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(54) Title: CELL ISOLATION AND SCREENING DEVICE AND METHOD OF USING SAME

(57) Abstract: The present invention provides methods and devices for screening a single cell or a small group of cells for a desired biological activity. In particular, the present invention provides for delivering cell(s) to a plurality of cell isolation regions of a cell isolation device, transferring cell(s) to a plurality of wells of a cell expansion device and then detecting the potential desired biological activity of the cell(s). Each of the cell isolation regions comprise a recess or a bioaffinity region sized to isolate a single cell or small group of cells and each of the wells encompass a cavity that provides sufficient volume for cell proliferation.

CELL ISOLATION AND SCREENING DEVICE AND METHOD OF USING SAME

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims priority to Provisional U.S. Application No. 60/334,593 filed December 3, 2001, Provisional U.S. Application No. 60/307,843 filed July 27, 2001, and U.S. Application 10/084,063 filed February 28, 2002, all of which are herein incorporated by reference in their entirety.

FIELD OF THE INVENTION

The present invention relates to devices and methods that are useful for isolating and screening cells for a desired biological activity. In particular, the present invention relates to devices and methods that are useful in isolating and screening hybridoma cells for specific antibody production.

BACKGROUND OF THE INVENTION

One of the major challenges for performing cell based screening is the isolation of small populations of cells in a manner that allows for subsequent screening procedures. Traditional devices and methods of isolating cells do not adequately provide for the isolation of small populations of cells without performing steps that potentially modify cellular function or activity. Isolation of cells is not only important in screening, but also in processes that involve the monitoring, measuring, and/or use of the output of cellular activity or function (e.g. antibody production) for small populations of cells.

For example, with respect to antibody production, one of the approaches used to produce antibodies is to manufacture hybridomas. These are cells created by the fusion of antibody-secreting B-cells and myeloma cells. This method is variable with respect to almost all stages of the process including the duration of screening and required needs for cell growth and proliferation. Much of the variability can be attributed to the immunogen and immune response mounted by the immunized animal. Moreover this process is time consuming and labor intensive.

In general, once the fusion is performed and the cells are plated, there are several issues that have to be addressed. First, the cells will grow at different rates, thus the point at which one must perform the assay for antibody production to assess positive pools of cells can vary and may require more than one assay point on the same pool of cells. During this process, the rapidly growing cells need to be passaged in order to promote viability and to prevent loss of potentially positive clones. The next step is to perform limiting dilution with the goal of achieving clonal populations. Successive rounds of this process may be required to achieve clonal or near clonal populations.

The value of specific monoclonal antibodies as useful tools for diagnostics and immunotherapies has meant that investigators have had no alternative but to tolerate long production periods and high development costs. Rapid and efficient ways of screening hybridoma cell lines for antibody production are not presently available. Therefore, there is a need for methods and devices to accelerate monoclonal antibody production for experimental, diagnostic and therapeutic applications. The devices and methods disclosed herein dramatically shorten the period for monoclonal antibody production from a period of weeks to a period of days. The invention provides a multicomponent device that also has application for other cell types requiring the isolation of a small population of cells and requiring the subsequent clonal expansion of this population of cells.

SUMMARY OF THE INVENTION

The present invention relates to devices and methods for screening a cell or small group of cells for a desired biological activity comprising a cell delivery step, a cell isolation device and a corresponding cell isolation step, a cell expansion device and a corresponding cell expansion step, and a detection device and a corresponding detection step.

In particular, the present invention provides a method of screening cells for a desired biological activity comprising providing a cell isolation device defining a plurality of cell isolation regions, each cell isolation region encompassing a recess, each of the cell isolation regions being sized to isolate about one to about five cells therein, and the cell isolation regions further defining a predetermined pitch with respect to one another. The method further comprises delivering the

about one cell to about five cells to each of the cell isolation regions. The method additionally comprises providing a cell expansion device defining a plurality of wells corresponding to respective ones of the plurality of cell isolation regions, the wells defining a predetermined pitch matching the predetermined pitch of the cell isolation regions. The method also comprises transferring the about one cell to about five cells from the cell isolation regions to the wells and allowing the about one cell to about five cells to proliferate and exhibit a desired biological activity in the wells. The method further comprises assaying the desired biological activity of the one cell to about five cells.

The present invention also provides a kit for screening cells for a desired biological activity, the kit comprising a cell isolation device defining a plurality of cell isolation regions, each cell isolation region encompassing a recess, each of the cell isolation regions being sized to isolate about one to about five cells therein, and the cell isolation regions further defining a predetermined pitch with respect to one another. The kit further comprises a cell expansion device defining a plurality of wells corresponding to respective ones of the plurality of cell isolation regions, and the wells defining a predetermined pitch matching the predetermined pitch of the cell isolation regions.

The present invention further provides a a method of screening cells for a desired biological activity comprising providing a cell isolation device defining a plurality of cell isolation regions, each cell isolation region being sized for isolating about one to about five cells therein, each cell isolation region encompassing a bioaffinity region, the cell isolation regions further defining a predetermined pitch with respect to one another. The method further comprises delivering the about one cell to about five cells to each of the cell isolation regions. The method moreover comprises providing a cell expansion device defining a plurality of wells corresponding to respective ones of the plurality of cell isolation regions and defining a predetermined pitch that matches the predetermined pitch of the cell isolation regions. The method additionally comprises transferring the about one to about five cells from the cell isolation regions to the wells and allowing about one cell to about five cells to proliferate and exhibit a desired biological activity. The method also comprises assaying the biological activity the about one to about five cells.

The present invention also provides a kit for screening cells for a desired biological activity, the kit comprising a cell isolation device defining a plurality of cell isolation regions, each of the cell isolation regions being sized to isolate about one cell to about five cells therein, and each encompassing a bioaffinity region, the cell isolation device further defining a predetermined pitch with respect to one another. The kit also comprises a cell expansion device defining a plurality of wells corresponding to respective ones of the plurality of cell isolation regions.

BRIEF DESCRIPTION OF THE DRAWINGS

The present invention will become more fully understood from the detailed description given hereinbelow and the accompanying drawings which are given by way of illustration only, and thus are not limitative of the present invention, and wherein:

FIG. 1 is a top perspective view of an embodiment of a cell delivery device and an embodiment of a cell isolation device according to the present invention.

FIG. 2 is a perspective view of an embodiment of a cell delivery device according to the present invention in contact with an embodiment of a cell isolation device according to the present invention.

FIG. 3 is a schematic view of the channel pathways of an embodiment of the cell delivery device according to the present invention.

FIG. 4 is an oblique perspective view of an embodiment of a cell isolation device according to the present invention.

FIG. 5 is a cross-sectional view of the device of **FIG. 4** along lines II-II.

FIG. 6 is a cross-sectional view of the device of **FIG. 4**.

FIG. 7 is an oblique perspective view of an alternative embodiment of a cell isolation device according to the present invention.

FIG. 8 is a partially cut-away perspective view of an embodiment of a cell isolation device according to the present invention.

FIG. 9A is a cross-sectional view of the cell isolation device of **FIG. 8** along lines III-III showing an embodiment of the cell isolation regions of the cell isolation device.

FIG. 9B is a view similar to **FIG. 8** showing an alternative embodiment of the cell isolation regions of the cell isolation device.

FIG. 10A-E shows respective stages of a method of fabricating an embodiment of the cell isolation device according to the present invention.

FIG. 11 depicts a method of fabricating an alternative embodiment of a cell isolation device according to the present invention.

FIG. 12A-C show respective stages of a method of fabricating an embodiment of a cell isolation device according to the present invention.

FIG. 13A-C shows respective stages of a method of fabricating an embodiment of a cell isolation device according to the present invention.

FIG. 14 is a cross-sectional view of an embodiment of a cell expansion device according to the present invention.

FIG. 15A shows a method of transferring cells from the cell isolation device to an embodiment of the cell expansion device according to the present invention.

FIG. 15B shows a method of transferring cells from the cell isolation device to an alternative embodiment of the cell expansion device according to the present invention.

FIG. 16 depicts a cross-sectional view of an embodiment of the detection device and a method of detecting antibody production according to the present invention.

FIG. 17 is a cross-section view of an alternative embodiment of a cell expansion device according to the present invention.

FIG. 18 shows a method of detecting antibody production according to the present invention.

DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to methods and devices for isolating a small population of cells and for screening a larger population of proliferated cells for biological activity with a minimal amount of manipulation of the cells. Although the present invention contemplates the screening of all types of cells for the detection of any desired biological activity, for purposes of illustration, the methods and devices described herein are occasionally described in the context

of isolating and screening hybridoma cells for specific antibody production. The present invention is not in any way limited to this application and may be used to isolate any types of cells, including all cell lines that secrete substances that can be isolated. Such cell lines include cell lines that secrete or express, for example, antibodies, proteins, carbohydrates, enzymes, peptides, hormones, and receptors. Other cells include, for example, B-cells; T-cells; non-adherent cells like epithelial cells and endothelial cells; oncogenic cell line; genetically engineered cells; and activated cells. Moreover, the present invention may be used to screen for a variety of biological activities including, but not limited to, the expression of surface receptor proteins, enzyme production, and peptide production. Furthermore, the present invention may be used to screen a variety of test agents to determine the effect of the test agents on the desired biological activity. Other types of cells desired to be isolated and screened, other types of biological activity desired to be detected, and specific test agents to be screened will be readily appreciated by one of skill in the art.

