



- (51) **International Patent Classification:**
C12P 19/44 (2006.01) *C07H 15/06* (2006.01)
C12P 7/64 (2006.01)
- (21) **International Application Number:**
PCT/US2016/012901
- (22) **International Filing Date:**
11 January 2016 (11.01.2016)
- (25) **Filing Language:** English
- (26) **Publication Language:** English
- (30) **Priority Data:**
- | | | |
|------------|------------------------------|----|
| 62/102,310 | 12 January 2015 (12.01.2015) | US |
| 62/141,679 | 1 April 2015 (01.04.2015) | US |
| 62/157,019 | 5 May 2015 (05.05.2015) | US |
| 62/198,736 | 30 July 2015 (30.07.2015) | US |
| 62/198,740 | 30 July 2015 (30.07.2015) | US |

BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IR, IS, JP, KE, KG, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

- (71) **Applicant:** LOGOS TECHNOLOGIES, LLC [US/US];
2701 Prosperity Avenue, Fairfax, VA 22031 (US).
- (72) **Inventors:** LOHITHARN, Nattaporn; 25260 Lake Shore
Sq Unit 205, Chantilly, VA 20152 (US). DERR, Daniel;
920 Loring Street, San Diego, CA 92109 (US).
- (74) **Agent:** AGRIS, Cheryl, H.; Agris & von Natzmer, LLP,
43 West 43rd Street, Suite 104, New York, NY 10036
(US).
- (81) **Designated States** (unless otherwise indicated, for every
kind of national protection available): AE, AG, AL, AM,
AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY,

(84) **Designated States** (unless otherwise indicated, for every
kind of regional protection available): ARIPO (BW, GH,
GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, ST, SZ,
TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU,
TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE,
DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU,
LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK,
SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ,
GW, KM, ML, MR, NE, SN, TD, TG).

Declarations under Rule 4.17:

- as to applicant's entitlement to apply for and be granted a patent (Rule 4.17(ii))
- as to the applicant's entitlement to claim the priority of the earlier application (Rule 4.17(iii))

Published:

- with international search report (Art. 21(3))
- before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments (Rule 48.2(h))

(54) **Title:** PRODUCTION OF RHAMNOLIPID COMPOSITIONS

(57) **Abstract:** Provided are organic solvent free methods for obtaining compositions comprising one or more rhamnolipids as well as compositions obtainable from said methods.



PRODUCTION OF RHAMNOLIPID COMPOSITIONS

TECHNICAL FIELD

5 Provided are organic solvent-free processes for obtaining composition comprising rhamnolipids as well as said compositions.

BACKGROUND

Rhamnolipids (RLs) are interface-active glycolipids produced by various bacterial species. RLs consist of one (monorhamnosylipids or mono-rhamnolipids) or
10 two rhamnose units (dirhamnosylipids or di-rhamnolipids) and one or two (predominantly two) 3-hydroxy fatty acid residues. Rhamnolipids have particular surface-active properties such as, for example, a strong foaming ability, and are of interest for highly diverse technical applications, particularly as surfactants (reviewed in Randhawa et al. (2014) "Rhamnolipid biosurfactants—past, present, and future
15 scenario of global market", *Frontiers in Microbiology* 5:1-7; Muller et al. (2012) "Rhamnolipids—Next generation surfactants?", *J Biotechnol* 162(4):366-80; Banat et al. (2010) "Microbial biosurfactants production, applications and future potential" *Appl Microbiol Biotechnol* 87:427-444). Examples of potential applications include: (1) bioremediation and enhanced oil recovery(EOR) since they efficiently remove crude
20 oil and heavy metals from contaminated soil and facilitate bioremediation of oil spills due to their emulsification properties; (2) cosmetics: including cosmeceuticals for example for wound healing, burns, psoriasis and wrinkles; (3) detergents and cleaners: particularly for laundry products, shampoos and soaps given their surface active and emulsification properties and (4) agriculture. Rhamnolipids are currently
25 available in an agricultural anti-fungal product marketed as ZONIX™ (Proptera LLC).

Interest in RLs has increased since they can be prepared by means of fermentation based on renewable raw materials. However, rhamnolipids have only been available on the market in small amounts and at high prices partially due to cumbersome downstream processing methods. Various methods have been disclosed
30 for producing rhamnolipids (reviewed in, for example, Heyd et al. (2008) "Development and trends of biosurfactant analysis and purification using rhamnolipids as an example", *Anal Bioanal Chem* 391:1579–1590 and Smyth et al. (2010) "Isolation and Analysis of Low Molecular Weight Microbial Glycolipids", in

Handbook of Hydrocarbon and Lipid Microbiology, K. N. Timmis (ed.), Springer-Verlag, Berlin, pp. 3705-3724; Desai et al. (1997) "Microbial Production of Surfactants and Their Commercial Potential", Microbiol. Mol. Biol. Rev. 61: 47-64 and also disclosed in for example US 5656747, US4628030, US20140148588, CN102796781,
5 CN102766172, CN101787057, CN101845468, CN102432643, CN1908180, KR1020060018783).

Currently, the most common isolation procedure for rhamnolipids involves autoclaving fermentation broth right after fermentation is complete, followed by acidifying to precipitate rhamnolipids out. The solid rhamnolipids that result are
10 contaminated with solid cellular material from the organism used in the fermentation. RLs are isolated from these cellular solids by extracting them into an organic solvent such as ethyl acetate. After stripping the ethyl acetate, a concentrated oily form of the product results. However, the solvent extraction process brings along any hydrophobic impurities with it, potentially including unconsumed triacyl glyceride oil and antifoam from the fermentation process. Examples of other methods disclosed
15 include (1) aluminum sulphate precipitation of fermentation broth followed by extraction with organic solvent; (2) continuous ultrafiltration; (3) column chromatography (see, for example, Lebrón-Paler A (2008) "Solution and interfacial characterization of rhamnolipid biosurfactant from *P. aeruginosa* ATCC 9027" PhD
20 Dissertation University of Arizona, CN101787057, CN101407831, CN1908180) using either adsorption, ion exchange, reversed phase or normal phase columns which may be performed in combination with the precipitation step; (4) countercurrent chromatography (Zhang et al. "Separation and purification of six biosurfactant rhamnolipids by high-speed countercurrent chromatography utilizing novel solvent
25 selection method", Separation Science and Technology, in press (posted Nov. 24, 2015)); (5) selective crystallization followed by extraction or (6) foam fractionation/adsorption (see, for example, US 2015/0011741; Sarachat et al., (2010) "Purification and concentration of a rhamnolipid biosurfactant produced by *Pseudomonas aeruginosa* SP4 using foam fractionation", Bioresource Technology 101:
30 324-330). However, all of these methods have various disadvantages as well (e.g., expensive, do not scale well, resource intensive, etc.).

