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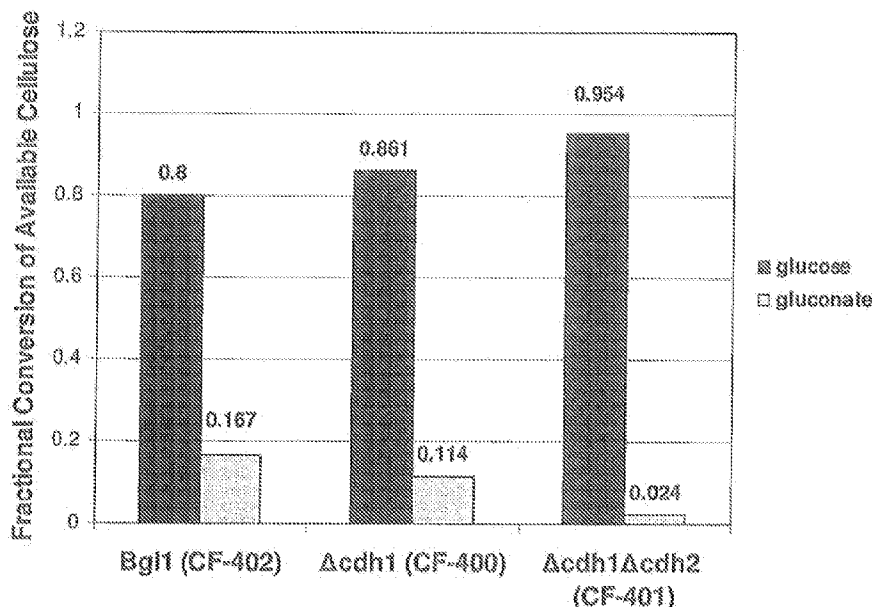
(72) Inventeurs/Inventors:
DHAWAN, ISH KUMAR, US;
BAIDYARROY, DIPNATH, US;
SHAW, ANDREW, US;
TANCHAK, OLEH, CA;
HILL, CHRISTOPHER, CA;
LIU, CHENGSONG, CA;
CHOKSHI, AMALA, CA;
SCOTT, BRIAN R., CA

(73) Propriétaire/Owner:
CODEXIS, INC., US

(74) Agent: SMART & BIGGAR LLP

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(54) Title: COMPOSITIONS AND METHODS FOR PRODUCTION OF FERMENTABLE SUGARS



(57) Abrégé/Abstract:

The present application provides genetically modified fungal organisms that produce enzyme mixtures exhibiting enhanced hydrolysis of cellulosic material to glucose, enzyme mixtures produced by the genetically modified fungal organisms, and processes for producing glucose from cellulose using such enzyme mixtures.

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(71) Applicant (for all designated States except US):
CODEXIS, INC. [US/US]; 200 Penobscot Drive, Redwood City, California 94063 (US).

(72) Inventors; and

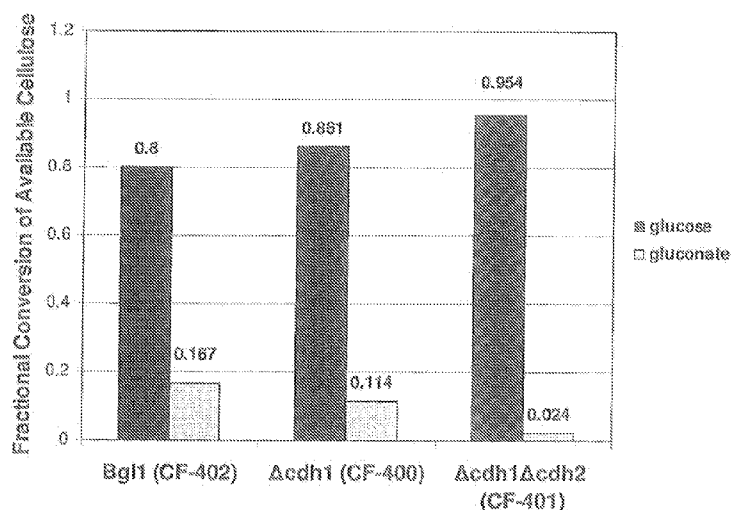
(75) Inventors/Applicants (for US only): **DHAWAN, Ish, Kumar** [US/US]; 200 Penobscot Drive, Redwood City, California 94063 (US). **BAIDYAROY, Dipnath** [IN/US]; 200 Penobscot Drive, Redwood City, California 94063 (US). **SHAW, Andrew** [US/US]; 200 Penobscot Drive, Redwood City, California 94063 (US). **TAN-CHAK, Oleh** [CA/CA]; 310 Hunt Club Road East, Ottawa, Ontario K1V 1C1 (CA). **HILL, Christopher** [CA/CA]; 310 Hunt Club Road East, Ottawa, Ontario K1V 1C1 (CA). **LIU, Chengsong** [CA/CA]; 310 Hunt Club Road East, Ottawa, Ontario K1V 1C1 (CA). **CHOKSHI, Amala** [CA/CA]; 310 Hunt Club Road East, Ottawa, Ontario K1V 1C1 (CA). **SCOTT, Brian, R.** [CA/CA]; 310 Hunt Club Road East, Ottawa, Ontario K1V 1C1 (CA).(74) Agents: **MACKNIGHT, Kamrin, T.** et al.; Codexis, Inc., 200 Penobscot Drive, Redwood City, California 94063 (US).

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Fig. 2



(57) Abstract: The present application provides genetically modified fungal organisms that produce enzyme mixtures exhibiting enhanced hydrolysis of cellulosic material to glucose, enzyme mixtures produced by the genetically modified fungal organisms, and processes for producing glucose from cellulose using such enzyme mixtures.

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COMPOSITIONS AND METHODS FOR PRODUCTION OF FERMENTABLE SUGARS

[0001] The present application claims priority to US Prov. Patent Appln. Ser. Nos. 61/409,186, 61/409,217, 61/409,472, and 61/409,480, all of which were filed on November 2, 2010.

FIELD OF THE INVENTION

[0002] The present invention provides compositions and methods for the production of fermentable sugars. In some embodiments, the present invention provides genetically modified fungal organisms. In some additional embodiments, the present invention provides enzymes that find use in enhancing hydrolysis of cellulosic material to fermentable sugars (*e.g.*, glucose), and methods for using the enzymes. In some further embodiments, the present invention provides enzyme mixtures useful for the hydrolysis of cellulosic materials.

BACKGROUND

[0003] Cellulose is a polymer of the simple sugar glucose linked by beta-1,4 glycosidic bonds. Many microorganisms produce enzymes that hydrolyze beta-linked glucans. These enzymes include endoglucanases, cellobiohydrolases, and beta-glucosidases. Endoglucanases digest the cellulose polymer at random locations, opening it to attack by cellobiohydrolases. Cellobiohydrolases sequentially release molecules of cellobiose from the ends of the cellulose polymer. Cellobiose is a water-soluble beta-1,4-linked dimer of glucose. Beta-glucosidases hydrolyze cellobiose to glucose.

[0004] The conversion of lignocellulosic feedstocks into ethanol has the advantages of the ready availability of large amounts of feedstock, the desirability of avoiding burning or land filling the materials, and lower overall greenhouse gas production. Wood, agricultural residues, herbaceous crops, and municipal solid wastes have been considered as feedstocks for ethanol production. These materials primarily consist of cellulose, hemicellulose, and lignin. Once the cellulose is converted to glucose, the glucose is easily fermented by yeast into ethanol.

[0005] Although progress has been made in increasing the efficiency of enzymatic degradation of lignocellulosic feedstocks, there remains a great need to improve yield of fermentable sugars using enzymatic processes.

SUMMARY OF THE INVENTION

[0006] The present invention provides genetically modified fungal organisms, as well as enzymes that enhance hydrolysis of cellulosic material to glucose, and methods for using the enzymes.

[0007] The present invention provides fungal cells that have been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is produced by the fungal cells. In some embodiments, the fungal cell is an Ascomycete belonging to the subdivision Pezizomycotina, and/or wherein the fungal cell is from the family Chaetomiaceae. In some embodiments, the fungal cell is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Neurospora*, *Sordaria*, *Podospora*, *Magnaporthe*, *Fusarium*, *Gibberella*, *Botryotinia*, *Humicola*, *Neosartorya*, *Pyrenophora*, *Phaeosphaeria*, *Sclerotinia*, *Chaetomium*, *Nectria*, *Verticillium*, *Corynascus*, *Acremonium*, *Ctenomyces*, *Chrysosporium*, *Scytalidium*, *Talaromyces*, *Thermoascus*, or *Aspergillus*. In some additional embodiments, the fungal cell is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Chrysosporium*, *Corynascus*, *Acremonium*, *Chaetomium*, *Ctenomyces*, *Scytalidium*, *Talaromyces*, or *Thermoascus*, while in some other embodiments, the fungal cell is *Sporotrichum thermophile*, *Sporotrichum cellulophilum*, *Thielavia heterothallica*, *Thielavia terrestris*, *Corynascus heterothallicus*, or *Myceliophthora thermophila*. In some embodiments, the fungal cell has been genetically modified to reduce the amount of endogenous glucose oxidase and/or cellobiose dehydrogenase that is produced by the fungal cell. In some additional embodiments, the fungal cell has been genetically modified to reduce the amount of endogenous glucose oxidase and/or cellobiose dehydrogenase that is produced by the fungal cell and to increase the production of at least one saccharide hydrolyzing enzyme. In some further embodiments, the fungal cell has been genetically modified to reduce the amount of endogenous glucose oxidase and/or cellobiose dehydrogenase that is produced by the fungal cell and to increase the production of at least one saccharide hydrolyzing enzyme, and wherein the fungal cell is a Basidiomycete

belonging to the class Agaricomycetes. In some embodiments, the Basidiomycete is a species of *Pleurotus*, *Peniophora*, *Trametes*, *Athelia*, *Sclerotium*, *Termitomyces*, *Flammulina*, *Coniophora*, *Ganoderma*, *Pycnoporus*, *Ceriporiopsis*, *Phanerochaete*, *Gloeophyllum*, *Hericium*, *Heterobasidion*, *Gelatoporia*, *Lepiota*, or *Irpex*. In some embodiments, the fungal cell has been genetically modified to reduce the amount of the endogenous glucose oxidase and/or cellobiose dehydrogenase that is secreted by the fungal cell. In some additional embodiments, the fungal cell has been genetically modified to disrupt the secretion signal peptide of the glucose and/or cellobiose oxidizing enzyme. In some further embodiments, the fungal cell has been genetically modified to reduce the amount of the endogenous glucose and/or cellobiose oxidizing enzyme that is expressed by the fungal cell. In still some additional embodiments, the fungal cell has been genetically modified to disrupt a translation initiation sequence in the transcript encoding the endogenous glucose and/or cellobiose oxidizing enzyme. In some additional embodiments, the fungal cell has been genetically modified to introduce a frameshift mutation in the transcript encoding the endogenous glucose and/or cellobiose oxidizing enzyme. In some further embodiments, the fungal cell has been genetically modified to reduce the transcription level of a gene encoding the endogenous glucose and/or cellobiose oxidizing enzyme. In some embodiments, the fungal cell has been genetically modified to disrupt the promoter of a gene encoding the endogenous glucose and/or cellobiose oxidizing enzyme. In still some additional embodiments, the fungal cell has been genetically modified to at least partially delete at least one gene encoding the endogenous glucose and/or cellobiose oxidizing enzyme. In some further embodiments, the fungal cell has been genetically modified to reduce the catalytic efficiency of the endogenous glucose and/or cellobiose oxidizing enzyme. In some additional embodiments, the fungal cell has been genetically modified to mutate one or more residues in an active site of the glucose and/or cellobiose oxidizing enzyme. In some further embodiments, the fungal cell has been genetically modified to mutate one or more residues in a heme binding domain of the glucose and/or cellobiose oxidizing enzyme. In some embodiments of the fungal cells provided herein, the glucose and/or cellobiose oxidizing enzyme is selected from cellobiose dehydrogenase (EC 1.1.99.18), glucose oxidase (EC 1.1.3.4), pyranose oxidase (EC 1.1.3.10), glucooligosaccharide oxidase (EC 1.1.99.B3), pyranose dehydrogenase (EC 1.1.99.29), and

glucose dehydrogenase (EC 1.1.99.10). In some additional embodiments, the glucose and/or cellobiose oxidizing enzyme comprises an amino acid sequence that is at least about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or about 99% identical to SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, and/or 16. In some further embodiments, the glucose and/or cellobiose oxidizing enzyme is cellobiose dehydrogenase (EC 1.1.99.18). In some embodiments, the fungal cell has been genetically modified to reduce the amount of glucose and/or cellobiose oxidizing enzyme activity of two or more endogenous glucose and/or cellobiose oxidizing enzymes that are produced by the fungal cell prior to genetic modification. In some further embodiments, the first of the two or more the glucose and/or cellobiose oxidizing enzymes comprises an amino acid sequence that is at least about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or about 99% identical to SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, and/or 16, and a second of the two or more the glucose and/or cellobiose oxidizing enzymes comprises an amino acid sequence that is at least about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or about 99% identical to SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14, and/or 16.

[0008] The present invention also provides enzyme mixtures comprising two or more cellulose hydrolyzing enzymes, wherein at least one of the two or more cellulose hydrolyzing enzymes is expressed by at least one of the fungal cells provided herein.

[0009] The present invention also provides enzyme mixtures comprising two or more cellulose hydrolyzing enzymes, wherein at least one of the two or more cellulose hydrolyzing enzymes is produced by a fungal cell that has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the fungal cell, and wherein the fungal cell is an Ascomycete belonging to the subdivision Pezizomycotina. In some embodiments, the fungal cell is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Corynascus*, *Acremonium*, *Chaetomium*, *Ctenomyces*, *Scytalidium*, *Talaromyces*, or *Thermoascus*.

[0010] The present invention also provides enzyme mixtures comprising two or more cellulose hydrolyzing enzymes, wherein at least one of the two or more cellulose hydrolyzing enzymes is produced by a fungal cell that has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the fungal cell and to increase the production of at least one saccharide hydrolyzing enzyme, wherein the fungal cell is a Basidiomycete belonging to the class Agaricomycetes.

[0011] In some embodiments, the enzyme mixtures are cell-free mixtures. In some additional embodiments, a substrate of the enzyme mixture comprises pretreated lignocellulose. In some further embodiments, the pretreated lignocellulose comprises lignocellulose treated by a treatment method selected from acid pretreatment, ammonium pretreatment, steam explosion and/or organic solvent extraction.

[0012] The present invention also provides enzyme mixtures comprising two or more cellulose hydrolyzing enzymes, wherein the fungal cellulase enzyme mixture is modified relative to a parental (or reference) enzyme mixture to be at least partially deficient in glucose and/or cellobiose oxidizing enzyme activity.

[0013] The present invention further provides enzyme mixtures comprising two or more cellulose hydrolyzing enzymes, at least one of the cellulose hydrolyzing enzymes being endogenous to a fungal cell, wherein the fungal cell is a Basidiomycete belonging to the class Agaricomycetes or an Ascomycete belonging to the subdivision Pezizomycotina and wherein the enzyme mixture is characterized in that, when the enzyme mixture is contacted with cellobiose and/or glucose, no more than about 10%, about 15% or about 20%, of the cellobiose and/or glucose is oxidized after 10 hours.

[0014] In some embodiments of the enzyme mixtures, the fungal cell has been genetically modified to reduce the amount of glucose and/or cellobiose oxidase enzyme activity that is secreted by the fungal cell. In some further embodiments, the enzyme mixture is a cell-free mixture. In some additional embodiments, the enzyme mixture comprises at least one beta-glucosidase. In some further embodiments, the enzyme mixture comprises at least one cellulase enzyme selected from endoglucanases (EGs), beta-glucosidases (BGLs), Type 1 cellobiohydrolases (CBH1s), Type 2 cellobiohydrolases (CBH2s), and /or glycoside hydrolase 61s (GH61s), and/or variants of the cellulase enzyme. In some embodiments, the enzyme mixture further comprises at least one cellobiose dehydrogenase. In some

embodiments, the celliobiose dehydrogenase is CDH1 and/or CDH2. In some additional embodiments, the enzyme mixture further comprises at least one cellulase enzyme and/or at least one additional enzyme. In some further embodiments, the enzyme mixture has been subjected to a purification process to selectively remove one or more glucose and/or cellobiose oxidizing enzymes from the enzyme mixture. In some embodiments, the purification process comprises selective precipitation to separate the glucose and/or cellobiose oxidizing enzymes from other enzymes present in the enzyme mixture. In some additional embodiments, the enzyme mixtures comprise at least one inhibitor of one or more glucose and/or cellobiose oxidizing enzymes.

[0015] The present invention also provides methods for generating cellobiose and/or glucose comprising contacting a cellulose substrate with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes to generate glucose and/or cellobiose, wherein at least one of the cellulose hydrolyzing enzymes is endogenous to a fungal cell that is an Ascomycete belonging to the subdivision Pezizomycotina, and wherein the enzyme mixture is characterized in that, when the enzyme mixture is contacted with cellobiose and/or glucose, no more than about 10%, about 15%, or about 20% of the cellobiose and/or glucose is oxidized after 10 hours. In some embodiments, the Ascomycete is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Neurospora*, *Sordaria*, *Podospora*, *Magnaporthe*, *Fusarium*, *Gibberella*, *Botryotinia*, *Humicola*, *Neosartorya*, *Pyrenophora*, *Phaeosphaeria*, *Sclerotinia*, *Chaetomium*, *Nectria*, *Verticillium*, or *Aspergillus*.

[0016] The present invention also provides methods for generating cellobiose and/or glucose comprising contacting a cellulose substrate with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes to generate glucose and/or cellobiose, wherein at least one of the cellulose hydrolyzing enzymes is endogenous to a fungal cell that is a Basidiomycete belonging to the class Agaricomycetes, and wherein the enzyme mixture is characterized in that, when the enzyme mixture is contacted with cellobiose and/or glucose, no more than about 10%, about 15% or about 20% of the cellobiose and/or glucose is oxidized after 10 hours. In some embodiments, the Basidiomycete is a species of *Pleurotus*, *Peniophora*, *Trametes*, *Athelia*, *Sclerotium*, *Termitomyces*, *Flammulina*, *Coniophora*, *Ganoderma*, *Pycnoporus*, *Ceriporiopsis*, *Phanerochaete*, *Gloeophyllum*, *Hericium*, *Heterobasidion*, *Gelatoporia*, *Lepiota*, or *Irpex*.

[0017] The present invention also provides methods for generating cellobiose and/or glucose comprising contacting a cellulose substrate with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes to generate glucose and/or cellobiose, wherein at least one of the cellulose hydrolyzing enzymes is endogenous to a fungal cell that is an Ascomycete belonging to the subdivision Pezizomycotina, and wherein, of the cellulose hydrolyzed by the enzyme mixture, at least about 80%, about 85%, or about 90% is present in the form of cellobiose and/or glucose. In some embodiments, the Ascomycete is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Neurospora*, *Sordaria*, *Podospora*, *Magnaporthe*, *Fusarium*, *Gibberella*, *Botryotinia*, *Humicola*, *Neosartorya*, *Pyrenophora*, *Phaeosphaeria*, *Sclerotinia*, *Chaetomium*, *Nectria*, *Verticillium*, or *Aspergillus*. In some embodiments, the Ascomycete is *Myceliophthora thermophila*, *Thielavia heterothallica* or *Sporotrichum thermophile*. In some embodiments, the fungal cell is *Myceliophthora thermophila*.

[0018] The present invention also provides methods for generating cellobiose and/or glucose comprising contacting a cellulose substrate with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes to generate glucose and/or cellobiose, wherein at least one of the cellulose hydrolyzing enzymes is endogenous to a fungal cell that is a Basidiomycete belonging to the class Agaricomycetes, and wherein, of the cellulose hydrolyzed by the enzyme mixture, at least about 80%, about 85%, or about 90% is present in the form of cellobiose and/or glucose. In some embodiments, the Basidiomycete is a species of *Pleurotus*, *Peniophora*, *Trametes*, *Athelia*, *Sclerotium*, *Termitomyces*, *Flammulina*, *Coniophora*, *Ganoderma*, *Pycnoporus*, *Ceriporiopsis*, *Phanerochaete*, *Gloeophyllum*, *Hericium*, *Heterobasidion*, *Gelatoporia*, *Lepiota*, or *Irpex*.

[0019] The present invention also provides methods for producing cellobiose and/or glucose from cellulose comprising treating a cellulose substrate with an enzyme mixture to generate glucose, wherein the enzyme mixture is modified relative to a secreted enzyme mixture from a reference (or parental) fungal cell to be at least partially deficient in glucose and/or cellobiose oxidizing enzyme activity. In some embodiments of the methods, the enzyme mixture is a cell-free mixture. In some additional embodiments, the cellulose substrate comprises pretreated lignocellulose. In some further embodiments, the pretreated lignocellulose comprises lignocellulose treated by a treatment method selected from acid

pretreatment, ammonium pretreatment, steam explosion and/or organic solvent extraction. In some further embodiments, the methods further comprise fermentation of the cellobiose and/or glucose to an end product. In some embodiments, the end product is at least one fuel alcohol and/or at least one precursor industrial chemical. In some additional embodiments, the fuel alcohol is ethanol or butanol. In some embodiments, the process for producing cellobiose and/or glucose from cellulose and said fermentation are conducted in a simultaneous saccharification and fermentation (SSF) process. In some further additional embodiments, the enzyme mixture is produced by a fungal cell that has been genetically modified to reduce the amount of one or more endogenous glucose and/or cellobiose oxidizing enzymes that is secreted by the fungal cell. In some embodiments, the enzyme mixture has been subjected to a purification process to selectively remove at least one glucose and/or cellobiose oxidizing enzyme from the enzyme mixture. In some further embodiments, the purification process comprises selective precipitation to separate the glucose and/or cellobiose oxidizing enzyme from other enzymes present in the enzyme mixture. In still some additional embodiments, the enzyme mixture comprises at least one inhibitor of the glucose and/or cellobiose oxidizing enzyme. In some embodiments, the inhibitor comprises a broad-spectrum oxidase inhibitor selected from sodium azide, potassium cyanide, a metal anion, and a combination thereof. In some embodiments, the inhibitor comprises a specific inhibitor of cellobiose dehydrogenase (EC 1.1.99.18) selected from cellobioimidazole, gentiobiose, lactobionol-1, 5-lactone, cellobionol-1, 5-lactone, tri-N-acetylchitotriose, methyl-beta-D cellobiosidase, 2,2-bipyridine, cytochrome C, and a combination thereof. In some embodiments, the method is a batch process, while in some other embodiments it is a continuous process, and in some further embodiments it is a fed-batch process and in still further embodiments, it is a combination of batch, continuous and/or fed-batch processes conducted in any order. In some embodiments, the method is conducted in a reaction volume of at least 10,000 liters, while in some other embodiments, the method is conducted in a reaction volume of at least 100,000 liters. In some embodiments, the enzyme mixture comprises at least one beta-glucosidase, while in some other embodiments, the enzyme mixture does not comprise a beta-glucosidase. In some embodiments, the enzyme mixture comprises at least one endoglucanase, while in some other embodiments, the enzyme mixture does not comprise an endoglucanase. In some

embodiments, the enzyme mixture comprises at least one cellulase enzyme selected from endoglucanases (EGs), beta-glucosidases (BGLs), Type 1 cellobiohydrolases (CBH1s), Type 2 cellobiohydrolases (CBH2s), and /or glycoside hydrolase 61s (GH61s), and/or variants of said cellulase enzyme.

[0020] The present invention also provides methods for generating glucose comprising contacting cellulose with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes, wherein at least one of the two or more cellulose hydrolyzing enzymes is produced by the fungal cells provided herein.

[0021] The present invention also provides methods for generating glucose comprising contacting cellulose with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes, wherein at least one of the two or more cellulose hydrolyzing enzymes is produced by a fungal cell that has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the fungal cell, wherein the fungal cell is an Ascomycete belonging to the subdivision Pezizomycotina. In some embodiments, the fungal cell is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Corynascus*, *Acremonium*, *Chaetomium*, *Ctenomyces*, *Scytalidium*, *Talaromyces*, or *Thermoascus*.

[0022] The present invention also provides methods for generating glucose comprising contacting cellulose with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes, wherein at least one of the two or more cellulose hydrolyzing enzymes is produced by a fungal cell that has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the fungal cell and to increase the production of at least one saccharide hydrolyzing enzyme, wherein the fungal cell is a Basidiomycete belonging to the class Agaricomycetes.

[0023] The present invention further provides methods for generating glucose comprising contacting cellulose with at least one enzyme mixture as provided herein. In some embodiments, the cellulose comprises pretreated lignocellulose. In some additional embodiments, the pretreated lignocellulose comprises lignocellulose treated by a treatment method selected from acid pretreatment, ammonium pretreatment, steam explosion and/or organic solvent extraction. In some additional embodiments, the enzyme mixture is a cell-free mixture. In some further embodiments, the methods further comprise fermentation of

the glucose to an end product. In some embodiments, the end product is a fuel alcohol or a precursor industrial chemical. In some embodiments, the fuel alcohol is ethanol or butanol.

[0024] The present invention further provides the fungal cells provided herein, as well as the enzyme mixtures provided herein, and the methods provided herein, further comprising a cellulose degrading enzyme that is heterologous to the fungal cell.

[0025] The present invention also provides fermentation media comprising at least one fungal cell provided herein.

[0026] The present invention also provides fermentation media comprising at least one enzyme mixture provided herein.

[0027] The present invention further provides fermentation media comprising at least one fungal cell and/or at least one enzyme mixture, as provided herein.

[0028] The present invention also provides methods of producing at least one cellulase, comprising at least one fungal cell provided herein, under conditions such that said at least one cellulase is produced. In some embodiments, the fungal cell is recombinant.

[0029] The present invention also provides compositions comprising at least one cellulase as provided herein.

DESCRIPTION OF THE DRAWINGS

[0030] Fig. 1 is a chart that shows the products of cellulose hydrolysis using enzyme mixtures obtained from strains CF-402, CF-403, and CF-401 as further described in Example 1 and Example 7. Dark bars represent measured glucose production. Light bars represent measured gluconate production. Numbers above horizontal bars indicate the sum of glucose and gluconate fractions.

[0031] Fig. 2 is a chart that shows the products of cellulose hydrolysis using enzyme mixtures produced by strain CF-400 (comprising a *cdh1* deletion); strain CF-401 (comprising the deletions of *cdh1* and *cdh2*) and strain CF-402 (comprising *cdh1* and *cdh2*), as further described in Example 8.

[0032] Figs. 3 and 4 provide the nucleotide and amino acid sequences of *M. thermophila* CDH1 and CDH2 (SEQ ID NOS:5-8).

[0033] Figs. 5 and 6 provide the nucleotide and amino acid sequences of *M. thermophila* GO1 and GO2 (SEQ ID NOS:1-4).

- [0034] Fig. 7 provides the nucleotide and amino acid sequences of *A. oryzae* pyranose oxidase (SEQ ID NOS:9-10).
- [0035] Fig. 8 provides the nucleotide and amino acid sequences of *A. strictum* glucooligosaccharide oxidase (SEQ ID NOS:11-12).
- [0036] Fig. 9 provides the nucleotide and amino acid sequences of *A. bisporus* pyranose dehydrogenase (SEQ ID NOS:13-14).
- [0037] Fig. 10 provides the nucleotide and amino acid sequences of *T. stipitatus* ATCC10500 glucose dehydrogenase (SEQ ID NOS:15-16).
- [0038] Fig. 11 provides a chart showing fractional recovery of available cellulose using an enzyme mixture containing cellobiose dehydrogenase activity. Dark bars represent glucose yield as measured using a horseradish peroxidase coupled enzymatic assay described in Example 1. Light bars represent expected glucose yield calculated using the IR method for determining cellulose conversion described in Example 9.
- [0039] Fig. 12A and 12B are HPLC chromatograms showing the effect of acid hydrolysis of cellotriose (Fig. 12A) or of cellulose hydrolysis products produced by an enzyme mixture containing cellobiose dehydrogenase (Fig. 12B) as described in Example 11.
- [0040] Fig. 13 provides an IR spectrum of cellulose hydrolysate obtained using enzyme mixtures lacking (Turbo) or containing (CF-402) cellobiose dehydrogenase activity. The vertical arrow indicates the carbonyl peak at 1715 cm^{-1} unique to the hydrolysate produced by the CF-402 enzyme mixture.
- [0041] Figs. 14A and 14B are HPLC chromatograms that identify an oxidized glucose product produced from glucose (Fig. 5A) or from cellulose hydrolysate using cellulase enzymes secreted by strain CF-402, as described in Example 13.

DESCRIPTION OF THE INVENTION

[0042] The present invention provides genetically modified fungal organisms, as well as enzymes that enhance hydrolysis of cellulosic material to glucose, and methods for using the enzymes.

[0043] Unless

otherwise indicated, the practice of the present invention involves conventional techniques commonly used in molecular biology, fermentation, microbiology, and related fields, which are known to those of skill in the art. Unless defined otherwise herein, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, some suitable methods and materials are described. Indeed, it is intended that the present invention not be limited to the particular methodology, protocols, and reagents described herein, as these may vary, depending upon the context in which they are used. The headings provided herein are not limitations of the various aspects or embodiments of the present invention.

[0044] Nonetheless, in order to facilitate understanding of the present invention, a number of terms are defined below. Numeric ranges are inclusive of the numbers defining the range. Thus, every numerical range disclosed herein is intended to encompass every narrower numerical range that falls within such broader numerical range, as if such narrower numerical ranges were all expressly written herein. It is also intended that every maximum (or minimum) numerical limitation disclosed herein includes every lower (or higher) numerical limitation, as if such lower (or higher) numerical limitations were expressly written herein.

[0045] As used herein, the term “comprising” and its cognates are used in their inclusive sense (*i.e.*, equivalent to the term “including” and its corresponding cognates).

[0046] As used herein and in the appended claims, the singular “a”, “an” and “the” include the plural reference unless the context clearly dictates otherwise. Thus, for example, reference to a “host cell” includes a plurality of such host cells.

[0047] Unless otherwise indicated, nucleic acids are written left to right in 5' to 3' orientation; amino acid sequences are written left to right in amino to carboxy orientation, respectively. The headings provided herein are not limitations of the various aspects or embodiments of the invention that can be had by reference to the specification as a whole. Accordingly, the terms defined below are more fully defined by reference to the specification as a whole.

[0048] As used herein, “substrate” refers to a substance or compound that is converted or meant to be converted into another compound by the action of an enzyme. The term includes not only a single compound but also combinations of compounds, such as solutions, mixtures and other materials which contain at least one substrate.

[0049] As used herein, “conversion” refers to the enzymatic transformation of a substrate to the corresponding product. “Percent conversion” refers to the percent of the substrate that is converted to the product within a period of time under specified conditions. Thus, for example, the “enzymatic activity” or “activity” of a cellobiose dehydrogenase (“CDH” or “cdh”) polypeptide can be expressed as “percent conversion” of the substrate to the product.

[0050] As used herein, “secreted activity” refers to enzymatic activity of glucose and/or cellobiose oxidizing enzymes produced by a fungal cell that is present in an extracellular environment. An extracellular environment can be, for example, an extracellular milieu such as a culture medium. The secreted activity is influenced by the total amount of glucose and/or cellobiose oxidizing enzyme secreted, and also is influenced by the catalytic efficiency of the secreted glucose and/or cellobiose oxidizing enzyme.

[0051] As used herein, a “reduction in catalytic efficiency” refers to a reduction in the activity of the glucose and/or cellobiose oxidizing enzyme, relative to unmodified glucose and/or cellobiose oxidizing enzyme, as measured using standard techniques, as provided herein or otherwise known in the art.

[0052] As used herein, the term “enzyme mixture” refers to a combination of at least two enzymes. In some embodiments, at least two enzymes are present in a composition. In some additional embodiments, the enzyme mixtures are present within a cell (*e.g.*, a fungal cell). In some embodiments, each or some of the enzymes present in an enzyme mixture are produced by different fungal cells and/or different fungal cultures. In some further embodiments, all of the enzymes present in an enzyme mixture are produced by the same cell. In some embodiments, the enzyme mixtures comprise cellulase enzymes, while in some additional embodiments, the enzyme mixtures comprise enzymes other than cellulases. In some embodiments, the enzyme mixtures comprise at least one cellulase and at least one enzyme other than a cellulase. In some embodiments, the enzyme mixtures comprise enzymes including, but not limited to endoxylanases (EC 3.2.1.8), beta-xylosidases (EC

3.2.1.37), alpha-L-arabinofuranosidases (EC 3.2.1.55), alpha-glucuronidases (EC 3.2.1.139), acetylxylanesterases (EC 3.1.1.72), feruloyl esterases (EC 3.1.1.73), coumaroyl esterases (EC 3.1.1.73), alpha-galactosidases (EC 3.2.1.22), beta-galactosidases (EC 3.2.1.23), beta-mannanases (EC 3.2.1.78), beta-mannosidases (EC 3.2.1.25), endo-polygalacturonases (EC 3.2.1.15), pectin methyl esterases (EC 3.1.1.11), endo-galactanases (EC 3.2.1.89), pectin acetyl esterases (EC 3.1.1.6), endo-pectin lyases (EC 4.2.2.10), pectate lyases (EC 4.2.2.2), alpha rhamnosidases (EC 3.2.1.40), exo-galacturonases (EC 3.2.1.82), exo-galacturonases (EC 3.2.1.67), exopolygalacturonate lyases (EC 4.2.2.9), rhamnogalacturonan endolyases EC (4.2.2.B3), rhamnogalacturonan acylesterases (EC 3.2.1.B11), rhamnogalacturonan galacturonohydrolases (EC 3.2.1.B11), endo-arabinanases (EC 3.2.1.99), laccases (EC 1.10.3.2), manganese-dependent peroxidases (EC 1.10.3.2), amylases (EC 3.2.1.1), glucoamylases (EC 3.2.1.3), lipases, lignin peroxidases (EC 1.11.1.14), and/or proteases.

[0053] In some additional embodiments, the present invention further provides enzyme mixtures comprising at least one expansin and/or expansin-like protein, such as a swollenin (*See e.g.*, Salheimo *et al.*, Eur. J. Biochem., 269:4202-4211 [2002]) and/or a swollenin-like protein. Expansins are implicated in loosening of the cell wall structure during plant cell growth. Expansins have been proposed to disrupt hydrogen bonding between cellulose and other cell wall polysaccharides without having hydrolytic activity. In this way, they are thought to allow the sliding of cellulose fibers and enlargement of the cell wall. Swollenin, an expansin-like protein contains an N-terminal Carbohydrate Binding Module Family 1 domain (CBD) and a C-terminal expansin-like domain. In some embodiments, an expansin-like protein and/or swollenin-like protein comprises one or both of such domains and/or disrupts the structure of cell walls (*e.g.*, disrupting cellulose structure), optionally without producing detectable amounts of reducing sugars. In some additional embodiments, the enzyme mixtures comprise at least one polypeptide product of a cellulose integrating protein, scaffoldin and/or a scaffoldin-like protein (*e.g.*, CipA or CipC from *Clostridium thermocellum* or *Clostridium cellulolyticum* respectively). In some additional embodiments, the enzyme mixtures comprise at least one cellulose induced protein and/or modulating protein (*e.g.*, as encoded by cip1 or cip2 gene and/or similar genes from *Trichoderma reesei*; *See e.g.*, Foreman *et al.*, J. Biol. Chem., 278:31988-31997 [2003]). In some additional embodiments, the enzyme mixtures comprise at least one member of each of

the classes of the polypeptides described above, several members of one polypeptide class, or any combination of these polypeptide classes to provide enzyme mixtures suitable for various uses.

[0054] Any combination of at least one two, three, four, five, or more than five enzymes and/or polypeptides find use in various enzyme mixtures provided herein. Indeed, it is not intended that the enzyme mixtures of the present invention be limited to any particular enzymes, polypeptides, nor combinations, as any suitable enzyme mixture finds use in the present invention.

[0055] As used herein, the term “saccharide” refers to any carbohydrate comprising monosaccharides (*e.g.*, glucose, ribose, fructose, galactose, etc.), disaccharides (*e.g.*, sucrose, lactose, maltose, cellobiose, trehalose, melibiose, etc.), oligosaccharides (*e.g.*, raffinose, stachyose, amylose, etc.), and polysaccharides (*e.g.*, starch, glycogen, cellulose, chitin, xylan, arabinoxylan, mannan, fucoidan, galactomannan, callose, laminarin, chrysolaminarin, amylopectin, dextran, dextrans, maltodextrins, inulin, oligofructose, polydextrose, etc.). The term encompasses simple carbohydrates, as well as complex carbohydrates. Indeed, it is not intended that the present invention be limited to any particular saccharide, as various saccharides and forms of saccharides find use in the present invention.

[0056] As used herein, the term “saccharide hydrolyzing enzyme” refers to any enzyme that hydrolyzes at least one saccharide.

[0057] As used herein, the terms “glucose oxidizing enzyme” and “cellobiose oxidizing enzyme” refer to enzymes that oxidize glucose and/or cellobiose. For example, glucose and/or cellobiose oxidizing enzymes include glucose oxidase (EC 1.1.3.4), cellobiose dehydrogenase (EC 1.1.99.18), pyranose oxidase (EC 1.1.3.10), glucooligosaccharide oxidase (EC 1.1.99.B3), pyranose dehydrogenase (EC 1.1.99.29), and glucose dehydrogenase (EC 1.1.99.10).

[0058] As used herein, the terms “glucose oxidase” and “GO” refer to an enzyme that is an oxido-reductase that catalyses the oxidation of β -D-glucose into D-glucono-1,5-lactone, which is a cyclic ester existing at a pH-dependent equilibrium in aqueous solution with gluconic acid or gluconate. Exemplary glucose oxidases fall into the enzyme classification (EC 1.1.3.4). In order to work as a catalyst, glucose oxidases typically utilize a co-substrate oxidant, such as flavin adenine dinucleotide (FAD). The enzyme is highly specific for β -D-

glucose. However, glucose oxidase also can demonstrate some lesser oxidase activity for substrates 2-deoxy-D-glucose, D-mannose and D-galactose (*See e.g.*, Bentley, Meth. Enzymol., 9:86 [1996]).

[0059] As used herein, the terms “cellobiose dehydrogenase” and “CDH” refer to a cellobiose:acceptor 1-oxidoreductase that catalyzes the conversion of cellobiose in the presence of an acceptor to cellobiono-1,5-lactone and a reduced acceptor. Examples of cellobiose dehydrogenases fall into the enzyme classification (E.C. 1.1.99.18). Typically 2,6-Dichloroindophenol can act as acceptor, as can iron, especially $\text{Fe}(\text{SCN})_3$, molecular oxygen, ubiquinone, or cytochrome C, and other polyphenolics, such as lignin. Substrates of the enzyme include cellobiose, cello-oligosaccharides, lactose, and D-glucosyl-1,4- β -D-mannose, glucose, maltose, mannobiose, thiocellobiose, galactosyl-mannose, xylobiose, xylose. Electron donors include beta-1-4 dihexoses with glucose or mannose at the reducing end, though alpha-1-4 hexosides, hexoses, pentoses, and beta-1-4 pentomers can act as substrates for at least some of these enzymes (*See e.g.*, Henriksson *et al*, Biochim. Biophys. Acta—Prot. Struct. Mol. Enzymol., 1383: 48-54 [1998]; and Schou *et al.*, Biochem. J., 330: 565-571 [1998]).

[0060] As used herein, the terms “oxidation”, “oxidize(d)” and the like as used herein refer to the enzymatic formation of one or more glucose or cellobiose oxidation products including, but not limited to, cellobionolactone, cellobionic acid, gluconolactone, gluconate and/or gluconic acid. When used in reference to a percentage of oxidized cellobiose and/or glucose, those percentages reflect a weight percent (w/w) relative to the initial amount of substrate. For example, when the enzyme mixture is contacted with cellobiose and/or glucose, the percentage of oxidized cellobiose and/or glucose reflects a weight percent (w/w) relative to the initial amount of cellobiose and/or glucose present in solution. Where the enzyme mixture is contacted with a cellulose substrate, the percentage of oxidized cellobiose and/or glucose reflects a weight percent (w/w) based on the maximum amount (wt %) of glucose that could be produced from the total hydrolyzed cellulose (*i.e.*, G_{max}).

[0061] As used herein, the terms “cellobiose dehydrogenase” and “CDH” refer to a cellobiose:acceptor 1-oxidoreductase that catalyzes the conversion of cellobiose in the presence of an acceptor to cellobiono-1,5-lactone and a reduced acceptor. Examples of cellobiose dehydrogenases are included in the enzyme classification (E.C. 1.1.99.18). In

some embodiments, the cellobiose dehydrogenase of interest in the present invention is CDH1, which is encoded by the *cdh1* gene. In some embodiments, the cellobiose dehydrogenase of interest in the present invention is CDH2, which is encoded by the *cdh2* gene. In some embodiments, both CDH1 and CDH2 are of interest.

[0062] As used herein, the terms “pyranose oxidase” and “PO” refer to an enzyme that catalyzes the conversion of D-glucose and O₂ to 2-dehydro-D-glucose and H₂O₂. Examples of pyranose oxidases fall into the enzyme classification (E.C. 1.1.3.10). The systematic name of this enzyme class is pyranose:oxygen 2-oxidoreductase. Other names in common use include glucose 2-oxidase, and pyranose-2-oxidase. Substrates of the enzyme include D-glucose, D-xylose, L-arabinose, L-sorbose, D-glucono-1,5-lactone, cellobiose and gentiobiose.

[0063] As used herein, the terms “glucooligosaccharide oxidase” and “GOOX” refer to an enzyme that catalyzes the oxidation of oligosaccharides with glucose on the reducing end and each sugar residue joined by an alpha- or beta-1,4 glucosidic bond. Examples of glucooligosaccharide oxidase fall into the enzyme classification (E.C. 1.1.99.B3). The systematic name of this enzyme class is carbohydrate:acceptor oxidoreductase. Substrates of the enzyme include maltose, lactose, cellobiose and maltose derivatives up to seven residues.

[0064] As used herein, the terms “pyranose dehydrogenase” and “PDH” refer to an enzyme that catalyzes the reaction of pyranose and an acceptor to yield 2-dehydropyranose (or 3-dehydropyranose or 2,3-didehydropyranose) and a reduced acceptor. PDH also catalyzes the reaction of a pyranoside and an acceptor to yield a 3-dehydropyranoside (or 3,4-didehydropyranoside) and a reduced acceptor. Examples of pyranose dehydrogenases fall into the enzyme classification (E.C. 1.1.99.29). The systematic name of this enzyme class is pyranose:acceptor oxidoreductase. Other names in common use include pyranose 2,3-dehydrogenase. PDH utilizes FAD as a cofactor. A number of aldoses and ketoses in pyranose form, as well as glycosides, gluco-oligosaccharides, sucrose and lactose can act as a donor. 1,4-Benzoquinone or ferricenium ion (ferrocene oxidized by removal of one electron) can serve as acceptor. Unlike EC 1.1.3.10 (pyranose oxidase), pyranose dehydrogenase does not interact with O₂ and exhibits extremely broad substrate tolerance with variable regioselectivity (C-3, C-2 or C-3 + C-2 or C-3 + C-4) for (di)oxidation of different sugars. D-Glucose is exclusively or preferentially oxidized at C-3 (depending on the enzyme source),

but can also be oxidized at C-2 + C-3. Pyranose dehydrogenase also acts on 1->4-alpha- and 1->4-beta-gluco-oligosaccharides, non-reducing gluco-oligosaccharides and L-arabinose, which are not substrates of EC 1.1.3.10. Sugars are oxidized by pyranose dehydrogenase in their pyranose but not in their furanose form.

[0065] As used herein, the terms “glucose dehydrogenase” and “GDH” refer to an enzyme that catalyzes the reaction of D-glucose and an acceptor to yield D-glucono-1,5-lactone and a reduced acceptor. Examples of glucose dehydrogenase fall into the enzyme classification (E.C. 1.1.99.10). The systematic name of this enzyme class is D-glucose:acceptor 1-oxidoreductase. GDH utilizes FAD as a cofactor.

[0066] As used herein, the term “cellulase” refers to any enzyme that is capable of degrading cellulose. Thus, the term encompasses enzymes capable of hydrolyzing cellulose (β -1,4-glucan or β -D-glucosidic linkages) to shorter cellulose chains, oligosaccharides, cellobiose and/or glucose. “Cellulases” are divided into three sub-categories of enzymes: 1,4- β -D-glucan glucanohydrolase (“endoglucanase” or “EG”); 1,4- β -D-glucan cellobiohydrolase (“exoglucanase,” “cellobiohydrolase,” or “CBH”); and β -D-glucoside-glucanohydrolase (“ β -glucosidase,” “cellobiase,” “BG,” or “BGL”). These enzymes act in concert to catalyze the hydrolysis of cellulose-containing substrates. Endoglucanases break internal bonds and disrupt the crystalline structure of cellulose, exposing individual cellulose polysaccharide chains (“glucans”). Cellobiohydrolases incrementally shorten the glucan molecules, releasing mainly cellobiose units (a water-soluble β -1,4-linked dimer of glucose) as well as glucose, cellotriose, and cellotetrose. Beta-glucosidases split the cellobiose into glucose monomers.

[0067] Cellulases often comprise a mixture of different types of cellulolytic enzymes (endoglucanases and cellobiohydrolases) that act synergistically to break down the cellulose to soluble di- or oligosaccharides such as cellobiose, which are then further hydrolyzed to glucose by beta-glucosidase. Cellulase enzymes are produced by a wide variety of microorganisms. Cellulases (and hemicellulases) from filamentous fungi and some bacteria are widely exploited for many industrial applications that involve processing of natural fibers to sugars.

[0068] As used herein, a “cellulase-producing fungal cell” is a fungal cell that produces at least one cellulase enzyme (*i.e.*, “cellulose hydrolyzing enzyme”). In some

embodiments, the cellulase-producing fungal cells provided herein express and secrete a mixture of cellulose hydrolyzing enzymes. As used herein, the terms “cellulose hydrolyzing enzyme,” “cellulolytic enzyme,” and like terms refer to an enzyme that acts in the process of breaking down cellulose to soluble di- or oligosaccharides such as cellobiose, which are then further hydrolyzed to glucose by beta-glucosidase. A mixture of cellulose hydrolyzing enzymes is also referred to herein as “cellulases,” a “cellulase-containing mixture,” and/or a “cellulase mixture.”

[0069] As used herein, the terms "endoglucanase" and "EG" refer to a category of cellulases (EC 3.2.1.4) that catalyze the hydrolysis of internal β -1,4 glucosidic bonds of cellulose. The term "endoglucanase" is further defined herein as an endo-1,4-(1,3;1,4)-beta-D-glucan 4-glucanohydrolase (E.C. 3.2.1.4), which catalyses endohydrolysis of 1,4-beta-D-glycosidic linkages in cellulose, cellulose derivatives (such as carboxymethyl cellulose and hydroxyethyl cellulose), lichenan, beta-1,4 bonds in mixed beta-1,3 glucans such as cereal beta-D-glucans or xyloglucans, and other plant material containing cellulosic components. Endoglucanase activity can be determined based on a reduction in substrate viscosity or increase in reducing ends determined by a reducing sugar assay (*See e.g.*, Zhang *et al.*, Biotechnol. Adv., 24:452-481 [2006]). For purposes of the present invention, endoglucanase activity is determined using carboxymethyl cellulose (CMC) hydrolysis (*See e.g.*, Ghose, Pur. Appl. Chem., 59:257-268 [1987]).

[0070] As used herein, "EG1" refers to a carbohydrate active enzyme expressed from a nucleic acid sequence coding for a glycohydrolase (GH) Family 7 catalytic domain classified under EC 3.2.1.4 or any protein, polypeptide or catalytically active fragment thereof. In some embodiments, the EG1 is functionally linked to a carbohydrate binding module (CBM), such as a Family 1 cellulose binding domain.

[0071] As used herein, the term "EG2" refers to a carbohydrate active enzyme expressed from a nucleic acid sequence coding for a glycohydrolase (GH) Family 5 catalytic domain classified under EC 3.2.1.4 or any protein, polypeptide or catalytically active fragment thereof. In some embodiments, the EG2 is functionally linked to a carbohydrate binding module (CBM), such as a Family 1 cellulose binding domain.

[0072] As used herein, the term “EG3” refers to a carbohydrate active enzyme expressed from a nucleic acid sequence coding for a glycohydrolase (GH) Family 12

catalytic domain classified under EC 3.2.1.4 or any protein, polypeptide or catalytically active fragment thereof. In some embodiments, the EG3 is functionally linked to a carbohydrate binding module (CBM), such as a Family 1 cellulose binding domain.

[0073] As used herein, the term “EG4” refers to a carbohydrate active enzyme expressed from a nucleic acid sequence coding for a glycohydrolase (GH) Family 61 catalytic domain classified under EC 3.2.1.4 or any protein, polypeptide or fragment thereof. In some embodiments, the EG4 is functionally linked to a carbohydrate binding module (CBM), such as a Family 1 cellulose binding domain.

[0074] As used herein, the term “EG5” refers to a carbohydrate active enzyme expressed from a nucleic acid sequence coding for a glycohydrolase (GH) Family 45 catalytic domain classified under EC 3.2.1.4 or any protein, polypeptide or fragment thereof. In some embodiments, the EG5 is functionally linked to a carbohydrate binding module (CBM), such as a Family 1 cellulose binding domain.

[0075] As used herein, the term “EG6” refers to a carbohydrate active enzyme expressed from a nucleic acid sequence coding for a glycohydrolase (GH) Family 6 catalytic domain classified under EC 3.2.1.4 or any protein, polypeptide or fragment thereof. In some embodiments, the EG6 is functionally linked to a carbohydrate binding module (CBM), such as a Family 1 cellulose binding domain.

[0076] As used herein, the terms "cellobiohydrolase" and "CBH" refer to a category of cellulases (EC 3.2.1.91) that hydrolyze glycosidic bonds in cellulose. The term "cellobiohydrolase" is further defined herein as a 1,4-beta-D-glucan cellobiohydrolase (E.C. 3.2.1.91), which catalyzes the hydrolysis of 1,4-beta-D-glucosidic linkages in cellulose, celooligosaccharides, or any beta-1,4-linked glucose containing polymer, releasing cellobiose from the reducing or non-reducing ends of the chain (*See e.g.*, Teeri, Tr. Biotechnol., 15:160-167 [1997]; and Teeri *et al.*, Biochem. Soc. Trans., 26:173-178 [1998]). In some embodiments, cellobiohydrolase activity is determined using a fluorescent disaccharide derivative 4-methylumbelliferyl-.beta.-D-lactoside (*See e.g.*, van Tilbeurgh *et al.*, FEBS Lett., 149:152-156 [1982]; and van Tilbeurgh and Claeysens, FEBS Lett., 187:283-288 [1985]).

[0077] As used herein, the terms "CBH1" and “type 1 cellobiohydrolase” refer to a carbohydrate active enzyme expressed from a nucleic acid sequence coding for a

glycohydrolase (GH) Family 7 catalytic domain classified under EC 3.2.1.91 or any protein, polypeptide or catalytically active fragment thereof. In some embodiments, the CBH1 is functionally linked to a carbohydrate binding module (CBM), such as a Family 1 cellulose binding domain.

[0078] As used herein, the terms "CBH2" and "type 2 cellobiohydrolase" refer to a carbohydrate active enzyme expressed from a nucleic sequence coding for a glycohydrolase (GH) Family 6 catalytic domain classified under EC 3.2.1.91 or any protein, polypeptide or catalytically active fragment thereof. Type 2 cellobiohydrolases are also commonly referred to as "the Cel6 family." In some embodiments, the CBH2 is functionally linked to a carbohydrate binding module (CBM), such as a Family 1 cellulose binding domain.

[0079] As used herein, the terms "beta-glucosidase," "cellobiase," and "BGL" refers to a category of cellulases (EC 3.2.1.21) that catalyze the hydrolysis of cellobiose to glucose. The term "beta-glucosidase" is further defined herein as a beta-D-glucoside glucohydrolase (E.C. 3.2.1.21), which catalyzes the hydrolysis of terminal non-reducing beta-D-glucose residues with the release of beta-D-glucose. Beta-glucosidase activity can be determined using any suitable method (*See e.g.*, J. Basic Microbiol., 42: 55-66 [2002]). One unit of beta-glucosidase activity is defined as 1.0 pmole of p-nitrophenol produced per minute at 40 °C, pH 5 from 1 mM p-nitrophenyl-beta-D-glucopyranoside as substrate in 100 mM sodium citrate containing 0.01% TWEEN® 20.

[0080] As used herein, the term "glycoside hydrolase 61" and "GH61" refers to a category of cellulases that enhance cellulose hydrolysis when used in conjunction with one or more additional cellulases. The GH61 family of cellulases is described, for example, in the Carbohydrate Active Enzymes (CAZY) database (*See e.g.*, Harris *et al.*, Biochem., 49(15):3305-16 [2010]).

[0081] A "hemicellulase" as used herein, refers to a polypeptide that can catalyze hydrolysis of hemicellulose into small polysaccharides such as oligosaccharides, or monomeric saccharides. Hemicelluloses include xylan, glucuronoxylan, arabinoxylan, glucomannan and xyloglucan. Hemicellulases include, for example, the following: endoxylanases, beta-xylosidases, alpha-L-arabinofuranosidases, alpha -D-glucuronidases, feruloyl esterases, coumaroyl esterases, alpha -galactosidases, beta-galactosidases, beta-mannanases, and beta-mannosidases.

[0082] As used herein, the terms "xylan degrading activity" and "xylanolytic activity" are defined herein as a biological activity that hydrolyzes xylan-containing material. The two basic approaches for measuring xylanolytic activity include: (1) measuring the total xylanolytic activity, and (2) measuring the individual xylanolytic activities (endoxylanases, beta-xylosidases, arabinofuranosidases, alpha-glucuronidases, acetylxylan esterases, feruloyl esterases, and alpha-glucuronyl esterases) (*See e.g.*, Biely and Puchard, J. Sci. Food Agr. 86:1636-1647 [2006]; Spanikova and Biely, FEBS Lett., 580:4597-4601 [2006]; and Herrmann *et al.*, Biochem. J., 321:375-381 [1997]).

[0083] Total xylan degrading activity can be measured by determining the reducing sugars formed from various types of xylan, including oat spelt, beechwood, and larchwood xylans, or by photometric determination of dyed xylan fragments released from various covalently dyed xylans. A common total xylanolytic activity assay is based on production of reducing sugars from polymeric 4-O-methyl glucuronoxylan (*See e.g.*, Bailey *et al.*, J. Biotechnol., 23:257-270 [1992]). In some embodiments, xylan degrading activity is determined by measuring the increase in hydrolysis of birchwood xylan (Sigma Chemical Co., Inc., St. Louis, Mo., USA) by xylan-degrading enzyme(s) under the following typical conditions: 1 mL reactions, 5 mg/mL substrate (total solids), 5 mg of xylanolytic protein/g of substrate, 50 mM sodium acetate pH 5, 50°C, 24 hours, sugar analysis using p-hydroxybenzoic acid hydrazide (PHBAH) assay (*See e.g.*, Lever, Anal. Biochem., 47:273-279 [1972]).

[0084] As used herein the term "xylanase activity" refers to a 1,4-beta-D-xylan-xylohydrolase activity (E.C. 3.2.1.8) that catalyzes the endo-hydrolysis of 1,4-beta-D-xylosidic linkages in xylans. In some embodiments, xylanase activity is determined using birchwood xylan as substrate. One unit of xylanase activity is defined as 1.0 μ mole of reducing sugar (measured in glucose equivalents; *See e.g.*, Lever, Anal. Biochem., 47:273-279 [1972]) produced per minute during the initial period of hydrolysis at 50°C, pH 5 from 2 g of birchwood xylan per liter as substrate in 50 mM sodium acetate containing 0.01% TWEEN® 20.

[0085] As used herein, the term "beta-xylosidase activity" refers to a beta-D-xyloside xylohydrolase (E.C. 3.2.1.37) that catalyzes the exo-hydrolysis of short beta (1→4)-xylooligosaccharides, to remove successive D-xylose residues from the non-reducing

termini. In some embodiments of the present invention, one unit of beta-xylosidase activity is defined as 1.0 μ mole of p-nitrophenol produced per minute at 40°C, pH 5 from 1 mM p-nitrophenyl-beta-D-xyloside as substrate in 100 mM sodium citrate containing 0.01% TWEEN® 20.

[0086] As used herein, the term "acetylxyylan esterase activity" refers to a carboxylesterase activity (EC 3.1.1.72) that catalyses the hydrolysis of acetyl groups from polymeric xylan, acetylated xylose, acetylated glucose, alpha-naphthyl acetate, and p-nitrophenyl acetate. In some embodiments of the present invention, acetylxyylan esterase activity is determined using 0.5 mM p-nitrophenylacetate as substrate in 50 mM sodium acetate pH 5.0 containing 0.01% TWEEN® 20. One unit of acetylxyylan esterase activity is defined as the amount of enzyme capable of releasing 1 pmole of p-nitrophenolate anion per minute at pH 5, 25°C.

[0087] As used herein, the term "feruloyl esterase activity" refers to a 4-hydroxy-3-methoxycinnamoyl-sugar hydrolase activity (EC 3.1.1.73) that catalyzes the hydrolysis of the 4-hydroxy-3-methoxycinnamoyl (feruloyl) group from an esterified sugar, which is usually arabinose in "natural" substrates, to produce ferulate (4-hydroxy-3-methoxycinnamate). Feruloyl esterase is also known as ferulic acid esterase, hydroxycinnamoyl esterase, FAE-III, cinnamoyl ester hydrolase, FAEA, cinnAE, FAE-I, or FAE-II. In some embodiments of the present invention, feruloyl esterase activity is determined using 0.5 mM p-nitrophenylferulate as substrate in 50 mM sodium acetate pH 5.0. One unit of feruloyl esterase activity equals the amount of enzyme capable of releasing 1 μ mole of p-nitrophenolate anion per minute at pH 5, 25 °C.

[0088] As used herein, the term "alpha-glucuronidase activity" refers to an alpha-D-glucosiduronate glucuronohydrolase activity (EC 3.2.1.139) that catalyzes the hydrolysis of an alpha-D-glucuronoside to D-glucuronate and an alcohol. One unit of alpha-glucuronidase activity equals the amount of enzyme capable of releasing 1 pmole of glucuronic or 4-O-methylglucuronic acid per minute at pH 5, 40 °C (*See e.g., de Vries, J. Bacteriol., 180:243-249 [1998]*).