In general, the present invention relates to a method and accompanying device for isolating and screening cells including a cell delivery step, a cell isolation step utilizing a cell isolation device, a cell expansion step utilizing a cell expansion device, and a detection step utilizing a detection device. In general, cells are delivered to cell isolation regions of the cell isolation device. By "cell isolation region" is meant any area capable of attracting, trapping, securing and/or binding a single cell or a small population of cells. Cells are then transferred to wells of a cell expansion device. By "well" is meant a cavity that is capable of retaining cells therein. Cells are then transferred from the cell expansion device to the detection device where biological activity is detected, or, in the alternative, the detection device is brought into contact with the cell expansion device.

With respect to particular details of the methods and devices according to the present invention, in the cell delivery step, cells may be delivered to the cell isolation regions of the cell isolation device (described in more detail below) by any means known in the art. For example, cells may be delivered by directly pipetting cells into or on the cell isolation regions of the cell isolation device either manually or by robotic liquid handling systems; bulk delivering the cells in fluid to the cell isolation device and removing the excess by pipetting so as to leave only cells in

or on the cell isolation regions; or by utilizing a delivery device with microfluidic channels to deliver the cells into or on the cell isolation regions.

FIG. 1 illustrates an example of a microfluidic delivery device **200** with microfluidic channels **220** to deliver the cells into or on cell isolation regions **20** of cell isolation device **10**. "Microfluidic delivery device," as used herein, refers to a system or device having fluidic conduits, such as channels, that are typically fabricated at the micron to submicron scale. Generally, the fluidic conduits have at least one cross-sectional dimension in the range of from about 0.1 μm to about 500 μm . As seen in **FIG. 1**, the microfluidic device **200** generally comprises a housing **210** defining a plurality of channels **220**. Each channel includes an entrance port **230** at one end and an exit port **240** at another end. Although **FIG. 1** depicts only two channels, microfluidic device **200** can include more than two channels **220** as seen in **FIG. 3** (wherein dashed lines indicate channel paths and arrows indicate potential flow in multiple directions through each channel). The diameter of channels **220** should be large enough to prevent clogging of channels **220**, preferably being about three times the diameter of the cell. A 100 μm x 100 μm channel is the preferred size yet alternate sizes may include, but are not limited to, 60 μm x 100 μm or 30 μm x 100 μm . Although a microfluidic device for use in the present invention is described briefly below, methods of employing microfluidics for cell delivery are well-known in the art and are described, for example, in Love, et al., MRS BULLETIN, pp.523-527 (July 2001) "Fabrication of Three-Dimensional Microfluidic Systems by Soft Lithography," Delamarche et al.; JOURNAL OF AMERICAN CHEMICAL SOCIETY, Vol. 120, pp.500-508 (1998), Delamarche et al.; SCIENCE, Vol.276, pp.779-781 (May 1997), Quake et al., SCIENCE , Vol. 290, pp. 1536-1540 (Nov.24,2000), all of which are hereby incorporated by reference.

With respect to the use of microfluidic device **200** according to the present invention, as seen in **FIG. 2**, microfluidic device **200** is contacted with the top surface of cell isolation device **10** to provide a substantially fluid-tight seal. Cell are delivered to the cell isolation regions by flowing medium containing the cells desired to be isolated through channels **220** over cell isolation regions **20**. The flowing of the medium containing the cells through channels **220** of microfluidic device **200** may be carried out by a number of mechanisms, including, for example,

pressure based flow, electrokinetic flow, or mechanisms that utilize a hybrid of the two. Microfluidic device **200** may also include integrated microfluidic structures, such as micropumps and microvalves, or external elements, e.g., pumps and switching valves, for the pumping and direction of the medium through device **200**. Medium transport and direction may also be accomplished through electroosmosis or electrokinesis. The selection and details of the above mechanisms would be well within the knowledge of a person skilled in the art.

In some embodiments of the present invention, preferably the flow of the cell containing medium into the cell isolation regions is constant, or continuous. Typically, a known and/or constant flow rate is useful, for example, when attempting to establish precise control over the delivery of cells to cell isolation regions **20**. Furthermore, constant flow facilitates feeding of cells and high throughput screening.

In one embodiment of the method of delivering cells to cell isolation device **10** according to the present invention, microfluidic device **200** is used to deliver about fifty to about hundred cells to each cell isolation region **20**. In another embodiment, microfluidic device **200** is used to deliver about twenty-five to about fifty cells to each cell isolation region **20**. In yet another embodiment, microfluidic device **200** is used to delivery about ten to about twenty-five cells to each cell isolation region **20**. In another embodiment, microfluidic device **200** is used to delivery about five to about ten cells to each cell isolation region **20**. In still another embodiment, microfluidic device is used to delivery about one to about five cells to each cell isolation region **20**. In a preferred embodiment, microfluidic device is used to deliver a single cell to each cell isolation region **20**. Typically, about 90% of each of the cell isolation regions **20** of the cell isolation device **10** are filled when the flow rate is adjusted or regulated. The flow rate by which the cell containing medium is delivered to the cell isolation regions **20** can be adjusted by known methods. A person of skill in the art would know how to adjust the flow rate of a microfluidic device to achieve a desired cell distribution per cell isolation region **20**.

Microfluidic device **200** may be fabricated from materials that are compatible with the conditions present in the particular experiment of interest. Such conditions include, but are not limited to, pH, temperature, ionic concentration, pressure, and application of electrical fields. The materials of device **200** may also be chosen for their inertness to components of the experiment

to be carried out in the device. Such materials include, but are not limited to, glass, silicon, fused silica, metal films, polymeric substrates, such as polystyrene, poly(methylacrylate), polydimethylsiloxane (PDMS) and polycarbonate, depending on the intended application.

Notwithstanding the method of delivery, according to embodiments of the present invention, cells are ultimately delivered to cell isolation device **10** where cells are isolated. As illustrated in **FIG. 4** and **FIG. 7**, cell isolation device **10 (a/b)** utilized in the cell isolation step generally includes a plurality of cell isolation regions **20 (a/b)** for receiving cells therein.

FIGS. 4-6 show one embodiment of a cell isolation device according to the present invention, where the cell isolation device **10a** generally includes a housing **11** defining a plurality of cell isolation regions **20a** into which cells **30** may be delivered. Cell isolation region **20a** presents a recess sized to isolate a single cell **30** or a small group of cells **30**. In one embodiment, cell isolation region **20a** is sized to isolate about fifty to about hundred cells. In another embodiment, cell isolation region **20a** is sized to isolate about twenty-five to about fifty cells. In yet another embodiment, cell isolation region **20a** is sized to isolate about ten to about twenty-five cells. In another embodiment, cell isolation region **20a** is sized to isolate about five to about ten cells. In still another embodiment, cell isolation region **20a** is sized to isolate about one to about five cells. In a preferred embodiment, cell isolation region **20a** is sized to isolate a single cell.

In another embodiment of the cell isolation device, as seen in **FIG. 7**, cell isolation device **10b** is generally defined by a surface **40** defining a plurality of cell isolation regions **20b** upon which cells **30** may be delivered. Preferably, surface **40** is planar. In the embodiment depicted in **FIG. 7**, cell isolation region **20b**, corresponds to a discrete area of surface **40** that has the ability to attract, trap, secure and/or bind a cell **30** or a small group of cells **30** thereon with respect to the non-attractive surrounding surface **40** (such discrete area referred to herein as a "bioaffinity region"). This ability to secure cells **30** to the bioaffinity region may be accomplished through the use of "bioaffinity ligands" as disclosed in more detail herein. With respect to the size of cell isolation region **20b**, in one embodiment, cell isolation region **20b** is sized to isolate about fifty to about hundred cells. In another embodiment, cell isolation region **20b** is sized to isolate about twenty-five to about fifty cells. In yet another embodiment, cell

isolation region **20b** is sized to isolate about ten to about twenty-five cells. In another embodiment, cell isolation region **20b** is sized to isolate about five to about ten cells. In still another embodiment, cell isolation region **20b** is sized to isolate about one to about five cells. In a preferred embodiment, cell isolation region **20b** is sized to isolate a single cell.

Notwithstanding the type of cell isolation region **20** utilized, preferably adjacent cell isolation regions **20** are disposed relative to one another to define a predetermine pitch. More, preferably, the cell isolation regions **20** are disposed relative to one another to match a pitch **P** of an industry standard microtiter plate such as, for example, a 24-, 96-, 384-, 768-, or a 1536-well microtiter plate. The term "pitch" **P** as used herein in reference to the cell isolation device, the cell expansion device, and the cell detection device, refers to the distance between respective vertical centerlines between adjacent cell isolation regions **20** in the test orientation of the particular device being utilized. By "test orientation" of the device is meant to refer to a spatial orientation of the cell isolation device of the particular device being utilized during its respective step. For example, the test orientation of the cell isolation device **10** is the orientation of the device when cells are isolated in their respective cell isolation regions **20**. The test orientation of the cell expansion device is the orientation of the cell expansion device when cells are allowed to proliferate in their respective wells. It is also noted that in the context of the present invention, "top," "bottom," and "lateral" are defined relative to the test orientation of the cell isolation device.

