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Elsberry et al.

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- [54] **TECHNIQUES OF TREATING NEURODEGENERATIVE DISORDERS BY BRAIN INFUSION**
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- [73] Assignee: **Medtronic Incorporated**, Minneapolis, Minn.
- [21] Appl. No.: **902,548**
- [22] Filed: **Jul. 29, 1997**

Related U.S. Application Data

- [62] Division of Ser. No. 640,358, Apr. 30, 1996, Pat. No. 5,735,814.
- [51] **Int. Cl.⁶** **A61M 3/00**
- [52] **U.S. Cl.** **604/43; 604/93**
- [58] **Field of Search** 604/43, 49, 50, 604/51, 93

References Cited

U.S. PATENT DOCUMENTS

3,957,036	5/1976	Normann .	
4,146,029	3/1979	Ellinwood, Jr. .	
4,256,437	3/1981	Brown .	
4,576,556	3/1986	Thompson .	
4,692,147	9/1987	Duggan .	
5,011,472	4/1991	Aebischer et al. .	
5,293,879	3/1994	Vonk et al. .	
5,474,547	12/1995	Aebischer .	
5,713,858	2/1998	Heruth et al.	604/93
5,713,888	2/1998	Neuenfeldt et al.	604/93
5,735,814	4/1998	Elsberry et al.	604/43

FOREIGN PATENT DOCUMENTS

WO 9401166 1/1994 European Pat. Off. .

OTHER PUBLICATIONS

Duncan et al., "Thalamic VPM Nucleus In The Behaving Monkey, III. Effects of Reversible Inactivation by Lidocaine on Thermal and Mechanical Discrimination," *J. Neurophysiology*, vol. 70, No. 5, pp. 2086-2096 (1994).

- Bobo et al., "Convection-enhanced delivery of macromolecules in the brain," *Proc. Natl. Acad. Sci. USA*, 91, pp. 2076-2080 (1994).
- Crossman et al., "Experimental Herniballismus In The Baboon Produced By Injection Of A Gamma-Aminobutyric Acid Antagonis Into The Basal Ganglia," *Neuroscience Letters*, 20 pp. 369-372 (1980).
- Graham et al., "Injection of Excitatory Amino Acid Antagonists Into The Medial Pallidal Segment of a 1-Methyl-4-Phenyl-1,2,3,6-Tetrahydropyridine (MPTP) Treated Primate Reverses Motor Symptoms of Parkinsonism", *Life Sciences*, 47, pp. 91-97 (1990).
- van Horne et al., "Multichannel Semiconductor-based electrodes for in vivo electrochemical and electrophysical studies in rat CNS", *Neuroscience Letters*, 120, pp. 249-252 (1990).
- Couratier et al., "Cell culture evidence for neuronal degeneration in amyotrophic lateral sclerosis being linked to glutamate AMPA/kainate receptors", *The Lancet*, vol. 341, pp. 265-268 (Jan. 30, 1993).

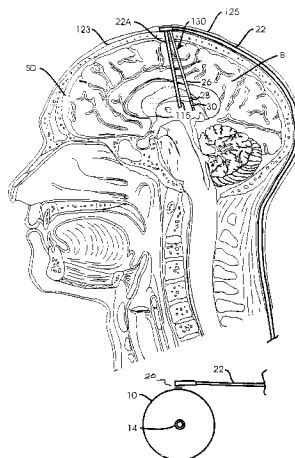
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[57] **ABSTRACT**

Techniques for infusing drugs into the brain to treat neurodegenerative disorders by system of an implantable pump and catheter. The drugs are capable of altering the level of excitation of neruons in the brain. A sensor is used to detect an attribute of the nervous system which reflects the hyperexcitation of the nerve cells projecting onto the degenerating nerve cells. A microprocessor algorithm analyzes the output from the sensor in order to regulate the amount of drug delivered to the brain.