SUMMARY

Provided is an organic solvent-free process for obtaining composition comprising rhamnolipids comprising:

- (a) providing an aqueous medium comprising at least one rhamnolipid (RL);
- 5 (b) non-filtration based sterilization of said medium provided in (a);
- (c) separating a RL containing phase from waste in said sterilized medium in (b) to obtain said composition

The final product using the method set forth above comes from a natural process, has not been chemically modified, and has not come in contact with any
10 petroleum based organic solvents. The process or method set forth above eliminates the use of petroleum derived chemical solvents to extract rhamnolipids from aqueous medium (e.g., fermented medium or broth) and thus, it significantly reduces the production cost. Furthermore, the final rhamnolipid solution is naturally derived.

In a particular embodiment, the composition comprising rhamnolipids is a
15 clarified broth comprising at least about 5%, 6%, 7%, 8%, 9%, or 10% rhamnolipids by weight, wherein said method further comprises after step (a) and before step (b), (1) aging said medium provided in step (a) to obtain aged and sterilized medium. Also provided is a composition comprising said aged and sterilized medium obtainable from the process set forth above. The method may further comprise after step (a) and
20 before step (b) removing solid waste from said medium provided in step (a). The composition in the above specific embodiments may comprise between about 5% to about 10% RL.

In yet another embodiment, the method may further comprise after the sterilization step (b) and before the separation step (c) aging said sterilized medium of
25 step (a). In yet another embodiment, after step (a) and before said medium is aged in (1), solid waste may be removed from medium provided.

In yet another embodiment, the method comprises two aging steps and may comprise the following steps:

- (a) providing an aqueous medium (e.g., fermentation medium) comprising at
30 least one rhamnolipid;
- (b) aging said aqueous medium (e.g., fermentation medium) provided;
- (c) non-filtration based sterilization of said medium provided in (b);
- (d) aging said sterilized medium of (c) and

(e) separating a RL containing liquid phase from waste in said aged and sterilized medium in (d) to obtain said composition.

In yet even another embodiment, a composition comprising rhamnolipids may be a clarified broth which comprises at least about 5%, 6%, 7%, 8% 9%, or 10% rhamnolipids by weight. In a particular embodiment, the method for obtaining the composition comprising RLs, particularly a clarified broth may, comprise before step (b) aging said medium provided in step (a) then subsequently (b) sterilizing said aged medium via chemical treatment, in particular, peroxide treatment (e.g., hydrogen peroxide, and organic peroxides such as benzoyl peroxide and peroxyacetic acid and inorganic peroxides such as lithium peroxide, sodium peroxide, barium peroxide), and/or ultraviolet light radiation treatment; (c) separating a RL containing phase from the solid waste in said sterilized medium and optionally further comprising (c) (1) additionally sterilizing said RL containing phase and (c)(2) separating a RL containing phase from solid waste in said sterilized RL containing phase to obtain said clarified broth. In a particular embodiment, said sterilization step in step (b) is via chemical treatment and/or ultraviolet radiation treatment and the sterilization step in step (c)(2) is via heat.

Also provided is a composition obtainable after sterilizing said aged medium. Further provided are sterilized compositions obtainable using steps (b) and/or (c)(1) set forth above.

The methods set forth above may further comprise a decolorization and/or deodorization step. The decolorization and/or deodorization treatment may be performed before and/or after the sterilization step. Also provided is a composition comprising decolorized and/or deodorized medium obtainable using the methods set forth above.

In even yet another embodiment, after step (c), fluid or liquid may be removed from the separated RL containing phase to obtain a solid composition for example, by evaporation. In a particular embodiment, said composition is in for example, powder form or granular form.

In even yet another embodiment, said composition comprising rhamnolipids is a concentrated clarified broth comprising at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50% or at least about 60% rhamnolipids by weight. In yet a more particular embodiment, said concentrated

clarified broth may comprise between about 30—60% RL, about 30-40% RL or about 50-60% RL by weight.

In a particular embodiment, said medium provided in step (a) in the method set forth above is sterilized in step (b) by treating said medium with acid and said acidic medium obtained comprises a solid, liquid and oily phase and at least the liquid phase is removed and before step (c), the solid and optionally oily phases are neutralized to obtain a solution and in step (c) insoluble waste material is removed from said solution to obtain said composition comprising said concentrated clarified broth set forth above. Thus a method for obtaining concentrated clarified broth may comprise:

(a) providing aqueous medium (e.g., fermentation medium) comprising rhamnolipids (RLs);

(b) sterilizing said fermentation or culture medium provided in (a) via acid treatment to obtain an acidified liquid, oily and solid phase;

(c) removing said liquid phase obtained in (b) and treating said solid and optionally oily phase with base to obtain a neutralized solution;

(d) removing insoluble material from said solution obtained in (c) to obtain a composition comprising concentrated clarified broth comprising one or more rhamnolipids.

In yet another embodiment, the method for obtaining a composition comprising concentrated clarified broth, may further comprise an additional sterilization step and optionally a further separation step to remove insoluble waste from said solution. Furthermore, said method may further comprise after sterilization, decolorizing and/or deodorizing said sterilized concentrated clarified broth. The method may also further comprise removing liquid or fluid from said solution by evaporation obtained in step (c) to obtain a solid composition. The medium may be treated with acid and then neutralized either before or after the additional sterilization step.

