (20 mL) and added hydrazine (580 mg, 11.6 mmol), reacted at room temperature for 18 hours. When the reaction is cooled to room temperature, some solids will precipitate out. The reaction was filtered, the filtrate was dried by rotary evaporation. The crude product was washed repeatedly with ethanol, and the filtrate was dried by rotary dryer to obtain 1.2 g crude product **UB-180758e**. LCMS $[M+H]^+ = 351$

Step 4: UB-180811f

Compound **UB-180811e** (1 g, 2.8 mmol) was dissolved in tetrahydrofuran (20 mL). Di-*tert*-butyl

dicarbonate (2 mL, 8.7 mmol) was added. Reaction was allowed at room temperature for 3 hours. After completion of the reaction, the filtrate was concentrated to obtain the crude product, which was isolated by silica gel column chromatography (dichloromethane/methanol = 0% to 5%) to obtain target product **UB-180811f** (580 mg, yield 42%) as a yellow transparent oil. LCMS $[M+H]^+ = 451$

Step 5: UB-180811g

Compound **UB-180811f** (580 mg, 1.28 mmol) was dissolved in dichloromethane (10 mL) and added trifluoroacetic acid (2 mL), reacted at room temperature for 3 hours. The target compound obtained by concentrating after completion of reaction was directly used in the next step. LCMS $[M+H]^+ = 351$

Step 6: UB-180811h

Triethylamine (84 mg, 0.84 mmol) was added dropwise to a solution of *tert*-butyl bromoacetate (82 mg, 0.42 mmol) in dichloromethane (10 mL) under ice bath, and then reacted for 10 minutes. Compound **UB-180811g** (150 mg, 0.42 mmol) was dissolved in dichloromethane (2 mL) and added to the above reaction, reacted at room temperature for 18 hours. The reaction was added saturated brine (10 mL) and extracted with dichloromethane (20 mL * 2). The organic phase was concentrated and purified by climbing large plate (dichloromethane/methanol = 10/1) to obtain the target product **UB-180811h** (40 mg, yield 20%) as a yellow transparent oil. LCMS $[M+H]^+ = 465$

Step 7: UB-180811i

Compound **UB-180811h** (450 mg, 0.97 mmol) was dissolved in tetrahydrofuran (10 mL), added di-*tert*-butyl dicarbonate (632 mg, 2.9 mmol) and sodium bicarbonate (243 mg, 2.9 mmol), and reacted at room temperature for 3 hours. The reaction solution was concentrated to obtain a crude product, which was isolated by silica gel column chromatography (dichloromethane/methanol=0% to 5%) to obtain the target product **UB-180811i** (500 mg, yield 91%) as yellow transparent oil. LCMS $[M+H]^+ = 565$

^1H NMR (400 MHz, DMSO- d_6) δ 7.43 – 7.28 (m, 5H), 7.16 – 7.07 (m, 1H), 5.01 (s, 2H), 3.84 (d, $J = 6.3$ Hz, 2H), 3.45 (dt, $J = 8.5, 5.8$ Hz, 4H), 3.30 (d, $J = 4.9$ Hz, 2H), 3.09 (q, $J = 6.5$ Hz, 2H), 2.48 – 2.19 (m, 12H), 1.45 – 1.32 (m, 18H).

Step 8: UB-180811j

Compound **UB-180811i** (500 mg, 0.88 mmol) was dissolved in ethanol (5 mL), added 2M NaOH (3 mL, 2 mmol), and reacted at room temperature for 18 hours. The reaction solution was concentrated, added water (3 mL), and then extracted with ether (10 mL * 3) to remove organic impurities. The aqueous phase was neutralized to pH~6 using 1M HCl and lyophilized to obtain product **UB-180811j** (408 mg) as white solid. LCMS $[M+H]^+ = 509$

Step 9: UB-180811k

Compound **UB-180811j** (405 mg, 0.8 mmol), **A1** (208 mg, 0.8 mmol), HATU (365 mg, 0.96 mmol), and DIPEA (0.4 mL) were dissolved in DMF (3 mL), reacted at room temperature for 18 hours. After completion of reaction, the reaction solution was concentrated to obtain a crude product, which was isolated by silica gel column chromatography (dichloromethane/methanol=0% to 20%) to obtain the target product **UB-180811k** (380 mg, yield 54%) as a yellow transparent oil. LCMS $[M+H]^+ = 750$

Step 10: UB-180811l

Compound **UB-180811k** (380 mg, 0.5 mmol) was dissolved in dichloromethane (6 mL) methanol (3 mL), added 10% palladium on carbon (50 mg) and reacted at room temperature for 2 hours under hydrogen. After the completion of reaction, the palladium on carbon was filtered off, and the filtrate was concentrated to obtain light yellow crude product **UB-180811l** (300 mg). LCMS $[M+H]^+ = 616$

Step 11: UB-180811m

Compound **UB-180811l** (123 mg, 0.2 mmol), **R1** (95 mg, 0.2 mmol), HATU (152 mg, 0.4 mmol), and

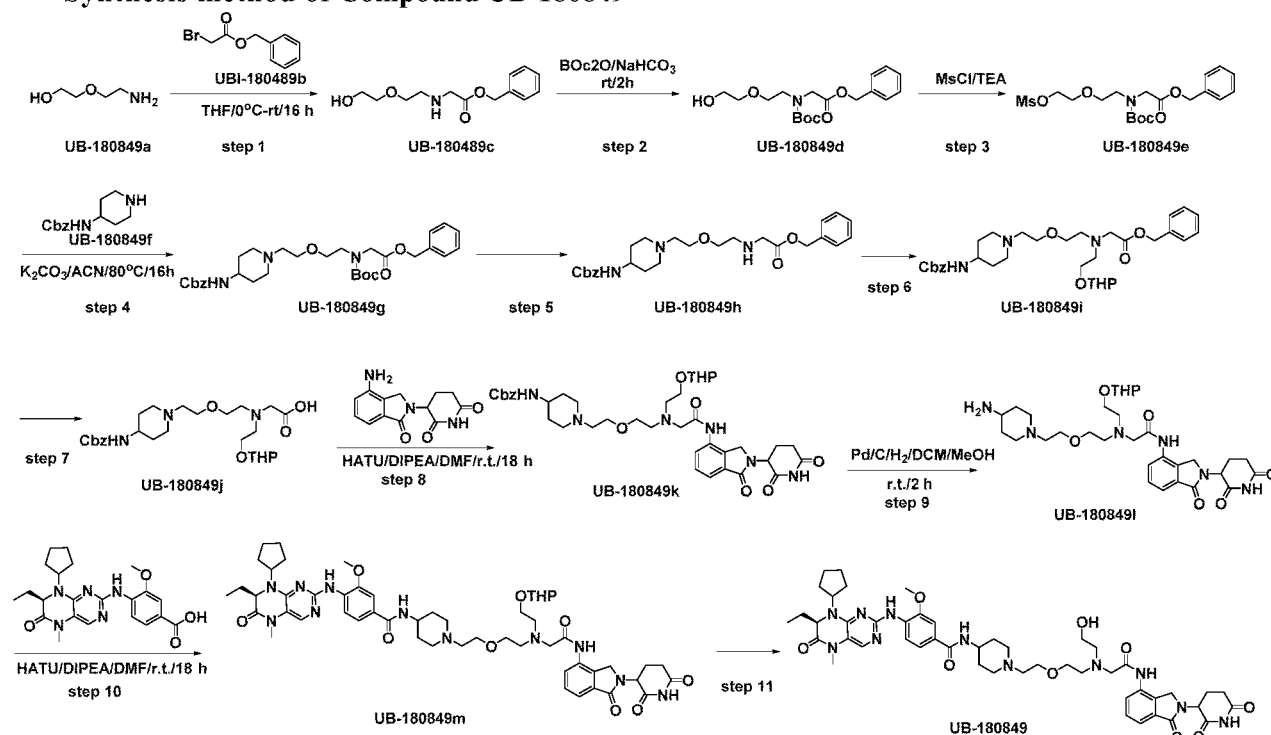
DIPEA (0.2 mL) were dissolved in DMF (3 mL), reacted at room temperature for 18 hours. The reaction was added saturated brine (10 mL), extracted with dichloromethane (20 mL*2). The organic phase was concentrated, and subjected to Prep-TLC (dichloromethane/methanol = 10/1) to obtain target product **UB-180811m** (100 mg, yield 46%) as yellow solid. LCMS $[M+H]^+ = 1074$

Step 12: UB-180811

Compound **UB-180811m** (100 mg) was dissolved in dichloromethane (1.5 mL) and added 4M HCl/dioxane (0.4 mL), reacted at room temperature for 5 minutes. The supernatant was discarded and the solid was washed with ether (10 mL*3), lyophilized to obtain product **UB-180811** (97 mg, yield 100%) as light yellow solid. LCMS $[M+H]^+ = 974.4$

^1H NMR (400 MHz, DMSO- d_6) δ 11.05 (s, 1H), 10.95 (s, 1H), 10.23 (s, 1H), 9.45 (s, 2H), 8.77 (m, 2H), 8.30 (d, $J = 8.5$ Hz, 1H), 8.04 – 7.83 (m, 6H), 7.55 (q, $J = 7.3$ Hz, 3H), 7.35 (d, $J = 2.2$ Hz, 1H), 7.18 (t, $J = 8.8$ Hz, 2H), 5.17 (dd, $J = 13.2, 5.2$ Hz, 1H), 4.54 (d, $J = 17.8$ Hz, 1H), 4.41 (d, $J = 17.7$ Hz, 1H), 4.15 (d, $J = 4.8$ Hz, 2H), 3.96 (m, 4H), 3.80 (m, 4H), 3.72 (m, 8H), 3.50 (m, 2H), 3.40 – 3.17 (m, 4H), 2.94 (m, 1H), 2.64 (m, 1H), 2.37 – 2.26 (m, 1H), 2.06 (m, 1H).

Synthesis method of Compound UB-180849



Step 1: UB-180849c

Compound **UB-180849b** (19.05g, 66.05mmol) was dissolved in tetrahydrofuran (250 mL), and added dropwise to a solution of Compound **UB-180849a** (22.95g, 218.7mmol) in tetrahydrofuran (550 mL) in ice bath. The reaction was carried out at room temperature for 16 hours. The reaction solution was concentrated and isolated by silica gel column chromatography (methanol/dichloromethane = 0% to 10%) to obtain product **UB-180849c** (17.4g, yield 63%) as colorless transparent oil. ^1H NMR (400 MHz, chloroform- d) δ 7.44 - 7.29 (m, 5H), 5.18 (s, 2H), 3.79 - 3.68 (m, 2H), 3.65 - 3.55 (m, 4H), 3.51 (s, 2H), 2.90 - 2.79 (m, 2H), 1.95 (s, 2H).

Step 2: UB-180849d

Compound **UB-180849c** (4.100g, 18.76mmol), (16.50 g, 65.14 mmol), (Boc) $_2$ O (28.43 g, 130.3 mmol) and NaHCO $_3$ (10.94 g, 130.3 mmol) were dissolved in tetrahydrofuran (600 mL), reacted at room temperature for 2 hours. The reaction was filtered. The filtrate was concentrated and isolated by silica gel

column chromatography (dichloromethane/methanol = 0% to 3%) to obtain product **UBI-180849d** (20.7g, yield 90%) as colorless transparent oil.

¹H NMR (400 MHz, chloroform-d) δ 7.43 - 7.29 (m, 5H), 5.17 (s, 2H), 4.06 (d, J = 33.2 Hz, 2H), 3.70 - 3.56 (m, 4H), 3.55 - 3.41 (m, 4H), 1.41 (d, J = 43.2 Hz, 9H).

5 Step 3: UB-180849e

Compound **UB-180849d** (119.78 g, 55.97 mmol) was dissolved in dichloromethane (500 mL), added TEA (11.33g, 111.9 mmol) and MsCl (7.692 g, 67.16 mmol) under ice bath, and reacted for 1 hour in the ice bath, followed by at room temperature for 2 hours. The reaction solution was washed with saturated brine, dried over anhydrous sodium sulfate, and concentrated by rotary evaporation under reduced pressure to obtain product **UB-180849f** (22.94g, yield 95%) as yellow oil.

¹H NMR (400 MHz, chloroform-d) δ 7.43 - 7.29 (m, 5H), 5.21 - 5.13 (m, 2H), 4.32 - 4.21 (m, 2H), 4.06 (d, J = 33.2 Hz, 2H), 3.66 - 3.56 (m, 4H), 3.53 - 3.43 (m, 2H), 3.01 (s, 3H), 1.41 (d, J = 42.8 Hz, 9H).

Step 4: UB-180849i

Compound **UBI-180849h** (12.00g, 44.32mmol), **UBI-180849e** (22.95g, 53.18mmol), and K₂CO₃ (15.31g, 110.8mmol) were added to acetonitrile (300 mL) and reacted at 80 °C for 16 hours under stirring. The reaction was filtered. The filtrate was concentrated and then isolated by silica gel column chromatography (methanol/dichloromethane = 0% to 10%) to obtain product **UBI-180849c** (23.4g, yield 93%) as colorless transparent oil.

¹H NMR (400 MHz, chloroform-d) δ 7.38 - 7.30 (m, 10H), 5.17 - 5.14 (m, 2H), 5.09 (s, 2H), 4.62 (s, 1H), 4.13 - 4.02 (m, 2H), 3.63 - 3.42 (m, 8H), 2.97 - 2.66 (m, 2H), 2.51 (s, 2H), 2.14 (s, 2H), 2.01 - 1.85 (m, 2H), 1.39 (d, J = 42.4 Hz, 9H).

Step 5: UB-180849h

4 mol/L solution of hydrogen chloride in dioxane (2ml) was added dropwise to a solution of UB-180849g (500mg, 0.87mmol) in dichloromethane (20ml), stirred at room temperature for 1 hour. After completion of reaction, the reaction solution was concentrated to obtain compound UB-180849h (350 mg, 85%) as a yellow solid. The product was directly used in the next step without further purification. LCMS [M+H]⁺ = 470.3

Step 6: UB-180849i

2-(2-Bromoethoxy) tetrahydro-2H-pyran (187mg, 0.89 mmol), **UB-180849h** (350mg, 0.75mmol), and potassium carbonate (206 mg, 1.49 mmol) were dissolved in CH₃CN (10 ml). The reaction solution was refluxed overnight. After completion of reaction, it was diluted with water and extracted with ethyl acetate. The organic layers were combined, dried, filtered, and concentrated. The crude product was isolated by silica gel column chromatography (methanol/dichloromethane=5%) to obtain target product **UB-180849I** (250mg, yield 56%) as yellow oil. LCMS [M+H]⁺ = 598.4

Step 7: UB-180849j

Compound **UB-180849i** (250 mg, 19.66 mmol) was dissolved in tetrahydrofuran (20 mL), added NaOH (20 mmol, 5 mL), and reacted overnight at room temperature under stirring. The reaction solution was added water, concentrated and extracted with ethyl acetate. The pH of the aqueous phase was adjusted to 6 using 1N HCl. After the aqueous phase was lyophilized, the solid was dissolved in dichloromethane (200 mL) and methanol (20 mL). It was filtered. The filtrate was concentrated to obtain product **UBI-180849j** (130g, yield 61%) as yellow oil. LCMS [M+H]⁺ = 342.9; ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.43 - 7.26 (m, 5H), 5.02 (s, 2H), 4.03 (q, J = 7.1 Hz, 1H), 3.87 (d, J = 9.8 Hz, 2H), 3.74 - 3.68 (m, 2H), 3.51 (q, J = 5.8 Hz, 3H), 3.37 (t, J = 5.8 Hz, 10H), 3.13 (s, 2H), 2.99 (s, 2H), 1.99 (s, 2H), 1.91 (s, 2H), 1.71 (d, J = 11.7 Hz, 2H), 1.34 (m, 6H).

Step 8: UB-180849k

Compound **UBI-180849j** (130mg, 0.26mmol), A1 (66mg, 0.26mmol), and HATU (144mg, 0.39 mmol) were dissolved in DMF (3 mL) and DIPEA (0.3 mL), and reacted at room temperature for 16 hours. The reaction was quenched with water (10 mL), then extracted with ethyl acetate (8 mL*3). The organic phases were combined, then dried over anhydrous sodium sulfate and concentrated to obtain crude product. The crude product was purified by reversed-phase column (methanol/water = 5% to 95%, collected at 70%) to obtain **UBI-180849k** (80mg, yield 62%) as white solid. LCMS $[M+H]^+ = 749.5$

Step 9: UB-180849l

Under hydrogen atmosphere, compound **UBI-180849k** (80mg, 0.10mmol), Pd/C (20mg) were dissolved in methanol (1 mL) and dichloromethane (10mL), and the reaction solution was reacted at room temperature for 1 hour. The reaction solution was filtered through Celite. The filtrate was concentrated and purified via reversed-phase column (methanol/water = 5% to 95%, collected at 15%) to obtain the product **UBI-180849l** (35mg, yield 53%) as yellow oil. LCMS $[M+H]^+ = 615.4$

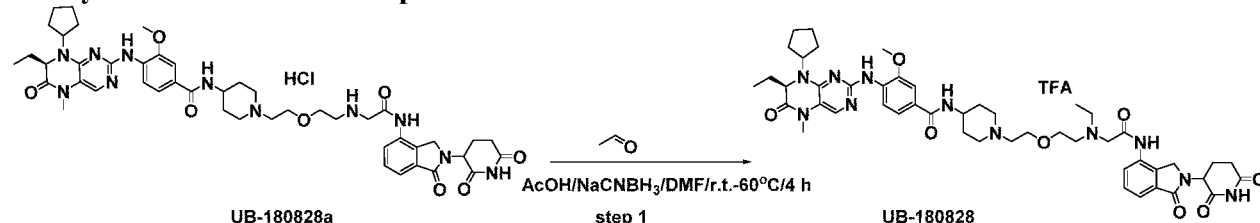
Step 10: UB-180849m

Compound **UBI-180849l** (35 mg, 0.06 mmol), P1 (24.2 mg, 0.06 mmol), and HATU (32 mg, 85.4 μ mol) were dissolved in DMF (2 mL) and DIPEA (0.3 mL), and reacted at room temperature for 18 hours. The reaction was quenched with water (20 mL), then extracted with ethyl acetate (20 mL*3). The organic phases were combined, then dried over anhydrous sodium sulfate and concentrated to obtain crude product. The crude product was isolated by silica gel column chromatography (methanol/dichloromethane=10%) to obtain **UB-180849m** (20 mg, yield 34%) as yellow solid. LCMS $[M+H]^+ = 1022.6$

Step 11: UB-180849

A solution of **UB-180849m** (20 mg, 20 μ mol) and *p*-toluenesulfonic acid monohydrate (5 mg) in methanol (2 ml) was stirred at room temperature for 6 hours, and added saturated aqueous sodium bicarbonate solution (1 ml), concentrated under reduced pressure to obtain crude product. The crude product was purified by reversed-phase column (methanol/water = 5% to 95%, collected at 65%) to obtain **UB-180849** (3mg, yield 16%) as yellow solid. ^1H NMR (400 MHz, Methanol- d_4) δ 8.48 (d, $J = 8.4$ Hz, 1H), 7.86 (dd, $J = 16.4, 7.8$ Hz, 1H), 7.78 (s, 1H), 7.64 (dd, $J = 11.9, 7.5$ Hz, 1H), 7.57 – 7.50 (m, 1H), 7.47 (d, $J = 7.8$ Hz, 2H), 4.51 (d, $J = 9.2$ Hz, 2H), 4.28 (dd, $J = 7.7, 3.6$ Hz, 1H), 3.99 (s, 3H), 3.92 (ddd, $J = 17.3, 11.7, 5.4$ Hz, 2H), 3.65 (dt, $J = 11.0, 5.2$ Hz, 5H), 3.59 (s, 3H), 3.52 (d, $J = 10.9$ Hz, 2H), 3.04 (d, $J = 12.3$ Hz, 2H), 2.88 (ddt, $J = 16.5, 11.2, 5.2$ Hz, 5H), 2.66 (dt, $J = 11.7, 5.6$ Hz, 2H), 2.49 (dt, $J = 19.8, 6.8$ Hz, 2H), 2.26 – 2.10 (m, 4H), 2.00 – 1.80 (m, 8H), 1.78 – 1.67 (m, 4H), 0.86 (t, $J = 7.5$ Hz, 3H)

Synthesis method of Compound UB-180828

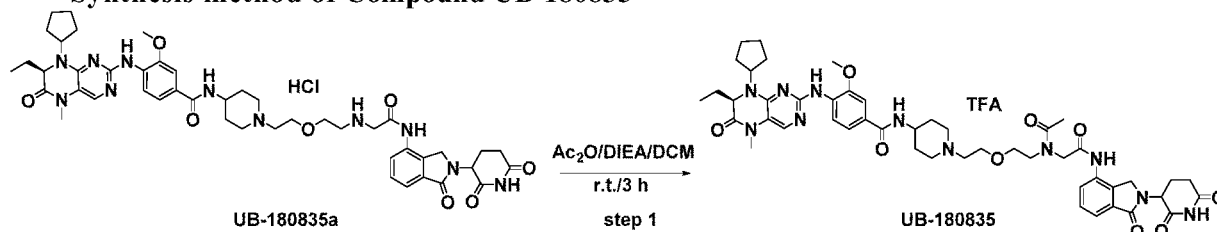


Step 1: UB-180828

Compound **UB-180828a** (34 mg, 0.036 mmol), acetaldehyde (0.02 mL, 0.1 mmol), glacial acetic acid (9 mg, 0.072 mmol) were dissolved in DMF (3 mL) and reacted for 1 hour at room temperature. Then NaBH_3CN (6.3 mg, 0.1 mmol) was added and reacted at 60°C for 3 hours. The reaction solution was prepared and purified to obtain product **UB-180828** (15 mg, yield 40%) as white solid. LCMS $[M+H]^+ =$

¹H NMR (400 MHz, DMSO-*d*₆) δ 11.05 (s, 1H), 10.55 (s, 1H), 9.69 (s, 3H), 8.37 (s, 1H), 8.21 (s, 1H), 7.88 (d, *J* = 7.6 Hz, 1H), 7.81 (s, 1H), 7.60 (t, *J* = 5.8 Hz, 2H), 7.51 (d, *J* = 7.2 Hz, 2H), 5.32 (t, *J* = 4.8 Hz, 1H), 5.19 (dd, *J* = 13.2, 5.2 Hz, 1H), 4.47 – 4.32 (m, 4H), 4.27 (d, *J* = 8.8 Hz, 3H), 4.02 (s, 2H), 3.93 (s, 3H), 3.79 (s, 5H), 3.24 (s, 3H), 2.66 (dt, *J* = 7.5, 2.5 Hz, 2H), 2.61 (s, 1H), 2.00 (dq, *J* = 13.1, 7.8, 6.2 Hz, 7H), 1.90 – 1.78 (m, 6H), 1.69 (dd, *J* = 14.4, 7.1 Hz, 3H), 1.53 (t, *J* = 6.3 Hz, 2H), 1.46 (d, *J* = 7.1 Hz, 2H), 0.81 (t, *J* = 7.0 Hz, 3H).

Synthesis method of Compound UB-180835

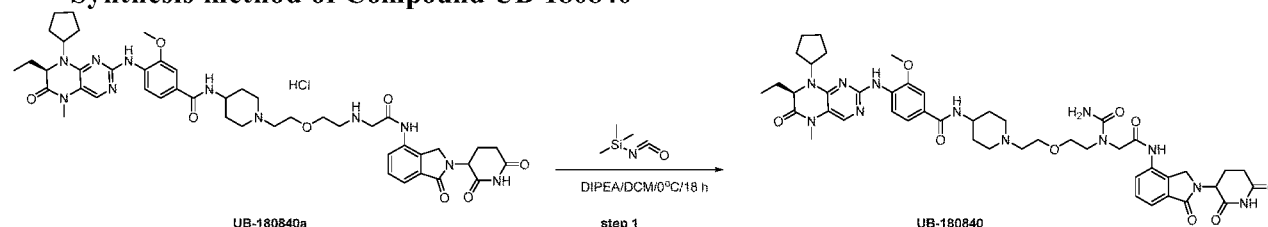


Step 1: UB-180835

Compound **UB-180835a** (26 mg, 0.028 mmol), acetic anhydride (1 drop), DIEA (1 drop) were dissolved in dichloromethane (3 mL) and reacted for 3 hours at room temperature. The reaction solution was prepared and purified to obtain product **UB-180835** (10 mg, yield 30%) as white solid. LCMS [*M*+*H*]⁺ = 936

¹H NMR (400 MHz, DMSO-*d*₆) δ 11.03 (s, 1H), 10.06 (s, 1H), 9.86 (s, 1H), 9.30 (s, 1H), 8.39 (d, *J* = 7.2 Hz, 1H), 8.13 (s, 1H), 7.87 – 7.78 (m, 2H), 7.53 (m, 4H), 5.16 (m, 1H), 4.38 (m, 3H), 4.28 (m, 2H), 4.15 (s, 2H), 4.01 (m, 2H), 3.93 (s, 3H), 3.79 (m, 2H), 3.73 (m, 2H), 3.62 (m, 2H), 3.55 (m, 2H), 3.24 (s, 3H), 3.13 (m, 2H), 2.92 (m, 2H), 2.67 – 2.59 (m, 2H), 2.33 (m, 1H), 2.12 (s, 2H), 2.02 (m, 4H), 1.98 (m, 2H), 1.83 (m, 5H), 1.64 (m, 2H), 1.52 (m, 2H), 0.76 (t, *J* = 7.4 Hz, 3H).

