



(86) Date de dépôt PCT/PCT Filing Date: 1996/06/06
(87) Date publication PCT/PCT Publication Date: 1996/12/19
(45) Date de délivrance/Issue Date: 2007/03/20
(85) Entrée phase nationale/National Entry: 1997/12/02
(86) N° demande PCT/PCT Application No.: US 1996/009294
(87) N° publication PCT/PCT Publication No.: 1996/040795
(30) Priorité/Priority: 1995/06/07 (US08/481,883)

(51) Cl.Int./Int.Cl. *C08B 37/00* (2006.01),
A61K 39/08 (2006.01), *A61K 39/09* (2006.01),
A61K 39/116 (2006.01), *A61K 39/385* (2006.01),
A61K 47/48 (2006.01), *C07K 16/12* (2006.01),
C07K 9/00 (2006.01), *G01N 33/569* (2006.01)

(72) Inventeurs/Inventors:
MICHON, FRANCIS, US;
UITZ, CATHERINE, US;
TAI, JOSEPH Y., US

(73) Propriétaire/Owner:
BAXTER HEALTHCARE S.A., CH

(74) Agent: SMART & BIGGAR

(54) Titre : FRAGMENTS ANTIGENIQUES DE POLYSACCHARIDES DE STREPTOCOQUES DE TYPES II ET III DU GROUPE B, PRESENTANT UNE STRUCTURE TERMINALE 2,5-ANHYDRO-D-MANNOSE ET VACCINS CONJUGUES A BASE DESDITS FRAGMENTS

(54) Title: ANTIGENIC GROUP B STREPTOCOCCUS TYPE II AND TYPE III POLYSACCHARIDE FRAGMENTS HAVING A 2,5-ANHYDRO-D-MANNOSE TERMINAL STRUCTURE AND CONJUGATE VACCINE THEREOF

(57) Abrégé/Abstract:

The process for depolymerizing Group B Types II and III Streptococcus is disclosed which results in polysaccharide fragments having a reducing end suitable for conjugating to protein. Conjugate molecules, vaccines and their use to immunize mammals including humans are also disclosed.



PCTWORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C08B 37/00, A61K 39/09, 39/385	A1	(11) International Publication Number: WO 96/40795 (43) International Publication Date: 19 December 1996 (19.12.96)
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(21) International Application Number: PCT/US96/09294

(22) International Filing Date: 6 June 1996 (06.06.96)

(30) Priority Data:
08/481,883 7 June 1995 (07.06.95) US(71) Applicant: NORTH AMERICAN VACCINE, INC. [CA/US];
12103 Indian Creek Court, Beltsville, MD 20705 (US).(72) Inventors: MICHON, Francis; 9735 Country Meadows Lane,
Laurel, MD 20723 (US). CATHERINE, Dong; 4126 North
34th Street, Arlington, VA 22207 (US). JOSEPH, Y., Tai;
1370 Cinnamon Drive, Fort Washington, PA 19034 (US).(74) Agent: SONNENFELD, Kenneth, H.; Morgan & Finnegan,
L.L.P., 345 Park Avenue, New York, NY 10154 (US).(81) Designated States: AL, AM, AT, AU, AZ, BB, BG, BR, BY,
CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS,
JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD,
MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD,
SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, UZ, VN, ARIPO
patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM,
AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT,
BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC,
NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA,
GN, ML, MR, NE, SN, TD, TG).**Published***With international search report.*(54) Title: ANTIGENIC GROUP B STREPTOCOCCUS TYPE II AND TYPE III POLYSACCHARIDE FRAGMENTS HAVING A 2,
5-ANHYDRO-D-MANNOSE TERMINAL STRUCTURE AND CONJUGATE VACCINE THEREOF

(57) Abstract

The process for depolymerizing Group B Types II and III Streptococcus is disclosed which results in polysaccharide fragments having a reducing end suitable for conjugating to protein. Conjugate molecules, vaccines and their use to immunize mammals including humans are also disclosed.

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**ANTIGENIC GROUP B STREPTOCOCCUS TYPE II AND TYPE III
POLYSACCHARIDE FRAGMENTS HAVING A 2, 5-ANHYDRO-D-MANNOSE
TERMINAL STRUCTURE AND CONJUGATE VACCINE THEREOF**

FIELD OF THE INVENTION

This invention relates to antigenic capsular polysaccharide fragments useful for conjugating to a protein to create immunogens which elicit protective antibodies. More specifically, the invention relates to Group B Streptococcus capsular polysaccharides (GBS CP) with analyzable reducing-end groups, their preparation, and their use to make conjugate vaccines.

BACKGROUND OF THE INVENTION

GBS bacteria are a recognized etiological agent for bacteremia and/or meningitis in infants, and for infections in adults. Baker, "Group B Streptococcal Infections" in Advances in Internal Medicine, 25:475-500 (1980). Accordingly, it is important to develop rapid and definitive assays for diagnosis of GBS infection, and methods of generating protection against GBS, particularly in infants and compromised individuals.

The capsular polysaccharides from GBS bacteria are known to be important to GBS virulence and the development of protective immunity. See Kasper et al. U.S. Patent 5,302,386. Moreover, the CP of recognized GBS types (I-V) are chemically related but antigenically distinct having repeating structures composed of galactose, glucose, N-acetyl glucosamine, and N-acetyl-neuraminic (sialic) acid.

Infants and young children have poor immunogenic response to polysaccharide antigens. These responses are characterized as being T cell independent and therefore are not associated with important attributes such as memory, isotype switching, or affinity maturation, which are necessary for conferring long term immunologic protection against subsequent infection. To circumvent this lack of an effective immunogenic response in infants and young children to polysaccharides, the art has developed means of converting the T cell independent

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response to T cell dependent response by covalently coupling polysaccharide bacterial antigens to a carrier protein to form a conjugate molecule. See, Jennings et al. U.S. patent 4,356,170.

5 Various procedures have been described in the art for conjugating capsular polysaccharides to proteins. For review, see Contributions to Microbiology and Immunology, vol 10, Conjugate Vaccines, volume editions J.M. Cruse and R.E. Lewis, Jr., 1989.

10 In one method, polysaccharide is subjected to mild acid hydrolysis to produce reducing end groups capable of reacting with protein to form a covalent bond. Anderson, P.A., Infect. Immun., 39:233-238 (1983). However, the terminal sugar
15 groups which participate in conjugating to protein exist in equilibrium between a hemiacetal and aldehyde and therefore couple to protein with poor efficiency. To overcome the poor reactivity of the terminal reducing sugar, the art turned to mild oxidation to introduce
20 stable aldehyde groups at terminal positions of polysaccharides used to conjugate to protein. Jennings et al. U.S. patent 4,356,170, supra.

 Kasper et al. U.S. patent 5,302,386 and International application WO 94/06467, respectively relate
25 to GBS type III and II conjugate vaccines.

 According to the 5,302,386 patent, endo- β -galactosidase is used to cleave the polysaccharide backbone to produce products suitable for conjugating to protein. Oxidation of at least two
30 terminal sialic acid groups to produce cross-linked conjugates is described in the WO 94/06467 application.

 Type III GBS capsular polysaccharides are composed of a backbone of repeating branched pentasaccharide units. Jennings et al., Canadian J. Biochem., 58:112-120 (1980). One study of type III GBS
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polysaccharides reports that the natural immunodeterminant site is located at the side chain-backbone junction. Jennings et al., Biochemistry, 20:4511-4518 (1980). The presence of the side chain terminal N-acetyl-neuraminic acid residues reportedly was critical for immunodeterminant expression.

Prior methods of depolymerizing GBS II or III polysaccharides rely on either costly enzymatic methods or on acid hydrolysis which may alter the antigenicity of the CP due to removal of the labile terminal sialic acid groups. Accordingly, there is a need for relatively inexpensive and mild chemical procedures which are effective for depolymerizing GBS type II and III CP in a manner which results in fragments which are useful for producing CP-protein conjugate vaccines.

SUMMARY OF THE INVENTION

This invention relates to a method of depolymerizing Group B Streptococcus type II (GBS-II) and type III (GBS-III) capsular polysaccharides (CP) by deaminative cleavage to generate products terminating with a 2,5-anhydro-D-mannose structure. According to this invention, the GBS-II CP and GBS-III CP are treated with sodium hydroxide and a nitrosation reagent such as sodium nitrite to depolymerize the GBS polysaccharides to produce fragments having a terminal aldehyde group located at the end of the polysaccharide backbone. The resulting CP fragments are antigenic and are also useful for conjugating to protein to produce immunogens which are effective for eliciting protective immune responses in mammals including neonates.

Another embodiment of this invention therefore is a method of making a conjugate molecule for use as a vaccine. The method comprises subjecting GBS-II or GBS-III CP to treatment by base and a diazonium salt forming reagent to form a fragment terminating with a 2,5-anhydro-

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° D-mannose residue. The 2,5-anhydro-D-mannose terminating fragment is then combined with a protein and subjected to reductive amination to form the conjugate molecules of the invention. Accordingly, another aspect of this invention are GBS-II and GBS-III CP conjugate molecules comprising GBS-II or GBS-III CP fragments linked to protein through a terminal 2,5-anhydro-D-mannose. Because the process of depolymerizing the GBS type II and type III polysaccharides generates fragments having a single reactive site at the terminal end of the backbone, this invention provides a means of producing conjugate molecules wherein each GBS type II or III polysaccharide chain is bound to a single protein, each by a secondary amine through the terminal reducing sugar.

The conjugates of this invention are useful as active vaccines for immunizing individuals against GBS-II and GBS-III bacterial infection. Also provided by this invention are multivalent vaccines comprising polysaccharides derived from different serotypes or species of bacteria.

In addition, this invention encompasses immune serum or antibodies raised in response to immunization with the conjugate molecules of this invention and which are useful as reagents for detecting the presence of GBS type II or III bacteria or as vaccines for conferring passive immunity.

Another embodiment of this invention are methods and compositions useful for separating and/or detecting GBS type II or type III antibodies. According to one mode of practicing this embodiment, the polysaccharide fragments prepared according to this invention are immobilized onto a solid support. By combining a source of antibody such as serum, with the polysaccharide fragment bound to the solid support, the antibody which bonds to the polysaccharide fragment may be detected by

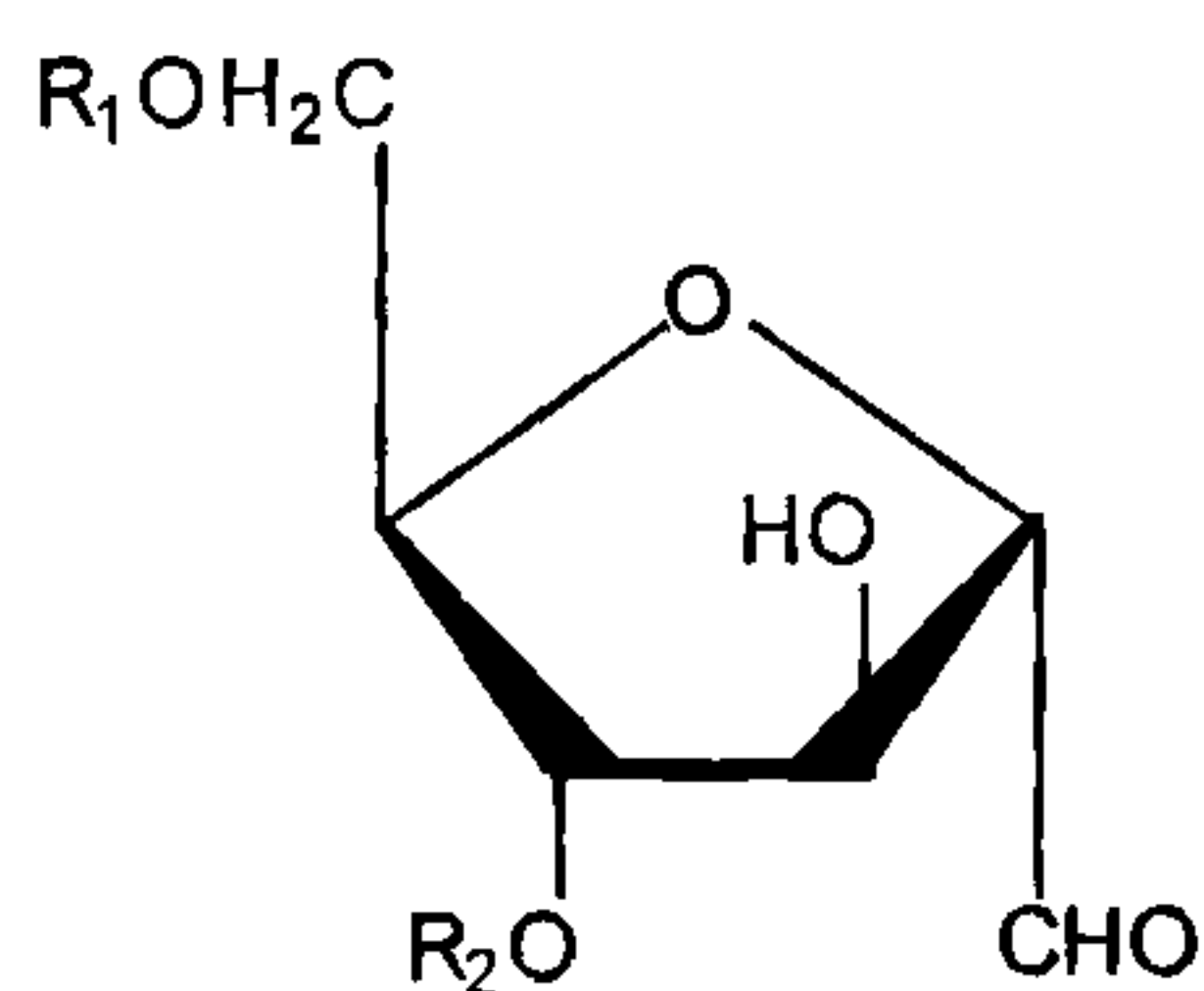
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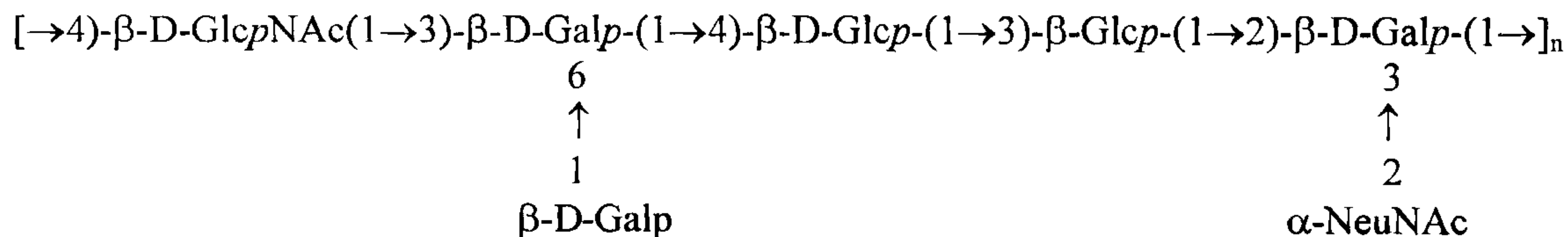
standard immunoassay techniques or separated from the starting material or serum.

An object of this invention is to provide methods for fragmenting GBS type II and III polysaccharides to
 5 produce fragments useful producing conjugate molecules. Another object of this invention is to produce GBS type II and type III polysaccharide molecules which are useful as vaccines to protect against infection and as immunoreagents.

According to one aspect of the present invention,
 10 there is provided a process for depolymerizing Group B Streptococcus (GBS) type II and type III polysaccharides to produce fragments having the following 2,5-anhydro-D mannose reducing-end structure,



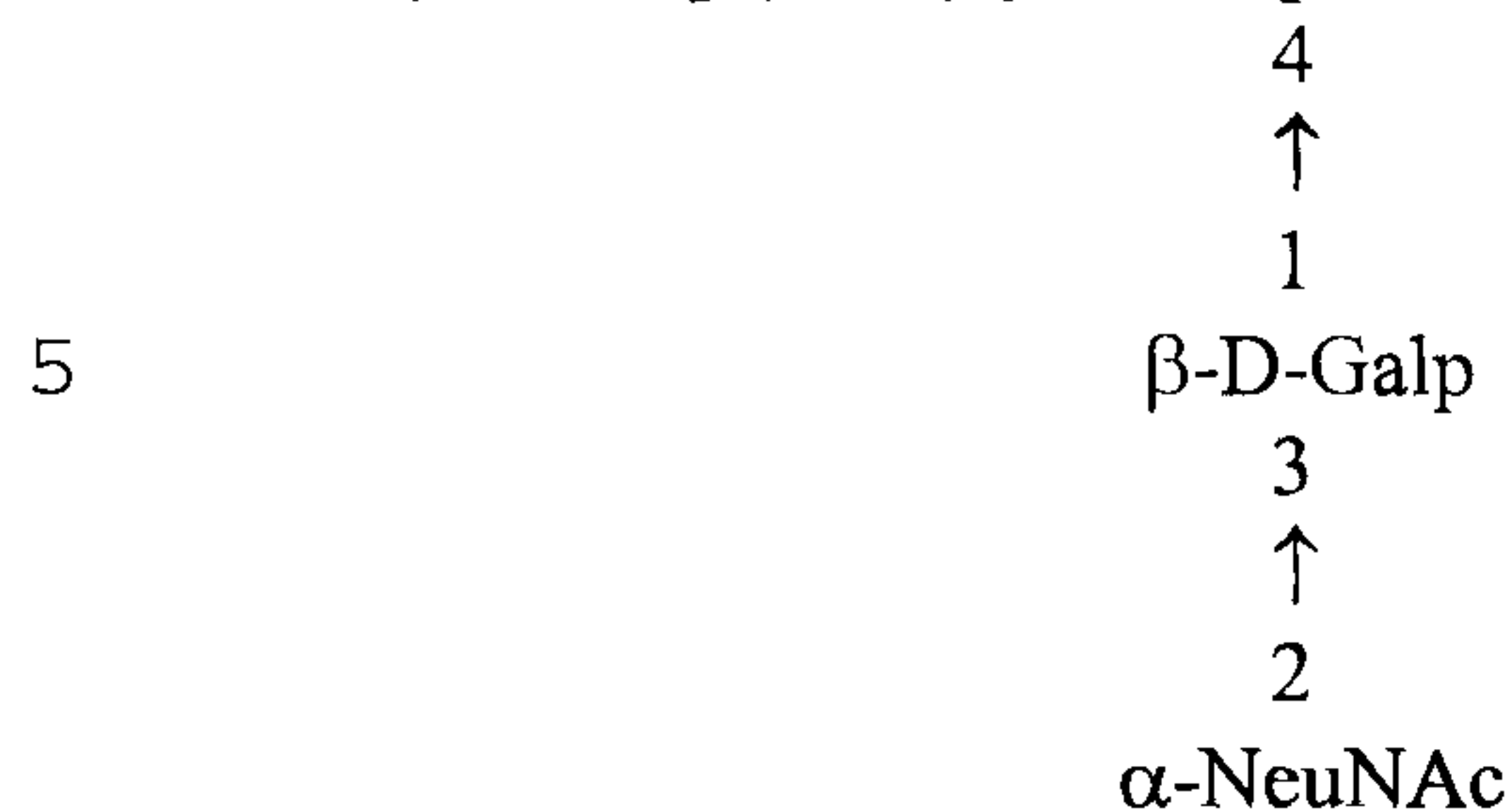
wherein R_1 is H and R_2 is sialylated
 15 heptasaccharide repeating-unit of formula



wherein n is 5 to 50 for the type II, and where R_1 is sialylated pentasaccharide repeating-units of formula

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- 5a -

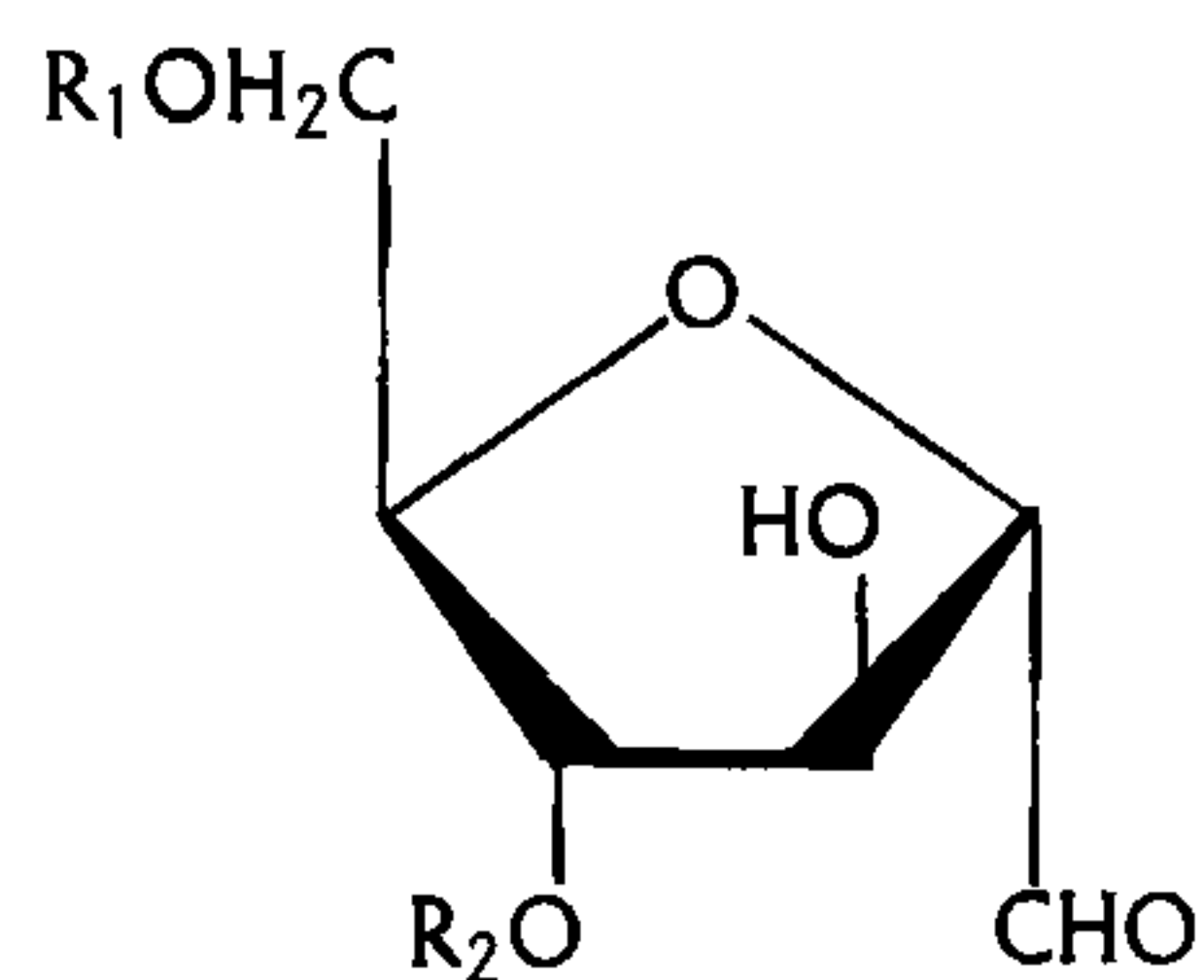


10 wherein n is 5 to 50 and R₂ is disaccharide
 $\alpha\text{NeuAc}(2\text{-}3)\beta\text{-D-Galp}1\text{-}$ for the type III,

said process comprising the steps of

providing a GBS type II or GBS type III
 polysaccharide to be depolymerized and reacting the
 15 polysaccharide in an aqueous medium with a base to form a
 partially de-N-acetylated polysaccharide product,
 depolymerizing the de-N-acetylated product with a
 nitrosation agent to form the GBS type II or type III
 fragments, and recovering the fragments.