[0089] As used herein the term "alpha-L-arabinofuranosidase activity" refers to an alpha-L-arabinofuranoside arabinofuranohydrolase activity (EC 3.2.1.55) that catalyzes the hydrolysis of terminal non-reducing alpha-L-arabinofuranoside residues in alpha-L-

arabinosides. The enzyme activity acts on alpha-L-arabinofuranosides, alpha-L-arabinans containing (1,3)- and/or (1,5)-linkages, arabinoxylans, and arabinogalactans. Alpha-L-arabinofuranosidase is also known as arabinosidase, alpha-arabinosidase, alpha-L-arabinosidase, alpha-arabinofuranosidase, arabinofuranosidase, polysaccharide alpha-L-arabinofuranosidase, alpha-L-arabinofuranoside hydrolase, L-arabinosidase and alpha-L-arabinanase. For purposes of the present invention, alpha-L-arabinofuranosidase activity is determined using 5 mg of medium viscosity wheat arabinoxylan (Megazyme International Ireland, Ltd., Bray, Co. Wicklow, Ireland) per mL of 100 mM sodium acetate pH 5 in a total volume of 200 μ L for 30 minutes at 40°C followed by arabinose analysis by AMINEX®. HPX-87H column chromatography (Bio-Rad Laboratories, Inc., Hercules, Calif., USA).

[0090] Enzymatic lignin depolymerization can be accomplished by lignin peroxidases, manganese peroxidases, laccases and cellobiose dehydrogenases (CDH), often working in synergy. These extracellular enzymes, essential for lignin degradation, are often referred to as “lignin-modifying enzymes” or “LMEs.” Three of these enzymes comprise two glycosylated heme-containing peroxidases: lignin peroxidase (LiP); Mn-dependent peroxidase (MnP); and, a copper-containing phenoloxidase laccase (LCC). Although the details of the reaction scheme of lignin biodegradation are not fully understood to date, without being bound by theory, it is suggested that these enzymes employ free radicals for depolymerization reactions.

[0091] As used herein, the term “laccase” refers to the copper containing oxidase enzymes that are found in many plants, fungi and microorganisms. Laccases are enzymatically active on phenols and similar molecules and perform a one electron oxidation. Laccases can be polymeric and the enzymatically active form can be a dimer or trimer.

[0092] As used herein, the term “Mn-dependent peroxidase” refers to peroxidases that require Mn. The enzymatic activity of Mn-dependent peroxidase (MnP) is dependent on Mn^{2+} . Without being bound by theory, it has been suggested that the main role of this enzyme is to oxidize Mn^{2+} to Mn^{3+} (See e.g., Glenn *et al.* Arch. Biochem. Biophys., 251:688-696 [1986]). Subsequently, phenolic substrates are oxidized by the Mn^{3+} generated.

[0093] As used herein, the term “lignin peroxidase” refers to an extracellular heme that catalyses the oxidative depolymerization of dilute solutions of polymeric lignin *in vitro*. Some of the substrates of LiP, most notably 3,4-dimethoxybenzyl alcohol (veratryl alcohol,

VA), are active redox compounds that have been shown to act as redox mediators. VA is a secondary metabolite produced at the same time as LiP by ligninolytic cultures of *P. chrysosporium* and without being bound by theory, has been proposed to function as a physiological redox mediator in the LiP-catalysed oxidation of lignin in vivo (*See e.g.*, Harvey *et al.*, FEBS Lett. 195:242–246 [1986]).

[0094] As used herein, the term “glucoamylase” (EC 3.2.1.3) refers to enzymes that catalyze the release of D-glucose from non-reducing ends of oligo- and poly-saccharide molecules. Glucoamylase is also generally considered a type of amylase known as amylo-glucosidase.

[0095] As used hererin, the term “amylase” (EC 3.2.1.1) refers to starch cleaving enzymes that degrade starch and related compounds by hydrolyzing the alpha-1,4 and/or alpha-1,6 glucosidic linkages in an endo- or an exo-acting fashion. Amylases include alpha-amylases (EC 3.2.1.1); beta-amylases (3.2.1.2), amylo-amylases (EC 3.2.1.3), alpha-glucosidases (EC 3.2.1.20), pullulanases (EC 3.2.1.41), and isoamylases (EC 3.2.1.68). In some embodiments, the amylase is an alpha-amylase.

[0096] As used herein, the term “pectinase” refers to enzymes that catalyze the hydrolysis of pectin into smaller units such as oligosaccharide or monomeric saccharides. In some embodiments, the enzyme mixtures comprise any pectinase, for example an endo-polygalacturonase, a pectin methyl esterase, an endo-galactanase, a pectin acetyl esterase, an endo-pectin lyase, pectate lyase, alpha rhamnosidase, an exo-galacturonase, an exo-polygalacturonate lyase, a rhamnogalacturonan hydrolase, a rhamnogalacturonan lyase, a rhamnogalacturonan acetyl esterase, a rhamnogalacturonan galacturonohydrolase and/or a xylogalacturonase.

[0097] As used herein, the term “endo-polygalacturonase” (EC 3.2.1.15) refers to enzymes that catalyze the random hydrolysis of 1,4-alpha-D-galactosiduronic linkages in pectate and other galacturonans. This enzyme may also be referred to as “polygalacturonase pectin depolymerase,” “pectinase,” “endopolygalacturonase,” “pectolase,” “pectin hydrolase,” “pectin polygalacturonase,” “poly-alpha-1,4-galacturonide glycanohydrolase,” “endogalacturonase,” “endo-D-galacturonase,” or “poly(1,4-alpha-D-galacturonide) glycanohydrolase.”

[0098] As used herein, the term “pectin methyl esterase” (EC 3.1.1.11) refers to enzymes that catalyze the reaction: pectin + n H₂O = n methanol + pectate. The enzyme may also be known as “pectin esterase,” “pectin demethoxylase,” “pectin methoxylase,” “pectin methylesterase,” “pectase,” “pectinoesterase,” or” pectin pectylhydrolase.”

[0099] As used herein, the term “endo-galactanase” (EC 3.2.1.89) refers to enzymes that catalyze the endohydrolysis of 1,4-beta-D-galactosidic linkages in arabinogalactans. The enzyme may also be known as “arabinogalactan endo-1,4-beta-galactosidase,” “endo-1,4-beta-galactanase,” galactanase,” “arabinogalactanase,” or “arabinogalactan 4-β-D-galactanohydrolase.”

[00100] As used herein, the term “pectin acetyl esterase” refers to enzymes that catalyze the deacetylation of the acetyl groups at the hydroxyl groups of GalUA residues of pectin.

[00101] As used herein, the term “one endo-pectin lyase” (EC 4.2.2.10) refers to enzymes that catalyze the eliminative cleavage of (1 →4)-alpha-D-galacturonan methyl ester to give oligosaccharides with 4-deoxy-6-O-methyl-alpha-D-galact-4-enuronosyl groups at their non- reducing ends. The enzyme may also be known as “pectin lyase,” “pectin trans-eliminase,” “endo-pectin lyase,” “polymethylgalacturonic transeliminase,” “pectin methyltranseliminase,” “pectolyase,” “PL,” “PNL,” ” PMGL,” or “(1 →4)-6-O-methyl-α-D-galacturonan lyase.”

[00102] As used herein, the term “pectate lyase” (EC 4.2.2.2) refers to enzymes that catalyze the eliminative cleavage of (1 →4)-alpha-D-galacturonan to give oligosaccharides with 4-deoxy-alpha-D-galact-4-enuronosyl groups at their non-reducing ends. The enzyme may also be known as “polygalacturonic transeliminase,” “pectic acid transeliminase,” “polygalacturonate lyase,” “endopectin methyltranseliminase,” “pectate transeliminase,” “endogalacturonate transeliminase,” “pectic acid lyase,” “pectic lyase,” “alpha-1,4-D-endopolygalacturonic acid lyase,” “PGA lyase,” “PPase-N,” “endo-alpha-1,4-polygalacturonic acid lyase,” “polygalacturonic acid lyase,” “pectin trans-eliminase,” “polygalacturonic acid trans-eliminase,” or “(1 →4)-alpha-D- galacturonan lyase.”

[00103] As used herein, the term “alpha-rhamnosidase” (EC 3.2.1.40) refers to enzymes that catalyze the hydrolysis of terminal non-reducing alpha-L-rhamnose residues in alpha-L- rhamnosides or alternatively in rhamnogalacturonan. This enzyme may also be

known as “alpha-L-rhamnosidase T,” “alpha-L-rhamnosidase N,” or “alpha-L-rhamnoside rhamnohydrolase.”

[00104] As used herein, the term “exo-galacturonase” (EC 3.2.1.82) refers to enzymes that hydrolyze pectic acid from the non-reducing end, releasing digalacturonate. The enzyme may also be known as “exo-poly-alpha-galacturonosidase,” “exopolygalacturonosidase,” or “exopolygalacturanosidase.”

[00105] As used herein, the term “exo-galacturan 1,4-alpha galacturonidase” (EC 3.2.1.67) refers to enzymes that catalyze reactions of the following types: $(1,4\text{-}\alpha\text{-D-galacturonide})_n + \text{H}_2\text{O} = (1,4\text{-}\alpha\text{-D-galacturonide})_{n-1} + \text{D-galacturonate}$. The enzyme may also be known as “poly [1→4] alpha-D-galacturonide] galacturonohydrolase,” “exopolygalacturonase,” “poly(galacturonate) hydrolase,” “exo-D-galacturonase,” “exo-D-galacturonanase,” “exopoly-D-galacturonase,” or “poly(1,4-alpha-D-galacturonide) galacturonohydrolase.”

[00106] As used herein, the term “exopolygalacturonate lyase” (EC 4.2.2.9) refers to enzymes that catalyze eliminative cleavage of 4-(4-deoxy- α -D-galact-4-enuronosyl)-D-galacturonate from the reducing end of pectate (*i.e.* de-esterified pectin). This enzyme may be known as “pectate disaccharide-lyase,” “pectate exo-lyase,” “exopectic acid transeliminase,” “exopectate lyase,” “exopolygalacturonic acid-trans-eliminase,” “PATE,” “exo-PATE,” “exo-PGL,” or “(1 →4)-alpha-D-galacturonan reducing-end-disaccharide-lyase.”

[00107] As used herein, the term “rhamnogalacturonanase” refers to enzymes that hydrolyze the linkage between galactosyluronic acid and rhamnopyranosyl in an endo-fashion in strictly alternating rhamnogalacturonan structures, consisting of the disaccharide [(1,2-alpha-L-rhamnoyl-(1,4)-alpha-galactosyluronic acid)].

[00108] As used herein, the term “rhamnogalacturonan lyase” refers to enzymes that cleave alpha-L-Rhap-(1 →4)-alpha-D-GalpA linkages in an endo-fashion in rhamnogalacturonan by beta-elimination.

[00109] As used herein, the term “rhamnogalacturonan acetyl esterase” refers to enzymes that catalyze the deacetylation of the backbone of alternating rhamnose and galacturonic acid residues in rhamnogalacturonan.

[00110] As used herein, the term “rhamnogalacturonan galacturonohydrolase” refers to enzymes that hydrolyze galacturonic acid from the non-reducing end of strictly alternating rhamnogalacturonan structures in an exo-fashion. This enzyme may also be known as “xylogalacturonan hydrolase.”

[00111] As used herein, the term “endo-arabinanase” (EC 3.2.1.99) refers to enzymes that catalyze endohydrolysis of 1,5- α -arabinofuranosidic linkages in 1,5-arabinans. The enzyme may also be known as “endo-arabinase,” “arabinan endo-1,5- α -L-arabinosidase,” “endo-1,5- α -L-arabinanase,” “endo- α -1,5-arabanase,” “endo-arabanase,” or “1,5- α -L-arabinan 1,5- α -L-arabinanohydrolase.”

[00112] As used herein, “protease” includes enzymes that hydrolyze peptide bonds (peptidases), as well as enzymes that hydrolyze bonds between peptides and other moieties, such as sugars (glycopeptidases). Many proteases are characterized under EC 3.4, and are suitable for use in the present invention. Some specific types of proteases include but are not limited to, cysteine proteases including pepsin, papain and serine proteases including chymotrypsins, carboxypeptidases and metalloendopeptidases.

[00113] As used herein, “lipase” includes enzymes that hydrolyze lipids, fatty acids, and acylglycerides, including phosphoglycerides, lipoproteins, diacylglycerols, and the like. In plants, lipids are used as structural components to limit water loss and pathogen infection. These lipids include waxes derived from fatty acids, as well as cutin and suberin.

[00114] As used herein, the terms “isolated” and “purified” are used to refer to a molecule (*e.g.*, an isolated nucleic acid, polypeptide [including, but not limited to enzymes], etc.) or other component that is removed from at least one other component with which it is naturally associated. It is intended that the term encompass any suitable method for removing at least one component with which the molecule is naturally associated. In some embodiments, the terms also encompass cells that are separated from other cells and/or media components. It is intended that any suitable separation method finds use in the present invention.

[00115] As used herein, the term “purification process” used in reference to an enzyme mixture encompasses any process that physically removes an undesired component of the enzyme mixture. Thus, in some embodiments, purification processes provided herein include purification methodologies that physically remove one or more glucose and/or cellobiose

oxidizing enzymes from the enzyme mixture or vice versa. It is contemplated that any suitable purification process known in the art will find use in the present invention. Indeed, it is not intended that the present invention be limited to any particular purification process.

[00116] As used herein, the term “cell-free enzyme mixture” comprises enzymes that have been separated from any cells, including the cells that secreted the enzymes. Cell-free enzyme mixtures can be prepared by any of a variety of methodologies that are known in the art, such as filtration or centrifugation methodologies. In some embodiments, the enzyme mixture can be, for example, partially cell-free, substantially cell-free, or entirely cell-free.

[00117] As used herein, “polynucleotide” refers to a polymer of deoxyribonucleotides or ribonucleotides in either single- or double-stranded form, and complements thereof.

[00118] The terms “protein” and “polypeptide” are used interchangeably herein to refer to a polymer of amino acid residues.

[00119] In addition, the terms “amino acid,” “polypeptide,” and “peptide” encompass naturally-occurring and synthetic amino acids, as well as amino acid analogs. Naturally occurring amino acids are those encoded by the genetic code, as well as those amino acids that are later modified (*e.g.*, hydroxyproline, γ -carboxyglutamate, and O-phosphoserine). As used herein, the term “amino acid analogs” refers to compounds that have the same basic chemical structure as a naturally occurring amino acid (*i.e.*, an α -carbon that is bound to a hydrogen, a carboxyl group, an amino group, and an R group, including but not limited to homoserine, norleucine, methionine sulfoxide, and methionine methyl sulfonium). In some embodiments, these analogs have modified R groups (*e.g.*, norleucine) and/or modified peptide backbones, but retain the same basic chemical structure as a naturally occurring amino acid.

[00120] Amino acids are referred to herein by either their commonly known three letter symbols or by the one-letter symbols recommended by the IUPAC-IUB Biochemical Nomenclature Commission. Nucleotides, likewise, may be referred to by their commonly accepted single-letter codes.

[00121] An amino acid or nucleotide base “position” is denoted by a number that sequentially identifies each amino acid (or nucleotide base) in the reference sequence based on its position relative to the N-terminus (or 5'-end). Due to deletions, insertions, truncations, fusions, and the like that must be taken into account when determining an optimal alignment,

the amino acid residue number in a test sequence determined by simply counting from the N-terminus will not necessarily be the same as the number of its corresponding position in the reference sequence. For example, in a case where a test sequence has a deletion relative to an aligned reference sequence, there will be no amino acid in the variant that corresponds to a position in the reference sequence at the site of deletion. Where there is an insertion in an aligned test sequence, that insertion will not correspond to a numbered amino acid position in the reference sequence. In the case of truncations or fusions there can be stretches of amino acids in either the reference or aligned sequence that do not correspond to any amino acid in the corresponding sequence.

[00122] As used herein, the terms "numbered with reference to" or "corresponding to," when used in the context of the numbering of a given amino acid or polynucleotide sequence, refers to the numbering of the residues of a specified reference sequence when the given amino acid or polynucleotide sequence is compared to the reference sequence.

[00123] As used herein, the term "reference enzyme" refers to an enzyme to which another enzyme of the present invention (*e.g.*, a "test" enzyme) is compared in order to determine the presence of an improved property in the other enzyme being evaluated. In some embodiments, a reference enzyme is a wild-type enzyme. In some embodiments, the reference enzyme is an enzyme to which a test enzyme of the present invention is compared in order to determine the presence of an improved property in the test enzyme being evaluated, including but not limited to improved thermoactivity, improved thermostability, and/or improved stability. In some embodiments, a reference enzyme is a wild-type enzyme.

[00124] As used herein, the term "biologically active fragment," refers to a polypeptide that has an amino-terminal and/or carboxy-terminal deletion(s) and/or internal deletion(s), but where the remaining amino acid sequence is identical to the corresponding positions in the sequence to which it is being compared and that retains substantially all of the activity of the full-length polypeptide.

[00125] As used herein, the term "recombinant" refers to a polynucleotide or polypeptide that does not naturally occur in a host cell. In some embodiments, recombinant molecules contain two or more naturally-occurring sequences that are linked together in a way that does not occur naturally. In some embodiments, "recombinant cells" express genes that are not found in identical form within the native (*i.e.*, non-recombinant) form of the cell

and/or express native genes that are otherwise abnormally over-expressed, under-expressed, and/or not expressed at all due to deliberate human intervention. Recombinant cells contain at least one recombinant polynucleotide or polypeptide. A nucleic acid construct, nucleic acid (*e.g.*, a polynucleotide), polypeptide, or host cell is referred to herein as "recombinant" when it is non-naturally occurring, artificial or engineered. "Recombination," "recombining" and generating a "recombined" nucleic acid generally encompass the assembly of at least two nucleic acid fragments.

[00126] The present invention also provides a recombinant nucleic acid construct comprising at least one CDH polynucleotide sequence that hybridizes under stringent hybridization conditions to the complement of a polynucleotide which encodes a polypeptide having the amino acid sequence of SEQ ID NOS:6 and/or 8.

[00127] Nucleic acids "hybridize" when they associate, typically in solution. Nucleic acids hybridize due to a variety of well-characterized physico-chemical forces, such as hydrogen bonding, solvent exclusion, base stacking and the like. As used herein, the term "stringent hybridization wash conditions" in the context of nucleic acid hybridization experiments, such as Southern and Northern hybridizations, are sequence dependent, and are different under different environmental parameters. An extensive guide to the hybridization of nucleic acids is found in Tijssen, 1993, "Laboratory Techniques in Biochemistry and Molecular Biology-Hybridization with Nucleic Acid Probes," Part I, Chapter 2 (Elsevier, New York). For polynucleotides of at least 100 nucleotides in length, low to very high stringency conditions are defined as follows: prehybridization and hybridization at 42°C in 5xSSPE, 0.3% SDS, 200 µg/ml sheared and denatured salmon sperm DNA, and either 25% formamide for low stringencies, 35% formamide for medium and medium-high stringencies, or 50% formamide for high and very high stringencies, following standard Southern blotting procedures. For polynucleotides of at least 100 nucleotides in length, the carrier material is finally washed three times each for 15 minutes using 2xSSC, 0.2% SDS 50°C (low stringency), at 55°C (medium stringency), at 60°C (medium-high stringency), at 65°C (high stringency), or at 70°C (very high stringency).

[00128] Moderately stringent conditions encompass those known in the art and described in various standard texts and include the use of washing solution and hybridization conditions (*e.g.*, temperature, ionic strength and %SDS). An example of moderately

stringent conditions involves overnight incubation at 37°C in a solution comprising: 20% formamide, 5 x SSC (150 mM NaCl, 15 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5 x Denhardt's solution, 10% dextran sulfate, and 20 mg/mL denatured sheared salmon sperm DNA, followed by washing the filters in 1 x SSC at about 37-50°C. The skilled artisan will recognize how to adjust the temperature, ionic strength, etc. as necessary to accommodate factors such as probe length and the like.

[00129] As used in some embodiments herein, stringent conditions or high stringency conditions utilize: (1) low ionic strength and high temperature for washing, for example 0.015 M sodium chloride/0.0015 M sodium citrate/0.1% sodium dodecyl sulfate at 50°C; (2) during hybridization a denaturing agent, such as formamide, for example, 50% (v/v) formamide with 0.1% bovine serum albumin/0.1% Ficoll/0.1% polyvinylpyrrolidone/50mM sodium phosphate buffer at pH 6.5 with 750 mM sodium chloride, 75 mM sodium citrate at 42°C; or (3) 50% formamide, 5 x SSC (0.75 M NaCl, 0.075 M sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5 x Denhardt's solution, sonicated salmon sperm DNA (50 µg/mL), 0.1% SDS, and 10% dextran sulfate at 42°C, with washes at 42°C in 0.2 x SSC (sodium chloride/sodium citrate) and 50% formamide at 55°C, followed by a high-stringency wash consisting of 0.1 x SSC containing EDTA at 55°C.

[00130] As used herein, "similarity" refers to an identical or conservative amino acid substitution thereof as defined below. Accordingly, a change to an identical or conservative substitution for the purposes of similarity is viewed as not comprising a change. A deletion of an amino acid or a non-conservative amino acid substitution is viewed herein as comprising a change. Calculation of percent similarity is performed in the same manner as performed for percent identity. A conservative amino acid substitution can be a substitution such as the conservative substitutions shown in Table A. The substitutions shown are based on amino acid physical-chemical properties, and as such, are independent of organism. In some embodiments, the conservative amino acid substitution is a substitution listed under the heading of exemplary substitutions.

Table A. Substitutions		
Original Residue	Conservative Substitutions	Exemplary Substitutions

Table A. Substitutions		
Original Residue	Conservative Substitutions	Exemplary Substitutions
Ala (A)	val; leu; ile	Val
Arg (R)	lys; gln; asn	Lys
Asn (N)	gln; his; lys; arg	Gln
Asp (D)	Glu	Glu
Cys (C)	Ser	Ser
Gln (Q)	Asn	Asn
Glu (E)	Asp	Asp
Gly (G)	pro; ala	Ala
His (H)	asn; gln; lys; arg	Arg
Ile (I)	leu; val; met; ala; phe	Leu
Leu (L)	ile; val; met; ala; phe	Ile
Lys (K)	arg; gln; asn	Arg
Met (M)	leu; phe; ile	Leu
Phe (F)	leu; val; ile; ala; tyr	Leu
Pro (P)	Ala	Ala
Ser (S)	Thr	Thr
Thr (T)	Ser	Ser
Trp (W)	tyr; phe	Tyr
Tyr (Y)	trp; phe; thr; ser	Phe
Val (V)	ile; leu; met; phe; ala	Leu

[00131] As used herein, "identity" and "percent identity," in the context of two or more polypeptide sequences, refers to two or more sequences or subsequences that are the same or have a specified percentage of amino acid residues that are the same (*e.g.*, share at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 88% identity, at least about 89%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, or at least about 99% identity) over a specified region to a reference sequence, when compared and aligned for maximum correspondence over a comparison window, or designated region as measured using a sequence comparison algorithms or by manual alignment and visual inspection.

[00132] In some embodiments, the terms "percent identity," "% identity", "percent identical," and "% identical," are used interchangeably herein to refer to the percent amino acid or polynucleotide sequence identity that is obtained by ClustalW analysis (version W 1.8 available from European Bioinformatics Institute, Cambridge, UK), counting the number of

identical matches in the alignment and dividing such number of identical matches by the length of the reference sequence, and using the following ClustalW parameters to achieve slow/more accurate pairwise optimal alignments – DNA/Protein Gap Open Penalty:15/10; DNA/Protein Gap Extension Penalty:6.66/0.1; Protein weight matrix: Gonnet series; DNA weight matrix: Identity.

[00133] Two sequences are "aligned" when they are aligned for similarity scoring using a defined amino acid substitution matrix (e.g., BLOSUM62), gap existence penalty and gap extension penalty so as to arrive at the highest score possible for that pair of sequences. Amino acid substitution matrices and their use in quantifying the similarity between two sequences are well known in the art (*See, e.g., Dayhoff et al., in Dayhoff [ed.], Atlas of Protein Sequence and Structure, " Vol. 5, Suppl. 3, Natl. Biomed. Res. Round., Washington D.C. [1978]; pp. 345-352; and Henikoff et al., Proc. Natl. Acad. Sci. USA, 89:10915-10919 [1992]*). The BLOSUM62 matrix is often used as a default scoring substitution matrix in sequence alignment protocols such as Gapped BLAST 2.0. The gap existence penalty is imposed for the introduction of a single amino acid gap in one of the aligned sequences, and the gap extension penalty is imposed for each additional empty amino acid position inserted into an already opened gap. The alignment is defined by the amino acid position of each sequence at which the alignment begins and ends, and optionally by the insertion of a gap or multiple gaps in one or both sequences so as to arrive at the highest possible score. While optimal alignment and scoring can be accomplished manually, the process is facilitated by the use of a computer-implemented alignment algorithm (e.g., gapped BLAST 2.0; *See, Altschul et al., Nucleic Acids Res., 25:3389-3402 [1997]*), and made available to the public at the National Center for Biotechnology Information Website). Optimal alignments, including multiple alignments can be prepared using readily available programs such as PSI-BLAST (*See e.g., Altschul et al., supra*).

[00134] The present invention also provides a recombinant nucleic acid construct comprising a CDH polynucleotide sequence that hybridizes under stringent hybridization conditions to the complement of a polynucleotide which encodes a polypeptide having the amino acid sequence of SEQ ID NO:6 and/or 8. Two nucleic acid or polypeptide sequences that have 100% sequence identity are said to be "identical." A nucleic acid or polypeptide

sequence is said to have "substantial sequence identity" to a reference sequence when the sequences have at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, or at least about 99%, or greater sequence identity as determined using the methods described herein, such as BLAST using standard parameters.

[00135] As used herein, a "secretion signal peptide" can be a propeptide, a prepeptide or both. For example, the term "propeptide" refers to a protein precursor that is cleaved to yield a "mature protein." The signal peptide is cleaved from the pre-protein by a signal peptidase prior to secretion to result in the "mature" or "secreted" protein. The terms "prepeptide" and "pre-protein" refer to a polypeptide synthesized with an N-terminal signal peptide that targets it for secretion. Accordingly, a "pre-pro-peptide" is a polypeptide that contains a signal peptide that targets the polypeptide for secretion and which is cleaved off to yield a mature polypeptide. Signal peptides are found at the N-terminus of the protein and are typically composed of between 6 to 136 basic and hydrophobic amino acids.

[00136] As used herein, "transcription" and like terms refer to the conversion of the information encoded in a gene to an RNA transcript. Accordingly, a reduction of the transcription level of a glucose and/or cellobiose oxidizing enzyme is a reduction in the amount of RNA transcript of an RNA coding for a glucose and/or cellobiose oxidizing enzyme.

[00137] As used herein, a "vector" is a polynucleotide construct for introducing a polynucleotide sequence into a cell. In some embodiments, the vector comprises a suitable control sequence operably linked to and capable of effecting the expression of the polypeptide encoded in the polynucleotide sequence in a suitable host. An "expression vector" has a promoter sequence operably linked to the polynucleotide sequence (*e.g.*, transgene) to drive expression in a host cell, and in some embodiments a transcription terminator sequence. In some embodiments, the vectors are deletion vectors. In some embodiments, vectors comprise polynucleotide sequences that produce small interfering RNA or antisense RNA transcripts that interfere with the translation of a target polynucleotide sequence.

[00138] As used herein, a “deletion vector” comprises polynucleotide sequences homologous to a polynucleotide sequences 5’ and 3’ to a target sequence to be deleted from a host genome so as to direct recombination and replacement of the target sequence with a polynucleotide between the 5’ and 3’ targeting sequences.

[00139] As used herein, the term “expression” includes any step involved in the production of the polypeptide including, but not limited to, transcription, post-transcriptional modification, translation, and post-translational modification. In some embodiments, the term also encompasses secretion of the polypeptide from a cell. In general the term, “expression” refers to conversion of the information encoded in a gene to the protein encoded by that gene. Thus, a “reduction of the amount of an expressed glucose and/or cellobiose oxidizing enzyme” is a reduction in the amount of the glucose and/or cellobiose oxidizing enzyme that is eventually translated by the cell.

[00140] As used herein, the term “overexpress” is intended to encompass increasing the expression of a protein to a level greater than the cell normally produces. It is intended that the term encompass overexpression of endogenous, as well as heterologous proteins. In some embodiments, overexpression includes an elevated transcription rate and/or level of the gene compared to the endogenous transcription rate and/or level for that gene. For example, in some embodiments, a heterologous gene is introduced into a fungal cell to express a gene encoding a heterologous enzyme such as a beta-glucosidase from another organism. In some other embodiments, a heterologous gene is introduced into a fungal cell to overexpress a gene encoding a homologous enzyme such as a beta-glucosidase.

[00141] In some embodiments, the heterologous gene is a gene that has been modified to overexpress the gene product. In some embodiments, “overexpression” refers to any state in which a gene is caused to be expressed at an elevated rate or level as compared to the endogenous expression rate or level for that gene. In some embodiments, overexpression includes elevated translation rate and/or level of the gene compared to the endogenous translation rate and/or level for that gene. As used herein, the term “produces” refers to the production of proteins and/or other compounds by cells. It is intended that the term encompass any step involved in the production of polypeptides including, but not limited to, transcription, post-transcriptional modification, translation, and post-translational

modification. In some embodiments, the term also encompasses secretion of the polypeptide from a cell.

[00142] As used herein, a “polynucleotide sequence that has been adapted for expression” is a polynucleotide sequence that has been inserted into an expression vector or otherwise modified to contain regulatory elements necessary for expression of the polynucleotide in the host cell, positioned in such a manner as to permit expression of the polynucleotide in the host cell. Such regulatory elements required for expression include promoter sequences, transcription initiation sequences and, optionally, enhancer sequences. For example, in some embodiments, a polynucleotide sequence is inserted into a plasmid vector adapted for expression in the fungal host cell.

[00143] As used herein, the term "operably linked" refers to a configuration in which a control sequence is appropriately placed at a position relative to the coding sequence of the DNA sequence such that the control sequence influences the expression of a polypeptide.

[00144] As used herein, an amino acid or nucleotide sequence (*e.g.*, a promoter sequence, signal peptide, terminator sequence, etc.) is "heterologous" to another sequence with which it is operably linked if the two sequences are not associated in nature.

[00145] As used herein, a “heterologous enzyme” refers to an enzyme that is encoded by a “heterologous gene.” However, it is also contemplated that a heterologous gene encodes an endogenous or homologous enzyme, as explained below. In general, the term “heterologous gene” refers to a gene that occurs in a form not found in a parental strain of the host fungal cell (including but not limited to wild-type). Thus, in some embodiments, a heterologous gene is a gene that is derived from a species that is different from the species of the fungal cell expressing the gene and recognized anamorphs, teleomorphs or taxonomic equivalents of the fungal cell expressing the gene. In some embodiments, a heterologous gene is a modified version of a gene that is endogenous to the host fungal cell, which endogenous gene has been subjected to manipulation and then introduced or transformed into the host cell. For example, in some embodiments, a heterologous gene has an endogenous coding sequence, but has modifications to the promoter sequence. Similarly, in some embodiments, a heterologous gene encodes the same amino acid sequence as an endogenous gene, but has modifications to the codon usage or to noncoding regions such as introns, or a combination thereof. For example, in some embodiments, a heterologous gene comprises

modifications to the coding sequence to encode a non-wild type polypeptide. In some other embodiments, a heterologous gene has the same promoter sequence, 5' and 3' untranslated regions and coding regions as a parental strain, but be located in another region of the same chromosome, or on an entirely different chromosome as compared to a parental strain of the host cell.

[00146] As used herein, an “endogenous” or “homologous” gene refers to a gene that is found in a parental strain of the host fungal cell (including, but not limited to wild-type).

[00147] As used herein, the term “introduced” used in the context of inserting a nucleic acid sequence into a cell, means transformation, transduction, conjugation, transfection, and/or any other suitable method(s) known in the art for inserting nucleic acid sequences into host cells. Any suitable means for the introduction of nucleic acid into host cells find use in the present invention.

[00148] As used herein, the terms “transformed” and “transformation” used in reference to a cell refer to a cell that has a non-native nucleic acid sequence integrated into its genome or has an episomal plasmid that is maintained through multiple generations.

[00149] As used herein, the terms “host cell” and “host strain” refer to suitable hosts for expression vectors comprising polynucleotide sequences (*e.g.*, DNA) as provided herein. In some embodiments, the host cells are prokaryotic or eukaryotic cells that have been transformed or transfected with vectors constructed using recombinant techniques as known in the art. Transformed hosts are capable of either replicating vectors encoding at least one protein of interest and/or expressing the desired protein of interest. In addition, reference to a cell of a particular strain refers to a parental cell of the strain as well as progeny and genetically modified derivatives. Genetically modified derivatives of a parental cell include progeny cells that contain a modified genome or episomal plasmids that confer for example, antibiotic resistance, improved fermentation, etc. In some embodiments, host cells are genetically modified to have characteristics that improve protein secretion, protein stability or other properties desirable for expression and/or secretion of a protein. For example, knockout of *Alp1* function results in a cell that is protease deficient. Knockout of *pyr5* function results in a cell with a pyrimidine deficient phenotype. In some embodiments, host cells are modified to delete endogenous cellulase protein-encoding sequences or otherwise eliminate expression of one or more endogenous cellulases. In some embodiments,

expression of one or more endogenous cellulases is inhibited to increase production of cellulases of interest. Genetic modification can be achieved by any suitable genetic engineering techniques and/or classical microbiological techniques (*e.g.*, chemical or UV mutagenesis and subsequent selection). Using recombinant technology, nucleic acid molecules can be introduced, deleted, inhibited or modified, in a manner that results in increased yields of enzyme within the organism or in the culture. For example, knockout of *Alp1* function results in a cell that is protease deficient. Knockout of *pyr5* function results in a cell with a pyrimidine deficient phenotype. In some genetic engineering approaches, homologous recombination is used to induce targeted gene modifications by specifically targeting a gene *in vivo* to suppress expression of the encoded protein. In an alternative approach, siRNA, antisense, and/or ribozyme technology finds use in inhibiting gene expression.

[00150] As used herein, “gene deletion” and “deletion mutation” refer to a mutation in which part of a gene is missing. Thus, a deletion is a loss or replacement of genetic material resulting in a complete or partial disruption of the sequence of the DNA making up the gene. Any number of nucleotides can be deleted, from a single base to an entire piece of a chromosome. In some embodiments, complete or near-complete deletion of the gene sequence is contemplated. However, a deletion mutation need not completely remove the entire gene sequence for the glucose and/or cellobiose oxidizing enzyme in order to reduce the endogenous glucose and/or cellobiose oxidizing enzyme activity secreted by the fungal cell. For example, a partial deletion that removes one or more nucleotides encoding an amino acid in a glucose and/or cellobiose oxidizing enzyme active site, encoding a secretion signal, or encoding another portion of the glucose and/or cellobiose oxidizing enzyme that plays a role in endogenous glucose and/or cellobiose oxidizing enzyme activity being secreted by the fungal cell.

[00151] As used herein, a “conditional mutation” is a mutation that has wild-type phenotype under certain environmental conditions and a mutant phenotype under certain other conditions.

[00152] As used herein, the terms “amplification” and “gene amplification” refer to a method by which specific DNA sequences are disproportionately replicated such that the amplified gene becomes present in a higher copy number than was initially present in the

genome. In some embodiments, selection of cells by growth in the presence of a drug (*e.g.*, an inhibitor of an inhibitable enzyme) results in the amplification of either the endogenous gene encoding the gene product required for growth in the presence of the drug or by amplification of exogenous (*i.e.*, input) sequences encoding this gene product, or both. "Amplification" is a special case of nucleic acid replication involving template specificity. It is to be contrasted with non-specific template replication (*i.e.*, replication that is template-dependent but not dependent on a specific template). Template specificity is here distinguished from fidelity of replication (*i.e.*, synthesis of the proper polynucleotide sequence) and nucleotide (ribo- or deoxyribo-) specificity. Template specificity is frequently described in terms of "target" specificity. Target sequences are "targets" in the sense that they are sought to be sorted out from other nucleic acid. Amplification techniques have been designed primarily for this sorting out.

[00153] As used herein, the term "primer" refers to an oligonucleotide, whether occurring naturally as in a purified restriction digest or produced synthetically, that is capable of acting as a synthesis initiation point when placed under conditions in which synthesis of a primer extension product which is complementary to a nucleic acid strand is induced (*i.e.*, in the presence of nucleotides and an inducing agent such as DNA polymerase and at a suitable temperature and pH). The primer is preferably single stranded for maximum efficiency in amplification, but may alternatively be double stranded. If double stranded, the primer is first treated to separate its strands before being used to prepare extension products. In some embodiments, the primer is an oligodeoxyribonucleotide. The primer must be sufficiently long to prime the synthesis of extension products in the presence of the inducing agent. As known in the art, the exact lengths of the primers will depend on many factors, including temperature, source of primer and the use of the method.

[00154] As used herein, the term "probe" refers to an oligonucleotide (*i.e.*, a sequence of nucleotides), whether occurring naturally as in a purified restriction digest or produced synthetically, recombinantly or by PCR amplification, that is capable of hybridizing to another oligonucleotide of interest. A probe may be single-stranded or double-stranded. Probes are useful in the detection, identification and isolation of particular gene sequences. It is contemplated that any probe used in the present invention will be labeled with any "reporter molecule," so that is detectable in any detection system, including, but not limited

to enzyme (*e.g.*, ELISA, as well as enzyme-based histochemical assays), fluorescent, radioactive, and luminescent systems. It is not intended that the present invention be limited to any particular detection system or label.

[00155] As used herein, the term "target," when used in reference to the polymerase chain reaction, refers to the region of nucleic acid bounded by the primers used for polymerase chain reaction. Thus, the "target" is sought to be sorted out from other nucleic acid sequences. A "segment" is defined as a region of nucleic acid within the target sequence.

[00156] As used herein, the term "polymerase chain reaction" (PCR) refers to the methods of U.S. Pat. Nos. 4,683,195 4,683,202, and 4,965,188, which include methods for increasing the concentration of a segment of a target sequence in a mixture of genomic DNA without cloning or purification. This method for amplifying the target sequence is well known in the art.

[00157] As used herein, the term "amplification reagents" refers to those reagents (deoxyribonucleotide triphosphates, buffer, etc.), needed for amplification except for primers, nucleic acid template and the amplification enzyme. Typically, amplification reagents along with other reaction components are placed and contained in a reaction vessel (test tube, microwell, etc.).

[00158] As used herein, the terms "restriction endonucleases" and "restriction enzymes" refer to bacterial enzymes, each of which cut double-stranded DNA at or near a specific nucleotide sequence.

[00159] A "restriction site" refers to a nucleotide sequence recognized and cleaved by a given restriction endonuclease and is frequently the site for insertion of DNA fragments. In some embodiments of the invention, restriction sites are engineered into the selective marker and into 5' and 3' ends of the DNA construct.

[00160] As used herein, "homologous recombination" means the exchange of DNA fragments between two DNA molecules or paired chromosomes at the site of identical or nearly identical nucleotide sequences. In some embodiments, chromosomal integration is homologous recombination.

[00161] As used herein, the term "C1" refers to *Myceliophthora thermophila*, including the fungal strain described by Garg (*See*, Garg, Mycopathol., 30: 3-4 [1966]). As used herein, "*Chrysosporium lucknowense*" includes the strains described in U.S. Pat. Nos.

6,015,707, 5,811,381 and 6,573,086; US Pat. Pub. Nos. 2007/0238155, US 2008/0194005, US 2009/0099079; International Pat. Pub. Nos., WO 2008/073914 and WO 98/15633, and include, without limitation, *Chrysosporium lucknowense* Garg 27K, VKM-F 3500 D (Accession No. VKM F-3500-D), C1 strain UV13-6 (Accession No. VKM F-3632 D), C1 strain NG7C-19 (Accession No. VKM F-3633 D), and C1 strain UV18-25 (VKM F-3631 D), all of which have been deposited at the All-Russian Collection of Microorganisms of Russian Academy of Sciences (VKM), Bakhurhina St. 8, Moscow, Russia, 113184, and any derivatives thereof. Although initially described as *Chrysosporium lucknowense*, C1 may currently be considered a strain of *Myceliophthora thermophila*. Other C1 strains include cells deposited under accession numbers ATCC 44006, CBS (Centraalbureau voor Schimmelcultures) 122188, CBS 251.72, CBS 143.77, CBS 272.77, CBS122190, CBS122189, and VKM F-3500D. Exemplary C1 derivatives include modified organisms in which one or more endogenous genes or sequences have been deleted or modified and/or one or more heterologous genes or sequences have been introduced. Derivatives include, but are not limited to UV18#100f Δ alp1, UV18#100f Δ pyr5 Δ alp1, UV18#100.f Δ alp1 Δ pep4 Δ alp2, UV18#100.f Δ pyr5 Δ alp1 Δ pep4 Δ alp2 and UV18#100.f Δ pyr4 Δ pyr5 Δ alp1 Δ pep4 Δ alp2, as described in WO 2008073914 and WO 2010107303.

[00162] As used herein, a “genetically modified” and/or “genetically engineered cell” (e.g., a “genetically engineered fungal cell” and/or a “genetically modified fungal cell”) is a cell whose genetic material has been altered using genetic engineering techniques. A genetically modified cell also refers to a derivative of or the progeny of a cell whose genetic material has been altered using genetic engineering techniques. An example of a genetic modification as a result of genetic engineering techniques includes a modification to the genomic DNA; another example of a genetic modification as a result of genetic engineering techniques includes introduction of a stable heterologous nucleic acid into the cell. For example, as provided herein, a genetically modified fungal cell as provided herein is a fungal cell that whose genetic material has been altered in such a way as to either reduce the amount of secreted glucose and/or cellobiose oxidizing enzyme activity, or to reduce the ability of the secreted enzyme to oxidize cellobiose or glucose.

[00163] As used herein, the term "culturing" refers to growing a population of microbial cells under suitable conditions in a liquid or solid medium. It is contemplated that the culturing be carried out in any suitable format, equipment (*e.g.*, shake flasks, fermentation tanks, bioreactors, etc.). It is also intended that the culturing be conducted using any suitable process methods, including but not limited to batch, fed-batch, and/or continuous culturing. Indeed, it is contemplated that any combination of suitable methods will find use.

[00164] In a "batch process," all the necessary materials, with the exception of oxygen for aerobic processes, are placed in a reactor at the start of the operation and the fermentation is allowed to proceed until completion, at which point the product is harvested. In some embodiments, batch processes for producing the fungal cells, enzymes, and/or enzyme mixtures of the present invention are carried out in a shake-flask or a bioreactor.

[00165] In a "fed-batch process," the culture is fed continuously or sequentially with one or more media components without the removal of the culture fluid.

[00166] In a "continuous process," fresh medium is supplied and culture fluid is removed continuously at volumetrically equal rates to maintain the culture at a steady growth rate. In reference to continuous processes, "steady state" refers to a state in which the concentration of reactants does not vary appreciably, and "quasi-steady state" refers to a state in which, subsequent to the initiation of the reaction, the concentration of reactants fluctuates within a range consistent with normal operation of the continuous hydrolysis process.

[00167] As used herein, the term "saccharification" refers to the process in which substrates (*e.g.*, cellulosic biomass) are broken down via the action of cellulases to produce fermentable sugars (*e.g.* monosaccharides such as but not limited to glucose).

[00168] As used herein, the term "fermentable sugars" refers to simple sugars (*e.g.*, monosaccharides, disaccharides and short oligosaccharides), including but not limited to glucose, xylose, galactose, arabinose, mannose and sucrose. Indeed, a fermentable sugar is any sugar that a microorganism can utilize or ferment.

[00169] As used herein the term "soluble sugars" refers to water-soluble pentose and hexose monomers and oligomers of up to about six monomer units. It is intended that the term encompass any water soluble mono- and/or oligosaccharides.

[00170] As used herein, the term "fermentation" is used broadly to refer to the process of obtaining energy from the oxidation of organic compounds (*e.g.*, carbohydrates). Indeed, "fermentation" broadly refers to the chemical conversion of a sugar source to an end product through the use of a fermenting organism. In some embodiments, the term encompasses cultivation of a microorganism or a culture of microorganisms that use sugars, such as fermentable sugars, as an energy source to obtain a desired product.

[00171] As used herein, the term "fermenting organism" refers to any organism, including prokaryotic, as well as eukaryotic organisms (*e.g.*, bacterial organisms, as well as fungal organisms such as yeast and filamentous fungi), suitable for producing a desired end product. Especially suitable fermenting organisms are able to ferment (*i.e.*, convert) sugars, including but not limited to glucose, fructose, maltose, xylose, mannose and/or arabinose, directly or indirectly into at least one desired end product. In some embodiments, yeast that find use in the present invention include, but are not limited to strains of the genus *Saccharomyces* (*e.g.*, strains of *Saccharomyces cerevisiae* and *Saccharomyces uvarum*), strains of the genus *Pichia* (*e.g.*, *Pichia stipitis* such as *Pichia stipitis* CBS 5773 and *Pichia pastoris*), and strains of the genus *Candida* (*e.g.*, *Candida utilis*, *Candida arabinofementans*, *Candida diddensii*, *Candida sonorensis*, *Candida shehatae*, *Candida tropicalis*, and *Candida boidinii*). Other fermenting organisms include, but are not limited to strains of *Zymomonas*, *Hansenula* (*e.g.*, *Hansenula polymorpha* and *Hansenula anomala*), *Kluyveromyces* (*e.g.*, *Kluyveromyces fragilis*), and *Schizosaccharomyces* (*e.g.*, *Schizosaccharomyces pombe*).

[00172] As used herein, the term "slurry" refers to an aqueous solution in which are dispersed one or more solid components, such as a cellulosic substrate. Thus, the term "slurry" refers to a suspension of solids in a liquid. In some embodiments, the cellulosic substrate is slurried in a liquid at a concentration that is thick, but can still be pumped. For example, in some embodiments, the liquid is water, a recycled process stream, and/or a treated effluent. However, it is not intended that the present invention be limited to any particular liquid and/or solid.

[00173] As used herein, "cellulose" refers to a polymer of the simple sugar glucose linked by beta-1,4 glycosidic bonds.

[00174] As used herein, "cellobiose" refers to a water-soluble beta-1,4-linked dimer of glucose.

[00175] The terms "biomass," and "biomass substrate," encompass any suitable materials for use in saccharification reactions. The terms encompass, but are not limited to, materials that comprise cellulose (*i.e.*, "cellulosic biomass," "cellulosic feedstock," and "cellulosic substrate"), as well as lignocellulosic biomass. Indeed, the term "biomass" encompasses any living or dead biological material that contains a polysaccharide substrate, including but not limited to cellulose, starch, other forms of long-chain carbohydrate polymers, and mixtures of such sources. In some embodiments, it is assembled entirely or primarily from glucose or xylose, and in some embodiments, optionally also contains various other pentose and/or hexose monomers. Biomass can be derived from plants, animals, or microorganisms, and includes, but is not limited to agricultural, industrial, and forestry residues, industrial and municipal wastes, and terrestrial and aquatic crops grown for energy purposes. Examples of biomass substrates include, but are not limited to, wood, wood pulp, paper pulp, corn fiber, corn grain, corn cobs, crop residues such as corn husks, corn stover, grasses, wheat, wheat straw, barley, barley straw, hay, rice, rice straw, switchgrass, waste paper, paper and pulp processing waste, woody or herbaceous plants, fruit or vegetable pulp, distillers grain, grasses, rice hulls, cotton, hemp, flax, sisal, sugar cane bagasse, sorghum, soy, switchgrass, components obtained from milling of grains, trees, branches, roots, leaves, wood chips, sawdust, shrubs and bushes, vegetables, fruits, and flowers and any suitable mixtures thereof. In some embodiments, the biomass comprises, but is not limited to cultivated crops (*e.g.*, grasses, including C4 grasses, such as switch grass, cord grass, rye grass, miscanthus, reed canary grass, or any combination thereof), sugar processing residues, for example, but not limited to, bagasse (*e.g.*, sugar cane bagasse, beet pulp [*e.g.*, sugar beet], or a combination thereof), agricultural residues (*e.g.*, soybean stover, corn stover, corn fiber, rice straw, sugar cane straw, rice, rice hulls, barley straw, corn cobs, wheat straw, canola straw, oat straw, oat hulls, corn fiber, hemp, flax, sisal, cotton, or any combination thereof), fruit pulp, vegetable pulp, distillers' grains, forestry biomass (*e.g.*, wood, wood pulp, paper pulp, recycled wood pulp fiber, sawdust, hardwood, such as aspen wood, softwood, or a combination thereof). Furthermore, in some embodiments, the biomass comprises cellulosic waste material and/or forestry waste materials, including but not limited to, paper and pulp processing waste, municipal paper waste, newsprint, cardboard and the like. In some embodiments, biomass comprises one species of fiber, while in some alternative

embodiments, the biomass comprises a mixture of fibers that originate from different biomasses. In some embodiments, the biomass also comprises transgenic plants that express ligninase and/or cellulase enzymes (*See e.g.*, US 2008/0104724 A1).

[00176] As used herein, “lignocellulose” refers to a matrix of cellulose, hemicellulose and lignin. Economic production of biofuels from lignocellulosic biomass typically involves conversion of the cellulose and hemicellulose components to fermentable sugars, typically monosaccharides such as glucose (from the cellulose) and xylose and arabinose (from the hemicelluloses). Nearly complete conversion can be achieved by a chemical pretreatment of the lignocellulose followed by enzymatic hydrolysis with cellulase enzymes. The chemical pretreatment step renders the cellulose more susceptible to enzymatic hydrolysis and, in some cases, also hydrolyzes the hemicellulose component. Numerous chemical pretreatment processes are known in the art, and include, but are not limited to, mild acid pretreatment at high temperatures and dilute acid, ammonium pretreatment or organic solvent extraction.

[00177] Lignin is a more complex and heterogeneous biopolymer than either cellulose or hemicellulose and comprises a variety of phenolic subunits. Enzymatic lignin depolymerization can be accomplished by lignin peroxidases, manganese peroxidases, laccases and cellobiose dehydrogenases (CDH), often working in synergy. However, as the name suggests, CDH enzymes also oxidize cellobiose to cellobionolactone. Several reports indicate that the oxidation of cellobiose by CDH enhances the rate of cellulose hydrolysis by cellulases by virtue of reducing the concentrations of cellobiose, which is a potent inhibitor of some cellulase components (Mansfield *et al.*, Appl. Environ. Microbiol., 63: 3804-3809 [1997]; and Igarishi *et al.*, Eur. J. Biochem., 253: 101-106 [1998]). Recently, it has been reported that CDHs can enhance the activity of cellulolytic enhancing proteins from Glycosyl Hydrolase family 61 (*See e.g.*, WO2010/080532A1).

[00178] As used herein, the term “lignocellulosic biomass” refers to any plant biomass comprising cellulose and hemicellulose, bound to lignin

[00179] In some embodiments, the biomass is optionally pretreated to increase the susceptibility of cellulose to hydrolysis by chemical, physical and biological pretreatments (such as steam explosion, pulping, grinding, acid hydrolysis, solvent exposure, and the like, as well as combinations thereof). Various lignocellulosic feedstocks find use, including those that comprise fresh lignocellulosic feedstock, partially dried lignocellulosic feedstock,

fully dried lignocellulosic feedstock, and/or any combination thereof. In some embodiments, lignocellulosic feedstocks comprise cellulose in an amount greater than about 20%, more preferably greater than about 30%, more preferably greater than about 40% (w/w). For example, in some embodiments, the lignocellulosic material comprises from about 20% to about 90% (w/w) cellulose, or any amount therebetween, although in some embodiments, the lignocellulosic material comprises less than about 19%, less than about 18%, less than about 17%, less than about 16%, less than about 15%, less than about 14%, less than about 13%, less than about 12%, less than about 11%, less than about 10%, less than about 9%, less than about 8%, less than about 7%, less than about 6%, or less than about 5% cellulose (w/w). Furthermore, in some embodiments, the lignocellulosic feedstock comprises lignin in an amount greater than about 10%, more typically in an amount greater than about 15% (w/w). In some embodiments, the lignocellulosic feedstock comprises small amounts of sucrose, fructose and/or starch. The lignocellulosic feedstock is generally first subjected to size reduction by methods including, but not limited to, milling, grinding, agitation, shredding, compression/expansion, or other types of mechanical action. Size reduction by mechanical action can be performed by any type of equipment adapted for the purpose, for example, but not limited to, hammer mills, tub-grinders, roll presses, refiners and hydropulpers. In some embodiments, at least 90% by weight of the particles produced from the size reduction have lengths less than between about 1/16 and about 4 in (the measurement may be a volume or a weight average length). In some embodiments, the equipment used to reduce the particle size is a hammer mill or shredder. Subsequent to size reduction, the feedstock is typically slurried in water, as this facilitates pumping of the feedstock. In some embodiments, lignocellulosic feedstocks of particle size less than about 6 inches do not require size reduction.

[00180] As used herein, the term “lignocellulosic feedstock” refers to any type of lignocellulosic biomass that is suitable for use as feedstock in saccharification reactions.

[00181] As used herein, the term “pretreated lignocellulosic feedstock,” refers to lignocellulosic feedstocks that have been subjected to physical and/or chemical processes to make the fiber more accessible and/or receptive to the actions of cellulolytic enzymes, as described above.

[00182] As used herein, the terms “lignocellulose-competent,” “lignocellulose-utilizing” and like terms refer to an organism that secretes enzymes that participate in lignin breakdown and hydrolysis. For example, in some embodiments, lignocellulose-competent fungal cells secrete one or more lignin peroxidases, manganese peroxidases, laccases and/or cellobiose dehydrogenases (CDH). These extracellular enzymes, essential for lignin degradation, are often referred to as “lignin-modifying enzymes” or “LMEs.”

[00183] A biomass substrate is said to be “pretreated” when it has been processed by some physical and/or chemical means to facilitate saccharification. As described further herein, in some embodiments, the biomass substrate is “pretreated,” or treated using methods known in the art, such as chemical pretreatment (*e.g.*, ammonia pretreatment, dilute acid pretreatment, dilute alkali pretreatment, or solvent exposure), physical pretreatment (*e.g.*, steam explosion or irradiation), mechanical pretreatment (*e.g.*, grinding or milling) and biological pretreatment (*e.g.*, application of lignin-solubilizing microorganisms) and combinations thereof, to increase the susceptibility of cellulose to hydrolysis.

[00184] In some embodiments, the substrate is slurried prior to pretreatment. In some embodiments, the consistency of the slurry is between about 2% and about 30% and more typically between about 4% and about 15%. In some embodiments, the slurry is subjected to a water and/or acid soaking operation prior to pretreatment. In some embodiments, the slurry is dewatered using any suitable method to reduce steam and chemical usage prior to pretreatment. Examples of dewatering devices include, but are not limited to pressurized screw presses (*See e.g.*, WO 2010/022511) pressurized filters and extruders.

[00185] In some embodiments, the pretreatment is carried out to hydrolyze hemicellulose, and/or a portion thereof present in lignocellulose to monomeric pentose and hexose sugars (*e.g.*, xylose, arabinose, mannose, galactose, and/or any combination thereof). In some embodiments, the pretreatment is carried out so that nearly complete hydrolysis of the hemicellulose and a small amount of conversion of cellulose to glucose occurs. In some embodiments, an acid concentration in the aqueous slurry from about 0.02% (w/w) to about 2% (w/w), or any amount therebetween, is typically used for the treatment of the cellulosic substrate. Any suitable acid finds use in these methods, including but not limited to, hydrochloric acid, nitric acid, and/or sulfuric acid. In some embodiments, the acid used

during pretreatment is sulfuric acid. Steam explosion is one method of performing acid pretreatment of biomass substrates (*See e.g.*, U.S. Patent No. 4,461,648). Another method of pretreating the slurry involves continuous pretreatment (*i.e.*, the cellulosic biomass is pumped through a reactor continuously). This methods are well-known to those skilled in the art (*See e.g.*, U.S. Patent No. 7,754,457).

[00186] In some embodiments, alkali is used in the pretreatment. In contrast to acid pretreatment, pretreatment with alkali may not hydrolyze the hemicellulose component of the biomass. Rather, the alkali reacts with acidic groups present on the hemicellulose to open up the surface of the substrate. In some embodiments, the addition of alkali alters the crystal structure of the cellulose so that it is more amenable to hydrolysis. Examples of alkali that find use in the pretreatment include, but are not limited to ammonia, ammonium hydroxide, potassium hydroxide, and sodium hydroxide. One method of alkali pretreatment is Ammonia Freeze Explosion, Ammonia Fiber Explosion or Ammonia Fiber Expansion ("AFEX" process; *See e.g.*, U.S. Patent Nos. 5,171,592; 5,037,663; 4,600,590; 6,106,888; 4,356,196; 5,939,544; 6,176,176; 5,037,663; and 5,171,592). During this process, the cellulosic substrate is contacted with ammonia or ammonium hydroxide in a pressure vessel for a sufficient time to enable the ammonia or ammonium hydroxide to alter the crystal structure of the cellulose fibers. The pressure is then rapidly reduced, which allows the ammonia to flash or boil and explode the cellulose fiber structure. In some embodiments, the flashed ammonia is then recovered using methods known in the art. In some alternative methods, dilute ammonia pretreatment is utilized. The dilute ammonia pretreatment method utilizes more dilute solutions of ammonia or ammonium hydroxide than AFEX (*See e.g.*, WO 2009/045651 and US 2007/0031953). This pretreatment process may or may not produce any monosaccharides.

[00187] An additional pretreatment process for use in the present invention includes chemical treatment of the cellulosic substrate with organic solvents, in methods such as those utilizing organic liquids in pretreatment systems (*See e.g.*, U.S. Patent No. 4,556,430). These methods have the advantage that the low boiling point liquids easily can be recovered and reused. Other pretreatments, such as the OrganosolvTM process, also use organic liquids (*See e.g.*, U.S. Patent No. 7,465,791). Subjecting the substrate to pressurized water may also be a suitable pretreatment method (*See e.g.*, Weil *et al.*, Appl. Biochem. Biotechnol., 68: 21-40

[1997]). In some embodiments, the pretreated cellulosic biomass is processed after pretreatment by any of several steps, such as dilution with water, washing with water, buffering, filtration, or centrifugation, or any combination of these processes, prior to enzymatic hydrolysis, as is familiar to those skilled in the art. The pretreatment produces a pretreated feedstock composition (*e.g.*, a “pretreated feedstock slurry”) that contains a soluble component including the sugars resulting from hydrolysis of the hemicellulose, optionally acetic acid and other inhibitors, and solids including unhydrolyzed feedstock and lignin. In some embodiments, the soluble components of the pretreated feedstock composition are separated from the solids to produce a soluble fraction. In some embodiments, the soluble fraction, including the sugars released during pretreatment and other soluble components (*e.g.*, inhibitors), is then sent to fermentation. However, in some embodiments in which the hemicellulose is not effectively hydrolyzed during the pretreatment one or more additional steps are included (*e.g.*, a further hydrolysis step(s) and/or enzymatic treatment step(s) and/or further alkali and/or acid treatment) to produce fermentable sugars. In some embodiments, the separation is carried out by washing the pretreated feedstock composition with an aqueous solution to produce a wash stream and a solids stream comprising the unhydrolyzed, pretreated feedstock. Alternatively, the soluble component is separated from the solids by subjecting the pretreated feedstock composition to a solids-liquid separation, using any suitable method (*e.g.*, centrifugation, microfiltration, plate and frame filtration, cross-flow filtration, pressure filtration, vacuum filtration, etc.). Optionally, in some embodiments, a washing step is incorporated into the solids-liquids separation. In some embodiments, the separated solids containing cellulose, then undergo enzymatic hydrolysis with cellulase enzymes in order to convert the cellulose to glucose. In some embodiments, the pretreated feedstock composition is fed into the fermentation process without separation of the solids contained therein. In some embodiments, the unhydrolyzed solids are subjected to enzymatic hydrolysis with cellulase enzymes to convert the cellulose to glucose after the fermentation process. In some embodiments, the pretreated cellulosic feedstock is subjected to enzymatic hydrolysis with cellulase enzymes.