In the methods set forth above, said solid and oily phases may be neutralized separately to obtain separate solutions. Alternatively, said solid and oily phases are combined and said combined solid and oily phases are neutralized to obtain a neutralized solution. Furthermore, in yet another embodiment, only the solid phase is neutralized.

The methods set forth above may further comprise decolorizing and/deodorizing said composition. Additionally, said composition may be dried.

In another embodiment, a method for obtaining a composition comprising rhamnolipids which is a concentrated clarified broth comprising at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50% or at least about 60% rhamnolipids by weight, may further comprise before and/or after said additional sterilization step aging said medium provided.

In particular, said method comprises:

- (a) providing an aqueous (e.g., fermentation) medium comprising one or more rhamnolipids;
- (b) optionally removing insoluble waste from said medium provided;
- (c) sterilizing said medium provided;
- (d) aging said sterilized medium of (c);
- (e) removing waste from said aged and sterilized medium;
- (f) treating medium obtained in (e) with acid, wherein said acidic medium obtained comprises a solid, liquid and oily phase and at least the liquid phase is removed and the solid and optionally oily phases are neutralized to obtain a neutralized solution;
- (g) removing solid waste from said neutralized solution of (f) to obtain said composition comprising said concentrated clarified broth.

Also provided are compositions comprising the neutralized solution obtainable according to the methods set forth above. Further provided are decolorized and/or deodorized compositions comprising at least about 30% RLs using or obtainable from methods set forth above.

The compositions obtained may be used as a surfactant in hand soaps, body washes, facial cleansers, tooth paste, shampoos, conditioners, conditioning shampoos, cosmetics, dish soap, laundry detergents, laundry pretreatment sprays, hard surface cleaners, porous surface cleaners, floor cleaners, automobile cleaners, enhanced oil recovery, wound dressings, agricultural anti-bacterials, anti-corrosion treatments, bioremediation, anti-stick films, anti-biofouling applications, fire extinguishing media, anti-virals, anti-molds, and anti-zoosporics. In one embodiment, the compositions containing higher amounts of RL (>30% wt/vol) or more particularly about (>50% wt/vol) obtained using the methods set forth above can be used to make a

rhamnolipid based product that can displace an anionic surfactant such as sodium lauryl sulfate (SLS) in personal care and household cleaner markets given that certain anionic surfactants are often sold at a concentration of about 30% for some application.

5

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 schematically shows an organic solvent free method for obtaining rhamnolipid rich clarified broth (CB).

10 Figure 2 schematically shows where the CB is concentrated by acid/base treatment to obtain concentrated clarified broth (CCB).

Figure 3 schematically shows obtaining CCB from hydrogen peroxide treated CB.

Figure 4 schematically shows obtaining CCB from hydrogen peroxide treated sterilized fermentation medium.

15 Figure 5 schematically shows obtaining CCB by first performing a separation step on the fermentation broth and performing the sterilization step later in the process.

Figure 6 schematically shows a method for obtaining CCB by performing separation, sterilization, aging and acidification steps.

20

DEFINITIONS

Where a range of values is provided, it is understood that each intervening value, to the tenth of the unit of the lower limit unless the context clearly dictates otherwise, between the upper and lower limit of that range and any other stated or
25 intervening value in that stated range is encompassed within the invention. The upper and lower limits of these smaller ranges may independently be included in the smaller ranges and are also encompassed within the invention, subject to any specifically excluded limit in the stated range. Where the stated range includes one or both of the limits, ranges excluding either or both of those included limits are also included in the
30 invention.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to

those described herein can also be used in the practice or testing of the present invention, the preferred methods and materials are now described.

All publications and patents cited in this disclosure are incorporated by reference in their entirety. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention. To the extent the material incorporated by reference contradicts or is inconsistent with this specification, the specification will supersede any such material.

It must be noted that as used herein and in the appended claims, the singular forms "a," "and" and "the" include plural references unless the context clearly dictates otherwise.

Unless otherwise indicated, the term "at least" preceding a series of elements is to be understood to refer to every element in the series. Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the present invention. Throughout this specification and the claims which follow, unless the context requires otherwise, the word "comprise", and variations such as "comprises" and "comprising", will be understood to imply the inclusion of a stated integer or step or group of integers or steps but not the exclusion of any other integer or step or group of integer or step. Thus the terms "comprising", "including," "containing", "having" etc. shall be read expansively or open-ended and without limitation. When used herein the term "comprising" can be substituted with the term "containing" or sometimes when used herein with the term "having".

As defined herein, a "rhamnolipid" refers to a glycolipid that has a lipid portion that includes one or more, typically linear, saturated or unsaturated β -hydroxy-carboxylic acid moieties and a saccharide portion of one or more units of rhamnose.

The saccharide portion and the lipid portion are linked via a β -glycosidic bond between the 1-OH group of a rhamnose moiety of the saccharide portion and the 3-OH group of a β -hydroxy-carboxylic acid of the lipid portion. Thus the carboxylic group of one carboxylic acid moiety defines the end of the rhamnolipid. Where more than one rhamnose-moiety is included in a rhamnolipid, each of the rhamnose moieties not linked to the lipid portion is linked to another rhamnose moiety via a 1,4 β -glycosidic bond. In embodiments where two or more β -hydroxy-carboxylic acids are present in a

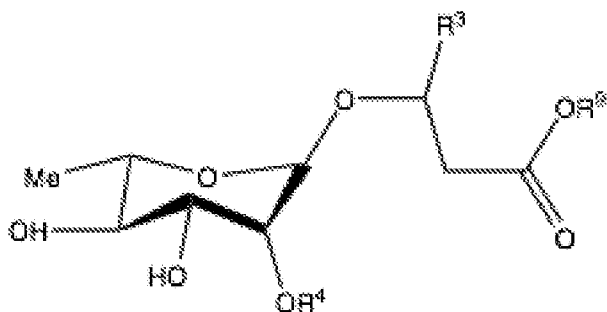
rhamnolipid, the β -hydroxy-carboxylic acid moieties are selected independently from each other. β -hydroxy carboxylic acid moieties of a respective plurality of β -hydroxy carboxylic acid moieties may in some embodiments be identical. In some embodiments they are different from each other.