7 Claims, 2 Drawing Sheets



OTHER PUBLICATIONS

- Shaw et al., "Studies on Cellular Free Radical Protection Mechanism in the Anterior Horn from Patients with Amyotrophic Lateral Sclerosis", *Neurodegeneration*, vol. 4, pp. 391–396 (1995).
- Yamauchi et al., "Corpus Callosum atrophy in amyotrophic lateral sclerosis", *J. Neurological Sciences*, 134, pp. 189–196 (1995).
- Hollander et al., "High-dose Dextromethorphan in Amyotrophic Lateral Sclerosis: Phase I Safety and Pharmacokinetic Studies", *Ann. Neurol.* 36, pp. 920–924 (1994).
- Nihei et al., "NPY Immunoreactive Neurons Are Involved In the Degeneration Of Amyotrophic Lateral Sclerosis", *Biomedical Research* 15, Suppl. 2, pp. 209–212 (1994).
- Lanius et al., "A Role for Amplified Protein Kinase C Activity in the Pathogenesis of Amyotrophic Lateral Sclerosis", *J. Neurochemistry*, pp. 927–930 (1995).
- Appel, "Excitotoxic neuronal cell death in amyotrophic lateral sclerosis", *TINS* vol. 16, No. 1, pp. 3–5 (1993).
- Rothman et al., "Excitotoxicity and the NMDA receptor", *TINS* vol. 10, No. 7, pp. 299–302 (1987).
- Benabid et al., "Long-term suppression of tremor by chronic stimulation of the ventral intermediate thalamic nucleus", *The Lancet*, vol. 337, pp. 403–406 (1991).
- Benabid et al., "Vim and STN Stimulation in Parkinson's Disease", *Movement Disorders*, vol. 9, Suppl. 1, Abstract M39 (1994).
- Limousin et al., "Effect on Parkinsonian signs and symptoms of bilateral subthalamic nucleus stimulation", *The Lancet*, vol. 345, pp. 91–95 (1995).
- Bensimon et al., "A Controlled Trial of Riluzole in Amyotrophic Lateral Sclerosis", *The New England Journal of Medicine*, vol. 330, No. 9, pp. 585–591 (1994).
- A. Depaulis et al., "Suppressive effects of intranigral injection of muscimol in three models of generalized non-convulsive epilepsy induced by chemical agents", *Brain Research*, 498 (1989) 64–72.
- Dill et al., "Dyskinesias in Rats following Chemical Stimulation of the Neostriatum", *Texas Reports on Biology and Medicine*, vol. 26, No. 1, Spring (1968).
- Godsey et al., "Omnitrode: A Simple Cannula for Chemical and Bipolar Electrical Stimulation", *Physiology and Behavior*, vol. 8, pp. 773–775, Brain Research Publications Inc. 1972 (Great Britain).
- Kroll et al., "Increasing volume of Distribution to the Brain with Interstitial Infusion: Dose, Rather Than convection, Might be the Most Important Factor", *Neurosurgery*, vol. 38, pp. 746–754 (1996).
- May et al., "Intrastratial Infusion of Lisuride—a Potential Treatment for Parkinson's disease? Behavioral and Autoradiographic Studies in 6-OHDA Lesioned Rats", *Neurodegeneration*, vol. 3, pp. 305–313 (1994).

