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(54) **Title:** COMPOSITIONS FOR GROWTH AND VIGOUR IN ORNAMENTALS

(57) **Abstract:** Coating composition for applying to plant structures of ornamental plants such as a seed of ornamental plants from which roots and shoots are capable of growing, wherein the said coating composition comprises organic material in particulate form having a melting point of > 50°Centigrade, methods of making such compositions and coated ornamental plant structures.



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COMPOSITIONS FOR GROWTH AND VIGOUR IN ORNAMENTALS

The present invention relates to coating compositions including an organic component for applying to ornamental plant structures from which roots and shoots are capable of growing, uses of coating compositions on ornamental plant structures, methods of producing such coating compositions and ornamental plant structures coated with such coating compositions. In particular, the invention relates to coating compositions that comprise an organic material that provides protection from environmental stresses to ornamental plant structures.

Young ornamental plants grown from ornamental plant structures are vulnerable to abiotic and environmental stresses, particularly in growing habitats that have low rainfall and/or sub-optimal soil quality. Losses due to sub-optimal soil quality are typically realised in the growth of young plants lacking plant vigour in which the plants do not become well established, such as where the rooting systems do not develop and in circumstances where essential elements in the soil are not readily available. Agronomic losses due to young ornamental plants not being well established remain unacceptably high on soils which are for example mineral deficient despite the employment of conventional inorganic ornamental structure coatings such as ornamental seed coatings that typically include essential elements for establishing young seedlings. A problem with the use of such conventional coatings is that they introduce nutrients to the soil in unbalanced quantities and this can have adverse effects on plant growth and vigour in unforeseen ways. Additionally, such conventional coatings are typically applied in the form of wet slurries to ornamental plant structures, such as seeds. Once applied, the coatings are typically dried on the ornamental plant structure and this drying may cause further abiotic stresses, which in turn may have deleterious consequences on the viability of young plants grown therefrom. Additionally, such conventionally applied coatings may not be applied to ornamental plant structures evenly, and as a consequence, such coatings tend to be susceptible to chipping and/or flaking. Furthermore, the degree of coating uniformity of such conventionally applied coatings typically is not optimal, with a percentage of ornamental plant structures of any one batch receiving little or no coating depending on the coating method being deployed.

In the following description, the terms "ornamental plant structure treatment" and "ornamental plant structure coating" are used interchangeably for the compositions of the invention and their uses to treat ornamental plant structures, by any of the specific methods described in the prior art that provide an improvement, typically an enhancement, of seedling vigour. The commonly used ingredients in ornamental plant structure treatment such as ornamental seed treatment compositions (sometimes designated as formulations) include antidotes and safeners; fertilisers, micronutrients and inoculants; bioregulators of natural or

synthetic origin which are either hormones or interfere in hormone metabolism and do not influence plant nutrition; and/or bioregulators which interfere with plant growth by enhancing nutrient uptake.

It has now been found that by using certain organic materials as components of coatings on ornamental plant structures, such as ornamental seeds, together with the application of inorganic components and/or biological agents, plant vigour and plant growth of plant seedlings grown from ornamental plant structures is improved relative to the plant vigour and plant growth of seedlings grown from conventionally treated ornamental plant structures, such as seeds. It has further been found that the quantity of additives, particularly inorganic fertilisers that is required per unit of ornamental plant structure weight, such as ornamental plant seed weight is less than that required using conventional formulations that are employed in current farming techniques.

It is an object of the present invention to supply improved ornamental plant structure coatings, for ornamental plant structures such as ornamental plant seeds comprising organic components.

It is a further object of the present invention to provide improved ornamental plant structure coatings, such as ornamental seed coatings comprising a minimum amount of additives.

These and other objects of the invention will become apparent from the following description and examples.

According to the present invention there is provided an ornamental plant structure coating composition that comprises

- i) at least one organic material selected from waxes having a melting point of $\geq 50^{\circ}\text{C}$ entigrade; and
- ii) at least one additive for enhancing seedling vigour and/or seedling growth from ornamental plant structures wherein the at least one additive is selected from one or more inorganic additives and/or one or more live biological agents.

The organic wax materials of use in the invention act as a carrier for desired additives for placing on or near to seeds.

Commercial growers of ornamental plants require ornamental plant structures for the provision of new ornamental plants for building up seed and energy storage organ stocks for harvest and eventual sale to outlets such as commercial growers and from there to garden centres and the like. For the purposes of the present invention "ornamental plant structures" are viable plant structures to which compositions of the invention may be applied

and include seeds, seed tubers, tuberous roots (sometimes referred to as 'tubers', e.g. dahlia 'tubers'), bulbs, corms and rhizomes. Viable ornamental plant structures as provided herein means that the plant structures are capable of germinating to conventional levels of germination typical of ornamental plant seeds or in the case of seed tubers, tuberous roots, bulbs, corms and rhizomes of growing roots and shoots. Thus, viable plant structures as herein defined may be used for the planting of ornamentals such as varieties of tulip, amaryllis, hyacinth, daffodil, narcissus, cyclamen, lily, lily of the valley, iris, gladiolus, crocus, crocosmia, dahlia, snowdrop, bluebell, dahlia, freesia, gloxinia, anemone, fritillaria, alstromeria ligu and hybrids thereof, camassia esculenta, arum italicum, muscari, agapanthus, begonia, acidanthera, ranunculus, ornamental allium and the like. Examples of ornamentals seeds that may be coated with compositions of the invention include seeds of viola (pansy), primula, petunia, polyanthus, tagetes, pelargoniums including *P. Peltatum*, begonia, cyclamen, achillea, ageratum, agrostemma, alyssum, amaranthus, antirrhinum, aquilegia, aster, calendula, campanula, carnation, chrysanthemum, helleborus, cineraria, clematis, convolvulus, Centaurea cyanus, cosmea, dahlia, delphinium, dianthus, digitalis, myosotis, freesia, geranium, godetia, impatiens, cheiranthus cheiri, dianthus barbatus, lathyrus odoratus, salvia, salpiglossis, verbena, zinnia and the like. The online Sutton Seeds catalogue 2010 herein incorporated by reference provides examples of many varieties of ornamental plant which may be treated with the coating compositions of the present invention. Woody ornamental plant structures may also be treated with compositions of the invention. Woody ornamental plant structures include seeds of ornamental plants such as members of the arborvitae, acer, azalea, Chamaecyparis, dogwood, euonymus, rose, forsythia, Fraser fir, hemlock, Japanese holly, juniper, Pieris, rhododendron, Taxus, white pine, maple, elm, aspen, ash, beech, and oak.

Reference to "seed" and "seeds" is used interchangeably herein and means seeds, typically viable seeds, to which compositions of the invention may be applied.

The organic material used in the present invention is selected from organic materials that can be applied to ornamental plant structures either as a powder wherein the powder particles are of a pre-determined volume mean diameter (VMD) or the powder particles are applied in liquid form, such as an oleaginous formulation or as an aqueous formulation.