5 As defined herein, the term “an aqueous medium” is a composition which comprises at least about 5% by weight of rhamnolipids or salts of rhamnolipids in water. In a particular embodiment, it comprises the product of a fermentation that has the ability to produce one or more rhamnolipids from at least one carbon source under fermentation conditions known in the art.

10 The terms “fermentation broth” and “fermentation medium” are synonymous and may be used interchangeably.

DETAILED DESCRIPTION

Provided herein is an organic solvent free-method for obtaining a composition comprising one or more rhamnolipids. In a particular embodiment, the rhamnolipid may have the structure (I)

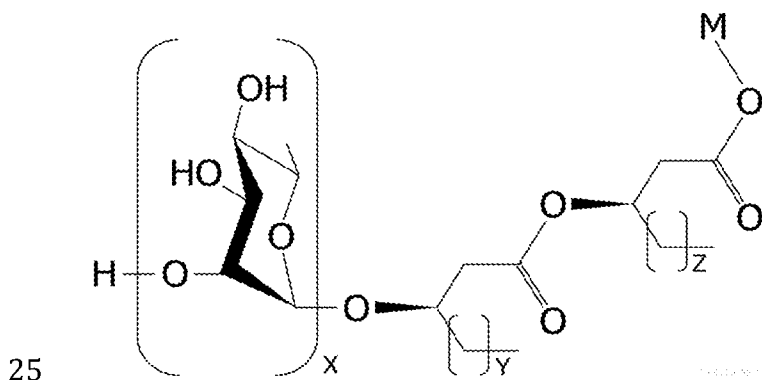


In this formula, R⁹ is a hydrogen atom (H) or an aliphatic group that has a main chain of one to about 46, such as one to about 42, one to about 40, one to about 38, one to about 36, one to about 34, one to about 30, one to about 28, including e.g. 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27 or 28 carbon atoms and one to about three, including two, oxygen atoms. In some embodiments, the main chain of the respective aliphatic group carries a terminal carboxylic acid group and/or an internal ester group. As an illustrative example in this regard, R⁹ may be of the formula —CH(R⁵)—CH₂—COOR⁶. In these illustrative moieties, R⁵ may be an aliphatic moiety with a main chain that has a length from 1 to about 19, such as from 1 to about 17, from 1 to about 15, from 1 to about 13, about 2 to about 13, about 3 to about 13 or about 4 to about 13, including e.g. 2, 3, 4, 5, 6, 7, 8, 9, 10, 11 or 12 carbon

atoms. R^4 in formula (I) is a hydrogen atom (H), or a rhamnopyranosyl moiety. R^6 is a hydrogen atom.

The term "aliphatic" means, unless otherwise stated, a straight or branched hydrocarbon chain, which may be saturated or mono- or poly-unsaturated and include
 5 heteroatoms. The term "heteroatom" as used herein means an atom of any element other than carbon or hydrogen. Herein, an unsaturated aliphatic group contains one or more double bonds (alkenyl moieties). The branches of the hydrocarbon chain may include linear chains as well as non-aromatic cyclic elements. The hydrocarbon chain, which may, unless otherwise stated, be of any length, and contain any number of
 10 branches. Typically, the hydrocarbon (main) chain includes 1 to about 5, to about 10, to about 15 or to about 20 carbon atoms. Examples of alkenyl moieties are straight-chain or branched hydrocarbon moieties which contain one or more double bonds. Alkenyl moieties generally contain about two to about twenty carbon atoms and one or more, for instance two, double bonds, such as about two to about ten carbon atoms,
 15 and one double bond. Examples of alkyl groups are methyl, ethyl, propyl, butyl, pentyl, hexyl, heptyl, octyl, nonyl, decyl, the n isomers of these radicals, isopropyl, isobutyl, isopentyl, sec-butyl, tert-butyl, neopentyl, 3,3-dimethylbutyl. Both the main chain as well as the branches may furthermore contain heteroatoms as for instance N, O, S, Se or Si or a carbon atom may be replaced by one of these heteroatoms. An aliphatic
 20 moiety may be substituted or unsubstituted with one or more functional groups. Substituents may be any functional group, as for example, but not limited to, amino, amido, carbonyl, carboxyl, hydroxyl, nitro, thio and sulfonyl.

In a more particular embodiment, the rhamnolipid(s) or rhamnolipid salts in said structure has the structure (II):



wherein x is 1 or 2, y is 4, 6 or 8 and M is H, or a metal, such as alkali metals Li, Na, or K, alkali earth metals Mg or Ca, or transition metals Mn, Fe, Cu, or Zn. In the cases of the alkali earth and transition metals, multiple rhamnolipid salt moieties may associate with each metal. In a specific embodiment, the composition comprises a mixture of mono (where x=1) and di (where x=2) rhamnolipids where y and z are 6 and M is H or Na. The mono-rhamnolipid may be referred to as Rha-C10-C10, with a formula of $C_{26}H_{48}O_9$. The IUPAC Name is 3-[3-[(2R,3R,4R,5R,6S)-3,4,5-trihydroxy-6-methyloxan-2-yl]oxydecanoyloxy]decanoic acid. The di-rhamnolipid may be referred to as RhaRha-C10-C10, with a formula of $C_{32}H_{58}O_{13}$. The IUPAC name is 3-[3-[4,5-dihydroxy-6-methyl-3-(3,4,5-tri hydroxy-6-methyloxan-2-yl)oxyoxan-2-yl]oxydecanoyloxy]decanoic acid. In a more specific embodiment, mono-rhamnolipid may be present in the amount of about from 30% of total rhamnolipid to about 50% of total and di-rhamnolipid may be present in the amount of about from 50% of total rhamnolipid to about 70% di-rhamnolipid. The composition in another embodiment comprises mono and di-rhamnolipids wherein the ratio of mono-rhamnolipids:di-rhamnolipids is from about 35:65 to about 55:45

In a most specific embodiment, the composition comprises the compounds set forth in Table 1.

