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(54) **CLEANING COMPOSITIONS INCLUDING ENZYME AND PLANT FIBER**

(57) Cleaning compositions that include a mannanase enzyme and water-insoluble plant fibers. Methods of making and using such cleaning compositions. Use of water-insoluble plant fibers.

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Description

REFERENCE TO A SEQUENCE LISTING

5 **[0001]** This application contains a Sequence Listing in computer readable form, which is incorporated herein by reference.

FIELD OF THE INVENTION

10 **[0002]** The present invention relates to cleaning compositions that include a mannanase enzyme and water-insoluble plant fibers. The present invention also relates to methods of making and using such cleaning compositions. The present invention also relates to the use of a mannanase enzyme and water-insoluble plant fibers.

BACKGROUND OF THE INVENTION

15 **[0003]** The laundry detergent formulator is constantly aiming to improve the performance of detergent compositions. Enzymes may be added to liquid detergent formulations in order to improve cleaning performance, but soils may remain on the targeted surface.

20 **[0004]** There is a need for improved cleaning compositions that provide improved soil removal benefits. There is also a need for effective deposition of benefit agents.

SUMMARY OF THE INVENTION

25 **[0005]** The present invention provides a cleaning composition comprising a mannanase enzyme comprising a polypeptide having mannan endo-1,4-beta-mannosidase activity (EC 3.2.1 .78) that catalyzes the hydrolysis of 1 ,4-3-D-mannosidic linkages in mannans, galactomannans and/or glucomannans, and/or optionally according to SEQ ID NO:1 herein, or a variant having at least 60% sequence identity to SEQ ID NO:1 or according to SEQ ID NO:2 herein, or a variant having at least 81% sequence identity to SEQ ID NO: 2; or according to SEQ ID NO:3 herein, or a variant having at least 75% sequence identity to SEQ ID NO: 3 or according to SEQ ID NO:4 herein, or a variant having at least 65% sequence identity to SEQ ID NO: 4 or according to SEQ ID NO:5 herein, or a variant having at least 75% sequence identity to SEQ ID NO: 5 and water-insoluble plant fibers. Preferably the composition comprises a benefit agent, most preferably a particulate benefit agent. Preferably the mannanase is a member of the glycoside hydrolase family 26.

30 **[0006]** The present invention also relates to a method of cleaning a surface, preferably a textile, where the method comprises mixing the cleaning composition described herein with water to form an aqueous liquor and contacting a surface, preferably a textile, with the aqueous liquor in a laundering step.

35 **[0007]** The present invention also relates to a use of a mannanase enzyme comprising a polypeptide having mannan endo-1,4-beta-mannosidase activity (EC 3.2.1 .78) that catalyzes the hydrolysis of 1 ,4-3-D-mannosidic linkages in mannans, galactomannans and/or glucomannans and water-insoluble plant fibers in a cleaning composition to enhance stain-removal and/or malodor-reducing benefits.

40 **[0008]** The invention also provides a method of enhancing deposition of a particulate benefit agent comprising contacting a surface, preferably a textile with an aqueous liquor comprising a composition comprising (i) a polypeptide having mannan endo-1,4-beta-mannosidase activity (EC 3.2.1 .78) that catalyzes the hydrolysis of 1 ,4-3-D-mannosidic linkages in mannans, galactomannans and/or glucomannans; (ii) a water-insoluble plant fiber; and (iii) a particulate benefit agent in a textile treatment step, preferably a laundering step, and optionally rinsing and drying the textile.

45 Preferably the mannanase is a member of the glycoside hydrolase family 26.

DETAILED DESCRIPTION OF THE INVENTION

50 **[0009]** The present invention relates to cleaning compositions, for example liquid cleaning compositions, comprising a specific mannanase enzyme and water-insoluble plant fiber. Without wishing to be bound by theory, it is believed that the water-insoluble plant fibers result in microabrasion of a target surface, such as a soiled fabric, thereby complementing the cleaning mechanism of the mannanase enzyme and enhancing the removal of the soil matrix. This effect may be particularly strong in instances of direct application of a neat liquid detergent onto the fabric surface, such as in a pretreatment process.

55 **[0010]** The components of the compositions and processes of the present invention are described in more detail below.

[0011] As used herein, the articles "a" and "an" when used in a claim, are understood to mean one or more of what is claimed or described. As used herein, the terms "include," "includes," and "including" are meant to be non-limiting. The compositions of the present invention can comprise, consist essentially of, or consist of, the components of the

present disclosure.

[0012] The terms "substantially free of" or "substantially free from" may be used herein. This means that the indicated material is at the very minimum not deliberately added to the composition to form part of it, or, preferably, is not present at analytically detectable levels. It is meant to include compositions whereby the indicated material is present only as an impurity in one of the other materials deliberately included. The indicated material may be present, if at all, at a level of less than 1%, or less than 0.1%, or less than 0.01%, or even 0%, by weight of the composition.

[0013] As used herein, "insoluble" means having a water solubility of less than 10% when 1g of the dry material is stirred in 100g of deionized water in a 250ml beaker for 15 minutes at 20°C using a magnetic stirrer set at 100rpm. The degree of solubility is calculated by comparing the mass of dry fiber before (m_i = 1g) and after (m_f) the solubility test as follows:

$$\% \text{ Solubility} = 100[1 - (m_f / m_i)]$$

[0014] Unless otherwise noted, all component or composition levels are in reference to the active portion of that component or composition, and are exclusive of impurities, for example, residual solvents or by-products, which may be present in commercially available sources of such components or compositions.

[0015] All temperatures herein are in degrees Celsius (°C) unless otherwise indicated. Unless otherwise specified, all measurements herein are conducted at 20°C and under the atmospheric pressure.

[0016] In all embodiments of the present disclosure, all percentages are by weight of the total composition, unless specifically stated otherwise. All ratios are weight ratios, unless specifically stated otherwise.

[0017] It should be understood that every maximum numerical limitation given throughout this specification includes every lower numerical limitation, as if such lower numerical limitations were expressly written herein. Every minimum numerical limitation given throughout this specification will include every higher numerical limitation, as if such higher numerical limitations were expressly written herein. Every numerical range given throughout this specification will include every narrower numerical range that falls within such broader numerical range, as if such narrower numerical ranges were all expressly written herein.

[0018] As used herein, the term "alkoxy" is intended to include C1-C8 alkoxy and C1-C8 alkoxy derivatives of polyols having repeating units such as butylene oxide, glycidol oxide, ethylene oxide or propylene oxide.

[0019] As used herein, unless otherwise specified, the terms "alkyl" and "alkyl capped" are intended to include C1-C18 alkyl groups, or even C1-C6 alkyl groups.

[0020] As used herein, unless otherwise specified, the term "aryl" is intended to include C3-12 aryl groups.

[0021] As used herein, unless otherwise specified, the term "arylalkyl" and "alkaryl" are equivalent and are each intended to include groups comprising an alkyl moiety bound to an aromatic moiety, typically having C1-C18 alkyl groups and, in one aspect, C1-C6 alkyl groups.

[0022] The terms "ethylene oxide," "propylene oxide" and "butylene oxide" may be shown herein by their typical designation of "EO," "PO" and "BO," respectively.

[0023] As used herein, the term "cleaning and/or treatment composition" includes, unless otherwise indicated, granular, powder, liquid, gel, paste, unit dose, bar form and/or flake type washing agents and/or fabric treatment compositions.

[0024] As used herein, "cellulosic substrates" are intended to include any substrate which comprises cellulose, either 100% by weight cellulose or at least 20% by weight, or at least 30 % by weight or at least 40 or at least 50 % by weight or even at least 60 % by weight cellulose. Cellulose may be found in wood, cotton, linen, jute, and hemp. Cellulosic substrates may be in the form of powders, fibers, pulp and articles formed from powders, fibers and pulp. Cellulosic fibers, include, without limitation, cotton, rayon (regenerated cellulose), acetate (cellulose acetate), triacetate (cellulose triacetate), and mixtures thereof. Typically cellulosic substrates comprise cotton. Articles formed from cellulosic fibers include textile articles such as fabrics. Articles formed from pulp include paper.

[0025] As used herein, the term "maximum extinction coefficient" is intended to describe the molar extinction coefficient at the wavelength of maximum absorption (also referred to herein as the maximum wavelength), in the range of 400 nanometers to 750 nanometers.

[0026] As used herein "average molecular weight" is reported as a weight average molecular weight, as determined by its molecular weight distribution; as a consequence of their manufacturing process, polymers disclosed herein may contain a distribution of repeating units in their polymeric moiety.

[0027] As used herein the term "variant" refers to a polypeptide that contains an amino acid sequence that differs from a wild type or reference sequence. A variant polypeptide can differ from the wild type or reference sequence due to a deletion, insertion, or substitution of a nucleotide(s) relative to said reference or wild type nucleotide sequence. The reference or wild type sequence can be a full-length native polypeptide sequence or any other fragment of a full-length polypeptide sequence. A polypeptide variant generally has at least about 70% amino acid sequence identity with the reference sequence, but may include at least 75% amino acid sequence identity within the reference sequence, at least

80% amino acid sequence identity within the reference sequence, at least 85% amino acid sequence identity with the reference sequence, at least 86% amino acid sequence identity with the reference sequence, at least 87% amino acid sequence identity with the reference sequence, at least 88% amino acid sequence identity with the reference sequence, at least 89% amino acid sequence identity with the reference sequence, at least 90% amino acid sequence identity with the reference sequence, at least 91% amino acid sequence identity with the reference sequence, at least 92% amino acid sequence identity with the reference sequence, at least 93% amino acid sequence identity with the reference sequence, at least 94% amino acid sequence identity with the reference sequence, at least 95% amino acid sequence identity with the reference sequence, at least 96% amino acid sequence identity with the reference sequence, at least 97% amino acid sequence identity with the reference sequence, at least 98% amino acid sequence identity with the reference sequence, at least 98.5% amino acid sequence identity with the reference sequence or at least 99% amino acid sequence identity with the reference sequence.

[0028] As used herein, the term "solid" includes granular, powder, bar and tablet product forms.

[0029] As used herein, the term "fluid" includes liquid, gel, paste, and gas product forms.

Cleaning Composition

[0030] The present disclosure relates to cleaning and/or treatment compositions. The cleaning composition may be selected from the group of light duty liquid detergents compositions, heavy duty liquid detergent compositions, solid, for example particulate/powder or "dry" cleaning compositions, hard surface cleaning compositions, detergent gels commonly used for laundry, bleaching compositions, laundry additives, fabric enhancer compositions, shampoos, body washes, other personal care compositions, and mixtures thereof. The cleaning composition may be a hard surface cleaning composition (such as a dishwashing composition) or a laundry composition (such as a heavy duty liquid or solid detergent composition).

[0031] The cleaning compositions may be in any suitable form. The composition can be selected from a liquid, solid, or combination thereof. As used herein, "liquid" includes free-flowing liquids, as well as pastes, gels, foams and mousses. Non-limiting examples of liquids include light duty and heavy duty liquid detergent compositions, fabric enhancers, detergent gels commonly used for laundry, bleach and laundry additives. Gases, e.g., suspended bubbles, or solids, e.g. particles, may be included within the liquids. A "solid" as used herein includes, but is not limited to, powders, agglomerates, and mixtures thereof. Non-limiting examples of solids include: granules, micro-capsules, beads, noodles, and pearlised balls. Solid compositions may provide a technical benefit including, but not limited to, through-the-wash benefits, pre-treatment benefits, and/or aesthetic effects.

[0032] The cleaning composition may be in the form of a unitized dose article, such as a tablet or in the form of a pouch. Such pouches typically include a water-soluble film, such as a polyvinyl alcohol water-soluble film, that at least partially encapsulates a composition. Suitable films are available from MonoSol, LLC (Indiana, USA). The composition can be encapsulated in a single or multi-compartment pouch. A multi-compartment pouch may have at least two, at least three, or at least four compartments. A multi-compartmented pouch may include compartments that are side-by-side and/or superposed. The composition contained in the pouch may be liquid, solid (such as powders), or combinations thereof. Preferably the composition of the invention is a liquid.

Mannanases

[0033] The composition comprises a mannanase enzyme. The term "mannanase" herein means a polypeptide having mannan endo-1,4- beta-mannosidase activity (EC 3.2.1.78) from the glycoside hydrolase family 26 that catalyzes the hydrolysis of 1,4-3-D-mannosidic linkages in mannans, galactomannans and glucomannans. Alternative names of mannan endo-1,4-beta-mannosidase are 1,4-3-D-mannan mannanohydrolase; endo-1,4-3-mannanase; endo- β -1,4-mannase; β -mannanase B; 3-1,4-mannan 4-mannanohydrolase; endo-3-mannanase; and β -D-mannanase. Preferred mannanases are members of the glycoside hydrolase family 26.

[0034] For purposes of the present disclosure, mannanase activity may be determined using the Reducing End Assay as described in the experimental section of WO 2015040159.

Suitable examples from class EC 3.2.1.78 are described in WO 2015040159, such as the mature polypeptide SEQ ID NO: 2 described therein.

[0035] Preferred mannanases are variants having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 81 %, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91 %, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99% or 100% sequence identity to the mature polypeptide SEQ ID NO: 1 from *Ascobolus stictoides*;

[0036] Preferred mannanases are variants having at least 81 %, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91 %, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99% or 100% sequence identity to the mature

polypeptide SEQ ID NO: 2 from

Chaetomium virescens.

- 5 **[0037]** Preferred mannanases are variants having at least 75%, at least 76%, at least 77%, at least 78%, at least 79%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91 %, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99% or 100% sequence identity to the mature polypeptide SEQ ID NO: 3 from *Preussia aemulans*.
- 10 **[0038]** Preferred mannanases are variants having at least at least 65%, at least 66%, at least 67%, at least 68%, at least 69%, at least 70%, at least 71%, at least 72%, at least 73%, at least 74%, at least 75%, at least 76%, at least 77%, at least 78%, at least 79%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91 %, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99% or 100% sequence identity to the mature polypeptide
- 15 SEQ ID NO: 4 from *Yunnania penicillata*.
- [0039]** Preferred mannanases are variants having at least at least 75%, at least 76%, at least 77%, at least 78%, at least 79%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91 %, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99% or 100% sequence identity to the mature polypeptide SEQ ID NO: 5 from
- 20 *Myrothecium roridum*. Preferably the mannanase is an isolated mannanase.
- [0040]** Preferably the mannanase enzyme is present in the cleaning compositions in an amount from 0.001 to 1 wt% based on active protein in the composition, or from 0.005 to 0.5 wt% or from 0.01 to 0.25 wt%. Preferably the mannanase enzyme is present in the laundering aqueous liquor in an amount of from 0.01ppm to 1000 ppm of the mannanase enzyme, or from 0.05 or from 0.1ppm to 750 or 500ppm. The mannanases or compositions comprising them may also
- 25 give rise to biofilm-disrupting effects.