Generally, the particles of use in coating compositions of the invention possess a volume mean diameter of a certain size as defined herein. To obtain particles of organic materials of a volume mean diameter applicable for use in the invention, organic materials in the form of, for example, 1 to 5 kilogram blocks or tablets may be broken up or kibbled into small millimetre-sized pieces (such as from 2mm – 8mm approximate diameter in size, for example from 4mm to 6mm) in a kibbling machine. The millimetre-sized pieces can then be

passed through a comminuting means such as a standard mill, e.g. an Apex Comminuting mill, and milled or comminuted into particles having an approximate diameter in the range from 100 μ m - 500 μ m, for example from 250 μ m - 300 μ m. The micron-sized comminuted particles can then be passed through a micronising apparatus, such as an AFG micronising air mill to obtain particles of a desired VMD range, such as from 15 μ m - 20 μ m, that is of use in the present invention. The skilled addressee will appreciate that such procedures for obtaining small particles are well known in the art. Preferably, dry powder compositions of the invention comprise composite particles having a volume mean diameter of $\geq 10\mu$ m, for example of 10 μ m, 11 μ m, 12 μ m, 13 μ m, 14 μ m, 15 μ m up to 40 μ m or any value therein between. As stated herein, the volume mean diameter of the composite particles is typically $\geq 10\mu$ m or $\geq 12\mu$ m and may lie in the range from 10 μ m to 200 μ m and may have a value that lies anywhere therein between, for example from $\geq 10\mu$ m to 100 μ m; or from $\geq 10\mu$ m to 40 μ m; or from $\geq 10\mu$ m to 30 μ m or any desired volume mean diameter value in between. Preferably, dry powder compositions of the invention comprise particles having a volume mean diameter of $\geq 10\mu$ m, for example of 10 μ m, 11 μ m, 12 μ m, 13 μ m, 14 μ m, 15 μ m and the like up to any volume mean diameter of choice, such as up to 200 μ m or any volume mean diameter in between for example 40 μ m or 30 μ m. Such compositions are considered to be less of a thoracic hazard to humans and are not thought to be allergenic.

In liquid formulations, particles of a pre-determined volume mean diameter are suspended therein forming a suspension formulation and applied to ornamental plant structures, which are then dried using conventional drying procedures. Preferably, the organic material is applied to ornamental plant structures in a dry powder form, the particles of the organic material may have a volume mean diameter of any conventional size as herein described. Such organic materials include additives as herein defined and may include added further components such as added UV blockers or added antioxidants or the like. Dry powders of the present invention may be made up of one or more organic materials that have a melting point at or above 50°C and which are of use in the present invention. Suitable organic materials of use in the invention include waxes selected from natural, synthetic and mineral waxes. Suitable waxes of use in the invention include waxes having a melting point of preferably $\geq 50^\circ\text{C}$, more preferably of $\geq 60^\circ\text{C}$, and most preferably are made up of hard waxes having a melting point of $\geq 70^\circ\text{C}$. In a preferment, the organic material is selected from carnauba wax, beeswax, montan wax, Chinese wax, shellac wax, spermaceti wax, myricyl palmitate, cetyl palmitate, candelilla wax, castor wax, ouricury wax, and rice bran wax; or a mixture of two or more thereof.

Synthetic waxes of use in the present invention include suitable waxes selected from paraffin wax, microcrystalline wax, Polyethylene waxes, Fischer-Tropsch waxes, substituted amide waxes, polymerized α -olefins and the like.

Mineral waxes of use in the invention include montan wax (e.g. Lumax® Bayer) ceresin wax, ozocerite, peat wax and the like.

Such waxes typically display a high enthalpy of lattice energy during melt. Preferably the organic material is carnauba wax which may be applied in liquid form, typically in the form of a liquid suspension or powder form as discrete particles. Preferably, the organic material is applied in dry powder form to ornamental plant structures. The skilled addressee will appreciate that the actual VMD of particles of use in the invention that are used on ornamental plant structures will be appropriate to the size of the structures to which the particles are to be applied. Furthermore, the skilled addressee will also appreciate that where the VMD is defined as being $\geq 10\mu\text{m}$ or $\geq 12\mu\text{m}$ the size of the particles will be governed by the size of the seed to which it is applied and such a range should be construed as being commensurate therewith. Thus, the size range of particles of use in the invention is not open-ended in respect of an upper size limit but only insofar as such a limit is applicable to ornamental plant structures, such as seed to which particles of the invention may be expected to adhere as a coating. The limit in the sizing of the particles of use on ornamental plant structures such as seed coatings of the invention will be apparent to the skilled addressee.

The one or more additives for enhancing seedling vigour and/or seedling growth from ornamental plant structures may be selected from one or more inorganic or chemical additives and/or one or more live biological agents.

Suitable inorganic agents include commercially available NPK fertilisers that may be added to ornamental plant structure coatings of the invention. These may be added in the form of dry powders of soluble ions that include the so-called primary macronutrients such as nitrogen, phosphorus, and potassium; the so-called secondary macronutrients such as calcium, sulphur, and magnesium; and the so-called "micronutrients" (trace minerals such as boron, chlorine, manganese, iron, zinc, copper, molybdenum, and selenium). "Macronutrients" are taken up in relatively large quantities and are present in plant tissue in quantities from about 0.2% - 4% on a dry weight basis. "Micronutrients" are taken up in smaller quantities and are present in plant tissue in quantities measured in parts per million (ppm), ranging from about 5 – 200 ppm, or less than 0.02% dry weight.

Additives may be selected from bioregulators commonly applied in the art such as brassinosteroids, cytokinines e.g. kinetin or zeatin, the auxins e.g. indolylacetic acid or indolylacetyl aspartate, the flavonoids and isoflavanoids e.g. formononetin or diosmetin, the phytoalexins e.g. glyceollin, phytoalexin-inducing oligosaccharides such as pectin, chitin, chitosan, polygalacturonic acid and oligogalacturonic acid, compounds such as the

gibellerins produced by rhizobial symbionts and endophytic microorganisms such as *acetobacter diazotrophicus* and *herbaspirillum seropedicae* and the like.

Species of bacteria and fungi of use could be those that are capable of acting on an inorganic and/or organic substrate to release phosphorus-containing compounds in soluble form from such substrates.

Species of bacteria include those from *Alcaligenes*, *Acinetobacter*, *Azospirillum*, *Bacillus*, *Enterobacter*, *Erwinia*, *Flavobacterium*, *Paenibacillus*, *Pseudomonas*, *Rhizobium*, *Burkholderia*, and *Serratia*. Examples of species of the *Bacillus* genus are *Bacillus megaterium*, *Bacillus coagulans*, species of the *Azospirillum* genus such as *Azospirillum brasilense*, species of the *Pseudomonas* genus, such as *Pseudomonas aeruginosa*, *Pseudomonas aurantiaca*, *Pseudomonas putida*, *Pseudomonas pseudoalcaligenes*, *Pseudomonas fluorescens*, *Pseudomonas poae*, and *Pseudomonas trivialis*, species of the *Rhizobium* genus such as *Bradyrhizobium* and *Rhizobium leguminosarum*, and species of the *Paenibacillus* genus (formerly considered as *Bacillus* genus) such as *Paenibacillus lautus*. Commonly used *Rhizobium* inoculants may be sourced from such companies as Becker Underwood and EMD Crop Bioscience.