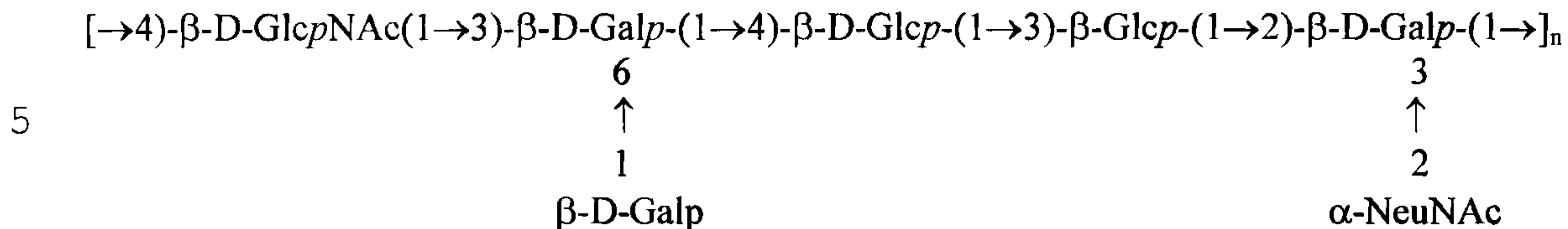
20 According to another aspect of the present
 invention, there is provided a GBS type II or type III
 polysaccharide fragment prepared according to the process
 described above, wherein the fragment has the 2,5-anhydro-D
 mannose reducing-end structure,



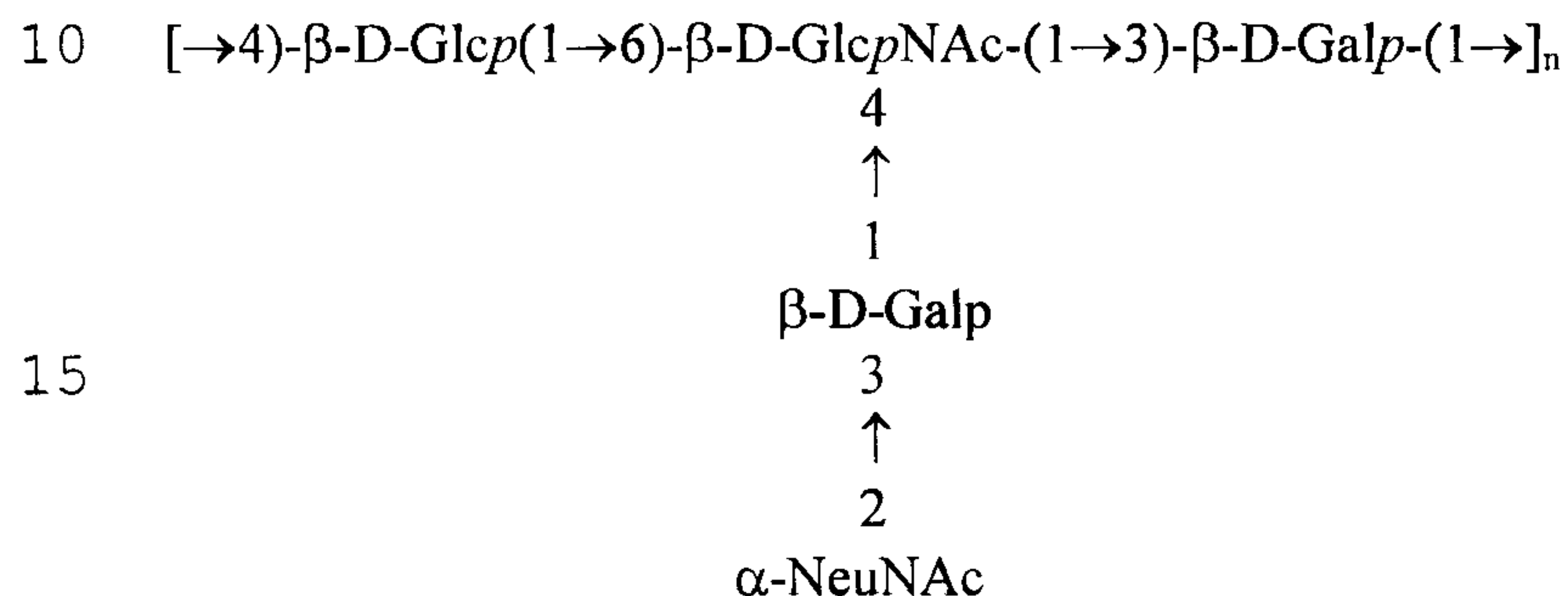
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wherein R_1 is H and R_2 is sialylated
heptasaccharide repeating-unit of formula



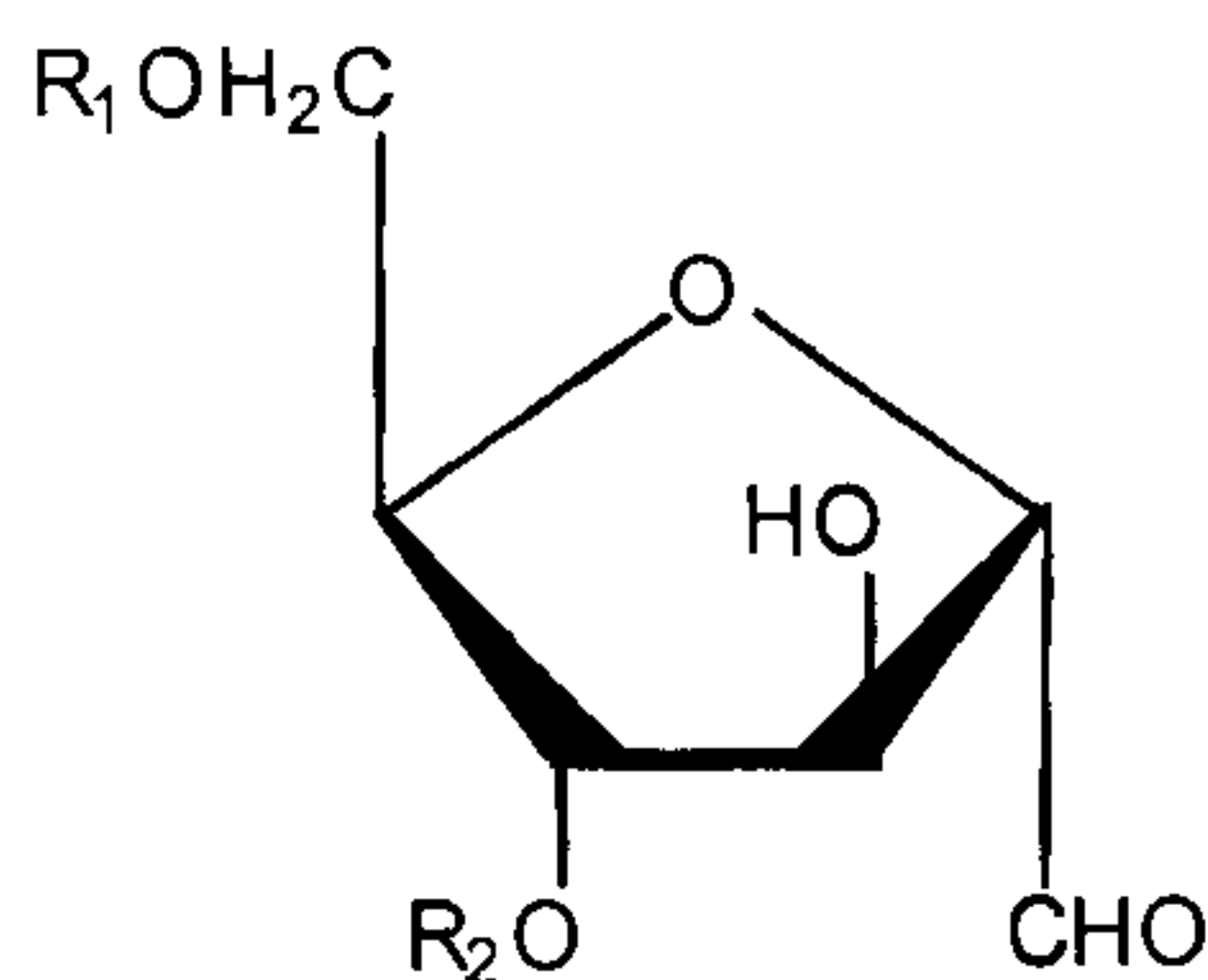
wherein n is 5 to 50 for the type II, and where R_1
is sialylated pentasaccharide repeating-units of formula



wherein n is 5 to 50 and R_2 is disaccharide

$\alpha\text{NeuAc(2-3)}\beta\text{-D-Galp1-}$ for the type III.

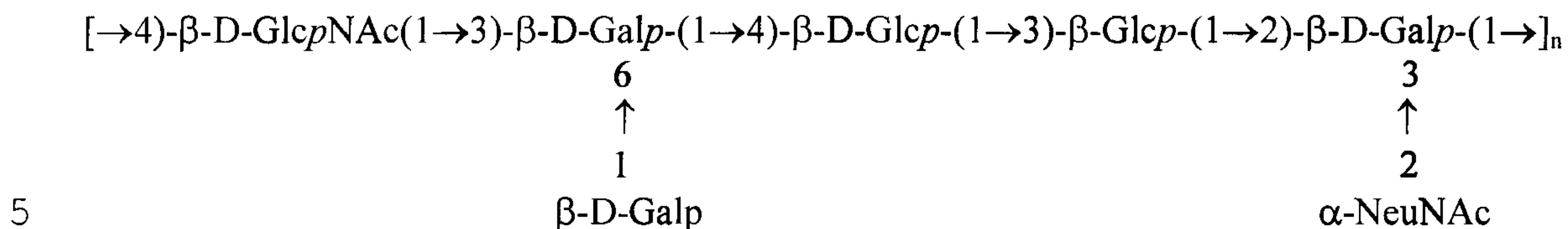
According to still another aspect of the present
invention, there is provided a Group B Streptococcus (GBS)
type II or type III polysaccharide fragment having the
following 2,5-anhydro-D mannose reducing-end structure,



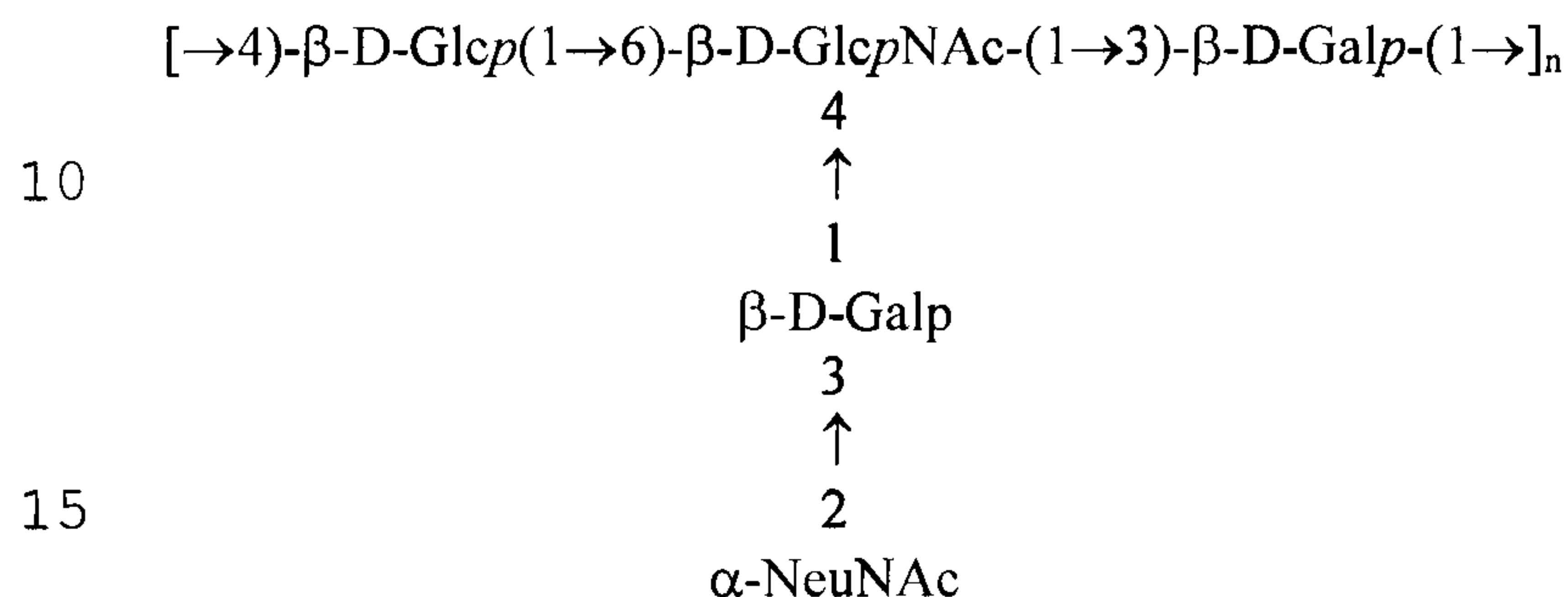
wherein R_1 is H and R_2 is sialylated
heptasaccharide repeating-unit of formula

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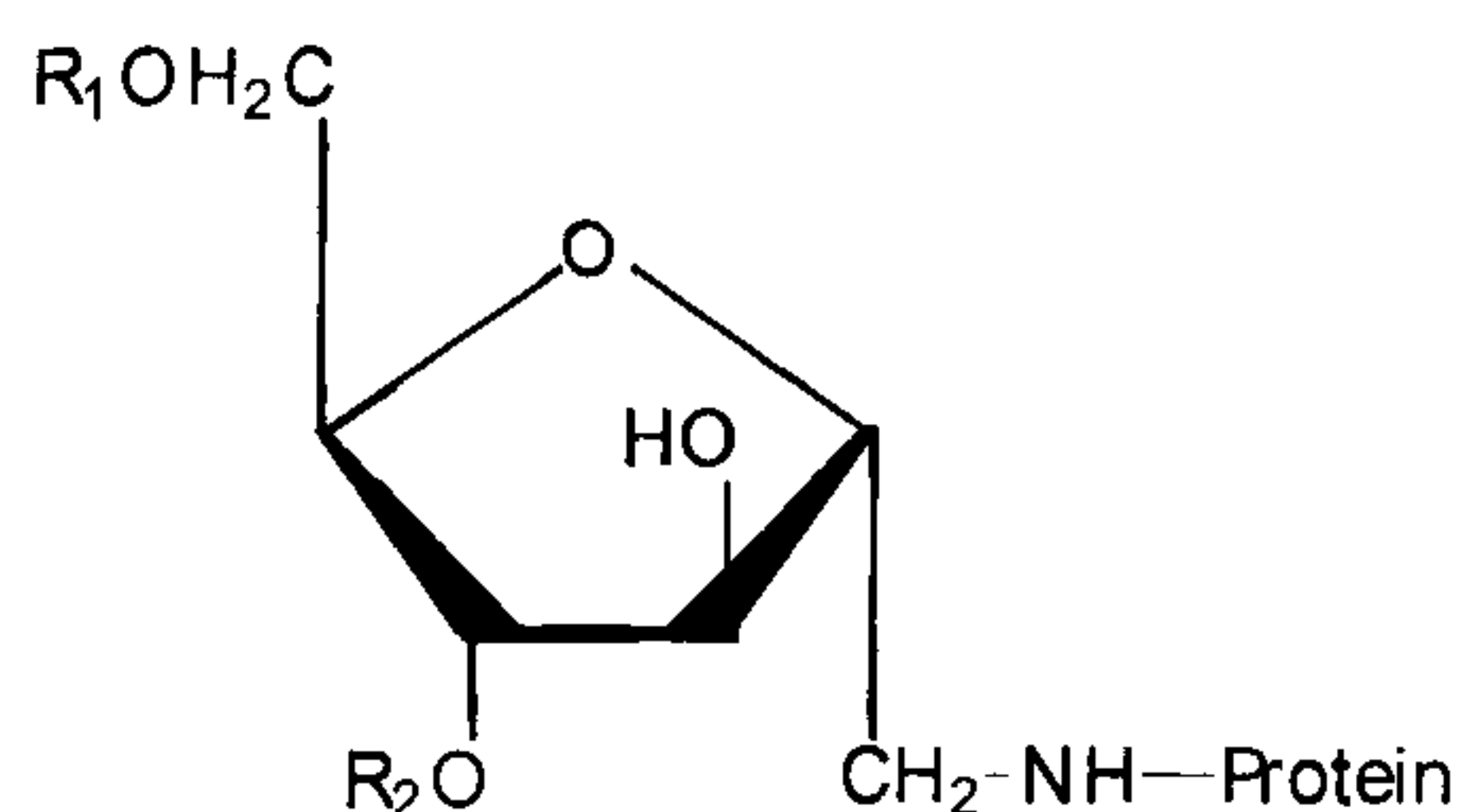


wherein n is 5 to 50 for the type II, and where R₁ is sialylated pentasaccharide repeating-units of formula



wherein n is 5 to 50 and R₂ is disaccharide $\alpha\text{NeuAc}(2\rightarrow 3)\beta\text{-D-Galp}1\text{-}$ for the type III.

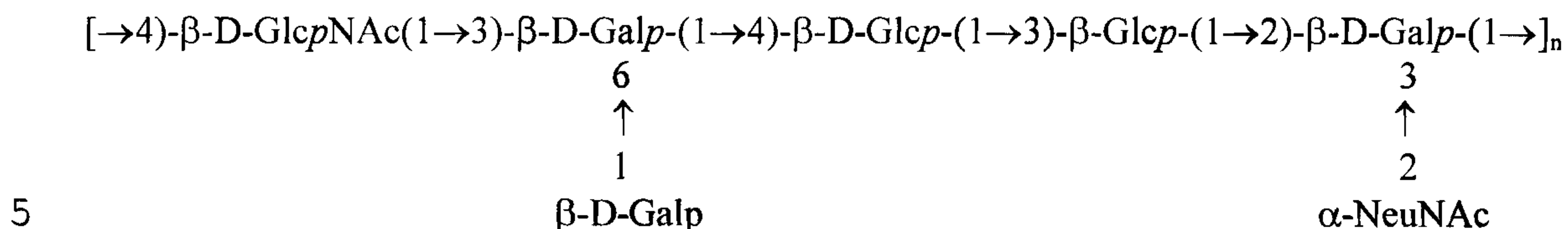
According to yet another aspect of the present invention, there is provided a conjugate molecule comprising at least one polysaccharide fragment selected from GBS type II and type III polysaccharide fragments, and wherein the fragment is covalently bound to a protein wherein the conjugate molecule has the structure



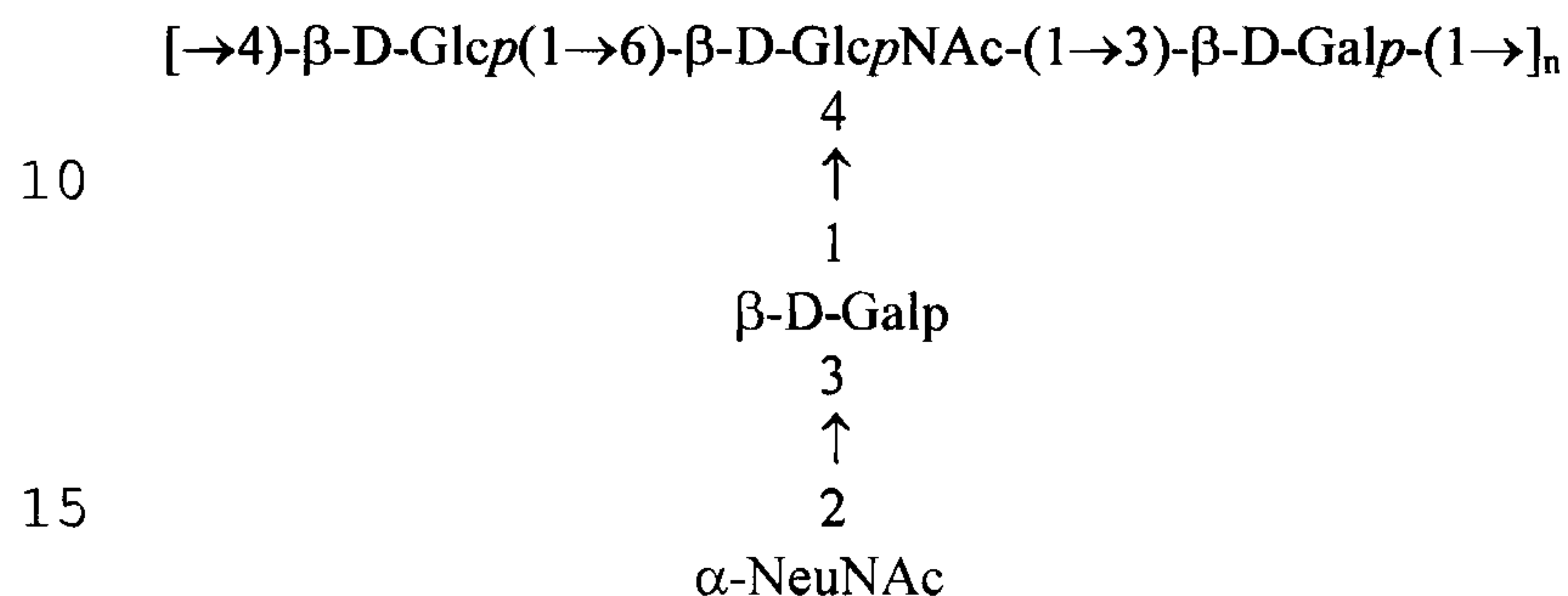
wherein R₁ is H and R₂ is sialylated heptasaccharide repeating-unit of formula

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- 5d -



wherein n is 5 to 50 for the type II, and where R₁ is sialylated pentasaccharide repeating-units of formula



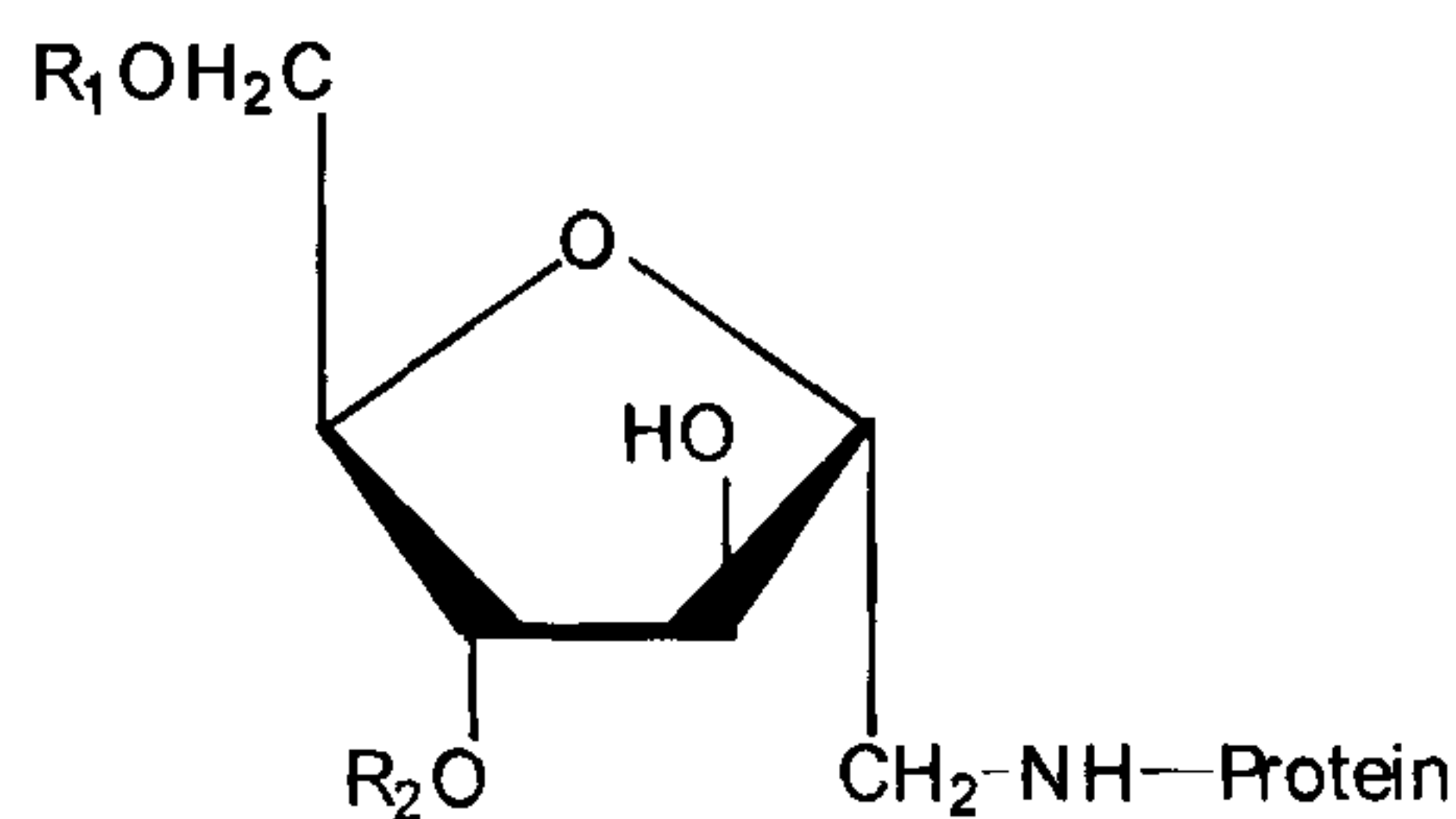
wherein n is 5 to 50 and R₂ is disaccharide $\alpha\text{NeuAc}(2\rightarrow 3)\beta\text{-D-Galp}1\text{-}$ for the type III.

