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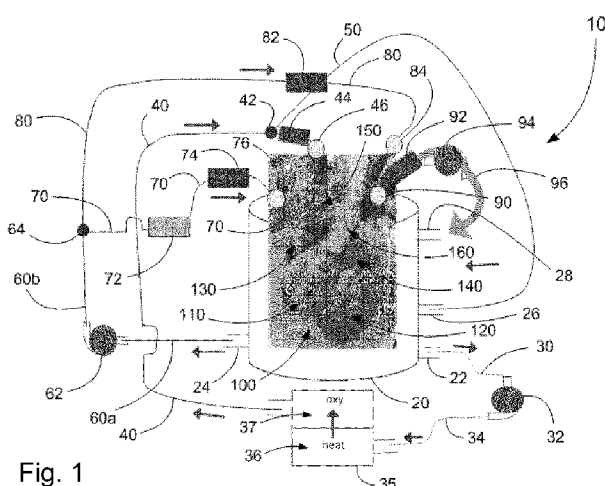


Fig. 1

(57) Abstract: An apparatus, a system, and methods for maintaining and monitoring an excised donor heart. The apparatus comprises a first component for receiving and submerging therein an excised heart in a constantly circulating perfusate solution and a second component comprising equipment for adjusting the temperature and oxygen content of the perfusate solution. The first component comprises an integral pair of defibrillating pads. A first conduit infrastructure interconnects the first module, the second module and an aorta of the excised donor heart pushing a perfusion solution from the first module through the second module into the aorta. The second conduit infrastructure connects the first module with the right atrium and the left atrium for pushing the perfusion solution from the first module into the atria. The third conduit infrastructure connects the first module with the pulmonary artery and provides an after pressure to the flow of the perfusion solution from the pulmonary artery.

TITLE: APPARATUS FOR MAINTENANCE OF HARVESTED HEARTS  
FOR TRANSPLANTING

FIELD OF THE INVENTION

The present invention pertains to apparatus, systems, and methods for *ex vivo* perfusion and maintenance of harvested donor hearts, and more particularly, to pre-transplant assessment of harvested donor hearts for their suitability for transplantation.

BACKGROUND OF THE INVENTION

Heart failure affects 10% of North Americans and is the leading hospital discharge diagnosis. The diagnosis of heart failure is accompanied by a survival outlook that is comparable to a major cancer. There are limited rehabilitation options available to patients who are suffering with heart failure, and few strategies actually re-power the heart. Cardiac transplantation remains the gold-standard therapeutic intervention for patients with end-stage heart failure, with an increasing number of individuals being added to the transplant wait list every year. However, wider application of this life-preserving intervention is limited by the availability of donors. Data from the International Society of Heart and Lung Transplantation Registry shows that cardiac transplantation is in progressive decline in suitable donors (2007, *Overall Heart and Adult Heart Transplantation Statistics*). Two hundred and fifty eight Canadians have died during the last decade (2000 - 2010; Heart and Stroke Foundation of Canada) while waiting for heart transplantation. Similarly, in the United States, 304 patients died in 2010 alone while waiting for heart transplantation (Organ Procurement and Transplantation Network, US Dept. of Health & Human Services). This phenomenon is primarily due to a shortage of suitable organ donors, and is being experienced across the globe.

Time is of the essence for removal of a heart from a donor and its successful transplantation into a recipient. The following principles generally apply for optimal donor heart preservation for the period of time between removal from the donor and transplantation: (i) minimization of cell swelling and edema, (ii) prevention of intracellular acidosis, (iii) prevention of injury caused by oxygen free radicals, and (iv) provision of substrate for regeneration of high-energy phosphate compounds and ATP during reperfusion. The two main sources of donor hearts for transplantation are breathing patients who have suffered irreversible loss of brain function as a result of blunt head trauma or intracerebral hemorrhage and are classified as "brainstem-dead" donors, and patients who have suffered circulatory death and are referred to as "non-heart-beating" donors.

Brainstem-dead organ donors can be maintained under artificial respiration for extended periods of time to provide relative hemodynamic stability up throughout their bodies until the point of organ retrieval. Therefore, cardiac perfusion is uncompromised and organ functionality is theoretically maintained. However, brainstem death itself can profoundly affect cardiac function. The humoral response to brainstem death is characterized by a marked rise in circulating catecholamines. Physiological responses to this "catecholamine storm" include vasoconstriction, hypertension and tachycardia, all of which increase myocardial oxygen demand. In the coronary circulation Significant increased levels of catecholamine circulating throughout the vascular system induce vasoconstriction which in turn, compromises myocardial oxygen supply and can lead to subendocardial ischemia. This imbalance between myocardial oxygen supply and demand is one factor implicated in the impairment of cardiac function observed following brainstem death (Halejcio-Delophont et al., 1998, *Increase in myocardial interstitial adenosine and net lactate production in brain-dead pigs: an in vivo microdialysis study*. Transplantation 66(10):1278-1284; Halejcio-Delophont et al., 1998, *Consequences of brain death on coronary blood flow and myocardial metabolism*. Transplant Proc. 30(6):2840-2841). Structural myocardial damage occurring after brainstem death is characterized by myocytolysis, contraction band necrosis, sub-endocardial hemorrhage, edema and

interstitial mononuclear cell infiltration (Baroldi et al., 1997, *Type and extent of myocardial injury related to brain damage and its significance in heart transplantation: a morphometric study*. J. Heart Lung Transplant 16(10):994-1000). In spite of no direct cardiac insult, brainstem-dead donors often exhibit  
5 reduced cardiac function and the current views are that only 25% of hearts can be recovered from this donor population for transplantation.