Water-Insoluble Plant Fiber

- 30 **[0041]** The cleaning compositions of the present invention include water-soluble plant fiber. Without wishing to be bound by theory, it is believed that the insoluble fibers may microabrade a target surface, thereby synergistically facilitating the cleaning benefits provided by other components of the composition. Additionally, such fibers may be particularly useful in liquid compositions, as they can provide structuring benefits.
- [0042]** The cleaning compositions of the present invention may include from about 0.01% to about 5%, by weight of the cleaning composition, of water-insoluble fiber.
- 35 **[0043]** The water-insoluble plant fiber may be derived from any member of the plant kingdom, including trees, herbaceous plants and the fruits of these plants. Example sources of such fibers are wood, chicory root, sugar beet and citrus or other fruits such as apple. The fibers may be produced as a side-stream from the processing of such crops for other purposes, for example in sugar refining, inulin production and fruit juice production.
- [0044]** Examples of suitable materials include parenchymal cellulose compositions and activated citrus fibers.
- 40 **[0045]** Suitable parenchymal cellulose compositions include particulate cellulose material containing, by dry weight of the particulate cellulose material, at least 70% cellulose, less than 10% pectin and at least 3% hemicellulose, wherein the particulate material has a volume-weighted median major particle dimension within the range of 25-75 μm , preferably within the range of 35-65 μm , as measured by laser light diffractometry in accordance with the established protocol ISO13320 (2009). The particulate cellulose material of this invention may contain particles of specific structure, shape
- 45 and size. Typically the material contains particles having the form of platelets comprising parenchymal cellulose structures or networks. It is preferred that the size distribution of the particulate material falls within certain limits. When the distribution is measured with a laser light scattering particle size analyzer, such as the Malvern Mastersizer or another instrument of equal or better sensitivity, the diameter data is preferably reported as a volume distribution. Thus the reported median for a population of particles will be volume-weighted, with about one-half of the particles, on a volume basis, having
- 50 diameters less than the median diameter for the population. Typically, the median major dimension of the particles of the parenchymal cellulose composition is within the range of 25-75 μm . More preferably the median major dimension of the particles of the parenchymal cellulose composition is within the range of 35-65 μm . Typically at least about 90%, on a volume basis, of the particles has a diameter less than about 120 μm , more preferably less than 110 μm , more preferably less than 100 μm . Preferably, the particulate cellulose material has a volume-weighted median minor dimension
- 55 larger than 0.5 μm , preferably larger than 1 μm .
- [0046]** The parenchymal cellulose is characterized by the fact that the majority of the cellulose material is present in the form of particles that are distinct from the nanofibrillised cellulose described in the prior art in that the cellulose nanofibrils are not substantially unraveled. Preferably, less than 10%, or more preferably less than 1% or less than 0.1%

by dry weight of the cellulose within the composition is in the form of nanofibrillated cellulose. This is advantageous as nanofibrillated cellulose negatively affects the redispersability of the material, as indicated herein before. By 'nanofibrils' we refer to the fibrils making up the cellulose fibers, typically having a width in the nanometer range (e.g., less than 1 μm) and a length of up to 20 μm . The nomenclature used in the field over the past decades has been somewhat inconsistent in that the terms 'microfibril' and 'nanofibril' have been used to denote the same material. In the context of this invention, the two terms are deemed to be fully interchangeable.

[0047] In accordance with the invention, the plant parenchymal cellulose material has been treated, modified and/or some components may have been removed but the cellulose at no time has been broken down to individual microfibrils, thereby losing the structure of plant cell wall sections. As mentioned before, the cellulose material of this invention has a reduced pectin content, as compared to the parenchymal cell wall material from which it is derived. Removal of some of the pectin is believed to result in enhanced thermal stability. The term "pectin" as used herein refers to a class of plant cell-wall heterogeneous polysaccharides that can be extracted by treatment with acids and chelating agents. Typically, 70-80% of pectin is found as a linear chain of α -(1,4)-linked D-galacturonic acid monomers. The smaller RG-I fraction of pectin is comprised of alternating (1-4)-linked galacturonic acid and (1,2)-linked L-rhamnose, with substantial arabinogalactan branching emanating from the L-rhamnose residue. Other monosaccharides, such as D-fucose, D-xylose, apiose, aceric acid, Kdo, Dha, 2-O-methyl-D-fucose, and 2-O-methyl-D-xylose, are found either in the RG-II pectin fraction (<2%), or as minor constituents in the RG-I fraction. Proportions of each of the monosaccharides in relation to D-galacturonic acid vary depending on the individual plant and its micro-environment, the species, and time during the growth cycle. For the same reasons, the homogalacturonan and RG-I fractions can differ widely in their content of methyl esters on GalA residues, and the content of acetyl residue esters on the C-2 and C-3 positions of GalA and neutral sugars. It is preferred that the particulate cellulose material of the invention comprises less than 5 wt.% of pectin, by dry weight of the particulate cellulose material, more preferably less than 2.5 wt.%. The presence of at least some pectin in the cellulose material is nevertheless desired. Without wishing to be bound by any theory it is assumed that pectin plays a role in the electrostatic interactions between particles contained in the material and/or in supporting the network/structure of the cellulose. Hence, it is preferred that the particulate cellulose material contains at least 0.5 wt% of pectin by dry weight of the particulate cellulose material, more preferably at least 1 wt.%.

[0048] The composition of the present invention preferably comprises, based on the total composition weight, from 0.01 to 5 %, preferably 0.05 to 1 %, more preferably from 0.1 to 0.75 % of water insoluble plant derived fibers.

[0049] By water insoluble plant derived fibers it is meant herein cellulose micro or nano fibrils and micro or nano crystals. The plant fibers can be extracted from plants, fruits or wood. Water insoluble plant derived fibers sources may be selected from the group consisting of citrus peels, such as lemons, oranges and/or grapefruit; fruits, such as apples, bananas and/or pear; vegetables such as carrots, peas, potatoes and/or chicory; plants such as bamboo, jute, abaca, flax, cotton and/or sisal, cereals, and different wood sources such as spruces, eucalyptus and/or oak. Preferably, the cellulose fibers source may be selected from the group consisting of wood or plants, in particular, spruce, eucalyptus, jute and sisal. After water-insoluble fibers have been activated by high pressure homogenizer (from 80 to 350 bars), the mean hydrodynamic diameter of such fibers is preferably from 3 microns to 130 microns (as measured using the hydrodynamic method), more preferably from 5 microns to 110 microns, even more preferably from 10 to 100 microns and the average diameter is from 1 nm to 1 micron, preferably from 10 nm to 850 nm, even more preferably from 15 to 350 nm. Without wishing to be bound by any theory, acid hydrolysis of water-insoluble plant derived fibers, would lead to micro or nanocrystals having an average diameter from 1 nm to 100 nm and a length from 200 nm to 3 microns (as measured using the AFM method). Such materials are commercially available from American process under the tradename of Bioplus.

[0050] The content of cellulose will vary depending on the source and treatment applied for the extraction of the fibers, and will range from 15 to 100%, preferably above 30%, more preferably above 50%, and even more preferably above 80%.

[0051] Such cellulose fibers may comprise pectin, hemicellulose, proteins, lignin and other impurities inherent to the cellulose based material source such as ash, metals, salts and combinations thereof. The cellulose fibers are preferably non-ionic.

[0052] Suitable activated citrus fruits may be produced from lemons and limes. These fruits may be de-juiced to leave an insoluble plant cell wall material with some internally contained sugars and pectin. The 'spongy microstructure', known as albedo, may be used to make acidic, powdered citrus fiber. The structure is dried, sieved and then washed to increase the fiber content. Dried materials are typically large (with cell fragments greater than 100 microns), consisting of tightly bound/bonded fibrils. After milling a powdered citrus fiber material is obtained. This procedure leaves much of the natural cell wall intact whilst sugars are removed. The resultant swellable citrus fiber materials are typically used as food additives and are often employed for example in low fat mayonnaise.

[0053] A preferred type of powdered citrus fiber for detergent formulations and used in accordance with the present invention is available from Herbafood Ingredients GmbH under the tradename, Herbacel™ AQ+ type N citrus fiber and Citri-Fi 100FG from Fiberstar. This citrus fiber has a total (soluble and insoluble) fiber content of greater than 80% by weight and soluble fiber content of greater than 20% by weight. It is supplied as a fine dried powder with low colour and

has a water binding capacity of about 20 kg water per kg of powder.

[0054] The citrus fiber useful in the present invention is preferably activated citrus fiber. To activate the citrus fibers, powdered citrus fiber may be activated (hydrated and opened up structurally) by using a high shear dispersion process at low concentration, in water. It is also advantageous to include a preservative into the premix as the dispersed activated citrus fiber is biodegradable.

[0055] A particularly preferred plant fiber may be provided by is Exilva® (from Borregaard).

Encapsulated Benefit Agent

[0056] The composition may further comprise an encapsulated benefit agent. The encapsulated benefit may comprise a shell surrounding a core. The core may comprise a benefit agent. The benefit agent may comprise perfume raw materials.

[0057] The shell may comprise a material selected from the group consisting of aminoplast copolymer, an acrylic, an acrylate, and mixtures thereof. The aminoplast copolymer may be melamine-formaldehyde, urea-formaldehyde, cross-linked melamine formaldehyde, or mixtures thereof.

[0058] The shell may be coated with one or more materials, such as a polymer, that aids in the deposition and/or retention of the perfume microcapsule on the site that is treated with the composition disclosed herein. The polymer may be a cationic polymer selected from the group consisting of polysaccharides, cationically modified starch, cationically modified guar, polysiloxanes, poly diallyl dimethyl ammonium halides, copolymers of poly diallyl dimethyl ammonium chloride and vinyl pyrrolidone, acrylamides, imidazoles, imidazolinium halides, imidazolium halides, poly vinyl amine, copolymers of poly vinyl amine and N-vinyl formamide, and mixtures thereof.

[0059] The core may comprise a benefit agent. Suitable benefit agents include a material selected from the group consisting of perfume raw materials, silicone oils, waxes, hydrocarbons, higher fatty acids, essential oils, lipids, skin coolants, vitamins, sunscreens, antioxidants, glycerine, catalysts, bleach particles, silicon dioxide particles, malodor reducing agents, odor-controlling materials, chelating agents, antistatic agents, softening agents, insect and moth repelling agents, colorants, antioxidants, chelants, bodying agents, drape and form control agents, smoothness agents, wrinkle control agents, sanitization agents, disinfecting agents, germ control agents, mold control agents, mildew control agents, antiviral agents, drying agents, stain resistance agents, soil release agents, fabric refreshing agents and freshness extending agents, chlorine bleach odor control agents, dye fixatives, dye transfer inhibitors, color maintenance agents, optical brighteners, color restoration/rejuvenation agents, anti-fading agents, whiteness enhancers, anti-abrasion agents, wear resistance agents, fabric integrity agents, anti-wear agents, anti-pilling agents, defoamers, anti-foaming agents, UV protection agents, sun fade inhibitors, anti-allergenic agents, enzymes, water proofing agents, fabric comfort agents, shrinkage resistance agents, stretch resistance agents, stretch recovery agents, skin care agents, glycerin, and natural actives, antibacterial actives, antiperspirant actives, cationic polymers, dyes and mixtures thereof. The benefit agent may comprise perfume raw materials.

[0060] The composition may comprise, based on total composition weight, from about 0.01% to about 10%, or from about 0.1% to about 5%, or from about 0.2% to about 1%, of encapsulated benefit agent. The encapsulated benefit agent may be friable and/or have a mean particle size of from about 10 microns to about 500 microns or from about 20 microns to about 200 microns.

[0061] Suitable encapsulated benefit agents may be obtained from Encapsys, LLC, of Appleton, Wisconsin USA.

[0062] Formaldehyde scavengers may also be used in or with such encapsulated benefit agents.

[0063] In a further preferred aspect of the invention, the composition is preferably liquid and comprises particulate benefit agents such as the encapsulated benefit agents mentioned above. The combination of the mannanase enzyme in addition to the plant fiber and particulate benefit agent has been found to provide the additional benefit of enhanced deposition of the particulate benefit agent. Thus, the present invention also provides a method of enhancing deposition of a particulate benefit agent comprising contacting a textile with an aqueous liquor comprising a composition defined herein, comprising a mannanase enzyme and a plant fiber and in addition a particulate benefit agent in a textile treatment step, preferably a laundering step, and optionally rinsing and drying the textile. In a preferred method, the aqueous liquor is an aqueous wash liquor. In a preferred method, the particulate benefit agent comprises an encapsulated perfume particle, most preferably comprising a shell which comprises a material selected from the group consisting of polymers or copolymers comprising acrylic acid and/or acrylates, and mixtures thereof.

Adjuncts

Adjuncts

[0064] The cleaning compositions described herein preferably include other adjunct components, for example selected from surfactants, fabric shading dyes, fabric care benefit agent; additional enzyme; deposition aid; rheology modifier;

builder; chelant; bleach; bleaching agent; bleach precursor; bleach booster; bleach activator, bleach catalyst; perfume and/or perfume microcapsules; perfume loaded zeolite; starch encapsulated accord; polyglycerol esters; whitening agent; pearlescent agent; enzyme stabilizing systems; scavenging agents including fixing agents for anionic dyes, complexing agents for anionic surfactants, and mixtures thereof; optical brighteners or fluoreseers; polymer including but not limited to soil release polymer and/or soil suspension polymer; dispersants; antifoam agents; non-aqueous solvent; fatty acid; suds suppressors, e.g., silicone suds suppressors; cationic starches; scum dispersants; substantive dyes; colorants; opacifier; antioxidant; hydrotropes such as toluenesulfonates, cumenesulfonates and naphthalenesulfonates; color speckles; colored beads, spheres or extrudates; clay softening agents; anti-bacterial agents. Additionally or alternatively, the compositions may comprise surfactants, and/or solvent systems. Quaternary ammonium compounds may be present, particularly in fabric enhancer compositions, such as fabric softeners, and comprise quaternary ammonium cations that are positively charged polyatomic ions of the structure NR_4^+ , where R is an alkyl group or an aryl group.

Additional Enzymes

[0065] Preferably the composition of the invention comprises additional enzymes, for example selected from lipases, amylases, proteases, nucleases, pectate lyases, cellulases, cutinases, and mixtures thereof. The cleaning compositions preferably comprise one or more additional enzymes from the group selected from nucleases. The cleaning compositions preferably comprises one or more additional enzymes selected from the group amylases, lipases, proteases, pectate lyases, cellulases, cutinases, and mixtures thereof. Preferably, the cleaning compositions comprises one or more additional enzymes selected from amylases and proteases and mixtures thereof. Preferably the cleaning compositions comprise one or more additional enzymes selected from lipases. The compositions may also comprise hemicellulases, peroxidases, xylanases, pectinases, keratinases, reductases, oxidases, phenoloxidases, lipoxigenases, ligninases, pullulanases, tannases, pentosanases, malanases, β -glucanases, arabinosidases, hyaluronidase, chondroitinase, lacase and mixtures thereof. When present in the composition, the aforementioned additional enzymes may be present at levels from about 0.00001% to about 2%, from about 0.0001% to about 1% or even from about 0.001% to about 0.5% enzyme protein by weight of the composition. Preferably the or each additional enzyme is present in the laundering aqueous liquor in an amount of from 0.01ppm to 1000 ppm of the active enzyme protein, or from 0.05 or from 0.1ppm to 750 or 500ppm.

Nucleases

[0066] Preferably the composition additionally comprises a nuclease enzyme. The nuclease enzyme is an enzyme capable of cleaving the phosphodiester bonds between the nucleotide subunits of nucleic acids. Suitable nuclease enzymes may be deoxyribonuclease or ribonuclease enzyme or a functional fragment thereof. By functional fragment or part is meant the portion of the nuclease enzyme that catalyzes the cleavage of phosphodiester linkages in the DNA backbone and so is a region of said nuclease protein that retains catalytic activity. Thus it includes truncated, but functional versions, of the enzyme and/or variants and/or derivatives and/or homologues whose functionality is maintained.