According to a further aspect of the present invention, there is provided the conjugate molecule as described above wherein the protein is derived from a bacteria selected from the group consisting of tetanus toxoid, diphtheria toxoid, CRM₁₉₇, a recombinant non IgA binding protein of the β -C antigen of type Ia/Ib Group B streptococcus, and recombinant class 3 outer membrane protein from Neisseria Meningitides.

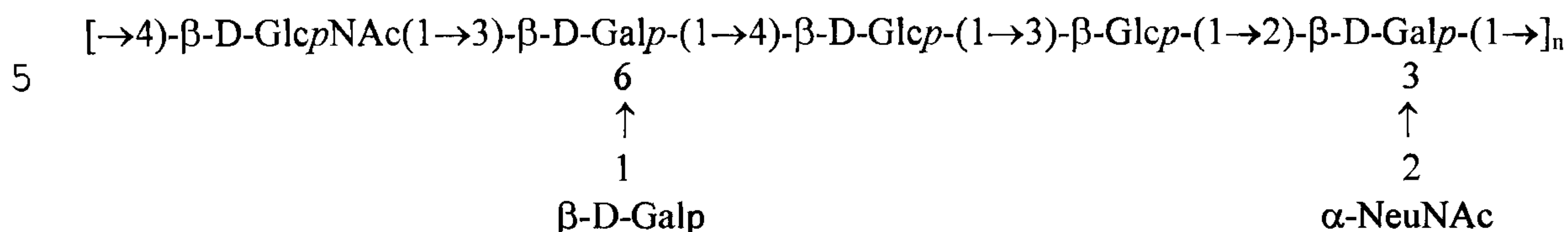
According to yet a further aspect of the present invention, there is provided a vaccine comprising conjugate molecules comprising a GBS type II and/or type III polysaccharide fragment covalently bound to a protein wherein the conjugate molecule has the structure

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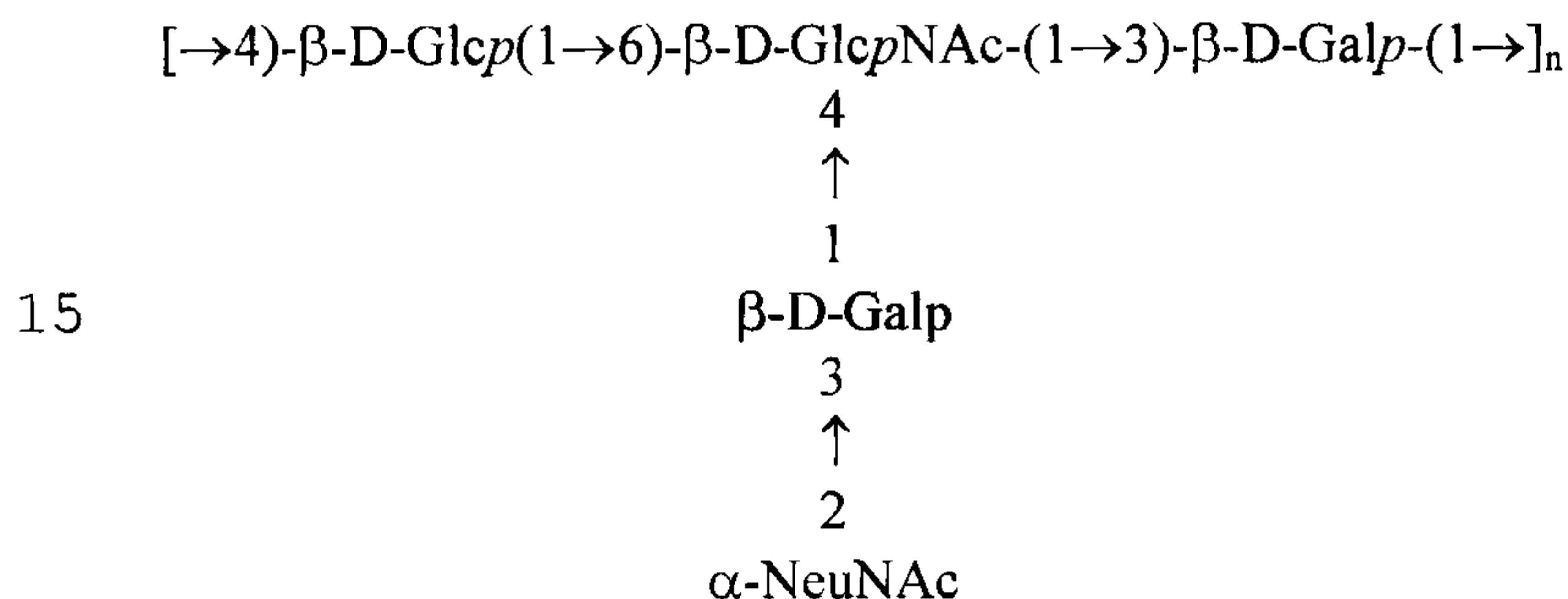
- 5e -



wherein R_1 is H and R_2 is sialylated
heptasaccharide repeating-unit of formula



wherein n is 5 to 50 for the type II, and where R_1
10 is sialylated pentasaccharide repeating-units of formula



20 wherein n is 5 to 50 and R_2 is disaccharide
 $\alpha\text{NeuAc}(2\text{-}3)\beta\text{-D-Galp}1\text{-}$ for the type III.

According to still a further aspect of the present
invention, there is provided an immune serum comprising
antibodies raised in a mammal immunized with the conjugate
25 as described above.

According to another aspect of the present
invention, there is provided a method of separating GBS
type II or III antibodies from serum comprising immobilizing
a polysaccharide fragment prepared as described above to a

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solid support, combining the solid support with bound polysaccharide with serum under conditions to allow binding of GBS type II or III antibodies to the bound polysaccharide fragment, and separating the remaining serum from the solid
5 support.

According to yet another aspect of the present invention, there is provided an immunoassay reagent comprising a GBS type II or III polysaccharide fragment prepared according to the method described above wherein the
10 polysaccharide fragment is immobilized on a solid support.

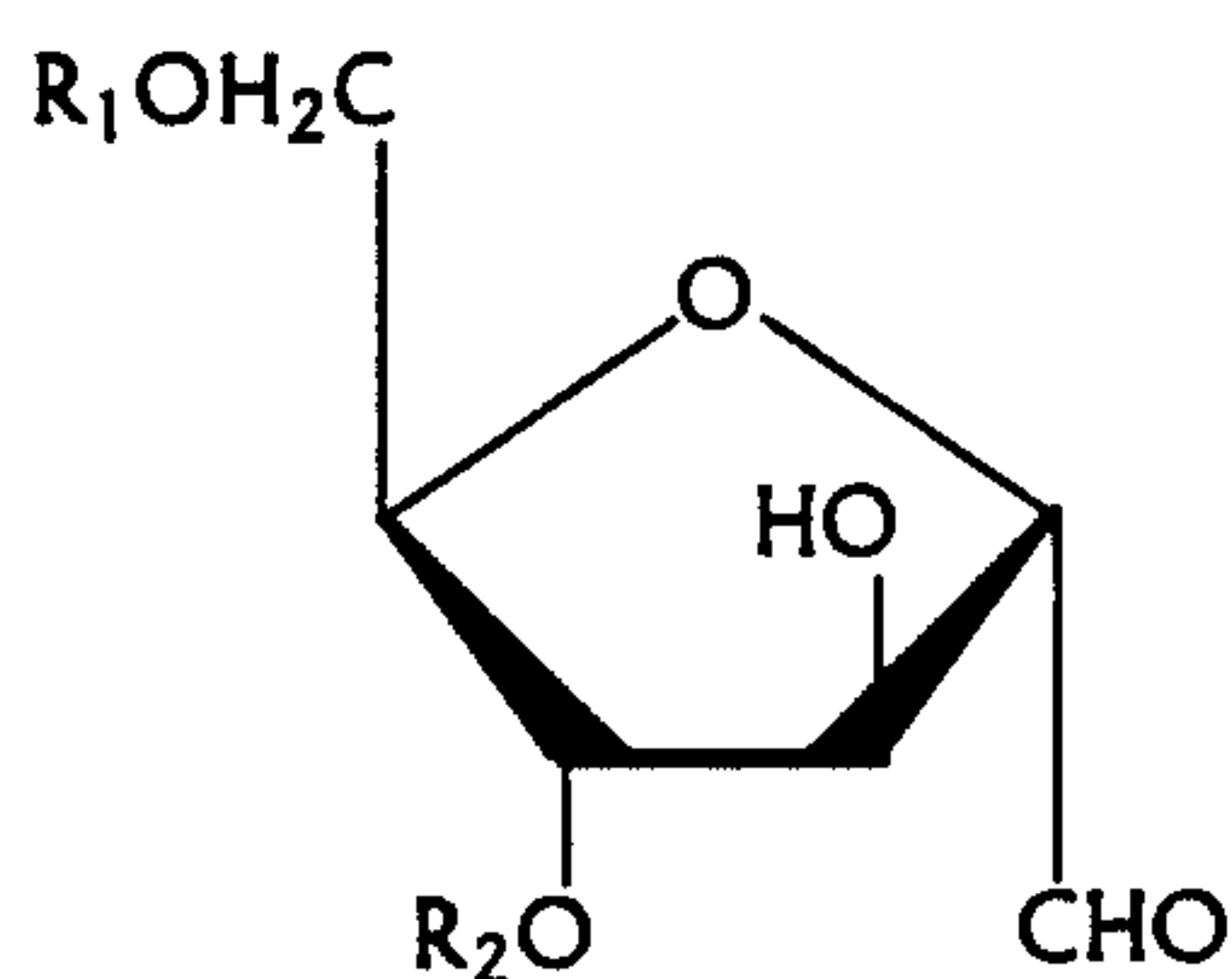
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Fig. 5. H-NMR spectrum of native GBS type III polysaccharide having a molecular weight of approximately 100 kDa.

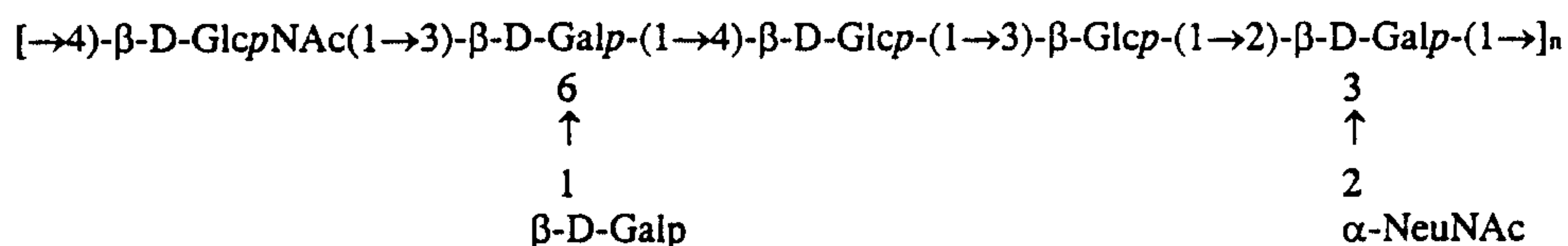
Fig. 6. H-NMR spectrum of GBS type III polysaccharide fragment having a molecular weight of approximately 9 kDa and showing the hydrogen peaks associated with the 2,5-anhydro-D-mannose prepared according to the method of this invention.

DETAILED DESCRIPTION OF THE INVENTION

This invention relates to Group B Streptococcus type II and type III antigenic polysaccharide-fragments having the following 2,5-anhydro-D-mannose reducing-end structure:



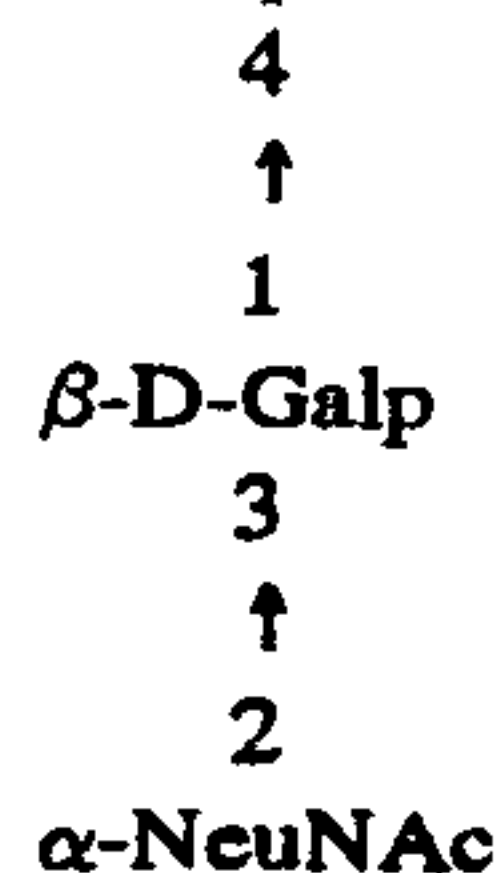
wherein R_1 is H and R_2 is a sialylated heptasaccharide repeating-unit of formula



wherein n is about 5 to about 50 for GBS type II; and

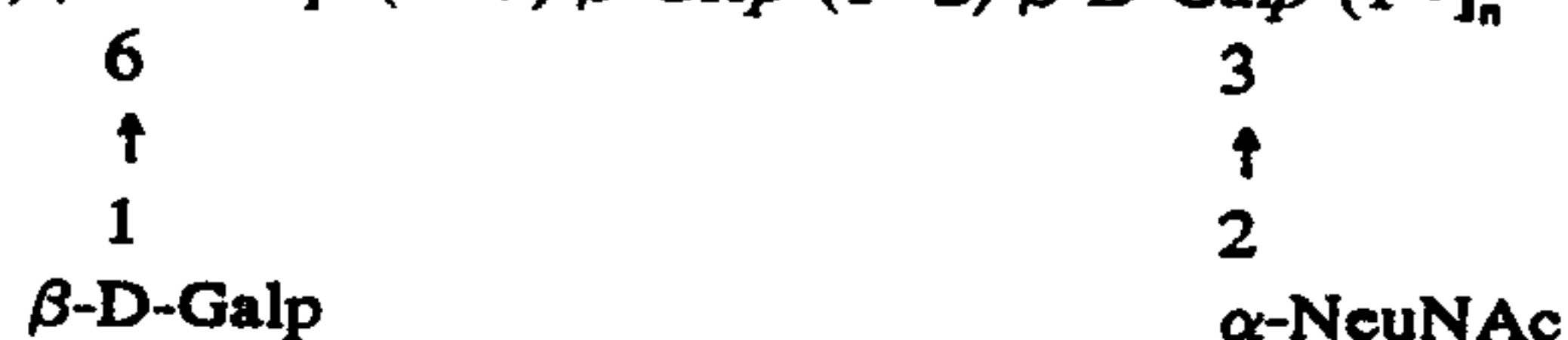
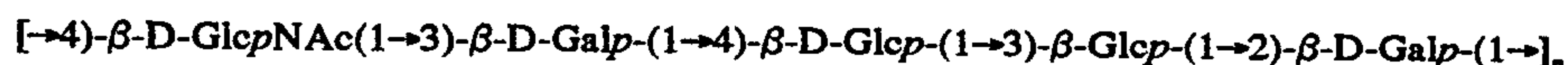
wherein R_1 is a sialylated pentasaccharide repeating-unit of formula

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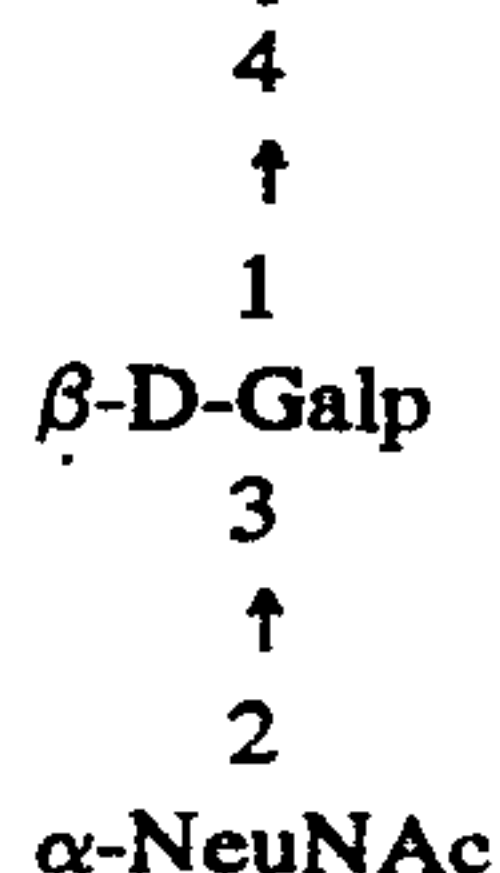
wherein n is about 5 to 50 and R₂ is disaccharide $\alpha\text{NeuAc}(2\rightarrow 3)\beta\text{-D-Galp}1-$ for GBS type III.

These fragments are produced according to this invention by depolymerizing larger molecular weight GBS type II and III polysaccharides having the structures shown below:



TYPE II

wherein n for the native polysaccharide is about 200; or the repeating unit for GBS type III



TYPE III

wherein n for the native polysaccharide is about 100.

As used herein, the term Group B Streptococcus or (GBS) bacteria has the same meaning as understood by those in the art, particularly with reference to Lancefield, J. Exp. Med., 108:329-341 (1938) and subsequent work further characterizing Group B serotypes, e.g. Russell-Jones, J. Exper. Med., 160:1476 (1984), to

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specifically include bacteria taxonomically designated *Streptococcus agalactiae*.

The process of this invention for fragmenting the GBS type II and III capsular polysaccharide to produce the novel fragments of the invention uses mild non-

5 denaturing conditions to obtain GBS type II and GBS type III polysaccharide fragments resulting from chemical depolymerization. These fragments may be obtained in high yield making this process economical for large scale production of vaccines.

