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<p>(21) International Application Number: PCT/US95/07156 (22) International Filing Date: 5 June 1995 (05.06.95) (71) Applicant (for all designated States except US): HUMAN GENOME SCIENCES, INC. [US/US]; 9410 Key West Avenue, Rockville, MD 20850-3338 (US). (72) Inventors; and (75) Inventors/Applicants (for US only): GREENE, John, M. [US/US]; 872 Diamond Drive, Gaithersburg, MD 20878 (US). ROSEN, Craig, A. [US/US]; 22400 Rolling Hill Road, Laytonsville, MD 20882 (US). (74) Agents: SQUIRE, William; Carella, Byrne, Bain, Gilfillan, Cecchi, Stewart & Olstein, 6 Becker Farm Road, Roseland, NJ 07068 (US) et al.</p>	<p>(81) Designated States: AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, ES, FI, GB, GE, HU, JP, KE, KG, KP, KR, KZ, LK, LT, LU, LV, MD, MG, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SI, SK, TJ, TT, UA, US, UZ, VN, ARIPO patent (KE, MW, SD, SZ, UG), European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).</p> <p>Published <i>With international search report.</i></p>	
<p>(54) Title: FIBROBLAST GROWTH FACTOR 15</p> <p>(57) Abstract</p> <p>Disclosed is a human Fibroblast growth factor 15 polypeptide and DNA (RNA) encoding such polypeptide. Also provided is a procedure for producing such polypeptide by recombinant techniques. Also disclosed are methods for utilizing such polypeptide for stimulating revascularization, for treating wounds and prevent neuronal damage. Antagonists against such polypeptides and their use as a therapeutic to prevent abnormal cellular proliferation, hyper-vascular diseases and epithelial lens cell proliferation are also disclosed. Diagnostic methods for detecting mutations in the coding sequence and alterations in the concentration of the polypeptides in a sample derived from a host are also disclosed.</p>		

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FIBROBLAST GROWTH FACTOR 15

This invention relates to newly identified polynucleotides, polypeptides encoded by such polynucleotides, the use of such polynucleotides and polypeptides, as well as the production of such polynucleotides and polypeptides. More particularly, the polypeptide of the present invention have been putatively identified as fibroblast growth factor/heparin binding growth factor, hereinafter referred to as "FGF-15". The invention also relates to inhibiting the action of such polypeptides.

Fibroblast growth factors are a family of proteins characteristic of binding to heparin and are, therefore, also called heparin binding growth factors (HBGF). Expression of different members of these proteins are found in various tissues and are under particular temporal and spatial control. These proteins are potent mitogens for a variety of cells of mesodermal, ectodermal, and endodermal origin, including fibroblasts, corneal and vascular endothelial cells, granulocytes, adrenal cortical cells, chondrocytes, myoblasts, vascular smooth muscle cells, lens epithelial cells, melanocytes, keratinocytes, oligodendrocytes, astrocytes, osteoblasts, and hematopoietic cells.

Each member has functions overlapping with others and also has its unique spectrum of functions. In addition to

the ability to stimulate proliferation of vascular endothelial cells, both FGF-1 and 2 are chemotactic for endothelial cells and FGF-2 has been shown to enable endothelial cells to penetrate the basement membrane. Consistent with these properties, both FGF-1 and 2 have the capacity to stimulate angiogenesis. Another important feature of these growth factors is their ability to promote wound healing. Many other members of the FGF family share similar activities with FGF-1 and 2 such as promoting angiogenesis and wound healing. Several members of the FGF family have been shown to induce mesoderm formation and to modulate differentiation of neuronal cells, adipocytes and skeletal muscle cells.

Other than these biological activities in normal tissues, FGF proteins have been implicated in promoting tumorigenesis in carcinomas and sarcomas by promoting tumor vascularization and as transforming proteins when their expression is deregulated.

The FGF family presently consists of eight structurally-related polypeptides: basic FGF, acidic FGF, int 2, hst 1/k-FGF, FGF-5, FGF-6, keratinocyte growth factor, AIGF (FGF-8) and recently a glia-activating factor has been shown to be a novel heparin-binding growth factor which was purified from the culture supernatant of a human glioma cell line (Miyamoto, M. et al., Mol. and Cell. Biol., 13(7):4251-4259 (1993)). The genes for each have been cloned and sequenced. Two of the members, FGF-1 and FGF-2, have been characterized under many names, but most often as acidic and basic fibroblast growth factor, respectively. The normal gene products influence the general proliferation capacity of the majority of mesoderm and neuroectoderm-derived cells. They are capable of inducing angiogenesis *in vivo* and may play important roles in early development (Burgess, W.H. and Maciag, T., Annu. Rev. Biochem., 58:575-606, (1989)).

Many of the above-identified members of the FGF family also bind to the same receptors and elicit a second message through binding to these receptors.

A eukaryotic expression vector encoding a secreted form of FGF-1 has been introduced by gene transfer into porcine arteries. This model defines gene function in the arterial wall *in vivo*. FGF-1 expression induced intimal thickening in porcine arteries 21 days after gene transfer (Nabel, E.G., et al., *Nature*, 362:844-6 (1993)). It has further been demonstrated that basic fibroblast growth factor may regulate glioma growth and progression independent of its role in tumor angiogenesis and that basic fibroblast growth factor release or secretion may be required for these actions (Morrison, R.S., et al., *J. Neurosci. Res.*, 34:502-9 (1993)).

Fibroblast growth factors, such as basic FGF, have further been implicated in the growth of Kaposi's sarcoma cells *in vitro* (Huang, Y.Q., et al., *J. Clin. Invest.*, 91:1191-7 (1993)). Also, the cDNA sequence encoding human basic fibroblast growth factor has been cloned downstream of a transcription promoter recognized by the bacteriophage T7 RNA polymerase. Basic fibroblast growth factors so obtained have been shown to have biological activity indistinguishable from human placental fibroblast growth factor in mitogenicity, synthesis of plasminogen activator and angiogenesis assays (Squires, C.H., et al., *J. Biol. Chem.*, 263:16297-302 (1988)).

U.S. Patent No. 5,155,214 discloses substantially pure mammalian basic fibroblast growth factors and their production. The amino acid sequences of bovine and human basic fibroblast growth factor are disclosed, as well as the DNA sequence encoding the polypeptide of the bovine species.

Newly discovered FGF-9 has around 30% sequence similarity to other members of the FGF family. Two cysteine residues and other consensus sequences in family members were also well conserved in the FGF-9 sequence. FGF-9 was found

to have no typical signal sequence in its N terminus like those in acidic and basic FGF. However, FGF-9 was found to be secreted from cells after synthesis despite its lack of a typical signal sequence FGF (Miyamoto, M. et al., Mol. and Cell. Biol., 13(7):4251-4259 (1993). Further, FGF-9 was found to stimulate the cell growth of oligodendrocyte type 2 astrocyte progenitor cells, BALB/c3T3, and PC-12 cells but not that of human umbilical vein endothelial cells (Naruo, K., et al., J. Biol. Chem., 268:2857-2864 (1993)).

Basic FGF and acidic FGF are potent modulators of cell proliferation, cell motility, differentiation, and survival and act on cell types from ectoderm, mesoderm and endoderm. These two FGFs, along with KGF and AIGF, were identified by protein purification. However, the other four members were isolated as oncogenes., expression of which was restricted to embryogenesis and certain types of cancers. FGF-9 was demonstrated to be a mitogen against glial cells. Members of the FGF family are reported to have oncogenic potency. FGF-9 has shown transforming potency when transformed into BALB/c3T3 cells (Miyamoto, M., et al., Mol. Cell. Biol., 13(7):4251-4259 (1993)).

Androgen induced growth factor (AIGF), also known as FGF-8, was purified from a conditioned medium of mouse mammary carcinoma cells (SC-3) simulated with testosterone. AIGF is a distinctive FGF-like growth factor, having a putative signal peptide and sharing 30-40% homology with known members of the FGF family. Mammalian cells transformed with AIGF shows a remarkable stimulatory effect on the growth of SC-3 cells in the absence of androgen. Therefore, AIGF mediates androgen-induced growth of SC-3 cells, and perhaps other cells, since it is secreted by the tumor cells themselves.

The polypeptide of the present invention has been putatively identified as a member of the FGF family as a

result of amino acid sequence homology with other members of the FGF family.

In accordance with one aspect of the present invention, there are provided novel mature polypeptides as well as biologically active and diagnostically or therapeutically useful fragments, analogs and derivatives thereof. The polypeptides of the present invention are of human origin.

In accordance with another aspect of the present invention, there are provided isolated nucleic acid molecules encoding the polypeptides of the present invention, including mRNAs, DNAs, cDNAs, genomic DNA, as well as antisense analogs thereof and biologically active and diagnostically or therapeutically useful fragments thereof.

In accordance with still another aspect of the present invention, there are provided processes for producing such polypeptides by recombinant techniques through the use of recombinant vectors, such as cloning and expression plasmids useful as reagents in the recombinant production of the polypeptides of the present invention, as well as recombinant prokaryotic and/or eukaryotic host cells comprising a nucleic acid sequence encoding a polypeptide of the present invention.

In accordance with a further aspect of the present invention, there is provided a process for utilizing such polypeptides, or polynucleotides encoding such polypeptides, for screening for agonists and antagonists thereto and for therapeutic purposes, for example, promoting wound healing for example as a result of burns and ulcers, to prevent neuronal damage due to neuronal disorders and promote neuronal growth, and to prevent skin aging and hair loss, to stimulate angiogenesis, mesodermal induction in early embryos and limb regeneration.

In accordance with yet a further aspect of the present invention, there are provided antibodies against such polypeptides.

In accordance with yet another aspect of the present invention, there are provided antagonists against such polypeptides and processes for their use to inhibit the action of such polypeptides, for example, in the treatment of cellular transformation, for example, tumors, to reduce scarring and treat hyper-vascular diseases.

In accordance with another aspect of the present invention, there are provided nucleic acid probes comprising nucleic acid molecules of sufficient length to specifically hybridize to a polynucleotide encoding a polypeptide of the present invention

In accordance with yet another aspect of the present invention, there are provided diagnostic assays for detecting diseases or susceptibility to diseases related to mutations in a nucleic acid sequence of the present invention and for detecting over-expression of the polypeptides encoded by such sequences.

In accordance with another aspect of the present invention, there is provided a process for utilizing such polypeptides, or polynucleotides encoding such polypeptides, for *in vitro* purposes related to scientific research, synthesis of DNA and manufacture of DNA vectors.

These and other aspects of the present invention should be apparent to those skilled in the art from the teachings herein.

The following drawings are meant only as illustrations of specific embodiments of the present invention and are not meant as limitations in any manner.

Figure 1 depicts the cDNA sequence and corresponding deduced amino acid sequence of FGF-15.

Figure 2 illustrates the amino acid sequence homology between FGF-15 and the other FGF family members. Conserved amino acids are readily ascertainable.

In accordance with one aspect of the present invention, there are provided isolated nucleic acids molecules

(polynucleotides) which encode for the mature polypeptide having the deduced amino acid sequence of Figure 1 (SEQ ID NOS:2) or for the mature polypeptide encoded by the cDNA of the clone deposited as ATCC Deposit No. 97146 on May 12, 1995.

The polynucleotide encoding FGF-15 of this invention was discovered initially in a cDNA library derived from human adrenal tumor tissue. It is structurally related to all members of the fibroblast growth factor family and contains an open reading frame encoding a polypeptide of 252 amino acids. Among the top matches are: 1) 41 % identity and 66 % sequence similarity to human FGF-9 over a stretch of 129 amino acids; and 2) 37 % identity and 59 % similarity to human KGF over a region of 88 amino acids.

The FGF/HBGF family signature, GXLX(S,T,A,G)X6(D,E)CXFXE is conserved in the polypeptide of the present invention, (X means any amino acid residue; (D,E) means either D or E residue; X6 means any 6 amino acid residues).