Well-defined criteria have been developed for harvesting organs for transplantation from non-heart-beating donors (Kootstra et al., 1995, *Categories of non-heart-beating donors*. Transplant Proc. 27(5):2893-2894; Bos, 2005,  
10 *Ethical and legal issues in non-heart-beating organ donation*. Transplantation, 2005. 79(9): p. 1143-1147). Non-heart-beating donors have minimal brain function but do not meet the criteria for brainstem death and therefore, cannot be legally declared brainstem dead. When it is clear that there is no hope for meaningful recovery of the patient, the physicians and family must be in  
15 agreement to withdraw supportive measures. Up to this point in care, non-heart-beating patients are often supported with mechanical ventilation as well as intravenous inotropic or vasopressor medication. However, only those with single system organ failure (neurologic system) can be considered for organ donation. Withdrawal of life support, most commonly the cessation of mechanical  
20 ventilation, is followed by anoxic cardiac arrest after which, the patient must remain asystolic for five minutes before organ procurement is allowed. Consequently, non-heart-beating donors are necessarily exposed to variable periods of warm ischemia after cardiac arrest which may result in various degrees of organ damage. However, provided that the time duration of warm ischemia is  
25 not excessive, many types organs, i.e., kidneys, livers, and lungs, harvested from non-heart-beating donors are able to recover function after transplantation with success rates that approximate those for transplanted organs from brainstem-dead beating donors.

Numerous perfusion apparatus, systems and methods have been developed  
30 for *ex vivo* maintenance and transportation of harvested organs. Most employ hypothermic conditions to reduce organ metabolism, lower organ energy

requirements, delay the depletion of high energy phosphate reserves, delay the accumulation of lactic acid, and retard morphological and functional deteriorations associated with disruption of oxygenated blood supply. Harvested organs are generally perfused in these systems with preservative solutions comprising antioxidants and pyruvate under low temperatures to maintain their physiological functionality. However, it has been found that increasing amounts of free radicals and catalytic enzymes are produced during extended maintenance of harvested organs in pulsing pressurized hypothermic systems. Fluctuating perfusion pressures in such systems can damage the organs by washing off their vascular endothelial lining and traumatize the underlying tissues. Furthermore, the harvested organs will elute increasing amounts of intracellular, endothelial and membrane constituents resulting in their further physiological debilitation.

The short-comings of hypothermic apparatus, systems and methods have been recognized by those skilled in these arts, and alternative apparatus, systems and methods have been developed for preservation and maintenance of harvested organs at temperatures in the range of about 25° C to about 35° C, commonly referred to as "normothermic" temperatures. Normothermic systems typically use perfusates based on the Viaspan formulation supplemented with one or more of serum albumin as a source of protein and colloid, trace elements to potentiate viability and cellular function, pyruvate and adenosine for oxidative phosphorylation support, transferrin as an attachment factor; insulin and sugars for metabolic support, glutathione to scavenge toxic free radicals as well as a source of impermeant, cyclodextrin as a source of impermeant, scavenger, and potentiator of cell attachment and growth factors, a high  $Mg^{++}$  concentration for microvessel metabolism support, mucopolysaccharides for growth factor potentiation and hemostasis, and endothelial growth factors (Viaspan comprises potassium lactobionate,  $KH_2PO_4$ ,  $MgSO_4$ , raffinose, adenosine, glutathione, allopurinol, and hydroxyethyl starch). Other normothermic perfusion solutions have been developed and used (Muhlbacher et al., 1999, *Preservation solutions for transplantation*. Transplant Proc. 31(5):2069-2070). While harvested kidneys and livers can be maintained beyond twelve hours in normothermic systems, it has

become apparent that normothermic bathing, and maintenance of harvested hearts by pulsed perfusion beyond 12 hours results in deterioration and irreversible debilitation of the hearts' physiological functionality. Another disadvantage of using normothermic continuous pulsed perfusion systems for maintenance of harvested hearts is the time required to excise the heart from a donor, mount it into the normothermic perfusion system and then initiate and stabilize the perfusion process. After the excised heart has been stabilized, its physiological functionality is determined and if transplantation criteria are met, then the excised heart is transported as quickly as possible to a transplant facility.

Current technologies employ occlusive roller pumps to provide flow of perfusate into an isolated aortic root. With this approach, the heart cannot eject against the pump without a significant rise in systolic stress. Furthermore, there currently is no device in the market that allows comprehensive assessment of right and left ventricular systolic and diastolic function, in addition to providing metabolic assessments of excised hearts.

## SUMMARY OF THE INVENTION

The present disclosure pertains to an apparatus, a system, and methods for maintenance and monitoring of the physiological functionality of an excised donor heart.

The apparatus comprises a first component for receiving and submerging an excised heart in a constantly circulating perfusate solution, a second component comprising equipment for adjusting the temperature and oxygen content of the perfusate solution, a third component comprising a non-occlusive centrifugal pump to pump perfusate into an isolated aortic root of an excised heart during preservation mode and to provide non-occlusive resistance to ejection (afterload) during working/assessment mode, and a fourth component comprising a non-occlusive centrifugal pump to provide filling of the excised heart (preload) during working/assessment mode. By positioning the pumps below the heart, coupled with the non-occlusive nature of the pumps, decompression of the excised

heart is provided in the event of poor cardiac function or arrhythmias. The need for gravity as an energy source for provision of preload or afterload to excised hearts is obviated in the current design, thus permitting a compact, portable design for the apparatus of the present disclosure.

5           The system generally comprises the apparatus into which an excised heart is installed, wherein the apparatus is interconnected with: (i) a perfusate pumping system, (ii) flow sensors for monitoring the flow of perfusate to and from the installed heart's aorta, pulmonary artery, pulmonary vein, and vena cava, (iii) an ECG apparatus interconnectable with the excised heart, and (vv) probes  
10           interconnecting the installed heart with instruments for monitoring the excised heart's physiological functionality using load independent indices and load dependent indices.

#### BRIEF DESCRIPTION OF THE DRAWINGS

          The present invention will be described in conjunction with reference to the  
15           following drawings in which:

          Fig. 1 is a schematic illustration of an exemplary maintenance apparatus for harvested donor hearts, according to one embodiment of the present disclosure; and

          Fig. 2 is a close-up partial view of exemplary embodiments of the  
20           pacemaker, ECG monitor, and defibrillator components of the harvested donor heart maintenance apparatus of the present disclosure.

#### DESCRIPTION OF THE INVENTION

          Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the  
25           art to which this invention belongs. In order that the invention herein described may be fully understood, the following terms and definitions are provided herein.

The word “comprise” or variations such as “comprises” or “comprising” will be understood to imply the inclusion of a stated integer or groups of integers but not the exclusion of any other integer or group of integers.

5 The term “about” or “approximately” means within 20%, preferably within 10%, and more preferably within 5% of a given value or range.

