A bone graft substitute in the form of an implantable three-dimensional scaffold that includes calcium phosphate and has pores. The scaffold is impregnated with a calcium and/or phosphate containing substance, and the dissolution rate $DR_s$ of the scaffold is slower than the dissolution rate $DR_d$ of the calcium and/or phosphate containing substance.
FIG. 1

y = 0.8388x
R² = 0.9723
FIG. 2
FIG. 3
Fig. 4
BONE GRAFT SUBSTITUTE

[0001] The invention relates to a bone graft substitute according to the preamble of claim 1 and a method for manufacturing a bone graft substitute according to the preamble of claim 2.

[0002] The following definitions shall be used throughout the description:

[0003] Resorption/degradation—process by which a material is removed from the human body.

[0004] Scaffold—matrix—porous material

[0005] Macropores—here, we define macropores as pores that have a diameter superior to 30-50 microns

[0006] Micropores—pores with a diameter in the range of 0.1 to 20-30 microns

[0007] Nanopores—pores with a diameter smaller than 100 nm

[0008] Tortuous—tortuous pores are pores that do not have a straight shape (e.g. cylindrical, spherical), but a complex shape, such as a helix, with a large aspect ratio (ratio between the longest and the shortest pore dimension).

[0009] Tortuosity—Tortuosity is defined as the ratio between the distance required to join two points in a porous structure through the porous network and the direct distance (with a straight line). Tortuosity values are by definition larger than 1 and often larger than 3.

[0010] Calcium phosphate bone graft substitutes have proved to be very good bone graft substitutes: the materials have an excellent biocompatibility and depending on their exact composition, might also be degraded over time and replaced by new bone. One particularly successful material is β-tricalcium phosphate [β-Ca₃(PO₄)₂] or shortly β-TCP.

[0011] In past years, many studies have shown the importance of calcium and phosphate ions on the cellular response of bone cells, such as osteoblasts (“bone-forming cells”) and osteoclasts (“bone-resorbing cells”). For example, it is known that a small increase of calcium concentration down-regulates osteoclast activity and up-regulates osteoblast activity. Also, it has been shown that increased calcium ion concentrations could trigger osteoblasts to produce bone morphogenetic proteins such as BMP-2 and BMP-4. We have therefore surprisingly found that calcium phosphate bone graft substitutes can be used as drug delivery systems (Ca and phosphate ions being the drugs). The control of calcium and/or phosphate ions release enables also a control of the in vivo properties of calcium phosphate materials.

[0012] Generally, it is desirable to have a cell-mediated degradation (e.g. osteoclasts) rather than having a purely physico-chemical degradation, i.e. dissolution, because a cell-mediated degradation ensures that material degradation is not too fast compared to bone formation. However, by just relying on cells to reach material degradation and hence calcium and phosphate release, it is not possible to control the up-regulation or down-regulation of cells in the close surroundings of the material.

[0013] It is an object of the invention to provide a bone graft substitute in the form of an implantable three-dimensional scaffold comprising calcium phosphate and having pores and which is impregnated with a calcium and/or phosphate containing substance whereby the dissolution rate DR₃ of said scaffold is slower than the dissolution rate DR₅ of said calcium and/or phosphate containing substance.

[0014] The advantage of the bone graft substitute according to the invention lies in the improved in vivo response of calcium phosphate bone substitutes through selective calcium or phosphate release.

[0015] It is a further object of the invention to provide a method for manufacturing a bone graft substitute characterized by impregnating a three-dimensional scaffold comprising calcium phosphate having interconnected pores with a calcium and/or phosphate containing substance, whereby the chemical composition and integrity of said scaffold remains essentially unaffected by said impregnation with said calcium and/or phosphate containing substance. The impregnation can be effected e.g. by spraying, soaking, tipping.

[0016] It is a further object of the invention therefore to load a matrix or scaffold that is degraded by cells like β-TCP with a compound that can spontaneously dissolve in vivo, like calcium chloride (CaCl₂). The main condition for that purpose is to use a compound that is soluble in vivo. Further in the text, the term of “scaffold” will be used to designate a material resorbed by cell-mediation and the term of “drug” when reference is made to the compound that is soluble in vivo and contains calcium and/or phosphate ions.

[0017] Typical calcium phosphate bone graft materials of interest for the scaffold (beside β-TCP) are hydroxyapatite (Ca₃(PO₄)₂·OH; HA; sintered or non-sintered), di-calcium phosphate (CaHPO₄·DCP), octacalcium phosphate (Ca₅H₂(PO₄)₆·5H₂O; OCP), α-tricalcium phosphate (α-Ca₃(PO₄)₂; α-TCP), α-calcium pyrophosphate (α-Ca₅P₂O₇·α-PPP), and β-calcium pyrophosphate (β-Ca₅P₂O₇·β-PPP). Of interest are also all calcium phosphates having the general apatite structure according to x-ray diffraction, but not having the exact stoichiometry of hydroxyapatite. This includes for example calcium-deficient hydroxyapatite (Ca₅(PO₄)₃·(HPO₄)·(OH); CDHA—sometimes called “tricalcium phosphate”), carbonated apatites, and more generally all ion-substituted apatites.

