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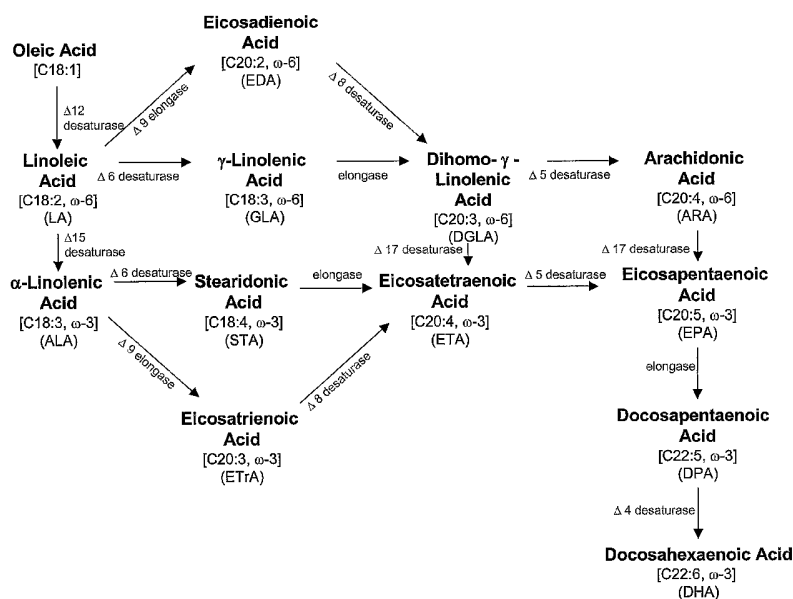
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(54) Title: PRODUCTION OF γ -LINOLENIC ACID IN OLEAGINOUS YEAST



(57) Abstract: The present invention relates to methods for the production of γ -linolenic acid (GLA) in oleaginous yeast. Thus, $\Delta 12$ and $\Delta 6$ desaturases able to catalyze the conversion of oleic acid to GLA have been introduced into the genome of *Yarrowia*, using zeta-directed integration. Transformed strains produced over 25% GLA in the total lipids, as opposed to wildtype *Yarrowia* that is unable to synthesize any GLA. Metabolic engineering and fermentation methods are provided to further enhance GLA productivity in oleaginous yeast.

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For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

TITLE

PRODUCTION OF γ -LINOLENIC ACID IN OLEAGINOUS YEAST

This application claims the benefit of U.S. Provisional Application No. 60/600270, filed August 10, 2004.

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FIELD OF THE INVENTION

This invention is in the field of biotechnology. More specifically, this invention pertains to the production of γ -linolenic acid in oleaginous yeast.

BACKGROUND OF THE INVENTION

Gamma-linolenic acid (GLA; γ -linolenic acid; *cis*-6, 9, 12-octadecatrienoic acid) is an important intermediate in the biosynthesis of biologically active prostaglandin from linoleic acid (LA). Additionally, GLA is recognized as an essential ω -6 polyunsaturated fatty acid (PUFA) having tremendous clinical and pharmaceutical value. For example, GLA has been shown to reduce increases in blood pressure associated with stress and to improve performance on arithmetic tests. GLA and dihomo- γ -linolenic acid (DGLA, another ω -6 PUFA) have been shown to inhibit platelet aggregation, cause vasodilation, lower cholesterol levels and inhibit proliferation of vessel wall smooth muscle and fibrous tissue (Brenner et al., *Adv. Exp. Med. Biol.* 83:85-101 (1976)). Administration of GLA or DGLA, alone or in combination with eicosapentaenoic acid (EPA, an ω -3 PUFA), has been shown to reduce or prevent gastrointestinal bleeding and other side effects caused by non-steroidal anti-inflammatory drugs (U.S. 4,666,701). Further, GLA and DGLA have been shown to prevent or treat endometriosis and premenstrual syndrome (U.S. 4,758,592) and to treat myalgic encephalomyelitis and chronic fatigue after viral infections (U.S. 5,116,871). Additionally, GLA is also used in products relating to functional foods (nutriceuticals), infant nutrition, bulk nutrition, cosmetics and animal health.

Although seeds of many plants contain GLA, the most common commercial sources of GLA are evening primrose (*Oenothera biennis* L.; containing 5-9% GLA of total seed oil), black currant (*Ribes nigrum*; containing 14% GLA of total seed oil) and borage (*Borago officinalis* L.; containing 17-25% GLA from a total seed oil content of 28-38%) (Simon, J.E. et al., In *Advances In New Crops*; Janick, J. and J.E. Simon, Eds.; Timber: Portland, OR (1990); p 528). There are several disadvantages associated with production of GLA from these sources, however. First, plant oils tend to have highly heterogeneous oil compositions that can

require extensive purification to separate or enrich one or more of the desired PUFAs. Natural sources are also subject to uncontrollable fluctuations in availability (e.g., due to weather or disease); and, crops that produce PUFAs often are not competitive economically with hybrid crops developed for food production. Specific challenges associated with large-scale commercial production of borage are encountered because of the plants' indeterminate vegetative growth, lack of concentrated flowering and seed set, non-uniform seed maturation, susceptibility to a wide range of insects and disease pests, and poor ability to compete with weeds.

To overcome these problems, microorganisms have been investigated as an alternative source of GLA and other PUFAs. Specifically, many microorganisms (including algae, bacteria, molds and yeast) can synthesize oils in the ordinary course of cellular metabolism. Thus, oil production involves cultivating the microorganism in a suitable culture medium to allow for oil synthesis, followed by separation of the microorganism from the fermentation medium and treatment for recovery of the intracellular oil. Microbial production of GLA has been investigated, although these efforts have been minimal in comparison to the work focused on the microbial production of longer-chain PUFAs such as arachidonic acid (ARA), EPA and docosahexaenoic acid (DHA). Some suitable strains have been proposed as perspective GLA producers, including *Mortierella ramanniana*, *Mucor* sp., *Cunninghamella japonica* and *Entomophthora exitalis*. Currently, however, only Japan is producing GLA commercially using the fungus *Mortierella* (Ratledge, C. *Trends Biotech.* 11:278-284 (1993)). Thus, there remains a need for an appropriate microbial system for economical production of commercial quantities of GLA.

One class of microorganisms that has not been previously examined as a production platform for PUFAs such as GLA are the oleaginous yeast. These organisms can accumulate oil up to 80% of their dry cell weight. The technology for growing oleaginous yeast with high oil content is well developed (for example, see EP 0 005 277B1; Ratledge, C., *Prog. Ind. Microbiol.* 16:119-206 (1982)) and may offer a cost advantage compared to commercial micro-algae fermentation for production of PUFAs. Whole yeast cells may also represent a convenient way of encapsulating PUFA-enriched oils for use in functional foods and animal feed supplements.

Despite the advantages noted above, oleaginous yeast are naturally deficient in PUFAs, since naturally produced PUFAs in these organisms are limited to 18:2 acids (and, less commonly, 18:3 acids). Thus, the problem to be solved is to develop an oleaginous yeast that accumulates oils enriched in GLA. Toward this end, it is necessary to introduce desaturases that allow for the synthesis and accumulation of GLA in oleaginous yeast. Although advances in the art of genetic engineering have been made, such techniques have not been developed or optimized for oleaginous yeast; and, one must overcome problems associated with the use of these particular host organisms for the production of GLA.

Applicants have solved the stated problem by engineering strains of *Yarrowia lipolytica* that are capable of producing over 25% and 34% GLA in the total lipids, respectively. Additional metabolic engineering and fermentation methods are provided to further enhance GLA productivity in oleaginous yeast.

SUMMARY OF THE INVENTION

The present invention provides methods for the expression of enzymes comprising a γ -linolenic acid (GLA) biosynthetic pathway in an oleaginous yeast host for the production of γ -linolenic acid. Accordingly the invention provides A method for producing γ -linolenic acid comprising:

- a) providing an oleaginous yeast comprising:
 - i) at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity;
 - ii) at least one genetic construct encoding a polypeptide having a $\Delta 12$ desaturase activity; and,
 - iii) a source of desaturase substrate selected from the group consisting of oleic acid and linoleic acid;
- b) growing the yeast of step (a) in the presence of a fermentable carbon source whereby at least about 15% γ -linolenic acid in the total lipids is produced; and
- c) optionally recovering the γ -linolenic acid.

Additionally the method makes use of a host cell optionally comprising a) at least one genetic construct encoding $\Delta 9$ desaturase polypeptide; b) at least one genetic construct encoding $C_{14/16}$ elongase polypeptide; and, c) at least one genetic construct encoding $C_{16/18}$

elongase polypeptide. Host cells preferred in the present invention are strains of *Yarrowia lipolytica*.

In a preferred embodiment the invention provides a method for producing γ -linolenic acid comprising:

- 5 a) providing a *Yarrowia lipolytica* comprising:
- i) at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity;
 - ii) at least one genetic construct encoding a polypeptide having a $\Delta 12$ desaturase activity; and,
 - 10 iii) a source of desaturase substrate selected from the group consisting of: oleic acid and linoleic acid;
- wherein the at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity and the at least one genetic construct encoding a polypeptide having a $\Delta 12$ desaturase activity are in multicopy and are integrated into a genetic locus selected from the group consisting of the *zeta* loci, the *Ura3* loci, the *Leu 2* loci, the *Lys5* loci, the *Aco2* loci, the *Pox3* loci, the *Lip1* loci and the *Lip2* loci of the *Yarrowia lipolytica*, and
- 15 wherein the at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity and the at least one genetic construct encoding a polypeptide having a $\Delta 12$ desaturase activity are under the control of a strong promoter isolated from genes selected from the group consisting of: alcohol dehydrogenase, glyceraldehyde-3-phosphate-dehydrogenase, phosphoglycerate mutase, fructose-bisphosphate aldolase, glycerol-3-phosphate o-acyltransferase, phosphoglucose-isomerase, phosphoglycerate kinase, acid phosphatase, lactase, metallothionein and glucoamylase;
- 20 b) growing the *Yarrowia lipolytica* of step (a) in the presence of a fermentable carbon source whereby at least about 15% γ -linolenic acid in the total lipids is produced; and,
- 25 c) optionally recovering the γ -linolenic acid.

- In another embodiment the invention provides a production host cell
- 35 for the production of γ -linolenic acid comprising:
- a) a background *Yarrowia* sp. comprising:

- i) at least one genetic construct encoding $\Delta 12$ desaturase polypeptide; and,
 - ii) at least one genetic construct encoding $\Delta 6$ desaturase peptide;
- 5 wherein at least one of said genetic constructs of (i) and (ii) is over-expressed and whereby at least about 15% γ -linolenic acid in the total lipids is produced.

BRIEF DESCRIPTION OF THE DRAWINGS AND SEQUENCE DESCRIPTIONS

- 10 Figure 1 illustrates the ω -3/ ω -6 fatty acid biosynthetic pathway.
Figure 2 illustrates the construction of intermediate plasmid vector pYZ2A.
- Figure 3 provides plasmid maps for the following: (A) pYD6S; (B) pY25-d12d; (C) pY25-d12d-PB; (D) pDMW208; and (E) pYSD17SCP.
- 15 Figure 4 provides plasmid maps for the following: (A) pYTS6; (B) pDMW208D6; (C) pYZGDG; (D) pELS; and (E) pY5-13.
- Figure 5 illustrates the construction of plasmid vector pDMW208D62 for GLA expression in *Yarrowia lipolytica*.
- Figure 6A, 6B and 6C are chromatograms illustrating the production
20 of GLA in wildtype *Y. lipolytica*, *Y. lipolytica* transformed with plasmid pDMW208 and *Y. lipolytica* transformed with plasmid pDMW208D62, respectively.
- The invention can be more fully understood from the following detailed description and the accompanying sequence descriptions, which
25 form a part of this application.
- The following sequences comply with 37 C.F.R. §1.821-1.825 ("Requirements for Patent Applications Containing Nucleotide Sequences and/or Amino Acid Sequence Disclosures - the Sequence Rules") and are consistent with World Intellectual Property Organization (WIPO) Standard
30 ST.25 (1998) and the sequence listing requirements of the EPO and PCT (Rules 5.2 and 49.5(a-bis), and Section 208 and Annex C of the Administrative Instructions). The symbols and format used for nucleotide and amino acid sequence data comply with the rules set forth in 37 C.F.R. §1.822.
- 35 SEQ ID NOs:1-4 correspond to primers YL127, YL128, YL129 and YL130, respectively, used for amplification of the *zeta* regions of *Y. lipolytica*.

SEQ ID NO:5 is a 265 bp 5'-sequence from the *Y. lipolytica* *zeta* region; in contrast, SEQ ID NO:6 is a 359 bp 3'-sequence from the *Y. lipolytica* *zeta* region. Together, these sequences are useful to direct integration of expression cassettes into the *zeta* loci of the *Yarrowia* genome.

SEQ ID NO:7 is the DNA sequence of the synthetic $\Delta 6$ desaturase gene codon-optimized for expression in *Y. lipolytica*, while SEQ ID NO:8 is the corresponding amino acid sequence.

SEQ ID NO:9 is the DNA sequence of a ~1.95 kB fragment comprising the synthetic $\Delta 6$ desaturase gene codon-optimized for expression in *Y. lipolytica*, flanked by a TEF promoter and an XPR terminator.

SEQ ID NOs:10 and 11 correspond to primers YL238 and YL239, respectively, used for site-directed mutagenesis to generate pDMW200-B.

SEQ ID NOs:12-15 correspond to primers YL242, YL243, YL240 and YL241, respectively, used for site-directed mutagenesis to generate pY25-d12d-PB.

SEQ ID NO:16 is the DNA sequence of a ~1.9 kB fragment comprising the *Y. lipolytica* $\Delta 12$ desaturase gene flanked by a TEF promoter and an XPR terminator.

SEQ ID NO:17 is the DNA sequence of GLA expression vector pDMW208.

SEQ ID NOs:18 and 19 correspond to primers YL253 and YL254, respectively, used for amplification of the *Saprolegnia diclina* $\Delta 6$ desaturase.

SEQ ID NOs:20 and 21 correspond to primers YL293 and YL317, respectively, used for amplification of the OCT terminator.

SEQ ID NO:22 is the DNA sequence of a ~2.2 kB fragment comprising the *S. diclina* $\Delta 6$ desaturase gene flanked by the TEF promoter and OCT terminator.

SEQ ID NOs:23-30 correspond to primers YL226, YL227, YL61, YL62, YL230, YL231, YL228 and YL229, respectively, used for site-directed mutagenesis to generate pYZDE1SB-CN.

SEQ ID NOs:31 and 32 correspond to primers YL179 and YL179A, respectively, used for amplification of the *Thraustochytrium aureum* T7091 elongase gene.

SEQ ID NOs:33-36 correspond to primers YL77, YL78, YL322 and YL323, respectively, used for site-directed mutagenesis to generate pGDE2S-PC.

5 SEQ ID NOs:37 and 38 correspond to primers YL325 and YL326, respectively, used for amplification of the ACO terminator.

SEQ ID NO:39 is the DNA sequence of a ~2.6 kB fragment comprising a $\Delta 12$ desaturase flanked by the GPD promoter and an Aco terminator.

10 SEQ ID NO:40 is the DNA sequence of GLA expression vector pDMW208D62.

SEQ ID NO:41 corresponds to the codon-optimized translation initiation site for genes optimally expressed in *Yarrowia sp.*

SEQ ID NO:42 comprises the FBA promoter.

SEQ ID NO:43 is the DNA sequence of the FBAIN promoter.

15 SEQ ID NO:44 is the DNA sequence of the *Fusarium moniliforme* $\Delta 12$ desaturase, while SEQ ID NO:45 is the corresponding amino acid sequence.

SEQ ID NO:46 is the DNA sequence of the synthetic $C_{16/18}$ elongase gene derived from *Rattus norvegicus* and codon-optimized for
20 expression in *Yarrowia lipolytica*. SEQ ID NO:47 is the DNA sequence of the native *Rattus norvegicus* $C_{16/18}$ elongase gene (GenBank Accession No. AB071986), while SEQ ID NO:48 is the corresponding amino acid sequence.

DETAILED DESCRIPTION OF THE INVENTION

25 The following patents, patent applications, and patent publications cited herein are incorporated by reference in their entirety; commonly owned USSN 10/840478, USSN 10/840579 USSN 10/840325, USSN 10/869630, USSN 10/985691, USSN 10/987548, U.S. Provisional Patent Application No. 60/610060 and U.S. Provisional Patent Application No.
30 60/624812..

In accordance with the subject invention, Applicants provide methods for the production of γ -linolenic acid (GLA) in oleaginous yeasts and production hosts useful in said methods. Accumulation of this particular polyunsaturated fatty acid (PUFA) is accomplished by
35 introduction of a functional ω -6 fatty acid biosynthetic pathway comprising proteins that encode $\Delta 12$ desaturase (responsible for conversion of oleic acid to linoleic acid (LA)) and $\Delta 6$ desaturase (responsible for conversion of

LA to GLA) activities into oleaginous yeast hosts for recombinant expression. Thus, this disclosure demonstrates that oleaginous yeast can be engineered to enable commercial production of GLA and derivatives thereof. Methods of production are also claimed.

5 The subject invention finds many applications. PUFAs, or derivatives thereof, made by the methodology disclosed herein can be used as dietary substitutes, or supplements, particularly infant formulas, for patients undergoing intravenous feeding or for preventing or treating malnutrition. Alternatively, the purified PUFAs (or derivatives thereof) may
10 be incorporated into cooking oils, fats or margarines formulated so that in normal use the recipient would receive the desired amount for dietary supplementation. The PUFAs may also be incorporated into infant formulas, nutritional supplements or other food products, and may find use as anti-inflammatory or cholesterol lowering agents. Optionally, the
15 compositions may be used for pharmaceutical use (human or veterinary). In this case, the PUFAs are generally administered orally but can be administered by any route by which they may be successfully absorbed, e.g., parenterally (e.g., subcutaneously, intramuscularly or intravenously), rectally, vaginally or topically (e.g., as a skin ointment or lotion).

20 Supplementation of humans or animals with PUFAs produced by recombinant means can result in increased levels of the added PUFAs, as well as their metabolic progeny. For example, treatment with GLA can result not only in increased levels of GLA, but also downstream products of GLA such as prostaglandins. Complex regulatory mechanisms can
25 make it desirable to combine various PUFAs, or add different conjugates of PUFAs, in order to prevent, control or overcome such mechanisms to achieve the desired levels of specific PUFAs in an individual.

Definitions

 In this disclosure, a number of terms and abbreviations are used.
30 The following definitions are provided.

 "Open reading frame" is abbreviated ORF.

 "Polymerase chain reaction" is abbreviated PCR.

 "American Type Culture Collection" is abbreviated ATCC.

 "Polyunsaturated fatty acid(s)" is abbreviated PUFA(s).

