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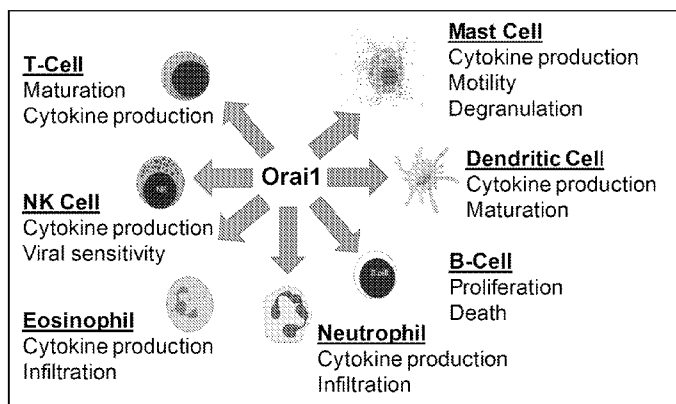
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(54) Title: MODIFIED PEPTIDOMIMETICS AND METHODS OF USE

FIG. 1

Orai1 – Ca²⁺ Channel and Master Regulator of Inflammation

- Orai1 is upstream of all cytokine responses
- Selective/local Orai1 inhibition is immunomodulatory



(57) Abstract: This invention relates synthetic modified polypeptides which bind to the Orai1 calcium channel, and their therapeutic use as lung immunomodulators in disorders such as, but not limited to, viral infections, bacterial infections, allergic responses, asthma, cystic fibrosis and other inflammatory disorders.

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MODIFIED PEPTIDOMIMETICS AND METHODS OF USE**STATEMENT OF PRIORITY**

5 This application claims the benefit, under 35 U.S.C. § 119(e), of U.S. Provisional Application Serial No. 63/164,132, filed March 22, 2021, the entire contents of which are incorporated by reference herein.

STATEMENT OF GOVERNMENT SUPPORT

10 This invention was made with government support under Grant Number AI155107 awarded by the National Institutes of Health. The government has certain rights in the invention.

STATEMENT REGARDING ELECTRONIC FILING OF A SEQUENCE LISTING

15 A Sequence Listing in ASCII text format, submitted under 37 C.F.R. § 1.821, entitled 5470-886WO_ST25.txt, 25,102 bytes in size, generated on March 22, 2022 and filed via EFS-Web, is provided in lieu of a paper copy. This Sequence Listing is hereby incorporated herein by reference into the specification for its disclosures.

FIELD OF THE INVENTION

20 This invention relates to synthetic polypeptides that bind to the Orai1 calcium channel, and their therapeutic use as lung immunomodulators in disorders such as, but not limited to, viral infections, bacterial infections, allergic responses, asthma, cystic fibrosis and other inflammatory disorders.

BACKGROUND OF THE INVENTION

25 Orai1 is a ubiquitously-expressed plasma membrane Ca^{2+} channel, and an essential, proximal component of the immune system/inflammatory cascade. Orai1 is tightly regulated, since too much Orai1 contributes to hyperinflammation, while body-wide inhibition of Orai1 is immunosuppressive. Thus, there is a need for fine-tuned, local modulation of Orai1
30 function.

The present invention overcomes previous shortcomings by providing synthetic polypeptides that bind to the Orai1 calcium channel and reduce its expression and functionality specifically in the lungs while leaving normal systemic Orai1 functioning

unaffected, as well as providing methods of using these synthetic polypeptides to treat disorders responsive to modulating Orai1.

SUMMARY OF THE INVENTION

5 A first aspect of the present invention provides a synthetic polypeptide comprising one or more (e.g., one or more, two or more, at least one, at least two, etc.) of the following amino acid modifications, wherein the numbering is based on the reference amino acid sequence of **SEQ ID NO:1**: 14R substitution; deletion of residues in positions 1-4; deletion of residue in position 16; deletion of valine residue(s); and/or insertion of D-ala residue(s),
10 wherein the synthetic polypeptide binds to a Orai1 plasma membrane Ca^{2+} channel.

Also provided herein are polynucleotides, vectors, cells, compositions, kits and delivery dosage devices comprising the synthetic polynucleotides of the present invention.

Another aspect of the present invention provides a method of inhibiting calcium influx through a calcium channel, comprising contacting the calcium channel with a synthetic
15 polypeptide, polynucleotide, vector and/or composition (e.g., pharmaceutical composition) of the present invention of the present invention, thereby inhibiting calcium influx through the calcium channel.

An additional aspect of the present invention provides a method of reducing expression of an Orai1 calcium channel, comprising contacting the calcium channel Orai1
20 with a synthetic polypeptide, polynucleotide, vector and/or composition (e.g., pharmaceutical composition) of the present invention, thereby reducing expression of the Orai1 calcium channel.

Another aspect of the present invention provides a method of inhibiting an immune response in a subject, comprising delivering to the subject a synthetic polypeptide,
25 polynucleotide, vector and/or composition (e.g., pharmaceutical composition) of the present invention of the present invention, thereby inhibiting the immune response.

Another aspect of the present invention provides a method of inhibiting inflammation in a subject, comprising delivering to the subject a synthetic polypeptide, polynucleotide,
vector and/or composition (e.g., pharmaceutical composition) of the present invention of the
30 present invention, thereby inhibiting the inflammation.

Another aspect of the present invention provides a method of treating or preventing a disorder responsive to inhibition of calcium influx in an airway in a subject in need thereof, comprising delivering to the airway of the subject a therapeutically or prophylactically effective amount of a synthetic polypeptide, polynucleotide, vector and/or composition (e.g.,

pharmaceutical composition) of the present invention of the present invention, thereby treating or preventing the disorder.

Another aspect of the present invention provides a method of treating or preventing a disorder responsive to inhibition of calcium influx in a subject in need thereof, comprising delivering to the subject a therapeutically or prophylactically effective amount of a synthetic polypeptide, polynucleotide, vector and/or composition (e.g., pharmaceutical composition) of the present invention, thereby treating or preventing the disorder.

These and other aspects of the invention are set forth in more detail in the description of the invention below.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows a schematic of cell types influenced by Orai1.

FIG. 2 shows a schematic of Orai1 related signaling. During infection, increased SOCE and Orai1 activity leads to greater phosphorylation of Ca^{2+} -sensitive kinases (e.g., calcineurin) and activation of transcription factors (e.g., NFAT) leading to a pro-inflammatory response. ELD607 binds to and internalizes Orai1, preventing SOCE, despite ongoing upstream receptor engagement.

FIG. 3 shows a schematic of ELD607 inhibitory activity on Orai1 as related to SARS-CoV-2 infection (COVID-19).

FIG. 4 shows a bar graph of Ca^{2+} levels for Alanine scan peptides after thapsigargin addition. The results were normalized to the baseline. Thapsigargin (Tg) at 1 μM was used as control. The inhibition of Ca^{2+} release after scaffold peptide (ELD100) addition is shown for comparison. $N=9$. * $p < 0.02$.

FIG. 5 shows a bar graph of Ca^{2+} levels for peptides that underwent conservative mutations upon thapsigargin addition. The results were normalized to the baseline. Thapsigargin (Tg) at 1 μM was used as control. The inhibition of Ca^{2+} release after ELD100 addition is shown for comparison. $N=9$. * $p < 0.02$.

FIG. 6 shows a bar graph of Ca^{2+} levels upon thapsigargin addition for peptides that were size-optimized. The results were normalized to the baseline. Thapsigargin (Tg) at 1 μM was used as control. The inhibition of Ca^{2+} release after ELD100 addition is shown for comparison. $N=9$. * $p < 0.0001$.

FIG. 7 shows a bar graph of Ca^{2+} levels upon thapsigargin addition for peptides that were size-optimized and capped for stability. The results were normalized to the baseline.

Thapsigargin (Tg) at 1 μ M was used as control. The inhibition of Ca^{2+} release after ELD100 addition is shown for comparison. N=9. * $p < 0.05$.

FIG. 8 shows a graph of cytoplasmic Ca^{2+} of HEK293T cells treated with the noted peptides.

FIG. 9 shows a graph of ELD604 and ELD607 resistance to neutrophil elastase-mediated degradation. Peptides were exposed to NE over time, and the reaction was stopped by addition of sivelestat. HPLC was then performed in order to determine peptide stability. N=3-4 replicates per time point.

FIG. 10 shows a 3D graph comparing elastase stability, efficacy, and size for the 600 series of compounds vs. the ELD100 scaffold.

FIG. 11 shows that ELD607 is stable in CF sputum. **FIG. 11 panel A** shows a graph of HPLC analysis demonstrating a peak at ~4.5 minutes that corresponds to ELD607. **FIG. 11 panel B** shows a bar graph of NL and CF sputum each pooled from 3 different donors. ELD607 was incubated in NL and CF sputum for 6 h at 37°C vs PBS (control) and samples were passed through a 10 kDa spin column to halt the reaction. The area under the curve was similar for each of the conditions. **FIG. 11 panel C** shows a graph of ELD607 or ELD100 incubated in either PBS or CF sputum and added to HEK293T cells. 1 h later, thapsigargin was added and the change in Fluo-4 fluorescence was measured. Data show the peak Fluo-4 response. Control is addition of thapsigargin alone. * $p < 0.001$ different to control.

FIG. 12A shows a graph indicating that ELD607 does not interact with mucus. A Quartz Crystal Microbalance with Dissipation (QCM-D) assay was used to measure human airway dissipation and frequency as indicators of mucus-cross linking and mass, respectively. ELD607 did not alter these properties. In contrast, poly-L-lysine (positive control) bound to mucus, which altered cross-linking and mass. **FIG. 12B** shows a bar graph of summary data for QCM-D frequency for the peptides indicated. **FIG. 12C** shows a bar graph of summary data for QCM-D dissipation for the peptides indicated.

FIG. 13 shows fluorescence measurements indicating that Orai1 knockdown prevents ELD607 from inhibiting SOCE. HEK293T cells were transfected with anti-Orai1 shRNA for 48 h and the change in Fluo4 fluorescence was measured. Cells were then exposed to vehicle or 10 mM ELD607 for 1 h. Thapsigargin was then added followed by extracellular Ca^{2+} to differentiate ER Ca^{2+} release and SOCE. $n=8$ cultures per condition.

FIG. 14 shows a graph of dose response curves for ELD607 and ELD100 based on Ca^{2+} influx rather than global Ca^{2+} . HEK293T cells were cultured in 384 well plates. Cells were then exposed to vehicle or 10 mM ELD607 for 1 h. 100 nM thapsigargin was then

added followed by extracellular Ca^{2+} to differentiate ER Ca^{2+} release and SOCE. $n=8-12$ cultures per data point.

FIGS. 15A-15D show (**FIG. 15A**) a schematic of a murine model of *P. aeruginosa* (top left panel), and bar graphs of (**FIG. 15B**) BAL monocytes, (**FIG. 15C**) BAL neutrophils, and (**FIG. 15D**) bacterial counts in whole lung at 24 hours quantified following experiments according to the mouse model as schematized and conditions as noted on the x-axis.

FIGS. 16A-16G show data from further *P. aeruginosa* infection experiments. **FIG. 16A** shows bar graphs of lung cytokine levels (left, MIP-2/CXCL2; middle, IL-17/CTLA-8; right KC/GROa/CXCL1) in the *P. aeruginosa* infection model with or without treatment of ELD607. **FIG. 16B** shows further bar graphs of lung cytokine levels (left, IL-1 β ; middle, TNF- α ; right BAL IL-6) in the *P. aeruginosa* infection model with or without treatment of ELD607. **FIG. 16C** shows images of lung histology from mice according to the indicated conditions: naïve (left), *P. aeruginosa*-infected (middle) or *P. aeruginosa*-infected and treated with ELD607. Increased inflammation is seen in infection as indicated in the alveolar/gas exchange regions of the lungs which show alveolar damage and thickening that prevent gas exchange. ELD607 treatment normalized alveoli histology similar to naïve lung conditions. **FIG. 16D** shows bar graphs of total bronchoalveolar lavage (BAL) protein levels (left), total BAL neutrophil elastase levels (middle), total lactate dehydrogenase (LDH) levels (right) in the *P. aeruginosa* infection model with or without treatment of ELD607.

FIG. 16E shows a bar graph of bacterial counts (CFU/ml) in the spleen of the *P. aeruginosa* infected mice 24 hours post ELD607 treatment. **FIG. 16F** shows a schematic of the infection protocol and a bar graph of % weight loss in the *P. aeruginosa* infected mice 24 hours post ELD607 treatment.

FIG. 16G shows histology and graphs of BAL analysis, suggesting that ELD607 reduces Orai1 in murine BAL macrophages. Mice were exposed to vehicle or *P. aeruginosa* $\pm$ ELD607 and BAL was obtained and stained with an anti-Orai1 antibody, fluorescent 2° and DAPI. **FIG. 16G panel A** shows representative images of macrophages from the different groups as indicated. **FIG. 16G panels B and C** show plots showing fluorescence intensities of Orai1 in macrophages and neutrophils, respectively. *** $p<0.001$ different as indicated. ND = no detectable neutrophils in these groups.

FIG. 17A shows a schematic of the infection protocol and a bar graph of % survival of the *P. aeruginosa* infected mice 48 hours post ELD607 treatment. **FIG. 17B** shows a bar graph of BAL neutrophils of the same mice.