[0018] All potential scaffolds can also contain some foreign ions in their structure (not only hydroxyapatite). Surprisingly it has been found that many ionic substitutions exist in calcium phosphates. Of particular interest are Mg, Sr, Zn, Si, Na, K, Li and CI as potential ions for b-TCP, b-CPP, a-CPP and b-CP. For HA, OCP, DCP and DCDP, the latter ions as well as CO₃⁻ ions or SO₄²⁻ ions can be used.

[0019] Typical calcium-containing ionic materials that can be used as calcium “drug” are calcium chloride (anhydrous: CaCl₂·monohydrate; CaCl₂·H₂O, dihydrate: CaCl₂·2H₂O, or hexahydrate: CaCl₂·6H₂O), di-calcium phosphate dihydrate (CaHPO₄·2H₂O; DCPD), calcium sulphate dihydrate (CaSO₄·2H₂O; GSP), calcium sulphate hemihydrate (CaSO₄·½H₂O; CSH), calcium sulphate (CaSO₄), calcium acetate (anhydrous: Ca(CH₃COO)₂), monohydrate: Ca(CH₃COO)₂·H₂O, or dihydrate Ca(CH₃COO)₂·2H₂O, calcium citrate (Ca₃(C₆H₅O₇)·4H₂O), calcium fumarate (Ca₃(C₆H₅O₇)·3H₂O), calcium glycerophosphate (Ca₃H₂(OH)₂PO₄), calcium lactate (Ca₃(C₆H₁₀O₇)·2H₂O), calcium malate (dl-malate: CaH₂C₆H₄O₇·3H₂O, 1-malate: CaH₂C₆H₄O₇·2H₂O, or malate dihydrogen: Ca(H₂C₆H₄O₇)·2H₂O, calcium maleate (Ca₃(C₆H₅O₇)·H₂O), calcium malonate (Ca₃(C₆H₅O₇)·4H₂O), calcium oxalate (CaC₂O₄), calcium oxalate hydrate (CaC₂O₄·H₂O), calcium salicylate (Ca(C₆H₄O₂)₂·H₂O), calcium succinate (Ca₃(C₆H₅O₇)·3H₂O), calcium tartrate (d-tartrate: CaC₄H₆O₆·4H₂O; dl-tartrate: CaC₄H₆O₆·4H₂O; mesotartrate: CaC₄H₆O₆·3H₂O), and calcium valerate (Ca₃(C₆H₁₄O₇)₂).
Typical phosphate-containing ionic materials that can be used as phosphate “drug” are DCPD, sodium phosphate (Na$_2$HPO$_4$, NaH$_2$PO$_4$, or a mixture thereof; non-hydrated or hydrated species like Na$_2$HPO$_4$·2H$_2$O, Na$_2$HPO$_4$·7H$_2$O, Na$_3$HPO$_4$·12H$_2$O, NaH$_2$PO$_4$·H$_2$O, NaH$_2$PO$_4$·2H$_2$O), calcium glycero phosphate (CaC$_6$H$_7$(OH)$_3$PO$_4$), potassium orthophosphate (K$_2$PO$_4$), dihydrogen potassium orthophosphate (K$_2$HPO$_4$), monohydrogen potassium orthophosphate (K$_2$HPO$_4$), and sodium hydrogen phosphate (Na$_2$HPO$_4$·10H$_2$O and Na$_2$PO$_4$·12H$_2$O).

Drug Solubility

The solubility of the drug in an aqueous solution having a physiological ionic strength (0.15M) and a pH of 7.4 at 37°C should be in an adequate range, typically superior to 2 mM, preferably superior to 10 mM. An adequate range appears to be between 10 mM and 1M.

It is particularly useful to have a rather low solubility in physiological conditions because the release rate is accordingly low. However, a low solubility is not adequate for loading because the loaded amount is limited. So, compounds that present a rather low solubility at physiological conditions and a high solubility in other conditions (e.g., Na$_2$HPO$_4$·12H$_2$O is much more soluble at 90°C than at 37°C.) are interesting because loading can be made in these advantageous conditions and release in physiological conditions is still slow.

Porosity and Pore Size

To load the scaffold with calcium and/or phosphate ions, it is necessary to have a porous scaffold, preferably a scaffold with interconnected pores to allow drug invasion into the pores. A porosity in the range of 40 to 95%, preferably of 55 to 80% is advantageous. It is important to have a slow release, hence implying that the pores should be relatively small (the smaller they are, the slower the release of calcium and phosphate ions will be). Therefore, the scaffold should preferably contain micropores or even nanopores. Ideally, at least 10% of the total volume (preferably 20%) should be constituted of micropores or nanopores. It is further advantageous to have tortuous pores. Tortuosity values larger than 5 are preferred.

