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Fortsættes ...

DESCRIPTION

TECHNICAL FIELD OF THE INVENTION

[0001] The invention generally relates to peptides that bind Tissue Factor Pathway Inhibitor (TFPI) and uses thereof.

BACKGROUND OF THE INVENTION

[0002] Hemostasis relies on the complex coagulation cascade, wherein a series of events mediated by blood clotting factors leads to conversion of prothrombin to thrombin. Factor X (FX) activation is the central event of both the intrinsic and extrinsic pathways of the coagulation cascade. The extrinsic pathway has been proposed as the primary activator of the coagulation cascade (Mackman et al., Arterioscler. Thromb. Casc. Biol., 27, 1687-1693 (2007)). Circulating Tissue Factor (TF) and activated Factor VII (FVIIa) interact to form the "extrinsic complex," which mediates activation of FX. The coagulation cascade is amplified by the intrinsic pathway, during which successive activation of factors XII, XI, IX, and VIII results in formation of the "intrinsic" FIXa-FVIIIa complex that also mediates FX activation. Activated FX promotes thrombin formation, which is required for the body to create fibrin and effectively curb bleeding.

[0003] Severe bleeding disorders, such as hemophilia, result from disruption of the blood coagulation cascade. Hemophilia A, the most common type of hemophilia, stems from a deficiency in factor VIII, while hemophilia B is associated with deficiencies in Factor IX (FIX). Hemophilia C is caused by a deficiency in Factor XI (FXI) (Cawthern et al., Blood, 91(12), 4581-4592 (1998)). There is currently no cure for hemophilia and other clotting diseases. Factor replacement therapy is the most common treatment for blood coagulation disorders. However, blood clotting factors typically are cleared from the bloodstream shortly after administration. To be effective, a patient must receive frequent intravenous infusions of plasma-derived or recombinant factor concentrates, which is uncomfortable, requires clinical settings, is expensive, and is time consuming. In addition, therapeutic efficacy of factor replacement therapy can diminish drastically upon formation of inhibitory antibodies. Approximately 30% of patients with severe hemophilia A develop inhibitory antibodies that neutralize Factor VIII (FVIII) (Peerlinck and Hermans, Haemophilia, 12, 579-590 (2006)). Few therapeutic options exist for patients with anti-Factor antibodies.

[0004] Thus, there exists a need in the art for compositions and methods for treating blood coagulation disorders. The invention provides such compositions and methods.

SUMMARY OF THE INVENTION

[0005] The invention provides a TFPI-binding peptide complex comprising a first peptide and a second peptide, wherein the first peptide comprises the structure of formula (XIII): X6001-X6002-X6003-X6004-X6005-X6006-X6007-X6008-X6009-X6010-X6011-X6012-X6013- X6014-X6015-X6016-X6017-X6018-X6019-X6020 (XIII) (SEQ ID NO: 3153); wherein X6001 is an amino acid selected from the group consisting of F, L, M, Y, 1Ni, Thi, Bta, Dopa, Bhf, C, D, G, H, I, K, N, Nmf, Q, R, T, V, and W; wherein X6002 is an amino acid selected from the group consisting of Q, G, and K; wherein X6003 is an amino acid selected from the group consisting of C, D, E, M, Q, R, S, T, Ede(O), Cmc, A, Aib, Bhs, F, G, H, I, K, L, N, P, V, W and Y; wherein X6004 is an amino acid selected from the group consisting of Aib, E, G, I, K, L, M, P, R, W, Y, A, Bhk, C, D, F, H, k, N, Nmk, Q, S, T and V; wherein X6005 is an amino acid selected from the group consisting of a,

A, Aib, C, D, d, E, G, H, K, k, M, N, Nmg, p, Q, R, NpropylG, aze, pip, tic, oic, hyp, nma, Ncg, Abg, Apg, thz, dtc, Bal, F, L, S, T, V, W and Y; wherein X6006 is an amino acid selected from the group consisting of A, C, C(NEM), D, E, G, H, K, M, N, Q, R, S, V, Cit, C(Acm), Nle, I, Ede(O), Cmc, Ecl, Eea, Eec, Eef, Nif, Eew, Aib, Btq, F, L, T, W and Y; wherein X6007 is an amino acid selected from the group consisting of I, V, T, Chg, Phg, Tie, A, F, G, K, L, Nmv, P, Q, S, W and Y; wherein X6008 is an amino acid selected from the group consisting of F, H, 1Ni, 2Ni, Pmy, Y, and W; wherein X6009 is an amino acid selected from the group consisting of Aib, V, Chg, Phg, Abu, Cpg, Tie, L-2-amino-4,4,4-trifluorobutyric acid, A, f, I, K, S, T and V; wherein X6010 is an amino acid selected from the group consisting of A, C, D, d, E, F, H, K, M, N, P, Q, R, S, T, V, W, Y, Nmd, C(NEM), Aib, G, I, L and Nmf; wherein X6011 is an amino acid selected from the group consisting of A, a, G, p, Sar, c, hcy, Aib, C, K, and Nmg; wherein X6012 is an amino acid selected from the group consisting of Y, Tym, Pty, Dopa and Pmy; wherein X6013 is an amino acid selected from the group consisting of Aib, C, F, 1Ni, Thi, Bta, A, E, G, H, K, L, M, Q, R, W and Y; wherein X6014 is an amino acid selected from the group consisting of A, Aib, C, C(NEM), D, E, K, L, M, N, Q, R, T, V, Hcy, Bhe, F, G, H, I, P, S, W and Y; wherein X6015 is an amino acid selected from the group consisting of R, (omega-methyl)- R, D, E and K; wherein X6016 is an amino acid selected from the group consisting of L, Hcy, Hle and Aml; wherein X6017 is an amino acid selected from the group consisting of A, a, Aib, C, c, Cha, Dab, Eag, Eew, H, Har, Hci, Hle, I, K, L, M, Nle, Nva, Opa, Orn, R, S, Deg, Ebc, Eca, Egz, Aic, Apc, Egt, (omega-methyl)-R, Bhr, Cit, D, Dap, E, F, G, N, Q, T, V, W and Y; wherein X6018 is an amino acid selected from the group consisting of A, Aib, Hcy, hcy, C, c, L, Nle, M, N, R, Bal, D, E, F, G, H, I, K, Q, S, T, V, W and Y; wherein X6019 is an amino acid selected from the group consisting of K, R, Har, Bhk and V; and wherein X6020 is an amino acid selected from the group consisting of K, L, Hcy, Aml, Aib, Bhl, C, F, G, H, I, Nml, Q, R, S, T, V, W and Y; and the second peptide comprises the structure of formula (XIV): X7001-X7002-X7003-X7004-X7005-X7006-[X7007-X7008-X7009-X7010-X7011-X7012- X7013-X7014-X7015-X7016-X7017-X7018]-X7019-X7020-X7021-X7022-X7023 (XIV) (SEQ ID NO: 3154), wherein X7001 is either present or absent, whereby in case X7001 is present it is an amino acid selected from the group consisting of A, C, C(NEM), D, E, F, G, H, I, K, L, P, R, S, T, V and W; wherein X7002 is either present or absent, whereby in case X7002 is present it is an amino acid selected from the group consisting of A, C, C(NEM), D, E, F, G, H, I, K, L, M, P, Q, R, S, T, V, W and Y; wherein X7003 is an amino acid selected from the group consisting of A, F, I, K, L, R, S, T, V, W and Y; wherein X7004 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, R, S, T, V and W; wherein X7005 is R or W; wherein X7006 is an amino acid selected from the group consisting of F, H, I, K, L, R, V and W; wherein X7007 is an amino acid selected from the group consisting of Orn, homoK, C, Hcy, Dap and K, preferably selected from the group consisting of C and Hcy; wherein X7008 is an amino acid selected from the group consisting of A, G, R, S and T; wherein X7009 is an amino acid selected from the group consisting of a, A, I, K, L, M, m, Moo, Nle, p, R, Sem and V; wherein X7010 is an amino acid selected from the group consisting of A, G, I, K, L, P, R, S, T and V; wherein X7011 is an amino acid selected from the group consisting of D, E, G, S and T; wherein X7012 is an amino acid selected from the group consisting of A, a, D, d, E, e, F, f, G, I, K, k, L, I, M, m, Moo, Nle, nle, P, p, R, r, S, s, Sem, T, t, V, v, W and w; wherein X7013 is an amino acid selected from the group consisting of A, C, C(NEM), Con, Con(Meox), D, d, E, e, Eag, F, G, I, K, L, N, R, S, s, T, V and W; wherein X7014 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, M, R, S, T, V and W; wherein X7015 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, M, Nle, R, S, T, V and W; wherein X7016 is an amino acid selected from the group consisting of A, D, E, F, I, K, L, M, Moo, Nle, R, S, Sem, T, V, W and Y; wherein X7017 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, R, S, T, V, W and Y; wherein X7018 is an amino acid selected from the group consisting of C and D, preferably C; wherein X7019 is an amino acid selected from the group consisting of A, F, I, L, S, T, V and W; wherein X7020 is an amino acid selected from the group consisting of F and W; wherein X7021 is an amino acid selected from the group consisting of I, L and V; wherein X7022 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, P, R, S, T, V and W; wherein X7023 is either present or absent, whereby in case X7023 is present it is an amino acid selected from the group consisting of A, C, C(NEM), Con, Con(Meox), D, E, Eag, F, G, I, K, L, R, S, T, V, W and Y; wherein the peptide comprises as a cyclic structure generated by a linkage between X7007 and X7018; wherein the C-terminus of the first peptide is linked to the N-terminus of the second peptide via a linker moiety; wherein the first peptide

comprises an amino acid sequence at least 80%, at least 85%, at least 90%, at least 95%, or 100% identical to SEQ ID NO: 178 or SEQ ID NO: 4261; and wherein the second peptide comprises an amino acid sequence at least 80%, at least 85%, at least 90%, at least 95%, or 100% identical to SEQ ID NO: 1044.

[0006] The invention also provides a peptide complex of the invention, for use in a method for the treatment of a subject.

[0007] The invention also provides a pharmaceutical composition comprising a peptide complex of the invention and a pharmaceutically acceptable carrier, and optionally further comprising a further pharmaceutically effective agent.

[0008] . In various embodiments, the peptide complex binds to at least two different TFPI epitopes and, optionally, inhibits at least two functions of TFPI. In some embodiments, the peptide complex of the invention binds TFPI-1 (e.g., TFPI-1α) and, optionally, improves TFPI-regulated thrombin generation in the absence of FVIII, FIX, and/or FXI. A composition (e.g., a pharmaceutical composition) comprising the peptide complex also is provided.

[0009] The invention provides an *in vitro* method of inhibiting a TFPI comprising contacting the TFPI with a peptide complex as described herein. The invention also provides a peptide complex, for use in a method of enhancing thrombin formation in a clotting factor-deficient subject, for use in a method for increasing blood clot formation in a subject, and for use in a method of treating a blood coagulation disorder in a subject. The methods comprise administering to the subject a peptide complex as provided herein in an amount effective to achieve a desired effect, e.g., an amount effective to enhance thrombin formation, an amount effective to enhance blood clot formation, or an amount effective to treat the blood coagulation disorder in the subject. Further aspects of the invention include use of the peptide complex of the invention, for the manufacture of a medicament, an *in vitro* method for targeting a cell displaying TFPI, for use in a method for treating or diagnosing a subject suffering from a disease or at risk of suffering from a disease, a method of purifying TFPI, and a method of identifying a TFPI-binding compound. Unless explicitly indicated to the contrary, the description provided herein with respect to one peptide complex of the invention or method of the invention applies to each and every peptide complex of the invention and method of the invention, respectively.

[0010] The following numbered paragraphs each succinctly define one or more exemplary variations of the invention:

1. 1. A peptide complex comprising a first peptide and a second peptide, wherein
    1. (a) the first peptide comprises the structure of formula (XIII):

    X6001-X6002-X6003-X6004-X6005-X6006-X6007-X6008-X6009-X6010-X6011-X6012-X6013-X6014-X6015-X6016-X6017-X6018-X6019-X6020 (XIII) (SEQ ID NO: 3153) ;

    wherein X6001 is an amino acid selected from the group consisting of F, L, M, Y, 1Ni, Thi, Bta, Dopa, Bhf, C, D, G, H, I, K, N, Nmf, Q, R, T, V, and W; wherein X6002 is an amino acid selected from the group consisting of Q, G, and K; wherein X6003 is an amino acid selected from the group consisting of C, D, E, M, Q, R, S, T, Ede(O), Cmc, A, Aib, Bhs, F, G, H, I, K, L, N, P, V, W and Y; wherein X6004 is an amino acid selected from the group consisting of Aib, E, G, I, K, L, M, P, R, W, Y, A, Bhk, C, D, F, H, k, N, Nmk, Q, S, T and V; wherein X6005 is an amino acid selected from the group consisting of a, A, Aib, C, D, d, E, G, H, K, k, M, N, Nmg, p, Q, R, NpropylG, aze, pip, tic, oic, hyp, nma, Ncg, Abg, Apg, thz, dtc, Bal, F, L, S, T, V, W and Y; wherein X6006 is an amino acid selected from the group consisting of A, C, C(NEM), D, E, G, H, K, M, N, Q, R, S, V, Cit, C(Acm), Nle, I, Ede(O), Cmc, Ecl, Eea, Eec, Eef, Nif, Eew, Aib, Btq, F, L, T, W and Y; wherein X6007 is an amino acid selected from the group consisting of I, V, T, Chg, Phg, Tie, A, F, G, K, L, Nmv, P, Q, S, W and Y; wherein X6008 is an amino acid selected from the

group consisting of F, H, 1Ni, 2Ni, Pmy, Y, and W; wherein X6009 is an amino acid selected from the group consisting of Aib, V, Chg, Phg, Abu, Cpg, Tie, L-2-amino-4,4,4-trifluorobutyric acid, A, f, I, K, S, and T; wherein X6010 is an amino acid selected from the group consisting of A, C, D, d, E, F, H, K, M, N, P, Q, R, S, T, V, W, Y, Nmd, C(NEM), Aib, G, I, L and Nmf; wherein X6011 is an amino acid selected from the group consisting of A, a, G, p, Sar, c, hcy, Aib, C, K, G and Nmg; wherein X6012 is an amino acid selected from the group consisting of Y, Tym, Pty, Dopa and Pmy; wherein X6013 is an amino acid selected from the group consisting of Aib, C, F, 1Ni, Thi, Bta, A, E, G, H, K, L, M, Q, R, W and Y; wherein X6014 is an amino acid selected from the group consisting of A, Aib, C, C(NEM), D, E, K, L, M, N, Q, R, T, V, Hcy, Bhe, F, G, H, I, P, S, W and Y; wherein X6015 is an amino acid selected from the group consisting of R, (omega-methyl)-R, D, E and K; wherein X6016 is an amino acid selected from the group consisting of L, Hcy, Hle and Aml; wherein X6017 is an amino acid selected from the group consisting of A, a, Aib, C, c, Cha, Dab, Eag, Eew, H, Har, Hci, Hle, I, K, L, M, Nle, Nva, Opa, Orn, R, S, Deg, Ebc, Eca, Egz, Aic, Apc, Egt, (omega-methyl)-R, Bhr, Cit, D, Dap, E, F, G, N, Q, T, V, W and Y; wherein X6018 is an amino acid selected from the group consisting of A, Aib, Hcy, hcy, C, c, L, Nle, M, N, R, Bal, D, E, F, G, H, I, K, Q, S, T, V, W and Y; wherein X6019 is an amino acid selected from the group consisting of K, R, Har, Bhk and V; and wherein X6020 is an amino acid selected from the group consisting of K, L, Hcy, Aml, Aib, Bhl, C, F, G, H, I, Nml, Q, R, S, T, V, W and Y, and

2. (b) the second peptide comprises the structure of formula (XIV):

X7001-X7002-X7003-X7004-X7005-X7006-[X7007-X7008-X7009-X7010-X7011-X7012-X7013-X7014-X7015-X7016-X7017-X7018]-X7019-X7020-X7021-X7022-X7023 (XIV) (SEQ ID NO: 3154),

wherein X7001 is either present or absent, whereby in case X7001 is present it is an amino acid selected from the group consisting of A, C, C(NEM), D, E, F, G, H, I, K, L, P, R, S, T, V and W; wherein X7002 is either present or absent, whereby in case X7002 is present it is an amino acid selected from the group consisting of A, C, C(NEM), D, E, F, G, H, I, K, L, M, P, Q, R, S, T, V, W and Y; wherein X7003 is an amino acid selected from the group consisting of A, F, I, K, L, R, S, T, V, W and Y; wherein X7004 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, R, S, T, V and W; wherein X7005 is R or W; wherein X7006 is an amino acid selected from the group consisting of F, H, I, K, L, R, V and W; wherein X7007 is an amino acid selected from the group consisting of Orn, homoK, C, Hcy, Dap and K, preferably selected from the group consisting of C and Hcy; wherein X7008 is an amino acid selected from the group consisting of A, G, R, S and T; wherein X7009 is an amino acid selected from the group consisting of a, A, I, K, L, M, m, Moo, Nle, p, R, Sem and V; wherein X7010 is an amino acid selected from the group consisting of A, G, I, K, L, P, R, S, T and V; wherein X7011 is an amino acid selected from the group consisting of D, E, G, S and T; wherein X7012 is an amino acid selected from the group consisting of A, a, D, d, E, e, F, f, G, I, K, k, L, 1, M, m, Moo, Nle, nle, P, p, R, r, S, s, Sem, T, t, V, v , W and w; wherein X7013 is an amino acid selected from the group consisting of A, C, C(NEM), Con, Con(Meox), D, d, E, e, Eag, F, G, I, K, L, N, R, S, s, T, V and W; wherein X7014 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, M, R, S, T, V and W; wherein X7015 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, M, Nle, R, S, T, V and W; wherein X7016 is an amino acid selected from the group consisting of A, D, E, F, I, K, L, M, Moo, Nle, R, S, Sem, T, V, W and Y; wherein X7017 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, R, S, T, V, W and Y; wherein X7018 is an amino acid selected from the group consisting of C and D, preferably C; wherein X7019 is an amino acid selected from the group consisting of A, F, I, L, S, T, V and W; wherein X7020 is an amino acid selected from the group consisting of F and W; wherein X7021 is an amino acid selected from the group consisting of I, L and V; wherein X7022 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, P, R, S, T, V and W; wherein X7023 is either present or absent, whereby in case X7023 is present it is an amino acid selected from the group

consisting of A, C, C(NEM), Con, Con(Meox), D, E, Eag, F, G, I, K, L, R, S, T, V, W and Y; and wherein the peptide comprises as a cyclic structure generated by a linkage between X7007 and X7018; wherein the C-terminus of the first peptide is linked to the N-terminus of the second peptide via a linker moiety; wherein the first peptide comprises an amino acid sequence at least 80%, at least 85%, at least 90%, at least 95%, or 100% identical to SEQ ID NO: 178 or SEQ ID NO: 4261; and wherein the second peptide comprises an amino acid sequence at least 80%, at least 85%, at least 90%, at least 95%, or 100% identical to SEQ ID NO: 1044.

2. 2. The peptide complex of paragraph 1,
wherein X6001 is an amino acid selected from the group consisting of 1Ni, Bta, Dopa, F, L, Y and M; wherein X6002 is Q; wherein X6003 is an amino acid selected from the group consisting of D, E, S, M, Q, R, T and C; wherein X6004 is an amino acid selected from the group consisting of K, Aib, L, P, R, E, G, I, Y, M and W; wherein X6005 is an amino acid selected from the group consisting of p, Nmg, NpropylG, aze, pip, tic, oic, hyp, a, Aib, D, d, G, H, K, k, N, Q, R, A, E, C and M; wherein X6006 is an amino acid selected from the group consisting of C, E, K, R, S, V, C(Acm), Nle, C(NEM), I, Cit, A, D, G, H, N, Q and M; wherein X6007 is an amino acid selected from the group consisting of Tie, V and I; wherein X6008 is an amino acid selected from the group consisting of H, 1Ni, 2Ni, Pmy, F and Y; wherein X6009 is V, Abu or Tle; wherein X6010 is an amino acid selected from the group consisting of D, P, C, T, A, E, K, M, N, Q, R, F, H, S, V, W and Y; wherein X6011 is G, a, c, hcy or Sar; wherein X6012 is Y; wherein X6013 is an amino acid selected from the group consisting of F, 1Ni, Bta and C; wherein X6014 is an amino acid selected from the group consisting of Aib, C, E, Hcy, A, D, K, L, M, N, Q, R, T, V and Aib; wherein X6015 is R; wherein X6016 is an amino acid selected from the group consisting of L, Aml, Hle and Hcy; wherein X6017 is an amino acid selected from the group consisting of A, Aib, C, c, Aic, Eca, Deg, Cha, Dab, Dap, Eag, Eew, H, Har, Hci, Hle, K, Nle, Nva, Opa, Orn, R, I, L, S and M; wherein X6018 is an amino acid selected from the group consisting of A, Aib, C, c, L, Hcy, N, M and R; wherein X6019 is K; and wherein X6020 is an amino acid selected from the group consisting of L, Aml, Hcy and K.

3. 3. The peptide complex of paragraph 1 or paragraph 2, wherein the first peptide and/or second peptide further comprises N-terminal amino acid(s) and/or moieties linked to X6001 and/or X7001 and selected from the group consisting of FAM-Ttds, PE, Palm, 2-phenyl acetyl, 3-phenyl propionyl, 2-(naphth-2-yl) acetyl, hexanoyl, 2-methyl propionyl, 3-methyl butanoyl, 2-naphthylsulfonyl, 1-naphthylsulfonyl, acetyl, Con, Con(Meox), AOA, Oxme-AOA, Meox-Lev, levulinic acid (Lev), and pentynoic acid (Pyn).

4. 4. The peptide complex of any one of paragraphs 1-3, wherein the first peptide and/or second peptide further comprises X6021 linked to X6020 or X7024 linked to X7023, respectively, wherein X6021 and/or X7024 comprises C-terminal amino acid(s) and/or moieties selected from the group consisting of Hly, K, Orn, Dab, Eag, Dap, Hcy, Pen, C, c, C(NEM), Con, Con(Meox), K(Ttds-maleimidopropionyl(EtSH)), K(Tdts-maleimid), K(AOA), K(Myr), K(Ttds-Myr), K(Ttds-Palm), K(Ttds-Ac), K(Ttds-yGlu-Myr), K(AlbuTag), K(4PBSA), Cea, and amide.

5. 5. The peptide complex of any one of paragraphs 1-4,
wherein X7001 is an amino acid selected from the group consisting of A, D, F, G, H, K, L and S; wherein X7002 is an amino acid selected from the group consisting of H, F, M and R; wherein X7003 is an amino acid selected from the group consisting of F and Y; wherein X7004 is K; wherein X7005 is W; wherein X7006 is an amino acid selected from the group consisting of F and H; wherein X7007 is C; wherein X7008 is an amino acid selected from the group consisting of A, G and S; wherein X7009 is an amino acid selected from the group consisting of M, Sem and V; wherein X7010 is an amino acid selected from the group consisting of K, P and R; wherein X7011 is D; wherein X7012 is an amino acid selected from the group consisting of F, L, 1, M and Sem; wherein X7013 is an amino acid selected from the group consisting of D, G, K and S; wherein X7014 is G; wherein X7015 is an amino acid selected from the group consisting of I and T; wherein X7016 is an amino acid selected from the group consisting of D, F, M, Sem and Y; wherein X7017 is an amino acid selected from the group consisting of S and T; wherein X7018 is C; wherein X7019 is an amino acid selected from the group consisting of

A and V; wherein X7020 is W; wherein X7021 is V; wherein X7022 is an amino acid selected from the group consisting of F, L, K, R, P and W; wherein X7023 is either present or absent, whereby in case X7023 is present it is an amino acid sequence selected from the group consisting of A, D, F, M, S and Y; and wherein the peptide comprises as a cyclic structure generated by a linkage between X7007 and X7018.

6. 6. The peptide complex of any one of paragraphs 2-5 wherein the first peptide and the second peptide is linked by linker moiety, preferably about 1-100 Å in length.
7. 7. The peptide complex of paragraph 6, wherein the linker moiety is about 5-50 Å in length.
8. 8. The peptide complex of paragraph 7, wherein the linker moiety is about 10-30 Å in length.
9. 9. The peptide complex of any one of paragraphs 6-8, wherein the linker moiety comprises the structure Z_{1-20}, wherein Z is an amino acid, hydroxy acid, ethylene glycol, propylene glycol, or a combination of any of the foregoing.
10. 10. The peptide complex of paragraph 9, wherein Z is G, s, S, a, A, Bal, Gaba, Ahx, Ttds, or a combination of any of the foregoing.
11. 11. The peptide complex of any one of paragraphs 6-10, wherein the linker moiety is attached to the first peptide and/or the second peptide via an oxime, a hydrazide, a succinimide, a thioether, a triazole, a secondary amine, an amide, or a disulfide.
12. 12. The peptide complex of any one of paragraphs 6-11, wherein the C-terminus of the first peptide is linked to the N-terminus of the second peptide via the linker moiety.
13. 13. The peptide complex of any one of paragraphs 6-12, wherein the first peptide comprises the structure of formula (XIII), and linker moiety attaches to the first peptide at the N-terminus, at the C-terminus, or at side chains of X6001, X6004, X6006, X6010, X6014, or X6020.
14. 14. The peptide complex of paragraph 1 or paragraph 2 comprising the amino acid sequence set forth in SEQ ID NO: 4260.
15. 15. The peptide complex of any one of paragraphs 1-13, wherein the peptide complex is conjugated to a polyethylene glycol (PEG) moiety, human serum albumin (HSA), an HSA-binding domain, an antibody or fragment thereof, hydroxyethyl starch, a proline-alanine-serine multimer (PASylation), a C_{12}-C_{18} fatty acid, or polysialic acid.
16. 16. A peptide complex of any one of paragraphs 1-15 for use in a method for the treatment of a subject.
17. 17. The peptide complex paragraph 16, wherein the method is for the treatment of a blood coagulation disorder.
18. 18. Use of the peptide complex of any one of paragraphs 1-15 for the manufacture of a medicament.
19. 19. Use of the peptide complex of any one of paragraphs 1-15 for the manufacture of a medicament for the treatment of a blood coagulation disorder.
20. 20. A pharmaceutical composition comprising the peptide complex of any one of paragraphs 1-15 and a pharmaceutically acceptable carrier.
21. 21. The pharmaceutical composition of paragraph 20, wherein the composition comprises a further pharmaceutically effective agent.
22. 22. The pharmaceutical composition of paragraph 21, wherein the pharmaceutical composition is for use in a method of treating a blood coagulation disorder.

DESCRIPTION OF THE FIGURES

[0011]

Figure 1 is an illustration of the blood coagulation cascade.

Figure 2 is an illustration of the secondary structure of Tissue Factor Pathway Inhibitor-1.

Figure 3 is an illustration of the formation of a quaternary complex comprising Tissue Factor, Factor Xa (FXa), Factor VIIa (FVIIa), and TFPI.

Figure 4 is a listing of amino acid sequences of various TFPI-binding peptides (e.g., TFPI-inhibitory peptides) denoting amino acid substitutions (bolded and underlined) in reference to peptide JBT0293.

Figure 5 is an illustration of mRNA display selection of TFPI-binding peptides (e.g., TFPI-inhibitory peptides).

Figure 6A is an illustration of the EC_{50} binding ELISA and Figure 6B is an illustration of the IC_{50} ELISA described in Example 1.

Figure 7 is a binding ELISA curve comparing % OD (y-axis) and concentration [nM] (x-axis) for biotinylated peptide JBT0132.

Figures 8A-8D are competition ELISA curves comparing % OD (y-axis) and concentration [nM] (x-axis) for exemplary peptides of the invention.

Figures 9A and 9B are sensorgrams plotting RU (y-axis) against time in seconds (x-axis) for peptides JBT0120 and JBT0132.

Figures 10A and 10B are sensorgrams plotting RU (y-axis) against time in seconds (x-axis) for peptide JBT0120 interaction with Tissue Factor Pathway Inhibitor-1 and Tissue Factor Pathway Inhibitor-2.

Figures 11A and 11B are graphs comparing amount of thrombin generated (nM) (y-axis) and time in minutes (x-axis) for peptide JBT0120 and peptide JBT0132 in a plasma-based assay.

Figures 12-18 are tables listing the amino acid sequences of various TFPI-binding peptides; EC_{50} and percent inhibition of TFPI observed in the FXa inhibition assay; EC_{50} and percent inhibition of TFPI observed in the extrinsic tenase inhibition assay; and FEIBA, Factor VIII (FVIII) Immunate, or Factor IX (FIX) equivalent activities (mU/mL) in plasma-based assays. "*" denotes negative controls.

Figures 19-21 are tables listing the results from BIAcore analysis of several TFPI-binding peptides. "*" denotes negative controls.

Figures 22-30 are tables listing the amino acid sequences of various TFPI-binding peptides; EC_{50} and percent inhibition of TFPI observed in the FXa inhibition assay; EC_{50} and percent inhibition of TFPI observed in the extrinsic tenase inhibition assay; and FEIBA, FVIII Immunate, or FIX equivalent activities (mU/mL) in plasma-based assays. "*" denotes negative controls.

Figure 31 is a graph comparing a pharmacokinetic characteristic (concentration of peptide (y-axis) versus time after administration (x-axis)) of a PEGylated TFPI-binding peptide to the pharmacokinetic characteristic of same peptide lacking PEG. The peptides were administered intravenously to C57Bl6 mice at a dose of 10 mg/kg. Three biological samples were analyzed for the presence of peptide at each time point.

Figures 32-39 are tables listing the amino acid sequences and IC_{50} or EC_{50} values of various peptides of the invention. "*" denotes negative controls.

Figure 40 is a graph illustrating a pharmacokinetic characteristic (concentration of peptide (nM) (y-axis) versus time after administration (minutes) (x-axis)) of a PEGylated TFPI-binding peptide following subcutaneous administration to mice at a dose of 10 mg/kg.

Figure 41 is a graph correlating the amount of thrombin generated (nM) (y-axis) with time (minutes) (x-axis) for peptide JBT1855 in a plasma-based assay of hemophilia A patient plasma.

Figure 42 is a graph illustrating the amount of blood loss (µl; y-axis) observed following a nail-clip in mice treated with JBT-1855 (intravenous or subcutaneous administration), anti-TFPI antibody (intravenous administration), or vehicle (intravenous administration) (x-axis).

Figure 43 is a graph plotting TFPI160 amino acid residue (x-axis) against the chemical shift differences of HSQC signals for free TFPI160 and TFPI160 bound to JBT0303 (y-axis).

Figure 44 is a ribbon model of the secondary structure of TFPI illustrating regions of chemical shift changes of HSQC signals when TFPI160 is complexed to JBT0303 compared to uncomplexed (free) TFPI160.

Figure 45 is a graph plotting TFPI160 amino acid residue (x-axis) against the chemical shift differences of HSQC signals for free TFPI160 and TFPI160 bound to JBT0122 (y-axis).

Figure 46 is a ribbon model of the secondary structure of TFPI protein illustrating regions of chemical shift changes of HSQC signals when TFPI160 is complexed to JBT0122 compared to uncomplexed (free) TFPI160.

Figure 47 is a table listing assignments for the carbonyl carbon (C), the alpha carbon (CA), the beta carbon (CB), the amide proton (H), and the amide nitrogen (N) of JBT0788 based on HSQC, HNCACB, HNCA, HNCO and HNN spectra.

Figure 48 is a ribbon model of the secondary structure of free JBT0788.

Figure 49 is a table listing assignments for the carbonyl carbon (C), the alpha carbon (CA), the beta carbon (CB), the amide proton (H), and the amide nitrogen (N) of JBT0788 complexed with TFPI160 based on HSQC, HNCACB, HNCA, HCCOCA, and HNCO spectra.

Figure 50 is a ribbon model of the secondary structure of JBT0788 when complexed with TFPI160.

Figure 51 is a table listing assignments for the carbonyl carbon (C), the alpha carbon (CA), the beta carbon (CB), the amide proton (H), and the amide nitrogen (N) of JBT0616 based on HSQC, HNCACB, and HNN spectra.

Figure 52 is a ribbon model of the secondary structure of free JBT0616.

Figure 53 is a table listing assignments for the carbonyl carbon (C), the alpha carbon (CA), the beta carbon (CB), the amide proton (H), and the amide nitrogen (N) of JBT0616 complexed with TFPI based on HSQC, HNCO, HNCA, and HNCOCA spectra.

Figure 54 is a ribbon model of the secondary structure of JBT0616 when complexed with TFPI160.

Figure 55 is a ribbon structure of the energetically minimized best model of KD1 (residues 22-79) in complex with JBT0303 with residues proposed to drive the protein-protein interaction displayed as sticks. Italicized and underlined residues belong to JBT0303; the remaining residues belong to KD1 of TFPI.

Figure 56 is a rotational thromboelastogram correlating sample elasticity (mm) with time in seconds (s) for JBT2317.

Figure 57 is a rotational thromboelastogram correlating sample elasticity (mm) with time in seconds (s) for JBT2329.

Figure 58 is an illustration of a computing device.

Figure 59 is an illustration of a three dimensional (3D) model of a KD1 protein.

Figure 60 is an illustration of a 3D model of a TFPI-binding peptide.

Figure 61 is an illustration of a method of modeling protein and peptide interaction.

Figure 62 is a table listing the amino acid sequences and IC_{50} or EC_{50} values of various peptides of the invention. Designation "n.a." is "not analyzed." Progression curve data were obtained using the FXa inhibition assay described in Example 3 with recombinant human full length TFPI. Assay concentration of progression curve assay was 0.0025% (0.1% Tween80 used in peptide dilution buffer).

Figure 63 is a graph correlating concentration of peptides JBT2325-JBT2329 (nM) (y-axis) with time following intravenous administration (hours) (x-axis). Peptides comprising higher weight PEG moieties exhibited a prolonged *in vivo* half life in mice. Each time point is represented by the mean of three independent samples quantified by ELISA.

Figure 64A-64C are graphs correlating concentration of peptides JBT2401, JBT2404 and JBT2410 (nM) (y-axis) with time following intravenous administration (hours) (x-axis). Each time point is represented by the mean of three independent samples quantified by ELISA. Solid circles symbolize intravenous data, solid triangles symbolize subcutaneous data.

Figure 65 is a table listing the amino acid sequences of various peptides of the invention.

Figures 66A and 66B depicts IC_{50} curves of TFPI-binding peptides JBT1837 (SEQ ID NO: 1044) and JBT1857 (SEQ ID NO: 178) and peptide complex JBT2547 (SEQ ID NO: 4260) using two tracers: JBT2271 (SEQ ID NO: 4033), a biotinylated derivative of JBT1857 (Figure 66A), and JBT2316 (SEQ ID NO: 1313), a biotinylated derivative of JBT1837 (Figure 66B).

Figure 67 is a bar graph illustrating the results of a K_{off} assay with JBT2547 (SEQ ID NO: 4260) and JBT1857 (SEQ ID NO: 178) (X-axis) presented as % inhibition of TFPI binding by the tracer peptide, JBT2271 (SEQ ID NO: 4033) (Y-axis).

Figures 68A-F illustrate the results of FXa inhibition assays using a TFPI-binding peptide complex. Figures 68A-68D are graphs correlating concentration of JBT1837 (triangles; SEQ ID NO: 1044), JBT1857 (circles; SEQ ID NO: 178), JBT2547 (diamonds; SEQ ID NO: 4260), and JBT1837+JBT1857 (squares) (μM) (X-axis) with percent inhibition of TFPI in a FXa inhibition assay performed with 0.5 nM full length human TFPI (Figure 68A), 0.5 nM human TFPI 1-160 (Figure 68B), 0.5 nM murine TFPI 1-160 (Figure 68C), or 0.5 nM cynomologus money TFPI 1-160 (Figure 68D). Figure 68E illustrates TFPI inhibition by JBT2547 at increasing TFPI concentrations (0.1 to 10 nM from left to right) in a FXa inhibition assay. Data points were fitted by a sigmoidal dose response equation resulting in EC_{50}s (nM) and maximal inhibition (%). Figure 68F is a graph correlating TFPI inhibition (%) by JBT2547 (diamonds), JBT2548 (circles), JBT1837 (triangles) and a combination of JBT1837 and JBT1857 (squares) (1 μM peptide) in the presence of increasing full length human TFPI concentrations.

Figure 69 illustrates the results of extrinsic tenase inhibition assays using a TFPI-binding peptide complex. Figure 69A is a graph correlating concentration of JBT1837 (triangles; SEQ ID NO: 1044), JBT1857 (circles; SEQ ID NO: 178), JBT2547 (diamonds; SEQ ID NO: 4260), and JBT1837+JBT1857 (squares) (μM) (X-axis) with percent inhibition of TFPI in an extrinsic tenase inhibition assay performed with 0.063 nM full length human TFPI. Figure 69B illustrates TFPI inhibition by JBT2547 at increasing TFPI concentrations (0.031 to 10 nM from left to right) in an extrinsic tenase inhibition assay. Data points were fitted by a sigmoidal dose response equation resulting in EC_{50}s (nM) and maximal inhibition (%). Figure 69C is a graph correlating percent maximum inhibition of TFPI (Y-axis) mediated by JBT1837 (triangles; SEQ ID NO: 1044), JBT2548 (circles; SEQ ID NO: 4261), JBT2547 (diamonds; SEQ ID NO: 4260), and JBT1837+JBT1857 (squares) (1 μM) with concentration of full length human TFPI used in the extrinsic tenase inhibition assay. Figure 69D is a graph correlating EC_{50} of JBT1837 (triangles; SEQ ID NO: 1044), JBT2548 (circles; SEQ ID NO: 4261), JBT2547 (diamonds); SEQ ID NO: 4260, and JBT1837+JBT1857 (squares) (1 μM) with concentration (nM) of

full length human TFPI used in the extrinsic tenase inhibition assay. EC_{50}s were calculated by fitting of peptide concentrations response at increasing TFPI concentrations vs. full length TFPI concentrations.

Figures 70A-70C illustrate the results of a thrombin generation assay performed in FVIII-inhibited plasma. The figures correlate peak Factor IIa (thrombin) generation (nM) (Y-axis) with the concentration (nM) of JBT1837 (triangle), JBT1857 (circle), a combination of JBT1837+JBT1857 (square), and JBT2547 (diamond) (X-axis) in the presence of 1.25 nM flTFPI (Figure 70A), 3.75 nM flTFPI (Figure 70B), and 10 nM flTFPI (Figure 70C). Dotted line - pooled normal plasma (PNP); solid line - FVIII-inhibited plasma (PNP + aFVIII).

Figure 71A-71B are graphs correlating percent inhibition of TFPI (Y-axis) achieved at various concentrations (X-axis) of JBT2547, JBT1837, and 1857 (Figure 71A) or JBT2547 (diamonds), JBT1837 (triangles), JBT1837+JBT1857 (squares), and JBT2548 (circles) (Figure 71B) in a cell-based extrinsic tenase assay.

Figures 72A-72C illustrate the procoagulant effect of increasing concentrations (10, 100, 1000 nM) of JBT2547 (Figure 72A), JBT1837 (Figure 72B), or JBT1857 (Figure 72C) in FVIII-inhibited whole blood in absence of additional external human full length TFPI (open circles) and presence of increasing amounts of external full length TFPI (2 nM, closed triangles; 10 nM, closed squares). Clot times of FVIII-inhibited whole blood and normal whole blood are given as reference. + = no clot time achieved.

Figure 73 is a chart listing non-limiting examples of nucleophilic or electrophilic reactive groups for peptide linkage.

Figure 74 is a sequence alignment of TFPI KD1-KD2 from different species. Residues with a contact surface of 10-25 $Å^2$ = 41, 56, 59, 60, 67, 71, 74, 96, 106, 130, 132, 133, 136, 137, 142; residues with a contact surface of 26-60 $Å^2$ =75, 80, 82, 84, 85, 87, 145; residues with a contact surface of 61-100 $Å^2$ = 63, 65, 131, 146; residues with a contact surface of greater than 100 $Å^2$ = 83, 134.

Figure 75 is a ribbon structure of the a model of KD1-KD2 in complex with JBT1857 and JBT1837.

Figure 76A-76D is a table listing the amino acid sequences of various peptides of the invention.

Figure 77 is a table listing the amino acid sequences of various peptides of the invention.

Figures 78A-78AA is a table listing the amino acid sequences of various peptides of the invention.

DETAILED DESCRIPTION OF THE INVENTION

[0012] The invention provides peptide complexes. The peptide complex binds Tissue Factor Pathway Inhibitor-1 and, in some instances, blocks the inhibitory activity of Tissue Factor Pathway Inhibitor-1 (herein referred to as TFPI) within the blood coagulation cascade. Upon vascular injury, Tissue Factor (TF) complexes with Factor VIIa to form the "extrinsic complex" or "extrinsic tenase complex," which activates Factors IX and X (Figure 1). TFPI is the main natural regulator of TF/FVIIa extrinsic complex activity and by extension, plays a role in controlling thrombin generation (Panteleev et al., Eur. J. Biochem., 249, 2016-2031 (2002)). TFPI is a 43 kDa serine protease inhibitor comprising three Kunitz-type inhibitory domains (Figure 2). Kunitz domain 1 of TFPI binds FVIIa and Kunitz domain 2 binds FXa, enabling the inhibitor to form a quaternary FXa-TFPI-FVIIa-TF complex that blocks activity of the TF/FVIIa extrinsic complex (Figure 3). TFPI binding of FXa also downregulates the common pathway of the coagulation cascade, during which FXa converts prothrombin to thrombin (Audu et al., Anesth. Analg., 103(4), 841-845 (2006)). The invention provides, e.g., TFPI-inhibitory peptide complexes that block TFPI's inhibitory action on the blood coagulation cascade, thereby enhancing thrombin formation.

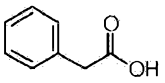
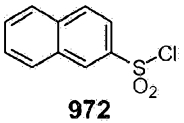
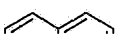
[0013] The amino acid sequences of several TFPI-binding peptides are provided herein. Conventional amino acids are identified according to their standard, one-letter or three-letter codes, as set forth in Table 1.

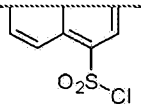
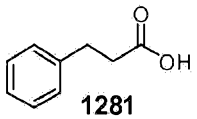
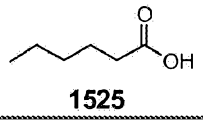
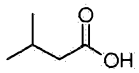
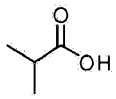
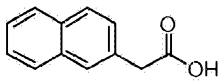
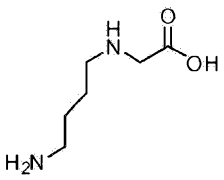
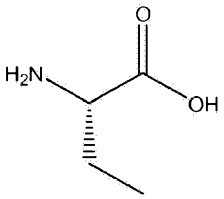
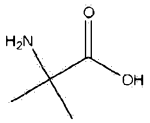
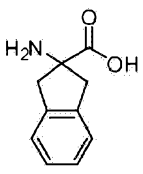
TABLE 1

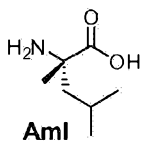
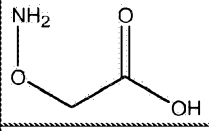
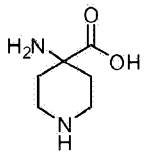
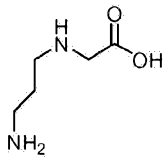
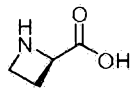
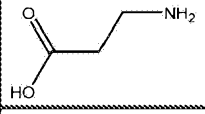
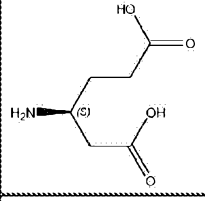
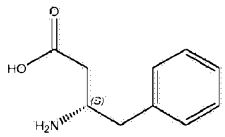
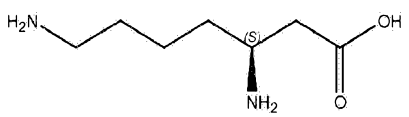
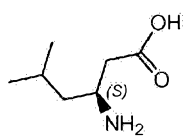
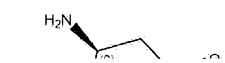
3-letter codes	1-letter code	Amino acids
Ala	A	Alanine
Cys	C	Cysteine
Asp	D	Aspartic acid
Glu	E	Glutamic acid
Phe	F	Phenylalanine
Gly	G	Glycine
His	H	Histidine
Ile	I	Isoleucine
Lys	K	Lysine
Leu	L	Leucine
Met	M	Methionine
Asn	N	Asparagine
Pro	P	Proline
Gln	Q	Glutamine
Arg	R	Arginine
Ser	S	Serine
Thr	T	Threonine
Val	V	Valine
Trp	W	Tryptophan
Tyr	Y	Tyrosine

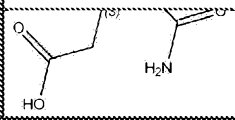
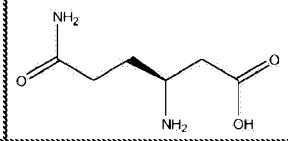
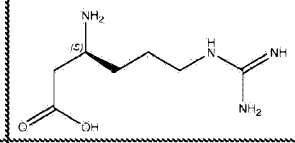
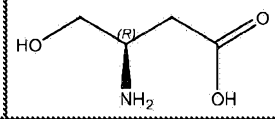
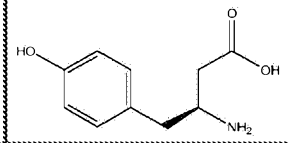
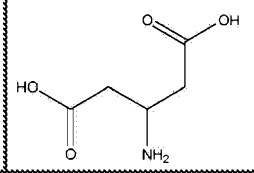
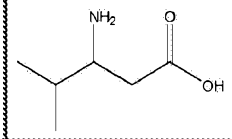
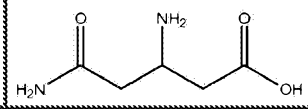
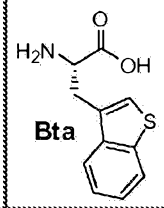
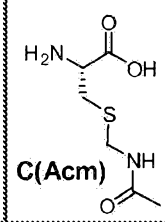
[0014] Examples of non-conventional amino acids and additional peptide building blocks are identified according to a three-letter code (with the exception of Ttds and Dopa, which are common four-letter abbreviations) found in Table 2. Additional building blocks designated by three-, four- or seven-number/letter designations or abbreviations also are listed in Table 2. The structures of some building blocks are depicted with an exemplary reagent for introducing the building block into a peptide (e.g., the structure provided for 2-naphthyl sulfonyl comprises a chloride).

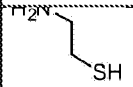
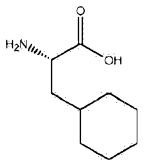
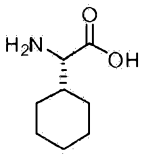
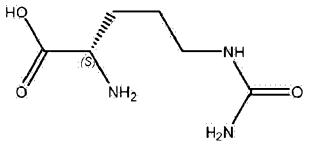
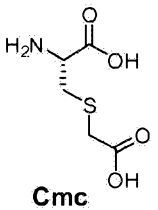
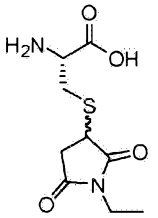
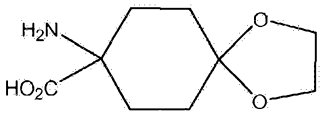
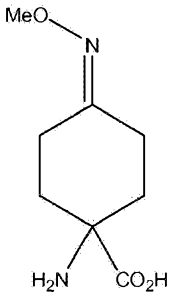
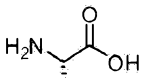
TABLE 2

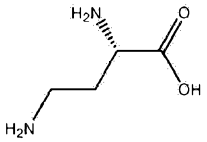
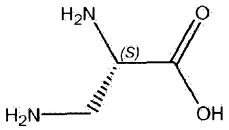
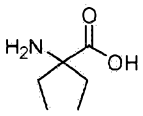
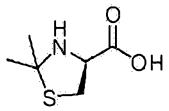
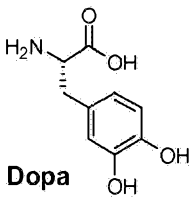
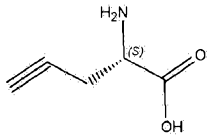
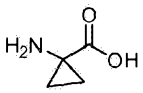
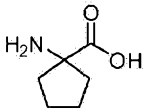
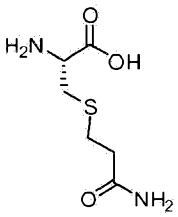
Name	Abbreviation	Structure
Phenyl acetyl	374	 **374**
2-Naphthyl sulfonyl	972	 **972**
1-Naphthyl sulfonyl	973	

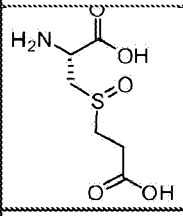
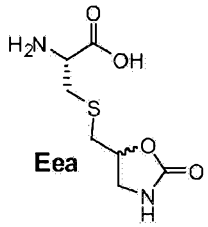
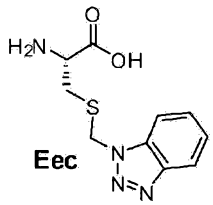
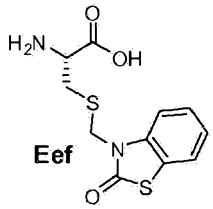
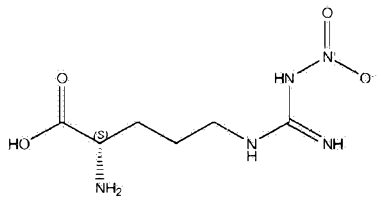
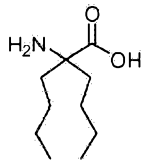
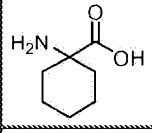
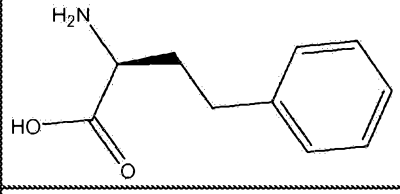
Name	Abbreviation	Structure
		 973
3-Phenyl propionyl	1281	 1281
Hexanoyl	1525	 1525
3-Methyl butanoyl	3067	 3067
2-Methyl propionyl	4635	 4635
2-(Naphth-2-yl) acetyl	5963	 5963
N-(4-aminobutyl)-glycine	Abg	
2-aminobutyric acid	Abu	
2-Amino-isobutyric acid	Aib	
2-Aminoindane-2-carboxylic acid	Aic	 Aic

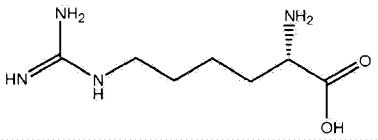
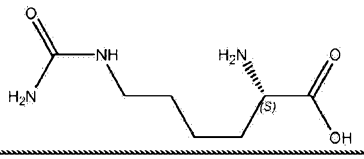
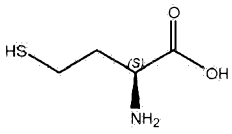
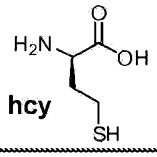
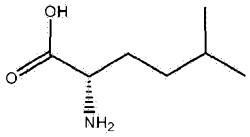
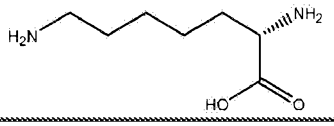
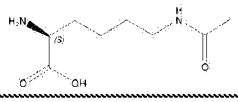
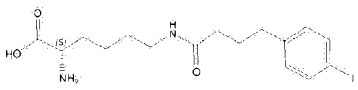
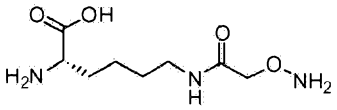
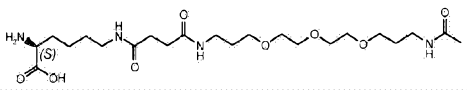
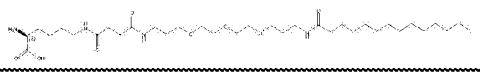
Name	Abbreviation	Structure
L-alpha-Methyl leucine	Aml	 Aml
Aminooxyacetic acid	AOA	
1-Amino-(4-N-piperidinyl)carboxylic acid	Apc	
N-(4-aminopropyl)-glycine	Apg	
D-Azetidine-2-carboxylic acid	aze	
β-Alanine	Bal	
β-Homoglutamatic acid	Bhe	
β-Homophenylalanine	Bhf	
β-Homo lysine	Bhk	
β-Homoleucine	Bhl	
β-Homoasparagine	Bhn	

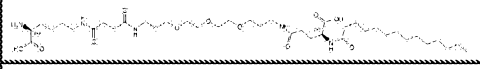
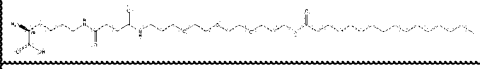
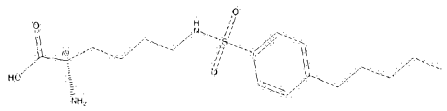
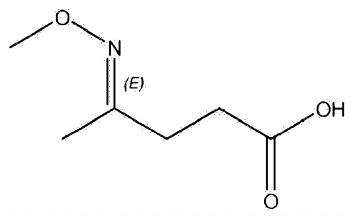
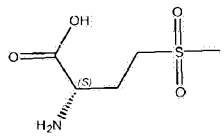
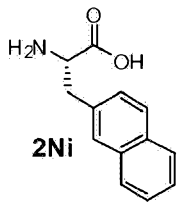
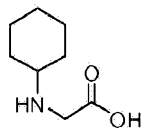
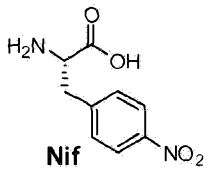
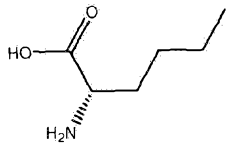
Name	Abbreviation	Structure
		
β-Homoglutamine	Bhq	
β-Homoarginine	Bhr	
β-Homoserine	Bhs	
β-Homotyrosine	Bhy	
β-Homoaspartic acid	Bhd	
β-Homovaline	Bhv, Btl	
β-Homoasparagin	Bhn, Btq	
L-3-Benzothienylalanine	Bta	
3-(Acetylamino-methylsulfanyl)-2-amino-propionic acid	C(Acm)	
Aminoethylthiol	Cea	

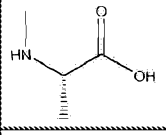
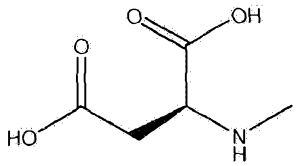
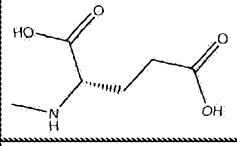
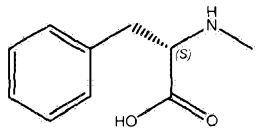
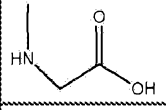
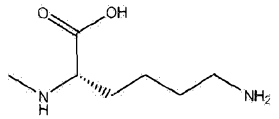
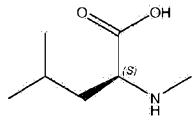
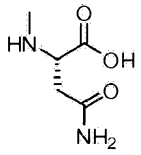
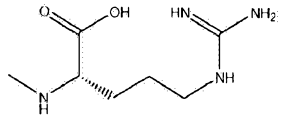
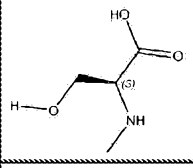
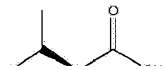
Name	Abbreviation	Structure
		
(S)-Cyclohexylalanine	Cha	
L-Cyclohexylglycine	Chg	
(S)-Citrullin	Cit	
Carboxymethylen cystein	Cmc	 Cmc
N-ethylmaleiimido cysteine	C(NEM)	
8-amino-1,4-dioxa-spiro[4,5]decane-8-carboxylic acid	Con	
1-Amino-(4-methoximino)-cyclohexane-1-carboxlic acid	Con(Meox)	
L-Cyclopentylglycine	Cpg	

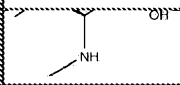
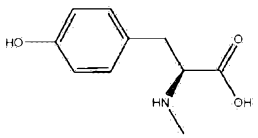
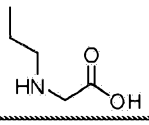
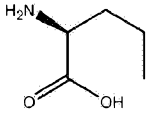
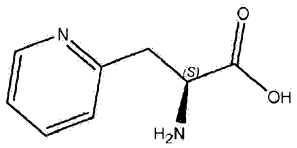
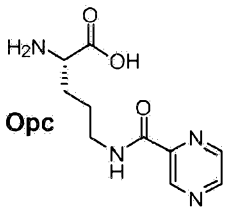
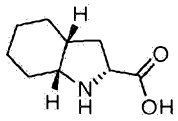
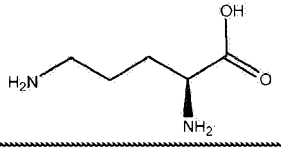
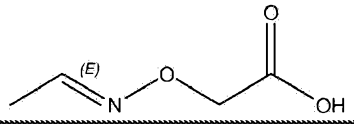
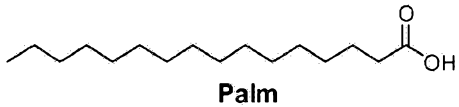
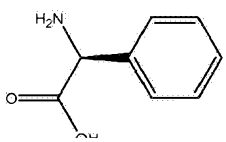
Name	Abbreviation	Structure
(S)-2,4-Diaminobutyric acid	Dab	
(S)-Diaminopropionic acid	Dap	
alpha,alpha-Diethylglycine	Deg	 Deg
5,5-Dimethyl-D-thiazolidine-4-carboxylic acid	dtc	
3,4-Dihydroxyphenylalanine	Dopa	 Dopa
(S)-2-Propargylglycine	Eag	
1-Amino-cyclopropane-1-carboxylic acid	Ebc	
1-Amino-cyclopentane-1-carboxylic acid	Eca	
Cys(3-propionic acid amide)	Ecl	
Sulfoxid of Carboxyethylcystein	Ede(O)	

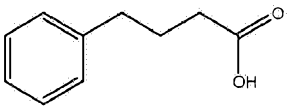
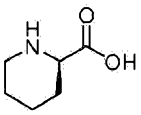
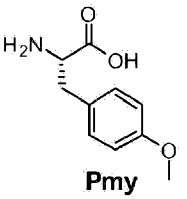
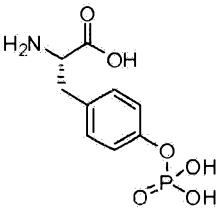
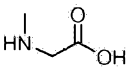
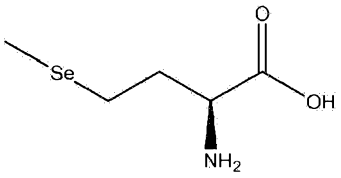
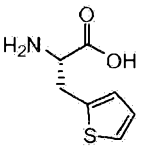
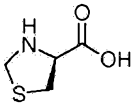
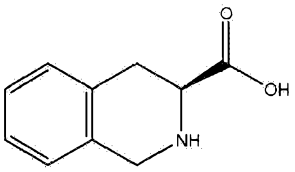
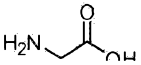
Name	Abbreviation	Structure
		
Cys(5-methylen-2-oxazolidinon)	Eea	
Cys(1-methylen-1H-benzotriazol)	Eec	
Cys(3-methylen-2-benzothiazolinon)	Eef	
(S)-N(omega)-nitro-arginine	Eew	
alpha,alpha-Dibutylglycine	Egt	
1-amino-cyclohexane-1-carboxylic acid	Egz	
L-homophenylalanine	Hfe	

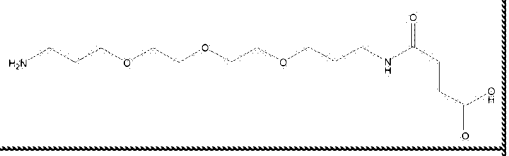
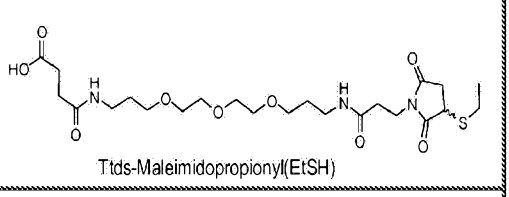
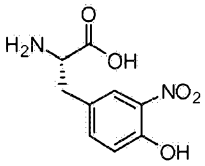
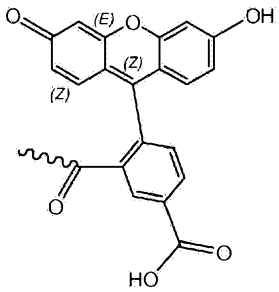
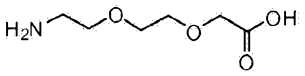
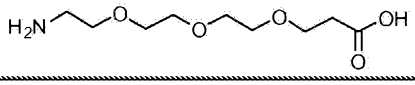
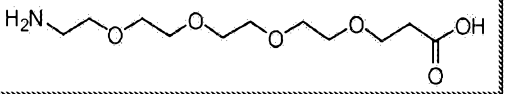
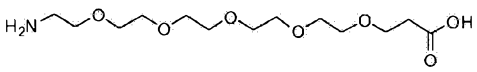
Name	Abbreviation	Structure
(S)-Homo-arginine	Har	
(S)-Homo-citrulline	Hci	
(S)-Homo-cysteine	Hcy	
D-Homo-cysteine	hcy	
(S)-2-Amino-5-methyl-hexanoic acid	Hle	
(S)-Homo-lysine	Hly	
ε-(Acetyl)-L-lysine	K(Ac)	
ε-(4-(p-Iodophenyl)butyryl))-L-lysine	K(AlbuTag)	
2-Amino-6-(2-aminooxy-acetylamino)-hexanoic acid	K(AOA)	
ε-(Myristyl)-L-lysine	K(Myr)	
ε-(Acetyl-Ttds)-L-lysine	K(TtdsAc)	
ε-(Myristyl-Ttds)-L-lysine	K(TtdsMyr)	

Name	Abbreviation	Structure
ε-(Myristyl-γ-glutamyl-Ttds)-L-lysine	K(Ttds-yGlu-Myr)	
ε-(Palmityl-Ttds)-L-lysine	K(TtdsPal)	
ε-(4-(Pentyl)-benzolsulfonamidyl)-L-lysine	K(4PBSA)	
4-Methoxyimino-pentanoic acid	Meox-Lev	
L-methionine-sulphone		
1-Naphthylalanine	1Ni	
2-Naphthylalanine	2Ni	
N-(cyclohexyl)-glycine	Ncg	
4-Nitrophenyl alanine	Nif	
(S)-Norleucine	Nle	

Name	Abbreviation	Structure
(S)-*N*-Methylalanine	Nma	
(S)-*N*-Methyl-Aspartic acid	Nmd	
(S)-*N*-Methyl-glutamic acid	Nme	
(S)-*N*-Methyl-phenylalanine	Nmf	
N-Methyl-glycine	Nmg	
(S)-*N*-Methyl-lysine	Nmk	
(S)-*N*-Methyl-leucine	Nml	
N-Methyl-asparagine	Nmn	
(S)-*N*-Methyl-arginine	Nmr	
(S)-*N*-Methyl-serine	Nms	
(S)-*N*-Methyl-valine	Nmv	

Name	Abbreviation	Structure
		
(S)-*N*-Methyl-tyrosine	Nmy	
N-propyl glycine	NpropylG	
(S)-2-Amino-pentanoic acid	Nva	
(S)-2-Pyridyl-alanine	Opa	
Ornithine-(pyrazin-carboxylate)	Opc	
D-Octahydroindol-2-carboxylic acid	oic	
(S)-Ornithine	Orn	
Ethylidene-aminooxy-acetic acid	Oxme-AOA	
Palmitoyl	Palm	
L-Phenylglycin	Phg	

Name	Abbreviation	Structure
4-Phenyl-butyric acid	PhPrCO	
Polyethylene glycol	PEG	
D-Pipecolic acid	pip	
L-Tyrosin(O-Methyl)-OH	Pmy	 **Pmy**
L-Phosphotyrosine	Pty	
N-Methylglycine	Sar	
Selenomethionine	Sem	
L-2-Thienylalanine	Thi	
D-thiazolidine-4-carboxylic acid	thz	
1,2,3,4-L-tetrahydroisoquinolinecarboxylic acid	Tic	
L-alpha-t-Butylglycine	Tle	

Name	Abbreviation	Structure
		Tle
(13-Amino-4,7,10-trioxa-tridecayl)-succinamic acid	Ttds	
Ttds-Maleimidopropionyl(EtSH))		 Ttds-Maleimidopropionyl(EtSH)
3-Nitro-L-tyrosine	Tym	
Carboxyfluorescein	FAM	
[2-(2-Amino-ethoxy)-ethoxy]-acetic acid	FA03202	
3-{2-[2-(2-Amino-ethoxy)-ethoxy]-ethoxy}-propionic acid	FA19203	
3-(2-{2-[2-(2-Amino-ethoxy)-ethoxy]-ethoxy}-ethoxy)-propionic acid	FA19204	
3-[2-(2-{2-[2-(2-Amino-ethoxy)-ethoxy]-ethoxy}-ethoxy)-ethoxy]-propionic acid	FA19205	

[0015] The amino acid sequences of the peptides provided herein are depicted in typical peptide sequence format, as would be understood by the ordinary skilled artisan. For example, the three-letter code or one-letter code of a conventional amino acid, or the three-, four-, or seven-number/letter code additional building blocks, indicates the presence of the amino acid or building block in a specified position within the peptide sequence. The code for each non-conventional amino acid or building block is connected to the code for the next and/or previous amino acid or building block in the sequence by a hyphen. Adjacent amino acids are

connected by a chemical bond (typically an amide bond). The formation of the chemical bond removes a hydroxyl group from the 1-carboxyl group of the amino acid when it is located to the left of the adjacent amino acid (e.g., Hle-adjacent amino acid), and removes a hydrogen from the amino group of the amino acid when it is located on the right of the adjacent amino acid (e.g., adjacent amino acid-Hle). It is understood that both modifications can apply to the same amino acid and apply to adjacent conventional amino acids present in amino acid sequences without hyphens explicitly illustrated. Where an amino acid contains more than one amino and/or carboxy group in the amino acid side chain, the 2- or 3-amino group and/or the 1-carboxy group generally are used for the formation of peptide bonds. For non-conventional amino acids, a 3-letter code was used where the first letter indicates the stereochemistry of the C-α-atom. For example, a capital first letter indicates that the L-form of the amino acid is present in the peptide sequence, while a lower case first letter indicating that the D-form of the correspondent amino acid is present in the peptide sequence. When one-letter code is used, a lower case letter represents a D-amino acid, while an upper case letter represents an L-amino acid. Unless indicated to the contrary, the amino acid sequences are presented herein in N- to C-terminus direction.

[0016] The C-termini of several TFPI-binding peptide sequences described herein are explicitly illustrated by inclusion of an OH, NH2, or an abbreviation for a specific terminating amine linked to the C-terminal amino acid code via a hyphen. The N-termini of several peptides described herein are explicitly illustrated by inclusion of a hydrogen (for a free N-terminus), or an abbreviation for a specific terminating carboxylic acid or other chemical group linked to the N-terminal amino acid code via a hyphen.

[0017] Optionally, the peptide complex of the invention comprises one or more amino acid substitutions (with reference to any of the amino acid sequences provided herein) that do not destroy the ability of the peptide to bind and/or inhibit TFPI.

[0018] Amino acid substitutions include, but are not limited to, those which: (1) reduce susceptibility to proteolysis, (2) reduce susceptibility to oxidation, (3) alter binding affinities, and/or (4) confer or modify other physiochemical or functional properties on a peptide. In one aspect, the substitution is a conservative substitution, wherein an amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined within the art, and include amino acids with basic side chains (e.g., lysine, arginine, and histidine), acidic side chains (e.g., aspartic acid and glutamic acid), uncharged polar side chains (e.g., glycine, asparagine, glutamine, serine, threonine, tyrosine, and cysteine), nonpolar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, and tryptophan), beta-branched side chains (e.g., threonine, valine, and isoleucine) and aromatic side chains (e.g., tyrosine, phenylalanine, tryptophan, and histidine). It will be appreciated, however, that a practitioner is not limited to creating conservative substitutions so long as the resulting peptide retains the ability to downregulate, in whole or in part, TFPI activity. The invention also embraces TFPI-binding peptides (e.g., TFPI-inhibitory peptides) comprising atypical, non-naturally occurring amino acids, which are well known in the art. Exemplary non-naturally occurring amino acids include ornithine, citrulline, hydroxyproline, homoserine, phenylglycine, taurine, iodotyrosine, 2,4-diaminobutyric acid, α-amino isobutyric acid, 4-aminobutyric acid, 2-amino butyric acid, y-amino butyric acid, 2-amino isobutyric acid, 3-amino propionic acid, norleucine, norvaline, sarcosine, homocitrulline, cysteic acid, t-butylglycine, t-butylalanine, phenylglycine, cyclohexylalanine, β-alanine, a fluoro-amino acid, a 3-methyl amino acid, α-C-methyl amino acid, a N-methyl amino acid, 2-amino-isobutyric acid, β-homoglutamatic acid, β-homophenylalanine, β-homolysine, β-homoleucine, β-homoasparagine, β-homoglutamine, β-homoarginine, β-homoserine, β-homotyrosine, β-homoaspartic acid, β-homovaline, β-homoasparagin, (S)-cyclohexylalanine, (S)-citrullin, (S)-2,4-diaminobutyric acid, (S)-2,4-diaminobutyric acid, (S)-diaminopropionic acid, (S)-2-propargylglycine, (S)-N(omega)-nitro-arginine, L-homophenylalanine, S)-homo-arginine, (S)-homo-citrulline, (S)-homo-cysteine, (S)-2-amino-5-methyl-hexanoic acid, (S)-homo-lysine, (S)-norleucine, (S)-*N*-methylalanine, (S)-*N*-methyl-aspartic acid, (S)-*N*-methyl-glutamic acid, (S)-*N*-methyl-phenylalanine, *N*-methyl-glycine, (S)-*N*-methyl-lysine, (S)-*N*-methyl-leucine, (S)-*N*-methyl-arginine, (S)-*N*-methyl-serine, (S)-*N*-

methyl-valine, (S)-*N*-methyl-tyrosine, (S)-2-amino-pentanoic acid, (S)-2-pyridyl-alanine, (S)-ornithine, L-phenylglycin, 4-phenyl-butyric acid and selenomethionine. The individual amino acids may have either L or D stereochemistry when appropriate, although the L stereochemistry is typically employed for all of the amino acids in the peptide.

[0019] The invention further includes TFPI-binding peptide (e.g., TFPI-inhibitory peptide) variants in a complex comprising one or more amino acids inserted within an amino acid sequence provided herein and/or attached to the N-terminus or C-terminus. In one aspect, the peptide further comprises one or more amino acids that facilitate synthesis, handling, or use of the peptide, including, but not limited to, one or two lysines at the N-terminus and/or C-terminus to increase solubility of the peptide. Suitable fusion proteins include, but are not limited to, proteins comprising a TFPI-binding peptide (e.g., TFPI-inhibitory peptide) linked to one or more polypeptides, polypeptide fragments, or amino acids not generally recognized to be part of the protein sequence. In one aspect, a fusion peptide comprises the entire amino acid sequences of two or more peptides or, alternatively, comprises portions (fragments) of two or more peptides. In addition to all or part of the TFPI-binding peptides (e.g., TFPI-inhibitory peptides) described herein, a fusion protein optionally includes all or part of any suitable peptide comprising a desired biological activity/function. Indeed, in some aspects, a TFPI-binding peptide (e.g., TFPI-inhibitory peptide) is operably linked to, for instance, one or more of the following: a peptide with long circulating half life, a marker protein, a peptide that facilitates purification of the TFPI-binding peptide (e.g., TFPI-inhibitory peptide), a peptide sequence that promotes formation of multimeric proteins, or a fragment of any of the foregoing. Suitable fusion partners include, but are not limited to, a His tag, a FLAG tag, a strep tag, and a myc tag. Optionally, the TFPI-binding peptide (e.g., TFPI-inhibitory peptide) is fused to one or more entities that enhance the half life of the peptide. Half life can be increased by, e.g., increasing the molecular weight of the TFPI-binding peptide to avoid renal clearance and/or incorporating a ligand for the nFc receptor-mediated recycling pathway. In one embodiment, the TFPI-binding peptide is fused to or chemically conjugated to (as described further below) an albumin polypeptide or a fragment thereof (e.g., human serum albumin (HSA) or bovine serum albumin (BSA)). The albumin fragment comprises 10%, 25%, 50%, or 75% of the full length albumin protein. Alternatively or in addition, the TFPI-binding peptide comprises an albumin binding domain or fatty acid that binds albumin when administered *in vivo.* An example of an albumin binding domain is "albu-tag," a moiety derived from on 4-(p-iodophenyl)-butanoic acid (Dumelin et al., Angew Chem Int Ed Engl 47:3196-3201 (2008)). Other suitable fusion partners include, but are not limited to, a proline-alanine-serine multimer (PASylation) and an antibody or fragment thereof (e.g., an Fc portion of an antibody).

[0020] Two or more TFPI-binding peptides are fused together, linked by a multimerization domain, or attached via chemical linkage to generate a TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex. The invention provides, a hetero-dimer (i.e., a dimer comprising two different TFPI-binding peptides), and hetero-multimer (i.e., a complex comprising three or more TFPI-binding peptides, wherein at least two of the TFPI-binding peptides are different), optionally attached by one or more linkers.

[0021] In this regard, the invention provides a peptide complex comprising a first peptide and a second peptide. In some The first peptide and the second peptide may bind different regions (epitopes) of human TFPI and, optionally, the peptide complex inhibits two or more TFPI functions, such as, but not limited to, TFPI binding to FXa and TFPI binding to FVIIa. The level of inhibition of at least one TFPI (e.g., TFPI-1) activity mediated by the peptide complex is greater than the level of inhibition achieved by the first peptide or the second peptide, alone or in combination, in various embodiments of the invention. The different regions (epitopes) on TFPI may be on the same TFPI polypeptide, or on different TFPI polypeptides. The functional characteristics and therapeutic and diagnostic applications of monomeric TFPI-binding peptides described herein also are applicable to the peptide complexes described herein. Similarly, descriptions of modifications to monomeric TFPI-binding peptides also relate to peptide complexes.

[0022] The invention provides a peptide complex comprising a first peptide comprising the structure of

formula (XIII): X6001-X6002-X6003-X6004-X6005-X6006-X6007-X6008-X6009-X6010-X6011-X6012-X6013-X6014-X6015-X6016-X6017-X6018-X6019-X6020 (XIII) (SEQ ID NO: 3153). In formula (XIII),

X6001 is an amino acid selected from the group consisting of F, L, M, Y, 1Ni, Thi, Bta, Dopa, Bhf, C, D, G, H, I, K, N, Nmf, Q, R, T, V, and W, such as an amino acid selected from the group consisting of 1Ni, Bta, Dopa, F, L, Y and M;

X6002 is an amino acid selected from the group consisting of Q, G, and K, such as Q;

X6003 is an amino acid selected from the group consisting of C, D, E, M, Q, R, S, T, Ede(O), Cmc, A, Aib, Bhs, F, G, H, I, K, L, N, P, V, W and Y, such as an amino acid selected from the group consisting of D, E, S, M, Q, R, T and C;

X6004 is an amino acid selected from the group consisting of Aib, E, G, I, K, L, M, P, R, W, Y, A, Bhk, C, D, F, H, k, N, Nmk, Q, S, T, and V, such as an amino acid selected from the group consisting of K, Aib, L, P, R, E, G, I, Y, M and W;

X6005 is an amino acid selected from the group consisting of a, A, Aib, C, D, d, E, G, H, K, k, M, N, Nmg, p, Q, R, NpropylG, aze, pip, tic, oic, hyp, nma, Ncg, Abg, Apg, thz, dtc, Bal, F, L, S, T, V, W and Y, such as an amino acid selected from the group consisting of p, Nmg, NpropylG, aze, pip, tic, oic, hyp, a, Aib, D, d, G, H, K, k, N, Q, R, A, E, C and M;

X6006 is an amino acid selected from the group consisting of A, C, C(NEM), D, E, G, H, K, M, N, Q, R, S, V, Cit, C(Acm), Nle, I, Ede(O), Cmc, Ecl, Eea, Eec, Eef, Nif, Eew, Aib, Btq, F, I, L, T, W and Y, such as an amino acid selected from the group consisting of C, E, K, R, S, V, C(Acm), Nle, C(NEM), I, Cit, A, D, G, H, N, Q and M;

X6007 is an amino acid selected from the group consisting of I, V, T, Chg, Phg, Tie, A, F, G, I, K, L, Nmv, P, Q, S, W and Y, such as Tie, V or I;

X6008 is an amino acid selected from the group consisting of F, H, 1Ni, 2Ni, Pmy, Y, and W, such as an amino acid selected from the group consisting of H, 1Ni, 2Ni, Pmy, F and Y;

X6009 is an amino acid selected from the group consisting of Aib, V, Chg, Phg, Abu, Cpg, Tie, L-2-amino-4,4,4-trifluorobutyric acid, A, f, I, K, S, T and V, such as V, Abu or Tle;

X6010 is an amino acid selected from the group consisting of A, C, D, d, E, F, H, K, M, N, P, Q, R, S, T, V, W, Y, Nmd, C(NEM), Aib, G, I, L and Nmf, such as an amino acid selected from the group consisting of D, P, C, T, A, E, K, M, N, Q, R, F, H, S, V, W and Y;

X6011 is an amino acid selected from the group consisting of A, a, G, p, Sar, c, hcy, Aib, C, K, G and Nmg, such as G, a, c, hcy or Sar;

X6012 is an amino acid selected from the group consisting of Y, Tym, Pty, Dopa, and Pmy, such as Y;

X6013 is an amino acid selected from the group consisting of Aib, C, F, 1Ni, Thi, Bta, A, E, G, H, K, L, M, Q, R, W and Y, such as an amino acid selected from the group consisting of F, 1Ni, Bta and C;

X6014 is an amino acid selected from the group consisting of A, Aib, C, C(NEM), D, E, K, L, M, N, Q, R, T, V, Hcy, Bhe, F, G, H, I, P, S, W and Y, such as an amino acid selected from the group consisting of Aib, C, E, Hcy, A, D, K, L, M, N, Q, R, T, V and Aib;

X6015 is an amino acid selected from the group consisting of R, (omega-methyl)-R, D, E and K, such as R;

X6016 is an amino acid selected from the group consisting of L, Hcy, Hle, and Aml, such as an amino acid selected from the group consisting of L, Aml, Hle and Hcy;

X6017 is an amino acid selected from the group consisting of A, a, Aib, C, c, Cha, Dab, Eag, Eew, H, Har, Hci, Hle, I, K, L, M, Nle, Nva, Opa, Orn, R, S, Deg, Ebc, Eca, Egz, Aic, Apc, Egt, (omega-methyl)-R, Bhr, Cit, D, Dap, E, F, G, N, Q, T, V, W and Y, such as an amino acid selected from the group consisting of A, Aib, C, c, Aic, Eca, Deg, Cha, Dab, Dap, Eag, Eew, H, Har, Hci, Hle, K, Nle, Nva, Opa, Orn, R, I, L, S and M;

X6018 is an amino acid selected from the group consisting of A, Aib, Hcy, hcy, C, c, L, Nle, M, N, R, Bal, D, E, F, G, H, I, K, Q, S, T, V, W and Y, such as an amino acid selected from the group consisting of A, Aib, C, c, L, Hcy, N, M and R;

X6019 is an amino acid selected from the group consisting of K, R, Har, Bhk and V, such as K; and

X6020 is an amino acid selected from the group consisting of K, L, Hcy, Aml, Aib, Bhl, C, F, G, H, I, Nml, Q, R, S, T, V, W and Y, such as an amino acid selected from the group consisting of L, Aml, Hcy and K.

[0023] The first peptide, in various embodiments disclosed herein, comprises (or consists of) an amino acid sequence selected from the group consisting of SEQ ID NOs: 8-978, 4022, 4024, 4032, 4036-4047, 4049-4078, 4086-4097, 4100-4127, 4129-4170, 4173-4195, 4200-4214, 4217-4225, 4228, 4230, 4231, 4238, 4239, 4002, 4013, 4021, 4023, 4025-4031, 4033-4035, 4048, 4079-4085, 4098, 4099, 4128, 4171, 4172, 4196-4199, 4215, 4216, 4226, 4277, 4229, 4232, and 4233.

[0024] The second peptide, in one aspect, comprises the structure of formula (XIV): X7001-X7002-X7003-X7004-X7005-X7006-[X7007-X7008-X7009-X7010-X7011-X7012-X7013-X7014-X7015-X7016-X7017-X7018]-X7019-X7020-X7021-X7022-X7023 (XIV) (SEQ ID NO: 3154). In formula (XIV),

X7001 is either present or absent, whereby in case X7001 is present it is an amino acid selected from the group consisting of A, C, C(NEM), D, E, F, G, H, I, K, L, P, R, S, T, V and W, such as an amino acid selected from the group consisting of A, D, F, G, H, K, L and S;

X7002 is either present or absent, whereby in case X7002 is present it is an amino acid selected from the group consisting of A, C, C(NEM), D, E, F, G, H, I, K, L, M, P, Q, R, S, T, V, W, and Y, such as an amino acid selected from the group consisting of H, F, M and R;

X7003 is an amino acid selected from the group consisting of A, F, I, K, L, R, S, T, V, W and Y, such as F or Y;

X7004 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, R, S, T, V and W, such as K;

X7005 is R or W, such as W;

X7006 is an amino acid selected from the group consisting of F, H, I, K, L, R, V and W, such as F or H;

X7007 is an amino acid selected from the group consisting of Orn, homoK, C, Hcy, Dap and K, such as C or Hcy;

X7008 is an amino acid selected from the group consisting of A, G, R, S and T, such as A, G or S;

X7009 is an amino acid selected from the group consisting of a, A, I, K, L, M, m, Moo, Nle, p, R, Sem and V, such as M, Sem, or V;

X7010 is an amino acid selected from the group consisting of A, G, I, K, L, P, R, S, T and V, such as K, P or R;

X7011 is an amino acid selected from the group consisting of D, E, G, S and T, such as D;

X7012 is an amino acid selected from the group consisting of A, a, D, d, E, e, F, f, G, I, K, k, L, 1, M, m, Moo, Nle, nle, P, p, R, r, S, s, Sem, T, t, V, v, W and w, such as an amino acid selected from the group consisting of F, L, I, M and Sem;

X7013 is an amino acid selected from the group consisting of A, C, C(NEM), Con, Con(Meox), D, d, E, e, Eag, F, G, I, K, L, N, R, S, s, T, V and W, such as an amino acid selected from the group consisting of D, G, K and S;

X7014 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, M, R, S, T, V and W, such as G;

X7015 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, M, Nle, R, S, T, V and W, such as I or T;

X7016 is an amino acid selected from the group consisting of A, D, E, F, I, K, L, M, Moo, Nle, R, S, Sem, T, V, W and Y, such as an amino acid selected from the group consisting of D, F, M, Sem and Y;

X7017 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, R, S, T, V, W and Y, such as S or T;

X7018 is an amino acid selected from the group consisting of C and D, such as C;

X7019 is an amino acid selected from the group consisting of A, F, I, L, S, T, V and W, such as A or V;

X7020 is an amino acid selected from the group consisting of F and W, such as W;

X7021 is an amino acid selected from the group consisting of I, L and V, such as V;

X7022 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, P, R, S, T, V and W, such as the group consisting of F, L, K, R, P and W;

X7023 is either present or absent, whereby in case X7023 is present it is an amino acid selected from the group consisting of A, C, C(NEM), Con, Con(Meox), D, E, Eag, F, G, I, K, L, R, S, T, V, W and Y, such as an amino acid selected from the group consisting of A, D, F, M, S and Y; and the peptide comprises as a cyclic structure generated by a linkage between X7007 and X7018;

wherein the C-terminus of the first peptide is linked to the N-terminus of the second peptide via a linker moiety;

wherein the first peptide comprises an amino acid sequence at least 80%, at least 85%, at least 90%, at least 95%, or 100% identical to SEQ ID NO: 178 or SEQ ID NO: 4261; and

wherein the second peptide comprises an amino acid sequence at least 80%, at least 85%, at least 90%, at least 95%, or 100% identical to SEQ ID NO: 1044.

[0025] The second peptide, in some embodiments disclosed herein, comprises (or consists of) an amino acid sequence selected from the group consisting of SEQ ID NOs: 1001-1336.

[0026] Examples of TFPI-binding peptides that bind different TFPI epitopes include JBT1857, an example of the JBT0047 class of peptides (represented as, e.g., formulas (I)-(IV) and (XI) and examples of which are set forth in Figures 32, 62, and 65), and JBT1837, an example of the JBT0120 class of peptides (represented as, e.g., formulas (V)-(VII) and examples of which are set forth in Figure 34). Thus, the invention provides a peptide complex comprising a peptide subunit (e.g., a first peptide) comprising an amino acid sequence at

least 80%, at least 85%, at least 90%, at least 95%, or 100% identical to SEQ ID NO: 178 (JBT1857) or SEQ ID NO: 4261 (JBT2548). Additionally, the invention provides a peptide complex comprising a peptide subunit (e.g., a second peptide) comprising an amino acid sequence at least 80%, at least 85%, at least 90%, at least 95%, or 100% identical to SEQ ID NO: 1044 (JBT1837). Exemplary TFPI-binding peptide dimers include JBT2496 (SEQ ID NO: 4211), JBT2547 (SEQ ID NO: 4260), JBT2521 (SEQ ID NO: 4242), JBT2522 (SEQ ID NO: 4243), JBT2523 (SEQ ID NO: 4244), and JBT2533 (SEQ ID NO: 4250).

[0027] The first peptide and the second peptide (and optionally additional peptides) are linked by a linker moiety. Any linker moiety is suitable for use in the context of the peptide complex. The linker moiety, in some aspects of the invention, bridges a distance of about 1 Å to about 100 Å, e.g., about 5 Å to about 80 Å (about 5 Å to about 50Å), about 10 Å to about 70 Å (about 10 Å to about 60 Å, about 10 Å to about 50 Å, about 10 Å to about 40 Å, or about 10 Å to about 30 Å), in one of its conformations. Thus, the linker is optionally about 1 Å to about 100 Å in length, e.g., about 5 Å to about 50Å or about 10 Å to about 30 Å in length in one of its conformations. Linkers of greater length (greater than about 100 Å) also are contemplated. For example, biocompatible polymers, optionally having a molecular weight of about 2 kDa to about 60 kDa, also are contemplated for use in the peptide complex. Examples of biocompatible polymers include, but are not limited to, PEG, PSA, proline-alanine-serine multimer, and hydroxyethyl starch.

[0028] In one non-limiting example, the linker moiety is a molecule with at least two reactive groups (before conjugation to the first and second peptides) capable of reacting with each of the first peptide and the second peptide. In some embodiments, the linker moiety has only two reactive groups and is bifunctional. The reactive groups are both nucleophilic, both electrophilic, or a combination of nucleophilic and electrophilic reactive groups. Nonlimiting combinations of reactive groups are shown in Figure 73. The linker moiety may contain structural elements resulting from a chemical ligation, such as, for instance, cystein, oxime, hydrazide, succinimide, thioether, triazole, secondary amine, amide, disulfide. In various embodiments, the linker moiety is attached to the first peptide and/or the second peptide via an oxime, a hydrazide, a succinimide, a thioether, a triazole, a secondary amine, an amide, or a disulfide. Additional description of linker moieties and reactive groups is provided in International Patent Publication No. WO 2011/143209.

[0029] Hydrophobic linkers also are suitable for use in the context of the invention. Hydrophobic linkers are known in the art. See, e.g., Bioconjugate Techniques, G. T. Hermanson (Academic Press, San Diego, CA, 1996). Suitable hydrophobic linker moieties include, for example, 8-hydroxyoctanoic acid and 8-mercaptooctanoic acid. Hydrophilic linker moieties such as, for example, polyalkylene glycol, also are suitable for use in the context of the invention. In some embodiments, the linker moiety comprises a chain of atoms from about 1 to about 60 atoms long. In some embodiments, the chain atoms are all carbon atoms. In some embodiments, the chain atoms in the backbone of the linker are selected from the group consisting of C, O, N, and S. Chain atoms and linkers may be selected according to their expected solubility (hydrophilicity) so as to provide a more soluble peptide complex.

[0030] In one aspect, the linker moiety comprises the structure $Z_{1\text{-}20}$, wherein Z is an oligomer building block. Examples of oligomer building blocks include, but are not limited to, an amino acid, hydroxy acid, ethylene glycol, propylene glycol, or a combination of any of the foregoing. For example, the linker moiety is optionally an amino acid, a dipeptide, a tripeptide, or a polypeptide comprising 4-20 amino acids. In some embodiments, Z is G, s, S, a, A, Bal, Gaba, Ahx, Ttds, or a combination of any of the foregoing (such as peptide ten-mer comprising A, S, or a combination of A and S). If desired, the linking moiety comprises an amine, ether, thioether, maleimide, disulfide, amide, ester, alkene, cycloalkene, alkyne, trizoyl, carbamate, carbonate, cathepsin B-cleavable, or hydrazone.

[0031] The terms "first peptide" and "second peptide" are not meant to imply a particular physical order of the peptides, but merely to distinguish different subunits of the peptide complex. The subunits of the peptide complex may be linked in any of a number of configurations so long as the first peptide and second peptide

interact with TFPI. For example, the C-terminus of the first peptide is connected to the N-terminus of the second peptide, the N-terminus of the first peptide is connected to the C-terminus of the second peptide, the N- or C-terminus of the first (or second) peptide is connected to an internal attachment point in the second (or first) peptide, or the first and second peptides are connected via internal attachment points (i.e., attachment points located within the amino acid sequence of the peptide and not at the N- or C- terminus). Exemplary attachment points for a linking moiety in the first peptide of formula (XIII) is the N-terminal amino group, the C-terminal carboxylic acid, X6004, X6006, X60010, or X60014 via an appropriate functional group in the amino acid side chain, such as, but not limited to, amine, carboxylic acid, thiol, alkene, alkyne, azide, carbonyl, aminooxy, hydrazine and halogens. More than one linker may be used, e.g., a first linking moiety is attached at the N-terminus of the first peptide and the C-terminus of the second peptide, and a second linking moiety (which may be the same type of moiety or a different type of moiety) is attached at the C-terminus of the first peptide and attached at the N-terminus of the second peptide. While the discussion of possible configurations refers to the first and second peptides, it will be appreciated that additional peptides may be linked to the first and/or second peptides as described herein.

[0032] "Derivatives" are included in the invention and include TFPI-binding peptide complexes that have been chemically modified in some manner distinct from addition, deletion, or substitution of amino acids. In this regard, a peptide of the invention provided herein is chemically bonded with polymers, lipids, other organic moieties, and/or inorganic moieties. Examples of peptide and protein modifications are given in Hermanson, Bioconjugate Techniques, Academic Press, (1996). The TFPI-binding peptides described herein optionally comprise a functional group that facilitates conjugation to another moiety (e.g., a peptide moiety). Exemplary functional groups include, but are not limited to, isothiocyanate, isocyanate, acyl azide, NHS ester, sulfonyl chloride, aldehyde, epoxide, oxirane, carbonate, arylating agent, imidoester, carbodiimide, anhydride, alkyl halide derivatives (e.g., haloacetyl derivatives), maleimide, aziridine, acryloyl derivatives, arylating agents, thiol-disulfide exchange reagents (e.g., pyridyl disulfides or TNB thiol), diazoalkane, carboyldiimadazole, N,N'-Disuccinyl carbonate, N-Hydroxysuccinimidyl chloroformate, and hydrazine derivatives. Maleimide is useful, for example, for generating a TFPI-binding peptide complex that binds with albumin *in vivo.*

[0033] Derivatives are prepared in some situations to increase solubility, stability, absorption, or circulating half life. Various chemical modifications eliminate or attenuate any undesirable side effect of the agent. In one aspect, the invention includes TFPI-binding peptides covalently modified to include one or more water soluble polymer attachments. A water soluble polymer (or other chemical moiety) is attached to any amino acid residue, although attachment to the N- or C-terminus is preferred in some embodiments. Optionally, a polymer is attached to the peptide via one or more amino acids or building blocks that offer functional groups that facilitate polymer attachment. For example, JBT2315 comprises a C-terminal cysteine (position X4021 with respect to formula (XI)), which facilitates the addition of, e.g., a maleimide polyethylene glycol (PEG). Useful polymers include, but are not limited to, PEG (e.g., PEG approximately 40 kD, 30 kD, 20 kD, 10, kD, 5 kD, or 1 kD in size), polyoxyethylene glycol, polypropylene glycol, monomethoxy-polyethylene glycol, dextran, hydroxyethyl starch, cellulose, poly-(N-vinyl pyrrolidone)-polyethylene glycol, propylene glycol homopolymers, a polypropylene oxide/ethylene oxide co-polymer, polysialic acid (PSA), polyoxyethylated polyols (e.g., glycerol) and polyvinyl alcohol, as well as mixtures of any of the foregoing. In one aspect, the peptide of the invention is a PEGylated peptide. PEG moieties are available in different shapes, e.g., linear or branched. For further discussion of water soluble polymer attachments, see U.S. Patent Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192; and 4,179,337. Other moieties useful for improving peptide half life or stability are described herein and include, for instance, albumin (optionally modified to allow conjugation to the inventive peptide), fatty acid chains (e.g., C12-C18 fatty acid, such as a C14 fatty acid), an antibody or fragment thereof (e.g., an Fc portion of an antibody), and proline-alanine-serine multimers.

[0034] In another aspect, a peptide derivative includes a targeting moiety specific for a particular cell type,

tissue, and/or organ. Alternatively, the peptide is linked to one or more chemical moieties that facilitate purification, detection, multimerization, binding with an interaction partner, and characterization of peptide activity. An exemplary chemical moiety is biotin. Other moieties suitable for conjugation to the TFPI-binding peptide complex of the invention include, but are not limited to, a photosensitizer, a dye, a fluorescence dye, a radionuclide, a radionuclide-containing complex, an enzyme, a toxin, and a cytotoxic agent. Photosensitizers include, e.g., Photofrin, Visudyne, Levulan, Foscan, Metvix, Hexvix®, Cysview™, Laserphyrin, Antrin, Photochlor, Photosens, Photrex, Lumacan, Cevira, Visonac, BF-200 ALA, and Amphinex. If desired, a His tag, a FLAG tag, a strep tag, or a myc tag is conjugated to the peptide.

[0035] In addition, in one aspect, the peptide complexes of the invention are acylated at the N-terminal amino acid of the peptide. In another aspect, the peptide complexes of the invention are amidated at the C-terminal amino acid of the peptide. In a still further aspect, the peptide complexes of the invention are acylated at the N-terminal amino acid of the peptide and are amidated at the C-terminal amino acid of the peptide.

[0036] Derivatives also include peptides comprising modified or non-proteinogenic amino acids or a modified linker group (see, e.g., Grant, Synthetic Peptides: A User's Guide, Oxford University Press (1992)). Modified amino acids include, for example, amino acids wherein the amino and/or carboxyl group is replaced by another group. Non-limiting examples include modified amino acids incorporating thioamides, ureas, thioureas, acylhydrazides, esters, olefines, sulfonamides, phosphoric acid amides, ketones, alcohols, boronic acid amides, benzodiazepines and other aromatic or non-aromatic heterocycles (see Estiarte et al., Burgers Medicinal Chemistry, 6th edition, Volume 1, Part 4, John Wiley & Sons, New York (2002)). Modified amino acids are often connected to the peptide with at least one of the above mentioned functional groups instead of an amide bond. Non-proteinogenic amino acids include, but are not limited, to β-alanine (Bal), norvaline (Nva), norleucine (Nle), 4-aminobutyric acid (γ-Abu), 2-aminoisobutyric acid (Aib), 6-aminohexanoic acid (ε-Ahx), ornithine (Orn), hydroxyproline (Hyp), taurine, sarcosine, citrulline (Cit), cysteic acid (Coh), cyclohexylalanine (Cha), methioninesulfoxide (Meo), methioninesulfone (Moo), homoserinemethylester (Hsm), propargylglycine (Eag), 5-fluorotryptophan (5Fw), 6-fluorotryptophan (6Fw), 3',4'-dimethoxyphenyl-alanine (Ear), 3',4'-difluorophenylalanine (Dff), 4'-fluorophenyl-alanine (Pff), 1-naphthyl-alanine (1Ni), 1-methyltryptophan (1Mw), penicillamine (Pen), homoserine (Hse), t-butylglycine, t-butylalanine, phenylglycine (Phg), benzothienylalanine (Bta), L-homo-cysteine (Hcy), N-methyl-phenylalanine (Nmf), 2-thienylalanine (Thi), 3,3-diphenylalanine (Ebw), homophenylalanine (Hfe) and S-benzyl-L-cysteine (Ece). The structures of many of the non-proteinogenic amino acids are provided in Table 2. These and other non-proteinogenic amino acids may exist as D- or L- isomers. Examples of modified linkers include, but are not limited to, the flexible linker 4,7,10-trioxa-1,13-tridecanediamine (Ttds), glycine, 6-aminohexanoic acid, beta-alanine (Bal), pentynoic acid (Pyn), and combinations of Ttds, glycine, 6-aminohexanoic acid and Bal.

[0037] Homologs of the amino acids constituting the peptides of the invention may be as set forth in Table 3. In any embodiment, one or more amino acids of the TFPI-binding peptide complex are substituted with a homolog.

TABLE 3

Amino Acid	Exemplary homologs
A	Aib, Bal, Eag, Nma, Abu, G, M, Nva, Nle
C	S, A, Hcy, M, L, I, V, Nmc, β-Cysteine
D	E, Homoglutamic acid, γ-Hydroxy-glutamic acid, γ-Carboxy-glutamic acid, Nmd, β-Aspartic acid, N, Q, Cysteic acid
E	D, Glu, Homoglutamic acid, γ-Hydroxy-glutamic acid, γ-Carboxy-glutamic acid, α-Aminoadipic acid, Nme, β-glutamic acid, Q, N, Cysteic acid
F	Hfe, Nmf, β-Phenylalanine, Phg, Bhf, Thienylalanine, Benzothienylalanine,

Amino Acid	Exemplary homologs
	Bromophenylalanine, Iodophenylalanione, Chlorophenylalanine, Methylphenylalanine, Nitrophenylalanine, Y, W, Naphtylalanine, Tic
G	A, Nmg
H	Nmh, 1-Methylhistidine, 3-Methylhistidine, Thienylalanine
I	L, V, Hle, Nva, Nle, β-Isoleucine, Nml, M, Nmi
K	Nmk, R, Nmr, β-Lysine, Dab, Dap, β-(1-Piperazinyl)-alanine, 2,6-Diamino-4-hexynoic acid, delta-Hydroxy-lysine, Har, omega-Hydroxy-norarginine, omega-Amino-arginine, omega-Methyl-arginine, β-(2-Pyridyl)-alanine, β-(3-Pyridyl)-alanine, 3-Amino-tyrosine, 4-Amino-phenylalanine, Hci, Cit
L	I, V, Hle, Nle, Nva, β-Isoleucine, Nml, M
M	I, V, Hle, Nva, R, Har, Nmm, Methioninesulfone
N	Nmn, β-Asparagine, Q, Nmq, β-Glutamine, Cys(3-propionic acid amide)-OH, Cys(O2-3-propionic acid amide)-OH
P	Azetidine-2-carboxylic acid, Hyp, α-Methyl-methionine, 4-Hydroxy-piperidine-2-carboxylic acid, Pip, α-Methyl-Pro
Q	N, Nmn, Nmq, β-Glutamine, Cys(3-propionic acid amide)-OH, Cys(O2-3-propionic acid amide)-OH
R	Nmk, K, Nmr, β-Lysine, Dab, Dap, Orn, β-(1-Piperazinyl)-alanine, 2,6-Diamino-4-hexynoic acid, delta-Hydroxy-lysine, Har, omega-Hydroxy-norarginine, omega-Amino-arginine, omega-Methyl-arginine, β-(2-Pyridyl)-alanine, β-(3-Pyridyl)-alanine, 3-Amino-tyrosine, 4-Amino-phenylalanine, Hci, Cit, Hle, L, Nle, M
S	T, Hse, β-Serine, C, β-Cyano-alanine, allo-Threonine
T	S, Homothreonine, β-Threonine, allo-Threonine
V	L, I, Hle, Nva, Nle, β-Valine, Nmv, M, Nmi, Nml
W	Nmw, β-Tryptophan, F, Hfe, Nmf, β-Phenylalanine, Phg, Bhf, Thienylalanine, Benzothienylalanine, Bromophenylalanine, Iodophenylalanine, Chlorophenylalanine, Methylphenylalanine, Nitrophenylalanine, Y, Naphtylalanine, Tic
Y	Nmy, β-Tyrosine,, F, Hfe, Nmf, β-Phenylalanine, Phg, Bhf, Thienylalanine, Benzothienylalanine, Bromophenylalanine, Iodophenylalanine, Chlorophenylalanine, Methylphenylalanine, Nitrophenylalanine, W, Naphtylalanine, Tic

[0038] Derivatives also include peptides comprising amino acids having modified substituents, such as amino acids modified by halogenation with, e.g., fluorine, chlorine, iodine, or bromine. In some embodiments, the TFPI-binding peptide complex comprises a halogenated aromatic amino acid, such as phenylalanine.

[0039] In some embodiments, the peptide (CO-NH) linkages joining amino acids within the peptide of the invention are reversed to create a "retro-modified" peptide, i.e., a peptide comprising amino acid residues assembled in the opposite direction (NH-CO bonds) compared to the reference peptide. The retro-modified peptide comprises the same amino acid chirality as the reference peptide. An "inverso-modified" peptide is a peptide of the invention comprising amino acid residues assembled in the same direction as a reference peptide, but the chirality of the amino acids is inverted. Thus, where the reference peptide comprises L-amino acids, the "inverso-modified" peptide comprises D-amino acids, and vice versa. Inverso-modified peptides comprise CO-NH peptide bonds. A "retro-inverso modified" peptide refers to a peptide comprising amino acid residues assembled in the opposite direction and which have inverted chirality. A retro-inverso analogue has reversed termini and reversed direction of peptide bonds (i.e., NH-CO), while approximately maintaining the side chain topology found in the reference peptide. Retro-inverso peptidomimetics are made

using standard methods, including the methods described in Meziere et al, J. Immunol., 159, 3230-3237 (1997). Partial retro-inverso peptides are peptides in which only part of the amino acid sequence is reversed and replaced with enantiomeric amino acid residues.

[0040] TFPI-binding peptides of the invention (including TFPI inhibitor peptides) are made in a variety of ways. In one aspect, the peptides are synthesized by solid phase synthesis techniques including those described in Merrifield, J. Am. Chem. Soc., 85, 2149 (1963); Davis et al., Biochem. Intl., 10, 394-414 (1985); Larsen et al., J. Am. Chem. Soc., 115, 6247 (1993); Smith et al., J. Peptide Protein Res., 44, 183 (1994); O'Donnell et al., J. Am. Chem. Soc., 118, 6070 (1996); Stewart and Young, Solid Phase Peptide Synthesis, Freeman (1969); Finn et al., The Proteins, 3rd ed., vol. 2, pp. 105-253 (1976); and Erickson et al., The Proteins, 3rd ed., vol. 2, pp. 257-527 (1976). Alternatively, the TFPI-binding peptide (e.g., the TFPI-inhibitory peptide) is expressed recombinantly by introducing a nucleic acid encoding a TFPI-binding peptide (e.g., a TFPI-inhibitory peptide) into host cells, which are cultured to express the peptide. Such peptides are purified from the cell culture using standard protein purification techniques.

[0041] Any of the peptide complexes of the invention or nucleic acids encoding the peptides also is provided in a composition (e.g., a pharmaceutical composition). In this regard, the peptide complex is formulated with a physiologically-acceptable (i.e., pharmacologically-acceptable) carrier, buffer, excipient, or diluent, as described further herein. Optionally, the peptide is in the form of a physiologically acceptable salt, which is encompassed by the invention. "Physiologically acceptable salts" means any salts that are pharmaceutically acceptable. Some examples of appropriate salts include acetate, hydrochloride, hydrobromide, sulfate, citrate, tartrate, glycolate, and oxalate. If desired, the composition comprises one or more additional pharmaceutically-effective agents.

[0042] The peptide complex provided herein optionally inhibits at least one TFPI-1 (e.g., TFPI-1α or TFPI-1β) activity such as, but not limited to, an activity that downregulates the blood coagulation cascade. Without being bound by any specific mechanism of action, a proposed mechanism of inhibition may involve preventing formation of the quaternary TF-FVIIA-FXA-TFPI complex. The peptide may inhibit binding (competitively or allosterically) of TFPI to FXa (e.g., inhibit binding of TFPI Kunitz domain 2 to Factor Xa or interrupt binding of TFPI Kunitz domain 1 to an exosite of Factor Xa), the TF/FVIIa complex (e.g., inhibit binding of TFPI Kunitz domain 1 to the TF/FVIIa complex), TF alone, and/or FVIIa alone. With TFPI activity diminished, TF and FVIIa are free to activate FX which, in turn, enhances conversion of prothrombin to thrombin. Surprisingly, in one embodiment, the peptide of the invention that binds Kunitz domain 1 interferes with TFPI-mediated inhibition of FXa. Thus, the invention provides a method of, e.g., inhibiting TFPI-mediated downregulation of the extrinsic and/or common pathway of the coagulation cascade and/or enhancing FXa-mediated conversion of prothrombin to thrombin, by administering to a subject a peptide complex described herein that binds Kunitz domain 1.

[0043] In one aspect, the peptide complex of the invention exhibits TFPI antagonistic activity in model and/or plasmatic systems. An exemplary model system for determining TFPI-inhibitory activity is the extrinsic tenase assay, which tests the ability of candidate peptides to restore extrinsic complex-mediated FX activation in the presence of TFPI (which is a natural inhibitor of the FX activation reaction) (see, e.g., Lindhout et al., Thromb. Haemost., 74, 910-915 (1995)). Another model system for characterizing TFPI-inhibitory activity is the FXa inhibition assay, wherein FXa activity is measured in the presence of TFPI (see Sprecher et al., PNAS, 91, 3353-3357 (1994)). The extrinsic tenase assay and the FXa inhibition assay are further described in Example 3. Optionally, the peptide complex of the invention enhances FX activation in the presence of TFPI with a half maximal effective concentration (EC_{50}) of less than or equal to 1×10^{-4} M, less than or equal to 1×10^{-5} M, less than or equal to 1×10^{-6} M, or less than or equal to 1×10^{-7} M.

[0044] In one aspect, TFPI-antagonist activity is characterized in a plasma-based assay. Thrombin formation

is triggered in plasma substantially lacking FVIII or FIX activity (e.g., the residual coagulation factor activity is lower than 1%) in the presence of a candidate peptide. Thrombin formation can be detected using a fluorogenic or chromogenic substrate, as described in Example 4. A system for measuring thrombin activity is provided by Thrombinoscope BV (Maastricht, The Netherlands). Prothrombin conversion is measured using, e.g., a Thrombograph™ (Thermo Scientific, Waltham, MA), and the resulting data is compiled into a Calibrated Automatic Thrombogram generated by Thrombinoscope™ software available from Thrombinoscope BV. In certain embodiments, the TFPI-inhibitory peptide complex increases the amount of peak thrombin generated during the assay and/or decreases the time required to achieve peak thrombin formation. For example, the peptide complex improves TFPI-regulated thrombin generation in the absence of FVIII (e.g., in FVIII-depleted plasma) to at least 1% of the level of TFPI-dependent thrombin generation in normal plasma. Generally, normal (unafflicted) plasma contains about 0.5 U/mL to about 2 U/mL Factor VIII. Accordingly, in some instances, a TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex will enhance thrombin formation in the absence of FVIII to at least about 1% of that observed in the presence of 0.5 U/mL to 2 U/mL FVIII. In further embodiments, the peptide enhances thrombin formation in the absence of Factor VIII to at least about 2%, at least about 3%, at least about 5%, at least about 7%, or at least about 10% of the level of thrombin formation in normal plasma, i.e., in the presence of physiological levels of Factor VIII. In various aspects, the peptide is administered to an animal model of thrombin deficiency or hemophilia to characterize TFPI inhibitory activity *in vivo.* Such *in vivo* models are known in the art and include for example, mice administered anti-FVIII antibodies to induce hemophilia A (Tranholm et al., Blood, 102, 3615-3620 (2003)); coagulation factor knock-out models such as, but not limited to, FVIII knock-out mice (Bi et al., Nat. Genet., 10(1), 119-121 (1995)) and FIX knock-out mice (Wang et al., PNAS, 94(21), 11563-66 (1997)); induced hemophilia-A in rabbits (Shen et al., Blood, 42(4), 509-521 (1973)); and Chapel Hill HA dogs (Lozier et al., PNAS, 99, 12991-12996 (2002)).

[0045] Various peptides bind TFPI from any source including, but not limited to, mouse, rat, rabbit, dog, cat, cow, horse, pig, guinea pig, and primate. In one embodiment, the peptide complex binds human TFPI. Optionally, the TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex binds TFPI from more than one species (i.e., the peptide is cross-reactive among multiple species). In certain aspects, the peptide binds TFPI with a dissociation constant (K_D) of less than or equal to 1 x 10^{-4} M, less than or equal to 1 x 10^{-5} M, less than or equal to 1 x 10^{-6} M, or less than or equal to 1 x 10^{-7} M. Affinity may be determined using, for example and without limitation, any one, two, or more of a variety of techniques, such as affinity ELISA assay, a competitive ELISA assay, and/or surface plasmon resonance (BIAcore™) assay. When characterized using a competitive (IC_{50}) ELISA assay, the peptide of the invention optionally demonstrates an IC_{50} of less than or equal to about 50,000 nM. For example, the peptide demonstrates an IC_{50} of less than or equal to about 10,000 nM, such as an IC_{50} of less than or equal to about 5,000 nM, less than or equal to about 1,000 nM, or less than or equal to about 500 nM. In one aspect, the peptide demonstrates an IC_{50} of less than or equal to about 250 nM, less than or equal to about 100 nM, less than or equal to about 50 nM, or less than or equal to about 10 nM. Exemplary peptides and their IC_{50} values are provided in Figures 32-39; in some instances, the peptides are classified into Groups A, B, C, D, E, F, and G (see Table 4 in Example 1) based on their IC_{50} values. In various aspects, the invention provides peptides falling within Groups A, B, C, D, E, F, and/or G as defined in Table 4. Affinity may also be determined by a kinetic method or an equilibrium/solution method. Such methods are described in further detail herein or known in the art.

[0046] Another suitable assay for characterizing the inventive peptides is a k_{off} assay, which examines a peptide's release from TFPI. The k_{off} assay result is not the dissociation rate constant, but a percentage of competitor peptide blocked from TFPI binding by a test peptide after an incubation period with TFPI. An exemplary k_{off} assay includes the following steps: 1) incubation of a TFPI-coated microtiter plate with an amount of test peptide resulting in approximately 90% TFPI occupation; 2) removal of unbound test peptide; 3) addition of a biotinylated tracer (i.e., competitor) peptide that competes with the test peptide for binding to

TFPI; 4) incubation for a period of time during which binding sites released by the test peptide is occupied by the tracer; 5) removal of unbound tracer and test peptide;, and 6) detection of bound tracer by a chromogenic reaction using streptavidin-horseradish peroxidase conjugate. The resulting signal is indicative of binding sites freed by the test peptide. A test peptide that does not dissociate from TFPI during the incubation period yields a weaker signal compared to an analyte that dissociates completely.

[0047] As with all binding agents and binding assays, one of skill in the art recognizes that the various moieties to which a binding agent should not detectably bind in order to be biologically (e.g., therapeutically) effective would be exhaustive and impractical to list. Therefore, the term "specifically binds" refers to the ability of a peptide to bind TFPI with greater affinity than it binds to an unrelated control protein that is not TFPI. For example, the peptide may bind to TFPI with an affinity that is at least, 5, 10, 15, 25, 50, 100, 250, 500, 1000, or 10,000 times greater than the affinity for a control protein. In some embodiments, the peptide binds TFPI with greater affinity than it binds to an "anti-target," a protein or other naturally occurring substance in humans to which binding of the peptide might lead to adverse effects. Several classes of peptides or proteins are potential anti-targets. Because TFPI-inhibitory peptides exert their activity in the blood stream and/or at the endothelium, plasma proteins represent potential anti-targets. Proteins containing Kunitz domains (KDs) are potential anti-targets because KDs of different proteins share a significant similarity. Tissue Factor Pathway Inhibitor-2 (TFPI-2) is highly similar to TFPI-1α and, like TFPI-1α, contains KDs (Sprecher et al., PNAS, 91, 3353-3357 (1994)). Thus, in one aspect, the peptide of the invention binds to TFPI with an affinity that is at least 5, 10, 15, 25, or 50 times greater than the affinity for an anti-target, such as TFPI-2.

[0048] Optionally, the TFPI-binding peptide complex demonstrates one or more desired characteristics described herein, and the amino acid sequence of a peptide can be modified to optimize binding, stability, and/or activity, if desired. An exemplary TFPI-binding peptide complex binds TFPI with a K_D of less than or equal to 20 nM and/or exhibits a binding affinity for TFPI that is at least 100 times greater than the binding affinity for an anti-target. Alternatively or in addition, the TFPI-binding peptide complex enhances FX activation in the presence of TFPI with an EC_{50} (as measured using any suitable assay, such as the assays described here) of less than or equal to 50 nM and/or enhances thrombin formation in the absence of Factor VIII to at least about 20% (e.g., 40%) of the level of thrombin formation in plasma containing physiological levels of Factor VIII. Alternatively or in addition, the TFPI-binding peptide complex achieves a desired level of plasma stability (e.g., 50% or more of a dose remains in plasma after 12 hours) and/or demonstrates a desired half life *in vivo* (e.g., at least two, three, four, five, six, seven, eight, nine, or ten hours). Alternatively or in addition, the TFPI-binding peptide complex exhibits a desired level of bioavailability, such as a desired level of bioavailability following subcutaneous administration (e.g., greater than or equal to 5%, 10%, 15%, 20%, 25%, 30%, or 50%) and/or demonstrates a desired level of TFPI-inhibitory activity at a given dose *in vivo*.

[0049] The invention further includes an *in vitro* method of inhibiting TFPI-1. The method comprises contacting TFPI with a TFPI-binding peptide complex as described herein. Any degree of TFPI-activity inhibition is contemplated. For example, a TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex reduces TFPI-inhibition of the extrinsic pathway at least about 5% (e.g., at least about 10%, at least about 25%, or at least about 30%). In some embodiments, the TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex reduces TFPI activity within the extrinsic pathway at least about 50%, at least about 75%, or at least about 90% compared to TFPI activity in the absence of the peptide.

[0050] In one aspect of the invention, TFPI-binding peptide complexes are used to detect and/or quantify TFPI *in vitro*. An exemplary method of detecting and/or quantifying TFPI in a sample comprises (a) contacting a sample with a TFPI-binding peptide complex of the invention, and (b) detecting binding of the TFPI-binding peptide complex to TFPI.

[0051] The invention further includes an *in vitro* method for targeting biological structures (including, but not limited to, cell surfaces and endothelial lining) where TFPI is located. The method comprises contacting the biological structure (e.g., including, without limitation, a cell displaying TFPI on the cell surface) with a TFPI-binding peptide complex described herein, optionally conjugated to a moiety that adds additional functionality to the peptide. The moiety can be a dye (such as a fluorescence dye), a radionuclide or a radionuclide-containing complex, a protein (e.g., an enzyme, a toxin, or an antibody) or a cytotoxic agent. For example, the peptide is linked or conjugated to an effector moiety that facilitates peptide detection and/or purification and/or comprises therapeutic properties. In one aspect, the TFPI-binding peptide complex or peptide complex conjugate is for use in an *in vivo* method, and is administered to a mammal to target a TFPI-displaying cell within the mammal. Optionally, the method further comprises detecting binding of the TFPI-binding peptide complex to TFPI. The method is useful for therapy and diagnosis of disease where TFPI is a suitable diagnostic marker or TFPI-expressing cells are a target for a therapeutic approach.