FIG. 18 shows a graph of *P. aeruginosa* growth in the presence of ELD100 (light circles), ELD607 (triangles), untreated (dark circles; negative control), or penicillin and streptomycin (diamonds; positive control), wherein the antibiotic treatment fully inhibited growth. ELD607 did not demonstrate antimicrobial activity.

FIG. 19 shows graphs of BAL, lung and spleen analyses from three different bacterial infections, suggesting that ELD607 broadly reduces neutrophilia and increases bacterial clearance from the lung. Mice were dosed intranasally with *P. aeruginosa* (PA), *S. aureus* (Sa), or *H. influenzae* followed by vehicle (saline) or 0.5 mg/kg of ELD100 or ELD607 per mouse. **FIG. 19 panels A, F, and K** show total cell counts; **FIG. 19 panels B, G, and L** show BAL macrophages; **FIG. 19 panels C, H, and M** show BAL neutrophils; **FIG. 19 panels D, I, and N** show whole lung CFUs; and **FIG. 19 panels E, J, and O** show spleen CFUs. Top row, *P. aeruginosa*, middle row, *S. aureus*, bottom row, *H. influenzae*. * $p < 0.05$ different from naïve mice; # $p < 0.05$ different from infected mice.

FIG. 20 shows bar graphs of results of house dust mite (HDM) allergy model experiments, indicating BAL total cell count (**FIG. 20 top left panel**), BAL eosinophils (**FIG. 20 bottom right panel**) BAL monocytes (**FIG. 20 bottom left panel**), and BAL neutrophils (**FIG. 20 top right panel**) in HDM-exposed mice treated with or without ELD100 and ELD607.

FIG. 21 shows graphs of BAL macrophages (**FIG. 21 panel A**) and neutrophils (**FIG. 21 panel B**) in SNCC1B mice, showing that ELD607 reduces pulmonary inflammation in SNCC1B mice. SCNN1B and littermate control mice were exposed to ELD607 or vehicle intranasally on a daily basis for 11 days, starting on day 1 post-partum. ELD607 had no effect on WT mice but significantly decreased BAL neutrophil levels in SCNN1B mice. * $p < 0.05$, *** $p < 0.001$ different as indicated.

FIG. 22 shows a bar graph of % infected cells by DsRed-positivity, showing that ELD607 reduces SARS-CoV-2 S1 pseudovirus infection in ACE2-expressing HEK293T cells. All $n = 3$ per group. * $p < 0.05$ different to S1 pseudovirus infection alone.

FIG. 23 shows a bar graph of % infected cells by DsRed-positivity. Experiments were performed in the same manner as **FIG. 22**, with additional peptides noted below each bar. All $n = 3$ per group. * $p < 0.05$ different to S1 pseudovirus infection alone.

FIG. 24 shows dot graphs indicating that ELD607 normalizes leukocyte levels and increases viral clearance from the lung. Mice were dosed intranasally with 10^6 PFU MHV-A59 followed by vehicle (saline) or 1.0 mg/kg of ELD607 per mouse 4 d p.i. BAL and lungs were collected 6 d p.i. **FIG. 24 panel A** shows BAL monocytes; **FIG. 24 panel B** shows

BAL macrophages; **FIG. 24 panel C** shows whole lung PFUs. * $p < 0.05$ different from naïve mice; # $p < 0.05$ different from infected mice.

FIG. 25 shows data plots and bar graphs of ELD607 in HEK293T cell studies. **FIG. 25, panel A** shows % peptide remaining over time; **FIG. 25, panel B** shows relative inhibition; **FIG. 25, panel C** shows store-operated calcium entry (SOCE) by relative fluorescence units (RFU); **FIG. 25, panel D** shows intensity weight harmonic mean particle size (Z-average) in the presence of OP1; **FIG. 25, panel E** shows Z-average in the presence of OP2.

FIG. 26 shows images of total internal reflection fluorescence (TIFR) microscopy and related data plots regarding Orai1 in the plasma membrane. **FIG. 26 panel A** shows cell culture images with vehicle, ELD607, and scrambled at 0h and 3h treatment. **FIG. 26 panel B** shows Orai1-YFP measured by relative fluorescence units, as related to **FIG. 26 panel A**. **FIG. 26 panel C** shows cell culture images of baseline, thapsigargin treatment, and thapsigargin + ELD607 treatment. **FIG. 26 panel D** shows NFAT1 activation levels, as related to **FIG. 26 panel C**.

FIG. 27 shows data plots and fluorescence images regarding ARDS BAL samples. **FIG. 27 panel A** shows a bar graph of % stable ELD607 remaining after 8 hours at 37 °C. **FIG. 27 panel B** shows % vehicle response. **FIG. 27 panel C** shows store-operated calcium entry (SOCE) score per relative fluorescence units. **FIG. 27 panel D** shows fluorescence images of vehicle, ELD607, and scrambled. **FIG. 27 panel E** shows Orai1 levels per relative fluorescence units in relation to **FIG. 27 panel D**.

FIG. 28 shows data plots and fluorescence images regarding experiments in HEK293 T cells exposed to 100 μ M ELD607 for 24 hours. **FIG. 28 panel A** shows a bar graph of cell viability (RFU). **FIG. 28 panel B** shows fluorescence images of vehicle and ELD607. **FIG. 28 panels C and D** show cell viability per arbitrary units (left) and transepithelial resistance (right) of vehicle vs. ELD607 before and after treatment.

FIG. 29 shows images of histology and related data plots regarding experiments performed in mice treated intratracheally at 10 mg/kg daily for 7 days. **FIG. 29 panel A** shows images of the lung. **FIG. 29 panel B** shows the lung histology score, in relation to **FIG. 29 panel A**. **FIG. 29 panel C** shows % weight change of the mice over time in days.

FIG. 30 shows images of histology and related data plots regarding experiments performed in mice treated intratracheally at 0.5 mM for 1 hour. **FIG. 30 panel A** shows images of light-sheet fluorescence microscopy in the airway lumen and interstitial. **FIG. 30**

panels B and C show % ELD607 recovered in BAL (FIG. 30 panel B) and blood (FIG. 30 panel C).

FIG. 31 shows bar graphs regarding experiments performed in Orai1 heterozygous mice that have about 70% reduced calcium influx, infected with 10^7 CFU/mouse *P.*

aeruginosa POA1 and treated intranasally 1 hour later with 0.5 mg/kg ELD607. FIG. 31 panel A shows Bal total cells/ml; FIG. 31 panel B shows BAL macrophages; FIG. 31 panel C shows BAL neutrophils; FIG. 31 panel D shows lung *P. aeruginosa* CFU count/ml; FIG. 31 panel E shows BAL macrophages; FIG. 31 panel F shows BAL neutrophils; FIG. 31 panel G shows % weight change; and FIG. 31 panel H shows lung *P. aeruginosa* CFU count/ml.

FIG. 32 shows a data plot and related survival plot regarding ELD607 reduction of BAL neutrophilia when added 24 h after infection and when chronically added to bENaC mice. FIG. 32 panel A: Mice were infected with *P. aeruginosa* for 24 h; then, 0.5 mg/kg ELD607 was added intratracheally. BAL was collected 24 h later to determine cell counts. SCNN1B mice were given 0.5 mg/kg ELD607 daily for 11 days before sacrifice and BAL collection. FIG. 32 panel B: Survival of bENaC mice.

FIG. 33 shows data plots regarding ELD607 reduction of allergic inflammation in the house dust mite (HDM) model. FIG. 33 panel A shows % eosinophils; FIG. 33 panel B shows % neutrophils; and FIG. 33 panel C shows % macrophages. Mice were exposed to HDM allergen for 14 days, with 3 μ g ELD100 or 2.5 μ g ELD607 instilled intranasally for 2 days before sacrifice.

FIG. 34 shows images of histology and related data plots regarding ELD607 reduction of inflammation in an ARDS model. FIG. 34 panel A shows lung histology after treatment with LPS or 6h of ELD607. FIG. 34 panels B-D show lung injury score (FIG. 34 panel B), % weight loss (FIG. 34 panel C), and % BAL neutrophils (FIG. 34 panel C) in relation to FIG. 34 panel A. Lung injury scoring system taken from Aeffner et al. Mouse Models of Acute Respiratory Distress Syndrome: A Review of Analytical Approaches, Pathologic Features, and Common Measurements. 2015 Tox Pathol.

FIG. 35 shows data plots regarding ELD607 reduction of inflammation in a sepsis model. FIG. 35 panel A shows % weight loss, FIG. 35 panel B shows % neutrophils, FIG. 35 panel C shows % macrophages, and FIG. 35 panel D shows % peptide remaining in experiments wherein mice were intraperitoneally injected with *E. coli* LPS, followed by 0.5 mg/kg ELD607 injected intravenously 1 h later.

DETAILED DESCRIPTION

The present invention now will be described hereinafter with reference to the accompanying drawings and examples, in which embodiments of the invention are shown. This description is not intended to be a detailed catalog of all the different ways in which the invention may be implemented, or all the features that may be added to the instant invention. For example, features illustrated with respect to one embodiment may be incorporated into other embodiments, and features illustrated with respect to a particular embodiment may be deleted from that embodiment. Thus, the invention contemplates that in some embodiments of the invention, any feature or combination of features set forth herein can be excluded or omitted. In addition, numerous variations and additions to the various embodiments suggested herein will be apparent to those skilled in the art in light of the instant disclosure, which do not depart from the instant invention. Hence, the following descriptions are intended to illustrate some particular embodiments of the invention, and not to exhaustively specify all permutations, combinations, and variations thereof.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. The terminology used in the description of the invention herein is for the purpose of describing particular embodiments only and is not intended to be limiting of the invention.

All publications, patent applications, patents and other references cited herein are incorporated by reference in their entireties for the teachings relevant to the sentence and/or paragraph in which the reference is presented.

Unless the context indicates otherwise, it is specifically intended that the various features of the invention described herein can be used in any combination. Moreover, the present invention also contemplates that in some embodiments of the invention, any feature or combination of features set forth herein can be excluded or omitted. To illustrate, if the specification states that a composition comprises components A, B and C, it is specifically intended that any of A, B or C, or a combination thereof, can be omitted and disclaimed singularly or in any combination.

As used in the description of the invention and the appended claims, the singular forms "a," "an" and "the" are intended to include the plural forms as well, unless the context clearly indicates otherwise.

Also as used herein, "and/or" refers to and encompasses any and all possible combinations of one or more of the associated listed items, as well as the lack of combinations when interpreted in the alternative ("or").

The term "about," as used herein when referring to a measurable value such as an amount or concentration and the like, is meant to encompass variations of $\pm 10\%$, $\pm 5\%$, $\pm 1\%$, $\pm 0.5\%$, or even $\pm 0.1\%$ of the specified value as well as the specified value. For example, "about X" where X is the measurable value, is meant to include X as well as variations of $\pm 10\%$, $\pm 5\%$, $\pm 1\%$, $\pm 0.5\%$, or even $\pm 0.1\%$ of X. A range provided herein for a measurable value may include any other range and/or individual value therein.

As used herein, phrases such as "between X and Y" and "between about X and Y" should be interpreted to include X and Y. As used herein, phrases such as "between about X and Y" mean "between about X and about Y" and phrases such as "from about X to Y" mean "from about X to about Y."

The term "comprise," "comprises" and "comprising" as used herein, specify the presence of the stated features, integers, steps, operations, elements, and/or components, but do not preclude the presence or addition of one or more other features, integers, steps, operations, elements, components, and/or groups thereof.

As used herein, the transitional phrase "consisting essentially of" means that the scope of a claim is to be interpreted to encompass the specified materials or steps recited in the claim and those that do not materially affect the basic and novel characteristic(s) of the claimed invention. Thus, the term "consisting essentially of" when used in a claim of this invention is not intended to be interpreted to be equivalent to "comprising."

The term "consists essentially of" (and grammatical variants), as applied to a peptide sequence of this invention, means a peptide that consists of both the recited sequence (*e.g.*, SEQ ID NO) and a total of ten or less (*e.g.*, 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10) additional amino acids on the N-terminal and/or C-terminal ends of the recited sequence such that the function of the peptide is not materially altered. The total of ten or less additional amino acids includes the total number of additional amino acids on both ends added together. The term "materially altered," as applied to peptides of the invention, refers to an increase or decrease in binding activity (*e.g.*, to a calcium channel) of at least about 50% or more as compared to the activity of a peptide consisting of the recited sequence.

Amino acids are represented herein in the manner recommended by the IUPAC-IUB Biochemical Nomenclature Commission, or (for amino acids) by either the one-letter code, or the three letter code, both in accordance with 37 C.F.R. §1.822 and established usage, *e.g.*, as shown in Table 6.

The term "modulate," "modulates," or "modulation" refers to enhancement (*e.g.*, an increase) or inhibition (*e.g.*, a decrease) in the specified level or activity.

The term "enhance" or "increase" refers to an increase in the specified parameter of at least about 1.25-fold, 1.5-fold, 2-fold, 3-fold, 4-fold, 5-fold, 6-fold, 8-fold, 10-fold, twelve-fold, or even fifteen-fold.

5 The term "inhibit" or "reduce" or grammatical variations thereof as used herein refers to a decrease or diminishment in the specified level or activity of at least about 15%, 25%, 35%, 40%, 50%, 60%, 75%, 80%, 90%, 95% or more. In particular embodiments, the inhibition or reduction results in little or essentially no detectable activity (at most, an insignificant amount, *e.g.*, less than about 10% or even 5%).