Although the present invention contemplates both the isolation of a single cell **30** and small groups of cells **30**, in certain embodiments it may be preferable to isolate a single cell **30**. Examples of such embodiments include the embodiments of **FIGS. 9A** and **9B**, which respectively show alternative embodiments of cell isolation region **20a**. Referring to **FIGS. 9A** and **9B**, the shown embodiments of cell isolation device **10a**, show cell isolation region **20a** that is defined by a microwell portion **52**. The microwell portion **52** may adjoin accessory delivery portions including a conical portion **51**. Additionally, in the embodiments of **FIGS. 8** and **9A**, another accessory delivery portion, top cylindrical portion **50**, adjoins conical portion **51**, which in turn, adjoins the bottom microwell portion **52**. The dimensions of the accessory delivery portions **51** and **52** and the cell isolation region, microwell portion **52**, may include any

dimensions that allow for cells to be added to the cell isolation regions **20a** *en masse* yet permit only a single cell to settle into microwell portion **52** of cell isolation region **20a**. Preferably, as depicted in **FIG. 9B**, the cylindrical portion **50** (in embodiments including portion **50**) has a diameter d_1 of about 2 millimeters (mm) and a depth D_1 of about 3 mm; the conical portion **51** has a depth D_2 of about 1 mm; and the microwell portion **52** has a diameter d_3 of about 10 microns (μm) to about 50 μm and a depth D_3 of about 10 μm to about 50 μm . In a preferred embodiment, the diameter d_3 of microwell portion **52** is about 20 μm and the depth D_3 of microwell portion is about 20 μm . The cell isolation regions **20a** may also be disposed relative to one another to match a pitch of a standard microtiter plate such as, for example, a 24-, 96-, 384-, 768-, or a 1536-well microtiter plate.

The present invention contemplates several methods of fabricating cell isolation device **10a** wherein cell isolation region **20a** of device **10a** is defined a microwell portion **52** and accessory delivery portions such as conical portion **51** and optionally cylindrical portion **50**. Referring to **FIGS. 10 (A-E)**, various stages of an embodiment of a fabrication method according to the present invention are shown. According to the shown embodiment of the fabrication method, device **10a** may be fabricated by standard photolithographic procedures. According to this procedure, a substrate **70**, as seen in **FIG. 10A**, defining a plurality of conical elements **71** thereon extending from a base **72** is fabricated via standard machining methods. The dimensions of each conical element **71** corresponds to the dimensions of a corresponding conical portion **51** and may be machined directly into base **72** or attached to base **72** by mechanical means, such as by press-fitting, or chemical means, such as by adhesive bonds. Alternatively, conical elements **71** may be fabricated by a multi-level photolithography technique resulting in a quasi-conical topology of sequentially decreasing layers of photoresist as seen in **FIG. 10E**. The final layer **77** of photoresist corresponds to the dimensions of microwell portion **52**. In a preferred embodiment and as illustrated in **FIGS. 10 A-E** and **11**, conical elements **71** are disposed relative to one another to match a pitch of a standard microtiter plate. In an alternative embodiment (not shown), substrate **70** is further defined by a plurality of cylindrical elements. With respect to a singly representative cylindrical element, the cylindrical element is positioned between base **72** and a conical element **71**. The dimensions of the cylindrical elements

correspond to the dimensions of cylindrical portions **50**. In such an embodiment, the substrate **70** may be used as the substrate in subsequent stages of **FIGS. 10A-D** described below.

As illustrated in **FIG. 10B**, in a further step of this fabrication procedure, substrate **70** is coated with a photoresist **73** and as illustrated in **FIG. 10C**, a mask **74** defining regions **75**, the dimensions of which correspond to the dimensions of microwell portion **52**, is placed over photoresist **73**. Ultraviolet (UV) light is then exposed through mask **74** onto photoresist **73** and the exposed photoresist is washed with a developer solution resulting in the formation of conical-microwell master **76** as seen in **FIG. 10D**. Material for the cell isolation device **10a** is then either spin cast, injected, or molded over conical-microwell master **76** and cured, resulting in formation of the conical-microwell member **80** as seen in **FIG. 11**. Photolithographic techniques are well-known to one of skill in the art and as such, substrate **70** materials, photoresist **73** materials, mask **74** materials, developer solutions, the use thereof, including variations of described technique, are well within the knowledge of one of skill in the art.

Referring to **FIG. 11**, in embodiments where it is desired to only have each cell isolation region **20a** defined by microwell portion **52** and to only have conical portion **51** as the accessory cell delivery portion, such as the embodiment of **FIG. 8B**, then conical-microwell member **80** comprises the entire single cell isolation device **10a**. In the alternative, in embodiments where it is desired to have accessory delivery portions, cylindrical portion **50** and conical portion **51**, such as the embodiment of **FIG. 8A**, then a member **81** defining a plurality of cylindrical through-holes **82** may be utilized as illustrated in **FIG. 11**. In this embodiment, member **81** is aligned with member **80** such that when member **81** is contacted with member **80**, cylindrical through-holes **82** are brought into registration with conical elements **71** to form the plurality of accessory delivery portions (cylindrical portion **50** and conical portion **51**) adjoining cell isolation region(microwell portion **52**).

As shown in **FIG. 12A-12C**, another embodiment of a method of fabricating a cell isolation device **10a** having cell isolation region **20a** adjoining accessory delivery portions of is through the use of surface tension driven epoxy molding. According to this embodiment, a substrate **70** similar to the one described in relation to **FIGS. 10A-E** may be used. Here, conical elements **71** of substrate **70** are immersed into a liquid **90** such as, for example, UV-

curable epoxy as shown in **FIG. 12A**. Liquid drops **72** form at the tips of conical elements **71** and these liquid drops **72** are contacted with a support **91** as shown in **FIG. 12B**. Surface tension forces cause the liquid drops **72** to take on a known geometry, such as a cylindrical geometry. It will be recognized by one of skill in the art that the conical elements **71** should be properly aligned and oriented in relation to support **91** to assure uniformity of the liquid cylindrical shape. The liquid **72** is then cured into solid form resulting in the formation of a conical-microwell master **92** as seen in **FIG. 12C**. Material for the cell isolation device **10a** is then either spin cast, injected, or molded over cell isolation region master **92** resulting in the formation a conical-microwell member **80** as shown by way of example in **FIG. 11**, and as further shown in the embodiment of **FIG. 9B**.

As shown in **FIGS. 13A-13C**, an alternative method of fabricating a cell isolation device **10a** according to the present invention, is through the use of epoxy molding in microfabricated wells. This method involves the use of a substrate **70** similar to the one described in relation to **FIGS. 12A-C** and a microwell housing **100** as seen in **FIG. 13A**. Microwell housing **100** may be fabricated of photoresist, or cast in an elastomer such as PIMS. Microwell housing **100** defines a plurality of microwell **101** therein, each of which has dimensions that correspond to the dimensions of microwell portion **52**. Preferably, the microwell **101** are disposed relative to one another to match a pitch of a standard microtiter plate. Microwell **101** are filled with liquid **102**, such as, for example, UV-curable epoxy. If needed, excess liquid can be removed from the areas of microwell housing **100** surrounding microwell **101** by any means known in the art such as, for example, using a "squeegee" or "doctor blade," or razor thereby leaving only liquid in microwell **101**. As shown in **FIG. 13B**, conical elements **71** of substrate **70** and microwell **101** of microwell member **100** are aligned such that conical elements **71** are in registration with their corresponding microwell **101**. Conical elements **71** are then immersed into their respective microwell **101** and the liquid inside microwell **101** is cured into a solid form. Conical elements **71** are then removed from their respective microwell **101** resulting in the formation of conical-microwell master **92** as seen in **FIG. 12C**. Material for the cell isolation device **10a** is then either spin cast, injected, or molded over cell isolation region master **92** to form conical-microwell member **80**.

Materials for cell isolations device **10a** include any rigid or flexible machinable material such as glass, co-polymer or polymer, most preferably urethane, rubber, molded plastic, polymethyl methacrylate (PMMU), polycarbonate, polytetrafluoroethylene (TEFLON), polyvinyl chloride (PVC), polydimethylsiloxane (PIMS), polysulfone, and the like.

Cell isolation device **10a** (whether designed to isolate a single cell or a group of cells) may be exposed to differential surface treatment in order to ensure that cells are isolated in cell isolation region **20a**. For example, the areas of housing **11** surrounding cell isolation regions **20a** may be treated to reduce protein adhesion while cell isolation regions **20a** may be treated to provide a more hydrophilic environment. The hydrophilic nature of cell isolation region **20a** versus the surrounding hydrophobic housing **11** would favor the beading of liquid containing cells **30** in cell isolation region **20a** and potentially hold cells **30** in place, limiting the exhibition of biological activity, such as antibody deposition, to the "wet" areas. Examples of some hydrophobic surface treatments known in the art are teflon, perfluorinated plastic, and polyethylene glycol.