Synthesis method of Compound UB-180840

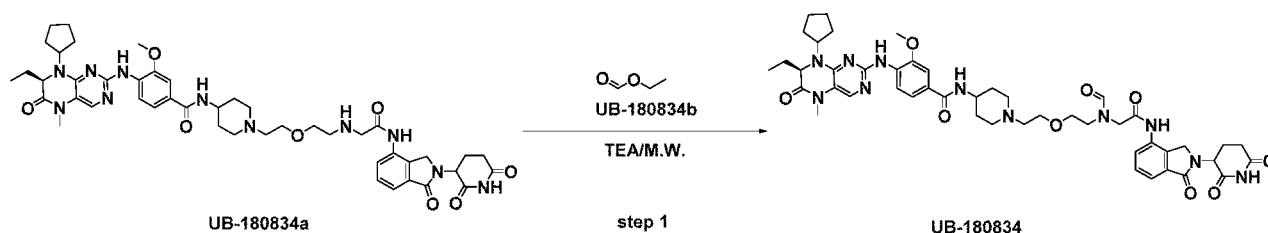


Step 1: UB-180840

Compound **UB-180840a** (30 mg, 0.03 mmol), trimethylsilyl isocyanate (7 mg, 0.06 mmol), DIPEA (7 mg) were dissolved in dichloromethane (10 mL) and reacted at room temperature for 18 hours. Reaction solution was prepared and purified to obtain product **UB-180840** (2 mg, yield 7%) as white solid. LCMS [*M*+*H*]⁺ = 937

¹H NMR (400 MHz, DMSO-*d*₆) δ 11.03 (s, 1H), 9.88 (s, 1H), 9.60 (s, 1H), 8.76 (s, 1H), 8.43 (d, *J* = 7.3 Hz, 1H), 8.07 (d, *J* = 8.5 Hz, 1H), 7.90 – 7.78 (m, 2H), 7.58 – 7.46 (m, 4H), 6.03 (s, 2H), 5.16 (dd, *J* = 13.3, 5.2 Hz, 1H), 4.46 – 4.34 (m, 3H), 4.32 – 4.19 (m, 2H), 4.11 (s, 2H), 4.07 – 3.96 (m, 2H), 3.93 (s, 3H), 3.78 (t, *J* = 5.0 Hz, 2H), 3.60 – 3.57 (m, 4H), 3.23 (s, 3H), 3.13 (s, 3H), 2.97 – 2.88 (m, 2H), 2.63 (d, *J* = 3.7 Hz, 1H), 2.33 (d, *J* = 3.8 Hz, 1H), 2.02 (ddd, *J* = 18.7, 11.1, 4.6 Hz, 7H), 1.84 (dq, *J* = 10.8, 3.8 Hz, 5H), 1.62 (s, 2H), 1.50 (t, *J* = 7.1 Hz, 2H), 0.76 (t, *J* = 7.4 Hz, 3H).

Synthesis method of Compound UB-180834(699-CHO)

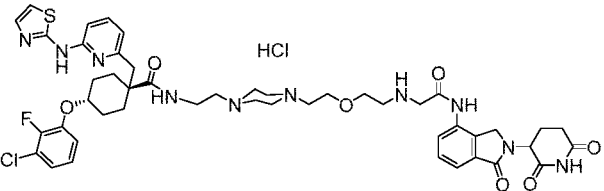
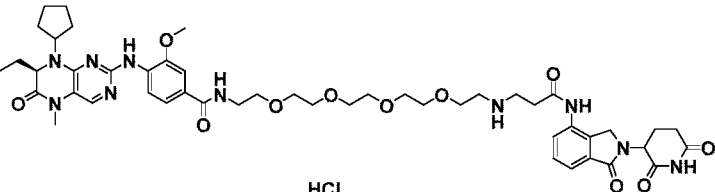
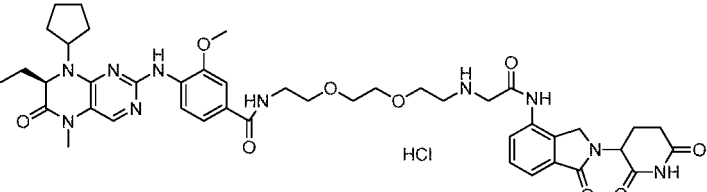
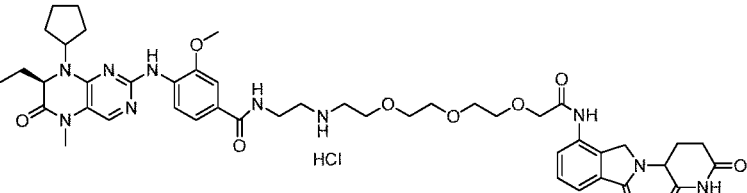
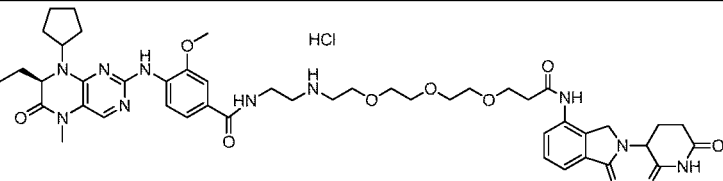


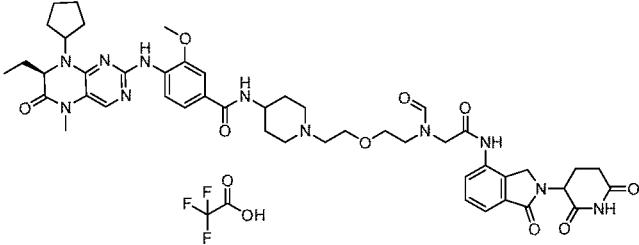
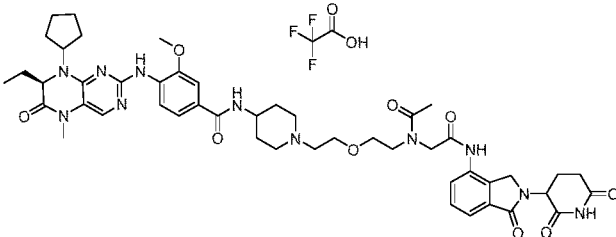
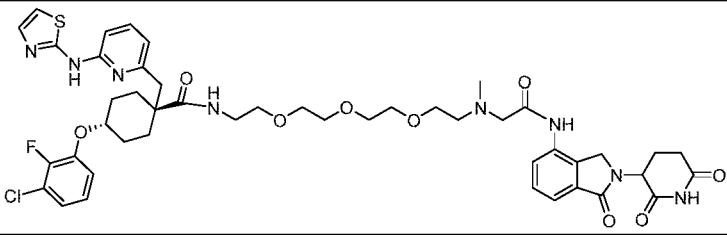
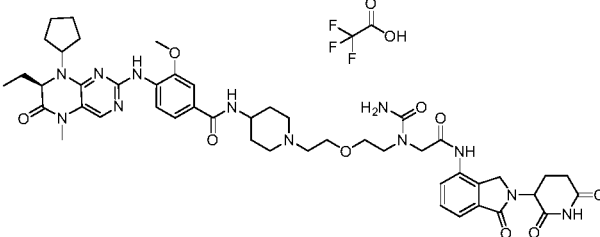
Step 1: UB-180834

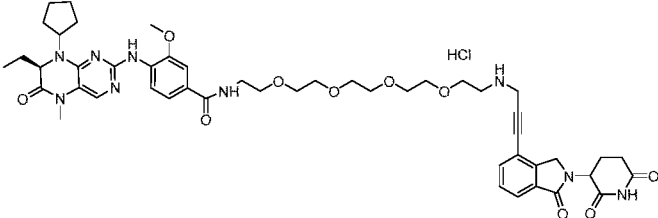
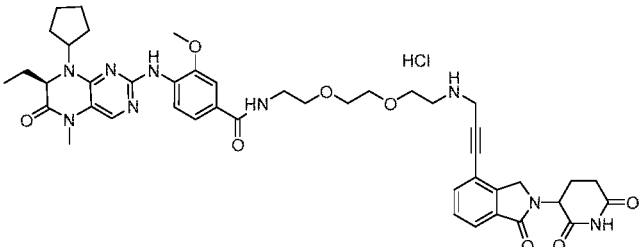
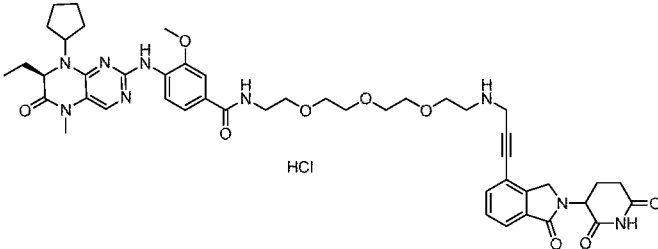
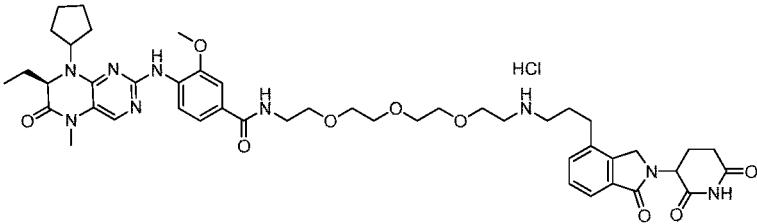
Compound **UB-180834a** (30 mg, 0.03 mmol) was dissolved in **UB-180834b** (5 mL), and then added triethylamine (5 mg, 0.05 mmol). The reaction was heated to 60 °C under microwave and reacted for 1 hour. The reaction solution was concentrated and prepared to obtain target product **UB-180834** (14.2 mg, yield 46%) as white solid.

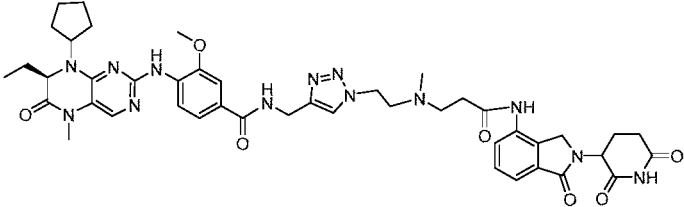
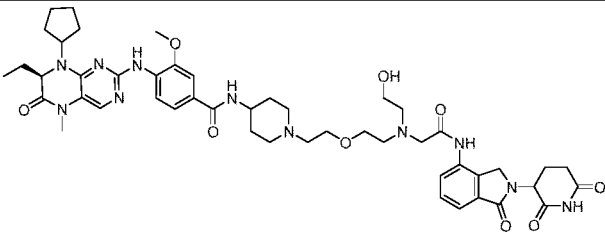
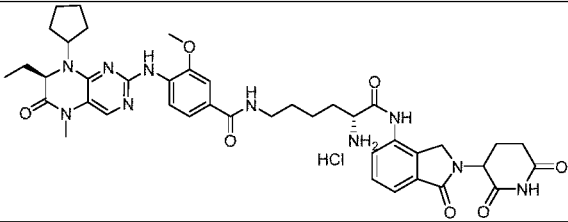
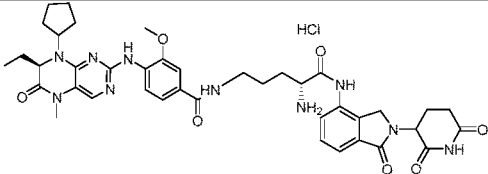
Table B11

Cmpd. No.	Structure and data analysis
UB-180809 (809)	^1H NMR (400 MHz, DMSO-d_6) δ 11.05 (s, 1H), 10.71 (s, 1H), 10.17 (s, 1H), 9.15 (s, 2H), 8.73 (s, 1H), 8.43 (t, J = 5.6 Hz, 1H), 8.30 (dd, J = 8.5, 2.7 Hz, 1H), 7.94 – 7.81 (m, 7H), 7.61 – 7.49 (m, 3H), 7.34 (d, J = 2.2 Hz, 1H), 7.17 (t, J = 8.8 Hz, 2H), 5.17 (dd, J = 13.3, 5.1 Hz, 1H), 4.52 – 4.33 (m, 2H), 4.06 (t, J = 5.6 Hz, 2H), 3.75 (d, J = 5.2 Hz, 2H), 3.56 (d, J = 6.0 Hz, 2H), 3.44 (q, J = 6.5, 6.1 Hz, 2H), 3.23 (q, J = 5.2 Hz, 2H), 3.01 – 2.86 (m, 2H), 2.67 – 2.58 (m, 1H), 2.26 (tt, J = 13.2, 6.5 Hz, 1H), 2.10 – 1.99 (m, 1H). LCMS $[\text{M}+\text{H}]^+ = 906.5$
UB-180810 (810)	^1H NMR (400 MHz, DMSO-d_6) δ 11.92 (s, 1H), 11.05 (s, 1H), 10.72 (s, 1H), 9.16 (s, 2H), 7.87 (dd, J = 7.5, 1.5 Hz, 1H), 7.74 (t, J = 5.5 Hz, 1H), 7.64 (t, J = 7.8 Hz, 1H), 7.60 – 7.51 (m, 2H), 7.46 (d, J = 3.8 Hz, 1H), 7.21 – 7.07 (m, 4H), 7.00 (d, J = 8.2 Hz, 1H), 6.75 (d, J = 7.4 Hz, 1H), 5.17 (dd, J = 13.3, 5.2 Hz, 1H), 4.54 (s, 1H), 4.50 – 4.33 (m, 2H), 4.06 (s, 2H), 3.59 – 3.52 (m, 5H), 3.37 (t, J = 6.3 Hz, 2H), 3.20 (dd, J = 12.0, 6.2 Hz, 4H), 2.64 (d, J = 3.6 Hz, 1H), 2.35 – 2.20 (m, 1H), 2.10 – 2.00 (m, 1H), 1.96 – 1.58 (m, 9H). LCMS $[\text{M}+\text{H}]^+ = 891.4$
UB-180811 (811)	^1H NMR (400 MHz, DMSO-d_6) δ 11.05 (s, 1H), 10.95 (s, 1H), 10.23 (s, 1H), 9.45 (s, 2H), 8.77 (m, 2H), 8.30 (d, J = 8.5 Hz, 1H), 8.04 – 7.83 (m, 6H), 7.55 (q, J = 7.3 Hz, 3H), 7.35 (d, J = 2.2 Hz, 1H), 7.18 (t, J = 8.8 Hz, 2H), 5.17 (dd, J = 13.2, 5.2 Hz, 1H), 4.54 (d, J = 17.8 Hz, 1H), 4.41 (d, J = 17.7 Hz, 1H), 4.15 (d, J = 4.8 Hz, 2H), 3.96 (m, 4H), 3.80 (m, 4H), 3.72 (m, 8H), 3.50 (m, 2H), 3.40 – 3.17 (m, 4H), 2.94 (m, 1H), 2.64 (m, 1H), 2.37 – 2.26 (m, 1H), 2.06 (m, 1H). LCMS $[\text{M}+\text{H}]^+ = 974.4$

UB-180812 (812)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.04 (s, 1H), 10.91 (s, 1H), 9.43 (s, 1H), 8.12 (s, 1H), 7.98 (d, J = 8.4 Hz, 0.5H), 7.88 (d, J = 7.3 Hz, 1H), 7.72 (m, 1.5H), 7.60 – 7.39 (m, 4H), 7.20 – 7.05 (m, 5H), 6.82 (d, J = 7.4 Hz, 1H), 5.16 (dd, J = 13.3, 5.2 Hz, 1H), 4.60 – 4.49 (m, 2H), 4.40 (d, J = 17.7 Hz, 1H), 4.13 (s, 2H), 3.82 (m, 12H), 3.24 (m, 8H), 3.00 (s, 2H), 2.96 – 2.89 (m, 1H), 2.68 – 2.60 (m, 1H), 2.36 – 2.26 (m, 1H), 2.10 – 1.96 (m, 3H), 1.86 – 1.59 (m, 6H). LCMS $[\text{M}+\text{H}]^+ = 959.4$
UB-180813 (813)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.03 (s, 1H), 10.30 (s, 1H), 9.69 (s, 1H), 9.02 (s, 2H), 8.67 (t, J = 5.7 Hz, 1H), 7.85 (t, J = 4.1 Hz, 3H), 7.63 (s, 1H), 7.61 – 7.39 (m, 3H), 5.15 (dd, J = 13.3, 5.1 Hz, 1H), 4.59 – 4.26 (m, 3H), 4.16 (t, J = 8.8 Hz, 1H), 3.90 (s, 3H), 3.72 (t, J = 5.2 Hz, 2H), 3.59 – 3.49 (m, 13H), 3.29 – 3.19 (m, 5H), 3.14 (q, J = 5.7 Hz, 3H), 2.92 (t, J = 7.3 Hz, 3H), 2.62 (d, J = 17.3 Hz, 1H), 2.33 (td, J = 13.3, 8.7 Hz, 1H), 2.10 – 1.76 (m, 7H), 1.60 – 1.37 (m, 4H), 1.29 (dd, J = 13.5, 6.7 Hz, 1H), 0.75 (t, J = 7.4 Hz, 3H). LCMS $[\text{M}+1]^+ = 957.7$
UB-180814 (814)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.04 (s, 1H), 10.85 (s, 1H), 9.72 (s, 1H), 9.23 (d, J = 7.8 Hz, 2H), 8.74 (d, J = 5.8 Hz, 1H), 8.01 – 7.76 (m, 3H), 7.65 (d, J = 1.8 Hz, 1H), 7.62 – 7.41 (m, 3H), 5.16 (dd, J = 13.3, 5.1 Hz, 1H), 4.56 – 4.32 (m, 3H), 4.15 (t, J = 8.8 Hz, 1H), 4.07 (t, J = 5.1 Hz, 2H), 3.90 (s, 3H), 3.75 (t, J = 5.1 Hz, 2H), 3.59 (d, J = 11.7 Hz, 5H), 3.22 (m, 4H), 2.94 (ddd, J = 17.8, 13.4, 5.3 Hz, 1H), 2.67 – 2.58 (m, 1H), 2.28 (qd, J = 13.2, 4.5 Hz, 1H), 2.16 – 1.67 (m, 7H), 1.60 – 1.38 (m, 4H), 1.39 – 1.26 (m, 3H), 0.75 (t, J = 7.4 Hz, 3H). LCMS $[\text{M}+1]^+ = 855.6$
UB-180815 (815)	 ^1H NMR (400 MHz, DMSO-d_6) δ 11.01 (s, 1H), 9.78 (s, 1H), 8.94 (d, J = 28.3 Hz, 3H), 7.98 (d, J = 7.3 Hz, 1H), 7.84 (s, 1H), 7.75 (dd, J = 7.7, 1.3 Hz, 1H), 7.69 (d, J = 1.9 Hz, 1H), 7.63 – 7.47 (m, 3H), 5.15 (dd, J = 13.3, 5.2 Hz, 1H), 4.52 – 4.28 (m, 3H), 4.21 (t, J = 8.3 Hz, 1H), 4.15 (s, 2H), 3.93 (s, 3H), 3.69 (dd, J = 5.9, 3.5 Hz, 3H), 3.61 (d, J = 11.1 Hz, 8H), 3.22 (s, 3H), 3.16 (q, J = 7.1, 6.1 Hz, 4H), 2.62 (dd, J = 20.8, 16.9 Hz, 1H), 2.38 – 2.30 (m, 1H), 2.07 – 1.71 (m, 8H), 1.67 – 1.36 (m, 4H), 1.36 – 1.26 (m, 1H), 0.75 (t, J = 7.4 Hz, 3H). LCMS $[\text{M}+1]^+ = 899.7$
UB-180820 (820)	

	¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 9.95 (d, <i>J</i> = 9.4 Hz, 1H), 9.47 (d, <i>J</i> = 27.8 Hz, 1H), 9.12 (s, 1H), 8.44 (d, <i>J</i> = 7.2 Hz, 1H), 7.96 (d, <i>J</i> = 8.2 Hz, 1H), 7.84 (t, <i>J</i> = 6.3 Hz, 1H), 7.79 (s, 1H), 7.53 (dd, <i>J</i> = 8.8, 6.7 Hz, 4H), 5.16 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.43 (m, 1H), 4.40 – 4.31 (m, 2H), 4.17 (m, 4H), 4.05 – 3.98 (m, 2H), 3.92 (s, 3H), 3.76 (m, 4H), 3.64 (m, 2H), 3.58 (m, 1H), 3.52 (m, 2H), 3.28 (m, 2H), 3.23 (s, 3H), 3.14 (m, 2H), 2.97 – 2.88 (m, 1H), 2.63 (m, 1H), 2.38 – 2.29 (m, 1H), 2.08 – 1.91 (m, 5H), 1.88 – 1.75 (m, 5H), 1.61 – 1.42 (m, 4H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 952.8
UB-180834 (834)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 10.12 (s, 1H), 9.99 (s, 1H), 9.35 (s, 1H), 8.40 (d, <i>J</i> = 7.2 Hz, 1H), 8.18 – 8.03 (m, 2H), 7.90 – 7.73 (m, 2H), 7.53 (t, <i>J</i> = 8.1 Hz, 4H), 5.16 (dd, <i>J</i> = 13.2, 4.7 Hz, 1H), 4.40 – 4.35 (m, 2H), 4.28 (s, 1H), 4.18 (s, 1H), 4.00 (s, 1H), 3.93 (s, 3H), 3.76 (dt, <i>J</i> = 12.0, 4.9 Hz, 3H), 3.64 – 3.52 (m, 4H), 3.24 (s, 3H), 3.14 (d, <i>J</i> = 10.9 Hz, 2H), 2.93 (ddd, <i>J</i> = 18.1, 13.5, 5.2 Hz, 2H), 2.63 (t, <i>J</i> = 15.5 Hz, 2H), 2.33 (t, <i>J</i> = 6.6 Hz, 2H), 2.01 (ddd, <i>J</i> = 21.6, 13.0, 6.1 Hz, 5H), 1.92 – 1.76 (m, 4H), 1.64 (s, 2H), 1.52 (d, <i>J</i> = 7.4 Hz, 2H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+1]⁺ = 922.8
UB-180835 (835)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 10.06 (s, 1H), 9.86 (s, 1H), 9.30 (s, 1H), 8.39 (d, <i>J</i> = 7.2 Hz, 1H), 8.13 (s, 1H), 7.87 – 7.78 (m, 2H), 7.53 (m, 4H), 5.16 (m, 1H), 4.38 (m, 3H), 4.28 (m, 2H), 4.15 (s, 2H), 4.01 (m, 2H), 3.93 (s, 3H), 3.79 (m, 2H), 3.73 (m, 2H), 3.62 (m, 2H), 3.55 (m, 2H), 3.24 (s, 3H), 3.13 (m, 2H), 2.92 (m, 2H), 2.67 – 2.59 (m, 2H), 2.33 (m, 1H), 2.12 (s, 2H), 2.02 (m, 4H), 1.98 (m, 2H), 1.83 (m, 5H), 1.64 (m, 2H), 1.52 (m, 2H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 936
UB-180836 (836)	 ¹H NMR (400 MHz, Methanol-<i>d</i>₄) δ 7.86 – 7.79 (m, 1H), 7.64 – 7.59 (m, 1H), 7.54 – 7.46 (m, 2H), 7.29 (dd, <i>J</i> = 3.7, 2.1 Hz, 1H), 7.03 (tt, <i>J</i> = 5.2, 2.2 Hz, 3H), 6.88 (dd, <i>J</i> = 3.7, 1.2 Hz, 1H), 6.82 (dd, <i>J</i> = 8.1, 3.9 Hz, 1H), 6.70 – 6.63 (m, 1H), 4.73 – 4.65 (m, 2H), 4.63 – 4.47 (m, 4H), 3.72 (d, <i>J</i> = 0.8 Hz, 1H), 3.63 – 3.55 (m, 8H), 3.52 – 3.46 (m, 4H), 3.41 – 3.37 (m, 2H), 3.28 (d, <i>J</i> = 5.6 Hz, 2H), 2.97 (s, 2H), 2.70 (t, <i>J</i> = 5.0 Hz, 2H), 2.37 (d, <i>J</i> = 3.4 Hz, 5H), 2.31 – 2.12 (m, 2H), 2.04 – 1.87 (m, 6H), 1.75 – 1.59 (m, 2H). LCMS [M+H]⁺ = 949.6
UB-180840 (840)	