[0067] Preferably the nuclease enzyme is a deoxyribonuclease, preferably selected from any of the classes E.C. 3.1.21.x, where x=1, 2, 3, 4, 5, 6, 7, 8 or 9, E.C. 3.1.22.y where y=1, 2, 4 or 5, E.C. 3.1.30.z where z= 1 or 2, E.C. 3.1.31.1 and mixtures thereof. Nuclease enzymes from class E.C. 3.1.21.x and especially where x=1 are particularly preferred. Nucleases in class E.C. 3.1.22.y cleave at the 5' hydroxyl to liberate 3' phosphomonoesters. Enzymes in class E.C. 3.1.30.z may be preferred as they act on both DNA and RNA and liberate 5'-phosphomonoesters. Suitable examples from class E.C. 3.1.31.2 are described in US2012/0135498A, such as SEQ ID NO:3 therein. Such enzymes are commercially available as DENARASE® enzyme from c-LECTA. Nuclease enzymes from class E.C. 3.1.31.1 produce 3' phosphomonoesters.

[0068] Preferably, the nuclease enzyme comprises a microbial enzyme. The nuclease enzyme may be fungal or bacterial in origin. Bacterial nucleases may be most preferred. Fungal nucleases may be most preferred.

[0069] The microbial nuclease is obtainable from *Bacillus*, such as a *Bacillus licheniformis* or *Bacillus subtilis* bacterial nucleases. A preferred nuclease is obtainable from *Bacillus licheniformis*, preferably from strain EI-34-6. A preferred deoxyribonuclease is a variant of *Bacillus licheniformis*, from strain EI-34-6 nucB deoxyribonuclease defined in SEQ ID NO:6 herein, or variant thereof, for example having at least 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identical thereto. Other suitable nucleases are defined in SEQ ID NO: 7 herein, or variant thereof, for example having at least 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identical thereto. Other suitable nucleases are defined in SEQ ID NO: 8 herein, or variant thereof, for example having at least 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identical thereto.

[0070] A fungal nuclease is obtainable from *Aspergillus*, for example *Aspergillus oryzae*. A preferred nuclease is obtainable from *Aspergillus oryzae* defined in SEQ ID NO: 9 herein, or variant thereof, for example having at least 60%

or 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identical thereto.

[0071] Another suitable fungal nuclease is obtainable from *Trichoderma*, for example *Trichoderma harzianum*. A preferred nuclease is obtainable from *Trichoderma harzianum* defined in SEQ ID NO: 10 herein, or variant thereof, for example having at least 60% or 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identical thereto.

[0072] Other fungal nucleases include those encoded by the DNA sequences of *Aspergillus oryzae* RIB40, *Aspergillus oryzae* 3.042, *Aspergillus flavus* NRRL3357, *Aspergillus parasiticus* SU-1, *Aspergillus nomius* NRRL13137, *Trichoderma reesei* QM6a, *Trichoderma virens* Gv29-8, *Oidiodendron maius* Zn, *Metarhizium guizhouense* ARSEF 977, *Metarhizium majus* ARSEF 297, *Metarhizium robertsii* ARSEF 23, *Metarhizium acridum* CQMa 102, *Metarhizium brunneum* ARSEF 3297, *Metarhizium anisopliae*, *Colletotrichum fiorinae* PJ7, *Colletotrichum sublineola*, *Trichoderma atroviride* IMI 206040, *Tolypocladium ophioglossoides* CBS 100239, *Beauveria bassiana* ARSEF 2860, *Colletotrichum higginsianum*, *Hirsutella minnesotensis* 3608, *Scedosporium apiospermum*, *Phaeomoniella chlamydospora*, *Fusarium verticillioides* 7600, *Fusarium oxysporum* f. sp. cubense race 4, *Colletotrichum graminicola* M1.001, *Fusarium oxysporum* FOSC 3-a, *Fusarium avenaceum*, *Fusarium langsethiae*, *Grosmannia clavigera* kw1407, *Claviceps purpurea* 20.1, *Verticillium longisporum*, *Fusarium oxysporum* f. sp. cubense race 1, *Magnaporthe oryzae* 70-15, *Beauveria bassiana* D1-5, *Fusarium pseudograminearum* CS3096, *Neonectria ditissima*, *Magnaportheopsis poae* ATCC 64411, *Cordyceps militaris* CM01, *Marssonina brunnea* f. sp. 'multigermtubi' MB_m1, *Diaporthe ampelina*, *Metarhizium album* ARSEF 1941, *Colletotrichum gloeosporioides* Nara gc5, *Madurella mycetomatis*, *Metarhizium brunneum* ARSEF 3297, *Verticillium alfalfae* VaMs.102, *Gaeumannomyces graminis* var. tritici R3-111a-1, *Nectria haematococca* mpVI 77-13-4, *Verticillium longisporum*, *Verticillium dahliae* VdLs.17, *Botrytis cinerea* B05.10, *Chaetomium globosum* CBS 148.51, *Metarhizium anisopliae*, *Stemphylium lycopersici*, *Sclerotinia borealis* F-4157, *Metarhizium robertsii* ARSEF 23, *Myceliophthora thermophila* ATCC 42464, *Phaeosphaeria nodorum* SN15, *Phialophora attae*, *Ustilagoidea virens*, *Diplodia seriata*, *Ophiostoma piceae* UAMH 11346, *Pseudogymnoascus pannorum* VKM F-4515 (FW-2607), *Bipolaris oryzae* ATCC 44560, *Metarhizium guizhouense* ARSEF 977, *Chaetomium thermophilum* var. thermophilum DSM 1495, *Pestalotiopsis fici* W106-1, *Bipolaris zeicola* 26-R-13, *Setosphaeria turcica* Et28A, *Arthroderma otae* CBS 113480 and *Pyrenophora tritici-repentis* Pt-1C-BFP.

[0073] Preferably the nuclease is an isolated nuclease.

[0074] Preferably the nuclease enzyme is present in the laundering aqueous liquor in an amount of from 0.01ppm to 1000 ppm of the nuclease enzyme, or from 0.05 or from 0.1ppm to 750 or 500ppm.

Acetylglucosaminidases.

[0075] Preferably the composition comprises an acetylglucosaminidase enzyme, preferably a β -N-acetylglucosaminidase enzyme from E.C. 3.2.1.52, preferably an enzyme having at least 70%, or at least 75% or at least 80% or at least 85% or at least 90% or at least 95% or at least 96% or at least 97% or at least 98% or at least 99% or at least or 100% identity to SEQ ID NO: 11.

Galactanase Enzyme

[0076] The compositions preferably additionally comprise an endo-beta-1,6-galactanase enzyme an extracellular polymer-degrading enzyme. The term "endo-beta-1,6-galactanase" or "a polypeptide having endo-beta-1,6-galactanase activity" means a endo-beta-1,6-galactanase activity (EC 3.2.1.164) that catalyzes the hydrolytic cleavage of 1,6-3-D-galactooligosaccharides with a degree of polymerization (DP) higher than 3, and their acidic derivatives with 4-O-methylglucosyluronate or glucosyluronate groups at the non-reducing terminals.

[0077] For purposes of the present disclosure, endo-beta-1,6-galactanase activity is determined according to the procedure described in WO 2015185689 in Assay I. Suitable examples from class EC 3.2.1.164 are described in WO 2015185689, such as the mature polypeptide SEQ ID NO: 2 described therein. Preferably the galactanase enzyme is selected from Glycoside Hydrolase (GH) Family 30.

[0078] Preferably, the endo-beta-1,6-galactanase comprises a microbial enzyme. The endo-beta-1,6-galactanase may be fungal or bacterial in origin. Bacterial endo-beta-1,6-galactanase may be most preferred. Fungal endo-beta-1,6-galactanase may be most preferred.

[0079] A bacterial endo-beta-1,6-galactanase is obtainable from *Streptomyces*, for example *Streptomyces davawensis*. A preferred endo-beta-1,6-galactanase is obtainable from *Streptomyces davawensis* JCM 4913 defined in SEQ ID NO: 12 herein, or a variant thereof, for example having at least 40% or 50% or 60% or 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identity thereto.

[0080] Other bacterial endo-beta-1,6-galactanase include those encoded by the DNA sequences of *Streptomyces avermitilis* MA-4680 with the amino acid sequence defined in SEQ ID NO: 13 herein, or a variant thereof, for example having at least 40% or 50% or 60% or 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identity thereto.

[0081] A fungal endo-beta-1,6-galactanase is obtainable from *Trichoderma*, for example *Trichoderma harzianum*. A preferred endo-beta-1,6-galactanase is obtainable from *Trichoderma harzianum* defined in SEQ ID NO: 14 herein, or a variant thereof, for example having at least 40% or 50% or 60% or 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identical thereto.

[0082] Other fungal endo-beta-1,6-galactanases include those encoded by the DNA sequences of *Ceratocystis fimbriata* f. sp. Platani, *Muscodor strobilii* WG-2009a, *Oculimacula yallundae*, *Trichoderma viride* GD36A, *Thermomyces stellatus*, *Myceliophthora thermophila*.

[0083] Preferably the galactanase has an amino acid sequence having at least 60%, or at least 80%, or at least 90% or at least 95% identity with the amino acid sequence shown in SEQ ID NO:12, SEQ ID NO:13 or SEQ ID NO:14. Preferably the galactanase is an isolated galactanase.

[0084] Preferably the galactanase enzyme is present in a laundering aqueous liquor in an amount of from 0.01ppm to 1000 ppm of the galactanase enzyme, or from 0.05 or from 0.1ppm to 750 or 500ppm.

[0085] The compositions of the invention comprising both mannanase and galactanase may be particularly effective against sticky soils and for improved cleaning. It is believed the two enzymes function together in a complementary way.

Further Glycosyl Hydrolases

[0086] The composition may comprise a glycoside hydrolase selected from GH family 39 and GH family 114 and mixtures thereof, for example as described in co-pending applications having applicants reference numbers CM4645FM and CM4646 FM, respectively.

Proteases.

[0087] Preferably the composition comprises one or more proteases. Suitable proteases include metalloproteases and serine proteases, including neutral or alkaline microbial serine proteases, such as subtilisins (EC 3.4.21.62). Suitable proteases include those of animal, vegetable or microbial origin. In one aspect, such suitable protease may be of microbial origin. The suitable proteases include chemically or genetically modified mutants of the aforementioned suitable proteases. In one aspect, the suitable protease may be a serine protease, such as an alkaline microbial protease or/and a trypsin-type protease. Examples of suitable neutral or alkaline proteases include:

(a) subtilisins (EC 3.4.21.62), preferably those derived from *Bacillus* sp., such as *B. lentus*, *B. alkalophilus*, *B. subtilis*, *B. amyloliquefaciens*, *B. pumilus* and *B. gibsonii* and *B. akibaii* described in WO2004067737, WO2015091989, WO2015091990, WO2015024739, WO2015143360, US 6,312,936 B1, US 5,679,630, US 4,760,025, US7,262,042 and WO09/021867, DE102006022216A1, DE102006022224A1, WO2015089447, WO2015089441, WO2016066756, WO2016066757, WO2016069557, WO2016069563, WO2016069569.

(b) trypsin-type or chymotrypsin-type proteases, such as trypsin (e.g., of porcine or bovine origin), including the *Fusarium* protease described in WO 89/06270 and the chymotrypsin proteases derived from *Cellulomonas* described in WO 05/052161 and WO 05/052146.

(c) metalloproteases, preferably those derived from *Bacillus amyloliquefaciens* described in WO 07/044993A2; from *Bacillus*, *Brevibacillus*, *Thermoactinomyces*, *Geobacillus*, *Paenibacillus*, *Lysinibacillus* or *Streptomyces* spp. Described in WO2014194032, WO2014194054 and WO2014194117; from *Kribella alluminosa* described in WO2015193488; and from *Streptomyces* and *Lysobacter* described in WO2016075078.

(d) protease having at least 90% identity to the subtilase from *Bacillus* sp. TY145, NCIMB 40339, described in WO92/17577 (*Novozymes* A/S), including the variants of this *Bacillus* sp TY145 subtilase described in WO2015024739, and WO2016066757.

[0088] Preferred proteases include those derived from *Bacillus gibsonii* or *Bacillus Lentus*.

[0089] Suitable commercially available protease enzymes include those sold under the trade names Alcalase®, Savinase®, Primase®, Durazym®, Polarzyme®, Kannase®, Liquanase®, Liquanase Ultra®, Savinase Ultra®, Ovozime®, Neutrase®, Everlase® and Esperase® by Novozymes A/S (Denmark), those sold under the tradename Maxatase®, Maxacal®, Maxapem®, Properase®, Purafect®, Purafect Prime®, Purafect Ox®, FN3®, FN4®, Excellase® and Purafect OXP® by Genencor International, those sold under the tradename Opticlean® and Optimase® by Solvay Enzymes, those available from Henkel/ Kemira, namely BLAP (sequence shown in Figure 29 of US 5,352,604 with the following mutations S99D + S101 R + S103A + V104I + G159S, hereinafter referred to as BLAP), BLAP R (BLAP with S3T + V4I + V199M + V205I + L217D), BLAP X (BLAP with S3T + V4I + V205I) and BLAP F49 (BLAP with S3T + V4I + A194P + V199M + V205I + L217D) - all from Henkel/Kemira; and KAP (*Bacillus alkalophilus* subtilisin with mutations A230V + S256G + S259N) from Kao, or as disclosed in WO2009/149144, WO2009/149145, WO2010/56653, WO2010/56640, WO2011/072117, US2011/0237487, WO2011/140316, WO2012/151480, EP2510092, EP2566960 OR EP2705145.

Amylases

[0090] Preferably the composition may comprise an amylase. Suitable alpha-amylases include those of bacterial or fungal origin. Chemically or genetically modified mutants (variants) are included. A preferred alkaline alpha-amylase is derived from a strain of *Bacillus*, such as *Bacillus licheniformis*, *Bacillus amyloliquefaciens*, *Bacillus stearothermophilus*, *Bacillus subtilis*, or other *Bacillus* sp., such as *Bacillus* sp. NCIB 12289, NCIB 12512, NCIB 12513, DSM 9375 (USP 7,153,818) DSM 12368, DSMZ no. 12649, KSM AP1378 (WO 97/00324), KSM K36 or KSM K38 (EP 1,022,334). Preferred amylases include:

- (a) the variants described in WO 94/02597, WO 94/18314, WO96/23874 and WO 97/43424, especially the variants with substitutions in one or more of the following positions versus the enzyme listed as SEQ ID No. 2 in WO 96/23874: 15, 23, 105, 106, 124, 128, 133, 154, 156, 181, 188, 190, 197, 202, 208, 209, 243, 264, 304, 305, 391, 408, and 444.
- (b) the variants described in USP 5,856,164 and WO99/23211, WO 96/23873, WO00/60060 and WO 06/002643, especially the variants with one or more substitutions in the following positions versus the AA560 enzyme listed as SEQ ID No. 12 in WO 06/002643:

26, 30, 33, 82, 37, 106, 118, 128, 133, 149, 150, 160, 178, 182, 186, 193, 203, 214, 231, 256, 257, 258, 269, 270, 272, 283, 295, 296, 298, 299, 303, 304, 305, 311, 314, 315, 318, 319, 339, 345, 361, 378, 383, 419, 421, 437, 441, 444, 445, 446, 447, 450, 461, 471, 482, 484, preferably that also contain the deletions of D183* and G184*.