[00188] As used herein, the term "chemical treatment" refers to any chemical pretreatment that promotes the separation and/or release of cellulose, hemicellulose, and/or lignin.

[00189] As used herein, the term "physical pretreatment" refers to any pretreatment that promotes the separation and/or release of cellulose, hemicellulose, and/or lignin from cellulosic material.

[00190] As used herein, the term "mechanical pretreatment" refers to any mechanical means for treating biomass, including but not limited to various types of grinding or milling (*e.g.*, dry milling, wet milling, or vibratory ball milling).

[00191] As used herein, the term "biological pretreatment" refers to any biological pretreatment that promotes the separation and/or release of cellulose, hemicellulose, and/or lignin from cellulosic material.

[00192] As used herein, the term "recovered" refers to the harvesting, isolating, collecting, or recovering of protein from a cell and/or culture medium. In the context of saccharification, it is used in reference to the harvesting of fermentable sugars produced during the saccharification reaction from the culture medium and/or cells. In the context of fermentation, it is used in reference to harvesting the fermentation product from the culture medium and/or cells. Thus, a process can be said to comprise "recovering" a product of a reaction (such as a soluble sugar recovered from saccharification) if the process includes separating the product from other components of a reaction mixture subsequent to at least some of the product being generated in the reaction.

[00193] As used herein, "increasing" the yield of a product (such as a fermentable sugar) from a reaction occurs when a particular component of interest is present during the reaction (*e.g.*, enzyme) causes more product to be produced, compared with a reaction conducted under the same conditions with the same substrate and other substituents, but in the absence of the component of interest (*e.g.*, without enzyme).

[00194] As used herein, a reaction is said to be "substantially free" of a particular enzyme if the amount of that enzyme compared with other enzymes that participate in catalyzing the reaction is less than about 2%, about 1%, or about 0.1% (wt/wt).

[00195] As used herein, "fractionating" a liquid (*e.g.*, a culture broth) means applying a separation process (*e.g.*, salt precipitation, column chromatography, size exclusion, and filtration) or a combination of such processes to provide a solution in which a desired protein (*e.g.*, a cellulase enzyme, and/or a combination thereof) comprises a greater percentage of total protein in the solution than in the initial liquid product.

[00196] As used herein, the term “enzymatic hydrolysis,” refers to the hydrolysis of a substrate by an enzyme. In some embodiments, the hydrolysis comprises methods in which at least one enzyme is contacted with at least one substrate to produce an end product. In some embodiments, the enzymatic hydrolysis methods comprise at least one cellulase and at least one glycosidase enzyme and/or a mixture glycosidases that act on polysaccharides, (*e.g.*, cellulose), to convert all or a portion thereof to fermentable sugars. “Hydrolyzing” and/or “hydrolysis” of cellulose or other polysaccharide occurs when at least some of the glycosidic bonds between two monosaccharides present in the substrate are hydrolyzed, thereby detaching from each other the two monomers that were previously bonded.

[00197] It is intended that the enzymatic hydrolysis be carried out with any suitable type of enzyme(s) capable of hydrolyzing at least one substrate to at least one end-product. In some embodiments, the substrate is cellulose, while in some other embodiments, it is lignocelluloses, and in still further embodiments, it is another composition (*e.g.*, starch). In some embodiments, the end-product comprises at least one fermentable sugar. It is further intended that the enzymatic hydrolysis encompass processes carried out with any suitable type of cellulase enzymes capable of hydrolyzing the cellulose to glucose, regardless of their source. It is intended that any suitable source of enzyme will find use in the present invention, including but not limited to enzymes obtained from fungi, such as *Trichoderma* spp., *Aspergillus* spp., *Hypocrea* spp., *Humicola* spp., *Neurospora* spp., *Orpinomyces* spp., *Gibberella* spp., *Emericella* spp., *Chaetomium* spp., *Chrysosporium* spp., *Fusarium* spp., *Penicillium* spp., *Magnaporthe* spp., *Phanerochaete* spp., *Trametes* spp., *Lentinula edodes*, *Gleophyllum trabeiu*, *Ophiostoma piliferum*, *Corpinus cinereus*, *Geomyces pannorum*, *Cryptococcus laurentii*, *Aureobasidium pullulans*, *Amorphotheca resinae*, *Leucosporidium scotti*, *Cunninghamella elegans*, *Thermomyces lanuginosus*, *Myceliophthora thermophila*, and *Sporotrichum thermophile*, as well as those obtained from bacteria of the genera *Bacillus*, *Thermomyces*, *Clostridium*, *Streptomyces* and *Thermobifida*.

[00198] In some embodiments, the enzymatic hydrolysis is carried out at a pH and temperature that is at or near the optimum for the cellulase enzymes being used. For example, in some embodiments, the enzymatic hydrolysis is carried out at about 30°C to about 75°C, or any suitable temperature therebetween, for example a temperature of about 30°C, about 35°C, about 40°C, about 45°C, about 50°C, about 55°C, about 60°C, about

65°C, about 70°C, about 75°C, or any temperature therebetween, and a pH of about 3.5 to about 7.5, or any pH therebetween (*e.g.*, about 3.5, about 4.0, about 4.5, about 5.0, about 5.5, about 6.0, about 6.5, about 7.0, about 7.5, or any suitable pH therebetween). In some embodiments, the initial concentration of cellulose, prior to the start of enzymatic hydrolysis, is preferably about 0.1% (w/w) to about 20% (w/w), or any suitable amount therebetween (*e.g.*, about 0.1%, about 0.5%, about 1%, about 2%, about 4%, about 6%, about 8%, about 10%, about 12%, about 14%, about 15%, about 18%, about 20%, or any suitable amount therebetween). In some embodiments, the combined dosage of all cellulase enzymes is about 0.001 to about 100 mg protein per gram cellulose, or any suitable amount therebetween (*e.g.*, about 0.001, about 0.01, about 0.1, about 1, about 5, about 10, about 15, about 20, about 25, about 30, about 40, about 50, about 60, about 70, about 80, about 90, about 100 mg protein per gram cellulose or any amount therebetween). The enzymatic hydrolysis is carried out for any suitable time period. In some embodiments, the enzymatic hydrolysis is carried out for a time period of about 0.5 hours to about 200 hours, or any time therebetween (*e.g.*, about 2 hours to about 100 hours, or any suitable time therebetween). For example, in some embodiments, it is carried out for about 0.5, about 1, about 2, about 5, about 7, about 10, about 12, about 14, about 15, about 20, about 25, about 30, about 35, about 40, about 45, about 50, about 55, about 60, about 65, about 70, about 75, about 80, about 85, about 90, about 95, about 100, about 120, about 140, about 160, about 180, about 200, or any suitable time therebetween.

[00199] In some embodiments, the enzymatic hydrolysis is batch hydrolysis, continuous hydrolysis, and/or a combination thereof. In some embodiments, the hydrolysis is agitated, unmixed, or a combination thereof. The enzymatic hydrolysis is typically carried out in a hydrolysis reactor. The cellulase enzyme composition is added to the pretreated lignocellulosic substrate prior to, during, or after the addition of the substrate to the hydrolysis reactor. Indeed it is not intended that reaction conditions be limited to those provided herein, as modifications are well-within the knowledge of those skilled in the art. In some embodiments, following cellulose hydrolysis, any insoluble solids present in the resulting lignocellulosic hydrolysate, including but not limited to lignin, are removed using conventional solid-liquid separation techniques prior to any further processing. In some embodiments, these solids are burned to provide energy for the entire process.

[00200] As used herein, the “total available cellulose” is the amount (wt %) of cellulose that is accessible to enzymatic hydrolysis. Total available cellulose is typically equal to, or very close to being equal to, the amount of initial cellulose present in a hydrolysis reaction.

[00201] As used herein, the “residual cellulose” is the portion (wt %) of the total available cellulose in the hydrolysis mixture that remains unhydrolyzed. Residual cellulose can be measured using any suitable method known in the art. For example, it can be directly measured using IR spectroscopy, or it can be measured by determining the amount of glucose generated by concentrated acid hydrolysis of the residual solids.

[00202] As used herein, the “total hydrolyzed cellulose” is the portion of the total available cellulose that is hydrolyzed in the hydrolysis mixture. For example, the total hydrolyzed cellulose can be calculated as the difference between the “total available cellulose” and the “residual cellulose.”

[00203] As used herein, the “theoretical maximum glucose yield” is the maximum amount (wt %) of glucose that could be produced under given conditions from the total available cellulose.

[00204] As used herein, “Gmax” refers to the maximum amount (wt %) of glucose that could be produced from the total hydrolyzed cellulose. Gmax can be calculated, for example, by directly measuring the amount of residual cellulose remaining at the end of a reaction under a given reaction conditions, subtracting the amount of residual cellulose from the total available cellulose to determine the total hydrolyzed cellulose, and then calculating the amount of glucose that could be produced from the total hydrolyzed cellulose.

[00205] It will be appreciated by those skilled in the art that when calculating theoretical values such as Gmax and theoretical maximum glucose yield, the mass of two hydrogen atoms and one oxygen atom that are added to the glucose molecule in the course of the hydrolysis reaction are taken into account. For example, when a polymer of “n” glucose units is hydrolyzed, (n-1) units of water are added to the glucose molecules formed in the hydrolysis, so the weight of the glucose produced is about 10% greater than the weight of cellulose consumed in the hydrolysis (*e.g.*, hydrolysis of 1 g cellulose would produce about 1.1 g glucose).

[00206] Thus, as an example, where 5 g of total available cellulose is present at the beginning of a hydrolysis reaction, and 2 g of residual cellulose remains after the reaction,

the total hydrolyzed cellulose is 3 g cellulose. A theoretical maximum glucose yield of 100% (w/w) under the reaction conditions is about 5.5 g of glucose. Gmax is calculated based on the 3 g of cellulose that was released or converted in the reaction by hydrolysis. Thus, in this example, a Gmax of 100% (w/w) is about 3.3 g of glucose. Cellulose levels, either the total available amount present in the substrate or the amount of unhydrolyzed or residual cellulose, can be quantified by any of a variety of methods known in the art, such as by IR spectroscopy or by measuring the amount of glucose generated by concentrated acid hydrolysis of the cellulose (*See e.g.*, U.S. Patent Nos. 6,090,595 and 7,419,809).

[00207] As used herein, the term “undissolved solids” refers to solid material which is suspended, but not dissolved, in a liquid. As is well known in the art, the concentration of suspended or undissolved solids can be determined by any suitable method (*e.g.*, by filtering a sample of the slurry using glass microfiber filter paper, washing the filter cake with water, and drying the cake overnight at about 105°C).

[00208] As used herein, the terms “unhydrolyzed solids,” “unconverted solids,” and the like refer to cellulose that is not digested by the cellulase enzyme(s), as well as non-cellulosic, or other, materials that are inert to the cellulase enzyme(s), present in the feedstock.

[00209] As used herein, the term “by-product” refers to an organic molecule that is an undesired product of a particular process (*e.g.*, saccharification).

[00210] In some embodiments, the present invention provides fungal organisms and methods for the conversion of cellulose to glucose. In some embodiments, the conversion is improved by genetically modifying a fungus to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell. Prior to this invention, it was generally believed that cellobiose dehydrogenase enhances the rate of cellulose hydrolysis by reducing the concentration of cellobiose, which is a potent inhibitor of some cellulase components (*See e.g.*, Mansfield *et al.*, Appl. Environ. Microbiol., 63: 3804-3809 [1997]; Igarishi *et al.*, Eur. J. Biochem., 253: 101-106 [1998]). Further, cellobiose dehydrogenase has been reported as playing a critical role contributing to synergistic enhancement in degradation of cellulose by preventing product inhibition of hydrolysis (*See e.g.*, Hai *et al.*, J. Appl. Glycosci., 49:9-17 [2002]). As a result, genetic modification of *Trametes versicolor* (*See*, Archibald, 7th International Conference on

Biotechnology in the Pulp and Paper Industry, Vol. B: B225-B228 [1998]) and *Coriolus hirsutus* (See, U.S. Pat. Appln. Publn. No. 2005/0181485) has been carried out in order to produce cellulase systems with reduced cellolytic activities for pulp and paper applications. It was also generally believed that cellobiose dehydrogenase was useful in delignifying lignocellulose, and thereby enhance cellulose degradation. Recently, it has been reported that cellobiose dehydrogenases can enhance the activity of cellulolytic enhancing proteins from Glycosyl Hydrolase Family 61 (See e.g., WO 2010/080532A1).

[00211] Contrary to general understanding in the art, the present invention provides fungal cells with genetic modification (such as deletion) of glucose and/or cellobiose oxidizing enzyme-encoding genes in cellulase-producing fungal cells that results in an improvement in the yield of fermentable sugars from enzyme mixtures secreted by the genetically modified cells. Thus, reduction of glucose and/or cellobiose oxidizing enzyme activity secreted by a cellulase-producing organism results in a mixture of cellulase enzymes that can improve yield of fermentable sugars during enzymatic hydrolysis of cellulose-containing substrates.

[00212] The present invention provides a fungal cell that has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell, wherein the fungal cell is an Ascomycete belonging to the subdivision Pezizomycotina, and where the fungal cell is capable of secreting a cellulase-containing enzyme mixture. Also provided herein is a fungal cell that has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell and to increase the expression of at least one saccharide hydrolyzing enzyme, wherein the fungal cell is a Basidiomycete belonging to the class Agaricomycetes, and where the fungal cell is capable of secreting a cellulase-containing enzyme mixture. Also provided herein is a fungal cell that has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell, wherein the fungal cell is a Basidiomycete, and where the fungal cell is capable of secreting a cellulase-containing enzyme mixture. In some embodiments, the endogenous glucose and/or cellobiose oxidizing enzyme is a cellobiose dehydrogenase, while in some alternative embodiments, the endogenous glucose and/or cellobiose oxidizing enzyme is an enzyme other than cellobiose dehydrogenase.

[00213] In some embodiments, the fungal cell is capable of secreting an enzyme mixture comprising at least two or more cellulase enzymes. In some embodiments, the Basidiomycete is a species of *Pleurotus*, *Peniophora*, *Trametes*, *Athelia*, *Sclerotium*, *Termitomyces*, *Flammulina*, *Chrysosporium*, *Coniphora*, *Ganoderma*, *Pycnoporus*, *Ceriporiopsis*, *Phanerochaete*, *Gloeophyllum*, *Hericium*, *Heterobasidion*, *Gelatoporia*, *Lepiota*, or *Irpex*. In some embodiments, the Ascomycete is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Neurospora*, *Sordaria*, *Podospora*, *Magnaporthe*, *Fusarium*, *Gibberella*, *Botryotinia*, *Humicola*, *Neosartorya*, *Pyrenophora*, *Phaeosphaeria*, *Sclerotinia*, *Chaetomium*, *Nectria*, *Verticillium*, or *Aspergillus*.

[00214] The present invention also provides a fungal cell that has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell, wherein the fungal cell is a Basidiomycete species *Pleurotus*, *Peniophora*, *Athelia*, *Sclerotium*, *Termitomyces*, *Flammulina*, *Coniphora*, *Ganoderma*, *Pycnoporus*, *Ceriporiopsis*, *Phanerochaete*, *Gloeophyllum*, *Hericium*, *Heterobasidion*, *Gelatoporia*, *Lepiota*, or *Irpex*.

[00215] In some embodiments, the fungal cell has been genetically modified to reduce the amount of endogenous cellobiose dehydrogenase activity that is secreted by the cell. In some embodiments, the fungal cell has been genetically modified to reduce the amount of endogenous glucose oxidase activity that is secreted by the cell. In some embodiments, the fungal cell has been genetically modified to reduce the amount of endogenous pyranose oxidase activity that is secreted by the cell. In some embodiments, the fungal cell has been genetically modified to reduce the amount of endogenous glucooligosaccharide oxidase activity that is secreted by the cell. In some embodiments, the fungal cell has been genetically modified to reduce the amount of endogenous pyranose dehydrogenase activity that is secreted by the cell. In some embodiments, the fungal cell has been genetically modified to reduce the amount of endogenous glucose dehydrogenase activity that is secreted by the cell.

[00216] In some embodiments, the fungal cell is a species of *Myceliophthora*, *Thielavia*, *Chrysosporium*, *Sporotrichum*, *Corynascus*, *Acremonium*, *Chaetomium*, *Ctenomyces*, *Scytalidium*, *Talaromyces*, or *Thermoascus*. In some embodiments the fungal cell is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Corynascus*, *Acremonium*,

Chaetomium, or Talaromyces. In some embodiments, the fungal cell is Sporotrichum thermophile, Sporotrichum cellulophilum, Thielavia terrestris, Corynascus heterothallicus, Thielavia heterothallica, Chaetomium globosum, Talaromyces stipitatus, or Myceliophthora thermophila. In some embodiments, the fungal cell is an isolated fungal cell.

[00217] In some embodiments, the fungal cell has been genetically modified to reduce the amount of the endogenous glucose and/or cellobiose oxidizing enzyme that is secreted by the cell. Thus, in some embodiments, the fungal cell has been genetically modified to disrupt the secretion signal peptide of the glucose and/or cellobiose oxidizing enzyme. In some embodiments, the fungal cell has been genetically modified to reduce the amount of the endogenous glucose and/or cellobiose oxidizing enzyme that is expressed by the cell. For example, the fungal cell can be genetically modified to disrupt a translation initiation sequence or to introduce a frameshift mutation in the transcript encoding the endogenous glucose and/or cellobiose oxidizing enzyme. In some other embodiments, the fungal cell has been genetically modified to reduce the transcription level of a gene encoding the endogenous glucose and/or cellobiose oxidizing enzyme. For example, the fungal cell can be genetically modified to disrupt the promoter of a gene encoding the endogenous glucose and/or cellobiose oxidizing enzyme. In some embodiments, the fungal cell has been genetically modified to at least partially delete a gene encoding the endogenous glucose and/or cellobiose oxidizing enzyme. In some other embodiments, the fungal cell has been genetically modified to reduce the catalytic efficiency of the endogenous glucose and/or cellobiose oxidizing enzyme. In some embodiments, the fungal cell has been genetically modified to mutate one or more residues in an active site of the glucose and/or cellobiose oxidizing enzyme. In some embodiments, the fungal cell has been genetically modified to mutate one or more residues in a heme binding domain of the glucose and/or cellobiose oxidizing enzyme.

[00218] In some embodiments, the glucose and/or cellobiose oxidizing enzyme is glucose oxidase (EC 1.1.3.4). In some other embodiments, the glucose and/or cellobiose oxidizing enzyme is cellobiose dehydrogenase (EC 1.1.99.18). In some other embodiments, the glucose and/or cellobiose oxidizing enzyme is pyranose oxidase (EC 1.1.3.10). In some other embodiments, the glucose and/or cellobiose oxidizing enzyme is glucooligosaccharide oxidase (EC 1.1.99.B3). In some additional embodiments, the glucose and/or cellobiose

oxidizing enzyme is pyranose dehydrogenase (EC 1.1.99.29). In some further embodiments, the glucose and/or cellobiose oxidizing enzyme is glucose dehydrogenase (EC 1.1.99.10). In some embodiments, the glucose and/or cellobiose oxidizing enzyme comprises an amino acid sequence that is at least about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or at least about 99% identical to SEQ ID NOS:2, 4, 6, 8, 10, 12, 14, and/or 16.

[00219] In some embodiments, the cell has been genetically modified to reduce the amount of glucose and/or cellobiose oxidizing enzyme activity of two or more endogenous glucose and/or cellobiose oxidizing enzymes that are secreted by the cell. In certain such embodiments, a first of the two or more the glucose and/or cellobiose oxidizing enzymes comprises an amino acid sequence that is at least about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or at least about 99% identical to SEQ ID NO:2, 4, 6, 8, 10, 12, 14, and/or 16, and a second of the two or more the glucose and/or cellobiose oxidizing enzymes comprises an amino acid sequence that is at least about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or at least about 99% identical to SEQ ID NOS:2, 4, 6, 8, 10, 12, 14, and/or 16.

[00220] In some embodiments, the fungal cell further comprises at least one gene encoding at least one cellulose degrading enzyme that is heterologous to the fungal cell. For example, the fungal cell can overexpress a homologous or heterologous gene encoding a cellulose degrading enzyme such as beta-glucosidase. In some embodiments, the fungal cell overexpresses beta-glucosidase and has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell.

[00221] The present invention also provides enzyme mixtures comprising two or more cellulose hydrolyzing enzymes, wherein at least one of the two or more cellulose hydrolyzing enzymes is expressed by a fungal cell as described herein. For example, in some embodiments, the fungal cell is a cell that has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell, wherein the fungal cell is an Ascomycete belonging to the subdivision Pezizomycotina. In some other embodiments, the fungal cell has been genetically modified

to reduce the activity of an endogenous glucose and/or cellobiose oxidizing enzyme that is secreted by the cell and to increase the expression of at least one saccharide hydrolyzing enzyme, wherein the fungal cell is a Basidiomycete belonging to the class Agaricomycetes. In some embodiments, the Basidiomycete is a species of *Pleurotus*, *Peniophora*, *Trametes*, *Athelia*, *Sclerotium*, *Termitomyces*, *Flammulina*, *Coniophora*, *Ganoderma*, *Pycnoporus*, *Ceriporiopsis*, *Phanerochaete*, *Gloeophyllum*, *Herichium*, *Heterobasidion*, *Gelatoporia*, *Lepiota*, or *Irpex*. In some embodiments, the Ascomycete is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Neurospora*, *Sordaria*, *Podospora*, *Magnaporthe*, *Fusarium*, *Gibberella*, *Botryotinia*, *Humicola*, *Neosartorya*, *Pyrenophora*, *Phacosphaeria*, *Sclerotinia*, *Chaetomium*, *Nectria*, *Verticillium*, or *Aspergillus*. In some embodiments, the fungal cell can be a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Corynascus*, *Acremonium*, *Chaetomium*, *Ctenomyces*, *Scytalidium*, *Talaromyces*, or *Thermoascus*. In some embodiments the fungal cell is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Corynascus*, *Acremonium*, or *Chaetomium*. In some embodiments, the fungal cell is *Sporotrichum cellulophilum*, *Thielavia terrestris*, *Corynascus heterothallicus*, *Thielavia heterothallica*, or *Myceliophthora thermophila*.

[00222] In some embodiments, the fungal cell is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Chrysosporium*, *Corynascus*, *Acremonium*, *Chaetomium*, *Ctenomyces*, *Scytalidium*, *Talaromyces*, or *Thermoascus*. In some embodiments the fungal cell is a species of *Myceliophthora*, *Chrysosporium*, *Thielavia*, *Sporotrichum*, *Corynascus*, *Acremonium*, or *Chaetomium*. In some embodiments, the fungal cell is *Sporotrichum cellulophilum*, *Thielavia terrestris*, *Corynascus heterothallicus*, *Thielavia heterothallica*, *Chaetomium globosum*, *Talaromyces stipitatus*, or *Myceliophthora thermophila*. In some embodiments, the fungal cell is an isolated fungal cell.

[00223] In some additional embodiments, the enzyme mixture is a cell-free mixture. In some embodiments, a substrate of the enzyme mixture comprises pretreated lignocellulose. In some additional embodiments, the pretreated lignocellulose comprises lignocellulose treated by a treatment method selected from acid pretreatment, ammonia pretreatment, steam explosion, organic solvent extraction, and/or any other suitable pretreatment method(s). In some embodiments, the enzyme mixture further comprises a cellulose degrading enzyme that is heterologous to the fungal cell. In some embodiments, at

least one of the two or more cellulose hydrolyzing enzymes is expressed by an isolated fungal cell.

[00224] The present invention also provides methods for generating cellobiose and/or glucose comprising contacting a cellulosic substrate with the enzyme mixture described herein. For example, in some embodiments, the methods comprise contacting cellulose with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes, wherein at least one of the two or more cellulose hydrolyzing enzymes is expressed by a fungal cell as described herein. In some embodiments, the methods comprise contacting cellulose with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes, wherein at least one of the two or more cellulose hydrolyzing enzymes is expressed by a cell that has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell, wherein the fungal cell is an Ascomycete belonging to the subdivision Pezizomycotina.

[00225] In some other embodiments, the fungal cell has been genetically modified to reduce the activity of an endogenous glucose and/or cellobiose oxidizing enzyme that is secreted by the cell and to increase the expression of at least one saccharide hydrolyzing enzyme, wherein the fungal cell is a Basidiomycete belonging to the class Agaricomycetes. In some embodiments, the Basidiomycete is a species of *Pleurotus*, *Peniophora*, *Trametes*, *Athelia*, *Sclerotium*, *Termitomyces*, *Flammulina*, *Coniophora*, *Ganoderma*, *Pycnoporus*, *Ceriporiopsis*, *Phanerochaete*, *Gloeophyllum*, *Herichium*, *Heterobasidion*, *Gelatoporia*, *Lepiota*, or *Irpex*. In some embodiments, the Ascomycete is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Neurospora*, *Sordaria*, *Podospora*, *Magnaporthe*, *Fusarium*, *Gibberella*, *Botryotinia*, *Humicola*, *Neosartorya*, *Pyrenophora*, *Phacosphaeria*, *Sclerotinia*, *Chaetomium*, *Nectria*, *Verticillium*, or *Aspergillus*. In some embodiments, the fungal cell is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Corynascus*, *Acremonium*, *Chaetomium*, *Ctenomyces*, *Scytalidium*, *Talaromyces*, or *Thermoascus*. In some embodiments the fungal cell is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Corynascus*, *Acremonium*, or *Chaetomium*. In some embodiments, the fungal cell is *Sporotrichum cellulophilum*, *Thielavia terrestris*, *Corynascus heterothallicus*, *Thielavia heterothallica*, or *Myceliophthora thermophila*.

[00226] In some embodiments, the fungal cell is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Corynascus*, *Acremonium*, *Chaetomium*, *Ctenomyces*, *Scytalidium*, *Talaromyces*, or *Thermoascus*. In some embodiments the fungal cell is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Corynascus*, *Acremonium*, or *Chaetomium*. In some embodiments, the fungal cell is *Sporotrichum cellulophilum*, *Thielavia terrestris*, *Corynascus heterothallicus*, *Thielavia heterothallica*, *Chaetomium globosum*, *Talaromyces stipitatus*, or *Myceliophthora thermophila*. In some embodiments, the fungal cell is an isolated fungal cell.

[00227] The present invention provides methods for generating cellobiose and/or glucose comprising contacting a cellulose substrate with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes to generate glucose and/or cellobiose, wherein at least one of the cellulose hydrolyzing enzymes is endogenous to a fungus that is an Ascomycete belonging to the subdivision Pezizomycotina, and wherein the enzyme mixture is characterized in that, when the enzyme mixture is contacted with cellobiose and/or glucose, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellobiose and/or glucose is oxidized after 10 hours.

[00228] The present invention also provides methods for generating cellobiose and/or glucose comprising contacting a cellulose substrate with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes to generate glucose and/or cellobiose, wherein at least one of the cellulose hydrolyzing enzymes is endogenous to a fungus that is a Basidiomycete belonging to the class Agaricomycetes, and wherein the enzyme mixture is characterized in that, when the enzyme mixture is contacted with cellobiose and/or glucose, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellobiose and/or glucose is oxidized after 10 hours.

[00229] The present invention further provides methods for generating cellobiose and/or glucose comprising contacting a cellulose substrate with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes to generate glucose and/or

cellobiose, wherein at least one of the cellulose hydrolyzing enzymes is endogenous to a fungus of a species of Myceliophthora, Thielavia, Sporotrichum, Corynascus, Acremonium, Chaetomium or Ctenomyces, Scytalidium or Thermoascus, and wherein the enzyme mixture is characterized in that, when the enzyme mixture is contacted with cellobiose and/or glucose, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellobiose and/or glucose is oxidized after 10 hours.

[00230] In some embodiments of the methods provided herein, when the enzyme mixture is contacted with a cellulose substrate, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or 20% (wt %) of the cellobiose and/or glucose resulting from the hydrolysis of the cellulose substrate is oxidized. For example, when the enzyme mixture is contacted with a cellulose substrate, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19% or about 20% (wt %) of the cellobiose and/or glucose resulting from the hydrolysis of the cellulose substrate is oxidized to form cellobionolactone, cellobionic acid, gluconolactone, gluconate or gluconic acid after a period of time during which hydrolysis occurs. For example, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of cellobiose and/or glucose is oxidized after about 1, about 5, about 10, about 20, about 30, about 40, about 50 or about 60 minutes, or after about 1.5, about 2, about 3, about 4, about 5, about 6, about 7, about 8, about 9, about 10, about 12, about 14, about 16, about 18, about 20, about 25, about 30, about 35, about 40, about 45, about 50, about 55, about 60, about 65, about 70, about 75, about 80, about 85, about 90, about 95, about 100, about 105, about 110, about 115, about 120, about 125, about 130, about 135, about 140, about 145, about 150, about 155, about 160, about 165, about 170, about 175, about 180, about 185, about 190, about 195, about 200, about 205, about 210, about 215, about 220, about 225, about 230, about 235, about 240, about 245, about

250, about 255, about 260, about 265, about 270, about 275, about 280, about 285, about 290, about 395, about 300 hours, or longer.

[00231] The present invention provides methods for generating cellobiose and/or glucose comprising contacting a cellulose substrate with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes to generate glucose and/or cellobiose, wherein at least one of the cellulose hydrolyzing enzymes is endogenous to a fungus that is an Ascomycete belonging to the subdivision Pezizomycotina, and wherein, of the cellulose hydrolyzed by the enzyme mixture, at least about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 98%, about 99% or 100% (wt %) is present in the form of cellobiose and/or glucose.

[00232] The present invention also provides methods for generating cellobiose and/or glucose comprising contacting a cellulose substrate with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes to generate glucose and/or cellobiose, wherein at least one of the cellulose hydrolyzing enzymes is endogenous to a fungus that is a Basidiomycete belonging to the class Agaricomycetes, and wherein, of the cellulose hydrolyzed by the enzyme mixture, at least about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 98%, about 99%, or about 100% (wt %) is present in the form of cellobiose and/or glucose.

[00233] In some embodiments, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellulose hydrolyzed by the enzyme mixture is present in the form of gluconolactone or gluconic acid. In some embodiments, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellulose hydrolyzed by the enzyme mixture is present in the form of gluconolactone, gluconic acid, cellobionolactone or cellobionic acid.

[00234] Further provided herein are methods for generating cellobiose and/or glucose comprising contacting a cellulose substrate with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes to generate glucose and/or cellobiose, wherein at least one of the cellulose hydrolyzing enzymes is endogenous to a fungus of a species of *Pleurotus*, *Peniophora*, *Trametes*, *Athelia*, *Sclerotium*, *Termitomyces*, *Flammulina*, *Coniophora*, *Ganoderma*, *Pycnoporus*, *Ceriporiopsis*, *Phanerochaete*, *Gloeophyllum*, *Herichium*, *Heterobasidion*, *Gelatoporia*, *Lepiota*, or *Irpex*, and wherein, of the cellulose hydrolyzed by the enzyme mixture, at least about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 98%, about 99%, or about 100% (wt %) is present in the form of cellobiose and/or glucose.

[00235] Further provided herein are methods for generating cellobiose and/or glucose comprising contacting a cellulose substrate with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes to generate glucose and/or cellobiose, wherein at least one of the cellulose hydrolyzing enzymes is endogenous to a fungus of a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Neurospora*, *Sordaria*, *Podospora*, *Magnaporthe*, *Fusarium*, *Gibberella*, *Botryotinia*, *Humicola*, *Neosartorya*, *Pyrenophora*, *Phaeosphaeria*, *Sclerotinia*, *Chaetomium*, *Nectria*, *Verticillium*, or *Aspergillus*, and wherein, of the cellulose hydrolyzed by the enzyme mixture, at least about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 98%, about 99%, or about 100% (wt %) is present in the form of cellobiose and/or glucose.

[00236] In some embodiments, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellulose hydrolyzed by the enzyme mixture is present in the form of gluconolactone or gluconic acid. In some embodiments, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellulose hydrolyzed by the enzyme mixture is present in the form of gluconolactone, gluconic acid, cellobionolactone or cellobionic acid.

[00237] In some embodiments, the methods result in an increased yield of glucose and/or cellobiose from the hydrolyzed cellulose and a decreased oxidation of the glucose and/or cellobiose to oxidized sugar products, such as gluconolactone, gluconate, gluconic acid, cellobionolactone, and/or cellobionic acid from the hydrolyzed cellulose. In some embodiments, the methods result in an increased yield of glucose and/or cellobiose from the hydrolyzed cellulose and a decreased oxidation of the glucose and/or cellobiose to oxidized sugar products, such as gluconolactone, gluconate, gluconic acid, cellobionolactone, and/or cellobionic acid from the hydrolyzed cellulose, relative to an enzyme mixture with an unmodified amount of glucose and/or cellobiose oxidizing enzyme activity, or relative to a parental enzyme mixture.

[00238] In some embodiments, the present invention provides methods for producing cellobiose and/or glucose from cellulose comprising treating a cellulose substrate with an enzyme mixture to generate glucose and/or cellobiose, wherein the enzyme mixture is modified relative to a secreted enzyme mixture from a wild type or reference (e.g., parental) fungal cell to be at least partially deficient in glucose and/or cellobiose oxidizing enzyme activity.

[00239] In some aspects of the above embodiments, the enzyme mixture is a cell-free mixture. In some other aspects, the cellulose substrate comprises pretreated lignocellulose. In some embodiments, the pretreated lignocellulose comprises lignocellulose treated by a treatment method selected from acid pretreatment, ammonia pretreatment, steam explosion, and organic solvent extraction.

[00240] In some aspects of the above embodiments, the methods further comprise fermentation of the glucose to an end product such as a fuel alcohol or a precursor industrial chemical. In some aspects, the fuel alcohol is ethanol or butanol. Accordingly, in some embodiments, increased glucose yield can result in lower fuel production costs. In some aspects, the methods comprise contacting cellulose with an enzyme mixture that further comprises a cellulose degrading enzyme that is heterologous to the fungal cell.

[00241] In some embodiments, the enzyme mixture is produced by a fungal cell that has been genetically modified to reduce the amount of one or more endogenous glucose and/or cellobiose oxidizing enzymes that is secreted by the cell.

[00242] In some embodiments, the enzyme mixture is subjected to a purification process to selectively remove one or more glucose and/or cellobiose oxidizing enzymes from the enzyme mixture. In some such aspects, the purification process comprises selective precipitation to separate the glucose and/or cellobiose oxidizing enzymes from other enzymes present in the enzyme mixture.

[00243] In some embodiments, the enzyme mixture comprises an inhibitor of one or more glucose and/or cellobiose oxidizing enzymes. In some embodiments, the inhibitor includes a broad-spectrum oxidase inhibitor selected from sodium azide, potassium cyanide and a number of metal anions such as Ag^+ , Hg^{2+} , Zn^{2+} . In some additional embodiments, the inhibitor includes a specific inhibitor of cellobiose dehydrogenase (EC 1.1.99.18) such as gentiobiose, lactobionolactone, 5-lactone, cellobionolactone, 5-lactone, tri-N-acetylchitotriose, methyl-beta-D-cellobioside, 2,2-bipyridine and cytochrome C.

[00244] In some embodiments, the enzyme mixture comprises at least one beta-glucosidase. In some additional embodiments, the enzyme mixture comprises at least one cellulase enzyme selected from endoglucanases (EGs), β -glucosidases (BGLs), Type 1 cellobiohydrolases (CBH1s), Type 2 cellobiohydrolases (CBH2s), and /or glycoside hydrolase 61s (GH61s), and/or variants of said cellulase enzyme.

[00245] The present invention also provides enzyme mixtures comprising two or more cellulose hydrolyzing enzymes, at least one of the cellulose hydrolyzing enzymes being endogenous to a fungal cell, wherein the fungal cell is an Ascomycete belonging to the subdivision Pezizomycotina and wherein the enzyme mixtures are characterized in that, when the enzyme mixtures are contacted with cellobiose and/or glucose, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellobiose and/or glucose is oxidized after 10 hours. In some aspects of the above embodiments, the fungal cell is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Corynascus*, *Acremonium*, *Chaetomium*, *Ctenomyces*, *Scytalidium*, *Talaromyces* or *Thermoascus*.

[00246] The present invention also provides enzyme mixtures comprising two or more cellulose hydrolyzing enzymes, at least one of the cellulose hydrolyzing enzymes being endogenous to a fungal cell, wherein the fungal cell is a Basidiomycete belonging to the class

Agaricomycetes and wherein the enzyme mixtures are characterized in that, when the enzyme mixtures are contacted with cellobiose and/or glucose, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellobiose and/or glucose is oxidized after 10 hours.

[00247] The present invention further provides enzyme mixtures comprising two or more cellulose hydrolyzing enzymes, at least one of the cellulose hydrolyzing enzymes being endogenous to a fungal cell, wherein the fungal cell is *Pleurotus*, *Peniophora*, *Trametes*, *Athelia*, *Sclerotium*, *Termitomyces*, *Flammulina*, *Coniophora*, *Ganoderma*, *Pycnoporus*, *Ceriporiopsis*, *Phanerochaete*, *Gloeophyllum*, *Herichium*, *Heterobasidion*, *Gelatoporia*, *Lepiota*, or *Irpex*, and wherein the enzyme mixtures are characterized in that, when the enzyme mixtures are contacted with cellobiose and/or glucose, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellobiose and/or glucose is oxidized after 10 hours.

[00248] The present invention also provides enzyme mixtures comprising two or more cellulose hydrolyzing enzymes, at least one of the cellulose hydrolyzing enzymes being endogenous to a fungal cell, wherein the fungal cell is *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Neurospora*, *Sordaria*, *Podospira*, *Magnaporthe*, *Fusarium*, *Gibberella*, *Botryotinia*, *Humicola*, *Neosartorya*, *Pyrenophora*, *Phaeosphaeria*, *Sclerotinia*, *Chaetomium*, *Nectria*, *Verticillium*, or *Aspergillus*, and wherein the enzyme mixtures are characterized in that, when the enzyme mixtures are contacted with cellobiose and/or glucose, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellobiose and/or glucose is oxidized after 10 hours.

[00249] In some aspects of the above embodiments, the fungal cell has been genetically modified to reduce the amount of one or more endogenous glucose and/or cellobiose oxidizing enzymes that is secreted by the cell. In some embodiments, the enzyme

mixtures are cell-free mixtures. In some embodiments, the enzyme mixtures contain a beta-glucosidase. In some additional embodiments, the enzyme mixtures comprise at least one cellulase enzyme selected from endoglucanases (EGs), β -glucosidases (BGLs), Type 1 cellobiohydrolases (CBH1s), Type 2 cellobiohydrolases (CBH2s), and /or glycoside hydrolase 61s (GH61s), and/or variants of said cellulase enzyme.

[00250] In some embodiments, the enzyme mixtures are subjected to a purification process to selectively remove one or more glucose and/or cellobiose oxidizing enzymes from the enzyme mixture. In some embodiments, the purification process comprises selective precipitation to separate the glucose and/or cellobiose oxidizing enzymes from other enzymes present in the enzyme mixture. In some embodiments, the enzyme mixtures comprise at least one inhibitor of one or more glucose and/or cellobiose oxidizing enzymes.

[00251] The present invention also provides compositions comprising the fungal cell of any of the above embodiments, and/or comprising the enzyme mixture derived from the fungal cell of any of the above embodiments.

[00252] In some embodiments, the present invention provides methods for the production of fungal cells. In some further embodiments, the present invention provides methods for the production of at least one enzyme from fungal cells. In some embodiments, these methods comprise fermentation methods, including but not limited to, batch process, continuous process, fed-batch and/or a combination of methods. In some embodiments, the methods are conducted in a reaction volume of at least about 0.01 mL, about 0.1 mL, about 1 mL, about 10 mL, about 100 mL, about 1000 mL, or at least about 10 L, about 50 L, about 100 L, about 200 L, about 300 L, about 400 L, about 500 L, about 600 L, about 700 L, about 800 L, about 900 L, about 1000 L, about 10,000 L, about 50,000 L, about 100,000 L, about 250,000 L, about 500,000 L, or greater than about 1,000,000 L.

DETAILED DESCRIPTION OF THE INVENTION

[00253] The present invention provides genetically modified cellulase-producing fungal cells that have reduced secreted activity of an endogenous glucose and/or cellobiose oxidizing enzyme, and which are therefore able to secrete enzyme mixtures that improve the yield of fermentable sugars from cellulose. Previous reports have indicated that the oxidation of cellobiose by cellobiose dehydrogenase enhances the rate of cellulose hydrolysis by

cellulases. In contrast to the traditional thinking in the art, the present invention provides fungal cells with genomic deletion(s) or other genetic modification(s) to reduce glucose and/or cellobiose oxidizing enzyme activity that results in improved yield of fermentable sugars from cellulose. Advantageously, the genetically modified cellulase-producing fungal cells provided herein secrete enzyme mixtures that result in vastly improved yields of fermentable sugars such as glucose from cellulose.

[00254] Among the cellulase-producing filamentous fungi, there are those that also produce a variety of enzymes involved in lignin degradation. For example, organisms of such genera as *Myceliophthora*, *Chrysosporium*, *Sporotrichum*, *Thielavia*, *Phanerochaete* and *Trametes* produce and secrete a mixture of cellulases, hemicellulases and lignin degrading enzymes. These types of organisms are commonly called “white rot fungi” by virtue of their ability to digest lignin and to distinguish them from the “brown rot” fungi (such as *Trichoderma*) which typically cannot digest lignin. The genera *Myceliophthora*, *Chrysosporium*, *Sporotrichum*, and *Thielavia* are closely related and in some cases different genus/species identifiers have been used interchangeably for strains of the same species (e.g., *M. thermophila* and *S. thermophile*). Continuing developments in the methods to establish the taxonomy of filamentous fungi has led to reclassification of some strains from one genus to another or has identified an “anamorph-teleomorph” relationship between strains of two genera (e.g., *M. thermophila* and *T. heterothallica*).

[00255] The present invention provides cells, enzyme mixtures and methods in which the activity of glucose and/or cellobiose oxidizing enzyme(s) is reduced so as to improve the yield of fermentable sugars from an enzymatic cellulose hydrolysis process. In view of this, as described herein, the present invention provides for removal or inactivation of glucose and/or cellobiose oxidizing enzyme from a mixture of cellulase enzymes to improve the yield of fermentable sugars from cellulose or biomass.

Genetically Modified Fungal Cells

[00256] The genetically modified fungal cells provided herein permit a reduction in the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell.

[00257] In some embodiments of the genetically modified fungal cells provided herein, glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell is reduced by at least about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or more, relative to the level of glucose and/or cellobiose oxidizing enzyme activity secreted by the unmodified parental fungal cell grown or cultured under essentially the same culture conditions.

[00258] In some embodiments, the genetic modification results in at least about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or about a 99% reduction in the total glucose and/or cellobiose oxidizing enzyme activity secreted by the fungal cell.

[00259] It will be readily appreciated that any genetic modification known in the art can be employed to reduce the secreted activity of the endogenous glucose and/or cellobiose oxidizing enzyme. For example, as described below, modifications contemplated herein include modifications that reduce the amount of glucose and/or cellobiose oxidizing enzyme secreted by the cell. Further contemplated are modifications that reduce the amount of glucose and/or cellobiose oxidizing enzyme that is expressed by the cell. Additional embodiments include modifications that reduce the transcription level of glucose and/or cellobiose oxidizing enzyme. Still further embodiments include the complete or partial deletion of a gene encoding glucose and/or cellobiose oxidizing enzyme. Other embodiments include modifications that reduce the catalytic efficiency of glucose and/or cellobiose oxidizing enzyme.

[00260] **Secreted Enzyme(s).** Accordingly, in some embodiments, the fungal cell has been genetically modified to reduce the amount of the endogenous glucose and/or cellobiose oxidizing enzyme that is secreted by the cell. The glucose and/or cellobiose oxidizing enzyme that is secreted by a cell is a glucose and/or cellobiose oxidizing enzyme produced by the cell in a manner such that the glucose and/or cellobiose oxidizing enzyme is exported across a cell membrane and then subsequently released into the extracellular milieu, such as

into culture media. Thus, a reduction in the amount of secreted glucose and/or cellobiose oxidizing enzyme can be a complete or partial reduction of the glucose and/or cellobiose oxidizing enzyme secreted to the extracellular milieu. Reduction in the amount of secreted glucose and/or cellobiose oxidizing enzyme can be accomplished by reducing the amount of glucose and/or cellobiose oxidizing enzyme produced by the cell and/or by reducing the ability of the cell to secrete the glucose and/or cellobiose oxidizing enzyme that is produced by the cell. Methods for reducing the ability of the cell to secrete a polypeptide can be performed according to any of a variety of methods known in the art (*See e.g.*, Fass and Engels, J. Biol. Chem., 271, 15244-15252 [1996]). For example, the gene encoding a secreted polypeptide can be modified to delete or inactivate a secretion signal peptide. Thus, in some embodiments, the fungal cell has been genetically modified to disrupt the N-terminal secretion signal peptide of the glucose and/or cellobiose oxidizing enzyme. The amount of glucose and/or cellobiose oxidizing enzyme that is secreted by the cell can be reduced by at least about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or more, relative to the secretion of glucose and/or cellobiose oxidizing enzyme in an unmodified organism grown or cultured under essentially the same culture conditions.

[00261] Furthermore, the total amount of glucose and/or cellobiose oxidizing enzyme activity can be reduced by at least about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99% or more, relative to the total amount of glucose and/or cellobiose oxidizing enzyme secreted in an unmodified organism grown or cultured under essentially the same culture conditions.

[00262] Decreased secretion of a glucose and/or cellobiose oxidizing enzyme can be determined by any of a variety of methods known in the art for detection of protein or enzyme levels. For example, the levels of glucose and/or cellobiose oxidizing enzyme in the supernatant of a fungal culture can be detected using Western blotting techniques or other protein detection techniques that use an antibody specific to the glucose and/or cellobiose

oxidizing enzyme. Similarly, secreted glucose and/or cellobiose oxidizing enzyme activity in the supernatant of a fungal culture can be measured using assays for glucose and/or cellobiose oxidizing enzyme activity as described in greater detail herein.

[00263] Expression Level. In some embodiments, the fungal cell has been genetically modified to reduce the amount of the endogenous glucose and/or cellobiose oxidizing enzyme that is expressed by the cell. In some embodiments, the reduction in the expression is accomplished by reducing the amount of mRNA that is transcribed from a gene encoding the glucose and/or cellobiose oxidizing enzyme. In some other embodiments, the reduction in the expression is accomplished by reducing the amount of protein that is translated from a mRNA encoding the glucose and/or cellobiose oxidizing enzyme.

[00264] The amount of glucose and/or cellobiose oxidizing enzyme that is expressed by the cell can be reduced by at least about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or more, relative to the expression of glucose and/or cellobiose oxidizing enzyme in an unmodified fungal cell. In some embodiments, the reduction in the expression is accomplished by reducing the amount of mRNA that is transcribed from a gene encoding cellobiose dehydrogenase or glucose oxidase in an unmodified organism grown or cultured under essentially the same culture conditions.

[00265] Furthermore, in some embodiments a reduction in the expression level of a glucose and/or cellobiose oxidizing enzyme will result in at least about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or about a 99% reduction in the total expression level of glucose and/or cellobiose oxidizing enzyme activity by the fungal cell relative to an unmodified fungal cell grown or cultured under essentially the same culture conditions.

[00266] Decreased expression of a glucose and/or cellobiose oxidizing enzyme can be determined by any of a variety of methods known in the art for detection of protein or enzyme levels. For example, the levels of glucose and/or cellobiose oxidizing enzyme in the

supernatant of a fungal culture can be detected using Western blotting techniques or other protein detection techniques that use an antibody specific to the glucose and/or cellobiose oxidizing enzyme.

[00267] Methods for reducing expression of a polypeptide are well known and can be performed using any of a variety of methods known in the art. For example, the gene encoding a secreted polypeptide can be modified to disrupt a translation initiation sequence such as a Shine-Delgarno sequence or a Kozak consensus sequence. Furthermore, the gene encoding a secreted polypeptide can be modified to introduce a frameshift mutation in the transcript encoding the endogenous glucose and/or cellobiose oxidizing enzyme. It will also be recognized that usage of uncommon codons can result in reduced expression of a polypeptide. It will be appreciated that in some embodiments, the gene encoding the glucose and/or cellobiose oxidizing enzyme can have a nonsense mutation that results in the translation of a truncated protein.

[00268] Other methods of reducing the amount of expressed polypeptide include post-transcriptional RNA silencing methodologies such as antisense RNA and RNA interference. Antisense techniques are well-established, and include using a nucleotide sequence complementary to the nucleic acid sequence of the gene. More specifically, in some embodiments, expression of the gene by a fungal cell is reduced or eliminated by introducing a nucleotide sequence complementary to the nucleic acid sequence, which is transcribed in the cell and is capable of hybridizing to the mRNA produced in the cell. Under conditions allowing the complementary anti-sense nucleotide sequence to hybridize to the mRNA, the amount of protein translated is thus reduced or eliminated (*See e.g.*, Ngiam *et al.*, Appl Environ. Microbiol., 66:775-82 [2000]; and Zrenner *et al.*, Planta 190:247-52 [1993]).

[00269] In some further embodiments, modification, downregulation and/or inactivation of the gene is achieved via any suitable RNA interference (RNAi) technique (*See e.g.*, Kadotani *et al.*, Mol. Plant Microbe Interact., 16:769-76 [2003]). RNA interference methodologies include double stranded RNA (dsRNA), short hairpin RNAs (shRNAs) and small interfering RNAs (siRNAs). Potent silencing using dsRNA may be obtained (*See e.g.*, Fire *et al.*, Nature 391:806-11 [1998]). Silencing using shRNAs is also well-established (*See e.g.*, Paddison *et al.*, Genes Dev. 16:948-958 [2002]). Silencing using siRNA techniques are also known (*See e.g.*, Miyagishi *et al.*, Nat. Biotechnol., 20:497-500 [2002]).

[00270] Transcription Level. In some embodiments, the fungal cell has been genetically modified to reduce the transcription level of a gene encoding the endogenous glucose and/or cellobiose oxidizing enzyme. The transcription level can be reduced by at least about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or more, relative to the transcription level of a glucose and/or cellobiose oxidizing enzyme in an unmodified organism grown or cultured under essentially the same culture conditions.

[00271] Furthermore, a reduction in the transcription level of a glucose and/or cellobiose oxidizing enzyme will result in at least about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or about a 99% reduction in the total glucose and/or cellobiose oxidizing enzyme activity secreted by the fungal cell relative to an unmodified organism grown or cultured under essentially the same culture conditions.

[00272] Decreased transcription can be determined by any of a variety of methods known in the art for detection of transcription levels. For example, the levels of transcription of a particular mRNA in a fungal cell can be detected using quantitative RT-PCR techniques or other RNA detection techniques that specifically detect a particular mRNA.

[00273] Methods for reducing transcription level of a gene can be performed according to any method known in the art, and include partial or complete deletion of the gene, and disruption or replacement of the promoter of the gene such that transcription of the gene is greatly reduced or even inhibited. For example, the promoter of the gene can be replaced with a weak promoter (*See e.g.*, U.S. Patent No. 6,933,133). Thus, where the weak promoter is operably linked with the coding sequence of an endogenous polypeptide, transcription of that gene will be greatly reduced or even inhibited.

[00274] Gene Deletion. In some embodiments, the fungal cell has been genetically modified to at least partially delete a gene encoding the endogenous glucose and/or cellobiose oxidizing enzyme. In some embodiments, this deletion reduces or eliminates the

total amount of endogenous glucose and/or cellobiose oxidizing enzyme activity secreted by the fungal cell.

[00275] A deletion in a gene encoding a glucose and/or cellobiose oxidizing enzyme in accordance with the embodiments provided herein can be a deletion of one or more nucleotides in the gene encoding the glucose and/or cellobiose oxidizing enzyme, and is often a deletion of at least about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% of the gene encoding the glucose and/or cellobiose oxidizing enzyme, where the amount of glucose and/or cellobiose oxidizing enzyme activity secreted by the cell is reduced.

[00276] Thus, for example, in some embodiments, the deletion results in at least about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or about a 99% reduction in the activity of the endogenous glucose and/or cellobiose oxidizing enzyme secreted by the fungal cell, relative to the activity of the glucose and/or cellobiose oxidizing enzyme secreted by an unmodified organism grown or cultured under essentially the same culture conditions.

[00277] Furthermore, in some embodiments, the deletion results in at least about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or about a 99% reduction in the total glucose and/or cellobiose oxidizing enzyme activity secreted by the fungal cell relative to an unmodified fungal cell grown or cultured under essentially the same culture conditions.

[00278] Deletion of a glucose and/or cellobiose oxidizing enzyme gene can be detected and confirmed by any of a variety of methods known in the art for detection of gene deletions. For example, as exemplified in the Example section below, gene deletion can be confirmed using PCR amplification of the modified genomic region. It will be appreciated that additional techniques for confirming deletion can be used and are well known, including

Southern blot techniques, DNA sequencing of the modified genomic region, and screening for positive or negative markers incorporated during recombination events.

[00279] Methods for complete and/or partial deletion of a gene are well-known and the genetically modified fungal cell described herein can be generated using any of a variety of deletion methods known in the art that permits a reduction in the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell. Such methods may advantageously include standard gene disruption using homologous flanking markers (*See e.g.*, Rothstein, *Meth. Enzymol.*, 101:202-211 [1983]). Another technique for gene deletion includes PCR-based methods for standard deletion (*See e.g.*, Davidson *et al.*, *Microbiol.*, 148:2607-2615 [2002], which describes a PCR-based strategy to generate integrative targeting alleles with large regions of homology).

[00280] Further gene deletion techniques include “positive-negative” cassettes; cre/lox based deletion, biolistic transformation to increase homologous recombination, and agrobacterium-mediated gene disruption. The “positive-negative” method employs cassettes which consist of one marker gene for positive screening and another marker gene for negative screening (*See e.g.*, Chang *et al.*, *Proc. Natl. Acad. Sci. USA* 84:4959-4963 [1987]). Cre/lox based methodologies employ elimination of marker genes using expression of Cre recombinase (*See e.g.*, Florea *et al.*, *Fung. Genet. Biol.*, 46:721-730 [2009]).

[00281] Methods to introduce DNA or RNA into fungal cells are known to those of skill in the art and include PEG-mediated transformation of protoplasts, electroporation, biolistic transformation, and *Agrobacterium*-mediated transformation. Biolistic transformation employs a unique process in which DNA or RNA is introduced into cells on micron-sized particles, thus increasing delivery of a deletion construct to the fungal cell (*See e.g.*, Davidson *et al.*, *Fung. Genet. Biol.*, 29:38-48 [2000]). Similarly, *Agrobacterium*-mediated transformation in conjunction with linear or split-marker deletion cassettes can facilitate delivery of deletion constructs to the target cell (*See e.g.*, Wang *et al.*, *Curr. Genet.*, 56:297-307 [2010]).

[00282] Additional methods for complete or partial gene deletion include, but are not limited to, disruption of the gene. Such gene disruption techniques are known to those of skill in the art, and include the use of, for example, insertional mutagenesis, the use of transposons and marked integration. However, it will be appreciated that any technique that

provides for disruption of the coding sequence or any other functional aspect of a gene can be utilized to generate the genetically modified fungal cells provided herein. Methods of insertional mutagenesis can be performed according to any such method known in the art (See e.g., Combier *et al.*, FEMS Microbiol. Lett., 220:141–8 [2003]). For example, *Agrobacterium*-mediated insertional mutagenesis can be used to insert a sequence that disrupts the function of the encoded gene, such as disruption of the coding sequence or any other functional aspect of the gene.

[00283] Transposon mutagenesis methodologies are another manner for disruption of a gene. Transposon mutagenesis is well known in the art, and can be performed using *in vivo* techniques (See e.g., Firon *et al.*, Eukaryot. Cell 2:247–55 [2003]); or by the use of *in vitro* techniques (See e.g., Adachi *et al.*, Curr Genet., 42:123–7 [2002]). Thus, targeted gene disruption using transposon mutagenesis can be used to insert a sequence that disrupts the function of the encoded gene, such as disruption of the coding sequence or any other functional aspect of the gene.

[00284] Restriction enzyme-mediated integration (REMI) is another methodology for gene disruption, and is well known in the art (See e.g., Thon *et al.*, Mol. Plant Microbe Interact., 13:1356–65 [2000]).

REMI generates insertions into genomic restriction sites in an apparently random manner, some of which cause mutations. Thus, insertional mutants that demonstrate a disruption in the gene encoding the endogenous glucose and/or cellobiose oxidizing enzyme can be selected and utilized as provided herein.

[00285] Catalytic Disruption. In some other embodiments, the fungal cell is genetically modified to reduce the catalytic efficiency of the endogenous glucose and/or cellobiose oxidizing enzyme. In some embodiments, a genetic modification that reduces catalytic efficiency can result in, for example, a translated protein product that has a reduction in enzymatic activity.

[00286] In some embodiments, a reduction in catalytic efficiency is a reduction of glucose and/or cellobiose oxidizing enzyme activity of about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%,

about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or more, relative to unmodified glucose and/or cellobiose oxidizing enzyme, as measured using standard techniques.

[00287] In some further embodiments, the genetic modification results in a reduction of glucose and/or cellobiose oxidizing enzyme activity of at least about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or about a 99% reduction in the total glucose and/or cellobiose oxidizing enzyme activity secreted by the fungal cell.

[00288] Methods for reducing catalytic efficiency of dehydrogenases and oxidases are well known, and as such, any of a variety of suitable methods known in the art for reducing catalytic efficiency can be utilized in the genetic modification of the fungal cells provided herein. Thus, for example, the fungal cell can be genetically modified to inactivate one or more residues in an active site of the glucose and/or cellobiose oxidizing enzyme (*See e.g.*, Frederik *et al.*, *Biochem.*, 42:4049–4056 [2003]). For example, one or more residues can be modified to decrease substrate binding, and/or one or more residues can be modified to decrease the catalytic activity of the glucose and/or cellobiose oxidizing enzyme. Accordingly, one or more residues in the electron acceptor (*e.g.*, flavin) binding domain, saccharide binding domain or other substrate binding domain of glucose and/or cellobiose oxidizing enzyme can be performed to reduce or inactivate the catalytic efficiency of the glucose and/or cellobiose oxidizing enzyme. Similarly, it will be apparent that mutation of residues outside an active site can result in allosteric change in the shape or activity of the glucose and/or cellobiose oxidizing enzyme.

