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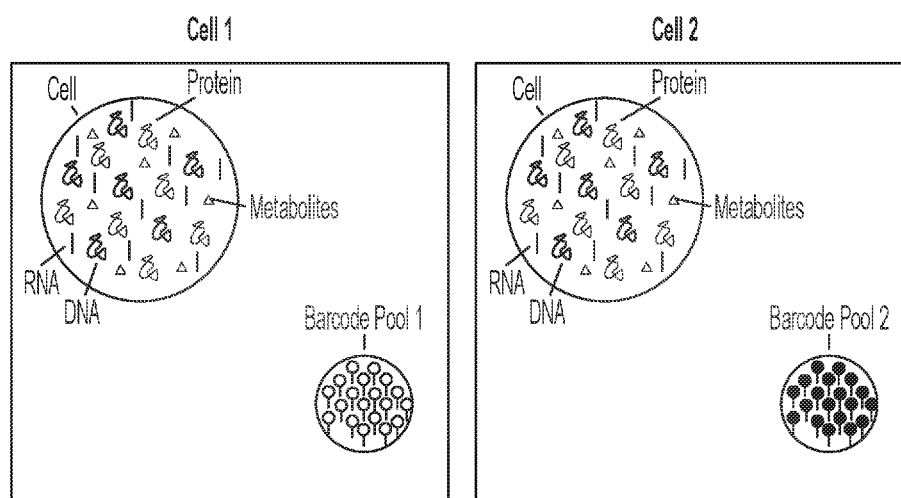


FIG. 1

(57) Abstract: Methods of polypeptide sequencing and/or nucleic acid sequencing which may be carried out on single cell samples and may use barcode components and which facilitate the direct sequencing without amplification. All proteins in the sample may be first labelled with one barcode, secondly the nucleic acids or metabolites are labelled with a second barcode. Also provided herein are compositions, kits and devices useful for the same.

METHODS OF SINGLE-CELL PROTEIN AND NUCLEIC ACID SEQUENCING

RELATED APPLICATIONS

This application claims the benefit under 35 U.S.C. § 119(e) of the filing date of U.S. Provisional Application Serial No. 62/926,991, filed October 28, 2019, and of U.S. Provisional Application Serial No. 62/991,425, filed March 18, 2020, the entire contents of each of which is incorporated herein by reference.

BACKGROUND OF INVENTION

Proteomics has emerged as an important and necessary complement to genomics and transcriptomics in the study of biological systems. However, unlike single-cell genomic and transcriptomic analyses, approaches for single-cell proteomic analysis have been limited to date.

SUMMARY OF INVENTION

Provided herein are methods of single-cell polypeptide sequencing and single-cell nucleic acid sequencing, which facilitate the direct sequencing of a single cell without amplification. Also provided herein are compositions, kits and devices useful for the same.

In some aspects, the disclosure relates to methods comprising directly sequencing, in parallel, the proteome of a single cell and/or sequencing the genome and/or transcriptome of the single cell, and/or optionally detecting one or more metabolite of the single cell. In some embodiments, the method comprises: (i) providing a cell sample comprising the composition of a single cell; (ii) contacting the cell sample with a barcode component to produce a labeled sample comprising barcoded molecules, wherein the barcoded molecules comprise barcoded polypeptides and/or barcoded nucleic acids; and (iii) sequencing the polypeptides and/or nucleic acids of the labeled sample. In some embodiments, the method comprises: (i) providing a cell sample comprising the composition of a single cell; (ii) contacting the cell sample with a barcode component to produce a labeled sample comprising barcoded molecules, wherein the barcoded molecules comprise barcoded polynucleic acids; and (iii) sequencing the polynucleic acids of the labeled sample.

In some embodiments, sequencing the polynucleic acids of the labeled sample comprises long-read sequencing applications, short-read sequencing applications, or hybrid assembly applications. In some embodiments, the barcoded polynucleic acids have a length of about 0.5-2 kb, 0.5-5 kb, 1-2 kb, 1-3 kb, 1-4 kb, 1-5 kb, 1-10 kb, 2-10 kb, 2-5 kb, 5-10 kb, 5-15 kb, 5-20 kb, 5-25 kb, 10-15 kb, 10-20 kb, or 10-25 kb. In some embodiments, the barcoded polynucleic acids have a length of about 700-3000, 1000-3000, 1000-2500, 1000-2400, 1000-2300, 1000-2200,

1000-2100, 1000-2000, 1000-1900, 1000-1800, 1000-1700, 1000-1600, 1000-1500, 1000-1400, 1000-1300, 1000-1200, 1500-3000, 1500-2500, 1500-2000, or 2000-3000 nucleotides in length.

In some embodiments, the composition of (i) comprises a living cell. In some embodiments, the composition of (i) comprises a lysed cell.

5 In some embodiments, the barcoded molecules of the labeled sample each comprise an identical barcode. In some embodiments, the barcoded molecules of the labeled sample further comprise barcoded DNA, barcoded RNA, barcoded cDNA, or barcoded metabolites. In some embodiments, the barcoded molecules of the labeled sample comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the method further comprises amplifying the
10 barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

In some embodiments, (ii) comprises: (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded polypeptides; (b) isolating the barcoded polypeptides from the first sample, thereby generating a second sample comprising the barcoded polypeptides and a third sample comprising the genome, transcriptome, and/or
15 metabolome of the single cell in the cell sample; and (c) contacting the third sample with an additional barcode component, to produce a fourth sample comprising barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites; wherein the labeled sample comprises the second sample and the fourth sample. In some embodiments, the third sample in (b) comprises: (i) a first subsample comprising the genome and transcriptome of the single cell and a second
20 subsample comprising the metabolome of the single cell; (ii) a first subsample comprising the genome and metabolome of the single cell and a second subsample comprising the transcriptome of the single cell; (iii) a first subsample comprising the metabolome and transcriptome of the single cell and a second subsample comprising the genome of the single cell; or (iv) a first
25 subsample comprising the genome of the single cell, a second subsample comprising the transcriptome of the single cell, and a third subsample comprising the metabolome of the single cell. In some embodiments, the additional barcode component in (c) comprises a second, third, fourth, or fifth barcode component. In some embodiments, the barcoded molecules of the fourth sample in (c) comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded
30 cDNA.

In some embodiments, (ii) comprises: (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA; (b) amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA of the first sample; and (c) contacting the first sample with a second barcode component, to

produce a second sample comprising barcoded polypeptides and/or barcoded metabolites; wherein the labeled sample comprises the second sample.

In some embodiments, (ii) comprises: (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA; (b) isolating the barcoded DNA, barcoded RNA, and/or barcoded cDNA from the first sample, thereby generating a second sample comprising the barcoded DNA and/or barcoded cDNA and a third sample comprising the proteome and/or metabolome of the single cell in the cell sample; and (c) contacting the third sample with an additional barcode component, to produce a fourth sample comprising barcoded polypeptides and/or the barcoded metabolites; wherein the labeled sample comprises the second sample and the fourth sample. In some embodiments, the third sample in (b) comprises a first subsample comprising the proteome of the single cell and a second subsample comprising the metabolome of the single cell. In some embodiments, the additional barcode component in (c) comprises a second or third barcode component. In some embodiments, the barcoded molecules of the first sample in (a) comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

In some embodiments, the method further comprises sequencing the barcoded polypeptides, the barcoded DNA, barcoded RNA, and/or barcoded cDNA of the labeled sample. In some embodiments, the sequencing comprises: (a) detecting the barcode identities of the barcoded molecules of the labeled sample, thereby determining the origins of the barcoded molecules; and (b) sequencing, in parallel, the barcoded polypeptides in the labeled sample, thereby determining at least the partial amino acid sequences of the barcoded polypeptides; wherein (a) occurs before, after, or concurrently with (b).

In some embodiments, the method further comprises detecting and optionally quantifying one or more of the barcoded metabolites of the labeled sample.

In some embodiments, the barcoded DNA, barcoded RNA, and/or barcoded cDNA have a length of about 0.5-2 kb, 0.5-5 kb, 1-2 kb, 1-3 kb, 1-4 kb, 1-5 kb, 1-10 kb, 2-10 kb, 2-5 kb, 5-10 kb, 5-15 kb, 5-20 kb, 5-25 kb, 10-15 kb, 10-20 kb, or 10-25 kb. In some embodiments, the barcoded DNA, barcoded RNA, and/or barcoded cDNA have a length of about 700-3000, 1000-3000, 1000-2500, 1000-2400, 1000-2300, 1000-2200, 1000-2100, 1000-2000, 1000-1900, 1000-1800, 1000-1700, 1000-1600, 1000-1500, 1000-1400, 1000-1300, 1000-1200, 1500-3000, 1500-2500, 1500-2000, or 2000-3000 nucleotides in length.

In some embodiments, the method further comprises combining the labeled sample with at least one supplemental sample comprising barcoded molecules, wherein the barcoded molecules of each sample are distinguishable, thereby producing a multiplexed sample. In some

embodiments, at least one supplemental sample is prepared by a method comprising: (a) providing a cell sample comprising the composition of a single cell; and (b) contacting the cell sample with a barcode component to produce a labeled sample comprising barcoded molecules. In some embodiments, the composition of (a) comprises a living cell. In some embodiments, the composition of (a) comprises a lysed cell. In some embodiments, the barcoded molecules of (b) each comprise an identical barcode. In some embodiments, the barcoded molecules of (b) comprise barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites.

In some embodiments, the method further comprises detecting, and optionally quantifying, the barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites of the multiplexed sample.

In some embodiments, sequencing the barcoded DNA, barcoded RNA, and/or barcoded cDNA comprises long-read sequencing applications, short-read sequencing applications, or hybrid assembly applications. In some embodiments, the barcoded DNA, barcoded RNA, and/or barcoded cDNA have a length of about 0.5-2 kb, 0.5-5 kb, 1-2 kb, 1-3 kb, 1-4 kb, 1-5 kb, 1-10 kb, 2-10 kb, 2-5 kb, 5-10 kb, 5-15 kb, 5-20 kb, 5-25 kb, 10-15 kb, 10-20 kb, or 10-25 kb. In some embodiments, the barcoded DNA, barcoded RNA, and/or barcoded cDNA have a length of about 700-3000, 1000-3000, 1000-2500, 1000-2400, 1000-2300, 1000-2200, 1000-2100, 1000-2000, 1000-1900, 1000-1800, 1000-1700, 1000-1600, 1000-1500, 1000-1400, 1000-1300, 1000-1200, 1500-3000, 1500-2500, 1500-2000, or 2000-3000 nucleotides in length.

In some embodiments, the method further comprises sequencing the barcoded polypeptides, barcoded DNA, barcoded RNA, and/or barcoded cDNA of the multiplexed sample. In some embodiments, the sequencing comprises: (a) detecting the barcode identities of the barcoded molecules of the multiplexed sample, thereby determining the origins of the barcoded molecules; and (b) sequencing, in parallel, the barcoded polypeptides in the multiplexed sample, thereby determining at least the partial amino acid sequences of the barcoded polypeptides; wherein (a) occurs before, after, or concurrently with (b). In some embodiments, the barcode identities are detected in (a) by DNA sequencing, protein sequencing, hybridization, luminescence, binding kinetics, and/or physical location on or within a solid substrate.

In some embodiments, the sequencing comprises: (a) contacting a single polypeptide molecule with one or more terminal amino acid recognition molecules; and (b) detecting a series of signal pulses indicative of association of the one or more terminal amino acid recognition molecules with successive amino acids exposed at a terminus of the single polypeptide while the single polypeptide is being degraded, thereby sequencing the single polypeptide molecule.

In some embodiments, the sequencing comprises: (a) contacting a single polypeptide molecule with a composition comprising one or more terminal amino acid recognition molecules and a cleaving reagent; and (b) detecting a series of signal pulses indicative of association of the one or more terminal amino acid recognition molecules with a terminus of the single polypeptide molecule in the presence of the cleaving reagent, wherein the series of signal pulses is indicative of a series of amino acids exposed at the terminus over time as a result of terminal amino acid cleavage by the cleaving reagent.

In some embodiments, the sequencing comprises: (a) identifying a first amino acid at a terminus of a single polypeptide molecule; (b) removing the first amino acid to expose a second amino acid at the terminus of the single polypeptide molecule, and (c) identifying the second amino acid at the terminus of the single polypeptide molecule, wherein (a)-(c) are performed in a single reaction mixture.

In some embodiments, the sequencing comprises: (a) contacting a single polypeptide molecule with one or more amino acid recognition molecules that bind to the single polypeptide molecule; (b) detecting a series of signal pulses indicative of association of the one or more amino acid recognition molecules with the single polypeptide molecule under polypeptide degradation conditions; and (c) identifying a first type of amino acid in the single polypeptide molecule based on a first characteristic pattern in the series of signal pulses.

In some embodiments, the sequencing comprises: (a) obtaining data during a polypeptide degradation process; (b) analyzing the data to determine portions of the data corresponding to amino acids that are sequentially exposed at a terminus of the polypeptide during the degradation process; and (c) outputting an amino acid sequence representative of the polypeptide.

In some embodiments, the sequencing comprises: (a) contacting a polypeptide with one or more labeled affinity reagents that selectively bind one or more types of terminal amino acids at a terminus of the polypeptide; and (b) identifying a terminal amino acid at the terminus of the polypeptide by detecting an interaction of the polypeptide with the one or more labeled affinity reagents.

In some embodiments, the sequencing comprises: (a) contacting a polypeptide with one or more labeled affinity reagents that selectively bind one or more types of terminal amino acids at a terminus of the polypeptide; (b) identifying a terminal amino acid at the terminus of the polypeptide by detecting an interaction of the polypeptide with the one or more labeled affinity reagents; (c) removing the terminal amino acid; and (d) repeating (a)-(c) one or more times at the terminus of the polypeptide to determine an amino acid sequence of the polypeptide. In some embodiments, the method further comprises: after (a) and before (b), removing any of the one or more labeled affinity reagents that do not selectively bind the terminal amino acid; and/or after

(b) and before (c), removing any of the one or more labeled affinity reagents that selectively bind the terminal amino acid. In some embodiments, (c) comprises modifying the terminal amino acid by contacting the terminal amino acid with an isothiocyanate, and: contacting the modified terminal amino acid with a protease that specifically binds and removes the modified terminal amino acid; or subjecting the modified terminal amino acid to acidic or basic conditions sufficient to remove the modified terminal amino acid. In some embodiments, identifying the terminal amino acid comprises: identifying the terminal amino acid as being one type of the one or more types of terminal amino acids to which the one or more labeled affinity reagents bind; or identifying the terminal amino acid as being a type other than the one or more types of terminal amino acids to which the one or more labeled affinity reagents bind.

In some embodiments, the one or more labeled affinity reagents comprise one or more labeled aptamers, one or more labeled peptidases, one or more labeled antibodies, one or more labeled degradation pathway protein, one or more aminotransferase, one or more tRNA synthetase, or a combination thereof. In some embodiments, the one or more labeled peptidases have been modified to inactivate cleavage activity; or wherein the one or more labeled peptidases retain cleavage activity for the removing of (c).

In some aspects, the disclosure relates to methods comprising: (i) providing a cell sample; (ii) contacting the cell sample with a barcode component to produce a labeled sample comprising barcoded molecules, wherein the barcoded molecules comprise barcoded polypeptides, barcoded DNA, barcoded RNA, and/or barcoded cDNA; and (iii) sequencing the barcoded polypeptides, barcoded DNA, barcoded RNA, and/or barcoded cDNA of the labeled sample; wherein the barcoded molecules of the labeled sample are not amplified prior to sequencing; optionally wherein the barcoded molecules in (ii) further comprise barcoded metabolites and optionally wherein the method further comprises detecting one or more of the barcoded metabolites.

In some embodiments, the cell sample comprises the composition of a single cell. In some embodiments, the composition of (i) comprises a living cell. In some embodiments, the composition of (i) comprises a lysed cell.

In some embodiments, the barcoded molecules of the labeled sample each comprise an identical barcode. In some embodiments, the barcoded molecules of the labeled sample comprise barcoded DNA, barcoded RNA, barcoded cDNA, and barcoded metabolites. In some embodiments, the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

In some embodiments, (ii) comprises: (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded polypeptides; (b) isolating the barcoded polypeptides from the first sample, thereby generating a second sample comprising the

barcoded polypeptides and a third sample comprising the genome, transcriptome, and/or metabolome of the single cell in the cell sample; and (c) contacting the third sample with an additional barcode component, to produce a fourth sample comprising barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites; wherein the labeled sample comprises the second sample and the fourth sample. In some embodiments, the third sample in (b) comprises: (i) a first subsample comprising the genome and transcriptome of the single cell and a second subsample comprising the metabolome of the single cell; (ii) a first subsample comprising the genome and metabolome of the single cell and a second subsample comprising the transcriptome of the single cell; (iii) a first subsample comprising the metabolome and transcriptome of the single cell and a second subsample comprising the genome of the single cell; or (iv) a first subsample comprising the genome of the single cell, a second subsample comprising the transcriptome of the single cell, and a third subsample comprising the metabolome of the single cell. In some embodiments, the additional barcode component in (c) comprises a second, third, fourth, or fifth barcode component. In some embodiments, the barcoded molecules of the fourth sample in (c) comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

In some embodiments, (ii) comprises: (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA; (b) amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA of the first sample; and (c) contacting the first sample with a second barcode component, to produce a second sample comprising barcoded polypeptides and/or barcoded metabolites; wherein the labeled sample comprises the second sample.

In some embodiments, (ii) comprises: (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA; (b) isolating the barcoded DNA, barcoded RNA, and/or barcoded cDNA from the first sample, thereby generating a second sample comprising the barcoded DNA and/or barcoded cDNA and a third sample comprising the proteome and/or metabolome of the single cell in the cell sample; and (c) contacting the third sample with an additional barcode component, to produce a fourth sample comprising barcoded polypeptides and/or the barcoded metabolites; wherein the labeled sample comprises the second sample and the fourth sample. In some embodiments, the third sample in (b) comprises a first subsample comprising the proteome of the single cell and a second subsample comprising the metabolome of the single cell. In some embodiments, the additional barcode component in (c) comprises a second or third barcode component. In some embodiments, the barcoded molecules of the first sample in (a) comprise

barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

In some embodiments, the method further comprises sequencing the barcoded polypeptides, barcoded DNA, barcoded RNA, and/or barcoded cDNA of the labeled sample. In some embodiments, the sequencing comprises: (a) detecting the barcode identities of the barcoded molecules of the labeled sample, thereby determining the origins of the barcoded molecules; and (b) sequencing, in parallel, the barcoded polypeptides in the labeled sample, thereby determining at least the partial amino acid sequences of the barcoded polypeptides; wherein (a) occurs before, after, or concurrently with (b).

In some embodiments, the method further comprises detecting, and optionally quantifying, the barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites of the labeled sample.

In some embodiments, sequencing the barcoded DNA, barcoded RNA, and/or barcoded cDNA comprises long-read sequencing applications, short-read sequencing applications, or hybrid assembly applications. In some embodiments, the barcoded DNA, barcoded RNA, and/or barcoded cDNA have a length of about 0.5-2 kb, 0.5-5 kb, 1-2 kb, 1-3 kb, 1-4 kb, 1-5 kb, 1-10 kb, 2-10 kb, 2-5 kb, 5-10 kb, 5-15 kb, 5-20 kb, 5-25 kb, 10-15 kb, 10-20 kb, or 10-25 kb. In some embodiments, the barcoded DNA, barcoded RNA, and/or barcoded cDNA have a length of about 700-3000, 1000-3000, 1000-2500, 1000-2400, 1000-2300, 1000-2200, 1000-2100, 1000-2000, 1000-1900, 1000-1800, 1000-1700, 1000-1600, 1000-1500, 1000-1400, 1000-1300, 1000-1200, 1500-3000, 1500-2500, 1500-2000, or 2000-3000 nucleotides in length.

In some embodiments, the method further comprises combining the labeled sample with at least one supplemental sample comprising barcoded molecules, wherein the barcoded molecules of each sample are distinguishable, thereby producing a multiplexed sample. In some embodiments, at least one supplemental sample is prepared by a method comprising: (a) providing a cell sample comprising the composition of a single cell; and (b) contacting the cell sample with a barcode component to produce a labeled sample comprising barcoded molecules. In some embodiments, the composition of (a) comprises a living cell. In some embodiments, the composition of (a) comprises a lysed cell.

In some embodiments, the barcoded molecules of (b) each comprise an identical barcode. In some embodiments, the barcoded molecules of (b) comprise barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites.

In some embodiments, the method further comprises detecting, and optionally quantifying, the barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites of the multiplexed sample.

In some embodiments, the method further comprises sequencing the barcoded polypeptides, barcoded DNA, barcoded RNA, and/or barcoded cDNA of the multiplexed sample. In some embodiments, the sequencing comprises: (a) detecting the barcode identities of the barcoded molecules of the multiplexed sample, thereby determining the origins of the barcoded molecules; and (b) sequencing, in parallel, the barcoded polypeptides in the multiplexed sample, thereby determining at least the partial amino acid sequences of the barcoded polypeptides; wherein (a) occurs before, after, or concurrently with (b). In some embodiments, the barcode identities are detected in (a) by DNA sequencing, protein sequencing, hybridization, luminescence, binding kinetics, and/or physical location on or within a solid substrate.

In some embodiments, the sequencing comprises: (a) contacting a single polypeptide molecule with one or more terminal amino acid recognition molecules; and (b) detecting a series of signal pulses indicative of association of the one or more terminal amino acid recognition molecules with successive amino acids exposed at a terminus of the single polypeptide while the single polypeptide is being degraded, thereby sequencing the single polypeptide molecule.

In some embodiments, the sequencing comprises: (a) contacting a single polypeptide molecule with a composition comprising one or more terminal amino acid recognition molecules and a cleaving reagent; and (b) detecting a series of signal pulses indicative of association of the one or more terminal amino acid recognition molecules with a terminus of the single polypeptide molecule in the presence of the cleaving reagent, wherein the series of signal pulses is indicative of a series of amino acids exposed at the terminus over time as a result of terminal amino acid cleavage by the cleaving reagent.

In some embodiments, the sequencing comprises: (a) identifying a first amino acid at a terminus of a single polypeptide molecule; (b) removing the first amino acid to expose a second amino acid at the terminus of the single polypeptide molecule, and (c) identifying the second amino acid at the terminus of the single polypeptide molecule, wherein (a)-(c) are performed in a single reaction mixture.

In some embodiments, the sequencing comprises: (a) contacting a single polypeptide molecule with one or more amino acid recognition molecules that bind to the single polypeptide molecule; (b) detecting a series of signal pulses indicative of association of the one or more amino acid recognition molecules with the single polypeptide molecule under polypeptide degradation conditions; and (c) identifying a first type of amino acid in the single polypeptide molecule based on a first characteristic pattern in the series of signal pulses.

In some embodiments, the sequencing comprises: (a) obtaining data during a polypeptide degradation process; (b) analyzing the data to determine portions of the data corresponding to

amino acids that are sequentially exposed at a terminus of the polypeptide during the degradation process; and (c) outputting an amino acid sequence representative of the polypeptide.

In some embodiments, the sequencing comprises: (a) contacting a polypeptide with one or more labeled affinity reagents that selectively bind one or more types of terminal amino acids at a terminus of the polypeptide; and (b) identifying a terminal amino acid at the terminus of the polypeptide by detecting an interaction of the polypeptide with the one or more labeled affinity reagents.

In some embodiments, the sequencing comprises: (a) contacting a polypeptide with one or more labeled affinity reagents that selectively bind one or more types of terminal amino acids at a terminus of the polypeptide; (b) identifying a terminal amino acid at the terminus of the polypeptide by detecting an interaction of the polypeptide with the one or more labeled affinity reagents; (c) removing the terminal amino acid; and (d) repeating (a)-(c) one or more times at the terminus of the polypeptide to determine an amino acid sequence of the polypeptide. In some embodiments, the method further comprises: after (a) and before (b), removing any of the one or more labeled affinity reagents that do not selectively bind the terminal amino acid; and/or after (b) and before (c), removing any of the one or more labeled affinity reagents that selectively bind the terminal amino acid. In some embodiments, (c) comprises modifying the terminal amino acid by contacting the terminal amino acid with an isothiocyanate, and: contacting the modified terminal amino acid with a protease that specifically binds and removes the modified terminal amino acid; or subjecting the modified terminal amino acid to acidic or basic conditions sufficient to remove the modified terminal amino acid. In some embodiments, identifying the terminal amino acid comprises: identifying the terminal amino acid as being one type of the one or more types of terminal amino acids to which the one or more labeled affinity reagents bind; or identifying the terminal amino acid as being a type other than the one or more types of terminal amino acids to which the one or more labeled affinity reagents bind.

In some embodiments, the one or more labeled affinity reagents comprise one or more labeled aptamers, one or more labeled peptidases, one or more labeled antibodies, one or more labeled degradation pathway protein, one or more aminotransferase, one or more tRNA synthetase, or a combination thereof. In some embodiments, the one or more labeled peptidases have been modified to inactivate cleavage activity; or wherein the one or more labeled peptidases retain cleavage activity for the removing of (c).

In some aspects, the disclosure relates to methods comprising: (i) providing a cell sample; (ii) contacting the cell sample with a barcode component to produce a labeled sample comprising barcoded molecules, wherein the barcoded molecules comprise barcoded polypeptides and barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites.

In some embodiments, (i) comprises: (a) providing a cell population; and (b) lysing the cell population. In some embodiments, the cell population: consists of a single cell; comprises a plurality of homogeneous cells; or comprises a plurality of heterogeneous cells. In some embodiments, the cell population is isolated from a subject. In some embodiments, the subject is a human, mouse, rat, or non-human primate subject.

In some embodiments, the barcoded molecules of (ii) each comprise an identical barcode.

In some embodiments, (ii) comprises: (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded polypeptides; (b) isolating the barcoded polypeptides from the first sample, thereby generating a second sample comprising the barcoded polypeptides and a third sample comprising the genome, transcriptome, and/or metabolome of the single cell in the cell sample; and (c) contacting the third sample with an additional barcode component, to produce a fourth sample comprising barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites; wherein the labeled sample comprises the second sample and the fourth sample. In some embodiments, the third sample in (b) comprises: (i) a first subsample comprising the genome and transcriptome of the single cell and a second subsample comprising the metabolome of the single cell; (ii) a first subsample comprising the genome and metabolome of the single cell and a second subsample comprising the transcriptome of the single cell; (iii) a first subsample comprising the metabolome and transcriptome of the single cell and a second subsample comprising the genome of the single cell; or (iv) a first subsample comprising the genome of the single cell, a second subsample comprising the transcriptome of the single cell, and a third subsample comprising the metabolome of the single cell. In some embodiments, the additional barcode component in (c) comprises a second, third, fourth, or fifth barcode component. In some embodiments, the barcoded molecules of the fourth sample in (c) comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

In some embodiments, (ii) comprises: (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA; (b) amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA of the first sample; and (c) contacting the first sample with a second barcode component, to produce a second sample comprising barcoded polypeptides and/or barcoded metabolites; wherein the labeled sample comprises the second sample.

In some embodiments, (ii) comprises: (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA; (b) isolating the barcoded DNA, barcoded RNA, and/or barcoded cDNA from

the first sample, thereby generating a second sample comprising the barcoded DNA and/or barcoded cDNA and a third sample comprising the proteome and/or metabolome of the single cell in the cell sample; and (c) contacting the third sample with an additional barcode component, to produce a fourth sample comprising barcoded polypeptides and/or the barcoded metabolites;

5 wherein the labeled sample comprises the second sample and the fourth sample. In some embodiments, the third sample in (b) comprises a first subsample comprising the proteome of the single cell and a second subsample comprising the metabolome of the single cell. In some embodiments, the additional barcode component in (c) comprises a second or third barcode component. In some embodiments, the barcoded molecules of the first sample in (a) comprise
10 barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

In some embodiments, the method further comprises detecting, and optionally quantifying, the barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites of the multiplexed sample.

15 In some embodiments, the method further comprises sequencing the barcoded polypeptides, the barcoded DNA, barcoded RNA, and/or barcoded cDNA of the labeled sample. In some embodiments, the sequencing comprises: (a) detecting the barcode identities of the barcoded molecules of the labeled sample, thereby determining the origins of the barcoded molecules; and (b) sequencing, in parallel, the barcoded polypeptides in the labeled sample,
20 thereby determining at least the partial amino acid sequences of the barcoded polypeptides; wherein (a) occurs before, after, or concurrently with (b).

In some embodiments, the method further comprises combining the labeled sample with at least one supplemental sample comprising barcoded molecules, wherein the barcoded molecules of each sample are distinguishable, thereby producing a multiplexed sample.

25 In some embodiments, at least one supplemental sample is prepared by a method comprising: (a) providing a cell sample; (b) contacting the cell sample with a barcode component to produce a labeled sample comprising barcoded molecules, wherein the barcoded molecules comprise barcoded polypeptides and barcoded DNA, barcoded cDNA, and/or barcoded metabolites.

In some embodiments, (a) comprises: i. providing a cell population; and ii. lysing the cell
30 population. In some embodiments, the cell population: consists of a single cell; comprises a plurality of homologous cells; or comprises a plurality of heterologous cells. In some embodiments, the cell population is isolated from a subject. In some embodiments, the subject is a human, mouse, rat, or non-human primate subject.

In some embodiments, the barcoded molecules of (b) each comprise an identical barcode. In some embodiments, the barcoded molecules of (b) comprise barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites.

In some embodiments, the method further comprises detecting, and optionally
5 quantifying, the barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites of the multiplexed sample.

In some embodiments, the method further comprises sequencing the barcoded polypeptides, the barcoded DNA, barcoded RNA, and/or barcoded cDNA of the multiplexed sample. In some embodiments, the sequencing comprises: (a) detecting the barcode identities of
10 the barcoded molecules of the multiplexed sample, thereby determining the origins of the barcoded molecules; and (b) sequencing, in parallel, the barcoded polypeptides in the multiplexed sample, thereby determining at least the partial amino acid sequences of the barcoded polypeptides; wherein (a) occurs before, after, or concurrently with (b). In some embodiments, the barcode identities are detected in (a) by DNA sequencing, protein sequencing,
15 hybridization, luminescence, binding kinetics, and/or physical location on or within a solid substrate.

In some embodiments, the sequencing comprises: (a) contacting a single polypeptide molecule with one or more terminal amino acid recognition molecules; and (b) detecting a series of signal pulses indicative of association of the one or more terminal amino acid recognition
20 molecules with successive amino acids exposed at a terminus of the single polypeptide while the single polypeptide is being degraded, thereby sequencing the single polypeptide molecule.

In some embodiments, the sequencing comprises: (a) contacting a single polypeptide molecule with a composition comprising one or more terminal amino acid recognition molecules and a cleaving reagent; and (b) detecting a series of signal pulses indicative of association of the
25 one or more terminal amino acid recognition molecules with a terminus of the single polypeptide molecule in the presence of the cleaving reagent, wherein the series of signal pulses is indicative of a series of amino acids exposed at the terminus over time as a result of terminal amino acid cleavage by the cleaving reagent.

In some embodiments, the sequencing comprises: (a) identifying a first amino acid at a
30 terminus of a single polypeptide molecule; (b) removing the first amino acid to expose a second amino acid at the terminus of the single polypeptide molecule, and (c) identifying the second amino acid at the terminus of the single polypeptide molecule, wherein (a)-(c) are performed in a single reaction mixture.

In some embodiments, the sequencing comprises: (a) contacting a single polypeptide
35 molecule with one or more amino acid recognition molecules that bind to the single polypeptide

molecule; (b) detecting a series of signal pulses indicative of association of the one or more amino acid recognition molecules with the single polypeptide molecule under polypeptide degradation conditions; and (c) identifying a first type of amino acid in the single polypeptide molecule based on a first characteristic pattern in the series of signal pulses.

5 In some embodiments, the sequencing comprises: (a) obtaining data during a polypeptide degradation process; (b) analyzing the data to determine portions of the data corresponding to amino acids that are sequentially exposed at a terminus of the polypeptide during the degradation process; and (c) outputting an amino acid sequence representative of the polypeptide.

10 In some embodiments, the sequencing comprises: (a) contacting a polypeptide with one or more labeled affinity reagents that selectively bind one or more types of terminal amino acids at a terminus of the polypeptide; and (b) identifying a terminal amino acid at the terminus of the polypeptide by detecting an interaction of the polypeptide with the one or more labeled affinity reagents.

15 In some embodiments, the sequencing comprises: (a) contacting a polypeptide with one or more labeled affinity reagents that selectively bind one or more types of terminal amino acids at a terminus of the polypeptide; (b) identifying a terminal amino acid at the terminus of the polypeptide by detecting an interaction of the polypeptide with the one or more labeled affinity reagents; (c) removing the terminal amino acid; and (d) repeating (a)-(c) one or more times at the terminus of the polypeptide to determine an amino acid sequence of the polypeptide. In some
20 embodiments, the method further comprises: after (a) and before (b), removing any of the one or more labeled affinity reagents that do not selectively bind the terminal amino acid; and/or after (b) and before (c), removing any of the one or more labeled affinity reagents that selectively bind the terminal amino acid. In some embodiments, (c) comprises modifying the terminal amino acid by contacting the terminal amino acid with an isothiocyanate, and: contacting the modified
25 terminal amino acid with a protease that specifically binds and removes the modified terminal amino acid; or subjecting the modified terminal amino acid to acidic or basic conditions sufficient to remove the modified terminal amino acid. In some embodiments, identifying the terminal amino acid comprises: identifying the terminal amino acid as being one type of the one or more types of terminal amino acids to which the one or more labeled affinity reagents bind; or
30 identifying the terminal amino acid as being a type other than the one or more types of terminal amino acids to which the one or more labeled affinity reagents bind.

 In some embodiments, the one or more labeled affinity reagents comprise one or more labeled aptamers, one or more labeled peptidases, one or more labeled antibodies, one or more labeled degradation pathway protein, one or more aminotransferase, one or more tRNA
35 synthetase, or a combination thereof. In some embodiments, the one or more labeled peptidases

have been modified to inactivate cleavage activity; or wherein the one or more labeled peptidases retain cleavage activity for the removing of (c).

In some aspects, the disclosure relates to kits for performing a method described herein, wherein the kit comprises a barcode component comprising a plurality of barcode molecules. In some embodiments, the barcode component further comprises a reaction component comprising one or more reagent for covalently attaching a barcode molecule to polypeptide. In some embodiments, the barcode component comprises one or more barcode molecules comprising a polynucleic acid portion, a polypeptide portion, and/or a fluorescent molecule portion.

In some embodiments, the polynucleic acid portion is 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, or 60 nucleotides in length. In some embodiments, the polynucleic acid portion comprises an aptamer.

In some embodiments, the polypeptide portion is 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 amino acids in length. In some embodiments, the polypeptide portion is an antibody or aptamer.

In some embodiments, the fluorescent molecule portion comprises an aromatic or heteroaromatic compound, such as a pyrene, anthracene, naphthalene, acridine, stilbene, indole, benzindole, oxazole, carbazole, thiazole, benzothiazole, phenanthridine, phenoxazine, porphyrin, quinoline, ethidium, benzamide, cyanine, carbocyanine, salicylate, anthranilate, coumarin, fluorescein, rhodamine, or the like. In some embodiments, the fluorescent molecule portion comprise a dye selected from the group consisting of a xanthene dye, a naphthalene dye, a coumarin dye, an acridine dye, a cyanine dye, a benzoxazole dye, a stilbene dye, a pyrene dye, a phthalocyanine dye, a phycobiliprotein dye, a squaraine dye, and a BODIPY dye.

In some embodiments, the kit further comprises a solid support. In some embodiments, the solid support comprises immobilized detector molecules comprising a polynucleic acid portion corresponding to a barcode molecule of the barcode component. In some embodiments, the solid support comprises immobilized detector molecules comprising a polypeptide portion corresponding to a barcode molecule of the barcode component. In some embodiments, the kit comprises a solid support that allows for the physical separation of populations of polypeptides of different origins.

In some aspects, the disclosure relates to devices comprising: at least one hardware processor; and at least one non-transitory computer-readable storage medium storing processor-executable instructions that, when executed by the at least one hardware processor, cause the at least one hardware processor to perform a method described herein.

In some aspects, the disclosure relates to non-transitory computer-readable storage mediums storing processor-executable instructions that, when executed by at least one hardware processor, cause the at least one hardware processor to perform a method described herein.

In some aspects, the disclosure relates to devices comprising: (i) a sample preparation module configured to interface with one or more cartridge, each cartridge comprising: (a) one or more reservoirs or reaction vessels configured to receive a complex sample; (b) one or more sequence sample preparation reagents, wherein the sample preparation reagents comprise a plurality of barcode molecules; and (c) a matrix comprising one or more immobilized capture probes. In some embodiments, the device further comprises (ii) a sequencing module comprising an array of pixels, wherein each pixel is configured to receive a sequencing sample from the sample preparation module and comprises: (a) a sample well; and (b) at least one photodetector.

In some embodiments, the sample preparation reagents further comprise a plurality of enrichment molecules. In some embodiments, at least a subset of the enrichment molecules in the plurality of enrichment molecules are covalently attached to an immobilized capture probe. In some embodiments, at least a subset of the enrichment molecules are covalently attached to a bead or particle that is capable of being bound by an immobilized capture probe. In some embodiments, each of the enrichment molecules in the plurality of enrichment molecules comprises an antibody, an aptamer, or an enzyme. In some embodiments, the enrichment molecules in a subset of the plurality of enrichment molecules comprise an antibody, an aptamer, or an enzyme.

In some embodiments, the sample preparation reagents comprise a modifying agent. In some embodiments, the modifying agent mediates polypeptide fragmentation, polypeptide denaturation, addition of a post-translational modification, and/or the blocking of one or more functional groups.

In some embodiments, the sequencing module further comprises a reservoir or reaction vessel configured to deliver sequencing reagents to the sample well of each pixel. In some embodiments, the sequencing reagents comprise a labeled affinity reagent. In some embodiments, the labeled affinity reagent comprises one or more labeled aptamers, one or more labeled peptidases, one or more labeled antibodies, one or more labeled degradation pathway protein, one or more aminotransferase, one or more tRNA synthetase, or a combination thereof.

BRIEF DESCRIPTION OF THE DRAWINGS

The skilled artisan will understand that the figures, described herein, are for illustration purposes only. It is to be understood that, in some instances, various aspects of the invention

may be shown exaggerated or enlarged to facilitate an understanding of the invention. In the drawings, like reference characters generally refer to like features, functionally similar and/or structurally similar elements throughout the various figures. The drawings are not necessarily to scale, emphasis instead being placed upon illustrating the principles of the teachings. The drawings are not intended to limit the scope of the present teachings in any way.

The features and advantages of the present invention will become more apparent from the detailed description set forth below when taken in conjunction with the drawings.

When describing embodiments in reference to the drawings, direction references (“above,” “below,” “top,” “bottom,” “left,” “right,” “horizontal,” “vertical,” etc.) may be used.

Such references are intended merely as an aid to the reader viewing the drawings in a normal orientation. These directional references are not intended to describe a preferred or only orientation of an embodied device. A device may be embodied in other orientations.

As is apparent from the detailed description, the examples depicted in the figures and further described for the purpose of illustration throughout the application describe non-limiting embodiments, and in some cases may simplify certain processes or omit features or steps for the purpose of clearer illustration.

FIG. 1 provides an exemplary illustration of a method for barcoding molecules (*e.g.*, polypeptides, polynucleotides, and/or metabolites) of single cells. The isolation of single cells can be done in various ways, included cell sorting. The barcode pool contacted with the first cell is different than the barcode pool contacted with the second cell.

FIG. 2 provides an exemplary illustration of multiplexed sample preparation and analysis. Samples 1-4 contain barcoded molecules, prepared as illustrated in FIG. 1. Samples 1-4 are then pooled, thereby generating a multiplexed sample. The origins of the barcoded molecules (*e.g.*, polypeptides, polynucleotides, and/or metabolites) are then determined and the barcoded molecules are grouped according to their origins. The barcoded molecules may also be analyzed by sequencing (*e.g.*, barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, etc.) or by detection and/or quantification (*e.g.*, barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, barcoded metabolites, etc.).

FIG. 3 provides an illustration depicting an exemplary workflow of preparing a multiplexed sample for polypeptide sequencing.

FIG. 4 provides an illustration depicting an exemplary workflow of preparing a multiplexed sample for polypeptide sequencing.

FIG. 5 provides an illustration depicting an exemplary workflow of preparing an enriched sample.

FIG. 6 provides an illustration depicting an exemplary workflow of preparing an enriched sample.

FIG. 7 provides an illustration depicting an exemplary workflow of preparing an enriched sample.

5 **FIG. 8** provides an illustration depicting an exemplary apparatus for preparing an enriched and/or multiplexed sample.

DETAILED DESCRIPTION

As described herein, the inventors have recognized and appreciated that differential binding interactions can provide an additional or alternative approach to conventional labeling strategies in polypeptide sequencing. Conventional polypeptide sequencing can involve labeling each type of amino acid with a uniquely identifiable label. This process can be laborious and prone to error, as there are at least twenty different types of naturally occurring amino acids in addition to numerous post-translational variations thereof. In some aspects, the disclosure relates to the discovery of techniques involving the use of amino acid recognition molecules which differentially associate with different types of amino acids to produce detectable characteristic signatures indicative of an amino acid sequence of a polypeptide.

15 In some aspects, the disclosure relates to the discovery that a polypeptide sequencing reaction can be monitored in real-time using only a single reaction mixture (e.g., without requiring iterative reagent cycling through a reaction vessel). Conventional polypeptide sequencing reactions can involve exposing a polypeptide to different reagent mixtures to cycle between steps of amino acid detection and amino acid cleavage. Accordingly, in some aspects, the disclosure relates to an advancement in next generation sequencing that allows for the analysis of polypeptides by amino acid detection throughout an ongoing degradation reaction in real-time.

25 Applicants have recognized that the ability to analyze the proteome of a single cell would provide insights into cellular processes and response patterns, leading to improved diagnostic and therapeutic strategies. However, unlike single-cell genomic and transcriptomic analyses, approaches for single-cell proteomic analysis have been limited to date at least because they are not scalable to analyzing single cell content. In some aspects, the disclosure relates to methods of single-cell sequencing, which facilitate the direct sequencing of the molecules of a single cell (e.g., polypeptides, DNA, and/or RNA) without amplification.

30 In some embodiments, the method comprises directly sequencing, in parallel, the proteome of a single cell and/or sequencing the genome and/or transcriptome of the single cell. In some embodiments, the proteome and genome of a single cell is sequenced simultaneously or

sequentially. In some embodiments, the proteome and transcriptome of a single cell is sequenced simultaneously or sequentially. In some embodiments, the proteome, genome, and transcriptome of a single cell is sequenced simultaneously or sequentially. In some embodiments, the metabolome of the single cell is also analyzed by detecting and optionally
5 quantifying one or more metabolites of the single cell.

Some embodiment may utilize molecular barcoding to facilitate multiplexed sample sequencing (*e.g.*, of a cell's proteome, genome, transcriptome, etc.) and analysis (*e.g.*, detection and/or quantification of a cell's proteome, genome, transcriptome, metabolome, etc.). For example, in some embodiments, the method comprises: (i) providing a cell sample (*e.g.*,
10 consisting of a single cell); and (ii) contacting the cell sample with a barcode component to produce a labeled sample comprising barcoded molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.).

In some embodiments, the cell sample contains only a single cell. In some embodiments, the cell of the cell sample is a living cell (*i.e.*, the barcode component is contacted with a living
15 cell). In other embodiments, the cell of the cell sample is a lysed cell (*i.e.*, the barcode component is contacted with the contents of the lysed cell).

In some embodiments, (i) comprises: (a) providing a cell population; and (b) lysing the cell population. In some embodiments, the cell population: consists of a single cell; comprises a plurality of homogeneous cells; or comprises a plurality of heterogeneous cells. In some
20 embodiments, the cell population is isolated from a subject. In some embodiments, the subject is a human, mouse, rat, or non-human primate subject.

The barcoded molecules of the labeled sample may comprise barcoded polypeptides, barcoded polynucleotides (*e.g.*, barcoded DNA, barcoded RNA, barcoded cDNA, etc.), barcoded metabolites, or a combination thereof. It is understood that the barcoding of polypeptides,
25 polynucleic acids (*e.g.*, DNA, RNA, cDNA, etc.), and/or metabolites may be performed in any order. In some instance, two or more of polypeptides, polynucleic acids (*e.g.*, DNA, RNA, cDNA, etc.), and metabolites are barcoded simultaneously.

For example, in some embodiments (ii) comprises: (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded polypeptides; (b)
30 isolating the barcoded polypeptides from the first sample, thereby generating a second sample comprising the barcoded polypeptides and a third sample comprising the genome, transcriptome, and/or metabolome of the cell sample; and (c) contacting the third sample with an additional barcode component, to produce a fourth sample comprising barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites; wherein the labeled sample comprises the second
35 sample and the fourth sample. In some embodiments, the third sample in (b) comprises: (i) a

first subsample comprising the genome and transcriptome of the single cell and a second subsample comprising the metabolome of the single cell; (ii) a first subsample comprising the genome and metabolome of the single cell and a second subsample comprising the transcriptome of the single cell; (iii) a first subsample comprising the metabolome and transcriptome of the single cell and a second subsample comprising the genome of the single cell; or (iv) a first subsample comprising the genome of the single cell, a second subsample comprising the transcriptome of the single cell, and a third subsample comprising the metabolome of the single cell. In some embodiments, the additional barcode component in (c) comprises a second, third, fourth, or fifth barcode component. In some embodiments, the barcoded molecules of the fourth sample in (c) comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

In other embodiments (ii) comprises: (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA; (b) amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA of the first sample; and (c) contacting the first sample with a second barcode component, to produce a second sample comprising barcoded polypeptides and/or barcoded metabolites; wherein the labeled sample comprises the second sample.

In other embodiments, (ii) comprises: (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA; (b) isolating the barcoded DNA, barcoded RNA, and/or barcoded cDNA from the first sample, thereby generating a second sample comprising the barcoded DNA and/or barcoded cDNA and a third sample comprising the proteome and/or metabolome of the single cell in the cell sample; and (c) contacting the third sample with an additional barcode component, to produce a fourth sample comprising barcoded polypeptides and/or the barcoded metabolites; wherein the labeled sample comprises the second sample and the fourth sample. In some embodiments, the third sample in (b) comprises a first subsample comprising the proteome of the single cell and a second subsample comprising the metabolome of the single cell. In some embodiments, the additional barcode component in (c) comprises a second or third barcode component. In some embodiments, the barcoded molecules of the first sample in (a) comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

In some embodiments, the method further comprises (iii) sequencing the barcoded polypeptides, barcoded DNA, barcoded RNA, and/or barcoded cDNA of the labeled sample (or multiplexed sample). In some embodiments, the barcoded molecules of the labeled sample are not amplified prior to sequencing. In some embodiments, the method further comprises

detecting and optionally quantifying the barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites of the labeled sample.

Also provide herein are compositions, kits and devices useful for the direct sequencing of the proteome (and optionally sequencing the genome, and/or transcriptome, and optionally analyzing the metabolome) of a single cell.

I. Methods of Preparing a Complex Sample

In some aspects, the disclosure relates to methods of preparing a complex sample (*e.g.*, a complex polypeptide sample). As used herein, the term “complex sample” refers to a sample comprising a plurality of molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.), at least two of which are chemically unique. In some embodiments, a complex sample comprises a plurality of polypeptides, wherein the plurality comprises at least two polypeptides that comprise different amino acid sequences. In some embodiments, a complex sample comprises a plurality of polynucleic acids, wherein the plurality comprises at least two polynucleic acids that comprise different nucleotide sequences.

Typically, the complex sample is derived from a population of cells (*e.g.*, produced by a population of cells). In some embodiments, the population of cells consists of a single cell. In other embodiments, the population of cells comprises two or more cells.

For example, in some embodiments the population of cells comprises at least 5, at least 10, at least 20, at least 30, at least 40, at least 50, at least 60, at least 70, at least 80, at least 90, at least 100, at least 150, at least 200, at least 250, at least 300, at least 350, at least 400, at least 450, at least 500, at least 600, at least 700, at least 800, at least 900, at least 1×10^3 , at least 1×10^4 , at least 1×10^5 , at least 1×10^6 , at least 1×10^7 , at least 1×10^8 , at least 1×10^9 , or at least 1×10^{10} cells.

In some embodiments, the population comprises 1-5, 1-10, 1-20, 1-30, 1-50, 1-60, 1-70, 1-80, 1-90, 1-100, 1-150, 1-200, 1-250, 1-300, 1-350, 1-400, 1-450, 1-500, 1-600, 1-700, 1-800, 1-900, $1-1 \times 10^3$, $1-1 \times 10^4$, $1-1 \times 10^5$, $1-1 \times 10^6$, $1-1 \times 10^7$, $1-1 \times 10^8$, $1-1 \times 10^9$, $1-1 \times 10^{10}$, 100-150, 100-200, 100-250, 100-300, 100-350, 100-400, 100-450, 100-500, 100-600, 100-700, 100-800, 100-900, $100-1 \times 10^3$, $100-1 \times 10^4$, $100-1 \times 10^5$, $100-1 \times 10^6$, $100-1 \times 10^7$, $100-1 \times 10^8$, $100-1 \times 10^9$, $100-1 \times 10^{10}$, $1 \times 10^3-1 \times 10^4$, $1 \times 10^3-1 \times 10^5$, $1 \times 10^3-1 \times 10^6$, $1 \times 10^3-1 \times 10^7$, $1 \times 10^3-1 \times 10^8$, $1 \times 10^3-1 \times 10^9$, $1 \times 10^3-1 \times 10^{10}$, $1 \times 10^4-1 \times 10^5$, $1 \times 10^4-1 \times 10^6$, $1 \times 10^4-1 \times 10^7$, $1 \times 10^4-1 \times 10^8$, $1 \times 10^4-1 \times 10^9$, $1 \times 10^4-1 \times 10^{10}$, $1 \times 10^5-1 \times 10^6$, $1 \times 10^5-1 \times 10^7$, $1 \times 10^5-1 \times 10^8$, $1 \times 10^5-1 \times 10^9$, or $1 \times 10^5-1 \times 10^{10}$ cells.

A population of cells may comprise prokaryotic cells and/or eukaryotic cells. A population of cells may comprise a plurality of homogeneous cells. Alternatively, a population of cells may comprise a plurality of heterogeneous cells.

A population of cells may be isolated from a subject (*e.g.*, a multicellular or symbiotic organism). In some embodiments, the subject is a mouse, rat, rabbit, guinea pig, hamster, pig, sheep, dog, primate, cat, or human.

Methods of isolating populations of cells are known to those having skill in the art. For example, a method of preparing a complex sample may comprise biopsy, dissection (*e.g.*, microdissection, such as laser capture), limited dilution, micromanipulation, immunomagnetic cell separation, fluorescence-activated cell sorting, density gradient centrifugation, immunodensity cell isolation, microfluidic cell sorting, sedimentation, adhesion, or a combination thereof.

In some embodiments, the method of preparing a complex sample comprises lysing a population of cells, thereby generating a lysis sample comprising a plurality of molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.). Methods of lysing a population of cells are known to those having ordinary skill in the art. In some embodiments, a sample comprising cells is lysed using any one of known physical or chemical methodologies to release a target molecule from said cells. In some embodiments, a sample may be lysed using an electrolytic method, an enzymatic method, a detergent-based method, and/or mechanical homogenization. In some embodiments, if a sample does not comprise cells or tissue (*e.g.*, a sample comprising purified polypeptides), a lysis step may be omitted.

Alternatively, or in addition, a method of preparing a complex sample may comprise subcellular fractionation (*i.e.*, the isolation of one or more cellular compartment, such as endosomes, snyaptosomes, cytoplasm, nucleoplasm, chromatin, mitochondria, peroxisomes, lysosomes, melanosomes, exosomes, Golgi apparatus, endoplasmic reticulum, centrosomes, pseudopodia, or a combination thereof).

Molecules derived from the same cell population are described herein as having the same “origin”.

II. Methods of Preparing a Multiplexed Sample

In some aspects, the disclosure relates to methods of preparing a multiplexed sample. As used herein, the term “multiplexed sample” refers to a sample comprising at least two subsamples having different origins (*e.g.*, two or more samples, each prepared from a different population of cells or plurality of molecules).

In some embodiments, a multiplexed sample comprises at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at least 17, at least 18, at least 19, at least 20, at least 25, at least 30, at least 35, at least 40, at least 45, at least 50, at least 60, at least 70, at least 80, at least 90, at least 100, at least 200, at least 300, at least 400, at least 500, at least 600, at least 700, at least 800, at least 900, or at least 1000 subsamples each having different origins.

In some embodiments, a multiplexed sample comprises 2-3, 2-4, 2-5, 2-6, 2-7, 2-8, 2-9, 2-10, 2-11, 2-12, 2-13, 2-14, 2-15, 2-16, 2-17, 2-18, 2-19, 2-20, 2-25, 2-30, 2-35, 2-40, 2-45, 2-50, 2-60, 2-70, 2-80, 2-90, 2-100, 2-200, 2-300, 2-400, 2-500, 2-600, 2-700, 2-800, 2-900, 2-1000, 5-10, 5-15, 5-20, 5-25, 5-30, 5-35, 5-40, 5-45, 5-50, 5-60, 5-70, 5-80, 5-90, 5-100, 5-200, 5-300, 5-400, 5-500, 5-600, 5-700, 5-800, 5-900, 10-15, 10-20, 10-25, 10-30, 10-35, 10-40, 10-45, 10-50, 10-60, 10-70, 10-80, 10-90, 10-100, 10-200, 10-300, 10-400, 10-500, 10-600, 10-700, 10-800, 10-900, 10-1000, 20-30, 20-40, 20-50, 20-60, 20-70, 20-80, 20-90, 20-100, 20-200, 20-300, 20-400, 20-500, 20-600, 20-700, 20-800, 20-900, 20-1000, 50-60, 50-70, 50-80, 50-90, 50-100, 50-200, 50-300, 50-400, 50-500, 50-600, 50-700, 50-800, 50-900, 50-1000, 100-200, 100-300, 100-400, 100-500, 100-600, 100-700, 100-800, 100-900, 100-1000, 500-600, 500-700, 1500-800, 500-900, or 500-1000 subsamples each having different origins.

In some embodiments, a multiplexed sample comprises 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50 subsamples each having different origins.

Each subsample in a multiplexed sample may comprise a plurality of molecules. In some embodiments, one or more of the subsamples in a multiplexed sample comprises: the molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.) of a complex sample prepared from a cell population (which may be a single cell) (see “Methods of Preparing a Complex Sample”); or the molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.) of an enriched sample (see “Methods of Preparing an Enriched Sample”). In some embodiments, the plurality of molecules of a subsample are derived from a single molecule (*e.g.*, through the fragmentation of a single polypeptide).

Each subsample in a multiplexed sample may comprises a single molecule (*e.g.*, a single polypeptide, a single polynucleic acid, a single metabolite, etc.). In some embodiments, one or more subsample in a multiplexed sample comprises a single molecule (*e.g.*, a single polypeptide, a single polynucleic acid, a single metabolite, etc.).

Typically, at least a subset of the molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.) in each subsample in a multiplexed sample can be distinguished from the molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.) of the other subsamples in the

multiplexed sample. For example, in some embodiments, at least a subset of the polypeptides in each subsample in a multiplexed sample can be distinguished from the polypeptides of the other subsamples in the multiplexed sample. In this way, the origins of at least a subset of the molecules in a multiplexed sample can be identified.

5 As such, in some embodiments, at least one of the subsamples in a multiplexed sample comprises barcoded molecules, each barcoded molecule comprising a barcode unique to the subsample (*i.e.*, a unique barcode). A barcode is considered unique to a subsample, if the barcode is not found on a molecule of any other subsample in the multiplexed sample.

10 In some embodiments, two or more of the subsamples in a multiplexed sample comprise barcoded molecules. In some embodiments, each of the subsamples in a multiplexed sample comprises barcoded molecules. In some embodiments, all but one of the subsamples in a multiplexed sample comprise barcoded molecules.

15 Within a multiplexed sample, the barcoded molecules of each subsample comprising barcoded molecules (*i.e.*, each “labeled subsample”) comprise unique barcodes. In some embodiments, each of the barcoded molecules in a labeled subsample comprise the same barcode. In some embodiments, the barcode molecules in a labeled subsample comprise a combination of unique barcodes. For example, in some embodiments, a labeled subsample comprises a unique combination of 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 barcoded molecules.

20 In some embodiments, a labeled subsample comprises barcoded polypeptides, barcoded DNA molecules, barcoded RNA molecules, barcoded cDNA molecules, barcoded metabolites, or a combination thereof, wherein: the barcoded polypeptides comprise a first barcode (or a first combination of barcodes); the barcoded DNA molecules comprise a second barcode (or a second combination of barcodes); the barcoded RNA molecules in the subsample comprise a third barcode (or a third combination of barcodes); the barcoded cDNA molecules comprise a fourth barcode (or a fourth combination of barcodes); the barcoded metabolites comprise a fifth barcode (or a fifth combination of barcodes); or a combination thereof.

25 In some embodiments, a method of preparing a multiplexed sample comprises: (i) contacting a population of cells with a barcode component to produce a sample (*i.e.*, a first labeled subsample) comprising barcoded molecules (*e.g.*, barcoded polypeptides, barcoded polynucleic acids, barcoded metabolites, or a combination thereof); and (ii) combining the sample of (i) with one or more supplemental sample (*i.e.*, one or more additional subsample) to generate a multiplexed sample.

35 In some embodiments, a method of preparing a multiplexed sample comprises: (i) contacting a plurality of molecules with a barcode component to produce a sample (*i.e.*, a first

labeled subsample) comprising barcoded molecules (*e.g.*, barcoded polypeptides, barcoded polynucleic acids, barcoded metabolites, or a combination thereof); and (ii) combining the sample of (i) with one or more supplemental sample (*i.e.*, one or more additional subsample) to generate a multiplexed sample.

5 In some of the embodiments described in the preceding two paragraphs, step (ii) further comprises depositing the multiplexed sample on or within a solid substrate. In some embodiments, the solid substrate comprises a plurality of immobilized (*e.g.*, covalently-attached) detector molecules, wherein one or more the detector molecules interacts with a barcode of a barcoded molecule of the multiplexed sample. In some embodiments, the solid substrate is a
10 chip array.

In some embodiments, a method of preparing a multiplexed sample comprises: (i) providing at least two populations of molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.); (ii) depositing the at least two populations of molecules of (i) on or within a solid substrate, wherein each population of molecules remains physically separated from the
15 other populations of molecules in (i); thereby preparing a multiplexed sample.

A. Methods of Molecule Barcoding

In some aspects, the disclosure relates to methods of barcoding molecules (*e.g.*, polypeptides, polynucleotides (such as DNA, RNA, cDNA, etc.) metabolites, etc.) of a sample.

20 In some embodiments, the sample comprises living cells. In some embodiments, the sample is a complex sample prepared from a cell population (which may be a single cell) (see “Methods of Preparing a Complex Sample”). In some embodiments, the sample is an enriched sample (see “Methods of Preparing an Enriched Sample”). In some embodiments, the sample comprises a single molecule (*e.g.*, a polypeptide, polynucleic acid, metabolite, etc.) or fragments derived
25 from a single molecule (*e.g.*, fragments of the polypeptide, fragments of a polynucleic acid, fragments of a metabolite, etc.).

Of particular relevance here, the disclosure relates to methods of barcoding molecules. Molecules may be barcoded by chemical modification and/or physical separation.