35 The term "fatty acids" refers to long-chain aliphatic acids (alkanoic acids) of varying chain lengths, from about C₁₂ to C₂₂ (although both longer and shorter chain-length acids are known). The predominant chain

lengths are between C₁₆ and C₂₂. The structure of a fatty acid is represented by a simple notation system of "X:Y", where X is the total number of carbon (C) atoms and Y is the number of double bonds. Information relating to the differentiation between "saturated fatty acids" versus "unsaturated fatty acids", "monounsaturated fatty acids" versus "polyunsaturated fatty acids" (or "PUFAs"), and "omega-6 fatty acids" (ω -6 or *n*-6) versus "omega -3 fatty acids" (ω -3 or *n*-3) are provided in USSN 10/840579.

Nomenclature used to describe PUFAs in the present disclosure is shown below in Table 1. In the column titled "Shorthand Notation", the omega-reference system is used to indicate the number of carbons, the number of double bonds and the position of the double bond closest to the omega carbon, counting from the omega carbon (which is numbered 1 for this purpose). The remainder of the Table summarizes the common names of ω -3 and ω -6 fatty acids and their precursors, the abbreviations that will be used throughout the specification and each compounds' chemical name.

Table 1

Nomenclature of Polyunsaturated Fatty Acids And Precursors

Common Name	Abbreviation	Chemical Name	Shorthand Notation
Myristic	--	tetradecanoic	14:0
Palmitate	--	hexadecanoic	16:0
Palmitoleic	--	9-hexadecenoic	16:1
Stearic	--	octadecanoic	18:0
Oleic	--	<i>cis</i> -9-octadecenoic	18:1
Linoleic	LA	<i>cis</i> -9, 12-octadecadienoic; CAS RN #60-33-3	18:2 ω -6
γ -Linolenic	GLA	<i>cis</i> -6, 9, 12-octadecatrienoic; CAS RN #506-26-3	18:3 ω -6
Eicosadienoic	EDA	<i>cis</i> -11, 14-eicosadienoic	20:2 ω -6
Dihomo- γ -Linolenic	DGLA	<i>cis</i> -8, 11, 14-eicosatrienoic	20:3 ω -6
Arachidonic	ARA	<i>cis</i> -5, 8, 11, 14-eicosatetraenoic	20:4 ω -6

α -Linolenic	ALA	<i>cis</i> -9, 12, 15-octadecatrienoic	18:3 ω -3
Stearidonic	STA	<i>cis</i> -6, 9, 12, 15-octadecatetraenoic	18:4 ω -3
Eicosatrienoic	ETrA	<i>cis</i> -11, 14, 17-eicosatrienoic	20:3 ω -3
Eicosa-tetraenoic	ETA	<i>cis</i> -8, 11, 14, 17-eicosatetraenoic	20:4 ω -3
Eicosa-pentaenoic	EPA	<i>cis</i> -5, 8, 11, 14, 17-eicosapentaenoic	20:5 ω -3
Docosa-pentaenoic	DPA	<i>cis</i> -7, 10, 13, 16, 19-docosapentaenoic	22:5 ω -3
Docosa-hexaenoic	DHA	<i>cis</i> -4, 7, 10, 13, 16, 19-docosahexaenoic	22:6 ω -3

The term "high-level GLA production" refers to production of at least about 15% GLA in the total lipids of the microbial host, preferably at least about 20% GLA in the total lipids, more preferably at least about 30% GLA in the total lipids, more preferably at least about 40% GLA in the total lipids and most preferably at least about 50% GLA in the total lipids. The structural form of GLA is not limiting; thus, for example, GLA may exist in the total lipids as free fatty acids or in esterified forms such as acylglycerols, phospholipids, sulfolipids or glycolipids.

"Percent (%) GLA in the total lipids" refers to the percent of GLA relative to the total fatty acids in the total lipid and oil fractions. The term "total lipid fraction" or "lipid fraction" both refer to the sum of all lipids (i.e., neutral and polar) within an oleaginous organism, thus including those lipids that are located in the phosphatidylcholine fraction, phosphatidyletanolamine fraction and triacylglycerol (TAG or oil) fraction. However, the terms "lipid" and "oil" will be used interchangeably throughout the specification herein.

"Microbial oils" or "single cell oils" are those oils naturally produced by microorganisms (e.g., algae, oleaginous yeasts and filamentous fungi) during their lifespan. The term "oil" refers to a lipid substance that is liquid at 25 °C and usually polyunsaturated. In contrast, the term "fat" refers to a lipid substance that is solid at 25 °C and usually saturated.

The term "PUFA biosynthetic pathway enzyme" refers to any of the following enzymes (and genes which encode said enzymes) associated with the biosynthesis of a PUFA, including: a $\Delta 4$ desaturase, a $\Delta 5$ desaturase, a $\Delta 6$ desaturase, a $\Delta 8$ desaturase, a $\Delta 9$ desaturase, a $\Delta 12$ desaturase, a $\Delta 15$ desaturase, a $\Delta 17$ desaturase, a $\Delta 9$ elongase, a $C_{14/16}$ elongase, a $C_{16/18}$ elongase, a $C_{18/20}$ elongase and/or a $C_{20/22}$ elongase. Similarly, the term " ω -3/ ω -6 fatty acid biosynthetic pathway" refers to a set of genes which, when expressed under the appropriate conditions encode enzymes that catalyze the production of either or both ω -3 and ω -6 fatty acids. Typically the genes involved in the ω -3/ ω -6 fatty acid biosynthetic pathway encode some or all of the following enzymes: $\Delta 12$ desaturase, $\Delta 6$ desaturase, $\Delta 5$ desaturase, $\Delta 17$ desaturase, $\Delta 15$ desaturase, $\Delta 8$ desaturase, $\Delta 4$ desaturase, a $\Delta 9$ elongase, a $C_{18/20}$ elongase and/or a $C_{20/22}$ elongase. A representative pathway is illustrated in Figure 1, providing for the conversion of myristic acid through various intermediates to DHA, which demonstrates how both ω -3 and ω -6 fatty acids may be produced from a common source. The pathway is naturally divided into two portions where one portion will generate ω -3 fatty acids and the other portion, only ω -6 fatty acids. That portion that only generates ω -3 fatty acids will be referred to herein as the ω -3 fatty acid biosynthetic pathway, whereas that portion that generates only ω -6 fatty acids will be referred to herein as the ω -6 fatty acid biosynthetic pathway.

The term "functional" as used herein in context with the ω -3/ ω -6 fatty acid biosynthetic pathway (or a portion thereof) means that some (or all of) the genes in the pathway express active enzymes, resulting in *in vivo* catalysis or substrate conversion. It should be understood that " ω -3/ ω -6 fatty acid biosynthetic pathway" or "functional ω -3/ ω -6 fatty acid biosynthetic pathway" does not imply that all the genes listed in the above paragraph are required, as a number of fatty acid products will only require the expression of a subset of the genes of this pathway.

The term "GLA biosynthetic pathway" refers to genes encoding enzymes providing for the conversion of oleic acid through GLA (i.e., $\Delta 12$ desaturase and $\Delta 6$ desaturase).

The term "desaturase" refers to a polypeptide that can desaturate one or more fatty acids to produce a mono- or polyunsaturated fatty acid or precursor of interest. Despite use of the omega-reference system throughout the specification to refer to specific fatty acids, it is more

convenient to indicate the activity of a desaturase by counting from the carboxyl end of the substrate using the delta-system. Of particular interest herein are: $\Delta 12$ desaturases that desaturate a fatty acid between the 12th and 13th carbon atom numbered from the carboxyl-terminal end of the molecule and which, for example, catalyze the conversion of oleic acid to LA; and, $\Delta 6$ desaturases that catalyze the conversion of LA to GLA and/or ALA to STA. Other PUFA desaturases include: 1.) $\Delta 17$ desaturases that catalyze the conversion of ARA to EPA and DGLA to ETA; 2.) $\Delta 5$ desaturases that catalyze the conversion of DGLA to ARA and/or ETA to EPA; 3.) $\Delta 4$ desaturases that catalyze the conversion of DPA to DHA; 4.) $\Delta 15$ desaturases that catalyze the conversion of LA to ALA; 5.) $\Delta 9$ desaturases that catalyze the conversion of palmitate to palmitoleic acid (16:1) and/or stearate to oleic acid (18:1); and 6.) $\Delta 8$ desaturases that catalyze the conversion of EDA to DGLA and ETrA to ETA.

The term "elongase system" refers to a suite of four enzymes that are responsible for elongation of a fatty acid carbon chain to produce a fatty acid that is 2 carbons longer than the fatty acid substrate that the elongase system acts upon. More specifically, the process of elongation occurs in association with fatty acid synthase, whereby CoA is the acyl carrier (Lassner et al., *The Plant Cell* 8:281-292 (1996)). In the first step, which has been found to be both substrate-specific and also rate-limiting, malonyl-CoA is condensed with a long-chain acyl-CoA to yield CO₂ and a β -ketoacyl-CoA (where the acyl moiety has been elongated by two carbon atoms). Examples of reactions catalyzed by elongase systems are the conversion of GLA to DGLA, LA to EDA, ALA to ETrA, STA to ETA and EPA to DPA.

For the purposes herein, an enzyme catalyzing the first condensation reaction (i.e., conversion of malonyl-CoA to β -ketoacyl-CoA) will be referred to generically as an "elongase". In general, the substrate selectivity of elongases is somewhat broad but segregated by both chain length and the degree of unsaturation. Accordingly, elongases can have different specificities. For example, a C_{14/16} elongase will utilize a C₁₄ substrate (e.g., myristic acid), a C_{16/18} elongase will utilize a C₁₆ substrate (e.g., palmitate), a C_{18/20} elongase will utilize a C₁₈ substrate (e.g., GLA, STA) and a C_{20/22} elongase will utilize a C₂₀ substrate (e.g., EPA). In like manner, a $\Delta 9$ elongase is able to catalyze the conversion of LA and ALA to EDA and ETrA, respectively. It is important to note that some

elongases have broad specificity and thus a single enzyme may be capable of catalyzing several elongase reactions (e.g., thereby acting as both a C_{16/18} elongase and a C_{18/20} elongase).

5 The terms "conversion efficiency" and "percent substrate conversion" refer to the efficiency by which a particular enzyme (e.g., a desaturase or elongase) can convert substrate to product. The conversion efficiency is measured according to the following formula:

([product]/[substrate+product])*100, where 'product' includes the immediate product and all products in the pathway derived from it.

10 The term "oleaginous" refers to those organisms that tend to store their energy source in the form of lipid (Weete, In: Fungal Lipid Biochemistry, 2nd Ed., Plenum, 1980). Generally, the cellular PUFA content of these microorganisms follows a sigmoid curve, wherein the concentration of lipid increases until it reaches a maximum at the late logarithmic or early stationary growth phase and then gradually decreases during the late stationary and death phases (Yongmanitchai and Ward, *Appl. Environ. Microbiol.* 57:419-25 (1991)).

15 The term "oleaginous yeast" refers to those microorganisms classified as yeast that can accumulate at least 25% of their dry cell weight as oil. Examples of oleaginous yeast include, but are no means limited to, the following genera: *Yarrowia*, *Candida*, *Rhodotorula*, *Rhodospiridium*, *Cryptococcus*, *Trichosporon* and *Lipomyces*.

25 The term "fermentable carbon source" means a carbon source that a microorganism will metabolize to derive energy. Typical carbon sources of the invention include, but are not limited to: monosaccharides, oligosaccharides, polysaccharides, alkanes, fatty acids, esters of fatty acids, monoglycerides, diglycerides, triglycerides, carbon dioxide, methanol, formaldehyde, formate and carbon-containing amines.

30 As used herein, an "isolated nucleic acid fragment" is a polymer of RNA or DNA that is single- or double-stranded, optionally containing synthetic, non-natural or altered nucleotide bases. An isolated nucleic acid fragment in the form of a polymer of DNA may be comprised of one or more segments of cDNA, genomic DNA or synthetic DNA.

35 A "substantial portion" of an amino acid or nucleotide sequence is that portion comprising enough of the amino acid sequence of a polypeptide or the nucleotide sequence of a gene to putatively identify that polypeptide or gene, either by manual evaluation of the sequence by one

skilled in the art, or by computer-automated sequence comparison and identification using algorithms such as BLAST (Basic Local Alignment Search Tool; Altschul, S. F., et al., *J. Mol. Biol.* 215:403-410 (1993)). In general, a sequence of ten or more contiguous amino acids or thirty or
5 more nucleotides is necessary in order to identify putatively a polypeptide or nucleic acid sequence as homologous to a known protein or gene. Moreover, with respect to nucleotide sequences, gene-specific oligonucleotide probes comprising 20-30 contiguous nucleotides may be used in sequence-dependent methods of gene identification (e.g.,
10 Southern hybridization) and isolation (e.g., *in situ* hybridization of microbial colonies or bacteriophage plaques). In addition, short oligonucleotides of 12-15 bases may be used as amplification primers in PCR in order to obtain a particular nucleic acid fragment comprising the primers. Accordingly, a "substantial portion" of a nucleotide sequence comprises
15 enough of the sequence to specifically identify and/or isolate a nucleic acid fragment comprising the sequence.

The term "complementary" is used to describe the relationship between nucleotide bases that are capable of hybridizing to one another. For example, with respect to DNA, adenosine is complementary to
20 thymine and cytosine is complementary to guanine.

"Codon degeneracy" refers to the nature in the genetic code permitting variation of the nucleotide sequence without affecting the amino acid sequence of an encoded polypeptide. The skilled artisan is well aware of the "codon-bias" exhibited by a specific host cell in usage of
25 nucleotide codons to specify a given amino acid. Therefore, when synthesizing a gene for improved expression in a host cell, it is desirable to design the gene such that its frequency of codon usage approaches the frequency of preferred codon usage of the host cell.

"Gene" refers to a nucleic acid fragment that expresses a specific
30 protein, including regulatory sequences preceding (5' non-coding sequences) and following (3' non-coding sequences) the coding sequence. "Native gene" refers to a gene as found in nature with its own regulatory sequences. "Chimeric gene" refers to any gene that is not a native gene, comprising regulatory and coding sequences that are not
35 found together in nature. Accordingly, a chimeric gene may comprise regulatory sequences and coding sequences that are derived from different sources, or regulatory sequences and coding sequences derived

from the same source, but arranged in a manner different than that found in nature. "Endogenous gene" refers to a native gene in its natural location in the genome of an organism. A "foreign" gene refers to a gene not normally found in the host organism, but that is introduced into the host organism by gene transfer. Foreign genes can comprise native genes inserted into a non-native organism, or chimeric genes. A "transgene" is a gene that has been introduced into the genome by a transformation procedure. A "codon-optimized gene" is a gene having its frequency of codon usage designed to mimic the frequency of preferred codon usage of the host cell.

"Coding sequence" refers to a DNA sequence that codes for a specific amino acid sequence. "Suitable regulatory sequences" refer to nucleotide sequences located upstream (5' non-coding sequences), within or downstream (3' non-coding sequences) of a coding sequence and which influence the transcription, RNA processing or stability, or translation of the associated coding sequence. Regulatory sequences may include promoters, translation leader sequences, introns, polyadenylation recognition sequences, RNA processing sites, effector binding sites and stem-loop structures.

"Promoter" refers to a DNA sequence capable of controlling the expression of a coding sequence or functional RNA. In general, a coding sequence is located 3' to a promoter sequence. Promoters may be derived in their entirety from a native gene, or be composed of different elements derived from different promoters found in nature, or even comprise synthetic DNA segments. It is understood by those skilled in the art that different promoters may direct the expression of a gene in different tissues or cell types, or at different stages of development, or in response to different environmental or physiological conditions. Promoters that cause a gene to be expressed in most cell types at most times are commonly referred to as "constitutive promoters". It is further recognized that since in most cases the exact boundaries of regulatory sequences have not been completely defined, DNA fragments of different lengths may have identical promoter activity.

The terms "3' non-coding sequences" or "transcription terminator" refer to DNA sequences located downstream of a coding sequence. This includes polyadenylation recognition sequences and other sequences encoding regulatory signals capable of affecting mRNA processing or

gene expression. The polyadenylation signal is usually characterized by affecting the addition of polyadenylic acid tracts to the 3' end of the mRNA precursor. The 3' region can influence the transcription, RNA processing or stability, or translation of the associated coding sequence.

5 “RNA transcript” refers to the product resulting from RNA polymerase-catalyzed transcription of a DNA sequence. When the RNA transcript is a perfect complementary copy of the DNA sequence, it is referred to as the primary transcript or it may be a RNA sequence derived from post-transcriptional processing of the primary transcript and is
10 referred to as the mature RNA. “Messenger RNA” or “mRNA” refers to the RNA that is without introns and that can be translated into protein by the cell. “cDNA” refers to a double-stranded DNA that is complementary to, and derived from, mRNA. “Sense” RNA refers to RNA transcript that includes the mRNA and so can be translated into protein by the cell.
15 “Antisense RNA” refers to a RNA transcript that is complementary to all or part of a target primary transcript or mRNA and that blocks the expression of a target gene (U.S. 5,107,065; WO 99/28508). The complementarity of an antisense RNA may be with any part of the specific gene transcript, i.e., at the 5' non-coding sequence, 3' non-coding sequence or the coding
20 sequence. “Functional RNA” refers to antisense RNA, ribozyme RNA or other RNA that is not translated and yet has an effect on cellular processes.

 The term “operably linked” refers to the association of nucleic acid sequences on a single nucleic acid fragment so that the function of one is
25 affected by the other. For example, a promoter is operably linked with a coding sequence when it is capable of affecting the expression of that coding sequence (i.e., the coding sequence is under the transcriptional control of the promoter). Coding sequences can be operably linked to regulatory sequences in sense or antisense orientation.

30 The term “expression”, as used herein, refers to the transcription and stable accumulation of sense (mRNA) or antisense RNA derived from the nucleic acid fragments of the invention. Expression may also refer to translation of mRNA into a polypeptide.

 “Transformation” refers to the transfer of a nucleic acid molecule
35 into a host organism, resulting in genetically stable inheritance. The nucleic acid molecule may be a plasmid that replicates autonomously, for example; or, it may integrate into the genome of the host organism. Host

organisms containing the transformed nucleic acid fragments are referred to as "transgenic" or "recombinant" or "transformed" organisms.

The terms "plasmid", "vector" and "cassette" refer to an extra chromosomal element often carrying genes that are not part of the central metabolism of the cell, and usually in the form of circular double-stranded DNA fragments. Such elements may be autonomously replicating sequences, genome integrating sequences, phage or nucleotide sequences, linear or circular, of a single- or double-stranded DNA or RNA, derived from any source, in which a number of nucleotide sequences have been joined or recombined into a unique construction that is capable of introducing a promoter fragment and DNA sequence for a selected gene product along with appropriate 3' untranslated sequence into a cell. "Expression cassette" refers to a specific vector containing a foreign gene and having elements in addition to the foreign gene that allow for enhanced expression of that gene in a foreign host.

The term "homologous recombination" refers to the exchange of DNA fragments between two DNA molecules (during cross over). The fragments that are exchanged are flanked by sites of identical nucleotide sequences between the two DNA molecules (i.e., "regions of homology"). The term "regions of homology" refer to stretches of nucleotide sequence on nucleic acid fragments that participate in homologous recombination that have homology to each other. Effective homologous recombination will generally take place where these regions of homology are at least about 10 bp in length where at least about 50 bp in length is preferred. Typically fragments that are intended for recombination contain at least two regions of homology where targeted gene disruption or replacement is desired.