[00289] In some additional embodiments, other domains are targeted for a mutation which results in reducing catalytic efficiency of the endogenous glucose and/or cellobiose oxidizing enzyme. For example, in some embodiments, a mutation to one or more residues in a heme binding domain of a glucose and/or cellobiose oxidizing enzyme can result in reduced catalytic efficiency (*See e.g.*, Rotsaert *et al.*, *Arch. Biochem. Biophys.*, 390:206-14 [2001]).

[00290] Similarly, in some embodiments, the genetic modification is a conditional mutation to a glucose and/or cellobiose oxidizing enzyme. In some embodiments, the glucose and/or cellobiose oxidizing enzyme has a temperature sensitive mutation that renders the protein non-functional (*i.e.*, inactive or less active) at (*e.g.* warm temperatures, such as 37–42°C.), and functional (*i.e.*, active) at colder temperatures.

Fungal Cells

[00291] In some embodiments, the present invention provides a fungal cell that is a Basidiomycete belonging to the class Agaricomycetes or an Ascomycete belonging to the subdivision Pezizomycotina that has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell, where the fungal cell is capable of secreting a cellulase-containing enzyme mixture. In some embodiments, the genetically modified fungal cell provided herein is a Basidiomycete belonging to the class Agaricomycetes or an Ascomycete belonging to the subdivision Pezizomycotina. In some embodiments, the Basidiomycete is a species of *Pleurotus*, *Peniophora*, *Trametes*, *Athelia*, *Sclerotium*, *Termitomyces*, *Flammulina*, *Coniophora*, *Ganoderma*, *Pycnoporus*, *Ceriporiopsis*, *Phanerochaete*, *Gloeophyllum*, *Hericium*, *Heterobasidion*, *Gelatoporia*, *Lepiota*, or *Irpex*. In some embodiments, the Ascomycete is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Neurospora*, *Sordaria*, *Podospora*, *Magnaporthe*, *Fusarium*, *Gibberella*, *Botryotinia*, *Humicola*, *Neosartorya*, *Pyrenophora*, *Phaeosphaeria*, *Sclerotinia*, *Chaetomium*, *Nectria*, *Verticillium*, or *Aspergillus*.

[00292] The classification of a given fungal cell as belonging to the Basidiomycete class Agaricomycetes or to the Ascomycetes subdivision Pezizomycotina is done as is recognized in the art, as exemplified in the NCBI taxonomy database.

[00293] In some embodiments, the fungal cell is a Chaetomiaceae family member. The Chaetomiaceae are a family of fungi in the Ascomycota, class Sordariomycetes. The family Chaetomiaceae includes the genera *Achaetomium*, *Aporothielavia*, *Chaetomidium*, *Chaetomium*, *Corylomyces*, *Corynascus*, *Farrowia*, *Thielavia*, *Zopfiella*, and *Myceliophthora*.

[00294] In some embodiments, the genetically modified fungal cell is an anamorph or teleomorph of a Basidiomycete belonging to the class Agaricomycetes or an Ascomycete

belonging to the subdivision Pezizomycotina. In some embodiments, the genetically modified fungal cell is an anamorph or teleomorph of a Chaetomiaceae family member selected from the genera *Myceliophthora*, *Thielavia*, *Corynascus*, *Chaetomium*. As such, the genetically modified fungal cell can also be selected from the genera *Sporotrichum*, *Chrysosporium*, *Paecilomyces*, *Talaromyces* or *Acremonium*. It is also contemplated that the genetically modified fungal cell be selected from the genera *Ctenomyces*, *Thermoascus*, and *Scytalidium*, including anamorphs and teleomorphs of fungal cells from those genera.

[00295] In some further embodiments, the genetically modified fungal cell is a thermophilic member of the genera *Acremonium*, *Arthroderma*, *Corynascus*, *Thielavia*, *Myceliophthora*, *Thermoascus*, *Chromocleista*, *Byssoschlamys*, *Sporotrichum*, *Chaetomium*, *Chrysosporium*, *Scytalidium*, *Ctenomyces*, *Paecilomyces*, and *Talaromyces*. By “thermophilic fungus” is meant any fungus which exhibits optimum growth at a temperature of at least about 37°C, and generally below about 80°C, such as for example between about 37-80°C, also between about 37-75°C, also between about 40-65°C, and also between about 40-60°C. In some embodiments, the optimum growth is exhibited at a temperature of at least 40°-60°C.

[00296] In some embodiments, the genetically modified fungal cell is selected from the strains of *Sporotrichum cellulophilum*, *Thielavia heterothallica*, *Corynascus heterothallicus*, *Thielavia terrestris*, and *Myceliophthora thermophila*, including anamorphs and teleomorphs thereof. It will be understood that for the aforementioned species, the genetically modified fungal cell presented herein encompasses both the perfect and imperfect states, and other taxonomic equivalents (*e.g.*, anamorphs), regardless of the species name by which they are known. For example, the following species are anamorphs or teleomorphs and may therefore be considered as synonymous: *Myceliophthora thermophila*, *Sporotrichum thermophile*, *Sporotrichum thermophilum*, *Sporotrichum cellulophilum*, *Chrysosporium thermophile*, *Corynascus heterothallicus*, and *Thielavia heterothallica*. Additionally, the following species may be considered synonymous with each other: *Thielavia terrestris*, *Allscheria terrestris*, and *Acremonium alabamense*. Further examples of taxonomic equivalents are known in the art (*See e.g.*, Cannon, Mycopathol., 111:75-83 [1990]; Moustafa *et al.*, Persoonia 14:173-175[1990]; Stalpers, Stud. Mycol., 24, [1984]; Upadhyay *et al.*, Mycopathol., 87:71-80 [1984]; Guarro *et al.*, Mycotaxon 23: 419-427 [1985]; Awao *et al.*,

Mycotaxon 16:436-440 [1983]; von Klopotek, Arch. Microbiol., 98:365-369 [1974]; and Long *et al.*, ATCC Names of Industrial Fungi, ATCC, Rockville MD [1994]). Those skilled in the art will readily recognize the identity of appropriate equivalents. Accordingly, it will be understood that, unless otherwise stated, the use of a particular species designation in the present disclosure also refers to species that are related by anamorphic or teleomorphic relationship.

[00297] In some embodiments, the genetically modified fungal cell is a cellulase-producing fungal cell that is a Basidiomycete belonging to the class Agaricomycetes or an Ascomycete belonging to the subdivision Pezizomycotina. For example, in some embodiments, the genetically modified fungal cell is a Basidiomycete belonging to the class Agaricomycetes or an Ascomycete belonging to the subdivision Pezizomycotina that secretes two or more cellulose hydrolyzing enzymes, such as, for example, endoglucanase, cellobiohydrolase, or beta-glucosidase. It will be appreciated that cellulase can include hemicellulose-hydrolyzing enzymes such as endoxylanase, beta-xylosidase, arabinofuranosidase, alpha-glucuronidase, acetylxylenesterase, feruloyl esterase, and alpha-glucuronylesterase. It will also be appreciated that a cellulase-producing fungal cell can produce two or more of these enzymes, in any combination.

[00298] Additionally, in some embodiments, the genetically modified fungal cell is derived from a lignocellulose-competent parental fungal cell. The present invention also provides a fungal culture in a vessel comprising a genetically modified fungal cell as described hereinabove. In some embodiments, the vessel comprises a liquid medium, such as fermentation medium. For example, the vessel can be a flask, bioprocess reactor, and the like. In some embodiments, the vessel comprises a solid growth medium. For example, the solid medium can be an agar medium such as potato dextrose agar, carboxymethylcellulose, cornmeal agar, and the like. In some embodiments, the fungal cell described herein is an isolated fungal cell.

Sugar Oxidizing Enzymes

[00299] The present invention provides fungal cells that have been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme

secreted by the fungal cell. Examples of some suitable glucose and/or cellobiose oxidizing enzymes that find use in the present invention are described in greater detail below.

[00300] Glucose Oxidase. As indicated herein, in some embodiments, glucose oxidase and fungal cells producing reduced levels of glucose oxidase activity find use in the present invention. Glucose oxidase is known to function via a so-called ping-pong mechanism of enzymatic catalysis, which involves successive binding on two different sites. One site is a saccharide binding domain that is capable of binding β -D-glucose. The other site is a relatively non-selective co-substrate site for binding an oxidant such as FAD.

[00301] One of skill in the art will appreciate that glucose oxidase enzyme activity typically employs the presence of oxygen or an equivalent redox acceptor (*e.g.*, lignin, molecular oxygen, cytochrome c, redox dyes, benzoquinones and Fe^{2+} complexes). Glucose oxidase (GO) activity can be measured using any of a variety of methods known in the art. For example, GO activity assays can be performed using any suitable method known in the art (*See e.g.*, Bergmeyer *et al.*, in Methods of Enzymatic Analysis (Bergmeyer, ed.) Volume I, 2nd Ed., pp. 457-458, Academic Press Inc., New York, NY [1974]; and U.S. Patent No. 3,953,295). For example, GO activity is determined by an increase in absorbance at 460 nm resulting from the oxidation of o-dianisidine through a peroxidase coupled system.

[00302] In some embodiments, the present invention provides fungal cells that have been genetically modified to reduce the secreted activity of a glucose oxidase and have reduced secreted activity of an endogenous glucose oxidase. Accordingly, one or more glucose oxidase enzymes from each of the fungal species described herein can be targeted for genetic modification.

[00303] In some embodiments, the glucose oxidase is from a fungal species from the division Basidiomycete and belonging to the class Agaricomycetes; or from the division Ascomycete and belonging to the subdivision Pezizomycotina. Some examples of glucose oxidase enzymes identified from division Basidiomycete belonging to the class Agaricomycetes; and division Ascomycete belonging to the subdivision Pezizomycotina are set forth in the Table B, below.

[00304] In some embodiments, the glucose oxidase is from a fungal species selected from *Chaetomium globosum*, *Thielavia heterothallica*, *Thielavia terrestris*, *Talaromyces stipitatus* and *Myceliophthora thermophila*. Some glucose oxidase enzymes identified from

these and other species are set forth in Table B, below. The proteins listed in Table B are examples of glucose oxidase that are known in the art, or identified herein as being a glucose oxidase.

Table B. Glucose Oxidase Sequences			
Accession Number	Organism	GMC oxred N Domain	GMC oxred C Domain
chr1-56652m21GM (SEQ ID NO.: 2)	<i>Myceliophthora thermophila</i>	36-356	495-634
XP_001227361.1	<i>Chaetomium globosum</i> CBS 148.51	36-355	465-604
JGITHite5217	<i>Thielavia terrestris</i>	36-355	466-605
XP_001223540.1	<i>Chaetomium globosum</i> CBS 148.51	185-380	514-655
XP_001910674.1	<i>Podospira anserina</i> S mat+	39-373	506-644
XP_001220376.1	<i>Chaetomium globosum</i> CBS 148.51	38-342	481-620
XP_001226009.1	<i>Chaetomium globosum</i> CBS 148.51	214-289	323-464
CBI59558.1	<i>Sordaria macrospora</i>	43-373	480-619
XP_383916.1	<i>Gibberella zeae</i> PH-1	35-345	466-603
CBI59559.1	<i>Sordaria macrospora</i>	43-356	440-579
JGITHite6377	<i>Thielavia terrestris</i>	40-363	490-630
XP_001549389.1	<i>Botryotinia fuckeliana</i> B05.10	39-347	438-578
XP_001903685.1	<i>Podospira anserina</i> S mat+	36-337	465-606
XP_002792207.1	<i>Paracoccidioides brasiliensis</i> Pb01	68-290	414-556
XP_003001656.1	<i>Verticillium albo-atrum</i> VaMs.102	46-359	488-627
XP_361250.1	<i>Magnaporthe oryzae</i> 70-15	113-324	450-594
JGITHite5048	<i>Thielavia terrestris</i>	45-354	480-619
XP_001226113.1	<i>Chaetomium globosum</i> CBS 148.51	44-353	480-619
XP_001906345.1	<i>Podospira anserina</i> S mat+	39-352	480-620
chr4-293m24GM (SEQ ID NO.: 4)	<i>Myceliophthora thermophila</i>	81-380	508-647
CAJ85791.1	<i>Fusarium oxysporum</i> f. sp. <i>Lycopersici</i>	34-344	465-602
XP_661610.1	<i>Aspergillus nidulans</i> FGSC A4	35-346	467-604
XP_003041895.1	<i>Nectria haematococca</i> mpVI 77-13-4	35-346	467-604
XP_001912227.1	<i>Podospira anserina</i> S mat+	27-333	443-584
XP_681267.1	<i>Aspergillus nidulans</i> FGSC A4	38-343	471-609
XP_001227424.1	<i>Chaetomium globosum</i> CBS 148.51	42-374	500-640
CBI58590.1	<i>Sordaria macrospora</i>	46-351	477-588
EEH49925.1	<i>Paracoccidioides brasiliensis</i> Pb18	176-397	NA
XP_001223186.1	<i>Chaetomium globosum</i> CBS 148.51	37-292	424-599
XP_001791201.1	<i>Phaeosphaeria nodorum</i> SN15	60-223	349-480
XP_001905375.1	<i>Podospira anserina</i> S mat+	49-370	495-633
XP_001592050.1	<i>Sclerotinia sclerotiorum</i> 1980	227-296	314-446
XP_003002940.1	<i>Verticillium albo-atrum</i> VaMs.102	43-355	462-600
XP_366260.2	<i>Magnaporthe oryzae</i> 70-15	42-351	491-631
XP_003048882.1	<i>Nectria haematococca</i> mpVI 77-13-4	121-295	394-530

Table B. Glucose Oxidase Sequences

Accession Number	Organism	GMC oxred N Domain	GMC oxred C Domain
XP_001796868.1	<i>Phaeosphaeria nodorum</i> SN15	38-350	475-615
XP_001805358.1	<i>Phaeosphaeria nodorum</i> SN15	20-328	454-593
XP_001931252.1	<i>Pyrenophora tritici-repentis</i> Pt-1C-BFP	23-318	508-595
XP_001218113.1	<i>Aspergillus terreus</i> NIH2624	43-354	475-612
XP_001224467.1	<i>Chaetomium globosum</i> CBS 148.51	218-318	446-552
XP_003049247.1	<i>Nectria haematococca</i> mpVI 77-13-4	25-323	451-589
XP_001906627.1	<i>Podospora anserina</i> S mat ⁺	37-349	473-618
XP_001804484.1	<i>Phaeosphaeria nodorum</i> SN15	29-319	449-586
JGITHite9772	<i>Thielavia terrestris</i>	33-357	460-599
EEH08138.1	<i>Ajellomyces capsulatus</i> G186AR	181-256	366-509
XP_002565293.1	<i>Penicillium chrysogenum</i> Wisconsin 54-1255	34-355	458-595
CBI52485.1	<i>Sordaria macrospora</i>	28-338	NA
XP_001273036.1	<i>Aspergillus clavatus</i> NRRL 1	14-322	421-560
AAF31169.1 AF143814_1	<i>Pleurotus pulmonarius</i>	30-342	446-585
EER39780.1	<i>Ajellomyces capsulatus</i> H143	228-301	411-554
XP_001558188.1	<i>Botryotinia fuckeliana</i> B05.10	37-358	482-619
XP_001904543.1	<i>Podospora anserina</i> S mat ⁺	27-344	453-596
XP_001544530.1	<i>Ajellomyces capsulatus</i> NAM1	228-301	411-554
JGITHite8281	<i>Thielavia terrestris</i>	5-301	456-624
XP_383802.1	<i>Gibberella zeae</i> PH-1	24-324	452-589
XP_001833868.1	<i>Coprinopsis cinerea</i> okayama7#130	35-341	430-568
XP_001836103.1	<i>Coprinopsis cinerea</i> okayama7#130	39-366	462-535
XP_001597615.1	<i>Sclerotinia sclerotiorum</i> 1980	134-290	295-422
ADD14021.1	<i>Pleurotus eryngii</i>	30-342	446-585
EEH50230.1	<i>Paracoccidioides brasiliensis</i> Pb18	131-252	366-484
JGITHite6435	<i>Thielavia terrestris</i>	100-404	509-658
XP_362999.2	<i>Magnaporthe oryzae</i> 70-15	31-327	431-575
XP_761104.1	<i>Ustilago maydis</i> 521	54-372	485-603
XP_002482522.1	<i>Talaromyces stipitatus</i> ATCC 10500	36-362	456-593
XP_001879270.1	<i>Laccaria bicolor</i> S238N-H82	35-345	451-581
XP_001584680.1	<i>Sclerotinia sclerotiorum</i> 1980	24-335	440-581
XP_001798598.1	<i>Phaeosphaeria nodorum</i> SN15	35-337	440-584
XP_661816.1	<i>Aspergillus nidulans</i> FGSC A4	33-350	461-599
XP_001883085.1	<i>Laccaria bicolor</i> S238N-H82	34-348	450-594
XP_001556658.1	<i>Botryotinia fuckeliana</i> B05.10	39-364	464-603
XP_001833865.1	<i>Coprinopsis cinerea</i> okayama7#130	34-345	432-570
XP_002390024.1	<i>Moniliophthora perniciosa</i> FA553	36-298	411-536
XP_001801353.1	<i>Phaeosphaeria nodorum</i> SN15	22-332	436-579
XP_001817515.1	<i>Aspergillus oryzae</i> RIB40	28-335	436-577
XP_568317.1	<i>Cryptococcus neoformans</i> var. <i>neoformans</i> JEC21	55-373	507-643
XP_001273087.1	<i>Aspergillus clavatus</i> NRRL 1	51-369	481-620

Table B. Glucose Oxidase Sequences

Accession Number	Organism	GMC oxred N Domain	GMC oxred C Domain
XP_002372599.1	<i>Aspergillus flavus</i> NRRL3357	28-340	441-582
XP_001806098.1	<i>Phaeosphaeria nodorum</i> SN15	62-382	494-634
XP_001835456.1	<i>Coprinopsis cinerea</i> okayama7#130	34-345	433-569
XP_001586361.1	<i>Sclerotinia sclerotiorum</i> 1980	32-346	446-586
XP_001884302.1	<i>Laccaria bicolor</i> S238N-H82	36-348	452-589
XP_001821530.1	<i>Aspergillus oryzae</i> RIB40	75-394	496-635
XP_760191.1	<i>Ustilago maydis</i> 521	59-366	497-628
XP_002375018.1	<i>Aspergillus flavus</i> NRRL3357	53-358	466-631
XP_391162.1	<i>Gibberella zeae</i> PH-1	21-316	442-579
XP_759762.1	<i>Ustilago maydis</i> 521	84-408	530-667
CBI51995.1	<i>Sordaria macrospora</i>	30-326	469-610
XP_002376612.1	<i>Aspergillus flavus</i> NRRL3357	22-321	457-599
XP_660833.1	<i>Aspergillus nidulans</i> FGSC A4	41-358	466-603
EDP53840.1	<i>Aspergillus fumigatus</i> A1163	36-347	448-589
XP_001911514.1	<i>Podospora anserina</i> S mat+	77-396	498-637
XP_002471526.1	<i>Postia placenta</i> Mad-698-R	86-400	517-660
XP_367669.2	<i>Magnaporthe oryzae</i> 70-15	21-323	427-568
XP_001389920.1	<i>Aspergillus niger</i>	5-289	391-528
XP_001732158.1	<i>Malassezia globosa</i> CBS 7966	52-370	481-618
XP_001826806.1	<i>Aspergillus oryzae</i> RIB40	27-338	439-580
XP_001216916.1	<i>Aspergillus terreus</i> NIH2624	26-338	439-580
XP_003000545.1	<i>Verticillium albo-atrum</i> VaMs.102	21-339	440-581
XP_391184.1	<i>Gibberella zeae</i> PH-1	264-389	491-631
XP_002148263.1	<i>Penicillium marneffei</i> ATCC 18224	30-351	469-606
XP_749312.1	<i>Aspergillus fumigatus</i> Af293	36-347	448-589
JGIThte9811	<i>Thielavia terrestris</i>	22-337	440-579
XP_001907031.1	<i>Podospora anserina</i> S mat+	29-322	451-590
XP_001550244.1	<i>Botryotinia fuckeliana</i> B05.10	24-336	451-484
XP_002479433.1	<i>Talaromyces stipitatus</i> ATCC 10500	42-367	467-606
XP_001390806.1	<i>Aspergillus niger</i>	38-356	466-605
XP_681081.1	<i>Aspergillus nidulans</i> FGSC A4	143-367	469-608
XP_001882478.1	<i>Laccaria bicolor</i> S238N-H82	33-345	451-588
XP_664049.1	<i>Aspergillus nidulans</i> FGSC A4	36-353	464-603
XP_002373928.1	<i>Aspergillus flavus</i> NRRL3357	41-353	465-604
XP_001592756.1	<i>Sclerotinia sclerotiorum</i> 1980	42-358	469-539
YP_914851.1	<i>Paracoccus denitrificans</i> PD1222	13-304	394-530
XP_002622246.1	<i>Ajellomyces dermatitidis</i> SLH14081	39-349	459-602
XP_002565328.1	<i>Penicillium chrysogenum</i> Wisconsin 54-1255	34-338	450-589
XP_001215452.1	<i>Aspergillus terreus</i> NIH2624	46-350	454-589
XP_001820476.1	<i>Aspergillus oryzae</i> RIB40	41-343	455-594
XP_001732090.1	<i>Malassezia globosa</i> CBS 7966	55-373	484-621
XP_002388554.1	<i>Moniliophthora perniciosa</i> FA553	46-367	487-624

Table B. Glucose Oxidase Sequences

Accession Number	Organism	GMC oxred N Domain	GMC oxred C Domain
XP_001265740.1	<i>Neosartorya fischeri</i> NRRL 181	34-345	446-587
XP_381957.1	<i>Gibberella zeae</i> PH-1	53-320	432-553
XP_001587168.1	<i>Sclerotinia sclerotiorum</i> 1980	33-351	464-603
XP_660308.1	<i>Aspergillus nidulans</i> FGSC A4	27-334	437-568
YP_001923964.1	<i>Methylobacterium populi</i> BJ001	67-360	428-563
XP_001559357.1	<i>Botryotinia fuckeliana</i> B05.10	50-370	482-621
XP_002389049.1	<i>Moniliophthora perniciosa</i> FA553	33-345	NA
XP_001394544.1	<i>Aspergillus niger</i>	27-338	439-580
XP_760250.1	<i>Ustilago maydis</i> 521	38-293	419-556
XP_359722.1	<i>Magnaporthe oryzae</i> 70-15	39-365	476-616
XP_001800211.1	<i>Phaeosphaeria nodorum</i> SN15	122-375	485-623
XP_002481914.1	<i>Talaromyces stipitatus</i> ATCC 10500	5-303	416-553
XP_001398576.1	<i>Aspergillus niger</i>	40-353	465-604
XP_003040786.1	<i>Nectria haematococca</i> mpVI 77-13-4	61-379	493-631
XP_001904483.1	<i>Podospora anserina</i> S mat ⁺	120-397	510-649
XP_759393.1	<i>Ustilago maydis</i> 521	40-366	478-618
XP_002388140.1	<i>Moniliophthora perniciosa</i> FA553	25-346	NA
XP_002373140.1	<i>Aspergillus flavus</i> NRRL3357	43-357	469-603
XP_002143250.1	<i>Penicillium marneffei</i> ATCC 18224	37-355	466-605
XP_001729093.1	<i>Malassezia globosa</i> CBS 7966	35-351	454-604
XP_001793977.1	<i>Phaeosphaeria nodorum</i> SN15	25-345	451-587
XP_002476910.1	<i>Postia placenta</i> Mad-698-R	5-307	431-563
XP_001559633.1	<i>Botryotinia fuckeliana</i> B05.10	33-352	465-604
XP_001732157.1	<i>Malassezia globosa</i> CBS 7966	49-368	480-617
XP_002149622.1	<i>Penicillium marneffei</i> ATCC 18224	43-324	400-532
XP_001910399.1	<i>Podospora anserina</i> S mat ⁺	51-369	480-620
XP_391404.1	<i>Gibberella zeae</i> PH-1	64-382	496-634
XP_002794971.1	<i>Paracoccidioides brasiliensis</i> Pb01	5-300	414-551
XP_001270826.1	<i>Aspergillus clavatus</i> NRRL 1	48-372	484-623
XP_001548196.1	<i>Botryotinia fuckeliana</i> B05.10	41-349	477-617
XP_001215424.1	<i>Aspergillus terreus</i> NIH2624	41-351	450-591
XP_758019.1	<i>Ustilago maydis</i> 521	36-343	457-608
BAI66412.1	<i>Fusarium oxysporum</i>	66-382	496-634
EDP52931.1	<i>Aspergillus fumigatus</i> A1163	46-370	482-621
XP_002387978.1	<i>Moniliophthora perniciosa</i> FA553	224-327	NA
XP_001216137.1	<i>Aspergillus terreus</i> NIH2624	27-340	441-525
XP_002375706.1	<i>Aspergillus flavus</i> NRRL3357	51-359	471-608
XP_001598641.1	<i>Sclerotinia sclerotiorum</i> 1980	50-370	482-621
gi 71002308 ref XP_755835.1	<i>Aspergillus fumigatus</i> Af293	38-354	466-602
gi 119481873 ref XP_001260965.1	<i>Neosartorya fischeri</i> NRRL 181	38-354	466-605
XP_001817967.1	<i>Aspergillus oryzae</i> RIB40	43-357	469-608

Table B. Glucose Oxidase Sequences			
Accession Number	Organism	GMC oxred N Domain	GMC oxred C Domain
XP_001275639.1	<i>Aspergillus clavatus</i> NRRL 1	39-356	468-607
XP_002148584.1	<i>Penicillium marneffei</i> ATCC 18224	54-377	490-629
XP_002485672.1	<i>Talaromyces stipitatus</i> ATCC 10500	54-377	476-605
EDP55006.1	<i>Aspergillus fumigatus</i> A1163	38-354	466-602
XP_754807.1	<i>Aspergillus fumigatus</i> Af293	46-370	482-621
CBF78527.1	<i>Aspergillus nidulans</i> FGSC A4	21-324	436-575
XP_002556907.1	<i>Penicillium chrysogenum</i> Wisconsin 54-1255	28-339	NA
XP_002567445.1	<i>Penicillium chrysogenum</i> Wisconsin 54-1255	53-377	485-622
XP_001396848.1	<i>Aspergillus niger</i>	60-366	454-593
XP_001397016.1	<i>Aspergillus niger</i>	51-381	502-641
XP_001023507.1	<i>Tetrahymena thermophila</i>	8-309	403-539
XP_001263633.1	<i>Neosartorya fischeri</i> NRRL 181	46-370	482-621
XP_001400283.1	<i>Aspergillus niger</i>	49-372	477-616
XP_001261659.1	<i>Neosartorya fischeri</i> NRRL 181	44-369	477-612
XP_001797048.1	<i>Phaeosphaeria nodorum</i> SN15	39-366	458-598
XP_001211074.1	<i>Aspergillus terreus</i> NIH2624	44-320	430-565
XP_001398522.1	<i>Aspergillus niger</i>	50-373	475-614

*Accession numbers for *Thielavia terrestris* refer to the U.S. Department of Energy (DOE) Joint Genome Institute (JGI) genome sequence

[00305] Some amino acid sequences encoding glucose oxidase are provided herein. For example, the nucleotide sequence encoding *Myceliophthora thermophila* GO1 is set forth herein as SEQ ID NO:1, and the encoded amino acid sequence of *Myceliophthora thermophila* GO1 is set forth as SEQ ID NO:2. Furthermore, the nucleotide sequence encoding *Myceliophthora thermophila* GO2 is set forth herein as SEQ ID NO:3, and the encoded amino acid sequence of *Myceliophthora thermophila* GO2 is set forth as SEQ ID NO:4.

[00306] In some embodiments, the glucose oxidase is glucose oxidase (EC 1.1.3.4). In some embodiments, the glucose oxidase is a glucose oxidase with the amino acid sequence of *Myceliophthora thermophila* GO1 as set forth in SEQ ID NO:2. In some embodiments, the glucose oxidase is a glucose oxidase with the amino acid sequence of *Myceliophthora thermophila* GO2 as set forth in SEQ ID NO:4. In other embodiments, the glucose oxidase

comprises an amino acid sequence provided in the GenBank entry of any one of the accession numbers set forth in Table B. In some embodiments, the glucose oxidase is encoded by a nucleic acid sequence that is at least about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to SEQ ID NOS:1 and/or 3. In some embodiments, the glucose oxidase is encoded by a nucleic acid sequence that is at least about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to a nucleic acid sequence encoding the amino acid sequence set forth as SEQ ID NOS:2 and/or 4, or an amino acid sequence provided in the GenBank entry of any one of the accession numbers set forth in Table B. In some embodiments, the glucose oxidase is encoded by a nucleic acid sequence that can selectively hybridize to SEQ ID NOS:1 and/or 3 under moderately stringent or stringent conditions, as described below. In some embodiments, the glucose oxidase is encoded by a nucleic acid sequence that can selectively hybridize under moderately stringent or stringent conditions to a nucleic acid sequence that encodes SEQ ID NOS:2 and/or 4, or an amino acid sequence provided in the GenBank entry of any one of the accession numbers set forth in Table B.

[00307] In some embodiments, the glucose oxidase comprises an amino acid sequence with at least about 50%, about 51%, about 52%, about 53%, about 54%, about 55%, about 56%, about 57%, about 58%, about 59%, about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%,

about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% similarity to the amino acid sequence set forth as SEQ ID NOS:2 and/or 4, or an amino acid sequence provided in the GenBank entry of any one of the accession numbers set forth in Table B.

[00308] Cellobiose Dehydrogenase. In some embodiments, the CDH contains both the conserved glucose-methanol-choline (GMC) oxido-reductase N and the GMC oxido-reductase C domains. In some other embodiments, a CDH contains the GMC oxido-reductase N domain alone. The GMC oxidoreductases are FAD flavoproteins oxidoreductases (*See e.g.*, Cavener, J. Mol. Biol., 223:811-814 [1992]; and Vrielink and Blow, Biochem., 32:11507-15 [1993]). The GMC oxidoreductases include a variety of proteins; choline dehydrogenase (CHD), methanol oxidase (MOX) and cellobiose dehydrogenase (CDH) which share a number of regions of sequence similarities. One of these regions, located in the N-terminal section, corresponds to the FAD ADP-binding domain, as further defined by the Pfam database under the entry GMC_oxred_N (PF00732). Similarly, the C-terminal conserved domain (GMC oxido-reductase C domain) is defined as set forth in the Pfam database under the entry GMC_oxred_C (PF05199).

[00309] Cellobiose dehydrogenases can be categorized into two families, where a first family contains a catalytic portion and a second family contains a catalytic portion and a cellulose binding motif (CBM). The three-dimensional structure of an example cellobiose dehydrogenase features two globular domains, each containing one of two cofactors: a heme or a flavin. The active site lies at a cleft between the two domains. Oxidation of cellobiose typically occurs via 2-electron transfer from cellobiose to the flavin, generating cellobiono-1,5-lactone and reduced flavin. Active FAD is regenerated by electron transfer to the heme group, leaving a reduced heme. The native state heme is regenerated by reaction with the oxidizing substrate at the second active site.

[00310] The acceptor is preferentially iron ferricyanide, cytochrome C, or an oxidized phenolic compound such as dichloroindophenol (DCIP), an acceptor commonly used for colorimetric assays. Metal ions and O₂ are also acceptors, but for most cellobiose dehydrogenases the reaction rate of cellobiose oxidase for these acceptors is several orders of magnitude lower than that observed for iron or organic oxidants. Following

cellobionolactone release, the product may undergo spontaneous ring-opening to generate cellobionic acid (Hallberg *et al.*, 2003, J. Biol. Chem. 278: 7160-7166).

[00311] Those of skill in the art will appreciate that cellobiose dehydrogenase enzyme activity typically employs the presence of oxygen or an equivalent redox acceptor (*e.g.*, lignin, molecular oxygen, cytochrome c, redox dyes, benzoquinones and Fe²⁺ complexes).

[00312] Cellobiose dehydrogenase activity can be measured using any of a variety of methods known in the art. For example, CDH activity assays can be performed using any suitable method known in the art (*See e.g.*, Schou *et al.*, Biochem J., 220:565-71 [1998]). For example, DCPIP (2,6-dichlorophenolindophenol) reduction by CDH activity in the presence of cellobiose can be monitored by absorbance at 530 nm.

[00313] In some embodiments, the fungal cells provided herein that have been genetically modified to reduce the secreted activity of a cellobiose dehydrogenase have reduced secreted activity of an endogenous cellobiose dehydrogenase. Accordingly, one or more cellobiose dehydrogenase enzymes from each of the fungal species described herein can be targeted for genetic modification.

[00314] In some embodiments, the cellobiose dehydrogenase is from a fungal species in the division Basidiomycete and belonging to the class Agaricomycetes; or in the division Ascomycete and belonging to the subdivision Pezizomycotina. Some examples of cellobiose dehydrogenase enzymes identified from division Basidiomycete belonging to the class Agaricomycetes; and division Ascomycete belonging to the subdivision Pezizomycotina are set forth in Table C, below.

[00315] In some embodiments, the cellobiose dehydrogenase is from a fungal species selected from *Thielavia heterothallica*, *Thielavia terrestris*, *Chaetomium globosum* and *Myceliophthora thermophila*. Some cellobiose dehydrogenase enzymes identified from these and other species are set forth in the table below. The proteins listed in Table C are examples of cellobiose dehydrogenase that are known in the art, or identified herein as being a cellobiose dehydrogenase.

Table C. Cellobiose Dehydrogenase Sequences
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Accession Number	Organism	GMC oxred N Domain	GMC oxred C Domain
AC26221 (SEQ ID NO.: 6)	<i>Myceliophthora thermophila</i>	251-554	645-781
AAC26221	<i>Myceliophthora thermophila</i>	251-554	645-781
ABS45566	<i>Myriococcum thermophilum</i>	251-554	645-781
ABS45567	<i>Myriococcum thermophilum</i>	251-554	645-781
CHGT_03380	<i>Chaetomium globosum</i>	226-529	620-757
XP_001229896.1	<i>Chaetomium globosum</i> CBS 148.51	226-529	620-757
JGIThte5441	<i>Thielavia terrestris</i>	253-555	647-783
CAP68427	<i>Podospora anserina</i>	247-550	643-779
CBI53519.1	<i>Sordaria macrospora</i>	252-554	645-782
XP_956591.1	<i>Neurospora crassa</i> OR74A	253-555	646-782
XP_360402.2	<i>Magnaporthe oryzae</i> 70-15	264-566	657-794
EDP55266	<i>Aspergillus fumigatus</i> A1163	265-568	661-796
BAE61169	<i>Aspergillus oryzae</i>	254-556	647-782
CBI59551.1	<i>Sordaria macrospora</i>	167-295	382-518
EAW14611	<i>Aspergillus clavatus</i> NRRL 1	254-556	647-783
XP_001209295.1	<i>Aspergillus terreus</i> NIH2624	253-474	586-720
JGIThte4524	<i>Thielavia terrestris</i>	36-337	NA
CHGT_08276	<i>Chaetomium globosum</i>	36-338	NA
XP_001225932.1	<i>Chaetomium globosum</i> CBS 148.51	36-338	NA
JGIThte6738	<i>Thielavia terrestris</i>	249-550	642-779
CAP61651	<i>Podospora anserina</i>	254-555	647-783
AAF69005	<i>Humicola insolens</i>	247-548	640-776
XP_389261.1	<i>Gibberella zeae</i> PH-1	213-516	607-743
XP_958234.1	<i>Neurospora crassa</i> OR74A	274-576	668-804
CDH2 derived from a C1 strain (SEQ ID NO.: 8)	<i>Myceliophthora thermophila</i>	249-550	NA
CBI54739.1	<i>Sordaria macrospora</i>	272-574	666-802
XP_001800470.1	<i>Phaeosphaeria nodorum</i> SN15	36-343	366-502
XP_001939778.1	<i>Pyrenophora tritici-repentis</i> Pt-1C-BFP	247-550	640-776
CHGT_08622	<i>Chaetomium globosum</i>	249-521	549-667
XP_001226549.1	<i>Chaetomium globosum</i> CBS 148.51	249-521	549-667
XP_001801490.1	<i>Phaeosphaeria nodorum</i> SN15	245-543	633-769
XP_001553707.1	<i>Botryotinia fuckeliana</i> B05.10	265-570	656-796
XP_002999803.1	<i>Verticillium albo-atrum</i> VaMs.102	393-534	590-681
XP_001591237.1	<i>Sclerotinia sclerotiorum</i> 1980	265-570	655-796
XP_001273175.1	<i>Aspergillus clavatus</i> NRRL 1	244-543	627-752
XP_749254.1	<i>Aspergillus fumigatus</i> Af293	247-546	637-755
ACF60617	<i>Ceriporiopsis subvermispora</i>	236-519	634-763

Table C. Cellobiose Dehydrogenase Sequences

Accession Number	Organism	GMC oxred N Domain	GMC oxred C Domain
BAC20641	<i>Grifola frondosa</i>	230-509	628-757
AAC50004	<i>Trametes versicolor</i>	230-512	628-757
BAD32781	<i>Coniophora puteana</i>	236-519	634-763
XP_367658.1	<i>Magnaporthe oryzae</i> 70-15	39-334	426-551
BAD36748	<i>Irpex lacteus</i>	239-516	637-766
AAB61455	<i>Phanerochaete chrysosporium</i>	235-517	633-762
CAA61359	<i>Phanerochaete chrysosporium</i>	234-516	632-761
AAO32063	<i>Trametes versicolor</i>	230-512	628-757
AAO64483	<i>Athelia rolfsii</i>	233-520	631-760
XP_001265679.1	<i>Neosartorya fischeri</i> NRRL 181	247-546	639-755
AAC32197	<i>Pycnoporus cinnabarinus</i>	231-517	628-758
XP_383093.1	<i>Gibberella zeae</i> PH-1	27-325	408-528
2118247A	<i>Phanerochaete chrysosporium</i>	234-515	631-759
XP_001937164.1	<i>Pyrenophora tritici-repentis</i> Pt-1C-BFP	245-542	625-754
XP_001400060.1	<i>Aspergillus niger</i>	33-329	415-542
CAP85828	<i>Penicillium chrysogenum</i> Wisconsin 54-1255	30-327	393-537
XP_001803287.1	<i>Phaeosphaeria nodorum</i> SN15	243-516	NA
XP_003006847.1	<i>Verticillium albo-atrum</i> VaMs.102	27-317	411-529
XP_001402432.1	<i>Aspergillus niger</i> CBS 513.88	245-495	598-726
XP_003042062.1	<i>Nectria haematococca</i> mpVI 77-13-4	241-540	597-754
XP_001210806.1	<i>Aspergillus terreus</i> NIH2624	32-329	413-541
XP_386159.1	<i>Gibberella zeae</i> PH-1	25-323	406-530
BAE63115	<i>Aspergillus oryzae</i>	24-317	401-527
XP_001559563.1	<i>Botryotinia fuckeliana</i> B05.10	232-534	620-746
XP_003003908.1	<i>Verticillium albo-atrum</i> VaMs.102	232-507	NA
XP_003042935.1	<i>Nectria haematococca</i> mpVI 77-13-4	233-531	614-739
XP_383918.1	<i>Gibberella zeae</i> PH-1	234-533	615-740
XP_001793048.1	<i>Phaeosphaeria nodorum</i> SN15	28-314	408-542
BAE79276	<i>Fusarium oxysporum</i> f. sp. <i>Lycopersici</i>	27-325	408-532
XP_364344.1	<i>Magnaporthe oryzae</i> 70-15	49-321	426-560
XP_001547235.1	<i>Botryotinia fuckeliana</i> B05.10	30-313	422-548
XP_001593342.1	<i>Sclerotinia sclerotiorum</i> 1980	232-536	622-748
XP_385048.1	<i>Gibberella zeae</i> PH-1	239-538	595-752
XP_362950.1	<i>Magnaporthe oryzae</i> 70-15	30-327	415-537
XP_003052041.1	<i>Nectria haematococca</i> mpVI 77-13-4	32-332	415-539

Table C. Cellobiose Dehydrogenase Sequences			
Accession Number	Organism	GMC oxred N Domain	GMC oxred C Domain
XP_001940494.1	<i>Pyrenophora tritici-repentis</i> Pt-1C-BFP	90-271	363-479

*Accession numbers for *Thielavia terrestris* refer to the U.S. Department of Energy (DOE) Joint Genome Institute (JGI) genome sequence

[00316] Some amino acid sequences encoding cellobiose dehydrogenase are provided herein. For example, the nucleotide sequence encoding *Myceliophthora thermophila* CDH1 is set forth herein as SEQ ID NO:5, and the encoded amino acid sequence of *Myceliophthora thermophila* CDH1 is set forth as SEQ ID NO:6. Furthermore, the nucleotide sequence encoding *Myceliophthora thermophila* CDH2 is set forth herein as SEQ ID NO:7, and the encoded amino acid sequence of *Myceliophthora thermophila* CDH2 is set forth as SEQ ID NO:8.

[00317] In some embodiments, the cellobiose dehydrogenase is cellobiose dehydrogenase (EC 1.1.99.18). In some embodiments, the cellobiose dehydrogenase is a cellobiose dehydrogenase with the amino acid sequence of *Myceliophthora thermophila* CDH1 as set forth in SEQ ID NO:6. In some embodiments, the cellobiose dehydrogenase is a cellobiose dehydrogenase with the amino acid sequence of *Myceliophthora thermophila* CDH2 as set forth in SEQ ID NO:8. In other embodiments, the cellobiose dehydrogenase comprises an amino acid sequence provided in the GenBank entry of any one of the accession numbers set forth in Table C. In some embodiments, the cellobiose dehydrogenase is encoded by a nucleic acid sequence that is at least about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to SEQ ID NOS:5 and/or 7. In some embodiments, the cellobiose dehydrogenase is encoded by a nucleic acid sequence that is at least about 60%, about 61%,

about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to a nucleic acid sequence encoding the amino acid sequence set forth as SEQ ID NOS:6 and/or 8, or an amino acid sequence provided in the GenBank entry of any one of the accession numbers set forth in Table D. In some embodiments, the cellobiose dehydrogenase is encoded by a nucleic acid sequence that can selectively hybridize to SEQ ID NOS:5 and/or 7 under moderately stringent or stringent conditions, as described hereinabove. In some embodiments, the cellobiose dehydrogenase is encoded by a nucleic acid sequence that can selectively hybridize under moderately stringent or stringent conditions to a nucleic acid sequence that encodes SEQ ID NOS:6 and/or 8, or an amino acid sequence provided in the GenBank entry of any one of the accession numbers set forth in Table C.

[00318] In some embodiments, the cellobiose dehydrogenase comprises an amino acid sequence with at least about 50%, about 51%, about 52%, about 53%, about 54%, about 55%, about 56%, about 57%, about 58%, about 59%, about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% similarity to the amino acid sequence set forth as SEQ ID NOS:6 and/or 8, or an amino acid sequence provided in the GenBank entry of any one of the accession numbers set forth in Table C. Similarity as used herein is described in greater detail hereinabove.

[00319] Cellobiose dehydrogenase sequences can be identified by any of a variety of methods known in the art. For example, a sequence alignment can be conducted against a database, for example against the NCBI database, and sequences with the lowest HMM E-value can be selected.

[00320] Pyranose Oxidase. As indicated herein, pyranose oxidases and fungal cells that have been modified to have reduced pyranose oxidase activity find use in the present invention. Pyranose oxidase activity can be measured using any of a variety of methods known in the art. For example, PO activity assays can be performed as described by Leitner et al., Appl Biochem Biotechnol 1998, 70-72:237-248).

For example, 2'-azinobis(3-ethylbenzthiazolinesulfonic acid) (ABTS) reduction by PO activity can be monitored by absorbance at 530 nm. In some additional embodiments, PO activity is determined by an increase in absorbance at 420 nm resulting from the oxidation of 2,2'-azinobis(3-ethylbenzthiazolinesulfonic acid) (ABTS) through a peroxidase coupled system.

[00321] In some embodiments, the fungal cells provided herein that have been genetically modified to reduce the secreted activity of a pyranose oxidase have reduced secreted activity of an endogenous pyranose oxidase. Accordingly, one or more pyranose oxidase enzymes from each of the fungal species described herein can be targeted for genetic modification.

[00322] In some embodiments, the pyranose oxidase is from a fungal species in the division Basidiomycete and belonging to the class Agaricomycetes; or in the division Ascomycete and belonging to the subdivision Pezizomycotina. Some examples of pyranose oxidase enzymes identified from division Basidiomycete belonging to the class Agaricomycetes; and division Ascomycete belonging to the subdivision Pezizomycotina are set forth in Table D, below.

[00323] In some embodiments, the pyranose oxidase is from a fungal species selected from *Peniophora gigantea*, *Phanerochaete chrysosporium*, *Trametes ochracea*, *Trametes pubescens*, *Emericella nidulans*, *Aspergillus oryzae*, *Gloeophyllum trabeum*, *Tricholoma matsutake*, *Trametes hirsute*, *Gloeophyllum trabeum*, *Phanerochaete chrysosporium*, *Peniophora sp.*, *Trametes versicolor*, *Lyophyllum shimeji*, *Trametes pubescens*, *Phlebiopsis gigantea*, *Aspergillus parasiticus*, *Auricularia polytricha*, *Coriolus hirsutus*, *Coriolus versicolor*, *Gloeophyllum sepiarum*, *Iridophycus faccidum*, *Irpex lactus*, *Oudemansiella mucida*, *Phanerochaete gigantea*, *Pleurotus ostreatus*, *Polyporus obtusus*, *Saxidomus giganteus*, *Todus multicolor*, *Trametes cinnabarinus* and *Trametes multicolor*. Some pyranose oxidase enzymes identified from these species are set forth in Table D, below. The

proteins listed in the table below are examples of pyranose oxidase that are known in the art, or identified herein as being a pyranose oxidase.

Table D. Pyranose Oxidase Sequences			
<u>Database</u>	<u>Accession Number</u>	<u>Organism</u>	<u>Reference</u>
Swiss Prot	Q6UG02	<i>Peniophora gigantea</i>	
Swiss Prot	Q6QWR1	<i>Phanerochaete chrysosporium</i>	
Swiss Prot	Q7ZA32	<i>Trametes ochracea</i>	
Swiss Prot	Q5G234	<i>Trametes pubescens</i>	
GenPept	Q5B2E9	<i>Emericella nidulans</i>	
GenPept	BAE56707.1 (SEQ ID NO: 10)	<i>Aspergillus oryzae</i>	
GenPept	ACJ54278.1	<i>Gloeophyllum trabeum</i>	
GenPept	BAC24805.1	<i>Tricholoma matsutake</i>	
GenPept	P59097	<i>Trametes hirsuta</i>	
GenPept	ACM47528.1	<i>Gloeophyllum trabeum</i>	
GenPept	AAS93628.1	<i>Phanerochaete chrysosporium</i>	
GenPept	AAO13382.1	<i>Peniophora sp.</i>	
GenPept	BAA11119.1	<i>Trametes versicolor</i>	
GenPept	BAD1079.1	<i>Lyophyllum shimeji</i>	
GenPept	AAW57304.1	<i>Trametes pubescens</i>	
GenPept	AAQ72486.1	<i>Phlebiopsis gigantea</i>	
		<i>Aspergillus parasiticus</i>	Giffhorn, Appl. Microbiol. Biotechnol., 54: 727-740 (2000)
		<i>Auricularia polytricha</i>	Izumi <i>et al.</i> , Agric. Biol. Chem., 54: 799-801 (1990)
		<i>Coriolus hirsutus</i>	Machida <i>et al.</i> , Agric. Biol. Chem., 48: 2463-2470 (1984)
		<i>Coriolus versicolor</i>	Taguchi <i>et al.</i> , J. Appl. Biochem., 7: 289-295 (1985)
		<i>Gloeophyllum sepiarium</i>	Izumi <i>et al.</i> , Agric. Biol. Chem., 54: 799-801 (1990)
		<i>Iridophycus faccidum</i>	Giffhorn, Appl. Microbiol. Biotechnol., 54: 727-740 (2000)
		<i>Irpex lactus</i>	Izumi <i>et al.</i> , Agric. Biol. Chem., 54: 799-801 (1990)
		<i>Oudemansiella mucida</i>	Giffhorn, Appl. Microbiol. Biotechnol., 54: 727-740 (2000)
		<i>Phanerochaete</i>	Giffhorn, Appl. Microbiol. Biotechnol., 54: 727-

Table D. Pyranose Oxidase Sequences			
<u>Database</u>	<u>Accession Number</u>	<u>Organism</u>	<u>Reference</u>
		<i>gigantea</i>	740 (2000)
		<i>Pleurotus ostreatus</i>	Giffhorn, Appl. Microbiol. Biotechnol., 54: 727-740 (2000)
		<i>Polyporus obtusus</i>	Janssen <i>et al.</i> , Methods Enzymol., 41B: 170-173 (1975)
		<i>Saxidomus giganteus</i>	Giffhorn, Appl. Microbiol. Biotechnol., 54: 727-740 (2000)
		<i>Todus multicolor</i>	Giffhorn, Appl. Microbiol. Biotechnol., 54: 727-740 (2000)
		<i>Trametes cinnabarinus</i>	Izumi <i>et al.</i> , Biol. Chem., 54: 799-801 (1990)
		<i>Trametes multicolor</i>	Tasca <i>et al.</i> , Electroanal., 19: 294-302 (2007)

[00324] Some amino acid sequences encoding pyranose oxidase are provided herein. For example, the nucleotide sequence encoding *Aspergillus oryzae* PO1 is set forth herein as SEQ ID NO:9, and the encoded amino acid sequence of *Aspergillus oryzae* PO1 is set forth as SEQ ID NO:10.

[00325] In some embodiments, the pyranose oxidase is pyranose oxidase (E.C. 1.1.3.10). In some embodiments, the pyranose oxidase is a pyranose oxidase with the amino acid sequence of *Aspergillus oryzae* PO1 as set forth in SEQ ID NO: 10. In other embodiments, the pyranose oxidase comprises an amino acid sequence provided in the literature reference and/or GenBank entry of any one of the accession numbers set forth in Table D. In some embodiments, the pyranose oxidase is encoded by a nucleic acid sequence that is at least about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to SEQ ID NO:9. In some embodiments, the pyranose oxidase is encoded by a nucleic acid sequence that is at least about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about

82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to a nucleic acid sequence encoding the amino acid sequence set forth as SEQ ID NO:10, or an amino acid sequence provided in the literature reference and/or GenBank entry of any one of the accession numbers set forth in Table D. In some embodiments, the pyranose oxidase is encoded by a nucleic acid sequence that can selectively hybridize to SEQ ID NO:9 under moderately stringent or stringent conditions, as described hereinabove. In some embodiments, the pyranose oxidase is encoded by a nucleic acid sequence that can selectively hybridize under moderately stringent or stringent conditions to a nucleic acid sequence that encodes SEQ ID NO:10, or an amino acid sequence provided in the literature reference and/or GenBank entry of any one of the accession numbers set forth in Table D.

[00326] In some embodiments, the pyranose oxidase comprises an amino acid sequence with at least about 50%, about 51%, about 52%, about 53%, about 54%, about 55%, about 56%, about 57%, about 58%, about 59%, about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% similarity to the amino acid sequence set forth as SEQ ID NO:10, or an amino acid sequence provided in the literature reference and/or GenBank entry of any one of the accession numbers set forth in Table D. Similarity as used herein is described in greater detail hereinabove.

[00327] Pyranose oxidase sequences can be identified by any of a variety of methods known in the art. For example, a sequence alignment can be conducted against a database, for example against the NCBI database, and sequences with the lowest HMM E-value can be selected.

[00328] Glucoligosaccharide Oxidase. As indicated herein, glucoligosaccharide oxidases and fungal cells modified to have reduced GOOX activity find use in the present invention. Glucoligosaccharide oxidase activity can be measured using any of a variety of

methods known in the art. For example, GOOX activity assays can be performed as described by Lin et al., (Biochim. Biophys. Acta. 1991, 11:417-427), or Lee et al. (Appl. Environ. Microbiol. 2005, 71:8881-8887). For example, activity can be measured by determining H₂O₂ production by coupling to a peroxidase enzyme assay.

[00329] In some embodiments, the fungal cells provided herein that have been genetically modified to reduce the secreted activity of at least one glucooligosaccharide oxidase have reduced secreted activity of an endogenous glucooligosaccharide oxidase. Accordingly, one or more glucooligosaccharide oxidase enzymes from each of the fungal species described herein can be targeted for genetic modification.

[00330] In some embodiments, the glucooligosaccharide oxidase is from a fungal species in the division Basidiomycete and belonging to the class Agaricomycetes; or in the division Ascomycete and belonging to the subdivision Pezizomycotina. Some examples of glucooligosaccharide oxidase identified from division Basidiomycete belonging to the class Agaricomycetes; and division Ascomycete belonging to the subdivision Pezizomycotina are set forth in the table below.

[00331] In some embodiments, the glucooligosaccharide oxidase is from a fungal species selected from *Acremonium strictum* and *Paraconiothyrium sp.* Some glucooligosaccharide oxidase enzymes identified from these species are set forth in Table E, below. The proteins listed in the table below are examples of glucooligosaccharide oxidase that are known in the art, or identified herein as being a glucooligosaccharide oxidase.

Table E. Glucooligosaccharide Oxidase Sequences			
Database	Accession Number	Organism	Reference
TrEMBL UniProt	Q6PW77 (SEQ ID NO: 12)	<i>Acremonium strictum</i>	
		<i>Paraconiothyrium sp.</i>	Kiryu <i>et al.</i> , 2008, Biosci. Biotechnol. Biochem., 72: 833-841 (2008)

[00332] Some amino acid sequences encoding glucooligosaccharide oxidase are provided herein. For example, the nucleotide sequence encoding *Acremonium strictum* GOOX1 is set forth herein as SEQ ID NO:11, and the encoded amino acid sequence of *Acremonium strictum* GOOX1 is set forth as SEQ ID NO:12.

[00333] In some embodiments, the glucooligosaccharide oxidase is glucooligosaccharide oxidase (E.C. 1.1.99.B3). In some embodiments, the glucooligosaccharide oxidase is a glucooligosaccharide oxidase with the amino acid sequence of *Acremonium strictum* GOOX1 as set forth in SEQ ID NO:12. In other embodiments, the glucooligosaccharide oxidase comprises an amino acid sequence provided in the literature reference and/or GenBank entry of any one of the accession numbers set forth in Table E. In some embodiments, the glucooligosaccharide oxidase is encoded by a nucleic acid sequence that is at least about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to SEQ ID NO:11. In some embodiments, the glucooligosaccharide oxidase is encoded by a nucleic acid sequence that is at least about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to a nucleic acid sequence encoding the amino acid sequence set forth as SEQ ID NO:12, or an amino acid sequence provided in the literature reference and/or the GenBank entry of any one of the accession numbers set forth in Table E. In some embodiments, the glucooligosaccharide oxidase is encoded by a nucleic acid sequence that can selectively hybridize to SEQ ID NO:11 under moderately stringent or stringent conditions, as described hereinabove. In some embodiments, the glucooligosaccharide oxidase is encoded by a nucleic acid sequence that can selectively hybridize under moderately stringent or stringent

conditions to a nucleic acid sequence that encodes SEQ ID NO:12, or an amino acid sequence provided in the literature reference and/or GenBank entry of any one of the accession numbers set forth in Table E.

[00334] In some embodiments, the glucooligosaccharide oxidase comprises an amino acid sequence with at least about 50%, about 51%, about 52%, about 53%, about 54%, about 55%, about 56%, about 57%, about 58%, about 59%, about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% similarity to the amino acid sequence set forth as SEQ ID NO:12, or an amino acid sequence provided in the literature reference and/or the GenBank entry of any one of the accession numbers set forth in Table E. Similarity as used herein is described in greater detail hereinabove.

[00335] Glucooligosaccharide oxidase sequences can be identified by any of a variety of methods known in the art. For example, a sequence alignment can be conducted against a database, for example against the NCBI database, and sequences with the lowest HMM E-value can be selected.

[00336] Pyranose Dehydrogenase. In some embodiments, pyranose dehydrogenases and fungal cells that have reduced pyranose dehydrogenase activity find use in the present invention. Pyranose dehydrogenases activity can be measured using any of a variety of methods known in the art. For example, PDH activity assays can be performed using any suitable method (*See e.g., Volc et al., Arch. Microbiol., 176:178–186 [2001]*).

[00337] In some embodiments, the fungal cells that have been genetically modified to reduce the secreted activity of a pyranose dehydrogenase have reduced secreted activity of an endogenous pyranose dehydrogenase. Accordingly, one or more pyranose dehydrogenase enzymes from each of the fungal species described herein can be targeted for genetic modification.

[00338] In some embodiments, the pyranose dehydrogenase is from a fungal species in the division Basidiomycete and belonging to the class Agaricomycetes; or in the division

Ascomycete and belonging to the subdivision Pezizomycotina. Some examples of pyranose dehydrogenase identified from division Basidiomycete belonging to the class Agaricomycetes; and division Ascomycete belonging to the subdivision Pezizomycotina are set forth in the table below.

[00339] In some embodiments, the pyranose dehydrogenase is from a fungal species selected from *Agaricus bisporus*, *Agaricus meleagris*, *Agaricus xanthoderma*, *Macroleptota rhacodes* and *Leucoagaricus meleagris*. Some pyranose dehydrogenase enzymes identified from these species are set forth in Table F, below. The proteins listed in the table below are examples of pyranose dehydrogenase that are known in the art, or identified herein as being a pyranose dehydrogenase.

Table F. Pyranose Dehydrogenase Sequences			
<u>Database</u>	<u>Accession Number</u>	<u>Organism</u>	<u>Reference</u>
TrEMBL UniProt	Q3L1D1 (SEQ ID NO:14)	<i>Agaricus bisporus</i>	
TrEMBL UniProt	Q3L245	<i>Agaricus meleagris</i>	
TrEMBL UniProt	Q0R4L2	<i>Agaricus meleagris</i>	
TrEMBL UniProt	Q3L243	<i>Agaricus meleagris</i>	
TrEMBL UniProt	Q3L1D2	<i>Agaricus xanthoderma</i>	
		<i>Macroleptota rhacodes</i>	Volc <i>et al.</i> , Arch. Microbiol., 176:178-186 (2001)
GenBank	AAW82996.1	<i>Leucoagaricus meleagris</i>	
GenBank	AAW82998.1	<i>Leucoagaricus meleagris</i>	
GenBank	AAZ94874.1	<i>Leucoagaricus meleagris</i>	

[00340] Some amino acid sequences encoding pyranose dehydrogenase are provided herein. For example, the nucleotide sequence encoding *Agaricus bisporus* PDH1 is set forth herein as SEQ ID NO:13, and the encoded amino acid sequence of *Agaricus bisporus* PDH1 is set forth as SEQ ID NO:14.