- (c) variants exhibiting at least 90% identity with SEQ ID No. 4 in WO06/002643, the wild-type enzyme from *Bacillus* SP722, especially variants with deletions in the 183 and 184 positions and variants described in WO 00/60060, which is incorporated herein by reference.

- (d) variants exhibiting at least 95% identity with the wild-type enzyme from *Bacillus* sp.707 (SEQ ID NO:7 in US 6,093, 562), especially those comprising one or more of the following mutations M202, M208, S255, R172, and/or M261. Preferably said amylase comprises one or more of M202L, M202V, M202S, M202T, M202I, M202Q, M202W, S255N and/or R172Q. Particularly preferred are those comprising the M202L or M202T mutations.

- (e) variants described in WO 09/149130, preferably those exhibiting at least 90% identity with SEQ ID NO: 1 or SEQ ID NO:2 in WO 09/149130, the wild-type enzyme from *Geobacillus Stearothermophilus* or a truncated version thereof;

- (f) variants as described in EP2540825 and EP2357220, EP2534233; (g) variants as described in WO2009100102 and WO2010115028.

[0091] Suitable commercially available alpha-amylases include DURAMYL®, LIQUEZYME®, TERMAMYL®, TERMAMYL ULTRA®, NATALASE®, SUPRAMYL®, STAINZYME®, STAINZYME PLUS®, FUNGAMYL® and BAN® (Novozymes A/S, Bagsvaerd, Denmark), KEMZYM® AT 9000 Biozym Biotech Trading GmbH Wehlstrasse 27b A-1200 Wien Austria, RAPIDASE®, PURASTAR®, ENZYSE®, OPTISIZE HT PLUS®, POWERASE® and PURASTAR OX-AM® (Genencor International Inc., Palo Alto, California) and KAM® (Kao, 14-10 Nihonbashi Kayabacho, 1-chome, Chuo-ku Tokyo 103-8210, Japan). In one aspect, suitable amylases include NATALASE®, STAINZYME® and STAINZYME PLUS® and mixtures thereof.

Lipases

[0092] Preferably the composition comprises one or more lipases, including "first cycle lipases" such as those described in U.S. Patent 6,939,702 B1 and US PA 2009/0217464. Preferred lipases are first-wash lipases. In one embodiment of the invention the composition comprises a first wash lipase. First wash lipases includes a lipase which is a polypeptide having an amino acid sequence which: (a) has at least 90% identity with the wild-type lipase derived from *Humicola lanuginosa* strain DSM 4109; (b) compared to said wild-type lipase, comprises a substitution of an electrically neutral or negatively charged amino acid at the surface of the three-dimensional structure within 15Å of E1 or Q249 with a positively charged amino acid; and (c) comprises a peptide addition at the C-terminal; and/or (d) comprises a peptide addition at the N-terminal and/or (e) meets the following limitations: i) comprises a negative amino acid in position E210 of said wild-type lipase; ii) comprises a negatively charged amino acid in the region corresponding to positions 90-101 of said wild-type lipase; and iii) comprises a neutral or negative amino acid at a position corresponding to N94 or said wild-type lipase and/or has a negative or neutral net electric charge in the region corresponding to positions 90-101 of said wild-type lipase. Preferred are variants of the wild-type lipase from *Thermomyces lanuginosus* comprising one or more of the T231R and N233R mutations. The wild-type sequence is the 269 amino acids (amino acids 23 - 291) of the Swissprot accession number Swiss-Prot O59952 (derived from *Thermomyces lanuginosus* (*Humicola lanuginosa*)).

Preferred lipases include those sold under the tradenames Lipex® and Lipolex® and Lipoclean®. Other suitable lipases include those described in European Patent Application No. 12001034.3 or EP2623586.

Endoglucanases

[0093] Other preferred enzymes include microbial-derived endoglucanases exhibiting endo-beta-1,4-glucanase activity (E.C. 3.2.1.4), including a bacterial polypeptide endogenous to a member of the genus *Bacillus* which has a sequence of at least 90%, 94%, 97% and even 99% identity to the amino acid sequence SEQ ID NO:2 in US7,141,403B2) and mixtures thereof. Suitable endoglucanases are sold under the tradenames Celluclean® and Whitezyme® (Novozymes A/S, Bagsvaerd, Denmark).

Pectate Lyases

[0094] Other preferred enzymes include pectate lyases sold under the tradenames Pectawash®, Pectaway®, Xpect® and mannanases sold under the tradenames Mannaway® (all from Novozymes A/S, Bagsvaerd, Denmark), and Purabrite® (Genencor International Inc., Palo Alto, California).

Cleaning Cellulase

[0095] The cleaning composition described herein may additionally comprise a cleaning cellulase. The cellulase may be an endoglucanase. The cellulase may have endo beta 1,4-glucanase activity and a structure which does not comprise a class A Carbohydrate Binding Module (CBM). A class A CBM is defined according to A. B. Boraston et al. Biochemical Journal 2004, Volume 382 (part 3) pages 769-781. In particular, the cellulase does not comprise a class A CBM from families 1, 2a, 3, 5 and 10.

[0096] The cellulase may be a glycosyl hydrolase having enzymatic activity towards amorphous cellulose substrates, wherein the glycosyl hydrolase is selected from GH families 5, 7, 12, 16, 44 or 74. Preferably, the cellulase is a glycosyl hydrolase selected from GH family 5. A preferred cellulase is Celluclean, supplied by Novozymes. This preferred cellulase is described in more detail in WO2002/099091. The glycosyl hydrolase (GH) family definition is described in more detail in Biochem J. 1991, v280, 309-316. Another preferred cellulase is a glycosyl hydrolase having enzymatic activity towards both xyloglucan and amorphous cellulose substrates, wherein the glycosyl hydrolase is selected from GH families 5, 12, 44 or 74. Preferably, the glycosyl hydrolase selected from GH family 44.

[0097] For purposes of the present invention, the degree of identity between two amino acid sequences is determined using the Needleman-Wunsch algorithm (Needleman and Wunsch, 1970, J. Mol. Biol. 48: 443-453) as implemented in the Needle program of the EMBOSS package (EMBOSS: The European Molecular Biology Open Software Suite, Rice et al., 2000, Trends in Genetics 16: 276-277), preferably version 3.0.0 or later. The optional parameters used are gap open penalty of 10, gap extension penalty of 0.5, and the EBLOSUM62 (EMBOSS version of BLOSUM62) substitution matrix. The output of Needle labeled "longest identity" (obtained using the -nobrief option) is used as the percent identity and is calculated as follows: (Identical Residues x 100)/(Length of Alignment - Total Number of Gaps in Alignment).

[0098] Suitable cleaning cellulase glycosyl hydrolases are selected from the group consisting of: GH family 44 glycosyl hydrolases from *Paenibacillus polyxyma* (wild-type) such as XYG1006 described in WO 01/062903 or are variants thereof; GH family 12 glycosyl hydrolases from *Bacillus licheniformis* (wild-type) such as Seq. No. ID: 1 described in WO 99/02663 or are variants thereof; GH family 5 glycosyl hydrolases from *Bacillus agaradhaerens* (wild type) or variants thereof; GH family 5 glycosyl hydrolases from *Paenibacillus* (wild type) such as XYG1034 and XYG 1022 described in WO 01/064853 or variants thereof; GH family 74 glycosyl hydrolases from *Jonesia* sp. (wild type) such as XYG1020 described in WO 2002/077242 or variants thereof; and GH family 74 glycosyl hydrolases from *Trichoderma Reesei* (wild type), such as the enzyme described in more detail in Sequence ID no. 2 of WO03/089598, or variants thereof.

[0099] Preferred glycosyl hydrolases are selected from the group consisting of: GH family 44 glycosyl hydrolases from *Paenibacillus polyxyma* (wild-type) such as XYG1006 or are variants thereof.

[0100] Typically, the cellulase modifies the fabric surface during the laundering process so as to improve the removal of soils adhered to the fabric after the laundering process during wearing and usage of the fabric, in subsequent wash cycles. Preferably, the cellulase modifies the fabric surface during the laundering process so as to improve the removal of soils adhered to the fabric after the laundering process during wearing and usage of the fabric, in the subsequent two, or even three wash cycles.

[0101] Typically, the cellulase is used at a concentration of 0.005ppm to 1.0ppm in the aqueous liquor during the first laundering process. Preferably, the cellulase is used at a concentration of 0.02ppm to 0.5ppm in the aqueous liquor during the first laundering process.

Surfactant system

[0102] The cleaning composition preferably comprises a surfactant system. The cleaning composition preferably comprises from about 1% to about 80%, or from 1% to about 60%, preferably from about 5% to about 50% more preferably from about 8% to about 40%, by weight of the cleaning composition, of a surfactant system.

[0103] Surfactants suitable for use in the surfactant system may be derived from natural and/or renewable sources.

[0104] The surfactant system may comprise an anionic surfactant, more preferably an anionic surfactant selected from the group consisting of, alkyl benzene sulfonate, alkyl sulfate, alkyl alkoxy sulfate, especially alkyl ethoxy sulfate, paraffin sulfonate and mixtures thereof, alkyl benzene sulfonates are particularly preferred. The surfactant system may further comprise a surfactant selected from the group consisting of nonionic surfactant, cationic surfactant, amphoteric surfactant, zwitterionic surfactant, and mixtures thereof. The surfactant system preferably comprises a nonionic surfactant, for example an ethoxylated nonionic surfactant. The surfactant system may comprise an amphoteric surfactant, for example an amine oxide surfactant, such as an alkyl dimethyl amine oxide. The surfactant system may comprise a zwitterionic surfactant, such as a betaine.

[0105] The most preferred surfactant system for the detergent composition of the present invention comprises from 1% to 40%, preferably 6% to 35%, more preferably 8% to 30% weight of the total composition of an anionic surfactant, preferably comprising an alkyl benzene sulphonate. The preferred surfactant system may optionally in addition comprise an alkyl alkoxy sulfate surfactant, more preferably an alkyl ethoxy sulfate, optionally combined with 0.5% to 15%, preferably from 1% to 12%, more preferably from 2% to 10% by weight of the composition of amphoteric and/or zwitterionic surfactant, more preferably an amphoteric and even more preferably an amine oxide surfactant, especially an alkyl dimethyl amine oxide.

[0106] Preferably the composition further comprises a nonionic surfactant, especially an alcohol alkoxyate in particular an alcohol ethoxyate nonionic surfactant. Most preferably the surfactant system comprises an anionic and a nonionic surfactant, preferably the weight ratio of the anionic to nonionic surfactant is from 25:1 to 1:2.

Anionic surfactant

[0107] Anionic surfactants may be in salt form or acid form, typically in the form of a water-soluble sodium, potassium, ammonium, magnesium or mono-, di- or tri- C2-C3 alkanolammonium salt, with the sodium cation being the usual one chosen.

Sulfonate Surfactant

[0108] Suitable anionic sulfonate surfactants for use herein include water-soluble salts of C8-C18 alkyl or hydroxyalkyl sulfonates; C11-C18 alkyl benzene sulfonates (LAS), modified alkylbenzene sulfonate (MLAS) as discussed in WO 99/05243, WO 99/05242, WO 99/05244, WO 99/05082, WO 99/05084, WO 99/05241, WO 99/07656, WO 00/23549, and WO 00/23548; methyl ester sulfonate (MES); and alpha-olefin sulfonate (AOS). Those also include the paraffin sulfonates may be monosulfonates and/or disulfonates, obtained by sulfonating paraffins of 10 to 20 carbon atoms. The sulfonate surfactant may also include the alkyl glyceryl sulfonate surfactants.

Sulfated anionic surfactant

[0109] Preferably the sulfated anionic surfactant is alkoxyated, more preferably, an alkoxyated branched sulfated anionic surfactant having an alkoxylation degree of from about 0.2 to about 4, even more preferably from about 0.3 to about 3, even more preferably from about 0.4 to about 1.5 and especially from about 0.4 to about 1. Preferably, the alkoxy group is ethoxy. When the sulfated anionic surfactant is a mixture of sulfated anionic surfactants, the alkoxylation degree is the weight average alkoxylation degree of all the components of the mixture (weight average alkoxylation degree). In the weight average alkoxylation degree calculation the weight of sulfated anionic surfactant components not having alkoxyated groups should also be included.

$$\text{Weight average alkoxylation degree} = (x_1 * \text{alkoxylation degree of surfactant 1} + x_2 * \text{alkoxylation degree of surfactant 2} + \dots) / (x_1 + x_2 + \dots)$$

wherein x_1 , x_2 , ... are the weights in grams of each sulfated anionic surfactant of the mixture and alkoxylation degree is the number of alkoxy groups in each sulfated anionic surfactant.

[0110] Preferably, the branching group is an alkyl. Typically, the alkyl is selected from methyl, ethyl, propyl, butyl,

pentyl, cyclic alkyl groups and mixtures thereof. Single or multiple alkyl branches could be present on the main hydrocarbyl chain of the starting alcohol(s) used to produce the sulfated anionic surfactant used in the detergent of the invention. Most preferably the branched sulfated anionic surfactant is selected from alkyl sulfates, alkyl ethoxy sulfates, and mixtures thereof.

[0111] The branched sulfated anionic surfactant can be a single anionic surfactant or a mixture of anionic surfactants. In the case of a single surfactant the percentage of branching refers to the weight percentage of the hydrocarbyl chains that are branched in the original alcohol from which the surfactant is derived.

[0112] In the case of a surfactant mixture the percentage of branching is the weight average and it is defined according to the following formula:

$$\text{Weight average of branching (\%)} = [(x_1 * \text{wt\% branched alcohol 1 in alcohol 1} + x_2 * \text{wt\% branched alcohol 2 in alcohol 2} + \dots) / (x_1 + x_2 + \dots)] * 100$$

wherein x_1 , x_2 , ... are the weight in grams of each alcohol in the total alcohol mixture of the alcohols which were used as starting material for the anionic surfactant for the detergent of the invention. In the weight average branching degree calculation the weight of anionic surfactant components not having branched groups should also be included.

[0113] Suitable sulfate surfactants for use herein include water-soluble salts of C8-C18 alkyl or hydroxyalkyl, sulfate and/or ether sulfate. Suitable counterions include alkali metal cation or ammonium or substituted ammonium, but preferably sodium.

[0114] The sulfate surfactants may be selected from C8-C18 primary, branched chain and random alkyl sulfates (AS); C8-C18 secondary (2,3) alkyl sulfates; C8-C18 alkyl alkoxy sulfates (AExS) wherein preferably x is from 1-30 in which the alkoxy group could be selected from ethoxy, propoxy, butoxy or even higher alkoxy groups and mixtures thereof.

[0115] Alkyl sulfates and alkyl alkoxy sulfates are commercially available with a variety of chain lengths, ethoxylation and branching degrees. Commercially available sulfates include, those based on Neodol alcohols ex the Shell company, Lial - Isalchem and Safol ex the Sasol company, natural alcohols ex The Procter & Gamble Chemicals company.

[0116] Preferred alkyl sulfates are those in which the anionic surfactant is an alkyl ethoxy sulfate with a degree of ethoxylation of from about 0.2 to about 3, more preferably from about 0.3 to about 2, even more preferably from about 0.4 to about 1.5, and especially from about 0.4 to about 1. They are also preferred anionic surfactant having a level of branching of from about 5% to about 40%, even more preferably from about 10% to 35% and especially from about 20% to 30%.