[0052] Peptide-TFPI complexes are directly or indirectly detected *in vitro.* Detection moieties are widely used in the art to identify biological substances and include, for example, dye (e.g., fluorescent dye), radionuclides and radionuclide-containing complexes, and enzymes. In some aspects, peptide-TFPI binding is detected indirectly. In this regard, the peptide is optionally contacted with an interaction partner that binds the peptide of invention without significantly interfering with peptide-TFPI binding, and the interaction partner is detected. Exemplary interaction partners include, but are not limited to, antibodies, antigen-binding antibody fragments, anticalins and antibody mimetics, aptamers, streptavidin, avidin, neutravidin, and spiegelmers. Optionally, the interaction partner comprises a detection moiety to facilitate detection of an interaction partner-peptide complex. The TFPI-binding peptide complex is, in some embodiments, modified to facilitate binding of an interaction partner. For example, in one aspect, the TFPI-binding peptide complex is conjugated to biotin, which is bound by an interaction partner comprising streptavidin. An exemplary interaction partner comprises strepavidin fused to horseradish peroxidase, which is detected in, e.g., an ELISA-like assay. Alternatively, the TFPI-binding peptide complex is modified to include an antibody epitope, and binding of the corresponding antibody to the peptide-TFPI complex is detected. Methods of detecting, e.g., antibodies and fragments thereof, are well understood in the art.

[0053] Peptide-TFPI complexes and interaction partner-peptide complexes are identified using any of a number of methods, such as, but not limited to, biochemical assays (e.g., enzymatic assays), spectroscopy (e.g., detection based on optical density, fluorescence, FRET, BRET, TR-FRET, fluorescence polarization, electrochemoluminescence, or NMR), positron emission tomography (PET), and single Photon Emission Computed Tomography (SPECT). Detectable moieties that facilitate fluorescence detection of peptide-TFPI complexes or interaction partner-peptide complexes include, but are not limited to, fluorescein, Alexa Fluor® 350, Marina Blue™, Cascade Yellow™, Alexa Fluor® 405, Pacific Blue™, Pacific Orange™, Alexa Fluor® 430, Alexa Fluor® 488, Oregon Green® 488, Alexa Fluor® 500, Oregon Green® 514, Alexa Fluor® 514, Alexa Fluor® 532, Alexa Fluor® 555, Tetramethylrhodamine, Alexa Fluor® 546, Rhodamine B, Rhodamine Red™-X, Alexa Fluor® 568, Alexa Fluor® 594, Texas Red®, Texas Red®-X, Alexa Fluor® 610, Alexa Fluor® 633, Alexa Fluor® 635, Alexa Fluor® 647, Alexa Fluor® 660, Alexa Fluor® 680, Alexa Fluor® 700, Alexa Fluor® 750, B-Phycoerythrin, R-Phycoerythrin, Allophycocyanin, BODIPY®, Cy3, Cy5, TAMRA, and fluorescent proteins (GFP and derivatives thereof). An example of a TFPI-binding peptide comprising a fluorescent detection moiety is JBT2454 (FAM-Ttds- FQSKpNVHVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 4171)), which is labeled with 5,6-carboxyfluoresceine.

[0054] Radioactive labels also are used to detect biological materials (e.g., TFPI, TFPI-binding peptides, or TFPI-binding peptide-TFPI complexes), and, in some instances, are attached to peptides or interaction partners using a chelator, such as (but not limited to) EDTA (ethylene diamine tetra-acetic acid), DTPA (diethylene triamine pentaacetic acid), CDTA (cyclohexyl 1,2-diamine tetra-acetic acid), EGTA (ethyleneglycol-O,O'-bis(2-aminoethyl)-N,N,N',N'-tetra-acetic), HBED (N,N-bis(hydroxybenzyl)-

ethylenediamine-N,N'-diacetic acid), TTHA (triethylene tetramine hexa-acetic acid), DOTA (1,4,7,10-tetraazacyclododecane-N,N',N",N"'-tetra-acetic acid), HEDTA (hydroxyethyldiamine triacetic acid), or TETA (1,4,8,11-tetra-azacyclotetradecane-N,N',N",N""-tetra-acetic acid). Examples of radioactive labels include ^{99m}Tc, ^{203}Pb, ^{66}Ga, ^{67}Ga, ^{68}Ga, ^{72}As, ^{111}In, ^{113m}In, ^{114m}In, ^{97}Ru, ^{62}Cu, ^{64}Cu, ^{52}Fe, ^{52m}Mn, ^{51}Cr, ^{186}Re. ^{188}Re, ^{77}As, ^{90}Y, ^{67}Cu, ^{169}Er, ^{117m}Sn, ^{121}Sn, ^{127}Te, ^{142}Pr, ^{143}Pr, ^{198}Au, ^{199}Au, ^{149}Tb, ^{161}Tb, ^{109}Pd, ^{165}Dy, ^{149}Pm, ^{151}Pm, ^{153}Sm, ^{157}Gd, ^{166}Ho, ^{172}Tm, ^{169}Yb, ^{175}Yb, ^{177}Lu, ^{105}Rh and ^{111}Ag. Paramagnetic metals also are detectable moieties that are suitable for attachment to TFPI-binding peptides or interaction partners, optionally via chelator complex. Examples of paramagnetic metals include, for example, Cr, Mn, Fe, Co, Ni, Cu, Pr, Nd, Sm, Yb, Gd, Tb, Dy, Ho, and Er.

[0055] TFPI-binding peptide complexes, themselves, are, in some aspects, modified to include one or more amino acids with detectable substituents or nuclides. In this regard, in one embodiment, the TFPI-binding peptide complex comprises at least one amino acid comprising a detectable isotope (e.g., ^{13}C, ^{14}C, ^{35}S, ^{3}H, ^{18}O or ^{15}N), and/or an amino acid that is halogenated with, e.g., ^{123}I, ^{124}I, ^{125}I, ^{131}I, ^{75}Br, ^{76}Br, ^{77}Br or ^{82}Br. Amino acids suitable for halogenation include, but are not limited to, tyrosine and tryptophan.

[0056] The invention also provides TFPI-binding peptide complexes for use in a method for diagnosing a subject suffering from a disease or disorder, or at risk of suffering from a disease or disorder, wherein the disease or disorder is associated with or caused by aberrant TFPI activity. The method comprises administering to the subject the TFPI-binding peptide complex and detecting the TFPI-peptide complex. In some instances, the peptide is conjugated to a detectable moiety, and the method comprises detecting the detectable moiety. Exemplary detectable moieties are described herein. In other instances, the method comprises administering to the subject a TFPI-binding peptide complex interaction partner that binds the TFPI-binding peptide, and detecting the interaction partner. If desired, the interaction partner comprises or is conjugated to a detectable moiety, and the detectable moiety is detected. The presence of the detectable moiety indicates the presence of TFPI, thereby allowing diagnosis of a disease or disorder associated with TFPI (e.g., a disease or disorder which (i) can be treated by inhibiting TFPI or (ii) comprises symptoms which can be ameliorated or prevented by inhibiting TFPI). If administration of the peptide to the subject is not desired, a biological sample is obtained from the subject, contacted with the TFPI-binding peptide complex as described herein, and TFPI-peptide complexes are detected.

[0057] The peptides of the invention bind TFPI and, therefore, are useful for purifying TFPI or recombinant TFPI from a biological sample (e.g., a biological fluid, such as serum), fermentation extract, tissue preparations, culture medium, and the like. The invention includes methods of using the TFPI-binding peptide complex in the commercial production of TFPI or in a method of characterizing TFPI molecules. For example, the invention includes a method of purifying TFPI. The method comprises contacting a sample containing TFPI with a peptide as defined herein under conditions appropriate to form a complex between TFPI and the peptide; removing the complex from the sample; and, optionally, dissociating the complex to release TFPI. Exemplary conditions appropriate to form a complex between TFPI and the peptide are disclosed in the Examples, and such conditions can be easily modified to dissociate the TFPI-peptide complex. In some embodiments, the peptide is immobilized to a support, e.g., a solid support, to facilitate recovery of TFPI. For example, in one embodiment, the peptide is immobilized to chromatography stationary phase (e.g., silica, affinity chromatography beads, or chromatography resins), a sample comprising TFPI is applied to the stationary phase such that TFPI-peptide complexes are formed, the remainder of the sample is removed from the stationary phase, and TFPI is eluted from the stationary phase. In this regard, the peptides of the invention are, in one aspect, suitable for use in affinity chromatography techniques.

[0058] The invention also provides TFPI-binding peptide complexes for use in a method of enhancing thrombin formation in a clotting factor-deficient subject. The method comprises administering to the subject a

peptide provided herein under conditions effective to inhibit TFPI. In this regard, the TFPI-binding peptide complex is administered in an amount and under conditions effective to enhance thrombin formation in the subject. By "clotting factor-deficient" is meant that the subject suffers from a deficiency in one or more blood factors required for thrombin formation, such as FVIII, FIX, or FXI. Indeed, in one embodiment, the subject is deficient in FVIII. Alternatively or in addition, the subject is deficient in Factor IX. Clotting factor deficiencies are identified by examining the amount of factor in a clinical sample. Practitioners classify hemophilia according to the magnitude of clotting factor deficiency. Subjects suffering from mild hemophilia have approximately 5% to 30% of the normal amount (1 U/ml) of Factor VIII or Factor IX. Moderate hemophilia is characterized by approximately 1% to 5% of normal Factor VIII, Factor IX, or Factor XI levels, while subjects suffering from severe hemophilia have less than 1% of the normal amount of Factor VIII, Factor IX, or Factor XI. Deficiencies can be identified indirectly by activated partial thromboplastin time (APTT) testing. APTT testing measures the length of time required for a blood clot to form, which is longer for patients with Factor VIII Deficiency (hemophilia A), Factor IX Deficiency (hemophilia B), and Factor XI Deficiency (hemophilia C) compared to patients with normal clotting factor levels. Almost 100% of patients with severe and moderate Factor VIII deficiency can be diagnosed with an APTT. The invention further includes enhancing thrombin formation in a subject that does not suffer from a clotting factor deficiency. The method comprises administering to a subject (e.g., a subject comprising normal, physiological levels of clotting factor) a peptide provided herein under conditions effective to enhance thrombin formation.

[0059] In one aspect, the TFPI-binding peptide complex is used for increasing blood clot formation in a subject. The method of increasing blood clot formation comprises administering to the subject a peptide described herein in an amount and under conditions effective to increase blood clot formation. It will be appreciated that the method need not completely restore the coagulation cascade to achieve a beneficial (e.g., therapeutic) effect. Any enhancement or increase in thrombin or blood clot formation that reduces the onset or severity of symptoms associated with clotting factor deficiencies is contemplated. Methods of determining the efficacy of the method in promoting thrombin formation and blood clotting are known in the art and described herein.

[0060] The invention further provides TFPI-binding peptide complexes for use in a method of treating a blood coagulation disorder in a subject, the method comprising administering to the subject one or more TFPI-binding peptide complexes, such as any one or more of the peptide complexes described herein, in an amount and under conditions effective to treat the blood coagulation disorder in the subject. In one aspect, the peptide complex is a recombinant or synthetic peptide complex that inhibits TFPI activity. "Coagulation disorders" include bleeding disorders caused by deficient blood coagulation factor activity and deficient platelet activity. Blood coagulation factors include, but are not limited to, Factor V (FV), FVII, FVIII, FIX, FX, FXI, FXIII, FII (responsible for hypoprothrombinemia), and von Willebrand's factor. Factor deficiencies are caused by, for instance, a shortened *in vivo*-half life of the factor, altered binding properties of the factor, genetic defects of the factor, and a reduced plasma concentration of the factor. Coagulation disorders can be congenital or acquired. Potential genetic defects include deletions, additions and/or substitution within a nucleotide sequence encoding a clotting factor whose absence, presence, and/or substitution, respectively, has a negative impact on the clotting factor's activity. Coagulation disorders also stem from development of inhibitors or autoimmunity (e.g., antibodies) against clotting factors. In one example, the coagulation disorder is hemophilia A. Alternatively, the coagulation disorder is hemophilia B or hemophilia C.

[0061] Platelet disorders are caused by deficient platelet function or abnormally low platelet number in circulation. Low platelet count may be due to, for instance, underproduction, platelet sequestration, or uncontrolled patent destruction. Thrombocytopenia (platelet deficiencies) may be present for various reasons, including chemotherapy and other drug therapy, radiation therapy, surgery, accidental blood loss, and other disease conditions. Exemplary disease conditions that involve thrombocytopenia are: aplastic anemia; idiopathic or immune thrombocytopenia (ITP), including idiopathic thrombocytopenic purpura associated with breast cancer; HIV-associated ITP and HIV-related thrombotic thrombocytopenic purpura;

metastatic tumors which result in thrombocytopenia; systemic lupus erythematosus, including neonatal lupus syndrome splenomegaly; Fanconi's syndrome; vitamin B12 deficiency; folic acid deficiency; May-Hegglin anomaly; Wiskott-Aldrich syndrome; chronic liver disease; myelodysplastic syndrome associated with thrombocytopenia; paroxysmal nocturnal hemoglobinuria; acute profound thrombocytopenia following C7E3 Fab (Abciximab) therapy; alloimmune thrombocytopenia, including maternal alloimmune thrombocytopenia; thrombocytopenia associated with antiphospholipid antibodies and thrombosis; autoimmune thrombocytopenia; drug-induced immune thrombocytopenia, including carboplatin-induced thrombocytopenia and heparin-induced thrombocytopenia; fetal thrombocytopenia; gestational thrombocytopenia; Hughes' syndrome; lupoid thrombocytopenia; accidental and/or massive blood loss; myeloproliferative disorders; thrombocytopenia in patients with malignancies; thrombotic thrombocytopenia purpura, including thrombotic microangiopathy manifesting as thrombotic thrombocytopenic purpura/hemolytic uremic syndrome in cancer patients; post-transfusion purpura (PTP); autoimmune hemolytic anemia; occult jejunal diverticulum perforation; pure red cell aplasia; autoimmune thrombocytopenia; nephropathia epidemica; rifampicin-associated acute renal failure; Paris-Trousseau thrombocytopenia; neonatal alloimmune thrombocytopenia; paroxysmal nocturnal hemoglobinuria; hematologic changes in stomach cancer; hemolytic uremic syndromes (e.g., uremic conditions in childhood); and hematologic manifestations related to viral infection including hepatitis A virus and CMV-associated thrombocytopenia. Platelet disorders also include, but are not limited to, Von Willebrand Disease, paraneoplastic platelet dysfunction, Glanzman's thrombasthenia, and Bernard-Soulier disease. Additional bleeding disorders amenable to treatment with a TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex include, but are not limited to, hemorrhagic conditions induced by trauma; a deficiency in one or more contact factors, such as FXI, FXII, prekallikrein, and high molecular weight kininogen (HMWK); vitamin K deficiency; a fibrinogen disorder, including afibrinogenemia, hypofibrinogenemia, and dysfibrinogenemia; and alpha2-antiplasmin deficiency. In one embodiment, the TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex is used to treat excessive bleeding, such as excessive bleeding caused by surgery, trauma, intracerebral hemorrhage, liver disease, renal disease, thrombocytopenia, platelet dysfunction, hematomas, internal hemorrhage, hemarthroses, hypothermia, menstruation, pregnancy, and Dengue hemorrhagic fever. All of the above are considered "blood coagulation disorders" in the context of the disclosure.

[0062] In one aspect, the TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex of the invention is used to reverse the effects (in whole or in part) of one or more anticoagulants in a subject. Numerous anticoagulants are known in the art and include, for instance, heparin; coumarin derivatives, such as warfarin or dicumarol; TFPI; AT III; lupus anticoagulant; nematode anticoagulant peptide (NAPc2); FVIIa inhibitors; active-site blocked FVIIa (FVIIai); active-site blocked FIXa (FIXai); FIXa inhibitors; FXa inhibitors, including fondaparinux, idraparinux, DX-9065a, and razaxaban (DPC906); active-site blocked FXa (FXai); inhibitors of FVa or FVIIIa, including activated protein C (APC) and soluble thrombomodulin; thrombin inhibitors, including hirudin, bivalirudin, argatroban, and ximelagatran; and antibodies or antibody fragments that bind a clotting factor (e.g., FV, FVII, FVIII, FIX, FX, FXIII, FII, FXI, FXII, von Willebrand factor, prekallikrein, or high molecular weight kininogen (HMWK)).

[0063] As used herein, "treating" and "treatment" refers to any reduction in the severity and/or onset of symptoms associated with a blood coagulation disorder. Accordingly, "treating" and "treatment" includes therapeutic and prophylactic measures. One of ordinary skill in the art will appreciate that any degree of protection from, or amelioration of, a blood coagulation disorder or symptom associated therewith is beneficial to a subject, such as a human patient. The quality of life of a patient is improved by reducing to any degree the severity of symptoms in a subject and/or delaying the appearance of symptoms. Accordingly, the method in one aspect is performed as soon as possible after it has been determined that a subject is at risk for developing a blood coagulation disorder (e.g., a deficiency in a clotting factor (e.g., FVIII, FIX, or FXI) is detected) or as soon as possible after a blood coagulation disorder (e.g., hemophilia A, hemophilia B, or hemophilia C) is detected. In an additional aspect, the peptide is administered to protect, in whole or in part, against excessive blood loss during injury or surgery.

[0064] In view of the above, the invention provides a peptide complex for use in a method for the treatment of a subject, such as a method for the treatment of a disease where the inhibition of TFPI is beneficial. In one aspect, the disease or disorder is a blood coagulation disorder. The subject is suffering from a disease or disorder or is at risk from suffering from a disease or disorder (or adverse biological event, such as excessive blood loss). The method comprises administering to the subject the peptide complex of the invention in an amount and under conditions effective to treat or prevent, in whole or in part, the disease or disorder. The invention further provides a peptide complex for use in the manufacture of a medicament. For example, the peptide complex can be used in the manufacture of a medicament for the treatment of a blood coagulation disorder, as described in detail herein.

[0065] In some embodiments, it is advantageous to administer to a subject a nucleic acid comprising a nucleic acid sequence encoding a peptide complex (e.g., TFPI-binding peptide, TFPI-inhibitory peptide) of the invention. Such a nucleic acid, in one aspect, is provided instead of, or in addition to, a peptide complex (e.g., TFPI-binding, TFPI-inhibitory peptide). Expression vectors, nucleic acid regulatory sequences, administration methods, and the like, are further described herein and in U.S. Patent Publication No. 20030045498.

[0066] A particular administration regimen for a particular subject will depend, in part, upon the TFPI-inhibitory peptide complex of the invention used, the amount of TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex administered, the route of administration, the particular ailment being treated, considerations relevant to the recipient, and the cause and extent of any side effects. The amount of peptide complex administered to a subject (e.g., a mammal, such as a human) and the conditions of administration (e.g., timing of administration, route of administration, dosage regimen) are sufficient to affect the desired biological response over a reasonable time frame. Dosage typically depends upon a variety of factors, including the particular TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex employed, the age and body weight of the subject, as well as the existence and severity of any disease or disorder in the subject. The size of the dose also will be determined by the route, timing, and frequency of administration. Accordingly, the clinician may titer the dosage and modify the route of administration to obtain the optimal therapeutic effect, and conventional range-finding techniques are known to those of ordinary skill in the art. Purely by way of illustration, in one aspect, the method comprises administering, e.g., from about 0.1 μg/kg to about 100 mg/kg or more, depending on the factors mentioned above. In other embodiments, the dosage may range from 1 μg/kg up to about 75 mg/kg; or 5 μg/kg up to about 50 mg/kg; or 10 μg/kg up to about 20 mg/kg. In certain embodiments, the dose comprises about 0.5 mg/kg to about 20 mg/kg (e.g., about 1 mg/kg, 1.5 mg/kg, 2 mg/kg, 2.3 mg/kg, 2.5 mg/kg, 3 mg/kg, 3.5 mg/kg, 4 mg/kg, 4.5 mg/kg, 5 mg/kg, 5.5 mg/kg, 6 mg/kg, 6.5 mg/kg, 7 mg/kg, 8 mg/kg, 9 mg/kg, or 10 mg/kg) of peptide. Given the chronic nature of many blood coagulation disorders, it is envisioned that a subject will receive the TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex over a treatment course lasting weeks, months, or years, and may require one or more doses daily or weekly. In other embodiments, the TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex is administered to treat an acute condition (e.g., bleeding caused by surgery or trauma, or factor inhibitor/autoimmune episodes in subjects receiving coagulation replacement therapy) for a relatively short treatment period, e.g., one to 14 days.

[0067] Suitable methods of administering a physiologically-acceptable composition, such as a pharmaceutical composition comprising a peptide described herein, are well known in the art. Although more than one route can be used to administer a peptide, a particular route can provide a more immediate and more effective reaction than another route. Depending on the circumstances, a pharmaceutical composition is applied or instilled into body cavities, absorbed through the skin or mucous membranes, ingested, inhaled, and/or introduced into circulation. In one aspect, a composition comprising a TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex is administered intravenously, intraarterially, or intraperitoneally to introduce

the peptide of the invention into circulation. Non-intravenous administration also is appropriate, particularly with respect to low molecular weight therapeutics. In certain circumstances, it is desirable to deliver a pharmaceutical composition comprising the TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex orally, topically, sublingually, vaginally, rectally, pulmonary; through injection by intracerebral (intra-parenchymal), intracerebroventricular, intramuscular, intra-ocular, intraportal, intralesional, intramedullary, intrathecal, intraventricular, transdermal, subcutaneous, intranasal, urethral, or enteral means; by sustained release systems; or by implantation devices. If desired, the TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex is administered regionally via intraarterial or intravenous administration feeding a region of interest, e.g., via the femoral artery for delivery to the leg. In one embodiment, the peptide is incorporated into a microparticle as described in, for example, U.S. Patents 5,439,686 and 5,498,421, and U.S. Patent Publications 2003/0059474, 2003/0064033, 2004/0043077, 2005/0048127, 2005/0170005, 2005/0142205, 2005/142201, 2005/0233945, 2005/0147689. 2005/0142206, 2006/0024379, 2006/0260777, 2007/0207210, 2007/0092452, 2007/0281031, and 2008/0026068. Alternatively, the composition is administered via implantation of a membrane, sponge, or another appropriate material on to which the desired molecule has been absorbed or encapsulated. Where an implantation device is used, the device in one aspect is implanted into any suitable tissue, and delivery of the desired molecule is in various aspects via diffusion, timed-release bolus, or continuous administration. In other aspects, the TFPI-inhibitory peptide is administered directly to exposed tissue during surgical procedures or treatment of injury, or is administered via transfusion of blood procedures. Therapeutic delivery approaches are well known to the skilled artisan, some of which are further described, for example, in U.S. Patent No. 5,399,363.

[0068] To facilitate administration, the TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex in one embodiment is formulated into a physiologically-acceptable composition comprising a carrier (i.e., vehicle, adjuvant, buffer, or diluent). The particular carrier employed is limited only by chemico-physical considerations, such as solubility and lack of reactivity with the peptide, and by the route of administration. Physiologically-acceptable carriers are well known in the art. Illustrative pharmaceutical forms suitable for injectable use include without limitation sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions (for example, see U.S. Patent No. 5,466,468). Injectable formulations are further described in, e.g., Pharmaceutics and Pharmacy Practice, J. B. Lippincott Co., Philadelphia. Pa., Banker and Chalmers. eds., pages 238-250 (1982), and ASHP Handbook on Injectable Drugs, Toissel, 4th ed., pages 622-630 (1986)). A pharmaceutical composition comprising a peptide provided herein is optionally placed within containers, along with packaging material that provides instructions regarding the use of such pharmaceutical compositions. Generally, such instructions include a tangible expression describing the reagent concentration, as well as, in certain embodiments, relative amounts of excipient ingredients or diluents that may be necessary to reconstitute the pharmaceutical composition.

[0069] When appropriate, the TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex of the invention is administered in combination with other substances and/or other therapeutic modalities to achieve an additional or augmented biological effect. Co-treatments include, but are not limited to, plasma-derived or recombinant coagulation factors, hemophilia prophylaxis treatments, immunosuppressants, plasma factor-inhibiting antibody antagonists (i.e., anti-inhibitors), antifibrinolytics, antibiotics, hormone therapy, anti-inflammatory agents (e.g., Non-Steroidal Anti-Inflammatory Drugs (NSAIDs) or steroidal anti-inflammatory substances), procoagulants, and pain relievers. In one aspect, the method is an adjunct therapy to traditional replacement factor treatment regimens involving administration of, e.g., FXIII, FXII, FXI (e.g., HEMOLEVEN® (Laboratoire francais du Fractionnement et des Biotechnologies, Les Ulis, France) and FXI concentrate (BioProducts Laboratory, Elstree, Hertfordshire, UK)), FX, FIX (e.g., BENEFIX® Coagulation Factor IX (Wyeth, Madison, NJ); ALPHANINE® SD (Grifols, Los Angeles, CA); MONONINE® (CSL Behring, King of Prussia, PA); BEBULIN-VH™ (Baxter, Deerfield, IL); PROFILNINE® SD (Grifols, Los Angeles, CA); or PROPLEX T™ (Baxter, Deerfield, IL)), FVIII (e.g., ADVATE™ (Baxter, Deerfield, IL); HELIXATE® FS (CSL Behring, King of Prussia, PA); REFACTO® (Wyeth, Madison, NJ), XYNTHA™ (Wyeth, Madison, NJ),

KOGENATE® and KOGENATE® FS (Bayer, Pittsburgh, PA); ALPHANATE® (Grifols, Los Angeles, CA); HEMOPHIL M™ (Baxter, Deerfield, IL); KOATE®-DVI (Talecris Biotherapeutics-USA, Research Triangle Park, NC); or MONARC-M™ (Baxter, Deerfield, IL)), FVIIa (e.g., NOVOSEVEN® FVIIa (Novo Nordisk, Princeton, NJ) and FVII concentrate (Baxter Bioscience, Vienna, Austria, or BioProducts Laboratory, Elstree, Hertfordshire, UK)), FV, FVa, FII, and/or FIII, to a subject. In some instances, the subject also receives FEIBA VH Immuno™ (Baxter BioScience, Vienna, Austria), which is a freeze-dried sterile human plasma fraction with Factor VIII inhibitor bypassing activity. FEIBA VH Immuno™ contains approximately equal units of Factor VIII inhibitor bypassing activity and Prothrombin Complex Factors (Factors II, VII, IX, and X and protein C). Other exemplary co-treatments include, but are not limited to, prekallikrein, high molecular weight kininogen (HMWK), Von Willebrand's factor, Tissue Factor, and thrombin. Alternatively or in addition, the TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex is co-formulated with one or more different TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complexes. In one aspect, administration of the TFPI-binding peptide allows a reduction in the dose of co-therapeutic required to achieve a desired biological response.

[0070] The invention thus includes a TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex for use in therapy comprising administering the complex to a subject, in combination with one or more additionally suitable substances(s), each being administered according to a regimen suitable for that medicament. Administration strategies include concurrent administration (i.e., substantially simultaneous administration) and non-concurrent administration (i.e., administration at different times, in any order, whether overlapping or not) of the TFPI-binding peptide (e.g., TFPI-inhibitory peptide) complex and one or more additionally suitable agents(s). It will be appreciated that different components are optionally administered in the same or in separate compositions, and by the same or different routes of administration.

[0071] In some embodiments, the peptide of the invention is conjugated to a moiety, e.g., a therapeutic or diagnostic moiety, such as the detection moieties and co-treatments described above. Alternatively or in addition, the peptide is administered in combination with an interaction partner (e.g., an antibody, antibody fragment, anticalin, aptamer, or spiegelmer) that (a) binds the peptide and (b) is therapeutically active and/or is linked to a moiety that provides additional functionality to the interaction partner (e.g., a therapeutic, diagnostic, or detection agent). Suitable moieties include, but are not limited to, photosensitizers, dyes, radionuclides, radionuclide-containing complexes, enzymes, toxins, antibodies, antibody fragments, and cytotoxic agents, and, in some instances, the moiety possesses therapeutic activity (i.e., achieves an advantageous or desired biological effect). The peptide conjugates or peptide-interaction partner pair is suitable for use in any of the methods described herein, such as methods of treating a subject suffering from a disease or disorder or at risk of suffering from a disease or disorder.

[0072] The invention further provides an *in vitro* method for inhibiting degradation of TFPI by a serine protease. Protease degradation can complicate handling of TFPI in research settings. Additionally, although TFPI inhibition is desired to improve various medical conditions (e.g., hemophilia), TFPI may be desired in other clinical embodiments to, for example, reduce coagulation. By "inhibiting" is meant protection, in whole in part, from degradation or cleavage by a protease. Serine proteases are well characterized and include, e.g., elastase, thrombin, plasmin, FXa, or chymase. The method comprises contacting TFPI with a peptide comples comprising a peptide comprising the structure of formula (XIV):

X7001-X7002-X7003-X7004-X7005-X7006-[X7007-X7008-X7009-X7010-X7011-X7012-X7013-X7014-X7015-X7016-X7017-X7018]-X7019-X7020-X7021-X7022-X7023 (XIV) (SEQ ID NO: 3154),

wherein X7001 is either present or absent, whereby in case X7001 is present it is an amino acid selected from the group consisting of A, C, C(NEM), D, E, F, G, H, I, K, L, P, R, S, T, V and W;

wherein X7002 is either present or absent, whereby in case X7002 is present it is an amino acid selected from the group consisting of A, C, C(NEM), D, E, F, G, H, I, K, L, M, P, Q, R, S, T, V, W and Y;

wherein X7003 is an amino acid selected from the group consisting of A, F, I, K, L, R, S, T, V, W and Y;

wherein X7004 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, R, S, T, V and W;

wherein X7005 is R or W;

wherein X7006 is an amino acid selected from the group consisting of F, H, I, K, L, R, V and W;

wherein X7007 is an amino acid selected from the group consisting of Orn, homoK, C, Hcy, Dap and K, preferably selected from the group consisting of C and Hcy;

wherein X7008 is an amino acid selected from the group consisting of A, G, R, S and T;

wherein X7009 is an amino acid selected from the group consisting of a, A, I, K, L, M, m, Moo, Nle, p, R, Sem and V;

wherein X7010 is an amino acid selected from the group consisting of A, G, I, K, L, P, R, S, T and V;

wherein X7011 is an amino acid selected from the group consisting of D, E, G, S and T;

wherein X7012 is an amino acid selected from the group consisting of A, a, D, d, E, e, F, f, G, I, K, k, L, 1, M, m, Moo, Nle, nle, P, p, R, r, S, s, Sem, T, t, V, v , W and w;

wherein X7013 is an amino acid selected from the group consisting of A, C, C(NEM), Con, Con(Meox), D, d, E, e, Eag, F, G, I, K, L, N, R, S, s, T, V and W;

wherein X7014 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, M, R, S, T, V and W;

wherein X7015 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, M, Nle, R, S, T, V and W;

wherein X7016 is an amino acid selected from the group consisting of A, D, E, F, I, K, L, M, Moo, Nle, R, S, Sem, T, V, W and Y;

wherein X7017 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, R, S, T, V, W and Y;

wherein X7018 is an amino acid selected from the group consisting of C and D, preferably C;

wherein X7019 is an amino acid selected from the group consisting of A, F, I, L, S, T, V and W;

wherein X7020 is an amino acid selected from the group consisting of F and W;

wherein X7021 is an amino acid selected from the group consisting of I, L and V;

wherein X7022 is an amino acid selected from the group consisting of A, D, E, F, G, I, K, L, P, R, S, T, V and W;

wherein X7023 is either present or absent, whereby in case X7023 is present it is an amino acid selected from the group consisting of A, C, C(NEM), Con, Con(Meox), D, E, Eag, F, G, I, K, L, R, S, T, V, W and Y; and

wherein the peptide comprises as a cyclic structure generated by a linkage between X7007 and X7018, whereby degradation of TFPI by the serine protease is inhibited.

[0073] Optionally, X7001 is an amino acid selected from the group consisting of A, D, F, G, H, K, L and S; X7002 is an amino acid selected from the group consisting of H, F, M and R; X7003 is an amino acid selected from the group consisting of F and Y; X7004 is K; X7005 is W; X7006 is an amino acid selected from the group consisting of F and H; X7007 is C; X7008 is an amino acid selected from the group consisting of A, G and S; X7009 is an amino acid selected from the group consisting of M, Sem and V; X7010 is an amino acid selected from the group consisting of K, P and R; X7011 is D; wherein X7012 is an amino acid selected from the group consisting of F, L, 1, M and Sem; wherein X7013 is an amino acid selected from the group consisting of D, G, K and S; wherein X7014 is G; wherein X7015 is an amino acid selected from the group consisting of I and T; wherein X7016 is an amino acid selected from the group consisting of D, F, M, Sem and Y; wherein X7017 is an amino acid selected from the group consisting of S and T; wherein X7018 is C; wherein X7019 is an amino acid selected from the group consisting of A and V; wherein X7020 is W; wherein X7021 is V; wherein X7022 is an amino acid selected from the group consisting of F, L, K, R, P and W; wherein X7023 is either present or absent, whereby in case X7023 is present it is an amino acid sequence selected from the group consisting of A, D, F, M, S and Y; and wherein the peptide comprises as a cyclic structure generated by a linkage between X7007 and X7018. The "contacting" may be performed *in vitro.* The dosage and route of administration considerations described herein are applicable to the method of inhibiting proteolytic degradation.

[0074] The peptide is part of a peptide complex that further comprises a peptide comprising the structure of formula (XIII):

X6001-X6002-X6003-X6004-X6005-X6006-X6007-X6008-X6009-X6010-X6011-X6012-X6013-X6014-X6015-X6016-X6017-X6018-X6019-X6020 (XIII) (SEQ ID NO: 3153);

wherein X6001 is an amino acid selected from the group consisting of F, L, M, Y, 1Ni, Thi, Bta, Dopa, Bhf, C, D, G, H, I, K, N, Nmf, Q, R, T, V, and W;

wherein X6002 is an amino acid selected from the group consisting of Q, G, and K;

wherein X6003 is an amino acid selected from the group consisting of C, D, E, M, Q, R, S, T, Ede(O), Cmc, A, Aib, Bhs, F, G, H, I, K, L, N, P, V, W and Y;

wherein X6004 is an amino acid selected from the group consisting of Aib, E, G, I, K, L, M, P, R, W, Y, A, Bhk, C, D, F, H, k, N, Nmk, Q, S, T and V;

wherein X6005 is an amino acid selected from the group consisting of a, A, Aib, C, D, d, E, G, H, K, k, M, N, Nmg, p, Q, R, NpropylG, aze, pip, tic, oic, hyp, nma, Ncg, Abg, Apg, thz, dtc, Bal, F, L, S, T, V, W and Y;

wherein X6006 is an amino acid selected from the group consisting of A, C, C(NEM), D, E, G, H, K, M, N, Q, R, S, V, Cit, C(Acm), Nle, I, Ede(O), Cmc, Ecl, Eea, Eec, Eef, Nif, Eew, Aib, Btq, F, I, L, T, W and Y;

wherein X6007 is an amino acid selected from the group consisting of I, V, T, Chg, Phg, Tie, A, F, G, I, K, L, Nmv, P, Q, S, W and Y;

wherein X6008 is an amino acid selected from the group consisting of F, H, 1Ni, 2Ni, Pmy, Y, and W;

wherein X6009 is an amino acid selected from the group consisting of Aib, V, Chg, Phg, Abu, Cpg, Tie, L-2-amino-4,4,4-trifluorobutyric acid, A, f, I, K, S, T and V;

wherein X6010 is an amino acid selected from the group consisting of A, C, D, d, E, F, H, K, M, N, P, Q, R, S, T, V, W, Y, Nmd, C(NEM), Aib, G, I, L and Nmf;

wherein X6011 is an amino acid selected from the group consisting of A, a, G, p, Sar, c, hcy, Aib, C, K, G and Nmg;

wherein X6012 is an amino acid selected from the group consisting of Y, Tym, Pty, Dopa and Pmy;

wherein X6013 is an amino acid selected from the group consisting of Aib, C, F, 1Ni, Thi, Bta, A, E, G, H, K, L, M, Q, R, W and Y;

wherein X6014 is an amino acid selected from the group consisting of A, Aib, C, C(NEM), D, E, K, L, M, N, Q, R, T, V, Hcy, Bhe, F, G, H, I, P, S, W and Y;

wherein X6015 is an amino acid selected from the group consisting of R, (omega-methyl)-R, D, E and K;

wherein X6016 is an amino acid selected from the group consisting of L, Hcy, Hle and Aml;

wherein X6017 is an amino acid selected from the group consisting of A, a, Aib, C, c, Cha, Dab, Eag, Eew, H, Har, Hci, Hle, I, K, L, M, Nle, Nva, Opa, Orn, R, S, Deg, Ebc, Eca, Egz, Aic, Apc, Egt, (omega-methyl)-R, Bhr, Cit, D, Dap, E, F, G, N, Q, T, V, W and Y;

wherein X6018 is an amino acid selected from the group consisting of A, Aib, Hcy, hcy, C, c, L, Nle, M, N, R, Bal, D, E, F, G, H, I, K, Q, S, T, V, W and Y;

wherein X6019 is an amino acid selected from the group consisting of K, R, Har, Bhk and V; and

wherein X6020 is an amino acid selected from the group consisting of K, L, Hcy, Aml, Aib, Bhl, C, F, G, H, I, Nml, Q, R, S, T, V, W and Y;

wherein the C-terminus of the first peptide is linked to the N-terminus of the second peptide via a linker moiety;

wherein the first peptide comprises an amino acid sequence at least 80%, at least 85%, at least 90%, at least 95%, or 100% identical to SEQ ID NO: 178 or SEQ ID NO: 4261; and

wherein the second peptide comprises an amino acid sequence at least 80%, at least 85%, at least 90%, at least 95%, or 100% identical to SEQ ID NO: 1044.

[0075] The invention further provides a method for identifying a TFPI-binding compound, such as a TFPI-binding peptide. In one aspect, the method comprises (a) contacting a peptide comprising TFPI Kunitz domain 1 (KD1) with a TFPI-binding peptide complex described herein and a test compound under conditions that allow formation of KD1-TFPI-binding peptide complexes. The method further comprises (b) measuring KD1-TFPI-binding peptide complexes formed in step (a), and (c) comparing the number of KD1-TFPI-binding peptide complexes formed in the presence of the test compound with the number of KD1-TFPI-binding peptide complexes formed in the absence of the test compound. A reduction in the number of KD1-TFPI-binding peptide complexes formed in the presence of the test compound compared to the number of KD1-TFPI-binding peptide complexes formed in the absence of the test compound indicates that the test compound is a TFPI-binding compound. In one aspect, the method further comprises forming KD1-TFPI-binding complexes in the absence of the test compound for comparison in step (c), although this is not required inasmuch as the information may be obtained separately (e.g., from previously prepared reference standards).

[0076] KD1, the TFPI-binding peptide, and the test compound are combined simultaneously or sequentially, optionally with washing steps before and/or after addition of the TFPI-binding peptide and/or the test compound. In one embodiment, the peptide comprising KD1 is contacted with a TFPI-binding peptide described herein under conditions that allow formation of KD1-TFPI-binding peptide complexes, unbound TFPI-binding peptide is removed, and the remaining KD-peptide complexes are contacted with a test

compound. Displacement of the TFPI-binding peptide from the TFPI-peptide complexes is detected, and indicates that the test compound is a TFPI-binding compound. Displacement is detected by, for example, measuring the number of KD1-TFPI-binding peptide complexes before and after exposure to the test compound.

[0077] KD1-TFPI-binding peptide complexes are detected and/or measured (quantified) using any suitable detection means, including detection means known in the art for detecting peptides in a sample. For example, in one embodiment of the invention, the TFPI-binding peptide comprises a label that generates a signal. Exemplary labels are described herein and include, e.g., radionuclides, fluorescent dyes, isotopes, enzyme substrates, and enzymes. The method comprises measuring signal generated by KD1-TFPI-binding peptide complexes and comparing signal generated by KD1-TFPI-binding peptide complexes formed in the presence of the test compound with signal generated by KD1-TFPI-binding peptide complexes formed in the absence of the test compound. A reduction in signal from a sample comprising KD1-TFPI-binding peptide complexes exposed to test compound (compared to signal generated by a similar sample of KD1-TFPI-binding peptide complexes not exposed to the test compound) indicates that complex formation has been inhibited or disrupted, and that the test compound is a TFPI-binding compound.

[0078] The entire document is intended to be related as a unified disclosure, and it should be understood that all combinations of features described herein are contemplated, even if the combination of features are not found together in the same sentence, or paragraph, or section of this document. For example, where protein therapy is described, embodiments involving polynucleotide therapy (using polynucleotides/vectors that encode the protein) are specifically contemplated, and the reverse also is true. The invention includes, for instance, all embodiments of the invention narrower in scope in any way than the variations specifically mentioned above. With respect to aspects of the invention described as a genus, all individual species are individually considered separate aspects of the invention. With respect to aspects of the invention described or claimed with "a" or "an," it should be understood that these terms mean "one or more" unless context unambiguously requires a more restricted meaning. With respect to elements described as one or more within a set, it should be understood that all combinations within the set are contemplated.

EXAMPLES

[0079] The invention, thus generally described, will be understood more readily by reference to the following examples, which are provided by way of illustration and are not intended to limit the invention.

Example 1

[0080] The following example describes production, identification, and screening of peptides for binding to TFPI.

[0081] Peptides candidates were obtained from commercial suppliers (e.g., PolyPeptide Laboratories SAS (Strasbourg, France) and JPT Peptide Technologies GmbH (Berlin, Germany)). Methods for synthesizing candidate peptides are provided above. Candidate peptides were synthesized as trifluoroacetate (TFA) salts with a purity >90% or >60%. All peptides were solved in DMSO to a stock concentration of 10 mM. TFPI-binding peptide sequences were identified using an mRNA display library. The mRNA display technology is superior to other library screening techniques for allowing for a diversity of 10^{14} different sequences within a starting pool and avoiding, e.g., the *in vivo* steps required for phage display. In brief, the technology involves directly linking mRNA to its encoded candidate peptide through a puromycin molecule (Figure 5). The mRNA display method is further described in International Patent Publication No. WO 2005/051985 and Liu et al.,

Methods in Enzymology, 318, 268-293 (2000). TFPI was immobilized to a solid support via biotin and exposed to candidate peptide-RNA complexes. TFPI-bound candidate peptide-RNA complexes were isolated, and the RNA reverse transcribed to obtain coding DNA. High affinity binders were obtained following six to ten selection rounds using a competitive elusion strategy. Many of the candidate peptides were 31 amino acids in length (27 randomized amino acids and 2 amino acids flanking both termini).

[0082] Selected peptides were synthesized and subjected to peptide optimization using a microarray-based scan analysis to identify peptide fragments retaining TFPI-binding affinity. For example, a microarray-based scan of JBT0047 was performed using a series of 20 amino acid fragments of the peptide, the sequences of which overlapped by 19 amino acids. Briefly, N-terminally, aminooxyacetate-modified peptides were printed on Corning epoxide glass slides. After washing and drying, the slides were treated in a TECAN HS400™ incubation station. Slides were washed for two minutes in Tris-buffered saline with 0.1% TWEEN 20® (TBST), and blocked for 30 minutes in Tris-based, T-20 SuperBlock™ buffer (5 mM $CaCl_2$) (Pierce). After blocking, the slides were washed for 2.5 minutes in TBST. The slides were subsequently incubated with DYLIGHT™ 649-labeled TFPI (1 µg/ml in Tris-based, T-20 SuperBlock™ buffer (5 mM $CaCl_2$)) for 45 minutes, and washed twice with continuous flow TBST for ten minutes. The slides were subjected to a final wash with saline-sodium citrate buffer for two minutes, and air-dried for four minutes. The slides were scanned in an Axon GenePix® 4000B scanner, and scans were analyzed using the GenePix® Pro software. N-and C-terminal truncation analysis supplemented the scan analysis. The microarray scan results demonstrated that peptide JBT0293 bound TFPI with the highest affinity. A series of substitution mutants based on the amino acid sequence of JBT0293 was generated and tested for TFPI binding properties.

[0083] The affinity of a subset of peptides for TFPI was demonstrated via an enzyme-linked immunosorbent assay (ELISA)-like assay (binding (EC_{50}) ELISA) performed with biotinylated peptides. Ninety-six well MaxiSorp plates (Nunc) were coated with 3 µg/mL TFPI in coating buffer (15 mM Na_2CO_3, 35 mM $NaHCO_3$, pH 9.6) over night. Plates were washed three times with 350 µl wash buffer (HNaT: 175 mM NaCl, 25 mM HEPES, 5 mM $CaCl_2$, 0.1% Tween 80, pH 7.35), and subsequently blocked with 200 µl 2% yeast extract in HNaT for 2 hours. Plates were then washed three times with 350 µl HNaT. Biotinylated candidate peptides were diluted from a DMSO stock 1/200 in HNaT. The initial peptide concentration was 50 µM if no precipitate appeared during the 1/200 dilution of the 10 mM peptide stock solution. Pre-dilutions of the peptide stock in DMSO were conducted if precipitates formed. The diluted peptides were applied to the Maxisorp plates, serial dilutions (1/3) were generated, and the dilutions were incubated for 1.5 hours at room temperature. Incubation was followed by three wash steps (350 µl HNaT). Bound peptide was detected by incubation with horseradish peroxidase-conjugated streptavidin (1 hour), followed by three wash steps with HNaT and a subsequent chromogenic conversion of added TMB (3,3'5,5'-Tetramethylbenzidin). The assay is illustrated in Figure 6A.

[0084] Generally, peptide binding to immobilized TFPI was significantly above background. EC_{50} values for biotinylated peptides are given in Figures 32-39. The binding curve of one TFPI-binding peptide, JBT0132, is depicted in Figure 7. The EC_{50} of JBT0132 was calculated to be about 2.2 nM.

[0085] In addition, a competition (IC_{50}) ELISA was performed using biotinylated TFPI-binding peptides as "tracers" to compete for TFPI-binding with non-biotinylated candidate peptides. The assay principle is depicted in Figure 6B. Ninety-six well MaxiSorp plates (Nunc) were coated with 3 µg/mL TFPI in coating buffer (15 mM Na_2CO_3, 35 mM $NaHCO_3$, pH 9.6) over night. The concentration of TFPI can be altered depending on the particular conditions of the assay; in other IC_{50} ELISA assays referenced herein, the coating buffer contained 0.05 µg/ml TFPI. Plates were washed three times with 350 µl wash buffer (HNaT: 175 mM NaCl, 25 mM HEPES, 5 mM $CaCl_2$, 0.1% Tween 80, pH 7.35), and blocked with 200 µl 2% yeast extract in HNaT for 2 hours. Plates were then washed three times with 350 µl HNaT. Biotinylated tracer

peptides were applied at a concentration corresponding to their respective EC_{90} values determined in the binding ELISA (median if n > 2). A competitor stock solution of peptide (10 mM) was diluted 1/33.3 in HNaT without HSA, and a serial 1/3 dilution was prepared with HNaT with 3% DMSO. The dilution strategy employed in a particular assay will depend on the affinity of the peptides. The dilution was further diluted with the biotinylated tracer peptide in a ratio of 1:6 (20 µl competitor dilution and 100 µl tracer peptide). The mixture of competitor and tracer peptide was applied to the TFPI-coated microtiter plate and incubated for 1.5 hours. The plates were washed three times with 350 µl HNaT. Peptide-TFPI binding was detected by applying HRP-conjugated streptavidin to the microtiter plate, incubating the mixture for one hour, washing the plate three times with 350 µl HNaT, applying TMB (3,3'5,5'-Tetramethylbenzidin), and detecting the subsequent chromogenic conversion of TMB by HRP. IC_{50} graphs for representative non-biotinylated peptides are provided in Figures 8A-8D. IC_{50} measurements of peptides JBT0303, JBT0120, and JBT0224 are set forth in Table 3.

TABLE 3

Peptide	IC_{50} [µM]	n	SD	Tracer Peptide	Tracer Concentration [µM]
JBT0303	0.119	2	0.064	JBT0131	0.0409
JBT0120	0.0189	3	0.0044	JBT0124	0.0718
JBT0224	n.a.	1		JBT0126	0.240

[0086] In addition to the competition ELISA (IC_{50}) assay, a screening assay was employed to measure higher numbers of peptides in parallel. The screening ELISA is similar to the competition IC_{50} ELISA with the exception that only three different concentrations of the competitor were employed (300 nM, 100 nM and 33.3 nM for the JBT0047 class, and 50000 nM, 16667 nM and 5556 nM for the JBT0122 class). In some instances, screening results were expressed as percent inhibition of the tracer signal in relation to a competitive peptide (competitive peptide JBT0477 for the JBT0047 family, and competitive peptide JBT1697 for the JBT0122 family). The competition IC_{50} assay results and the screening assay results of peptides prepared and screened in accordance with the methods set forth herein are provided in Figures 32-39. The mean IC_{50} values presented in Figures 32-39 are based on a greater number of assays than the values presented in Table 3 and, therefore, the values may differ slightly. The results of the screening ELISA are presented as percent inhibition of tracer peptide JBT0131 binding. Several peptides that were analyzed using the IC_{50} ELISA are classified in Figures 32-39 according to their binding affinity as set forth in Table 4.

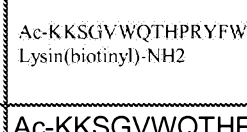
TABLE 4

TFPI competition ELISA IC_{50} [nM]	Group
< 50 nM	A
50 ≤ x <100 nM	B
100 ≤ x < 250 nM	C
250 ≤ x < 1000 nM	D
1000 ≤ x < 5000 nM	E
5000 ≤ x < 10000 nM	F
10000 ≤ x < 50000 nM	G

[0087] Exemplary TFPI-binding peptides identified using the methods described herein are presented in Table 5. Some peptides were biotinylated, and many comprise N- and C-terminal lysines to promote solubility. Several peptides exhibited TFPI-inhibitory activity in model and/or plasmatic assay systems, as described below.

TABLE 5

Peptide	Parent	Sequence	SEQ ID
	JBT0047	QSKKNVFVFGYFERLRAK	1
JBT0047	JBT0047	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-NH2	253
JBT0051	JBT0047	Biotinyl-Ttds-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-NH2	962
JBT0055	JBT0047	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-Ttds-Lys(Biotinyl)-NH2	963
JBT0131	JBT0047	Biotinyl-Ttds-AFQSKKNVFVFGYFERLRAK-NH2	964
JBT0132	JBT0047	Biotinyl-Ttds-FQSKKNVFVFGYFERLRAKL-NH2	965
JBT0133	JBT0047	Biotinyl-Ttds-QSKKNVFVFGYFERLRAKLT-NH2	966
JBT0155	JBT0047	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2	8
JBT0158	JBT0047	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKKK-NH2	9
JBT0162	JBT0047	Ac-KKGRLQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2	10
JBT0163	JBT0047	Ac-KKQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2	11
JBT0164	JBT0047	Ac-KKFQSKKNVFVFGYFERLRAKLTSKK-NH2	12
JBT0166	JBT0047	Biotinyl-Ttds-KKFQSKKNVFVFGYFERLRAKLKK-NH2	968
JBT0169	JBT0047	Ac-KKAFQSKKNVFVFGYFERLRAKKK-NH2	254
JBT0170	JBT0047	Ac-KKFQSKKNVFVFGYFERLRAKLKK-NH2	13
JBT0171	JBT0047	Ac-KKQSKKNVFVFGYFERLRAKLTKK-NH2	255
JBT0174	JBT0047	Ac-KKAFQSKKNVFVFGYFERLRAKLKK-NH2	14
JBT0175	JBT0047	Ac-KKAFQSKKNVFVFGYFERLRAKLTKK-NH2	182
JBT0293	JBT0047	Ac-FQSKKNVFVFGYFERLRAKL-NH2	256
		$X_3X_4X_5KX_7NVFX_{11}X_{12}GYX_{15}X_{16}RLRAKX_{22}$	2
JBT0294	JBT0047	Ac-YQSKKNVFVFGYFERLRAKL-NH2	257
JBT0295	JBT0047	Ac-FSSKKNVFVFGYFERLRAKL-NH2	713
JBT0296	JBT0047	Ac-FQNKKNVFVFGYFERLRAKL-NH2	407
JBT0297	JBT0047	Ac-FQSKNNVFVFGYFERLRAKL-NH2	183
JBT0298	JBT0047	Ac-FQSKQNVFVFGYFERLRAKL-NH2	747
JBT0299	JBT0047	Ac-FQSKKNVFAFGYFERLRAKL-NH2	408
JBT0300	JBT0047	Ac-FQSKKNVFSFGYFERLRAKL-NH2	409
JBT0301	JBT0047	Ac-FQSKKNVFTFGYFERLRAKL-NH2	470
JBT0302	JBT0047	Ac-FQSKKNVFVAGYFERLRAKL-NH2	258
JBT0303	JBT0047	Ac-FQSKKNVFVDGYFERLRAKL-NH2	184
JBT0304	JBT0047	Ac-FQSKKNVFVLGYFERLRAKL-NH2	259
JBT0305	JBT0047	Ac-FQSKKNVFVQGYFERLRAKL-NH2	260
JBT0306	JBT0047	Ac-FQSKKNVFVSGYFERLRAKL-NH2	185
JBT0307	JBT0047	Ac-FQSKKNVFVYGYFERLRAKL-NH2	261
JBT0308	JBT0047	Ac-FQSKKNVFVFGYKERLRAKL-NH2	411
JBT0309	JBT0047	Ac-FQSKKNVFVFGYYERLRAKL-NH2	412
JBT0310	JBT0047	Ac-FQSKKNVFVFGYFDRLRAKL-NH2	262

Peptide	Parent	Sequence	SEQ ID
JBT0311	JBT0047	Ac-FQSKKNVFVFGYFERLRAKN-NH2	748
		TFVDERLLYFLTIGNMGMYAAQLKF	3
JBT0049	JBT0049	Ac-SGNTFVDERLLYFLTIGNMGMYAAQLKFRTS-NH2	3025
JBT0053	JBT0049	Biotinyl-Ttds-SGNTFVDERLLYFLTIGNMGMYAAQLKFRTS-NH2	3006
JBT0057	JBT0049	Ac-SGNTFVDERLLYFLTIGNMGMYAAQLKFRTS-Ttds-Lysin(biotin)-NH2	3018
JBT0190	JBT0049	Ac-KKSGNTFVDERLLYFLTIGNMGMYAAQLKFRTSKK-NH2	3031
JBT0193	JBT0049	Ac-KKSGNTFVDERLLYFLTIGNMGMYAAQLKFKK-NH2	3073
JBT0197	JBT0049	Ac-KKTFVDERLLYFLTIGNMGMYAAQLKFRTSKK-NH2	3076
		VIVFTFRHNKLIGYERRY	4
JBT0050	JBT0050	Ac-SGRGCTKVIVFTFRHNKLIGYERRYNCTS-NH2	3047
JBT0054	JBT0050	Biotinyl-Ttds-SGRGCTKVIVFTFRHNKLIGYERRYNCTS-NH2	3002
JBT0058	JBT0050	Ac-SGRGCTKVIVFTFRHNKLIGYERRYNCTS-Ttds-Lysin(biotin)-NH2	3003
JBT0129	JBT0050	Ac-SGRG[CTKVIVFTFRHNKLIGYERRYNC]TS-NH2	3026
JBT0130	JBT0050	Biotinyl-Ttds-SGRG[CTKVIVFTFRHNKLIGYERRYNC]TS-NH2	3001
JBT0205	JBT0050	Ac-KKSGRGCTKVIVFTFRHNKLIGYERRYNCTSKK-NH2	3029
JBT0208	JBT0050	Ac-KKSGRGCTKVIVFTFRHNKLIGYERRYNKK-NH2	3027
JBT0211	JBT0050	Ac-KKGCTKVIVFTFRHNKLIGYERRYNCTSKK-NH2	3032
JBT0212	JBT0050	Ac-KKKVIVFTFRHNKLIGYERRYNCTSKK-NH2	3033
JBT0217	JBT0050	Ac-KKTKVIVFTFRHNKLIGYERRYKK-NH2	3062
JBT0218	JBT0050	Ac-KKKVIVFTFRHNKLIGYERRYNKK-NH2	3063
JBT0219	JBT0050	Ac-KKVIVFTFRHNKLIGYERRYNCKK-NH2	3030
		GVWQTHPRYFWTMWPDIKGEVIVLFGT	5
JBT0101	JBT0101	Ac-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTSKK-NH2	3036
JBT0052	JBT0101	Biotinyl-Ttds-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTSKK-NH2	3004
JBT0103	JBT0101	 Ac-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTS-Ttds-KK-Lysin(biotinyl)-NH2	3005
JBT0178	JBT0101	Ac-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTKK-NH2	3028
JBT0182	JBT0101	Ac-KKGVWQTHPRYFWTMWPDIKGEVIVLFGTSKK-NH2	3037
		KWFCGMRDMKGTMSCVWVKF	6
JBT0120	JBT0120	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2	1047
JBT0124		Biotinyl-Ttds-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2	1290
JBT0247	JBT0120	Ac-SGASRYKWFCGMRDMKGTMSCVWVKFRYDTS-NH2	1213
JBT0248	JBT0120	Ac-KKSGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTSKK-NH2	1001
JBT0251	JBT0120	Ac-KKKWFCGMRDMKGTMSCV WVKFKK-NH2	1202

Peptide	Parent	Sequence	SEQ ID
JBT0252	JBT0120	Ac-KKCGMRDMKGTMSCVWVKFRYDKK-NH2	1215
		ASFPLAVQLHVSKRSKEMA	7
JBT0122	JBT0122	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2	2002
JBT0126	JBT0122	Biotinyl-Ttds-SGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2	2498
JBT0221	JBT0122	Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKTSKK-NH2	2003
JBT0224	JBT0122	Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKK-NH2	2298
JBT0225	JBT0122	Ac-KKSGYASFPLAVQLHVSKRSKEMALARKK-NH2	2128
JBT0226	JBT0122	Ac-KKSGYASFPLAVQLHVSKRSKEMAKK-NH2	2299
JBT0228	JBT0122	Ac-KKASFPLAVQLHVSKRSKEMALARLYYKTSKK-NH2	2016
JBT0232	JBT0122	Ac-KKGYASFPLAVQLHVSKRSKEMKK-NH2	2303
JBT0233	JBT0122	Ac-KKYASFPLAVQLHVSKRSKEMAKK-NH2	2304

[0088] This example provides exemplary methods of generating and characterizing TFPI-binding peptides (e.g., TFPI-inhibitory peptides). All peptides in Table 5 were found to bind human TFPI-1α. Mutation analysis demonstrated that at least one amino acid in a TFPI-binding peptide may be substituted while retaining affinity for TFPI. The peptides of Table 5 tested in ELISA assays bound TFPI-la with an EC_{50} of less than 10 μM (1 x 10^{-5} M) and an IC_{50} of less than 50 μM.

Example 2

[0089] Selected TFPI-binding peptides were further characterized in terms of "anti-target" binding. This example demonstrates that TFPI-binding peptides exhibit reduced affinity for non-TFPI-1 proteins.

[0090] TFPI-2 was selected as an anti-target because of its similarity to TFPI-1. The binding kinetics of TFPI-binding peptides to human TFPI-1 (residues 29-282 fused at the C-terminus to a 10 His-tag; MW 41 kDa (R&D Systems, Minneapolis, MN; catalog number 2974-PI)) murine TFPI-1 (residues 29-289 fused at the C-terminus to a 10 His-tag; MW 41kDa (R&D Systems; catalogue number 2975-PI)), and TFPI-2 (R&D Systems, Minneapolis, MN) were studied using a BIAcore 3000™ surface plasmon resonance assay (GE Healthcare, Chalfont St. Giles, UK). TFPI proteins were immobilized on a C1 chip (GE Healthcare, Order Code: BR-1005-40) by amine coupling chemistry aiming for 500 RU. Several TFPI-binding peptides were employed as analytes for interacting with the immobilized TFPI proteins. A flow rate of 30 μl/min was utilized. After 180 seconds, 180 μl of peptide solution was injected at six different concentrations ranging from 3.84 nM to 656.25 nM, followed by a dissociation time of 480 seconds. The chip was regenerated with 45 μl 10 mM NaOH. Each binding experiment was preceded and followed by four measurements with HBS-P buffer (10 mM HEPES, pH 7.4, 150 mM NaCl, 0.005% P20) plus 1% DMSO and 0.8% P80. BIAevaluation® Version 4.1 software (GE Healthcare) was employed to analyze the data. Sensorgrams were fitted to a 1:1 Langmuir binding curve to determine k_{on} and k_{off} and calculate K_D.

[0091] Certain tested peptides, e.g., JBT0050, JBT0121, JBT0205 and JBT0211, bound to the blank cell and binding constants from those sensorgrams could not be determined. JBT0133 showed weak binding to TFPI-1. Sensorgrams from other peptides gave reliable binding constants. Results from BIAcore analysis of several TFPI-binding peptides is provided in Table 6 and Figures 19-21. Each of the peptides listed in Table 6 presented a K_D of less than 10 μM. In addition to the peptides listed below, JBT0375 and JBT0477, substitution mutants of JBT0293 at amino acid position 5 (JBT0375) or amino acid positions 5 and 10

(JBT0477), also exhibited a K_D of less than 10 μM. JBT1837 (SEQ ID NO: 1044) also demonstrated a K_D of less than 10 μM (K_D = 0.5 nM; k_{on} = 8 x 10^3 1/Ms; k_{off} = 1.3 x 10^{-6} 1/s). Sensorgrams of two of the peptides are provided as Figures 9A and 9B.

TABLE 6

Peptide	k_{on} (1/Ms)	k_{off} (1/s)	K_D (M)
JBT0047	4.0 x 10^5	1.9 x 10^{-2}	4.7 x 10^{-8}
JBT0120	1.17 x 10^6	4.78 x 10^{-2}	4.08 x 10^{-8}
JBT0131	1.4 x 10^5	6.0 x 10^{-2}	4.31 x 10^{-7}
JBT0132	3.55 x 10^4	3.26 x 10^{-2}	9.17 x 10^{-7}
JBT0224	6.39 x 10^4	1.95 x 10^{-2}	3.05 x 10^{-7}
JBT0293	6.0 x 10^5	5.6 x 10^{-2}	9.5 x 10^{-8}
JBT0297	5.0 x 10^5	1.4 x 10^{-2}	2.9 x 10^{-8}
JBT0303	8.13 x 10^5	2.75 x 10^{-2}	3.4 x 10^{-8}
JBT0305	7.5 x 10^5	3.1 x 10^{-2}	6.1 x 10^{-8}

[0092] Interaction with the TFPI-2 anti-target also was examined. The maximum signal generated from candidate peptide interaction with human TFPI-2 was much lower than the signals obtained with TFPI-1 as an interaction partner. Kinetic analysis of the low TFPI-2 binding signals was prone to error; therefore, visual comparison of sensorgrams was used to estimate binding affinity. A sensorgram illustrating JBT0120 binding to TFPI-1 and TFPI-2 is provided as Figures 10A and 10B. JBT0120 binds TFPI-2 with 10-fold lower affinity compared to its binding affinity for TFPI-1. JBT0132 also was found to exhibit at least 10-fold greater affinity for TFPI-1 than TFPI-2.

[0093] The data provided by this example confirm that TFPI-binding peptides specifically bind TFPI-1.

Example 3

[0094] The following example describes the characterization of TFPI-inhibitory activity of select peptides identified in Example 1 using FXa inhibition and extrinsic tenase inhibition assays. Both assays are predictive of activity in plasmatic systems. The extrinsic tenase assay gives insight into the influence of the peptides on (a) the interaction of FXa and TFPI and (b) the interaction of the FXa-TFPI complex with the TF-FVIIa complex. The FXa inhibition assay measures a peptide's influence on the interaction of FXa and TFPI only.

[0095] The extrinsic tenase complex is responsible for FX and FIX activation upon initiation of the coagulation process. The extrinsic complex is composed of FVIIa, Tissue Factor (TF), and FX substrate. To determine the influence of peptides on the TFPI-mediated inhibition of the extrinsic tenase complex, a coupled enzyme assay was established. Peptides were diluted 1/6.25 from 10 mM stocks (in DMSO) and further diluted by serial 1/4 dilutions in buffer or DMSO to prevent unwanted precipitation. TFPI was diluted in HNaCa-HSA or BSA (25mM HEPES; 175mM NaCl; 5mM $CaCl_2$; 0.1% HSA or BSA; pH 7.35). FVIIa, lipidated TF, phospholipid vesicles (DOPC/ POPS 80/20), and chromogenic substrate specific for FXa (S-2222 (available from DiaPharma, West Chester, OH)), all diluted in HNaCa-HSA, were added to 96-well plates. After an incubation period, TFPI and peptide dilutions were added, resulting in a final concentration of 2.5% DMSO (if present in the peptide stock). FX activation was initiated by adding FX to the wells. FXa-mediated

chromogenic substrate conversion was determined by observing an increase in absorbance using a micro-plate reader. The amount of FXa generated at certain time points was calculated from the OD readings. FXa generated at 20 minutes after start of the reaction was considered for calculation of EC_{50} from plots of peptide concentration versus the inhibition of TFPI (%).

[0096] The functional inhibition of TFPI also was examined using a FXa inhibition assay. A FXa-specific chromogenic substrate (S-2222) and TFPI, both diluted in HNaCa-HSA, were added to 96 well plates. Peptides were diluted 1/6.25 from 10 mM stocks (in DMSO or Aqua-Dest) and further diluted by serial 1/4 dilutions in buffer or DMSO to prevent unwanted precipitation. The peptide dilutions (2.5 μl) were added to the 96 well plates, resulting in a final concentration of 2.5% DMSO (if present in the peptide stock). The conversion of chromogenic substrate was triggered by the addition of FXa, and the kinetics of the conversion were measured in a micro-plate reader. Because TFPI inhibits FXa slowly, OD readings after 115 minutes were considered for calculation of the EC_{50} from plots of peptide concentration versus the inhibition of TFPI (%).

[0097] Results from the extrinsic tenase assay and FXa inhibition assay are provided in Table 7 and Figures 22-27.

TABLE 7

	FXa Inhibition Assay		Extrinsic Tenase Assay	
	EC_{50} [μM]	**% inhibition @ 2.5 μM**	**EC_{50} [μM]**	**% inhibition @ 2.5 μM**
JBT0120	0.9	45	0.9	45
JBT0132	1.2	36	0.1	10
JBT0224	n.a.	26	3.5	18
JBT0303	1.2	61	n.a.	8

[0098] Referring to Table 7, JBT0120, JBT0132, and JBT0224 restored extrinsic complex-mediated FX activation in the presence of TFPI-1 with an EC_{50} of < 2 μM, resulting in between about 20% to about 60% inhibition of TFPI activity. JBT0047 (EC_{50}=1.4 μM), JBT0131 (EC_{50} =2.2 μM), and JBT0293 (EC_{50}=2.9 μM) also restored extrinsic complex activity in the presence of TFPI-1. In addition, JBT0120, JBT0132, JBT0224, and JBT0303 restored FXa activity in the presence of TFPI-1 with an EC_{50} of < 5 μM, resulting in between about 5% to about 50% inhibition of TFPI activity, in the FXa inhibition assay. JBT0047 (EC_{50}=0.7 μM), JBT0131 (EC_{50} =8.2 μM), JBT0293 (EC_{50}=1.3 μM), JBT0297 (EC_{50}=0.6 μM), and JBT0305 (EC_{50}=2.3 μM) also restored activity of FXa in the presence of TFPI-1 in the FXa inhibition assay. This example confirms that peptides of the invention are TFPI antagonists.

Example 4

[0099] In this example, the TFPI inhibitory activity of peptides is established using a plasma-based assay.

[0100] The influence of peptides on thrombin generation was measured in duplicate via calibrated automated thrombography in a Fluoroskan Ascent® reader (Thermo Labsystems, Helsinki, Finland; filters 390 nm excitation and 460 nm emission) following the slow cleavage of the thrombin-specific fluorogenic substrate Z-Gly-Gly-Arg-AMC (Hemker, Pathophysiol. Haemost. Thromb., 33, 4-15 (2003)). Plasma from patients with FVIII or FIX deficiency (George King Bio-Medical Inc., Overland Park, KN) was obtained for testing. The residual coagulation factor activity for each of the plasmas was lower than 1%. As a model for antibody-mediated FVIII deficiency, frozen pooled normal plasma (George King Bio-Medical Inc., Overland Park, KN)

was incubated with high titer, heat inactivated, antihuman FVIII plasma raised in goat (4490 BU/ml; Baxter BioScience, Vienna, Austria) giving rise to 50 BU/mL. The plasmas were mixed with corn trypsin inhibitor (CTI) (Hematologic Technologies, Inc., Essex Junction, VT) to inhibit Factor XIIa contamination, resulting in a final concentration of 40 µg/mL.

[0101] Pre-warmed (37°C) plasma (80 µL) was added to each well of a 96 well micro-plate (Immulon 2HB, clear U-bottom; Thermo Electron, Waltham, MA). To trigger thrombin generation by Tissue Factor, 10 µL of PPP low reagent containing low amounts (12 pM) of recombinant human Tissue Factor and phospholipid vesicles composed of phosphatidylserine, phosphatidylcholine and phosphatidylethanolamine (48 µM) (Thrombinoscope BV, Maastricht, The Netherlands) were added. Peptides were diluted 1/7.5 from 10 mM stocks with DMSO, and further diluted 1/8.33 with Aqua-Dest resulting in a DMSO concentration of 12%, providing a 0.5% DMSO concentration in the final assay mix. Just prior putting the plate into the pre-warmed (37°C) reader, 5 µL of HEPES buffered saline with 5 mg/mL human serum albumin (Sigma-Aldrich Corporation, St. Louis, Missouri, USA) or 12% DMSO in Aqua-Dest was added, followed by addition of the peptide dilutions or reference proteins (FVIII Immunate reference standard (Baxter BioScience, Vienna, Austria); Factor VIII Inhibitor By-Passing Activity (FEIBA) reference standard (Baxter BioScience, Vienna, Austria); NovoSeven (Novo Nordisk, Denmark); and purified human plasma FIX (Enzyme Research Laboratories, South Bend, IL)). Thrombin generation was initiated by dispensing into each well 20 µL of FluCa reagent (Thrombinoscope BV, Maastricht, The Netherlands) containing a fluorogenic substrate and HEPES-buffered $CaCl_2$ (100 mM). Fluorescence intensity was recorded at 37°C.

[0102] The parameters of the resulting thrombin generation curves were calculated using Thrombinoscope™ software (Thrombinoscope BV, Maastricht, The Netherlands) and thrombin calibrator to correct for inner filter and substrate consumption effects (Hemker, Pathophysiol. Haemost. Thromb., 33, 4-15 (2003)). For calculating the thrombin generating activity of certain peptide concentrations equivalent to the reference proteins (e.g., FVIII Immunate® reference standard, FEIBA reference standard), the thrombin amounts at the peak of each thrombin generation curve (peak thrombin, nM) were plotted against the standard concentrations, and fitted by a non-linear algorithm. Based on this calibration, FVIII Immunate, FIX, FEIBA or NovoSeven equivalent activities were calculated. Results for various peptides are provided in Figures 12-18 and 28-30. Representative results are provided in Table 8. (* denotes that FVIII deficient plasma was obtained from a different donor.)

TABLE 8

	% FVIII-equivalent activity in FVIII deficient plasma @ 10 µM peptide	**FEIBA-equivalent activity in FVIII inhibited plasma @ 10 µM peptide [mU/ml]**
JBT0120	37.4*	298
JBT0132	5.3	41
JBT0224	16.2	191
JBT0303	20.8	253

[0103] With reference to Table 8, JBT0120, JBT0132, JBT0224, and JBT0303 improved TFPI-dependent thrombin generation in FVIII-depleted plasma to levels exceeding 1% of the level of thrombin generation in plasma containing FVIII (% FVIII-equivalent activity). The tested peptides exhibited approximately 5%-40% FVIII-equivalent activity in FVIII-deficient plasma. JBT0120 and JBT0132 improved peak thrombin and peak time, dose dependently, as illustrated in Figures 11A and 11B.

[0104] Substitution mutants based on the amino acid sequence of JBT0293 also were tested in a plasma-based assay, as well as the FXa inhibition and extrinsic tenase inhibition assay described in Example 3.

Representative results are provided in Table 9.

TABLE 9

	Biacore K_D (nM)	FXa Inhibition EC_{50} (μM)	Extrinsic Tenase Inhibition EC_{50} (μM)	FVIII-equivalent activity (mU/ml) in Hem A plasma @ 1 μM peptide
JBT0047	47	0.7	1.4	45
JBT0293	97	1.3	2.9	48
JBT0303	34	1.2	NA	125
JBT0500	8.2	0.12	----	372
JBT0740	2.4	0.07	----	333
JBT1584	0.3	0.01	----	489

[0105] Additionally, JBT0477, which comprises the amino acid sequence of JBT0293 but for substitutions at amino acid positions 5 and 10 of the JBT0293 sequence, improves thrombin generation equivalent to 413 mU/ml of FVIII (at 1 μM of peptide) in FVIII-deficient plasma. Substitution mutation of JBT0293 resulted in highly optimized peptides with respect to affinity for TFPI and improved activity in FXa inhibition, extrinsic tenase inhibition, and plasma-based assays.

Example 5

[0106] The following example demonstrates that the peptides of the invention can be modified by the addition of moieties that enhance physicochemical or pharmacokinetic properties of the peptides. As illustrated below, the addition of 40 kDa PEG to peptides described herein dramatically improved the pharmacokinetic behavior of the peptides. The example also describes optimization of a TFPI-binding peptide, JBT1857, to reduce susceptibility to proteolysis.

[0107] Methods of conjugating chemical or biological moieties to peptides are known in the art. To add PEG (polyethylene glycol) to the peptides describe herein, a functional group (AOA = aminooxy acetate) was added to the N-terminus of the peptides for coupling to aldehydes and ketones. Alternatively, a cysteine was added to the C-terminal part of the peptide for coupling with maleimid (Hermanson, Bioconjugate Techniques, Academic Press (1996)). The peptides (JBT1586) AOA-FQSKGNVFVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 166) and (JBT1587) Ac-FQSKGNVFVDGYFERL-Aib-AKLC-NH2 (SEQ ID NO: 167) were used for N-terminal and C-terminal modification with PEG, respectively. AOA-FQSKGNVFVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 166) and Ac-FQSKGNVFVDGYFERL-Aib-AKLC-NH2 (SEQ ID NO: 167) were incubated with excess 40 kDa mPEG-Propionaldehyde (SUNBRIGHT ME-400AL2, NOF, Japan) and 40 kDa mPEG-maleimide (SUNBRIGHT ME-400MA, NOF, Japan), respectively. The resulting PEGylated peptides, JBT1852 and JBT1855, show similar affinities compared to the starting structure Ac-FQSKGNVFVDGYFERL-Aib-AKL-NH2 (JBT0740) (SEQ ID NO: 66).

[0108] The resulting PEGylated peptides demonstrated significantly increased plasma stability and prolonged plasma half-life in mice. Figure 31 illustrates the results from a pharmacokinetic analysis of the free peptide JBT0740 (Ac-FQSKGNVFVDGYFERL-Aib-AKL-NH2) (SEQ ID NO: 66) compared to the C-terminally PEGylated peptide JBT1855 (Ac-FQSKGNVFVDGYFERL-Aib-AKLC(PEG(40kD))-NH2) (SEQ ID NO: 252) following intravenous administration to mice. In contrast to the unPEGylated peptide, the PEGylated peptide is present at high concentrations in mouse plasma at 100 minutes post-administration. The unPEGylated peptide is rapidly cleared from the plasma. Figure 40 illustrates the results from a pharmacokinetic analysis of JBT1855 following subcutaneous injection. JBT1855 also strongly improved thrombin generation in the

assay described in Example 4 (Figure 41).

[0109] The JBT1852 and JBT1855 peptides also were characterized in the assays described in Examples 1-4 and compared to JBT0740 and other peptides in the JBT0047 family. Representative results are provided in Table 10 set forth below.

TABLE 10

	TFPI-1α Affinity (nM) **Biacore K_D**	**FXa Inhibition IC_{50}** (μM)	**FVIII-equivalent activity** (mU/ml) **in FVIII deficient plasma @ 1 μM peptide**	**Solubility** (mg/ml; PBS without Ca^{2+} and Mg^{2+})	**Plasma Stability** (half life in minutes)	
					mouse	human
JBT0717	1.1	0.05	421	0.97	24	>120
JBT0740	2.4	0.06	333	0.92	50	>120
JBT1584	0.3	0.02	486	2.66	40	>120
JBT1852	11.1	0.17	>1000	>1.00*	>120	>120
JBT1855	10.5	0.07	>1000	>1.00*	>120	>120
*formulated in 25 mm HEPES, pH 7.35, 175 mM NaCl						

[0110] The peptides listed in Table 10 also were assayed for interaction with the TFPI-2 anti-target, and generated signals too low for reliable affinity measurement. The data suggest that PEGylation does not ablate the inhibitory activity of the inventive peptides or negatively affect selectivity for TFPI-1.

Cell-based extrinsic tenase assay

[0111] The ability of the TFPI-binding peptides described above to restore extrinsic tenase complex-mediated conversion of FX to FXa also was determined using a cell-based extrinsic tenase assay. The cell-based extrinsic tenase assay also was employed to explore the influence of PEGylation on an exemplary TFPI-binding peptide of the invention, JBT0740. Human umbilical vein endothelial cells (HUVEC) were counted and seeded in complete growth medium in a 96-well plate (black flat with clear bottom) at a density of $1.5x10^4$ cells per well. Cells were grown overnight (for approximately 16 to 18 hours), washed twice with pre-warmed basal medium, stimulated with 1 ng/ml recombinant TNFα (Sigma Aldrich (Cat.No. T6674)) in 200 μl of basal medium for four hours at 37°C, and washed twice with 200 μl of pre warmed cell culture buffer. Buffer (50 μl) containing FVIIa (Enzyme Research Laboratories), TFPI-binding peptides (dissolved in either DMSO or Hepes buffered saline with or without 0.1% Tween-80), or αTFPI antibodies were applied to the cells and incubated for 20 minutes at 37°C, allowing FVIIa/TF complex formation and binding of TFPI antagonists to TFPI. After the incubation period, 50 μl of cell culture buffer containing FX and a FXa-specific substrate (Fluophen FXa (HYPHEN BioMed)) was applied, resulting in a final volume of 100 μl cell culture buffer mix on the cells. The final concentrations were: 39 pM FVIIa; 170 nM FX; 250 μM Fluophen FXa, and 2.5% DMSO (when peptides were dissolved in DMSO).

[0112] The 96 well plate was transferred to a pre-warmed (37°C) fluorescence reader for detecting FXa-specific fluorogenic substrate conversion by FXa, which is generated by the TF/FVIIa complex on the surface of stimulated HUVECs. Readings taken after nine minutes of incubation were used for calculation of the TFPI inhibitory effect of the TFPI-binding peptides or antibodies. The approximate percent inhibition of TFPI observed at various concentrations of the following peptides (belonging to the JBT0047 family) is set forth in Table 11: JBT0717 (Ac-FQSK-Nmg-NVFVDGYFERLRAKL-NH_2) (SEQ ID NO: 61), JBT0740 (Ac-

FQSKGNVFVDGYFERL-Aib-AKL-NH_2) (SEQ ID NO: 66), JBT1584 (Ac-FQSK-Nmg-NVFVDGYFERL-Aib-AKL-NH_2) (SEQ ID NO: 164), and JBT1857 (Ac-FQSKpNVHVDGYFERL-Aib-AKL-NH2) (SEQ ID NO: 178).

TABLE 11

	% TFPI inhibition				
	40 μM	8 μM	1.6 μM	0.32 μM	64 nM
JBT0717	60%	50%	32%	28%	20%
JBT0740	70%	39%	28%	14%	3%
JBT1584	73%	62%	51%	40%	29%
JBT1857	80%	57%	41%	35%	15%

[0113] PEGylated peptides also were tested using the cell-based extrinsic tenase assay. JBT0740 (SEQ ID NO: 66) was conjugated to a 1 kD PEG moiety at the N-terminus to produce JBT1853 or at the C-terminus to produce JBT1854. JBT1853 and JBT1854 inhibited TFPI by 20% or less depending on the amount of peptide used in the assay. JBT1855, which comprises a 40 kD PEG moiety at the C-terminus (parent peptide, JBT0740) performed better in the cell-based assay than JBT1852, which comprises a 40kD PEG moiety at the N-terminus. JBT1855 mediated 20-30% TFPI inhibition, while JBT1852 inhibited TFPI activity by 10% or less.

[0114] Peptides of the JBT0120 family, JBT0120, JBT0415, JBT0444, JBT1426, and JBT1837, also were tested in the cell-based extrinsic tenase assay and found to inhibit TFPI to a lesser degree compared to peptides of the JBT0047 family. The reduced or partial inhibitory activity may be desired in some embodiments of the invention. Similar to the peptides of the JBT0047 family, peptide optimization increased TFPI inhibitor activity of JBT0120 family peptides.

[0115] In the course of examining the stability and inhibitory activity of JBT1857, it was determined that the amino acid sequence of the peptide contained a protease cleavage site between Val9 and Asp10. Substitution of Tle at position 9 (generating JBT2431) and substitution of Pro at position 10 (generating JBT2432) blocked cleavage of the peptide and enhanced the plasma stability of the peptide by about three-fold from 27% (JBT1857) to 82% (JBT2431) and 76% (JBT2432). An additional putative cleavage site was identified between Gly11 and Tyr12. A G11a substitution (generating JBT2414) further improved the stability of the peptide to 100%. All stabilities were determined by quantitative ELISA after 24 hour incubation in human plasma.

[0116] The results described above demonstrate that optimization of the TFPI-binding peptides described herein utilizing non-conventional amino acids improved TFPI inhibition and plasma stability. Additionally, PEGylated peptides of the invention inhibit TFPI activity in a cell-based extrinsic tenase assay, with C-terminal PEGylated peptides performing better than N-terminal PEGylated peptides. The TFPI-binding peptides of the invention inhibit the activity of both free TFPI and cell-bound TFPI.

Example 6

[0117] The following example illustrates the ability of peptides described herein to reduce bleeding in an animal model.

[0118] Ten week old C57B1/6NCr1 mice were housed for two weeks prior to the study. Thirty minutes before the nail clip, the animals were administered (a) JBT1855 (10 mg/kg) intravenously (i.v.) via the tail vein or subcutaneously (s.c.) in the neck region, (b) anti-TFPI antibody (18 mg/kg; i.v.), or (c) vehicle (175 mM NaCl,

25 mM HEPES, pH 7.35; 10 ml/kg; i.v.). The animals were anaesthetized with 80 mg/kg pentobarbital ten minutes prior to the nail clip. To achieve bleeding, the nail of the small toe of the right hind paw was removed. The paw was submerged in a 0.9% NaCl solution for blood collection for a period of 60 minutes. Blood loss was quantified after lysis by spectrophotometry. The temperature was kept constant at 37°C over the course of the experiment. The results of the study are illustrated in Figure 42 and summarized in Table 12.

TABLE 12

	JBT1855 i.v.	**JBT1855 s.c.**	**α-TFPI i.v.**	**Vehicle** i.v.
Mean (in µl) (SD)	29.9 (71.4)	31.7 (31.5)	1.9 (1.4)	74.9 (74.6)
# of mice	12	12	12	12
p-value	0.07	0.04	0.001	

[0119] Intravenous or subcutaneous administration of JBT1855, a PEGylated peptide of the invention, reduced blood loss in mice compared to treatment with vehicle alone.

Example 7

[0120] The following example describes characterization of TFPI-peptide interactions via nuclear magnetic resonance and x-ray crystallography. In particular, the TFPI binding site of the antagonistic peptides JBT0303, JBT0122 and JBT0415; the residues of JBT0303, JBT0122 and JBT0415 interacting with TFPI160; and the secondary structure of complexed and free JBT0303, JBT0122 and JBT0415 were investigated at a molecular level using 2D ^{15}N-heteronuclear single quantum coherence (HSQC) spectra. The interaction of JBT1857 and KD1 of TFPI was examined using x-ray crystallography, and the residues of TFPI KD1 that mediate JBT1857 binding were mapped.

Identification of the binding site of JBT0303 on TFPI160

[0121] A ^{15}N-labelled preparation of TFPI160 was used for titration experiments of TFPI160 with JBT0303. HSQC spectra of a ~500 µM ^{15}N-TFPI160 sample without and with increasing amounts of peptide were recorded at 30°C on a Varian 600 MHz spectrometer. The peptide-protein interaction showed slow exchange behavior ($k_{ex} << \Delta\omega$), meaning that each TFPI residue results in a defined signal for the free protein and the protein-peptide complex. Unlike fast exchange behavior ($k_{ex} >> \Delta\omega$), where a mixture results in only one peak with averaged position according to the population of the species, slow exchange behavior does not allow tracking of the signals upon peptide binding. Thus, in order to locate the binding site, the shifted peaks of the TFPI160-JBT0303 complex needed to be assigned. This required the preparation of a sample of ^{13}C/^{15}N-TFPI160 and JBT0303.

[0122] Initially, a sample was prepared with 992 µM ^{13}C/^{15}N-TFPI160 and 1190 µM JBT0303. However, the NMR sample resulted in poor quality spectra which did not allow assignment of the complex. The sample gelled, likely due to the formation of high molecular weight aggregates. Thus, the acquired NMR data predominantly showed signals arising from the most flexible parts of the isotope-labeled TFPI160. Therefore, sample conditions were reinvestigated for further experiments. From a series of ^{15}N-HSQC experiments conducted on the TFPI160-JBT0303 complex, it was concluded that gel formation could be avoided by sample dilution and data acquisition at elevated temperature. The final concentration of ^{13}C/^{15}N-TFP1160 was 331 µM and that of JBT0303 was 397 µM. Spectra quality was improved. Due to the lower concentration

and reduced signal-to-noise ratio, assignment had to be performed based on HNCA, HNCO and HNCOCA experiments.

[0123] Except for four previously assigned residues, all residues that could be assigned in the apo-TFPI160 could be assigned in the TFPI160-JBT0303 complex. Assignment of some residues was ambiguous due to the lack of peaks in the 3D spectra. Furthermore, the peaks of three residues were only visible in the HSQC spectrum from the original titration experiment. However, all peaks in the vicinity where unambiguously assigned and, therefore, the assignment of these residues is likely to be correct.

[0124] Chemical shift changes of the HSQC signals of ^{15}N-TFPI160 bound to JBT0303 compared to free TFPI160 is illustrated in Figure 43. Residues undergoing the strongest chemical shift were exclusively on Kunitz domain 1. Chemical shifts of residues F25, F28, D32, A37, T48 and Y56 shifted the most (> 2 ppm). Residues I38, I46, F47 and F54 also shifted more than 1.5 ppm. It is unclear whether residues N-terminal of F25 are involved the interaction with JBT0303 because residues 20-24 are not assigned. L19 shows a change of chemical shift amounting to ~ 0.6 ppm. Thus, in contrast to previous beliefs that amino acids within residues 1-18 of TFPI are involved in peptide binding, the present data suggest that there is little, if any, peptide binding to the N-terminal tail of TFPI. A ribbon model of the secondary structure of TFPI protein illustrating regions of chemical shift changes of HSQC signals of TFPI160 bound to JBT0303 compared to free TFPI160 is set forth in Figure 44.

[0125] To more particularly identify the binding site of JBT0303 on TFPI160, the amide exchange rates of ^{15}N-TFPI160 and ^{15}N-TFPI160+JBT0303 were determined. The amide exchange experiment mainly detects changes in the environment of the peptide backbone by measuring H exchange of amide groups. The H_2O frequency is irradiated with a power high enough that it is not dissipated by relaxation, resulting in a complete saturation and suppression of the H_2O signal. A side effect of this method of H_2O signal suppression is that the suppression is transferred to exchangeable amide NHs which exchange with solvent (H/H exchange). The saturation transfer is dependent on the H/H exchange rate which is semiquantitative. The effect is reduced for more protected NH groups (i.e., unprotected NHs are attenuated more than protected NHs). If a protected NH lies in proximity to H-alphas of a ligand, a higher exchange rate is observed compared to the apo form. Similarly, H exchanges can be mediated by the OH groups of Ser, Thr or Tyr.

[0126] HSQC spectra without and with water suppression of apo ^{15}N-TFPI160 and the ^{15}N-TFPI160-JBT0303 complex were recorded. The relative exchange rate of each residue of TFPI160 was determined by calculating the ratio of the peak intensities in the HSQC spectra with and without water suppression. A comparison of the data sets of ^{15}N-TFPI160 and ^{15}N-TFPI160+JBT0303 revealed that TFPI residues 25, 26, 36, 62, 63, 127, 132 and 152 exhibited greater than 10% decreased amide exchange rate in the complex, whereas residues 29, 30, 42, 45, 49, 50, 56, 66 and 98 exhibited more than 10% increased exchange rate.

[0127] Constraints derived from the amide exchange experiment were included for the calculation of refined HADDOCK models: (a) torsion angles are taken from the calculations of TALOS for K4, K5, V7, F8, Y12-A18 of JBT0303 (chemical shift experiments); (b) residues of KD1 with chemical shift changes of more than 1.5 ppm are involved in binding JBT0303: F25, F28, D32, A37, I38, I46, F47, T48, F54 and Y56 (chemical shift experiments); (c) the hydrophobic side of the amphipathic helix of JBT0303 is bound to KD1: Y12 or L16 or L20 of JBT0303 bind to D32 or A37 or 138 or F54 or Y56 of KD1 (chemical shift experiments); (d) R15 or K19 of JBT0303 bind to D31 or D32 or E60 of KD1 (chemical shift experiments); (e) F8 or V9 of JBT0303 bind to F25 or F28 of KD1 (chemical shift experiments); (f) Y12 or F13 of JBT0303 bind to 146 or F47 or T48 of KD1 (chemical shift experiments); (g) Q2 of JBT0303 binds to Y56 of KD1 (chemical shift experiments); (h) F1 of JBT0303 binds to M39 or F66 of KD1 (amide exchange experiments); (i) S3 or K4 or K5 of JBT0303 bind to F66 (amide exchange experiments); (j) V7 or F8 or V9 of JBT0303 bind to F25 or C26 or N62 or Q63

of KD1 (amide exchange experiments); (k) V9 or D10 or G11 or R15 of JBT0303 bind to F28 or K29 or A30 of KD1 (amide exchange experiments); (1) Y12 or F13 of JBT0303 bind to N45 of KD1 (amide exchange experiments); (m) Y12 or F13 or E14 or R15 bind to R49 or Q50 of KD1 (amide exchange experiments); and (n) L20 of JBT0303 binds to K36 of KD1 (amide exchange experiments). The data converged to essentially one model of the KD1+JBT0303 complex.

Identification of the binding site of JBT0122 on TFPI160

[0128] As with the $^{13}C/^{15}N$-TFPI160 + JBT0303 complex, the $^{13}C/^{15}N$-TFPI160 + JBT0122 NMR sample resulted in spectra of poor quality due to the formation of a gel. The concentration of 723 µM $^{13}C/^{15}N$-TFPI160 + JBT0122 lead to formation of higher order aggregates. The sample was diluted to 361.5 µM and spectra recorded at 37°C, resulting in improved spectra quality. HNCO, HNCA and HNCOCA spectra were acquired. Except for five residues, all of the previously assigned peaks of apo-TFPI160 could be assigned in the TFPI160-JBT0122 complex. Residues undergoing the strongest chemical shift changes and likely to interact with the peptide often did not result in peaks in the 3D spectra. Peaks in the linker region between Kunitz domain 1 (KD1) and Kunitz domain 2 (KD2), however, also exhibited low intensities. Hence, the assignment of these peaks is ambiguous. Some peaks were only visible in the HSQC of the original titration experiment. Their assignment was in most cases certain, as the peaks overlapped in the TFPI160 and the TFPI160 + JBT0122 HSCQ spectra.

[0129] Chemical shift changes of the HSQC signals of ^{15}N-TFPI160 bound to JBT0122 compared to free TFPI160 is illustrated in Figure 45. Significant chemical shift changes were exclusively found for residues of KD2. In general, the extent of the chemical shift changes caused by binding of JBT0122 to TFPI160 was less pronounced than that of JBT0303. Residues with the strongest perturbation of chemical shift were F96, G128, G129, G132, N133 and N136. C97, E101, T111, F114, N135 and F137 were perturbed, exhibiting chemical shift changes of more than 0.5 ppm. A ribbon model of the secondary structure of TFPI protein illustrating regions of chemical shift changes of HSQC signals of TFPI160 bound to JBT0122 compared to free TFPI160 is set forth in Figure 46.

Identification residues of JBT0122 that interact with TFPI160

[0130] For the sequential backbone signal assignment of JBT0122, $^{13}C/^{15}N$-labelled peptide was produced recombinantly. Briefly, the peptide was expressed as a fusion protein with thioredoxin in *E. coli.* $^{13}C/^{15}N$-labelled peptide was prepared using M9 medium containing 3.0 g/l ^{13}C-glucose and 1.0 g/l $^{15}NH_4Cl$. The fusion protein was affinity purified using a Ni-chelating column and a poly-histidine tag. The peptide was cleaved by thrombin. The thioredoxin/his-tag and thrombin was removed using a Ni-chelating column and a benzamidine column, respectively. The peptide was then purified by reverse phase chromatography. Purity, integrity, and identity were verified by SDS-PAGE, RP-HPLC and mass spectrometry. Recombinant JBT0122 was named JBT0788 and had two additional residues at its N-terminus, glycine and serine, which represent the remains of the thrombin cleavage site.

[0131] The assignment of JBT0788 was done on the basis of HSQC, HNCACB, HNCA, HNCO and HNN spectra recorded at 10°C on a Varian 600 MHz spectrometer and assigned using the SPARKY software. The temperature was reduced compared to NMR experiments with TFPI160 to improve spectra quality. From the recorded spectra, the carbonyl carbon (C), the alpha carbon (CA), the beta carbon (CB), the amide proton (H), and the amide nitrogen (N) of most residues were assigned. The assignment for residues H13 and R17

was ambiguous. An HNCOCA led to an unambiguous assignment for these residues.

[0132] An assignment table for JBT0788 is provided in Figure 47. Two sets of signals for residues 4-12 were observed in the spectra of JBT0788. Considering that the primary structure of JBT0788 is not compromised, the two sets of signals likely result from a cis/trans isomerization of the peptide bond between F6 and P7. A ratio of 76:24 was determined for major:minor conformation based on the intensities of the corresponding signals in the HSQC spectrum. As judged from the Cα shift of the proline, the major conformation is likely trans, as its Cα value of 63.16 ppm is higher than of the minor conformation (62.49 ppm).

[0133] One purpose of the assignment was to extract the secondary structure of the peptide from Cα chemical shifts. Cα chemical shifts are influenced by the angles ϕ and ψ and, thus, by the secondary structure of the peptide. In β-strands, Cα are generally shifted to lower ppm; in α-helices, Cα are generally shifted to higher ppm. By subtracting the measured Cα value from a tabulated random coil value, negative values are calculated for residues in β-strands and positive values for residues in α-helices. Thus, a batch of consecutive negative values indicates a β-strand while a batch of consecutive positive values indicates an α-helix.

[0134] JBT0788 exhibited a broad patch of increased Cα values ($\Delta\delta(C\alpha) = C\alpha_{measured} - C\alpha_{random\ coil}$) indicating an α-helix comprising residues 8 to 26. Δδ(Cα) values for stable α-helices within tertiary structures of native proteins are typically between 3-4 ppm. Δδ(Cα) values of the α-helix of JBT0788 rise up to about 1.7 ppm, indicating more flexibility than an average helix within a protein. Another feature of JBT0788 is the proline at position 7, directly N-terminal to the α-helix, which fits well as α-helices in proteins are frequently terminated by a proline at the N-terminus. Residue 6 has a strong negative value, which is caused by the neighboring proline known to force its N-terminal neighbor into a β-strand-like conformation. The strong positive value of C-terminal residue 31 is also typical for residues without a C-terminal neighbor. The peptide bond between F6 and P7 in JBT0788 adopts two conformations, a trans (76%) and a cis conformation (24%). The conformation at this position impacts the conformation of the consecutive residues. In the trans isoform, the α-helix starts immediately after P7; the α-helix of the cis isoform does not start until residue L12. A ribbon model illustrating the secondary structure of free JBT0788 is set forth in Figure 48.

[0135] The chemical shifts within JBT0788 can also be employed to calculate the torsion angles using TALOS software. TALOS is a database system for empirical prediction of ϕ and ψ backbone torsion angles using a combination of five kinds (HA, CA, CB, CO, N) of chemical shift assignments for a given protein or peptide sequence. The TALOS approach is an extension of the observation that many kinds of secondary chemical shifts (i.e., differences between chemical shifts and their corresponding random coil values) are correlated with aspects of protein secondary structure. The goal of TALOS is to use secondary shift and sequence information in order to make quantitative predictions for the protein backbone angles ϕ and ψ, and to provide a measure of the uncertainties in these predictions. TALOS uses the secondary shifts of a given residue to predict ϕ and ψ angles for that residue. TALOS also includes the information from the next and previous residues when making predictions for a given residue. The idea behind TALOS is that if one can find a triplet of residues in a protein of known structure with similar secondary shifts and sequence to a triplet in a target protein, then the ϕ and ψ angles in the known structure will be useful predictors for the angles in the target. In practice, TALOS searches a database for the 10 best matches to a given triplet in the target protein.

[0136] In order to assign the HSQC spectrum of JBT0788 complexed with TFPI160, a sample consisting of 400 μM $^{13}C/^{15}N$-JBT0788 and 400 μM TFPI160 was prepared. As with previous NMR samples of peptide and TFPI160, the sample gelled. The sample was diluted and the pellet dissolved in deuterated DMSO, resulting in a final concentration of ~300 μM $^{13}C/^{15}N$-JBT0788 + TFPI160 and 5% DMSO. Measurements were performed at 40°C. This improved the quality of the acquired spectra. Experiments were acquired in the

TROSY mode to account for the relaxation properties of a partially aggregated sample. Cryo-probe technology on the Varian 600 MHz spectrometer was employed due to the low concentration of the protein-peptide complex in the sample. The resulting data quality was sufficient to obtain the backbone shifts of JBT0788 when utilizing the cryo-probe technology and acquiring the triple-resonance experiments in duplicate. The assignment of JBT0788 in complex with TFPI160 was performed on the basis of HNCA, HNCOCA and HNCO spectra. From the recorded spectra, the carbonyl carbon (CO), the alpha carbon (CA), the amide proton (H), and the amide nitrogen (N) of most residues were assigned. An assignment table for JBT0788 complexed to TFPI160 is provided in Figure 49.

[0137] A feature of apo-JBT0788 was the presence of two sets of signals for amino acid residues 4-12, likely resulting from a cis/trans isomerization of the peptide bond between F6 and P7. In the JBT0788-TFPI160 complex, only one set of peaks is observed, implying that only one of the conformations binds to TFPI160. Apo-JBT0788 also exhibited a broad patch of increased Cα values ($\Delta\delta(C\alpha) = C\alpha_{measured} - C\alpha_{random\ coil}$ = positive) indicating an α-helix reaching from residue 8 to residue 26. As mentioned above, Δδ(Cα) values for stable α-helices within tertiary structures of native proteins are typically between 3-4 ppm. Δδ(Cα) values of the α-helix of apo-JBT0788 increase to about 1.7 ppm, indicating more flexibility than an average helix within a protein. When complexed with TFPI, residues 8 to 26 exhibited values of between 3-5 ppm, indicating the formation of a stable α-helix or helices. A ribbon model illustrating the secondary structure of JBT0788 when complexed with TFPI160 is set forth in Figure 50. Large chemical shift changes within JBT0788 caused by binding with TFPI160 are evenly distributed over the length of the peptide. Residues undergoing the strongest perturbation of chemical shift were residues S5, A9, Q11, Y28, and K29 with more than 4 ppm. Residues Y3, A4, V10, L12, S15, M21, A22, L23, and A24 were perturbed by more than 3 ppm.

Identification of residues of JBT0303 that interact with TFPI160

[0138] JBT0303 was produced recombinantly using the same procedure as described above for JBT0122 and isotope-labeled with ^{13}C and ^{15}N. The recombinant JBT0303 was named JBT0616 and had an additional glycine and serine at its N-terminus. The assignment of JBT0616 was performed on the basis of HSQC, HNCACB and HNN spectra, which were recorded at 10°C on a Varian 500 MHz spectrometer and assigned using SPARKY software. The quality of the spectra of JBT0616 was better than that of JBT0788, although the experimental conditions with respect to buffer, temperature, NMR tube, and NMR parameters were identical. The alpha carbon (CA), the beta carbon (CB), the amide proton (H), and the amide nitrogen (N) of most residues were assigned. The assignment was mainly based on the less sensitive but more informative HNCACB instead of the HNCA. In combination with the HNN spectrum, this resulted in an unambiguous assignment of all JBT0303 derived residues.

[0139] An assignment table for JBT0616 is provided in Figure 51. The secondary structure was extracted from Cα chemical shifts and determined by TALOS using the assignments of H, CA, CB, CO and N. Like JBT0788, JBT0616 exhibited a patch of positive Δδ(Cα) values indicative of α-helical conformation. The helix was located at the C-terminal part of the peptide and comprised residues 10-18. As for JBT0788, Δδ(Cα) values up to about 1.8 ppm were calculated, qualifying this helix as relatively stable for such a short peptide. A ribbon model illustrating the secondary structure of JBT0616 is set forth in Figure 52. The strong positive value of the C-terminal residue 20 is, like residue 31 in JBT0788, typical for residues without a C-terminal neighbor. The N-terminal residues 1-9 of JBT0616 exhibited slightly positive Aγ(Cα) values, suggesting a preference for an α-helical structure.

[0140] The assignment of JBT0616 in complex with TFPI160 was performed using a $^{13}C/^{15}N$-labelled peptide sample with an excess of unlabelled TFPI160. HSQC, HNCA, HNCOCA, and HNCO spectra were recorded on a Varian 800 MHz spectrometer and assigned using the SPARKY software. The spectra were recorded at

30°C. Using these spectra, the alpha carbon (CA), the beta carbon (CO), the amide proton (H), and the amide nitrogen (N) of most residues were assigned, as set forth in the table in Figure 53. The secondary structure of JBT0616 in complex with TFPI160 was extracted from Cα chemical shifts and calculated by TALOS. Like the free peptide, JBT0616 in complex with TFPI160 exhibited a C-terminal patch of positive Δδ(Cα) values indicative of α-helical conformation. The stability of the α-helix is increased upon complex formation. This finding suggests that the C-terminal region of JBT0616 is the core binding motif. The Δδ(Cα) values for the N-terminal residues also changed, but to a lesser extent. The secondary structure of JBT0616 when complexed with TFPI is illustrated in the ribbon model in Figure 54.

[0141] The most significant changes of chemical shifts upon complex formation were observed for residues Q2, K5, F8, V9 and A18 of JBT0616 with more than 7 ppm. Residues F13, R17, K19 and L20 also were perturbed and demonstrated chemical shift changes of more than 4 ppm. The strong chemical shift changes of residues at the N-terminus indicated that it is not only the amphipathic C-terminal α-helix which drives binding of the peptide to TFPI160.

[0142] Results from the NMR experiments in combination with analysis of JBT0477 substitutions were used to create a model of KD1 in complex with JBT0303 using HADDOCK (High Ambiguity Driven protein-protein DOCKing) software. HADDOCK is an information-driven flexible docking approach for the modeling of biomolecular complexes. HADDOCK distinguishes itself from *ab-initio* docking methods in the fact that it encodes information from identified or predicted protein interfaces in ambiguous interaction restraints (AIRs) to drive the docking process. Identification of the binding site on TFPI160 and the peptides as revealed by chemical shift data, the torsion angles of the peptides as determined by the software TALOS, and the substitution analysis of JBT0477 provide the restraints for the calculation of the models.

[0143] For the calculation of the KD1-JBT0303 HADDOCK models, the following restraints were employed: (a) torsion angles were taken from the calculations of TALOS for K4, K5, V7, F8, Y12-A18 of JBT0303; (b) residues of KD1 with chemical shift changes of more than 1.5 ppm are involved in binding to JBT0303: F25, F28, D32, A37, I38, I46, F47, T48, F54 and Y56; (c) the hydrophobic side of the amphipathic helix of JBT0303 is bound to KD1: Y12 or L16 or L20 of JBT0303 bind to D32 or A37 or 138 or F54 or Y56 of KD1; (d) R15 or K19 of JBT0303 binds to D31 or D32 or E60 of KD1; (e) F8 or V9 of JBT0303 binds to F25 or F28 of KD1; (f) Y12 or F13 of JBT0303 binds to 146 or F47 or T48 of KD1; and (g) Q2 of JBT0303 binds to Y56 of KD1. The Q2 JBT0303 - Y56 KD1 interaction also was taken as a restraint for model calculation.

[0144] Strong chemical shift changes were observed for K5 of JBT0303 upon complex formation. For the remaining residues of JBT0303 considered to drive the peptide-protein interaction, the models are in good agreement with the data. The model of KD1-JBT0303 with the lowest energy places F8 of JBT0303 in proximity to F25 and F28 of TFPI, explaining the observed chemical shift changes and the data from the substitution analysis. V9 of JBT0303 interacts with the hydrophobic patch of the KD1 including F54. Y12, F13, L16 and L20 of JBT0303 also face the hydrophobic patch of the KD1. The proximity of Y12 to F28, I46, T48 of F13 to F47, T48, of L16 to F54 and of L20 to A37, I38 causes the observed perturbations of NMR chemical shift of those residues in the complex; the conservation of Y12 and L16 may be due to the extensive interactions of these residues with the protein. K19 of JBT0303 is in a position allowing interaction with D32 of KD1. The role of R15 of JBT0303 seems to be an interaction with the hydrophobic patch of KD1 as well as with D32. Moreover, the model explains why a negatively charged aspartate is preferred at position 10 of JBT0303; it can interact with the positively charged K29 of KD1. A glycine at position 11 of JBT0303 is present due to the steric and conformational restraints at this position. A HADDOCK model of KD1 (TFPI residues 22-79 comprising KD1) in complex with JBT0303 is provided in Figure 55.

Models of JBT0740 and JBT1857 bound to KD1

[0145] Peptides JBT0740 and JBT1857 (FQSK-dP-NBHBDGYFERL-Aib-AKL (SEQ ID NO: 178)), both derivatives of JBT0303, demonstrate significantly enhanced EC_{50} values in the FXa-TFPI inhibition assay (0.11 µM and 0.0023 µM, respectively) and lower K_d's as determined by Biacore. Models of JBT0740 and JBT1857 in complex with TFPI KD1 (residues 22-79 of TFPI160) were calculated by HADDOCK using similar constraints as for JBT0303: (a) the constraints for the torsion angles of residues 4 and 5 of JBT0740 and JBT1857 were amended in order to take account of the substitutions at position 5 of the JBT0303 derivatives; (b) torsion angles were taken from the calculations of TALOS for V7, F8, Y12-A18 of JBT0303 and, in contrast to JBT0303, no fixed values for Phi and Psi were given for K4 and for NmetG5/dP5; (c) NmetG5 and dP5 are in the cis conformation; (d) residues of KD1 with chemical shift changes of more than 1.5 ppm are involved in binding to JBT0303: F25, F28, D32, A37, I38, I46, F47, T48, F54 and Y56; (e) the hydrophobic side of the amphipathic helix of JBT0303 is bound to KD1; (f) Y12 or L16 or L20 of JBT0303 bind to D32 or A37 or 138 or F54 or Y56 of KD1; (g) residues R15 or K19 of JBT0303 bind to D31 or D32 or E60 of KD1; (h) residues F8 or V9 of JBT0303 bind to F25 or F28 of KD1; (i) residues Y12 or F13 of JBT0303 bind to 146 or F47 or T48 of KD1; and (j) residue Q2 of JBT0303 binds to Y56 of KD1.

[0146] The energetically most favorable HADDOCK models of JBT0740 and JBT1857 illustrated a different mode of binding compared to JBT0303. The most obvious differences were in the region of residues 5 to 11. Less dramatic deviations were observed at the N-terminus and the C-terminus of the peptides. However, the different binding of the termini also might contribute to the optimized binding of the JBT0303 derivatives to TFPI.

X-ray Crystal Structure of JBT1857 bound to KD1

[0147] The crystal structure of KD1 in complex with a KD1 binding peptide, JBT1857, was determined. TFPI was recombinantly expressed in *E.coli* and oxidatively refolded from inclusion bodies. TFPI amino acids 1-150 comprising a thrombin cleavage site within the TFPI linker sequence joining KD1 and KD2 (TFPI1-150-Thrombin (MADSEEDEEHTIITDTELPPLKLMHSF**CAFKADDGPCKAIMKRFFFNIFTRQCEEFI YGGCEGNQNRFESLEECKKMC**TRDNANR*LVPRGS*QQEKPDF**CFLEEDPGICRGYI TRYFYNNQTKQCERFKYGGCLGNMNNFETLEECKNIC**EDG (SEQ ID NO: 4235)) was cloned into an *E. coli* expression vector (pET19b). The TFPI 1-150-Thrombin sequence comprises two amino acids at the N-terminus that are artifacts of recombinant expression, and are not part of the wild-type TFPI amino acid sequence. The sequences encoding Kunitz domain 1 and 2 are bolded. *E. coli* (BL21(DE3) pLysS) was cultivated in MagicMedia™ and TFPI 1-150-Thrombin was expressed as insoluble inclusion bodies. Inclusion bodies were harvested by lysis of *E. coli* by incubation with BugBuster Master Mix and purified upon washing with 50 mM Tris/HCl pH 8, 0.1% Tween 20. Inclusion bodies were dissolved in 8M urea, 50 mM Tris/HCl pH 8.0 and TFPI 1-150-Thrombin was reduced upon addition of 20 mM DTT. Oxidative refolding was performed by rapid 1/10 dilution into a buffer containing 50 mM Tris/HCl pH 10 and 1.1 mM oxidized Glutathion, followed by excessive dialysis against 20 mM Tris/HCl pH 7. Refolded TFPI1-150-Thrombin was purified by a sequential purification protocol using a Q Sepharose FF anion exchange and a peptide affinity (JBT131) media. Purified TFPI1-150-TFPI was proteolytically digested by incubation with thrombin (1U thrombin / mg TFPI1-150-Thrombin, cleavage site, LVPR/GS) resulting in the generation of Nterm KD1-Thrombin (MADSEEDEEHTIITDTELPPLKLMHSF**CAFKADDGPCKAIMKRFFFNIFTRQCEEFI GGCEGNQNRFESLEECKKMC**TRDNANR*LVPR* (SEQ ID NO: 4236)) and KD2-Thrombin (*GS*QQEKPDF**CFLEEDPGICRGYITRYFYNNQTKQCERFKYGGCLGNMNNFETLE** ECKNICEDG (SEQ ID NO: 4237)). Nterm KD1-Thrombin was purified from the digestion mixture using benzamidin sepharose for removal of thrombin, followed by a JBT131 peptide affinity column. Purified Nterm KD1-Thrombin was used for complex formation with JBT1857 and further crystallization.

[0148] The antagonistic peptide, JBT1857, was prepared by solid phase synthesis. Successful co-

crystallization of equimolar complexes was obtained under 100 mM MES pH 6.5, 20% PEG 4000, 600 mM NaCl. Crystals diffracted to better than 2.5 Å resolution, albeit with some non-merohedral twinning. Diffraction data were processed with iMosflm and SCALA from the CCP4 program package, revealing a monoclinic crystal form with unit cells dimensions of a= 113.67 Å, b= 69.32 Å, c= 42.37 Å, α= 90.0°, β= 92.97°, γ= 90.0°, spacegroup C2 (Leslie, Acta Crystallogr D Biol Crystallogr, 62(Pt 1), 48-57 (2006); Evans, Acta Crystallogr D Biol Crystallogr, 62(Pt 1), 72-82 (2006)). Self-rotation calculations indicated an approximately two-fold non-crystallographic symmetry. Consistent herewith, two molecules were localized in the asymmetric unit related by a 170° rotation. The Patterson search was carried out by using the program PHASER and a structure ensemble of the available Kunitz domain 2 crystal structures as search model (McCoy et al., J Appl Crystallogr, 40(Pt 4), 658-674 (2007)). The unit cell contained approximately 64% solvent. Non-crystallographic electron density averaging and model building and model refinement was carried out with Coot, Refmac, MAIN and CNS programs. The current model was completely defined for both copies of the JBT1857 peptide and the interaction with the protein with current R=0.257, Rfree=0.298, deviation from ideal geometry rms(bond) = 0.008 Å, rms(angle) = 1.8°.

[0149] *JBT1857 structure:* The structure of JBT1857 can be segmented into (i) the N-terminal anchor consisting of acetylated $Phe1_{AP}$-$Gln2_{AP}$; (ii) an Ω-shaped loop comprising $Ser3_{AP}$-$Asn6_{AP}$; (iii) an intermediate segment built from $Val7_{AP}$ and $His8_{AP}$; (iv) a tight glycine-loop containing $Val9_{AP}$-$Gly11_{AP}$; and (v) the C-terminal α-helix comprising $Tyr12_{AP}$-$Leu20_{AP}$. As used herein, the subscript $_{AP}$ indicates the sequence numbering in the "antagonistic peptide" JBT1857. The conformation of the α-helix is stabilized by a non-natural α-methyl alanine positioned at the center of the helix (position 17_{AP}); a C-terminal amide that completes the 1-4 hydrogen bonding pattern of the α-helix; and a stacked cluster by the aromatic side chains of $His8_{AP}$, $Tyr12_{AP}$ and $Phe13_{AP}$. These effects cooperate to stabilize the C-terminal α-helix spontaneously in solution, consistent with circular dichroism data on the peptide. The observed aromatic side chain stacking ($His8_{AP}$, $Tyr12_{AP}$, $Phe13_{AP}$) enforces a tight turn that can be only accomplished by glycine at position 11_{AP}. This structural constraint is reflected by dramatic losses in binding affinity upon replacement of $Gly11_{AP}$ by any other amino acid. The conformation of the N-terminal loop segment is partly stabilized by a D-proline, known to induce a tight turn conformation, and a 1-4 hydrogen bond by the carbonyl oxygen of $Ser3_{AP}$ with the amide nitrogen of $Asn6_{AP}$. All ring side chains ($Tyr1_{AP}$, $Pro5_{AP}$, $His8_{AP}$, $Tyr12_{AP}$, $Phe13_{AP}$) point towards the same direction, enabling them to interact with the KD1 domain of TFPI.

[0150] *Interaction of JBT1857 and KD1*: The interactions between JBT1857 and KD1 were determined. Hydrophobic contacts are interactions having an intermolecular distance of ≤ 4 Å, while hydrogen bonds have a distance between 2.6 - 3.2 Å. $Phe1_{AP}$ interacts non-specifically with TFPI making contacts with Phe2 and Ala27. In contrast, $Gln2_{AP}$ contacts a deeply buried pocket of TFPI and makes hydrophobic interactions with Phe28, Lys29, Ile46 and Phe47. Moreover, the amide group of $Gln2_{AP}$ forms three H-bonds with Phe28-CO, Phe44-CO and Ile46-NH. The Ω-loop of JBT1857, comprising $Ser3_{AP}$-$Asn6_{AP}$, mediates rather limited hydrophobic interactions with the protein; $Ser3_{AP}$, $Pro5_{AP}$ and $Asn6_{AP}$ interact with Lys29 and Phe47. $Val7_{AP}$ of JBT1857's intermediate segment also binds to Lys29 and Phe47. $His8_{AP}$ mainly contributes intramolecular aromatic stacking interactions with $Tyr12_{AP}$ and partly with $Phel3_{AP}$, and exhibits a hydrophobic interaction with Ala30 of TFPI. Similarly, the glycine-loop $Val9_{AP}$-$Gly11_{AP}$ contributes few contacts with the Kunitz domain. $Val9_{AP}$ interacts directly with KD1 by forming a hydrogen bond with the carbonyl group of Ala30 and a hydrophobic interaction with Asp32. $Tyr12_{AP}$ mediates a hydrogen bond via its hydroxyl group with the amide nitrogen of Ile55 and a hydrophobic interaction with Asp30. $Leu16_{AP}$ is part of a hydrophobic contact with Ile55. Beside the largely hydrophobic interactions of the C-terminal helix of the peptide with the protein, there are electrostatic interactions between $Arg15_{AP}$ and Asp32. Furthermore, $Lys19_{AP}$ contributes to binding with TFPI by forming a hydrogen bond to the carbonyl group of Ala37 and contacts with Lys36 and

Ile38. The TFPI contact surface has an overall hydrophobic character with some charted hot spots, and a driving force of complex formation with JBT1857 is the steric surface complementarity.