Standard recombinant DNA and molecular cloning techniques used herein are well known in the art and are described by Sambrook, J., Fritsch, E. F. and Maniatis, T., Molecular Cloning: A Laboratory Manual, 2nd ed., Cold Spring Harbor Laboratory: Cold Spring Harbor, NY (1989) (hereinafter "Maniatis"); by Silhavy, T. J., Bannan, M. L. and Enquist, L. W., Experiments with Gene Fusions, Cold Spring Harbor Laboratory: Cold Spring Harbor, NY (1984); and by Ausubel, F. M. et al., Current Protocols in Molecular Biology, published by Greene Publishing Assoc. and Wiley-Interscience (1987).

Microbial Biosynthesis of Fatty Acids

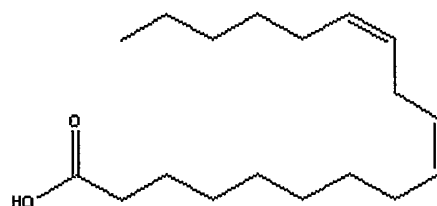
The process of *de novo* synthesis of palmitate (16:0) in oleaginous microorganisms is described in 10/840579. This fatty acid is the precursor of longer-chain saturated and unsaturated fatty acid derivatives, which are formed through the action of elongases and desaturases. For example, palmitate is converted to its unsaturated derivative [palmitoleic acid (16:1)] by the action of a Δ^9 desaturase; alternatively, palmitate is elongated by a $C_{16/18}$ elongase to form stearic acid (18:0), which can be converted to its unsaturated derivative by a Δ^9 desaturase to thereby yield oleic acid (18:1). Since the primary fate of palmitate is elongation, however, it is concluded that $C_{16/18}$ elongases play an important role in determining overall carbon flux into the fatty acid biosynthetic pathway since they control the amount of stearic acid produced in the microbial cell.

Triacylglycerols or "TAGs" (the primary storage unit for fatty acids) are formed by the esterification of two molecules of acyl-CoA to glycerol-3-phosphate to yield 1,2-diacylglycerol phosphate (commonly identified as phosphatidic acid (PA)) (and CoA as a by-product)). The phosphate is then removed, by phosphatidic acid phosphatase, to yield 1,2-diacylglycerol (DAG). Triacylglycerol is formed upon the addition of a third fatty acid to the *sn*-3 position of DAG by the action of a DAG transferase (e.g., phospholipid:diacylglycerol acyltransferase [PDAT] and diacylglycerol acyltransferase [DGAT1 or DGAT2]).

Biosynthesis Of GLA, An ω -6 Fatty Acid

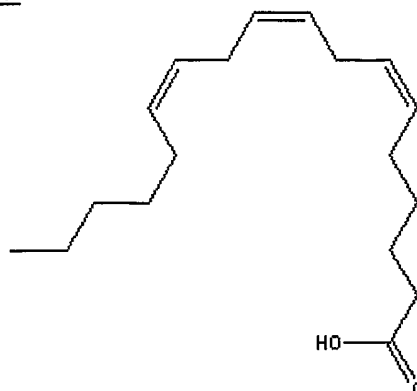
The metabolic process wherein oleic acid is converted to LA and then GLA requires desaturation of the molecule through the addition of double bonds. Specifically, oleic acid (18:1) is converted to LA (18:2; Formula I below), the first of the ω -6 fatty acids, by the activity of a Δ^{12} desaturase.

Formula I:



Then, LA is converted to GLA (Formula II below) by the activity of a $\Delta 6$ desaturase.

Formula II:



5 Thus, the construction of a *Yarrowia lipolytica* strain producing greater than 15% GLA in the total oil fraction, or more preferably greater than 20% GLA in the total oil fraction, or even more preferably greater than 30% GLA in the total oil fraction, or most preferably greater than 40-50% GLA in the total oil fraction requires at least the following genes: a
 10 $\Delta 12$ desaturase and a $\Delta 6$ desaturase. However, it may be desirable to additionally include a $\Delta 9$ desaturase, a $C_{14/16}$ elongase and/or a $C_{16/18}$ elongase in the host strain (*infra*).

A variety of other PUFAs can be produced using GLA as substrate. The complete ω -6 and ω -3 pathways for fatty acid biosynthesis are shown
 15 in Figure 1. Specifically, GLA can be converted to DGLA by the activity of a $C_{18/20}$ elongase; and DGLA is subsequently converted to ARA by the activity of a $\Delta 5$ desaturase or ETA by the activity of a $\Delta 17$ desaturase. Downstream products may also include e.g., EPA, DPA and DHA, depending on the presence of additional desaturases and elongases with
 20 appropriate specificity.

Microbial Production of GLA

Microbial production of GLA and other PUFAs has several advantages over purification from natural plant sources. For example: (1) many microbes are known with greatly simplified oil compositions
 25 compared with those of higher organisms, making purification of desired components easier; (2) microbial production is not subject to fluctuations caused by external variables, such as weather and disease; (3) microbially produced oil is substantially free of contamination by environmental pollutants; and, (4) microbial oil production can be manipulated by
 30 controlling culture conditions, notably by providing particular substrates for

microbially expressed enzymes, or by addition of compounds or genetic engineering approaches to suppress undesired biochemical pathways. With respect to the production of ω -3 and/or ω -6 fatty acids in particular (e.g., GLA), and TAGs containing those PUFAs, additional advantages are incurred since microbes can provide fatty acids in particular forms that may have specific uses; and, recombinant microbes provide the ability to alter the naturally occurring microbial fatty acid profile by providing new biosynthetic pathways in the host or by suppressing undesired pathways, thereby increasing levels of desired PUFAs or conjugated forms thereof and decreasing levels of undesired PUFAs.

Microbial Genes for GLA Production

Many microorganisms, including algae, bacteria, molds and yeast, can synthesize PUFAs and omega fatty acids in the ordinary course of cellular metabolism. Particularly well-studied are fungi including *Schizochytrium aggregatum*, species of the genus *Thraustochytrium* and *Mortierella alpina*. Additionally, many dinoflagellates (Dinophyceae) naturally produce high concentrations of PUFAs. As such, a variety of genes involved in oil production have been identified through genetic means; and, the DNA sequences of some of the genes required for GLA biosynthesis (i.e., Δ 12 and Δ 6 desaturases) are publicly available (non-limiting examples are shown below in Table 2):

Table 2
Some Publicly Available Genes Involved In GLA Production

Genbank Accession No.	Description
AY131238	<i>Argania spinosa</i> Δ 6 desaturase
Y055118	<i>Echium pitardii</i> var. <i>pitardii</i> Δ 6 desaturase
AY055117	<i>Echium gentianoides</i> Δ 6 desaturase
AF296076	<i>Mucor rouxii</i> Δ 6 desaturase
AF007561	<i>Borago officinalis</i> Δ 6 desaturase
L11421	<i>Synechocystis</i> sp. Δ 6 desaturase
NM_031344	<i>Rattus norvegicus</i> Δ 6 fatty acid desaturase
AF465283, AF465281, AF110510	<i>Mortierella alpina</i> Δ 6 fatty acid desaturase

Genbank Accession No.	Description
AF465282	<i>Mortierella isabellina</i> $\Delta 6$ fatty acid desaturase
AF419296	<i>Pythium irregulare</i> $\Delta 6$ fatty acid desaturase
AB052086	<i>Mucor circinelloides</i> D6d mRNA for $\Delta 6$ fatty acid desaturase
AJ250735	<i>Ceratodon purpureus</i> mRNA for $\Delta 6$ fatty acid desaturase
AF126799	<i>Homo sapiens</i> $\Delta 6$ fatty acid desaturase
AF126798	<i>Mus musculus</i> $\Delta 6$ fatty acid desaturase
AF251842	<i>Helianthus annuus</i> $\Delta 12$ fatty acid desaturase
AY165023	<i>Phaeodactylum tricornutum</i> $\Delta 12$ fatty acid desaturase
AF343065	<i>Calendula officinalis</i> $\Delta 12$ fatty acid desaturase FAD2
AF417245	<i>Mortierella isabellina</i> M6-22 $\Delta 12$ fatty acid desaturase
AF417244	<i>Mortierella alpina</i> ATCC #16266 $\Delta 12$ fatty acid desaturase gene
AB052087	<i>Mucor circinelloides</i> D12d mRNA for $\Delta 12$ fatty acid desaturase
AAG36933	<i>Emericella nidulans</i> oleate $\Delta 12$ desaturase
AF110509, AB020033	<i>Mortierella alpina</i> $\Delta 12$ fatty acid desaturase mRNA
AAL13300	<i>Mortierella alpina</i> $\Delta 12$ fatty acid desaturase
AF161219	<i>Mucor rouxii</i> $\Delta 12$ desaturase mRNA

Additionally, the patent literature provides many additional DNA sequences of genes (and/or details concerning several of the genes above and their methods of isolation) involved in PUFA production. See, for example: U.S. 5,968,809 and U.S. Patent Application Number 60/468718
5 (Δ6 desaturases); WO 94/11516, U.S. 5,443,974, WO 03/099216, U.S. Patent Application Number 60/484209 and U.S. Patent Application Number - (Δ12 desaturases).

Microbial Gene Selection

As is well known in the art, the particular functionalities required to be introduced into a host organism for production of a particular PUFA final product will depend on the host cell (and its native PUFA profile and/or desaturase/elongase profile), the availability of substrate and the desired end product(s). For GLA production from oleic acid, LA and GLA may both be produced in oleaginous yeast by introducing a $\Delta 12$ desaturase and a $\Delta 6$ desaturase and subsequent PUFA derivatives, including DGLA, ARA, STA, ETA, EPA, DPA and DHA, may also be produced if the host possesses endogenous or exogenous $\Delta 4$, $\Delta 5$, $\Delta 15$ and/or $\Delta 17$ desaturase activities and/or $C_{18/20}$ and/or $C_{20/22}$ activities). One skilled in the art will be able to identify various candidate genes encoding $\Delta 12$ desaturase and/or $\Delta 6$ desaturase activities, according to publicly available literature (e.g., GenBank), the patent literature and experimental analysis of microorganisms having the ability to produce PUFAs. The sequences may be derived from any source, e.g., isolated from a natural source (from bacteria, algae, fungi, plants, animals, etc.), produced via a semi-synthetic route or synthesized *de novo*. In some embodiments, manipulation of genes endogenous to the host is preferred; for other purposes, it is necessary to introduce heterologous genes.

Although the particular source of the desaturase gene introduced into the host is not critical to the present invention, considerations for choosing a specific polypeptide having desaturase activity include: 1.) the substrate specificity of the polypeptide; 2.) whether the polypeptide or a component thereof is a rate-limiting enzyme; 3.) whether the desaturase is essential for synthesis of a desired PUFA; and/or 4.) co-factors required by the polypeptide. The expressed polypeptide preferably has parameters compatible with the biochemical environment of its location in the host cell. For example, the polypeptide may have to compete for substrate with other enzymes in the host cell. Analyses of the K_M and specific activity of the polypeptide are therefore considered in determining the suitability of a given polypeptide for modifying PUFA production, and specifically GLA production, in a given host cell. The polypeptide used in a particular host cell is one that can function under the biochemical conditions present in the intended host cell but otherwise can be any polypeptide having desaturase activity capable of converting a particular substrate into the desired PUFA.

Endogenous PUFA Genes

In some cases, the host organism in which it is desirable to produce GLA will possess endogenous genes encoding some PUFA biosynthetic pathway enzymes. For example, oleaginous yeast can typically produce 18:2 acids (and some have the additional capability of synthesizing 18:3 acids); thus, oleaginous yeast typically possess native $\Delta 12$ desaturase activity and may also have $\Delta 15$ desaturases. In some embodiments, therefore, expression of the native desaturase enzyme is preferred over a heterologous (or "foreign") enzyme since: 1.) the native enzyme is optimized for interaction with other enzymes and proteins within the cell; and 2.) heterologous genes are unlikely to share the same codon preference in the host organism.

In one embodiment of the present invention, *Yarrowia lipolytica* was selected as a host organism for production of GLA. This oleaginous yeast is able to synthesize 18:2 fatty acids (but not 18:3), as a result of an endogenous $\Delta 12$ fatty acid desaturase (contained within SEQ ID NO:16; see commonly owned USSN 10/840325).

Heterologous PUFA Genes

In many instances, the appropriate desaturases (and optionally, elongases) are not present in the host organism of choice to enable production of the desired PUFA products. Thus, it is necessary to introduce heterologous genes.

For the purposes of the present invention herein, it was desirable to demonstrate high-level production of GLA in an oleaginous host organism; and thus, a native $\Delta 12$ desaturase, a synthetic *Mortierella alpina* $\Delta 6$ desaturase codon-optimized for expression in *Yarrowia lipolytica* and a *Saprolegnia diclina* $\Delta 6$ desaturase were introduced into the host, *Y. lipolytica*. However, the specific enzymes (and genes encoding those enzymes) introduced into the host organism are by no means limiting to the invention herein.

If one desired to produce GLA, as demonstrated herein, it is clear that numerous other genes derived from different sources would be suitable to introduce $\Delta 12$ desaturase and $\Delta 6$ desaturase activity into the preferred microbial host. Thus, in one embodiment of the present invention, other DNAs that are substantially identical to the *Y. lipolytica* $\Delta 12$ desaturase, synthetically produced $\Delta 6$ desaturase (codon-optimized for expression in *Y. lipolytica*) and the *S. diclina* $\Delta 6$ desaturase also could

be used for production of GLA in oleaginous yeast. By "substantially identical" is intended an amino acid sequence or nucleic acid sequence exhibiting in order of increasing preference at least 80%, 90% or 95% homology to the selected polypeptides, or nucleic acid sequences encoding the amino acid sequence. For polypeptides, the length of comparison sequences generally is at least 16 amino acids, preferably at least 20 amino acids, or most preferably 35 amino acids. For nucleic acids, the length of comparison sequences generally is at least 50 nucleotides, preferably at least 60 nucleotides, more preferably at least 75 nucleotides, and most preferably 110 nucleotides.

Homology typically is measured using sequence analysis software, wherein the term "sequence analysis software" refers to any computer algorithm or software program that is useful for the analysis of nucleotide or amino acid sequences. "Sequence analysis software" may be commercially available or independently developed. Typical sequence analysis software will include, but is not limited to: 1.) the GCG suite of programs (Wisconsin Package Version 9.0, Genetics Computer Group (GCG), Madison, WI); 2.) BLASTP, BLASTN, BLASTX (Altschul et al., *J. Mol. Biol.* 215:403-410 (1990)); 3.) DNASTAR (DNASTAR, Inc., Madison, WI); and 4.) the FASTA program incorporating the Smith-Waterman algorithm (W. R. Pearson, *Comput. Methods Genome Res.*, [Proc. Int. Symp.] (1994), Meeting Date 1992, 111-20. Suhai, Sandor, Ed. Plenum: New York, NY). Within the context of this application it will be understood that where sequence analysis software is used for analysis, the results of the analysis will be based on the "default values" of the program referenced, unless otherwise specified. As used herein "default values" will mean any set of values or parameters that originally load with the software when first initialized. In general, such computer software matches similar sequences by assigning degrees of homology to various substitutions, deletions and other modifications.

Additionally, it will be appreciated by one of skill in the art that polypeptides may have amino acids conservatively substituted in a manner such that the function of the polypeptide is not altered or compromised. Polypeptides having the desaturase activities as described herein and possessing such conservative substitutions are considered within the scope of the present invention. Conservative substitutions typically include substitutions within the following groups: 1.) glycine and

alanine; 2.) valine, isoleucine and leucine; 3.) aspartic acid, glutamic acid, asparagine and glutamine; 4.) serine and threonine; 5.) lysine and arginine; and 6.) phenylalanine and tyrosine. Substitutions may also be made on the basis of conserved hydrophobicity or hydrophilicity (Kyte and Doolittle, *J. Mol. Biol.* 157:105-132 (1982)), or on the basis of the ability to assume similar polypeptide secondary structure (Chou and Fasman, *Adv. Enzymol.* 47:45-148 (1978)).

One alternative $\Delta 12$ desaturase that is specifically contemplated for overexpression in *Yarrowia lipolytica* is that from *Fusarium moniliforme* (WO2005/047485; SEQ ID NOs:44 and 45 herein). And, in some cases, it may prove advantageous to replace the native *Y. lipolytica* $\Delta 12$ desaturase with the *F. moniliforme* $\Delta 12$ desaturase, since the latter shows increased percent substrate conversion. More specifically, although both $\Delta 12$ desaturases catalyze the conversion of oleic acid to LA, the two enzymes differ in their overall specificity (which thereby affects each enzyme's percent substrate conversion). The Applicants have determined that the *F. moniliforme* $\Delta 12$ desaturase has a higher loading capacity of LA onto the *sn*-2 position of a phosphatidylcholine substrate (thereby facilitating the subsequent reaction by $\Delta 6$ desaturase) than the *Y. lipolytica* $\Delta 12$ desaturase. On this basis, overexpression of the *F. moniliforme* $\Delta 12$ desaturase in conjunction with a knockout of the *Y. lipolytica* $\Delta 12$ desaturase may result in increased product for subsequent conversion to EPA.

In alternate embodiments of the invention, other DNAs that, although not substantially identical to the *Yarrowia lipolytica* $\Delta 12$ desaturase, synthetically produced $\Delta 6$ desaturase (codon-optimized for expression in *Y. lipolytica*) and the *S. diclina* $\Delta 6$ desaturase, also can be used for the purposes herein (e.g., for production of GLA). For example, DNA sequences encoding $\Delta 6$ desaturase polypeptides that would be useful for introduction into an oleaginous yeast according to the teachings of the present invention may be obtained from microorganisms having an ability to produce GLA or STA. Such microorganisms include, for example, those belonging to the genera *Mortierella*, *Conidiobolus*, *Pythium*, *Phytophthora*, *Penicillium*, *Porphyridium*, *Coidosporium*, *Mucor*, *Fusarium*, *Aspergillus*, *Rhodotorula* and *Entomophthora*. Within the genus *Porphyridium*, of particular interest is *P. cruentum*. Within the genus *Mortierella*, of particular interest are *M. elongata*, *M. exigua*, *M. hygrophila*,

M. ramanniana var. *angulispora* and *M. alpina*. Within the genus *Mucor*, of particular interest are *M. circinelloides* and *M. javanicus*.

Alternatively, a related desaturase that is not substantially identical to e.g., the *S. diclina* $\Delta 6$ desaturase, but which can desaturate a fatty acid molecule at carbon 6 from the carboxyl end of the molecule would also be
5 useful in the present invention as a $\Delta 6$ desaturase, assuming the desaturase can still effectively convert LA to GLA. As such, related desaturases can be identified (or created) by their ability to function substantially the same as the desaturases disclosed herein.