The term “modulate” as used herein means to regulate the operation of a device by increasing a signal to the device in order to increase an output by the device, or by decreasing a signal to the device in order to decrease an output by the device

10 The term “afterload” means the mean tension produced by a chamber of the heart in order to contract. It can also be considered as the ‘load’ that the heart must eject blood against. Afterload is therefore a consequence of aortic large vessel compliance, wave reflection and small vessel resistance (left ventricular afterload) or similar pulmonary artery parameters (right ventricular afterload).

15 The term “preload” refers to the stretching of a single cardiac myocyte immediately prior to contraction and is therefore related to the sarcomere length. Since sarcomere length cannot be determined in the intact heart, other indices of preload such as ventricular end diastolic volume or pressure are used. As an example, preload increases when venous return is increased.

20 The term “cardiac myocyte” means a cardiac muscle cell.

The term “stroke volume” (SV) means the volume of blood ejected by the right/left ventricle in a single contraction. It is the difference between the end diastolic volume (EDV) and the end systolic volume (ESV). Mathematically,  $SV = EDV - ESV$ . The stroke volume is affected by changes in preload, afterload and  
25 inotropy (contractility). In normal hearts, the SV is not strongly influenced by afterload whereas in failing hearts, the SV is highly sensitive to afterload changes.

The term “stroke work” (SW) refers to the work performed by the left or right ventricle to eject the stroke volume into the aorta or pulmonary artery,



respectively. The area enclosed by the pressure/volume loop is a measure of the ventricular stroke work, which is a product of the stroke volume and the mean aortic or pulmonary artery pressure (afterload), depending on whether one is considering the left or the right ventricle.

5           The term “ejection fraction” (EF) means the fraction of end diastolic volume that is ejected out of the ventricle during each contraction. Mathematically,  $EF = SV/EDV$ . Healthy ventricles typically have ejection fractions greater than 0.55. Low EF usually indicates systolic dysfunction and severe heart failure can result in EF lower than 0.2. EF is also used as a clinical  
10 indicator of the inotropy (contractility) of the heart. Increasing inotropy leads to an increase in EF, while decreasing inotropy decreases EF.

          The term “end systolic pressure volume relationship” (ESPVR) describes the maximal pressure that can be developed by the left ventricle at any given left ventricular volume, or alternatively, by the right ventricle at any given right  
15 ventricular volume. This implies that the PV loop cannot cross over the line defining ESPVR for any given contractile state. The slope of ESPVR ( $E_{es}$ ) represents the end-systolic elastance, which provides an index of myocardial contractility. The ESPVR is relatively insensitive to changes in preload, afterload and heart rate. This makes it an improved index of systolic function over other  
20 hemodynamic parameters like ejection fraction, cardiac output and stroke volume. The ESPVR becomes steeper and shifts to the left as inotropy (contractility) increases. The ESPVR becomes flatter and shifts to the right as inotropy decreases.

          The term “preload recruitable stroke work relationship” (PRSW) means a  
25 measure of cardiac contractility, and is the linear relationship between SW and EDV.

          The term “pressure-volume area” (PVA) means the total mechanical energy generated by ventricular contraction. This is equal to the sum of the stroke

work (SW), encompassed within the PV loop, and the elastic potential energy (PE). Mathematically,  $PVA = PE + SW$ .

5 The term “Langendorff perfusion” refers to a method of perfusing an excised heart with a nutrient-rich oxygenated solution in a reverse fashion via the aorta. The backwards pressure causes the aortic valve to shut thereby forcing the solution into the coronary vessels, which normally supply the heart tissue with blood. This feeds nutrients and oxygen to the cardiac muscle, allowing it to continue beating for several hours after its removal from the animal.

10 The term “working heart” as used herein, refers to clinical *ex vivo* coronary perfusion throughout a excised heart by ventricular filling via the left atrium and ejection from the left ventricle via the aorta driven by the heart’s contractile function and regular cardiac rhythm. The excised heart is attached by cannulae to a perfusate reservoir and circulatory pumps in a Langendorff preparation. The flow of perfusate through the excised heart in “working heart”  
15 mode is in the direction opposite to the flow of perfusate during Langedorff perfusion.

The term “ischemia” means a condition that occurs when blood flow and oxygen are kept from the heart.

The term “conduit” as used herein means tubing and/or cannula.

20 The present disclosure pertains to apparatus, systems and methods for maintaining an excised heart under continuous Langendorff perfusion until transplantation. The apparatus and systems are communicable and cooperable with cardiac monitoring equipment and microprocessors for monitoring the physiological condition and functioning of the excised heart.

25 One embodiment of the present disclosure pertains to an exemplary modular apparatus for receiving and maintaining an excised heart under continuous Langendorff perfusion until transplantation. The exemplary apparatus comprises two modules. The first module comprises a hard-shell reservoir, also

referred to herein as a reservoir, for housing therein an excised heart under constant bathing with a suitable perfusate solution. The excised heart is mounted onto a stand and submerged within the hard-shell reservoir. The hard-shell reservoir is provided with four ports (i.e., two egress ports and two ingress ports) that are sealingly engageable by conduits that have been interconnected to the excised heart's right atrium, left atrium, aorta, and pulmonary artery. The second module is a perfusate conditioning apparatus comprising: (i) a heat-exchanger for warming and maintaining the perfusate solution at a user-specified temperature (typically referred to as a normothermic temperature), and (ii) an oxygenator for maintaining the dissolved oxygen levels in the perfusate solution above 95% saturation, and maintaining the pH balance through addition of carbon dioxide. The two modules are interconnected by a conduit infrastructure that is engageable by a pump such as those exemplified by centrifugal pumps. Suitable centrifugal pumps are exemplified by ROTAFLOW<sup>®</sup> centrifugal pumps (ROTAFLOW is a registered trademark of Maquet Cardiopulmonary AG Corp., Hirrlingen, Fed. Rep. Ger.), by Medtronic's centrifugal blood BIO-PUMP<sup>®</sup>s BIO-PUMP is a registered trademark of Medtronic Bio-Medicus Inc., Minnetonka, MN, USA), by Sorin's RevOlution 5 blood pump (Sorin Group USA, Arvada, CO, USA). In operation, the centrifugal pump provides a constant flow of perfusate solution from the first module (i.e., the hard-shell reservoir) to the second module (i.e., the perfusate conditioning apparatus). The first module is additionally provided with ports for receiving therethrough leads from cardiac monitoring equipment for engaging specific sites on and/or in the excised heart. Each module can be separately assembled and prepared for use multiple units, thereby facilitating rapid assembly and configuration of the apparatus as needed to receive and maintain an excised heart.