Species of mycorrhizal fungus are also capable of augmenting levels of available nutrients in the soil with further organic and inorganic nutrients that are assimilable by a crop plant. Suitable species of mycorrhizal fungus include those that are capable of colonising a host plant's roots, either intracellularly as in arbuscular mycorrhizal fungi (AMF), or extracellularly as in ectomycorrhizal (EcM) or ericoid mycorrhizal (EM) fungi.

AMF mycorrhizae include *Glomus*, *Gigaspora*, *Acaulospora* and *Sclerocystis*. Suitable species include *Glomus fasciculatum*, *G. intraradices*, *G. claroideum*; *G. intra*, *G. clarum*, *G. brasilianum*, *G. deserticola*, *G. monosporus*, *G. mosseae*, *G. tortuosum*, *G. sinuosum*, *Gigaspora margarita*, *Gigaspora gigantea* and *Acaulospora longula*.

Ectomycorrhizae (EcM) are typically formed between the roots of around 10% of plant families, mostly woody plants including the birch, dipterocarp, eucalyptus, oak, pine, and rose families, and fungi belonging to the genera Basidiomycota, Ascomycota, and Zygomycota. Nutrients move between different plants through the fungal network (sometimes called 'the wood wide web'). Genera of EcM of potential use on woody plants include *Suillus*, *Boletus*, *Lactarius*, *Laccaria*, *Pisolithus* and *Rhizopogon*. Examples include *Pisolithus tictorus*, *Laccaria laccata*, *L. bicolor*, *Rhizopogon villosuli*, *R. rubescens*, *R. fulvigleba*, *R. luteolus*, and *R. amylopogon*.

Ericoid mycorrhizas (EM) are the third of the three more ecologically important types of

mycorrhizal fungus. Ericoid mycorrhizae are known to have saprotrophic capabilities and these are thought to enable plants to receive nutrients from not-yet-decomposed materials via the decomposing actions of their ericoid partners. A suitable genus of EM of potential use is *Pezizella*.

A further live biological inoculant that may be useful for ornamental plant structure coating is *Trichoderma*, a fungus that is capable of making available, and in the adsorption of, mineral nutrients from the soil such as by solubilising insoluble phosphorus and zinc in the soil. Other capabilities of the fungus include the decomposition of organic matter thereby releasing calcium, potassium, and nitrogen available for plant use. By such capabilities certain *Trichoderma* species can be used to contribute to a balanced fertilisation of ornamental plants in the field and thereby the requirement for adding large amounts of artificial fertilisers may be reduced by as much as 50% depending on crop type. *Trichoderma* strains are known in the art, for example, useful strains are known from the University of the Philippines Los Baños (UPLB), Institute of Biological Sciences.

Examples of conventional additives for increasing fertiliser efficiency, plant productivity, growth, and nutrient accumulation may be sourced from such commercial sources as Incotec Inc., Germains, Bayer CropScience, and Becker Underwood. Suitable additives may be selected from commercially available products such as Auxigrow(R) (Auxein Corp., Lansing, Mich., USA) and Amisorb(R) (Donlar Corp., Chicago) or the so-called phytochelates described by A. M. Kinnersley in *Plant Growth Regul.* (1993), 12(3), 207-18, which are thought to influence the availability to the plant of minimal amounts of certain metals such as Zn, Fe, Cu and the like for optimal growth and productivity. Examples of the latter include polymers of L-lactic acid, L-lactoyllactic acid and water-soluble polyaspartates. Other additives that may be applied to ornamental plant structure coatings, such as ornamental seed coatings of the invention include the kinds of adjuvant that are found in conventional commercial agrochemical formulations. Suitable additives for inclusion into ornamental plant structure coatings of the invention may be selected from those described by Chester L. Foy, *Pestic. Sci.*(1993) 38, pp.65-76; and in EP 0357559. Seed coating compositions of the invention may further include conventional additives such as agents having wetting, dispersing and de-foaming modes of action. Suitable surface-active compounds are non-ionic, cationic and/or anionic surfactants having good emulsifying, dispersing and wetting properties. Such adjuvants for crop protection formulations are obtainable from fine chemicals producers [e.g. by Clariant AG (Muttenz, Switzerland)] and include (fatty)alcohol alkylphenol ethoxylates, polyarylphenol ethoxylates, dispersing phosphates, taurides and/or alcohol monosuccinates. The term "surfactants" also comprises mixtures of two or more surfactants and natural or synthetic phospholipids of the cephatin

and lecithin series, e.g. phosphatidyl-ethanolamine, phosphatidylserine, phosphatidylglycerol, lysolecithin sugar esters. A typical de-foaming agent is Fluowet PL80B(R) (Clariant AG) and typical antifreeze compounds are glycols and polyethylene glycols. Further ingredients may include solid or liquid substances ordinarily employed in formulation technology, e.g. natural or regenerated minerals, tackifiers, thickeners or binders. Other suitable additives are emulgating protein hydrolysates, e.g. as described in EP 0297426 (Bayer AG). Dyes that may be used in ornamental seed treatment compositions include water-insoluble or water-soluble dyes. Examples of dyes that may be added to compositions of the invention include Colanyl Red(R) (Clariant AG, Muttentz), Rhodamin B, white pigment (titanium dioxide) or Luconyl(R) (BASF AG). Altogether additives may be used to ensure that the formulation disperses well, does not settle or freeze and differentiates the seeds from untreated seeds. Other special additives which are known to enhance seedling vigour in particular in combination with certain pesticides, e.g. fungicides in combination with 3',4',5',6'-tetrachloro-2,4,5,7-tetraiodo-fluorescein (EP0297426), may be applied to the seeds in a combined amount that is effective, preferably synergistically effective, to increase seedling vigour and plant growth.

Additionally, the organic particles of use in compositions of the invention may contain other further components such as additives selected from UV blockers such as beta-carotene or p-amino benzoic acid, colouring agents such as optical brighteners and commercially available colouring agents such as food colouring agents, plasticisers such as glycerine or soy oil, antimicrobials such as potassium sorbate, nitrates, nitrites, propylene oxide and the like, antioxidants such as vitamin E, butylated hydroxyl anisole (BHA), butylated hydroxytoluene (BHT), and other antioxidants that may be present, or mixtures thereof. The skilled addressee will appreciate that the selection of such commonly included additives will be made depending on end purpose, and perceived need.

Ornamental plant structure compositions, such as seed compositions of the invention may be applied to ornamental plant structures or plant seed at a rate of application from 0.1 g to 500 g, preferably from 1g to 100g, most preferably from 5g to 50g of the active ingredient (a.i.) per 100kg of seed.

Liquid formulations of the invention may be formulated as an aqueous formulation or as an oleaginous formulation, depending on design. Aqueous formulations may include surfactants selected from commercially available surfactants such as Tween 20, Silwet L77, Tween 80, Torpedo II, Newmans T80, Fortune, Guard, Rhino, Biopower, and the like.

Oleaginous formulations, that is to say oil-based formulations, may contain any oil suitable for use in the present invention which may be selected from petroleum oils, such as paraffin

oil, and vegetable oils such as rapeseed oil, soybean oil, sunflower oil, palm oil and the like. Oil formulations of use in the invention contain organic particles of the invention and as described herein and these in turn may be admixed with flow agents such as hydrophilic precipitated silicas, for example Sipernat 383 DS, Sipernat 320, EXP 4350, and Sipernat D-17 and the like. Such free-flowing agents may be dispersed in oils, for example, for anti-foaming purposes.