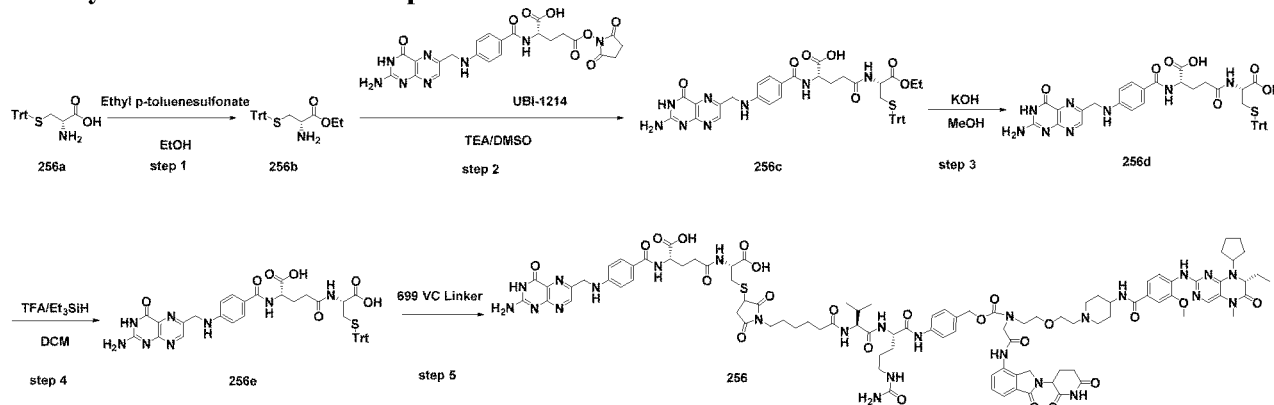
	¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 9.88 (s, 1H), 9.60 (s, 1H), 8.76 (s, 1H), 8.43 (d, <i>J</i> = 7.3 Hz, 1H), 8.07 (d, <i>J</i> = 8.5 Hz, 1H), 7.90 – 7.78 (m, 2H), 7.58 – 7.46 (m, 4H), 6.03 (s, 2H), 5.16 (dd, <i>J</i> = 13.3, 5.2 Hz, 1H), 4.46 – 4.34 (m, 3H), 4.32 – 4.19 (m, 2H), 4.11 (s, 2H), 4.07 – 3.96 (m, 2H), 3.93 (s, 3H), 3.78 (t, <i>J</i> = 5.0 Hz, 2H), 3.60 – 3.57 (m, 4H), 3.23 (s, 3H), 3.13 (s, 3H), 2.97 – 2.88 (m, 2H), 2.63 (d, <i>J</i> = 3.7 Hz, 1H), 2.33 (d, <i>J</i> = 3.8 Hz, 1H), 2.02 (ddd, <i>J</i> = 18.7, 11.1, 4.6 Hz, 7H), 1.84 (dq, <i>J</i> = 10.8, 3.8 Hz, 5H), 1.62 (s, 2H), 1.50 (t, <i>J</i> = 7.1 Hz, 2H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 937
UB-180844 (844)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.05 (s, 1H), 9.59 (m, 3H), 8.63 (t, <i>J</i> = 5.5 Hz, 1H), 7.91 – 7.79 (m, 3H), 7.74 (d, <i>J</i> = 7.5 Hz, 1H), 7.63 – 7.57 (m, 2H), 7.54 (dd, <i>J</i> = 8.3, 1.8 Hz, 1H), 5.18 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.57 (d, <i>J</i> = 18.0 Hz, 1H), 4.47 (m, 1H), 4.38 (d, <i>J</i> = 17.9 Hz, 1H), 4.19 (m, 3H), 3.90 (s, 3H), 3.75 (t, <i>J</i> = 5.1 Hz, 2H), 3.58 (m, 2H), 3.56 (m, 2H), 3.53 (m, 4H), 3.51 (s, 3H), 3.25 (m, 2H), 3.22 (s, 3H), 2.95 (m, 1H), 2.62 (m, 1H), 2.42 – 2.35 (m, 1H), 2.09 – 1.73 (m, 8H), 1.57 – 1.41 (m, 4H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 924
UB-180845 (845)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.05 (s, 1H), 9.37 (m, 3H), 8.59 (m, 1H), 7.97 (m, 1H), 7.85 – 7.71 (m, 3H), 7.64 – 7.49 (m, 3H), 5.18 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.57 – 4.49 (m, 1H), 4.39 (t, <i>J</i> = 15.8 Hz, 2H), 4.22 (m, 3H), 3.91 (s, 2H), 3.73 (m, 2H), 3.61 (s, 3H), 3.56 (m, 2H), 3.47 – 3.43 (m, 5H), 3.23 (s, 3H), 2.94 (m, 1H), 2.63 (m, 1H), 2.39 (m, 1H), 2.07 – 1.79 (m, 8H), 1.62 – 1.45 (m, 4H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 836
UB-180846 (846)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.04 (s, 1H), 9.79 (s, 2H), 9.51 (s, 1H), 8.68 (d, <i>J</i> = 5.6 Hz, 1H), 7.91 (d, <i>J</i> = 8.4 Hz, 1H), 7.87 (s, 1H), 7.81 (d, <i>J</i> = 7.6 Hz, 1H), 7.74 (d, <i>J</i> = 7.5 Hz, 1H), 7.65 – 7.42 (m, 3H), 5.17 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.60 (d, <i>J</i> = 18.1 Hz, 1H), 4.46 (dd, <i>J</i> = 6.6, 3.3 Hz, 1H), 4.38 (d, <i>J</i> = 18.1 Hz, 1H), 4.19 (d, <i>J</i> = 10.0 Hz, 3H), 3.90 (s, 3H), 3.76 (t, <i>J</i> = 5.1 Hz, 2H), 3.56 (d, <i>J</i> = 9.0 Hz, 10H), 3.24 (s, 1H), 3.22 (s, 3H), 2.94 (ddd, <i>J</i> = 18.0, 13.6, 5.5 Hz, 1H), 2.63 (dd, <i>J</i> = 18.4, 14.6 Hz, 2H), 2.42 (dd, <i>J</i> = 13.3, 4.5 Hz, 1H), 2.07 – 1.77 (m, 8H), 1.55 – 1.45 (m, 4H), 1.35 – 1.25 (m, 1H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+1]⁺ = 880.7
UB-180847 (847)	

	¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (s, 1H), 9.62 (s, 1H), 9.23 (s, 0H), 8.98 (s, 2H), 8.67 (t, <i>J</i> = 5.6 Hz, 1H), 7.92 – 7.81 (m, 2H), 7.67 – 7.57 (m, 2H), 7.55 (dd, <i>J</i> = 8.3, 1.8 Hz, 1H), 7.51 – 7.45 (m, 2H), 5.15 (dd, <i>J</i> = 13.3, 5.1 Hz, 1H), 4.58 – 4.41 (m, 2H), 4.32 (d, <i>J</i> = 17.2 Hz, 1H), 4.17 (t, <i>J</i> = 8.8 Hz, 1H), 3.90 (s, 3H), 3.82 – 3.64 (m, 3H), 3.65 – 3.58 (m, 3H), 3.46 – 3.37 (m, 5H), 3.22 (s, 3H), 3.11 (ddt, <i>J</i> = 12.3, 9.1, 4.9 Hz, 3H), 2.94 (td, <i>J</i> = 16.1, 14.4, 5.3 Hz, 4H), 2.73 (t, <i>J</i> = 7.7 Hz, 2H), 2.63 (dd, <i>J</i> = 17.4, 13.8 Hz, 2H), 2.45 – 2.26 (m, 1H), 2.11 – 1.70 (m, 9H), 1.56 – 1.43 (m, 4H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+1]⁺ = 884.7
UB-180848 (848)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.01 (s, 1H), 9.83 (d, <i>J</i> = 19.2 Hz, 1H), 8.87 (s, 1H), 8.42 (d, <i>J</i> = 8.8 Hz, 1H), 7.95 (d, <i>J</i> = 5.3 Hz, 1H), 7.82 (d, <i>J</i> = 13.2 Hz, 2H), 7.60 (s, 1H), 7.53 – 7.38 (m, 4H), 5.21 – 4.73 (m, 1H), 4.42 (s, 2H), 4.33 (d, <i>J</i> = 7.7 Hz, 2H), 4.24 (dd, <i>J</i> = 7.5, 3.5 Hz, 1H), 3.92 (d, <i>J</i> = 4.8 Hz, 3H), 2.93 – 2.67 (m, 4H), 2.27 (d, <i>J</i> = 6.0 Hz, 3H), 2.09 – 1.84 (m, 5H), 1.78 (s, 4H), 1.67 – 1.49 (m, 4H), 0.76 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 876.9
UB-180849 (849)	 ¹H NMR (400 MHz, Methanol-<i>d</i>₄) δ 8.48 (d, <i>J</i> = 8.4 Hz, 1H), 7.86 (dd, <i>J</i> = 16.4, 7.8 Hz, 1H), 7.78 (s, 1H), 7.64 (dd, <i>J</i> = 11.9, 7.5 Hz, 1H), 7.57 – 7.50 (m, 1H), 7.47 (d, <i>J</i> = 7.8 Hz, 2H), 4.51 (d, <i>J</i> = 9.2 Hz, 2H), 4.28 (dd, <i>J</i> = 7.7, 3.6 Hz, 1H), 3.99 (s, 3H), 3.92 (ddd, <i>J</i> = 17.3, 11.7, 5.4 Hz, 2H), 3.65 (dt, <i>J</i> = 11.0, 5.2 Hz, 5H), 3.59 (s, 3H), 3.52 (d, <i>J</i> = 10.9 Hz, 2H), 3.04 (d, <i>J</i> = 12.3 Hz, 2H), 2.88 (ddt, <i>J</i> = 16.5, 11.2, 5.2 Hz, 5H), 2.66 (dt, <i>J</i> = 11.7, 5.6 Hz, 2H), 2.49 (dt, <i>J</i> = 19.8, 6.8 Hz, 2H), 2.26 – 2.10 (m, 4H), 2.00 – 1.80 (m, 8H), 1.78 – 1.67 (m, 4H), 0.86 (t, <i>J</i> = 7.5 Hz, 3H). LCMS [M+H]⁺ = 938.7
UB-180855 (855)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.03 (s, 1H), 9.49 (s, 1H), 8.62 (m, 1H), 8.45 (m, 2H), 7.90 – 7.71 (m, 3H), 7.63 – 7.46 (m, 4H), 5.16 (dd, <i>J</i> = 13.1, 5.2 Hz, 1H), 4.64 – 4.33 (m, 3H), 4.17 (t, <i>J</i> = 8.6 Hz, 2H), 3.89 (s, 3H), 3.22 (s, 3H), 2.99 – 2.83 (m, 1H), 2.62 (d, <i>J</i> = 17.0 Hz, 1H), 2.32 (dt, <i>J</i> = 12.9, 6.4 Hz, 1H), 2.08 – 1.71 (m, 11H), 1.63 – 1.38 (m, 8H), 0.76 (t, <i>J</i> = 7.3 Hz, 3H). LCMS [M+H]⁺ = 795.6
UB-180856 (856)	 ¹H NMR (400 MHz, DMSO-<i>d</i>₆) δ 11.02 (d, <i>J</i> = 3.3 Hz, 1H), 8.94 – 8.68 (m, 1H), 8.42 (s, 3H), 8.07 – 7.73 (m, 3H), 7.71 – 7.31 (m, 4H), 5.15 (dt, <i>J</i> = 12.0, 5.7 Hz, 1H), 4.55 – 4.37 (m, 2H), 4.18 (dd, <i>J</i> = 21.9, 12.6 Hz, 2H), 3.89 (d, <i>J</i> = 1.9 Hz, 3H), 3.49 (d, <i>J</i> = 2.4 Hz, 1H), 3.22 (s, 3H), 3.00 – 2.84 (m, 1H), 2.59 (m, 1H), 2.28 (p, <i>J</i> = 10.1, 9.4 Hz, 1H), 2.16 – 1.65 (m, 12H), 1.48 (d, <i>J</i> = 35.1 Hz, 5H), 1.24 (s, 2H), 0.75 (t, <i>J</i> = 7.4 Hz, 3H). LCMS [M+H]⁺ = 781.6

Example 3 Synthesis of ACTED Compound

Example 3.1 Synthesis of TED with linker

Synthesis method of Compound 256



Step 1: 256b (V1318-051)

5 Compound **156a** (19 g, 52 mmol) and ethyl *p*-toluenesulfonate (21 g, 105 mmol) were dissolved in ethanol (150 mL) and reacted under reflux for 16 hours. The reaction solution was concentrated to obtain crude product. The crude product was isolated by silica gel column chromatography (ethyl acetate/petroleum ether = 1/3) to obtain target product **256b** (23 g, yield 100 %) as yellow oil. LCMS $[M+H]^+ = 392.0$

10 Step 2: 256c (V1318-053)

Compound **256b** (2.0 g, 3.7 mmol) was dissolved in DMSO (30 mL), then added **UBI-1214** (1.45 g, 3.7 mmol) and TEA (1.9 g, 18.6 mmol), and the reaction was reacted at 25°C for 16 hours. The reaction solution was slurried with ether (50 mL*3), then filtered to obtain target product **156c** (2.3 g, yield 77%) as brown solid. The solid was directly used in the next reaction.

15 Step 3: 256d (V1318-062)

Compound **156c** (3 g, 3.7 mmol) was dissolved in methanol (30 mL) and water (5 mL) and then added KOH (825 mg, 14.7 mmol). The reaction was carried out at 25°C for 18 hours. The crude product was purified by a reversed-phase column (methanol/water=0% to 95%, 45 minutes, collected at 3%) to obtain target product **256d** (750 mg, yield 26 %) as yellow solid. LCMS $[M+H]^+ = 787.2$

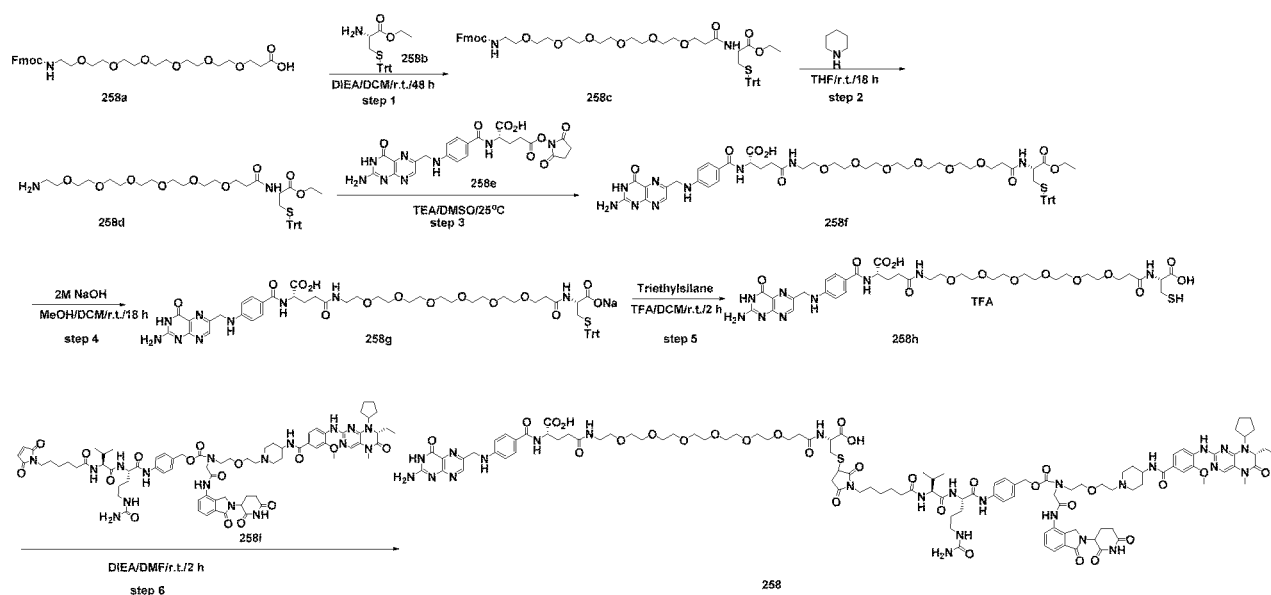
20 Step 4: 256e (V1318-064)

Compound **256d** (100 mg, 0.13 mmol) was dissolved in TFA (3 mL) and dichloromethane (3 mL), followed by adding a catalytic amount of Et_3SiH , and then reacted at room temperature for 1 hour. The reaction solution was concentrated to obtain target crude product **256e** (100 mg, yield 100%) as a yellow solid. The solid was directly used in the next step. LCMS $[M+H]^+ = 545.0$

25 Step 5: 256 (V1318-065)

Compound **156e** (5 mg, 0.11 mmol) was dissolved in *N,N*-dimethylformamide (1 mL) and DIEA (6 mg, 0.04 mmol) and then added **1699-Vc-linker** (10 mg, 0.01 mmol), and then reacted at room temperature for 16 hours. The reaction solution was purified by Prep-HPLC to obtain target product **256** (7.6 mg, yield 40%) as yellow solid. LCMS $[M/2+H]^+ = 1018.8$

30 Synthesis method of Compound 258



Step 1: 258c (V1480-015)

Compound **258a** (2 g, 3.47 mmol), compound **158b** (2 g, 5.21 mmol) and DCC (1.43 g, 6.94 mmol), DIEA (2 mL) were dissolved in dry dichloromethane (50 mL) and reacted at 25°C for 48 hours. After the completion of reaction, water (20 mL) was added, extracted with dichloromethane (20 mL *3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na₂SO₄, concentrated under reduced pressure and isolated by reversed-phase column (MeOH/H₂O = 2 % to 97%) to obtain target product **258c** as yellow transparent oil (1.5 g, yield 45%). LCMS [M+H]⁺ = 949

Step 2: 258d (V1480-018)

Compound **258c** was dissolved in dry tetrahydrofuran (15 mL), added piperidine (9 mL, 90 mmol) and reacted at room temperature for 18 hours. After the completion of reaction, water (20 mL) was added, extracted with ethyl acetate (20 mL *3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na₂SO₄, concentrated under reduced pressure and isolated by silica gel column chromatography (dichloromethane/MeOH=0% to 50%) to obtain target compound **258d** (900 mg, yield 78%) as yellow oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.36 (d, *J* = 7.8 Hz, 1H), 7.42 – 7.20 (m, 15H), 4.10 (td, *J* = 8.1, 5.4 Hz, 1H), 4.02 – 3.97 (m, 2H), 3.58 (t, *J* = 6.5 Hz, 2H), 3.53 – 3.45 (m, 19H), 3.40 (d, *J* = 5.6 Hz, 2H), 2.73 (t, *J* = 5.6 Hz, 2H), 2.35 (tt, *J* = 7.1, 2.9 Hz, 3H), 1.10 (t, *J* = 7.1 Hz, 3H). LCMS [M+H]⁺ = 727

Step 3: 258f (V1480-020)

Compound **258e** (1.65 g, 3.0 mmol), compound **258d** (900 mg, 1.23 mmol) and triethylamine (500 mg) were dissolved in dry DMSO (10 mL) and reacted at 25 °C for 24 hours. Ether (50 mL *3) was added and stirring for 3 minutes. The ether layer was removed and centrifuging with acetonitrile (100 mL). The solid was collected to obtain the target product **258f** (2 g) as red transparent oil. LCMS [M+H]⁺ = 1150.

Step 4: 258g (V1480-022)

Compound **158f** (500 mg, 0.43 μmol) were dissolved in dichloromethane (20 mL) and MeOH (10 mL) and then added 2M NaOH (1 mL). The reaction was reacted at room temperature for 18 hours. The reaction solution was concentrated under reduced pressure and isolated by a reversed-phase column (MeOH/H₂O = 2% to 30%) to obtain target product **258g** (30 mg, purity 70%) as yellow solid. LCMS [M+H]⁺ = 1122

Step 5: 258h (V1480-027)

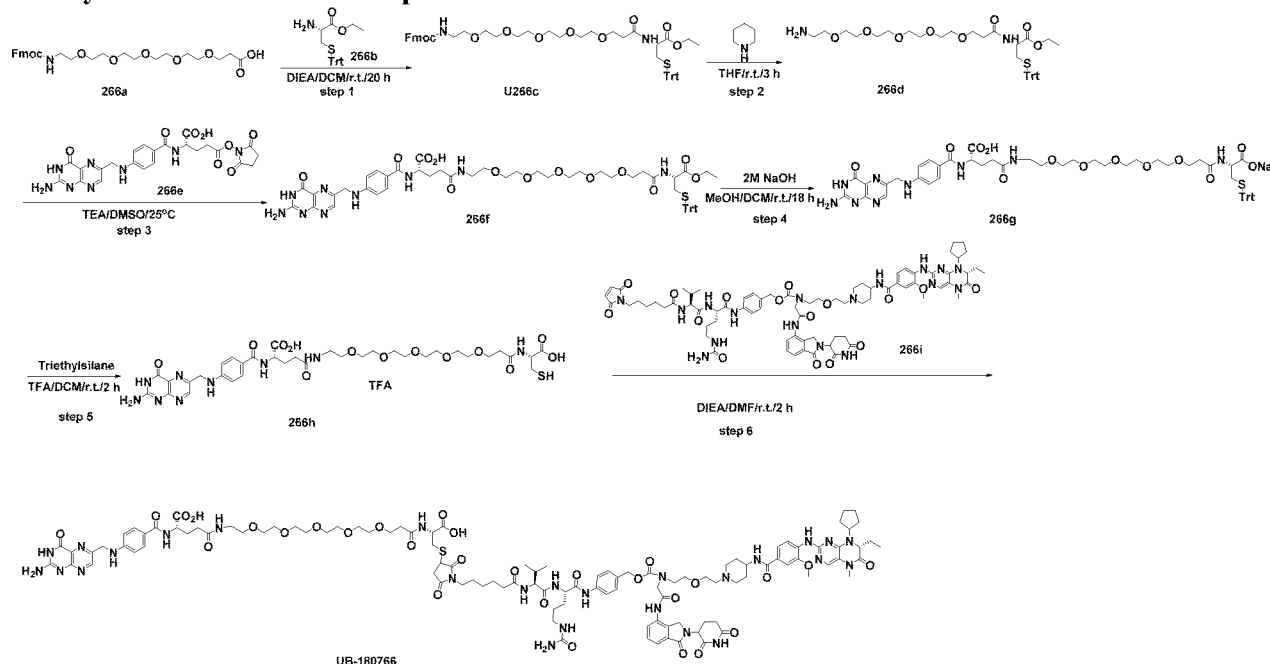
Compound **258g** (40 mg, 0.035 mmol) was dissolved in dry dichloromethane (4 mL) and TFA (0.5 mL) and triethylsilane (3 drops). The reaction was reacted at room temperature for 2 hours. The reaction was concentrated under reduced pressure, added ether (10 mL) and stirred. Solids were formed. The solids

is filtered to obtain yellow target product **258h** (45 mg, purity 70%). ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.65 (d, *J* = 1.6 Hz, 1H), 8.23 – 8.15 (m, 2H), 7.88 (q, *J* = 7.6, 6.6 Hz, 1H), 7.69 – 7.61 (m, 2H), 6.64 (dd, *J* = 8.9, 2.3 Hz, 2H), 4.49 (s, 2H), 4.44 – 4.23 (m, 4H), 3.60 (td, *J* = 6.5, 1.6 Hz, 3H), 3.52 – 3.43 (m, 19H), 3.41 – 3.28 (m, 3H), 3.18 (t, *J* = 5.7 Hz, 2H), 2.89 – 2.79 (m, 1H), 2.78 – 2.65 (m, 2H), 2.43 – 2.35 (m, 3H), 2.26 – 2.13 (m, 2H), 2.10 – 1.80 (m, 3H). LCMS [M+H]⁺ = 880.

Step 6: **258** (V1480-029)

Compound **258h** (7 mg, 8*10⁻³ mmol), compound **258i** (12 mg, 8*10⁻³ mmol), DIEA (5 mg) were dissolved in dry DMF (0.7 mL), reacted at room temperature for 2 hours. The reaction solution was filtered, prepared by Prep-HPLC to obtain target product **258** (10.4 mg, 55% yield) as yellow solid. LCMS [M/2+H]⁺ = 1186

Synthesis method of Compound **266**



Step 1: **266c** (V1480-023)

Compound **266a** (1.5 g, 2.82 mmol), compound **266b** (1.65 g, 4.23 mmol) and (1.16 g, 5.64 mmol), DIEA (1.5 mL) were dissolved in dry dichloromethane (50 mL) and reacted at 25°C for 48 hours. After the completion of reaction, water (20 mL) was added, extracted with dichloromethane (20 mL * 3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na₂SO₄, concentrated under reduced pressure and isolated by reversed-phase column (MeOH/H₂O = 2 % to 97%) to obtain target product **266c** (900 mg, yield 35%) as yellow transparent oil. LCMS [M+H]⁺ = 905

Step 2: **266d** (V1480-026)

Compound **266c** (900 mg, 1 mmol) was dissolved in dry tetrahydrofuran (15 mL), added piperidine (5 mL, 68.8 mmol) and reacted at room temperature for 3 hours. After the completion of reaction, water (20 mL) was added, extracted with ethyl acetate (20 mL * 3). The organic phases were combined, then washed with saturated brine, dried over anhydrous Na₂SO₄, concentrated under reduced pressure and isolated by reversed-phase column (MeOH/H₂O = 2 % to 80%) to obtain target compound **266d** (460 mg, yield 67%) as yellow oil.

LCMS [M+H]⁺ = 683.