[0151] This example describes characterization of the secondary structure of exemplary peptides of the invention and correlates the structure with inhibitory function of the peptides. The example also identifies the TFPI amino acid residues that interact with JBT1857, a TFPI-binding peptide that inhibits TFPI activity.

Example 8

[0152] The following example describes additional TFPI-binding peptides modified by the addition of moieties that enhance physicochemical or pharmacokinetic properties of the peptides. The example further describes a method for assessing clot formation in whole blood using rotation thromboelastography.

[0153] JBT1857 (JBT0047 peptide family) was conjugated to different PEG moieties, and the binding affinity and TFPI inhibitory activity of the PEGylated peptides were examined. JBT1857 was modified by addition of a C-terminal cysteine to produce JBT2315 (Ac-FQSKpNVHVDGYFERL-Aib-AKLC-NH2 (SEQ ID NO: 4077)), which was conjugated at the C-terminus with linear maleimide PEG moieties of increasing size: 5 kD, 12 kD, 20 kD, 30 kD, and 40 kD, using the methods described in Example 5. The resulting PEGylated peptides were designated as follows:

TABLE 13

Peptide	PEG (kD)	Sequence	SEQ ID NO
JBT1857	---	Ac-FQSKpNVHVDGYFERL-Aib-AKL-NH2	4020
JBT2317	---	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(NEM)-NH2	4078
JBT2325	5.3	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(PEG)-NH2	4086
JBT2326	12.1	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(PEG)-NH2	4087
JBT2327	21.0	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(PEG)-NH2	4088
JBT2328	29.1	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(PEG)-NH2	4089
JBT2329	41.5	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(PEG)-NH2	4090

Stability, binding affinity, and TFPI-inhibitory activity of PEGylated peptides

[0154] The PEGylated peptides demonstrated significantly increased plasma stability in mouse and human plasma. The peptides were added to samples of mouse or human plasma, and the percentage of the initial amount of peptide remaining in plasma 24 hours after the addition was measured by IC_{50} ELISA on Maxisorp plates coated with 0.05 mg/ml TFPI (2.26 nM tracer peptide JBT2271). Less than approximately 10% of the initial amount of JBT1857 and JBT2317 remained in plasma, while 40% or more of the initial amount of the PEGylated TFPI-binding peptides remained after 24 hours. Approximately 60% or more of JBT2327 and JBT2329 was detected. PEGylated peptides also are significantly more stable in human plasma compared to unmodified peptides. Approximately 60% or more of PEGylated peptide remained after 24 hours. The unmodified peptides were more stable in human plasma than mouse plasma; about 20% or more of the initial amount remained after 24 hours of incubation.

[0155] The PEGylated peptides also were characterized in the assays described in Examples 1-4 and compared to JBT1857. Representative results are provided in Table 14 set forth below. The thrombin generation assay was performed as described in Example 4, and the results are provided as EC_{50}, corresponding to the concentration of peptide which improved peak thrombin (nM) half maximal.

TABLE 14

	PEG (kD)	Biacore K_D (nM)	Competition ELISA IC_{50} (nM)	FXa Inhibition EC_{50} (nM)	Extrinsic Tenase Inhibition EC_{50} (nM)	Thrombin generation in human FVIII-inhibited plasma EC_{50} (nM)
JBT1857		0.061	3.0	3.7	6.9	
JBT2317		0.054	2.9	3.8	7.8	88
JBT2325	5.3	0.71	6.6	10.7	10.7	35
JBT2326	12.1	1.1	9.3	9.3	9.3	34
JBT2327	21.0	1.3	10.9	7.2	7.2	24
JBT2328	29.1	1.6	12.3	6.0	6.0	19
JBT2329	41.5	1.1	12.6	6.0	12.8	19
*Competition ELISA performed with tracer JBT2271 (1 nM) and 0.05 µg/ml TFPI in the coating buffer.						

[0156] Addition of the C-terminal cysteine blocked with NEM did not significantly influence the binding affinity of JBT2317 or the activity of the peptide in the FXa inhibition, extrinsic tenase assay, or thrombin generation assay compared to JBT1857. PEG size did not significantly impact the TFPI-binding peptides' ability to restore activity of FXa in the presence of TFPI-1. In the extrinsic tenase assay of Example 3, inhibitory activity increased with higher molecular weight PEG moieties up to 20 kD PEG. Activity did not further improve for 30 kD or 40 kD PEG moieties. In the thrombin generation assay of Example 4 using human plasma, EC_{50} decreased with PEG size, and maximal inhibition of TFPI (as measured by peak FIIa (nM)) increased with PEG size. In mouse plasma, attachment of 40 kD PEG to a TFPI binding peptide increased maximal inhibition of TFPI.

[0157] The ability of PEGylated TFPI-binding peptides to restore extrinsic tenase complex activity for converting FX to FXa also was determined using a cell-based extrinsic tenase assay using the method of Example 5. Addition of the C-terminal cysteine blocked with NEM did not significantly influence the activity of JBT2317 in the cell-based extrinsic tenase assay compared to JBT1857. Conjugation of PEG moieties (5 kD, 20 kD, 30 kD, or 40 kD) to JBT2317 increased TFPI inhibitory activity by 5-20%.

Rotational thromboelastography

[0158] Continuous visco-elastic assessment of human whole blood clot formation and firmness was performed by rotation thromboelastography with whole blood preparations in the presence or absence of peptides. Blood samples from a healthy individual were drawn into citrated Sarstedt Mono S (0.106 M or 3.2% (w/v) Na-citrate) (5 ml), mixing one part of citrate with nine parts blood, using a 21 gauge butterfly needle. A portion of the blood samples was incubated with high titer, heat-inactivated anti-human FVIII antiserum raised in goat (3876 BU/ml; Baxter BioScience, Vienna, Austria) resulting in 51 BU/mL. Test samples were prepared by dissolving quantities of peptides in either DMSO or HEPES buffered saline (with or without 0.1% Tween 80).

[0159] Recordings were made using a ROTEM thromboelastography coagulation analyzer (Pentapharm, Munich, Germany) at 37°C. Briefly, blood is added into a disposable cuvette in a heated cuvette holder. A disposable pin (sensor) is fixed on the tip of a rotating axis. The axis is guided by a high precision ball bearing system and rotates back and forth. The axis is connected with a spring for the measurement of elasticity. The exact position of the axis is detected by the reflection of light on a small mirror on the axis. The loss of elasticity when the sample clots leads to a change in the rotation of the axis. The data obtained are computer analyzed and visualized in a thromboelastogram. The thromboelastogram shows elasticity (mm) versus time (s). An elasticity of approximately zero is observed before clot formation begins. Mirror image traces above and below the zero line indicate the effect of clot formation on the rotation of the axis.

[0160] Before starting each experiment, the citrated whole blood was mixed with corn trypsin inhibitor (CTI) (Hematologic Technologies, Inc., Essex Junction, VT, USA) providing a final concentration 62 µg/mL for specific inhibition of FXIIa, in order to inhibit FXIIa-mediated contact activation. The analytical set-up was as follows: to 20 µL of test sample or control, 300 µL of pre-warmed (37°C) CTI treated citrated whole blood was added, followed by 20 µL of a 1:15 dilution of TF PRP reagent containing recombinant human tissue factor (rTF, 3 pM) (TS40, Thrombinoscope BV, Maastricht, The Netherlands). Coagulation was initiated by the addition of 20 µL 200 mM $CaCl_2$ (star-TEM®, Pentapharm, Munich, Germany) and recordings were allowed to proceed for at least 120 min. The final concentration of rTF in the assay was 11 or 44 fM.

[0161] The thromboelastographic parameters of clotting time (CT), clot formation time (CFT) and maximum clot firmness (MCF) were recorded in accordance with the manufacturer's instructions. CT is defined as the time from the start of measurement to the start of clot formation. CFT is defined as the time from the start of clot formation until an amplitude of 20 mm is reached. MCF is the maximum difference in amplitude between the two traces during the assay. The first derivative of the data of the thromboelastogram are plotted to obtain a graph of velocity (mm/s) against time (s). From this graph, the maximum velocity (maxV) is determined. The time at which the maximum velocity is obtained (maxV-t) is also determined.

[0162] Exemplary results are illustrated in Figures 56 and 57. JBT1857 and JBT2317 restored coagulation parameters in Hem A blood. PEGylated (40 kD) TFPI-binding peptide JBT2329 also restored prolonged coagulation parameters in Hem A blood, as illustrated in Figure 57. PEGylation of JBT2317 reduces clot time and clot formation time.

Nail Clip Study

[0163] The effect of JBT2329 on blood loss in naive mice also was studied. C57BL6 mice were administered vehicle, 1 mg/kg JBT2329, or 0.1 mg/kg JBT2329 (N=19 or 20 for each group) intravenously 30 minutes prior

to nail clip at 10 ml/kg. Animals were anaesthetized 10 minutes before nail clip with 80 mg/kg pentobarbital (i.p.). At time=0 minutes, the nail of the small toe of the right hind paw was cut just before the nail bed. The paw was transferred to a vial prefilled with 0.9% NaCl solution. Samples of blood were collected for analysis during the first 30 minutes following the nail clip and the next 30 minutes thereafter, and mean collected volume for the groups was calculated and compared. Mean blood loss in vehicle treated mice was about 30.5 µl over the first 30 minutes, 52.1 µl over the second 30 minute period, resulting in about 82.6 µl of blood loss over 60 minutes. In contrast, administration of 0.1 mg/kg JBT2329 reduced blood loss by about 50% over the first 30 minutes (16.0 µl) and about 64% over the second 30 minute period (18.7 µl), resulting in about a 60% reduction of total blood loss over 60 minutes (34.7 µl) compared to vehicle-treated mice. Increasing the dose of JBT2329 to 1.0 mg/kg further reduced blood loss by at least about 10%; 12.2 µl was collected over the first 30 minutes, 10.6 µl was collected over the second 30 minute period, resulting in 22.8 µl collected over the entire 60 minute collection period. JBT2329 also efficiently reduced bleeding when administered subcutaneously compared to vehicle-treated naive mice; subcutaneous injection of 10 mg/kg JBT2329 reduced blood loss during the 60 minutes following nail clip by approximately 58% compared to vehicle-treated subjects.

[0164] The results described above were generated using a JBT1857 derivative comprising a linear PEG moiety attached to the C-terminus of the peptide and a JBT1586 derivative comprising a PEG moiety at the N-terminus. Peptides comprising an alternate conjugation site or alternative chemical moiety also were generated. A 40 kD linear PEG moiety was conjugated to residue 14 of JBT1857 to generate JBT2404. The linear 40 kD PEG moiety of JBT2329 was replaced with a 40 kD branched PEG moiety to generate JBT2401. JBT1857 also was modified to comprise K(Ttds-Maleimidopropionyl) (JBT2374) at the C-terminus. JBT2374 was used to generate JBT2410, an HSA conjugate of JBT2374. JBT2375, a K(AOA)-comprising derivative of JBT1857, was used to couple PSA aldehyde to the peptide JBT1857, resulting in JBT2430. JBT2401, JBT2404, JBT2410 and JBT2430 were characterized using the assays described above. Representative results are summarized in Table 15:

TABLE 15

	Biacore K_D (nM)	**ELISA affinity EC_{50}** (nM)	**FXa Inhibition EC_{50}** (nM)	**Human Plasma Stability, 24 hour** (%)	**Thrombin generation in human FVIII-inhibited plasma EC_{50} (nM)**
JBT2329	< 1	12.6	6	67.3	1.4
JBT2401	< 1	22.4	7.7	85.1	1.4
JBT2404	< 1	18.2	13.4	96.7	1.7
JBT2410	n.d.	5.1	4.7	65.7	1.8
JBT2430	n.d.	5.6	9.0	135.6	

[0165] This example demonstrates that an exemplary TFPI-binding peptide of the invention, JBT1857, is a potent inhibitor of TFPI and can be functionalized and conjugated with PEG without loss of activity. PEGylation increased TFPI-inhibitory activity in several functional assays. Surprisingly, peptides conjugated to higher weight PEG moieties demonstrated enhanced TFPI inhibitory activity. JBT2329, comprising a 40 kD linear PEG moiety, significantly reduced blood loss in a clinically-relevant animal model. PEG conjugation within the amino acid sequence of JBT1857, use of a branched PEG moiety, and attachment of HSA and PSA did not destroy the activity of the peptide.

Example 9

[0166] The following example describes the characterization of two TFPI-binding peptides of the invention,

JBT1837 and JBT1857. JBT1837 (Ac-SYYKWH[CAMRDMKGTMTC]VWVKF-NH) (SEQ ID NO: 1044) is a cyclic peptide of the JBT0120 family that binds KD1 and KD2 of TFPI. JBT1857 (Ac-FQSKpNVHVDGYFERL-Aib-AKL-NH2) (SEQ ID NO: 178) is a linear peptide of the JBT0047 family that binds KD1 of TFPI. The affinity and TFPI-inhibitory activity of JBT1837 and JBT1857 were examined using the assays described in Examples 1-4, the results of which are summarized in Table 16.

TABLE 16

	Biacore K_D (nM)	**ELISA affinity EC_{50}** (nM)	**FXa Inhibition; R&D TFPI/f1TFPI EC_{50}** (nM)	**Extrinsic Tenase Inhibition; R&D TFPI/f1TFPI EC_{50} (μM)**	**Thrombin generation in human FVIII-inhibited plasma EC_{50} (nM)**	**Thrombin generation in human FIX-deficient plasma EC_{50} (nM)**
JBT1837	< 1	4.8	3.2/5.9	0.5/0.9	10	16
JBT1857	< 1	3.0	3.7/21.9	6.9/13.6	69	51

[0167] Affinity of the peptides to human TFPI measured via BiaCore was less than 1 nM. Affinity measured by ELISA (IC_{50}) was 4.8 nM for JBT1837 and 2.5 nM for JBT1857. JBT1837 dissociated from human TFPI more slowly than JBT1857 (i.e., JBT1837 remained bound to human TFPI for a longer period of time compared to JBT1857). A FXa inhibition assay was performed using both full length human TFPI ("flTFPI") and truncated human TFPI (254 amino acids "R&D TFPI") (0.1 nM FXa, 0.5 nM TFPI, 0.25% DMSO). Activity of the truncated TFPI was fully inhibited by both JBT1837 and JBT1857 at 0.5 nM TFPI, while full length TFPI was inhibited 85% and 95% by JBT1857 and JBT1837, respectively. At higher concentrations of flTFPI (e.g., 10 nM flTFPI), JBT1837 fully inhibited TFPI activity, while JBT1857 partially inhibited TFPI activity. EC_{50}'s also were higher when flTFPI was used in the FXa inhibition study.

[0168] In the extrinsic tenase assay, about 85% of truncated TFPI was inhibited by both peptides. Full length TFPI activity was inhibited about 56% and 48% by JBT1837 and JBT1857, respectively. Surprisingly, in the cell-based extrinsic tenase assay, JBT1837 inhibited the activity cell-associated TFPI by about 50% whereas JBT1857 almost fully inhibited cell-bound TFPI activity. In the plasma-based functional assay, JBT1837 inhibited TFPI more efficiently than JBT1857 in human FVIII-inhibited plasma and FIX-deficient plasma. JBT1837 corrected blood coagulation parameters in FVIII-inhibited blood in the ROTEM assay described in Example 8. JBT1857 also positively impacted blood coagulation parameters, but performed less efficiently than JBT1837 in the assay.

[0169] This example compared the affinity and TFPI-inhibitory activity of cyclic and linear TFPI-binding peptides that target different regions of the TFPI protein. JBT1837 (a cyclic peptide belonging to family JBT0120) and JBT1857 (a linear peptide belonging to family JBT0047) efficiently bind human TFPI with affinities less than 1 nM and are potent inhibitors. FXa-TFPI interaction is fully blocked at low TFPI concentrations by both peptides, while TFPI inhibition by JBT1857 is reduced in the presence of higher concentrations of TFPI. Both peptides partially inhibit the activity of full-length TFPI in the extrinsic tenase assay, and JBT1857 inhibits TFPI activity to a greater degree in the cell-based extrinsic tenase assay compared to JBT1837. Compared to JBT1857, JBT1837 more efficiently inhibits TFPI in FVIII-deficient plasma. Both peptides improve coagulation parameters of FVIII-inhibited human whole blood by reducing clot time, while JBT1857 improves clot formation velocity to a lesser degree compared to JBT1837.

Example 10

[0170] This example illustrates the *in vivo* activity of TFPI-binding peptides of the invention in a clinically-

relevant animal model. As described below, an exemplary TFPI-binding peptide significantly reduced blood loss in an animal when administered with suboptimal doses of FVIII and FIX.

[0171] JBT2329, a PEGylated (40 kD) TFPI-binding peptide (JBT0047 family) that cross-reacts with human and murine TFPI, was tested in tail-tip bleeding model in FVIII knock-out mice and FIX knock-out mice. FVIII knock-out mice closely mirror the condition of hemophilia A patients, and the tail-tip bleeding model is widely used in research to assess efficacy of drugs by measuring, e.g., bleeding time, blood loss or survival. ADVATE, a commercially available rFVIII, served as a reference, and ADVATE buffer-treated animals served as negative controls. Each group contained 16 FVIII knock-out mice (8 female + 8 male). JBT2329 (1 mg/kg or 0.1 mg/kg) or anti-TFPI antibody (maTFPI; 18 mg/kg) was administered 30 minutes before the tail-tip was cut. ADVATE (10 IU/kg or 50 IU mg/kg) or ADVATE buffer was administered five minutes before the tail was cut off. Test and control substances were administered as an intravenous bolus via a lateral tail vein injection. Animals were anaesthetized by an intraperitoneal injection of 100 mg/kg ketamine and 10 mg/kg xylazine. Approximately 10 minutes later, 2 mm of the tail-tip was cut off. The tail-tips were placed in warm saline (approximately 37 °C) and blood was collected over an observation period of 60 minutes. The amount of blood was determined gravimetrically. At the end of the observation period of 60 minutes the animals were humanely killed by cervical dislocation before recovery from anesthesia.

[0172] Median total blood loss in buffer-treated animals was 930 mg. Median total blood loss in subjects treated with murine anti-TFPI antibody (maTFPI) was 724 mg. The reduction in median total blood loss was more pronounced when the subjects were administered maTFPI with ADVATE. A combination of maTFPI + 10 IU/kg ADVATE led to a median total blood loss of 136 mg, animals treated with maTFPI + 50 IU/kg ADVATE experienced a median total blood loss of 13 mg. Median blood losses of animals treated with either 10 or 50 IU/kg ADVATE alone experienced median blood loss of 798 and 364 mg, respectively. The superiority of the combination treatment of maTFPI + ADVATE over ADVATE alone was statistically shown for maTFPI + 50 IU/kg ADVATE versus 50 IU/kg ADVATE ($p = 0.0010$). Although not statistically significantly superior, blood loss in animals treated with maTFPI + 10 IU/kg ADVATE was distinctively lower than in animals treated with 10 IU/kg ADVATE alone.

[0173] Efficacy, defined as statistically significant superiority over buffer at a 2.5% level, was shown for JBT2329 dosed at 1 mg/kg in combination with 10 and 50 IU/kg ADVATE and dosed at 0.1 mg/kg in combination with 50 IU/kg ADVATE ($p < 0.0004$). Animals treated with JBT2329 in combination with ADVATE showed a clinically-relevant reduction in blood loss, although the results were not statistically significant ($p \geq 0.0506$). Administration of 1 mg/kg JBT2329 without ADVATE did not reduce median total blood loss over that observed in buffer-treated animals (930 mg).

[0174] JBT2329 also was tested in a FIX knock-out tail-tip bleed mouse model, which is a clinically-relevant model for hemophilia B human patients. The methodology was substantially similar to that described above with respect to the FVIII knock-out model. Instead of ADVATE, a recombinant FIX (rFIX) served as a reference. Median total blood loss in buffer-treated animals was 935 mg. Median total blood loss in animals treated with a murine anti-TFPI antibody (maTFPI) was 774 mg. Median total blood loss was reduced further when the animals received combined treatment of maTFPI and rFIX. A combination of maTFPI + 10 IU/kg rFIX led to a median total blood loss of 25 mg, while animals treated with maTFPI + 50 IU/kg rFIX exhibited a median total blood loss of 10 mg. Median blood loss of animals treated with either 10 or 25 IU/kg rFIX alone experienced a median blood loss of 888 and 774 mg, respectively.

[0175] Efficacy, defined as statistically significant superiority over buffer at a 2.5% level, was shown for JBT2329 when dosed at 1mg/kg in combination with 10 IU/kg rFIX and at 0.1 mg/kg in combination with 10 IU/kg rFIX. The superiority of JBT2329 in combination with rFIX over administration of rFIX alone was observed ($p < 0.0172$), while treatment with 1 mg/kg JBT2329 alone did not lead to a significant reduction in median total blood loss compared with buffer-treated animals ($p = 0.321$).

[0176] In summary, JBT2329 promoted a clinically-relevant reduction of blood loss when co-administered with suboptimal doses of FVIII and rFIX at all doses tested. Furthermore, intravenous administration of JBT2329 was well tolerated in all subjects across all treatment groups without any signs of acute toxicity.

Example 11

[0177] The TFPI-binding peptides described herein are suitable for detecting TFPI in a sample, such as biological sample. This example describes a method for detecting TFPI using the inventive peptides in an ELISA-like assay format.

[0178] The peptide sequence of JBT1857 was N-terminally modified by the addition of a biotinyl-Ttds moiety to generate JBT2271 (Biotinyl-Ttds-FQSKpNVHVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 4033)). A 96-well microtiter plate (Maxisorp, Nunc) was coated with 50 µ1per well coating buffer (15 mM Na_2CO_3, 35 mM $NaHCO_3$, pH 9.3) containing a range of TFPI concentrations (0-3 µg/ml, human recombinant TFPI, R&D Systems) for 1 hour at room temperature. The plate was washed three times with 350 µl/well wash buffer (175 mM NaCl, 5 mM $CaCl_2$, 25 mM HEPES, 0.1% Tween 80, pH 7.35). The plate was then blocked with 100 µl blocking buffer (2% yeast extract, 175 mM NaCl, 5 mM $CaCl_2$, 25 mM HEPES, 0.1% Tween 80, pH 7.35) for 1 hour at room temperature. The plate was then washed three times with 350 µl wash buffer. Fifty µl of differently concentrated JBT2271 solutions in wash buffer (100-0 nM) were added to each well. The plate was incubated for 1 hour and washed three times with 350 µl wash buffer. To each well, 50 µl streptavidin-horseradish peroxidase conjugate (R&D Systems, 1:200 in wash buffer) is added. After an incubation period of 1 hour at room temperature, the plate was washed three times with wash buffer. Fifty µl TMB solution (SeramunBlau fast, Seramun) was added to each well. After a 1.5 minute incubation at room temperature, the reaction was stopped by adding 50 µl 1 M H_2SO_4 per well. Absorbance was measured in a photometer (Molecular Devices Spectramax M5) at 450 and 620 nm.

[0179] JBT2271 allowed detection of as little as 4.1×10^{-14} mole of TFPI per well. The results of the assay described above illustrate that the inventive peptides are powerful tools for identifying and/or quantifying TFPI in a sample.

Example 12

[0180] This example describes conditions for an exemplary k_{off} assay for characterizing TFPI-binding peptides.

[0181] Wells of a microtiter plate (96 wells, Maxisorp, Nunc) are coated with 1.6 nM TFPI in coating buffer (15 mM Na_2CO_3, 35 mM $NaHCO_3$, pH 9.3) for two hours at room temperature. The plate is then washed three times with 350 µl wash buffer (175 mM NaCl, 5 mM $CaCl_2$, 25 mM HEPES, 0.1% Tween 80, pH 7.35), and wells are blocked with 100 µl blocking buffer (2% yeast extract, 175 mM NaCl, 5 mM $CaCl_2$, 25 mM HEPES, 0.1% Tween 80, pH 7.35). If an incubation period of 24 hours is employed, the wells are blocked for at least one hour. Control wells used for a 15 minute incubation period are blocked for an additional 23.5 hours.

[0182] For a 24 hour incubation period, the wells are washed three times with 350 µl wash buffer and are incubated with 50 µl test peptide in wash buffer. The concentration of test peptide depends on the individual

IC_{90} concentration determined in, e.g., the TFPI IC_{50} ELISA assay described herein. The TFPI-coated wells are exposed to test peptide for approximately 15 minutes. The wells are subsequently washed three times with 350 µl wash buffer and 50 µl tracer peptide (competitor) is added. An exemplary tracer peptide is JBT2271 (1.13 nM in wash buffer). Control wells (maximum signal) are incubated with tracer only. Blank wells lacking TFPI are incubated with tracer only. Addition of the tracer peptide commences the 24 hour incubation period.

[0183] A 15 minute incubation period is employed as a control if the IC_{90} concentration of the test peptide leads to a 90% reduction of the maximum signal. Wells blocked for an additional 23.5 hours are washed three times with 350 µl wash buffer to remove the blocking buffer. Subsequently, 50 µl analyte in wash buffer is added and the wells are incubated for 15 min. The concentration of test peptide utilized depends on the peptide's IC_{90} concentration determined using, e.g., a TFPI IC_{50} ELISA assay. The 15 minute incubation is followed by three washes with 350 µl wash buffer and addition of 50 µl tracer peptide. Control wells (maximum signal) are incubated with tracer only. Blank wells lacking TFPI also are incubated with tracer only.

[0184] The plate is washed three times with 350 µl wash buffer, and 50 µl streptavidin-horseradish peroxidase conjugate (R&D Systems, 1:200 in wash buffer) is added to each well. After an incubation period of one hour at room temperature, the plate is washed three times with wash buffer. TMB solution (50 µl per well; SeramunBlau fast, Seramun) is added. After a 1.5 minute incubation at room temperature, the reaction is stopped by the addition of 50 µl 1 M H_2SO_4 per well. Absorbance is measured using a photometer (Spectramax M5, Molecular Devices) at 450 and 620 nm. The assay results are presented as a percentage of the corrected optical density (OD450-OD620) of wells exposed to test peptide and tracer peptide in relation to TFPI-coated wells exposed only to tracer.

Example 13

[0185] TFPI inhibits FVIIa/TF activity by binding to FVIIa via Kunitz domain 1 (KD1). This example describes an exemplary method for evaluating the influence of TFPI-binding peptides on TFPI's inhibition of FVIIa/TF.

[0186] Kinetic measurements were performed in 25 mM HEPES, 175 mM NaCl, 5 mM $CaCl_2$, 0.1% BSA, pH 7.3 at 25°C in 96-well microtiter plates. Twenty µl soluble tissue factor (residues 33-251; Creative Biomart) and 20 µl FVIIa (ERL) at final concentrations of 100 nM and 5 nM, respectively, were mixed and incubated for 15 minutes. Twenty µl of TFPI-binding peptide in varying final concentrations (0-2 µM) were added to the mixture and incubated for a further 15 minutes. In order to measure the residual activity of the FVIIa/sTF complex, the reaction mixture was incubated for 60 minutes with 20 µl TFPI (200 nM). The reaction was initiated by the addition of a chromogenic substrate, Chromozym-tPA (Roche) (1 mM). The change in absorbance at 405 nm was monitored by using a Labsystems iEMS ELISA Reader for 30 minutes. FVIIa/sTF activity measured in the absence of TFPI was considered "100% activity" in the context of the assay. By plotting peptide concentration against residual activity, EC_{50} values were determined.

[0187] JBT1857 and JBT1837 were screened against TFPI160, TFPI1-150-Thrombin, NTermKD1, KD1, and KD2 (negative control). JBT1857 demonstrated an EC_{50} of approximately 0.21-0.23 µM for TFPI160, TFPI1-150-Thrombin, NTermKD1, and KD1. JBT1837, which binds KD1 and KD2, demonstrated an EC_{50} of approximately 0.17-0.19 µM for TFPI160 and TFPI1-150-Thrombin, while activity in assays involving NTermKD1 and KD1 was approximately background.

[0188] The results described above demonstrate that TFPI-binding peptides efficiently inhibit TFPI-FVIIa/TF interaction. JBT1857 efficiently inhibited TFPI fragments containing KD1 as a minimal functional entity. Thus,

this enzymatic assay confirms X-ray crystallographic data placing the binding site of JBT1857 within KD1. JBT1837 inhibited TFPI fragments containing the first two Kunitz domains, suggesting that the JBT1837 binding site(s) are located within KD1-linker-KD2 region of TFPI. A combination of Kunitz domains and fragments of a thrombin cleaved TFPI (1-150) did not restore inhibitory activity of JBT1837 in the chromogenic assay. The enzymatic assay described herein is a suitable surrogate for detecting binding of a TFPI-binding peptide (or a test compound) to TFPI, and is useful for examining the TFPI-inhibitory effect of TFPI-binding compounds.

Example 14

[0189] This example describes the influence of PEG and HSA conjugation on exemplary TFPI-binding peptides *in vivo.*

[0190] For pharmacokinetic analysis, C57B16 mice were treated with various TFPI-binding peptides conjugated to different molecular weight PEGs and HSA. The dose of the peptide-PEG and peptide-HSA conjugates was normalized to 1 mg/kg (peptide content). Normalization assures comparability between the conjugates of different molecular weight. The peptide conjugates were dissolved in 175 mM NaCl, 25 mM HEPES pH 7.35 and administered intravenously via the tail vein or subcutaneously in the neck region. Blood draws were taken from three animals (retro bulbar) and collected in heparinized vials at several time points following administration. The samples were centrifuged, and the peptide-conjugate content in plasma was quantified by ELISA.

[0191] Figure 63 illustrates the concentration of PEGylated TFPI-peptides detected in plasma at several time points following administration, and Table 17 provides detailed information about the terminal half life and bioavailability of JBT2325-JBT2329, JBT2401, JBT2404 and JBT2410.

TABLE 17

	JBT2325	JBT2326	JBT2327	JBT2328	JBT2329	JBT2401	JBT2404	JBT2410
HE_λ_z [h] (intravenous)	0.16	0.35	4.2	10.1	19.8	20.7	12.3	7.8
Bioavailability [%] (s.c.)	58.2	76.0	89.7	52.0	73.3	58.4	59.3	46.6

[0192] JBT2329, JBT2401 and JBT2404 are peptides conjugated to 40 kDa linear PEG (JBT2329 and JBT2404) or 40 kDa branched PEG (JBT2401). The 40 kDa conjugates exhibited a longer terminal half-life (HL_λ_z) compared to peptides conjugated to smaller PEGs following intravenous administration. The area under curve (AUC) of the concentration-time curve resulting from subcutaneous administration of the peptides was compared to the AUC generated following intravenous administration to calculate the bioavailability of the peptides. Results are shown in Table 16. The data demonstrate that TFPI-binding peptide conjugation to higher molecular weight molecules allows a subcutaneous bioavailability of more than 30%.

[0193] Figures 64A-C illustrate the pharmacokinetic profile of JBT2401, JBT2404, and JBT2410 resulting from subcutaneous and intravenous administration of the peptides to mice. JBT2404 comprises a PEG conjugated to cysteine in position X4014 relative to formula (XI). JBT2401 comprises a branched PEG, and JBT2410 is conjugated to HSA. Figure 64A demonstrates that fusion of a higher molecular weight molecule to a TFPI-binding peptide at an internal position increases half life. Half life also is increased if using branched PEG (JBT2401) and HSA, which increased the *in vivo* half life of JBT2410 compared to conjugates having smaller-sized PEGs (e.g., JBT2325) or free peptide (see Figure 31).

[0194] This example illustrates that the *in vivo* properties of various peptides described herein can be improved by conjugation with higher molecular weight molecules (like PEG) and/or with nFcR ligands (like HSA).

Example 15

[0195] The following example describes the characterization of JBT1837's interaction with KD1-KD2 of TFPI, alone and in the presence of JBT1857, via X-ray crystallography.

Native-PAGE

[0196] Native-PAGE was performed to confirm that TFPI-binding peptides that bind different regions of TFPI could bind TFPI KD1-KD2 simultaneously. Fifteen percent polyacrylamide-gels (without SDS) were used for the native gel-electrophoresis on samples comprising (1) a TFPI fragment comprising KD1-KD2, (2) KD1-KD2 and JBT1857, (3) KD1-KD2 and JBT1837, and (4) KD1-KD2 with both JBT1857 and JBT1837. Glycerine and Ponceau red were added to the protein samples. The anode buffer was obtained by dissolving 25 mM imidazole in 1 liter water and adjusting the pH to 7 with HCl. Clear cathode buffer was prepared by dissolving 7.5 mM imidazole and 50 mM tricine in one liter of water. Water was added to each solution to reach a final volume of two liters. Serva Blue G (0.02% w/v final) was added to two liters of cathode buffer to generate blue stock 10X cathode buffer (blue stock 10X). Gels were run for 30 minutes at 80 V, then for about three hours at 250 V. The gels were stained and de-stained as described in Chakavarti et al., J Vis Exp., 12;(16) (2008).

[0197] KD1-KD2 was visualized in the gel as a single band. Samples comprising K1-K2 in combination JBT1857 and/or JBT1837 resulted in a shift of the initial KD1-KD2 band to a higher molecular weight to charge ratio. After mixing of KD1-KD2 and JBT1837, partial precipitation was observed, indicating a decrease in solubility after complex formation. JBT1857 shifted the complete KD1-KD2 band, while JBT1837 shifted approximately 1/3 of the initial band. Additionally, in the presence of both JBT1837 and JBT1857, the KD1-KD2 band shifts to the highest molecular weight to charge ratio, indicating the formation of a ternary complex consisting of KD1-KD2, JBT1837 and JBT1857. Thus, JBT1837 and JBT1857 can bind TFPI simultaneously.

Materials and Methods - Interaction of JBT1837 with KD1-KD2

[0198] Crystal Growth: Initial crystals were formed at 20°C by mixing 0.4 µl of a 1:1 complex (1.5 mg/ml; 10 mM Tris/8.0, 100 mM NaCl) with 0.2 µl precipitant (20% w/v PEG 6000, 50 mM Imidazole pH 8.0). Crystals were reproduced at 20°C by mixing 1 µl protein with 0.5 µl precipitant (20% w/v PEG 6000, 50 mM Imidazole pH 8.0). A cryoprotectant (30% w/v PEG 6000, 34% glycerol) was added to the crystal drop (final concentration of 25% glycerol) to protect resulting crystals from disorder and ice formation during flash cooling with liquid nitrogen.

[0199] Data Processing: Diffraction data were collected from 87.26Å to 1.95Å and evaluated using the MOSFLM and iMOSFLM packages (Leslie, (2006) Acta Crystallogr D Biol Crystallogr 62, 48-57), followed by merging and scaling using CCP4 suite (McCoy et al., (2007) J Appl Crystallogr 40, 658-674). Crystals belong to the orthorhombic space group $P\,2_12_12_1$ with unit cell dimensions of a= 42.35 Å, b= 44.13 Å, c= 174.53 Å, $\alpha = \beta = \gamma = 90.0°$ and 2 complexes in the asymmetric unit. The Patterson search was carried out using the PHASER program (McCoy, *supra*) and two search ensembles, a crystal structure of KD2 (1TFX) (Burgering

et al. (1997) J Mol Biol 269, 395-407) and a previously solved KD1 crystal structure. The unit cell contained approximately 54% solvent. Non-crystallographic electron density averaging, model building and model refinement was carried out with the programs Coot and Refmac (McCoy, *supra;* Murshudov et al, (1997) Acta Crystallogr D Biol Crystallogr 53, 240-55). The current model is completely defined for both copies of the JBT1837 peptide (23AA) and the interaction with the protein with current R=0.213, R_{free}=0.245, deviation from ideal geometry rms(bond) = 0.0098 Å, rms(angle) = 1.28°.

[0200] Analysis of the Interaction Surface: The interaction surface of KD1-KD2 and JBT1837 was analyzed with PDBePISA. The impact of single amino acids was weighted by the total reduction of their solvent accessible area after complex formation (buried solvent accessible area BSAA) and considered by generating a 2D-interaction-matrix.

Materials and Methods - Interaction of KD1-KD2 with JBT1837 and JBT1857

[0201] Crystal Growth: Initial crystals where formed at 20°C by mixing 0.4 µl of a 1:1:1 complex (1.5 mg/ml; 10 mM Tris/8.0, 100 mM NaCl) with 0.2 µl precipitant (2M $(NH_4)_2SO_4$, 5% PEG 400, 100 mM MES pH 6.5). Crystals were reproduced and optimized at 20°C by mixing 1 µl protein with 0.5 µl precipitant (1.5 M $(NH_4)_2SO_4$, 5% PEG 400, 100 mM MES pH 6.5). A cryoprotectant (2M $(NH_4)_2SO_4$, 25% glycerol) was added stepwise to the crystal drop (final concentration of 2.3M $(NH_4)_2SO_4$ and 20% glycerol) to protect the resulting crystals from disorders and ice formation during flash cooling with liquid nitrogen. The drop was covered with cryo-oil and crystals were harvested.

[0202] Data Processing: Diffraction data were collected from 74.3Å to 2.7Å and evaluated using the MOSFLM and iMOSFLM packages, followed by merging and scaling using CCP4 suite (Leslie, (2006) Acta Crystallogr D Biol Crystallogr 62, 48-57; Acta Crystallogr D Biol Crystallogr 50, 760-3 (1994)). Crystals belong to the tetragonal space group $P\,4_32_12$ with unit cell dimensions of a= 79.07 Å, b= 79.07 Å, c= 218.17 Å, α= β= γ= 90.0°.

X-Ray Crystallography Results

[0203] The KD1-KD2 structure is defined in the electron density from L21 through E148; while chemically present, residues D149 and G150 are conformationally disordered in two crystallographically-independent copies of the molecule.

[0204] Both domains show a Kunitz-type structure. Only ~ 1/3 of the structure is engaged in secondary structure elements; these are two short α-helical elements at S24-A27(KD1)/D95-F98(KD2) (α1/α3) and L69-M75/L140-E148 (α2/α4) and a two-stranded β sheet comprising M39-N45/I110-N116 (β1/β3) and R49-I55/K120-K126 (β2/β4). These elements form the topological framework that is stabilized by the three canonical disulfide bonds involving C26-C76, C35-C59, and C51-C72 in KD1 and C95-C147, C106-C130, and C122-C143 in KD2.

[0205] This is the first structure of TFPI consisting of KD1, KD2 and their linker elucidated by X-ray crystallography. Remarkably, JBT1837 locks KD1-KD2 in a distinct conformational state in which both Kunitz-domains are related via a two-fold symmetry. Additionally the conformation of TFPI is intrinsically stabilized by two turns, a β-turn (tβ) from T77-A81 and a γ-turn (tγ) from Q90-K93. tβ is stabilized by three hydrogen bonds (O K74-N N82, O N80-Nζ K74, O N82-Nε H23) and leads to a shortening of α2 in KD1 by two residues compared to homolog Kunitz-domains and the crystal structure of KD1 alone, tγ is stabilized by four

hydrogen bonds (O T88-N D95, O Q90-N K93, O K93-N Q90, Oγ T88-Oδ D95).

[0206] The 23mer peptide JBT1837 assumes a β-hairpin like structure which can be segmented into (i) the pin, a two-stranded β sheet comprising $Y2_{AP}$-$A8_{AP}$ and $T17_{AP}$-$F23_{AP}$; (ii) and the needle eye, a β-turn comprising $D11_{AP}$-$T15_{AP}$. (The subscript $_{AP}$ indicates the sequence numbering in the antagonistic peptide (JBT1837).) The β-sheet is stabilized by a disulfide bridge ($C7_{AP}$ and $C18_{AP}$) and a hydrophobic zipper comprising the side chains of $Y3_{AP}$, $W5_{AP}$ and $W20_{AP}$.

[0207] Analyzing the interaction between KD1-KD2 and JBT1837 with the PISA server resulted in a total interaction surface of 1340$Å^2$. More than 2/3 of the interaction surface consists of a hydrophobic anchor in JBT1837 interacting with residues spread over TFPI, including KD1, KD2, and the linker, as illustrated by an interaction matrix, a summary of which is provided in Table 18, and Figure 74.

TABLE 18

JBT 1837	BSAA		KD1	LINKER	KD2
	$Å^2$	%			
Ser1	50.7	35	C59, E60		
Tyr2	83.9	72	R41, Y56		
Tyr3	11.7	12	R65		
Lys4	48.0	72	E67		E142
Trp5	143.5	96	Q63, M75	184	
His6	83.9	89			M134, F137, E142, I146
Cys7	26.8	65	E71		
Ala8	36.9	89			M134, N136
Met9	95.0	63		R83	L131, G132, N133, M134
Arg10	52.5	31			L131
Asp11	41.2	61		R83	L131
Met12	55.2	28			C106, C130
Lys13	34.7	19		N80	
Gly14	17.2	57			
Thr15	75.7	94	K74	N80, N82, R83	
Met16	33.26	42		N82, R83	
Thr17	71.5	89		R83, 185	M134
Cys18	60.9	100	M75	N82, R83, I84, I85	
Val19	74.65	99		185, T87	F96, M134
Trp20	89.4	68	Q63, R65	T87	
Val 21	54.25	99		T87	I146
Lys22	37.1	27			
Phe23	108.5	64			N145, I146

[0208] In addition to the hydrophobic contacts, the matrix identifies polar interactions that stabilize the TFPI-JBT1837 complex, e.g., by hydrogen bonds between the side chain of $H6_{AP}$ with the carbonyl oxygen of M134 and Nε of $W5_{AP}$ with the carbonyl oxygen of Q63, as well as a short β-strand comprising $M16_{AP}$-

$C18_{AP}$ and R83-I85. Additionally Nζ of $K4_{AP}$ is equidistantly coordinated by the side chains of E67 and E142.

[0209] Beside its role in JBT1837 stabilization, the disulfide bridge of $C7_{AP}$-$C18_{AP}$ perfectly fits into a hydrophobic cavity formed by E71, M75, N82, and I84, thus playing a significant role in stabilizing the TFPI-JBT1837 complex.

[0210] Although TFPI is mainly conserved through different species, the key residues within the linker and KD2 (M134, I146) are only shared by close relatives to the human species (Figure 74). Additionally, the A81V substitution in macaque destabilizes the β-turn of the linker by steric hindrance, impairing subsequent interaction of the linker with JBT1837.

[0211] KD1 of the previously solved KD1+JBT1857 complex was superimposed on the KD1-KD2+JBT1837 complex to provide a model of the arrangement of the ternary structure. The model suggested that JBT1837 and JBT1857 bind to opposing sites of KD1. The C-terminus of JBT1857 and the N-terminus of JBT1837 were 20 Å apart; thus, a linker moiety approximately 20 Å in length, corresponding to about 10 amino acids, would connect JBT1857 and JBT1837 and allow binding to the subunits respective binding sites on TFPI.

[0212] KD1-KD2+JBT1837+JBT1857 crystals belong to the tetragonal space group $P\,4_32_12$ and diffract to a maximum resolution of 2.7 Å. Screw axes were confirmed by axial extinctions and cell content analysis indicated a mass of 72 kDa in the asymmetric unit. Assuming that the ternary complex crystallized, this corresponds to either three (62 kDa; 61% solvens) or four (82 kDa; 46% solvens) copies in the asymmetric unit. However, the structure could not be completely solved, possibly due to the inherent flexibility of the KD1-KD2 molecule structure. Analysis of the interactions between TFPI and JBT1837, as well as the peptide-induced conformation of TFPI, suggests that JBT1837 may not bind to TFPI of species other than human. The high selectivity of JBT1837 is advantageous as it minimizes cross-reactivity and thus unwanted side effects. JBT1857 binds TFPI at a different site, and native PAGE demonstrated that both peptides can bind to TFPI at the same time. This analysis is further confirmed by a crystal structure-based model of KD1-KD2+JBT1837+JBT1857 (Figure 75). The C-terminus of JBT1857 and the N-terminus of JBT1837 lie only 20 Å apart, and a linker of e.g., ten Ala (or Ser) would allow binding of both JBT1837 and JBT1857 to their binding sites within TFPI.

Example 16

[0213] The following example describes the characterization of a peptide complex that binds TFPI.

[0214] TFPI-binding peptides were produced as described herein. The peptides were synthesized as trifluoroacetate (TFA) salts with a purity >90%, and solved in DMSO to a stock concentration of 10 mM.

[0215] Competition (IC_{50}) ELISAs were performed using biotinylated TFPI-binding peptides as "tracers" to compete for TFPI-binding with non-biotinylated candidate peptides. The assay principle is depicted in Figure 6B. Ninety-six well Maxisorp plates (Nunc) were coated with 0.05 µg/mL TFPI in coating buffer (15 mM Na_2CO_3, 35 mM $NaHCO_3$, pH 9.6) overnight. Plates were washed three times with 350 µl wash buffer (HNaT: 175 mM NaCl, 25 mM HEPES, 5 mM $CaCl_2$, 0.1% Tween 80, pH 7.35), and blocked with 200 µl 2% yeast extract in HNaT for two hours. Plates were then washed three times with 350 µl HNaT. Biotinylated tracer peptides were applied at a concentration corresponding to their respective EC_{90} values determined in a binding ELISA (mean value if n > 2). A competitor stock solution of candidate peptide (10 mM or prediluted in DMSO) was diluted 1/33.3 in HNaT, and a serial 1/3 dilution was prepared with HNaT with 3% DMSO. The dilution strategy employed in a particular assay was adjusted based on the affinity of the peptide. The dilution

was further diluted with the biotinylated tracer peptide in a ratio of 1:6 (20 µl competitor dilution and 100 µl tracer peptide). The mixture of competitor and tracer peptide was applied to the TFPI-coated microtiter plate and incubated for 1.5 hours. The plates were washed three times with 350 µl HNaT. Peptide-TFPI binding was detected by applying horseradish peroxidase (HRP)-conjugated streptavidin to the microtiter plate, incubating the mixture for one hour, washing the plate three times with 350 µl HNaT, applying TMB (3,3'5,5'-Tetramethylbenzidin), and detecting the subsequent chromogenic conversion of TMB by HRP. IC_{50} graphs for the representative peptides JBT1857 (SEQ ID NO: 178), JBT1837 (SEQ ID NO: 1044) and the peptide complex JBT2547 (Ac-FQSKpNVHVDGYFERL-Aib-AKLSSSSSSSSSSSYYKWH[CAMRDMKGTMTC]VWVKF-NH2 (SEQ ID NO: 4260)) are depicted in Figures 66A and 66B.

[0216] The TFPI-inhibitory peptide complex JBT2547, comprising two different TFPI-binding peptides (JBT1857 and JBT1837) that bind two different sites within TFPI, demonstrated an IC_{50} of $1.33x10^{-9}$ M (Tracer JBT2271) and an IC_{50} of $5.57x10^{-10}$ M (tracer JBT2316), which was comparable or lower than the IC_{50} of the peptide subunits.

[0217] Additionally, JBT2547 binding to full length TFPI was characterized in a surface plasmon resonance assay (BIAcore T200™, GE Healthcare, Chalfont St. Giles, UK). Recombinant, full length TFPI-1 was immobilized on a CM5 chip aiming at 500 RU. Peptide was injected at a flow rate of 30 µL/minutc at concentrations ranging from 1 to 16 nM in HBS-P buffer, pH 7.4, 0.1% DMSO. Subsequently, peptide was dissociated for 600 seconds. BIAcore T200™ Evaluation Software was utilized to analyze the data, which revealed that JBT2547 tightly binds full length TFPI with a binding constant < 1 nM. The K_D was calculated to be 7.49×10^{-12}M.

Example 17

[0218] This example describes an exemplary method for characterizing the binding affinity of a TFPI-binding peptide complex using a k_{off} assay.

[0219] Wells of a microtiter plate (96 wells, Maxisorp, Nunc) were coated with 1.6 nM TFPI in coating buffer (15 mM Na_2CO_3, 35 mM $NaHCO_3$, pH 9.3) for two hours at room temperature. The plate was then washed three times with 350 µl wash buffer (175 mM NaCl, 5 mM $CaCl_2$, 25 mM HEPES, 0.1% Tween 80, pH 7.35), and the wells were blocked with 100 µl blocking buffer (2% yeast extract, 175 mM NaCl, 5 mM $CaCl_2$, 25 mM HEPES, 0.1% Tween 80, pH 7.35). If an incubation period of 24 hours was employed, the wells were blocked for at least one hour. Control wells used for a 15 minute incubation period were blocked for additional 23.5 hours.

[0220] For a 24 hour incubation period, the wells were washed three times with 350 µl wash buffer and incubated with 50 µl test peptide (JBT2547 (SEQ ID NO: 4260) or JBT1857 (SEQ ID NO: 178)) in wash buffer. The concentration of test peptide was adjusted based on the individual IC_{90} concentration determined in, e.g., the TFPI IC_{50} ELISA assay described herein. The TFPI-coated wells were exposed to test peptide for approximately 15 minutes. The wells were subsequently washed three times with 350 µl wash buffer, and 50 µl tracer (competitor) peptide (JBT2271 (SEQ ID NO: 4033)) was added. Control wells (maximum signal) were incubated with tracer only. Blank wells lacking TFPI-coating were incubated with tracer only. Addition of the tracer peptide started the 24 hour incubation period.

[0221] A 15 minute incubation period was employed as a control if the IC_{90} concentration of the test peptide led to a 90% reduction of the maximum signal. Wells blocked for an additional 23.5 hours were washed three

times with 350 µl wash buffer to remove the blocking buffer. Subsequently, 50 µl analyte in wash buffer was added, and the wells were incubated for 15 minutes. The concentration of test peptide utilized was adjusted based on the peptide's IC_{90} concentration determined using, e.g., a TFPI IC_{50} ELISA assay. The 15 minute incubation was followed by three wash steps with 350 µl wash buffer and the addition of 50 µl tracer peptide. Control wells (maximum signal) were incubated with tracer only. Blank wells lacking TFPI also were incubated with tracer only.

[0222] The plate was washed three times with 350 µl wash buffer, and 50 µl HRP-conjugated streptavidin (R&D Systems, 1:200 in wash buffer) was added to each well. After an incubation period of one hour at room temperature, the plate was washed three times with wash buffer. TMB solution (50 µl per well; SeramunBlau fast, Seramun) was added. After a 1.5 minute incubation at room temperature, the reaction was stopped by the addition of 50 µl 1 M H_2SO_4 per well. Absorbance was measured using a photometer (Spectramax M5, Molecular Devices) at 450 nm and 620 nm.

[0223] The assay results are presented in Figure 67 as a percentage of the corrected optical density (OD450-OD620) of wells exposed to test peptide and tracer peptide in relation to TFPI-coated wells exposed only to tracer. After 24 hours, the TFPI-binding peptide complex continued to block binding of the tracer to TFPI. In contrast, JBT1857 dissociated from TFPI during the 24 hour incubation period. Thus, JBT2547 dissociated significantly slower from TFPI than one of its peptide subunit, JBT1857.

[0224] The affinity of JBT2528 for TFPI also was determined using the methods described herein: K_D = 1.6 nM; k_{on} = 6.5 x 10^5 1/Ms; k_{off} = 9.7 x 10^{-4} 1/s. JBT2528 binds TFPI KD1.

Example 18

[0225] This example describes the plasma stability of the TFPI-binding peptide complex JBT-2547 in mouse and human plasma. JBT2547 (SEQ ID NO: 4260) and JBT1857 (SEQ ID NO: 178) were added to samples of mouse or human plasma and incubated for 24 hour at 37°C. The percentage of the initial amount of peptide remaining after incubation was determined by IC_{50} ELISA on Maxisorp plates coated with 0.05 mg/ml TFPI (2.26 nM tracer peptide JBT2271). Less than 5% of the initial amount of JBT1857 remained in mouse plasma after the incubation period, while 15% of the initial amount of JBT-2547 remained after the same incubation period. Similarly, JBT2547 was more stable in human plasma compared to JBT1857 and JBT1837; 27% of the original amount of JBT1857 and 46% of JBT1837 remained after 24 hours while 54% of the original amount of JBT2547 was detected. Thus, the complex of JBT1857 and JBT1837 increased stability and resistance to plasma proteases. JBT2528 (Hex-FQSKp-C(Acm)-VH-Tle-DaYFERL-Aib-AKL-NH2 (SEQ ID NO: 4246)) also exhibited enhanced stability: 93% of the original amount of remained after 24 hours in human plasma, and 35% of the original amount of remained after 24 hours in mouse plasma.

Example 19

[0226] The following example describes the characterization of the TFPI-inhibitory activity of a TFPI-binding peptide complex and TFPI-binding peptide monomers using FXa inhibition and extrinsic tenase inhibition assays described in Example 3.

[0227] Peptides were diluted in 1.25X reaction buffer + 0.1% Tween-80 (31.25 mM HEPES; 218.75 mM NaCl; 6.25 mM $CaCl_2$; 0.125% BSA; pH 7.35) from 1 or 10 mM stocks (in DMSO). TFPI, FVIIa, and lipidated TF were diluted in 1.25X reaction buffer. Phospholipid vesicles (DOPC/ POPS 80/20) and chromogenic substrate

specific for FXa (S-2222 (available from DiaPharma, West Chester, OH)), all diluted in Aqua dest., were added to 96-well plates. After an incubation period, TFPI and peptide dilutions were added. The TFPI concentration in the extrinsic tenase inhibition assay was 0.0625 nM. FX activation was initiated by adding FX to the wells. FXa-mediated chromogenic substrate conversion was determined by observing an increase in absorbance using a micro-plate reader. The amount of FXa generated at certain time points was calculated from the OD readings. FXa generated at 20 minutes after start of the reaction was considered for calculation of EC_{50} from plots of peptide concentration versus the inhibition of TFPI (%).

[0228] The functional inhibition of TFPI also was examined using a FXa inhibition assay. A FXa-specific chromogenic substrate (S-2222) and phospholipid vesicles (DOPC/ POPS 80/ 20), both diluted in Aqua dest., and TFPI proteins (full length human TFPI, human TFPI 1-160, murine TFPI 1-160, and cynomolgus TFPI 1-160) diluted in 1.25X reaction buffer, were added to 96 well plates. The TFPI concentration in the FXa inhibition assay was 0.5 nM. Peptides were diluted from 1 or 10 mM stocks (in DMSO) in 1.25 x reaction buffer + 0.1% Tween-80. The peptide dilutions (2.5 µl) were added to the 96 well plates. The conversion of chromogenic substrate was triggered by the addition of FXa, and the kinetics of the conversion was measured in a micro-plate reader. OD readings after 115 minutes were considered for calculation of the EC_{50} from plots of peptide concentration versus the inhibition of TFPI.

[0229] Results from the FXa inhibition assay and extrinsic tenase assay are provided in Table 19 and Figures 68A and 68B.

TABLE 19

	FXa Inhibition Assay		Extrinsic Tenase Assay	
	EC_{50} [µM]	**Maximal inhibition (%)**	**EC_{50} [µM]**	**Maximal inhibition (%)**
JBT1837	1.3	97	0.2	72
JBT1857	4.3	90	8.8	59
JBT2528	2.9	89	6.0	71
JBT2547	1.0	98	0.3	99
JBT1837 + JBT1857	0.8	98	0.3	84

[0230] JBT1837, JBT2547, JBT2528, and the combination of JBT1837 and JBT1857 very efficiently inhibited 0.5 nM full length and C-terminally truncated TFPI 1-160 (Table 19 and Figures 68A and 68B) in the FXa inhibition assay. JBT1857 less efficiently inhibited both TFPIs with EC_{50}s of 4.3 and 3.1 nM, respectively. At concentrations above 100 nM, JBT1837, JBT2547, and the combination of JBT1837 and JBT1857 fully blocked TFPI activity, as indicated by a nearly 100% maximal inhibition. JBT1857 (and JBT2528) is a partial inhibitor of TFPI and demonstrated some residual TFPI inhibitory activity at high and saturating concentrations. Inhibition of both full length and C-terminally truncated TFPIs confirms that binding epitopes of the peptides are within Kunitz Domains 1 and 2, and that the C-terminal region of TFPI is not required for efficient inhibition of TFPI activity.

[0231] The candidate peptides also were analyzed for inhibition of murine and cynomolgus monkey TFPI. C-terminally truncated murine and monkey TFPI proteins (TFPI 1-160) were used in a FXa inhibition assay performed as described herein. The results are summarized in Table 20 and Figures 68C and 68D.

TABLE 20

FXa Inhibition Assay	Mouse TFPI 1-160		Cynomolgus Monkey TFPI 1-160	
	EC_{50} [µM]	Maximal inhibition (%)	EC_{50} [µM]	Maximal inhibition (%)
JBT1837	-	-	-	-
JBT1857	7.2	93	2060*	-*
JBT2547	3.1	85	10	100
JBT1837 + JBT1857	7.0	89	100	89
* no maximum reached for fitting				

[0232] JBT1837 did not inhibit mouse TFPI 1-160 up to 10 µM (Figure 68C). Cynomolgus monkey TFPI was only weakly inhibited at µM concentrations, likely due to inter-species sequence differences which are incompatible with binding of JBT1837 (Figure 68D). JBT1857 efficiently inhibited mouse TFPI resulting in an EC_{50} of 7.2 nM, which is comparable to inhibition of human TFPI. Cynomolgus monkey TFPI was inefficiently inhibited by JBT1857, likely due to an Ala to Pro substitution within the binding site of JBT1857.

[0233] JBT2547 most efficiently inhibited both mouse and cynomolgus monkey TFPI. JBT2547 mediated an approximately 200-fold and 10-fold greater reduction of EC_{50}s compared to JBT1857 and to the combination of JBT1837 and JBT1857. This further indicates that the molecular fusion of the two peptides entities increases its TFPI inhibitory activity.

[0234] To demonstrate that the peptide complex efficiently inhibits high concentrations of TFPI, peptides were titrated at increasing human TFPI concentrations. JBT2547 stoichiometrically and fully (100%) inhibited TFPI activity at all concentrations tested (up to 10 nM) (Figures 68E and 68F). In contrast, peptides JBT1837 and JBT2548 (a derivative of JBT1857) are partial inhibitors of TFPI at high concentrations (Figure 68D). Linking TFPI-inhibitory peptides to form a peptide complex enhances inhibition of TFPI activity.

[0235] In the extrinsic tenase assay, JBT2547 restored extrinsic complex-mediated FX activation in the presence of TFPI with an EC_{50} of 0.3 nM, resulting in nearly 100% inhibition of TFPI activity at low TFPI concentration (0.063 nM). JBT1837, JBT1857 and the combination of JBT1837 + JBT1857 less efficiently inhibited TFPI, as indicated by higher EC_{50} and reduced maximal inhibition (Figure 69A). To demonstrate that the peptide complex efficiently inhibits high concentrations of TFPI, JBT2547 was titrated at increasing TFPI concentrations. JBT2547 efficiently inhibited TFPI activity at all concentrations tested (up to 10 nM) with lowest EC_{50}s of the peptides tested (Figures 69B-69D). In contrast, peptides JBT1837, JBT2548, and the combination of peptides JBT1837 + JBT1857 partially inhibited TFPI at high concentrations (Figures 69C-69D) with higher EC_{50} compared to JBT2547.

[0236] TFPI antagonist activities of JBT1837, JBT1857, a mixture of JBT1837 and JBT1857, and JBT2547 also was determined in reactions systems in which FXa was inhibited by TFPI in the presence of Ca^{2+}, phospholipid (PL) + Ca^{2+}, and PL + Ca^{2+} + protein S. The effectivity by which the peptides block the activity of TFPI increases in the order JBT1857, JBT1837, the mixture of JBT1837 and JBT1857, and JBT2547. JBT1837 and JBT1857 did not completely block TFPI, particularly not in the presence of cofactor protein S. A mixture of JBT1837 and JBT1857 was more potent as TFPI antagonist than the individual peptides. The fusion peptide JBT2547 was by far the best TFPI antagonist; at a concentration of 50 nM the peptide complex almost completely blocked TFPI even in the presence of PL + Ca^{2+} + protein S.

[0237] This example demonstrates that the linkage of two TFPI-binding peptides of the invention by, e.g., molecular fusion, improves TFPI inhibitory activity. JBT1837 prevents the formation of the primary TFPI-FXa complex (encounter complex) and at high concentrations can fully block TFPI inhibition of FXa. JBT1857 prevents the transition of the weak encounter to the tight TFPI-FXa complex and partially inhibits TFPI since in the presence of JBT1857 it is still possible to form the primary TFPI-FXa complex (encounter complex). A molecular fusion of the two TFPI binding peptides demonstrated a great degree of inhibitory activity than each peptide alone and a mixture of the peptide subunits (i.e., the effect was greater than additive).

Example 20

[0238] In this example, the TFPI inhibitory activity of a TFPI-binding peptide complex is characterized using the plasma-based assay of Example 4. TFPI-binding peptides were tested under physiological conditions (~0.2 nM full length TFPI) as well conditions mimicking elevated full length TFPI plasma levels (up to 10 nM full length TFPI).

[0239] The influence of peptides on thrombin generation in the absence or in the presence of exogenous flTFPI was measured in duplicate via calibrated automated thrombography in a Fluoroskan Ascent® reader (Thermo Labsystems, Helsinki, Finland; filters 390 nm excitation and 460 nm emission) following the slow cleavage of a thrombin-specific fluorogenic substrate (Hemker, Pathophysiol. Haemost. Thromb., 33, 4-15 (2003)). As a model for antibody-mediated FVIII deficiency, frozen pooled normal plasma (George King BioMedical Inc., Overland Park, KN) was incubated with high titer, heat inactivated, anti-human FVIII plasma raised in goat (4490 BU/ml; Baxter BioScience, Vienna, Austria) giving rise to 50 BU/mL. Assays were also performed with cynomolgus monkey and marmoset monkey plasma. The plasmas were mixed with corn trypsin inhibitor (CTI) (Hematologic Technologies, Inc., Essex Junction, VT) to inhibit Factor XIIa contamination, resulting in a final concentration of 40 µg/mL.

[0240] Pre-warmed (37°C) plasma (80 µL) was added to each well of a 96 well micro-plate (Immulon 2HB, clear U-bottom; Thermo Electron, Waltham, MA). To trigger thrombin generation by Tissue Factor, 10 µL of PPP low reagent containing recombinant human Tissue Factor (12 pM) and phospholipid vesicles composed of phosphatidylserine, phosphatidylcholine and phosphatidylethanolamine (48 µM) (Thrombinoscope BV, Maastricht, The Netherlands) were added. Just prior to putting the plate into the pre-warmed (37°C) reader, 5 µL of peptide solutions were added, resulting in plasma concentrations of 1-100 nM. Finally, 5 µL HEPES buffered saline with 5 mg/mL bovine serum albumin (Sigma-Aldrich Corporation, St. Louis, Missouri, USA) or 5 µL full length TFPI (flTFPI) dilution were added. The flTFPI protein (3557 nM) had been expressed in SK Hep cells and purified. Plasma concentrations of flTFPI varied between 0.31 and 10 nM, which is equivalent to a ~2 to 50-fold increase in endogenous flTFPI plasma concentration. Thrombin generation was initiated by addition of 20 µL of FluCa reagent (Thrombinoscope BV, Maastricht, The Netherlands) containing a fluorogenic substrate and HEPES-buffered $CaCl_2$ (100 mM). Fluorescence intensity was recorded at 37°C. The parameters of the resulting thrombin generation curves were calculated using Thrombinoscope™ software (Thrombinoscope BV, Maastricht, The Netherlands) and thrombin calibrator to correct for inner filter and substrate consumption effects (Hemker, Pathophysiol. Haemost. Thromb., 33, 4-15 (2003)). The final plasma dilution, TF concentration, and assay temperature for the human plasma assay was 1:1.5, 1 pM, and 37°C, respectively. The final plasma dilution, TF concentration, and assay temperature for the mouse C57B16 plasma assay and mouse FVII -/- plasma assay was 1:2.4, 0.4 pM, and 33°C respectively. The final plasma dilution, TF concentration, and assay temperature for the cynomolgous and marmoset monkey plasma assays was 1:1.5, 0.6 pM, and 37°C respectively.

[0241] Representative results for improvement of thrombin generation of human FVIII inhibited plasma are provided in Table 21.

TABLE 21

	EC_{50} (nM)	Maximal improvement of thrombin generation relative to an inhibitory polyclonal anti TFPI antibody (%)
JBT1834	4.8	55.9
JBT1857	6.8	37.0
JBT1837+JBT1857	7.4	104.6
JBT2547	7.8	112.7

[0242] JBT1837, JBT1857, a mixture of JBT1837 + JBT1857, and the complex of JBT1837 and JBT1857 (JBT2547) improved thrombin generation of FVIII- inhibited plasma with similar EC_{50}s. The maximal efficacy relative to a polyclonal anti-TFPI antibody was highest for JBT2547, demonstrating that fusion of two TFPI-binding peptides described herein enhances inhibition of TFPI activity in plasma.

[0243] Figures 70A-70C illustrate the inhibitory activity of 1-100 nM of the fusion peptide JBT2547 (diamond) toward elevated levels of flTFPI plasma levels (1.25, 3.75 and 10 nM) in the thrombin generation assay. In comparison with JBT1837 (triangle) and JBT1857 (circle), JBT2547 shows a substantially greater ability to increase the thrombin peak even in the presence of very high flTFPI plasma levels. Although combining JBT1837 and JBT1857 (square) improved the response over each monomeric peptide alone, the combination of monomers did not achieve the level of activation achieved by the peptide complex. The TFPI-antagonistic potential of JBT2547 also was tested in the presence of a wide concentration range of f1TFPI of up to 10 nM, which is equivalent to 50-fold higher than physiological flTFPI plasma concentration. Concentrations of 50-100 nM JBT2547 fully compensated the anticoagulant effect of 10 nM flTFPI, and thrombin peak values reached normal plasma levels or above. JBT2547 concentrations below 50 nM improved thrombin generation of FVIII-inhibited plasma in a flTFPI-dependent manner.

[0244] The results of thrombin generation experiments with several animal plasmas are summarized in Tables 22 and 23.

TABLE 22

	Mouse C57B16 plasma		Mouse FVIII knock out	
	EC_{50} [nM]	Maximal inhibition (%)	EC_{50} [µM]	Maximal inhibition (%)
JBT1837	No binding	No binding	No binding	No binding
JBT1857	64.1	115.8	55.1	134.4
JBT2547	41.0	109.3	56.7	138.3
JBT1837 + JBT1857	52.7	104.7	61.1	144.0

TABLE 23

	EC_{50} [nM]; Cynomologus Monkey	EC_{50} [µM]; Marmoset Plasma
JBT1837	-	No binding
JBT1857	1003.7	35.1
JBT2547	-	38.4
JBT1837 + JBT1857	-	34.8
* "-" indicates no curve fit		

[0245] JBT1837 did not inhibit mouse and marmoset monkey TFPI at relevant concentrations. Cynomolgus

monkey TFPI is a poorly inhibited only at µM concentrations. JBT1857 efficiently inhibited mouse and marmoset monkey TFPIs, whereas cynomolgus monkey TFPI was less efficiently inhibited by JBT1857, likely due to an Ala to Pro amino acid substitution within the binding site of JBT1857 which is conserved as an Ala in human, mouse and marmoset monkey TFPI. Cynomolgus monkey TFPI is most efficiently inhibited by the fusion peptide JBT2547.

Example 21

[0246] This example describes the ability of the TFPI-binding peptides described above to restore extrinsic tenase complex-mediated conversion of FX to FXa in a cell-based extrinsic tenase assay. The assay was performed as described in Example 5. FXa concentrations obtained after nine minutes of reaction in the presence of 100nM polyclonal anti-human TFPI antibody (R&D Systems, AF2974) were set to 100%, and the samples that were not exposed to antibody were set to 0% TFPI inhibitory effect, allowing for assessment of peptide's inhibitory activity. The results of the assay are illustrated in Figures 71A and 71B. JBT2547 demonstrated improved inhibition of HUVEC TFPI compared to JBT1837, JBT1857, JBT2548, and a mixture of JBT1857 and JBT1837. Full inhibition of TFPI was achieved at concentrations as low as about 10nM.

Example 22

[0247] This example describes a method for assessing clot formation in whole blood using rotation thromboelastography (ROTEM). The assay was performed as described in Example 8. The analytical set-up was as follows: 300 µL of pre-warmed (37°C) CTI treated citrated whole blood was added to a peptide sample (10, 100, 1000 nM final assay concentration) or a control, followed by addition of a 1:15 dilution of TF PRP reagent containing recombinant human tissue factor (rTF, 3 pM) (TS40, Thrombinoscope BV, Maastricht, The Netherlands). In certain experiments, exogenous full length TFPI (flTFPI) at final concentrations of 2 or 10 nM was added to simulate flTFPI plasma levels of up to 50-fold over normal. Coagulation was initiated by the addition of 20 µL 200 mM $CaCl_2$ (star-TEM®, Pentapharm, Munich, Germany) and recordings were allowed to proceed for at least 120 min. The final concentration of rTF in the assay was 44 fM. The thromboelastographic parameters of clotting time (CT), clot formation time (CFT) and maximum clot firmness (MCF) were recorded in accordance with the manufacturer's instructions. CT is defined as the time from the start of measurement to the start of clot formation. CFT is defined as the time from the start of clot formation until an amplitude of 20 mm is reached. MCF is the maximum difference in amplitude between the two traces during the assay.

[0248] Addition of full length TFPI to FVIII-inhibited whole blood in the absence of TFPI inhibitory peptides substantially inhibited coagulation as indicated by a marked increased in clot time (Figures 72A-72C). JBT2528 and JBT1837 improved global hemostatic parameters of FVIII-inhibited plasma (EC_{50}=11 nM and 4 nM, respectively). JBT1837 and JBT2548 improved coagulation by reducing the clot time to normal levels in a concentration dependent manner (Figures 72B and 72C; open circles). These peptides failed to reach clot times of FVIII-inhibited blood at increased TFPI concentrations (e.g., 10 nM). In contrast, concentrations of JBT2547 above 100 nM fully restored normal coagulation of FVIII-inhibited whole blood to which 2 nM and 10 nM full length TFPI was added.

[0249] This example further confirms that TFPI-binding peptide complexes efficiently neutralize TFPI and restore normal coagulation even at increased TFPI concentrations.

Example 23

[0250] Platelets contain full length TFPI, which is released to plasma upon platelet activation and results in a platelet TFPI concentration in plasma that approximates 50-75% of the full length TFPI in plasma at sites of injury. This example demonstrates inhibition of platelet TFPI by peptides of the invention.

[0251] *Blood collection and plasma preparation*: Blood samples were collected from a number of different donors. Nine volumes of venous blood was drawn by venipuncture into 1 volume of 1.09 tri-sodium citrate with or without 500 µg/ml CTI and centrifuged at 250xg for 15 minutes for the preparation of platelet-rich plasma (PRP) or at 2860xg for the preparation of platelet-poor plasma (PPP). PPP was aliquoted, snap-frozen and stored at -80 °C until use. In experiments in which a normal plasma pool (NP) was used, blood collected from more than 25 healthy individuals was centrifuged at 2000xg for 15 minutes to separate plasma from blood cells, and again at 11000xg for 5 min to obtain platelet-poor plasma.

[0252] *Platelet isolation:* Blood (45 ml) was collected on 7.5 ml acid citrate/dextrose (ACD, 80 mM trisodium citrate, 52 mM citric acid, 183 mM glucose) and centrifuged during 15 min at 248xg. To remove residual erythrocytes, the supernatant (platelet rich plasma, PRP) was centrifuged for an additional 5 minutes at 248xg. The PRP was subsequently centrifuged during 15 minutes at 2760 rpm (1360xg) to spin down the platelets. The platelet pellet was washed twice by resuspending in 20 ml (first wash) and 15 ml (second wash) platelet buffer (10 mM HEPES, 136 mM NaCl, 2.7 mM KCl, 2.0 mM $MgCl_2$, 0.5% bovine serum albumin and 0.2% glucose, pH 6.6), followed by centrifugation during 15 minutes at 2760 rpm (1360xg). After the second wash step, the platelets were resuspended in 3.5 ml platelet buffer (pH 7.5) and the platelet concentration was determined in a Coulter counter. Finally, the platelet suspension was diluted to the required platelet concentration by dilution with platelet buffer (pH 7.5) and 40 µl of a 25 mg/ml solution of the synthetic peptide Arg-Gly-Asp-Ser (RGDS) was added per 3.5 ml platelet suspension to inhibit platelet aggregation during storage at room temperature.

[0253] *Functional Characterization of Platelet TFPI:* TFPI was released from isolated platelets by activating the platelets at 37 °C during 15 min with 100 ng/ml convulxin. The platelets were spun down by centrifugation (15 min at 2800xg) in an Eppendorf centrifuge tube, and the supernatant was collected as source of platelet TFPI. Platelet TFPI was quantified with a full length TFPI ELISA using recombinant TFPI as standard. The functional activity of platelet TFPI was compared to the activity of recombinant TFPI in two assay systems: 1) inhibition of FVIIa-catalysed FX activation in a model system and 2) inhibition of thrombin generation in TFPI-deficient plasma.

[0254] In the model system assay, the effect of varying concentrations recombinant or platelet TFPI on TF-FVIIa catalysed FX activation was determined at 37 °C in a 25 mM HEPES (pH 7.7), 175 mM NaCl, 5 mg/ml bovine serum albumin (BSA) buffer containing 2 pM FVIIa, 5 nM TF, 100 nM FX, and 400 uM Fluophen FXa. In this assay FVIIa, TF, TFPI and Fluophen FXa were preincubated for 7 minutes at 37 °C in the HEPES buffer and FX activation was initiated by adding FX. Progress curves of fluophen FXa conversion by the FXa generated were determined in Fluoroskan Ascent® reader (Thermo Labsystems, Helsinki, Finland) and corrected for consumption of Fluophen FXa and so-called inner filter effects (Udenfriend S, Fluorescence Assay in Biology and Medicine. New York, Academic Press, 1996, vol 1, pp13, 118, vol 2, pp182-185) by the methodology that is also used for the correction of thrombin generation curves for substrate consumption (Hemker et al., Pathophysiol Haemost Thromb, 32, 249-53 (2002)). Time courses of FXa generation and the effects of TFPI thereon were obtained by taking the first derivative of the corrected progress curves of Fluophen FXa conversion. Alternatively, the effect of TFPI on TF-FVIIa-catalysed FX activation was also followed with a FXa-specific chromogenic substrate, e.g., 125 µM CS-11(65).

[0255] In the thrombin generation assay, TFPI-depleted plasma was reconstituted with varying amounts of recombinant TFPI or platelet-derived TFPI, and thrombin generation was determined using the Calibrated

Automated Thrombogram (CAT) method described by Hemker et al., *supra.* Thrombin generation was initiated in plasma by addition of varying concentrations of recombinant TF (0.1-10 pM), 16 mM $CaCl_2$, 30 µM phospholipid vesicles (1,2-dioleoyl-sn-glycero-3-phosphoserine/1,2-dioleoyl-sn-glycero-3-phosphoethanolamine/1,2-dioleoyl-sn-glycero-3-phosphatidylcholine, 20/20/60, M/M/M) and 30-50 µg/ml corn trypsin inhibitor (CTI). In experiments in PRP, no phospholipid vesicles were added to the plasma. The thrombin activity in plasma was monitored continuously with the fluorogenic substrate Z-Gly-Gly-Arg-AMC (BACHEM, Bubendorf, Switzerland). Fluorescence was read in a Fluoroskan Ascent® reader (Thermo Labsystems, Helsinki, Finland) and thrombin generation curves were calculated using Thrombinoscope™ software (Thrombinoscope, Maastricht, The Netherlands).

[0256] *Analysis of the effect of peptides of the invention on recombinant-, plasma-, and platelet TFPI:* The effects of TFPI binding peptides on the anticoagulant activity of TFPI was tested in model systems (inhibition of FXa and TF-FVIIa-catalysed FX activation by TFPI) and in plasma (NP, PPP, PRP or TFPI-depleted plasma reconstituted with recombinant- or plate let-TFPI or platelets). The effect of peptides on the inhibition of FXa by TFPI was followed in a HNBSA buffer (50 mM HEPES pH 7.7, 175 mM NaCl, 5 mg/mL BSA) containing 125 µM CS-11(65), 1 mM EDTA or 3 mM $CaCl_2$ and, if present, varying concentrations of peptide and TFPI, 80 nM protein S and/or 30 µM phospholipid vesicles (20:60:20 DOPS/DOPC/DOPE), which was preincubated for 10 minutes at 37 °C. hFXa was added and the increase in absorbance at 405 nm was followed in an Ultra Microplate Reader (Bio-Tek, Burlington, VT, USA) until a steady state rate of chromogenic substrate conversion was achieved (~ 60 min). Progress curves of chromogenic substrate conversion were fitted to the integrated rate equation for slow-tight binding inhibition (Huang et al., J Biol Chem, 268, 26950-55 (1993)):

$$A_t = A_0 + (v_s.t) + (v_0 - v_s).(1 - \exp(-k_{obs}.t))/k_{obs}$$

in which A_t is absorbance at 405 nm at time t; A_0 is initial absorbance at 405 nm; v_s is final steady-state velocity; v_0 is initial velocity; k_{obs} is apparent rate constant for the transition from v_0 to v_s (FXa-TFPI to FXa-TFPI*). The values v_0 and v_s relative to rates of chromogenic substrate conversion by FXa in the absence of TFPI, represent the extent of loose and tight FXa-TFPI complex formation.

[0257] The effect of peptides on the inhibition of TF-FVIIa-catalysed FX activation by TFPI was determined at 37 °C in a 25 mM HEPES (pH 7.7), 175 mM NaCl, 5 mg/ml bovine serum albumin (BSA) buffer containing 2 pM FVIIa, 5 nM TF, 100 nM FX, 400 µM Fluophen FXa and varying amounts of peptide and TFPI (recombinant- or platelet-derived). In this assay FVIIa, TF, TFPI, peptide, and Fluophen FXa were preincubated for 7 minutes at 37 °C in the HEPES buffer, and FX activation was initiated by adding FX. Progress curves of Fluophen FXa conversion by the FXa generated were determined in Fluoroskan Ascent® reader (Thermo Labsystems, Helsinki, Finland) and corrected for consumption of Fluophen FXa and so-called inner filter effects by the methodology that is also used for the correction of thrombin generation curves for substrate. The time courses of FXa generation and the effects of TFPI thereon were obtained by taking the first derivative of the corrected progress curves of Fluophen FXa conversion. Alternatively, the effect of TFPI on TF-FVIIa-catalysed FX activation was also followed with a FXa-specific chromogenic substrate e.g. 125 µM CS-11(65).

[0258] The effect of peptides on plasma- and platelet TFPI was assessed by measuring their effects on thrombin generation determined as described below in NP, PPP, PRP or in TFPI-depleted plasma reconstituted with recombinant- or platelet-TFPI or platelets. In experiments in PRP or in TFPI-depleted plasma reconstituted with platelets, the platelets were either not-activated or activated with 80 ng/ml convulxin or 40 ug/ml Horm collagen (final concentrations in the well). To simulate haemophilia plasma, thrombin generation experiments were performed in the presence of a goat inhibitor plasma of which 1 µl was added to 80 µl plasma to neutralize FVIII.

[0259] In thrombin generation experiments, the plasma was added to the microtiter plate and, if present,

mixed with proper amounts of phospholipids, platelets, TFPI, anti-FVIII, anti-TFPI antibodies, peptide and incubated for 7 minutes at 37 °C. After this preincubation, TF and platelets activator (if present) were added immediately, followed by a mixture fluorogenic substrate/$CaCl_2$ mixture (FluCa) which initiates thrombin generation.

[0260] *Results:* The platelet supernatant TFPI inhibited TF-FVIIa-catalysed FX inactivation in a concentration-dependent manner. The inhibition by platelet supernatant was prevented by a cocktail of anti-TFPI antibodies, which underscores that it was TFPI in the platelet supernatant that is the inhibitor of TF-FVIIa-catalysed FX activation. However, rates of FX activation in the presence of platelet supernatant and anti-TFPI were higher than the rate of FX activation without platelet supernatant, suggesting that the platelet supernatant contained a small amount of a FX activator, e.g., FVIIa. This suggests that the data is an underestimation of the platelet TFPI concentration due to the fact that more FVIIa has to be inhibited when platelet supernatant is present in the assay mixture than in the assay mixture in which the calibration curve is made with recombinant TFPI in buffer.

[0261] The functional activity of platelet TFPI was also tested in TFPI-deficient plasma and compared to the activity of the recombinant non-glycosylated TFPI. The concentrations of platelet TFPI from different donors were determined with an ELISA in which the recombinant non-glycosylated TFPI was used as calibrator. Although no titrations were made with the different TFPI preparations in TFPI-deficient plasma, inspection of the effects of platelet- and recombinant TFPI on thrombin generation showed that, on a concentration basis, platelet and recombinant TFPI exhibit similar anticoagulant activities.

[0262] JBT1837, JBT1857, JBT1837+JBT1857, and JBT2547 (50 nM) blocked the ability of platelet TFPI to inhibit TF-FVIIa-catalysed FX activation . The fusion peptide was most effective and completely blocked the anticoagulant activity of TFPI. JBT1857 was the least effective followed by JBT1837 and the mixture of JBT1837 and JBT1857. Titration with JBT2547 demonstrated that a concentration of 10 nM almost completely inhibited the activity of platelet TFPI.

[0263] An additional study was performed to further elucidate the effects of plasma- and platelet-TFPI on the inhibition of thrombin generation in platelet-rich plasma (PRP) and platelet-poor plasma (PPP). To discriminate between the down-regulation of thrombin generation by platelet TFPI and plasma-derived TFPI, experiments were performed in TFPI-depleted plasma supplemented with platelets (source of platelet TFPI) and/or varying amounts of added purified TFPI (simulating variation of plasma TFPI levels). A preliminary experiment in TFPI-depleted plasma supplemented with convulxin-activated platelets showed that anti-TFPI antibodies enhanced thrombin generation, which indicated that TFPI released from the platelets inhibits thrombin generation.

[0264] It appeared that thrombin generation in PRP required TF. In PRP without CTI and in PRP to which CTI was added later, the ETP and thrombin peak height were hardly affected by the TF concentration (TF was not required) and only the lag time was shortened at increasing TF. When PRP was prepared from blood that was collected in citrate plus CTI, both the lag time and the thrombin peak were affected by TF concentration, indicating that experiments performed in PRP require that blood has to be collected in CTI.

[0265] In the experiments described above, the plasma was pre-activated with convulxin to promote the release of platelet TFPI before initiating thrombin generation with TF plus Ca^{2+}. Since convulxin is not a physiological trigger, a comparison of thrombin generation in PRP with non-activated platelets and platelets pre-activated with convulxin, collagen and Ca-ionophore was performed. It appeared that the thrombin-generating capacity of the platelets increased in the order no activator = ionophore < collagen < convulxin.

[0266] Additionally, in the experiments described above, the platelets were preincubated with activator (pre-

activated) for 7 minutes in the presence of TF, and thrombin generation was initiated with a fluorogenic substrate/Ca^{2+} mixture. When the platelet activator convulxin is added to the PRP 10-15 seconds before initiating thrombin generation with the fluorogenic substrate/Ca^{2+} mixture, substantially less thrombin is generated than in PRP in which the platelets were pre-activated with convulxin before thrombin generation was started. In view of these results, future studies will not include platelet pre-activation, and collagen or no activator will be primarily used in experiments in PRP.

[0267] The effects of TFPI antagonists were initially tested in PRP prepared from blood collected on CTI. Thrombin generation triggered in PRP with 0.1 pM TF + collagen was substantially enhanced by both an anti-TFPI cocktail and JBT2547. This shows that TFPI is a major regulator of thrombin generation on platelets, and that JBT2547 is as effective as an anti-TFPI cocktail in neutralizing the anticoagulant activity of TFPI. TFPI hardly downregulated thrombin generation in PRP triggered with a high amount (10 pM) of TF.

[0268] The major part of the thrombin generated in PRP at low TF (1 pM) appeared be formed via the intrinsic Xase and, thus, requires FVIII. Both JBT2547 and an anti-TFPI cocktail substantially increased thrombin generation in PRP in which FVIII was neutralized with anti-FVIII, a condition representative for hemophilia plasma. JBT2547 also enhanced thrombin generation in TFPI-depleted plasma supplemented with platelets that were not activated or that were activated with collagen.

[0269] The experiments with JBT2547 were carried out with 4 µM peptide, which, considering the affinity of JBT2547 for TFPI (sub-nM range), likely is a large excess. Indeed, both in PPP and in PRP, 5-10 nM JBT2547 was sufficient to fully inhibit TFPI.

[0270] To test whether TFPI in plasma, either plasma- or platelet-derived, is fully inhibited by the TFPI antibody cocktail or JBT2547, a TFPI titration was performed in PPP and in PRP in the absence and presence of a TFPI antibody cocktail. Increasing TFPI concentrations on top of the TFPI that is present in plasma plus what is released from platelets gradually inhibited thrombin generation. In the presence of an TFPI antibody cocktail, all thrombin generation curves were the same, indicating that the TFPI antibody cocktail is not a partial TFPI inhibitor but fully neutralizes TFPI.

[0271] In PPP, low concentrations of TFPI added (0.05 - 0.5 nM) substantially inhibited thrombin generation and plots of thrombin generation parameters (log lag time or In ETP) as a function of total TFPI (plasma TFPI + added TFPI) could be fitted to a straight line assuming that plasma contained ~ 0.3 nM TFPI. This indicated that the anticoagulant activity of added TFPI is similar to that of the TFPI that is present in plasma. In PRP, much larger amounts of added TFPI were required to inhibit thrombin generation.

[0272] The peptide complex JBT2547 enhanced thrombin generation 3-4-fold in PPP with an effective concentration (EC_{50}) of 2 nM and enhanced thrombin generation 2-fold in PRP with an EC_{50} of 8 nM. Thrombin generation in PRP to which an inhibitory FVIII antibody was added to simulate PRP of a hemophilia patient was enhanced 3-fold by the peptide complex. The data described herein establish that peptides of the invention bind plasma-derived, recombinant, and platelet-derived TFPI, and enhance thrombin generation by blocking the anticoagulant effects of platelet and plasma-TFPI.

Example 24

[0273] This example describes the hemostatic effect of peptides of the invention in a murine model of hemophilic joint bleeding.

[0274] The goal of the study was to evaluate the effect of an exemplary PEGylated peptide of the invention

on puncture-induced hemarthrosis in a murine model of hemophilia A. Peptide JBT2329 was administered at a dose of 1 mg/kg to FVIII-deficient mice with severe hemophilia (E16 FVIII B6; 129S4-F8 $^{tm\ I\ kaz}$/J) via intravenous tail vein injection. Various doses of recombinant FVIII (ADVATE; 10, 50, and 100 IU/kg) also were administered alone or in combination with JBT2329. After each product administration, the right knee joint capsule was punctured with a 30 gauge needle to induce hemorrhage. Animals were sacrificed three days following injury, and bleeding was assessed via gross and histological methods. The animal model and methods of evaluating bleeding are further described in Hakobyan et al., Haemophilia, 14, 804-809 (2008). Summary bleeding scores (SBS), which include visual and histological bleeding scores, were assigned to each joint. Administration of a PEGylated peptide of the invention prior to injury significantly reduced joint bleeding. The protective effect of 1 mg/kg of JBT2329 was significantly greater than the effect of 10 IU/kg ADVATE, but less than 100 IU/kg ADVATE, as determined by SBS. There was no significant difference in the protective effect conferred by 1 mg/kg JBT2329 and 50 IU/kg ADVATE. The results described herein further confirm that peptides of the invention provide a prophylactic or protective effect against bleeding in an *in vivo* model of hemophilia A.

Example 25

[0275] This example describes the impact of peptides of the invention on TFPI's ability to bind receptors that mediate clearance and cellular degradation. The pharmacokinetics of peptide-bound TFPI also is described.

[0276] The binding full length TFPI (fl-TFPI) to low density lipoprotein receptor-related protein (LRP) was studied using a BIAcore T200™ surface plasmon resonance assay (GE Healthcare, Chalfont St. Giles, UK) at 37°C. LRP was biotinylated using a biotinylation kit according to the manufacturer's protocol (Thermo Scientific). Following Neutravidin immobilization (2500 RU) to a Series S Sensor chip C1 (GE Healthcare) using standard amine coupling chemistry according to manufacturer's protocols, biotinylated recombinant human LRP-1 Cluster II Fc Chimera protein (R&D Systems) was bound to the surface via biotin - NeutrAvidin interactions resulting in 450 RUs. Following immobilization, fl-TFPI was injected at a flow rate of 30 µL/min in a single concentration of 10 nM diluted in running buffer (HBS-N, 0.1% P80, 5mM $CaCl_2$). fl-TFPI was subsequently dissociated by changing the flow to running buffer conditions. When interaction of fl-TFPI and LRP was studied in the presence of peptides (JBT1837, JBT1857, JBT2329 (JBT1857+40 kD PEG), or JBT2547 (JBT1837+JBT1857)) or polyethylene glycol (40 kD), 1 µM final concentration of the peptide or PEG was added to fl-TFPI.

[0277] TFPI interacted efficiently with immobilized LRP with fast on- and off-rates. Truncated TFPI lacking kunitz domain 3 (KD3) and the C-terminus does not interact with LRP. By visual inspection, it was apparent that fl-TFPI bound to JBT1837, JBT1857 or JBT2329 still interacts with LRP. TFPI complexed to the fusion peptide JBT2547 resulted in a higher response than that observed for TFPI complexed with single peptides. The unchanged association and dissociation kinetics of the LRP-fl-TFPI interaction in the presence and absence of a peptide complex (JBT2547) suggests that differing results are associated with the increased molecular weight of the fl-TFPI-JBT2547 complex and potential simultaneous binding of LRP and peptide to fl-TFPI by JBT2547. A slight decrease in fl-TFPI LRP interaction was observed when TFPI was bound to JBT2329 (PEGylated JBT1857), whereas a 40 kD PEG alone does not influence the assay system, suggesting that a highly hydrated PEG- linked peptide slightly interferes with fl-TFPI LRP interaction.

[0278] TFPI interaction with asialoglycoprotein receptor (ASGPR) also was evaluated using a BIAcore 3000™ surface plasmon resonance assay (GE Healthcare, Chalfont St. Giles, UK) at 25°C. Following immobilization of ASGPR (Novus Biologicals, 470 RU) to a Series S Sensor chip CM5 (GE Healthcare) using standard amine coupling chemistry according to manufacturer's protocols, fl-TFPI was injected at a flow rate of 30 µL/min in the single cycle analysis mode at concentrations ranging from 3.84 nM to 150 nM diluted in

running buffer (HBS pH 7.4, 0.1% P80, 5mM $CaCl_2$). fl-TFPI was subsequently dissociated by changing the flow to running buffer conditions. When interaction of fl-TFPI and ASGPR was studied in the presence of JBT1837, JBT1857, JBT2329, or JBT2547, 1 µM final concentration of peptide was added to the fl-TFPI.

[0279] Fl-TFPI interacted efficiently with immobilized ASGPR with fast on- and off-rates. C-terminally truncated TFPI does not interact with ASGPR. It was apparent from visual inspection that fl-TFPI bound to JBT1857 or JBT2547 interacted with ASGPR. Like LRP, TFPI complexed to the fusion peptide JBT2547 generated in a higher response. Also similar to the interaction of fl-TFPI to LRP, JBT2329 diminished the ASGPR-fl-TFPI interaction, whereas a 40 kDa PEG alone did not influence the assay system. The data suggests that highly hydrated PEG-linked peptides of the invention interfere with TFPI-ASGPR interaction via steric hindrance.

[0280] To study the impact of TFPI-binding peptides on the pharmacokinetics of TFPI in vivo, groups of mice (C57B16, male, 20 - 25 g) were treated with either human fl-TFPI (775 nM, 5 mL/kg, i.v.), human fl-TFPI complexed to a 10-fold molar excess of JBT2528 (775 nM hu fl-TFPI, 7752 nM JBT2528, 5 mL/kg, i.v.) or human fl-TFPI complexed to a 10-fold molar excess of JBT2534 (775 nM hu fl-TFPI, 7752 nM JBT2534, 5 mL/kg, i.v.). JBT2528 has the same peptide sequence as JBT2534, but lacks the 40 kD PEG-modification. As such, JBT2528 serves as a control for a possible impact of PEGylation in the assay. At various time points, three mice were sacrificed, blood was taken by heart puncture, and plasma was isolated and stored frozen (< -60°C) for further analysis. The sampling time points were as follows: fl-TFPI, 0.5, 1, 2 minutes; fl-TFPI-JBT2528, 0.5, 1, 2, 3, 5, 8 minutes; and fl-TFPI-JBT2534, 1, 2, 5, 10, 20, 35 minutes.

[0281] Plasma samples were analyzed for human TFPI with an ELISA which is specific for human TFPI. For quantification, wells of a microtiter plate (Nunc Maxisorp) were coated with 1 µg/mL of a monoclonal anti-human KD2 specific TFPI antibody (Sanquin, White label; MW1845) overnight at 4°C, followed by three wash cycles with TBS containing 0.1 % Tween 20 (TBST). Wells were blocked for 1 hour at room temperature with TBS containing 2 % non-fat dry milk (BioRad). 100 µL of diluted sample were applied to the wells and incubated for 2 hours at room temperature. After washing with TBST (5x), wells were incubated for 1 hour with 0.5 µg/mL of a polyclonal rabbit anti hTFPI antibody (ADG72; American Diagnostica), washed 5x with TBST, and incubated further for 1 hour with 0.2 µg/mL of a goat anti rabbit HRP labelled antibody (A0545; Sigma). Color was developed by addition of 100 µL of substrate (SureBlue TMP, KLP) and stopping with 50µL of 1 M HCl. Absorbance at 450 nm was measured with a microtiter plate reader (Thermo Appliskan Reader). Purified endogenous fl-TFPI, expressed by SKHep cells, was used as standard protein at concentrations of 0.25-16 ng/ml for quantification (Baxter Innovations GmbH). Plasma samples were diluted from 1/20 to 1/800 depending on the expected human TFPI concentration.

[0282] Human fl-TFPI had a very short half-life and very poor *in vivo* recovery. At the earliest time point (0.5 min), only one tenth of the expected TFPI level was observed. The recovery and the half-life of fl-TFPI remained unchanged when fl-TFPI was complexed with the tested peptides. This experiment indicates that PEGylation of a TFPI-binding peptide likely does not confer a longer half-life to fl-TFPI and does not affect TFPI clearance.

[0283] This example demonstrates that representative peptides of the invention do not significantly diminish TFPI interaction with clearance receptors and do not increase the half life of TFPI *in vivo.*

Example 26

[0284] This example demonstrates the ability of peptides of the invention to inhibit proteolytic degradation of TFPI.

[0285] TFPI is proteolytically inactivated by several enzymes, including elastase, thrombin, plasmin, FXa, and chymase. See, e.g., Hamuro et al., FEBS Journal, 274, 3056-3077 (2007). The impact of fusion protein JBT2547, as well as the individual peptide subunits JBT1837 and JBT1857, on the proteolytic degradation of fl-TFPI by neutrophil elastase was studied. Neutrophil elastase is a representative protease cleaving TFPI within Lys86 and Gln90 located between the Kunitz 1 and Kunitz 2 domains.

[0286] For proteolytic digest of fl-TFPI, 5 nM fl-TFPI were incubated with 5 nM neutrophil elastase (purified from human neutrophils, Calbiochem) in a reaction buffer (50 mM Tris, 300 mM NaCl, 10 mM $CaCl_2$, pH 7.5) at 37°C. The reaction was performed with and without 1 µM JBT1837, JBT1857, or JBT2547. Proteolysis was monitored by Western blot analysis. Aliquots were taken from the reaction mixture after 0, 5, 15, 30, and 60 minutes and immediately heated at 96°C for 5 minutes in SDS-loading buffer at reducing conditions. Samples were separated on a 4-20% Tris-glycine SDS-polyacrylamide gel, and the proteins were transferred to a PVDF membrane. After blocking the membrane with non-fat dry milk, TFPI proteins were detected with a rabbit polyclonal antibody against human TFPI (AF2974, R&D Systems) and a secondary anti-rabbit-HRP conjugated antibody (Sigma). SuperSignal West Femto chemiluminescent substrate (Thermo Scientific) was applied to develop a chemiluminescent signal that was captured on film. The developed bands were quantified by densitometry.

[0287] In the absence of peptides, gradual degradation of fl-TFPI by neutrophil elastase was observed within 1 hour. The ~43 kD band visualized in the Western Blot corresponding to intact fl-TFPI almost completely disappeared, while two cleavage products were formed by cleavage of the peptide bond T87-T88. When JBT2547 was present, the proteolysis of fl-TFPI was blocked during the monitored time, indicating that JBT2547 protects fl-TFPI from degradation by elastase *in vitro.* The individual peptide subunits mediated different effects on fl-TFPI cleavage. JBT1837 protected fl-TFPI from proteolysis for one hour, similar to the results observed for JBT2547. In contrast, JBT1857 had only a minor or no effect on elastase cleavage.

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Patentkrav

1. TFPI-bindende peptidkompleks omfattende et første peptid og et andet peptid, hvor

(a) det første peptid omfatter strukturen med formel (XIII):

X6001-X6002-X6003-X6004-X6005-X6006-X6007-X6008-X6009-X6010-X6011-X6012-X6013-X6014-X6015-X6016-X6017-X6018-X6019-X6020 (XIII) (SEQ ID NO: 3153);

hvor X6001 er en aminosyre valgt fra gruppen bestående af F, L, M, Y, 1Ni, Thi, Bta, Dopa, Bhf, C, D, G, H, I, K, N, Nmf, Q, R, T, V, og W;

hvor X6002 er en aminosyre valgt fra gruppen bestående af Q, G, og K;

hvor X6003 er en aminosyre valgt fra gruppen bestående af C, D, E, M, Q, R, S, T, Ede(O), Cmc, A, Aib, Bhs, F, G, H, I, K, L, N, P, V, W og Y;

hvor X6004 er en aminosyre valgt fra gruppen bestående af Aib, E, G, I, K, L, M, P, R, W, Y, A, Bhk, C, D, F, H, k, N, Nmk, Q, S, T og V;

hvor X6005 er en aminosyre valgt fra gruppen bestående af a, A, Aib, C, D, d, E, G, H, K, k, M, N, Nmg, p, Q, R, NpropylG, aze, pip, tic, oic, hyp, nma, Ncg, Abg, Apg, thz, dtc, Bal, F, L, S, T, V, W og Y;

hvor X6006 er en aminosyre valgt fra gruppen bestående af A, C, C(NEM), D, E, G, H, K, M, N, Q, R, S, V, Cit, C(Acm), Nle, I, Ede(O), Cmc, Ecl, Eea, Eec, Eef, Nif, Eew, Aib, Btq, F, L, T, W og Y;

hvor X6007 er en aminosyre valgt fra gruppen bestående af I, V, T, Chg, Phg, Tie, A, F, G, K, L, Nmv, P, Q, S, W og Y;

hvor X6008 er en aminosyre valgt fra gruppen bestående af F, H, 1Ni, 2Ni, Pmy, Y, og W;

hvor X6009 er en aminosyre valgt fra gruppen bestående af Aib, V, Chg, Phg, Abu, Cpg, Tie, L-2-amino-4,4,4-trifluorbutanonsyre, A, f, I, K, S, T og V;

hvor X6010 er en aminosyre valgt fra gruppen bestående af A, C, D, d, E, F, H, K, M, N, P, Q, R, S, T, V, W, Y, Nmd, C(NEM), Aib, G, I, L og Nmf;

hvor X6011 er en aminosyre valgt fra gruppen bestående af A, a, G, p, Sar, c, hcy, Aib, C, K, og Nmg;

hvor X6012 er en aminosyre valgt fra gruppen bestående af Y, Tym, Pty, Dopa og Pmy;

hvor X6013 er en aminosyre valgt fra gruppen bestående af Aib, C, F, 1Ni, Thi, Bta, A, E, G, H, K, L, M, Q, R, W og Y;

hvor X6014 er en aminosyre valgt fra gruppen bestående af A, Aib, C, C(NEM), D, E, K, L, M, N, Q, R, T, V, Hcy, Bhe, F, G, H, I, P, S, W og Y;

hvor X6015 er en aminosyre valgt fra gruppen bestående af R, (omega-methyl)-R, D, E og K;

hvor X6016 er en aminosyre valgt fra gruppen bestående af L, Hcy, Hle og Aml;

hvor X6017 er en aminosyre valgt fra gruppen bestående af A, a, Aib, C, c, Cha, Dab, Eag, Eew, H, Har, Hci, Hle, I, K, L, M, Nle, Nva, Opa, Orn, R, S, Deg, Ebc, Eca, Egz, Aic, Apc, Egt, (omega-methyl)-R, Bhr, Cit, D, Dap, E, F, G, N, Q, T, V, W og Y;

hvor X6018 er en aminosyre valgt fra gruppen bestående af A, Aib, Hcy, hcy, C, c, L, Nle, M, N, R, Bal, D, E, F, G, H, I, K, Q, S, T, V, W og Y;

hvor X6019 er en aminosyre valgt fra gruppen bestående af K, R, Har, Bhk og V; og

hvor X6020 er en aminosyre valgt fra gruppen bestående af K, L, Hcy, Aml, Aib, Bhl, C, F, G, H, I, Nml, Q, R, S, T, V, W og Y
og

(b) det andet peptid omfatter strukturen med formel (XIV):

X7001-X7002-X7003-X7004-X7005-X7006-[X7007-X7008-X7009-X7010-X7011-X7012-X7013-X7014-X7015-X7016-X7017-X7018]-X7019-X7020-X7021-X7022-X7023 (XIV) (SEQ ID NO: 3154),

hvor X7001 er enten til stede eller fraværende, hvorved i tilfælde X7001 er det tilstede som en aminosyre valgt fra gruppen bestående af A, C, C(NEM), D, E, F, G, H, I, K, L, P, R, S, T, V og W;

hvor X7002 er enten til stede eller fraværende, hvorved i tilfælde X7002 er det tilstede som en aminosyre valgt fra gruppen bestående af A, C, C(NEM), D, E, F, G, H, I, K, L, M, P, Q, R, S, T, V, W og Y;

hvor X7003 er en aminosyre valgt fra gruppen bestående af A, F, I, K, L, R, S, T, V, W og Y;

hvor X7004 er en aminosyre valgt fra gruppen bestående af A, D, E, F, G, I, K, L, R, S, T, V og W;

hvor X7005 er R eller W;

hvor X7006 er en aminosyre valgt fra gruppen bestående af F, H, I, K, L, R, V og W;

hvor X7007 er en aminosyre valgt fra gruppen bestående af Orn, homoK, C, Hcy, Dap og K, fortrinsvis valgt fra gruppen bestående af C og Hcy;

hvor X7008 er en aminosyre valgt fra gruppen bestående af A, G, R, S og T;

hvor X7009 er en aminosyre valgt fra gruppen bestående af a, A, I, K, L, M, m, Moo, Nle, p, R, Sem og V;

hvor X7010 er en aminosyre valgt fra gruppen bestående af A, G, I, K, L, P, R, S, T og V;

hvor X7011 er en aminosyre valgt fra gruppen bestående af D, E, G, S og T;

hvor X7012 er en aminosyre valgt fra gruppen bestående af A, a, D, d, E, e, F, f, G, l, K, k, L, l, M, m, Moo, Nle, nle, P, p, R, r, S, s, Sem, T, t, V, v, W og w;

hvor X7013 er en aminosyre valgt fra gruppen bestående af A, C, C(NEM), Con, Con(Meox), D, d, E, e, Eag, F, G, I, K, L, N, R, S, s, T, V og W;

hvor X7014 er en aminosyre valgt fra gruppen bestående af A, D, E, F, G, I, K, L, M, R, S, T, V og W;

hvor X7015 er en aminosyre valgt fra gruppen bestående af A, D, E, F, G, I, K, L, M, Nle, R, S, T, V og W;

hvor X7016 er en aminosyre valgt fra gruppen bestående af A, D, E, F, I, K, L, M, Moo, Nle, R, S, Sem, T, V, W og Y;

hvor X7017 er en aminosyre valgt fra gruppen bestående af A, D, E, F, G, I, K, L, R, S, T, V, W og Y;

hvor X7018 er en aminosyre valgt fra gruppen bestående af C og D, fortrinsvis C;

hvor X7019 er en aminosyre valgt fra gruppen bestående af A, F, I, L, S, T, V og W;

hvor X7020 er en aminosyre valgt fra gruppen bestående af F og W; hvor X7021 er en aminosyre valgt fra gruppen bestående af I, L og V;

hvor X7022 er en aminosyre valgt fra gruppen bestående af A, D, E, F, G, I, K, L, P, R, S, T, V og W;

hvor X7023 er enten til stede eller fraværende, hvorved i tilfælde X7023 er det tilstede som en aminosyre valgt fra gruppen bestående af A, C, C(NEM), Con, Con(Meox), D, E, Eag, F, G, I, K, L, R, S, T, V, W og Y;

hvor peptidet omfatter som en cyklisk struktur genereret ved en binding mellem X7007 og X7018;

hvor C-terminus af det første peptid er bundet til N-terminus af det andet peptid via en linkermolekyldel;

hvor det første peptid omfatter en aminosyresekvens mindst 80%, mindst 85%, mindst 90%, mindst 95%, eller 100% identisk med SEQ ID NO: 178 eller SEQ ID NO: 4261; og

hvor det andet peptid omfatter en aminosyresekvens mindst 80%, mindst 85%, mindst 90%, mindst 95%, eller 100% identisk med SEQ ID NO: 1044.

2. Peptidkomplekset ifølge krav 1,
hvor X6001 er en aminosyre valgt fra gruppen bestående af 1Ni, Bta, Dopa, F, L, Y og M;
hvor X6002 er Q;
hvor X6003 er en aminosyre valgt fra gruppen bestående af D, E, S, M, Q, R, T og C;
hvor X6004 er en aminosyre valgt fra gruppen bestående af K, Aib, L, P, R, E, G, I, Y, M og W;
hvor X6005 er en aminosyre valgt fra gruppen bestående af p, Nmg, NpropylG, aze, pip, tic, oic, hyp, a, Aib, D, d, G, H, K, k, N, Q, R, A, E, C og M;
hvor X6006 er en aminosyre valgt fra gruppen bestående af C, E, K, R, S, V, C(Acm), Nle, C(NEM), I, Cit, A, D, G, H, Q og M;
hvor X6007 er en aminosyre valgt fra gruppen bestående af Tle, V og I; hvor X6008 er en aminosyre valgt fra gruppen bestående af H, 1Ni, 2Ni, Pmy, F og Y;
hvor X6009 er V, Abu eller Tle;
hvor X6010 er en aminosyre valgt fra gruppen bestående af D, P, C, T, A, E, K, M, N, Q, R, F, H, S, V, W og Y;
hvor X6011 er G, a, c, hcy eller Sar;
hvor X6012 er Y;
hvor X6013 er en aminosyre valgt fra gruppen bestående af F, 1Ni, Bta og C; hvor X6014 er en aminosyre valgt fra gruppen bestående af Aib, C, E, Hcy, A, D, K, L, M, N, Q, R, T, V og Aib;
hvor X6015 er R;

hvor X6016 er en aminosyre valgt fra gruppen bestående af L, Aml, Hle og Hcy; hvor X6017 er en aminosyre valgt fra gruppen bestående af A, Aib, C, c, Aic, Eca, Deg, Cha, Dab, Dap, Eag, Eew, H, Har, Hci, Hle, K, Nle, Nva, Opa, Orn, R, I, L, S og M; hvor X6018 er en aminosyre valgt fra gruppen bestående af A, Aib, C, c, L, Hcy, N, M og R;
hvor X6019 er K; og
hvor X6020 er en aminosyre valgt fra gruppen bestående af L, Aml, Hcy og K.

3. Peptidkomplekset ifølge krav 1 eller krav 2, hvor det første peptid og/eller andet peptid yderligere omfatter N-terminale aminosyre(r) og/eller molekyldele bundet til X6001 og/eller X7001 og valgt fra gruppen bestående af FAM-Ttds, PE, Palm, 2-phenylacetyl, 3-phenylpropionyl, 2-(naphth-2-yl) acetyl, hexanoyl, 2-methylpropionyl, 3-methylbutanoyl, 2-naphthylsulfonyl, 1-naphthylsulfonyl, acetyl, Con, Con(Meox), AOA, Oxme-AOA, Meox-Lev, levulinsyre (Lev), og pentinsyre (Pyn).

4. Peptidkomplekset ifølge et hvilket som helst af kravene 1-3, hvor det første peptid og/eller andet peptid yderligere omfatter X6021 bundet til X6020 eller X7024 bundet til X7023, respektivt, hvor X6021 og/eller X7024 omfatter C-terminal aminosyre(r) og/eller molekyldele valgt fra gruppen bestående af Hly, K, Orn, Dab, Dap, Eag, Hcy, Pen, C, c, C(NEM), Con, Con(Meox), K(Ttds-maleimidopropionyl(EtSH)), K(Tdts-maleimid), K(AOA), K(Myr), K(Ttds-Myr), K(Ttds-Palm), K(Ttds-Ac), K(Ttds-yGlu-Myr), K(AlbuTag), K(4PBSA), Cea, og amid.

5. Peptidkomplekset ifølge et hvilket som helst af kravene 1-4,
hvor X7001 er en aminosyre valgt fra gruppen bestående af A, D, F, G, H, K, L og S;
hvor X7002 er en aminosyre valgt fra gruppen bestående af H, F, M og R;
hvor X7003 er en aminosyre valgt fra gruppen bestående af F og Y;
hvor X7004 er K;
hvor X7005 er W;
hvor X7006 er en aminosyre valgt fra gruppen bestående af F og H;
hvor X7007 er C;
hvor X7008 er en aminosyre valgt fra gruppen bestående af A, G og S;

hvor X7009 er en aminosyre valgt fra gruppen bestående af M, Sem og V;
hvor X7010 er en aminosyre valgt fra gruppen bestående af K, P og R;
hvor X7011 er D;
hvor X7012 er en aminosyre valgt fra gruppen bestående af F, L, I, M og Sem;
hvor X7013 er en aminosyre valgt fra gruppen bestående af D, G, K og S;
hvor X7014 er G;
hvor X7015 er en aminosyre valgt fra gruppen bestående af I og T;
hvor X7016 er en aminosyre valgt fra gruppen bestående af D, F, M, Sem og Y;
hvor X7017 er en aminosyre valgt fra gruppen bestående af S og T;
hvor X7018 er C;
hvor X7019 er en aminosyre valgt fra gruppen bestående af A og V;
hvor X7020 er W;
hvor X7021 er V;
hvor X7022 er en aminosyre valgt fra gruppen bestående af F, L, K, R, P og W;
hvor X7023 er enten til stede eller fraværende, hvorved i tilfælde X7023 er det tilstede som en aminosyresekvens valgt fra gruppen bestående af A, D, F, M, S og Y; og
hvor peptidet omfatter en cyklisk struktur genereret ved en binding mellem X7007 og X7018.

6. Peptidkomplekset ifølge et hvilket som helst af kravene 1-5 hvor linkermolekyldelen er 1-100 Å i længde, 5-50 Å i længde, eller 10-30 Å i længde.

7. Peptidkomplekset ifølge krav 6, hvor linkermolekyldelen omfatter strukturen Z1-20, hvor Z er en aminosyre, hydroxysyre, ethylenglycol, propylenglycol, eller en kombination af hvilke som helst af de foregående.

8. Peptidkomplekset ifølge krav 7, hvor Z er G, s, S, a, A, Bal, Gaba, Ahx, Ttds, eller en kombination af hvilke som helst af de foregående.

9. Peptidkomplekset ifølge krav 1 omfattende aminosyresekvensen som beskrevet i SEQ ID NO: 4260.

10. Peptidkomplekset ifølge krav 1, hvor det andet peptid omfatter aminosyresekvensen som beskrevet i SEQ ID NO: 1334.

11. Peptidkomplekset ifølge et hvilket som helst af kravene 1-10, hvor peptidkomplekset er konjugeret til en polyethylenglycol (PEG)-molekyldel, human serum albumin (HSA), et HSA-bindende domæne, et antistof eller fragment deraf, hydroxyethylstivelse, en prolin-alanin-serin multimer (PASylation), en C12-C18 fedtsyre, eller polysialinsyre.

12. Peptidekompleks ifølge et hvilket som helst af kravene 1-11 til anvendelse i en fremgangsmåde til behandlingen af et individ.

13. Peptidkomplekset til anvendelse ifølge krav 12, hvor fremgangsmåden er til behandlingen af en blodkoagulationslidelse.

14. Farmaceutisk sammensætning omfattende peptidkomplekset ifølge et hvilket som helst af kravene 1 til 11 og en farmaceutisk acceptabel bærer, og eventuelt yderligere omfattende et yderligere farmaceutisk effektivt middel.