The polynucleotide of the present invention may be in the form of RNA or in the form of DNA, which DNA includes cDNA, genomic DNA, and synthetic DNA. The DNA may be double-stranded or single-stranded. The coding sequence which encodes the mature polypeptide may be identical to the coding sequence shown in Figure 1 (SEQ ID NO:1) or that of the deposited clone or may be a different coding sequence, as a result of the redundancy or degeneracy of the genetic code, encodes the same, mature polypeptide as the DNA of Figure 1, (SEQ ID NO:1) or the deposited cDNA.

The polynucleotides which encodes for the mature polypeptide of Figure 1 (SEQ ID NO:2) or for the mature polypeptides encoded by the deposited cDNA(s) may include: only the coding sequence for the mature polypeptide; the coding sequence for the mature polypeptide and additional coding sequence such as a leader or secretory sequence or a proprotein sequence; the coding sequence for the mature

polypeptide (and optionally additional coding sequence) and non-coding sequence, such as introns or non-coding sequence 5' and/or 3' of the coding sequence for the mature polypeptide.

Thus, the term "polynucleotide encoding a polypeptide" encompasses a polynucleotide which includes only coding sequence for the polypeptide as well as a polynucleotide which includes additional coding and/or non-coding sequence.

The present invention further relates to variants of the hereinabove described polynucleotides which encode for fragments, analogs and derivatives of the polypeptides having the deduced amino acid sequence of Figure 1 (SEQ ID NO:2) or the polypeptides encoded by the cDNA(s) of the deposited clone(s). The variants of the polynucleotide may be a naturally occurring allelic variant of the polynucleotide or a non-naturally occurring variant of the polynucleotide.

Thus, the present invention includes polynucleotides encoding the same mature polypeptide as shown in Figure 1 (SEQ ID NO:2) or the same mature polypeptides encoded by the cDNA(s) of the deposited clone(s) as well as variants of such polynucleotides which variants encode for a fragment, derivative or analog of the polypeptide of Figure 1 (SEQ ID NO:2) or the polypeptides encoded by the cDNA(s) of the deposited clone(s). Such nucleotide variants include deletion variants, substitution variants and addition or insertion variants.

As hereinabove indicated, the polynucleotide may have a coding sequence which is a naturally occurring allelic variant of the coding sequence shown in Figure 1 (SEQ ID NO:1) or of the coding sequence of the deposited clone(s). As known in the art, an allelic variant is an alternate form of a polynucleotide sequence which may have a substitution, deletion or addition of one or more nucleotides, which does not substantially alter the function of the encoded polypeptides.

The present invention also includes polynucleotides, wherein the coding sequence for the mature polypeptides may be fused in the same reading frame to a polynucleotide sequence which aids in expression and secretion of a polypeptide from a host cell, for example, a leader sequence which functions as a secretory sequence for controlling transport of a polypeptide from the cell. The polypeptide having a leader sequence is a preprotein and may have the leader sequence cleaved by the host cell to form the mature form of the polypeptide. The polynucleotides may also encode for a proprotein which is the mature protein plus additional 5' amino acid residues. A mature protein having a prosequence is a proprotein and is an inactive form of the protein. Once the prosequence is cleaved an active mature protein remains.

Thus, for example, the polynucleotides of the present invention may encode for a mature protein, or for a protein having a prosequence or for a protein having both a prosequence and a presequence (leader sequence).

The polynucleotides of the present invention may also have the coding sequence fused in frame to a marker sequence which allows for purification of the polypeptide of the present invention. The marker sequence may be a hexahistidine tag supplied by a pQE-9 vector to provide for purification of the mature polypeptide fused to the marker in the case of a bacterial host, or, for example, the marker sequence may be a hemagglutinin (HA) tag when a mammalian host, e.g. COS-7 cells, is used. The HA tag corresponds to an epitope derived from the influenza hemagglutinin protein (Wilson, I., et al., Cell, 37:767 (1984)).

The term "gene" means the segment of DNA involved in producing a polypeptide chain; it includes regions preceding and following the coding region (leader and trailer) as well as intervening sequences (introns) between individual coding segments (exons).

Fragments of the full length FGF-15 gene may be used as a hybridization probe for a cDNA library to isolate the full length gene and to isolate other genes which have a high sequence similarity to the gene or similar biological activity. Probes of this type preferably have at least 30 bases and may contain, for example, 50 or more bases. The probe may also be used to identify a cDNA clone corresponding to a full length transcript and a genomic clone or clones that contain the complete FGF-15 gene including regulatory and promotor regions, exons, and introns. An example of a screen comprises isolating the coding region of the FGF-15 gene by using the known DNA sequence to synthesize an oligonucleotide probe. Labeled oligonucleotides having a sequence complementary to that of the gene of the present invention are used to screen a library of human cDNA, genomic DNA or mRNA to determine which members of the library the probe hybridizes to.

The present invention further relates to polynucleotides which hybridize to the hereinabove-described sequences if there is at least 70%, preferably at least 90%, and more preferably at least 95% identity between the sequences. The present invention particularly relates to polynucleotides which hybridize under stringent conditions to the hereinabove-described polynucleotides. As herein used, the term "stringent conditions" means hybridization will occur only if there is at least 95% and preferably at least 97% identity between the sequences. The polynucleotides which hybridize to the hereinabove described polynucleotides in a preferred embodiment encode polypeptides which either retain substantially the same biological function or activity as the mature polypeptide encoded by the cDNAs of Figure 1 (SEQ ID NO:1) or the deposited cDNA(s).

Alternatively, the polynucleotide may have at least 20 bases, preferably 30 bases, and more preferably at least 50 bases which hybridize to a polynucleotide of the present

invention and which has an identity thereto, as hereinabove described, and which may or may not retain activity. For example, such polynucleotides may be employed as probes for the polynucleotide of SEQ ID NO:1, for example, for recovery of the polynucleotide or as a diagnostic probe or as a PCR primer.

Thus, the present invention is directed to polynucleotides having at least a 70% identity, preferably at least 90% and more preferably at least a 95% identity to a polynucleotide which encodes the polypeptide of SEQ ID NO:2 as well as fragments thereof, which fragments have at least 30 bases and preferably at least 50 bases and to polypeptides encoded by such polynucleotides.

The deposit(s) referred to herein will be maintained under the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the purposes of Patent Procedure. These deposits are provided merely as a convenience and are not an admission that a deposit is required under 35 U.S.C. § 112. The sequence of the polynucleotides contained in the deposited materials, as well as the amino acid sequence of the polypeptides encoded thereby, are incorporated herein by reference and are controlling in the event of any conflict with the description of sequences herein. A license may be required to make, use or sell the deposited materials, and no such license is hereby granted.

The present invention further relates to an FGF polypeptide which has the deduced amino acid sequence of Figure 1 (SEQ ID NO:2) or which has the amino acid sequence encoded by the deposited cDNA(s), as well as fragments, analogs and derivatives of such polypeptides.

The terms "fragment," "derivative" and "analog" when referring to the polypeptide of Figure 1 (SEQ ID NO:2) or those encoded by the deposited cDNA(s), means polypeptides which retains essentially the same biological function or

activity as such polypeptides. Thus, an analog includes a proprotein which can be activated by cleavage of the proprotein portion to produce an active mature polypeptide.

The polypeptides of the present invention may be recombinant polypeptides, natural polypeptides or synthetic polypeptides, preferably recombinant polypeptides.

The fragment, derivative or analog of the polypeptide of Figure 1 (SEQ ID NO:2) or that encoded by the deposited cDNA(s) may be (i) one in which one or more of the amino acid residues are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code, or (ii) one in which one or more of the amino acid residues includes a substituent group, or (iii) one in which the mature polypeptide is fused with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol), or (iv) one in which the additional amino acids are fused to the mature polypeptide, such as a leader or secretory sequence or a sequence which is employed for purification of the mature polypeptide or a proprotein sequence. Such fragments, derivatives and analogs are deemed to be within the scope of those skilled in the art from the teachings herein.

The polypeptides and polynucleotides of the present invention are preferably provided in an isolated form, and preferably are purified to homogeneity.

The term "isolated" means that the material is removed from its original environment (e.g., the natural environment if it is naturally occurring). For example, a naturally-occurring polynucleotide or polypeptide present in a living animal is not isolated, but the same polynucleotide or DNA or polypeptide, separated from some or all of the coexisting materials in the natural system, is isolated. Such polynucleotide could be part of a vector and/or such

polynucleotide or polypeptide could be part of a composition, and still be isolated in that such vector or composition is not part of its natural environment.

The polypeptides of the present invention include the polypeptide of SEQ ID NO:2 (in particular the mature polypeptide) as well as polypeptides which have at least 70% similarity (preferably at least a 70% identity) to the polypeptide of SEQ ID NO:2 and more preferably at least a 90% similarity (more preferably at least a 90% identity) to the polypeptide of SEQ ID NO:2 and still more preferably at least a 95% similarity (still more preferably a 95% identity) to the polypeptide of SEQ ID NO:2 and also include portions of such polypeptides with such portion of the polypeptide generally containing at least 30 amino acids and more preferably at least 50 amino acids.

As known in the art "similarity" between two polypeptides is determined by comparing the amino acid sequence and its conserved amino acid substitutes of one polypeptide to the sequence of a second polypeptide.

Fragments or portions of the polypeptides of the present invention may be employed for producing the corresponding full-length polypeptide by peptide synthesis; therefore, the fragments may be employed as intermediates for producing the full-length polypeptides. Fragments or portions of the polynucleotides of the present invention may be used to synthesize full-length polynucleotides of the present invention.

The present invention also relates to vectors which include polynucleotides of the present invention, host cells which are genetically engineered with vectors of the invention and the production of polypeptides of the invention by recombinant techniques.

Host cells may be genetically engineered (transduced or transformed or transfected) with the vectors of this invention which may be, for example, a cloning vector or an

expression vector. The vector may be, for example, in the form of a plasmid, a viral particle, a phage, etc. The engineered host cells can be cultured in conventional nutrient media modified as appropriate for activating promoters, selecting transformants or amplifying the FGF genes. The culture conditions, such as temperature, pH and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

The polynucleotide of the present invention may be employed for producing a polypeptide by recombinant techniques. Thus, for example, the polynucleotide sequence may be included in any one of a variety of expression vehicles, in particular vectors or plasmids for expressing a polypeptide. Such vectors include chromosomal, nonchromosomal and synthetic DNA sequences, e.g., derivatives of SV40; bacterial plasmids; phage DNA; yeast plasmids; vectors derived from combinations of plasmids and phage DNA, viral DNA such as vaccinia, adenovirus, fowl pox virus, and pseudorabies. However, any other vector or plasmid may be used as long as they are replicable and viable in the host.

The appropriate DNA sequence may be inserted into the vector by a variety of procedures. In general, the DNA sequence is inserted into an appropriate restriction endonuclease sites by procedures known in the art. Such procedures and others are deemed to be within the scope of those skilled in the art.

The DNA sequence in the expression vector is operatively linked to an appropriate expression control sequence(s) (promoter) to direct mRNA synthesis. As representative examples of such promoters, there may be mentioned: LTR or SV40 promoter, the E. coli. lac or trp, the phage lambda P_L promoter and other promoters known to control expression of genes in prokaryotic or eukaryotic cells or their viruses.

The expression vector also contains a ribosome binding site for translation initiation and a transcription terminator. The vector may also include appropriate sequences for amplifying expression.

In addition, the expression vectors preferably contain a gene to provide a phenotypic trait for selection of transformed host cells such as dihydrofolate reductase or neomycin resistance for eukaryotic cell culture, or such as tetracycline or ampicillin resistance in E. coli.

The vector containing the appropriate DNA sequence as herein above described, as well as an appropriate promoter or control sequence, may be employed to transform an appropriate host to permit the host to express the protein. As representative examples of appropriate hosts, there may be mentioned: bacterial cells, such as E. coli, Salmonella typhimurium, Streptomyces; fungal cells, such as yeast; insect cells, such as Drosophila S2 and Spodoptera Sf9; animal cells such as CHO, COS or Bowes melanoma; adenoviruses; plant cells, etc. The selection of an appropriate host is deemed to be within the scope of those skilled in the art from the teachings herein.