10 The term "contact" or grammatical variations thereof as used with respect to a polypeptide and a calcium channel, refers to bringing the polypeptide and the calcium channel in sufficiently close proximity to each other for one to exert a biological effect on the other. In some embodiments, the term contact means binding of the polypeptide to the calcium channel.

15 By the terms "treat," "treating," or "treatment of," it is intended that the severity of the subject's condition is reduced or at least partially improved or modified and that some alleviation, mitigation or decrease in at least one clinical symptom is achieved.

20 The terms "prevent," "preventing," and "prevention" (and grammatical variations thereof) refer to prevention and/or delay of the onset of a disease, disorder and/or a clinical symptom(s) in a subject and/or a reduction in the severity of the onset of the disease, disorder and/or clinical symptom(s) relative to what would occur in the absence of the methods of the invention. The prevention can be complete, *e.g.*, the total absence of the disease, disorder and/or clinical symptom(s). The prevention can also be partial, such that the occurrence of the disease, disorder and/or clinical symptom(s) in the subject and/or the severity of onset is less than what would occur in the absence of the present invention.

25 A "therapeutically effective" or "effective" amount as used herein is an amount that provides some improvement or benefit to the subject. Alternatively stated, a "therapeutically effective" amount or an "effective amount" is an amount that will provide some alleviation, mitigation, or decrease in at least one clinical symptom in the subject. Those skilled in the art will appreciate that the effects need not be complete or curative, as long as some benefit is provided to the subject.

30 A "prophylactically effective" amount as used herein is an amount that is sufficient to prevent and/or delay the onset of a disease, disorder and/or clinical symptoms in a subject and/or to reduce and/or delay the severity of the onset of a disease, disorder and/or clinical symptoms in a subject relative to what would occur in the absence of the methods of the

invention. Those skilled in the art will appreciate that the level of prevention need not be complete, as long as some benefit is provided to the subject.

The term "fragment," as applied to a peptide, will be understood to mean an amino acid sequence of reduced length relative to a reference peptide or amino acid sequence and comprising, consisting essentially of, and/or consisting of an amino acid sequence of contiguous amino acids identical to the reference peptide or amino acid sequence. Such a peptide fragment according to the invention may be, where appropriate, included in a larger polypeptide of which it is a constituent. In some embodiments, such fragments can comprise, consist essentially of, and/or consist of peptides having a length of at least about 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15 or more consecutive amino acids of a peptide or amino acid sequence according to the invention. In other embodiments, such fragments can comprise, consist essentially of, and/or consist of peptides having a length of less than about 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, or less consecutive amino acids of a peptide or amino acid sequence according to the invention.

As used herein, the terms "protein" and "polypeptide" are used interchangeably and encompass both peptides and proteins, unless indicated otherwise.

The "N-terminus" of a polypeptide is any portion of the polypeptide that starts from the N-terminal amino acid residue and continues to a maximum of the midpoint of the polypeptide.

The "C-terminus" of a polypeptide is any portion of the polypeptide that starts from the C-terminal amino acid residue and continues to a maximum of the midpoint of the polypeptide.

As used herein, a "functional" peptide or "functional fragment" is one that substantially retains at least one biological activity normally associated with that peptide (*e.g.*, binding to or inhibiting a calcium channel). In particular embodiments, the "functional" peptide or "functional fragment" substantially retains all of the activities possessed by the unmodified peptide. By "substantially retains" biological activity, it is meant that the peptide retains at least about 50%, 60%, 75%, 85%, 90%, 95%, 97%, 98%, 99%, or more, of the biological activity of the native polypeptide (and can even have a higher level of activity than the native peptide). A "non-functional" peptide is one that exhibits little or essentially no detectable biological activity normally associated with the peptide (*e.g.*, at most, only an insignificant amount, *e.g.*, less than about 10% or even 5%). Biological activities such as protein binding and calcium channel inhibitory activity can be measured using assays that are well known in the art and as described herein.

A "fusion protein" is a polypeptide produced when two heterologous nucleotide sequences or fragments thereof coding for two (or more) different polypeptides not found fused together in nature are fused together in the correct translational reading frame. Illustrative fusion polypeptides include fusions of a peptide of the invention (or a fragment thereof) to all or a portion of glutathione-S-transferase, maltose-binding protein, or a reporter protein (*e.g.*, Green Fluorescent Protein, β -glucuronidase, β -galactosidase, luciferase, *etc.*), hemagglutinin, c-myc, FLAG epitope, *etc.*

As used herein, "nucleic acid molecule," "nucleotide sequence," and "polynucleotide" are used interchangeably and encompass both RNA and DNA, including cDNA, genomic DNA, mRNA, synthetic (*e.g.*, chemically synthesized) DNA or RNA and chimeras of RNA and DNA. The term nucleic acid refers to a chain of nucleotides without regard to length of the chain. The nucleic acid can be double-stranded or single-stranded. Where single-stranded, the nucleic acid can be a sense strand or an antisense strand. The nucleic acid can be synthesized using oligonucleotide analogs or derivatives (*e.g.*, inosine or phosphorothioate nucleotides). Such oligonucleotides can be used, for example, to prepare nucleic acids that have altered base-pairing abilities or increased resistance to nucleases. The present invention further provides a nucleic acid that is the complement (which can be either a full complement or a partial complement) of a nucleic acid or nucleotide sequence of this invention.

An "isolated polynucleotide" is a nucleotide sequence (*e.g.*, DNA or RNA) that is not immediately contiguous with nucleotide sequences with which it is immediately contiguous (one on the 5' end and one on the 3' end) in the naturally occurring genome of the organism from which it is derived. Thus, in one embodiment, an isolated nucleic acid includes some or all of the 5' non-coding (*e.g.*, promoter) sequences that are immediately contiguous to a coding sequence. The term therefore includes, for example, a recombinant DNA that is incorporated into a vector, into an autonomously replicating plasmid or virus, or into the genomic DNA of a prokaryote or eukaryote, or which exists as a separate molecule (*e.g.*, a cDNA or a genomic DNA fragment produced by PCR or restriction endonuclease treatment), independent of other sequences. It also includes a recombinant DNA that is part of a hybrid nucleic acid encoding an additional polypeptide or peptide sequence. An isolated polynucleotide that includes a gene is not a fragment of a chromosome that includes such gene, but rather includes the coding region and regulatory regions associated with the gene, but no additional genes naturally found on the chromosome.

The term "isolated" can refer to a nucleic acid, nucleotide sequence or polypeptide that is substantially free of cellular material, viral material, and/or culture medium (when produced by recombinant DNA techniques), or chemical precursors or other chemicals (when chemically synthesized). Moreover, an "isolated fragment" is a fragment of a nucleic acid, nucleotide sequence or polypeptide that is not naturally occurring as a fragment and would not be found in the natural state. "Isolated" does not mean that the preparation is technically pure (homogeneous), but it is sufficiently pure to provide the polypeptide or nucleic acid in a form in which it can be used for the intended purpose.

An isolated cell refers to a cell that is separated from other components with which it is normally associated in its natural state. For example, an isolated cell can be a cell in culture medium and/or a cell in a pharmaceutically acceptable carrier of this invention. Thus, an isolated cell can be delivered to and/or introduced into a subject. In some embodiments, an isolated cell can be a cell that is removed from a subject and manipulated as described herein *ex vivo* and then returned to the subject.

A "vector" is any nucleic acid molecule for the cloning of and/or transfer of a nucleic acid into a cell. A vector may be a replicon to which another nucleotide sequence may be attached to allow for replication of the attached nucleotide sequence. A "replicon" can be any genetic element (*e.g.*, plasmid, phage, cosmid, chromosome, viral genome) that functions as an autonomous unit of nucleic acid replication *in vivo*, *i.e.*, capable of replication under its own control. The term "vector" includes both viral and nonviral (*e.g.*, plasmid) nucleic acid molecules for introducing a nucleic acid into a cell *in vitro*, *ex vivo*, and/or *in vivo*. A large number of vectors known in the art may be used to manipulate nucleic acids, incorporate response elements and promoters into genes, *etc.* For example, the insertion of the nucleic acid fragments corresponding to response elements and promoters into a suitable vector can be accomplished by ligating the appropriate nucleic acid fragments into a chosen vector that has complementary cohesive termini. Alternatively, the ends of the nucleic acid molecules may be enzymatically modified or any site may be produced by ligating nucleotide sequences (linkers) to the nucleic acid termini. Such vectors may be engineered to contain sequences encoding selectable markers that provide for the selection of cells that contain the vector and/or have incorporated the nucleic acid of the vector into the cellular genome. Such markers allow identification and/or selection of host cells that incorporate and express the proteins encoded by the marker. A "recombinant" vector refers to a viral or non-viral vector that comprises one or more heterologous nucleotide sequences (*i.e.*, transgenes), *e.g.*, two, three, four, five or more heterologous nucleotide sequences.

Viral vectors have been used in a wide variety of gene delivery applications in cells, as well as living animal subjects. Viral vectors that can be used include, but are not limited to, retrovirus, lentivirus, adeno-associated virus, poxvirus, alphavirus, baculovirus, vaccinia virus, herpes virus, Epstein-Barr virus, and adenovirus vectors. Non-viral vectors include plasmids, liposomes, electrically charged lipids (cytofectins), nucleic acid-protein complexes, and biopolymers. In addition to a nucleic acid of interest, a vector may also comprise one or more regulatory regions, and/or selectable markers useful in selecting, measuring, and monitoring nucleic acid transfer results (delivery to specific tissues, duration of expression, etc.).

Vectors may be introduced into the desired cells by methods known in the art, *e.g.*, transfection, electroporation, microinjection, transduction, cell fusion, DEAE dextran, calcium phosphate precipitation, lipofection (lysosome fusion), use of a gene gun, or a nucleic acid vector transporter (*see, e.g.*, Wu *et al.*, *J. Biol. Chem.* 267:963 (1992); Wu *et al.*, *J. Biol. Chem.* 263:14621 (1988); and Hartmut *et al.*, Canadian Patent Application No. 2,012,311, filed Mar. 15, 1990). In various embodiments, other molecules can be used for facilitating delivery of a nucleic acid *in vivo*, such as a cationic oligopeptide (*e.g.*, WO95/21931), peptides derived from nucleic acid binding proteins (*e.g.*, WO96/25508), and/or a cationic polymer (*e.g.*, WO95/21931). It is also possible to introduce a vector *in vivo* as naked nucleic acid (see U.S. Patent Nos. 5,693,622, 5,589,466 and 5,580,859). Receptor-mediated nucleic acid delivery approaches can also be used (Curiel *et al.*, *Hum. Gene Ther.* 3:147 (1992); Wu *et al.*, *J. Biol. Chem.* 262:4429 (1987)).

The term "transfection" or "transduction" means the uptake of exogenous or heterologous nucleic acid (RNA and/or DNA) by a cell. A cell has been "transfected" or "transduced" with an exogenous or heterologous nucleic acid when such nucleic acid has been introduced or delivered inside the cell. A cell has been "transformed" by exogenous or heterologous nucleic acid when the transfected or transduced nucleic acid imparts a phenotypic change in the cell and/or a change in an activity or function of the cell. The transforming nucleic acid can be integrated (covalently linked) into chromosomal DNA making up the genome of the cell or it can be present as a stable plasmid.

By the term "express" or "expression" of a polynucleotide coding sequence, it is meant that the sequence is transcribed, and optionally, translated. Typically, according to the present invention, expression of a coding sequence of the invention will result in production of the polypeptide of the invention. The entire expressed polypeptide can also function in intact cells without purification.

Compositions

The present invention relates to the design of peptidomimetics that bind to and inhibit the Orai1 calcium channel. Orai1 is a Ca^{2+} channel and master regulator of inflammation upstream of cytokine responses (**FIG. 1**). Therefore, selective inhibition of Orai1 can be immunomodulatory.

Global Orai1 expression presents a problem of potential off-target effects when drug-targeted. Orai1 regulates Ca^{2+} homeostasis in immune cells, where activation of the endoplasmic reticulum (ER) calcium sensor stromal interaction molecule 1 (STIM1) via ER calcium depletion downstream of T Cell Receptor (TCR) stimulation leads to Orai1-dependent calcium transport into the cell, which in turn activates calcineurin (a calcium and calmodulin dependent serine/threonine protein phosphatase)-dependent immune responses such as interleukin (IL)-8, IL-2, and tumor necrosis factor-alpha (TNF- α) (**FIG. 2**).

While not wishing to be bound to theory, it is believed that the synthetic polypeptides of the present invention such as ELD607 may inhibit Orai1 and thereby attenuate Ca^{2+} -dependent inflammation. ELD607 may also modulate immune responses against viruses which stimulate immune responses which depend on calcium fluxes. For example, COVID-19, the infection caused by SARS-CoV-2, causes damage to the infected subject via cytokine storm, pulmonary infiltrates, neutrophilia, and/or lung injury such as acute respiratory distress syndrome (ARDS). Treatment with synthetic polypeptides of the present invention, therefore, may reduce the levels of lung cytokines, decrease neutrophilia, and/or reduce lung damage, resulting in an improved infectious outcome (**FIG. 3**).