Table 1

Fatty acid chains	Mono- (x = 1)	Di- (x = 2)	y and z
C10-C10	28-40%	42-60%	6
Other	0-12%	0-18%	y and/or z = 4 or 8 (and if 8, there may be a point of unsaturation)

20

As set forth above, the method comprises: (a) providing an aqueous medium comprising at least one rhamnolipid (RL); (b) non-mechanical sterilization of said medium provided in (a) and (c) separating a RL containing phase in the sterilized medium of (b) from waste to obtain said composition. Furthermore, in specific embodiments, aging, acidification, neutralization, drying, decolorization and/or deodorization steps may also be used.

25

Source of medium

The aqueous medium provided comprising one or more rhamnolipids may be obtainable from a fermentation broth derived from and in particular, may be a result of a recombinant host cell producing rhamnolipids using methods known in the art or
5 from a rhamnolipid producing microorganism. A recombinant host cell producing rhamnolipids may be a host cell, such as a bacterial cell that expresses a RhlA gene or ortholog thereof and/or a RhlB gene or ortholog thereof, and/or a RhlC gene or ortholog thereof, and/or RhlR gene or ortholog thereof, and/or RhlI gene or ortholog thereof, and/or RhlG gene or ortholog thereof and others.

10 The term "rhamnolipid-producing microorganism" as used herein, refers to any microorganism, such as bacteria, which has the capacity to synthesize/produce rhamnolipids under suitable conditions which includes but is not limited to bacterium of the phyla Actinobacteria, Firmicutes and Proteobacteria. In a particular embodiment, the rhamnolipid-producing microorganism is a bacterium of the
15 *Gammaproteobacteria* class. In a further embodiment, the rhamnolipid-producing microorganism is a bacterium of the *Pseudomonadales* order. In yet another further embodiment, the rhamnolipid producing microorganism is a bacterium of the Pseudomonadaceae family. In a further embodiment, the rhamnolipid-producing microorganism is a bacterium of the *Pseudomonas* genus, such as *P. alcaligenes*, *P. aeruginosa*, *P. chlororaphis*, *P. clemancea*, *P. collierea*, *P. fluorescens*, *P. luteola*, *P. putida*, *P. stutzeri* and *P. teessidea*. In a further embodiment, the rhamnolipid-producing microorganism is *P. aeruginosa*.

The host cell is cultured under conditions that allow RL production. A variety of carbon sources may be used such as a monosaccharide, *e.g.* glucose, a disaccharide, *e.g.*
25 sucrose, an alcohol, *e.g.* glycerol, an alkane, *e.g.*, n-hexadecane, a fatty acid such as caprylic acid (also termed octanoate), vegetable oils (fresh or waste; *e.g.*, soybean oil) or mixtures thereof, organic acids (*e.g.* lactic acid, acetic acid, citric acid, propionic acid), alcohols (*e.g.* ethanol, glycerin), and mixtures of these; nitrogen sources such as ammonium sulfate, ammonium phosphate, urea, yeast extract, meat extract, peptone,
30 and corn steep liquor; and other nutritional sources such as mineral salts and vitamins. The bacterial host cell will typically be exposed to a fermentation process. The bacterial host cell may for instance be in the logarithmic growth phase or in the stationary phase.

Sterilization

The aqueous (e.g., fermentation) medium provided is sterilized. In particular, the medium in the the methods set forth above may be sterilized by non filtration methods using methods known in the art. These methods may be either heat based, chemical based or ultraviolet light radiation based. In a particular embodiment, the heat based treatment may be via moist heat sterilization, particularly autoclaving.

Alternatively, the fermentation medium may be treated chemically. In one particular embodiment, the chemical treatment may be via acidification, removal of solid waste and neutralization using procedures set forth below.

In another particular embodiment, the aqueous (e.g. fermentation) medium may be treated with peroxides such as hydrogen peroxide, lithium peroxide, sodium peroxide, barium peroxide, peroxyacetic acid, benzoyl peroxide, in particular, hydrogen peroxide using methods known in the art and set forth below.

In yet another embodiment, the aqueous medium may be irradiated with, for example, ultraviolet light irradiation.

In one embodiment, the aqueous medium (e.g., fermentation medium or broth) may be sterilized by one of the above procedures. In another embodiment, the fermentation media may be sterilized by more than one of the procedures set forth above and these sterilizations could be in any order.

Separation/Removal of Waste Products (Solid/liquid (S/L) separation)

Waste from the compositions and/or medium obtained using the medium set forth herein may be removed or separated from the composition using separation procedures known in the art. These procedures include but are not limited to, settling batch or continuous centrifugation or ultracentrifugation, which may be followed by decantation or filtration using procedures known in the art. The waste can be either a solid phase or a liquid phase, depending on where in the process the separation occurs.

30 Aging

The culture or fermentation medium may be aged by incubating for at least about 1 day and between about 24-72 hrs. at between about 0-30C. This aging step may occur before and/or after one or more of the sterilization steps set forth above.

Acidification/Neutralization

Aqueous (e.g. Fermentation) medium may be concentrated or sterilized by treating with acid, removing liquid waste and then neutralizing the remaining solid and optionally oily phase or layer. In a particular embodiment, the aqueous medium
5 may be treated with acid so the culture medium is adjusted to a pH of between about 1.5 to 2.5, preferentially, about 2.05 to about 2.15. The acid can be an organic acid such as acetic acid, or a mineral acid. In a preferred embodiment, the acid is a mineral acid, e.g. HCl, H₂SO₄, HNO₃, or H₃ClO₄. As a result, a liquid, oily and solid phase is generated. The liquid phase is removed using procedures known in the art and in a
10 specific embodiment using methods set forth above (e.g., filtration, or centrifugation or settling combined with decanting).