The skilled addressee will appreciate that where an aqueous or an oil formulation may be used, the liquid element should be removed from the coated ornamental plant structures such as ornamental plant seeds after coating is achieved, for example by drying off using conventional drying processes.

Coatings of organic materials of use in the present invention also serve to protect immediately planted ornamental plant structures from soil borne pathogens, that is to say, pathogens that are able to colonise the ornamental plant structure, such as the seed cuticle and/or pathogens that populate the soil and which are capable of acting on ornamental plant structures, such as seeds. Such soil borne pathogens are typically bacteria and/or fungi. Examples of soil borne bacterial and fungal pathogens that attack ornamental plants include species of *Rhizoctonia solani* (e.g. active against verbena; anemone; dahlia; impatiens; primula; poinsettia), *Chalara elegans* (e.g. active against verbena; cyclamen), *Cylindrocarpon destructans* (e.g. active against cyclamen), *Cylindrocladiella peruviana* (e.g. active against cyclamen), *Erwinia chrysanthemi* (e.g. active against *Euphorbia pulcherrima* (poinsettia)), *Pythium spp* (e.g. active against impatiens; primula), *Agrobacterium tumefaciens* (e.g. active against anemone; carnation; dahlia; geraniums (pelargonium); primula; poinsettia), *Phytophthora spp.* (e.g. active against impatiens; geraniums (pelargonium)); species such as *phytophthora tentaculata* (e.g. active against Chrysanthemum, Verbena, and Delphinium species, *Sclerotinia sclerotiorum*, *Fusarium spp.*, and *Verticillium spp.*, such as *Verticillium albo-atrum* (e.g. active against woody ornamentals such as maple, elm, aspen, ash, beech, and oak; chrysanthemum) and the like.

According to a further aspect of the invention there is provided use of an organic material in the form of particles wherein the particles are selected from at least one wax having a melting point of $\geq 50^{\circ}\text{C}$ Centigrade in the manufacture of an ornamental plant structure coating composition as defined herein. The organic materials are selected from one or more organic materials having a melting point of $\geq 50^{\circ}\text{C}$ Centigrade, more preferably of $\geq 60^{\circ}\text{C}$ and most preferably are made up of hard waxes having a melting point of $\geq 70^{\circ}\text{C}$. Suitable organic materials include carnauba wax, beeswax, montan wax, Chinese wax, shellac wax, spermaceti wax, candelilla wax, castor wax, ouricury wax, and rice bran wax or a mixture of two or more thereof, and preferably, the seed coating that is used includes carnauba wax.

Preferably, in this aspect of the invention, the organic particles have a mean volume diameter $\geq 10\mu\text{m}$, more preferably in the range selected from $\geq 10\mu\text{m}$ to $200\mu\text{m}$, as herein described and as appropriate for the ornamental plant structure to which the particles are to be applied. Naturally, the skilled addressee will appreciate that the size of the organic particles to be applied to ornamental plant structures, such as ornamental plant seeds will depend on the size of the structure or seed, and the type or form of such ornamental plant structures that are contemplated for coating.

In a third aspect of the invention there is provided a method of manufacturing an ornamental plant structure coating composition as herein described that comprises

- 1) selecting solid organic material wherein the solid organic material is wax having a melting point of $\geq 50^\circ\text{C}$;
- 2) machining said organic material into particles of a mean volume diameter of $\geq 10\mu\text{m}$; and
- 3) adding one or more additives for enhancing seedling vigour and/or seedling growth to the machined organic material of 2), wherein the said additives are selected from one or more inorganic additives and/or one or more live biological agents.

The organic material in this aspect of the invention may be selected from organic materials such as from organic waxes having a melting point of $\geq 50^\circ\text{C}$, more preferably of $\geq 60^\circ\text{C}$, and most preferably are made up of hard waxes having a melting point of $\geq 70^\circ\text{C}$. Suitable waxes for use in the invention include carnauba wax, beeswax, montan wax, Chinese wax, shellac wax, spermaceti wax, myricyl palmitate, cetyl palmitate, candelilla wax, castor wax, ouricury wax, and rice bran wax; or a mixture of two or more thereof. Preferably, the selected organic material includes a substantial proportion of carnauba wax up to 100%, for example 1%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% or more or any proportion therein between, the rest being made up of is at least one other organic material as herein defined. Preferably, the selected organic material is solely carnauba wax which may contain further additives as herein described, and further components such as UV blockers, antioxidants such as vitamin E and the like.

In a further aspect of the invention, there is provided an ornamental plant structure coating composition produced by the method as described herein.

In a further aspect of the invention there is provided a coating composition as described herein for use on ornamental plant structures.

In a further aspect of the invention there is provided a method of coating ornamental plant structures with a coating composition that comprises at least one organic material wherein

the organic material is selected from waxes having a melting point of $\geq 50^{\circ}\text{Centigrade}$ as herein defined, the method comprising

- i) obtaining a population of particles of wax of a pre-determined VMD; and
- ii) applying the said population of particles to ornamental plant structures, such as ornamental plant seeds. Preferably, the coating composition of the invention comprises a particulate composition that is applied to ornamental plant structures in dry particulate form.

The skilled addressee will appreciate that environmental factors that may affect ornamental plant structure viability includes such factors as extremes of heat, loss of moisture and the presence of pathogens such as bacteria and/or fungi. The skilled addressee will also appreciate that the pre-determined VMD will be appropriate to the size of the ornamental plant structures to which the coating is to be applied.

In a variant of this aspect of the invention there is provided a method of coating ornamental plant structures with a coating composition that comprises an organic material that is selected from waxes having a melting point of $\geq 50^{\circ}\text{Centigrade}$, the method comprising

- i) obtaining said organic material;
- ii) heating the organic material of i) so as to form a liquid phase or a gaseous phase;
- iii) cooling the liquid phase or gaseous phase of ii) to below the melting point of the organic material, forming a solid;
- iv) adding one or more additives to the solid formed in iii);
- v) machining the solid organic material of iii) into particles of a pre-determined VMD; and
- vi) applying the particles of v) to ornamental plant structures.

In a second variant of the above aspect of the invention there is provided a method of coating ornamental plant structures with a coating composition of the invention that comprises an organic material wherein the organic material is selected from waxes having a melting point of $\geq 50^{\circ}\text{Centigrade}$, the method comprising

- i) obtaining said organic material;
- ii) heating the organic material so as to form a liquid phase or a gaseous phase;
- iii) adding one or more additives to the liquid phase or gaseous phase of ii);
- iv) cooling the liquid phase or gaseous phase of iii) to below the melting point of the

organic material, forming a solid;

v) machining the solid organic material of iv) into particles of a pre-determined VMD as herein defined; and

vi) applying the particles of v) to ornamental plant structures.