An exemplary apparatus 10 according to one embodiment of the present disclosure is shown in Fig. 1. The apparatus comprises: (i) a first component which is a hard-shell reservoir 20 housing a removable support (not shown) for mounting thereon and therein an excised heart 100, and (ii) a second component which is a perfusate solution conditioning device 35 comprising a heat-exchanger

36 and an oxygenator 37. The hard-shell reservoir 20 may additionally have a level sensor (not shown) for monitoring the level of perfusate solution in the hard-shell reservoir 20. The two components are interconnected by a first conduit infrastructure comprising: (i) an egress line 30 that is sealably engageable at one end with a port 22 provided therefore near the bottom of the hard-shell reservoir 20 and is sealably engageable at its other end with the inlet of a first centrifugal pump 32. The outlet of the first centrifugal pump 32 is sealably engaged with a line 34 that is sealably engageable with an inlet to the heat exchanger 36 of the perfusate solution conditioning device 35. A line 40 is sealably engageable with an outlet from the oxygenator 37 of the perfusate solution conditioning device 35. The other end of line 40 is sealably engageable with a Y-connector 42 which diverts a portion of the flow of conditioned perfusion solution from the perfusion solution conditioning device 35 into a purge line 50 that is sealably engageable with a first ingress port 26 provided therefore on the hard-shell reservoir 20. The remainder of the flow of perfusate solution conditioning device 35 is diverted by the Y-connector 42 into a flow sensor 44 interconnected with an integrated pressure port 46 that is clampable into the aorta 150 of the harvested heart. In operation, the flow sensor 44 measures aortic flow of the conditioned perfusion solution from the perfusate solution conditioning device 35 into the aorta 150. Perfusion solution egressing from the hard-shell reservoir 20 into the perfusate solution conditioning device 35 is conditioned by heating in the heat exchanger 36 to a normothermic temperature from the range of about 20° C to about 37° C and then is oxygenated by oxygenator 37 prior to flowing into line 40 for conveyance into the aorta 150. The diastolic pressure in the aorta 150 can be specified and tightly regulated by computer controlled feedback to modulate the centrifugal pump 32. During assessment mode, with provision of flow into the left atrium the heart ejects the perfusion solution back through line 40 with the centrifugal pump 32 providing resistance (afterload). In this manner the heart can beat against an afterload pressure that is delivered by the flow of perfusate solution from the centrifugal pump 32.

A second conduit infrastructure comprises a line 60a sealably engageable at one end with a second egress port 24 provided therefore near the bottom of the hard-shell reservoir 20, and its other end sealably engageable with the inlet into a second centrifugal pump 62. The outlet of the second centrifugal pump 62 is sealably engageable with a line 60b that terminates in a Y-connector 64. Y-connector 64 splits the pressurized flow of perfusion solution into two lines 70, 80. Line 70 is interconnected with, firstly, an occlusion clamp 72, secondly, a flow sensor 74, and thirdly, an integrated pressure port 76. The terminal end of line 70 is insertable into the right atrium 130 of the harvested heart 100. It should be noted that occlusion clamp 72 is preferably a servo-actuated partial occlusion clamp whose variable positions enables regulation of the rate of flow of the perfusion solution into the right atrium 130 and therefore, can also be used to modulate pressure delivered to the harvested heart 100. Line 80 is interconnected with, firstly, a flow sensor 82, and secondly, an integrated pressure port 84. The terminal end of line 80 is insertable into the left atrium 140 of the harvested heart 100. It should be noted that lines 70, 80 are additionally provided with bubble detectors (not shown). During the assessment mode, pump 62 provides flow of the perfusate solution into the right atrium and left atrium (preload pressure) under a feedback loop from pressure ports 84, 76 with differential control of flow into the right atrium and left atrium being provided by modulation of clamp 72. In the event of overpressurization of the heart as a consequence, for example, of arrhythmia or poor cardiac function, the flow of perfusate solution from pump 62 is decreased thereby allowing decompression of the heart to occur through passive retrograde flow of the perfusate solution back through the pump 62.

A third conduit infrastructure comprises a line 96 that is clampable into the pulmonary artery 160 of the harvested heart 100. The line 96 is sequentially sealably engageable with an integrated pressure port 90, a flow sensor 92, and a third centrifugal pump 94. The terminal end of the line 96 is sealably engageable with the second ingress port 28 provided therefore on the hard-shell reservoir 20. Pump 94 provides resistance (afterload pressure) to the right ventricle, through

computer-controlled modulation of the pump 94 in reference to feedback from pressure port 90.

Fig. 2 illustrates exemplary monitoring and maintenance equipment for maintaining a harvested heart in a functional condition during storage and transport in the exemplary apparatus of the present disclosure. Leads 172 from an ECG monitoring device 170 are engageable with, for example, the right ventricle 110 and the left ventricle 120 of a harvested heart 100 for monitoring the electrical activity of the harvested heart 100. Alternatively, the ECG leads may be integrally incorporated into the walls of the hard-shell reservoir 20. Leads 192 from a dual-chamber pacemaker 190 are engageable with the right atrium 130 and the right ventricle 110 of the harvested heart 100. Although a dual-chamber pacemaker is preferable for use with the apparatus 10 of the present disclosure, it is optional to substitute a single-chamber pacemaker having a single lead that is engageable with the right atrium or the right ventricle. Two defibrillator pads 184 are integrally provided opposite each other on the inner surfaces of the hard-shell reservoir 20 and are connected by leads 182 to a defibrillator. The ECG monitoring device 170, the pacemaker 190 and the defibrillator 180 may be mounted on a support provided therefore (not shown) that is an integral component of the hard-shell reservoir 20. Alternatively, the ECG monitoring device 170, the pacemaker 190 and the defibrillator 180 may be integrally incorporated into the housing of a transportation container configured to receive therein the hard-shell reservoir 20.