10 GBS type II and III CP are depolymerized according to the method of this invention as follows. The backbone 2-deoxy-2N-acetamido- β -D glucopyranosyl residues in the type II and type III GBS CP (Formula I) is partially de-N-acetylated with mild base in an aqueous

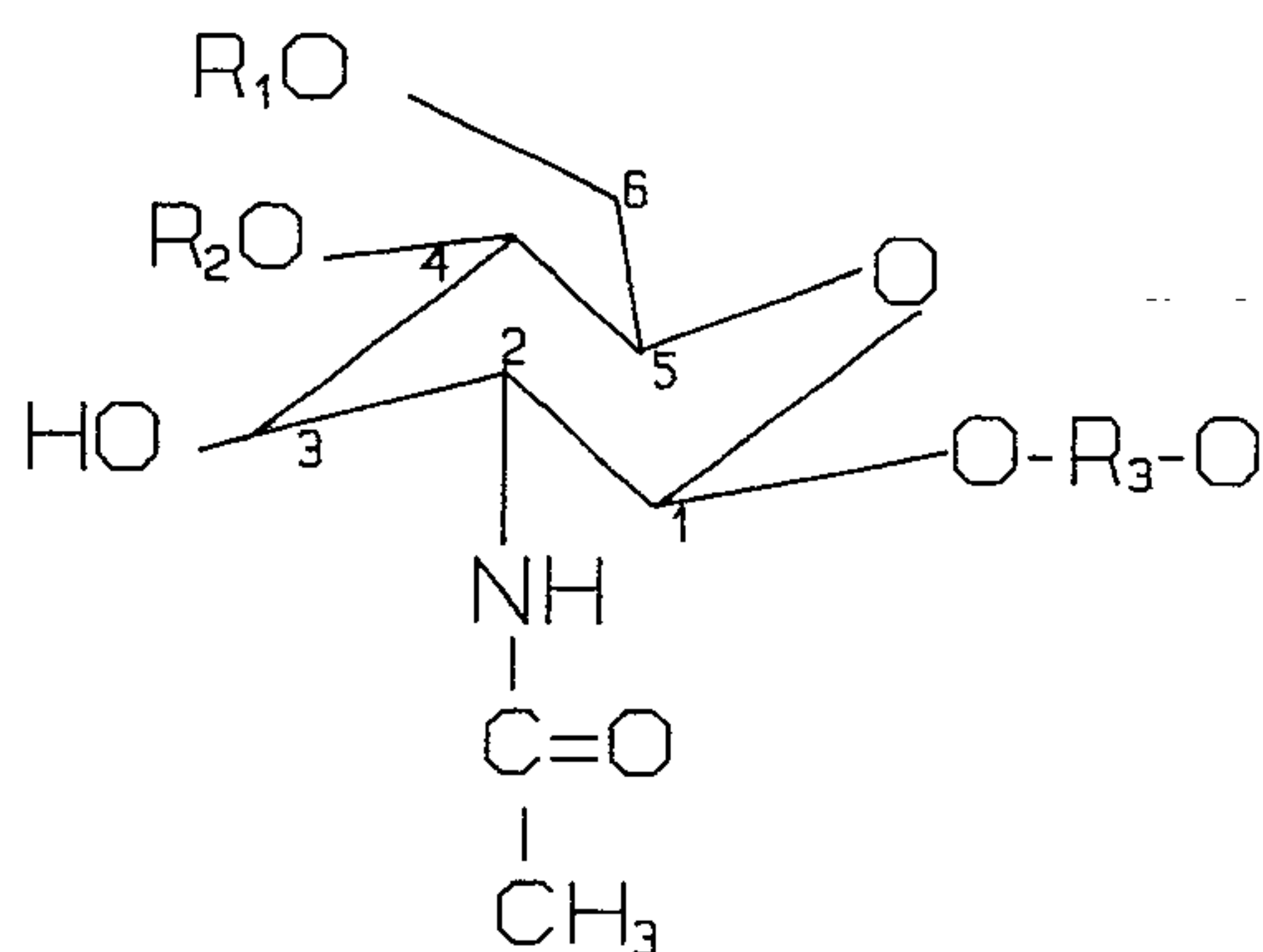
15 solution. Examples of bases which are suitable for use in the process of this invention include, but are not limited to aqueous alkali metal hydroxide solutions for example, sodium hydroxide or potassium hydroxide, or other bases such as ammonium hydroxide, hydrazine, sodium carbonate

20 and sodium bicarbonate. Following base treatment, the resulting glucosamine residue (Formula II) is then susceptible to nitrosation using an appropriate reagent such as sodium nitrite or nitrous acid, for example, to form an unstable N-nitroso derivative (Formula III).

25 Rearrangement due to nucleophilic attack by the ring oxygen on carbon 2, results in ring contraction and cleavage of the adjacent glycosidic linkage. (Formula IV). This reaction has been used to study the structure of heparin and various glycosaminoglycans. Barnett U.S.

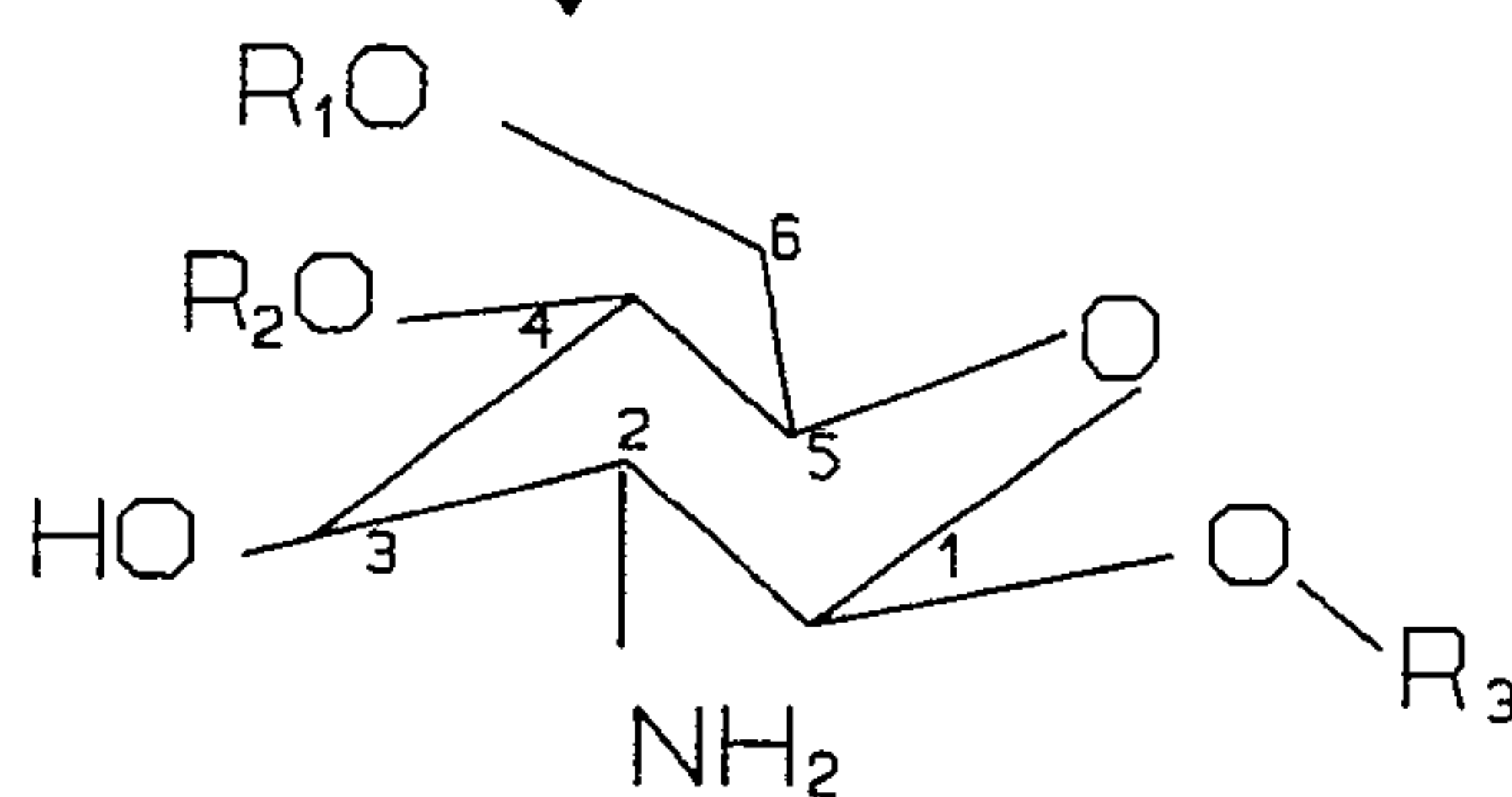
30 Patent 4,438,261.

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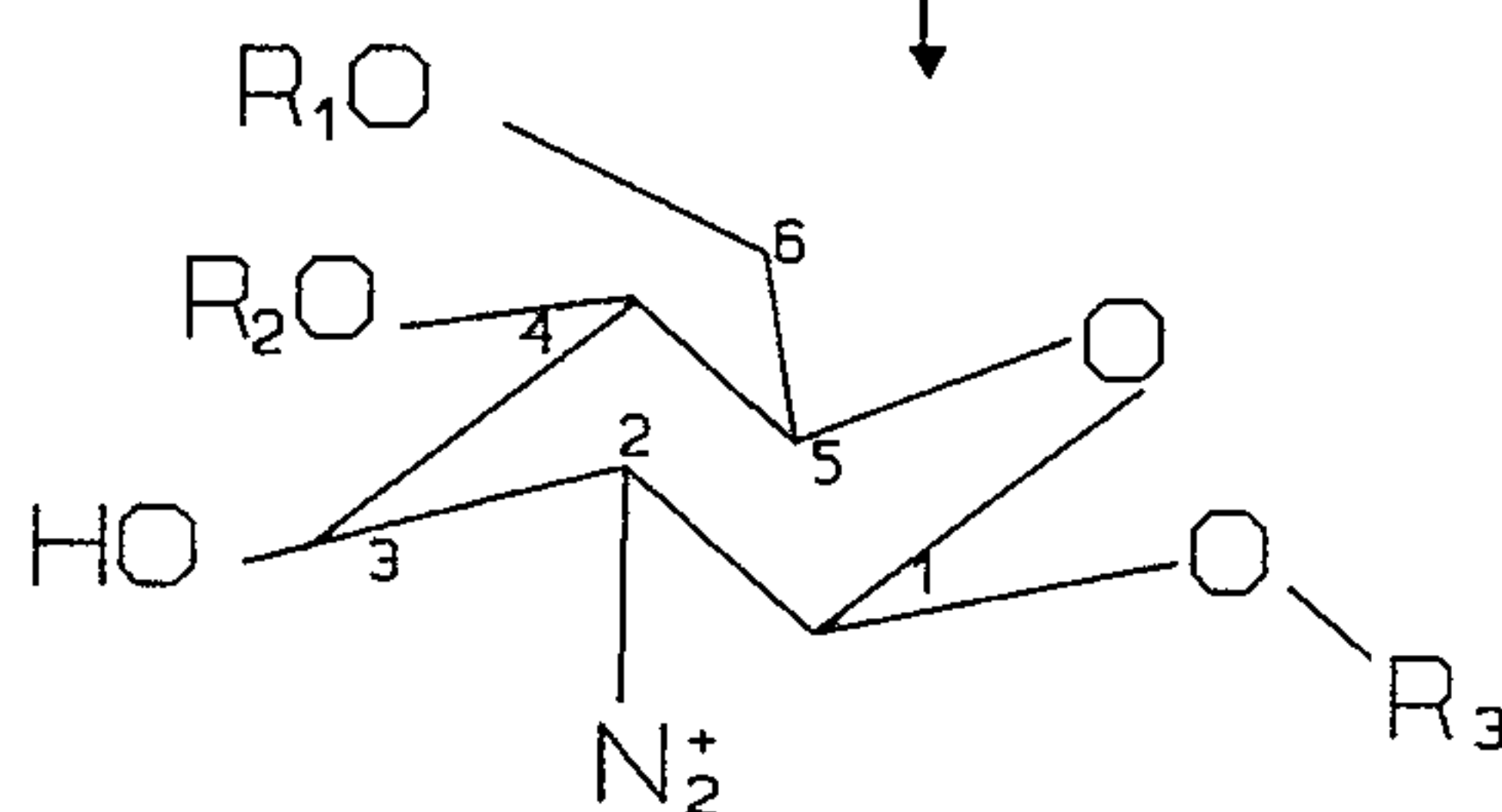


(Formula I)

1) Base

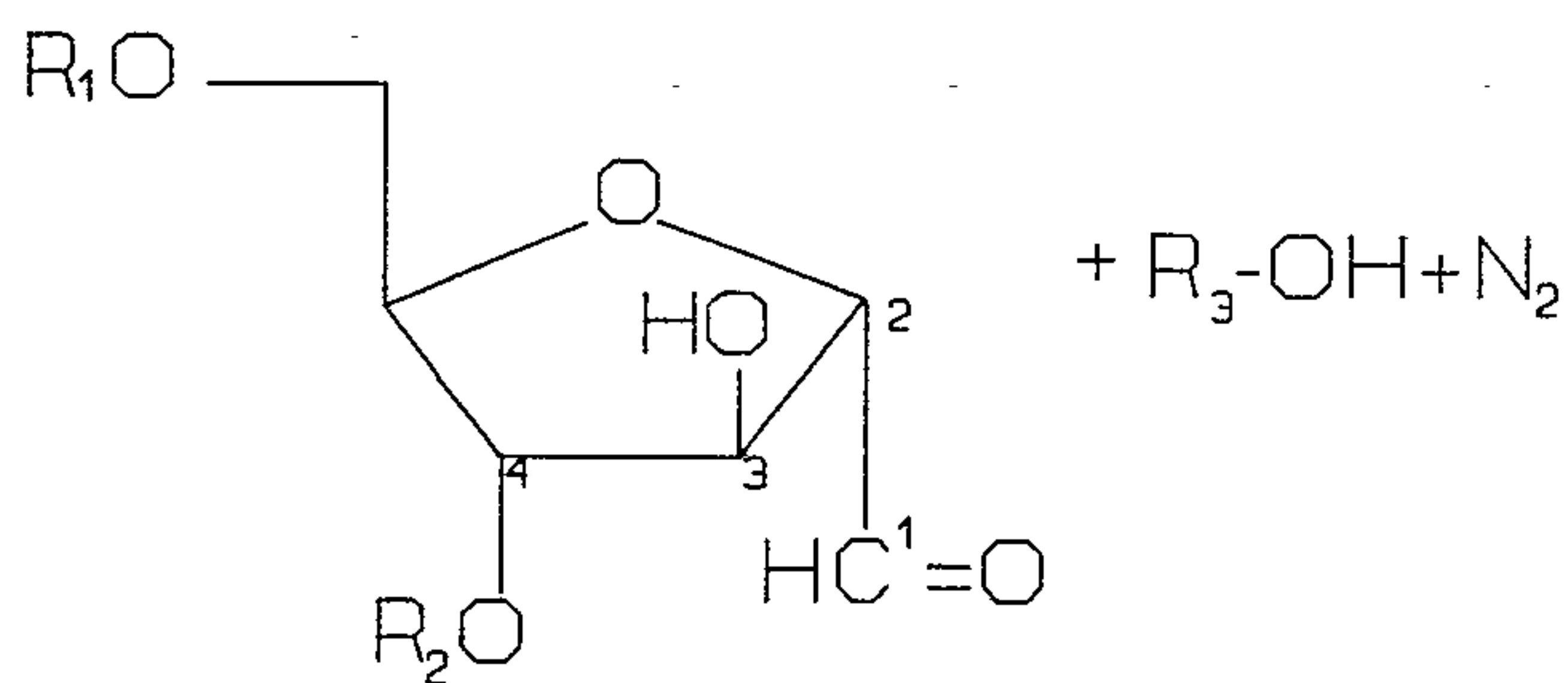


(Formula II)

2) $NaNO_2$ 

(Formula III)

Rearrangement



(Formula IV)

2,5-anhydro-D-mannose

- 10 -

0 The aldehyde group in the resulting 2,5-anhydro-D-mannose (Formula IV) residue formed at the reducing end of the polysaccharide fragment can be used directly, without further chemical manipulation (e.g. use of a spacer arm), for linking through reductive amination to an amino group containing polymer, preferably a protein.

5 More specifically, to carry out the depolymerisation of the GBS polysaccharides according to the invention, the reaction is carried out in a convenient size vessel in an aqueous solution. To begin the reaction, an appropriate amount of polysaccharide in an aqueous solution is treated with a base to partially de-N-acetylate the backbone glucopyranosyl residue. Briefly, the reaction can be carried out in a basic aqueous medium at elevated temperatures, for example about 50°C to 110°C, and at a pH of about 13 to 14. The amount of base be optimized empirically. Preferably the ratio of base to N-acetyl groups is between about 10 to 50 meq. More preferably the ratio is about 20-25 meq. The rate and extent of reaction may be optimized by adjusting the base concentration, reaction temperature or time of reaction. The extent of de-N-acetylation may be monitored by ¹H-NMR.

15 To achieve fragments of between about 5 to 60 kDa, the degree of de-N-acetylation should be between about 20 to about 2 percent of the total number of available sites. To stop the deacetylation reaction, the reaction may be cooled, i.e., chilled on ice, or acidified to about pH 4. Acidification must be done carefully to avoid hydrolysis of remaining sialic acid groups.

30 The nitrosation reaction to form the aldehyde is then achieved by addition of sodium nitrite, or other suitable reagent, such as dilute nitrous acid, to the de-N-acetylated polysaccharide. The nitrosation reagent such as sodium nitrite preferably is added in molar excess compared to the moles of de-N-acetylated groups. Reaction

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of the polysaccharide with the nitrosation reagent is carried out at cold temperatures, for example about 4°C with stirring, for approximately 2 hours or until completion. The extent of the reaction may be monitored by assaying for the presence of aldehyde groups. Over the course of the reaction, the concentration of aldehyde groups should increase until a plateau is reached. Termination of the reaction may be accomplished by dilution of the reaction and by raising the pH to about 7 with dilute base such as NaOH. Removal of excess reagents may be accomplished by dialysis using standard procedures.

After completion of the reaction, polysaccharide fragments having a terminal aldehydic group at the end of the backbone may be sized and collected using standard chromatography procedures. Preferred sizes for conjugation to protein are between 5 kDa and 50 kDa for the GBS type II polysaccharide and between about 5 kDa and 50 kDa for the GBS type III polysaccharide. More preferred sizes are between 5 and 20 kDa for GBS type II and between 10 and 50 kDa for GBS type III polysaccharides. The sized fragments may be used for conjugation reactions using standard reductive amination procedures previously described (See for example U.S. patent 4,356,170 and International application WO 94/06467) or may be stored for later use.

Deaminative cleavage and conjugation applied to GBS type II may be accomplished according to the invention as follows:



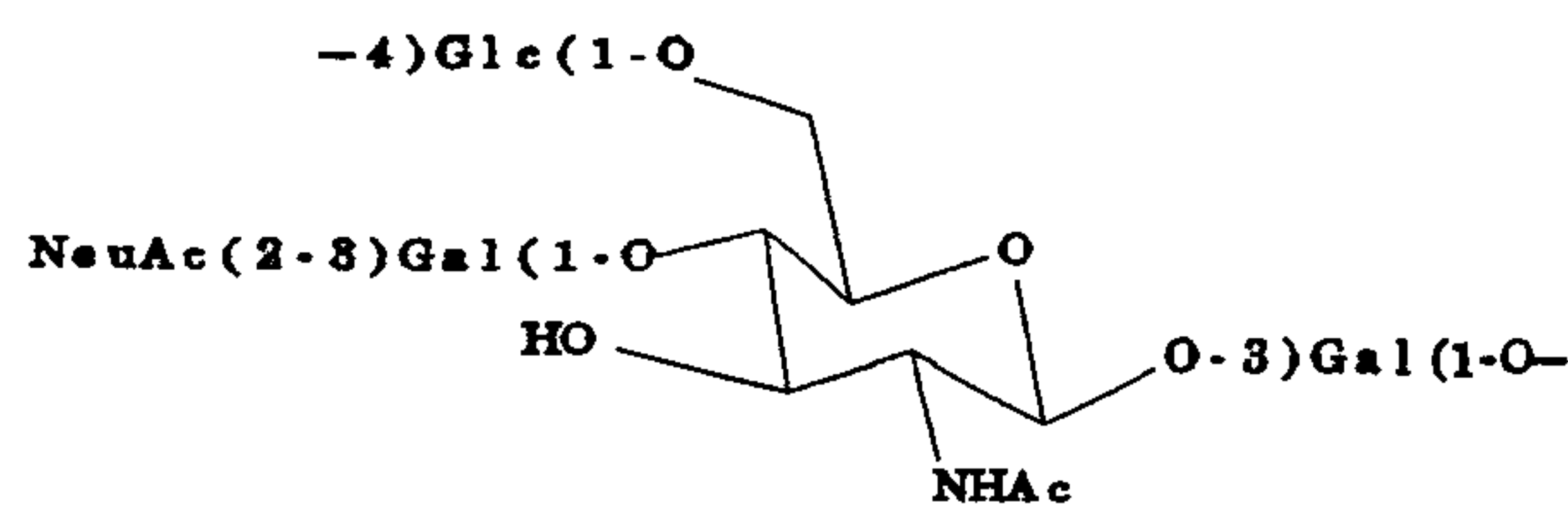
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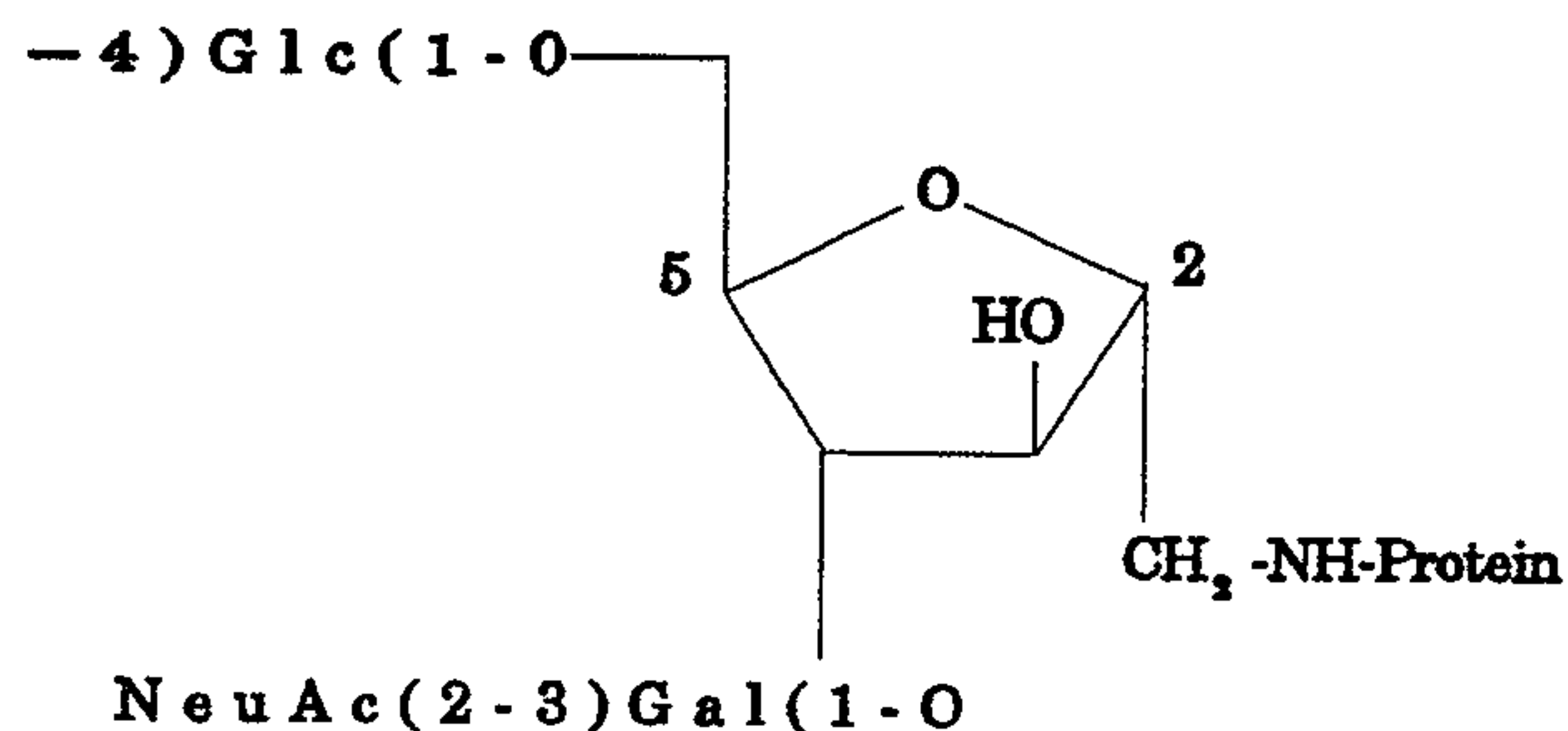
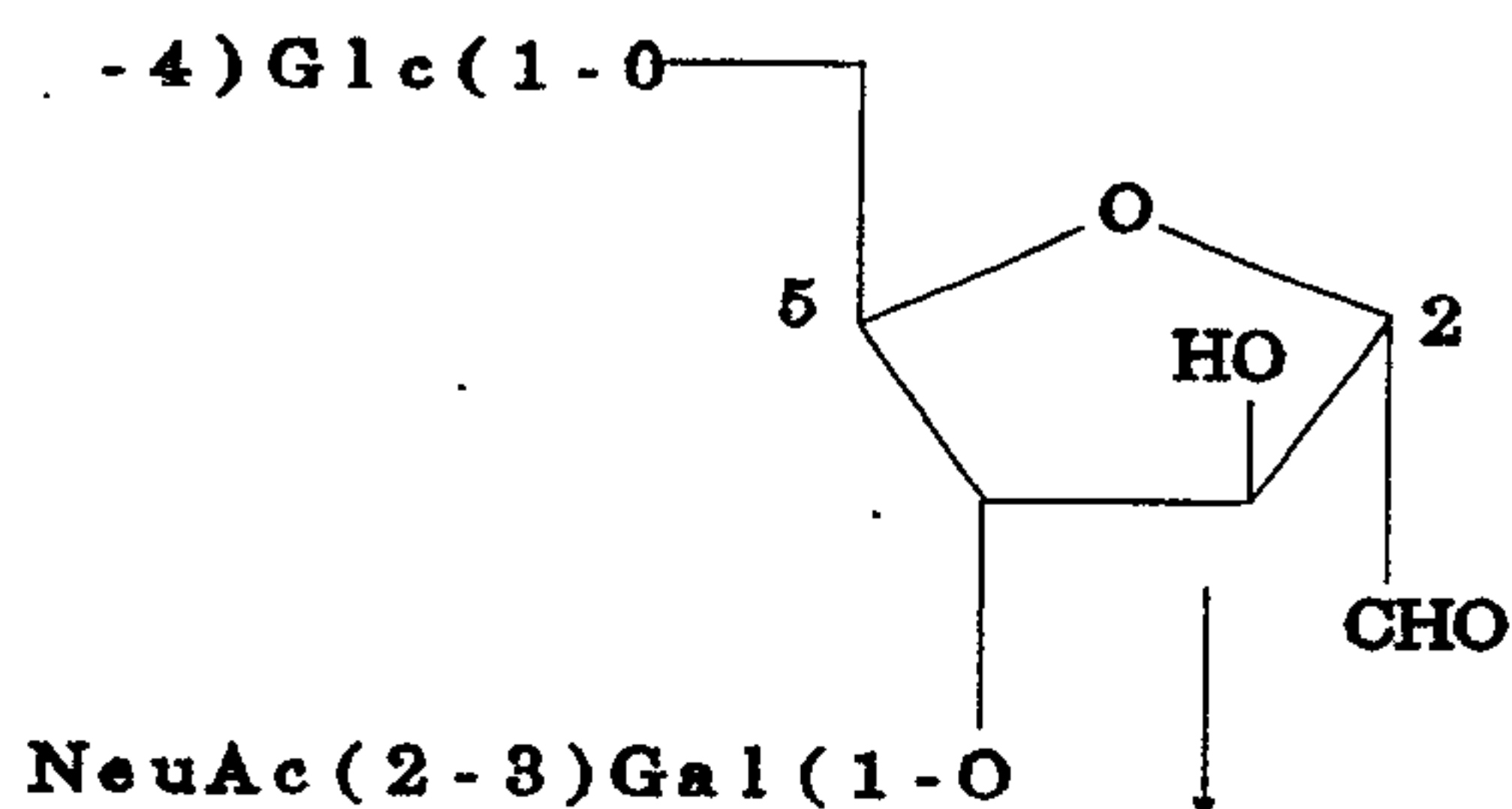
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SUBSTITUTE SHEET (RULE 26)