The ideal pore size depends on the purpose of the bone graft substitute. Small pores (or a large specific surface area) will favor a rapid resorption. So, the resorption rate will increase in the order micropore-micropore-nanopore. However, since the scaffold is meant to be resorbed by cells, a fast resorption also requires the presence of cells. In other words, scaffolds that have a size superior to a few millimeters should preferably have an interconnected porous network with interconnections larger than 30 to 50 microns to allow bone vessel ingrowth hence leading to rapid bone ingrowth and scaffold resorption. In this case, it is important to have about 30 to 70% of the scaffold volume constituted of macropores, preferably 40 to 60%.

Loading Method

Two main loading methods can be used for manufacturing the bone graft substitute, i.e., depending on the invention.

(i) Soaking in a Concentrated Drug Solution

The first possibility is to create a solution containing calcium and/or phosphate ions, soak the porous scaffold into this solution, and let it dry. The pores are then filled with the salt used for the preparation of the calcium and/or phosphate containing solution.

In that respect, it is advantageous to soak the scaffold in a small amount of solution (for example by placing the scaffold vertically into a solution—the solution reaching only the bottom third of the scaffold) and let it dry. During drying, there is constantly a capillary rise from the solution to the top of the scaffold, leading finally to a very large loading of the scaffold with the soluble calcium and/or phosphate entities.

The temperature of the soaking solution is important. Some compounds are much more soluble at low or at high temperature in water. So, it can be advantageous to prepare a solution at e.g. 80°C and perform the impregnation and drying at the latter temperature. For other compounds, it can be advantageous to soak the sample with a cold solution (e.g., of 5°C.) and then perform drying at e.g. 60°C. So the solution, soaking and drying temperatures can be varied and the temperature at which the impregnation solution is prepared is of some importance due to the temperature dependence of some solubilities.

(ii) Soaking the Scaffold with a Slurry

The second possibility is to create a slurry containing drug particles and soak the scaffold with the slurry. A requirement is to have drug particles small enough to penetrate the scaffold porosity. Impregnation may be performed under vacuum or under varying pressure cycles, e.g., vacuum—room pressure cycles.

Drying

The procedures of impregnation and drying can require different conditions. Impregnation is preferably performed at slower rate than drying (drying may start when there is no more liquid surrounding the scaffold, but still some liquid within the scaffold).

Impregnation Geometry

Dimension and shape of the impregnation will vary depending on the loading method and the pore size. Micropores are likely to be completely filled with the soluble calcium and/or phosphate compound. On the other side, macropores are likely to be only partially filled. So, the geometry will vary depending on the pore size.

Release Rate

The dissolution rate DR$_D$ of the scaffold should preferably be null in serum or “simulated body fluid” whereas the dissolution rate DR$_D$ of the drug should be superior to zero in such conditions.

By “simulated body fluid” an aqueous solution is understood which has (at pH 7.4, 37°C, and in equilibrium with a gas atmosphere containing 5% CO$_2$) the ionic strength as serum and the same supersaturation towards hydroxyapatite precipitation than serum.

Not all calcium phosphates can be used as scaffolds because not all of them are insoluble in serum (or simulated body fluid) in physiological conditions (pH 7.4, 37°C, 5% CO$_2$ gas atmosphere). Only calcium phosphates insoluble in serum can be considered as scaffold material. All other calcium phosphates which are soluble in serum (or simulated body fluid) in physiological conditions (pH 7.4, 37°C, 5% CO$_2$ gas atmosphere) can be considered as drug material. When a material is insoluble in serum (or simulated body fluid) in physiological conditions (pH 7.4, 37°C, 5% CO$_2$ gas atmosphere).
In special embodiments said calcium and/or phosphate containing substance is ionic and/or crystalline. In a further embodiment said calcium and/or phosphate containing substance has a degree of crystallinity higher than 80%, preferably higher than 90%.

In a special embodiment said calcium and/or phosphate containing substance is not chemically bound to said scaffold and/or is adhering only physically to said scaffold. The dissolution rate $DR_s$ as measured in a phosphate-buffered solution (PBS) at pH 7.4 is preferably at least 10 times larger than said dissolution rate $DR_p$. Alternatively the dissolution rate $DR_p$ as measured in an aqueous citric acid solution at pH 3.0 is at least 10 times larger than said dissolution rate $DR_p$. Preferably the dissolution rate $DR_p$ and the dissolution rate $DR_s$ of 0 in serum or simulated body fluid having a pH 7.4 at 37°C and 5% CO₂ in the atmosphere.