In some embodiments, synthetic polypeptides of the present invention may comprise modifications such as, but not limited to, substitution(s) of residue(s) such as but not limited to alanine substitution(s) and/or arginine substitution(s), deletion of residues (e.g., deletion at the C-terminal end, at the N-terminal end, and/or at intermediate positions, e.g., positions 1-4; deletion of a residue in a specific position, e.g., position 16; and/or deletion of specific amino acid residue(s), e.g., deletion of valine(s)); and/or insertion(s) of one or more residue(s) including non-natural amino acid residue(s) such as D-ala residue(s) and/or modifications of terminal ends (e.g., C-terminal amidation and/or N-terminal amidation, C-terminal acetylation and/or N-terminal acetylation, etc.). These modifications may be single modifications, or any combination of modifications, e.g., at least one, at least two, at least three, at least four, or more modifications in any combination.

Amino acid substitutions other than those disclosed herein may be achieved by changing the codons of the DNA sequence (or RNA sequence), according to **Table 6**.

Alternatively, the amino acid can be a modified amino acid residue (nonlimiting examples are shown in **Table 7**) or can be an amino acid that is modified by post-translation modification (e.g., acetylation, amidation, formylation, hydroxylation, methylation, phosphorylation or sulfatation). Further, the non-naturally occurring amino acid can be an "unnatural" amino acid as described by Wang *et al.*, (2006) *Annu. Rev. Biophys. Biomol. Struct.* 35:225-49.

One aspect of the present invention provides a synthetic polypeptide comprising one or more (e.g., at least one) of the following amino acid modifications, wherein the numbering is based on the reference amino acid sequence of an $\alpha 6$ helix peptide of a Short Palate Lung and Nasal Epithelial Clone 1 (SPLUNC1) protein: 14R substitution; deletion of residues in positions 1-4; deletion of residue in position 16; deletion of valine residue(s); and/or insertion of D-ala residue(s), wherein the synthetic polypeptide binds to a Orai1 plasma membrane Ca^{2+} channel.

Another aspect of the present invention provides a synthetic polypeptide comprising one or more (e.g., at least one) of the following amino acid modifications, wherein the numbering corresponds to the amino acid sequence of **SEQ ID NO:1**: 14R substitution; deletion of residues in positions 1-4; deletion of residue in position 16; deletion of valine residue(s); and/or insertion of D-ala residue(s), wherein the synthetic polypeptide binds to a Orai1 plasma membrane Ca^{2+} channel.

SEQ ID NO:1. ELD100 peptide.

DITLVHDIVNMLIHGL

The SPLUNC1 protein may be a SPLUNC1 protein of any organism, for example, a mammal. The term "mammal" as used herein includes, but is not limited to, humans, primates, non-human primates (e.g., monkeys and baboons), cattle, sheep, goats, pigs, horses, cats, dogs, rabbits, rodents (e.g., rats, mice, hamsters, and the like), *etc.* In some embodiments, the SPLUNC1 protein may be of a mouse, i.e., a murine SPLUNC protein. In some embodiments, the SPLUNC1 protein is human SPLUNC1.

The amino acid sequence of human SPLUNC1 is disclosed below.

SEQ ID NO:2. Human SPLUNC1.

	10	20	30	40	50	60
30	MFQTGGILIVF	YGLLAQTMAQ	FGGLPVPLDQ	TLPLNVNPAL	PLSPTGLAGS	LTNALSNGLL
	70	80	90	100	110	120
	SGGLLGILEN	LPLLDILKPG	GGTSGGILLGG	LLGKVTSVIP	GLNNIIDIKV	TDPQLLELGL
	130	140	150	160	170	180
	VQSPDGHRLY	VTIPLGIKLQ	VNTPLVGASL	LRLAVKLDIT	AEILAVRDKQ	ERIHVLGDC
35	190	200	210	220	230	240
	THSPGSLQIS	LLDGLGPLPI	QGLLDLSLTGI	LNKVLPELVQ	GNVCPLVNEV	LRGLDITLVH
	250					
	DIVNMLIHGL	QFVIKV				

In certain embodiments, the synthetic polypeptide mimics the Orail calcium channel binding domain of a SPLUNC1 protein. The calcium channel binding domain is the minimal fragment required to have substantially the same binding activity to the calcium channel as the full length SPLUNC1 protein. The term "substantially the same binding activity" refers to an activity that is at least about 50% of the binding activity of the full length protein, *e.g.*, at least about 60%, 70%, 80%, or 90% of the binding activity. In some embodiments, the peptide has at least the same binding activity as the full length SPLUNC1 protein.

The length of the synthetic polypeptide is not critical as long as it substantially retains the biological activity of the polypeptide (*e.g.*, calcium channel binding activity). In some embodiments, the synthetic polypeptide comprises no more than about 30, 25, 20, 15, 12, 10, or 8 contiguous amino acids of a SPLUNC1 protein or an amino acid sequence that has at least 70% sequence identity thereto, *e.g.*, at least 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% sequence identity thereto. In certain embodiments, the fragment is about 5 to about 30 amino acids, about 5 to about 25 amino acids, about 5 to about 20 amino acids, about 10 to about 30 amino acids, about 10 to about 25 amino acids, about 10 to about 20 amino acids, about 15 to about 30 amino acids, about 15 to about 25 amino acids, or about 15 to about 20 amino acids. In some embodiments, synthetic polypeptide may be about 16 amino acid residues in length or less, *e.g.*, about 16, 15, 14, 13, 12, 11, 10, 9, or 8 amino acid residues in length or less.

In certain embodiments, the polypeptide of the invention may comprise at least one modified terminus, *e.g.*, to protect the polypeptide against degradation. In some embodiments, the synthetic polypeptide may be resistant to cleavage by proteases such as endogenous mammalian proteases found in lung tissue. In some embodiments, the synthetic polypeptide of the present invention may be resistant to cleavage by, but not limited to, elastase and/or trypsin.

In some embodiments, a synthetic polypeptide of the present invention may be a polypeptide which binds to a Orail plasma membrane Ca^{2+} channel and comprises the following amino acid modifications, wherein the numbering corresponds to the amino acid sequence of **SEQ ID NO:1**: deletion of DITL residues in positions 1-4, and L16 deletion.

In some embodiments, a synthetic polypeptide of the present invention may comprise, consist essentially of, or consist of the amino acid sequence **SEQ ID NO:3**.

SEQ ID NO:3. ELD302.

VHDIVNMLIHG

In some embodiments, a synthetic polypeptide of the present invention may be a polypeptide which binds to a Orail plasma membrane Ca^{2+} channel and comprises the

following amino acid modifications, wherein the corresponds to the amino acid sequence of **SEQ ID NO:1**: deletion of DITL residues in positions 1-4; L16 deletion; V5 deletion; and V9 deletion.

In some embodiments, a synthetic polypeptide of the present invention may comprise, consist essentially of, or consist of the amino acid sequence **SEQ ID NO:4**.

SEQ ID NO:4. ELD601.

HDINMLIHG

In some embodiments, a synthetic polypeptide of the present invention may be a polypeptide which binds to a Orai1 plasma membrane Ca^{2+} channel and comprises the following amino acid modifications, wherein the numbering corresponds to the amino acid sequence of **SEQ ID NO:1**: deletion of DITL residues in positions 1-4; L16 deletion; C-terminal amidation; and N-terminal amidation.

In some embodiments, a synthetic polypeptide of the present invention may comprise, consist essentially of, or consist of the amino acid sequence **SEQ ID NO:5**.

SEQ ID NO:5. ELD621.

n-VHDIVNMLIHG-n

In some embodiments, a synthetic polypeptide of the present invention may be a polypeptide which binds to a Orai1 plasma membrane Ca^{2+} channel and comprises the following amino acid modifications, wherein the numbering corresponds to the amino acid sequence of **SEQ ID NO:1**: deletion of DITL residues in positions 1-4; L16 deletion; V5 deletion; G15 deletion; C-terminal D-ala insertion; and N-terminal D-ala insertion.

In some embodiments, a synthetic polypeptide of the present invention may comprise, consist essentially of, or consist of the amino acid sequence **SEQ ID NO:6**.

SEQ ID NO:6. ELD604.

a-HDIVNMLIH-a

In some embodiments, a synthetic polypeptide of the present invention may be a polypeptide which binds to a Orai1 plasma membrane Ca^{2+} channel and comprises the following amino acid modifications, wherein the corresponds to the amino acid sequence of **SEQ ID NO:1**: H14R substitution, C-terminal amidation, and N-terminal amidation.

In some embodiments, a synthetic polypeptide of the present invention may comprise, consist essentially of, or consist of the amino acid sequence **SEQ ID NO:7**.

SEQ ID NO:7. ELD142.

DITLVHDIVNMLIRGL-NH₂

In some embodiments, a synthetic polypeptide of the present invention may be a polypeptide which binds to a Orai1 plasma membrane Ca^{2+} channel and comprises the following amino acid modifications, wherein the numbering corresponds to the amino acid sequence of **SEQ ID NO:1**: H14R substitution; deletion of DITL residues in positions 1-4; and L16 deletion.

In some embodiments, a synthetic polypeptide of the present invention may comprise, consist essentially of, or consist of the amino acid sequence **SEQ ID NO:8**.

SEQ ID NO:8. ELD607.

VHDIVNMLIRG

It is understood that these examples are not intended to be limited and that any of the above-described modifications can be combined with any other of the above-referenced modifications in a synthetic polypeptide which binds to a Orai1 plasma membrane Ca^{2+} channel.

Likewise, those skilled in the art will appreciate that the present invention also encompasses fusion polypeptides (and nucleic acid molecules encoding the same) comprising a synthetic polypeptide of the present invention. For example, it may be useful to express the synthetic polypeptide as a fusion protein that can be recognized by a commercially available antibody (*e.g.*, FLAG motifs) or as a fusion protein that can otherwise be more easily purified (*e.g.*, by addition of a poly-His tail). Additionally, fusion proteins that enhance the stability of the synthetic polypeptide may be produced, *e.g.*, fusion proteins comprising maltose binding protein (MBP) or glutathione-S-transferase. As another alternative, the fusion protein can comprise a reporter molecule. In other embodiments, the fusion protein can comprise a polypeptide that provides a function or activity that is the same as or different from the activity of the synthetic polypeptide of the present invention, *e.g.*, a targeting, binding, or enzymatic activity or function.

The present invention further provides an isolated nucleic acid molecule encoding the synthetic polypeptide of this invention. In some embodiments, a nucleic acid molecule of this invention may be a cDNA molecule. In some embodiments, a nucleic acid molecule of this invention may be an mRNA molecule. It will be appreciated by those skilled in the art that there can be variability in the nucleic acid molecules that encode the polypeptides (and fragments thereof) of the present invention due to the degeneracy of the genetic code. The degeneracy of the genetic code, which allows different nucleic acid sequences to code for the same polypeptide, is well known in the literature (*See, e.g.*, **Table 6**).

As is known in the art, a number of different programs can be used to identify whether a nucleic acid molecule or polypeptide has sequence identity or similarity to a known sequence. Sequence identity or similarity may be determined using standard techniques known in the art, including, but not limited to, the local sequence identity algorithm of Smith & Waterman, *Adv. Appl. Math.* 2:482 (1981), by the sequence identity alignment algorithm of Needleman & Wunsch, *J. Mol. Biol.* 48:443 (1970), by the search for similarity method of Pearson & Lipman, *Proc. Natl. Acad. Sci. USA* 85:2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Drive, Madison, WI), the Best Fit sequence program described by Devereux *et al.*, *Nucl. Acid Res.* 12:387 (1984), preferably using the default settings, or by inspection.

An example of a useful algorithm is PILEUP. PILEUP creates a multiple sequence alignment from a group of related sequences using progressive, pairwise alignments. It can also plot a tree showing the clustering relationships used to create the alignment. PILEUP uses a simplification of the progressive alignment method of Feng & Doolittle, *J. Mol. Evol.* 35:351 (1987); the method is similar to that described by Higgins & Sharp, *CABIOS* 5:151 (1989).

Another example of a useful algorithm is the BLAST algorithm, described in Altschul *et al.*, *J. Mol. Biol.* 215:403 (1990) and Karlin *et al.*, *Proc. Natl. Acad. Sci. USA* 90:5873 (1993). A particularly useful BLAST program is the WU-BLAST-2 program which was obtained from Altschul *et al.*, *Meth. Enzymol.*, 266:460 (1996); blast.wustl.edu/blast/README.html. WU-BLAST-2 uses several search parameters, which are preferably set to the default values. The parameters are dynamic values and are established by the program itself depending upon the composition of the particular sequence and composition of the particular database against which the sequence of interest is being searched; however, the values may be adjusted to increase sensitivity. An additional useful algorithm is gapped BLAST as reported by Altschul *et al.*, *Nucleic Acids Res.* 25:3389 (1997).

A percentage amino acid sequence identity value is determined by the number of matching identical residues divided by the total number of residues of the "longer" sequence in the aligned region. The "longer" sequence is the one having the most actual residues in the aligned region (gaps introduced by WU-Blast-2 to maximize the alignment score are ignored).

In a similar manner, percent nucleic acid sequence identity with respect to the coding sequence of the polypeptides disclosed herein is defined as the percentage of nucleotide residues in the candidate sequence that are identical with the nucleotides in the polynucleotide specifically disclosed herein.