Nonionic surfactant

[0117] Preferably the surfactant system comprises a nonionic surfactant, in an amount of from 0.1% to 40%, preferably 0.2% to 20%, most preferably 0.5% to 10% by weight of the composition. Suitable nonionic surfactants include the condensation products of aliphatic alcohols with from 1 to 25 moles of ethylene oxide. The alkyl chain of the aliphatic alcohol can either be straight or branched, primary or secondary, and generally contains from 8 to 22 carbon atoms. Particularly preferred are the condensation products of alcohols having an alkyl group containing from 10 to 18 carbon atoms, preferably from 10 to 15 carbon atoms with from 2 to 18 moles, preferably 2 to 15, more preferably 5-12 of ethylene oxide per mole of alcohol. Highly preferred nonionic surfactants are the condensation products of guerbet alcohols with from 2 to 18 moles, preferably 2 to 15, more preferably 5-12 of ethylene oxide per mole of alcohol.

[0118] Other suitable non-ionic surfactants for use herein include fatty alcohol polyglycol ethers, alkylpolyglucosides and fatty acid glucamides.

Amphoteric surfactant

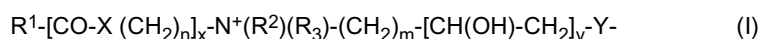
[0119] The surfactant system may include amphoteric surfactant, such as amine oxide. Preferred amine oxides are alkyl dimethyl amine oxide or alkyl amido propyl dimethyl amine oxide, more preferably alkyl dimethyl amine oxide and especially coco dimethyl amino oxide. Amine oxide may have a linear or mid-branched alkyl moiety. Typical linear amine oxides include water-soluble amine oxides containing one R1 C8-18 alkyl moiety and 2 R2 and R3 moieties selected from the group consisting of C1-3 alkyl groups and C1-3 hydroxyalkyl groups. Preferably amine oxide is characterized by the formula $R_1 - N(R_2)(R_3)O$ wherein R1 is a C8-18 alkyl and R2 and R3 are selected from the group consisting of methyl, ethyl, propyl, isopropyl, 2-hydroxyethyl, 2-hydroxypropyl and 3-hydroxypropyl. The linear amine oxide surfactants in particular may include linear C10-C18 alkyl dimethyl amine oxides and linear C8-C12 alkoxy ethyl dihydroxy ethyl amine oxides. Preferred amine oxides include linear C10, linear C10-C12, and linear C12-C14 alkyl dimethyl amine oxides. As used herein "mid-branched" means that the amine oxide has one alkyl moiety having n_1 carbon atoms with

one alkyl branch on the alkyl moiety having n_2 carbon atoms. The alkyl branch is located on the α carbon from the nitrogen on the alkyl moiety. This type of branching for the amine oxide is also known in the art as an internal amine oxide. The total sum of n_1 and n_2 is from 10 to 24 carbon atoms, preferably from 12 to 20, and more preferably from 10 to 16. The number of carbon atoms for the one alkyl moiety (n_1) should be approximately the same number of carbon atoms as the one alkyl branch (n_2) such that the one alkyl moiety and the one alkyl branch are symmetric. As used herein "symmetric" means that $|n_1 - n_2|$ is less than or equal to 5, preferably 4, most preferably from 0 to 4 carbon atoms in at least 50 wt%, more preferably at least 75 wt% to 100 wt% of the mid-branched amine oxides for use herein.

[0120] The amine oxide may further comprise two moieties, independently selected from a C1-3 alkyl, a C1-3 hydroxy-alkyl group, or a polyethylene oxide group containing an average of from about 1 to about 3 ethylene oxide groups. Preferably the two moieties are selected from a C1-3 alkyl, more preferably both are selected as a C1 alkyl.

Zwitterionic surfactant

[0121] Other suitable surfactants include betaines, such as alkyl betaines, alkylamidobetaine, amidazoliniumbetaine, sulfobetaine (INCI Sultaines) as well as the Phosphobetaine and preferably meets formula (I):



wherein

R^1 is a saturated or unsaturated C6-22 alkyl residue, preferably C8-18 alkyl residue, in particular a saturated C10-16 alkyl residue, for example a saturated C12-14 alkyl residue;

X is NH, NR^4 with C1-4 Alkyl residue R^4 , O or S,

n a number from 1 to 10, preferably 2 to 5, in particular 3,

x 0 or 1, preferably 1,

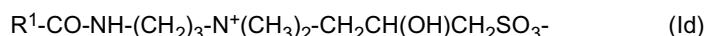
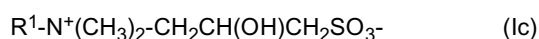
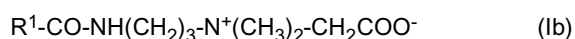
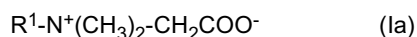
R^2 , R^3 are independently a C1-4 alkyl residue, potentially hydroxy substituted such as a hydroxyethyl, preferably a methyl.

m a number from 1 to 4, in particular 1, 2 or 3,

y 0 or 1 and

Y is COO, SO₃, OPO(OR⁵)O or P(O)(OR⁵)O, whereby R^5 is a hydrogen atom H or a C1-4 alkyl residue.

[0122] Preferred betaines are the alkyl betaines of the formula (Ia), the alkyl amido propyl betaine of the formula (Ib), the Sulfo betaines of the formula (Ic) and the Amido sulfobetaine of the formula (Id);



in which R^1 has the same meaning as in formula I. Particularly preferred betaines are the Carbobetaine [wherein $Y^- = COO^-$], in particular the Carbobetaine of the formula (Ia) and (Ib), more preferred are the Alkylamidobetaine of the formula (Ib).

[0123] Examples of suitable betaines and sulfobetaine are the following [designated in accordance with INCI]: Almondamidopropyl of betaines, Apricotamidopropyl betaines, Avocamidopropyl of betaines, Babassuamidopropyl of betaines, Behenamidopropyl betaines, Behenyl of betaines, betaines, Canolamidopropyl betaines, Capryl/Capramidopropyl betaines, Carnitine, Cetyl of betaines, Cocamidopropyl of betaines, Cocamidopropyl betaines, Cocamidopropyl Hydroxysultaine, Coco betaines, Coco Hydroxysultaine, Coco/Oleamidopropyl betaines, Coco Sultaine, Decyl of betaines, Dihydroxyethyl Oleyl Glycinate, Dihydroxyethyl Soy Glycinate, Dihydroxyethyl Stearyl Glycinate, Dihydroxyethyl Tallow Glycinate, Dimethicone Propyl of PG-betaines, Erucamidopropyl Hydroxysultaine, Hydrogenated Tallow of betaines, Isostearamidopropyl betaines, Lauramidopropyl betaines, Lauryl of betaines, Lauryl Hydroxysultaine, Lauryl Sultaine, Milkamidopropyl betaines, Minkamidopropyl of betaines, Myristamidopropyl betaines, Myristyl of betaines, Oleamidopropyl betaines, Oleamidopropyl Hydroxysultaine, Oleyl of betaines, Olivamidopropyl of betaines, Palmamidopropyl betaines, Palm itamidopropyl betaines, Palmitoyl Carnitine, Palm Kernelamidopropyl betaines, Polytetrafluoroethylene Acetoxypromyl of betaines, Ricinoleamidopropyl betaines, Sesamidopropyl betaines, Soyamidopropyl betaines, Stearamidopropyl betaines, Stearyl of betaines, Tallowamidopropyl betaines, Tallowamidopropyl Hydrox-

ysultaine, Tallow of betaines, Tallow Dihydroxyethyl of betaines, Undecylenam idopropyl betaines and Wheat German idopropyl betaines. A preferred betaine is, for example, Cocoamidopropylbetaine.

Fatty Acid

[0124] Especially when in liquid form, preferably, the detergent composition comprises between 1.5% and 20%, more preferably between 2% and 15%, even more preferably between 3% and 10%, most preferably between 4% and 8% by weight of the liquid detergent composition of soap, preferably a fatty acid salt, more preferably an amine neutralized fatty acid salt, wherein preferably the amine is an alkanolamine more preferably selected from monoethanolamine, diethanolamine, triethanolamine or a mixture thereof, more preferably monoethanolamine.

Perfume

[0125] Preferred compositions of the invention comprise perfume. Typically the composition comprises a perfume that comprises one or more perfume raw materials, selected from the group as described in WO08/87497. However, any perfume useful in a detergent may be used. A preferred method of incorporating perfume into the compositions of the invention is via an encapsulated perfume particle comprising either a water-soluble hydroxylic compound or melamine-formaldehyde or modified polyvinyl alcohol. In one aspect the encapsulate comprises (a) an at least partially water-soluble solid matrix comprising one or more water-soluble hydroxylic compounds, preferably starch; and (b) a perfume oil encapsulated by the solid matrix. In a further aspect the perfume may be pre-complexed with a polyamine, preferably a polyethylenimine so as to form a Schiff base.

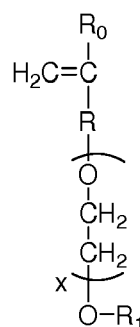
Polymers

[0126] The detergent composition may comprise one or more polymers for example for cleaning and/or care. Examples are optionally modified carboxymethylcellulose, poly (ethylene glycol), poly(vinyl alcohol), polycarboxylates such as polyacrylates, maleic/acrylic acid copolymers and lauryl methacrylate/acrylic acid co-polymers and carboxylate polymers.

[0127] Suitable carboxylate polymers include maleate/acrylate random copolymer or polyacrylate homopolymer. The carboxylate polymer may be a polyacrylate homopolymer having a molecular weight of from 4,000 Da to 9,000 Da, or from 6,000 Da to 9,000 Da. Other suitable carboxylate polymers are co-polymers of maleic acid and acrylic acid, and may have a molecular weight in the range of from 4,000 Da to 90,000 Da.

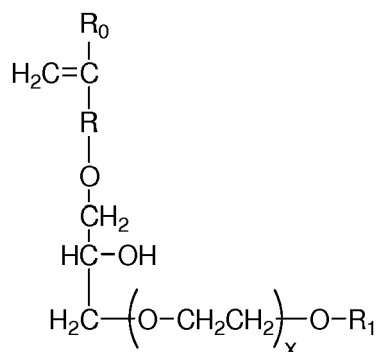
[0128] Other suitable carboxylate polymers are co-polymers comprising: (i) from 50 to less than 98 wt% structural units derived from one or more monomers comprising carboxyl groups; (ii) from 1 to less than 49 wt% structural units derived from one or more monomers comprising sulfonate moieties; and (iii) from 1 to 49 wt% structural units derived from one or more types of monomers selected from ether bond-containing monomers represented by formulas (I) and (II):

formula (I):



wherein in formula (I), R_0 represents a hydrogen atom or CH_3 group, R represents a CH_2 group, CH_2CH_2 group or single bond, X represents a number 0-5 provided X represents a number 1-5 when R is a single bond, and R_1 is a hydrogen atom or C1 to C20 organic group;

formula (II)



in formula (II), R_0 represents a hydrogen atom or CH_3 group, R represents a CH_2 group, CH_2CH_2 group or single bond, X represents a number 0-5, and R_1 is a hydrogen atom or C1 to C20 organic group.

[0129] The composition may comprise one or more amphiphilic cleaning polymers such as the compound having the following general structure: $bis((C_2H_5O)(C_2H_4O)_n)(CH_3)-N^+-C_xH_{2x}-N^+-(CH_3)-bis((C_2H_5O)(C_2H_4O)_n)$, wherein n = from 20 to 30, and x = from 3 to 8, or sulphated or sulphonated variants thereof. In one aspect, this polymer is sulphated or sulphonated to provide a zwitterionic soil suspension polymer.

[0130] The composition preferably comprises amphiphilic alkoxyated grease cleaning polymers which have balanced hydrophilic and properties such that they remove grease particles from fabrics and surfaces. Preferred amphiphilic alkoxyated grease cleaning polymers comprise a core structure and a plurality of alkoxyate groups attached to that core structure. These may comprise alkoxyated polyalkylenimines, preferably having an inner polyethylene oxide block and an outer polypropylene oxide block. Typically these may be incorporated into the compositions of the invention in amounts of from 0.005 to 10 wt%, generally from 0.5 to 8 wt%.

[0131] Alkoxyated polycarboxylates such as those prepared from polyacrylates are useful herein to provide additional grease removal performance. Such materials are described in WO 91/08281 and PCT 90/01815. Chemically, these materials comprise polyacrylates having one ethoxy side-chain per every 7-8 acrylate units. The side-chains are of the formula $-(CH_2CH_2O)_m(CH_2)_nCH_3$ wherein m is 2-3 and n is 6-12. The side-chains are ester-linked to the polyacrylate "backbone" to provide a "comb" polymer type structure. The molecular weight can vary, but is typically in the range of about 2000 to about 50,000. Such alkoxyated polycarboxylates can comprise from about 0.05% to about 10%, by weight, of the compositions herein.

[0132] The composition may comprise polyethylene glycol polymers and these may be particularly preferred in compositions comprising mixed surfactant systems. Suitable polyethylene glycol polymers include random graft co-polymers comprising: (i) hydrophilic backbone comprising polyethylene glycol; and (ii) side chain(s) selected from the group consisting of: C4-C25 alkyl group, polypropylene, polybutylene, vinyl ester of a saturated C1-C6 mono-carboxylic acid, C1-C6 alkyl ester of acrylic or methacrylic acid, and mixtures thereof. Suitable polyethylene glycol polymers have a polyethylene glycol backbone with random grafted polyvinyl acetate side chains. The average molecular weight of the polyethylene glycol backbone can be in the range of from 2,000 Da to 20,000 Da, or from 4,000 Da to 8,000 Da. The molecular weight ratio of the polyethylene glycol backbone to the polyvinyl acetate side chains can be in the range of from 1:1 to 1:5, or from 1:1.2 to 1:2. The average number of graft sites per ethylene oxide units can be less than 1, or less than 0.8, the average number of graft sites per ethylene oxide units can be in the range of from 0.5 to 0.9, or the average number of graft sites per ethylene oxide units can be in the range of from 0.1 to 0.5, or from 0.2 to 0.4. A suitable polyethylene glycol polymer is Sokalan HP22.

[0133] Typically these polymers when present are each incorporated into the compositions of the invention in amounts from 0.005 to 10 wt%, more usually from 0.05 to 8 wt%.

[0134] Preferably the composition comprises one or more carboxylate polymer, such as a maleate/acrylate random copolymer or polyacrylate homopolymer. In one aspect, the carboxylate polymer is a polyacrylate homopolymer having a molecular weight of from 4,000 Da to 9,000 Da, or from 6,000 Da to 9,000 Da. Typically these are incorporated into the compositions of the invention in amounts from 0.005 to 10 wt%, or from 0.05 to 8 wt%.

[0135] Preferably the composition comprises one or more soil release polymers.

[0136] Suitable soil release polymers are polyester soil release polymers such as Repel-o-tex polymers, including Repel-o-tex SF, SF-2 and SRP6 supplied by Rhodia. Other suitable soil release polymers include Texcare polymers, including Texcare SRA100, SRA300, SRN100, SRN170, SRN240, SRN260, SRN300 and SRN325 supplied by Clariant. Other suitable soil release polymers are Marloquest polymers, such as Marloquest SL supplied by Sasol.