DRAWINGS

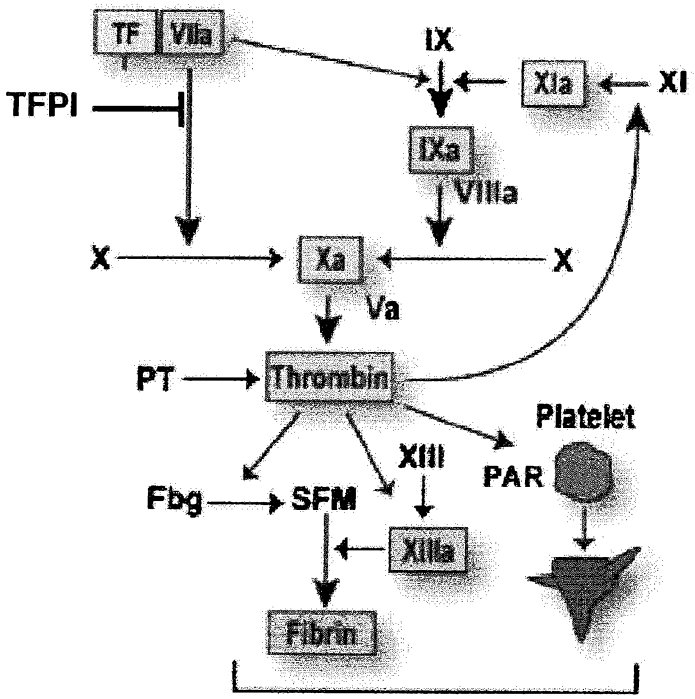

FIGURE 1

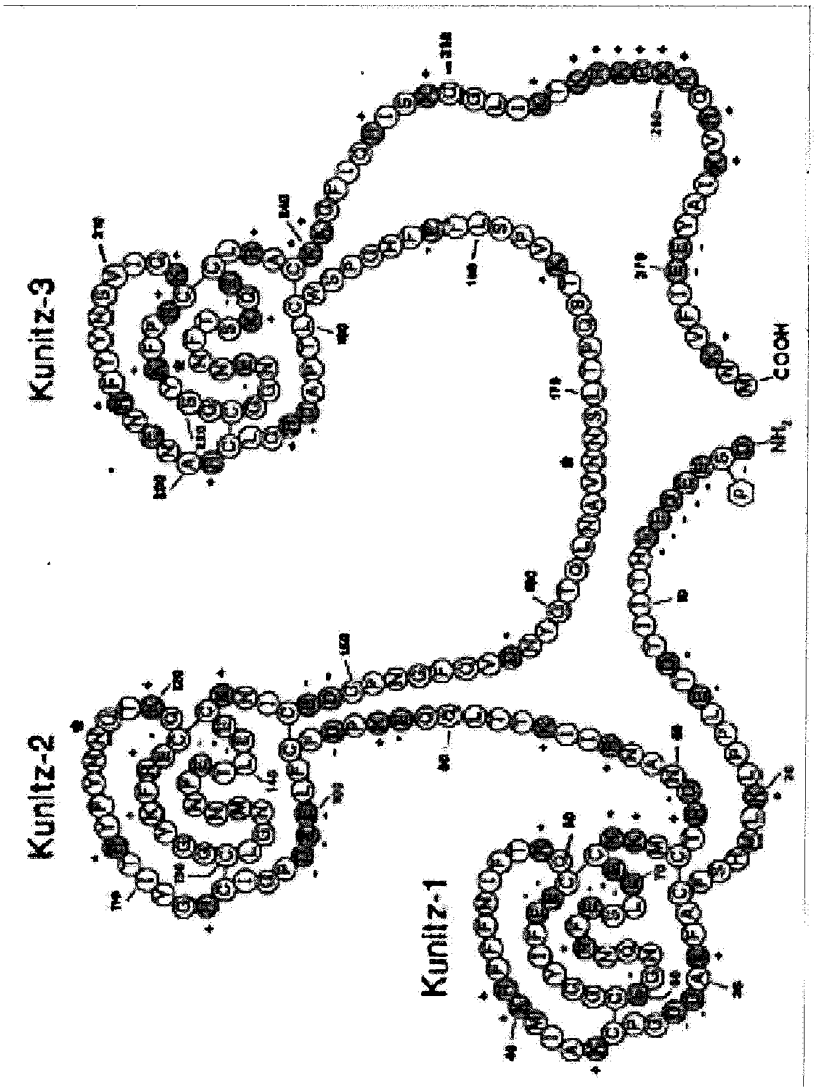

FIGURE 2

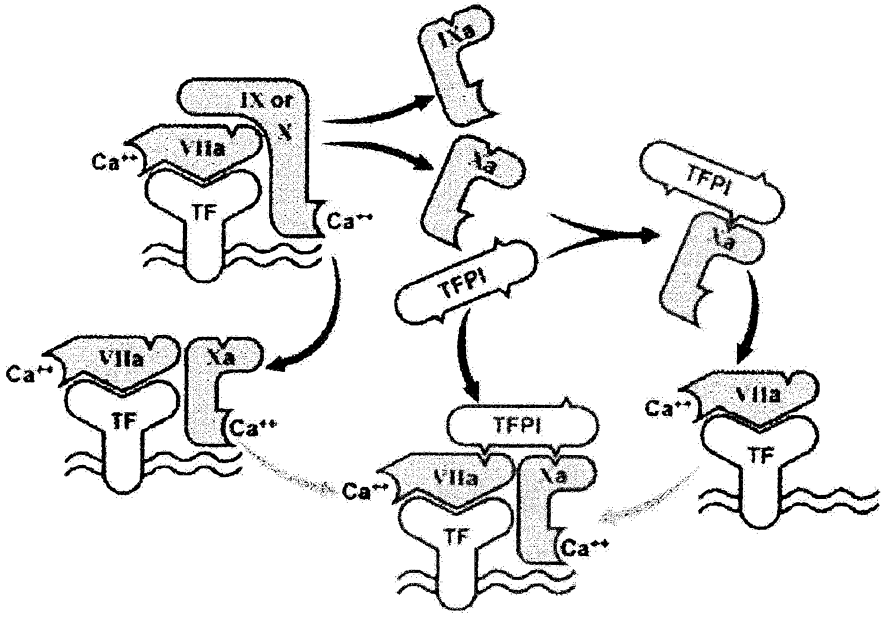
IXa
IX or X
VIIa
Ca++
TF
Ca++
Xa
TFPI
TFPI
Xa
VIIa
Ca++
TF
Ca++
TFPI
VIIa
Xa
Ca++
TF
Ca++
VIIa
Ca++
TF

FIGURE 3

Base Sequence JBT0293	FQSKKNVFVFGYFERLRAKL	(SEQ ID NO: 256)
JBT0294	Ac-**Y**QSKKNVFVFGYFERLRAKL-NH2	(SEQ ID NO: 257)
JBT0295	Ac-F**S**SKKNVFVFGYFERLRAKL-NH2	(SEQ ID NO: 713)
JBT0296	Ac-FQ**N**KKNVFVFGYFERLRAKL-NH2	(SEQ ID NO: 407)
JBT0297	Ac-FQSK**N**NVFVFGYFERLRAKL-NH2	(SEQ ID NO: 183)
JBT0298	Ac-FQSK**Q**NVFVFGYFERLRAKL-NH2	(SEQ ID NO: 747)
JBT0299	Ac-FQSKKNVF**A**FGYFERLRAKL-NH2	(SEQ ID NO: 408)
JBT0300	Ac-FQSKKNVF**S**FGYFERLRAKL-NH2	(SEQ ID NO: 409)
JBT0301	Ac-FQSKKNVF**T**FGYFERLRAKL-NH2	(SEQ ID NO: 410)
JBT0302	Ac-FQSKKNVFV**A**GYFERLRAKL-NH2	(SEQ ID NO: 258)
JBT0303	Ac-FQSKKNVFV**D**GYFERLRAKL-NH2	(SEQ ID NO: 184)
JBT0304	Ac-FQSKKNVFV**L**GYFERLRAKL-NH2	(SEQ ID NO: 259)
JBT0305	Ac-FQSKKNVFV**Q**GYFERLRAKL-NH2	(SEQ ID NO: 260)
JBT0306	Ac-FQSKKNVFV**S**GYFERLRAKL-NH2	(SEQ ID NO: 185)
JBT0307	Ac-FQSKKNVFV**Y**GYFERLRAKL-NH2	(SEQ ID NO: 261)
JBT0308	Ac-FQSKKNVFVFGY**K**ERLRAKL-NH2	(SEQ ID NO: 411)
JBT0309	Ac-FQSKKNVFVFGY**Y**ERLRAKL-NH2	(SEQ ID NO: 412)
JBT0310	Ac-FQSKKNVFVFGYF**D**RLRAKL-NH2	(SEQ ID NO: 262)
JBT0311	Ac-FQSKKNVFVFGYFERLRAK**N**-NH2	(SEQ ID NO: 748)

FIGURE 4

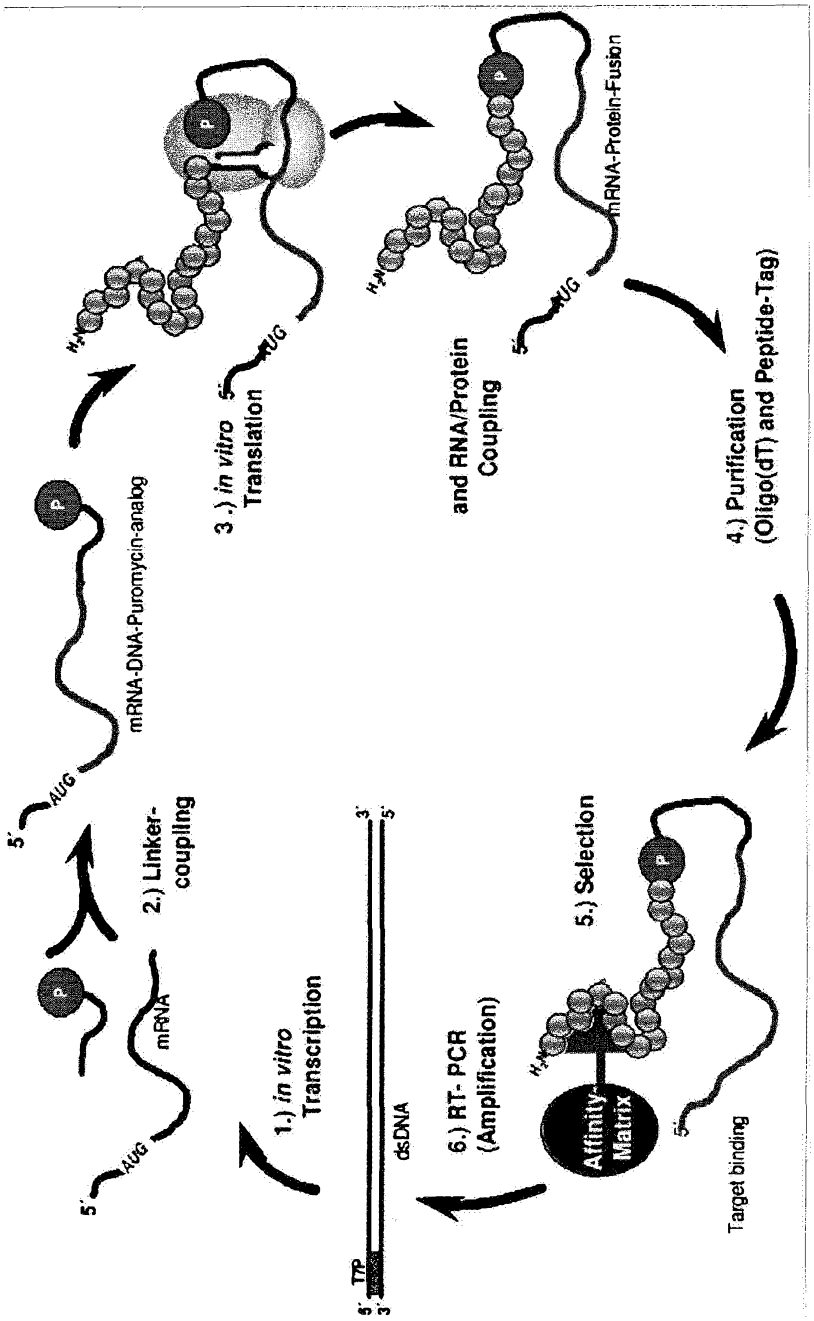
1.) in vitro Transcription
2.) Linker-coupling
3.) in vitro Translation
and RNA/Protein Coupling
4.) Purification (Oligo(dT) and Peptide-Tag)
5.) Selection
6.) RT- PCR (Amplification)
mRNA
mRNA-DNA-Puromycin-analog
mRNA-Protein-Fusion
dsDNA
Affinity-Matrix
Target binding
AUG
TTP

FIGURE 5

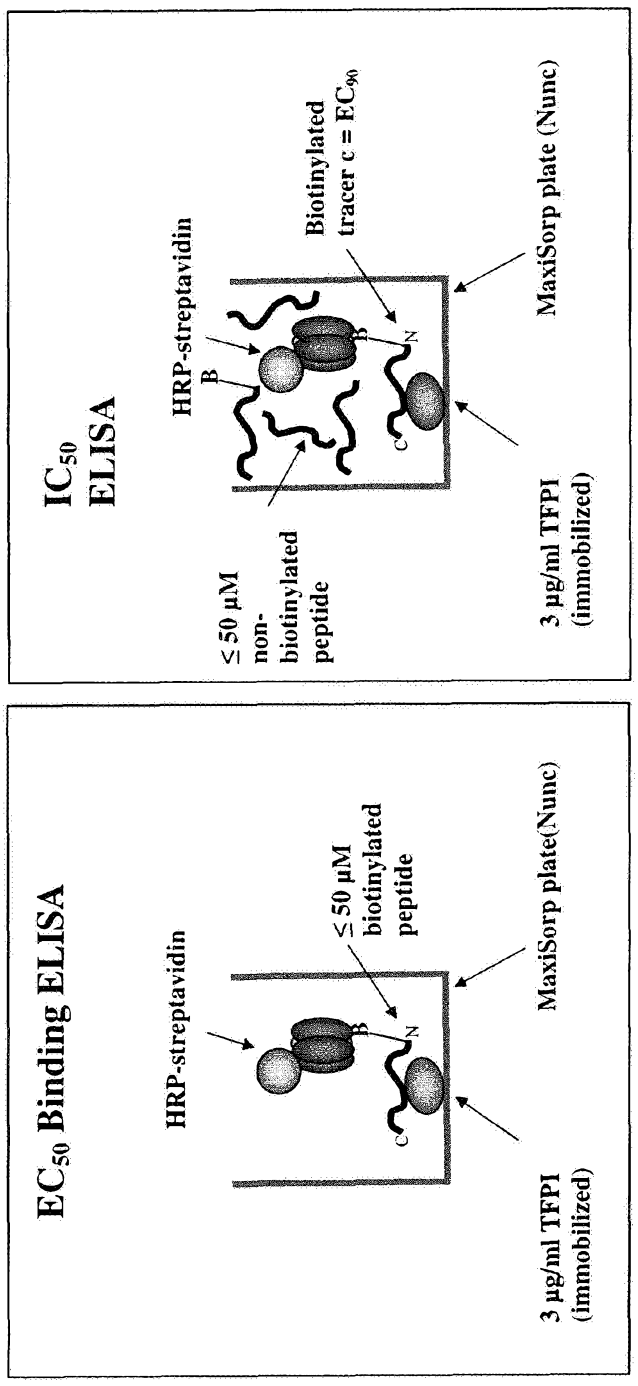

FIGURE 6A

FIGURE 6B

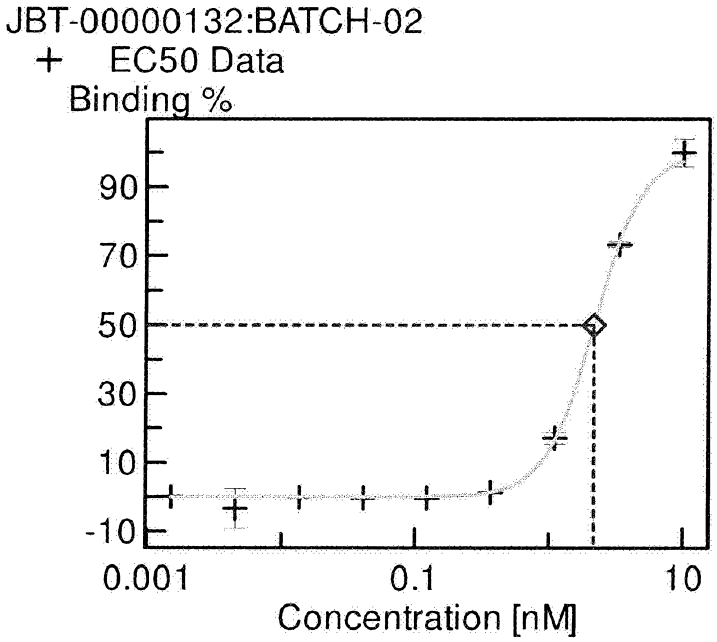
JBT-00000132:BATCH-02
EC50 Data
Binding %
90
70
50
30
10
-10
0.001
0.1
10
Concentration [nM]

FIGURE 7

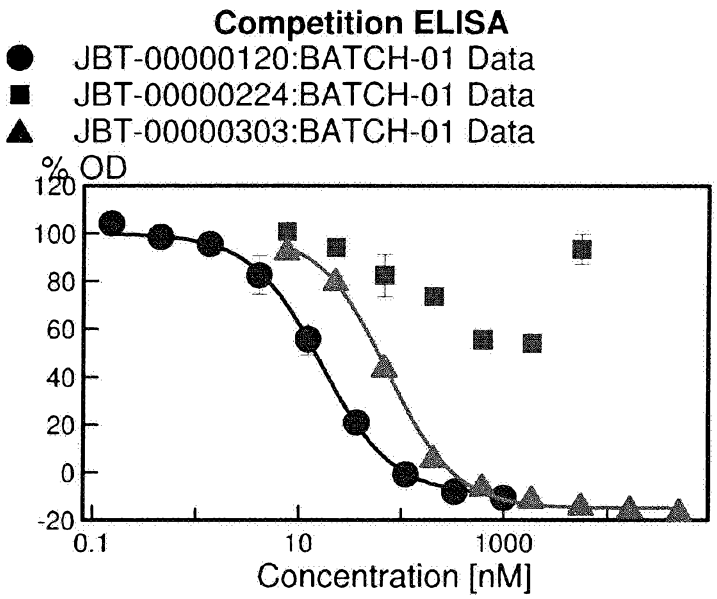
Competition ELISA
JBT-00000120:BATCH-01 Data
JBT-00000224:BATCH-01 Data
JBT-00000303:BATCH-01 Data
% OD
120
100
80
60
40
20
0
-20
0.1
10
1000
Concentration [nM]

FIGURE 8A

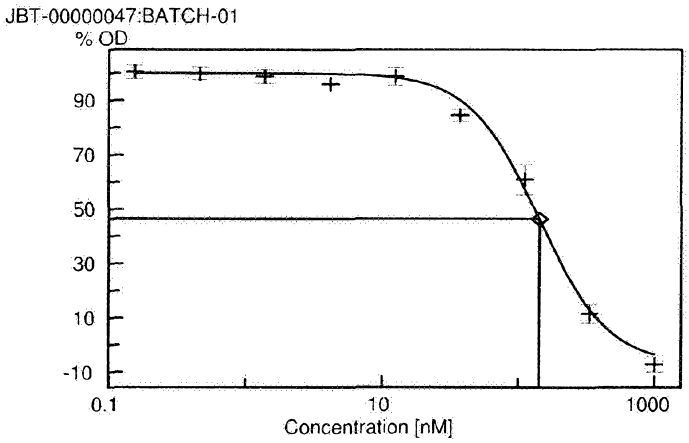

FIGURE 8B

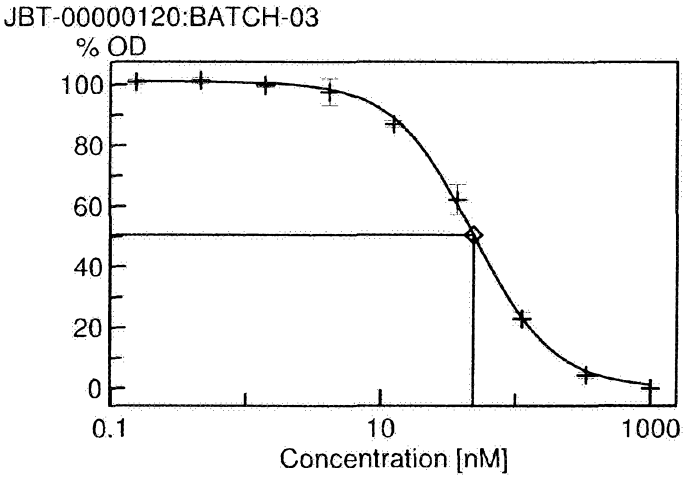

FIGURE 8C

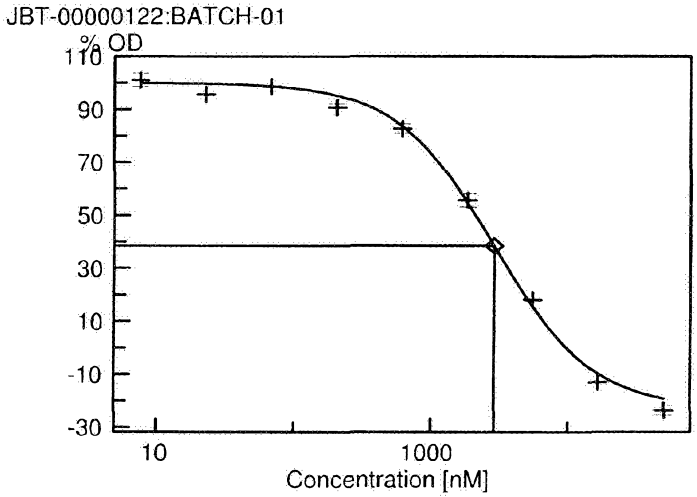

FIGURE 8D

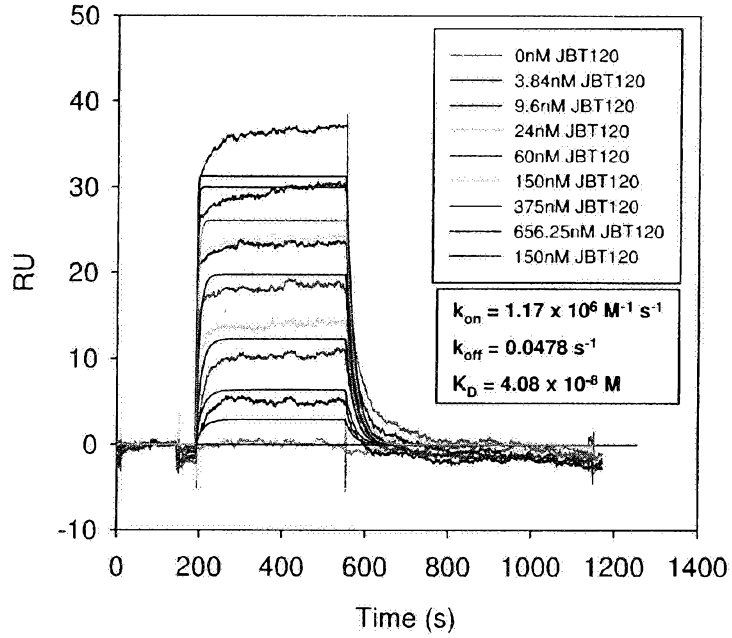

FIGURE 9A

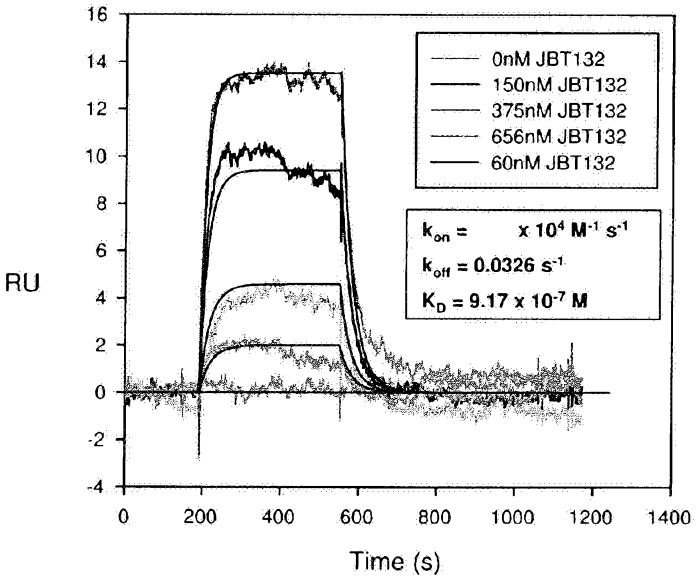

FIGURE 9B

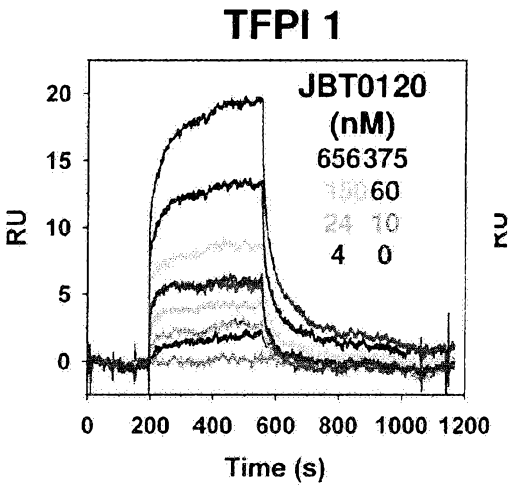

FIGURE 10A

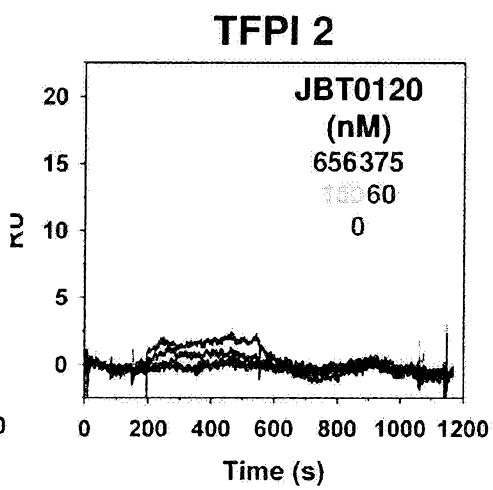

FIGURE 10B

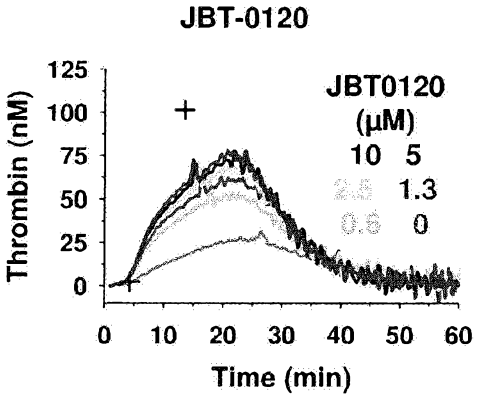

FIGURE 11A

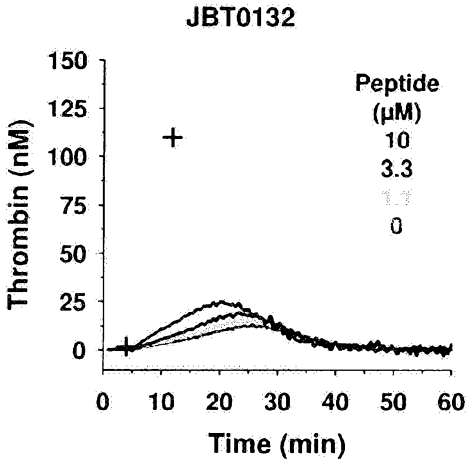

FIGURE 11B

FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0047	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-NH2 (SEQ ID NO: 253)	---		NA	---	NA	123	122	
		---		NA	1.23	NA			
		---		NA	---	NA			
		---		NA	2.98	16			
		0.72	82.4	68					
				NA					
		---		49					
JBT0047	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-NH2 (SEQ ID NO: 253)	---		NA	1.37	20	128	289	68
		1.33	63.10	35	1.46	26			
							160		
							155		
JBT0047	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-NH2*HCL (SEQ ID NO: 253)								
JBT0051	Biotin-Ttds-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-	---		NA	2.49	NA			

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
	NH2 (SEQ ID NO: 962)								
		0.53	86.9	66	1.04	19			
JBT0055	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-Ttds-Lysin(biotin)-NH2 (SEQ ID NO: 963)	---		NA	---	NA			
		---		NA	---	NA			
		---		NA	---	NA			
		---		NA		18			
		0.53	88.4	72	1.67	25			
		---		NA					
JBT0055	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-Ttds-Lysin(biotin)-NH2 (SEQ ID NO: 963)	---		NA					
JBT0131	Biotin-Ttds-AFQSKKNVFVFGYFERLRAK-NH2 (SEQ ID NO: 964)	---		NA	2.02	11	65	173	
		8.20	77.80	23					
JBT0131	Biotin-Ttds-AFQSKKNVFVFGYFERLRAK-NH2*HCl (SEQ ID NO: 964)	10.52	72.06	12	1.67	14	106	45	
		---		14	2.96	14			
JBT0132	Biotin-Ttds-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 965)	---		NA	0.06	10	41	102	

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	SEQUENCE	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
		---		44	---	16	28		
		1.15	49.37	36					
JBT0132	Biotin-Ttds-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 965)	---		23		11	109		
		---		27	---	13	123	53	
							112		
JBT0132	Biotin-Ttds-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 965)	---		30	---	11	117		
JBT0133	Biotin-Ttds-QSKKNVFVFGYFERLRAKLT-NH2 (SEQ ID NO: 966)	---		NA	0.34	9	28	67	
		---		15					
JBT0134	Biotin-Ttds-QSKKNVFVFGYFERLRAK-NH2 (SEQ ID NO: 967)	---		NA	---	3	11	--	
		---		5					
JBT0155	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKL TSKK-NH2 (SEQ ID NO: 8)	---		74	0.38	33	49		
		0.97	77.70	54			67		
JBT0156*	Ac-VVEKLTFVQLSFLNRRRFSQYAGFKGAGKV-NH2 (SEQ ID NO: 742)	---		0	---	18	--		

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
					1.49	14			
JBT0157*	Ac-RVFLYFSGKAGGLVKLVERQAFQTNVSKFR-NH2 (SEQ ID NO: 743)	---		0	---	12	--		
					---	9			
					2.77	9			
JBT0158	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKK K-NH2 (SEQ ID NO: 9)	---		34	---	23	30		
		4.26	88.20	35					
JBT0159	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLKK-NH2 (SEQ ID NO: 744)	---		0	---	18	inhib		
					---	22			
JBT0160	Ac-KKSGVGRLQVAFQSKKNVFVFGYFKK-NH2 (SEQ ID NO: 745)	---		0	---	22	inhib		
JBT0161	Ac-KKSGVGRLQVAFQSKKNVFVFKK-NH2 (SEQ ID NO: 746)	---		0	---	6	inhib		
JBT0162	Ac-KKGRLQVAFQSKKNVFVFGYFERLRAKLTSK K-NH2 (SEQ ID NO: 10)	---		68	0.28	34	54		
		0.74	70.70	54					

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0163	Ac-KKQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 11)	---		66	0.89	31	69		
		1.54	79.30	48					
JBT0164	Ac-KKFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 12)	---		55	2.68	24	39		
		---		50					
		1.74	71.24	43					
JBT0165	Ac-KKKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 712)	---		0	---	15	inhib		
JBT0166	Biotinyl-Ttds-KKFQSKKNVFVFGYFERLRAKLKK-NH2 (SEQ ID NO: 968)	---		71	1.95	36			
		---		58					
		2.60	85.60	50					
JBT0167*	Biotinyl-Ttds-GKNAKFYLFESLRQVKFVFR-NH2 (SEQ ID NO: 969)	---		0	---	21	--		
					---	19			
					---	15			
JBT0168*	Biotinyl-Ttds-YKFSFNKELFKQARLRFVGV-NH2 (SEQ ID NO: 970)	---		0	---	9	--		

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
					---	2			
JBT0169	Ac-KKAFQSKKNVFVFGYFERLRAKKK-NH2 (SEQ ID NO: 254)	---		16	2.35	20	21		
		---		21					
JBT0170	Ac-KKFQSKKNVFVFGYFERLRAKLKK-NH2 (SEQ ID NO: 13)	---		61	1.90	30	68		
		---		49					
		---		43					
JBT0171	Ac-KKQSKKNVFVFGYFERLRAKLTKK -NH2 (SEQ ID NO: 255)	---		25	4.12	13	33		
		---		29					
JBT0172	Ac-KKQSKKNVFVFGYFERLRAKKK-NH2 (SEQ ID NO: 406)	---		7	3.98	8	--		
		---		14					
JBT0173	Biotinyl-Ttds-SGVGRLQVAFQSKKNVFVFG-NH2 (SEQ ID NO: 971)	---		0		3			
JBT0174	Ac-KKAFQSKKNVFVFGYFERLRAKLKK-NH2 (SEQ ID NO: 14)	---		47	2.90	26	49		
		---		39					
JBT0175	Ac-KKAFQSKKNVFVFGYFERLRAKLTKK-NH2 (SEQ ID NO: 182)	---		45	2.29	26	49		

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
		2.71	78.67	40					
					---	1			
					---	3			
JBT0293	Ac-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 256)	---		28		12	119		
		1.32	40.6	27	2.93	14	147	71	
		1.69	44.11	27	---	13	135		
		---		29	---	15	132		
		1.94	39.5	22	---	16			
JBT0293	Ac-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 256)	---		27	1.62	16			
		5.37	78.1	26					
JBT0294	Ac-YQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 257)	---		46		12	122		
		4.07	101.07	40	5.31	9	169	82	
JBT0295	Ac-FSSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 713)	---		2		13	inhib		
					1.80	10			
JBT0296	Ac-FQNKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 407)	---		25		6	79		
		4.91	69.35	19	7.83	10			

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0297	Ac-FQSKNNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 183)	---		67	---	13	147		
		0.63	67.87	59	---	14	206	113	
		0.44	58.5	56	---	7	170		
		1.1	60.6	41	---	14	197		
JBT0298	Ac-FQSKQNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 747)	---		45	---	15	109		
		1.83	71.69	42	---	17			
JBT0299	Ac-FQSKKNVFAFGYFERLRAKL-NH2 (SEQ ID NO: 408)	---		13	---	5	63		
		15.31	73.69	9	---	6			
JBT0300	Ac-FQSKKNVFSFGYFERLRAKL-NH2 (SEQ ID NO: 409)	---		7	---	3	40		
JBT0301	Ac-FQSKKNVFTFGYFERLRAKL-NH2 (SEQ ID NO: 410)	---		9	---	0	49		
JBT0302	Ac-FQSKKNVFVAGYFERLRAKL-NH2 (SEQ ID NO: 258)	---		55	---	3	220		
		2.10	90.94	47	---	6	249	185	
		2.6	68.2	33	---	8	218		
JBT0303	Ac-FQSKKNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 184)	---		61	---	7	253		

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
		1.16	93.12	61	---	8	250	218	
		0.92	78.01	54	---	7	244		
					11.16	7	239		
JBT0303	Ac-FQSKKNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 184)	1.76	77.5	45	8.8	9			
JBT0304	Ac-FQSKKNVFVLGYFERLRAKL-NH2 (SEQ ID NO: 259)	---		35	---	7	138		
		2.95	75.15	32	12.87	7			
JBT0305	Ac-FQSKKNVFVQGYFERLRAKL-NH2 (SEQ ID NO: 260)	---		46	---	0	207		
		2.28	87.92	45	---	5	223	205	
		3.15	65.6	27	---	4	207		
JBT0306	Ac-FQSKKNVFVSGYFERLRAKL-NH2 (SEQ ID NO: 185)	---		42	---	3	197		
		3.56	94.28	37	---	2	204	174	
		3.89	71.4	28	---	5	247		
JBT0307	Ac-FQSKKNVFVYGYFERLRAKL-NH2 (SEQ ID NO: 261)	---		38	---	7	151		
		3.42	85.15	34	8.72	4	182	168	
		3.01	60.2	26	---	9	182		
JBT0308	Ac-FQSKKNVFVFGYKERLRAKL-NH2 (SEQ ID	---		9	---	4	133		

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	SEQUENCE	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
	NO: 411)								
JBT0309	Ac-FQSKKNVFVFGYYERLRAKL-NH2 (SEQ ID NO: 412)	---		21	---	7	123		
		4.50	66.79	19	8.55	7			
JBT0310	Ac-FQSKKNVFVFGYFDRLRAKL-NH2 (SEQ ID NO: 262)	---		31	---	5	142		
		3.86	70.81	26	16.98	3	152		
		2.11	47.68	24	---	4	163		
		3.37	48.8	19	---	5			
JBT0311	Ac-FQSKKNVFVFGYFERLRAKN-NH2 (SEQ ID NO: 748)	---		5	---	5	27		
		44.92	68.81	5	3.72	10			
JBT0335	Ac-FQSKKNVFVFGYFERLRAK-NH2 (SEQ ID NO: 749)	6.2	16.4	2	10.48	5	28		
JBT0336	Ac-FQSKNNVFVAGYFDRLRAKL-OH (SEQ ID NO: 263)	6.52	63	19	12.13	5	186		
							181		
JBT0337	Ac-FQSKNNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 16)	0.71	69.1	51	13.33	15	219		
JBT0338	Ac-FQSKNNVFVQGYFDRLRAKL-NH2 (SEQ ID NO: 17)	1.12	60.9	40	8.42	9	197		

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0339	Ac-FQSKNNVFVSGYFDRLRAKL-NH2 (SEQ ID NO: 18)	1.22	66.9	42	19.1	9	230		
							231		
JBT0340	Ac-FQSKNNVFVYGYFDRLRAKL-NH2 (SEQ ID NO: 186)	1.87	66	37	26.12	9	197		
JBT0341	Ac-FQSKNNVFVFGYFDRLRAKL-NH2 (SEQ ID NO: 187)	1.08	54.1	38	8.68	10	193		
JBT0342	Ac-FQSKNNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 19)	0.38	65.8	58	7.15	18	244		
							241		
JBT0343	Ac-FQSKKNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 188)	1.08	54.1	34	42.24	8	239		
		2.21	82	42					
JBT0374	Ac-FQSKDNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 20)	---		47	---	5	216		
		0.64	56.9	49	20.22	19			
JBT0375	Ac-FQSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 190)	---		57	---	14	224		
		---		70	2.93	30			
		0.31	62.1	61					
JBT0375	Ac-FQSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 190)	---		53	---	19			

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0376	Ac-FQSKKNAFVFGYFERLRAKL-NH2 (SEQ ID NO: 414)	---		11	---	3	101		
JBT0377	Ac-FQSKKNQFVFGYFERLRAKL-NH2 (SEQ ID NO: 264)	---		20	---	8	180		
JBT0378	Ac-FQSKKNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 21)	---		41	---	7	245		
		1.59	79.1	44	9.03	12			
JBT0379	Ac-FQSKKNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 22)	---		31	---	6	226		
		3.13	81.8	36	47.85 ??	12			
JBT0380	Ac-FQSKKNVFVVGYFERLRAKL-NH2 (SEQ ID NO: 191)	---		27	---	6	199		
		2.59	73.1	34	7.58	10			
JBT0381	Ac-FQSPKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 265)	---		30	---	9	173		
		no fit		34	no fit	11			
JBT0385	Ac-FQSKKNNFVFGYFERLRAKL-NH2 (SEQ ID NO: 672)	---		0	---	2	---		
JBT0386	Ac-FQSKKNPFVFGYFERLRAKL-NH2 (SEQ ID NO: 415)	---		6	---	5	85		
JBT0388	Ac-FQSKKNVHVFGYFERLRAKL-NH2 (SEQ ID	---		25	---	6	185		

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	SEQUENCE	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
	NO: 192)								
		4.34	72.5	26	no fit	9			
JBT0389	Ac-FQSKKNVVVFGYFERLRAKL-NH2 (SEQ ID NO: 673)	---		0	---	8	34		
JBT0390	Ac-FQSKKNVFQFGYFERLRAKL-NH2 (SEQ ID NO: 674)	---		0	---	5	28		
JBT0391	Ac-FQSKKNVFVGGYFERLRAKL-NH2 (SEQ ID NO: 266)	---		12	---	1	178		
		7.02	73.8	18	---	1			
JBT0392	Ac-FQSKKNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 23)	---		32	---	4	228		
		2.48	74.4	37	---	6			
JBT0393	Ac-FQSKKNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 193)	---		20	---	3	268		
		3.02	64.1	28	---	1			
JBT0394	Ac-FQSKKNVFVMGYFERLRAKL-NH2 (SEQ ID NO: 24)	---		10	---	7	115		
		6.83	57.3	12	---	8			
JBT0395	Ac-FQSKKNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 25)	---		28	---	7	235		
		2.80	72.1	32	no fit	7			

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	SEQUENCE	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0396	Ac-FQSKKNVFVPGYFERLRAKL-NH2 (SEQ ID NO: 194)	---		22	---	6	196		
		4.33	77.2	30	8.71	6			
JBT0397	Ac-FQSKKNVFVRGYFERLRAKL-NH2 (SEQ ID NO: 195)	---		24	---	4	240		
		5.09	72.3	23	9.80	0			
JBT0398	Ac-FQSKKNVFVFGYFEELRAKL-NH2 (SEQ ID NO: 196)	---		4	---	8	--		
JBT0399	Ac-FQSKKNVFVFGYFELLRAKL-NH2 (SEQ ID NO: 750)	---		0	---	7	--		
JBT0400	Ac-FQSKKNVFVFGYFLRLRAKL-NH2 (SEQ ID NO: 267)	---		1	---	13	--		
JBT0401	Ac-FQSKKNVFVFGYFERLRAVL-NH2 (SEQ ID NO: 416)	---		0	---	4	--		
JBT0402	Biotin-Ttds-FQSKNNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 973)	0.32	45.41	48	---	11	185		
JBT0403	Biotin-Ttds-FQSKKNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 972)	0.49	64.39	59	---	9	253		
		---		62					
JBT0404	Biotin-Ttds-FQSKKNVFVFGYFDRLRAKL-NH2 (SEQ ID NO: 974)	1.89	42.89	24	3.38	14	115		
JBT0405	Biotin-FQSKKNVFVFGYFERLRAKL-NH2	0.86	81.33	59	---	15	142		

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FIGURE 12

JBT-0047		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	SEQUENCE	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
	(SEQ ID NO: 975)								
JBT0406	Ac-Ttds-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 268)	---		28	---	9	114		

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FIGURE 13

JBT-0049		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0049	Ac-SGNTFVDERLLYFLTIGNMGMYAAQLKFRTS-NH2 (SEQ ID NO: 3025)	---		NA	---	NA		12	12
		---		NA	---	17	36	44	
		---		NA	---	11			
		---		NA					
		---		17					
JBT0049	Ac-SGNTFVDERLLYFLTIGNMGMYAAQLKFRTS-NH2 (SEQ ID NO: 3025)	---		10	---	13	49	17	
		---		19	---	8	42		
							53		
JBT0053	Biotin-Ttds-SGNTFVDERLLYFLTIGNMGMYAAQLKFRTS-NH2 (SEQ ID NO: 3006)	---		NA	---	11			
		---		NA					
		---		18					
JBT0057	Ac-SGNTFVDERLLYFLTIGNMGMYAAQLKFRTS-Ttds-Lysin(biotin)-NH2 (SEQ ID NO: 3018)	---		NA	---	15			
		---		NA					
		---		15					
JBT0135	Biotin-Ttds-LLYFLTIGNMGMYAAQLKFR-NH2 (SEQ ID NO: 3021)	---		NA	---	10 (0.625μ			

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FIGURE 13

JBT-0049		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
						M)			
		---		0					
JBT0136	Biotin-Ttds-LYFLTIGNMGMYAAQLKFRT-NH2 (SEQ ID NO: 3014)	---		NA	---	11 (0.625μM)			
		---		0					
JBT0137	Biotin-Ttds-YFLTIGNMGMYAAQLKFRTS-NH2 (SEQ ID NO: 3013)	---		0	---	2			
JBT0138	Biotin-Ttds-YFLTIGNMGMYAAQLKFR-NH2 (SEQ ID NO: 3012)	---		0	---	4			
JBT0190	Ac-KKSGNTFVDERLLYFLTIGNMGMYAAQLKFRTSKK-NH2 (SEQ ID NO: 3031)	---		21	0.49	32	80	72	
		---		27			74		
JBT0191*	Ac-MLGVLMRGISALTGDYTARFEFYLNKQTFN-NH2 (SEQ ID NO: 3054)	---		0	---	0			
JBT0192*									
JBT0193	Ac-KKSGNTFVDERLLYFLTIGNMGMYAAQLKFKK-NH2 (SEQ ID NO: 3073)			51	1.26	38	140	114	
		2.30	104.85	52			149		
JBT0194	Ac-KKSGNTFVDERLLYFLTIGNMGMYAAQKK-NH2 (SEQ ID NO: 3074)	---		0	---	8			

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FIGURE 13

JBT-0049		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0195	Ac-KKSGNTFVDERLLYFLTIGNMGMYKK-NH2 (SEQ ID NO: 3075)	---		0	---	5			
JBT0196	Ac-KKSGNTFVDERLLYFLTIGNMKK-NH2 (SEQ ID NO: 3048)	---		0	---	12			
JBT0197	Ac-KKTFVDERLLYFLTIGNMGMYAAQLKFRTSKK-NH2 (SEQ ID NO: 3076)	---		38	0.38	35	146		
		0.91	57.40	39					
JBT0198	Ac-KKDERLLYFLTIGNMGMYAAQLKFRTSKK-NH2 (SEQ ID NO: 3077)	---		0	---	17			
JBT0199	Ac-KKLLYFLTIGNMGMYAAQLKFRTSKK-NH2 (SEQ ID NO: 3078)	---		0	---	20			
					---	17			
JBT0200	Ac-KKFLTIGNMGMYAAQLKFRTSKK-NH2 (SEQ ID NO: 3079)	---		0	---	9			
JBT0201	Ac-KKLLYFLTIGNMGMYAAQLKFRKK-NH2 (SEQ ID NO: 3080)	---		2	---	23			
					---	16			
JBT0202	Ac-KKLYFLTIGNMGMYAAQLKFRTKK-NH2 (SEQ ID NO: 3081)	---		0	---	15			
JBT0203	Ac-KKYFLTIGNMGMYAAQLKFRTSKK-NH2 (SEQ ID NO: 3082)	---		0	---	7			
JBT0204	Ac-KKSGNTFVDERLLYFLTIGNMGKK-NH2 (SEQ ID NO: 3060)	---		0	---	5			

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FIGURE 13

JBT-0049		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
					---	0			
JBT0243	Ac-SGDTFVDERLLYFLTIGNMRMYAVQLKFRTS-NH2 (SEQ ID NO: 3085)			39	---	37			
		0.51	41.53	37					
JBT0244	Ac-SGDTFVDERLLYFLTTGNMRMYAVQLKFRTS-NH2 (SEQ ID NO: 3086)	---		50	---	37			
		---		49					
JBT0245	Ac-SGDTFVDERLLYFLTIGDMRMYAVQLKFRTS-NH2 (SEQ ID NO: 3087)	---		45	---	NA			
		0.44	50.53	44	---	21			
					---	24			
JBT0344	Ac-TFVDERLLYFLTIGNMGMYAAQLKF-NH2 (SEQ ID NO: 3038)	---		2	---	1	---		
JBT0345	Ac-FVDERLLYFLTIGNMGMYAAQLKF-NH2 (SEQ ID NO: 3039)	---		2	---	2	---		
JBT0346	Ac-VDERLLYFLTIGNMGMYAAQLKF-NH2 (SEQ ID NO: 3034)	---		0	---	2	---		
JBT0347	Ac-TFVDERLLYFLTIGNMGMYAAQLK-NH2 (SEQ ID NO: 3040)	---		2	---	4	---		
JBT0348	Ac-TFVDERLLYFLTIGNMGMYAAQ-NH2 (SEQ ID NO: 3084)	---		0	---	2	---		

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FIGURE 14

JBT-0050		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0050	Ac-SGRGCTKVIVFTFRHNKLIGYERRYNCTS-NH2 (SEQ ID NO: 3047)	---		NA	---	NA		81	21
		---		NA	0.14	25	32	38	
		---		NA	---	NA	46		
		---		NA	---	NA	46		
		0.95	56.6	39	0.05	33	49		
		---		NA					
		1.14	58.99	35					
JBT0050	Ac-SGRGCTKVIVFTFRHNKLIGYERRYNCTS-NH2 (SEQ ID NO: 3047)	---		33	---	28	23		
		1.07	35,,6	26	0.14	19	27	36	
JBT0054	Biotin-Ttds-SGRGCTKVIVFTFRHNKLIGYERRYNCTS-NH2 (SEQ ID NO: 3002)	---		NA	0.16	NA			
		0.99	56.1	41	0.45	31			
JBT0058	Ac-SGRGCTKVIVFTFRHNKLIGYERRYNCTS-Ttds-Lysin(biotin)-NH2 (SEQ ID NO: 3003)	---		NA	---	NA			
		1.07	60.9	42	0.37	28			
JBT0129	Ac-SGRGCTKVIVFTFRHNKLIGYERRYNCTS-NH2; Cycl. C5-C27 (SEQ ID NO: 3026)	---		NA	0.16	21	27		

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FIGURE 14

JBT-0050		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
		---		32			31	18	
JBT0130	Biotin-Ttds-SGRGCTKVIVFTFRHNKLIGYERRYNCTS-NH2; cycl. C6-C28 (SEQ ID NO: 3001)	---		NA	0.15	21			
		---		34					
JBT0139	Biotin-Ttds-GRGCTKVIVFTFRHNKLIGY-NH2 (SEQ ID NO: 3008)	---		NA	---	9	--	--	
		---		7 (0,625μM)	0.10	15			
JBT0140	Biotin-Ttds-GCTKVIVFTFRHNKLIGYER-NH2 (SEQ ID NO: 3010)	---		0	---	10	18	25	
					0.05	NA			
JBT0141	Biotin-Ttds-CTKVIVFTFRHNKLIGYERR-NH2 (SEQ ID NO: 3009)	---		0	---	14	34	55	
					0.04	NA			
JBT0142	Biotin-Ttds-TKVIVFTFRHNKLIGYERRY-NH2 (SEQ ID NO: 3022)	---		0	---	7	12	--	
					1.96	12			
JBT0143	Biotin-Ttds-KVIVFTFRHNKLIGYERRYN-NH2 (SEQ ID NO: 3023)	---		7	---	16	19	--	
		---		6	0.42	17			
JBT0144	Biotin-Ttds-VIVFTFRHNKLIGYERRYNC-NH2 (SEQ ID NO: 3011)	---		NA	---	18	--	--	

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FIGURE 14

JBT-0050		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
		---		0	0.03	---			
JBT0145	Biotin-Ttds-VIVFTFRHNKLIGYER-NH2 (SEQ ID NO: 3024)	---		0	---	5	--	--	
JBT0205	Ac-KKSGRGCTKVIVFTFRHNKLIGYERRYNCTSKK-NH2 (SEQ ID NO: 3029)	---		40	---	34	134		
		1.64	87.46	49	0.11	36			
JBT0206*	Ac-TNYTGSEKCIRFVTRRYLGVRINCFHKGS-NH2 (SEQ ID NO: 3049)	---		0	---	22	--		
					---	17			
					0.22	20			
JBT0207*	Ac-TRNVVRRYECFGSTGCIKYFIHSRTGLNK-NH2 (SEQ ID NO: 3050)	---		9	---	8	--		
					---	6			
JBT0208	Ac-KKSGRGCTKVIVFTFRHNKLIGYERRYNKK-NH2 (SEQ ID NO: 3027)	---		7	---	18			
					---	24			
JBT0209	Ac-KKSGRGCTKVIVFTFRHNKLIGYERKK-NH2 (SEQ ID NO: 3051)	---		0	---	17			
JBT0210	Ac-KKSGRGCTKVIVFTFRHNKLIGKK-NH2 (SEQ ID NO: 3061)	---		0	---	17			
					---	13			
JBT0211	Ac-	---		42	---	31	182		

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FIGURE 14

JBT-0050		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	SEQUENCE	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
	KKGCTKVIVFTFRHNKLIGYERRYNCTSKK-NH2 (SEQ ID NO: 3032)								
		---		48	0.04	35	(250)	(650)	
JBT0212	Ac-KKKVIVFTFRHNKLIGYERRYNCTSKK-NH2 (SEQ ID NO: 3033)	---		7	---	23	49		
		---		19	0.05	24			
JBT0213	Ac-KKVFTFRHNKLIGYERRYNCTSKK-NH2 (SEQ ID NO: 3083)	---		0	---	15			
JBT0214	Ac-KKGRGCTKVIVFTFRHNKLIGYKK-NH2 (SEQ ID NO: 3052)	---		0	---	15			
JBT0215	Ac-KKGCTKVIVFTFRHNKLIGYERKK-NH2 (SEQ ID NO: 3055)	---		1	---	16			
JBT0216	Ac-KKCTKVIVFTFRHNKLIGYERRKK-NH2 (SEQ ID NO: 3053)	---		6	---	18			
JBT0217	Ac-KKTKVIVFTFRHNKLIGYERRYKK-NH2 (SEQ ID NO: 3062)	---		1	1.06	18			
		---		11					
JBT0218	Ac-KKKVIVFTFRHNKLIGYERRYNKK-NH2 (SEQ ID NO: 3063)	---		4	1.18	12			
		---		12					
JBT0219	Ac-KKVIVFTFRHNKLIGYERRYNCKK-NH2 (SEQ ID NO: 3030)	---		8	---	17	29		
		---		22	0.10	17			
JBT0220	Ac-KKIVFTFRHNKLIGYERRYNCTKK-NH2 (SEQ ID NO: 3064)	---		0	---	15			

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FIGURE 14

JBT-0050		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	SEQUENCE	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
					---	16			
JBT0349	Ac-GCTKVIVFTFRHNKLIGYERRYNCTS-NH2 (SEQ ID NO: 3041)	---		19	---	28	54		
		1.63	27.6	17	0.18	13			
JBT0350	Ac-CTKVIVFTFRHNKLIGYERRYNCTS-NH2 (SEQ ID NO: 3042)	---		21	---	26	55		
		1.65	33.8	18	0.27	16			
JBT0351	Ac-TKVIVFTFRHNKLIGYERRYNCTS-NH2 (SEQ ID NO: 3043)	---		1	---	19	30		
		no fit		0	0.21	8			
JBT0352	Ac-GCTKVIVFTFRHNKLIGYERRYNCT-NH2 (SEQ ID NO: 3044)	---		24	---	24	65		
		1.34	26.3	19	0.35	18			
JBT0353	Ac-GCTKVIVFTFRHNKLIGYERRYNC-NH2 (SEQ ID NO: 3045)	---		28	---	30	76		
		0.72	24.5	21	0.61	20			

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FIGURE 15

JBT-0101		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0052	Biotin-Ttds-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTSKK-NH2 (SEQ ID NO: 3004)	---		NA	0.22	20			
		---		NA	0.39	23			
		1.47	57.24	52					
JBT0101	Ac-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTSKK-NH2 (SEQ ID NO: 3036)	---		NA	---	NA	65	164	40
		---		NA	1.42	22			
		2.25	50.13	26	0.24	21			
					---	20			
					0.17	22			
JBT0101	Ac-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTSKK-NH2 (SEQ ID NO: 3036)	2.01	45.27	26	---	19	68	38	
		---		NA			60		
		---		NA					
		---		NA					
		1.51	44.8	31					
JBT0101	Ac-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTSKK-NH2 (SEQ ID NO: 3036)	---		29	---	19			

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FIGURE 15

JBT-0101		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
		---		18	0.25	14			
		1.94	37.47	19	0.61	19			
JBT0103	Ac-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTS-Ttds-KKLysin(biotin)-NH2 (SEQ ID NO: 3005)	---		NA	0.19	21			
		1.69	38.91	24	0.17	17			
JBT0176*	Ac-KKVGSVTRWSMYGPIFIKFTWTLEQPVGWDHKK-NH2 (SEQ ID NO: 3056)	---		0	---	17	150		
					---	16			
JBT0177*	Ac-KKLTGDWTYFWSKVIWGPGVIERQMPVSTFHKK-NH2 (SEQ ID NO: 3065)	---		4	---	15	--		
					---	10			
					---	10			
JBT0178	Ac-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTKK-NH2 (SEQ ID NO: 3028)			42	---	23	190	80	
		1.51	65.93	42	0.20	26	111		
JBT0179	Ac-KKSGVWQTHPRYFWTMWPDIKGEVIVLKK-NH2 (SEQ ID NO: 3066)	---		0	---	14			
JBT0180	Ac-KKSGVWQTHPRYFWTMWPDIKGEVKK-NH2 (SEQ ID NO: 3067)	---		0	---	5			

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FIGURE 15

JBT-0101		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0181	Ac-KKSGVWQTHPRYFWTMWPDIKKK-NH2 (SEQ ID NO: 3068)	---		0	---	2			
JBT0182	Ac-KKGVWQTHPRYFWTMWPDIKGEVIVLFGTSKK-NH2 (SEQ ID NO: 3037)	---		39	0.26	27	146		
		0.62	52.16	40					
JBT0183	Ac-KKQTHPRYFWTMWPDIKGEVIVLFGTSKK-NH2 (SEQ ID NO: 3069)	---		0	---	19			
JBT0184	Ac-KKPRYFWTMWPDIKGEVIVLFGTSKK-NH2 (SEQ ID NO: 3070)	---		0	---	26			
					---	16			
JBT0185	Ac-KKFWTMWPDIKGEVIVLFGTSKK-NH2 (SEQ ID NO: 3057)	---		0	---	14			
JBT0186	Ac-KKSGVWQTHPRYFWTMWPDIKGKK-NH2 (SEQ ID NO: 3071)	---		0	---	3			
JBT0187	Ac-KKWQTHPRYFWTMWPDIKGEVIKK-NH2 (SEQ ID NO: 3072)	---		0	---	5			
JBT0188	Ac-KKHPRYFWTMWPDIKGEVIVLFKK-NH2 (SEQ ID NO: 3058)	---		0	---	21			
					---	17			
JBT0189	Ac-KKYFWTMWPDIKGEVIVLFGTSKK-NH2 (SEQ ID NO: 3059)	---		0	---	19			
JBT0354	Ac-KKGVWQTHPRYFWTMWPDIKGEVIVLFGTK	---		26	0.62	14			

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FIGURE 15

JBT-0101		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
	K-NH2 (SEQ ID NO: 3088)								
		2.25	50.46	25	0.56	18			
JBT0355	Ac-KKVWQTHPRYFWTMWPDIKGEVIVLFGTKK-NH2 (SEQ ID NO: 3089)	---		12	---	14			
JBT0356	Ac-KKWQTHPRYFWTMWPDIKGEVIVLFGTKK-NH2 (SEQ ID NO: 3090)	---		11	no fit	11			
JBT0357	Ac-KKGVWQTHPRYFWTMWPDIKGEVIVLFGKK-NH2 (SEQ ID NO: 3035)	---		30	---	25			
		2.51	55.65	26	0.58	20			
JBT0358	Ac-KKGVWQTHPRYFWTMWPDIKGEVIVLFKK-NH2 (SEQ ID NO: 3046)	---		19	---	27			
		3.26	41.21	15	0.46	16			

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FIGURE 16

JBT-0120		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0120	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1047)	---		NA	0.94	35	298	374	162
		0.90	85.30	58	1.84	31			
		1.50	88.65	48					
JBT0120	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1047)	1.50	74.75	43	0.86	45	355		
							258		
		1.29	69.17	43	0.88	45			
JBT0120	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1047)	1.18	50.68	34	1.18	46	326	288	
		---		37	---	48	383		
		---		38			299		
		1.26	46.06	32	0.38	40			
JBT0120	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2***HCL** (SEQ ID NO: 1047)	2.64	93.62	43	0.98	43			
JBT0124	Biotin-Ttds-SGASRYKWFCGMRDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1290)	1.59	73.59	41	1.26	36			
JBT0247	Ac-	---		43	---	44	273		

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FIGURE 16

JBT-0120		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
	SGASRYKWFCGMRDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1213)								
		0.79	50.9	42	0.61	42			
JBT0248	Ac-KKSGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTSKK-NH2 cyc. (SEQ ID NO: 1001)	---		54	---	60	164		
		0.82	76.13	53	0.27	52			
JBT0249	Ac-KKSGASRYKWFCGMRDMKGTMSKK-NH2 (SEQ ID NO: 1214)	---		0	---	3	inhib		
					---	17			
					1.06	16			
JBT0250	Ac-KKSRYKWFCGMRDMKGTMSCVWKK-NH2 (SEQ ID NO: 1201)	---		0		15	inhib		
		---		11	---	20	inhib		
					0.61	19			
					---	17			
JBT0251	Ac-KKKWFCGMRDMKGTMSCVWVKFKK-NH2 (SEQ ID NO: 1202)	---		17	0.63	23	inhib		
		0.66	24.03	16			inhib		
		---		11	---	18			
JBT0252	Ac-KKCGMRDMKGTMSCVWVKFRYDKK-NH2 (SEQ ID NO: 1215)	---		13	1.02	18	inhib		
		6.27	37.09	11					

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FIGURE 16

JBT-0120		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0253	Ac-KKMRDMKGTMSCVWVKFRYDTSKK-NH2 (SEQ ID NO: 1216)	---		0	---	15	inhib		
					---	23			
					0.55	23			
JBT0319	Ac-SGASRYKWFCGMRDMKGTMSCVWVKFRY-NH2, cyc (SEQ ID NO: 1002)	0.51	71.07	59	---	56	383	293	
							294		
JBT0319	Ac-SGASRYKWFCGMRDMKGTMSCVWVKFRY-NH2, cyc (SEQ ID NO: 1002)	0.30	82.38	72	---	60	466		
JBT0320	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVK-NH2 (SEQ ID NO: 1103)	18.80	73.34	9	6.40	7	153		
JBT0321	Ac-SGASRYKWFCGMRDMKGTMSCV-NH2 (SEQ ID NO: 1217)	---		0	---	1	--		
JBT0322	Ac-SGASRYKWFCGMRDMKGTM-NH2 (SEQ ID NO: 1218)	---		0	---	5	--		
JBT0322	Ac-SGASRYKWFCGMRDMKGTM-NH2 (SEQ ID NO: 1218)	---		0	---	3	--		
JBT0323	Ac-SRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1076)	0.72	51.55	42	0.77	48	355	314	
							285		
JBT0324	Ac-KWFCGMRDMKGTMSCVWVKFRYDTS-	---		0	2.44	18	91		

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FIGURE 16

JBT-0120		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	SEQUENCE	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
	NH2 (SEQ ID NO: 1155)								
JBT0325	Ac-[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1219)	---		0	---	4	--		
JBT0326	Ac-RDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1220)	---		0	---	3	--		
JBT0326	Ac-RDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1220)						--		
JBT0327	Ac-SGASRYKWFCGMRDMKGTMS-NH2 (SEQ ID NO: 1222)	---		0	---	0	--		
JBT0328	Ac-SRYKWFCGMRDMKGTMSCVW-NH2 (SEQ ID NO: 1223)	---		0	---	1	--		
JBT0329	Ac-KWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1156)	---		2	---	8	70	66	
		---		4	---	13	40		
		no fit		7	3.53	6	84		
JBT0330	Ac-CGMRDMKGTMSCVWVKFRYD-NH2 (SEQ ID NO: 1224)	---		0	---	9	--		
JBT0330	Ac-[CGMRDMKGTMSC]VWVKFRYD-NH2 (SEQ ID NO: 1224)	---		0	---	7			
JBT0331	Ac-MRDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1225)	---		0	---	5	--		
JBT0331	Ac-MRDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1225)	---		0	---	5	--		
JBT0332	Ac-SRYKWFCGMRDMKGTMSCVW-NH2 (SEQ ID NO: 1206)	---		1	---	5	--		

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FIGURE 16

JBT-0120		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	SEQUENCE	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0333	Ac-KWFCGMRDMKGTMSCVWVKF-NH2 (SEQ ID NO: 1207)	---		3	---	12	40	--	
							--		
JBT0333	Ac-KWFCGMRDMKGTMSCVWVKF-NH2 (SEQ ID NO: 1207)	---		4	---	11	114		
JBT0334	Ac-CGMRDMKGTMSCVWVKFRYD-NH2 (SEQ ID NO: 1227)	---		0	---	3	--		
JBT0409	Ac-SGASRYKWFSGMRDMKGTMSSVWVKFRYDTS-NH2 (SEQ ID NO: 1204)	---		0	---	5	inhib		
JBT0410	Ac-SGASRYKWFSG-Nle-RD-Nle-KGT-Nle-SSVWVKFRYDTS-NH2 (SEQ ID NO: 1208)	---		0	---	7	inhib		
JBT0411	Ac-SGASRYKWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKFRYDTS-NH2 (SEQ ID NO: 1104)	---		11	---	24	97		
JBT0412	Ac-KWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKF-NH2 (SEQ ID NO: 1209)	---		7	---	12	46		
JBT0413	Ac-KWFSGMRDMKGTMSSVWVKF-NH2 (SEQ ID NO: 1228)	---		0	---	2	inhib		
JBT0414	Ac-KWFSG-Nle-RD-Nle-KGT-Nle-SSVWVKF-NH2 (SEQ ID NO: 1205)	---		4	---	8	inhib		
JBT0415	Ac-SRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1003)	---		59	---	43	359		
		---		66					
		0.38	74.56	61	0.19	52			

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FIGURE 16

JBT-0120		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0416	Ac-SRYKWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKF-NH2 (SEQ ID NO: 1077)	---		52	---	46	256		
JBT0417	Ac-SRYKWFSGMRDMKGTMSSVWVKF-NH2 (SEQ ID NO: 1229)	---		2	---	11	inhib		
		no fit		0	no fit	6			
JBT0418	Ac-SRYKWFSG-Nle-RD-Nle-KGT-Nle-SSVWVKF-NH2 (SEQ ID NO: 1210)	---		4	---	11	inhib		
JBT0435*	Ac-RKRDVSGKM[CWSGFGSKWFRC]ADMMTYYSVT-NH2 (SEQ ID NO: 1211)	---		0	---	5			
JBT0436*	Ac-RWSGKSTYS[CDAMVRRMGMKC]FSFGWTDVYK-NH2 (SEQ ID NO: 1212)	---		0	---	0			

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FIGURE 17

JBT-0121		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	SEQUENCE	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0121	Ac-SGGRKHKHFLRSNGKPSRALCSMHFWRWSTS-NH2 (SEQ ID NO: 3091)	---		NA	---	15	22	37	25
		0.08	14.44	14	0.17	11	inhib		
		---		8	---	19			
JBT0125	Biotin-Ttds-SGGRKHLHFLRSNGKPSRALCSMHFWRWSTS-NH2 (SEQ ID NO: 3092)	0.10	17.22	15	0.45	9			
JBT0419	Ac-SGGRKHKHFLRSNGKPSRALSSMHFWRWSTS-NH2 (SEQ ID NO: 3093)	---		6	---	7			
JBT0420*	Ac-KGSSRSGHNALKTHSWMSPFRRRGSWSLHFK-NH2 (SEQ ID NO: 3094)	---		3	---	10	inhib		
JBT0421*	Ac-KSRSGFWHAWSHRTRPMKSRLHLGKFSNSSG-NH2 (SEQ ID NO: 3095)	---		3	---	5			
JBT0422	Ac-SGGRKHKHFLRSNGKPSRALSSMHFWRW-NH2 (SEQ ID NO: 3096)	---		4	---	11	inhib		
JBT0423	Ac-SGGRKHKHFLRSNGKPSRALSSMHF-NH2 (SEQ ID NO: 3097)	---		2	---	1	inhib		
JBT0424	Ac-SGGRKHKHFLRSNGKPSRALSS-NH2 (SEQ	---		1	---	0	--		

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FIGURE 17

JBT-0121		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
	ID NO: 3098)								
JBT0425	Ac-SGGRKHKHFLRSNGKPSRA-NH2 (SEQ ID NO: 3099)	---		3	---	0			
JBT0426	Ac-RKHKHFLRSNGKPSRALSSMHFWRWSTS-NH2 (SEQ ID NO: 3100)	---		2	---	8	inhib		
JBT0427	Ac-KHFLRSNGKPSRALSSMHFWRWSTS-NH2 (SEQ ID NO: 3101)	---		0	---	0			
JBT0428	Ac-LRSNGKPSRALSSMHFWRWSTS-NH2 (SEQ ID NO: 3102)	---		2	---	1	--		
JBT0429	Ac-NGKPSRALSSMHFWRWSTS-NH2 (SEQ ID NO: 3103)	---		0	---	0			
JBT0430	Ac-SGGRKHKHFLRSNGKPSRAL-NH2 (SEQ ID NO: 3104)	---		0	---	0			
JBT0431	Ac-RKHKHFLRSNGKPSRALSSM-NH2 (SEQ ID NO: 3105)	---		2	---	8			
JBT0432	Ac-KHFLRSNGKPSRALSSMHFW-NH2 (SEQ ID NO: 3106)	---		3	---	1	--		
JBT0433	Ac-LRSNGKPSRALSSMHFWRWS-NH2 (SEQ ID NO: 3107)	---		0	---	1	--		
JBT0434	Ac-SNGKPSRALSSMHFWRWSTS-NH2 (SEQ ID NO: 3108)	---		0	---	0			

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FIGURE 18

JBT-0122		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0122	AC-SGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2002)	---		NA	12.7	5	115	139	60
		15.5	93.8	15	39.5	8	175		
		7.92	78.52	23			183	160	
		---		9	---	9	210		
		---		11					
JBT0122	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2*HCL (SEQ ID NO: 2002)	15.81	64.12	6	6.70	7			
JBT0126	Biotin-Ttds-SGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2498)	---		NA	5.05	5			
		14.1	88.4	17					
JBT0221	Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2003)	---		10	3.37	19	218		
		---		29					
JBT0222*	Ac-MKSRSLGLAYFAKHSSLEVLQTRKVAAPYY-NH2 (SEQ ID NO: 2127)	---		0	---	4	--		

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FIGURE 18

JBT-0122		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
					---	5			
JBT0223*	Ac-KMQLRVYASTAHSRYLLGSSLFPKYAEVKA-NH2 (SEQ ID NO: 2297)	---		0	---	4	--		
					---	1			
					---	3			
JBT0224	Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKK-NH2 (SEQ ID NO: 2298)	---		14	3.49	18	191		
		---		26			203	162	
JBT0225	Ac-KKSGYASFPLAVQLHVSKRSKEMALARKK-NH2 (SEQ ID NO: 2128)	---		9	---	0	55		
		---		12					
JBT0226	Ac-KKSGYASFPLAVQLHVSKRSKEMAKK-NH2 (SEQ ID NO: 2299)	---		4	---	0			
		---		6					
JBT0227	Ac-KKSGYASFPLAVQLHVSKRSKKK-NH2 (SEQ ID NO: 2300)	---		0	---	5			
JBT0228	Ac-KKASFPLAVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2016)	---		4	6.93	9			

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FIGURE 18

JBT-0122		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
		---		21					
JBT0229	Ac-KKPLAVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2129)	---		0	---	0			
JBT0230	Ac-KKVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2301)	---		0	---	0			
					---	0			
JBT0231	Ac-KKHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2302)	---		0	---	0			
JBT0232	Ac-KKGYASFPLAVQLHVSKRSKEMKK-NH2 (SEQ ID NO: 2303)	---		4	---	2			
		13.25	38.12	6					
JBT0233	Ac-KKYASFPLAVQLHVSKRSKEMAKK-NH2 (SEQ ID NO: 2304)	---		6	---	6			
		---		8					
JBT0234	Ac-KKVQLHVSKRSKEMALARLYYKKK-NH2 (SEQ ID NO: 2305)	---		0	---	3			
JBT0235	Ac-KKQLHVSKRSKEMALARLYYKTKK-NH2 (SEQ ID NO: 2306)	---		0	---	0			
					---	0			
JBT0236	Ac-KKLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2307)	---		0	---	0			

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FIGURE 18

JBT-0122		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
JBT0237	Ac-KKSGYASFPLAVQLHVSKRSKEKK-NH2 (SEQ ID NO: 2308)	---		0	---	2			
JBT0359	Ac-ASFPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2309)	---		0	---	6	---		
JBT0360	Ac-SFPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2310)	---		1	---	2	---		
JBT0361	Ac-FPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2311)	---		0	---	9	---		
JBT0362	Ac-ASFPLAVQLHVSKRSKEM-NH2 (SEQ ID NO: 2312)	---		0	---	6	---		
JBT0363	Ac-ASFPLAVQLHVSKRSKE-NH2 (SEQ ID NO: 2313)	---		0	---	9	---		
JBT0364	Ac-ASFPLAVQLHVSKRSKEMAL-NH2 (SEQ ID NO: 2314)	---		2	---	12	---		
JBT0365	Ac-ASFPLAVQLHVSKRSKEMALA-NH2 (SEQ ID NO: 2315)	---		0	---	9	---		
JBT0366	Ac-ASFPLAVQLHVSKRSKEMALAR-NH2 (SEQ ID NO: 2316)	---		0	---	11	---		
JBT0367	Ac-ASFPLAVQLHVSKRSKEMALARL-NH2 (SEQ ID NO: 2317)	---		2	---	10	28		
JBT0368	Ac-ASFPLAVQLHVSKRSKEMALARLY-NH2 (SEQ ID NO: 2130)	---		0	---	8	36		
JBT0369	Ac-ASFPLAVQLHVSKRSKEMALARLYY-NH2	---		2	---	9	68		

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FIGURE 18

JBT-0122		Progress Curves of Inhibition of FXa			Extrinsic Tenase Inhibition by TFPI		Factor IIa Generation		
PEPTIDE	**SEQUENCE**	EC_{50} (μM)	% Inhibit (Max)	% Inhibit (2.5μM)	EC_{50} (μM)	% Inhibit (2.5μM)	Induced FVIII Deficiency FEIBA EA (mU/mL) Conc: 10μM	FVIII Deficient (mU/mL) Conc: 10μM	FIX Deficient (mU/mL) Conc: 10μM
	(SEQ ID NO: 2017)								
JBT0370	Ac-YASFPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2318)	---		2	---	0	39		
JBT0371	Ac-GYASFPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2319)	---		8	---	1	60		

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FIGURE 19

JBT0047 class		BiaCore		
Peptide	**Sequence**	**k_{on} (1/Ms)**	**k_{off} (1/s)**	**K_D (M)**
JBT0047	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-NH2 (SEQ ID NO: 253)	3.98E+05	1.88E-02	4.72E-08
JBT0131	Biotin-Ttds-AFQSKKNVFVFGYFERLRAK-NH2 (SEQ ID NO: 964)	1.38E+05	5.94E-02	4.31E-07
JBT0132	Biotin-Ttds-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 965)	3.55E+04	3.26E-02	9.17E-07
JBT0155	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 8)	1.64E+05	7.25E-03	4.41E-08
JBT0158	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKKK-NH2 (SEQ ID NO: 9)	4.49E+05	3.27E-02	7.29E-08
JBT0164	Ac-KKFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 12)	2.26E+05	1.05E-02	4.65E-08
JBT0170	Ac-KKFQSKKNVFVFGYFERLRAKLKK-NH2 (SEQ ID NO: 13)	4.09E+05	9.34E-03	2.28E-08
JBT0175	Ac-KKAFQSKKNVFVFGYFERLRAKLTKK-NH2 (SEQ ID NO: 182)	2.70E+05	1.67E-02	6.18E-08
JBT0293	Ac-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 256)	2.17E+05	2.12E-02	9.74E-08
JBT0294	Ac-YQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 257)	5.69E+05	3.92E-02	6.90E-08
JBT0297	Ac-FQSKNNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 183)	5.69E+05	3.92E-02	6.90E-08
JBT0302	Ac-FQSKKNVFVAGYFERLRAKL-NH2 (SEQ ID NO: 258)	1.02E+06	8.47E-02	8.27E-08
JBT0303	Ac-FQSKKNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 184)	8.13E+05	2.75E-02	3.38E-08
JBT0305	Ac-FQSKKNVFVQGYFERLRAKL-NH2 (SEQ ID NO: 260)	7.45E+05	3.07E-02	4.11E-08
JBT0306	Ac-FQSKKNVFVSGYFERLRAKL-NH2 (SEQ ID NO: 185)	6.56E+05	3.61E-02	5.50E-08
JBT0307	Ac-FQSKKNVFVYGYFERLRAKL-NH2 (SEQ ID NO: 261)	5.95E+05	3.15E-02	5.30E-08
JBT0310	Ac-FQSKKNVFVFGYFDRLRAKL-NH2 (SEQ ID NO: 262)	5.62E+04	1.05E-02	1.87E-07
JBT0337	Ac-FQSKNNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 16)	2.90E+05	5.30E-03	1.83E-08
JBT0342	Ac-FQSKNNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 19)	3.92E+05	6.55E-03	1.67E-08
JBT0343	Ac-FQSKKNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 188)	3.28E+05	1.46E-02	4.46E-08
JBT0375	Ac-FQSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 190)	4.00E+05	1.21E-02	3.04E-08
JBT0471	Ac-FQSKGNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 26)	7.03E+05	9.34E-03	1.33E-08
JBT0474	Ac-FQSKGNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 29)	9.95E+05	1.49E-02	1.50E-08
JBT0477	Ac-FQSKGNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 32)	3.72E+05	3.02E-03	8.10E-09
JBT0500	Ac-FQSK-NmetGly-NVFVFGYFERLRAKL-NH2 (SEQ ID NO: 46)	2.52E+05	2.06E-03	8.19E-09
JBT0683	Ac-FQSK-Nmg-AVFVDGYFARLRAKL-NH2 (SEQ ID NO: 55)	1.00E+06	1.07E-03	1.07E-09

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FIGURE 19

JBT0047 class		BiaCore		
Peptide	**Sequence**	**k_{on} (1/Ms)**	**k_{off} (1/s)**	**K_D (M)**
JBT0765	Ac-FQSKGNVFVDGYFERL-Eew-AKL-NH2 (SEQ ID NO: 68)	2.31E+05	1.85E-03	8.00E-09
JBT0789	Ac-FQSK-Nmg-NVFVDGYFERL-Nle-AKL-NH2 (SEQ ID NO: 71)	1.92E+05	4.06E-04	2.12E-09
JBT1122	Ac-FQSK-Nmg-NVFVDGYFARL-Nle-AKL-NH2 (SEQ ID NO: 113)	6.99E+04	9.61E-04	1.38E-08
JBT1141	Ac-FQSKkAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 120)	1.36E+05	3.16E-03	2.33E-08
JBT1144	Ac-FQSKdAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 121)	3.05E+05	1.00E-02	3.28E-08

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FIGURE 20

JBT0120 class		BiaCore		
Peptide	**Sequence**	**k_{on} (1/Ms)**	**k_{off} (1/s)**	**K_D (M)**
JBT0120	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1047)	1.17E+06	4.78E-02	4.08E-08
JBT0320	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVK-NH2 (SEQ ID NO: 1103)	1.78E+05	4.25E-02	2.38E-07
JBT0324	Ac-KWFCGMRDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1155)	1.54E+04	6.36E-03	4.14E-07
JBT0415	Ac-SRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1003)	1.75E+05	2.24E-02	1.28E-07
JBT0444	Ac-SRYKWF[CAMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1004)	4.18E+05	1.21E-02	2.90E-08
JBT0447	Ac-SRYKWF[CGMRAMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1158)	6.86E+03	5.60E-03	8.16E-07
JBT0455	Ac-SRYKWF[CGMRDMKGTMSC]AWVKF-NH2 (SEQ ID NO: 1049)	4.77E+05	3.03E-02	6.35E-08
JBT0643	Ac-SRYKWF[CGMPDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1006)	1.78E+05	3.22E-02	1.81E-07

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FIGURE 21

JBT0122 class		BiaCore		
Peptide	**Sequence**	**k_{on} (1/Ms)**	**k_{off} (1/s)**	**K_D (M)**
JBT0122	AC-SGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2002)	1.13E+04	2.23E-02	1.97E-06
JBT0221	Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2003)	6.31E+04	1.19E-02	1.88E-07
JBT0224	Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKK-NH2 (SEQ ID NO: 2298)	6.39E+04	1.95E-02	3.05E-07

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FIGURE 22

JBT0047 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0047	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-NH2 (SEQ ID NO: 253)	0.72		68	
JBT0051	Biotin-Ttds-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-NH2 (SEQ ID NO: 962)	0.53		66	
JBT0055	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-Ttds-Lysin(biotin)-NH2 (SEQ ID NO: 963)	0.53		72	
JBT0131	Biotin-Ttds-AFQSKKNVFVFGYFERLRAK-NH2 (SEQ ID NO: 964)	8.20		23	
JBT0132	Biotin-Ttds-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 965)	1.15		36	
JBT0133	Biotin-Ttds-QSKKNVFVFGYFERLRAKLT-NH2 (SEQ ID NO: 966)			15	
JBT0134	Biotin-Ttds-QSKKNVFVFGYFERLRAK-NH2 (SEQ ID NO: 967)			5	
JBT0155	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 8)	0.97		54	
JBT0156*	Ac-VVEKLTFVQLSFLNRRRFSQYAGFKGAGKV-NH2 (SEQ ID NO: 742)			0	
JBT0157*	Ac-RVFLYFSGKAGGLVKLVERQAFQTNVSKFR-NH2 (SEQ ID NO: 743)			0	
JBT0158	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKKK-NH2 (SEQ ID NO: 9)	4.26		35	
JBT0159	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLKK-NH2 (SEQ ID NO: 744)			0	
JBT0160	Ac-KKSGVGRLQVAFQSKKNVFVFGYFKK-NH2 (SEQ ID NO: 745)			0	
JBT0161	Ac-KKSGVGRLQVAFQSKKNVFVFKK-NH2 (SEQ ID NO: 746)			0	
JBT0162	Ac-KKGRLQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 10)	0.74		54	
JBT0163	Ac-KKQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 11)	1.54		48	
JBT0164	Ac-KKFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 12)	1.74		43	
JBT0165	Ac-KKKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 712)			0	
JBT0166	Biotinyl-Ttds-KKFQSKKNVFVFGYFERLRAKLKK-NH2 (SEQ ID NO: 968)	2.60		50	
JBT0167*	Biotinyl-Ttds-GKNAKFYLFESLRQVKFVFR-NH2 (SEQ ID NO: 969)			0	
JBT0168*	Biotinyl-Ttds-YKFSFNKELFKQARLRFVGV-NH2 (SEQ ID NO: 970)			0	

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FIGURE 22

JBT0047 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0169	Ac-KKAFQSKKNVFVFGYFERLRAKKK-NH2 (SEQ ID NO: 254)			16	
JBT0170	Ac-KKFQSKKNVFVFGYFERLRAKLKK-NH2 (SEQ ID NO: 13)			61	
JBT0171	Ac-KKQSKKNVFVFGYFERLRAKLTKK -NH2 (SEQ ID NO: 255)			25	
JBT0172	Ac-KKQSKKNVFVFGYFERLRAKKK-NH2 (SEQ ID NO: 406)			7	
JBT0173	Biotinyl-Ttds-SGVGRLQVAFQSKKNVFVFG-NH2 (SEQ ID NO: 971)			0	
JBT0174	Ac-KKAFQSKKNVFVFGYFERLRAKLKK-NH2 (SEQ ID NO: 14)			47	
JBT0175	Ac-KKAFQSKKNVFVFGYFERLRAKLTKK-NH2 (SEQ ID NO: 182)	2.71		40	
JBT0293	Ac-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 256)	3.11	61	26	8
JBT0294	Ac-YQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 257)	4.07		40	
JBT0295	Ac-FSSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 713)			2	
JBT0296	Ac-FQNKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 407)	4.91		19	
JBT0297	Ac-FQSKNNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 183)	0.63		59	
JBT0298	Ac-FQSKQNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 747)	1.83		42	
JBT0299	Ac-FQSKKNVFAFGYFERLRAKL-NH2 (SEQ ID NO: 408)	15.31		9	
JBT0300	Ac-FQSKKNVFSFGYFERLRAKL-NH2 (SEQ ID NO: 409)			7	
JBT0301	Ac-FQSKKNVFTFGYFERLRAKL-NH2 (SEQ ID NO: 410)			9	
JBT0302	Ac-FQSKKNVFVAGYFERLRAKL-NH2 (SEQ ID NO: 258)	2.10		47	
JBT0303	Ac-FQSKKNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 184)	0.88	69	50	26
JBT0304	Ac-FQSKKNVFVLGYFERLRAKL-NH2 (SEQ ID NO: 259)	2.95		32	
JBT0305	Ac-FQSKKNVFVQGYFERLRAKL-NH2 (SEQ ID NO: 260)	2.28		45	
JBT0306	Ac-FQSKKNVFVSGYFERLRAKL-NH2 (SEQ ID NO: 185)	3.56		37	
JBT0307	Ac-FQSKKNVFVYGYFERLRAKL-NH2 (SEQ ID NO: 261)	3.42		34	
JBT0308	Ac-FQSKKNVFVFGYKERLRAKL-NH2 (SEQ ID NO: 411)			9	
JBT0309	Ac-FQSKKNVFVFGYYERLRAKL-NH2 (SEQ ID NO: 412)	4.50		19	

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FIGURE 22

JBT0047 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0310	Ac-FQSKKNVFVFGYFDRLRAKL-NH2 (SEQ ID NO: 262)	3.86		26	
JBT0311	Ac-FQSKKNVFVFGYFERLRAKN-NH2 (SEQ ID NO: 748)			5	
JBT0335	Ac-FQSKKNVFVFGYFERLRAK-NH2 (SEQ ID NO: 749)	6.20		2	
JBT0336	Ac-FQSKNNVFVAGYFDRLRAKL-OH (SEQ ID NO: 263)	6.52		19	
JBT0337	Ac-FQSKNNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 16)	0.71		51	
JBT0338	Ac-FQSKNNVFVQGYFDRLRAKL-NH2 (SEQ ID NO: 17)	1.12		40	
JBT0339	Ac-FQSKNNVFVSGYFDRLRAKL-NH2 (SEQ ID NO: 18)	1.22		42	
JBT0340	Ac-FQSKNNVFVYGYFDRLRAKL-NH2 (SEQ ID NO: 186)	1.87		37	
JBT0341	Ac-FQSKNNVFVFGYFDRLRAKL-NH2 (SEQ ID NO: 187)	1.08		38	
JBT0342	Ac-FQSKNNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 19)	0.38		58	
JBT0343	Ac-FQSKKNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 188)	1.08		34	
JBT0374	Ac-FQSKDNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 20)	0.75	66	52	27
JBT0375	Ac-FQSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 190)	0.41	73	65	40
JBT0376	Ac-FQSKKNAFVFGYFERLRAKL-NH2 (SEQ ID NO: 414)			11	
JBT0377	Ac-FQSKKNQFVFGYFERLRAKL-NH2 (SEQ ID NO: 264)			20	
JBT0378	Ac-FQSKKNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 21)	1.59	79	44	
JBT0379	Ac-FQSKKNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 22)	3.13	81	36	
JBT0380	Ac-FQSKKNVFVVGYFERLRAKL-NH2 (SEQ ID NO: 191)	2.59	73	34	
JBT0381	Ac-FQSPKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 265)	3.28	81	34	
JBT0385	Ac-FQSKKNNFVFGYFERLRAKL-NH2 (SEQ ID NO: 672)			0	
JBT0386	Ac-FQSKKNPFVFGYFERLRAKL-NH2 (SEQ ID NO: 415)			6	
JBT0388	Ac-FQSKKNVHVFGYFERLRAKL-NH2 (SEQ ID NO: 192)	4.34	73	26	
JBT0389	Ac-FQSKKNVVVFGYFERLRAKL-NH2 (SEQ ID NO: 673)			0	
JBT0390	Ac-FQSKKNVFQFGYFERLRAKL-NH2 (SEQ ID NO: 674)			0	

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FIGURE 22

JBT0047 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0391	Ac-FQSKKNVFVGGYFERLRAKL-NH2 (SEQ ID NO: 266)	7.02	74	18	
JBT0392	Ac-FQSKKNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 23)	2.48	74	37	
JBT0393	Ac-FQSKKNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 193)	3.02	64	28	
JBT0394	Ac-FQSKKNVFVMGYFERLRAKL-NH2 (SEQ ID NO: 24)	6.83	57	12	
JBT0395	Ac-FQSKKNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 25)	2.80	72	32	
JBT0396	Ac-FQSKKNVFVPGYFERLRAKL-NH2 (SEQ ID NO: 194)	4.33	77	30	
JBT0397	Ac-FQSKKNVFVRGYFERLRAKL-NH2 (SEQ ID NO: 195)	5.09	72	23	
JBT0398	Ac-FQSKKNVFVFGYFEELRAKL-NH2 (SEQ ID NO: 196)			4	
JBT0399	Ac-FQSKKNVFVFGYFELLRAKL-NH2 (SEQ ID NO: 750)			0	
JBT0400	Ac-FQSKKNVFVFGYFLRLRAKL-NH2 (SEQ ID NO: 267)			1	
JBT0401	Ac-FQSKKNVFVFGYFERLRAVL-NH2 (SEQ ID NO: 416)			0	
JBT0402	Biotin-Ttds-FQSKNNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 973)	0.32	45	48	
JBT0403	Biotin-Ttds-FQSKKNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 972)	0.49	64	59	
JBT0404	Biotin-Ttds-FQSKKNVFVFGYFDRLRAKL-NH2 (SEQ ID NO: 974)	1.89	43	24	
JBT0405	Biotin-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 975)	0.86	81	59	
JBT0406	Ac-Ttds-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 268)			28	
JBT0471	Ac-FQSKGNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 26)	0.41	77	64	45
JBT0472	Ac-FQSKGNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 27)	0.37	60	49	34
JBT0473	Ac-FQSKGNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 28)	0.25	47	45	31
JBT0474	Ac-FQSKGNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 29)	0.29	75	68	50
JBT0475	Ac-FQSKGNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 30)	0.25	60	50	37
JBT0476	Ac-FQSKGNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 31)	0.22	54	49	38
JBT0477	Ac-FQSKGNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 32)	0.23	72	68	51
JBT0478	Ac-FQSKDNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 33)	0.76	51	39	21

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FIGURE 22

JBT0047 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0479	Ac-FQSKDNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 34)	1.10	58	40	21
JBT0480	Ac-FQSKDNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 35)	1.11	65	42	24
JBT0481	Ac-FQSKDNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 36)	0.77	53	41	23
JBT0482	Ac-FQSKDNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 37)	0.53	86	69	43
JBT0483	Ac-FQSKDNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 38)	0.53	84	68	41
JBT0484	Ac-FQSKDNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 976)	0.79	73	56	29
JBT0485	Ac-FQSKNNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 39)	0.75	86	63	38
JBT0486	Ac-FQSKNNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 40)			44	26
JBT0487	Ac-FQSKNNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 41)			58	36
JBT0488	Ac-FQSKNNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 42)	0.56	78	64	40
JBT0489	Ac-FQSKNNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 43)	0.42	78	68	48
JBT0490	Ac-FQSKQNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 44)			53	25
JBT0491	Ac-FQSKQNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 197)			53	29
JBT0492	Ac-FQSKQNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 198)			45	24
JBT0493	Ac-FQSKQNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 199)			45	21
JBT0494	Ac-FQSKQNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 45)			60	36
JBT0495	Ac-FQSKQNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 200)			59	36
JBT0497	Ac-NmetPhe-QSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 201)	0.47		40	24
JBT0498	Ac-FQ-NmetSer-KGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 675			3	1
JBT0499	Ac-FQS-NmetLys-GNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 202)	0.83	75	56	30
JBT0500	Ac-FQSK-NmetGly-NVFVFGYFERLRAKL-NH2 (SEQ ID NO: 46)	0.13	71	70	60
JBT0501	Ac-FQSKGN-NmetV-FVFGYFERLRAKL-NH2 (SEQ ID NO: 417)			16	6
JBT0502	Ac-FQSKGNV-NmetPhe-VFGYFERLRAKL-NH2 (SEQ ID NO: 751)			0	0
JBT0503	Ac-FQSKGNVF-NmetV-FGYFERLRAKL-NH2 (SEQ ID NO: 676)			0	0

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FIGURE 22

JBT0047 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0504	Ac-FQSKGNVFV-NmetPhe-GYFERLRAKL-NH2 (SEQ ID NO: 418)			7	2
JBT0505	Ac-FQSKGNVFVF-NmetGly-YFERLRAKL-NH2 (SEQ ID NO: 269)			21	7
JBT0506	Ac-FQSKGNVFVFG-NmetTyr-FERLRAKL-NH2 (SEQ ID NO: 714)			0	0
JBT0507	Ac-FQSKGNVFVFGY-NmetPhe-ERLRAKL-NH2 (SEQ ID NO: 677)			0	0
JBT0508	Ac-FQSKGNVFVFGYF-NmetGlu-RLRAKL-NH2 (SEQ ID NO: 678)			0	0
JBT0509	Ac-FQSKGNVFVFGYFE-Nmr-LRAKL-NH2 (SEQ ID NO: 752)			0	0
JBT0510	Ac-FQSKGNVFVFGYFER-NmetLeu-RAKL-NH2 (SEQ ID NO: 753)			0	0
JBT0511	Ac-FQSKGNVFVFGYFERL-Nmr-AKL-NH2 (SEQ ID NO: 754)			0	0
JBT0512	Ac-FQSKGNVFVFGYFERLR-NmetAla-KL-NH2 (SEQ ID NO: 679)			0	0
JBT0513	Ac-FQSKGNVFVFGYFERLRA-NmetLys-L-NH2 (SEQ ID NO: 755)			0	0
JBT0514	Ac-FQSKGNVFVFGYFERLRAK-NmetLeu-NH2 (SEQ ID NO: 419)			1	0
JBT0515	Ac-bHomoPhe-QSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 420)			22	11
JBT0516	Ac-F-bHomoGln-SKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 680)			7	4
JBT0517	Ac-FQ-bHomoSer-KGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 421)			7	2
JBT0518	Ac-FQS-bHomoLys-GNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 422)			9	2
JBT0519	Ac-FQSK-bAla-NVFVFGYFERLRAKL-NH2 (SEQ ID NO: 270)			20	15
JBT0520	Ac-FQSKG-bGln-VFVFGYFERLRAKL-NH2 (SEQ ID NO: 271)			24	9
JBT0521	Ac-FQSKGN-bLeu-FVFGYFERLRAKL-NH2 (SEQ ID NO: 756)			3	3
JBT0522	Ac-FQSKGNV-bHomoPhe-VFGYFERLRAKL-NH2 (SEQ ID NO: 681)			5	3
JBT0523	Ac-FQSKGNVF-bLeu-FGYFERLRAKL-NH2 (SEQ ID NO: 757)			1	2
JBT0524	Ac-FQSKGNVFV-bHomoPhe-GYFERLRAKL-NH2 (SEQ ID NO: 682)			0	4
JBT0525	Ac-FQSKGNVFVF-bAla-YFERLRAKL-NH2 (SEQ ID NO: 758)			0	3
JBT0526	Ac-FQSKGNVFVFG-bHomoTyr-FERLRAKL-NH2 (SEQ ID NO: 759)			0	0
JBT0527	Ac-FQSKGNVFVFGY-bHomoPhe-ERLRAKL-NH2 (SEQ ID NO: 683)			9	2

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FIGURE 22

JBT0047 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0528	Ac-FQSKGNVFVFGYF-bE-RLRAKL-NH2 (SEQ ID NO: 272)			25	20
JBT0529	Ac-FQSKGNVFVFGYFE-bHomoArg-LRAKL-NH2 (SEQ ID NO: 760)			0	1
JBT0530	Ac-FQSKGNVFVFGYFER-Btl-RAKL-NH2 (SEQ ID NO: 684)			3	0
JBT0532	Ac-FQSKGNVFVFGYFERLR-bAla-KL-NH2 (SEQ ID NO: 423)			14	7
JBT0533	Ac-FQSKGNVFVFGYFERLRA-bHomoK-L-NH2 (SEQ ID NO: 424)			17	4
JBT0534	Ac-FQSKGNVFVFGYFERLRAK-Btl-NH2 (SEQ ID NO: 204)	0.47	48	39	28
JBT0535	Ac-FQSKGNVFVFGYFE-Cit-LRAKL-NH2 (SEQ ID NO: 685)			13	5
JBT0536	Ac-FQSKGNVFVFGYFERL-Cit-AKL-NH2 (SEQ ID NO: 273)			52	32
JBT0537	Ac-FQSKGNVFVFGYFE-Nle-LRAKL-NH2 (SEQ ID NO: 761)			0	0
JBT0538	Ac-FQSKGNVFVFGYFERL-Nle-AKL-NH2 (SEQ ID NO: 425)			16	20
JBT0564	Ac-FQSKKNVFVFGYFKRLRAKL-NH2 (SEQ ID NO: 205)	4.12	54	19	6
JBT0578	Ac-FQSKKNVFVFGYFFRLRAKL-NH2 (SEQ ID NO: 441)			1	0
JBT0613	Ac-FQSKGNVFVFGYFERLRAKL-OH (SEQ ID NO: 283)			21	8
JBT0614	NH2-FQSKGNVFVFGYFERLRAKL-OH (SEQ ID NO: 720)			3	0
JBT0615	NH2-FQSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 284)			14	5
JBT0616	NH2-GSFQSKKNVFVDGYFERLRAKL-OH (SEQ ID NO: 285)	0.85	30	25	
JBT0636	Ac-FQSK-Nmg-NVFVDGYFARLRAKL-NH2 (SEQ ID NO: 47)	0.10	90	86	76
JBT0651	Ac-FQSKGNVHVKGYFERLRAKL-NH2 (SEQ ID NO: 48)	0.59	50	40	26
JBT0652	Ac-FQSKGNVHVDGYFERLRAKL-NH2 (SEQ ID NO: 49)	0.34	52	48	34
JBT0653	Ac-FQSPGNVHVKGYFERLRAKL-NH2 (SEQ ID NO: 50)	0.44	57	47	33
JBT0654	Ac-FQSPGNVHVDGYFERLRAKL-NH2 (SEQ ID NO: 51)	0.37	60	50	36
JBT0655	Ac-FQSKGNIFVFGYFERLRAKL-NH2 (SEQ ID NO: 52)	1.32	65	41	22
JBT0656	Ac-FQSKGNLFVFGYFERLRAKL-NH2 (SEQ ID NO: 286)			21	10
JBT0657	Ac-FQSKGNVFIFGYFERLRAKL-NH2 (SEQ ID NO: 287)			24	10

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FIGURE 22

JBT0047 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0658	Ac-FQSKGNVFLFGYFERLRAKL-NH2 (SEQ ID NO: 694)			6	3
JBT0663	Ac-FQSKaNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 53)	0.29	86	81	60
JBT0668	Ac-FQSKaNVFVTGYFARLRAKL-NH2 (SEQ ID NO: 54)	0.52	84	70	43
JBT0681	Ac-FQSK-Nmg-AVFVFGYFARLRAKL-NH2 (SEQ ID NO: 206)			52	52
JBT0683	Ac-FQSK-Nmg-AVFVDGYFARLRAKL-NH2 (SEQ ID NO: 55)	0.13	91	86	74
JBT0696	Ac-FQSKKAVFVFGYFERLRAKL-NH2 (SEQ ID NO: 288)	0.71	56	48	26
JBT0697	Ac-FQSKGNVFVDGYFERL-Dap-AKL-NH2 (SEQ ID NO: 56)	0.50	89	73	52
JBT0699	Ac-FQSKGNVFVDGYFERL-Orn-AKL-NH2 (SEQ ID NO: 57)	0.41	89	76	57
JBT0700	Ac-FQSKGNVFVDGYFERL-Nva-AKL-NH2 (SEQ ID NO: 58)	0.19	83	77	64
JBT0704	Ac-FQSKKNVFVFGYFARLRAKL-NH2 (SEQ ID NO: 289)			19	8
JBT0708	Ac-FQSKaNVFVAGYFERLRAKL-NH2 (SEQ ID NO: 59)	0.25	89	77	59
JBT0714	Ac-FQSKaAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 60)	0.18	81	75	59
JBT0717	Ac-FQSK-Nmg-NVFVDGYFERLRAKL-NH2 (SEQ ID NO: 61)	0.06	91	87	84
JBT0720	Ac-FQSK-Nmg-NVFVTGYFARLRAKL-NH2 (SEQ ID NO: 62)	0.22	85	77	62
JBT0732	Ac-FQSKGNVFVDGYFERL-Hci-AKL-NH2 (SEQ ID NO: 63)	0.18	83	75	62
JBT0733	Ac-FQSKGNVFVDGYFERL-Har-AKL-NH2 (SEQ ID NO: 64)	0.18	81	75	62
JBT0739	Ac-FQSKGNVFVDGYFERL-Opa-AKL-NH2 (SEQ ID NO: 65)	0.28	82	74	51
JBT0740	Ac-FQSKGNVFVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 66)	0.09	87	84	78
JBT0754	Ac-FQSK-Nmg-AVFVTGYFARLRAKL-NH2 (SEQ ID NO: 67)	0.24	80	74	57
JBT0757	Ac-FQsKKNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 699)			3	1
JBT0759	Ac-FQSKkNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 207)	0.65	60	53	29
JBT0765	Ac-FQSKGNVFVDGYFERL-Eew-AKL-NH2 (SEQ ID NO: 68)	0.16	95	87	73
JBT0775	Ac-FQSKGNVFVTGYFDRLRAKL-NH2 (SEQ ID NO: 69)	0.77	77	57	34
JBT0778	Ac-FQSKGNVFVKGYFDRLRAKL-NH2 (SEQ ID NO: 209)	1.48	77	46	23

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FIGURE 22

JBT0047 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0779	Ac-FQSKGNVFVEGYFDRLRAKL-NH2 (SEQ ID NO: 210)	0.77	82	59	37
JBT0780	Ac-FQSK-Nmg-NVFVFGYFARLRAKL-NH2 (SEQ ID NO: 70)			45	54
JBT0781	Ac-FQSKGNVFVDGYFKRLRAKL-NH2 (SEQ ID NO: 211)	1.25	81	48	25
JBT0789	Ac-FQSK-Nmg-NVFVDGYFERL-Nle-AKL-NH2 (SEQ ID NO: 71)	0.06	86	84	77
JBT0806	Ac-FQSK-Nmg-NVFVAGYFARLRAKL-NH2 (SEQ ID NO: 74)	0.13	79	75	65
JBT0837	Ac-FQSK-Nmg-NVFVTGYFERL-Nle-AKL-NH2 (SEQ ID NO: 213)	0.20	76	70	54
JBT0844	Ac-FQSKaNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 300)	0.32	84	72	51
JBT0850	Ac-FQSKaAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 302)	0.19	75	70	55
JBT0854	Ac-FQSKaAVFVAGYFARL-Nle-AKL-NH2 (SEQ ID NO: 490)	0.29	49	47	36
JBT0870	Ac-FQSKaAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 214)	0.31	84	80	57
JBT0886	Ac-FQSKaNVFVDGYFARLRAKL-NH2 (SEQ ID NO: 215)	0.36	83	71	48
JBT0894	Ac-FQSKGNVFVDGYFERLHAKL-NH2 (SEQ ID NO: 76)	0.39	95	80	56
JBT0919	Ac-FQSKaNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 79)	0.36	86	74	51
JBT0931	Ac-FQSKGNVFVDGYFERL-Eag-AKL-NH2 (SEQ ID NO: 879)	0.42	91	75	52
JBT0946	Ac-FQSKGNVFVDGYFERL-Dab-AKL-NH2 (SEQ ID NO: 85)	0.27	85	74	56
JBT0950	Ac-FQSKaNVFVFGYFARLRAKL-NH2 (SEQ ID NO: 217)			47	31
JBT0973	Ac-FQSKGNVFVDGYFERL-Cha-AKL-NH2 (SEQ ID NO: 92)	0.16	70	65	58
JBT1006	Ac-FQSKGNVFVDGYFERL-Hle-AKL-NH2 (SEQ ID NO: 96)	0.17	77	71	61
JBT1035	Ac-FQSK-Nmg-AVFVAGYFARL-Nle-AKL-NH2 (SEQ ID NO: 218)				53
JBT1037	Ac-FQSKGNVFVDGYFERL-Nle-AKL-NH2 (SEQ ID NO: 104)	0.22	74	68	55
JBT1043	Ac-FQSKaNVFVDGYFERL-Nle-AKL-NH2 (SEQ ID NO: 106)	0.30	80	70	51
JBT1082	Ac-FQSKaAVFVFGYFARLRAKL-NH2 (SEQ ID NO: 565)	0.20	47	45	38
JBT1084	Ac-FQSK-Nmg-AVFVAGYFARLRAKL-NH2 (SEQ ID NO: 109)	0.21	66	62	51
JBT1106	Ac-FQSKGNVFVDGYFWRLRAKL-NH2 (SEQ ID NO: 220)	0.28	94	79	65

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FIGURE 22

JBT0047 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT1122	Ac-FQSK-Nmg-NVFVDGYFARL-Nle-AKL-NH2 (SEQ ID NO: 113)			67	65
JBT1131	Biotin-Ttds-FQSKGNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 977)	0.20	94	86	69
JBT1133	Ac-FQSKkAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 115)	0.22	80	72	57
JBT1134	Ac-FQSKGAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 116)	0.24	72	66	54
JBT1135	Ac-FQSKDAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 221)	0.39	68	61	38
JBT1136	Ac-FQSKdAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 117)	0.17	77	72	58
JBT1137	Ac-FQSKkAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 118)	0.32	71	58	40
JBT1138	Ac-FQSKGAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 222)	0.25	50	47	38
JBT1139	Ac-FQSKDAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 223)			48	26
JBT1140	Ac-FQSKdAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 119)	0.21	62	58	47
JBT1141	Ac-FQSKkAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 120)	0.37	62	54	38
JBT1142	Ac-FQSKGAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 224)	0.39	49	47	32
JBT1143	Ac-FQSKDAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 225)	0.84	54	40	19
JBT1144	Ac-FQSKdAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 121)	0.32	63	57	38
JBT1145	Ac-FQSKkAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 122)	0.39	66	60	38
JBT1146	Ac-FQSKGAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 226)			41	28
JBT1147	Ac-FQSKDAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 227)			48	24
JBT1148	Ac-FQSKdAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 123)	0.33	68	61	44
JBT1149	Ac-FQSKGNVFvFGYFERLRAKL-NH2 (SEQ ID NO: 936)			0	0
JBT1151	Ac-FQSKKNVFVFGYFERLRAKD-NH2 (SEQ ID NO: 937)			0	0
JBT1152	Ac-FQSKKNVFFFGYFERLRAKL-NH2 (SEQ ID NO: 735)			0	0
JBT1153	Ac-FQSKKNVFVFGYFERLGAKL-NH2 (SEQ ID NO: 705)	5.25	51	12	0
JBT1155	Ac-FQSK-Nmg-NVFVTGYFERLRAKL-NH2 (SEQ ID NO: 125)	0.21	87	77	61
JBT1156	Ac-FQSKaNVFVDGYFARL-Nle-AKL-NH2 (SEQ ID NO: 126)			67	44

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FIGURE 22

JBT0047 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT1157	Ac-FQSKaNVFVAGYFARLRAKL-NH2 (SEQ ID NO: 127)	0.27	77	69	51
JBT1158	Ac-FQSKGNVFVFGYFERLRAKL-N-methyl (SEQ ID NO: 228)			73	45
JBT1159	Ac-FQSKGNVFVFGYFERLRAKL-N-ethyl (SEQ ID NO: 128)			71	41
JBT1160	Ac-FQSKGNVFVFGYFERLRAKL-N-propyl (SEQ ID NO: 978)			76	54
JBT1161	Ac-FQSK-Aib-NVFVFGYFERLRAKL-NH2 (SEQ ID NO: 338)			71	46
JBT1162	Ac-FQSKpNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 129)	0.03	94	92	90
JBT1396	Ac-FQSK-Nmg-NVFVAGYFERLRAKL-NH2 (SEQ ID NO: 161)	0.05	95	93	87
JBT1584	Ac-FQSK-Nmg-NVFVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 164)	0.02	93	93	92
JBT1585	Ac-FQSK-Nmg-NVFVTGYFERL-Aib-AKL-NH2 (SEQ ID NO: 165)	0.04	90	90	89
JBT1587	Ac-FQSKGNVFVDGYFERL-Aib-AKLC-NH2 (SEQ ID NO: 167)	0.05	90	88	83
JBT1590	Ac-FQSKGNVFV[CGYFERL-Aib-AKLC]-NH2 (SEQ ID NO: 401)			53	21
JBT1591	Ac-FQSKGNVFVDGYF[CRL-Aib-AKLC]-NH2 (SEQ ID NO: 168)	0.67	81	72	41
JBT1592	Ac-[CFQSKGNVFVDGYFERLC]AKL-NH2 (SEQ ID NO: 402)	1.55	86	54	24
JBT1593	Ac-FQSK[CNVFVDGYFERLC]AKL-NH2 (SEQ ID NO: 670)			43	18
JBT1594	Ac-FQSKGNVFV[CGYFERLC]AKL-NH2 (SEQ ID NO: 671)			9	5
JBT1595	Ac-[CFQSKGNVFVC]GYFERL-Aib-AKL-NH2 (SEQ ID NO: 403)			32	14
JBT1596	Ac-[CFQSKGC]VFVEGYFERL-Aib-AKL-NH2 (SEQ ID NO: 404)			52	24
JBT1597	Ac-DGYFERLRAKL-NH2 (SEQ ID NO: 960)			0	0
JBT1598	Ac-FQSKKNV-NH2 (SEQ ID NO: 961)			0	0
JBT1843	Ac-FQSKGNIFVDGYFERLHAKL-NH2 (SEQ ID NO: 169)	0.81	94	70	42
JBT1844	Ac-FQSKNNVFVDGYFKRLRAKL-NH2 (SEQ ID NO: 170)	0.73	91	70	42
JBT1845	Ac-FQSYKHVFVDGYFERLRAKL-NH2 (SEQ ID NO: 249)	1.24	94	61	33
JBT1846	Ac-FQSKGIVFVDGYFKRLRAKL-NH2 (SEQ ID NO: 250)	0.87	91	65	38
JBT1847	Ac-YQTKGNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 171)	0.12	87	82	73

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FIGURE 22

JBT0047 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT1852	PEG(1kD)-FQSKGNVFVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 174)	0.11	93	88	76
JBT1853	PEG(40kD)-FQSKGNVFVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 175)	0.17	99	91	75
JBT1854	Ac-FQSKGNVFVDGYFERL-Aib-AKLC(PEG1kD)-NH2 (SEQ ID NO: 176)	0.07	93	90	83
JBT1855	Ac-FQSKGNVFVDGYFERL-Aib-AKLC(PEG40kD)-NH2 (SEQ ID NO: 252)	0.10	97	91	79

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FIGURE 23

JBT0120 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0120	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1047)	1.21	75	47	26
JBT0124	Biotin-Ttds-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1290)	1.59		41	
JBT0247	Ac-SGASRYKWFCGMRDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1213)	0.79		42	
JBT0248	Ac-KKSGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTSKK-NH2 (SEQ ID NO: 1001)	0.82		53	
JBT0249	Ac-KKSGASRYKWFCGMRDMKGTMSKK-NH2 (SEQ ID NO: 1214)			0	
JBT0250	Ac-KKSRYKWF[CGMRDMKGTMSC]VWKK-NH2 (SEQ ID NO: 1201)			11	
JBT0251	Ac-KKKWF[CGMRDMKGTMSC]VWVKFKK-NH2 (SEQ ID NO: 1202)			17	
JBT0252	Ac-KK[CGMRDMKGTMSC]VWVKFRYDKK-NH2 (SEQ ID NO: 1215)			13	
JBT0253	Ac-KKMRDMKGTMSCVWVKFRYDTSKK-NH2 (SEQ ID NO: 1216)			0	
JBT0319	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRY-NH2 (SEQ ID NO: 1002)	0.30		72	
JBT0320	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVK-NH2 (SEQ ID NO: 1103)			9	
JBT0321	Ac-SGASRYKWF[CGMRDMKGTMSC]V-NH2 (SEQ ID NO: 1217)			0	
JBT0322	Ac-SGASRYKWFCGMRDMKGTM-NH2 (SEQ ID NO: 1218)			0	
JBT0322	Ac-SGASRYKWFCGMRDMKGTM-NH2 (SEQ ID NO: 1218)			0	
JBT0323	Ac-SRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1076)	0.72		42	
JBT0324	Ac-KWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1155)			0	
JBT0325	Ac-[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1219)			0	
JBT0326	Ac-RDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1220)			0	
JBT0327	Ac-SGASRYKWFCGMRDMKGTMS-NH2 (SEQ ID NO: 1222)			0	
JBT0328	Ac-SRYKWF[CGMRDMKGTMSC]VW-NH2 (SEQ ID NO: 1223)			0	
JBT0329	Ac-KWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1156)			2	

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FIGURE 23

JBT0120 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0330	Ac-[CGMRDMKGTMSC]VWVKFRYD-NH2 (SEQ ID NO: 1224)			0	
JBT0331	Ac-MRDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1225)			0	
JBT0332	Ac-SRYKWFCGMRDMKGTMSCVW-NH2 (SEQ ID NO: 1206)			1	
JBT0333	Ac-KWFCGMRDMKGTMSCVWVKF-NH2 (SEQ ID NO: 1207)			4	
JBT0334	Ac-CGMRDMKGTMSCVWVKFRYD-NH2 (SEQ ID NO: 1227)			0	
JBT0409	Ac-SGASRYKWFSGMRDMKGTMSSVWVKFRYDTS-NH2 (SEQ ID NO: 1204)			0	
JBT0410	Ac-SGASRYKWFSG-Nle-RD-Nle-KGT-Nle-SSVWVKFRYDTS-NH2 (SEQ ID NO: 1208)			0	
JBT0411	Ac-SGASRYKWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKFRYDTS-NH2 (SEQ ID NO: 1104)			11	
JBT0412	Ac-KWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKF-NH2 (SEQ ID NO: 1209)			7	
JBT0413	Ac-KWFSGMRDMKGTMSSVWVKF-NH2 (SEQ ID NO: 1228)			0	
JBT0414	Ac-KWFSG-Nle-RD-Nle-KGT-Nle-SSVWVKF-NH2 (SEQ ID NO: 1205)			4	
JBT0415	Ac-SRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1003)	0.52	73	59	39
JBT0416	Ac-SRYKWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKF-NH2 (SEQ ID NO: 1077)			52	
JBT0417	Ac-SRYKWFSGMRDMKGTMSSVWVKF-NH2 (SEQ ID NO: 1229)			0	
JBT0418	Ac-SRYKWFSG-Nle-RD-Nle-KGT-Nle-SSVWVKF-NH2 (SEQ ID NO: 1210)			4	
JBT0419	Ac-SGGRKHKHFLRSNGKPSRALSSMHFWRWSTS-NH2 (SEQ ID NO: 1293)			6	
JBT0435	Ac-RKRDVSGKM[CWSGFGSKWFRC]ADMMTYYSVT-NH2 (SEQ ID NO: 1211)			0	
JBT0436	Ac-RWSGKSTYS[CDAMVRRMGMKC]FSFGWTDVYK-NH2 (SEQ ID NO: 1212)			0	
JBT0437	Ac-ARYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1048)			71	
JBT0438	Ac-SAYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1078)			54	
JBT0439	Ac-SRAKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1105)			35	
JBT0440	Ac-SRYAWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1106)			37	
JBT0441	Ac-SRYKAF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1230)			0	

FIGURE 23

JBT0120 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0442	Ac-SRYKWA[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1231)			8	
JBT0443	Ac-SRYKWFAGMRDMKGTMSCVWVKF-NH2 (SEQ ID NO: 1157)			0	
JBT0444	Ac-SRYKWF[CAMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1004)	0.24	90	80	62
JBT0445	Ac-SRYKWF[CGARDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1107)			18	
JBT0446	Ac-SRYKWF[CGMADMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1079)			50	
JBT0447	Ac-SRYKWF[CGMRAMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1158)			11	
JBT0448	Ac-SRYKWF[CGMRDAKGTMSC]VWVKF-NH2 (SEQ ID NO: 1108)			44	
JBT0449	Ac-SRYKWF[CGMRDMAGTMSC]VWVKF-NH2 (SEQ ID NO: 1080)	1.26	82	46	30
JBT0450	Ac-SRYKWF[CGMRDMKATMSC]VWVKF-NH2 (SEQ ID NO: 1159)			33	
JBT0451	Ac-SRYKWF[CGMRDMKGAMSC]VWVKF-NH2 (SEQ ID NO: 1081)			54	
JBT0452	Ac-SRYKWF[CGMRDMKGTASC]VWVKF-NH2 (SEQ ID NO: 1109)			39	
JBT0453	Ac-SRYKWF[CGMRDMKGTMAC]VWVKF-NH2 (SEQ ID NO: 1110)	1.18	62	38	23
JBT0454	Ac-SRYKWFCGMRDMKGTMSAVWVKF-NH2 (SEQ ID NO: 1160)			0	
JBT0455	Ac-SRYKWF[CGMRDMKGTMSC]AWVKF-NH2 (SEQ ID NO: 1049)	1.02	65	40	26
JBT0456	Ac-SRYKWFCGMRDMKGTMSCVAVKF-NH2 (SEQ ID NO: 1232)			3	
JBT0457	Ac-SRYKWF[CGMRDMKGTMSC]VWAKF-NH2 (SEQ ID NO: 1161)			16	
JBT0458	Ac-SRYKWF[CGMRDMKGTMSC]VWVAF-NH2 (SEQ ID NO: 1082)	1.34	66	39	24
JBT0459	Ac-SRYKWF[CGMRDMKGTMSC]VWVKA-NH2 (SEQ ID NO: 1083)			47	
JBT0460	Ac-RYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1050)			57	
JBT0461	Ac-YKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1111)			32	
JBT0462	Ac-SRYKWFGM[CRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1233)			0	
JBT0463	Ac-SRYKWFGMRD[CMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1234)			0	
JBT0464	Ac-SRYKWFGMRDMK[CGTMSC]VWVKF-NH2 (SEQ ID NO: 1235)			0	
JBT0465	Ac-SRYKWF[CGMRDMKGTC]MSVWVKF-NH2 (SEQ ID NO: 1236)			0	
JBT0466	Ac-SRYKWF[CGMRDMKC]GTMSVWVKF-NH2 (SEQ ID NO: 1237)			0	