(i) *Chemical Modification*

A molecule (*e.g.*, a polypeptide, polynucleic acid, metabolite, etc.) or a plurality of molecules may be barcoded by chemical modification. Chemical modification of a molecule changes the chemical composition of the molecule and can occur during synthesis of the molecule (*in vivo* or *in vitro*) or after synthesis of the molecule. A molecule may be modified at
35 any position. Methods of performing chemical modification (*e.g.*, chemical conjugation) that can be used arrive at a barcoded molecule have been previously described, and are known to those

having ordinary skill in the art. See e.g., Corey et al., *Science*, 1987; 238: 1401–1403; Kukolka et al., *Org. Biomol. Chem.*, 2004; 2: 2203–2206; Debets et al., *Chem. Commun.*, 2010; 46: 97–99; Takeda et al., *Bioorg. Med. Chem. Lett.*, 2004; 14: 2407–2410; Yang et al., *Bioconjug. Chem.*, 2015; 26: 1381–1395; Rosen et al., *Nat. Chem.*, 2014; 6: 804–809; Cong et al.,
5 *Bioconjug. Chem.*, 2012; 23: 248–263; Mattson, G., et al. *Molecular Biology Reports*, 1993; 17:167-183.

In some embodiments, a molecule (*e.g.*, a polypeptide, polynucleic acid, metabolite, etc.) or a plurality of molecules is barcoded through a method comprising contacting a population of cells with a barcode component to produce a sample comprising barcoded molecules. In such an
10 instance, the molecule (or plurality of molecules) may be modified during synthesis or after synthesis.

In some embodiments, a molecule (*e.g.*, a polypeptide, polynucleic acid, metabolite, etc.) or a plurality of molecules is barcoded through a method comprising contacting the molecule (or the plurality of molecules) with a barcode component to produce a sample comprising barcoded
15 molecules. In such an instance, the molecule (or plurality of molecules) would be modified after synthesis.

A barcode component may comprise a modifying agent. The modifying agent may comprise an endoprotease having a distinct cleavage pattern. Examples of endoproteases are known to those having ordinary skill in the art and include, but are not limited to, trypsin,
20 chymotrypsin, elastase, thermolysin, pepsin, glutamyl endopeptidase, neprilysin, Lys-C, Arg-C, Asp-N, Lys-N, Glu-C, WaLP, and MaLP. See e.g., Giansanti et al., *Nat. Protoc.*, 2016 Apr. 28; 11(5): 993-1006. The modifying agent may comprise an enzyme capable of modifying polypeptides with a post-translational modification. Examples of post-translational modifications are known to those having skill in the art and include, but are not limited to,
25 acetylation, adenylation, ADP-ribosylation, alkylation (*e.g.*, methylation), amidation, arginylation, biotinylation, butyrylation, carbamylation, carbonylation, carboxylation, citrullination, deamidation, eliminination, formylation, glycosylation (*e.g.*, N-linked glycosylation, O-linked glycosylation), glypiation, glycation, hydroxylation, iodination, ISGylation, isoprenylation, lipoylation, malonylation, myristoylation, neddylation, nitration,
30 oxidation, palmitoylation, pegylation, phosphorylation, phosphopantetheinylation, polyglutamylation, prenylation, propionylation, pupylation, S-glutathionylation, S-nitrosylation, S-sulfenylation, S-sulfinylation, S-sulfonylation, succinylation, sulfation, SUMOylation, and ubiquitination. Enzymes responsible for modifying polypeptides in these ways are also known to those having skill in the art.

Alternatively or in addition, a barcode component may comprises a plurality of barcode molecules. In some embodiments, a barcode component consists of a plurality of barcode molecules. In some embodiments, a barcode component may further comprise one or more reagents (*e.g.*, enzymes, compounds, small molecules, buffers, and the like) to facilitate the covalently attachment of a barcode molecule to a molecule (*e.g.*, a polypeptide, polynucleic acid, metabolite, etc.) of a sample.

Barcode molecules may be covalently attached to a molecule at any position. For example, in some embodiments, a barcode molecule is covalently attached to a polypeptide at an amino acid position within 10, 9, 8, 7, 6, 5, 4, 3, or 2 amino acids of its terminus (N-terminus or C-terminus). In some embodiments, a barcode molecule is covalently attached to a polypeptide at its N-terminus. In some embodiments, a barcode is covalently attached to a polypeptide at its C-terminus. In some embodiments, a barcode is covalently attached to the 5' end of a polynucleic acid. In some embodiments, a barcode is covalently attached to the 3' end of a polynucleic acid.

In some embodiments, each of the barcode molecules of a barcode component are chemically identical. In some embodiments, a barcode component comprises two or more chemically distinct barcode molecules. For example, a barcode component may comprise 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 chemically distinct barcode molecules.

A barcode molecule of a barcode component may be an unnatural amino acid (*i.e.*, non-canonical amino acid). Examples of unnatural amino acids are known to those having skill in the art and include, but are not limited to, homoallylglycine (Hag), homopropargylglycine (Hpg), azidohomoalanine (Aha), azidonorleucine (Anl), azidophenylalanine (Azf), acetylphenylalanine (Acf), and propargyloxyphenylalanine (Pxf). In some embodiments, wherein the barcode component comprises unnatural amino acid barcode molecules, the barcode component further comprises one or more non-natural tRNA (or a nucleic acid encoding an expressible form of a non-natural tRNA). Examples of non-natural tRNAs are known to those having skill in the art. A barcode molecule of a barcode component may be an unnatural nucleotide (*i.e.*, nucleotide analog). Examples of unnatural nucleotides are known to those having ordinary skill in the art and include, but are not limited to, d5SICS, dNaM, 2-Aminopurine, 5-Nitroindole, Iso-dC, Iso-dG, and 5-Bromo dU. See *e.g.*, Malyshev D.A. et al., Efficient and sequence-independent replication of DNA containing a third base pair establishes a functional six-letter genetic alphabet, *Pro. Natl. Acad. Sci. U.S.A.*, 2012 Jul 24; 109(30): 12005-10.

Alternatively, or in addition, a barcode molecule of a barcode component may comprise a polynucleic acid portion, a polypeptide portion, a small molecule portion, a linker (*e.g.*, a peg-like linker), a dendrimer, a scaffold, or a combination thereof. In some embodiments, a barcode

molecule of a barcode component comprises a polynucleic acid portion, a polypeptide portion, a small molecule portion, a linker (*e.g.*, a peg-like linker), a dendrimer, a scaffold, or a combination thereof.

In some embodiments, a barcode molecule comprises a polynucleic acid portion. In some embodiments, a barcode molecule comprises two or more polynucleic acid portions. In embodiments wherein a barcode molecule comprises multiple polynucleic acid portions: each polynucleic acid portion may be identical; a subset of the polynucleic acid portions may be identical; or each polynucleic acid portion may be chemically distinct.

In some embodiment, the polynucleic acid portion is 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, or 60 nucleotides in length.

In some embodiment, the polynucleic acid portion is at least 5, at least 10, at least 15, at least 20, at least 25, at least 30, at least 40, at least 50, at least 60, at least 70, at least 80, at least 90, at least 100, at least 150, at least 200, at least 250, at least 300, at least 350, at least 400, at least 450, or at least 500 nucleotides in length.

In some embodiments, the polynucleic acid portion is 5-10, 5-15, 5-20, 5-25, 5-30, 5-40, 5-50, 5-60, 5-70, 5-80, 5-90, 5-100, 5-150, 5-200, 5-250, 5-300, 5-350, 5-400, 5-450, 5-500, 10-15, 10-20, 10-25, 10-30, 10-40, 10-50, 10-60, 10-70, 10-80, 10-90, 10-100, 10-150, 10-200, 10-250, 10-300, 10-350, 10-400, 10-450, 10-500, 20-30, 20-40, 20-50, 20-60, 20-70, 20-80, 20-90, 20-100, 20-150, 20-200, 20-250, 20-300, 20-350, 20-400, 20-450, 20-500, 50-75, 50-100, 50-150, 50-200, 50-250, 50-500, 50-350, 50-400, 50-450, 50-500, 100-200, 100-250, 100-500, 100-350, 100-400, 100-450, or 100-500 nucleotides in length.

In some embodiment, the polynucleic acid portion is an aptamer.

In some embodiments, a barcode molecule comprises a polypeptide portion. In some embodiments, a barcode molecule comprises two or more polypeptide portions. In embodiments wherein a barcode molecule comprises multiple polypeptide portions: each polypeptide portion may be identical; a subset of the polypeptide portions may be identical; or each polypeptide portion may be chemically distinct.

In some embodiment, the polypeptide portion is 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 amino acids in length. In some embodiments, the polypeptide portion is at least 5, at least 10, at least 15, at least 20, at least 25, at least 30, at least 40, at least 50, at least 60, at least 70, at least 80, at least 90, at least 100, at least 150, at least 200, at least 250, at least 300, at least 350, at least 400, at least 450, or at least 500 amino acids in length. In some embodiments, the polypeptide portion is 5-10, 5-15, 5-20, 5-25, 5-30, 5-40, 5-50, 5-60, 5-70, 5-

80, 5-90, 5-100, 5-150, 5-200, 5-250, 5-300, 5-350, 5-400, 5-450, 5-500, 10-15, 10-20, 10-25, 10-30, 10-40, 10-50, 10-60, 10-70, 10-80, 10-90, 10-100, 10-150, 10-200, 10-250, 10-300, 10-350, 10-400, 10-450, 10-500, 20-30, 20-40, 20-50, 20-60, 20-70, 20-80, 20-90, 20-100, 20-150, 20-200, 20-250, 20-300, 20-350, 20-400, 20-450, 20-500, 50-75, 50-100, 50-150, 50-200, 50-250, 50-500, 50-350, 50-400, 50-450, 50-500, 100-200, 100-250, 100-500, 100-350, 100-400, 100-450, or 100-500 amino acids in length.

In some embodiments, the polypeptide portion is an aptamer. In some embodiment, the polypeptide portion is an antibody. In some embodiments, the polypeptide portion is an antigen.

In some embodiments, a barcode molecule comprises a small molecule portion. In some 10 embodiments, a barcode molecule comprises two or more small molecule portions. In embodiments wherein a barcode molecule comprises multiple small molecule portions: each small molecule portion may be identical; a subset of the small molecule portions may be identical; or each small molecule portion may be chemically distinct.

In some embodiments, the small molecule portion comprises biotin.

15 In some embodiments, the small molecule portion comprises a drug or a luminescent molecule (or a fluorescent molecule). Examples of drugs and luminescent molecules suitable for the methods described herein are known to those having skill in the art. As used herein, a luminescent molecule is a molecule that absorbs one or more photons and may subsequently emit one or more photons after one or more time durations.

20 In some embodiments, a luminescent molecule may comprise a first and second chromophore. In some embodiments, an excited state of the first chromophore is capable of relaxation via an energy transfer to the second chromophore. In some embodiments, the energy transfer is a Förster resonance energy transfer (FRET). Such a FRET pair may be useful for providing a luminescent label with properties that make the label easier to differentiate from 25 amongst a plurality of luminescent labels in a mixture. In yet other embodiments, a FRET pair comprises a first chromophore of a first luminescent label and a second chromophore of a second luminescent label. In certain embodiments, the FRET pair may absorb excitation energy in a first spectral range and emit luminescence in a second spectral range.

In some embodiments, a luminescent molecule refers to a fluorophore or a dye.

30 Typically, a luminescent molecule comprises an aromatic or heteroaromatic compound and can be a pyrene, anthracene, naphthalene, naphthylamine, acridine, stilbene, indole, benzindole, oxazole, carbazole, thiazole, benzothiazole, benzoxazole, phenanthridine, phenoxazine, porphyrin, quinoline, ethidium, benzamide, cyanine, carbocyanine, salicylate, anthranilate, coumarin, fluorescein, rhodamine, xanthene, or other like compound.

In some embodiments, a luminescent molecule comprises a dye selected from one or more of the following: 5/6-Carboxyrhodamine 6G, 5-Carboxyrhodamine 6G, 6-Carboxyrhodamine 6G, 6-TAMRA, Abberior® STAR 440SXP, Abberior® STAR 470SXP, Abberior® STAR 488, Abberior® STAR 512, Abberior® STAR 520SXP, Abberior® STAR 580, Abberior® STAR 600, Abberior® STAR 635, Abberior® STAR 635P, Abberior® STAR RED, Alexa Fluor® 350, Alexa Fluor® 405, Alexa Fluor® 430, Alexa Fluor® 480, Alexa Fluor® 488, Alexa Fluor® 514, Alexa Fluor® 532, Alexa Fluor® 546, Alexa Fluor® 555, Alexa Fluor® 568, Alexa Fluor® 594, Alexa Fluor® 610-X, Alexa Fluor® 633, Alexa Fluor® 647, Alexa Fluor® 660, Alexa Fluor® 680, Alexa Fluor® 700, Alexa Fluor® 750, Alexa Fluor® 790, AMCA, ATTO 390, ATTO 425, ATTO 465, ATTO 488, ATTO 495, ATTO 514, ATTO 520, ATTO 532, ATTO 542, ATTO 550, ATTO 565, ATTO 590, ATTO 610, ATTO 620, ATTO 633, ATTO 647, ATTO 647N, ATTO 655, ATTO 665, ATTO 680, ATTO 700, ATTO 725, ATTO 740, ATTO Oxa12, ATTO Rho101, ATTO Rho11, ATTO Rho12, ATTO Rho13, ATTO Rho14, ATTO Rho3B, ATTO Rho6G, ATTO Thio12, BD Horizon™ V450, BODIPY® 493/501, BODIPY® 530/550, BODIPY® 558/568, BODIPY® 564/570, BODIPY® 576/589, BODIPY® 581/591, BODIPY® 630/650, BODIPY® 650/665, BODIPY® FL, BODIPY® FL-X, BODIPY® R6G, BODIPY® TMR, BODIPY® TR, CAL Fluor® Gold 540, CAL Fluor® Green 510, CAL Fluor® Orange 560, CAL Fluor® Red 590, CAL Fluor® Red 610, CAL Fluor® Red 615, CAL Fluor® Red 635, Cascade® Blue, CF™350, CF™405M, CF™405S, CF™488A, CF™514, CF™532, CF™543, CF™546, CF™555, CF™568, CF™594, CF™620R, CF™633, CF™633-V1, CF™640R, CF™640R-V1, CF™640R-V2, CF™660C, CF™660R, CF™680, CF™680R, CF™680R-V1, CF™750, CF™770, CF™790, Chromeo™ 642, Chromis 425N, Chromis 500N, Chromis 515N, Chromis 530N, Chromis 550A, Chromis 550C, Chromis 550Z, Chromis 560N, Chromis 570N, Chromis 577N, Chromis 600N, Chromis 630N, Chromis 645A, Chromis 645C, Chromis 645Z, Chromis 678A, Chromis 678C, Chromis 678Z, Chromis 770A, Chromis 770C, Chromis 800A, Chromis 800C, Chromis 830A, Chromis 830C, Cy®3, Cy®3.5, Cy®3B, Cy®5, Cy®5.5, Cy®7, DyLight® 350, DyLight® 405, DyLight® 415-Co1, DyLight® 425Q, DyLight® 485-LS, DyLight® 488, DyLight® 504Q, DyLight® 510-LS, DyLight® 515-LS, DyLight® 521-LS, DyLight® 530-R2, DyLight® 543Q, DyLight® 550, DyLight® 554-R0, DyLight® 554-R1, DyLight® 590-R2, DyLight® 594, DyLight® 610-B1, DyLight® 615-B2, DyLight® 633, DyLight® 633-B1, DyLight® 633-B2, DyLight® 650, DyLight® 655-B1, DyLight® 655-B2, DyLight® 655-B3, DyLight® 655-B4, DyLight® 662Q, DyLight® 675-B1, DyLight® 675-B2, DyLight® 675-B3, DyLight® 675-B4, DyLight® 679-C5, DyLight® 680, DyLight® 683Q, DyLight® 690-B1, DyLight® 690-B2, DyLight® 696Q, DyLight® 700-B1, DyLight® 700-B1, DyLight® 730-B1, DyLight® 730-B2, DyLight® 730-

B3, DyLight® 730-B4, DyLight® 747, DyLight® 747-B1, DyLight® 747-B2, DyLight® 747-B3, DyLight® 747-B4, DyLight® 755, DyLight® 766Q, DyLight® 775-B2, DyLight® 775-B3, DyLight® 775-B4, DyLight® 780-B1, DyLight® 780-B2, DyLight® 780-B3, DyLight® 800, DyLight® 830-B2, Dyomics-350, Dyomics-350XL, Dyomics-360XL, Dyomics-370XL,
5 Dyomics-375XL, Dyomics-380XL, Dyomics-390XL, Dyomics-405, Dyomics-415, Dyomics-430, Dyomics-431, Dyomics-478, Dyomics-480XL, Dyomics-481XL, Dyomics-485XL, Dyomics-490, Dyomics-495, Dyomics-505, Dyomics-510XL, Dyomics-511XL, Dyomics-520XL, Dyomics-521XL, Dyomics-530, Dyomics-547, Dyomics-547P1, Dyomics-548, Dyomics-549, Dyomics-549P1, Dyomics-550, Dyomics-554, Dyomics-555, Dyomics-556,
10 Dyomics-560, Dyomics-590, Dyomics-591, Dyomics-594, Dyomics-601XL, Dyomics-605, Dyomics-610, Dyomics-615, Dyomics-630, Dyomics-631, Dyomics-632, Dyomics-633, Dyomics-634, Dyomics-635, Dyomics-636, Dyomics-647, Dyomics-647P1, Dyomics-648, Dyomics-648P1, Dyomics-649, Dyomics-649P1, Dyomics-650, Dyomics-651, Dyomics-652, Dyomics-654, Dyomics-675, Dyomics-676, Dyomics-677, Dyomics-678, Dyomics-679P1,
15 Dyomics-680, Dyomics-681, Dyomics-682, Dyomics-700, Dyomics-701, Dyomics-703, Dyomics-704, Dyomics-730, Dyomics-731, Dyomics-732, Dyomics-734, Dyomics-749, Dyomics-749P1, Dyomics-750, Dyomics-751, Dyomics-752, Dyomics-754, Dyomics-776, Dyomics-777, Dyomics-778, Dyomics-780, Dyomics-781, Dyomics-782, Dyomics-800, Dyomics-831, eFluor® 450, Eosin, FITC, Fluorescein, HiLyte™ Fluor 405, HiLyte™ Fluor 488,
20 HiLyte™ Fluor 532, HiLyte™ Fluor 555, HiLyte™ Fluor 594, HiLyte™ Fluor 647, HiLyte™ Fluor 680, HiLyte™ Fluor 750, IRDye® 680LT, IRDye® 750, IRDye® 800CW, JOE, LightCycler® 640R, LightCycler® Red 610, LightCycler® Red 640, LightCycler® Red 670, LightCycler® Red 705, Lissamine Rhodamine B, Naphthofluorescein, Oregon Green® 488, Oregon Green® 514, Pacific Blue™, Pacific Green™, Pacific Orange™, PET, PF350, PF405,
25 PF415, PF488, PF505, PF532, PF546, PF555P, PF568, PF594, PF610, PF633P, PF647P, Quasar® 570, Quasar® 670, Quasar® 705, Rhodamine 123, Rhodamine 6G, Rhodamine B, Rhodamine Green, Rhodamine Green-X, Rhodamine Red, ROX, Seta™ 375, Seta™ 470, Seta™ 555, Seta™ 632, Seta™ 633, Seta™ 650, Seta™ 660, Seta™ 670, Seta™ 680, Seta™ 700, Seta™ 750, Seta™ 780, Seta™ APC-780, Seta™ PerCP-680, Seta™ R-PE-670, Seta™ 646,
30 SeTau 380, SeTau 425, SeTau 647, SeTau 405, Square 635, Square 650, Square 660, Square 672, Square 680, Sulforhodamine 101, TAMRA, TET, Texas Red®, TMR, TRITC, Yakima Yellow™, Zenon®, Zy3, Zy5, Zy5.5, and Zy7.

(ii) *Physical Separation*

A molecule (*e.g.*, a polypeptide, polynucleic acid, metabolite, etc.) or plurality of molecules may be barcoded by physical separation. In some embodiments, a molecule (or plurality of molecules) is deposited on or within a solid substrate such that the molecule (or plurality of molecules) remains physically separated from additional molecules (or additional pluralities of molecules).

In some embodiments, the solid substrate is a chip array.

In some embodiments, the chip array comprises a plurality of compartments (*e.g.*, wells) and/or injection ports. For example, in some embodiments, the chip array comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 compartments. In some embodiments, the chip array comprises 1-2, 1-3, 1-4, 1-5, 1-6, 1-7, 1-8, 1-9, 1-10, 1-11, 1-12, 1-13, 1-14, 1-15, 1-16, 1-17, 1-18, 1-19, 1-20, 2-3, 2-4, 2-5, 2-6, 2-7, 2-8, 2-9, 2-10, 2-11, 2-12, 2-13, 2-14, 2-15, 2-16, 2-17, 2-18, 2-19, 2-20, 3-4, 3-5, 3-6, 3-7, 3-8, 3-9, 3-10, 3-11, 3-12, 3-13, 3-14, 3-15, 3-16, 3-17, 3-18, 3-19, 3-20, 5-6, 5-7, 5-8, 5-9, 5-10, 5-11, 5-12, 5-13, 5-14, 5-15, 5-16, 5-17, 5-18, 5-19, 5-20, 10-15, or 15-20 compartments. In some embodiments, the chip array comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 injection ports. In some embodiments, the chip array comprises 1-2, 1-3, 1-4, 1-5, 1-6, 1-7, 1-8, 1-9, 1-10, 1-11, 1-12, 1-13, 1-14, 1-15, 1-16, 1-17, 1-18, 1-19, 1-20, 2-3, 2-4, 2-5, 2-6, 2-7, 2-8, 2-9, 2-10, 2-11, 2-12, 2-13, 2-14, 2-15, 2-16, 2-17, 2-18, 2-19, 2-20, 3-4, 3-5, 3-6, 3-7, 3-8, 3-9, 3-10, 3-11, 3-12, 3-13, 3-14, 3-15, 3-16, 3-17, 3-18, 3-19, 3-20, 5-6, 5-7, 5-8, 5-9, 5-10, 5-11, 5-12, 5-13, 5-14, 5-15, 5-16, 5-17, 5-18, 5-19, 5-20, 10-15, or 15-20 injection ports.

In some embodiments, the chip array comprises a plurality of physically separated spots (or regions) comprising immobilized (*e.g.*, covalently-attached) detector molecules, as described herein. For example, in some embodiments, the chip array comprises at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at least 17, at least 18, at least 19, at least 20, at least 25, at least 30, at least 35, at least 40, at least 45, at least 50, at least 55, at least 60, at least 65, at least 70, at least 75, at least 80, at least 85, at least 90, at least 95, at least 100, at least 150, at least 200, at least 250, at least 300, at least 400, at least 450, at least 500, at least 550, at least 600, at least 700, at least 800, at least 900, at least 1000, at least 5000, or at least 10,000 physically separated spots. In some embodiments, a chip array comprises 2-10, 2-20, 2-30, 2-40, 2-50, 2-60, 2-70, 2-80, 2-90, 2-100, 10-20, 10-30, 10-40, 10-50, 10-60, 10-70, 10-80, 10-90, 10-100, 50-100, 50-150, 50-200, 50-250, 50-300, 50-350, 50-400, 50-450, 50-500, 50-550, 50-600, 50-650, 50-700, 50-750, 50-800, 50-850, 50-900, 50-950, 50-1000, 500-1000, 500-2000, 500-3000, 500-4000, 500-5000, 500-6000, 500-7000, 500-8000, 500-9000, or 500-10,000 physically separated spots.

B. Methods of Determining the Origin of a Barcoded Molecule in a Multiplexed Sample

In some aspects, the disclosure relates to methods of determining the origin(s) of a barcoded molecule(s) (*e.g.*, polypeptides, polynucleic acids (such as DNA, RNA, cDNA, etc.) metabolites, etc.) in a multiplexed sample. The origin of a barcoded molecule (or origins of a plurality of barcoded molecules) is determined through the identification of the barcode(s) of the molecule(s). Barcode identities may be detected by sequencing (*e.g.*, polypeptide and/or polynucleic acid sequencing), luminescence, hybridization, binding kinetics, physical location on or within a solid substrate, or a combination thereof.

In some embodiments, a barcoded molecule (*i.e.*, a barcoded polypeptide or a barcoded polynucleic acid) or plurality of barcoded molecules of a multiplexed sample may be sequenced (*e.g.*, sequenced in parallel) to determine the sequence(s) of the molecule(s). In such embodiments, the origin(s) of the barcoded molecule(s) may be determined before, after, or concurrently with the sequencing of the molecule(s) of the multiplexed sample. In some embodiments, the origin(s) of the barcoded molecule(s) is determined before the sequencing of the molecule(s). In some embodiments, the origin(s) of the barcoded molecule(s) is determined after the sequencing of the molecule(s). In some embodiments, the origin(s) of the barcoded molecule(s) is determined concurrently with the sequencing of the molecule(s). In some embodiments, the sequences of barcoded molecules of a multiplexed sample are grouped according to their origins (as determined by their barcode identities).

In some aspects the disclosure relates to methods of sequencing molecules (*e.g.*, a polypeptide or polynucleic acid) and/or detecting/quantifying molecules (*e.g.*, a polypeptide, polynucleic acid, or metabolite). Many methods of sequencing and detecting/quantifying molecules are known to those having ordinary skill in the art. In addition, previously undescribed methods of sequencing molecules are described herein. See “Sequencing Methodologies”.

(i) *Detector Molecules*

In some embodiment, a method of determining the origin of a barcoded molecule (or the origins of a plurality of barcoded molecules) comprises detecting the barcode identity of the molecule (or barcode identities of the barcoded molecules) indirectly using detector molecules. For example, in some embodiments, barcode identity is detected in a method comprising: (i) contacting a barcoded molecule (or plurality of barcoded molecules) with a plurality of detector molecules, wherein one or more of the detector molecules in the plurality interacts with the barcode of the barcoded molecule (or interacts with one or more barcode of the barcoded molecules); and (ii) detecting any interaction between a barcoded molecule and a detector

molecule. An interaction between a barcoded molecule and a detector molecule may be identified through luminescence, hybridization, binding kinetics, or physical location. Detector molecules may also be used to quantify barcoded molecules.

In some embodiments, each of the detector molecules of the plurality of detector molecules are chemically identical. In some embodiments, a plurality of detector molecules comprises two or more chemically distinct detector molecules.

For example, in some embodiments, a plurality of detector molecules comprises 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 chemically distinct detector molecules.

In some embodiments, a plurality of detector molecules comprises at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at least 17, at least 18, at least 19, at least 20, at least 25, at least 30, at least 35, at least 40, at least 45, at least 50, at least 60, at least 70, at least 80, at least 90, at least 100, at least 200, at least 300, at least 400, at least 500, at least 600, at least 700, at least 800, at least 900, or at least 1000 chemically distinct detector molecules.

In some embodiments, a plurality of detector molecules comprises 2-3, 2-4, 2-5, 2-6, 2-7, 2-8, 2-9, 2-10, 2-11, 2-12, 2-13, 2-14, 2-15, 2-16, 2-17, 2-18, 2-19, 2-20, 2-25, 2-30, 2-35, 2-40, 2-45, 2-50, 2-60, 2-70, 2-80, 2-90, 2-100, 2-200, 2-300, 2-400, 2-500, 2-600, 2-700, 2-800, 2-900, 2-1000, 5-10, 5-15, 5-20, 5-25, 5-30, 5-35, 5-40, 5-45, 5-50, 5-60, 5-70, 5-80, 5-90, 5-100, 5-200, 5-300, 5-400, 5-500, 5-600, 5-700, 5-800, 5-900, 10-15, 10-20, 10-25, 10-30, 10-35, 10-40, 10-45, 10-50, 10-60, 10-70, 10-80, 10-90, 10-100, 10-200, 10-300, 10-400, 10-500, 10-600, 10-700, 10-800, 10-900, 10-1000, 20-30, 20-40, 20-50, 20-60, 20-70, 20-80, 20-90, 20-100, 20-200, 20-300, 20-400, 20-500, 20-600, 20-700, 20-800, 20-900, 20-1000, 50-60, 50-70, 50-80, 50-90, 50-100, 50-200, 50-300, 50-400, 50-500, 50-600, 50-700, 50-800, 50-900, 50-1000, 100-200, 100-300, 100-400, 100-500, 100-600, 100-700, 100-800, 100-900, 100-1000, 500-600, 500-700, 1500-800, 500-900, or 500-1000 chemically distinct detector molecules.

A detector molecule may comprise a polynucleic acid portion, a polypeptide portion, a small molecule portion, or a combination thereof.

In some embodiments, a detector molecule comprises a polynucleic acid portion. In some embodiments, a detector molecule comprises two or more polynucleic acid portions. In some embodiments wherein a detector molecule comprises multiple polynucleic acid portions: each polynucleic acid portion may be identical; a subset of the polynucleic acid portions may be identical; or each polynucleic acid portion may be chemically distinct.

In some embodiment, the polynucleic acid portion is 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39,

40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, or 60 nucleotides in length.

In some embodiment, the polynucleic acid portion is at least 5, at least 10, at least 15, at least 20, at least 25, at least 30, at least 40, at least 50, at least 60, at least 70, at least 80, at least 90, at least 100, at least 150, at least 200, at least 250, at least 300, at least 350, at least 400, at least 450, or at least 500 nucleotides in length.

In some embodiments, the polynucleic acid portion is 5-10, 5-15, 5-20, 5-25, 5-30, 5-40, 5-50, 5-60, 5-70, 5-80, 5-90, 5-100, 5-150, 5-200, 5-250, 5-300, 5-350, 5-400, 5-450, 5-500, 10-15, 10-20, 10-25, 10-30, 10-40, 10-50, 10-60, 10-70, 10-80, 10-90, 10-100, 10-150, 10-200, 10-250, 10-300, 10-350, 10-400, 10-450, 10-500, 20-30, 20-40, 20-50, 20-60, 20-70, 20-80, 20-90, 20-100, 20-150, 20-200, 20-250, 20-300, 20-350, 20-400, 20-450, 20-500, 50-75, 50-100, 50-150, 50-200, 50-250, 50-500, 50-350, 50-400, 50-450, 50-500, 100-200, 100-250, 100-500, 100-350, 100-400, 100-450, or 100-500 nucleotides in length.

In some embodiment, the polynucleic acid portion is an aptamer.

In some embodiments, a detector molecule comprises a polypeptide portion. In some embodiments, a detector molecule comprises two or more polypeptide portions. In embodiments wherein a detector molecule comprises multiple polypeptide portions: each polypeptide portion may be identical; a subset of the polypeptide portions may be identical; or each polypeptide portion may be chemically distinct.

In some embodiment, the polypeptide portion is 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 amino acids in length.

In some embodiments, the polypeptide portion is at least 5, at least 10, at least 15, at least 20, at least 25, at least 30, at least 40, at least 50, at least 60, at least 70, at least 80, at least 90, at least 100, at least 150, at least 200, at least 250, at least 300, at least 350, at least 400, at least 450, or at least 500 amino acids in length.

In some embodiments, the polypeptide portion is 5-10, 5-15, 5-20, 5-25, 5-30, 5-40, 5-50, 5-60, 5-70, 5-80, 5-90, 5-100, 5-150, 5-200, 5-250, 5-300, 5-350, 5-400, 5-450, 5-500, 10-15, 10-20, 10-25, 10-30, 10-40, 10-50, 10-60, 10-70, 10-80, 10-90, 10-100, 10-150, 10-200, 10-250, 10-300, 10-350, 10-400, 10-450, 10-500, 20-30, 20-40, 20-50, 20-60, 20-70, 20-80, 20-90, 20-100, 20-150, 20-200, 20-250, 20-300, 20-350, 20-400, 20-450, 20-500, 50-75, 50-100, 50-150, 50-200, 50-250, 50-500, 50-350, 50-400, 50-450, 50-500, 100-200, 100-250, 100-500, 100-350, 100-400, 100-450, or 100-500 amino acids in length.

In some embodiments, the polypeptide portion is an aptamer. In some embodiment, the polypeptide portion is an antibody. In some embodiment, the polypeptide portion is an antigen.

In some embodiments, the polypeptide portion is streptavidin.

In some embodiments, a detector molecule comprises a small molecule portion, such as a drug portion or a luminescent molecule portion (of fluorescent molecule portion). In some embodiments, a detector molecule comprises two or more small molecule portions. In embodiments wherein a detector molecule comprises multiple small molecule portions: each small molecule portion may be identical; a subset of the small molecule portions may be identical; or each small molecule portion may be chemically distinct.

Examples of drugs and luminescent molecules suitable for the methods described herein are known to those having skill in the art. As used herein, a luminescent molecule is a molecule that absorbs one or more photons and may subsequently emit one or more photons after one or more time durations.

In some embodiments, a luminescent molecule may comprise a first and second chromophore. In some embodiments, an excited state of the first chromophore is capable of relaxation via an energy transfer to the second chromophore. In some embodiments, the energy transfer is a Förster resonance energy transfer (FRET). Such a FRET pair may be useful for providing a luminescent label with properties that make the label easier to differentiate from amongst a plurality of luminescent labels in a mixture. In yet other embodiments, a FRET pair comprises a first chromophore of a first luminescent label and a second chromophore of a second luminescent label. In certain embodiments, the FRET pair may absorb excitation energy in a first spectral range and emit luminescence in a second spectral range.

In some embodiments, a luminescent molecule refers to a fluorophore or a dye. Typically, a luminescent molecule comprises an aromatic or heteroaromatic compound and can be a pyrene, anthracene, naphthalene, naphthylamine, acridine, stilbene, indole, benzindole, oxazole, carbazole, thiazole, benzothiazole, benzoxazole, phenanthridine, phenoxazine, porphyrin, quinoline, ethidium, benzamide, cyanine, carbocyanine, salicylate, anthranilate, coumarin, fluorescein, rhodamine, xanthene, or other like compound.

In some embodiments, a luminescent molecule comprises a dye selected from one or more of the following: 5/6-Carboxyrhodamine 6G, 5-Carboxyrhodamine 6G, 6-Carboxyrhodamine 6G, 6-TAMRA, Abberior® STAR 440SXP, Abberior® STAR 470SXP, Abberior® STAR 488, Abberior® STAR 512, Abberior® STAR 520SXP, Abberior® STAR 580, Abberior® STAR 600, Abberior® STAR 635, Abberior® STAR 635P, Abberior® STAR RED, Alexa Fluor® 350, Alexa Fluor® 405, Alexa Fluor® 430, Alexa Fluor® 480, Alexa Fluor® 488, Alexa Fluor® 514, Alexa Fluor® 532, Alexa Fluor® 546, Alexa Fluor® 555, Alexa Fluor® 568, Alexa Fluor® 594, Alexa Fluor® 610-X, Alexa Fluor® 633, Alexa Fluor® 647, Alexa Fluor® 660, Alexa Fluor® 680, Alexa Fluor® 700, Alexa Fluor® 750, Alexa Fluor® 790, AMCA, ATTO 390, ATTO 425, ATTO 465, ATTO 488, ATTO 495, ATTO 514, ATTO 520,

ATTO 532, ATTO 542, ATTO 550, ATTO 565, ATTO 590, ATTO 610, ATTO 620, ATTO 633, ATTO 647, ATTO 647N, ATTO 655, ATTO 665, ATTO 680, ATTO 700, ATTO 725, ATTO 740, ATTO Oxa12, ATTO Rho101, ATTO Rho11, ATTO Rho12, ATTO Rho13, ATTO Rho14, ATTO Rho3B, ATTO Rho6G, ATTO Thio12, BD Horizon™ V450, BODIPY® 493/501, BODIPY® 530/550, BODIPY® 558/568, BODIPY® 564/570, BODIPY® 576/589, BODIPY® 581/591, BODIPY® 630/650, BODIPY® 650/665, BODIPY® FL, BODIPY® FL-X, BODIPY® R6G, BODIPY® TMR, BODIPY® TR, CAL Fluor® Gold 540, CAL Fluor® Green 510, CAL Fluor® Orange 560, CAL Fluor® Red 590, CAL Fluor® Red 610, CAL Fluor® Red 615, CAL Fluor® Red 635, Cascade® Blue, CF™350, CF™405M, CF™405S, CF™488A, CF™514, CF™532, CF™543, CF™546, CF™555, CF™568, CF™594, CF™620R, CF™633, CF™633-V1, CF™640R, CF™640R-V1, CF™640R-V2, CF™660C, CF™660R, CF™680, CF™680R, CF™680R-V1, CF™750, CF™770, CF™790, Chromeo™ 642, Chromis 425N, Chromis 500N, Chromis 515N, Chromis 530N, Chromis 550A, Chromis 550C, Chromis 550Z, Chromis 560N, Chromis 570N, Chromis 577N, Chromis 600N, Chromis 630N, Chromis 645A, Chromis 645C, Chromis 645Z, Chromis 678A, Chromis 678C, Chromis 678Z, Chromis 770A, Chromis 770C, Chromis 800A, Chromis 800C, Chromis 830A, Chromis 830C, Cy®3, Cy®3.5, Cy®3B, Cy®5, Cy®5.5, Cy®7, DyLight® 350, DyLight® 405, DyLight® 415-Co1, DyLight® 425Q, DyLight® 485-LS, DyLight® 488, DyLight® 504Q, DyLight® 510-LS, DyLight® 515-LS, DyLight® 521-LS, DyLight® 530-R2, DyLight® 543Q, DyLight® 550, DyLight® 554-R0, DyLight® 554-R1, DyLight® 590-R2, DyLight® 594, DyLight® 610-B1, DyLight® 615-B2, DyLight® 633, DyLight® 633-B1, DyLight® 633-B2, DyLight® 650, DyLight® 655-B1, DyLight® 655-B2, DyLight® 655-B3, DyLight® 655-B4, DyLight® 662Q, DyLight® 675-B1, DyLight® 675-B2, DyLight® 675-B3, DyLight® 675-B4, DyLight® 679-C5, DyLight® 680, DyLight® 683Q, DyLight® 690-B1, DyLight® 690-B2, DyLight® 696Q, DyLight® 700-B1, DyLight® 700-B1, DyLight® 730-B1, DyLight® 730-B2, DyLight® 730-B3, DyLight® 730-B4, DyLight® 747, DyLight® 747-B1, DyLight® 747-B2, DyLight® 747-B3, DyLight® 747-B4, DyLight® 755, DyLight® 766Q, DyLight® 775-B2, DyLight® 775-B3, DyLight® 775-B4, DyLight® 780-B1, DyLight® 780-B2, DyLight® 780-B3, DyLight® 800, DyLight® 830-B2, Dyomics-350, Dyomics-350XL, Dyomics-360XL, Dyomics-370XL, Dyomics-375XL, Dyomics-380XL, Dyomics-390XL, Dyomics-405, Dyomics-415, Dyomics-430, Dyomics-431, Dyomics-478, Dyomics-480XL, Dyomics-481XL, Dyomics-485XL, Dyomics-490, Dyomics-495, Dyomics-505, Dyomics-510XL, Dyomics-511XL, Dyomics-520XL, Dyomics-521XL, Dyomics-530, Dyomics-547, Dyomics-547P1, Dyomics-548, Dyomics-549, Dyomics-549P1, Dyomics-550, Dyomics-554, Dyomics-555, Dyomics-556, Dyomics-560, Dyomics-590, Dyomics-591, Dyomics-594, Dyomics-601XL, Dyomics-605,

Dyomics-610, Dyomics-615, Dyomics-630, Dyomics-631, Dyomics-632, Dyomics-633, Dyomics-634, Dyomics-635, Dyomics-636, Dyomics-647, Dyomics-647P1, Dyomics-648, Dyomics-648P1, Dyomics-649, Dyomics-649P1, Dyomics-650, Dyomics-651, Dyomics-652, Dyomics-654, Dyomics-675, Dyomics-676, Dyomics-677, Dyomics-678, Dyomics-679P1, Dyomics-680, Dyomics-681, Dyomics-682, Dyomics-700, Dyomics-701, Dyomics-703, Dyomics-704, Dyomics-730, Dyomics-731, Dyomics-732, Dyomics-734, Dyomics-749, Dyomics-749P1, Dyomics-750, Dyomics-751, Dyomics-752, Dyomics-754, Dyomics-776, Dyomics-777, Dyomics-778, Dyomics-780, Dyomics-781, Dyomics-782, Dyomics-800, Dyomics-831, eFluor® 450, Eosin, FITC, Fluorescein, HiLyte™ Fluor 405, HiLyte™ Fluor 488, HiLyte™ Fluor 532, HiLyte™ Fluor 555, HiLyte™ Fluor 594, HiLyte™ Fluor 647, HiLyte™ Fluor 680, HiLyte™ Fluor 750, IRDye® 680LT, IRDye® 750, IRDye® 800CW, JOE, LightCycler® 640R, LightCycler® Red 610, LightCycler® Red 640, LightCycler® Red 670, LightCycler® Red 705, Lissamine Rhodamine B, Naphthofluorescein, Oregon Green® 488, Oregon Green® 514, Pacific Blue™, Pacific Green™, Pacific Orange™, PET, PF350, PF405, PF415, PF488, PF505, PF532, PF546, PF555P, PF568, PF594, PF610, PF633P, PF647P, Quasar® 570, Quasar® 670, Quasar® 705, Rhodamine 123, Rhodamine 6G, Rhodamine B, Rhodamine Green, Rhodamine Green-X, Rhodamine Red, ROX, Seta™ 375, Seta™ 470, Seta™ 555, Seta™ 632, Seta™ 633, Seta™ 650, Seta™ 660, Seta™ 670, Seta™ 680, Seta™ 700, Seta™ 750, Seta™ 780, Seta™ APC-780, Seta™ PerCP-680, Seta™ R-PE-670, Seta™ 646, SeTau 380, SeTau 425, SeTau 647, SeTau 405, Square 635, Square 650, Square 660, Square 672, Square 680, Sulforhodamine 101, TAMRA, TET, Texas Red®, TMR, TRITC, Yakima Yellow™, Zenon®, Zy3, Zy5, Zy5.5, and Zy7.

In some embodiments, a detector molecule is bound (e.g., covalently bound) to a substrate. The substrate may be a surface (e.g., a solid surface), a bead (e.g., a magnetic bead), a particle (e.g., a magnetic particle), or a gel.

(ii) *Luminescence*

In some embodiment, a method of determining the origin of a barcoded molecule (or the origins of a plurality of barcoded molecules) comprises detecting the barcode identity of the molecule (or plurality of barcoded molecules) by luminescence. Detection of barcode identity may be direct or indirect (e.g., by detecting luminescence of a detector molecule).

In some embodiments, barcode identity is identified based on luminescence lifetime, luminescence intensity, brightness, absorption spectra, emission spectra, luminescence quantum yield, or a combination of two or more thereof. In some embodiments, a plurality of barcode identities can be distinguished from each other based on different luminescence lifetimes,

luminescence intensities, brightnesses, absorption spectra, emission spectra, luminescence quantum yields, or combinations of two or more thereof.

In some embodiments, luminescence is detected by exposing a luminescent molecule to a series of separate light pulses and evaluating the timing or other properties of each photon that is emitted from the molecule. In some embodiments, a luminescence lifetime of a molecule is determined from a plurality of photons that are emitted sequentially from the molecule, and the luminescence lifetime can be used to identify the molecule. In some embodiments, a luminescence intensity of a molecule is determined from a plurality of photons that are emitted sequentially from the molecule, and the luminescence intensity can be used to identify the molecule. In some embodiments, a luminescence lifetime and luminescence intensity of a molecule is determined from a plurality of photons that are emitted sequentially from the molecule, and the luminescence lifetime and luminescence intensity can be used to identify the molecule.

In certain embodiments, a luminescent molecule absorbs one photon and emits one photon after a time duration. In some embodiments, the luminescence lifetime of a molecule can be determined or estimated by measuring the time duration. In some embodiments, the luminescence lifetime of a molecule can be determined or estimated by measuring a plurality of time durations for multiple pulse events and emission events. In some embodiments, the luminescence lifetime of a molecule can be differentiated amongst the luminescence lifetimes of a plurality of types of molecules by measuring the time duration. In some embodiments, the luminescence lifetime of a molecule can be differentiated amongst the luminescence lifetimes of a plurality of types of molecules by measuring a plurality of time durations for multiple pulse events and emission events. In certain embodiments, a molecule is identified or differentiated amongst a plurality of types of labels by determining or estimating the luminescence lifetime of the label. In certain embodiments, a molecule is identified or differentiated amongst a plurality of types of molecules by differentiating the luminescence lifetime of the molecule amongst a plurality of the luminescence lifetimes of a plurality of types of molecules.

Determination of a luminescence lifetime of a luminescent molecule can be performed using any suitable method (*e.g.*, by measuring the lifetime using a suitable technique or by determining time-dependent characteristics of emission). In some embodiments, determining the luminescence lifetime of a molecule comprises determining the lifetime relative to another label. In some embodiments, determining the luminescence lifetime of a molecule comprises determining the lifetime relative to a reference. In some embodiments, determining the luminescence lifetime of a molecule comprises measuring the lifetime (*e.g.*, fluorescence lifetime). In some embodiments, determining the luminescence lifetime of a molecule comprises

determining one or more temporal characteristics that are indicative of lifetime. In some embodiments, the luminescence lifetime of a molecule can be determined based on a distribution of a plurality of emission events (*e.g.*, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 30, 40, 50, 60, 70, 80, 90, 100, or more emission events) occurring across one or more time-gated windows relative to an excitation pulse. For example, a luminescence lifetime of a molecule can be distinguished from a plurality of molecules having different luminescence lifetimes based on the distribution of photon arrival times measured with respect to an excitation pulse.

It should be appreciated that a luminescence lifetime of a luminescent molecule is indicative of the timing of photons emitted after the label reaches an excited state and the label can be distinguished by information indicative of the timing of the photons. Some embodiments may include distinguishing a molecule from a plurality of molecules based on the luminescence lifetime of the label by measuring times associated with photons emitted by the molecule. The distribution of times may provide an indication of the luminescence lifetime which may be determined from the distribution. In some embodiments, the molecule is distinguishable from the plurality of molecules based on the distribution of times, such as by comparing the distribution of times to a reference distribution corresponding to a known molecule. In some embodiments, a value for the luminescence lifetime is determined from the distribution of times.

As used herein, in some embodiments, luminescence intensity refers to the number of emitted photons per unit time that are emitted by a luminescent molecule which is being excited by delivery of a pulsed excitation energy. In some embodiments, the luminescence intensity refers to the detected number of emitted photons per unit time that are emitted by a molecule which is being excited by delivery of a pulsed excitation energy, and are detected by a particular sensor or set of sensors.

As used herein, in some embodiments, brightness refers to a parameter that reports on the average emission intensity per luminescent molecule. Thus, in some embodiments, “emission intensity” may be used to generally refer to brightness of a composition comprising one or more molecules. In some embodiments, brightness of a molecule is equal to the product of its quantum yield and extinction coefficient.

As used herein, in some embodiments, luminescence quantum yield refers to the fraction of excitation events at a given wavelength or within a given spectral range that lead to an emission event, and is typically less than 1. In some embodiments, the luminescence quantum yield of a luminescent label described herein is between 0 and about 0.001, between about 0.001 and about 0.01, between about 0.01 and about 0.1, between about 0.1 and about 0.5, between

about 0.5 and 0.9, or between about 0.9 and 1. In some embodiments, a molecule is identified by determining or estimating the luminescence quantum yield.

As used herein, in some embodiments, an excitation energy is a pulse of light from a light source. In some embodiments, an excitation energy is in the visible spectrum. In some
5 embodiments, an excitation energy is in the ultraviolet spectrum. In some embodiments, an excitation energy is in the infrared spectrum. In some embodiments, an excitation energy is at or near the absorption maximum of a luminescent label from which a plurality of emitted photons are to be detected. In certain embodiments, the excitation energy is between about 500 nm and about 700 nm (*e.g.*, between about 500 nm and about 600 nm, between about 600 nm and about
10 700 nm, between about 500 nm and about 550 nm, between about 550 nm and about 600 nm, between about 600 nm and about 650 nm, or between about 650 nm and about 700 nm). In certain embodiments, an excitation energy may be monochromatic or confined to a spectral range. In some embodiments, a spectral range has a range of between about 0.1 nm and about 1 nm, between about 1 nm and about 2 nm, or between about 2 nm and about 5 nm. In some
15 embodiments, a spectral range has a range of between about 5 nm and about 10 nm, between about 10 nm and about 50 nm, or between about 50 nm and about 100 nm.

(iii) *Physical Separation*

In some embodiment, a method of determining the origin of a barcoded molecule (or the
20 origins of a plurality of barcoded molecules) comprises detecting the barcode identity of the molecule (or plurality of barcoded molecules) by physical separation. Detection of barcode identity by physical separation may comprise determining the location of a barcoded molecule on a substrate (*e.g.*, a microarray chip).

For example, a substrate may comprise a plurality of detector molecules (as described
25 herein) that are organized at discrete locations on the substrate. In such instances, barcoded molecules comprising a barcode that hybridizes to, binds to, or is bound by a detector molecule on the substrate can be positioned at the location of the detector molecule. As such, in some embodiments, a method of determining the origin of a barcoded molecule (or the origins of a plurality of barcoded molecules) comprises contacting the polypeptide (or plurality of
30 polypeptides) with a substrate comprising a plurality of detector molecules.

As described above, in some embodiments, a molecule (or plurality of molecules) is barcoded by depositing the molecule (or plurality of molecules) on or within a solid substrate such that the molecule (or plurality of molecules) remains physically separated from additional molecules (or additional pluralities of molecules). In such embodiments, a method of
35 determining the origin of a barcoded molecule (or the origins of a plurality of barcoded

molecules) comprises detecting the location of the barcoded molecule (or the plurality of barcoded molecules) on the solid substrate.

C. Exemplary Embodiments

5 In some embodiments, a barcode molecule comprises a polynucleic acid portion, which is identified by DNA sequencing.

In some embodiments, a barcode molecule comprises a polynucleic acid portion, which is identified via hybridization using a detector molecule comprising a polynucleic acid portion. In some embodiments, the detector molecule further comprises a luminescent molecule portion. In
10 some embodiments, the detector molecule is immobilized on (*e.g.*, covalently attached to) a substrate.

In some embodiments, a barcode molecule comprises a polynucleic acid portion, which is identified via hybridization using a detector molecule comprising a polypeptide portion (*e.g.*, a DNA binding protein, an aptamer, etc.). In some embodiments, the detector molecule further
15 comprises a luminescent molecule portion. In some embodiments, the detector molecule is immobilized on (*e.g.*, covalently attached to) a substrate.

In some embodiments, a barcode molecule comprises a polypeptide portion (*e.g.*, a short polypeptide tag), which is identified by polypeptide sequencing.

In some embodiments, a barcode molecule comprises a polypeptide portion (*e.g.*, a DNA
20 binding protein, or portion thereof), which is identified using a detector molecule comprising a polynucleic acid portion (*e.g.*, a polynucleic acid sequence bound by the DNA binding protein, or portion thereof). In some embodiments, the detector molecule further comprises a luminescent molecule portion. In some embodiments, the detector molecule is immobilized on (*e.g.*, covalently attached to) a substrate.

25 In some embodiments, a barcode molecule comprises a polypeptide portion, which is identified using a detector molecule comprising a polynucleic acid portion (*e.g.*, an aptamer). In some embodiments, the detector molecule further comprises a luminescent molecule portion. In some embodiments, the detector molecule is immobilized on (*e.g.*, covalently attached to) a substrate.

30 In some embodiments, a barcode molecule comprises an amino acid modification that is made to a polypeptide after it has been translated.

In some embodiments, a barcode molecule comprises a polypeptide portion (*e.g.*, an antibody, antigen, aptamer, etc.), which is identified using a detector molecule comprising a polypeptide portion (*e.g.*, an antigen, antibody, or substrate, etc.). In some embodiments, the

detector molecule further comprises a luminescent molecule portion. In some embodiments, the detector molecule is immobilized on (*e.g.*, covalently attached) to a substrate.

In some embodiments, a barcode component comprise an endoprotease with distinct cutting profiles, which can be detected by polypeptide sequencing.

III. Methods of Preparing an Enriched Sample

In some embodiments, a sample is enriched prior to, concurrently with, subsequent to, or in the absence of barcoding. Accordingly, in some aspects, the disclosure relates to methods of preparing an enriched sample. As used herein, the term “enrichment” refers to a process wherein the abundance of one or more molecule of interest (*e.g.*, polypeptide, polynucleic acid, metabolite, etc.) is increased relative to the abundance of one or more reference molecule (*e.g.*, a molecule in a complex sample that is not of interest). The term “molecule of interest” as used herein, refers to a molecule (*e.g.*, polypeptide, polynucleic acid, metabolite, etc.) that one seeks to enrich.

For example, a polypeptide of interest may comprise a specific amino acid sequence. Alternatively, or in addition, a polypeptide of interest may comprise a specific polypeptide modification (*e.g.*, a post-translational modification). These methods facilitate proteomic analysis of complex samples, which are made up of many different polypeptides, only some of which may be of interest.

In some embodiments, a polynucleic acid of interest may comprise a specific nucleotide sequence. Alternatively, or in addition, a polynucleic acid of interest may comprise a specific nucleotide modification (*e.g.*, a non-natural nucleotide). These methods facilitate genomic analysis of complex samples, which are made up of many different polynucleic acids, only some of which may be of interest.

In some embodiments, a method for enrichment comprises using a plurality of enrichment molecules to select a subset of molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.) from a plurality of molecules, thereby generating an enriched sample comprising the subset of molecules. In some embodiments, the method comprises contacting a plurality of molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.) with a plurality of enrichment molecules to produce an enriched sample comprising a subset of the molecules in the plurality of molecules.

In some embodiments, a method for enrichment comprises: (a) contacting a plurality of molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.) with a plurality of enrichment molecules, wherein at least a subset of the enrichment molecules in the plurality of enrichment molecules binds to a subset of the molecules in the plurality of molecules, thereby generating a

bound subset of molecules and an unbound subset of molecules; and (b) isolating the bound subset of molecules to produce an enriched sample comprising a subset of the molecules in the plurality of molecules.

In some embodiments, a method for enrichment comprises: (a) contacting a plurality of molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.) with a plurality of enrichment molecules, wherein at least a subset of the enrichment molecules in the plurality of enrichment molecules binds to a subset of the molecules in the plurality of molecules, thereby generating a bound subset of molecules and an unbound subset of molecules; and (b) isolating the unbound subset of molecules to produce an enriched sample comprising a subset of the molecules in the plurality of molecules.

In the embodiments described in the preceding paragraphs, it is understood that the binding of an enrichment molecule to a molecule is equivalent to the binding of the molecule to the enrichment molecule. Accordingly, step (a) in the embodiments described above can be equivalently describe as: (a) contacting a plurality of molecules with a plurality of enrichment molecules, wherein at least a subset of the enrichment molecules in the plurality of enrichment molecules is *bound by* a subset of the molecules in the plurality of molecules, thereby generating a bound subset of molecules and an unbound subset of molecules.

It is also understood that steps (a) and (b) of the embodiments described above may be repeated one or more times using additional pluralities of enrichment molecules to produce a further enriched sample. For example, in some embodiments, the method comprises: (a) contacting a plurality of molecules with a first plurality of enrichment molecules, wherein at least a subset of the enrichment molecules in the first plurality of enrichment molecules binds to a subset of the molecules in the plurality of molecules, thereby generating a first bound subset of molecules and a first unbound subset of molecules; (b) isolating the first bound subset of molecules or the first unbound subset of molecules of (a); and (c) iteratively repeating steps (a) and (b) with one or more additional plurality of enrichment molecules to produce an enriched sample comprising a subset of the molecules in the plurality of molecules. In some embodiments, steps (a) and (b) are repeated using a second, third, fourth, fifth, sixth, seventh, eighth, ninth, tenth, or any number of additional plurality of enrichment molecules.

For example, in some embodiments the method comprises: (a) contacting a plurality of molecules with a first plurality of enrichment molecules, wherein at least a subset of the enrichment molecules in the first plurality of enrichment molecules binds to a subset of the molecules in the plurality of molecules, thereby generating a first bound subset of molecules and a first unbound subset of molecules; (b) isolating the first bound subset of molecules or the first unbound subset of molecules of (a); (c) contacting the isolated molecules of (b) with a second

plurality of enrichment molecules, wherein at least a subset of the enrichment molecules in the second plurality of enrichment molecules binds to a subset of the molecules isolated in (b), thereby generating a second bound subset of molecules and a second unbound subset of molecules; (d) isolating the second bound subset of molecules or the second unbound subset of molecules of (c) to produce an enriched sample comprising a subset of the molecules in the plurality of molecules.

Alternatively, or in addition, a method of enrichment may comprise chromatography (*e.g.*, size exclusion, ion exchange, etc.), isoelectric focusing, membrane filtration, molecular sieve filtration, concentration, precipitation (*e.g.*, cryoprecipitation), dry down, dialysis, or a combination thereof.

In some embodiments, the method comprises contacting a complex sample with a kit or device described herein. See “Kits for Sample Preparation” and “Devices for Sample Preparation and Sample Sequencing”.

In some embodiments, the molecules in an enriched sample are chemically identical (*e.g.*, contain the same amino acid sequence or the same nucleotide sequence). In some embodiments, an enriched sample comprises at least two chemically unique molecules (*e.g.*, having differing amino acid sequences or differing nucleic acid sequences). For example, in some embodiments, an enriched sample comprises at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at least 17, at least 18, at least 19, at least 20, at least 25, at least 30, at least 40, at least 50, at least 60, at least 70, at least 80, at least 90, or at least 100 chemically unique molecules. In some embodiments, an enriched sample comprises 1-2, 1-5, 1-10, 1-15, 1-20, 1-30, 1-40, 1-50, 1-60, 1-70, 1-80, 1-90, 1-100, 2-5, 2-10, 2-15, 2-20, 2-30, 2-40, 2-50, 2-60, 2-70, 2-80, 2-90, 2-100, 5-10, 5-15, 5-20, 5-30, 5-40, 5-50, 5-60, 5-70, 5-80, 5-90, 5-100, 10-15, 10-20, 10-30, 10-40, 10-50, 10-60, 10-70, 10-80, 10-90, 10-100, 15-20, 20-30, 20-40, 20-50, 20-60, 20-70, 20-80, 20-90, 20-100, 20-30, 20-40, 20-50, 20-60, 20-70, 20-80, 20-90, 20-100, 30-40, 30-50, 30-60, 30-70, 30-80, 30-90, 30-100, 40-50, 40-60, 40-70, 40-80, 40-90, 40-100, 50-60, 50-70, 50-80, 50-90, or 50-100 chemically unique molecules.

In some embodiments, an enriched sample comprises a polynucleic acid that can be subjected to short-read sequencing applications, long-read sequencing applications, or a hybrid assembly application. In some embodiments, an enriched sample comprises a polynucleic acid having a length of about 0.5-2 kb, 0.5-5 kb, 1-2 kb, 1-3 kb, 1-4 kb, 1-5 kb, 1-10 kb, 2-10 kb, 2-5 kb, 5-10 kb, 5-15 kb, 5-20 kb, 5-25 kb, 10-15 kb, 10-20 kb, or 10-25 kb. In some embodiments, an enriched sample comprises a polynucleic acid comprising at least 700, 800, 900, 1000, 1100, 1200, 1300, 1400, 1500, 1600, 1700, 1800, 1900, 2000, 2100, 2200, 2300, 2400, 2500, 2600,

2700, 2800, 2900, or 3000 nucleotides in length. In some embodiments, an enriched sample comprises a polynucleic acid comprising 700-3000, 1000-3000, 1000-2500, 1000-2400, 1000-2300, 1000-2200, 1000-2100, 1000-2000, 1000-1900, 1000-1800, 1000-1700, 1000-1600, 1000-1500, 1000-1400, 1000-1300, 1000-1200, 1500-3000, 1500-2500, 1500-2000, or 2000-3000 nucleotides in length.