Step 3: **266f** (V1480-028)

Compound **266e** (725 mg, 1.34 mmol), compound **266d** (460 mg, 0.67 mmol) and triethylamine (270 mg) were dissolved in dry DMSO (4 mL) and reacted at 25 °C for 18 hours, added ether (50 mL * 3) and

stirred for 3 minutes. The ether layer was removed and centrifuging with acetonitrile (100 mL). The solid was collected to obtain the target product **266f** (1 g) as red transparent oil. LCMS $[M+H]^+ = 1107$

Step 4: **266g** (V1480-030)

Compound **266f** (1 g, 0.9 μ mol) were dissolved in dichloromethane (10 mL) and MeOH (20 mL) and then added 2M NaOH (2 mL). The reaction was reacted at room temperature for 3 hours. The reaction solution was concentrated under reduced pressure and isolated by a reversed-phase column (MeOH/H₂O = 2% to 30%) to obtain target product **266g** (170 mg, yield 17%) as yellow solid. LCMS $[M+H]^+ = 1078$

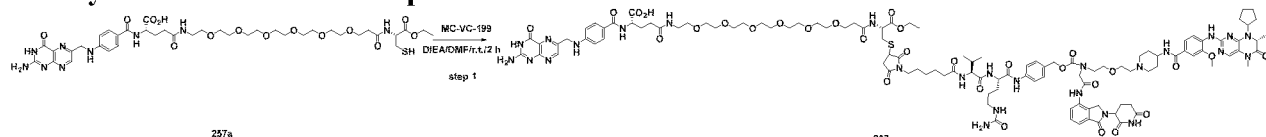
Step 5: **266h** (V1480-031)

Compound **266g** (40 mg, 0.037 mmol) was dissolved in dry dichloromethane (4 mL) and TFA (0.5 mL) and triethylsilane (3 drops). The reaction was reacted at room temperature for 2 hours. The reaction was concentrated under reduced pressure, added ether (10 mL) and stirred. Solid was formed. The solid was filtered to obtain yellow target product **266h** (25 mg, yield 80%). LCMS $[M+H]^+ = 836$

Step 6: **266** (V1480-032)

Compound **266h** (10 mg, 12×10^{-3} mmol), compound **266i** (12 mg, 8×10^{-3} mmol), DIEA (5 mg) were dissolved in dry DMF (0.7 mL), reacted at room temperature for 2 hours. The reaction solution was filtered, then purified by Prep-HPLC to obtain target product **266** (6.2 mg, 33% yield) as yellow solid. LCMS $[M/2+H]^+ = 1165$

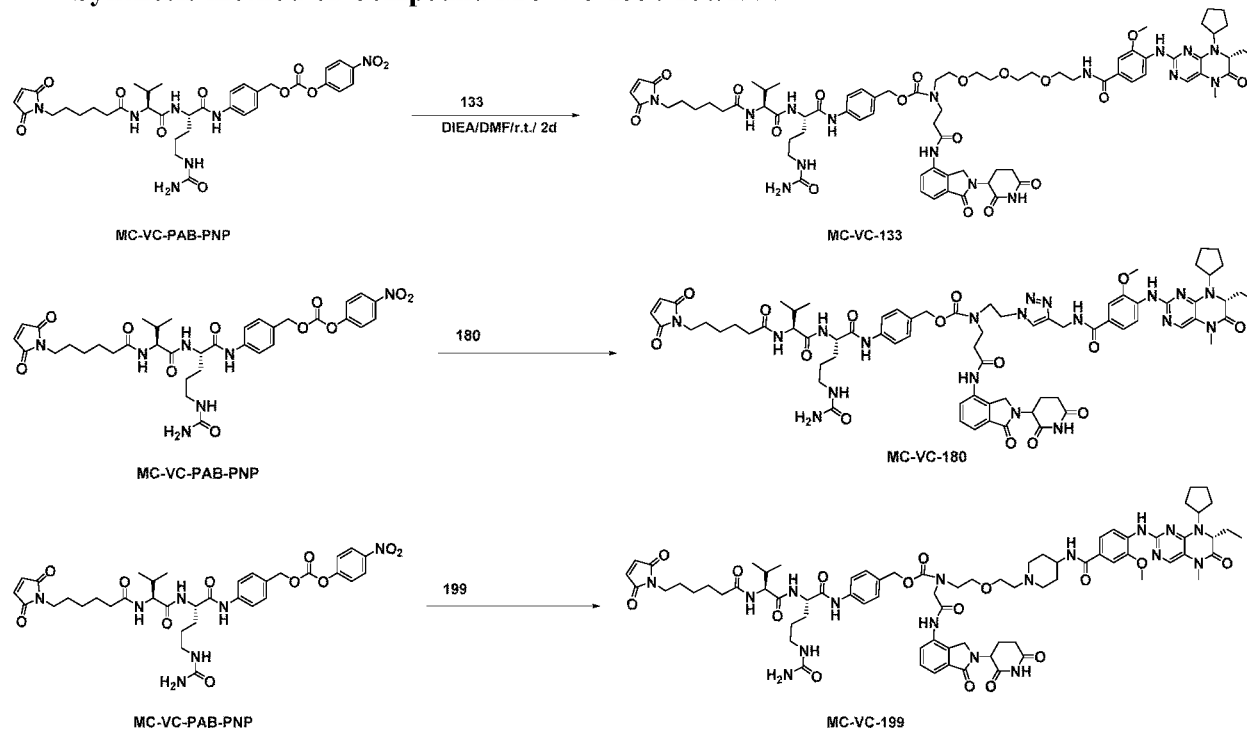
Synthesis method of Compound 157



Step 1: **257** (V1162-111)

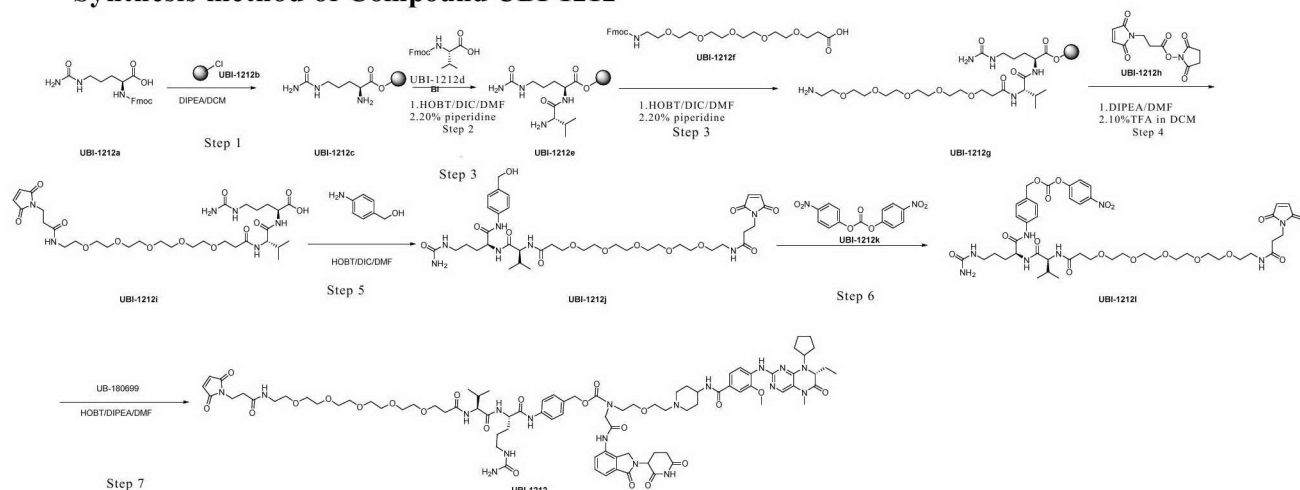
The solution of Compound **257a** (5 mg, 8×10^{-3} mmol), (12 mg, 8×10^{-3} mmol), TEA (5 mg) in N,N-dimethylformamide (0.7 mL) was reacted at room temperature for 2 hours. The reaction solution was isolated and purified by Prep-HPLC to obtain target compound (4.0 mg, yield 30.7%) as yellow solid. LCMS $[M/2+H]^+ = 1200.5$

Synthesis method of Compound MC-VC-133 / 180/199:



Compound 133/ 180/ 199 (40 mg, 1eq.), **MC-VC-PAB-PNP** (1 eq.), **HOBT** (2eq.) were dissolved in **DMF** (0.8 mL) and **DIEA** (2eq.), reacted at room temperature for 48 hours under nitrogen protection. The reaction solution was directly isolated by a reversed-phase preparative chromatography column to obtain the target compound **MC-Vc-133/180/199**.

Synthesis method of Compound UBI-1212



Step 1: UBI-1212c (V1162-120)

MC-Val-Cit-OHCl-TrtCl-resin (UBI-1212b) was placed in the filter, added 10mL of dichloromethane, stirred for one hour and then filtered. Then a solution of **Fmoc-Cit-OH (UBI-1212a)** (600mg, 1.5mmol) and **DIPEA** (0.66ml) in dichloromethane was added, reacted at room temperature for one hour. The reaction solution was added methanol and stirred for 15 minutes, and then filtered to obtain solids. The solids was washed with the following solvents: dichloromethane (5 x 50mL x 0.5 min), *N,N*-dimethylformamide (5 x 50 mL x 0.5 min), piperidine : *N,N*-dimethylformamide (1 :4, 1 x 1 min, 2 x 10 min), *N,N*-dimethylformamide (5 x 50 mL x 0.5 min), dichloromethane (5 x 50 mL x 0.5 min) to obtain the target compound resin (**UBI-1212c**).

Step 2: UBI-1212e (V1162-121)

Compound UBI-1212c washed with *N,N*-dimethylformamide (5 x 50 mL x 0.5 min) and then added **Fmoc-Val-OH (UBI-1212d)** (950mg, 2.8mmol), **HOBT** (567mg, 4.2mmol) in *N,N*-dimethylformamide (20 mL). After stirring for 10 minutes, **DIC** (538 mg, 4.2 mmol) was added and reacted at room temperature for 1.5 hours. The reaction solution was filtered to obtain solids, which was washed with the following solvent: *N,N*-dimethylformamide (5 x 50 mL x 0.5 min), piperidine : *N,N*-dimethylformamide (1 :4, 1 x 1 min, 2 x 10 min), *N,N*-dimethylformamide (5 x 50 mL x 0.5 min) to obtain target compound resin (**UBI-1212e**).

Step 3: UBI-1212g (V1162-122)

Compound UBI-1212e was washed with *N,N*-dimethylformamide (5 x 50 mL x 0.5 min) and then added **UBI-1212f** (740mg, 1.4mmol), **HOBT** (567mg, 4.2mmol) in *N,N*-dimethylformamide (20 mL). After stirring for 10 minutes, **DIC** (538 mg, 4.2 mmol) was added and reacted at room temperature for 1.5 hours. The reaction solution was filtered to obtain solids, which was washed with the following solvent: *N,N*-dimethylformamide (5 x 50 mL x 0.5 min), piperidine : *N,N*-dimethylformamide (1 :4, 1 x 1 min, 2 x 10 min), *N,N*-dimethylformamide (5 x 50 mL x 0.5 min) to obtain target compound resin (**UBI-1212g**).

Step 4: UBI-1212i (V1162-123)

Compound UBI-1212g was washed with *N,N*-dimethylformamide (5 x 50 mL x 0.5 min) and then added **UBI-1212h** (740mg, 2.8mmol) in *N,N*-dimethylformamide (20mL) and **DIPEA** (0.66mL) solution,

reacted at room temperature for 1 hour. The reaction solution was washed with N,N-dimethylformamide (5 x 50 mL x 0.5 min), then added dichloromethane (10 mL) and TFA (1 mL) were added, and reacted at room temperature for 2 hours. The reaction solution was filtered, and the filtrate was concentrated to obtain crude product as oil. Then it was isolated and purified by reversed-phase column (MeOH/H₂O=5% to 95%, collected at 45%, detected by UV 210) to obtain target compound **UBI-1212i** (600mg) as colorless oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.19 (d, *J* = 7.3 Hz, 1H), 8.02 (d, *J* = 5.7 Hz, 1H), 7.83 (d, *J* = 9.1 Hz, 1H), 7.35 – 7.11 (m, 2H), 7.00 (s, 2H), 5.95 (s, 1H), 4.25 (dd, *J* = 9.0, 6.7 Hz, 1H), 4.12 (t, *J* = 7.1 Hz, 1H), 3.81 (m, 8H), 3.59 (t, *J* = 7.3 Hz, 4H), 3.49 (t, *J* = 5.3 Hz, 8H), 3.36 (t, *J* = 5.9 Hz, 2H), 3.15 (q, *J* = 5.8 Hz, 2H), 2.95 (t, *J* = 6.7 Hz, 2H), 2.37 – 2.19 (m, 4H), 2.01 – 1.88 (m, 1H), 1.70 (d, *J* = 12.4 Hz, 1H), 1.60 – 1.52 (m, 1H), 1.43 – 1.30 (m, 2H), 0.84 (dd, *J* = 15.4, 6.8 Hz, 6H). LC- MS: (M+H)⁺=717.5

Step 5: **UBI-1212j** (V1162-129)

A solution of **Compound UBI-1212i** (442 mg, 0.61 mmol), 4-aminobenzyl alcohol (152mg, 1.2 mmol) in N,N-dimethylformamide (4 mL) was added HOBT (166 mg, 1.2 mmol) and DIC (194ul, 1.2mmol). The reaction was reacted at room temperature for 2 hours. The reaction solution was purified and isolated by reversed-phase column (MeOH/H₂O = 5% to 95%, collected at 40%) to obtain **UBI-1212j** (140mg, yield 28%) LC-MS: (M+H)⁺=822.4

Step 6: **UBI-1212l** (V1162-134)

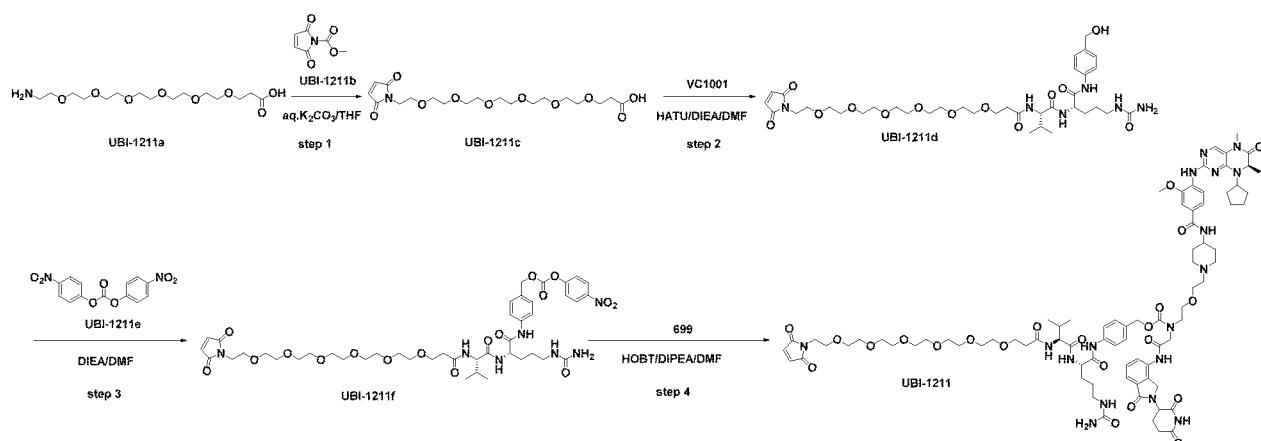
Compound UBI-1212j (380 mg, 0.46 mmol), **UBI-1212k** (281 mg, 0.92 mmol) were added to a solution of DIPEA (0.3 mL) and N,N-dimethylformamide (2 mL), and reacted at room temperature for 18 hours. The reaction solution was purified and isolated by a reversed-phase column (acetonitrile/water = 5% to 50%, collected at 40%) to obtain **UBI-1212l** (130 mg, yield 28.5%) as light yellow solid.

¹H NMR (400 MHz, DMSO-*d*₆) δ 10.06 (s, 1H), 8.38 – 8.23 (m, 2H), 8.14 (d, *J* = 7.4 Hz, 1H), 8.01 (t, *J* = 5.6 Hz, 1H), 7.88 (d, *J* = 8.6 Hz, 1H), 7.70 – 7.63 (m, 2H), 7.60 – 7.53 (m, 2H), 7.44 – 7.37 (m, 2H), 7.00 (s, 2H), 6.15 – 5.96 (m, 1H), 5.25 (s, 2H), 4.98 (s, 4H), 4.43 – 4.36 (m, 1H), 4.24 (dd, *J* = 8.6, 6.6 Hz, 1H), 3.64 – 3.56 (m, 4H), 3.52 – 3.45 (m, 16H), 3.36 (t, *J* = 5.9 Hz, 2H), 3.15 (q, *J* = 5.8 Hz, 2H), 3.06 – 2.91 (m, 2H), 2.33 (dd, *J* = 7.9, 6.7 Hz, 2H), 1.97 (h, *J* = 6.7 Hz, 1H), 1.78 – 1.32 (m, 4H), 0.85 (dd, *J* = 13.3, 6.7 Hz, 6H). LC-MS : (M+H)⁺=987.4

Step 7: **UBI-1212** (V1162-136)

Compound UBI-1212l (40 mg, 0.04 mmol) and compound **199** (37 mg, 0.04 mmol), and HOBT (5.4 mg, 0.04 mmol) were added to a solution of DIEA (15ul, 0.08 mmol) and N,N-dimethylformamide (500uL), reacted at room temperature for 36 hours. The reaction solution was purified by Prep-HPLC to obtain target compound **UBI-1212** (45 mg, yield 65.2 %) as white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.02 (d, *J* = 7.0 Hz, 1H), 10.04 – 9.87 (m, 2H), 9.38 (s, 1H), 8.99 (s, 1H), 8.43 (d, *J* = 7.3 Hz, 1H), 8.10 (d, *J* = 6.7 Hz, 2H), 8.03 – 7.98 (m, 1H), 7.86 (d, *J* = 7.8 Hz, 2H), 7.79 (s, 1H), 7.73 (d, *J* = 7.7 Hz, 1H), 7.62 (d, *J* = 8.3 Hz, 1H), 7.56 – 7.50 (m, 6H), 7.34 (d, *J* = 8.4 Hz, 1H), 7.25 (d, *J* = 8.2 Hz, 1H), 7.00 (s, 2H), 6.00 (s, 2H), 5.14 (td, *J* = 12.6, 5.1 Hz, 1H), 5.04 (d, *J* = 14.2 Hz, 1H), 4.44 – 4.12 (m, 2H), 3.98 – 3.86 (m, 12H), 3.80 – 3.68 (m, 10H), 3.64 – 3.52 (m, 12H), 3.38 – 3.33 (m, 12H), 3.26 – 3.19 (m, 3H), 3.14 (q, *J* = 5.8 Hz, 5H), 2.37 – 2.30 (m, 4H), 2.09 – 1.74 (m, 4H), 1.62 – 1.40 (m, 9H), 0.88 – 0.79 (m, 9H), 0.76 (t, *J* = 7.4 Hz, 3H). LC-MS: M+H=1742.9

Synthesis method of Compound **UBI-1211**



Step 1: UBI-1211c (V1318-111)

Compound **UBI-1211a** (2 g, 5.7 mmol) was dissolved in tetrahydrofuran (20 mL), then added **UBI-1211b** (1.1 g, 6.8 mmol) and K_2CO_3 aqueous solution (10 mL). The reaction was carried out at room temperature for 1 hours. After tetrahydrofuran was removed, the reaction solution was purified by a reversed-phase column (methanol/water = 0% to 25%, 45 minutes, collected at 0%) to obtain crude product as white solid. The crude product was acidified with 3N HCl and then purified by reversed-phase column (methanol/water = 0% to 80%, 45 minutes, collected at 25%) to obtain target product **UBI-1211c** (1g, yield 41%) as yellow oil. LCMS $[M+H]^+ = 434.3$

Step 2: UBI-1211d (V1318-112)

UBI-1211c (200 mg, 0.46 mmol) was dissolved in N,N-dimethylformamide (3 mL) and then added HATU (191 mg, 0.50 mmol) and DIEA (163 mg, 1.26 mmol). After reacting at room temperature for 20 minutes, VC1001 (**UBI-1207**) (159 mg, 0.42 mmol) was added, and then reacting at room temperature for 16 hours. The reaction solution was concentrated and purified by a reversed-phase column (methanol/water = 0% to 95%, 45 minutes, collected at 50%) to obtain 200 mg crude product as colorless oil. The crude product was subjected to Prep-TLC (methanol/dichloromethane = 1/10) to obtain target product **UBI-1211d** (100 mg, yield 27%) as colorless oil. ^1H NMR (400 MHz, Chloroform-*d*) δ 7.59 – 7.47 (m, 2H), 7.25 (d, J = 8.6 Hz, 2H), 6.71 (s, 2H), 4.57 (s, 2H), 4.23 (s, 1H), 3.91 – 3.37 (m, 26H), 3.19 – 2.98 (m, 2H), 2.68 (m, 12H), 1.49 – 1.36 (m, 2H), 1.12 – 0.70 (m, 6H).

Step 3: UBI-1211f(V1318-114)

Compound **UBI-1211d** (100 mg, 0.13 mmol), **UBI-1211e** (76 mg, 0.25 mmol) were dissolved in DIPEA (0.3 mL) and N,N-dimethylformamide (2 mL), and then reacted at room temperature for 18 hours. The reaction was purified by a reversed-phase column (acetonitrile/water = 5% to 50%, 40min, collected at 40%). After lyophilization, target product **UBI-1211f** (27 mg, yield 22%) as light yellow solid was obtained. LCMS $[M+H]^+ = 960.3$

Step 4: UBI-1211 (V1318-115)

Compound **UB-1211f** (27 mg, 0.03 mmol) and **199** (26 mg, 0.03 mmol), HOBT (4 mg, 0.03 mmol), DIEA (7 mg, 0.06 mmol) were dissolved in N,N-dimethylformamide (0.3 mL), then reacted at room temperature for 36 hours. The reaction solution was purified by Prep-HPLC to obtain target product **UBI-1211** (10 mg, yield 21%) as white solid. LCMS $[M+H]^+ = 1715.6$.

Example 3.2 Synthesis of ACTED Compound

Example 3.2.1

Compound TECP (Tris(2-chloroethyl) phosphate) (67 μ L, 5mmol/L in PBS buffer, pH=7.4) was added to a solution of anti-CD33 antibody (2.5mg, 5mg/mL) in PBS buffer solution at 0°C, incubated for 2 hours

at 37°C after well-mixed. The reaction solution was ultrafiltered for 5 times (500rpm, 5min) and replaced with PBS buffer solution. Then the reaction solution was cooled to 4 °C, and added TED with linker (53ul, 5mmol/L N,N-dimethylformamide solution), and reacted at 4 °C for 1 hour. L-cysteine was added and stirred for 10 minutes. After ultrafiltration for 5 times (14 krpm, 5min), clear ACTED solution was obtained by concentration.

Analysis Conditions:

The elution method of HIC column:

Time (min)	% B
0.0	0
12.0	100
12.1	0
18.0	0

Mobile Phase A: 1.5M (NH₄)₂SO₄ buffer solution: pH=6.9-7.1.

198.21g (NH₄)₂SO₄ was dissolved in 800 mL of water, and diluted to 1L. Adjusted to pH=7 using Na₂HPO₄ (25mM).

Mobile Phase B: 0.025M K₂HPO₄ and 0.025M KH₂PO₄ buffer solution: pH=6.9-7.1.

a: S1: 0.025M K₂HPO₄: 4.35g K₂HPO₄ was dissolved in 800mL of water, and diluted to 1L

b: S2: 0.025M KH₂PO₄: 3.4g K₂HPO₄ was dissolved in 800mL of water, and diluted to 1L

c: S1 (610ml) and S2 (390mL) were mixed and then added 0.025M K₂HPO₄ buffer solution, adjusted to pH=6.9-7.1.

d: Mobile phase B was prepared using c 75% + isopropanol (25%)

Notes: All mobile phases need to be filtered by a filter membrane.

Examples of ACTED compounds:

Table C: ACTED Compound

Example	Structure	DAR (Drug Antibody ratio)
AC-721(UB-180721)	Anti-CD33-Ab-MC-VC-633	2.0
AC-252(UB-180752)	Anti-CD33-Ab-MC-VC-680	4.0
AC-222(UB-180722)	Anti-CD33-Ab-MC-VC-680	2.0
AC-253(UB-180753)	Anti-CD33-Ab-MC-VC-699	3.6
AC-305(UB-180805)	Anti-CD33-Ab-PEG5-VC-699	2.3
AC-306 (UB-180806)	Anti-CD33-Ab-PEG5-VC-699	3.6
AC-825(UB-180825)	Anti-CD33-Ab-PEG6-VC-699	6.5

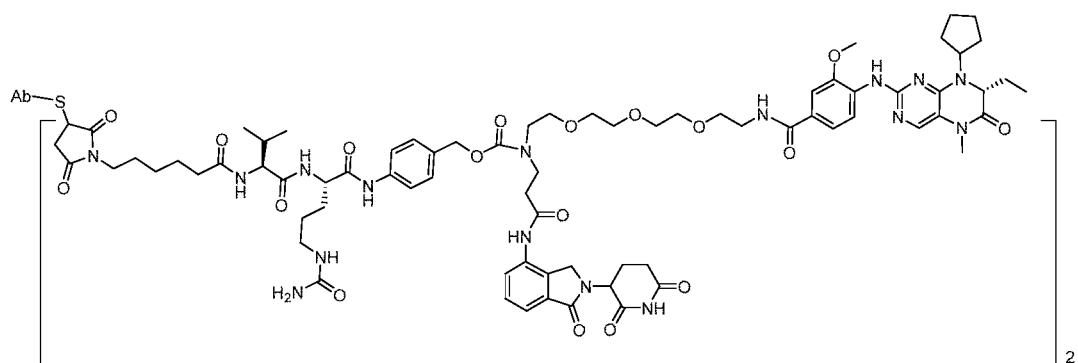
MC and VC are abbreviations for amino acids, and their structure is as follows:

MC	VC (Val-Cit)	VC-PAB (Val-Cit-PAB)

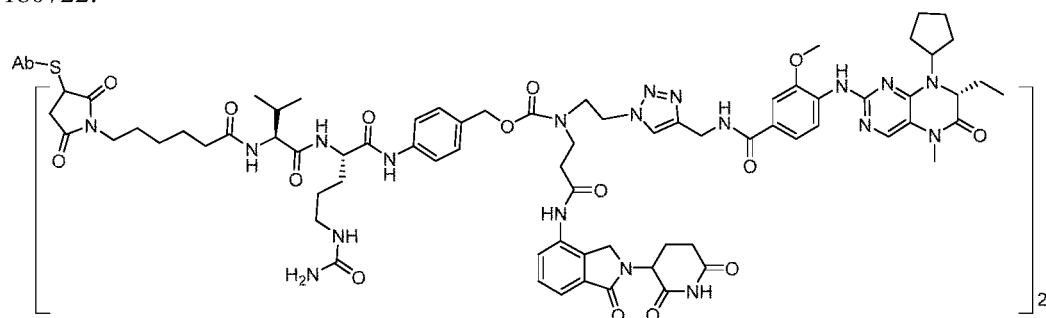
PEG5 and PEG6 refer to (-CH₂-CH₂-O-)₅ and (-CH₂-CH₂-O-)₆;

Anti-CD33-Ab refers to anti-CD33 antibody.