Fig. 1

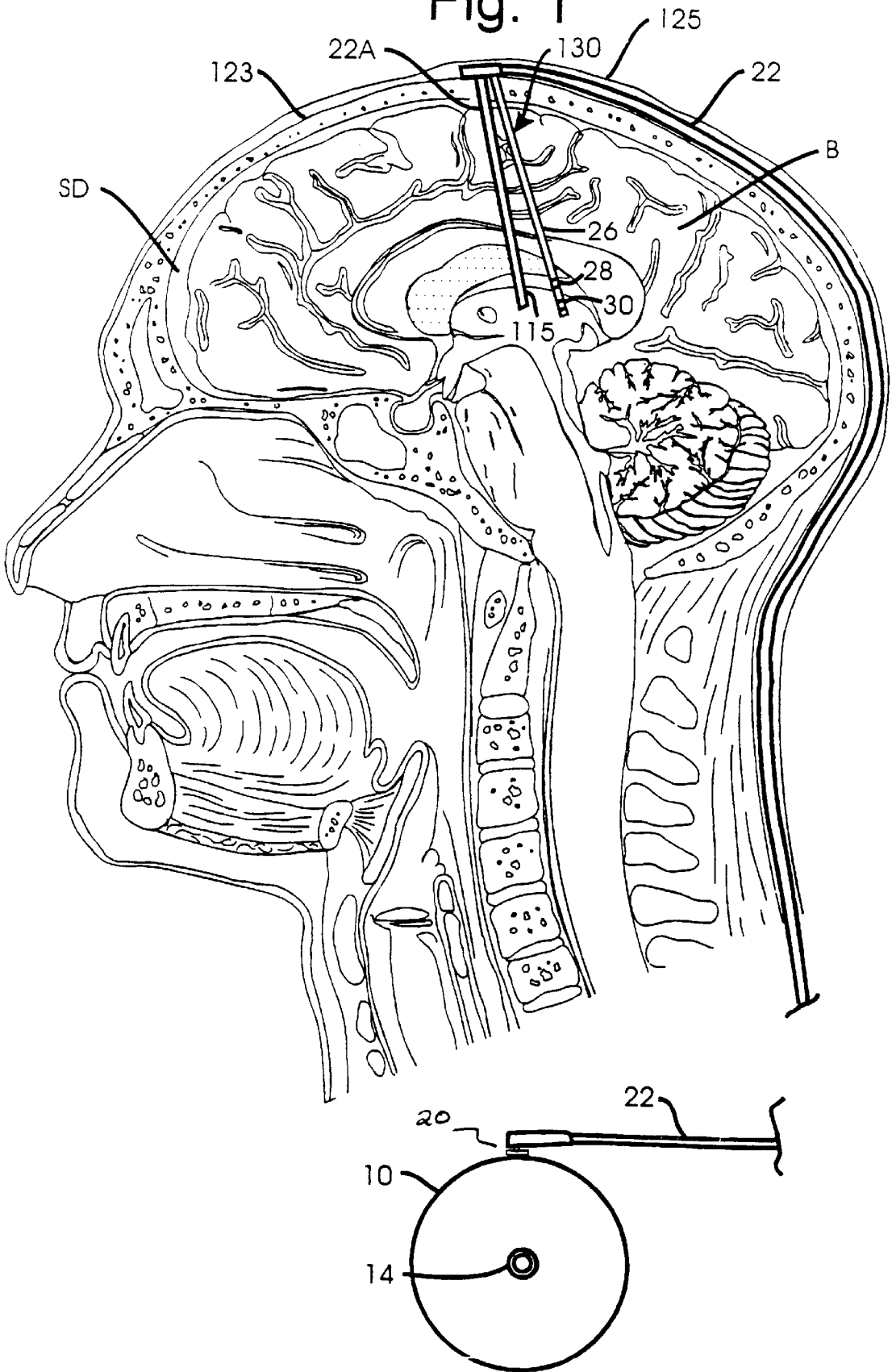


Fig. 2

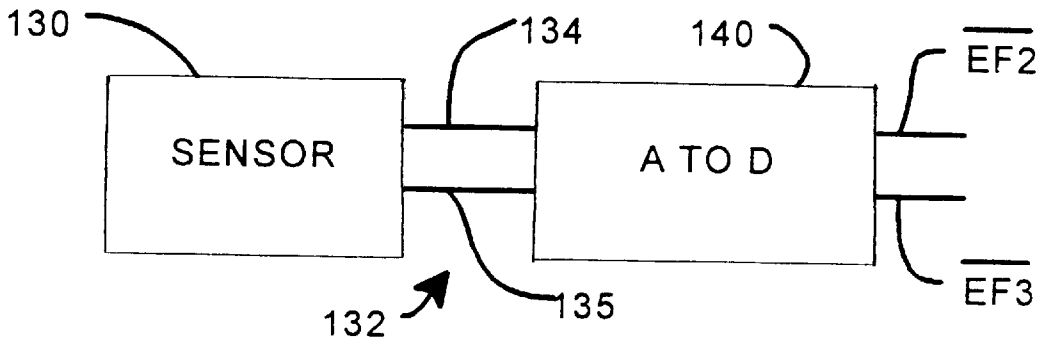
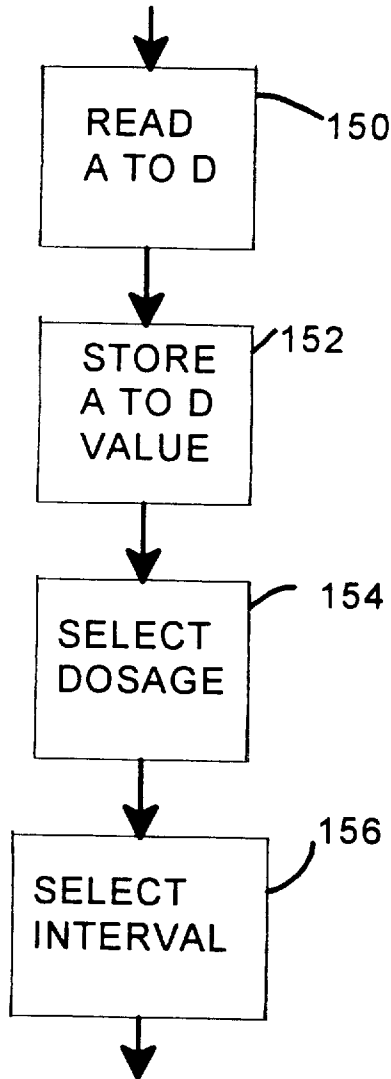