Said calcium and/or phosphate containing substance is preferably rigid. The degree of porosity of said scaffold is preferably 40-95%, and most preferably 55 to 80%.

The scaffold may have micropores with a mean diameter $D_{microp}$ smaller than 10 microns, preferably smaller than 1 micron. Further said scaffold may have macropores with a mean diameter $D_{macrop}$ in the range of 0.03 to 1 mm. The specific surface area of said scaffold preferably is superior to 1 m²/g, preferably superior to 4 m²/g. A scaffold with a high SSA value is indicative of a fine micro or nanostructure. Such a fine structure leads to a slow release rate.

The tortuosity of the pores of said scaffold is preferably larger than 3, most preferably larger than 5. Preferably the pores of said scaffold are interconnected and the size of interconnections between the pores is larger than 30 to 50 microns.

In a special embodiment said calcium and/or phosphate containing substance comprises calcium chloride. In a further embodiment said calcium and/or phosphate containing substance comprises $Ca\textsubscript{HPO}_4$.

The ratio WD/WS between the weight WD of said calcium and/or phosphate containing substance and the weight WS of said scaffold preferably is comprised in the range of 0.1 to 10, preferably of 0.5 to 2.

In a special embodiment said calcium phosphate of said scaffold is beta-tricalcium phosphate ($\beta$-TCP).

Preferably said scaffold has a volume larger than 10 mm³, more preferably larger than 50 mm³.

In a special embodiment the pores of said scaffold are filled with said calcium and/or phosphate containing substance to an extent of 1% to 50 vol.-%, preferably of 5% to 25 vol.-%.

In a special embodiment of the method according to the invention the calcium and/or phosphate containing substance is calcium chloride and the deposition is performed by applying an aqueous solution of calcium chloride to said scaffold. In another embodiment the calcium and/or phosphate containing substance is $Ca\textsubscript{HPO}_4$ and that the deposition is performed by applying an aqueous solution of calcium chloride to said scaffold. The aqueous solution applied to said scaffold may be dried in an atmosphere having a relative humidity below 10%, preferably below 5%. Preferably said aqueous solution applied to said scaffold is dried for more than 1 day. The aqueous solution purposely has a concentration of 12.5% to 50%.

In a special embodiment the impregnation is performed in two successive steps. One step may comprise the impregnation with a calcium-containing substance and another step may comprise the impregnation with a phosphate-containing substance. It can be purposeful to use different solvents to this effect and to first load the scaffold with a calcium-containing salt and second load the scaffold with a phosphate-containing salt using a solvent or loading conditions (e.g. T in/at which the calcium-containing salt is poorly-soluble. It is also possible to load twice the same block, for example by placing the block upside down, since the distribution of the crystalline calcium and/or phosphate containing substances present within the block will not be homogeneously distributed (it is more close to the solution and the block walls, and less in the center and at the top).

A BRIEF DESCRIPTION OF THE DRAWINGS

Several embodiments of the invention will be described in the following by way of examples and with reference to the accompanying drawings in which:

FIG. 1 shows the relationship between expected and measured CaCl₂ loading in the samples. The symbol (▲) represents the calcium chloride loading. The dotted line represents the curve “Expected amount—measured amount”. The results obtained with 7 mL of 0.50 g/mL solution are not shown because the samples were very difficult to dry.

FIG. 2 shows the relationship between expected and measured $Ca\textsubscript{HPO}_4$ loading in the samples. (▲) Measured amount of $Ca\textsubscript{HPO}_4$ added into the flask+block—measured by determining the difference of weight of the flask+block before and after adding the $Ca\textsubscript{HPO}_4$ solution (including drying); (Δ) $Ca\textsubscript{HPO}_4$ loading in the block—measured by determining the difference of weight of the block before and after impregnation. The dotted line represents the expected amount of $Ca\textsubscript{HPO}_4$ added into the flask and that could potentially “load” the block.

FIG. 3 shows the relationship between expected and measured $Na\textsubscript{2}HPO\textsubscript{4}2H\textsubscript{2}O$ loading in the samples. (▲) Measured amount of $Na\textsubscript{2}HPO\textsubscript{4}2H\textsubscript{2}O$ added into the flask+block—measured by determining the difference of weight of the flask+block before and after adding the $Na\textsubscript{2}HPO\textsubscript{4}2H\textsubscript{2}O$ solution (including drying); (LS) $Na\textsubscript{2}HPO\textsubscript{4}2H\textsubscript{2}O$ loading in the block—measured by determining the difference of weight of the block before and after impregnation. The upper dotted line represents the expected amount of $Na\textsubscript{2}HPO\textsubscript{4}2H\textsubscript{2}O$ added into the flask and that could potentially “load” the block. The lower dotted line represents the expected amount of $Na\textsubscript{2}HPO\textsubscript{4}$ added into the flask and that could potentially “load” the block, assuming that $Na\textsubscript{2}HPO\textsubscript{4}2H\textsubscript{2}O$ is transformed into $Na\textsubscript{2}HPO\textsubscript{4}$ at 95°C.