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FIGURE 23

JBT0120 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0467	Ac-SRYKWF[CGMRDC]MKGTMSVWVKF-NH2 (SEQ ID NO: 1238)			0	
JBT0468	Ac-SRYKWFG[CMRDMKGTMC]SVWVKF-NH2 (SEQ ID NO: 1239)			2	
JBT0469	Ac-SRYKWFGM[CRDMKGTC]MSVWVKF-NH2 (SEQ ID NO: 1240)			5	
JBT0470	Ac-SRYKWFGMR[CDMKGC]TMSVWVKF-NH2 (SEQ ID NO: 1162)			0	
JBT0617	Ac-SRYKWF[homoC-GMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1112)	1.91	35	18	8
JBT0618	Ac-SRYKWF[CGMRDMKGTMS-homoC]VWVKF-NH2 (SEQ ID NO: 1163)	3.44	18	5	1
JBT0619	Ac-SRYKWF[homoC-GMRDMKGTMS-homoC]VWVKF-NH2 (SEQ ID NO: 1241)			0	0
JBT0620	Ac-SRYKWF[Dap-GMRDMKGTMS-D]VWVKF-NH2 (SEQ ID NO: 1242)			0	0
JBT0623	Ac-SRYKWF[KGMRDMKGTMSD]VWVKF-NH2 (SEQ ID NO: 1243)			3	2
JBT0625	Ac-SRYKWF[CMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1244)			0	0
JBT0626	Ac-SRYKWF[CGMRDKGTMSC]VWVKF-NH2 (SEQ ID NO: 1164)			3	0
JBT0627	Ac-SRYKWF[CGMRDMGTMSC]VWVKF-NH2 (SEQ ID NO: 1165)			2	0
JBT0628	Ac-SRYKWF[CGMRDMKGMSC]VWVKF-NH2 (SEQ ID NO: 1166)			0	0
JBT0629	Ac-SRYKWF[cGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1167)			0	0
JBT0631	Ac-SRYKWF[CGmRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1113)	1.89	62	33	15
JBT0632	Ac-SRYKWF[CGMrDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1245)			0	0
JBT0633	Ac-SRYKWF[CGMRdMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1168)			0	0
JBT0634	Ac-SRYKWF[CGMRDmKGTMSC]VWVKF-NH2 (SEQ ID NO: 1051)	0.69	63	47	26
JBT0635	Ac-SRYKWF[CGMRDMkGTMSC]VWVKF-NH2 (SEQ ID NO: 1114)	5.14	48	13	2
JBT0637	Ac-SRYKWF[CGMRDMKGtMSC]VWVKF-NH2 (SEQ ID NO: 1169)			0	0
JBT0638	Ac-SRYKWF[CGMRDMKGTmSC]VWVKF-NH2 (SEQ ID NO: 1246)			0	0
JBT0639	Ac-SRYKWF[CGMRDMKGTMsC]VWVKF-NH2 (SEQ ID NO: 1170)			2	0
JBT0640	Ac-SRYKWF[CGMRDMKGTMSc]VWVKF-NH2 (SEQ ID NO: 1171)			3	0
JBT0641	Ac-SRYKWF[CPMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1247)			4	2
JBT0642	Ac-SRYKWF[CGPRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1248)			2	1

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FIGURE 23

JBT0120 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0643	Ac-SRYKWF[CGMPDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1006)			45	26
JBT0644	Ac-SRYKWF[CGMRPMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1172)	4.55	35	12	3
JBT0645	Ac-SRYKWF[CGMRDPKGTMSC]VWVKF-NH2 (SEQ ID NO: 1084)	3.79	62	27	12
JBT0646	Ac-SRYKWF[CGMRDMPGTMSC]VWVKF-NH2 (SEQ ID NO: 1173)			4	1
JBT0647	Ac-SRYKWF[CGMRDMKPTMSC]VWVKF-NH2 (SEQ ID NO: 1249)			4	2
JBT0648	Ac-SRYKWF[CGMRDMKGPMSC]VWVKF-NH2 (SEQ ID NO: 1174)			4	0
JBT0649	Ac-SRYKWF[CGMRDMKGTPSC]VWVKF-NH2 (SEQ ID NO: 1175)			5	2
JBT0650	Ac-SRYKWF[CGMRDMKGTMPC]VWVKF-NH2 (SEQ ID NO: 1250)			0	0
JBT0659	Biotinyl-Ttds-SRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1291)			63	38
JBT0787	Ac-SRYKWF[CG-SeMet-RD-SeMet-KGT-SeMet-SC]VWVKF-NH2 (SEQ ID NO: 1292)			68	56
JBT1416	Ac-DRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1007)	0.58	93	68	46
JBT1417	Ac-SDYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1116)			34	13
JBT1418	Ac-SRDKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1176)			15	5
JBT1419	Ac-SRYDWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO:1117)			31	10
JBT1420	Ac-SRYKDF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1251)			0	0
JBT1421	Ac-SRYKWD[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1252)			2	2
JBT1422	Ac-SRYKWF[CDMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1253)			0	0
JBT1423	Ac-SRYKWF[CGDRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1177)			1	0
JBT1424	Ac-SRYKWF[CGMDDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1254)			4	1
JBT1425	Ac-SRYKWF[CGMRDDKGTMSC]VWVKF-NH2 (SEQ ID NO: 1085)			53	28
JBT1426	Ac-SRYKWF[CGMRDMDGTMSC]VWVKF-NH2 (SEQ ID NO: 1008)	0.19	96	81	66
JBT1427	Ac-SRYKWF[CGMRDMKDTMSC]VWVKF-NH2 (SEQ ID NO: 1052)			57	31
JBT1428	Ac-SRYKWF[CGMRDMKGDMSC]VWVKF-NH2 (SEQ ID NO: 1118)			28	6
JBT1429	Ac-SRYKWF[CGMRDMKGTDSC]VWVKF-NH2 (SEQ ID NO: 1086)			49	21

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FIGURE 23

JBT0120 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT1430	Ac-SRYKWF[CGMRDMKGTMDC]VWVKF-NH2 (SEQ ID NO: 1119)			37	15
JBT1431	Ac-SRYKWF[CGMRDMKGTMSC]DWVKF-NH2 (SEQ ID NO: 1255)			0	0
JBT1432	Ac-SRYKWF[CGMRDMKGTMSC]VDVKF-NH2 (SEQ ID NO: 1256)			0	0
JBT1433	Ac-SRYKWF[CGMRDMKGTMSC]VWDKF-NH2 (SEQ ID NO: 1257)			0	0
JBT1434	Ac-SRYKWF[CGMRDMKGTMSC]VWVDF-NH2 (SEQ ID NO: 1120)			28	9
JBT1435	Ac-SRYKWF[CGMRDMKGTMSC]VWVKD-NH2 (SEQ ID NO: 1009)			61	34
JBT1436	Ac-FRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1053)	0.68	95	71	45
JBT1437	Ac-SFYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1010)	0.27	95	80	64
JBT1438	Ac-SRFKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1011)	0.69	88	64	43
JBT1439	Ac-SRYFWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1121)			35	14
JBT1440	Ac-SRYKFF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1258)			3	2
JBT1441	Ac-SRYKWF[CFMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1259)			7	4
JBT1442	Ac-SRYKWF[CGFRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1778)			10	3
JBT1443	Ac-SRYKWF[CGMFDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1779)			22	7
JBT1444	Ac-SRYKWF[CGMRFMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1260)			7	5
JBT1445	Ac-SRYKWF[CGMRDFKGTMSC]VWVKF-NH2 (SEQ ID NO: 1012)	0.77	89	64	41
JBT1446	Ac-SRYKWF[CGMRDMFGTMSC]VWVKF-NH2 (SEQ ID NO: 1054)	0.68	93	69	45
JBT1447	Ac-SRYKWF[CGMRDMKFTMSC]VWVKF-NH2 (SEQ ID NO: 1227)			46	21
JBT1448	Ac-SRYKWF[CGMRDMKGFMSC]VWVKF-NH2 (SEQ ID NO: 1123)	0.99	86	60	34
JBT1449	Ac-SRYKWF[CGMRDMKGTFSC]VWVKF-NH2 (SEQ ID NO: 1013)	0.40	89	74	53
JBT1450	Ac-SRYKWF[CGMRDMKGTMFC]VWVKF-NH2 (SEQ ID NO: 1124)			44	17
JBT1451	Ac-SRYKWF[CGMRDMKGTMSC]FWVKF-NH2 (SEQ ID NO: 1087)			34	11
JBT1452	Ac-SRYKWF[CGMRDMKGTMSC]VFVKF-NH2 (SEQ ID NO: 1125)			27	5
JBT1453	Ac-SRYKWF[CGMRDMKGTMSC]VWFKF-NH2 (SEQ ID NO: 1261)			6	1
JBT1454	Ac-SRYKWF[CGMRDMKGTMSC]VWVFF-NH2 (SEQ ID NO: 1262)			47	22

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FIGURE 23

JBT0120 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT1455	Ac-GRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1014)	0.79	87	62	40
JBT1456	Ac-SGYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1126)			41	15
JBT1457	Ac-SRGKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1180)			15	4
JBT1458	Ac-SRYGWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1127)			32	10
JBT1459	Ac-SRYKGF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1263)			0	0
JBT1460	Ac-SRYKWG[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1264)			0	0
JBT1461	Ac-SRYKWF[CGGRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1181)			8	0
JBT1462	Ac-SRYKWF[CGMGDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1128)			33	15
JBT1463	Ac-SRYKWF[CGMRGMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1129)			23	10
JBT1464	Ac-SRYKWF[CGMRDGKGTMSC]VWVKF-NH2 (SEQ ID NO: 1088)			50	25
JBT1465	Ac-SRYKWF[CGMRDMGGTMSC]VWVKF-NH2 (SEQ ID NO: 1015)	0.84	92	60	41
JBT1466	Ac-SRYKWF[CGMRDMKGGMSC]VWVKF-NH2 (SEQ ID NO: 1055)			53	26
JBT1467	Ac-SRYKWF[CGMRDMKGTGSC]VWVKF-NH2 (SEQ ID NO: 1130)			27	11
JBT1468	Ac-SRYKWF[CGMRDMKGTMGC]VWVKF-NH2 (SEQ ID NO: 1131)			25	9
JBT1469	Ac-SRYKWF[CGMRDMKGTMSC]GWVKF-NH2 (SEQ ID NO: 1182)			4	0
JBT1470	Ac-SRYKWF[CGMRDMKGTMSC]VGVKF-NH2 (SEQ ID NO: 1265)			0	0
JBT1471	Ac-SRYKWF[CGMRDMKGTMSC]VWGKF-NH2 (SEQ ID NO: 1266)			4	0
JBT1472	Ac-SRYKWF[CGMRDMKGTMSC]VWVGF-NH2 (SEQ ID NO: 1089)			48	21
JBT1473	Ac-SRYKWF[CGMRDMKGTMSC]VWVKG-NH2 (SEQ ID NO: 1090)			40	16
JBT1474	Ac-KRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1016)	0.37	90	75	55
JBT1475	Ac-SKYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1056)			63	37
JBT1476	Ac-SRKKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1132)			31	11
JBT1477	Ac-SRYKKF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1267)			0	0
JBT1478	Ac-SRYKWK[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1091)			45	18
JBT1479	Ac-SRYKWF[CKMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1183)			14	2

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FIGURE 23

JBT0120 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT1480	Ac-SRYKWF[CGKRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1133)			27	9
JBT1481	Ac-SRYKWF[CGMKDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1017)	0.37	88	71	54
JBT1482	Ac-SRYKWF[CGMRKMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1184)			9	1
JBT1483	Ac-SRYKWF[CGMRDKKGTMSC]VWVKF-NH2 (SEQ ID NO: 1057)	0.99	83	60	32
JBT1484	Ac-SRYKWF[CGMRDMKKTMSC]VWVKF-NH2 (SEQ ID NO: 1134)			30	10
JBT1485	Ac-SRYKWF[CGMRDMKGKMSC]VWVKF-NH2 (SEQ ID NO: 1058)			56	32
JBT1486	Ac-SRYKWF[CGMRDMKGTKSC]VWVKF-NH2 (SEQ ID NO: 1059)			58	34
JBT1487	Ac-SRYKWF[CGMRDMKGTMKC]VWVKF-NH2 (SEQ ID NO: 1135)			30	9
JBT1488	Ac-SRYKWF[CGMRDMKGTMSC]KWVKF-NH2 (SEQ ID NO: 1268)			3	0
JBT1489	Ac-SRYKWF[CGMRDMKGTMSC]VKVKF-NH2 (SEQ ID NO: 1269)			0	0
JBT1490	Ac-SRYKWF[CGMRDMKGTMSC]VWKKF-NH2 (SEQ ID NO: 1270)			0	1
JBT1491	Ac-SRYKWF[CGMRDMKGTMSC]VWVKK-NH2 (SEQ ID NO: 1136)			26	6
JBT1492	Ac-LRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1018)	0.47	91	72	52
JBT1493	Ac-SLYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1060)	0.48	96	74	54
JBT1494	Ac-SRLKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1137)			22	6
JBT1495	Ac-SRYLWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1138)			30	10
JBT1496	Ac-SRYKLF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1271)			0	0
JBT1497	Ac-SRYKWL[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1061)			58	33
JBT1498	Ac-SRYKWF[CLMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1272)			3	1
JBT1499	Ac-SRYKWF[CGLRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1139)			21	7
JBT1500	Ac-SRYKWF[CGMLDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1062)			60	34
JBT1501	Ac-SRYKWF[CGMRLMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1185)			14	3
JBT1502	Ac-SRYKWF[CGMRDLKGTMSC]VWVKF-NH2 (SEQ ID NO: 1019)	0.33	93	76	59
JBT1503	Ac-SRYKWF[CGMRDMLGTMSC]VWVKF-NH2 (SEQ ID NO: 1063)	0.53	83	66	44
JBT1504	Ac-SRYKWF[CGMRDMKLTMSC]VWVKF-NH2 (SEQ ID NO: 1140)			37	13

FIGURE 23

JBT0120 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT1505	Ac-SRYKWF[CGMRDMKGLMSC]VWVKF-NH2 (SEQ ID NO: 1092)			66	42
JBT1506	Ac-SRYKWF[CGMRDMKGTLSC]VWVKF-NH2 (SEQ ID NO: 1064)	0.89	91	63	39
JBT1507	Ac-SRYKWF[CGMRDMKGTMLC]VWVKF-NH2 (SEQ ID NO: 10141)			35	11
JBT1508	Ac-SRYKWF[CGMRDMKGTMSC]LWVKF-NH2 (SEQ ID NO: 1093)			52	27
JBT1509	Ac-SRYKWF[CGMRDMKGTMSC]VLVKF-NH2 (SEQ ID NO: 1273)			3	0
JBT1510	Ac-SRYKWF[CGMRDMKGTMSC]VWLKF-NH2 (SEQ ID NO: 1094)			51	27
JBT1511	Ac-SRYKWF[CGMRDMKGTMSC]VWVLF-NH2 (SEQ ID NO: 1142)			52	27
JBT1512	Ac-SRYKWF[CGMRDMKGTMSC]VWVKL-NH2 (SEQ ID NO: 1020)	0.37	91	72	55
JBT1513	Ac-SSYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1065)			63	40
JBT1514	Ac-SRSKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1066)			51	27
JBT1515	Ac-SRYSWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1143)			32	11
JBT1516	Ac-SRYKSF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1274)			0	0
JBT1517	Ac-SRYKWS[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1275)			1	0
JBT1518	Ac-SRYKWF[CSMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1021)			62	39
JBT1519	Ac-SRYKWF[CGSRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1186)			11	3
JBT1520	Ac-SRYKWF[CGMSDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1095)			45	20
JBT1521	Ac-SRYKWF[CGMRSMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1096)			44	21
JBT1522	Ac-SRYKWF[CGMRDSKGTMSC]VWVKF-NH2 (SEQ ID NO: 1067)			56	32
JBT1523	Ac-SRYKWF[CGMRDMSGTMSC]VWVKF-NH2 (SEQ ID NO: 1022)	0.50	91	70	51
JBT1524	Ac-SRYKWF[CGMRDMKSTMSC]VWVKF-NH2 (SEQ ID NO: 1144)			39	16
JBT1525	Ac-SRYKWF[CGMRDMKGSMSC]VWVKF-NH2 (SEQ ID NO: 1068)			68	46
JBT1526	Ac-SRYKWF[CGMRDMKGTSSC]VWVKF-NH2 (SEQ ID NO: 1069)			53	32
JBT1527	Ac-SRYKWF[CGMRDMKGTMSC]SWVKF-NH2 (SEQ ID NO: 1097)			43	20
JBT1528	Ac-SRYKWF[CGMRDMKGTMSC]VSVKF-NH2 (SEQ ID NO: 1276)			2	0
JBT1529	Ac-SRYKWF[CGMRDMKGTMSC]VWSKF-NH2 (SEQ ID NO: 1277)			1	0

FIGURE 23

JBT0120 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT1530	Ac-SRYKWF[CGMRDMKGTMSC]VWVSF-NH2 (SEQ ID NO: 1070)			54	31
JBT1531	Ac-SRYKWF[CGMRDMKGTMSC]VWVKS-NH2 (SEQ ID NO: 1071)			50	28
JBT1532	Ac-PRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1072)			62	38
JBT1533	Ac-SPYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1145)			31	10
JBT1534	Ac-SRPKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1187)			2	0
JBT1535	Ac-SRYPWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1278)			0	1
JBT1536	Ac-SRYKPF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1279)			0	0
JBT1537	Ac-SRYKWP[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1280)			0	0
JBT1538	Ac-SRYKWF[CGMRDMKGTMSC]PWVKF-NH2 (SEQ ID NO: 1281)			2	2
JBT1539	Ac-SRYKWF[CGMRDMKGTMSC]VPVKF-NH2 (SEQ ID NO: 1282)			0	0
JBT1540	Ac-SRYKWF[CGMRDMKGTMSC]VWPKF-NH2 (SEQ ID NO: 1283)			0	0
JBT1541	Ac-SRYKWF[CGMRDMKGTMSC]VWVPF-NH2 (SEQ ID NO: 1023)	0.47	89	69	49
JBT1542	Ac-SRYKWF[CGMRDMKGTMSC]VWVKP-NH2 (SEQ ID NO: 1188)			15	3
JBT1543	Ac-SRYKWF[CGMRDMKGTMSC]VWVKF-OH (SEQ ID NO: 1098)			44	20
JBT1544	H-SRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1024)	0.64	88	67	44
JBT1545	H-SRYKWF[CGMRDMKGTMSC]VWVKF-OH (SEQ ID NO: 1099)			40	20
JBT1546	Ac-SRYKWF[CEMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1284)			3	2
JBT1547	Ac-SRYKWF[CHMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1285)			2	0
JBT1548	Ac-SRYKWF[CIMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1286)			6	3
JBT1549	Ac-SRYKWF[CMMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1189)			9	3
JBT1550	Ac-SRYKWF[CNMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1190)			20	10
JBT1551	Ac-SRYKWF[CQMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1146)			31	13
JBT1552	Ac-SRYKWF[CRMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1147)			32	12
JBT1553	Ac-SRYKWF[CTMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1148)			36	15
JBT1554	Ac-SRYKWF[CVMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1287)			6	2

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FIGURE 23

JBT0120 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT1555	Ac-SRYKWF[CWMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1191)			8	2
JBT1556	Ac-SRYKWF[CYMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1288)			5	0
JBT1557	H-RYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 173)			53	27
JBT1558	H-YKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1149)			17	3
JBT1559	Ac-SRYKWF[CGaRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1150)			29	10
JBT1560	Ac-SRYKWF[CGdRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1289)			1	1
JBT1561	Ac-SRYKWF[CGfRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1192)			9	1
JBT1562	Ac-SRYKWF[CGkRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1193)			5	1
JBT1563	Ac-SRYKWF[CGlRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1194)			14	6
JBT1564	Ac-SRYKWF[CGpRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1100)			65	40
JBT1565	Ac-SRYKWF[CGsRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1195)			13	3
JBT1566	Ac-SRYKWF[CGMRDaKGTMSC]VWVKF-NH2 (SEQ ID NO: 1151)			33	13
JBT1567	Ac-SRYKWF[CGMRDdKGTMSC]VWVKF-NH2 (SEQ ID NO: 1101)			45	21
JBT1568	Ac-SRYKWF[CGMRDfKGTMSC]VWVKF-NH2 (SEQ ID NO: 1074)			63	38
JBT1569	Ac-SRYKWF[CGMRDkKGTMSC]VWVKF-NH2 (SEQ ID NO: 1102)			50	23
JBT1570	Ac-SRYKWF[CGMRDlKGTMSC]VWVKF-NH2 (SEQ ID NO: 1025)	0.74	88	62	43
JBT1571	Ac-SRYKWF[CGMRDpKGTMSC]VWVKF-NH2 (SEQ ID NO: 1196)			15	5
JBT1572	Ac-SRYKWF[CGMRDsKGTMSC]VWVKF-NH2 (SEQ ID NO: 1152)			28	9
JBT1573	Ac-SRYKWF[CGMRDMaGTMSC]VWVKF-NH2 (SEQ ID NO: 1197)			20	4
JBT1574	Ac-SRYKWF[CGMRDMdGTMSC]VWVKF-NH2 (SEQ ID NO: 1153)			39	15
JBT1575	Ac-SRYKWF[CGMRDMfGTMSC]VWVKF-NH2 (SEQ ID NO: 1198)			18	3
JBT1576	Ac-SRYKWF[CGMRDMlGTMSC]VWVKF-NH2 (SEQ ID NO: 1199)			19	6
JBT1577	Ac-SRYKWF[CGMRDMpGTMSC]VWVKF-NH2 (SEQ ID NO: 1200)			14	2
JBT1578	Ac-SRYKWF[CGMRDMsGTMSC]VWVKF-NH2 (SEQ ID NO: 1154)			30	10

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FIGURE 24

JBT0122 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0122	AC-SGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2002)	7.92		23	
JBT0126	Biotin-Ttds-SGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2498)	14.10		17	
JBT0221	Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2003)			29	
JBT0222*	Ac-MKSRSLGLAYFAKHSSLEVLQTRKVAAPYY-NH2 (SEQ ID NO: 2127)			0	
JBT0223*	Ac-KMQLRVYASTAHSRYLLGSSLFPKYAEVKA-NH2 (SEQ ID NO: 2297)			0	
JBT0224	Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKK-NH2 (SEQ ID NO: 2298)			26	
JBT0225	Ac-KKSGYASFPLAVQLHVSKRSKEMALARKK-NH2 (SEQ ID NO: 2128)			12	
JBT0226	Ac-KKSGYASFPLAVQLHVSKRSKEMAKK-NH2 (SEQ ID NO: 2299)			6	
JBT0227	Ac-KKSGYASFPLAVQLHVSKRSKKK-NH2 (SEQ ID NO: 2300)			0	
JBT0228	Ac-KKASFPLAVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2016)			21	
JBT0229	Ac-KKPLAVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2129)			0	
JBT0230	Ac-KKVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2301)			0	
JBT0231	Ac-KKHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2302)			0	
JBT0232	Ac-KKGYASFPLAVQLHVSKRSKEMKK-NH2 (SEQ ID NO: 2303)			6	
JBT0233	Ac-KKYASFPLAVQLHVSKRSKEMAKK-NH2 (SEQ ID NO: 2304)			8	
JBT0234	Ac-KKVQLHVSKRSKEMALARLYYKKK-NH2 (SEQ ID NO: 2305)			0	
JBT0235	Ac-KKQLHVSKRSKEMALARLYYKTKK-NH2 (SEQ ID NO: 2306)			0	
JBT0236	Ac-KKLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2307)			0	
JBT0237	Ac-KKSGYASFPLAVQLHVSKRSKEKK-NH2 (SEQ ID NO: 2308)			0	
JBT0359	Ac-ASFPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2309)			0	
JBT0360	Ac-SFPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2310)			1	

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FIGURE 24

JBT0122 class		FXa Inhibition assay			
Peptide	**Sequence**	**EC50 (μM)**	**Maximal inhibition (%)**	**% Inhibition @ 2.5μM**	**% Inhibition @ 0.63μM**
JBT0361	Ac-FPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2311)			0	
JBT0362	Ac-ASFPLAVQLHVSKRSKEM-NH2 (SEQ ID NO: 2312)			0	
JBT0363	Ac-ASFPLAVQLHVSKRSKE-NH2 (SEQ ID NO: 2313)			0	
JBT0364	Ac-ASFPLAVQLHVSKRSKEMAL-NH2 (SEQ ID NO: 2314)			2	
JBT0365	Ac-ASFPLAVQLHVSKRSKEMALA-NH2 (SEQ ID NO: 2315)			0	
JBT0366	Ac-ASFPLAVQLHVSKRSKEMALAR-NH2 (SEQ ID NO: 2316)			0	
JBT0367	Ac-ASFPLAVQLHVSKRSKEMALARL-NH2 (SEQ ID NO: 2317)			2	
JBT0368	Ac-ASFPLAVQLHVSKRSKEMALARLY-NH2 (SEQ ID NO: 2130)			0	
JBT0369	Ac-ASFPLAVQLHVSKRSKEMALARLYY-NH2 (SEQ ID NO: 2017)			2	
JBT0370	Ac-YASFPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2318)			2	
JBT0371	Ac-GYASFPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2319)			8	
JBT0660	Ac-AGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2004)			5	2
JBT0661	Ac-SAYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2005)			13	5
JBT0662	Ac-SGAASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2131)			4	0
JBT0664	Ac-SGYAAFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2006)			7	2
JBT0665	Ac-SGYASAPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2132)			2	0
JBT0666	Ac-SGYASFALAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2001)			12	6
JBT0667	Ac-SGYASFPAAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2018)			2	0
JBT0669	Ac-SGYASFPLAAQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2133)			0	0
JBT0670	Ac-SGYASFPLAVALHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2007)			6	0
JBT0671	Ac-SGYASFPLAVQAHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2019)			4	1
JBT0672	Ac-SGYASFPLAVQLAVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2134)			2	0
JBT0673	Ac-SGYASFPLAVQLHASKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2135)			0	0
JBT0674	Ac-SGYASFPLAVQLHVAKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2008)			10	3
JBT0675	Ac-SGYASFPLAVQLHVSARSKEMALARLYYKTS-NH2 (SEQ ID NO: 2136)			0	0

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FIGURE 24

JBT0122 class		FXa Inhibition assay			
Peptide	Sequence	EC50 (μM)	Maximal inhibition (%)	% Inhibition @ 2.5μM	% Inhibition @ 0.63μM
JBT0676	Ac-SGYASFPLAVQLHVSKASKEMALARLYYKTS-NH2 (SEQ ID NO: 2137)			0	0
JBT0677	Ac-SGYASFPLAVQLHVSKRAKEMALARLYYKTS-NH2 (SEQ ID NO: 2009)			4	0
JBT0678	Ac-SGYASFPLAVQLHVSKRSAEMALARLYYKTS-NH2 (SEQ ID NO: 2010)			6	2
JBT0679	Ac-SGYASFPLAVQLHVSKRSKAMALARLYYKTS-NH2 (SEQ ID NO: 2020)			2	0
JBT0680	Ac-SGYASFPLAVQLHVSKRSKEAALARLYYKTS-NH2 (SEQ ID NO: 2138)			0	0
JBT0682	Ac-SGYASFPLAVQLHVSKRSKEMAAARLYYKTS-NH2 (SEQ ID NO: 2139)			2	0
JBT0684	Ac-SGYASFPLAVQLHVSKRSKEMALAALYYKTS-NH2 (SEQ ID NO: 2011)			5	1
JBT0685	Ac-SGYASFPLAVQLHVSKRSKEMALARAYYKTS-NH2 (SEQ ID NO: 2021)			4	0
JBT0686	Ac-SGYASFPLAVQLHVSKRSKEMALARLAYKTS-NH2 (SEQ ID NO: 2140)			3	0
JBT0687	Ac-SGYASFPLAVQLHVSKRSKEMALARLYAKTS-NH2 (SEQ ID NO: 2141)			3	0
JBT0688	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYATS-NH2 (SEQ ID NO: 2012)			8	2
JBT0689	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYKAS-NH2 (SEQ ID NO: 2013)			10	3
JBT0690	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYKTA-NH2 (SEQ ID NO: 2014)			10	3
JBT1579	Ac-GYASFPLAVQLHVAKRSKEMA-NH2 (SEQ ID NO: 2015)			9	4
JBT1580	Ac-GYASFALSVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2320)			3	1
JBT1581	Ac-GYASFALAVQLHVAKRSKEMA-NH2 (SEQ ID NO: 2321)			11	4
JBT1582	Ac-GYASFPLAVQLHVMKRSKEMA-NH2 (SEQ ID NO: 2022)			10	3
JBT1583	Ac-GYASFALAVQLHVMKRSKEMA-NH2 (SEQ ID NO: 2322)			18	5
JBT1599	Ac-FPLAVQLHVSKRSKEMALA-NH2 (SEQ ID NO: 2323)			0	0
JBT1600	Ac-QLHVSKRSKEMALA-NH2 (SEQ ID NO: 2324)			0	0
JBT1601	Ac-SGYASFP-NH2 (SEQ ID NO: 2325)			0	0
JBT1602	Ac-LAVQLHVSKRSKEMALARL-NH2 (SEQ ID NO: 2326)			0	0

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FIGURE 25

JBT0047 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0047	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-NH2 (SEQ ID NO: 253)	1.37	20
JBT0051	Biotin-Ttds-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-NH2 (SEQ ID NO: 962)	1.04	19
JBT0055	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-Ttds-Lysin(biotin)-NH2 (SEQ ID NO: 963)	1.67	25
JBT0131	Biotin-Ttds-AFQSKKNVFVFGYFERLRAK-NH2 (SEQ ID NO: 964)	2.02	11
JBT0132	Biotin-Ttds-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 965)		11
JBT0133	Biotin-Ttds-QSKKNVFVFGYFERLRAKLT-NH2 (SEQ ID NO: 966)		9
JBT0134	Biotin-Ttds-QSKKNVFVFGYFERLRAK-NH2 (SEQ ID NO: 967)		3
JBT0155	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 8)	0.38	33
JBT0158	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKKK-NH2 (SEQ ID NO: 9)		23
JBT0159	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLKK-NH2 (SEQ ID NO: 744)		18
JBT0160	Ac-KKSGVGRLQVAFQSKKNVFVFGYFKK-NH2 (SEQ ID NO: 745)		22
JBT0161	Ac-KKSGVGRLQVAFQSKKNVFVFKK-NH2 (SEQ ID NO: 746)		6
JBT0162	Ac-KKGRLQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 10)	0.28	34
JBT0163	Ac-KKQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 11)	0.89	31
JBT0164	Ac-KKFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 12)	2.68	24
JBT0165	Ac-KKKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 712)		15
JBT0166	Biotinyl-Ttds-KKFQSKKNVFVFGYFERLRAKLKK-NH2 (SEQ ID NO: 968)	1.95	36
JBT0169	Ac-KKAFQSKKNVFVFGYFERLRAKKK-NH2 (SEQ ID NO: 254)	2.35	20
JBT0170	Ac-KKFQSKKNVFVFGYFERLRAKLKK-NH2 (SEQ ID NO: 13)	1.90	30
JBT0171	Ac-KKQSKKNVFVFGYFERLRAKLTKK -NH2 (SEQ ID NO: 255)	4.12	13

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FIGURE 25

JBT0047 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0172	Ac-KKQSKKNVFVFGYFERLRAKKK-NH2 (SEQ ID NO: 406)	3.98	8
JBT0173	Biotinyl-Ttds-SGVGRLQVAFQSKKNVFVFG-NH2 (SEQ ID NO: 971)		3
JBT0174	Ac-KKAFQSKKNVFVFGYFERLRAKLKK-NH2 (SEQ ID NO: 14)	2.90	26
JBT0175	Ac-KKAFQSKKNVFVFGYFERLRAKLTKK-NH2 (SEQ ID NO: 182)	2.29	26
JBT0293	Ac-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 256)	1.62	16
JBT0294	Ac-YQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 257)	5.31	9
JBT0295	Ac-FSSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 713)	1.80	10
JBT0296	Ac-FQNKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 407)	7.83	10
JBT0297	Ac-FQSKNNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 183)		13
JBT0298	Ac-FQSKQNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 747)		15
JBT0299	Ac-FQSKKNVFAFGYFERLRAKL-NH2 (SEQ ID NO: 408)		5
JBT0300	Ac-FQSKKNVFSFGYFERLRAKL-NH2 (SEQ ID NO: 409)		3
JBT0301	Ac-FQSKKNVFTFGYFERLRAKL-NH2 (SEQ ID NO: 410)		0
JBT0302	Ac-FQSKKNVFVAGYFERLRAKL-NH2 (SEQ ID NO: 258)		3
JBT0303	Ac-FQSKKNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 184)	8.80	9
JBT0304	Ac-FQSKKNVFVLGYFERLRAKL-NH2 (SEQ ID NO: 259)		7
JBT0305	Ac-FQSKKNVFVQGYFERLRAKL-NH2 (SEQ ID NO: 260)		0
JBT0306	Ac-FQSKKNVFVSGYFERLRAKL-NH2 (SEQ ID NO: 185)		3
JBT0307	Ac-FQSKKNVFVYGYFERLRAKL-NH2 (SEQ ID NO: 261)	8.72	4
JBT0308	Ac-FQSKKNVFVFGYKERLRAKL-NH2 (SEQ ID NO: 411)		4
JBT0309	Ac-FQSKKNVFVFGYYERLRAKL-NH2 (SEQ ID NO: 412)	8.55	7

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FIGURE 25

JBT0047 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0310	Ac-FQSKKNVFVFGYFDRLRAKL-NH2 (SEQ ID NO: 262)		5
JBT0311	Ac-FQSKKNVFVFGYFERLRAKN-NH2 (SEQ ID NO: 748)	3.72	10
JBT0335	Ac-FQSKKNVFVFGYFERLRAK-NH2 (SEQ ID NO: 749)	10.48	5
JBT0336	Ac-FQSKNNVFVAGYFDRLRAKL-OH (SEQ ID NO: 263)	12.13	5
JBT0337	Ac-FQSKNNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 16)	13.33	15
JBT0338	Ac-FQSKNNVFVQGYFDRLRAKL-NH2 (SEQ ID NO: 17)	8.42	9
JBT0339	Ac-FQSKNNVFVSGYFDRLRAKL-NH2 (SEQ ID NO: 18)		9
JBT0340	Ac-FQSKNNVFVYGYFDRLRAKL-NH2 (SEQ ID NO: 186)		9
JBT0341	Ac-FQSKNNVFVFGYFDRLRAKL-NH2 (SEQ ID NO: 187)	8.68	10
JBT0342	Ac-FQSKNNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 19)	7.15	18
JBT0343	Ac-FQSKKNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 188)		8
JBT0374	Ac-FQSKDNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 20)		19
JBT0375	Ac-FQSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 190)	2.93	30
JBT0376	Ac-FQSKKNAFVFGYFERLRAKL-NH2 (SEQ ID NO: 414)		3
JBT0377	Ac-FQSKKNQFVFGYFERLRAKL-NH2 (SEQ ID NO: 264)		8
JBT0378	Ac-FQSKKNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 21)	9.03	12
JBT0379	Ac-FQSKKNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 22)		6
JBT0380	Ac-FQSKKNVFVVGYFERLRAKL-NH2 (SEQ ID NO: 191)	7.58	10
JBT0381	Ac-FQSPKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 265)		9
JBT0385	Ac-FQSKKNNFVFGYFERLRAKL-NH2 (SEQ ID NO: 672)		2
JBT0386	Ac-FQSKKNPFVFGYFERLRAKL-NH2 (SEQ ID NO: 415)		5

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FIGURE 25

JBT0047 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0388	Ac-FQSKKNVHVFGYFERLRAKL-NH2 (SEQ ID NO: 192)		6
JBT0389	Ac-FQSKKNVVVFGYFERLRAKL-NH2 (SEQ ID NO: 673)		8
JBT0390	Ac-FQSKKNVFQFGYFERLRAKL-NH2 (SEQ ID NO: 674)		5
JBT0391	Ac-FQSKKNVFVGGYFERLRAKL-NH2 (SEQ ID NO: 266)		1
JBT0392	Ac-FQSKKNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 23)		4
JBT0393	Ac-FQSKKNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 193)		3
JBT0394	Ac-FQSKKNVFVMGYFERLRAKL-NH2 (SEQ ID NO: 24)	4.38	8
JBT0395	Ac-FQSKKNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 25)		7
JBT0396	Ac-FQSKKNVFVPGYFERLRAKL-NH2 (SEQ ID NO: 194)	8.71	6
JBT0397	Ac-FQSKKNVFVRGYFERLRAKL-NH2 (SEQ ID NO: 195)		4
JBT0398	Ac-FQSKKNVFVFGYFEELRAKL-NH2 (SEQ ID NO: 196)		8
JBT0399	Ac-FQSKKNVFVFGYFELLRAKL-NH2 (SEQ ID NO: 750)		7
JBT0400	Ac-FQSKKNVFVFGYFLRLRAKL-NH2 (SEQ ID NO: 267)		13
JBT0401	Ac-FQSKKNVFVFGYFERLRAVL-NH2 (SEQ ID NO: 416)		4
JBT0402	Biotin-Ttds-FQSKNNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 973)		11
JBT0403	Biotin-Ttds-FQSKKNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 972)		9
JBT0404	Biotin-Ttds-FQSKKNVFVFGYFDRLRAKL-NH2 (SEQ ID NO: 974)	3.38	14
JBT0405	Biotin-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 975)		15
JBT0406	Ac-Ttds-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 268)		9
JBT0471	Ac-FQSKGNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 26)		32
JBT0472	Ac-FQSKGNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 27)		22

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FIGURE 25

JBT0047 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0473	Ac-FQSKGNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 28)		24
JBT0474	Ac-FQSKGNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 29)		33
JBT0475	Ac-FQSKGNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 30)		30
JBT0476	Ac-FQSKGNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 31)		33
JBT0477	Ac-FQSKGNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 32)	2.22	36
JBT0478	Ac-FQSKDNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 33)		19
JBT0479	Ac-FQSKDNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 34)		18
JBT0480	Ac-FQSKDNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 35)		16
JBT0481	Ac-FQSKDNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 36)		15
JBT0482	Ac-FQSKDNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 37)		23
JBT0483	Ac-FQSKDNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 38)		24
JBT0484	Ac-FQSKDNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 976)	4.89	19
JBT0485	Ac-FQSKNNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 39)		21
JBT0486	Ac-FQSKNNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 40)		19
JBT0487	Ac-FQSKNNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 41)		19
JBT0488	Ac-FQSKNNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 42)		18
JBT0489	Ac-FQSKNNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 43)		21
JBT0490	Ac-FQSKQNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 44)		9
JBT0491	Ac-FQSKQNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 197)		10
JBT0492	Ac-FQSKQNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 198)		7
JBT0493	Ac-FQSKQNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 199)		7

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FIGURE 25

JBT0047 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0494	Ac-FQSKQNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 45)		15
JBT0495	Ac-FQSKQNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 200)		15
JBT0497	Ac-NmetPhe-QSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 201)	1.13	15
JBT0498	Ac-FQ-NmetSer-KGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 675)		9
JBT0499	Ac-FQS-NmetLys-GNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 202)		11
JBT0500	Ac-FQSK-NmetGly-NVFVFGYFERLRAKL-NH2 (SEQ ID NO: 46)	1.11	45
JBT0501	Ac-FQSKGN-NmetV-FVFGYFERLRAKL-NH2 (SEQ ID NO: 417)		0
JBT0502	Ac-FQSKGNV-NmetPhe-VFGYFERLRAKL-NH2 (SEQ ID NO: 751)		1
JBT0503	Ac-FQSKGNVF-NmetV-FGYFERLRAKL-NH2 (SEQ ID NO: 676)		0
JBT0504	Ac-FQSKGNVFV-NmetPhe-GYFERLRAKL-NH2 (SEQ ID NO: 418)		5
JBT0505	Ac-FQSKGNVFVF-NmetGly-YFERLRAKL-NH2 (SEQ ID NO: 269)		9
JBT0506	Ac-FQSKGNVFVFG-NmetTyr-FERLRAKL-NH2 (SEQ ID NO: 714)		0
JBT0507	Ac-FQSKGNVFVFGY-NmetPhe-ERLRAKL-NH2 (SEQ ID NO: 677)		0
JBT0508	Ac-FQSKGNVFVFGYF-NmetGlu-RLRAKL-NH2 (SEQ ID NO: 678)		2
JBT0509	Ac-FQSKGNVFVFGYFE-Nmr-LRAKL-NH2 (SEQ ID NO: 752)		2
JBT0510	Ac-FQSKGNVFVFGYFER-NmetLeu-RAKL-NH2 (SEQ ID NO: 753)		3
JBT0511	Ac-FQSKGNVFVFGYFERL-Nmr-AKL-NH2 (SEQ ID NO: 754)		7
JBT0512	Ac-FQSKGNVFVFGYFERLR-NmetAla-KL-NH2 (SEQ ID NO: 679)		1
JBT0513	Ac-FQSKGNVFVFGYFERLRA-NmetLys-L-NH2 (SEQ ID NO: 755)		3
JBT0514	Ac-FQSKGNVFVFGYFERLRAK-NmetLeu-NH2 (SEQ ID NO: 419)		5
JBT0515	Ac-bHomoPhe-QSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 420)		0

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FIGURE 25

JBT0047 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0516	Ac-F-bHomoGln-SKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 680)		0
JBT0517	Ac-FQ-bHomoSer-KGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 421)		0
JBT0518	Ac-FQS-bHomoLys-GNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 422)		0
JBT0519	Ac-FQSK-bAla-NVFVFGYFERLRAKL-NH2 (SEQ ID NO: 270)		0
JBT0520	Ac-FQSKG-bGln-VFVFGYFERLRAKL-NH2 (SEQ ID NO: 271)		0
JBT0521	Ac-FQSKGN-bLeu-FVFGYFERLRAKL-NH2 (SEQ ID NO: 756)		0
JBT0522	Ac-FQSKGNV-bHomoPhe-VFGYFERLRAKL-NH2 (SEQ ID NO: 681)		0
JBT0523	Ac-FQSKGNVF-bLeu-FGYFERLRAKL-NH2 (SEQ ID NO: 757)		0
JBT0524	Ac-FQSKGNVFV-bHomoPhe-GYFERLRAKL-NH2 (SEQ ID NO: 682)		0
JBT0525	Ac-FQSKGNVFVF-bAla-YFERLRAKL-NH2 (SEQ ID NO: 758)		0
JBT0526	Ac-FQSKGNVFVFG-bHomoTyr-FERLRAKL-NH2 (SEQ ID NO: 759)		0
JBT0527	Ac-FQSKGNVFVFGY-bHomoPhe-ERLRAKL-NH2 (SEQ ID NO: 683)		0
JBT0528	Ac-FQSKGNVFVFGYF-bE-RLRAKL-NH2 (SEQ ID NO: 272)		8
JBT0529	Ac-FQSKGNVFVFGYFE-bHomoArg-LRAKL-NH2 (SEQ ID NO: 760)		0
JBT0530	Ac-FQSKGNVFVFGYFER-Btl-RAKL-NH2 (SEQ ID NO: 684)		0
JBT0531	Ac-FQSKGNVFVFGYFERL-bHomoArg-AKL-NH2 (SEQ ID NO: 203)		0
JBT0532	Ac-FQSKGNVFVFGYFERLR-bAla-KL-NH2 (SEQ ID NO: 423)		0
JBT0533	Ac-FQSKGNVFVFGYFERLRA-bHomoK-L-NH2 (SEQ ID NO: 424)		0
JBT0534	Ac-FQSKGNVFVFGYFERLRAK-Btl-NH2 (SEQ ID NO: 204)		8
JBT0535	Ac-FQSKGNVFVFGYFE-Cit-LRAKL-NH2 (SEQ ID NO: 685)		1
JBT0536	Ac-FQSKGNVFVFGYFERL-Cit-AKL-NH2 (SEQ ID NO: 273)		11

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FIGURE 25

JBT0047 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0537	Ac-FQSKGNVFVFGYFE-Nle-LRAKL-NH2 (SEQ ID NO: 761)		7
JBT0538	Ac-FQSKGNVFVFGYFERL-Nle-AKL-NH2 (SEQ ID NO: 425)		15
JBT0564	Ac-FQSKKNVFVFGYFKRLRAKL-NH2 (SEQ ID NO: 205)		0
JBT0578	Ac-FQSKKNVFVFGYFFRLRAKL-NH2 (SEQ ID NO: 441)		0
JBT0613	Ac-FQSKGNVFVFGYFERLRAKL-OH (SEQ ID NO: 283)		6
JBT0614	NH2-FQSKGNVFVFGYFERLRAKL-OH (SEQ ID NO: 720)		4
JBT0615	NH2-FQSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 284)		6
JBT0651	Ac-FQSKGNVHVKGYFERLRAKL-NH2 (SEQ ID NO: 48)		6
JBT0652	Ac-FQSKGNVHVDGYFERLRAKL-NH2 (SEQ ID NO: 49)		8
JBT0653	Ac-FQSPGNVHVKGYFERLRAKL-NH2 (SEQ ID NO: 50)		9
JBT0654	Ac-FQSPGNVHVDGYFERLRAKL-NH2 (SEQ ID NO: 51)		11
JBT0655	Ac-FQSKGNIFVFGYFERLRAKL-NH2 (SEQ ID NO: 52)		9
JBT0656	Ac-FQSKGNLFVFGYFERLRAKL-NH2 (SEQ ID NO: 286)		5
JBT0657	Ac-FQSKGNVFIFGYFERLRAKL-NH2 (SEQ ID NO: 287)		8
JBT0658	Ac-FQSKGNVFLFGYFERLRAKL-NH2 (SEQ ID NO: 694)		3
JBT0663	Ac-FQSKaNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 53)		22
JBT0668	Ac-FQSKaNVFVTGYFARLRAKL-NH2 (SEQ ID NO: 54)		16
JBT0681	Ac-FQSK-Nmg-AVFVFGYFARLRAKL-NH2 (SEQ ID NO: 206)		25
JBT0683	Ac-FQSK-Nmg-AVFVDGYFARLRAKL-NH2 (SEQ ID NO: 55)		38
JBT0708	Ac-FQSKaNVFVAGYFERLRAKL-NH2 (SEQ ID NO: 59)		25
JBT0714	Ac-FQSKaAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 60)		29

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FIGURE 25

JBT0047 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0717	Ac-FQSK-Nmg-NVFVDGYFERLRAKL-NH2 (SEQ ID NO: 61)		50
JBT0720	Ac-FQSK-Nmg-NVFVTGYFARLRAKL-NH2 (SEQ ID NO: 62)		30
JBT0740	Ac-FQSKGNVFVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 66)		36
JBT0754	Ac-FQSK-Nmg-AVFVTGYFARLRAKL-NH2 (SEQ ID NO: 67)		27
JBT0765	Ac-FQSKGNVFVDGYFERL-Eew-AKL-NH2 (SEQ ID NO: 68)		37
JBT0780	Ac-FQSK-Nmg-NVFVFGYFARLRAKL-NH2 (SEQ ID NO: 70)	1.92	35
JBT0789	Ac-FQSK-Nmg-NVFVDGYFERL-Nle-AKL-NH2 (SEQ ID NO: 71)	0.79	51
JBT0806	Ac-FQSK-Nmg-NVFVAGYFARLRAKL-NH2 (SEQ ID NO: 74)	2.42	38
JBT0837	Ac-FQSK-Nmg-NVFVTGYFERL-Nle-AKL-NH2 (SEQ ID NO: 213)		33
JBT0844	Ac-FQSKaNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 300)		23
JBT0850	Ac-FQSKaAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 302)		24
JBT0854	Ac-FQSKaAVFVAGYFARL-Nle-AKL-NH2 (SEQ ID NO: 490)		14
JBT0870	Ac-FQSKaAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 214)		20
JBT0886	Ac-FQSKaNVFVDGYFARLRAKL-NH2 (SEQ ID NO: 215)		21
JBT0919	Ac-FQSKaNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 79)		24
JBT0931	Ac-FQSKGNVFVDGYFERL-Eag-AKL-NH2 (SEQ ID NO: 82)		24
JBT0946	Ac-FQSKGNVFVDGYFERL-Dab-AKL-NH2 (SEQ ID NO: 85)		29
JBT0950	Ac-FQSKaNVFVFGYFARLRAKL-NH2 (SEQ ID NO: 217)		14
JBT0973	Ac-FQSKGNVFVDGYFERL-Cha-AKL-NH2 (SEQ ID NO: 92)	1.21	32
JBT1006	Ac-FQSKGNVFVDGYFERL-Hle-AKL-NH2 (SEQ ID NO: 96)	1.35	35
JBT1035	Ac-FQSK-Nmg-AVFVAGYFARL-Nle-AKL-NH2 (SEQ ID NO: 218)		25

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FIGURE 25

JBT0047 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT1037	Ac-FQSKGNVFVDGYFERL-Nle-AKL-NH2 (SEQ ID NO: 104)		30
JBT1043	Ac-FQSKaNVFVDGYFERL-Nle-AKL-NH2 (SEQ ID NO: 106)		27
JBT1082	Ac-FQSKaAVFVFGYFARLRAKL-NH2 (SEQ ID NO: 565)		16
JBT1084	Ac-FQSK-Nmg-AVFVAGYFARLRAKL-NH2 (SEQ ID NO: 109)		28
JBT1122	Ac-FQSK-Nmg-NVFVDGYFARL-Nle-AKL-NH2 (SEQ ID NO: 113)	0.81	43
JBT1133	Ac-FQSKkAVFVDGYFARLRAKL-NH2(SEQ ID NO: 115)		31
JBT1134	Ac-FQSKGAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 116)		28
JBT1135	Ac-FQSKDAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 221)		22
JBT1136	Ac-FQSKdAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 117)		36
JBT1137	Ac-FQSKkAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 118)		23
JBT1138	Ac-FQSKGAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 222)		19
JBT1139	Ac-FQSKDAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 223)		16
JBT1140	Ac-FQSKdAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 119)		29
JBT1141	Ac-FQSKkAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 120)		13
JBT1142	Ac-FQSKGAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 224)		11
JBT1143	Ac-FQSKDAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 225)		6
JBT1144	Ac-FQSKdAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 121)		15
JBT1145	Ac-FQSKkAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 122)		13
JBT1146	Ac-FQSKGAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 226)		12
JBT1147	Ac-FQSKDAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 227)		14
JBT1148	Ac-FQSKdAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 123)		27

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FIGURE 25

JBT0047 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT1149	Ac-FQSKGNVFvFGYFERLRAKL-NH2 (SEQ ID NO: 936)		0
JBT1151	Ac-FQSKKNVFVFGYFERLRAKD-NH2 (SEQ ID NO: 937)		0
JBT1152	Ac-FQSKKNVFFFGYFERLRAKL-NH2 (SEQ ID NO: 735)		0
JBT1153	Ac-FQSKKNVFVFGYFERLGAKL-NH2 (SEQ ID NO: 705)		4
JBT1155	Ac-FQSK-Nmg-NVFVTGYFERLRAKL-NH2 (SEQ ID NO: 125)		32
JBT1156	Ac-FQSKaNVFVDGYFARL-Nle-AKL-NH2 (SEQ ID NO: 126)		22
JBT1157	Ac-FQSKaNVFVAGYFARLRAKL-NH2 (SEQ ID NO: 127)		22
JBT1162	Ac-FQSKpNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 129)		51
JBT1396	Ac-FQSK-Nmg-NVFVAGYFERLRAKL-NH2 (SEQ ID NO: 161)		34
JBT1584	Ac-FQSK-Nmg-NVFVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 164)		57
JBT1585	Ac-FQSK-Nmg-NVFVTGYFERL-Aib-AKL-NH2 (SEQ ID NO: 165)		48
JBT1590	Ac-FQSKGNVFV[CGYFERL-Aib-AKLC]-NH2 (SEQ ID NO: 401)		22
JBT1591	Ac-FQSKGNVFVDGYF[CRL-Aib-AKLC]-NH2 (SEQ ID NO: 168)		23
JBT1592	Ac-[CFQSKGNVFVDGYFERLC]AKL-NH2 (SEQ ID NO: 402)		11
JBT1593	Ac-FQSK[CNVFVDGYFERLC]AKL-NH2 (SEQ ID NO: 670)		13
JBT1594	Ac-FQSKGNVFV[CGYFERLC]AKL-NH2 (SEQ ID NO: 671)		5
JBT1595	Ac-[CFQSKGNVFVC]GYFERL-Aib-AKL-NH2 (SEQ ID NO: 403)		12
JBT1596	Ac-[CFQSKGC]VFVEGYFERL-Aib-AKL-NH2 (SEQ ID NO: 404)		13
JBT1843	Ac-FQSKGNIFVDGYFERLHAKL-NH2 (SEQ ID NO: 169)		7
JBT1844	Ac-FQSKNNVFVDGYFKRLRAKL-NH2 (SEQ ID NO: 170)		6
JBT1845	Ac-FQSYKHVFVDGYFERLRAKL-NH2 (SEQ ID NO: 249)		6

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FIGURE 25

JBT0047 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT1846	Ac-FQSKGIVFVDGYFKRLRAKL-NH2 (SEQ ID NO: 250)		7
JBT1847	Ac-YQTKGNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 171)		16
JBT1853	PEG(40kD)-FQSKGNVFVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 175)		56
JBT1855	Ac-FQSKGNVFVDGYFERL-Aib-AKLC(PEG40kD)-NH2 (SEQ ID NO: 252)		59

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FIGURE 26

JBT0120 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0120	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1047)	0.88	45
JBT0124	Biotin-Ttds-SGASRYKWFCGMRDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1290)	1.26	36
JBT0247	Ac-SGASRYKWFCGMRDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1213)	0.61	42
JBT0248	Ac-KKSGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTSKK-NH2 (SEQ ID NO: 1001)	0.27	52
JBT0249	Ac-KKSGASRYKWFCGMRDMKGTMSKK-NH2 (SEQ ID NO: 1214)	1.06	16
JBT0250	Ac-KKSRYKWF[CGMRDMKGTMSC]VWKK-NH2 (SEQ ID NO: 1201)	0.61	19
JBT0251	Ac-KKKWF[CGMRDMKGTMSC]VWVKFKK-NH2 (SEQ ID NO: 1202)	0.63	23
JBT0252	Ac-KK[CGMRDMKGTMSC]VWVKFRYDKK-NH2 (SEQ ID NO: 1215)	1.02	18
JBT0253	Ac-KKMRDMKGTMSCVWVKFRYDTSKK-NH2 (SEQ ID NO: 1216)	0.55	23
JBT0319	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRY-NH2 (SEQ ID NO: 1002)		60
JBT0320	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVK-NH2 (SEQ ID NO: 1103)	6.40	7
JBT0321	Ac-SGASRYKWF[CGMRDMKGTMSC]V-NH2 (SEQ ID NO: 1217)		1
JBT0322	Ac-SGASRYKWFCGMRDMKGTM-NH2 (SEQ ID NO: 1218)		3
JBT0323	Ac-SRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1076)	0.77	48
JBT0324	Ac-KWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1155)	2.44	18
JBT0325	Ac-[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1219)		4
JBT0326	Ac-RDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1220)		3
JBT0327	Ac-SGASRYKWFCGMRDMKGTMS-NH2 (SEQ ID NO: 1222)		0
JBT0328	Ac-SRYKWF[CGMRDMKGTMSC]VW-NH2 (SEQ ID NO: 1223)		1
JBT0329	Ac-KWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1156)		8
JBT0330	Ac-[CGMRDMKGTMSC]VWVKFRYD-NH2 (SEQ ID NO: 1224)		7
JBT0331	Ac-MRDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1225)		5

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FIGURE 26

JBT0120 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0332	Ac-SRYKWFCGMRDMKGTMSCVW-NH2 (SEQ ID NO: 1206)		5
JBT0333	Ac-KWFCGMRDMKGTMSCVWVKF-NH2 (SEQ ID NO: 1207)		11
JBT0334	Ac-CGMRDMKGTMSCVWVKFRYD-NH2 (SEQ ID NO: 1227)		3
JBT0409	Ac-SGASRYKWFSGMRDMKGTMSSVWVKFRYDTS-NH2 (SEQ ID NO: 1204)		5
JBT0410	Ac-SGASRYKWFSG-Nle-RD-Nle-KGT-Nle-SSVWVKFRYDTS-NH2 (SEQ ID NO: 1208)		7
JBT0411	Ac-SGASRYKWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKFRYDTS-NH2 (SEQ ID NO: 1104)		24
JBT0412	Ac-KWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKF-NH2 (SEQ ID NO: 1209)		12
JBT0413	Ac-KWFSGMRDMKGTMSSVWVKF-NH2 (SEQ ID NO: 1228)		2
JBT0414	Ac-KWFSG-Nle-RD-Nle-KGT-Nle-SSVWVKF-NH2 (SEQ ID NO: 1205)		8
JBT0415	Ac-SRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1003)	0.19	52
JBT0416	Ac-SRYKWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKF-NH2 (SEQ ID NO: 1077)		46
JBT0417	Ac-SRYKWFSGMRDMKGTMSSVWVKF-NH2 (SEQ ID NO: 1229)		11
JBT0418	Ac-SRYKWFSG-Nle-RD-Nle-KGT-Nle-SSVWVKF-NH2 (SEQ ID NO: 1210)		11
JBT0419	Ac-SGGRKHKHFLRSNGKPSRALSSMHFWRWSTS-NH2 (SEQ ID NO: 1293)		7
JBT0435*	Ac-RKRDVSGKM[CWSGFGSKWFRC]ADMMTYYSVT-NH2 (SEQ ID NO: 1211)		5
JBT0436*	Ac-RWSGKSTYS[CDAMVRRMGMKC]FSFGWTDVYK-NH2 (SEQ ID NO: 1212)		0
JBT0437	Ac-ARYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1048)		52
JBT0438	Ac-SAYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1078)		46
JBT0439	Ac-SRAKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1105)		38
JBT0440	Ac-SRYAWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1106)		34
JBT0441	Ac-SRYKAF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1230)		1
JBT0442	Ac-SRYKWA[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1231)		17
JBT0443	Ac-SRYKWFAGMRDMKGTMSCVWVKF-NH2 (SEQ ID NO: 1157)		7
JBT0444	Ac-SRYKWF[CAMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1004)		63

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FIGURE 26

JBT0120 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0445	Ac-SRYKWF[CGARDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1107)		12
JBT0446	Ac-SRYKWF[CGMADMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1079)		46
JBT0447	Ac-SRYKWF[CGMRAMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1158)		15
JBT0448	Ac-SRYKWF[CGMRDAKGTMSC]VWVKF-NH2 (SEQ ID NO: 1108)		33
JBT0449	Ac-SRYKWF[CGMRDMAGTMSC]VWVKF-NH2 (SEQ ID NO: 1080)		55
JBT0450	Ac-SRYKWF[CGMRDMKATMSC]VWVKF-NH2 (SEQ ID NO: 1159)		34
JBT0451	Ac-SRYKWF[CGMRDMKGAMSC]VWVKF-NH2 (SEQ ID NO: 1081)		46
JBT0452	Ac-SRYKWF[CGMRDMKGTASC]VWVKF-NH2 (SEQ ID NO: 1109)		31
JBT0453	Ac-SRYKWF[CGMRDMKGTMAC]VWVKF-NH2 (SEQ ID NO: 1110)	0.97	40
JBT0454	Ac-SRYKWFCGMRDMKGTMSAVWVKF-NH2 (SEQ ID NO: 1160)		0
JBT0455	Ac-SRYKWF[CGMRDMKGTMSC]AWVKF-NH2 (SEQ ID NO: 1049)	0.95	30
JBT0456	Ac-SRYKWFCGMRDMKGTMSCVAVKF-NH2 (SEQ ID NO: 1232)		0
JBT0457	Ac-SRYKWF[CGMRDMKGTMSC]VWAKF-NH2 (SEQ ID NO: 1161)		17
JBT0458	Ac-SRYKWF[CGMRDMKGTMSC]VWVAF-NH2 (SEQ ID NO: 1082)		52
JBT0459	Ac-SRYKWF[CGMRDMKGTMSC]VWVKA-NH2 (SEQ ID NO: 1083)		38
JBT0460	Ac-RYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1050)		51
JBT0461	Ac-YKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1111)		40
JBT0462	Ac-SRYKWFGM[CRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1233)		0
JBT0463	Ac-SRYKWFGMRD[CMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1234)		11
JBT0464	Ac-SRYKWFGMRDMK[CGTMSC]VWVKF-NH2 (SEQ ID NO: 1235)		10
JBT0465	Ac-SRYKWF[CGMRDMKGTC]MSVWVKF-NH2 (SEQ ID NO: 1236)		7
JBT0466	Ac-SRYKWF[CGMRDMKC]GTMSVWVKF-NH2 (SEQ ID NO: 1237)		11
JBT0467	Ac-SRYKWF[CGMRDC]MKGTMSVWVKF-NH2 (SEQ ID NO: 1238)		11
JBT0468	Ac-SRYKWFG[CMRDMKGTMC]SVWVKF-NH2 (SEQ ID NO: 1239)		7

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FIGURE 26

JBT0120 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0469	Ac-SRYKWFGM[CRDMKGTC]MSVWVKF-NH2 (SEQ ID NO: 1240)		11
JBT0470	Ac-SRYKWFGMR[CDMKGC]TMSVWVKF-NH2 (SEQ ID NO: 1162)		2
JBT0617	Ac-SRYKWF[homoC-GMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1112)		21
JBT0618	Ac-SRYKWF[CGMRDMKGTMS-homoC]VWVKF-NH2 (SEQ ID NO: 1163)		11
JBT0619	Ac-SRYKWF[homoC-GMRDMKGTMS-homoC]VWVKF-NH2 (SEQ ID NO: 1241)		1
JBT0620	Ac-SRYKWF[Dap-GMRDMKGTMS-D]VWVKF-NH2 (SEQ ID NO: 1242)		0
JBT0625	Ac-SRYKWF[CMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1244)		0
JBT0626	Ac-SRYKWF[CGMRDKGTMSC]VWVKF-NH2 (SEQ ID NO: 1164)		7
JBT0627	Ac-SRYKWF[CGMRDMGTMSC]VWVKF-NH2 (SEQ ID NO: 1165)		3
JBT0628	Ac-SRYKWF[CGMRDMKGMSC]VWVKF-NH2 (SEQ ID NO: 1166)		0
JBT0629	Ac-SRYKWF[cGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1167)		3
JBT0631	Ac-SRYKWF[CGmRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1113)		18
JBT0632	Ac-SRYKWF[CGMrDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1245)		0
JBT0633	Ac-SRYKWF[CGMRdMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1168)		2
JBT0634	Ac-SRYKWF[CGMRDmKGTMSC]VWVKF-NH2 (SEQ ID NO: 1051)	0.47	44
JBT0635	Ac-SRYKWF[CGMRDMkGTMSC]VWVKF-NH2 (SEQ ID NO: 1114)		10
JBT0637	Ac-SRYKWF[CGMRDMKGtMSC]VWVKF-NH2 (SEQ ID NO: 1169)		0
JBT0638	Ac-SRYKWF[CGMRDMKGTmSC]VWVKF-NH2 (SEQ ID NO: 1246)		0
JBT0639	Ac-SRYKWF[CGMRDMKGTMsC]VWVKF-NH2 (SEQ ID NO: 1170)		3
JBT0640	Ac-SRYKWF[CGMRDMKGTMSc]VWVKF-NH2 (SEQ ID NO: 1171)		1
JBT0641	Ac-SRYKWF[CPMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1247)		0
JBT0642	Ac-SRYKWF[CGPRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1248)		0
JBT0643	Ac-SRYKWF[CGMPDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1006)		31
JBT0644	Ac-SRYKWF[CGMRPMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1172)		17

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FIGURE 26

JBT0120 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0645	Ac-SRYKWF[CGMRDPKGTMSC]VWVKF-NH2 (SEQ ID NO: 1084)		23
JBT0646	Ac-SRYKWF[CGMRDMPGTMSC]VWVKF-NH2 (SEQ ID NO: 1173)		2
JBT0647	Ac-SRYKWF[CGMRDMKPTMSC]VWVKF-NH2 (SEQ ID NO: 1249)		0
JBT0648	Ac-SRYKWF[CGMRDMKGPMSC]VWVKF-NH2 (SEQ ID NO: 1174)		3
JBT0649	Ac-SRYKWF[CGMRDMKGTPSC]VWVKF-NH2 (SEQ ID NO: 1175)		2
JBT0650	Ac-SRYKWF[CGMRDMKGTMPC]VWVKF-NH2 (SEQ ID NO: 1250)		0
JBT0787	Ac-SRYKWF[CG-SeMet-RD-SeMet-KGT-SeMet-SC]VWVKF-NH2 (SEQ ID NO: 1292)		55
JBT1416	Ac-DRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1007)		47
JBT1426	Ac-SRYKWF[CGMRDMDGTMSC]VWVKF-NH2 (SEQ ID NO: 1008)		40
JBT1436	Ac-FRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1053)		51
JBT1437	Ac-SFYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO:1010)		53
JBT1438	Ac-SRFKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1011)		49
JBT1445	Ac-SRYKWF[CGMRDFKGTMSC]VWVKF-NH2 (SEQ ID NO: 1012)		39
JBT1446	Ac-SRYKWF[CGMRDMFGTMSC]VWVKF-NH2 (SEQ ID NO: 1054)		46
JBT1448	Ac-SRYKWF[CGMRDMKGFMSC]VWVKF-NH2 (SEQ ID NO: 1123)		47
JBT1449	Ac-SRYKWF[CGMRDMKGTFSC]VWVKF-NH2 (SEQ ID NO: 1013)		55
JBT1455	Ac-GRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1014)		49
JBT1474	Ac-KRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1016)		59
JBT1481	Ac-SRYKWF[CGMKDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1017)		55
JBT1483	Ac-SRYKWF[CGMRDKKGTMSC]VWVKF-NH2 (SEQ ID NO: 1057)		50
JBT1492	Ac-LRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1018)		52
JBT1493	Ac-SLYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1060)		43
JBT1502	Ac-SRYKWF[CGMRDLKGTMSC]VWVKF-NH2 (SEQ ID NO: 1019)		46
JBT1506	Ac-SRYKWF[CGMRDMKGTLSC]VWVKF-NH2 (SEQ ID NO:1064)		45

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FIGURE 26

JBT0120 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT1512	Ac-SRYKWF[CGMRDMKGTMSC]VWVKL-NH2 (SEQ ID NO: 1020)		44
JBT1541	Ac-SRYKWF[CGMRDMKGTMSC]VWVPF-NH2 (SEQ ID NO: 1023)		41
JBT1544	H-SRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1024)		59
JBT1570	Ac-SRYKWF[CGMRDIKGTMSC]VWVKF-NH2 (SEQ ID NO: 1025)		41

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FIGURE 27

JBT0122 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0122	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2002)	6.70	7
JBT0126	Biotin-Ttds-SGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2498)	5.05	5
JBT0221	Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2003)	3.37	19
JBT0222	Ac-MKSRSLGLAYFAKHSSLEVLQTRKVAAPYY-NH2 (SEQ ID NO: 2127)		4
JBT0223	Ac-KMQLRVYASTAHSRYLLGSSLFPKYAEVKA-NH2 (SEQ ID NO: 2297)		4
JBT0224	Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKK-NH2 (SEQ ID NO: 2298)	3.49	18
JBT0225	Ac-KKSGYASFPLAVQLHVSKRSKEMALARKK-NH2 (SEQ ID NO: 2128)		0
JBT0226	Ac-KKSGYASFPLAVQLHVSKRSKEMAKK-NH2 (SEQ ID NO: 2299)		0
JBT0227	Ac-KKSGYASFPLAVQLHVSKRSKKK-NH2 (SEQ ID NO: 2300)		5
JBT0228	Ac-KKASFPLAVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2016)	6.93	9
JBT0229	Ac-KKPLAVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2129)		0
JBT0230	Ac-KKVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2301)		0
JBT0231	Ac-KKHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2302)		0
JBT0232	Ac-KKGYASFPLAVQLHVSKRSKEMKK-NH2 (SEQ ID NO: 2303)		2
JBT0233	Ac-KKYASFPLAVQLHVSKRSKEMAKK-NH2 (SEQ ID NO: 2304)		6
JBT0234	Ac-KKVQLHVSKRSKEMALARLYYKKK-NH2 (SEQ ID NO: 2305)		3
JBT0235	Ac-KKQLHVSKRSKEMALARLYYKTKK-NH2 (SEQ ID NO: 2306)		0
JBT0236	Ac-KKLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2307)		0
JBT0237	Ac-KKSGYASFPLAVQLHVSKRSKEKK-NH2 (SEQ ID NO: 2308)		2
JBT0359	Ac-ASFPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2309)		6
JBT0360	Ac-SFPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2310)		2
JBT0361	Ac-FPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2311)		9
JBT0362	Ac-ASFPLAVQLHVSKRSKEM-NH2 (SEQ ID NO: 2312)		6
JBT0363	Ac-ASFPLAVQLHVSKRSKE-NH2 (SEQ ID NO: 2313)		9

FIGURE 27

JBT0122 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0364	Ac-ASFPLAVQLHVSKRSKEMAL-NH2 (SEQ ID NO: 2314)		12
JBT0365	Ac-ASFPLAVQLHVSKRSKEMALA-NH2 (SEQ ID NO: 2315)		9
JBT0366	Ac-ASFPLAVQLHVSKRSKEMALAR-NH2 (SEQ ID NO: 2316)		11
JBT0367	Ac-ASFPLAVQLHVSKRSKEMALARL-NH2 (SEQ ID NO: 2317)		10
JBT0368	Ac-ASFPLAVQLHVSKRSKEMALARLY-NH2 (SEQ ID NO: 2130)		8
JBT0369	Ac-ASFPLAVQLHVSKRSKEMALARLYY-NH2 (SEQ ID NO: 2017)		9
JBT0370	Ac-YASFPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2318)		0
JBT0371	Ac-GYASFPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2319)		5
JBT0660	Ac-AGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2004)		0
JBT0661	Ac-SAYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2005)		3
JBT0662	Ac-SGAASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2131)		0
JBT0664	Ac-SGYAAFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2006)		3
JBT0665	Ac-SGYASAPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2132)		0
JBT0666	Ac-SGYASFALAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2001)		5
JBT0667	Ac-SGYASFPAAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2018)		0
JBT0669	Ac-SGYASFPLAAQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2133)		0
JBT0670	Ac-SGYASFPLAVALHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2007)		2
JBT0671	Ac-SGYASFPLAVQAHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2019)		7
JBT0672	Ac-SGYASFPLAVQLAVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2134)		3
JBT0673	Ac-SGYASFPLAVQLHASKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2135)		5
JBT0674	Ac-SGYASFPLAVQLHVAKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2008)		5
JBT0675	Ac-SGYASFPLAVQLHVSARSKEMALARLYYKTS-NH2 (SEQ ID NO: 2136)		7
JBT0676	Ac-SGYASFPLAVQLHVSKASKEMALARLYYKTS-NH2 (SEQ ID NO: 2137)		1
JBT0677	Ac-SGYASFPLAVQLHVSKRAKEMALARLYYKTS-NH2 (SEQ ID NO: 2009)		2

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FIGURE 27

JBT0122 class		Extrinsic Tenase Inhibition assay	
Peptide	**Sequence**	**EC50 (μM)**	**% Inhibition @ 2.5μM**
JBT0678	Ac-SGYASFPLAVQLHVSKRSAEMALARLYYKTS-NH2 (SEQ ID NO: 2010)		2
JBT0679	Ac-SGYASFPLAVQLHVSKRSKAMALARLYYKTS-NH2 (SEQ ID NO: 2020)		3
JBT0680	Ac-SGYASFPLAVQLHVSKRSKEAALARLYYKTS-NH2 (SEQ ID NO: 2138)		3
JBT0682	Ac-SGYASFPLAVQLHVSKRSKEMAAARLYYKTS-NH2 (SEQ ID NO: 2139)		0
JBT0684	Ac-SGYASFPLAVQLHVSKRSKEMALAALYYKTS-NH2 (SEQ ID NO: 2011)		2
JBT0685	Ac-SGYASFPLAVQLHVSKRSKEMALARAYYKTS-NH2 (SEQ ID NO: 2021)		1
JBT0686	Ac-SGYASFPLAVQLHVSKRSKEMALARLAYKTS-NH2 (SEQ ID NO: 2140)		1
JBT0687	Ac-SGYASFPLAVQLHVSKRSKEMALARLYAKTS-NH2 (SEQ ID NO: 2141)		0
JBT0688	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYATS-NH2 (SEQ ID NO: 2012)		2
JBT0689	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYKAS-NH2 (SEQ ID NO: 2013)		2
JBT0690	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYKTA-NH2 (SEQ ID NO: 2014)		3
JBT1579	Ac-GYASFPLAVQLHVAKRSKEMA-NH2 (SEQ ID NO: 2015)		5
JBT1580	Ac-GYASFALSVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2320)		6
JBT1581	Ac-GYASFALAVQLHVAKRSKEMA-NH2 (SEQ ID NO: 2321)		13
JBT1582	Ac-GYASFPLAVQLHVMKRSKEMA-NH2 (SEQ ID NO: 2022)		7
JBT1583	Ac-GYASFALAVQLHVMKRSKEMA-NH2 (SEQ ID NO: 2322)		12
JBT1602	Ac-LAVQLHVSKRSKEMALARL-NH2 (SEQ ID NO: 2326)		0

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FIGURE 28

JBT0047 class		Thrombin generation assay				
Peptide	**Sequence**	**FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]**	**FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]**	**EC_{50} (μM), FVIII-inhibited plasma**	**% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide**	**% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide**
JBT0047	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-NH2 (SEQ ID NO: 253)	156	87	0.89	24.6	20.0
JBT0131	Biotin-Ttds-AFQSKKNVFVFGYFERLRAK-NH2 (SEQ ID NO: 964)	106			4.5	
JBT0132	Biotin-Ttds-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 965)	123			5.3	
JBT0133	Biotin-Ttds-QSKKNVFVFGYFERLRAKLT-NH2 (SEQ ID NO: 966)	28			6.7	
JBT0134	Biotin-Ttds-QSKKNVFVFGYFERLRAK-NH2 (SEQ ID NO: 967)	11				
JBT0155	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 8)	49				
JBT0158	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKKK-NH2 (SEQ ID NO: 9)	30				
JBT0162	Ac-KKGRLQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 10)	54				
JBT0163	Ac-KKQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 11)	69				
JBT0164	Ac-KKFQSKKNVFVFGYFERLRAKLTSKK-NH2 (SEQ ID NO: 12)	39				
JBT0169	Ac-KKAFQSKKNVFVFGYFERLRAKKK-NH2 (SEQ ID NO: 254)	21				
JBT0170	Ac-KKFQSKKNVFVFGYFERLRAKLKK-NH2 (SEQ ID NO: 13)	68				
JBT0171	Ac-KKQSKKNVFVFGYFERLRAKLTKK -NH2 (SEQ ID NO: 255)	33				
JBT0174	Ac-KKAFQSKKNVFVFGYFERLRAKLKK-NH2 (SEQ ID NO: 14)	49				

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FIGURE 28

JBT0047 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT0175	Ac-KKAFQSKKNVFVFGYFERLRAKLTKK-NH2 (SEQ ID NO: 182)	49				
JBT0293	Ac-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 256)	106	34	1.60	7.1	
JBT0294	Ac-YQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 257)	122			8.2	
JBT0296	Ac-FQNKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 407)	79				
JBT0297	Ac-FQSKNNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 183)	147			11.3	
JBT0298	Ac-FQSKQNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 747)	109				
JBT0299	Ac-FQSKKNVFAFGYFERLRAKL-NH2 (SEQ ID NO: 408)	63				
JBT0300	Ac-FQSKKNVFSFGYFERLRAKL-NH2 (SEQ ID NO: 409)	40				
JBT0301	Ac-FQSKKNVFTFGYFERLRAKL-NH2 (SEQ ID NO: 410)	49				
JBT0302	Ac-FQSKKNVFVAGYFERLRAKL-NH2 (SEQ ID NO: 258)	220			18.5	
JBT0303	Ac-FQSKKNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 184)	220	126	0.85	21.8	
JBT0304	Ac-FQSKKNVFVLGYFERLRAKL-NH2 (SEQ ID NO: 259)	138				
JBT0305	Ac-FQSKKNVFVQGYFERLRAKL-NH2 (SEQ ID NO: 260)	207	113		20.5	
JBT0306	Ac-FQSKKNVFVSGYFERLRAKL-NH2 (SEQ ID NO: 185)	247	113		17.4	
JBT0307	Ac-FQSKKNVFVYGYFERLRAKL-NH2 (SEQ ID NO: 261)	182	81		16.8	
JBT0308	Ac-FQSKKNVFVFGYKERLRAKL-NH2 (SEQ ID NO: 411)	133				
JBT0309	Ac-FQSKKNVFVFGYYERLRAKL-NH2 (SEQ ID NO: 412)	123				
JBT0310	Ac-FQSKKNVFVFGYFDRLRAKL-NH2 (SEQ ID NO: 262)	163	50			

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FIGURE 28

JBT0047 class		Thrombin generation assay				
Peptide	**Sequence**	**FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]**	**FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]**	**EC_{50} (μM), FVIII-inhibited plasma**	**% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide**	**% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide**
JBT0311	Ac-FQSKKNVFVFGYFERLRAKN-NH2 (SEQ ID NO: 748)	27				
JBT0335	Ac-FQSKKNVFVFGYFERLRAK-NH2 (SEQ ID NO: 749)	28				
JBT0336	Ac-FQSKNNVFVAGYFDRLRAKL-OH (SEQ ID NO: 263)	186	62			
JBT0337	Ac-FQSKNNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 16)	219	144			
JBT0338	Ac-FQSKNNVFVQGYFDRLRAKL-NH2 (SEQ ID NO: 17)	197	133			
JBT0339	Ac-FQSKNNVFVSGYFDRLRAKL-NH2 (SEQ ID NO: 18)	230	136			
JBT0340	Ac-FQSKNNVFVYGYFDRLRAKL-NH2 (SEQ ID NO: 186)	197	122			
JBT0341	Ac-FQSKNNVFVFGYFDRLRAKL-NH2 (SEQ ID NO: 187)	193	100			
JBT0342	Ac-FQSKNNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 19)	244	178		29.7	28.1
JBT0343	Ac-FQSKKNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 188)	239	126			
JBT0374	Ac-FQSKDNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 20)	216	128			
JBT0375	Ac-FQSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 190)	180	140	0.47	22.9	22.9
JBT0376	Ac-FQSKKNAFVFGYFERLRAKL-NH2 (SEQ ID NO: 414)	101	22			
JBT0377	Ac-FQSKKNQFVFGYFERLRAKL-NH2 (SEQ ID NO: 264)	180	75			
JBT0378	Ac-FQSKKNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 21)	245	148		30.7	21.2
JBT0379	Ac-FQSKKNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 22)	226	123			
JBT0380	Ac-FQSKKNVFVVGYFERLRAKL-NH2 (SEQ ID NO: 191)	199	102			
JBT0381	Ac-FQSPKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 265)	173	76			

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FIGURE 28

JBT0047 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT0386	Ac-FQSKKNPFVFGYFERLRAKL-NH2 (SEQ ID NO: 415)	85				
JBT0388	Ac-FQSKKNVHVFGYFERLRAKL-NH2 (SEQ ID NO: 192)	185	82			
JBT0389	Ac-FQSKKNVVVFGYFERLRAKL-NH2 (SEQ ID NO: 673)	34				
JBT0390	Ac-FQSKKNVFQFGYFERLRAKL-NH2 (SEQ ID NO: 674)	28				
JBT0391	Ac-FQSKKNVFVGGYFERLRAKL-NH2 (SEQ ID NO: 266)	178	62			
JBT0392	Ac-FQSKKNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 23)	228	112			
JBT0393	Ac-FQSKKNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 193)	268	161		38.8	30.1
JBT0394	Ac-FQSKKNVFVMGYFERLRAKL-NH2 (SEQ ID NO: 24)	115	43			
JBT0395	Ac-FQSKKNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 25)	235	107			
JBT0396	Ac-FQSKKNVFVPGYFERLRAKL-NH2 (SEQ ID NO: 194)	196	85			
JBT0397	Ac-FQSKKNVFVRGYFERLRAKL-NH2 (SEQ ID NO: 195)	240	128			
JBT0402	Biotin-Ttds-FQSKNNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 973)	185	159			
JBT0403	Biotin-Ttds-FQSKKNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 972)	253	188			
JBT0404	Biotin-Ttds-FQSKKNVFVFGYFDRLRAKL-NH2 (SEQ ID NO: 974)	115	52			
JBT0405	Biotin-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 975)	142	152			
JBT0406	Ac-Ttds-FQSKKNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 268)	114	63			
JBT0471	Ac-FQSKGNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 26)	193	164		42.5	34.7
JBT0472	Ac-FQSKGNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 27)	197	159			

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FIGURE 28

JBT0047 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT0473	Ac-FQSKGNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 28)	188	145			
JBT0474	Ac-FQSKGNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 29)	200	191		50.5	45.2
JBT0475	Ac-FQSKGNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 30)	202	165			
JBT0476	Ac-FQSKGNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 31)	202	164		43.2	41.5
JBT0477	Ac-FQSKGNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 32)	231	194	0.26	47.2	41.3
JBT0478	Ac-FQSKDNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 33)	161	104			
JBT0479	Ac-FQSKDNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 34)	161	105			
JBT0480	Ac-FQSKDNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 35)	180	125			
JBT0481	Ac-FQSKDNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 36)	168	124			
JBT0482	Ac-FQSKDNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 37)	183	133			
JBT0483	Ac-FQSKDNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 38)	191	136			
JBT0484	Ac-FQSKDNVFVDGYFDRLRAKL-NH2 (SEQ ID NO: 976)	178	111			
JBT0485	Ac-FQSKNNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 39)	186	150			
JBT0486	Ac-FQSKNNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 40)	171	112			
JBT0487	Ac-FQSKNNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 41)	178	138			
JBT0488	Ac-FQSKNNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 42)	193	153			
JBT0489	Ac-FQSKNNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 43)	185	148			
JBT0490	Ac-FQSKQNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 44)	162	79			

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FIGURE 28

JBT0047 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT0491	Ac-FQSKQNVFVNGYFERLRAKL-NH2 (SEQ ID NO: 197)	169	96			
JBT0492	Ac-FQSKQNVFVHGYFERLRAKL-NH2 (SEQ ID NO: 198)	124	69			
JBT0493	Ac-FQSKQNVFVKGYFERLRAKL-NH2 (SEQ ID NO: 199)	151	92			
JBT0494	Ac-FQSKQNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 45)	163	102			
JBT0495	Ac-FQSKQNVFVEGYFERLRAKL-NH2 (SEQ ID NO: 200)	156	93			
JBT0497	Ac-NmetPhe-QSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 201)	165	106			
JBT0499	Ac-FQS-NmetLys-GNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 202)	192	129			
JBT0500	Ac-FQSK-NmetGly-NVFVFGYFERLRAKL-NH2 (SEQ ID NO: 46)	254	239	0.17		51.1
JBT0501	Ac-FQSKGN-NmetV-FVFGYFERLRAKL-NH2 (SEQ ID NO: 417)	95	21			
JBT0504	Ac-FQSKGNVFV-NmetPhe-GYFERLRAKL-NH2 (SEQ ID NO: 418)	62				
JBT0505	Ac-FQSKGNVFVF-NmetGly-YFERLRAKL-NH2 (SEQ ID NO: 269)	204	90			
JBT0508	Ac-FQSKGNVFVFGYF-NmetGlu-RLRAKL-NH2 (SEQ ID NO: 678)	82				
JBT0514	Ac-FQSKGNVFVFGYFERLRAK-NmetLeu-NH2 (SEQ ID NO: 419)	69				
JBT0515	Ac-bHomoPhe-QSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 420)	86	31			
JBT0517	Ac-FQ-bHomoSer-KGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 421)	34				
JBT0518	Ac-FQS-bHomoLys-GNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 422)	38				
JBT0519	Ac-FQSK-bAla-NVFVFGYFERLRAKL-NH2 (SEQ ID NO: 270)	105	37			
JBT0520	Ac-FQSKG-bGln-VFVFGYFERLRAKL-NH2 (SEQ ID NO: 271)	100	42			

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FIGURE 28

JBT0047 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT0522	Ac-FQSKGNV-bHomoPhe-VFGYFERLRAKL-NH2 (SEQ ID NO: 681)	40				
JBT0524	Ac-FQSKGNVFV-bHomoPhe-GYFERLRAKL-NH2 (SEQ ID NO: 682)	36				
JBT0527	Ac-FQSKGNVFVFGY-bHomoPhe-ERLRAKL-NH2 (SEQ ID NO: 683)	44				
JBT0528	Ac-FQSKGNVFVFGYF-bE-RLRAKL-NH2 (SEQ ID NO: 272)	151	100			
JBT0531	Ac-FQSKGNVFVFGYFERL-bHomoArg-AKL-NH2 (SEQ ID NO: 203)	170	99			
JBT0532	Ac-FQSKGNVFVFGYFERLR-bAla-KL-NH2 (SEQ ID NO: 423)	118	53			
JBT0533	Ac-FQSKGNVFVFGYFERLRA-bHomoK-L-NH2 (SEQ ID NO: 424)	97	29			
JBT0534	Ac-FQSKGNVFVFGYFERLRAK-Btl-NH2 (SEQ ID NO: 204)	156	130			
JBT0535	Ac-FQSKGNVFVFGYFE-Cit-LRAKL-NH2 (SEQ ID NO: 685)	97	28			
JBT0536	Ac-FQSKGNVFVFGYFERL-Cit-AKL-NH2 (SEQ ID NO: 273)	196	133			
JBT0538	Ac-FQSKGNVFVFGYFERL-Nle-AKL-NH2 (SEQ ID NO: 425)	118	59			
JBT0564	Ac-FQSKKNVFVFGYFKRLRAKL-NH2 (SEQ ID NO: 205)	120	61			
JBT0613	Ac-FQSKGNVFVFGYFERLRAKL-OH (SEQ ID NO: 283)	156	55			
JBT0614	NH2-FQSKGNVFVFGYFERLRAKL-OH (SEQ ID NO: 720)	54				
JBT0615	NH2-FQSKGNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 284)	105	33			
JBT0616	NH2-GSFQSKKNVFVDGYFERLRAKL-OH (SEQ ID NO: 285)	162	77			
JBT0616	NH2-GSFQSKKNVFVDGYFERLRAKL-OH (SEQ ID NO: 285)	147	29			
JBT0651	Ac-FQSKGNVHVKGYFERLRAKL-NH2 (SEQ ID NO: 48)	250	216			

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FIGURE 28

JBT0047 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT0652	Ac-FQSKGNVHVDGYFERLRAKL-NH2 (SEQ ID NO: 49)		207			
JBT0653	Ac-FQSPGNVHVKGYFERLRAKL-NH2 (SEQ ID NO: 50)		237			
JBT0654	Ac-FQSPGNVHVDGYFERLRAKL-NH2 (SEQ ID NO: 51)		214			
JBT0655	Ac-FQSKGNIFVFGYFERLRAKL-NH2 (SEQ ID NO: 52)	150	120			
JBT0656	Ac-FQSKGNLFVFGYFERLRAKL-NH2 (SEQ ID NO: 286)	86	42			
JBT0657	Ac-FQSKGNVFIFGYFERLRAKL-NH2 (SEQ ID NO: 287)	92	47			
JBT0663	Ac-FQSKaNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 53)		180			
JBT0668	Ac-FQSKaNVFVTGYFARLRAKL-NH2 (SEQ ID NO: 54)		167			
JBT0681	Ac-FQSK-Nmg-AVFVFGYFARLRAKL-NH2 (SEQ ID NO: 206)		158			
JBT0683	Ac-FQSK-Nmg-AVFVDGYFARLRAKL-NH2 (SEQ ID NO: 55)		228			
JBT0696	Ac-FQSKKAVFVFGYFERLRAKL-NH2 (SEQ ID NO: 288)	64	60			
JBT0697	Ac-FQSKGNVFVDGYFERL-Dap-AKL-NH2 (SEQ ID NO: 56)		188			
JBT0699	Ac-FQSKGNVFVDGYFERL-Orn-AKL-NH2 (SEQ ID NO: 57)		207			
JBT0700	Ac-FQSKGNVFVDGYFERL-Nva-AKL-NH2 (SEQ ID NO: 58)		218			
JBT0704	Ac-FQSKKNVFVFGYFARLRAKL-NH2 (SEQ ID NO: 289)	96	35			
JBT0708	Ac-FQSKaNVFVAGYFERLRAKL-NH2 (SEQ ID NO: 59)		200			
JBT0714	Ac-FQSKaAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 60)		209			
JBT0717	Ac-FQSK-Nmg-NVFVDGYFERLRAKL-NH2 (SEQ ID NO: 61)	271	241	0.11		42.1

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FIGURE 28

JBT0047 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT0720	Ac-FQSK-Nmg-NVFVTGYFARLRAKL-NH2 (SEQ ID NO: 62)		271			
JBT0732	Ac-FQSKGNVFVDGYFERL-Hci-AKL-NH2 (SEQ ID NO: 63)		258			
JBT0733	Ac-FQSKGNVFVDGYFERL-Har-AKL-NH2 (SEQ ID NO: 64)		213			
JBT0739	Ac-FQSKGNVFVDGYFERL-Opa-AKL-NH2 (SEQ ID NO: 65)		193			
JBT0740	Ac-FQSKGNVFVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 66)	236	184	0.29		33.3
JBT0754	Ac-FQSK-Nmg-AVFVTGYFARLRAKL-NH2 (SEQ ID NO: 67)		224			
JBT0759	Ac-FQSKkNVFVFGYFERLRAKL-NH2 (SEQ ID NO: 207)	159	131			
JBT0765	Ac-FQSKGNVFVDGYFERL-Eew-AKL-NH2 (SEQ ID NO: 68)		205			
JBT0775	Ac-FQSKGNVFVTGYFDRLRAKL-NH2 (SEQ ID NO: 69)		157			
JBT0778	Ac-FQSKGNVFVKGYFDRLRAKL-NH2 (SEQ ID NO: 209)		176			
JBT0779	Ac-FQSKGNVFVEGYFDRLRAKL-NH2 (SEQ ID NO: 210)		145			
JBT0780	Ac-FQSK-Nmg-NVFVFGYFARLRAKL-NH2 (SEQ ID NO: 70)		227			
JBT0781	Ac-FQSKGNVFVDGYFKRLRAKL-NH2 (SEQ ID NO: 211)		156			
JBT0789	Ac-FQSK-Nmg-NVFVDGYFERL-Nle-AKL-NH2 (SEQ ID NO: 71)	266	230	0.26		
JBT0806	Ac-FQSK-Nmg-NVFVAGYFARLRAKL-NH2 (SEQ ID NO: 74)		281			
JBT0837	Ac-FQSK-Nmg-NVFVTGYFERL-Nle-AKL-NH2 (SEQ ID NO: 213)		259			
JBT0844	Ac-FQSKaNVFVDGYFERLRAKL-NH2 (SEQ ID NO: 300)		195			
JBT0850	Ac-FQSKaAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 302)		174			

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FIGURE 28

JBT0047 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT0854	Ac-FQSKaAVFVAGYFARL-Nle-AKL-NH2 (SEQ ID NO: 490)		34			
JBT0870	Ac-FQSKaAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 214)		167			
JBT0886	Ac-FQSKaNVFVDGYFARLRAKL-NH2 (SEQ ID NO: 215)		186			
JBT0919	Ac-FQSKaNVFVTGYFERLRAKL-NH2 (SEQ ID NO: 79)		204			
JBT0931	Ac-FQSKGNVFVDGYFERL-Eag-AKL-NH2 (SEQ ID NO: 82)		172			
JBT0946	Ac-FQSKGNVFVDGYFERL-Dab-AKL-NH2 (SEQ ID NO: 85)		177			
JBT0950	Ac-FQSKaNVFVFGYFARLRAKL-NH2 (SEQ ID NO: 217)		115			
JBT0973	Ac-FQSKGNVFVDGYFERL-Cha-AKL-NH2 (SEQ ID NO: 92)		193			
JBT1006	Ac-FQSKGNVFVDGYFERL-Hle-AKL-NH2 (SEQ ID NO: 96)		210			
JBT1035	Ac-FQSK-Nmg-AVFVAGYFARL-Nle-AKL-NH2 (SEQ ID NO: 218)		172			
JBT1037	Ac-FQSKGNVFVDGYFERL-Nle-AKL-NH2 (SEQ ID NO: 104)		192			
JBT1043	Ac-FQSKaNVFVDGYFERL-Nle-AKL-NH2 (SEQ ID NO: 106)		203			
JBT1082	Ac-FQSKaAVFVFGYFARLRAKL-NH2 (SEQ ID NO: 565)		90			
JBT1084	Ac-FQSK-Nmg-AVFVAGYFARLRAKL-NH2 (SEQ ID NO: 109)		192			
JBT1122	Ac-FQSK-Nmg-NVFVDGYFARL-Nle-AKL-NH2 (SEQ ID NO: 113)		353			
JBT1133	Ac-FQSKkAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 115)	233	236			
JBT1134	Ac-FQSKGAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 116)	240	204			
JBT1135	Ac-FQSKDAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 221)	227	146			

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FIGURE 28

JBT0047 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT1136	Ac-FQSKdAVFVDGYFARLRAKL-NH2 (SEQ ID NO: 117)	283	227			
JBT1137	Ac-FQSKkAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 118)	218	231			
JBT1138	Ac-FQSKGAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 222)	185	161			
JBT1139	Ac-FQSKDAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 223)	205	146			
JBT1140	Ac-FQSKdAVFVAGYFARLRAKL-NH2 (SEQ ID NO: 119)	203	124			
JBT1141	Ac-FQSKkAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 120)	223	217	0.12		
JBT1142	Ac-FQSKGAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 224)	299	233			
JBT1143	Ac-FQSKDAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 225)	214	170			
JBT1144	Ac-FQSKdAVFVKGYFARLRAKL-NH2 (SEQ ID NO: 121)	200	164	0.21		
JBT1145	Ac-FQSKkAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 122)	274	243			
JBT1146	Ac-FQSKGAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 226)	231	185			
JBT1147	Ac-FQSKDAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 227)	230	146			
JBT1148	Ac-FQSKdAVFVTGYFARLRAKL-NH2 (SEQ ID NO: 123)	231	185			
JBT1151	Ac-FQSKKNVFVFGYFERLRAKD-NH2 (SEQ ID NO: 937)	27				
JBT1153	Ac-FQSKKNVFVFGYFERLGAKL-NH2 (SEQ ID NO: 705)	125	45			
JBT1155	Ac-FQSK-Nmg-NVFVTGYFERLRAKL-NH2 (SEQ ID NO: 125)		258			
JBT1156	Ac-FQSKaNVFVDGYFARL-Nle-AKL-NH2 (SEQ ID NO: 126)		173			
JBT1157	Ac-FQSKaNVFVAGYFARLRAKL-NH2 (SEQ ID NO: 127)		178			

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FIGURE 28

JBT0047 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT1584	Ac-FQSK-Nmg-NVFVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 164)	330	301	0.16		48.6
JBT1585	Ac-FQSK-Nmg-NVFVTGYFERL-Aib-AKL-NH2 (SEQ ID NO: 165)	300	269	0.13		
JBT1587	Ac-FQSKGNVFVDGYFERL-Aib-AKLC-NH2 (SEQ ID NO: 167)		189	0.32		
JBT1590	Ac-FQSKGNVFV[CGYFERL-Aib-AKLC]-NH2 (SEQ ID NO: 401)	35				
JBT1591	Ac-FQSKGNVFVDGYF[CRL-Aib-AKLC]-NH2 (SEQ ID NO: 168)	148	75			
JBT1592	Ac-[CFQSKGNVFVDGYFERLC]AKL-NH2 (SEQ ID NO: 402)	138	46			
JBT1593	Ac-FQSK[CNVFVDGYFERLC]AKL-NH2 (SEQ ID NO: 670)	110	39			
JBT1594	Ac-FQSKGNVFV[CGYFERLC]AKL-NH2 (SEQ ID NO: 671)	35				
JBT1595	Ac-[CFQSKGNVFVC]GYFERL-Aib-AKL-NH2 (SEQ ID NO: 403)	82	22			
JBT1596	Ac-[CFQSKGC]VFVEGYFERL-Aib-AKL-NH2 (SEQ ID NO: 404)	116	36			
JBT1853	PEG(40kD)-FQSKGNVFVDGYFERL-Aib-AKL-NH2 (SEQ ID NO: 175)		605			
JBT1855	Ac-FQSKGNVFVDGYFERL-Aib-AKLC(PEG40kD)-NH2 (SEQ ID NO: 252)		642			

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FIGURE 29

JBT0120 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT0120	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1047)	269	95	2.78	28.8	8.6
JBT0247	Ac-SGASRYKWFCGMRDMKGTMSCVWVKFRYDTS-NH2 (SEQ ID NO: 1213)	273				
JBT0248	Ac-KKSGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTSKK-NH2 (SEQ ID NO: 1001)	164				
JBT0319	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRY-NH2 (SEQ ID NO: 1075)	383	187		29.3	
JBT0320	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVK-NH2 (SEQ ID NO: 1103)	153	21			
JBT0323	Ac-SRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1076)	355	142		31.4	
JBT0324	Ac-KWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2 (SEQ ID NO: 1155)	91				
JBT0329	Ac-KWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1156)	70			6.6	
JBT0333	Ac-KWFCGMRDMKGTMSCVWVKF-NH2 (SEQ ID NO: 1207)	40				
JBT0333	Ac-KWFCGMRDMKGTMSCVWVKF-NH2 (SEQ ID NO: 1207)	114				
JBT0411	Ac-SGASRYKWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKFRYDTS-NH2 (SEQ ID NO: 1104)	97				
JBT0412	Ac-KWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKF-NH2 (SEQ ID NO: 1209)	46				
JBT0415	Ac-SRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1003)	291	182	0.57	47.2	36.5
JBT0416	Ac-SRYKWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKF-NH2 (SEQ ID NO:	256	95		36.6	22.3

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FIGURE 29

JBT0120 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
	1077)					
JBT0437	Ac-ARYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1048)	362	225			
JBT0438	Ac-SAYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1078)	300	137			
JBT0439	Ac-SRAKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1105)	266	88			
JBT0440	Ac-SRYAWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1106)	175	33			
JBT0441	Ac-SRYKAF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1230)	18	5			
JBT0442	Ac-SRYKWA[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1231)	53	8			
JBT0444	Ac-SRYKWF[CAMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1004)	331	229	0.42	53.4	45.7
JBT0445	Ac-SRYKWF[CGARDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1107)	157	40			
JBT0446	Ac-SRYKWF[CGMADMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1079)	287	140			
JBT0447	Ac-SRYKWF[CGMRAMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1158)	101				
JBT0448	Ac-SRYKWF[CGMRDAKGTMSC]VWVKF-NH2 (SEQ ID NO: 1108)	324	138			
JBT0449	Ac-SRYKWF[CGMRDMAGTMSC]VWVKF-NH2 (SEQ ID NO: 1080)	362	217		54.1	34.2
JBT0450	Ac-SRYKWF[CGMRDMKATMSC]VWVKF-NH2 (SEQ ID NO: 1159)	232	68			
JBT0451	Ac-SRYKWF[CGMRDMKGAMSC]VWVKF-NH2 (SEQ ID NO: 1081)	339	189			
JBT0452	Ac-SRYKWF[CGMRDMKGTASC]VWVKF-NH2 (SEQ ID NO: 1109)	265	119			
JBT0453	Ac-SRYKWF[CGMRDMKGTMAC]VWVKF-NH2 (SEQ ID NO: 1110)	282	111			
JBT0455	Ac-SRYKWF[CGMRDMKGTMSC]AWVKF-NH2 (SEQ ID NO: 1049)	316	203			

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FIGURE 29

JBT0120 class		Thrombin generation assay				
Peptide	**Sequence**	**FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]**	**FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]**	**EC_{50} (μM), FVIII-inhibited plasma**	**% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide**	**% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide**
JBT0456	Ac-SRYKWFCGMRDMKGTMSCVAVKF-NH2 (SEQ ID NO: 1232)	16				
JBT0457	Ac-SRYKWF[CGMRDMKGTMSC]VWAKF-NH2 (SEQ ID NO: 1161)	76				
JBT0458	Ac-SRYKWF[CGMRDMKGTMSC]VWVAF-NH2 (SEQ ID NO: 1082)	292	161			
JBT0459	Ac-SRYKWF[CGMRDMKGTMSC]VWVKA-NH2 (SEQ ID NO: 1083)	266	137			
JBT0460	Ac-RYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1050)	262	138			
JBT0461	Ac-YKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1111)	197	47			
JBT0625	Ac-SRYKWF[CMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1244)	5	3			
JBT0626	Ac-SRYKWF[CGMRDKGTMSC]VWVKF-NH2 (SEQ ID NO: 1164)	81	10			
JBT0627	Ac-SRYKWF[CGMRDMGTMSC]VWVKF-NH2 (SEQ ID NO: 1165)	62	15			
JBT0628	Ac-SRYKWF[CGMRDMKGMSC]VWVKF-NH2 (SEQ ID NO: 1166)	54	16			
JBT0631	Ac-SRYKWF[CGmRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1113)	171	60			
JBT0634	Ac-SRYKWF[CGMRDmKGTMSC]VWVKF-NH2 (SEQ ID NO: 1051)	311	156			
JBT0635	Ac-SRYKWF[CGMRDMkGTMSC]VWVKF-NH2 (SEQ ID NO: 1114)	123				
JBT0641	Ac-SRYKWF[CPMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1247)	21				
JBT0642	Ac-SRYKWF[CGPRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1248)	32				
JBT0643	Ac-SRYKWF[CGMPDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1006)	289	192	0.63		
JBT0644	Ac-SRYKWF[CGMRPMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1172)	31				
JBT0645	Ac-SRYKWF[CGMRDPKGTMSC]VWVKF-NH2 (SEQ ID NO: 1084)		109			

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FIGURE 29

JBT0120 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT0646	Ac-SRYKWF[CGMRDMPGTMSC]VWVKF-NH2 (SEQ ID NO: 1173)	84				
JBT0648	Ac-SRYKWF[CGMRDMKGPMSC]VWVKF-NH2 (SEQ ID NO: 1174)	42				
JBT0649	Ac-SRYKWF[CGMRDMKGTPSC]VWVKF-NH2 (SEQ ID NO: 1175)	64				
JBT1416	Ac-DRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1007)	253	102	1.38		
JBT1417	Ac-SDYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1116)	140	30	4.79		
JBT1418	Ac-SRDKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1176)	67		3.15		
JBT1419	Ac-SRYDWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1117)	120		3.80		
JBT1426	Ac-SRYKWF[CGMRDMDGTMSC]VWVKF-NH2 (SEQ ID NO: 1008)	287	135	0.94		
JBT1436	Ac-FRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1053)	228	72	2.54		
JBT1437	Ac-SFYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1010)	284	157	0.85		
JBT1438	Ac-SRFKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1011)	218	81	1.63		
JBT1445	Ac-SRYKWF[CGMRDFKGTMSC]VWVKF-NH2 (SEQ ID NO: 1012)	213	89	1.75		
JBT1446	Ac-SRYKWF[CGMRDMFGTMSC]VWVKF-NH2 (SEQ ID NO: 1054)	169	64	1.53		
JBT1448	Ac-SRYKWF[CGMRDMKGFMSC]VWVKF-NH2 (SEQ ID NO: 1123)	184	34			
JBT1449	Ac-SRYKWF[CGMRDMKGTFSC]VWVKF-NH2 (SEQ ID NO: 1013)	276	116	1.45		
JBT1455	Ac-GRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1014)	226	66	2.50		
JBT1465	Ac-SRYKWF[CGMRDMGGTMSC]VWVKF-NH2 (SEQ ID NO: 1015)	235	74	2.46		
JBT1474	Ac-KRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1016)	275	136	1.06		

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FIGURE 29

JBT0120 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT1481	Ac-SRYKWF[CGMKDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1017)	303	123	1.48		
JBT1483	Ac-SRYKWF[CGMRDKKGTMSC]VWVKF-NH2 (SEQ ID NO: 1057)	275	88			
JBT1492	Ac-LRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1018)	239	100	1.55		
JBT1493	Ac-SLYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1060)	261	122	1.17		
JBT1502	Ac-SRYKWF[CGMRDLKGTMSC]VWVKF-NH2 (SEQ ID NO: 1019)	307	175	0.77		
JBT1503	Ac-SRYKWF[CGMRDMLGTMSC]VWVKF-NH2 (SEQ ID NO: 1063)	202	105	0.95		
JBT1506	Ac-SRYKWF[CGMRDMKGTLSC]VWVKF-NH2 (SEQ ID NO: 1064)	191	49	2.99		
JBT1512	Ac-SRYKWF[CGMRDMKGTMSC]VWVKL-NH2 (SEQ ID NO: 1020)	294	152	0.89		
JBT1523	Ac-SRYKWF[CGMRDMSGTMSC]VWVKF-NH2 (SEQ ID NO: 1022)	255	115	1.30		
JBT1541	Ac-SRYKWF[CGMRDMKGTMSC]VWVPF-NH2 (SEQ ID NO: 1023)	291	155	0.82		
JBT1543	Ac-SRYKWF[CGMRDMKGTMSC]VWVKF-OH (SEQ ID NO: 1098)	192	39	3.10		
JBT1544	H-SRYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1024)	235	98			
JBT1545	H-SRYKWF[CGMRDMKGTMSC]VWVKF-OH (SEQ ID NO: 1099)	152	21	7.92		
JBT1557	H-RYKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 173)	208	91			
JBT1558	H-YKWF[CGMRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1149)	115	20			
JBT1559	Ac-SRYKWF[CGaRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1150)	131	42			
JBT1562	Ac-SRYKWF[CGkRDMKGTMSC]VWVKF-NH2 (SEQ ID NO: 1193)	54				
JBT1567	Ac-SRYKWF[CGMRDdKGTMSC]VWVKF-NH2 (SEQ ID NO: 1101)	226	76			

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FIGURE 29

JBT0120 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT1570	Ac-SRYKWF[CGMRDlKGTMSC]VWVKF-NH2 (SEQ ID NO: 1025)	250	76			
JBT1574	Ac-SRYKWF[CGMRDMdGTMSC]VWVKF-NH2 (SEQ ID NO: 1153)	173	34			

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FIGURE 30

JBT0122 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT0122	AC-SGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2002)	170	31		160	
JBT0221	Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKTSKK-NH2 (SEQ ID NO: 2003)	206	150			
JBT0224	Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKK-NH2 (SEQ ID NO: 2298)	152	58	1.10	162	
JBT0225	Ac-KKSGYASFPLAVQLHVSKRSKEMALARKK-NH2 (SEQ ID NO: 2128)	55				
JBT0367	Ac-ASFPLAVQLHVSKRSKEMALARL-NH2 (SEQ ID NO: 2317)	28				
JBT0368	Ac-ASFPLAVQLHVSKRSKEMALARLY-NH2 (SEQ ID NO: 2130)	36				
JBT0369	Ac-ASFPLAVQLHVSKRSKEMALARLYY-NH2 (SEQ ID NO: 2017)	68				
JBT0370	Ac-YASFPLAVQLHVSKRSKEMA-NH2 (SEQ ID NO: 2318)	39				
JBT0371	Ac-GYASFPLAVQLHVSKRSKEMA-NH2(SEQ ID NO: 2319)	25				
JBT0660	Ac-AGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2004)	141	27			
JBT0661	Ac-SAYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2005)	215	49			
JBT0662	Ac-SGAASFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2131)	43				

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FIGURE 30

JBT0122 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
JBT0664	Ac-SGYAAFPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2006)	178	26			
JBT0665	Ac-SGYASAPLAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2132)	49				
JBT0666	Ac-SGYASFALAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2001)	99	23			
JBT0667	Ac-SGYASFPAAVQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2018)	95				
JBT0669	Ac-SGYASFPLAAQLHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2133)	20				
JBT0670	Ac-SGYASFPLAVALHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2007)	101				
JBT0671	Ac-SGYASFPLAVQAHVSKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2019)	120				
JBT0673	Ac-SGYASFPLAVQLHASKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2135)	39				
JBT0674	Ac-SGYASFPLAVQLHVAKRSKEMALARLYYKTS-NH2 (SEQ ID NO: 2008)	234	90			
JBT0675	Ac-SGYASFPLAVQLHVSARSKEMALARLYYKTS-NH2 (SEQ ID NO:	20				

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FIGURE 30

JBT0122 class		Thrombin generation assay				
Peptide	Sequence	FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]	FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]	EC_{50} (μM), FVIII-inhibited plasma	% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide	% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide
	2136)					
JBT0677	Ac-SGYASFPLAVQLHVSKRAKEMALARLYYKTS-NH2 (SEQ ID NO: 2009)	178	37			
JBT0678	Ac-SGYASFPLAVQLHVSKRSAEMALARLYYKTS-NH2 (SEQ ID NO: 2010)	153	23			
JBT0679	Ac-SGYASFPLAVQLHVSKRSKAMALARLYYKTS-NH2 (SEQ ID NO: 2020)	20				
JBT0682	Ac-SGYASFPLAVQLHVSKRSKEMAAARLYYKTS-NH2 (SEQ ID NO: 2139)	133	27			
JBT0684	Ac-SGYASFPLAVQLHVSKRSKEMALAALYYKTS-NH2 (SEQ ID NO: 2011)	159	28			
JBT0685	Ac-SGYASFPLAVQLHVSKRSKEMALARAYYKTS-NH2 (SEQ ID NO: 2021)	131	21			
JBT0686	Ac-SGYASFPLAVQLHVSKRSKEMALARLAYKTS-NH2 (SEQ ID NO: 2140)	109				
JBT0687	Ac-SGYASFPLAVQLHVSKRSKEMALARLYAKTS-NH2 (SEQ ID NO: 2141)	124				
JBT0688	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYATS-NH2 (SEQ ID NO: 2012)	148	29			

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FIGURE 30

JBT0122 class		Thrombin generation assay				
Peptide	**Sequence**	**FEIBA-equivalent activity in FVIII-inhibited plasma @ 10μM peptide [mU/mL]**	**FEIBA-equivalent activity in FVIII-inhibited plasma @ 1.1μM peptide [mU/mL]**	**EC_{50} (μM), FVIII-inhibited plasma**	**% FVIII-equivalent activity in FVIII deficient plasma @ 10μM peptide**	**% FVIII-equivalent activity in FVIII deficient plasma @ 1.1μM peptide**
JBT0689	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYKAS-NH2 (SEQ ID NO: 2013)	184	40			
JBT0690	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYKTA-NH2 (SEQ ID NO: 2014)	187	41			
JBT1579	Ac-GYASFPLAVQLHVAKRSKEMA-NH2 (SEQ ID NO: 2015)	66				
JBT1581	Ac-GYASFALAVQLHVAKRSKEMA-NH2 (SEQ ID NO: 2321)	67				
JBT1582	Ac-GYASFPLAVQLHVMKRSKEMA-NH2 (SEQ ID NO: 2022)	73				
JBT1583	Ac-GYASFALAVQLHVMKRSKEMA-NH2 (SEQ ID NO: 2322)	68				

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FIGURE 31

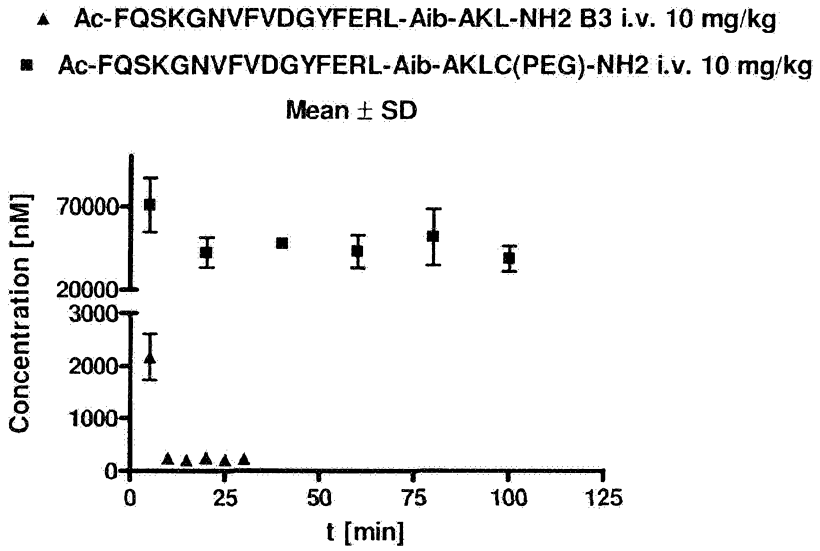
▲ Ac-FQSKGNVFVDGYFERL-Aib-AKL-NH2 B3 i.v. 10 mg/kg
■ Ac-FQSKGNVFVDGYFERL-Aib-AKLC(PEG)-NH2 i.v. 10 mg/kg
Mean ± SD
Concentration [nM]
70000
20000
3000
2000
1000
0
0
25
50
75
100
125
t [min]

FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0047		115	C	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-NH2	253
JBT0155		30	A	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2	8
JBT0156*		>4170		Ac-VVEKLTFVQLSFLNRRRFSQYAGFKGAGKV-NH2	742
JBT0157*		>4170		Ac-RVFLYFSGKAGGLVKLVERQAFQTNVSKFR-NH2	743
JBT0158		41	A	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKKK-NH2	9
JBT0159		>8300		Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLKK-NH2	744
JBT0160		>8300		Ac-KKSGVGRLQVAFQSKKNVFVFGYFKK-NH2	745
JBT0161		>8300		Ac-KKSGVGRLQVAFQSKKNVFVFKK-NH2	746
JBT0162		36	A	Ac-KKGRLQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2	10
JBT0163		18	A	Ac-KKQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2	11
JBT0164		43	A	Ac-KKFQSKKNVFVFGYFERLRAKLTSKK-NH2	12
JBT0165		6134	F	Ac-KKKKNVFVFGYFERLRAKLTSKK-NH2	712
JBT0169		120	C	Ac-KKAFQSKKNVFVFGYFERLRAKKK-NH2	254
JBT0170		39	A	Ac-KKFQSKKNVFVFGYFERLRAKLKK-NH2	13
JBT0171		123	C	Ac-KKQSKKNVFVFGYFERLRAKLTKK -NH2	255
JBT0172		295	D	Ac-KKQSKKNVFVFGYFERLRAKKK-NH2	406
JBT0174		44	A	Ac-KKAFQSKKNVFVFGYFERLRAKLKK-NH2	14
JBT0175		65	B	Ac-KKAFQSKKNVFVFGYFERLRAKLTKK-NH2	182
JBT0291		46	A	Ac-K(FAM)KSGVGRLQVAFQSKKNVFVFGYFERLRAKLTSKK-NH2	15
JBT0293	61	179	C	Ac-FQSKKNVFVFGYFERLRAKL-NH2	256
JBT0294		188	C	Ac-YQSKKNVFVFGYFERLRAKL-NH2	257
JBT0295	37	2777	F	Ac-FSSKKNVFVFGYFERLRAKL-NH2	713
JBT0296		575	D	Ac-FQNKKNVFVFGYFERLRAKL-NH2	407
JBT0297		63	B	Ac-FQSKNNVFVFGYFERLRAKL-NH2	183
JBT0298		>206		Ac-FQSKQNVFVFGYFERLRAKL-NH2	747
JBT0299	25	446	D	Ac-FQSKKNVFAFGYFERLRAKL-NH2	408