Fig. 3



TECHNIQUES OF TREATING NEURODEGENERATIVE DISORDERS BY BRAIN INFUSION

This is a divisional of application Ser. No. 08/640,358, filed Apr. 30, 1996, now U.S. Pat. No. 5,735,814 for which priority is claimed.

BACKGROUND OF THE INVENTION

1. Field of the Invention

This invention relates to brain infusion techniques, and more particularly relates to such techniques for treating neurodegenerative disorders.

2. Description of Related Art

Neuroscientists have recognized and continue to explore excitotoxicity, a phenomenon referring to excessive excitation of nerve cells leading to degeneration of the nervous system. This phenomena has been used to explain cell loss after stroke or some other hypoxic event. The research has focused on nerve cells that have glutamate neurotransmitter receptors especially susceptible to the sustained insult. Hyperexcitation of these nerve cells is fundamental to the mechanism (Rothman, S. M., Olney, J. W. (1987) *Trends Neurosci.* 10, 299-302). Researchers have also used excitotoxicity to explain the observed cell loss in the CA1 region of the Horn of Ammon in the dentate gyrus of hippocampus in patients and animal subjects that have suffered from seizure activity. Seizures can be viewed as a form of abnormal over excitation of the nerve cells in this region.

Typically, neuroscientists have focused on nerve cells that use the transmitter substance glutamate to communicate with target nerve cells; however, other excitatory amino acids (EAA) are included. When nerve cells are abnormally active, experiencing excessive action potentials, they are believed to release excessive amounts of glutamate or other EAA at their synaptic terminals. The presence of excessive amounts of glutamate leads to toxic effects on the secondary nerve cells targeted by the hyperactive ones. These toxic effects are believed to be mediated by an accumulation of calcium.

Benabid et al. (*The Lancet*, Vol 337:Feb. 16, 1991, pp 403-406) has shown that stimulation of the Vim nucleus of the Thalamus will block tremor. In this instance, stimulation at frequencies around 100 to 185 pulses per second accomplishes the same physiological response as a lesion of this region. Thus, it appears that stimulation inhibits the output of these cells. Benabid's research team has extended this work to stimulation of the subthalamus ("Vim and STN Stimulation in Parkinson's disease", *Movement Disorders*, Vol. 9, Supplement 1 (1994); "Effect on Parkinsonian signs and symptoms of bilateral subthalamic nucleus stimulation", *The Lancet*, Vol 345, Jan. 14, 1995).

Parkinson's disease is the result of degeneration of the substantia nigra pars compacta. The cells of subthalamus have been shown to use glutamate as the neurotransmitter effecting communication with their target cells of the basal ganglia. The state of hyperexcitation that exists in Parkinson's disease will cause an excessive release of glutamate. This, in theory, will lead to further degeneration via the mechanism described above.

Benabid has proposed a method of arresting degeneration of the substantia nigra by high frequency electrical pulsing of the subthalamic nucleus to block stimulation of the subthalamic nucleus, thereby inhibiting excessive release of glutamate at the terminal ends of the axons projecting from the subthalamic nucleus to the substantia nigra.

Amotrophic Lateral Sclerosis (ALS), sometimes referred to as Lou Gerhig's disease, is a progressive neurodegenerative disease affecting the voluntary motor system. The degeneration of the spinal cord and cortical motor neurons results in paralysis, respiratory depression and death. Glutamate neurons appear to be overactive in this disease state and are suspected of causing the neurodegeneration (S. H. Apel, "Excitotoxic neuronal cell death in amyotrophic lateral sclerosis", *TINS*, Vol. 16, No. 1, 1993).