In some embodiments, the enriched sample comprises polypeptides and/or polynucleic acids that share at least 50%, 60%, 70%, 80%, 90%, 95%, or 99% sequence identity. In some embodiments, the enriched sample comprises polypeptides that share one or more polypeptide modification (*e.g.*, post-translational modification). Examples of post-translational modifications are known to those having skill in the art and include, but are not limited to, acetylation, adenylation, ADP-ribosylation, alkylation (*e.g.*, methylation), amidation, arginylation, biotinylation, butyrylation, carbamylation, carbonylation, carboxylation, citrullination, deamidation, eliminylation, formylation, glycosylation (*e.g.*, N-linked glycosylation, O-linked glycosylation), glypyatyon, glycation, hydroxylation, iodination, ISGylation, isoprenylation, lipoylation, malonylation, myristoylation, neddylation, nitration, oxidation, palmitoylation, pegylation, phosphorylation, phosphopantetheinylation, polyglcylation, polyglutaminylation, prenylation, propionylation, pupylation, S-glutathionylation, S-nitrosylation, S-sulfenylation, S-sulfinylation, S-sulfonylation, succinylation, sulfation, SUMOylation, and ubiquitination.

A. Enrichment Molecules

As used herein, the term “enrichment molecule” refers to a molecule that exhibits preferentially binding to (or by) one or more target molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.). An enrichment molecule may bind to (or be bound by) a target molecule directly (*e.g.*, through a direct interaction with the amino acid sequence of a target polypeptide). Alternatively, or in addition, an enrichment molecule may bind to (or be bound by) a target molecule through an interaction with a modification of the target molecule (*e.g.*, through an interaction with a post-translational modification of a target polypeptide). The binding of an enrichment molecule to (or by) a target molecule may be mediated through electrostatic interactions, hydrophobic interactions, complementary shape, or a combination thereof.

In some embodiments, a target molecule is a molecule of interest. In other embodiments, a target molecule is not a molecule of interest.

Exemplary enrichment molecules that preferentially *bind to* one or more target molecules (or target molecule variants) include immunoglobulins, anticalins, lipocalins, DARPins, aptamers, enzymes, lectins, and peptide interaction domains.

As used herein, the term “immunoglobulin” refers to polypeptides characterized as having an immunoglobulin fold and which function as antibodies and bind to one or more substrates (*e.g.*, target molecules). As such, the term “immunoglobulin” encompasses conventional immunoglobulins (*i.e.*, IgA, IgD, IgE, IgG, and IgM), single-chain variable fragments (scFv), antigen-binding fragments (Fab), affibodies, and single domain antibodies (sdAb), such as Nanobodies, VHHs and VNARs.

The term “aptamer” as used herein refers to a polynucleic acid (*e.g.*, DNA or RNA) or polypeptide that preferentially binds to one or more target molecules (*e.g.*, target molecules). Although there are examples found in nature, aptamers are usually engineered through repeated rounds of *in vitro* selection.

As used herein, the term “enzyme” refers to a macromolecular biological catalyst that accelerates a chemical reaction upon binding one or more substrates (*e.g.*, target molecules). Typically, an enzyme will release its substrate after completion of a chemical reaction. As such, in some embodiments, wherein an enrichment molecule comprises an enzyme, the enzyme is catalytically inactivated so as to increase the likelihood that the enzyme remains the substrate. Catalytic inactivation may be performed via mutagenesis and/or depletion of one or more enzymatic cofactor (*i.e.*, a non-protein chemical compound or metallic ion that is required for an enzyme’s activity as a catalyst).

The term “peptide interaction domain” as used herein, refers to a polypeptide (or a portion of a polypeptide) that interacts with one or more polypeptides (*e.g.*, target polypeptides). For example, a peptide interaction domain may be a scaffold protein, a polypeptide of a multiprotein complex, or a portion thereof.

In some embodiments, an enrichment molecule comprises an immunoglobulin, an aptamer, an enzyme, and/or a peptide interaction domain.

Exemplary enrichment molecules that are preferentially *bound* by one or more target molecules include oligonucleotides (*e.g.*, double-stranded DNA, single-stranded DNA, double-stranded RNA, single-stranded RNA, or the like), oligosaccharides (or polysaccharides), lipids, glycoproteins, receptor ligands, receptor agonists, receptor antagonists, enzyme substrates, and enzyme cofactors.

In some embodiments, an enrichment molecule comprises an oligonucleotide (*e.g.*, double-stranded DNA, single-stranded DNA, double-stranded RNA, single-stranded RNA, or the like), an oligosaccharide, a lipid, a receptor ligand, a receptor agonist, a receptor antagonist, an enzyme substrate, and/or an enzyme cofactor.

Preferential binding is used herein to characterize enrichment molecules to emphasize: (i) that an enrichment molecule need not exhibit high specificity (*i.e.*, only bind to (or be bound by)

a single target molecule to an appreciable level); (ii) that an enrichment molecule may exhibit some degree of off-target binding (*i.e.*, bind to (or be bound by) an off-target molecule to a detectable level); and (iii) that an enrichment molecule need not bind to a target molecule with 100% efficiency (*i.e.*, not all target polypeptides in a complex sample need necessarily be bound, even in the presence of excess enrichment molecules).

In some embodiments, an enrichment molecule preferentially binds to (or is preferentially bound by) a single target molecule. However, in other embodiments, an enrichment molecule preferentially binds to (or is preferentially bound by) two or more target molecules.

In some embodiments, an enrichment molecule exhibits preferential binding to (or is preferentially bound by) at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at least 17, at least 18, at least 19, at least 20, at least 25, at least 30, at least 40, at least 50, at least 60, at least 70, at least 80, at least 90, or at least 100, at least 200, at least 300, at least 400, at least 500, at least 600, at least 700, at least 800, at least 900, at least 1000, at least 2000, at least 3000, at least 4000, at least 5000, or at least 10,000 target molecules.

In some embodiments, an enrichment molecule exhibits preferential binding to (or is preferentially bound by) two, three, four, five, six, seven, eight, nine, ten, eleven, twelve, thirteen, fourteen, or fifteen target molecules.

In some embodiments, an enrichment molecule exhibits preferential binding to (or is preferentially bound by) 1-2, 1-5, 1-10, 1-15, 1-20, 1-30, 1-40, 1-50, 1-60, 1-70, 1-80, 1-90, 1-100, 2-5, 2-10, 2-15, 2-20, 2-30, 2-40, 2-50, 2-60, 2-70, 2-80, 2-90, 2-100, 5-10, 5-15, 5-20, 5-30, 5-40, 5-50, 5-60, 5-70, 5-80, 5-90, 5-100, 10-15, 10-20, 10-30, 10-40, 10-50, 10-60, 10-70, 10-80, 10-90, 10-100, 15-20, 20-30, 20-40, 20-50, 20-60, 20-70, 20-80, 20-90, 20-100, 20-30, 20-40, 20-50, 20-60, 20-70, 20-80, 20-90, 20-100, 30-40, 30-50, 30-60, 30-70, 30-80, 30-90, 30-100, 40-50, 40-60, 40-70, 40-80, 40-90, 40-100, 50-60, 50-70, 50-80, 50-90, or 50-100, 100-200, 100-300, 100-400, 100-500, 100-600, 100-700, 100-800, 100-900, 100-1000, 100-5000, 100-10,000, 500-600, 500-700, 500-800, 500-900, 500-1000, 500-5000, 500-10,000, 1000-5000, or 1000-10,000 target molecules.

In some embodiments, an enrichment molecule exhibits preferential binding to (or is preferentially bound by) a plurality of related target molecules (*e.g.*, 2, 3, 4, 5, 6, 7, 8, 9, 10, 20, 30, 40, 50, or more related molecules) that share at least 50%, 60%, 70%, 80%, 90%, 95%, or 99% sequence homology.

In some embodiments, an enrichment molecule exhibits preferential binding to (or is preferentially bound by) a post-translational modification, such as acetylation, adenylation, ADP-ribosylation, alkylation (*e.g.*, methylation), amidation, arginylation, biotinylation,

butyrylation, carbamylation, carbonylation, carboxylation, citrullination, deamidation, eliminination, formylation, glycosylation (e.g., N-linked glycosylation, O-linked glycosylation), glypiation, glycation, hydroxylation, iodination, ISGylation, isoprenylation, lipoylation, malonylation, myristoylation, neddylation, nitration, oxidation, palmitoylation, pegylation, phosphorylation, phosphopantetheinylation, polyglutamylation, polyglutamylation, prenylation, propionylation, pupylation, S-glutathionylation, S-nitrosylation, S-sulfenylation, S-sulfinylation, S-sulfonylation, succinylation, sulfation, SUMOylation, and ubiquitination

An enrichment molecule may be immobilized on (e.g., covalently attached to) a substrate (e.g., a capture probe as described in “Devices for Sample Preparation and Sample Sequencing”).

The substrate may be a surface (e.g., a solid surface), a bead (e.g., a magnetic bead), a particle (e.g., a magnetic particle), or a gel.

(i) *Pluralities of Enrichment Molecules*

Typically, the enrichment methods described herein utilize a plurality of enrichment molecules. The enrichment molecules in a plurality may be chemically identical (i.e., a plurality having one enrichment molecule “type”). Alternatively, pluralities of enrichment molecules may contain a combination of different enrichment molecules (i.e., have two or more enrichment molecule “types”).

In some embodiments, a plurality of enrichment molecules contains a single enrichment molecule type. In other embodiments, a plurality of enrichment molecules comprises a combination of two or more, three or more, four or more, five or more, six or more, seven or more, eight or more, nine or more, ten or more, eleven or more, twelve or more, thirteen or more, fourteen or more, or fifteen or more enrichment molecule types. In some embodiments, a plurality of enrichment molecules comprises at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at least 17, at least 18, at least 19, at least 20, at least 25, at least 30, at least 40, at least 50, at least 60, at least 70, at least 80, at least 90, or at least 100, at least 200, at least 300, at least 400, at least 500 enrichment molecule types.

In some embodiments, a plurality of enrichment molecules comprises a combination of two, three, four, five, six, seven, eight, nine, ten, eleven, twelve, thirteen, fourteen, or fifteen enrichment molecule types.

In some embodiments, a plurality of enrichment molecules contains a combination of 1-2, 1-5, 1-10, 1-15, 1-20, 1-30, 1-40, 1-50, 1-60, 1-70, 1-80, 1-90, 1-100, 2-5, 2-10, 2-15, 2-20, 2-30, 2-40, 2-50, 2-60, 2-70, 2-80, 2-90, 2-100, 5-10, 5-15, 5-20, 5-30, 5-40, 5-50, 5-60, 5-70, 5-80, 5-90, 5-100, 10-15, 10-20, 10-30, 10-40, 10-50, 10-60, 10-70, 10-80, 10-90, 10-100, 15-20, 20-30,

20-40, 20-50, 20-60, 20-70, 20-80, 20-90, 20-100, 20-30, 20-40, 20-50, 20-60, 20-70, 20-80, 20-90, 20-100, 30-40, 30-50, 30-60, 30-70, 30-80, 30-90, 30-100, 40-50, 40-60, 40-70, 40-80, 40-90, 40-100, 50-60, 50-70, 50-80, 50-90, or 50-100, 100-200, 100-300, 100-400, or 100-500 enrichment molecule types.

5 In some embodiments, each of the enrichment molecules in the plurality of enrichment molecules preferentially binds to (or is preferentially bound by) a single target molecule. In other embodiments, one or more (*e.g.*, a subset) of the enrichment molecules in a plurality of enrichment molecules exhibits preferential binding to (or is preferentially bound by) two or more target molecules. In yet other embodiments, each of the enrichment molecules in the plurality of enrichment molecules exhibits preferential binding to (or is preferentially bound by) two or more target molecules.

10 In some embodiments, one or more (*e.g.*, a subset) of the enrichment molecules in the plurality of enrichment molecules binds to a post-translational polypeptide modification. In other embodiments, each of the enrichment molecules in a plurality of enrichment molecules exhibits preferential binding to two or more post-translational polypeptide modifications.

15 In some embodiments, each of the enrichment molecules in the plurality of enrichment molecules is immobilized on (*e.g.*, covalently attached to) a substrate (*e.g.*, a capture probe as described in “Devices for Sample Preparation and Sample Sequencing”), such as a surface (*e.g.*, a solid surface), a bead (*e.g.*, a magnetic bead), a particle (*e.g.*, a magnetic particle, or a gel). In some embodiments, one or more (*e.g.*, a subset) of the plurality of enrichment molecules is immobilized on (*e.g.*, covalently attached to) a substrate. As such, in some embodiments, the contacting of the plurality of molecules with the plurality of enrichment molecules occurs when a sample comprising the plurality of molecules contacts the substrate.

25 For example, in some embodiments, the enrichment molecules are immobilized on (*e.g.*, covalently attached or crosslinked to) a gel and the sample is pulled through the gel. In some embodiments, the enrichment molecules are immobilized on (*e.g.*, covalently attached to) a bead (*e.g.*, a magnetic bead), which are then pulled down.

(ii) *Multiple Enrichment Molecule Pluralities*

30 As described above, in some embodiments, the method comprises: (a) contacting a plurality of molecules with a first plurality of enrichment molecules, wherein at least a subset of the enrichment molecules in the first plurality of enrichment molecules binds to a subset of the molecules in the plurality of molecules, thereby generating a first bound subset of molecules and a first unbound subset of molecules; (b) isolating the first bound subset of molecules or the first unbound subset of molecules of (a); and (c) iteratively repeating steps (a) and (b) with one or

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more additional plurality of enrichment molecules to produce an enriched sample comprising a subset of the molecules in the plurality of molecules. In some embodiments, steps (a) and (b) are repeated using a second, third, fourth, fifth, sixth, seventh, eighth, ninth, tenth, or any number of additional plurality of enrichment molecules.

5 In some embodiments, each plurality of enrichment molecules utilized in the method of enrichment is unique (*i.e.*, each comprises a different plurality of enrichment molecules). In other embodiments, two or more of the pluralities are identical. In some embodiments, at least one of the pluralities of enrichment molecules targets a post-translational polypeptide modification and at least one of the pluralities of enrichment molecules does not target a post-
10 translational modification.

For example, the first enrichment step (utilizing a first plurality of enrichment molecules) may enrich for a particular post-translational polypeptide modification, and a second enrichment step (utilizing a second plurality of enrichment molecules) may enrich for a particular polypeptide (and variants of that polypeptide). Alternatively, the first enrichment step (utilizing
15 a first plurality of enrichment molecules) may enrich for a particular polypeptide (and variants of that polypeptide), and a second enrichment step (utilizing a second plurality of enrichment molecules) may enrich for a particular post-translational modification.

In some embodiments, the first enrichment step (utilizing a first plurality of enrichment molecules) may enrich for a particular nucleic acid modification, and a second enrichment step
20 (utilizing a second plurality of enrichment molecules) may enrich for a particular polynucleic acid (and variants of that polynucleic acid). Alternatively, the first enrichment step (utilizing a first plurality of enrichment molecules) may enrich of a particular polynucleic acid (and variants of that polynucleic acid), and a second enrichment step (utilizing a second plurality of enrichment molecules) may enrich for a particular nucleic acid modification.

25 B. Molecule Modifications

One or more of the molecules of a complex sample may be modified *in vitro* prior to, concurrently with, and/or subsequent to the enrichment described above. For example, in some embodiments, a complex sample is contacted with a modifying agent prior to, concurrently with,
30 and/or subsequent to performance of enrichment. Among other things, a modifying agent may mediate fragmentation, denaturation, addition of a post-translational modification, and/or the blocking of one or more functional groups.

In some embodiments, one or more molecules of a complex sample are modified by fragmentation. In some embodiments, fragmentation comprises enzymatic digestion. For
35 example, in some embodiments, digestion is carried out by contacting a polypeptide with an

endopeptidase (*e.g.*, trypsin) under digestion conditions. In some embodiments, fragmentation comprises chemical digestion. Examples of suitable reagents for chemical and enzymatic digestion are known in the art and include, without limitation, trypsin, chymotrypsin, Lys-C, Arg-C, Asp-N, Lys-N, BNPS-Skatole, CNBr, caspase, formic acid, glutamyl endopeptidase, hydroxylamine, iodosobenzoic acid, neutrophil elastase, pepsin, proline-endopeptidase, proteinase K, staphylococcal peptidase I, thermolysin, and thrombin.

In some embodiments, one or more molecules of a complex sample are modified by denaturation (*e.g.*, by heat and/or chemical means).

In some embodiments, one or more polypeptides of a complex sample are modified by *in vitro* post-translational modification, such as by acetylation, adenylation, ADP-ribosylation, alkylation (*e.g.*, methylation), amidation, arginylation, biotinylation, butyrylation, carbamylation, carbonylation, carboxylation, citrullination, deamidation, eliminination, formylation, glycosylation (*e.g.*, N-linked glycosylation, O-linked glycosylation), glypylation, glycation, hydroxylation, iodination, ISGylation, isoprenylation, lipoylation, malonylation, myristoylation, neddylation, nitration, oxidation, palmitoylation, pegylation, phosphorylation, phosphopantetheinylation, polyglutamylation, polyglutamylation, prenylation, propionylation, pupylation, S-glutathionylation, S-nitrosylation, S-sulfenylation, S-sulfinylation, S-sulfonylation, succinylation, sulfation, SUMOylation, or ubiquitination.

In some embodiments, one or more molecules of a complex sample are modified by the blocking of one or more functional groups (*e.g.*, free carboxylate groups and/or thiol groups).

In some embodiments, blocking free carboxylate groups refers to a chemical modification of these groups which alters chemical reactivity relative to an unmodified carboxylate. Suitable carboxylate blocking methods are known in the art and should modify side-chain carboxylate groups to be chemically different from a carboxy-terminal carboxylate group of a polypeptide to be functionalized. In some embodiments, blocking free carboxylate groups comprises esterification or amidation of free carboxylate groups of a polypeptide. In some embodiments, blocking free carboxylate groups comprises methyl esterification of free carboxylate groups of a polypeptide, *e.g.*, by reacting the polypeptide with methanolic HCl. Additional examples of reagents and techniques useful for blocking free carboxylate groups include, without limitation, 4-sulfo-2,3,5,6-tetrafluorophenol (STP) and/or a carbodiimide such as N-(3-Dimethylaminopropyl)-N'-ethylcarbodiimide hydrochloride (EDAC), uronium reagents, diazomethane, alcohols and acid for Fischer esterification, the use of N-hydroxysuccinimide (NHS) to form NHS esters (potentially as an intermediate to subsequent ester or amine formation), or reaction with carbonyldiimidazole (CDI) or the formation of mixed anhydrides, or

any other method of modifying or blocking carboxylic acids, potentially through the formation of either esters or amides.

In some embodiments, blocking free thiol groups refers to a chemical modification of these groups which alters chemical reactivity relative to an unmodified thiol. In some
5 embodiments, blocking free thiol groups comprises reducing and alkylating free thiol groups of a polypeptide. In some embodiments, reduction and alkylation is carried out by contacting a polypeptide with dithiothreitol (DTT) and one or both of iodoacetamide and iodoacetic acid. Examples of additional and alternative cysteine-reducing reagents which may be used are well known and include, without limitation, 2-mercaptoethanol, Tris (2-carboxyethyl) phosphine
10 hydrochloride (TCEP), tributylphosphine, dithiobutylamine (DTBA), or any reagent capable of reducing a thiol group. Examples of additional and alternative cysteine-blocking (*e.g.*, cysteine-alkylating) reagents which may be used are well known and include, without limitation, acrylamide, 4-vinylpyridine, N-Ethylmaleimide (NEM), N- ϵ -maleimidocaproic acid (EMCA), or any reagent that modifies cysteines so as to prevent disulfide bond formation.

15 In some embodiments, the N-terminal amino acid or the C-terminal amino acid of a polypeptide is modified.

In some embodiments, a carboxy-terminus of a polypeptide is modified in a method comprising: (i) blocking free carboxylate groups of the polypeptide; (ii) denaturing the polypeptide (*e.g.*, by heat and/or chemical means); (iii) blocking free thiol groups of the
20 polypeptide; (iv) digesting the polypeptide to produce at least one polypeptide fragment comprising a free C-terminal carboxylate group; and (v) conjugating (*e.g.*, chemically) a functional moiety to the free C-terminal carboxylate group. In some embodiments, the method further comprises, after (i) and before (ii), dialyzing a sample comprising the polypeptide.

In some embodiments, a carboxy-terminus of a polypeptide is modified in a method
25 comprising: (i) denaturing the polypeptide (*e.g.*, by heat and/or chemical means); (ii) blocking free thiol groups of the polypeptide; (iii) digesting the polypeptide to produce at least one polypeptide fragment comprising a free C-terminal carboxylate group; (iv) blocking the free C-terminal carboxylate group to produce at least one polypeptide fragment comprising a blocked C-terminal carboxylate group; and (v) conjugating (*e.g.*, enzymatically) a functional moiety to
30 the blocked C-terminal carboxylate group. In some embodiments, the method further comprises, after (iv) and before (v), dialyzing a sample comprising the polypeptide.

In some embodiments, a complex sample is contacted with a modifying agent prior to enrichment to mediate polypeptide fragmentation, polypeptide denaturation, addition of a post-translational modification, and/or the blocking of one or more functional groups. Alternatively,
35 or in addition, in some embodiments, a complex sample with a modifying agent concurrently

with enrichment to mediate polypeptide fragmentation, polypeptide denaturation, addition of a post-translational modification, and/or the blocking of one or more functional groups.

Alternatively, or in addition, in some embodiments, a complex sample (or a sample derived therefrom, comprising the one or more polypeptides of interest) with a modifying agent after enrichment to mediate polypeptide fragmentation, polypeptide denaturation, addition of a post-translational modification, and/or the blocking of one or more functional groups.

In some embodiments, the 5' terminal end or the 3' terminal end of a polynucleic acid is modified. In some embodiments, an internal nucleotide of a polynucleic acid is modified (e.g., by methylation or using DNA damage methods).

IV. Sequencing and Detection/Quantification Methodologies

In some embodiments, molecules (*i.e.*, polypeptides and/or polynucleic acids) of a multiplexed sample are sequenced. As such, in some aspects, the disclosure relates to methods of sequencing and identification. Various methods of sequencing are known to those having ordinary skill in the art. For example, methods of polypeptide sequencing include mass spectrometry (*e.g.*, peptide mass fingerprinting and tandem mass spectrometry) and Edman degradation. Additional, previously undescribed methods of sequencing are described herein.

In some embodiments, molecules (*e.g.*, polypeptides, polynucleic acids, metabolites, etc.) are detected, and optionally quantified). Various methods of detecting and/or quantifying molecules are known to those having skill in the art.

A. Polypeptide Sequencing and Detection/Quantification Methodologies

As used herein, “sequencing,” “sequence determination,” “determining a sequence,” and like terms, in reference to a polypeptide include determination of partial amino acid sequence information as well as full amino acid sequence information of the polypeptide. That is, the terminology includes sequence comparisons, fingerprinting, and like levels of information about a target molecule, as well as the express identification and ordering of each amino acid of the target molecule within a region of interest. The terminology includes identifying a single amino acid (or the probability of a single amino acid) of a polypeptide. In some embodiments, more than one amino acid (or the probability of more than one amino acid) of a polypeptide is identified. Accordingly, in some embodiments, the terms “amino acid sequence” and “polypeptide sequence” as used herein may refer to the polypeptide material itself and is not restricted to the specific sequence information (*e.g.*, the succession of letters representing the order of amino acids from one terminus to another terminus) that biochemically characterizes a specific polypeptide.

In some embodiments, the probability of an amino acid at a specific position within a polypeptide is determined and illustrated in a probability array. For example, for a polypeptide consisting of two amino acids, the terms “sequencing,” “sequence determination,” “determining a sequence,” and like terms may involve determining the probability of an amino at position 1 and/or position 2, such as [[0.80, 0.12, 0.05, 0.01, 0.01, 0.01, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00], [0.00, 0.10, 0.90, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00, 0.00]] where the probabilities in the array correspond to A, R, N, D, C, Q, E, G, H, I, L, K, M, F, P, S, T, W, Y, and V, respectively. One having ordinary skill in the art will understand that this example (and exemplary probability array) can be expanded to accommodate the analysis of additional amino acid identities (e.g., modified amino acids), such as those described herein.

In some embodiments, sequencing of a polypeptide molecule comprises identifying at least two (e.g., at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at least 17, at least 18, at least 19, at least 20, at least 25, at least 30, at least 35, at least 40, at least 45, at least 50, at least 60, at least 70, at least 80, at least 90, at least 100, or more) amino acids (or amino acid probabilities) in the polypeptide molecule. In some embodiments, the at least two amino acids are contiguous amino acids. In some embodiments, the at least two amino acids are non-contiguous amino acids.

In some embodiments, sequencing of a polypeptide molecule comprises identification of less than 100% (e.g., less than 99%, less than 95%, less than 90%, less than 85%, less than 80%, less than 75%, less than 70%, less than 65%, less than 60%, less than 55%, less than 50%, less than 45%, less than 40%, less than 35%, less than 30%, less than 25%, less than 20%, less than 15%, less than 10%, less than 5%, less than 1% or less) of all amino acids in the polypeptide molecule. For example, in some embodiments, sequencing of a polypeptide molecule comprises identification of less than 100% of one type of amino acid in the polypeptide molecule (e.g., identification of a portion of all amino acids of one type in the polypeptide molecule). In some embodiments, sequencing of a polypeptide molecule comprises identification of less than 100% of each type of amino acid in the polypeptide molecule.

In some embodiments, sequencing of a polypeptide molecule comprises identification of at least 1, at least 5, at least 10, at least 15, at least 20, at least 25, at least 30, at least 35, at least 40, at least 45, at least 50, at least 55, at least 60, at least 65, at least 70, at least 75, at least 80, at least 85, at least 90, at least 95, at least 100 or more types of amino acids in the polypeptide.

In some embodiments, the application provides compositions and methods for sequencing a polypeptide by identifying a series of amino acids that are present at a terminus of a

polypeptide over time (*e.g.*, by iterative detection and cleavage of amino acids at the terminus). In yet other embodiments, the application provides compositions and methods for sequencing a polypeptide by identifying labeled amino content of the polypeptide and comparing to a reference sequence database.

5 In some embodiments, the application provides compositions and methods for sequencing a polypeptide by sequencing a plurality of fragments of the polypeptide. In some embodiments, sequencing a polypeptide comprises combining sequence information for a plurality of polypeptide fragments to identify and/or determine a sequence for the polypeptide. In some embodiments, combining sequence information may be performed by computer
10 hardware and software. See “Devices for Sample Preparation and Sample Sequencing.” The methods described herein may allow for a set of related polypeptides, such as an entire proteome of an organism, to be sequenced. In some embodiments, a plurality of single molecule sequencing reactions are performed in parallel (*e.g.*, on a single chip) according to aspects of the present application. For example, in some embodiments, a plurality of single molecule
15 sequencing reactions are each performed in separate sample wells on a single chip or array.

In some embodiments, methods provided herein may be used for the sequencing and identification of an individual polypeptide in a sample comprising a complex mixture or an enriched mixture of polypeptides. In some embodiments, the application provides methods of uniquely identifying an individual polypeptide in a complex mixture or an enriched mixture of
20 polypeptides. In some embodiments, an individual polypeptide is detected in a mixed sample by determining a partial amino acid sequence of the polypeptide. In some embodiments, the partial amino acid sequence of the polypeptide is within a contiguous stretch of approximately 5 to 50 amino acids.

Without wishing to be bound by any particular theory, it is believed that most human
25 proteins can be identified using incomplete sequence information with reference to proteomic databases. For example, simple modeling of the human proteome has shown that approximately 98% of proteins can be uniquely identified by detecting just four types of amino acids within a stretch of 6 to 40 amino acids (*see, e.g.*, Swaminathan, et al. PLoS Comput Biol. 2015, 11(2):e1004080; and Yao, et al. Phys. Biol. 2015, 12(5):055003). Therefore, a complex mixture
30 or enriched mixture of polypeptides can be degraded (*e.g.*, chemically degraded, enzymatically degraded) into short polypeptide fragments of approximately 6 to 40 amino acids, and sequencing of this polypeptide library would reveal the identity and abundance of each of the polypeptides present in the original complex mixture or enriched mixture. Compositions and methods for selective amino acid labeling and identifying polypeptides by determining partial
35 sequence information are described in in detail in U.S. Pat. Application No. 15/510,962, filed

September 15, 2015, titled “SINGLE MOLECULE PEPTIDE SEQUENCING,” which is incorporated by reference in its entirety.

Embodiments are capable of sequencing single polypeptide molecules with high accuracy, such as an accuracy of at least about 50%, 60%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, 99.9%, 99.99%, 99.999%, or 99.9999%. In some embodiments, the target molecule used in single molecule sequencing is a polypeptide that is immobilized to a surface of a solid support such as a bottom surface or a sidewall surface of a sample well. The sample well also can contain any other reagents needed for a sequencing reaction in accordance with the application, such as one or more suitable buffers, co-factors, labeled affinity reagents, and enzymes (*e.g.*, catalytically active or inactive exopeptidase enzymes, which may be luminescently labeled or unlabeled).

Sequencing in accordance with the application, in some aspects, may involve immobilizing a polypeptide on a surface of a substrate (*e.g.*, of a solid support, for example a chip, for example an integrated device as described herein). In some embodiments, a polypeptide may be immobilized on a surface of a sample well (*e.g.*, on a bottom surface of a sample well) on a substrate. In some embodiments, the N-terminal amino acid of the polypeptide is immobilized (*e.g.*, attached to the surface). In some embodiments, the C-terminal amino acid of the polypeptide is immobilized (*e.g.*, attached to the surface). In some embodiments, one or more non-terminal amino acids are immobilized (*e.g.*, attached to the surface). The immobilized amino acid(s) can be attached using any suitable covalent or non-covalent linkage, for example as described in this application. In some embodiments, a plurality of polypeptides are attached to a plurality of sample wells (*e.g.*, with one polypeptide attached to a surface, for example a bottom surface, of each sample well), for example in an array of sample wells on a substrate.

Sequencing in accordance with the application, in some aspects, may be performed using a system that permits single molecule analysis. The system may include a sequencing device and an instrument configured to interface with the sequencing device. See “Devices for Sample Preparation and Sample Sequencing”.

(i) *Labeled Affinity Reagents and Methods of Use*

In some embodiments, methods provided herein comprise contacting a polypeptide with a labeled affinity reagent (also referred to herein as an amino acid recognition molecule, which may or may not comprise a label) that selectively binds one type of terminal amino acid. As used herein, in some embodiments, a terminal amino acid may refer to an amino-terminal amino acid of a polypeptide or a carboxy-terminal amino acid of a polypeptide. In some embodiments,

a labeled affinity reagent selectively binds one type of terminal amino acid over other types of terminal amino acids. In some embodiments, a labeled affinity reagent selectively binds one type of terminal amino acid over an internal amino acid of the same type. In yet other embodiments, a labeled affinity reagent selectively binds one type of amino acid at any position of a polypeptide, *e.g.*, the same type of amino acid as a terminal amino acid and an internal amino acid.

As used herein, in some embodiments, a type of amino acid refers to one of the twenty naturally occurring amino acids or a subset of types thereof. In some embodiments, a type of amino acid refers to a modified variant of one of the twenty naturally occurring amino acids or a subset of unmodified and/or modified variants thereof. Examples of modified amino acid variants include, without limitation, post-translationally-modified variants (*e.g.*, acetylation, ADP-ribosylation, caspase cleavage, citrullination, formylation, N-linked glycosylation, O-linked glycosylation, hydroxylation, methylation, myristoylation, neddylation, nitration, oxidation, palmitoylation, phosphorylation, prenylation, S-nitrosylation, sulfation, sumoylation, and ubiquitination), chemically modified variants, unnatural amino acids, and proteinogenic amino acids such as selenocysteine and pyrrolysine. In some embodiments, a subset of types of amino acids includes more than one and fewer than twenty amino acids having one or more similar biochemical properties. For example, in some embodiments, a type of amino acid refers to one type selected from amino acids with charged side chains (*e.g.*, positively and/or negatively charged side chains), amino acids with polar side chains (*e.g.*, polar uncharged side chains), amino acids with nonpolar side chains (*e.g.*, nonpolar aliphatic and/or aromatic side chains), and amino acids with hydrophobic side chains.

In some embodiments, methods provided herein comprise contacting a polypeptide with one or more labeled affinity reagents that selectively bind one or more types of terminal amino acids. As an illustrative and non-limiting example, where four labeled affinity reagents are used in a method of the application, any one reagent selectively binds one type of terminal amino acid that is different from another type of amino acid to which any of the other three selectively binds (*e.g.*, a first reagent binds a first type, a second reagent binds a second type, a third reagent binds a third type, and a fourth reagent binds a fourth type of terminal amino acid). For the purposes of this discussion, one or more labeled affinity reagents in the context of a method described herein may be alternatively referred to as a set of labeled affinity reagents.

In some embodiments, a set of labeled affinity reagents comprises at least one and up to six labeled affinity reagents. For example, in some embodiments, a set of labeled affinity reagents comprises one, two, three, four, five, or six labeled affinity reagents. In some embodiments, a set of labeled affinity reagents comprises ten or fewer labeled affinity reagents.

In some embodiments, a set of labeled affinity reagents comprises eight or fewer labeled affinity reagents. In some embodiments, a set of labeled affinity reagents comprises six or fewer labeled affinity reagents. In some embodiments, a set of labeled affinity reagents comprises four or fewer labeled affinity reagents. In some embodiments, a set of labeled affinity reagents comprises three or fewer labeled affinity reagents. In some embodiments, a set of labeled affinity reagents comprises two or fewer labeled affinity reagents. In some embodiments, a set of labeled affinity reagents comprises four labeled affinity reagents. In some embodiments, a set of labeled affinity reagents comprises at least two and up to twenty (*e.g.*, at least two and up to ten, at least two and up to eight, at least four and up to twenty, at least four and up to ten) labeled affinity reagents. In some embodiments, a set of labeled affinity reagents comprises more than twenty (*e.g.*, 20 to 25, 20 to 30) affinity reagents. It should be appreciated, however, that any number of affinity reagents may be used in accordance with a method of the application to accommodate a desired use.

In accordance with the application, in some embodiments, one or more types of amino acids are identified by detecting luminescence of a labeled affinity reagent (*e.g.*, an amino acid recognition molecule comprising a luminescent label). In some embodiments, a labeled affinity reagent comprises an affinity reagent that selectively binds one type of amino acid and a luminescent label having a luminescence that is associated with the affinity reagent. In this way, the luminescence (*e.g.*, luminescence lifetime, luminescence intensity, and other luminescence properties described elsewhere herein) may be associated with the selective binding of the affinity reagent to identify an amino acid of a polypeptide. In some embodiments, a plurality of types of labeled affinity reagents may be used in a method according to the application, wherein each type comprises a luminescent label having a luminescence that is uniquely identifiable from among the plurality. Suitable luminescent labels may include luminescent molecules, such as fluorophore dyes, and are described elsewhere herein.

In some embodiments, one or more types of amino acids are identified by detecting one or more electrical characteristics of a labeled affinity reagent. In some embodiments, a labeled affinity reagent comprises an affinity reagent that selectively binds one type of amino acid and a conductivity label that is associated with the affinity reagent. In this way, the one or more electrical characteristics (*e.g.*, charge, current oscillation color, and other electrical characteristics) may be associated with the selective binding of the affinity reagent to identify an amino acid of a polypeptide. In some embodiments, a plurality of types of labeled affinity reagents may be used in a method according to the application, wherein each type comprises a conductivity label that produces a change in an electrical signal (*e.g.*, a change in conductance, such as a change in amplitude of conductivity and conductivity transitions of a characteristic

pattern) that is uniquely identifiable from among the plurality. In some embodiments, the plurality of types of labeled affinity reagents each comprises a conductivity label having a different number of charged groups (*e.g.*, a different number of negatively and/or positively charged groups). Accordingly, in some embodiments, a conductivity label is a charge label.

5 Examples of charge labels include dendrimers, nanoparticles, nucleic acids and other polymers having multiple charged groups. In some embodiments, a conductivity label is uniquely identifiable by its net charge (*e.g.*, a net positive charge or a net negative charge), by its charge density, and/or by its number of charged groups.

In some embodiments, an affinity reagent (*e.g.*, an amino acid recognition molecule) may
10 be engineered by one skilled in the art using conventionally known techniques. In some embodiments, desirable properties may include an ability to bind selectively and with high affinity to one type of amino acid only when it is located at a terminus (*e.g.*, an N-terminus or a C-terminus) of a polypeptide. In yet other embodiments, desirable properties may include an ability to bind selectively and with high affinity to one type of amino acid when it is located at a
15 terminus (*e.g.*, an N-terminus or a C-terminus) of a polypeptide and when it is located at an internal position of the polypeptide.

As used herein, in some embodiments, the terms “selective” and “specific” (and variations thereof, *e.g.*, selectively, specifically, selectivity, specificity) refer to a preferential binding interaction. For example, in some embodiments, a labeled affinity reagent that
20 selectively binds one type of amino acid preferentially binds the one type over another type of amino acid. A selective binding interaction will discriminate between one type of amino acid (*e.g.*, one type of terminal amino acid) and other types of amino acids (*e.g.*, other types of terminal amino acids), typically more than about 10- to 100-fold or more (*e.g.*, more than about 1,000- or 10,000-fold). Accordingly, it should be appreciated that a selective binding interaction
25 can refer to any binding interaction that is uniquely identifiable to one type of amino acid over other types of amino acids. For example, in some aspects, the application provides methods of polypeptide sequencing by obtaining data indicative of association of one or more amino acid recognition molecules with a polypeptide molecule. In some embodiments, the data comprises a series of signal pulses corresponding to a series of reversible amino acid recognition molecule
30 binding interactions with an amino acid of the polypeptide molecule, and the data may be used to determine the identity of the amino acid. As such, in some embodiments, a “selective” or “specific” binding interaction refers to a detected binding interaction that discriminates between one type of amino acid and other types of amino acids. In some embodiments, a labeled affinity reagent (*e.g.*, an amino acid recognition molecule) selectively binds one type of amino acid with
35 a dissociation constant (K_D) of less than about 10^{-6} M (*e.g.*, less than about 10^{-7} M, less than

about 10^{-8} M, less than about 10^{-9} M, less than about 10^{-10} M, less than about 10^{-11} M, less than about 10^{-12} M, to as low as 10^{-16} M) without significantly binding to other types of amino acids. In some embodiments, a labeled affinity reagent selectively binds one type of amino acid (*e.g.*, one type of terminal amino acid) with a K_D of less than about 100 nM, less than about 50 nM, less than about 25 nM, less than about 10 nM, or less than about 1 nM. In some embodiments, a labeled affinity reagent selectively binds one type of amino acid with a K_D between about 50 nM and about 50 μ M (*e.g.*, between about 50 nM and about 500 nM, between about 50 nM and about 5 μ M, between about 500 nM and about 50 μ M, between about 5 μ M and about 50 μ M, or between about 10 μ M and about 50 μ M). In some embodiments, an amino acid recognition molecule binds one type of amino acid with a K_D of about 50 nM.

In some embodiments, a labeled affinity reagent (*e.g.*, an amino acid recognition molecule) binds two or more types of amino acids with a K_D of less than about 10^{-6} M (*e.g.*, less than about 10^{-7} M, less than about 10^{-8} M, less than about 10^{-9} M, less than about 10^{-10} M, less than about 10^{-11} M, less than about 10^{-12} M, to as low as 10^{-16} M). In some embodiments, an amino acid recognition molecule binds two or more types of amino acids with a K_D of less than about 100 nM, less than about 50 nM, less than about 25 nM, less than about 10 nM, or less than about 1 nM. In some embodiments, an amino acid recognition molecule binds two or more types of amino acids with a K_D of between about 50 nM and about 50 μ M (*e.g.*, between about 50 nM and about 500 nM, between about 50 nM and about 5 μ M, between about 500 nM and about 50 μ M, between about 5 μ M and about 50 μ M, or between about 10 μ M and about 50 μ M). In some embodiments, an amino acid recognition molecule binds two or more types of amino acids with a K_D of about 50 nM.

In some embodiments, a labeled affinity reagent (*e.g.*, an amino acid recognition molecule) binds at least one type of amino acid with a dissociation rate (k_{off}) of at least 0.1 s^{-1} . In some embodiments, the dissociation rate is between about 0.1 s^{-1} and about $1,000\text{ s}^{-1}$ (*e.g.*, between about 0.5 s^{-1} and about 500 s^{-1} , between about 0.1 s^{-1} and about 100 s^{-1} , between about 1 s^{-1} and about 100 s^{-1} , or between about 0.5 s^{-1} and about 50 s^{-1}). In some embodiments, the dissociation rate is between about 0.5 s^{-1} and about 20 s^{-1} . In some embodiments, the dissociation rate is between about 2 s^{-1} and about 20 s^{-1} . In some embodiments, the dissociation rate is between about 0.5 s^{-1} and about 2 s^{-1} .

In some embodiments, the value for K_D or k_{off} can be a known literature value, or the value can be determined empirically. For example, the value for K_D or k_{off} can be measured in a single-molecule assay or an ensemble assay. In some embodiments, the value for k_{off} can be determined empirically based on signal pulse information obtained in a single-molecule assay as described elsewhere herein. For example, the value for k_{off} can be approximated by the

reciprocal of the mean pulse duration. In some embodiments, an amino acid recognition molecule binds two or more types of amino acids with a different K_D or k_{off} for each of the two or more types. In some embodiments, a first K_D or k_{off} for a first type of amino acid differs from a second K_D or k_{off} for a second type of amino acid by at least 10% (e.g., at least 25%, at least 50%, at least 100%, or more). In some embodiments, the first and second values for K_D or k_{off} differ by about 10-25%, 25-50%, 50-75%, 75-100%, or more than 100%, for example by about 2-fold, 3-fold, 4-fold, 5-fold, or more.

In some embodiments, a labeled affinity reagent comprises a luminescent label (e.g., a label) and an affinity reagent (shown as stippled shapes) that selectively binds one or more types of terminal amino acids of a polypeptide. In some embodiments, an affinity reagent is selective for one type of amino acid or a subset (e.g., fewer than the twenty common types of amino acids) of types of amino acids at a terminal position or at both terminal and internal positions.

As described herein, an affinity reagent (also known as a “recognition molecule”) may be any biomolecule capable of selectively or specifically binding one molecule over another molecule (e.g., one type of amino acid over another type of amino acid, as with an “amino acid recognition molecule” referred to herein). Affinity reagents (e.g., recognition molecules) include, for example, proteins and nucleic acids, which may be synthetic or recombinant. In some embodiments, an affinity reagent or recognition molecule may be an antibody or an antigen-binding portion of an antibody, or an enzymatic biomolecule, such as a peptidase, an aminotransferase, a ribozyme, an aptazyme, or a tRNA synthetase, including aminoacyl-tRNA synthetases and related molecules described in U.S. Pat. Application No. 15/255,433, filed September 2, 2016, titled “MOLECULES AND METHODS FOR ITERATIVE POLYPEPTIDE ANALYSIS AND PROCESSING”.

In some embodiments, an affinity reagent or recognition molecule of the application is a degradation pathway protein. Examples of degradation pathway proteins suitable for use as recognition molecules include, without limitation, N-end rule pathway proteins, such as Arg/N-end rule pathway proteins, Ac/N-end rule pathway proteins, and Pro/N-end rule pathway proteins. In some embodiments, a recognition molecule is an N-end rule pathway protein selected from a Gid4 protein, a Ubr1 UBR box protein, and a ClpS protein (e.g., ClpS2).

A peptidase, also referred to as a protease or proteinase, is an enzyme that catalyzes the hydrolysis of a peptide bond. Peptidases digest polypeptides into shorter fragments and may be generally classified into endopeptidases and exopeptidases, which cleave a polypeptide chain internally and terminally, respectively. In some embodiments, labeled affinity reagent comprises a peptidase that has been modified to inactivate exopeptidase or endopeptidase activity. In this way, labeled affinity reagent selectively binds without also cleaving the amino acid from a

polypeptide. In yet other embodiments, a peptidase that has not been modified to inactivate exopeptidase or endopeptidase activity may be used. For example, in some embodiments, a labeled affinity reagent comprises a labeled exopeptidase.

In accordance with certain embodiments of the application, polypeptide sequencing methods may comprise iterative detection and cleavage at a terminal end of a polypeptide. In some embodiments, labeled exopeptidase may be used as a single reagent that performs both steps of detection and cleavage of an amino acid. As generically depicted, in some embodiments, labeled exopeptidase has aminopeptidase or carboxypeptidase activity such that it selectively binds and cleaves an N-terminal or C-terminal amino acid, respectively, from a polypeptide. It should be appreciated that, in certain embodiments, labeled exopeptidase may be catalytically inactivated by one skilled in the art such that labeled exopeptidase retains selective binding properties for use as a non-cleaving labeled affinity reagent, as described herein.

An exopeptidase generally requires a polypeptide substrate to comprise at least one of a free amino group at its amino-terminus or a free carboxyl group at its carboxy-terminus. In some embodiments, an exopeptidase in accordance with the application hydrolyses a bond at or near a terminus of a polypeptide. In some embodiments, an exopeptidase hydrolyses a bond not more than three residues from a polypeptide terminus. For example, in some embodiments, a single hydrolysis reaction catalyzed by an exopeptidase cleaves a single amino acid, a dipeptide, or a tripeptide from a polypeptide terminal end.

In some embodiments, an exopeptidase in accordance with the application is an aminopeptidase or a carboxypeptidase, which cleaves a single amino acid from an amino- or a carboxy-terminus, respectively. In some embodiments, an exopeptidase in accordance with the application is a dipeptidyl-peptidase or a peptidyl-dipeptidase, which cleave a dipeptide from an amino- or a carboxy-terminus, respectively. In yet other embodiments, an exopeptidase in accordance with the application is a tripeptidyl-peptidase, which cleaves a tripeptide from an amino-terminus. Peptidase classification and activities of each class or subclass thereof is well known and described in the literature (see, *e.g.*, Gurupriya, V. S. & Roy, S. C. *Proteases and Protease Inhibitors in Male Reproduction. Proteases in Physiology and Pathology* 195–216 (2017); and Brix, K. & Stöcker, W. *Proteases: Structure and Function*. Chapter 1).

An exopeptidase in accordance with the application may be selected or engineered based on the directionality of a sequencing reaction. For example, in embodiments of sequencing from an amino-terminus to a carboxy-terminus of a polypeptide, an exopeptidase comprises aminopeptidase activity. Conversely, in embodiments of sequencing from a carboxy-terminus to an amino-terminus of a polypeptide, an exopeptidase comprises carboxypeptidase activity. Examples of carboxypeptidases that recognize specific carboxy-terminal amino acids, which

may be used as labeled exopeptidases or inactivated to be used as non-cleaving labeled affinity reagents described herein, have been described in the literature (see, *e.g.*, Garcia-Guerrero, M.C., et al. (2018) *PNAS* 115(17)).

Suitable peptidases for use as cleaving reagents and/or affinity reagents (*e.g.*, recognition molecules) include aminopeptidases that selectively bind one or more types of amino acids. In some embodiments, an aminopeptidase recognition molecule is modified to inactivate aminopeptidase activity. In some embodiments, an aminopeptidase cleaving reagent is non-specific such that it cleaves most or all types of amino acids from a terminal end of a polypeptide. In some embodiments, an aminopeptidase cleaving reagent is more efficient at cleaving one or more types of amino acids from a terminal end of a polypeptide as compared to other types of amino acids at the terminal end of the polypeptide. For example, an aminopeptidase in accordance with the application specifically cleaves alanine, arginine, asparagine, aspartic acid, cysteine, glutamine, glutamic acid, glycine, histidine, isoleucine, leucine, lysine, methionine, phenylalanine, proline, selenocysteine, serine, threonine, tryptophan, tyrosine, and/or valine. In some embodiments, an aminopeptidase is a proline aminopeptidase. In some embodiments, an aminopeptidase is a proline iminopeptidase. In some embodiments, an aminopeptidase is a glutamate/aspartate-specific aminopeptidase. In some embodiments, an aminopeptidase is a methionine-specific aminopeptidase. In some embodiments, an aminopeptidase is an aminopeptidase set forth in TABLE 1. In some embodiments, an aminopeptidase cleaving reagent cleaves a peptide substrate set forth in TABLE 1.

In some embodiments, an aminopeptidase is a non-specific aminopeptidase. In some embodiments, a non-specific aminopeptidase is a zinc metalloprotease. In some embodiments, a non-specific aminopeptidase is an aminopeptidase set forth in TABLE 2. In some embodiments, a non-specific aminopeptidase cleaves a peptide substrate set forth in TABLE 2.

Accordingly, in some embodiments, the application provides an aminopeptidase (*e.g.*, an aminopeptidase recognition molecule, an aminopeptidase cleaving reagent) having an amino acid sequence selected from TABLE 1 or TABLE 2 (or having an amino acid sequence that has at least 50%, at least 60%, at least 70%, at least 80%, 80-90%, 90-95%, 95-99%, or higher, amino acid sequence identity to an amino acid sequence selected from TABLE 1 or TABLE 2). In some embodiments, an aminopeptidase has 25-50%, 50-60%, 60-70%, 70-80%, 80-90%, 90-95%, or 95-99%, or higher, amino acid sequence identity to an aminopeptidase listed in TABLE 1 or TABLE 2. In some embodiments, an aminopeptidase is a modified aminopeptidase and includes one or more amino acid mutations relative to a sequence set forth in TABLE 1 or TABLE 2.

TABLE 1. Non-limiting examples of aminopeptidases

Name	SEQ ID NO:	Sequence
<i>L. pneumophila</i> M1 Aminopeptidase (Glu/Asp Specific)	1	MGSSHHHHHHSSGLVPRGSHMMVKQGVFMKTDQSKVKKLSYKSLDYF VIHVDLQIDLSKKPVESKARLTVVPNLNVDSHSNDLVLDGENMTLVSLQ MNDNLLKENEYELTKDSLIIKNIPQNTPTIEMTSLLGENTDLFGLYETEGV ALVKAESEGLRRVFYLPDRPDNLATYKTTIANQEDYPVLLSNGVLIEKKE LPLGLHSVTWLDDVPKPSYLFALVAGNLQRSVTYYQTKSGRELPIEFYVP PSATSKCDFAKEVLKEAMAWDERTFNLECALRQHMVAGVDKYASGASE PTGLNLFNTENLFASPETKTDLGILRVLEVVAHEFFHYWSGDRVTIRDWF NLPLKEGLTTFRAAMFREELFGTDLIRLLDGKNLDERAPRQSAYTAVRSL YTAAAYEKSADIFRMMMLFIGKEPFIEAVAKFFKDNDGGAVTLEDIESIS NSSGKDLRSFLSWFTESGIPELIVTDELNPDTKQYFLKIKTVNGRNRPIIL MGLLDSSGAIEIVADKLLIVDQEEIEFQFENIQTRPIPSLLRSFSAPVHMKYE YSYQDLLLLMQFDTNLYNRCEAAKQLISALINDFCIGKKIELSPQFFAVYK ALLSDNSLNEWMLAELITLSLEELIENQDKPDFEKLNEGRQLIQNALANE LKTDIFYNLLFRIQISGDDDKQKLKGFDLKQAGLRRLKSVCFSYLLNVDFE KTKEKLILQFEDALGKNMTETALALSMLCEINCEEADVLEDYYHYWKN DPGAVNNWFSIQALAHSPDVIERVKKLMRHGDFDLSNPNKVYALLGSFIK NPFGFHSVTGEGYQLVADAIFDLKINPTLAANLTKFTYWDKYDVNRQ AMMISTLKIIYSNATSSDVRTMAKKGLDKVKEDLPLPIHLTFFHGGSTMQD RTAQLIADGNKENAYQLH
<i>E. coli</i> methionine aminopeptidase (Met specific)	2	MAHHHHHHMGT AISIKTPEDIEKMRVAGR LA AEVLEMI EPYVKPGVSTGE LDRICNDYIVNEQH AVSACLG YHGYPKSV CISINEV VCHGIPDDAKLLKD GDIVNIDVTVIKDGFGHGDTSKM FIVGKPTIMGERLCRITQESLYLALRMVK PGINLREIGAAIQKFVEAEGFSV VREYCGHGIGRGFHEEPQVLHYDSRETN VVLKPGMTFTIEPMVNAGKKEIRTMKDGWTVKTKDRSLSAQYEHTIVVT DNGCEILTLRKDDTIPAIISHD
<i>M. smegmatis</i> Proline iminopeptidase (Pro specific)	3	MAHHHHHHMGTLEANTNGPGSMLS RMPVSSRTVPFGDHETWVQVTTPE NAQPHALPLIVLHGGPGMAHNYVANIAALADETGRTVIHYDQVGCNST HLPDAPADFWTPQLFVDEFHAVCTALGIERYHVLGQSWGMLGAIEAVR QPSGLVSLAICNSPASMR LWSEAAGDLRAQLPAETRAALDRHEAAGTITH PDYLQAAAEFYRHRVCRV VPTPQDFADSV AQMEA EPTVYHTMNGPNEF HVVGTLGDWSVIDRLPDVTAPVLVIAGEHDEATPKTWQPFVDHIPDVRS VFPGTSHCTHLEKPEEFRAVVAQFLHQHDLAADARV
<i>Y. pestis</i> Proline iminopeptidase (Pro Specific)	4	MTQQEYQNRQALLAKMAPGSAAIIFAPEATRSADSEYPYRQNSDFSYL TGFNEPEAVLILVKSDETHNHSVLFNRIRDLTAEIWFGRRLGQEAAPTCLA VDRALPFDEINEQLYLLLNRLDVIYHAQQQYAYADNIVFAALEKLRHGFR KNLRAPATLTDWRPWLHEMRLFKSAEEI AVLRRAGEISALAHTRAMEKC RPGMF EYQLEGEILHEFTRHGARYPA YNTIVGGGENG CILHYTENECELR DGDVLIDAGCEYRGYAGDITRTFPVNGKFTPAQRAVYDIVLAAINKSLT LFRPGTSIREVTEEVVRIMV VGLVELGILKGDIEQLIAEQAHRPFFMHGLSH WLGMDVHDVGDY GSSDRGRILEPGMVLTVEPGLYIAPDADVPPQYRGIGI RIEDDIVITATGNENLTASVVKDPDDIEALMALNHAGENLYFQEHHHHHH
<i>P. furiosus</i> Methionine aminopeptidase	5	MDTEKLMKAGEIAKKVREKAIKLARPGMLLLELAESIEKMIMELGGKPAF PVNLSINEIAAHYTPYKGD TTVLKEGDY LKIDVG VHI DGFIA DTA VTVRVG MEEDELMEAAKEALNAAISVARAGVEIKELGKAIENEIRKRGFKPIVNL SG HKIERYKLHAGISIPNIYRPHDNYVLKEGDVFAIEPFATIGAGQVIEVPPTLI YMYVRDVPVRVAQARFLLAKIKREYGTLPFAYRWLQNDMPEGQLKLAL KTLEKAGAIYGYPV LKEIRNGIVAQFEHTIIVEKDSVIVTQDMINKSTLE
<i>Aeromonas sobria</i> Proline aminopeptidase	6	HMSSPLHYVLDGIHCEPHFFTVP LLDHQQPDDEETITLFGRTL CRKDRLLDDE LPWLLYLQGGPGFGAPRPSANGGWIKRALQEFRVLLLDQRGTGHSTPIHA ELLAHLNPRQQADYLSHFRADSIVRDAELIREQLSPDHPWSLLGQSFGGFC SLTYLSLFPDSLHEVYLTGGVAPIGRSADEVYRATYQRVADKNRAFFARF PHAQAIANRLATHLQRHDVRLPNGQRLTVEQLQQGLDLGASGA FEELY YLLEDAFIGEKLNPAFLYQVQAMQPFNTNPVFAILHELIYCEGAASHWAA ERVRGEFPALAWAQGKDAFTGEMIFPWFMEQFRELIPLKEAAHLLAEKA DWGPLYDPVQLARNKVPVACAVYAEDMYVEFDYSRETLKGLSNSRAWI TNEYEHNGLRVDGEQILDRLRLNRDCLE

<i>Pyrococcus furiosus</i> Proline Aminopeptidase (X- /-Pro)	7	MKERLEKLVKFMDENSIDRVFIKPVNVYYFSGTSPLGGGYIIVDGDEATL YVPELEYEMAKEESKLPVVKFKKFDEIYEILKNTETLGIETLSYSMVENF KEKSNVKEFKKIDDVIKDLRIIKTKEEIEIEKACEIADKAVMAAIEEITEGK REREVA AKVEYLMKMNGAEKPAFDITIIASGHRALPHGVASDKRIERGDL VVIDLGALYNHYNSDITRTIVVGSPNEKQREIYEIVLEAQKRAVEAAKPG MTAKELDSIAREIIKEYGYGDYFIHSLGHGVGLEIHEWPRISQYDETVLKE GMVITIEPGIYIPKLGGVRIEDTVLITENGAKRLTKTERELL
<i>Elizabethkingia meningoseptica</i> Proline aminopeptidase	8	MIPITTPVGNFKVWTKRFGTNP KIKVLLHGGPAMTHEYMECFETFFQRE GFEFYEDQLGSYYSDQPTDEKLWNIDRFVDEVEQVRKAHADKENFYV LGNSWGGILAMEYALKYQQNLKGLIVANMMASAPYVVKYAEVLSKQM KPEVLAEVRAIEAKKDYANPRYTELLFPNYYAQHICRLKEWPDALNRSK HVNSTVYTLMQGPSELGMSSDARLAKWDIKNRLHEIATPTLMIGARYDT MDPKAMEEQSKLVQKGRYLYCPNGSHLAMWDDQKVFMGDGVKFIKDV DTKSFN
<i>Aeromonas sobria</i> Proline aminopeptidase	9	HMSSPLHYVLDGIHCEPHFFTVPLDHQQPDDEETITLFGRTLCKDRDLDE LPWLLYLQGGPGFGAPRPSANGGWIKRALQEFRVLLLDQRGTGHSTPIHA ELLAHLNPRQQADYLSHFRADSIVRDAELIREQLSPDHPWSLLGQSFGGFC SLTYLSLFPDSLHEVYLTGGVAPIGRSADEVYRATYQRVADKNRAFFARF PHAQAIANRLATHLQRHVDRLPNQGRLTVEQLQQGLDLGASGAFEELY YLLEDAFIGEKLNPFLYQVQAMQPFNTNPVFALHELIYCEGAASHWAA ERVRGEFPALAWAQGKDFATGEMIFPWFMEQFRELIPLKEAAHLLAEKA DWGPLYDPVQLARNKVPVACAVYAEDMYVEFDYSRETLKGLSNSRAWI TNEYEHNGLRVDGEQILDRLRLNRDCLE
<i>N. gonorrhoeae</i> Proline Iminopeptidase	10	MYEIKQPFHSGYLQVSEIHQIYWEESGNPDGVPVIFLHGGPGAGASPECRG FFNPDVFRIVIIDQRCGRSHPYACAEDNTTWDLVADIEKVREMLGIGKW LVFGGSWGSTLSLAYAQTHPERVKGLVLRGIFLCRSETAWLNEAGGVSR IYPEQWQKFVAPIAENRRNRLLIEAYHGLLFHQDEEVCLSAKAWADWES YLIRFEPEGVDEDAYASLAARLENHYFVNGGWLQGDKAILNNIGKIRHIP TVIVQGRYDLCTPMQSAWELSKAFPEAELRVVQAGHCAFDPPLADALVQ AVEDILPRLL