The ornamental plant structures are typically selected from ornamental plants as herein defined. The organic material of use in the invention may comprise one or more organic materials selected from organic materials as herein defined. Preferably, the organic material is carnauba wax. Where two or more organic materials of use in the invention are employed as the organic material in for example, an ornamental plant seed coating composition of the invention they may be heated together so as to form a liquid phase or a gaseous phase during which phases the organic material may be mixed, if required. Once the organic materials are mixed they may be cooled to below the melting point of the organic material possessing the lowest melting point in the liquid phase (where a gas phase is employed, this will be cooled to a liquid phase), forming a solid which may then be machined, such as by comminution, into particles of a pre-determined VMD as herein defined using conventional procedures. As described above, one or more additives may be added to the organic materials at points indicated above. It will be appreciated that the person skilled in the art will understand at what point or points in the described processes additives may be added to the organic material, depending on the additive material to be added to the organic material forming particles of use in the invention.

Once the organic material is in the form of particles of a known VMD, the particles may be applied to ornamental plant structures using conventional means as employed in the art.

The treatment composition is applied to ornamental plant structures in dry particulate form or liquid form as hereinbefore described, preferably in dry particulate form. The organic material in the above aspect and variant aspect of the invention may be selected from organic materials selected from organic waxes having a melting point of $\geq 50^{\circ}\text{C}$, more preferably of $\geq 60^{\circ}\text{C}$, and most preferably are made up of hard waxes having a melting point of $\geq 70^{\circ}\text{C}$. Suitable waxes of use in the invention are those as described herein before. Preferably, the selected organic material includes a substantial proportion of carnauba wax up to 100%, for example 1%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% or more or any proportion therein between, the rest being made up of at least one other organic material as herein defined. Preferably, the selected organic material is solely carnauba wax which may contain further added components as herein defined, such as UV blockers, antioxidants such as vitamin E and the like.

Generally, the particles of use in the above aspect of the invention and the variant aspect of the invention possess a volume mean diameter of $\geq 10\mu\text{m}$, such as $\geq 12\mu\text{m}$ such as in the range of from $\geq 10\mu\text{m}$ to $200\mu\text{m}$, for example from $\geq 10\mu\text{m}$ to $100\mu\text{m}$; or from $\geq 10\mu\text{m}$ to $40\mu\text{m}$; or from $\geq 10\mu\text{m}$ to $30\mu\text{m}$ or any desired volume mean diameter value in between. Preferably, dry powder compositions of the invention comprise particles having a volume mean diameter of $\geq 10\mu\text{m}$, for example of $10\mu\text{m}$, $11\mu\text{m}$, $12\mu\text{m}$, $13\mu\text{m}$, $14\mu\text{m}$, $15\mu\text{m}$ and the like up to any volume mean diameter of choice, such as up to $200\mu\text{m}$ or any volume mean diameter in between for example $40\mu\text{m}$ or $30\mu\text{m}$. More preferably compositions of the invention comprise particles having a volume mean diameter of from about $12\mu\text{m}$ to $200\mu\text{m}$.

There now follow examples that illustrate the invention. It is to be understood that the examples are not to be construed as limiting the invention in any way.

Example 1: Growth and Vigour in Viola

Viola tricolor seed provided by Herbiseeds (Twyford, UK)

Combination of Carnauba wax particles and Inoculant

Rock Phosphate

Rock Phosphate (Garden Direct, UK) with a 30% P_2O_5 content is crushed using a pestle and mortar and then passed through a 32 micron mesh sieve.

Carnauba Wax Sizing Method

Steps in Air Milling in Boyes Micronisation Process (for carnauba wax particles with a VMD of approx. $75\mu\text{m}$)

1. 2kg carnauba wax blocks are first kibbled into approximately 4 to 6mm pieces in a KT Handling Ltd Model 04 kibbler (serial no. 729/C) following the manufacturer's instructions.
2. The kibbled pieces are then passed through an Apex Construction Ltd Model 314.2 Comminuting Mill (serial no. A21306) and reduced further in size to a range of 250 to $300\mu\text{m}$.
3. The comminuted particles are then passed through a Hosokawa Micron Ltd Alpine 100AFG jet mill (serial no. 168092) following the manufacturer's instructions, setting the mill at a speed of 2,500rpm with a positive system pressure of 0.03bar.
4. The grinding air is to be kept to 6 bar, the system rinsing air flow and Classifying Wheel gap rinsing air are both to be set at a minimum of 0.5 bar and no more than 0.75bar, the

cleaning air filter is to register a delta of no more than 5bar to achieve a final particle size with a VMD of approx. 75µm.

Rock phosphate is combined with Carnauba wax particles (VMD 75µm) at a ratio of 1:3 (Rock Phosphate:Carnauba wax particles). A homogeneous mix of is attained through tumbling seed and carnauba wax formulation in a cylinder, adapted to produce lateral mixing/tumbling through the inclusion of angled interior vanes, placed on a Wheaton roller for 5 minutes.

Dry Powder Formulation of Mycorrhizae – International Culture Collection of VA Mycorrhizal Fungi (INVAM).

Mycorrhizae concentration is measured by diluting 1gm in 1l of water, before further diluting by taking 1ml of the suspension and making it up to 1000ml. A 20µl sample is then added to an Improved Neubauer Counting Slide and a count made of 4 large squares (0.1mm³) in both of the grids. The mean for each square is calculated and the mean of the two grids used to produce a measurement of spores per 100nl of water. The dilution factor is then applied to produce an approximation of the number of spores per gram.

Carnauba wax particles (obtained using the milling protocol described above with the exception that the milling speed was set at 8000rpm to obtain a VMD of approx. 16µm) are combined with at a ratio of 1:3 (Mycorrhizae:Carnauba wax particles) in a 50ml tube using a Stuart roller mixer set at 25rpm for 5 minutes. This can then be used to calculate the quantity of spore/Carnauba wax particles powder mix required for the seed coating based on a standard of 1x10⁴ spores gram⁻¹ of seed.

A homogeneous mix of is attained through tumbling seed and carnauba wax formulation in a cylinder, adapted to produce lateral mixing/tumbling through the inclusion of angled interior vanes, placed on a Wheaton roller for 5 minutes.

Chitosan

Chitosan (>75% Deacetylated chitin, Poly(D-glucosamine)) (Sigma Aldrich,UK) is crushed using a pestle and mortar and then passed through a 32 micron mesh sieve.

This is combined with Carnauba wax particles (VMD 75µm) at a ratio of 1:19 (Chitosan:Carnauba wax particles). A homogeneous mix of is attained through tumbling seed and carnauba wax formulation in a cylinder, adapted to produce lateral mixing/tumbling through the inclusion of angled interior vanes, placed on a Wheaton roller for 5 minutes.

Treatments:

1. Carnauba wax particles and Mycorrhizae
2. Carnauba wax particles and Rock Phosphate

3. Carnauba wax particles and Chitosan
4. Mycorrhizae control
5. Rock Phosphate control
6. Chitosan control
7. Carnauba wax particle control (vehicle control)
8. Untreated Control

Seeds are planted in two 84 well plug trays using moist seed potting compost (John Innes No.2). The trays are placed in a Vitopod propagator (Greenhouse Sensations, UK) at 18°C. Moisture content (Brannan Soil Moisture Meter, Fisher Scientific, UK) and pH levels (Brannan Soil pH meter, Fisher Scientific, UK) are checked to ensure that the conditions are consistent across the tray. The order of the treatments is randomised (by row units) to reduce any unforeseen bias.