The solid and optionally the oily phase are neutralized to a pH greater than about 5.5, preferably between about 6.5-7.5, more preferably between about 6.9-7.1. In a specific embodiment, the solid and optionally oily phase is treated with a solid,
15 dry mineral base (e.g., NaOH, KOH, LiOH, NaHCO₃). In the event that both the solid and oily phases are neutralized, these phases may be combined and neutralized or alternatively neutralized separately. After neutralization, solid waste is removed by methods set forth above, including but not limited to filtration, settling or centrifugation and decantation. The neutralized material may also be subjected to a
20 further sterilization step, as set forth above.

Decolorization/Deodorization

The aqueous medium (e.g., fermentation medium or broth) may be decolorized and/or deodorized using procedures know in the art. In a particular embodiment,
25 chemical and/or ultraviolet or visible radiation treatment is used.

In a specific embodiment, said decolorization and/or deodorization step may be accomplished via peroxide treatment using either hydrogen peroxide or organic (e.g, benzoyl peroxide or peroxyacetic acid) or inorganic peroxide (e.g., lithium peroxide, sodium peroxide, barium peroxide). In a particular embodiment, the
30 aqueous medium (e.g., fermentation medium or broth) may be treated with peroxide, particularly, hydrogen peroxide, at a concentration of about 1-5% v/v for about 48-120 hours and more particularly between about 3-4% v/v for about 72-96 hours.

Alternatively, the aqueous medium may be decolorized and/or deodorized via ultraviolet or visible radiation from 350 – 550 nm, preferentially 400-450 nm. The dose of radiation needs to be at least about 100 J/g of rhamnolipids, preferentially at least about 1 kJ/g, more preferentially at least about 10 kJ/g.

5 This aqueous medium may be decolorized and/or deodorized before and/or after sterilization and before or after the acidification and neutralization process described above.

Liquid Removal

10 In another embodiment, the liquid may be removed after removal of solid waste using methods known in the art, with the proviso that it does not follow decolorization and/or deodorization by peroxide treatment. These include but are not limited to heat, evaporation, freeze drying (lyophilization), drum drying.

15 DESCRIPTION OF SPECIFIC EMBODIMENTS

Example 1: A process to recover a rhamnolipid (RL) rich solution (clarified broth) from fermentation broth without the use of organic solvent

20 Provided herein is a process to recover rhamnolipids from fermentation broth in a dilute solution containing at least 50 g/L of a mixture of di- and mono-rhamnolipids without the use of organic solvents. The general scheme that is used is depicted in Figure 1.

The process utilizes aging at room temperature, followed by autoclaving, and standard solid/liquid separation techniques to separate clarified rhamnolipid rich
25 solution contained at least 50 g/L from bacterial cell matter. In particular, the rhamnolipid containing broth is generated from the fermentation by *Pseudomonas aeruginosa* at pH 6.2-6.5 at 37C for 48-72 hours with soybean oil as the feedstock. The final pH of fermentation broth prior to aging is in the range of 6.4-6.5. After aging for ~48 hours at room temperature, the fermentation broth is sterilized by steam heating
30 or autoclaving. After solid/liquid separation, the rhamnolipid solution is clear and ready to be used. The dark brown color of the rhamnolipid solution can be reduced with the addition of hydrogen peroxide at 2-3% v/v over 72-96 hrs. at room temperature. Other peroxides can also be used in this step, including organic (e.g,

benzoyl peroxide or peroxyacetic acid) or inorganic peroxide (e.g., lithium peroxide, sodium peroxide, barium peroxide). Rhamnolipid rich solution (>5% active) is obtained.

5 The clarified rhamnolipid rich solution, if not treated with peroxide, can also be lyophilized to produce a lightly colored powder. A rhamnolipid rich powder (>75% active) is obtained.

Example 2: Isolation of an aqueous solution of rhamnolipids by precipitation from clarified fermentation broth and redissolution in base and the composition that results from the process

10 The current process starts with the clarified broth obtained in Example 1 and is depicted schematically in Figure 2. As described in Example 1, clarified broth is made by allowing fermentation broth that ends at a pH of 6.0 to 6.5 to age under ambient conditions for about 2 days. The biomass settles to the bottom of the vessel used for this aging process and the clear supernatant, after removal, is clarified broth. The next step in the process is to add acid, such as concentrated sulfuric acid, until the pH is about 2.1. The rhamnolipids precipitate out of solution and form a solid phase and an oily liquid phase at the bottom of the vessel used for this step. The separation of the solid and oily liquid phases can be sped up by centrifugation. The solid and oily liquid phases are separated from the aqueous top phase or layer, which can be discarded or recycled. The combined solid and oily liquid rhamnolipid containing phases are then treated with a solid base, such as sodium hydroxide, until the pH reaches about 7.0. This produces a viscous solution of rhamnolipids in water with a concentration of about 25 to 30%. The final step is to centrifuge the 25-30% rhamnolipid solution and decant the liquid final product away from the pellet formed from any remaining solids. This liquid can be lyophilized (freeze dried) to produce a dry solid product.

Example 3: Non-organic solvent concentrated process to produce 35-40% rhamnolipid solution using a pH shift

30 Provided is a process for the production of 30-40% rhamnolipid solution using a pH shift starting with "reduced color and odor clarified broth" as described in Example 1 or "reduced color and odor sterilized fermentation broth" and are schematically depicted in Figures 3 and 4.

The “reduced color and odor clarified broth” and “reduced color and odor sterilized fermentation broth” can be obtained by adding 3-5 % v/v hydrogen peroxide to clarified broth or sterilized fermentation broth respectively. After 72-96 hours at room temperature, the color of the clarified rhamnolipid rich solution is slightly yellow with no loss in rhamnolipids and odor is reduced. After that, the reduced color and odor clarified broth is filtered at 1 micron by cross flow filtration or solids are removed by centrifugation followed by decanting. The concentrated acid such as sulfuric acid is added into the filtrate (permeate) or supernatant until the pH is about 2.1. The mixture is then allowed to separate into a rhamnolipid rich layer made up of a solid phase and an oily liquid phase at the bottom of the container. The combined solid and oily liquid rhamnolipid containing phases are then separated from the aqueous top layer and treated with a solid base, such as sodium hydroxide, until the pH reaches about 7.0. This produces a viscous solution of rhamnolipids in water.