In another embodiment of the cell isolation device **10** according to the present invention, cell isolation device **10b** comprises a surface **40** having a plurality of cell isolation regions **20b** as seen in **FIG. 7** for receiving cells **30** thereon. In particular, cell isolation region **20b** corresponds to a bioaffinity region upon which cells are immobilized. The ability to secure cells **30** to the bioaffinity region according to the embodiment of **FIG. 7** may be accomplished through the use of bioaffinity ligands that are immobilized on the surface of the bioaffinity region and that are capable of binding to cells **30**. The term "bioaffinity ligand" is defined herein to mean any biological or other organic molecule capable of specific or nonspecific binding or interaction with another biological molecule (including biological molecules on the surface of cells). Such binding or interaction may be referred to as "ligand/ligate" binding or interaction and is exemplified by, but not limited to, antibody/antigen, antibody/hapten, enzyme/substrate, enzyme/inhibitor, enzyme/cofactor, binding protein/substrate, carrier protein/substrate, lectin/carbohydrate, receptor/hormone, nucleic acid/nucleic acid, oligonucleotides/nucleic acid, receptor/effector or repressor/inducer bindings or interactions. Examples of bioaffinity ligands used herein, include but are not limited to antibodies, Self-Assembled Monolayers ("SAMs") with biospecific ligands,

lectins (e.g. Ulex europaeus I lectin which binds to the terminal L-fucosyl residues present on the surface of human endothelial cell; peanut agglutinin, agglutinin-I, and phytohaem agglutinin), carbohydrates, antigens, and lipid bilayers. The bioaffinity region of cell isolation region **20b** may be coated with bioaffinity ligands to either specifically or non-specifically bind the plasma membrane of cells **30**.

With respect to non-specific affinity, exemplary bioaffinity ligands having a general non-specific affinity to cells are the lanthanides and in particular, erbium, which is known to bind cell surface glycoproteins as well as calcium-receptors. Another bioaffinity ligand having a general non-specific affinity to cells is ferritin.

In another embodiment, the bioaffinity ligand is an antibody specific for mammalian cells. For example, anti-Ig kappa light chain antibody, anti-CD45R antibody, or anti-syndecan, may be used to differentially bind activated B-cells. Preferably, an antibody specific for anti-Ig kappa light chain antibody cells is used. Any of the methods known in the art for conjugating an antibody to a solid phase support, such as surface **40** described herein, can be used in the present invention.

The use of SAMs provides a preferred method for binding and isolating cells **30** in the bioaffinity region of cell isolation region **20b**. SAMs are the most widely studied and best developed examples of nonbiological, self-assembling systems. They form spontaneously by chemisorption and self-organization of functionalized, long-chain organic molecules onto the surfaces of appropriate substrates. SAMs are usually prepared by immersing a substrate in the solution containing a ligand that is reactive toward the surface, or by exposing the substrate to the vapor of the reactive species. As is well-known to one of skill in the art, there are many systems known in the art to produce SAMs. In certain embodiments, it may be desirable to pattern the SAMs to have an arrayed surface. Patterning SAMs on a planar surface has been achieved by a wide variety of techniques, including micro-contact printing, photo-oxidation, photo-cross-linking, photo-activation, photolithography/plating, electron beam writing, focused ion beam writing, neutral metastable atom writing, SPM lithography, micro-machining, micro-pen writing. A preferred method is micro-contact printing is described in U.S. Patent 5,776,748 and is herein incorporated by reference in its entirety.

With respect to the dimensions of cell isolation region **20b** according to the embodiment of **FIG. 7**, cell isolation regions **20b** of cell delivery device **10b** may be between 10 μ m to 30 μ m in diameter and preferably between about 10 μ m to about 15 μ m in diameter and may be disposed relative to one another to match a pitch of a standard microtiter plate.

As with cell isolation device **10a**, cell isolation device **10b** may be exposed to differential surface treatment in order to ensure that cells are isolated in cell isolation region **20b**. For example, the areas of surface **40** surrounding cell isolation regions **20b** may be treated, for example, with a hydrophobic surface treatment, to reduce protein adhesion. The bioaffinity region of cell isolation region **20a** and the surrounding hydrophobic area of surface **40** would favor the binding of cells **30** on cell isolation region **20b**. Examples of some hydrophobic surface treatments known in the art are teflon, perfluorinated plastic, and polyethylene glycol.

Although the description of the foregoing embodiments has focused on the use of bioaffinity ligands to bind cells **30** onto cell isolation region **20b** of cell isolation device **10b**, it is also contemplated by the present invention that bioaffinity ligands may be immobilized on the bottom surface of cell isolation regions **20a** of cell isolation device **10a** to potentially bind cells **30**.

With respect to surface **40** of cell delivery device **10b**, surface **40** may be fabricated of any material capable of having bioaffinity ligands immobilized therein and such materials are readily known to one of skill in the art. Preferably, surface **40** is fabricated of any rigid or flexible machinable material such as glass, co-polymer or polymer, most preferably urethanes, rubber, molded plastic, polymethylmethacrylate (PMMA), polycarbonate, polytetrafluoroethylene (TEFLONTM), polyvinylchloride (PVC), polydimethylsiloxane (PDMS), polysulfone, and the like. Such substrates are readily manufactured from microfabricated masters, using well known molding techniques, such as injection molding, embossing or stamping, or by polymerizing the polymeric precursor material within a mold. Standard soft lithography techniques may also be used to fabricate surface **40** (see e.g., Love, et al., MRS BULLETIN, pp.523-527 (July 2001) "Fabrication of Three-Dimensional Microfluidic Systems by Soft Lithography," Delamarche et al., JOURNAL OF AMERICAN CHEMICAL SOCIETY, Vol. 120, pp.500-508 (1998), Delamarche et al., SCIENCE, Vol.276, pp.779-781 (May 1997),

Quake et al., SCIENCE , Vol. 290, pp. 1536-1540 (Nov.24,2000), U.S. Patent 6,090,251, all of which are hereby incorporated by reference). Such surface materials are preferred for their ease of manufacture, low cost and disposability, as well as their general inertness to most extreme reaction conditions. These materials may include treated surfaces, such as, derivatized or coated surfaces, to enhance their utility in the fluidic, preferably microfluidic, system, to provide enhanced fluid direction (*See e.g.*, U.S. Patent No. 6,238,538, and which is incorporated herein by reference).

Once cells **30** are isolated in cell isolation regions **20** of cell isolation device **10**, another aspect of the invention allows for the proliferation of the cells **30** through the use of a cell expansion device. As illustrated in **FIG. 14**, the cell expansion device **120** comprises a structure **121** defining a plurality of wells **122** with adjacent wells having the same pitch as the pitch defined by adjacent cell isolation regions of the cell isolation device **10**. In particular, each well **122** in a test orientation of the cell expansion device **120**, encompasses a cavity surrounded by two lateral sides connected to a bottom side. Preferably, each well is also characterized by having a greater volume than respective ones of the cell isolation region **20**. "Wells" as used herein are specific to the cell expansion device, whereas cell isolation regions as used herein are specific to the cell isolation device. In general as seen in **FIG. 14**, cell expansion device **120** is utilized by orienting structure **121** with housing **11** of cell isolation device **10** such that wells **122** overlie corresponding ones of the cell isolation regions **20**. Structure **121** is then placed in direct contact with housing **11** to form a seal such that fluid communication between cells **30** in adjacent cell isolation regions is inhibited. As seen in **FIG. 15A**, the mated cell isolation device/cell expansion device is then inverted and cells **30** are transferred, for example by centrifugal force, from cell isolation regions **20** to wells **122**.

Wells **122** may be of any shape, but wells with circular or square shaped top plan view (or transverse cross-sections) are preferred as these shapes are commonly used in the industry. Notwithstanding the shape, wells **122** of expansion device **120** preferably have a greater volume than cell isolation regions **20** such as, for example, having a greater diameter (as illustrated in **FIG. 15A**) or a greater depth (as illustrated in **FIG. 15B**). Preferably wells **122** have the same diameter as cell isolation regions **20** of cell delivery device **10**. More preferably,

the wells **122** of the expansion device **120** have a depth of about 2mm. In one embodiment, the lateral surfaces of well **122** are canted relative to one another or to the bottom surface of the well **122** in a test orientation of cell expansion device **120** as shown in **FIG. 16**. Canted wells **122** permit easier access by allowing for lateral movement when performing downstream experiments on cells **30** in wells **122** and may create a more uniform interface with a small cell isolation region **20**. On the other hand, a well **122** that has lateral sides that are at a 90° angle to the bottom side of the well **122** in a test orientation of the cell expansion device **120** provide a greater volume, which may increase the time available for the detection step to be described below.