[00341] In some embodiments, the pyranose dehydrogenase is pyranose dehydrogenase (E.C. 1.1.99.29). In some embodiments, the pyranose dehydrogenase is a pyranose dehydrogenase with the amino acid sequence of *Agaricus bisporus* PDH1 as set forth in SEQ ID NO:14. In some other embodiments, the pyranose dehydrogenase comprises an amino acid sequence provided in the literature reference and/or GenBank entry of any one of the accession numbers set forth in Table F. In some embodiments, the pyranose dehydrogenase is encoded by a nucleic acid sequence that is at least about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to SEQ ID NO:13. In some embodiments, the pyranose dehydrogenase is encoded by a nucleic acid sequence that is at least about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to a nucleic acid sequence encoding the amino acid sequence set forth as SEQ ID NO:14, or an amino acid sequence provided in the literature reference and/or GenBank entry of any one of the accession numbers set forth in Table F. In some embodiments, the pyranose dehydrogenase is encoded by a nucleic acid sequence that can selectively hybridize to SEQ ID NO:13 under moderately stringent or stringent conditions, as described hereinabove. In some embodiments, the pyranose dehydrogenase is encoded by a nucleic acid sequence that can selectively hybridize under moderately stringent or stringent conditions to a nucleic acid sequence that encodes SEQ ID NO:14, or an amino acid sequence provided in the literature reference and/or GenBank entry of any one of the accession numbers set forth in Table F.

[00342] In some embodiments, the pyranose dehydrogenase comprises an amino acid sequence with at least about 50%, about 51%, about 52%, about 53%, about 54%, about

55%, about 56%, about 57%, about 58%, about 59%, about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% similarity to the amino acid sequence set forth as SEQ ID NO:14, or an amino acid sequence provided in the literature reference and/or the GenBank entry of any one of the accession numbers set forth in Table F. Similarity as used herein is described in greater detail hereinabove.

[00343] Pyranose dehydrogenase sequences can be identified by any of a variety of methods known in the art. For example, a sequence alignment can be conducted against a database, for example against the NCBI database, and sequences with the lowest HMM E-value can be selected.

[00344] Glucose Dehydrogenase. As indicated herein, glucose dehydrogenases and fungal cells that have been modified to have reduced glucose dehydrogenase activity find use in the present invention. Glucose dehydrogenase activity can be measured using any of a variety of methods known in the art (*See e.g.*, Strecker, Meth. Enzymol., 1:335 [1955]). In some embodiments, GDH activity is determined by an increase in absorbance at 340 nm resulting from the generation of NADH from NAD when beta-D-glucose is provided as a substrate and NAD as an acceptor.

[00345] In some embodiments, the fungal cells provided herein that have been genetically modified to reduce the secreted activity of a glucose dehydrogenase have reduced secreted activity of an endogenous glucose dehydrogenase. Accordingly, one or more glucose dehydrogenase enzymes from each of the fungal species described herein can be targeted for genetic modification.

[00346] In some embodiments, the glucose dehydrogenase is from a fungal species in the division Basidiomycete and belonging to the class Agaricomycetes; or in the division Ascomycete and belonging to the subdivision Pezizomycotina. Some examples of glucose dehydrogenase identified from division Basidiomycete belonging to the class

Agaricomycetes; and division Ascomycete belonging to the subdivision Pezizomycotina are set forth in the table below.

[00347] In some embodiments, the glucose dehydrogenase is from a fungal species selected from *Aspergillus niger*, *Aspergillus oryzae*, *Aspergillus terreus* and *Talaromyces stipitatus*. Some glucose dehydrogenase enzymes identified from these species are set forth in Table G, below. The proteins listed in the table below are examples of glucose dehydrogenase that are known in the art, or identified herein as being a glucose dehydrogenase.

Table G. Glucose Dehydrogenase Sequences			
Database	Accession Number	Organism	Reference
		<i>Aspergillus niger</i>	Muller, Zentralbl. Bacteriol. Parasienkd. Infektionskr Hyg., 132(a): 14-24 (1977)
		<i>Aspergillus oryzae</i>	Bak, Biochim. Biophys. Acta 139: 277-293 (1967)
		<i>Aspergillus terreus</i>	Tsujimura <i>et al.</i> , (2006) Biosci. Biotechnol. Biochem., 70: 654-659 (2006)
GenBank	XP_002482522.1 (SEQ ID NO: 16)	<i>Talaromyces stipitatus</i> ATCC 10500	
GenBank	XP_002479433.1	<i>Talaromyces stipitatus</i> ATCC 10500	
GenBank	XP_002481914.1	<i>Talaromyces stipitatus</i> ATCC 10500	

[00348] Some amino acid sequences encoding glucose dehydrogenase are provided herein. For example, the nucleotide sequence encoding *Myceliophthora thermophila* GDH1 is set forth herein as SEQ ID NO:15, and the encoded amino acid sequence of *Myceliophthora thermophila* GDH1 is set forth as SEQ ID NO:16.

[00349] In some embodiments, the glucose dehydrogenase is glucose dehydrogenase (E.C. 1.1.99.10). In some embodiments, the glucose dehydrogenase is a glucose dehydrogenase with the amino acid sequence of *Myceliophthora thermophila* GDH1 as set forth in SEQ ID NO:16. In some other embodiments, the glucose dehydrogenase comprises an amino acid sequence provided in the literature reference and/or GenBank entry of any one

of the accession numbers set forth in Table G. In some embodiments, the glucose dehydrogenase is encoded by a nucleic acid sequence that is at least about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to SEQ ID NO:15. In some embodiments, the glucose dehydrogenase is encoded by a nucleic acid sequence that is at least about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to a nucleic acid sequence encoding the amino acid sequence set forth as SEQ ID NO:16, or an amino acid sequence provided in the literature reference and/or GenBank entry of any one of the accession numbers set forth in Table G. In some embodiments, the glucose dehydrogenase is encoded by a nucleic acid sequence that can selectively hybridize to SEQ ID NO:15 under moderately stringent or stringent conditions, as described hereinabove. In some embodiments, the glucose dehydrogenase is encoded by a nucleic acid sequence that can selectively hybridize under moderately stringent or stringent conditions to a nucleic acid sequence that encodes SEQ ID NO:16, or an amino acid sequence provided in the literature reference and/or GenBank entry of any one of the accession numbers set forth in Table G.

[00350] In some embodiments, the glucose dehydrogenase comprises an amino acid sequence with at least about 50%, about 51%, about 52%, about 53%, about 54%, about 55%, about 56%, about 57%, about 58%, about 59%, about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%,

about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% similarity to the amino acid sequence set forth as SEQ ID NO:16, or an amino acid sequence provided in the literature reference or the GenBank entry of any one of the accession numbers set forth in Table G. Similarity as used herein is described in greater detail hereinabove.

[00351] Glucose dehydrogenase sequences can be identified by any of a variety of methods known in the art. For example, a sequence alignment can be conducted against a database, for example against the NCBI database, and sequences with the lowest HMM E-value selected, as desired.

[00352] In some embodiments, the cell has been genetically modified to reduce the amount of glucose and/or cellobiose oxidizing enzyme activity from two or more endogenous glucose and/or cellobiose oxidizing enzymes that are secreted by the cell. In certain such embodiments, a first of the two or more the glucose and/or cellobiose oxidizing enzymes comprises an amino acid sequence that is at least about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to any one of SEQ ID NOS:2, 4, 6, 8, 10, 12, 14 and/or 16 and a second of the two or more the glucose and/or cellobiose oxidizing enzymes comprises an amino acid sequence that is at least about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identical to any one of SEQ ID NOS: 2, 4, 6, 8, 10, 12, 14 and/or 16.

Enzyme Mixtures

[00353] Also provided herein are enzyme mixtures that comprise at least one or more cellulose hydrolyzing enzymes expressed by a fungal cell that has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell, as described hereinabove.

[00354] In some embodiments the enzyme mixture is in a vessel comprising a genetically modified fungal cell as described hereinabove. In some embodiments, the vessel comprises a liquid medium. For example, the vessel can be a flask, bioprocess reactor, and the like. In some embodiments, the enzyme mixture is in a liquid volume. For example, the liquid volume can be greater than about 0.01 mL, 0.1 mL, 1 mL, 10 mL, 100 mL, 1000 mL, or greater than about 10 L, 50 L, 100 L, 200 L, 300 L, 400 L, 500 L, 600 L, 700 L, 800 L, 900 L, 1000 L, 10,000 L, 50,000 L, 100,000 L, 250,000 L, and 500,000 L or greater than about 1,000,000 L.

[00355] In some embodiments, the fungal cell is a lignocellulose-utilizing cell that is a Basidiomycete belonging to the class Agaricomycetes or an Ascomycete belonging to the subdivision Pezizomycotina, and where the fungal cell is capable of secreting a cellulase-containing enzyme mixture. In some embodiments, the fungal cell is capable of secreting an enzyme mixture comprising two or more cellulase enzymes. In some embodiments, the Basidiomycete is a species of *Pleurotus*, *Peniophora*, *Trametes*, *Athelia*, *Sclerotium*, *Termitomyces*, *Flammulina*, *Coniophora*, *Ganoderma*, *Pycnoporus*, *Ceriporiopsis*, *Phanerochaete*, *Gloeophyllum*, *Hericium*, *Heterobasidion*, *Gelatoporia*, *Lepiota*, or *Irpex*. In some embodiments, the Ascomycete is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Neurospora*, *Sordaria*, *Podospora*, *Magnaporthe*, *Fusarium*, *Gibberella*, *Botryotinia*, *Humicola*, *Neosartorya*, *Pyrenophora*, *Phaeosphaeria*, *Sclerotinia*, *Chaetomium*, *Nectria*, *Verticillium*, or *Aspergillus*.

[00356] In some embodiments, the fungal cell is a lignocellulose-utilizing cell from the family Chaetomiaceae. In some embodiments, the genetically modified fungal cell provided herein is a Chaetomiaceae family member selected from the genera *Myceliophthora*, *Thielavia*, *Corynascus*, or *Chaetomium*. The genetically modified fungal cell can also be an anamorph or teleomorph of a Chaetomiaceae family member selected from the genera *Myceliophthora*, *Thielavia*, *Corynascus*, or *Chaetomium*. As such, the genetically modified fungal cell can also be selected from the genera *Sporotrichum*,

Acremonium or *Talaromyces*. It is also contemplated that the genetically modified fungal cell be selected from the genera *Ctenomyces*, *Thermoascus*, and *Scytalidium*, including anamorphs and teleomorphs of fungal cells from those genera. In some embodiments, the fungal cell is a species selected from *Sporotrichum cellulophilum*, *Thielavia heterothallica*, *Corynascus heterothallicus*, *Thielavia terrestris*, *Chaetomium globosum*, *Talaromyces stipitatus* and *Myceliophthora thermophila*, including anamorphs and teleomorphs thereof.

[00357] In some embodiments, the present invention provides enzyme mixtures that are cell-free. In some embodiments, two or more cellulases and any additional enzymes and/or other components present in the cellulase enzyme mixtures are produced by a single type of genetically modified fungal cells, while in some embodiments the cellulases and/or other enzymes and/or other components are produced by different microbes. In some embodiments, the fermentations comprise single genetically modified cells and/or different microorganisms in combination, while in some other embodiments, the cells are grown in separate fermentations. Similarly, in some embodiments, the two or more cellulases and/or any additional enzymes and/or other components present in the cellulase enzyme mixture are expressed individually or in sub-groups from different strains of different organisms and the enzymes combined *in vitro* to produce the cellulase enzyme mixture. In some embodiments, cellulases and/or any additional enzymes and/or other components in the enzyme mixture are expressed individually or in sub-groups from different strains of a single organism, and the enzymes combined to make the cellulase enzyme mixture. In some embodiments, all of the enzymes and/or other components are expressed from a single host organism, such the genetically modified fungal cell as describe herein above.

[00358] In some embodiments, the enzyme mixtures comprise at least one or more cellulose hydrolyzing enzymes expressed by a fungal cell that has been genetically modified to reduce the amount of endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the cell, as described hereinabove.

[00359] In some embodiments, the fungal cell is a lignocellulose-utilizing cell that is a Basidiomycete belonging to the class Agaricomycetes or an Ascomycete belonging to the subdivision Pezizomycotina, and where the fungal cell is capable of secreting a cellulase-containing enzyme mixture. In some embodiments, the fungal cell is capable of secreting an enzyme mixture comprising two or more cellulase enzymes. In some embodiments, the

Basidiomycete is a species of *Pleurotus*, *Peniophora*, *Trametes*, *Athelia*, *Sclerotium*, *Termitomyces*, *Flammulina*, *Coniophora*, *Ganoderma*, *Pycnoporus*, *Ceriporiopsis*, *Phanerochaete*, *Gloeophyllum*, *Hericium*, *Heterobasidion*, *Gelatoporia*, *Lepiota*, or *Irpex*. In some embodiments, the Ascomycete is a species of *Myceliophthora*, *Thielavia*, *Sporotrichum*, *Neurospora*, *Sordaria*, *Podospira*, *Magnaporthe*, *Fusarium*, *Gibberella*, *Botryotinia*, *Humicola*, *Neosartorya*, *Pyrenophora*, *Phaeosphaeria*, *Sclerotinia*, *Chaetomium*, *Nectria*, *Verticillium*, or *Aspergillus*.

[00360] In some embodiments, the fungal cell is a lignocellulose-utilizing cell from the family Chaetomiaceae. In some embodiments, the genetically modified fungal cell provided herein is a Chaetomiaceae family member selected from the genera *Myceliophthora*, *Thielavia*, *Corynascus*, and *Chaetomium*. The genetically modified fungal cell can also be an anamorph or teleomorph of a Chaetomiaceae family member selected from the genera *Myceliophthora*, *Thielavia*, *Corynascus*, and *Chaetomium*. As such, the genetically modified fungal cell can also be selected from the genera *Sporotrichum* or *Acremonium*. It is also contemplated that the genetically modified fungal cell can also be selected from the genera *Ctenomyces*, *Scytalidium* and *Thermoascus*, including anamorphs and teleomorphs of fungal cells from those genera. Typically, the fungal cell is a species selected from *Sporotrichum cellulophilum*, *Thielavia heterothallica*, *Corynascus heterothallicus*, *Thielavia terrestris*, *Chaetomium globosum*, *Talaromyces stipitatus*, and *Myceliophthora thermophila*, including anamorphs and teleomorphs thereof.

[00361] Some cellulase mixtures for efficient enzymatic hydrolysis of cellulose that are known (See e.g., Viikari *et al.*, Adv. Biochem. Eng. Biotechnol., 108:121-45 [2007]; and US Pat. Appln. Publn. Nos. US 2009/0061484, US 2008/0057541, and US 2009/0209009) find use as components of some enzyme mixtures provided herein. In some embodiments, mixtures of purified naturally occurring or recombinant enzymes are combined with cellulosic feedstock or a product of cellulose hydrolysis. Alternatively or in addition, one or more cell populations, each producing one or more naturally occurring or recombinant cellulases, are combined with cellulosic feedstock or a product of cellulose hydrolysis. In some embodiments, the enzyme mixture comprises commercially available purified cellulases. Commercial cellulases are known and available to the art. In some embodiments, the enzyme mixtures do not comprise an endoglucanase.

[00362] In some embodiments, the enzyme mixture comprises at least 5%, at least 10%, at least 15%, at least 20%, at least 25%, at least 30%, at least 35%, at least 40%, at least 45%, or at least 50% GH61. In some embodiments, the enzyme mixture further comprises a cellobiohydrolase1a (*e.g.*, CBH1a) and GH61, wherein the enzymes together comprise at least 25%, at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, or at least 80% of the enzyme mixture. In some embodiments, the enzyme mixture further comprises a β -glucosidase (Bgl), GH61, and CBH, wherein the three enzymes together comprise at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, or at least 85% of the enzyme mixture. In some embodiments, the enzyme mixture further comprises an endoglucanase (EG), GH61, CBH2b, CBH1a, Bgl, wherein the five enzymes together comprise at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, or at least 90% of the enzyme mixture. In some embodiments, the enzyme mixture comprises GH61, CBH2b, CBH1, Bgl, and at least one EG, in any suitable proportion for the desired reaction.

[00363] In some embodiments, the enzyme mixture composition comprises isolated cellulases in the following proportions by weight (wherein the total weight of the cellulases is 100%), about 20%-10% of Bgl, about 30%-25% of CBH1a, about 10%-30% of GH61, about 20%-10% of EG, and about 20%-25% of CBH2b. In some embodiments, the enzyme mixture composition comprises isolated cellulases in the following proportions by weight: about 20%-10% of GH61, about 25%-15% of Bgl, about 20%-30% of CBH1a, about 10%-15% of EG, and about 25%-30% of CBH2b. In some embodiments, the enzyme mixture composition comprises isolated cellulases in the following proportions by weight: about 30%-20% of GH61, about 15%-10% of Bgl, about 25%-10% of CBH1a, about 25%-10% of CBH2b, about 15%-10% of EG. In some embodiments, the enzyme mixture composition comprises isolated cellulases in the following proportions by weight: about 40%-30% of GH61, about 15%-10% of Bgl, about 20%-10% of CBH1a, about 20%-10% of CBH2b, and about 15%-10% of EG. In some embodiments, the enzyme mixture composition comprises isolated cellulases in the following proportions by weight: about 50%-40% of GH61, about 15%-10% of Bgl, about 20%-10% of CBH1a, about 15%-10% of CBH2b, and about 10%-

5% of EG. In some embodiments, the enzyme mixture composition comprises isolated cellulases in the following proportions by weight: about 10%-15% of GH61, about 20%-25% of Bgl, about 30%-20% of CBH1a, about 15%-5% of EG, and about 25%-35% of CBH2b. In some embodiments, the enzyme mixture composition comprises isolated cellulases in the following proportions by weight: about 15%-5% of GH61, about 15%-10% of Bgl, about 45%-30% of CBH1a, about 25%-5% of EG, and about 40%-10% of CBH2b. In some embodiments, the enzyme mixture composition comprises isolated cellulases in the following proportions by weight: about 10% of GH61, about 15% of Bgl, about 40% of CBH1a, about 25% of EG, and about 10% of CBH2b.

[00364] In some embodiments, the enzyme component comprises more than one CBH1a, CBH2b, EG, Bgl, and/or GH61 enzyme (*e.g.*, 2, 3, 4, or more different variants), in any suitable combination. In some embodiments, an enzyme mixture composition of the invention further comprises at least one additional protein and/or enzyme. In some embodiments, enzyme mixture compositions of the present invention further comprise at least one additional enzyme other than the GH61, Bgl, CBH1a, GH61, and/or CBH2b. In some embodiments, the enzyme mixture compositions of the invention further comprise at least one additional cellulase, other than the GH61, Bgl, CBH1a, GH61, and/or CBH2b variant recited herein. In some embodiments, the GH61 polypeptide of the invention is also present in mixtures with non-cellulase enzymes that degrade cellulose, hemicellulose, pectin, and/or lignocellulose.

[00365] In some embodiments, GH61 polypeptide of the present invention is used in combination with other optional ingredients such as at least one buffer, surfactant, and/or scouring agent. In some embodiments, at least one buffer is used with the GH61 polypeptide of the present invention (optionally combined with other enzymes) to maintain a desired pH within the solution in which the GH61 is employed. The exact concentration of buffer employed depends on several factors which the skilled artisan can determine. Suitable buffers are well known in the art. In some embodiments, at least one surfactant is used in with the GH61 of the present invention. Suitable surfactants include any surfactant compatible with the GH61 and, optionally, with any other enzymes being used in the mixture. Exemplary surfactants include an anionic, a non-ionic, and ampholytic surfactants. Suitable anionic surfactants include, but are not limited to, linear or branched

alkylbenzenesulfonates; alkyl or alkenyl ether sulfates having linear or branched alkyl groups or alkenyl groups; alkyl or alkenyl sulfates; olefinsulfonates; alkanesulfonates, and the like. Suitable counter ions for anionic surfactants include, for example, alkali metal ions, such as sodium and potassium; alkaline earth metal ions, such as calcium and magnesium; ammonium ion; and alkanolamines having from 1 to 3 alkanol groups of carbon number 2 or 3. Ampholytic surfactants suitable for use in the practice of the present invention include, for example, quaternary ammonium salt sulfonates, betaine-type ampholytic surfactants, and the like. Suitable nonionic surfactants generally include polyoxalkylene ethers, as well as higher fatty acid alkanolamides or alkylene oxide adduct thereof, fatty acid glycerine monoesters, and the like. Mixtures of surfactants also find use in the present invention, as is known in the art.

[00366] In some embodiments, the cellulase enzyme mixtures of the present invention are produced in a fermentation process in which the fungal cell described herein above is grown in submerged liquid culture fermentation. It is intended that any suitable fermentation medium and process will find use in the present invention. In some embodiments, submerged liquid fermentations of fungal cells are conducted as a batch, fed-batch and/or continuous process. It is not intended that the present invention be limited to any particular fermentation medium, protocol, process, and/or equipment. In some embodiments, the fermentation medium is a liquid comprising a carbon source, a nitrogen source, and other nutrients, vitamins and minerals which can be added to the fermentation media to improve growth and enzyme production of the host cell. In some embodiments, these other media components are added prior to, simultaneously with or after inoculation of the culture with the host cell. In some embodiments, the carbon source comprises a carbohydrate that induces the expression of the cellulase enzymes from the fungal cell. For example, in some embodiments, the carbon source comprises one or more of cellulose, cellobiose, sophorose, xylan, xylose, xylobiose and related oligo- or poly-saccharides known to induce expression of cellulases and beta-glucosidase in such fungal cells. In some embodiments, the media comprise cellulose, while in some other embodiments, the media do not comprise cellulose (*i.e.*, measurable concentrations of cellulose). In some further embodiments, the media comprise carbon sources such as glucose, dextrose, etc. However, it is not intended that the present invention be limited to any specific carbon and/or nitrogen source, as any suitable

carbon and/or nitrogen source finds use in the present invention. Indeed, it is not intended that the present invention be limited to any particular medium, as any suitable medium will find use in the desired setting.

[00367] In some embodiments utilizing batch fermentation, the carbon source is added to the fermentation medium prior to or simultaneously with inoculation. In some other embodiments utilizing fed-batch and/or continuous operations, the carbon source is also supplied continuously or intermittently during the fermentation process. For example, in some embodiments, the carbon source is supplied at a carbon feed rate of between about 0.2 and about 2.5 g carbon/L of culture/h, or any amount therebetween. In some additional embodiments, the carbon source is supplied at a feed rate of between about 0.1 and about 10 g carbon/L of culture/hour or at any suitable rate therebetween (*e.g.*, about 0.1, about 0.2, about 0.3, about 0.4, about 0.5, about 0.6, about 0.7, about 0.8, about 0.9, about 1, about 2, about 3, about 4, about 5, about 6, about 7, about 8, about 9, or about 10 g carbon/L of culture/h).

[00368] In some embodiments, the process for producing the enzyme mixture of the present invention is performed at a temperature from about 20°C to about 80°C, or any temperature therebetween, for example from about 25°C to about 65°C, or any temperature therebetween, or from about 20°C, about 21°C, about 22°C, about 23°C, about 24 °C, about 25°C, about 26°C, about 27°C, about 28°C, about 29°C, about 30°C, about 31°C, about 32°C, about 33°C, about 34°C, about 35°C, about 36°C, about 37°C, about 38°C, about 39°C, about 40°C, about 41°C, about 42°C, about 43°C, about 44°C, about 45°C, about 46°C, about 47°C, about 48°C, about 49°C, about 50°C, about 51°C, about 52°C, about 53°C, about 54°C, about 55°C, about 56°C, about 57°C, about 58°C, about 59°C, about 60°C, about 61°C, about 62°C, about 63°C, about 64°C, about 65°C, about 66°C, about 67°C, about 68 °C, about 69°C, about 70°C, about 71°C, about 72°C, about 73°C, about 74°C, about 75°C, about 76°C, about 77°C, about 78°C, about 79°C, about 80°C, or any temperature therebetween.

[00369] In some embodiments, the methods for producing enzyme mixtures of the present invention are carried out at a pH from about 3.0 to about 8.0, or any pH therebetween, for example from about pH 3.5 to about pH 6.8, or any pH therebetween, for example from about pH 3.0, about 3.1, about 3.2, about 3.3, about 3.4, about 3.5, about 3.6,

about 3.7, about 3.8, about 3.9, about 4.0, about 4.1, about 4.2, about 4.3, about 4.4, about 4.5, about 4.6, about 4.7, about 4.8, about 4.9, about 5.0, about 5.1, about 5.2, about 5.3, about 5.4, about 5.5, about 5.6, about 5.7, about 5.8, about 5.9, about 6.0, about 6.1, about 6.2, about 6.3, about 6.4, 6.5, about 6.6, about 6.7, about 6.8, about 6.9, about 7.0, about 7.1, about 7.2, about 7.3, about 7.4, 7.5, about 7.6, about 7.7, about 7.8, about 7.9, about 8.0, or any pH therebetween.

[00370] In some embodiments, the fermentation medium containing the fungal cells are used following fermentation, while in some other embodiments, the fermentation medium containing the fungal cells and the enzyme mixture are used, while in some additional embodiments, an enzyme mixture is separated from the fungal cells (*e.g.*, by filtration and/or centrifugation), and the enzyme mixture in the fermentation medium is used, and in still additional embodiments, the fungal cells, enzyme(s), and/or enzyme mixtures are separated from the fermentation medium and then used. Low molecular solutes such as unconsumed components of the fermentation medium may be removed by ultrafiltration or any other suitable method. Any suitable method for separating cells, enzyme(s), and/or enzyme mixtures find use in the present invention. Indeed, it is not intended that the present invention be limited to any particular purification/separation method. In some additional embodiments, the fungal cells, enzyme(s) and/or enzyme mixtures are concentrated (*e.g.*, by evaporation, precipitation, sedimentation and/or filtration). In some embodiments, stabilizers are added to the compositions comprising fungal cells, enzyme(s), and/or enzyme mixtures. In some embodiments, chemicals such as glycerol, sucrose, sorbitol and the like find use to stabilize the enzyme mixtures. In some additional embodiments, other chemicals (*e.g.*, sodium benzoate and/or potassium sorbate), are added to the enzyme mixture to prevent growth of microbial contamination. In some additional embodiments, additional components are present in the compositions provided herein. It is not intended that the present invention be limited to any particular chemical and/or other components, as various components will find use in different settings. Indeed, it is contemplated that any suitable component will find use in the compositi~~osn~~ of the present invention.

Methods for Generating Fermentable Sugars

[00371] The present invention provides methods for generating fermentable sugars, including but not limited to glucose. In some embodiments, the methods for generating glucose comprise contacting cellulose with fungal cells producing at least one enzyme, at least one enzyme, and/or at least one enzyme mixture described herein. For example, in some embodiments, the process comprises contacting cellulose with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes, wherein at least one of the two or more cellulose hydrolyzing enzymes is produced by a fungal cell as described herein.

[00372] In some embodiments, the method for generating fermentable sugars such as glucose from cellulose using the enzyme mixture is batch hydrolysis, fed-batch hydrolysis, continuous hydrolysis, and/or a combination thereof. In some embodiments, the hydrolysis reaction is agitated, stirred, unmixed, and/or a combination thereof.

[00373] The methods for generating fermentable sugars such as glucose from cellulose are carried out at any suitable temperature known in the art. In some embodiments, a temperature of about 30°C to about 80°C, or any temperature therebetween (*e.g.*, a temperature of about 30°C, about 31°C, about 32°C, about 33°C, about 34°C, about 35°C, about 36°C, about 37°C, about 38°C, about 39°C, about 40°C, about 41°C, about 42°C, about 43°C, about 44°C, about 45°C, about 46°C, about 47°C, about 48°C, about 49°C, about 50°C, about 51°C, about 52°C, about 53°C, about 54°C, about 55°C, about 56°C, about 57°C, about 58°C, about 59°C, about 60°C, about 61°C, about 62°C, about 63°C, about 64°C, about 65°C, about 66°C, about 67°C, about 68 °C, about 69°C, about 70°C, about 71°C, about 72°C, about 73°C, about 74°C, about 75°C, about 76°C, about 77°C, about 78°C, about 79°C, about 80°C, or any temperature therebetween) find use.

[00374] In addition, any suitable pH finds use in the present invention. In some embodiments, a pH of about 3.0 to about 8.0, or any pH therebetween (*e.g.*, about pH 3.5 to about pH 6.8, or any pH therebetween, for example from about pH 3.0, about 3.1, about 3.2, about 3.3, about 3.4, about 3.5, about 3.6, about 3.7, about 3.8, about 3.9, about 4.0, about 4.1, about 4.2, about 4.3, about 4.4, about 4.5, about 4.6, about 4.7, about 4.8, about 4.9, about 5.0, about 5.1, about 5.2, about 5.3, about 5.4, about 5.5, about 5.6, about 5.7, about 5.8, about 5.9, about 6.0, about 6.1, about 6.2, about 6.3, about 6.4, 6.5, about 6.6, about 6.7, about 6.8, about 6.9, about 7.0, about 7.1, about 7.2, about 7.3, about 7.4, 7.5, about 7.6,

about 7.7, about 7.8, about 7.9, about 8.0, or any pH therebetween) finds use in the present invention,

[00375] In some embodiments, the initial concentration of cellulose in the hydrolysis reactor, prior to the start of hydrolysis, is about 0% (w/w), to about 0.1% (w/w), to about 15% (w/w), or any amount therebetween, for example about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 10%, about 11%, about 12%, about 13%, about 14%, or about 15% or any amount therebetween. However, it is not intended that the present invention be limited to any particular temperature, pH, time, nor cellulose concentration in the hydrolysis reaction, as any suitable temperature, pH, time, cellulose concentration, as well as other parameters find use in the present invention.

[00376] In some embodiments, the dosage of the cellulase enzyme and/or cellulase enzyme mixture used in the hydrolysis reaction is from about 0.1 to about 100 mg protein per gram cellulose, or any suitable amount therebetween (*e.g.*, about 0.1, about 0.2, about 0.3, about 0.4, about 0.5, about 0.6, about 0.7, about 0.8, about 0.9, about 1, about 2, about 3, about 4, about 5, about 6, about 7, about 8, about 9, about 10, about 11, about 12, about 13, about 14, about 15, about 16, about 17, about 18, about 19, about 20, about 21, about 22, about 23, about 24, about 25, about 26, about 27, about 28, about 29, about 30, about 31, about 32, about 33, about 34, about 35, about 36, about 37, about 38, about 39, about 40, about 41, about 42, about 43, about 44, about 45, about 46, about 47, about 48, about 49, about 50, about 51, about 52, about 53, about 54, about 55, about 56, about 57, about 58, about 59, 60, about 61, about 62, about 63, about 64, about 65, about 66, about 67, about 68, about 69, about 70, about 71, about 72, about 73, about 74, about 75, about 76, about 77, about 78, about 79, about 80, about 81, about 82, about 83, about 84, about 85, about 86, about 87, about 88, about 89%, about 90, about 91, about 100 mg protein per gram cellulose, or any amount therebetween. The hydrolysis is carried out for any suitable time period. In some embodiments, the hydrolysis is performed from about 0.5 hours to about 300 hours, from about 15 hours to 100 hours, or any time therebetween. In some embodiments, the hydrolysis reaction is performed for about 0.5, about 1, about 2, 3, about 4, about 5, about 6, about 7, about 8, about 9, about 10, about 11, about 12, about 13, about 14, about 15, about 16, about 17, about 18, about 19, about 20, about 25, about 30, about 35, about 40, about 45, about 50, about 55, about 60, about 65, about 70, about 75, about 80, about 85, about 90,

about 95, about 100, about 105, about 110, about 115, about 120, about 125, about 130, about 135, about 140, about 145, about 150, about 155, about 160, about 165, about 170, about 175, about 180, about 185, about 190, about 195, about 200, about 205, about 210, about 215, about 220, about 225, about 230, about 235, about 240, about 245, about 250, about 255, about 260, about 265, about 270, about 275, about 280, about 285, about 290, about 295, about 300 hours, or any time therebetween. It should be appreciated that the reaction conditions are not meant to limit the invention in any manner and may be adjusted as desired by those of skill in the art. Indeed, it is not intended that the present invention be limited to any particular hydrolysis reaction time, protein concentration, or any other specific reaction parameter, as various reaction parameters and components find use in the present invention.

[00377] In some embodiments, the enzymatic hydrolysis is carried out in a hydrolysis reactor. In some embodiments, the enzyme and/or enzyme mixture is added to the pretreated lignocellulosic feedstock (also referred to as the “substrate”) prior to, during, or after the addition of the substrate to the hydrolysis reactor.

[00378] In some embodiments, various environmental conditions are adjusted according to any variety of methods known in the art in order to maximize the formation of a hydrolysis product such as glucose. For example, temperature, pH, % dissolved oxygen, stirring speed can each be independently adjusted. In some embodiments, the enzyme mixture is a cell-free mixture, as described herein.

[00379] In some embodiments, the methods for generating glucose utilizing enzymes and/or enzyme mixtures comprising reduced glucose oxidase and/or reduced cellobiose dehydrogenase activity, as described herein, provide higher yields of glucose from the enzymatically hydrolyzed cellulose than a corresponding method using an enzyme mixture with its full complement of glucose oxidase or cellobiose dehydrogenase activity. Further, some embodiments of the methods provided herein result in decreased conversion of the cellobiose and glucose products in the enzymatic hydrolyzate to oxidized products such as gluconolactone, gluconate, gluconic acid cellobionolactone, and/or cellobionic acid.

[00380] In some embodiments of the methods provided herein utilizing the genetically modified fungal cell(s), enzyme(s), and/or enzyme mixture(s) provided herein, improved glucose yield is measured and/or quantified. As described herein, glucose yield can be

described in terms of the amount of generated glucose per theoretical maximum glucose yield, or in terms of Gmax.

[00381] For example, as described in U.S. Patent No. 6,090,595 and 7,419,809, the cellulose content can be determined by acid hydrolysis of the cellulose, followed by determination of glucose concentration, taking into account the water necessary to hydrolyze the cellulose. In one specific example, a slurry of feedstock is centrifuged, washed with water, and suspended in sulfuric acid at a net sulfuric acid concentration of 70%. The slurry is incubated at 40°C for 30 minutes, followed by diluting in deionized water to 2% sulfuric acid. At this time point, the samples are steam-autoclaved at 121°C for 1 hour, to convert the oligomers to monomeric glucose. The glucose concentration is measured by HPLC or enzymatic assay as described below.

[00382] Alternatively, cellulose content can be analyzed by infrared spectroscopy as described in Example 1. For example, solids can be washed and placed on the detection crystal of an infrared spectrometer and their absorbance measured between 500-4000 cm⁻¹.

[00383] Glucose levels can be quantified by any of a variety of methods known in the art (*See e.g.*, U.S. Patent Nos. 6,090,595 and 7,419,809). For example, glucose concentrations can be determined using a coupled enzymatic assay based on glucose oxidase and horseradish peroxidase (*See e.g.*, Trinder, *Ann. Clin. Biochem.*, 6:24-27 [1969]). Additional methods of glucose quantification include chromatographic methods, for example by HPLC (*See e.g.*, U.S. Patent Nos. 6,090,595 and 7,419,809). Cellobiose levels can be measured by any number of HPLC methods known to those skilled in the art (*See e.g.*, Kotiranta *et al.*, *Appl. Biochem. Biotechnol.*, 81: 81-90 [1999]).

[00384] Similarly, decreased conversion of cellobiose and glucose products to oxidized products such as cellobionolactone and gluconolactone can be quantified using any suitable method known in the art. For example, products of glucose or cellobiose oxidation can be detected and quantified using infrared spectroscopy, or by chromatographic methodologies such as HPLC (*See e.g.*, Rakotomanga *et al.*, *J. Chromatog. B.* 4:277-284 [1991]; and Mansfield *et al.*, *App. Environ. Microbiol.*, 64:3804-3809 [1997]). Accordingly, total oxidation of glucose or cellobiose can be determined, for example, as a function of total oxidation products per theoretical maximum glucose yield, or as a function of Gmax.

[00385] The methods, fungal cells, enzymes, and enzyme mixtures provided herein include reduction or removal of glucose and/or cellobiose oxidizing enzyme activity from a cellulose hydrolyzing enzyme mixture, thereby improving the yield of fermentable sugars such as glucose, xylose, and/or cellobiose during hydrolysis of cellulose. Advantageously, the processes and enzyme mixtures provided herein result in an increased yield of glucose and/or cellobiose from the hydrolyzed cellulose and a decreased oxidation of the glucose and/or cellobiose to oxidized sugar products, such as gluconolactone, gluconate, gluconic acid, cellobionolactone, and/or cellobionic acid from the hydrolyzed cellulose, relative to an enzyme mixture with an unmodified amount of glucose and/or cellobiose oxidizing enzyme activity, or relative to a parental enzyme mixture.

[00386] In some embodiments, the methods provided herein comprise contacting a cellulose substrate with fungal cells producing at least one cellulose hydrolyzing enzyme, at least one cellulose hydrolyzing enzyme, and/or an enzyme mixture comprising two or more cellulose hydrolyzing enzymes. In some embodiments, the enzyme mixtures are characterized in that oxidation of cellobiose and/or glucose is reduced during hydrolysis of cellulose, as described in greater detail herein, relative to an enzyme mixture with an unmodified amount of glucose and/or cellobiose oxidizing enzyme activity, or relative to a parental enzyme mixture. In some embodiments, the processes provided herein comprise providing a cellulose substrate, typically as an aqueous slurry, and providing at least one enzyme mixture comprising at least two cellulose hydrolyzing enzymes, at least one cellulose hydrolyzing enzyme, and/or fungal cells producing at least one cellulose hydrolyzing enzyme. The cellulose-containing slurry is introduced into a reaction vessel such as a hydrolysis reactor, and the at least one enzyme mixture comprising at least two cellulose hydrolyzing enzymes, at least one cellulose hydrolyzing enzyme, and/or fungal cells producing at least one cellulose hydrolyzing enzyme is added to the vessel, in any order. After a period during which hydrolysis occurs, hydrolysis product in the form of fermentable sugars such as glucose and/or cellobiose is produced, and, if desired, is recovered.

[00387] In some embodiments, the cellulosic substrate is provided in an aqueous slurry and added to a reaction vessel. The concentration of cellulosic feedstock in the slurry depends on the material. In some embodiments, the concentration of cellulosic feedstock in the slurry is between about 1% to about 30% (w/w) undissolved solids, or any concentration

therebetween, for example, from about 5% to about 20%, or from about 10% to about 20% undissolved solids, or any amount therebetween. In some embodiments, the concentration of cellulosic feedstock in the slurry comprises at least, at least about, up to, or about 1, about 2, about 3, about 4, about 5, about 6, about 7, about 8, about 9, about 10, about 11, about 12, 13, 14, about 15, about 16, about 17, about 18, about 19, about 20, about 21, about 22, about 23, about 24, about 25, about 26, about 27, about 28, about 29, or about 30% undissolved solids (w/w).

[00388] Any suitable method known in the art for generating glucose from cellulose using fungal cells producing at least one cellulose-hydrolyzing enzyme, at least one cellulose-hydrolyzing enzyme, and/or at least one enzyme mixtures find use in the present invention, including but not limited to batch hydrolysis, fed-batch hydrolysis, or continuous hydrolysis, as well as any suitable combination thereof. In some embodiments of batch processes, all the necessary materials are placed in a reactor at the start of the operation and the process is allowed to proceed until completion or until a desired endpoint, at which point the hydrolysis reaction is ended and in some embodiments, the product is harvested. In any batch process, one or more enzymes, the fungal cells producing at least one cellulose-hydrolyzing enzyme, and/or at least one enzyme mixture are added to the cellulose substrate before, during or after the introduction of the cellulose substrate to the reaction vessel. Thus, in some embodiments, the fungal cells, enzyme(s), and/or enzyme mixture(s) is added to the reaction vessel before introducing cellulosic substrate to the reaction vessel. In some other embodiments, the fungal cells, enzyme(s), and/or enzyme mixture(s) is added to the reaction vessel simultaneously with cellulosic substrate. In some other embodiments, the fungal cells, enzyme(s) and/or enzyme mixture(s) is added after introducing cellulosic substrate to the reaction vessel.

[00389] In some embodiments utilizing continuous process, cellulosic substrate is supplied and hydrolysis product is removed periodically or continuously at roughly volumetrically equal rates to maintain the hydrolysis reaction at a steady rate. Continuous processes can be performed according to any of a variety of methods known in the art, including but not limited to upflow hydrolysis processes (*See e.g.*, U.S. Pat. No. 7,727,746). However, it will be appreciated that any other suitable continuous process finds use in the present invention.

[00390] In some embodiments, following their withdrawal from the reactor, at least a portion of the unconverted solids are separated from the soluble hydrolysis liquor. Removal of the unconverted solids is accomplished using any suitable method, including but not limited to solids-liquid separation(*e.g.*, by use of a filter press, belt filter, drum filter, vacuum filter, and/or membrane filter), centrifugation, settling (*e.g.*, by use of a settling tank or an inclined settler for example, as disclosed in Knutsen and Davis, Appl. Biochem. Biotech., 98-100:1161-1172[2002]; and Mores *et al.*, Appl. Biochem. Biotech., 91-93:297-309 [2001]), clarification, or any other suitable process as would be known in the art. Clarification may be carried out using any suitable method known in the art. In some embodiments, a clarifier comprising a number of inclined plates to facilitate the separation of the solids and liquid or other features that are known in the art of solids-liquid separation find use. The soluble glucose, essentially free of undissolved solids, is then suitable for fermentation to ethanol. The unconverted solids are primarily lignin, which can be further utilized. For example, the unconverted solids can be burned and used as fuel or converted to generate electrical power.

[00391] In some embodiments, the unconverted solids comprise lignin, silica and/or other solid material. It is not intended that the present invention be limited to any particular unconverted solid(s). As the cellulose in the feedstock is hydrolyzed and released from the solid particles, the proportion of unconverted solids within the cellulose-containing solid particles increases. Depending on the density and particle size, the unconverted solids may be removed with the products at or settle to the bottom of the reaction vessel in a sediment or sludge. If a sludge layer forms at the bottom of the reactor due to very heavy particles, any means known in the art may be employed to remove the sludge or sediment. For example, in some embodiments, a scraper is used to remove the sludge. In some alternative embodiments, the bottom of the reactor is tapered to provide a path in which the heaviest solids settle, and then be removed and sent for processing, as desired.

[00392] In some embodiments, the fungal cells, enzyme(s), and/or enzyme mixture(s) are recovered and reused after the hydrolysis is completed or during the reaction. Recovery of the fungal cells, enzyme(s), and/or enzyme mixture(s) is accomplished using any suitable method known in the art. For example, in some embodiments, the fungal cells, enzyme(s), and/or enzyme mixture is removed from the hydrolysis liquor by precipitation (*e.g.*, pH

precipitation, salt precipitation, and/or temperature precipitation), extraction (*e.g.*, solvent extraction), and/or filtration (*e.g.*, ultrafiltration and/or microfiltration). It is not intended that the present invention be limited to any particular recovery method nor components. In some embodiments, the removed fungal cells, enzyme(s), and/or enzyme mixture(s) are added back to a hydrolysis reaction. In some embodiments utilizing ultrafiltration, the membrane has a molecular weight (MW) cut off of about 1,000, to about 20,000. In some embodiments, the MW cut off is from about 5,000 to about 10,000. In some embodiments, following recovery, the fungal cells, enzyme(s), and/or enzyme mixture(s) is recycled back into a reactor for further hydrolysis of additional feedstock. In some embodiments, the recycled enzyme(s) and/or enzyme mixture(s) are concentrated (*e.g.*, by evaporation, precipitation, sedimentation and/or filtration). In some embodiments, chemicals such as glycerol, sucrose, sorbitol and the like are added to stabilize the enzyme mixture. In some additional embodiments, other chemicals, such as sodium benzoate or potassium sorbate, are added to the enzyme(s) and/or enzyme mixture(s) to prevent growth of microbial contamination.

[00393] In embodiments, the methods are conducted in a reaction volume within a suitable vessel. Any suitable vessel finds use in the present invention, including but not limited to flasks, bioprocess reactors, hydrolysis reactors, and the like. As used herein, the term “hydrolysis tower,” “hydrolysis reactor,” bioprocess reactor,” “hydrolysis tank,” and the like refer to a reaction vessel of appropriate construction to accommodate the hydrolysis of cellulosic slurry by at least one cellulase enzyme. It should be appreciated that one or more hydrolysis reactors may be utilized, such as one or more batch or continuous stirred reactors. In some embodiments, in which more than one hydrolysis reactor is employed, the reactors are run in a series of two or more than two reactors, in which case the outlet of a first reactor feeds the inlet of a second reactor. Alternatively, in some embodiments, the reactors are run in parallel. Furthermore, in some embodiments, some of the reactors in the sequence are run in series, while others are run in parallel. Indeed, it is not intended that the present invention be limited to any particular reactor vessel, number of reactor vessels, nor any configuration of multiple reactors.

[00394] In some embodiments, the cellulose hydrolysis reaction volume is e greater than about 0.01 mL, about 0.1 mL, about 1 mL, about 10 mL, about 100 mL, about 1000 mL, or greater than about 5L, about 10 L, about 25 L, about 50 L, about 75L, about 100 L, about

150 L, about 200 L, about 250L, about 300 L, about 350L, about 400 L, about 450L, about 500 L, about 550L, about 600 L, about 650L, about 700 L, about 750L, about 800 L, about 850L, about 900 L, about 950L, about 1000 L, about 5000L, about 10,000 L, about 50,000 L, about 100,000 L, about 200,000L, about 250,000 L, about 500,000 L, about 750,000L, about 1,000,000L, or greater than about 1,000,000 L. Indeed, it is not intended that the present invention be limited to any particular reaction volume, as any suitable/desired reaction volume finds use in the present invention. In some embodiments, the hydrolysis reaction mixture is agitated, unmixed, or a combination thereof. For example, in some embodiments in which the hydrolysis reaction mixture is agitated, one or more impellers, agitators, eductors, and the like are used to mix the slurry. In some other embodiments, the hydrolysis reaction mixture is “unmixed,” in that no mixing (*i.e.*, no agitation) of the reactor contents takes place during the hydrolysis reaction. In some additional embodiments, other factors, such as the percentage of dissolved oxygen and/or stirring speed are monitored and independently adjusted as needed.

Cellulosic Material

[00395] The cellulosic material used in the present invention can be any material containing cellulose. The predominant polysaccharide in the primary cell wall of biomass is cellulose, the second most abundant is hemicellulose, and the third is pectin. The secondary cell wall, produced after the cell has stopped growing, also contains polysaccharides and is strengthened by polymeric lignin covalently cross-linked to hemicellulose. Cellulose is a homopolymer of anhydrocellobiose and thus a linear beta-(1-4)-D-glucan, while hemicelluloses include a variety of compounds, such as xylans, xyloglucans, arabinoxylans, and mannans in complex branched structures with a spectrum of substituents. Although generally polymorphous, cellulose is found in plant tissue primarily as an insoluble crystalline matrix of parallel glucan chains. Hemicelluloses usually hydrogen bond to cellulose, as well as to other hemicelluloses, which help stabilize the cell wall matrix.

[00396] Cellulose is generally found, for example, in the stems, leaves, hulls, husks, and cobs of plants or leaves, branches, and wood of trees. The cellulosic material can be, but is not limited to, herbaceous material, agricultural residue, forestry residue, municipal solid waste, waste paper, and pulp and paper mill residue (*See e.g.*, Wiseloge *et al.*, in Handbook

on Bioethanol, (Wyman, ed.), pp. 105-118, Taylor & Francis, Washington D.C. [1995]; Wyman, *Biores. Technol.*, 50: 3-16 [1994]; Lynd, *Appl. Biochem. Biotechnol.*, 24/25: 695-719 [1990]; Mosier *et al.*, in Advances in Biochemical Engineering/Biotechnology, (Scheper, ed.), Vol. 65, pp. 23-40, Springer-Verlag, New York [1999]). It is understood herein that the cellulose used in the present invention may be in the form of lignocellulose, a plant cell wall material containing lignin, cellulose, and/or hemicellulose in a mixed matrix. In some embodiments, the cellulosic material is lignocellulose.

[00397] In some embodiments, the pretreated lignocellulose used in the methods of the present invention is a material of plant origin that, prior to pretreatment, contains at least about 10%, about 20%, about 30%, or about 40% cellulose (dry weight). In some embodiments, the lignocellulose comprises about 20, about 21%, about 22%, about 23%, about 24%, about 25%, about 26%, about 27%, about 28%, about 29%, about 30%, about 31%, about 32%, about 33%, about 34%, about 35%, about 36%, about 37%, about 38%, about 39%, about 40%, about 41%, about 42%, about 43%, about 44%, about 45%, about 46%, about 47%, about 48%, about 49%, about 50%, about 51%, about 52%, about 53%, about 54%, about 55%, about 56%, about 57%, about 58%, about 59%, about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 85, about 90% cellulose (dry weight) or any percent therebetween, and at least about 10% lignin (dry wt) to about at least 12% (dry weight). However, it is not intended that the present invention be limited to lignocellulosic material comprising any particular percentage of cellulose and/or lignin. In some embodiments, the lignocellulose is subjected to physical and/or chemical processes to make the fiber more accessible and/or receptive to the actions of cellulolytic enzymes. In some embodiments, the lignocellulosic feedstock contains higher levels of cellulose following pre-treatment, while in other embodiments, the cellulose level is not altered during the pretreatment process. For example, if acid pretreatment is employed, the hemicellulose component is hydrolyzed, which increases the relative level of cellulose. In this case, in some embodiments, the pretreated feedstock contains greater than about 20% cellulose and greater than about 12% lignin.

[00398] Lignocellulosic feedstocks that find use in the present the present invention include, but are not limited to, agricultural residues such as corn stover, wheat straw, barley straw, rice straw, oat straw, canola straw, sugarcane straw and soybean stover; fiber process residues such as corn fiber, sugar beet pulp, pulp mill fines and rejects or sugar cane bagasse; forestry residues such as aspen wood, other hardwoods, softwood, and sawdust; or grasses such as switch grass, miscanthus, cord grass, and reed canary grass. In some embodiments, the lignocellulosic feedstock is first subjected to size reduction by any of a variety of methods including, but not limited to, milling, grinding, agitation, shredding, compression/expansion, or other types of mechanical action. Size reduction by mechanical action can be performed by any type of equipment adapted for the purpose, for example, but not limited to, a hammer mill.

Glucose and Cellobiose Oxidation

[00399] Use of the compositions and methods provided herein result in hydrolysis reactions having decreased oxidation of cellobiose and/or glucose during hydrolysis of cellulose, relative to an enzyme mixture with an unmodified amount of glucose and/or cellobiose oxidizing enzyme activity, or relative to a parental enzyme mixture. Therefore, in some embodiments, the enzyme(s) and/or enzyme mixtures used herein are characterized in that oxidation of cellobiose and/or glucose is reduced or eliminated.

[00400] In some embodiments, in some methods and enzyme mixtures of the present invention, the enzyme mixture is characterized in that, when the enzyme mixture is contacted with cellobiose and/or cellulose and/or glucose (*e.g.*, a cellobiose and/or cellulose and/or glucose substrate) no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellobiose and/or glucose is oxidized. For example, when the enzyme mixture is contacted with cellobiose or glucose no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellobiose and/or glucose is oxidized to form cellobionolactone, cellobionic acid, gluconolactone, gluconate or gluconic acid. For example, no more than about 1%,

about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of cellobiose and/or glucose is oxidized after about 1, about 5, about 10, about 15, about 20, about 25, about 30, about 35, about 40, about 45, about 50, about 55, or about 60 minutes, or after about 1.5, about 2, about 3, about 4, about 5, about 6, about 7, about 8, about 9, about 10, about 11, about 12, about 14, about 15, about 16, about 71, about 18, about 19, about 20, about 25, about 30, about 35, about 40, about 45, about 50, about 55, about 60, about 65, about 70, about 75, about 80, about 85, about 90, about 95, about 100, about 105, about 110, about 115, about 120, about 125, about 130, about 135, about 140, about 145, about 150, about 155, about 160, about 165, about 170, about 175, about 180, about 185, about 190, about 195, about 200, about 205, about 210, about 215, about 220, about 225, about 230 about 235, about 240, about 245, about 250, about 255, about 260, about 265, about 270, about 275, about 280, about 285, about 290, about 295, or about 300 hours or more. In some embodiments of the methods, the enzyme mixture is contacted with a cellulose and/or glucose substrate for a set period of time under reaction conditions at or about the optimal for enzymatic cellulose hydrolysis activity,

[00401] In some embodiments in which the cellulose hydrolysis reaction is performed in batch mode, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellobiose and/or glucose resulting from the hydrolysis of the cellulose substrate is oxidized after the termination of the batch mode cellulose hydrolysis reaction. In some embodiments in which the cellulose hydrolysis reaction is performed in continuous mode, no more than about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, or about 20% (wt %) of the cellobiose and/or glucose resulting from the hydrolysis of the cellulose substrate is oxidized at the time the cellulose hydrolysis reaction reaches steady state or quasi-steady state. In some embodiments,, the initiation of the reaction can be the initial about 1, about 5, about 10, about 15, about 20, about 25, about 30, about 35, about 40, about 45, about 50, about 55, or 60 minutes, or about 1.5, about 2, about 3, about 4, about 5, about 6, about 7, about 8, about 9, about 10, about 11,

about 12, about 13, about 14, about 15, about 16, about 17, about 18, about 19, about 20, about 25, about 30, about 35, about 40, about 45, about 50, about 55, about 60, about 65, about 70, about 75, about 80, about 85, about 90, about 95, or about 100 hours, after the cellulose substrate and cellulase enzymes are first admixed.

[00402] Decreased conversion of cellobiose and glucose products to oxidized products such as cellobionolactone and gluconolactone can be quantified by any of a variety of suitable methods known in the art. For example, products of glucose or cellobiose oxidation can be detected and quantified using infrared spectroscopy and/or chromatographic methods such as those as described in the Examples (*See also*, Rakotomanga *et al.*, J. Chromatog. B. 4:277-284 [1991]; and Mansfield *et al.*, App. Environ. Microbiol., 64:3804-3809 [1997]). Thus, in some embodiments, total oxidation of glucose and/or cellobiose are determined, for example, as a function of total oxidation products per theoretical maximum glucose yield, or as a function of Gmax, as described herein.

[00403] In some embodiments, the assessment of the glucose and/or cellobiose oxidizing activity in an enzyme mixture is carried out under similar conditions of pH and temperature to those employed for the process of hydrolyzing cellulose as described above. For example, in some embodiments, the assessment of the glucose and/or cellobiose oxidizing activity of the enzyme mixture is carried out at a pH of about 3.0 to about 8.0, or at a pH of about 5.0 to 6.0; and at a temperature of about 30°C to about 80°C, or at a temperature of about 50°C to about 60°C. The concentration of glucose and/or cellobiose in the assessment is typically in a range of glucose and/or cellobiose concentrations that would be expected to be generated during the process of hydrolyzing cellulose as described herein. For example, in some embodiments, the glucose and/or cellobiose concentrations are from about 1 g/L to about 500 g/L, or from about 10 g/L to about 200 g/L, or from about 30 g/L to about 100 g/L. In some embodiments, an enzyme mixture is mixed with a solution containing both about 50% w/w glucose and about 5% w/w cellobiose or a solution containing about 50% w/w glucose alone, at about pH 5.0 and about 60°C for about 24 hr, as set forth in the Examples, after which glucose and/or cellobiose oxidation products are quantified, for example by IR or by HPLC as described in the Examples. In some embodiments, an enzyme mixture is mixed with a solution containing about 100 g/L glucose at about pH 5.0 and about 55°C for about 24 hr.

[00404] Additionally, when comparing the glucose and/or cellobiose oxidizing activity in an enzyme mixture to a reference (*e.g.*, parental) enzyme mixture, the conditions of pH, temperature, and glucose/cellobiose concentration will depend upon such properties as the pH and temperature optima and stability, as well as the substrate affinity, of the particular glucose and/or cellobiose oxidizing enzymes that are present in the reference mixture and removed or inactivated in the enzyme mixture of interest. In some embodiments, the comparison is carried out at a pH and temperature range that is optimal for the reference enzyme mixture. In some other embodiments, the comparison is carried out at about pH and within temperature range that is optimal for cellulose hydrolysis reaction of the modified enzyme mixture. In some embodiments, the assessment of the glucose and/or cellobiose oxidizing activity of the enzyme mixture is carried out at a pH of about 3.0 to about 8.0, or at a pH of about 5.0 to about 6.0; and at a temperature of about 30°C to about 80°C, or about 50°C to about 60°C. Further, it will be appreciated that the concentration of cellobiose and/or glucose substrate in such a comparison will generally be within a range so as to readily detect oxidation products of the cellobiose and/or glucose substrate using the reference enzyme mixture. For example, in some embodiments, the concentration of cellobiose and/or glucose substrate is below a concentration that would cause substrate inhibition of the glucose and/or cellobiose oxidizing enzymes in the reference enzyme mixture. Thus, in some embodiments, the glucose and/or cellobiose concentrations generally range from about 1 g/L to about 300 g/L, or from about 10 g/L to about 100 g/L, or from about 30 g/L to about 70 g/L. In some embodiments, A analysis of glucose and/or cellobiose oxidizing activity in an enzyme mixture is carried out under similar conditions, including, pH, temperature and glucose and/or cellobiose concentrations, to those employed for the process of hydrolyzing cellulose. Further, it will be appreciated that identical conditions should be used to analyze the glucose and/or cellobiose oxidizing activity of both a modified enzyme mixture and of a reference enzyme mixture.

[00405] In some embodiments, conversion of cellobiose and glucose products to oxidized products such as cellobionolactone and gluconolactone is indirectly quantified (*e.g.*, by, measuring the total amount of glucose and cellobiose produced relative to the amount of cellulose consumed). In some cellulose hydrolysis reactions, the only significant by-products of the cellulose degradation reaction are oxidized products of cellobiose or glucose, or

transglycosylation products. The presence of transglycosylation products can be distinguished from the presence of oxidized products of cellobiose or glucose using a variety of methods known in the art or otherwise provided in the Examples herein. For example in some embodiments, only transglycosylation products, but not oxidized products, are acid-hydrolyzed to form only glucose. Thus, in some embodiments, the difference between the amount of cellulose consumed and the amount of cellobiose and/or glucose present is determined, and reflects the amount of oxidized products of cellobiose or glucose, or reflects the amount of oxidized products of cellobiose or glucose and the amount of transglycosylation products. Methods of quantifying cellulose and glucose are known in the art or are otherwise provided elsewhere herein.

Cellulose Conversion to Cellobiose and/or Glucose

[00406] In some embodiments, the methods for generating glucose, as described herein, using the enzyme mixture with reduced or removed glucose and/or cellobiose oxidizing enzyme activity, and the enzyme mixture itself, are characterized as providing a higher yield of cellobiose and/or glucose from the enzymatically hydrolyzed cellulose than a corresponding process using an enzyme mixture, or an enzyme mixture itself, with an unmodified amount of glucose and/or cellobiose oxidizing enzyme activity. In some embodiments, the methods for generating glucose provided herein using the enzyme mixture with reduced or removed glucose and/or cellobiose oxidizing enzyme activity, and the enzyme mixture itself, are characterized in providing a higher yield of cellobiose and/or glucose from the enzymatically hydrolyzed cellulose than a corresponding process using a reference enzyme mixture under essentially the same pH, temperature and other conditions including, but not limited to, feedstock concentration, mineral concentration, stir rate, percent oxygenation, and other conditions relevant to those skilled in the art for reproducing cellulose hydrolysis reactions.