As soon as an excised heart 100 is mounted onto the removable support and placed into the hard-shell reservoir 20, the terminal end of line 40 is clamped into the aorta, 150, line 70 is inserted into the right atrium 130, line 80 is inserted into the left atrium 140, and line 96 is clamped into the pulmonary artery 160. Then, a suitable perfusion solution exemplified by whole blood, whole blood amended with citrate and/or phosphate and/or dextrose, modified Krebs solutions, Viaspan, modified Viaspan solutions, and the like, is added into the hard-shell reservoir 20 until the heart 100 is completely submerged. It should be noted that the hard-shell reservoir 20 may be additionally provided with a level sensor (not

shown) and a supplementary supply of the perfusion solution (not shown) for conveyance into the hard-shell reservoir 20 as need to maintain the excised heart 100 fully submerged during storage and transport in the apparatus of the present disclosure.

5           When in operation, the pump 32 continuously draws the perfusate solution from the hard-shell reservoir 20 from egress port 22 into line 30 into the perfusate solution conditioning device 35 wherein the perfusate solution is conditioned by warming to a normothermic temperature and then, is oxygenated. The conditioned and pressurized conditioned perfusate solution is then conveyed to Y-connector  
10 42 that diverts a portion of the conditioned perfusate solution into purge line 50 for conveyance through ingress port 26 back into the hard-shell reservoir 20 where it circulates about and baths the heart 100. The remaining flow of pressurized conditioned perfusate solution is conveyed through flow sensor 44 and integrated pressure port 46 into the aorta 150. It is to be noted that the purge  
15 line 50 is positioned to be the highest point in the assembled apparatus 10 when an excised heart 100 is mounted therein so that any air that is ejected by the heart immediately goes out via the purge line 50 and back to the hard-shell reservoir 20.

          A preload centrifugal pump 62 draws the perfusion solution out of the hard-shell reservoir through egress port 24 into line 60b and then pushes the  
20 perfusion solution to Y-connector 64 where its flow is split into two lines 70,80. The perfusion solution is pushed through line 70 through a computer-controlled servo-actuated partial occlusion clamp 72, a flow sensor 74, and an integrated pressure port 76 into the right atrium 130. The variable positions of the servo-actuated partial occlusion clamp 72 enables precise regulation of the rate of flow  
25 of the perfusion solution into the right atrium 130. The perfusion solution is concurrently pushed through line 80 through a flow sensor 82, and an integrated pressure port 84 into the left atrium 140.

          The pressurised perfusion solution flowing into the aorta 150, right atrium 130, and left atrium 120 flows into the right ventricle 140, and then out through  
30 the pulmonary artery 160 into line 96 through, firstly, an integrated pressure port

90, secondly, a flow meter 92, thirdly, an afterload centrifugal pump 94 to regulate the right ventricular afterload pressure (which is measured by the flow meter 92), and finally, back into the hard-shell reservoir 20 through ingress port 28. The pressurized flow of conditioned perfusion solution into the aorta 150 via line 40 is supplied by centrifugal pump 32 and is monitored by aortic flow sensor 44. The pressurized flow of conditioned perfusion solution into the aorta 150 and then out of the pulmonary artery 160 will maintain the heart 100 in a Langendorff, isolated root perfusion state. To maintain and assess the heart's function in working mode, tight regulation of preload is required. Therefore lines 70, 80 connected to the right atrium and left atrium, respectively, comprise 3/8" tubing and receive pressurized flow of perfusion solution from the preload pump 62. Right atrial flow pressure is monitored by flow sensor 74 while left atrial flow pressure is monitored by flow sensor 82. The computer-controlled servo-actuated partial occlusion clamp 72 enables precise control over the rate of perfusion solution to the right atrium 130 and the left atrium 140, and therefore, the pressure applied to the receiving chamber. The flow meters 44, 74, 82, 92 and the integrated pressure points 46, 76, 84, 90 are connectable to and communicable with a computer for constant monitoring and integrating of the flow rates and pressures to enable constant assessment of cardiac function, i.e., the right ventricular stroke work and the left ventricular stroke work while varying resistance to the flow of perfusion solution (i.e., afterload). It should be noted that the levels of haematocrit,  $\text{Ca}^{++}$ ,  $\text{K}^+$ ,  $\text{NaHCO}_3$ ,  $\text{Na}^+$ ,  $\text{pO}_2$ ,  $\text{CO}_2$ , and glucose in the perfusion solution must be balanced before perfusion starts. In the case of using bank CPD donor blood, deranged  $\text{K}^+$  and  $\text{Ca}^{++}$  concentrations may not allow for a homeostatic prime. This can be adjusted by haemofiltration using Ringers solution as the rinse. All these values should ideally start within normal physiological ranges and should be monitored by inline continuous blood gas analysis. The primary purpose for the perfusion solution is to avoid causing tissue edema and to maintain ion homeostasis to preserve cardiac function.

Another exemplary embodiment of the present disclosure relates to a support for mounting thereon and dismounting therefrom of the modules and the



pumps. The support may additionally have mounts for installation of cardiac monitoring equipment and/or computer equipment and/or monitors for displaying the physiological condition and functioning of the excised heart. The support may be a racking system mounted on wheels so that the apparatus is transportable  
5 within a medical facility, for example between surgical theatres, staging rooms, assembly rooms and disassembly rooms. The support may be cabinet with two opposing side walls and with other two sides having opening doors. Alternatively, the support may be a cabinet with three fixed side walls being opposing walls and having one side with opening doors. The side walls and doors may be insulated  
10 and/or cushioned. The support may be configured for transport by vehicles or by airplanes.

Another exemplary embodiment of the present disclosure relates to a system for receiving, perfusing and maintaining and assessing an excised donor heart. The system generally comprises the above-disclosed apparatus  
15 interconnected with: (i) a perfusate-processing system, (ii) a perfusate pumping system, (iii) flow sensors for monitoring the flow of perfusate to and from an installed heart's aorta, right atrium, left atrium, and pulmonary artery vena cava, (iv) an ECG apparatus interconnectable with the installed heart, (v) a pacemaker interconnectable with the installed heart, (vi) a defibrillator interconnectable with  
20 the pair of defibrillator pads integral with the inner surface of the hard-shell reservoir component of the apparatus, and (vii) probes interconnecting the installed heart with instruments for monitoring the heart's physiological functionality using load independent indices and load dependent indices. Suitable perfusion-processing systems are exemplified by heart-lung machines commonly  
25 used for coronary bypass surgeries.