TABLE 2. Non-limiting example of non-specific aminopeptidases

Name	SEQ ID NO:	Sequence
<i>E. coli</i> Aminopeptidase N (Zinc Metalloprotease)*	11	MGSSHHHHHHSSGENLYFQGHMTQQPQAKYRHDYRAPDYQITDIDLTFD LDAQKTVVTAVSQAVRHGASDAPLRLNGEDLKLVSVINDEPWTAWKE EEGALVISNLPERFTLKIINEISPAANTALEGLYQSGDALCTQCEAEGFRHIT YYLDRPDVLARFTTKIADKIKYPFLLSNGNRVAQGELENGRHWVQWQD PFPKPCYLFALVAGDFDVLRTFTTTRSGREVALELYVDRGNLDRAPWAM TSLKNSMKWDEERFGLEYDLDIYMIVAVDFFNMGAMENKGLNIFNSKYV LARTDTATDKDYLDIERVIGHEYFHNWTGNRVTCRDWFQLSLKEGLTVF RDQEFSSDLGSRAVNRINNVRTMRGLQFAEDASPMAPHPDPMVIEMNNF YTLTVYEKGAEVIRMIHTLLGEENFQKGMQLYFERHDGSAATCDDFVQA MEDASNVDLSHFRRWYSQSGTPIVTVKDDYNPETEQYTLTISQRTPATPD QAEKQPLHIPFAIELYDNEGKVIPLQKGGHPVNSVLNVTQAEQTFVFDNV YFQVPALLCEFSAPVKLEYKWSDDQLTFLMRHARNDFSRWDAQAQSLLA TYIKLNVARHQGGQPLSLPVHVADAFRAVLLDEKIDPALAAEILTLPSVNE MAELFDIIDPIAIAEVREALTRTLATELADELLAIYNANYQSEYRVEHEDIA KRTLRLNACLRLAFGETHLADVLSKQFHEANNMTDALAALSAVAAQL PCRDALMQEYDDKWHQNGLVMDKWFILQATSPAANVLETVRGLLQHRS FTMSNPNRIRSLIGAFAGSNPAAFHAEDGSGYFLVEMLTDLNSRNPQVAS RLIEPLIRLKR YDAKRQEKMR AALEQLKGLNLSGDLYEKITKALA
<i>P. falciparum</i> M1 aminopeptidase**	12	PKIHYRKDYKPSGFIINQVTLNINIHDQETIVRSVLDMDISKHNVGEDLVFD GVGLKINEISINNKKLVEGEEYTYDNEFLTIFSKFVPSKFAFSSEVVIHPET NYALTGLYKSKNIIVSQCEATGFRRITFFIDRPDMMAKYDVTVTADKEY PVLLSNGDKVNEFEIPGGRHGARFNDPPLKPCYLFVAVAGDLKHLSATYI TKYTKKKVLEYVFSEEKYVSKLQWALECLKKSMAFDEYFGLEYDLSRL NLVAVSDFNVGAMENKGLNIFNANSLLASKKNSIDFSYARILTVVGHEYF HQYTGNRVTLRDWFQTLTKEGLTVHRENLFSEEMTKTVTTRLSHVDLLR SVQFLEDSSPLSHPIRPESYVSMENFYTTTVYDKGSEVMRMYLTILGEEYY

		KKGFDIYIKKNDGNTATCEDFNYAMEQAYKMKKADNSANLNQYLLWFS QSGTPHVSFKYNYDAEKKQYSIHVNQYTKPDENQKEKKPLFIPISVGLINP ENGKEMISQTTLELTKESTDFVFNNIAVKPIPSLFRGFSAPVYIEDQLTDEE RILLKKYDSDAFVRYNSCTNIYMKQILMNYNEFLKAKNEKLESFQLTPVN AQFIDAIKYLLEDPHADAGFKSYIVSLPQDRYIINFVSNLDTVDLADTKEYI YKQIGDKLNDVYYKMFKSLEAKADDLTTFNDESHVDFDQMNMRTRLNT LLSLLSKAQYPNILEIIEHSKSPYPSNWLTSLSVSAYFDKYFELYDKTYKL SKDDELLQEWLKTVSRSDRKDIYEILKKLENEVLKDSKNPNDIRAVYLPF TNNLRRFHDISGKGKYLIAEVITKTDKFNPMVATQLCEPFKLWNKLDTKR QELMLNEMNTMLQEPQISNNLKEYLLRLTNK
NPEPPS	13	MGSSHHHHHHSSGMWLAAPSLARRLLFLGPPPPPLLLLVSFRSSRRRL HSLGLAAMPEKRPFERLPADVSPINYSCLCKPDLLDFTFEGKLEAAQVR QATNQIVMNCADIDIITASYAPEGDEEIHATGFNYQNEDEKVTLSPSTLQ TGTGTLKIDFVGELNDKMKGFYRSKYTTSPSEVRYAAVTQFEATDARRA FPCWDEPAIKATFDISLVVPKDRV ALSNMNVIDRKYPDDENLVEVKFAR TPVMSTYLVAFVVG EYDFVETR SKDGV CVRVYTPVGKAEQ GKFALEVA AKTLPFYKDYFNVPYPLPKIDLIAIADFAAGAMENWGLVTYRETALLIDPK NSCSSSRQWVALVVGHELAHQWFGNLVTMEWWTHLWLN EGFASWIEY LCVDHCFPEYDIWTQFVSADYTRAQELDALDNSHPIEVSVGHPSEVDEIFD AISYSGGASVIRMLHDYIGDKDFKKGMNMYLTKFQQKNAATEDLWESLE NASGKPIAAVMNTWTKQMGFPLIYVEAEQVEDDRLLRLSQQKFCAGGSY VGEDCPQWMVPITISTSEDPNQAKLKILMDKPEMNVVLKNVKPDQWVKL NLGTVGFYRTQYSSAMLESLLPGIRDLSLPPVDRLGLQNDLFLSLARAGHST VEVLKVMEAFVNPNYTVWSDLSCNLGILSTLLSHTDFYEEIQEFVKDVFS PIGERLGWDPKPGEGHLDALLRGLVLGKLGKAGHKATLEEARRRFKDHV EGKQILSADLRSPVYLTVLKHGDTTLDIMLKLHKQADMQEEKNRIERVL GATLLPDLIQKVLTFALSEEVRPQDTVSVIGGVAGGSKHGRKAAWKFIKD NWEELYNRYQGGFLISRLIKLSVEGFAVDKMAGEVKAFFESH PAPS AERTI QQCCENILLNAAWLKRDAESIHQYLLQRKASPTV
NPEPPS E366V	14	MGSSHHHHHHSSGMWLAAPSLARRLLFLGPPPPPLLLLVSFRSSRRRL HSLGLAAMPEKRPFERLPADVSPINYSCLCKPDLLDFTFEGKLEAAQVR QATNQIVMNCADIDIITASYAPEGDEEIHATGFNYQNEDEKVTLSPSTLQ TGTGTLKIDFVGELNDKMKGFYRSKYTTSPSEVRYAAVTQFEATDARRA FPCWDEPAIKATFDISLVVPKDRV ALSNMNVIDRKYPDDENLVEVKFAR TPVMSTYLVAFVVG EYDFVETR SKDGV CVRVYTPVGKAEQ GKFALEVA AKTLPFYKDYFNVPYPLPKIDLIAIADFAAGAMENWGLVTYRETALLIDPK NSCSSSRQWVALVVGHVLAHQWFGNLVTMEWWTHLWLN EGFASWIEY LCVDHCFPEYDIWTQFVSADYTRAQELDALDNSHPIEVSVGHPSEVDEIFD AISYSGGASVIRMLHDYIGDKDFKKGMNMYLTKFQQKNAATEDLWESLE NASGKPIAAVMNTWTKQMGFPLIYVEAEQVEDDRLLRLSQQKFCAGGSY VGEDCPQWMVPITISTSEDPNQAKLKILMDKPEMNVVLKNVKPDQWVKL NLGTVGFYRTQYSSAMLESLLPGIRDLSLPPVDRLGLQNDLFLSLARAGHST VEVLKVMEAFVNPNYTVWSDLSCNLGILSTLLSHTDFYEEIQEFVKDVFS PIGERLGWDPKPGEGHLDALLRGLVLGKLGKAGHKATLEEARRRFKDHV EGKQILSADLRSPVYLTVLKHGDTTLDIMLKLHKQADMQEEKNRIERVL GATLLPDLIQKVLTFALSEEVRPQDTVSVIGGVAGGSKHGRKAAWKFIKD NWEELYNRYQGGFLISRLIKLSVEGFAVDKMAGEVKAFFESH PAPS AERTI QQCCENILLNAAWLKRDAESIHQYLLQRKASPTV
<i>Francisella tularensis</i> Aminopeptidase N	15	MIYEFVMTDPKIKYLKDYKPSNYLIDETHLIFELDESKTRVTANLYIVANR ENRENNTLVLDGVELKLLSIKLNKHLSPAFAVNENQLIINNVPKFLVQ TVVEINPSANTSLEGLYKSGDVFSQTCEATGFRKITYYLDPRPDVMAAFV KIIADKKKYPIILSNGDKIDSGDISDNQHFVWKDPFKKPCYLFALVAGDL ASIKDTYITKSQRKVSLEIYAFKQDIDKCHYAMQAVKDSMKWDEDRFGL EYDLDTFMIVAVPDFNAGAMENKGLNIFNTKYIMASNKATDKDFELVQ SVVGHEYFHNWTGDRVTCRDWFQLSLKEGLTVFRDQEFTSDLNSRDVVR IDDVRIIRSAQFAEDASPM SHPIRPESYIEMNNFYTVTVYNKGAEIIRMIHTL LGEEGFQKGMKLYFERHDGQAVTCDDFVNAMADANNRDFS LFKRWYA QSGTPNIKVSENYDASSQTYSLTLEQTTLPTADQKEKQALHIPVKMGLINP EGKNIAEQVIELKEQKQTYTFENIAAKPVASLFRDFSAPVKVEHKRSEKDL LHIVKYDNNAFNRWDSLQQIATNIILNADLNDEFLNAFKSILHDKDLK ALISNALLIPIESTIAEAMRVIMVDDIVLSRKNVNVQLADKLKDDWLAVY QQCNDNKPYSLSAEQIAKRKLKGVCLSYLMNASDQKVGTDLAQQLFDN

		ADNMTDQQTAFTELLKSNDKQVRDNAINEFYNRWRHEDLVVNKWLSSQ AQISHESALDIVKGLVNHPAYNPKNPNKVYSLIGGFGANFLQYHCKDGLG YAFMADTVLALDKFNHQAARMARNLMSWKRYDSRQAMMKNALEKI KASNPSKNVFEIVSKSLES
<i>Pyrococcus horikoshii</i> TET Aminopeptidase	16	MGSSHHHHHHSSGMEVRNMVDYELLKKVVEAPGVSGYEFLGIRDVVIEE IKDYVDEVKVDKLGNVIAHKKGEGPKVMIAAHMDQIGLMVTHIEKNGL RVAPIGGVDPKTLIAQRFKVIDKKGFIYGVGASVPPHIQKPEDRKKAPD WDQIFIDIGAESKEEAEDMGVKIGTVITWDGRLERLGKHRFVSIAFDDRIA VYTILEVAKQLKDAKADVYFVATVQEEVGLRGARTSAFGIEPDYGFADIV TIAADIPGTPEHKQVTHLGKGTAIKIMDRSVICHPTIVRWLEELAKKHEIPY QLEILLGGGTADAGAIHLTKAGVPTGALSVPARYIHSNTEVVDERDVDATV ELMTKALENIHELKI
<i>T. aquaticus</i> Aminopeptidase T	17	MDAFTENLNKLAELAIRVGLNLEEGQEIVATAPIEAVDFVRLLAEKAYEN GASLFTVLYGDNLIARKRLALVPEAHLDRAPAWLYEGMAKAFHEGAARL AVSGNDPKALEGLPPERVGRAQQAQSRAYRPTLSAITEFVTNWTIVPFAH PGWAKAVFPGLPEEEAVQRLWQAIFQATRVDQEDPVAWEAHNRVLHA KVAFLENEKRFHALHFQGGPTDLTVGLAEGHLWQGGATPTKKGRCLNPNL PTEEVFTAPHRRERVEGVVRASRPLALSGQLVEGLWARFEGGVAVEVGAE KGEEVLKLLDDEGARRLGEVALVPADNPIAKTGLVFFDTLFDENAASH IAFGQAY AENLEGRPSGEEFRRRGGNESMVHVDWMIGSEEDVDGLLED GTRVPLMRRGRWVI
<i>Bacillus stearothermophilus</i> Peptidase M28	18	MAKLDETTLMLKALTDAGKVPGNEREARDVMKTYIAPYADEVTTDGLG SLIAKKEGKSGGPKVMIAHGLDEVGFMVTQIDDKGFIRFQTLGGWWSQV MLAQRVTIVTKKGDTGVIGSKPPHILPSEARKKPEIKDMFIDIGATSREE AMEWGVPRGDMIVPYFEFTVLNNEKMLLAKAWDNRIGCAVAIDVLKQL KGVDHPNTVYGVGTVQEEVGLRGARTAAQFIQPDIAFAVDVGIAGDTPG VSEKEAMGKLGAGPHIVLYDATMVSHRGLREFVIEVAEELNIPHHFDAMP GVGTDAGAIHLTGIGVPSLTIAIPTRYIHSAAILHRDDYENTVKLLVEVIK RLDADKVKQLTFDE
<i>Vibrio cholera</i> Aminopeptidase	19	MEDKVWISMGADAVGSLNPALSESLLPHSFASGSQVWIGEV AIDELAELS HTMHEQHNRCCGYMVHTSAQGAMAALMMPESIANFTIPAPSQQDLVNA WLPQVSADQITNTIRALSSFNRFYTTTSGAQASDWLANEWRSLISSLPGS RIEQIKHSGYNQKSVVLTIQGSEKPDEWVIVGGHLDSTLGSHTNEQSIAPG ADDDASGIALSEIIRVLRDNNFRPKRSVALMAYAAEEVGLRGSQDLANQ YKAQGKKVSVLQLDMTNYRGS AEDIVFITDYTDSNLTQFLTTLIDEYLP ELTYGYDRCGYACSDHASWHKAGFSAAMPFESKFKDYNPKIHTSQDTLA NSDPTGNHAVKFTKLGLAYVIEMANAGSSQVPDDSVLQDGTAKINLSGA RGTQKRFTFELSQSKPLTIQTYGGSGDVDLYVKYGSAPSQSNWDCRPYQN GNRETCSFNNAQPGIYHVMLDGYTNYNDVALKASTQHHHHHH
<i>Photobacterium halotolerans</i> Aminopeptidasej	20	MEDKVWISIGSDASQTVKSVMQSNARSLLPESLASNGPVWVGQVDYSQ AELSHMHEDHQRCGYMVHSSPESAIASNMPQSLVAFSIPISQQDQTV NAWLQVNSQAITGTITSLTSFINRFYTTTSGAQASDWLANEWRSLASLP NASVRQVSHFGYNQKSVVLTITGSEKPDEWIVLGGHLDSTIGSHTNEQSV APGADDDASGIASVTEIIRVLSENNFPKRSIAFMAYAAEEVGLRGSQDLA NQYKAEGKQVISALQLDMTNYKGSVEDIVFITDYTDSNLTFLSGLVDEY LPSLTYGFDTCGYACSDHASWHKAGFSAAMPFEAKFNDYNPMIHTPNDT LQNSDPTASHAVKFTKLGLAYAIEMASTTGGTPPPTGNVLKDGVPVNGLS GATGSQVHYSELFPAQKNLQISTAGGSGDVDLYVSFGSEATKQNWDCRP YRNGNNEVCTFAGATPGTYSIMLDGYRQFSGVTLKASTQHHHHHH
<i>Yersinia pestis</i> AminopeptidaseN	21	MTQQPQAKYRHDYRAPDYTTIDIDLDFALDAQKTTVTAVSKVKRQGTDV TPLILNGEDLTLSVSDGQAWPHYRQQDNTLVIEQLPADFTLTIVNDIHPA TNSALEGLYLSGEALCTQCEAEGFRHITYYLD RPDVLRFTTRIVADKSRY PYLLSNGNRVGGQELDDGRHWVKWEDPFPKPSYLFALVAGDFDVLQDK FITRSGREVALEIFVDRGNLDRADWAMTSLKNSMKWDETRFGLEYDLDI YMIVAVDFFNMGAMENKGLNVFNSKYVLAKAETATDKDYLNIEAVIGHE YFHNWTGNRVTCRDWFQLSLKEGLTVFRDQEFSSDLGSRSVNRIENVRV MRAAQFAEDASPMHAIRPDKVIEMNNFYTLTVYEKGSEVIRMMHTLLG EQQFQAGMRLYFERHDGSAATCDDFVQAMEDVSNVDLSLFRRWYSQSG TPLLTVHDDYDVEKQYHLFVSQKTLPTADQPEKLPLHIPLDIELYDSKGN VIPLQHNGLPVHHVLNVTEAEQTFDFDVAQKPIPSLLREFSAPVKLDYPY SDQQLTFLMQHARNEFSRWDAQAQSLLATYIKLNVAKYQQQQPLSLPAHV ADAFRAILLDEHLDPALAAQILTLPSNEMAELFTTIDPQAISTVHEAITRC

		LAQELSDELLAVYVANMTPVYRIEHGDI AKRALRNTCLNYLAFGDEEFAN KLVSLQYHQADNMTDSLAALAAVAAQLPCRDELLAAFDVRWNHDGL VMDKWFALQATSPAANVLVQVRTLLKHPAFSLSNPNRTRSLIGSFASGNP AAFHAADGSGYQFLVEILSDLNTRNPQVAARLIEPLIRLKRYDAGRQALM RKALEQLKTLNLSGDLYEKITKALAAHHHHHH
<i>Vibrio anguillarum</i> Aminopeptidase	22	MEEKVWISIGGDATQTALRSGAQSLLENINQTSVWVGQVPVSELATLS HEMHENHQRCCGYMVHPSAQSA MSVSAMPLNLNAFSAPETQQT VNA WLPSVSAQQITSTITLTQFKNRFYTTSTGAQASNWIADHWRSLSASLPAS KVEQITHSGYNQKSVM LTTITGSEKPDEWV VIGGHL DSTLGSRTNESSIAPG ADDDASGIAGVTEIIRLLSEQNFRPKRSIAFMAYAAEEVGLRGSQDLANRF KAEGKKVMSVMQLDMTNYQGSREDIVFITDYTDSNFTQYLTQLLDEYLP SLTYGFDTCGYACSDHASWHAVGYPAAMPFESKFNDYNPNHSPQDTLQ NSDPTGFHAVKFTKLGLAYV VEMGNASTPPTPSNQLKNGVPVNGLSASR NSKTWYQFELQEAGNLSIVLSGGSGDADLYVKYQTDADLQQYDCRPYRS GNNETCQFSNAQPGRY SILLHGYNNYSNASLVANAQH HHHHHH
<i>Salinivibrio</i> <i>sp</i> YCSC6 Aminopeptidase	23	MEDKKVWISIGADAQQTALSSGAQPLLAQSV AHNGQAWIGEVSSELAA LSHEMHENHHRCCGYIVHSSAQSA MAASN MPLSRASFIAP AISQQALVTP WISQIDSALIVNTIDRLTDFPNRFYTTTSGAQASDWIKQRWQSL SAGLAGA SVTQISHSGYNQASVM LTTIEGSESPDEWV VGGHL DSTIGSRTNEQSIAPG ADDDASGIAAVTEVIRVLAQNNFQPKRSIAFVAYAAEEVGLRGSQDVAN QFKQAGKDVRGVLQ LDMTNYQGS AEDIVFITDYTDNQLTQYLTQLLDEY LPTLNYGFDTCGYACSDHASWHQVGYPAAMPFEAKFNDYNPNIH TPQDT LANS DSEGAHAAKFTKLGLAYTVELANADSSPNPGNELKLGEPI NGLSGA RGNEKYFNYRLDQSGELVIRTYGGSGD VDLVVKANGDVSTGNWDCRPY RSGNDEVCRFDNATPGNYAVMLRGYRTYDNVSLIVEH HHHHHH
<i>Vibrio proteolyticus</i> Aminopeptidase I	24	<u>G</u> MPPITQQATVTAWLPQVDASQITGTISSLESFTNRFYTTTSGAQASDWIA SEWQ <u>F</u> LSASLPNASVKQVSHSGYNQKSVM LTTITGSEAPDEWIVIGGHLDS TIGSHTNEQSVAPGADDDASGIAAVTEVIRV LSENNFQPKRSIAFMAYAAE EVGLRGSQDLANQYKSEGKNVVSALQ LDMTNYKGS AQDVVFITDYTDS NFTQYLTQLMDEYLP SLTYGFDTCGYACSDHASWHNAGYPAAMPFESKF NDYNPRIHTTQDTLANS DPTGSHAKKFTQLGLAYA IEMGSATGDTPTPGN QLEH HHHHHH
<i>P. furiosus</i> Aminopeptidase I	25	<u>MVDWELMKKIIESPGVSGYEHLGIRDLVVDILKDV ADEVKIDKLGNVIAH</u> <u>FKGSAPKVMVA AHMDKIGLMVN HIDKDG YLRVVP IGGVLPETLIAQKIRF</u> <u>FTEKGERYGVVGVLP PHLRREAKDQGGKIDWDSIIVDV GASSREEAEEMG</u> <u>FRIGTIGEFAPNFTRLSEHRFATPYLDDRICLYAMIEAARQLGEHEAD IYIV</u> <u>ASVQEEIGLRGARVASFAIDPEVGIAMDVTF AKQPNDKGKIVPELGKGPV</u> <u>MDVGPININPKLRQFADEVAKKYEIPLQVEPSRPRTGTDANVMQINREGVA</u> <u>TAVLSIPIRYMHSQVELADARDVDNTIKLAKALLEELKPMDFTPLEHHHH</u> <u>HH</u>

*Cleavage efficiency (from most to least): arginine > lysine > hydrophobic residues (including alanine, leucine, methionine, and phenylalanine) > proline (see, e.g., Matthews Biochemistry 47, 2008, 5303-5311).

**Cleavage efficiency (from most to least): leucine > alanine > arginine > phenylalanine > proline; does not cleave after glutamate and aspartate.

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For the purposes of comparing two or more amino acid sequences, the percentage of “sequence identity” between a first amino acid sequence and a second amino acid sequence (also referred to herein as “amino acid identity”) may be calculated by dividing [the number of amino acid residues in the first amino acid sequence that are identical to the amino acid residues at the corresponding positions in the second amino acid sequence] by [the total number of amino acid residues in the first amino acid sequence] and multiplying by [100], in which each deletion, insertion, substitution or addition of an amino acid residue in the second amino acid sequence compared to the first amino acid sequence is considered as a difference at a single amino acid residue (position). Alternatively, the degree of sequence identity between two amino acid

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sequences may be calculated using a known computer algorithm (*e.g.*, by the local homology algorithm of Smith and Waterman (1970) *Adv. Appl. Math.* 2:482c, by the homology alignment algorithm of Needleman and Wunsch, *J. Mol. Biol.* (1970) 48:443, by the search for similarity method of Pearson and Lipman. *Proc. Natl. Acad. Sci. USA* (1998) 85:2444, or by computerized implementations of algorithms available as Blast, Clustal Omega, or other sequence alignment algorithms) and, for example, using standard settings. Usually, for the purpose of determining the percentage of “sequence identity” between two amino acid sequences in accordance with the calculation method outlined hereinabove, the amino acid sequence with the greatest number of amino acid residues will be taken as the “first” amino acid sequence, and the other amino acid sequence will be taken as the “second” amino acid sequence.

Additionally, or alternatively, two or more sequences may be assessed for the identity between the sequences. The terms “identical” or percent “identity” in the context of two or more nucleic acids or amino acid sequences, refer to two or more sequences or subsequences that are the same. Two sequences are “substantially identical” if two sequences have a specified percentage of amino acid residues or nucleotides that are the same (*e.g.*, at least 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, 99.5%, 99.6%, 99.7%, 99.8%, or 99.9% identical) over a specified region or over the entire sequence, when compared and aligned for maximum correspondence over a comparison window, or designated region as measured using one of the above sequence comparison algorithms or by manual alignment and visual inspection. Optionally, the identity exists over a region that is at least about 25, 50, 75, or 100 amino acids in length, or over a region that is 100 to 150, 150 to 200, 100 to 200, or 200 or more, amino acids in length.

Additionally, or alternatively, two or more sequences may be assessed for the alignment between the sequences. The terms “alignment” or percent “alignment” in the context of two or more nucleic acids or amino acid sequences, refer to two or more sequences or subsequences that are the same. Two sequences are “substantially aligned” if two sequences have a specified percentage of amino acid residues or nucleotides that are the same (*e.g.*, at least 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, 99.5%, 99.6%, 99.7%, 99.8% or 99.9% identical) over a specified region or over the entire sequence, when compared and aligned for maximum correspondence over a comparison window, or designated region as measured using one of the above sequence comparison algorithms or by manual alignment and visual inspection. Optionally, the alignment exists over a region that is at least about 25, 50, 75, or 100 amino acids in length, or over a region that is 100 to 150, 150 to 200, 100 to 200, or 200 or more amino acids in length.

In addition to polypeptide molecules, nucleic acid molecules possess a variety of advantageous properties for use as affinity reagents (*e.g.*, amino acid recognition molecules) in accordance with the application.

Nucleic acid aptamers are nucleic acid molecules that have been engineered to bind desired targets with high affinity and selectivity. Accordingly, nucleic acid aptamers may be engineered to selectively bind a desired type of amino acid using selection and/or enrichment techniques known in the art. Thus, in some embodiments, an affinity reagent comprises a nucleic acid aptamer (*e.g.*, a DNA aptamer, an RNA aptamer). In some embodiments, a labeled affinity reagent is a labeled aptamer that selectively binds one type of terminal amino acid. For example, in some embodiments, labeled aptamer selectively binds one type of amino acid (*e.g.*, a single type of amino acid or a subset of types of amino acids) at a terminus of a polypeptide, as described herein. Although not shown, it should be appreciated that labeled aptamer may be engineered to selectively bind one type of amino acid at any position of a polypeptide (*e.g.*, at a terminal position or at terminal and internal positions of a polypeptide) in accordance with a method of the application.

In some embodiments, a labeled affinity reagent comprises a label having binding-induced luminescence. For example, in some embodiments, a labeled aptamer comprises a donor label and an acceptor label and functions. In yet other embodiments, labeled aptamer comprises a quenching moiety and functions analogously to a molecular beacon, wherein luminescence of labeled aptamer is internally quenched as a free molecule and restored as a selectively bound molecule (see, *e.g.*, Hamaguchi, et al. (2001) Analytical Biochemistry 294, 126-131). Without wishing to be bound by theory, it is thought that these and other types of mechanisms for binding-induced luminescence may advantageously reduce or eliminate background luminescence to increase overall sensitivity and accuracy of the methods described herein.

In addition to methods of identifying a terminal amino acid of a polypeptide, the application provides methods of sequencing polypeptides using labeled affinity reagents. In some embodiments, methods of sequencing may involve subjecting a polypeptide terminus to repeated cycles of terminal amino acid detection and terminal amino acid cleavage. For example, in some embodiments, the application provides a method of determining an amino acid sequence of a polypeptide comprising contacting a polypeptide with one or more labeled affinity reagents described herein and subjecting the polypeptide to Edman degradation.

Conventional Edman degradation involves repeated cycles of modifying and cleaving the terminal amino acid of a polypeptide, wherein each successively cleaved amino acid is identified to determine an amino acid sequence of the polypeptide. As an illustrative example of a conventional Edman degradation, the N-terminal amino acid of a polypeptide is modified using phenyl isothiocyanate (PITC) to form a PITC-derivatized N-terminal amino acid. The PITC-derivatized N-terminal amino acid is then cleaved using acidic conditions, basic conditions,

and/or elevated temperatures. It has also been shown that the step of cleaving the PITC-derivatized N-terminal amino acid may be accomplished enzymatically using a modified cysteine protease from the protozoa *Trypanosoma cruzi*, which involves relatively milder cleavage conditions at a neutral or near-neutral pH. Non-limiting examples of useful enzymes are described in U.S. Pat. Application No. 15/255,433, filed September 2, 2016, titled “MOLECULES AND METHODS FOR ITERATIVE POLYPEPTIDE ANALYSIS AND PROCESSING”.

In some embodiments, sequencing by Edman degradation comprises providing a polypeptide that is immobilized to a surface of a solid support (*e.g.*, immobilized to a bottom or sidewall surface of a sample well) through a linker. In some embodiments, as described herein, polypeptide is immobilized at one terminus (*e.g.*, an amino-terminal amino acid or a carboxy-terminal amino acid) such that the other terminus is free for detecting and cleaving of a terminal amino acid. Accordingly, in some embodiments, the reagents used in Edman degradation methods described herein preferentially interact with terminal amino acids at the non-immobilized (*e.g.*, free) terminus of polypeptide. In this way, polypeptide remains immobilized over repeated cycles of detecting and cleaving. To this end, in some embodiments, linker may be designed according to a desired set of conditions used for detecting and cleaving, *e.g.*, to limit detachment of polypeptide from surface under chemical cleavage conditions. Suitable linker compositions and techniques for immobilizing a polypeptide to a surface are described in detail elsewhere herein.

In accordance with the application, in some embodiments, a method of sequencing by Edman degradation comprises a step (i) of contacting a polypeptide with one or more labeled affinity reagents that selectively bind one or more types of terminal amino acids. In some embodiments, a labeled affinity reagent interacts with the polypeptide by selectively binding the terminal amino acid. In some embodiments, step (i) further comprises removing any of the one or more labeled affinity reagents that do not selectively bind the terminal amino acid (*e.g.*, the free terminal amino acid) of polypeptide.

In some embodiments, the method further comprises identifying the terminal amino acid of the polypeptide by detecting labeled affinity reagent. In some embodiments, detecting comprises detecting a luminescence from labeled affinity reagent. As described herein, in some embodiments, the luminescence is uniquely associated with labeled affinity reagent, and the luminescence is thereby associated with the type of amino acid to which labeled affinity reagent selectively binds. As such, in some embodiments, the type of amino acid is identified by determining one or more luminescence properties of labeled affinity reagent.

In some embodiments, a method of sequencing by Edman degradation comprises a step (ii) of removing the terminal amino acid of the polypeptide. In some embodiments, step (ii) comprises removing labeled affinity reagent (*e.g.*, any of the one or more labeled affinity reagents that selectively bind the terminal amino acid) from the polypeptide. In some

5 embodiments, step (ii) comprises modifying the terminal amino acid (*e.g.*, the free terminal amino acid) of the polypeptide by contacting the terminal amino acid with an isothiocyanate (*e.g.*, PITC) to form an isothiocyanate-modified terminal amino acid. In some embodiments, an isothiocyanate-modified terminal amino acid is more susceptible to removal by a cleaving reagent (*e.g.*, a chemical or enzymatic cleaving reagent) than an unmodified terminal amino acid.

10 In some embodiments, step (ii) comprises removing the terminal amino acid by contacting the polypeptide with a protease that specifically binds and cleaves the isothiocyanate-modified terminal amino acid. In some embodiments, the protease comprises a modified cysteine protease. In some embodiments, the protease comprises a modified cysteine protease, such as a cysteine protease from *Trypanosoma cruzi* (see, *e.g.*, Borgo, et al. (2015) Protein

15 Science 24:571-579). In yet other embodiments, step (ii) comprises removing the terminal amino acid by subjecting the polypeptide to chemical (*e.g.*, acidic, basic) conditions sufficient to cleave the isothiocyanate-modified terminal amino acid.

In some embodiments, a method of sequencing by Edman degradation comprises a step (iii) of washing the polypeptide following terminal amino acid cleavage. In some embodiments,

20 washing comprises removing the protease. In some embodiments, washing comprises restoring the polypeptide to neutral pH conditions (*e.g.*, following chemical cleavage by acidic or basic conditions). In some embodiments, a method of sequencing by Edman degradation comprises repeating steps (i) through (iii) for a plurality of cycles.

In some embodiments, a sample containing a complex mixture or enriched mixture of

25 polypeptides (*e.g.*, a mixture of polypeptides) can be degraded using common enzymes into short polypeptide fragments of approximately 6 to 40 amino acids. In some embodiments, sequencing of this polypeptide library in accordance with methods of the application would reveal the identity and abundance of each of the polypeptides present in the original complex mixture or enriched mixture. As described herein and in the literature, most polypeptides in the size range

30 of 6 to 40 amino acids can be uniquely identified by determining the number and location of just four amino acids within a polypeptide chain.

Accordingly, in some embodiments, a method of sequencing by Edman degradation may be performed using a set of labeled aptamers comprising four DNA aptamer types, each type recognizing a different N-terminal amino acid. Each aptamer type may be labeled with a

35 different luminescent label, such that the different aptamer types can be distinguished based on

one or more luminescence properties. For illustrative purposes, the example set of labeled aptamers includes: a cysteine-specific aptamer labeled with a first luminescent label (“dye 1”); a lysine-specific aptamer labeled with a second luminescent label (“dye 2”); a tryptophan-specific aptamer labeled with a third luminescent label (“dye 3”); and a glutamate-specific aptamer
5 labeled with a fourth luminescent label (“dye 4”).

In some embodiments, prior to step (i), single polypeptide molecules from a polypeptide library are immobilized to a surface of a solid support, *e.g.*, at a bottom or sidewall surface of a sample well of an array of sample wells. In some embodiments, as described elsewhere herein, moieties that enable surface immobilization (*e.g.*, biotin) or improve solubility (*e.g.*,
10 oligonucleotides) may be chemically or enzymatically attached to the C-terminus of the polypeptides. To determine the sequence of each polypeptide, in some embodiments, immobilized polypeptides are subjected to repeated cycles of N-terminal amino acid detection and N-terminal amino acid cleavage. In some embodiments, the process comprises reagent addition and wash steps which are performed by injection into a flowcell above the detection
15 surface using an automated fluidic system. In some embodiments, steps (i) through (iv) illustrate one cycle of detection and cleavage using labeled aptamers.

In some embodiments, a method of sequencing by Edman degradation comprises a step (i) of flowing in a mixture of four orthogonally labeled DNA aptamers and incubating to allow the aptamers to bind to any immobilized polypeptides (*e.g.*, polypeptides immobilized within a
20 sample well of an array) that contain one of the four correct amino acids at the N-terminus. In some embodiments, the method further comprises washing the immobilized polypeptides to remove unbound aptamers. In some embodiments, the method further comprises imaging the immobilized polypeptides (“Imaging step (i)”). In some embodiments, the acquired images contain enough information to determine the location of aptamer-bound polypeptides (*e.g.*,
25 location within an array of sample wells) and which of the four aptamers is bound at each location. In some embodiments, the method further comprises washing the immobilized polypeptides using an appropriate buffer to remove the aptamers from the immobilized polypeptides.

In some embodiments, a method of sequencing comprises a step (ii) of flowing in a
30 solution containing a reactive molecule (*e.g.*, PITC, as shown) that specifically modifies the N-terminal amine group. An isothiocyanate molecule such as PITC, in some embodiments, modifies the N-terminal amino acid into a substrate for cleavage by a modified protease such as the cysteine protease cruzain from *Trypanosoma Cruzi*.

In some embodiments, a method of sequencing according comprises a step (iii) of washing the immobilized polypeptides before flowing in a suitable modified protease that recognizes and cleaves the modified N-terminal amino acid from the immobilized polypeptide.

In some embodiments, the method comprises a step (iv) of washing the immobilized polypeptides after enzymatic cleavage. In some embodiments, steps (i) through (iv) depict one cycle of Edman degradation. Accordingly, step (i') as shown is the start of the next reaction cycle which proceeds as steps (i') through (iv') performed as described above for steps (i) through (iv). In some embodiments, steps (i) through (iv) are repeated for approximately 20-40 cycles.

In some embodiments, a labeled isothiocyanate (*e.g.*, a dye-labeled PITC) may be used to monitor sample loading. For example, in some embodiments, prior to subjecting a polypeptide sample to a method of sequencing, the polypeptide sample is pre-conjugated with a luminescent label at a terminal end by modification of the terminal end using a dye-labeled PITC. In this way, loading of the polypeptide sample into an array of sample wells may be monitored by detecting luminescence from the labels prior to step (i) described above. In some embodiments, the luminescence is used to determine single occupancy of sample wells in the array (*e.g.*, a fraction of sample wells containing a single polypeptide molecule), which may advantageously increase the amount of information reliably obtained for a given sample. Once a desired sample loading status is determined by luminescence, chemical or enzymatic cleavage may be performed, as described, before proceeding with step (i).

In some embodiments, a labeled isothiocyanate (*e.g.*, a dye-labeled PITC) may be used to monitor reaction progress for a polypeptide sample in an array. For example, in some embodiments, step (ii) comprises flowing in a solution containing a dye-labeled PITC that specifically modifies and labels N-terminal amine groups of polypeptides in the sample. In some embodiments, luminescence from the labels may be detected during or after step (ii) to evaluate N-terminal PITC modification of polypeptides in the sample. Accordingly, in some embodiments, luminescence is used to determine whether or when to proceed from step (ii) to step (iii). In some embodiments, luminescence from the labels may be detected during or after step (iii) to evaluate N-terminal amino acid cleavage of polypeptides in the sample—*e.g.*, to determine whether or when to proceed from step (iii) to step (iv).

A method of sequencing may utilize separate reagents for detecting and cleaving a terminal amino acid of a polypeptide. Nonetheless, in some aspects, the application provides a method of sequencing in which a single reagent comprising a peptidase (such as a labeled exopeptidase that selectively binds and cleaves a different type of terminal amino acid) may be used for detecting and cleaving a terminal amino acid of a polypeptide.

Labeled exopeptidases may comprise a lysine-specific exopeptidase comprising a first luminescent label, a glycine-specific exopeptidase comprising a second luminescent label, an aspartate-specific exopeptidase comprising a third luminescent label, and a leucine-specific exopeptidase comprising a fourth luminescent label. In accordance with certain embodiments described herein, each of labeled exopeptidases selectively binds and cleaves its respective amino acid only when that amino acid is at an amino- or carboxy-terminus of a polypeptide. Accordingly, as sequencing by this approach proceeds from one terminus of a peptide toward the other, labeled exopeptidases are engineered or selected such that all reagents of the set will possess either aminopeptidase or carboxypeptidase activity.

In some aspects, the application provides methods of polypeptide sequencing in real-time by evaluating binding interactions of terminal amino acids with labeled amino acid recognition molecules (*e.g.*, labeled affinity reagents) and a labeled cleaving reagent (*e.g.*, a labeled non-specific exopeptidase). Without wishing to be bound by theory, a labeled affinity reagent selectively binds according to a binding affinity (K_D) defined by an association rate, or an “on” rate, of binding (k_{on}) and a dissociation rate, or an “off” rate, of binding (k_{off}). The rate constants k_{off} and k_{on} are the critical determinants of pulse duration (*e.g.*, the time corresponding to a detectable binding event) and interpulse duration (*e.g.*, the time between detectable binding events), respectively. In some embodiments, these rates can be engineered to achieve pulse durations and pulse rates (*e.g.*, the frequency of signal pulses) that give the best sequencing accuracy.

A sequencing reaction mixture may further comprise a labeled non-specific exopeptidase comprising a luminescent label that is different than that of labeled affinity reagent. In some embodiments, a labeled non-specific exopeptidase is present in the mixture at a concentration that is less than that of the labeled affinity reagent. In some embodiments, the labeled non-specific exopeptidase displays broad specificity such that it cleaves most or all types of terminal amino acids.

In some embodiments, terminal amino acid cleavage by a labeled non-specific exopeptidase gives rise to a signal pulse, and these events occur with lower frequency than the binding pulses of a labeled affinity reagent. In this way, amino acids of a polypeptide may be counted and/or identified in a real-time sequencing process. In some embodiments, a plurality of labeled affinity reagents may be used, each with a diagnostic pulsing pattern (*e.g.*, characteristic pattern) which may be used to identify a corresponding terminal amino acid. For example, in some embodiments, different characteristic patterns correspond to the association of more than one labeled affinity reagent with different types of terminal amino acids. As described herein, it should be appreciated that a single affinity reagent that associates with more than one type of

amino acid may be used in accordance with the application. Accordingly, in some embodiments, different characteristic patterns correspond to the association of one labeled affinity reagent with different types of terminal amino acids.

As detailed above, a real-time sequencing process can generally involve cycles of terminal amino acid recognition and terminal amino acid cleavage, where the relative occurrence of recognition and cleavage can be controlled by a concentration differential between a labeled affinity reagent and a labeled non-specific exopeptidase. In some embodiments, the concentration differential can be optimized such that the number of signal pulses detected during recognition of an individual amino acid provides a desired confidence interval for identification. For example, if an initial sequencing reaction provides signal data with too few signal pulses between cleavage events to permit determination of characteristic patterns with a desired confidence interval, the sequencing reaction can be repeated using a decreased concentration of non-specific exopeptidase relative to affinity reagent. The inventors have recognized further techniques for controlling real-time sequencing reactions, which may be used in combination with, or alternatively to, the concentration differential approach as described.

In some embodiments, a sequencing reaction involves cycles of temperature-dependent terminal amino acid recognition and terminal amino acid cleavage. Each cycle of the sequencing reaction may be carried out over two temperature ranges: a first temperature range (“T₁”) that is optimal for affinity reagent activity over exopeptidase activity (*e.g.*, to promote terminal amino acid recognition), and a second temperature range (“T₂”) that is optimal for exopeptidase activity over affinity reagent activity (*e.g.*, to promote terminal amino acid cleavage). The sequencing reaction may progress by alternating the reaction mixture temperature between the first temperature range T₁ (to initiate amino acid recognition) and the second temperature range T₂ (to initiate amino acid cleavage). Accordingly, progression of a temperature-dependent sequencing process is controllable by temperature, and alternating between different temperature ranges (*e.g.*, between T₁ and T₂) which may be carried through manual or automated processes. In some embodiments, affinity reagent activity (*e.g.*, binding affinity (K_D) for an amino acid) within the first temperature range T₁ as compared to the second temperature range T₂ is increased by at least 10-fold, at least 100-fold, at least 1,000-fold, at least 10,000-fold, at least 100,000-fold, or more. In some embodiments, exopeptidase activity (*e.g.*, rate of substrate conversion to cleavage product) within the second temperature range T₂ as compared to the first temperature range T₁ is increased by at least 2-fold, 10-fold, at least 25-fold, at least 50-fold, at least 100-fold, at least 1,000-fold, or more.

In some embodiments, the first temperature range T₁ is lower than the second temperature range T₂. In some embodiments, the first temperature range T₁ is between about 15

°C and about 40 °C (*e.g.*, between about 25 °C and about 35 °C, between about 15 °C and about 30 °C, between about 20 °C and about 30 °C). In some embodiments, the second temperature range T_2 is between about 40 °C and about 100 °C (*e.g.*, between about 50 °C and about 90 °C, between about 60 °C and about 90 °C, between about 70 °C and about 90 °C). In some
5 embodiments, the first temperature range T_1 is between about 20 °C and about 40 °C (*e.g.*, approximately 30 °C), and the second temperature range T_2 is between about 60 °C and about 100 °C (*e.g.*, approximately 80 °C).

In some embodiments, the first temperature range T_1 is higher than the second temperature range T_2 . In some embodiments, the first temperature range T_1 is between about 40
10 °C and about 100 °C (*e.g.*, between about 50 °C and about 90 °C, between about 60 °C and about 90 °C, between about 70 °C and about 90 °C). In some embodiments, the second temperature range T_2 is between about 15 °C and about 40 °C (*e.g.*, between about 25 °C and about 35 °C, between about 15 °C and about 30 °C, between about 20 °C and about 30 °C). In some
15 embodiments, the first temperature range T_1 is between about 60 °C and about 100 °C (*e.g.*, approximately 80 °C), and the second temperature range T_2 is between about 20 °C and about 40 °C (*e.g.*, approximately 30 °C).

In some embodiments, the application provides a luminescence-dependent sequencing process using luminescence-activated reagents. In some embodiments, a luminescence-dependent sequencing process involves cycles of luminescence-dependent amino acid
20 recognition and cleavage. Each cycle of the sequencing reaction may be carried out by exposing a sequencing reaction mixture to two different luminescent conditions: a first luminescent condition that is optimal for affinity reagent activity over exopeptidase activity (*e.g.*, to promote amino acid recognition), and a second luminescent condition that is optimal for exopeptidase activity over affinity reagent activity (*e.g.*, to promote amino acid cleavage). The sequencing
25 reaction progresses by alternating between exposing the reaction mixture to the first luminescent condition (to initiate amino acid recognition) and exposing the reaction mixture to the second luminescent condition (to initiate amino acid cleavage). By way of example and not limitation, in some embodiments, the two different luminescent conditions comprise a first wavelength and a second wavelength.

In some aspects, the application provides methods of polypeptide sequencing in real-time
30 by evaluating binding interactions of one or more labeled affinity reagents with terminal and internal amino acids and binding interactions of a labeled non-specific exopeptidase with terminal amino acids. In some embodiments, a labeled affinity reagent is used that selectively binds to and dissociates from one type of amino acid at both terminal and internal positions. The
35 selective binding gives rise to a series of pulses in signal output. In this approach, however, the

series of pulses occur at a rate that is determined by the number of the type of amino acid throughout the polypeptide. Accordingly, in some embodiments, the rate of pulsing corresponding to binding events would be diagnostic of the number of cognate amino acids currently present in the polypeptide.

5 A labeled non-specific peptidase may be present at a relatively lower concentration than the labeled affinity reagent, *e.g.*, to give optimal time windows in between cleavage events. Additionally, in certain embodiments, uniquely identifiable luminescent label of labeled non-specific peptidase would indicate when cleavage events have occurred. As the polypeptide undergoes iterative cleavage, the rate of pulsing corresponding to binding by the labeled affinity
10 reagent would drop in a step-wise manner whenever a terminal amino acid is cleaved by the labeled non-specific peptidase. Thus, in some embodiments, amino acids may be identified—and polypeptides thereby sequenced—in this approach based on a pulsing pattern and/or on the rate of pulsing that occurs within a pattern detected between cleavage events.

15 (ii) *Sequencing by Degradation of Labeled Polypeptides*

In some aspects, the application provides methods of sequencing a polypeptide by identifying a unique combination of amino acids corresponding to a known polypeptide sequence. In some embodiments, the method comprises detecting selectively labeled amino acids of a labeled polypeptide. In some embodiments, the labeled polypeptide comprises
20 selectively modified amino acids such that different amino acid types comprise different luminescent labels. As used herein, unless otherwise indicated, a labeled polypeptide refers to a polypeptide comprising one or more selectively labeled amino acid sidechains. Methods of selective labeling and details relating to the preparation and analysis of labeled polypeptides are known in the art (see, *e.g.*, Swaminathan, et al. PLoS Comput Biol. 2015, 11(2):e1004080).

25 As described herein, in some aspects, the application provides methods of sequencing a polypeptide by obtaining data during a polypeptide degradation process, and analyzing the data to determine portions of the data corresponding to amino acids that are sequentially exposed at a terminus of the polypeptide during the degradation process. In some embodiments, the portions of the data comprise a series of signal pulses indicative of association of one or more amino acid
30 recognition molecules with successive amino acids exposed at the terminus of the polypeptide (*e.g.*, during a degradation). In some embodiments, the series of signal pulses corresponds to a series of reversible single molecule binding interactions at the terminus of the polypeptide during the degradation process.

In some aspects, the polypeptide sequencing techniques described herein generate data
35 indicating how a polypeptide interacts with a binding means (*e.g.*, one or more amino acid

recognition molecules) while the polypeptide is being degraded by a cleaving means (*e.g.*, one or more cleaving reagents). As discussed above, the data can include a series of characteristic patterns corresponding to association events at a terminus of a polypeptide in between cleavage events at the terminus. In some embodiments, methods of sequencing described herein comprise contacting a single polypeptide molecule with a binding means and a cleaving means, where the binding means and the cleaving means are configured to achieve at least 10 association events prior to a cleavage event. In some embodiments, the means are configured to achieve the at least 10 association events between two cleavage events.

As described herein, in some embodiments, a plurality of single-molecule sequencing reactions are performed in parallel in an array of sample wells. In some embodiments, an array comprises between about 10,000 and about 1,000,000 sample wells. The volume of a sample well may be between about 10^{-21} liters and about 10^{-15} liters, in some implementations. Because the sample well has a small volume, detection of single-molecule events may be possible as only about one polypeptide may be within a sample well at any given time. Statistically, some sample wells may not contain a single-molecule sequencing reaction and some may contain more than one single polypeptide molecule. However, an appreciable number of sample wells may each contain a single-molecule reaction (*e.g.*, at least 30% in some embodiments), so that single-molecule analysis can be carried out in parallel for a large number of sample wells. In some embodiments, the binding means and the cleaving means are configured to achieve at least 10 association events prior to a cleavage event in at least 10% (*e.g.*, 10-50%, more than 50%, 25-75%, at least 80%, or more) of the sample wells in which a single-molecule reaction is occurring. In some embodiments, the binding means and the cleaving means are configured to achieve at least 10 association events prior to a cleavage event for at least 50% (*e.g.*, more than 50%, 50-75%, at least 80%, or more) of the amino acids of a polypeptide in a single-molecule reaction.

In some embodiments, a labeled polypeptide is immobilized and exposed to an excitation source. An aggregate luminescence from the labeled polypeptide may be detected and, in some embodiments, exposure to luminescence over time may result in a loss in detected signal due to luminescent label degradation (*e.g.*, degradation due to photobleaching). In some embodiments, the labeled polypeptide comprises a unique combination of selectively labeled amino acids that give rise to an initial detected signal. Degradation of luminescent labels over time results in a corresponding decrease in a detected signal for the photobleached labeled polypeptide. In some embodiments, the signal can be deconvoluted by analysis of one or more luminescence properties (*e.g.*, signal deconvolution by luminescence lifetime analysis). In some embodiments, the unique combination of selectively labeled amino acids of the labeled polypeptide have been computationally precomputed and empirically verified - *e.g.*, based on known polypeptide

sequences of a proteome. In some embodiments, the combination of detected amino acid labels are compared against a database of known sequences of a proteome of an organism to identify a particular polypeptide of the database corresponding to the labeled polypeptide.

In some embodiments, an optimal sample concentration is determined for performing a sequencing reaction that maximizes sampling in massively parallel analysis. In some
5 embodiments, the concentration is selected so that a desired fraction of the sample wells of an array (*e.g.*, 30%) are occupied at any given time. Without wishing to be bound by theory, it is thought that while a polypeptide is bleached over a period of time, the same well continues to be available for further analysis. Through diffusion, approximately 30% of the sample wells of an
10 array can be used for analysis every 3 minutes. As an illustrative example, in a million sample well chip, 6,000,000 polypeptides per hour may be sampled, or 24,000,000 over a 4 hour period.

In some aspects, the application provides a method of sequencing a polypeptide by detecting luminescence of a labeled polypeptide which is subjected to repeated cycles of terminal amino acid modification and cleavage. In some embodiments, the method generally proceeds as
15 described herein for other methods of sequencing by Edman degradation.

In some embodiments, the method comprises a step of (i) modifying the terminal amino acid of a labeled polypeptide. As described elsewhere herein, in some embodiments, modifying comprises contacting the terminal amino acid with an isothiocyanate (*e.g.*, PITC) to form an isothiocyanate-modified terminal amino acid. In some embodiments, an isothiocyanate
20 modification converts the terminal amino acid to a form that is more susceptible to removal by a cleaving reagent (*e.g.*, a chemical or enzymatic cleaving reagent, as described herein). Accordingly, in some embodiments, the method comprises a step of (ii) removing the modified terminal amino acid using chemical or enzymatic means detailed elsewhere herein for Edman degradation.

In some embodiments, the method comprises repeating steps (i) through (ii) for a plurality of cycles, during which luminescence of the labeled polypeptide is detected, and cleavage events corresponding to the removal of a labeled amino acid from the terminus may be detected as a decrease in detected signal. In some embodiments, no change in signal following step (ii) identifies an amino acid of unknown type. Accordingly, in some embodiments, partial
30 sequence information may be determined by evaluating a signal detected following step (ii) during each sequential round by assigning an amino acid type by a determined identity based on a change in detected signal or identifying an amino acid type as unknown based on no change in a detected signal.

In some aspects, a method of sequencing a polypeptide in accordance with the application
35 comprises sequencing by processive enzymatic cleavage of a labeled polypeptide. In some

embodiments, a labeled polypeptide is subjected to degradation using a modified processive exopeptidase that continuously cleaves a terminal amino acid from one terminus to another terminus. Exopeptidases are described in detail elsewhere herein. In some embodiments, a labeled polypeptide is subjected to degradation by an immobilized processive exopeptidase. In some embodiments, an immobilized labeled polypeptide is subjected to degradation by a processive exopeptidase.

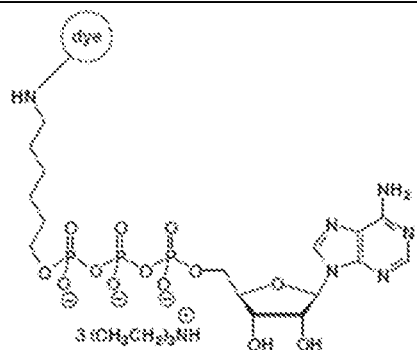
In some embodiments, the rate of processivity of processive exopeptidase is known, such that the timing between a detected decrease in signal may be used to calculate the number of unlabeled amino acids between each detection event. For example, if a polypeptide of 40 amino acids was cleaved in such a way that an amino acid was removed every second, a labeled polypeptide having 3 signals would show all 3 initially, then 2, then 1, and finally no signal. In this way, the order of the labeled amino acids can be determined. Accordingly, these methods may be used to determine partial sequence information, *e.g.*, for proteomic analysis based on polypeptide fragment sequencing.

In some embodiments, single molecule polypeptide sequencing can be achieved using an ATP-based Förster resonance energy transfer (FRET) scheme (*e.g.*, with one or more labeled cofactors). In some embodiments, sequencing by cofactor-based FRET can be performed using an immobilized ATP-dependent protease, donor-labeled ATP, and acceptor-labeled amino acids of a polypeptide substrate. In some embodiments, amino acids can be labeled with acceptors, and the one or more cofactors can be labeled with donors.

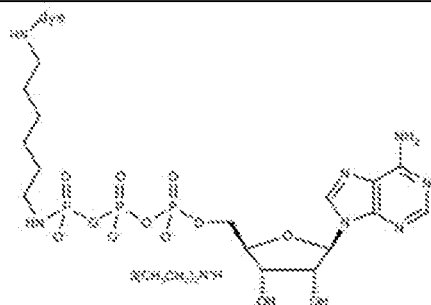
For example, in some embodiments, extracted polypeptides are denatured, and cysteines and lysines are labeled with fluorescent dyes. In some embodiments, an engineered version of a protein translocase (*e.g.*, bacterial ClpX) is used to bind to individual substrate polypeptides, unfold them, and translocate them through its nano-channel. In some embodiments, the translocase is labeled with a donor dye, and FRET occurs between the donor on the translocase and two or more distinct acceptor dyes on a substrate when the substrate passes through the nano-channel. The order of the labeled amino acids can then be determined from the FRET signal. In some embodiments, one or more of the following non-limiting labeled ATP analogues shown in Table 3 can be used.

TABLE 3. Non-limiting examples of labeled ATP analogues

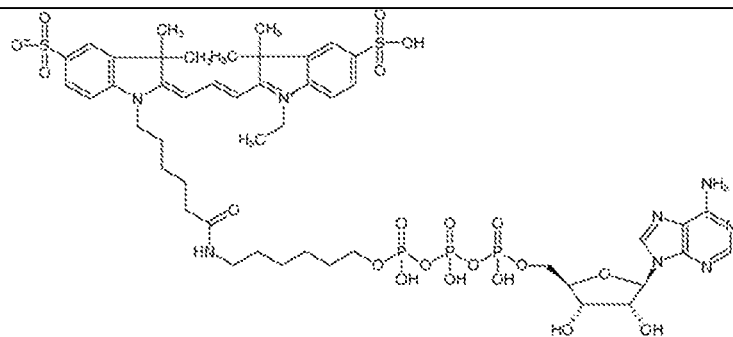
<i>Phosphate-labeled ATP:</i>



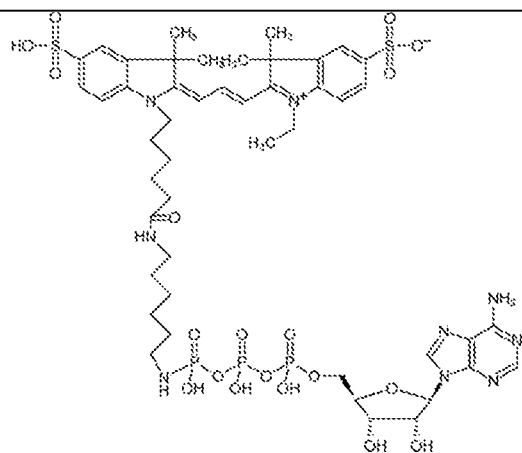
(γ-[(6-Amino)hexyl]-ATP)



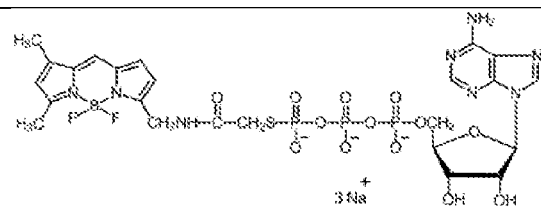
(γ-[(6-Amino)hexyl]imido)-ATP)



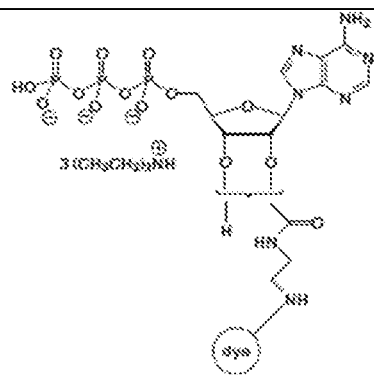
(γ-(6-Amino)hexyl)-ATP-Cy3)



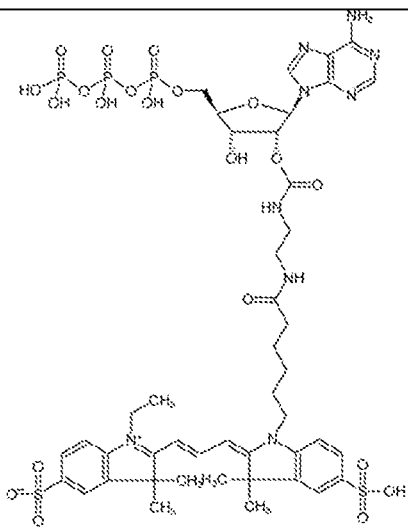
(γ-[(6-Amino)hexyl]imido)-ATP-Cy3)

(BODIPY FL ATP_γS)

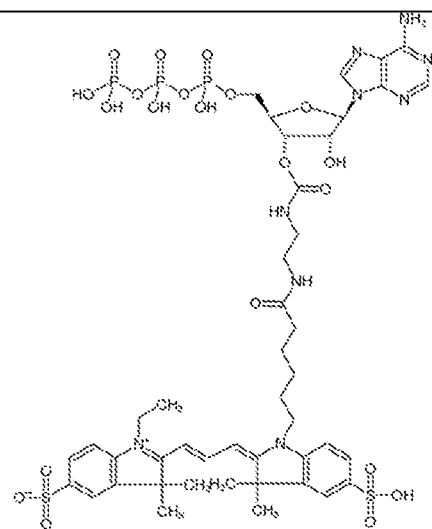
Ribose-labeled ATP:



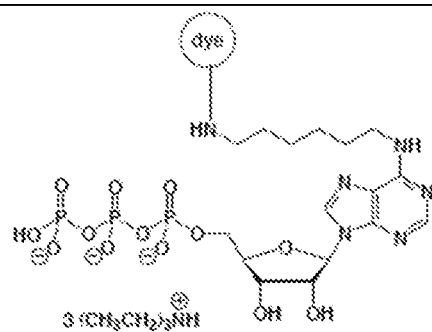
(EDA-ATP)

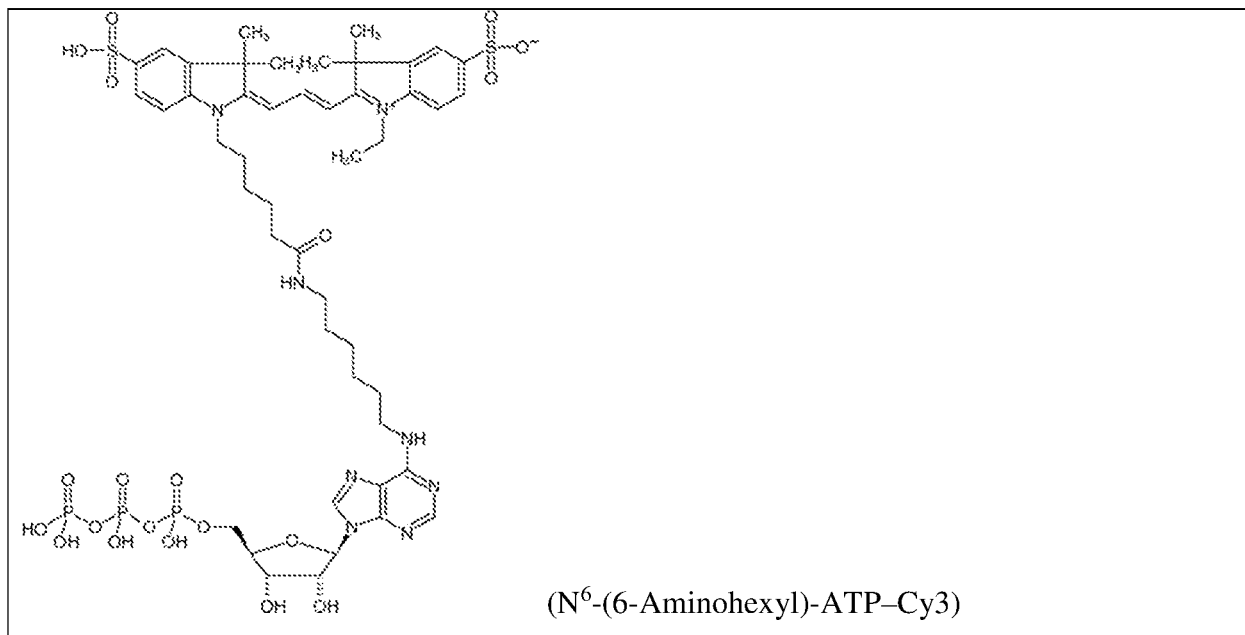


(EDA-ATP - Cy3)



(EDA-ATP - Cy3)

Base-labeled ATP:(N⁶-(6-Amino)hexyl-ATP)



(iii) Preparation of Samples for Sequencing

A polypeptide sample (*e.g.*, an enriched polypeptide sample) can be modified prior to sequencing.