[00407] In reference to an “unmodified amount of glucose and/or cellobiose oxidizing enzyme activity,” this phrase refers to an enzyme mixture obtained from a biological source organism in which the biological source organism has not been genetically modified or otherwise modified in such a way as to specifically target and thereby reduce the secretion, expression or activity of a glucose and/or cellobiose oxidizing enzyme, and/or the enzyme

mixture has not been manipulated in such a way as to thereby reduce the amount or activity of a glucose and/or cellobiose oxidizing enzyme. In reference to a reference enzyme mixture, this term refers to an enzyme mixture obtained from a reference biological source organism that is the same biological source organism as the biological source organism that provides the enzyme mixture with reduced or removed glucose and/or cellobiose oxidizing enzyme activity, where the biological source organism has not been genetically modified in such a way as to specifically target and thereby reduce the secretion, expression or activity of a glucose and/or cellobiose oxidizing enzyme, and the enzyme mixture has not been manipulated in such a way as to thereby reduce the amount or activity of a glucose and/or cellobiose oxidizing enzyme.

[00408] As used herein in reference to a percentage of cellulose hydrolyzed by the enzyme mixture present in the form of cellobiose and/or glucose, these percentages reflect a weight percent based on the dry weight of the hydrolyzed cellulose.

[00409] Thus, in some embodiments of the methods provided herein, when the fungal cells producing at least one enzyme, at least one enzyme, and/or at least one enzyme mixture is contacted with cellulose, at least about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% (wt %) of the cellulose hydrolyzed by the enzyme mixture is present in the form of cellobiose and/or glucose. In some embodiments, the fungal cells producing at least one enzyme, at least one enzyme, and/or at least one enzyme mixture is contacted with cellulose for a period of time of at least about 1, about 5 about 10, about 15, about 20, about 25, about 30, about 35, about 40, about 45, about 50, about 55, or about 60 minutes, or at least about 1.5, about 2, about 3, about 4, about 5, about 6, about 7, about 8, about 9, about 10, about 11, about 12, about 13, about 14, about 15, about 16, about 17, about 18, about 19, about 20, about 25, about 30, about 35, about 40, about 45, about 50, about 55, about 60, about 65, about 70, about 75, about 80, about 85, about 90, about 95, about 100, about 105, about 110, about 115, about 120, about 125, about 130, about 135, about 140, about 145, about 150, about 155, about 160, about 165, about 170, about 175, about 180, about 185, about 190, about 195, about 200, about 205, about 210, about 215, about 220, about 225, about 230, about 235, about 240, about 245, about 250, about 255, about 260, about 265,

about 270, about 275, about 280, about 285, about 290, about 295, or about 300 hours or more.

[00410] In some embodiments in which the cellulose hydrolysis reaction is performed in batch mode, at least about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, 9 about 8%, about 99%, or about 100% (wt %) of the cellulose hydrolyzed by the enzyme mixture is present in the form of cellobiose and/or glucose after the termination of the batch mode cellulose hydrolysis reaction. In some embodiments in which the cellulose hydrolysis reaction is performed in continuous mode, at least about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99% or about 100% (wt %) of the cellulose hydrolyzed by the enzyme mixture is present in the form of cellobiose and/or glucose at the time the cellulose hydrolysis reaction reaches steady state or quasi-steady state. In some embodiments, the initiation of the reaction can be the initial about 1, about 5, about 10, about 15, about 20, about 25, about 30, about 35, about 40, about 45, about 50, about 55, or about 60 minutes, or about 1.5, about 2, about 3, about 4, about 5, about 6, about 7, about 8, about 9, about 10, about 12, about 13, about 14, about 15, about 16, about 17, about 18, about 19, about 20, about 25, about 30, about 35, about 40, about 45, about 50, about 55, about 60, about 65, about 70, about 75, about 80, about 85, about 90, about 95, or about 100 hours, after the cellulose substrate and cellulase enzymes are first admixed.

Enzyme Mixtures with Reduced Glucose and/or Cellobiose Oxidizing Enzyme Activity

[00411] The present invention provides enzyme mixtures with reduced glucose and/or cellobiose oxidizing enzyme activity, which, when contacted with cellulose, result in a higher yield of glucose from the hydrolysis of cellulose than a corresponding method using an enzyme mixture with an unmodified amount of glucose and/or cellobiose oxidizing enzyme activity. In some embodiments, the enzyme mixture is characterized as providing a higher yield of cellobiose and/or glucose from the enzymatically hydrolyzed cellulose than a

corresponding process using a reference enzyme mixture. In some embodiments, the enzyme mixture is characterized as causing decreased conversion of the cellobiose and glucose products in the enzymatic hydrolysate to oxidized products such as cellobionolactone and gluconolactone relative to an enzyme mixture with an unmodified amount of glucose and/or cellobiose oxidizing enzyme activity, or relative to a reference enzyme mixture.

[00412] In some embodiments, an enzyme mixture with reduced glucose and/or cellobiose oxidizing enzyme activity is treated to reduce the amount of glucose and/or cellobiose oxidizing enzyme in the enzyme mixture. It will be readily appreciated that any of a variety of technologies known in the art can be employed to reduce the amount of glucose and/or cellobiose oxidizing enzyme from the enzyme mixture, including but not limited to purification processes that selectively remove one or more glucose and/or cellobiose oxidizing enzyme activity from the enzyme mixture. In some embodiments, inhibitors of glucose and/or cellobiose oxidizing enzyme activity are added to the enzyme mixture. Additional embodiments include the use of genetically modified fungal cells to reduce the amount of one or more endogenous glucose and/or cellobiose oxidizing enzymes secreted by the fungal cell.

[00413] Purification Processes. In some embodiments, the enzyme mixture is subjected to a purification process to selectively separate one or more glucose and/or cellobiose oxidizing enzymes from the enzyme mixture. In some embodiments, the purification process comprises removing the glucose and/or cellobiose oxidizing enzyme from the enzyme mixture using an affinity-based stationary phase. Affinity-based purification technologies are well known in the art, and include any method to selectively bind a component of a biological mixture to a solid support based on a highly specific biological interaction such as that between antigen and antibody or enzyme and substrate. Thus, affinity-based methodologies include contacting the enzyme mixture with beads or any other suitable solid support that comprises antibodies or other molecules that selectively bind to and immobilize the glucose and/or cellobiose oxidizing enzyme, while the remaining components of the enzyme mixture remain in solution. Some examples include chromatography methods either in batch form or column form. The solid support can comprise individual particles (*e.g.*, chromatography resin beads) or contiguous supports (*e.g.*, arrays). Ligands immobilized on a solid support matrix can then be employed to purify

targets from complex solutions. Conventional chromatography supports, as well as standard methods for grafting antibodies well known in the art and are described in numerous standard textbooks. These methods include the use of tubes, particles such as beads and any other suitable solid support. For example, particles are available in a large variety of different materials, including silica, glass, cellulose, agarose, and a wide variety of different polymers, including polystyrene polymethylmethacrylate, polyacrylamide, agarose, hydrogel, acrylic resins, and other types of gels used for electrophoresis. These supports may be purchased with ligands pre-attached or alternatively, the ligands can be indirectly attached or directly immobilized on the support using standard methods well known to those skilled in the art (*See e.g.*, Biancali *et al.*, *Lett. Peptide Sci.*, 7:297[2000]; MacBeath *et al.*, *Science*, 289:1760-1763 [2000]; Cass *et al.*, (eds.), *Proc. Thirteenth Am. Peptide Symp.*, . Leiden, Escom, 975-979 [1994]; U.S. Pat. No. 5,576,220; Cook *et al.*, *Tetrahed. Lett.*, 35:6777-6780 [1994]).

[00414] In some embodiments, the stationary phase comprises antibodies and/or any other molecule that selectively binds the glucose and/or cellobiose oxidizing enzyme, such as antibody fragments (*e.g.*, a Fab, Fab' or F(ab')₂). Strategies for depletion of specific proteins in a complex mixture of proteins are well known (*See e.g.*, Bjorhall *et al.* *Proteomics* 5:307-317 [2005]). In some embodiments, the stationary phase comprises antibodies directed toward glucose oxidase (EC 1.1.3.4), cellobiose dehydrogenase (EC 1.1.99.18), pyranose oxidase (EC 1.1.3.10), glucooligosaccharide oxidase (EC 1.1.99.B3), pyranose dehydrogenase (EC 1.1.99.29) or glucose dehydrogenase (EC 1.1.99.10).

[00415] In some embodiments, the stationary phase comprises molecules that selectively bind the glucose and/or cellobiose oxidizing enzyme(s). For example, in some embodiments, a binding protein, substrate, substrate analogue or other small molecule is coupled to the stationary phase to selectively bind the enzyme of interest. In some embodiments, the stationary phase comprises a glucose and/or cellobiose linked to the stationary phase. In some other embodiments, the stationary phase comprises a flavin adenine dinucleotide (FAD) linked to the stationary phase.

[00416] It will be appreciated that any of a variety of other purification methodologies are useful in the selective removal of one or more glucose and/or cellobiose oxidizing enzymes from the enzyme mixture. For example, in some embodiments, the purification

methodology comprises fractionation methods including selective precipitation such as ammonium sulfate precipitation, isoelectric precipitation, selective thermal denaturation, or any other method which selectively precipitates glucose and/or cellobiose oxidizing enzymes from the enzyme mixture, while leaving other components of the enzyme mixture in solution, or vice versa. In some other embodiments, the purification methodologies comprise chromatographic methods including gel filtration, size exclusion, anionic exchange, cationic exchange, gel electrophoresis, and/or other chromatic separation method known in the art for physically separating proteins.

[00417] Oxidase Inhibitors. In some embodiments, reducing the amount of glucose and/or cellobiose oxidizing enzyme activity from the enzyme mixture employs the addition of one or more glucose and/or cellobiose oxidizing enzyme inhibitor(s) to the enzyme mixture. Inhibitors of glucose and/or cellobiose oxidizing enzymes range from broad-spectrum oxidase inhibitors to specific inhibitors of glucose and/or cellobiose oxidizing enzymes, as described herein. .

[00418] In some embodiments, a broad-spectrum oxidase inhibitor is added to the enzyme mixture. Broad-spectrum oxidase inhibitors are well-known in the art. Some examples include but are not limited to mercuric chloride, silver sulphate, hydrazine compounds such as aminoguanidine, semicarbazide, benserazide, oxalic dihydrazide, hydralazine, phenylhydrazine, carbidopa, diaminoguanidine, and copper chelators such as desferrioxamine, EDTA, sodium azide, potassium cyanide, triene 5, o-phenanthroline, histidine and a number of metal anions such as Ag^+ , Hg^{2+} , and Zn^{2+} .

[00419] In some embodiments, a specific inhibitor of glucose and/or cellobiose oxidizing enzymes is added to the enzyme mixture. Specific inhibitors of cellobiose dehydrogenase include but are not limited to substrate analogues and other specific inhibitors such as cellobioimidazole, gentiobiose, lactobiono-1,5-lactone, celliobono-1,5-lactone, tri-N-acetylchitotriose, methyl-beta-D-cellobiosidase, 2,2-bipyridine, and/or cytochrome C. Specific inhibitors of glucose oxidase, pyranose oxidase, glucooligosaccharide oxidase, pyranose dehydrogenase, and glucose dehydrogenase include but are not limited to substrate analogues and other specific inhibitors.

[00420] Genetically Modified Fungal Cells. In some embodiments, the enzyme mixture is produced by a fungal cell that has been genetically modified to reduce the amount

of the endogenous glucose and/or cellobiose oxidizing enzyme activity that is secreted by the fungal cell.

[00421] Genetic modifications contemplated herein reduce the enzymatic activity of glucose and/or cellobiose oxidizing enzymes produced by a fungal cell. Any glucose and/or cellobiose oxidizing enzyme known in the art can be targeted for reduction of activity by genetic modification. For example, glucose and/or cellobiose oxidizing enzymes include glucose oxidase (EC 1.1.3.4), cellobiose dehydrogenase (EC 1.1.99.18), pyranose oxidase (EC 1.1.3.10), glucooligosaccharide oxidase (EC 1.1.99.B3), pyranose dehydrogenase (EC 1.1.99.29), and glucose dehydrogenase (EC 1.1.99.10). Each of these glucose and/or cellobiose oxidizing enzymes are described in the four Provisional applications to which the present application claims priority (*e.g.*, US Prov. Patent Appln. Ser. Nos. 61/409,186, 61/409,217, 61/409,472, and 61/409,480, all of which were filed on November 2, 2010).

Pretreatment.

[00422] In some embodiments, a substrate of the enzyme mixture comprises pretreated cellulosic material. Thus, for example, in processes described herein, any pretreatment process known in the art can be used to disrupt plant cell wall components of cellulosic material (*See e.g.*, Chandra *et al.*, Adv. Biochem. Engin./Biotechnol., 108: 67-93 [2007]; Galbe and Zacchi, Adv. Biochem. Engin./Biotechnol., 108: 41-65 [2007]; Hendriks and Zeeman, Biores. Technol., 100:10-18 [2009]; Mosier *et al.*, Biores. Technol., 96: 673-686 [2005]; Taherzadeh and Karimi, Int. J. Mol. Sci., 9:1621-1651 [2008]; and Yang and Wyman, Biofuels Bioprod. Bioref.-Biofpr. 2: 26-40 [2008]).

[00423] In some embodiments, the cellulosic material is subjected to particle size reduction, pre-soaking, wetting, washing, and/or conditioning prior to pretreatment, using any of a variety of methods known in the art.

[00424] In some embodiments, conventional pretreatments that find use in the present invention include, but are not limited to, steam pretreatment (with or without explosion), dilute acid pretreatment, hot water pretreatment, alkaline pretreatment, lime pretreatment, wet oxidation, wet explosion, ammonia fiber expansion, dilute ammonia pretreatment, organosolv pretreatment, and/or biological pretreatment. Additional pretreatments include,

but are not limited to ammonia percolation, ultrasound, electroporation, microwave, supercritical CO₂, supercritical H₂O, ozone, and gamma irradiation pretreatments.

[00425] In some embodiments, the cellulosic material is pretreated before hydrolysis and/or fermentation. In some embodiments, pretreatment is performed prior to the hydrolysis. In some alternative embodiments, the pretreatment is carried out simultaneously with enzyme hydrolysis to release fermentable sugars, such as glucose, xylose, and/or cellobiose. In some embodiments, the pretreatment step itself results in some conversion of biomass to fermentable sugars (even in absence of enzymes).

[00426] Steam Pretreatment. In steam pretreatment, cellulosic material is heated to disrupt the plant cell wall components, including lignin, hemicellulose, and cellulose to make the cellulose and other fractions (*e.g.*, hemicelluloses), accessible to enzymes. Cellulosic material is passed to or through a reaction vessel where steam is injected to increase the temperature to the required temperature and pressure and is retained therein for the desired reaction time. Steam pretreatment is preferably done at about 140°C to about 230°C, or from about 160°C to about 200°C, or about 170°C to about 190°C, where the optimal temperature range depends on any addition of a chemical catalyst. In some embodiments, residence time for the steam pretreatment is from about 1 to about 15 minutes, or about 3 to about 12 minutes, or about 4 to about 10 minutes, where the optimal residence time depends on temperature range and any addition of a chemical catalyst. Steam pretreatment allows for relatively high solids loadings, so that cellulosic material is generally only moist during the pretreatment. The steam pretreatment is often combined with an explosive discharge of the material after the pretreatment, which is known as steam explosion, that is, rapid flashing to atmospheric pressure and turbulent flow of the material to increase the accessible surface area by fragmentation (*See e.g.*, U.S. Patent No. 4,451,648; Duff and Murray, *Biores. Technol.*, 855: 1-33 [1996]; Galbe and Zacchi, *Appl. Microbiol. Biotechnol.*, 59:618-628 [2002]; and U.S. Pat. Appln. Pub. No. 2002/0164730). During steam pretreatment, hemicellulose acetyl groups are cleaved and the resulting acid autocatalyzes partial hydrolysis of the hemicellulose to monosaccharides and oligosaccharides. Lignin is removed to only a limited extent.

[00427] A catalyst such as H₂SO₄ or SO₂ (typically 0.3 to 3% w/w) is often added prior to steam pretreatment, which decreases the time and temperature, increases the

recovery, and improves enzymatic hydrolysis (*See e.g.*, Ballesteros *et al.*, Appl. Biochem. Biotechnol., 129-132: 496-508 [2006]; Varga *et al.*, Appl. Biochem. Biotechnol., 113-116:509-523 [2004]; Sassner *et al.*, Enzyme Microb. Technol., 39:756-762 [2006]).

[00428] Chemical Pretreatment. Examples of suitable chemical pretreatment processes include, but are not limited to, dilute acid pretreatment, dilute alkali pretreatment (*See e.g.*, U.S. Pat. Appln. Pub. Nos. 2007/0031918 and 2007/0037259), lime pretreatment, wet oxidation, ammonia fiber/freeze explosion or expansion (AFEX), ammonia percolation (APR), and organosolv pretreatments.

[00429] In dilute acid pretreatment, cellulosic material is mixed with dilute acid, typically H₂SO₄, and water to form a slurry, heated by steam to the desired temperature, and after a residence time flashed to atmospheric pressure. The dilute acid pretreatment can be performed with a number of reactor designs (*e.g.*, plug-flow reactors, counter-current reactors, or continuous counter-current shrinking bed reactors; *See e.g.*, Duff and Murray, Biores. Technol., 855: 1-33 [1996]; Schell *et al.*, Biores. Technol., 91:179-188 [2004]; and Lee *et al.*, Adv. Biochem. Eng. Biotechnol., 65: 93-115 [1999]).

[00430] Any suitable methods for pretreatment under alkaline conditions also find use. These alkaline pretreatments include, but are not limited to, lime pretreatment, wet oxidation, ammonia percolation (APR), ammonia fiber/freeze expansion (AFEX) and dilute ammonia pretreatment.

[00431] Lime pretreatment is performed with calcium carbonate, sodium hydroxide, or ammonia at low temperatures of 85-150°C and residence times from 1 hour to several days (*See e.g.*, Wyman *et al.*, Biores. Technol., 96:1959-1966 [2005]; Mosier *et al.*, Biores. Technol., 96:673-686 [2005]; WO 2006/110891; WO 2006/11899; WO 2006/11900; and WO 2006/110901).

[00432] Wet oxidation is a thermal pretreatment performed typically at 180-200°C for 5-15 minutes with addition of an oxidative agent such as hydrogen peroxide or over-pressure of oxygen (*See e.g.*, Schmidt and Thomsen, Biores. Technol., 64:139-151 [1998]; Palonen *et al.*, Appl. Biochem. Biotechnol., 117:1-17 [2004]; Varga *et al.*, Biotechnol. Bioeng., 88:567-574 [2004]; Martin *et al.*, J. Chem. Technol. Biotechnol., 81:1669-1677 [2006]). In some embodiments, the pretreatment is performed at about 1% to about 40% dry matter, or about

2% to about 30% dry matter, or about 5% to about 20% dry matter, and often the initial pH is increased by the addition of alkali such as sodium carbonate.

[00433] A modification of the wet oxidation pretreatment method, known as “wet explosion” (*i.e.*, the combination of wet oxidation and steam explosion), can handle dry matter up to 30%. In wet explosion, the oxidizing agent is introduced during pretreatment after a certain residence time. The pretreatment is then ended by flashing to atmospheric pressure (*See e.g.*, WO 2006/032282).

[00434] Ammonia fiber expansion (AFEX) involves treating cellulosic material with liquid or gaseous ammonia at moderate temperatures such as 90-100°C and high pressure such as 17-20 bar for 5-10 minutes, where the dry matter content can be as high as 60% (*See e.g.*, Gollapalli *et al.*, Appl. Biochem. Biotechnol., 98:23-35 [2002]; Chundawat *et al.*, Biotechnol. Bioeng., 96:219-231 [2007]; Alizadeh *et al.*, Appl. Biochem. Biotechnol., 121:1133-1141 [2005]; Teymouri *et al.*, Biores. Technol., 96:2014-2018 [2005]). AFEX pretreatment results in the depolymerization of cellulose and partial hydrolysis of hemicellulose. Lignin-carbohydrate complexes are cleaved. Dilute ammonia pretreatment utilizes more dilute solutions of ammonia than AFEX and may be conducted at a temperature of about 100-150°C, or any temperature therebetween (*See e.g.*, U.S. Pat. Appln. Pub. Nos. 2007/0031918 and 2007/0037259). The duration of the dilute ammonia pretreatment may be 1-20 minutes, or any duration therebetween.

[00435] Organosolv pretreatment delignifies cellulosic material by extraction using aqueous ethanol (40-60% ethanol) at 160-200°C for 30-60 minutes (*See e.g.*, Pan *et al.*, Biotechnol. Bioeng., 90:473-481 [2005]; Pan *et al.*, Biotechnol. Bioeng., 94:851-861 [2006]; and Kurabi *et al.*, Appl. Biochem. Biotechnol., 121:219-230 [2005]). Sulphuric acid is usually added as a catalyst. In organosolv pretreatment, the majority of hemicellulose is removed.

[00436] There are various other suitable methods for pretreatment that find use in the present invention (*See e.g.*, Schell *et al.*, Appl. Biochem. Biotechnol., 105-108:69-85 [2003]; and Mosier *et al.*, Biores. Technol., 96:673-686 [2005]; and U.S. Pat. Appln. Publ. No. 2002/0164730).

[00437] In some embodiments, the chemical pretreatment is carried out as an acid treatment. In some alternative embodiments, it is a continuous dilute and/or mild acid

treatment. In some embodiments, the acid is sulfuric acid, but other acids also find use, including but not limited to nitric acid, phosphoric acid, hydrogen chloride, and/or mixtures thereof. Mild acid treatment is conducted in the pH range of about 1 to about 5, or about 1 to about 4, or about 1 to about 3. In some embodiments, the acid concentration is in the range from about 0.01 to about 20 wt % acid, or about 0.05 to about 10 wt % acid, or about 0.1 to about 5 wt % acid, or about 0.2 to about 2.0 wt % acid. The acid is contacted with cellulosic material and held at a temperature in the range of about 160°C to about 220°C, or for about 165°C to about 195°C, for periods ranging from seconds to minutes (*e.g.*, about 1 second to about 60 minutes).

[00438] In some other embodiments, pretreatment is carried out as an ammonia fiber expansion step (AFEX pretreatment step).

[00439] In some embodiments, pretreatment takes place in an aqueous slurry. In some embodiments, cellulosic material is present during pretreatment in amounts between about 10 to about 80 wt %, or about 20 to about 70 wt %, or between about 30 to about 60 wt %, such as around 50 wt %. In some embodiments, the pretreated cellulosic material is unwashed, while in some other embodiments, it is washed using any method known in the art (*e.g.*, washed with water).

[00440] Mechanical Pretreatment. Any suitable methods of mechanical pretreatment find use in the present invention.

[00441] Physical Pretreatment. As used herein, the term "physical pretreatment" refers to any pretreatment that promotes the separation and/or release of cellulose, hemicellulose, and/or lignin from cellulosic material. For example, in some embodiments, physical pretreatment involves irradiation (*e.g.*, microwave irradiation), steaming/steam explosion, hydrothermolysis, and combinations thereof.

[00442] In some embodiments, physical pretreatment involves high pressure and/or high temperature (steam explosion). In some embodiments, "high pressure" means pressure in the range of about 300 to about 600 psi, or about 350 to about 550 psi, or about 400 to about 500 psi, such as around 450 psi. In some other embodiments, "high temperature" means temperatures in the range of about 100°C to about 300°C, or about 140°C to about 235°C. In some embodiments, mechanical pretreatment is performed in a batch-process, steam gun

hydrolyzer system that uses high pressure and high temperature as defined above (*e.g.*, a Sunds Hydrolyzer available from Sunds Defibrator AB, Sweden).

[00443] Combined Physical and Chemical Pretreatment. In some embodiments, cellulosic material is pretreated both physically and chemically. For instance, in some embodiments, the pretreatment step involves dilute or mild acid treatment and high temperature and/or pressure treatment. The physical and chemical pretreatments can be carried out sequentially or simultaneously, as desired. In some embodiments, a mechanical pretreatment is also included.

[00444] Accordingly, in some embodiments, cellulosic material is subjected to mechanical, chemical, and/or physical pretreatment, or any combination thereof, to promote the separation and/or release of cellulose, hemicellulose, and/or lignin.

[00445] Biological Pretreatment. In some embodiments, biological pretreatment processes find use in the present invention. Biological pretreatment techniques can involve applying lignin-solubilizing microorganisms (*See e.g.*, Hsu, "Pretreatment of Biomass," *in Handbook on Bioethanol: Production and Utilization* (Wyman, ed.), Taylor & Francis, Washington, D.C., pp. 179-212 [1996]; Ghosh and Singh, *Adv. Appl. Microbiol.*, 39:295-333 [1993]; McMillan, "Pretreating Lignocellulosic Biomass: a Review, *in Enzymatic Conversion of Biomass for Fuels Production* (Himmel *et al.* eds.), ACS Symposium Series 566, American Chemical Society, Washington, D.C., chapter 15 [1994]; Gong *et al.*, 65: 207-241 [1999]; Olsson and Hahn-Hagerdal, *Enz. Microb. Tech.*, 18:312-331 [1996]; and Vallander and Eriksson, *Adv. Biochem. Eng./Biotechnol.*, 42:63-95 [1990]).

[00446] In some embodiments, the soluble compounds derived from pretreatment process are subsequently separated from the solids. For example, in some embodiments, the separation step comprises one or more of standard mechanical means such as screening, sieving, centrifugation, and/or filtration to achieve the separation. In some other embodiments, the soluble compounds are not separated from the solids following pretreatment. It will be appreciated that pretreatment may be conducted as a batch, fed-batch or continuous process. It will also be appreciated that pretreatment may be conducted at low, medium or high solids consistency (*See e.g.*, WO 2010/022511).

Fermentation

[00447] In some embodiments, the methods for generating glucose provided herein further comprise fermentation of the resultant fermentable sugars (e.g., glucose) to an end product. Especially suitable fermenting organisms are able to ferment (i.e., convert), sugars, such as glucose, fructose, maltose, xylose, mannose and/or arabinose, directly or indirectly into at least one desired end product.

[00448] In some embodiments, yeast that find use in the present invention include, but are not limited to strains of the genus *Saccharomyces* (e.g., strains of *Saccharomyces cerevisiae* and *Saccharomyces uvarum*), strains of the genus *Pichia* (e.g., *Pichia stipitis*, such as *Pichia stipitis* CBS 5773 and *Pichia pastoris*), strains of the genus *Candida* (e.g., *Candida utilis*, *Candida arabinofementans*, *Candida diddensii*, *Candida sonorensis*, *Candida shehatae*, *Candida tropicalis*, and *Candida boidinii*). Other fermenting organisms include, but are not limited to strains of *Zymomonas*, *Hansenula* (e.g., *Hansenula polymorpha* and *Hansenula anomala*), *Kluyveromyces* (e.g., *Kluyveromyces fragilis*), and *Schizosaccharomyces* (e.g., *Schizosaccharomyces pombe*).

[00449] Suitable bacterial fermenting organisms include, but are not limited to strains of *Escherichia* (e.g., *Escherichia coli*), strains of *Zymomonas* (e.g., *Zymomonas mobilis*), strains of *Zymobacter* (e.g., *Zymobacter palmae*), strains of *Klebsiella* (e.g., *Klebsiella oxytoca*), strains of *Leuconostoc* (e.g., *Leuconostoc mesenteroides*), strains of *Clostridium* (e.g., *Clostridium butyricum*), strains of *Enterobacter* (e.g., *Enterobacter aerogenes*), and strains of *Thermoanaerobacter* (e.g., *Thermoanaerobacter* BG1L1; See, Appl. Microbiol, Biotech. 77: 61-86; *Thermoanaerobacter ethanolicus*, *Thermoanaerobacter thermosaccharolyticum*, and *Thermanaerobacter mathranii*). Strains of *Lactobacillus* also find use in the present invention, as well as strains of *Corynebacterium glutamicum* R, *Bacillus thermoglucosidasius*, and *Geobacillus thermoglucosidasius*. Indeed, it is not intended that the present invention be limited to any particular fermenting organism.

[00450] The fermentation conditions depend on the desired fermentation product and can easily be determined by one of ordinary skill in the art. In some embodiments involving ethanol fermentation by yeast, the fermentation occurs for between about 1 and about 120 hours, or between about 12 and about 96 hours. In some embodiments, the fermentation is carried out at a temperature between about 20 to about 40°C, or about 26 to about 34°C, or

about 32°C. In some embodiments, the fermentation pH is from pH about 3 to about 7, or about pH about 4 to about 6.

[00451] In some embodiments, enzymatic hydrolysis and fermentation are conducted in separate vessels so that each biological reaction can occur under its respective optimal conditions (*e.g.*, temperature). In some other embodiments, the process for producing glucose from cellulose described herein is conducted simultaneously with fermentation in a simultaneous saccharification and fermentation (SSF). SSF is typically carried out at temperatures of about 28°C to about 50°C, or about 30°C to about 40°C, or about 35°C to about 38°C, which is a compromise between the about 50°C optimum for most cellulase enzyme mixtures and the about 28°C to about 30°C optimum for growth of most yeast.

[00452] Accordingly, in some embodiments, the methods for generating glucose further comprise fermentation of the glucose to at least one end product. It is not intended that the present invention be limited to any particular end product, as the methods of the present invention are suitable to produce a variety of end products. In some embodiments, the end products include, but are not limited to fuel alcohols and/or precursor industrial chemicals. For example, in some embodiments, the fermentation products include precursor industrial chemicals such as alcohols (*e.g.*, ethanol, methanol, butanol); organic acids (*e.g.*, butyric acid, citric acid, acetic acid, itaconic acid, lactic acid, gluconic acid); ketones (*e.g.*, acetone); amino acids (*e.g.*, glutamic acid); gases (*e.g.*, H₂ and CO₂); antibiotics (*e.g.*, penicillin and tetracycline); enzymes; vitamins (*e.g.*, riboflavin, 812, beta-carotene); and/or hormones. In some embodiments, the end product is a fuel alcohol. Suitable fuel alcohols are known in the art and include, but are not limited to lower alcohols such as methanol, ethanol, butanol, and propyl alcohols.

Increased Expression of Saccharide Hydrolysis Enzymes

[00453] In some embodiments provided herein, the fungal cell is further genetically modified to increase its production of one or more saccharide hydrolysis enzymes. In some embodiments, the fungal cell overexpresses at least one homologous and/or heterologous gene encoding a saccharide hydrolysis enzyme (*e.g.*, beta-glucosidase). It is not intended that the present invention be limited to any particular enzyme(s), as numerous enzymes find use in the present invention. In some embodiments, the enzyme is any one of a variety of

endoglucanases, cellobiohydrolases, beta-glucosidases, endoxylanases, beta-xylosidases, arabinofuranosidases, alpha-glucuronidases, acetylxyylan esterases, feruloyl esterases, alpha-glucuronyl esterases, and/or any other enzyme involved in saccharide hydrolysis. In some embodiments, the fungal cell is genetically modified to increase expression of beta-glucosidase. Thus, in some embodiments, a fungal cell comprises a polynucleotide sequence for increased expression of beta-glucosidase-encoding polynucleotide and further can be genetically modified to delete polynucleotides encoding one or more endogenous glucose and/or cellobiose oxidizing enzymes.

[00454] In some embodiments, the saccharide hydrolysis enzyme is endogenous to the fungal cell. In some embodiments, the saccharide hydrolysis enzyme is exogenous to the fungal cell. In some other embodiments, the enzyme mixture further comprises a saccharide hydrolysis enzyme that is heterologous to the fungal cell. Still further, in some embodiments, the process for generating glucose comprises contacting cellulose with an enzyme mixture that comprises a saccharide hydrolysis enzyme that is heterologous to the fungal cell.

[00455] In some embodiments, the fungal cells of the present invention are genetically modified to increase the expression of a saccharide hydrolysis enzyme using any of a variety of methods that are known to those of skill in the art. In some embodiments, the hydrolysis enzyme-encoding polynucleotide sequence is adapted for increased expression in a host fungal cell.

EXPERIMENTAL

[00456] The following examples, including experiments and results achieved, are provided for illustrative purposes only and are not to be construed as limiting the present invention.

[00457] In the experimental disclosure below, the following abbreviations apply: ppm (parts per million); M (molar); mM (millimolar), uM and μ M (micromolar); nM (nanomolar); mol (moles); gm and g (gram); mg (milligrams); ug and μ g (micrograms); L and l (liter); ml and mL (milliliter); cm (centimeters); mm (millimeters); um and μ m (micrometers); sec. (seconds); min(s) (minute(s)); h(s) and hr(s) (hour(s)); U (units); MW (molecular weight); rpm (rotations per minute); °C (degrees Centigrade); DNA

(deoxyribonucleic acid); RNA (ribonucleic acid); HPLC (high pressure liquid chromatography); MES (2-N-morpholino ethanesulfonic acid); FIOPC (fold improvements over positive control); YPD (10g/L yeast extract, 20g/L peptone, and 20g/L dextrose); SOE-PCR (splicing by overlapping extension PCR); ARS (ARS Culture Collection or NRRL Culture Collection, Peoria, IL); Axygen (Axygen, Inc., Union City, CA); Lallemand (Lallemand Ethanol Technology, Milwaukee, WI); Dual Biosystems (Dual Biosystems AG, Schlieven, Switzerland); Alphalyse (Alphalyse Inc., Palo Alto, CA); Dyadic (Dyadic International, Inc., Jupiter, FL); Promega (Promega, Inc., Madison, WI); Megazyme (Megazyme International Ireland, Ltd., Wicklow, Ireland); McMaster (McMaster Regional Centre for Mass Spectrometry in Hamilton, Ontario, Canada); Sigma-Aldrich (Sigma-Aldrich, St. Louis, MO); Dasgip (Dasgip Biotools, LLC, Shrewsbury, MA); Difco (Difco Laboratories, BD Diagnostic Systems, Detroit, MI); PCRdiagnostics (PCRdiagnostics, by *E. coli* SRO, Slovak Republic); Agilent (Agilent Technologies, Inc., Santa Clara, CA); Molecular Devices (Molecular Devices, Sunnyvale, CA); Symbio (Symbio, Inc., Menlo Park, CA); Sartorius (Sartorius AG, Goettingen, Germany); Finnzymes (part of Thermo Fisher Scientific, Lafayette, CO); Dionex (now part of Thermo Fisher Scientific, Lafayette, CO); Idex (Idex Health and Science Group, Oak Harbor, WA); Microbeads (Microbeads A/S, Skedsmokorset, Norway); Calbiochem (Calbiochem-Novabiochem International, Inc., La Jolla, CA); Newport (Newport Scientific, Australia); and Bio-Rad (Bio-Rad Laboratories, Hercules, CA).

[00458] THE FOLLOWING POLYNUCLEOTIDE AND POLYPEPTIDE SEQUENCES FIND USE IN THE PRESENT INVENTION. AS SHOWN BELOW, THE POLYNUCLEOTIDE SEQUENCE IS FOLLOWED BY THE ENCODED POLYPEPTIDE.

M. thermophila GO1:

ATGGGCTTCCTCGCCGCCACTCTTGTGTCCTGTGCCGCTCTCGCGAGCGCAGCAA
GCATCCACGTCCCATGCCAAGCGCCAGGTCTCCAGCTTCGCGACGATTATGA
CTTCGTGATCGTTGGCGGTGGAAGTACGCGCCTCACTGTAGCCGATCGGCTGACA
GAGGCCTTTCAGCCAAGAACGTCCTTGTGTCATTGAGTATGGAGACGTCCACTACG
CCCCGGGAACCTTCGATCCGCCGACGGACTGGATCACACCTCAGCCTGATGCCC
CCCCTTCCTGGTCTTTCAATCCCTCCCCAACCCAGACATGGCAAACACAACAGC
GTTTGTGCTAGCCGGCCAAGTGGTGGGTGGAAGCAGTGCCGTGAACGGCATGTT
CTTTGACCGCGCATCCCGCCACGACTACGATGCGTGGACCGCGGTTCGGCGGGTC
CGGGTTCGAACAGTCCAGCCACAAGTGGGACTGGGAGGGGCTGTTCCCTTTCTTC

CAGAAGAGCGTCACGTTACGGAACCGCCGGCCGACATCGTCCAGAAGTATCAC
TACACCTGGGACCTGTCTGCCTACGGCAATGGCTCAACCCCCATCTACAGCAGCT
ATCCGGTCTTCCAGTGGGCGGACCAGCCGTTACTTAACCAGGCATGGCAGGAGA
TGGGAATCAATCCGGTGACCGAATGCGCCGGCGGCGACAAGGAGGGTGTCTGCT
GGGTTCCTCCGCTCGCAGCACCCCTGTACGGCGAGGAGGTCGCACGCCGGGCTCG
GCCACTACGCCGATGTGCTCCCGCGAGCCAATTACGACCTCCTCGTTCAACACCA
GGTTGTCAAGGTAGTATTCCCCAATGGGCCGAGCCACGGACCGCCGCTTGTCGA
GGCGCGGTCCCTGGCCGACAACCACCTGTTCAACGTGACTGTGAAGGGCGAAGT
CATCATCTCGGCGGGCGCTCTGCACACCCCGACCGTCCTTCAACGGAGCGGCATC
GGCCCGGCATCCTTCTTGACGACGCCGGGATCCCCGTGACGCTTGACCTGCCGG
GCGTCGGCGCCAACCTCCAGGACCACTGCGGTCCGCCCCGTACGTGGAATAACA
CCGAGCCCTACACCGGCTTCTTCCCGCTCCCCTCCGAGATGGTCAACAACGCGAC
CTTCAAAGCCGAAGCCATCACCGGCTTCGACGAGGTCCCGGCCCGCGGCCCTA
CACGCTCGCCGGGGGCAACAACGCCATCTTCGTATCGCTCCACACCTCACGGCC
GACTACGGCGCCATCACCGCAAATATCCGCGCCATGGTCGCCGACGGAACCGCC
GCCTCCTATCTCGCGGCCGACGTCCGCAACATCCCGGGGATGGTGGCCGGCTAC
GAGGCCAGCTCCTCGTGCTCGCCGACCTGCTCGACAACCCGGAGGCGCCAGC
CTGGAGACGCCGTGGGCGACGAGCGAGGCGCCGACGTCGTCGGTCTTGCC
TTCCTGCTGCACCCGCTCAGCCGCGGCAGCGTGCGGCTCAACCTCAGCGACCCGC
TCGCGCAGCCCGTGCTCGACTACCGCTCCGGGTCCAACCCGGTCGACATCGACCT
GCACCTCGCCACGTGCGCTTCTGCGCGGCCTGCTCGACACGCCACCATGCAG
GCCCCGCGGGGCGCTCGAGACGGCCCCCGGCTCGGCCGTGGCCGACAGCGACGAG
GCGCTGGGGGAGTACGTGCGCTCGCACAGCACGCTGTCCTTCATGCACCCGTGCT
GCACGGCCGCCATGCTGCCCCGAGGACCGGGGCGGCGTCGTCGGGCGGACCTCA
AGGTGCACGGGGCCGAGGGCCTGAGGGTCGTGGACATGAGCGTGATGCCGCTGT
TGCCGGGGGCGCACCTGAGCGCCACTGCTTATGCGGTGGGGGAGAAAGCTGCGG
ATATTATCATCCAGGAGTGGATGGACAAGGAGCAGTGA (SEQ ID NO:1)

MGFLAATLVSCAALASAASIPRPHAKRQVSQLRDDYDFVIVGGGTSGLTVADRLTE
AFPAKNVLVIEYGDVHYAPGTFDPPTDWITPQPDAPPSWSFNSLPNPDMAANTTAFVL
AGQVVGSSAVNGMFFDRASRDYDAWTA VGGSGFEQSSHKWDWEGLPFFQKS
VTFTEPPADIVQKYHYTWDLSAYGNGSTPIYSSYPVFQWADQPLLNQAWQEMGINP
VTECAGGDKEGVCWVPASQHPVTARRSHAGLGHYADVLPRANYDLLVQHQQVVRV
VFPNGPSHGPPLVEARSLADNHLFNVTVKGEVVISAGALHTPTVLQRSIGPASFLDD
AGIPVTLDLPGVGANLQDHCPPVTWNYTEPYTGFFPLPSEMVN NATFKAEAITGFD
EVPARGPYTLAGGNNAIFVSLPHLTADYGAITANIRAMVADGTAASYLAADVRTIPG
MVAGYEAQLLVADLLDNPEAPSLETPWATSEAPQTSSVLAFLHPLSRGSVRLNLS
DPLAQPVLDYRSGSNPVDIDLHLAHVRFLRGLLDTPTMQARGALETAPGSAVADSD
EALGEYVRSHSTLSFMHPCCTAAML PEDRGGVVGPD LKVHGAEGLRVVDM SVMPL
LPGAHL SATAYAVGEKAADIIIQEWMDKEQ (SEQ ID NO:2)

M. thermophila GO2:

ATGGAGCTGCTTCGAGTCTCCCTCGCCGCTGTTGCACTCTCCCCATTAATATTATT
CGGCGTTGCAGCCGCCCCACCTACCGCCCGATCCATTGCCCGCTCCACGATTCTT
GACGGAGCCGATGGCCTTCTTCCGGAGTATGACTACATCATCATCGGGGGCGGC

ACGTCCGGATTGACTGTCGCCGACAGACTCACGGAGAATAGAAAGCGCAAGTTT
TCCCGCTCTCCCCTCCCAACGTCACCCGCCCGATCGTCACCGGCGTGGTGTTATT
CTGTTCTTGTTTTGGAAAGAGGCATTTTCCAGAACTCTAGCTCGGTGACCACCAT
TTCTGGGGGAAGCAGAGGCCTCTTCGATCCAAGTCTGACCTTCAACATCAACTCC
GTTCCCAAGCTGGGCTGGACAACCGCAGCATTGCCGTCATTGGCGGGTTGATCC
TCGGCGGCAGCTCCGGCGTCAACGGGCTTCAAGTCCTCCGTGGACAAAGAGAAG
ACTATGACCGCTGGGGATCGTACTTTGGGGCCAACTCTGACTGGAGTTGGAAAG
GTCTCCTGCCGTATTTCAAGAAGGCATGGAATTTCCATCCGCCCAGGCCAGAGCT
GGTCAGTCAGTTCGACATCAAGTACGACCCAGCTACTGGGGCAACACGTCTGA
CGTGCACGCATCTTTCCCAACCACTTTCTGGCCGGTGCTCAAATTGGAGATGGCT
GCATTTGGTGACATCCCTGGGGTTCGAATATCCGCCCAGCTCTGCTTCTGGCGAGA
CCGGGGCGTATTGGCACCCAGCGTCCGTTGACCCAGCGACAGTCCTCCGCTCCTT
CGCTCGGCCCGCGCATTGGGACAACATTGAGGCGGCACGTCCCAATTACCACAC
CCTGACCGGGCAACGCGTATTGAAGGTCGCATTTGATGGCAATCGAGCGACCAG
CGTCGCTTTCGTGCCGGCGAATGCAACGGATCACAGCACTGCCAGGTCCGTGAA
GGCCAAGAAGGAGATCGTCTTGGCCGCCGGCGCCATTACACGCCCCAAATCCT
ACAGGCGAGCGGAGTAGGGCCGAAGCAGGTCCTGAAGGAAGCAGGCGTGCCGC
TTGTCTGTGACGCTCCCGGTGTCGGCAGCAATTTCCAAGACCAGCCGTATGTGGT
TGCTCCACCTTCAATTTTACCAAGTTCCCTTCCACCCGGACTTCTACGACATGA
TTCTGAACCAGACTTTTATCGCCGAGGCTCAGGCCAGTTTGAAAAGGACCGTAC
CGGACCTCACACCATCGCATCCGGCTATTGCGGCAGCTGGCTCCCCCTCCAGATC
ATTGCCCCAAATTCGTGGAAGGACATCGCTAGGCGGTACGAATCCCAAGACCCA
GCCGCCTACCTCCCCGCCGGCACCGATGAGACCGTCATCGAGGGGTACAGGGCG
CAGCAGAAAGCACTAGCGAGGTCCATGAGGAGCAAGCAATCGGCAATGTATAA
CTTCTTCTGAGGGGCGGCTACGAAGAGGGTTCTGTCGTCTACTTGCACCCAACC
AGCCGTGGCACCGTTTCGCATCAACCGATCCGACCCCTTCTTCTCGCCGCCCGAGG
TCGACTACAGGGCACTGAGCAACCCTACCGACCTGGAGGTCTGCTCGAATTCA
CTCCCTTACCCCGCAGGTACTTCTTGGAGACGAGGTTGAAGTCCCTCGACCCGGT
CGAGCTGTCGCCCCGGTGCCAACGTCACGGCGCCCGCCGACATCGAGGCCTGGCT
TCGCAGCGTCATGATCCCGTCCTCCTTCCATCCCATCGGCACGGCCGCCATGTTG
CCTAGGCACCTCGGTGGTGTCTGTGGACGAGAACCTTCTGGTGTACGGGGTCGAA
GGCTTGAGTGTCTGTCGACGCCAGCGTCATGCCCGACTTGCCGGGCTCATACACGC
AGCAGACCGTGTATGCTATTGCTGAGAAGGCCGCGGATCTCATTAAAGAGCAGG
CTTGA (SEQ ID NO:3)

MELLRVSLAAVALSPLILFGVAAAHPTARSIARSTILDGADGILLPEYDYIIIGGGTSGL
TVADRLTENRKRKFSRSPPTSPARSSPAWCYSVLVLERGIFQNSSSVTTISGGSRLGF
DPSLTFNINSVPQAGLDNRSIAVIGGLILGGSSGVNGLQVLRGQREDYDRWGSYFGP
NSDWSWKGLLPYFKKAWNFHPPPELVSQFDIKYDPSYWGNTSDVHASFPTTFWPV
LKLEMAAFGDIPGVEYPPDSASGETGAYWHPASVDPATVLRSFARPAHWDNIEAAR
PNYHTLTGQRVLKVAFDGNRATSVVFVPANATDHSTARSVKAKKEIVLAAGAIHTP
QILQASGVGPKQVLKEAGVPLVVDAPGVGSNFQDQPYVVAPTENFTKFPFHPDFYD
MILNQTFIAEAQAQFEKDRTGPHTIASGYCGSWLPLQIIAPNSWKDIARRYESQDPAA
YLPAGTDETVIEGYRAQQKALARSMRSKQSAMYNFFLRGGYEEGSVVYLHPTSRGT
VRINRSDPFFSPPEVDYRALSNPDTLEVLLEFTPFTRRYFLETRLKSLDPVELSPGANV

TAPADIEAWLRSVMIPSSFHPIGTAAMLPRHLGGVVDENLLVYGVEGLSVVDASVM
PDLPGSYTQQTQVY AIAEKAADLIKSRA (SEQ ID NO:4)

M. thermophila CDH1:

ATGAGGACCTCCTCTCGTTTAATCGGTGCCCTTGCGGCGGCACTCTTGCCGTCTG
CCCTTGCGCAGAACAAACGCGCCGGTAACCTTCACCGACCCGGACTCGGGCATTAC
CCTTCAACACGTGGGGTCTCGCCGAGGATTCTCCCCAGACTAAGGGCGGTTTCAC
TTTTGGTGTGCTCTGCCCTCTGATGCCCTCACGACAGACGCCAAGGAGTTCATC
GGTTACTTGAAATGCGCGAGGAACGATGAGAGCGGTTGGTGCGGTGTCTCCCTG
GGCGGCCCCATGACCAACTCGCTCCTCATCGCGGCCTGGCCCCACGAGGACACC
GTCTACACCTCTCTCCGCTTCGCCACCGGCTATGCCATGCCGGATGTCTACCAGG
GGGACGCCGAGATCACCCAGGTCTCCTCCTCTGTCAACTCGACGCACTTCAGCCT
CATCTTCAGGTGCGAGAACTGCCTGCAATGGAGTCAAAGCGGCGCCACCGGCGG
TGCCTCCACCTCGAACGGCGTGTTGGTCTCGGCTGGGTCCAGGCATTCGCCGAC
CCCGGCAACCCGACCTGCCCCGACCAGATCACCTCGAGCAGCACGACAACGGC
ATGGGTATCTGGGGTGCCCAGCTCAACTCCGACGCCGCCAGCCCGTCTACACC
GAGTGGGCGCGCCAGGCCACCAAGACCGTCACGGGTGACTGCGGCGGTCCCAAC
GAGACCTCTGTCTCGTGGTGTCCCCGTTCCGACGGGCGTCTCGTTCGATTACATCG
TCGTGGGCGGCGGTGCCGTGGCATCCCCGCCGCCGACAAGCTCAGCGAGGCCG
GCAAGAGTGTGCTGCTCATCGAGAAGGGCTTTGCCTCGACCGCCAACACCGGAG
GCACTCTCGGCCCCGAGTGGCTCGAGGGCCACGACCTTACCCGCTTTGACGTGCC
GGGTCTGTGCAACCAGATCTGGGTTGACTCCAAGGGGATCGCTTGCGAGGATAC
CGACCAGATGGCTGGCTGTGTCTCGGCGGCGGTACCGCCGTGAATGCCGGCCT
GTGGTTCAAGCCCTACTCGCTCGACTGGGACTACCTCTTCCCTAGTGGTTGGAAG
TACAAAGACGTCCAGCCGGCCATCAACCGCGCCCTCTCGCGCATCCCGGGCACC
GATGCTCCCTCGACCGACGGCAAGCGCTACTACCAACAGGGGCTTCGACGTCCTCT
CCAAGGGCCTGGCCGGCGGCGGCTGGACCTCGGTACAGGCCAATAACGCGCCAG
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GCAACGGCCCCGCTGGGCACCTACTTCCAGACCGCCAAGAAGCGCAGCAACTTCA
AGCTCTGGCTCAACACGTCTGGTCAAGCGCGTCAATCCGCCAGGGCGGCCACATCA
CCGGCGTTCGAGGTTCGAGCCGTTCCGCGACGGCGGTTACCAAGGCATCGTCCCCG
TCACCAAGGTTACGGGCGCGTTCATCTCTCTGCCGGTACCTTTGGCAGTGCAAA
GATCCTGCTGAGGAGCGGTATCGGTCCGAACGATCAGCTGCAGGTGTGCGCGC
CTCGGAGAAGGATGGCCCTACCATGATCAGCAACTCGTCCTGGATCAACCTGCC
TGTCGGCTACAACCTGGATGACCACCTCAACACCGACACTGTCATCTCCACCCCC
GACGTCGTGTTCTACGACTTCTACGAGGCGTGGGACAATCCCATCCAGTCTGACA
AGGACAGCTACCTCAACTCGCGCACGGGCATCCTCGCCCAAGCCGCTCCCAACA
TTGGGCCTATGTTCTGGGAAGAGATCAAGGGTGCGGACGGCATTGTTCCGCCAGC
TCCAGTGGACTGCCCCGTGTGAGGGCAGCCTGGGTGCCCCCAACGGCAAGACCA
TGACCATGTCGCAGTACCTCGGTCTGTTGGTGCACCTCGCGCGGCCGCATGACCAT
CACCCCGTCCCTGACAACTGTCGTCTCGGACGTGCCCTACCTCAAGGACCCCAAC
GACAAGGAGGCCGTATCCAGGGCATCATCAACCTGCAGAACGCCCTCAAGAAC
GTCGCCAACCTGACCTGGCTCTTCCCCAACTCGACCATCACGCCGCGCCAATACG
TTGACAGCATGGTCTGTCTCCCCGAGCAACCGGCGCTCCAACCACTGGATGGGCA
CCAACAAGATCGGCACCGACGACGGGCGCAAGGGCGGCTCCGCCGTCGTGAC

TCAACACCAAGGTCTACGGCACCGACAACCTCTTCGTCATCGACGCCTCCATCTT
 CCCC GGCGTGCCCCACCACCAACCCACCTCGTACATCGTGACGGCGTCGGAGCA
 CGCCTCGGCCCCGCATCCTCGCCCTGCCCCGACCTCACGCCCCGTCCCCAAGTACGGG
 CAGTGCGGCGGCCGCGAATGGAGCGGCAGCTTCGTCTGCGCCGACGGCTCCACG
 TGCCAGATGCAGAACGAGTGGTACTCGCAGTGCTTGTGA (SEQ ID NO:5)

MRTSSRLIGALAAALLPSALAQNNAPVTFTDPDSGITFNTWGLAEDSPQTKGGFTFG
 VALPSDALTTDAKEFIGYLKCARNDESGWCGVSLGGPMTNSLLIAAWPHEDTVYTS
 LRFATGYAMPDVYQGD AEITQVSSSVNSTHFSLIFRCENCLQWSQSGATGGASTSNG
 VLVLGWVQAFADPGNPTCPDQITLEQHDNGMGIWGAQLNSDAASPSYTEWAAQAT
 KTVTGDCCGPTETSVVGVVPVPTGVSFYIVVGGGAGGIPAADKLSEAGKSVLLIEKG
 FASTANTGGTLGPEWLEGHDLTRFDVPGLCNQIWVDSKGIACEDTDQMAGCVLGG
 GTAVNAGLWFKPYSLDWDYLFPSGWKYKDVQPAINRALSRIPTDAPSTDGKRYYY
 QQGFDVLSKGLAGGGWTSVTANNAPDKKNRTFSHAPFMFAGGERNGPLGTYFQTA
 KKRSNFKLWLNTSVKRVIRQGGHITGVEVEPF RDGGYQGIVPVTKVTGRVILSAGTF
 GSAKILLRSGIGPNDQLQVVAASEKDGPTMISNSSWINLPVGYNLDDHLNTDTVISHP
 DVVFYDFYEAWDNPIQSDKDSYLSNRTGILAQAAPNIGPMFWEEIKGADGIVRQLQ
 WTARVEGSLGAPNGKTMMSQYLGRGATSRGRMTITPSLTTVVSDVPYLKDPNDK
 EAVIQGIINLQNALKNVANLTWLFPNSTITPRQYVDSMVVSPSNRRSNHWMGTNKG
 TDDGRKGGS AVVDLNTKVYGTDLNFVIDASIFPGVPTTNPTSIVTASEHASARILAL
 PDLTPVPKYGQCGGREWSGSFVCADGSTCQMONEWYSQCL (SEQ ID NO:6)

M. thermophila CDH2:

ATGAAGCTACTCAGCCGCGTTGGGGCGACCGCCCTAGCGGCGACGTTGTCACTG
 CAGCAATGTGCAGCCCAGATGACCGAGGGGACCTACACCGATGAGGCTACCGGT
 ATCCAATTCAAGACGTGGACCGCCTCCGAGGGCGCCCCCTTTCACGTTTGGCTTGA
 CCCTCCCCGCGGACGCGCTGGAAAAGGATGCCACCGAGTACATTGGTCTCCTGC
 GTTGCCAAATCACCGATCCCGCCTCGCCCAGCTGGTGCGGTATCTCCCACGGCCA
 GTCCGGCCAGATGACGCAGGCGCTGCTGCTGGTGCCTGGGCCAGCGAGGACAC
 CGTCTACACGTCGTTCCGCTACGCCACCGGCTACACGCTCCCCGGCCTCTACACG
 GCGACGCCAAGCTGACCCAGATCTCCTCCTCGGTCAGCGAGGACAGCTTCGAG
 GTGCTGTTCCGCTGCGAAACTGCTTCTCCTGGGACCAGGATGGCACCAAGGGC
 AACGTCTCGACCAGCAACGGCAACCTGGTCCTCGGCCGCGCCGCCGCGCAAGGAT
 GGTGTGACGGGGCCCCACGTGCCCCGACACGGCCGAGTTCGGTTTCCATGATAAC
 GGTTTCGGACAGTGGGGTGCCGTGCTTGAGGGTGCTACTTCGGACTCGTACGAG
 GAGTGGGCTAAGCTGGCCACGACCACGCCCGAGACCACCTGCGATGGCACTGGC
 CCCGGCGACAAGGAGTGCGTTCCGGCTCCCGAGGACACGTATGATTACATCGTT
 GTCGGTGCCGGGCGCCGGTGGTATCACCGTCGCCGACAAGCTCAGCGAGGCCGGC
 CACAAGGTCCTTCTCATCGAGAAGGGACCCCTTCGACCGGCCTGTGGAACGGG
 ACCATGAAGCCCGAGTGGCTCGAGAGCACCGACCTTACCCGCTTCGACGTTCCC
 GGCCTGTGCAACCAGATCTGGGTCGACTCTGCCGGCATCGCCTGCACCGATAACC
 GACCAGATGGCGGGCTGCGTTCTCGGCGGTGGCACCGCTGTCAACGCTGGTTTGT
 GGTGGAAGCCCCACCCGCTGACTGGGATGAGAACTCCCCGAAGGGTGGAAGT
 CGAGCGATCTCGCGGATGCGACCGAGCGTGTCTTCAAGCGCATCCCCGGCACGT
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GCAAGGGCCTGGCCAACGCCGGCTGGAAGGAAATCAGCGCCAACGAGGCGCCC
 AGCGAGAAGAACCACACCTATGCACACACCGAGTTCATGTTCTCGGGCGGTGAG
 CGTGGCGGCCCCCTGGCGACGTACCTTGCCCTCGGCTGCCGAGCGCAGCAACTTC
 AACCTGTGGCTCAACACTGCCGTCCGGAGGGCCGTCCGCAGCGGCAGCAAGGTC
 ACCGGCGTCGAGCTCGAGTGCCTCACGGACGGTGGCTTCAGCGGGACCGTCAAC
 CTGAATGAGGGCGGTGGTGTTCATCTTCTCGGCCGGCGCTTTCGGCTCGGCCAAGC
 TGCTCCTTCGCAGCGGTATCGGTCTTGAGGACCAGCTCGAGATTGTGGCGAGCTC
 CAAGGACGGCGAGACCTTCACTCCCAAGGACGAGTGGATCAACCTCCCCGTCGG
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 GTTTTCTATGACTTCTATGCGGCCTGGGACGAGCCCATCACGGAGGATAAGGAG
 GCCTACCTGAACTCGCGGTCCGGCATTCTCGCCCAGGCGGCGCCCAATATCGGCC
 CTATGATGTGGGATCAAGTCACGCCGTCCGACGGCATCACCCGCCAGTTCCAGT
 GGACATGCCGTGTTGAGGGCGACAGCTCCAAGACCAACTCGACCCACGCCATGA
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 CCTCCGGGCTGAGCACGACGGTGGCCGAGCACCCGTACCTGCACAACAACGGCG
 ACCTGGAGGCGGTTCATCCAGGGGATCCAGAACGTGGTGGACGCGCTCAGCCAGG
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 TCAACAGCCTGATCGTCTCGCCGGCCAACCGCCGGGCCAACCCTGGATGGGCA
 CGGCCAAGCTGGGCACCGACGACGGCCGCTCGGGCGGCACCTCGGTCTGCGACC
 TCGACACCAAGGTGTACGGCACCGACAACCTGTTCTGTCGTCGACGCGTCCGTCTT
 CCCC GG CATGTGACGGGCAACCCGTGGCCATGATCGTCATCGTGGCCGAGCA
 GCGGCGCAGCGCATCCTGGCCCTGCGGTCTTAA (SEQ ID NO:7)