5 The alignment may include the introduction of gaps in the sequences to be aligned. In addition, for sequences which contain either more or fewer amino acids than the polypeptides specifically disclosed herein, it is understood that in one embodiment, the percentage of sequence identity will be determined based on the number of identical amino acids in relation to the total number of amino acids. Thus, for example, sequence identity of sequences
10 shorter than a sequence specifically disclosed herein, will be determined using the number of amino acids in the shorter sequence, in one embodiment. In percent identity calculations relative weight is not assigned to various manifestations of sequence variation, such as insertions, deletions, substitutions, *etc.*

 In one embodiment, only identities are scored positively (+1) and all forms of
15 sequence variation including gaps are assigned a value of "0," which obviates the need for a weighted scale or parameters as described below for sequence similarity calculations. Percent sequence identity can be calculated, for example, by dividing the number of matching identical residues by the total number of residues of the "shorter" sequence in the aligned region and multiplying by 100. The "longer" sequence is the one having the most actual
20 residues in the aligned region.

 Those skilled in the art will appreciate that the isolated nucleic acid molecules encoding the polypeptides of the invention will typically be associated with appropriate expression control sequences, *e.g.*, transcription/translation control signals and polyadenylation signals.

25 It will further be appreciated that a variety of promoter/enhancer elements can be used depending on the level and tissue-specific expression desired. The promoter can be constitutive or inducible, depending on the pattern of expression desired. The promoter can be native or foreign and can be a natural or a synthetic sequence. By foreign, it is intended that the transcriptional initiation region is not found in the wild-type host into which the
30 transcriptional initiation region is introduced. The promoter is chosen so that it will function in the target cell(s) of interest.

 To illustrate, the polypeptide coding sequence can be operatively associated with a cytomegalovirus (CMV) major immediate-early promoter, an albumin promoter, an

Elongation Factor 1- α (EF1- α) promoter, a PyK promoter, a MFG promoter, or a Rous sarcoma virus promoter.

Inducible promoter/enhancer elements include hormone-inducible and metal-inducible elements, and other promoters regulated by exogenously supplied compounds, including without limitation, the zinc-inducible metallothionein (MT) promoter; the dexamethasone (Dex)-inducible mouse mammary tumor virus (MMTV) promoter; the T7 polymerase promoter system (*see* WO 98/10088); the ecdysone insect promoter (No *et al.*, *Proc. Natl. Acad. Sci. USA* 93:3346 (1996)); the tetracycline-repressible system (Gossen *et al.*, *Proc. Natl. Acad. Sci. USA* 89:5547 (1992)); the tetracycline-inducible system (Gossen *et al.*, *Science* 268:1766 (1995); *see also* Harvey *et al.*, *Curr. Opin. Chem. Biol.* 2:512 (1998)); the RU486-inducible system (Wang *et al.*, *Nat. Biotech.* 15:239 (1997); Wang *et al.*, *Gene Ther.*, 4:432 (1997)); and the rapamycin-inducible system (Magari *et al.*, *J. Clin. Invest.* 100:2865 (1997)).

Moreover, specific initiation signals are generally required for efficient translation of inserted polypeptide coding sequences. These translational control sequences, which can include the ATG initiation codon and adjacent sequences, can be of a variety of origins, both natural and synthetic.

The present invention further provides cells comprising the isolated nucleic acid molecules and/or synthetic polypeptides of the invention, or a vector comprising the same. The cell may be a cultured cell or a cell *in vivo*, *e.g.*, for use in therapeutic methods, diagnostic methods, screening methods, methods for studying the biological action of the Orai1 calcium channel, in methods of producing the synthetic polypeptides, or in methods of maintaining or amplifying the nucleic acid molecules of the invention, *etc.* In another embodiment, the cell is an *ex vivo* cell that has been isolated from a subject. The *ex vivo* cell may be modified and then reintroduced into the subject for diagnostic or therapeutic purposes. In some embodiments, the cell may be an untransformed airway smooth muscle cell or a cell from an airway smooth muscle cell line.

Human subjects include neonates, infants, juveniles, and adults. Optionally, the subject is "in need of" the methods of the present invention, *e.g.*, because the subject has or is believed at risk for a disorder including those described herein or that would benefit from the delivery of a synthetic polypeptide including those described herein. As a further option, the subject can be a laboratory animal and/or an animal model of disease.

Also provided is a vector, plasmid or other nucleic acid construct comprising the isolated nucleic acid molecule of this invention.

A vector can be any suitable means for delivering a polynucleotide to a cell. A vector of this invention can be an expression vector that contains all of the genetic components required for expression of the nucleic acid in cells into which the vector has been introduced, as are well known in the art. For example, an expression vector may typically contain an "expression cassette," which includes, in the 5' to 3' direction, a promoter, a coding sequence encoding a polypeptide operatively associated with the promoter, and, optionally, a termination sequence including a stop signal for RNA polymerase and a polyadenylation signal for polyadenylase.

Non-limiting examples of promoters of this invention include CYC1, HIS3, GAL1, GAL4, GAL10, ADH1, PGK, PHO5, GAPDH, ADC1, TRP1, URA3, LEU2, ENO, TPI, and alkaline phosphatase promoters (useful for expression in *Saccharomyces*); AOX1 promoter (useful for expression in *Pichia*); β -lactamase, lac, ara, tet, trp, IPL, IPR, T7, tac, and trc promoters (useful for expression in *Escherichia coli*); light regulated-, seed specific-, pollen specific-, ovary specific-, pathogenesis or disease related-promoters, cauliflower mosaic virus 35S, CMV 35S minimal, cassava vein mosaic virus (CsVMV), chlorophyll a/b binding protein, ribulose 1,5-bisphosphate carboxylase, shoot-specific promoters, root specific promoters, chitinase, stress inducible promoters, rice tungro bacilliform virus, plant super-promoter, potato leucine aminopeptidase, nitrate reductase, mannopine synthase, nopaline synthase, ubiquitin, zein protein, and anthocyanin promoters (useful for expression in plant cells). Further examples of animal and mammalian promoters known in the art include, but are not limited to, the SV40 early (SV40e) promoter region, the promoter contained in the 3' long terminal repeat (LTR) of Rous sarcoma virus (RSV), the promoters of the E1A or major late promoter (MLP) genes of adenoviruses (Ad), the cytomegalovirus (CMV) early promoter, the herpes simplex virus (HSV) thymidine kinase (TK) promoter, baculovirus IE1 promoter, elongation factor 1 alpha (EF1) promoter, phosphoglycerate kinase (PGK) promoter, ubiquitin (Ubc) promoter, an albumin promoter, the regulatory sequences of the mouse metallothionein-L promoter and transcriptional control regions, the ubiquitous promoters (HPRT, vimentin, α -actin, tubulin and the like), the promoters of the intermediate filaments (desmin, neurofilaments, keratin, GFAP, and the like), the promoters of therapeutic genes (of the MDR, CFTR or factor VIII type, and the like), pathogenesis and/or disease-related promoters, and promoters that exhibit tissue specificity, such as the elastase I gene control region, which is active in pancreatic acinar cells; the insulin gene control region

active in pancreatic beta cells, the immunoglobulin gene control region active in lymphoid cells, the mouse mammary tumor virus control region active in testicular, breast, lymphoid and mast cells; the albumin gene promoter, the Apo AI and Apo AII control regions active in liver, the alpha-fetoprotein gene control region active in liver, the alpha 1-antitrypsin gene control region active in the liver, the beta-globin gene control region active in myeloid cells, the myelin basic protein gene control region active in oligodendrocyte cells in the brain, the myosin light chain-2 gene control region active in skeletal muscle, and the gonadotropic releasing hormone gene control region active in the hypothalamus, the pyruvate kinase promoter, the villin promoter, the promoter of the fatty acid binding intestinal protein, the promoter of smooth muscle cell α -actin, and the like. In addition, any of these expression sequences of this invention can be modified by addition of enhancer and/or regulatory sequences and the like.

Enhancers that may be used in embodiments of the invention include but are not limited to: an SV40 enhancer, a cytomegalovirus (CMV) enhancer, an elongation factor I (EF1) enhancer, yeast enhancers, viral gene enhancers, and the like.

Termination control regions, *i.e.*, terminator or polyadenylation sequences, may be derived from various genes native to the preferred hosts. In some embodiments of the invention, the termination control region may comprise or be derived from a synthetic sequence, a synthetic polyadenylation signal, an SV40 late polyadenylation signal, an SV40 polyadenylation signal, a bovine growth hormone (BGH) polyadenylation signal, viral terminator sequences, or the like.

It will be apparent to those skilled in the art that any suitable vector can be used to deliver the polypeptide and/or polynucleotide to a cell or subject. The vector can be delivered to cells *in vivo*. In other embodiments, the vector can be delivered to cells *ex vivo*, and then cells containing the vector are delivered to the subject. The choice of delivery vector can be made based on a number of factors known in the art, including age and species of the target host, *in vitro* versus *in vivo* delivery, level and persistence of expression desired, intended purpose (*e.g.*, for therapy or screening), the target cell or organ, route of delivery, size of the isolated polynucleotide, safety concerns, and the like.

The vector can be a commercial expression vector or it can be constructed in the laboratory according to standard molecular biology protocols. The vector can comprise viral nucleic acid including, but not limited to, poxvirus, vaccinia virus, adenovirus, retrovirus, alphavirus and/or adeno-associated virus nucleic acid. The nucleic acid molecule or vector of this invention can also be in a liposome or a delivery vehicle, which can be taken up by a cell

via receptor-mediated or other type of endocytosis. The nucleic acid molecule of this invention can be in a cell, which can be a cell expressing the nucleic acid whereby a synthetic polynucleotide of this invention is produced in the cell (e.g., a host cell). In addition, the vector of this invention can be in a cell, which can be a cell expressing the nucleic acid of the vector whereby a synthetic polynucleotide of this invention is produced in the cell. It is also contemplated that the nucleic acid molecules and/or vectors of this invention can be present in a host organism (e.g., a transgenic organism), which expresses the nucleic acids of this invention and produces the synthetic polynucleotide of this invention. Suitable vectors include plasmid vectors, viral vectors (e.g., retrovirus, alphavirus; vaccinia virus; adenovirus, adeno-associated virus and other parvoviruses, lentivirus, poxvirus, or herpes simplex virus), lipid vectors, poly-lysine vectors, synthetic polyamino polymer vectors, and the like. Any viral vector that is known in the art can be used in the present invention. Protocols for producing recombinant viral vectors and for using viral vectors for nucleic acid delivery can be found in Ausubel *et al.*, Current Protocols in Molecular Biology (Green Publishing Associates, Inc. and John Wiley & Sons, Inc., New York) and other standard laboratory manuals (e.g., Vectors for Gene Therapy. In: *Current Protocols in Human Genetics*. John Wiley and Sons, Inc.: 1997). In some embodiments, the vector is a plasmid, a viral vector, a bacterial vector, an expression cassette, a transformed cell, or a nanoparticle.

Non-viral transfer methods can also be employed. Many non-viral methods of nucleic acid transfer rely on normal mechanisms used by mammalian cells for the uptake and intracellular transport of macromolecules. In particular embodiments, non-viral nucleic acid delivery systems rely on endocytic pathways for the uptake of the nucleic acid molecule by the targeted cell. Exemplary nucleic acid delivery systems of this type include liposomal derived systems, poly-lysine conjugates, and artificial viral envelopes.

In some embodiments, plasmid vectors are used in the practice of the present invention. For example, naked plasmids can be introduced into muscle cells by injection into the tissue. Expression can extend over many months, although the number of positive cells is typically low (Wolff *et al.*, *Science* 247:247 (1989)). Cationic lipids have been demonstrated to aid in introduction of nucleic acids into some cells in culture (Felgner and Ringold, *Nature* 337:387 (1989)). Injection of cationic lipid plasmid DNA complexes into the circulation of mice has been shown to result in expression of the DNA in lung (Brigham *et al.*, *Am. J. Med. Sci.* 298:278 (1989)). One advantage of plasmid DNA is that it can be introduced into non-replicating cells.

In a representative embodiment, a nucleic acid molecule (*e.g.*, a plasmid) can be entrapped in a lipid particle bearing positive charges on its surface and, optionally, tagged with antibodies against cell surface antigens of the target tissue (Mizuno *et al.*, *No Shinkei Geka* 20:547 (1992); PCT publication WO 91/06309; Japanese patent application 1047381; and European patent publication EP-A-43075).

Liposomes that consist of amphiphilic cationic molecules are useful as non-viral vectors for nucleic acid delivery *in vitro* and *in vivo* (reviewed in Crystal, *Science* 270:404 (1995); Blaese *et al.*, *Cancer Gene Ther.* 2:291 (1995); Behr *et al.*, *Bioconjugate Chem.* 5:382 (1994); Remy *et al.*, *Bioconjugate Chem.* 5:647 (1994); and Gao *et al.*, *Gene Therapy* 2:710 (1995)). The positively charged liposomes are believed to complex with negatively charged nucleic acids via electrostatic interactions to form lipid:nucleic acid complexes. The lipid:nucleic acid complexes have several advantages as nucleic acid transfer vectors. Unlike viral vectors, the lipid:nucleic acid complexes can be used to transfer expression cassettes of essentially unlimited size. Since the complexes lack proteins, they can evoke fewer immunogenic and inflammatory responses. Moreover, they cannot replicate or recombine to form an infectious agent and have low integration frequency. A number of publications have demonstrated that amphiphilic cationic lipids can mediate nucleic acid delivery *in vivo* and *in vitro* (Felgner *et al.*, *Proc. Natl. Acad. Sci. USA* 84:7413 (1987); Loeffler *et al.*, *Meth. Enzymol.* 217:599 (1993); Felgner *et al.*, *J. Biol. Chem.* 269:2550 (1994)).