At the true leaf stage the plants are carefully transplanted from the plugs to 7cm square pots filled with a sterilised top soil. The macro-nutrient (nitrates, phosphates and potassium) content of the top soil is measured using a La Motte Model STH-4 soil testing kit and recorded. Six replicates for each treatment (48 plants) are randomly assigned to each of three propagators, and further randomised within the propagator (total = 144 plants). The propagators are set at 13.5°C, 18°C and 22.5°C.. Light is provided on a 16:8 Light:Dark cycle using a twin bulb T5 lighting array suspended 150mm above the propagator (Lightwave T5, 48w, 3300 lumens). T5 tubes (6500 Kelvin) deliver the bright blue/white light required by the plant for growth without emitting much heat which may scorch tender seedlings

Moisture content and pH levels are checked to ensure that the conditions are consistent across the propagator by measuring six random plants along a pathway (alternating between a W and Z). This is repeated for each propagator.

Plants are watered as required based on conditions to maintain consistent soil moisture content of 18% throughout all plants.

The lids of the propagators are removed at such time as required due to the plant height.

After 21 days the plants are removed from the propagators and the following measurements recorded:

Root weight (fresh)

Shoot weight (fresh)

% Mycorrhizal root colonisation (by microscopic examination)

Plant tissue is measured for macro-nutrient content using the instructions provided with a La Motte Model PT-3R Plant Tissue Test kit.

Analysis

The percentage data (root colonisation data) were arcsine transformed. The influence of the factors and their interactions are tested with a two-way analysis of variance. Where the ANOVA reveals significant effects by the factors, the differences between treatments are separated using a *post hoc* least significant difference (LSD), multiple comparison test ($p \leq 0.05$).

The influence of the factors and their interactions are tested with a 2-way ANOVA. The analysis was done for each temperature separately and with temperature as a factor. For the ANOVA with temperature as a factor, treatments were used as a sub-plot factor. Fisher's Least Significance Differences were calculated at the 5% significance level to compare treatment means. Shapiro-Wilks's test was performed to test for non-normality.

The above procedures are followed to apply rock phosphate, chitosan and *Glomus* sp. to seeds of Marigold (*Tagetes* sp.).

The above procedures are followed to apply rock phosphate, chitosan and *Glomus* sp. to seeds of Cyclamen (*Cyclamen* sp.).

The above procedures are followed to apply rock phosphate, chitosan and *Glomus* sp. to seeds of Verbena (*Verbena* sp.).

Delivery of Macronutrients using Carnauba wax particles as a seed coating on Cyclamen

Aim: to assess the potential for formulating essential macronutrients into carnauba wax particles and using this as a seed coating to provide the germinating seed and seedling with supplementary nutrients to aid in early stage growth.

Macronutrients selected:

Phosphorus (P)

- Phosphorus (P) is an essential part of the process of photosynthesis.
- Involved in the formation of all oils, sugars, starches, etc.
- Helps with the transformation of solar energy into chemical energy; proper plant maturation; withstanding stress.
- Effects rapid growth.
- Encourages blooming and root growth.

Potassium (K)

- Potassium is absorbed by plants in larger amounts than any other mineral element except nitrogen and, in some cases, calcium.
- Helps in the building of protein, photosynthesis, fruit quality and reduction of diseases.
- Potassium is supplied to plants by soil minerals, organic materials, and fertilizer.

Both Potassium and Phosphorus can be found in soluble form in Monobasic Potassium Phosphate or MKP(KH_2PO_4), a soluble salt commonly used as a fertiliser and plant growth supplement.

Formulation Method

Carnauba is heated on a hotplate at 100°C to a molten state. Monopotassium phosphate (MKP) is dissolved in deionised water to the required concentration. The MKP solution is slowly added to the molten wax under stirring at 1500rpm. Stirring continues for 5 minutes before the water/wax emulsion is poured onto a metal sheet to cool. The resulting solid wax including micro-droplets of MKP is then micronized in an air mill as outlined above with the exception the milling speed was set at 12,500rpm to obtain a VMD for the wax particles of approx. 10.3µm.

Experimental Method

Wax particles containing 10% MKP are added to 10g of Cyclamen seed, cv. Pripyat (Soya UK, Hampshire) at loadings of 0.1% and 1% by mass. Seed is well mixed to ensure a homogenous distribution across the seed. A third batch of seed is combined with unformulated carnauba wax particles as a control.

10 seeds for each treatment are sown in 20 cell modular seed trays with an individual cell size: Length 37mm x Width 37mm x Depth 65mm, with each tray representing a single sample. Each treatment is replicated four times.

The pots are filled with a sieved, heat-sterilised seed mix (Levingtons F1 Seed and Modular Compost – Low Nutrient)) to level with the top of the cell. Low Conductivity: 250-280 µS, Standard pH: 5.3-5.7, Mg/litre added: N – 100, P – 200, K – 200.

They are then lightly tamped and 30ml of deionised water added to each cell through a course filter. A single seed is then placed on the surface of the soil and covered with a thin layer of vermiculite to a depth of 2-3x the diameter of the seed, as per supplier recommendation. The trays are placed within plastic gravel trays (two per tray) which are

lined with capillary matting to aid watering.

The gravel trays are then placed in a thermostatically controlled plant growth chamber (Fitotron SGC120, Weiss Gallenkamp, Loughborough, UK). Temperature cycling is set at 20°C/10°C on a 16/8hr schedule. Lighting at 150 $\mu\text{mol m}^{-2} \text{s}^{-1}$ on a 16/8hr photoperiod is introduced at first emergence.

Plants are watered daily from the bottom in order to maintain a compost moisture level of approximately 40% in the cells

After 10 days the plants are removed from the individual cells and the compost mix separated from the root structure. Plants from each 10 cell tray are combined and separated into shoots, made up of the first true leaves and growing tip, and roots.

PLANT TISSUE ANALYSIS: (conducted to the following method by NRM Laboratories (Bracknell, UK)

Total Phosphorus (P), Potassium (K), Magnesium (Mg), Calcium (Ca), Sodium (Na), Manganese (Mn), Copper (Cu), Iron (Fe), Zinc (Zn), Boron (B) determination using Inductively Coupled Plasma-Optical Emission Spectroscopy (ICP-OES) method

EQUIPMENT

1. ICP Emission Spectrograph
2. Autosampler
3. Digital Dilutor

REAGENTS

1. 3N Hydrochloric Acid: Dilute 250 ml. of concentrated hydrochloric acid to 1 litre using deionized water and mix well.
2. Nitric Acid: HNO_3

STANDARDS

1. Stock Solutions: Use 1000 PPM certified, NIST traceable, plasma grade standards for the 16 listed elements.
2. Instrument Calibration Standards:
 - a. WAT 1 - deionized water
 - b. WAT 2 - Mn, Fe, Al, B, Cu, Zn, Na, Pb, Cd, Ni, Cr, Mo - Pipet 10 ml. of stock solution of each element into a 1 litre volumetric flask. Add 5 ml. of nitric acid.

Dilute to volume with deionized water and mix well.

- c. PLN2 - P, K, Ca, Mg - Pipet the designated ml. of stock solution into a 1 litre volumetric flask. Add 5 ml. of nitric acid. Dilute to volume with deionized water and mix well.