10 As suggested above, in another embodiment one skilled in the art could create a fusion protein having both $\Delta 12$ desaturase and $\Delta 6$ desaturase activities suitable for the purposes herein. This would be possible, for example, by fusing together a $\Delta 12$ desaturase and $\Delta 6$ desaturase with an adjoining linker. Either the $\Delta 12$ desaturase or the $\Delta 6$
15 desaturase could be at the N-terminal portion of the fusion protein. Means to design and synthesize an appropriate linker molecule are readily known by one of skill in the art; for example, the linker can be a stretch of alanine or lysine amino acids and will not affect the fusion enzyme's activity.

In alternate embodiments, one skilled in the art could isolate a gene
20 encoding a protein having both $\Delta 12$ desaturase and $\Delta 6$ desaturase activities. Such a gene could also be used to produce GLA.

Finally, it is well known in the art that methods for synthesizing sequences and bringing sequences together are well established in the literature. Thus, *in vitro* mutagenesis and selection, site-directed
25 mutagenesis or other means can be employed to obtain mutations of naturally occurring desaturase genes. This would permit production of a polypeptide having desaturase activity, respectively, *in vivo* with more desirable physical and kinetic parameters for functioning in the host cell (e.g., a longer half-life or a higher rate of production of a desired PUFA).

30 If desired, a $\Delta 6$ desaturase could be mutated to possess both $\Delta 6$ and $\Delta 12$ desaturase activities, or be completely changed into a $\Delta 12$ desaturase. Likewise, a $\Delta 12$ desaturase could be mutated to possess both $\Delta 12$ and $\Delta 6$ desaturase activities, or be completely changed into a $\Delta 6$ desaturase. Such mutated genes could also be used to produce GLA.

35 If desired, the regions of a polypeptide of interest (i.e., a $\Delta 6$ and/ or $\Delta 12$ desaturase) important for enzymatic activity can be determined through routine mutagenesis, expression of the resulting mutant

polypeptides and determination of their activities. Mutants may include deletions, insertions and point mutations, or combinations thereof. A typical functional analysis begins with deletion mutagenesis to determine the N- and C-terminal limits of the protein necessary for function, and then
5 internal deletions, insertions or point mutants are made to further determine regions necessary for function. Other techniques such as cassette mutagenesis or total synthesis also can be used. Deletion mutagenesis is accomplished, for example, by using exonucleases to sequentially remove the 5' or 3' coding regions. Kits are available for such
10 techniques. After deletion, the coding region is completed by ligating oligonucleotides containing start or stop codons to the deleted coding region after the 5' or 3' deletion, respectively. Alternatively, oligonucleotides encoding start or stop codons are inserted into the coding region by a variety of methods including site-directed mutagenesis,
15 mutagenic PCR or by ligation onto DNA digested at existing restriction sites. Internal deletions can similarly be made through a variety of methods including the use of existing restriction sites in the DNA, by use of mutagenic primers via site-directed mutagenesis or mutagenic PCR. Insertions are made through methods such as linker-scanning
20 mutagenesis, site-directed mutagenesis or mutagenic PCR, while point mutations are made through techniques such as site-directed mutagenesis or mutagenic PCR.

Chemical mutagenesis also can be used for identifying regions of a desaturase polypeptide important for activity. A mutated construct is
25 expressed, and the ability of the resulting altered protein to function as a desaturase is assayed. Such structure-function analysis can determine which regions may be deleted, which regions tolerate insertions and which point mutations allow the mutant protein to function in substantially the same way as the native desaturase.

30 Preferred Microbial Hosts for GLA Production

Host cells for production of GLA may include microbial hosts that grow on a variety of feedstocks, including simple or complex carbohydrates, organic acids and alcohols, and/or hydrocarbons over a wide range of temperature and pH values.

35 Preferred microbial hosts, however, are oleaginous yeast. These organisms are naturally capable of oil synthesis and accumulation, wherein the oil can comprise greater than about 25% of the cellular dry

weight, more preferably greater than about 30% of the cellular dry weight and most preferably greater than about 40% of the cellular dry weight. Genera typically identified as oleaginous yeast include, but are not limited to: *Yarrowia*, *Candida*, *Rhodotorula*, *Rhodospiridium*, *Cryptococcus*,
5 *Trichosporon* and *Lipomyces*. More specifically, illustrative oil-synthesizing yeast include: *Rhodospiridium toruloides*, *Lipomyces starkeyii*, *L. lipoferus*, *Candida revkaufi*, *C. pulcherrima*, *C. tropicalis*, *C. utilis*, *Trichosporon pullans*, *T. cutaneum*, *Rhodotorula glutinus*, *R. graminis* and *Yarrowia lipolytica* (formerly classified as *Candida*
10 *lipolytica*).

Most preferred is the oleaginous yeast *Yarrowia lipolytica*; and, in a further embodiment, most preferred are the *Y. lipolytica* strains designated as ATCC #20362, ATCC #8862, ATCC #18944, ATCC #76982 and/or LGAM S(7)1 (Papanikolaou S., and Aggelis G., *Bioresour. Technol.*
15 82(1):43-9 (2002)).

Metabolic Engineering For High-Level GLA Biosynthesis In Oleaginous Yeast

Methods for manipulating biochemical pathways are well known to those skilled in the art; and, it is expected that numerous manipulations
20 will be possible to maximize GLA biosynthesis in oleaginous yeast, and particularly, in *Yarrowia lipolytica*. As is well known to one of skill in the art, merely inserting a gene (e.g., a desaturase) into a cloning vector does not ensure that it will be successfully expressed at the level needed. It may be desirable to manipulate a number of different genetic elements
25 that control aspects of transcription, translation, protein stability, oxygen limitation and secretion from the host cell. More specifically, gene expression may be controlled by altering any of the following: the nature of the relevant transcriptional promoter and terminator sequences; the number of copies of the cloned gene; whether the gene is plasmid-borne
30 or integrated into the genome of the host cell; the final cellular location of the synthesized foreign protein; the efficiency of translation in the host organism; the intrinsic stability of the cloned gene protein within the host cell; and the codon usage within the cloned gene, such that its frequency approaches the frequency of preferred codon usage of the host cell. Each
35 of these types of modifications are encompassed in the present invention, as a means to increase the yield of GLA. Additionally, one must consider that metabolic engineering directly within the PUFA biosynthetic pathway

may also be supplemented by additional manipulation of pathways that contribute carbon to the PUFA biosynthetic pathway and global regulators that affect GLA production. Several of these methods of modifying gene expression will be discussed below, as a means to increase GLA

5 production in oleaginous microbial host cells following selection of a particular $\Delta 12$ and $\Delta 6$ desaturase(s).

1) The Nature Of The Relevant Transcriptional Promoter And Terminator Sequence: Expression of the desaturase gene(s) can be increased at the transcriptional level through the use of a stronger promoter (either
10 regulated or constitutive) to cause increased expression, by removing/deleting destabilizing sequences from either the mRNA or the encoded protein, or by adding stabilizing sequences to the mRNA (U.S. 4,910,141).

Initiation control regions or promoters which are useful to drive
15 expression of desaturases in the desired host cell are numerous and familiar to those skilled in the art. Virtually any promoter capable of directing expression of these genes in the selected host cell is suitable for the present invention. Expression in a host cell can be accomplished in a transient or stable fashion. Transient expression
20 can be accomplished by inducing the activity of a regulatable promoter operably linked to the gene of interest; alternatively, stable expression can be achieved by the use of a constitutive promoter operably linked to the gene of interest. As an example, when the host cell is yeast, transcriptional and translational regions functional in yeast cells are
25 provided, particularly from the host species. The transcriptional initiation regulatory regions can be obtained, for example, from: 1.) genes in the glycolytic pathway, such as alcohol dehydrogenase, glyceraldehyde-3-phosphate-dehydrogenase (WO 2005/003310), phosphoglycerate mutase (WO 2005/003310), fructose-bisphosphate aldolase (WO 2005/049805), glycerol-3-phosphate o-acyltransferase
30 (U.S. Patent Application Number 60/610060), phosphoglucose-isomerase, phosphoglycerate kinase, etc.; or 2.) regulatable genes, such as acid phosphatase, lactase, metallothionein, glucoamylase, the translation elongation factor EF1- α (TEF) protein (U.S. 6,265,185),
35 ribosomal protein S7 (U.S. 6,265,185), etc. Any one of a number of regulatory sequences can be used, depending upon whether constitutive or induced transcription is desired, the efficiency of the

promoter in expressing the ORF of interest, the ease of construction and the like.

5 Nucleotide sequences surrounding the translational initiation codon 'ATG' have been found to affect expression in yeast cells. If the desired polypeptide is poorly expressed in yeast, the nucleotide sequences of exogenous genes can be modified to include an efficient yeast translation initiation sequence to obtain optimal gene expression. For expression in yeast, this can be done by site-directed mutagenesis of an inefficiently expressed gene by fusing it in-frame to an
10 endogenous yeast gene, preferably a highly expressed gene. Alternatively, one can determine the consensus translation initiation sequence in the host and engineer this sequence into heterologous genes for their optimal expression in the host of interest (see, e.g., WO2004/101753 for specific teachings applicable for *Yarrowia*
15 *lipolytica*).

The termination region can be derived from the 3' region of the gene from which the initiation region was obtained or from a different gene. A large number of termination regions are known and function satisfactorily in a variety of hosts (when utilized both in the same and
20 different genera and species from where they were derived). The termination region usually is selected more as a matter of convenience rather than because of any particular property. Preferably, the termination region is derived from a yeast gene, particularly *Saccharomyces*, *Schizosaccharomyces*, *Candida*, *Yarrowia* or
25 *Kluyveromyces*. The 3'-regions of mammalian genes encoding γ -interferon and α -2 interferon are also known to function in yeast. Termination control regions may also be derived from various genes native to the preferred hosts. Optionally, a termination site may be unnecessary; however, it is most preferred if included. Termination
30 regions used in the disclosure herein include the extracellular protease gene terminator (XPR), the 1-aminocyclopropane-1-carboxylate oxidase (Aco3) terminator and the 3-oxoacyl-coA thiolase (OCT) terminator, although these examples are not intended to be limiting.

2) The Number Of Copies Of The Cloned Gene: Additional copies of the
35 desaturase gene(s) may be introduced into the host to increase the output of the ω -6 fatty acid biosynthetic pathway and thereby increase GLA production. Specifically, additional copies may be cloned within a

single expression construct; and/or, additional copies of the cloned gene may be introduced into the host cell by increasing the plasmid copy number or by multiple integration of the cloned gene into the genome (*infra*).

- 5 3) Whether The Gene(s) Is Plasmid-Borne Or Integrated Into The Genome Of The Host Cell: Once the DNA encoding a desaturase polypeptide suitable for expression in an oleaginous yeast has been obtained, it is placed in a plasmid vector capable of autonomous replication in a host cell; or, it is directly integrated into the genome of
10 the host cell. Integration of expression cassettes can occur randomly within the host genome or can be targeted through the use of constructs containing regions of homology with the host genome sufficient to target recombination with the host locus. Where
15 constructs are targeted to an endogenous locus, all or some of the transcriptional and translational regulatory regions can be provided by the endogenous locus.

The preferred method of expressing genes in oleaginous yeast is by integration of linear DNA into the genome of the host; and, integration into multiple locations within the genome can be particularly
20 useful when high level expression of genes is desired. Toward this end, it is desirable to identify a sequence within the genome that is present in multiple copies.

Schmid-Berger et al. (*J. Bact.* 176(9):2477-2482 (1994)) discovered the first retrotransposon-like element *Ylt1* in *Yarrowia lipolytica*. This
25 retrotransposon is characterized by the presence of long terminal repeats (LTRs; each approximately 700 bp in length) called *zeta* regions. *Ylt1* and solo *zeta* elements were present in a dispersed manner within the genome in at least 35 copies/genome and 50-60 copies/genome, respectively; both elements were determined to
30 function as sites of homologous recombination. Further, work by Juretzek et al. (*Yeast* 18:97-113 (2001)) demonstrated that gene expression could be dramatically increased by targeting plasmids into the repetitive regions of the yeast genome (using linear DNA with LTR *zeta* regions at both ends), as compared to the expression obtained
35 using low-copy plasmid transformants. Thus, *zeta*-directed integration can be ideal as a means to ensure multiple integration of plasmid DNA into *Y. lipolytica*, thereby permitting high-level gene expression.

Unfortunately, however, not all strains of *Y. lipolytica* possess *zeta* regions (e.g., the strain identified as ATCC #20362). When the strain lacks such regions, it is also possible to integrate plasmid DNA comprising expression cassettes into alternate loci to reach the desired copy number for the expression cassette. For example, preferred alternate loci include: the *Ura3* locus (GenBank Accession No. AJ306421), the *Leu2* gene locus (GenBank Accession No. AF260230), the *Lys5* gene (GenBank Accession No. M34929), the *Aco2* gene locus (GenBank Accession No. AJ001300), the *Pox3* gene locus (Pox3: GenBank Accession No. XP_503244; or, Aco3: GenBank Accession No. AJ001301), the $\Delta 12$ desaturase gene locus, the *Lip1* gene locus (GenBank Accession No. Z50020) and/or the *Lip2* gene locus (GenBank Accession No. AJ012632).

4) The Final Cellular Location Of The Synthesized Foreign Protein:
Mounting evidence appears to indicate that exogenously supplied fatty acids enter the yeast cell by concomitant conversion to acyl-CoAs before their use for various metabolic purposes (Domergue, F. et al. *J. Biol. Chem.* 278(37):35115-35126 (2003)). Different desaturases, however, are preferentially able to perform desaturation in the acyl-CoA pool versus the phosphatidylcholine pool (acting preferentially on the *sn*-2 position); the functionality of the desaturase appears to be determined, in part, according to its origin (e.g., lower plants, worms, fungi, algae, human, etc.).

For some applications, it will be useful to direct the instant proteins to different cellular compartments. For the purposes described herein, GLA may as be found as free fatty acids or in esterified forms such as acylglycerols, phospholipids, sulfolipids or glycolipids. It is envisioned that the chimeric genes described above encoding polypeptides that permit GLA biosynthesis may be further engineered to include appropriate intracellular targeting sequences.

5) The Efficiency Of Translation In The Host Organism/ Codon Usage Within The Cloned Gene(s): Yet another approach to increase expression of the desaturase genes required for GLA production is to increase the translational efficiency of the encoded mRNAs by replacement of codons in the native gene with those for optimal gene expression in the selected host microorganism. Specifically, it is frequently useful to modify a portion of the codons encoding a

particular polypeptide that is to be expressed in a foreign host, such that the modified polypeptide uses codons that are preferred by the alternative host. Use of host preferred codons can substantially enhance the expression of the foreign gene encoding the polypeptide.

5 In general, host preferred codons can be determined within a particular host species of interest by examining codon usage in proteins (preferably those expressed in the largest amount) and determining which are used with highest frequency. Then, the coding sequence for a polypeptide of interest can be synthesized in whole or
10 in part using the codons preferred in the host species. All (or portions) of the DNA also can be synthesized to remove any destabilizing sequences or regions of secondary structure that would be present in the transcribed mRNA. And, all (or portions) of the DNA also can be synthesized to alter the base composition to one more preferable in the
15 desired host cell.

As an example of this technique, in the present invention it was desirable to utilize a synthetic codon-optimized $\Delta 6$ desaturase wherein a portion of the codons encoding the polypeptide had been modified to correspond to those preferred by the alternative host, *Yarrowia*
20 *lipolytica*. Specifically, in addition to modifications within the translation initiation site, 152 bp of the 1374 bp coding region of the *Mortierella alpina* $\Delta 6$ desaturase were modified (corresponding to 144 codons) to enhance the percent substrate conversion in *Y. lipolytica* from 30% to 42% (see WO 2004/101753, Example 8). The skilled artisan will
25 appreciate that these optimization methods will be equally applicable to other genes that one would like to express in a host for high-level expression of GLA.

6) The Intrinsic Stability Of The Cloned Gene and/or Expressed Protein Within The Host Cell: Juretzek et al. (*Yeast* 18:97-113 (2001)) note
30 that the stability of integrated plasmid copy number in *Yarrowia lipolytica* is dependent on the individual transformants, the recipient strain and the targeting platform used. Thus, the skilled artisan will recognize that multiple transformants must be screened in order to obtain a strain displaying the desired expression level and pattern.
35 Such screening may be accomplished by Southern analysis of DNA blots (Southern, *J. Mol. Biol.* 98:503 (1975)), Northern analysis of mRNA expression (Krocze, *J. Chromatogr. Biomed. Appl.*, 618

(1-2):133-145 (1993)), Western analysis of protein expression, phenotypic analysis or GC analysis of the PUFA products.

- 7) Manipulations Within The ω -3/ ω -6 Biosynthetic Pathway: In the case of manipulations within the ω -3/ ω -6 biosynthetic pathway, it may be desirable to increase the production of LA to enable increased production of GLA. This may be accomplished by introducing (and/or amplifying) the genes encoding Δ 6 and/or Δ 12 desaturases.

Additionally, it is well known to one skilled in the art that GLA production is favored in a host microorganism that is substantially free of ALA. Thus, preferably, the host is selected or obtained by removing or inhibiting Δ 15 or ω -3 type desaturase activity that permits conversion of LA to ALA. The endogenous desaturase activity can be reduced or eliminated by, for example: 1.) providing a cassette for transcription of antisense sequences to the Δ 15 desaturase transcription product; 2.) disrupting the Δ 15 desaturase gene through insertion, substitution and/or deletion of all or part of the target gene; or 3.) using a host cell which naturally has [or has been mutated to have] low or no Δ 15 desaturase activity. Inhibition of undesired desaturase pathways can also be accomplished through the use of specific desaturase inhibitors such as those described in U.S. 4,778,630.

- 8) Manipulation Of Pathways Contributing Carbon To The ω -3/ ω -6 Biosynthetic Pathway: Beyond the immediate PUFA biosynthetic pathway, it is expected that manipulation of several other enzymatic pathways leading to the biosynthesis of precursor fatty acids may contribute to the overall net biosynthesis of specific PUFAs. For example, it may be useful to increase the flow of carbon into the PUFA biosynthetic pathway by increasing the availability of the precursors of longer chain saturated and unsaturated fatty acids, such as palmitate (16:0), stearic acid (18:0) and oleic acid (18:1). The synthesis of the palmitate is dependent on the activity of a $C_{14/16}$ elongase, while the synthesis of stearic acid is dependent on the activity of a $C_{16/18}$ elongase; oleic acid is the product of the reaction catalyzed by Δ 9 desaturase. Thus, in some embodiments of the present invention, it may be useful to over-express a Δ 9 desaturase, $C_{14/16}$ elongase and/or a $C_{16/18}$ elongase, in addition to the GLA biosynthetic pathway enzymes to thereby increase the substrate availability for those PUFA enzymes. The work of Inagaki, K. et al. (*Biosci. Biotech. Biochem.*

66(3):613-621 (2002)) specifically suggests advantages associated with co-expression of a heterologous C_{16/18} elongase from *Rattus norvegicus* (GenBank Accession No. AB071986). And, subsequent work by the Applicants has confirmed that over-expression of a codon-
5 optimized C_{16/18} elongase from *R. norvegicus* can substantially increase carbon flow into the PUFA pathway, by increasing conversion of C16 to C18 fatty acids. Thus, although a *Yarrowia lipolytica* host strain must minimally be manipulated to express a $\Delta 6$ desaturase and $\Delta 12$ desaturase for GLA biosynthesis, in a further preferred
10 embodiment the host strain additionally includes at least one of the following: a $\Delta 9$ desaturase, C_{14/16} elongase and/or a C_{16/18} elongase. Up-regulation of selected biochemical pathways and gene products may be controlled using the techniques described above. In contrast, it may be necessary to down-regulate portions of certain biochemical
15 pathways competing with the fatty acid biosynthetic pathway for energy or carbon. If so, these undesirable biosynthetic pathways may be eliminated by gene disruption or down-regulated by other means (e.g., antisense technology, mutagenesis with radiation or chemical agents or by use of transposable elements/ transposons), as reviewed in
20 WO2004/101757.