[0137] Preferably the composition comprises one or more cellulosic polymer, including those selected from alkyl cellulose, alkyl alkoxyalkyl cellulose, carboxyalkyl cellulose, alkyl carboxyalkyl cellulose. Preferred cellulosic polymers are selected from the group comprising carboxymethyl cellulose, methyl cellulose, methyl hydroxyethyl cellulose, methyl carboxymethyl cellulose, and mixtures thereof. In one aspect, the carboxymethyl cellulose has a degree of carboxymethyl substitution from 0.5 to 0.9 and a molecular weight from 100,000 Da to 300,000 Da.

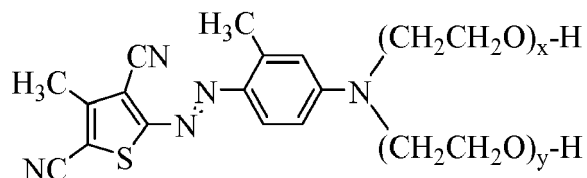
[0138] The composition preferably comprises a cationically-modified polysaccharide polymer. Preferably, the cationic polysaccharide polymer is selected from cationically modified hydroxyethyl cellulose, cationically modified hydroxypropyl cellulose, cationically and hydrophobically modified hydroxyethyl cellulose, cationically and hydrophobically modified hydroxypropyl cellulose, or a mixture thereof, more preferably cationically modified hydroxyethyl cellulose, cationically and hydrophobically modified hydroxyethyl cellulose, or a mixture thereof.

Amines

[0139] The cleaning compositions described herein preferably contain an amine, preferably in an amount from about 0.1% to about 10%, or from about 0.2% to about 5%, or from about 0.5% to about 4%, or from about 0.1% to about 4%, or from about 0.1% to about 2%, by weight of the composition, of an amine. The amine can be subjected to protonation depending on the pH of the cleaning medium in which it is used. Non-limiting examples of preferred amines include, but are not limited to, etheramines, cyclic amines, polyamines, oligoamines (e.g., triamines, diamines, pentamines, tetraamines), or combinations thereof. The compositions described herein may comprise an amine selected from the group consisting of oligoamines, etheramines, cyclic amines, and combinations thereof. In some aspects, the amine is not an alkanolamine. In some aspects, the amine is not a polyalkyleneimine. Examples of suitable oligoamines include tetraethylenepentamine, triethylenetetraamine, diethylenetriamine, and mixtures thereof. Etheramines and cyclic amines are particularly preferred.

Fabric Shading Dye

[0140] The composition preferably comprises a fabric shading agent. Suitable fabric shading agents include dyes, dye-clay conjugates, and pigments. Suitable dyes include small molecule dyes and polymeric dyes. Suitable small molecule dyes include small molecule dyes selected from the group consisting of dyes falling into the Colour Index (C.I.) classifications of Direct Blue, Direct Red, Direct Violet, Acid Blue, Acid Red, Acid Violet, Basic Blue, Basic Violet and Basic Red, or mixtures thereof. Preferred dyes include alkoxyated azothiophenes, Solvent Violet 13, Acid Violet 50 and Direct Violet 9. Particularly preferred dyes are polymeric dyes, particularly comprising polyalkoxy, most preferably polyethoxy groups, for example:



wherein the index values x and y are independently selected from 1 to 10.

Dye Transfer Inhibitors

[0141] Suitable dye transfer inhibitors include polyamine N-oxide polymers, copolymers of N-vinylpyrrolidone and N-vinylimidazole, polyvinylpyrrolidone, polyvinylloxazolidone, polyvinylimidazole and mixtures thereof. Preferred are poly(vinyl pyrrolidone), poly(vinylpyridine betaine), poly(vinylpyridine N-oxide), poly(vinyl pyrrolidone-vinyl imidazole) and mixtures thereof. Suitable commercially available dye transfer inhibitors include PVP-K15 and K30 (Ashland), Sokalan® HP165, HP50, HP53, HP59, HP56K, HP56, HP66 (BASF), Chromabond® S-400, S403E and S-100 (Ashland).

Chelant

[0142] The composition may comprise chelant for example selected from phosphonic, sulphonic, succinic and acetic chelants or mixtures thereof. Suitable examples include HEDP, DTPA, EDTA, MGDA, GLDA, EDDS and 4,5-dihydroxy-1,3-benzenedisulfonic acids and salts thereof.

Methods of Making the Composition

[0143] The present invention relates to methods of making the compositions described herein. The compositions of the invention may be solid (for example granules or tablets) or liquid form. Preferably the compositions are in liquid form. They may be made by any process chosen by the formulator, including by a batch process, a continuous loop process, or combinations thereof.

[0144] When in the form of a liquid, the compositions of the invention may be aqueous (typically above 2 wt% or even above 5 or 10 wt% total water, up to 90 or up to 80wt% or 70 wt% total water) or non-aqueous (typically below 2 wt% total water content). Typically the compositions of the invention will be in the form of an aqueous solution or uniform dispersion or suspension of optical brightener, DTI and optional additional adjunct materials, some of which may normally be in solid form, that have been combined with the normally liquid components of the composition, such as the liquid alcohol ethoxylate nonionic, the aqueous liquid carrier, and any other normally liquid optional ingredients. Such a solution, dispersion or suspension will be acceptably phase stable. When in the form of a liquid, the detergents of the invention preferably have viscosity from 1 to 1500 centipoises (1-1500 mPa*s), more preferably from 100 to 1000 centipoises (100-1000 mPa*s), and most preferably from 200 to 500 centipoises (200-500 mPa*s) at 20s-1 and 21°C. Viscosity can be determined by conventional methods. Viscosity may be measured using an AR 550 rheometer from TA instruments using a plate steel spindle at 40 mm diameter and a gap size of 500 μ m. The high shear viscosity at 20s-1 and low shear viscosity at 0.05-1 can be obtained from a logarithmic shear rate sweep from 0.1-1 to 25-1 in 3 minutes time at 21°C. The preferred rheology described therein may be achieved using internal existing structuring with detergent ingredients or by employing an external rheology modifier. More preferably the detergents, such as detergent liquid compositions have a high shear rate viscosity of from about 100 centipoise to 1500 centipoise, more preferably from 100 to 1000 cps. Unit Dose detergents, such as detergent liquid compositions have high shear rate viscosity of from 400 to 1000cps. Detergents such as laundry softening compositions typically have high shear rate viscosity of from 10 to 1000, more preferably from 10 to 800 cps, most preferably from 10 to 500 cps. Hand dishwashing compositions have high shear rate viscosity of from 300 to 4000 cps, more preferably 300 to 1000 cps.

[0145] The cleaning and/or treatment compositions in the form of a liquid herein can be prepared by combining the components thereof in any convenient order and by mixing, e.g., agitating, the resulting component combination to form a phase stable liquid detergent composition. In a process for preparing such compositions, a liquid matrix is formed containing at least a major proportion, or even substantially all, of the liquid components, e.g., nonionic surfactant, the non-surface active liquid carriers and other optional liquid components, with the liquid components being thoroughly admixed by imparting shear agitation to this liquid combination. For example, rapid stirring with a mechanical stirrer may usefully be employed. While shear agitation is maintained, substantially all of any anionic surfactants and the solid form ingredients can be added. Agitation of the mixture is continued, and if necessary, can be increased at this point to form a solution or a uniform dispersion of insoluble solid phase particulates within the liquid phase. After some or all of the solid-form materials have been added to this agitated mixture, particles of any enzyme material to be included, e.g., enzyme granulates, are incorporated. As a variation of the composition preparation procedure hereinbefore described, one or more of the solid components may be added to the agitated mixture as a solution or slurry of particles premixed with a minor portion of one or more of the liquid components. After addition of all of the composition components, agitation of the mixture is continued for a period of time sufficient to form compositions having the requisite viscosity and phase stability characteristics. Frequently this will involve agitation for a period of from about 30 to 60 minutes.

[0146] The adjunct ingredients in the compositions of this invention may be incorporated into the composition as the product of the synthesis generating such components, either with or without an intermediate purification step. Where there is no purification step, commonly the mixture used will comprise the desired component or mixtures thereof (and percentages given herein relate to the weight percent of the component itself unless otherwise specified) and in addition unreacted starting materials and impurities formed from side reactions and/or incomplete reaction. For example, for an ethoxylated or substituted component, the mixture will likely comprise different degrees of ethoxylation/substitution.

Method of Use

[0147] The present invention relates to methods of using the cleaning compositions of the present invention to clean a surface, such as a textile. In general, the method includes mixing the cleaning composition as described herein with water to form an aqueous liquor and contacting a surface, preferably a textile, with the aqueous liquor in a laundering step. The target surface may include a greasy soil such as a body soil. The compositions herein, typically prepared as hereinbefore described, can be used to form aqueous washing/treatment solutions for use in the laundering/treatment of fabrics and/or hard surfaces. Generally, an effective amount of such a composition is added to water, for example in a conventional fabric automatic washing machine, to form such aqueous liquor laundering solutions. The aqueous liquor so formed is then contacted, typically under agitation, with the fabrics to be laundered/treated therewith. An effective amount of the cleaning composition herein added to water to form aqueous liquors for washing can comprise amounts

sufficient to form from about 500 to 25,000 ppm, or from 500 to 15,000 ppm of composition in aqueous liquor, or from about 1,000 to 3,000 ppm of the cleaning compositions herein will be provided in aqueous liquor.

[0148] Typically, the aqueous liquor is formed by contacting the detergent with (wash) water in such an amount that the concentration of the cleaning composition in the aqueous liquor is from above 0.1 g/l to 5g/l, or from 1g/l, and to 4.5g/l, or to 4.0g/l, or to 3.5g/l, or to 3.0g/l, or to 2.5g/l, or even to 2.0g/l, or even to 1.5g/l. The method of laundering fabric or textile may be carried out in a top-loading or front-loading automatic washing machine, or can be used in a hand-wash laundry application. In these applications, the aqueous liquor formed and concentration of laundry detergent composition in the aqueous liquor is that of the main wash cycle. Any input of water during any optional rinsing step(s) is not included when determining the volume of the aqueous liquor.

[0149] The aqueous liquor may comprise 40 litres or less of water, or 30 litres or less, or 20 litres or less, or 10 litres or less, or 8 litres or less, or even 6 litres or less of water. The wash liquor may comprise from above 0 to 15 litres, or from 2 litres, and to 12 litres, or even to 8 litres of water. Typically from 0.01kg to 2kg of fabric per litre of aqueous liquor is dosed into said aqueous liquor. Typically from 0.01kg, or from 0.05kg, or from 0.07kg, or from 0.10kg, or from 0.15kg, or from 0.20kg, or from 0.25kg fabric per litre of aqueous liquor is dosed into said aqueous liquor. Optionally, 50g or less, or 45g or less, or 40g or less, or 35g or less, or 30g or less, or 25g or less, or 20g or less, or even 15g or less, or even 10g or less of the composition is contacted to water to form the aqueous liquor. Such compositions are typically employed at concentrations of from about 500 ppm to about 15,000 ppm in solution. When the wash solvent is water, the water temperature typically ranges from about 5 °C to about 90 °C and, when the situs comprises a fabric, the water to fabric ratio is typically from about 1:1 to about 30:1. Typically the aqueous liquor comprising the detergent of the invention has a pH of from 3 to 11.5.

[0150] In one aspect, such method comprises the steps of optionally washing and/or rinsing said surface or fabric, contacting said surface or fabric with any composition disclosed in this specification then optionally washing and/or rinsing said surface or fabric is disclosed, with an optional drying step.

[0151] Drying of such surfaces or fabrics may be accomplished by any one of the common means employed either in domestic or industrial settings: machine drying or open-air drying. The fabric may comprise any fabric capable of being laundered in normal consumer or institutional use conditions, and the invention is particularly suitable for synthetic textiles such as polyester and nylon and especially for treatment of mixed fabrics and/or fibres comprising synthetic and cellulosic fabrics and/or fibres. As examples of synthetic fabrics are polyester, nylon, these may be present in mixtures with cellulosic fibres, for example, polycotton fabrics. The solution typically has a pH of from 7 to 11, more usually 8 to 10.5. The compositions are typically employed at concentrations from 500 ppm to 5,000 ppm in solution. The water temperatures typically range from about 5 °C to about 90 °C. The water to fabric ratio is typically from about 1:1 to about 30:1.

Use of Water-Insoluble Plant Fiber

[0152] The present invention further relates to a use of water-insoluble plant fiber in a cleaning composition to enhance the stain-removal and/or malodor-reducing benefits of a mannanase enzyme.

Measurement Methods for Fibers

[0153] Sample preparation:

A) Cellulose fibers raw material: A cellulose fibers sample is prepared by adding 1% dry matter of cellulose fibers to water and activating it with a high pressure homogenizer (PANDA from GEA, 350 bars, 10 passes). Obtained sample is analyzed.

B) Composition comprising cellulose fibers:

[0154] The composition sample is centrifuged at 4,000 rpm for 10 minutes using a 5804 centrifuge from Eppendorf, in order to remove potential particles to avoid interference in the measurement of the fiber size. The clarified composition is then decanted as the supernatant. The water insoluble plant derived fibers present in the composition (supernatant) are redispersed in ethanol using an Ultra Turrax device from IKA, T25 S 25 N - 25 G - ST, at a speed of 21,000rpm for 10 minutes. Then, sample is centrifuged at 4,000 rpm for 10 minutes using a 5804 centrifuge from Eppendorf and supernatant is removed. Remaining cellulose fibers at the bottom are analyzed. Repeat the process as many times as needed to have enough amount for the analysis.

Measuring hydrodynamic diameter:

[0155] The instrument cell is cleaned and then filled with demineralised water. If the background has a laser intensity

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above 79%, the system is considered clean and the sample can be added to the vessel until the desired obscuration is achieved. Then ultrasounds are switched on for 30 seconds and once the sample is well dispersed, the measurement can start.

Then, the hydrodynamic diameter (volume weight mean [4,3]) is measured. The hydrodynamic diameter is the diameter of the equivalent sphere that has the same translational diffusion coefficient as the fiber being measured assuming a hydration layer surrounding the fiber.

Sampler selection: Hydro 2000MU

Sampler settings: Pump/stir speed: 2500rpm; Ultrasonics: 30 seconds

Material: Refractive Index of the material: 1.53; Dispersant used: demineralised water in an amount as needed; Particle shape: Irregular.

Measurement: Measurement cycles: 3 measurements per aliquot with a delay of 10 seconds; Measurement time: 10 seconds; Measurement snaps: 10,000; Background time: 10 seconds; o Background snaps: 10,000; Lower obscuration limit: 5; Upper obscuration limit: 15

Method for determining average cellulose fiber diameter:

[0156] The average water-insoluble plant derived fiber diameter can be determined directly from the cellulose fiber raw material or from the composition comprising cellulose fibers following sample preparation described above.

[0157] Water-insoluble plant derived fiber diameter is analysed using Atomic force microscopy (AFM). A 0,02% w/w cellulose fiber dispersion in demineralized water is prepared, and a drop of this dispersion is deposited onto freshly cleaved mica (highest grade VI Mica, 15x15mm - TED PELLA , INC., or equivalent). The sample is then allowed to dry in an oven at 40°C.

[0158] The mica sheet is mounted in an AFM (Nanosurf Flex AFM, ST Instruments or equivalent) and imaged in air under ambient conditions using a Si cantilever in dynamic mode with dynamic mode tip (ACTA -50 - APPNANO or equivalent). The image dimensions are 20 micron by 20 micron, and 256 points per line are captured.