<i>Element</i>	<i>Stock ml.</i>	<i>Solution Concentration ppm.</i>	<i>Final Instrument Readout %</i>
P	10	10	1.00
K	50	50	5.00
Ca	20	20	2.00
Mg	10	10	1.00

3. Instrument Calibration Verification Standards:

- a. A second set of calibration standards obtained from a different manufacturer.

SAMPLE PREPARATION

Samples are dried and ground to pass through a 1mm screen.

The elements in the residue remaining after the destruction of the organic matter by ashing at 550°C are dissolved in hydrochloric acid:

1. Dry Ash

- Weigh 1g sample into a 10 ml. glazed, high-form porcelain crucible.
- Ash in a muffle furnace for 4 hours at 500 C.
- Let cool and add 5 ml. of 3N HCl.
- Place on a hot plate and boil gently for 5 minutes.
- Let cool and transfer to a 100 ml. volumetric flask. Dilute to volume with deionized water and mix well. Use this solution for the analysis of Mn, Fe, Al, B, Cu, Zn, Na, Pb, Cd, Ni, Cr and Mo.
- Dilute the solution obtained in 1e. one to ten with deionized water using a digital dilutor. Use this solution for the analysis of P, K, Ca and Mg.

ICP PROCEDURE

1. Set up and operate the ICP Emission Spectrograph in accordance with manufacturer's specifications.
2. Mn, Fe, B, Cu, Zn, Na, Ni analysis.
 - a. Choose PLANT from the method menu.
 - b. Calibrate the instrument using WAT1 and WAT2 instrument calibration standards.
 - c. Analyze the sample digests obtained in 1e. of the sample preparation section.
3. P, K, Ca, Mg analysis.
 - a. Choose PLANTDIL from the method menu.
 - b. Calibrate the instrument using WAT1 and PLN2 instrument calibration standards.
 - c. Analyze the digests obtained in 1f. of the sample preparation section.

QUALITY CONTROL

1. Following calibration, analyze one high instrument calibration standard, one instrument calibration verification standard and one quality control sample.
 - a. Instrument Calibration Standard: Values must be within 3% of the known value for K and Mo. All other elements must be within 2% of the known value.
 - b. Instrument Calibration Verification Standard: Values must be within 10% of the certified values.
 - c. Quality Control Sample: Values for all elements must be within limits established by the Extension chemist.
2. Analyze a high instrument calibration standard after each tenth sample and at the end of the set of samples.
 - a. Values must be within 8% of the known values.
 - b. If any of the values are greater than 8% from the known values, recalibrate the instrument and begin sample analysis from the last "good" instrument calibration standard.
3. Prepare one duplicate sample for each 10 samples. If the set contains less than 10 samples, prepare one duplicate per set.
 - a. Results on the duplicate sample should agree within 20% of the average value of the two samples.

REFERENCES

1. Isaac, R.A. and W.C. Johnson, 1985, *Elemental Analysis of Plant Tissue by Plasma Emission Spectroscopy: Collaborative Study*. JAOAC. 68(3), pp 499- 505.

2. AOAC Official Method 985.01, in *Official Methods of Analysis of AOAC International*, 16th edition, Volume I Chapter 3, p. 4.
3. AOAC Official Method 968.08 D(a), in *Official Methods of Analysis of AOAC International*, 16th edition, Volume I Chapter 4, p. 23.

Phosphate Solubilisation using Beneficial Microbes

Several bacterial species are able to impart a beneficial effect upon plant growth. Mostly they are associated with the plant rhizosphere, so they are named rhizobacteria. This group of bacteria has been termed plant growth promoting rhizobacteria, and among them are strains from genera such as *Alcaligenes*, *Acinetobacter*, *Arthrobacter*, *Azospirillum*, *Bacillus*, *Burkholderia*, *Enterobacter*, *Erwinia*, *Flavobacterium*, *Paenibacillus*, *Pseudomonas*, *Rhizobium*, and *Serratia*.

The production of organic acids by phosphate solubilizing bacteria has been well documented and identified as the main mechanism for phosphate solubilisation. Gluconic acid seems to be the most frequent agent of phosphate solubilisation (*Pseudomonas* sp.), and 2-ketogluconic acid is also identified in strains with phosphate solubilizing ability (*Rhizobium* sp.).

Saprophytic fungi are also known to solubilise both organic and inorganic phosphates. Several genus, including *Trichoderma*, *Penicillium*, and *Gliocladium* have exhibited potential as biofertilisers. Morales et al (2011) demonstrated that *Penicillium albidum* was able to solubilise 64mg of organic/inorganic phosphate per gram of fungi.

Experiment to assess the potential for delivery of Phosphate Solubilising Organisms as a seed coating using Carnauba wax particles

Using a dry spore powder of a phosphate solubilising organism, such as *Penicillium bilaii*. Spores are combined with carnauba wax particles with a VMD of approximately 10µm at a ratio of 1:3. The powders are agitated to create a homogenous mix and applied to sterilised cyclamen seed at a loading of 0.1% (by mass). Additional batches of seed are treated with spores only (0.1%), Entostat only (0.1%) and untreated seed.

Phosphate Solubilising Activity Screening

Plate screening using Pikovskays' medium (see below) is used to demonstrate phosphate solubilising activity of the treated seed. 9cm petri dishes are divided into quadrants and a seed is placed in the centre of each quadrant. Plates are incubated at 20°C for 4 days.

Active phosphate solubilising agents produce clear zones around the seed as they solubilise

the insoluble mineral phosphates within the media. The radius of the clear zones is measured and compared to the mean results achieved for each treatment. Differences are analysed using one-way ANOVA and Tukey Post-Hoc diagnostic test where significance is found.

Phosphate uptake by plant

Seeds are treated as described above.

$\text{Ca}_3(\text{PO}_4)_2$ is used as a source of insoluble phosphate.

Sure to Grow PET grow cubes (25x25x38mm) are soaked in deionised water containing 1% $\text{Ca}_3(\text{PO}_4)_2$ in suspension until saturated. Cubes are placed in free draining plant trays on a level surface to prevent nutrient run-off and migration whilst taking care to avoid pooling of water at the root zone. 10 cubes are used per tray and the mean of these represents one replicate. Each treatment is replicated 8 times.

A single cyclamen seed is placed in the cross-cut X in the top of each cube. Seed trays are then covered to maintain a humid environment and regularly top watered with the 1% $\text{Ca}_3(\text{PO}_4)_2$ suspension to maintain a moist cube. Trays are incubated at 20°C and 10°C on a 16/8hr thermal cycle. On germination the cover is removed and the seedling exposed to lighting on a 16/8hr photoperiod.

After 15 days the plants are removed from the grow cube and nutrient content of the plant tissue is analysed using the ICP method described above.

Differences in the Phosphate content between treatments are assessed statistically using one-way ANOVA.