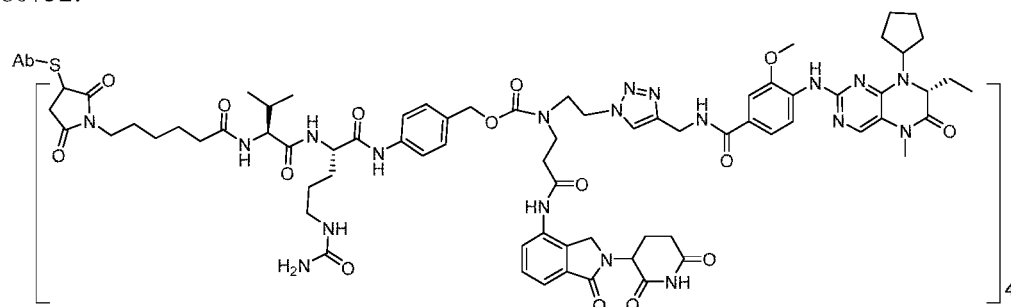
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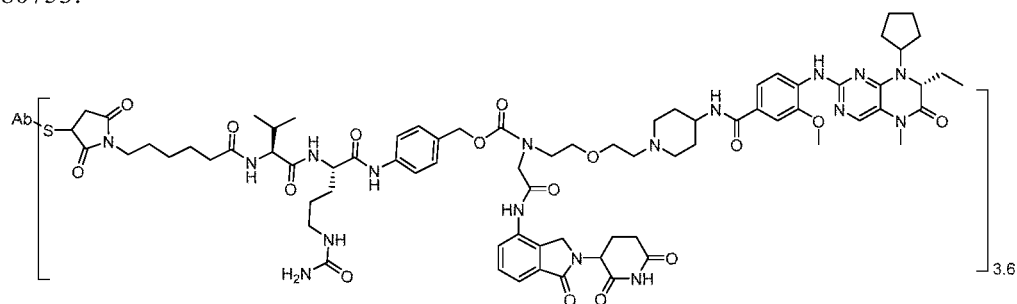
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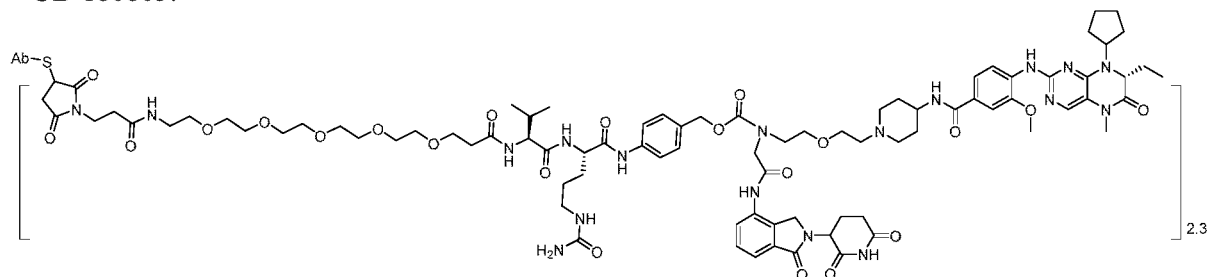
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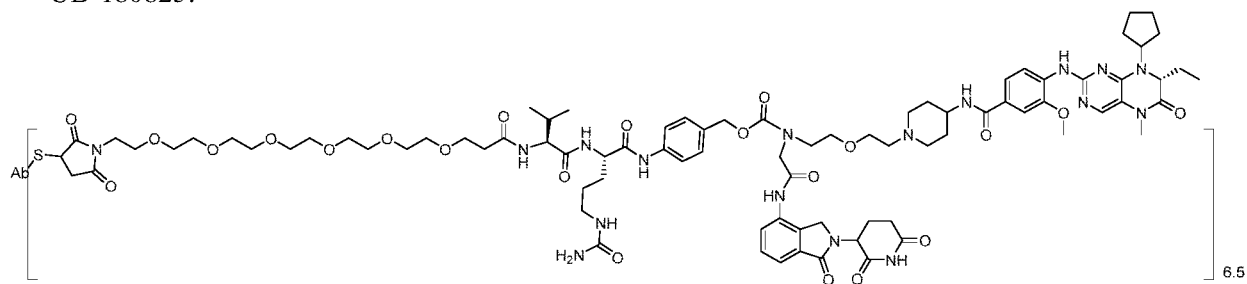
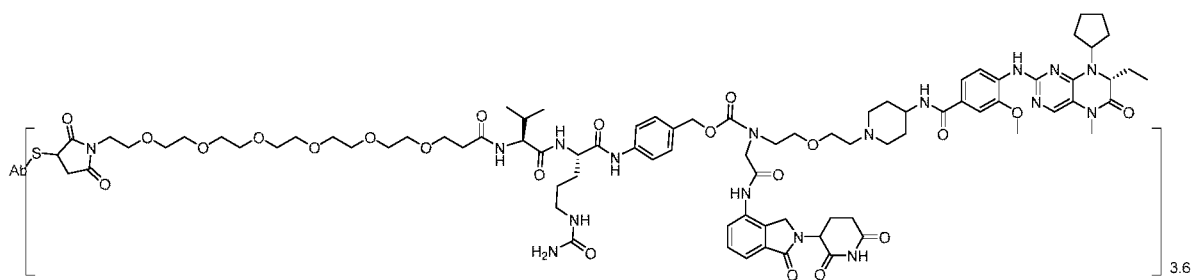
UB-180753:



UB-180805:



UB-180806:

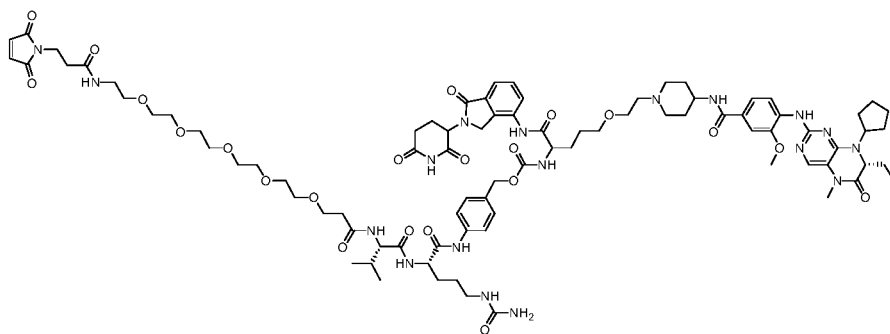


in each formula, Ab refers to antibody.

5

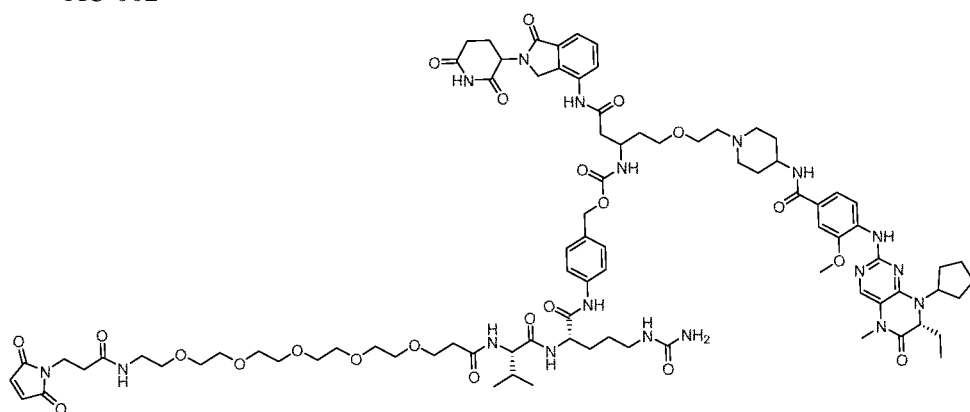
Similarly, a series of ACTED compounds with DAR ranging from 2 to 5 were synthesized using AC-001~AC-006 as TED with linker.

AC-001

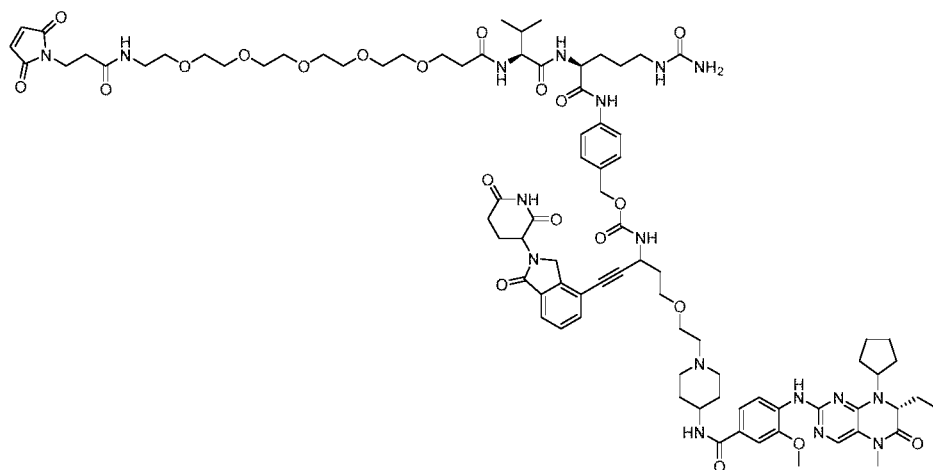


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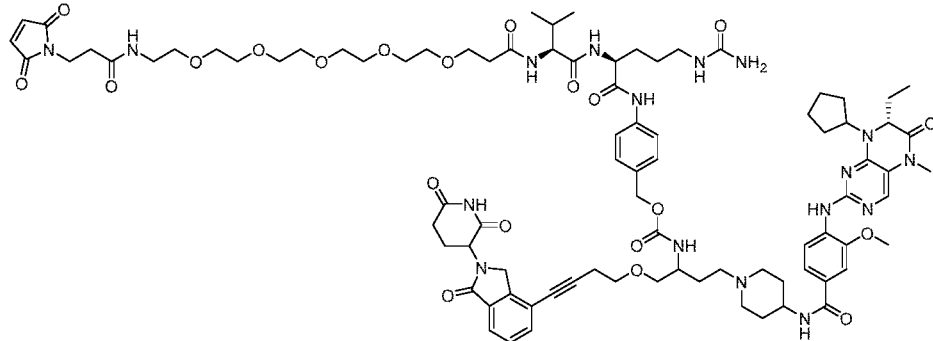
AC-002



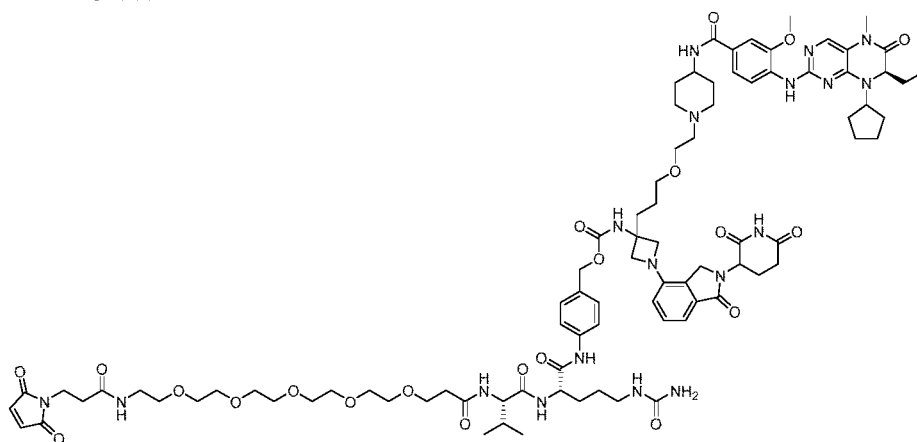
AC-003



AC-004



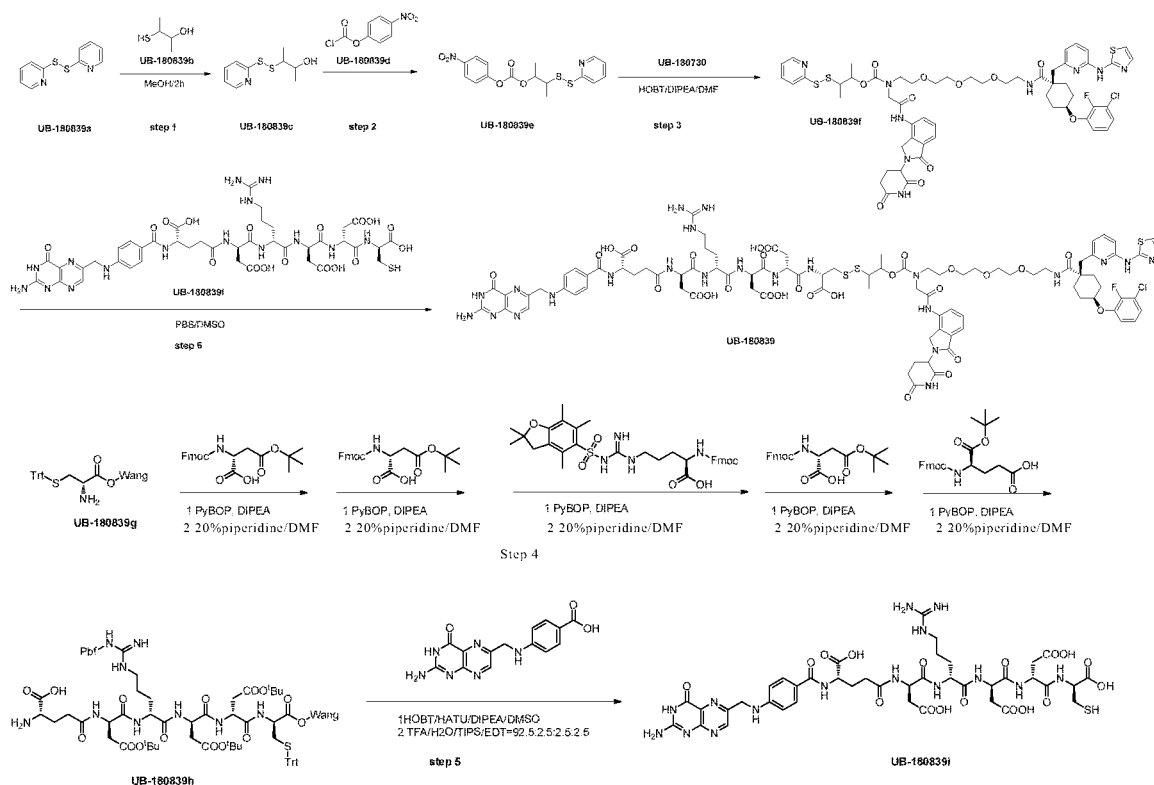
AC-005



AC-006

Example 4: Preparation of T ED conjugated with Folic Acid

Synthesis method of Compound UB-180839 FR-Peptide-Linker S- S-730



Step 1: UB-180839c

- 5 Compound UB-180839a (500 mg, 2.3 mmol) was dissolved in methanol (10 mL) and then added UB-180839b (80 mg, 0.76 mmol). The reaction was carried out at room temperature overnight under nitrogen protection. The reaction solution was concentrated to obtain crude product. The crude product was isolated by column chromatography (ethyl acetate/dichloromethane = 1/10) to obtain crude product UB-180839c (300 mg, yield 61%) as colorless oil.

10 Step 2: UB-180839e

- Compound UB-180839c (300 mg, 1.4 mmol) was dissolved in dichloromethane (10 mL), and then added UB-180839d (309 mg, 1.5 mmol) and TEA (282 mg, 2.8 mmol). The reaction was carried out at room temperature overnight under nitrogen protection. The reaction solution was concentrated and purified via reversed-phase column (methanol/water = 5 % to 95%, collected at 70%) to obtain target product UB-180839e (80 mg, yield 15%) as yellow oil.

Step 3: UB-180839f

- Compound UB-180839e (20 mg, 0.05 mmol) and UB-180730 (49 mg, 0.05 mmol), HOBT (7 mg, 0.05 mmol), DIEA (14 mg, 0.11 mmol) were dissolved in N,N-dimethylformaldehyde amide (2 mL). The reaction was allowed to react at room temperature for 36 hours. The reaction solution was prepared to obtain UB-180839f (10 mg, yield 16%) as white solid product.

Step 4: UB-180839h

- The resin of compound UB-180839g was placed in a reactor with a filter device, added 50 mL dichloromethane, and reacted for 1 hour on the shaker. The resin was obtained after filtration and was added to the piperidine/DMF (1:4) solution to react for 15 minutes, and then washed with anhydrous DMF (5 x 30

mL x 0.5 min) to obtain the resin.

N-Fluorenylmethoxycarbonyl-D-aspartate-4-tert-butyl ester (3.8g, 9.2 mmol), PyBOP (7.2g, 13.8 mmol), DIPEA (2.4mL, 6.9mL) were dissolved in DMF, and then added Resin. The mixture was reacted for 2h on a shaker. Then mixture was washed with DMF (5 x 30 mL x 0.5 min). The resulting resin was added to piperidine/DMF (1:4) solution and reacted for 15 minutes, then washed with anhydrous DMF (5 x 30 mL x 0.5 min) to obtain the resin.

N-Fluorenylmethoxycarbonyl-D-aspartate-4-tert-butyl ester (3.8g, 9.2 mmol), PyBOP (7.2g, 13.8 mmol), DIPEA (2.4mL, 6.9mL) were dissolved in DMF, and then added Resin. The mixture was reacted for 2h on a shaker. Then mixture was washed with DMF (5 x 30 mL x 0.5 min). The resulting resin was added to piperidine/DMF (1:4) solution and reacted for 15 minutes, then washed with anhydrous DMF (5 x 30 mL x 0.5 min) to obtain the resin.

Fmoc-D-Arg(Pbf)-OH(6g, 9.2 mmol), PyBOP (7.2g, 13.8 mmol), DIPEA (2.4mL, 6.9mL) were dissolved in DMF, and then added resin. The mixture was reacted for 2h on a shaker. Then mixture was washed with DMF (5 x 30 mL x 0.5 min). The resulting resin was added to piperidine/DMF (1:4) solution and reacted for 15 minutes, then washed with anhydrous DMF (5 x 30 mL x 0.5 min) to obtain the resin.

N-Fluorenylmethoxycarbonyl-D-aspartate-4-tert-butyl ester (3.8g, 9.2 mmol), PyBOP (7.2g, 13.8 mmol), DIPEA (2.4mL, 6.9mL) were dissolved in DMF, and then added Resin. The mixture was reacted for 2h on a shaker. Then mixture was washed with DMF (5 x 30 mL x 0.5 min). The resulting resin was added to piperidine/DMF (1:4) solution and reacted for 15 minutes, then washed with anhydrous DMF (5 x 30 mL x 0.5 min) to obtain the resin.

N-(9-Fluorenylmethoxycarbonyl)-D-glutamate-1-tert-butyl ester (3.8g, 9.2 mmol), PyBOP (7.2g, 13.8 mmol), DIPEA (2.4mL, 6.9mL) were dissolved in DMF, and then added resin. The mixture was reacted for 2h on a shaker. Then mixture was washed with DMF (5 x 30 mL x 0.5 min). The resulting resin was added to piperidine/DMF (1:4) solution and reacted for 15 minutes, then washed with anhydrous DMF (5 x 30 mL x 0.5 min) to obtain the resin.

Step 5: UB-180839i

Pterioic acid (1.75g, 4.6mmol) was added to DMSO (1L), then heated to 120 °C to dissolve, and slowly cooled to room temperature to obtain a clear solution. Then added UBI-180839h (20g) resin, HATU (5.2g, 13.8mmol), and DIPEA (7.2mL). The mixture was reacted for 16h on a shaker. After the reaction solution was filtered, the resin was washed with DMSO (50mLx3), DMF (50mLx3), DCM (50mLx3), respectively, to obtain crude product. The crude product was added to a solution of TFA/H₂O/TIPS/EDT=92.5:2.5:2.5:2.5(100 mL)and stirred for 1 h. The filtrate was obtained by filtration, concentrated and purified by reversed-phase column to obtain UB-180839i (80mg).

Step 6: UB-180839

Compound UB-180839g (10 mg, 0.01 mmol) was dissolved in DMSO (1 mL) and added UB-180839f (27 mg, 0.03 mmol) in PBS (0.5 mL) and DMSO (0.5 mL). The reaction solution was maintained at room temperature overnight. The reaction solution was prepared to obtain product UB-180839 (1.0 mg, yield 2%) as white solid.

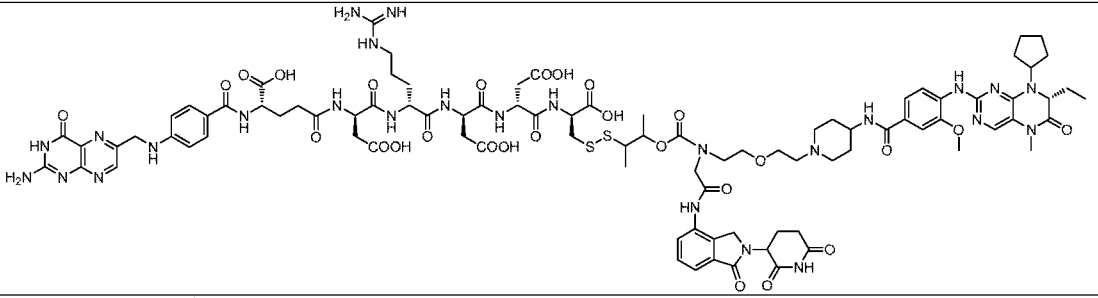
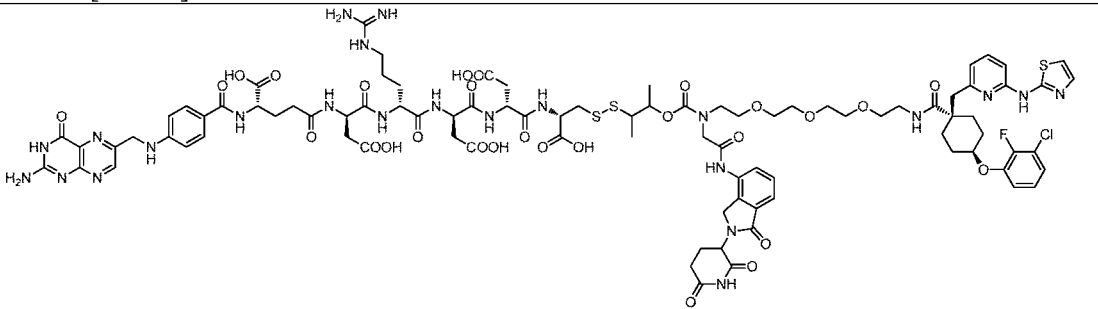
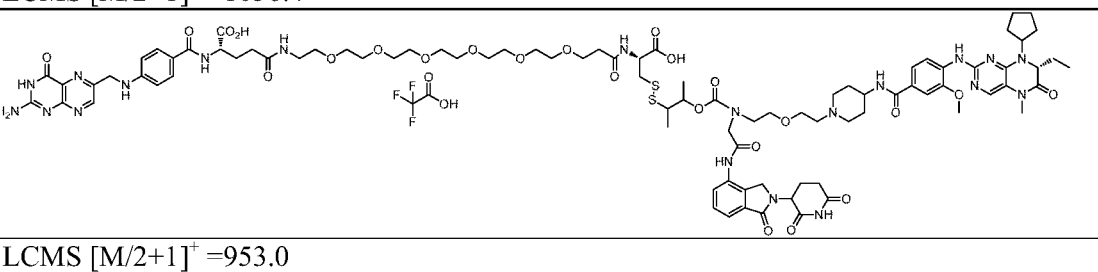
Synthesis method of Compound UB-180838 (FR-Peptide-Linker S-S-699) was the same as UB-180839.

Synthesis method of Compound UB-180829 (FR-Peptide-Linker S-S-699) was the same as UB-180839.

5

Table D shows the Analytical data for TED conjugated with folic acid molecule 829, 838, and 839.

Table D

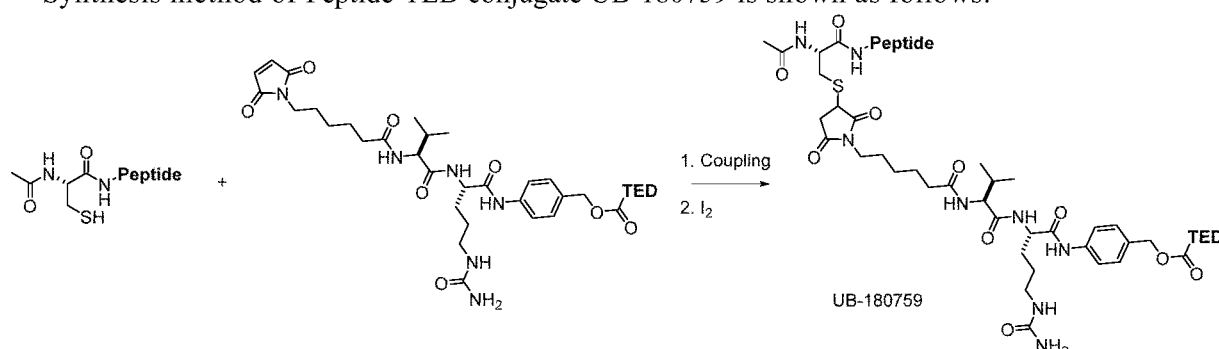
838	
LCMS $[M/2+1]^+ = 1036.0$	
839	
LCMS $[M/2+1]^+ = 1056.4$	
829	
LCMS $[M/2+1]^+ = 953.0$	

10

Example 5: Preparation of TED conjugated with polypeptide molecule

Polypeptide CAADGCTWEVWGRECWY-NH₂ was synthesized using conventional polypeptide synthesis procedures.

Synthesis method of Peptide-TED conjugate UB-180759 is shown as follows:



15 in the formula, Peptide (polypeptide) refers to any peptide drug (for example, a polypeptide with the

sequence of CAADGCTWEVWGRECWY).

The coupling reaction operation of polypeptide and TED is the same as that of antibody and TED; the molecule after the peptide coupling with TED needs to be further oxidized. The oxidation operation is as follows:

40 mg of polypeptide (75% purity, i.e. $40 \times 0.75 / 3710.7 = 0.008$ mmol) was taken and added following solvents to make the concentration be about 1 mg/mL: 50% methanol in water 15 mL, 50% acetic acid 15 mL, 1 N HCl 1 mL. Iodine (253) 8eq was dissolved in a very small amount of methanol; finally added iodine solution under nitrogen protection. After reacting 30 minutes, ascorbic acid (176.12) was added: 18mg (added about ratio of 1:1 according to calculation of remaining molar amount of iodine) was directly added, dissolved in 5mL of water, which was added dropwise till the reaction become colorless. After the reaction was completed, the system was slightly turbid white. The pH was adjusted to weakly acidic condition with aqueous ammonia. After centrifugation, the supernatant was lyophilized and purified to obtain compound UB-180759. MS=3567 was detected.

Test Example 1 Biological Test

1.1 Biological assay of the inhibitory effect of the compound of the present invention on MV4;11 cell proliferation

Experimental Materials:

Reagent	Supplier	Article No.	Batch number
MV4;11 cell line	Viva	/	/
Iscove's modified D μ L becco's medium	Gibco	12200-036	1946620
FBS	Gibco	10091-148	1735161
pen-strep	Gibco	15140-122	1953108
Trypsin-EDTA (0.25%), Phenol Red	Thermo	25200056	1951208
DPBS	Thermo	14190144	1952086
Cell Titer -Glo	Promega	G7573	0000310975

Experimental method:

1) Preparation of buffer

MV4;11 cell culture medium: Iscove's modified Dulbecco's medium was added 10% FBS and 1/100pen-strep.