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0300		943	D	Ac-FQSKKNVFSFGYFERLRAKL-NH2	409
JBT0301		671	D	Ac-FQSKKNVFTFGYFERLRAKL-NH2	410
JBT0302	55	133	C	Ac-FQSKKNVFVAGYFERLRAKL-NH2	258
JBT0303	66	85	B	Ac-FQSKKNVFVDGYFERLRAKL-NH2	184
JBT0304	54	231	C	Ac-FQSKKNVFVLGYFERLRAKL-NH2	259
JBT0305		154	C	Ac-FQSKKNVFVQGYFERLRAKL-NH2	260
JBT0306	50	89	B	Ac-FQSKKNVFVSGYFERLRAKL-NH2	185
JBT0307		108	C	Ac-FQSKKNVFVYGYFERLRAKL-NH2	261
JBT0308		736	D	Ac-FQSKKNVFVFGYKERLRAKL-NH2	411
JBT0309		565	D	Ac-FQSKKNVFVFGYYERLRAKL-NH2	412
JBT0310	66	146	C	Ac-FQSKKNVFVFGYFDRLRAKL-NH2	262
JBT0311		>50000		Ac-FQSKKNVFVFGYFERLRAKN-NH2	748
JBT0335		>50000		Ac-FQSKKNVFVFGYFERLRAK-NH2	749
JBT0336		232	C	Ac-FQSKNNVFVAGYFDRLRAKL-OH	263
JBT0337		32	A	Ac-FQSKNNVFVDGYFDRLRAKL-NH2	16
JBT0338		32	A	Ac-FQSKNNVFVQGYFDRLRAKL-NH2	17
JBT0339		46	A	Ac-FQSKNNVFVSGYFDRLRAKL-NH2	18
JBT0340		68	B	Ac-FQSKNNVFVYGYFDRLRAKL-NH2	186
JBT0341		75	B	Ac-FQSKNNVFVFGYFDRLRAKL-NH2	187
JBT0342	90	47	A	Ac-FQSKNNVFVDGYFERLRAKL-NH2	19
JBT0343		83	B	Ac-FQSKKNVFVDGYFDRLRAKL-NH2	188
JBT0372		313	D	Fam-Ttds-AFQSKKNVFVFGYFERLRAK-NH2	413
JBT0373		97	B	Fam-AFQSKKNVFVFGYFERLRAK-NH2	189
JBT0374		25	A	Ac-FQSKDNVFVFGYFERLRAKL-NH2	20
JBT0375	101	56	B	Ac-FQSKGNVFVFGYFERLRAKL-NH2	190
JBT0376	32	254	D	Ac-FQSKKNAFVFGYFERLRAKL-NH2	414
JBT0377		146	C	Ac-FQSKKNQFVFGYFERLRAKL-NH2	264

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0378		39	A	Ac-FQSKKNVFVEGYFERLRAKL-NH2	21
JBT0379		27	A	Ac-FQSKKNVFVTGYFERLRAKL-NH2	22
JBT0380		81	B	Ac-FQSKKNVFVVGYFERLRAKL-NH2	191
JBT0381	40	154	C	Ac-FQSPKNVFVFGYFERLRAKL-NH2	265
JBT0385		1765	E	Ac-FQSKKNNFVFGYFERLRAKL-NH2	672
JBT0386	42	526	D	Ac-FQSKKNPFVFGYFERLRAKL-NH2	415
JBT0388		51	B	Ac-FQSKKNVHVFGYFERLRAKL-NH2	192
JBT0389		1165	E	Ac-FQSKKNVVVFGYFERLRAKL-NH2	673
JBT0390		1393	E	Ac-FQSKKNVFQFGYFERLRAKL-NH2	674
JBT0391	22	107	C	Ac-FQSKKNVFVGGYFERLRAKL-NH2	266
JBT0392		32	A	Ac-FQSKKNVFVHGYFERLRAKL-NH2	23
JBT0393	87	56	B	Ac-FQSKKNVFVKGYFERLRAKL-NH2	193
JBT0394		48	A	Ac-FQSKKNVFVMGYFERLRAKL-NH2	24
JBT0395		31	A	Ac-FQSKKNVFVNGYFERLRAKL-NH2	25
JBT0396	28	80	B	Ac-FQSKKNVFVPGYFERLRAKL-NH2	194
JBT0397		69	B	Ac-FQSKKNVFVRGYFERLRAKL-NH2	195
JBT0398		69	B	Ac-FQSKKNVFVFGYFEELRAKL-NH2	196
JBT0399	-24	>185 >5000.000 >5000.000 >5000.000		Ac-FQSKKNVFVFGYFELLRAKL-NH2	750
JBT0400	40	216	C	Ac-FQSKKNVFVFGYFLRLRAKL-NH2	267
JBT0401		359	D	Ac-FQSKKNVFVFGYFERLRAVL-NH2	416
JBT0406		210	C	Ac-Ttds-FQSKKNVFVFGYFERLRAKL-NH2	268
JBT0471	93	29	A	Ac-FQSKGNVFVTGYFERLRAKL-NH2	26
JBT0472	96	38	A	Ac-FQSKGNVFVNGYFERLRAKL-NH2	27
JBT0473	106	39	A	Ac-FQSKGNVFVHGYFERLRAKL-NH2	28

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM)	Affinity Class	Sequence	SEQ ID NO:
JBT0474	100	27	A	Ac-FQSKGNVFVKGYFERLRAKL-NH2	29
JBT0475	106	25	A	Ac-FQSKGNVFVEGYFERLRAKL-NH2	30
JBT0476	56	23	A	Ac-FQSKGNVFVDGYFDRLRAKL-NH2	31
JBT0477	99	32	A	Ac-FQSKGNVFVDGYFERLRAKL-NH2	32
JBT0478		29	A	Ac-FQSKDNVFVTGYFERLRAKL-NH2	33
JBT0479		48	A	Ac-FQSKDNVFVNGYFERLRAKL-NH2	34
JBT0480		35	A	Ac-FQSKDNVFVHGYFERLRAKL-NH2	35
JBT0481		49	A	Ac-FQSKDNVFVKGYFERLRAKL-NH2	36
JBT0482		38	A	Ac-FQSKDNVFVEGYFERLRAKL-NH2	37
JBT0483	74	39	A	Ac-FQSKDNVFVDGYFERLRAKL-NH2	38
JBT0485		23	A	Ac-FQSKNNVFVTGYFERLRAKL-NH2	39
JBT0486		40	A	Ac-FQSKNNVFVNGYFERLRAKL-NH2	40
JBT0487		40	A	Ac-FQSKNNVFVHGYFERLRAKL-NH2	41
JBT0488		29	A	Ac-FQSKNNVFVKGYFERLRAKL-NH2	42
JBT0489		30	A	Ac-FQSKNNVFVEGYFERLRAKL-NH2	43
JBT0490		42	A	Ac-FQSKQNVFVTGYFERLRAKL-NH2	44
JBT0491		59	B	Ac-FQSKQNVFVNGYFERLRAKL-NH2	197
JBT0492		68	B	Ac-FQSKQNVFVHGYFERLRAKL-NH2	198
JBT0493		86	B	Ac-FQSKQNVFVKGYFERLRAKL-NH2	199
JBT0494	74	48	A	Ac-FQSKQNVFVDGYFERLRAKL-NH2	45
JBT0495		52	B	Ac-FQSKQNVFVEGYFERLRAKL-NH2	200
JBT0497		100	B	Ac-Nmf-QSKGNVFVFGYFERLRAKL-NH2	201
JBT0498		1512	E	Ac-FQ-Nms-KGNVFVFGYFERLRAKL-NH2	675
JBT0499		74	B	Ac-FQS-Nmk-GNVFVFGYFERLRAKL-NH2	202
JBT0500		24	A	Ac-FQSK-Nmg-NVFVFGYFERLRAKL-NH2	46
JBT0501		267	D	Ac-FQSKGN-Nmv-FVFGYFERLRAKL-NH2	417
JBT0502		>5000		Ac-FQSKGNV-Nmf-VFGYFERLRAKL-NH2	751

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0503		2038	E	Ac-FQSKGNVF-Nmv-FGYFERLRAKL-NH2	676
JBT0504		861	D	Ac-FQSKGNVFV-Nmf-GYFERLRAKL-NH2	418
JBT0505		118	C	Ac-FQSKGNVFVF-Nmg-YFERLRAKL-NH2	269
JBT0506		3106	F	Ac-FQSKGNVFVFG-Nmy-FERLRAKL-NH2	714
JBT0507		2091	E	Ac-FQSKGNVFVFGY-Nmf-ERLRAKL-NH2	677
JBT0508		1192	E	Ac-FQSKGNVFVFGYF-Nme-RLRAKL-NH2	678
JBT0509		>5000		Ac-FQSKGNVFVFGYFE-Nmr-LRAKL-NH2	752
JBT0510		>5000		Ac-FQSKGNVFVFGYFER-Nml-RAKL-NH2	753
JBT0511		>5000		Ac-FQSKGNVFVFGYFERL-Nmr-AKL-NH2	754
JBT0512		2218	E	Ac-FQSKGNVFVFGYFERLR-Nma-KL-NH2	679
JBT0513		>5000		Ac-FQSKGNVFVFGYFERLRA-Nmk-L-NH2	755
JBT0514		891	D	Ac-FQSKGNVFVFGYFERLRAK-Nml-NH2	419
JBT0515		366	D	Ac-Bhf-QSKGNVFVFGYFERLRAKL-NH2	420
JBT0516		1191	E	Ac-F-Bhq-SKGNVFVFGYFERLRAKL-NH2	680
JBT0517		781	D	Ac-FQ-Bhs-KGNVFVFGYFERLRAKL-NH2	421
JBT0518		767	D	Ac-FQS-Bhk-GNVFVFGYFERLRAKL-NH2	422
JBT0519		207	C	Ac-FQSK-Bal-NVFVFGYFERLRAKL-NH2	270
JBT0520		188	C	Ac-FQSKG-Btq-VFVFGYFERLRAKL-NH2	271
JBT0521		>5000		Ac-FQSKGN-Btl-FVFGYFERLRAKL-NH2	756
JBT0522		1424	E	Ac-FQSKGNV-Bhf-VFGYFERLRAKL-NH2	681
JBT0523		>5000		Ac-FQSKGNVF-Btl-FGYFERLRAKL-NH2	757
JBT0524		1573	E	Ac-FQSKGNVFV-Bhf-GYFERLRAKL-NH2	682
JBT0525		>5000		Ac-FQSKGNVFVF-Bal-YFERLRAKL-NH2	758
JBT0526		>5000		Ac-FQSKGNVFVFG-Bhy-FERLRAKL-NH2	759
JBT0527		1348	E	Ac-FQSKGNVFVFGY-Bhf-ERLRAKL-NH2	683
JBT0528		107	C	Ac-FQSKGNVFVFGYF-Bte-RLRAKL-NH2	272
JBT0529		>5000		Ac-FQSKGNVFVFGYFE-Bhr-LRAKL-NH2	760

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0530		1684	E	Ac-FQSKGNVFVFGYFER-Bhl-RAKL-NH2	684
JBT0531		93	B	Ac-FQSKGNVFVFGYFERL-Bhr-AKL-NH2	203
JBT0532		334	D	Ac-FQSKGNVFVFGYFERLR-Bal-KL-NH2	423
JBT0533		756	D	Ac-FQSKGNVFVFGYFERLRA-Bhk-L-NH2	424
JBT0534		84	B	Ac-FQSKGNVFVFGYFERLRAK-Bhl-NH2	204
JBT0535		1779	E	Ac-FQSKGNVFVFGYFE-Cit-LRAKL-NH2	685
JBT0536		229	C	Ac-FQSKGNVFVFGYFERL-Cit-AKL-NH2	273
JBT0537		>5000		Ac-FQSKGNVFVFGYFE-Nle-LRAKL-NH2	761
JBT0538		534	D	Ac-FQSKGNVFVFGYFERL-Nle-AKL-NH2	425
JBT0539	-1	542	D	Ac-GQSKKNVFVFGYFERLRAKL-NH2	426
JBT0540	3	479	D	Ac-FGSKKNVFVFGYFERLRAKL-NH2	427
JBT0541	14	292	D	Ac-FQGKKNVFVFGYFERLRAKL-NH2	428
JBT0542	14	129	C	Ac-FQSGKNVFVFGYFERLRAKL-NH2	274
JBT0543	67	184	C	Ac-FQSKKGVFVFGYFERLRAKL-NH2	275
JBT0544	-7	746	D	Ac-FQSKKNGFVFGYFERLRAKL-NH2	429
JBT0545	-19	1780	E	Ac-FQSKKNVGVFGYFERLRAKL-NH2	686
JBT0546	-15	>5000		Ac-FQSKKNVFGFGYFERLRAKL-NH2	762
JBT0547	-18	>5000		Ac-FQSKKNVFVFGGFERLRAKL-NH2	763
JBT0548	4	885	D	Ac-FQSKKNVFVFGYGERLRAKL-NH2	430
JBT0549	35	572	D	Ac-FQSKKNVFVFGYFGRLRAKL-NH2	431
JBT0550	-14	>5000		Ac-FQSKKNVFVFGYFEGLRAKL-NH2	764
JBT0551	-22	1640	E	Ac-FQSKKNVFVFGYFERGRAKL-NH2	687
JBT0552	10	591	D	Ac-FQSKKNVFVFGYFERLRGKL-NH2	432
JBT0553	-19	1101	E	Ac-FQSKKNVFVFGYFERLRAGL-NH2	688
JBT0554	-5	314	D	Ac-FQSKKNVFVFGYFERLRAKG-NH2	433
JBT0555	8	350	D	Ac-KQSKKNVFVFGYFERLRAKL-NH2	434
JBT0556	2	336	D	Ac-FKSKKNVFVFGYFERLRAKL-NH2	435

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0557	27	245	C	Ac-FQKKKNVFVFGYFERLRAKL-NH2	276
JBT0558	44	125	C	Ac-FQSKKKVFVFGYFERLRAKL-NH2	277
JBT0559	23	203	C	Ac-FQSKKNKFVFGYFERLRAKL-NH2	278
JBT0560	2	688	D	Ac-FQSKKNVKVFGYFERLRAKL-NH2	436
JBT0561	7	653	D	Ac-FQSKKNVFKFGYFERLRAKL-NH2	437
JBT0562	10	413	D	Ac-FQSKKNVFVFKYFERLRAKL-NH2	438
JBT0563	-6	1151	E	Ac-FQSKKNVFVFGKFERLRAKL-NH2	689
JBT0564	69	95	B	Ac-FQSKKNVFVFGYFKRLRAKL-NH2	205
JBT0565	-5	383	D	Ac-FQSKKNVFVFGYFEKLRAKL-NH2	439
JBT0566	-7	3310	F	Ac-FQSKKNVFVFGYFERKRAKL-NH2	715
JBT0567	51	225	C	Ac-FQSKKNVFVFGYFERLKAKL-NH2	279
JBT0568	49	147	C	Ac-FQSKKNVFVFGYFERLRKKL-NH2	280
JBT0569	13			Ac-FQSKKNVFVFGYFERLRAKK-NH2	765
JBT0570	4	>5000		Ac-FFSKKNVFVFGYFERLRAKL-NH2	766
JBT0571	4	>5000		Ac-FQFKKNVFVFGYFERLRAKL-NH2	767
JBT0572	12	>5000		Ac-FQSFKNVFVFGYFERLRAKL-NH2	768
JBT0573	5	1370	E	Ac-FQSKFNVFVFGYFERLRAKL-NH2	690
JBT0574	-8	>5000		Ac-FQSKKFVFVFGYFERLRAKL-NH2	769
JBT0575	3	578	D	Ac-FQSKKNFFVFGYFERLRAKL-NH2	440
JBT0576	12	>1666		Ac-FQSKKNVFVFFYFERLRAKL-NH2	770
JBT0577	14	>5000		Ac-FQSKKNVFVFGFFERLRAKL-NH2	771
JBT0578	58	292	D	Ac-FQSKKNVFVFGYFFRLRAKL-NH2	441
JBT0579	4	>5000		Ac-FQSKKNVFVFGYFEFLRAKL-NH2	772
JBT0580	12	>5000		Ac-FQSKKNVFVFGYFERFRAKL-NH2	773
JBT0581	17	>5000		Ac-FQSKKNVFVFGYFERLFAKL-NH2	774
JBT0582	45	>5000	E	Ac-FQSKKNVFVFGYFERLRFKL-NH2	691
JBT0583	13	>5000		Ac-FQSKKNVFVFGYFERLRAFL-NH2	775

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0584	50	401	D	Ac-FQSKKNVFVFGYFERLRAKF-NH2	442
JBT0585	10	609	D	Ac-DQSKKNVFVFGYFERLRAKL-NH2	443
JBT0586	5	>1667		Ac-FDSKKNVFVFGYFERLRAKL-NH2	776
JBT0587	50	172	C	Ac-FQDKKNVFVFGYFERLRAKL-NH2	281
JBT0588	5	>5000		Ac-FQSDKNVFVFGYFERLRAKL-NH2	777
JBT0589	58	241	C	Ac-FQSKKDVFVFGYFERLRAKL-NH2	282
JBT0590	2	>5000		Ac-FQSKKNDFVFGYFERLRAKL-NH2	778
JBT0591	1	>5000		Ac-FQSKKNVDVFGYFERLRAKL-NH2	779
JBT0592	-1	>5000		Ac-FQSKKNVFDFGYFERLRAKL-NH2	780
JBT0593	6	2982	F	Ac-FQSKKNVFVFDYFERLRAKL-NH2	716
JBT0594	-1	>5000		Ac-FQSKKNVFVFGDFERLRAKL-NH2	781
JBT0595	10	3557	F	Ac-FQSKKNVFVFGYDERLRAKL-NH2	717
JBT0596	1	549	D	Ac-FQSKKNVFVFGYFEDLRAKL-NH2	444
JBT0597	-5	>5000		Ac-FQSKKNVFVFGYFERDRAKL-NH2	782
JBT0598	-2	>5000		Ac-FQSKKNVFVFGYFERLDAKL-NH2	783
JBT0599	30	301	D	Ac-FQSKKNVFVFGYFERLRDKL-NH2	445
JBT0600	-12	3232	F	Ac-FQSKKNVFVFGYFERLRADL-NH2	718
JBT0601	-1	1531	E	Ac-LQSKKNVFVFGYFERLRAKL-NH2	692
JBT0602	8	>1000 >1000 >5000.000		Ac-FLSKKNVFVFGYFERLRAKL-NH2	784
JBT0603	4	>5000		Ac-FQLKKNVFVFGYFERLRAKL-NH2	785
JBT0604	11	>1668		Ac-FQSLKNVFVFGYFERLRAKL-NH2	786
JBT0605	18	>1669		Ac-FQSKLNVFVFGYFERLRAKL-NH2	787
JBT0606	1	>5000		Ac-FQSKKLVFVFGYFERLRAKL-NH2	788
JBT0607	19	718	D	Ac-FQSKKNLFVFGYFERLRAKL-NH2	446
JBT0608	4	2673	F	Ac-FQSKKNVLVFGYFERLRAKL-NH2	719

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0609	0	>5000		Ac-FQSKKNVFLFGYFERLRAKL-NH2	789
JBT0610	-4	>5000		Ac-FQSKKNVFVFLYFERLRAKL-NH2	790
JBT0611	-5	>5000		Ac-FQSKKNVFVFGLFERLRAKL-NH2	791
JBT0612	9	1378	E	Ac-FQSKKNVFVFGYLERLRAKL-NH2	693
JBT0613		237	C	Ac-FQSKGNVFVFGYFERLRAKL-OH	283
JBT0614		2575	F	H-FQSKGNVFVFGYFERLRAKL-OH	720
JBT0615		249	C	H-FQSKGNVFVFGYFERLRAKL-NH2	284
JBT0616		124	C	H-GSFQSKKNVFVDGYFERLRAKL-OH	285
JBT0636		24	A	Ac-FQSK-Nmg-NVFVDGYFARLRAKL-NH2	47
JBT0651		18	A	Ac-FQSKGNVHVKGYFERLRAKL-NH2	48
JBT0652	103	36	A	Ac-FQSKGNVHVDGYFERLRAKL-NH2	49
JBT0653		23	A	Ac-FQSPGNVHVKGYFERLRAKL-NH2	50
JBT0654		16	A	Ac-FQSPGNVHVDGYFERLRAKL-NH2	51
JBT0655		38	A	Ac-FQSKGNIFVFGYFERLRAKL-NH2	52
JBT0656		155	C	Ac-FQSKGNLFVFGYFERLRAKL-NH2	286
JBT0657		142	C	Ac-FQSKGNVFIFGYFERLRAKL-NH2	287
JBT0658		1062	E	Ac-FQSKGNVFLFGYFERLRAKL-NH2	694
JBT0663		45	A	Ac-FQSKaNVFVFGYFERLRAKL-NH2	53
JBT0668		49	A	Ac-FQSKaNVFVTGYFARLRAKL-NH2	54
JBT0681		68	B	Ac-FQSK-Nmg-AVFVFGYFARLRAKL-NH2	206
JBT0683		25	A	Ac-FQSK-Nmg-AVFVDGYFARLRAKL-NH2	55
JBT0691	32		F	Ac-AQSKKNVFVFGYFERLRAKL-NH2	721
JBT0692	49		E	Ac-FASKKNVFVFGYFERLRAKL-NH2	695
JBT0693	60		D	Ac-FQAKKNVFVFGYFERLRAKL-NH2	447
JBT0694	58		D	Ac-FQSAKNVFVFGYFERLRAKL-NH2	448
JBT0695	52		D	Ac-FQSKANVFVFGYFERLRAKL-NH2	449
JBT0696	94	101	C	Ac-FQSKKAVFVFGYFERLRAKL-NH2	288

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0697		47	A	Ac-FQSKGNVFVDGYFERL-Dap-AKL-NH2	56
JBT0698	-1			Ac-FQSKKNVAVFGYFERLRAKL-NH2	792
JBT0699		37	A	Ac-FQSKGNVFVDGYFERL-Orn-AKL-NH2	57
JBT0700		26	A	Ac-FQSKGNVFVDGYFERL-Nva-AKL-NH2	58
JBT0701	29			Ac-FQSKKNVFVFAYFERLRAKL-NH2	793
JBT0702	10			Ac-FQSKKNVFVFGAFERLRAKL-NH2	794
JBT0703	22			Ac-FQSKKNVFVFGYAERLRAKL-NH2	795
JBT0704	96	116	C	Ac-FQSKKNVFVFGYFARLRAKL-NH2	289
JBT0705	22			Ac-FQSKKNVFVFGYFEALRAKL-NH2	796
JBT0706	10			Ac-FQSKKNVFVFGYFERARAKL-NH2	797
JBT0707	50	502	D	Ac-FQSKKNVFVFGYFERLAAKL-NH2	450
JBT0708		30	A	Ac-FQSKaNVFVAGYFERLRAKL-NH2	59
JBT0709	6			Ac-FQSKKNVFVFGYFERLRAAL-NH2	798
JBT0710	-2			Ac-FQSKKNVFVFGYFERLRAKA-NH2	799
JBT0711	2			Ac-PQSKKNVFVFGYFERLRAKL-NH2	800
JBT0712	19			Ac-FPSKKNVFVFGYFERLRAKL-NH2	801
JBT0713	5	679	D	Ac-FQPKKNVFVFGYFERLRAKL-NH2	451
JBT0714		23	A	Ac-FQSKaAVFVDGYFARLRAKL-NH2	60
JBT0715	40		F	Ac-FQSKPNVFVFGYFERLRAKL-NH2	722
JBT0716	42		E	Ac-FQSKKPVFVFGYFERLRAKL-NH2	696
JBT0717		24	A	Ac-FQSK-Nmg-NVFVDGYFERLRAKL-NH2	61
JBT0718	14			Ac-FQSKKNVPVFGYFERLRAKL-NH2	802
JBT0719	31		F	Ac-FQSKKNVFPFGYFERLRAKL-NH2	723
JBT0720		27	A	Ac-FQSK-Nmg-NVFVTGYFARLRAKL-NH2	62
JBT0721	29			Ac-FQSKKNVFVFPYFERLRAKL-NH2	803
JBT0722	12			Ac-FQSKKNVFVFGPFERLRAKL-NH2	804
JBT0723	3			Ac-FQSKKNVFVFGYPERLRAKL-NH2	805

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0724	50		D	Ac-FQSKKNVFVFGYFPRLRAKL-NH2	452
JBT0725	21			Ac-FQSKKNVFVFGYFEPLRAKL-NH2	806
JBT0726	2		D	Ac-FQSKKNVFVFGYFERPRAKL-NH2	453
JBT0727	-1			Ac-FQSKKNVFVFGYFERLPAKL-NH2	807
JBT0728	12			Ac-FQSKKNVFVFGYFERLRPKL-NH2	808
JBT0729	4			Ac-FQSKKNVFVFGYFERLRAPL-NH2	809
JBT0730	3		D	Ac-FQSKKNVFVFGYFERLRAKP-NH2	454
JBT0731	21			Ac-SQSKKNVFVFGYFERLRAKL-NH2	810
JBT0732		20	A	Ac-FQSKGNVFVDGYFERL-Hci-AKL-NH2	63
JBT0733		26	A	Ac-FQSKGNVFVDGYFERL-Har-AKL-NH2	64
JBT0734	26		D	Ac-FQSSKNVFVFGYFERLRAKL-NH2	455
JBT0735	19			Ac-FQSKSNVFVFGYFERLRAKL-NH2	811
JBT0736	68		D	Ac-FQSKKSVFVFGYFERLRAKL-NH2	456
JBT0737	34		F	Ac-FQSKKNSFVFGYFERLRAKL-NH2	724
JBT0738	20			Ac-FQSKKNVSVFGYFERLRAKL-NH2	812
JBT0739		31	A	Ac-FQSKGNVFVDGYFERL-Opa-AKL-NH2	65
JBT0740	113	18	A	Ac-FQSKGNVFVDGYFERL-Aib-AKL-NH2	66
JBT0741	44		E	Ac-FQSKKNVFVFSYFERLRAKL-NH2	697
JBT0742	12			Ac-FQSKKNVFVFGSFERLRAKL-NH2	813
JBT0743	5			Ac-FQSKKNVFVFGYSERLRAKL-NH2	814
JBT0744	66		D	Ac-FQSKKNVFVFGYFSRLRAKL-NH2	457
JBT0745	9			Ac-FQSKKNVFVFGYFESLRAKL-NH2	815
JBT0746	-1			Ac-FQSKKNVFVFGYFERSRAKL-NH2	816
JBT0747	19			Ac-FQSKKNVFVFGYFERLSAKL-NH2	817
JBT0748	43		D	Ac-FQSKKNVFVFGYFERLRSKL-NH2	458
JBT0749	19		D	Ac-FQSKKNVFVFGYFERLRASL-NH2	459
JBT0750	10			Ac-FQSKKNVFVFGYFERLRAKS-NH2	818

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0751	33		F	Ac-FQSKKNVFVFGYFERLLAKL-NH2	725
JBT0752	46		E	Ac-FQSKKNVFVFGYFERLRLKL-NH2	698
JBT0753	26		D	Ac-FQSKKNVFVFGYFERLRALL-NH2	460
JBT0754		33	A	Ac-FQSK-Nmg-AVFVTGYFARLRAKL-NH2	67
JBT0755	34	>5000	F	Ac-fQSKKNVFVFGYFERLRAKL-NH2	726
JBT0756	35	>5000	F	Ac-FqSKKNVFVFGYFERLRAKL-NH2	727
JBT0757		2212	E	Ac-FQsKKNVFVFGYFERLRAKL-NH2	699
JBT0758	52	101	C	Ac-FQSkKNVFVFGYFERLRAKL-NH2	290
JBT0759	78	52	B	Ac-FQSKkNVFVFGYFERLRAKL-NH2	207
JBT0760	37		F	Ac-FQSKKnVFVFGYFERLRAKL-NH2	728
JBT0761	35		F	Ac-FQSKKNvFVFGYFERLRAKL-NH2	729
JBT0762	37		F	Ac-FQSKKNVfVFGYFERLRAKL-NH2	730
JBT0763	58	129	C	Ac-FQSKKNVFfFGYFERLRAKL-NH2	291
JBT0764	53	296	D	Ac-FQSKKNVFVfGYFERLRAKL-NH2	461
JBT0765		26	A	Ac-FQSKGNVFVDGYFERL-Eew-AKL-NH2	68
JBT0766	32		D	Ac-FQSKKNVFVFGyFERLRAKL-NH2	462
JBT0767	40		E	Ac-FQSKKNVFVFGYfERLRAKL-NH2	700
JBT0768	27		D	Ac-FQSKKNVFVFGYFeRLRAKL-NH2	463
JBT0769	22		D	Ac-FQSKKNVFVFGYFErLRAKL-NH2	464
JBT0770	20			Ac-FQSKKNVFVFGYFERlRAKL-NH2	819
JBT0771	29	423	D	Ac-FQSKKNVFVFGYFERLrAKL-NH2	465
JBT0772	21			Ac-FQSKKNVFVFGYFERLRaKL-NH2	820
JBT0773	21			Ac-FQSKKNVFVFGYFERLRAkL-NH2	821
JBT0774	26			Ac-FQSKKNVFVFGYFERLRAKl-NH2	822
JBT0775	73	22	A	Ac-FQSKGNVFVTGYFDRLRAKL-NH2	69
JBT0776	51	82	B	Ac-FQSKGNVFVNGYFDRLRAKL-NH2	208
JBT0777	49	101	C	Ac-FQSKGNVFVHGYFDRLRAKL-NH2	292

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0778	68	66	B	Ac-FQSKGNVFVKGYFDRLRAKL-NH2	209
JBT0779	51	71	B	Ac-FQSKGNVFVEGYFDRLRAKL-NH2	210
JBT0780		44	A	Ac-FQSK-Nmg-NVFVFGYFARLRAKL-NH2	70
JBT0781	82	63	B	Ac-FQSKGNVFVDGYFKRLRAKL-NH2	211
JBT0782	44	>556 >555.556	D	Ac-fqskknvfvfgyferlrakl-NH2	466
JBT0783	39	>556 >1666.670	D	Ac-lkarlrefygfvfvnkksqf-NH2	467
JBT0784		>5000		Ac-FQSKKNVFVFGYFE-(omega-methyl-R)-L-(omega-methyl-R)-AKL-NH2	823
JBT0785		163	C	Ac-FQSKKNVFVFGYFERL-(omega-methyl-R)-AKL-NH2	293
JBT0786		616	D	Ac-FQSKKNVFVFGYFE-(omega-methyl-R)-LRAKL-NH2	468
JBT0789		20	A	Ac-FQSK-Nmg-NVFVDGYFERL-Nle-AKL-NH2	71
JBT0791	42		D	Ac-AQSKGNVFVDGYFERLRAKL-NH2	469
JBT0792	5		D	Ac-FASKGNVFVDGYFERLRAKL-NH2	470
JBT0793	88		C	Ac-FQAKGNVFVDGYFERLRAKL-NH2	294
JBT0794	90	156	C	Ac-FQSAGNVFVDGYFERLRAKL-NH2	295
JBT0795	92	34	A	Ac-FQSKGAVFVDGYFERLRAKL-NH2	72
JBT0796	70		D	Ac-FQSKGNAFVDGYFERLRAKL-NH2	471
JBT0797	-2			Ac-FQSKGNVAVDGYFERLRAKL-NH2	824
JBT0798	39		F	Ac-FQSKGNVFADGYFERLRAKL-NH2	731
JBT0799	8			Ac-FQSKGNVFVDAYFERLRAKL-NH2	825
JBT0800	-2			Ac-FQSKGNVFVDGAFERLRAKL-NH2	826
JBT0801	64		D	Ac-FQSKGNVFVDGYAERLRAKL-NH2	472
JBT0802	101	35	A	Ac-FQSKGNVFVDGYFARLRAKL-NH2	73
JBT0803	7		D	Ac-FQSKGNVFVDGYFEALRAKL-NH2	473
JBT0804	10			Ac-FQSKGNVFVDGYFERARAKL-NH2	827
JBT0805	98	64	B	Ac-FQSKGNVFVDGYFERLAAKL-NH2	212

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0806		42	A	Ac-FQSK-Nmg-NVFVAGYFARLRAKL-NH2	74
JBT0807	-6			Ac-FQSKGNVFVDGYFERLRAAL-NH2	828
JBT0808	39		F	Ac-FQSKGNVFVDGYFERLRAKA-NH2	732
JBT0809	10			Ac-DQSKGNVFVDGYFERLRAKL-NH2	829
JBT0810	-1			Ac-FDSKGNVFVDGYFERLRAKL-NH2	830
JBT0811	69		D	Ac-FQDKGNVFVDGYFERLRAKL-NH2	474
JBT0812	81		C	Ac-FQSDGNVFVDGYFERLRAKL-NH2	296
JBT0813	76		D	Ac-FQSKGDVFVDGYFERLRAKL-NH2	475
JBT0814	4			Ac-FQSKGNDFVDGYFERLRAKL-NH2	831
JBT0815	2			Ac-FQSKGNVDVDGYFERLRAKL-NH2	832
JBT0816	1			Ac-FQSKGNVFDDGYFERLRAKL-NH2	833
JBT0817	5			Ac-FQSKGNVFVDDYFERLRAKL-NH2	834
JBT0818	3		D	Ac-FQSKGNVFVDGDFERLRAKL-NH2	476
JBT0819	31		D	Ac-FQSKGNVFVDGYDERLRAKL-NH2	477
JBT0820	0			Ac-FQSKGNVFVDGYFEDLRAKL-NH2	835
JBT0821	-3			Ac-FQSKGNVFVDGYFERDRAKL-NH2	836
JBT0822	51		D	Ac-FQSKGNVFVDGYFERLDAKL-NH2	478
JBT0823	82		C	Ac-FQSKGNVFVDGYFERLRDKL-NH2	297
JBT0824	3			Ac-FQSKGNVFVDGYFERLRADL-NH2	837
JBT0825	9			Ac-FQSKGNVFVDGYFERLRAKD-NH2	838
JBT0826	22		D	Ac-EQSKGNVFVDGYFERLRAKL-NH2	479
JBT0827	-3			Ac-FESKGNVFVDGYFERLRAKL-NH2	839
JBT0828	90		C	Ac-FQEKGNVFVDGYFERLRAKL-NH2	298
JBT0829	104		A	Ac-FQSEGNVFVDGYFERLRAKL-NH2	75
JBT0830	75		D	Ac-FQSKGEVFVDGYFERLRAKL-NH2	480
JBT0831	18		D	Ac-FQSKGNEFVDGYFERLRAKL-NH2	481
JBT0832	-5			Ac-FQSKGNVEVDGYFERLRAKL-NH2	840

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0833	11			Ac-FQSKGNVFEDGYFERLRAKL-NH2	841
JBT0834	13			Ac-FQSKGNVFVDEYFERLRAKL-NH2	842
JBT0835	5			Ac-FQSKGNVFVDGEFERLRAKL-NH2	843
JBT0836	51		D	Ac-FQSKGNVFVDGYEERLRAKL-NH2	482
JBT0837		87	B	Ac-FQSK-Nmg-NVFVTGYFERL-Nle-AKL-NH2	213
JBT0838	4		D	Ac-FQSKGNVFVDGYFEELRAKL-NH2	483
JBT0839	4			Ac-FQSKGNVFVDGYFERERAKL-NH2	844
JBT0840	71		D	Ac-FQSKGNVFVDGYFERLEAKL-NH2	484
JBT0841	82		C	Ac-FQSKGNVFVDGYFERLREKL-NH2	299
JBT0842	3			Ac-FQSKGNVFVDGYFERLRAEL-NH2	845
JBT0843	21			Ac-FQSKGNVFVDGYFERLRAKE-NH2	846
JBT0844		112	C	Ac-FQSKaNVFVDGYFERLRAKL-NH2	300
JBT0845	5		D	Ac-FFSKGNVFVDGYFERLRAKL-NH2	485
JBT0846	66		D	Ac-FQFKGNVFVDGYFERLRAKL-NH2	486
JBT0847	82		C	Ac-FQSFGNVFVDGYFERLRAKL-NH2	301
JBT0848	69		D	Ac-FQSKGFVFVDGYFERLRAKL-NH2	487
JBT0849	48		D	Ac-FQSKGNFFVDGYFERLRAKL-NH2	488
JBT0850		150	C	Ac-FQSKaAVFVAGYFARLRAKL-NH2	302
JBT0851	6		D	Ac-FQSKGNVFFDGYFERLRAKL-NH2	489
JBT0852	18			Ac-FQSKGNVFVDFYFERLRAKL-NH2	847
JBT0853	13			Ac-FQSKGNVFVDGFFERLRAKL-NH2	848
JBT0854		593	D	Ac-FQSKaAVFVAGYFARL-Nle-AKL-NH2	490
JBT0855	57		D	Ac-FQSKGNVFVDGYFFRLRAKL-NH2	491
JBT0856	4		D	Ac-FQSKGNVFVDGYFEFLRAKL-NH2	492
JBT0857	13			Ac-FQSKGNVFVDGYFERFRAKL-NH2	849
JBT0858	85		C	Ac-FQSKGNVFVDGYFERLFAKL-NH2	303
JBT0859	81		C	Ac-FQSKGNVFVDGYFERLRFKL-NH2	304

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0860	1			Ac-FQSKGNVFVDGYFERLRAFL-NH2	850
JBT0861	94		C	Ac-FQSKGNVFVDGYFERLRAKF-NH2	305
JBT0862	23			Ac-GQSKGNVFVDGYFERLRAKL-NH2	851
JBT0863	7		D	Ac-FGSKGNVFVDGYFERLRAKL-NH2	493
JBT0864	60		D	Ac-FQGKGNVFVDGYFERLRAKL-NH2	494
JBT0865	69		D	Ac-FQSGGNVFVDGYFERLRAKL-NH2	495
JBT0866	79		D	Ac-FQSKGGVFVDGYFERLRAKL-NH2	496
JBT0867	2			Ac-FQSKGNGFVDGYFERLRAKL-NH2	852
JBT0868	4			Ac-FQSKGNVGVDGYFERLRAKL-NH2	853
JBT0869	-1			Ac-FQSKGNVFGDGYFERLRAKL-NH2	854
JBT0870		57	B	Ac-FQSKaAVFVTGYFARLRAKL-NH2	214
JBT0871	1			Ac-FQSKGNVFVDGGFERLRAKL-NH2	855
JBT0872	24		D	Ac-FQSKGNVFVDGYGERLRAKL-NH2	497
JBT0873	64		D	Ac-FQSKGNVFVDGYFGRLRAKL-NH2	498
JBT0874	1		D	Ac-FQSKGNVFVDGYFEGLRAKL-NH2	499
JBT0875	-5			Ac-FQSKGNVFVDGYFERGRAKL-NH2	856
JBT0876	75		D	Ac-FQSKGNVFVDGYFERLGAKL-NH2	500
JBT0877	50		E	Ac-FQSKGNVFVDGYFERLRGKL-NH2	701
JBT0878	9			Ac-FQSKGNVFVDGYFERLRAGL-NH2	857
JBT0879	29			Ac-FQSKGNVFVDGYFERLRAKG-NH2	858
JBT0880	56		D	Ac-HQSKGNVFVDGYFERLRAKL-NH2	501
JBT0881	-2			Ac-FHSKGNVFVDGYFERLRAKL-NH2	859
JBT0882	85		C	Ac-FQHKGNVFVDGYFERLRAKL-NH2	306
JBT0883	93		C	Ac-FQSHGNVFVDGYFERLRAKL-NH2	307
JBT0884	71		D	Ac-FQSKGHVFVDGYFERLRAKL-NH2	502
JBT0885	50		D	Ac-FQSKGNHFVDGYFERLRAKL-NH2	503
JBT0886		50	B	Ac-FQSKaNVFVDGYFARLRAKL-NH2	215

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0887	0			Ac-FQSKGNVFHDGYFERLRAKL-NH2	860
JBT0888	35		D	Ac-FQSKGNVFVDHYFERLRAKL-NH2	504
JBT0889	-8			Ac-FQSKGNVFVDGHFERLRAKL-NH2	861
JBT0890	71		D	Ac-FQSKGNVFVDGYHERLRAKL-NH2	505
JBT0891	93		C	Ac-FQSKGNVFVDGYFHRLRAKL-NH2	308
JBT0892	-8			Ac-FQSKGNVFVDGYFEHLRAKL-NH2	862
JBT0893	-10			Ac-FQSKGNVFVDGYFERHRAKL-NH2	863
JBT0894	103	42	A	Ac-FQSKGNVFVDGYFERLHAKL-NH2	76
JBT0895	69		D	Ac-FQSKGNVFVDGYFERLRHKL-NH2	506
JBT0896	-12			Ac-FQSKGNVFVDGYFERLRAHL-NH2	864
JBT0897	71		D	Ac-FQSKGNVFVDGYFERLRAKH-NH2	507
JBT0898	87		C	Ac-IQSKGNVFVDGYFERLRAKL-NH2	309
JBT0899	7		D	Ac-FISKGNVFVDGYFERLRAKL-NH2	508
JBT0900	88		C	Ac-FQIKGNVFVDGYFERLRAKL-NH2	310
JBT0901	96		A	Ac-FQSIGNVFVDGYFERLRAKL-NH2	77
JBT0902	94		C	Ac-FQSKGIVFVDGYFERLRAKL-NH2	311
JBT0903	97		A	Ac-FQSKGNIFVDGYFERLRAKL-NH2	78
JBT0904	2			Ac-FQSKGNVIVDGYFERLRAKL-NH2	865
JBT0905	58		D	Ac-FQSKGNVFIDGYFERLRAKL-NH2	509
JBT0906	-2			Ac-FQSKGNVFVDIYFERLRAKL-NH2	866
JBT0907	1			Ac-FQSKGNVFVDGIFERLRAKL-NH2	867
JBT0908	39		D	Ac-FQSKGNVFVDGYIERLRAKL-NH2	510
JBT0909	92		C	Ac-FQSKGNVFVDGYFIRLRAKL-NH2	312
JBT0910	0			Ac-FQSKGNVFVDGYFEILRAKL-NH2	868
JBT0911	15			Ac-FQSKGNVFVDGYFERIRAKL-NH2	869
JBT0912	104	57	B	Ac-FQSKGNVFVDGYFERLIAKL-NH2	216
JBT0913	65		D	Ac-FQSKGNVFVDGYFERLRIKL-NH2	511

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0914	-7			Ac-FQSKGNVFVDGYFERLRAIL-NH2	870
JBT0915	80		D	Ac-FQSKGNVFVDGYFERLRAKI-NH2	512
JBT0916	59		D	Ac-KQSKGNVFVDGYFERLRAKL-NH2	513
JBT0917	-3			Ac-FKSKGNVFVDGYFERLRAKL-NH2	871
JBT0918	65		D	Ac-FQKKGNVFVDGYFERLRAKL-NH2	514
JBT0919		42	A	Ac-FQSKaNVFVTGYFERLRAKL-NH2	79
JBT0920	115	23	A	Ac-FQSKGKVFVDGYFERLRAKL-NH2	80
JBT0921	11			Ac-FQSKGNKFVDGYFERLRAKL-NH2	872
JBT0922	-4			Ac-FQSKGNVKVDGYFERLRAKL-NH2	873
JBT0923	3		D	Ac-FQSKGNVFKDGYFERLRAKL-NH2	515
JBT0924	17			Ac-FQSKGNVFVDKYFERLRAKL-NH2	874
JBT0925	-6			Ac-FQSKGNVFVDGKFERLRAKL-NH2	875
JBT0926	20		D	Ac-FQSKGNVFVDGYKERLRAKL-NH2	516
JBT0927	-8			Ac-FQSKGNVFVDGYFEKLRAKL-NH2	876
JBT0928	4			Ac-FQSKGNVFVDGYFERKRAKL-NH2	877
JBT0929	107	29	A	Ac-FQSKGNVFVDGYFERLKAKL-NH2	81
JBT0930	44		E	Ac-FQSKGNVFVDGYFERLRKKL-NH2	702
JBT0931		33	A	Ac-FQSKGNVFVDGYFERL-Eag-AKL-NH2	82
JBT0932	47		E	Ac-FQSKGNVFVDGYFERLRAKK-NH2	703
JBT0933	75		D	Ac-LQSKGNVFVDGYFERLRAKL-NH2	517
JBT0934	-7			Ac-FLSKGNVFVDGYFERLRAKL-NH2	878
JBT0935	72		D	Ac-FQLKGNVFVDGYFERLRAKL-NH2	518
JBT0936	97	41	A	Ac-FQSLGNVFVDGYFERLRAKL-NH2	83
JBT0937	90		C	Ac-FQSKGLVFVDGYFERLRAKL-NH2	313
JBT0938	44		D	Ac-FQSKGNLFVDGYFERLRAKL-NH2	519
JBT0939	27			Ac-FQSKGNVLVDGYFERLRAKL-NH2	879
JBT0940	4			Ac-FQSKGNVFLDGYFERLRAKL-NH2	880

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM)	Affinity Class	Sequence	SEQ ID NO:
JBT0941	28		D	Ac-FQSKGNVFVDLYFERLRAKL-NH2	520
JBT0942	4		D	Ac-FQSKGNVFVDGLFERLRAKL-NH2	521
JBT0943	83		C	Ac-FQSKGNVFVDGYLERLRAKL-NH2	314
JBT0944	103		A	Ac-FQSKGNVFVDGYFLRLRAKL-NH2	84
JBT0945	1		D	Ac-FQSKGNVFVDGYFELLRAKL-NH2	522
JBT0946		38	A	Ac-FQSKGNVFVDGYFERL-Dab-AKL-NH2	85
JBT0947	105		A	Ac-FQSKGNVFVDGYFERLLAKL-NH2	86
JBT0948	108	46	A	Ac-FQSKGNVFVDGYFERLRLKL-NH2	87
JBT0949	2			Ac-FQSKGNVFVDGYFERLRALL-NH2	881
JBT0950		73	B	Ac-FQSKaNVFVFGYFARLRAKL-NH2	217
JBT0951	97		A	Ac-MQSKGNVFVDGYFERLRAKL-NH2	88
JBT0953	109	43	A	Ac-FQMKGNVFVDGYFERLRAKL-NH2	89
JBT0956	27			Ac-FQSKGNMFVDGYFERLRAKL-NH2	882
JBT0961	77		D	Ac-FQSKGNVFVDGYMERLRAKL-NH2	523
JBT0962	107	40	A	Ac-FQSKGNVFVDGYFMRLRAKL-NH2	90
JBT0964	28			Ac-FQSKGNVFVDGYFERMRAKL-NH2	883
JBT0966	100		A	Ac-FQSKGNVFVDGYFERLRMKL-NH2	91
JBT0969	65		D	Ac-NQSKGNVFVDGYFERLRAKL-NH2	524
JBT0970	7		D	Ac-FNSKGNVFVDGYFERLRAKL-NH2	525
JBT0971	83		C	Ac-FQNKGNVFVDGYFERLRAKL-NH2	315
JBT0972	94		C	Ac-FQSNGNVFVDGYFERLRAKL-NH2	316
JBT0973		37	A	Ac-FQSKGNVFVDGYFERL-Cha-AKL-NH2	92
JBT0974	29			Ac-FQSKGNNFVDGYFERLRAKL-NH2	884
JBT0975	-3			Ac-FQSKGNVNVDGYFERLRAKL-NH2	885
JBT0976	-2			Ac-FQSKGNVFNDGYFERLRAKL-NH2	886
JBT0977	15			Ac-FQSKGNVFVDNYFERLRAKL-NH2	887
JBT0978	-10			Ac-FQSKGNVFVDGNFERLRAKL-NH2	888

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0979	44		D	Ac-FQSKGNVFVDGYNERLRAKL-NH2	526
JBT0980	93	40	A	Ac-FQSKGNVFVDGYFNRLRAKL-NH2	93
JBT0982	-2			Ac-FQSKGNVFVDGYFERNRAKL-NH2	889
JBT0983	81		C	Ac-FQSKGNVFVDGYFERLNAKL-NH2	317
JBT0984	100	34	A	Ac-FQSKGNVFVDGYFERLRNKL-NH2	94
JBT0985	4			Ac-FQSKGNVFVDGYFERLRANL-NH2	890
JBT0986	32		F	Ac-FQSKGNVFVDGYFERLRAKN-NH2	733
JBT0987	27			Ac-PQSKGNVFVDGYFERLRAKL-NH2	891
JBT0988	0			Ac-FPSKGNVFVDGYFERLRAKL-NH2	892
JBT0989	5			Ac-FQPKGNVFVDGYFERLRAKL-NH2	893
JBT0990	106	33	A	Ac-FQSPGNVFVDGYFERLRAKL-NH2	95
JBT0991	5			Ac-FQSKGPVFVDGYFERLRAKL-NH2	894
JBT0992	65		D	Ac-FQSKGNPFVDGYFERLRAKL-NH2	527
JBT0993	1			Ac-FQSKGNVPVDGYFERLRAKL-NH2	895
JBT0994	2			Ac-FQSKGNVFPDGYFERLRAKL-NH2	896
JBT0995	4		D	Ac-FQSKGNVFVDPYFERLRAKL-NH2	528
JBT0996	3			Ac-FQSKGNVFVDGPFERLRAKL-NH2	897
JBT0997	8		D	Ac-FQSKGNVFVDGYPERLRAKL-NH2	529
JBT0998	48		D	Ac-FQSKGNVFVDGYFPRLRAKL-NH2	530
JBT0999	3		D	Ac-FQSKGNVFVDGYFEPLRAKL-NH2	531
JBT1000	3			Ac-FQSKGNVFVDGYFERPRAKL-NH2	898
JBT1001	1			Ac-FQSKGNVFVDGYFERLPAKL-NH2	899
JBT1002	15			Ac-FQSKGNVFVDGYFERLRPKL-NH2	900
JBT1003	4			Ac-FQSKGNVFVDGYFERLRAPL-NH2	901
JBT1004	8			Ac-FQSKGNVFVDGYFERLRAKP-NH2	902
JBT1005	55		D	Ac-QQSKGNVFVDGYFERLRAKL-NH2	532
JBT1006		28	A	Ac-FQSKGNVFVDGYFERL-Hlc-AKL-NH2	96

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1007	98	28	A	Ac-FQQKGNVFVDGYFERLRAKL-NH2	97
JBT1008	83		C	Ac-FQSQGNVFVDGYFERLRAKL-NH2	318
JBT1009	99	38	A	Ac-FQSKGQVFVDGYFERLRAKL-NH2	98
JBT1010	46		D	Ac-FQSKGNQFVDGYFERLRAKL-NH2	533
JBT1011	8			Ac-FQSKGNVQVDGYFERLRAKL-NH2	903
JBT1012	14		D	Ac-FQSKGNVFQDGYFERLRAKL-NH2	534
JBT1013	23		D	Ac-FQSKGNVFVDQYFERLRAKL-NH2	535
JBT1014	4			Ac-FQSKGNVFVDGQFERLRAKL-NH2	904
JBT1015	55		D	Ac-FQSKGNVFVDGYQERLRAKL-NH2	536
JBT1016	102	27	A	Ac-FQSKGNVFVDGYFQRLRAKL-NH2	99
JBT1017	8		D	Ac-FQSKGNVFVDGYFEQLRAKL-NH2	537
JBT1018	12			Ac-FQSKGNVFVDGYFERQRAKL-NH2	905
JBT1019	76		D	Ac-FQSKGNVFVDGYFERLQAKL-NH2	538
JBT1020	88		C	Ac-FQSKGNVFVDGYFERLRQKL-NH2	319
JBT1021	2			Ac-FQSKGNVFVDGYFERLRAQL-NH2	906
JBT1022	55		D	Ac-FQSKGNVFVDGYFERLRAKQ-NH2	539
JBT1023	60		D	Ac-RQSKGNVFVDGYFERLRAKL-NH2	540
JBT1024	11		D	Ac-FRSKGNVFVDGYFERLRAKL-NH2	541
JBT1025	93	46	A	Ac-FQRKGNVFVDGYFERLRAKL-NH2	100
JBT1026	102	38	A	Ac-FQSRGNVFVDGYFERLRAKL-NH2	101
JBT1027	110	29	A	Ac-FQSKGRVFVDGYFERLRAKL-NH2	102
JBT1028	15		D	Ac-FQSKGNRFVDGYFERLRAKL-NH2	542
JBT1029	18		D	Ac-FQSKGNVRVDGYFERLRAKL-NH2	543
JBT1030	18		D	Ac-FQSKGNVFRDGYFERLRAKL-NH2	544
JBT1031	27			Ac-FQSKGNVFVDRYFERLRAKL-NH2	907
JBT1032	6			Ac-FQSKGNVFVDGRFERLRAKL-NH2	908
JBT1033	60		D	Ac-FQSKGNVFVDGYRERLRAKL-NH2	545

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1034	103	36	A	Ac-FQSKGNVFVDGYFRRLRAKL-NH2	103
JBT1035		92	B	Ac-FQSK-Nmg-AVFVAGYFARL-Nle-AKL-NH2	218
JBT1036	29		D	Ac-FQSKGNVFVDGYFERRRAKL-NH2	546
JBT1037		27	A	Ac-FQSKGNVFVDGYFERL-Nle-AKL-NH2	104
JBT1038	107		A	Ac-FQSKGNVFVDGYFERLRRKL-NH2	105
JBT1039	95		C	Ac-FQSKGNVFVDGYFERLRARL-NH2	320
JBT1040	84		C	Ac-FQSKGNVFVDGYFERLRAKR-NH2	321
JBT1041	37		F	Ac-SQSKGNVFVDGYFERLRAKL-NH2	734
JBT1042	2		D	Ac-FSSKGNVFVDGYFERLRAKL-NH2	547
JBT1043		40	A	Ac-FQSKaNVFVDGYFERL-Nle-AKL-NH2	106
JBT1044	78		D	Ac-FQSSGNVFVDGYFERLRAKL-NH2	548
JBT1045	109	50	A	Ac-FQSKGSVFVDGYFERLRAKL-NH2	107
JBT1046	57		D	Ac-FQSKGNSFVDGYFERLRAKL-NH2	549
JBT1047	5			Ac-FQSKGNVSVDGYFERLRAKL-NH2	909
JBT1048	38		D	Ac-FQSKGNVFSDGYFERLRAKL-NH2	550
JBT1049	18		D	Ac-FQSKGNVFVDSYFERLRAKL-NH2	551
JBT1050	2			Ac-FQSKGNVFVDGSFERLRAKL-NH2	910
JBT1051	47		D	Ac-FQSKGNVFVDGYSERLRAKL-NH2	552
JBT1052	84		C	Ac-FQSKGNVFVDGYFSRLRAKL-NH2	322
JBT1053	-7			Ac-FQSKGNVFVDGYFESLRAKL-NH2	911
JBT1054	1			Ac-FQSKGNVFVDGYFERSRAKL-NH2	912
JBT1055	78		D	Ac-FQSKGNVFVDGYFERLSAKL-NH2	553
JBT1056	78		D	Ac-FQSKGNVFVDGYFERLRSKL-NH2	554
JBT1057	-9			Ac-FQSKGNVFVDGYFERLRASL-NH2	913
JBT1058	58		D	Ac-FQSKGNVFVDGYFERLRAKS-NH2	555
JBT1059	69		D	Ac-TQSKGNVFVDGYFERLRAKL-NH2	556
JBT1060	-1			Ac-FTSKGNVFVDGYFERLRAKL-NH2	914

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM)	Affinity Class	Sequence	SEQ ID NO:
JBT1061	73		D	Ac-FQTKGNVFVDGYFERLRAKL-NH2	557
JBT1062	51		D	Ac-FQSTGNVFVDGYFERLRAKL-NH2	558
JBT1063	72		D	Ac-FQSKGTVFVDGYFERLRAKL-NH2	559
JBT1064	49		D	Ac-FQSKGNTFVDGYFERLRAKL-NH2	560
JBT1065	-12			Ac-FQSKGNVTVDGYFERLRAKL-NH2	915
JBT1066	24		D	Ac-FQSKGNVFTDGYFERLRAKL-NH2	561
JBT1067	-2			Ac-FQSKGNVFVDTYFERLRAKL-NH2	916
JBT1068	-10			Ac-FQSKGNVFVDGTFERLRAKL-NH2	917
JBT1069	46		D	Ac-FQSKGNVFVDGYTERLRAKL-NH2	562
JBT1070	97		A	Ac-FQSKGNVFVDGYFTRLRAKL-NH2	108
JBT1071	-9			Ac-FQSKGNVFVDGYFETLRAKL-NH2	918
JBT1072	9			Ac-FQSKGNVFVDGYFERTRAKL-NH2	919
JBT1073	82		C	Ac-FQSKGNVFVDGYFERLTAKL-NH2	323
JBT1074	71		D	Ac-FQSKGNVFVDGYFERLRTKL-NH2	563
JBT1075	-15			Ac-FQSKGNVFVDGYFERLRATL-NH2	920
JBT1076	69		D	Ac-FQSKGNVFVDGYFERLRAKT-NH2	564
JBT1077	81		C	Ac-VQSKGNVFVDGYFERLRAKL-NH2	324
JBT1078	-3			Ac-FVSKGNVFVDGYFERLRAKL-NH2	921
JBT1079	93		C	Ac-FQVKGNVFVDGYFERLRAKL-NH2	325
JBT1080	92		C	Ac-FQSVGNVFVDGYFERLRAKL-NH2	326
JBT1081	95		C	Ac-FQSKGVVFVDGYFERLRAKL-NH2	327
JBT1082		250	D	Ac-FQSKaAVFVFGYFARLRAKL-NH2	565
JBT1083	17			Ac-FQSKGNVVVDGYFERLRAKL-NH2	922
JBT1084		45	A	Ac-FQSK-Nmg-AVFVAGYFARLRAKL-NH2	109
JBT1085	-10			Ac-FQSKGNVFVDVYFERLRAKL-NH2	923
JBT1086	-15			Ac-FQSKGNVFVDGVFERLRAKL-NH2	924
JBT1087	40		D	Ac-FQSKGNVFVDGYVERLRAKL-NH2	566

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1088	99		A	Ac-FQSKGNVFVDGYFVRLRAKL-NH2	110
JBT1089	-4			Ac-FQSKGNVFVDGYFEVLRAKL-NH2	925
JBT1090	37		D	Ac-FQSKGNVFVDGYFERVRAKL-NH2	567
JBT1091	91	53	B	Ac-FQSKGNVFVDGYFERLVAKL-NH2	219
JBT1092	85		C	Ac-FQSKGNVFVDGYFERLRVKL-NH2	328
JBT1093	6			Ac-FQSKGNVFVDGYFERLRAVL-NH2	926
JBT1094	68		D	Ac-FQSKGNVFVDGYFERLRAKV-NH2	568
JBT1095	92		C	Ac-WQSKGNVFVDGYFERLRAKL-NH2	329
JBT1096	-3			Ac-FWSKGNVFVDGYFERLRAKL-NH2	927
JBT1097	55		D	Ac-FQWKGNVFVDGYFERLRAKL-NH2	569
JBT1099	58		D	Ac-FQSKGWVFVDGYFERLRAKL-NH2	570
JBT1100	45		D	Ac-FQSKGNWFVDGYFERLRAKL-NH2	571
JBT1101	55		D	Ac-FQSKGNVWVDGYFERLRAKL-NH2	572
JBT1102	4		D	Ac-FQSKGNVFWDGYFERLRAKL-NH2	573
JBT1103	15		D	Ac-FQSKGNVFVDWYFERLRAKL-NH2	574
JBT1104	0			Ac-FQSKGNVFVDGWFERLRAKL-NH2	928
JBT1105	85		C	Ac-FQSKGNVFVDGYWERLRAKL-NH2	330
JBT1106	82	59	B	Ac-FQSKGNVFVDGYFWRLRAKL-NH2	220
JBT1107	-7			Ac-FQSKGNVFVDGYFEWLRAKL-NH2	929
JBT1108	9		D	Ac-FQSKGNVFVDGYFERWRAKL-NH2	575
JBT1109	78		D	Ac-FQSKGNVFVDGYFERLWAKL-NH2	576
JBT1110	93		C	Ac-FQSKGNVFVDGYFERLRWKL-NH2	331
JBT1111	-6			Ac-FQSKGNVFVDGYFERLRAWL-NH2	930
JBT1112	87		C	Ac-FQSKGNVFVDGYFERLRAKW-NH2	332
JBT1113	101		A	Ac-YQSKGNVFVDGYFERLRAKL-NH2	111
JBT1114	12			Ac-FYSKGNVFVDGYFERLRAKL-NH2	931
JBT1115	75		D	Ac-FQYKGNVFVDGYFERLRAKL-NH2	577

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1117	83		C	Ac-FQSKGYVFVDGYFERLRAKL-NH2	333
JBT1118	82		C	Ac-FQSKGNYFVDGYFERLRAKL-NH2	334
JBT1119	99		A	Ac-FQSKGNVYVDGYFERLRAKL-NH2	112
JBT1120	11		D	Ac-FQSKGNVFYDGYFERLRAKL-NH2	578
JBT1121	12			Ac-FQSKGNVFVDYYFERLRAKL-NH2	932
JBT1122		33	A	Ac-FQSK-Nmg-NVFVDGYFARL-Nle-AKL-NH2	113
JBT1123	84		C	Ac-FQSKGNVFVDGYYERLRAKL-NH2	335
JBT1124	85		C	Ac-FQSKGNVFVDGYFYRLRAKL-NH2	336
JBT1125	8			Ac-FQSKGNVFVDGYFEYLRAKL-NH2	933
JBT1126	11			Ac-FQSKGNVFVDGYFERYRAKL-NH2	934
JBT1127	92		C	Ac-FQSKGNVFVDGYFERLYAKL-NH2	337
JBT1128	65		D	Ac-FQSKGNVFVDGYFERLRYKL-NH2	579
JBT1129	-7			Ac-FQSKGNVFVDGYFERLRAYL-NH2	935
JBT1130	76		D	Ac-FQSKGNVFVDGYFERLRAKY-NH2	580
JBT1132		20	A	Fam-FQSKGNVFVDGYFERLRAKL-NH2	114
JBT1133		31	A	Ac-FQSKkAVFVDGYFARLRAKL-NH2	115
JBT1134		39	A	Ac-FQSKGAVFVDGYFARLRAKL-NH2	116
JBT1135		82	B	Ac-FQSKDAVFVDGYFARLRAKL-NH2	221
JBT1136		40	A	Ac-FQSKdAVFVDGYFARLRAKL-NH2	117
JBT1137		40	A	Ac-FQSKkAVFVAGYFARLRAKL-NH2	118
JBT1138		64	B	Ac-FQSKGAVFVAGYFARLRAKL-NH2	222
JBT1139		87	B	Ac-FQSKDAVFVAGYFARLRAKL-NH2	223
JBT1140		36	A	Ac-FQSKdAVFVAGYFARLRAKL-NH2	119
JBT1141		34	A	Ac-FQSKkAVFVKGYFARLRAKL-NH2	120
JBT1142		53	B	Ac-FQSKGAVFVKGYFARLRAKL-NH2	224
JBT1143		58	B	Ac-FQSKDAVFVKGYFARLRAKL-NH2	225
JBT1144		34	A	Ac-FQSKdAVFVKGYFARLRAKL-NH2	121

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1145		45	A	Ac-FQSKkAVFVTGYFARLRAKL-NH2	122
JBT1146		65	B	Ac-FQSKGAVFVTGYFARLRAKL-NH2	226
JBT1147		95	B	Ac-FQSKDAVFVTGYFARLRAKL-NH2	227
JBT1148		46	A	Ac-FQSKdAVFVTGYFARLRAKL-NH2	123
JBT1149		>5000		Ac-FQSKGNVFvFGYFERLRAKL-NH2	936
JBT1150		1154	E	Ac-FQSKaNVFVTGYFERL-Nle-AKL-NH2	704
JBT1151		>5000		Ac-FQSKKNVFVFGYFERLRAKD-NH2	937
JBT1152		4119	F	Ac-FQSKKNVFFFGYFERLRAKL-NH2	735
JBT1153		1294	E	Ac-FQSKKNVFVFGYFERLGAKL-NH2	705
JBT1154		27	A	Fam-Ttds-FQSKGNVFVDGYFERLRAKL-NH2	124
JBT1155		31	A	Ac-FQSK-Nmg-NVFVTGYFERLRAKL-NH2	125
JBT1156		41	A	Ac-FQSKaNVFVDGYFARL-Nle-AKL-NH2	126
JBT1157		34	A	Ac-FQSKaNVFVAGYFARLRAKL-NH2	127
JBT1158		51	B	Ac-FQSKGNVFVFGYFERLRAKL-N-methyl	228
JBT1159		38	A	Ac-FQSKGNVFVFGYFERLRAKL-N-ethyl	128
JBT1161		122	C	Ac-FQSK-Aib-NVFVFGYFERLRAKL-NH2	338
JBT1162		28	A	Ac-FQSKpNVFVFGYFERLRAKL-NH2	129
JBT1164		28	A	Ac-FQSKGNVFVDGYFERLRAKLC-NH2	130
JBT1166	70	129	C	Ac-FQSKANVFVDGYFERLRAKL-NH2	339
JBT1167	57		D	Ac-FQSKENVFVDGYFERLRAKL-NH2	581
JBT1168	86	94	B	Ac-FQSKFNVFVDGYFERLRAKL-NH2	229
JBT1169	70	127	C	Ac-FQSKHNVFVDGYFERLRAKL-NH2	340
JBT1170	37		D	Ac-FQSKINVFVDGYFERLRAKL-NH2	582
JBT1171	81		C	Ac-FQSKLNVFVDGYFERLRAKL-NH2	341
JBT1173	11		D	Ac-FQSKPNVFVDGYFERLRAKL-NH2	583
JBT1174	83	115	C	Ac-FQSKRNVFVDGYFERLRAKL-NH2	342
JBT1175	63		D	Ac-FQSKSNVFVDGYFERLRAKL-NH2	584

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1176	79		D	Ac-FQSKTNVFVDGYFERLRAKL-NH2	585
JBT1177	61		D	Ac-FQSKVNVFVDGYFERLRAKL-NH2	586
JBT1178	54		D	Ac-FQSKWNVFVDGYFERLRAKL-NH2	587
JBT1179	86	100	C	Ac-FQSKYNVFVDGYFERLRAKL-NH2	343
JBT1180	101	98	B	Ac-FQSKGNVFVAGYFERLRAKL-NH2	230
JBT1181	86	107	C	Ac-FQSKGNVFVGGYFERLRAKL-NH2	344
JBT1182	71		D	Ac-FQSKGNVFVIGYFERLRAKL-NH2	588
JBT1183	52		D	Ac-FQSKGNVFVLGYFERLRAKL-NH2	589
JBT1184	97	52	B	Ac-FQSKGNVFVMGYFERLRAKL-NH2	231
JBT1185	105	50	B	Ac-FQSKGNVFVPGYFERLRAKL-NH2	232
JBT1186	60		D	Ac-FQSKGNVFVQGYFERLRAKL-NH2	590
JBT1188	104		A	Ac-FQSKGNVFVSGYFERLRAKL-NH2	131
JBT1189	124	48	A	Ac-FQSKGNVFVVGYFERLRAKL-NH2	132
JBT1190	116	101	C	Ac-FQSKGNVFVWGYFERLRAKL-NH2	345
JBT1191	112	158	C	Ac-FQSKGNVFVYGYFERLRAKL-NH2	346
JBT1192	81	113	C	Ac-FQSKANVFVDGYFERLAAKL-NH2	347
JBT1193	73		D	Ac-FQSKFNVFVDGYFERLAAKL-NH2	591
JBT1194	65		D	Ac-FQSKYNVFVDGYHERLAAKL-NH2	592
JBT1195	44		D	Ac-FQSSHNVFVDGYFERLAAKL-NH2	593
JBT1196	85		C	Ac-FQSRENVFVDGYFERLAAKL-NH2	348
JBT1197	100	63	B	Ac-FQSKGNVFVDGYFMRLAAKL-NH2	233
JBT1198	60		D	Ac-FQSAKNVFVDGYFERLRAKL-NH2	594
JBT1199	62		D	Ac-FQSFKNVFVDGYFERLAAKL-NH2	595
JBT1200	53	226	C	Ac-FQSKDNVFVHGYFERLAAKL-NH2	349
JBT1201	44		D	Ac-FQDPKNVFVDGYFERLAAKL-NH2	596
JBT1202	34		D	Ac-FQSKVNVFVDGYFERLAAKL-NH2	597
JBT1203	50		D	Ac-FQSIKNVFVDGYFERLAAKL-NH2	598

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1204	106	98	B	Ac-YQSKNNVFVDGYFERLAAKL-NH2	234
JBT1205	85	128	C	Ac-FQSKERVFVDGYFERLAAKL-NH2	350
JBT1206	104	74	B	Ac-FQSKGHVFVDGYFERLAAKL-NH2	235
JBT1207	48		D	Ac-FQSKTNIFVDGYFERLAAKL-NH2	599
JBT1208	37		D	Ac-FQSHYNVFVDGYFERLAAKL-NH2	600
JBT1209	32		D	Ac-FQSKENVFVDGYFDRLAAKL-NH2	601
JBT1210	61	137	C	Ac-FQSKQNVFVDGYFERLAIKL-NH2	351
JBT1211	79	105	C	Ac-FQSKYNVFVDGYFERLAVKL-NH2	352
JBT1212	86		C	Ac-FQSRNNVFVDGYFERLAAKL-NH2	353
JBT1213	64	122	C	Ac-FQSRQNVFVDGYFERLAAKL-NH2	354
JBT1214	64	132	C	Ac-FQSKQNVFVDGYFERLAAKL-NH2	355
JBT1215	72	171	C	Ac-FQSSKNVFVDGYFERLAAKL-NH2	356
JBT1216	70	117	C	Ac-FQSKENVFVDGYFERLAAKL-NH2	357
JBT1217	63		D	Ac-FQSHHNVFVDGYFERLAAKL-NH2	602
JBT1218	72		D	Ac-FQSYKNVFVDGYFERLAAKL-NH2	603
JBT1219	76	90	B	Ac-FQSKKNVFVDGYFERLAAKL-NH2	236
JBT1220	85	85	B	Ac-FQTKHNVFVDGYFERLAAKL-NH2	237
JBT1221	67		D	Ac-FQSKHNVFVDGYFERLAFKL-NH2	604
JBT1222	85	82	B	Ac-FQSKYNVFVDGYFERLAAKL-NH2	238
JBT1223	54		D	Ac-FQSFKHVFVDGYFERLAAKL-NH2	605
JBT1224	57		D	Ac-FQSKANVHVDGYFERLAAKL-NH2	606
JBT1225	77	107	C	Ac-FQSKDNVFVDGYFERLAAKL-NH2	358
JBT1226	26			Ac-LQSKTNVFVDGYFERLAAKL-NH2	938
JBT1227	99	76	B	Ac-FQSKNNVFVDGYFERLAAKL-NH2	239
JBT1228	99	55	B	Ac-FQSKDHVFVDGYFERLAAKL-NH2	240
JBT1229	93	92	B	Ac-FQSKHNVFVDGYFERLARKL-NH2	241
JBT1230	52		D	Ac-FQSKSNVFVDGYFERLAAKL-NH2	607

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1231	68	92	B	Ac-FQSKHNVFVDGYFERLAAKL-NH2	242
JBT1232	47		D	Ac-FQSHKNVFVDGYFERLAAKL-NH2	608
JBT1233	11			Ac-FQSKFNVFVDGYTERLAAKL-NH2	939
JBT1234	63		D	Ac-FQSKDNVFVDGYFERLAAKF-NH2	609
JBT1235	65		D	Ac-FQSKMNVFVDGYFERLAAKL-NH2	610
JBT1236	41		E	Ac-FQSVKNVFVDGYFERLAAKL-NH2	706
JBT1238	14			Ac-FQSKHNVFVDGHFERLAAKL-NH2	940
JBT1240	61		D	Ac-FQSAYNVFVDGYFERLAAKL-NH2	611
JBT1241	53		D	Ac-FQSKGNWFVDGYFERLAAKL-NH2	612
JBT1242	49		E	Ac-FQSKVAVFVDGYFERLAAKL-NH2	707
JBT1243	59		D	Ac-FQSAKNVFVDGYFVRLRAKL-NH2	613
JBT1244	62		D	Ac-FQSKTNVFVDGYFERLAAKL-NH2	614
JBT1245	34		F	Ac-FQSKKDVFVDGYFERLAAKL-NH2	736
JBT1246	68		D	Ac-FQSKHVVFVDGYFERLAAKL-NH2	615
JBT1247	39		F	Ac-FQSKSNVFVDGYFERLAARL-NH2	737
JBT1248	6			Ac-FQSVHNVFVDGYFERPAAKL-NH2	941
JBT1249	7			Ac-FQSKYNVFVDEYFERLAAKL-NH2	942
JBT1250	66	216	C	Ac-FQSKDNVFVDGYFERLAARL-NH2	359
JBT1251	60		D	Ac-FQSKKHVFVDGYFERLAAKL-NH2	616
JBT1252	37		F	Ac-FQSTKNVFVDGYFERLAAKL-NH2	738
JBT1253	30		D	Ac-FQSDKNVFVDGYFERLAAKL-NH2	617
JBT1254	93	47	A	Ac-FQSKHHVFVDGYFERLAAKL-NH2	133
JBT1255	41		D	Ac-FQSNKNVFVDGYFERLAAKL-NH2	618
JBT1256	88	125	C	Ac-FQSKHNVYVDGYFERLAAKL-NH2	360
JBT1257	40		D	Ac-FQSHKNVFVEGYFERLAAKL-NH2	619
JBT1258	13			Ac-FQSKPNVFVDGYFERLAAKL-NH2	943
JBT1259	23		D	Ac-FQSYKDVFVDGYFERLATKL-NH2	620

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1260	113	50	A	Ac-FQSRRGVFVDGYFERLAAKL-NH2	134
JBT1261	81	98	B	Ac-FQSKLNVFVDGYFERLAAKL-NH2	243
JBT1262	15		D	Ac-FQSKNNFFVDGYFERLAARL-NH2	621
JBT1263	78	152	C	Ac-FQSKYNIFVDGYFERLAAKL-NH2	361
JBT1264	39		F	Ac-YQSKQNVFVDGYFERLAAKL-NH2	739
JBT1265	58		D	Ac-FQSEQNVFVDGYFERLAAKL-NH2	622
JBT1266	53		D	Ac-FQSEHNVFVDGYFERLAAKL-NH2	623
JBT1267	55	219	C	Ac-FQSKNDVFVDGYFERLAAKL-NH2	362
JBT1268	-8			Ac-HQQFKNVFVDGYFERLAAKL-NH2	944
JBT1269	58	155	C	Ac-FQSGGNVFVDGYFERLAAKL-NH2	363
JBT1270	61		D	Ac-FQSEKNVFVDGYFERLAAKL-NH2	624
JBT1271	21		D	Ac-FQSKKNVFVDGYFERLAFKL-NH2	625
JBT1272	58		D	Ac-FQSKRNVFVDGYFERLAAKL-NH2	626
JBT1273	74	106	C	Ac-FQSPKNVFVDGYFERLAAKL-NH2	364
JBT1274	28		D	Ac-FQSGKNVFVDGYFERLAAKL-NH2	627
JBT1275	68	123	C	Ac-FQSPKNVFVDGYFERLRAKL-NH2	365
JBT1277	-4			Ac-FQSKHNVFVDAYFERLRAKL-NH2	945
JBT1278	74	73	B	Ac-FQSVGNVFVDGYFERLSAKL-NH2	244
JBT1279	13		D	Ac-FQSYKNVFVDGYFERLFAKL-NH2	628
JBT1280	56	160	C	Ac-FQSRKNVFVDGYFERLRAKL-NH2	366
JBT1281	-5			Ac-FQSKKNVFVDGYSERLRAKL-NH2	946
JBT1282	52	176	C	Ac-FQSKKRVFVDGYFERLRAKL-NH2	367
JBT1283	36		F	Ac-FQSFENVFVDGYFERLHAKL-NH2	740
JBT1285	100	186	C	Ac-FQSIKNVFVDGYFERLRAKL-NH2	368
JBT1286	113	69	B	Ac-FQSYKNVFVDGYFERLRAKL-NH2	245
JBT1287	86		C	Ac-CQSKGNVFVDGYFERLRAKL-NH2	369
JBT1288	32		D	Ac-FCSKGNVFVDGYFERLRAKL-NH2	629

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1289	96		A	Ac-FQCKGNVFVDGYFERLRAKL-NH2	135
JBT1290	64		D	Ac-FQSCGNVFVDGYFERLRAKL-NH2	630
JBT1292	109	30	A	Ac-FQSKGCVFVDGYFERLRAKL-NH2	136
JBT1293	17		D	Ac-FQSKGNCFVDGYFERLRAKL-NH2	631
JBT1294	-6			Ac-FQSKGNVCVDGYFERLRAKL-NH2	947
JBT1295	1			Ac-FQSKGNVFCDGYFERLRAKL-NH2	948
JBT1296	65		D	Ac-FQSKGNVFVCGYFERLRAKL-NH2	632
JBT1297	66		D	Ac-FQSKGNVFVDCYFERLRAKL-NH2	633
JBT1298	-20			Ac-FQSKGNVFVDGCFERLRAKL-NH2	949
JBT1299	111	28	A	Ac-FQSKGNVFVDGYCERLRAKL-NH2	137
JBT1300	113		A	Ac-FQSKGNVFVDGYFCRLRAKL-NH2	138
JBT1301	-18			Ac-FQSKGNVFVDGYFECLRAKL-NH2	950
JBT1302	27		D	Ac-FQSKGNVFVDGYFERCRAKL-NH2	634
JBT1303	97		A	Ac-FQSKGNVFVDGYFERLCAKL-NH2	139
JBT1304	85		C	Ac-FQSKGNVFVDGYFERLRCKL-NH2	370
JBT1305	-6			Ac-FQSKGNVFVDGYFERLRACL-NH2	951
JBT1306	65		D	Ac-FQSKGNVFVDGYFERLRAKC-NH2	635
JBT1307	103		A	Ac-FQSKANVFVDGYFERLCAKL-NH2	140
JBT1308	86		C	Ac-FQSKFNVFVDGYFERLCAKL-NH2	371
JBT1309	92		C	Ac-FQSKYNVFVDGYHERLCAKL-NH2	372
JBT1310	95		C	Ac-FQSSHNVFVDGYFERLCAKL-NH2	373
JBT1311	83		C	Ac-FQSRENVFVDGYFERLCAKL-NH2	374
JBT1312	99		A	Ac-FQSKGNVFVDGYFMRLCAKL-NH2	141
JBT1313	87		C	Ac-FQSCKNVFVDGYFERLRAKL-NH2	375
JBT1314	55		D	Ac-FQSFKNVFVDGYFERLCAKL-NH2	636
JBT1315	95	35	A	Ac-FQSKDNVFVHGYFERLCAKL-NH2	142
JBT1316	90		C	Ac-FQDPKNVFVDGYFERLCAKL-NH2	376

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1317	76		D	Ac-FQSKVNVFVDGYFERLCAKL-NH2	637
JBT1318	73		D	Ac-FQSIKNVFVDGYFERLCAKL-NH2	638
JBT1319	108	20	A	Ac-YQSKNNVFVDGYFERLCAKL-NH2	143
JBT1320	110	271	D	Ac-FQSKERVFVDGYFERLCAKL-NH2	639
JBT1321	117	31	A	Ac-FQSKGHVFVDGYFERLCAKL-NH2	144
JBT1322	89		C	Ac-FQSKTNIFVDGYFERLCAKL-NH2	377
JBT1323	87	171	C	Ac-FQSHYNVFVDGYFERLCAKL-NH2	378
JBT1324	92	424	D	Ac-FQSKENVFVDGYFDRLCAKL-NH2	640
JBT1325	62		D	Ac-FQSKQNVFVDGYFERLCIKL-NH2	641
JBT1326	61		D	Ac-FQSKYNVFVDGYFERLCVKL-NH2	642
JBT1328	93		C	Ac-FQSRQNVFVDGYFERLCAKL-NH2	379
JBT1329	96	20	A	Ac-FQSKQNVFVDGYFERLCAKL-NH2	145
JBT1330	86		C	Ac-FQSSKNVFVDGYFERLCAKL-NH2	380
JBT1331	100		A	Ac-FQSKENVFVDGYFERLCAKL-NH2	146
JBT1332	87		C	Ac-FQSHHNVFVDGYFERLCAKL-NH2	381
JBT1333	72		D	Ac-FQSYKNVFVDGYFERLCAKL-NH2	643
JBT1334	103	23	A	Ac-FQSKKNVFVDGYFERLCAKL-NH2	147
JBT1335	111	247	C	Ac-FQTKHNVFVDGYFERLCAKL-NH2	382
JBT1336	61		D	Ac-FQSKHNVFVDGYFERLCFKL-NH2	644
JBT1337	74		D	Ac-FQSKYNVFVDGYFERLCAKL-NH2	645
JBT1338	46		D	Ac-FQSFKHVFVDGYFERLCAKL-NH2	646
JBT1339	88		C	Ac-FQSKANVHVDGYFERLCAKL-NH2	383
JBT1340	7			Ac-FQSCKNVFVDGYFERLCAKL-NH2	952
JBT1341	98	49	A	Ac-FQSKDNVFVDGYFERLCAKL-NH2	148
JBT1342	75		D	Ac-LQSKTNVFVDGYFERLCAKL-NH2	647
JBT1343	99		A	Ac-FQSKNNVFVDGYFERLCAKL-NH2	149
JBT1344	95		A	Ac-FQSKDHVFVDGYFERLCAKL-NH2	150

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1345	91		C	Ac-FQSKHNVFVDGYFERLCRKL-NH2	384
JBT1346	20		D	Ac-FQSKCNVFVDGYFERLCAKL-NH2	648
JBT1347	95	24	A	Ac-FQSKHNVFVDGYFERLCAKL-NH2	151
JBT1348	91		C	Ac-FQSKSNVFVDGYFERLCAKL-NH2	385
JBT1349	82		C	Ac-FQSHKNVFVDGYFERLCAKL-NH2	386
JBT1350	35		D	Ac-FQSKFNVFVDGYTERLCAKL-NH2	649
JBT1351	85		C	Ac-FQSKDNVFVDGYFERLCAKF-NH2	387
JBT1352	85		C	Ac-FQSKMNVFVDGYFERLCAKL-NH2	388
JBT1353	51		D	Ac-FQSVKNVFVDGYFERLCAKL-NH2	650
JBT1354	86		C	Ac-FQSAKNVFVDGYFERLCAKL-NH2	389
JBT1356	49		E	Ac-FQSKENVFVDGYFERLCYKL-NH2	708
JBT1357	52		D	Ac-FQSKGNWFVDGYFERLCAKL-NH2	651
JBT1358	20		D	Ac-FQSKVAVFVDGYFERLCAKL-NH2	652
JBT1359	24			Ac-FQSAYNVFVDGYFERLCAKL-NH2	953
JBT1360	25		D	Ac-FQSCKNVFVDGYFVRLRAKL-NH2	653
JBT1361	75		D	Ac-FQSKTNVFVDGYFERLCAKL-NH2	654
JBT1362	76		D	Ac-FQSKKDVFVDGYFERLCAKL-NH2	655
JBT1363	88		C	Ac-FQSKHVVFVDGYFERLCAKL-NH2	390
JBT1364	64		D	Ac-FQSKSNVFVDGYFERLCARL-NH2	656
JBT1365	-4			Ac-FQSVHNVFVDGYFERPCAKL-NH2	954
JBT1366	21			Ac-FQSKYNVFVDEYFERLCAKL-NH2	955
JBT1367	83		C	Ac-FQSKDNVFVDGYFERLCARL-NH2	391
JBT1368	89		C	Ac-FQSKKHVFVDGYFERLCAKL-NH2	392
JBT1369	40		E	Ac-FQSTKNVFVDGYFERLCAKL-NH2	709
JBT1370	52		D	Ac-FQSDKNVFVDGYFERLCAKL-NH2	657
JBT1371	96		A	Ac-FQSKHHVFVDGYFERLCAKL-NH2	152
JBT1372	83		C	Ac-FQSNKNVFVDGYFERLCAKL-NH2	393

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1373	96		A	Ac-FQSKHNVYVDGYFERLCAKL-NH2	153
JBT1374	71		D	Ac-FQSHKNVFVEGYFERLCAKL-NH2	658
JBT1375	45		D	Ac-FQSKPNVFVDGYFERLCAKL-NH2	659
JBT1376	47		D	Ac-FQSYKDVFVDGYFERLCTKL-NH2	660
JBT1377	101		A	Ac-FQSRRGVFVDGYFERLCAKL-NH2	154
JBT1378	70		D	Ac-FQSKLNVFVDGYFERLCAKL-NH2	661
JBT1379	40		D	Ac-FQSKNNFFVDGYFERLCARL-NH2	662
JBT1380	66		D	Ac-FQSKYNIFVDGYFERLCAKL-NH2	663
JBT1381	106	19	A	Ac-YQSKQNVFVDGYFERLCAKL-NH2	155
JBT1383	103		A	Ac-FQSEHNVFVDGYFERLCAKL-NH2	156
JBT1384	105	23	A	Ac-FQSKNDVFVDGYFERLCAKL-NH2	157
JBT1385	54		D	Ac-HQQFKNVFVDGYFERLCAKL-NH2	664
JBT1386	107	76	B	Ac-FQSGGNVFVDGYFERLCAKL-NH2	246
JBT1387	92		C	Ac-FQSEKNVFVDGYFERLCAKL-NH2	394
JBT1388	90		C	Ac-FQSKKNVFVDGYFERLCFKL-NH2	395
JBT1389	106	23	A	Ac-FQSKRNVFVDGYFERLCAKL-NH2	158
JBT1390	96		A	Ac-FQSPKNVFVDGYFERLCAKL-NH2	159
JBT1391	99		A	Ac-FQSGKNVFVDGYFERLCAKL-NH2	160
JBT1392	68	235	C	Ac-FQSKQNVFVDGYFERLSAKL-NH2	396
JBT1393	67	205	C	Ac-FQSKKNVFVDGYFERLSAKL-NH2	397
JBT1394	88	191	C	Ac-FQSKDNVFVDGYFERLSAKL-NH2	398
JBT1395	100	173	C	Ac-FQSKHNVFVDGYFERLSAKL-NH2	399
JBT1396		29	A	Ac-FQSK-Nmg-NVFVAGYFERLRAKL-NH2	161
JBT1397	5	1569	E	Ac-F-Aib-SKGNVFVDGYFERL-Aib-AKL-NH2	710
JBT1398	5	660	D	Ac-FQ-Aib-KGNVFVDGYFERL-Aib-AKL-NH2	665
JBT1399	109	40	A	Ac-FQS-Aib-GNVFVDGYFERL-Aib-AKL-NH2	162
JBT1400	92	46	A	Ac-FQSK-Aib-NVFVDGYFERL-Aib-AKL-NH2	163

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1401	-7	913	D	Ac-FQSKG-Aib-VFVDGYFERL-Aib-AKL-NH2	666
JBT1402	-8	1144	E	Ac-FQSKGN-Aib-FVDGYFERL-Aib-AKL-NH2	711
JBT1403	-6	>5000		Ac-FQSKGNV-Aib-VDGYFERL-Aib-AKL-NH2	956
JBT1404	13	329	D	Ac-FQSKGNVF-Aib-DGYFERL-Aib-AKL-NH2	667
JBT1405	6	313	D	Ac-FQSKGNVFV-Aib-GYFERL-Aib-AKL-NH2	668
JBT1406	3	610	D	Ac-FQSKGNVFVD-Aib-YFERL-Aib-AKL-NH2	669
JBT1407	6	>5000		Ac-FQSKGNVFVDG-Aib- -Aib-AKL-NH2	957
JBT1408	78	92	B	Ac-FQSKGNVFVDGY-Aib-ERL-Aib-AKL-NH2	247
JBT1409	104	65	B	Ac-FQSKGNVFVDGYF-Aib-RL-Aib-AKL-NH2	248
JBT1410	-3	>5000		Ac-FQSKGNVFVDGYFE-Aib-L-Aib-AKL-NH2	958
JBT1413	-1	>5000		Ac-FQSKGNVFVDGYFERL-Aib-A-Aib-L-NH2	959
JBT1414	86	119	C	Ac-FQSKGNVFVDGYFERL-Aib-AK-Aib-NH2	400
JBT1415	5	3668	F	Ac-Aib-QSKGNVFVDGYFERL-Aib-AKL-NH2	741
JBT1584		9	A	Ac-FQSK-Nmg-NVFVDGYFERL-Aib-AKL-NH2	164
JBT1585		12	A	Ac-FQSK-Nmg-NVFVTGYFERL-Aib-AKL-NH2	165
JBT1586		25	A	AO-FQSKGNVFVDGYFERL-Aib-AKL-NH2	166
JBT1587		45	A	Ac-FQSKGNVFVDGYFERL-Aib-AKLC-NH2	167
JBT1590		106	C	Ac-FQSKGNVFV[CGYFERL-Aib-AKLC]-NH2	401
JBT1591		40	A	Ac-FQSKGNVFVDGYF[CRL-Aib-AKLC]-NH2	168
JBT1592		105	C	Ac-[CFQSKGNVFVDGYFERLC]AKL-NH2	402
JBT1593		462	D	Ac-FQSK[CNVFVDGYFERLC]AKL-NH2	670
JBT1594		952	D	Ac-FQSKGNVFV[CGYFERLC]AKL-NH2	671
JBT1595		249	C	Ac-[CFQSKGNVFVC]GYFERL-Aib-AKL-NH2	403
JBT1596		129	C	Ac-[CFQSKGC]VFVEGYFERL-Aib-AKL-NH2	404
JBT1597		>5000		Ac-DGYFERLRAKL-NH2	960
JBT1598		>5000		Ac-FQSKKNV-NH2	961
JBT1843		35	A	Ac-FQSKGNIFVDGYFERLHAKL-NH2	169

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FIGURE 32

Object ID	% Inhibition (relative to JBT-0477)	IC50 Mean (nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1844		41	A	Ac-FQSKNNVFVDGYFKRLRAKL-NH2	170
JBT1845		89	B	Ac-FQSYKHVFVDGYFERLRAKL-NH2	249
JBT1846		54	B	Ac-FQSKGIVFVDGYFKRLRAKL-NH2	250
JBT1847		30	A	Ac-YQTKGNVFVDGYFERLRAKL-NH2	171
JBT1848		112	C	Ac-FQSKYNVFVDGYFERLFAKL-NH2	405
JBT1849		44	A	Ac-FQTKDNVHVDGYFERLRAKL-NH2	172
JBT1850		56	B	Ac-FQSYKRVFVDGYFERLRAKL-NH2	251
JBT1851		49	A	Ac-LQQKGNVFVDGYFERLRAKL-NH2	173
JBT1852		19	A	PEG-FQSKGNVFVDGYFERL-Aib-AKL-NH2	174
JBT1853		42	A	PEG-FQSKGNVFVDGYFERL-Aib-AKL-NH2	175
JBT1854		16	A	Ac-FQSKGNVFVDGYFERL-Aib-AKLC(PEG)-NH2	176
JBT1855		96	B	Ac-FQSKGNVFVDGYFERL-Aib-AKLC(PEG)-NH2	252
JBT1856		17	A	Ac-FQSKpNVFVDGYFERL-Aib-AKL-NH2	177
JBT1857		14	A	Ac-FQSKpNVHVDGYFERL-Aib-AKL-NH2	178
JBT2266		20	A	Ac-PFQSKGNVFVDGYFERLRAKL-NH2	179
JBT2267		19	A	Ac-PEFQSKGNVFVDGYFERLRAKL-NH2	180
JBT2268		18	A	Ac-PFQSK-Nme-NVFVDGYFERL-Aib-AKL-NH2	181

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FIGURE 33

Object ID	EC50 (nM]	Sequence	SEQ ID NO:
JBT0051	1.0	Biotinyl-Ttds-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-NH2	962
JBT0055	1.7	Ac-SGVGRLQVAFQSKKNVFVFGYFERLRAKLTS-Ttds-Lysin(biotin)-NH2	963
JBT0131	16.8	Biotinyl-Ttds-AFQSKKNVFVFGYFERLRAK-NH2	964
JBT0132	2.2	Biotinyl-Ttds-FQSKKNVFVFGYFERLRAKL-NH2	965
JBT0133	>25000	Biotinyl-Ttds-Ttds-QSKKNVFVFGYFERLRAKLT-NH2	966
JBT0134	>1000	Biotinyl-Ttds-QSKKNVFVFGYFERLRAK-NH2	967
JBT0166	0.9	Biotinyl-Ttds-KKFQSKKNVFVFGYFERLRAKLKK-NH2	968
JBT0167*	574.8	Biotinyl-Ttds-GKNAKFYLFESLRQVKFVFR-NH2	969
JBT0168*	152.3	Biotinyl-Ttds-YKFSFNKELFKQARLRFVGV-NH2	970
JBT0173	>5000	Biotinyl-Ttds-SGVGRLQVAFQSKKNVFVFG-NH2	971
JBT0403	1.4	Biotinyl-Ttds-FQSKKNVFVDGYFERLRAKL-NH2	972

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FIGURE 34

Object ID	IC50 [nm]	Affinity Class	Sequence	SEQ ID NO:
JBT0120	59	B	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2	1047
JBT0247	>206		Ac-SGASRYKWFCGMRDMKGTMSCVWVKFRYDTS-NH2	1213
JBT0248	30	A	Ac-KKSGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTSKK-NH2	1001
JBT0249	>50000		Ac-KKSGASRYKWFCGMRDMKGTMSKK-NH2	1214
JBT0250	8766	F	Ac-KKSRYKWFCGMRDMKGTMSCVWKK-NH2	1201
JBT0251	5207	F	Ac-KKKWFCGMRDMKGTMSCVWVKFKK-NH2	1202
JBT0252	>50000		Ac-KKCGMRDMKGTMSCVWVKFRYDKK-NH2	1215
JBT0253	>207		Ac-KKMRDMKGTMSCVWVKFRYDTSKK-NH2	1216
JBT0319	32	A	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRY-NH2	1002
JBT0319	109	C	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVKFRY-NH2	1075
JBT0320	977	D	Ac-SGASRYKWF[CGMRDMKGTMSC]VWVK-NH2	1103
JBT0321	>50000		Ac-SGASRYKWF[CGMRDMKGTMSC]V-NH2	1217
JBT0322	>50000		Ac-SGASRYKWFCGMRDMKGTM-NH2	1218
JBT0323	118	C	Ac-SRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2	1076
JBT0324	1931	E	Ac-KWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2	1155
JBT0325	>50000		Ac-[CGMRDMKGTMSC]VWVKFRYDTS-NH2	1219
JBT0326	>10000		Ac-RDMKGTMSCVWVKFRYDTS-NH2	1220
JBT0326	>50000		Ac-RDMKGTMSCVWVKFRYDTS-NH2	1221
JBT0327	>50000		Ac-SGASRYKWFCGMRDMKGTMS-NH2	1222
JBT0328	>50000		Ac-SRYKWF[CGMRDMKGTMSC]VW-NH2	1223
JBT0329	4574	E	Ac-KWF[CGMRDMKGTMSC]VWVKF-NH2	1156
JBT0329	9396	F	Ac-KWF[CGMRDMKGTMSC]VWVKF-NH2	1203
JBT0330	>50000		Ac-[CGMRDMKGTMSC]VWVKFRYD-NH2	1224
JBT0331	>10001		Ac-MRDMKGTMSCVWVKFRYDTS-NH2	1225
JBT0331	>50000		Ac-MRDMKGTMSCVWVKFRYDTS-NH2	1226
JBT0332	29746	G	Ac-SRYKWFCGMRDMKGTMSCVW-NH2	1206
JBT0333	18840	G	Ac-KWFCGMRDMKGTMSCVWVKF-NH2	1207
JBT0334	>10002		Ac-CGMRDMKGTMSCVWVKFRYD-NH2	1227
JBT0409	6413	F	Ac-SGASRYKWFSGMRDMKGTMSSVWVKFRYDTS-NH2	1204

FIGURE 34

Object ID	IC50 [nm]	Affinity Class	Sequence	SEQ ID NO:
JBT0410	24801	G	Ac-SGASRYKWFSG-Nle-RD-Nle-KGT-Nle-SSVWVKFRYDTS-NH2	1208
JBT0411	685	D	Ac-SGASRYKWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKFRYDTS-NH2	1104
JBT0412	16103	G	Ac-KWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKF-NH2	1209
JBT0413	>50000		Ac-KWFSGMRDMKGTMSSVWVKF-NH2	1228
JBT0414	5693	F	Ac-KWFSG-Nle-RD-Nle-KGT-Nle-SSVWVKF-NH2	1205
JBT0415	50	A	Ac-SRYKWF[CGMRDMKGTMSC]VWVKF-NH2	1003
JBT0416	187	C	Ac-SRYKWF[CG-Nle-RD-Nle-KGT-Nle-SC]VWVKF-NH2	1077
JBT0417	>50000		Ac-SRYKWFSGMRDMKGTMSSVWVKF-NH2	1229
JBT0418	43941	G	Ac-SRYKWF SG-Nle-RD-Nle-KGT-Nle-SS VWVKF-NH2	1210
JBT0435*	31732	G	Ac-RKRDVSGKM[CWSGFGSKWFRC]ADMMTYYSVT-NH2	1211
JBT0436*	45925	G	Ac-RWSGKSTYS[CDAMVRRMGMKC]FSFGWTDVYK-NH2	1212
JBT0437	60	B	Ac-ARYKWF[CGMRDMKGTMSC]VWVKF-NH2	1048
JBT0438	194	C	Ac-SAYKWF[CGMRDMKGTMSC]VWVKF-NH2	1078
JBT0439	291	D	Ac-SRAKWF[CGMRDMKGTMSC]VWVKF-NH2	1105
JBT0440	393	D	Ac-SRYAWF[CGMRDMKGTMSC]VWVKF-NH2	1106
JBT0441	>5000		Ac-SRYKAF[CGMRDMKGTMSC]VWVKF-NH2	1230
JBT0442	>5000		Ac-SRYKWA[CGMRDMKGTMSC]VWVKF-NH2	1231
JBT0443	2861	E	Ac-SRYKWF AGMRDMKGTMSC VWVKF-NH2	1157
JBT0444	18	A	Ac-SRYKWF[CAMRDMKGTMSC]VWVKF-NH2	1004
JBT0444	31	A	Ac-SRYKWF[CAMRDMKGTMSC]VWVKF-NH2	1005
JBT0445	760	D	Ac-SRYKWF[CGARDMKGTMSC]VWVKF-NH2	1107
JBT0446	152	C	Ac-SRYKWF[CGMADMKGTMSC]VWVKF-NH2	1079
JBT0447	3249	E	Ac-SRYKWF[CGMRAMKGTMSC]VWVKF-NH2	1158
JBT0448	315	D	Ac-SRYKWF[CGMRDAKGTMSC]VWVKF-NH2	1108
JBT0449	104	C	Ac-SRYKWF[CGMRDMAGTMSC]VWVKF-NH2	1080
JBT0450	1061	E	Ac-SRYKWF[CGMRDMKATMSC]VWVKF-NH2	1159
JBT0451	131	C	Ac-SRYKWF[CGMRDMKGAMSC]VWVKF-NH2	1081
JBT0452	313	D	Ac-SRYKWF[CGMRDMKGTASC]VWVKF-NH2	1109
JBT0453	343	D	Ac-SRYKWF[CGMRDMKGTMAC]VWVKF-NH2	1110
JBT0454	4963	E	Ac-SRYKWF CGMRDMKGTMSA VWVKF-NH2	1160
JBT0455	99	B	Ac-SRYKWF[CGMRDMKGTMSC]AWVKF-NH2	1049
JBT0456	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VAVKF-NH2	1232

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FIGURE 34

Object ID	IC50 [nm]	Affinity Class	Sequence	SEQ ID NO:
JBT0457	2896	E	Ac-SRYKWF[CGMRDMKGTMSC]VWAKF-NH2	1161
JBT0458	148	C	Ac-SRYKWF[CGMRDMKGTMSC]VWVAF-NH2	1082
JBT0459	150	C	Ac-SRYKWF[CGMRDMKGTMSC]VWVKA-NH2	1083
JBT0460	88	B	Ac-RYKWF[CGMRDMKGTMSC]VWVKF-NH2	1050
JBT0461	503	D	Ac-YKWF[CGMRDMKGTMSC]VWVKF-NH2	1111
JBT0462	>5000		Ac-SRYKWFGM[CRDMKGTMSC]VWVKF-NH2	1233
JBT0463	>5000		Ac-SRYKWFGMRD[CMKGTMSC]VWVKF-NH2	1234
JBT0464	>5000		Ac-SRYKWFGMRDMK[CGTMSC]VWVKF-NH2	1235
JBT0465	>5000		Ac-SRYKWF[CGMRDMKGTC]MSVWVKF-NH2	1236
JBT0466	>5000		Ac-SRYKWF[CGMRDMKC]GTMSVWVKF-NH2	1237
JBT0467	>5000		Ac-SRYKWF[CGMRDC]MKGTMSVWVKF-NH2	1238
JBT0468	>5000		Ac-SRYKWFG[CMRDMKGTMC]SVWVKF-NH2	1239
JBT0469	>5000		Ac-SRYKWFGM[CRDMKGTC]MSVWVKF-NH2	1240
JBT0470	1472	E	Ac-SRYKWFGMR[CDMKGC]TMSVWVKF-NH2	1162
JBT0617	489	D	Ac-SRYKWF[homoC-GMRDMKGTMSC]VWVKF-NH2	1112
JBT0618	2073	E	Ac-SRYKWF[CGMRDMKGTMS-homoC]VWVKF-NH2	1163
JBT0619	>5000		Ac-SRYKWF[homoC-GMRDMKGTMS-homoC]VWVKF-NH2	1241
JBT0620	>5000		Ac-SRYKWF[Dap-GMRDMKGTMS-D]VWVKF-NH2	1242
JBT0623	>5000		Ac-SRYKWF[K-GMRDMKGTMS-D]VWVKF-NH2	1243
JBT0625	>5000		Ac-SRYKWF[CMRDMKGTMSC]VWVKF-NH2	1244
JBT0626	1078	E	Ac-SRYKWF[CGMRDKGTMSC]VWVKF-NH2	1164
JBT0627	2169	E	Ac-SRYKWF[CGMRDMGTMSC]VWVKF-NH2	1165
JBT0628	3611	E	Ac-SRYKWF[CGMRDMKGMSC]VWVKF-NH2	1166
JBT0629	4076	E	Ac-SRYKWF[cGMRDMKGTMSC]VWVKF-NH2	1167
JBT0631	343	D	Ac-SRYKWF[CGmRDMKGTMSC]VWVKF-NH2	1113
JBT0632	>5000		Ac-SRYKWF[CGMrDMKGTMSC]VWVKF-NH2	1245
JBT0633	2131	E	Ac-SRYKWF[CGMRdMKGTMSC]VWVKF-NH2	1168
JBT0634	61	B	Ac-SRYKWF[CGMRDmKGTMSC]VWVKF-NH2	1051
JBT0635	433	D	Ac-SRYKWF[CGMRDMkGTMSC]VWVKF-NH2	1114
JBT0637	3590	E	Ac-SRYKWF[CGMRDMKGtMSC]VWVKF-NH2	1169
JBT0638	>5000		Ac-SRYKWF[CGMRDMKGTmSC]VWVKF-NH2	1246
JBT0639	1220	E	Ac-SRYKWF[CGMRDMKGTMsC]VWVKF-NH2	1170

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FIGURE 34

Object ID	IC50 [nm]	Affinity Class	Sequence	SEQ ID NO:
JBT0640	3470	E	Ac-SRYKWF[CGMRDMKGTMSc]VWVKF-NH2	1171
JBT0641	>5000		Ac-SRYKWF[CPMRDMKGTMSC]VWVKF-NH2	1247
JBT0642	>5000		Ac-SRYKWF[CGPRDMKGTMSC]VWVKF-NH2	1248
JBT0643	23	A	Ac-SRYKWF[CGMPDMKGTMSC]VWVKF-NH2	1006
JBT0644	2873	E	Ac-SRYKWF[CGMRPMKGTMSC]VWVKF-NH2	1172
JBT0645	165	C	Ac-SRYKWF[CGMRDPKGTMSC]VWVKF-NH2	1084
JBT0646	3169	E	Ac-SRYKWF[CGMRDMPGTMSC]VWVKF-NH2	1173
JBT0647	>5000		Ac-SRYKWF[CGMRDMKPTMSC]VWVKF-NH2	1249
JBT0648	3682	E	Ac-SRYKWF[CGMRDMKGPMSC]VWVKF-NH2	1174
JBT0649	2989	E	Ac-SRYKWF[CGMRDMKGTPSC]VWVKF-NH2	1175
JBT0650	>5000		Ac-SRYKWF[CGMRDMKGTMPC]VWVKF-NH2	1250
JBT0790	255	D	H-GSSRYKWF[CGMRDMKGTMSC]VWVKF-OH	1115
JBT1416	39	A	Ac-DRYKWF[CGMRDMKGTMSC]VWVKF-NH2	1007
JBT1417	344	D	Ac-SDYKWF[CGMRDMKGTMSC]VWVKF-NH2	1116
JBT1418	1469	E	Ac-SRDKWF[CGMRDMKGTMSC]VWVKF-NH2	1176
JBT1419	536	D	Ac-SRYDWF[CGMRDMKGTMSC]VWVKF-NH2	1117
JBT1420	>5000		Ac-SRYKDF[CGMRDMKGTMSC]VWVKF-NH2	1251
JBT1421	>5000		Ac-SRYKWD[CGMRDMKGTMSC]VWVKF-NH2	1252
JBT1422	>5000		Ac-SRYKWF[CDMRDMKGTMSC]VWVKF-NH2	1253
JBT1423	4671	E	Ac-SRYKWF[CGDRDMKGTMSC]VWVKF-NH2	1177
JBT1424	>5000		Ac-SRYKWF[CGMDDMKGTMSC]VWVKF-NH2	1254
JBT1425	101	C	Ac-SRYKWF[CGMRDDKGTMSC]VWVKF-NH2	1085
JBT1426	17	A	Ac-SRYKWF[CGMRDMDGTMSC]VWVKF-NH2	1008
JBT1427	87	B	Ac-SRYKWF[CGMRDMKDTMSC]VWVKF-NH2	1052
JBT1428	478	D	Ac-SRYKWF[CGMRDMKGDMSC]VWVKF-NH2	1118
JBT1429	107	C	Ac-SRYKWF[CGMRDMKGTDSC]VWVKF-NH2	1086
JBT1430	265	D	Ac-SRYKWF[CGMRDMKGTMDC]VWVKF-NH2	1119
JBT1431	>5000		Ac-SRYKWF[CGMRDMKGTMSC]DWVKF-NH2	1255
JBT1432	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VDVKF-NH2	1256
JBT1433	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VWDKF-NH2	1257
JBT1434	515	D	Ac-SRYKWF[CGMRDMKGTMSC]VWVDF-NH2	1120
JBT1435	34	A	Ac-SRYKWF[CGMRDMKGTMSC]VWVKD-NH2	1009

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FIGURE 34

Object ID	IC50 [nm]	Affinity Class	Sequence	SEQ ID NO:
JBT1436	54	B	Ac-FRYKWF[CGMRDMKGTMSC]VWVKF-NH2	1053
JBT1437	33	A	Ac-SFYKWF[CGMRDMKGTMSC]VWVKF-NH2	1010
JBT1438	40	A	Ac-SRFKWF[CGMRDMKGTMSC]VWVKF-NH2	1011
JBT1439	572	D	Ac-SRYFWF[CGMRDMKGTMSC]VWVKF-NH2	1121
JBT1440	>5000		Ac-SRYKFF[CGMRDMKGTMSC]VWVKF-NH2	1258
JBT1441	>5000		Ac-SRYKWF[CFMRDMKGTMSC]VWVKF-NH2	1259
JBT1442	2701	E	Ac-SRYKWF[CGFRDMKGTMSC]VWVKF-NH2	1178
JBT1443	1218	E	Ac-SRYKWF[CGMFDMKGTMSC]VWVKF-NH2	1179
JBT1444	>5000		Ac-SRYKWF[CGMRFMKGTMSC]VWVKF-NH2	1260
JBT1445	43	A	Ac-SRYKWF[CGMRDFKGTMSC]VWVKF-NH2	1012
JBT1446	69	B	Ac-SRYKWF[CGMRDMFGTMSC]VWVKF-NH2	1054
JBT1447	344	D	Ac-SRYKWF[CGMRDMKFTMSC]VWVKF-NH2	1122
JBT1448	296	D	Ac-SRYKWF[CGMRDMKGFMSC]VWVKF-NH2	1123
JBT1449	23	A	Ac-SRYKWF[CGMRDMKGTFSC]VWVKF-NH2	1013
JBT1450	475	D	Ac-SRYKWF[CGMRDMKGTMFC]VWVKF-NH2	1124
JBT1451	236	C	Ac-SRYKWF[CGMRDMKGTMSC]FWVKF-NH2	1087
JBT1452	546	D	Ac-SRYKWF[CGMRDMKGTMSC]VFVKF-NH2	1125
JBT1453	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VWFKF-NH2	1261
JBT1454	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VWVFF-NH2	1262
JBT1455	39	A	Ac-GRYKWF[CGMRDMKGTMSC]VWVKF-NH2	1014
JBT1456	265	D	Ac-SGYKWF[CGMRDMKGTMSC]VWVKF-NH2	1126
JBT1457	1866	E	Ac-SRGKWF[CGMRDMKGTMSC]VWVKF-NH2	1180
JBT1458	430	D	Ac-SRYGWF[CGMRDMKGTMSC]VWVKF-NH2	1127
JBT1459	>5000		Ac-SRYKGF[CGMRDMKGTMSC]VWVKF-NH2	1263
JBT1460	>5000		Ac-SRYKWG[CGMRDMKGTMSC]VWVKF-NH2	1264
JBT1461	2099	E	Ac-SRYKWF[CGGRDMKGTMSC]VWVKF-NH2	1181
JBT1462	370	D	Ac-SRYKWF[CGMGDMKGTMSC]VWVKF-NH2	1128
JBT1463	682	D	Ac-SRYKWF[CGMRGMKGTMSC]VWVKF-NH2	1129
JBT1464	150	C	Ac-SRYKWF[CGMRDGKGTMSC]VWVKF-NH2	1088
JBT1465	50	A	Ac-SRYKWF[CGMRDMGGTMSC]VWVKF-NH2	1015
JBT1466	85	B	Ac-SRYKWF[CGMRDMKGGMSC]VWVKF-NH2	1055
JBT1467	290	D	Ac-SRYKWF[CGMRDMKGTGSC]VWVKF-NH2	1130

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FIGURE 34

Object ID	IC50 [nm]	Affinity Class	Sequence	SEQ ID NO:
JBT1468	778	D	Ac-SRYKWF[CGMRDMKGTMGC]VWVKF-NH2	1131
JBT1469	2366	E	Ac-SRYKWF[CGMRDMKGTMSC]GWVKF-NH2	1182
JBT1470	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VGVKF-NH2	1265
JBT1471	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VWGKF-NH2	1266
JBT1472	158	C	Ac-SRYKWF[CGMRDMKGTMSC]VWVGF-NH2	1089
JBT1473	176	C	Ac-SRYKWF[CGMRDMKGTMSC]VWVKG-NH2	1090
JBT1474	31	A	Ac-KRYKWF[CGMRDMKGTMSC]VWVKF-NH2	1016
JBT1475	69	B	Ac-SKYKWF[CGMRDMKGTMSC]VWVKF-NH2	1056
JBT1476	282	D	Ac-SRKKWF[CGMRDMKGTMSC]VWVKF-NH2	1132
JBT1477	>5000		Ac-SRYKKF[CGMRDMKGTMSC]VWVKF-NH2	1267
JBT1478	162	C	Ac-SRYKWK[CGMRDMKGTMSC]VWVKF-NH2	1091
JBT1479	1110	E	Ac-SRYKWF[CKMRDMKGTMSC]VWVKF-NH2	1183
JBT1480	392	D	Ac-SRYKWF[CGKRDMKGTMSC]VWVKF-NH2	1133
JBT1481	20	A	Ac-SRYKWF[CGMKDMKGTMSC]VWVKF-NH2	1017
JBT1482	1354	E	Ac-SRYKWF[CGMRKMKGTMSC]VWVKF-NH2	1184
JBT1483	59	B	Ac-SRYKWF[CGMRDKKGTMSC]VWVKF-NH2	1057
JBT1484	520	D	Ac-SRYKWF[CGMRDMKKTMSC]VWVKF-NH2	1134
JBT1485	67	B	Ac-SRYKWF[CGMRDMKGKMSC]VWVKF-NH2	1058
JBT1486	54	B	Ac-SRYKWF[CGMRDMKGTKSC]VWVKF-NH2	1059
JBT1487	434	D	Ac-SRYKWF[CGMRDMKGTMKC]VWVKF-NH2	1135
JBT1488	>5000		Ac-SRYKWF[CGMRDMKGTMSC]KWVKF-NH2	1268
JBT1489	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VKVKF-NH2	1269
JBT1490	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VWKKF-NH2	1270
JBT1491	335	D	Ac-SRYKWF[CGMRDMKGTMSC]VWVKK-NH2	1136
JBT1492	38	A	Ac-LRYKWF[CGMRDMKGTMSC]VWVKF-NH2	1018
JBT1493	51	B	Ac-SLYKWF[CGMRDMKGTMSC]VWVKF-NH2	1060
JBT1494	679	D	Ac-SRLKWF[CGMRDMKGTMSC]VWVKF-NH2	1137
JBT1495	729	D	Ac-SRYLWF[CGMRDMKGTMSC]VWVKF-NH2	1138
JBT1496	>5000		Ac-SRYKLF[CGMRDMKGTMSC]VWVKF-NH2	1271
JBT1497	72	B	Ac-SRYKWL[CGMRDMKGTMSC]VWVKF-NH2	1061
JBT1498	>5000		Ac-SRYKWF[CLMRDMKGTMSC]VWVKF-NH2	1272
JBT1499	636	D	Ac-SRYKWF[CGLRDMKGTMSC]VWVKF-NH2	1139

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FIGURE 34

Object ID	IC50 [nm]	Affinity Class	Sequence	SEQ ID NO:
JBT1500	68	B	Ac-SRYKWF[CGMLDMKGTMSC]VWVKF-NH2	1062
JBT1501	3449	E	Ac-SRYKWF[CGMRLMKGTMSC]VWVKF-NH2	1185
JBT1502	38	A	Ac-SRYKWF[CGMRDLKGTMSC]VWVKF-NH2	1019
JBT1503	54	B	Ac-SRYKWF[CGMRDMLGTMSC]VWVKF-NH2	1063
JBT1504	757	D	Ac-SRYKWF[CGMRDMKLTMSC]VWVKF-NH2	1140
JBT1505	155	C	Ac-SRYKWF[CGMRDMKGLMSC]VWVKF-NH2	1092
JBT1506	57	B	Ac-SRYKWF[CGMRDMKGTLSC]VWVKF-NH2	1064
JBT1507	411	D	Ac-SRYKWF[CGMRDMKGTMLC]VWVKF-NH2	10141
JBT1508	216	C	Ac-SRYKWF[CGMRDMKGTMSC]LWVKF-NH2	1093
JBT1509	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VLVKF-NH2	1273
JBT1510	221	C	Ac-SRYKWF[CGMRDMKGTMSC]VWLKF-NH2	1094
JBT1511	330	D	Ac-SRYKWF[CGMRDMKGTMSC]VWVLF-NH2	1142
JBT1512	18	A	Ac-SRYKWF[CGMRDMKGTMSC]VWVKL-NH2	1020
JBT1513	56	B	Ac-SSYKWF[CGMRDMKGTMSC]VWVKF-NH2	1065
JBT1514	92	B	Ac-SRSKWF[CGMRDMKGTMSC]VWVKF-NH2	1066
JBT1515	411	D	Ac-SRYSWF[CGMRDMKGTMSC]VWVKF-NH2	1143
JBT1516	>5000		Ac-SRYKSF[CGMRDMKGTMSC]VWVKF-NH2	1274
JBT1517	>5000		Ac-SRYKWS[CGMRDMKGTMSC]VWVKF-NH2	1275
JBT1518	50	A	Ac-SRYKWF[CSMRDMKGTMSC]VWVKF-NH2	1021
JBT1519	1825	E	Ac-SRYKWF[CGSRDMKGTMSC]VWVKF-NH2	1186
JBT1520	202	C	Ac-SRYKWF[CGMSDMKGTMSC]VWVKF-NH2	1095
JBT1521	226	C	Ac-SRYKWF[CGMRSMKGTMSC]VWVKF-NH2	1096
JBT1522	92	B	Ac-SRYKWF[CGMRDSKGTMSC]VWVKF-NH2	1067
JBT1523	35	A	Ac-SRYKWF[CGMRDMSGTMSC]VWVKF-NH2	1022
JBT1524	512	D	Ac-SRYKWF[CGMRDMKSTMSC]VWVKF-NH2	1144
JBT1525	56	B	Ac-SRYKWF[CGMRDMKGSMSC]VWVKF-NH2	1068
JBT1526	80	B	Ac-SRYKWF[CGMRDMKGTSSC]VWVKF-NH2	1069
JBT1527	190	C	Ac-SRYKWF[CGMRDMKGTMSC]SWVKF-NH2	1097
JBT1528	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VSVKF-NH2	1276
JBT1529	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VWSKF-NH2	1277
JBT1530	83	B	Ac-SRYKWF[CGMRDMKGTMSC]VWVSF-NH2	1070
JBT1531	63	B	Ac-SRYKWF[CGMRDMKGTMSC]VWVKS-NH2	1071