Within the context of the present invention, it may be useful to modulate the expression of a GLA biosynthetic pathway by any one of the methods described above. For example, the present invention provides
25 methods whereby genes encoding $\Delta 6$ desaturase and $\Delta 12$ desaturase activities are introduced into oleaginous yeast such as *Yarrowia lipolytica* for the high-level production of GLA. It will be particularly useful to express these genes in oleaginous yeast that do not naturally possess ω -3 and/or ω -6 fatty acid biosynthetic pathways and to coordinate the
30 expression of these genes to maximize production of GLA using various means of metabolic engineering within the host organism.

Expression Systems, Cassettes, Vectors and Transformation

Microbial expression systems and expression vectors containing regulatory sequences that direct high-level expression of foreign proteins
35 such as GLA are well known to those skilled in the art. Any of these could be used to construct chimeric genes encoding the preferred desaturase sequences. These chimeric genes could then be introduced into

appropriate heterologous microbial host cells, particularly in the cells of oleaginous yeast (e.g., *Yarrowia lipolytica*), via transformation to provide high-level expression of the encoded enzymes.

Vectors or DNA cassettes useful for the transformation of suitable host cells are well known in the art. The specific choice of sequences present in the construct is dependent upon the desired expression products, the nature of the host cell, and the proposed means of separating transformed cells versus non-transformed cells. Typically, however, the vector or cassette contains sequences directing transcription and translation of the relevant gene(s), a selectable marker and sequences allowing autonomous replication or chromosomal integration. Suitable vectors comprise a region 5' of the gene that controls transcriptional initiation and a region 3' of the DNA fragment that controls transcriptional termination. It is most preferred when both control regions are derived from genes from the transformed host cell, although it is to be understood that such control regions need not be derived from the genes native to the specific species chosen as a production host.

Where two or more genes are expressed from separate replicating vectors, it is desirable that each vector has a different means of selection and should lack homology to the other constructs to maintain stable expression and prevent reassortment of elements among constructs. Judicious choice of regulatory regions, selection means and method of propagation of the introduced construct(s) can be experimentally determined so that all introduced genes are expressed at the necessary levels to provide for synthesis of the desired products.

Constructs comprising the gene(s) of interest may be introduced into a host cell by any standard technique. These techniques include transformation (e.g., lithium acetate transformation [*Methods in Enzymology*, 194:186-187 (1991)]), protoplast fusion, bolistic impact, electroporation, microinjection, or any other method that introduces the gene(s) of interest into the host cell. More specific teachings applicable for oleaginous yeast (i.e., *Yarrowia lipolytica*) include U.S. Patents No. 4,880,741 and No. 5,071,764 and Chen, D. C. et al. (*Appl Microbiol Biotechnol.* 48(2):232-235 (1997)).

For convenience, a host cell that has been manipulated by any method to take up a DNA sequence (e.g., an expression cassette) will be referred to as "transformed" or "recombinant" herein. The transformed

host will have at least one copy of the expression construct and may have two or more, depending upon whether the gene is integrated into the genome, amplified, or is present on an extrachromosomal element having multiple copy numbers. The transformed host cell can be identified by
5 various selection techniques, as described in WO2004/101757. Preferred selection methods for use herein are resistance to kanamycin, hygromycin and the amino glycoside G418, as well as ability to grow on media lacking uracil, leucine, lysine, tryptophan or histidine.

Accordingly, it is expected that introduction of chimeric genes
10 encoding a GLA biosynthetic pathway (e.g., a $\Delta 12$ desaturase and $\Delta 6$ desaturase), under the control of appropriate promoters will result in production of GLA when the microbial host cell is provided with oleic acid as the desaturase substrate. The level of $\Delta 12$ desaturase gene expression and $\Delta 6$ desaturase gene expression required will depend on
15 the host cell, its ability to synthesize PUFAs using endogenous desaturases/elongases and the availability of substrate (which may be produced endogenously or be supplied exogenously). Using the teachings provided herein, transformant oleaginous microbial hosts will produce at least about 15% GLA in the total lipids, preferably at least
20 about 20% GLA in the total lipids, more preferably at least about 30% GLA in the total lipids, more preferably at least about 40% GLA in the total lipids and most preferably at least about 50% GLA in the total lipids.

Fermentation Processes For GLA Production

The transformed microbial host cell is grown under conditions that
25 optimize desaturase activities and produce the greatest and the most economical yield of GLA. In general, media conditions that may be optimized include the type and amount of carbon source, the type and amount of nitrogen source, the carbon-to-nitrogen ratio, the oxygen level, growth temperature, pH, length of the biomass production phase, length of
30 the oil accumulation phase and the time of cell harvest. Microorganisms of interest, such as oleaginous yeast, are grown in complex media (e.g., yeast extract-peptone-dextrose broth (YPD)) or a defined minimal media that lacks a component necessary for growth and thereby forces selection of the desired expression cassettes (e.g., Yeast Nitrogen Base (DIFCO
35 Laboratories, Detroit, MI)).

Fermentation media in the present invention must contain a suitable carbon source. Suitable carbon sources may include, but are not limited

to: monosaccharides (e.g., glucose, fructose), disaccharides (e.g., lactose, sucrose), oligosaccharides, polysaccharides (e.g., starch, cellulose or mixtures thereof), sugar alcohols (e.g., glycerol) or mixtures from renewable feedstocks (e.g., cheese whey permeate, cornsteep liquor, sugar beet molasses, barley malt). Additionally, carbon sources may include alkanes, fatty acids, esters of fatty acids, monoglycerides, diglycerides, triglycerides, phospholipids and various commercial sources of fatty acids including vegetable oils (e.g., soybean oil) and animal fats. Additionally, the carbon source may include one-carbon sources (e.g., carbon dioxide, methanol, formaldehyde, formate, carbon-containing amines) for which metabolic conversion into key biochemical intermediates has been demonstrated. Hence it is contemplated that the source of carbon utilized in the present invention may encompass a wide variety of carbon-containing sources and will only be limited by the choice of host organism. Although all of the above-mentioned carbon sources and mixtures thereof are expected to be suitable in the present invention, preferred carbon sources are sugars and/or fatty acids. Most preferred is glucose and/or fatty acids containing between 10-22 carbons.

Nitrogen may be supplied from an inorganic (e.g., $(\text{NH}_4)_2\text{SO}_4$) or organic (e.g., urea or glutamate) source. In addition to appropriate carbon and nitrogen sources, the fermentation media must also contain suitable minerals, salts, cofactors, buffers, vitamins and other components known to those skilled in the art, appropriate for the growth of the microorganism and promotion of the enzymatic pathways necessary for GLA production. Particular attention is given to several metal ions (e.g., Mn^{+2} , Co^{+2} , Zn^{+2} , Mg^{+2}) that promote synthesis of lipids and PUFAs (Nakahara, T. et al. *Ind. Appl. Single Cell Oils*, D. J. Kyle and R. Colin, eds. pp 61-97 (1992)).

Preferred growth media in the present invention are common commercially prepared media, such as Yeast Nitrogen Base (DIFCO Laboratories, Detroit, MI). Other defined or synthetic growth media may also be used and the appropriate medium for growth of the particular microorganism will be known by one skilled in the art of microbiology or fermentation science. A suitable pH range for the fermentation is typically between about pH 4.0 to pH 8.0, wherein pH 5.5 to pH 7.0 is preferred as the range for the initial growth conditions. The fermentation may be conducted under aerobic or anaerobic conditions, wherein microaerobic conditions are preferred.

Typically, accumulation of high levels of PUFAs such as GLA in oleaginous yeast cells requires a two-stage process, since the metabolic state must be "balanced" between growth and synthesis/storage of fats. Thus, most preferably, a two-stage fermentation process is necessary for the production of PUFAs in oleaginous yeast. This approach is described in WO 2004/101757, as are various suitable fermentation process designs (i.e., batch, fed-batch and continuous) and considerations during growth.

Purification of PUFAs

The PUFAs may be found in the host microorganism as free fatty acids or in esterified forms such as acylglycerols, phospholipids, sulfolipids or glycolipids, and may be extracted from the host cell through a variety of means well-known in the art. One review of extraction techniques, quality analysis and acceptability standards for yeast lipids is that of Z. Jacobs (*Critical Reviews in Biotechnology* 12(5/6):463-491 (1992)). A brief review of downstream processing is also available by A. Singh and O. Ward (*Adv. Appl. Microbiol.* 45:271-312 (1997)).

In general, means for the purification of PUFAs such as GLA may include extraction with organic solvents, sonication, supercritical fluid extraction (e.g., using carbon dioxide), saponification, and physical means such as presses, or combinations thereof. One is referred to the teachings of WO 2004/101757 for additional details.

DESCRIPTION OF PREFERRED EMBODIMENTS

The present invention demonstrates the feasibility of engineering high-level production of GLA in oleaginous yeast. Toward this end, a GLA biosynthetic pathway was introduced into *Yarrowia lipolytica* by the introduction of genes encoding $\Delta 12$ and $\Delta 6$ desaturase activities. Specific modifications that were designed to increase production of GLA comprised: (1) use of *zeta*-directed integration to ensure multiple integration of expression cassettes into the *zeta* loci of the *Y. lipolytica* genome; (2) introduction of $\Delta 6$ desaturase activity by incorporation of a TEF:: $\Delta 6$::XPR chimeric gene, wherein the $\Delta 6$ desaturase was a synthetic gene (originally derived from *Mortierella alpina*) that was codon-optimized for expression in *Yarrowia*; (3) up-regulation of endogenous $\Delta 12$ desaturase activity, by creation of a TEF:: $\Delta 12$::XPR chimeric gene; optionally, (4) addition of a second $\Delta 6$ desaturase, in the form of a TEF:: $\Delta 6$::OCT chimeric gene, wherein the $\Delta 6$ desaturase gene was isolated from *Saprolegnia diclina*; and optionally, (5) insertion of a

GPD:: $\Delta 12$::Aco3 chimeric gene, wherein expression of the *Yarrowia* $\Delta 12$ desaturase was driven by a glyceraldehyde-3-phosphate-dehydrogenase (GPD) promoter of *Yarrowia*.

Upon transformation of plasmid pDMW208, wherein a single copy
5 each of a $\Delta 12$ desaturase gene and a $\Delta 6$ desaturase gene were integrated into the *zeta* loci of the *Y. lipolytica* genome, GC analysis of the host cells determined that those transformants produced 25.9% GLA in the total lipids. In contrast, recombinant *Y. lipolytica* that had been transformed with plasmid pDMW208D62, such that two copies each of a $\Delta 12$
10 desaturase gene and a $\Delta 6$ desaturase gene were integrated into the *zeta* loci of the *Y. lipolytica* genome, produced 34% GLA in the total lipids.

Although the Applicants demonstrate production of 25.9% and 34% GLA in these particular recombinant strains of *Y. lipolytica*, it will be apparent that the concentration of GLA in the host cells could be
15 dramatically increased via additional genetic modifications, according to the invention herein and as suggested in U.S. Patent Application No. 60/624812. Furthermore, on the basis of the teachings and results described herein, it is expected that one skilled in the art will recognize the feasibility and commercial utility created by using oleaginous yeast as a
20 production platform for the synthesis of a variety of ω -3 and/or ω -6 PUFAs.

EXAMPLES

The present invention is further defined in the following Examples. It should be understood that these Examples, while indicating preferred embodiments of the invention, are given by way of illustration only. From
25 the above discussion and these Examples, one skilled in the art can ascertain the essential characteristics of this invention, and without departing from the spirit and scope thereof, can make various changes and modifications of the invention to adapt it to various usages and conditions.

GENERAL METHODS

Standard recombinant DNA and molecular cloning techniques used in the Examples are well known in the art and are described by:

- 1.) Sambrook, J., Fritsch, E. F. and Maniatis, T. *Molecular Cloning: A Laboratory Manual*; Cold Spring Harbor Laboratory: Cold Spring Harbor,
35 NY (1989) (hereinafter "Maniatis"); 2.) T. J. Silhavy, M. L. Bennis, and L. W. Enquist, *Experiments with Gene Fusions*; Cold Spring Harbor Laboratory: Cold Spring Harbor, NY (1984); and 3.) Ausubel, F. M. et al.,

Current Protocols in Molecular Biology, published by Greene Publishing Assoc. and Wiley-Interscience (1987).

Materials and methods suitable for the maintenance and growth of microbial cultures are well known in the art. Techniques suitable for use in the following examples may be found as set out in Manual of Methods for General Bacteriology (Phillipp Gerhardt, R. G. E. Murray, Ralph N. Costilow, Eugene W. Nester, Willis A. Wood, Noel R. Krieg and G. Briggs Phillips, Eds), American Society for Microbiology: Washington, D.C. (1994)); or by Thomas D. Brock in Biotechnology: A Textbook of Industrial Microbiology, 2nd ed., Sinauer Associates: Sunderland, MA (1989). All reagents, restriction enzymes and materials used for the growth and maintenance of microbial cells were obtained from Aldrich Chemicals (Milwaukee, WI), DIFCO Laboratories (Detroit, MI), GIBCO/BRL (Gaithersburg, MD) or Sigma Chemical Company (St. Louis, MO), unless otherwise specified.

E. coli (XL1-Blue) competent cells were purchased from the Stratagene Company (San Diego, CA). *E. coli* strains were typically grown at 37°C on Luria Bertani (LB) plates.

A leucine autotrophic strain of *Yarrowia lipolytica* was purchased from the American Type Culture Collection (Rockville, MD; ATCC #76982) and used for functional assays. *Y. lipolytica* strains were usually grown at 28°C on YPD agar (1% yeast extract, 2% bactopectone, 2% glucose, 2% agar). For selection of transformants, minimal medium (0.17% yeast nitrogen base (DIFCO, Detroit, MI) without ammonium sulfate or amino acids, 2% glucose, 0.1% proline, pH 6.1) was used. Supplements of adenine, leucine, lysine and/or uracil were added to a final concentration of 0.01%.

General molecular cloning was performed according to standard methods (Sambrook et al., *supra*). Oligonucleotides were synthesized by Sigma-Genosys (Spring, TX). Site-directed mutagenesis was performed using Stratagene's QuikChange™ Site-Directed Mutagenesis kit, per the manufacturer's instructions. When polymerase chain reaction (PCR) or site-directed mutagenesis was involved in subcloning, the constructs were sequenced to confirm that no errors had been introduced to the sequence. PCR products were cloned into Promega's pGEM-T-easy vector (Madison, WI).

Manipulations of genetic sequences were accomplished using the suite of programs available from the Genetics Computer Group, Inc.

(Wisconsin Package Version 9.0, Genetics Computer Group (GCG), Madison, WI). The GCG program "Pileup" was used with the gap creation default value of 12, and the gap extension default value of 4. The GCG "Gap" or "Bestfit" programs were used with the default gap creation
5 penalty of 50 and the default gap extension penalty of 3. Unless otherwise stated, in all other cases GCG program default parameters were used.

The meaning of abbreviations is as follows: "sec" means second(s), "min" means minute(s), "hr" means hour(s), "d" means day(s), "μL" means microliter(s), "mL" means milliliter(s), "L" means liter(s), "μM"
10 means micromolar, "mM" means millimolar, "M" means molar, "mmol" means millimole(s), "μmole" mean micromole(s), "g" means gram(s), "μg" means microgram(s), "ng" means nanogram(s), "U" means unit(s), "bp" means base pair(s) and "kB" means kilobase(s).

EXAMPLE 1

Construction Of A Zeta Integration Plasmid

Plasmid pYZ2A (Figure 2) was constructed as a derivative of pYZV5 to facilitate integration of the plasmid into the *zeta* regions of the *Yarrowia lipolytica* genome. Briefly, pYZV5 (WO2004/101757, Example 4) comprised the following: (1) an ampicillin-resistance gene (Amp^R); (2)
20 401 bp of 5'-sequence and 568 bp of 3'-sequence from the *Y. lipolytica* URA3 gene to direct integration of expression cassettes into the Ura loci of the *Yarrowia* genome; (3) a *Yarrowia* isopropylmalate isomerase (LEU2) gene; (4) the *Y. lipolytica* translation elongation factor EF1- α (TEF) protein promoter (U.S. 6,265,185); (5) a *Mortierella alpina* (GenBank Accession
25 #AF067654; U.S. 6,075,183) $\Delta 5$ desaturase; and (6) the extracellular protease gene terminator (XPR). Plasmid pYZ2A was constructed by substituting fragments of the *zeta* region for the fragments directing integration into the URA3 gene of the pYZV5 construct.

Specifically, genomic DNA was purified from *Y. lipolytica* ATCC
30 #79682 using Qiagen's DNeasy Tissue Kit in the following manner: a 5 mL overnight culture of ATCC #79682 was spun down and washed in 5 mL SCE buffer (1 M sorbital, 0.1 M sodium citrate and 60 mM EDTA), before being resuspended again in 50 μL SCE containing 2 mM β -mercaptoethanol and 10 units zymolyase (ICN Biomedicals, Inc.). The
35 cells were then incubated at 37 °C for 1 hr, prior to 15 min incubation at 70 °C to inactivate the zymolyase. Qiagen's protocol was resumed following the inactivation step and was followed for the remainder of the DNA

isolation. PCR amplification was carried out in a 50 µl total volume containing: 100 ng *Yarrowia* genomic DNA, PCR buffer containing 10 mM KCl, 10 mM (NH₄)₂SO₄, 20 mM Tris-HCl (pH 8.75), 2 mM MgSO₄, 0.1% Triton X-100, 100 µg/mL BSA (final concentration), 200 µM each deoxyribonucleotide triphosphate, 10 pmole of each primer (*infra*) and 1 µl of PfuTurbo DNA polymerase (Stratagene, San Diego, CA). Amplification was carried out as follows: initial denaturation at 95 °C for 3 min, followed by 35 cycles of the following: 95 °C for 1 min, 56 °C for 30 sec, 72 °C for 1 min. A final extension cycle of 72 °C for 10 min was carried out, followed by reaction termination at 4 °C.

Primers for the amplification of the *zeta* regions were designed according to the sequence of *Ylt1*, a highly repetitive retrotransposon in the genome of *Yarrowia lipolytica* (Schmid-Berger, N. et al., *J Bacteriol.* 176(9):2477-82 (1994)). These primers are shown below in Table 3; underlined regions within each primer sequence correspond with a specific restriction enzyme site that was designed to enable downstream cloning reactions.