Huntington's disease is a degenerative disorder characterized by choreathetosis which is at first slight. Usually, the choreathetosis is accompanied by hypotonus. The motor dysfunction, associated with excitatory transmitter neurotoxicity, may be accompanied by subtle forms of mental disorder that progress to a deterioration of cognitive function.

SUMMARY OF THE INVENTION

A preferred form of the invention can treat a neurodegenerative disorder, such as Parkinson's disease Huntington's or Amyotrophic Lateral Sclerosis (ALS), by means of an implantable pump and a catheter having a proximal end coupled to the pump and having a discharge portion for infusing therapeutic dosages of the one or more drugs capable of altering the level of excitation of neurons of the brain. The catheter is implanted in the brain so that the discharge portion lies adjacent to a predetermined infusion site in the basal ganglia or thalamus of the brain. Alternatively, the catheter may be implanted in a ventricle of the brain or subdurally so that the discharge portion lies within the cerebral spinal fluid (CSF). The pump is operated to discharge a predetermined dosage of the one or more drugs through the discharge portion of the catheter into the infusion site. By using the foregoing method, the neurodegeneration that occurs in diseases, such as Parkinson's disease, Huntington's disease and Amyotrophic Lateral Sclerosis, can be alleviated or prevented.

According to one embodiment of the invention, one or more drugs can decrease excitement of the subthalamus or increase inhibition of the subthalamus. According to another embodiment of the invention, the one or more drugs can increase excitement of the thalamus or decrease inhibition of the thalamus.

Another form of the invention uses a sensor in combination with the implantable pump and catheter to administer one or more drugs capable of altering the level of excitation of neurons of the brain to treat a neurodegenerative disorder. In this form of the invention, the sensor generates a signal relating to an attribute of the nervous system which indicates the hyperexcitation of the degenerating neurons or the neurons related to the degenerating neurons. Control means responsive to the sensor signal regulate the therapeutic dosage. For example, the dosage can be increased in response to an increase in the hyperexcitation of the neurons and is decreased in response to a decrease in the hyperexcitation of the neurons.

By using the foregoing techniques, neurodegeneration can be controlled to a degree unattainable by prior art methods or apparatus.

BRIEF DESCRIPTION OF THE DRAWINGS

These and other advantages and features of the invention will become apparent upon reading the following detailed description and referring to the accompanying drawings in which like numbers refer to like parts throughout and in which:

FIG. 1 is a diagrammatic illustration of a portion of the nervous system of the human body in which a preferred form of hyperexcitation sensor, pump and catheter have been implanted;

FIG. 2 is a schematic block diagram of a sensor and analog to digital converter circuit used in the preferred embodiment of the invention; and

FIG. 3 is a flow chart illustrating a preferred form of a microprocessor program for utilizing the sensor to control the dosage of drug administered to the brain.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

Referring to FIG. 1, a system or device **10** made in accordance with the preferred embodiment may be implanted below the skin of a patient. The device has a port **14** into which a hypodermic needle can be inserted through the skin to inject a quantity of a liquid agent, such as a medication or drug. The liquid agent is delivered from device **10** through a catheter port **20** into a catheter **22**. Catheter **22** is positioned to deliver the agent to specific infusion sites in a brain (B). Device **10** may take the form of the like-numbered device shown in U.S. Pat. No. 4,692,147 (Duggan), assigned to Medtronic, Inc., Minneapolis, Minn., which is incorporated by reference.

The distal end of catheter **22** terminates in a cylindrical hollow tube **22A** having a distal end **115** implanted into a portion of the basal ganglia of the brain by conventional stereotactic surgical techniques. Additional details about end **115** may be obtained from pending U.S. application Ser. No. 08/430,960 entitled "Intraparenchymal Infusion Catheter System," filed Apr. 28, 1995 in the name of Dennis Elsberry et al. and assigned to the same assignee as the present application. Tube **22A** is surgically implanted through a hole in the skull **123** and catheter **22** is implanted between the skull and the scalp **125** as shown in FIG. 1. Catheter **22** is joined to implanted device **10** in the manner shown. Device **10** is implanted in a human body in a subcutaneous pocket located in the chest below the clavicle. Alternatively, device **10** may be implanted in the abdomen.