FIG. 4 shows the amount of $Na\textsubscript{2}HPO\textsubscript{4}2H\textsubscript{2}O$ released during (Δ) 1 h or (▲) 13 h incubation in 1L deionized water according to example 3.

FIG. 5 shows the release of calcium chloride amount (in %) as a function of incubation time in deionized water as described in example 4.

FIG. 6 is a schematic representation of the impregnation test setup (profile) of example 5.
Example 1

[0064] The aim was to perform impregnation tests of porous β-TCP blocks to assess how such blocks can be loaded with a calcium and/or phosphate containing salt.

[0065] The porosity of the porous β-TCP blocks was in the range of 69 to 77%. The porosity consisted of roughly 54% macropores (mean diameter close to 0.3-0.4 mm) and 25-33% micropores (mean diameter in the range of 1-10 micrometers).

[0066] The samples were calcined at 500°C for 1 h prior to the impregnation tests to remove organic residues present on the block surface (without calcination, the samples were so hydrophobic that they were floating in aqueous solutions).

[0067] Impregnation tests were performed with a 0.25 g/mL and 0.50 g/mL calcium chloride solution.

[0068] A 2^2 factorial design of experiments with three repeats was performed with the following factors: Factor A: CaCl₂ concentration (0.25 or 0.50 g/mL); Factor B: Liquid amount (3.5 or 7.0 mL). Each of the 14x14 mm cylinders was placed standing in a snap cap flask. The solution was then slowly injected at the bottom of the snap-cap flask to allow impregnation through capillarity. The cylinder top was always protruding out of the solution. The samples were then inserted into the drying cupboard tempered at 60°C. The ventilation of the drying cupboard was set at its maximum.

[0069] The weight of the flask+solution+samples was measured at regular interval to determine the point at which constant weight was reached. After 24 h, most samples but 3 (those made with 7 mL of 0.50 g/mL solution) appeared to be dry. Therefore, these 9 samples were removed from the flask and the weight was determined without flask. The 3 samples made with 7 mL of 0.50 g/mL solution were kept for a longer time. The drying temperature was increased to 80°C, 36 h after the experiment start.

[0070] Both Ca solutions were sucked by the porous block within seconds. Since an excess of liquid was used, some solution was left besides the samples.

[0071] After drying, a different picture was revealed depending on the soaking solution and the amount: the blocks soaked in calcium chloride appeared mostly "clean"; whereas the blocks soaked in 7 mL of 0.50 g/mL CaCl₂ solution were encrusted in large CaCl₂ residues, suggesting that too much CaCl₂ was used.

[0072] Interestingly, drying was very slow and as soon as the samples were retrieved from the drying cupboard and left in the lab, the block surface became wet, suggesting that the samples were very hygroscopic due the presence of calcium chloride. After 3.5 days, a liquid could still be found below the crust formed around the samples produced with 7 mL of 0.50 g/mL CaCl₂.

[0073] The amount of calcium chloride present within the samples was slightly lower than expected, perhaps because the calcium chloride crystals used to produce the Ca chloride solution contained water and/or because some of the Ca chloride was left in the snap cap flask. The loading efficiency was close to 84%, with noticeable difference between 7 mL of 0.25 g/mL solution and 3.5 mL of 0.50 g/mL solution.

Example 2

[0074] Perform impregnation tests of porous β-TCP blocks to assess how such blocks can be loaded with a phosphate containing salt.

[0075] Materials and Methods

[0076] The porosity of the porous β-TCP blocks was in the range of 69 to 77%. The porosity consisted of roughly 54% macropores (mean diameter close to 0.3-0.4 mm) and 25-33% micropores (mean diameter in the range of 1-10 micrometers).

[0077] The samples were calcined at 500°C for 1 h prior to the impregnation tests to remove organic residues present on the block surface (without calcination, the samples were so hydrophobic that they were floating in aqueous solutions).

[0078] Impregnation tests were performed with a 0.50 g/mL di-potassium hydrogen phosphate solution (K₂HPO₄).

[0079] Each of the 14x14 mm cylinders was placed standing in a snap cap flask. The solution was then slowly injected at the bottom of the snap-cap flask to allow impregnation through capillarity. 6 different volumes were injected: 1.8, 2.7, 3.6, 4.5, 5.6 and 6.3 mL. These volumes correspond to an expected K₂HPO₄ amount of 0.9, 1.35, 1.8, 2.25, 2.8 and 3.15 g. Two cylinders were prepared for each solution volume. The cylinder top was always protruding out of the solution. The samples were then inserted into the drying cupboard tempered at 95°C. The ventilation of the drying cupboard was set at its maximum.