More particularly, the present invention also includes recombinant constructs comprising one or more of the sequences as broadly described above. The constructs comprise a vector, such as a plasmid or viral vector, into which a sequence of the invention has been inserted, in a forward or reverse orientation. In a preferred aspect of this embodiment, the construct further comprises regulatory sequences, including, for example, a promoter, operably linked to the sequence. Large numbers of suitable vectors and promoters are known to those of skill in the art, and are commercially available. The following vectors are provided by way of example. Bacterial: pQE70, pQE60, pQE-9 (Qiagen), pBS, phagescript, psiX174, pBluescript SK, pBsKS, pNH8a, pNH16a, pNH18a, pNH46a (Stratagene); pTRC99A, pKK223-3,

pKK233-3, pDR540, pRIT5 (Pharmacia). Eukaryotic: pWLneo, pSV2cat, pOG44, pXT1, pSG (Stratagene) pSVK3, pBPV, pMSG, pSVL (Pharmacia). However, any other plasmid or vector may be used as long as they are replicable and viable in the host.

Promoter regions can be selected from any desired gene using CAT (chloramphenicol transferase) vectors or other vectors with selectable markers. Two appropriate vectors are pKK232-8 and pCM7. Particular named bacterial promoters include lacI, lacZ, T3, T7, gpt, lambda P_R, P_L and trp. Eukaryotic promoters include CMV immediate early, HSV thymidine kinase, early and late SV40, LTRs from retrovirus, and mouse metallothionein-I. Selection of the appropriate vector and promoter is well within the level of ordinary skill in the art.

In a further embodiment, the present invention relates to host cells containing the above-described construct. The host cell can be a higher eukaryotic cell, such as a mammalian cell, or a lower eukaryotic cell, such as a yeast cell, or the host cell can be a prokaryotic cell, such as a bacterial cell. Introduction of the construct into the host cell can be effected by calcium phosphate transfection, DEAE-Dextran mediated transfection, or electroporation (Davis, L., Dibner, M., Battey, I., Basic Methods in Molecular Biology, 1986)).

The constructs in host cells can be used in a conventional manner to produce the gene product encoded by the recombinant sequence. Alternatively, the polypeptides of the invention can be synthetically produced by conventional peptide synthesizers.

Mature proteins can be expressed in mammalian cells, yeast, bacteria, or other cells under the control of appropriate promoters. Cell-free translation systems can also be employed to produce such proteins using RNAs derived from the DNA constructs of the present invention.

Appropriate cloning and expression vectors for use with prokaryotic and eukaryotic hosts are described by Sambrook, et al., *Molecular Cloning: A Laboratory Manual*, Second Edition, (Cold Spring Harbor, N.Y., 1989), the disclosure of which is hereby incorporated by reference.

Transcription of a DNA encoding the polypeptides of the present invention by higher eukaryotes is increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp, that act on a promoter to increase its transcription. Examples include the SV40 enhancer on the late side of the replication origin (bp 100 to 270), a cytomegalovirus early promoter enhancer, a polyoma enhancer on the late side of the replication origin, and adenovirus enhancers.

Generally, recombinant expression vectors will include origins of replication and selectable markers permitting transformation of the host cell, e.g., the ampicillin resistance gene of *E. coli* and *S. cerevisiae* TRP1 gene, and a promoter derived from a highly-expressed gene to direct transcription of a downstream structural sequence. Such promoters can be derived from operons encoding glycolytic enzymes such as 3-phosphoglycerate kinase (PGK), α factor, acid phosphatase, or heat shock proteins, among others. The heterologous structural sequence is assembled in appropriate phase with translation, initiation and termination sequences, and preferably, a leader sequence capable of directing secretion of translated protein into the periplasmic space or extracellular medium. Optionally, the heterologous sequence can encode a fusion protein including an N-terminal identification peptide imparting desired characteristics, e.g., stabilization or simplified purification of expressed recombinant product.

Useful expression vectors for bacterial use are constructed by inserting a structural DNA sequence encoding a desired protein together with suitable translation,

initiation and termination signals in operable reading phase with a functional promoter. The vector will comprise one or more phenotypic selectable markers and an origin of replication to ensure maintenance of the vector and to, if desirable, provide amplification within the host. Suitable prokaryotic hosts for transformation include E. coli, Bacillus subtilis, Salmonella typhimurium and various species within the genera Pseudomonas, Streptomyces, and Staphylococcus, although others may also be employed as a matter of choice.

As a representative but nonlimiting example, useful expression vectors for bacterial use can comprise a selectable marker and bacterial origin of replication derived from commercially available plasmids comprising genetic elements of the well known cloning vector pBR322 (ATCC 37017). Such commercial vectors include, for example, pKK223-3 (Pharmacia Fine Chemicals, Uppsala, Sweden) and GEM1 (Promega Biotec, Madison, WI, USA). These pBR322 "backbone" sections are combined with an appropriate promoter and the structural sequence to be expressed.

Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, the selected promoter is derepressed by appropriate means (e.g., temperature shift or chemical induction) and cells are cultured for an additional period.

Cells are typically harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract retained for further purification.

Microbial cells employed in expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents.

Various mammalian cell culture systems can also be employed to express recombinant protein. Examples of mammalian expression systems include the COS-7 lines of

monkey kidney fibroblasts, described by Gluzman, Cell, 23:175 (1981), and other cell lines capable of expressing a compatible vector, for example, the C127, 3T3, CHO, HeLa and BHK cell lines. Mammalian expression vectors will comprise an origin of replication, a suitable promoter and enhancer, and also any necessary ribosome binding sites, polyadenylation site, splice donor and acceptor sites, transcriptional termination sequences, and 5' flanking nontranscribed sequences. DNA sequences derived from the SV40 viral genome, for example, SV40 origin, early promoter, enhancer, splice, and polyadenylation sites may be used to provide the required nontranscribed genetic elements.

The polypeptide of the present invention may be recovered and purified from recombinant cell cultures by methods used heretofore, including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxyapatite chromatography and lectin chromatography. Protein refolding steps can be used, as necessary, in completing configuration of the mature protein. Finally, high performance liquid chromatography (HPLC) can be employed for final purification steps.

The polypeptide of the present invention may be a naturally purified product, or a product of chemical synthetic procedures, or produced by recombinant techniques from a prokaryotic or eukaryotic host (for example, by bacterial, yeast, higher plant, insect and mammalian cells in culture). Depending upon the host employed in a recombinant production procedure, the polypeptides of the present invention may be glycosylated with mammalian or other eukaryotic carbohydrates or may be non-glycosylated. Polypeptides of the invention may also include an initial methionine amino acid residue.

The polypeptide of the present invention, as a result of the ability to stimulate vascular endothelial cell growth, may be employed in treatment for stimulating re-vascularization of ischemic tissues due to various disease conditions such as thrombosis, arteriosclerosis, and other cardiovascular conditions. These polypeptide may also be employed to stimulate angiogenesis and limb regeneration.

The polypeptide may also be employed for treating wounds due to injuries, burns, post-operative tissue repair, and ulcers since they are mitogenic to various cells of different origins, such as fibroblast cells and skeletal muscle cells, and therefore, facilitate the repair or replacement of damaged or diseased tissue.

The polypeptide of the present invention may also be employed stimulate neuronal growth and to treat and prevent neuronal damage which occurs in certain neuronal disorders or neuro-degenerative conditions such as Alzheimer's disease, Parkinson's disease, and AIDS-related complex. FGF-15 has the ability to stimulate chondrocyte growth, therefore, they may be employed to enhance bone and periodontal regeneration and aid in tissue transplants or bone grafts.

The polypeptide of the present invention may be also be employed to prevent skin aging due to sunburn by stimulating keratinocyte growth.

The FGF-15 polypeptide may also be employed for preventing hair loss, since FGF family members activate hair-forming cells and promotes melanocyte growth. Along the same lines, the polypeptides of the present invention may be employed to stimulate growth and differentiation of hematopoietic cells and bone marrow cells when used in combination with other cytokines.

The FGF-15 polypeptide may also be employed to maintain organs before transplantation or for supporting cell culture of primary tissues.

The polypeptide of the present invention may also be employed for inducing tissue of mesodermal origin to differentiate in early embryos.

In accordance with yet a further aspect of the present invention, there is provided a process for utilizing such polypeptides, or polynucleotides encoding such polypeptides, for *in vitro* purposes related to scientific research, synthesis of DNA, manufacture of DNA vectors and for the purpose of providing diagnostics and therapeutics for the treatment of human disease.

This invention provides a method for identification of the receptors for the polypeptides of the present invention. The genes encoding the receptor can be identified by numerous methods known to those of skill in the art, for example, ligand panning and FACS sorting (Coligan, et al., *Current Protocols in Immun.*, 1(2), Chapter 5, (1991)). Preferably, expression cloning is employed wherein polyadenylated RNA is prepared from a cell responsive to the polypeptides, for example, NIH3T3 cells which are known to contain multiple receptors for the FGF family proteins, and SC-3 cells, and a cDNA library created from this RNA is divided into pools and used to transfect COS cells or other cells that are not responsive to the polypeptides. Transfected cells which are grown on glass slides are exposed to the the polypeptide of the present invention, after they have been labelled. The polypeptides can be labeled by a variety of means including iodination or inclusion of a recognition site for a site-specific protein kinase.

Following fixation and incubation, the slides are subjected to auto-radiographic analysis. Positive pools are identified and sub-pools are prepared and re-transfected using an iterative sub-pooling and re-screening process, eventually yielding a single clones that encodes the putative receptor.

As an alternative approach for receptor identification, the labeled polypeptides can be photoaffinity linked with cell membrane or extract preparations that express the receptor molecule. Cross-linked material is resolved by PAGE analysis and exposed to X-ray film. The labeled complex containing the receptors of the polypeptides can be excised, resolved into peptide fragments, and subjected to protein microsequencing. The amino acid sequence obtained from microsequencing would be used to design a set of degenerate oligonucleotide probes to screen a cDNA library to identify the genes encoding the putative receptors.

This invention provides a method of screening compounds to identify those which modulate the action of the polypeptide of the present invention. An example of such an assay comprises combining a mammalian fibroblast cell, a the polypeptide of the present invention, the compound to be screened and ³[H] thymidine under cell culture conditions where the fibroblast cell would normally proliferate. A control assay may be performed in the absence of the compound to be screened and compared to the amount of fibroblast proliferation in the presence of the compound to determine if the compound stimulates proliferation by determining the uptake of ³[H] thymidine in each case. The amount of fibroblast cell proliferation is measured by liquid scintillation chromatography which measures the incorporation of ³[H] thymidine. Both agonist and antagonist compounds may be identified by this procedure.

In another method, a mammalian cell or membrane preparation expressing a receptor for a polypeptide of the present invention is incubated with a labeled polypeptide of the present invention in the presence of the compound. The ability of the compound to enhance or block this interaction could then be measured. Alternatively, the response of a known second messenger system following interaction of a compound to be screened and the FGF-15 receptor is measured

and the ability of the compound to bind to the receptor and elicit a second messenger response is measured to determine if the compound is a potential agonist or antagonist. Such second messenger systems include but are not limited to, cAMP guanylate cyclase, ion channels or phosphoinositide hydrolysis.

Examples of antagonist compounds include antibodies, or in some cases, oligonucleotides, which bind to the receptor for the polypeptide of the present invention but elicit no second messenger response or bind to the FGF-15 polypeptide itself. Alternatively, a potential antagonist may be a mutant form of the polypeptide which binds to the receptors, however, no second messenger response is elicited and, therefore, the action of the polypeptide is effectively blocked.