Viral vectors have been used in a wide variety of gene delivery applications in cells, as well as living animal subjects. Viral vectors that can be used include, but are not limited to, retrovirus, lentivirus, adeno-associated virus, poxvirus, alphavirus, baculovirus, vaccinia virus, herpes virus, Epstein-Barr virus, adenovirus, geminivirus, and caulimovirus vectors. Non-viral vectors include plasmids, liposomes, electrically charged lipids (cytofectins), nucleic acid-protein complexes, and biopolymers. In addition to a nucleic acid of interest, a vector may also comprise one or more regulatory regions, and/or selectable markers useful in selecting, measuring, and monitoring nucleic acid transfer results (delivery to specific tissues, duration of expression, *etc.*).

In addition to the regulatory control sequences discussed above, the recombinant expression vector can contain additional nucleotide sequences. For example, the recombinant expression vector can encode a selectable marker gene to identify host cells that have incorporated the vector.

Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and

"transfection" refer to a variety of art-recognized techniques for introducing foreign nucleic acids (*e.g.*, DNA and RNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, electroporation, microinjection, DNA-loaded liposomes, lipofectamine-DNA complexes, cell sonication, gene bombardment using high velocity microprojectiles, and viral-mediated transfection. Suitable methods for transforming or transfecting host cells can be found in Sambrook *et al.*, Molecular Cloning: A Laboratory Manual 2nd Ed. (Cold Spring Harbor, NY, 1989), and other laboratory manuals.

If stable integration is desired, often only a small fraction of cells (in particular, mammalian cells) integrate the foreign DNA into their genome. In order to identify and select integrants, a nucleic acid that encodes a selectable marker (*e.g.*, resistance to antibiotics) can be introduced into the host cells along with the nucleic acid of interest. Preferred selectable markers include those that confer resistance to drugs, such as G418, hygromycin and methotrexate. Nucleic acids encoding a selectable marker can be introduced into a host cell on the same vector as that comprising the nucleic acid of interest or can be introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by drug selection (*e.g.*, cells that have incorporated the selectable marker gene will survive, while the other cells die).

A further aspect of the invention relates to a composition comprising the synthetic polypeptide of the invention and a carrier. In some embodiments, the composition is a pharmaceutical composition comprising the synthetic polypeptide of the invention and a pharmaceutically acceptable carrier.

By "pharmaceutically acceptable" it is meant a material that is not toxic or otherwise undesirable, *i.e.*, the material may be administered to a subject without causing any undesirable biological effects. For injection, the carrier will typically be a liquid. For other methods of administration (*e.g.*, such as, but not limited to, administration to the mucous membranes of a subject (*e.g.*, via intranasal administration, buccal administration and/or inhalation)), the carrier may be either solid or liquid. For inhalation administration, the carrier will be respirable, and will preferably be in solid or liquid particulate form. The formulations may be conveniently prepared in unit dosage form and may be prepared by any of the methods well known in the art. In some embodiments, that pharmaceutically acceptable carrier can be a sterile solution or composition.

In some embodiments, the present invention provides a pharmaceutical composition comprising the synthetic polypeptide, nucleic acid molecule, vector, and/or composition of

the present invention, a pharmaceutically acceptable carrier, and optionally, other medicinal agents, therapeutic agents, pharmaceutical agents, stabilizing agents, buffers, carriers, adjuvants, diluents, etc., which can be included in the composition singly or in any combination and/or ratio.

5 In some embodiments, a pharmaceutical composition of the present invention may further comprise a therapeutic agent selected from, but not limited to, the group consisting of dexamethasone, remdesivir, elixacaftor, ivacaftor, lumacaftor, tezacaftor, ibuprofen, acebilustat, lenabasum, levofloxacin, piperacillin-taxobactam, vancomycin, azithromycin, ciprofloxacin, cephalixin, doxycycline hyclate, and any combination thereof.

10 Immunogenic compositions comprising a synthetic polypeptide, nucleic acid molecule, vector, and/or composition of the present invention may be formulated by any means known in the art. Such compositions are typically prepared as injectables, either as liquid solutions or suspensions. Solid forms suitable for solution in, or suspension in, liquid prior to injection may also be prepared. Lyophilized preparations are also suitable. The
15 active immunogenic ingredients are often mixed with excipients and/or carriers that are pharmaceutically acceptable and/or compatible with the active ingredient. Suitable excipients include but are not limited to sterile water, saline, dextrose, glycerol, ethanol, or the like and combinations thereof, as well as stabilizers, *e.g.*, HSA or other suitable proteins and reducing sugars. In addition, if desired, immunogenic compositions may contain minor
20 amounts of auxiliary substances such as wetting and/or emulsifying agents, pH buffering agents, and/or adjuvants that enhance the effectiveness of the vaccine or immunogenic composition.

Additionally provided here is a dosage delivery device comprising a pharmaceutical composition of the present invention. In some embodiments, the dosage delivery device is an
25 inhaler, *e.g.*, for delivery of the composition to the airways of a subject, *e.g.*, by oral and/or nasal inhalation.

Another aspect of the invention relates to a kit comprising the synthetic polypeptide, nucleic acid molecule, vector, cell, and/or composition of the invention and useful for carrying out the methods of the invention. The kit may further comprise additional reagents
30 for carrying out the methods (*e.g.*, buffers, containers, additional therapeutic agents) as well as instructions.

Methods

The methods of the invention relate to the ability of synthetic polypeptides to bind to an Orai1 calcium channel, *e.g.*, a human Orai1 calcium channel.

Thus, one aspect of the present invention provides a method of inhibiting calcium influx through a calcium channel, comprising contacting the calcium channel with a synthetic polypeptide or a pharmaceutical composition of the present invention, thereby inhibiting calcium influx through the calcium channel.

5 The inhibition of calcium channel activity can be measured by any method known in the art or disclosed herein, including, without limitation, measuring calcium flow or change in potential across a membrane, across a cell, or across a natural or artificial lining. The inhibition can be at least about 20%, *e.g.*, at least about 30%, 40%, 50%, 60%, 70%, 80%, 90%, or 100%.

Another aspect of the invention relates to a method of inhibiting airway smooth muscle
10 contraction, comprising contacting the airway with the polypeptide of the invention, thereby inhibiting airway smooth muscle contraction. Inhibition of smooth muscle contraction can be measured by any technique known in the art or disclosed herein. Inhibition of contraction is measured relative to the level of contraction in the absence of contact with the polypeptide of the invention. In some embodiments, contraction is inhibited by at least about 10%, *e.g.*, at least
15 about 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, or more.

Another aspect of the invention relates to a method of inhibiting airway hyperreactivity, comprising contacting the airway with the polypeptide of the invention, thereby inhibiting airway hyperreactivity. Airway reactivity can be measured by any technique known in the art or disclosed herein. Inhibition of hyperreactivity is measured relative to the level of reactivity in
20 the absence of contact with the polypeptide of the invention. "Hyperreactivity," as used herein, refers to increased reactivity of the airway to calcium relative to the level of reactivity of normal airway. In some embodiments, hyperreactivity is inhibited by at least about 10%, *e.g.*, at least about 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, or more.

Another aspect of the present invention provides a method of reducing expression of
25 an Orai1 calcium channel, comprising contacting the calcium channel Orai1 with a synthetic polypeptide or a pharmaceutical composition of the present invention. While not wishing to be bound by theory, the synthetic polypeptides of the invention may induce allosteric in Orai1 that causes Orai1 to be internalized and degraded, *e.g.*, in the cell.

The methods of the present invention can be carried out, *e.g.*, on an isolated Orai1
30 calcium channel, an Orai1 calcium channel in an artificial membrane, or an Orai1 calcium channel in a cell. In one embodiment, the calcium channel is present in an isolated cell, *e.g.*, a cultured primary cell or cell line. In certain embodiments, the cell is an airway smooth muscle cell, *e.g.*, part of an airway smooth muscle cell culture. In another embodiment, the isolated cell is part of an epithelial cell culture, *e.g.*, a natural or artificial epithelial lining, *e.g.*, a cell culture

in a device (such as an Ussing chamber) in which characteristics such as ion flow and/or potential can be measured across a lining. In another embodiment, the isolated cell is an immune system cell, *e.g.*, a leukocyte, lymphocyte, T cell, mast cell, macrophage, *etc.* In another embodiment, the isolated cell is a cancer cell. In another embodiment, the cell is part of an isolated tissue or a tissue culture. In a further embodiment, the cell can be present in an animal, *e.g.*, an animal that is a disease model or a subject in need of treatment.

In some embodiments, contacting the calcium channel with the synthetic polypeptide in the animal inhibits calcium influx only in the lungs (*e.g.*, does not reduce global Ca^{2+}) of the animal. In some embodiments, the animal is a disease model, *e.g.*, a murine disease model.

In some embodiments, contacting the calcium channel with the synthetic polypeptide comprises delivering the synthetic polypeptide to a cell comprising the calcium channel.

In some embodiments, contacting the calcium channel with the synthetic polypeptide comprises delivering a polynucleotide encoding the synthetic polypeptide to a cell comprising the calcium channel.

In some embodiments, contacting the calcium channel with the synthetic polypeptide comprises delivering the synthetic polypeptide directly to the lungs (*e.g.*, administering the synthetic polypeptide, polynucleotide, vector, and/or composition (*e.g.*, pharmaceutical composition) of the present invention directly to the lungs), optionally by intranasal, intratracheal, inhalation, and/or intrapulmonary administration).

In some embodiments, contacting the calcium channel with the synthetic polypeptide comprises delivering the synthetic polypeptide to a target organ system (*e.g.*, administering the synthetic polypeptide, polynucleotide, vector, and/or composition of the present invention to a target organ system), optionally systemically, optionally by intravenous administration.

Target organ systems contemplated in the present invention may be any mammalian organ system to which the compositions and methods of the invention may provide benefit. Non-limiting examples of target organ systems include the nervous system (*e.g.*, PNS, CNS), the muscular system, the respiratory system (*e.g.*, the lungs, *e.g.*, the airways), the reproductive system, the lymphatic system, the renal system, the digestive system, systemically, or any combination thereof. In some embodiments, the target organ system is the lungs (*e.g.*, the airways). In some embodiments, the target organ system is the organism (*e.g.*, systemically).

Another aspect of the present invention provides a method of inhibiting an immune response in a subject, comprising delivering to the subject a synthetic polypeptide,

polynucleotide, vector, and/or composition (e.g., pharmaceutical composition) of the present invention, thereby inhibiting the immune response. In some embodiments, the immune response is inhibited by at least about 10%, *e.g.*, at least about 10%, 25%, 50%, 75%, or more. Inhibition of the immune response can be quantitated by methods known in the art, *e.g.*, by measurement of the level of immune system cells and/or antibodies in the blood or in the tissue of a subject.

Another aspect of the present invention provides a method of inhibiting inflammation in a subject, comprising delivering to the subject a synthetic polypeptide, polynucleotide, vector, and/or composition (e.g., pharmaceutical composition) of the present invention, thereby inhibiting the inflammation. In some embodiments, inflammation is inhibited by at least about 10%, *e.g.*, at least about 10%, 25%, 50%, 75%, or more. Inhibition of inflammation can be quantitated by methods known in the art, *e.g.*, by measurement of the level of immune system cells, interleukins, chemokines, or other biological effector molecules in the blood or in the tissue of a subject.

In some embodiments, delivering to the subject a synthetic polypeptide, polynucleotide, vector, and/or composition (e.g., pharmaceutical composition) of the present invention may comprise delivering the synthetic polypeptide, polynucleotide, vector, and/or composition (e.g., pharmaceutical composition) directly to the respiratory target organ system (e.g., the lungs, the airways), optionally via intranasal, intratracheal, inhalation, and/or intrapulmonary administration/delivery.

In some embodiments, delivering to the subject a synthetic polypeptide, polynucleotide, vector, and/or composition (e.g., pharmaceutical composition) of the present invention may comprise delivering the synthetic polypeptide, polynucleotide, vector, and/or composition (e.g., pharmaceutical composition) to a target organ system, optionally systemically, optionally via intravenous administration/delivery.

Another aspect of the present invention provides a method of treating or preventing a disorder responsive to inhibition of calcium influx in an airway in a subject in need thereof, comprising delivering to the airway of the subject a synthetic polypeptide, polynucleotide, vector, and/or composition (e.g., pharmaceutical composition) of the present invention, thereby treating or preventing the disorder. As used herein, the term "disorder responsive to inhibition of calcium influx in an airway," refers to any disease, disorder, or condition that can be treated and/or prevented by inhibiting calcium influx in an airway. The disorder in the methods of the invention can be, in non-limiting examples, asthma or respiratory allergies. In certain embodiments, the polypeptide is delivered by inhalation, *e.g.*, using an inhaler or nebulizer for

delivery by oral and/or nasal inhalation. In some embodiments, the polypeptide is delivered to the airway of the subject via intranasal, intratracheal, intrapulmonary, and/or inhalation delivery.