[0159] The AFM image is opened using suitable AFM data analysis software (such as Mountainsmap SPM 7.3, ST Instruments, or equivalent). Each image is leveled line by line. One or more profiles are extracted crossing perpendicularly one or multiple fibers avoiding bundles of fibers, and from each profile, a distance measurement is performed to obtain the diameter of the fibers. Ten diameter measurements are performed per picture counting each fiber only once. Three sets of measurements (sample preparation, AFM measurement and image analysis) are made. The arithmetic mean of all fibers measured in all images is the Average water-insoluble plant derived fiber Diameter.

TEST METHODS

Enzymatic activity towards xyloglucan substrates

[0160] An enzyme is deemed to have activity towards xyloglucan if the pure enzyme has a specific activity of greater than 50000 XyloU/g according to the following assay at pH 7.5.

[0161] The xyloglucanase activity is measured using AZCL-xyloglucan from Megazyme, Ireland as substrate (blue substrate).

[0162] A solution of 0.2% of the blue substrate is suspended in a 0.1M phosphate buffer pH 7.5, 20°C under stirring in a 1.5ml Eppendorf tubes (0.75ml to each), 50 microlitres enzyme solution is added and they are incubated in an Eppendorf Thermomixer for 20 minutes at 40°C, with a mixing of 1200 rpm. After incubation the coloured solution is separated from the solid by 4 minutes centrifugation at 14,000 rpm and the absorbance of the supernatant is measured at 600nm in a 1cm cuvette using a spectrophotometer. One XyloU unit is defined as the amount of enzyme resulting in an absorbance of 0.24 in a 1cm cuvette at 600nm.

[0163] Only absorbance values between 0.1 and 0.8 are used to calculate the XyloU activity. If an absorbance value is measured outside this range, optimization of the starting enzyme concentration should be carried out accordingly.

Enzymatic activity towards amorphous cellulose substrates

[0164] An enzyme is deemed to have activity towards amorphous cellulose if the pure enzyme has a specific activity of greater than 20000 EBG/g according to the following assay at pH 7.5. Chemicals used as buffers and substrates were commercial products of at least reagent grade. Endoglucanase Activity Assay Materials:

- 0.1M phosphate buffer pH 7.5
- Cellazyme C tablets, supplied by Megazyme International, Ireland.

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- Glass microfiber filters, GF/C, 9cm diameter, supplied by Whatman.

Method:

- 5 **[0165]** In test tubes, mix 1ml pH 7.5 buffer and 5ml deionised water.
- [0166]** Add 100 microliter of the enzyme sample (or of dilutions of the enzyme sample with known weight:weight dilution factor). Add 1 Cellazyme C tablet into each tube, cap the tubes and mix on a vortex mixer for 10 seconds. Place the tubes in a thermostated water bath, temperature 40°C. After 15, 30 and 45 minutes, mix the contents of the tubes by inverting the tubes, and replace in the water bath. After 60 minutes, mix the contents of the tubes by inversion and then
- 10 **[0167]** filter through a GF/C filter. Collect the filtrate in a clean tube.
- [0167]** Measure Absorbance (Aenz) at 590nm, with a spectrophotometer. A blank value, Awater, is determined by adding 100µl water instead of 100 microliter enzyme dilution.

15 Calculate $\Delta A = A_{enz} - A_{water}$.

[0168] ΔA must be <0.5. If higher results are obtained, repeat with a different enzyme dilution factor. Determine DFO.1, where DFO.1 is the dilution factor needed to give $\Delta A = 0.1$.

- 20 **[0169]** Unit Definition: 1 Endo-Beta-Glucanase activity unit (1 EBG) is the amount of enzyme that gives $\Delta A = 0.10$, under the assay conditions specified above. Thus, for example, if a given enzyme sample, after dilution by a dilution factor of 100, gives $\Delta A = 0.10$, then the enzyme sample has an activity of 100 EBG/g.

EXAMPLES

- 25 **[0170]** The following are illustrative examples of cleaning compositions according to the present invention and are not intended to be limiting.

Examples 1-7: Heavy Duty Liquid laundry detergent compositions.

30 **[0171]**

Ingredients	1	2	3	4	5	6	7
	% weight						
AE _{1.8} S	6.77	5.16	1.36	1.30	-	-	-
AE ₃ S	-	-	-	-	0.45	-	-
LAS	0.86	2.06	2.72	0.68	0.95	1.56	3.55
HSAS	1.85	2.63	1.02	-	-	-	-
AE9	6.32	9.85	10.20	7.92			
AE8							35.45
AE7					8.40	12.44	
C ₁₂₋₁₄ dimethyl Amine Oxide	0.30	0.73	0.23	0.37	-	-	-
C ₁₂₋₁₈ Fatty Acid	0.80	1.90	0.60	0.99	1.20	-	15.00
Citric Acid	2.50	3.96	1.88	1.98	0.90	2.50	0.60
Optical Brightener 1	1.00	0.80	0.10	0.30	0.05	0.50	0.001
Optical Brightener 3	0.001	0.05	0.01	0.20	0.50	-	1.00
Sodium formate	1.60	0.09	1.20	0.04	1.60	1.20	0.20
DTI 1	0.32	0.05	-	0.60	0.10	0.60	0.01
DTI 2	0.32	0.10	0.60	0.60	0.05	0.40	0.20
Sodium hydroxide	2.30	3.80	1.70	1.90	1.70	2.50	2.30

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(continued)

	<u>Ingredients</u>	1	2	3	4	5	6	7
		% weight						
5	Monoethanolamine	1.40	1.49	1.00	0.70	-	-	-
	Diethylene glycol	5.50	-	4.10	-	-	-	-
	Chelant 1	0.15	0.15	0.11	0.07	0.50	0.11	0.80
10	4-formyl-phenylboronic acid	-	-	-	-	0.05	0.02	0.01
	Sodium tetraborate	1.43	1.50	1.10	0.75	-	1.07	-
	Ethanol	1.54	1.77	1.15	0.89	-	3.00	7.00
15	Polymer 1	0.10	-	-	-	-	-	2.00
	Polymer 2	0.30	0.33	0.23	0.17	-	-	-
	Polymer 3	-	-	-	-	-	-	0.80
	Polymer 4	0.80	0.81	0.60	0.40	1.00	1.00	-
20	1,2-Propanediol	-	6.60	-	3.30	0.50	2.00	8.00
	Structurant	0.10	-	-	-	-	- 0.10	0.10
	Perfume	1.60	1.10	1.00	0.80	0.90	1.50	1.60
25	Perfume encapsulate	0.10	0.05	0.01	0.02	0.10	0.05	0.10
	Protease	0.80	0.60	0.70	0.90	0.70	0.60	1.50
	Mannanase	0.07	-	-	0.06	0.04	0.045	0.10
	Amylase 1	0.30	-	0.30	0.10	-	0.40	0.10
30	Amylase 2	-	0.20	0.10	0.15	0.07	-	0.10
	Xyloglucanase	0.20	0.10	-	-	0.05	0.05	0.20
	Lipase	0.40	0.20	0.30	0.10	0.20	-	-
35	Polishing enzyme	-	0.04	-	-	-	0.004	-
	Mannanase of SEQ ID NO: 1 as active protein	0.05	0.03	0.01	0.03	0.03	0.003	0.003
	Dispersin B	-	-	-	0.05	0.03	0.001	0.001
40	Acid Violet 50	0.05	-	-	-	-	-	0.005
	Direct Violet 9	-	-	-	-	-	0.05	-
	Violet DD	-	0.035	0.02	0.037	0.04	-	-
	Water insoluble plant fiber	0.2	0.6	0.2	0.03	1.2	0.3	0.3
45	Water, dyes & minors	Balance						
	pH	8.2						

[0172] Based on total cleaning and/or treatment composition weight. Enzyme levels are reported as raw material.

50 Examples 8 to 18: Unit Dose Compositions.

[0173] These examples provide various formulations for unit dose laundry detergents. Compositions 8 to 12 comprise a single unit dose compartment. The film used to encapsulate the compositions is polyvinyl alcohol-based film.

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Ingredients	8	9	10	11	12
	% weight				
LAS	19.09	16.76	8.59	6.56	3.44
AE3S	1.91	0.74	0.18	0.46	0.07
AE7	14.00	17.50	26.33	28.08	31.59
Citric Acid	0.6	0.6	0.6	0.6	0.6
C12-15 Fatty Acid	14.8	14.8	14.8	14.8	14.8
Polymer 3	4.0	4.0	4.0	4.0	4.0
Chelant 2	1.2	1.2	1.2	1.2	1.2
Optical Brightener 1	0.20	0.25	0.01	0.01	0.50
Optical Brightener 2	0.20	-	0.25	0.03	0.01
Optical Brightener 3	0.18	0.09	0.30	0.01	-
DTI 1	0.10	-	0.20	0.01	0.05
DTI 2	-	0.10	0.20	0.25	0.05
Glycerol	6.1	6.1	6.1	6.1	6.1
Monoethanol amine	8.0	8.0	8.0	8.0	8.0
Tri-isopropanol amine	-	-	2.0	-	-
Tri-ethanol amine	-	2.0	-	-	-
Cumene sulfonate	-	-	-	-	2.0
Protease	0.80	0.60	0.07	1.00	1.50
Mannanase of SEQ ID NO: 1 as active protein	0.07	-	0.05	-	0.01
Amylase 1	0.20	0.11	0.30	0.50	0.05
Amylase 2	0.11	0.20	0.10	-	0.50
Polishing enzyme	0.005	0.05	-	-	-
Galactanase	0.005	0.05	0.005	0.010	0.005
Dispersin B	0.010	0.05	0.005	0.005	-
Cyclohexyl dimethanol	-	-	-	2.0	-
Acid violet 50	0.03	0.02			
Violet DD			0.01	0.05	0.02
Structurant	0.14	0.14	0.14	0.14	0.14
Perfume	1.9	1.9	1.9	1.9	1.9
Water insoluble plant fiber	0.02	0.3	0.02	0.03	0.3
Water and miscellaneous	To 100%				
pH	7.5-8.2				

[0174] Based on total cleaning and/or treatment composition weight. Enzyme levels are reported as raw material.

[0175] In the following examples the unit dose has three compartments, but similar compositions can be made with two, four or five compartments. The film used to encapsulate the compartments is polyvinyl alcohol.

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<u>Base compositions Ingredients</u>	13	14	15	16
	% weight			
HLAS	26.82	16.35	7.50	3.34
AE7	17.88	16.35	22.50	30.06
Citric Acid	0.5	0.7	0.6	0.5
C12-15 Fatty acid	16.4	6.0	11.0	13.0
Polymer 1	2.9	0.1	-	-
Polymer 3	1.1	5.1	2.5	4.2
Cationic cellulose polymer	-	-	0.3	0.5
Polymer 6	-	1.5	0.3	0.2
Chelant 2	1.1	2.0	0.6	1.5
Optical Brightener 1	0.20	0.25	0.01	0.005
Optical Brightener 3	0.18	0.09	0.30	0.005
DTI 1	0.1	-	0.2	-
DTI 2	-	0.1	0.2	-
Glycerol	5.3	5.0	5.0	4.2
Monoethanolamine	10.0	8.1	8.4	7.6
Polyethylene glycol	-	-	2.5	3.0
Potassium sulfite	0.2	0.3	0.5	0.7
Protease	0.80	0.60	0.40	0.80
Amylase 1	0.20	0.20	0.200	0.30
Polishing enzyme	-	-	0.005	0.005
Mannanase of SEQ ID NO: 2 as active protein	0.05	0.010	0.005	0.005
Dispersin B	-	0.010	0.010	0.010
MgCl ₂	0.2	0.2	0.1	0.3
Structurant	0.2	0.1	0.2	0.2
Acid Violet 50	0.04	0.03	0.05	0.03
Perfume / encapsulates	0.10	0.30	0.01	0.05
Water-insoluble plant fiber	0.2	0.03	0.4	2.0
Solvents and misc.	To 100%			
pH	7.0-8.2			

<u>Finishing compositions</u>	17			18		
Compartment	A	B	c	A	B	c
Volume of each compartment	40 ml	5 ml	5 ml	40 ml	5 ml	5 ml
<u>Ingredients</u>	Active material in Wt. %					
Perfume	1.6	1.6	1.6	1.6	1.6	1.6
Violet DD	0	0.006	0	0	0.004	-
TiO ₂	-	-	0.1	-		0.1

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(continued)

<u>Ingredients</u>	Active material in Wt. %					
Sodium Sulfite	0.4	0.4	0.4	0.3	0.3	0.3
Polymer 5	-			2	-	-
Hydrogenated castor oil	0.14	0.14	0.14	0.14	0.14	0.14
Base Composition 13, 14, 15 or 16	Add to 100%					

[0176] Based on total cleaning and/or treatment composition weight, enzyme levels are reported as raw material.

Examples 19 to 24:

[0177] Granular laundry detergent compositions for hand washing or washing machines, typically top-loading washing machines.

<u>Ingredient</u>	19	20	21	22	23	24
	% weight					
LAS	11.33	10.81	7.04	4.20	3.92	2.29
Quaternary ammonium	0.70	0.20	1.00	0.60	-	-
AE3S	0.51	0.49	0.32	-	0.08	0.10
AE7	8.36	11.50	12.54	11.20	16.00	21.51
Sodium Tripolyphosphate	5.0	-	4.0	9.0	2.0	-
Zeolite A	-	1.0	-	1.0	4.0	1.0
Sodium silicate 1.6R	7.0	5.0	2.0	3.0	3.0	5.0
Sodium carbonate	20.0	17.0	23.0	14.0	14.0	16.0
Polyacrylate MW 4500	1.0	0.6	1.0	1.0	1.5	1.0
Polymer 6	0.1	0.2	-	-	0.1	-
Carboxymethyl cellulose	1.0	0.3	1.0	1.0	1.0	1.0
Acid Violet 50	0.05	-	0.02	-	0.04	-
Violet DD	-	0.03	-	0.03	-	0.03
Protease 2	0.10	0.10	0.10	0.10	-	0.10
Amylase	0.03	-	0.03	0.03	0.03	0.03
Lipase	0.03	0.07	0.30	0.10	0.07	0.40
Polishing enzyme	0.002	-	0.05	-	0.02	-
Mannanase of SEQ ID NO: 1 as active protein	0.001	0.001	0.01	0.05	0.002	0.02
Dispersin B	0.001	0.001	0.05	-	0.001	-
Optical Brightener 1	0.200	0.001	0.300	0.650	0.050	0.001
Optical Brightener 2	0.060	-	0.650	0.180	0.200	0.060
Optical Brightener 3	0.100	0.060	0.050	-	0.030	0.300
Chelant 1	0.60	0.80	0.60	0.25	0.60	0.60
DTI 1	0.32	0.15	0.15	-	0.10	0.10
DTI 2	0.32	0.15	0.30	0.30	0.10	0.20
Sodium Percarbonate	-	5.2	0.1	-	-	-

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(continued)

<u>Ingredient</u>	19	20	21	22	23	24
	% weight					
Sodium Perborate	4.4	-	3.85	2.09	0.78	3.63
Nonanoyloxybenzensulfonate	1.9	0.0	1.66	0.0	0.33	0.75
Tetraacetylenehtylenediamine	0.58	1.2	0.51	0.0	0.015	0.28
Photobleach	0.0030	0.0	0.0012	0.0030	0.0021	-
S-ACMC	0.1	0.0	0.0	0.0	0.06	0.0
Water-insoluble plant fiber	1.5	3.2	2.4	2.2	2.8	0.9
Sulfate/Moisture	Balance					

Examples 25-30

[0178] Granular laundry detergent compositions typically for front-loading automatic washing machines.