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1) Base
2) NaNO_2



The protein component of the conjugate molecules of the invention may be any physiologically tolerated protein or polypeptide of sufficient length to evoke a T cell dependent response. Examples of such proteins include, but are not limited to bacterial proteins, or polypeptides, including tetanus toxin or toxoid, cross reactive materials such as CRM₁₉₇, a recombinant non IgA binding protein of the β -C antigen of type Ia/Ib Group B streptococcus, and recombinant class 3 Outer Membrane Protein and (OMP) from Neisseria Meningitides.

The molar ratio of polysaccharide to protein in the conjugate molecules of the invention is preferably

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° between about 1 mole to about 10 moles polysaccharide per mole protein. More preferably the ratio is between 3 and 10 polysaccharide fragments per mole of protein. Variations in protein/polysaccharide ratio may be achieved by adjusting the ratio of the starting components in the conjugation reaction.

In addition to providing conjugate molecules comprising GBS type II or III polysaccharides conjugated to protein, this invention also contemplates multivalent conjugates and their vaccines wherein different types of polysaccharides are conjugated to a single protein. For example, the polysaccharides of GBS types I, II, III, IV or V may be bound to protein in various combinations, as well as polysaccharides derived from other bacteria such as, for example, Haemophilus influenzae type b, or meningococcus types A, B or C as well. A preferred combination would be polysaccharides of GBS type II and III.

The conjugate molecules prepared according to this invention typically comprise a protein to which is bound at least one GBS type II or III polysaccharide fragment through a single binding site at the terminal end of the backbone of the polysaccharide fragment. Thus, this invention provides the ability, if desired, to produce GBS type II or III conjugate molecules wherein the polysaccharide component, except for one end, is unobscured by protein. Other methods of conjugating GBS type II and III polysaccharides to protein through the terminal sialic acids of the branches may result in crosslinking, and attachment of polysaccharide to protein at a plurality of sites. This invention also contemplates conjugate molecules which may be made using a combination of methods. For example, conjugates synthesized according to the invention by producing a single reactive 2,5-annhydro-D-mannose terminal group may be further reacted

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with polysaccharides which have been activated at multiple sites.

The process of preparing vaccines according to this invention provides useful vaccines which are important for providing protection against GBS type II and III infection in mammals, and in particular females of child bearing age, neonates, immunocompromised adults, and children who are at risk for GBS infection. These vaccines are expected to be especially useful for administration to pregnant women as a means of evoking an immunogenic response in the fetus prior to birth.

Vaccines are administered in amounts sufficient to provoke an immunogenic response. Typically a dose of between about 1 and 50 μ g of polysaccharide for generating such a response. Dosages may be adjusted based on the size, weight or age of the individual receiving the vaccine. The antibody response in an individual can be monitored by assaying for antibody titer or bactericidal activity and boosted if necessary to enhance the response.

Vaccines may comprise standard carriers, buffers or preservatives known to those in the art which are suitable for vaccines. In addition, adjuvants such as alum or stearyl tyrosine may also be included in the formulation to enhance the immunogenic response.

The polysaccharide fragments prepared according to this invention are also useful for preparing various immuno reagents for use in immunoassays and separations of GBS type II or III antibodies. For example, for immunoassays the polysaccharide fragments may be immobilized either directly or through a protein linker as in the conjugates of this invention to a solid support. The solid support can then be used in various immunoassay systems known to those in the art including radioimmuno and ELISA assays for detecting the presence of antibodies to GBS type II or III bacteria. Such assays may be used for diagnosing the presence of infection in individuals by

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assaying for the presence of GBS type II or III antibodies in serum.

For use in separation chemistry, the polysaccharide fragments may be immobilized to a solid support to prepare an affinity column. In a preferred embodiment, the polysaccharide fragment is first conjugated to protein according to the method of this invention and the resultant conjugate is then coupled to a support matrix. Methods of coupling protein to affinity columns are known to those skilled in the art. Common supports for affinity columns are prepared from agarose and are commercially available, e.g. activated Sepharose* (Pharmacia). Such affinity columns may then be used for separating GBS type II or III antibodies from sources such as serum. Antibody may then be separated from serum by combining the immobilized polysaccharide fragments with serum suspected of containing GBS type II or III antibodies under conditions which allow for antibodies to bind to immobilized fragments. The bound antibody may then either be detected using conventional assay techniques, or separated and recovered from the polysaccharide fragment following separation of the remaining serum components from the immobilized support.

The invention will now be described with reference to the following, non-limiting examples.

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EXAMPLESEXAMPLE 1

5 Base Depolymerization and Sodium Nitrite Mediated Ring
Contraction To Produce GBS Type II And III 2,5-anhydro-D-
Mannose Terminated Fragments

GBS Type II

10 Native type II GBS CP (75 mg) of average
molecular weight about 200,000 was dissolved in 3 ml of
0.5 N NaOH and the solution was then divided in 3 parts (1
ml each). The samples (S1-S3) were heated at 70°C for
60, 90 and 180 min respectively, then chilled in an ice-
water bath. 125 mcL of glacial acetic acid was added to
15 each sample to bring their pH to 4. Following addition of
200 mcL of 5% (w/v) NaNO_2 , the samples were kept under
stirring at 4°C for 2 hrs. S1-S3 samples were then
diluted to 5 ml with DI water and their pH adjusted to 7
with 0.5 N NaOH. Excess reagents were dialysed out by
20 diafiltration with DI water through a Diaflo*
ultrafiltration membrane (Amicon YM 10) and the solutions
were lyophilized. Three type II polysaccharide fragments
(II-1-II-3) were obtained.

GBS Type III

25 Native type III GBS PCP (125 mg) of average
molecular weight about 100,000 was dissolved in 5 ml of
0.5 N NaOH and the solution was then divided in 5 parts (1
ml each). The samples (S1-S5) were heated at 70°C for 60,
30 90, 120, 180 and 240 min respectively, then chilled in an
ice-water bath. 125 mcL of glacial acetic acid was added
to each sample to bring their pH to 4. Following addition
of 200 mcL of 5% (w/v) NaNO_2 , the samples were kept under
stirring at 4°C for 3 hrs. S1-S5 samples were then diluted
35 to 5 ml with DI water and their pH adjusted to 7 with 0.5

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N NaOH. Excess reagents were dialysed out by diafiltration with DI water through a Diaflo* ultrafiltration membrane (Amicon YM 10) and the solutions were lyophilized. Five type III polysaccharide fragments (III-1-III-5) were obtained.

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Sizing of CP Fragments

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The average molecular weight (avMw) of each fragment was estimated by HPLC using a Superose*-12 size exclusion column (Pharmacia) with a dextran series (Pharmacia) of average molecular weight ranging from 10,000 to 2,000 daltons. Void volume (Vo) and total volume (Vt) were determined with Dextran 2,000 and sodium azide respectively. The average molecular weight (avMw) of each fragment determined by this method is as follows:

15

Fragment	Kav (range)	AvMw Kilodaltons	(range)
II-1	0.23 (0.13-0.34)	51	(99-26)
II-2	0.30 (0.19-0.40)	33	(68-17)
II-3	0.43 (0.30-0.50)	15	(33-9)
III-1	0.21 (0.11-0.27)	41	(81-28)
III-2	0.26 (0.17-0.38)	30	(53-13)
III-3	0.29 (0.18-0.40)	24	(50-12)
III-4	0.37 (0.23-0.44)	14	(36-9)
III-5	0.41 (0.31-0.47)	11	(21-8)

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Physico-chemical analysis of the polysaccharide fragments

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The structural integrity of each fragment with respect to their parent native polysaccharide was established by high resolution one-dimensional H-NMR spectroscopy at 500 MHz on a Bruker* AM500 spectrometer. Comparison of the H-NMR spectra of the type II and type III fragments with those of their respective native polysaccharides indicated that no structural change had occurred during the chemical processes, and most

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importantly that terminal sialic acid residues had been preserved during the nitrosation treatment.

Example 2

5 **Conjugation of the type II and type III polysaccharide haptens to tetanus toxoid**

Tetanus toxoid (SSI, Denmark) was first purified to its monomeric form by gel filtration through a Biogel-A column (Biorad Laboratories). The tetanus toxoid monomer
 10 (TTm) thus obtained (150,000 Daltons) was solubilized in 0.2 M phosphate buffer, pH 7.5, at a concentration of 25 mg/ml and added to dried GBS type II (II-1 to II-3) or type III (III-1 to III-5) polysaccharides and recrystallized sodium cyanoborohydride NaCNBH₃ in the
 15 amounts shown below:

		PS(mg)	TTm(mg)	NaCNBH ₃ (mg)	Final vol (ul)
	II-1	10	4	8	200
	II-2	10	4	8	200
20	II-3	11	4.5	9	220
	III-1	18	7.2	14	360
	III-2	10	4	8	200
	III-3	7.2	3	6	150
	III-4	6.4	2.5	5	130
25	III-5	8	4	8	130

The reaction mixtures were then incubated at 37°C for 4 days. The progress of the conjugation reaction was monitored by HPLC of small aliquots of the reaction
 30 mixtures analyzed on Superose^{*}-12 (Pharmacia). The conjugates were purified by molecular exclusion chromatography on a column of Superdex^{*} G-200 (Pharmacia) using PBS containing 0.01% thimerosal as an eluant. Fractions eluting from the column were monitored by a
 35 Waters R403 differential refractometer and by UV

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° spectroscopy at 280 nm. The fractions containing the
conjugates were pooled sterile-filtered through a 0.22 μ m
Millipore membrane and analyzed, respectively, for their
protein and sialic acid contents by the method of Bradford
(Bradford, M.M., 1976. Anal. Biochem., 72:248-254) and by
5 the resorcinol assay. The average total amount of
carbohydrate in each type II and type III individual
conjugate molecule was calculated by taking the sialic
acid content and multiplying them by a correction factor
of 4 and 3.3 respectively based on the composition of
10 their repeating-unit. The analyses for each individual
conjugate are as shown in Table 1 below:

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Table 1

	Conjugate	AvMw PS chains	Protein (mcg/ml)	CHO (mcg/ml)	%CHO in conjugate	#PS chains
	II-1-TT	51,000	120	15	11	0.4
	II-2-TT	33,000	140	42	23	1.4
5	II-3-TT	15,000	110	26	19	2.4
	III-1-TT	41,000	190	70	27	1.2
	III-2-TT	30,000	140	61	29	1.8
	III-3-TT	24,000	100	39	28	2.0
	III-4-TT	14,000	70	17	20	2.0
10	III-5-TT	11,000	80	21	21	3.0

The immunochemical specificity of rabbit polyclonal antibodies for the type II and type III polysaccharide-conjugates as compared to those observed for the native capsular polysaccharides epitopes was measured by ELISA and is reported in Figures 1 (GBS-type II) and 2 (GBS type III).

Example 3

Immunogenic Response of Female Mice To Immunization With GBS Type II and Type III - Tetanus Toxoid Conjugate Vaccines

a. Immunizations Groups of 10 Swiss Webster female mice (4-6 weeks old) were immunized subcutaneously with 2 μ g of either native type II or III polysaccharide or their corresponding Tetanus-toxoid conjugates. The vaccine were absorbed on aluminum hydroxide (Alhydrogel; Superfos, Denmark) at a concentration of 1 mg of elemental aluminum/ml of 10mM PBS containing 0.01% thimerosal. Mice received the vaccine at days 0, 21 and 42 and finally were exsanguinated at day 52. Sera were collected and stored at 70°C.

b. ELISAs and opsonic activity of conjugate antisera

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° **ELISAs:** Microtiter plates (Nunc Polysorb ELISA plates) were sensitized by adding 100 μ L of native type II or III polysaccharide-HSA conjugate (1 μ g/ml) in PBS with 0.02% azide per well. The plates were incubated at 37°C for one hour. The plates were washed with PBS containing 0.05% between 20 (PBS-T) and blocked with 0.5% BSA in PBS for one hour at r.t. The wells were then filled with 100 μ L of serial two-fold dilutions in PBS-T of mice antiserum and the plates were incubated at r.t. for one hour. After washing with PBS-T, plates were filled with 100 μ L of peroxidase labeled goat anti-mouse IgG(H+L) (Kirkegaard & Perry Laboratories) and then washed five times with PBS-T. Finally, 50 μ L of TMB peroxidase substrate (Kirkegaard & Perry Laboratories) were added to each well and following incubation of the plates for 10 min. at r.t. the reaction was stopped by the addition of 50 μ L of 1M H_3PO_4 . The plates were read at 450 nm with a Molecular Device Amex microplate reader using 650nm as a reference wavelength.

Type III polysaccharide-specific antibody titers of mice vaccinated with native type III polysaccharide or type III fragment polysaccharide-Tetanus toxoid conjugates as shown in Table II.

Table II

	Average MW of fragment	ELISA titer at day 52*	OP Titer ⁺
25	13,000	2,500	405
	18,000	2,500	610
	26,000	3,000	1,050
	34,000	3,000	520
30	48,000	12,000	2,600
	Native PS	<100	<100

*Mean titer of serum pooled from 10 mice.

+Opsonophagocytic killing of pooled serum at day 52.

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Type II polysaccharide-specific antibody titers of mice vaccinated with native type II polysaccharide or type II fragment polysaccharide-Tetanus toxoid conjugates is shown in Table III.

Table III

5	Average MW of fragment	ELISA titer at day 52*	OP Titer ⁺
	15,000	123,000	9,300
	33,000	15,000	1,400
	51,000	2,600	<500
10	Native PS	<500	<100

*Mean titer of serum pooled from 10 mice.

+Opsonophagocytic killing of pooled serum at day 52.

Opsonic Activity of Conjugate Antisera: The opsonic ability of mice antisera to the GPS type II or type III polysaccharide-fragment-tetanus toxoid conjugates was tested in an in vitro opsonophagocytic killing assay using the human promyelocytic leukemia HL-60 cell line (ATCC No. CCL 240). Briefly, 200 cfu of GBS type II strain 18RS21 cells or type III strain M781 cells were mixed in equal volume with serum antibodies and incubated under shaking 15 min. at 35°C in a 5% CO₂ incubator. Baby rabbit complement and HL-60 cells (5X10⁵) cultured 5 days in the presence of 90 mM DMF were added to the mixture and incubated at 37°C for 1 hour under shaking. Aliquots were removed for quantitative culture. Titers were determined by extrapolating the antibody dilution corresponding to fifty percent live bacteria.

c. ELISA binding experiments: Direct binding of rabbit anti-GBS type II or type III capsular polysaccharide specific antibodies (obtained from rabbits hyper-immunized with type III or type II GBS whole cells) to the various tetanus-toxoid conjugates was carried out as follows:

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Microtiter plates were sensitized with 100 μ l of various sizes of the GBS type II or type III polysaccharide-tetanus-toxoid (TT) conjugates (1 μ g/ml) in PBS containing 0.01% thimerosal per well. The plates were incubated at r.t. for one hour, processed as described above, and filled with 100 μ l of serial two fold dilutions of rabbit antiserum diluted in PBS-T. The remaining ELISA steps are as described above except for the addition of the secondary peroxidase labeled goat anti-rabbit IgG (H&L) (Kirkegaard & Perry Laboratories).

The binding of the rabbit GBS type II or III polysaccharide-specific antibodies to the various tetanus-toxoid conjugates are expressed in percent relative to the binding of the native type II or type III polysaccharide-tetanus-toxoid conjugates as illustrated in Figs. 1 and 2 respectively.

Example 4

Immunization of Female Mice To Confer Protection Against GBS Infection In Neonatal Mice

Female CD-1 mice (n=3), 6-8 weeks old, from Charles River Laboratories, Wilmington, MA, were injected i.p. with two doses of conjugate vaccines (containing 2 μ g (equalized for polysaccharide 1 μ g/mouse) of polysaccharide-fragment (average MW about 11,000 Daltons) in Alhydrogel 1.3% (Superfos Biosector a/s, batch #2043), total volume 0.5 ml I.P.) on day 0 and 21. control mice (n=3) received vaccines containing 2 μ g of native type III GBSCP. Mice were bred at day 21 and pups (<36h old) were challenged i.p. with lethal doses of type III GBS M781 bacteria (6×10^5 CFU). Survival was assessed 48h after bacterial challenge.

As shown in Table IV vaccination of the female mice with GBS type III-TT conjugate prior to breeding

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conferred protection in 94% of the pups subsequently born to them.

Table IV

	Vaccine	No. of pups (No. of dams)	No. (%) Surviving 48h
5	GBS III-TT	33 (3)	31 (94)+
	GBS III-PS	39 (3)	0 (0)
	Saline	34 (3)	0 0

+Statistically significant ($P < 0.0001$) from control

10 Refs: Lawrence C. Madoff et. al., *Infection and Immunity*, 60:4989-4994 (1992)

Rodewald, A.K., et. al., *Journal of Infectious Disease*, 166:635-639 (1992)

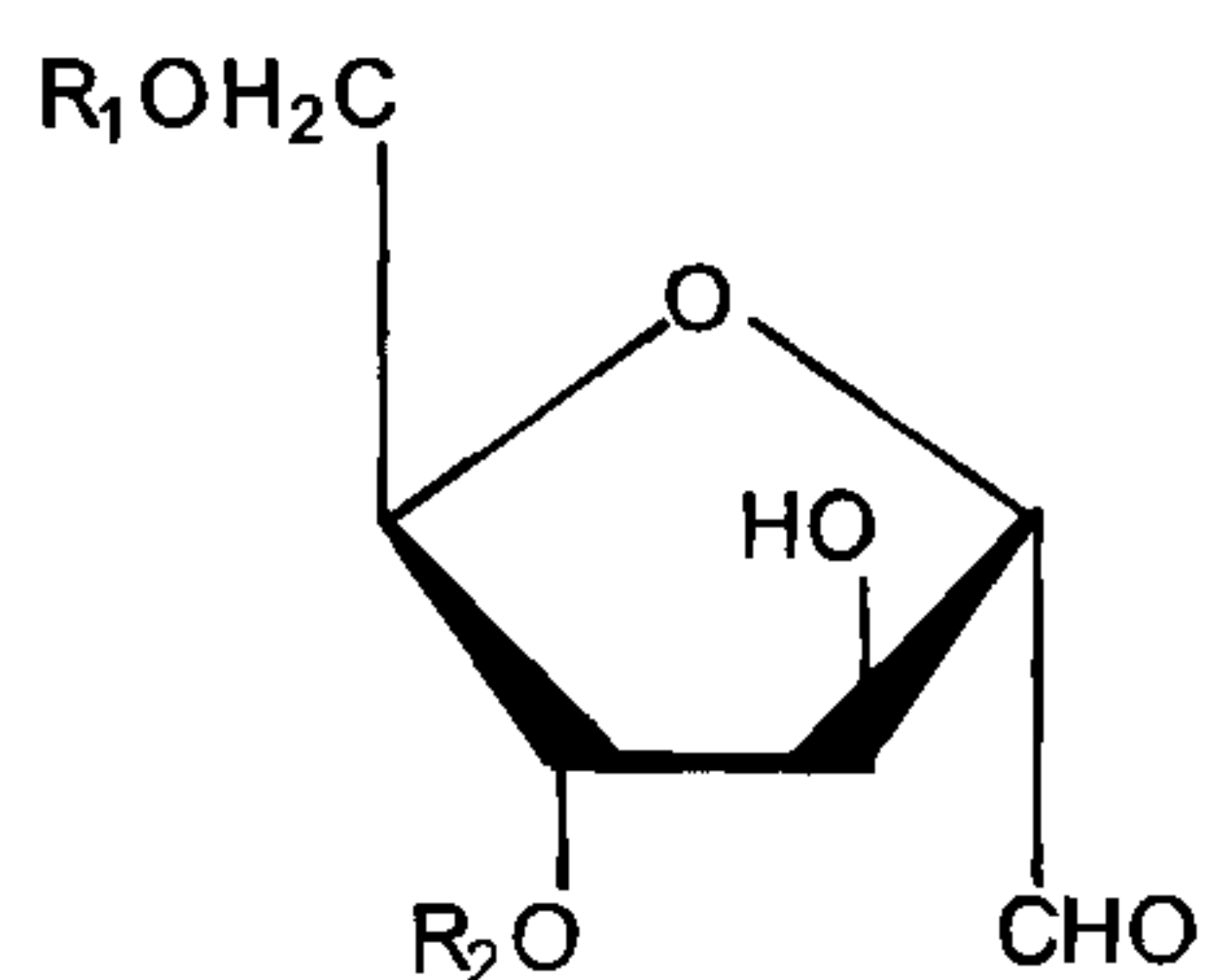
15 Although the invention has been described in conjunction with the specific embodiments, it is evident that many alternatives and variations will be apparent to those skilled in the art in light of the foregoing description. Accordingly, the invention is intended to embrace all of the alternatives and variations that fall
20 within the spirit and scope of the appended claims.