5 In some embodiments, the N-terminal amino acid or the C-terminal amino acid of a polypeptide is modified. In some embodiments, a terminal end of a polypeptides is modified with moieties that enable immobilization to a surface (*e.g.*, a surface of a sample well on a chip used for polypeptide analysis). In some embodiments, such methods comprise modifying a terminal end of a labeled polypeptide to be analyzed in accordance with the application. In yet
10 other embodiments, such methods comprise modifying a terminal end of a protein or enzyme that degrades or translocates a polypeptide substrate in accordance with the application.

In some embodiments, a carboxy-terminus of a polypeptide is modified in a method comprising: (i) blocking free carboxylate groups of the polypeptide; (ii) denaturing the polypeptide (*e.g.*, by heat and/or chemical means); (iii) blocking free thiol groups of the
15 polypeptide; (iv) digesting the polypeptide to produce at least one polypeptide fragment comprising a free C-terminal carboxylate group; and (v) conjugating (*e.g.*, chemically) a functional moiety to the free C-terminal carboxylate group. In some embodiments, the method further comprises, after (i) and before (ii), dialyzing a sample comprising the polypeptide.

In some embodiments, a carboxy-terminus of a polypeptide is modified in a method
20 comprising: (i) denaturing the polypeptide (*e.g.*, by heat and/or chemical means); (ii) blocking free thiol groups of the polypeptide; (iii) digesting the polypeptide to produce at least one polypeptide fragment comprising a free C-terminal carboxylate group; (iv) blocking the free C-terminal carboxylate group to produce at least one polypeptide fragment comprising a blocked C-terminal carboxylate group; and (v) conjugating (*e.g.*, enzymatically) a functional moiety to

the blocked C-terminal carboxylate group. In some embodiments, the method further comprises, after (iv) and before (v), dialyzing a sample comprising the polypeptide.

In some embodiments, blocking free carboxylate groups refers to a chemical modification of these groups which alters chemical reactivity relative to an unmodified carboxylate. Suitable carboxylate blocking methods are known in the art and should modify side-chain carboxylate groups to be chemically different from a carboxy-terminal carboxylate group of a polypeptide to be functionalized. In some embodiments, blocking free carboxylate groups comprises esterification or amidation of free carboxylate groups of a polypeptide. In some embodiments, blocking free carboxylate groups comprises methyl esterification of free carboxylate groups of a polypeptide, *e.g.*, by reacting the polypeptide with methanolic HCl. Additional examples of reagents and techniques useful for blocking free carboxylate groups include, without limitation, 4-sulfo-2,3,5,6-tetrafluorophenol (STP) and/or a carbodiimide such as N-(3-Dimethylaminopropyl)-N'-ethylcarbodiimide hydrochloride (EDAC), uronium reagents, diazomethane, alcohols and acid for Fischer esterification, the use of N-hydroxysuccinimide (NHS) to form NHS esters (potentially as an intermediate to subsequent ester or amine formation), or reaction with carbonyldiimidazole (CDI) or the formation of mixed anhydrides, or any other method of modifying or blocking carboxylic acids, potentially through the formation of either esters or amides.

In some embodiments, blocking free thiol groups refers to a chemical modification of these groups which alters chemical reactivity relative to an unmodified thiol. In some embodiments, blocking free thiol groups comprises reducing and alkylating free thiol groups of a polypeptide. In some embodiments, reduction and alkylation is carried out by contacting a polypeptide with dithiothreitol (DTT) and one or both of iodoacetamide and iodoacetic acid. Examples of additional and alternative cysteine-reducing reagents which may be used are well known and include, without limitation, 2-mercaptoethanol, Tris (2-carboxyethyl) phosphine hydrochloride (TCEP), tributylphosphine, dithiobutylamine (DTBA), or any reagent capable of reducing a thiol group. Examples of additional and alternative cysteine-blocking (*e.g.*, cysteine-alkylating) reagents which may be used are well known and include, without limitation, acrylamide, 4-vinylpyridine, N-Ethylmaleimide (NEM), N- ϵ -maleimidocaproic acid (EMCA), or any reagent that modifies cysteines so as to prevent disulfide bond formation.

In some embodiments, digestion comprises enzymatic digestion. In some embodiments, digestion is carried out by contacting a polypeptide with an endopeptidase (*e.g.*, trypsin) under digestion conditions. In some embodiments, digestion comprises chemical digestion. Examples of suitable reagents for chemical and enzymatic digestion are known in the art and include, without limitation, trypsin, chymotrypsin, Lys-C, Arg-C, Asp-N, Lys-N, BNPS-Skatole, CNBr,

caspase, formic acid, glutamyl endopeptidase, hydroxylamine, iodosobenzoic acid, neutrophil elastase, pepsin, proline-endopeptidase, proteinase K, staphylococcal peptidase I, thermolysin, and thrombin.

In some embodiments, the functional moiety comprises a biotin molecule. In some
5 embodiments, the functional moiety comprises a reactive chemical moiety, such as an alkynyl. In some embodiments, conjugating a functional moiety comprises biotinylation of carboxy-terminal carboxy-methyl ester groups by carboxypeptidase Y, as known in the art.

In some embodiments, a solubilizing moiety is added to a polypeptide. Accordingly, in
10 some embodiments methods and compositions provided herein are useful for modifying terminal ends of polypeptides with moieties that increase their solubility. In some embodiments, a solubilizing moiety is useful for small polypeptides that result from fragmentation (*e.g.*, enzymatic fragmentation, for example using trypsin) and that are relatively insoluble. For example, in some embodiments, short polypeptides in a polypeptide pool can be solubilized by conjugating a polymer (*e.g.*, a short oligo, a sugar, or other charged polymer) to the polypeptides.

15 (iv) *Luminescent Labels*

As used herein, a luminescent label is a molecule that absorbs one or more photons and may subsequently emit one or more photons after one or more time durations. In some
20 embodiments, the term is used interchangeably with “label” or “luminescent molecule” depending on context. A luminescent label in accordance with certain embodiments described herein may refer to a luminescent label of a labeled affinity reagent, a luminescent label of a labeled peptidase (*e.g.*, a labeled exopeptidase, a labeled non-specific exopeptidase), a luminescent label of a labeled peptide, a luminescent label of a labeled cofactor, or another labeled composition described herein. In some embodiments, a luminescent label in accordance
25 with the application refers to a labeled amino acid of a labeled polypeptide comprising one or more labeled amino acids.

In some embodiments, a luminescent label may comprise a first and second
chromophore. In some embodiments, an excited state of the first chromophore is capable of relaxation via an energy transfer to the second chromophore. In some embodiments, the energy
30 transfer is a Förster resonance energy transfer (FRET). Such a FRET pair may be useful for providing a luminescent label with properties that make the label easier to differentiate from amongst a plurality of luminescent labels in a mixture. In yet other embodiments, a FRET pair comprises a first chromophore of a first luminescent label and a second chromophore of a second luminescent label. In certain embodiments, the FRET pair may absorb excitation energy in a
35 first spectral range and emit luminescence in a second spectral range.

In some embodiments, a luminescent label refers to a fluorophore or a dye. Typically, a luminescent label comprises an aromatic or heteroaromatic compound and can be a pyrene, anthracene, naphthalene, naphthylamine, acridine, stilbene, indole, benzindole, oxazole, carbazole, thiazole, benzothiazole, benzoxazole, phenanthridine, phenoxazine, porphyrin, quinoline, ethidium, benzamide, cyanine, carbocyanine, salicylate, anthranilate, coumarin, fluorescein, rhodamine, xanthene, or other like compound.

In some embodiments, a luminescent label comprises a dye selected from one or more of the following: 5/6-Carboxyrhodamine 6G, 5-Carboxyrhodamine 6G, 6-Carboxyrhodamine 6G, 6-TAMRA, Abberior® STAR 440SXP, Abberior® STAR 470SXP, Abberior® STAR 488, Abberior® STAR 512, Abberior® STAR 520SXP, Abberior® STAR 580, Abberior® STAR 600, Abberior® STAR 635, Abberior® STAR 635P, Abberior® STAR RED, Alexa Fluor® 350, Alexa Fluor® 405, Alexa Fluor® 430, Alexa Fluor® 480, Alexa Fluor® 488, Alexa Fluor® 514, Alexa Fluor® 532, Alexa Fluor® 546, Alexa Fluor® 555, Alexa Fluor® 568, Alexa Fluor® 594, Alexa Fluor® 610-X, Alexa Fluor® 633, Alexa Fluor® 647, Alexa Fluor® 660, Alexa Fluor® 680, Alexa Fluor® 700, Alexa Fluor® 750, Alexa Fluor® 790, AMCA, ATTO 390, ATTO 425, ATTO 465, ATTO 488, ATTO 495, ATTO 514, ATTO 520, ATTO 532, ATTO 542, ATTO 550, ATTO 565, ATTO 590, ATTO 610, ATTO 620, ATTO 633, ATTO 647, ATTO 647N, ATTO 655, ATTO 665, ATTO 680, ATTO 700, ATTO 725, ATTO 740, ATTO Oxa12, ATTO Rho101, ATTO Rho11, ATTO Rho12, ATTO Rho13, ATTO Rho14, ATTO Rho3B, ATTO Rho6G, ATTO Thio12, BD Horizon™ V450, BODIPY® 493/501, BODIPY® 530/550, BODIPY® 558/568, BODIPY® 564/570, BODIPY® 576/589, BODIPY® 581/591, BODIPY® 630/650, BODIPY® 650/665, BODIPY® FL, BODIPY® FL-X, BODIPY® R6G, BODIPY® TMR, BODIPY® TR, CAL Fluor® Gold 540, CAL Fluor® Green 510, CAL Fluor® Orange 560, CAL Fluor® Red 590, CAL Fluor® Red 610, CAL Fluor® Red 615, CAL Fluor® Red 635, Cascade® Blue, CF™350, CF™405M, CF™405S, CF™488A, CF™514, CF™532, CF™543, CF™546, CF™555, CF™568, CF™594, CF™620R, CF™633, CF™633-V1, CF™640R, CF™640R-V1, CF™640R-V2, CF™660C, CF™660R, CF™680, CF™680R, CF™680R-V1, CF™750, CF™770, CF™790, Chromeo™ 642, Chromis 425N, Chromis 500N, Chromis 515N, Chromis 530N, Chromis 550A, Chromis 550C, Chromis 550Z, Chromis 560N, Chromis 570N, Chromis 577N, Chromis 600N, Chromis 630N, Chromis 645A, Chromis 645C, Chromis 645Z, Chromis 678A, Chromis 678C, Chromis 678Z, Chromis 770A, Chromis 770C, Chromis 800A, Chromis 800C, Chromis 830A, Chromis 830C, Cy®3, Cy®3.5, Cy®3B, Cy®5, Cy®5.5, Cy®7, DyLight® 350, DyLight® 405, DyLight® 415-Co1, DyLight® 425Q, DyLight® 485-LS, DyLight® 488, DyLight® 504Q, DyLight® 510-LS, DyLight® 515-LS, DyLight® 521-LS, DyLight® 530-R2, DyLight® 543Q, DyLight® 550, DyLight® 554-R0, DyLight® 554-R1,

DyLight® 590-R2, DyLight® 594, DyLight® 610-B1, DyLight® 615-B2, DyLight® 633, DyLight® 633-B1, DyLight® 633-B2, DyLight® 650, DyLight® 655-B1, DyLight® 655-B2, DyLight® 655-B3, DyLight® 655-B4, DyLight® 662Q, DyLight® 675-B1, DyLight® 675-B2, DyLight® 675-B3, DyLight® 675-B4, DyLight® 679-C5, DyLight® 680, DyLight® 683Q, DyLight® 690-B1, DyLight® 690-B2, DyLight® 696Q, DyLight® 700-B1, DyLight® 700-B1, DyLight® 730-B1, DyLight® 730-B2, DyLight® 730-B3, DyLight® 730-B4, DyLight® 747, DyLight® 747-B1, DyLight® 747-B2, DyLight® 747-B3, DyLight® 747-B4, DyLight® 755, DyLight® 766Q, DyLight® 775-B2, DyLight® 775-B3, DyLight® 775-B4, DyLight® 780-B1, DyLight® 780-B2, DyLight® 780-B3, DyLight® 800, DyLight® 830-B2, Dyomics-350, Dyomics-350XL, Dyomics-360XL, Dyomics-370XL, Dyomics-375XL, Dyomics-380XL, Dyomics-390XL, Dyomics-405, Dyomics-415, Dyomics-430, Dyomics-431, Dyomics-478, Dyomics-480XL, Dyomics-481XL, Dyomics-485XL, Dyomics-490, Dyomics-495, Dyomics-505, Dyomics-510XL, Dyomics-511XL, Dyomics-520XL, Dyomics-521XL, Dyomics-530, Dyomics-547, Dyomics-547P1, Dyomics-548, Dyomics-549, Dyomics-549P1, Dyomics-550, Dyomics-554, Dyomics-555, Dyomics-556, Dyomics-560, Dyomics-590, Dyomics-591, Dyomics-594, Dyomics-601XL, Dyomics-605, Dyomics-610, Dyomics-615, Dyomics-630, Dyomics-631, Dyomics-632, Dyomics-633, Dyomics-634, Dyomics-635, Dyomics-636, Dyomics-647, Dyomics-647P1, Dyomics-648, Dyomics-648P1, Dyomics-649, Dyomics-649P1, Dyomics-650, Dyomics-651, Dyomics-652, Dyomics-654, Dyomics-675, Dyomics-676, Dyomics-677, Dyomics-678, Dyomics-679P1, Dyomics-680, Dyomics-681, Dyomics-682, Dyomics-700, Dyomics-701, Dyomics-703, Dyomics-704, Dyomics-730, Dyomics-731, Dyomics-732, Dyomics-734, Dyomics-749, Dyomics-749P1, Dyomics-750, Dyomics-751, Dyomics-752, Dyomics-754, Dyomics-776, Dyomics-777, Dyomics-778, Dyomics-780, Dyomics-781, Dyomics-782, Dyomics-800, Dyomics-831, eFluor® 450, Eosin, FITC, Fluorescein, HiLyte™ Fluor 405, HiLyte™ Fluor 488, HiLyte™ Fluor 532, HiLyte™ Fluor 555, HiLyte™ Fluor 594, HiLyte™ Fluor 647, HiLyte™ Fluor 680, HiLyte™ Fluor 750, IRDye® 680LT, IRDye® 750, IRDye® 800CW, JOE, LightCycler® 640R, LightCycler® Red 610, LightCycler® Red 640, LightCycler® Red 670, LightCycler® Red 705, Lissamine Rhodamine B, Naphthofluorescein, Oregon Green® 488, Oregon Green® 514, Pacific Blue™, Pacific Green™, Pacific Orange™, PET, PF350, PF405, PF415, PF488, PF505, PF532, PF546, PF555P, PF568, PF594, PF610, PF633P, PF647P, Quasar® 570, Quasar® 670, Quasar® 705, Rhodamine 123, Rhodamine 6G, Rhodamine B, Rhodamine Green, Rhodamine Green-X, Rhodamine Red, ROX, Seta™ 375, Seta™ 470, Seta™ 555, Seta™ 632, Seta™ 633, Seta™ 650, Seta™ 660, Seta™ 670, Seta™ 680, Seta™ 700, Seta™ 750, Seta™ 780, Seta™ APC-780, Seta™ PerCP-680, Seta™ R-PE-670, Seta™ 646, SeTau 380, SeTau 425, SeTau 647, SeTau 405, Square 635,

Square 650, Square 660, Square 672, Square 680, Sulforhodamine 101, TAMRA, TET, Texas Red®, TMR, TRITC, Yakima Yellow™, Zenon®, Zy3, Zy5, Zy5.5, and Zy7.

(v). *Luminescence*

5 In some aspects, the application relates to polypeptide sequencing and/or identification based on one or more luminescence properties of a luminescent label. In some embodiments, a luminescent label is identified based on luminescence lifetime, luminescence intensity, brightness, absorption spectra, emission spectra, luminescence quantum yield, or a combination of two or more thereof. In some embodiments, a plurality of types of luminescent labels can be distinguished from each other based on different luminescence lifetimes, luminescence
10 intensities, brightnesses, absorption spectra, emission spectra, luminescence quantum yields, or combinations of two or more thereof. Identifying may mean assigning the exact identity and/or quantity of one type of amino acid (*e.g.*, a single type or a subset of types) associated with a luminescent label, and may also mean assigning an amino acid location in a polypeptide relative to other types of amino acids.
15

In some embodiments, luminescence is detected by exposing a luminescent label to a series of separate light pulses and evaluating the timing or other properties of each photon that is emitted from the label. In some embodiments, information for a plurality of photons emitted sequentially from a label is aggregated and evaluated to identify the label and thereby identify an
20 associated type of amino acid. In some embodiments, a luminescence lifetime of a label is determined from a plurality of photons that are emitted sequentially from the label, and the luminescence lifetime can be used to identify the label. In some embodiments, a luminescence intensity of a label is determined from a plurality of photons that are emitted sequentially from the label, and the luminescence intensity can be used to identify the label. In some
25 embodiments, a luminescence lifetime and luminescence intensity of a label is determined from a plurality of photons that are emitted sequentially from the label, and the luminescence lifetime and luminescence intensity can be used to identify the label.

In some aspects of the application, a single polypeptide molecule is exposed to a plurality of separate light pulses and a series of emitted photons are detected and analyzed. In some
30 embodiments, the series of emitted photons provides information about the single polypeptide molecule that is present and that does not change in the reaction sample over the time of the experiment. However, in some embodiments, the series of emitted photons provides information about a series of different molecules that are present at different times in the reaction sample (*e.g.*, as a reaction or process progresses). By way of example and not limitation, such

information may be used to sequence and/or identify a polypeptide subjected to chemical or enzymatic degradation in accordance with the application.

In certain embodiments, a luminescent label absorbs one photon and emits one photon after a time duration. In some embodiments, the luminescence lifetime of a label can be determined or estimated by measuring the time duration. In some embodiments, the luminescence lifetime of a label can be determined or estimated by measuring a plurality of time durations for multiple pulse events and emission events. In some embodiments, the luminescence lifetime of a label can be differentiated amongst the luminescence lifetimes of a plurality of types of labels by measuring the time duration. In some embodiments, the luminescence lifetime of a label can be differentiated amongst the luminescence lifetimes of a plurality of types of labels by measuring a plurality of time durations for multiple pulse events and emission events. In certain embodiments, a label is identified or differentiated amongst a plurality of types of labels by determining or estimating the luminescence lifetime of the label. In certain embodiments, a label is identified or differentiated amongst a plurality of types of labels by differentiating the luminescence lifetime of the label amongst a plurality of the luminescence lifetimes of a plurality of types of labels.

Determination of a luminescence lifetime of a luminescent label can be performed using any suitable method (*e.g.*, by measuring the lifetime using a suitable technique or by determining time-dependent characteristics of emission). In some embodiments, determining the luminescence lifetime of one label comprises determining the lifetime relative to another label. In some embodiments, determining the luminescence lifetime of a label comprises determining the lifetime relative to a reference. In some embodiments, determining the luminescence lifetime of a label comprises measuring the lifetime (*e.g.*, fluorescence lifetime). In some embodiments, determining the luminescence lifetime of a label comprises determining one or more temporal characteristics that are indicative of lifetime. In some embodiments, the luminescence lifetime of a label can be determined based on a distribution of a plurality of emission events (*e.g.*, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 30, 40, 50, 60, 70, 80, 90, 100, or more emission events) occurring across one or more time-gated windows relative to an excitation pulse. For example, a luminescence lifetime of a label can be distinguished from a plurality of labels having different luminescence lifetimes based on the distribution of photon arrival times measured with respect to an excitation pulse.

It should be appreciated that a luminescence lifetime of a luminescent label is indicative of the timing of photons emitted after the label reaches an excited state and the label can be distinguished by information indicative of the timing of the photons. Some embodiments may include distinguishing a label from a plurality of labels based on the luminescence lifetime of the

label by measuring times associated with photons emitted by the label. The distribution of times may provide an indication of the luminescence lifetime which may be determined from the distribution. In some embodiments, the label is distinguishable from the plurality of labels based on the distribution of times, such as by comparing the distribution of times to a reference
5 distribution corresponding to a known label. In some embodiments, a value for the luminescence lifetime is determined from the distribution of times.

As used herein, in some embodiments, luminescence intensity refers to the number of emitted photons per unit time that are emitted by a luminescent label which is being excited by delivery of a pulsed excitation energy. In some embodiments, the luminescence intensity refers
10 to the detected number of emitted photons per unit time that are emitted by a label which is being excited by delivery of a pulsed excitation energy, and are detected by a particular sensor or set of sensors.

As used herein, in some embodiments, brightness refers to a parameter that reports on the average emission intensity per luminescent label. Thus, in some embodiments, “emission
15 intensity” may be used to generally refer to brightness of a composition comprising one or more labels. In some embodiments, brightness of a label is equal to the product of its quantum yield and extinction coefficient.

As used herein, in some embodiments, luminescence quantum yield refers to the fraction of excitation events at a given wavelength or within a given spectral range that lead to an
20 emission event, and is typically less than 1. In some embodiments, the luminescence quantum yield of a luminescent label described herein is between 0 and about 0.001, between about 0.001 and about 0.01, between about 0.01 and about 0.1, between about 0.1 and about 0.5, between about 0.5 and 0.9, or between about 0.9 and 1. In some embodiments, a label is identified by determining or estimating the luminescence quantum yield.

As used herein, in some embodiments, an excitation energy is a pulse of light from a light
25 source. In some embodiments, an excitation energy is in the visible spectrum. In some embodiments, an excitation energy is in the ultraviolet spectrum. In some embodiments, an excitation energy is in the infrared spectrum. In some embodiments, an excitation energy is at or near the absorption maximum of a luminescent label from which a plurality of emitted photons
30 are to be detected. In certain embodiments, the excitation energy is between about 500 nm and about 700 nm (*e.g.*, between about 500 nm and about 600 nm, between about 600 nm and about 700 nm, between about 500 nm and about 550 nm, between about 550 nm and about 600 nm, between about 600 nm and about 650 nm, or between about 650 nm and about 700 nm). In
35 certain embodiments, an excitation energy may be monochromatic or confined to a spectral range. In some embodiments, a spectral range has a range of between about 0.1 nm and about 1

nm, between about 1 nm and about 2 nm, or between about 2 nm and about 5 nm. In some embodiments, a spectral range has a range of between about 5 nm and about 10 nm, between about 10 nm and about 50 nm, or between about 50 nm and about 100 nm.

5 B. Polynucleic Acid Sequencing and Detection/Quantification Methodologies

In some aspects, the disclosure relates to methods of sequencing polynucleic acids (e.g., DNA, RNA, cDNA, etc.). In some embodiments, a method of polynucleic acid sequencing comprises the steps of: (i) exposing a complex in a target volume to one or more labeled nucleotides, the complex comprising a target polynucleic acid or a plurality of polynucleic acids present in a sample, at least one primer, and a polymerizing enzyme; (ii) directing one or more excitation energies, or a series of pulses of one or more excitation energies, towards a vicinity of the target volume; (iii) detecting a plurality of emitted photons from the one or more labeled nucleotides during sequential incorporation into a polynucleic acid comprising one of the at least one primers; and (iv) identifying the sequence of incorporated nucleotides by determining one or more characteristics of the emitted photons.

In some embodiments, a primer is a sequencing primer. In some embodiments, a sequencing primer can be annealed to a polynucleic acid (e.g., a target polynucleic acid) that may or may not be immobilized to a solid support. A solid support can comprise, for example, a sample well (e.g., a nanoaperture, a reaction chamber) on a chip or cartridge used for polynucleic acid sequencing. In some embodiments, a sequencing primer may be immobilized to a solid support and hybridization of the polynucleic acid (e.g., the target nucleic acid) further immobilizes the nucleic acid molecule to the solid support. In some embodiments, a polymerase (e.g., RNA Polymerase) is immobilized to a solid support and soluble sequencing primer and polynucleic acid are contacted to the polymerase. In some embodiments a complex comprising a polymerase, a polynucleic acid (e.g., a target nucleic acid) and a primer is formed in solution and the complex is immobilized to a solid support (e.g., via immobilization of the polymerase, primer, and/or target polynucleic acid). In some embodiments, none of the components are immobilized to a solid support. For example, in some embodiments, a complex comprising a polymerase, a target polynucleic acid, and a sequencing primer is formed in situ and the complex is not immobilized to a solid support.

In some embodiments, a plurality of single molecule sequencing reactions are performed in parallel (e.g., on a single chip or cartridge) according to aspects of the instant disclosure. For example, in some embodiments, a plurality of single molecule sequencing reactions are each performed in separate sample wells (e.g., nanoapertures, reaction chambers) on a single chip or cartridge.

In some embodiments, the disclosure provides methods of sequencing target nucleic acids or a plurality of target nucleic acids present in a sample by sequencing a plurality of nucleic acid fragments, wherein the target nucleic acid(s) comprises the fragments. In certain embodiments, the method comprises combining a plurality of fragment sequences to provide a sequence or
5 partial sequence for the parent nucleic acid (e.g., parent target nucleic acid). In some embodiments, the step of combining is performed by computer hardware and software. The methods described herein may allow for a set of related nucleic acids (e.g., two or more nucleic acids present in a sample), such as an entire chromosome or genome to be sequenced.

In some embodiments, sequencing by synthesis methods can include the presence of a
10 population of target nucleic acid molecules (e.g., copies of a target nucleic acid) and/or a step of amplification (e.g., polymerase chain reaction (PCR)) of a target nucleic acid to achieve a population of target nucleic acids. However, in some embodiments, sequencing by synthesis is used to determine the sequence of a single nucleic acid molecule in any one reaction that is being evaluated and nucleic acid amplification may not be required to prepare the target nucleic acid.

15 In some embodiments, a plurality of single molecule sequencing reactions are performed in parallel (e.g., on a single chip or cartridge) according to aspects of the instant disclosure. For example, in some embodiments, a plurality of single molecule sequencing reactions are each performed in separate sample wells (e.g., nanoapertures, reaction chambers) on a single chip or cartridge.

20 In some embodiments, sequencing of a target nucleic acid molecule comprises identifying at least two (e.g., at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at least 17, at least 18, at least 19, at least 20, at least 25, at least 30, at least 35, at least 40, at least 45, at least 50, at least 60, at least 70, at least 80, at least 90, at least 100, or more) nucleotides of the target
25 nucleic acid. In some embodiments, the at least two nucleotides are contiguous nucleotides. In some embodiments, the at least two nucleotides are non-contiguous nucleotides.

In some embodiments, sequencing of a target nucleic acid comprises identification of less than 100% (e.g., less than 99%, less than 95%, less than 90%, less than 85%, less than 80%, less than 75%, less than 70%, less than 65%, less than 60%, less than 55%, less than 50%, less than
30 45%, less than 40%, less than 35%, less than 30%, less than 25%, less than 20%, less than 15%, less than 10%, less than 5%, less than 1% or less) of all nucleotides in the target nucleic acid. For example, in some embodiments, sequencing of a target nucleic acid comprises identification of less than 100% of one type of nucleotide in the target nucleic acid. In some embodiments, sequencing of a target nucleic acid comprises identification of less than 100% of each type of
35 nucleotide in the target nucleic acid.

In some embodiments, methods of polynucleic acid sequencing comprise or enable long-read sequencing applications. In some embodiments, long-read sequencing applications involve sequencing of nucleic acids having a length of up to and about 10+ kilobases. In some embodiments, target nucleic acids for long-read sequencing applications have a length of about 0.5-2 kb, 0.5-5 kb, 1-2 kb, 1-3 kb, 1-4 kb, 1-5 kb, 1-10 kb, 2-10 kb, 2-5 kb, 5-10 kb, 5-15 kb, 5-20 kb, 5-25 kb, 10-15 kb, 10-20 kb, or 10-25 kb. In some embodiments, target nucleic acids for long-read sequencing applications comprise at least 700, 800, 900, 1000, 1100, 1200, 1300, 1400, 1500, 1600, 1700, 1800, 1900, 2000, 2100, 2200, 2300, 2400, 2500, 2600, 2700, 2800, 2900, or 3000 nucleotides in length. In some embodiments, target nucleic acids for long-read sequencing applications comprise 700-3000, 1000-3000, 1000-2500, 1000-2400, 1000-2300, 1000-2200, 1000-2100, 1000-2000, 1000-1900, 1000-1800, 1000-1700, 1000-1600, 1000-1500, 1000-1400, 1000-1300, 1000-1200, 1500-3000, 1500-2500, 1500-2000, or 2000-3000 nucleotides in length.

In some embodiments, long-read sequencing applications may be combined with short-read sequencing applications (*e.g.*, hybrid assembly). Long-read target nucleic acids can enable assembly of a series of short-read nucleic acids into a single contig or nucleic acid scaffold. Hybrid assembly, in some embodiments, allows for multiple long-read sequences to be aligned, thereby enabling the identification of sequence overlaps or gaps that can be ‘stitched’ together using short-read sequences.

Additional polynucleic acid sequencing methodologies are known to those having skill in the art.

C. Metabolite Detection/Quantification Methodologies

Methods of metabolite detection/quantification (*i.e.*, metabolite profiling) are known to those having ordinary skill in the art and include, but are not limited to, mass spectrometry (*e.g.*, LC-MS, GC-MS, diMS, etc.) and NMR (*e.g.*, LC-NMR).

V. **Kits for Sample Preparation**

In some aspects, the disclosure relates to kits for preparing a sample (*e.g.*, a multiplexed sample). A kit may be sufficient to prepare one or more samples (*e.g.*, multiplexed samples). In some embodiments, a kit is sufficient to prepare a single sample. In other embodiments, a kit is sufficient to prepare, at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 20, at least 25, at least 30, at least 40, at least 50, at least 60, at least 70, at least 80, at least 90, or at least 100 samples.

In some embodiments, a kit comprises a barcode component comprising a plurality of barcode molecules, as described herein. See “Methods of Preparing a Multiplexed Sample.” In some embodiments, a kit comprises one or more detector molecules, as described herein. See “Methods of Preparing a Multiplexed Sample.” In some embodiments, a kit comprises a solid support that allows for the physical separation of population of molecules of different origins, as described herein. See “Methods of Preparing a Multiplexed Sample.” In some embodiments, a kit comprises an enrichment component comprising a plurality of enrichment molecules, as described herein. See “Methods of Polypeptide Enrichment.” In some embodiments, a kit comprises a modifying agent, as described herein. See “Methods of Polypeptide Enrichment.”

10 In some embodiments, a kit comprises an affinity reagent, as described herein. See “Polypeptide Sequencing Methodologies.” In some embodiments, a kit comprises a labeled peptidase, as described herein. See “Sequencing Methodologies”.

A kit may be specific for one or more organisms (e.g., one or more single-cellular and/or multicellular organisms). In some embodiments, a kit comprises components (e.g., barcode molecules, detector molecules, enrichment molecules, or a combination thereof) that modify, bind to, are bound by, etc., polypeptides of one or more organisms. For example, in some embodiments, a kit comprises components that modify, bind to, are bound by, etc., one or more known polypeptides in the human proteome.

In some embodiments, a kit is specific for one or more disease or condition. For example, a kit may be an oncology kit, a cardiology kit, an inherited disease kit, or a combination thereof.

An oncology kit may comprise enrichment molecules that bind to (or are bound by) the amino acid sequence or the nucleotide sequence of ABL1, ABL2, ACSL3, ACVR2A, ADAMTS20, ADGRA2, ADGRB3, ADGRL3, AFF1, AFF3, AKAP9, AKT1, AKT2, AKT3, ALK, AMER1, APC, AR, ARID1A, ARID2, ARNT, ASXL1, ATF1, ATM, ATR, ATRX, AURKA, AURKB, AURKC, AXL, BAP1, BCL10, BCL11A, BCL11B, BCL2, BCL2L1, BCL2L2, BCL3, BCL6, BCL7A, BCL9, BCR, BIRC2, BIRC3, BIRC5, BLM, BLNK, BMPR1A, BRAF, BRCA1, BRCA2, BRD3, BRIP1, BTK, BUB1B, CACNA1D, CARD11, CASC5, CASP8, CBFA2T3, CBF3, CBL, CCND1, CCND2, CCNE1, CD79A, CD79B, CDC73, CDH1, CDH11, CDH2, CDH20, CDH5, CDK12, CDK4, CDK6, CDK8, CDKN2A, CDKN2B, CDKN2C, CEBPA, CHEK1, CHEK2, CIC, CKS1B, CMPK1, COL1A1, CRBN, CREB1, CREBBP, CRKL, CRLF2, CRTC1, CSF1R, CSMD3, CTNNA1, CTNNB1, CYLD, CYP2C19, CYP2D6, DAXX, DCC, DDB2, DDIT3, DDR2, DEK, DICER1, DNMT3A, DPYD, DST, EGFR, EML4, EP300, EP400, EPHA3, EPHA7, EPHB1, EPHB4, EPHB6, ERBB2, ERBB3, ERBB4, ERCC1, ERCC2, ERCC3, ERCC4, ERCC5, ERG, ESR1, ETS1, ETV1,

ETV4, EXT1, EXT2, EZH2, FANCA, FANCC, FANCD2, FANCF, FANCG, FAS, FBXW7, FCGR2B, FGFR1, FGFR2, FGFR3, FGFR4, FH, FLCN, FLI1, FLT1, FLT3, FLT4, FN1, FOXA1, FOXL2, FOXO1, FOXO3, FOXP1, FOXP4, FZR1, G6PD, GATA1, GATA2, GATA3, GDNF, GNA11, GNAQ, GNAS, GPC3, GRM8, GUCY1A2, HCAR1, HEY1, HIF1A, HIST1H3B, HLF, HMGA1, HNF1A, HOOK3, HOXA13, HOXD11, HRAS, HSP90AA1, HSP90AB1, ICK, IDH1, IDH2, IGF1R, IGF2, IGF2R, IKBKB, IKBKE, IKZF1, IL2, IL21R, IL6ST, IL7R, ING4, IRF4, IRS2, ITGA10, ITGA9, ITGB2, ITGB3, JAK1, JAK2, JAK3, JUN, KAT6A, KAT6B, KDM5C, KDM6A, KDR, KEAP1, KIAA1549, KIT, KLF6, KMT2A, KMT2C, KMT2D, KRAS, LAMP1, LCK, LIFR, LPP, LRP1B, LTF, LTK, MAF, MAFB, MAGEA1, MAGI1, MALT1, MAML2, MAP2K1, MAP2K2, MAP2K4, MAP3K7, MAPK1, MAPK8, MARK1, MARK4, MBD1, MCL1, MDM2, MDM4, MEN1, MET, MITF, MLH1, MLLT10, MLLT4, MLLT6, MMP2, MN1, MPL, MRE11A, MSH2, MSH6, MTCP1, MTOR, MTR, MTRR, MUC1, MUTYH, MYB, MYC, MYCL, MYCN, MYD88, MYH11, MYH9, NBN, NCOA1, NCOA2, NCOA4, NF1, NF2, NFE2L2, NFKB1, NFKB2, NIN, NKX2-1, NLRP1, NOTCH1, NOTCH2, NOTCH4, NPM1, NR4A3, NRAS, NSD1, NTRK1, NTRK3, NUMA1, NUP214, NUP98, NUTM2A, NUTM2B, OMD, P2RY8, PAK3, PALB2, PARP1, PAX3, PAX5, PAX7, PAX8, PBRM1, PBX1, PDE4DIP, PDGFB, PDGFRA, PDGFRB, PER1, PGAP3, PHOX2B, PIK3C2B, PIK3CA, PIK3CB, PIK3CD, PIK3CG, PIK3R1, PIK3R2, PIM1, PKHD1, PLAG1, PLCG1, PLEKHG5, PML, PMS1, PMS2, POT1, POU5F1, PPARG, PPP2R1A, PRDM1, PRKAR1A, PRKDC, PSIP1, PTCH1, PTEN, PTGS2, PTPN11, PTPRD, PTPRT, RAD50, RAF1, RALGDS, RAP1GDS1, RARA, RB1, RECQL4, REL, RET, RHOH, RNASEL, RNF2, RNF213, ROS1, RPS6KA2, RRM1, RUNX1, RUNX1T1, SAMD9, SBDS, SDHA, SDHB, SDHC, SDHD, SET, SETBP1, SETD2, SF3B1, SGK1, SH2D1A, SH3GL1, SMAD2, SMAD4, SMARCA4, SMARCB1, SMO, SMUG1, SOCS1, SOX11, SOX2, SRC, SSX1, SSX2, SSX4, STAT5B, STK11, STK36, SUFU, SYK, SYNE1, TAF1, TAF1L, TAL1, TBL1XR1, TBX22, TCF12, TCF3, TCF7L1, TCF7L2, TCL1A, TERT, TET1, TET2, TFE3, TGFBR2, TGM7, THBS1, TIMP3, TLR4, TLX1, TMPRSS2, TNFAIP3, TNFRSF14, TNK2, TOP1, TP53, TPR, TRIM24, TRIM33, TRIP11, TRRAP, TSC1, TSC2, TSHR, TTL, UBR5, UGT1A1, USP9X, VHL, WAS, WHSC1, WRN, WT1, XPA, XPC, XPO1, XRCC2, ZNF384, ZNF521, or any combination thereof.

A cardiology kit may comprise enrichment molecules that bind to (or are bound by) the amino acid sequence or the nucleotide sequence of ABCC9, ABCG5, ABCG8, ACTA1, ACTA2, ACTC1, ACTN2, AKAP9, ALMS1, ANK2, ANKRD1, APOA4, APOA5, APOB, APOC2, APOE, BAG3, BRAF, CACNA1C, CACNA2D1, CACNB2, CALM1, CALR3, CASQ2, CAV3, CBL, CBS, CETP, COL3A1, COL5A1, COL5A2, COX15, CREB3L3,

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PEX1, PEX10, PEX13, PEX14, PEX19, PEX26, PEX3, PEX5, PINK1, PKD1, PKD2, PKHD1, PKP2, PLEC, PLN, PLOD1, PMM2, PMP22, POLG, PPT1, PRCD, PRKAG2, PROM1, PRPF31, PRPF8, PRPH2, PSEN1, PSEN2, PTCH1, PTPN11, RAF1, RAG1, RAG2, RAI1, RAPSN, RB1, RDH12, RET, RHO, ROR2, RP9, RPE65, RPGR, RPGRIP1, RPL11, RPL35A, RPS10, RPS19, RPS24, RPS26, RPS6KA3, RPS7, RS1, RSPH4A, RSPH9, RYR1, RYR2, SALL4, SCN1B, SCN3B, SCN4B, SCN5A, SCN9A, SEMA4A, SERPINA1, SERPING1, SGCD, SH3BP2, SIX1, SIX5, SLC25A13, SLC25A4, SLC26A4, SMAD3, SMAD4, SNCA, SNRNP200, SNTA1, SOD1, SOS1, SOX9, SPATA7, SPG7, STARD3, TAF1, TAZ, TBX5, TCOF1, TGFBR1, TGFBR2, TMEM43, TNNC1, TNNI3, TNNT1, TNNT2, TNXB, TOPORS, TP53, TPM1, TSC1, TSC2, TTPA, TTR, TULP1, TWIST1, TYR, USH1C, USH2A, VCL, VHL, WAS, WRN, WT1, or any combination thereof.

In some embodiments, at least one component in the kit is provided in a desiccated or lyophilized form. In other embodiments, at least one component of the kit is provided in a solubilized form.

The kits provided herein are in suitable packaging. Suitable packaging includes, but is not limited to, vials, bottles, jars, flexible packaging, and the like. Also contemplated are packages for use in combination with a specific device. See “Devices for Sample Preparation and Sample Sequencing.” A kit may have a sterile access port (for example, the container may be an intravenous solution bag or a vial having a stopper pierceable by a hypodermic injection needle). The container may also have a sterile access port.

Kits optionally may provide additional components such as buffers and interpretive information. In some embodiments, the kit further comprises at least one buffer. Buffers suitable for the methods described herein have been described previously. In some embodiments, the kit can additionally comprise instructions for use in any of the methods described herein.

In some embodiment, the disclosure provides articles of manufacture comprising contents of the kits described above.

VI. Devices for Sample Preparation and Sample Sequencing

In some aspects, the disclosure relates to devices for sample preparation and/or sample sequencing. In some embodiments, the device comprises a sample preparation module. In some embodiments, the device comprises a sample sequencing module. In some embodiments, the device comprises a sample preparation module and a sample sequencing module.

A. Device for Sample Preparation

Devices including apparatuses, cartridges (*e.g.*, comprising channels (*e.g.*, microfluidic channels)), and/or pumps (*e.g.*, peristaltic pumps) for use in a process of preparing a sample for analysis are generally provided. Devices can be used in accordance with the instant disclosure to enable enrichment, concentration, manipulation, and/or detection of a target molecule from a biological sample. In some embodiments, devices and related methods are provided for automated processing of a sample to produce material for next generation sequencing and/or other downstream analytical techniques. Devices and related methods may be used for performing chemical and/or biological reactions, including reactions for nucleic acid and/or polypeptide processing in accordance with sample preparation or sample analysis processes described elsewhere herein.

In some embodiments, a sample preparation device is positioned to deliver or transfer to a sequencing module or device a target molecule or sample comprising a plurality of molecules (*e.g.*, a target nucleic acid or a target polypeptide). In some embodiments, a sample preparation device is connected directly to (*e.g.*, physically attached to) or indirectly to a sequencing device.

In some embodiments, a device comprise a sequence preparation module that is configured to receive one or more cartridges. In some embodiments, a cartridge comprises one or more reservoirs or reaction vessels configured to receive a fluid and/or contain one or more reagents used in a sample preparation process. In some embodiments, a cartridge comprises one or more channels (*e.g.*, microfluidic channels) configured to contain and/or transport a fluid (*e.g.*, a fluid comprising one or more reagents) used in a sample preparation process. Reagents include buffers, enzymatic reagents, polymer matrices, barcode components (*e.g.*, barcode molecules), detector molecules, enrichment molecules, capture reagents, size-specific selection reagents, sequence-specific selection reagents, and/or purification reagents. Additional reagents for use in a sample preparation process are described elsewhere herein.

In some embodiments, a cartridge includes one or more stored reagents (*e.g.*, of a liquid or lyophilized form suitable for reconstitution to a liquid form). The stored reagents of a cartridge include reagents suitable for carrying out a desired process and/or reagents suitable for processing a desired sample type. In some embodiments, a cartridge is a single-use cartridge (*e.g.*, a disposable cartridge) or a multiple-use cartridge (*e.g.*, a reusable cartridge). In some embodiments, a cartridge is configured to receive a user-supplied sample. The user-supplied sample may be added to the cartridge before or after the cartridge is received by the device, *e.g.*, manually by the user or in an automated process.

In some embodiments, the device may facilitate the preparation of a multiplexed sample in a process in accordance with the instant disclosure. See “Methods of Preparing a Multiplexed Sample”.

In some embodiments, the device may facilitate enrichment of a target molecule in a process in accordance with the instant disclosure. See “Methods of Polypeptide Enrichment.” In this way, the device enables the leveraging of molecules to enrich for polypeptides of interest in a highly multiplexed fashion.

In some embodiments, a sample is enriched for a target molecule using an electrophoretic method. In some embodiments, a sample is enriched for a target molecule using affinity SCODA. In some embodiments, a sample is enriched for a target molecule using field inversion gel electrophoresis (FIGE). In some embodiments, a sample is enriched for a target molecule using pulsed field gel electrophoresis (PFGE).

In some embodiments, a device comprises sample preparation module comprising a matrix used during enrichment (*e.g.*, a porous media, electrophoretic polymer gel) comprising immobilized capture probes that bind (directly or indirectly) to target molecules present in the sample. In some embodiments, a matrix used during enrichment comprises 1, 2, 3, 4, 5, or more unique immobilized capture probes, each of which binds to a unique target molecule and/or bind to the same target molecule with different binding affinities.

In some embodiments, an immobilized capture probe is a polypeptide capture probe that binds to a target polypeptide or polypeptide fragment. For example, in some embodiments, an immobilized capture probe is an enrichment molecule as described herein.

In some embodiments, a polypeptide capture probe binds to a target polypeptide (or polypeptide fragment) with a binding affinity of 10^{-9} to 10^{-8} M, 10^{-8} to 10^{-7} M, 10^{-7} to 10^{-6} M, 10^{-6} to 10^{-5} M, 10^{-5} to 10^{-4} M, 10^{-4} to 10^{-3} M, or 10^{-3} to 10^{-2} M. In some embodiments, the binding affinity is in the picomolar to nanomolar range (*e.g.*, between about 10^{-12} and about 10^{-9} M). In some embodiments, the binding affinity is in the nanomolar to micromolar range (*e.g.*, between about 10^{-9} and about 10^{-6} M). In some embodiments, the binding affinity is in the micromolar to millimolar range (*e.g.*, between about 10^{-6} and about 10^{-3} M). In some embodiments, the binding affinity is in the picomolar to micromolar range (*e.g.*, between about 10^{-12} and about 10^{-6} M). In some embodiments, the binding affinity is in the nanomolar to millimolar range (*e.g.*, between about 10^{-9} and about 10^{-3} M).

In some embodiments, an immobilized capture probe is an oligonucleotide capture probe that hybridizes to a target nucleic acid. In some embodiments, an oligonucleotide capture probe is at least 50%, 60%, 70%, 80%, 90% 95%, or 100% complementary to a target nucleic acid. In some embodiments, a single oligonucleotide capture probe may be used to enrich a plurality of related target nucleic acids (*e.g.*, 2, 3, 4, 5, 6, 7, 8, 9, 10, 20, 30, 40, 50, or more related target nucleic acids) that share at least 50%, 60%, 70%, 80%, 90% 95%, or 99% sequence identity.

Enrichment of a plurality of related target nucleic acids may allow for the generation of a

metagenomic library. In some embodiments, an oligonucleotide capture probe may enable differential enrichment of related target nucleic acids. In some embodiments, an oligonucleotide capture probe may enable enrichment of a target nucleic acid relative to a nucleic acid of identical sequence that differs in its modification state (*e.g.*, methylation state, acetylation state).

5 In some embodiments, for the purposes of enriching nucleic acid target molecules with a length of 0.5-2 kilobases, oligonucleotide capture probes may be covalently immobilized in an acrylamide matrix using a 5' Acrydite moiety. In some embodiments, for the purposes of enriching larger nucleic acid target molecules (*e.g.*, with a length of >2 kilobases), oligonucleotide capture probes may be immobilized in an agarose matrix. In some
10 embodiments, oligonucleotide capture probes may be immobilized in an agarose matrix using thiol-epoxide chemistries (*e.g.*, by covalently attached thiol-modified oligonucleotides to crosslinked agarose beads). Oligonucleotide capture probes linked to agarose beads can be combined and solidified within standard agarose matrices (*e.g.*, at the same agarose percentage).

In some embodiments, multiple capture probes (*e.g.*, populations of multiple capture
15 probe types, *e.g.*, that bind to deterministic target molecules of infectious agents such as adenovirus, staphylococcus, pneumonia, or tuberculosis) may be immobilized in an enrichment matrix. Application of a sample to an enrichment matrix with multiple deterministic capture probes may result in diagnosis of a disease or condition (*e.g.*, presence of an infectious agent).

In some embodiments, a device may facilitate release of a target molecule from the
20 enrichment matrix after removal of non-target molecules, in a process in accordance with the instant disclosure. In some embodiments, a target molecule may be released from the enrichment matrix by increasing the temperature of the enrichment matrix. Adjusting the temperature of the matrix further influences migration rate as increased temperatures provide a higher capture probe stringency, requiring greater binding affinities between the target molecule
25 and the capture probe. In some embodiments, when enriching related target molecules, the matrix temperature may be gradually increased in a step-wise manner in order to release and isolate target molecules in steps of ever-increasing homology. This may allow for the sequencing of target polypeptides or target nucleic acids that are increasingly distant in their relation to an initial reference target molecule, enabling discovery of novel proteins (*e.g.*,
30 enzymes) or functions (*e.g.*, enzymatic function or gene function). In some embodiments, when using multiple capture probes (*e.g.*, multiple deterministic capture probes), the matrix temperature may be increased in a step-wise or gradient fashion, permitting temperature-dependent release of different target molecules and resulting in generation of a series of barcoded release bands that represent the presence or absence of control and target
35 molecules.

Devices in accordance with the instant disclosure generally contain mechanical and electronic and/or optical components which can be used to operate a cartridge as described herein. In some embodiments, the device components operate to achieve and maintain specific temperatures on a cartridge or on specific regions of the cartridge. In some embodiments, the device components operate to apply specific voltages for specific time durations to electrodes of a cartridge. In some embodiments, the device components operate to move liquids to, from, or between reservoirs and/or reaction vessels of a cartridge. In some embodiments, the device components operate to move liquids through channel(s) of a cartridge, *e.g.*, to, from, or between reservoirs and/or reaction vessels of a cartridge. In some embodiments, the device components move liquids via a peristaltic pumping mechanism (*e.g.*, apparatus) that interacts with an elastomeric, reagent-specific reservoir or reaction vessel of a cartridge. In some embodiments, the device components move liquids via a peristaltic pumping mechanism (*e.g.*, apparatus) that is configured to interact with an elastomeric component (*e.g.*, surface layer comprising an elastomer) associated with a channel of a cartridge to pump fluid through the channel. Device components can include computer resources, for example, to drive a user interface where sample information can be entered, specific processes can be selected, and run results can be reported.

The following non-limiting example is meant to illustrate aspects of the devices, methods, and compositions described herein. The use of a sample preparation device in accordance with the instant disclosure may proceed with one or more of the following described steps. A user may open the lid of the device and insert a cartridge that supports the desired process. The user may then add a sample, which may be combined with a specific lysis solution, to a sample port on the cartridge. The user may then close the device lid, enter any sample specific information via a touch screen interface on the device, select any process specific parameters (*e.g.*, range of desired size selection, desired degree of homology for target molecule capture, etc.), and initiate the sample preparation process run.

Following the run, the user may receive relevant run data (*e.g.*, confirmation of successful completion of the run, run specific metrics, etc.), as well as process specific information (*e.g.*, amount of sample generated, presence or absence of specific target sequence, etc.). Data generated by the run may be subjected to subsequent bioinformatics analysis, which can be either local or cloud based. Depending on the process, a finished sample may be extracted from the cartridge for subsequent use (*e.g.*, genomic sequencing, qPCR quantification, cloning, etc.). The device may then be opened, and the cartridge may then be removed.

FIG. 8 provides an illustration depicting an exemplary apparatus for preparing a sample (*e.g.*, an enriched or multiplexed sample). See *e.g.*, U.S. Pat. No. 8608929, the entirety of which is incorporated herein by reference.

B. Device for Sequencing

Devices including apparatuses, cartridges (*e.g.*, comprising channels (*e.g.*, microfluidic channels)), and/or pumps (*e.g.*, peristaltic pumps) for use in a process of sequencing a sample (*e.g.*, a multiplexed sample) comprising polypeptides are also generally provided. Sequencing of nucleic acids or polypeptides in accordance with the instant disclosure, in some aspects, may be performed using a system that permits single molecule analysis and/or the sequencing of single molecules in parallel. The system may include a sequencing device and an instrument configured to interface with the sequencing device.

The sequencing device may include a sequencing module comprising an array of pixels, where individual pixels include a sample well and at least one photodetector. The sample wells of the sequencing device may be formed on or through a surface of the sequencing device and be configured to receive a sample placed on the surface of the sequencing device. In some embodiments, the sample wells are a component of a cartridge (*e.g.*, a disposable or single-use cartridge) that can be inserted into the device. Collectively, the sample wells may be considered as an array of sample wells. The plurality of sample wells may have a suitable size and shape such that at least a portion of the sample wells receive a single target molecule or sample comprising a plurality of molecules (*e.g.*, a target nucleic acid or a target polypeptide). In some embodiments, the number of molecules within a sample well may be distributed among the sample wells of the sequencing device such that some sample wells contain one molecule (*e.g.*, a target nucleic acid or a target polypeptide) while others contain zero, two, or a plurality of molecules.

In some embodiments, a sequencing device is positioned to receive a sample comprising a plurality of molecules (*e.g.*, one or more polypeptides of interest) from a sample preparation device. In some embodiments, a sequencing device is connected directly (*e.g.*, physically attached to) or indirectly to a sample preparation device.

The sequencing device may include an array of pixels, where individual pixels include a sample well and at least one photodetector. The sample wells of the sequencing device may be formed on or through a surface of the sequencing device and be configured to receive a sample placed on the surface of the sequencing device. Collectively, the sample wells may be considered as an array of sample wells. The plurality of sample wells may have a suitable size and shape such that at least a portion of the sample wells receive a single sample (*e.g.*, a single molecule, such as a polypeptide). In some embodiments, the number of samples within a sample well may be distributed among the sample wells of the sequencing device such that some sample wells contain one sample while others contain zero, two or more samples.

Excitation light is provided to the sequencing device from one or more light source, which may be external or internal to the sequencing device. Optical components of the sequencing device may receive the excitation light from the light source and direct the light towards the array of sample wells of the sequencing device and illuminate an illumination region within the sample well. In some embodiments, a sample well may have a configuration that allows for the sample to be retained in proximity to a surface of the sample well, which may ease delivery of excitation light to the sample and detection of emission light from the sample. A sample positioned within the illumination region may emit emission light in response to being illuminated by the excitation light. For example, the sample may be labeled with a fluorescent marker, which emits light in response to achieving an excited state through the illumination of excitation light. Emission light emitted by a sample may then be detected by one or more photodetectors within a pixel corresponding to the sample well with the sample being analyzed. When performed across the array of sample wells, which may range in number between approximately 10,000 pixels to 1,000,000 pixels according to some embodiments, multiple samples can be analyzed in parallel.

The sequencing device may include an optical system for receiving excitation light and directing the excitation light among the sample well array. The optical system may include one or more grating couplers configured to couple excitation light to the sequencing device and direct the excitation light to other optical components. The optical system may include optical components that direct the excitation light from a grating coupler towards the sample well array. Such optical components may include optical splitters, optical combiners, and waveguides. In some embodiments, one or more optical splitters may couple excitation light from a grating coupler and deliver excitation light to at least one of the waveguides. According to some embodiments, the optical splitter may have a configuration that allows for delivery of excitation light to be substantially uniform across all the waveguides such that each of the waveguides receives a substantially similar amount of excitation light. Such embodiments may improve performance of the sequencing device by improving the uniformity of excitation light received by sample wells of the sequencing device. Examples of suitable components, *e.g.*, for coupling excitation light to a sample well and/or directing emission light to a photodetector, to include in a sequencing device are described in U.S. Pat. Application No. 14/821,688, filed August 7, 2015, titled "INTEGRATED DEVICE FOR PROBING, DETECTING AND ANALYZING MOLECULES," and U.S. Pat. Application No. 14/543,865, filed November 17, 2014, titled "INTEGRATED DEVICE WITH EXTERNAL LIGHT SOURCE FOR PROBING, DETECTING, AND ANALYZING MOLECULES," both of which are incorporated by reference in their entirety. Examples of suitable grating couplers and waveguides that may be

implemented in the sequencing device are described in U.S. Pat. Application No. 15/844,403, filed December 15, 2017, titled "OPTICAL COUPLER AND WAVEGUIDE SYSTEM," which is incorporated by reference in its entirety.

Additional photonic structures may be positioned between the sample wells and the photodetectors and configured to reduce or prevent excitation light from reaching the photodetectors, which may otherwise contribute to signal noise in detecting emission light. In some embodiments, metal layers which may act as a circuitry for the sequencing device, may also act as a spatial filter. Examples of suitable photonic structures may include spectral filters, a polarization filters, and spatial filters and are described in U.S. Pat. Application No. 16/042,968, filed July 23, 2018, titled "OPTICAL REJECTION PHOTONIC STRUCTURES," which is incorporated by reference in its entirety.

Components located off of the sequencing device may be used to position and align an excitation source to the sequencing device. Such components may include optical components including lenses, mirrors, prisms, windows, apertures, attenuators, and/or optical fibers.

Additional mechanical components may be included in the instrument to allow for control of one or more alignment components. Such mechanical components may include actuators, stepper motors, and/or knobs. Examples of suitable excitation sources and alignment mechanisms are described in U.S. Pat. Application No. 15/161,088, filed May 20, 2016, titled "PULSED LASER AND SYSTEM," which is incorporated by reference in its entirety. Another example of a beam-steering module is described in U.S. Pat. Application No. 15/842,720, filed December, 14, 2017, titled "COMPACT BEAM SHAPING AND STEERING ASSEMBLY," which is incorporated herein by reference. Additional examples of suitable excitation sources are described in U.S. Pat. Application No. 14/821,688, filed August 7, 2015, titled "INTEGRATED DEVICE FOR PROBING, DETECTING AND ANALYZING MOLECULES," which is incorporated by reference in its entirety.

The photodetector(s) positioned with individual pixels of the sequencing device may be configured and positioned to detect emission light from the pixel's corresponding sample well. Examples of suitable photodetectors are described in U.S. Pat. Application No. 14/821,656, filed August 7, 2015, titled "INTEGRATED DEVICE FOR TEMPORAL BINNING OF RECEIVED PHOTONS," which is incorporated by reference in its entirety. In some embodiments, a sample well and its respective photodetector(s) may be aligned along a common axis. In this manner, the photodetector(s) may overlap with the sample well within the pixel.

Characteristics of the detected emission light may provide an indication for identifying the marker associated with the emission light. Such characteristics may include any suitable type of characteristic, including an arrival time of photons detected by a photodetector, an amount of

photons accumulated over time by a photodetector, and/or a distribution of photons across two or more photodetectors. In some embodiments, a photodetector may have a configuration that allows for the detection of one or more timing characteristics associated with a sample's emission light (*e.g.*, luminescence lifetime). The photodetector may detect a distribution of photon arrival times after a pulse of excitation light propagates through the sequencing device, and the distribution of arrival times may provide an indication of a timing characteristic of the sample's emission light (*e.g.*, a proxy for luminescence lifetime). In some embodiments, the one or more photodetectors provide an indication of the probability of emission light emitted by the marker (*e.g.*, luminescence intensity). In some embodiments, a plurality of photodetectors may be sized and arranged to capture a spatial distribution of the emission light. Output signals from the one or more photodetectors may then be used to distinguish a marker from among a plurality of markers, where the plurality of markers may be used to identify a sample within the sample. In some embodiments, a sample may be excited by multiple excitation energies, and emission light and/or timing characteristics of the emission light emitted by the sample in response to the multiple excitation energies may distinguish a marker from a plurality of markers.