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FIGURE 34

Object ID	IC50 [nm]	Affinity Class	Sequence	SEQ ID NO:
JBT1532	54	B	Ac-PRYKWF[CGMRDMKGTMSC]VWVKF-NH2	1072
JBT1533	263	D	Ac-SPYKWF[CGMRDMKGTMSC]VWVKF-NH2	1145
JBT1534	2398	E	Ac-SRPKWF[CGMRDMKGTMSC]VWVKF-NH2	1187
JBT1535	>5000		Ac-SRYPWF[CGMRDMKGTMSC]VWVKF-NH2	1278
JBT1536	>5000		Ac-SRYKPF[CGMRDMKGTMSC]VWVKF-NH2	1279
JBT1537	>5000		Ac-SRYKWP[CGMRDMKGTMSC]VWVKF-NH2	1280
JBT1538	>5000		Ac-SRYKWF[CGMRDMKGTMSC]PWVKF-NH2	1281
JBT1539	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VPVKF-NH2	1282
JBT1540	>5000		Ac-SRYKWF[CGMRDMKGTMSC]VWPKF-NH2	1283
JBT1541	29	A	Ac-SRYKWF[CGMRDMKGTMSC]VWVPF-NH2	1023
JBT1542	1052	E	Ac-SRYKWF[CGMRDMKGTMSC]VWVKP-NH2	1188
JBT1543	202	C	Ac-SRYKWF[CGMRDMKGTMSC]VWVKF-OH	1098
JBT1544	40	A	H-SRYKWF[CGMRDMKGTMSC]VWVKF-NH2	1024
JBT1545	239	C	H-SRYKWF[CGMRDMKGTMSC]VWVKF-OH	1099
JBT1546	>5000		Ac-SRYKWF[CEMRDMKGTMSC]VWVKF-NH2	1284
JBT1547	>5000		Ac-SRYKWF[CHMRDMKGTMSC]VWVKF-NH2	1285
JBT1548	>5000		Ac-SRYKWF[CIMRDMKGTMSC]VWVKF-NH2	1286
JBT1549	1736	E	Ac-SRYKWF[CMMRDMKGTMSC]VWVKF-NH2	1189
JBT1550	1086	E	Ac-SRYKWF[CNMRDMKGTMSC]VWVKF-NH2	1190
JBT1551	682	D	Ac-SRYKWF[CQMRDMKGTMSC]VWVKF-NH2	1146
JBT1552	400	D	Ac-SRYKWF[CRMRDMKGTMSC]VWVKF-NH2	1147
JBT1553	292	D	Ac-SRYKWF[CTMRDMKGTMSC]VWVKF-NH2	1148
JBT1554	>5000		Ac-SRYKWF[CVMRDMKGTMSC]VWVKF-NH2	1287
JBT1555	3382	E	Ac-SRYKWF[CWMRDMKGTMSC]VWVKF-NH2	1191
JBT1556	>5000		Ac-SRYKWF[CYMRDMKGTMSC]VWVKF-NH2	1288
JBT1557	60	B	H-RYKWF[CGMRDMKGTMSC]VWVKF-NH2	173
JBT1558	975	D	H-YKWF[CGMRDMKGTMSC]VWVKF-NH2	1149
JBT1559	286	D	Ac-SRYKWF[CGaRDMKGTMSC]VWVKF-NH2	1150
JBT1560	>5000		Ac-SRYKWF[CGdRDMKGTMSC]VWVKF-NH2	1289
JBT1561	3031	E	Ac-SRYKWF[CGfRDMKGTMSC]VWVKF-NH2	1192
JBT1562	1783	E	Ac-SRYKWF[CGkRDMKGTMSC]VWVKF-NH2	1193
JBT1563	1821	E	Ac-SRYKWF[CGlRDMKGTMSC]VWVKF-NH2	1194

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FIGURE 34

Object ID	IC50 [nm]	Affinity Class	Sequence	SEQ ID NO:
JBT1564	170	C	Ac-SRYKWF[CGpRDMKGTMSC]VWVKF-NH2	1100
JBT1565	1712	E	Ac-SRYKWF[CGsRDMKGTMSC]VWVKF-NH2	1195
JBT1566	367	D	Ac-SRYKWF[CGMRDaKGTMSC]VWVKF-NH2	1151
JBT1567	212	C	Ac-SRYKWF[CGMRDdKGTMSC]VWVKF-NH2	1101
JBT1568	73	B	Ac-SRYKWF[CGMRDfKGTMSC]VWVKF-NH2	1074
JBT1569	155	C	Ac-SRYKWF[CGMRDkKGTMSC]VWVKF-NH2	1102
JBT1570	47	A	Ac-SRYKWF[CGMRDlKGTMSC]VWVKF-NH2	1025
JBT1571	1320	E	Ac-SRYKWF[CGMRDpKGTMSC]VWVKF-NH2	1196
JBT1572	650	D	Ac-SRYKWF[CGMRDsKGTMSC]VWVKF-NH2	1152
JBT1573	1154	E	Ac-SRYKWF[CGMRDMaGTMSC]VWVKF-NH2	1197
JBT1574	350	D	Ac-SRYKWF[CGMRDMdGTMSC]VWVKF-NH2	1153
JBT1575	2377	E	Ac-SRYKWF[CGMRDMfGTMSC]VWVKF-NH2	1198
JBT1576	1004	E	Ac-SRYKWF[CGMRDMlGTMSC]VWVKF-NH2	1199
JBT1577	1121	E	Ac-SRYKWF[CGMRDMpGTMSC]VWVKF-NH2	1200
JBT1578	633	D	Ac-SRYKWF[CGMRDMsGTMSC]VWVKF-NH2	1154
JBT1735	9	A	Ac-SMYKWH[CGMRDMKGTYSC]VWVKF-NH2	1026
JBT1772	11	A	Ac-FHYKWH[CGMRDMKGTYSC]VWVKF-NH2	1027
JBT1808	12	A	Ac-SYYKWH[CGMRDMKGIMSC]AWVKF-NH2	1028
JBT1811	11	A	Ac-SYYKWH[CGMRDMKGIMSC]VWVKY-NH2	1029
JBT1812	11	A	Ac-SYYKWH[CGMRDMKGIDSC]VWVRF-NH2	1030
JBT1813	9	A	Ac-SYYKWH[CGMRDMKGIMSC]VWVKA-NH2	1031
JBT1815	10	A	Ac-SYYKWH[CGMRDMKGTMTC]VWVKF-NH2	1032
JBT1816	9	A	Ac-SYYKWH[CGMRDMKGTMSC]VWVKS-NH2	1033
JBT1817	9	A	Ac-SHYKWH[CAMRDMKGTYSC]VWVKF-NH2	1034
JBT1821	11	A	Ac-SYYKWH[CGVRDMKGTMSC]VWVKS-NH2	1035
JBT1822	18	A	Ac-GHYKWH[CGMRDMKGTFSC]VWVF-NH2	1036
JBT1825	11	A	Ac-AYYKWH[CGMRDLKGTMSC]VWVKS-NH2	1037
JBT1826	11	A	Ac-SYYKWH[CGMRDMKGTYSC]VWVKM-NH2	1038
JBT1828	22	A	Ac-SHYKWH[CGMRDMKGIMSC]VWVF-NH2	1039
JBT1829	8	A	Ac-FYYKWH[CGMRDMKGTMSC]AWVKF-NH2	1040
JBT1830	10	A	Ac-SYYKWH[CGMRDMKGTDSC]VWVWY-NH2	1041
JBT1831	13	A	Ac-SHYKWH[CGMRDMKGTMSC]AWVKF-NH2	1042

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FIGURE 34

Object ID	IC50 [nm]	Affinity Class	Sequence	SEQ ID NO:
JBT1832	11	A	Ac-HYYKWH[CGMRDMKGTMSC]VWVKS-NH2	1043
JBT1837	10	A	Ac-SYYKWH[CAMRDMKGTMTC]VWVKF-NH2	1044
JBT1840	11	A	Ac-SYYKWH[CGMRDMKGTMSC]VWVLF-NH2	1045
JBT1842	9	A	Ac-SHYKWH[CAMRDMKGTMSC]AWVKF-NH2	1046

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FIGURE 35

Object ID	EC50(nM)	Sequence	SEQ ID NO:
JBT0124	1.8	Biotinyl-Ttds-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH2	1290
JBT0659	2.1	Biotinyl-Ttds-SRYKWF[CGMRDMKGTMSC]VWVKF-NH2	1291

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0122	3084	E	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2	2002
JBT0221	1412	E	Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKTSKK-NH2	2003
JBT0222*	36272	G	Ac-MKSRSLGLAYFAKHSSLEVLQTRKVAAPYY-NH2	2127
JBT0223*	>50000		Ac-KMQLRVYASTAHSRYLLGSSLFPKYAEVKA-NH2	2297
JBT0224	>50000		Ac-KKSGYASFPLAVQLHVSKRSKEMALARLYYKK-NH2	2298
JBT0225	25395	G	Ac-KKSGYASFPLAVQLHVSKRSKEMALARKK-NH2	2128
JBT0226	>50000		Ac-KKSGYASFPLAVQLHVSKRSKEMAKK-NH2	2299
JBT0227	>50000		Ac-KKSGYASFPLAVQLHVSKRSKKK-NH2	2300
JBT0228	5436	F	Ac-KKASFPLAVQLHVSKRSKEMALARLYYKTSKK-NH2	2016
JBT0229	16971	G	Ac-KKPLAVQLHVSKRSKEMALARLYYKTSKK-NH2	2129
JBT0230	>50000		Ac-KKVQLHVSKRSKEMALARLYYKTSKK-NH2	2301
JBT0231	>50000		Ac-KKHVSKRSKEMALARLYYKTSKK-NH2	2302
JBT0232	>50000		Ac-KKGYASFPLAVQLHVSKRSKEMKK-NH2	2303
JBT0233	>50000		Ac-KKYASFPLAVQLHVSKRSKEMAKK-NH2	2304
JBT0234	>50000		Ac-KKVQLHVSKRSKEMALARLYYKKK-NH2	2305
JBT0235	>50000		Ac-KKQLHVSKRSKEMALARLYYKTKK-NH2	2306
JBT0236	>50000		Ac-KKLHVSKRSKEMALARLYYKTSKK-NH2	2307
JBT0237	>50000		Ac-KKSGYASFPLAVQLHVSKRSKEKK-NH2	2308
JBT0359	>50000		Ac-ASFPLAVQLHVSKRSKEMA-NH2	2309
JBT0360	>50000		Ac-SFPLAVQLHVSKRSKEMA-NH2	2310
JBT0361	>50000		Ac-FPLAVQLHVSKRSKEMA-NH2	2311
JBT0362	>50000		Ac-ASFPLAVQLHVSKRSKEM-NH2	2312
JBT0363	>50000		Ac-ASFPLAVQLHVSKRSKE-NH2	2313
JBT0364	>50000		Ac-ASFPLAVQLHVSKRSKEMAL-NH2	2314
JBT0365	>50000		Ac-ASFPLAVQLHVSKRSKEMALA-NH2	2315
JBT0366	>50000		Ac-ASFPLAVQLHVSKRSKEMALAR-NH2	2316
JBT0367	>50000		Ac-ASFPLAVQLHVSKRSKEMALARL-NH2	2317
JBT0368	27346	G	Ac-ASFPLAVQLHVSKRSKEMALARLY-NH2	2130
JBT0369	5107	F	Ac-ASFPLAVQLHVSKRSKEMALARLYY-NH2	2017
JBT0370	>50000		Ac-YASFPLAVQLHVSKRSKEMA-NH2	2318
JBT0371	>50000		Ac-GYASFPLAVQLHVSKRSKEMA-NH2	2319
JBT0660	2645	E	Ac-AGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2	2004

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT0661	1383	E	Ac-SAYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2	2005
JBT0662	13759	G	Ac-SGAASFPLAVQLHVSKRSKEMALARLYYKTS-NH2	2131
JBT0664	2739	E	Ac-SGYAAFPLAVQLHVSKRSKEMALARLYYKTS-NH2	2006
JBT0665	25704	G	Ac-SGYASAPLAVQLHVSKRSKEMALARLYYKTS-NH2	2132
JBT0666	915	D	Ac-SGYASFALAVQLHVSKRSKEMALARLYYKTS-NH2	2001
JBT0667	9314	F	Ac-SGYASFPAAVQLHVSKRSKEMALARLYYKTS-NH2	2018
JBT0669	33021	G	Ac-SGYASFPLAAQLHVSKRSKEMALARLYYKTS-NH2	2133
JBT0670	2506	E	Ac-SGYASFPLAVALHVSKRSKEMALARLYYKTS-NH2	2007
JBT0671	5836	F	Ac-SGYASFPLAVQAHVSKRSKEMALARLYYKTS-NH2	2019
JBT0672	38327	G	Ac-SGYASFPLAVQLAVSKRSKEMALARLYYKTS-NH2	2134
JBT0673	16425	G	Ac-SGYASFPLAVQLHASKRSKEMALARLYYKTS-NH2	2135
JBT0674	1173	E	Ac-SGYASFPLAVQLHVAKRSKEMALARLYYKTS-NH2	2008
JBT0675	18317	G	Ac-SGYASFPLAVQLHVSARSKEMALARLYYKTS-NH2	2136
JBT0676	37412	G	Ac-SGYASFPLAVQLHVSKASKEMALARLYYKTS-NH2	2137
JBT0677	1905	E	Ac-SGYASFPLAVQLHVSKRAKEMALARLYYKTS-NH2	2009
JBT0678	4071	E	Ac-SGYASFPLAVQLHVSKRSAEMALARLYYKTS-NH2	2010
JBT0679	6196	F	Ac-SGYASFPLAVQLHVSKRSKAMALARLYYKTS-NH2	2020
JBT0680	20935	G	Ac-SGYASFPLAVQLHVSKRSKEAALARLYYKTS-NH2	2138
JBT0682	10271	G	Ac-SGYASFPLAVQLHVSKRSKEMAAARLYYKTS-NH2	2139
JBT0684	4567	E	Ac-SGYASFPLAVQLHVSKRSKEMALAALYYKTS-NH2	2011
JBT0685	9134	F	Ac-SGYASFPLAVQLHVSKRSKEMALARAYYKTS-NH2	2021
JBT0686	13474	G	Ac-SGYASFPLAVQLHVSKRSKEMALARLAYKTS-NH2	2140
JBT0687	11783	G	Ac-SGYASFPLAVQLHVSKRSKEMALARLYAKTS-NH2	2141
JBT0688	4196	E	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYATS-NH2	2012
JBT0689	3127	E	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYKAS-NH2	2013
JBT0690	3000	E	Ac-SGYASFPLAVQLHVSKRSKEMALARLYYKTA-NH2	2014
JBT0788	16205	G	H-GSSGYASFPLAVQLHVSKRSKEMALARLYYKTS-OH	2142
JBT1579	2593	E	Ac-GYASFPLAVQLHVAKRSKEMA-NH2	2015
JBT1580	>50000		Ac-GYASFALSVQLHVSKRSKEMA-NH2	2320
JBT1581	>50000		Ac-GYASFALAVQLHVAKRSKEMA-NH2	2321
JBT1582	5841	F	Ac-GYASFPLAVQLHVMKRSKEMA-NH2	2022
JBT1583	>50000		Ac-GYASFALAVQLHVMKRSKEMA-NH2	2322

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1599	>50000		Ac-FPLAVQLHVSKRSKEMALA-NH2	2323
JBT1600	>50000		Ac-QLHVSKRSKEMALA-NH2	2324
JBT1601	>50000		Ac-SGYASFP-NH2	2325
JBT1602	>50000		Ac-LAVQLHVSKRSKEMALARL-NH2	2326
JBT1644			Ac-VYASFFLTVQLHVSKRSKEMA-NH2	2327
JBT1645			Ac-GYWSFPQAVQLHVSKRSKEMA-NH2	2328
JBT1646			Ac-GYASFILAVQLHVSKRSKEMA-NH2	2329
JBT1647			Ac-GYWSFYLAVQLHVSKRSKEMA-NH2	2330
JBT1648			Ac-SYASFFLAVQLHVSKRSEEMA-NH2	2331
JBT1649	10282	G	Ac-GYWSFPHAVWHHVSKRSKEMA-NH2	2143
JBT1650			Ac-GYASFILAVQLHIIKRSKEMA-NH2	2332
JBT1651			Ac-GMASFFLARDLHWSKVFKEMA-NH2	2333
JBT1653			Ac-EYAQFWLAVQLHVSKRSKEMA-NH2	2334
JBT1654			Ac-GYASFPLIVQLHVSKRSKEMA-NH2	2335
JBT1655			Ac-GYASFPIHVQHHVSKRSKEMA-NH2	2336
JBT1656			Ac-GYASFALMVQLHVSKRSKEMA-NH2	2337
JBT1657			Ac-GYASFHQAVQRHVSKRSKEMA-NH2	2338
JBT1658			Ac-GYASFALMVQHHVSKRSKEMA-NH2	2339
JBT1659	14473	G	Ac-GYASFWQAVQLHVWKRSKEIA-NH2	2144
JBT1660	14063	G	Ac-GYASFPLIVWLHVSKRSKEMA-NH2	2145
JBT1661			Ac-GYASFSLAVQLHVSKRSKEMA-NH2	2340
JBT1662			Ac-GYHSFKLAVQLHVSKRSKEIA-NH2	2341
JBT1663	12912	G	Ac-GYASFWQAVQLHVSKRSKEIA-NH2	2146
JBT1664			Ac-GYASYVLAVQHHVSWRSWEMA-NH2	2342
JBT1665			Ac-HYHSLPKQVQLHVSKRSKEMA-NH2	2343
JBT1666			Ac-GYASFSLAVQLHVHKRSYEMA-NH2	2344
JBT1668			Ac-GYASFELLVQLHVSKRSKEMA-NH2	2345
JBT1669			Ac-SYSSFKLAVQLHVSKRSKEMA-NH2	2346
JBT1670			Ac-GYAWFPLNVWLHVHKRSHEMA-NH2	2347
JBT1671			Ac-GYASFWLSVQTHVSKRSKEMA-NH2	2348

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1672			Ac-GYASFPLMVQLHVIKRSKEMA-NH2	2349
JBT1673			Ac-GYWSFNLVVQLHVSKRSKEMA-NH2	2350
JBT1674			Ac-GYASFHLAVQLHVWKRSKEMA-NH2	2351
JBT1675			Ac-GYASFWLVVQLHVSKRSKEMA-NH2	2352
JBT1676			Ac-GYWSFPLAVQLHVWKRSWEMA-NH2	2353
JBT1677			Ac-GYASFPWYVQLHVSKRSKEMA-NH2	2354
JBT1678			Ac-GYTSFQLAVQLHVSKRSKEMA-NH2	2355
JBT1679			Ac-RYASFPLAVYLHVTKRSKEMA-NH2	2356
JBT1680			Ac-GYWSFPLAVQLHVSKRLKEMA-NH2	2357
JBT1681			Ac-GHASFQTAVQQHVSKRSKEMA-NH2	2358
JBT1682			Ac-FHVSKRSKEMA-NH2	2359
JBT1683			Ac-GYASFWHAVQLHVSKRSKEMA-NH2	2360
JBT1684			Ac-GIASFILAVQLHVYKRSKEMA-NH2	2361
JBT1685			Ac-GYTSFMQAVQHHVSKRSKEIA-NH2	2362
JBT1686			Ac-HYHSFYLAVQLHVWERSYEMA-NH2	2363
JBT1687			Ac-GYASFWHAVQHHVTKRSREMA-NH2	2364
JBT1688			Ac-GYASFTLTVQLHVSKRSKEMA-NH2	2365
JBT1689			Ac-GYASFILTVQLHVSKRSKEMA-NH2	2366
JBT1690			Ac-GYASFWLAVQIHVSKRSKEMA-NH2	2367
JBT1691			Ac-GYASFILAVQHHVSKRSKEMA-NH2	2368
JBT1692			Ac-SYASFPPAVMLHVWKRSYEMA-NH2	2369
JBT1693			Ac-GYASFNLAVQLHVSKRSKEMA-NH2	2370
JBT1694			Ac-GYWSFNLAVQLHVSKRSKEMA-NH2	2371
JBT1695			Ac-GYASFFLAVQLHVSKRSKEMA-NH2	2372
JBT1696			Ac-GYASFWLAVQMHVWKRSKEMA-NH2	2373
JBT1697	8481	F	Ac-GYASFPWFVQLHVHKRSWEMA-NH2	223
JBT1698			Ac-GYASFSLAIQLHVSKRSKEMA-NH2	2374
JBT1858		F	Ac-AYASFPWFVQLHVHKRSWEMA-NH2	2024
JBT1859			Ac-GAASFPWFVQLHVHKRSWEMA-NH2	2375
JBT1860		F	Ac-GYAAFPWFVQLHVHKRSWEMA-NH2	2025

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1861			Ac-GYASAPWFVQLHVHKRSWEMA-NH2	2376
JBT1862			Ac-GYASFAWFVQLHVHKRSWEMA-NH2	2377
JBT1863		G	Ac-GYASFPAFVQLHVHKRSWEMA-NH2	2147
JBT1864		F	Ac-GYASFPWAVQLHVHKRSWEMA-NH2	2026
JBT1865			Ac-GYASFPWFAQLHVHKRSWEMA-NH2	2378
JBT1866		F	Ac-GYASFPWFVALHVHKRSWEMA-NH2	2027
JBT1867		G	Ac-GYASFPWFVQAHVHKRSWEMA-NH2	2148
JBT1868			Ac-GYASFPWFVQLAVHKRSWEMA-NH2	2379
JBT1869		G	Ac-GYASFPWFVQLHAHKRSWEMA-NH2	2149
JBT1870		G	Ac-GYASFPWFVQLHVAKRSWEMA-NH2	2150
JBT1871			Ac-GYASFPWFVQLHVHARSWEMA-NH2	2380
JBT1872			Ac-GYASFPWFVQLHVHKASWEMA-NH2	2381
JBT1874			Ac-GYASFPWFVQLHVHKRSAEMA-NH2	2382
JBT1875		G	Ac-GYASFPWFVQLHVHKRSWAMA-NH2	2151
JBT1876		G	Ac-GYASFPWFVQLHVHKRSWEAA-NH2	2152
JBT1877		F	Ac-CYASFPWFVQLHVHKRSWEMA-NH2	2028
JBT1878		F	Ac-GCASFPWFVQLHVHKRSWEMA-NH2	2029
JBT1879		F	Ac-GYCSFPWFVQLHVHKRSWEMA-NH2	2030
JBT1880		F	Ac-GYACFPWFVQLHVHKRSWEMA-NH2	2031
JBT1881		F	Ac-GYASCPWFVQLHVHKRSWEMA-NH2	2032
JBT1882		G	Ac-GYASFCWFVQLHVHKRSWEMA-NH2	2153
JBT1883		F	Ac-GYASFPCFVQLHVHKRSWEMA-NH2	2033
JBT1884		F	Ac-GYASFPWCVQLHVHKRSWEMA-NH2	2034
JBT1885		F	Ac-GYASFPWFCQLHVHKRSWEMA-NH2	2035
JBT1886		F	Ac-GYASFPWFVCLHVHKRSWEMA-NH2	2236
JBT1888		G	Ac-GYASFPWFVQLCVHKRSWEMA-NH2	2154
JBT1889		F	Ac-GYASFPWFVQLHCHKRSWEMA-NH2	2037
JBT1890		F	Ac-GYASFPWFVQLHVCKRSWEMA-NH2	2038
JBT1891			Ac-GYASFPWFVQLHVHCRSWEMA-NH2	2383
JBT1892			Ac-GYASFPWFVQLHVHKCSWEMA-NH2	2384

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1893		F	Ac-GYASFPWFVQLHVHKRCWEMA-NH2	2039
JBT1894		F	Ac-GYASFPWFVQLHVHKRSCEMA-NH2	2040
JBT1895		F	Ac-GYASFPWFVQLHVHKRSWCMA-NH2	2041
JBT1896		F	Ac-GYASFPWFVQLHVHKRSWECA-NH2	2042
JBT1897		F	Ac-GYASFPWFVQLHVHKRSWEMC-NH2	2043
JBT1898		F	Ac-DYASFPWFVQLHVHKRSWEMA-NH2	2044
JBT1899		G	Ac-GDASFPWFVQLHVHKRSWEMA-NH2	2155
JBT1900		G	Ac-GYDSFPWFVQLHVHKRSWEMA-NH2	2156
JBT1901		G	Ac-GYADFPWFVQLHVHKRSWEMA-NH2	2157
JBT1902		G	Ac-GYASDPWFVQLHVHKRSWEMA-NH2	2158
JBT1903			Ac-GYASFDWFVQLHVHKRSWEMA-NH2	2385
JBT1904			Ac-GYASFPDFVQLHVHKRSWEMA-NH2	2386
JBT1905			Ac-GYASFPWDVQLHVHKRSWEMA-NH2	2387
JBT1906			Ac-GYASFPWFDQLHVHKRSWEMA-NH2	2388
JBT1907			Ac-GYASFPWFVDLHVHKRSWEMA-NH2	2389
JBT1908			Ac-GYASFPWFVQDHVHKRSWEMA-NH2	2390
JBT1909			Ac-GYASFPWFVQLDVHKRSWEMA-NH2	2391
JBT1910			Ac-GYASFPWFVQLHDHKRSWEMA-NH2	2392
JBT1911			Ac-GYASFPWFVQLHVDKRSWEMA-NH2	2393
JBT1912			Ac-GYASFPWFVQLHVHDRSWEMA-NH2	2394
JBT1913			Ac-GYASFPWFVQLHVHKDSWEMA-NH2	2395
JBT1914			Ac-GYASFPWFVQLHVHKRDWEMA-NH2	2396
JBT1915			Ac-GYASFPWFVQLHVHKRSDEMA-NH2	2397
JBT1916		G	Ac-GYASFPWFVQLHVHKRSWDMA-NH2	2159
JBT1917			Ac-GYASFPWFVQLHVHKRSWEDA-NH2	2398
JBT1918			Ac-GYASFPWFVQLHVHKRSWEMD-NH2	2399
JBT1920			Ac-GEASFPWFVQLHVHKRSWEMA-NH2	2400
JBT1921		G	Ac-GYESFPWFVQLHVHKRSWEMA-NH2	2160
JBT1922		G	Ac-GYAEFPWFVQLHVHKRSWEMA-NH2	2161
JBT1923			Ac-GYASEPWFVQLHVHKRSWEMA-NH2	2401

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1924			Ac-GYASFEWFVQLHVHKRSWEMA-NH2	2402
JBT1925			Ac-GYASFPEFVQLHVHKRSWEMA-NH2	2403
JBT1926			Ac-GYASFPWEVQLHVHKRSWEMA-NH2	2404
JBT1927			Ac-GYASFPWFEQLHVHKRSWEMA-NH2	2405
JBT1928			Ac-GYASFPWFVELHVHKRSWEMA-NH2	2406
JBT1929			Ac-GYASFPWFVQEHVHKRSWEMA-NH2	2407
JBT1930			Ac-GYASFPWFVQLEVHKRSWEMA-NH2	2408
JBT1931			Ac-GYASFPWFVQLHEHKRSWEMA-NH2	2409
JBT1932			Ac-GYASFPWFVQLHVEKRSWEMA-NH2	2410
JBT1933			Ac-GYASFPWFVQLHVHERSWEMA-NH2	2411
JBT1934			Ac-GYASFPWFVQLHVHKESWEMA-NH2	2412
JBT1935			Ac-GYASFPWFVQLHVHKREWEMA-NH2	2413
JBT1936			Ac-GYASFPWFVQLHVHKRSEEMA-NH2	2414
JBT1937			Ac-GYASFPWFVQLHVHKRSWEEA-NH2	2415
JBT1938			Ac-GYASFPWFVQLHVHKRSWEME-NH2	2416
JBT1939		G	Ac-FYASFPWFVQLHVHKRSWEMA-NH2	2162
JBT1940		G	Ac-GFASFPWFVQLHVHKRSWEMA-NH2	2163
JBT1941		G	Ac-GYFSFPWFVQLHVHKRSWEMA-NH2	2164
JBT1942		G	Ac-GYAFFPWFVQLHVHKRSWEMA-NH2	2165
JBT1943			Ac-GYASFFWFVQLHVHKRSWEMA-NH2	2417
JBT1944		G	Ac-GYASFPFFVQLHVHKRSWEMA-NH2	2166
JBT1945		G	Ac-GYASFPWFFQLHVHKRSWEMA-NH2	2167
JBT1946		G	Ac-GYASFPWFVFLHVHKRSWEMA-NH2	2168
JBT1948			Ac-GYASFPWFVQLFVHKRSWEMA-NH2	2418
JBT1949		G	Ac-GYASFPWFVQLHFHKRSWEMA-NH2	2169
JBT1950		F	Ac-GYASFPWFVQLHVFKRSWEMA-NH2	2045
JBT1951			Ac-GYASFPWFVQLHVHFRSWEMA-NH2	2419
JBT1952		G	Ac-GYASFPWFVQLHVHKFSWEMA-NH2	2170
JBT1953		G	Ac-GYASFPWFVQLHVHKRFWEMA-NH2	2171
JBT1954		G	Ac-GYASFPWFVQLHVHKRSFEMA-NH2	2172

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1955		G	Ac-GYASFPWFVQLHVHKRSWFMA-NH2	2173
JBT1956		G	Ac-GYASFPWFVQLHVHKRSWEFA-NH2	2174
JBT1959		F	Ac-GYGSFPWFVQLHVHKRSWEMA-NH2	2046
JBT1960		F	Ac-GYAGFPWFVQLHVHKRSWEMA-NH2	2047
JBT1961		G	Ac-GYASGPWFVQLHVHKRSWEMA-NH2	2175
JBT1962			Ac-GYASFGWFVQLHVHKRSWEMA-NH2	2420
JBT1963		G	Ac-GYASFPGFVQLHVHKRSWEMA-NH2	2176
JBT1964		G	Ac-GYASFPWGVQLHVHKRSWEMA-NH2	2177
JBT1965		G	Ac-GYASFPWFGQLHVHKRSWEMA-NH2	2178
JBT1966		G	Ac-GYASFPWFVGLHVHKRSWEMA-NH2	2179
JBT1967		G	Ac-GYASFPWFVQGHVHKRSWEMA-NH2	2180
JBT1968			Ac-GYASFPWFVQLGVHKRSWEMA-NH2	2421
JBT1969		G	Ac-GYASFPWFVQLHGHKRSWEMA-NH2	2181
JBT1970		G	Ac-GYASFPWFVQLHVGKRSWEMA-NH2	2182
JBT1971			Ac-GYASFPWFVQLHVHGRSWEMA-NH2	2422
JBT1972			Ac-GYASFPWFVQLHVHKGSWEMA-NH2	2423
JBT1973		F	Ac-GYASFPWFVQLHVHKRGWEMA-NH2	2048
JBT1974			Ac-GYASFPWFVQLHVHKRSGEMA-NH2	2424
JBT1975		G	Ac-GYASFPWFVQLHVHKRSWGMA-NH2	2183
JBT1976		G	Ac-GYASFPWFVQLHVHKRSWEGA-NH2	2184
JBT1979		G	Ac-GHASFPWFVQLHVHKRSWEMA-NH2	2185
JBT1980		F	Ac-GYHSFPWFVQLHVHKRSWEMA-NH2	2049
JBT1981		F	Ac-GYAHFPWFVQLHVHKRSWEMA-NH2	2050
JBT1982		F	Ac-GYASHPWFVQLHVHKRSWEMA-NH2	2051
JBT1984		G	Ac-GYASFPHFVQLHVHKRSWEMA-NH2	2186
JBT1985		F	Ac-GYASFPWHVQLHVHKRSWEMA-NH2	2052
JBT1986		G	Ac-GYASFPWFHQLHVHKRSWEMA-NH2	2187
JBT1987		G	Ac-GYASFPWFVHLHVHKRSWEMA-NH2	2188
JBT1989		G	Ac-GYASFPWFVQLHHHKRSWEMA-NH2	2189
JBT1990			Ac-GYASFPWFVQLHVHHRSWEMA-NH2	2425

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT1991			Ac-GYASFPWFVQLHVHKHSWEMA-NH2	2426
JBT1993			Ac-GYASFPWFVQLHVHKRSHEMA-NH2	2427
JBT1994		F	Ac-GYASFPWFVQLHVHKRSWHMA-NH2	2053
JBT1995		G	Ac-GYASFPWFVQLHVHKRSWEHA-NH2	2190
JBT1996		F	Ac-GYASFPWFVQLHVHKRSWEMH-NH2	2054
JBT1997		G	Ac-IYASFPWFVQLHVHKRSWEMA-NH2	2191
JBT1999		F	Ac-GYISFPWFVQLHVHKRSWEMA-NH2	2055
JBT2000		F	Ac-GYAIFPWFVQLHVHKRSWEMA-NH2	2056
JBT2001		G	Ac-GYASIPWFVQLHVHKRSWEMA-NH2	2192
JBT2002			Ac-GYASFIWFVQLHVHKRSWEMA-NH2	2428
JBT2003		G	Ac-GYASFPIFVQLHVHKRSWEMA-NH2	2193
JBT2005		G	Ac-GYASFPWFIQLHVHKRSWEMA-NH2	2194
JBT2006		G	Ac-GYASFPWFVILHVHKRSWEMA-NH2	2195
JBT2007		F	Ac-GYASFPWFVQIHVHKRSWEMA-NH2	2057
JBT2008		G	Ac-GYASFPWFVQLIVHKRSWEMA-NH2	2196
JBT2010		F	Ac-GYASFPWFVQLHVIKRSWEMA-NH2	2058
JBT2011			Ac-GYASFPWFVQLHVHIRSWEMA-NH2	2429
JBT2012			Ac-GYASFPWFVQLHVHKISWEMA-NH2	2430
JBT2013		F	Ac-GYASFPWFVQLHVHKRIWEMA-NH2	2059
JBT2014		F	Ac-GYASFPWFVQLHVHKRSIEMA-NH2	2060
JBT2017		F	Ac-GYASFPWFVQLHVHKRSWEMI-NH2	2061
JBT2018		F	Ac-KYASFPWFVQLHVHKRSWEMA-NH2	2062
JBT2019		F	Ac-GKASFPWFVQLHVHKRSWEMA-NH2	2063
JBT2020		F	Ac-GYKSFPWFVQLHVHKRSWEMA-NH2	2064
JBT2021		F	Ac-GYAKFPWFVQLHVHKRSWEMA-NH2	2065
JBT2022		F	Ac-GYASKPWFVQLHVHKRSWEMA-NH2	2066
JBT2023		G	Ac-GYASFKWFVQLHVHKRSWEMA-NH2	2197
JBT2024		G	Ac-GYASFPKFVQLHVHKRSWEMA-NH2	2198
JBT2025		F	Ac-GYASFPWKVQLHVHKRSWEMA-NH2	2067
JBT2027		F	Ac-GYASFPWFVKLHVHKRSWEMA-NH2	2068

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT2028		F	Ac-GYASFPWFVQKHVHKRSWEMA-NH2	2069
JBT2029		G	Ac-GYASFPWFVQLKVHKRSWEMA-NH2	2199
JBT2030		F	Ac-GYASFPWFVQLHKHKRSWEMA-NH2	2070
JBT2032			Ac-GYASFPWFVQLHVHKKSWEMA-NH2	2431
JBT2033		F	Ac-GYASFPWFVQLHVHKRKWEMA-NH2	2071
JBT2034		G	Ac-GYASFPWFVQLHVHKRSKEMA-NH2	2200
JBT2035		F	Ac-GYASFPWFVQLHVHKRSWKMA-NH2	2072
JBT2036		G	Ac-GYASFPWFVQLHVHKRSWEKA-NH2	2201
JBT2037		F	Ac-GYASFPWFVQLHVHKRSWEMK-NH2	2073
JBT2038		F	Ac-LYASFPWFVQLHVHKRSWEMA-NH2	2074
JBT2040		F	Ac-GYLSFPWFVQLHVHKRSWEMA-NH2	2075
JBT2041		F	Ac-GYALFPWFVQLHVHKRSWEMA-NH2	2076
JBT2042		G	Ac-GYASLPWFVQLHVHKRSWEMA-NH2	2202
JBT2043			Ac-GYASFLWFVQLHVHKRSWEMA-NH2	2432
JBT2044		G	Ac-GYASFPLFVQLHVHKRSWEMA-NH2	2203
JBT2045		G	Ac-GYASFPWLVQLHVHKRSWEMA-NH2	2204
JBT2046		G	Ac-GYASFPWFLQLHVHKRSWEMA-NH2	2205
JBT2047		F	Ac-GYASFPWFVLLHVHKRSWEMA-NH2	2077
JBT2048		G	Ac-GYASFPWFVQLLVHKRSWEMA-NH2	2206
JBT2049		G	Ac-GYASFPWFVQLHLHKRSWEMA-NH2	2207
JBT2050		G	Ac-GYASFPWFVQLHVLKRSWEMA-NH2	2208
JBT2051			Ac-GYASFPWFVQLHVHLRSWEMA-NH2	2433
JBT2052			Ac-GYASFPWFVQLHVHKLSWEMA-NH2	2434
JBT2053		F	Ac-GYASFPWFVQLHVHKRLWEMA-NH2	2078
JBT2054		G	Ac-GYASFPWFVQLHVHKRSLEMA-NH2	2209
JBT2055		G	Ac-GYASFPWFVQLHVHKRSWLMA-NH2	2210
JBT2056		G	Ac-GYASFPWFVQLHVHKRSWELA-NH2	2211
JBT2057		F	Ac-GYASFPWFVQLHVHKRSWEML-NH2	2079
JBT2058		F	Ac-MYASFPWFVQLHVHKRSWEMA-NH2	2080
JBT2059		G	Ac-GMASFPWFVQLHVHKRSWEMA-NH2	2212

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT2060		F	Ac-GYMSFPWFVQLHVHKRSWEMA-NH2	2081
JBT2061		F	Ac-GYAMFPWFVQLHVHKRSWEMA-NH2	2082
JBT2062		G	Ac-GYASMPWFVQLHVHKRSWEMA-NH2	2213
JBT2063			Ac-GYASFMWFVQLHVHKRSWEMA-NH2	2435
JBT2064			Ac-GYASFPMFVQLHVHKRSWEMA-NH2	2436
JBT2065		G	Ac-GYASFPWMVQLHVHKRSWEMA-NH2	2214
JBT2066		G	Ac-GYASFPWFMQLHVHKRSWEMA-NH2	2215
JBT2067		G	Ac-GYASFPWFVMLHVHKRSWEMA-NH2	2216
JBT2068		F	Ac-GYASFPWFVQMHVHKRSWEMA-NH2	2083
JBT2069			Ac-GYASFPWFVQLMVHKRSWEMA-NH2	2437
JBT2070		G	Ac-GYASFPWFVQLHMHKRSWEMA-NH2	2217
JBT2071		G	Ac-GYASFPWFVQLHVMKRSWEMA-NH2	2218
JBT2072			Ac-GYASFPWFVQLHVHMRSWEMA-NH2	2438
JBT2073			Ac-GYASFPWFVQLHVHKMSWEMA-NH2	2439
JBT2075		G	Ac-GYASFPWFVQLHVHKRSMEMA-NH2	2219
JBT2076			Ac-GYASFPWFVQLHVHKRSWMMA-NH2	2440
JBT2077		F	Ac-GYASFPWFVQLHVHKRSWEMM-NH2	2084
JBT2078		F	Ac-NYASFPWFVQLHVHKRSWEMA-NH2	2085
JBT2079		G	Ac-GNASFPWFVQLHVHKRSWEMA-NH2	2220
JBT2080		G	Ac-GYNSFPWFVQLHVHKRSWEMA-NH2	2221
JBT2081		F	Ac-GYANFPWFVQLHVHKRSWEMA-NH2	2086
JBT2082		G	Ac-GYASNPWFVQLHVHKRSWEMA-NH2	2222
JBT2083			Ac-GYASFNWFVQLHVHKRSWEMA-NH2	2441
JBT2084		G	Ac-GYASFPNFVQLHVHKRSWEMA-NH2	2223
JBT2085		G	Ac-GYASFPWNVQLHVHKRSWEMA-NH2	2224
JBT2086			Ac-GYASFPWFNQLHVHKRSWEMA-NH2	2442
JBT2087		G	Ac-GYASFPWFVNLHVHKRSWEMA-NH2	2225
JBT2088		G	Ac-GYASFPWFVQNHVHKRSWEMA-NH2	2226
JBT2089			Ac-GYASFPWFVQLNVHKRSWEMA-NH2	2443
JBT2090			Ac-GYASFPWFVQLHNHKRSWEMA-NH2	2444

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT2091		G	Ac-GYASFPWFVQLHVNKRSWEMA-NH2	2227
JBT2092			Ac-GYASFPWFVQLHVHNRSWEMA-NH2	2445
JBT2093			Ac-GYASFPWFVQLHVHKNSWEMA-NH2	2446
JBT2094		F	Ac-GYASFPWFVQLHVHKRNWEMA-NH2	2087
JBT2095			Ac-GYASFPWFVQLHVHKRSNEMA-NH2	2447
JBT2096		G	Ac-GYASFPWFVQLHVHKRSWNMA-NH2	2228
JBT2097		G	Ac-GYASFPWFVQLHVHKRSWENA-NH2	2229
JBT2098		F	Ac-GYASFPWFVQLHVHKRSWEMN-NH2	2088
JBT2099		F	Ac-PYASFPWFVQLHVHKRSWEMA-NH2	2089
JBT2100		G	Ac-GPASFPWFVQLHVHKRSWEMA-NH2	2230
JBT2101		G	Ac-GYPSFPWFVQLHVHKRSWEMA-NH2	2231
JBT2103		G	Ac-GYASPPWFVQLHVHKRSWEMA-NH2	2232
JBT2104			Ac-GYASFPPFVQLHVHKRSWEMA-NH2	2448
JBT2105		G	Ac-GYASFPWPVQLHVHKRSWEMA-NH2	2233
JBT2106			Ac-GYASFPWFPQLHVHKRSWEMA-NH2	2449
JBT2107		G	Ac-GYASFPWFVPLHVHKRSWEMA-NH2	2234
JBT2108			Ac-GYASFPWFVQPHVHKRSWEMA-NH2	2450
JBT2109			Ac-GYASFPWFVQLPVHKRSWEMA-NH2	2451
JBT2110			Ac-GYASFPWFVQLHPHKRSWEMA-NH2	2452
JBT2111			Ac-GYASFPWFVQLHVPKRSWEMA-NH2	2453
JBT2112			Ac-GYASFPWFVQLHVHPRSWEMA-NH2	2454
JBT2113			Ac-GYASFPWFVQLHVHKPSWEMA-NH2	2455
JBT2114			Ac-GYASFPWFVQLHVHKRPWEMA-NH2	2456
JBT2115			Ac-GYASFPWFVQLHVHKRSPEMA-NH2	2457
JBT2116		G	Ac-GYASFPWFVQLHVHKRSWPMA-NH2	2235
JBT2118		G	Ac-GYASFPWFVQLHVHKRSWEMP-NH2	2236
JBT2119		G	Ac-QYASFPWFVQLHVHKRSWEMA-NH2	2237
JBT2120			Ac-GQASFPWFVQLHVHKRSWEMA-NH2	2458
JBT2121		G	Ac-GYQSFPWFVQLHVHKRSWEMA-NH2	2238
JBT2122		G	Ac-GYAQFPWFVQLHVHKRSWEMA-NH2	2239

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT2123		G	Ac-GYASQPWFVQLHVHKRSWEMA-NH2	2240
JBT2124			Ac-GYASFQWFVQLHVHKRSWEMA-NH2	2459
JBT2125			Ac-GYASFPQFVQLHVHKRSWEMA-NH2	2460
JBT2126		G	Ac-GYASFPWQVQLHVHKRSWEMA-NH2	2241
JBT2127			Ac-GYASFPWFQQLHVHKRSWEMA-NH2	2461
JBT2128		G	Ac-GYASFPWFVQQHVHKRSWEMA-NH2	2242
JBT2129			Ac-GYASFPWFVQLQVHKRSWEMA-NH2	2462
JBT2130			Ac-GYASFPWFVQLHQHKRSWEMA-NH2	2463
JBT2131		G	Ac-GYASFPWFVQLHVQKRSWEMA-NH2	2243
JBT2132			Ac-GYASFPWFVQLHVHQRSWEMA-NH2	2464
JBT2133			Ac-GYASFPWFVQLHVHKQSWEMA-NH2	2465
JBT2134		F	Ac-GYASFPWFVQLHVHKRQWEMA-NH2	2090
JBT2135		G	Ac-GYASFPWFVQLHVHKRSQEMA-NH2	2244
JBT2136		G	Ac-GYASFPWFVQLHVHKRSWQMA-NH2	2245
JBT2137		G	Ac-GYASFPWFVQLHVHKRSWEQA-NH2	2246
JBT2138		F	Ac-GYASFPWFVQLHVHKRSWEMQ-NH2	2091
JBT2139		F	Ac-RYASFPWFVQLHVHKRSWEMA-NH2	2092
JBT2140		F	Ac-GRASFPWFVQLHVHKRSWEMA-NH2	2093
JBT2141		F	Ac-GYRSFPWFVQLHVHKRSWEMA-NH2	2094
JBT2142		F	Ac-GYARFPWFVQLHVHKRSWEMA-NH2	2095
JBT2143		F	Ac-GYASRPWFVQLHVHKRSWEMA-NH2	2096
JBT2144		G	Ac-GYASFRWFVQLHVHKRSWEMA-NH2	2247
JBT2145		F	Ac-GYASFPRFVQLHVHKRSWEMA-NH2	2097
JBT2146		F	Ac-GYASFPWRVQLHVHKRSWEMA-NH2	2098
JBT2147		F	Ac-GYASFPWFRQLHVHKRSWEMA-NH2	2099
JBT2148		F	Ac-GYASFPWFVRLHVHKRSWEMA-NH2	2100
JBT2149		F	Ac-GYASFPWFVQRHVHKRSWEMA-NH2	2101
JBT2150		G	Ac-GYASFPWFVQLRVHKRSWEMA-NH2	2248
JBT2151		F	Ac-GYASFPWFVQLHRHKRSWEMA-NH2	2102
JBT2152		F	Ac-GYASFPWFVQLHVRKRSWEMA-NH2	2103

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT2153		F	Ac-GYASFPWFVQLHVHRRSWEMA-NH2	2104
JBT2154		F	Ac-GYASFPWFVQLHVHKRRWEMA-NH2	2105
JBT2155		F	Ac-GYASFPWFVQLHVHKRSREMA-NH2	2106
JBT2156		F	Ac-GYASFPWFVQLHVHKRSWRMA-NH2	2107
JBT2157		G	Ac-GYASFPWFVQLHVHKRSWERA-NH2	2249
JBT2158		F	Ac-GYASFPWFVQLHVHKRSWEMR-NH2	2108
JBT2159		F	Ac-SYASFPWFVQLHVHKRSWEMA-NH2	2109
JBT2160		G	Ac-GSASFPWFVQLHVHKRSWEMA-NH2	2250
JBT2161		F	Ac-GYSSFPWFVQLHVHKRSWEMA-NH2	2110
JBT2162		G	Ac-GYASSPWFVQLHVHKRSWEMA-NH2	2251
JBT2163			Ac-GYASFSWFVQLHVHKRSWEMA-NH2	2466
JBT2164		G	Ac-GYASFPSFVQLHVHKRSWEMA-NH2	2252
JBT2165		G	Ac-GYASFPWSVQLHVHKRSWEMA-NH2	2253
JBT2166		G	Ac-GYASFPWFSQLHVHKRSWEMA-NH2	2254
JBT2167		F	Ac-GYASFPWFVSLHVHKRSWEMA-NH2	2111
JBT2168		G	Ac-GYASFPWFVQSHVHKRSWEMA-NH2	2255
JBT2169			Ac-GYASFPWFVQLSVHKRSWEMA-NH2	2467
JBT2170		G	Ac-GYASFPWFVQLHSHKRSWEMA-NH2	2256
JBT2171		G	Ac-GYASFPWFVQLHVSKRSWEMA-NH2	2257
JBT2172			Ac-GYASFPWFVQLHVHSRSWEMA-NH2	2468
JBT2173			Ac-GYASFPWFVQLHVHKSSWEMA-NH2	2469
JBT2174			Ac-GYASFPWFVQLHVHKRSSEMA-NH2	2470
JBT2175			Ac-GYASFPWFVQLHVHKRSWSMA-NH2	2471
JBT2176			Ac-GYASFPWFVQLHVHKRSWESA-NH2	2472
JBT2177			Ac-GYASFPWFVQLHVHKRSWEMS-NH2	2473
JBT2178		F	Ac-TYASFPWFVQLHVHKRSWEMA-NH2	2112
JBT2179		G	Ac-GTASFPWFVQLHVHKRSWEMA-NH2	2258
JBT2180		F	Ac-GYTSFPWFVQLHVHKRSWEMA-NH2	2113
JBT2181		F	Ac-GYATFPWFVQLHVHKRSWEMA-NH2	2114
JBT2182		G	Ac-GYASTPWFVQLHVHKRSWEMA-NH2	2259

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT2183			Ac-GYASFTWFVQLHVHKRSWEMA-NH2	2474
JBT2184		G	Ac-GYASFPTFVQLHVHKRSWEMA-NH2	2260
JBT2185		G	Ac-GYASFPWTVQLHVHKRSWEMA-NH2	2261
JBT2186		G	Ac-GYASFPWFTQLHVHKRSWEMA-NH2	2262
JBT2187		G	Ac-GYASFPWFVTLHVHKRSWEMA-NH2	2263
JBT2188		G	Ac-GYASFPWFVQTHVHKRSWEMA-NH2	2264
JBT2189			Ac-GYASFPWFVQLTVHKRSWEMA-NH2	2475
JBT2190			Ac-GYASFPWFVQLHTHKRSWEMA-NH2	2476
JBT2191		G	Ac-GYASFPWFVQLHVTKRSWEMA-NH2	2265
JBT2192			Ac-GYASFPWFVQLHVHTRSWEMA-NH2	2477
JBT2193			Ac-GYASFPWFVQLHVHKTSWEMA-NH2	2478
JBT2194		F	Ac-GYASFPWFVQLHVHKRTWEMA-NH2	2115
JBT2195			Ac-GYASFPWFVQLHVHKRSTEMA-NH2	2479
JBT2196			Ac-GYASFPWFVQLHVHKRSWTMA-NH2	2480
JBT2197			Ac-GYASFPWFVQLHVHKRSWETA-NH2	2481
JBT2198		G	Ac-GYASFPWFVQLHVHKRSWEMT-NH2	2266
JBT2199			Ac-VYASFPWFVQLHVHKRSWEMA-NH2	2482
JBT2200			Ac-GVASFPWFVQLHVHKRSWEMA-NH2	2483
JBT2201			Ac-GYVSFPWFVQLHVHKRSWEMA-NH2	2484
JBT2202		G	Ac-GYAVFPWFVQLHVHKRSWEMA-NH2	2267
JBT2203		G	Ac-GYASVPWFVQLHVHKRSWEMA-NH2	2268
JBT2204			Ac-GYASFVWFVQLHVHKRSWEMA-NH2	2485
JBT2205			Ac-GYASFPVFVQLHVHKRSWEMA-NH2	2486
JBT2206		F	Ac-GYASFPWVVQLHVHKRSWEMA-NH2	2116
JBT2207		G	Ac-GYASFPWFVVLHVHKRSWEMA-NH2	2269
JBT2208		F	Ac-GYASFPWFVQVHVHKRSWEMA-NH2	2117
JBT2209			Ac-GYASFPWFVQLVVHKRSWEMA-NH2	2487
JBT2210		G	Ac-GYASFPWFVQLHVVKRSWEMA-NH2	2270
JBT2211			Ac-GYASFPWFVQLHVHVRSWEMA-NH2	2488
JBT2212			Ac-GYASFPWFVQLHVHKVSWEMA-NH2	2489

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT2213		F	Ac-GYASFPWFVQLHVHKRVWEMA-NH2	2118
JBT2214		G	Ac-GYASFPWFVQLHVHKRSVEMA-NH2	2271
JBT2215		G	Ac-GYASFPWFVQLHVHKRSWVMA-NH2	2272
JBT2216		G	Ac-GYASFPWFVQLHVHKRSWEVA-NH2	2273
JBT2217		F	Ac-GYASFPWFVQLHVHKRSWEMV-NH2	2119
JBT2218		G	Ac-WYASFPWFVQLHVHKRSWEMA-NH2	2274
JBT2219		F	Ac-GWASFPWFVQLHVHKRSWEMA-NH2	2120
JBT2220		F	Ac-GYWSFPWFVQLHVHKRSWEMA-NH2	2121
JBT2221		G	Ac-GYAWFPWFVQLHVHKRSWEMA-NH2	2275
JBT2222		F	Ac-GYASWPWFVQLHVHKRSWEMA-NH2	2122
JBT2223			Ac-GYASFWWFVQLHVHKRSWEMA-NH2	2490
JBT2224		F	Ac-GYASFPWWVQLHVHKRSWEMA-NH2	2123
JBT2225		G	Ac-GYASFPWFWQLHVHKRSWEMA-NH2	2276
JBT2226		G	Ac-GYASFPWFVWLHVHKRSWEMA-NH2	2277
JBT2227		F	Ac-GYASFPWFVQWHVHKRSWEMA-NH2	2124
JBT2228			Ac-GYASFPWFVQLWVHKRSWEMA-NH2	2491
JBT2229		G	Ac-GYASFPWFVQLHWHKRSWEMA-NH2	2278
JBT2230		G	Ac-GYASFPWFVQLHVWKRSWEMA-NH2	2279
JBT2231			Ac-GYASFPWFVQLHVHWRSWEMA-NH2	2492
JBT2232			Ac-GYASFPWFVQLHVHKWSWEMA-NH2	2493
JBT2233		G	Ac-GYASFPWFVQLHVHKRWWEMA-NH2	2280
JBT2234		G	Ac-GYASFPWFVQLHVHKRSWWMA-NH2	2281
JBT2235		G	Ac-GYASFPWFVQLHVHKRSWEWA-NH2	2282
JBT2236		F	Ac-GYASFPWFVQLHVHKRSWEMW-NH2	2125
JBT2238		G	Ac-GYYSFPWFVQLHVHKRSWEMA-NH2	2283
JBT2239		G	Ac-GYAYFPWFVQLHVHKRSWEMA-NH2	2284
JBT2240		G	Ac-GYASYPWFVQLHVHKRSWEMA-NH2	2285
JBT2241			Ac-GYASFYWFVQLHVHKRSWEMA-NH2	2494
JBT2242		G	Ac-GYASFPYFVQLHVHKRSWEMA-NH2	2286
JBT2243		F	Ac-GYASFPWYVQLHVHKRSWEMA-NH2	2126

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FIGURE 36

Object ID	IC50 [nM]	Affinity Class	Sequence	SEQ ID NO:
JBT2244		G	Ac-GYASFPWFYQLHVHKRSWEMA-NH2	2287
JBT2245		G	Ac-GYASFPWFVYLHVHKRSWEMA-NH2	2288
JBT2246		G	Ac-GYASFPWFVQYHVHKRSWEMA-NH2	2289
JBT2247			Ac-GYASFPWFVQLYVHKRSWEMA-NH2	2495
JBT2248		G	Ac-GYASFPWFVQLHYHKRSWEMA-NH2	2290
JBT2249		G	Ac-GYASFPWFVQLHVYKRSWEMA-NH2	2291
JBT2250			Ac-GYASFPWFVQLHVHYRSWEMA-NH2	2496
JBT2251			Ac-GYASFPWFVQLHVHKYSWEMA-NH2	2497
JBT2252		G	Ac-GYASFPWFVQLHVHKRYWEMA-NH2	2292
JBT2253		G	Ac-GYASFPWFVQLHVHKRSYEMA-NH2	2293
JBT2254		G	Ac-GYASFPWFVQLHVHKRSWYMA-NH2	2294
JBT2255		G	Ac-GYASFPWFVQLHVHKRSWEYA-NH2	2295
JBT2256		G	Ac-GYASFPWFVQLHVHKRSWEMY-NH2	2296

FIGURE 37

Object ID	EC50 [nM]		Sequence	SEQ ID NO:
JBT0126	55.6868		Biotinyl-Ttds-SGYASFPLAVQLHVSKRSKEMALARLYYKTS-NH2	2498

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FIGURE 38

Object ID	IC50 Mean (nM)	Affinity Class	Sequence	SEQ ID NO:
JBT0049	ND		Ac-SGNTFVDERLLYFLTIGNMGMYAAQLKFRTS-NH2	3025
JBT0050	2745	E	Ac-SGRGCTKVIVFTFRHNKLIGYERRYNCTS-NH2	3047
JBT0101	251	D	Ac-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTSKK-NH2	3036
JBT0129	ND		Ac-SGRG[CTKVIVFTFRHNKLIGYERRYNC]TS-NH2	3026
JBT0176*	39444	G	Ac-KKVGSVTRWSMYGPIFIKFTWTLEQPVGWDHKK-NH2	3056
JBT0177*	>50000		Ac-KKLTGDWTYFWSKVIWGPGVIERQMPVSTFHKK-NH2	3065
JBT0178	72	B	Ac-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTKK-NH2	3028
JBT0179	>50000		Ac-KKSGVWQTHPRYFWTMWPDIKGEVIVLKK-NH2	3066
JBT0180	>50000		Ac-KKSGVWQTHPRYFWTMWPDIKGEVKK-NH2	3067
JBT0181	>50000		Ac-KKSGVWQTHPRYFWTMWPDIKKK-NH2	3068
JBT0182	351	D	Ac-KKGVWQTHPRYFWTMWPDIKGEVIVLFGTSKK-NH2	3037
JBT0183	>50000		Ac-KKQTHPRYFWTMWPDIKGEVIVLFGTSKK-NH2	3069
JBT0184	>50000		Ac-KKPRYFWTMWPDIKGEVIVLFGTSKK-NH2	3070
JBT0185	45487	G	Ac-KKFWTMWPDIKGEVIVLFGTSKK-NH2	3057
JBT0186	>50000		Ac-KKSGVWQTHPRYFWTMWPDIKGKK-NH2	3071
JBT0187	>50000		Ac-KKWQTHPRYFWTMWPDIKGEVIKK-NH2	3072
JBT0188	25532	G	Ac-KKHPRYFWTMWPDIKGEVIVLFKK-NH2	3058
JBT0189	35115	G	Ac-KKYFWTMWPDIKGEVIVLFGTSKK-NH2	3059
JBT0190	100	C	Ac-KKSGNTFVDERLLYFLTIGNMGMYAAQLKFRTSKK-NH2	3031
JBT0191*	8935	F	Ac-MLGVLMRGISALTGDYTARFEFYLNKQTFN-NH2	3054
JBT0193	>50000		Ac-KKSGNTFVDERLLYFLTIGNMGMYAAQLKFKK-NH2	3073
JBT0194	>50000		Ac-KKSGNTFVDERLLYFLTIGNMGMYAAQKK-NH2	3074
JBT0195	>50000		Ac-KKSGNTFVDERLLYFLTIGNMGMYKK-NH2	3075
JBT0196	3457	E	Ac-KKSGNTFVDERLLYFLTIGNMKK-NH2	3048
JBT0197	>50000		Ac-KKTFVDERLLYFLTIGNMGMYAAQLKFRTSKK-NH2	3076
JBT0198	>50000		Ac-KKDERLLYFLTIGNMGMYAAQLKFRTSKK-NH2	3077
JBT0199	>50000		Ac-KKLLYFLTIGNMGMYAAQLKFRTSKK-NH2	3078
JBT0200	>50000		Ac-KKFLTIGNMGMYAAQLKFRTSKK-NH2	3079
JBT0201	>50000		Ac-KKLLYFLTIGNMGMYAAQLKFRKK-NH2	3080
JBT0202	>50000		Ac-KKLYFLTIGNMGMYAAQLKFRTKK-NH2	3081
JBT0203	>50000		Ac-KKYFLTIGNMGMYAAQLKFRTSKK-NH2	3082

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FIGURE 38

Object ID	IC50 Mean (nM)	Affinity Class	Sequence	SEQ ID NO:
JBT0204	25693	G	Ac-KKSGNTFVDERLLYFLTIGNMGKK-NH2	3060
JBT0205	72	B	Ac-KKSGRGCTKVIVFTFRHNKLIGYERRYNCTSKK-NH2	3029
JBT0206*	3594	E	Ac-TNYTGSEKCIRFVTRRYLGVRINCFHKGS-NH2	3049
JBT0207*	4316	E	Ac-TRNVVRRYECFGSTGCIKYFIHSRTGLNK-NH2	3050
JBT0208	ND		Ac-KKSGRGCTKVIVFTFRHNKLIGYERRYNKK-NH2	3027
JBT0209	4756	E	Ac-KKSGRGCTKVIVFTFRHNKLIGYERKK-NH2	3051
JBT0210	20471	G	Ac-KKSGRGCTKVIVFTFRHNKLIGKK-NH2	3061
JBT0211	211	C	Ac-KKGCTKVIVFTFRHNKLIGYERRYNCTSKK-NH2	3032
JBT0212	117	C	Ac-KKKVIVFTFRHNKLIGYERRYNCTSKK-NH2	3033
JBT0213	>50000		Ac-KKVFTFRHNKLIGYERRYNCTSKK-NH2	3083
JBT0214	2277	E	Ac-KKGRGCTKVIVFTFRHNKLIGYKK-NH2	3052
JBT0215	6146	F	Ac-KKGCTKVIVFTFRHNKLIGYERKK-NH2	3055
JBT0216	2123	E	Ac-KKCTKVIVFTFRHNKLIGYERRKK-NH2	3053
JBT0217	47024	G	Ac-KKTKVIVFTFRHNKLIGYERRYKK-NH2	3062
JBT0218	32865	G	Ac-KKKVIVFTFRHNKLIGYERRYNKK-NH2	3063
JBT0219	78	B	Ac-KKVIVFTFRHNKLIGYERRYNCKK-NH2	3030
JBT0220	31914	G	Ac-KKIVFTFRHNKLIGYERRYNCTKK-NH2	3064
JBT0344	629	D	Ac-TFVDERLLYFLTIGNMGMYAAQLKF-NH2	3038
JBT0345	271	D	Ac-FVDERLLYFLTIGNMGMYAAQLKF-NH2	3039
JBT0346	214	C	Ac-VDERLLYFLTIGNMGMYAAQLKF-NH2	3034
JBT0347	434	D	Ac-TFVDERLLYFLTIGNMGMYAAQLK-NH2	3040
JBT0348	>5000		Ac-TFVDERLLYFLTIGNMGMYAAQ-NH2	3084
JBT0349	552	D	Ac-GCTKVIVFTFRHNKLIGYERRYNCTS-NH2	3041
JBT0350	527	D	Ac-CTKVIVFTFRHNKLIGYERRYNCTS-NH2	3042
JBT0351	948	D	Ac-TKVIVFTFRHNKLIGYERRYNCTS-NH2	3043
JBT0352	848	D	Ac-GCTKVIVFTFRHNKLIGYERRYNCT-NH2	3044
JBT0353	668	D	Ac-GCTKVIVFTFRHNKLIGYERRYNC-NH2	3045
JBT0357	125	C	Ac-KKGVWQTHPRYFWTMWPDIKGEVIVLFGKK-NH2	3035
JBT0358	802	D	Ac-KKGVWQTHPRYFWTMWPDIKGEVIVLFKK-NH2	3046

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FIGURE 39

Object ID	EC50 [nM]	Object Name	SEQ ID NO:
JBT0052	2.8	Biotinyl-Ttds-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTSKK-NH2	3004
JBT0053	50.3	Biotinyl-Ttds-SGNTFVDERLLYFLTIGNMGMYAAQLKFRTS-NH2	3006
JBT0054	2.1	Biotinyl-Ttds-SGRGCTKVIVFTFRHNKLIGYERRYNCTS-NH2	3002
JBT0057	>5000	Ac-SGNTFVDERLLYFLTIGNMGMYAAQLKFRTS-Ttds-Lysin(biotin)-NH2	3018
JBT0058	2.4	Ac-SGRGCTKVIVFTFRHNKLIGYERRYNCTS-Ttds-Lysin(biotin)-NH2	3003
JBT0103	4.8	Ac-KKSGVWQTHPRYFWTMWPDIKGEVIVLFGTS-Ttds-KK-Lysin(biotin)-NH2	3005
JBT0105	>10000	Biotinyl-Ttds-SGQFWHGLLLPKGIVLNWTLKPWWMIGAFTS-NH2	3015
JBT0106	>10000	Ac-SGQFWHGLLLPKGIVLNWTLKPWWMIGAFTS-Ttds-Lysin(biotin)-NH2	3016
JBT0113	458.7	Biotinyl-Ttds-LSLVGSFAVLCA-NH2	3007
JBT0117	>50000	Biotinyl-Ttds-SGRFTTRLWYFSWDRLPWYMPFKQYVLSSTS-NH2	3019
JBT0118	>50000	Biotinyl-Ttds-SGSYQWWARPYRMWFGLPYWNQRVIFPWNTS-NH2	3020
JBT0128	>5000	Ac-Ttds-SGRVSYYWGVKWSQQMPMSWWPDTWYTFDTS-NH2	3017
JBT0130	1.2	Biotinyl-Ttds-SGRG[CTKVIVFTFRHNKLIGYERRYNC]TS-NH2	3001
JBT0135	>50000	Biotinyl-Ttds-LLYFLTIGNMGMYAAQLKFR-NH2	3021
JBT0136	16461.9	Biotinyl-Ttds-LYFLTIGNMGMYAAQLKFRT-NH2	3014
JBT0137	4120.3	Biotinyl-Ttds-YFLTIGNMGMYAAQLKFRTS-NH2	3013
JBT0138	2354.6	Biotinyl-Ttds-YFLTIGNMGMYAAQLKFR-NH2	3012
JBT0139	755.5	Biotinyl-Ttds-GRGCTKVIVFTFRHNKLIGY-NH2	3008
JBT0140	924.0	Biotin-Ttds-GCTKVIVFTFRHNKLIGYER-NH2	3010
JBT0141	800.3	Biotin-Ttds-CTKVIVFTFRHNKLIGYERR-NH2	3009
JBT0142	>50000	Biotinyl-Ttds-TKVIVFTFRHNKLIGYERRY-NH2	3022
JBT0143	4229.9	Biotinyl-Ttds-KVIVFTFRHNKLIGYERRYN-NH2	3023
JBT0144	1327.0	Biotin-Ttds-VIVFTFRHNKLIGYERRYNC-NH2	3011
JBT0145	650.9	Biotin-Ttds-VIVFTFRHNKLIGYER-NH2	3024

DK/EP 2827883 T3

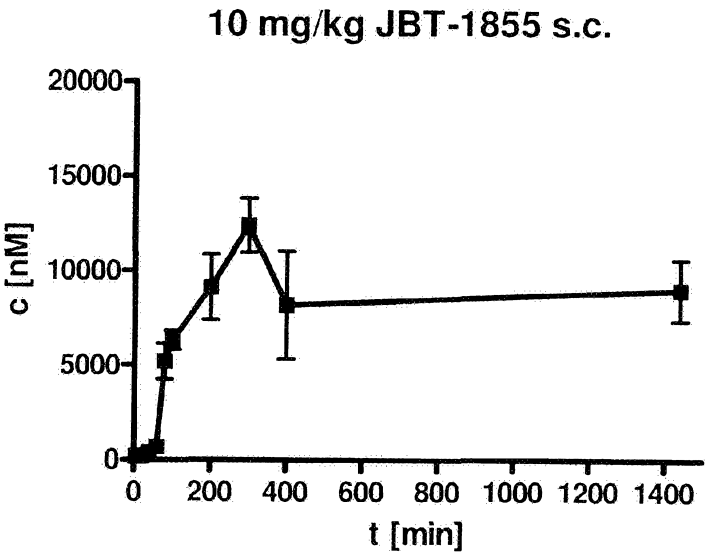

10 mg/kg JBT-1855 s.c.
20000
15000
10000
5000
0
c [nM]
0
200
400
600
800
1000
1200
1400
t [min]

FIGURE 40

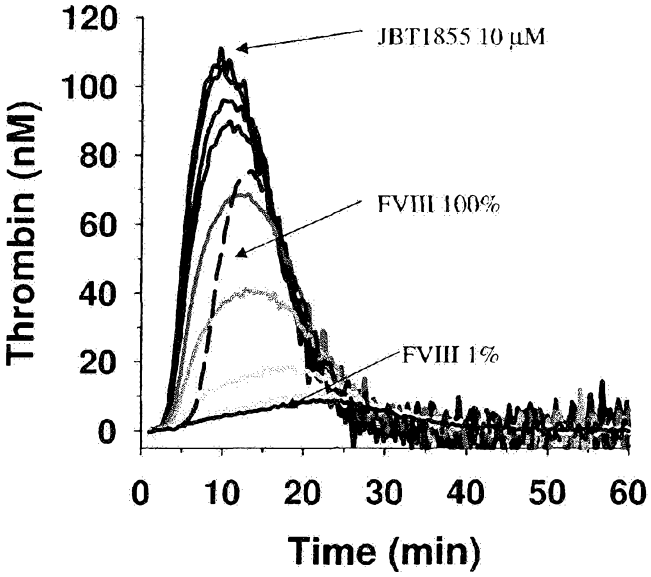

120
100
80
60
40
20
0
Thrombin (nM)
JBT1855 10 µM
FVIII 100%
FVIII 1%
0
10
20
30
40
50
60
Time (min)

FIGURE 41

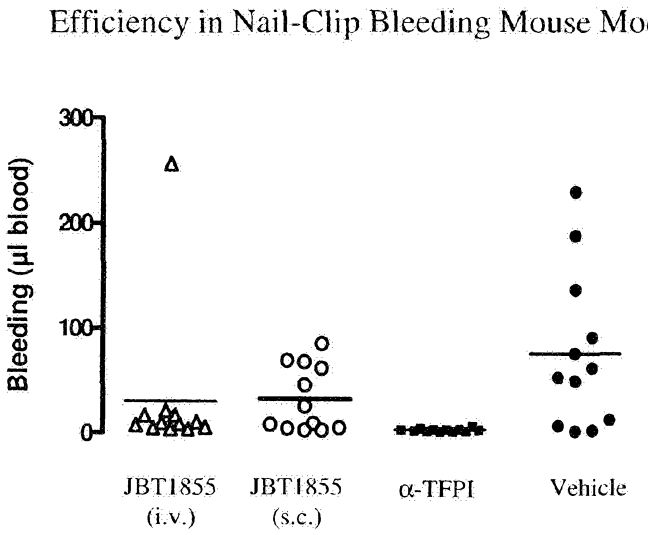

Efficiency in Nail-Clip Bleeding Mouse Model
Bleeding (µl blood)
300
200
100
0
JBT1855 (i.v.)
JBT1855 (s.c.)
α-TFPI
Vehicle

FIGURE 42

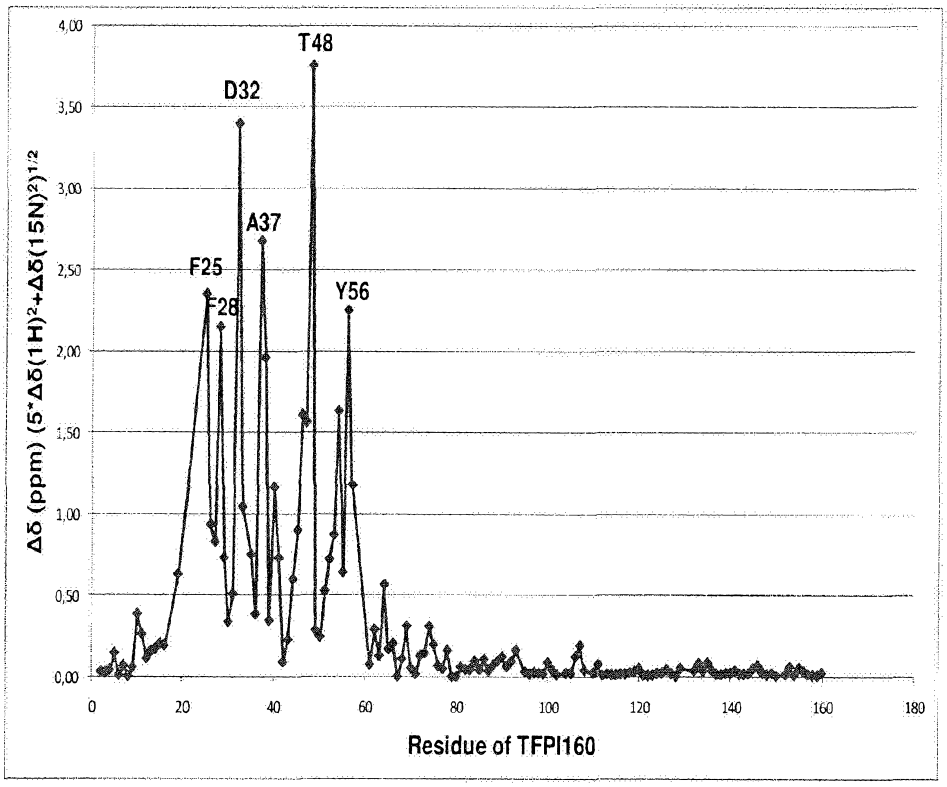

T48
D32
A37
F25
F28
Y56
Δδ (ppm) (5*Δδ(1H)²+Δδ(15N)²)¹/²
Residue of TFPI160
4,00
3,50
3,00
2,50
2,00
1,50
1,00
0,50
0,00
0
20
40
60
80
100
120
140
160
180

FIGURE 43

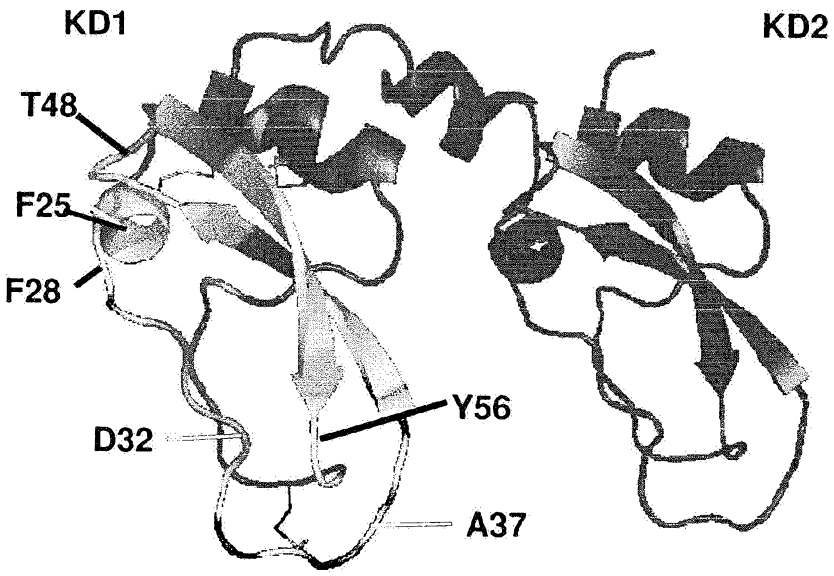


FIGURE 44

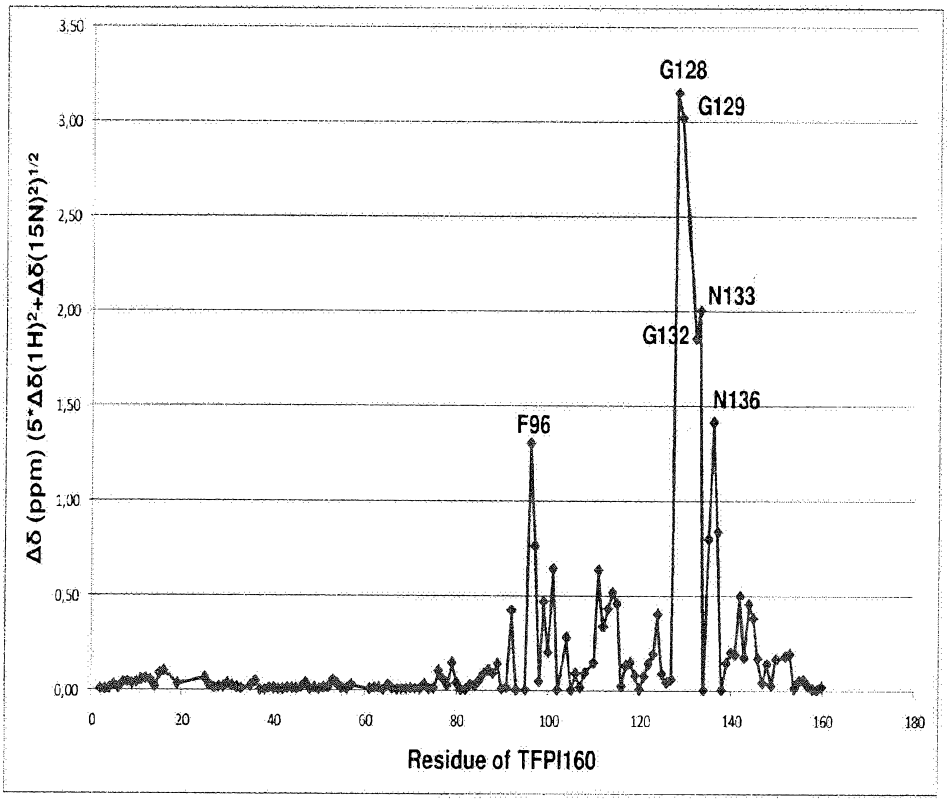

G128
G129
N133
G132
N136
F96
Δδ (ppm) (5*Δδ(1H)²+Δδ(15N)²)¹/²
Residue of TFPI160
3,50
3,00
2,50
2,00
1,50
1,00
0,50
0,00
0
20
40
60
80
100
120
140
160
180

FIGURE 45

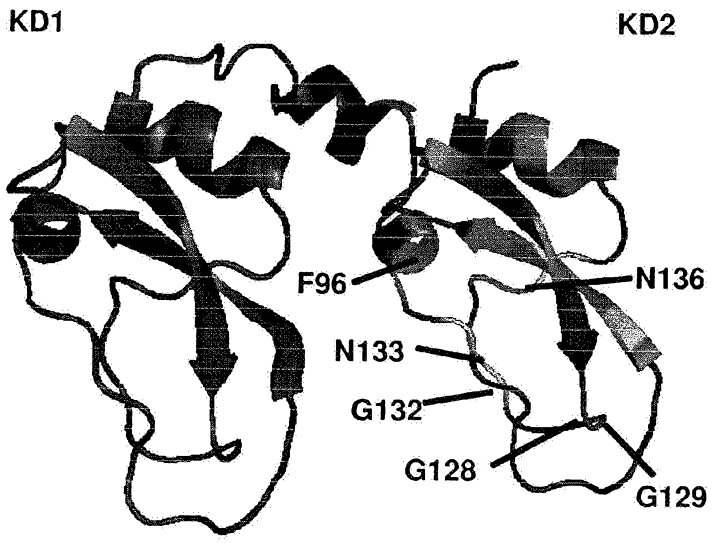


FIGURE 46

	C	CA	CB	H	N
S1	174.9	58.60	-	-	-
G2	173.8	45.18	-	8.447	110.7
Y3	175.6	57.86	38.68	8.116	120.3
A4iso	177.0	52.26	-	8.307	126.1
A4	177.1	52.36	-	8.282	125.9
S5	173.6	58.07	63.54	8.091	115.0
S5iso	-	58.16	-	8.151	115.3
F6	-	55.75	38.85	8.220	122.8
F6iso	-	-	-	8.146	122.6
P7	176.8	63.24	-	-	-
P7iso	175.9	62.53	-	-	-
L8iso	177.3	55.43	-	8.467	123.1
L8	177.4	55.31	42.28	8.354	122.4
A9iso	177.6	52.34	-	8.406	125.4
A9	178.0	52.74	19.15	8.350	124.8
V10	176.3	62.62	32.70	8.082	119.5
V10iso	176.1	62.34	-	8.149	120.0
Q11iso	175.7	55.52	-	8.511	124.6
Q11	176.0	55.88	29.17	8.453	124.0
L12	177.2	55.48	42.23	8.372	123.6
L12iso	-	-	-	8.370	124.2
H13	175.5	56.53	30.30	8.381	120.7
V14	176.3	62.74	32.70	8.142	122.4
S15	174.9	58.71	-	8.570	119.8
K16	177.2	57.38	-	8.531	123.8
R17	177.1	56.86	-	8.351	121.2
S18	175.7	59.62	-	8.330	117.1
K19	177.7	57.91	-	8.435	123.5
E20	178.0	57.92	29.74	8.404	120.6
M21	177.0	56.54	32.63	8.333	120.7
A22	178.9	53.59	18.57	8.164	123.9
L23	177.9	55.94	42.12	8.053	120.3
A24	178.3	53.27	18.79	8.048	123.2
R25	176.5	56.61	30.39	8.008	118.6
L26	177.1	55.50	42.17	7.946	121.8
Y27	175.3	57.84	38.73	8.010	119.5
Y28	175.1	57.66	38.75	7.940	121.5
K29	176.3	56.17	33.20	8.187	123.6
T30	173.9	61.74	69.80	8.316	115.8
S31	-	59.99	64.51	8.012	123.2

FIGURE 47

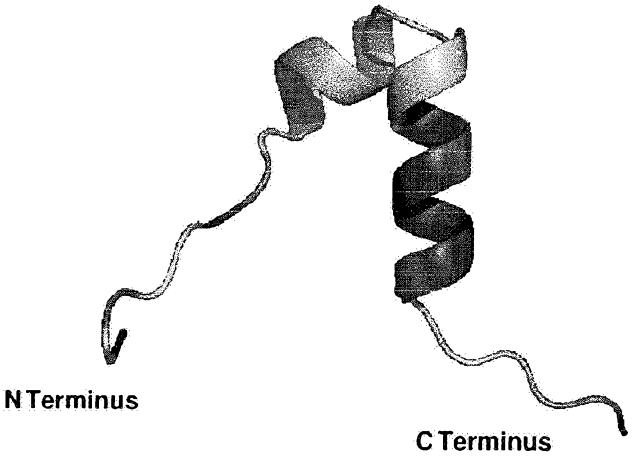


FIGURE 48

	C	CA	H	N
S1	-	61.76	-	-
G2	-	45.19	7.867	109.6
Y3	178.1	56.77	7.856	123.6
A4	179.0	53.67	8.101	121.9
S5	176.2	56.58	7.895	119.9
F6	-	-	-	-
P7	173.7	63.32	-	-
L8	176.6	60.00	8.143	122.0
A9	181.2	55.75	7.938	118.1
V10	177.0	66.76	7.675	115.9
Q11	179.4	59.70	7.519	118.6
L12	179.6	57.91	8.864	119.6
H13	176.3	60.46	7.684	120.4
V14	179.8	65.76	8.178	117.4
S15	176.3	61.89	8.102	116.0
K16	179.7	59.89	7.930	121.8
R17	178.7	56.68	8.718	120.2
S18	177.0	62.62	8.340	115.0
K19	178.3	59.41	7.388	122.2
E20	179.0	59.18	7.566	119.5
M21	176.7	58.97	8.277	116.8
A22	178.1	53.69	7.445	120.4
L23	176.6	57.07	7.745	117.1
A24	177.3	55.82	7.793	119.6
R25	175.5	58.20	7.787	118.5
L26	175.0	57.73	7.868	119.7
Y27	-	56.29	8.038	122.4
Y28	178.1	-	8.161	72.13
K29	-	-	8.897	118.2
T30	173.8	61.71	-	-
S31	-	60.02	7.879	122.7

FIGURE 49

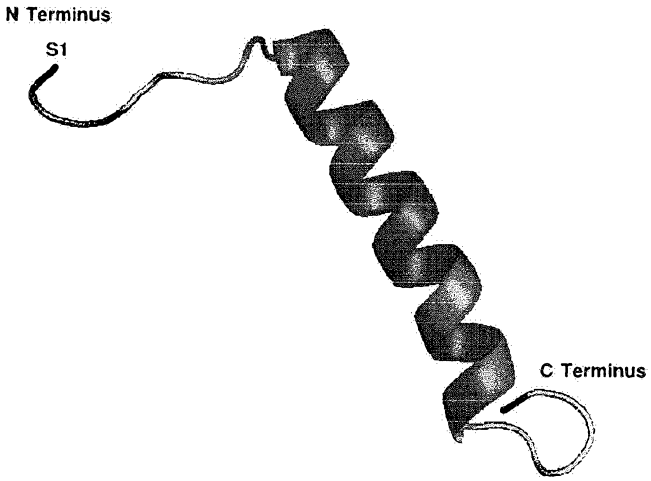


FIGURE 50

	CA	CB	H	N
S-1	56,24	63,73	-	-
F1	58,27	39,61	8,569	122,7
Q2	55,71	29,50	8,381	123,1
S3	58,48	63,67	8,391	117,9
K4	56,41	33,00	8,481	123,6
K5	56,44	33,04	8,371	122,5
N6	53,26	38,80	8,539	120,4
V7	62,31	32,71	8,005	119,9
F8	57,66	39,60	8,395	124,1
V9	62,13	33,05	8,099	122,2
D10	55,06	41,21	8,479	124,1
G11	45,73	-	8,495	110,2
Y12	59,71	38,88	8,110	121,0
F13	59,22	39,25	8,170	120,3
E14	57,68	29,76	8,343	121,8
R15	57,05	30,44	8,130	121,0
L16	55,81	42,18	8,049	121,9
R17	56,35	30,54	8,121	121,1
A18	52,68	19,41	8,144	124,6
K19	56,27	32,87	8,237	121,0
L20	56,70	43,25	7,956	129,8

FIGURE 51

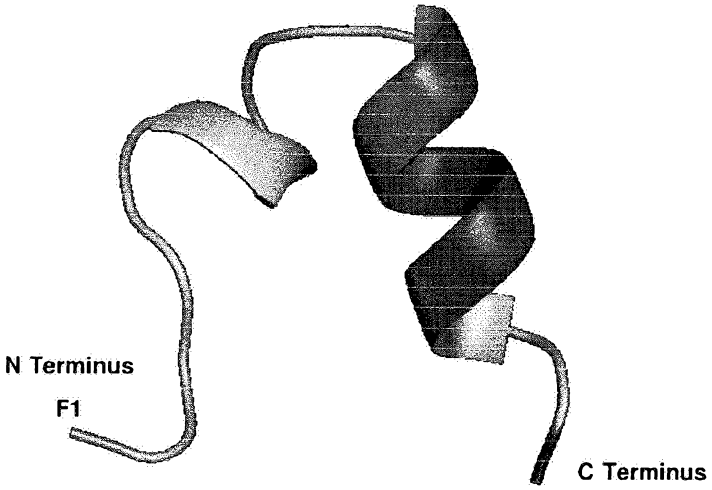


FIGURE 52

	C	CA	H	N
F1	174.5	57.69	8.559	122.1
Q2	176.5	56.60	8.079	116.3
S3	174.2	58.06	8.690	120.0
K4	177.5	58.13	8.519	122.3
K5	176.2	57.27	7.942	116.6
N6	174.1	53.48	8.008	119.2
V7	173.4	62.72	7.775	120.7
F8	177.0	56.11	7.712	117.9
V9	176.0	62.14	8.056	115.2
D10	-	56.04	8.619	123.6
G11	174.2	-	8.496	110.2
Y12	177.3	62.74	8.444	119.2
F13	178.2	62.61	8.342	116.9
E14	179.9	60.23	8.326	122.0
R15	179.4	60.05	8.205	119.5
L16	178.8	57.88	7.770	122.5
R17	178.1	58.84	7.782	117.5
A18	178.0	74.53	7.161	118.7
K19	175.6	57.01	7.722	117.7
L20	-	56.67	7.195	126.2

FIGURE 53

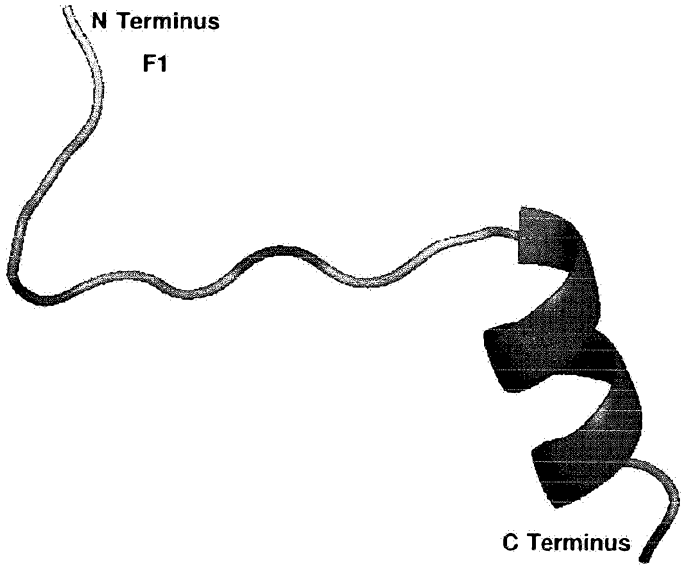


FIGURE 54

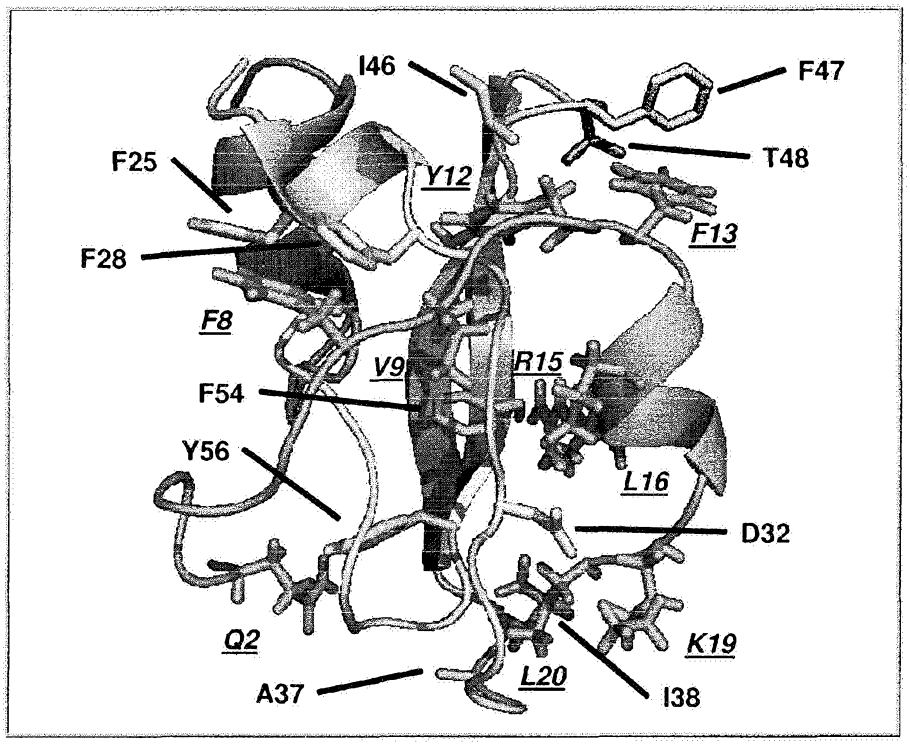
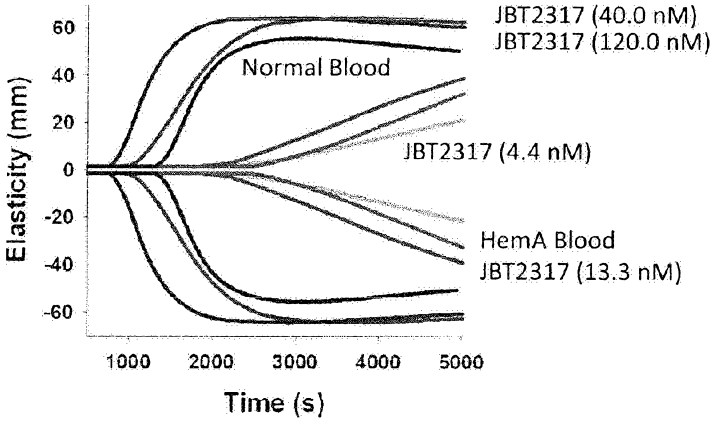
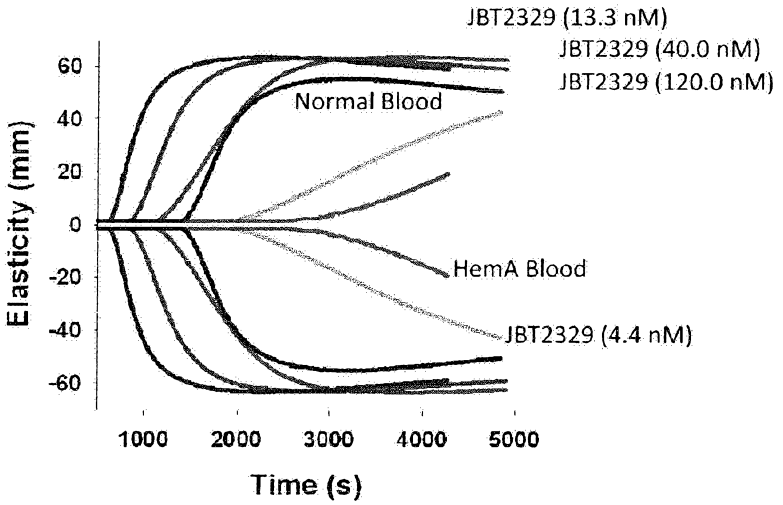

I46
F47
F25
T48
Y12
F13
F28
F8
V9
R15
F54
Y56
L16
D32
Q2
K19
L20
A37
I38

FIGURE 55



	CT (s)	CFT (s)	MCF (mm)	Activity (%)
Hem A blood	2907	1540	-	0
+4.4nM JBT2317	2901	2488	-	0
+13.3nM JBT2317	2249	1480	-	44
+40nM JBT2317	1143	457	64	119
+120nM JBT2317	853	219	64	139
Normal blood	1427	312	56	100

FIGURE 56



	CT (s)	CFT (s)	MCF (mm)	Activity (%)
Hem A blood	3168	1672	-	0
+4.4nM JBT2329	2296	1159	63.5	52
+13.3nM JBT2329	1267	421	63.5	114
+40nM JBT2329	930	238	63.5	134
+120nM JBT2329	674	154	63.5	150
Normal blood	1502	266	55	100

FIGURE 57

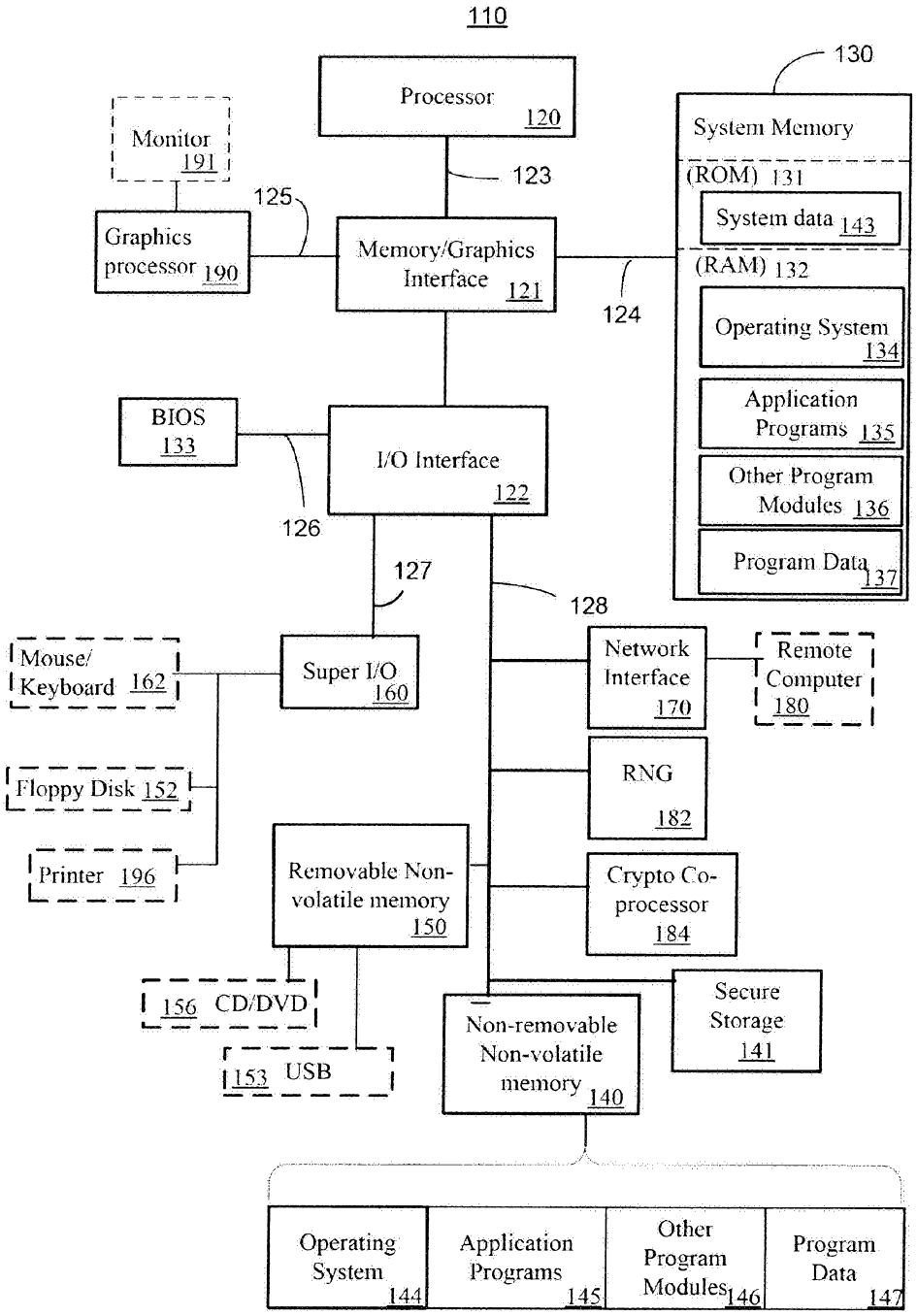

110
Processor 120
130
System Memory
(ROM) 131
System data 143
(RAM) 132
Operating System 134
Application Programs 135
Other Program Modules 136
Program Data 137
Monitor 191
123
125
Graphics processor 190
Memory/Graphics Interface 121
124
BIOS 133
I/O Interface 122
126
127
128
Mouse/ Keyboard 162
Super I/O 160
Network Interface 170
Remote Computer 180
Floppy Disk 152
RNG 182
Printer 196
Removable Non-volatile memory 150
Crypto Co-processor 184
156 CD/DVD
153 USB
Non-removable Non-volatile memory 140
Secure Storage 141
Operating System 144
Application Programs 145
Other Program Modules 146
Program Data 147

FIGURE 58

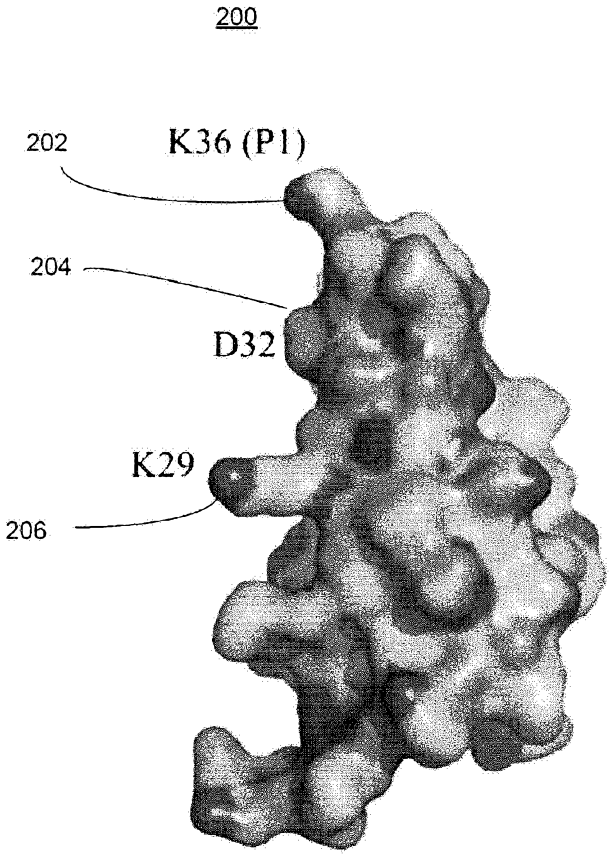


FIGURE 59

300

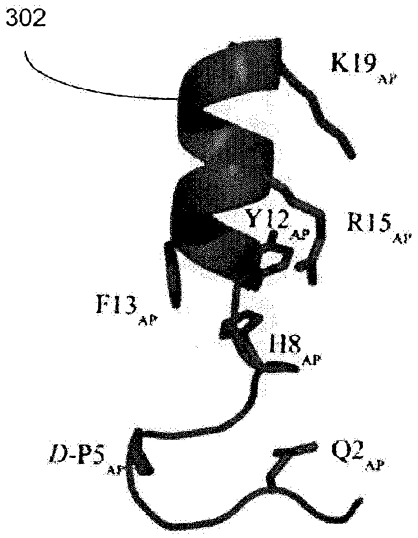


FIGURE 60

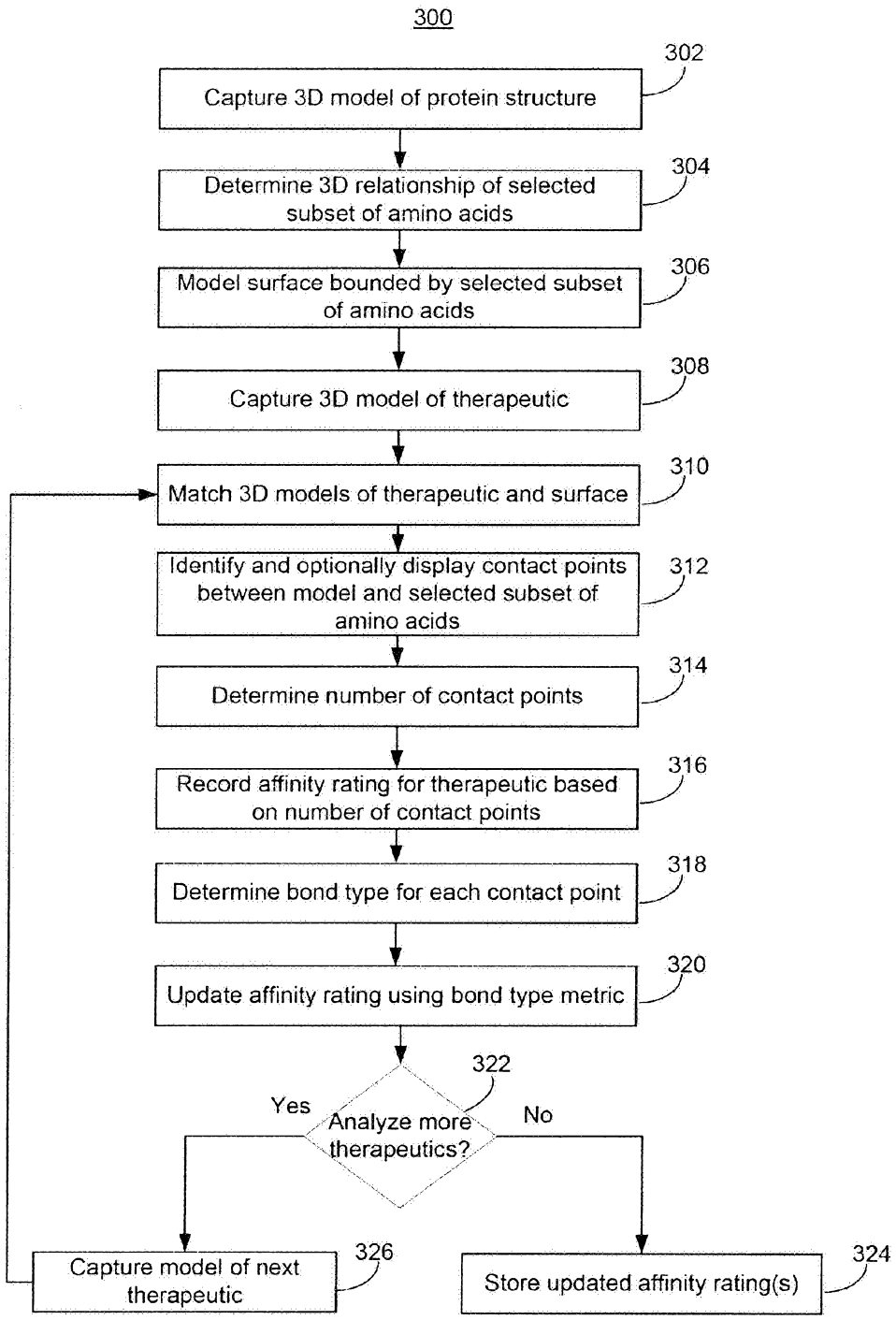

300
302
Capture 3D model of protein structure
304
Determine 3D relationship of selected subset of amino acids
306
Model surface bounded by selected subset of amino acids
308
Capture 3D model of therapeutic
310
Match 3D models of therapeutic and surface
312
Identify and optionally display contact points between model and selected subset of amino acids
314
Determine number of contact points
316
Record affinity rating for therapeutic based on number of contact points
318
Determine bond type for each contact point
320
Update affinity rating using bond type metric
322
Analyze more therapeutics?
Yes
No
326
Capture model of next therapeutic
324
Store updated affinity rating(s)

FIGURE 61

FIGURE 62

		IC50 ELISA	Progression curve		
Compound ID	Sequence	IC50 [nM]	EC50 (µM)	Max. Inh%	SEQ ID NO
JBT0293	Ac-FQSKKNVFVFGYFERLRAKL-NH2	20353.31	---	---	4003
JBT0303	Ac-FQSKKNVFVDGYFERLRAKL-NH2	439.39	---	---	4004
JBT0375	Ac-FQSKGNVFVFGYFERLRAKL-NH2	440.11	---	---	4007
JBT0477	Ac-FQSKGNVFVDGYFERLRAKL-NH2	100.73	---	---	4010
JBT0740	Ac-FQSKGNVFVDGYFERL-Aib-AKL-NH2	34.51	0.245	94	4012
JBT1156	Ac-FQSKaNVFVDGYFARL-Nle-AKL-NH2	416.11	---	---	4014
JBT1319	Ac-YQSKNNVFVDGYFERLCAKL-NH2	137.2	---	---	4015
JBT1320	Ac-FQSKERVFVDGYFERLCAKL-NH2	364.74	---	---	4016
JBT1584	Ac-FQSK-Nmg-NVFVDGYFERL-Aib-AKL-NH2	5.3	---	---	4017
JBT1585	Ac-FQSK-Nmg-NVFVTGYFERL-Aib-AKL-NH2	15.14	---	---	4018
JBT1855	Ac-FQSKGNVFVDGYFERL-Aib-AKLC(PEG)-NH2	121.04	---	---	4019
JBT1857	Ac-FQSKpNVHVDGYFERL-Aib-AKL-NH2	3	0.0037	90	4020
JBT2258	Ac-FQSKKNVFV-NH2		---	---	4022
JBT2262	FAM-Ttds-FQSK-Nmg-NVFVDGYFERL-Aib-AKL-NH2	13.4	---	---	4024
JBT2270	Ac-PEFQSK-Nmg-NVFVDGYFERL-Aib-AKL-NH2	1.94	---	---	4032
JBT2274	Ac-FQSK-NpropylG-NVHVDGYFERL-Aib-AKL-NH2	4.34	---	---	4036
JBT2275	Ac-FQSK-aze-NVHVDGYFERL-Aib-AKL-NH2	2.74	---	---	4037
JBT2276	Ac-FQSK-pip-NVHVDGYFERL-Aib-AKL-NH2	3.27	---	---	4038
JBT2277	Ac-FQSK-tic-NVHVDGYFERL-Aib-AKL-NH2	3.86	---	---	4039
JBT2278	Ac-FQSK-oic-NVHVDGYFERL-Aib-AKL-NH2	2.27	---	---	4040
JBT2279	Ac-FQSK-hyp-NVHVDGYFERL-Aib-AKL-NH2	2.72	---	---	4041
JBT2280	Ac-FQSK-nma-NVHVDGYFERL-Aib-AKL-NH2	4.6	---	---	4042
JBT2281	Ac-FQSK-Ncg-NVHVDGYFERL-Aib-AKL-NH2	6.08	---	---	4043
JBT2282	Ac-FQSK-Abg-NVHVDGYFERL-Aib-AKL-NH2	9.98	---	---	4044
JBT2283	Ac-FQSK-Apg-NVHVDGYFERL-Aib-AKL-NH2	20.12	---	---	4045
JBT2284	Ac-FQSK-thz-NVHVDGYFERL-Aib-AKL-NH2	671.59	---	---	4046
JBT2285	Ac-FQSK-dtc-NVHVDGYFERL-Aib-AKL-NH2	3.44	---	---	4047
JBT2287	Ac-FQSKp-Nmn-VHVDGYFERL-Aib-AKL-NH2		---	---	4049
JBT2288	Ac-FQSKp-Nma-VHVDGYFERL-Aib-AKL-NH2		---	---	4050
JBT2289	Ac-FQSKp-Nmk-VHVDGYFERL-Aib-AKL-NH2		---	---	4051
JBT2290	Ac-FQSKp-Nmr-VHVDGYFERL-Aib-AKL-NH1		---	---	4052
JBT2291	Ac-FQSKp-Nms-VHVDGYFERL-Aib-AKL-NH2	644.27	---	---	4053
JBT2292	Ac-FQSKpnVHVDGYFERL-Aib-AKL-NH2	1124.91	---	---	4054
JBT2293	Ac-FQSKpN-Chg-HVDGYFERL-Aib-AKL-NH2	45.13	---	---	4055
JBT2294	Ac-FQSKpNVH-Chg-DGYFERL-Aib-AKL-NH2	332.48	---	---	4056

FIGURE 62

		IC50 ELISA	Progression curve		
Compound ID	Sequence	IC50 [nM]	EC50 (µM)	Max. Inh%	SEQ ID NO
JBT2295	Ac-FQSKpN-Chg-H-Chg-DGYFERL-Aib-AKL-NH2		---	---	4057
JBT2296	Ac-FQSKpN-Phg-HVDGYFERL-Aib-AKL-NH2	639.17	---	---	4058
JBT2297	Ac-FQSKpNVH-Phg-DGYFERL-Aib-AKL-NH2	66.94	---	---	4059
JBT2298	Ac-FQSKpN-Phg-H-Phg-DGYFERL-Aib-AKL-NH2		---	---	4060
JBT2299	Ac-FQSKpNVHVDGYFERL-Deg-AKL-NH2	3.01	---	---	4061
JBT2300	Ac-FQSKpNVHVDGYFERL-Ebc-AKL-NH2	8.39	---	---	4062
JBT2301	Ac-FQSKpNVHVDGYFERL-Eca-AKL-NH2	3.36	---	---	4063
JBT2302	Ac-FQSKpNVHVDGYFERL-Egz-AKL-NH2	2.7	---	---	4064
JBT2303	Ac-FQSKpNVHVDGYFERL-Aic-AKL-NH2	3.04	---	---	4065
JBT2304	Ac-FQSKpNVHVDGYFERL-Apc-AKL-NH2	2.22	---	---	4066
JBT2305	Ac-FQSKpNVHVDGYFERL-Egt-AKL-NH2	4.48	---	---	4067
JBT2306	Ac-1Ni-QSKpNVHVDGYFERL-Aib-AKL-NH2	3.13	---	---	4068
JBT2307	Ac-Thi-QSKpNVHVDGYFERL-Aib-AKL-NH2	3.41	---	---	4069
JBT2308	Ac-Bta-QSKpNVHVDGYFERL-Aib-AKL-NH2	3.04	---	---	4070
JBT2309	Ac-FQSKpNVHVDGY-1Ni-ERL-Aib-AKL-NH2	1.63	---	---	4071
JBT2310	Ac-FQSKpNVHVDGY-Thi-ERL-Aib-AKL-NH2	3.81	---	---	4072
JBT2311	Ac-FQSKpNVHVDGY-Bta-ERL-Aib-AKL-NH2	1.94	---	---	4073
JBT2312	Ac-FQSKpNVHVDG-Tym-FERL-Aib-AKL-NH2	194.26	---	---	4074
JBT2313	Ac-FQSKpNVHVDG-Pty-FERL-Aib-AKL-NH2	257.97	---	---	4075
JBT2314	Ac-FQSKpNVHVDG-Pmy-FERL-Aib-AKL-NH2	5.14	---	---	4076
JBT2315	Ac-FQSKpNVHVDGYFERL-Aib-AKLC-NH2	2.49	---	---	4077
JBT2317	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(NEM)-NH2	2.91	0.0038	89	4078
JBT2325	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(PEG)-NH2	6.6	---	---	4086
JBT2326	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(PEG)-NH2	9.3	---	---	4087
JBT2327	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(PEG)-NH2	10.92	---	---	4088
JBT2328	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(PEG)-NH2	12.29	---	---	4089
JBT2329	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(ME-400MA)-NH2	12.62	0.0060	93	4090
JBT2366	Ac-Bta-QSKpNVHVDGY-1Ni-ERL-Aib-AKL-NH2	1.99	0.0033	83	4091
JBT2367	Ac-Bta-QSKpNVHVDGY-Bta-ERL-Aib-AKL-NH2	2.05	0.0026	81	4092
JBT2368	Ac-1Ni-QSKpNVHVDGY-1Ni-ERL-Aib-AKL-NH2	2.32	0.0032	78	4093
JBT2369	Ac-1Ni-QSKpNVHVDGY-Bta-ERL-Aib-AKL-NH2	2.03	0.0023	81	4094
JBT2370	Ac-FQSKpCVHVDGYFERL-Aib-AKL-NH2	6.33	---	---	4238
JBT2371	Ac-FQSKpNVHVCGYFERL-Aib-AKL-NH2	10.67	---	---	4095
JBT2372	Ac-FQSKpNVHVDGYCERL-Aib-AKL-NH2	11.68	---	---	4096
JBT2373	Ac-FQSKpNVHVDGYFCRL-Aib-AKL-NH2	4.07	---	---	4097