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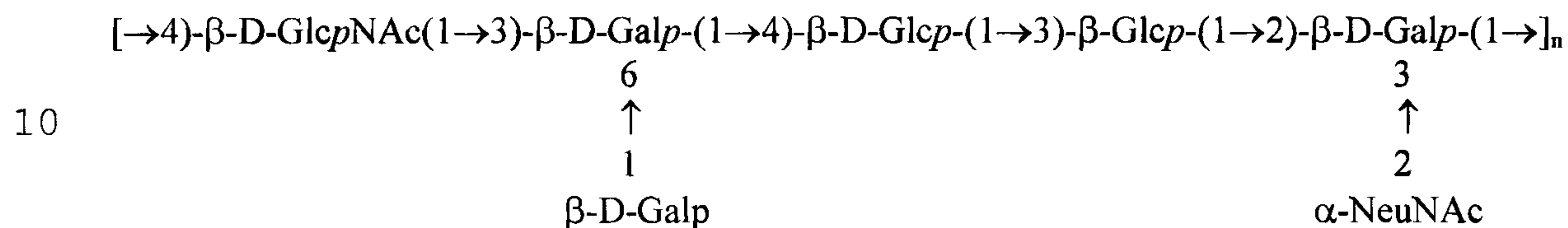
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CLAIMS:

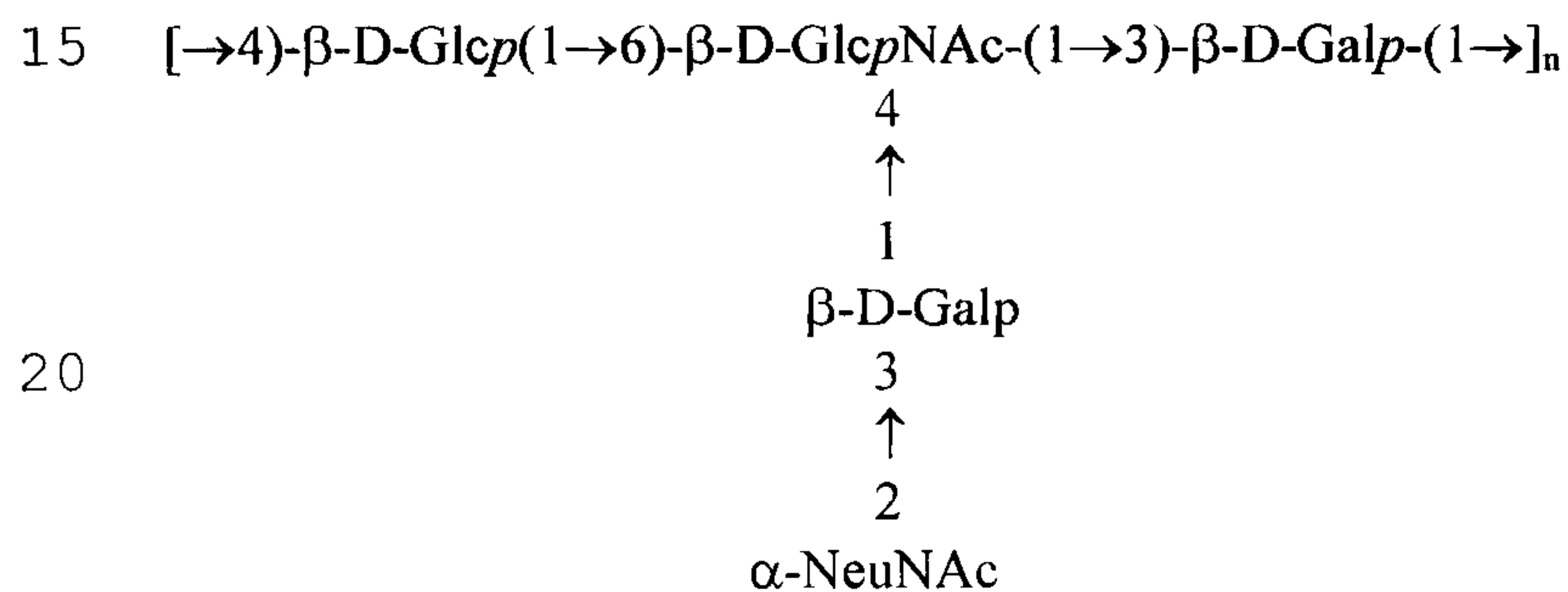
1. A process for depolymerizing Group B Streptococcus (GBS) type II and type III polysaccharides to produce fragments having the following 2,5-anhydro-D mannose reducing-end structure,



wherein R_1 is H and R_2 is sialylated heptasaccharide repeating-unit of formula



wherein n is 5 to 50 for the type II, and where R_1 is sialylated pentasaccharide repeating-units of formula



wherein n is 5 to 50 and R_2 is disaccharide $\alpha\text{NeuAc}(2\text{-}3)\beta\text{-D-Galp}1\text{-}$ for the type III,

said process comprising the steps of

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providing a GBS type II or GBS type III polysaccharide to be depolymerized and reacting the polysaccharide in an aqueous medium with a base to form a partially de-N-acetylated polysaccharide product,
5 depolymerizing the de-N-acetylated product with a nitrosation agent to form the GBS type II or type III fragments, and recovering the fragments.

2. The process according to claim 1 wherein the base is selected from the group consisting of sodium hydroxide,
10 potassium hydroxide, ammonium hydroxide, sodium carbonate, sodium bicarbonate, and hydrazine.

3. The process according to claim 1 or claim 2 wherein the nitrosation reagent is sodium nitrite or nitrous acid.

15 4. The process according to claim 1 wherein the base is sodium hydroxide and the nitrosation reagent is sodium nitrite.

5. The process according to claim 4 wherein the polysaccharide is obtained from GBS type II bacteria.

20 6. The process according to claim 5 wherein the resulting GBS type II fragments have a molecular weight of 5 kDa to 50 kDa.

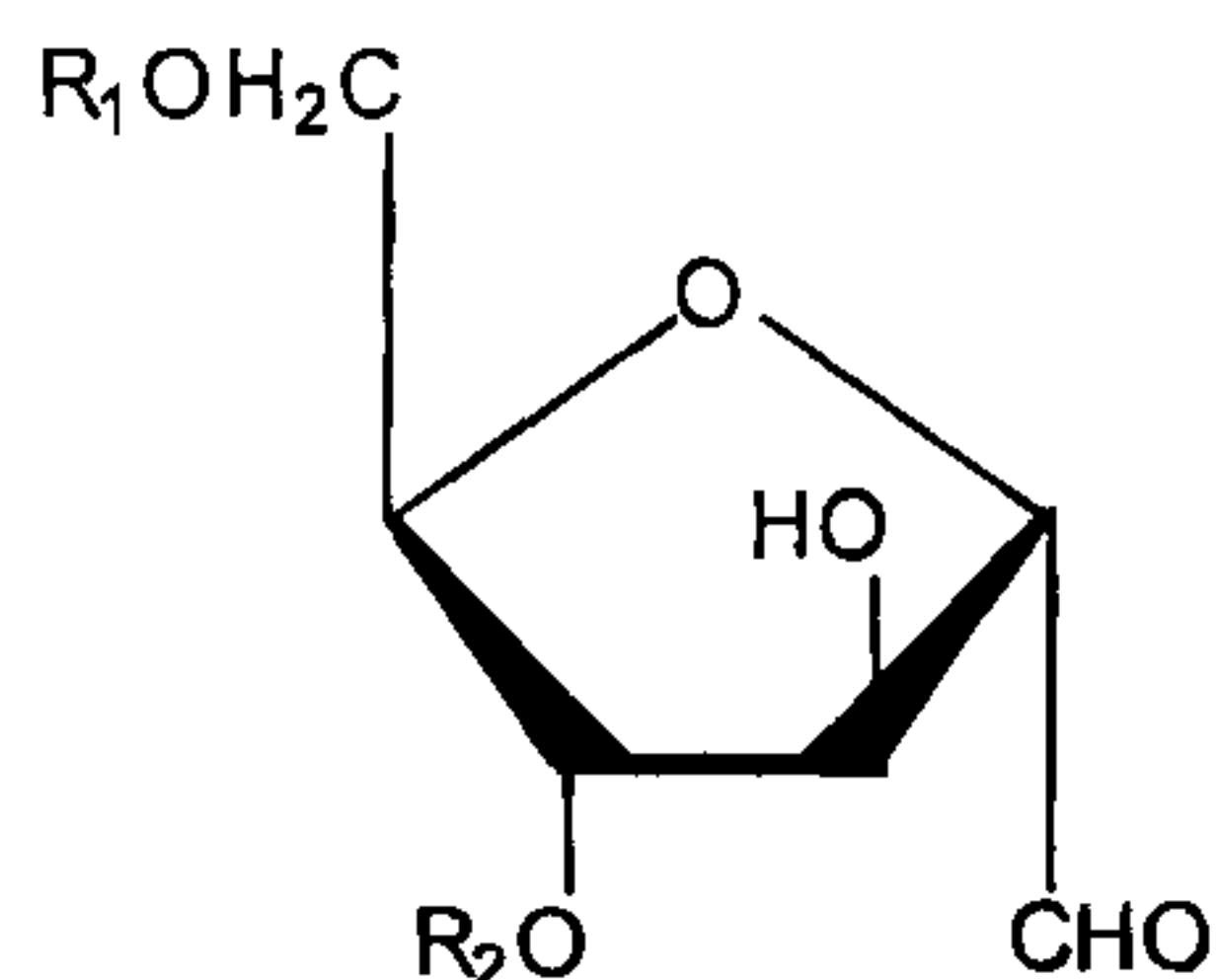
7. The process according to claim 4 wherein the polysaccharide is obtained from GBS type III bacteria.

25 8. The process according to claim 7 wherein the resulting fragments have a molecular weight of 5 kDa to 50 kDa.

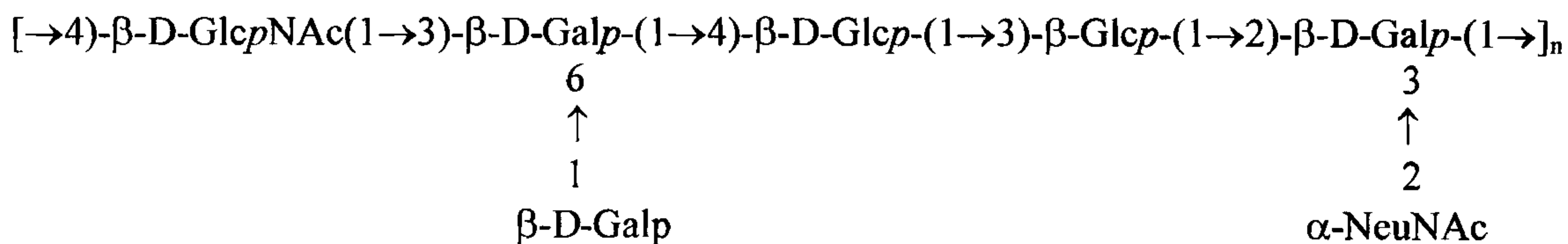
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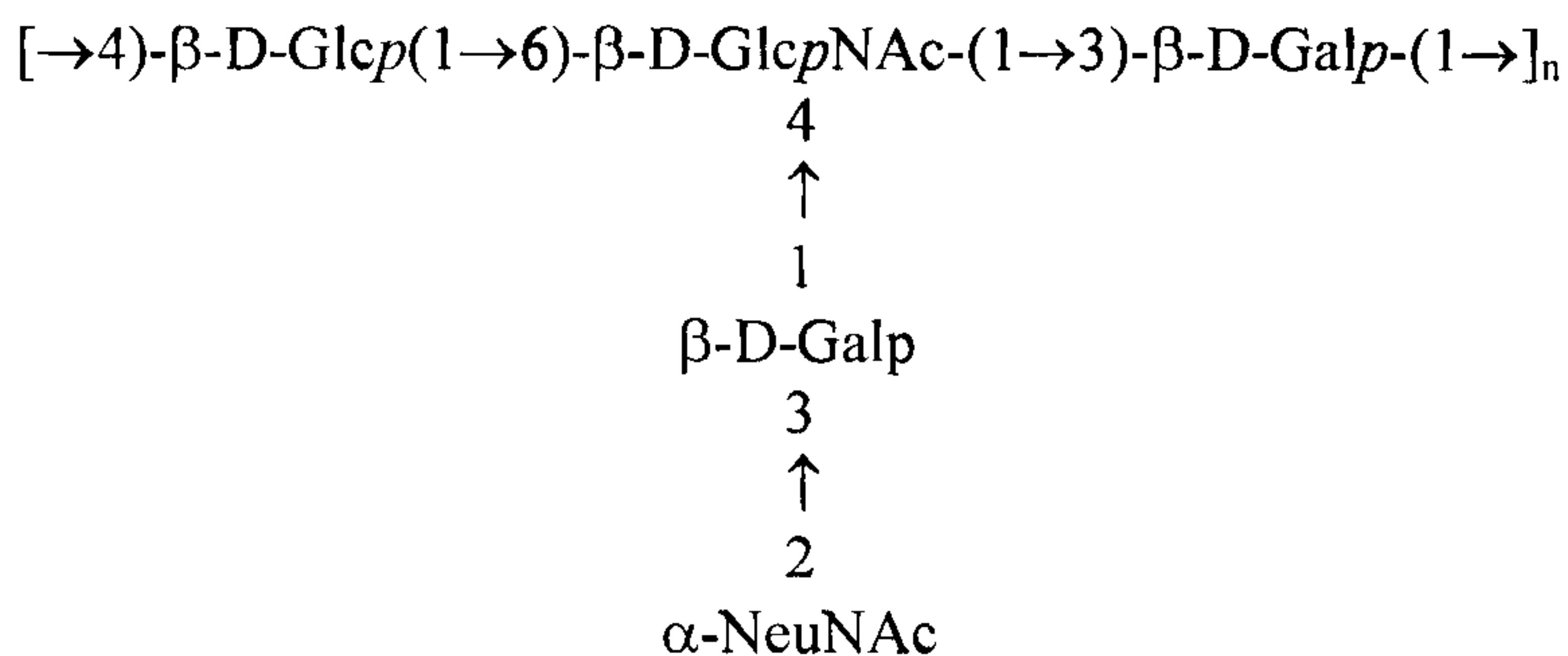
9. A GBS type II or type III polysaccharide fragment prepared according to the process of claim 1, wherein the fragment has the 2,5-anhydro-D mannose reducing-end structure,



5 wherein R_1 is H and R_2 is sialylated heptasaccharide repeating-unit of formula



wherein n is 5 to 50 for the type II, and where R_1 is sialylated pentasaccharide repeating-units of formula



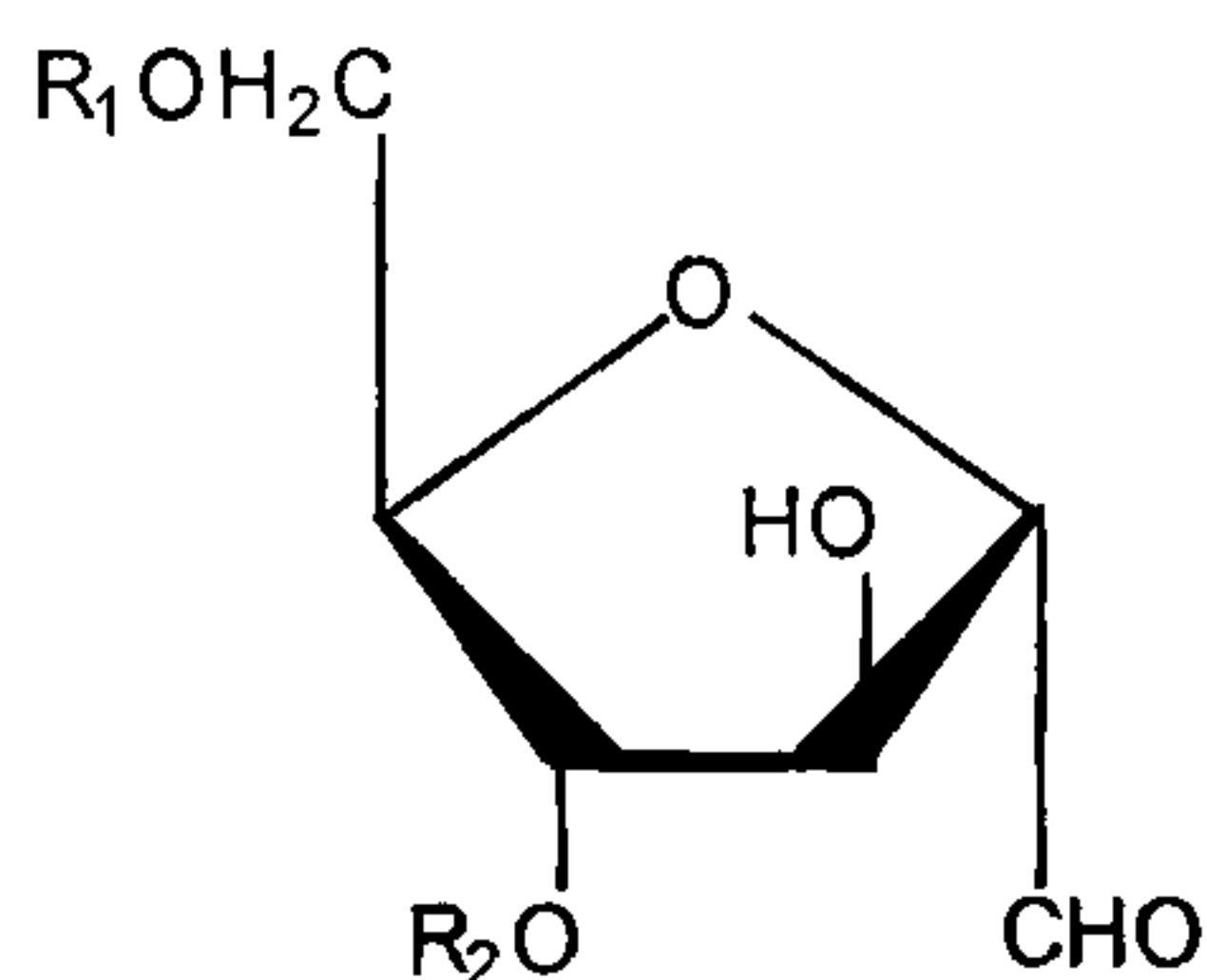
wherein n is 5 to 50 and R_2 is disaccharide $\alpha\text{NeuAc}(2\text{-}3)\beta\text{-D-Galp}1\text{-}$ for the type III.

10. The GBS type II or type III polysaccharide fragment according to claim 9 wherein the base for de-N-acetylation is sodium hydroxide and the nitrosation reagent is sodium nitrite.

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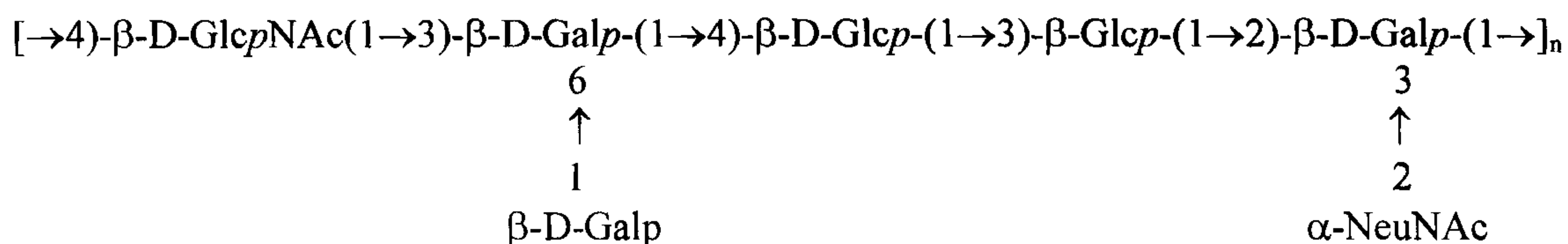
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11. A Group B Streptococcus (GBS) type II or type III polysaccharide fragment having the following 2,5-anhydro-D mannose reducing-end structure,

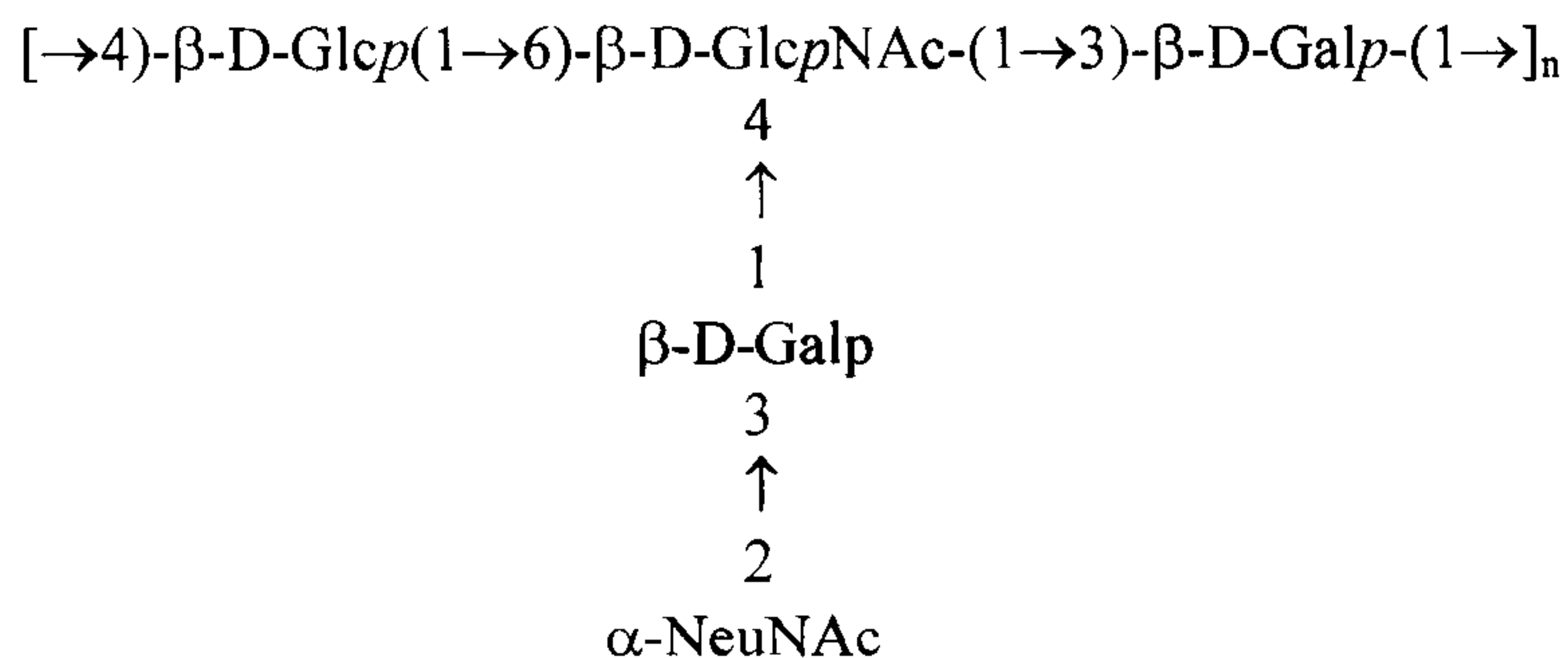


wherein R_1 is H and R_2 is sialylated

5 heptasaccharide repeating-unit of formula



wherein n is 5 to 50 for the type II, and where R_1 is sialylated pentasaccharide repeating-units of formula



wherein n is 5 to 50 and R_2 is disaccharide $\alpha\text{NeuAc}(2\text{-}3)\beta\text{-D-Galp}1\text{-}$ for the type III.