The final step is to centrifuge the resulting rhamnolipid solution and remove the liquid final product from the pellet formed from any remaining solids. The final liquid product is 30-40% rhamnolipids.

Example 4: Process for isolating rhamnolipids from fermentation broth with sterilization at end of the process and materials produced during the process

The process, schematically depicted in Figure 5, begins by doing a solid/liquid separation process such as centrifugation or filtration of fermentation broth that still contains a living organism that produces rhamnolipids. The clarified liquid product is then acidified to pH 2.1 using concentrated sulfuric acid, or another concentrated, strong mineral acid. After settling or centrifugation, the water layer is decanted away from the denser product, which is an oily layer and a solid layer. This oily and/or solid layer is neutralized to pH 7 with a solid base such as sodium hydroxide, resulting in a solution. A final solid/liquid separation step is done to remove any remaining undissolved impurities. The liquid is then sterilized by, for example, autoclaving.

Example 5: Non-organic solvent concentrated process to produce 50-60% rhamnolipid solution using a pH shift

Provided is a modification of the process disclosed in Example 3, "Non-organic solvent concentrated process to produce 30-40% rhamnolipid solution using a pH shift" to produce 50-60% rhamnolipid solution.

5 A preparation of clarified broth and the acid precipitation of the clarified broth with concentrated sulfuric acid (95-98 wt%) to pH 2.1 remain the same as disclosed in Example 3. After adding 95-98 wt% sulfuric acid to the clarified broth until the pH is at 2.1, the mixture is then allowed to separate into a rhamnolipid rich layer made up of a solid phase and an oily liquid phase at the bottom of the container. However, to obtain 50-60% rhamnolipid solution, the oily material and the solid, are separated using, for 10 example centrifugation. After centrifugation, the solid is obtained at the bottom and the oily material is at the middle while the aqueous top layer is at the top of the vessel. The solids are then separated from oily liquid rhamnolipid containing phases and are treated with a solid base, such as sodium hydroxide until the pH reaches about 7.0 at which the product liquefies.

15 The final step is to centrifuge the resulting rhamnolipid solution and remove the liquid final product from the pellet formed from any remaining solids. The final liquid product is 50-60% rhamnolipids and is part of the invention. This liquid may be lyophilized (freeze dried) to produce a dry solid product, which is also part of the invention.

20

Example 6: Process for isolating rhamnolipid from fermentation broth before sterilization to obtain high rhamnolipid concentration without solvent extraction

25 This process is a modification of the process described in Example 5: "Non-organic solvent concentrated process to produce 50-60% rhamnolipid solution using a pH shift" to produce 50-60% rhamnolipid solution but at a higher yield since it includes the oily layer obtained after acid precipitation and thus, increasing the recovery of rhamnolipids.

30 The process is depicted in Figure 6. It begins with fermentation broth that contains rhamnolipids and biomass from cells that produced the rhamnolipids, preferentially *Pseudomonas aeruginosa*. First, centrifugation is performed at at least 7500 *g* for at least 5 min. A clarified broth (CB) is then separated from microbial cells (solid) and is sterilized via steam autoclave. The CB is then allowed to settle,

preferably overnight, prior to being centrifuged or filtered to remove any additional solids that precipitated out during settling.

5 This further clarified CB is then acidified to pH 2.1 using concentrated sulfuric acid, or another concentrated, strong mineral acid. The acidified clarified broth is part of the invention. After settling or centrifugation, the water layer is decanted away from a solid layer. The solid is then treated with a solid base, such as sodium hydroxide, until the pH reaches about 7.0. The result is a liquid with 50-60% rhamnolipids. This liquid can be lyophilized (freeze dried) to produce a dry solid product.

10 By centrifuging immediately after fermentation is stopped and then ageing the product as described in Example 1, a Clarified Broth (CB) is produced that can be further processed. This solves the problem that solid / liquid separation of fermentation broth can be difficult.

15 The overall process produces a product that is a high concentration of rhamnolipid biosurfactant cost effectively with a recovery rate of the rhamnolipids in the original fermentation broth of about >80%. This solves the problem that it is perceived that isolating products from rhamnolipid fermentations is too expensive to be commercially viable.

WHAT IS CLAIMED IS:

1. An organic solvent-free process for obtaining a composition comprising one or more rhamnolipids (RLs) comprising:

5 (a) providing an aqueous medium comprising at least one rhamnolipid;
(b) non-filtration based sterilization of said medium provided in (a) and
(c) separating a RL containing phase from waste in said sterilized medium in
(b) to obtain said composition and which optionally further comprises (1) after step
(a) and before step (b) removing solid waste from said medium provided in step (a)
10 and/or (2) after the sterilization step (b) and before the separation step (c) aging said
sterilized medium of step (a) and/or (3) after step (c), fluid is optionally removed
from the separated RL containing phase to obtain a solid composition and/or (4)
before and/or after said sterilization step, decolorizing and/or deodorizing medium
provided.

15 2. The method according to claim 1, wherein said method further comprises
after step (a) and before step (b) and/or after step (b) and before step (c), (1) aging
said medium provided in step (a) to obtain aged and sterilized medium and wherein
after step (a) and before said medium is aged in (1) solid waste is optionally removed
from medium provided in step (1).

20 3. The method according to claim 2, wherein said medium is aged for at least
about one day at between about 0-30C.

4. The method according to claims 1-3, wherein said sterilization is via heat
treatment, chemical treatment and/or irradiation.

25 5. The method according to claim 4, wherein said chemical treatment is via
acidification and/or peroxide treatment.

6. The method according to claims 1-5, wherein said method comprises before
step (b) aging said medium provided in step (a); (b) sterilizing said aged medium; (c)
separating a RL containing phase from waste in said sterilized medium and further
optionally comprises (c) (1) additionally sterilizing said RL containing phase and
30 (c)(2) separating a RL containing phase from waste in said sterilized RL containing
phase to obtain said composition.