Another antagonist compound to the FGF-15 gene and gene product is an antisense construct prepared using antisense technology. Antisense technology can be used to control gene expression through triple-helix formation or antisense DNA or RNA, both of which methods are based on binding of a polynucleotide to DNA or RNA. For example, the 5' coding portion of the polynucleotide sequence, which encodes for the mature polypeptides of the present invention, is used to design an antisense RNA oligonucleotide of from about 10 to 40 base pairs in length. A DNA oligonucleotide is designed to be complementary to a region of the gene involved in transcription (triple helix -see Lee et al., Nucl. Acids Res., 6:3073 (1979); Cooney et al, Science, 241:456 (1988); and Dervan et al., Science, 251: 1360 (1991)), thereby preventing transcription and the production of the polypeptides of the present invention. The antisense RNA oligonucleotide hybridizes to the mRNA *in vivo* and blocks translation of the mRNA molecule into the polypeptide (Antisense - Okano, J. Neurochem., 56:560 (1991); Oligodeoxynucleotides as Antisense Inhibitors of Gene

Expression, CRC Press, Boca Raton, FL (1988)). The oligonucleotides described above can also be delivered to cells such that the antisense RNA or DNA may be expressed *in vivo* to inhibit production of the polypeptide.

Potential antagonist compounds also include small molecules which bind to and occupy the binding site of the receptors thereby making the receptor inaccessible to its polypeptide such that normal biological activity is prevented. Examples of small molecules include, but are not limited to, small peptides or peptide-like molecules.

Antagonist compounds may be employed to inhibit the cell growth and proliferation effects of the polypeptides of the present invention on neoplastic cells and tissues, i.e. stimulation of angiogenesis of tumors, and, therefore, retard or prevent abnormal cellular growth and proliferation, for example, in tumor formation or growth.

The antagonists may also be employed to prevent hyper-vascular diseases, and prevent the proliferation of epithelial lens cells after extracapsular cataract surgery. Prevention of the mitogenic activity of the polypeptides of the present invention may also be desirable in cases such as restenosis after balloon angioplasty.

The antagonists may also be employed to prevent the growth of scar tissue during wound healing.

The antagonists may be employed in a composition with a pharmaceutically acceptable carrier, e.g., as hereinafter described.

The polypeptides, agonists and antagonists of the present invention may be employed in combination with a suitable pharmaceutical carrier to comprise a pharmaceutical composition for parenteral administration. Such compositions comprise a therapeutically effective amount of the polypeptide, agonist or antagonist and a pharmaceutically acceptable carrier or excipient. Such a carrier includes but is not limited to saline, buffered saline, dextrose, water,

glycerol, ethanol, and combinations thereof. The formulation should suit the mode of administration.

The invention also provides a pharmaceutical pack or kit comprising one or more containers filled with one or more of the ingredients of the pharmaceutical compositions of the invention. Associated with such container(s) can be a notice in the form prescribed by a governmental agency regulating the manufacture, use or sale of pharmaceuticals or biological products, which notice reflects approval by the agency of manufacture, use or sale for human administration. In addition, the polypeptides, agonists and antagonists of the present invention may be employed in conjunction with other therapeutic compounds.

The pharmaceutical compositions may be administered in a convenient manner such as by the oral, topical, intravenous, intraperitoneal, intramuscular, subcutaneous, intranasal or intradermal routes. The pharmaceutical compositions are administered in an amount which is effective for treating and/or prophylaxis of the specific indication. In general, they are administered in an amount of at least about 10 $\mu\text{g}/\text{kg}$ body weight and in most cases they will be administered in an amount not in excess of about 8 mg/Kg body weight per day. In most cases, the dosage is from about 10 $\mu\text{g}/\text{kg}$ to about 1 mg/kg body weight daily, taking into account the routes of administration, symptoms, etc. In the specific case of topical administration, dosages are preferably administered from about 0.1 μg to 9 mg per cm^2 .

The polypeptide of the invention and agonist and antagonist compounds which are polypeptides, may also be employed in accordance with the present invention by expression of such polypeptide *in vivo*, which is often referred to as "gene therapy."

Thus, for example, cells may be engineered with a polynucleotide (DNA or RNA) encoding for the polypeptide *ex vivo*, the engineered cells are then provided to a patient to

be treated with the polypeptide. Such methods are well-known in the art. For example, cells may be engineered by procedures known in the art by use of a retroviral particle containing RNA encoding for the polypeptide of the present invention.

Similarly, cells may be engineered *in vivo* for expression of the polypeptide *in vivo*, for example, by procedures known in the art. As known in the art, a producer cell for producing a retroviral particle containing RNA encoding the polypeptide of the present invention may be administered to a patient for engineering cells *in vivo* and expression of the polypeptide *in vivo*. These and other methods for administering a polypeptide of the present invention by such methods should be apparent to those skilled in the art from the teachings of the present invention. For example, the expression vehicle for engineering cells may be other than a retroviral particle, for example, an adenovirus, which may be used to engineer cells *in vivo* after combination with a suitable delivery vehicle.

Retroviruses from which the retroviral plasmid vectors hereinabove mentioned may be derived include, but are not limited to, Moloney Murine Leukemia Virus, spleen necrosis virus, retroviruses such as Rous Sarcoma Virus, Harvey Sarcoma Virus, avian leukosis virus, gibbon ape leukemia virus, human immunodeficiency virus, adenovirus, Myeloproliferative Sarcoma Virus, and mammary tumor virus. In one embodiment, the retroviral plasmid vector is derived from Moloney Murine Leukemia Virus.

The vector includes one or more promoters. Suitable promoters which may be employed include, but are not limited to, the retroviral LTR; the SV40 promoter; and the human cytomegalovirus (CMV) promoter described in Miller, et al., Biotechniques, Vol. 7, No. 9, 980-990 (1989), or any other promoter (e.g., cellular promoters such as eukaryotic cellular promoters including, but not limited to, the

histone, pol III, and β -actin promoters). Other viral promoters which may be employed include, but are not limited to, adenovirus promoters, thymidine kinase (TK) promoters, and B19 parvovirus promoters. The selection of a suitable promoter will be apparent to those skilled in the art from the teachings contained herein.

The nucleic acid sequence encoding the polypeptide of the present invention is under the control of a suitable promoter. Suitable promoters which may be employed include, but are not limited to, adenoviral promoters, such as the adenoviral major late promoter; or heterologous promoters, such as the cytomegalovirus (CMV) promoter; the respiratory syncytial virus (RSV) promoter; inducible promoters, such as the MMT promoter, the metallothionein promoter; heat shock promoters; the albumin promoter; the ApoAI promoter; human globin promoters; viral thymidine kinase promoters, such as the Herpes Simplex thymidine kinase promoter; retroviral LTRs (including the modified retroviral LTRs hereinabove described); the β -actin promoter; and human growth hormone promoters. The promoter also may be the native promoter which controls the gene encoding the polypeptide.

The retroviral plasmid vector is employed to transduce packaging cell lines to form producer cell lines. Examples of packaging cells which may be transfected include, but are not limited to, the PE501, PA317, ψ -2, ψ -AM, PA12, T19-14X, VT-19-17-H2, ψ CRE, ψ CRIP, GP+E-86, GP+envAm12, and DAN cell lines as described in Miller, Human Gene Therapy, Vol. 1, pgs. 5-14 (1990), which is incorporated herein by reference in its entirety. The vector may transduce the packaging cells through any means known in the art. Such means include, but are not limited to, electroporation, the use of liposomes, and CaPO_4 precipitation. In one alternative, the retroviral plasmid vector may be encapsulated into a liposome, or coupled to a lipid, and then administered to a host.

The producer cell line generates infectious retroviral vector particles which include the nucleic acid sequence(s) encoding the polypeptides. Such retroviral vector particles then may be employed, to transduce eukaryotic cells, either *in vitro* or *in vivo*. The transduced eukaryotic cells will express the nucleic acid sequence(s) encoding the polypeptide. Eukaryotic cells which may be transduced include, but are not limited to, embryonic stem cells, embryonic carcinoma cells, as well as hematopoietic stem cells, hepatocytes, fibroblasts, myoblasts, keratinocytes, endothelial cells, and bronchial epithelial cells.

This invention is also related to the use of the genes of the present invention as part of a diagnostic assay for detecting diseases or susceptibility to diseases related to the presence of mutations in the nucleic acid sequences encoding the polypeptide of the present invention.

Individuals carrying mutations in a gene of the present invention may be detected at the DNA level by a variety of techniques. Nucleic acids for diagnosis may be obtained from a patient's cells, such as from blood, urine, saliva, tissue biopsy and autopsy material. The genomic DNA may be used directly for detection or may be amplified enzymatically by using PCR (Saiki et al., *Nature*, 324:163-166 (1986)) prior to analysis. RNA or cDNA may also be used for the same purpose. As an example, PCR primers complementary to the nucleic acid encoding a polypeptide of the present invention can be used to identify and analyze mutations. For example, deletions and insertions can be detected by a change in size of the amplified product in comparison to the normal genotype. Point mutations can be identified by hybridizing amplified DNA to radiolabeled RNA or alternatively, radiolabeled antisense DNA sequences. Perfectly matched sequences can be distinguished from mismatched duplexes by RNase A digestion or by differences in melting temperatures.

Genetic testing based on DNA sequence differences may be achieved by detection of alteration in electrophoretic mobility of DNA fragments in gels with or without denaturing agents. Small sequence deletions and insertions can be visualized by high resolution gel electrophoresis. DNA fragments of different sequences may be distinguished on denaturing formamide gradient gels in which the mobilities of different DNA fragments are retarded in the gel at different positions according to their specific melting or partial melting temperatures (see, e.g., Myers et al., Science, 230:1242 (1985)).

Sequence changes at specific locations may also be revealed by nuclease protection assays, such as RNase and S1 protection or the chemical cleavage method (e.g., Cotton et al., PNAS, USA, 85:4397-4401 (1985)).

Thus, the detection of a specific DNA sequence may be achieved by methods such as hybridization, RNase protection, chemical cleavage, direct DNA sequencing or the use of restriction enzymes, (e.g., Restriction Fragment Length Polymorphisms (RFLP)) and Southern blotting of genomic DNA.

In addition to more conventional gel-electrophoresis and DNA sequencing, mutations can also be detected by *in situ* analysis.

The present invention also relates to a diagnostic assay for detecting altered levels of FGF-15 proteins in various tissues since an over-expression of the proteins compared to normal control tissue samples may detect the presence of abnormal cellular proliferation, for example, a tumor. Assays used to detect levels of protein in a sample derived from a host are well-known to those of skill in the art and include radioimmunoassays, competitive-binding assays, Western Blot analysis, ELISA assays and "sandwich" assay. An ELISA assay (Coligan, et al., Current Protocols in Immunology, 1(2), Chapter 6, (1991)) initially comprises preparing an antibody specific to an antigen to the

polypeptides of the present invention, preferably a monoclonal antibody. In addition a reporter antibody is prepared against the monoclonal antibody. To the reporter antibody is attached a detectable reagent such as radioactivity, fluorescence or, in this example, a horseradish peroxidase enzyme. A sample is removed from a host and incubated on a solid support, e.g. a polystyrene dish, that binds the proteins in the sample. Any free protein binding sites on the dish are then covered by incubating with a non-specific protein like bovine serum albumen. Next, the monoclonal antibody is incubated in the dish during which time the monoclonal antibodies attach to any polypeptides of the present invention attached to the polystyrene dish. All unbound monoclonal antibody is washed out with buffer. The reporter antibody linked to horseradish peroxidase is now placed in the dish resulting in binding of the reporter antibody to any monoclonal antibody bound to the protein of interest.

Unattached reporter antibody is then washed out. Peroxidase substrates are then added to the dish and the amount of color developed in a given time period is a measurement of the amount of a polypeptide of the present invention present in a given volume of patient sample when compared against a standard curve.

A competition assay may be employed wherein antibodies specific to a polypeptide of the present invention are attached to a solid support and labeled FGF-13 and a sample derived from the host are passed over the solid support and the amount of label detected, for example by liquid scintillation chromatography, can be correlated to a quantity of a polypeptide of the present invention in the sample.