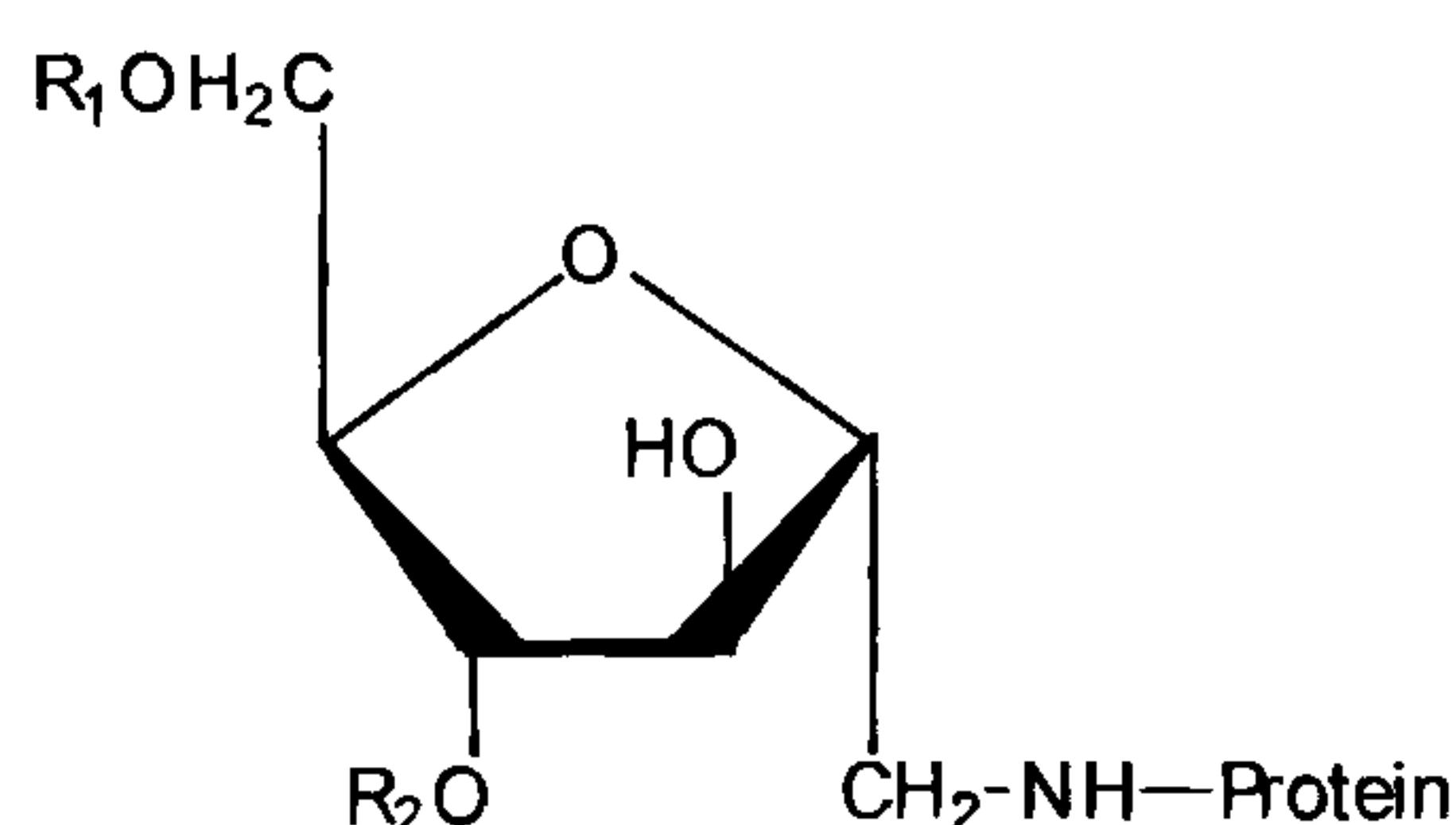
12. The GBS type II polysaccharide fragment according to claim 11 wherein the molecular weight is in the range of 5 kDa to 50 kDa.

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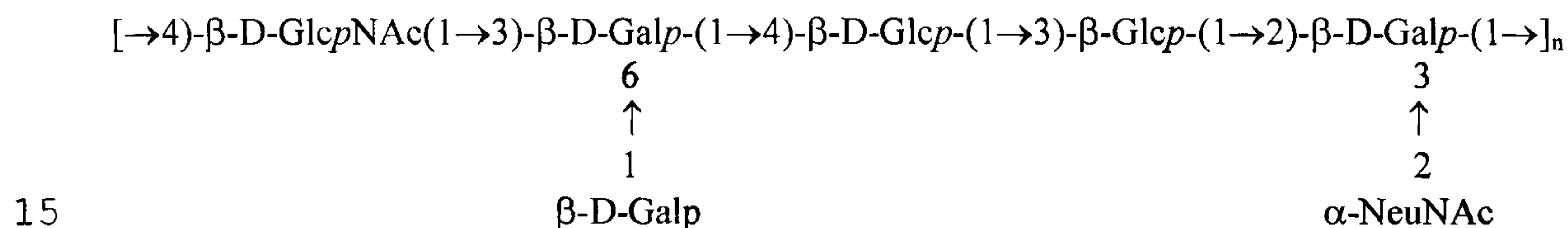
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13. The GBS type III polysaccharide fragment according to claim 11 wherein the molecular weight is in the range of 5 kDa to 50 kDa.

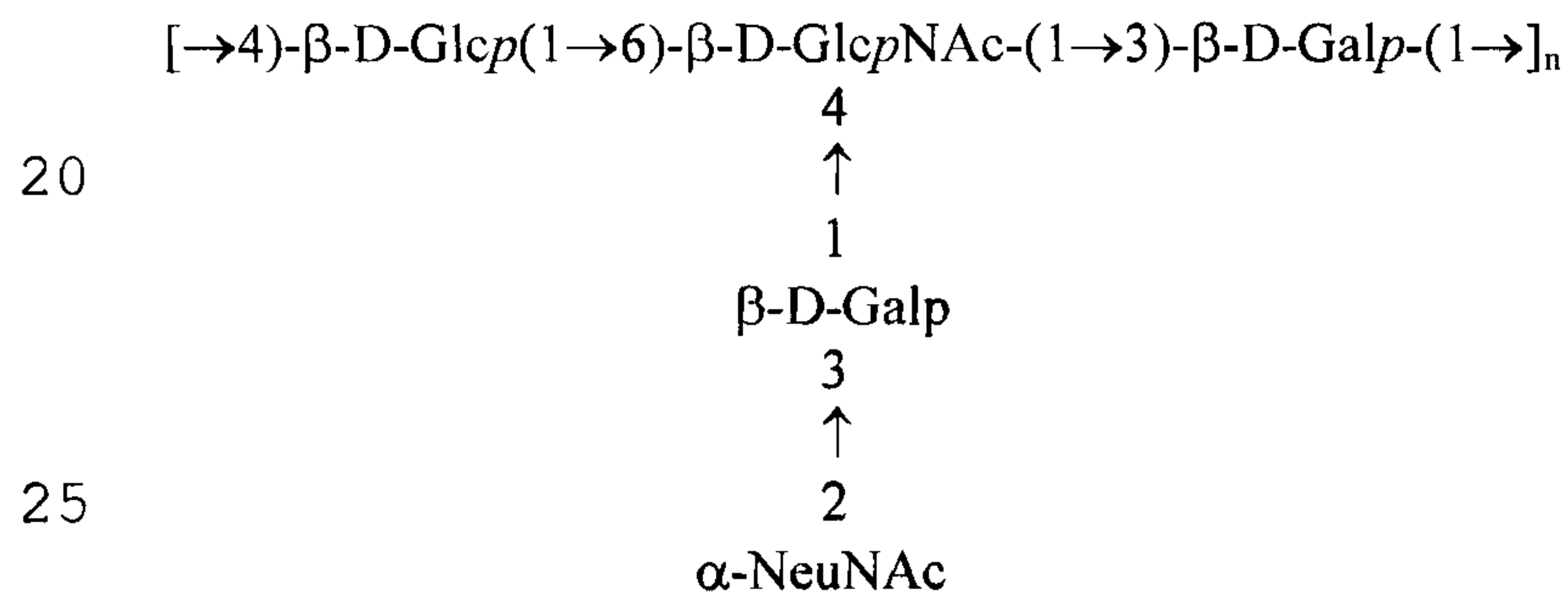
14. A conjugate molecule comprising at least one polysaccharide fragment selected from GBS type II and type III polysaccharide fragments, and wherein the fragment is covalently bound to a protein wherein the conjugate molecule has the structure



wherein R_1 is H and R_2 is sialylated
 10 heptasaccharide repeating-unit of formula



wherein n is 5 to 50 for the type II, and where R_1 is sialylated pentasaccharide repeating-units of formula



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wherein n is 5 to 50 and R₂ is disaccharide α NeuAc(2-3) β -D-Galp1- for the type III.

15. The conjugate molecule according to claim 14 wherein the protein is derived from a bacteria.

5 16. The conjugate molecule according to claim 15 wherein the protein is derived from a bacteria selected from the group consisting of tetanus toxoid, diphtheria toxoid, CRM₁₉₇, a recombinant non IgA binding protein of the β -C antigen of type Ia/Ib Group B streptococcus, and recombinant
10 class 3 outer membrane protein from Neisseria Meningitides.

17. The conjugate molecule according to claim 16 wherein the polysaccharide fragment is a GBS type II polysaccharide, the protein is tetanus toxoid and the molecular weight of the polysaccharide fragment is
15 between 5 kDa and 50 kDa.

18. The conjugate molecule according to claim 16 wherein the polysaccharide fragment is a GBS type III polysaccharide, the protein is tetanus toxoid and the molecular weight of the polysaccharide fragment is
20 between 5 kDa and 50 kDa.

19. The conjugate molecules according to claim 16 wherein the molar ratio of polysaccharide to protein is between 1 and 10.

20. The conjugate molecules according to claim 19
25 wherein the molar ratio of polysaccharide to protein is between 3 and 10.

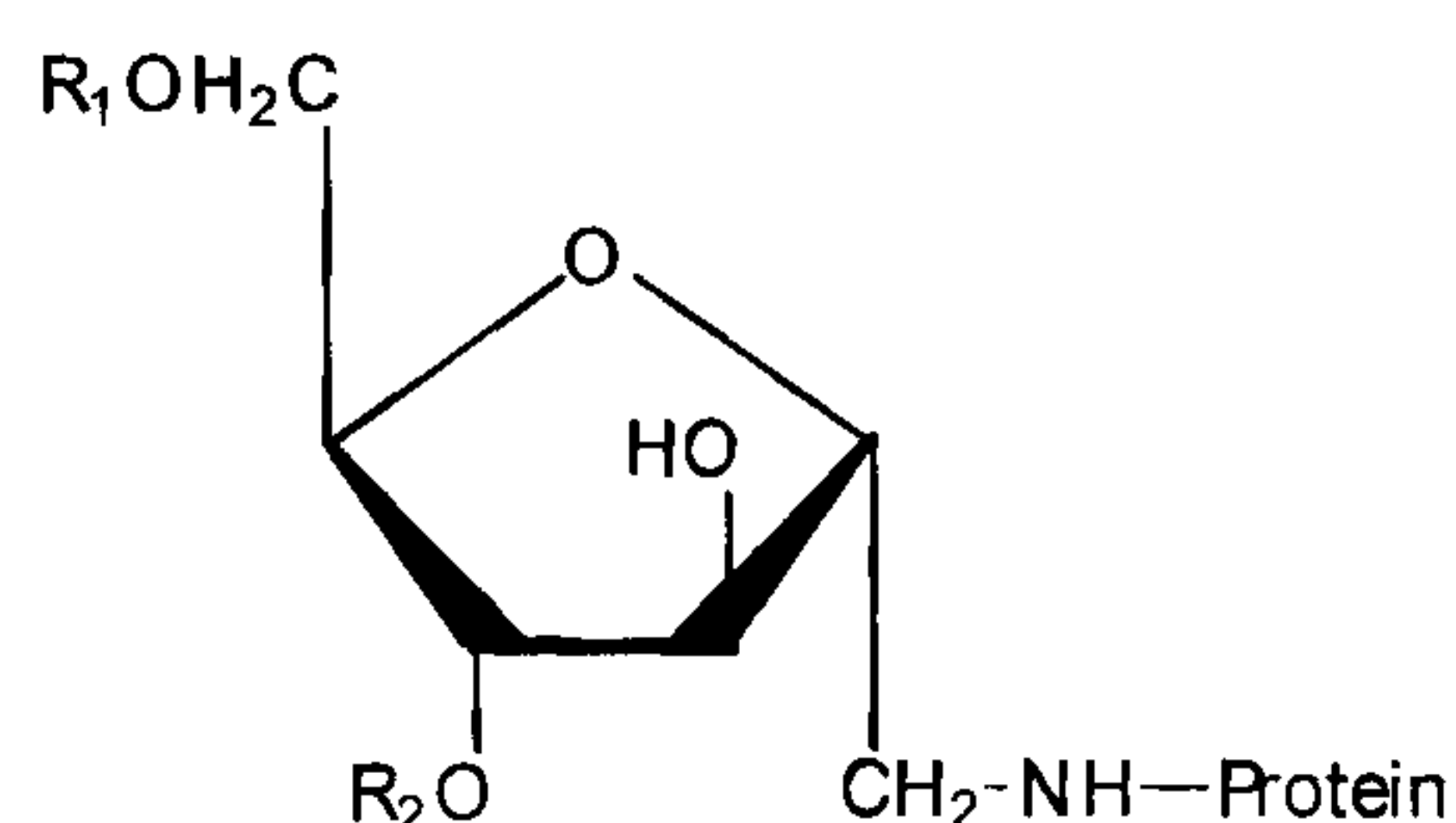
21. The conjugate molecule according to claim 14 comprising GBS type II and type III fragments.

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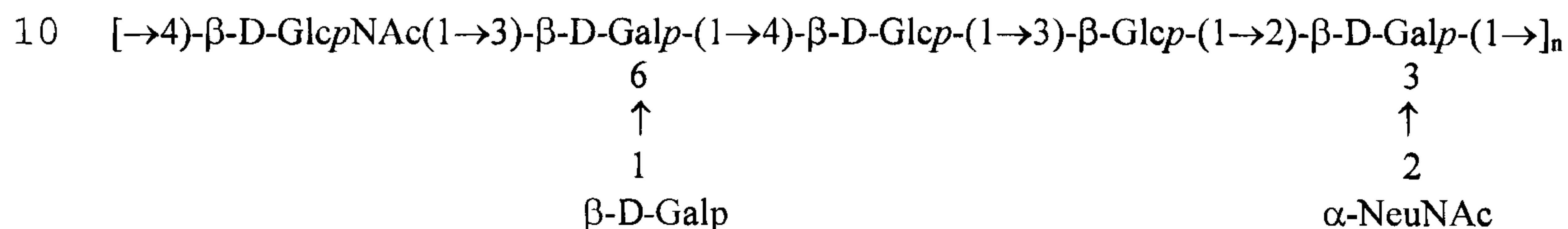
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22. The conjugate molecule according to claim 21 wherein the protein is a recombinant non-IgA binding protein of the β -C antigen of type Ia/Ib Group B streptococcus.

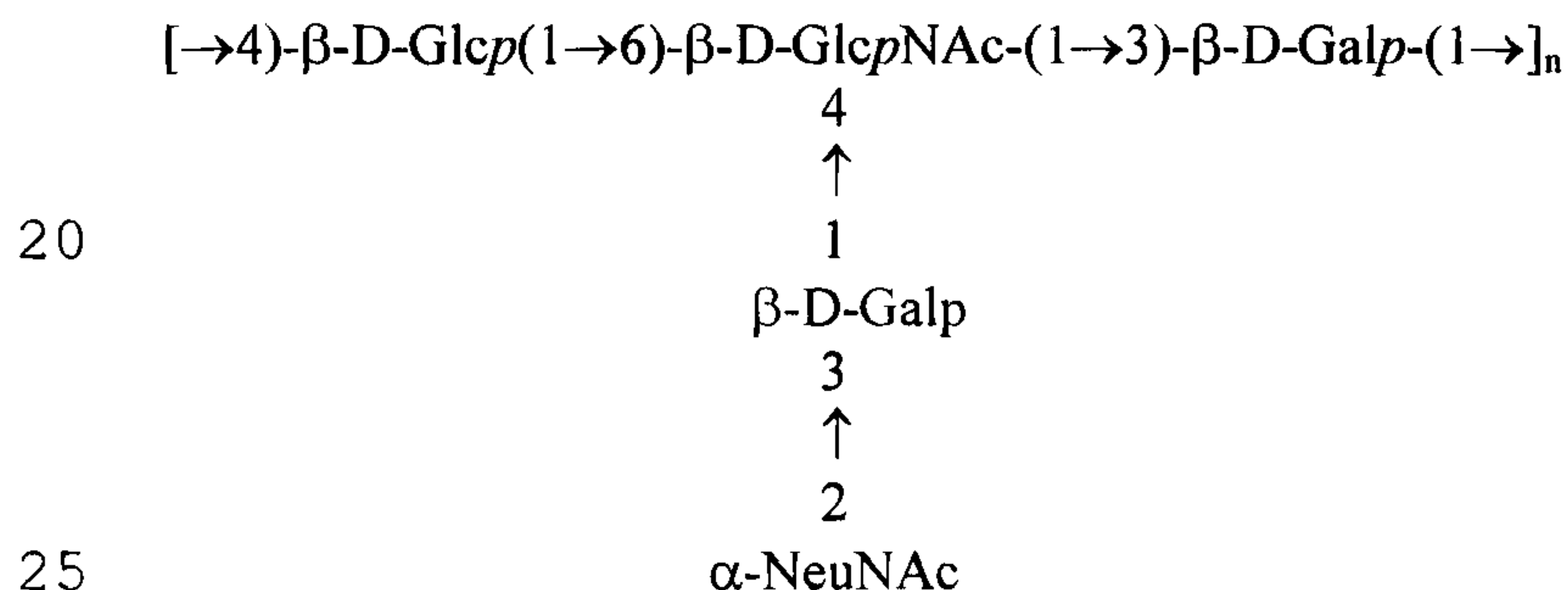
23. A vaccine comprising conjugate molecules
5 comprising a GBS type II or type III polysaccharide fragment or a combination thereof covalently bound to a protein wherein the conjugate molecule has the structure



wherein R_1 is H and R_2 is sialylated heptasaccharide repeating-unit of formula



15 wherein n is 5 to 50 for the type II, and where R_1 is sialylated pentasaccharide repeating-units of formula



wherein n is 5 to 50 and R_2 is disaccharide $\alpha\text{NeuAc}(2\rightarrow 3)\beta\text{-D-Galp}1\text{-}$ for the type III.

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24. The vaccine according to claim 23 wherein the fragment is a GBS type II polysaccharide and the molecular weight of the polysaccharide fragment is between 5 kDa and 50 kDa.

5 25. The vaccine according to claim 23 wherein the fragment is a GBS type III polysaccharide and the molecular weight of the polysaccharide fragment is between 5 kDa and 50 kDa.

26. The vaccine according to claim 23 comprising
10 conjugates with GBS type II and III polysaccharide fragments.

27. The vaccine according to any one of claims 23 to 26 wherein the protein component is derived from a bacteria selected from the group consisting of tetanus
15 toxoid, diphtheria toxoid, CRM₁₉₇, a recombinant non IgA binding protein of the β -C antigen of type Ia/Ib Group B streptococcus, and recombinant class 3 outer membrane protein from Neisseria Meningitides.

28. The vaccine according to claim 27 wherein the
20 protein is a recombinant non IgA binding protein of the β -C antigen of type Ia/Ib Group B streptococcus, and recombinant class 3 outer membrane protein from Neisseria Meningitides.

29. An immune serum comprising antibodies raised in a mammal immunized with the conjugate according to any one of
25 claims 14 to 22.

30. The immune serum according to claim 29 wherein the polysaccharides of the conjugate are fragments of GBS type II polysaccharides.

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31. The immune serum according to claim 29 wherein the polysaccharides of the conjugate are fragments of GBS type III polysaccharides.

32. Use of the vaccine according to any one of
5 claims 23 to 28 to immunize a mammal against GBS type II or type III infection.

33. Use of the vaccine according to any one of claims 23 to 28 to immunize a pregnant woman or neonate against GBS type II or type III infection.

10 34. A commercial package comprising the vaccine according to any one of claims 23 to 28, together with instructions for its use in immunizing a mammal against GBS type II or type III.

35. A method of separating GBS type II or III
15 antibodies from serum comprising immobilizing a polysaccharide fragment prepared according to claim 1 to a solid support, combining the solid support with bound polysaccharide with serum under conditions to allow binding of GBS type II or III antibodies to the bound polysaccharide
20 fragment, and separating the remaining serum from the solid support.

36. The method according to claim 35 wherein the polysaccharide fragment is GBS type II.

37. The method according to claim 35 wherein the
25 polysaccharide fragment is GBS type III.

38. An immunoassay reagent comprising a GBS type II or III polysaccharide fragment prepared according to the method of claim 1 wherein the polysaccharide fragment is immobilized on a solid support.