2) Experimental Procedure:

(1) MV4;11 cells were subcultured with cell culture medium, and the well-growing cells are seeded in a 96-well plate, 100 μ L per well, the number of cells per well is 20,000, and the cells are cultured overnight in a cell incubator at 37°C and 5% CO₂.

(2) Drugs were prepared as 10mM stock solution with dimethyl sulfoxide (DMSO). The compounds were diluted (which ensure the concentration of DMSO in the culture system being 1%), each concentration in duplicate. Diluted compounds were added to cell culture wells (final concentration was 100 μ M, 33 μ M, 11 μ M...), shaking gently to mix. Two negative control wells were set only added cells (ensuring that the DMSO concentration in the culture system was 1%).

3) Measurement for Result:

(1) After incubating for 72 hours, 50 μ L Cell Titer-Glo was added to each well and incubating at room temperature for 10 minutes.

(2)The chemiluminescence signal values were read by using Envision.

(3) Data were analysed using Dose-response-inhibition equation in software GraphPad Prism to obtain

the IC50 value.

1.2 PLK1 protein expression biological assay of the compound of the present invention

Experimental Materials:

Reagent	Supplier	Article No.	Batch number
MV4;11 cell line	Viva	/	/
RIPA Lysis and Extraction Buffer	Thermo	89900	TD265751
Complete Tablets EDTA-free	Roche	4693132001	27423500
Novex™ 4 - 20% Triglycine Mini Gel, WedgeWell™ format, 12 wells	Invitrogen	XP04202BOX	18052448
PageRuler™ added pre-stained protein ladder	Thermo		
Novex™ Tris-Glycine SDS electrophoretic buffer (10X)	Thermo	LC2675	1952401
Novex™ Tris-Glycine Transfer Buffer (25X)	Thermo	LC3675	1934476
Pierce™ 20X TBS Tween™ 20 buffer	Thermo	28360	TD265259
SuperSignal™ West Femto Maximum Sensitivity Substrate	Thermo	34095	TE265690
Nitrocellulose membrane	Amersham	10600001	A14115264
PLK1 (208G4) Rabbit mAb	Cell Signaling	4513	/
GAPDH (D16H11) XP® Rabbit mAb	Cell Signaling	5174	/
Anti -Rabbit IgG, HRP-linked antibody	Cell Signaling	7074P2	/

Experimental method:

1) Preparation of buffer

1x electrophoretic buffer	Novex™ Tris-Glycine SDS electrophoretic buffer (10X) was diluted to 1X
1x transfer buffer	Novex™ Tris-Glycine transfer buffer (25X) was diluted to 1X, added 20% methanol
1x TBST	Pierce™ 20X TBS Tween™ 20 Buffer was diluted to 1X
Confining Liquid	1x TBST with 5% skimmed milk

2) Experimental procedure:

(1) MV4;11 cells were subcultured with cell culture medium, and the well-growing cells were seeded in a 6-well plate, 2ml per well, the number of cells per well is 1 million, and the cells were cultured overnight in a cell incubator at 37°C and 5% CO₂.

(2) Drugs were prepared as 10mM stock solution with dimethyl sulfoxide (DMSO). Diluted compounds were added to cell culture wells (ensuring that the concentration of DMSO in the culture system was 1%), shaking gently to mix. Negative control well was set in addition (by adding the same amount of DMSO).

(3) After incubating 6 hours, the cells were lysed using RIPA cells lysis buffer, proteins were extracted, the concentration of the proteins measured by BCA kits. 2X protein loading buffer was added, heated at 100°C for 10 minutes, and the samples were stored at -20°C.

(4) Samples were loaded, in amount of protein 25µg per well, onto polyacrylamide gel for electrophoresis.

(5) Proteins transferred from polyacrylamide gel to PVDF membrane, 5% skimmed milk was added to seal at room temperature for 1 hour.

(6) Primary antibodies (PLK1 (208G4) rabbit mAb and GAPDH (D16H11) XP® rabbit mAb) were incubated overnight at 4°C, the membranes were washed three times with TBST solution, 10 minutes for each.

(7) Secondary antibodies (Anti-rabbit IgG, HRP-linked antibody) were incubated at room temperature for 1 hour, and then the membranes were washed with TBST solution three times, 10 minutes for each.

3) Measurement for Result:

Finally, chromogenic solution (SuperSignal™ West Femto Maximum Sensitivity Substrate) was added for color development, photos were taken using automatic chemiluminescence instrument (ChemiDoc XRS+ (BIO-RAD)), pictures were collected and analyzed.

1.3 Biological assay of inhibitory effect of the compound of the present invention on PLK1 enzyme activity

Experimental Materials:

Reagent	Supplier	Article No.	Batch number
ADP-Glo™ Kinase Assay	promega	V9101	0000312764
ADP-Glo™ reagent	promega	V912A	0000234146
Kinase Dection Buffer	promega	V913A	0000310185
Kinase Dection Substrate	promega	V914A	0000233450
Ultra Pure ATP	promega	V915A	0000301398
ADP	promega	V916A	0000307789
PLK1 Kinase Enzyme System	promega	V2841	0000322411
PLK1, active	promega	/	P1590-7 C306
Casein, dephosphorylated	promega	/	C2090-7 D205
5x reaction buffer A	promega	/	M2814-6 C523
DTT	promega	/	G1379-9

Experimental method:

1) Preparation of buffer

Enzyme reaction buffer: 5X buffer were diluted to 1X using ddH₂O, 50 uM DTT was added.

2) Experimental Procedure:

(1) Enzyme, substrate and ATP were diluted using 1X buffer.

(2) 2μL of 2.5X enzyme solution was added to each well.

(3) 1μL of diluted compound was added to each well, and incubated at room temperature for 15 minutes.

(4) 2μL of 2.5X substrate/ATP mixture was added to each well, and incubated at room temperature for 1 hour.

(5) 5μL ADP-glo was added to each well, and incubated at room temperature for 40 minutes.

(6) 10μL detection reagent was added, incubated at room temperature for 30 minutes.

3) Measurement for Result:

(1) The chemiluminescence signal values were read using Envision.

(2) Data were analysed using Dose-response-inhibition equation in software GraphPad Prism to obtain the IC₅₀ value.

1.4 BRD4 (Bromodomain containing protein 4) protein expression biological assay of the compounds of the present invention

The test method is similar to that in 1.2 PLK1 protein expression biological assays.

1.5 PLK1 (Polo-like kinase 1) protein expression biological assay of the compounds of the present invention

The test method was similar to that in 1.2 PLK1 protein expression biological assay.

1.6 Results of Biological test

Results for protein expression test of 1.2, 1.4 and 1.5 are shown in Figure 2.

The inhibitory activity IC_{50} of following molecules on MV4;11 cells are between 0.1nM-10M:
UB-180510, UB-180519, UB-180523, UB-180525, UB-180529, UB-180537, UB-180539, UB-180541,
UB-180545, UB-180552, UB-180553, UB-180554, UB-180564, UB-180566, UB-180594, UB-180601, and
UB-180602.

The inhibitory activity IC_{50} of following molecules on Hela cells are between 0.1nM-10M:
UB-180512, and UB-180525.

The above results show that the present invention provides a class of Targeted Enzyme Degradation (TED) molecules, which are consisted of a unit of small molecule compound targeting to target proteins, a unit binding E3 ubiquitin ligases and linker unit, which is able to bind with PLK and/or BRD4 proteins, promoting PLK1 and/or BRD4 protein to become more vulnerably degradable by proteases, thereby inhibiting cell proliferation, and can be used as a protein degradation drug. The compound provided in the present invention has a good inhibitory effect on the proliferation of MV4-11 cells, indicating that the compound of the present invention can be prepared as antitumor drugs, especially those for the treatment of acute myeloid leukemia, AML).

Test Example 2

(2.1) Cell Proliferation Experiment:

Reagents: RPMI-1640 medium, McCoy's 5A medium, IMDM medium, MEM medium, L-15 medium, fetal bovine serum, Penicillin-Streptomycin double antibody, trypsin, etc., 2-mercaptoethanol, NEAA, pyruvate, etc.

The tumor cell lines used in this experiment are shown in Table 1.1:

Table1 .1 List of Tumor Cell Lines

No.	Cell Name	Cell Source
1	Human Colon Cancer HT-29	Shanghai Cell Bank of Chinese Academy of Sciences (CAS)
2	Human Colon Cancer HCT-116	Shanghai Cell Bank of Chinese Academy of Sciences (CAS)
3	Human Small Cell Lung Cancer NCI-H82	American Type Culture Collection (ATCC)
4	Human Monocytic Leukemia THP-1	Shanghai Cell Bank of Chinese Academy of Sciences (CAS)
5	Human Acute Myelomonocytic Leukemia HL-60	Shanghai Cell Bank of Chinese Academy of Sciences (CAS)
6	Human Glioma Cells U-87 MG	Shanghai Cell Bank of Chinese Academy of Sciences (CAS)
7	Human cervical adenocarcinoma cells Hela	Shanghai Cell Bank of Chinese Academy of Sciences (CAS)
8	Human Breast Cancer MDA-MB-231	Shanghai Cell Bank of Chinese Academy of Sciences (CAS)
9	Human Acute Lymphoblastic Leukemia Cells MOLT-4	Shanghai Cell Bank of Chinese Academy of Sciences (CAS)
10	Human Ovarian Cancer SK-OV-3	Shanghai Cell Bank of Chinese Academy of Sciences (CAS)
11	Human Monocytic Leukemia HL-60	Shanghai Cell Bank of Chinese Academy of Sciences (CAS)
12	Human Monocytic Leukemia MV4-11	American Type Culture Collection (ATCC)

The cells were cultured in conventional ways, and the cells should be passed at least for at least 2 generations before plating. Cells at the logarithmic growth phase are collected, prepared into single cell suspensions and counted. The concentration of cells was adjusted to the desired concentration, and cells were inoculated into a 96-well cell culture plate at 100 μ L per well. 100 μ L of complete medium of the test compound were added to each well, set up 2 duplicates for each concentration, and diluted with a 5-fold gradient, and continue to culture for 72 h. All cells were subjected to EC₅₀ test for corresponding samples.

The fluorescence intensity of each well was detected using the Alarm blue method, and EC₅₀ was calculated.

EC₅₀ was calculated according to the following formula:

$$Y = \text{Max} + (\text{Min} - \text{Max}) / [1 + (X / \text{EC}_{50}) \times \text{Slope}]$$

wherein Min, Max and Slope represent the minimum, maximum and slope respectively.

(2.2) Flow Cytometry Experiment

In order to test the binding ability of Ab. and ACTED to receptors (CD33, DLL3) on cell surface, cells were plated on 96-well plate, after that added primary antibody and incubated for 1 hour at 4 °C, and then wash with PBS containing 1% BSA twice, then added secondary antibody and incubated at 4 °C for half an hour in the dark, washed three times and analyzed using a flow cytometer.

In order to test endocytosis of Ab. and ACTED, cells were first treated with antibody or antibody protein degradation agent conjugate for a period of time, then washed with PBS containing 1% BSA twice, and then added secondary antibody and incubated for half an hour in the dark at 4 °C. In order to test the level of target receptors on the cell surface, cells were incubated with FITC-anti-human CD33 or DLL3 antibody at 4 °C in the dark for half an hour, and finally washed three times and tested with a flow cytometer.

(2.3) Western Blot

Cells were treated with the compound for a period of time, after that the cells were collected by centrifugation. After washing with PBS, the cells were lysed by adding RIPA buffer; the cell lysate were added to the loading buffer and then appropriate volume were taken and slowly added to the corresponding wells of the gel plate, and run the SDS-PAGE gel (4%-12%). After running the gel, transferred to the PVDF membrane and sealed with 5% skimmed milk powder at room temperature for 1 hour. The membrane was placed in the primary antibody diluted with 5% skimmed milk powder and shaken slowly overnight at 4 °C. After incubation with primary antibody, the membrane was washed 3 times using a TBST shaker; added secondary antibody diluted with 5% skimmed milk powder corresponding to the primary antibody, and shaken slowly at room temperature for 1 hour. After incubation with secondary antibody, the membrane was washed 3 times using the TBST shaker again. The PVDF membrane was flattened in the cassette, the strip was evenly infiltrated with ECL developer solution, and placed in ChemDoc XRS+ gel imager for taking pictures. The intensity of protein bands was analyzed quantitatively using ImageJ software.

DC50 was calculated.

(2.4) *In vitro* kinase activity test

The compound, enzyme, substrate and ATP were diluted using 1x reaction buffer to the desired concentration. 1 μ L of compounds with different concentrations, 2 μ L of enzyme, and 2 μ L of substrate/ATP mixed solution were added to the 384-well plate, and incubated for 1 hour at room temperature. Then 5 μ L of ADP-Glo™ reagent was added to each well and incubated at room temperature for 40 minutes. Finally, 10 μ L of detection reagents were added, incubated at room temperature for 30 minutes, and chemiluminescence signals were detected using Envision.

IC50 was calculated.

The results of test are shown in Table 1.2

In the table, A≤10 nM, 10 nM<B≤100 nM, 100 nM<C≤1000 nM, D>1000 nM

UB No.	MV4;11 EC50 nM	HCT116 EC50 nM	NCIH82 EC50 nM	HL-60 EC50 nM	HT-29 EC50 (nM)	U-87MG EC50 nM	MDA-MB-231 EC50 nM	THP-1 EC50 nM	Hela EC50 nM	MOL4-4 EC50(nM)	HUVEC EC50 nM	PBMC EC50 nM	Daudi EC50 (nM)	SKO3 EC50 (nM)	U937 EC50 (nM)	A549 EC50 (nM)	PLK1 DC50 (nM) (24h)	PLK1 DC50 (nM) (6h)	BRD4 DC50 (nM) (24h)	BRD4 DC50 (nM) (6h)	BRD4 IC50 nM
UB-180501	D	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180502	B	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180503	C	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180504	B	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180505	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180506	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180509	D	D	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180510	D	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	-	-	-	-
UB-180511	C	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	-	-	-	-
UB-180512	C	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	-	-	-	-
UB-180513	B	B	A	B	A	B	C	B	A	B	C	C	B	A	A	A	B	-	-	-	B
UB-180514	B	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180515	D	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	-	-	-	-
UB-180516	D	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	-	-	-	-
UB-180517	B	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	-	-	-	-
UB-180518	D	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	-	-	-	-
UB-180519	B	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	-	-	-	-
UB-180520	B	D	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180521	D	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	-	-	-	-
UB-180522	C	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	-	-	-	-
UB-180523	B	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	D	-	D	-
UB-180524	B	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	-	-	-	-
UB-180525	B	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	D	-	D	-
UB-180526	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	D	-	D	-
UB-180527	D	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180528	B	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	-	-	-	-
UB-180529	B	-	-	-	-	-	-	-	D	-	-	-	-	-	-	-	-	D	-	D	-
UB-180530	B	D	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180531	B	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180532	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

UB-180843	A	-	A	-	A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180844	A	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	B	-	-
UB-180845	A	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	A	-	-
UB-180846	A	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	B	-	-
UB-180847	C	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180848	A	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180849	C	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180850	C	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180851	C	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180852	C	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180853	C	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180854	C	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180855	C	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180856	C	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
UB-180857	B	-	C	-	C	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

Test Example 3 Animal Experiment

Female BALB/c nude mice (number: 58; age: 5-6 weeks) were purchased from Beijing Weitong Lihua Laboratory Animal Technology Co., Ltd. and raised in the SPF animal room of Suzhou Shengsu New Pharmaceutical Development Co., Ltd., temperature was 20-25 °C, relative humidity was 40%-70%, light and dark light for 12 hours each; animals were free to drink and eat freely. After normal feeding for about 5 days, mice with good physical signs can be selected for this experiment after veterinary inspection. Before grouping, animals were identified at their base of tail using maker pens. After grouping, each animal was identified by ear clipping.

Human acute monocytic leukemia cells MV-4-11 were obtained from American Type Culture Collection (ATCC, cryopreserved in liquid nitrogen in our laboratory).

Under culture conditions of 5% CO₂ and 37 °C, MV-4-11 cells were subjected to routine cell culture in IMDM medium with 10% fetal bovine serum. Passaged with 0.25% trypsin digestion. The passage was performed according to cell growth, passaging ratio is 1:3 or 1:6. MV-4-11 cells at logarithmic growth phase were collected, the cells were resuspended in 50% PBS (pH 7.4, 0.01 M) and 50% Matrigel after counting, the concentration of cells was adjusted to 7.0×10^7 cells/mL; the cells were placed in ice box, the cell suspension was suck up using 1 mL syringe, and injected into the Right anterior axillary subcutaneous of nude mice. Each animal was inoculated with 200 μL (14×10^6 cells/mouse) to establish MV-4-11 xenograft tumor model. The animal status was regularly observed, the tumor diameter was measured using the electronic vernier caliper, the data were entered into Excel, the tumor volume was calculated, and the tumor growth was monitored. When the tumor volume reached 100-300 mm³, animals with tumors that are too large, too small, or of uncertain formation were eliminated, and tumor-bearing mice with good health and similar tumor volumes were selected, and divided into 4 groups(n = 6~8) using randomized block method. After the start of experiment, the tumor diameter was measured twice a week, the tumor volume was calculated, and the animal weight was weighed and recorded at the same time.

Calculating formula for tumor volume (TV) is as follows:

$$TV (mm^3) = l \times w^2 / 2$$

wherein, *l* means the long diameter of the tumor (mm); *w* means the short diameter (mm) of the tumor.

Before each administration the compounds were taken and prepared into dosage formulation with certain concentration.

Animal grouping and dosage regimen see Table 2.

Table. 2 The dosage regimen of the pharmacodynamic experiment of the nude mouse xenograft tumor model

Group	Sample	Number of animals	Dose (mg·kg ⁻¹)	Dosing volume (mL·kg ⁻¹)	Route of administration	Dosing frequency
A	0.01 M PBS (pH 7.4)	6	-	10	i.v.	dosing once every other 2 days
B	UB-180752(AC-252)	6	10	10	i.v.	dosing once every other 2 days
C	UB-180753(AC-253)	6	10	10	i.v.	dosing once every other 2 days
D	UB-180699(199)	8	10	10	i.v.	dosing once every other 1 day
E	UB-180699(199)	8	10	10	i.g.	dosing once every other 1 day

During the experiment, experimenters and veterinarians need take continuous observation on the

physical signs and health of the experimental animals, weigh their weight and measure the diameter of the tumor on at regular intervals.

Calculating formula for relative tumor volume (RTV) is:

$$RTV = TV_t / TV_{initial}$$

5 wherein $TV_{initial}$ is the tumor volume measured when grouped administration; TV_t is the tumor volume at each measurement during the administration period.

Calculating formula for the relative increasing rate of tumor (%T/C) is:

$$\%T/C = 100\% \times (RTV_T / RTV_C)$$

wherein RTV_T means the RTV of treatment group; RTV_C means the RTV of solvent control group.

10 Calculating formula for tumor growth inhibition rate TGI (%) is:

$$TGI = 100\% \times [1 - (TV_{t(T)} - TV_{initial(T)}) / (TV_{t(C)} - TV_{initial(C)})]$$

wherein $TV_{t(T)}$ means the tumor volume of the treatment group at each measurement; $TV_{initial(T)}$ means the tumor volume of the treatment group when grouped administration; $TV_{t(C)}$ means tumor volume of solvent control group at each measurement; $TV_{initial(C)}$ means the tumor volume of solvent control group when grouped administration.

15 Calculating formula for loss rate of animal weight is:

$$\text{Loss rate of animal weight} = 100\% \times (BW_{initial} - BW_t) / BW_{initial}$$

wherein, BW_t means the animal weight at each measurement during administration; $BW_{initial}$ means the the animal weight when grouped administration.

20 Calculating formula for inhibit rate of tumor weight IR (%) is:

$$IR = 100\% \times (W_C - W_T) / W_C$$

wherein W_C means the tumor weight of control group; W_T means the tumor weight of treatment group.

Result Analysis

25 Referring to Figure 3, TED compound 199 was administered to MV4;11 tumor model mice at a dose of 30mpk, injected every other day, and showed tumor shrinking effect. The weight of the mice was stable, and no other abnormalities of mice were observed.

ACTED compounds AC-252 and AC-253 were administered via injection to MV4;11 tumor model mice at a dose of 10mpk every other 2 days, and all showed tumor growth inhibitory effects (64%). The weight of the mice was stable, and no other abnormalities of mice were observed.

30 The amount of corresponding TED molecules carried, calculated based on ACTED, is an Extremely small amount. Taking 253 as an example, the administration dose is 10 mpk, which is equivalent to 0.014 mpk after converting into dose of TED molecule 199. This dose is far below the dose of 199 itself of 30mpk. At such a small amount of administration dose, it is difficult to observe expected tumor growth inhibitory effect if TED molecule 199 is used alone. The ACTED compounds of the present invention (as shown in Table C) all show excellent anti-tumor activity and excellent pharmacokinetic properties similar to AC-252 and AC-253.

40 All documents mentioned in the present invention are cited as references in this application, just as each document is individually cited as a reference. In addition, it should be understood that, after reading the above teaching content of the present invention, those skilled in the art can make various changes or modifications to the present invention, and these equivalent forms also fall within the scope defined by the appended claims of the present application.

Claims

1. A conjugate of formula I:



wherein

A is a monovalent moiety of a targeting molecule;

B is a monovalent moiety of an E3 ligase ligand;

L1 is a linker connecting A and B; and L1 has a formula II:



wherein

when X or Y is NH or NR, X or Y and part of L2 chain to which X or Y is attached are combined together to form a 4 - 8 membered aromatic or non-aromatic heterocyclic ring; wherein R is substituted or unsubstituted C1-C10 alkyl, $-(\text{C}=\text{O})-\text{R}'$, $(\text{C}=\text{O})\text{NH}-\text{R}'$, $-\text{NH}(\text{C}=\text{O})-\text{R}'$, $-\text{SO}_2-\text{R}'$, $-\text{NHSO}_2-\text{R}'$, $-\text{SO}_2\text{NH}-\text{R}'$, $-\text{SO}-\text{R}'$, $-\text{NHSO}-\text{R}'$, $-\text{SONH}-\text{R}'$, $-\text{PO}_3-\text{R}'$, $-\text{NHCOO}-\text{R}'$, $-\text{COO}-\text{R}'$, or $-\text{NH}-\text{CO}-\text{NH}-\text{R}'$, $-\text{NH}-\text{CO}-\text{O}-\text{R}'$, or $-\text{X}'-\text{L3}-\text{Z}$; wherein L3 is linker, and Z is a polypeptide element or target molecule T; wherein R' is selected from the group consisting of H, substituted or unsubstituted C1-C6 alkyl, substituted or unsubstituted C3-C8 cycloalkyl, amino protecting group, and $-\text{X}'-\text{L3}-\text{Z}$;

or

each X and Y is independently selected from the group consisting of $-\text{O}-$, $-\text{S}-$, $-\text{NH}-$, $-\text{NR}-$, $-(\text{C}=\text{O})\text{NH}-$, $-\text{NH}(\text{C}=\text{O})-$, $-\text{SO}_2-$, $-\text{NHSO}_2-$, $-\text{SO}_2\text{NH}-$, $-\text{SO}-$, $-\text{NHSO}-$, $-\text{SONH}-$, $-\text{PO}_3-$, $-\text{NHCOO}-$, $-\text{COO}-$, $-\text{NHCOCH}_2\text{O}-$, and $-\text{NH}-\text{CO}-\text{NH}-$; wherein R is substituted or unsubstituted C1-C10 alkyl, $-(\text{C}=\text{O})-\text{R}'$, $-(\text{C}=\text{O})\text{NH}-\text{R}'$, $-\text{NH}(\text{C}=\text{O})-\text{R}'$, $-\text{SO}_2-\text{R}'$, $-\text{NHSO}_2-\text{R}'$, $-\text{SO}_2\text{NH}-\text{R}'$, $-\text{SO}-\text{R}'$, $-\text{NHSO}-\text{R}'$, $-\text{SONH}-\text{R}'$, $-\text{PO}_3-\text{R}'$, $-\text{NHCOO}-\text{R}'$, $-\text{COO}-\text{R}'$ or $-\text{NH}-\text{CO}-\text{NH}-\text{R}'$, or $-\text{X}'-\text{L3}-\text{Z}$;

wherein L3 is linker, and Z is a peptide element or target molecule T; wherein R' is selected from the group consisting of H, substituted or unsubstituted C1-C6 alkyl, substituted or unsubstituted C3-C8 cycloalkyl, protecting group, and $-\text{X}'-\text{L3}-\text{Z}$;

X' is selected from the group consisting of $-\text{O}-$, $-\text{S}-$, $-\text{NH}-$, $-(\text{C}=\text{O})\text{NH}-$, $-\text{NH}(\text{C}=\text{O})-$, $-\text{SO}_2-$, $-\text{NHSO}_2-$, $-\text{SO}_2\text{NH}-$, $-\text{SO}-$, $-\text{NHSO}-$, $-\text{SONH}-$, $-\text{PO}_3-$, $-\text{NHCOO}-$, $-\text{COO}-$, $-\text{COOCH}_2-$, and $-\text{NH}-\text{CO}-\text{NH}-$;

each L2 and L3 is independently linker selected from the group consisted of

(a) substituted or unsubstituted 5-20 membered carbon chain;