Structure **121** is intended to define any number of wells **122**, but preferably defines the number of wells in a standard microtiter plate such as a 24-, 96-, 384-, 768-, or 1536-well plate. More preferably, structure **121** defines the number of wells of a 385-well plate or 1536-well plate. Approximately 2 µl of liquid can be held per well **122** in a standard 1536 well plate. Larger well volumes can be achieved by extending the depth of the well **122** without compromising the correspondence between the pitch between adjacent wells **122** and the pitch between adjacent cell isolation regions **20**.

In another embodiment of cell expansion device **120** as seen in **FIG. 17**, cell expansion device **120a** is configured such that the respective wells **122a** are accessible from a bottom side **127** thereof in a test orientation of cell expansion device **120a**. Structure **123a** further defines an entrance port **125** and optionally, an exit port **126**. The wells' **122a** accessibility from the bottom side **127** permits exchange of media, thereby facilitating cell expansion and refeeding. Cells **30** are retained from passing through the access to the bottom side **127** of well **122a** by a block such as a size constraint or semi-permeable membrane, such as a dialysis membrane, nitrocellulose or perforated polydimethylsialoxane, for example. The semi-permeable membrane of this embodiment also allows for gas exchange while reducing the evaporation of liquid contained in each well **122a**. The initial flow-through of old medium may be drained by positive pressure, vacuum, or gravity to capture liquid potentially containing antibody or another product of the desired biological activity, on a surface designed for sampling/detection, e.g., nitrocellulose, glass, urethanes, rubber, molded plastic, or polydimethylsialoxane.

Preferably, the material used for the manufacture of cell expansion device **120** comprises any rigid or flexible material such as glass, urethanes, rubber, molded plastic, co-polymer or polymer, more preferably urethanes, rubber, molded plastic, polymethylmethacrylate (PMMA), polycarbonate, polytetrafluoroethylene (TEFLON™), polyvinylchloride (PVC), polydimethylsiloxane (PDMS), polysulfone, and the like, and most preferably PDMS. Such materials are readily manufactured from fabricated masters, using well known molding techniques, such as injection molding, embossing or stamping, or by polymerizing the polymeric precursor material within the mold. Such materials are preferred for their ease of manufacture, low cost and disposability, as well as their general inertness to most extreme reaction conditions.

Once the cells **30** are transferred from cell isolation device **10** to cell expansion device **122**, the cells **30** are incubated in the cell expansion device **122**, for a sufficient amount of time to allow for proliferation of cells **30**. Cell expansion device **122** is intended to be used at any biologically viable temperature. Lowering the incubation temperature of the cells (i.e., from 37° C to 18° C) may, however, slow the metabolic processes of the cells **30** and reduce cell cloning time, thus extending the time for assay. Alternatively, media that is optimal for exhibiting the desired biological activity that is to be screened but not optimal for cell proliferation could also be used to extend the time for assay.

As illustrated in **FIG. 16**, after sufficient time has been allowed for cells **30** to grow and divide, a detection device **140** is utilized to detect the desired biological activity exhibited by the cells, such as antibody production. In the case of detecting antibody production, in one embodiment as illustrated in **FIG. 16**, detection device **140** generally comprises a surface **141** defining an array of prongs **142** upon which antigens are immobilized. In a preferred embodiment, surface **141** is a substantially planar surface. In one embodiment, prongs **142** are coated with the specific antigen(s) used to immunize animals from which the cells being tested are derived. In use, detection device **140** is placed over structure **121** of cell expansion device **120** such that prongs **142** are in registration with wells **122**. Prongs **142** are then immersed into wells **122** of cell expansion device **120** in order to allow antigens immobilized on prongs **142** to potentially bind to antibodies secreted/produced by cells, such as hybridoma cells in wells **120**. Prongs **142** of detection device **140** are then removed from well **122** of cell expansion device

120 and then immersed into a solution of detectable secondary antibody. Secondary antibodies may be visualized by standard techniques known in the art such as by enzymatic color reactions or fluorescence. A prong surface may also be interfaced with mass spectrometer for additional assay flexibility. Other methods of labeling and detecting the antibodies are well within the knowledge of one of skill in the art and therefore are not discussed in detail herein.

The antibodies may be coated on prongs **142** through non-specific physisorption or through the use of SAMs on prongs **142**. For example, a particular antigen may be specifically patterned on prong **142** by first evaporating gold on a polyurethane prong, then adding a self-assembling monolayer onto which an antigen can be covalently linked. (See, e.g., Mrksich et al., *Annu. Rev. Biophys. Biomol. Struct.*, vol. 25 (1996) incorporated herein by reference). Additionally, other strategies for immobilizing protein may be employed such as lysine linkage, cutinase linkage, GST-fusion or GPI linkage.

In another embodiment of a detection device used to detect the production of antibodies from cells, such as, for example, hybridoma cells, cell isolation device **10** additionally serves as a detection device as shown in **FIG. 18**. In this embodiment, the bottoms of cell isolation regions **20** of cell isolation device **10** are coated with the specific antigen(s) used to immunize animals from which the hybridomas being tested are derived. If the desired biological activity being screened is the production of a biological product, other than antibodies, the bottoms of cell isolation regions **20** could be coated with a ligand that binds to the biological product (ligate) produced by the desired biological activity that is being screened. Ligands include, for example, a peptide, hormone, enzyme, or carbohydrate. Cells **30** are then delivered to cell isolation regions **20** and the cells **30** will begin to produce antibodies that will bind the antigens coated on cell isolation regions **20**. Cells **30** are then transferred from the cell isolation regions **20** of cell isolation device **10** to the wells **122** of cell expansion device **120**. The antibodies bound to the antigens on the bottoms of cell isolation regions **20** are then detected using an ELISA sandwich or a variety of detection methods known in the art such as secondary antibody coupled to alkaline phosphatase, horse radish peroxidase, etc.

Although in the embodiment illustrated in **FIG. 16**, cell isolation region **20** of the cell isolation device **10** actually becomes the detection device, it is well within the scope of the

present invention to utilize a separate detection device. In such an embodiment, the detection device comprises a structure defining a plurality of indentations sized to receive cells from cell expansion device **120**, wherein the bottom of the indentations are coated with a ligand that binds to the biological product (ligate) produced by the desired biological activity that is being screened. Such ligates include, for example, a protein, enzyme, peptide, carbohydrate or antibody. After cells are transferred from cell isolation device **10** to the cell expansion device **120**, cells are, in turn, transferred to the indentations of the cell detection device where they will potentially express the biological product of the desired biological activity that is being screened and such biological product binds its complementary binding pair member coated on the indentations.

Although the foregoing description of detection methods have been described in reference to production of a biological product produced by the desired biological activity being screened, the present invention contemplates the detection of any biological activity by any types of cells. Such detection methods are well known to one of skill in the art.

It will be appreciated that the present disclosure is intended to set forth the exemplifications of the invention, and the exemplifications set forth are not intended to limit the invention to the specific embodiments illustrated. The disclosure is intended to cover, by the appended claims, all such modifications as fall within the spirit and scope of the claims.

We Claim:

1. A method of screening cells for a desired biological activity comprising:
 - providing a cell isolation device defining a plurality of cell isolation regions, each cell isolation region being sized for isolating about one to about five cells therein, each cell isolation region encompassing a bioaffinity region, the cell isolation regions further defining a predetermined pitch with respect to one another;
 - delivering the about one cell to about five cells to each of the cell isolation regions;
 - providing a cell expansion device defining a plurality of wells corresponding to respective ones of the plurality of cell isolation regions and defining a predetermined pitch that matches the predetermined pitch of the cell isolation regions;
 - transferring the about one to about five cells from the cell isolation regions to the wells;
 - allowing about one cell to about five cells to proliferate and exhibit a desired biological activity; and
 - assaying the biological activity the about one to about five cells.
2. The method of claim 1, wherein delivering comprises delivering about one cells to about five cells through a microfluidic channel.
3. The method of claim 2, wherein the channel has a diameter of about 100 microns.

4. The method of claim 1, wherein each of the bioaffinity regions comprises bioaffinity ligands immobilized therein.
5. The method of claim 4, wherein the bioaffinity ligands are selected from the group consisting of antibodies, antigens, self-assembled monolayers (SAMs), lectins, carbohydrates, transmembrane proteins, and transmembrane protein receptors.
6. The method of claim 1, wherein the predetermined pitch of the cell isolation regions matches a pitch of a standard microtiter plate.
7. The method of claim 1, wherein each of the cell isolation regions has a diameter of about 10 to about 30 microns.
8. The method of claim 1, wherein transferring comprises transferring about one cell to about five cells from the cell isolation device to the cell expansion device by centrifugal force.
9. The method of claim 1, wherein the cells are hybridoma cells and the biological activity exhibited by the cells is antibody production.
10. The method of claim 1, wherein assaying comprises assaying hybridoma cells for specific antibody production by:
 - providing a detection device comprising a member defining a plurality of prongs, the prongs coated with specific antigens;
 - immersing the prongs into the wells to allow potential binding between specific antibodies produced by the hybridoma cells and the antigens;
 - removing the prongs from the wells;
 - immersing the prongs into a detection solution;

detecting the presence of specific antibodies.