In operation, parallel analyses of samples within the sample wells are carried out by exciting some or all of the samples within the wells using excitation light and detecting signals from sample emission with the photodetectors. Emission light from a sample may be detected by a corresponding photodetector and converted to at least one electrical signal. The electrical signals may be transmitted along conducting lines in the circuitry of the sequencing device, which may be connected to an instrument interfaced with the sequencing device. The electrical signals may be subsequently processed and/or analyzed. Processing or analyzing of electrical signals may occur on a suitable computing device either located on or off the instrument.

The instrument may include a user interface for controlling operation of the instrument and/or the sequencing device. The user interface may be configured to allow a user to input information into the instrument, such as commands and/or settings used to control the functioning of the instrument. In some embodiments, the user interface may include buttons, switches, dials, and a microphone for voice commands. The user interface may allow a user to receive feedback on the performance of the instrument and/or sequencing device, such as proper alignment and/or information obtained by readout signals from the photodetectors on the sequencing device. In some embodiments, the user interface may provide feedback using a speaker to provide audible feedback. In some embodiments, the user interface may include indicator lights and/or a display screen for providing visual feedback to a user.

In some embodiments, the instrument may include a computer interface configured to connect with a computing device. The computer interface may be a USB interface, a FireWire

interface, or any other suitable computer interface. A computing device may be any general purpose computer, such as a laptop or desktop computer. In some embodiments, a computing device may be a server (*e.g.*, cloud-based server) accessible over a wireless network via a suitable computer interface. The computer interface may facilitate communication of information between the instrument and the computing device. Input information for controlling and/or configuring the instrument may be provided to the computing device and transmitted to the instrument via the computer interface. Output information generated by the instrument may be received by the computing device via the computer interface. Output information may include feedback about performance of the instrument, performance of the sequencing device, and/or data generated from the readout signals of the photodetector.

In some embodiments, the instrument may include a processing device configured to analyze data received from one or more photodetectors of the sequencing device and/or transmit control signals to the excitation source(s). In some embodiments, the processing device may comprise a general purpose processor, a specially-adapted processor (*e.g.*, a central processing unit (CPU) such as one or more microprocessor or microcontroller cores, a field-programmable gate array (FPGA), an application-specific integrated circuit (ASIC), a custom integrated circuit, a digital signal processor (DSP), or a combination thereof). In some embodiments, the processing of data from one or more photodetectors may be performed by both a processing device of the instrument and an external computing device. In other embodiments, an external computing device may be omitted and processing of data from one or more photodetectors may be performed solely by a processing device of the sequencing device.

According to some embodiments, the instrument that is configured to analyze samples based on luminescence emission characteristics may detect differences in luminescence lifetimes and/or intensities between different luminescent molecules, and/or differences between lifetimes and/or intensities of the same luminescent molecules in different environments. The inventors have recognized and appreciated that differences in luminescence emission lifetimes can be used to discern between the presence or absence of different luminescent molecules and/or to discern between different environments or conditions to which a luminescent molecule is subjected. In some cases, discerning luminescent molecules based on lifetime (rather than emission wavelength, for example) can simplify aspects of the system. As an example, wavelength-discriminating optics (such as wavelength filters, dedicated detectors for each wavelength, dedicated pulsed optical sources at different wavelengths, and/or diffractive optics) may be reduced in number or eliminated when discerning luminescent molecules based on lifetime. In some cases, a single pulsed optical source operating at a single characteristic wavelength may be used to excite different luminescent molecules that emit within a same wavelength region of the

optical spectrum but have measurably different lifetimes. An analytic system that uses a single pulsed optical source, rather than multiple sources operating at different wavelengths, to excite and discern different luminescent molecules emitting in a same wavelength region can be less complex to operate and maintain, more compact, and may be manufactured at lower cost.

Although analytic systems based on luminescence lifetime analysis may have certain benefits, the amount of information obtained by an analytic system and/or detection accuracy may be increased by allowing for additional detection techniques. For example, some embodiments of the systems may additionally be configured to discern one or more properties of a sample based on luminescence wavelength and/or luminescence intensity. In some implementations, luminescence intensity may be used additionally or alternatively to distinguish between different luminescent labels. For example, some luminescent labels may emit at significantly different intensities or have a significant difference in their probabilities of excitation (*e.g.*, at least a difference of about 35%) even though their decay rates may be similar. By referencing binned signals to measured excitation light, it may be possible to distinguish different luminescent labels based on intensity levels.

According to some embodiments, different luminescence lifetimes may be distinguished with a photodetector that is configured to time-bin luminescence emission events following excitation of a luminescent label. The time binning may occur during a single charge-accumulation cycle for the photodetector. A charge-accumulation cycle is an interval between read-out events during which photo-generated carriers are accumulated in bins of the time-binning photodetector. Examples of a time-binning photodetector are described in U.S. Pat. Application No. 14/821,656, filed August 7, 2015, titled "INTEGRATED DEVICE FOR TEMPORAL BINNING OF RECEIVED PHOTONS," which is incorporated herein by reference. In some embodiments, a time-binning photodetector may generate charge carriers in a photon absorption/carrier generation region and directly transfer charge carriers to a charge carrier storage bin in a charge carrier storage region. In such embodiments, the time-binning photodetector may not include a carrier travel/capture region. Such a time-binning photodetector may be referred to as a "direct binning pixel." Examples of time-binning photodetectors, including direct binning pixels, are described in U.S. Pat. Application No. 15/852,571, filed December, 22, 2017, titled "INTEGRATED PHOTODETECTOR WITH DIRECT BINNING PIXEL," which is incorporated herein by reference.

In some embodiments, different numbers of fluorophores of the same type may be linked to different reagents in a sample, so that each reagent may be identified based on luminescence intensity. For example, two fluorophores may be linked to a first labeled affinity reagent and four or more fluorophores may be linked to a second labeled affinity reagent. Because of the

different numbers of fluorophores, there may be different excitation and fluorophore emission probabilities associated with the different affinity reagents. For example, there may be more emission events for the second labeled affinity reagent during a signal accumulation interval, so that the apparent intensity of the bins is significantly higher than for the first labeled affinity reagent.

The inventors have recognized and appreciated that distinguishing nucleotides or any other biological or chemical samples based on fluorophore decay rates and/or fluorophore intensities may enable a simplification of the optical excitation and detection systems. For example, optical excitation may be performed with a single-wavelength source (*e.g.*, a source producing one characteristic wavelength rather than multiple sources or a source operating at multiple different characteristic wavelengths). Additionally, wavelength discriminating optics and filters may not be needed in the detection system. Also, a single photodetector may be used for each sample well to detect emission from different fluorophores. The phrase “characteristic wavelength” or “wavelength” is used to refer to a central or predominant wavelength within a limited bandwidth of radiation (*e.g.*, a central or peak wavelength within a 20 nm bandwidth output by a pulsed optical source). In some cases, “characteristic wavelength” or “wavelength” may be used to refer to a peak wavelength within a total bandwidth of radiation output by a source.

EQUIVALENTS AND SCOPE

In the claims articles such as “a,” “an,” and “the” may mean one or more than one unless indicated to the contrary or otherwise evident from the context. Claims or descriptions that include “or” between one or more members of a group are considered satisfied if one, more than one, or all of the group members are present in, employed in, or otherwise relevant to a given product or process unless indicated to the contrary or otherwise evident from the context. The invention includes embodiments in which exactly one member of the group is present in, employed in, or otherwise relevant to a given product or process. The invention includes embodiments in which more than one, or all of the group members are present in, employed in, or otherwise relevant to a given product or process.

Furthermore, the invention encompasses all variations, combinations, and permutations in which one or more limitations, elements, clauses, and descriptive terms from one or more of the listed claims is introduced into another claim. For example, any claim that is dependent on another claim can be modified to include one or more limitations found in any other claim that is dependent on the same base claim. Where elements are presented as lists, *e.g.*, in Markush group format, each subgroup of the elements is also disclosed, and any element(s) can be removed from

the group. It should be understood that, in general, where the invention, or aspects of the invention, is/are referred to as comprising particular elements and/or features, certain embodiments of the invention or aspects of the invention consist, or consist essentially of, such elements and/or features. For purposes of simplicity, those embodiments have not been specifically set forth *in haec verba* herein.

The phrase “and/or,” as used herein in the specification and in the claims, should be understood to mean “either or both” of the elements so conjoined, *i.e.*, elements that are conjunctively present in some cases and disjunctively present in other cases. Multiple elements listed with “and/or” should be construed in the same fashion, *i.e.*, “one or more” of the elements so conjoined. Other elements may optionally be present other than the elements specifically identified by the “and/or” clause, whether related or unrelated to those elements specifically identified. Thus, as a non-limiting example, a reference to “A and/or B”, when used in conjunction with open-ended language such as “comprising” can refer, in one embodiment, to A only (optionally including elements other than B); in another embodiment, to B only (optionally including elements other than A); in yet another embodiment, to both A and B (optionally including other elements); etc.

As used herein in the specification and in the claims, “or” should be understood to have the same meaning as “and/or” as defined above. For example, when separating items in a list, “or” or “and/or” shall be interpreted as being inclusive, *i.e.*, the inclusion of at least one, but also including more than one, of a number or list of elements, and, optionally, additional unlisted items. Only terms clearly indicated to the contrary, such as “only one of” or “exactly one of,” or, when used in the claims, “consisting of,” will refer to the inclusion of exactly one element of a number or list of elements. In general, the term “or” as used herein shall only be interpreted as indicating exclusive alternatives (*i.e.* “one or the other but not both”) when preceded by terms of exclusivity, such as “either,” “one of,” “only one of,” or “exactly one of.” “Consisting essentially of,” when used in the claims, shall have its ordinary meaning as used in the field of patent law.

As used herein in the specification and in the claims, the phrase “at least one,” in reference to a list of one or more elements, should be understood to mean at least one element selected from any one or more of the elements in the list of elements, but not necessarily including at least one of each and every element specifically listed within the list of elements and not excluding any combinations of elements in the list of elements. This definition also allows that elements may optionally be present other than the elements specifically identified within the list of elements to which the phrase “at least one” refers, whether related or unrelated to those elements specifically identified. Thus, as a non-limiting example, “at least one of A and B” (or, equivalently, “at least one of A or B,” or, equivalently “at least one of A and/or B”) can refer, in

one embodiment, to at least one, optionally including more than one, A, with no B present (and optionally including elements other than B); in another embodiment, to at least one, optionally including more than one, B, with no A present (and optionally including elements other than A); in yet another embodiment, to at least one, optionally including more than one, A, and at least
5 one, optionally including more than one, B (and optionally including other elements); etc.

It should also be understood that, unless clearly indicated to the contrary, in any methods claimed herein that include more than one step or act, the order of the steps or acts of the method is not necessarily limited to the order in which the steps or acts of the method are recited.

In the claims, as well as in the specification above, all transitional phrases such as
10 “comprising,” “including,” “carrying,” “having,” “containing,” “involving,” “holding,” “composed of,” and the like are to be understood to be open-ended, *i.e.*, to mean including but not limited to. Only the transitional phrases “consisting of” and “consisting essentially of” shall be closed or semi-closed transitional phrases, respectively, as set forth in the United States Patent Office Manual of Patent Examining Procedures, Section 2111.03. It should be appreciated that
15 embodiments described in this document using an open-ended transitional phrase (*e.g.*, “comprising”) are also contemplated, in alternative embodiments, as “consisting of” and “consisting essentially of” the feature described by the open-ended transitional phrase. For example, if the application describes “a composition comprising A and B,” the application also contemplates the alternative embodiments “a composition consisting of A and B” and “a
20 composition consisting essentially of A and B.”

Where ranges are given, endpoints are included. Furthermore, unless otherwise indicated or otherwise evident from the context and understanding of one of ordinary skill in the art, values that are expressed as ranges can assume any specific value or sub-range within the stated ranges in different embodiments of the invention, to the tenth of the unit of the lower limit of the range,
25 unless the context clearly dictates otherwise.

This application refers to various issued patents, published patent applications, journal articles, and other publications, all of which are incorporated herein by reference. If there is a conflict between any of the incorporated references and the instant specification, the specification shall control. In addition, any particular embodiment of the present invention that
30 falls within the prior art may be explicitly excluded from any one or more of the claims. Because such embodiments are deemed to be known to one of ordinary skill in the art, they may be excluded even if the exclusion is not set forth explicitly herein. Any particular embodiment of the invention can be excluded from any claim, for any reason, whether or not related to the existence of prior art.

Those skilled in the art will recognize or be able to ascertain using no more than routine experimentation many equivalents to the specific embodiments described herein. The scope of the present embodiments described herein is not intended to be limited to the above Description, but rather is as set forth in the appended claims. Those of ordinary skill in the art will appreciate
5 that various changes and modifications to this description may be made without departing from the spirit or scope of the present invention, as defined in the following claims.

The recitation of a listing of chemical groups in any definition of a variable herein includes definitions of that variable as any single group or combination of listed groups. The recitation of an embodiment for a variable herein includes that embodiment as any single
10 embodiment or in combination with any other embodiments or portions thereof. The recitation of an embodiment herein includes that embodiment as any single embodiment or in combination with any other embodiments or portions thereof.

CLAIMS

1. A method comprising directly sequencing, in parallel, the proteome of a single cell and optionally sequencing the genome and/or transcriptome of the single cell, and/or optionally
5 detecting one or more metabolite of the single cell.

2. The method of claim 1, wherein the method comprises:

- (i) providing a cell sample comprising the composition of a single cell;
- (ii) contacting the cell sample with a barcode component to produce a labeled sample
10 comprising barcoded molecules, wherein the barcoded molecules comprise barcoded polypeptides; and
- (iii) sequencing the polypeptides of the labeled sample.

3. The method of claim 1, wherein the method comprises:

- (i) providing a cell sample comprising the composition of a single cell;
- (ii) contacting the cell sample with a barcode component to produce a labeled sample
15 comprising barcoded molecules, wherein the barcoded molecules comprise barcoded polynucleic acids; and
- (iii) sequencing the polynucleic acids of the labeled sample.

4. The method of claim 3, wherein (iii) comprises long-read sequencing applications, short-read sequencing applications, or hybrid assembly applications.

5. The method of claim 3 or claim 4, wherein the barcoded polynucleic acids have a length
25 of about 0.5-2 kb, 0.5-5 kb, 1-2 kb, 1-3 kb, 1-4 kb, 1-5 kb, 1-10 kb, 2-10 kb, 2-5 kb, 5-10 kb, 5-15 kb, 5-20 kb, 5-25 kb, 10-15 kb, 10-20 kb, or 10-25 kb.

6. The method of claim 3 or claim 4, wherein the barcoded polynucleic acids have a length of about 700-3000, 1000-3000, 1000-2500, 1000-2400, 1000-2300, 1000-2200, 1000-2100,
30 1000-2000, 1000-1900, 1000-1800, 1000-1700, 1000-1600, 1000-1500, 1000-1400, 1000-1300, 1000-1200, 1500-3000, 1500-2500, 1500-2000, or 2000-3000 nucleotides in length.

7. The method of any one of claims 2-6, wherein the composition of (i) comprises a living cell.

8. The method of any one of claims 2-6, wherein the composition of (i) comprises a lysed cell.

9. The method of any one of claims 2-8, wherein the barcoded molecules of the labeled sample each comprise an identical barcode.

10. The method of any one of claims 2-9, wherein the barcoded molecules of the labeled sample further comprise barcoded DNA, barcoded RNA, barcoded cDNA, or barcoded metabolites.

11. The method of claim 10, wherein the barcoded molecules of the labeled sample comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

12. The method of any one of claims 2-11, wherein (ii) comprises:

(a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded polypeptides;

(b) isolating the barcoded polypeptides from the first sample, thereby generating a second sample comprising the barcoded polypeptides and a third sample comprising the genome, transcriptome, and/or metabolome of the single cell in the cell sample; and

(c) contacting the third sample with an additional barcode component, to produce a fourth sample comprising barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites;

wherein the labeled sample comprises the second sample and the fourth sample.

13. The method of claim 12, wherein the third sample in (b) comprises:

(i) a first subsample comprising the genome and transcriptome of the single cell and a second subsample comprising the metabolome of the single cell;

(ii) a first subsample comprising the genome and metabolome of the single cell and a second subsample comprising the transcriptome of the single cell;

(iii) a first subsample comprising the metabolome and transcriptome of the single cell and a second subsample comprising the genome of the single cell; or

- (iv) a first subsample comprising the genome of the single cell, a second subsample comprising the transcriptome of the single cell, and a third subsample comprising the metabolome of the single cell.

5 14. The method of claim 12 or claim 13, wherein the additional barcode component in (c) comprises a second, third, fourth, or fifth barcode component.

15. The method of any one of claims 12-14, wherein the barcoded molecules of the fourth sample in (c) comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the
10 method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

16. The method of any one of claims 2-11, wherein (ii) comprises:

- (a) contacting the cell sample with a first barcode component to produce a first
15 sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA;
- (b) amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA of the first sample; and
- (c) contacting the first sample with a second barcode component, to produce a second sample comprising barcoded polypeptides and/or barcoded metabolites;
20 wherein the labeled sample comprises the second sample.

17. The method of any one of claims 2-11, wherein (ii) comprises:

- (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA;
- 25 (b) isolating the barcoded DNA, barcoded RNA, and/or barcoded cDNA from the first sample, thereby generating a second sample comprising the barcoded DNA and/or barcoded cDNA and a third sample comprising the proteome and/or metabolome of the single cell in the cell sample; and
- (c) contacting the third sample with an additional barcode component, to produce a
30 fourth sample comprising barcoded polypeptides and/or the barcoded metabolites;
wherein the labeled sample comprises the second sample and the fourth sample.

18. The method of claim 17, wherein the third sample in (b) comprises a first subsample comprising the proteome of the single cell and a second subsample comprising the metabolome
35 of the single cell.

19. The method of claim 17 or claim 18, wherein the additional barcode component in (c) comprises a second or third barcode component.

20. The method of any one of claims 16-19, wherein the barcoded molecules of the first sample in (a) comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

21. The method of any one of claims 10-20, further comprising sequencing the barcoded polypeptides, the barcoded DNA, barcoded RNA, and/or barcoded cDNA of the labeled sample.

22. The method of any one of claims 10-20, further comprising detecting and optionally quantifying one or more of the barcoded metabolites of the labeled sample.

23. The method of any one of claims 10-22, wherein the barcoded DNA, barcoded RNA, and/or barcoded cDNA have a length of about 0.5-2 kb, 0.5-5 kb, 1-2 kb, 1-3 kb, 1-4 kb, 1-5 kb, 1-10 kb, 2-10 kb, 2-5 kb, 5-10 kb, 5-15 kb, 5-20 kb, 5-25 kb, 10-15 kb, 10-20 kb, or 10-25 kb.

24. The method of any one of claims 10-22, wherein the barcoded DNA, barcoded RNA, and/or barcoded cDNA have a length of about 700-3000, 1000-3000, 1000-2500, 1000-2400, 1000-2300, 1000-2200, 1000-2100, 1000-2000, 1000-1900, 1000-1800, 1000-1700, 1000-1600, 1000-1500, 1000-1400, 1000-1300, 1000-1200, 1500-3000, 1500-2500, 1500-2000, or 2000-3000 nucleotides in length.

25. The method of claim 21, wherein the sequencing comprises:

(a) detecting the barcode identities of the barcoded molecules of the labeled sample, thereby determining the origins of the barcoded molecules; and

(b) sequencing, in parallel, the barcoded polypeptides in the labeled sample, thereby determining at least the partial amino acid sequences of the barcoded polypeptides;

wherein (a) occurs before, after, or concurrently with (b).

26. The method of any one of claims 2-20, wherein the method further comprises combining the labeled sample with at least one supplemental sample comprising barcoded molecules,

wherein the barcoded molecules of each sample are distinguishable, thereby producing a multiplexed sample.

27. The method of claim 26, wherein at least one supplemental sample is prepared by a method comprising:

- (a) providing a cell sample comprising the composition of a single cell; and
- (b) contacting the cell sample with a barcode component to produce a labeled sample comprising barcoded molecules.

28. The method of claim 27, wherein the composition of (a) comprises a living cell.

29. The method of claim 27, wherein the composition of (a) comprises a lysed cell.

30. The method of any one of claims 27-29, wherein the barcoded molecules of (b) each comprise an identical barcode.

31. The method of any one of claims 27-29, wherein the barcoded molecules of (b) comprise barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites.

32. The method of claim 31, further comprising sequencing the barcoded polypeptides, barcoded DNA, barcoded RNA, and/or barcoded cDNA of the multiplexed sample.

33. The method of claim 31 or claim 32, further comprising detecting, and optionally quantifying, the barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites of the multiplexed sample.

34. The method of claim 32, wherein the sequencing comprises:

- (a) detecting the barcode identities of the barcoded molecules of the multiplexed sample, thereby determining the origins of the barcoded molecules; and
- (b) sequencing, in parallel, the barcoded polypeptides in the multiplexed sample, thereby determining at least the partial amino acid sequences of the barcoded polypeptides;

wherein (a) occurs before, after, or concurrently with (b).

35. The method of claim 25 or claim 34, wherein the barcode identities are detected in (a) by DNA sequencing, protein sequencing, hybridization, luminescence, binding kinetics, and/or physical location on or within a solid substrate.

36. The method of any one of claims 1-35, wherein the sequencing comprises:

- (a) contacting a single polypeptide molecule with one or more terminal amino acid recognition molecules; and
- (b) detecting a series of signal pulses indicative of association of the one or more terminal amino acid recognition molecules with successive amino acids exposed at a terminus of the single polypeptide while the single polypeptide is being degraded, thereby sequencing the single polypeptide molecule.

37. The method of any one of claims 1-35, wherein the sequencing comprises:

- (a) contacting a single polypeptide molecule with a composition comprising one or more terminal amino acid recognition molecules and a cleaving reagent; and
- (b) detecting a series of signal pulses indicative of association of the one or more terminal amino acid recognition molecules with a terminus of the single polypeptide molecule in the presence of the cleaving reagent, wherein the series of signal pulses is indicative of a series of amino acids exposed at the terminus over time as a result of terminal amino acid cleavage by the cleaving reagent.

38. The method of any one of claims 1-35, wherein the sequencing comprises:

- (a) identifying a first amino acid at a terminus of a single polypeptide molecule;
- (b) removing the first amino acid to expose a second amino acid at the terminus of the single polypeptide molecule, and
- (c) identifying the second amino acid at the terminus of the single polypeptide molecule,

wherein (a)-(c) are performed in a single reaction mixture.

39. The method of any one of claims 1-35, wherein the sequencing comprises:

- (a) contacting a single polypeptide molecule with one or more amino acid recognition molecules that bind to the single polypeptide molecule;
- (b) detecting a series of signal pulses indicative of association of the one or more amino acid recognition molecules with the single polypeptide molecule under polypeptide degradation conditions; and

- (c) identifying a first type of amino acid in the single polypeptide molecule based on a first characteristic pattern in the series of signal pulses.

40. The method of any one of claims 1-35, wherein the sequencing comprises:

- (a) obtaining data during a polypeptide degradation process;
- (b) analyzing the data to determine portions of the data corresponding to amino acids that are sequentially exposed at a terminus of the polypeptide during the degradation process; and
- (c) outputting an amino acid sequence representative of the polypeptide.

41. The method of any one of claims 1-35, wherein the sequencing comprises:

- (a) contacting a polypeptide with one or more labeled affinity reagents that selectively bind one or more types of terminal amino acids at a terminus of the polypeptide; and
- (b) identifying a terminal amino acid at the terminus of the polypeptide by detecting an interaction of the polypeptide with the one or more labeled affinity reagents.

42. The method of any one of claims 1-35, wherein the sequencing comprises:

- (a) contacting a polypeptide with one or more labeled affinity reagents that selectively bind one or more types of terminal amino acids at a terminus of the polypeptide;
- (b) identifying a terminal amino acid at the terminus of the polypeptide by detecting an interaction of the polypeptide with the one or more labeled affinity reagents;
- (c) removing the terminal amino acid; and
- (d) repeating (a)-(c) one or more times at the terminus of the polypeptide to determine an amino acid sequence of the polypeptide.

43. The method of claim 42, wherein the method further comprises:

after (a) and before (b), removing any of the one or more labeled affinity reagents that do not selectively bind the terminal amino acid; and/or
after (b) and before (c), removing any of the one or more labeled affinity reagents that selectively bind the terminal amino acid.

44. The method of claim 42, wherein (c) comprises modifying the terminal amino acid by contacting the terminal amino acid with an isothiocyanate, and:

contacting the modified terminal amino acid with a protease that specifically binds and removes the modified terminal amino acid; or
subjecting the modified terminal amino acid to acidic or basic conditions sufficient to remove the modified terminal amino acid.

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45. The method of claim 42, wherein identifying the terminal amino acid comprises:
identifying the terminal amino acid as being one type of the one or more types of terminal amino acids to which the one or more labeled affinity reagents bind; or
identifying the terminal amino acid as being a type other than the one or more
10 types of terminal amino acids to which the one or more labeled affinity reagents bind.

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46. The method of claim 42, wherein the one or more labeled affinity reagents comprise one or more labeled aptamers, one or more labeled peptidases, one or more labeled antibodies, one or more labeled degradation pathway protein, one or more aminotransferase, one or more tRNA
15 synthetase, or a combination thereof.

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47. The method of claim 46, wherein the one or more labeled peptidases have been modified to inactivate cleavage activity; or wherein the one or more labeled peptidases retain cleavage activity for the removing of (c).

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48. A method comprising:
(i) providing a cell sample;
(ii) contacting the cell sample with a barcode component to produce a labeled sample comprising barcoded molecules, wherein the barcoded molecules comprise
25 barcoded polypeptides, barcoded DNA, barcoded RNA, and/or barcoded cDNA;
and
(iii) sequencing the barcoded polypeptides, barcoded DNA, barcoded RNA, and/or barcoded cDNA of the labeled sample;
wherein the barcoded molecules of the labeled sample are not amplified prior to
30 sequencing;
optionally wherein the barcoded molecules in (ii) further comprise barcoded metabolites
and optionally wherein the method further comprises detecting one or more of the barcoded metabolites.

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49. The method of claim 48, wherein the cell sample comprises the composition of a single cell.

50. The method of claim 49, wherein the composition of (i) comprises a living cell.

51. The method of claim 49, wherein the composition of (i) comprises a lysed cell.

52. The method of any one of claims 49-51, wherein the barcoded molecules of the labeled sample each comprise an identical barcode.

53. The method of any one of claims 49-52, wherein the barcoded molecules of the labeled sample comprise barcoded DNA, barcoded RNA, barcoded cDNA, and barcoded metabolites.

54. The method of claim 53, wherein the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

55. The method of any one of claims 49-54, wherein (ii) comprises:

(a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded polypeptides;

(b) isolating the barcoded polypeptides from the first sample, thereby generating a second sample comprising the barcoded polypeptides and a third sample comprising the genome, transcriptome, and/or metabolome of the single cell in the cell sample; and

(c) contacting the third sample with an additional barcode component, to produce a fourth sample comprising barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites;

wherein the labeled sample comprises the second sample and the fourth sample.

56. The method of claim 55, wherein the third sample in (b) comprises:

(i) a first subsample comprising the genome and transcriptome of the single cell and a second subsample comprising the metabolome of the single cell;

(ii) a first subsample comprising the genome and metabolome of the single cell and a second subsample comprising the transcriptome of the single cell;

(iii) a first subsample comprising the metabolome and transcriptome of the single cell and a second subsample comprising the genome of the single cell; or

- (iv) a first subsample comprising the genome of the single cell, a second subsample comprising the transcriptome of the single cell, and a third subsample comprising the metabolome of the single cell.

5 57. The method of claim 55 or claim 56, wherein the additional barcode component in (c) comprises a second, third, fourth, or fifth barcode component.

58. The method of any one of claims 55-57, wherein the barcoded molecules of the fourth sample in (c) comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the
10 method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

59. The method of any one of claims 49-54, wherein (ii) comprises:

- (a) contacting the cell sample with a first barcode component to produce a first
15 sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA;
 - (b) amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA of the first sample; and
 - (c) contacting the first sample with a second barcode component, to produce a second sample comprising barcoded polypeptides and/or barcoded metabolites;
- 20 wherein the labeled sample comprises the second sample.

60. The method of any one of claims 49-54, wherein (ii) comprises:

- (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA;
 - 25 (b) isolating the barcoded DNA, barcoded RNA, and/or barcoded cDNA from the first sample, thereby generating a second sample comprising the barcoded DNA and/or barcoded cDNA and a third sample comprising the proteome and/or metabolome of the single cell in the cell sample; and
 - (c) contacting the third sample with an additional barcode component, to produce a
30 fourth sample comprising barcoded polypeptides and/or the barcoded metabolites;
- wherein the labeled sample comprises the second sample and the fourth sample.

61. The method of claim 60, wherein the third sample in (b) comprises a first subsample comprising the proteome of the single cell and a second subsample comprising the metabolome
35 of the single cell.

62. The method of claim 60 or claim 61, wherein the additional barcode component in (c) comprises a second or third barcode component.

63. The method of any one of claims 59-62, wherein the barcoded molecules of the first sample in (a) comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

64. The method of any one of claims 49-63, further comprising sequencing the barcoded polypeptides, barcoded DNA, barcoded RNA, and/or barcoded cDNA of the labeled sample.

65. The method of any one of claims 49-63, further comprising detecting, and optionally quantifying, the barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites of the labeled sample.

66. The method of claim 64, wherein sequencing the barcoded DNA, barcoded RNA, and/or barcoded cDNA comprises long-read sequencing applications, short-read sequencing applications, or hybrid assembly applications.

67. The method of any one of claims 63-66, wherein the barcoded DNA, barcoded RNA, and/or barcoded cDNA have a length of about 0.5-2 kb, 0.5-5 kb, 1-2 kb, 1-3 kb, 1-4 kb, 1-5 kb, 1-10 kb, 2-10 kb, 2-5 kb, 5-10 kb, 5-15 kb, 5-20 kb, 5-25 kb, 10-15 kb, 10-20 kb, or 10-25 kb.

68. The method of any one of claims 63-66, wherein the barcoded DNA, barcoded RNA, and/or barcoded cDNA have a length of about 700-3000, 1000-3000, 1000-2500, 1000-2400, 1000-2300, 1000-2200, 1000-2100, 1000-2000, 1000-1900, 1000-1800, 1000-1700, 1000-1600, 1000-1500, 1000-1400, 1000-1300, 1000-1200, 1500-3000, 1500-2500, 1500-2000, or 2000-3000 nucleotides in length.

69. The method of claim 64, wherein the sequencing comprises:

- (a) detecting the barcode identities of the barcoded molecules of the labeled sample, thereby determining the origins of the barcoded molecules; and

- (b) sequencing, in parallel, the barcoded polypeptides in the labeled sample, thereby determining at least the partial amino acid sequences of the barcoded polypeptides;

wherein (a) occurs before, after, or concurrently with (b).

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70. The method of any one of claims 49-63, wherein the method further comprises combining the labeled sample with at least one supplemental sample comprising barcoded molecules, wherein the barcoded molecules of each sample are distinguishable, thereby producing a multiplexed sample.

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71. The method of claim 70, wherein at least one supplemental sample is prepared by a method comprising:

- (a) providing a cell sample comprising the composition of a single cell; and
- (b) contacting the cell sample with a barcode component to produce a labeled sample comprising barcoded molecules.

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72. The method of claim 71, wherein the composition of (a) comprises a living cell.

73. The method of claim 71, wherein the composition of (a) comprises a lysed cell.

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74. The method of any one of claims 71-73, wherein the barcoded molecules of (b) each comprise an identical barcode.

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75. The method of any one of claims 71-73, wherein the barcoded molecules of (b) comprise barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites.

76. The method of claim 75, further comprising sequencing the barcoded polypeptides, barcoded DNA, barcoded RNA, and/or barcoded cDNA of the multiplexed sample.

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77. The method of claim 75 or claim 76, further comprising detecting, and optionally quantifying, the barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites of the multiplexed sample.

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78. The method of claim 76, wherein the sequencing comprises:

- (a) detecting the barcode identities of the barcoded molecules of the multiplexed sample, thereby determining the origins of the barcoded molecules; and
- (b) sequencing, in parallel, the barcoded polypeptides in the multiplexed sample, thereby determining at least the partial amino acid sequences of the barcoded polypeptides;

wherein (a) occurs before, after, or concurrently with (b).

79. The method of claim 69 or claim 78, wherein the barcode identities are detected in (a) by DNA sequencing, protein sequencing, hybridization, luminescence, binding kinetics, and/or physical location on or within a solid substrate.

80. The method of any one of claims 48-79, wherein the sequencing comprises:

- (a) contacting a single polypeptide molecule with one or more terminal amino acid recognition molecules; and
- (b) detecting a series of signal pulses indicative of association of the one or more terminal amino acid recognition molecules with successive amino acids exposed at a terminus of the single polypeptide while the single polypeptide is being degraded, thereby sequencing the single polypeptide molecule.

81. The method of any one of claims 48-79, wherein the sequencing comprises:

- (a) contacting a single polypeptide molecule with a composition comprising one or more terminal amino acid recognition molecules and a cleaving reagent; and
- (b) detecting a series of signal pulses indicative of association of the one or more terminal amino acid recognition molecules with a terminus of the single polypeptide molecule in the presence of the cleaving reagent, wherein the series of signal pulses is indicative of a series of amino acids exposed at the terminus over time as a result of terminal amino acid cleavage by the cleaving reagent.

82. The method of any one of claims 48-79, wherein the sequencing comprises:

- (a) identifying a first amino acid at a terminus of a single polypeptide molecule;
- (b) removing the first amino acid to expose a second amino acid at the terminus of the single polypeptide molecule, and
- (c) identifying the second amino acid at the terminus of the single polypeptide molecule,

wherein (a)-(c) are performed in a single reaction mixture.

83. The method of any one of claims 48-79, wherein the sequencing comprises:
- (a) contacting a single polypeptide molecule with one or more amino acid recognition molecules that bind to the single polypeptide molecule;
 - 5 (b) detecting a series of signal pulses indicative of association of the one or more amino acid recognition molecules with the single polypeptide molecule under polypeptide degradation conditions; and
 - (c) identifying a first type of amino acid in the single polypeptide molecule based on a first characteristic pattern in the series of signal pulses.

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84. The method of any one of claims 48-79, wherein the sequencing comprises:
- (a) obtaining data during a polypeptide degradation process;
 - (b) analyzing the data to determine portions of the data corresponding to amino acids that are sequentially exposed at a terminus of the polypeptide during the
 - 15 degradation process; and
 - (c) outputting an amino acid sequence representative of the polypeptide.

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85. The method of any one of claims 48-79, wherein the sequencing comprises:
- (a) contacting a polypeptide with one or more labeled affinity reagents that
 - 20 selectively bind one or more types of terminal amino acids at a terminus of the polypeptide; and
 - (b) identifying a terminal amino acid at the terminus of the polypeptide by detecting an interaction of the polypeptide with the one or more labeled affinity reagents.

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86. The method of any one of claims 48-79, wherein the sequencing comprises:
- (a) contacting a polypeptide with one or more labeled affinity reagents that
 - 25 selectively bind one or more types of terminal amino acids at a terminus of the polypeptide;
 - (b) identifying a terminal amino acid at the terminus of the polypeptide by detecting
 - 30 an interaction of the polypeptide with the one or more labeled affinity reagents;
 - (c) removing the terminal amino acid; and
 - (d) repeating (a)-(c) one or more times at the terminus of the polypeptide to determine an amino acid sequence of the polypeptide.

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87. The method of claim 86, wherein the method further comprises:

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after (a) and before (b), removing any of the one or more labeled affinity reagents that do not selectively bind the terminal amino acid; and/or

after (b) and before (c), removing any of the one or more labeled affinity reagents that selectively bind the terminal amino acid.

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88. The method of claim 86, wherein (c) comprises modifying the terminal amino acid by contacting the terminal amino acid with an isothiocyanate, and:

contacting the modified terminal amino acid with a protease that specifically binds and removes the modified terminal amino acid; or

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subjecting the modified terminal amino acid to acidic or basic conditions sufficient to remove the modified terminal amino acid.

89. The method of claim 86, wherein identifying the terminal amino acid comprises:

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identifying the terminal amino acid as being one type of the one or more types of terminal amino acids to which the one or more labeled affinity reagents bind; or

identifying the terminal amino acid as being a type other than the one or more types of terminal amino acids to which the one or more labeled affinity reagents bind.

90. The method of claim 86, wherein the one or more labeled affinity reagents comprise one or more labeled aptamers, one or more labeled peptidases, one or more labeled antibodies, one or more labeled degradation pathway protein, one or more aminotransferase, one or more tRNA synthetase, or a combination thereof.

91. The method of claim 90, wherein the one or more labeled peptidases have been modified to inactivate cleavage activity; or wherein the one or more labeled peptidases retain cleavage activity for the removing of (c).

92. A method comprising:

- (i) providing a cell sample;
- (ii) contacting the cell sample with a barcode component to produce a labeled sample comprising barcoded molecules, wherein the barcoded molecules comprise barcoded polypeptides and barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites.

93. The method of claim 92, wherein (i) comprises:

- (a) providing a cell population; and
- (b) lysing the cell population.

94. The method of claim 93, wherein the cell population:

consists of a single cell;
comprises a plurality of homogeneous cells; or
comprises a plurality of heterogeneous cells.

95. The method of claim 93 or claim 94, wherein the cell population is isolated from a subject.

96. The method of claim 95, wherein the subject is a human, mouse, rat, or non-human primate subject.

97. The method of any one of claims 92-96, wherein the barcoded molecules of (ii) each comprise an identical barcode.

98. The method of any one of claims 92-97, wherein (ii) comprises:

- (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded polypeptides;
- (b) isolating the barcoded polypeptides from the first sample, thereby generating a second sample comprising the barcoded polypeptides and a third sample comprising the genome, transcriptome, and/or metabolome of the single cell in the cell sample; and
- (c) contacting the third sample with an additional barcode component, to produce a fourth sample comprising barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites;

wherein the labeled sample comprises the second sample and the fourth sample.

99. The method of claim 98, wherein the third sample in (b) comprises:

- (i) a first subsample comprising the genome and transcriptome of the single cell and a second subsample comprising the metabolome of the single cell;
- (ii) a first subsample comprising the genome and metabolome of the single cell and a second subsample comprising the transcriptome of the single cell;

- (iii) a first subsample comprising the metabolome and transcriptome of the single cell and a second subsample comprising the genome of the single cell; or
- (iv) a first subsample comprising the genome of the single cell, a second subsample comprising the transcriptome of the single cell, and a third subsample comprising the metabolome of the single cell.

100. The method of claim 98 or claim 99, wherein the additional barcode component in (c) comprises a second, third, fourth, or fifth barcode component.

101. The method of any one of claims 98-100, wherein the barcoded molecules of the fourth sample in (c) comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

102. The method of any one of claims 92-97, wherein (ii) comprises:

- (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA;
- (b) amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA of the first sample; and
- (c) contacting the first sample with a second barcode component, to produce a second sample comprising barcoded polypeptides and/or barcoded metabolites; wherein the labeled sample comprises the second sample.

103. The method of any one of claims 92-97, wherein (ii) comprises:

- (a) contacting the cell sample with a first barcode component to produce a first sample comprising barcoded DNA, barcoded RNA, and/or barcoded cDNA;
- (b) isolating the barcoded DNA, barcoded RNA, and/or barcoded cDNA from the first sample, thereby generating a second sample comprising the barcoded DNA and/or barcoded cDNA and a third sample comprising the proteome and/or metabolome of the single cell in the cell sample; and
- (c) contacting the third sample with an additional barcode component, to produce a fourth sample comprising barcoded polypeptides and/or the barcoded metabolites; wherein the labeled sample comprises the second sample and the fourth sample.

104. The method of claim 103 wherein the third sample in (b) comprises a first subsample comprising the proteome of the single cell and a second subsample comprising the metabolome of the single cell.

5 105. The method of claim 103 or claim 104, wherein the additional barcode component in (c) comprises a second or third barcode component.

106. The method of any one of claims 102-105, wherein the barcoded molecules of the first sample in (a) comprise barcoded DNA, barcoded RNA, and/or barcoded cDNA and wherein the
10 method further comprises amplifying the barcoded DNA, barcoded RNA, and/or the barcoded cDNA.

107. The method of any one of claims 92-106, further comprising sequencing the barcoded polypeptides, the barcoded DNA, barcoded RNA, and/or barcoded cDNA of the labeled sample.

15 108. The method of any one of claims 92-107, further comprising detecting, and optionally quantifying, the barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites of the multiplexed sample.

20 109. The method of claim 107, wherein sequencing the barcoded DNA, barcoded RNA, and/or barcoded cDNA comprises long-read sequencing applications, short-read sequencing applications, or hybrid assembly applications.

110. The method of any one of claims 106-109, wherein the barcoded DNA, barcoded RNA,
25 and/or barcoded cDNA have a length of about 0.5-2 kb, 0.5-5 kb, 1-2 kb, 1-3 kb, 1-4 kb, 1-5 kb, 1-10 kb, 2-10 kb, 2-5 kb, 5-10 kb, 5-15 kb, 5-20 kb, 5-25 kb, 10-15 kb, 10-20 kb, or 10-25 kb.

111. The method of any one of claims 106-109, wherein the barcoded DNA, barcoded RNA,
and/or barcoded cDNA have a length of about 700-3000, 1000-3000, 1000-2500, 1000-2400,
30 1000-2300, 1000-2200, 1000-2100, 1000-2000, 1000-1900, 1000-1800, 1000-1700, 1000-1600, 1000-1500, 1000-1400, 1000-1300, 1000-1200, 1500-3000, 1500-2500, 1500-2000, or 2000-3000 nucleotides in length.

112. The method of claim 107, wherein the sequencing comprises:

- (a) detecting the barcode identities of the barcoded molecules of the labeled sample, thereby determining the origins of the barcoded molecules; and
- (b) sequencing, in parallel, the barcoded polypeptides in the labeled sample, thereby determining at least the partial amino acid sequences of the barcoded polypeptides;

wherein (a) occurs before, after, or concurrently with (b).

113. The method of any one of claims 92-106, wherein the method further comprises combining the labeled sample with at least one supplemental sample comprising barcoded molecules, wherein the barcoded molecules of each sample are distinguishable, thereby producing a multiplexed sample.

114. The method of claim 113, wherein at least one supplemental sample is prepared by a method comprising:

- (a) providing a cell sample;
- (b) contacting the cell sample with a barcode component to produce a labeled sample comprising barcoded molecules, wherein the barcoded molecules comprise barcoded polypeptides and barcoded DNA, barcoded cDNA, and/or barcoded metabolites.

115. The method of claim 114, wherein (a) comprises:

- i. providing a cell population; and
- ii. lysing the cell population.

116. The method of claim 115, wherein the cell population: consists of a single cell; comprises a plurality of homologous cells; or comprises a plurality of heterologous cells.

117. The method of claim 115 or claim 116, wherein the cell population is isolated from a subject.

118. The method of claim 117, wherein the subject is a human, mouse, rat, or non-human primate subject.

119. The method of any one of claims 114-118, wherein the barcoded molecules of (b) each comprise an identical barcode.

120. The method of any one of claims 114-119, wherein the barcoded molecules of (b) comprise barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites.

121. The method of any one of claims 113-120, further comprising sequencing the barcoded polypeptides, the barcoded DNA, barcoded RNA, and/or barcoded cDNA of the multiplexed sample.

122. The method of any one of claims 113-121, further comprising detecting, and optionally quantifying, the barcoded polypeptides, barcoded DNA, barcoded RNA, barcoded cDNA, and/or barcoded metabolites of the multiplexed sample.

123. The method of claim 121, wherein the sequencing comprises:

- (a) detecting the barcode identities of the barcoded molecules of the multiplexed sample, thereby determining the origins of the barcoded molecules; and
- (b) sequencing, in parallel, the barcoded polypeptides in the multiplexed sample, thereby determining at least the partial amino acid sequences of the barcoded polypeptides;

wherein (a) occurs before, after, or concurrently with (b).

124. The method of claim 112 or claim 123, wherein the barcode identities are detected in (a) by DNA sequencing, protein sequencing, hybridization, luminescence, binding kinetics, and/or physical location on or within a solid substrate.

125. The method of any one of claims 112, 123, or 124, wherein the sequencing comprises:

- (a) contacting a single polypeptide molecule with one or more terminal amino acid recognition molecules; and
- (b) detecting a series of signal pulses indicative of association of the one or more terminal amino acid recognition molecules with successive amino acids exposed at a terminus of the single polypeptide while the single polypeptide is being degraded, thereby sequencing the single polypeptide molecule.

126. The method of any one of claims 112, 123, or 124, wherein the sequencing comprises:

- (a) contacting a single polypeptide molecule with a composition comprising one or more terminal amino acid recognition molecules and a cleaving reagent; and
- (b) detecting a series of signal pulses indicative of association of the one or more terminal amino acid recognition molecules with a terminus of the single polypeptide molecule in the presence of the cleaving reagent, wherein the series of signal pulses is indicative of a series of amino acids exposed at the terminus over time as a result of terminal amino acid cleavage by the cleaving reagent.

127. The method of any one of claims 112, 123, or 124, wherein the sequencing comprises:

- (a) identifying a first amino acid at a terminus of a single polypeptide molecule;
- (b) removing the first amino acid to expose a second amino acid at the terminus of the single polypeptide molecule, and
- (c) identifying the second amino acid at the terminus of the single polypeptide molecule,

wherein (a)-(c) are performed in a single reaction mixture.

128. The method of any one of claims 112, 123, or 124, wherein the sequencing comprises:

- (a) contacting a single polypeptide molecule with one or more amino acid recognition molecules that bind to the single polypeptide molecule;
- (b) detecting a series of signal pulses indicative of association of the one or more amino acid recognition molecules with the single polypeptide molecule under polypeptide degradation conditions; and
- (c) identifying a first type of amino acid in the single polypeptide molecule based on a first characteristic pattern in the series of signal pulses.

129. The method of any one of claims 112, 123, or 124, wherein the sequencing comprises:

- (a) obtaining data during a polypeptide degradation process;
- (b) analyzing the data to determine portions of the data corresponding to amino acids that are sequentially exposed at a terminus of the polypeptide during the degradation process; and
- (c) outputting an amino acid sequence representative of the polypeptide.

130. The method of any one of claims 112, 123, or 124, wherein the sequencing comprises:

- (a) contacting a polypeptide with one or more labeled affinity reagents that selectively bind one or more types of terminal amino acids at a terminus of the polypeptide; and
- (b) identifying a terminal amino acid at the terminus of the polypeptide by detecting an interaction of the polypeptide with the one or more labeled affinity reagents.

131. The method of any one of claims 112, 123, or 124, wherein the sequencing comprises:

- (a) contacting a polypeptide with one or more labeled affinity reagents that selectively bind one or more types of terminal amino acids at a terminus of the polypeptide;
- (b) identifying a terminal amino acid at the terminus of the polypeptide by detecting an interaction of the polypeptide with the one or more labeled affinity reagents;
- (c) removing the terminal amino acid; and
- (d) repeating (a)-(c) one or more times at the terminus of the polypeptide to determine an amino acid sequence of the polypeptide.

132. The method of claim 131, wherein the method further comprises:

after (a) and before (b), removing any of the one or more labeled affinity reagents that do not selectively bind the terminal amino acid; and/or

after (b) and before (c), removing any of the one or more labeled affinity reagents that selectively bind the terminal amino acid.

133. The method of claim 131, wherein (c) comprises modifying the terminal amino acid by contacting the terminal amino acid with an isothiocyanate, and:

- contacting the modified terminal amino acid with a protease that specifically binds and removes the modified terminal amino acid; or
- subjecting the modified terminal amino acid to acidic or basic conditions sufficient to remove the modified terminal amino acid.

134. The method of claim 131, wherein identifying the terminal amino acid comprises:

- identifying the terminal amino acid as being one type of the one or more types of terminal amino acids to which the one or more labeled affinity reagents bind; or
- identifying the terminal amino acid as being a type other than the one or more types of terminal amino acids to which the one or more labeled affinity reagents bind.

135. The method of claim 131, wherein the one or more labeled affinity reagents comprise one or more labeled aptamers, one or more labeled peptidases, one or more labeled antibodies, one or more labeled degradation pathway protein, one or more aminotransferase, one or more tRNA synthetase, or a combination thereof.

5

136. The method of claim 135, wherein the one or more labeled peptidases have been modified to inactivate cleavage activity; or wherein the one or more labeled peptidases retain cleavage activity for the removing of (c).

10 137. A kit for performing the method of any one of claims 1-136, wherein the kit comprises a barcode component comprising a plurality of barcode molecules.

138. The kit of claim 137, wherein the barcode component further comprises a reaction component comprising one or more reagent for covalently attaching a barcode molecule to
15 polypeptide.

139. The kit of claim 137 or claim 138, wherein the barcode component comprises one or more barcode molecules comprising a polynucleic acid portion, a polypeptide portion, and/or a fluorescent molecule portion.

20

140. The kit of claim 139, wherein the polynucleic acid portion is 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, or 60 nucleotides in length.

25 141. The kit of claim 139, wherein the polynucleic acid portion comprises an aptamer.

142. The kit of claim 139, wherein the polypeptide portion is 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 amino acids in length.

30 143. The kit of claim 139, wherein the polypeptide portion is an antibody or aptamer.

144. The kit of claim 139, wherein the fluorescent molecule portion comprises an aromatic or heteroaromatic compound, such as a pyrene, anthracene, naphthalene, acridine, stilbene, indole, benzindole, oxazole, carbazole, thiazole, benzothiazole, phenanthridine, phenoxazine, porphyrin,

quinoline, ethidium, benzamide, cyanine, carbocyanine, salicylate, anthranilate, coumarin, fluorescein, rhodamine, or the like.

145. The kit of claim 139 or claim 144, wherein the fluorescent molecule portion comprise a dye selected from the group consisting of a xanthene dye, a naphthalene dye, a coumarin dye, an acridine dye, a cyanine dye, a benzoxazole dye, a stilbene dye, a pyrene dye, a phthalocyanine dye, a phycobiliprotein dye, a squaraine dye, and a BODIPY dye.

146. The kit of any one of claims 137-145, further comprising a solid support.

147. The kit of claim 146, wherein the solid support comprises immobilized detector molecules comprising a polynucleic acid portion corresponding to a barcode molecule of the barcode component.

148. The kit of claim 146 or claim 147, wherein the solid support comprises immobilized detector molecules comprising a polypeptide portion corresponding to a barcode molecule of the barcode component.

149. A kit for performing the method of any one of claims 1-136, wherein the kit comprises a solid support that allows for the physical separation of populations of polypeptides of different origins.

150. A device comprising:

at least one hardware processor; and

at least one non-transitory computer-readable storage medium storing processor-executable instructions that, when executed by the at least one hardware processor, cause the at least one hardware processor to perform the method of any of claims 1-136.

151. At least one non-transitory computer-readable storage medium storing processor-executable instructions that, when executed by at least one hardware processor, cause the at least one hardware processor to perform the method of any of claims 1-136.

152. A device comprising a sample preparation module configured to interface with one or more cartridge, each cartridge comprising: (a) one or more reservoirs or reaction vessels configured to receive a complex sample; (b) one or more sequence sample preparation reagents,

wherein the sample preparation reagents comprise a plurality of barcode molecules; and (c) a matrix comprising one or more immobilized capture probes.

153. The device of claim 152, further comprising a sequencing module comprising an array of pixels, wherein each pixel is configured to receive a sequencing sample from the sample preparation module and comprises: (a) a sample well; and (b) at least one photodetector.

154. The device of claim 152 or claim 153, wherein the sample preparation reagents further comprise a plurality of enrichment molecules.

155. The device of claim 154, wherein at least a subset of the enrichment molecules in the plurality of enrichment molecules are covalently attached to an immobilized capture probe.

156. The device of claim 154 or claim 155, wherein at least a subset of the enrichment molecules are covalently attached to a bead or particle that is capable of being bound by an immobilized capture probe.

157. The device of any one of claims 154-156, wherein each of the enrichment molecules in the plurality of enrichment molecules comprises an antibody, an aptamer, or an enzyme.

158. The device of any one of claims 154-156, wherein the enrichment molecules in a subset of the plurality of enrichment molecules comprise an antibody, an aptamer, or an enzyme.

159. The device of any one of claims 152-158, wherein the sample preparation reagents comprise a modifying agent.

160. The device of claim 159, wherein the modifying agent mediates polypeptide fragmentation, polypeptide denaturation, addition of a post-translational modification, and/or the blocking of one or more functional groups.

161. The device of any one of claims 152-160, wherein the sequencing module further comprises a reservoir or reaction vessel configured to deliver sequencing reagents to the sample well of each pixel.

162. The device of claim 161, wherein the sequencing reagents comprise a labeled affinity reagent.

163. The device of claim 162, wherein the labeled affinity reagent comprises one or more
5 labeled aptamers, one or more labeled peptidases, one or more labeled antibodies, one or more
labeled degradation pathway protein, one or more aminotransferase, one or more tRNA
synthetase, or a combination thereof.