[0080] The weight of the flask+solution+samples was measured at regular interval to determine the point at which constant weight was reached. The sample weight was determined to assess how much di-potassium hydrogen phosphate was present in the block pores.

[0081] Results

[0082] The solution was sucked by the porous block within seconds. Since an excess of liquid was used, some solution was left besides the samples, especially with a high liquid amount. The samples looked very good, with hardly any crystals protruding at the sample surface.

[0083] The experimental results show a good agreement between measured and expected amount (FIG. 1) of K₂HPO₄ present in the β-TCP block. However, it appears that beyond an expected amount of ~2.5 g, most of the additional K₂HPO₄ amount remains in the flask and does not load the β-TCP block.

Example 3

Aim

[0084] Perform impregnation tests of porous β-TCP blocks to assess how such blocks can be loaded with a phosphate containing salt.

[0085] Materials and Methods

[0086] The porosity of the porous β-TCP blocks was in the range of 69 to 77%. The porosity consisted of roughly 54% macropores (mean diameter close to 0.3-0.4 mm) and 25-33% micropores (mean diameter in the range of 1-10 micrometers).

[0087] The samples were calcined at 500°C for 1 h prior to the impregnation tests to remove organic residues present on the block surface (without calcination, the samples were so hydrophobic that they were floating in aqueous solutions).
Impregnation tests were performed with a 0.50 g/mL di-sodium hydrogen phosphate solution (Na$_2$HPO$_4$·2H$_2$O). Since the solubility of di-sodium hydrogen phosphate solution is relatively low at room temperature, the solution was heated up at 95°C. At that temperature, a clear solution was obtained.

Each of the 14×14 mm cylinders was placed standing in a snap cap flask. The solution (kept at 95°C) was then slowly injected at the bottom of the snap-cap flask to allow impregnation through capillarity. 3 different volumes were injected: 2.7, 4.5, and 6.3 mL, corresponding to 1.35, 2.25 and 3.15 g of Na$_2$HPO$_4$·2H$_2$O. Two cylinders were prepared for each solution volume. The cylinder top was always protruding out of the solution. The samples were then inserted into the drying cupboard tempered at 95°C. The ventilation of the drying cupboard was set at its maximum.

The weight of the flask+solution+samples was measured at regular interval to determine the point at which constant weight was reached. The sample weight was determined to assess how much di-potassium hydrogen phosphate was present in the block pores.

Each sample was then placed into a small porous cage produced by joining together two BD Falcon filters (Mesh size: 70 micrometers). The cage was lowered into a 1 L deionized water. Stirring was performed with a large magnetic bar (6 cm in length) at 50 RPM. The samples were removed from the solution after 1 h or 3 h (1 sample per time and per composition). The samples were then dried at 95°C and weighed to determine the amount of Na$_2$HPO$_4$·2H$_2$O released during the test.

As control group, 6 β-TCP blocks that had not been impregnated in sodium hydrogen phosphate solution were also tested.

The solution was sucked by the porous block within seconds. Since an excess of liquid was used, some solution was left besides the samples, particularly at a high loading. Interestingly, the sample surface was not “clean” but covered with a thick material layer. The amount of material left after drying the flask was lower than expected assuming that all Na$_2$HPO$_4$·2H$_2$O present in the Na$_2$HPO$_4$·2H$_2$O solution is left in the flask. This suggests that either the solution concentration was too long (for example due to adsorbed water into the initial powder) or the Na$_2$HPO$_4$·2H$_2$O was transformed into Na$_2$HPO$_4$ during drying. Since there is a very good correlation between the experimental points and the predictions made assuming the presence of Na$_2$HPO$_4$ (FIG. 1), the latter explanation is probably correct.

The results also show that the loading efficiency decreases with an increase in added Na$_2$HPO$_4$·2H$_2$O amount, suggesting that the block pores cannot be fully filled with Na$_2$HPO$_4$·2H$_2$O. The point at which the loading efficiency drastically decreases is close to 1.35 g.

All release solutions contained particles in suspension after the release test suggesting that some particles precipitated in the release solution. This could explain why released Na$_2$HPO$_4$·2H$_2$O amount was not affected by the initial loading and the release duration (FIG. 2). In any case, this experiment shows that the release rate of Na$_2$HPO$_4$·2H$_2$O proceeds relatively slowly, for example compared to the results obtained with CaCl$_2$, in the following example 4.

The mean weight loss of pure β-TCP blocks (without loading) was slightly positive after 3 h (0.005±0.012 g) but not significantly larger than zero, which means that the dissolution rate DR$_p$ of sodium hydrogen phosphate was two order of magnitude larger than the dissolution rate DR$_p$ of β-TCP (since 0.3 to 0.8 grams of sodium hydrogen phosphate were dissolved in the release medium within the same duration).