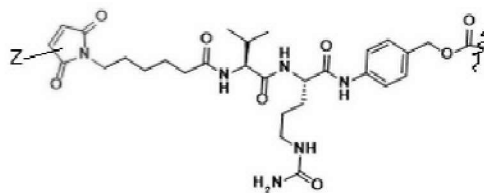
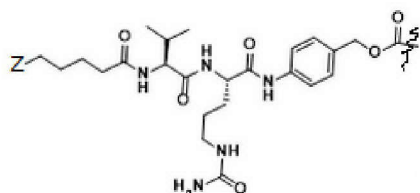
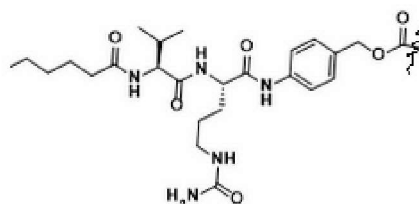
(b) substituted or unsubstituted 5-20 membered heterocarbon chain containing 1 to 5 heteroatoms selected from N, O and S;

(c) substituted or unsubstituted 5-20 membered carbon chain; and 1 to 4 chain atoms in the chain are replaced with heterocyclic ring selected from the group consisting of 4-10 membered saturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S, and 5-10 membered partially unsaturated or fully unsaturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S;

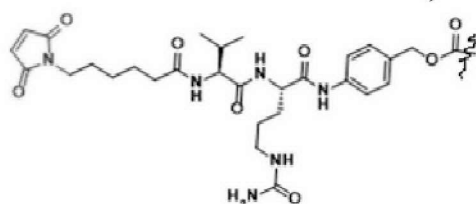
(d) substituted or unsubstituted 5-20 membered heterocarbon chain containing 1 to 5 heteroatoms selected from N, O and S; and 1 to 4 chain atoms in the chain are replaced with heterocyclic ring selected from the group consisting of 4-10 membered saturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S, and 5-10 membered partially unsaturated or fully unsaturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S;

wherein the "substituted" means one or more (preferably, 1, 2, 3 or 4) hydrogen atoms in the group is substituted with substituent selected from the group consisting of C1-C10 alkyl, C3-C8 cycloalkyl, $-\text{COOH}$,

-OH, -SH, -NH₂, -NHR, -N(R₁)R₂, -Z, -X'-L3-Z,

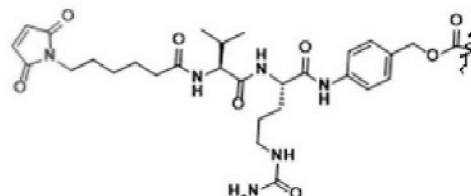
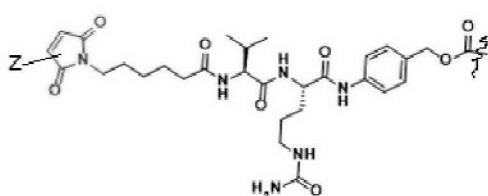
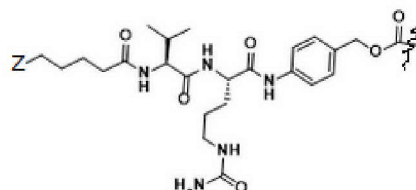
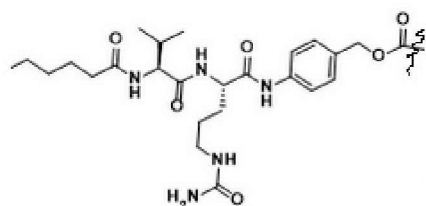


, and



; wherein Z is a polypeptide element or target molecule T;

wherein when the substituent is -Z, -Z is located only on N atom; wherein each R₁ and R₂ is indenpedently H, C1-C6 alkyl, C3-C8 cycloalkyl, amino protecting group, -X'-L3-Z,



, or

; wherein R is

substituted or unsubstituted C1-C10 alkyl, -(C=O)-R', (C=O)NH-R', -NH(C=O)-R', -SO₂-R', -NHSO₂-R', -SO₂NH-R', -SO-R', -NHSO-R', -SONH-R', -PO₃-R', -NHCOO-R', -COO-R', or -NH-CO-NH-R',

-NH-CO-O-R', or -X'-L3-Z; wherein L3 is linker, and Z is a polypeptide element or target molecule T; wherein R' is selected from the group consisting of H, substituted or unsubstituted C1-C6 alkyl, substituted or unsubstituted C3-C8 cycloalkyl, protecting group, and X'-L3-Z.

2. The conjugate of Claim 1, wherein L2 is linker selected from the group consisted of

(a) substituted 5-20 membered carbon chain;

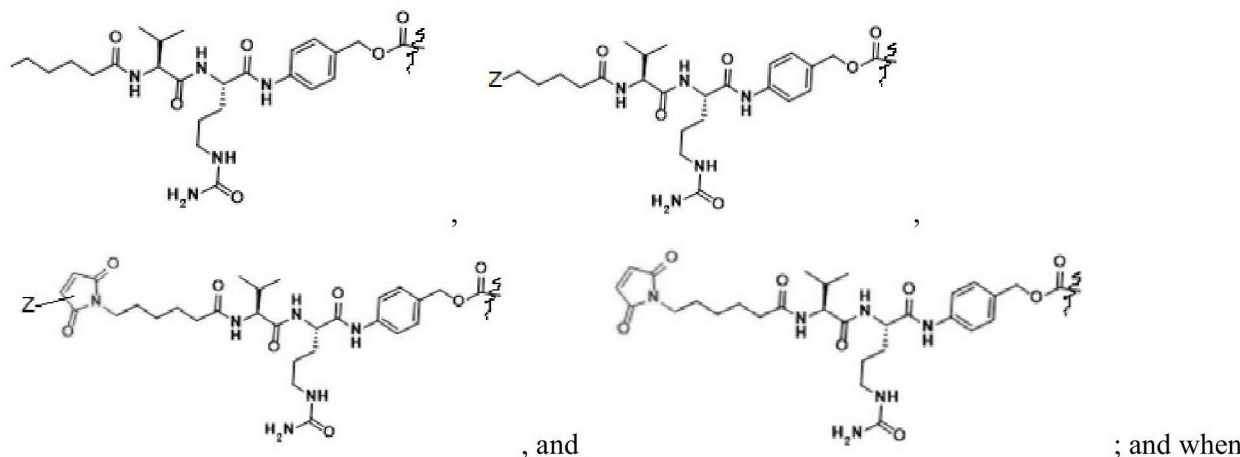
(b) substituted 5-20 membered heterocarbon chain containing 1 to 5 heteroatoms selected from N, O and S;

(c) substituted 5-20 membered carbon chain; and 1 to 4 chain atoms in the chain are replaced with heterocyclic ring selected from the group consisting of 4-10 membered saturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S, and 5-10 membered partially unsaturated or fully unsaturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S;

(d) substituted 5-20 membered heterocarbon chain containing 1 to 5 heteroatoms selected from N, O and S; and 1 to 4 chain atoms in the chain are replaced with heterocyclic ring selected from the group

consisting of 4-10 membered saturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O and S, and 5-10 membered partially unsaturated or fully unsaturated heterocyclic rings containing 1- 4 heteroatoms selected from N, O and S;

wherein, the “substituted” means at least one hydrogen atom in the group is substituted with substituent selected from the group consisting of -NHR, -N(R₁)R₂, -Z, -X-L3-Z,



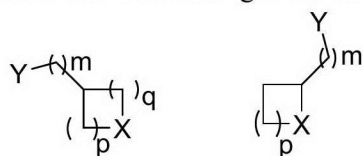
the substituent is -Z, -Z is located only on N atom;

and if the remaining hydrogen atoms are substituted, the definition of substituted is the same as that of claim 1.

3. The conjugate of Claim 1, wherein L2 and/or L3 are linkers selected from the group consisted of
 - (a) substituted or unsubstituted -(CH₂)_s;
 - (b) substituted or unsubstituted -(CH₂)_s-, wherein 1-5 of -CH₂- are replaced with O;
 - (c) substituted or unsubstituted -(CH₂)_s-, wherein 1-2 of -CH₂- are replaced with NR and optionally 1-2 of -CH₂- are replaced with O; wherein R is H, amino protecting group or X'-L3-Z;
 - (d) substituted or unsubstituted -(CH₂)_s-, wherein 1-4 of -CH₂- are replaced with 5 membered unsaturated heterocyclic ring containing 1-4 heteroatoms selected from N, O, S and P;
 - (e) substituted or unsubstituted -(CH₂)_s-, wherein 1-4 of -CH₂- are replaced with 4-6 membered saturated heterocyclic ring containing 1- 4 heteroatoms selected from N, O, S and P;
 - (f) substituted or unsubstituted -(CH₂)_s-, wherein 1-4 of -CH₂- are replaced with O and 4-6 membered saturated heterocyclic ring containing 1-4 heteroatoms selected from N, O, S and P;
 - (g) divalent 4-6 membered heterocyclic group containing 1-2 nitrogen atoms;

wherein s is an integer of 5-20.

4. The conjugate of Claim 1, wherein when X is NH or NR, X and part of L2 chain to which X is attached are combined together to form a 4-8 membered heterocyclic ring,



wherein Y and R are defined as above; m is an integer of 1-15; each p and q is independently 0, 1, 2, or 3, and p + q ≥ 2.

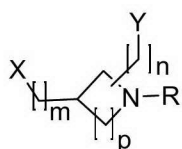
5. The conjugate of Claim 1, wherein L1 is selected from the group consisted of

in the formula, "Y, X" means this terminal is X or Y; with the proviso that one end is X, and the other end is Y;

X and Y are defined as above, each m and n is independently an integer of 1-15;

each T, V, Q and W is independently CH, C=O, S, O, NH or NR; R is C1-C10 alkyl, -(C=O)R',
 5 -(C=O)NH-R', -NH(C=O)-R', -SO₂-R', -NHSO₂-R', -SO₂NH-R', -SO-R', -NHSO-R', -SONH-R', -PO₃-R',
 -NHCOO-R', -COO-R', -NH-CO-NH-R', -NH-CO-O-R', or N-X'-L3-Z; wherein L3 is linker, and Z is a
 polypeptide element or target molecule T; R' and X' are defined as in Claim 1 or 2;

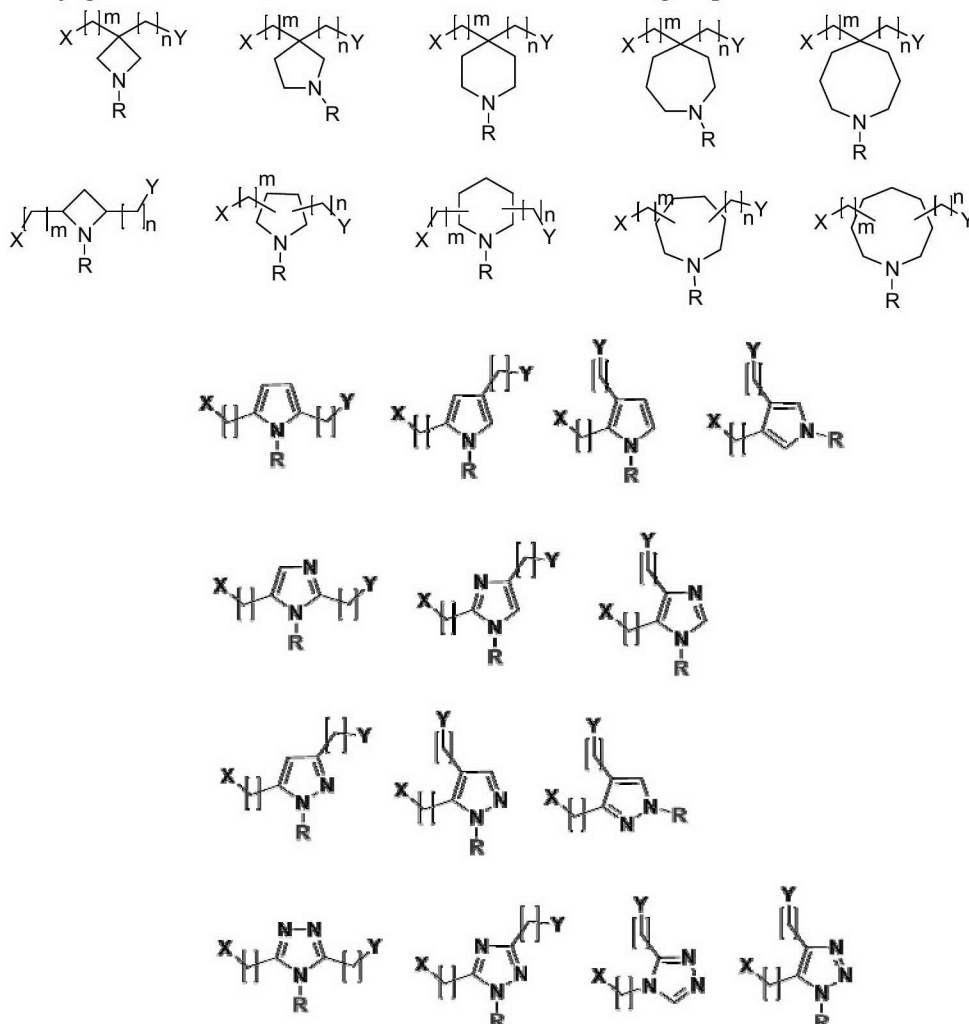
or, L1 is



10 wherein X, Y and R are defined as above;

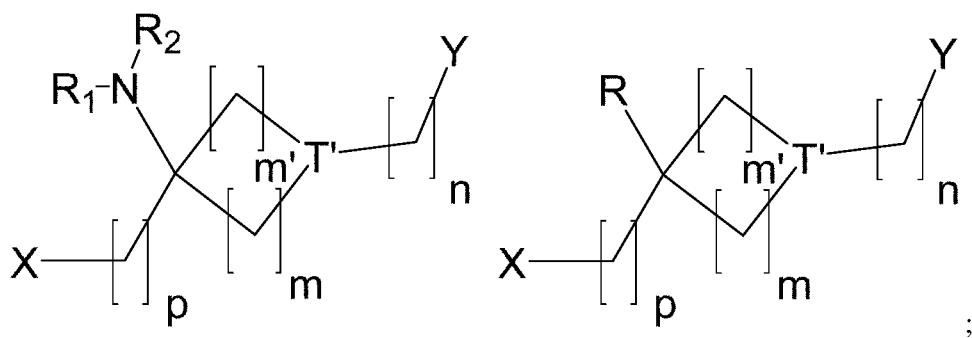
each m and n is independently an integer of 1-15; p is 1, 2 or 3.

6. The conjugate of Claim 1, wherein L1 is selected from the group consisted of



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7. The conjugate of Claim 1, wherein L1 is selected from the group consisting of

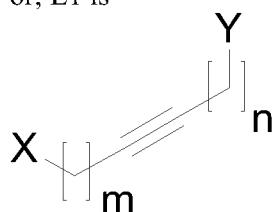


wherein each m' and m is independently 0, 1, 2, 3, 4, or 5 (preferably 0, 1, 2, or 3), and m' and m are not 0 at the same time;

each p and n is independently an integer of 1-15; T' is N or CR_b; wherein R_b is a group selected from the group consisting of H, substituted or unsubstituted C1-C6 alkyl;

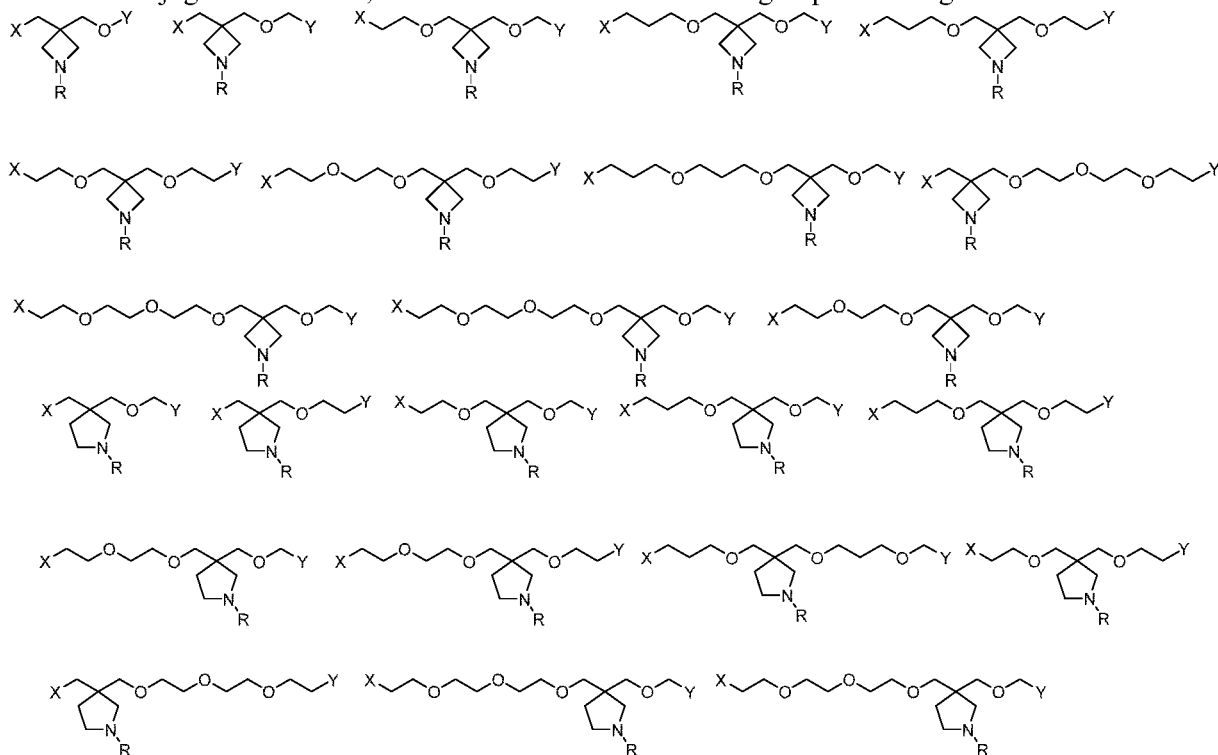
R, R₁ and R₂ are as defined in claim 1 or 2;

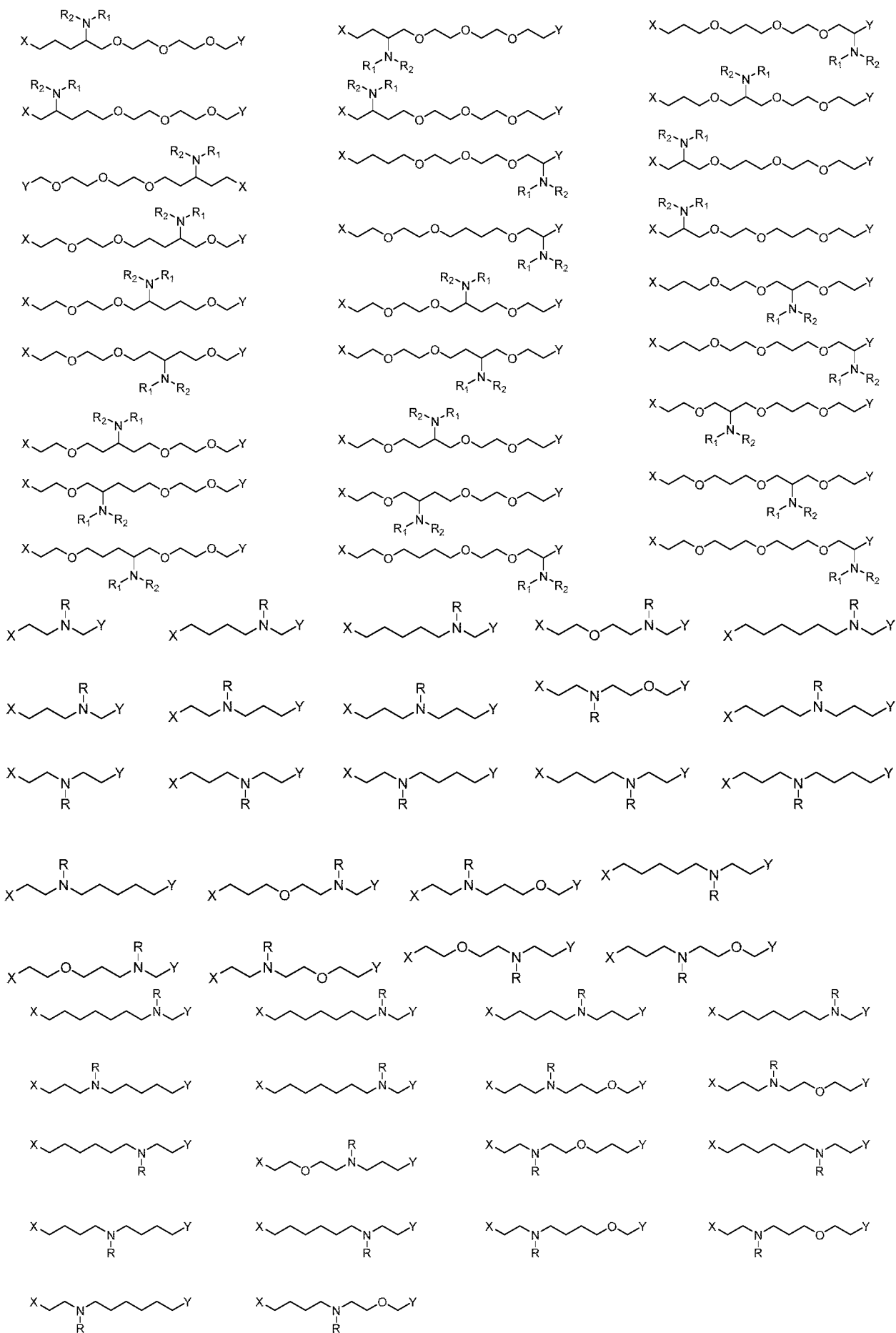
or, L1 is

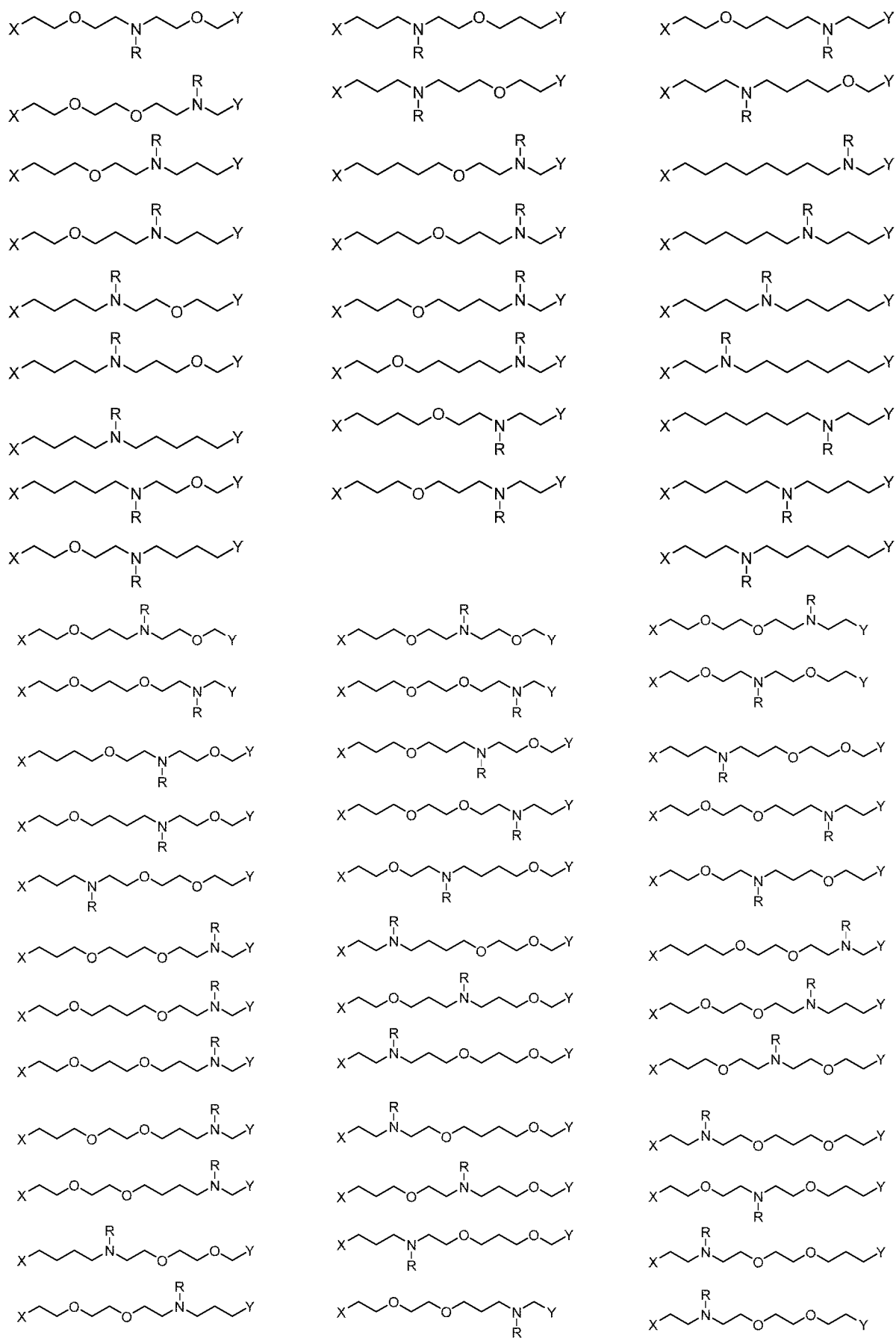


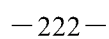
wherein each m and n is independently an integer of 1-15.