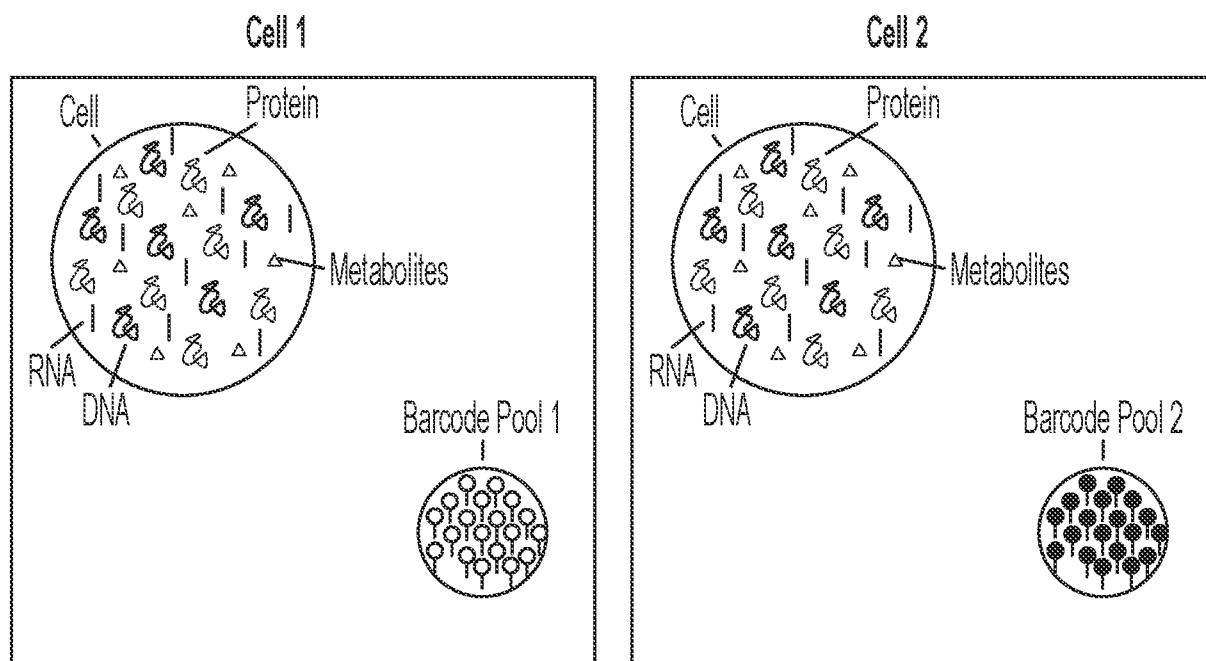
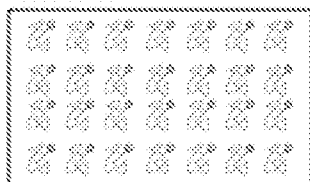


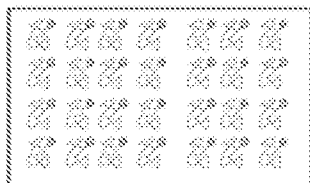
FIG. 1

1. Label Multiple Cells

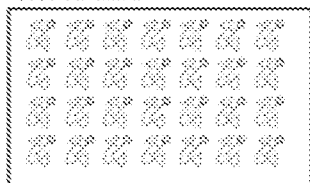
Labeled Cell 1



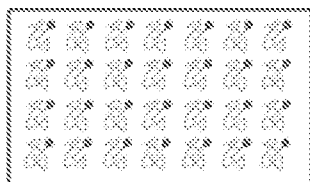
Labeled Cell 2



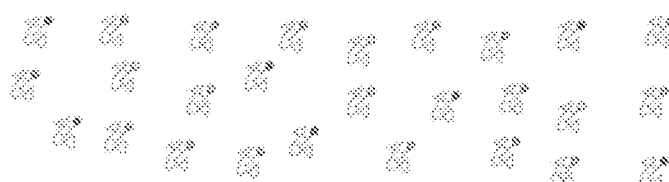
Labeled Cell 3



Labeled Cell 4



2. Pool Barcoded Cells



3. Sequence and Identify Barcodes

- DYRAGP (SEQ ID NO: 26)
- LPVVNC (SEQ ID NO: 27)
- REPIGN (SEQ ID NO: 28)
- ILSTEP (SEQ ID NO: 29)
- DAGMNV (SEQ ID NO: 30)
- SPIGRYP (SEQ ID NO: 31)
- QWCVRE (SEQ ID NO: 32)
- WVDYER (SEQ ID NO: 33)

4. Deconvolute Cell Origin

Cell 1

- DYRAGP (SEQ ID NO: 26)
- REPIGN (SEQ ID NO: 28)

Cell 2

- LPVVNC (SEQ ID NO: 27)
- DAGMNV (SEQ ID NO: 30)

Cell 3

- ILSTEP (SEQ ID NO: 29)
- QWCVRE (SEQ ID NO: 32)

Cell 4

- SPIGRYP (SEQ ID NO: 31)
- WVDYER (SEQ ID NO: 33)

FIG. 2

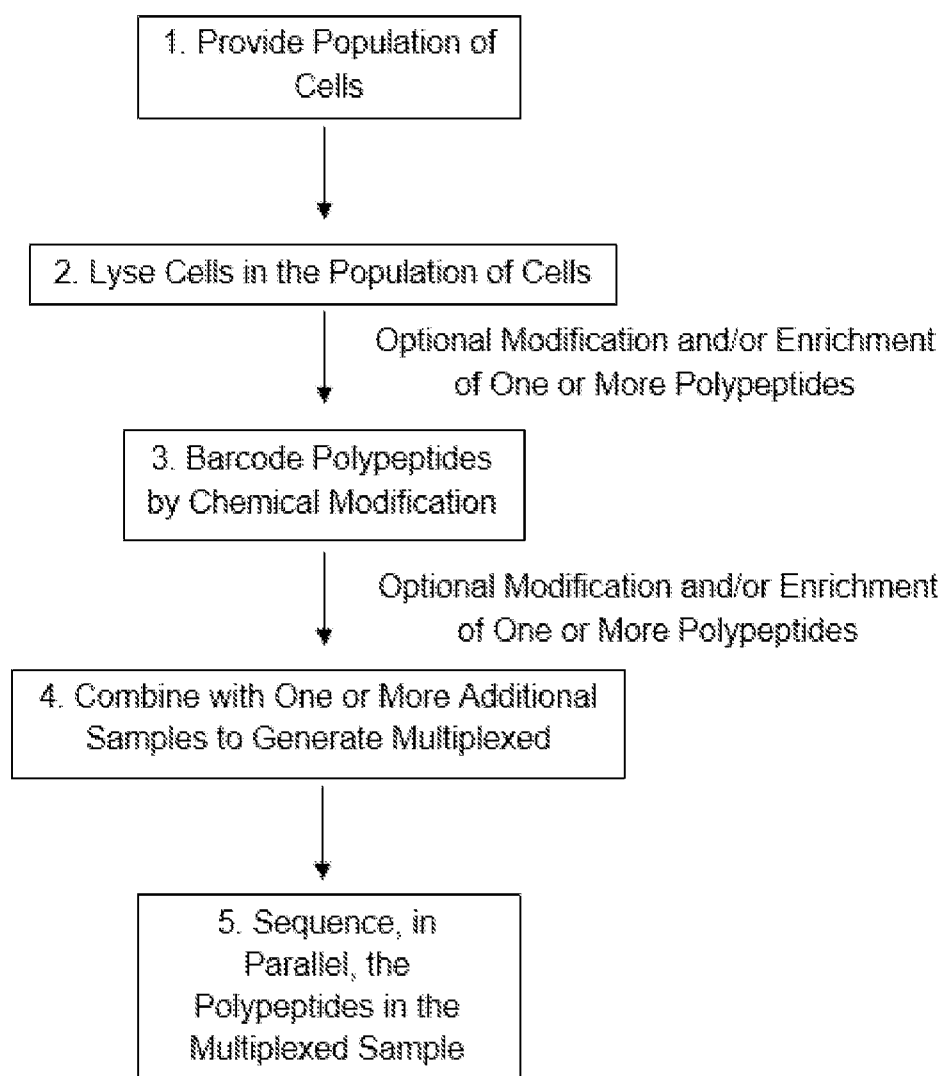


FIG. 3

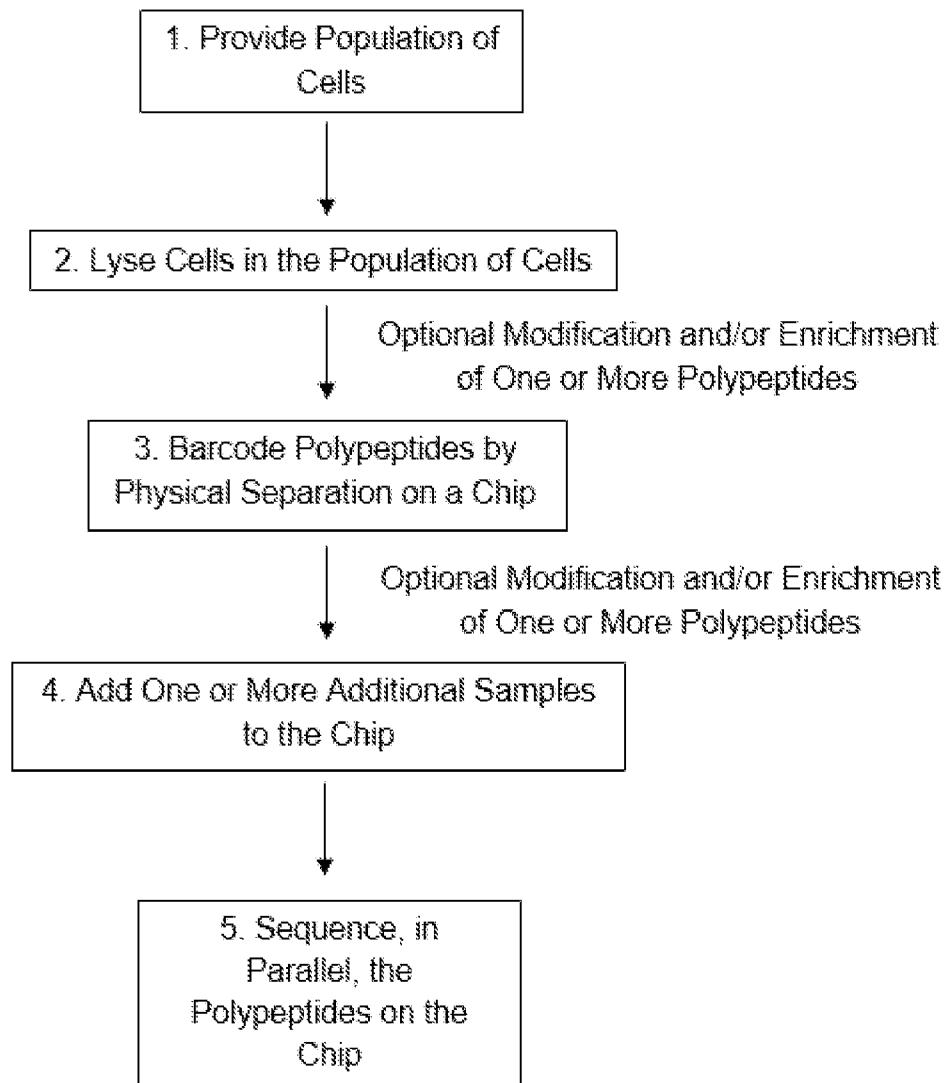


FIG. 4

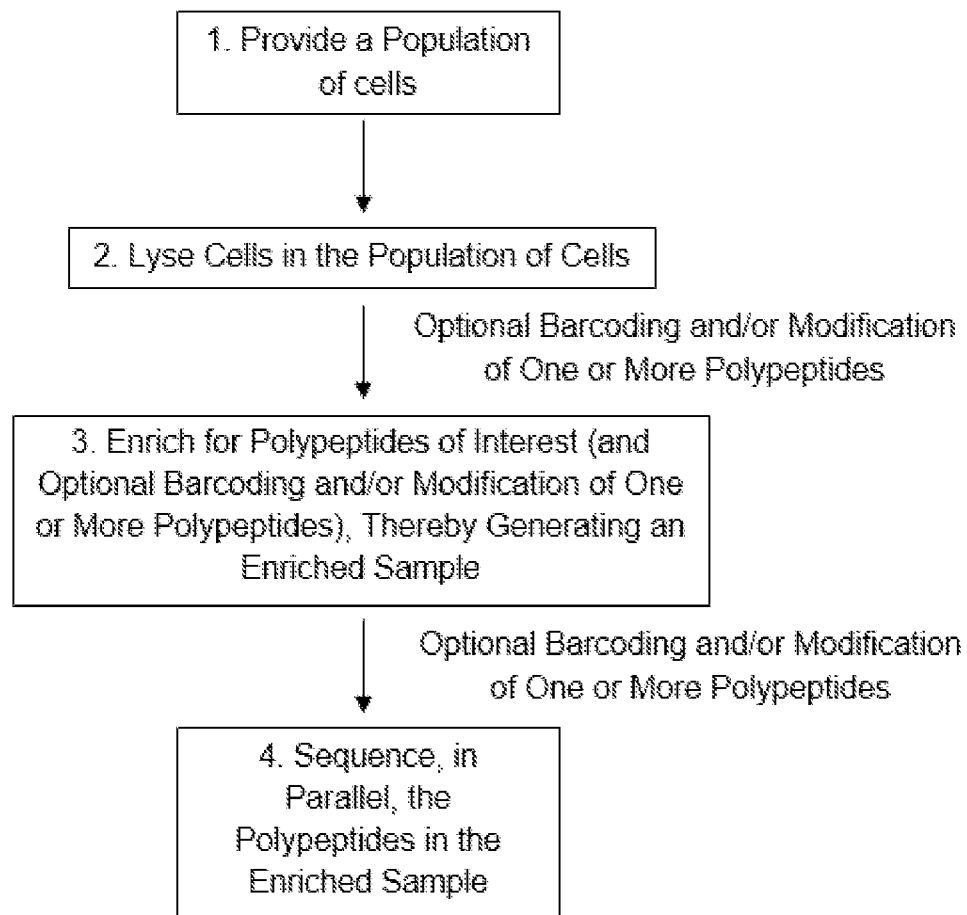
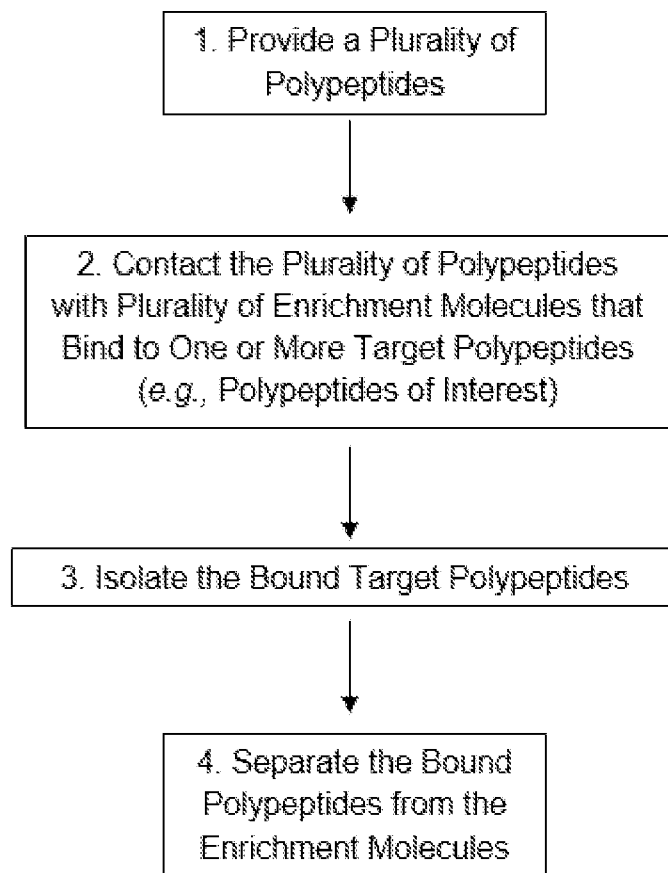
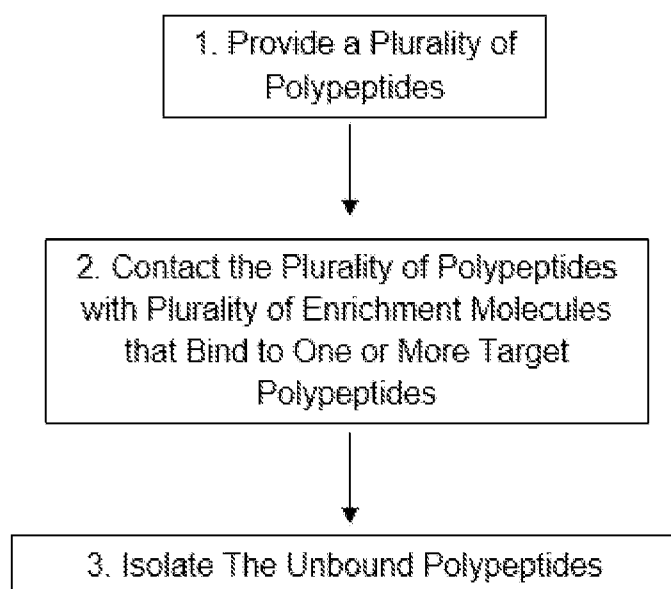


FIG. 5

**FIG. 6**

**FIG. 7**

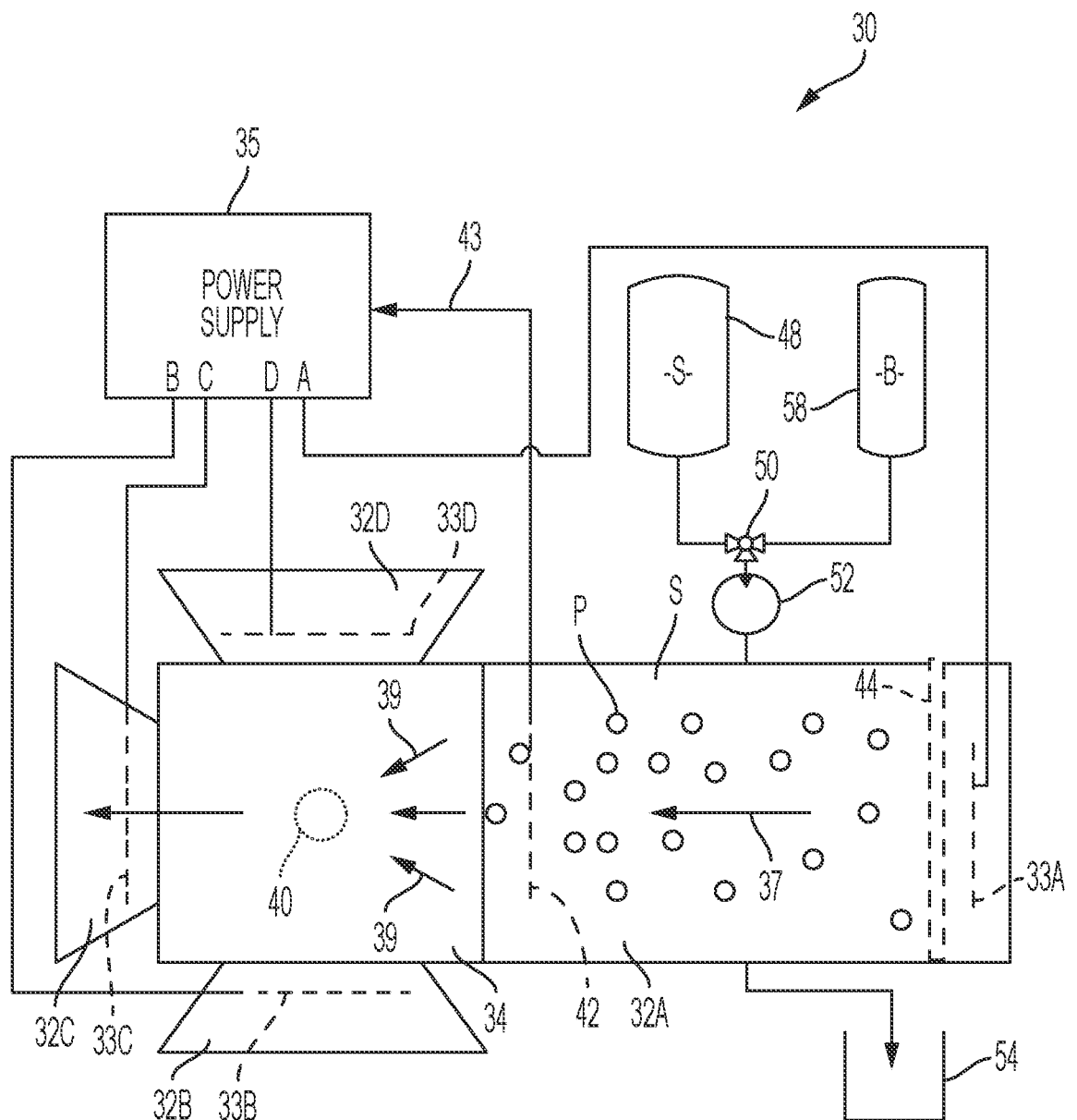


FIG. 8

SEQUENCE LISTING

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<120> METHODS OF SINGLE-CELL SEQUENCING

<130> R0708.70078W000

<140> Not Yet Assigned

<141> Concurrently Herewith

<150> US 62/991,425

<151> 2020-03-18

<150> US 62/926,991

<151> 2019-10-28

<160> 33

<170> PatentIn version 3.5

<210> 1

<211> 921

<212> PRT

<213> Artificial Sequence

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<223> Synthetic

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20 25 30

Gln Ser Lys Val Lys Lys Leu Ser Asp Tyr Lys Ser Leu Asp Tyr Phe
35 40 45

Val Ile His Val Asp Leu Gln Ile Asp Leu Ser Lys Lys Pro Val Glu
50 55 60

Ser Lys Ala Arg Leu Thr Val Val Pro Asn Leu Asn Val Asp Ser His
65 70 75 80

Ser Asn Asp Leu Val Leu Asp Gly Glu Asn Met Thr Leu Val Ser Leu
85 90 95

Gln Met Asn Asp Asn Leu Leu Lys Glu Asn Glu Tyr Glu Leu Thr Lys
100 105 110

Asp Ser Leu Ile Ile Lys Asn Ile Pro Gln Asn Thr Pro Phe Thr Ile
115 120 125

Glu Met Thr Ser Leu Leu Gly Glu Asn Thr Asp Leu Phe Gly Leu Tyr
130 135 140

Glu Thr Glu Gly Val Ala Leu Val Lys Ala Glu Ser Glu Gly Leu Arg

145		150		155		160
Arg Val Phe Tyr	Leu Pro Asp Arg Pro Asp Asn Leu Ala Thr Tyr Lys	165		170		175
Thr Thr Ile Ile	Ala Asn Gln Glu Asp Tyr Pro Val Leu Leu Ser Asn	180		185		190
Gly Val Leu Ile	Glu Lys Lys Glu Leu Pro Leu Gly Leu His Ser Val	195		200		205
Thr Trp Leu Asp	Asp Val Pro Lys Pro Ser Tyr Leu Phe Ala Leu Val	210		215		220
Ala Gly Asn Leu	Gln Arg Ser Val Thr Tyr Tyr Gln Thr Lys Ser Gly	225		230		235
Arg Glu Leu Pro	Ile Glu Phe Tyr Val Pro Pro Ser Ala Thr Ser Lys	245		250		255
Cys Asp Phe Ala	Lys Glu Val Leu Lys Glu Ala Met Ala Trp Asp Glu	260		265		270
Arg Thr Phe Asn	Leu Glu Cys Ala Leu Arg Gln His Met Val Ala Gly	275		280		285
Val Asp Lys Tyr	Ala Ser Gly Ala Ser Glu Pro Thr Gly Leu Asn Leu	290		295		300
Phe Asn Thr Glu	Asn Leu Phe Ala Ser Pro Glu Thr Lys Thr Asp Leu	305		310		315
Gly Ile Leu Arg	Val Leu Glu Val Val Ala His Glu Phe Phe His Tyr	325		330		335
Trp Ser Gly Asp	Arg Val Thr Ile Arg Asp Trp Phe Asn Leu Pro Leu	340		345		350
Lys Glu Gly Leu	Thr Thr Phe Arg Ala Ala Met Phe Arg Glu Glu Leu	355		360		365
Phe Gly Thr Asp	Leu Ile Arg Leu Leu Asp Gly Lys Asn Leu Asp Glu	370		375		380
Arg Ala Pro Arg	Gln Ser Ala Tyr Thr Ala Val Arg Ser Leu Tyr Thr	385		390		395
Ala Ala Ala Tyr	Glu Lys Ser Ala Asp Ile Phe Arg Met Met Met Leu	405		410		415
Phe Ile Gly Lys	Glu Pro Phe Ile Glu Ala Val Ala Lys Phe Phe Lys	420		425		430

Asp Asn Asp Gly Gly Ala Val Thr Leu Glu Asp Phe Ile Glu Ser Ile
435 440 445

Ser Asn Ser Ser Gly Lys Asp Leu Arg Ser Phe Leu Ser Trp Phe Thr
450 455 460

Glu Ser Gly Ile Pro Glu Leu Ile Val Thr Asp Glu Leu Asn Pro Asp
465 470 475 480

Thr Lys Gln Tyr Phe Leu Lys Ile Lys Thr Val Asn Gly Arg Asn Arg
485 490 495

Pro Ile Pro Ile Leu Met Gly Leu Leu Asp Ser Ser Gly Ala Glu Ile
500 505 510

Val Ala Asp Lys Leu Leu Ile Val Asp Gln Glu Glu Ile Glu Phe Gln
515 520 525

Phe Glu Asn Ile Gln Thr Arg Pro Ile Pro Ser Leu Leu Arg Ser Phe
530 535 540

Ser Ala Pro Val His Met Lys Tyr Glu Tyr Ser Tyr Gln Asp Leu Leu
545 550 555 560

Leu Leu Met Gln Phe Asp Thr Asn Leu Tyr Asn Arg Cys Glu Ala Ala
565 570 575

Lys Gln Leu Ile Ser Ala Leu Ile Asn Asp Phe Cys Ile Gly Lys Lys
580 585 590

Ile Glu Leu Ser Pro Gln Phe Phe Ala Val Tyr Lys Ala Leu Leu Ser
595 600 605

Asp Asn Ser Leu Asn Glu Trp Met Leu Ala Glu Leu Ile Thr Leu Pro
610 615 620

Ser Leu Glu Glu Leu Ile Glu Asn Gln Asp Lys Pro Asp Phe Glu Lys
625 630 635 640

Leu Asn Glu Gly Arg Gln Leu Ile Gln Asn Ala Leu Ala Asn Glu Leu
645 650 655

Lys Thr Asp Phe Tyr Asn Leu Leu Phe Arg Ile Gln Ile Ser Gly Asp
660 665 670

Asp Asp Lys Gln Lys Leu Lys Gly Phe Asp Leu Lys Gln Ala Gly Leu
675 680 685

Arg Arg Leu Lys Ser Val Cys Phe Ser Tyr Leu Leu Asn Val Asp Phe
690 695 700

Glu Lys Thr Lys Glu Lys Leu Ile Leu Gln Phe Glu Asp Ala Leu Gly
705 710 715 720

Lys Asn Met Thr Glu Thr Ala Leu Ala Leu Ser Met Leu Cys Glu Ile
725 730 735

Asn Cys Glu Glu Ala Asp Val Ala Leu Glu Asp Tyr Tyr His Tyr Trp
740 745 750

Lys Asn Asp Pro Gly Ala Val Asn Asn Trp Phe Ser Ile Gln Ala Leu
755 760 765

Ala His Ser Pro Asp Val Ile Glu Arg Val Lys Lys Leu Met Arg His
770 775 780

Gly Asp Phe Asp Leu Ser Asn Pro Asn Lys Val Tyr Ala Leu Leu Gly
785 790 795 800

Ser Phe Ile Lys Asn Pro Phe Gly Phe His Ser Val Thr Gly Glu Gly
805 810 815

Tyr Gln Leu Val Ala Asp Ala Ile Phe Asp Leu Asp Lys Ile Asn Pro
820 825 830

Thr Leu Ala Ala Asn Leu Thr Glu Lys Phe Thr Tyr Trp Asp Lys Tyr
835 840 845

Asp Val Asn Arg Gln Ala Met Met Ile Ser Thr Leu Lys Ile Ile Tyr
850 855 860

Ser Asn Ala Thr Ser Ser Asp Val Arg Thr Met Ala Lys Lys Gly Leu
865 870 875 880

Asp Lys Val Lys Glu Asp Leu Pro Leu Pro Ile His Leu Thr Phe His
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Gly Gly Ser Thr Met Gln Asp Arg Thr Ala Gln Leu Ile Ala Asp Gly
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Asn Lys Glu Asn Ala Tyr Gln Leu His
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Thr Pro Glu Asp Ile Glu Lys Met Arg Val Ala Gly Arg Leu Ala Ala
 20 25 30

Glu Val Leu Glu Met Ile Glu Pro Tyr Val Lys Pro Gly Val Ser Thr
 35 40 45

Gly Glu Leu Asp Arg Ile Cys Asn Asp Tyr Ile Val Asn Glu Gln His
 50 55 60

Ala Val Ser Ala Cys Leu Gly Tyr His Gly Tyr Pro Lys Ser Val Cys
 65 70 75 80

Ile Ser Ile Asn Glu Val Val Cys His Gly Ile Pro Asp Asp Ala Lys
 85 90 95

Leu Leu Lys Asp Gly Asp Ile Val Asn Ile Asp Val Thr Val Ile Lys
 100 105 110

Asp Gly Phe His Gly Asp Thr Ser Lys Met Phe Ile Val Gly Lys Pro
 115 120 125

Thr Ile Met Gly Glu Arg Leu Cys Arg Ile Thr Gln Glu Ser Leu Tyr
 130 135 140

Leu Ala Leu Arg Met Val Lys Pro Gly Ile Asn Leu Arg Glu Ile Gly
 145 150 155 160

Ala Ala Ile Gln Lys Phe Val Glu Ala Glu Gly Phe Ser Val Val Arg
 165 170 175

Glu Tyr Cys Gly His Gly Ile Gly Arg Gly Phe His Glu Glu Pro Gln
 180 185 190

Val Leu His Tyr Asp Ser Arg Glu Thr Asn Val Val Leu Lys Pro Gly
 195 200 205

Met Thr Phe Thr Ile Glu Pro Met Val Asn Ala Gly Lys Lys Glu Ile
 210 215 220

Arg Thr Met Lys Asp Gly Trp Thr Val Lys Thr Lys Asp Arg Ser Leu
 225 230 235 240

Ser Ala Gln Tyr Glu His Thr Ile Val Val Thr Asp Asn Gly Cys Glu
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Ile Leu Thr Leu Arg Lys Asp Asp Thr Ile Pro Ala Ile Ile Ser His
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<212> PRT
<213> Artificial Sequence

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<400> 3

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Val Pro Phe Gly Asp His Glu Thr Trp Val Gln Val Thr Thr Pro Glu
35 40 45

Asn Ala Gln Pro His Ala Leu Pro Leu Ile Val Leu His Gly Gly Pro
50 55 60

Gly Met Ala His Asn Tyr Val Ala Asn Ile Ala Ala Leu Ala Asp Glu
65 70 75 80

Thr Gly Arg Thr Val Ile His Tyr Asp Gln Val Gly Cys Gly Asn Ser
85 90 95

Thr His Leu Pro Asp Ala Pro Ala Asp Phe Trp Thr Pro Gln Leu Phe
100 105 110

Val Asp Glu Phe His Ala Val Cys Thr Ala Leu Gly Ile Glu Arg Tyr
115 120 125

His Val Leu Gly Gln Ser Trp Gly Gly Met Leu Gly Ala Glu Ile Ala
130 135 140

Val Arg Gln Pro Ser Gly Leu Val Ser Leu Ala Ile Cys Asn Ser Pro
145 150 155 160

Ala Ser Met Arg Leu Trp Ser Glu Ala Ala Gly Asp Leu Arg Ala Gln
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Leu Pro Ala Glu Thr Arg Ala Ala Leu Asp Arg His Glu Ala Ala Gly
180 185 190

Thr Ile Thr His Pro Asp Tyr Leu Gln Ala Ala Ala Glu Phe Tyr Arg
195 200 205

Arg His Val Cys Arg Val Val Pro Thr Pro Gln Asp Phe Ala Asp Ser
210 215 220

Val Ala Gln Met Glu Ala Glu Pro Thr Val Tyr His Thr Met Asn Gly
225 230 235 240

Pro Asn Glu Phe His Val Val Gly Thr Leu Gly Asp Trp Ser Val Ile
245 250 255

Asp Arg Leu Pro Asp Val Thr Ala Pro Val Leu Val Ile Ala Gly Glu
260 265 270

His Asp Glu Ala Thr Pro Lys Thr Trp Gln Pro Phe Val Asp His Ile
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Pro Asp Val Arg Ser His Val Phe Pro Gly Thr Ser His Cys Thr His
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Tyr Leu Thr Gly Phe Asn Glu Pro Glu Ala Val Leu Ile Leu Val Lys
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Ser Asp Glu Thr His Asn His Ser Val Leu Phe Asn Arg Ile Arg Asp
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Leu Thr Ala Glu Ile Trp Phe Gly Arg Arg Leu Gly Gln Glu Ala Ala
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Pro Thr Lys Leu Ala Val Asp Arg Ala Leu Pro Phe Asp Glu Ile Asn
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Glu Gln Leu Tyr Leu Leu Leu Asn Arg Leu Asp Val Ile Tyr His Ala
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Gln Gly Gln Tyr Ala Tyr Ala Asp Asn Ile Val Phe Ala Ala Leu Glu

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Lys Leu Arg His Gly Phe Arg Lys Asn Leu Arg Ala Pro Ala Thr Leu				
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Thr Asp Trp Arg Pro Trp Leu His Glu Met Arg Leu Phe Lys Ser Ala				
		165		170
				175
Glu Glu Ile Ala Val Leu Arg Arg Ala Gly Glu Ile Ser Ala Leu Ala				
		180		185
				190
His Thr Arg Ala Met Glu Lys Cys Arg Pro Gly Met Phe Glu Tyr Gln				
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				205
Leu Glu Gly Glu Ile Leu His Glu Phe Thr Arg His Gly Ala Arg Tyr				
		210		215
				220
Pro Ala Tyr Asn Thr Ile Val Gly Gly Gly Glu Asn Gly Cys Ile Leu				
		225		230
				235
His Tyr Thr Glu Asn Glu Cys Glu Leu Arg Asp Gly Asp Leu Val Leu				
		245		250
				255
Ile Asp Ala Gly Cys Glu Tyr Arg Gly Tyr Ala Gly Asp Ile Thr Arg				
		260		265
				270
Thr Phe Pro Val Asn Gly Lys Phe Thr Pro Ala Gln Arg Ala Val Tyr				
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				285
Asp Ile Val Leu Ala Ala Ile Asn Lys Ser Leu Thr Leu Phe Arg Pro				
		290		295
				300
Gly Thr Ser Ile Arg Glu Val Thr Glu Glu Val Val Arg Ile Met Val				
		305		310
				315
Val Gly Leu Val Glu Leu Gly Ile Leu Lys Gly Asp Ile Glu Gln Leu				
		325		330
				335
Ile Ala Glu Gln Ala His Arg Pro Phe Phe Met His Gly Leu Ser His				
		340		345
				350
Trp Leu Gly Met Asp Val His Asp Val Gly Asp Tyr Gly Ser Ser Asp				
		355		360
				365
Arg Gly Arg Ile Leu Glu Pro Gly Met Val Leu Thr Val Glu Pro Gly				
		370		375
				380
Leu Tyr Ile Ala Pro Asp Ala Asp Val Pro Pro Gln Tyr Arg Gly Ile				
		385		390
				395
Gly Ile Arg Ile Glu Asp Asp Ile Val Ile Thr Ala Thr Gly Asn Glu				
		405		410
				415

Asn Leu Thr Ala Ser Val Val Lys Asp Pro Asp Asp Ile Glu Ala Leu
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His His His His
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Leu Ala Glu Ser Ile Glu Lys Met Ile Met Glu Leu Gly Gly Lys Pro
35 40 45

Ala Phe Pro Val Asn Leu Ser Ile Asn Glu Ile Ala Ala His Tyr Thr
50 55 60

Pro Tyr Lys Gly Asp Thr Thr Val Leu Lys Glu Gly Asp Tyr Leu Lys
65 70 75 80

Ile Asp Val Gly Val His Ile Asp Gly Phe Ile Ala Asp Thr Ala Val
85 90 95

Thr Val Arg Val Gly Met Glu Glu Asp Glu Leu Met Glu Ala Ala Lys
100 105 110

Glu Ala Leu Asn Ala Ala Ile Ser Val Ala Arg Ala Gly Val Glu Ile
115 120 125

Lys Glu Leu Gly Lys Ala Ile Glu Asn Glu Ile Arg Lys Arg Gly Phe
130 135 140

Lys Pro Ile Val Asn Leu Ser Gly His Lys Ile Glu Arg Tyr Lys Leu
145 150 155 160

His Ala Gly Ile Ser Ile Pro Asn Ile Tyr Arg Pro His Asp Asn Tyr
165 170 175

Val Leu Lys Glu Gly Asp Val Phe Ala Ile Glu Pro Phe Ala Thr Ile

180

185

190

Gly Ala Gly Gln Val Ile Glu Val Pro Pro Thr Leu Ile Tyr Met Tyr
 195 200 205

Val Arg Asp Val Pro Val Arg Val Ala Gln Ala Arg Phe Leu Leu Ala
 210 215 220

Lys Ile Lys Arg Glu Tyr Gly Thr Leu Pro Phe Ala Tyr Arg Trp Leu
 225 230 235 240

Gln Asn Asp Met Pro Glu Gly Gln Leu Lys Leu Ala Leu Lys Thr Leu
 245 250 255

Glu Lys Ala Gly Ala Ile Tyr Gly Tyr Pro Val Leu Lys Glu Ile Arg
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Asn Gly Ile Val Ala Gln Phe Glu His Thr Ile Ile Val Glu Lys Asp
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 20 25 30

Glu Thr Ile Thr Leu Phe Gly Arg Thr Leu Cys Arg Lys Asp Arg Leu
 35 40 45

Asp Asp Glu Leu Pro Trp Leu Leu Tyr Leu Gln Gly Gly Pro Gly Phe
 50 55 60

Gly Ala Pro Arg Pro Ser Ala Asn Gly Gly Trp Ile Lys Arg Ala Leu
 65 70 75 80

Gln Glu Phe Arg Val Leu Leu Leu Asp Gln Arg Gly Thr Gly His Ser
 85 90 95

Thr Pro Ile His Ala Glu Leu Leu Ala His Leu Asn Pro Arg Gln Gln
 100 105 110

Ala Asp Tyr Leu Ser His Phe Arg Ala Asp Ser Ile Val Arg Asp Ala
115 120 125

Glu Leu Ile Arg Glu Gln Leu Ser Pro Asp His Pro Trp Ser Leu Leu
130 135 140

Gly Gln Ser Phe Gly Gly Phe Cys Ser Leu Thr Tyr Leu Ser Leu Phe
145 150 155 160

Pro Asp Ser Leu His Glu Val Tyr Leu Thr Gly Gly Val Ala Pro Ile
165 170 175

Gly Arg Ser Ala Asp Glu Val Tyr Arg Ala Thr Tyr Gln Arg Val Ala
180 185 190

Asp Lys Asn Arg Ala Phe Phe Ala Arg Phe Pro His Ala Gln Ala Ile
195 200 205

Ala Asn Arg Leu Ala Thr His Leu Gln Arg His Asp Val Arg Leu Pro
210 215 220

Asn Gly Gln Arg Leu Thr Val Glu Gln Leu Gln Gln Gln Gly Leu Asp
225 230 235 240

Leu Gly Ala Ser Gly Ala Phe Glu Glu Leu Tyr Tyr Leu Leu Glu Asp
245 250 255

Ala Phe Ile Gly Glu Lys Leu Asn Pro Ala Phe Leu Tyr Gln Val Gln
260 265 270

Ala Met Gln Pro Phe Asn Thr Asn Pro Val Phe Ala Ile Leu His Glu
275 280 285

Leu Ile Tyr Cys Glu Gly Ala Ala Ser His Trp Ala Ala Glu Arg Val
290 295 300

Arg Gly Glu Phe Pro Ala Leu Ala Trp Ala Gln Gly Lys Asp Phe Ala
305 310 315 320

Phe Thr Gly Glu Met Ile Phe Pro Trp Met Phe Glu Gln Phe Arg Glu
325 330 335

Leu Ile Pro Leu Lys Glu Ala Ala His Leu Leu Ala Glu Lys Ala Asp
340 345 350

Trp Gly Pro Leu Tyr Asp Pro Val Gln Leu Ala Arg Asn Lys Val Pro
355 360 365

Val Ala Cys Ala Val Tyr Ala Glu Asp Met Tyr Val Glu Phe Asp Tyr
370 375 380

Ser Arg Glu Thr Leu Lys Gly Leu Ser Asn Ser Arg Ala Trp Ile Thr

385 390 395 400

Asn Glu Tyr Glu His Asn Gly Leu Arg Val Asp Gly Glu Gln Ile Leu
405 410 415

Asp Arg Leu Ile Arg Leu Asn Arg Asp Cys Leu Glu
420 425

<210> 7
<211> 348
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 7

Met Lys Glu Arg Leu Glu Lys Leu Val Lys Phe Met Asp Glu Asn Ser
1 5 10 15

Ile Asp Arg Val Phe Ile Ala Lys Pro Val Asn Val Tyr Tyr Phe Ser
20 25 30

Gly Thr Ser Pro Leu Gly Gly Gly Tyr Ile Ile Val Asp Gly Asp Glu
35 40 45

Ala Thr Leu Tyr Val Pro Glu Leu Glu Tyr Glu Met Ala Lys Glu Glu
50 55 60

Ser Lys Leu Pro Val Val Lys Phe Lys Lys Phe Asp Glu Ile Tyr Glu
65 70 75 80

Ile Leu Lys Asn Thr Glu Thr Leu Gly Ile Glu Gly Thr Leu Ser Tyr
85 90 95

Ser Met Val Glu Asn Phe Lys Glu Lys Ser Asn Val Lys Glu Phe Lys
100 105 110

Lys Ile Asp Asp Val Ile Lys Asp Leu Arg Ile Ile Lys Thr Lys Glu
115 120 125

Glu Ile Glu Ile Ile Glu Lys Ala Cys Glu Ile Ala Asp Lys Ala Val
130 135 140

Met Ala Ala Ile Glu Glu Ile Thr Glu Gly Lys Arg Glu Arg Glu Val
145 150 155 160

Ala Ala Lys Val Glu Tyr Leu Met Lys Met Asn Gly Ala Glu Lys Pro
165 170 175

Ala Phe Asp Thr Ile Ile Ala Ser Gly His Arg Ser Ala Leu Pro His
180 185 190

Gly Val Ala Ser Asp Lys Arg Ile Glu Arg Gly Asp Leu Val Val Ile
195 200 205

Asp Leu Gly Ala Leu Tyr Asn His Tyr Asn Ser Asp Ile Thr Arg Thr
210 215 220

Ile Val Val Gly Ser Pro Asn Glu Lys Gln Arg Glu Ile Tyr Glu Ile
225 230 235 240

Val Leu Glu Ala Gln Lys Arg Ala Val Glu Ala Ala Lys Pro Gly Met
245 250 255

Thr Ala Lys Glu Leu Asp Ser Ile Ala Arg Glu Ile Ile Lys Glu Tyr
260 265 270

Gly Tyr Gly Asp Tyr Phe Ile His Ser Leu Gly His Gly Val Gly Leu
275 280 285

Glu Ile His Glu Trp Pro Arg Ile Ser Gln Tyr Asp Glu Thr Val Leu
290 295 300

Lys Glu Gly Met Val Ile Thr Ile Glu Pro Gly Ile Tyr Ile Pro Lys
305 310 315 320

Leu Gly Gly Val Arg Ile Glu Asp Thr Val Leu Ile Thr Glu Asn Gly
325 330 335

Ala Lys Arg Leu Thr Lys Thr Glu Arg Glu Leu Leu
340 345

<210> 8
<211> 298
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 8

Met Ile Pro Ile Thr Thr Pro Val Gly Asn Phe Lys Val Trp Thr Lys
1 5 10 15

Arg Phe Gly Thr Asn Pro Lys Ile Lys Val Leu Leu Leu His Gly Gly
20 25 30

Pro Ala Met Thr His Glu Tyr Met Glu Cys Phe Glu Thr Phe Phe Gln
35 40 45

Arg Glu Gly Phe Glu Phe Tyr Glu Tyr Asp Gln Leu Gly Ser Tyr Tyr
50 55 60

Ser Asp Gln Pro Thr Asp Glu Lys Leu Trp Asn Ile Asp Arg Phe Val
65 70 75 80

Asp Glu Val Glu Gln Val Arg Lys Ala Ile His Ala Asp Lys Glu Asn
85 90 95

Phe Tyr Val Leu Gly Asn Ser Trp Gly Gly Ile Leu Ala Met Glu Tyr
100 105 110

Ala Leu Lys Tyr Gln Gln Asn Leu Lys Gly Leu Ile Val Ala Asn Met
115 120 125

Met Ala Ser Ala Pro Glu Tyr Val Lys Tyr Ala Glu Val Leu Ser Lys
130 135 140

Gln Met Lys Pro Glu Val Leu Ala Glu Val Arg Ala Ile Glu Ala Lys
145 150 155 160

Lys Asp Tyr Ala Asn Pro Arg Tyr Thr Glu Leu Leu Phe Pro Asn Tyr
165 170 175

Tyr Ala Gln His Ile Cys Arg Leu Lys Glu Trp Pro Asp Ala Leu Asn
180 185 190

Arg Ser Leu Lys His Val Asn Ser Thr Val Tyr Thr Leu Met Gln Gly
195 200 205

Pro Ser Glu Leu Gly Met Ser Ser Asp Ala Arg Leu Ala Lys Trp Asp
210 215 220

Ile Lys Asn Arg Leu His Glu Ile Ala Thr Pro Thr Leu Met Ile Gly
225 230 235 240

Ala Arg Tyr Asp Thr Met Asp Pro Lys Ala Met Glu Glu Gln Ser Lys
245 250 255

Leu Val Gln Lys Gly Arg Tyr Leu Tyr Cys Pro Asn Gly Ser His Leu
260 265 270

Ala Met Trp Asp Asp Gln Lys Val Phe Met Asp Gly Val Ile Lys Phe
275 280 285

Ile Lys Asp Val Asp Thr Lys Ser Phe Asn
290 295

<210> 9
<211> 428
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 9

His Met Ser Ser Pro Leu His Tyr Val Leu Asp Gly Ile His Cys Glu
1 5 10 15

Pro His Phe Phe Thr Val Pro Leu Asp His Gln Gln Pro Asp Asp Glu
 20 25 30

Glu Thr Ile Thr Leu Phe Gly Arg Thr Leu Cys Arg Lys Asp Arg Leu
 35 40 45

Asp Asp Glu Leu Pro Trp Leu Leu Tyr Leu Gln Gly Gly Pro Gly Phe
 50 55 60

Gly Ala Pro Arg Pro Ser Ala Asn Gly Gly Trp Ile Lys Arg Ala Leu
 65 70 75 80

Gln Glu Phe Arg Val Leu Leu Leu Asp Gln Arg Gly Thr Gly His Ser
 85 90 95

Thr Pro Ile His Ala Glu Leu Leu Ala His Leu Asn Pro Arg Gln Gln
 100 105 110

Ala Asp Tyr Leu Ser His Phe Arg Ala Asp Ser Ile Val Arg Asp Ala
 115 120 125

Glu Leu Ile Arg Glu Gln Leu Ser Pro Asp His Pro Trp Ser Leu Leu
 130 135 140

Gly Gln Ser Phe Gly Gly Phe Cys Ser Leu Thr Tyr Leu Ser Leu Phe
 145 150 155 160

Pro Asp Ser Leu His Glu Val Tyr Leu Thr Gly Gly Val Ala Pro Ile
 165 170 175

Gly Arg Ser Ala Asp Glu Val Tyr Arg Ala Thr Tyr Gln Arg Val Ala
 180 185 190

Asp Lys Asn Arg Ala Phe Phe Ala Arg Phe Pro His Ala Gln Ala Ile
 195 200 205

Ala Asn Arg Leu Ala Thr His Leu Gln Arg His Asp Val Arg Leu Pro
 210 215 220

Asn Gly Gln Arg Leu Thr Val Glu Gln Leu Gln Gln Gln Gly Leu Asp
 225 230 235 240

Leu Gly Ala Ser Gly Ala Phe Glu Glu Leu Tyr Tyr Leu Leu Glu Asp
 245 250 255

Ala Phe Ile Gly Glu Lys Leu Asn Pro Ala Phe Leu Tyr Gln Val Gln
 260 265 270

Ala Met Gln Pro Phe Asn Thr Asn Pro Val Phe Ala Ile Leu His Glu
 275 280 285

Leu Ile Tyr Cys Glu Gly Ala Ala Ser His Trp Ala Ala Glu Arg Val
290 295 300

Arg Gly Glu Phe Pro Ala Leu Ala Trp Ala Gln Gly Lys Asp Phe Ala
305 310 315 320

Phe Thr Gly Glu Met Ile Phe Pro Trp Met Phe Glu Gln Phe Arg Glu
325 330 335

Leu Ile Pro Leu Lys Glu Ala Ala His Leu Leu Ala Glu Lys Ala Asp
340 345 350

Trp Gly Pro Leu Tyr Asp Pro Val Gln Leu Ala Arg Asn Lys Val Pro
355 360 365

Val Ala Cys Ala Val Tyr Ala Glu Asp Met Tyr Val Glu Phe Asp Tyr
370 375 380

Ser Arg Glu Thr Leu Lys Gly Leu Ser Asn Ser Arg Ala Trp Ile Thr
385 390 395 400

Asn Glu Tyr Glu His Asn Gly Leu Arg Val Asp Gly Glu Gln Ile Leu
405 410 415

Asp Arg Leu Ile Arg Leu Asn Arg Asp Cys Leu Glu
420 425

<210> 10
<211> 310
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 10

Met Tyr Glu Ile Lys Gln Pro Phe His Ser Gly Tyr Leu Gln Val Ser
1 5 10 15

Glu Ile His Gln Ile Tyr Trp Glu Glu Ser Gly Asn Pro Asp Gly Val
20 25 30

Pro Val Ile Phe Leu His Gly Gly Pro Gly Ala Gly Ala Ser Pro Glu
35 40 45

Cys Arg Gly Phe Phe Asn Pro Asp Val Phe Arg Ile Val Ile Ile Asp
50 55 60

Gln Arg Gly Cys Gly Arg Ser His Pro Tyr Ala Cys Ala Glu Asp Asn
65 70 75 80

Thr Thr Trp Asp Leu Val Ala Asp Ile Glu Lys Val Arg Glu Met Leu
85 90 95

Gly Ile Gly Lys Trp Leu Val Phe Gly Gly Ser Trp Gly Ser Thr Leu
100 105 110

Ser Leu Ala Tyr Ala Gln Thr His Pro Glu Arg Val Lys Gly Leu Val
115 120 125

Leu Arg Gly Ile Phe Leu Cys Arg Pro Ser Glu Thr Ala Trp Leu Asn
130 135 140

Glu Ala Gly Gly Val Ser Arg Ile Tyr Pro Glu Gln Trp Gln Lys Phe
145 150 155 160

Val Ala Pro Ile Ala Glu Asn Arg Arg Asn Arg Leu Ile Glu Ala Tyr
165 170 175

His Gly Leu Leu Phe His Gln Asp Glu Glu Val Cys Leu Ser Ala Ala
180 185 190

Lys Ala Trp Ala Asp Trp Glu Ser Tyr Leu Ile Arg Phe Glu Pro Glu
195 200 205

Gly Val Asp Glu Asp Ala Tyr Ala Ser Leu Ala Ile Ala Arg Leu Glu
210 215 220

Asn His Tyr Phe Val Asn Gly Gly Trp Leu Gln Gly Asp Lys Ala Ile
225 230 235 240

Leu Asn Asn Ile Gly Lys Ile Arg His Ile Pro Thr Val Ile Val Gln
245 250 255

Gly Arg Tyr Asp Leu Cys Thr Pro Met Gln Ser Ala Trp Glu Leu Ser
260 265 270

Lys Ala Phe Pro Glu Ala Glu Leu Arg Val Val Gln Ala Gly His Cys
275 280 285

Ala Phe Asp Pro Pro Leu Ala Asp Ala Leu Val Gln Ala Val Glu Asp
290 295 300

Ile Leu Pro Arg Leu Leu
305 310

<210> 11
<211> 891
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 11

Met Gly Ser Ser His His His His His Ser Ser Gly Glu Asn Leu

1	5	10	15
Tyr Phe Gln Gly His Met Thr Gln Gln Pro Gln Ala Lys Tyr Arg His	20	25	30
Asp Tyr Arg Ala Pro Asp Tyr Gln Ile Thr Asp Ile Asp Leu Thr Phe	35	40	45
Asp Leu Asp Ala Gln Lys Thr Val Val Thr Ala Val Ser Gln Ala Val	50	55	60
Arg His Gly Ala Ser Asp Ala Pro Leu Arg Leu Asn Gly Glu Asp Leu	65	70	75
Lys Leu Val Ser Val His Ile Asn Asp Glu Pro Trp Thr Ala Trp Lys	85	90	95
Glu Glu Glu Gly Ala Leu Val Ile Ser Asn Leu Pro Glu Arg Phe Thr	100	105	110
Leu Lys Ile Ile Asn Glu Ile Ser Pro Ala Ala Asn Thr Ala Leu Glu	115	120	125
Gly Leu Tyr Gln Ser Gly Asp Ala Leu Cys Thr Gln Cys Glu Ala Glu	130	135	140
Gly Phe Arg His Ile Thr Tyr Tyr Leu Asp Arg Pro Asp Val Leu Ala	145	150	155
Arg Phe Thr Thr Lys Ile Ile Ala Asp Lys Ile Lys Tyr Pro Phe Leu	165	170	175
Leu Ser Asn Gly Asn Arg Val Ala Gln Gly Glu Leu Glu Asn Gly Arg	180	185	190
His Trp Val Gln Trp Gln Asp Pro Phe Pro Lys Pro Cys Tyr Leu Phe	195	200	205
Ala Leu Val Ala Gly Asp Phe Asp Val Leu Arg Asp Thr Phe Thr Thr	210	215	220
Arg Ser Gly Arg Glu Val Ala Leu Glu Leu Tyr Val Asp Arg Gly Asn	225	230	235
Leu Asp Arg Ala Pro Trp Ala Met Thr Ser Leu Lys Asn Ser Met Lys	245	250	255
Trp Asp Glu Glu Arg Phe Gly Leu Glu Tyr Asp Leu Asp Ile Tyr Met	260	265	270
Ile Val Ala Val Asp Phe Phe Asn Met Gly Ala Met Glu Asn Lys Gly	275	280	285