Example 4

Aim

Assess the rate of calcium chloride release

Materials and Methods

The porosity of the porous β-TCP blocks was in the range of 69 to 77%. The porosity consisted of roughly 54% macropores (mean diameter close to 0.3-0.4 mm) and 25-33% micropores (mean diameter in the range of 1-10 micrometers).

The samples were calcined at 500°C for 1 h prior to the impregnation tests to remove organic residues present on the block surface (without calcination, the samples were more hydrophilic that they were floating in aqueous solutions).

Impregnation tests were performed with a 0.25 g/mL calcium chloride solution (CaCl$_2$).

Each of the 14×14 mm cylinders was placed standing in a snap cap flask. The solution (kept at 95°C) was then slowly injected at the bottom of the snap-cap flask to allow impregnation through capillarity. 2 different volumes were injected: 10.5 and 14 mL. Three cylinders were prepared for each solution volume. The cylinder top was always protruding out of the solution. The samples were then inserted into the drying cupboard tempered at 95°C. The ventilation of the drying cupboard was set at its maximum.

The weight of the flask+solution+samples was measured at regular interval to determine the point at which constant weight was reached. After 24 h, the samples were dry. The sample weight was determined to assess how much di-potassium hydrogen phosphate was present in the block pores.

Each sample was then placed into a small porous cage produced by joining together two BD Falcon filters (Mesh size: 70 micrometers). The cage was lowered into a 1 L deionized water. Stirring was performed with a large magnetic bar (6 cm in length) at 50 RPM. The samples were removed from the solution after 10 min, 1 h or 6 h (1 sample per time and per composition). The samples were then dried at 95°C and weighed to determine the amount of CaCl$_2$ released during the incubation.

As control group, 6 β-TCP blocks that had not been impregnated in calcium chloride solution were also tested.

Results

The solution was sucked by the porous block within seconds. Since an excess of liquid was used, some solution was left besides the samples. Between 1.38 and 1.64 g calcium chloride could be loaded into the blocks. Despite a difference in the initial volume (10.5 or 14 mL), the loaded calcium chloride did not vary much, suggesting that a loading plateau is reached. Using the density of calcium chloride, it could be calculated that 42 to 47% of the pore volume was filled with calcium chloride.

The release rate was very fast, with roughly 50% released within 10 minutes and 80-85% after 1 h. At 6 h, the samples were free of calcium chloride. In other words, 1.38 to
1.64 grams of calcium chloride were dissolved within 6 h. Contrarily, the mean weight loss of pure β-TCP blocks (without calcium chloride) was slightly positive after 6 h (0.01±0.01 g) but not significantly larger than zero, which means that the dissolution rate DRp of calcium chloride was more than 100 times larger than the dissolution rate DRs of β-TCP.

Example 5

Aim

[0110] Load porous β-TCP blocks with Na₂HPO₄·2H₂O and measure the Na₂HPO₄·2H₂O release rate

[0111] Materials and Methods

[0112] The porosity of the porous β-TCP blocks was in the range of 69 to 77%. The porosity consisted of roughly 54% macro pores (mean diameter close to 0.3-0.4 mm) and 25-33% micropores (mean diameter in the range of 1-10 micrometers)

[0113] The samples were calcined at 500°C for 1 h prior to the impregnation tests to remove organic residues present on the block surface (without calcination, the samples were so hydrophobic that they were floating in aqueous solutions).

[0114] Impregnation tests were performed with a 0.50 g/mL Na₂HPO₄·2H₂O release rate (Na₂HPO₄·2H₂O).

[0115] Each of the 14×14 mm cylinders was placed standing in a Petri dish (inner dimensions: diameter: 8.7 cm; height: 1.1 cm). 24.3 mL of the solution (kept at 95°C) was slowly injected at the bottom of the petri dish. The blocks were then press-fitted into a plastic membrane covering the Petri dish (FIG. 1). The bottom of the samples touched the bottom of the Petri dish, whereas the cylinder top protruded out of the membrane covering the Petri dish. The Petri dish samples were then inserted into the drying cupboard tempered at 95°C. The ventilation of the drying cupboard was set at its maximum.

[0116] The weight of the flask solution samples was measured at regular intervals to determine the point at which constant weight was reached. The sample weight was determined to assess how much Na₂HPO₄·2H₂O was present in the block pores.

[0117] Results

[0118] The solution was sucked by the porous block within seconds. The samples could be loaded with the Na₂HPO₄·2H₂O crystals, but large agglomerates were present on the block surface and quite some crystals remained at the bottom of the Petri dish. Large differences of loaded amount were observed, since values ranged from 0.7 to 1.2 g.

[0119] While various descriptions of the present invention are described above, it should be understood that the various features can be used singly or in any combination thereof. The scope of the present invention is accordingly defined as set forth in the appended claim.