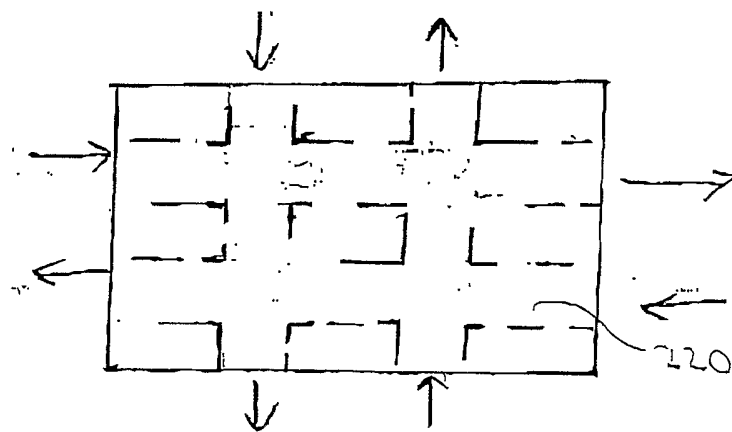
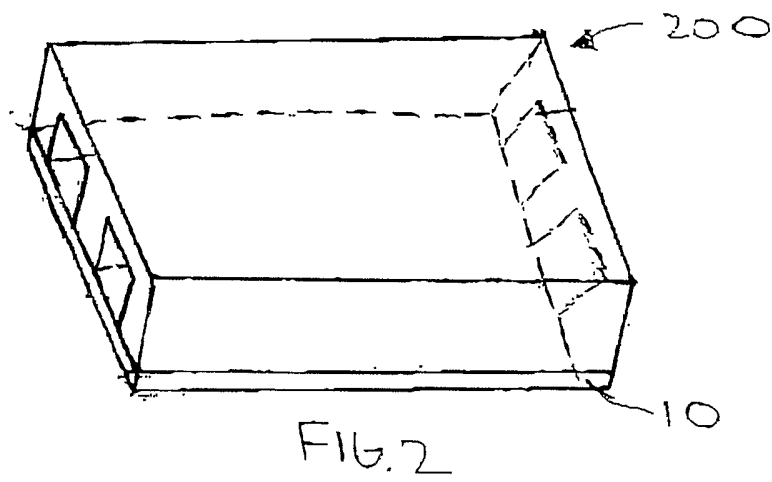
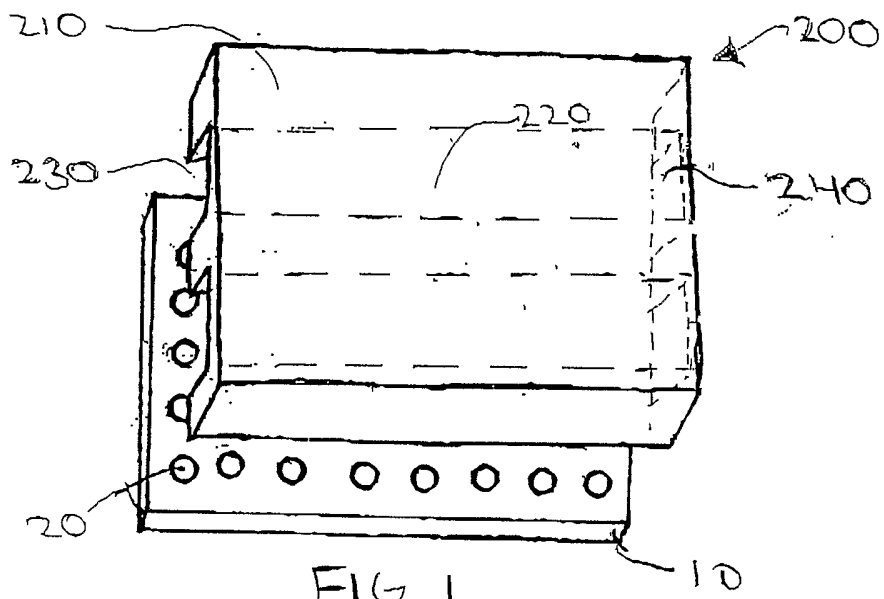
11. A kit for screening cells for a desired biological activity, the kit comprising:
 - a cell isolation device defining a plurality of cell isolation regions, each of the cell isolation regions being sized to isolate about one cell to about five cells therein, and each encompassing a bioaffinity region, the cell isolation device further defining a predetermined pitch with respect to one another;
 - a cell expansion device defining a plurality of wells corresponding to respective ones of the plurality of cell isolation regions;
12. The kit of claim 11, further comprising a detection device for screening the about one to about five cells for the desired biological activity.
13. The kit of claim 11, wherein the bioaffinity regions comprise bioaffinity ligands immobilized therein.
14. The kit of claim 12, wherein the bioaffinity ligands are selected from the group consisting of antibodies, antigens, self-assembled monolayer (SAMs), lectins, carbohydrates, transmembrane proteins, and protein receptors.
15. The kit of claim 11, wherein the surface of the cell isolation device has a substantially planar surface and wherein the bioaffinity regions comprise respective discrete areas of the substantially planar surface.
16. The kit of claim 11, wherein each of the cell isolation regions has a diameter of about 10 microns to about 30 microns in diameter.

17. The kit of claim 11, wherein the cell expansion device comprises lateral sides partially defining respective ones of the plurality of wells, the lateral sides of each of the wells being canted relative to one another.
18. The device of claim 11, wherein the predetermined pitch of the cell isolation regions matches a pitch of a standard microtiter plate.
19. A method of screening cells for a desired biological activity comprising:
 - providing a cell isolation device defining a plurality of cell isolation regions, each cell isolation region encompassing a recess, each of the cell isolation regions being sized to isolate about one to about five cells therein, and the cell isolation regions further defining a predetermined pitch with respect to one another;
 - delivering the about one cell to about five cells to each of the cell isolation regions;
 - providing a cell expansion device defining a plurality of wells corresponding to respective ones of the plurality of cell isolation regions, the wells defining a predetermined pitch matching the predetermined pitch of the cell isolation regions;
 - transferring the about one cell to about five cells from the cell isolation regions to the wells;
 - allowing the about one cell to about five cells to proliferate and exhibit a desired biological activity in the wells;
 - assaying the desired biological activity of the one cell to about five cells.
20. The method of claim 19, wherein delivering comprises delivering about one cell to about five cells through a microfluidic channel.

21. The method of claim 19, wherein the channel has a diameter of about 100 microns.
22. The method of claim 19, wherein the recess of each of the cell isolation regions defines a top conical portion, and a bottom microwell portion adjoining the top conical portion in a test orientation of the cell isolation device.
23. The method of claim 19, wherein the recess of each of the cell isolation regions defines a top cylindrical portion, an intermediate conical portion, and a bottom microwell portion, the intermediate conical portion being disposed between and adjoining the top cylindrical portion and the bottom microwell portion.
24. The method of claim 19, wherein bioaffinity ligands are immobilized in the cell isolation regions.
25. The method of claim 24, wherein the bioaffinity ligands are selected from the group consisting of antibodies, self-assembled monolayers (SAMs), lectin, carbohydrate, transmembrane proteins, and antigens.
26. The method of claim 19, wherein the predetermine pitch of the cell isolation regions and the predetermined pitch of the wells matches a pitch of a standard microtiter plate.
27. The method of claim 19, wherein the cells are transferred from the cell isolation device to the cell expansion device by centrifugal force.
28. The method of claim 19, wherein the cells are hybridoma cells and the biological activity exhibited by the cells is antibody production.

29. The method of claim 28, wherein assaying comprises assaying the hybridoma cells for specific antibody production by:
- providing a detection device comprising a member defining a plurality of prongs, the prongs being coated with specific antigens;
 - immersing the prongs into the wells to allow potential binding between specific antibodies produced by the hybridoma cells and the antigens;
 - removing the prongs from the wells;
 - immersing the prongs into a detection solution;
 - detecting the presence of specific antibodies.
30. A kit for screening cells for a desired biological activity, the kit comprising:
- a cell isolation device defining a plurality of cell isolation regions, each cell isolation region encompassing a recess, each of the cell isolation regions being sized to isolate about one to about five cells therein, and the cell isolation regions further defining a predetermined pitch with respect to one another;
 - a cell expansion device defining a plurality of wells corresponding to respective ones of the plurality of cell isolation regions, and the wells defining a predetermined pitch matching the predetermined pitch of the cell isolation regions.
31. The kit of claim 30, further comprising a detection device for screening the cells for the desired biological activity.
32. The kit of claim 30, wherein the recess of each of the cell isolation regions defines a top conical portion, and a bottom microwell portion adjoining the top conical portion in a test orientation of the cell isolation device.
33. The kit of claim 32, wherein the top conical portion has a depth of about 1 millimeter.