An exemplary use of the apparatus, system and methods of the present disclosure generally comprises the steps of selection, preparation, and balancing of a perfusate solution, setting up the system by interconnecting the perfusate-processing system and the bi-directional perfusate pumping system with  
30 cannulae that are subsequently interconnected with the appropriate ports on the lid of the receiving, maintaining, and assessing apparatus, priming the interconnected

- system with the perfusate solution, installing an excised heart onto the support provided with the apparatus and then installing the appropriate cannulae into the heart's aorta, pulmonary artery, pulmonary vein, and vena cava, expressing all air from within the heart and the cannulae, and then commencing the Langendorff
- 5 perfusion at a normothermic temperature from the range of about 25° C to about 35° C.

## CLAIMS:

1. A modular perfusion apparatus for maintenance and transport of an excised donor heart, comprising:

5 a first module comprising a hard-shell reservoir with a removable support for positioning and mounting thereon the excised heart, said hard-shell reservoir having a pair of opposing defibrillator pads engaged with an inner surface of the hard-shell reservoir;

a second module comprising a heat-exchanger in communication with an oxygenator;

10 a support for disengagably mounting thereon the first module and the second module;

a first conduit infrastructure interconnecting the first module, the second module and an aorta of the excised donor heart, said first conduit infrastructure having a first centrifugal pump for pushing a perfusion solution from the first  
15 module to the second module;

a second conduit infrastructure for connecting the first module with a right atrium and a left atrium of the excised donor heart, said second conduit infrastructure having a second centrifugal pump for pushing the perfusion solution from the first module to the right atrium and the left atrium; and

20 a third conduit infrastructure for connecting the first module with a pulmonary artery of the excised donor heart, said third conduit infrastructure having a third centrifugal pump for providing an after pressure to a flow of the perfusion solution from the pulmonary artery.

2. The modular perfusion apparatus of claim 1, additionally comprising a  
25 support for disengagably mounting thereon the first module, the second module, the first centrifugal pump, the second centrifugal pump, and the third centrifugal pump.

3. The modular perfusion apparatus of claim 2, wherein the support comprises a housing for encasing the mounted first module, the second module, the first centrifugal pump, the second centrifugal pump, and the third centrifugal pump.
- 5 4. The modular perfusion apparatus of claim 1, additionally comprising an ECG monitoring device.
5. The modular perfusion apparatus of claim 1, additionally comprising cardiac pacemaker.
6. The modular perfusion apparatus of claim 1, additionally comprising a  
10 defibrillator for communicating with the pair of defibrillator pads.