A "sandwich" assay is similar to an ELISA assay. In a "sandwich" assay a polypeptide of the present invention is passed over a solid support and binds to antibody attached to a solid support. A second antibody is then bound to the

polypeptide of interest. A third antibody which is labeled and specific to the second antibody is then passed over the solid support and binds to the second antibody and an amount can then be quantified.

The sequences of the present invention are also valuable for chromosome identification. The sequence is specifically targeted to and can hybridize with a particular location on an individual human chromosome. Moreover, there is a current need for identifying particular sites on the chromosome. Few chromosome marking reagents based on actual sequence data (repeat polymorphism's) are presently available for marking chromosomal location. The mapping of DNAs to chromosomes according to the present invention is an important first step in correlating those sequences with genes associated with disease.

Briefly, sequences can be mapped to chromosomes by preparing PCR primers (preferably 15-25 bp) from the cDNA. Computer analysis of the 3' untranslated region is used to rapidly select primers that do not span more than one exon in the genomic DNA, thus complicating the amplification process. These primers are then used for PCR screening of somatic cell hybrids containing individual human chromosomes. Only those hybrids containing the human gene corresponding to the primer will yield an amplified fragment.

PCR mapping of somatic cell hybrids is a rapid procedure for assigning a particular DNA to a particular chromosome. Using the present invention with the same oligonucleotide primers, sublocalization can be achieved with panels of fragments from specific chromosomes or pools of large genomic clones in an analogous manner. Other mapping strategies that can similarly be used to map to its chromosome include *in situ* hybridization, prescreening with labeled flow-sorted chromosomes and preselection by hybridization to construct chromosome specific-cDNA libraries.

Fluorescence *in situ* hybridization (FISH) of a cDNA clone to a metaphase chromosomal spread can be used to provide a precise chromosomal location in one step. This technique can be used with cDNA as short as 50 or 60 bases. For a review of this technique, see Verma et al., Human Chromosomes: a Manual of Basic Techniques, Pergamon Press, New York (1988).

Once a sequence has been mapped to a precise chromosomal location, the physical position of the sequence on the chromosome can be correlated with genetic map data. (Such data are found, for example, in V. McKusick, Mendelian Inheritance in Man (available on line through Johns Hopkins University Welch Medical Library). The relationship between genes and diseases that have been mapped to the same chromosomal region are then identified through linkage analysis (coinheritance of physically adjacent genes).

Next, it is necessary to determine the differences in the cDNA or genomic sequence between affected and unaffected individuals. If a mutation is observed in some or all of the affected individuals but not in any normal individuals, then the mutation is likely to be the causative agent of the disease.

With current resolution of physical mapping and genetic mapping techniques, a cDNA precisely localized to a chromosomal region associated with the disease could be one of between 50 and 500 potential causative genes. (This assumes 1 megabase mapping resolution and one gene per 20 kb).

The polypeptides, their fragments or other derivatives, or analogs thereof, or cells expressing them can be used as an immunogen to produce antibodies thereto. These antibodies can be, for example, polyclonal or monoclonal antibodies. The present invention also includes chimeric, single chain, and humanized antibodies, as well as Fab fragments, or the product of an Fab expression library. Various procedures

known in the art may be used for the production of such antibodies and fragments.

Antibodies generated against the polypeptides corresponding to a sequence of the present invention can be obtained by direct injection of the polypeptides into an animal or by administering the polypeptides to an animal, preferably a nonhuman. The antibody so obtained will then bind the polypeptides itself. In this manner, even a sequence encoding only a fragment of the polypeptides can be used to generate antibodies binding the whole native polypeptides. Such antibodies can then be used to isolate the polypeptide from tissue expressing that polypeptide.

For preparation of monoclonal antibodies, any technique which provides antibodies produced by continuous cell line cultures can be used. Examples include the hybridoma technique (Kohler and Milstein, 1975, *Nature*, 256:495-497), the trioma technique, the human B-cell hybridoma technique (Kozbor et al., 1983, *Immunology Today* 4:72), and the EBV-hybridoma technique to produce human monoclonal antibodies (Cole, et al., 1985, in *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, Inc., pp. 77-96).

Techniques described for the production of single chain antibodies (U.S. Patent 4,946,778) can be adapted to produce single chain antibodies to immunogenic polypeptide products of this invention. Also, transgenic mice may be used to express humanized antibodies to immunogenic polypeptide products of this invention.

The present invention will be further described with reference to the following examples; however, it is to be understood that the present invention is not limited to such examples. All parts or amounts, unless otherwise specified, are by weight.

In order to facilitate understanding of the following examples, certain frequently occurring methods and/or terms will be described.

"Plasmids" are designated by a lower case p preceded and/or followed by capital letters and/or numbers. The starting plasmids herein are either commercially available, publicly available on an unrestricted basis, or can be constructed from available plasmids in accord with published procedures. In addition, equivalent plasmids to those described are known in the art and will be apparent to the ordinarily skilled artisan.

"Digestion" of DNA refers to catalytic cleavage of the DNA with a restriction enzyme that acts only at certain sequences in the DNA. The various restriction enzymes used herein are commercially available and their reaction conditions, cofactors and other requirements were used as would be known to the ordinarily skilled artisan. For analytical purposes, typically 1 μ g of plasmid or DNA fragment is used with about 2 units of enzyme in about 20 μ l of buffer solution. For the purpose of isolating DNA fragments for plasmid construction, typically 5 to 50 μ g of DNA are digested with 20 to 250 units of enzyme in a larger volume. Appropriate buffers and substrate amounts for particular restriction enzymes are specified by the manufacturer. Incubation times of about 1 hour at 37°C are ordinarily used, but may vary in accordance with the supplier's instructions. After digestion the reaction is electrophoresed directly on a polyacrylamide gel to isolate the desired fragment.

Size separation of the cleaved fragments is performed using 8 percent polyacrylamide gel described by Goeddel, D. et al., *Nucleic Acids Res.*, 8:4057 (1980).

"Oligonucleotides" refers to either a single stranded polydeoxynucleotide or two complementary polydeoxynucleotide strands which may be chemically synthesized. Such synthetic oligonucleotides have no 5' phosphate and thus will not ligate to another oligonucleotide without adding a phosphate with an ATP in the presence of a kinase. A synthetic

oligonucleotide will ligate to a fragment that has not been dephosphorylated.

"Ligation" refers to the process of forming phosphodiester bonds between two double stranded nucleic acid fragments (Maniatis, T., et al., Id., p. 146). Unless otherwise provided, ligation may be accomplished using known buffers and conditions with 10 units of T4 DNA ligase ("ligase") per 0.5 μ g of approximately equimolar amounts of the DNA fragments to be ligated.

Unless otherwise stated, transformation was performed as described by the method of Graham, F. and Van der Eb, A., Virology, 52:456-457 (1973).

Example 1

Bacterial Expression and Purification of FGF-15 protein

The DNA sequence encoding FGF-15 ATCC # 97146, is initially amplified using PCR oligonucleotide primers corresponding to the 5' sequences of the processed protein (minus the signal peptide sequence) and the vector sequences 3' to the gene. Additional nucleotides corresponding to the gene are added to the 5' and 3' sequences. The 5' oligonucleotide primer has the sequence 5' GCCAGACCATGGTAAAACCGTGCCCTC 3' (SEQ ID NO:3) and contains an NcoI restriction enzyme site (in bold). The 3' sequence 5' GGCAGGAGATCTTGTTGTCCTACTCTTGTTGAC 3' (SEQ ID NO:4) contains complementary sequences to a BglII site (in bold) and is followed by 21 nucleotides of FGF-15 coding sequence.

The restriction enzyme sites correspond to the restriction enzyme sites on the bacterial expression vector pQE60 (Qiagen, Inc. Chatsworth, CA 91311). pQE-60 encodes antibiotic resistance (Amp^R), a bacterial origin of replication (ori), an IPTG-regulatable promoter operator (P/O), a ribosome binding site (RBS), a 6-His tag and restriction enzyme sites. pQE-60 was then digested with NcoI and BglII. The amplified sequences are ligated into pQE-60

and are inserted in frame with the sequence encoding for the histidine tag and the ribosome binding site (RBS). The ligation mixture is then used to transform E. coli strain M15/rep 4 (Qiagen, Inc.) by the procedure described in Sambrook, J. et al., Molecular Cloning: A Laboratory Manual, Cold Spring Laboratory Press, (1989). M15/rep4 contains multiple copies of the plasmid pREP4, which expresses the lacI repressor and also confers kanamycin resistance (Kan^r). Transformants are identified by their ability to grow on LB plates and ampicillin/kanamycin resistant colonies were selected. Plasmid DNA is isolated and confirmed by restriction analysis. Clones containing the desired constructs are grown overnight (O/N) in liquid culture in LB media supplemented with both Amp (100 ug/ml) and Kan (25 ug/ml). The O/N culture is used to inoculate a large culture at a ratio of 1:100 to 1:250. The cells are grown to an optical density 600 (O.D.⁶⁰⁰) of between 0.4 and 0.6. IPTG ("Isopropyl-B-D-thiogalacto pyranoside") is then added to a final concentration of 1 mM. IPTG induces by inactivating the lacI repressor, clearing the P/O leading to increased gene expression. Cells are grown an extra 3 to 4 hours. Cells are then harvested by centrifugation. The cell pellet is solubilized in the chaotropic agent 6 Molar Guanidine HCl. After clarification, solubilized FGF-15 is purified from this solution by chromatography on a Nickel-Chelate column under conditions that allow for tight binding by proteins containing the 6-His tag (Hochuli, E. et al., J. Chromatography 411:177-184 (1984)). The proteins are eluted from the column in 6 molar guanidine HCl pH 5.0 and for the purpose of renaturation adjusted to 3 molar guanidine HCl, 100mM sodium phosphate, 10 mmolar glutathione (reduced) and 2 mmolar glutathione (oxidized). After incubation in this solution for 12 hours the proteins are dialyzed to 10 mmolar sodium phosphate.

Example 2Cloning and expression of FGF-15 using the baculovirus expression system

The DNA sequence encoding the full length FGF-15 protein, ATCC # 97146, is amplified using PCR oligonucleotide primers corresponding to the 5' and 3' sequences of the gene:

The FGF-15 5' primer has the sequence 5' CTAGTGGATCCGCC **ATCATGGTAAAACCGGTGCCC** 3' (SEQ ID NO:5) and contains a BamHI restriction enzyme site (in bold) followed by 4 nucleotides resembling an efficient signal for the initiation of translation in eukaryotic cells (Kozak, M., J. Mol. Biol., 196:947-950 (1987) which is just behind the first 18 nucleotides of the gene (the initiation codon for translation "ATG" is underlined).

The 3' primer has the sequence 5' CGACTGGTACCAGCCACGGA GCAGGAATGTCT 3' (SEQ ID NO:6) and contains the cleavage site for the restriction endonuclease Asp718 (in bold) and 21 nucleotides complementary to the 3' non-translated sequence of the gene.

The amplified sequences are isolated from a 1% agarose gel using a commercially available kit ("GeneClean," BIO 101 Inc., La Jolla, Ca.). The fragment is then digested with the respective endonucleases and purified again on a 1% agarose gel. This fragment is designated F2.

The vector pA2 (modifications of pVL941 vector, discussed below) is used for the expression of the proteins using the baculovirus expression system (for review see: Summers, M.D. and Smith, G.E. 1987, A manual of methods for baculovirus vectors and insect cell culture procedures, Texas Agricultural Experimental Station Bulletin No. 1555). This expression vector contains the strong polyhedrin promoter of the Autographa californica nuclear polyhedrosis virus (AcMNPV) followed by the recognition sites for the restriction endonucleases BamHI and XbaI. The polyadenylation site of the simian virus (SV)40 is used for

efficient polyadenylation. For an easy selection of recombinant virus the beta-galactosidase gene from *E. coli* is inserted in the same orientation as the polyhedrin promoter followed by the polyadenylation signal of the polyhedrin gene. The polyhedrin sequences are flanked at both sides by viral sequences for the cell-mediated homologous recombination of co-transfected wild-type viral DNA. Many other baculovirus vectors could be used in place of pA2 such as pRG1, pAc373, pVL941 and pAcIM1 (Luckow, V.A. and Summers, M.D., *Virology*, 170:31-39).