7. The method according to claim 6, wherein said sterilization step in step (b) is via chemical treatment and/or irradiation and the sterilization step in step (c)(2) is via heat.

8. The method according to claims 1-6, wherein said composition comprising
5 rhamnolipids obtained is a clarified broth comprising at least about 5% rhamnolipids by weight.

9. The method according to claim 1, wherein said decolorization and/or deodorization step is via chemical and/or radiation treatment.

10. The method according to claim 1, wherein in said method, said method
10 further comprises (i) before sterilizing said medium provided, removing solid waste from said medium provided in step (a) and in said method, said medium provided in step (a) is sterilized in step (b) by treating said medium with acid and said acidic medium obtained comprises a solid, liquid and oily phase and at least the liquid phase is removed and before step (c), the solid and optionally oily phases are neutralized to
15 obtain a neutralized solution, wherein said solid and oily phases are neutralized separately to obtain separate solutions or wherein said solid and oily phases are combined and said combined solid and oily phases are neutralized to obtain a neutralized solution and in step (c) insoluble waste material is removed from said solution to obtain said composition comprising rhamnolipids wherein said
20 composition comprising rhamnolipids is a concentrated clarified broth comprising at least about 30% rhamnolipids by weight and wherein said method optionally further comprises an additional sterilization step before and/or after said medium is treated with acid in step (b) and wherein said method optionally further comprises (1) an additional separation step to remove insoluble waste from said solution before and/or
25 after said additional sterilization step and/or (2) after sterilization, decolorizing and/or deodorizing said sterilized concentrated clarified broth and/or (3) removing fluid from said solution obtained in step (c) and/or (4) removing water from said solution obtained in step (c).

11. The method according to claim 10, wherein said method further comprises
30 before and/or after said additional sterilization step aging said medium provided.

12. The method according to claims 1-11, wherein said composition comprises mono- and di-rhamnolipids wherein the ratio of mono-rhamnolipids:di-rhamnolipids is from about 35:65 to about 55:45.

13. A composition comprising a neutralized solution obtainable according to the method of claims 10-12.

14. A composition comprising a decolorized and/or deodorized medium obtainable according to the method of claims 1-12.

5 15. A composition comprising the aged medium obtainable according to the method of claim 1-12.

Figure 1



Figure 2



Figure 3

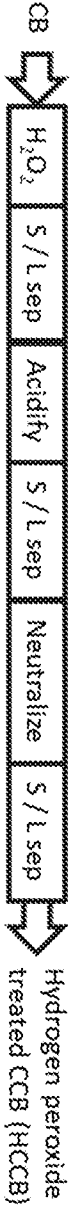


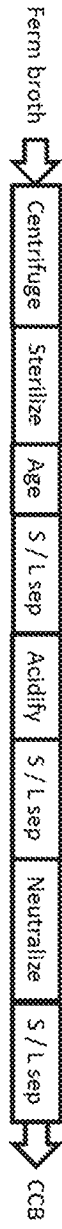
Figure 4



Figure 5



Figure 6



A. CLASSIFICATION OF SUBJECT MATTER

C12P 19/44(2006.01)i, C12P 7/64(2006.01)i, C07H 15/06(2006.01)i

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHEDMinimum documentation searched (classification system followed by classification symbols)
C12P 19/44; C07G 3/00; C07H 15/04; C12P 7/64; C12N 1/20; C07H 15/06Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
Korean utility models and applications for utility models
Japanese utility models and applications for utility modelsElectronic data base consulted during the international search (name of data base and, where practicable, search terms used)
eKOMPASS(KIPO internal) & Keywords: organic solvent free, rhamnolipid, sterilization, aging, acid, neutralize**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 6060287 A (ROCHA, CARLOS ALI et al.) 09 May 2000 See abstract; columns 10-11; claims 1, 17, 19-22.	1-3,9
A		10-11
A	US 2014-0148588 A1 (SCHILLING, MARTIN et al.) 29 May 2014 See abstract; claims 1-11.	1-3,9-11
A	US 7202063 B1 (GUNTHER, NEREUS W. et al.) 10 April 2007 See abstract; columns 5-6.	1-3,9-11
A	RIKALOVIC, MILENA G. et al., 'Production and characterization of rhamnolipids from Pseudomonas aeruginosa san-ai', Journal of the Serbian Chemical Society, 2012, Vol.77, No.1, pp.27-42 See the whole document.	1-3,9-11
A	WITEK-KROWIAK, ANNA et al., 'Ultrafiltrative separation of rhamnolipid from culture medium', World Journal of Microbiology and Biotechnology, 2011, Vol.27, No.8, pp.1961-1964 See the whole document.	1-3,9-11

 Further documents are listed in the continuation of Box C. See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

25 May 2016 (25.05.2016)

Date of mailing of the international search report

26 May 2016 (26.05.2016)

Name and mailing address of the ISA/KR

International Application Division

Korean Intellectual Property Office

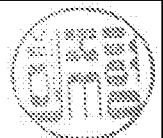
189 Cheongsa-ro, Seo-gu, Daejeon, 35208, Republic of Korea

Facsimile No. +82-42-481-8578

Authorized officer

HEO, Joo Hyung

Telephone No. +82-42-481-8150



Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. Claims Nos.: 5,7
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
Claims 5 and 7 are unclear since they each refer to a multiple dependent claim which does not comply with PCT Rule 6.4(a).

3. Claims Nos.: 4,6,8,12-15
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.

2. As all searchable claims could be searched without effort justifying an additional fees, this Authority did not invite payment of any additional fees.

3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.
- The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.
- No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No.

PCT/US2016/012901

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US 6060287 A	09/05/2000	US 5866376 A US 6060287 A	02/02/1999 09/05/2000
US 2014-0148588 A1	29/05/2014	CA 2835098 A1 CN 103833800 A EP 2735605 A1 JP 2014-111595 A	26/05/2014 04/06/2014 28/05/2014 19/06/2014
US 7202063 B1	10/04/2007	None	