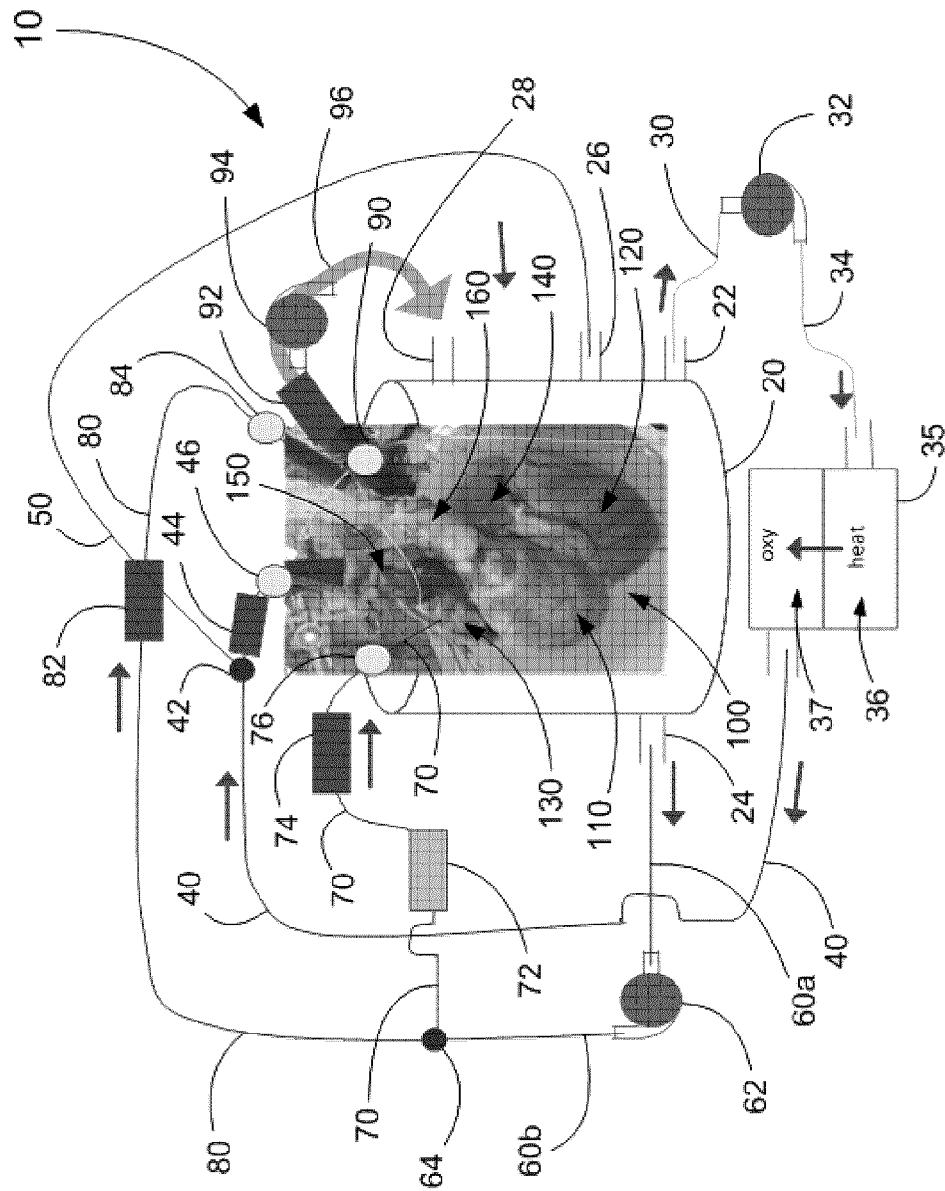


Fig. 1

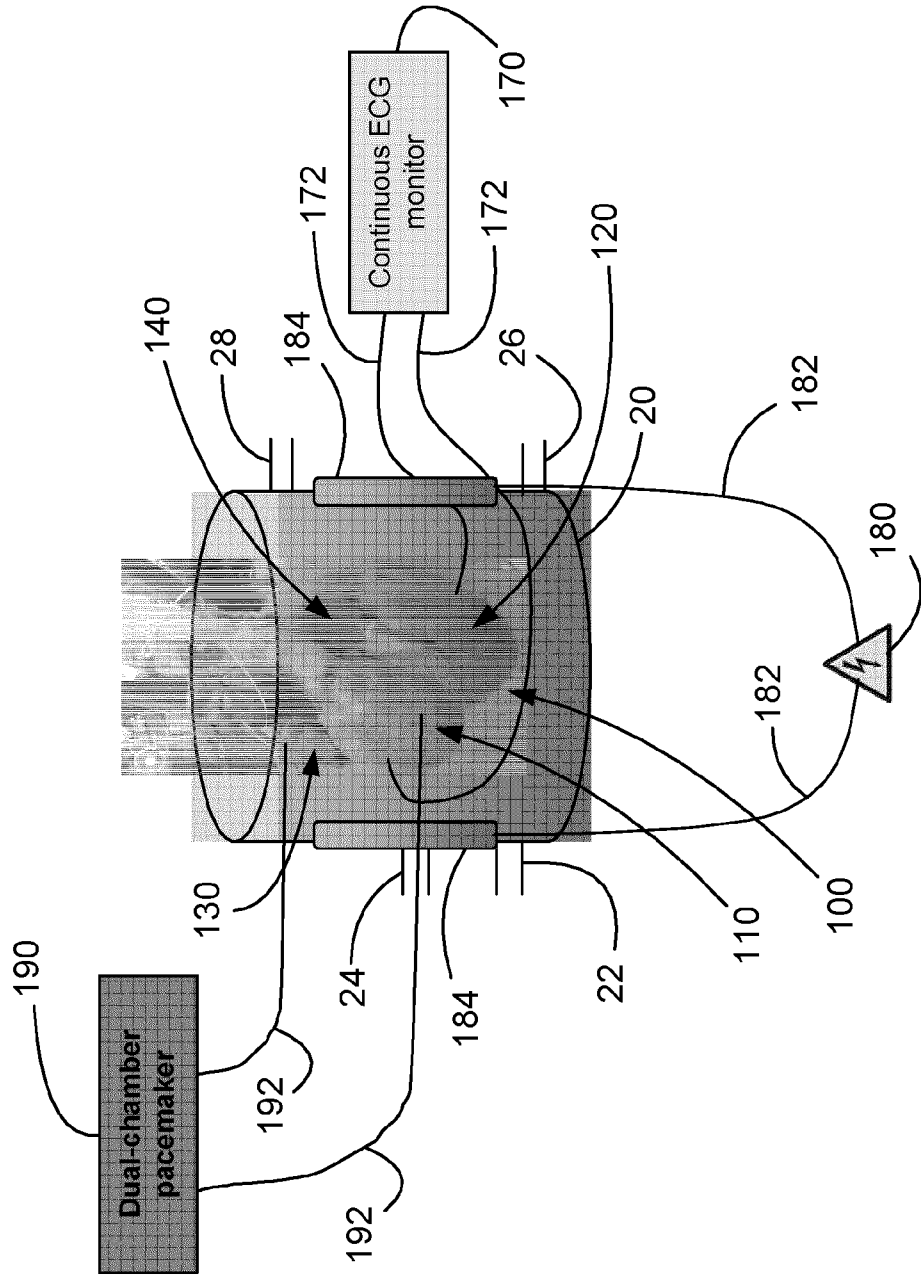


Fig. 2

## INTERNATIONAL SEARCH REPORT

International application No.

**PCT/CA2015/050201**

## A. CLASSIFICATION OF SUBJECT MATTER

IPC: *A01N 1/02* (2006.01), *A61N 1/362* (2006.01), *A61N 1/39* (2006.01)

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC: *A01N 1/02* (2006.01), *A61N 1/362* (2006.01), *A61N 1/39* (2006.01)

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic database(s) consulted during the international search (name of database(s) and, where practicable, search terms used)

Google Patent Search: heart AND preservation AND centrifugal AND (modul\* OR portable OR cassette)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X Y	CA 2861545 A1 (FREED, D. H. et al.) 25 July 2013 (25-07-2013) *see entire document*	1 4, 6 5
Y	US 5807737 A (SCHILL, D. M. et al.) 15 September 1998 (15-09-1998) *see entire document*	5
A	US 2013/0295552 A1 (HASSANEIN, W. et al.) 7 November 2013 (07-11-2013) *see entire document*	1 – 6
A	US 7811808 B2 (VAN DER PLAATS, A. et al.) 12 October 2010 (12-10-2010) *see entire document*	1 – 6
A	US 8585380 B2 (HASSANEIN, W. et al.) 19 November 2013 (19-11-2013) *see entire document*	1 - 6

<input type="checkbox"/>	Further documents are listed in the continuation of Box C.	<input checked="" type="checkbox"/>	See patent family annex.
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* “A” “E” “L” “O” “P”	Special categories of cited documents: document defining the general state of the art which is not considered to be of particular relevance earlier application or patent but published on or after the international filing date document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) document referring to an oral disclosure, use, exhibition or other means document published prior to the international filing date but later than the priority date claimed	“T” “X” “Y” “&”	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art document member of the same patent family
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Date of the actual completion of the international search 08 June 2015 (08-06-2015)	Date of mailing of the international search report 10 June 2015 (10-06-2015)
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Name and mailing address of the ISA/CA Canadian Intellectual Property Office Place du Portage I, C114 - 1st Floor, Box PCT 50 Victoria Street Gatineau, Quebec K1A 0C9 Facsimile No.: 001-819-953-2476	Authorized officer  Steven Kolodziejczyk (819) 997-3239
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**INTERNATIONAL SEARCH REPORT**  
Information on patent family members