The plasmid is digested with the restriction enzymes and dephosphorylated using calf intestinal phosphatase by procedures known in the art. The DNA is then isolated from a 1% agarose gel using the commercially available kit ("Geneclean" BIO 101 Inc., La Jolla, Ca.). This vector DNA is designated V2.

Fragment F2 and the dephosphorylated plasmid V2 are ligated with T4 DNA ligase. *E. coli* DH5 α cells are then transformed and bacteria identified that contained the plasmid (pBacFGF-15) using the respective restriction enzymes. The sequence of the cloned fragment are confirmed by DNA sequencing.

5 μ g of the plasmid pBacFGF-15 is co-transfected with 1.0 μ g of a commercially available linearized baculovirus ("BaculoGold™ baculovirus DNA", Pharmingen, San Diego, CA.) using the lipofection method (Felgner et al. *Proc. Natl. Acad. Sci. USA*, 84:7413-7417 (1987)).

1 μ g of BaculoGold™ virus DNA and 5 μ g of the plasmid is mixed in a sterile well of microtiter plates containing 50 μ l of serum free Grace's medium (Life Technologies Inc., Gaithersburg, MD). Afterwards 10 μ l Lipofectin plus 90 μ l Grace's medium are added, mixed and incubated for 15 minutes at room temperature. Then the transfection mixture is added drop-wise to the Sf9 insect cells (ATCC CRL 1711) seeded in 35 mm tissue culture plates with 1 ml Grace's medium without

serum. The plates are rocked back and forth to mix the newly added solution. The plates are then incubated for 5 hours at 27°C. After 5 hours the transfection solution is removed from the plate and 1 ml of Grace's insect medium supplemented with 10% fetal calf serum is added. The plates are put back into an incubator and cultivation continued at 27°C for four days.

After four days the supernatant is collected and plaque assays performed similar as described by Summers and Smith (supra). As a modification an agarose gel with "Blue Gal" (Life Technologies Inc., Gaithersburg) is used which allows an easy isolation of blue stained plaques. (A detailed description of a "plaque assay" can also be found in the user's guide for insect cell culture and baculovirology distributed by Life Technologies Inc., Gaithersburg, page 9-10).

Four days after the serial dilution the virus is added to the cells and blue stained plaques are picked with the tip of an Eppendorf pipette. The agar containing the recombinant viruses is then resuspended in an Eppendorf tube containing 200 μ l of Grace's medium. The agar is removed by a brief centrifugation and the supernatant containing the recombinant baculovirus is used to infect Sf9 cells seeded in 35 mm dishes. Four days later the supernatants of these culture dishes are harvested and then stored at 4°C.

Sf9 cells are grown in Grace's medium supplemented with 10% heat-inactivated FBS. The cells are infected with the recombinant baculovirus V-FGF-15 at a multiplicity of infection (MOI) of 2. Six hours later the medium is removed and replaced with SF900 II medium minus methionine and cysteine (Life Technologies Inc., Gaithersburg). 42 hours later 5 μ Ci of 35 S-methionine and 5 μ Ci 35 S cysteine (Amersham) are added. The cells are further incubated for 16 hours before they are harvested by centrifugation and the labelled proteins visualized by SDS-PAGE and autoradiography.

Example 4

Expression of Recombinant FGF-15 in COS cells

The expression of plasmids, FGF-15-HA derived from a vector pcDNA3/Amp (Invitrogen) containing: 1) SV40 origin of replication, 2) ampicillin resistance gene, 3) E.coli replication origin, 4) CMV promoter followed by a polylinker region, an SV40 intron and polyadenylation site. DNA fragments encoding the entire FGF-15 precursor and an HA tag fused in frame to the 3' end is cloned into the polylinker region of the vector, therefore, the recombinant protein expression is directed under the CMV promoter. The HA tag corresponds to an epitope derived from the influenza hemagglutinin protein as previously described (I. Wilson, H. Niman, R. Heighten, A. Cherenson, M. Connolly, and R. Lerner, 1984, Cell 37:767, (1984)). The infusion of HA tag to the target protein allows easy detection of the recombinant protein with an antibody that recognizes the HA epitope.

The plasmid construction strategy is described as follows:

The DNA sequence encoding FGF-15, ATCC # 97146, is constructed by PCR using two primers: the 5' primer 5' CTAG TGGATCCGCCATCATGGTAAAACCGGTGCC 3' (SEQ ID NO:7) contains a BamHI site followed by 18 nucleotides of coding sequence starting from the initiation codon; the 3' sequence 5' GTCGACCTCGAGTGTGTGCTTACTCTTGTT 3' (SEQ ID NO:8) contains complementary sequences to an XhoI site, translation stop codon, HA tag and the last 18 nucleotides of the FGF-15 coding sequence (not including the stop codon). Therefore, the PCR product contains a BamHI site, coding sequence followed by HA tag fused in frame, a translation termination stop codon next to the HA tag, and an XhoI site.

The PCR amplified DNA fragments and the vector, pcDNA3/Amp, are digested with the respective restriction enzymes and ligated. The ligation mixture is transformed into E. coli strain SURE (available from Stratagene Cloning

Systems, La Jolla, CA 92037) the transformed culture is plated on ampicillin media plates and resistant colonies are selected. Plasmid DNA is isolated from transformants and examined by restriction analysis for the presence of the correct fragment. For expression of the recombinant FGF-15 COS cells are transfected with the expression vector by DEAE-DEXTRAN method (J. Sambrook, E. Fritsch, T. Maniatis, Molecular Cloning: A Laboratory Manual, Cold Spring Laboratory Press, (1989)). The expression of the FGF-15-HA protein is detected by radiolabelling and immunoprecipitation method (E. Harlow, D. Lane, Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory Press, (1988)). Cells are labelled for 8 hours with ³⁵S-cysteine two days post transfection. Culture media is then collected and cells are lysed with detergent (RIPA buffer (150 mM NaCl, 1% NP-40, 0.1% SDS, 1% NP-40, 0.5% DOC, 50mM Tris, pH 7.5) (Wilson, I. et al., Id. 37:767 (1984)). Both cell lysate and culture media are precipitated with an HA specific monoclonal antibody. Proteins precipitated are analyzed on 15% SDS-PAGE gels.

Numerous modifications and variations of the present invention are possible in light of the above teachings and, therefore, within the scope of the appended claims, the invention may be practiced otherwise than as particularly described.

SEQUENCE LISTING

- (1) GENERAL INFORMATION:
- (i) APPLICANT: GREENE, ET AL.
- (ii) TITLE OF INVENTION: Fibroblast Growth Factor-15
- (iii) NUMBER OF SEQUENCES: 8
- (iv) CORRESPONDENCE ADDRESS:
- (A) ADDRESSEE: CARELLA, BYRNE, BAIN, GILFILLAN,
CECCHI, STEWART & OLSTEIN
- (B) STREET: 6 BECKER FARM ROAD
- (C) CITY: ROSELAND
- (D) STATE: NEW JERSEY
- (E) COUNTRY: USA
- (F) ZIP: 07068
- (v) COMPUTER READABLE FORM:
- (A) MEDIUM TYPE: 3.5 INCH DISKETTE
- (B) COMPUTER: IBM PS/2
- (C) OPERATING SYSTEM: MS-DOS
- (D) SOFTWARE: WORD PERFECT 5.1
- (vi) CURRENT APPLICATION DATA:
- (A) APPLICATION NUMBER:
- (B) FILING DATE: Concurrently
- (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA
- (A) APPLICATION NUMBER: 08/207,412
- (B) FILING DATE: 8 MAR 1994
- (viii) ATTORNEY/AGENT INFORMATION:
- (A) NAME: FERRARO, GREGORY D.
- (B) REGISTRATION NUMBER: 36,134
- (C) REFERENCE/DOCKET NUMBER: 325800-364
- (ix) TELECOMMUNICATION INFORMATION:
- (A) TELEPHONE: 201-994-1700
- (B) TELEFAX: 201-994-1744
- (2) INFORMATION FOR SEQ ID NO:1:
- (i) SEQUENCE CHARACTERISTICS
- (A) LENGTH: 680 BASE PAIRS
- (B) TYPE: NUCLEIC ACID
- (C) STRANDEDNESS: SINGLE
- (D) TOPOLOGY: LINEAR

- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:
 - (2) INFORMATION FOR SEQ ID NO:2:
 - (i) SEQUENCE CHARACTERISTICS
 - (A) LENGTH: 225 AMINO ACIDS
 - (B) TYPE: AMINO ACID
 - (C) STRANDEDNESS:
 - (D) TOPOLOGY: LINEAR
 - (ii) MOLECULE TYPE: PROTEIN
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:
 - (2) INFORMATION FOR SEQ ID NO:3:
 - (i) SEQUENCE CHARACTERISTICS
 - (A) LENGTH: 32 BASE PAIRS
 - (B) TYPE: NUCLEIC ACID
 - (C) STRANDEDNESS: SINGLE
 - (D) TOPOLOGY: LINEAR
 - (ii) MOLECULE TYPE: Oligonucleotide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:
 - (2) INFORMATION FOR SEQ ID NO:4:
 - (i) SEQUENCE CHARACTERISTICS
 - (A) LENGTH: 33 BASE PAIRS
 - (B) TYPE: NUCLEIC ACID
 - (C) STRANDEDNESS: SINGLE
 - (D) TOPOLOGY: LINEAR
 - (ii) MOLECULE TYPE: Oligonucleotide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:
 - (2) INFORMATION FOR SEQ ID NO:5:
 - (i) SEQUENCE CHARACTERISTICS
 - (A) LENGTH: 33 BASE PAIRS
 - (B) TYPE: NUCLEIC ACID

- (C) STRANDEDNESS: SINGLE
 - (D) TOPOLOGY: LINEAR
 - (ii) MOLECULE TYPE: Oligonucleotide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:
- (2) INFORMATION FOR SEQ ID NO:6:
- (i) SEQUENCE CHARACTERISTICS
 - (A) LENGTH: 30 BASE PAIRS
 - (B) TYPE: NUCLEIC ACID
 - (C) STRANDEDNESS: SINGLE
 - (D) TOPOLOGY: LINEAR
 - (ii) MOLECULE TYPE: Oligonucleotide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:
- (2) INFORMATION FOR SEQ ID NO:7:
- (i) SEQUENCE CHARACTERISTICS
 - (A) LENGTH: 33 BASE PAIRS
 - (B) TYPE: NUCLEIC ACID
 - (C) STRANDEDNESS: SINGLE
 - (D) TOPOLOGY: LINEAR
 - (ii) MOLECULE TYPE: Oligonucleotide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:
- (2) INFORMATION FOR SEQ ID NO:8:
- (i) SEQUENCE CHARACTERISTICS
 - (A) LENGTH: 33 BASE PAIRS
 - (B) TYPE: NUCLEIC ACID
 - (C) STRANDEDNESS: SINGLE
 - (D) TOPOLOGY: LINEAR
 - (ii) MOLECULE TYPE: Oligonucleotide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

WHAT IS CLAIMED IS:

1. An isolated polynucleotide comprising a member selected from the group consisting of:
 - (a) a polynucleotide encoding the polypeptide as set forth in Figure 1;
 - (b) a polynucleotide capable of hybridizing to and which is at least 70% identical to the polynucleotide of (a); and
 - (c) a polynucleotide fragment of the polynucleotide of (a) or (b).
2. The polynucleotide of Claim 1 encoding the polypeptide comprising amino acid 1 to amino acid 252 as set forth in SEQ ID NO:2.
3. The polynucleotide of Claim 1 wherein the polynucleotide is DNA.
4. The polynucleotide of Claim 1 wherein said polynucleotide is RNA.
5. The polynucleotide of Claim 1 wherein the polynucleotide is genomic DNA.
6. The polynucleotide of Claim 1 comprising from nucleotide 1 to nucleotide 759 as set forth in SEQ ID NO:1.
7. An isolated polynucleotide comprising a member selected from the group consisting of:
 - (a) a polynucleotide encoding a mature polypeptide encoded by the DNA contained in ATCC Deposit No. 97146;
 - (b) a polynucleotide encoding the polypeptide expressed by the DNA contained in ATCC Deposit No. 97146;

- (c) a polynucleotide capable of hybridizing to and which is at least 70% identical to the polynucleotide of (a) or (b); and
- (d) a polynucleotide fragment of the polynucleotide of (a), (b) or (c).