Pikovskays' Medium

Components	Quantities (g l ⁻¹)
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Glucose	10
$\text{Ca}_3(\text{PO}_4)_2$	5
$(\text{NH}_4)_2\text{SO}_4$	0.5
NaCl	0.2
$\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$	0.1
KCl	0.2
Yeast Extract	0.5
$\text{MnSO}_4 \cdot \text{H}_2\text{O}$	0.002
$\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	0.002
pH	7.0

CLAIMS

1. An ornamental plant structure coating composition in the form of particles that comprises
 - i) at least one organic material selected from waxes having a melting point of $\geq 50^{\circ}\text{Centigrade}$; and
 - ii) at least one additive for enhancing seedling vigour and/or seedling growth from ornamental plant structures wherein the at least one additive is selected from one or more inorganic additives and/or one or more live biological agents.
2. A coating composition according to claim 1 wherein the particles have a volume mean diameter of $\geq 5\mu\text{m}$.
3. A coating composition according to claim 1 or claim 2 wherein the particles have a mean volume diameter in the range from $\geq 10\mu\text{m}$ to $200\mu\text{m}$.
4. A coating composition according to any one of claims 1 to 3 wherein the organic material is selected from carnauba wax, beeswax, montan wax, Chinese wax, shellac wax, spermaceti wax, myricyl palmitate, cetyl palmitate, candelilla wax, castor wax, ouricury wax, and rice bran wax; or a mixture of two or more thereof.
5. A coating composition according to any one of the preceding claims wherein the particles are carnauba wax particles.
6. Use of an organic material in the form of particles wherein the particles are selected from at least one wax having a melting point of $\geq 50^{\circ}\text{Centigrade}$ in the manufacture of a coating composition for ornamental plant structures according to any one of claims 1 to 5.
7. Use according to claim 6 wherein the at least one wax is selected from carnauba wax, beeswax, montan wax, Chinese wax, shellac wax, spermaceti wax, myricyl palmitate, cetyl palmitate, candelilla wax, castor wax, ouricury wax, and rice bran wax; or a mixture of two or more thereof.
8. Use according to claim 6 or claim 7 wherein the ornamental plant structure coating composition comprises particles of carnauba wax.
9. Use according to any one of claims 4 to 8 wherein the particles have a volume mean diameter $\geq 10\mu\text{m}$.
10. A method of manufacturing an ornamental plant structure coating composition according to any one of claims 1 to 5 that comprises

- 1) selecting solid organic material wherein the solid organic material is wax having a melting point of $\geq 50^{\circ}\text{C}$ Centigrade;
- 2) machining said organic material into particles of a mean volume diameter $\geq 10\mu\text{m}$; and
- 3) adding one or more additives for enhancing seedling vigour and/or seedling growth to the machined organic material of 2), wherein the said additives are selected from one or more inorganic additives and/or one or more live biological agents.

11. A method according to claim 10 wherein the organic material is selected from carnauba wax, beeswax, montan wax, Chinese wax, shellac wax, spermaceti wax, myricyl palmitate, cetyl palmitate, candelilla wax, castor wax, ouricury wax, and rice bran wax; or a mixture of two or more thereof.

12. A method according to claim 10 or claim 11 wherein the said material is carnauba wax.

13. A coating composition according to any one of claims 1 to 5 wherein the ornamental plant structure is selected from seeds of an ornamental plant.

14. A coating composition according to claim 13 wherein the seeds of the ornamental plant are selected from viola (pansy), primula, petunia, polyanthus, tagetes, pelargoniums including *P. Peltatum*, begonia, cyclamen, achillea, ageratum, agrostemma, alyssum, amaranthus, antirrhinum, aquilegia, aster, calendula, campanula, carnation, chrysanthemum, helleborus, cineraria, clematis, convolvulus, Centaurea cyanus, cosmea, dahlia, delphinium, dianthus, digitalis, myosotis, freesia, geranium, godetia, impatiens, cheiranthus cheiri, dianthus barbatus, lathyrus odoratus, salvia, salpiglossis, verbena, and zinnia; and woody seeds of ornamental plants such as from the arborvitae, acer, azalea, Chamaecyparis, dogwood, euonymus, rose, forsythia, Fraser fir, hemlock, Japanese holly, juniper, Pieris, rhododendron, Taxus, white pine, maple, elm, aspen, ash, beech, and oak.

15. A method of coating ornamental plant structures with a coating composition that comprises at least one organic material wherein the organic material is selected from waxes having a melting point of $\geq 50^{\circ}\text{C}$ Centigrade, the method comprising:

- i) obtaining a population of particles of wax of a pre-determined VMD; and
- ii) applying the said population of particles to ornamental plant structures, such as ornamental plant seeds.

16. A method according to claim 15 wherein the coating composition comprises a

particulate composition that is applied to ornamental plant structures in dry particulate form.

17. A method according to claim 15 or claim 16 wherein the coating composition comprises a wax in particulate form selected from carnauba wax, beeswax, montan wax, Chinese wax, shellac wax, spermaceti wax, myricyl palmitate, cetyl palmitate, candelilla wax, castor wax, ouricury wax, and rice bran wax; or a mixture of two or more thereof.

18. A method of coating ornamental plant structures according to any one of claims 15 – 17 with a coating composition that comprises an organic material that is selected from waxes having a melting point of $\geq 50^{\circ}\text{C}$ entigrade, the method comprising

- i) obtaining said organic material;
- ii) heating the organic material so as to form a liquid phase or a gaseous phase;
- iii) cooling the liquid phase or gaseous phase of ii) to below the melting point of the organic material, forming a solid;
- iv) adding one or more additives to the solid formed in iii);
- v) machining the solid organic material of iii) into particles of a pre-determined VMD; and
- vi) applying the particles of v) to ornamental plant structures.

19. A method of coating ornamental plant structures according to any one of claims 15 – 18 with a coating composition that comprises an organic material that is selected from waxes having a melting point of $\geq 50^{\circ}\text{C}$ entigrade, the method comprising

- i) obtaining said organic material;
- ii) heating the organic material so as to form a liquid phase or a gaseous phase;
- iii) adding one or more additives to the liquid phase or gaseous phase of ii);
- iv) cooling the liquid phase or gaseous phase of iii) to below the melting point of the organic material, forming a solid;
- v) machining the solid organic material of iv) into particles of a pre-determined VMD; and
- vi) applying the particles of v) to ornamental plant structures.

20. A method according to any one of claims 15 to 19 wherein the organic material is carnauba wax.

21. Ornamental plant structures comprising a coating composition according to any one

of claims 1 to 5.

22. Ornamental plant structures according to claim 21 that are selected from coated seeds and coated bulbs.

23. Ornamental plant structures according to claim 21 or claim 22 that are ornamental plant seeds.

24. Ornamental plant structures according to any one of claims 21 to 23 that are selected from ornamental plant seeds selected from viola (pansy), primula, petunia, polyanthus, tagetes, pelargoniums including *P. Peltatum*, begonia, cyclamen, achillea, ageratum, agrostemma, alyssum, amaranthus, antirrhinum, aquilegia, aster, calendula, campanula, carnation, chrysanthemum, helleborus, cineraria, clematis, convolvulus, Centaurea cyanus, cosmea, dahlia, delphinium, dianthus, digitalis, myosotis, freesia, geranium, godetia, impatiens, cheiranthus cheiri, dianthus barbatus, lathyrus odoratus, salvia, salpiglossis, verbenas, and zinnia; and woody seeds of ornamental plants such as from the arborvitae, acer, azalea, Chamaecyparis, dogwood, euonymus, rose, forsythia, Fraser fir, hemlock, Japanese holly, juniper, Pieris, rhododendron, Taxus, white pine, maple, elm, aspen, ash, beech, and oak.