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39. The immunoassay reagent according to claim 38 wherein the polysaccharide fragment is GBS type II.

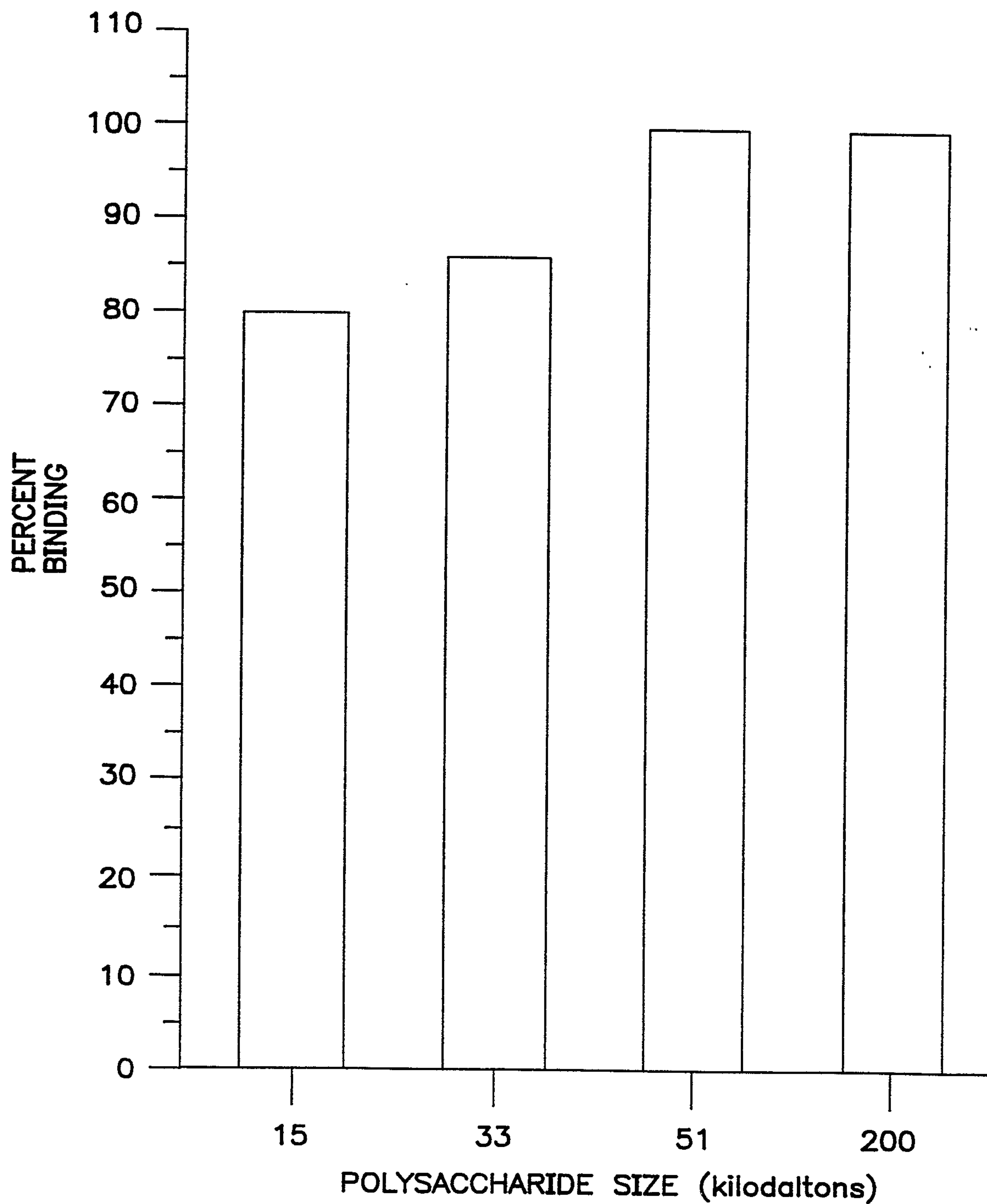
40. The immunoassay reagent according to claim 38 wherein the polysaccharide fragment is GBS type III.

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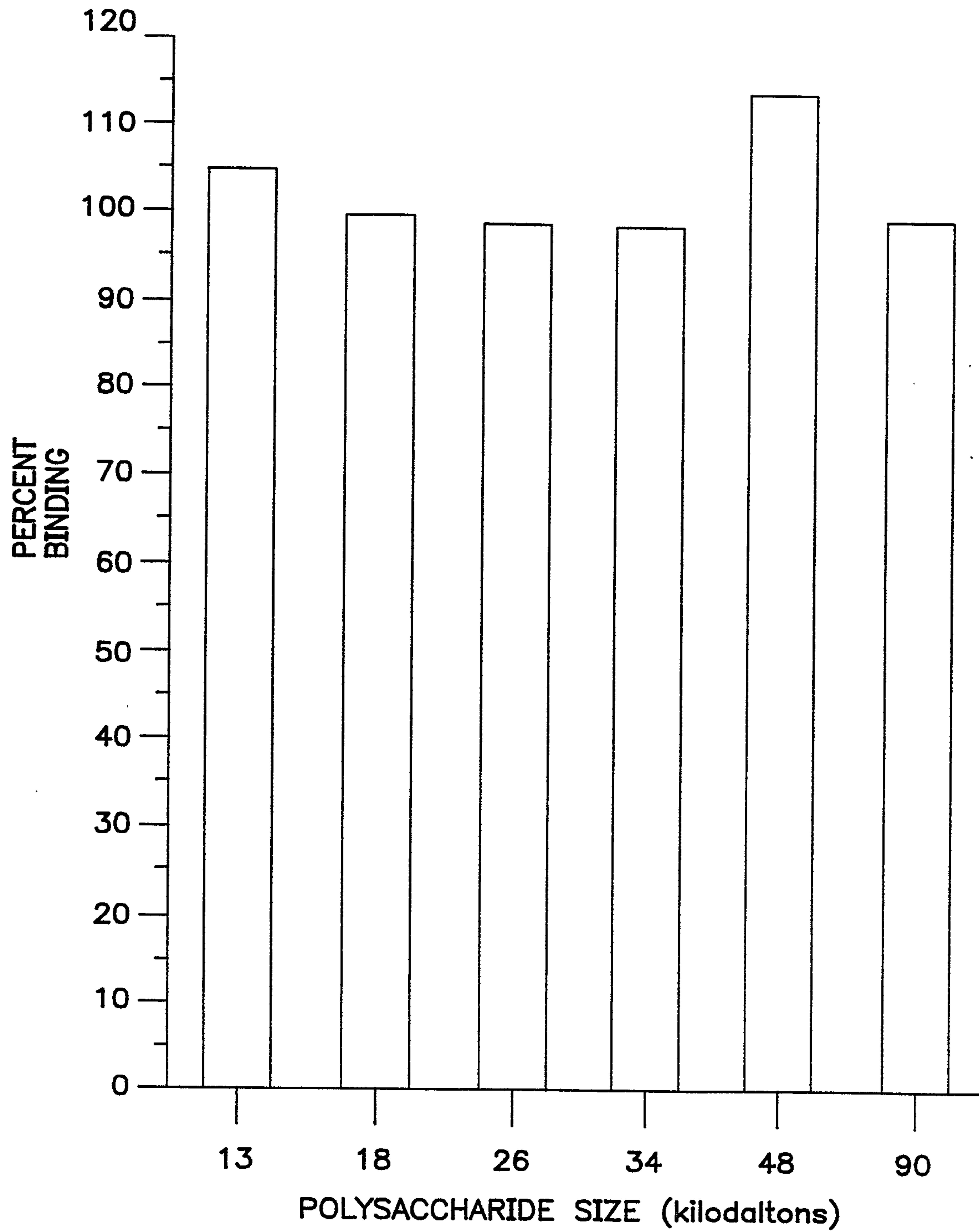
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**FIG. 1**

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**FIG. 2**

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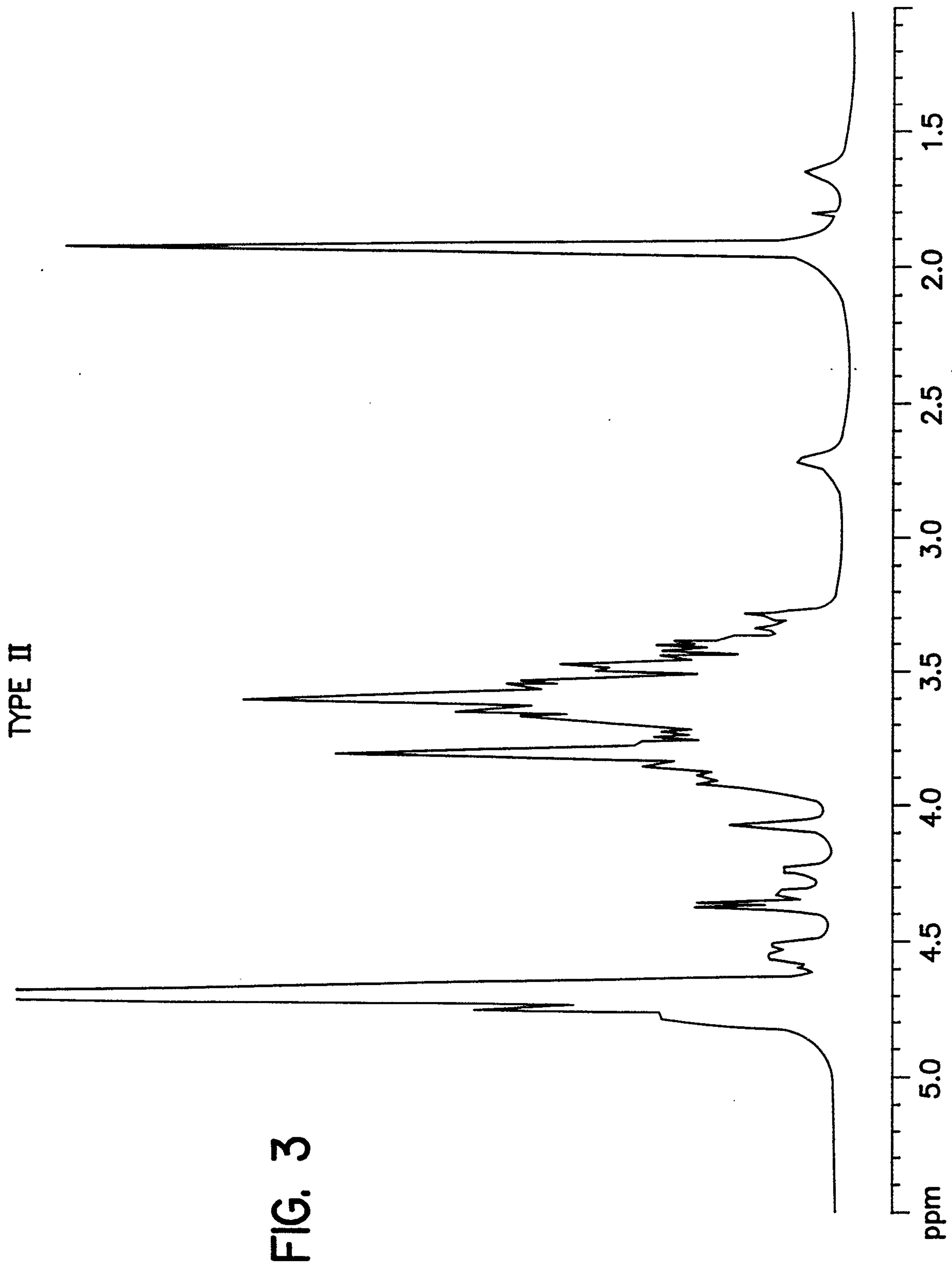


FIG. 3
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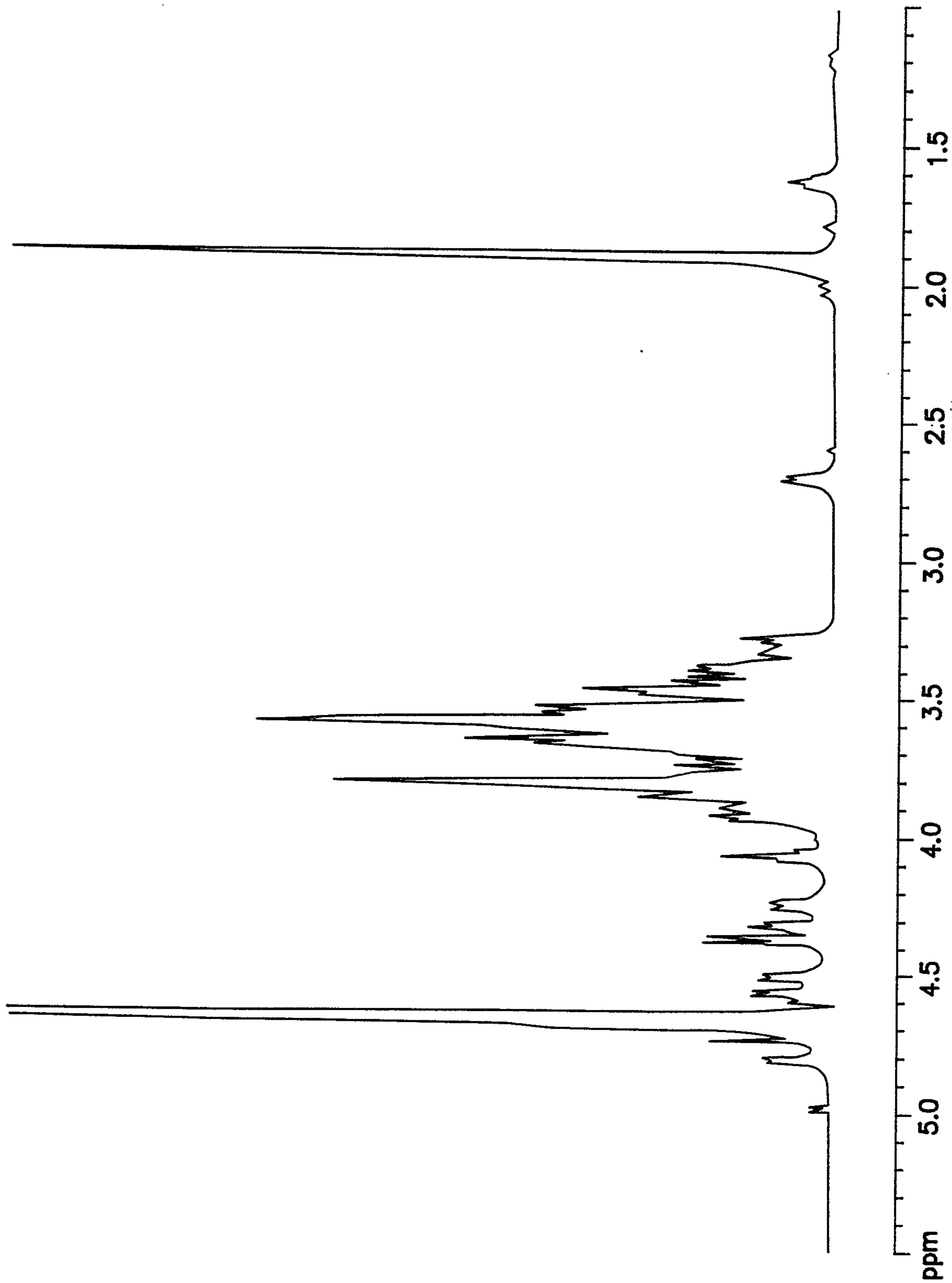


FIG. 4

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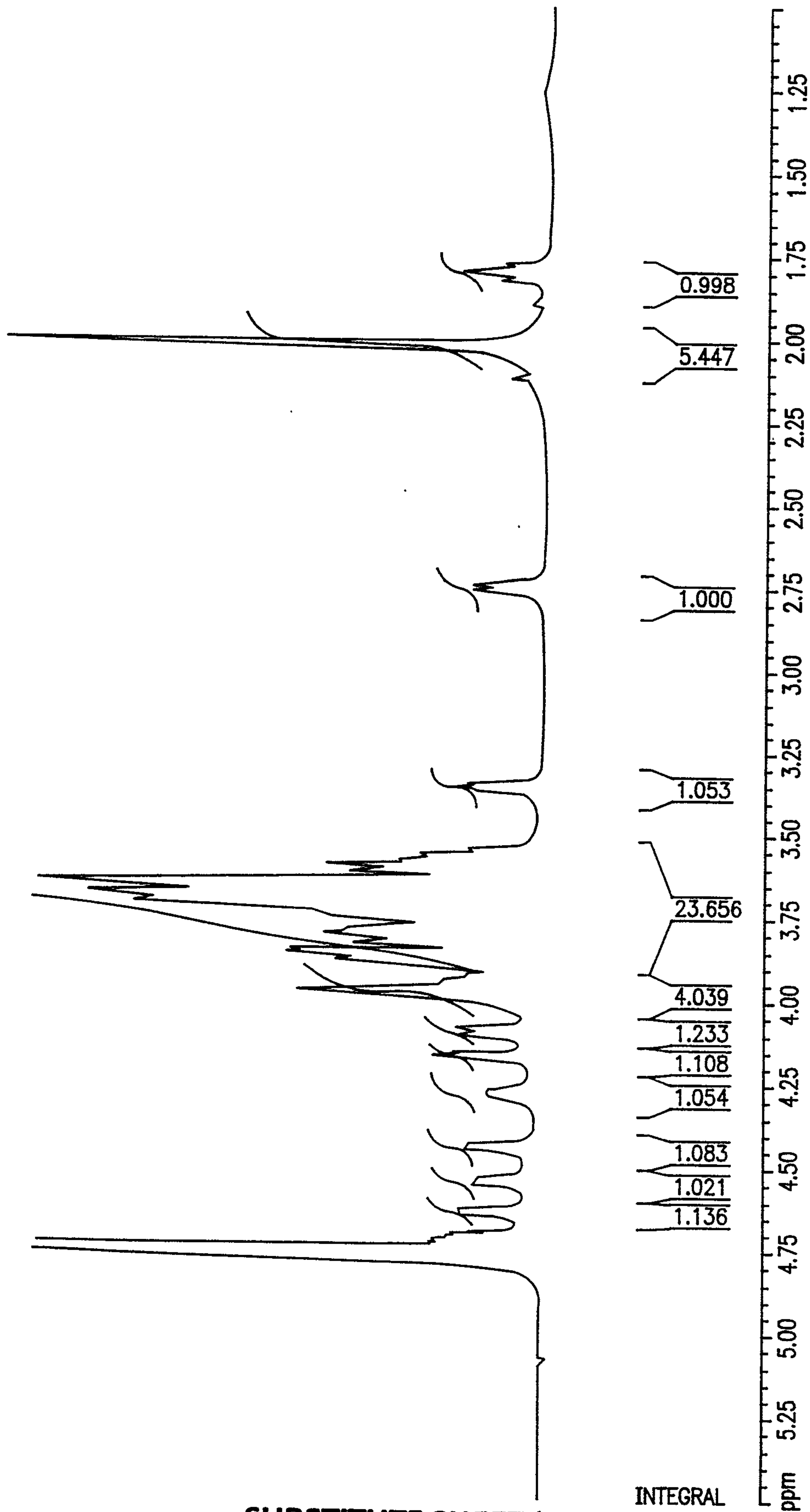
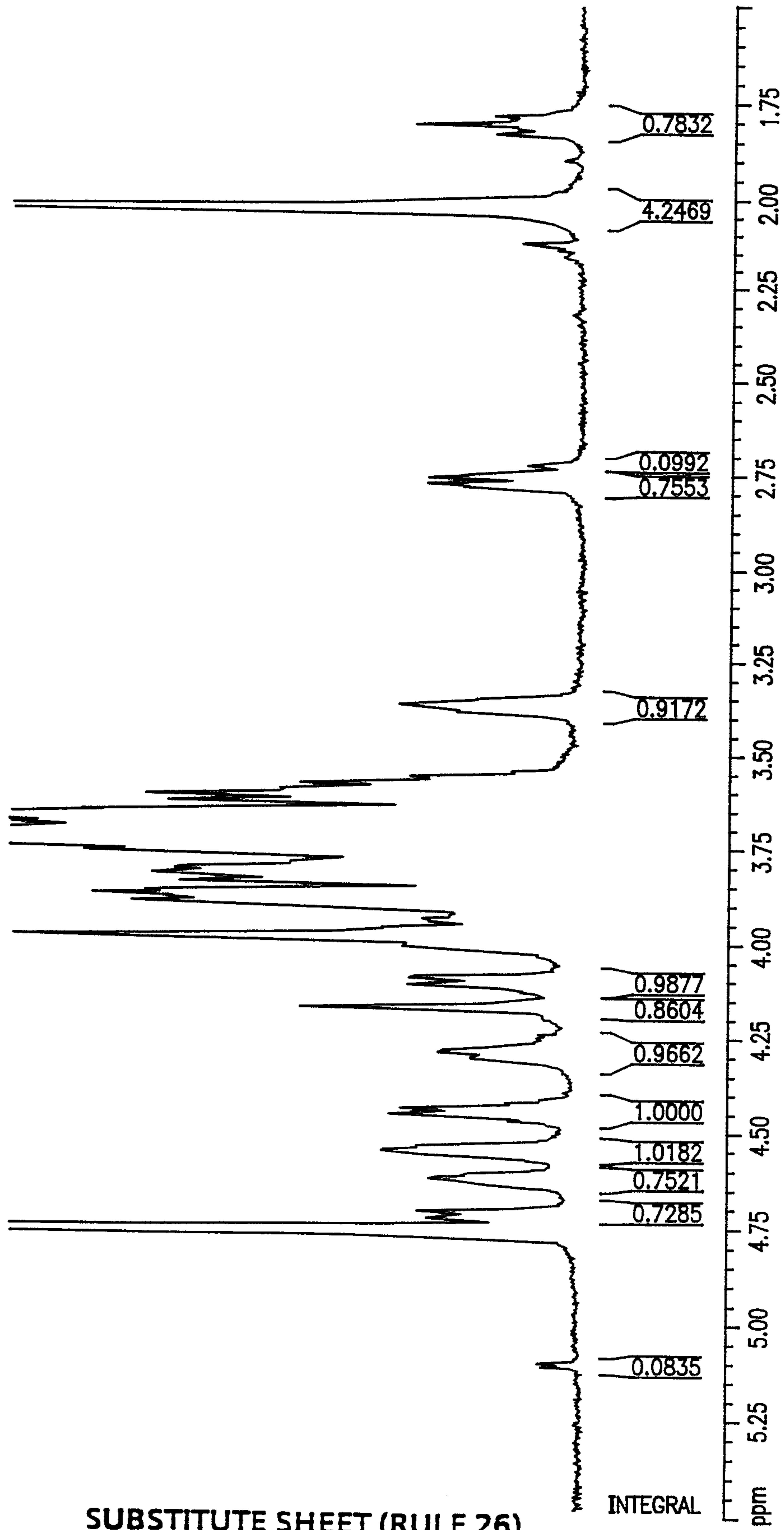


FIG. 5

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FIG. 6