MKLLSRVGATALAATLSLQQCAAQMTEGTYTDEATGIQFKTWTASEGAPFTFGLTL
 PADALEKDATEYIGLLRCQITDPASPSWCGISHGQSGQMTQALLLVAWASEDTVYTS
 FRYATGYTLPLGLYTDAKLTQISSVSSEDSFEVLFRCECFSWDQDGTGKGNVSTSNG
 NLVLGRAAAKDGVTGPTCPDTAEFGFHDNGFGQWGAVLEGATSDSYEEWAKLATT
 TPETTCDDGTGPGDKECVPAPEPTYDYIVVGAGAGGITVADKLSEAGHKVLLIEKGPP
 STGLWNGTMKPEWLESTDLTRFDVPGLCNQIWVDSAGIACTDTDQMAGCVLGGGT
 AVNAGLWWKPHPADWDENFPEGWKSSDLADATERVFKRIPGTSHPSQDGKLYRQE
 GFEVISKGLANAGWKEISANEAPSEKNHTYAHTEFMFSGGERGGPLATYLASAAERS
 NFNLWLNTAVRRRAVRSGSKVTGVELECLTDGGFSGTVNLNEGGGVIFSAGAFGSAK
 LLLRSGIGPEDQLEIVASSKDGETFTPKDEWINLPVGHNLIDHLNTDLIITHPDVVFYD
 FYAAWDEPITEDKEAYLNSRSGILAQAPNIGPMMWDQVTPSDGITRQFQWTCRVE
 GDSSKTNSTHAMTSLQYLGRGVVSRGRMGITSLSTTVAEHPYLNNGDLEAVIQGI
 QNVVDALSQVADLEWVLPDPDGTVDYVNSLIVSPANRRANHWMGTAKLGTDDG
 RSGGTSVVDLDTKVYGTDLNFVVDASVFGPMSTGNPSAMIVIVAEQAAQRILALRS
 (SEQ ID NO:8)

Aspergillus oryzae pyranose oxidase:

ATGTCCATGACATCAGGACGTCAAGCGTTTACTTCCGAGTGCAGAGATTCAAATA
 CCACAAATTCATTTTGGTTGGCTAATTCACCGACTCTCACACTTGGCTCTACGAT
 GCAGGTCGTGGGGTCCGGCCCCATCGGCGCCACCTATGCCAAGATTCTAGCTGA
 CGCCGGCAAGGATGTCCTCATGGTTGAGACTGGCACCCAGGAAAGTAAGATTGC
 TGGAGAGCATAAGAAGAATGCTATCAACTACCAGAAAGATATCGATGCCTTTGT

GCATGTCATTAAGGTAATCAGCTCAAGAATTAGCACCTTTGAGTGTATTTCTCTA
 ACTTTTCGATCTTCTCCTCTTTTCAGGGAAGTCTACACTACACGTCTGTACCGACCA
 ACAAAGCCGCGCTTCTACACTGGCTCCGATCTCCTGGAAAGCGAACGGCCAAA
 TTTTCAACGGACAGAATCCCCGCCAGGATCCAAACGTAAACCTGGATGCCAATG
 GTGTGGCACGTAATGTGGGCGGCATGTCTACCCACTGGACTTGTGCGACTCCCCG
 ACAGAAAGAGAAGGTTGAACGCAGCGATATATTCAGTGGTGACGAATGGGATA
 GCCTGTACAAGGAGGCAGAAAAGTTGATCGGAACCAGCAAGACTGTGCTGAATG
 ACTCGATCCGGCAAGAATTGGTCATGGAGATTCTGAATGACGAGTACGGGAAGC
 GATCAGCCGAACCACTACCTTTGGCTGCAAAGAGGAATGGCAATACGGCCTACA
 TCACTTGGTCATCCTCGTCAACTATCCTTGACGCGATGAACTGTAAGAAGAAATT
 TACACTATGGCCCCGAGCACCCTGTGAGAAGTTTAAAGTCGAGGAAACAGATAA
 CGGGCCACAGGTACCAAGGCTATAATCCGCAAACCTGCCACAGATAAACTGAT
 TACAGTTAAGGCGAAAGTATTTATCGCTTGCAGGGGGGCCTATACTTACACCCAG
 CTACTTTTCAATTCGGGCTTCGTGCCGACAAAGCCCAACAGGGATCCCAGAACCC
 AAATACCATTAGAAGACGACGAGAAAGGCATCCCACCTCCACCGGATACTCTGG
 AGCATCTCAAGCTTCCTGCTCTAGGACGCTATCTGACAGAGCAAAGCATGTGCTT
 CTGCCAAATTGTTCTGAAAAAAGAATGGATTGAGGCAGTGGCTAATCCAAAAA
 GAACCCTTATCAAAGCGATGGGGTGAAACGCAAAAAGTGGGAGAAGCTCAAGG
 AAGGGTGGAAGGAAAGGGTCCAGGAACATATGAAAAGGTTTAATGACCCTATTC
 CCTTCCCGTTTCGATGATTTGGACCCTCAGGTTACTCTACCCTTGGACTATCACCAT
 CCGTGGCATAACCAAATCCATCGCGATGCCTTCTCCTATGGCGCAGCACCCCCAG
 CCATTGATAAGCGGACCATTGTTGACCTCCGATTCTTCGGAACGGTTGAGCCGGA
 CTGGAAGAACTATGTGACCTTTGAAACCGACATCAGGGATGCGTACGGCATGCC
 CCAGCCCACCTTCCGCTACAAGCTGAACGATGAGGATCGCAAACGGTCGCACCA
 GATGATGAAAGATATGGAAGAGGCCGCTGGTGTCTCTGGGTGGCTACCTCCCAGG
 GTCGGAGCCTCAATTTCTAGCTCCTGGCCTTGCACTGCACGTCTGTGGTACCCT
 AGAGCTCAGAAGAAGGAGAAAGAGTGTGACCCTGATCCCAAAGAGACCTCGTG
 CTGCGATGAGAACTCCAAGATCTGGGGTATCCACAACCTGTACGTGGGTGGGT
 AAATGTGATCCCTGGTGCCAATGGGTCCAACCCTACCTTGACAGCAATGTGCTTC
 GCCATCAAAAGCGCGAAGAGTATCCTTGAAGGGAATTCTTAG (SEQ ID NO:9)

MSMTSGRQFTSECRDSNTTNSFWLANSPTLTLGSTMQVVGSGPIGATYAKILADA
 GKDVLMVETGTQESKIAGEHKKNAINYQKDIDAFVHVIVISSRISTFECISLTFDILL
 FQGS LHYSVPTNKA AVPTLAPISWKANGQIFNGQNPRQDPNVNLDANGVARNVGG
 MSTHWTCATPRQKEKVERSDIFSGDEWDSLYKEAEKLIGTSKTVLNDLSIRQELVMEI
 LNDEYGKRSAPLPLAAKRNGNTAYITWSSSSTILDAMNCKKKFTLWPEHHCEKFK
 VEETDNGPQVTKAIIRKLATDKLITVKAKVFIACGGPILTPQLLFNSGFVPTKPNRDP
 TQIPLEDDEKGIPPPDTLEHLKLPALGRYLTEQSMCFQCIVLKKEWIEAVANPKKNP
 YQSDGVKRKKWEKLKEGWKERVQEHMKRFNDPIPFPPDLDLPQVTLPLDYHHPWH
 TQIHRDAFSYGAAPPAIDKRTIVDLRFFGTVEPDWKNYVTFETDIRDAYGMPQPTFR
 YKLNDEDRKRSHQMMKDMEEAAGALGGYLPGSEPQFLAPGLALHVC GTTRAQKK
 EKECDPDPKETSCCDENSKIWGIHNLVVGGLNVIPGANGSNPTLTAMCFAIKSAKSIL
 EGNS (SEQ ID NO:10)

Acremonium strictum glucooligosaccharide oxidase:

GCCTTCCAACAAGGACGTCTTCGAAACACGGGTCCCTGGCCTTTCTTCGGAACTC
 CGGCCACGTTTTGATTGGAATTATACAACGATTCCTCAAGATGCTCTCGGTGGCA
 GGAGCCTGAATTACTCGAGGGCGAAGCTCTTAGGCGGTTGCAGTAGCCATAATG
 GGATGGTTTACACACGATGTTTCGAGAGACGATTGGGACAATTATGCCGAAATCA
 CCGGTAATCAAGCATTTAGCTGGGACAGCATCCTACCTGTCATGAAGAGGGCTG
 AGAAATTCAGTAAAGATTCTCTCATAAACCGGTAAAGGGCCATATTGACCCCTC
 CGTGCACGGTGGTGACGGAAAATTGTCCGTGGTCGCATCATACACCAACGCCTC
 TTCAATGACTTATTACTTGAAACCGCGAAAGAATTAAGCGGTGAATTTCCGTTC
 AAATTGGATATGAATGACGGGGCGGCTCTTGGAATTAAGTGGACTCAGTATACG
 ATTGATCAACGCGGGGAGCGGAGCAGCTCTGCAACAGCGTATTTAGAGGGTACT
 GGAAATAACGTCCATGTCTTGGTTAACTCTTGTACCCGTATAGTCTCAGCAG
 AAAATGGGACCGACTTCCGAAGCGTCGAGTTTGCTACTGATGCCGACAGCCCAA
 AGATCCAATTACGAGCGAAAAAGGAAGTCATTGTATCTGGAGGAGTCATCAATT
 CGCCTCAGATCCTCATGAATTCCGGCATTGGGGGCCGAGAGGTGCTTGGAGCTA
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 AGGTGCGCTAAACCACCTTACATGGGTACGATTGCCTGATGACAAGCTGGATGG
 ACTTGATCCTTCAAGTGGCGAAAATTCGCCACATATTGAGTTCCAATTCGGGCAA
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 GTTCTCCAATTCGCGCGTTGATCAACCTCTACACTGTTTCGCGGGGTTCTATTTCT
 CTCAGTAACAACGATCCGTTCTCCACCCACTCATCGATCTCAACATGTTTGGAG
 AGGAAATAGATCCCGCTATTCTGCGTGAGGGTATTCGCAGTGCCCCGAAGAATGC
 TTTCTTCCCAAGCATTCAAAGGCTTTGTTCGGTGAAACGGTGTTCCTCCAAGCGA
 CGCTACCTCTGATGAAGATTTGGATACTTCTCCTCAAACGTCAACGTTTTCTTAC
 GTGCATGGTGTGGGAACGTTGTCTATGTCTCCTCAGAGTGCCTCGTGGGGTGTCTG
 TTAACCTGATTTCCGTGTCAAAGGAACCAGTGGCCTGCGGGTTGTCTGACGCGTC
 TGTGATTCCATTTCGCTCCGGCGGGGCACACTCAAGAACCTGTTTATGCATTTGCT
 GAGCATGCAAGTGTGTTAATAGCGAAGAGCTACAGCTAA (SEQ ID NO:13)

MIPRVAKFNFRLLSLALLGIQVARSAITYQNPTDLPDGDVDYDFIVAGGGTAGLVVAS
 RLSNPEWNVLVIEAGPSNKDVFETRVPGLSSELPRFDWNYTTIPQDALGGRSLNY
 SRAKLLGGCSSHNGMVYTRCSRDDWDNYAEITGNQAFSWDSILPVMKRAEKFSKD
 SSHKPVKGHIDPSVHGGDGKLSVVASYTNASFNDLLLETAKELSGEFPFKLDMNDG
 RPLGLTWTQYTIDQGERSSSATAYLEGTGNNVHVLVNTLVTRIVSAENGTDFRSVE
 FATDADSPKIQLRKKEVIVSGGVINSPQILMNSGIGGREVLGANGIDTLVDNPSVGK
 NLSDQAATIIMLDTTLPITDYDVDAALIEWKKSHTGPLAQGGRLNHLTWVRLPDDKL
 DGLDPSSGENSPHIEFQFGQISHQLPPSGLTRFSFYRHCSPIPLINLYTVSRGSISLNN
 DPFSLPLIDLNMFGEEIDPAILREGIRSARRMLSSQAFKGFVGETVFPPSDATSDLDL
 TFLKTSTFSYVHGVGTLSMSPQSASWGVVNPDFRVKGTSGLRVVDASVIPFAPAGHT
 QEPVYAFAEHASVLIAKSYS (SEQ ID NO:14)

Talaromyces stipitatus (ATCC 10500) glucose dehydrogenase:

ATGCGACTTGGCTCTATCGGCGCAGGCCTCGCTCTCCTCGCTGCCCTCGCTGTCC
 TCGCTGCCCCACGTGCACGCCTTGGCACCGCGCACCCAGATTGCCGAGGAATACG

ATTTTGTCTGTCGTTGGCGGCGGCCAGGCTGGTCTCGTGATCGGAGCTCGTCTGTC
GGAGATTGCAAATTATACAGTTCTCGTGCTGGAGGCAGGGACGAATGGAGACGA
ATTTTCGAGAACGAATAGGCACGTACAACCTTTTATACTCCCGCATATTCCTACTAC
GAGTCACTATGGACGACACCAATGAATTGGGCATACTATACTGTGCCTCAATCCC
ATGCCGAGAATCGTCAAATTGAGTGGACCCGTGGTAAGGGGGCTGGGCGGAAGTT
CTGCGATCAACGGATTGTACCTGACTCGCCCCGGTAAAGAGGAGATCAATGCAT
GGAAAGACCTGCTAGGAGACATGGACGGGGCGGACAATTGGTCGTGGGATTTCGT
TCTATGCTGCAATGAAGAAGAGCGAGACTTTTACTCCCCCGTCAATGAGATTGC
TACAGAAGGGAAACATTACATGGGACCTTTCTACTCGTGGTATTCAGGGACCGATT
CAGGCAACGTATCCCGGCTATACCTTCCCCCAAGTCGGCGAATGGGTGCATGTCTC
TGGAAGCAATGGGCATTGCTAGTTCTAACGATATGTACGGTGGTGAGGTGTATG
GCGCCGAAGTCTCGACGTCGAGTATCAATCCACGAACTGGACACGCTCGTACA
GCCGGACGGGATATCTCGACCCGCTCGCAGACAACGGCAATTACGACGTTGTGG
CCGATGCGTTTGTACGCGCATTCTCTTTGATGCTTCTTCTCCGTCGAATAATCTG
ACAGCAAACGGCGTGCAGTATACTCTTGACAACGGCAAGACAACTGCACGGTC
AAGGTCAAGAAAGAGGTGATCTTATCAGCTGGGACGGTTGGCAGTCTGCGGTA
CTGCTCCACAGCGGTGTCGGTCCGAAAGATGTTCTTTCAGATGCTGGAGTTGAGC
TGGTGTCTGAACTTCCTGGTGTGGGTACACCTTCAGGATCATTTTAAACAACAC
CCTTTATCTCTCCTACATCGATTACGCCATCGCCTACATCAATTCCACGCTGATGT
ACGGCGATAATCTGGACGCACTACAGAAGAACATCACCCTCAAATCAACCAAT
TCGTGCTGAACACGACTTACGATGCTGGTGTGATTGCAGGATACAAAGCAATTGC
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TTCATGAATAGTGACTTAAACGGCGATATTGGTATCACTGCTGCTCTTCAACATC
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CATTGATCCGAATTATCTTGCTGTTTCTGCTGACTATGAAATCCTCCGCGACGGC
CTCAATCTAGCCCGCCAACTCGGCAACACACAACCCCTAAGCAGCTGTCTAATA
GCCGAAACAATCCCCGGTCCCAGCGTCAAAACCGACGACGACTGGCTTGAATGG
ATCCGCGAAGCGACGGGGACAGAGTTCCACCCTTCATCGTCCTGTGCGATGCTA
CCCCGAGAGCAAGGCGGAGTAGTCGATGCCAACCTGCGCGTCTACGGTCTTGCC
AATGTTTCGTGTTGCGGATGCCAGCGTTGTCCCGATTTTCATTGTCGACGCATCTTAT
GGCGTCGACGTATGGAGTCGCAGAACAGGCTAGTAATATCATTCGTGCGCACTA
CACGGATAGTAGGACTACAGGCACGAGTAGTTCCGATCCTGGCTCTGCGTCGTC
ACCGACAAGCAGTGCATTGGGCGCTGAAGGGACTACTGGGGCGATTAGTGCTCA
TACAGCGCCTTCTGGTGGTGTACGAAGCGTTTCTGCGGTATCCGCTTGGGTTGCT
GTTGTGTTGCTGCGCTGCAGCTGTTTCCATTTCCATTCCTTGCATTGA (SEQ ID NO:15)

MRLGSIGAGLALLAALAVLAAHVHALAPRTQIAEEYDFVVVGGGQAGLVIGARLSEI
ANYTVLVLEAGTNGDEFRRIGTYNFYTPAYSYYESLWTPMNBWAYYTVPQSHAE
NRQIEWTRGKGLGGSSAINGLYLTRPGKEEINAWKDLLGDMDGADNWSWDSFYAA
MKKSETFTPPSNEIATEGNITWDLSTRGIQGPIQATYPGYTFPQVGEWVMSLEAMGIA
SSNDMYGGEVYGAEVSTSSINPTNWTRSYRTGYLDPLADNGNYDVVADAFVTRIL
FDASSPSNNLTANGVQYTLDNKTNCTVKVKKEVILSAGTVGSPAVLLHSGVGPKD
VLSDAGVELVSELPVGVHHLQDHFNNLYLSYIDSAIAYINSTLMYGDNLDAQKNI
TTQINQFVLNTTYDAGVIAGYKAIANMTATTILSSSIGQIELLFMNSDLNGDIGITAAL
QHPYSHGRIYINSSNPLDYPVIDPNYLAVSADYEILRDGLNLARQLGNTQPLSSCLIAE
TIPGPSVKTDDDWLEWIREATGTEFHPSSSCAMLPREQGGVVDANLRVYGLANVRV

ADASVVPISLSTHLMASTYGVAEQASNIIRAHYTDSRTTGTSSSDPGSASSPTSSALGA
EGTTGAISAHTAPSGGVRSVSAVSAWVAVVFAAAVSIFHSLH (SEQ ID NO:16)

EXAMPLE 1

FUNGAL STRAINS AND METHODS

[00459] As described below, variants of fungal strain C1 were prepared. In addition, the *Trichoderma reesei* cellulase enzyme mixture (“Turbo”) used in the following Examples is produced by a strain modified to produce and secrete high levels of the TrCel3A beta-glucosidase, encoded by *T. reesei bglI*, as described in U.S. Patent No. 6,015,703.

Strains and Nomenclature

[00460] Strain CF-400 ($\Delta cdh1$) is a derivative of C1 strain (“UV18#100f $\Delta alpl\Delta pyr5$ ”) further modified with a deletion of *cdh1*, wherein *cdh1* comprises the polynucleotide sequence of SEQ ID NO:5. Strain CF-401 ($\Delta cdh1\Delta cdh2$), is a derivative of the C1 strain further modified with a deletion of both a *cdh1* and a *cdh2*, wherein *cdh2* comprises the polynucleotide sequence of SEQ ID NO:7. Strain CF-402 (+*BglI*) is a derivative of the C1 strain further modified for overexpression of an endogenous beta-glucosidase 1 enzyme (*BglI*). Strain CF-403 is a derivative of the C1 strain modified with a deletion of *cdh1* and further modified to overexpress *bglI*. Strain CF-404 is a derivative of the C1 strain further modified to overexpress *bglI* with a deletion of both *cdh1* and *cdh2*.

[00461] Cellulolytic enzymes from strain CF-400, CF-401, CF-402, CF-403 and CF-404, were produced by submerged liquid culture fermentation using methods well-known in the art.

[00462] The *T. reesei* Turbo cellulose was produced by submerged liquid culture fermentation using methods described in U.S. Pat. Appln. Publ. No. 2010/0304438. The filtered fermentation broth was desalted using Biospin columns (Biorad) following the manufacturer’s protocol. Total protein concentration of the desalted enzyme was assayed using a BCA kit (Sigma) with a bovine serum albumin (Sigma) control.

Hydrolysis Reaction Conditions

[00463] Wheat straw (“WS”) was pretreated using the methods described in US Pat. 4,461,648. Following pretreatment, sodium benzoate was added at a concentration of 0.5% as a preservative. The pretreated material was then washed with six volumes of lukewarm (~35°C) tap water using a Buchner funnel and filter paper to produce the substrate for subsequent hydrolysis reactions (“pretreated WS”).

[00464] The cellulosic portion of the pretreated WS was hydrolyzed using the cellulolytic enzyme systems obtained as described above. Pretreated WS was hydrolyzed with 30 mg of cellulase per g of cellulose in reactions with 50 g/L cellulose at 50°C and pH 5.0, with 250 rpm orbital shaking, in total reaction volumes of 50 mL unless specified. For hydrolysis reactions containing the cellulase enzyme mixtures produced by C1 strains CF-400 and CF-401, beta-glucosidase purified from the Turbo or CF-402 cellulases was added at a dose of 125 IU per gm cellulose.

Detection of Glucose Yield in Hydrolysis Reaction

[00465] In this reaction, 1 mL aliquots of reaction mixture were sampled periodically from reaction flasks. Each reaction was well mixed during sampling to avoid removing a disproportionate amount of solid or supernatant. The reaction was stopped by incubating the aliquot in a 100°C hot block for at least 5 minutes. The supernatants of each reaction were analyzed for glucose concentration to determine the extent of conversion. The conversion calculation included correction terms for the effect of glucose on the density of the solution and the volume exclusion effect of non-hydrolyzable lignin present in the reaction. Glucose concentration was determined using a coupled enzymatic assay based on glucose oxidase and horseradish peroxidases using methods known in the art (*See e.g.*, Trinder, Ann. Clin. Biochem., 6:24–27 [1969]).

Detection of Cellulose Conversion

[00466] Residual solids from each of the 50 mL reactions were recovered, washed and analyzed by infrared spectroscopy. Aliquots of the samples were centrifuged at 2500 rpm for 5 min in an Eppendorf microfuge to sediment the solids, the supernatant was decanted, and the solids were resuspended back to the original volume in water. This procedure was repeated 5 times. Washed solids were placed on the detection crystal of a Golden Gate ATR

cell installed on a Bruker Vertex 70 infrared spectrometer and absorbance was measured between 500-4000 cm^{-1} .

EXAMPLE 2

PURIFICATION OF C1 CDH1

[00467] First, 400 mL of C1 supernatant was concentrated to 140 mL using a rotary evaporator. Then, 63 mL of the concentrate was buffer-exchanged into 20 mM MOPS buffer pH 7.0 using 4 in-line Hi-Prep 26/10 desalting columns (GE Healthcare, 17-5087-02). The resulting buffer-exchanged supernatant (~150 g/L total protein) was loaded onto a column containing 500 mL DEAE Fast Flow resin (GE Healthcare, 17-0709-01) pre-equilibrated with 20 mM pH7.0 MOPS buffer. The column was rinsed with 1 column volume (CV) of 20 mM MOPS (pH7.0) and then a 0-300 mM sodium chloride gradient was run over 12 column volumes. Fractions were collected and analyzed by NuPage® Novex® Bis-tris SDS-PAGE gels (Invitrogen, NP0322BOX). The SDS-PAGE bands corresponding to the apparent molecular weight of CDH1 were analyzed by MS (performed by Alphalyse). The mass-mapping analysis confirmed the presence of CDH1 in late-eluting fractions. Fractions containing CDH1, as demonstrated by SDS-PAGE gel, and confirmed by MS were pooled and concentrated by ultrafiltration using Sartorius centrifugal 10 kDa filter (Sartorius-Stedim, VS2002). Then, 10 mL 500 mM piperazine (pH 5.6) and 45 mL saturated ammonium sulfate were added to 45 mL of the CDH1-containing pool and the resulting mixture was loaded onto a Phenyl FF (high sub) 16/10 column (GE Healthcare, 28-9365-45) pre-equilibrated with 1.6 M ammonium sulfate in 50 mM piperazine, pH 5.6. A gradient of 1.6 M to 0 M ammonium sulfate in 50 mM piperazine, pH 5.6, was run over 30 CV. Fractions were collected and SDS-PAGE gel analysis was performed on the selected fractions as described above, revealing CDH1 eluted in the final rinse step with approximately 80-90% purity.

[00468] CDH1 activity was measured using a DCPIP (2,6-dichlorophenolindophenol) reduction assay similar to that described by Schou *et al.* (Schou *et al.*, Biochem J., 330:565-71 [1998]). Briefly, In a UV-transparent flat-bottom 96-well plate, 50 μL CDH1-containing fractions were added to 150 μL of a solution of 1.0 g/L cellobiose and 100 μM DCPIP in 100 mM sodium acetate, pH 5.0. Samples were agitated briefly at room temperature and then

absorbance at 530 nm (A_{530}) was measured for 10 minutes. C1 CDH1-containing fractions displayed a rapid drop in absorbance at 530 nm. DCPIP assays were performed using varying amounts of glucose or cellobiose with purified CDH1. Serial dilutions of cellobiose (1.0 g/L to 7.8 mg/L) and glucose (10 g/L to 78 mg/L) were prepared in a 96-well shallow-well plate. 20 μ L glucose and cellobiose standards were added to 160 μ L/well 200 mM DCPIP (in 100 mM pH 5.0 sodium acetate). Reactions were initiated by addition of 20 μ L CDH1 solution. Absorbance at 530 nm was monitored for 30 minutes. Comparisons of the rates of decrease in absorbance at 530 nm indicate that C1 CDH1 is approximately 10-fold more active on cellobiose than glucose.

EXAMPLE 3

MAKING OF *CDH1* SPLIT MARKER DELETION CONSTRUCTS

[00469] Genomic DNA was isolated from the C1 strain using standard procedures. Briefly, hyphal inoculum was seeded into a growth medium and allowed to grow for 72 hours at 35°C. The mycelial mat was collected by centrifugation, washed, and 50 μ L DNA extraction buffer (200 mM Tris, pH 8.0; 250 mM NaCl; 125 mM EDTA; 0.5% SDS) was added. The mycelia were ground with conical grinder, re-extracted with 250 μ L extraction buffer, and the suspension was centrifuged. The supernatant was transferred to a new tube containing 300 μ L isopropanol. DNA was collected by centrifugation, washed twice with 70% ethanol, and diluted in 100 μ L of water.

[00470] Genomic DNA fragments flanking the *cdh1* gene were cloned using primers cf09067 and cf09068 (*cdh1* upstream homology) and primers cf09069 and cf09070 (*cdh1* downstream homology). PCR reactions were performed by using the GoTaq® Polymerase (Promega) following the manufacturer's instructions using 0.2 μ M of each primer. Amplification conditions were 95°C 2 minutes, followed by 35 cycles of 95°C for 30 seconds, 55°C for 30 seconds (for upstream homology) or 53°C for 30 seconds (for downstream homology), and 72°C for 1 minute, and followed by final extension at 72°C for 5 minutes. The *pyr5* gene was PCR amplified as a split marker from a vector using primers cf09024 and cf09025 (for the 5' portion of the gene) and cf09026 and cf09027 (for the 3' portion of the gene). PCR reactions were performed using the GoTaq® Polymerase (Promega) following the manufacturer's instructions using 0.2 μ M of each primer.

Amplification conditions were 95°C for 2 minutes, followed by 35 cycles of 95°C for 30 seconds, 53°C for 30 seconds, and 72°C for 1 minute, followed by a final extension at 72°C for 5 minutes. Primers are shown in Table 3-1. In separate strand overlap extension reactions (Horton *et al.*, Meth. Enzymol., 217:270-279 [1993]), the PCR products resulting from primers cf09067 and cf09068 and primers cf09026 and cf09027 were fused as were the PCR products resulting from primers cf09069 and cf09070 and primers cf09024 and cf09025. PCR reactions were performed by using Finnzymes' Phusion® DNA Polymerases following the manufacturer's instructions including 3% DMSO and using 0.2 µM of each primer. Amplification conditions were 98°C for 1 minute, followed by 35 cycles of 98°C for 10 seconds, 62°C for 20 seconds, 72°C for 2 minutes, and followed by final extension at 72°C for 5 minutes. The strand overlap extension products were used for *cdh1* deletion.

Table 3-1. Primer Sequences	
Primer Name	Sequence (5'-3')
cf09067	CACGCGGGGTTCTTTCTCCATCTC (SEQ ID NO:17)
cf09068	TGAGGAAAACGCCGAGACTGAGCTCGACTCTGCCGGCCT ACCTACGA (SEQ ID NO:18)
cf09069	ATCAGTTGGGTGCACGAGTGGGTTTTGATGGGGAGTTGA GTTTGTGAA (SEQ ID NO:19)
cf09070	GGATGGATGAGGTTGTTTTTGAGC (SEQ ID NO:20)
cf09024	AACCCACTCGTGCACCCAACTGAT (SEQ ID NO:21)
cf09025	GACCACGATGCCGGCTACGATACC (SEQ ID NO:22)
cf09026	ACATGGCCCCACTCGCTTCTTACA (SEQ ID NO:23)

EXAMPLE 4

TRANSFORMATION METHOD

[00471] C1 cells and derivative strains were inoculated into 100 mL growth medium in a 500 mL Erlenmeyer flask using 10^6 spores/mL. The culture was incubated for 48 hours at 35°C, 250 rpm. To harvest the mycelia, the culture was filtered over a sterile Myracloth filter (Calbiochem) and washed with 100 mL 1700 mM NaCl/CaCl₂ solution (0.6 M NaCl, 0.27 M

CaCl₂*H₂O). The washed mycelia were transferred into a 50 mL tube and weighed. Caylase (20 mg/gram mycelia) was dissolved in 1700 mM NaCl/CaCl₂ and UV-sterilized for 90 sec. Then 3 mL of sterile Caylase solution was added into the tube containing washed mycelia and mixed. Then 15 mL of 1700 mM NaCl/CaCl₂ solution was added into the tube and mixed. The mycelium/Caylase suspension was incubated at 30°C, 70 rpm for 2 hours. Protoplasts were harvested by filtering through a sterile Myracloth filter into a sterile 50 mL tube. 25 mL cold STC (1.2 M sorbitol, 50 mM CaCl₂*H₂O, 35 mM NaCl, 10 mM Tris-HCl) was added to the flow through and spun down at 2720 rpm (1500xg) for 10 min at 4°C. The pellet was resuspended in 50 mL STC and centrifuged again. After the washing steps the pellet was resuspended in 1 mL STC.

[00472] Into the bottom of a 15 mL sterile tube, 2 µg DNA of each *pyr5::Acdh1* strand overlap extension product was pipetted and 1 µL aurintricarboxylic acid and 100 µL protoplasts were added. The content was mixed and the protoplasts with the DNA were incubated at room temperature for 25 min. 1.7 mL PEG4000 solution (60% PEG4000 (polyethylene glycol, average molecular weight 4000 daltons), 50 mM CaCl₂*H₂O, 35 mM NaCl, 10 mM Tris-HCl) was added and mixed thoroughly. The solution was kept at room temperature for 20 min. The tube was filled with STC, mixed and centrifuged at 2500 rpm (1250 xg) for 10 min at 4°C. The STC was poured off and the pellet was resuspended in the remaining STC and plated on minimal selective media plates, lacking uracil, but containing 0.67 M sucrose as an osmotic stabilizer. The plates were incubated for 5 days at 35°C. Colonies were restreaked and checked for the deletion of *cdh1*, designated as strain "CF-400."

EXAMPLE 5

CONFIRMATION OF CDH1 DELETION

[00473] Genomic DNA was prepared as described in Example 3. Primer pairs cf09112 and cf09113 (PCR reactions were performed by using the GoTaq® Polymerase (Promega) following the manufacturer's instructions using 0.2 µM of each primer. Amplification conditions were 95°C for 2 minutes, followed by 35 cycles of 95°C for 30 seconds, 54°C for 30 seconds, 72°C for 30 seconds, and followed by final extension at 72°C for 5 minutes) as

well as cf09110 and cf09111 (reactions were performed by using the GoTaq® Polymerase (Promega) following the manufacturer's instructions using 0.2 µM of each primer. Amplification conditions were 95°C for 2', followed by 35 cycles of 95°C for 30", 55.4°C for 30", 72°C for 30", and followed by final extension at 72°C for 5') were used in separate PCR reactions to confirm absence of the *cdh1* gene. Primers cf09181 and cf09091 were used in PCR to confirm proper junction structure and targeting of the pyr5 marker construct (See, Table 5-1). The PCR reaction was performed by using the GoTaq® Polymerase (Promega) following the manufacturer's instructions using 0.2 µM of each primer. Amplification conditions were 95°C for 2 minutes, followed by 35 cycles of 95°C for 30 seconds, 54.4°C for 30 seconds, 72°C for for 3 minutes 30 seconds, and followed by final extension at 72°C for 5 minutes. PCR products were run on agarose gel to confirm a banding pattern indicative of *cdh1* deletion.

Table 5-1. Primer Sequences

Primer Name	Sequence (5'-3')
cf09110	AAGCGTGCCGATTTTCCTGATTTC (SEQ ID NO:24)
cf09111	GCATTTCTGGGGCGGTTAGCA (SEQ ID NO:25)
cf09112	TCATCGACGCCTCCATCTTCC (SEQ ID NO:26)
cf09113	TTTCGGTTGTCGTGTTTCCATTAT (SEQ ID NO:27)
cf09181	GGAGATCCTGGAGGATTTCC (SEQ ID NO:28)
cf09091	CAGGCGGTGTGCGTTATCAAAA (SEQ ID NO:29)

[00474] A colorimetric dichlorophenolindophenol (DCPIP) assay was used to test for deletion of *cdh1* in CF-400. Deletion of *cdh1* was determined by decreased ability to reduce the DCPIP substrate compared to a cellulase enzyme mixture (or culture filtrate) produced by the parent strain. Cells of the parental C1 strain and putative *cdh1* delete strain were grown and supernatant tested for DCPIP activity. Combined in microtiter plates were 160 µL of freshly made DCPIP reagent solution (0.2 mM DCPIP in 100 mM sodium acetate, pH 5.0), 20 µL cellobiose solution (1 g/L cellobiose in deionized water), and 20 mLs of undiluted cell supernatant. The absorbance of the solution was immediately measured over time at 530 nm

in kinetic mode for 30 minutes to track loss of absorbance as a result of DCPIP reduction. Supernatant from strains displaying decreased ability to reduce the DCPIP substrate were run on SDS-PAGE to confirm the absence of CDH1.

[00475] Proteins from culture supernatants of submerged liquid culture fermentations of CF-400 and the untransformed parent were separated by SDS-PAGE using standard protocols. The proteins were visualized by staining with Simply Blue Safe Stain (Invitrogen), as per manufacturer's instructions. The Cdh1 protein was observed as a ~90 kD band in the untransformed parent but was absent in CF-400.

EXAMPLE 6

DELETION OF CDH2

[00476] Genomic DNA was isolated as described in Example 3. Genomic DNA fragments flanking the *cdh2* gene were cloned using primers cf10340 and cf10341 (*cdh2* upstream homology) and primers cf10342 and cf10343 (*cdh2* downstream homology). PCR reactions were performed by using the GoTaq® Polymerase (Promega) following the manufacturer's instructions using 0.2 µM of each primer. Amplification conditions were 95°C for 2 minutes, followed by 35 cycles of 95°C for 30 seconds, 58°C for 30 seconds (for upstream homology) or 58.4°C for 30 seconds (for downstream homology), 72°C for 1 minute, and followed by final extension at 72°C for 5 minutes. The hygromycin gene was PCR amplified as a split marker from a vector using primers cf10176 and cf10177 (for the 5' region of the gene) and cf10178 and cf10179 (for the 3' region of the gene). PCR reactions were performed by using the GoTaq® Polymerase (Promega) following the manufacturer's instructions using 0.2 µM of each primer. Amplification conditions were 95°C for 2 minutes, followed by 35 cycles of 95°C for 30 seconds, 56.3°C for 30 seconds, 72°C for 1 minute 30 seconds, and followed by final extension at 72°C for 5 minutes. Primers are shown in Table 5. In separate strand overlap extension reactions (Example 3) the PCR products resulting from primers cf10340 and cf10341 and primers cf10178 and cf10179 were fused as were the PCR products resulting from primers cf10342 and cf10343 and primers cf10176 and cf10177. PCR reactions were performed by using Phusion® DNA Polymerases (Finnzymes) following the manufacturer's instructions and including 3% DMSO and 0.2 µM of each primer. Amplification conditions were 98°C 1 minute, 35 cycles of 98°C 10 seconds, 62°C

20 seconds, 72°C 2 minutes and final extension at 72°C 5 minutes. The strand overlap extension products were used for *cdh2* deletion.

Table 5-1. Primer Sequences	
Primer Name	Sequence (5'-3')
cf10340	TTCAGCACGGCCGGGGATTTTATCCA (SEQ ID NO:30)
cf10341	GTAACACCCAATACGCCGGCCGAACATAAGAGCGGAGGTCAGGAATAA (SEQ ID NO:31)
cf10342	CCGTCTCTCCGCATGCCAGAAAGAGCTGTCAACGCTGGTTTGTGGTGG (SEQ ID NO:32)
cf10343	AATGCCGACCGCGAGTTCAGGTA (SEQ ID NO:33)
cf10176	TCTTTCTGGCATGCGGAGAGACGG (SEQ ID NO:34)
cf10177	TGTTGGCGACCTCGTATTGGGAAT (SEQ ID NO:35)
cf10178	TCTCGGAGGGCGAAGAATCTCGTG (SEQ ID NO:36)
cf10179	TTCGGCCGGCGTATTGGGTGTTAC (SEQ ID NO:37)

[00477] Strain CF-401 cells were grown and transformed as described in Example 4. Transformed colonies of CF-401 were restreaked and checked for deletion of *cdh2*.

[00478] Genomic DNA was prepared as described in Example 3. Deletion of *cdh2* in CF-401 was confirmed by PCR. Primers cf10326 and cf10327 were used in PCR to confirm absence of the *cdh2* gene. The PCR reaction was performed by using the GoTaq® Polymerase (Promega) following the manufacturer's instructions using 0.2 µM of each primer. Amplification conditions were 95°C for 2 minutes, followed by 35 cycles of 95°C for 30 seconds, 59.3°C for 30 seconds, 72°C for 30 seconds and followed by final extension at 72°C for 5 minutes. Primers cf10364 and cf10295 were used in PCR to confirm proper junction structure and targeting of the hygromycin marker construct (Table 6-2). The PCR reaction was performed by using the GoTaq® Polymerase (Promega) following the manufacturer's instructions using 0.2 µM of each primer. Amplification conditions were 95°C 2 minutes, 35 cycles of 95°C 30 seconds, 56°C 30 seconds, 72°C 3 minutes and final extension at 72°C 5 minutes. PCR products were run on agarose gel to confirm a banding pattern indicative of *cdh2* deletion. Dichlorophenolindophenol (DCPIP) assay and SDS-PAGE confirmation of *cdh2* deletion were performed as described in Example 5. Deletion of

cdh2 from strain CF-401 was determined by decreased ability of the resulting culture filtrate (or cellulase enzyme mixture) to reduce the DCPIP substrate compared to that produced by the parent strain CF-400.

Table 6-2. Primer Sequences	
Primer Name	Sequence (5'-3')
cf10326	GCGCTGGAAAAGGATGCCACCGAGT (SEQ ID NO:38)
cf10327	GCACCCCACTGTCCGAAACCGTTA (SEQ ID NO:39)
cf10295	AGCGCGTCTGCTGCTCCATACAAG (SEQ ID NO:40)
cf10364	CAAAGCCACGTCCAGGTTGATAGA (SEQ ID NO:41)

EXAMPLE 7

HYDROLYSIS OF PRETREATED WHEAT STRAW BY AN ENZYME MIXTURE LACKING CDH1

[00479] The ability of CF-402 and CF-403 enzyme mixtures to saccharify the cellulose present in pretreated wheat straw (WS) as measured by glucose and gluconate production was compared in a hydrolysis assay as described in Example 1. Each enzyme mixture was assessed using both a single enzyme dose of 30 mg enzyme/g cellulose and in parallel using a second 30 mg enzyme/g cellulose dose added after a 24 hr hydrolysis to the original 30 mg/g enzyme load. The preparation of pretreated WS used in these experiments had a maximum convertibility of 95%; conversions of all enzymes were normalized. Hydrolysis assay results are depicted in Fig. 1.

[00480] In the single dose experiment, the total soluble products measured, equaled a cellulose conversion of 79% and 87% of theoretical maximum glucose yield with enzyme mixtures from CF-402 and CF-403 respectively. About 5% of soluble products were measured to be gluconate for CF-403 as compared to 10% for CF-402. The results demonstrate that removing the CDH1 enzyme from the cellulase mixture produced by C1 derived strains improves glucose yield.

[00481] In the multidose experiments, the total soluble products measured with enzyme mixtures from CF-402 indicated a 90% cellulose conversion as compared to 95%

with enzyme mixtures from CF-403. The conversion of glucose to gluconate was higher for CF-402 (14%) compared to CF-403 (7%).

EXAMPLE 8

HYDROLYSIS OF PRETREATED WHEAT STRAW BY AN ENZYME MIXTURE LACKING BOTH CDH1 AND CDH2

[00482] The ability of enzyme mixtures to saccharify the cellulose present in pretreated WS was compared for enzyme mixtures obtained from CF-401, CF-400, and CF-402 in the following hydrolysis assay.

[00483] Enzyme mixtures derived from CF-400, CF-401 and CF-402 were produced by fermentation as described in Example 1. Enzyme mixtures derived from CF-400, CF-401 and CF-402 were added to the pretreated wheat straw at enzyme loads of 3.83- 6% w.r.t cellulose. The cellulose concentration was 110 g/L. *Aspergillus niger* beta-glucosidase (ANBG, Sigma) at an enzyme load of 2% w.r.t cellulose, was supplemented to CF-400 and CF-401 enzyme mixtures as these strains lack added beta-glucosidase activity. Glucose and gluconate yields were compared for the sample withdrawn at 48-70 hrs.

[00484] For glucose analysis, the samples were diluted 1:10 using 10 mM H₂SO₄ and then analyzed using an Agilent HPLC 1200 equipped with HPX-87H Ion exclusion column (300 mm x 7.8 mm) with 5 mM H₂SO₄ as a mobile phase at a flow rate of 0.6 mL/min at 65°C. The retention time of the glucose was 9.1 minutes. The gluconate analysis was carried out using LC-MS. The LC/MS/MS used was a API2000 triple quadrupole system (AB Sciex) equipped with Agilent 1100 HPLC, and CTC PAL autosampler. The column used was a HYPERCARB 50 X 2.1 mm, 3 µm at 80°C temperature. The chromatography method was a 2 minutes isocratic (95%A) run with a flow rate of 350 µL/min. The two mobile phases contained 1.5 % NH₄OH. The mobile phase A was aqueous, and the B phase was a 50:50 solution of CH₃CN: IPA. The MS/MS transition monitored for gluconate was 194.99/161.10. The analytical methods and controls for measuring gluconate and glucose were slightly different for CF-402. However, the methods and controls used are well known in the art.

[00485] The results are provided in Fig. 2 and in Table 7-1 below.

Table 7.1. Glucose Conversion			
Enzyme Mixture	Fractional Conversion of Available Cellulose		
	Glucose	Gluconate	Sum
<i>Bgl1</i> (CF-402)	0.800	0.167	0.967
$\Delta cdh1$ (CF-400)	0.861	0.114	0.974
$\Delta cdh1 \Delta cdh2$ (CF-401)	0.954	0.024	0.978

[00486] Results from the experiments show the total soluble products obtained from the enzyme mixtures derived from strains CF-400, CF-401 and CF-402 were similar. However the ratios of glucose:gluconate were different in the soluble products produced by the enzyme mixtures from the different strains. CF-402-derived enzyme mixtures produced about 23 g/L of gluconate. In comparison, CF-400-derived enzyme mixtures (the deletion of the *cdh1* gene) reduced the gluconate production to about 16 g/L. The enzyme mixture resulting from the additional deletion of the *cdh2* gene in CF-401 significantly reduced (3.3 g/L) the gluconate production. This reduction in gluconate production represented an 86% reduction compared to the enzyme mixtures derived from CF-402 strain. Correspondingly the glucose yields increased from 112 g/L with CF-402 to 133.6 g/L with CF-401-derived enzyme mixtures. The results demonstrate that the removal of CDH1 and CDH2 enzymes by deletion of both *cdh1* and *cdh2* genes significantly increases glucose yield with a concomitant decrease in gluconate production in the saccharification reaction.

EXAMPLE 9

ENZYMATIC HYDROLYSIS OF CELLULOSE

[00487] The cellulosic portion of pretreated WS was hydrolyzed according to the methods described in Example 1 using enzyme mixture derived from CF-402. In each reaction, an initial 30 mg/g dose of CF-402 was allowed to react with the pretreated WS substrate. After 24 hours, an additional dose of 30 mg/g of CF-402 enzyme mixture was added to one reaction flask.

[00488] Aliquots of 1 mL were sampled periodically from all these flasks, and glucose concentrations were determined using a coupled enzymatic assay as described in Example 1. By glucose yield, a single 30 mg/g dose of the CF-402 enzyme mixture produces 69.5% of theoretical maximum glucose yield (Gmax). Dosing additional CF-402 increased this yield to 75.6% as shown in Fig. 11.

EXAMPLE 10

CONVERSION OF GLUCOSE BY C1-DERIVED ENZYME MIXTURES

[00489] Fifty-two mg of the C1-derived CF-402 enzyme mixture or the *T. reesei* “Turbo” enzyme mixture was mixed with a solution containing both 50% w/w glucose and 5% w/w cellobiose or a solution containing 50% w/w glucose alone, at pH 5.0 and 60°C for 24 hr and 48 hr.

[00490] An unknown species (detected in HPLC chromatograms as described in more detail herein) was produced in a time-dependent manner in these reactions (Table 2). The peak areas of the unknown species are normalized to the peak area of the unknown species in the 24 hr reaction of the CF-402 enzyme mixture and glucose only. These data indicate that the insoluble cellulose substrate, primarily comprising cellulose and lignin, need not be present for the formation of this species. The presence of glucose alone was sufficient for the formation of the species, although production of the species was enhanced by the addition of cellobiose.

Table 10-1. Production of Unknown Compound by Enzyme Mixtures

[Glucose] (w/w)	[Cellobiose] (w/w)	Enzyme	Peak Area of Unknown (normalized)	
			24h	48h
50%	-	Turbo	0.08	0.09
		CF-402	1.00	1.44
	5%	Turbo	0.13	0.14
		CF-402	1.32	1.70

EXAMPLE 11
ANALYSIS OF CELLULOSE HYDROLYSIS PRODUCTS USING
CONCENTRATED ACID HYDROLYSIS

[00491] The unknown species observed in Example 10 was subjected to acid conditions known to hydrolyse oligosaccharides to test if it represented a glucose oligomer. Such a glucose oligomer could be present as either a direct product of cellulose hydrolysis or from synthesis reaction of the direct products through a reaction such as transglycosylation. Cellotriose, a beta 1-4 trimer of glucose, was used as a positive control. Acid hydrolysis was performed by mixing an equal volume of a sample and 98% sulphuric acid. Acid hydrolysis completely abolished the cellotriose peak in a HPLC chromatogram collected using the method described herein (Fig. 12A) but it did not significantly alter the peak area of the unknown species, though it did have a minor effect on its retention time (Fig. 12B). These results indicate that the unknown species is not a glucose oligomer.

EXAMPLE 12
INFRARED (IR) SPECTROSCOPY OF LIGNOCELLULOSIC HYDROLYSATE

[00492] IR spectroscopy was used to analyze the hydrolysate from reactions with pretreated wheat straw in Example 11. Hydrolysates were selected from reactions which had reached their endpoint (*i.e.* no additional glucose formation was observed). Hydrolysate was applied dropwise to the ATR detection crystal of a Bruker Vertex 70 Infrared spectrometer, and each drop was allowed to evaporate to form a cast film. A peak at a wavenumber of 1715 cm^{-1} in the spectrum of CF-402 hydrolysate was not observed in the Turbo hydrolysate (Fig. 13). This wavenumber is associated with a vibrational mode of a carbonyl group. In conjunction with other observations described herein that the unknown species can be produced by the CF-402 enzyme mixture from glucose alone, this indicates that the unknown species is an oxidized form of glucose.

EXAMPLE 13
IDENTIFICATION AND QUANTIFICATION OF GLUCONO-LACTONE IN
ENZYMATIC CELLULOSE HYDROLYSATES

[00493] There are three forms of oxidized glucose: gluconic acid formed by oxidation at carbon 1, glucuronic acid formed by oxidation at carbon 6 and glucaric acid which is oxidized at carbons 1 and 6. Gluconic acid exists in pH-dependent equilibrium with its lactone form (gluconolactone or 1,5-gluconolactone, or D-glucono-1,5-lactone) produced via esterification. High-performance liquid chromatography (HPLC) was performed to characterize these compounds in enzymatic cellulose hydrolysates.

[00494] Glucuronic acid, glucaric acid and the lactone form of gluconic acid were analyzed by anion-exchange HPLC. The HPLC system was a Dionex DX-500 modular chromatography system consisting of a GP50 gradient pump, a Rheodyne (Idex) six port sampling valve, an AS40 automated sampler, an ED40 or ED50 electrochemical detector with a gold working electrode and Ag/AgCl reference electrode for pulsed amperometric detection (PAD). The CarboPacTM PA1 column (4 x 250 mm; Dionex) and guard (4 x 50 mm) consist of a 10 µm diameter polystyrene/divinylbenzene substrate agglomerated with 580 nm MicroBead quaternary ammonium functionalized latex (2% cross linkage; Microbeads).

[00495] Samples of glucuronic acid, glucaric acid and gluconolactone were diluted in water to bring them within range of the standards (0.003 – 0.030 g/L) and injected in a volume of 25 µL. Samples were subjected to an isocratic separation in a solution of 6 mM NaOH for 28 minutes after injection. A 6 minute gradient, raising the NaOH concentration linearly from 10 mM to 300 mM, was then applied. Next, a 5 minute gradient, raising the NaOH concentration from 300 mM to 1 M, was applied. The column was then re-equilibrated with 6 mM NaOH prior to the next run. Peak integration and quantification of standards and unknowns was performed with CHROMELEON® software (Dionex).

[00496] Glucose was mixed with a secreted enzyme mixture harvested from CF-402. Specifically, fifty-two mg of the CF-402 enzyme mixture was mixed with a solution containing both 50% w/w glucose, at pH 5.0 and 60°C for 24 hr and 48 hr. The product of this reaction was analyzed using the above HPLC conditions. One component from the product of this reaction co-eluted with both glucaric acid and gluconolactone (Fig. 14A).

[00497] A second HPLC method was used to analyze the glucaric acid, gluconic acid lactone, and the product of the reaction of glucose with the secreted enzyme mixture harvested from CF-402. This method employs a IonPac®AS11-HC column (4 x 250 mm;

Dionex) and AG11-HC guard (4 x 50 mm) consisting of a 9 µm diameter ethylvinylbenzene polymer cross linked with 55% divinylbenzene polymer agglomerated with a 70 nm alkanol quaternary ammonium latex (6% latex cross linkage) and a capacity of 290 µeq/column (4x250 mm). The HPLC system used for this method comprised a dual gradient pump (DP) with vacuum degassing, dual Rheodyne (Idex) six port sampling valves and an AS automated sampler with diverter valve. The system was equipped with both electrochemical and conductivity detectors. An anion trap column (ATC) was installed in-line between the serial pump and the DC injection valve to remove any anionic species in the eluent and a carbonate removal device which was installed between the chemical suppressor and the conductivity detector to partially remove any carbonate in the mobile phase.

[00498] Samples of glucaric acid, gluconic acid lactone, and the product of the reaction of glucose with the secreted enzyme mixture harvested from CF-402 were diluted in water to bring them within range of the standards (0.003 – 0.030 g/L) and injected in a volume of 25 µL. The samples were subjected to an isocratic separation in a solution of 1 mM NaOH for 6 minutes after injection. A six-minute gradient, raising the NaOH concentration linearly from 1 mM to 60 mM was then applied. The column was subsequently cleaned with a short 1 M NaOH step prior to equilibration in 1 mM NaOH. Peak integration and quantification of standards and unknowns was performed with CHROMELEON® software (Dionex).

[00499] Using this second HPLC method, gluconolactone, but not glucaric acid, co-elutes with a component of the product of the reaction of glucose with the secreted enzyme mixture harvested from CF-402 (Fig. 14B). Therefore, the unknown species is gluconolactone, which is produced by the action of one or more enzymes within the C-1 enzyme system.

EXAMPLE 14

DECREASE IN GLUCOSE YIELD FROM CELLULOSE HYDROLYSIS IN THE PRESENCE OF OXIDOREDUCTASES

[00500] Wheat straw was pretreated and washed as described in Example 1. The cellulose in this pretreated material was hydrolysed by 30 mg *T. reesei* Turbo cellulase per

gram of cellulose. The reaction was performed at 50°C and pH 5.0, with 250 rpm orbital shaking.

[00501] Enzymes from the E.C. 1 group of enzymes, the oxidoreductases, were added to the hydrolysate of this reaction in separate reactions. Glucose oxidase from *Aspergillus niger* (E.C. 1.1.3.4), pyranose oxidase from *Coriolus* sp. (E.C. 10.1.3.10) and glucose dehydrogenase from *Pseudomonas* sp. (10.1.1.47), all purchased from Sigma-Aldrich, were tested. Enzymatic reactions were carried out over 72 h at the pH and temperature optima of each enzyme: pH 8.0 and 37°C for glucose oxidase and glucose dehydrogenase; pH 7.0 and 37°C for pyranose oxidase. For glucose dehydrogenase, a supplement of one equivalent (with respect to glucose) of beta-nicotinamide adenine dinucleotide hydrate (Sigma) was added to the reaction mixture. Enzymes were dosed at 0.45 mg per gram of starting cellulose. This dose corresponds to 1.5% of the full dose of 30 mg/g, simulating the presence of a low-abundance oxidative enzyme in a mixed enzyme system. The glucose concentrations were determined in each reaction flask and in no-enzyme controls by HPLC (Dionex ICS5000 with a CarboPacTM PA1, 4 x 250 mm column (Dionex), with an eluent of 6% 200 mM NaOH in degassed ddH₂O, run time of 25 minutes at a rate of 1.5 ml/min in which glucose retention time is 11.2 minutes) to determine the glucose yield loss. *P*-values were determined from a *t*-statistic calculated for the case of unequal sample sizes and equal variances; there were two to four plus-enzyme and one to two no-enzyme flasks.

Table 14-1. Glucose Yield Loss Due to Enzymatic Action of Oxidoreductases		
Enzyme	Glucose Yield Loss	<i>P</i>-value
Glucose Oxidase	4.38%	<0.01
Pyranose Oxidase	3.81%	<0.01
Glucose Dehydrogenase	2.27%	<0.05

[00502] These data indicate that a significant decrease in glucose yield can be produced by oxidoreductases present in very low abundance in a cellulolytic enzyme system. Elimination or reduction of these enzymatic activities by any means would therefore result in improved glucose recovery.

EXAMPLE 15

SACCHARIFICATION OF ACID-PRETREATED CORN STOVER

[00503] In this Example, experiments conducted using CF-404 on acid-pretreated corn stover are described. In these experiments, 0.81% to 6% CF-404 (with regard to glucan concentration) was added to a mixture of water and pre-treated corn stover (NREL) (90 g/kg glucan) at a cellulose concentration of 9% and pH 5. The mixture was incubated at 55°C, with shaking for 73 hours. During this incubation, samples were periodically taken and the sugar concentration determined using by HPLC, using methods known in the art. Cellulose and residual xylan were found to have been converted to their monomeric sugars in high yield.

SEQUENCE LISTING IN ELECTRONIC FORM

In accordance with Section 111(1) of the Patent Rules, this description contains a sequence listing in electronic form in ASCII text format (file: 81770100 Seq 26-FEB-19 v1.txt).

A copy of the sequence listing in electronic form is available from the Canadian Intellectual Property Office.

CLAIMS:

1. A method for generating at least one fermentable sugar comprising contacting a substrate comprising cellulose with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes, wherein at least one of the two or more cellulose hydrolyzing enzymes is produced by a *Myceliophthora* fungal cell that has been genetically modified to reduce the amount of endogenous cellobiose oxidizing enzyme activity that is produced by the fungal cell, wherein the fungal cell has been genetically modified by deleting endogenous cellobiose dehydrogenase enzymes having at least 90%, about 90%, at least 95%, or at about 95% sequence identity to the sequences of SEQ ID NOS:6 and 8.
2. The method of claim 1, wherein the *Myceliophthora* fungal cell is a *Myceliophthora thermophila* fungal cell.
3. The method of claim 1, wherein the cellulose comprises pretreated lignocellulose.
4. The method of claim 1, wherein the substrate is selected from wood, wood pulp, wood chips, sawdust, hardwood, softwood, newsprint, cardboard, paper pulp, corn fiber, corn grain, corn cobs, corn husks, corn stover, grasses, wheat, wheat straw, barley, barley straw, oat straw, oat hulls, hay, rice, rice straw, switchgrass, cord grass, rye grass, miscanthus, reed carary grass, waste paper, paper, fruit pulp, vegetable pulp, distillers grain, rice hulls, rice straw, cotton, hemp, flax, sisal, sugar cane bagasse, sugar cane straw, beet pulp, sorghum, soy, soybean stover, canola straw, flowers, and any suitable mixtures thereof.
5. The method of claim 1, wherein the enzyme mixture comprises a cell-free mixture.
6. The method of claim 1, wherein said at least one fermentable sugar comprises glucose.
7. The method of claim 1, wherein said at least one fermentable sugar comprises cellobiose.
8. The method of claim 1, wherein the enzyme mixture comprises at least one inhibitor of said two or more cellulose hydrolyzing enzymes.

9. The method of claim 1, wherein the enzyme mixture comprises at least one cellulase enzyme selected from endoglucanases (EGs), beta-glucosidases (BGLs), Type 1 cellobiohydrolases (CBH1s), Type 2 cellobiohydrolases (CBH2s), and/or glycoside hydrolase 61s (GH61s), and/or variants of said cellulase enzyme.
10. The method of claim 9, wherein at least one cellulase enzyme is an enzyme that is endogenous to the *Myceliophthora* fungal cell.
11. The method of claim 10, wherein the enzyme mixture comprises at least one beta-glucosidase.
12. The method of claim 1, wherein the enzyme mixture comprises at least one cellobiose dehydrogenase.
13. The method of claim 12, wherein the cellobiose dehydrogenase comprises CDH1 and/or CDH2.
14. The method of claim 1, further comprising at least one cellulase enzyme and/or at least one additional enzyme.
15. The method of claim 1, wherein the enzyme mixture is subjected to a purification process to selectively remove one or more glucose and/or cellobiose oxidizing enzymes from the enzyme mixture.
16. The method of claim 15, wherein the purification process comprises selective precipitation to separate the glucose and/or cellobiose oxidizing enzymes from other enzymes present in the enzyme mixture.
17. The method of claim 1, wherein the enzyme mixture comprises at least one inhibitor of the cellobiose oxidizing enzyme.
18. The method of claim 1, wherein at least 80%, about 80%, at least 85%, about 85%, at least 90%, or about 90% of the cellulose hydrolyzed by the enzyme mixture is present in the form of cellobiose and/or glucose.