TABLE 3
Primers For Amplification Of The *Zeta* Regions

Primer	Sequence	Encoded Restriction Enzyme Site	SEQ ID NO
YL127	TTT <u>G</u> GCGCGCCTGTGCGTACCAG GGATAGGGTAGG	<i>Asc1</i>	SEQ ID NO:1
YL128	TTTC <u>G</u> TACGGACCACTCCCTTGC ACTTCTTG	<i>BsiW1</i>	SEQ ID NO:2
YL129	GGGTTAATTA <u>A</u> CGCCGATTCTCTC TGCAGTCG	<i>Pac1</i>	SEQ ID NO:3
YL130	TTTGCATGCTCTGGAGAGTTAGTC ATCCGAC	<i>Sph1</i>	SEQ ID NO:4

Amplification of the 3' *zeta* region of *Y. lipolytica* utilized primers YL129 and YL130 and the genomic DNA described above. The product was digested with *Pac1* and *Sph1*, gel-purified, and the resulting 359 bp

PCR fragment (SEQ ID NO:6) was cloned into *Pac1* and *Sph1* digested pYZV5 (WO2004/101757). The plasmid was named pYZ1A (Figure 2).

A second PCR reaction was conducted with primers YL127 and YL128 to amplify the 5' *zeta* region of *Y. lipolytica*. The PCR product was
5 digested with *Asc1* and *BsiW1*, gel-purified, and the 265 bp fragment (SEQ ID NO:5) was cloned into *Asc1*- and *BsiW1*-digested pYZ1A, to produce pYZ2A (Figure 2).

Thus, pYZ2A possessed a 265 bp 5'-sequence (SEQ ID NO:5) from the *Y. lipolytica* *zeta* region and a 359 bp 3'-sequence (SEQ ID NO:6) from
10 the *Y. lipolytica* *zeta* region suitable to direct integration of expression cassettes into the *zeta* loci of the *Yarrowia* genome.

EXAMPLE 2

Construction Of A High GLA Producing Plasmid, pDMW208, Comprising A $\Delta 6$ Desaturase And A $\Delta 12$ Desaturase

15 In order to produce high levels of GLA, a variety of modifications to plasmid pYZ2A (from Example 1) were engineered. Specifically, this Example describes the insertion of a synthetic $\Delta 6$ desaturase gene (responsible for conversion of LA to GLA) and a native $\Delta 12$ desaturase gene (responsible for conversion of oleic acid to LA) to ultimately produce
20 plasmid pDMW208. The latter enzymatic activity was utilized to increase the carbon flux through the GLA biosynthetic pathway; the former activity was required for the synthesis of the desired product, GLA.

Cloning Of A Synthetic Codon-Optimized $\Delta 6$ Desaturase

Plasmid pDMW200 was synthesized by cloning the $\Delta 6$ desaturase
25 from pYD6S (described in WO2004/101753 and illustrated herein as Figure 3A; $\Delta 6$ desaturase gene labeled as "D6Syn") into *Not1*- and *Nco1*-digested pYZ2A (Example 1). Specifically, pYD6S contains a synthetic DNA fragment encoding $\Delta 6$ desaturase (SEQ ID NO:7) that is codon-optimized for expression in *Yarrowia*, wherein 152 bp of the 1374 bp
30 coding region of the wildtype *Mortierella alpina* $\Delta 6$ desaturase (GenBank Accession #AF46528) were modified (corresponding to 144 codons), in addition to modifications within the translation initiation site. This optimization led to an increase in percent substrate conversion (calculated as [product/(substrate + product)]*100) of the gene from 30% to 42%,
35 upon expression in *Yarrowia* under the control of the TEF promoter. As a result of cloning, pDMW200 comprised a 1949 bp TEF:: $\Delta 6$::XPR chimeric gene (SEQ ID NO:9).

pDMW200-B was then constructed as a derivative of pDMW200 to facilitate subcloning and heterologous gene expression in *Y. lipolytica*. Specifically, pDMW200-B was constructed by a single round of site-directed mutagenesis using pDMW200 as a template. A *Bgl*III site was introduced into pDMW200 in the 3' end of the leucine gene, 44 bases upstream of the existing *Pme*I site using oligonucleotides YL238 (SEQ ID NO:10) and YL239 (SEQ ID NO:11).

Cloning Of A *Yarrowia lipolytica* Δ 12 Desaturase

Although wildtype *Yarrowia* has a Δ 12 desaturase gene (sequenced and biochemically characterized in WO2004/104167), a copy of this gene was introduced into pDMW200-B to amplify the endogenous Δ 12 desaturase activity. Specifically, the Δ 12 desaturase gene was derived from plasmid pY25-d12d (illustrated herein as Figure 3B; see WO2004/104167, Example 4 for further details). Using oligonucleotides YL242 (SEQ ID NO:12) and YL243 (SEQ ID NO:13) as primers and pY25-d12d as template, a *Pme*I site was introduced into pY25-d12d by site-directed mutagenesis to generate pY25-d12d-P. A second round of mutagenesis was then conducted using oligonucleotides YL240 (SEQ ID NO:14) and YL241 (SEQ ID NO:15) as primers to introduce a *Bgl*III site into pY25-d12d-P, thereby generating pY25-d12d-PB (Figure 3C).

The Δ 12 desaturase gene, flanked by a TEF promoter and an XPR terminator, was isolated from *Bgl*III/*Pme*I-digested pY25-d12d-PB. The ~1.9 kB fragment (SEQ ID NO:16) was gel purified and ligated into *Bgl*III/*Pme*I-digested pDMW200-B. The resulting plasmid was called pDMW208 (SEQ ID NO:17).

As shown in Figure 3D, the pDMW208 expression vector is a 9541 bp plasmid comprising: a Δ 12 desaturase gene ("d12") flanked by a TEF promoter and an XPR terminator, a Δ 6 desaturase gene flanked by a TEF promoter and an XPR terminator and a leucine selection marker. These genes are flanked by sequences specific for the *zeta* regions of *Yarrowia*. After a *Sph*I/*Asc*I digestion, this plasmid was targeted to integrate into the *zeta* regions of *Yarrowia* chromosomes. GC analyses of the transformants selected on minimal media confirmed production of GLA (see Example 5).

EXAMPLE 3

Construction Of A High GLA Producing Plasmid, pDMW208D62, Comprising Two $\Delta 6$ Desaturases And Two $\Delta 12$ Desaturases

As is well known in the art, gene expression can frequently be
5 increased by amplifying the number of copies of the cloned gene(s). To
test this hypothesis and possibly up-regulate GLA production with respect
to that obtainable by expression of pDMW208 (Example 2), this Example
describes the insertion of a second $\Delta 6$ desaturase gene and a second $\Delta 12$
desaturase gene into pDMW208 to produce plasmid pDMW208D62.

10 Cloning Of A *Saprolegnia diclina* $\Delta 6$ Desaturase

An additional copy of a $\Delta 6$ desaturase gene cloned under control of
the TEF promoter and the OCT terminator was cloned into plasmid
pDMW208. First, the 1366 bp *S. diclina* $\Delta 6$ desaturase gene was
amplified using plasmid pRSP1 (Abbott Laboratories, U.S. Patent
15 6,635,451; GenBank Accession No. AX576988) as the template and
oligonucleotides YL253 (SEQ ID NO:18) and YL254 (SEQ ID NO:19) as
primers. PCR amplification was carried out in a 50 μ l total volume
containing 10 ng pRSP1 plasmid DNA, using the components and
amplification conditions described in Example 1. The PCR product was
20 digested with *Nco*1 and *Not*1 and gel purified.

The OCT terminator of the gene encoding 3-oxoacyl-coA thiolase
(GenBank X69988) was amplified from *Y. lipolytica* genomic DNA using
YL293 (SEQ ID NO:20) and YL317 (SEQ ID NO:21) as primers. The 372
bp PCR product was digested with *Not*1 and *Cla*1 and gel purified.

25 Finally, the TEF promoter was obtained from plasmid pYSD17SPC
(WO2004/101757, Example 4). Specifically, plasmid pYSD17SPC
(illustrated herein as Figure 3E) was linearized with *Nco*1 and *Cla*1. Then,
the *S. diclina* $\Delta 6$ desaturase gene, the 372 bp PCR product encoding the
OCT terminator and pYSD17SPC were all ligated together in a single
30 standard overnight ligation reaction. The resulting plasmid was named
pYTS6 (Figure 4A). pYTS6 was digested with *Cla*1 and *Pme*1 to release
a ~2.2 kB fragment (SEQ ID NO:22) containing the *S. diclina* $\Delta 6$
desaturase gene, flanked by the TEF promoter and OCT terminator. This
fragment was cloned into *Cla*1 and *Pme*1-digested pDMW208 to yield
35 plasmid pDMW208D6 (Figure 4B).

Cloning Of A Second *Yarrowia lipolytica* $\Delta 12$ Desaturase

An additional copy of the *Yarrowia* $\Delta 12$ desaturase gene, cloned under control of a *Yarrowia* glyceraldehyde-3-phosphate dehydrogenase (GPD) promoter and an Aco3 terminator, was introduced into pDMW208D6. The GPD promoter, isolated and biochemically characterized in WO 2005/003310, was determined to be about 3 times stronger than the bench-marker TEF promoter in *Y. lipolytica* (activity measured according to GUS expression). Thus, the $\Delta 12$ desaturase gene from pY25-d12d-PB (Example 2) was modified to replace the existing TEF promoter with a GPD promoter and to replace the existing XPR terminator with an Aco3 terminator. This modified construct was then introduced into pDMW208D6 (*supra*) to yield plasmid pDMW208D62. The cloning scheme for the construction of pDMW208D62 is described below. Specifically, the construction of pDMW208D62 incorporated promoters and transcription terminators from pre-existing plasmids that contained genes unrelated to the purposes described herein. These unnecessary genes were removed from the construct prior to completion of pDMW208D62 synthesis.

First, the *Yarrowia* GPD promoter fragment was isolated from pYZGDG (WO 2005/003310, Example 5; illustrated herein as Figure 4C). pYZGDG was completely digested with *Sph1* and then partially digested with *Nco1*. The 3.2 kB *Sph1*/*Nco1* fragment containing the chimeric GPD::LEU2 gene of pYZGDG was used to replace the *Sph1*/*Nco1* fragment of pELS containing the chimeric TEF::LEU2 gene (WO2004/101753, Example 6; illustrated herein as Figure 4D) to generate pYZDEL1S.

Four consecutive rounds of site-directed mutagenesis were then conducted using pYZDEL1S as a template to ultimately produce pYZDE1SB-CN. Specifically, a first round of site-directed mutagenesis allowed for the introduction of a *Swa1* site into pYZDEL1S using primers YL226 and YL227 (SEQ ID NOs:23 and 24) to generate pYZDEL1S-S. A second round of site-directed mutagenesis with primers YL61 and YL62 (SEQ ID NOs:25 and 26) introduced a *BsiW1* site into pYZDEL1S-S to generate pYDDE1-SB. The third round of mutagenesis eliminated the *Cla1* site in the GPD promoter region of pYDDE1-SB, using primers YL230 and YL231 (SEQ ID NOs:27 and 28), to create pYZDE1SB. The final round of mutagenesis eliminated the *Nco1* site within the GPD

promoter region of pYZDE1SB with primers YL228 and YL229 (SEQ ID NOs:29 and 30), to thereby generate pYZDE1SB-CN.

Using pRAT-4-A7 (Abbott Laboratories, U.S. Patent 6,677,145) as template and oligonucleotides YL179 and YL179A (SEQ ID NOs:31 and 32) as primers, an elongase gene from *Thraustochytrium aureum* T7091 (abbreviated as "EL2") was PCR amplified using standard conditions and inserted into the *NcoI/NotI* site of pY5-13 (WO2004/101757, Example 1; see Figure 4E herein) to generate pELS2, comprising the chimeric TEF::EL2 gene. The *NotI/NcoI* fragment from pELS2 was then introduced into *NotI/NcoI* digested pYZDE1SB-CN to generate pGDE2S, comprising the chimeric GPD::EL2::XPR gene.

pGDE2S was subjected to two consecutive rounds of mutagenesis. The first site-directed mutagenesis step used primers YL77 and YL78 (SEQ ID NOs:33 and 34) to add a *PacI* site to pGDE2S, generating pGDE2S-P. The second step used primers YL322 and Y323 (SEQ ID NOs:35 and 36) to add a *ClaI* site to pGDE2S-P to generate pGDE2S-PC.

The final step in the construction of pGDE2ST (Figure 5) was addition of the ACO terminator. The terminator was amplified from *Y. lipolytica* genomic DNA by PCR with primers YL325 and YL326 (SEQ ID NOs:37 and 38) using standard PCR conditions, digested with *NotI* and *PacI* and ligated with *NotI/PacI*-digested pGDE2S-PC to generate pGDE2ST. Thus, pGDE2ST comprised a chimeric GPD::EL2::Aco gene.

Final Synthesis Of Plasmid pDMW208D62

As shown in Figure 5, the $\Delta 12$ desaturase gene contained on pY25-d12d-PB (Example 2) was excised from the plasmid by *NcoI/NotI* digestion and cloned into the *NcoI/NotI* digested pGDE2ST plasmid to generate pZGD12-PC, comprising the chimeric GPD:: $\Delta 12$::Aco gene. The ~2.6 kbp *ClaI/PacI* fragment (SEQ ID NO:39) of pZGD12-PC containing the GPD:: $\Delta 12$::Aco chimeric gene was then inserted into the *ClaI/PacI*-digested pDMW208D6 to generate pDMW208D62 (SEQ ID NO:40).

EXAMPLE 4

Transformation Of *Yarrowia lipolytica*

The plasmids pDMW208 and pDMW208D62 were digested with *AscI* and *SphI* restriction endonucleases and transformed into a leucine auxotroph mutant of *Y. lipolytica* ATCC #76982 cells according to the lithium acetate method of Chen, D. C. et al. (*Appl Microbiol Biotechnol.* 48(2):232-235 (1997)). Briefly, *Y. lipolytica* was streaked onto a YPD plate and grown at 30 °C for approximately 18

hr. Several large loopfuls of cells were scraped from the plate and resuspended in 1 mL of transformation buffer containing: 2.25 mL of 50% PEG, average MW 3350; 0.125 mL of 2 M Li acetate, pH 6.0; 0.125 mL of 2 M DTT; and, 50 μ g sheared salmon sperm DNA.

- 5 About 500 ng of plasmid DNA was incubated in 100 μ l of resuspended cells, and maintained at 39 °C for 1 hr with vortex mixing at 15 min intervals. The cells were plated onto minimal media (with 0.1 g/L L-adenine and 0.1 g/L L-lysine) plates lacking leucine and maintained at 30 °C for 2 to 3 days.

10

EXAMPLE 5

GLA Production In *Yarrowia*

- Colonies of transformant *Y. lipolytica* containing pDMW208 or pDMW208D62 were each grown in 20 mL minimal media in a 125 mL Erlenmeyer flask at 30 °C. After about 24 hr, when the cultures reached
- 15 an OD₆₀₀ ~1.0, one 5 mL aliquot from each culture was removed and harvested. The pellet was resuspended in 4 mL of FRF media (7 g KH₂PO₄, 2 g K₂HPO₄, 1 g MgSO₄ • 7H₂O, 1.5 g Na-glutamate, 21.7 g glucose, 0.1 g lysine). The culture (3 mL) was then added to a 14 mL Falcon culture tube (Becton Dickinson, Franklin Lakes, NJ) and
- 20 subsequently incubated at 30 °C on a shaking incubator for an additional 24 hrs.

- Cells were collected from the entire 3 mL culture by centrifugation and lipids were extracted as described in Bligh, E. G. & Dyer, W. J. (*Can. J. Biochem. Physiol.* 37:911-917 (1959)). Fatty acid methyl esters were
- 25 prepared by transesterification of the lipid extract with sodium methoxide (Roughan, G., and Nishida I. *Arch Biochem Biophys.* 276(1):38-46 (1990)) and subsequently analyzed with a Hewlett-Packard 6890 GC fitted with a 30-m X 0.25 mm (i.d.) HP-INNOWAX (Hewlett-Packard) column. The oven temperature was from 170 °C (25 min hold) to 185 °C at 3.5 °C/min.

- 30 Duplicate GC analyses of one transformant containing pDMW208 showed that the average GLA content was 25.9% of the total lipids, with a standard deviation of 0.42. Duplicate GC analysis of two transformants containing pDMW208D62 showed that the average GLA content was 34% of the total lipids, with a standard deviation of 2.2. There was no GLA
- 35 detected in the wildtype *Yarrowia* strain ATCC #76982. The results are shown in Figure 6 and in Table 4 below. Fatty acids are identified as 16:0,

16:1, 18:0, 18:1 (oleic acid), 18:2 (LA) and GLA; and the composition of each is presented as a % of the total fatty acids.

TABLE 4
GC Analysis Of Fatty Acids In *Yarrowia lipolytica*

Strain	% 16:0	% 16:1	% 18:0	% 18:1	% 18:2	% GLA
Wildtype	14	11	3.5	34.8	31	0
pDMW208	11.9	8.6	1.5	24.4	17.8	25.9
pDMW208D62	16.2	1.5	0.1	17.8	22.2	34.0

These results demonstrate that *Yarrowia lipolytica* strains transformed with two copies each of the $\Delta 6$ desaturase and $\Delta 12$ desaturase genes (i.e., pDMW208D62) produced about 30.8% more GLA than the strains transformed a single copy each of $\Delta 6$ desaturase and $\Delta 12$ desaturase genes (i.e., pDMW208).

EXAMPLE 6 (Prophetic)

Synthesis Of A Codon-Optimized $\Delta 6$ Desaturase Gene

Significant improvements in GLA production could readily be obtained by codon-optimizing the *S. diclina* $\Delta 6$ desaturase (contained within SEQ ID NO:22) to enhance its expression in *Yarrowia*. This would be possible by designing a codon-optimized $\Delta 6$ desaturase gene based on the codon usage and signature of structural genes in *Y. lipolytica* and then synthesizing the gene *in vitro* to enable its increased efficiency in the host (with respect to the wildtype gene), according to the methodology presented in WO2004/ 101753, Example 4.

Specifically, both the preferred codon usage in *Y. lipolytica* and an analysis of the consensus sequence (i.e., SEQ ID NO:41) around the 'ATG' initiation codon of 79 genes in *Y. lipolytica* is presented in WO2004/101753. Design of a codon-optimized $\Delta 6$ desaturase gene should rely on these teachings, to further optimize expression of the gene.

EXAMPLE 7 (Prophetic)

Increased GLA Production Using Fructose-Bisphosphate Aldolase Regulatory Sequences

To further improve the production of GLA in *Yarrowia lipolytica*, fructose-bisphosphate aldolase (FBA) regulatory sequences could be used to drive expression of the $\Delta 12$ and/or $\Delta 6$ desaturase genes, instead of the TEF and GPD promoters used in the chimeric genes described in

Examples 2 and 3. Specifically, WO 2005/049805 describes a *Y. lipolytica* FBA and FBAIN promoter; the FBA promoter (contained within SEQ ID NO:42) was 2.2 times stronger than the GPD promoter in *Yarrowia*, while the FBAIN promoter (SEQ ID NO:43, comprising 826 bp upstream of the *fba1* gene and 169 bp downstream of the putative initiation codon) was about 6.6 times stronger than the GPD promoter (activity was measured according to GUS expression). One of skill in the art of molecular cloning could readily modify plasmid pDMW208 and/or pDMW208D62 to encompass either of these strong promoters, or a portion thereof, as a means to increase GLA expression.