International application No.  
**PCT/CA2015/050201**

Patent Document Cited in Search Report	Publication Date	Patent Family Member(s)	Publication Date
CA2861545A1	25 July 2013 (25-07-2013)	CA2861545A1 EP2809153A1 US2015017710A1 WO2013106908A1	25 July 2013 (25-07-2013) 10 December 2014 (10-12-2014) 15 January 2015 (15-01-2015) 25 July 2013 (25-07-2013)
US2013295552A1	07 November 2013 (07-11-2013)	US2013295552A1 AT253819T AU728233B2 AU9504298A CA2304598A1 CA2304598C DE69819759D1 DE69819759T2 DK1017274T3 EP1017274A1 EP1017274B1 ES2210825T3 JP2001516768A JP2011157374A JP2014040491A JP2014148553A JP2014177484A JP2015025023A PT1017274E US6046046A US6100082A US6953655B1 US2005147958A1 US8409846B2 WO9915011A1	07 November 2013 (07-11-2013) 15 November 2003 (15-11-2003) 04 January 2001 (04-01-2001) 12 April 1999 (12-04-1999) 01 April 1999 (01-04-1999) 10 March 2009 (10-03-2009) 18 December 2003 (18-12-2003) 23 September 2004 (23-09-2004) 22 March 2004 (22-03-2004) 12 July 2000 (12-07-2000) 12 November 2003 (12-11-2003) 01 July 2004 (01-07-2004) 02 October 2001 (02-10-2001) 18 August 2011 (18-08-2011) 06 March 2014 (06-03-2014) 21 August 2014 (21-08-2014) 25 September 2014 (25-09-2014) 05 February 2015 (05-02-2015) 30 April 2004 (30-04-2004) 04 April 2000 (04-04-2000) 08 August 2000 (08-08-2000) 11 October 2005 (11-10-2005) 07 July 2005 (07-07-2005) 02 April 2013 (02-04-2013) 01 April 1999 (01-04-1999)
US5807737A	15 September 1998 (15-09-1998)	None	
US7811808B2	12 October 2010 (12-10-2010)	US2007184545A1 AT373952T CA2534112A1 DE602004009189D1 DE602004009189T2 EP1648228A1 EP1648228B1 ES2294540T3 NL1024022C2 WO2005009125A1	09 August 2007 (09-08-2007) 15 October 2007 (15-10-2007) 03 February 2005 (03-02-2005) 08 November 2007 (08-11-2007) 19 June 2008 (19-06-2008) 26 April 2006 (26-04-2006) 26 September 2007 (26-09-2007) 01 April 2008 (01-04-2008) 01 February 2005 (01-02-2005) 03 February 2005 (03-02-2005)
US8585380B2	19 November 2013 (19-11-2013)	US2006160204A1 AT470354T AU2005294206A1 AU2005294206B2 AU2008260409A1 AU2008260409B2 CA2584066A1 CA2685302A1 CN101072500A CN101072500B CN101778560A CN101778560B CN103120154A CN103931605A DE602005021788D1	20 July 2006 (20-07-2006) 15 June 2010 (15-06-2010) 20 April 2006 (20-04-2006) 14 June 2012 (14-06-2012) 11 December 2008 (11-12-2008) 05 September 2013 (05-09-2013) 20 April 2006 (20-04-2006) 11 December 2008 (11-12-2008) 14 November 2007 (14-11-2007) 04 December 2013 (04-12-2013) 14 July 2010 (14-07-2010) 12 June 2013 (12-06-2013) 29 May 2013 (29-05-2013) 23 July 2014 (23-07-2014) 22 July 2010 (22-07-2010)

(Continued on Supplementary Sheet)



## INTERNATIONAL SEARCH REPORT

International application No.

**PCT/CA2015/050201**

(Continuation of Patent Family Members)

DK1768490T3	11 October 2010 (11-10-2010)
EP1768490A2	04 April 2007 (04-04-2007)
EP1768490B1	09 June 2010 (09-06-2010)
EP2150105A2	10 February 2010 (10-02-2010)
EP2150105A4	21 May 2014 (21-05-2014)
ES2348736T3	13 December 2010 (13-12-2010)
HK1101890A1	21 January 2011 (21-01-2011)
HK1145942A1	07 March 2014 (07-03-2014)
IL182403D0	19 August 2007 (19-08-2007)
IL182403A	31 March 2011 (31-03-2011)
IL201739D0	16 June 2010 (16-06-2010)
IL211084D0	28 April 2011 (28-04-2011)
IL211085D0	28 April 2011 (28-04-2011)
JP2008515914A	15 May 2008 (15-05-2008)
JP5113522B2	09 January 2013 (09-01-2013)
JP2014028845A	13 February 2014 (13-02-2014)
JP5462406B2	02 April 2014 (02-04-2014)
JP2010525076A	22 July 2010 (22-07-2010)
JP2012255003A	27 December 2012 (27-12-2012)
NZ554543A	31 March 2011 (31-03-2011)
NZ580648A	27 July 2012 (27-07-2012)
NZ591524A	28 September 2012 (28-09-2012)
NZ597482A	26 July 2013 (26-07-2013)
NZ600702A	20 December 2013 (20-12-2013)
NZ608461A	29 August 2014 (29-08-2014)
NZ614472A	27 March 2015 (27-03-2015)
US2007190636A1	16 August 2007 (16-08-2007)
US7651835B2	26 January 2010 (26-01-2010)
US2007275364A1	29 November 2007 (29-11-2007)
US8304181B2	06 November 2012 (06-11-2012)
US2006154358A1	13 July 2006 (13-07-2006)
US8465970B2	18 June 2013 (18-06-2013)
US2006148062A1	06 July 2006 (06-07-2006)
US2006154357A1	13 July 2006 (13-07-2006)
US2006154359A1	13 July 2006 (13-07-2006)
US2006292544A1	28 December 2006 (28-12-2006)
US2013078710A1	28 March 2013 (28-03-2013)
WO2006042138A2	20 April 2006 (20-04-2006)
WO2006042138A8	28 September 2006 (28-09-2006)
WO2008150587A2	11 December 2008 (11-12-2008)
WO2008150587A3	26 February 2009 (26-02-2009)