Leu Asn Ile Phe Asn Ser Lys Tyr Val Leu Ala Arg Thr Asp Thr Ala
290 295 300

Thr Asp Lys Asp Tyr Leu Asp Ile Glu Arg Val Ile Gly His Glu Tyr
305 310 315 320

Phe His Asn Trp Thr Gly Asn Arg Val Thr Cys Arg Asp Trp Phe Gln
325 330 335

Leu Ser Leu Lys Glu Gly Leu Thr Val Phe Arg Asp Gln Glu Phe Ser
340 345 350

Ser Asp Leu Gly Ser Arg Ala Val Asn Arg Ile Asn Asn Val Arg Thr
355 360 365

Met Arg Gly Leu Gln Phe Ala Glu Asp Ala Ser Pro Met Ala His Pro
370 375 380

Ile Arg Pro Asp Met Val Ile Glu Met Asn Asn Phe Tyr Thr Leu Thr
385 390 395 400

Val Tyr Glu Lys Gly Ala Glu Val Ile Arg Met Ile His Thr Leu Leu
405 410 415

Gly Glu Glu Asn Phe Gln Lys Gly Met Gln Leu Tyr Phe Glu Arg His
420 425 430

Asp Gly Ser Ala Ala Thr Cys Asp Asp Phe Val Gln Ala Met Glu Asp
435 440 445

Ala Ser Asn Val Asp Leu Ser His Phe Arg Arg Trp Tyr Ser Gln Ser
450 455 460

Gly Thr Pro Ile Val Thr Val Lys Asp Asp Tyr Asn Pro Glu Thr Glu
465 470 475 480

Gln Tyr Thr Leu Thr Ile Ser Gln Arg Thr Pro Ala Thr Pro Asp Gln
485 490 495

Ala Glu Lys Gln Pro Leu His Ile Pro Phe Ala Ile Glu Leu Tyr Asp
500 505 510

Asn Glu Gly Lys Val Ile Pro Leu Gln Lys Gly Gly His Pro Val Asn
515 520 525

Ser Val Leu Asn Val Thr Gln Ala Glu Gln Thr Phe Val Phe Asp Asn
530 535 540

Val Tyr Phe Gln Pro Val Pro Ala Leu Leu Cys Glu Phe Ser Ala Pro
545 550 555 560

Val Lys Leu Glu Tyr Lys Trp Ser Asp Gln Gln Leu Thr Phe Leu Met
565 570 575

Arg His Ala Arg Asn Asp Phe Ser Arg Trp Asp Ala Ala Gln Ser Leu
580 585 590

Leu Ala Thr Tyr Ile Lys Leu Asn Val Ala Arg His Gln Gln Gly Gln
595 600 605

Pro Leu Ser Leu Pro Val His Val Ala Asp Ala Phe Arg Ala Val Leu
610 615 620

Leu Asp Glu Lys Ile Asp Pro Ala Leu Ala Ala Glu Ile Leu Thr Leu
625 630 635 640

Pro Ser Val Asn Glu Met Ala Glu Leu Phe Asp Ile Ile Asp Pro Ile
645 650 655

Ala Ile Ala Glu Val Arg Glu Ala Leu Thr Arg Thr Leu Ala Thr Glu
660 665 670

Leu Ala Asp Glu Leu Leu Ala Ile Tyr Asn Ala Asn Tyr Gln Ser Glu
675 680 685

Tyr Arg Val Glu His Glu Asp Ile Ala Lys Arg Thr Leu Arg Asn Ala
690 695 700

Cys Leu Arg Phe Leu Ala Phe Gly Glu Thr His Leu Ala Asp Val Leu
705 710 715 720

Val Ser Lys Gln Phe His Glu Ala Asn Asn Met Thr Asp Ala Leu Ala
725 730 735

Ala Leu Ser Ala Ala Val Ala Ala Gln Leu Pro Cys Arg Asp Ala Leu
740 745 750

Met Gln Glu Tyr Asp Asp Lys Trp His Gln Asn Gly Leu Val Met Asp
755 760 765

Lys Trp Phe Ile Leu Gln Ala Thr Ser Pro Ala Ala Asn Val Leu Glu
770 775 780

Thr Val Arg Gly Leu Leu Gln His Arg Ser Phe Thr Met Ser Asn Pro
785 790 795 800

Asn Arg Ile Arg Ser Leu Ile Gly Ala Phe Ala Gly Ser Asn Pro Ala
805 810 815

Ala Phe His Ala Glu Asp Gly Ser Gly Tyr Leu Phe Leu Val Glu Met
820 825 830

Leu Thr Asp Leu Asn Ser Arg Asn Pro Gln Val Ala Ser Arg Leu Ile
835 840 845

Glu Pro Leu Ile Arg Leu Lys Arg Tyr Asp Ala Lys Arg Gln Glu Lys
850 855 860

Met Arg Ala Ala Leu Glu Gln Leu Lys Gly Leu Glu Asn Leu Ser Gly
865 870 875 880

Asp Leu Tyr Glu Lys Ile Thr Lys Ala Leu Ala
885 890

<210> 12
<211> 889
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 12

Pro Lys Ile His Tyr Arg Lys Asp Tyr Lys Pro Ser Gly Phe Ile Ile
1 5 10 15

Asn Gln Val Thr Leu Asn Ile Asn Ile His Asp Gln Glu Thr Ile Val
20 25 30

Arg Ser Val Leu Asp Met Asp Ile Ser Lys His Asn Val Gly Glu Asp
35 40 45

Leu Val Phe Asp Gly Val Gly Leu Lys Ile Asn Glu Ile Ser Ile Asn
50 55 60

Asn Lys Lys Leu Val Glu Gly Glu Glu Tyr Thr Tyr Asp Asn Glu Phe
65 70 75 80

Leu Thr Ile Phe Ser Lys Phe Val Pro Lys Ser Lys Phe Ala Phe Ser
85 90 95

Ser Glu Val Ile Ile His Pro Glu Thr Asn Tyr Ala Leu Thr Gly Leu
100 105 110

Tyr Lys Ser Lys Asn Ile Ile Val Ser Gln Cys Glu Ala Thr Gly Phe
115 120 125

Arg Arg Ile Thr Phe Phe Ile Asp Arg Pro Asp Met Met Ala Lys Tyr
130 135 140

Asp Val Thr Val Thr Ala Asp Lys Glu Lys Tyr Pro Val Leu Leu Ser
145 150 155 160

Asn Gly Asp Lys Val Asn Glu Phe Glu Ile Pro Gly Gly Arg His Gly
165 170 175

Ala Arg Phe Asn Asp Pro Pro Leu Lys Pro Cys Tyr Leu Phe Ala Val
180 185 190

Val Ala Gly Asp Leu Lys His Leu Ser Ala Thr Tyr Ile Thr Lys Tyr
195 200 205

Thr Lys Lys Lys Val Glu Leu Tyr Val Phe Ser Glu Glu Lys Tyr Val
210 215 220

Ser Lys Leu Gln Trp Ala Leu Glu Cys Leu Lys Lys Ser Met Ala Phe
225 230 235 240

Asp Glu Asp Tyr Phe Gly Leu Glu Tyr Asp Leu Ser Arg Leu Asn Leu
245 250 255

Val Ala Val Ser Asp Phe Asn Val Gly Ala Met Glu Asn Lys Gly Leu
260 265 270

Asn Ile Phe Asn Ala Asn Ser Leu Leu Ala Ser Lys Lys Asn Ser Ile
275 280 285

Asp Phe Ser Tyr Ala Arg Ile Leu Thr Val Val Gly His Glu Tyr Phe
290 295 300

His Gln Tyr Thr Gly Asn Arg Val Thr Leu Arg Asp Trp Phe Gln Leu
305 310 315 320

Thr Leu Lys Glu Gly Leu Thr Val His Arg Glu Asn Leu Phe Ser Glu
325 330 335

Glu Met Thr Lys Thr Val Thr Thr Arg Leu Ser His Val Asp Leu Leu
340 345 350

Arg Ser Val Gln Phe Leu Glu Asp Ser Ser Pro Leu Ser His Pro Ile
355 360 365

Arg Pro Glu Ser Tyr Val Ser Met Glu Asn Phe Tyr Thr Thr Thr Val
370 375 380

Tyr Asp Lys Gly Ser Glu Val Met Arg Met Tyr Leu Thr Ile Leu Gly
385 390 395 400

Glu Glu Tyr Tyr Lys Lys Gly Phe Asp Ile Tyr Ile Lys Lys Asn Asp
405 410 415

Gly Asn Thr Ala Thr Cys Glu Asp Phe Asn Tyr Ala Met Glu Gln Ala
420 425 430

Tyr Lys Met Lys Lys Ala Asp Asn Ser Ala Asn Leu Asn Gln Tyr Leu
435 440 445

Leu Trp Phe Ser Gln Ser Gly Thr Pro His Val Ser Phe Lys Tyr Asn
450 455 460

Tyr Asp Ala Glu Lys Lys Gln Tyr Ser Ile His Val Asn Gln Tyr Thr
465 470 475 480

Lys Pro Asp Glu Asn Gln Lys Glu Lys Lys Pro Leu Phe Ile Pro Ile
485 490 495

Ser Val Gly Leu Ile Asn Pro Glu Asn Gly Lys Glu Met Ile Ser Gln
500 505 510

Thr Thr Leu Glu Leu Thr Lys Glu Ser Asp Thr Phe Val Phe Asn Asn
515 520 525

Ile Ala Val Lys Pro Ile Pro Ser Leu Phe Arg Gly Phe Ser Ala Pro
530 535 540

Val Tyr Ile Glu Asp Gln Leu Thr Asp Glu Glu Arg Ile Leu Leu Leu
545 550 555 560

Lys Tyr Asp Ser Asp Ala Phe Val Arg Tyr Asn Ser Cys Thr Asn Ile
565 570 575

Tyr Met Lys Gln Ile Leu Met Asn Tyr Asn Glu Phe Leu Lys Ala Lys
580 585 590

Asn Glu Lys Leu Glu Ser Phe Gln Leu Thr Pro Val Asn Ala Gln Phe
595 600 605

Ile Asp Ala Ile Lys Tyr Leu Leu Glu Asp Pro His Ala Asp Ala Gly
610 615 620

Phe Lys Ser Tyr Ile Val Ser Leu Pro Gln Asp Arg Tyr Ile Ile Asn
625 630 635 640

Phe Val Ser Asn Leu Asp Thr Asp Val Leu Ala Asp Thr Lys Glu Tyr
645 650 655

Ile Tyr Lys Gln Ile Gly Asp Lys Leu Asn Asp Val Tyr Tyr Lys Met
660 665 670

Phe Lys Ser Leu Glu Ala Lys Ala Asp Asp Leu Thr Tyr Phe Asn Asp
675 680 685

Glu Ser His Val Asp Phe Asp Gln Met Asn Met Arg Thr Leu Arg Asn
690 695 700

Thr Leu Leu Ser Leu Leu Ser Lys Ala Gln Tyr Pro Asn Ile Leu Asn
705 710 715 720

Glu Ile Ile Glu His Ser Lys Ser Pro Tyr Pro Ser Asn Trp Leu Thr

725

730

735

Ser Leu Ser Val Ser Ala Tyr Phe Asp Lys Tyr Phe Glu Leu Tyr Asp
740 745 750

Lys Thr Tyr Lys Leu Ser Lys Asp Asp Glu Leu Leu Leu Gln Glu Trp
755 760 765

Leu Lys Thr Val Ser Arg Ser Asp Arg Lys Asp Ile Tyr Glu Ile Leu
770 775 780

Lys Lys Leu Glu Asn Glu Val Leu Lys Asp Ser Lys Asn Pro Asn Asp
785 790 795 800

Ile Arg Ala Val Tyr Leu Pro Phe Thr Asn Asn Leu Arg Arg Phe His
805 810 815

Asp Ile Ser Gly Lys Gly Tyr Lys Leu Ile Ala Glu Val Ile Thr Lys
820 825 830

Thr Asp Lys Phe Asn Pro Met Val Ala Thr Gln Leu Cys Glu Pro Phe
835 840 845

Lys Leu Trp Asn Lys Leu Asp Thr Lys Arg Gln Glu Leu Met Leu Asn
850 855 860

Glu Met Asn Thr Met Leu Gln Glu Pro Gln Ile Ser Asn Asn Leu Lys
865 870 875 880

Glu Tyr Leu Leu Arg Leu Thr Asn Lys
885

<210> 13
<211> 932
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 13

Met Gly Ser Ser His His His His His His Ser Ser Gly Met Trp Leu
1 5 10 15

Ala Ala Ala Ala Pro Ser Leu Ala Arg Arg Leu Leu Phe Leu Gly Pro
20 25 30

Pro Pro Pro Pro Leu Leu Leu Leu Val Phe Ser Arg Ser Ser Arg Arg
35 40 45

Arg Leu His Ser Leu Gly Leu Ala Ala Met Pro Glu Lys Arg Pro Phe
50 55 60

Glu Arg Leu Pro Ala Asp Val Ser Pro Ile Asn Tyr Ser Leu Cys Leu
65 70 75 80

Lys Pro Asp Leu Leu Asp Phe Thr Phe Glu Gly Lys Leu Glu Ala Ala
85 90 95

Ala Gln Val Arg Gln Ala Thr Asn Gln Ile Val Met Asn Cys Ala Asp
100 105 110

Ile Asp Ile Ile Thr Ala Ser Tyr Ala Pro Glu Gly Asp Glu Glu Ile
115 120 125

His Ala Thr Gly Phe Asn Tyr Gln Asn Glu Asp Glu Lys Val Thr Leu
130 135 140

Ser Phe Pro Ser Thr Leu Gln Thr Gly Thr Gly Thr Leu Lys Ile Asp
145 150 155 160

Phe Val Gly Glu Leu Asn Asp Lys Met Lys Gly Phe Tyr Arg Ser Lys
165 170 175

Tyr Thr Thr Pro Ser Gly Glu Val Arg Tyr Ala Ala Val Thr Gln Phe
180 185 190

Glu Ala Thr Asp Ala Arg Arg Ala Phe Pro Cys Trp Asp Glu Pro Ala
195 200 205

Ile Lys Ala Thr Phe Asp Ile Ser Leu Val Val Pro Lys Asp Arg Val
210 215 220

Ala Leu Ser Asn Met Asn Val Ile Asp Arg Lys Pro Tyr Pro Asp Asp
225 230 235 240

Glu Asn Leu Val Glu Val Lys Phe Ala Arg Thr Pro Val Met Ser Thr
245 250 255

Tyr Leu Val Ala Phe Val Val Gly Glu Tyr Asp Phe Val Glu Thr Arg
260 265 270

Ser Lys Asp Gly Val Cys Val Arg Val Tyr Thr Pro Val Gly Lys Ala
275 280 285

Glu Gln Gly Lys Phe Ala Leu Glu Val Ala Ala Lys Thr Leu Pro Phe
290 295 300

Tyr Lys Asp Tyr Phe Asn Val Pro Tyr Pro Leu Pro Lys Ile Asp Leu
305 310 315 320

Ile Ala Ile Ala Asp Phe Ala Ala Gly Ala Met Glu Asn Trp Gly Leu
325 330 335

Val Thr Tyr Arg Glu Thr Ala Leu Leu Ile Asp Pro Lys Asn Ser Cys

340

345

350

Ser Ser Ser Arg Gln Trp Val Ala Leu Val Val Gly His Glu Leu Ala
 355 360 365

His Gln Trp Phe Gly Asn Leu Val Thr Met Glu Trp Trp Thr His Leu
 370 375 380

Trp Leu Asn Glu Gly Phe Ala Ser Trp Ile Glu Tyr Leu Cys Val Asp
 385 390 395 400

His Cys Phe Pro Glu Tyr Asp Ile Trp Thr Gln Phe Val Ser Ala Asp
 405 410 415

Tyr Thr Arg Ala Gln Glu Leu Asp Ala Leu Asp Asn Ser His Pro Ile
 420 425 430

Glu Val Ser Val Gly His Pro Ser Glu Val Asp Glu Ile Phe Asp Ala
 435 440 445

Ile Ser Tyr Ser Lys Gly Ala Ser Val Ile Arg Met Leu His Asp Tyr
 450 455 460

Ile Gly Asp Lys Asp Phe Lys Lys Gly Met Asn Met Tyr Leu Thr Lys
 465 470 475 480

Phe Gln Gln Lys Asn Ala Ala Thr Glu Asp Leu Trp Glu Ser Leu Glu
 485 490 495

Asn Ala Ser Gly Lys Pro Ile Ala Ala Val Met Asn Thr Trp Thr Lys
 500 505 510

Gln Met Gly Phe Pro Leu Ile Tyr Val Glu Ala Glu Gln Val Glu Asp
 515 520 525

Asp Arg Leu Leu Arg Leu Ser Gln Lys Lys Phe Cys Ala Gly Gly Ser
 530 535 540

Tyr Val Gly Glu Asp Cys Pro Gln Trp Met Val Pro Ile Thr Ile Ser
 545 550 555 560

Thr Ser Glu Asp Pro Asn Gln Ala Lys Leu Lys Ile Leu Met Asp Lys
 565 570 575

Pro Glu Met Asn Val Val Leu Lys Asn Val Lys Pro Asp Gln Trp Val
 580 585 590

Lys Leu Asn Leu Gly Thr Val Gly Phe Tyr Arg Thr Gln Tyr Ser Ser
 595 600 605

Ala Met Leu Glu Ser Leu Leu Pro Gly Ile Arg Asp Leu Ser Leu Pro
 610 615 620

Pro Val Asp Arg Leu Gly Leu Gln Asn Asp Leu Phe Ser Leu Ala Arg
 625 630 635 640

Ala Gly Ile Ile Ser Thr Val Glu Val Leu Lys Val Met Glu Ala Phe
 645 650 655

Val Asn Glu Pro Asn Tyr Thr Val Trp Ser Asp Leu Ser Cys Asn Leu
 660 665 670

Gly Ile Leu Ser Thr Leu Leu Ser His Thr Asp Phe Tyr Glu Glu Ile
 675 680 685

Gln Glu Phe Val Lys Asp Val Phe Ser Pro Ile Gly Glu Arg Leu Gly
 690 695 700

Trp Asp Pro Lys Pro Gly Glu Gly His Leu Asp Ala Leu Leu Arg Gly
 705 710 715 720

Leu Val Leu Gly Lys Leu Gly Lys Ala Gly His Lys Ala Thr Leu Glu
 725 730 735

Glu Ala Arg Arg Arg Phe Lys Asp His Val Glu Gly Lys Gln Ile Leu
 740 745 750

Ser Ala Asp Leu Arg Ser Pro Val Tyr Leu Thr Val Leu Lys His Gly
 755 760 765

Asp Gly Thr Thr Leu Asp Ile Met Leu Lys Leu His Lys Gln Ala Asp
 770 775 780

Met Gln Glu Glu Lys Asn Arg Ile Glu Arg Val Leu Gly Ala Thr Leu
 785 790 795 800

Leu Pro Asp Leu Ile Gln Lys Val Leu Thr Phe Ala Leu Ser Glu Glu
 805 810 815

Val Arg Pro Gln Asp Thr Val Ser Val Ile Gly Gly Val Ala Gly Gly
 820 825 830

Ser Lys His Gly Arg Lys Ala Ala Trp Lys Phe Ile Lys Asp Asn Trp
 835 840 845

Glu Glu Leu Tyr Asn Arg Tyr Gln Gly Gly Phe Leu Ile Ser Arg Leu
 850 855 860

Ile Lys Leu Ser Val Glu Gly Phe Ala Val Asp Lys Met Ala Gly Glu
 865 870 875 880

Val Lys Ala Phe Phe Glu Ser His Pro Ala Pro Ser Ala Glu Arg Thr
 885 890 895

Ile Gln Gln Cys Cys Glu Asn Ile Leu Leu Asn Ala Ala Trp Leu Lys
900 905 910

Arg Asp Ala Glu Ser Ile His Gln Tyr Leu Leu Gln Arg Lys Ala Ser
915 920 925

Pro Pro Thr Val
930

<210> 14
<211> 932
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 14

Met Gly Ser Ser His His His His His His Ser Ser Gly Met Trp Leu
1 5 10 15

Ala Ala Ala Ala Pro Ser Leu Ala Arg Arg Leu Leu Phe Leu Gly Pro
20 25 30

Pro Pro Pro Pro Leu Leu Leu Leu Val Phe Ser Arg Ser Ser Arg Arg
35 40 45

Arg Leu His Ser Leu Gly Leu Ala Ala Met Pro Glu Lys Arg Pro Phe
50 55 60

Glu Arg Leu Pro Ala Asp Val Ser Pro Ile Asn Tyr Ser Leu Cys Leu
65 70 75 80

Lys Pro Asp Leu Leu Asp Phe Thr Phe Glu Gly Lys Leu Glu Ala Ala
85 90 95

Ala Gln Val Arg Gln Ala Thr Asn Gln Ile Val Met Asn Cys Ala Asp
100 105 110

Ile Asp Ile Ile Thr Ala Ser Tyr Ala Pro Glu Gly Asp Glu Glu Ile
115 120 125

His Ala Thr Gly Phe Asn Tyr Gln Asn Glu Asp Glu Lys Val Thr Leu
130 135 140

Ser Phe Pro Ser Thr Leu Gln Thr Gly Thr Gly Thr Leu Lys Ile Asp
145 150 155 160

Phe Val Gly Glu Leu Asn Asp Lys Met Lys Gly Phe Tyr Arg Ser Lys
165 170 175

Tyr Thr Thr Pro Ser Gly Glu Val Arg Tyr Ala Ala Val Thr Gln Phe
180 185 190

Glu Ala Thr Asp Ala Arg Arg Ala Phe Pro Cys Trp Asp Glu Pro Ala
 195 200 205

Ile Lys Ala Thr Phe Asp Ile Ser Leu Val Val Pro Lys Asp Arg Val
 210 215 220

Ala Leu Ser Asn Met Asn Val Ile Asp Arg Lys Pro Tyr Pro Asp Asp
 225 230 235 240

Glu Asn Leu Val Glu Val Lys Phe Ala Arg Thr Pro Val Met Ser Thr
 245 250 255

Tyr Leu Val Ala Phe Val Val Gly Glu Tyr Asp Phe Val Glu Thr Arg
 260 265 270

Ser Lys Asp Gly Val Cys Val Arg Val Tyr Thr Pro Val Gly Lys Ala
 275 280 285

Glu Gln Gly Lys Phe Ala Leu Glu Val Ala Ala Lys Thr Leu Pro Phe
 290 295 300

Tyr Lys Asp Tyr Phe Asn Val Pro Tyr Pro Leu Pro Lys Ile Asp Leu
 305 310 315 320

Ile Ala Ile Ala Asp Phe Ala Ala Gly Ala Met Glu Asn Trp Gly Leu
 325 330 335

Val Thr Tyr Arg Glu Thr Ala Leu Leu Ile Asp Pro Lys Asn Ser Cys
 340 345 350

Ser Ser Ser Arg Gln Trp Val Ala Leu Val Val Gly His Val Leu Ala
 355 360 365

His Gln Trp Phe Gly Asn Leu Val Thr Met Glu Trp Trp Thr His Leu
 370 375 380

Trp Leu Asn Glu Gly Phe Ala Ser Trp Ile Glu Tyr Leu Cys Val Asp
 385 390 395 400

His Cys Phe Pro Glu Tyr Asp Ile Trp Thr Gln Phe Val Ser Ala Asp
 405 410 415

Tyr Thr Arg Ala Gln Glu Leu Asp Ala Leu Asp Asn Ser His Pro Ile
 420 425 430

Glu Val Ser Val Gly His Pro Ser Glu Val Asp Glu Ile Phe Asp Ala
 435 440 445

Ile Ser Tyr Ser Lys Gly Ala Ser Val Ile Arg Met Leu His Asp Tyr
 450 455 460

Ile Gly Asp Lys Asp Phe Lys Lys Gly Met Asn Met Tyr Leu Thr Lys
465 470 475 480

Phe Gln Gln Lys Asn Ala Ala Thr Glu Asp Leu Trp Glu Ser Leu Glu
485 490 495

Asn Ala Ser Gly Lys Pro Ile Ala Ala Val Met Asn Thr Trp Thr Lys
500 505 510

Gln Met Gly Phe Pro Leu Ile Tyr Val Glu Ala Glu Gln Val Glu Asp
515 520 525

Asp Arg Leu Leu Arg Leu Ser Gln Lys Lys Phe Cys Ala Gly Gly Ser
530 535 540

Tyr Val Gly Glu Asp Cys Pro Gln Trp Met Val Pro Ile Thr Ile Ser
545 550 555 560

Thr Ser Glu Asp Pro Asn Gln Ala Lys Leu Lys Ile Leu Met Asp Lys
565 570 575

Pro Glu Met Asn Val Val Leu Lys Asn Val Lys Pro Asp Gln Trp Val
580 585 590

Lys Leu Asn Leu Gly Thr Val Gly Phe Tyr Arg Thr Gln Tyr Ser Ser
595 600 605

Ala Met Leu Glu Ser Leu Leu Pro Gly Ile Arg Asp Leu Ser Leu Pro
610 615 620

Pro Val Asp Arg Leu Gly Leu Gln Asn Asp Leu Phe Ser Leu Ala Arg
625 630 635 640

Ala Gly Ile Ile Ser Thr Val Glu Val Leu Lys Val Met Glu Ala Phe
645 650 655

Val Asn Glu Pro Asn Tyr Thr Val Trp Ser Asp Leu Ser Cys Asn Leu
660 665 670

Gly Ile Leu Ser Thr Leu Leu Ser His Thr Asp Phe Tyr Glu Glu Ile
675 680 685

Gln Glu Phe Val Lys Asp Val Phe Ser Pro Ile Gly Glu Arg Leu Gly
690 695 700

Trp Asp Pro Lys Pro Gly Glu Gly His Leu Asp Ala Leu Leu Arg Gly
705 710 715 720

Leu Val Leu Gly Lys Leu Gly Lys Ala Gly His Lys Ala Thr Leu Glu
725 730 735

Glu Ala Arg Arg Arg Phe Lys Asp His Val Glu Gly Lys Gln Ile Leu
740 745 750

Ser Ala Asp Leu Arg Ser Pro Val Tyr Leu Thr Val Leu Lys His Gly
755 760 765

Asp Gly Thr Thr Leu Asp Ile Met Leu Lys Leu His Lys Gln Ala Asp
770 775 780

Met Gln Glu Glu Lys Asn Arg Ile Glu Arg Val Leu Gly Ala Thr Leu
785 790 795 800

Leu Pro Asp Leu Ile Gln Lys Val Leu Thr Phe Ala Leu Ser Glu Glu
805 810 815

Val Arg Pro Gln Asp Thr Val Ser Val Ile Gly Gly Val Ala Gly Gly
820 825 830

Ser Lys His Gly Arg Lys Ala Ala Trp Lys Phe Ile Lys Asp Asn Trp
835 840 845

Glu Glu Leu Tyr Asn Arg Tyr Gln Gly Gly Phe Leu Ile Ser Arg Leu
850 855 860

Ile Lys Leu Ser Val Glu Gly Phe Ala Val Asp Lys Met Ala Gly Glu
865 870 875 880

Val Lys Ala Phe Phe Glu Ser His Pro Ala Pro Ser Ala Glu Arg Thr
885 890 895

Ile Gln Gln Cys Cys Glu Asn Ile Leu Leu Asn Ala Ala Trp Leu Lys
900 905 910

Arg Asp Ala Glu Ser Ile His Gln Tyr Leu Leu Gln Arg Lys Ala Ser
915 920 925

Pro Pro Thr Val
930

<210> 15
<211> 864
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 15

Met Ile Tyr Glu Phe Val Met Thr Asp Pro Lys Ile Lys Tyr Leu Lys
1 5 10 15

Asp Tyr Lys Pro Ser Asn Tyr Leu Ile Asp Glu Thr His Leu Ile Phe
20 25 30

Glu Leu Asp Glu Ser Lys Thr Arg Val Thr Ala Asn Leu Tyr Ile Val
35 40 45

Ala Asn Arg Glu Asn Arg Glu Asn Asn Thr Leu Val Leu Asp Gly Val
50 55 60

Glu Leu Lys Leu Leu Ser Ile Lys Leu Asn Asn Lys His Leu Ser Pro
65 70 75 80

Ala Glu Phe Ala Val Asn Glu Asn Gln Leu Ile Ile Asn Asn Val Pro
85 90 95

Glu Lys Phe Val Leu Gln Thr Val Val Glu Ile Asn Pro Ser Ala Asn
100 105 110

Thr Ser Leu Glu Gly Leu Tyr Lys Ser Gly Asp Val Phe Ser Thr Gln
115 120 125

Cys Glu Ala Thr Gly Phe Arg Lys Ile Thr Tyr Tyr Leu Asp Arg Pro
130 135 140

Asp Val Met Ala Ala Phe Thr Val Lys Ile Ile Ala Asp Lys Lys Lys
145 150 155 160

Tyr Pro Ile Ile Leu Ser Asn Gly Asp Lys Ile Asp Ser Gly Asp Ile
165 170 175

Ser Asp Asn Gln His Phe Ala Val Trp Lys Asp Pro Phe Lys Lys Pro
180 185 190

Cys Tyr Leu Phe Ala Leu Val Ala Gly Asp Leu Ala Ser Ile Lys Asp
195 200 205

Thr Tyr Ile Thr Lys Ser Gln Arg Lys Val Ser Leu Glu Ile Tyr Ala
210 215 220

Phe Lys Gln Asp Ile Asp Lys Cys His Tyr Ala Met Gln Ala Val Lys
225 230 235 240

Asp Ser Met Lys Trp Asp Glu Asp Arg Phe Gly Leu Glu Tyr Asp Leu
245 250 255

Asp Thr Phe Met Ile Val Ala Val Pro Asp Phe Asn Ala Gly Ala Met
260 265 270

Glu Asn Lys Gly Leu Asn Ile Phe Asn Thr Lys Tyr Ile Met Ala Ser
275 280 285

Asn Lys Thr Ala Thr Asp Lys Asp Phe Glu Leu Val Gln Ser Val Val
290 295 300

Gly His Glu Tyr Phe His Asn Trp Thr Gly Asp Arg Val Thr Cys Arg
305 310 315 320

Asp Trp Phe Gln Leu Ser Leu Lys Glu Gly Leu Thr Val Phe Arg Asp
325 330 335

Gln Glu Phe Thr Ser Asp Leu Asn Ser Arg Asp Val Lys Arg Ile Asp
340 345 350

Asp Val Arg Ile Ile Arg Ser Ala Gln Phe Ala Glu Asp Ala Ser Pro
355 360 365

Met Ser His Pro Ile Arg Pro Glu Ser Tyr Ile Glu Met Asn Asn Phe
370 375 380

Tyr Thr Val Thr Val Tyr Asn Lys Gly Ala Glu Ile Ile Arg Met Ile
385 390 395 400

His Thr Leu Leu Gly Glu Glu Gly Phe Gln Lys Gly Met Lys Leu Tyr
405 410 415

Phe Glu Arg His Asp Gly Gln Ala Val Thr Cys Asp Asp Phe Val Asn
420 425 430

Ala Met Ala Asp Ala Asn Asn Arg Asp Phe Ser Leu Phe Lys Arg Trp
435 440 445

Tyr Ala Gln Ser Gly Thr Pro Asn Ile Lys Val Ser Glu Asn Tyr Asp
450 455 460

Ala Ser Ser Gln Thr Tyr Ser Leu Thr Leu Glu Gln Thr Thr Leu Pro
465 470 475 480

Thr Ala Asp Gln Lys Glu Lys Gln Ala Leu His Ile Pro Val Lys Met
485 490 495

Gly Leu Ile Asn Pro Glu Gly Lys Asn Ile Ala Glu Gln Val Ile Glu
500 505 510

Leu Lys Glu Gln Lys Gln Thr Tyr Thr Phe Glu Asn Ile Ala Ala Lys
515 520 525

Pro Val Ala Ser Leu Phe Arg Asp Phe Ser Ala Pro Val Lys Val Glu
530 535 540

His Lys Arg Ser Glu Lys Asp Leu Leu His Ile Val Lys Tyr Asp Asn
545 550 555 560

Asn Ala Phe Asn Arg Trp Asp Ser Leu Gln Gln Ile Ala Thr Asn Ile
565 570 575

Ile Leu Asn Asn Ala Asp Leu Asn Asp Glu Phe Leu Asn Ala Phe Lys

580

585

590

Ser Ile Leu His Asp Lys Asp Leu Asp Lys Ala Leu Ile Ser Asn Ala
 595 600 605

Leu Leu Ile Pro Ile Glu Ser Thr Ile Ala Glu Ala Met Arg Val Ile
 610 615 620

Met Val Asp Asp Ile Val Leu Ser Arg Lys Asn Val Val Asn Gln Leu
 625 630 635 640

Ala Asp Lys Leu Lys Asp Asp Trp Leu Ala Val Tyr Gln Gln Cys Asn
 645 650 655

Asp Asn Lys Pro Tyr Ser Leu Ser Ala Glu Gln Ile Ala Lys Arg Lys
 660 665 670

Leu Lys Gly Val Cys Leu Ser Tyr Leu Met Asn Ala Ser Asp Gln Lys
 675 680 685

Val Gly Thr Asp Leu Ala Gln Gln Leu Phe Asp Asn Ala Asp Asn Met
 690 695 700

Thr Asp Gln Gln Thr Ala Phe Thr Glu Leu Leu Lys Ser Asn Asp Lys
 705 710 715 720

Gln Val Arg Asp Asn Ala Ile Asn Glu Phe Tyr Asn Arg Trp Arg His
 725 730 735

Glu Asp Leu Val Val Asn Lys Trp Leu Leu Ser Gln Ala Gln Ile Ser
 740 745 750

His Glu Ser Ala Leu Asp Ile Val Lys Gly Leu Val Asn His Pro Ala
 755 760 765

Tyr Asn Pro Lys Asn Pro Asn Lys Val Tyr Ser Leu Ile Gly Gly Phe
 770 775 780

Gly Ala Asn Phe Leu Gln Tyr His Cys Lys Asp Gly Leu Gly Tyr Ala
 785 790 795 800

Phe Met Ala Asp Thr Val Leu Ala Leu Asp Lys Phe Asn His Gln Val
 805 810 815

Ala Ala Arg Met Ala Arg Asn Leu Met Ser Trp Lys Arg Tyr Asp Ser
 820 825 830

Asp Arg Gln Ala Met Met Lys Asn Ala Leu Glu Lys Ile Lys Ala Ser
 835 840 845

Asn Pro Ser Lys Asn Val Phe Glu Ile Val Ser Lys Ser Leu Glu Ser
 850 855 860

<210> 16
<211> 366
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 16

Met Gly Ser Ser His His His His His Ser Ser Gly Met Glu Val
1 5 10 15

Arg Asn Met Val Asp Tyr Glu Leu Leu Lys Lys Val Val Glu Ala Pro
20 25 30

Gly Val Ser Gly Tyr Glu Phe Leu Gly Ile Arg Asp Val Val Ile Glu
35 40 45

Glu Ile Lys Asp Tyr Val Asp Glu Val Lys Val Asp Lys Leu Gly Asn
50 55 60

Val Ile Ala His Lys Lys Gly Glu Gly Pro Lys Val Met Ile Ala Ala
65 70 75 80

His Met Asp Gln Ile Gly Leu Met Val Thr His Ile Glu Lys Asn Gly
85 90 95

Phe Leu Arg Val Ala Pro Ile Gly Gly Val Asp Pro Lys Thr Leu Ile
100 105 110

Ala Gln Arg Phe Lys Val Trp Ile Asp Lys Gly Lys Phe Ile Tyr Gly
115 120 125

Val Gly Ala Ser Val Pro Pro His Ile Gln Lys Pro Glu Asp Arg Lys
130 135 140

Lys Ala Pro Asp Trp Asp Gln Ile Phe Ile Asp Ile Gly Ala Glu Ser
145 150 155 160

Lys Glu Glu Ala Glu Asp Met Gly Val Lys Ile Gly Thr Val Ile Thr
165 170 175

Trp Asp Gly Arg Leu Glu Arg Leu Gly Lys His Arg Phe Val Ser Ile
180 185 190

Ala Phe Asp Asp Arg Ile Ala Val Tyr Thr Ile Leu Glu Val Ala Lys
195 200 205

Gln Leu Lys Asp Ala Lys Ala Asp Val Tyr Phe Val Ala Thr Val Gln
210 215 220

Glu Glu Val Gly Leu Arg Gly Ala Arg Thr Ser Ala Phe Gly Ile Glu

225 230 235 240

Pro Asp Tyr Gly Phe Ala Ile Asp Val Thr Ile Ala Ala Asp Ile Pro
245 250 255

Gly Thr Pro Glu His Lys Gln Val Thr His Leu Gly Lys Gly Thr Ala
260 265 270

Ile Lys Ile Met Asp Arg Ser Val Ile Cys His Pro Thr Ile Val Arg
275 280 285

Trp Leu Glu Glu Leu Ala Lys Lys His Glu Ile Pro Tyr Gln Leu Glu
290 295 300

Ile Leu Leu Gly Gly Gly Thr Asp Ala Gly Ala Ile His Leu Thr Lys
305 310 315 320

Ala Gly Val Pro Thr Gly Ala Leu Ser Val Pro Ala Arg Tyr Ile His
325 330 335

Ser Asn Thr Glu Val Val Asp Glu Arg Asp Val Asp Ala Thr Val Glu
340 345 350

Leu Met Thr Lys Ala Leu Glu Asn Ile His Glu Leu Lys Ile
355 360 365

<210> 17
<211> 408
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 17

Met Asp Ala Phe Thr Glu Asn Leu Asn Lys Leu Ala Glu Leu Ala Ile
1 5 10 15

Arg Val Gly Leu Asn Leu Glu Glu Gly Gln Glu Ile Val Ala Thr Ala
20 25 30

Pro Ile Glu Ala Val Asp Phe Val Arg Leu Leu Ala Glu Lys Ala Tyr
35 40 45

Glu Asn Gly Ala Ser Leu Phe Thr Val Leu Tyr Gly Asp Asn Leu Ile
50 55 60

Ala Arg Lys Arg Leu Ala Leu Val Pro Glu Ala His Leu Asp Arg Ala
65 70 75 80

Pro Ala Trp Leu Tyr Glu Gly Met Ala Lys Ala Phe His Glu Gly Ala
85 90 95

Ala Arg Leu Ala Val Ser Gly Asn Asp Pro Lys Ala Leu Glu Gly Leu
100 105 110

Pro Pro Glu Arg Val Gly Arg Ala Gln Gln Ala Gln Ser Arg Ala Tyr
115 120 125

Arg Pro Thr Leu Ser Ala Ile Thr Glu Phe Val Thr Asn Trp Thr Ile
130 135 140

Val Pro Phe Ala His Pro Gly Trp Ala Lys Ala Val Phe Pro Gly Leu
145 150 155 160

Pro Glu Glu Glu Ala Val Gln Arg Leu Trp Gln Ala Ile Phe Gln Ala
165 170 175

Thr Arg Val Asp Gln Glu Asp Pro Val Ala Ala Trp Glu Ala His Asn
180 185 190

Arg Val Leu His Ala Lys Val Ala Phe Leu Asn Glu Lys Arg Phe His
195 200 205

Ala Leu His Phe Gln Gly Pro Gly Thr Asp Leu Thr Val Gly Leu Ala
210 215 220

Glu Gly His Leu Trp Gln Gly Gly Ala Thr Pro Thr Lys Lys Gly Arg
225 230 235 240

Leu Cys Asn Pro Asn Leu Pro Thr Glu Glu Val Phe Thr Ala Pro His
245 250 255

Arg Glu Arg Val Glu Gly Val Val Arg Ala Ser Arg Pro Leu Ala Leu
260 265 270

Ser Gly Gln Leu Val Glu Gly Leu Trp Ala Arg Phe Glu Gly Gly Val
275 280 285

Ala Val Glu Val Gly Ala Glu Lys Gly Glu Glu Val Leu Lys Lys Leu
290 295 300

Leu Asp Thr Asp Glu Gly Ala Arg Arg Leu Gly Glu Val Ala Leu Val
305 310 315 320

Pro Ala Asp Asn Pro Ile Ala Lys Thr Gly Leu Val Phe Phe Asp Thr
325 330 335

Leu Phe Asp Glu Asn Ala Ala Ser His Ile Ala Phe Gly Gln Ala Tyr
340 345 350

Ala Glu Asn Leu Glu Gly Arg Pro Ser Gly Glu Glu Phe Arg Arg Arg
355 360 365

Gly Gly Asn Glu Ser Met Val His Val Asp Trp Met Ile Gly Ser Glu

370

375

380

Glu Val Asp Val Asp Gly Leu Leu Glu Asp Gly Thr Arg Val Pro Leu
 385 390 395 400

Met Arg Arg Gly Arg Trp Val Ile
 405

<210> 18

<211> 362

<212> PRT

<213> Artificial Sequence

<220>

<223> Synthetic

<400> 18

Met Ala Lys Leu Asp Glu Thr Leu Thr Met Leu Lys Ala Leu Thr Asp
 1 5 10 15

Ala Lys Gly Val Pro Gly Asn Glu Arg Glu Ala Arg Asp Val Met Lys
 20 25 30

Thr Tyr Ile Ala Pro Tyr Ala Asp Glu Val Thr Thr Asp Gly Leu Gly
 35 40 45

Ser Leu Ile Ala Lys Lys Glu Gly Lys Ser Gly Gly Pro Lys Val Met
 50 55 60

Ile Ala Gly His Leu Asp Glu Val Gly Phe Met Val Thr Gln Ile Asp
 65 70 75 80

Asp Lys Gly Phe Ile Arg Phe Gln Thr Leu Gly Gly Trp Trp Ser Gln
 85 90 95

Val Met Leu Ala Gln Arg Val Thr Ile Val Thr Lys Lys Gly Asp Ile
 100 105 110

Thr Gly Val Ile Gly Ser Lys Pro Pro His Ile Leu Pro Ser Glu Ala
 115 120 125

Arg Lys Lys Pro Val Glu Ile Lys Asp Met Phe Ile Asp Ile Gly Ala
 130 135 140

Thr Ser Arg Glu Glu Ala Met Glu Trp Gly Val Arg Pro Gly Asp Met
 145 150 155 160

Ile Val Pro Tyr Phe Glu Phe Thr Val Leu Asn Asn Glu Lys Met Leu
 165 170 175

Leu Ala Lys Ala Trp Asp Asn Arg Ile Gly Cys Ala Val Ala Ile Asp
 180 185 190

Val Leu Lys Gln Leu Lys Gly Val Asp His Pro Asn Thr Val Tyr Gly
195 200 205

Val Gly Thr Val Gln Glu Glu Val Gly Leu Arg Gly Ala Arg Thr Ala
210 215 220

Ala Gln Phe Ile Gln Pro Asp Ile Ala Phe Ala Val Asp Val Gly Ile
225 230 235 240

Ala Gly Asp Thr Pro Gly Val Ser Glu Lys Glu Ala Met Gly Lys Leu
245 250 255

Gly Ala Gly Pro His Ile Val Leu Tyr Asp Ala Thr Met Val Ser His
260 265 270

Arg Gly Leu Arg Glu Phe Val Ile Glu Val Ala Glu Glu Leu Asn Ile
275 280 285

Pro His His Phe Asp Ala Met Pro Gly Val Gly Thr Asp Ala Gly Ala
290 295 300

Ile His Leu Thr Gly Ile Gly Val Pro Ser Leu Thr Ile Ala Ile Pro
305 310 315 320

Thr Arg Tyr Ile His Ser His Ala Ala Ile Leu His Arg Asp Asp Tyr
325 330 335

Glu Asn Thr Val Lys Leu Leu Val Glu Val Ile Lys Arg Leu Asp Ala
340 345 350

Asp Lys Val Lys Gln Leu Thr Phe Asp Glu
355 360

<210> 19
<211> 490
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 19

Met Glu Asp Lys Val Trp Ile Ser Met Gly Ala Asp Ala Val Gly Ser
1 5 10 15

Leu Asn Pro Ala Leu Ser Glu Ser Leu Leu Pro His Ser Phe Ala Ser
20 25 30

Gly Ser Gln Val Trp Ile Gly Glu Val Ala Ile Asp Glu Leu Ala Glu
35 40 45

Leu Ser His Thr Met His Glu Gln His Asn Arg Cys Gly Gly Tyr Met
50 55 60

Val His Thr Ser Ala Gln Gly Ala Met Ala Ala Leu Met Met Pro Glu
65 70 75 80

Ser Ile Ala Asn Phe Thr Ile Pro Ala Pro Ser Gln Gln Asp Leu Val
85 90 95

Asn Ala Trp Leu Pro Gln Val Ser Ala Asp Gln Ile Thr Asn Thr Ile
100 105 110

Arg Ala Leu Ser Ser Phe Asn Asn Arg Phe Tyr Thr Thr Thr Ser Gly
115 120 125

Ala Gln Ala Ser Asp Trp Leu Ala Asn Glu Trp Arg Ser Leu Ile Ser
130 135 140

Ser Leu Pro Gly Ser Arg Ile Glu Gln Ile Lys His Ser Gly Tyr Asn
145 150 155 160

Gln Lys Ser Val Val Leu Thr Ile Gln Gly Ser Glu Lys Pro Asp Glu
165 170 175

Trp Val Ile Val Gly Gly His Leu Asp Ser Thr Leu Gly Ser His Thr
180 185 190

Asn Glu Gln Ser Ile Ala Pro Gly Ala Asp Asp Asp Ala Ser Gly Ile
195 200 205

Ala Ser Leu Ser Glu Ile Ile Arg Val Leu Arg Asp Asn Asn Phe Arg
210 215 220

Pro Lys Arg Ser Val Ala Leu Met Ala Tyr Ala Ala Glu Glu Val Gly
225 230 235 240

Leu Arg Gly Ser Gln Asp Leu Ala Asn Gln Tyr Lys Ala Gln Gly Lys
245 250 255

Lys Val Val Ser Val Leu Gln Leu Asp Met Thr Asn Tyr Arg Gly Ser
260 265 270

Ala Glu Asp Ile Val Phe Ile Thr Asp Tyr Thr Asp Ser Asn Leu Thr
275 280 285

Gln Phe Leu Thr Thr Leu Ile Asp Glu Tyr Leu Pro Glu Leu Thr Tyr
290 295 300

Gly Tyr Asp Arg Cys Gly Tyr Ala Cys Ser Asp His Ala Ser Trp His
305 310 315 320

Lys Ala Gly Phe Ser Ala Ala Met Pro Phe Glu Ser Lys Phe Lys Asp
325 330 335

Tyr Asn Pro Lys Ile His Thr Ser Gln Asp Thr Leu Ala Asn Ser Asp
340 345 350

Pro Thr Gly Asn His Ala Val Lys Phe Thr Lys Leu Gly Leu Ala Tyr
355 360 365

Val Ile Glu Met Ala Asn Ala Gly Ser Ser Gln Val Pro Asp Asp Ser
370 375 380

Val Leu Gln Asp Gly Thr Ala Lys Ile Asn Leu Ser Gly Ala Arg Gly
385 390 395 400

Thr Gln Lys Arg Phe Thr Phe Glu Leu Ser Gln Ser Lys Pro Leu Thr
405 410 415

Ile Gln Thr Tyr Gly Gly Ser Gly Asp Val Asp Leu Tyr Val Lys Tyr
420 425 430

Gly Ser Ala Pro Ser Lys Ser Asn Trp Asp Cys Arg Pro Tyr Gln Asn
435 440 445

Gly Asn Arg Glu Thr Cys Ser Phe Asn Asn Ala Gln Pro Gly Ile Tyr
450 455 460

His Val Met Leu Asp Gly Tyr Thr Asn Tyr Asn Asp Val Ala Leu Lys
465 470 475 480

Ala Ser Thr Gln His His His His His His
485 490

<210> 20
<211> 494
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 20

Met Glu Asp Lys Val Trp Ile Ser Ile Gly Ser Asp Ala Ser Gln Thr
1 5 10 15

Val Lys Ser Val Met Gln Ser Asn Ala Arg Ser Leu Leu Pro Glu Ser
20 25 30

Leu Ala Ser Asn Gly Pro Val Trp Val Gly Gln Val Asp Tyr Ser Gln
35 40 45

Leu Ala Glu Leu Ser His His Met His Glu Asp His Gln Arg Cys Gly
50 55 60

Gly Tyr Met Val His Ser Ser Pro Glu Ser Ala Ile Ala Ala Ser Asn
65 70 75 80

Met Pro Gln Ser Leu Val Ala Phe Ser Ile Pro Glu Ile Ser Gln Gln
 85 90 95

Asp Thr Val Asn Ala Trp Leu Pro Gln Val Asn Ser Gln Ala Ile Thr
 100 105 110

Gly Thr Ile Thr Ser Leu Thr Ser Phe Ile Asn Arg Phe Tyr Thr Thr
 115 120 125

Thr Ser Gly Ala Gln Ala Ser Asp Trp Leu Ala Asn Glu Trp Arg Ser
 130 135 140

Leu Ser Ala Ser Leu Pro Asn Ala Ser Val Arg Gln Val Ser His Phe
 145 150 155 160

Gly Tyr Asn Gln Lys Ser Val Val Leu Thr Ile Thr Gly Ser Glu Lys
 165 170 175

Pro Asp Glu Trp Ile Val Leu Gly Gly His Leu Asp Ser Thr Ile Gly
 180 185 190

Ser His Thr Asn Glu Gln Ser Val Ala Pro Gly Ala Asp Asp Asp Ala
 195 200 205

Ser Gly Ile Ala Ser Val Thr Glu Ile Ile Arg Val Leu Ser Glu Asn
 210 215 220

Asn Phe Gln Pro Lys Arg Ser Ile Ala Phe Met Ala Tyr Ala Ala Glu
 225 230 235 240

Glu Val Gly Leu Arg Gly Ser Gln Asp Leu Ala Asn Gln Tyr Lys Ala
 245 250 255

Glu Gly Lys Gln Val Ile Ser Ala Leu Gln Leu Asp Met Thr Asn Tyr
 260 265 270

Lys Gly Ser Val Glu Asp Ile Val Phe Ile Thr Asp Tyr Thr Asp Ser
 275 280 285

Asn Leu Thr Thr Phe Leu Ser Gln Leu Val Asp Glu Tyr Leu Pro Ser
 290 295 300

Leu Thr Tyr Gly Phe Asp Thr Cys Gly Tyr Ala Cys Ser Asp His Ala
 305 310 315 320

Ser Trp His Lys Ala Gly Phe Ser Ala Ala Met Pro Phe Glu Ala Lys
 325 330 335

Phe Asn Asp Tyr Asn Pro Met Ile His Thr Pro Asn Asp Thr Leu Gln
 340 345 350

Asn Ser Asp Pro Thr Ala Ser His Ala Val Lys Phe Thr Lys Leu Gly
355 360 365

Leu Ala Tyr Ala Ile Glu Met Ala Ser Thr Thr Gly Gly Thr Pro Pro
370 375 380

Pro Thr Gly Asn Val Leu Lys Asp Gly Val Pro Val Asn Gly Leu Ser
385 390 395 400

Gly Ala Thr Gly Ser Gln Val His Tyr Ser Phe Glu Leu Pro Ala Gln
405 410 415

Lys Asn Leu Gln Ile Ser Thr Ala Gly Gly Ser Gly Asp Val Asp Leu
420 425 430

Tyr Val Ser Phe Gly Ser Glu Ala Thr Lys Gln Asn Trp Asp Cys Arg
435 440 445

Pro Tyr Arg Asn Gly Asn Asn Glu Val Cys Thr Phe Ala Gly Ala Thr
450 455 460

Pro Gly Thr Tyr Ser Ile Met Leu Asp Gly Tyr Arg Gln Phe Ser Gly
465 470 475 480

Val Thr Leu Lys Ala Ser Thr Gln His His His His His His
485 490

<210> 21
<211> 877
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 21

Met Thr Gln Gln Pro Gln Ala Lys Tyr Arg His Asp Tyr Arg Ala Pro
1 5 10 15

Asp Tyr Thr Ile Thr Asp Ile Asp Leu Asp Phe Ala Leu Asp Ala Gln
20 25 30

Lys Thr Thr Val Thr Ala Val Ser Lys Val Lys Arg Gln Gly Thr Asp
35 40 45

Val Thr Pro Leu Ile Leu Asn Gly Glu Asp Leu Thr Leu Ile Ser Val
50 55 60

Ser Val Asp Gly Gln Ala Trp Pro His Tyr Arg Gln Gln Asp Asn Thr
65 70 75 80

Leu Val Ile Glu Gln Leu Pro Ala Asp Phe Thr Leu Thr Ile Val Asn
85 90 95

Asp Ile His Pro Ala Thr Asn Ser Ala Leu Glu Gly Leu Tyr Leu Ser
100 105 110

Gly Glu Ala Leu Cys Thr Gln Cys Glu Ala Glu Gly Phe Arg His Ile
115 120 125

Thr Tyr Tyr Leu Asp Arg Pro Asp Val Leu Ala Arg Phe Thr Thr Arg
130 135 140

Ile Val Ala Asp Lys Ser Arg Tyr Pro Tyr Leu Leu Ser Asn Gly Asn
145 150 155 160

Arg Val Gly Gln Gly Glu Leu Asp Asp Gly Arg His Trp Val Lys Trp
165 170 175

Glu Asp Pro Phe Pro Lys Pro Ser Tyr Leu Phe Ala Leu Val Ala Gly
180 185 190

Asp Phe Asp Val Leu Gln Asp Lys Phe Ile Thr Arg Ser Gly Arg Glu
195 200 205

Val Ala Leu Glu Ile Phe Val Asp Arg Gly Asn Leu Asp Arg Ala Asp
210 215 220

Trp Ala Met Thr Ser Leu Lys Asn Ser Met Lys Trp Asp Glu Thr Arg
225 230 235 240

Phe Gly Leu Glu Tyr Asp Leu Asp Ile Tyr Met Ile Val Ala Val Asp
245 250 255

Phe Phe Asn Met Gly Ala Met Glu Asn Lys Gly Leu Asn Val Phe Asn
260 265 270

Ser Lys Tyr Val Leu Ala Lys Ala Glu Thr Ala Thr Asp Lys Asp Tyr
275 280 285

Leu Asn Ile Glu Ala Val Ile Gly His Glu Tyr Phe His Asn Trp Thr
290 295 300

Gly Asn Arg Val Thr Cys Arg Asp Trp Phe Gln Leu Ser Leu Lys Glu
305 310 315 320

Gly Leu Thr Val Phe Arg Asp Gln Glu Phe Ser Ser Asp Leu Gly Ser
325 330 335

Arg Ser Val Asn Arg Ile Glu Asn Val Arg Val Met Arg Ala Ala Gln
340 345 350

Phe Ala Glu Asp Ala Ser Pro Met Ala His Ala Ile Arg Pro Asp Lys
355 360 365

Val Ile Glu Met Asn Asn Phe Tyr Thr Leu Thr Val Tyr Glu Lys Gly
370 375 380

Ser Glu Val Ile Arg Met Met His Thr Leu Leu Gly Glu Gln Gln Phe
385 390 395 400

Gln Ala Gly Met Arg Leu Tyr Phe Glu Arg His Asp Gly Ser Ala Ala
405 410 415

Thr Cys Asp Asp Phe Val Gln Ala Met Glu Asp Val Ser Asn Val Asp
420 425 430

Leu Ser Leu Phe Arg Arg Trp Tyr Ser Gln Ser Gly Thr Pro Leu Leu
435 440 445

Thr Val His Asp Asp Tyr Asp Val Glu Lys Gln Gln Tyr His Leu Phe
450 455 460

Val Ser Gln Lys Thr Leu Pro Thr Ala Asp Gln Pro Glu Lys Leu Pro
465 470 475 480

Leu His Ile Pro Leu Asp Ile Glu Leu Tyr Asp Ser Lys Gly Asn Val
485 490 495

Ile Pro Leu Gln His Asn Gly Leu Pro Val His His Val Leu Asn Val
500 505 510

Thr Glu Ala Glu Gln Thr Phe Thr Phe Asp Asn Val Ala Gln Lys Pro
515 520 525

Ile Pro Ser Leu Leu Arg Glu Phe Ser Ala Pro Val Lys Leu Asp Tyr
530 535 540

Pro Tyr Ser Asp Gln Gln Leu Thr Phe Leu Met Gln His Ala Arg Asn
545 550 555 560

Glu Phe Ser Arg Trp Asp Ala Ala Gln Ser Leu Leu Ala Thr Tyr Ile
565 570 575

Lys Leu Asn Val Ala Lys Tyr Gln Gln Gln Gln Pro Leu Ser Leu Pro
580 585 590

Ala His Val Ala Asp Ala Phe Arg Ala Ile Leu Leu Asp Glu His Leu
595 600 605

Asp Pro Ala Leu Ala Ala Gln Ile Leu Thr Leu Pro Ser Glu Asn Glu
610 615 620

Met Ala Glu Leu Phe Thr Thr Ile Asp Pro Gln Ala Ile Ser Thr Val
625 630 635 640

His Glu Ala Ile Thr Arg Cys Leu Ala Gln Glu Leu Ser Asp Glu Leu

655

<400> 22

Met Glu Glu Lys Val Trp Ile Ser Ile Gly Gly Asp Ala Thr Gln Thr
1 5 10 15

Ala Leu Arg Ser Gly Ala Gln Ser Leu Leu Pro Glu Asn Leu Ile Asn
20 25 30

Gln Thr Ser Val Trp Val Gly Gln Val Pro Val Ser Glu Leu Ala Thr
35 40 45

Leu Ser His Glu Met His Glu Asn His Gln Arg Cys Gly Gly Tyr Met
50 55 60

Val His Pro Ser Ala Gln Ser Ala Met Ser Val Ser Ala Met Pro Leu
65 70 75 80

Asn Leu Asn Ala Phe Ser Ala Pro Glu Ile Thr Gln Gln Thr Thr Val
85 90 95

Asn Ala Trp Leu Pro Ser Val Ser Ala Gln Gln Ile Thr Ser Thr Ile
100 105 110

Thr Thr Leu Thr Gln Phe Lys Asn Arg Phe Tyr Thr Thr Ser Thr Gly
115 120 125

Ala Gln Ala Ser Asn Trp Ile Ala Asp His Trp Arg Ser Leu Ser Ala
130 135 140

Ser Leu Pro Ala Ser Lys Val Glu Gln Ile Thr His Ser Gly Tyr Asn
145 150 155 160

Gln Lys Ser Val Met Leu Thr Ile Thr Gly Ser Glu Lys Pro Asp Glu
165 170 175

Trp Val Val Ile Gly Gly His Leu Asp Ser Thr Leu Gly Ser Arg Thr
180 185 190

Asn Glu Ser Ser Ile Ala Pro Gly Ala Asp Asp Ala Ser Gly Ile
195 200 205

Ala Gly Val Thr Glu Ile Ile Arg Leu Leu Ser Glu Gln Asn Phe Arg
210 215 220

Pro Lys Arg Ser Ile Ala Phe Met Ala Tyr Ala Ala Glu Glu Val Gly
225 230 235 240

Leu Arg Gly Ser Gln Asp Leu Ala Asn Arg Phe Lys Ala Glu Gly Lys
245 250 255

Lys Val Met Ser Val Met Gln Leu Asp Met Thr Asn Tyr Gln Gly Ser
260 265 270

Arg Glu Asp Ile Val Phe Ile Thr Asp Tyr Thr Asp Ser Asn Phe Thr

275

280

285

Gln Tyr Leu Thr Gln Leu Leu Asp Glu Tyr Leu Pro Ser Leu Thr Tyr
 290 295 300

Gly Phe Asp Thr Cys Gly Tyr Ala Cys Ser Asp His Ala Ser Trp His
 305 310 315 320

Ala Val Gly Tyr Pro Ala Ala Met Pro Phe Glu Ser Lys Phe Asn Asp
 325 330 335

Tyr Asn Pro Asn Ile His Ser Pro Gln Asp Thr Leu Gln Asn Ser Asp
 340 345 350

Pro Thr Gly Phe His Ala Val Lys Phe Thr Lys Leu Gly Leu Ala Tyr
 355 360 365

Val Val Glu Met Gly Asn Ala Ser Thr Pro Pro Thr Pro Ser Asn Gln
 370 375 380

Leu Lys Asn Gly Val Pro Val Asn Gly Leu Ser Ala Ser Arg Asn Ser
 385 390 395 400

Lys Thr Trp Tyr Gln Phe Glu Leu Gln Glu Ala Gly Asn Leu Ser Ile
 405 410 415

Val Leu Ser Gly Gly Ser Gly Asp Ala Asp Leu Tyr Val Lys Tyr Gln
 420 425 430

Thr Asp Ala Asp Leu Gln Gln Tyr Asp Cys Arg Pro Tyr Arg Ser Gly
 435 440 445

Asn Asn Glu Thr Cys Gln Phe Ser Asn Ala Gln Pro Gly Arg Tyr Ser
 450 455 460

Ile Leu Leu His Gly Tyr Asn Asn Tyr Ser Asn Ala Ser Leu Val Ala
 465 470 475 480

Asn Ala Gln His His His His His
 485

<210> 23
 <211> 488
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Synthetic

<400> 23

Met Glu Asp Lys Lys Val Trp Ile Ser Ile Gly Ala Asp Ala Gln Gln
 1 5 10 15

Thr Ala Leu Ser Ser Gly Ala Gln Pro Leu Leu Ala Gln Ser Val Ala
20 25 30

His Asn Gly Gln Ala Trp Ile Gly Glu Val Ser Glu Ser Glu Leu Ala
35 40 45

Ala Leu Ser His Glu Met His Glu Asn His His Arg Cys Gly Gly Tyr
50 55 60

Ile Val His Ser Ser Ala Gln Ser Ala Met Ala Ala Ser Asn Met Pro
65 70 75 80

Leu Ser Arg Ala Ser Phe Ile Ala Pro Ala Ile Ser Gln Gln Ala Leu
85 90 95

Val Thr Pro Trp Ile Ser Gln Ile Asp Ser Ala Leu Ile Val Asn Thr
100 105 110

Ile Asp Arg Leu Thr Asp Phe Pro Asn Arg Phe Tyr Thr Thr Thr Ser
115 120 125

Gly Ala Gln Ala Ser Asp Trp Ile Lys Gln Arg Trp Gln Ser Leu Ser
130 135 140

Ala Gly Leu Ala Gly Ala Ser Val Thr Gln Ile Ser His Ser Gly Tyr
145 150 155 160

Asn Gln Ala Ser Val Met Leu Thr Ile Glu Gly Ser Glu Ser Pro Asp
165 170 175

Glu Trp Val Val Val Gly Gly His Leu Asp Ser Thr Ile Gly Ser Arg
180 185 190

Thr Asn Glu Gln Ser Ile Ala Pro Gly Ala Asp Asp Asp Ala Ser Gly
195 200 205

Ile Ala Ala Val Thr Glu Val Ile Arg Val Leu Ala Gln Asn Asn Phe
210 215 220

Gln Pro Lys Arg Ser Ile Ala Phe Val Ala Tyr Ala Ala Glu Glu Val
225 230 235 240

Gly Leu Arg Gly Ser Gln Asp Val Ala Asn Gln Phe Lys Gln Ala Gly
245 250 255

Lys Asp Val Arg Gly Val Leu Gln Leu Asp Met Thr Asn Tyr Gln Gly
260 265 270

Ser Ala Glu Asp Ile Val Phe Ile Thr Asp Tyr Thr Asp Asn Gln Leu
275 280 285

Thr Gln Tyr Leu Thr Gln Leu Leu Asp Glu Tyr Leu Pro Thr Leu Asn

290

295

300

Tyr Gly Phe Asp Thr Cys Gly Tyr Ala Cys Ser Asp His Ala Ser Trp
305 310 315 320

His Gln Val Gly Tyr Pro Ala Ala Met Pro Phe Glu Ala Lys Phe Asn
325 330 335

Asp Tyr Asn Pro Asn Ile His Thr Pro Gln Asp Thr Leu Ala Asn Ser
340 345 350

Asp Ser Glu Gly Ala His Ala Ala Lys Phe Thr Lys Leu Gly Leu Ala
355 360 365

Tyr Thr Val Glu Leu Ala Asn Ala Asp Ser Ser Pro Asn Pro Gly Asn
370 375 380

Glu Leu Lys Leu Gly Glu Pro Ile Asn Gly Leu Ser Gly Ala Arg Gly
385 390 395 400

Asn Glu Lys Tyr Phe Asn Tyr Arg Leu Asp Gln Ser Gly Glu Leu Val
405 410 415

Ile Arg Thr Tyr Gly Gly Ser Gly Asp Val Asp Leu Tyr Val Lys Ala
420 425 430

Asn Gly Asp Val Ser Thr Gly Asn Trp Asp Cys Arg Pro Tyr Arg Ser
435 440 445

Gly Asn Asp Glu Val Cys Arg Phe Asp Asn Ala Thr Pro Gly Asn Tyr
450 455 460

Ala Val Met Leu Arg Gly Tyr Arg Thr Tyr Asp Asn Val Ser Leu Ile
465 470 475 480

Val Glu His His His His His
485

<210> 24
<211> 308
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 24

Gly Met Pro Pro Ile Thr Gln Gln Ala Thr Val Thr Ala Trp Leu Pro
1 5 10 15

Gln Val Asp Ala Ser Gln Ile Thr Gly Thr Ile Ser Ser Leu Glu Ser
20 25 30

Phe Thr Asn Arg Phe Tyr Thr Thr Thr Ser Gly Ala Gln Ala Ser Asp
35 40 45

Trp Ile Ala Ser Glu Trp Gln Phe Leu Ser Ala Ser Leu Pro Asn Ala
50 55 60

Ser Val Lys Gln Val Ser His Ser Gly Tyr Asn Gln Lys Ser Val Val
65 70 75 80

Met Thr Ile Thr Gly Ser Glu Ala Pro Asp Glu Trp Ile Val Ile Gly
85 90 95

Gly His Leu Asp Ser Thr Ile Gly Ser His Thr Asn Glu Gln Ser Val
100 105 110

Ala Pro Gly Ala Asp Asp Asp Ala Ser Gly Ile Ala Ala Val Thr Glu
115 120 125

Val Ile Arg Val Leu Ser Glu Asn Asn Phe Gln Pro Lys Arg Ser Ile
130 135 140

Ala Phe Met Ala Tyr Ala Ala Glu Glu Val Gly Leu Arg Gly Ser Gln
145 150 155 160

Asp Leu Ala Asn Gln Tyr Lys Ser Glu Gly Lys Asn Val Val Ser Ala
165 170 175

Leu Gln Leu Asp Met Thr Asn Tyr Lys Gly Ser Ala Gln Asp Val Val
180 185 190

Phe Ile Thr Asp Tyr Thr Asp Ser Asn Phe Thr Gln Tyr Leu Thr Gln
195 200 205

Leu Met Asp Glu Tyr Leu Pro Ser Leu Thr Tyr Gly Phe Asp Thr Cys
210 215 220

Gly Tyr Ala Cys Ser Asp His Ala Ser Trp His Asn Ala Gly Tyr Pro
225 230 235 240

Ala Ala Met Pro Phe Glu Ser Lys Phe Asn Asp Tyr Asn Pro Arg Ile
245 250 255

His Thr Thr Gln Asp Thr Leu Ala Asn Ser Asp Pro Thr Gly Ser His
260 265 270

Ala Lys Lys Phe Thr Gln Leu Gly Leu Ala Tyr Ala Ile Glu Met Gly
275 280 285

Ser Ala Thr Gly Asp Thr Pro Thr Pro Gly Asn Gln Leu Glu His His
290 295 300

His His His His

305

<210> 25
<211> 354
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 25

Met Val Asp Trp Glu Leu Met Lys Lys Ile Ile Glu Ser Pro Gly Val
1 5 10 15

Ser Gly Tyr Glu His Leu Gly Ile Arg Asp Leu Val Val Asp Ile Leu
20 25 30

Lys Asp Val Ala Asp Glu Val Lys Ile Asp Lys Leu Gly Asn Val Ile
35 40 45

Ala His Phe Lys Gly Ser Ala Pro Lys Val Met Val Ala Ala His Met
50 55 60

Asp Lys Ile Gly Leu Met Val Asn His Ile Asp Lys Asp Gly Tyr Leu
65 70 75 80

Arg Val Val Pro Ile Gly Gly Val Leu Pro Glu Thr Leu Ile Ala Gln
85 90 95

Lys Ile Arg Phe Phe Thr Glu Lys Gly Glu Arg Tyr Gly Val Val Gly
100 105 110

Val Leu Pro Pro His Leu Arg Arg Glu Ala Lys Asp Gln Gly Gly Lys
115 120 125

Ile Asp Trp Asp Ser Ile Ile Val Asp Val Gly Ala Ser Ser Arg Glu
130 135 140

Glu Ala Glu Glu Met Gly Phe Arg Ile Gly Thr Ile Gly Glu Phe Ala
145 150 155 160

Pro Asn Phe Thr Arg Leu Ser Glu His Arg Phe Ala Thr Pro Tyr Leu
165 170 175

Asp Asp Arg Ile Cys Leu Tyr Ala Met Ile Glu Ala Ala Arg Gln Leu
180 185 190

Gly Glu His Glu Ala Asp Ile Tyr Ile Val Ala Ser Val Gln Glu Glu
195 200 205

Ile Gly Leu Arg Gly Ala Arg Val Ala Ser Phe Ala Ile Asp Pro Glu
210 215 220

Val Gly Ile Ala Met Asp Val Thr Phe Ala Lys Gln Pro Asn Asp Lys
225 230 235 240

Gly Lys Ile Val Pro Glu Leu Gly Lys Gly Pro Val Met Asp Val Gly
245 250 255

Pro Asn Ile Asn Pro Lys Leu Arg Gln Phe Ala Asp Glu Val Ala Lys
260 265 270

Lys Tyr Glu Ile Pro Leu Gln Val Glu Pro Ser Pro Arg Pro Thr Gly
275 280 285

Thr Asp Ala Asn Val Met Gln Ile Asn Arg Glu Gly Val Ala Thr Ala
290 295 300

Val Leu Ser Ile Pro Ile Arg Tyr Met His Ser Gln Val Glu Leu Ala
305 310 315 320

Asp Ala Arg Asp Val Asp Asn Thr Ile Lys Leu Ala Lys Ala Leu Leu
325 330 335

Glu Glu Leu Lys Pro Met Asp Phe Thr Pro Leu Glu His His His His
340 345 350

His His

<210> 26
<211> 6
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 26

Asp Tyr Arg Ala Gly Pro
1 5

<210> 27
<211> 6
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic

<400> 27

Leu Phe Trp Val Met Cys
1 5

<210> 28
<211> 7
<212> PRT
<213> Artificial Sequence

<220>

<223> Synthetic

<400> 28

Arg Glu Pro Ile Leu Gln Asn

1 5

<210> 29

<211> 6

<212> PRT

<213> Artificial Sequence

<220>

<223> Synthetic

<400> 29

Ile Leu Ser Thr Glu Pro

1 5

<210> 30

<211> 6

<212> PRT

<213> Artificial Sequence

<220>

<223> Synthetic

<400> 30

Asp Ala Gly Met Cys Val

1 5

<210> 31

<211> 7

<212> PRT

<213> Artificial Sequence

<220>

<223> Synthetic

<400> 31

Ser Pro Ile Gln Arg Tyr Pro

1 5

<210> 32

<211> 6

<212> PRT

<213> Artificial Sequence

<220>

<223> Synthetic

<400> 32

Gln Trp Cys Val Arg Glu

1 5

<210> 33

<211> 6

<212> PRT

<213> Artificial Sequence

<220>

<223> Synthetic

<400> 33

Trp	Val	Asp	Tyr	Glu	Arg
1				5	