EXAMPLE 8 (Prophetic)

Increased GLA Production Using A Heterologous $\Delta 12$ Desaturase

To further improve the production of GLA in *Yarrowia lipolytica*, the $\Delta 12$ desaturase gene from *Fusarium moniliforme* can be over-expressed, instead of, or in addition to, the endogenous $\Delta 12$ desaturase gene of *Y. lipolytica*. Specifically, previous work has demonstrated that the *F. moniliforme* $\Delta 12$ desaturase was more efficient than the *Y. lipolytica* $\Delta 12$ desaturase in producing 18:2 in *Y. lipolytica* (i.e., 68% product accumulation of LA versus 59% product accumulation of LA, which corresponds to a difference in percent substrate conversion [calculated as $([18:2]/[18:1+18:2])*100$] of 85% versus 74%; WO 2005/047485). In this cited application, expression of the *F. moniliforme* $\Delta 12$ desaturase gene under the control of the TEF promoter in *Y. lipolytica* was compared directly to expression of a chimeric gene encoding the *Y. lipolytica* $\Delta 12$ desaturase under the control of the TEF promoter. On the basis of these results, expression of the fungal *F. moniliforme* $\Delta 12$ desaturase (SEQ ID NOs:44 and 45) is preferred relative to the *Y. lipolytica* $\Delta 12$ desaturase as a means to engineer an oleaginous yeast that accumulates oils enriched in GLA. One of skill in the art of molecular cloning could readily modify plasmid pDMW208 and/or pDMW208D62 such that the $\Delta 12$ desaturase from *F. moniliforme* was expressed, as a means to increase GLA expression.

Furthermore, in some cases, it may prove advantageous to replace the native *Yarrowia lipolytica* $\Delta 12$ desaturase with the *F. moniliforme* $\Delta 12$ desaturase. Although both $\Delta 12$ desaturases catalyze the conversion of oleic acid to LA, the two enzymes differ in their overall specificity; specifically, the Applicants have determined that the *F. moniliforme* $\Delta 12$

desaturase has a higher loading capacity of LA onto the *sn*-2 position of a phosphatidylcholine substrate (thereby facilitating the subsequent reaction by $\Delta 6$ desaturase) than the *Y. lipolytica* $\Delta 12$ desaturase. On this basis, overexpression of the *F. moniliforme* $\Delta 12$ desaturase in conjunction with a knockout of the *Y. lipolytica* $\Delta 12$ desaturase may result in increased product for subsequent conversion to GLA.

EXAMPLE 9 (Prophetic)

Increased GLA Production Using A Heterologous C_{16/18} Elongase Gene

To further improve the production of GLA in *Yarrowia lipolytica*, a gene encoding a C_{16/18} elongase can be used to push the carbon from palmitate (16:0) to stearic acid (18:0), thereby increasing the substrate availability of stearic acid. A $\Delta 9$ desaturase then converts stearic acid directly to oleic acid (18:1), the substrate of the $\Delta 12$ desaturase (Figure 1). This strategy for increasing the amount of C18 fatty acids in the total lipids has been successfully utilized in a co-pending application (U.S. Patent Application No. 60/624812), to enable increased production of EPA. Specifically, the C_{16/18} elongase from *Rattus norvegicus* [GenBank Accession No. AB071986] was codon-optimized for expression in *Y. lipolytica* ("rELO2S"; SEQ ID NOs:46 and 48) and then expressed as a chimeric TEF::codon-optimized C_{16/18} elongase::Pex20 gene.

It would be expected that over-expression of the codon-optimized C_{16/18} elongase in conjunction with both a $\Delta 6$ desaturase and $\Delta 12$ desaturase will result in increased production of GLA in the host. Furthermore, the source of the C_{16/18} elongase is not limiting in the invention herein.

CLAIMS

What is claimed is:

1. A method for producing γ -linolenic acid comprising:
 - a) providing an oleaginous yeast comprising:
 - i) at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity;
 - ii) at least one genetic construct encoding a polypeptide having a $\Delta 12$ desaturase activity; and,
 - iii) a source of desaturase substrate selected from the group consisting of oleic acid and linoleic acid;
 - b) growing the yeast of step (a) in the presence of a fermentable carbon source whereby at least about 15% γ -linolenic acid in the total lipids is produced; and
 - c) optionally recovering the γ -linolenic acid.
2. A method according to Claim 1 wherein said oleaginous yeast optionally comprises additional genes selected from the group consisting of:
 - a) at least one genetic construct encoding $\Delta 9$ desaturase polypeptide;
 - b) at least one genetic construct encoding $C_{14/16}$ elongase polypeptide; and,
 - c) at least one genetic construct encoding $C_{16/18}$ elongase polypeptide
3. A method according to Claim 1 wherein the at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity is in multicopy in the oleaginous yeast.
4. A method according to Claim 1 wherein the at least one genetic construct encoding a polypeptide having a $\Delta 12$ desaturase activity is in multicopy in the oleaginous yeast.

5 A method according to Claim 1 wherein the at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity is codon-optimized for expression in the oleaginous yeast.

6 A method according to Claim 1 wherein the at least one genetic construct encoding a polypeptide having a $\Delta 12$ desaturase activity is codon-optimized for expression in the oleaginous yeast.

7 A method according to Claim 1 wherein the oleaginous yeast is selected from the group consisting of: *Yarrowia*, *Candida*, *Rhodotorula*, *Rhodospiridium*, *Cryptococcus*, *Trichosporon* and *Lipomyces*.

8 A method according to Claim 7 wherein the oleaginous yeast is *Yarrowia lipolytica*.

9 A method according to Claim 8 wherein the at least one genetic construct encoding a polypeptide having a $\Delta 12$ desaturase activity is integrated into at least one genetic loci selected from the group consisting of the *zeta* loci, the *Ura3* loci, the *Leu 2* loci, the *Lys5* loci, the *Aco2* loci, the *Pox3* loci, the *Lip1* loci and the *Lip2* loci.

10. A method according to Claim 8 wherein the at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity is integrated into at least one genetic loci selected from the group consisting of the *zeta* loci, the *Ura3* loci, the *Leu 2* loci, the *Lys5* loci, the *Aco2* loci, the *Pox3* loci, the *Lip1* loci and the *Lip2* loci.

11. A method according to Claim 1 wherein the at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity is under the control of a strong promoter isolated from genes selected from the group consisting of: alcohol dehydrogenase, glyceraldehyde-3-phosphate-dehydrogenase, phosphoglycerate mutase, fructose-bisphosphate aldolase, glycerol-3-phosphate o-acyltransferase, phosphoglucose-isomerase, phosphoglycerate kinase, acid phosphatase, lactase, metallothionein and glucoamylase.

12. A method according to Claim 1 wherein the at least one genetic construct encoding a polypeptide having a $\Delta 12$ desaturase activity is under the control of a strong promoter isolated from genes selected from the group consisting of: alcohol dehydrogenase, glyceraldehyde-3-phosphate-dehydrogenase, phosphoglycerate mutase, fructose-bisphosphate aldolase, glycerol-3-phosphate o-acyltransferase, phosphoglucose-isomerase, phosphoglycerate kinase, acid phosphatase, lactase, metallothionein and glucoamylase.

13. A method for producing γ -linolenic acid comprising:

a) providing a *Yarrowia lipolytica* comprising:

- i) at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity;
- ii) at least one genetic construct encoding a polypeptide having a $\Delta 12$ desaturase activity; and,
- iii) a source of desaturase substrate selected from the group consisting of: oleic acid and linoleic acid;

wherein the at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity and the at least one genetic construct encoding a polypeptide having a $\Delta 12$ desaturase activity are in multicopy and are integrated into a genetic locus selected from the group consisting of the *zeta* loci, the *Ura3* loci, the *Leu 2* loci, the *Lys5* loci, the *Aco2* loci, the *Pox3* loci, the *Lip1* loci and the *Lip2* loci of the *Yarrowia lipolytica*, and

wherein the at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity and the at least one genetic construct encoding a polypeptide having a $\Delta 12$ desaturase activity are under the control of a strong promoter isolated from genes selected from the group consisting of: alcohol dehydrogenase, glyceraldehyde-3-phosphate-dehydrogenase, phosphoglycerate mutase, fructose-bisphosphate aldolase, glycerol-3-phosphate o-acyltransferase, phosphoglucose-isomerase, phosphoglycerate kinase, acid phosphatase, lactase, metallothionein and glucoamylase;

- b) growing the *Yarrowia lipolytica* of step (a) in the presence of a fermentable carbon source whereby at least about 15% γ -linolenic acid in the total lipids is produced; and,
- c) optionally recovering the γ -linolenic acid.

14. A method according to Claim 13 wherein the at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity and the at least one genetic construct encoding a polypeptide having a $\Delta 12$ desaturase activity are codon-optimized for expression in *Yarrowia lipolytica*.

15. A method according to either of Claims 1 or 13 wherein at least about 20% γ -linolenic acid in the total lipids is produced.

16. A method according to either of Claims 1 or 13 wherein at least about 30% γ -linolenic acid in the total lipids is produced.

17. A method according to either of Claims 1 or 13 wherein at least about 40% γ -linolenic acid in the total lipids is produced.

18. A method according to either of Claims 1 or 13 wherein at least about 50% γ -linolenic acid in the total lipids is produced.

19. A method according to either of Claims 1 or 13 wherein the source of desaturase substrate is endogenous to the oleaginous yeast.

20. A method according to either of Claims 1 or 13 wherein the source of desaturase substrate is exogenous to the oleaginous yeast.

21. A method according to either of Claims 1 or 13 wherein the at least one genetic construct encoding a polypeptide having a $\Delta 6$ desaturase activity encodes a polypeptide having the amino acid sequence selected from the group consisting of SEQ ID NOs:9 and 22.

22. A method according to either of Claims 1 or 13 wherein the at least one genetic construct encoding a polypeptide having a $\Delta 12$

desaturase activity encodes a polypeptide having the amino acid sequence selected from the group consisting of SEQ ID NOs:16 and 39.

23. A method according to either of Claims 8 or 13 wherein the *Yarrowia lipolytica* is a strain selected from the group consisting of: ATCC #20362, ATCC #8862, ATCC #18944 and ATCC #76982.

24. A production host cell for the production of γ -linolenic acid comprising:

a) a background *Yarrowia* sp. comprising:

- i) at least one genetic construct encoding $\Delta 12$ desaturase polypeptide; and,
- ii) at least one genetic construct encoding $\Delta 6$ desaturase peptide;

wherein at least one of said genetic constructs of (i) and (ii) is over-expressed and whereby at least about 15% γ -linolenic acid in the total lipids is produced.

25. A production host cell according to Claim 24 wherein:

- a) the $\Delta 6$ desaturase polypeptide has an amino acid sequence selected from the group consisting of SEQ ID NOs:9 and 22; and,
- b) the $\Delta 12$ desaturase polypeptide has an amino acid sequence selected from the group consisting of SEQ ID NOs:16 and 39.

26. A production host cell according to claim 24 optionally comprising a genetic construct selected from the group consisting of:

- a) at least one genetic construct encoding $\Delta 9$ desaturase polypeptide;
- b) at least one genetic construct encoding $C_{14/16}$ elongase polypeptide; and,
- c) at least one genetic construct encoding $C_{16/18}$ elongase polypeptide.

27. A production host according to Claim 26 wherein said $C_{16/18}$ elongase has the amino acid sequence as set forth in SEQ ID NO:48.

28. A production host according to claim 24 wherein the background *Yarrowia* sp. is devoid of any native gene encoding a polypeptide having $\Delta 12$ desaturase activity.

29. A production host according to claim 24 wherein at least one of said genetic constructs are integrated into the *Yarrowia* sp. genome at a gene locus selected from the group consisting of: *zeta*, *Ura3*, *Leu2*, *Lys5*, *Aco2*, *Pox3*, *Lip1*, and the *Lip2* gene locus.

30. A production host according to Claim 24 wherein the background *Yarrowia* sp. is selected from the group consisting of *Yarrowia lipolytica* ATCC #20362, *Yarrowia lipolytica* ATCC #8862, *Yarrowia lipolytica* ATCC #18944, *Yarrowia lipolytica* ATCC #76982 and *Yarrowia lipolytica* LGAM S(7)1.