In a second embodiment, distal end **115** of cylindrical hollow tube **22A** may be implanted in a ventricle. Alternatively, the distal tip may be located in subdural area SD beneath the dura under the skull **123** but outside the brain B.

Catheter **22** may be divided into twin tubes **22A** and **22B** (not shown) that are implanted into the brain bilaterally. Alternatively, tube **22B** (not shown) implanted on the other side of the brain may be supplied with drugs from a separate catheter and pump.

A sensor **130** is implanted into a portion of a patient's central nervous system. As shown in FIG. 1, sensor **130** comprises a sensing lead **26** having two sensing electrodes **28** and **30** located in the subthalamic region, substantia nigra or other brain region whose electrical activity indicates the activity of the degenerating neurons, i.e., the neurons exhibiting hyperexcitation. Alternatively, electrodes **28** and **30** could be carried by tube **22A**. Electrodes **28** and **30** are connected to an analog to digital converter **140** (FIG. 2) by conductors **134** and **135** which are located within catheter **22**. The potentials sensed by electrodes **28** and **30** indicate the electrical activity in the subthalamic nucleus and consequently the substantia nigra. Electrodes **28** and **30** transmit a signal related to the excitation of the portion of the brain exhibiting hyperexcitation. More specifically, electrodes **28** and **30** sense an attribute of the nervous system which

indicates the hyperexcitation of the nerve cells projecting onto the degenerating nerve cells or the hyperexcitation of the degenerating nerve cells. Sensor **130** may take the form of a device capable of detecting nerve cell electrical activity that is related to the hyperexcitation. Such a sensor may be located deep in the brain. For such detecting, sensor **130** may take the form of an electrode inserted into one of the nuclei of the basal ganglia, the thalamus, the internal capsule or the cortex of the brain. Alternatively, such a sensor may be located outside the dura in the bone of the cranium. Signals that are received by the sensor may be amplified before transmission to circuitry contained within device **10**.

Alternatively, sensor **130** may electronically transduce the concentration of a transmitter substance infused into the brain or released endogenously. A paper describing such a sensor is entitled "Multichannel Semiconductor-based Electrodes for In Vivo Electrochemical and Electrophysiological Studies in Rat CNS", by van Horne et al., 120 *Neuroscience Letters* 249-252 (Elsevier Scientific Publishers Ireland Ltd. 1990).

Referring to FIG. 2, the output of sensor **130** is coupled by a cable **132** comprising conductors **134** and **135** to the input of analog to digital converter **140**. The output of the analog to digital converter is connected to terminals EF2 BAR and EF3 BAR shown in FIG. 11A of U.S. Pat. No. 4,692,147 ("147 Patent"). Before converter **140** is connected to the terminals, the demodulator **101** currently shown in FIG. 11A would be disconnected.

The present invention may be implemented by providing seven different drug dosages from 0 dosage to a 1.0 ml dosage with 0.1 ml increments between choices. The time interval between dosages can be selected between one and twelve hours in seven choices. This is the same type of dosage and interval described in connection with device **10** shown in the '147 Patent (column 5, beginning at line 63). The seven drug dosages and corresponding time increments may be loaded into RAM **102a** (FIG. 11B) of the '147 Patent. The appropriate drug dosage and interval is selected by a computer algorithm that reads the output of converter **140** and makes the appropriate selection.

One exemplary computer algorithm is shown in FIG. 3. Microprocessor **100** included within device **10** reads converter **140** in step **150**, and stores one or more values in RAM **102a** in step **152**. One of seven dosages is selected in step **154**, and an appropriate time interval is selected in step **156**. The selected dosage and interval of a drug is then delivered through catheter **22** and tube **22A** to the basal ganglia of the brain by the pump of the type as described in the '147 Patent.

For some types of sensor, a microprocessor and analog to digital converter will not be necessary. The output from sensor **130** can be filtered by an appropriate electronic filter in order to provide a control signal for a pump of the type shown in the '147 Patent.