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FIGURE 62

		IC50 ELISA	Progression curve		
Compound ID	Sequence	IC50 [nM]	EC50 (µM)	Max. Inh%	SEQ ID NO
JBT2381	Ac-FQSKpNVH-Abu-DGYFERL-Aib-AKLC(NEM)-NH2	3.45	n.a.	---	4100
JBT2382	Ac-FQSKpNVH-(L-2-Amino-4,4,4-trifluorobutyric acid)-DGYFERL-Aib-AKLC(NEM)-NH2	62.74	n.a.	---	4101
JBT2383	Ac-FQSKpNVH-Cpg-DGYFERL-Aib-AKLC(NEM)-NH2	52.18	n.a.	---	4102
JBT2384	Ac-FQSKpNVHvDGYFERL-Aib-AKLC(NEM)-NH2		n.a.	---	4103
JBT2385	Ac-FQSKpNVH-Aib-DGYFERL-Aib-AKLC(NEM)-NH2	24.23	n.a.	---	4104
JBT2386	Ac-FQSKpNVH-Tle-DGYFERL-Aib-AKLC(NEM)-NH2	4.23	0.0052	94	4105
JBT2387	Ac-FQSKpNVH-Nmv-DGYFERL-Aib-AKLC(NEM)-NH2		n.a.	---	4106
JBT2388	Ac-FQSKpNVHV-Nmd-GYFERL-Aib-AKLC(NEM)-NH2	9.62	n.a.	---	4107
JBT2389	Ac-FQSKpNVHVHGYFERL-Aib-AKLC(NEM)-NH2	4.4	n.a.	---	4108
JBT2390	Ac-FQSKpNVHVSGYFERL-Aib-AKLC(NEM)-NH2	2.97	n.a.	---	4109
JBT2391	Ac-FQSKpNVHVTGYFERL-Aib-AKLC(NEM)-NH2	3.05	n.a.	---	4110
JBT2392	Ac-FQSKpNVHVPGYFERL-Aib-AKLC(NEM)-NH2	2.62	n.a.	---	4111
JBT2393	Ac-FQSKpNVHVdGYFERL-Aib-AKLC(NEM)-NH2	32.75	n.a.	---	4112
JBT2394	Ac-FQSKpNVH-Tle-PGYFERL-Aib-AKLC(NEM)-NH2	5.53	n.a.	---	4113
JBT2395	Ac-FQSKpNVH-Tle-TGYFERL-Aib-AKLC(NEM)-NH2	4.37	n.a.	---	4114
JBT2396	Ac-FQSKpNVHVDAYFERL-Aib-AKLC(NEM)-NH2	31.54	n.a.	---	4115
JBT2397	Ac-FQSKp-C(NEM)-VHVDGYFERL-Aib-AKL-NH2	2.44	0.0015	87	4116
JBT2398	Ac-FQSKpNVHV-C(NEM)-GYFERL-Aib-AKL-NH2	4.29	0.0164	93	4117
JBT2399	Ac-FQSKpNVHVDGYF-C(NEM)-RL-Aib-AKL-NH2	4.23	0.0131	88	4118
JBT2400	Ac-FQSKpNVHVDGYFERL-Aib-AKLK(Ttds-Maleimidopropionyl(EtSH))-NH2	4.02	0.0465	86	4119
JBT2401	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(Y-MAL-40K)-NH2	22.42	0.0077	91	4120
JBT2402	Ac-FQSKp-C(ME-400MA)-VHVDGYFERL-Aib-AKL-NH2	20.46	0.0026	82	4121
JBT2403	Ac-FQSKpNVHV-C(ME-400MA)-GYFERL-Aib-AKL-NH2	22.89	0.0153	80	4122
JBT2404	Ac-FQSKpNVHVDGYF-C(ME-400MA)-RL-Aib-AKL-NH2	18.18	0.0134	86	4123
JBT2405	[FQSKpNVHVDGYFERL-Aib-AKL-FA19205]	38.76	---	---	4124
JBT2406	[FQSKpNVHVDGYFERL-Aib-AKL-FA19204]	75.35	---	---	4125
JBT2407	[FQSKpNVHVDGYFERL-Aib-AKL-FA19203]	28.48	---	---	4126
JBT2408	[FQSKpNVHVDGYFERL-Aib-AKL-FA03202]	65.52	---	---	4127
JBT2410	Ac-FQSKpNVHVDGYFERL-Aib-AKLK(Ttds-Maleimid-HSA)-NH2	5.09	0.0047	87	4129
JBT2411	Ac-FQSKpKVH-Tle-DGYFERL-Aib-AKL-NH2	4.61	0.0035	90	4130
JBT2412	Ac-FQSKp-Cit-VH-Tle-DGYFERL-Aib-AKL-NH2	6.09	0.0085	90	4131
JBT2413	Ac-FQSKpVVH-Tle-DGYFERL-Aib-AKL-NH2	8.6	0.0224	90	4132
JBT2414	Ac-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	5.99	0.0130	88	4133

FIGURE 62

		IC50 ELISA	Progression curve		
Compound ID	Sequence	IC50 [nM]	EC50 (µM)	Max. Inh%	SEQ ID NO
JBT2415	Ac-FQSKp-C(Acm)-VH-Tle-DGYFERL-Aib-AKL-NH2	3.78	0.0046	89	4134
JBT2416	Ac-FQSKp-Nle-VH-Tle-DGYFERL-Aib-AKL-NH2	7.72	0.0164	85	4135
JBT2418	Ac-FQSKpRVH-Tle-DGYFERL-Aib-AKL-NH2	3.12	0.0049	89	4136
JBT2419	Ac-FQSKpSVH-Tle-DGYFERL-Aib-AKL-NH2	3.23	0.0048	89	4137
JBT2420	Ac-FQSKpIVH-Tle-DGYFERL-Aib-AKL-NH2	7.33	0.0259	94	4138
JBT2421	Ac-FQSKpNVH-Tle-D-Sar-YFERL-Aib-AKL-NH2	19.51	n.a.	---	4139
JBT2422	Ac-FQSKpNVH-Tle-DpYFERL-Aib-AKL-NH2	28.02	n.a.	---	4140
JBT2423	Ac-FQSKpNVH-Tle-DGYFERL-Aib-LKL-NH2	3.56	0.0057	89	4141
JBT2424	Ac-FQSKpNVH-Tle-DGYFERL-Aib--Nle-KL-NH2	4.37	0.0088	85	4142
JBT2425	Ac-FQSKpNVH-Tle-DGYFERL-Aib-A-Nmk-L-NH2		n.a.	---	4143
JBT2426	Ac-FQDKpNVH-Tle-DGYFERL-Aib-AKL-NH2	10.46	n.a.	---	4144
JBT2427	Ac-FQEKpNVH-Tle-DGYFERL-Aib-AKL-NH2	5.24	0.0031	85	4145
JBT2428	Ac-FQVH-Tle-DGYFERL-Aib-AKL-NH2		n.a.	---	4146
JBT2429	Ac-FqVH-Tle-DGYFERL-Aib-AKL-NH2		n.a.	---	4147
JBT2430	Ac-FQSKpNVHVDGYFERL-Aib-AKLK(AOA-40kDa PSA)-NH2	5.6	0.0009	86	4148
JBT2431	Ac-FQSKpNVH-Tle-DGYFERL-Aib-AKL-NH2	4.74	0.0063	83	4149
JBT2432	Ac-FQSKpNVHVPGYFERL-Aib-AKL-NH2	3.76	0.0046	80	4150
JBT2433	Ac-FQSKpNVHVPaYFERL-Aib-AKL-NH2	5.71	0.0178	96	4151
JBT2434	Ac-FQSKpNVHVPGYF-Aib-RL-Aib-AKL-NH2	5.53	0.0106	92	4152
JBT2435	Ac-FQSKpNVH-Tle-DaYFERL-Aic-AKL-NH2	8	0.0125	87	4153
JBT2436	Ac-FQSKpNVH-Tle-DaYFERL-Eca-AKL-NH2	5.69	0.0116	91	4154
JBT2437	Ac-FQSKpNVH-Tle-DaYFERL-Deg-AKL-NH2	5.69	0.0175	89	4155
JBT2438	Ac-FQSKpNVH-Tle-DaYFERL-Aib--Aib-KL-NH2	8.55	0.0214	88	4156
JBT2439	Ac-FQSKpNV-2Ni--Tle-DaYFERL-Aib-AKL-NH2	8.01	0.0059	91	4157
JBT2440	Palm-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2		n.a.	---	4158
JBT2441	Ac-FQSKpNVH-Tle-DaYFER[-Hcy--Aib-AK-Hcy-]-NH2	55.25	n.a.	---	4159
JBT2442	Ac-FQSKpNV-1Ni--Tle-DaYFERL-Aib-AKL-NH2	15.29	0.0270	86	4239
JBT2443	Ac-FQCKpCVH-Tle-DaYFERL-Aib-AKL-NH2	237.49	n.a.	---	4160
JBT2444	Ac-FQSKpNVH-Tle-DaYFER-Hle--Aib-AKL-NH2	12.54	n.a.	---	4161
JBT2445	Ac-FQSKpN-Tle-H-Tle-DaYFERL-Aib-AKL-NH2	129.64	n.a.	---	4162
JBT2446	Ac-FQSKpNVH-Tle-DaYFERL-Aib-A-Har-L-NH2	57.48	n.a.	---	4163
JBT2447	Ac-FQSKpNVH-Tle-DaYFERL-Aib-A-Opc-L-NH2		n.a.	---	4164
JBT2448	Ac-FQSKpNVH-Tle-DaYFERLA-Aib-KL-NH2	9.36	n.a.	---	4165
JBT2449	Ac-FQSKpNVH-Tle-DaYFERLa-Aib-KL-NH2	43.23	n.a.	---	4166
JBT2450	Ac-FQSKpNVH-Tle-DaYF[-Hcy-RLA-Hcy-]KL-NH2	398.31	n.a.	---	4167

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FIGURE 62

		IC50 ELISA	Progression curve		
Compound ID	Sequence	IC50 [nM]	EC50 (µM)	Max. Inh%	SEQ ID NO
JBT2451	Ac-FQSKpEVH-Tle-DaYFERL-Aib-AKL-NH2	7.36	n.a.	---	4168
JBT2452	Ac-FQSKp-Ede(O)-VH-Tle-DaYFERL-Aib-AKL-NH2	5.75	n.a.	---	4169
JBT2453	Ac-FQ-Ede(O)-KpNVH-Tle-DaYFERL-Aib-AKL-NH2	7.9	n.a.	---	4170
JBT2458	374-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	6.17	0.0062	90	4173
JBT2459	1281-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	8.45	0.0096	86	4174
JBT2460	5963-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	10.9	n.a.	---	4175
JBT2461	1525-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	3.74	0.0031	87	4176
JBT2462	4635-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	6.3	0.0093	88	4177
JBT2463	3067-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	4.15	0.0035	86	4178
JBT2464	972-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	7.72	0.0043	84	4179
JBT2465	973-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	6.41	0.0027	82	4180
JBT2466	Ac-FQSKpNVH-Tle-DaYFERL-Aib-[CKL-Cea-]	8.27	0.0212	71	4181
JBT2467	Ac-FQSKpNVH-Tle-DaYFERL-Aib-[cKL-Cea-]	6.4	0.0173	60	4182
JBT2468	Ac-FQSKpNVH-Tle-DaYFERL[CAKL-Cea-]	60.46	n.a.	---	4183
JBT2469	Ac-FQSKpNVH-Tle-DaYFERL[cAKL-Cea-]	14.71	0.0187	73	4184
JBT2470	Ac-FQSKpNVH-Tle-DaYFERL-Aib-AK-Aml--NH2	19.61	n.a.	---	4185
JBT2471	Ac-FQSKpNVH-Tle-DaYFER-Aml-AAKL-NH2	9.34	0.0222	85	4186
JBT2472	Ac-FQSKpNVH-Tle-DaYFERL-Aib-LKL-NH2	7.59	0.0201	87	4187
JBT2473	Ac-FQSKpNVH-Tle-DaYF-Aib-RL-Aib-LKL-NH2	11.92	0.0621	83	4188
JBT2474	Ac-FQSKpNVH-Tle-DaYF-Aib-RL-Aib-AKL-NH2	7.27	0.0383	85	4189
JBT2475	Ac-FQSKpNVHVPaYF-Aib-RL-Aib-AKL-NH2	7.66	0.0350	86	4190
JBT2476	Ac-FQSKpNVHVPaYFERL-Aib-LKL-NH2	5.17	0.0169	81	4191
JBT2477	Ac-FQSKpNVHVPaYF-Aib-RL-Aib-LKL-NH2	13.85	n.a.	---	4192
JBT2478	Ac-FQSKpNVH-Tle-Da-Dopa-FERL-Aib-AKL-NH2	38.36	n.a.	---	4193
JBT2479	Ac-YQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	4.41	0.0092	81	4194
JBT2480	Ac--Dopa-QSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	6.33	n.a.	---	4195
JBT2485	Ac-FQSKpNV-Pmy--Tle-DaYFERL-Aib-AKL-NH2	5.55	0.0084	92	4200
JBT2486	Ac-FQSKpNTH-Tle-DaYFERL-Aib-AKL-NH2	8.11	0.0358	87	4201
JBT2487	Ac-FQSKp-Cmc-VH-Tle-DaYFERL-Aib-AKL-NH2	9.1	0.0150	88	4202
JBT2488	Ac-FQ-Cmc-KpNVH-Tle-DaYFERL-Aib-AKL-NH2	9.04	0.0173	87	4203
JBT2489	Ac-FQSKpNVH-Tle-DaYF[-Hcy-RL-Aib--Hcy-]KL-NH2		n.a.	---	4204
JBT2490	Ac-FQSKpNVH-Tle-DaYF[cRLA-Hcy-]KL-NH2	6165.16	n.a.	---	4205
JBT2491	Ac-FQSKpNVH-Tle-DaYF[CRLA-Hcy-]KL-NH2		n.a.	---	4206
JBT2492	Ac-FQSKpNVH-Tle-DaYF[cRLC]AKL-NH2		n.a.	---	4207
JBT2493	Ac-FQSKpNVH-Tle-DaYF[CRLC]AKL-NH2		n.a.	---	4208

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FIGURE 62

		IC50 ELISA	Progression curve		
Compound ID	Sequence	IC50 [nM]	EC50 (µM)	Max. Inh%	SEQ ID NO
JBT2494	Ac-FQSKpNVH-Tle-D[cYF-Hcy-]RL-Aib-AKL-NH2	2.84	0.0020	88	4209
JBT2495	Ac-FQSKpNVH-Tle-D[cYFC]RL-Aib-AKL-NH2	3.48	0.0021	90	4210
JBT2496	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(Ac-FQSKpNVHVDGYFERL-Aib-AKLK(Ttds-Maleiimidopropionyl)-NH2)--NH2	0.75			4211
JBT2497	Ac-FQSKpNVH-Tle-D[-hcy-YF-Hcy-]RL-Aib-AKL-NH2	7.16			4212
JBT2498	Ac-FQSKpNVH-Tle-D[-hcy-YFC]RL-Aib-AKL-NH2	7.59			4213
JBT2499	Ac-FQSKpNVHVP[cYF-Hcy-]RL-Aib-AKL-NH2	3.89			4214
JBT2500	Ac-FQSKpNVH-Tle-DcY-Bta--Hcy-RL-Aib-AKL-NH2	6.03			4215
JBT2501	Ac-FQSKp-C(Acm)-VH-Tle-DcY-Bta--Hcy-RL-Aib-AKL-NH2	3.89			4216
JBT2502	Ac-FQSKp-C(NEM)-VH-Tle-DaY-Bta-ERL-Aib-AKL-NH2	4.1			4217
JBT2503	Ac-FQSKp-C(NEM)-VHVPaY-Bta-ERL-Aib-AKL-NH2	5.04			4218
JBT2504	Ac-FQSKpNVH-Tle-DaYFER-Aml--Aib-AKL-NH2	9.43			4219
JBT2505	Ac-FQSKp-Ecl-VHVPaYFERL-Aib-AKL-NH2	5.82			4220
JBT2506	Ac-FQSKp-Eea-VHVPaYFERL-Aib-AKL-NH2	6.01			4221
JBT2507	Ac-FQSKp-Eec-VHVPaYFERL-Aib-AKL-NH2	9.71			4222
JBT2508	Ac-FQSKp-Eef-VHVPaYFERL-Aib-AKL-NH2	31.05			4223
JBT2509	Ac-FQSKp-Nif-VHVPaYFERL-Aib-AKL-NH2	19.69			4224
JBT2510	Ac-FQSKp-Eew-VHVPaYFERL-Aib-AKL-NH2	8.86			4225
JBT2511	Ac-FQSKpNVH-Tle-DaY-Bta-ERL-Aib-cKL-Cea-	189.24			4226
JBT2512	Ac-FQSKp-C(Acm)-VH-Tle-DaY-Bta-ERL-Aib-cKL-Cea-	157.63			4227
JBT2513	Ac-FQSKpNVH-Tle-DaYFERL-Aib-[-hcy-KL-Cea-]	317.99			4228
JBT2514	Ac-FQSKpNVH-Tle-DaYFERL-Aib-cKL-Hcy--NH2	132.62			4229
JBT2515	Ac-FQSKpNVH-Tle-DaYFERL-Aib-[cKLC]-NH2	579.9			4230
JBT2516	Ac-FQSKpNVH-Tle-DaYFERL-Aib-[cKLc]-NH2	472.87			4231
JBT2517	Ac-FQSKpNVHVPaY-Bta-ERL-Aib-cKL-Cea-	269.09			4232
JBT2518	Ac-FQSKpNVHVPcY-Bta--Hcy-RL-Aib-AKL-NH2	10.78			4233

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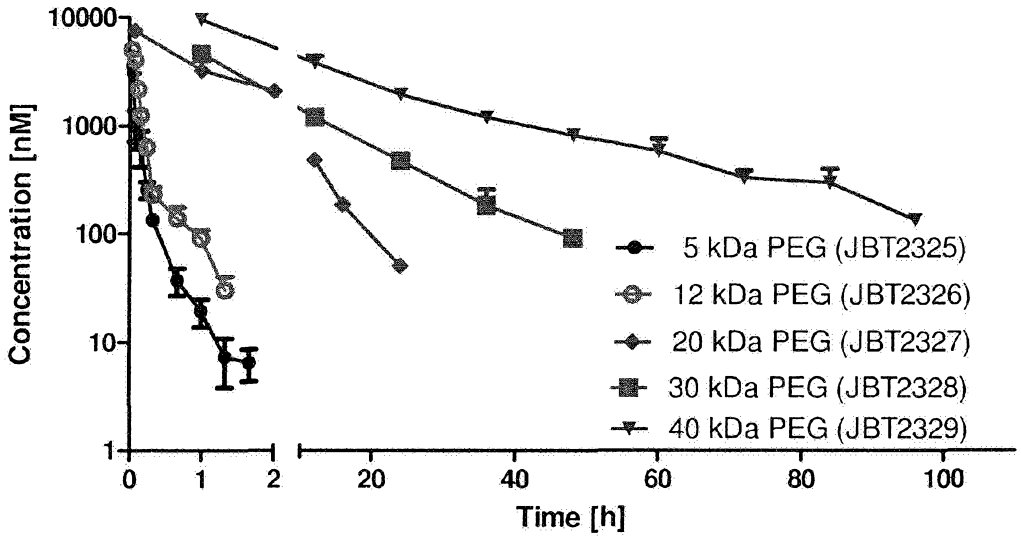

FIGURE 63

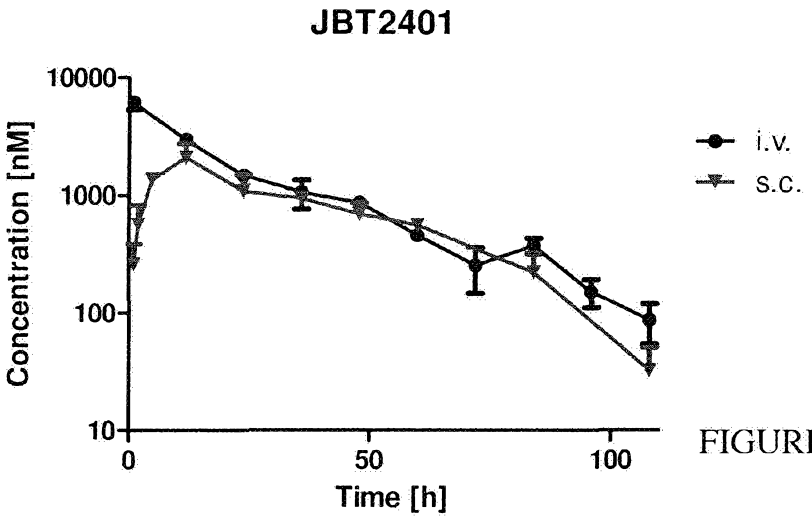

FIGURE 64A

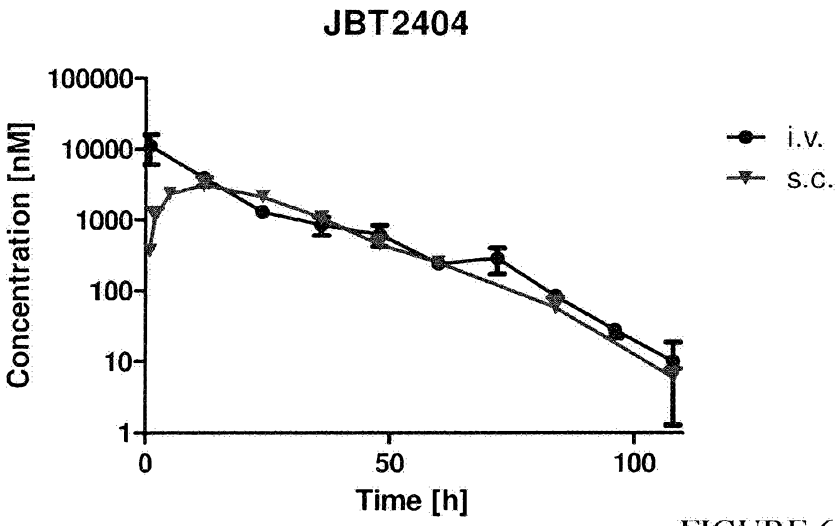

FIGURE 64B

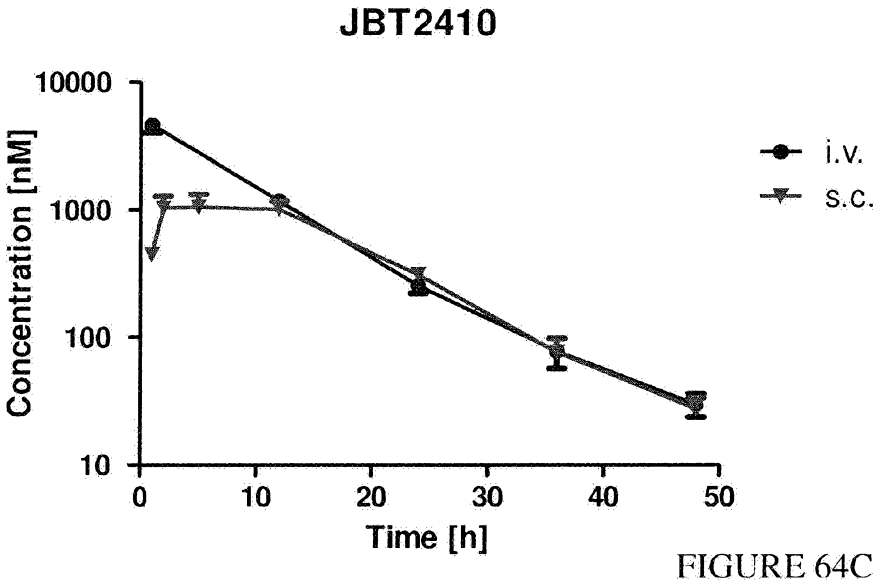

FIGURE 64C

FIGURE 65

JBT #	SEQ ID NO	Sequence
0292	4002	Ac-KKSGVGRLQVAFQSKKNVFVFGYFERLRAKLTSK-Lys(FAM)-NH2
2257	4021	Biotinyl-Ttds-QKVRFVLDKEAGSFLYRGFN-NH2
2261	4023	Biotinyl-Ttds-FQSK-Nmg-NVFVDGYFERL-Aib-AKL-NH2
2263	4025	Ac-FK-Nmg-VSAEYDVGLL-Aib-KFFQRN-NH2
2264	4026	Biotin-Ttds-FK-Nmg-VSAEYDVGLL-Aib-KFFQRN-NH2
2265	4027	FAM-Ttds-FK-Nmg-VSAEYDVGLL-Aib-KFFQRN-NH2
2266	4028	Ac-PFQSKGNVFVDGYFERLRAKL-NH2
2267	4029	Ac-PEFQSKGNVFVDGYFERLRAKL-NH2
2268	4030	Ac-PFQSK-Nmg-NVFVDGYFERL-Aib-AKL-NH2
2269	4031	Ac-YQTK-Nmg-NVFVDGYFERL-Aib-AKL-NH2
2271	4033	Biotinyl-Ttds-FQSKpNVHVDGYFERL-Aib-AKL-NH2
2272	4034	Ac-GKRLKFD-Nmg-YLFNEAVQSF-Aib-V-NH2
2273	4035	Ac-Nmg-LYFKANF-Aib-SVDQFEVGRLK-NH2
2286	4048	Ac-FQSKpPVHVDGYFERL-Aib-AKL-NH2
2318	4079	Ac-FQSKpNVHVDGYFERL-Aib-AKL-Ttds-Lys(biotinyl)-NH2
2319	4080	Ac-VF-Aib-EVLKFGHALKpNSQDRY-NH2
2320	4081	Ac-FpDYVER-Aib-KGFLVHQKLASN-NH2
2321	4082	Biotinyl-Ttds-VF-Aib-EVLKFGHALKpNSQDRY-NH2
2322	4083	Biotinyl-Ttds-FpDYVER-Aib-KGFLVHQKLASN-NH2
2323	4084	Ac-VF-Aib-EVLKFGHALKpNSQDRY-Ttds-Lys(biotinyl)-NH2
2324	4085	Ac-FpDYVER-Aib-KGFLVHQKLASN-Ttds-Lys(biotinyl)-NH2
2374	4098	Ac-FQSKpNVHVDGYFERL-Aib-AKL-K(Ttds-Maleimidopropionyl)--NH2
2375	4099	Ac-FQSKpNVHVDGYFERL-Aib-AKLK(Aoa)-NH2
2409	4128	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(20 kDa PSA)-NH2
2454	4171	FAM-Ttds-FQSKpNVHVDGYFERL-Aib-AKL-NH2
2455	4172	FAM-Ttds-VF-Aib-EVLKFGHALKpNSQDRY-NH2
2481	4196	Ac-FQSKp-C(NEM)-VH-Tle-DaY-Bta-ERL-Aib-AKL-NH2
2482	4197	Ac-FQSK-oic--C(NEM)-VH-Tle-DaY-Bta-ERL-Aib-AKL-NH2
2483	4198	Ac-FQSKp-C(NEM)-VHVPaY-Bta-ERL-Aib-AKL-NH2
2484	4199	Ac-FQSK-oic--C(NEM)-VHVPaY-Bta-ERL-Aib-AKL-NH2
0621	1294	Ac-SRYKWF[Dab-GMRDMKGTMS-D]VWVKF-NH2
0622	1295	Ac-SRYKWF[Orn-GMRDMKGTMS-D]VWVKF-NH2
0624	1296	Ac-SRYKWF[homoK-GMRDMKGTMS-D]VWVKF-NH2
1809	1297	Ac-SHFKWH[CAMRDMKGTMSC]VWVKF-NH2
1810	1298	Ac-SYYKWH[CGMRDMKGTMSC]VWVKF-NH2
1814	1299	Ac-SQYKWH[CAMRDMKGTMSC]VWVKW-NH2
1818	1300	Ac-SYYKWH[CGMRDMKGTYSC]VWVKF-NH2

FIGURE 65

1819	1301	Ac-SHYKWH[CAMRDMKGTMSC]VWVKF-NH2
1820	1302	Ac-SHYKWH[CAMRDMKGTMSC]VWVKS-NH2
1823	1303	Ac-SHYKWH[CAMRDMNGTMSC]VWVKF-NH2
1824	1304	Ac-SYYKRH[CGMRDMKGTMSC]VWVKF-NH2
1827	1305	Ac-SRYKWH[CAMRDMAGTMSC]VWVKE-NH2
1833	1306	Ac-SYYKWH[CGMRDMKGTMSC]AWVKF-NH2
1834	1307	Ac-SHYKWH[CAMRDMTGTMSC]VWVKF-NH2
1835	1308	Ac-HYYKWH[CGMLDMKGTMTC]VWVKF-NH2
1836	1309	Ac-HYYKWH[CGMRDMKGTMSC]VWVKF-NH2
1838	1310	Ac-HYYKWH[CGMRDMKGTYSC]VWVKF-NH2
1839	1311	Ac-SYYKWH[CGMRDMKGIYSC]VWVKF-NH2
1841	1312	Ac-SYYKWH[CAMRDMKGTMSC]VWVKF-NH2
2316	1313	Biotinyl-Ttds-SYYKWH[CAMRDMKGTMTC]VWVKF-NH2
2339	1314	Ac-C(NEM)-YYKWH[CAMRDMKGTMTC]VWVKF-NH2
2340	1315	Ac-C(NEM)-YKWH[CAMRDMKGTMTC]VWVKF-NH2
2343	1316	Pyn-SYYKWH[CAMRDMKGTMTC]VWVKF-NH2
2344	1317	Pyn-YYKWH[CAMRDMKGTMTC]VWVKF-NH2
2345	1318	Pyn-YKWH[CAMRDMKGTMTC]VWVKF-NH2
2349	1319	H-YYKWH[CAMRDMKGTMTC]VWVKF-NH2
2350	1320	H-SYYKWH[CAMRDMKGTMTC]VWVKF-NH2
2351	1321	Ac-SYYKWH[CA-Nle-RDMKGTMTC]VWVKF-NH2
2352	1322	Ac-SYYKWH[CAMRD-Nle-KGTMTC]VWVKF-NH2
2353	1323	Ac-SYYKWH[CAMRDMKGT-Nle-TC]VWVKF-NH2
2354	1324	Ac-SYYKWH[CA-Nle-RD-Nle-KGT-Nle-TC]VWVKF-NH2
2355	1325	Ac-SYYKWH[CA-Moo-RDMKGTMTC]VWVKF-NH2
2356	1326	Ac-SYYKWH[CAMRD-Moo-KGTMTC]VWVKF-NH2
2357	1327	Ac-SYYKWH[CAMRDMKGT-Moo-TC]VWVKF-NH2
2358	1328	Ac-SYYKWH[CA-Moo-RD-Moo-KGT-Moo-TC]VWVKF-NH2
2359	1329	Ac-SYYKWH[CAMRDLKGTMTC]VWVKF-NH2
2360	1330	Ac-SYYKWH[CAMRDMKGTFTC]VWVKF-NH2
2376	1331	Ac-SYYKWH[CAMRDLKGTFTC]VWVKF-NH2
2377	1332	Ac-SYYKWH[CA-Nle-RDLKGTFTC]VWVKF-NH2
2378	1333	Ac-SYYKWH[CA-Nle-RD-Nle-KGTFTC]VWVKF-NH2
2379	1334	Ac-SYYKWH[CA-Moo-RDLKGTFTC]VWVKF-NH2
2380	1335	Ac-SYYKWH[CA-Moo-RDLKGT-Nle-TC]VWVKF-NH2
2417	1336	PEG2000-CH2-YYKWH[CAMRDMKGTMTC]VWVKF-NH2

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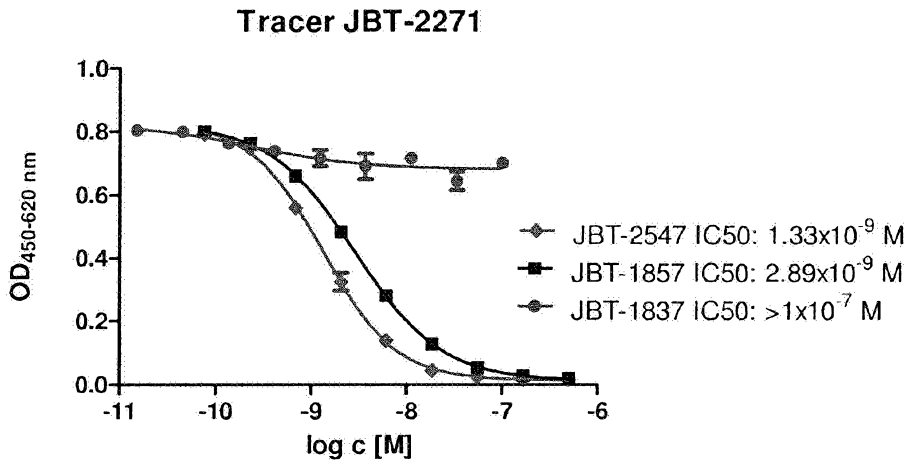


FIGURE 66A

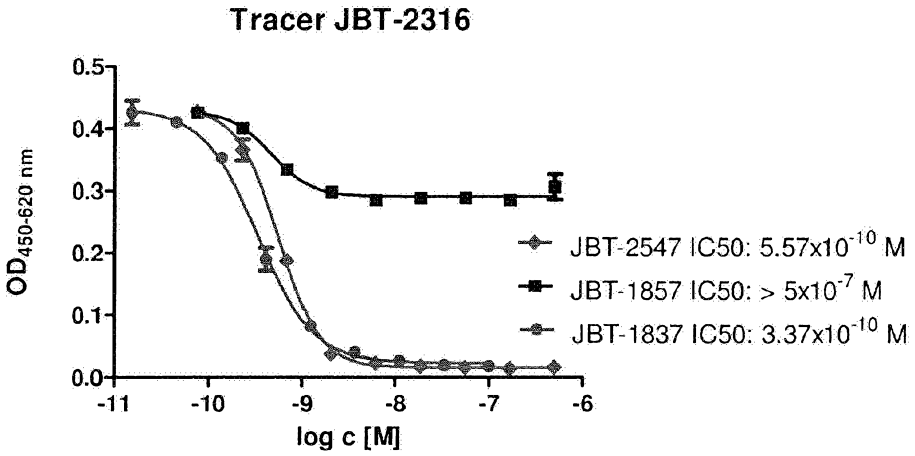


FIGURE 66B

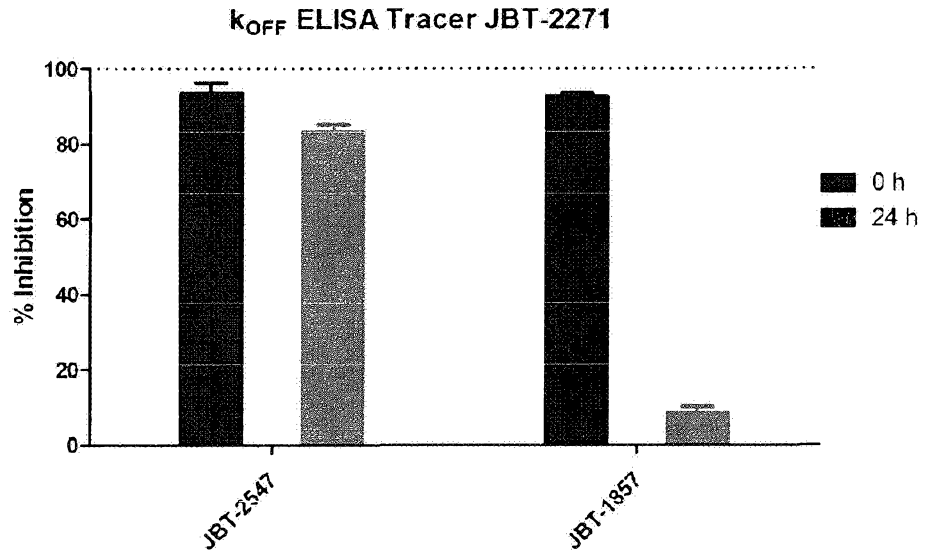

k_{OFF} ELISA Tracer JBT-2271
% Inhibition
100
80
60
40
20
0
0 h
24 h
JBT-2547
JBT-1857

FIGURE 67

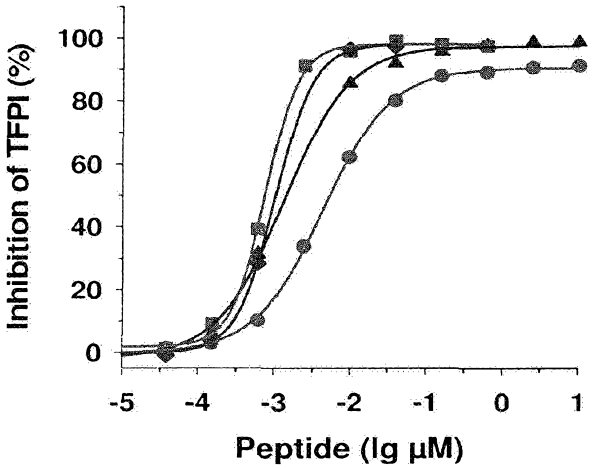


FIGURE 68A

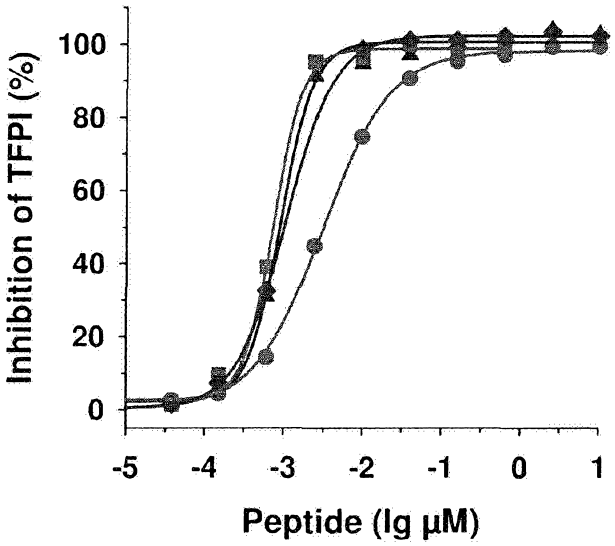


FIGURE 68B

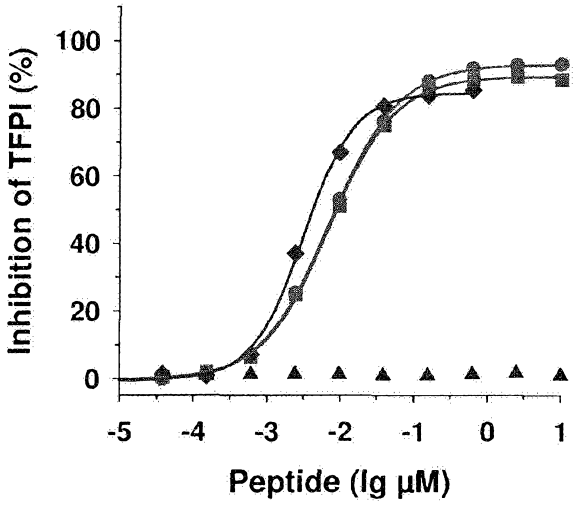


FIGURE 68C

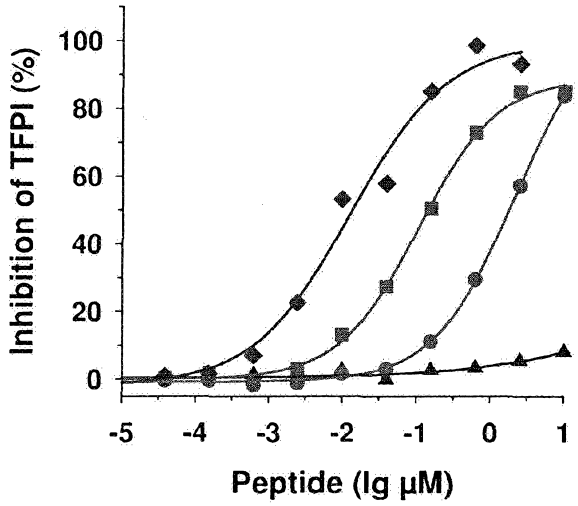
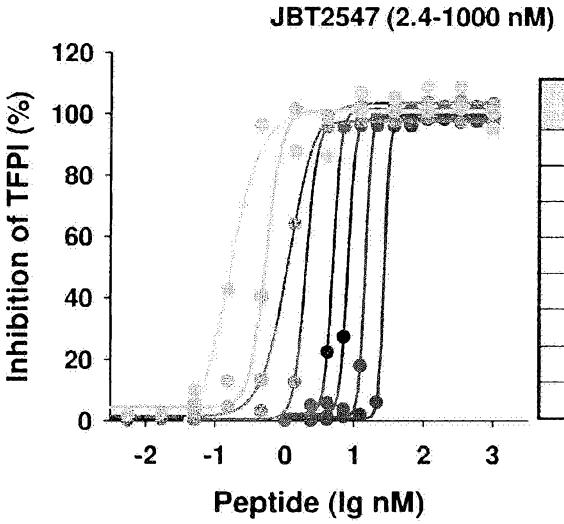


FIGURE 68D



TFPI (nM)	EC50 (nM)	Max. Inhibition (%)
0.1	0.2	101
0.3	0.5	101
0.5	1.0	104
1	2.0	98
2	5.0	101
3	8.1	98
5	14.6	100
10	27.1	98

FIGURE 68E

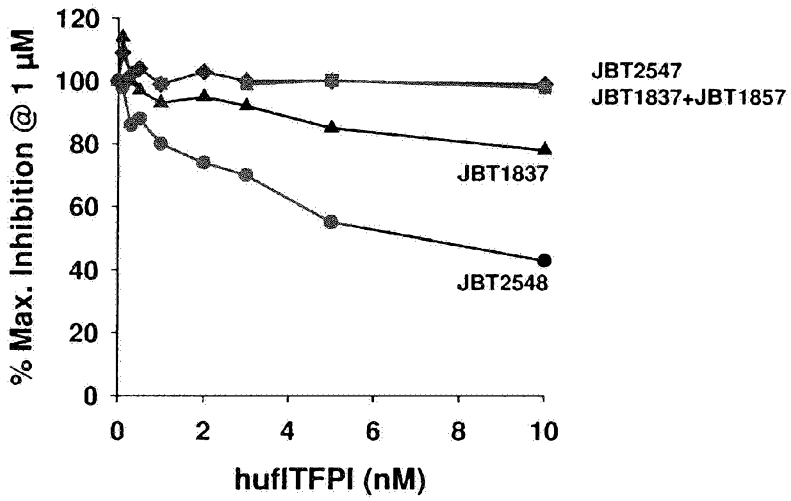


FIGURE 68F

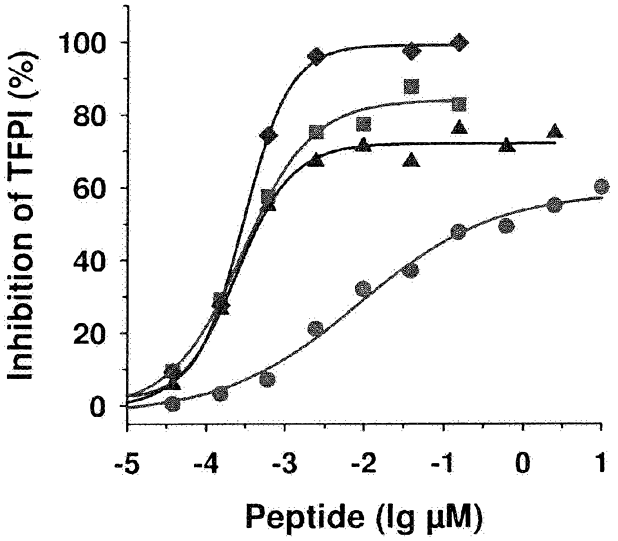
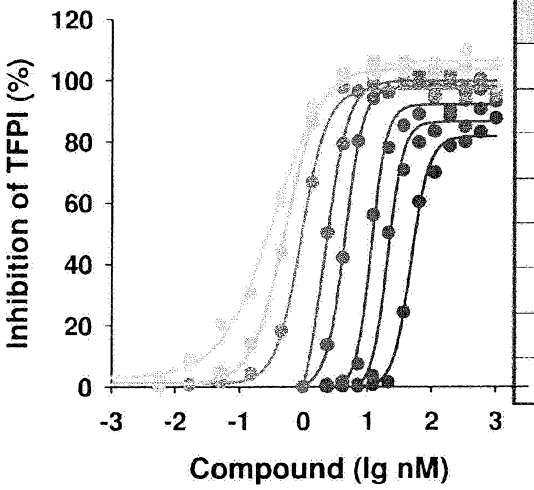


FIGURE 69A

JBT2547 (2.4-1000 nM)



TFPI (nM)	EC50 (nM)	Max. Inhibition (%)
0.0313	0.3	106
0.0625	0.5	104
0.25	0.9	97
0.5	2.1	100
1	4.4	98
3	11.5	92
5	20.9	87
10	48.7	82

FIGURE 69B

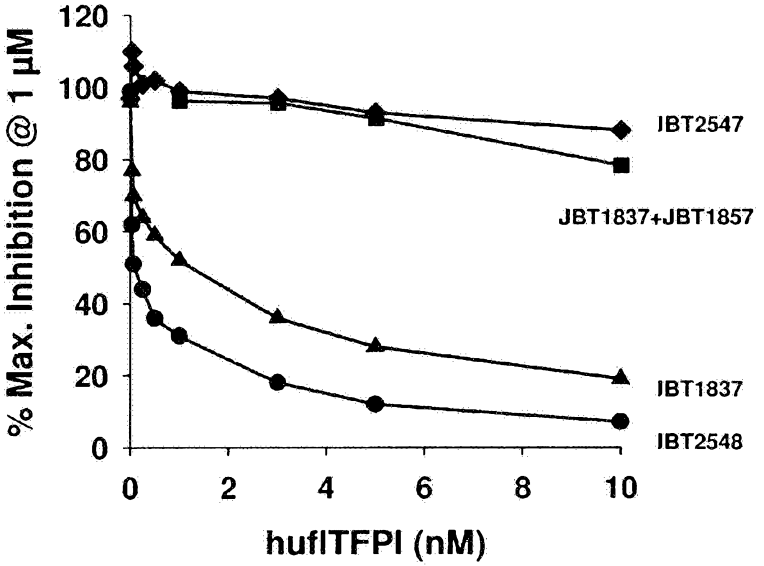


FIGURE 69C

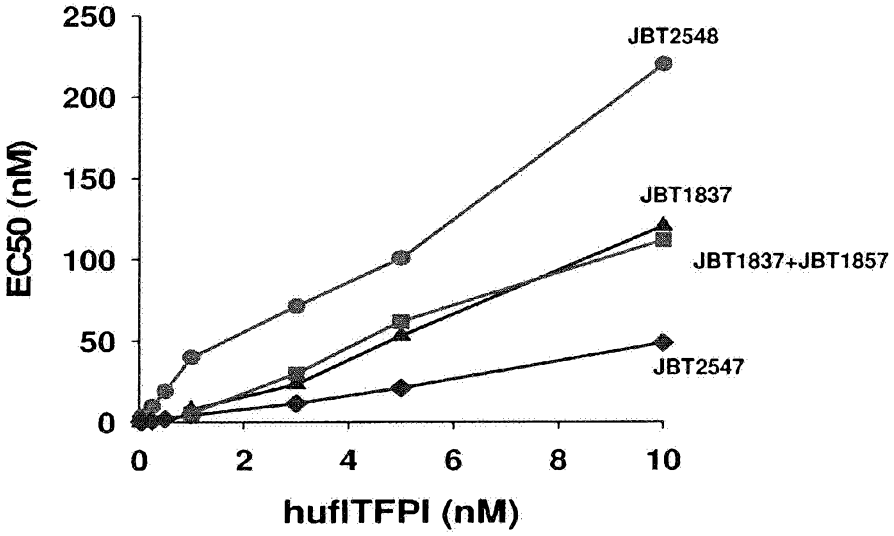


FIGURE 69D

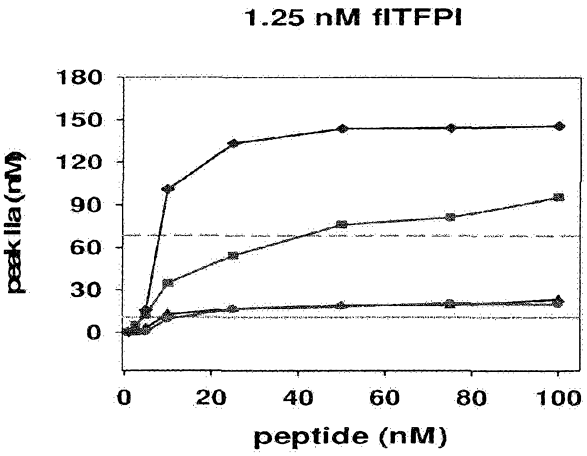


FIGURE 70A

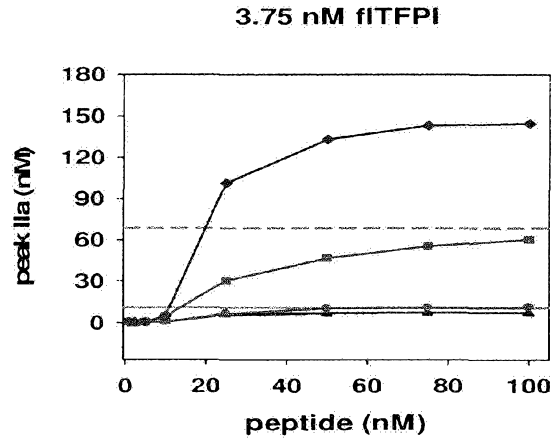


FIGURE 70B

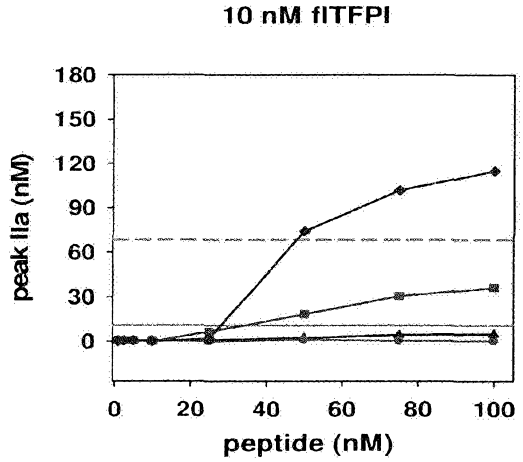


FIGURE 70C

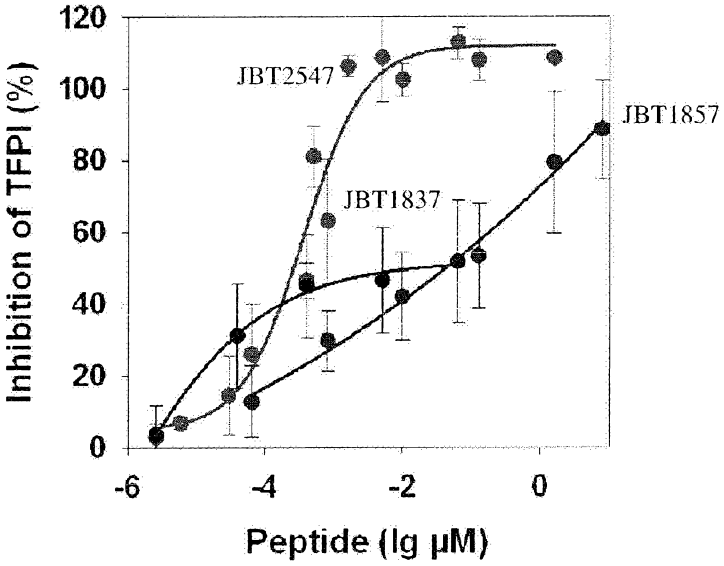


FIGURE 71A

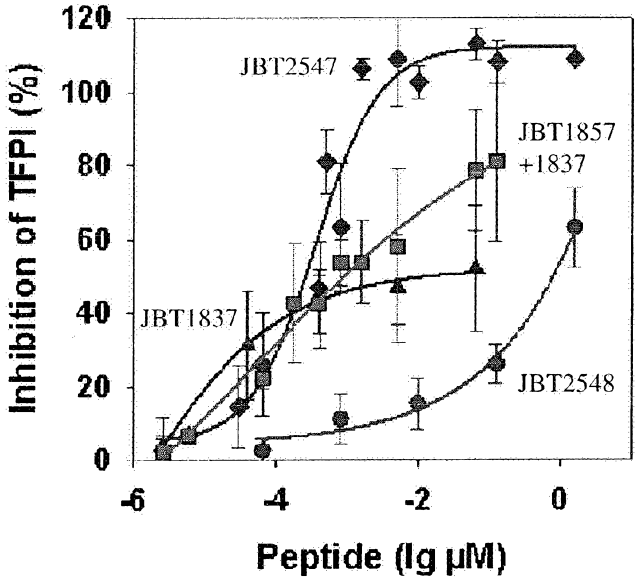


FIGURE 71B

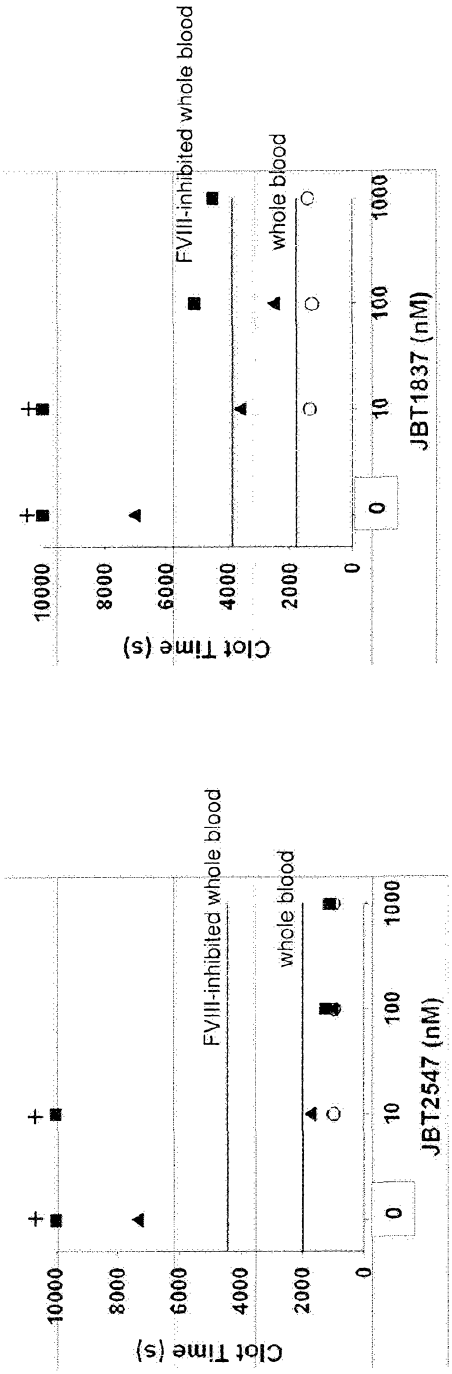


FIGURE 72A

FIGURE 72B

FIGURE 72C

FIGURE 73

Both Nucleophilic		Both Electrophilic		Nucleophilic/Electrophilic	
Reactive Group 1	**Reactive Group 2**	**Reactive Group 1**	**Reactive Group 2**	**Reactive Group 1**	**Reactive Group 2**
amino	Amino	carboxyl	carboxyl	amino	carboxyl
amino	Thiol	carboxyl	acyl chloride	amino	acyl chloride
amino	hydroxyl	carboxyl	anhydride	amino	anhydride
thiol	Amino	carboxyl	Ester	amino	ester
thiol	Thiol	carboxyl	NHS	amino	NHS
thiol	hydroxyl	carboxyl	Halogen	amino	halogen
hydroxyl	Amino	carboxyl	sulfonate ester	amino	sulfonate ester
hydroxyl	Thiol	carboxyl	maleimido	amino	maleimido
hydroxyl	hydroxyl	carboxyl	haloacetyl	amino	haloacetyl
		carboxyl	isocyanate	amino	isocyanate
		acyl chloride	carboxyl	thiol	carboxyl
		acyl chloride	acyl chloride	thiol	acyl chloride
		acyl chloride	anhydride	thiol	anhydride
		acyl chloride	Ester	thiol	ester
		acyl chloride	NHS	thiol	NHS
		acyl chloride	Halogen	thiol	halogen
		acyl chloride	sulfonate ester	thiol	sulfonate ester
		acyl chloride	maleimido	thiol	maleimido
		acyl chloride	haloacetyl	thiol	haloacetyl
		acyl chloride	isocyanate	thiol	isocyanate
		anhydride	carboxyl	hydroxyl	carboxyl
		anhydride	acyl chloride	hydroxyl	acyl chloride
		anhydride	anhydride	hydroxyl	anhydride
		anhydride	Ester	hydroxyl	ester
		anhydride	NHS	hydroxyl	NHS
		anhydride	Halogen	hydroxyl	halogen
		anhydride	sulfonate ester	hydroxyl	sulfonate ester
		anhydride	maleimido	hydroxyl	maleimido
		anhydride	haloacetyl	hydroxyl	haloacetyl
		anhydride	isocyanate	hydroxyl	isocyanate
		ester	carboxyl		
		ester	acyl chloride		
		ester	anhydride		
		ester	Ester		
		ester	NHS		
		ester	Halogen		
		ester	sulfonate ester		
		ester	maleimido		
		ester	haloacetyl		
		ester	isocyanate		

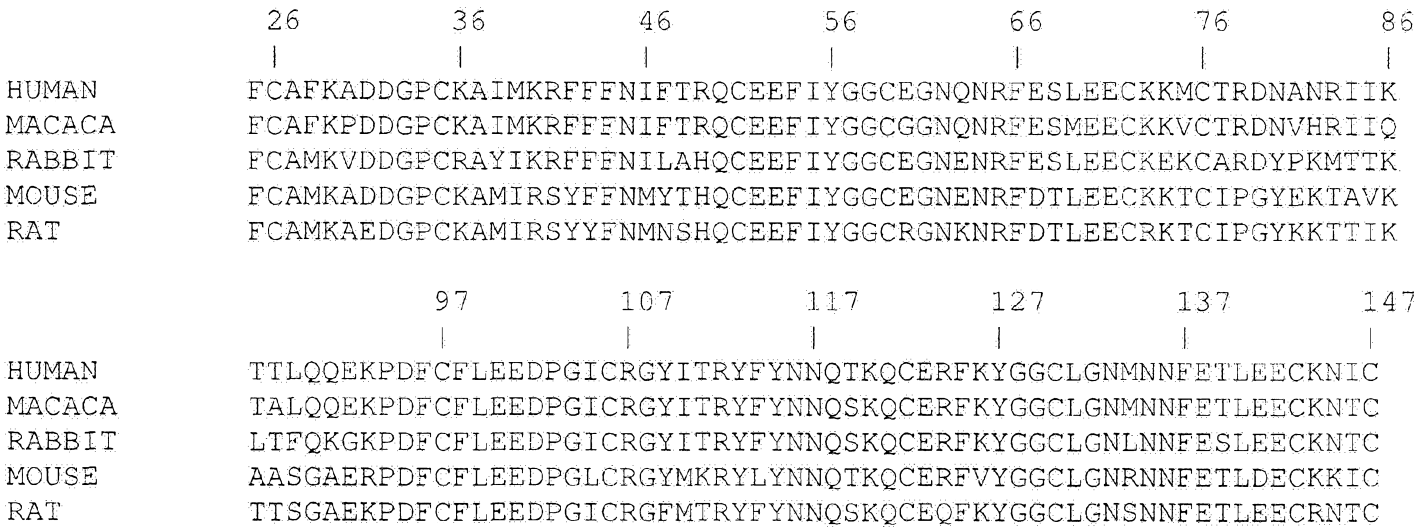
FIGURE 73 Continued…

Both Nucleophilic		Both Electrophilic		Nucleophilic/Electrophilic	
Reactive Group 1	**Reactive Group 2**	**Reactive Group 1**	**Reactive Group 2**	**Reactive Group 1**	**Reactive Group 2**
		NHS	carboxyl		
		NHS	acyl chloride		
		NHS	anhydride		
		NHS	Ester		
		NHS	NHS		
		NHS	Halogen		
		NHS	sulfonate ester		
		NHS	maleimido		
		NHS	haloacetyl		
		NHS	isocyanate		
		halogen	carboxyl		
		halogen	acyl chloride		
		halogen	anhydride		
		halogen	Ester		
		halogen	NHS		
		halogen	Halogen		
		halogen	sulfonate ester		
		halogen	maleimido		
		halogen	haloacetyl		
		halogen	isocyanate		
		sulfonate ester	carboxyl		
		sulfonate ester	acyl chloride		
		sulfonate ester	anhydride		
		sulfonate ester	Ester		
		sulfonate ester	NHS		
		sulfonate ester	Halogen		
		sulfonate ester	sulfonate ester		
		sulfonate ester	maleimido		
		sulfonate ester	haloacetyl		
		sulfonate ester	isocyanate		
		maleimido	carboxyl		
		maleimido	acyl chloride		
		maleimido	anhydride		
		maleimido	Ester		
		maleimido	NHS		
		maleimido	Halogen		
		maleimido	sulfonate ester		
		maleimido	maleimido		
		maleimido	haloacetyl		
		maleimido	isocyanate		
		haloacetyl	carboxyl		

FIGURE 73 Continued…

Both Nucleophilic		Both Electrophilic		Nucleophilic/Electrophilic	
Reactive Group 1	**Reactive Group 2**	**Reactive Group 1**	**Reactive Group 2**	**Reactive Group 1**	**Reactive Group 2**
		haloacetyl	acyl chloride		
		haloacetyl	anhydride		
		haloacetyl	Ester		
		haloacetyl	NHS		
		haloacetyl	Halogen		
		haloacetyl	sulfonate ester		
		haloacetyl	maleimido		
		haloacetyl	haloacetyl		
		haloacetyl	isocyanate		
		isocyanate	carboxyl		
		isocyanate	acyl chloride		
		isocyanate	anhydride		
		isocyanate	Ester		
		isocyanate	NHS		
		isocyanate	Halogen		
		isocyanate	sulfonate ester		
		isocyanate	maleimido		
		isocyanate	haloacetyl		
		isocyanate	isocyanate		

FIGURE 74



HUMAN (SEQ ID NO: 3146)
MACACA (SEQ ID NO: 3147)
RABBIT (SEQ ID NO: 3148)
MOUSE (SEQ ID NO: 3149)
RAT (SEQ ID NO: 3150)

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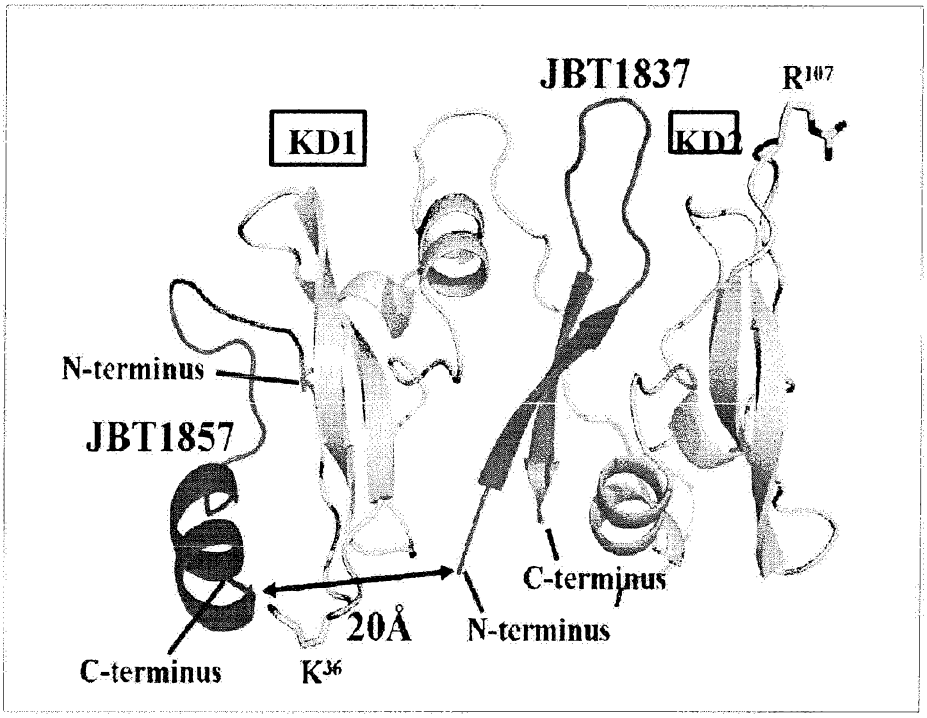
JBT1837
R107
KD1
KD1
N-terminus
JBT1857
C-terminus
20Å
N-terminus
C-terminus
K36

FIGURE 75

FIGURE 76A

Compound ID	Name	TRACER_CONC: 0.5 IC50_NUMERIC Arithmetic Mean	TRACER_CONC: 0.5 IC50_NUMERIC SD	TRACER_CONC: 0.5 IC50_NUMERIC Values	SEQ ID NO:
JBT00000415	Ac-SRYKWF[CGMRDMKGTMSC]VWVKF-NH_2	278.44	37.77	247.36 251.81 285.62 328.97	1003
JBT00001735	Ac-SMYKWH[CGMRDMKGTYSC]VWVKF-NH_2	2.06	0.30	1.85 2.27	1026
JBT00001809	Ac-SHFKWH[CAMRDMKGTMSC]VWVKF-NH_2	2.04	0.74	1.52 2.56	1297
JBT00001810	Ac-SYYKWH[CGMRDMKGTMSC]VWVKF-NH_2	0.92	0.15	0.82 1.03	1298
JBT00001816	Ac-SYYKWH[CGMRDMKGTMSC]VWVKS-NH_2	1.63	0.49	1.28 1.98	1033
JBT00001818	Ac-SYYKWH[CGMRDMKGTYSC]VWVKF-NH_2	0.87	0.08	0.81 0.92	1300
JBT00001819	Ac-SHYKWH[CAMRDMKGTMSC]VWVKF-NH_2	1.34	0.21	1.19 1.48	1301
JBT00001820	Ac-SHYKWH[CAMRDMKGTMSC]VWVKS-NH_2	2.20	0.72	1.70 2.71	1302
JBT00001822	Ac-GHYKWH[CGMRDMKGTFSC]VWVF-NH_2	110.47	44.03	79.33 141.61	1036
JBT00001833	Ac-SYYKWH[CGMRDMKGTMSC]AWVKF-NH_2	1.32	0.25	1.14 1.49	1306

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FIGURE 76B

Compound ID	Name	TRACER_CONC: 0.5 IC50_NUMERIC Arithmetic Mean	TRACER_CONC: 0.5 IC50_NUMERIC SD	TRACER_CONC: 0.5 IC50_NUMERIC Values	SEQ ID NO:
JBT00001837	Ac-SYYKWH[CAMRDMKGTMTC]VWVKF-NH_2	0.52	0.09	0.34 0.40 0.41 0.45 0.50 0.52 0.52 0.56 0.57 0.57 0.58 0.59 0.62 0.63	1044
JBT00001838	Ac-HYYKWH[CGMRDMKGTYSC]VWVKF-NH_2	1.04	0.24	0.87 1.20	1310
JBT00001839	Ac-SYYKWH[CGMRDMKGIYSC]VWVKF-NH_2	0.71	0.15	0.60 0.81	1311
JBT00001841	Ac-SYYKWH[CAMRDMKGTMSC]VWVKF-NH_2	0.78	0.01	0.78 0.79	1312
JBT00002339	Ac-C(NEM)-YYKWH[CAMRDMKGTMTC]VWVKF-NH_2	1.06		1.06	1314
JBT00002340	Ac-C(NEM)-YKWH[CAMRDMKGTMTC]VWVKF-NH_2	1.75		1.75	1315
JBT00002343	Pyn-SYYKWH[CAMRDMKGTMTC]VWVKF-NH_2	0.61	0.06	0.56 0.65	1316
JBT00002344	Pyn-YYKWH[CAMRDMKGTMTC]VWVKF-NH_2	0.72	0.05	0.69 0.76	1317
JBT00002345	Pyn-YKWH[CAMRDMKGTMTC]VWVKF-NH_2	7.75	2.61	5.90 9.59	1318

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FIGURE 76C

Compound ID	Name	TRACER_CONC: 0.5 IC50_NUMERIC Arithmetic Mean	TRACER_CONC: 0.5 IC50_NUMERIC SD	TRACER_CONC: 0.5 IC50_NUMERIC Values	SEQ ID NO:
JBT00002347	Ac-SYYKWH[CAMRDMKGTMTC]VWVKF-Eag-NH_2	0.94		0.937	1351
JBT00002348	Ac-SYYKWH[CAMRDMKGTMTC]VWVK-Eag-NH_2	0.88		0.878	1352
JBT00002349	H-YYKWH[CAMRDMKGTMTC]VWVKF-NH_2	1.22		1.22	1319
JBT-00002350	H-SYYKWH[CAMRDMKGTMTC]VWVKF-NH_2	0.61	0.01	0.60 0.62	1320
JBT00002351	Ac-SYYKWH[CA-Nle-RDMKGTMTC]VWVKF-NH_2	0.65	0.02	0.63 0.67	1321
JBT-00002352	Ac-SYYKWH[CAMRD-Nle-KGTMTC]VWVKF-NH_2	0.57	0.07	0.52 0.62	1322
JBT00002353	Ac-SYYKWH[CAMRDMKGT-Nle-TC]VWVKF-NH_2	0.56	0.06	0.53 0.60	1323
JBT00002354	Ac-SYYKWH[CA-Nle-RD-Nle-KGT-Nle-TC]VWVKF-NH_2	0.96	0.03	0.94 0.98	1324
JBT00002355	Ac-SYYKWH[CA-Moo-RDMKGTMTC]VWVKF-NH_2	0.58	0.06	0.54 0.63	1325
JBT00002356	Ac-SYYKWH[CAMRD-Moo-KGTMTC]VWVKF-NH_2	0.66	0.10	0.59 0.73	1326
JBT00002357	Ac-SYYKWH[CAMRDMKGT-Moo-TC]VWVKF-NH_2	0.60	0.12	0.52 0.69	1327
JBT00002358	Ac-SYYKWH[CA-Moo-RD-Moo-KGT-Moo-TC]VWVKF-NH_2	1.86	0.12	1.77 1.94	1328
JBT00002359	Ac-SYYKWH[CAMRDLKGTMTC]VWVKF-NH_2	0.57	0.03	0.55 0.59	1329
JBT00002360	Ac-SYYKWH[CAMRDMKGTFTC]VWVKF-NH_2	0.64	0.04	0.61 0.66	1330
JBT00002376	Ac-SYYKWH[CAMRDLKGTFTC]VWVKF-NH_2	0.74		0.74	1331
JBT00002377	Ac-SYYKWH[CA-Nle-RDLKGTFTC]VWVKF-NH_2	1.01		1.01	1332

FIGURE 76D

Compound ID	Name	TRACER_CONC: 0.5 IC50_NUMERIC Arithmetic Mean	TRACER_CONC: 0.5 IC50_NUMERIC SD	TRACER_CONC: 0.5 IC50_NUMERIC Values	SEQ ID NO:
JBT000002378	Ac-SYYKWH[CA-Nle-RD-Nle-KGTFTC]VWVKF-NH_2	1.14		1.14	1333
JBT000002379	Ac-SYYKWH[CA-Moo-RDLKGTFTC]VWVKF-NH_2	0.66		0.66	1334
JBT000002380	Ac-SYYKWH[CA-Moo-RDLKGT-Nle-TC]VWVKF-NH_2	0.72		0.72	1335
JBT000002417	PEG2000-CH2-YYKWH[CAMRDMKGTMTC]VWVKF-NH_2	3.68	0.11	3.60 3.76	1336

FIGURE 77

Compound ID	Name	SEQ ID NO:
JBT00000124	Biotinyl-Ttds-SGASRYKWF[CGMRDMKGTMSC]VWVKFRYDTS-NH_2	1290
JBT00000496	Ac-SRYKWF[CG-Sem-RD-Sem-KGT-Sem-SC]VWVKF-NH2	1337
JBT00001165	AOA-SRYKWF[CGMRDMKGTMSC]VWVKF-NH2	1338
JBT00002330	Ac-SYYKWH[CAMRDMKGTMTC]VWVKF-Con(Meox)-NH_2	1339
JBT00002331	Ac-SYYKWH[CAMRDMKGTMTC]VWVK-Con(Meox)-NH_2	1340
JBT00002332	Ac-SYYKWH[CAMRDM-Con(Meox)-GTMTC]VWVKF-NH_2	1341
JBT00002333	Oxme-Aoa-SYYKWH[CAMRDMKGTMTC]VWVKF-NH_2	1342
JBT00002334	Oxme-Aoa-YYKWH[CAMRDMKGTMTC]VWVKF-NH_2	1343
JBT00002335	Oxme-Aoa-YKWH[CAMRDMKGTMTC]VWVKF-NH_2	1344
JBT00002336	Meox-Lev-SYYKWH[CAMRDMKGTMTC]VWVKF-NH_2	1345
JBT00002337	Meox-Lev-YYKWH[CAMRDMKGTMTC]VWVKF-NH_2	1346
JBT00002338	Meox-Lev-YKWH[CAMRDMKGTMTC]VWVKF-NH_2	1347
JBT00002346	Ac-SYYKWH[CAMRDM-Eag-GTMTC]VWVKF-NH_2	1350
JBT00002347	Ac-SYYKWH[CAMRDMKGTMTC]VWVKF-Eag-NH_2	1351
JBT00002348	Ac-SYYKWH[CAMRDMKGTMTC]VWVK-Eag-NH_2	1352
JBT00002456	FAM-Ttds-SYYKWH[CAMRDMKGTMTC]VWVKF-NH_2	1353
JBT00002457	FAM-Ttds-ASMTKF[CKWVMKTYRMYC]HGVDW-NH_2	1354
JBT00002538	Ac-SYYKWH[CAMRD-SeMet-KGT-SeMet-TC]VWVKF-NH_2	1355

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FIGURE 78A

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00000293	Ac-FQSKKNVFVFGYFERLRAKL-NH2	18468.82	>1000.00 3396.67 >5000.00 8513.19 9961.37 12061.10 13202.76 16369.51 16609.04 38403.84 47701.88						
JBT00000303	Ac-FQSKKNVFVDGYFERLRAKL-NH2	439.39	314.91 335.86 432.05 674.75						
JBT00000375	Ac-FQSKGNVFVFGYFERLRAKL-NH2	440.11	326.63 331.22 384.44 718.16						
JBT00000477	Ac-FQSKGNVFVDGYFERLRAKL-NH2	86.96	40.42 74.01 79.67 112.19 128.50						
JBT00000477	Ac-FQSKGNVFVDGYFERLRAKL-NH2	77.64	77.64						
JBT00000477	Ac-FQSKGNVFVDGYFERLRAKL-NH2	146.70	72.95 220.46						

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FIGURE 78B

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00000500	Ac-FQSK-Nmg-NVFVFGYFERLRAKL-NH2	93.36	85.86 92.90 93.89 100.79						
JBT00000740	Ac-FQSKGNVFVDGYFERL-Aib-AKL-NH2	34.51	21.88 25.82 27.37 29.95 46.60 55.46						
JBT00000758	Ac-FQSkKNVFVFGYFERLRAKL-NH2	2286.18	>500.00 2194.24 2231.70 2432.61						
JBT00000759	Ac-FQSKkNVFVFGYFERLRAKL-NH2	632.32	>500.00 552.42 661.62 682.92						
JBT00001156	Ac-FQSKaNVFVDGYFARL-Nle-AKL-NH2	416.11	294.77 537.44						
JBT00001319	Ac-YQSKNNVFVDGYFERLCAKL-NH2	137.20	137.20						
JBT00001320	Ac-FQSKERVFVDGYFERLCAKL-NH2	364.74	364.74						
JBT00001584	Ac-FQSK-Nmg-NVFVDGYFERL-Aib-AKL-NH2	5.30	3.04 3.59 7.08 7.48						

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FIGURE 78C

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00001585	Ac-FQSK-Nmg-NVFVTGYFERL-Aib-AKL-NH2	15.14	8.67 9.82 20.55 21.54						
JBT00001855	Ac-FQSKGNVFVDGYFERL-Aib-AKLC(PEG)-NH2	121.04	60.78 66.26 78.60 121.81 162.52 236.24						
JBT00001856	Ac-FQSKpNVFVDGYFERL-Aib-AKL-NH2	4.57	3.98 4.23 4.24 5.85						

FIGURE 78D

		IC50 ELISA		24 h Koff ELISA	24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00001857	Ac-FQSKpNVHVDGYFERL-Aib-AKL-NH2	3.08	1.48	11.14	27.30	26.30	1.40	1.40
			2.07	4.30		28.30		
			2.10	4.60				
			2.16	5.90				
			2.20	6.90				
			2.53	7.90				
			2.69	11.10				
			2.99	22.10				
			3.03	26.30				
			3.11					
			3.13					
			3.21					
			3.36					
			3.51					
			3.52					
			3.69					
			3.70					
			4.23					
			5.90					

FIGURE 78E

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00001857	Ac-FQSKpNVHVDGYFERL-Aib-AKL-NH2	3.17	1.69 2.32 2.41 2.65 2.83 2.89 2.97 3.03 3.11 3.15 3.26 3.39 4.09 4.13 4.20 4.55	12.58	4.70 7.30 9.20 11.20 12.30 13.20 14.20 14.20 14.30 16.10 16.40 17.80				
JBT00002258	Ac-FQSKKNVFV-NH2		>5000.00						
JBT00002262	FAM-Ttds-FQSK-Nmg-NVFVDGYFERL-Aib-AKL-NH2	13.40	7.45 10.09 17.56 18.50						
JBT00002270	Ac-PEFQSK-Nmg-NVFVDGYFERL-Aib-AKL-NH2	1.94	1.72 2.15	9.80	9.80				
JBT00002274	Ac-FQSK-NpropylG-NVHVDGYFERL-Aib-AKL-NH2	4.34	3.72 4.96	2.30	2.30				

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FIGURE 78F

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002275	Ac-FQSK-aze-NVHVDGYFERL-Aib-AKL-NH2	2.74	1.25 2.07 2.69 3.69 4.02	8.60	8.60				
JBT00002276	Ac-FQSK-pip-NVHVDGYFERL-Aib-AKL-NH2	3.27	1.49 2.06 2.95 3.43 3.59 6.08	4.70	4.70				
JBT00002277	Ac-FQSK-tic-NVHVDGYFERL-Aib-AKL-NH2	3.86	2.23 3.06 4.49 4.74 4.78	0.30	0.30				
JBT00002278	Ac-FQSK-oic-NVHVDGYFERL-Aib-AKL-NH2	2.75	1.82 3.69	25.00	15.10 34.90				
JBT00002279	Ac-FQSK-hyp-NVHVDGYFERL-Aib-AKL-NH2	2.72	1.54 2.19 3.03 3.16 3.68	9.00	9.00				
JBT00002280	Ac-FQSK-nma-NVHVDGYFERL-Aib-AKL-NH2	4.60	4.56 4.65	3.40	3.40				
JBT00002281	Ac-FQSK-Ncg-NVHVDGYFERL-Aib-AKL-NH2	6.08	5.77 6.38	1.80	1.80	17.40	17.40		
JBT00002282	Ac-FQSK-Abg-NVHVDGYFERL-Aib-AKL-NH2	9.98	9.95	5.90	5.90				

FIGURE 78G

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
			10.01						
JBT00002283	Ac-FQSK-Apg-NVHVDGYFERL-Aib-AKL-NH2	20.12	16.62 23.63	6.50	6.50	46.70	46.70		
JBT00002284	Ac-FQSK-thz-NVHVDGYFERL-Aib-AKL-NH2	671.59	671.59	1.00	1.00				
JBT00002285	Ac-FQSK-dtc-NVHVDGYFERL-Aib-AKL-NH2	3.44	2.80 4.07	19.40	19.40				
JBT00002287	Ac-FQSKp-Nmn-VHVDGYFERL-Aib-AKL-NH2		>500.00						
JBT00002288	Ac-FQSKp-Nma-VHVDGYFERL-Aib-AKL-NH2		>5000.00						
JBT00002289	Ac-FQSKp-Nmk-VHVDGYFERL-Aib-AKL-NH2		>5000.00						
JBT00002290	Ac-FQSKp-Nmr-VHVDGYFERL-Aib-AKL-NH1		>5000.00						
JBT00002291	Ac-FQSKp-Nms-VHVDGYFERL-Aib-AKL-NH2	644.27	515.17 773.37						
JBT00002292	Ac-FQSKpnVHVDGYFERL-Aib-AKL-NH2	1124.91	>1000.00 1124.91						
JBT00002293	Ac-FQSKpN-Chg-HVDGYFERL-Aib-AKL-NH2	45.13	43.57 46.69	4.60	4.60			4.10	4.10
JBT00002294	Ac-FQSKpNVH-Chg-DGYFERL-Aib-AKL-NH2	332.48	191.09 335.09 471.25			27.60	27.60	5.40	5.40
JBT00002295	Ac-FQSKpN-Chg-H-Chg-DGYFERL-Aib-AKL-NH2		>5000.00						
JBT00002296	Ac-FQSKpN-Phg-HVDGYFERL-Aib-AKL-NH2	639.17	515.14 763.19						
JBT00002297	Ac-FQSKpNVH-Phg-DGYFERL-Aib-AKL-NH2	66.94	41.08 74.17 85.57			34.70	34.70	4.80	4.80
JBT00002298	Ac-FQSKpN-Phg-H-Phg-DGYFERL-Aib-AKL-NH2		>5000.00						
JBT00002299	Ac-FQSKpNVHVDGYFERL-Deg-AKL-NH2	3.01	2.25 3.76	16.90	16.90				

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FIGURE 78H

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002300	Ac-FQSKpNVHVDGYFERL-Ebc-AKL-NH2	8.39	5.58 6.03 8.20 10.39 11.75	-1.60	-1.60				
JBT00002301	Ac-FQSKpNVHVDGYFERL-Eca-AKL-NH2	3.36	2.57 2.60 4.92	12.10	12.10				
JBT00002302	Ac-FQSKpNVHVDGYFERL-Egz-AKL-NH2	2.70	2.56 2.84	10.90	10.90				
JBT00002303	Ac-FQSKpNVHVDGYFERL-Aic-AKL-NH2	3.04	2.64 3.43	13.20	13.20				
JBT00002304	Ac-FQSKpNVHVDGYFERL-Apc-AKL-NH2	2.22	1.90 2.54	14.20	14.20				
JBT00002305	Ac-FQSKpNVHVDGYFERL-Egt-AKL-NH2	4.48	3.74 5.22	8.30	8.30				
JBT00002306	Ac-1Ni-QSKpNVHVDGYFERL-Aib-AKL-NH2	3.13	2.58 3.68	19.50	19.50	19.30	19.30	0.00	0.00
JBT00002307	Ac-Thi-QSKpNVHVDGYFERL-Aib-AKL-NH2	3.41	3.25 3.58	12.60	12.60	19.80	19.80	1.20	1.20
JBT00002308	Ac-Bta-QSKpNVHVDGYFERL-Aib-AKL-NH2	3.04	2.22 3.87	18.10	18.10	18.65	15.70 21.60	0.00	0.00
JBT00002309	Ac-FQSKpNVHVDGY-1Ni-ERL-Aib-AKL-NH2	1.97	1.39 1.44 1.77 1.91 3.36	21.70	21.70			6.30	6.30

FIGURE 78I

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002310	Ac-FQSKpNVHVDGY-Thi-ERL-Aib-AKL-NH2	3.81	1.82 3.80 4.27 4.57 4.60	10.00	10.00				
JBT00002311	Ac-FQSKpNVHVDGY-Bta-ERL-Aib-AKL-NH2	2.30	1.65 1.71 2.44 3.38	28.30	28.30	46.64	35.90 38.70 41.80 43.60 73.20	6.10	6.10
JBT00002312	Ac-FQSKpNVHVDG-Tym-FERL-Aib-AKL-NH2	194.26	156.94 231.59						
JBT00002313	Ac-FQSKpNVHVDG-Pty-FERL-Aib-AKL-NH2	257.97	230.29 285.65						
JBT00002314	Ac-FQSKpNVHVDG-Pmy-FERL-Aib-AKL-NH2	5.14	4.21 6.07	7.40	7.40			7.30	7.30
JBT00002315	Ac-FQSKpNVHVDGYFERL-Aib-AKLC-NH2	6.12	2.49 5.36 6.90 7.04 8.80						

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FIGURE 78J

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002317	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(NEM)-NH2	3.53	2.26 2.37 2.54 3.08 3.32 3.53 3.58 3.72 3.97 4.31 4.42 5.26	12.73	9.60 10.70 17.90	23.77	18.20 21.00 32.10	15.95	3.40 28.50
JBT00002319	Ac-VF-Aib-EVLKFGHALKpNSQDRY-NH2		>50000.00 >50000.00 >50000.00 >50000.00						
JBT00002320	Ac-FpDYVER-Aib-KGFLVHQKLASN-NH2		>50000.00 >50000.00 >50000.00 >50000.00						
JBT00002325	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(ME-50MA)-NH2	6.60	5.96 6.62 6.64 7.16	3.70	3.70	67.60	67.60	44.70	44.70
JBT00002326	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(ME-120MA)-NH2	9.30	7.91 9.57 10.43	4.30	4.30	58.70	58.70	57.20	57.20

FIGURE 78K

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002327	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(ME-200MA0B)-NH2	10.92	8.98 11.80 11.99	5.00	5.00	76.50	76.50	65.50	65.50
JBT00002328	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(ME-300MA)-NH2	12.29	8.99 13.92 13.97	2.70	2.70	68.40	68.40	51.00	51.00
JBT00002329	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(ME-400MA)-NH2	16.43	8.41 10.74 11.39 11.93 12.24 13.46 13.88 15.67 16.98 18.48 18.97 19.77 21.20 36.94	14.27	11.60 15.10 16.10	69.50	64.10 74.90	69.50	69.50

FIGURE 78L

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002329	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(ME-400MA)-NH2	14.77	7.24 7.29 7.43 7.43 7.49 12.33 16.91 17.73 18.80 20.15 20.52 33.87			62.90	62.90	81.80	81.80
JBT00002329	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(ME-400MA)-NH2	19.03	13.25 16.78 27.06						
JBT00002329	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(ME-400MA)-NH2	15.57	10.82 11.20 11.91 12.87 13.35 13.74 16.31 21.18 28.75						
JBT00002329	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(ME-400MA)-NH2	18.54	14.70 17.79 23.13						

FIGURE 78M

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002329	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(ME-400MA)-NH2	20.76	20.57 20.96			59.30	59.30	56.70	56.70
JBT00002366	Ac-Bta-QSKpNVHVDGY-1Ni-ERL-Aib-AKL-NH2	1.99	1.85 2.13	8.30	8.30				
JBT00002367	Ac-Bta-QSKpNVHVDGY-Bta-ERL-Aib-AKL-NH2	2.05	1.83 2.28	12.80	12.80				
JBT00002368	Ac-1Ni-QSKpNVHVDGY-1Ni-ERL-Aib-AKL-NH2	2.32	2.10 2.55	11.10	11.10				
JBT00002369	Ac-1Ni-QSKpNVHVDGY-Bta-ERL-Aib-AKL-NH2	2.03	1.90 2.16	12.00	12.00				
JBT00002370	Ac-FQSKpCVHVDGYFERL-Aib-AKL-NH2	6.33	4.04 8.62						
JBT00002371	Ac-FQSKpNVHVCGYFERL-Aib-AKL-NH2	10.67	5.48 15.86			31.20	31.20		
JBT00002372	Ac-FQSKpNVHVDGYCERL-Aib-AKL-NH2	11.68	8.14 15.23						
JBT00002373	Ac-FQSKpNVHVDGYFCRL-Aib-AKL-NH2	4.07	3.12 5.02						
JBT00002381	Ac-FQSKpNVH-Abu-DGYFERL-Aib-AKLC(NEM)-NH2	3.45	2.34 4.56	1.90	1.90	17.30	17.30		
JBT00002382	Ac-FQSKpNVH-(L-2-Amino-4,4,4-trifluorobutyric acid)-DGYFERL-Aib-AKLC(NEM)-NH2	62.74	43.29 82.19	-2.20	-2.20	20.50	20.50		
JBT00002383	Ac-FQSKpNVH-Cpg-DGYFERL-Aib-AKLC(NEM)-NH2	52.18	40.44 63.91	-4.20	-4.20	31.00	31.00		
JBT00002384	Ac-FQSKpNVHvDGYFERL-Aib-AKLC(NEM)-NH2		>500.00						
JBT00002385	Ac-FQSKpNVH-Aib-DGYFERL-Aib-AKLC(NEM)-NH2	24.23	18.11 30.36	-0.80	-0.80	51.50	51.50		

FIGURE 78N

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002386	Ac-FQSKpNVH-Tle-DGYFERL-Aib-AKLC(NEM)-NH2	4.92	1.97 3.64 3.66 3.68 4.03 4.22 6.17 6.49 10.46	11.93	6.50 9.30 11.50 12.10 15.70 16.50	52.00	52.00		
JBT00002387	Ac-FQSKpNVH-Nmv-DGYFERL-Aib-AKLC(NEM)-NH2		>500.00						
JBT00002388	Ac-FQSKpNVHV-Nmd-GYFERL-Aib-AKLC(NEM)-NH2	9.62	7.02 12.22	1.60	1.60	46.50	46.50		
JBT00002389	Ac-FQSKpNVHVHGYFERL-Aib-AKLC(NEM)-NH2	4.40	2.97 5.83	16.50	16.50	18.60	18.60		
JBT00002390	Ac-FQSKpNVHVSGYFERL-Aib-AKLC(NEM)-NH2	2.97	2.26 3.68	10.00	10.00	33.40	33.40		
JBT00002391	Ac-FQSKpNVHVTGYFERL-Aib-AKLC(NEM)-NH2	3.05	2.90 3.19	11.20	11.20	41.40	41.40		
JBT00002392	Ac-FQSKpNVHVPGYFERL-Aib-AKLC(NEM)-NH2	2.62	2.05 3.20	12.80	12.80	58.00	58.00		
JBT00002393	Ac-FQSKpNVHVdGYFERL-Aib-AKLC(NEM)-NH2	32.75	24.71 40.80	-0.50	-0.50	70.90	70.90		
JBT00002394	Ac-FQSKpNVH-Tle-PGYFERL-Aib-AKLC(NEM)-NH2	5.53	4.04 7.02	5.80	5.80	55.90	55.90		
JBT00002395	Ac-FQSKpNVH-Tle-TGYFERL-Aib-AKLC(NEM)-NH2	4.37	3.32 5.43	9.70	9.70	52.60	52.60		
JBT00002396	Ac-FQSKpNVHVDAYFERL-Aib-AKLC(NEM)-NH2	31.54	23.49 39.59	-0.50	-0.50	17.90	17.90		

FIGURE 78O

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002397	Ac-FQSKp-C(NEM)-VHVDGYFERL-Aib-AKL-NH2	3.30	1.93 3.52 4.44	39.73	27.60 40.30 43.30 47.70	10.60	10.60		
JBT00002397	Ac-FQSKp-C(NEM)-VHVDGYFERL-Aib-AKL-NH2	3.45	3.45	43.30	43.30				
JBT00002398	Ac-FQSKpNVHV-C(NEM)-GYFERL-Aib-AKL-NH2	4.29	3.26 5.33	4.10	4.10	39.05	33.90 44.20	21.60	21.60
JBT00002399	Ac-FQSKpNVHVDGYF-C(NEM)-RL-Aib-AKL-NH2	4.23	3.35 5.12	11.20	11.20	10.70	10.70		
JBT00002400	Ac-FQSKpNVHVDGYFERL-Aib-AKLK(Ttds-Maleimidopropionyl(EtSH))-NH2	4.02	2.17 3.13 3.57 4.63 6.59	23.40	23.10 23.70	60.00	60.00		
JBT00002401	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(Y-MAL-40K)-NH2	31.42	14.47 18.58 44.37 48.26	21.40	21.40	83.70	77.40 90.00	112.10	112.10
JBT00002401	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(Y-MAL-40K)-NH2	35.52	35.52			87.90	87.90		
JBT00002402	Ac-FQSKp-C(ME-400MA)-VHVDGYFERL-Aib-AKL-NH2	20.46	12.29 12.91 36.18	28.90	28.90	75.30	53.30 73.70 98.90	99.70	99.70
JBT00002403	Ac-FQSKpNVHV-C(ME-400MA)-GYFERL-Aib-AKL-NH2	22.89	15.60 17.55 35.52	6.30	6.30	96.70	74.30 102.90 112.90	92.50	92.50

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FIGURE 78P

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002404	Ac-FQSKpNVHVDGYF-C(ME-400MA)-RL-Aib-AKL-NH2	24.31	13.60 16.36 33.41 33.87	14.90	14.90	96.60	70.70 122.50	93.20	93.20
JBT00002404	Ac-FQSKpNVHVDGYF-C(ME-400MA)-RL-Aib-AKL-NH2	31.65	31.65			96.90	96.90		
JBT00002405	[FQSKpNVHVDGYFERL-Aib-AKL-FA19205]	38.76	38.76			55.90	55.90		
JBT00002406	[FQSKpNVHVDGYFERL-Aib-AKL-FA19204]	75.35	67.04 83.66			62.80	62.80	38.10	38.10
JBT00002407	[FQSKpNVHVDGYFERL-Aib-AKL-FA19203]	28.48	20.94 36.01	24.00	24.00	45.20	45.20		
JBT00002408	[FQSKpNVHVDGYFERL-Aib-AKL-FA03202]	65.52	56.17 74.88			61.60	61.60	39.40	39.40
JBT00002410	Ac-FQSKpNVHVDGYFERL-Aib-AKLK(Ttds-Maleimid-HSA)-NH2	6.90	4.72 6.03 9.95						
JBT00002410	Ac-FQSKpNVHVDGYFERL-Aib-AKLK(Ttds-Maleimid-HSA)-NH2	6.21	4.33 4.83 6.07 9.62	31.80	31.80	65.70	25.90 76.80 94.40	79.15	55.40 102.90
JBT00002410	Ac-FQSKpNVHVDGYFERL-Aib-AKLK(Ttds-Maleimid-HSA)-NH2	4.59	4.59						
JBT00002411	Ac-FQSKpKVH-Tle-DGYFERL-Aib-AKL-NH2	4.61	3.05 5.36 5.41	16.80	14.10 19.50				
JBT00002412	Ac-FQSKp-Cit-VH-Tle-DGYFERL-Aib-AKL-NH2	6.09	4.43 7.75	4.40	4.40				
JBT00002413	Ac-FQSKpVVH-Tle-DGYFERL-Aib-AKL-NH2	8.60	5.98 11.22	-2.80	-2.80				

FIGURE 78Q

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002414	Ac-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	6.22	4.03 4.92 5.15 5.32 6.31 6.78 7.52 7.89 8.06	3.17	-3.60 1.50 11.60	100.50	93.10 107.90	47.25	46.80 47.70
JBT00002415	Ac-FQSKp-C(Acm)-VH-Tle-DGYFERL-Aib-AKL-NH2	4.45	3.35 3.41 3.66 4.58 5.04 6.64	11.40	4.80 6.20 9.30 25.30	81.20	81.20	36.10	36.10
JBT00002416	Ac-FQSKp-Nle-VH-Tle-DGYFERL-Aib-AKL-NH2	7.72	5.89 9.54	5.40	5.40				
JBT00002418	Ac-FQSKpRVH-Tle-DGYFERL-Aib-AKL-NH2	3.12	3.12	20.60	20.60	66.10	66.10		
JBT00002419	Ac-FQSKpSVH-Tle-DGYFERL-Aib-AKL-NH2	3.23	3.23	13.00	13.00				
JBT00002420	Ac-FQSKpIVH-Tle-DGYFERL-Aib-AKL-NH2	7.33	7.33	0.90	0.90				
JBT00002421	Ac-FQSKpNVH-Tle-D-Sar-YFERL-Aib-AKL-NH2	19.51	19.51	1.60	1.60				
JBT00002422	Ac-FQSKpNVH-Tle-DpYFERL-Aib-AKL-NH2	28.02	28.02	1.10	1.10				
JBT00002423	Ac-FQSKpNVH-Tle-DGYFERL-Aib-LKL-NH2	3.56	3.56	8.90	8.90	78.70	78.70	13.70	13.70
JBT00002424	Ac-FQSKpNVH-Tle-DGYFERL-Aib--Nle-KL-NH2	4.37	4.37	8.20	8.20	90.10	90.10	17.00	17.00
JBT00002425	Ac-FQSKpNVH-Tle-DGYFERL-Aib-A-Nmk-L-NH2		>500.00						
JBT00002426	Ac-FQDKpNVH-Tle-DGYFERL-Aib-AKL-NH2	10.46	10.46	0.10	0.10				
JBT00002427	Ac-FQEKpNVH-Tle-DGYFERL-Aib-AKL-NH2	5.24	5.24	14.10	14.10				
JBT00002428	Ac-FQVH-Tle-DGYFERL-Aib-AKL-NH2		>500.00						

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FIGURE 78R

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002429	Ac-FqVH-Tle-DGYFERL-Aib-AKL-NH2		>500.00						
JBT00002430	Ac-FQSKpNVHVDGYFERL-Aib-AKLK(AOA-40kDa PSA)-NH2	15.45	15.45						
JBT00002430	Ac-FQSKpNVHVDGYFERL-Aib-AKLK(AOA-40kDa PSA)-NH2			40.95	39.50 42.40	135.60	124.20 147.00	122.60	119.10 126.10
JBT00002431	Ac-FQSKpNVH-Tle-DGYFERL-Aib-AKL-NH2	4.74	4.67 4.80	5.10	5.10	81.90	81.90	42.60	42.60
JBT00002432	Ac-FQSKpNVHVPGYFERL-Aib-AKL-NH2	3.76	3.45 4.06	6.70	6.70	76.30	76.30		
JBT00002433	Ac-FQSKpNVHVPaYFERL-Aib-AKL-NH2	5.71	5.01 6.40	17.80	17.80	99.30	99.30	27.30	27.30
JBT00002434	Ac-FQSKpNVHVPGYF-Aib-RL-Aib-AKL-NH2	5.53	5.22 5.84	13.75	7.70 8.60 9.60 29.10	99.55	77.50 121.60	31.20	31.20
JBT00002435	Ac-FQSKpNVH-Tle-DaYFERL-Aic-AKL-NH2	8.00	7.47 8.54	4.80	4.80	103.95	87.90 120.00	53.00	53.00
JBT00002436	Ac-FQSKpNVH-Tle-DaYFERL-Eca-AKL-NH2	5.69	5.69	3.10	3.10				
JBT00002437	Ac-FQSKpNVH-Tle-DaYFERL-Deg-AKL-NH2	5.69	5.69	15.20	15.20			62.90	62.90
JBT00002438	Ac-FQSKpNVH-Tle-DaYFERL-Aib--Aib-KL-NH2	8.55	8.55	16.80	16.80	99.20	90.10 108.30	85.90	85.90
JBT00002439	Ac-FQSKpNV-2Ni--Tle-DaYFERL-Aib-AKL-NH2	8.01	7.03 9.00	13.00	6.40 19.60	88.80	88.80	51.10	51.10
JBT00002440	Palm-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2		>500.00						
JBT00002441	Ac-FQSKpNVH-Tle-DaYFER[-Hcy--Aib-AK-Hcy-]-NH2	55.25	50.25 60.25	1.15	-8.10 10.40			65.50	65.50
JBT00002442	Ac-FQSKpNV-1Ni--Tle-DaYFERL-Aib-AKL-NH2	15.29	15.29	-6.80	-6.80				

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FIGURE 78S

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002443	Ac-FQ[CKpC]VH-Tle-DaYFERL-Aib-AKL-NH2	237.49	191.11 283.87	2.35	-3.70 8.40				
JBT00002444	Ac-FQSKpNVH-Tle-DaYFER-Hle--Aib-AKL-NH2	12.54	12.54	7.40	7.40				
JBT00002445	Ac-FQSKpN-Tle-H-Tle-DaYFERL-Aib-AKL-NH2	129.64	129.64						
JBT00002446	Ac-FQSKpNVH-Tle-DaYFERL-Aib-A-Har-L-NH2	57.48	55.65 59.31	2.05	-2.30 6.40				
JBT00002447	Ac-FQSKpNVH-Tle-DaYFERL-Aib-A-Opc-L-NH2		>500.00						
JBT00002448	Ac-FQSKpNVH-Tle-DaYFERLA-Aib-KL-NH2	9.36	9.36	7.90	7.90				
JBT00002449	Ac-FQSKpNVH-Tle-DaYFERLa-Aib-KL-NH2	43.23	35.79 50.66	6.80	6.80				
JBT00002450	Ac-FQSKpNVH-Tle-DaYF[-Hcy-RLA-Hcy-]KL-NH2	398.31	190.47 413.89 590.57	10.05	4.80 15.30				
JBT00002451	Ac-FQSKpEVH-Tle-DaYFERL-Aib-AKL-NH2	7.36	7.36	8.50	8.50				
JBT00002452	Ac-FQSKp-Ede(O)-VH-Tle-DaYFERL-Aib-AKL-NH2	5.75	5.75	8.10	8.10				
JBT00002453	Ac-FQ-Ede(O)-KpNVH-Tle-DaYFERL-Aib-AKL-NH2	7.90	7.90	4.40	4.40				
JBT00002458	374-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	6.17	5.36 6.98	9.90	9.90				
JBT00002459	1281-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	8.45	7.20 9.70	6.10	6.10				
JBT00002460	5963-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	10.90	9.14 12.66	12.40	12.40	107.60	107.60		

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FIGURE 78T

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002461	Hex-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	4.63	3.20 4.27 4.28 4.47 5.26 5.30 5.63	10.24	8.10 9.20 11.20 11.30 11.40	97.40	88.00 90.40 113.80	39.85	34.30 45.40
JBT00002462	4635-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	6.30	5.15 7.45	4.00	4.00				
JBT00002463	3067-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	4.15	3.30 5.00	9.20	9.20				
JBT00002464	972-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	7.72	6.64 8.79	10.20	10.20	104.70	104.70		
JBT00002465	973-FQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	6.72	5.51 7.30 7.34	15.40	15.40	107.10	107.10	52.00	52.00
JBT00002466	Ac-FQSKpNVH-Tle-DaYFERL-Aib-[CKL-Cea-]	8.27	6.39 8.65 9.78	33.00	27.00 28.70 43.30	16.00	16.00		
JBT00002467	Ac-FQSKpNVH-Tle-DaYFERL-Aib-[cKL-Cea-]	6.65	4.18 5.42 5.82 6.59 7.22 7.63 8.16 8.22	52.90	44.50 48.70 56.30 62.10	12.10	12.10	34.30	34.30
JBT00002468	Ac-FQSKpNVH-Tle-DaYFERL[CAKL-Cea-]	60.46	60.46						

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FIGURE 78U

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002469	Ac-FQSKpNVH-Tle-DaYFERL[cAKL-Cea-]	14.71	14.71	1.00	1.00				
JBT00002470	Ac-FQSKpNVH-Tle-DaYFERL-Aib-AK-Aml--NH2	19.61	19.61	-4.60	-4.60				
JBT00002471	Ac-FQSKpNVH-Tle-DaYFER-Aml-AAKL-NH2	9.34	9.34	-2.90	-2.90				
JBT00002472	Ac-FQSKpNVH-Tle-DaYFERL-Aib-LKL-NH2	7.59	7.59	-3.90	-3.90				
JBT00002473	Ac-FQSKpNVH-Tle-DaYF-Aib-RL-Aib-LKL-NH2	11.92	11.92	-9.50	-9.50				
JBT00002474	Ac-FQSKpNVH-Tle-DaYF-Aib-RL-Aib-AKL-NH2	7.27	7.27	-8.90	-8.90				
JBT00002475	Ac-FQSKpNVHVPaYF-Aib-RL-Aib-AKL-NH2	7.66	7.66	0.80	0.80				
JBT00002476	Ac-FQSKpNVHVPaYFERL-Aib-LKL-NH2	5.17	5.17	-1.00	-1.00				
JBT00002477	Ac-FQSKpNVHVPaYF-Aib-RL-Aib-LKL-NH2	13.85	13.61 14.09	5.40	5.40				
JBT00002478	Ac-FQSKpNVH-Tle-Da-Dopa-FERL-Aib-AKL-NH2	38.36	38.36	-6.10	-6.10				
JBT00002479	Ac-YQSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	4.41	4.23 4.60	0.40	0.40				
JBT00002480	Ac--Dopa-QSKpNVH-Tle-DaYFERL-Aib-AKL-NH2	6.33	6.33	5.50	5.50				
JBT00002485	Ac-FQSKpNV-Pmy--Tle-DaYFERL-Aib-AKL-NH2	5.55	4.85 6.25	-0.40	-0.40				
JBT00002486	Ac-FQSKpNTH-Tle-DaYFERL-Aib-AKL-NH2	8.11	8.11	-2.20	-2.20				
JBT00002487	Ac-FQSKp-Cmc-VH-Tle-DaYFERL-Aib-AKL-NH2	9.10	9.10	-2.90	-2.90				
JBT00002488	Ac-FQ-Cmc-KpNVH-Tle-DaYFERL-Aib-AKL-NH2	9.04	9.04	-1.10	-1.10				
JBT00002489	Ac-FQSKpNVH-Tle-DaYF[-Hcy-RL-Aib--Hcy-]KL-NH2		>500.00						
JBT00002490	Ac-FQSKpNVH-Tle-DaYF[cRLA-Hcy-]KL-NH2	6165.16	>500.00 6165.16						
JBT00002491	Ac-FQSKpNVH-Tle-DaYF[CRLA-Hcy-]KL-NH2		>500.00						
JBT00002492	Ac-FQSKpNVH-Tle-DaYF[cRLC]AKL-NH2		>500.00						
JBT00002493	Ac-FQSKpNVH-Tle-DaYF[CRLC]AKL-NH2		>500.00						
JBT00002494	Ac-FQSKpNVH-Tle-D[cYF-Hcy-]RL-Aib-AKL-NH2	2.84	2.54 2.81	17.60	13.00 22.20	61.50	61.50	11.10	11.10

FIGURE 78V

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
			3.18						
JBT00002495	Ac-FQSKpNVH-Tle-D[cYFC]RL-Aib-AKL-NH2	3.36	3.14 3.37 3.59	12.50	12.50	49.30	49.30		
JBT00002496	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(Ac-FQSKpNVHVDGYFERL-Aib-AKLK(Ttds-Maleiimidopropionyl)-NH2)--NH2	0.85	0.56 0.59 0.62 0.88 1.06 1.09 1.19	55.63	49.30 52.20 56.50 64.50	56.60	56.60	25.80	25.80
JBT00002496	Ac-FQSKpNVHVDGYFERL-Aib-AKLC(Ac-FQSKpNVHVDGYFERL-Aib-AKLK(Ttds-Maleiimidopropionyl)-NH2)--NH2	1.18	0.76 0.93 1.02 1.07 1.13 1.18 1.19 1.22 1.23 1.30 1.36 1.44 1.48	53.62	47.00 50.50 52.80 55.30 62.50			16.80	16.80
JBT00002497	Ac-FQSKpNVH-Tle-D-hcy-YF-Hcy-RL-Aib-AKL-NH2	7.16	7.16	2.50	2.50				
JBT00002498	Ac-FQSKpNVH-Tle-D-hcy-YFCRL-Aib-AKL-NH2	7.59	7.59	6.00	6.00				

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FIGURE 78W

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002499	Ac-FQSKpNVHVPcYF-Hcy-RL-Aib-AKL-NH2	3.89	3.30 4.48	17.20	17.20	40.70	40.70		
JBT00002500	Ac-FQSKpNVH-Tle-DcY-Bta--Hcy-RL-Aib-AKL-NH2	5.26	4.91 5.05 5.82	9.10	9.10				
JBT00002501	Ac-FQSKp-C(Acm)-VH-Tle-D[cY-Bta--Hcy-]RL-Aib-AKL-NH2	6.31	5.92 6.13 6.87	15.10	15.10				
JBT00002502	Ac-FQSKp-C(NEM)-VH-Tle-DaY-Bta-ERL-Aib-AKL-NH2	3.89	3.68 4.10	12.40	12.40				
JBT00002503	Ac-FQSKp-C(NEM)-VHVPaY-Bta-ERL-Aib-AKL-NH2	5.04	5.04	2.80	2.80				
JBT00002504	Ac-FQSKpNVH-Tle-DaYFER-Aml--Aib-AKL-NH2	9.43	9.43	-4.90	-4.90				
JBT00002505	Ac-FQSKp-Ecl-VHVPaYFERL-Aib-AKL-NH2	5.82	5.82	-2.80	-2.80				
JBT00002506	Ac-FQSKp-Eea-VHVPaYFERL-Aib-AKL-NH2	6.01	6.01	-2.60	-2.60				
JBT00002507	Ac-FQSKp-Eec-VHVPaYFERL-Aib-AKL-NH2	9.71	9.71	-3.80	-3.80				
JBT00002508	Ac-FQSKp-Eef-VHVPaYFERL-Aib-AKL-NH2	31.05	31.05	-5.20	-5.20				
JBT00002509	Ac-FQSKp-Nif-VHVPaYFERL-Aib-AKL-NH2	19.69	19.69	-4.10	-4.10				
JBT00002510	Ac-FQSKp-Eew-VHVPaYFERL-Aib-AKL-NH2	8.86	8.86	2.90	2.90				
JBT00002511	Ac-FQSKpNVH-Tle-DaY-Bta-ERL-Aib-[cKL-Cea-]	189.24	183.33 195.16						
JBT00002512	Ac-FQSKp-C(Acm)-VH-Tle-DaY-Bta-ERL-Aib-[cKL-Cea-]	157.63	134.39 180.87						
JBT00002513	Ac-FQSKpNVH-Tle-DaYFERL-Aib-[-hcy-KL-Cea-]	317.99	304.79 331.19						
JBT00002514	Ac-FQSKpNVH-Tle-DaYFERL-Aib-[cKL-Hcy-]-NH2	132.62	131.69 133.55						
JBT00002515	Ac-FQSKpNVH-Tle-DaYFERL-Aib-[cKLC]-NH2	579.90	>500.00 579.90						

FIGURE 78X

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002516	Ac-FQSKpNVH-Tle-DaYFERL-Aib-[cKLc]-NH2	472.87	402.05 543.69						
JBT00002517	Ac-FQSKpNVHVPaY-Bta-ERL-Aib-[cKL-Cea-]	269.09	228.94 309.24						
JBT00002518	Ac-FQSKpNVHVP[cY-Bta--Hcy-]RL-Aib-AKL-NH2	11.56	10.49 11.07 12.02 12.66	37.50	24.30 50.70	77.70	77.70	28.30	28.30
JBT00002519	Ac-FQSKpNVH-Tle-DaYFERL-Aib-AKL-K(Ttds-Maleimidopropionyl)--NH2	5.92	4.79 5.92 7.06						
JBT00002520	Ac-FQSKpNVH-Tle-DaYFERL-Aib-AKL-K(Ttds-Maleimidopropionyl(EtSH))--NH2	6.01	4.60 5.02 6.29 6.34 7.78						
JBT00002521	(Ac-FQSKpNVHVDGYFERL-Aib-AKLC-NH2)2-M2Hex	1.81	1.34 1.53 1.93 2.43	39.00	38.60 39.40				
JBT00002522	(Ac-FQSKpNVHVDGYFERL-Aib-AKLC-NH2)2-M2TriO	0.82	0.53 0.64 0.81 0.87 0.89 0.92 1.11	57.43	54.70 56.00 61.60	46.90	46.90	18.05	17.50 18.60

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FIGURE 78Y

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
JBT00002523	(Ac-FQSKpNVHVDGYFERL-Aib-AKLC-NH2)2-M2PEG2000	1.03	0.92 0.92 1.05 1.09 1.17	61.40	58.00 59.10 60.60 67.90				
JBT00002527	Ac-FQSKp-Cys(Acm)-VH-Tle-DaYFERL-Aib-AKL-NH2	5.43	5.40 5.46	2.90	1.60 3.20 3.90	86.15	75.90 96.40	43.00	38.90 47.10
JBT00002528	Hex-FQSKp-C(Acm)-VH-Tle-DaYFERL-Aib-AKL-NH2	4.78	3.07 3.34 4.00 4.06 4.57 4.73 5.06 5.37 5.48 5.80 5.90 5.99	12.28	8.20 10.40 11.20 13.20 18.40	90.13	83.80 83.90 102.70		
JBT00002531	Hex-FQSKp-Cys(Acm)-VH-Tle-DaYFERL-Aib-AKLK(Ttds-Myr)--NH2	27.24	12.54 13.22 22.62 26.31 26.93 27.39 27.68 42.78	10.25	10.10 10.40	96.50	86.60 106.40	67.50	42.90 92.10

FIGURE 78Z

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
			45.67						
JBT00002531	Hex-FQSKp-Cys(Acm)-VH-Tle-DaYFERL-Aib-AKLK(Ttds-Myr)--NH2	26.97	26.97						
JBT00002532	Hex-FQSKp-Cys(Acm)-VH-Tle-DaYFERL-Aib-AKLK(Ttds-Palm)--NH2	36.68	32.09 35.38 36.51 39.35 40.08	13.30	13.20 13.40	107.70	107.70	67.50	67.50
JBT00002533	(Hex-FQSKp-Cys(Acm)-VH-Tle-DaYFERL-Aib-AKLC-NH2)2-M2TriO	3.10	2.76 2.79 2.97 3.88	77.70	77.30 78.10	185.13	157.50 172.30 225.60	52.65	51.70 53.60
JBT00002534	Hex-FQSKp-Cys(Acm)-VH-Tle-DaYFERL-Aib-AKLC(ME-400MA)-NH2	15.32	13.96 14.46 15.74 17.12	13.35	11.80 14.90	59.83	46.90 59.70 60.90 71.80	69.87	66.50 69.00 74.10
JBT00002537	Hex-FQSKp-Cys(Acm)-VH-Tle-DaYFERL-Aib-AKLC(NEM)-NH2	7.13	4.79 6.41 6.45 7.59 10.41	14.40	9.60 15.70 17.90	106.65	88.70 124.60		
JBT00002539	Hex-FQSKp-Cys(Acm)-VH-Tle-DaYFERL-Aib-AKL-Ttds-K(Ttds-Myr)--NH2	10.07	10.07						
JBT00002540	Hex-FQSKp-Cys(Acm)-VH-Tle-DaYFERL-Aib-AKLK(Ttds-Ac)--NH2	3.99	3.99						
JBT00002541	Hex-FQSKp-Cys(Acm)-VH-Tle-DaYFERL-Aib-AKLK(Ttds-gGlu-Myr)--NH2	24.34	24.34						
JBT00002543	Hex-FQSKp-Cys(Acm)-VH-Tle-DaYFERL-Aib-AK-OH	71.08	68.82						

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FIGURE 78AA

		IC50 ELISA		24 h Koff ELISA		24 h Human Plasma Stability		24 h Mouse Plasma Stability	
Compound ID	Name	Mean IC50 [nM]	IC50 [nM]	INHIBITION [%]	INHIBITION [%]	Mean Remaining [%]	Remaining [%]	Mean Remaining [%]	Remaining [%]
			73.35						
JBT00002544	(Hex-FQSKp-Cys(Acm)-VH-Tle-DaYFERL-Aib-AKLC-NH2)-Mal-mPEG23	3.61	3.61			54.00	54.00	46.20	46.20
JBT00002546	Hex-FQSKpSVH-Tle-DaYFERL-Aib-AKLC(NEM)-NH2	7.08	5.77 8.39	10.35	9.10 11.60	87.66	65.40 71.70 86.40 98.00 116.80	72.98	53.30 65.30 83.10 90.20
JBT00002548	Hex-FQSKpSVH-Tle-DaYFERL-Aib-AKL-NH2	3.62	2.07 3.83 3.96 4.63			98.85	85.00 112.70	50.25	45.00 55.50
JBT00002549	Hex-FQSKpSVH-Tle-DaYFERL-Aib-AKL-K(Myr)--NH2	89.96	89.96						
JBT00002550	Hex-FQSKpSVH-Tle-DaYF-K(Myr)-RL-Aib-AKL-NH2	218.04	218.04						
JBT00002551	Hex-FQSKpSVH-Tle--K(Myr)-aYFERL-Aib-AKL-NH2		>500.00						
JBT00002552	Hex-FQSKp-K(Myr)-VH-Tle-DaYFERL-Aib-AKL-NH2		>500.00						
JBT00002553	Hex-FQS-K(Myr)-pSVH-Tle-DaYFERL-Aib-AKL-NH2		>500.00						
JBT00002554	Hex-FQSKpSVH-Tle-DaYFERL-Aib-AKL-K(AlbuTag)-NH2	19.57	19.57						
JBT00002555	Hex-FQSKpSVH-Tle-DaYFERL-Aib-AKL-K(4PBSA)-NH2	49.77	49.77						