1. A bone graft substitute in the form of an implantable three-dimensional scaffold comprising calcium phosphate and having pores, wherein:
   (i) the scaffold is impregnated with a calcium and/or phosphate containing substance;
   (ii) the dissolution rate DRₗ of said scaffold being slower than the dissolution rate DRₛ of said calcium and/or phosphate containing substance used for the impregnation of the scaffold, whereby and (iii) the chemical composition and integrity of the scaffold remains essentially unaffected by said impregnation with said calcium and/or phosphate containing substance.

2. The bone graft substitute according to claim 1, wherein said calcium and/or phosphate containing substance is ionic.

3. The bone graft substitute according to claim 1, wherein said calcium and/or phosphate containing substance is crystalline.

4. The bone graft substitute according to claim 3, wherein said calcium and/or phosphate containing substance has a degree of crystallinity higher than 80%.

5. The bone graft substitute according to claim 1, wherein said calcium and/or phosphate containing substance is not chemically bound to said scaffold.

6. The bone graft substitute according to claim 1, wherein said calcium and/or phosphate containing substance only adheres physically to said scaffold.

7. The bone graft substitute according to claim 1, wherein said dissolution rate DRₛ as measured in a phosphate-buffered solution (PBS) at pH 7.4 is at least 10 times larger than said dissolution rate DRₛ.

8. The bone graft substitute according to claim 1, wherein said dissolution rate DRₛ as measured in an aqueous citric acid solution at pH 3.0 is at least 10 times larger than said dissolution rate DRₛ.

9. The bone graft substitute according to claim 1, wherein said dissolution rate DRₛ and said dissolution rate DRₛ in serum or simulated body fluid having a pH 7.4 at 37°C and 5% CO₂ in the atmosphere.

10. The bone graft substitute according to claim 1, wherein said calcium and/or phosphate containing substance is rigid.

11. The bone graft substitute according to claim 1, wherein the degree of porosity of said scaffold is 40-95%.

12. The bone graft substitute according to claim 1, wherein said scaffold has micropores with a mean diameter Dₘicrop smaller than 10 microns.

13. The bone graft substitute according to claim 1, wherein said scaffold has macropores with a mean diameter Dₘacro in the range of 0.03 to 1 mm.

14. The bone graft substitute according to claim 1, wherein said scaffold has a specific surface area superior to 1 m²/g.

15. The bone graft substitute according to claim 1, wherein the tortuosity of the pores of said scaffold is larger than 5.

16. The bone graft substitute according to claim 1, wherein said pores are interconnected.

17. The bone graft substitute according to claim 13, wherein the size of interconnections between the pores is larger than 30 microns.

18. The bone graft substitute according to claim 1, wherein said calcium and/or phosphate containing substance comprises calcium chloride.

19. The bone graft substitute according to claim 1, wherein said calcium and/or phosphate containing substance comprises KH₂PO₄.

20. The bone graft substitute according to claim 1, wherein the ratio WD/WS between the weight WD of said calcium and/or phosphate containing substance and the weight WS of said scaffold is comprised in the range of 0.1 to 0.

21. The bone graft substitute according to claim 1, wherein said calcium phosphate of said scaffold is beta-tricalcium phosphate (β-TCP).

22. The bone graft substitute according to claim 1, wherein said scaffold has a volume larger than 10 mm³.
23. The bone graft substitute according to claim 1, wherein said pores are filled with said calcium and/or phosphate containing substance to an extent of 1% to 50 vol.-%.

24. A method for manufacturing a bone graft substitute, the method comprising impregnating a three-dimensional scaffold comprising calcium phosphate having interconnected pores with a calcium and/or phosphate containing substance, wherein the chemical composition and integrity of said scaffold remains essentially unaffected by said impregnation with said calcium and/or phosphate containing substance.

25. The method according to claim 24, wherein the calcium and/or phosphate containing substance is calcium chloride and the impregnating is performed by applying an aqueous solution of calcium chloride to said scaffold.

26. The method according to claim 24, wherein the calcium and/or phosphate containing substance is K₂HPO₄ and the impregnating is performed by applying an aqueous solution of calcium chloride to said scaffold.

27. The method according to claim 24, wherein said aqueous solution applied to said scaffold is dried in an atmosphere having a relative humidity of 0%.

28. The method according to claim 24, wherein said aqueous solution applied to said scaffold is dried for more than 1 day.

29. The method according to claim 24, wherein said aqueous solution has a concentration of 12.5% to 50%.

30. The method according to claim 24, wherein said impregnating is performed in two successive steps.

31. The method according to claim 30, wherein one step of the two successive steps comprises impregnating the scaffold with a calcium-containing substance and another step of the two successive steps comprises impregnating the scaffold with a phosphate-containing substance.

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