Figure 1

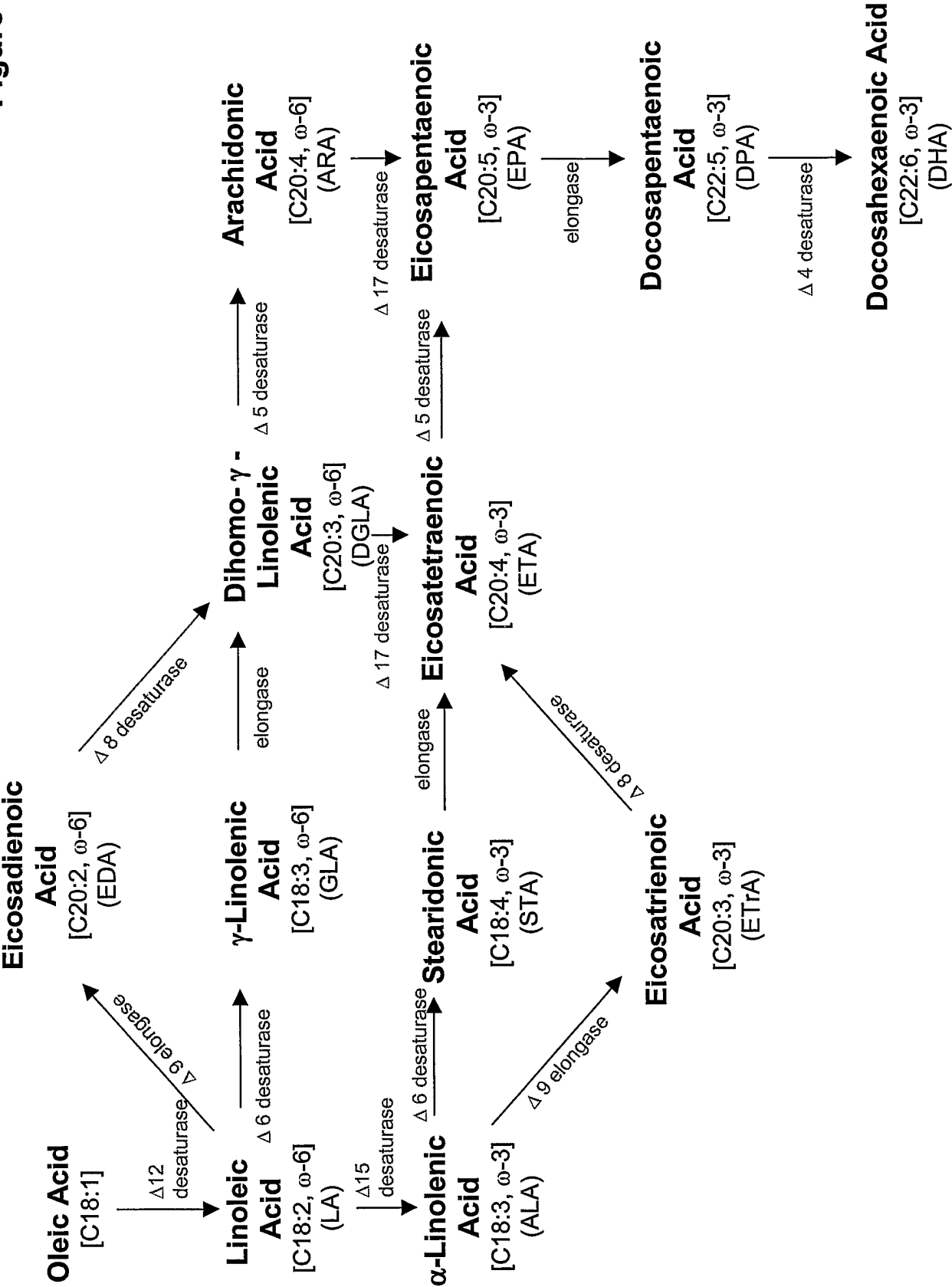


Figure 2

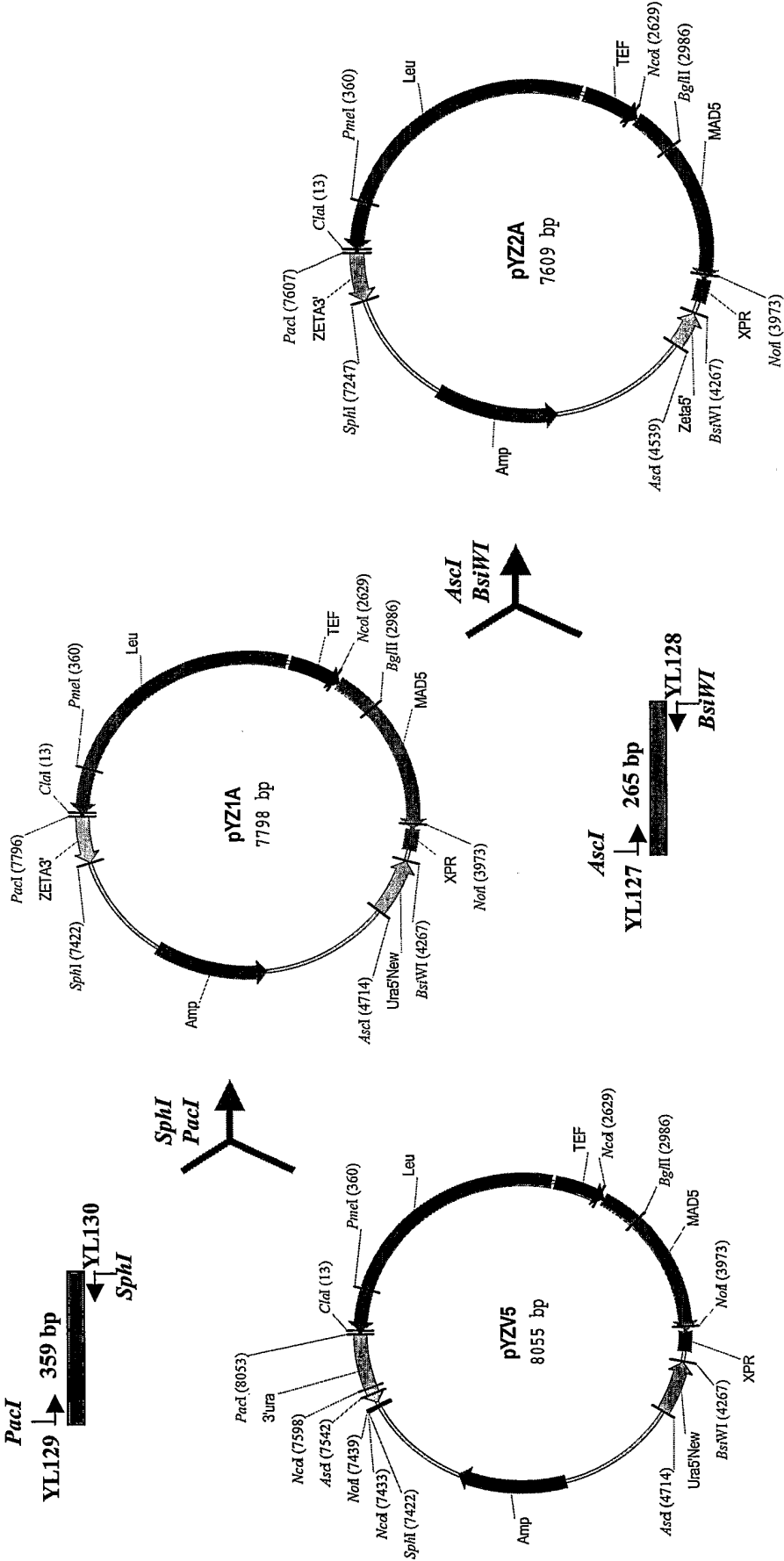


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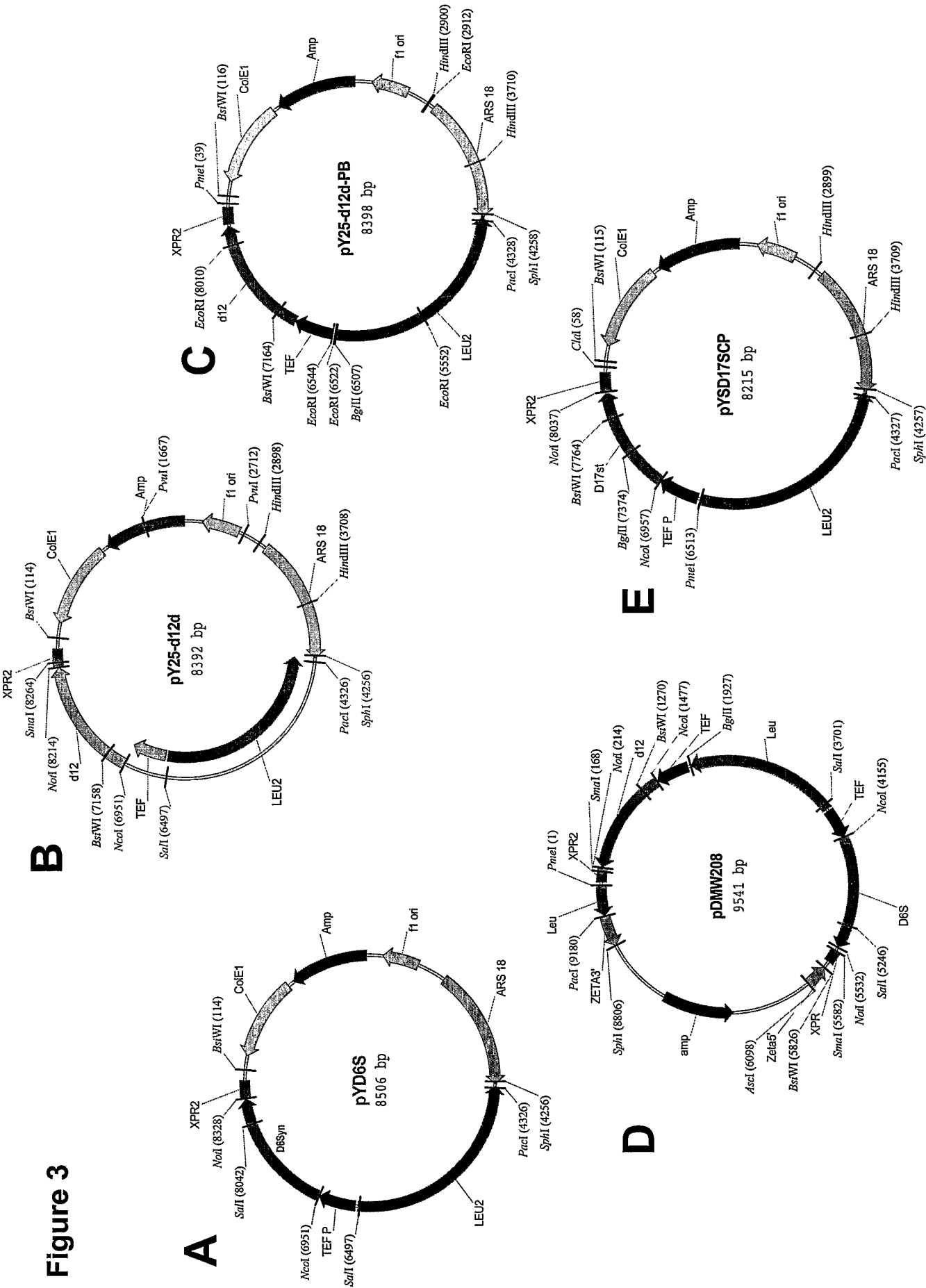


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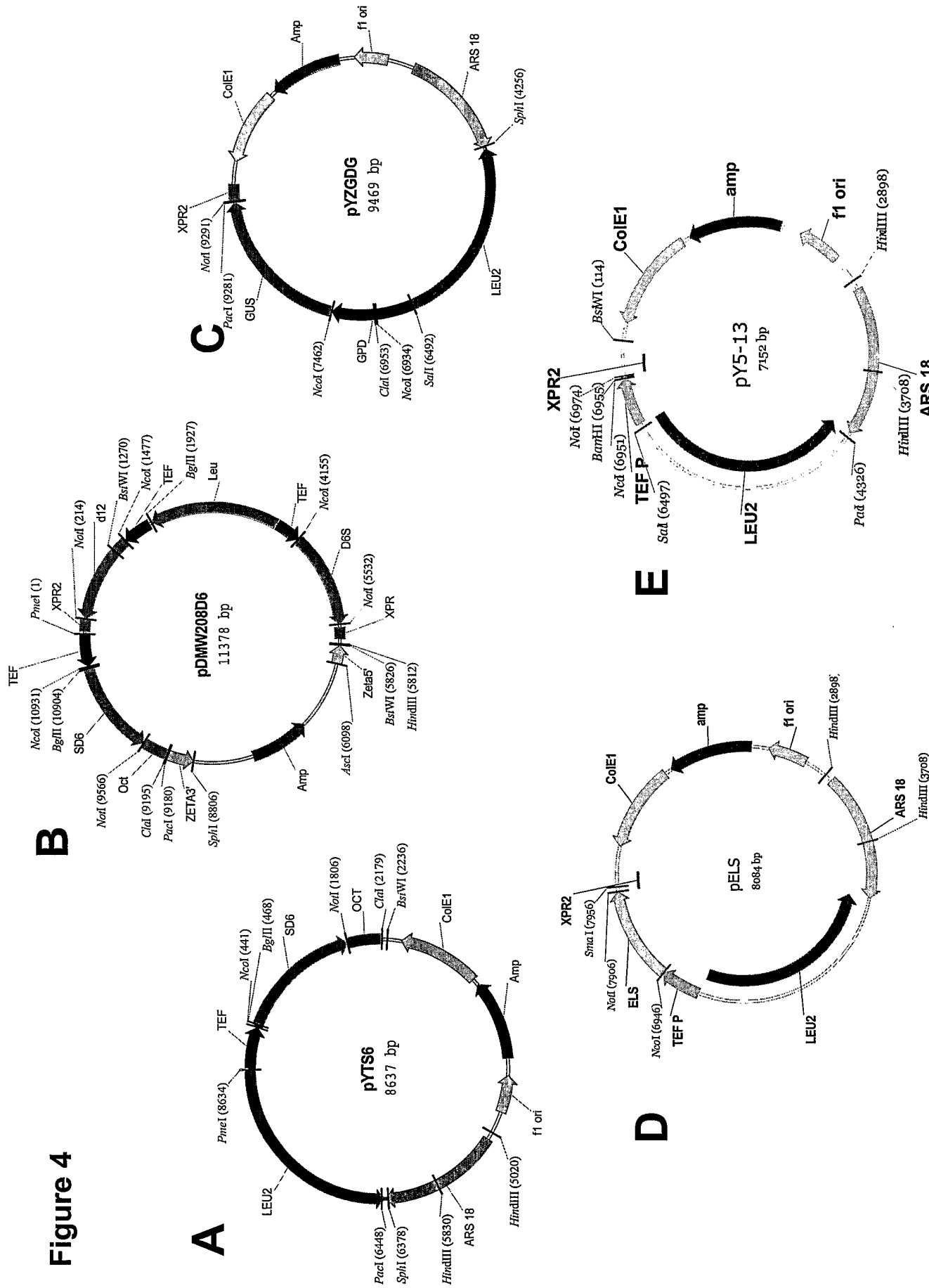
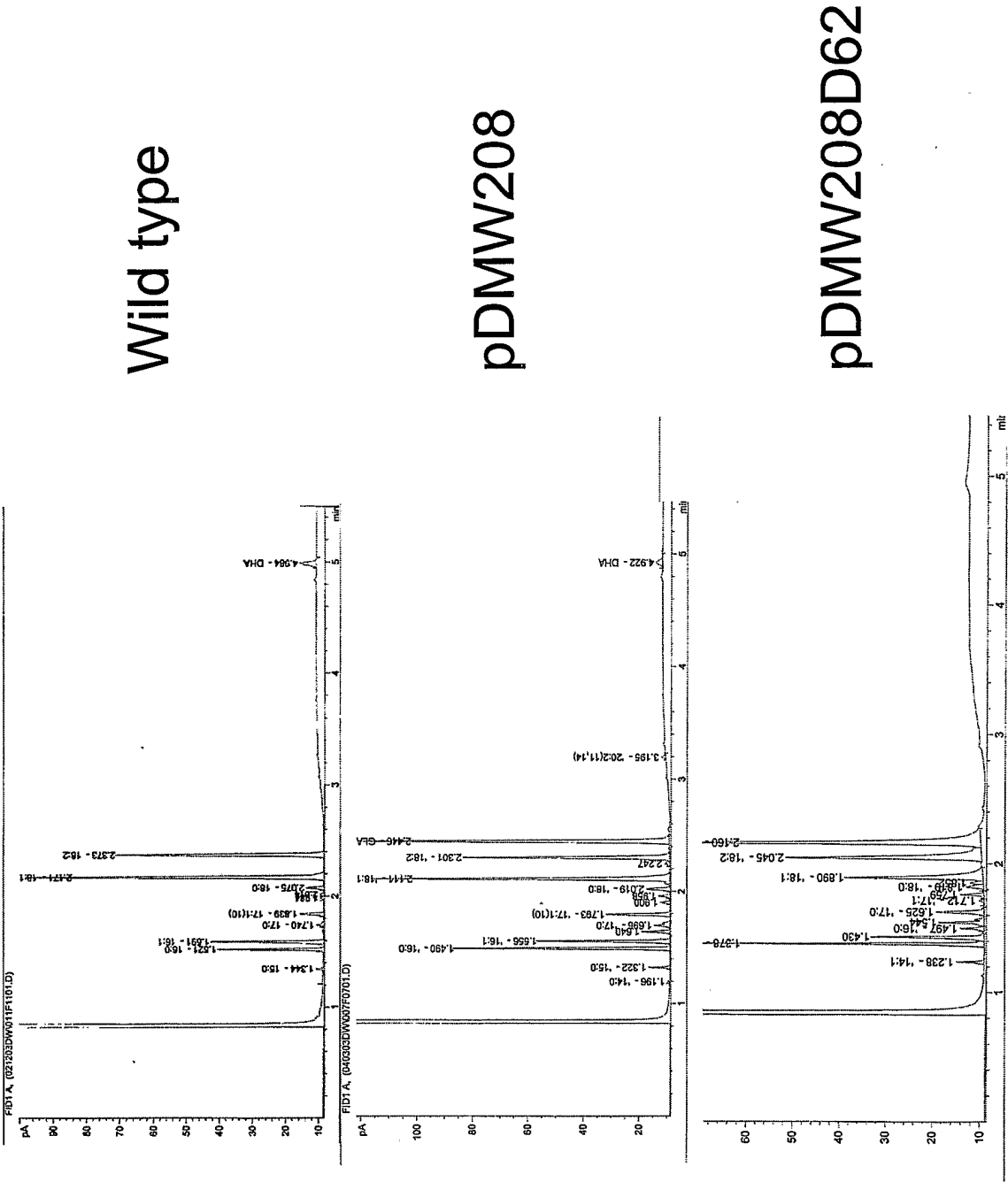




Figure 6



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<210> 45
 <211> 477
 <212> PRT
 <213> *Fusarium moniliforme*

<400> 45

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 35 40 45

Met Ser Glu Val Asp Ile Ala Lys Pro Lys Ser Glu Tyr Gly Val Met
 50 55 60

Leu Asp Thr Tyr Gly Asn Gln Phe Glu Val Pro Asp Phe Thr Ile Lys
 65 70 75 80

Asp Ile Tyr Asn Ala Ile Pro Lys His Cys Phe Lys Arg Ser Ala Leu
 85 90 95

Lys Gly Tyr Gly Tyr Ile Leu Arg Asp Ile Val Leu Leu Thr Thr Thr
 100 105 110

Phe Ser Ile Trp Tyr Asn Phe Val Thr Pro Glu Tyr Ile Pro Ser Thr
 115 120 125

Pro Ala Arg Ala Gly Leu Trp Ala Val Tyr Thr Val Leu Gln Gly Leu
 130 135 140

Phe Gly Thr Gly Leu Trp Val Ile Ala His Glu Cys Gly His Gly Ala
 145 150 155 160

Phe Ser Asp Ser Arg Ile Ile Asn Asp Ile Thr Gly Trp Val Leu His
 165 170 175

Ser Ser Leu Leu Val Pro Tyr Phe Ser Trp Gln Ile Ser His Arg Lys
 180 185 190

His His Lys Ala Thr Gly Asn Met Glu Arg Asp Met Val Phe Val Pro
 195 200 205

Arg Thr Arg Glu Gln Gln Ala Thr Arg Leu Gly Lys Met Thr His Glu
 210 215 220

Leu Ala His Leu Thr Glu Glu Thr Pro Ala Phe Thr Leu Leu Met Leu
 225 230 235 240

Val Leu Gln Gln Leu Val Gly Trp Pro Asn Tyr Leu Ile Thr Asn Val
 245 250 255

Thr Gly His Asn Tyr His Glu Arg Gln Arg Glu Gly Arg Gly Lys Gly
 260 265 270

Lys His Asn Gly Leu Gly Gly Gly Val Asn His Phe Asp Pro Arg Ser
 275 280 285

Pro Leu Tyr Glu Asn Ser Asp Ala Lys Leu Ile Val Leu Ser Asp Ile
 290 295 300

Gly Ile Gly Leu Met Ala Thr Ala Leu Tyr Phe Leu Val Gln Lys Phe
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 <212> DNA
 <213> Rattus norvegicus (GenBank Accession No. AB071986)

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 caa gaa tat gaa ttc gag aag cag ttc aac gag aat gaa gcc atc caa 161
 Gln Glu Tyr Glu Phe Glu Lys Gln Phe Asn Glu Asn Glu Ala Ile Gln
 10 15 20
 tgg atg cag gaa aac tgg aag aaa tct ttc ctg ttt tct gcg ctg tac 209
 Trp Met Gln Glu Asn Trp Lys Lys Ser Phe Leu Phe Ser Ala Leu Tyr
 25 30 35 40
 gct gcc ttt ata ttt ggt ggt cgg cat ctg atg aac aag cgg gcc aag 257
 Ala Ala Phe Ile Phe Gly Gly Arg His Leu Met Asn Lys Arg Ala Lys
 45 50 55
 ttt gaa ctg cgg aag ccg ctc gtg ctc tgg tca ctg act ctt gcg gtc 305
 Phe Glu Leu Arg Lys Pro Leu Val Leu Trp Ser Leu Thr Leu Ala Val
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 ttc agt ata ttc ggt gct ctt cga act ggt gct tac atg ctg tac att 353
 Phe Ser Ile Phe Gly Ala Leu Arg Thr Gly Ala Tyr Met Leu Tyr Ile
 75 80 85
 ttg atg acc aaa ggc ctg aag cag tca gtt tgt gac cag agt ttt tac 401
 Leu Met Thr Lys Gly Leu Lys Gln Ser Val Cys Asp Gln Ser Phe Tyr
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 Asn Gly Pro Val Ser Lys Phe Trp Ala Tyr Ala Phe Val Leu Ser Lys
 105 110 115 120
 gca ccc gaa cta ggt gat acg ata ttc atc att ctg agg aag cag aag 497
 Ala Pro Glu Leu Gly Asp Thr Ile Phe Ile Ile Leu Arg Lys Gln Lys
 125 130 135
 ctg atc ttc ctg cac tgg tac cac cac atc act gtg ctc ctg tac tct 545
 Leu Ile Phe Leu His Trp Tyr His His Ile Thr Val Leu Leu Tyr Ser
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<211> 267

<212> PRT

<213> Rattus norvegicus (GenBank Accession No. AB071986)

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Phe Asn Glu Asn Glu Ala Ile Gln Trp Met Gln Glu Asn Trp Lys Lys
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Ser Phe Leu Phe Ser Ala Leu Tyr Ala Ala Phe Ile Phe Gly Gly Arg
35 40 45

His Leu Met Asn Lys Arg Ala Lys Phe Glu Leu Arg Lys Pro Leu Val
50 55 60

Leu Trp Ser Leu Thr Leu Ala Val Phe Ser Ile Phe Gly Ala Leu Arg
65 70 75 80

Thr Gly Ala Tyr Met Leu Tyr Ile Leu Met Thr Lys Gly Leu Lys Gln
85 90 95

Ser Val Cys Asp Gln Ser Phe Tyr Asn Gly Pro Val Ser Lys Phe Trp
100 105 110

Ala Tyr Ala Phe Val Leu Ser Lys Ala Pro Glu Leu Gly Asp Thr Ile
115 120 125

Phe Ile Ile Leu Arg Lys Gln Lys Leu Ile Phe Leu His Trp Tyr His
130 135 140

His Ile Thr Val Leu Leu Tyr Ser Trp Tyr Ser Tyr Lys Asp Met Val
145 150 155 160

Ala Gly Gly Gly Trp Phe Met Thr Met Asn Tyr Gly Val His Ala Val
165 170 175

Met Tyr Ser Tyr Tyr Ala Leu Arg Ala Ala Gly Phe Arg Val Ser Arg
180 185 190

Lys Phe Ala Met Phe Ile Thr Leu Ser Gln Ile Thr Gln Met Leu Met
195 200 205

Gly Cys Val Ile Asn Tyr Leu Val Phe Asn Trp Met Gln His Asp Asn
210 215 220

Asp Gln Cys Tyr Ser His Phe Gln Asn Ile Phe Trp Ser Ser Leu Met
225 230 235 240

Tyr Leu Ser Tyr Leu Leu Leu Phe Cys His Phe Phe Phe Glu Ala Tyr
245 250 255

Ile Gly Lys Val Lys Lys Ala Thr Lys Ala Glu
260 265