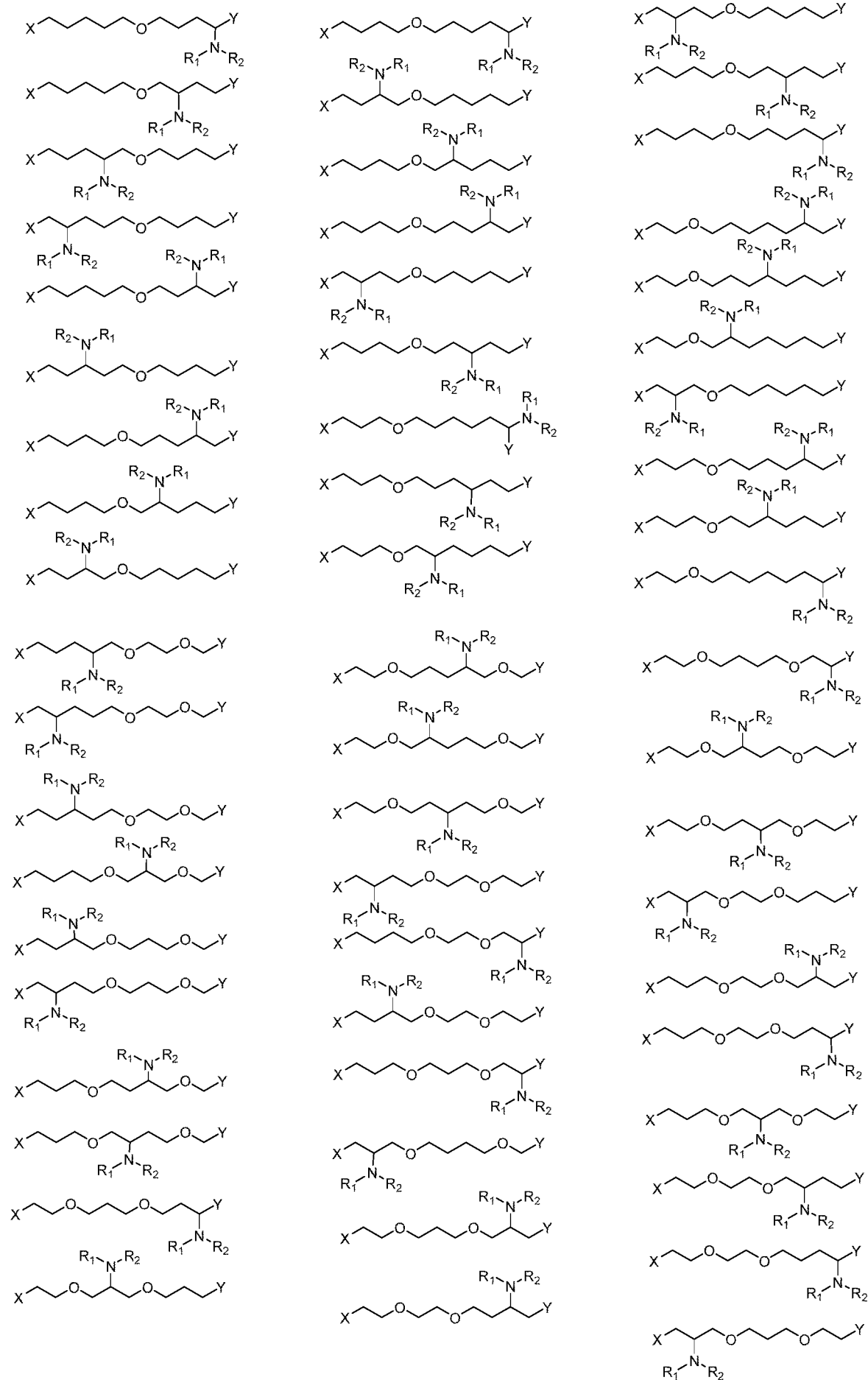
8. The conjugate of Claim 1, wherein L1 is selected from the group consisting of

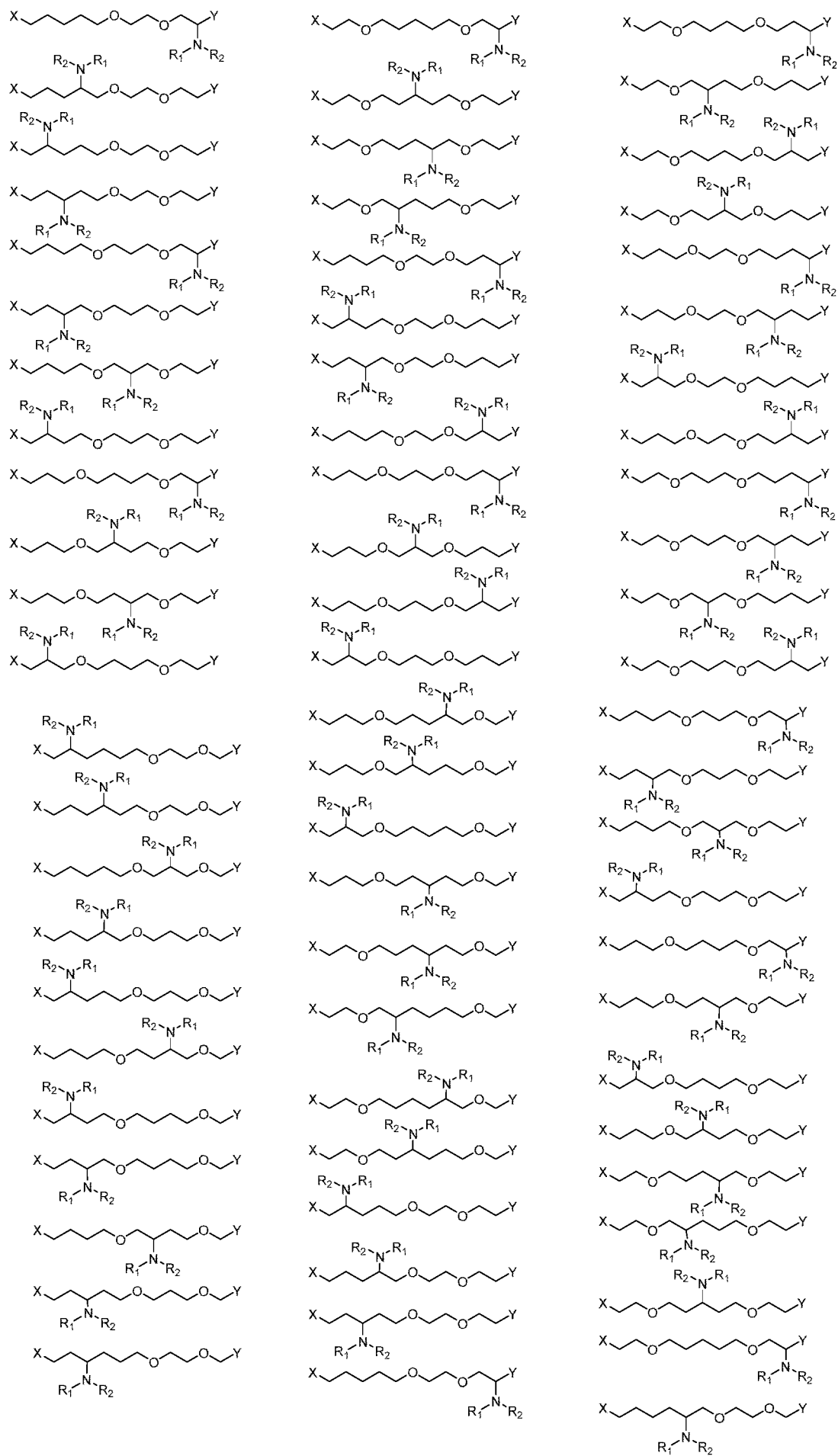


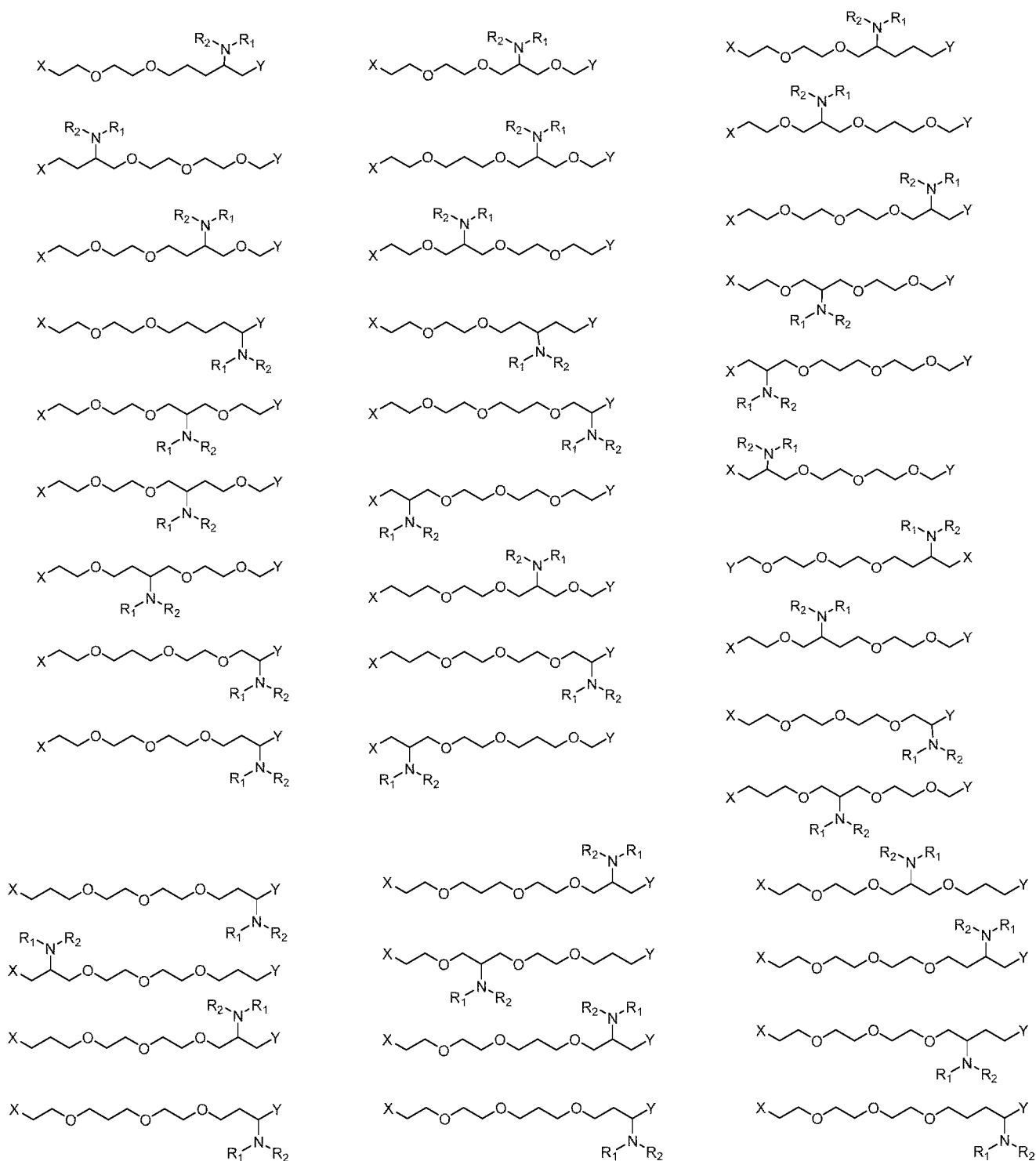










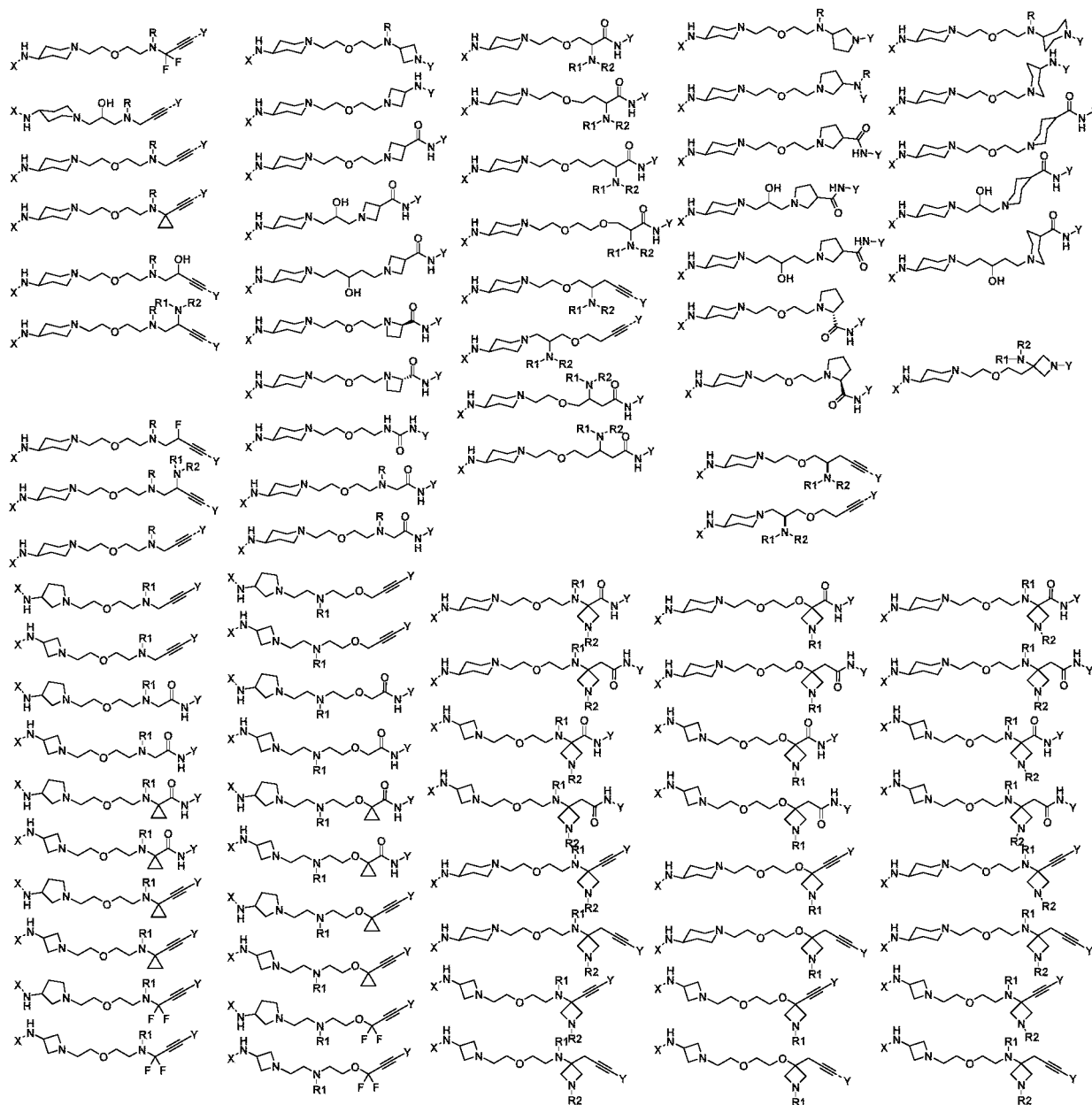


wherein, R or one of R₁ and R₂ is -X'-L3-Z;

the other one of R₁ and R₂, and X', X and Y are as defined in claim 1.

9. The conjugate of Claim 1, wherein L1 is selected from the group consisting of

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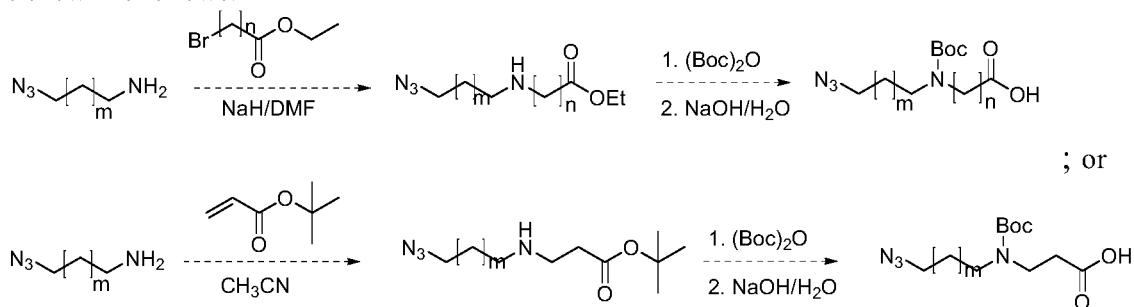


wherein, R or one of R_1 and R_2 is $-X'-L3-Z$;

the other one of R_1 and R_2 , and X' , X and Y are as defined in claim 1.

- 5 10. A pharmaceutical composition wherein the pharmaceutical composition comprises the conjugate of any one of Claims 1-8 and pharmaceutically acceptable carrier.

11. A preparation method of the conjugate of any one of Claims 1-8, wherein the reaction route of the method is shown as follows:



10 wherein m, and n are defined as above.

12. A use of the conjugate of any one of Claims 1-9 in the preparation of pharmaceutical compositions

for the treatment of diseases associated with excess of the target protein.

13. A method for reducing the content of target proteins in a cell, wherein the cell is contacted with the conjugate of any one of Claims 1-9, thereby reducing the content of the target proteins in the cell.

5 14. A method for reducing the content of target proteins in a subject or treating diseases associated with excess of target proteins, wherein comprising step of administering the conjugate of any one of Claims 1-9 to a subject in need thereof.

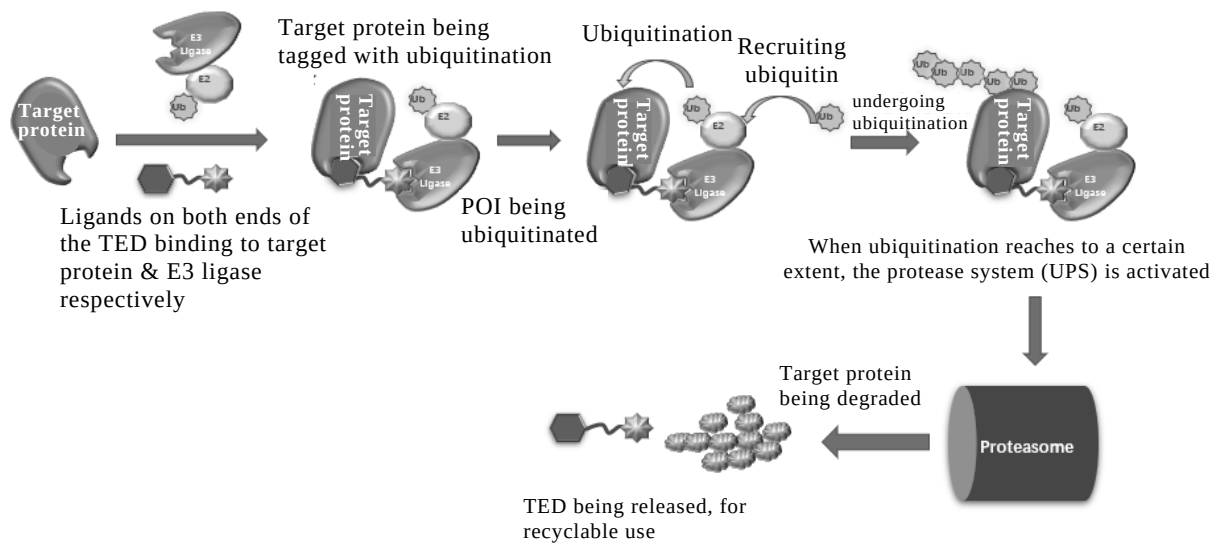


Figure 1

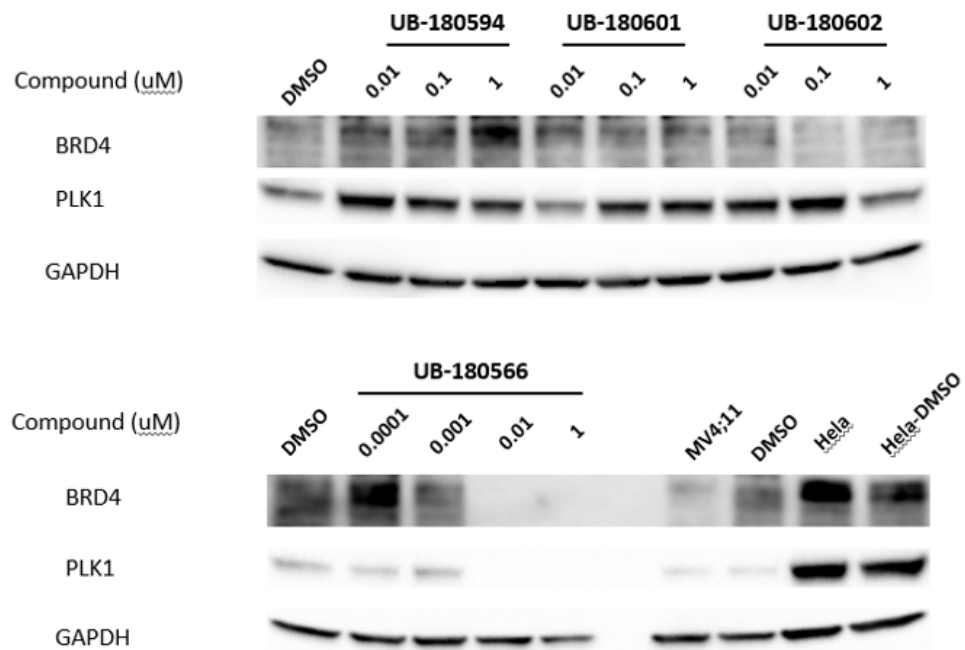


Figure 2

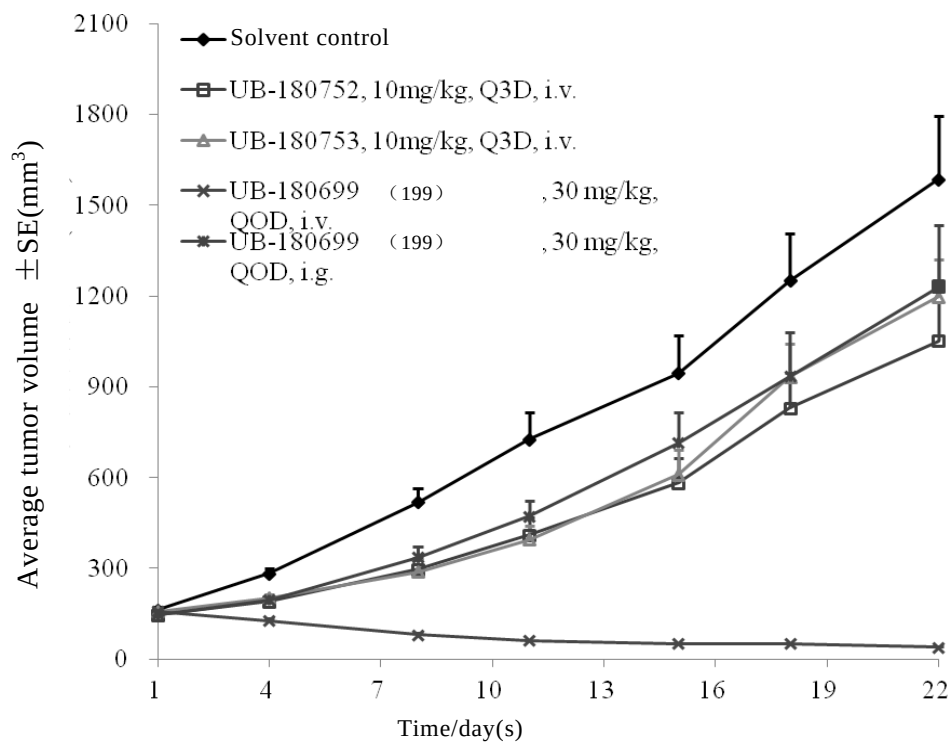


Figure 3

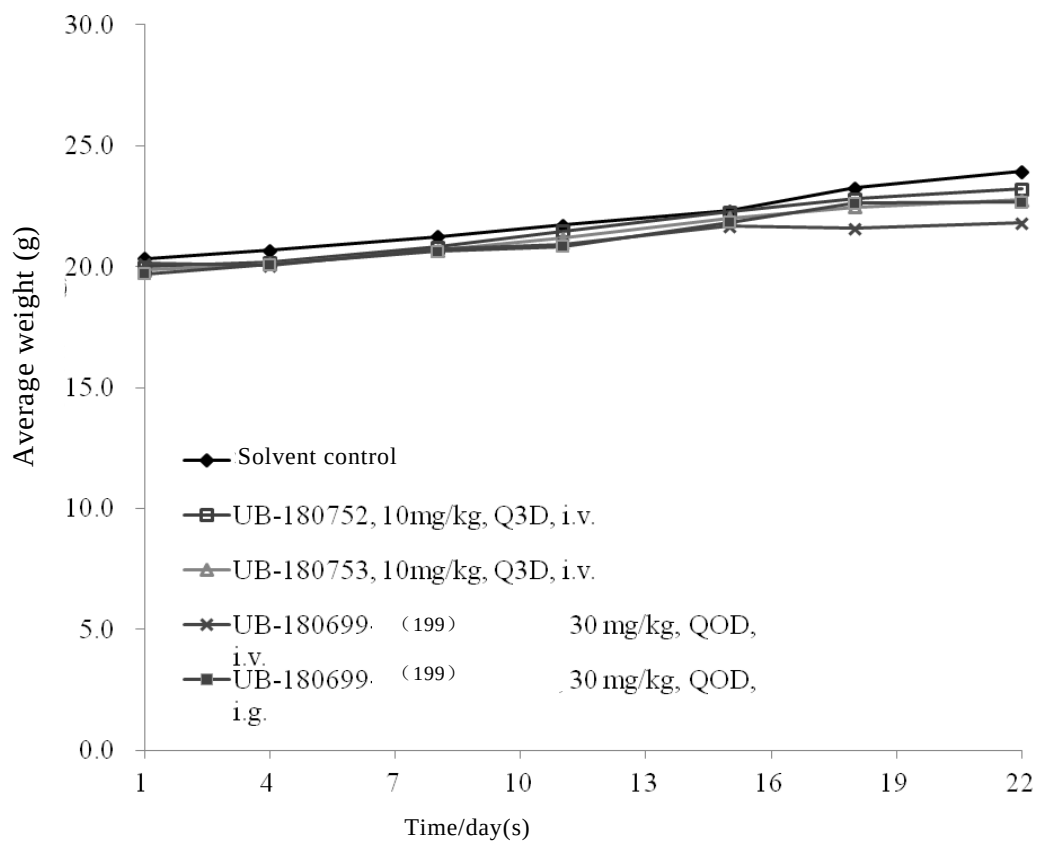


Figure 4