Disorders of application by the methods of the present invention may include, but are not limited to, inflammatory lung disorders such as asthma, chronic obstructive pulmonary disease, idiopathic pulmonary fibrosis, cystic fibrosis, non-cystic fibrosis bronchiectasis, and acute or chronic bronchitis. In some embodiments, the disorder is asthma. In some
5 embodiments, the disorder is chronic obstructive pulmonary disease ((COPD), e.g., chronic bronchitis or emphysema). In some embodiments, the disorder is an allergy (e.g., a dust allergy, pollen allergy, mold allergy (e.g., *Aspergillus fumigatus*), etc.

10 In some embodiments, the disorder may be a genetic inflammatory condition of the lungs. In some embodiments, the disorder may be cystic fibrosis.

In some embodiments, the disorder may be fibrosis, such as, but not limited to, idiopathic pulmonary fibrosis.

In some embodiments, the disorder may pneumonia, such as, but not limited to,
15 ventilator associated pneumonia (VAP), hospital acquired pneumonia (HAP), viral associated pneumonia and/or bacterial associated pneumonia or COVID-19.

In some embodiments, the disorder may be an infection, such as viral, bacterial, and/or fungal infection. Thus, in some embodiments, the infection may be a respiratory infection wherein the infectious agent is *S. aureus*, methicillin-resistant *Staphylococcus aureus* (MRSA), *P. aeruginosa*, *H. influenzae*, *K. pneumoniae*, *E. coli*, *Acinetobacter*,
20 *Enterobacter*, *S. pneumoniae*, *M. catarrhalis*, *M. pneumoniae*, *mycobacterium*, *mycoplasma pneumoniae*, fungi (e.g., *Aspergillus fumigates*, *Cryptococcus*, *Pneumocystis*, *Candida*), or any combination thereof. In some embodiments, the infection may be a viral respiratory infection wherein the infectious agent is a coronavirus (e.g., MHV-A59, SARS CoV, SARS-CoV-2, or
25 MERS-CoV), dengue virus, hepatitis virus, influenza virus (e.g., influenza A, influenza B), respiratory syncytial virus (RSV), rhinovirus, parainfluenza virus, metapneumovirus, or any other respiratory virus or any combination thereof.

Another aspect of the invention provides a method of treating or preventing a disorder responsive to inhibition of calcium influx in a subject in need thereof, comprising delivering
30 to the subject a therapeutically or prophylactically effective amount of a synthetic polypeptide, polynucleotide, vector, and/or composition (e.g., pharmaceutical composition) of the present invention, thereby treating or preventing the disorder. In some embodiments, the synthetic polypeptide, polynucleotide, vector, and/or composition of the present invention may be delivered directly to a target organ system. In some embodiments, the synthetic

polypeptide, polynucleotide, vector, and/or composition of the present invention may be delivered directly to the subject, e.g., systemically, optionally via intravenous delivery.

Accordingly, further disorders applicable to the present invention include, but are not limited to, acute pancreatitis, arthritis, allergy, Bell's Palsy, cardiovascular disease (CVD; e.g., including but not limited to vasculitis), dermatomyositis, diabetes, Guillain-Barré Syndrome, inflammatory bowel disease (e.g., Ulcerative Colitis And Crohn's Disease), Lupus, Myasthenia Gravis, psoriasis, reactive arthritis, renal inflammation, rheumatoid arthritis, sarcoidosis, scleroderma, infection (e.g., sepsis), Sjögren's Syndrome, temporal arteritis, Kawasaki disease, or any combination thereof.

In some embodiments, the disorder may be an infection, e.g., an infection in a particular target organ system and/or a systemic infection (e.g., sepsis). In some embodiments, the infection may be a bacterial, viral, and/or fungal infection. In some embodiments, the disorder may be sepsis.

In some embodiments, the infection may be a systemic and/or local target organ system infection wherein the infectious agent is bacterial (e.g., *S. aureus*, methicillin-resistant *Staphylococcus aureus* (MRSA), *P. aeruginosa*, *H. influenzae*, *K. pneumoniae*, *E. coli*, *Acinetobacter*, *Enterobacter*, *S. pneumoniae*, *M. catarrhalis*, *M. pneumoniae*, *mycobacterium*, *mycoplasma pneumonia*, group B streptococci, *Neisseria meningitidis*), fungal (e.g., *Aspergillus fumigates*, *Cryptococcus*, *Pneumocystis*, *Candida*), viral (e.g., coronavirus (e.g., MHV-A59, SARS CoV, SARS-CoV-2, or MERS-CoV), dengue virus, hepatitis virus, influenza virus (e.g., influenza A, influenza B), respiratory syncytial virus (RSV), rhinovirus, parainfluenza virus, metapneumovirus), or any combination thereof.

In some embodiments, the synthetic polypeptide, polynucleotide, vector, and/or composition (e.g., pharmaceutical composition) of the present invention may be delivered/administered systemically to the subject via intravenous delivery/administration.

In some embodiments, delivering the synthetic polypeptide, polynucleotide, vector, and/or composition (e.g., pharmaceutical composition) of the present invention to the subject does not inhibit calcium influx in the lungs of the subject.

In some embodiments of the present invention, the polypeptides of the invention are administered directly to a subject. Generally, the compounds of the invention will be suspended in a pharmaceutically-acceptable carrier (e.g., physiological saline) and administered orally or by intravenous infusion, or administered subcutaneously, intramuscularly, intrathecally, intraperitoneally, intrarectally, intravaginally, intranasally, intragastrically, intratracheally, or intrapulmonarily. In another embodiment, the intratracheal or intrapulmonary delivery can be

accomplished using a standard nebulizer, jet nebulizer, wire mesh nebulizer, dry powder inhaler, or metered dose inhaler. They can be delivered directly to the site of the disease or disorder, such as lungs, kidney, or intestines. The dosage required depends on the choice of the route of administration; the nature of the formulation; the nature of the patient's illness; the subject's size, weight, surface area, age, and sex; other drugs being administered; and the judgment of the attending physician. Suitable dosages are in the range of 0.01-100.0 µg/kg.

In some embodiments, the synthetic polypeptide is administered/delivered in an amount of about 0.1 mg/kg to about 10 mg/kg of the subject, e.g., about 0.1 mg/kg, 0.2 mg/kg, 0.3 mg/kg, 0.4 mg/kg, 0.5 mg/kg, 0.6 mg/kg, 0.7 mg/kg, 0.8 mg/kg, 0.9 mg/kg, 1 mg/kg, 1.25 mg/kg, 1.5 mg/kg, 1.75 mg/kg, 2 mg/kg, 3 mg/kg, 4 mg/kg, 5 mg/kg, 6 mg/kg, 7 mg/kg, 8 mg/kg, 9 mg/kg, or 10 mg/kg or any value or range therein. For example, in some embodiments, the synthetic polypeptide of the present invention may be administered/delivered in an amount of about 0.1 mg/kg to about 9.75 mg/kg, about 2 mg/kg to about 5 mg/kg, about 0.25 mg/kg to about 2.5 mg/kg, about 5 mg/kg to about 10 mg/kg, or about 0.50 mg/kg to about 1 mg/kg of the subject. In some embodiments, the synthetic polypeptides of the present invention may be administered/delivered in an amount of about 0.1 mg/kg, about 0.25 mg/kg, about 0.50 mg/kg, about 1 mg/kg, or about 4 mg/kg of the subject.

Wide variations in the needed dosage are to be expected in view of the variety of polypeptides available and the differing efficiencies of various routes of administration. The synthetic polypeptide of the present invention and/or a nucleic acid molecule, vector, and/or pharmaceutical composition comprising the same may be delivered in any number of doses and according to any regimen or schedule as needed to achieve the desired therapeutic effect. For example, oral administration would be expected to require higher dosages than administration by i.v. injection. Variations in these dosage levels can be adjusted using standard empirical routines for optimization as is well understood in the art. Administrations can be single or multiple (e.g., 2-, 3-, 4-, 6-, 8-, 10-, 20-, 50-, 100-, 150-, or more fold. For example, the synthetic polypeptide may be delivered in one or more doses, e.g., two or more, three or more, four or more doses, etc., or in other words, at least one, at least two, at least three, at least four or more doses, etc. In some embodiments, a single dose of the synthetic polypeptide is delivered. In some embodiments, multiple doses of the synthetic polypeptide are delivered.

In some embodiments, a synthetic polypeptide of the present invention may be delivered according to a regimen/schedule of continual dosing over an extended period of time (e.g., chronic dosing). Aspects of an appropriate regimen/schedule of dosing may be

ascertainable by a person of skill in the art, e.g., a physician, and adjusted according to relevant parameters of the subject and disorder such as noted above.

Encapsulation of the polypeptides in a suitable delivery vehicle (*e.g.*, polymeric microparticles or implantable devices) may increase the efficiency of delivery, particularly for oral delivery.

In some embodiments, the synthetic polypeptide is delivered by inhalation.

In some embodiments, delivering the synthetic polypeptide or pharmaceutical composition to the subject inhibits calcium influx only in the lungs (*e.g.*, does not reduce global Ca^{2+}) of the subject.

The polypeptide of the present invention can optionally be delivered in conjunction with other therapeutic agents. The additional therapeutic agents can be delivered concurrently with the polypeptide of the invention. As used herein, the word "concurrently" means sufficiently close in time to produce a combined effect (that is, concurrently can be simultaneously, or it can be two or more events occurring within a short time period before or after each other). Thus, in some embodiments, methods of the present invention may further comprise delivering to the subject a therapeutic agent as known in the art for the treatment of the disorders of the invention. In one embodiment of the invention, the polypeptide is delivered to a patient concurrently with a compound that treats and/or prevents asthma, *e.g.*, a bronchodilator such as a β agonist or a steroid. In other embodiments, the is delivered to a patient concurrently with a compound that treats and/or prevents allergies, *e.g.*, an antihistamine. In some embodiments, the combined activity of the polypeptide and the other therapeutic agent is superior to the other therapeutic agent alone.

Additional therapeutic agents as known in the art for the treatment of the disorders of the invention which may be delivered in conjunction with a polypeptide of the present invention include, but not limited to, dexamethasone, corticosteroids (*e.g.*, Fluticasone (Flovent HFA), Budesonide (Pulmicort Flexhaler), Mometasone (Asmanex Twisthaler), Beclomethasone (Qvar RediHaler), Ciclesonide (Alvesco)); remdesivir (*e.g.*, for COVID19); ellexacaftor, ivacaftor, lumacaftor, tezacaftor, ibuprofen, acebilustat, lenabasum (*e.g.*, for cystic fibrosis); hypertonic saline, dornase alfa (Pulmozyme®), amikacin, amoxicillin, augmentin, ampicillin, bactrim (sulfamethoxazole and/or trimethoprim) levofloxacin, doxycycline, tetracycline, piperacillin-taxobactam, vancomycin, clindamycin, azithromycin, clarithromycin, ciprofloxacin (*e.g.*, for pneumonia such as HAP and/or VAP), cephalexin, doxycycline hyclate, mepolizumab, reslizumab, Lebrikizumab, Tralokinumab, Tezepelumab, benralizumab, and any other antibiotic known for the treatment of pneumonia.

As a further aspect, the invention provides pharmaceutical formulations and methods of administering the same to achieve any of the therapeutic effects (*e.g.*, modulation of calcium influx) discussed above. The pharmaceutical formulation may comprise any of the reagents discussed above in a pharmaceutically acceptable carrier, *e.g.*, a polypeptide and/or nucleic acid and/or vector comprising the same. By "pharmaceutically acceptable" it is meant a material that is not biologically or otherwise undesirable, *i.e.*, the material can be administered to a subject without causing any undesirable biological effects such as toxicity.

The formulations of the invention can optionally comprise medicinal agents, pharmaceutical agents, carriers, adjuvants, dispersing agents, diluents, and the like.

The peptides of the invention can be formulated for administration in a pharmaceutical carrier in accordance with known techniques. *See, e.g.*, Remington, *The Science And Practice of Pharmacy* (9th Ed. 1995). In the manufacture of a pharmaceutical formulation according to the invention, the polypeptide (including the physiologically acceptable salts thereof) is typically admixed with, *inter alia*, an acceptable carrier. The carrier can be a solid or a liquid, or both, and may be formulated with the polypeptide as a unit-dose formulation, for example, a metered dose inhaler, which can contain from 0.01 or 0.5% to 95% or 99% by weight of the polypeptide. One or more polypeptide or a functional fragment or homolog thereof can be incorporated in the formulations of the invention, which can be prepared by any of the well-known techniques of pharmacy.

A further aspect of the invention is a method of treating subjects *in vivo*, comprising administering to a subject a pharmaceutical composition comprising a polypeptide of the invention in a pharmaceutically acceptable carrier, wherein the pharmaceutical composition is administered in a therapeutically effective amount. Administration of the polypeptide of the present invention to a human subject or an animal in need thereof can be by any means known in the art for administering compounds.

The formulations of the invention include those suitable for oral, rectal, topical, buccal (*e.g.*, sub-lingual), vaginal, parenteral (*e.g.*, subcutaneous, intramuscular including skeletal muscle, cardiac muscle, diaphragm muscle and smooth muscle, intradermal, intravenous, intraperitoneal), topical (*i.e.*, both skin and mucosal surfaces, including airway surfaces), intranasal, transdermal, intraarticular, intrathecal, and inhalation administration, administration to the liver by intraportal delivery, as well as direct organ injection (*e.g.*, into the liver, into the brain for delivery to the central nervous system, into the pancreas, or into a tumor or the tissue surrounding a tumor). The most suitable route in any given case will depend on the nature and

severity of the condition being treated and on the nature of the particular peptide which is being used.