The type of drugs administered by device **10** into the brain depend on the specific location at which distal end **115** of tube **22A** is surgically implanted. The appropriate drugs for use in connection with the portion of the basal ganglia or thalamus in which tube **22A** terminates, together with the effect of the drug on that portion of the brain for reducing the hyperexcitation of the subthalamic nucleus for the purpose of treating Parkinson's degeneration is provided in Table I:

TABLE I

EFFECT	PORTION OF BRAIN	DRUG
INCREASE EXCITATION	VL THALAMUS	glutamate agonist
DECREASE INHIBITION	VL THALAMUS	GABA antagonist
INCREASE INHIBITION	GPI/SNr	GABA agonist
DECREASE EXCITATION	GPI/SNr	Glutamate antagonist
INCREASE INHIBITION	STN	GABA agonist
DECREASE EXCITATION	STN	Glutamate antagonist
INCREASE EXCITATION	GPe	glutamate agonist
DECREASE INHIBITION	GPe	GABA antagonist
INCREASE DOPAMINE	NEOSTRIATUM	Dopamine agonist

In the foregoing Table I, VL Thalamus means ventrolateral thalamus; GPI means internal segment of globus pallidus; SNr means substantia nigra pars reticulata, STN means subthalamic nucleus; and GPe means external segment of globus pallidus.

Alternative targets and infusion strategies are appropriate for treatment of degenerative disorders that are hyperkinetic in nature. For treatment of amyotrophic lateral sclerosis appropriate drugs for use in connection with the portion of the spinal cord, cerebral motor cortex, basal ganglia, and thalamus in which tube 22A terminates, together with the effect of the drug on that portion of the nervous system for decreasing excitation of the thalamic pathway is provided in Table II.

TABLE II

EFFECT	TARGET STRUCTURE	DRUG
DECREASE EXCITATION	MOTOR CORTEX	GLUTAMATE ANTAGONIST
DECREASE EXCITATION	SPINAL CORD ANTERIOR HORN	PKC INHIBITOR
DECREASE EXCITATION	SPINAL CORD ANTERIOR HORN	GLUTAMATE ANTAGONIST
DECREASE EXCITATION	VL THALAMUS	GLUTAMATE ANTAGONIST
INCREASE INHIBITION	VL THALAMUS	GABA AGONIST
DECREASE INHIBITION	GPI/SNr	GABA ANTAGONIST
INCREASE EXCITATION	GPI/SNr	GLUTAMATE AGONIST
DECREASE INHIBITION	STN	GABA ANTAGONIST
INCREASE EXCITATION	STN	GLUTAMATE AGONIST
DECREASE EXCITATION	GPe	GLUTAMATE ANTAGONIST
INCREASE INHIBITION	GPe	GABA AGONIST
DECREASE DOPAMINE	NEOSTRIATUM	DOPAMINE ANTAGONIST

Stereotaxic coordinates based on a normal brain for the portions of the brain described in Tables I and II are identified in the following Table III:

TABLE III

BRAIN REGION	MEDIAL-LATERAL DIMENSION	DORSAL-VENTRAL DIMENSION	ANTERIOR-POSTERIOR DIMENSION
VL Thalamus	0.7 to 1.8	1.5 to -0.2	0.0 to -1.0
Gpi	0.5 to 2.0	0.5 to -0.7	0.7 to 2.0
SNr	0.5 to 1.5	-0.6 to -1.5	0.7 to -0.7
STN	0.5 to 2.0	0.0 to -1.0	0.6 to -1.0
GPe	1.6 to 2.7	1.0 to -1.0	2.0 to -1.0

TABLE III-continued

BRAIN REGION	MEDIAL-LATERAL DIMENSION	DORSAL-VENTRAL DIMENSION	ANTERIOR-POSTERIOR DIMENSION
<u>Striatum:</u>			
Caudate	0.5 to 2.0	1.5 to 3.0	1.5 to 3.0
Putamen	1.2 to 3.3	1.5 to -1.0	2.5 to -1.2

In the foregoing table: the medial-lateral dimensions are relative to midline of the brain; the anterior-posterior dimensions are relative to the midpoint between the anterior commissure and posterior commissure with negative indicating the posterior direction; the dorsal-ventral dimensions are relative to a line connecting the midpoints of the anterior and posterior commissures with negative being ventral to said line; all dimension are in centimeters.