34. The kit of claim 32, wherein the bottom microwell portion has a diameter of about 10 microns to about 50 microns and a depth of about 10 microns to about 50 microns.
35. The kit of claim 34, wherein the bottom microwell portion has a diameter of about 20 microns and a depth of about 20 microns.
36. The kit of claim 30, wherein the recess of each of the cell isolation regions defines a top cylindrical portion, an intermediate conical portion, and a bottom microwell portion, the intermediate conical portion being disposed between and adjoining the top cylindrical portion and the bottom microwell portion.
37. The kit of claim 36, wherein the cylindrical portion has a diameter of about 2 millimeters and a depth of about 3 millimeters.
38. The kit of claim 36, wherein the conical portion has a depth of about 1 millimeters.
39. The kit of claim 36, wherein the bottom microwell portion has a diameter of about
10 microns to about 50 microns and a depth of about 10 microns to about 50 microns.
40. The kit of claim 30, wherein the pitch of the cell isolation regions and the pitch of the wells match a pitch of a standard microtiter plate.



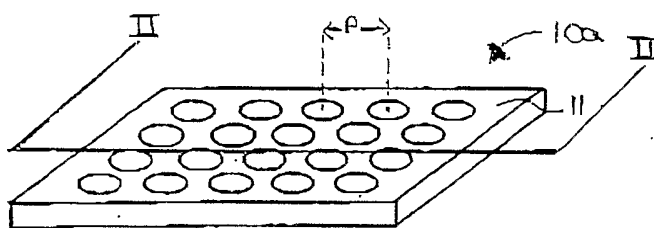


FIG 4

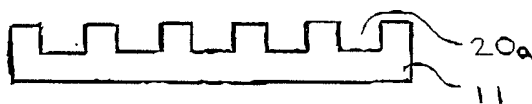


FIG 5

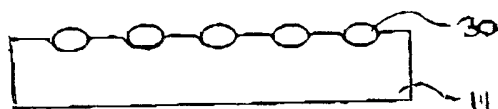


FIG 6

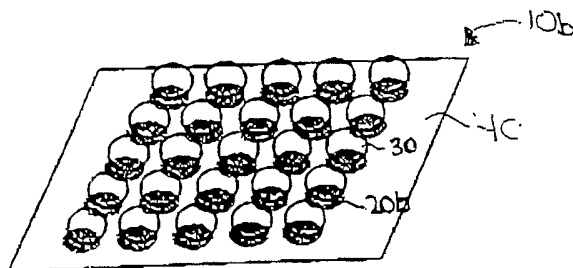


FIG 7

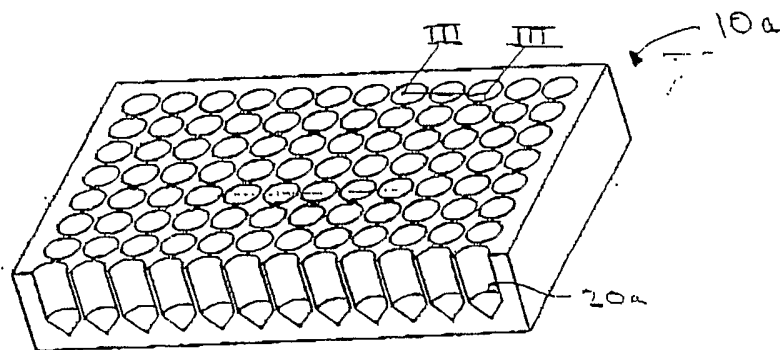


FIG 8

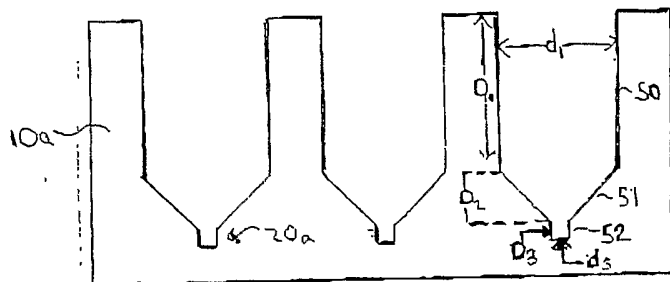


FIG 9A

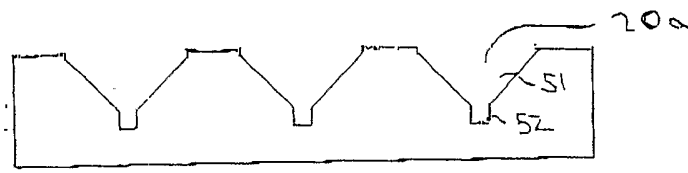


FIG. 9B

FIG. 10A



FIG. 10B

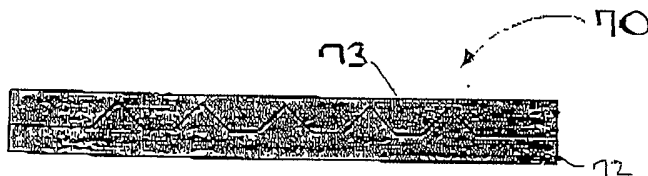


FIG. 10C

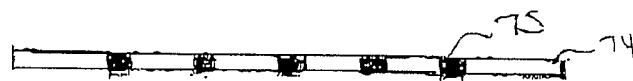


FIG. 10D



FIG. 10E

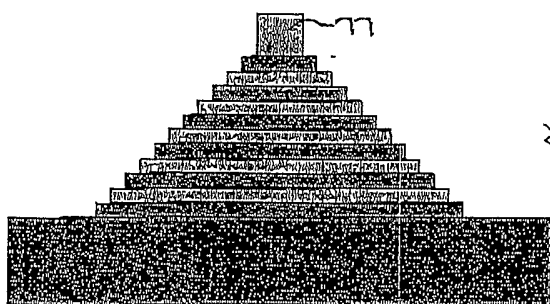


FIG. 10E

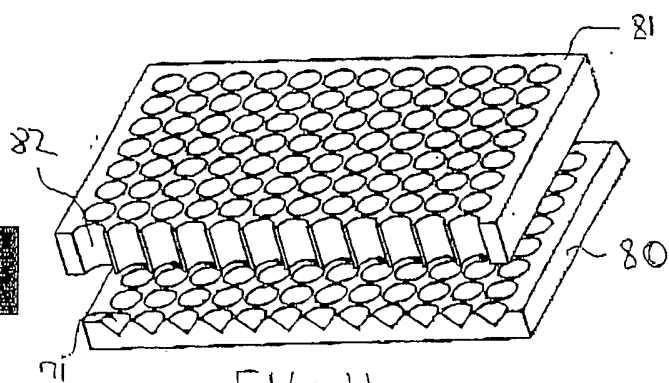
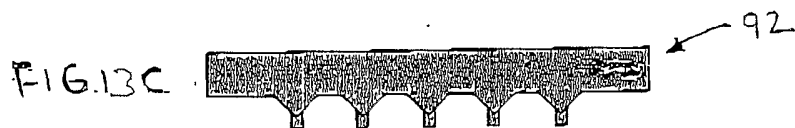
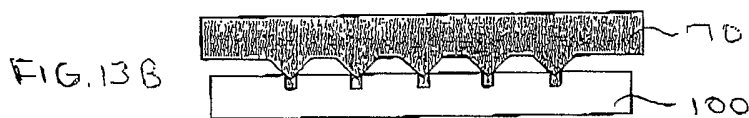
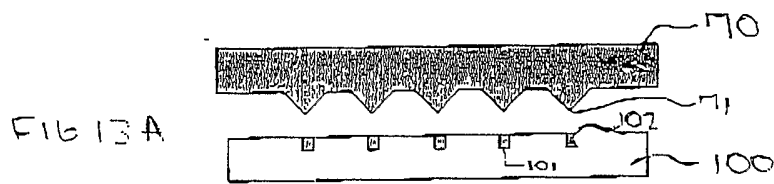
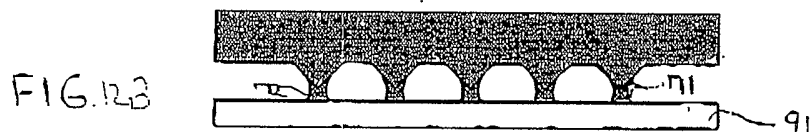
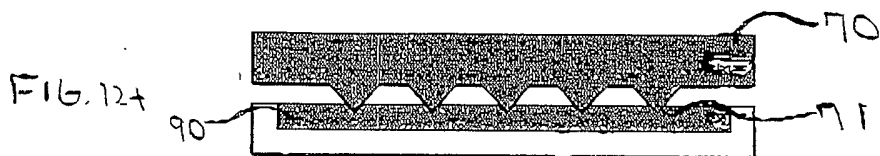