19. The method of claim 1, wherein the method is conducted in a reaction volume of at least 10,000 liters.

20. The method of claim 1, further comprising fermentation of the at least one fermentable sugar to at least one end product.

21. The method of claim 20, wherein said method for producing at least one fermentable sugar and said fermentation of said at least one fermentable sugar are conducted in a simultaneous saccharification and fermentation process.

22. The method of claim 20, wherein the enzyme mixture comprises a cell-free mixture.

23. The method of claim 20, wherein the cellulose comprises pretreated lignocellulose.

24. The method of claim 20, wherein the substrate is selected from wood, wood pulp, wood chips, sawdust, hardwood, softwood, newsprint, cardboard, paper pulp, corn fiber, corn grain, corn cobs, corn husks, corn stover, grasses, wheat, wheat straw, barley, barley straw, oat straw, oat hulls, hay, rice, rice straw, switchgrass, cord grass, rye grass, miscanthus, reed carary grass, waste paper, paper, fruit pulp, vegetable pulp, distillers grain, rice hulls, rice straw, cotton, hemp, flax, sisal, sugar cane bagasse, sugar cane straw, beet pulp, sorghum, soy, soybean stover, canola straw, flowers, and any suitable mixtures thereof.

25. The method of claim 20, wherein the enzyme mixture comprises at least one inhibitor of the cellobiose oxidizing enzyme.

26. The method of claim 20, wherein said at least one fermentable sugar comprises glucose.

27. The method of claim 20, wherein said at least one fermentable sugar comprises cellobiose.

28. The method of claim 20, wherein the enzyme mixture comprises at least one inhibitor of said two or more cellulose hydrolyzing enzymes.

29. The method of claim 20, wherein the enzyme mixture comprises at least one cellulase enzyme selected from endoglucanases (EGs), beta-glucosidases (BGLs), Type 1 cellobiohydrolases (CBH1s), Type 2 cellobiohydrolases (CBH2s), and/or glycoside hydrolase 61s (GH61s), and/or variants of said cellulase enzyme.

30. The method of claim 29, wherein at least one cellulase enzyme is an enzyme that is endogenous to the *Myceliophthora* fungal cell.

31. The method of claim 30, wherein the enzyme mixture comprises at least one beta-glucosidase.

32. The method of claim 20, wherein the enzyme mixture comprises at least one cellobiose dehydrogenase.

33. The method of claim 32, wherein the cellobiose dehydrogenase comprises CDH1 and/or CDH2.

34. The method of claim 20, further comprising at least one cellulase enzyme and/or at least one additional enzyme.

35. The method of claim 20, wherein the enzyme mixture is subjected to a purification process to selectively remove one or more glucose and/or cellobiose oxidizing enzymes from the enzyme mixture.

36. The enzyme mixture of claim 35, wherein the purification process comprises selective precipitation to separate the glucose and/or cellobiose oxidizing enzymes from other enzymes present in the enzyme mixture.

37. The method of claim 20, wherein the enzyme mixture comprises at least one inhibitor of the cellobiose oxidizing enzyme.

38. The method of claim 20, wherein at least 80%, about 80%, at least 85%, about 85%, at least 90%, or about 90% of the cellulose hydrolyzed by the enzyme mixture is present in the form of cellobiose and/or glucose.

39. The method of claim 20, wherein the method is conducted in a reaction volume of at least 10,000 liters.

40. The method of claim 20, wherein the end product is a fuel alcohol or a precursor industrial chemical.

41. The method of claim 40, wherein the fuel alcohol is ethanol or butanol.

42. A *Myceliophthora thermophila* fungal cell that has been genetically modified to reduce the amount of endogenous cellobiose oxidizing enzyme activity that is produced by the fungal cell, wherein the fungal cell has been genetically modified by deleting endogenous cellobiose dehydrogenase enzymes having at least 90%, about 90%, at least 95%, or about 95% sequence identity to the sequences of SEQ ID NOS:6 and 8.

43. The fungal cell of claim 42, wherein the fungal cell has been genetically modified to reduce the amount of endogenous cellobiose dehydrogenase produced by the fungal cell and to increase the production of at least one saccharide hydrolyzing enzyme, wherein said at least one saccharide hydrolyzing enzyme is selected from beta-glucosidases, endoglucanases, cellobiohydrolases, esterases, beta-xylosidases, xylanases, arabinofuranosidases, alpha-glucuronidases, acetylxyylan esterases, feruloyl esterases, alpha-glucuronyl esterases, and GH61 enzymes.

44. The fungal cell of claim 43, wherein said at least one saccharide hydrolyzing enzyme is a *Myceliophthora thermophila* enzyme.

45. The fungal cell of claim 43, wherein said at least one saccharide hydrolyzing enzyme is overexpressed by said fungal cell.

46. The fungal cell of claim 43, wherein said overexpressed saccharide hydrolyzing enzyme is beta-glucosidase.

47. A *Myceliophthora thermophila* fungal cell that has been genetically modified to reduce the amount of endogenous cellobiose oxidizing enzyme activity of two or more endogenous cellobiose oxidizing enzymes that are produced by the fungal cell, wherein the fungal cell has been genetically modified to at least partially delete the genes encoding the two or more

endogenous cellobiose oxidizing enzymes, wherein the first of the two or more cellobiose oxidizing enzymes comprises an amino acid sequence that is at least 90% or about 90% identical to SEQ ID NO: 6, and wherein the second of the two or more cellobiose oxidizing enzymes comprises an amino acid sequence that is at least 90% or about 90% identical to SEQ ID NO: 8.

48. The fungal cell of claim 47, wherein the fungal cell has been genetically modified to reduce the amount of endogenous cellobiose dehydrogenase that is produced by the fungal cell and to increase the production of at least one saccharide hydrolyzing enzyme.

49. The fungal cell of claim 48, wherein the at least one saccharide hydrolyzing enzyme is selected from endoglucanase, cellobiohydrolase, beta-glucosidase, endoxylanase, beta-xylosidase, arabinofuranosidase, alpha-glucuronidase, acetylxyylan esterase, feruloyl esterase and alpha-glucuronyl esterase.

50. The fungal cell of claim 48 or 49, wherein the at least one more saccharide hydrolyzing enzyme is: (i) endogenous to the fungal cell and/or (ii) is overexpressed by the fungal cell.

51. The fungal cell of claim 50, wherein the saccharide hydrolyzing enzyme overexpressed by the fungal cell is beta-glucosidase.

52. A method for generating cellobiose and/or glucose comprising contacting cellulose with an enzyme mixture comprising two or more cellulose hydrolyzing enzymes, wherein the enzyme mixture is produced by the fungal cell according to any one of claims 47-51, and wherein the enzyme mixture comprises a glycoside hydrolase 61 (GH61), a Type 2b cellobiohydrolase (CBH2b), a Type 1 cellobiohydrolase (CBH1), a beta-glucosidase (Bgl), and at least one endoglucanase (EG).

53. The method of any one of claim 52, wherein the cellulose comprises pretreated lignocellulose.

54. The method of claim 53, wherein the pretreated lignocellulose comprises lignocellulose treated by a treatment method selected from acid pretreatment, ammonium pretreatment, steam explosion and/or organic solvent extraction.

55. The method of any one of claims 52-54, wherein the enzyme mixture is a cell-free mixture.

56. The method of any one of claims 52-55, further comprising fermentation of the glucose to an end product.

57. The method of claim 56, wherein the end product is a fuel alcohol or a precursor industrial chemical, and wherein the fuel alcohol is optionally ethanol or butanol.

58. The method of any one of claims 56-57, wherein said method for producing cellobiose and/or glucose from cellulose and said fermentation are conducted in a simultaneous saccharification and fermentation (SSF) process.

59. The method of any one of claims 52-58, wherein the method is a batch, fed-batch or continuous process, or a combination of batch, continuous, and/or fed-batch processes conducted in any order.

60. The method of any one of claims 52-59, wherein the method is conducted in a reaction volume of: a) at least 10,000 liters or b) at least 100,000 liters.

61. The fungal cell of any one of claims 47-51, or the method of any one of claims 52-60, further comprising a cellulose degrading enzyme that is heterologous to the fungal cell.

62. A fermentation medium comprising the fungal cell of any one of claims 47-51.

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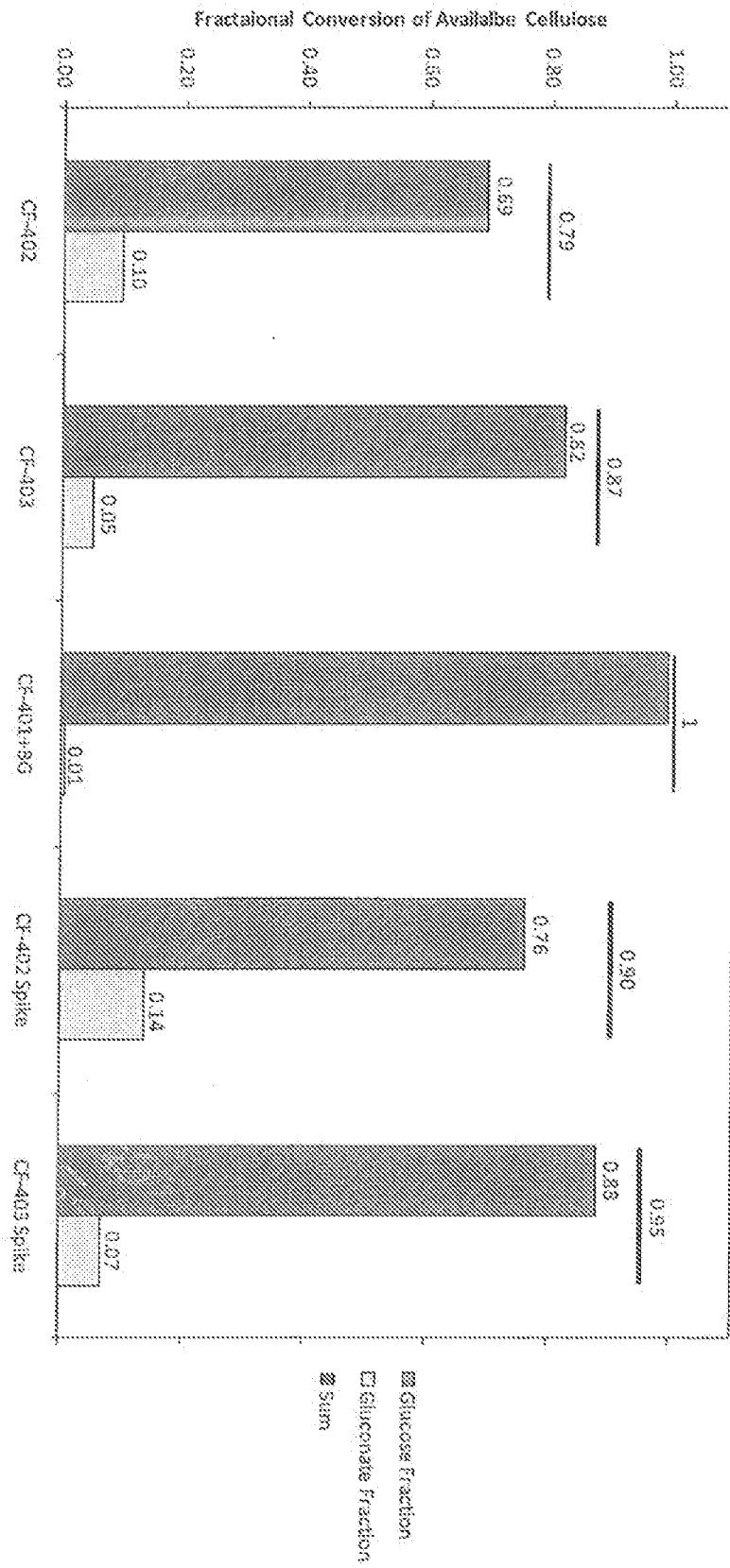
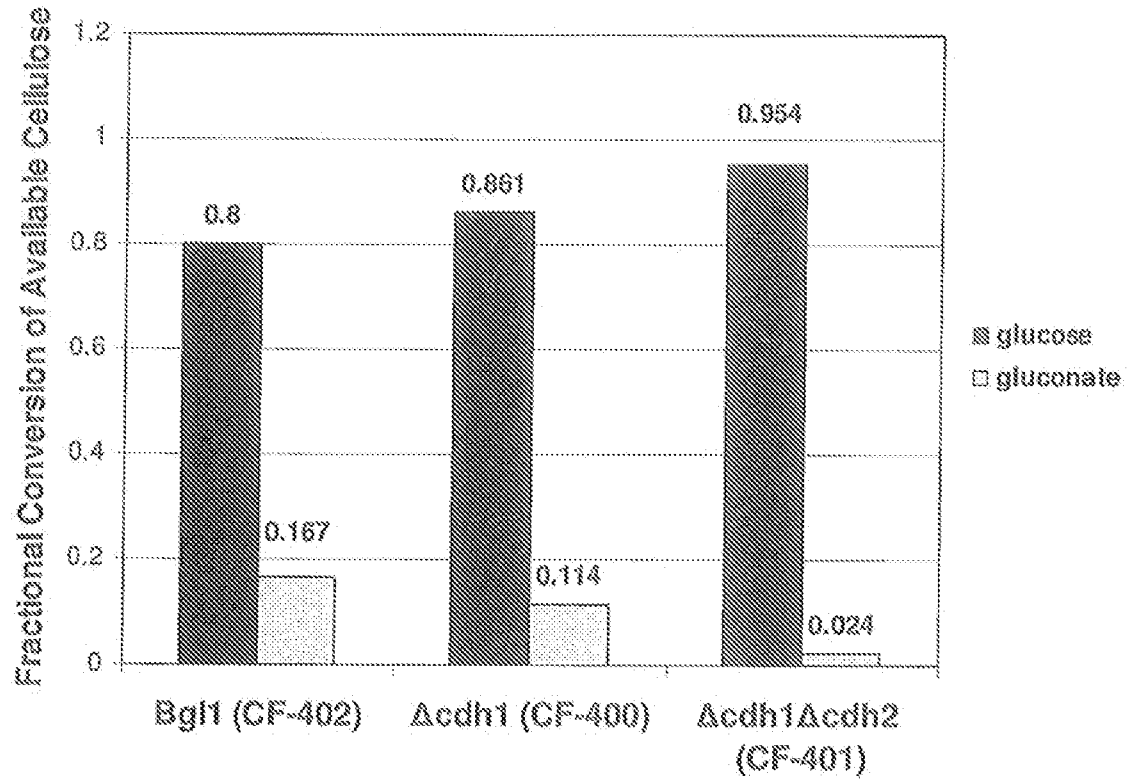


Fig. 1

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Fig. 2



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Fig. 3

*(M. thermophila -- CDH1)*Nucleotide sequence (SEQ ID NO:5)

ATGAGGAACCTCTCTCGTTTAATCGGTGCCCTTGGGCGGCACTCTTGGCGTCTGCCCTTGGCAGAACACGGGGCCGGTAAC
 OTTCAACCGACCGGACTCGGGCATTAGCTTCAACACCTGGGGTCTGCCCGAGGATTCTGCCCAGACTAAGGGCGGTTTCACTTT
 TGGTGTGTCTCTGCCCTCTGATGCCCTCACGACAGACGCCAAGGAGTTGATCGGTTAGTTGAAATGCGCGAGGAACGATGAGA
 GCGGTTGGTGGCGGTGTCTCCCTGGCGGGCCGATGACCAACTCGGTCTCATGCGGGCTGGCCCCACGAGGACACGCTOTA
 CAGCTCTCTCGGCTTGCCCAACCGGCTATGCCATGCGGATGTGTACCAGGGGGACGCGGAGATCACCCAGGTCTCTCTCTGTG
 TCAACTCGACGCACTTCAGGCTCATCTTCAGGTGGGAGAACTGCTGCAATGGAGTCAAAGCGGGCGCCACGGGGCGGTGCTCTC
 ACCTCGAACGGGCTGTTGGTCTCTCGGCTGGGTCCAGGCATTGGCGACCCCGGGAACCCGACCTGCCCGGACCGAGATCACCC
 TCGAGCAGCACGACAAACGGGATGCGTATCTGGGGTGGCCAGCTCAACTCGGACGGCGGCAACCCGCTCTTACACCGAGTGGG
 CGCCACGGCCACCAAGACCGTACCGGGTGAAGTGGCGGGTCCACCGAGAGCTCTGTCTGTGGTGTCTCCCGTTCCGACGGGG
 GTCTGTGTGATTACATCTCTGTGGGCGGGCGGTGGCGGTGGCATCCCGCGCGCGGACAAAGGTGAGCGAGGGCGGCAAGAGTG
 TGCTGCTCATGAGAAAGGGCTTTGGTCTGACCGGCCAACCGGAGGGGACTCTCGGCGCGGAGTGGCTCGAGGGGCCACGACCT
 TACCCGCTTTGACGTGCCGGGTCTGTGTCAACAGATGTGGGTTGACTCCAGGGGATCGGTTGGGAGGATACCGACGAGATGG
 CTGGCTGTGTCTGTGGCGGGGTACCGCGGTGAATGGCGGCTGTGGTTCAAGGCGTACTCGCTGAGTGGGACTACCTGTTCT
 CTAAGTGGTTGGAAGTACAAAGACGTCCAGCGGGCCATCAACCGCGCGCTCTCGCGCATCCCGGGCACCGATGCTCTCTCGAC
 CGAGGGCAAGGGCTACTACCAACAGGGCTTCGAGCTCTCTCAAGGGGCTGGCGGGCGGGGCTGGAGCTCGGTACAGGGCC
 AATAACGGCGCGCAGAACGAAGAAAGACGCTTCTCCCATCGCGCGCGCGCATGTTCTGTTCCCGCGCGGGAGCGCAAGCGCGCTGTG
 GCACCTACTTCCAGACCGCAAGAAGCGCAGCAACTTCAAGCTCTGGCTCAACAGTGGGTCAAGCGGGTCACTCCCGCAGGGC
 GGCCACATCAACGGCGCTCGAGGTGAGCGGTTCCGCGACGGCGGTACCAAGGCGATCGTCCCGCTCACCAAGGTTACGGGGCC
 GCGTCATCTCTCTCGCGGTACCTTTGGCAGTGCAAAGATCTCTGCTGAGGAGCGGTATCGGTCCGAACGATCAGCTGCGAGGT
 QTCGCGGCTCGGAGAAAGGATGGGCGTACCATGATCAGCAAGCTGCTCGGATCAAGCTGCGTCTCGGCTACAACCTGGATGA
 CCACCTCAACACCGACACTGTCTCTCCACCGCGAGCTGTGTCTACGACTTCTAAGAGGGCTGGGACAATCCCATCGAGT
 TGACAAGGACAGCTACCTCAACTCGGCGACGGGGCATCTCGGCCAAGCGGCTCCCAACATTGGGCGTATGTTGTGGGAAGAGA
 TCAAGGGTGGCGAAGGCTATTGTTCCCGAGCTCCAGTGGAGTGGCGGTCTCGAGGGCAGGCTGGGTGCCCCCAAGGGCAAGAG
 TGAACCATGTCTGAGTACCTGGTCTGGTGGCACTCGGCGCGCGCATGAGCATCACCGGCTCGCTGACAAGCTGCTGTCT
 CGGAGGTGCGCTACCTCAAGGACCCCAACGACAAGGAGGCGGTCTATCCAGGGCATCATCAACCTGCAGAACGCGCTCAAGAAC
 GTGCGCAACCTGAGCTGGCTCTTCCCGCACTCGACCATCAAGCGCGCGCAATAAGTTGACAGCATGCTGTCTCTCCCGAGGCAA
 CCGGCGCTCCAGCACTGGATGGGCAACCAAGATCGGACCGAGCAGCGGGCGCAAGGGCGGCTCGGCGGTCTGTGCACTC
 AACACCAAGGTCTACGGCACCGACAACCTCTTGTGATCGAGCGCTCATCTTCCCGGGGTGCGCAACCAACCAACCGCTGTG
 TACATCGTGAAGCGGTGAGGACAGCGCTGGGCGCGCATCTGCGCTGCGCGACCTACGCGCGTCCCAAGTACGGGCGGT
 GCGCGCGCGGCAATGAGAGCGGCGAGTTCTGTCTGCGCGCAGCGGTCCAGCTGCCAGATGACAGAACGAGTGGTACTCGGAGTG
 CTTGTGA

AA sequence (SEQ ID NO:6)

MRTSSRLIGALAAALLPSALAQNNAPVFTDPDGGITFNTWGLAEDSPQTKGGFTFGVALPSDALTTDAKEFIGYLLKGRNDESGWCG
 VSLGGPMTNLSLIAAWPHEDTVYTSRFAIDYAMPDYYQGDAEITQVSSSVNSTHFLIFRCENGLQWSQSGATGGASTGNGVLVLQ
 WVQAFADPGNPTCPDQITLQHDNGMGIWGAOLNSDAASPSTEWAAQATKTVTGDCGGPTETSVVGVPVPTQVSFDYIVVGGGA
 GGIPAADKLEAGKSVLLIEKGFASTANTGSLGPEWLEGHDLTRFDVPGLCNQIWWVDSKGIACEDTDMAGCVLGGGTAVNAGLWF
 KPYSLDWDYLFPSGWKYKDVQPAIRHLSRIPGTDAPSTDGKRYQQGFQVLSKGLAGGGWTSVTANNAPDKKRTFSHAPFMFAG
 GERNGPLGTIFYQTAKRSNFKLWLNTRYKRVIRQGGHITGVEVEFFRDGGYQGIVPVTKVTRVILSAGTFQSAKILLRSGIGPNDOL
 QVVAASEKDGPMTISNSSWINLPVGYNLDDHLNTDTVISHPDVVFYDFYEAWDNPIQSDKDSYLSNRTGHLAQAAAPNIGPMFWEEIKG
 ADGIVRQLQWTARVEGSLGAPNGKMTMSQYLRGATSRGRMTITPSLTTVVGDVPYLLKDPNOKEAVIQGIINLQNALKNVANLTWLF
 PNSTITPRQYVDSMVVSPSNRRSNHWMGTNKGTDGGRKGGSAVVDLNTKVYGTDLNLFVIDASIFPGVPTTNPSTSYIVTAGEHASARIL
 ALPDLTVPKYGGCGGREGWGGGFVCAAGSTCCMQMONEWYSGL*

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Fig. 4

*(M. thermophila -- CDH2)*Nucleotide sequence (SEQ ID NO:7)

ATGAAGCTACTCAGCCGGGTTGGGGCACCAGCCCTAGCGGGCAGCGTTGTCACTGCAGCAATGTGCAGCCGAGATGAOCGAGG
 GGAOCTACACCGATGAGGCTAGCGGTATCCAATTCAAGACGTGGACCGGCTCCGAGGGGGCCCGTTTCAAGTTTGGGTTGACC
 CTCCCGCGGACGCGCTGGAAAAGGATGCCACCGAGTACATTGGTCTGCTGCGTTGCCAAATCACCGATCCCGCTCGCCAG
 CTGGTGGGATCTCCACGGGAGTCCGGCCAGATGACGACGGCGGCTGCTGCTGGTGGCTGGGGCAGGAGGACACCGTC
 TACACGCTCGTTCCGCTACGCCACCGGCTACACGCTCCCGCGGCTCTACACGGGCGACGCCAAGCTGACCGAGATCTGCTGCTC
 GGTACGCGAGGACAGCTTCCGAGGTGCTGTTCCGCTGCGAAACTGCTTCTGCTGGACCGGATGGACCAAGGCGAAAGCTGT
 CGACCAAGCAACGGCAACCTGGTCTGCGCGCGCGCGCGCGGAAGGATGGTGTGACGGGCCCCACGTCGCCCGACACGGCGCG
 AGTTCCGTTTCCATGATAACGGTTTCCGACAGTGGGGTCCCGTGGTTBAGGGTCTACTTCCGACTGTAACGAGGAGTGGGCT
 AAGCTGGCCACGACCGCGCGGAGACCGTGGATGGGACTGGCCCGCGGACAAAGGAGTGGCTTCCGGCTCCCGAGGACA
 CGTATGATTACATCGTTGTGCGGTGCGCGCGCGCGGTGGTATCACCGTGGCGGACAAAGCTCAGCGAGGCGCGCGGACAAAGGCTGTT
 CTCATCGAGAAAGGACCGCGCTTCGACCGCGCGCTGTGGAACGGGACCATGAAGCGCGAGTGGCTCGAGAGGCAACCGACCTTACCG
 GCTTGGACGTTCCCGCGCTGTGCAACCGATCTGGGTGCGACTGCGCGCGCATCGCTGCACCGATACCGACCGAGTGGCGCG
 CTGCGTTCTCGCGCGTGGCAGCGCTGTCAACCGCTGGTTTGTGGTGGAAAGCGCGCGCGCTGACTGGGATGAGAACTTCCCGG
 AAGGGTGGAAAGTGGAGCGATCTCGCGGATGGGACCGAGCGTGTCTTCAAGCGCATCCCGCGACGTCGACCGCGTGGAGGA
 CGGCAAGTTGTACCGCGAGGAGGCTTCCAGGTCATCAGCAAGGGCGCTGGCGAAACCGCGCGTGGAAAGGAAATCAGCGCGCAAC
 GAGGCGCGCGAGGAGAGAAACCGACCTATGCAGACACCGAGTTTCACTGTTCTCGCGCGGTGAGCGTGGCGCGCGCTGGCGA
 CGTACCTTGCCTCGGCTGCGGAGCGGACGCAACTTCAACCTGTGGCTGAAAGCTGCGGTCCGGAGGGCGCTCCCGAGCGGCGAG
 CAAGGTCAACCGCGCTCGAGGCTCGAGTGGCTCAACCGACCGTGGCTTCAACCGGAGCGTCAACCTGAATGAGGGCGGTTGGTGT
 ATCTTCTCGCGCGCGCGCTTCCGCTCGCGCAAGGCTGCTCTTCCGAGCGGTATCGGTCTCGAGGACCGCTCGAGATTGTGGC
 GAGCTCCAAAGGACCGGAGACGTTCACTCCCAAGGACGAGTGGATCAACCTCCCGCTCGCGCGACAACTGATCGACCATCTCA
 ACACTGACCTCATTATCAGCGACCGCGGATGTGCTTTTCTATGACTTCTATGCGCGCTGGGACGAGCGCATCACGGAGGATAAGG
 AGGCTACCTGAACCTCGCGCTCGCGGCTTCTCGCGCGAGCGCGCGCGCAATATCGCGCGCTATGATGTGGGATCAAGTCAACCGG
 TCCGACCGCATCAACCGCGAGTTCAGTGGAGATGCGGTGTTGAGGGCGGACAGCTGCAAGACCAACTCGACCGACCGCATGAC
 CCTCAGCGAGTACCTCGCGCGCTGGCGTCTGCTCGCGCGCGCGGATGGGATCACCTCGCGGCTGAGCAACCGGCTGGCGG
 GCACCGCTACCTGCAACAACCGCGGACCTGGAGGCGGTGATCCAGGGGATCCAGAACGCTGGTGGAGCGGCTCAGCGAGGTG
 GCGACCTCGAGTGGGTGCTCCCGCGCGCGGACGGGACGGTGGCGGACTACGTCAACAGCGCTGATCGTCTCGCGCGCGCAACG
 GCGCGGCGCAACCGCTGGATGGGACCGCGCAACGCTGGGACCGGACGCGCGGCTCGGGCGGACCTCGCTCGTGGACCTCG
 ACACCAAGGTGTACCGGCAACGCAACCTGTTCTGCTGCTGACCGCTGCTGCTTCCCGCGCATGTCGACCGGCAACCGCTGGG
 ATGATCGTCATGCTGGCGAGCGAGGCGCGGACGCGCATGCTGGCGCTGCGGTCTTAA

AA sequence (SEQ ID NO:8)

MKLLSRVGATALAATLSLQCCAAQMTGTYTDEATGIGFKTWTASEGAPFTFGLTLPADALEKDATEYIGLLRCQITDPASPSWOGISH
 GGSQMTQALLVWASEDTVYTSFRYATGYTLPLGYTGDAKLTQISSVSSEDSFEVLFRCEGCFSWDQDGTGKGNVSTNGLVLG
 RAAAKDGVGTGFTCPDTEAFGFHONGFGQWGAIVLEGATSSYEAWKLATTTPETTCDGTGPDKECVPAPEDTYDYIVVAGAGGI
 TVADKLSEAGHKVLLIEKPPSTGLWNGTMRPEWLESTDLTRFDVPLCNQWVDSAGIACDTDQMGAGCVLGGGTAVNAGLWVKP
 HPADWDENFPEQWKSSDLADATERVFKRIPOTSHPGQDKLYRQSGFEVSKGLANAGWKEISANEAPSEKNIHTYAHTFPMFSGGE
 HGGPLATYLAASAEKSNFNLWLNATVRRVRSKVTGVELECLTGGGFGTGNLNEGGGVIFSAAGFSAKLLLRSGIGPEDQLEIV
 ASSKDGETFTPKDEWNLVQHNLDHNLNLTDLIHPDVVFYDFYAAWDEPITDKAYLNSRSGILAAAPNIGPMWQDQVTPSDGITH
 QFQWTCRVEGDSKTNSTHMTLSOYLGRGVVSRGRMGITSGLSITVAEHPYLNHNGDLEAVIGGIGNVVDALSQVADLEWVLPFP
 DGTVDYVNSLIVSPANRRANHWMTAKLGTDDGRSGQTSVVDLDTKVYGTGNLNVVDASVFFPMSTGNPSAMIVVAEQAAQRILA
 LRS*

MGFLAATLVSCAALASASIPRPHAKRQVSQLRDDYDFVIVGGSTSQLTVAORLTEAPPKKNVLVIEYGGDVHYAPGTFDPPPTDWITPQ
PDAPPSWSPNSLPNDMANTAFVLAAQQVVGGSAAVYGMFFDRASRHDYDAWTAUVGGSGFEQSSHKWCWEGLPFPFQKSVTFTE
PPADIVQKHYDITWDLRSAYNGNSTPIYSSYPVQWQADQPLLNQAWQEMGINPVTACAGQKQGVQWVSPASQHPVPTARKSHAGLOHY
ADVLPRANYHYLVQHOVVRVYFVFNPGSHGPPLEARSLLADNHLPNVTVKGEVISAQALHTPTVLQSGIGFASFLDQAGIPVTLDLQ
VGANLQDHGGPPVTWNYTEPYTOFFPLPSEMVNNTATFKAEAITGFDEVPARGPYTLACGNNAIFVSLPHLTADYGAITANIRAMVADG
TAASYLAADVRTPGMVAGYEAQILLGLADLLNPEAPSLTEPWTATSEAPQTSSVLAFLLLPLSRGSGVRLNLSPLAQPVLDYRSGSNP
VDIDLHLHVRFLRLGLDTPMTMQARGALETAPGSAVDSDEALGEYVRSHSLTSFMHPCCTAAMLPEDRGGVVGPDCLKVHGAEGRLV
VDMSVMPPLPGHLSATAYAVGEKAAIDHCEWMDKEQ*

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Fig.6

*(M. thermophila -- GO2)*Nucleotide sequence (SEQ ID NO:3)

ATGAGAGCTGCTTGGAGTCTCCCTCGCGGTGTTGCACTCTCCCATTAATATTATTCCGGGTTGCAGCGCGCCACGGTACCGGCG
 CGATCCATTGCCCGCTCCACGATTCTTGACGGAGCGGATGGGCTTCTTCCGGAGTATGACTACATCATCAGGGGGCGGCAG
 GTCCGGATTGACTGTGCGCGACAGACTCACGGAGAATAGAAAGCGCAAGTTTTCCCGCTCTCCCTCCCAACGTACCGCGCG
 GATCGTCACCGCGGTGGTGTATTCTGTTCTTCTTTTGGAAAGAGGCGATTTCCAGAACTCTAGCTCGGTGACCAACATTCTGG
 GGGAAAGCAGAGGCGCTTTCGATCCAAAGTCTGACCTTCAACATCAACTCCGTTCCCGAAGCTGGGGTGGACAACCGCAGCATTGC
 CGTCATTGGCGGGTTGATCGTCGGCGGCGAGCTCCGGGCTCAACGGGCTTCAAGTCTCTCGTGGACAAGAGAGACTATGACC
 GCTCGCGGATCGTACTTTGGGDCAAACTCTGACTGGAGTTGGAAGGTTCTCCTGCCGTATTTCAGAAGGCGATGGAATTTCCATC
 CGCCAGGGCCAGAGCTGGTCAGTCAGTTGACATCAAGTACGACGCCAGCTACTGGGGCAACAGCTGTGAGCTGCAAGCCTCT
 TTCCCAACCACTTTCTGGCGGGTGTCAAAATTGGAGATGGCTGCATTGGTGACATGCTGCGGTGCAATATCCGGCGACTCT
 GCTTCTGGCGAGACCGCGCGGTATTTGGACCCAGCGTGGTTGACCCAGCGACAGTCTCGGTCTGCTTCTGCTGGCGCGCGCG
 ATTGGGACAACATTGAGCGCGCACGTCCCAATTACACACCCCTGACCGGGCAACCGGTATTGAAGGTGGCATTGATGGCAATC
 GAGGACACAGCGTCTGTTCTGTCGGCGGAATGCAACGATCACAGCACTGCCAGGTCTGAGGCGCAAGAGGAGATCGT
 CTGGCGCGCGCGCGGATTACACAGCGCGCAATCTACAGGCGAGCGGAGTAGGGCGCAAGCAGGTCTGAGGGAAGCAGGCG
 GTGCGCGTGTGTTGAGCGTCTCGGTGTGCGCGCAATTTCAAGACCGAGCGGTATGTTGCTGCTCCGACCTTCAATTTACG
 AAGTTCCGCTTCCAGCGGAGTCTTACGACATGATTCTGAACGAGCTTTTATCGCGGAGGCTCAGGCGCAGTTTGAAGAGGAC
 CBTACCGGACCTCACAGCATCGCATCGGCTATTGCGGCGAGTGGGTCCCGCTCCAGATCATTGCCGCAAAATTGGTGAAGGA
 CATCGGTAGCGCGTACGAATCCCAAGACCGAGCGCGTACCTCCCGCGCGGCAACCGATGAGACCGTCATCGAGGGGTACAGG
 GCGCAGCAGAAAGCACTAGCGAGGTCCATGAGGAGCAAGCAATCGGCAATGTATAACTTCTTCTGAGCGGGCGGCTACGAAGA
 GGGTCTGTGCTCTACTTGCACCGAAGCAGCGGTGGCACCGTTCGATCAACCGATCGAGCGGCTTCTTCTGCGCGCGCGAGGT
 CGACTACAGGGCACTGAGCAACCGTACCGAAGCTGGAGGTCTGCTGGAATTCACCTCCCTTACCGCGCAGGTACTTCTTGGAGAC
 GAGGTTGAAGTCCCTGAGCGCGGTGAGGCTGTGCGCGCGGTGCCAACGTCACGGCGCGCGCGGACATCGAGGCGTGGCTTGGC
 AGCGTCATGATCCGCTCTCTTCCATCGGATCGGACCGCGCGCATGTTGCTAGGACCGTGGGTGCTGCTGCGGAGAGAA
 CCTTCTGCTGATCGGGGTGGAAGGCTGAGTGTGCTGAGCGCGCGGTGATGCGCGGCTGATACCGGAGCAGAA
 CCGTGTATGCTATTGCTGAGAAAGCGCGGATCTCATTAGAGCAGGGCTTGA

AA sequence (SEQ ID NO:4)

MELLRYSLAAVALSPLILFQVAAAHPTARSIRSTILDGAGLLPEYDVIHGGGTSGLTVADRLTENRKRKFSRSPLTSPARSSPAWCV
 SVLVLERGIFONSSSVTTISGSSRGLFDPSTLFNINSVPQAGLDNRSAVIGGLILGGSSGVNGLQVLRGOREDYCRWGSYFGPNSDW
 SWKGLLPYFKKAWNFHPPRPPELVSCFDIKYDPSYWGNTSDVHASFFTFWVPLKLEMAAFGDIPOVEYPPDSASGETGAYWHPASV
 DPATVLRGFAHPAHWDNIEAARPNYHTLTGQRVLKVAFDGNRATSVFVFPANATDHSTARSVKAKKEIVLAAGAIHTPCILQASGVGP
 KQVLKEAGVPLVVDAPGVGSNFDQDPYVVPFTFNFTKFPFHPDFYDMILNOTFIAEAQAQFEKDRTPHTIASGYCGSWLPLOIAPNS
 WKDIARRYESQDPAAAYLPAGTDETVIEGYRAQKALARSMSRKSQSAMYNFFLRGGYEEGSSVYVYLPHTSRGTVRINRSDPFFSPPEVD
 YRALSNPTDLEVLLEFPTFRYFLETRLKSLDPVELSPQANYTAPADIEAWLRSMIPSGFHPIGTAAMLPRHLOGVVDENLLVYGVE
 GLSVVDASVMPDLPGSYTQQTYYIAEKAADLIKSR*

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Fig. 7

*(Aspergillus oryzae -- Pyranose Oxidase)*Nucleotide sequence (SEQ ID NO:9)

ATGTCATGACATCAGGACGTCAAGCGTTTACTTCCGAGTGCAGAGATTCAAATACCAAAATTCATTTTGGTTGGCTAATTCAQD
 GACTCTCACACTTGGCTCTACGATGCAGGTCTGCGGGTCCGGCCCATCGGGCCACCTATGCCAAQATTCTAGCTGACCGCG
 GCAAGGATGTCCTCATGGTTGAGACTGGCACCCAGGAAAGTAAGATTGCTGGAGAGCATAAGAAGAATGCTATCAACTACCAGA
 AAGATATCGATGCGTTTGTGCATGTCATTAAGGTAATCAGCTCAAGAATTAGCACTTTGAGTGTATTTCTCTAAGTTTCGATCTTC
 TCCTCTTTTCAGGGAAGTCTACACTACACGTCTGTACCGACCAACAAAGCCGCCCTTCTACACTGGCTCCGATCTCCTGGAAG
 CGAACCGGCCAAATTTTCAACGGACAGAAATCCGGCCAGGATCCAAACGTAAACCTGGATGCCAATGGTGTGGCACGTAATGTGG
 GCGGCATGTCTACCCACTGGACTTGTGGCACTCCCGACAGAAAGAGAAGGTTGAACGCAGCGATATATTCAGTGGTGACGAAT
 GGGATAGCCTGTACAAGGAGGACAGAAAAGTTGATCGGAACGACCAAGACTGTGCTGAATGACTGGATCCGGGAAGAATTGGTC
 ATGGAGATTCTGAATGACGAGTACGGGAAGCGATCAGCCGAACCACTACCTTTGGCTGCAAGAGAGGAATGGCAATACGGCCTA
 CATCACTTGGTCACTCTGCTCAACTATCCTTGAAGGATGAAGTGAAGAAGAAATTTACAGTATGGCCCGAGGACCACTGTGAG
 AAGTTTAAAGTGGAGGAACAGATAACGGGGCCACAGGTCAACCAAGGCTATAATCGGCAAACTCGCCACAGATAAAGTATTACA
 GTTAAGGGGAAAGTATTATGCTTGGCGGGGGCCCTATACCTACACCCAGCTACTTTTCAATTGGGGTTCTGTGGGACAAAG
 CCCAAGAGGGATCCAGAAACCCAAATACCATTAAGAGACGAGAGAAAGGATCCCACTCCACCGGATCTCTGGAGCATCTG
 AAGCTTCTGCTCTAGGACGCTATCTGACAGAGCAAGGATGTGCTTCTGCCAAATTGTTCTGAAAAAGAAATGGATTGAGGCAG
 TGGCTAATCCAAAAAGAACCTTATCAAGCGATGGGGTGAAGCGCAAAAGTGGGAGAAGCTCAAGGAAGGGTGGAGGAA
 AGGGTCCAGGAACATATGAAAAGGTTTAAAGACCTATTCCCTTCCCGTTGATGATTGGGACCTCAGGTTACTCTACCTTGG
 ACTATACCATCCGTGGGATACCCAAATCCATCGCGATGCTTCTCCTATGGGCGAGCAGCCCGAGCAATTGATAAGCGGAGCA
 TTGTTGACCTCCGATTCTTGGAAAGGTTGAGCGGACTGGAAGAACTATGTGACCTTTGAAACCGACATCAGGGATGCGTACG
 GCATGCCCGAGCCACCTTCCGCTACAAGCTGAACGATGAGGATGGCAAGCGTGGACACAGATGATGAAGATATGGAAGAG
 GCGGCTGGTGTCTGGGTGGCTACCTCCAGGGTGGGAGCTCAATTTCTAGCTCCTGGCCTTCCACTGGACGTCTGTGGTAC
 CACTAGAGGTCAGAAGAAGGAGAAAGAGTGTGACCTGATCCCAAGAGAGCTCGTGGTGGGATGAGAAGTCCAAAGATCTGGG
 GTATCCACAACTGTACGTGGGTGGGTTAAATGTGATCCGTGGTGGCAATGGGTCCAACTACCTTGACAGCAATGTGCTTGG
 CCATCAAAAGCGCGAAGATATCCTTGAAGGGAATTCTTAG

AA sequence (SEQ ID NO:10)

MGMTSGRQAFSTSECRDSNTTNSFWLANSPTLTGSTMVVGSGPIGATYAKILADAGKDVLMVETGTGIESKIAGEHKQNAINYQKDID
 AFVHVIVKISSRISTFECISLTFDLLLLFQGSLSHYTSVPTNKAAPVTLAPISWKANGQIFNGCNPRQDPNVNLDANGVARNVQGMSTHWT
 CATPRQKEKVERSDIFSQDEWDSLYKEAEKLIGTSKTVLNDSSIRGELYMEILNDEYGRSAEPLPLAANRNGNTAYITWSSGSLDAM
 NOKKFTLWPEIHCEKFKVEETDNGPQVTKAIRKLATDKLITVYKAKVFIACGGHILTPQLLPNSGFVPTKPNRDPRTQIPLEDDEKGIFF
 PPDLEHLKLPALGRYLTEQSMCFQGVKKKEWIEAVANPKKNFYQSDGVKRRKWEKLKEGWKERVGEHMKRFNDPIPPFDLDLPQ
 VTLPLDYHHPWHTQIHRAAFSYGAAPPADIKRTIVDLRFPGTVEPDWIKNYVTFETDIRDAYGMPQPTFRYKLNDEQKRSHQMMKDM
 EEAAGALGGYLPQSEPOFLAPGLALHVCCTTRAGKKEKECDPDKETSCCDENSKIWGIIHNLVQGLNVIPGANGSNPTLTAMCFAIK
 SAKSILEGNS

MYRICELTAAALSLASVVQASWICKRNSINACI.AAADVEFHEEEDSEGWDMDGTAFLNRVDYDPAIAIAPRSTEDIAAAVOCGLDAGVQIS
AKGGGHSYSGYGGFAGGDSHLMLELDPMYRVSDNNVATIQGGARLGTYALELDQGNRLASHGTCAPVAGVYGLVLSSGGYGFATH
THGLTLDWLQIATVVLADAGHIVHSETENADLFWALRGQGGQFAIVSEFEFNITFEAPEINTYQVTTIWNRRKHVAGLKALODWAONT
MPRELSMRLEINANALNVEGNFFGNAGKDLKILQPIMKKAGGKSTISKLVETDWWYQGQINTLYGADLNITYNYDVHEFYFANSLTAPRL
SDEAICAFVDYKFDNSSVRFGWVOWDWFHGGKNSALAAVSNDEATAYAHRDQLWLWOFYDSYDYENNTSPYSESGFEFMGGFV
AETIETLDPEDRKGKGFYNADTTLTKEEAQKLWYRGNLEKLQAIKYQPEDVFGNVVSVPEIA

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Fig. 9

*(Agaricus bisporus -- Pyranose Dehydrogenase)*Nucleotide sequence (SEQ ID NO:13)

ATGATACCTCGAGTGGCCAAATTCAACTTTGACTCTTGTCTCTGCGATTATTGGGGATTGAGGTTGCACGGCACTGCCATCACAT
 AOCAAAACCCGAOCGATTTACCTGGTGACGTTGACTATGATTTTCATCGTTGCTGGGGGTGGAACTGCAGGTTTAGTTGTGGCCT
 CTGCTCTCAGTGAGAATCCGGAATGGAATGTACTGGTTCATCGAGGCCGGGGCCTTCCAAACAAGGAOCGTCTTCGAAAACAGGGGTG
 OCTGGCCTTTCTTCGGAATCCGGCCACGTTTTGATTGGAATTATACAAAGATTCCTCAAGATGCTCTCGGTGGCAGGAGCCTG
 AATTACTCGAGGGCGAAGCTCTTAGGGCGTTGCAGTAGCCATAATGGGATGGTTACACAOCGATGTTTCGAGAGACGATTGGGAC
 AATTATGCCGAAATCAACGGTAATCAAGCATTAGCTGGGACAGGATCCTACCTGTCATGAAGAGGGCTGAGAAATTCAGTAAAG
 ATTCCCTCTCATAAACCGGTAAAGGGCCATATTGACCCCTCCGTGCACGGTGGTGACGGGAAATTTGTCGGTGGTGGCATCATACA
 CCAAGGCTCTTTCAATGACTTATTACTTGAACCGCCAAAGAATTAAAGCGGTGAATTTCCGTTCAAATGGATATGAATGACGG
 GCGGGCCTCTTGGATTAACTTGGACTCAGTATACGATTGATCAACGCCGGGAGCGGAGCAGCTCTGCAACAGCGTATTAGAGG
 GTACTGGAAATAACGTCATGCTCTTGGTTAACTCTTGTACCGGTATAGTCTCAGCAGAAATGGGACCGACTTCGAAAGCGT
 CGAGTTTGGTACTGATGCGGACAGCCCAAGATCCAATTACGAGCGGAAAGGAAGTCATTGTATCTGGAGGAGTCATCAATTC
 GCCTCAGATCCTCATGAATTCGGGCATTGGGGGCCGAGAGGTTGCTTGGAGCTAATGGAATTGACACATTGGTGGATAATCCGAG
 TGTGGGAAAAATTTATCGGACCGAGGCTGCAAGATTATAATGCTCGATACAAACACTCCCTATTACTGATTATGATGTTGATGCGAG
 CGCTTATTGAATGGAAGAGTGGCAGCTGGACCTCTAGCCCAAGGAGGTGGCTAAACACCTTACATGGGTAGGATTGCTG
 ATGACAAGCTGGATGGACTTGATCCCTCAAGTGCGGAAAAATTGCGCCACATATTGAGTTCCAAATTGGGGCAAATTAGCCACGAGCT
 CCTGCCAGTGCTCTAACACGTTTTAGCTTGTATCGACACTGTTCTCCAATTCCGCCGTTGATGAACCTCTACACTGTTTGGGG
 GGTCTGATTTCTCTCAGTAACAACGATCGGTTCTGCCAGCCAGTCATCGATCTCAACATGTTTGGAGAGGAAATAGATCCCGCTAT
 TGTGGGTGAGGATATTCCAGTGCCCCAAGAATGCTTTCTCCCAAGCATTCAAAGGCTTTGTCGGTGAACGGTGTCTTCTCTCA
 AGCGACCTAGCTCTGATGAAGATTGGATACCTTCTCAAAACGTCACGCTTTCTTACGTGCATGCTGTGGGAAACGTTGCTCTA
 TGTCTCTCAGAGTGGCTGCTGGGGTGTGCTTAAACCTGATTTCCGTGTCAAAGGAACAGTGGCTGCGGGTGTCTGACCGG
 TCTGTGATTCCATTGCTCCGGCGGGGCAOACTCAAGAACTGTTTATGCAATTGCTGAGGATGCAAGTGTGTTAATAGCGAAGA
 GGTACAGCTAA

AA sequence (SEQ ID NO:14)

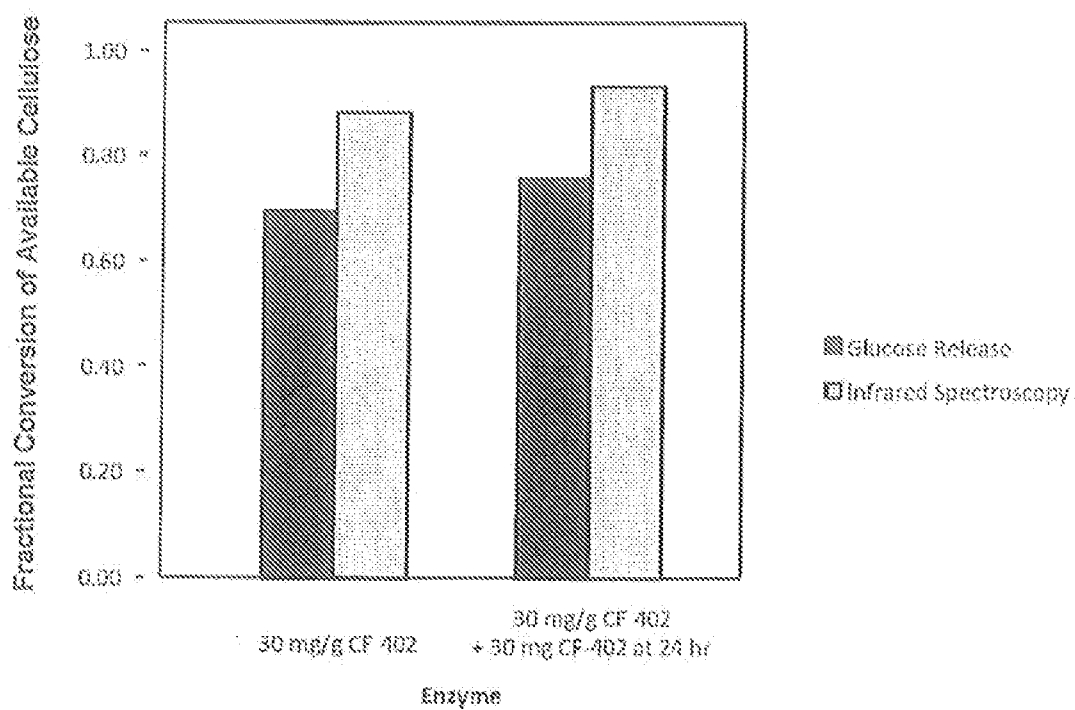
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 KGHIDPSVHGGDGKLSVVASYNASFNLLLETAKELSGEFPFKLDMNDGRPLGLTWTQYTDQGRERSSSATAYLEGTGNNVHLV
 NTLVTRIVSAENGDFRSVEFATDADSPKICLRKKEVIVSGGVINSPOILMNSGIGGREVLGANGIDTLVDNPSVSKNLSDOAATIMLD
 TTLPTDQVDVDAALIEWIKKSHTGPLACGGRLNHLTWVRLPDDKLDGLDPSSGGENSPHIEFGGCHSHQLPPSGLTRFSFYRHGSPIPPLI
 NLYTVSRGGSLSNNDPFSHPLIDLNMFGEEIDPAILREGIRSAARMILSSQAFKQFVGETVFPSPDATSDLDLDFLKTSTFSYVHGVGT
 LSMSPQSAWGVVNPDRVKGTGSLRVVDASVIFAPAGHTQEPVYAFAEHASVLIAKSYS

Fig. 10

MRLQSSIGAGLALLAALAVLAHVHALAPRTQIAEEYDFVVGSSQAGLVIGARLSEIANYTVLVLEAGTNGDEFRERIGTYNFYTPAYS
YVESLWTTIPMNWAYTYVPSQSHAGNPDIEWTROKGLGSSAINGLYLTRPGKEEINAWKDLLGDMDGADNWSWDSFYAAMKKSETP
TPPSSNATEGNTWDLSTRGHIQGPQIATYPGYTFQVGEWMSLEAMGIASSNDMAGYGGVEYSGAEVTS76SINPTNWTYSRSTGYLD
PLADNGNYDVVADAFVTRILFDASSPSNLTANGVQYTLDNQKTNCTYVGVKKEVILSAGTGVSPAVLHSGVGPKDVLSDAGVELYSE
LPGVGHHLGDHFNNTLYLSYIDSAIAYINSTLMYQDNLDALCKNITTOINOFVLNTTYDAGVIAGYKAJANMTATTILSSSIGGIELFMNS
DLNGDIQITAAALGPYSHGRIYINSSNPLDYPVIDPNVYLASADYEILRGLNLARQLGNTGPLSSCUAETIPGPSVKTDDDWLEWIREA
TOTEFHPSSGACMLPREQQGVVDANLRYVGLANVRVADASYRISLSTHLMASTYGVAEQASNIIRAHYTDGRTTGTSSSDPQGSASSP
TSSALCAEGTGTGAISHTAPSGGVRSGVAWSAWVAVVFAAAVSPHISHLH

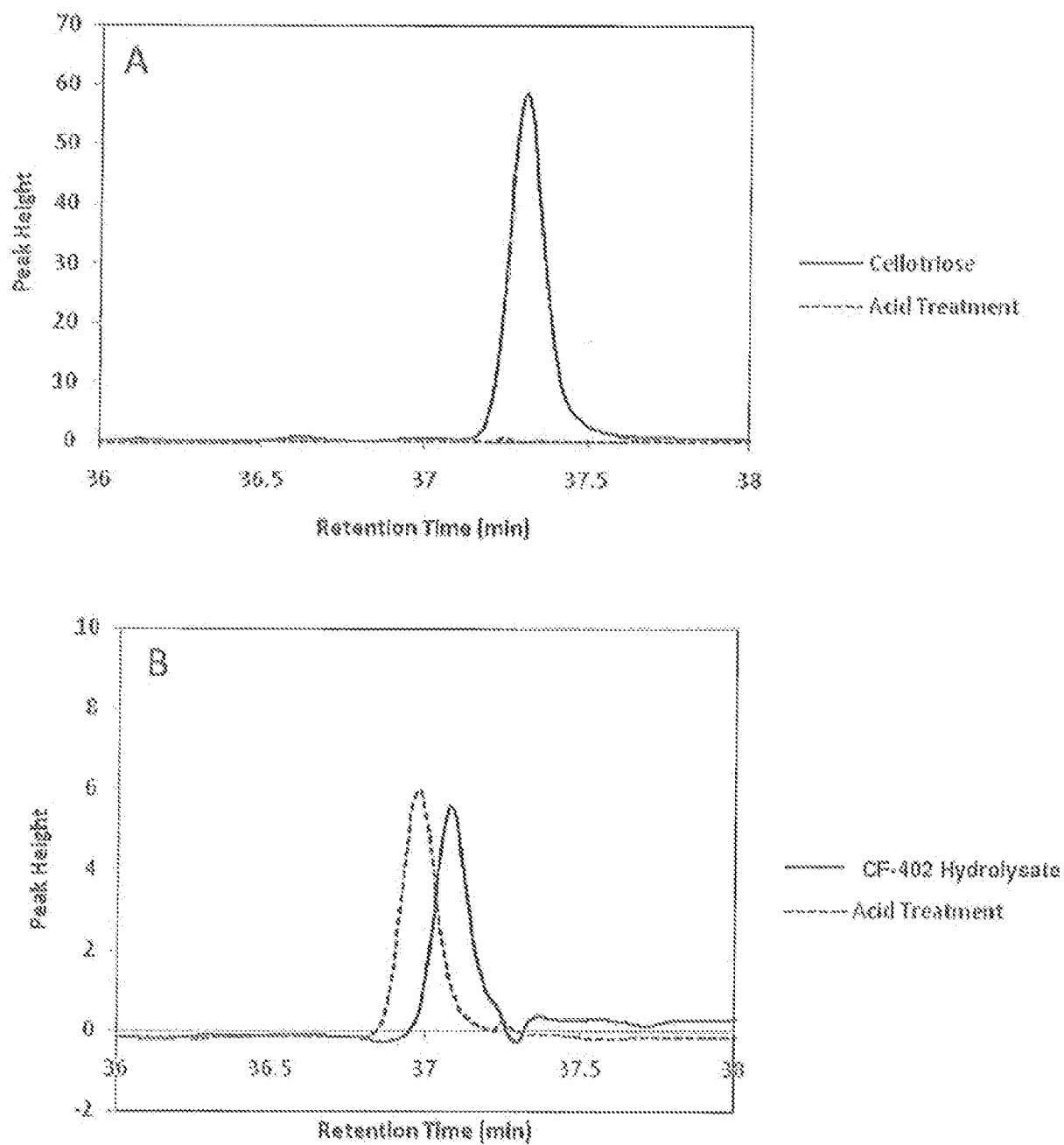
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Fig. 11



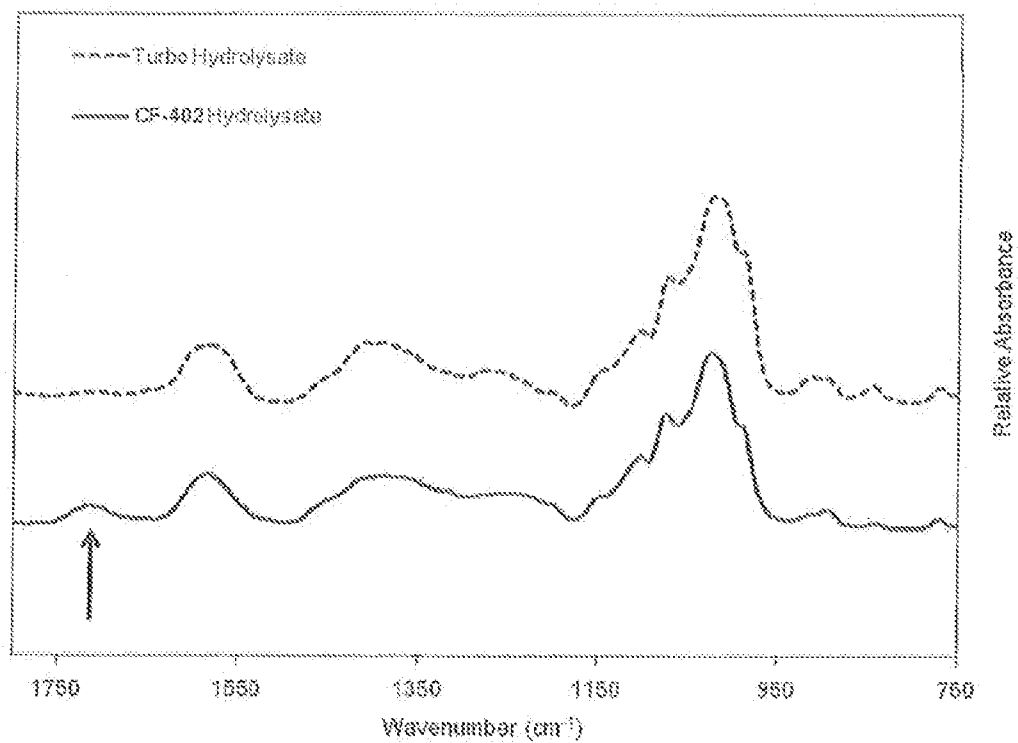
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Fig. 12



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Fig. 13



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Fig. 14