<u>Ingredient</u>	25	26	27	28	29	30
	% weight					
LAS	6.08	5.05	4.27	3.24	2.30	1.09
AE3S	-	0.90	0.21	0.18	-	0.06
AS	0.34	-	-	-	-	-
AE7	4.28	5.95	6.72	7.98	9.20	10.35
Quaternary ammonium	0.5	-	-	0.3	-	-
Crystalline layered silicate	4.1	-	4.8	-	-	-
Zeolite A	5.0	-	2.0	-	2.0	2.0
Citric acid	3.0	4.0	3.0	4.0	2.5	3.0
Sodium carbonate	11.0	17.0	12.0	15.0	18.0	18.0
Sodium silicate 2R	0.08	-	0.11	-	-	-
Optical Brightener 1	-	0.25	0.05	0.01	0.10	0.02
Optical Brightener 2	-	-	0.25	0.20	0.01	0.08
Optical Brightener 3	-	0.06	0.04	0.15	-	0.05
DTI 1	0.08	-	0.04	-	0.10	0.01
DTI 2	0.08	-	0.04	0.10	0.10	0.02
Soil release agent	0.75	0.72	0.71	0.72	-	-
Acrylic /maleic acid copolymer	1.1	3.7	1.0	3.7	2.6	3.8
Carboxymethyl cellulose	0.2	1.4	0.2	1.4	1.0	0.5
Protease 3	0.20	0.20	0.30	0.15	0.12	0.13
Amylase 3	0.20	0.15	0.20	0.30	0.15	0.15
Lipase	0.05	0.15	0.10	-	-	-
Amylase 2	0.03	0.07	-	-	0.05	0.05
Cellulase 2	-	-	-	-	0.10	0.10
Polishing enzyme	0.003	0.005	0.020	-	-	-

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(continued)

	Ingredient	25	26	27	28	29	30
		% weight					
5	Mannanase of SEQ ID NO: 1 as active protein	0.002	0.010	0.020	0.020	0.010	0.003
	Dispersin B	0.002	0.010	0.020	0.020	0.010	0.002
	Tetraacetylenediamine	3.6	4.0	3.6	4.0	2.2	1.4
10	Sodium percarbonate	13.0	13.2	13.0	13.2	16.0	14.0
	Chelant 3	-	0.2	-	0.2	-	0.2
	Chelant 2	0.2	-	0.2	-	0.2	0.2
15	MgSO ₄	-	0.42	-	0.42	-	0.4
	Perfume	0.5	0.6	0.5	0.6	0.6	0.6
	Suds suppressor agglomerate	0.05	0.10	0.05	0.10	0.06	0.05
	Soap	0.45	0.45	0.45	0.45	-	-
20	Acid Violet 50	0.04	-	0.05	-	0.04	-
	Violet DD	-	0.04	-	0.05	-	0.04
	S-ACMC	0.01	0.01	-	0.01	-	-
25	Direct Violet 9 (active)	-	-	0.0001	0.0001	-	-
	Water-insoluble plant fiber	1.23	2.2	0.87	4.4	2.6	2.8
	Sulfate/ Water & Miscellaneous	Balance					
30	AE1.8S	is C ₁₂₋₁₅ alkyl ethoxy (1.8) sulfate					
	AE3S	is C ₁₂₋₁₅ alkyl ethoxy (3) sulfate					
	AE7	is C ₁₂₋₁₃ alcohol ethoxylate, with an average degree of ethoxylation of 7					
	AE8	is C ₁₂₋₁₃ alcohol ethoxylate, with an average degree of ethoxylation of 8					
	AE9	is C ₁₂₋₁₃ alcohol ethoxylate, with an average degree of ethoxylation of 9					
35	Amylase 1	is Stainzyme®, 15 mg active/g					
	Amylase 2	is Natalase®, 29 mg active/g					
	Amylase 3	is Stainzyme Plus®, 20 mg active/g,					
	AS	is C ₁₂₋₁₄ alkylsulfate					
	Cellulase 2	is Celluclean™, 15.6 mg active/g					
40	Xyloglucanase	is Whitezyme®, 20mg active/g					
	Chelant 1	is diethylene triamine pentaacetic acid					
	Chelant 2	is 1-hydroxyethane 1,1-diphosphonic acid					
	Chelant 3	is sodium salt of ethylenediamine-N,N'-disuccinic acid, (S,S) isomer (EDDS)					
	Dispersin B	is a glycoside hydrolase, reported as 1000mg active/g					
45	DTI 1	is poly(4-vinylpyridine-1-oxide) (such as Chromabond S-403E®),					
	DTI 2	is poly(1-vinylpyrrolidone-co-1-vinylimidazole) (such as Sokalan HP56®).					
	HSAS	is mid-branched alkyl sulfate as disclosed in US 6,020,303 and US6,060,443					
	LAS	is linear alkylbenzenesulfonate having an average aliphatic carbon chain length C ₉ -C ₁₅ (HLAS is acid form).					
50	Galactanase	is SEQ ID NO: 5, as active protein.					
	Lipase	is Lipex®, 18 mg active/g					
	Optical Brightener 1	is disodium 4,4'-bis[[4-anilino-6-morpholino-s-triazin-2-yl]-amino]-2,2'-stilbenedisulfonate					
	Optical Brightener 2	is disodium 4,4'-bis-(2-sulfostyryl)biphenyl (sodium salt)					
	Optical Brightener 3	is Optiblanc SPL10® from 3V Sigma					
55	Perfume encapsulate	is a core-shell melamine formaldehyde perfume microcapsules.					
	Photobleach	is a sulfonated zinc phthalocyanine					
	Polishing enzyme	is Para-nitrobenzyl esterase, reported as 1000mg active/g					
	Polymer 1	is bis((C ₂ H ₅ O)(C ₂ H ₄ O) _n)(CH ₃)-N ⁺ -C _x H _{2x} -N ⁺ -(CH ₃)-bis((C ₂ H ₅ O)(C ₂ H ₄ O) _n), wherein n =					

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	20-30, x = 3 to 8 or sulphated or sulfonated variants thereof
Polymer 2	is ethoxylated (EO ₁₅) tetraethylene pentamine
Polymer 3	is ethoxylated polyethylenimine
Polymer 4	is ethoxylated hexamethylene diamine
5 Polymer 5	is Acusol 305, provided by Rohm&Haas
Polymer 6	is a polyethylene glycol polymer grafted with vinyl acetate side chains, provided by BASF.
Protease	is Purafect Prime®, 40.6 mg active/g
Protease 2	is Savinase®, 32.89 mg active/g
Protease 3	is Purafect®, 84 mg active/g
10 Quaternary ammonium	is C ₁₂₋₁₄ Dimethylhydroxyethyl ammonium chloride
S-ACMC	is Reactive Blue 19 Azo-CM-Cellulose provided by Megazyme
Soil release agent	is Repel-o-tex® SF2
Structurant	is Hydrogenated Castor Oil
Violet DD	is a thiophene azo dye provided by Milliken
15 Water insoluble plant fiber	Water insoluble plant fiber in accordance with the present disclosure, for example Herbacel AQ+ Type N, supplied by Herbafood Ingredients GmbH, Werder, Germany and activated as a 2% aqueous slurry using a high pressure homogenizer PandaPlus from GEA (350 bars, 10 passes), then this slurry is added in the last step by using a Ultra-turrax with S 25 N - 18 G - ST Dispersing element from IKA.

[0179] The dimensions and values disclosed herein are not to be understood as being strictly limited to the exact numerical values recited. Instead, unless otherwise specified, each such dimension is intended to mean both the recited value and a functionally equivalent range surrounding that value. For example, a dimension disclosed as "40 mm" is intended to mean "about 40 mm."

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SEQUENCE LISTING

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Thr Gly Gln Gln Glu Leu His Ala His Gln Trp Val Glu Lys Asn Val
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Ala	Lys	Val	Pro	Ile	Leu	Phe	Arg	Pro	Leu	His	Glu	Pro	Asp	Gly	Ala
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Ile Cys Thr Ile Asp Asp Gly Ala Asp Lys Arg Arg Glu Glu Ser Leu
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Lys Gly Ile Pro Thr Lys Pro Gly Tyr Asp Arg Asp Glu Trp Pro Met
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Ala Val Cys Glu Glu Gly Gly Ala Gly Ala Asp Val Arg Tyr Val Thr
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Pro Ser Asp Asn Arg Gly Ala Gly Ser Trp Val Gly Asn Gln Met Ser
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Ser Tyr Pro Asp Gly Thr Arg Val Leu Phe Ile Val Gln
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Cys Thr Ile Glu Arg Ser Gly Ala Asp Lys Arg Arg Gln Glu Ser Leu
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Lys Gly Ile Pro Thr Lys Pro Gly Phe Asp Arg Asp Glu Trp Pro Met
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Ala Met Cys Glu Glu Gly Gly Lys Gly Ala Ser Val Arg Tyr Val Ser
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Ser Ser Asp Asn Arg Gly Ala Gly Ser Trp Val Gly Asn Arg Leu Ser

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	Ile	Leu	Cys	Lys	Gly	Ala	Pro	Asn	Val	Leu	Gln	Arg	Val	Asn	Glu	Lys
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Asn Cys Asn Ala Arg Glu Tyr Val Leu Lys Arg Asp Gly Glu Gly Val
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25

Gln Val Asn Asn Ala Cys Glu Ser Gln Ser Gly Thr Trp Ile Ser Pro
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Tyr Asp Asn Ala Ser Phe Thr Asn Ala Ser Ser Leu Asp Ile Asp His
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Thr Ala Gln Arg Glu Ala Leu Ala Asn Asp Val Ser Arg Pro Gln Leu
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Trp Ala Val Ser Ala Ser Ala Asn Arg Ser Lys Gly Asp Arg Ser Pro
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Asp Gln Trp Lys Pro Pro Leu Thr Ser Phe Tyr Cys Thr Tyr Ala Lys
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40	Pro	Glu	Leu	Asp	Ser	Pro	Asn	His	Met	Thr	Ala	Ile	Phe	Lys	Leu	Val
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		290					295					300					
40																	
	Leu	Leu	Leu	Asp	Phe	Thr	Trp	Leu	His	Pro	Thr	Ala	Trp	Val	Tyr	Trp	
	305					310					315					320	
	Gln	Ala	Ile	Asp	Gly	Ser	Gly	Trp	Gly	Leu	Ile	Val	Gly	Asp	Asn	Asp	
45					325					330					335		
	Gln	Leu	Thr	Leu	Ser	Ser	Ala	Ser	Thr	Lys	Tyr	Phe	Val	Leu	Ala	Gln	
50				340					345					350			
	Leu	Thr	Arg	His	Ile	Arg	Pro	Gly	Met	Gln	Ile	Leu	Thr	Thr	Pro	Asp	
			355					360					365				
55	Gly	Asn	Thr	Val	Ala	Ala	Tyr	Asp	Ser	Gly	Ser	Gln	Lys	Leu	Val	Ile	
		370					375					380					

Val Ala Ala Asn Trp Gly Ser Ala Gln Thr Ile Thr Phe Asp Leu Thr
385 390 395 400

Arg Ala Lys Thr Ala Gly Ser Asn Gly Ala Thr Val Pro Arg Trp Ser
405 410 415

Thr Gln Thr Ser Gly Gly Asp Gln Tyr Lys Ser Tyr Ser Asp Thr Lys
420 425 430

Ile Asn Asn Gly Lys Phe Ser Val Ser Phe Ser Thr Gly Gln Val Gln
435 440 445

Thr Phe Glu Ile Ser Gly Val Val Leu Lys
450 455

Claims

1. A cleaning composition comprising:

a mannanase enzyme comprising a polypeptide having mannan endo-1,4-beta-mannosidase activity (EC 3.2.1.78) that catalyzes the hydrolysis of 1,4-3-D-mannosidic linkages in mannans, galactomannans and/or glucomannans; and
from about 0.01% to about 5%, by weight of the cleaning composition, of water-insoluble plant fiber; and
optionally a particulate benefit agent.

2. A cleaning composition according to claim 1, wherein the mannanase enzyme is selected from Glycoside Hydrolase Family 26.

3. A cleaning composition according to claim 1 or claim 2, wherein the mannanase enzyme has at least 60% sequence identity to SEQ ID NO:1, and/or at least 81% sequence identity to SEQ ID NO: 2, and/or having at least 75% identity to SEQ ID NO: 3, and/or having at least 65% identity to SEQ ID NO: 4, and/or having at least 75% identity to SEQ ID NO: 5.

4. A cleaning composition according to any preceding claim, wherein the composition further comprises a β -N-acetylglucosaminidase enzyme from E.C. 3.2.1.52, preferably an enzyme having at least 70% identity to SEQ ID NO: 11.

5. A cleaning composition according to any preceding claim, wherein the water-insoluble plant fiber is selected from the group consisting of particulate cellulose material, activated citrus fiber, and mixtures thereof.

6. A cleaning composition according to claim 5, wherein the water-insoluble plant fiber comprises particulate cellulose material containing, by dry weight of the particulate cellulose material, at least 70% cellulose.

7. A cleaning composition according to claim 5 or claim 6, wherein the particulate cellulose material further comprises less than about 10% pectin, and at least about 3% hemicellulose.

8. A cleaning composition according to any of claims 5 to 7, wherein the water-insoluble plant fiber comprises particulate cellulose material having a volume-weighted median major particle dimension of from about 25 μ m to about 100 μ m, preferably from about 35 μ m to about 65 μ m, as measured by laser light diffractometry.

9. A cleaning composition according to any of claims 5 to 8, wherein the water-insoluble plant fiber comprises particulate cellulose material, wherein less than about 10% by dry weight of the cellulose material is in the form of nanofibrillated cellulose.

10. A cleaning composition according to any preceding claim, wherein the composition further comprises fabric shading agent and/or an additional enzyme selected from lipases, amylases, proteases, galactanases, pectate lyases, cellulases, cutinases, and mixtures thereof.

5 11. A method of cleaning a surface, preferably a textile, comprising mixing the cleaning composition according to any preceding claim with water to form an aqueous liquor and contacting a surface, preferably a textile, with the aqueous liquor in a laundering step.

10 12. A method of enhancing deposition of a particulate benefit agent comprising contacting a surface, preferably a textile with an aqueous liquor comprising a composition comprising a mannanase enzyme comprising a polypeptide having mannan endo-1,4-beta-mannosidase activity (EC 3.2.1 .78) that catalyzes the hydrolysis of 1 ,4-3-D-mannosidic linkages in mannans, galactomannans and/or glucomannans; a water-insoluble plant fiber and a particulate benefit agent in a textile treatment step, preferably a laundering step, and optionally rinsing and drying the textile.

15 13. A composition or method according to any preceding claim wherein the encapsulated benefit agent comprises a shell surrounding a core, the core comprising a benefit agent, preferably the benefit agent comprising perfume raw materials.

20 14. The use of a mannanase enzyme comprising a polypeptide having mannan endo-1,4-beta-mannosidase activity (EC 3.2.1 .78) that catalyzes the hydrolysis of 1 ,4-3-D-mannosidic linkages in mannans, galactomannans and/or glucomannans and a water-insoluble plant fiber in a cleaning composition to enhance stain-removal and/or malodor-reducing benefits.

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EUROPEAN SEARCH REPORT

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Place of search Munich		Date of completion of the search 12 January 2018	Examiner Vernier, Frédéric
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