8. The polynucleotide of Claim 7 wherein said polynucleotide encodes a polypeptide expressed by the DNA contained in ATCC Deposit No. 97146.

9. The polynucleotide of Claim 7 wherein said polynucleotide encodes a polypeptide expressed by the DNA contained in ATCC Deposit No. 97146.

10. The polynucleotide of Claim 7 wherein said polynucleotide encodes a polypeptide expressed by the DNA contained in ATCC Deposit No. 97146.

11. A vector containing the DNA of Claim 2.

12. A host cell genetically engineered with the vector of Claim 11.

13. A process for producing a polypeptide comprising: expressing from the host cell of Claim 12 the polypeptide encoded by said DNA.

14. A process for producing cells capable of expressing a polypeptide comprising genetically engineering cells with the vector of Claim 11.

15. A polypeptide selected from the group consisting of (i) a polypeptide having the deduced amino acid sequence of Figure 1 and fragments, analogs and derivatives thereof; and (ii) a polypeptide encoded by the cDNA of ATCC Deposit

No. 97146 and fragments, analogs and derivatives of said polypeptide.

16. The polypeptide of Claim 15 wherein the polypeptide has the deduced amino acid sequence of Figure 1.

17. An antibody against the polypeptide of claim 15.

18. A compound which inhibits the polypeptide of claim 15.

19. A compound which activates a receptor to the polypeptide of claim 15.

20. A method for the treatment of a patient having need of an FGF-15 polypeptide comprising: administering to the patient a therapeutically effective amount of the polypeptide of claim 15.

21. A method for the treatment of a patient having need to inhibit an FGF-15 polypeptide comprising: administering to the patient a therapeutically effective amount of the compound of claim 18.

22. The method of claim 20 wherein said therapeutically effective amount of said polypeptide is administered by providing to the patient DNA encoding said polypeptide and expressing said polypeptide *in vivo*.

23. The method of claim 21 wherein said compound is a polypeptide and a therapeutically effective amount of the compound is administered by providing to the patient DNA encoding said antagonist and expressing said antagonist *in vivo*.

24. A process for identifying compounds active as agonists to the polypeptide of claim 15 comprising:
- (a) combining a compound to be screened and a reaction mixture containing cells under conditions where the cells are normally stimulated by said polypeptide, said reaction mixture containing a label incorporated into the cells as they proliferate; and
 - (b) determining the extent of proliferation of the cells to identify if the compound is an effective agonist.
25. A process for identifying compounds active as antagonists to the polypeptide of claim 15 comprising:
- (a) combining a compound to be screened, the polypeptide and a reaction mixture containing cells under conditions where the cells are normally stimulated by said polypeptide, said reaction mixture containing a label incorporated into the cells as they proliferate; and
 - (b) determining the extent of proliferation of the cells to identify if the compound is an effective antagonist.
26. A process for diagnosing a disease or a susceptibility to a disease related to an under-expression of the polypeptide of claim 15 comprising:
- determining a mutation in the nucleic acid sequence encoding said polypeptide.
27. A diagnostic process comprising:
- analyzing for the presence of the polypeptide of claim 15 in a sample derived from a host.

FGF15 coding sequence
(Sequenced Strand)

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1  ATGGTAAAAC CGGTGGCCCT CTTCAGGAGA ACTGATTTC
   M V K P V P L F R R T D F
41  AATTATTATT ATGCAACCAC AAGGATCTCT TCTTTCTCAG
   K L L L C N H K D L F F L R
81  GGIGICTAAG CTGCTGGATT GCTTTTCGCC CAAATCAATG
   V S K L L D C F S P K S M
121  TGGTTCCTTT GGAACATTTT CAGCAAAGGA ACGCATATGC
   W F L W N I F S K G T H M
161  TGCAGTGTCT TTGTGGCAAG AGTCTTAAGA AAAACAAGAA
   L Q C L C G K S L K K N K N
201  CCCAACTGAT CCCAGCTCA AGGGTATAGT GACCAGGTTA
   P T D P Q L K G I V T R L
241  TATTGCAGGC AAGGCTACTA CTIGCAAATG CACCCCGATG
   Y C R Q G Y Y L Q M H P D
281  GAGCTCTCGA TGGAACCAAG GGIGACAGCA CTAATTCTAC
   G A L D G T K G D S T N S T
321  ACTCTTCAAC CTCATACCAG TGGGACTACG TGTGTGTGCC
   L F N L I P V G L R V V A
361  ATCCAGGGAG TGAAAACAGG GTTGTATATA ACCATGAATG
   I Q G V K T G L Y I T M N
401  GAGAAGGTTA CCTCTACCCA TCAGAACTTT TTACCCCTGA
   G E G Y L Y P S E L F T P E
441  ATGCAAGTIT AAAGAATCTG TTTTIGAAA TTATTATGTA
   C K F K E S V F E N Y Y V
481  ATCTACTCAT CCATGTGTGA CAGACAACAG GAATCTGGTA
   I Y S S M L Y R Q Q E S G
521  GAGCCITGGIT TTTGGGATTA AATAAGGAAG GGCAAGCTAT
   R A W F L G L N K E G Q A M
561  GAAAGGGAAC AGAGTAAAGA AAACCAAACC AGCAGCTCAT
   K G N R V K K T K P A A H
601  TTTCTACCCA AGCCATTTGA AGTTGCCATG TACCGAGAAC
   F L P K P L E V A M Y R E
641  CATCTTTTGA TGATGTGTGG GAAACGGTCC CGAAGCCTGG
   P S L H D V G E T V P K P G

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FGF15 coding sequence
(Sequenced Strand)

681 GGIGACGCCA AGTAAAAGCA CAAGTGGGTC TGCAATAATG
V T P S K S T S A S A I M
701 AATGGAGGCA AACCAGTCAA CAAGAGTAAG ACAACATAG
N G G K P V N K S K T T .

Align. report of HGS FGF ALIGN ITS, using Clustal method with PAM250 residue weight
Tuesday, April 25, 1995 11:07 AM

M A												L G L		Majority																							
		10					20					30																									
1	MA	-	-	-	E	-	-	-	-	-	-	-	-	-	-	E	aFGF																				
1	MA	-	-	-	A	-	-	-	-	-	-	-	-	-	-	S	bFGF																				
1	MA	G	L	I	W	L	L	-	-	-	-	-	-	-	L	S	FGF3																				
1	MA	S	-	-	-	-	-	-	-	-	G	P	G	T	A	A	V	A	L	L	P	A	V	L	A	I	FGF4										
1	MA	S	L	S	F	L	L	L	F	F	S	H	-	-	-	-	-	-	-	-	-	-	-	-	-	L	I	FGF5									
1	MA	E	L	G	Q	K	L	F	I	T	M	S	R	G	A	G	R	L	Q	G	T	L	W	A	L	V	F	I	FGF6								
1	MA	-	H	K	W	I	L	T	-	-	-	W	I	L	P	T	L	L	-	-	Y	R	S	C	F	H	I	I	C	FGF7							
1	MA	G	S	P	R	S	-	-	-	-	-	-	-	-	-	-	-	-	-	-	A	L	S	C	L	L	L	H	L	L	V	I	C	FGF8			
1	MA	-	-	-	P	L	G	E	V	G	N	Y	F	G	V	Q	D	A	-	-	-	-	-	-	-	V	P	F	S	N	-	-	FGF9				
1	MA	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF10			
1	MA	S	-	-	-	-	-	-	-	-	-	-	-	-	-	G	K	V	T	K	P	K	E	E	K	D	A	S	K	V	-	-	FGF11				
1	MA	-	W	K	W	I	L	T	H	C	A	S	A	F	P	H	L	P	G	C	C	C	C	C	F	L	L	F	L	-	-	-	-	FGF12			
1	MA	R	L	L	P	N	L	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF13		
1	MA	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF14	
1	MA	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF15

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		40					50					60																									
6	IT	T	F	T	A	-	-	-	-	L	T	E	K	F	N	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	aFGF		
6	IT	T	T	L	P	A	-	-	-	-	L	P	E	D	G	G	-	-	-	S	G	A	F	-	-	-	-	-	-	-	-	-	-	-	-	bFGF	
12	L	E	P	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	G	W	P	A	A	G	P	G	A	R	L	R	-	-	-	-	FGF3		
20	L	A	P	W	A	-	-	-	-	-	G	R	G	G	A	A	A	P	T	A	P	N	G	T	L	E	A	E	L	-	-	-	-	-	-	FGF4	
16	L	S	A	W	A	H	G	E	K	R	L	A	P	K	G	Q	P	G	P	A	A	T	D	R	N	P	R	G	S	S	-	-	-	-	-	FGF5	
31	L	V	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF6	
25	V	G	T	I	S	L	A	C	N	D	M	T	P	E	Q	M	A	T	N	V	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF7	
21	Q	A	Q	V	T	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF8	
22	V	P	V	L	P	V	D	S	P	V	L	L	S	D	H	L	G	Q	S	E	A	G	G	L	-	-	-	-	-	-	-	-	-	-	-	FGF9	
2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF10	
18	L	D	D	A	P	P	G	T	Q	E	Y	I	M	L	R	Q	D	S	I	Q	S	A	E	L	K	K	K	E	-	-	-	-	-	-	-	FGF11	
30	V	S	S	V	P	V	T	C	Q	A	L	G	Q	D	M	V	S	P	E	A	T	N	S	S	S	S	S	F	S	-	-	-	-	-	-	-	FGF12
18	Q	T	Q	G	E	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF13	
18	P	G	G	S	R	P	V	S	A	Q	R	R	V	C	P	R	G	T	-	-	K	S	L	C	Q	K	Q	L	L	I	-	-	-	-	FGF14		
16	L	-	-	-	L	C	N	H	K	D	L	F	F	L	R	V	S	K	L	L	D	C	F	S	P	K	S	M	W	F	-	-	-	-	-	FGF15	

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		70					80					90																									
19	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	aFGF	
22	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	bFGF
27	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF3
44	-	-	E	R	R	W	E	S	L	V	A	L	S	L	A	R	L	P	V	A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF4
46	S	R	Q	S	S	S	S	S	A	M	S	S	S	S	A	S	S	S	P	A	A	S	L	S	Q	G	S	G	L	E	-	-	-	-	-	-	FGF5
49	L	D	S	R	G	W	G	T	L	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF6
45	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF7
26	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF8
46	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF9
2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF10
46	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF11
59	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF12
23	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF13
46	L	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF14
43	L	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	FGF15

Alignment Report of HGS FGF ALIGNMENTS, using Clustal method with PAM250 residue weight
Tuesday, April 25, 1995 11:07 AM