FIG. 11



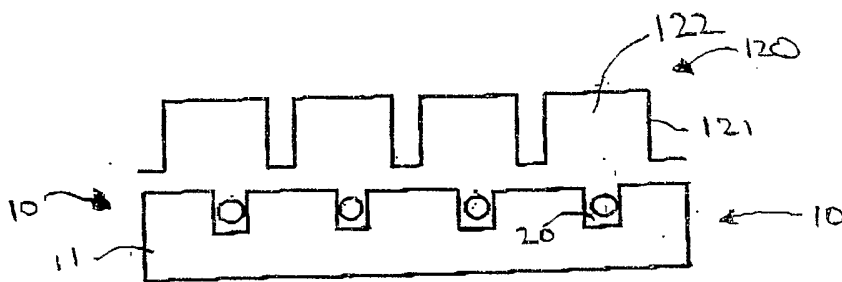


FIG. 14

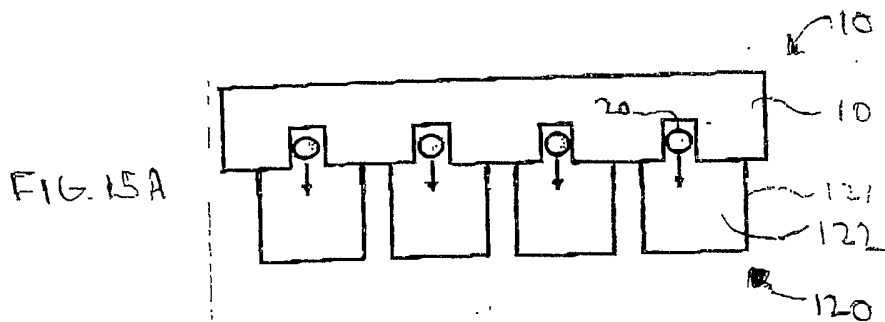


FIG. 15A

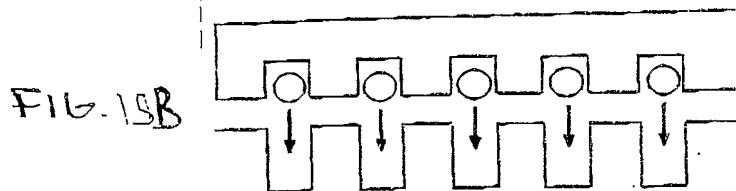


FIG. 15B

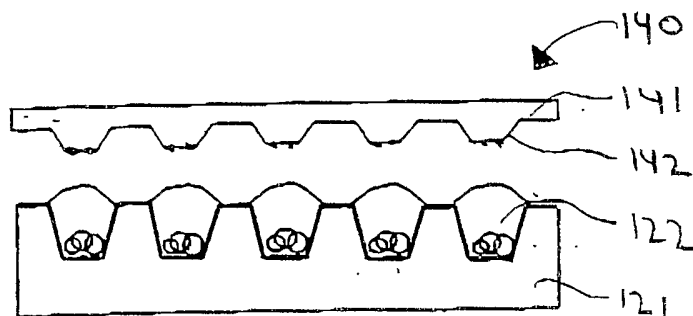


FIG. 16

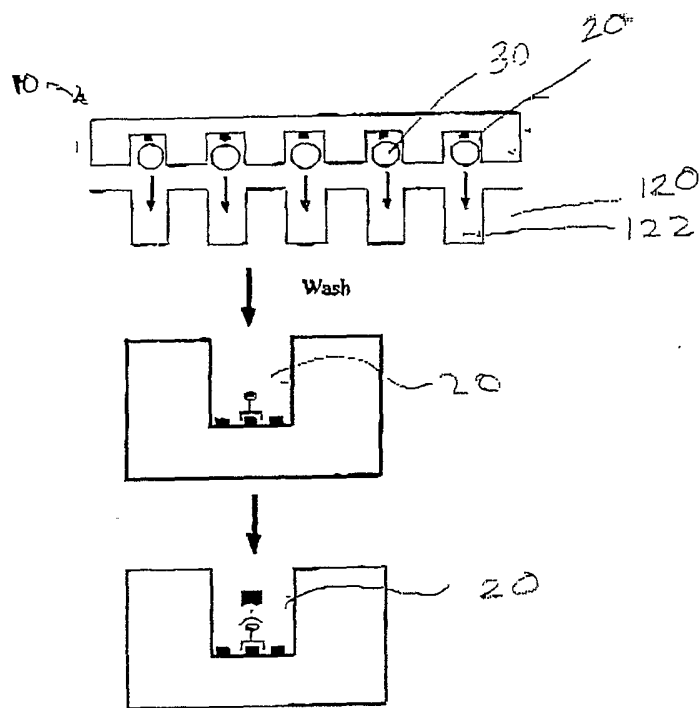


FIG. 18

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US02/24068

<p>A. CLASSIFICATION OF SUBJECT MATTER IPC(7) : B01J 19/00; B01L 3/00; C12M1/34; G01N 15/14, 33/53 US CL : 204/454; 422/ 68.1, 82.05; 435/7.1; 436/180,501, 518, 536, 807-809 According to International Patent Classification (IPC) or to both national classification and IPC</p>																																						
<p>B. FIELDS SEARCHED</p> <p>Minimum documentation searched (classification system followed by classification symbols) U.S. : 204/454; 422/ 68.1, 82.05; 435/7.1; 436/180,501, 518, 536, 807-809</p> <p>Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched</p> <p>Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) Please See Continuation Sheet</p>																																						
<p>C. DOCUMENTS CONSIDERED TO BE RELEVANT</p> <table border="1"> <thead> <tr> <th>Category *</th> <th>Citation of document, with indication, where appropriate, of the relevant passages</th> <th>Relevant to claim No.</th> </tr> </thead> <tbody> <tr> <td>X</td> <td>WO 00/17624 A2 (CELLOMICS, INC) 30 March 2000, see entire document.</td> <td>1-40</td> </tr> <tr> <td>X</td> <td>US 6,083,761 A (KEDAR et al.) 04 July 2000, see entire document.</td> <td>11-12, 14-18, 30-40</td> </tr> <tr> <td>---</td> <td></td> <td>-----</td> </tr> <tr> <td>Y</td> <td></td> <td>1-40</td> </tr> <tr> <td>X</td> <td>US 5,603,351 A (CHERUKURI et al.) 18 February 1997, see entire document.</td> <td>11-12,14-18,30-31</td> </tr> <tr> <td>---</td> <td></td> <td>-----</td> </tr> <tr> <td>Y</td> <td></td> <td>1-40</td> </tr> <tr> <td>X</td> <td>US 6,167,910 B1 (CHOW) 02 January 2001, see entire document.</td> <td>11-12,14-18,30-31</td> </tr> <tr> <td>---</td> <td></td> <td>-----</td> </tr> <tr> <td>Y</td> <td></td> <td>1-40</td> </tr> <tr> <td>Y</td> <td>DELAMARCHE et al., "Microfluidic Networks for Chemical Patterning of Substrates: Design and Application to Bioassays", J. Am. Chem. Soc., Vol. 120, No. 3 (1998), pages 500-508, see entire article.</td> <td>1-40</td> </tr> </tbody> </table>			Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.	X	WO 00/17624 A2 (CELLOMICS, INC) 30 March 2000, see entire document.	1-40	X	US 6,083,761 A (KEDAR et al.) 04 July 2000, see entire document.	11-12, 14-18, 30-40	---		-----	Y		1-40	X	US 5,603,351 A (CHERUKURI et al.) 18 February 1997, see entire document.	11-12,14-18,30-31	---		-----	Y		1-40	X	US 6,167,910 B1 (CHOW) 02 January 2001, see entire document.	11-12,14-18,30-31	---		-----	Y		1-40	Y	DELAMARCHE et al., "Microfluidic Networks for Chemical Patterning of Substrates: Design and Application to Bioassays", J. Am. Chem. Soc., Vol. 120, No. 3 (1998), pages 500-508, see entire article.	1-40
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<p><input checked="" type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.</p>																																						
<p>* Special categories of cited documents:</p> <table border="0"> <tr> <td>"A" document defining the general state of the art which is not considered to be of particular relevance</td> <td>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</td> </tr> <tr> <td>"E" earlier application or patent published on or after the international filing date</td> <td>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone</td> </tr> <tr> <td>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</td> <td>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art .</td> </tr> <tr> <td>"O" document referring to an oral disclosure, use, exhibition or other means</td> <td>"&" document member of the same patent family</td> </tr> <tr> <td>"P" document published prior to the international filing date but later than the priority date claimed</td> <td></td> </tr> </table>			"A" document defining the general state of the art which is not considered to be of particular relevance	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention	"E" earlier application or patent published on or after the international filing date	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone	"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art .	"O" document referring to an oral disclosure, use, exhibition or other means	"&" document member of the same patent family	"P" document published prior to the international filing date but later than the priority date claimed																											
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<p>Date of the actual completion of the international search 30 October 2002 (30.10.2002)</p>		<p>Date of mailing of the international search report 04 DEC 2002</p>																																				
<p>Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230</p>		<p>Authorized officer <i>Valerie Bell-Harris for</i> Bennett Celsa Telephone No. (703) 308-0196</p>																																				

INTERNATIONAL SEARCH REPORT

PCT/US02/24068

Continuation of B. FIELDS SEARCHED Item 3:
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STN: CAPLUS, BIOSIS, MEDLINE, WPIDS, EMBASE