Preferred ranges of dosages and specific drugs for the brain infusion sites identified in Tables I and II are provided in the following Table IV:

TABLE IV

DRUG CLASS	SPECIFIC DRUG	DOSING RANGE
Glutamate Agonist	D-Cycloserine	1-10 muM
	L-AP4	1-10 muM
	Carboxyphenylglycine	10-500 muM
	L-glutamic acid	1-100 muM
	cis-Piperidine-2,3-dicarboxylic acid	1-10 muM
Glutamate Antagonists	(+/-)-trans-ACPD	1-10 muM
	L-AP4	1-10 muM
	MK801(dizocilpine)	1-20 muM
	ketamine Hcl	5-50 muM
	AP-3	1-10muM
GABA Agonists	Dextromethorphan	1-100 muM
	MCPD	0.02-10 muM
	dextropran tartrate	1-100 muM
	CNOX	1-100 muM
	baclofen	0.1-10 muM
GABA Antagonists	muscinol HBr	100-500 muM
	Gabazine	1-50 muM
	Saclofen	0.5-25 muM
	Biculline	1-100 muM
	picrotoxin	10-100 muM
Dopamine Antagonist	(+) apomorphine Hcl	5-20 muM
	spiperone Hcl	0.1-10 muM
	haloperidol	10-100 muM
	(-) Sulpiride	0.05-1 muM
	(-) apomorphine pergolide	1-10 muM
Anesthetic	Lidocaine hydrochloride	5-20 muM

In the preceding table, muM means micromolar.

Alternatively, these agents might be infused into the lateral ventricle or third ventricle of the brain or just beneath the dura above the cortex or in the intrathecal space. In this instance the drug would diffuse to the appropriate site of action.

Microprocessor 100 within device 10 can be programmed so that a controlled amount of drug can be delivered to the specific brain sites described in Tables I and II. Alternatively, sensor 130 can be used with a closed loop feedback system in order to automatically determine the level of drug delivery necessary to alleviate the hyperexcitation as described in connection with FIG. 3.

By using the foregoing techniques, neurodegenerative disorders can be controlled in a manner previously unattainable.

Those skilled in that art will recognize that the preferred embodiments may be altered or amended without departing

from the true spirit and scope of the invention, as defined in the accompanying claims.

We claim:

1. A system for treating a neurodegenerative disorder resulting in degenerating neurons forming part of a central nervous system comprising in combination:

an implantable pump;

a catheter having a proximal end coupled to said pump and a discharge portion for infusing into a predetermined infusion site in said central nervous system a therapeutic dosage of at least one drugs capable of altering the level of excitation of neurons in said brain related to a degenerating neurons;

a sensor for generating a signal related to an attribute of said nervous system which indicates the hyperexcitation of said degenerating neurons or of neurons related to said degenerating neurons; and

control means responsive to said sensor signal for regulating said therapeutic dosage.

2. A system, as claimed in claim 1, wherein said sensor comprises means for detecting changes in electromagnetic waves generated by muscle or nerve tissue.

3. A system, as claimed in claim 1, wherein said sensor comprises means for detecting the extent of the hyperexcitation of the glutamatergic neurons of said brain.

4. A system, as claimed in claim 1, wherein said control means comprises a microprocessor.

5. A system, as claimed in claim 1, wherein said control means comprises an electrical filter.

6. A system, as claimed in claim 1, wherein said control means comprises means for increasing said therapeutic dosage in response to an increase in said hyperexcitation and for decreasing said therapeutic dosage in response to a decrease in said hyperexcitation.

7. A system, as claimed in claim 1, wherein said predetermined infusion site is selected from the group consisting of the brain, the cerebral ventricle, the subdural space or the intrathecal space.

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UNITED STATES PATENT AND TRADEMARK OFFICE
CERTIFICATE OF CORRECTION

PATENT NO. : 5,814,014
DATED : Sep.29, 1998
INVENTOR(S) : Dennis D. Elsberry et al.

It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

- C. 7 L. 11 "at least one drugs" to be changed to "at least one drug"
C. 8 L. 13 "dosage in response" to be change to "dosage in response"

C. 7 L. 12 "neurons in said brain" to be changed to "neurons in a brain"
C. 7 L. 16 "said nervous system" to be changed to "said central nervous system"

Signed and Sealed this
Sixteenth Day of November, 1999

Attest:



Q. TODD DICKINSON

Attesting Officer

Acting Commissioner of Patents and Trademarks