	H L K G - - L - G I V R - R R L Y C R T G - G F H L Q I L P Majority																														
	100					110					120																				
22	N	Y	I	-	-	-	-	-	-	K	P	K	L	-	-	C	S	N	G	-	C	H	F	L	R	I	L	E	aFGF		
25	H	F	R	-	-	-	-	-	-	D	P	K	L	-	-	L	Y	C	K	N	-	C	F	F	L	R	I	H	bFGF		
38	H	L	-	G	-	-	-	-	-	A	P	F	R	F	K	-	L	Y	C	A	K	-	-	Y	H	L	Q	L	H	FGF3	
71	S	G	A	G	D	Y	I	L	G	I	K	F	L	F	R	L	Y	C	N	V	I	G	F	H	L	Q	A	L	F	FGF4	
76	Q	S	S	F	Q	W	S	L	A	-	T	G	S	-	L	Y	C	H	V	G	I	G	F	H	L	Q	I	Y	FGF5		
74	W	E	S	G	-	Y	L	V	G	I	K	R	Q	R	F	L	Y	C	N	V	G	I	G	F	H	L	Q	V	L	FGF6	
58	Y	M	E	-	-	-	-	-	D	I	P	V	P	R	L	F	C	R	T	Q	-	-	W	Y	I	R	I	D	K	FGF7	
41	L	V	T	D	Q	L	S	R	R	L	I	P	T	Y	Q	-	L	Y	S	R	T	S	-	C	R	H	V	L	V	FGF8	
56	H	L	K	G	-	-	-	-	-	I	L	F	R	P	Q	-	L	Y	C	R	T	G	-	-	F	H	L	E	I	F	FGF9
2	E	S	K	E	P	Q	L	K	G	I	V	-	-	T	R	L	F	S	Q	Q	-	-	Y	F	L	Q	M	H	F	FGF10	
70	E	P	E	E	P	Q	L	K	G	I	V	-	-	T	K	L	S	P	Q	-	-	-	Y	H	L	Q	L	Q	A	FGF11	
72	H	L	Q	G	-	-	-	-	D	V	P	W	F	K	L	F	S	F	K	-	-	-	Y	F	L	K	I	E	K	FGF12	
38	A	M	T	D	Q	L	S	R	R	O	I	P	E	Y	Q	L	Y	S	R	T	S	-	C	R	H	V	Q	V	-	FGF13	
62	R	G	P	E	P	Q	L	K	G	I	V	-	-	T	K	L	F	C	H	Q	-	-	G	E	Y	L	Q	A	N	F	FGF14
67	N	P	T	D	P	Q	L	K	G	I	V	-	-	T	R	L	Y	C	R	Q	-	-	G	Y	L	Q	M	H	F	FGF15	

	D G R I D G T K E D N S P Y S L L E L I P V E V G V - V A I Majority																														
	130					140					150																				
43	D	G	T	V	D	G	T	R	D	R	S	D	O	H	I	Q	L	Q	L	S	A	E	S	V	G	E	-	V	Y	I	aFGF
46	D	G	R	V	D	G	V	R	E	K	S	D	F	H	I	K	L	Q	L	Q	A	E	E	R	G	V	-	V	S	I	bFGF
61	S	C	R	V	N	G	S	L	E	-	H	S	A	Y	S	I	L	E	I	T	A	V	E	V	G	I	-	V	A	I	FGF3
101	D	G	R	I	G	G	A	H	A	D	T	-	R	D	S	L	L	E	L	S	P	V	E	R	G	V	-	V	S	I	FGF4
105	D	G	K	V	N	G	S	H	E	A	-	-	M	L	S	V	L	E	I	F	A	S	Q	D	I	-	V	G	I	FGF5	
103	D	G	R	I	S	G	T	H	E	A	-	-	P	Y	S	L	L	E	I	S	T	V	E	R	G	V	-	V	S	L	FGF6
92	R	C	K	V	K	G	T	Q	E	M	K	N	N	Y	N	I	M	E	I	R	T	A	G	I	-	V	A	I	FGF7		
70	N	K	P	I	N	A	M	A	E	D	G	D	I	F	A	K	L	I	V	E	T	D	T	F	S	R	V	R	V	FGF8	
79	N	G	T	I	Q	G	T	R	K	D	H	S	R	F	G	I	L	E	F	I	S	I	A	G	L	-	V	S	I	FGF9	
28	D	G	T	I	D	G	T	K	E	D	S	D	Y	T	L	F	N	L	I	P	V	G	L	R	V	-	V	A	I	FGF10	
96	D	G	T	I	D	G	T	K	E	D	S	T	Y	T	L	F	N	L	I	P	V	G	L	R	V	-	V	A	I	FGF11	
95	N	K	V	S	G	T	K	E	H	C	P	Y	S	I	L	E	I	T	S	E	I	Q	-	V	A	I	FGF12				
66	G	R	I	S	A	T	A	E	D	G	N	K	F	A	K	L	I	V	E	T	D	T	F	S	R	V	R	V	FGF13		
88	D	G	S	I	Q	G	T	P	E	D	T	S	S	F	T	H	F	N	L	I	P	V	G	L	R	V	-	V	A	I	FGF14
93	D	G	A	L	D	G	T	K	E	D	S	T	N	S	T	L	F	N	L	I	P	V	G	L	R	V	-	V	A	I	FGF15

	K G V E S G L Y L A M N K K G K L Y A S E L F T D - E C K F Majority																														
	160					170					180																				
72	K	S	T	E	T	Q	Y	L	A	H	D	T	D	-	L	L	Y	G	S	Q	T	P	N	E	-	E	C	L	S	aFGF	
75	K	G	V	C	A	N	R	Y	L	A	H	K	E	D	-	R	R	L	A	S	K	C	V	L	-	E	C	F	E	bFGF	
89	R	G	L	F	S	G	R	Y	L	A	M	N	R	-	R	L	Y	A	S	E	H	Y	S	A	-	E	C	E	S	FGF3	
129	F	G	V	A	S	R	F	F	V	A	H	S	S	E	S	L	Y	G	S	P	F	F	T	D	-	E	C	K	F	FGF4	
133	R	Q	V	F	S	N	K	F	L	A	H	S	E	K	G	L	H	A	S	A	K	F	T	D	-	D	C	K	F	FGF5	
131	F	Q	V	R	C	A	-	F	V	A	H	S	E	S	G	R	L	Y	A	T	P	S	Q	E	-	E	C	K	F	FGF6	
111	K	G	V	E	S	E	F	-	L	A	M	N	K	K	G	K	L	Y	A	S	E	L	F	T	D	-	E	C	K	F	FGF7
100	R	G	A	F	T	G	L	I	C	H	N	K	K	G	K	L	Y	A	S	E	L	F	T	D	-	D	C	V	E	FGF8	
108	R	G	V	D	S	G	L	Y	L	G	H	E	F	G	E	L	Y	G	S	E	F	K	L	Q	-	E	C	V	E	FGF9	
57	Q	G	V	K	A	S	-	V	A	H	H	E	G	E	-	Y	L	Y	S	D	D	V	E	T	P	-	E	C	K	F	FGF10
125	Q	G	V	Q	T	K	-	L	A	M	N	S	E	-	Y	L	Y	T	S	E	L	F	T	P	-	E	C	K	F	FGF11	
124	K	A	I	N	-	N	Y	L	A	M	N	K	K	G	K	L	Y	A	S	E	L	F	T	D	-	D	C	V	E	FGF12	
96	K	G	A	E	S	E	K	-	I	C	H	N	R	S	-	L	I	G	K	P	S	G	K	S	K	D	-	V	E	FGF13	
117	Q	S	A	K	L	E	H	-	M	A	H	H	A	E	S	-	L	Y	S	E	P	H	F	T	A	-	E	C	K	F	FGF14
122	Q	G	V	K	T	-	L	I	T	H	G	E	S	-	Y	L	Y	P	S	E	L	F	T	P	-	E	C	K	F	FGF15	

Alignment Report of HGS FGF ALIGNMENTS, using Clustal method with PAM250 residue weight table.
 Tuesday, April 25, 1995 11:07 AM

	280	290	300	Majority
155				aFGF
155				bFGF
206	V Q P R R R R Q K Q S P D N L E P S H V Q A S R L G S Q L E			FGF3
206				FGF4
242	I K S K I P - - - - - - - - - - L S A P R K N T N S V K Y R L K			FGF5
208				FGF6
194				FGF7
195	N Y P P F T R - - - - - - - - - - S L R G S Q R T W A P E			FGF8
205	- -			FGF9
161	- - S R K S S - - - - - - - - - - G T P T M N G G K V V N Q D -			FGF10
232	T P T K S R S - - - - - - - - - - V S G V L N G G K S M S H N E			FGF11
208				FGF12
195	G S A P T R R - - - - - - - - - - T - - - - - K R T R R P Q			FGF13
224	A P .			FGF14
229	T P S K S T S - - - - - - - - - - A S A I M N G G K P V N K S K			FGF15

		Majority
155		aFGF
155		bFGF
236	A S A H	FGF3
206		FGF4
264	F R F G	FGF5
208		FGF6
194		FGF7
214	P - - R	FGF8
205	L S Q S	FGF9
180	- S - T	FGF10
254	- S - T	FGF11
208		FGF12
210	P L T .	FGF13
226		FGF14
251	T T .	FGF15

Decoration 'Decoration #1': Shade (with solid black) residues that match the Consensus exactly.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US95/07156

A. CLASSIFICATION OF SUBJECT MATTER IPC(6) :C12N 15/12, 15/18; C07K 14/50 US CL :Please See Extra Sheet. According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) U.S. : 435/6, 69.1, 172.3, 240.2, 320.1; 530/350, 399; 536/23.5, 24.3, 24.31, 24.33 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) APS and DIALOG (files 5, 155, 351, 357, 358) search terms: FGF, heparin, fibroblast growth factor		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	PROGRESS IN GROWTH FACTOR RESEARCH, Volume 1, issued 1989, M. Klagsbrun, "The Fibroblast Growth Factor Family: Structural and Biological Properties", pages 207-235, see entire document.	1-16
A	J. BRUGGE et al, "ORIGINS OF HUMAN CANCER: A COMPREHENSIVE REVIEW" published 1991 by Cold Spring Harbor Laboratory Press (N.Y.), pages 675-683, see pages 675-683.	1-16
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.		
*	Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A"	document defining the general state of the art which is not considered to be of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E"	earlier document published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L"	document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&" document member of the same patent family
"O"	document referring to an oral disclosure, use, exhibition or other means	
"P"	document published prior to the international filing date but later than the priority date claimed	
Date of the actual completion of the international search	Date of mailing of the international search report	
03 AUGUST 1995	11 SEP 1995	
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231	Authorized officer	
Facsimile No. (703) 305-3230	MARIANNE PORTA ALLEN	
	Telephone No. (703) 308-0196	

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US95/07156

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

1-16

Remark on Protest

The additional search fees were accompanied by the applicant's protest.

No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US95/07156

A. CLASSIFICATION OF SUBJECT MATTER:

US CL :

435/6, 69.1, 172.3, 240.2, 320.1; 530/350, 399; 536/23.5, 24.3, 24.31, 24.33

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be examined, the appropriate additional examination fees must be paid.

Group I, claims 1-16, drawn to nucleic acid sequences and the encoded proteins.

Group II, claim 17, drawn to antibodies.

Group III, claim 18, drawn to inhibitory compounds.

Group IV, claim 19, drawn to receptor activating compounds.

Group V, claim 20, drawn to a method of treatment by administering the protein.

Group VI, claim 21, drawn to a method of treatment by administering the inhibitory compound.

Group VII, claim 22, drawn to a method of treatment by administering a nucleic acid sequence encoding the protein.

Group VIII, claim 23, drawn to a method of treatment by administering a nucleic acid sequence encoding the inhibitory compound.

Group IX, claims 24-25, drawn to a method for identifying compounds.

Group X, claim 26, drawn to a method of diagnosis using nucleic acid sequences.

Group XI, claim 27, drawn to a method of diagnosis using the protein.

The inventions listed as Groups I-XI do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: Groups I-IV are drawn to structurally different products which do not share the same or a corresponding special technical feature. Groups V-XI are drawn to methods having different goals, method steps, and starting materials which do not share the same or a corresponding special technical feature. Note that PCT Rule 13 does not provide for multiple products or methods within a single application.