For injection, the carrier will typically be a liquid, such as sterile pyrogen-free water, sterile normal saline, hypertonic saline, pyrogen-free phosphate-buffered saline solution, bacteriostatic water, or Cremophor EL[R] (BASF, Parsippany, N.J.). For other methods of administration, the carrier can be either solid or liquid.

For oral administration, the polypeptide can be administered in solid dosage forms, such as capsules, tablets, and powders, or in liquid dosage forms, such as elixirs, syrups, and suspensions. Polypeptides can be encapsulated in gelatin capsules together with inactive ingredients and powdered carriers, such as glucose, lactose, sucrose, mannitol, starch, cellulose or cellulose derivatives, magnesium stearate, stearic acid, sodium saccharin, talcum, magnesium carbonate and the like. Examples of additional inactive ingredients that can be added to provide desirable color, taste, stability, buffering capacity, dispersion or other known desirable features are red iron oxide, silica gel, sodium lauryl sulfate, titanium dioxide, edible white ink and the like. Similar diluents can be used to make compressed tablets. Both tablets and capsules can be manufactured as sustained release products to provide for continuous release of medication over a period of hours. Compressed tablets can be sugar coated or film coated to mask any unpleasant taste and protect the tablet from the atmosphere, or enteric-coated for selective disintegration in the gastrointestinal tract. Liquid dosage forms for oral administration can contain coloring and flavoring to increase patient acceptance.

Formulations suitable for buccal (sub-lingual) administration include lozenges comprising the compound in a flavored base, usually sucrose and acacia or tragacanth; and pastilles comprising the compound in an inert base such as gelatin and glycerin or sucrose and acacia.

Formulations of the present invention suitable for parenteral administration comprise sterile aqueous and non-aqueous injection solutions of the polypeptide, which preparations are preferably isotonic with the blood of the intended recipient. These preparations can contain antioxidants, buffers, bacteriostats and solutes which render the formulation isotonic with the blood of the intended recipient. Aqueous and non-aqueous sterile suspensions can include suspending agents and thickening agents. The formulations can be presented in unit/dose or multi-dose containers, for example sealed ampoules and vials, and can be stored in a freeze-dried (lyophilized) condition requiring only the addition of the sterile liquid carrier, for example, saline or water-for-injection immediately prior to use.

Extemporaneous injection solutions and suspensions can be prepared from sterile powders, granules and tablets of the kind previously described. For example, in one aspect of the present invention, there is provided an injectable, stable, sterile composition comprising a polypeptide or a functional fragment or homolog thereof of the invention, in a unit dosage form in a sealed container. The polypeptide or salt is provided in the form of a lyophilizate which is capable of being reconstituted with a suitable pharmaceutically acceptable carrier to form a liquid composition suitable for injection thereof into a subject. The unit dosage form typically comprises from about 1 mg to about 10 grams of the polypeptide or salt. When the polypeptide or salt is substantially water-insoluble, a sufficient amount of emulsifying agent which is pharmaceutically acceptable can be employed in sufficient quantity to emulsify the peptide or salt in an aqueous carrier. One such useful emulsifying agent is phosphatidyl choline.

Formulations suitable for rectal administration are preferably presented as unit dose suppositories. These can be prepared by admixing the polypeptide with one or more conventional solid carriers, for example, cocoa butter, and then shaping the resulting mixture.

Formulations suitable for topical application to the skin preferably take the form of an ointment, cream, lotion, paste, gel, spray, aerosol, or oil. Carriers which can be used include petroleum jelly, lanoline, polyethylene glycols, alcohols, transdermal enhancers, and combinations of two or more thereof.

Formulations suitable for transdermal administration can be presented as discrete patches adapted to remain in intimate contact with the epidermis of the recipient for a prolonged period of time. Formulations suitable for transdermal administration can also be delivered by iontophoresis (*see*, for example, Tyle, *Pharm. Res.* 3:318 (1986)) and typically take the form of an optionally buffered aqueous solution of the polypeptide. Suitable formulations comprise citrate or bis/tris buffer (pH 6) or ethanol/water and contain from 0.1 to 0.2M of the compound.

The polypeptide can alternatively be formulated for nasal administration or otherwise administered to the lungs of a subject by any suitable means, *e.g.*, administered by an aerosol suspension of respirable particles comprising the polypeptide, which the subject inhales. The respirable particles can be liquid or solid. The term "aerosol" includes any gas-borne suspended phase, which is capable of being inhaled into the bronchioles or nasal passages. Specifically, aerosol includes a gas-borne suspension of droplets, as can be produced in a metered dose inhaler or nebulizer, or in a mist sprayer. Aerosol also includes a dry powder composition suspended in air or other carrier gas, which can be delivered by insufflation from an inhaler device, for example. *See* Ganderton & Jones, *Drug Delivery to the Respiratory Tract*, Ellis Horwood (1987); Gonda (1990) *Critical Reviews in Therapeutic Drug Carrier Systems* 6:273-

313; and Raeburn *et al.*, *J. Pharmacol. Toxicol. Meth.* 27:143 (1992). Aerosols of liquid particles comprising the polypeptide can be produced by any suitable means, such as with a pressure-driven aerosol nebulizer or an ultrasonic nebulizer, as is known to those of skill in the art. *See, e.g.*, U.S. Patent No. 4,501,729. Aerosols of solid particles comprising the polypeptide
5 can likewise be produced with any solid particulate medicament aerosol generator, by techniques known in the pharmaceutical art.

Alternatively, one can administer the polypeptide in a local rather than systemic manner, for example, in a depot or sustained-release formulation.

Further, the present invention provides liposomal formulations of the polypeptide
10 disclosed herein and salts thereof. The technology for forming liposomal suspensions is well known in the art. When the polypeptide or salt thereof is an aqueous-soluble salt, using conventional liposome technology, the same can be incorporated into lipid vesicles. In such an instance, due to the water solubility of the polypeptide or salt, the polypeptide or salt will be substantially entrained within the hydrophilic center or core of the liposomes. The lipid layer
15 employed can be of any conventional composition and can either contain cholesterol or can be cholesterol-free. When the polypeptide or salt of interest is water-insoluble, again employing conventional liposome formation technology, the salt can be substantially entrained within the hydrophobic lipid bilayer which forms the structure of the liposome. In either instance, the liposomes which are produced can be reduced in size, as through the use of standard sonication
20 and homogenization techniques.

The liposomal formulations containing the polypeptide disclosed herein or salts thereof, can be lyophilized to produce a lyophilizate which can be reconstituted with a pharmaceutically acceptable carrier, such as water, to regenerate a liposomal suspension.

In the case of a water-insoluble polypeptide or a functional fragment or homolog thereof,
25 a pharmaceutical composition can be prepared containing the water-insoluble polypeptide or a functional fragment or homolog thereof, such as for example, in an aqueous base emulsion. In such an instance, the composition will contain a sufficient amount of pharmaceutically acceptable emulsifying agent to emulsify the desired amount of the polypeptide. Particularly useful emulsifying agents include phosphatidyl cholines and lecithin.

30 In particular embodiments, the polypeptide is administered to the subject in a therapeutically effective amount, as that term is defined above. Dosages of pharmaceutically active polypeptides can be determined by methods known in the art, *see, e.g., Remington's Pharmaceutical Sciences* (Mack Publishing Co., Easton, Pa). The therapeutically effective dosage of any specific polypeptide or a functional fragment or homolog thereof will vary

somewhat from compound to compound, and patient to patient, and will depend upon the condition of the patient and the route of delivery. As a general proposition, a dosage from about 0.1 to about 50 mg/kg will have therapeutic efficacy, with all weights being calculated based upon the weight of the polypeptide, including the cases where a salt is employed. Toxicity concerns at the higher level can restrict intravenous dosages to a lower level such as up to about 10 mg/kg, with all weights being calculated based upon the weight of the polypeptide, including the cases where a salt is employed. A dosage from about 10 mg/kg to about 50 mg/kg can be employed for oral administration. Typically, a dosage from about 0.5 mg/kg to 5 mg/kg can be employed for intramuscular injection. Particular dosages are about 1 μ mol/kg to 50 μ mol/kg, and more particularly to about 22 μ mol/kg and to 33 μ mol/kg of the polypeptide for intravenous or oral administration, respectively.

In particular embodiments of the invention, more than one administration (*e.g.*, two, three, four, or more administrations) can be employed over a variety of time intervals (*e.g.*, hourly, daily, weekly, monthly, *etc.*) to achieve therapeutic effects.

The present invention finds use in veterinary and medical applications. Suitable subjects include both avians and mammals, with mammals being preferred. The term "avian" as used herein includes, but is not limited to, chickens, ducks, geese, quail, turkeys, and pheasants. The term "mammal" as used herein includes, but is not limited to, humans, bovines, ovines, caprines, equines, felines, canines, lagomorphs, *etc.* Human subjects include neonates, infants, juveniles, and adults.

The invention will now be described with reference to the following examples. It should be appreciated that these examples are not intended to limit the scope of the claims to the invention but are rather intended to be exemplary of certain embodiments. Any variations in the exemplified methods that occur to the skilled artisan are intended to fall within the scope of the invention.

EXAMPLES

Example 1: Novel Immunomodulatory Peptides for Lung Diseases and Disorders

A multi-platform approach was used to study the lung's peptidome for novel immunomodulatory and antiviral therapies, including *in vitro* optimization and *in vitro* and *in vivo* validation of lead compounds in acute and chronic disease models. These studies identified ELD607 as an Orai1/ Ca^{2+} inhibitor with broad spectrum efficacy as an immunomodulator in murine infection, allergic, and chronic inflammation models. This study

shows that ELD607 inhibits Orai1 in the lung, but not systemically, leading to effective local immunomodulatory action.

ELD100 peptide was incubated overnight in cystic fibrosis (CF) patient sputum, size-fractionated and then analyzed by mass spectrometry without tryptic digest. Three major fragments were found: namely, DITLV, HDIV and NMLIHG, suggesting that ELD100 was cleaved after valines by neutrophil elastase. ELD100 contained 16 residues. A series of peptides were generated including N-terminal, C-terminal and mid-peptide deletions to fully-optimize the peptide's size. This study found that N-terminal deletion of DITLV and removal of all valines was permissible, but that C-terminal deletions, beyond the terminal glycine were not. Using this approach, a library was generated to fully optimize ELD100. The ELD100 parent peptide sequence was extensively modified and superior peptidomimetics were developed that showed enhanced activity with shorter, more potent and more stable peptide sequences. The amino acid sequence modifications included alanine scanning, conservative mutations, shorter sequences and N- and C- terminus modifications (D-ala and Ser additions).

Alanine scan: To identify the key amino acids that retain the thapsigargin-induced inhibition of Ca^{2+} release seen with ELD100, single point changes were introduced to the ELD100 sequence by switching each amino acid to Ala, one at a time. **Table 1** shows the 16 sequences of peptides that resulted from this Ala scan, including the ELDEC ID and the alternative nomenclature. Peptides were pre-incubated for 3 h before adding thapsigargin and measuring fluorescence every 30 s for 7 m in HEK293T cells. The peptides were tested to evaluate the inhibition of Ca^{2+} release upon addition of thapsigargin. **FIG. 4** shows the Ca^{2+} levels for each peptide, including the controls (vehicle and thapsigargin) and the response of ELD100 for comparison.

Conservative mutations (e.g., replacing Q for N, K for R, etc.) maintain electrostatic charges in residues but alter their size. Conservative mutations were tested to evaluate whether these modifications improved potency of the resultant peptide (**FIG. 5** and **Table 2**). In addition, by testing shorter sequences, an improved peptide, ELD607, was found that is cheaper to synthesize and more soluble in aqueous solution (**FIG. 6** and **Table 3**). Amino (N) and carboxy (C) terminus optimization was also performed (**FIG. 7** and **Table 4**).

Based on these data, five peptides from the "600" series were optimized for resistance to proteolysis. HEK293T cells were plated and incubated for 2 hours with Fluo-4 plus or minus ELD peptides, with fluorescent images taken every 30 seconds for 2-6 minutes at excitation/emission wavelengths of 488/530 nm. **FIG. 8** shows measured cytoplasmic Ca^{2+}

after incubation with various ELD peptides, where several peptides including ELD604, 607, 100, and 621 reduced cytoplasmic calcium levels as compared to a control scrambled peptide. Candidate peptides ELD604, 607, 601, and 100 were tested for resistance against protease degradation, wherein the peptides were incubated with neutrophil elastase, treated with the elastase inhibitor sivelestat at various timepoints to halt ongoing reactions, and then analyzed by HPLC (**FIG. 9**).

These peptides were not degraded by trypsin. Neutrophil elastase, which is one of the major proteases in the CF lung, was also tested. ELD601 was the shortest peptides generated and lacked valines, which are a substrate for neutrophil elastase. Also made was ELD604, which is ELD601 flanked with D-alanines to prevent exoproteases. Additionally, a series of ELD601-based peptides were generated that incorporated the conservative mutations of **Table 2** that showed increased efficacy including ELD607, where histidine was substituted for an arginine. Despite lacking valines, ELD601 was still degraded by elastase. In contrast, ELD604 was significantly more stable. Surprisingly, despite containing valines and not being flanked by D-alanines, arginine substitution resulted in a significant increase in stability for ELD607 (**FIG. 9**).

Of the peptides tested in the mouse model, ELD607 was chosen as a lead peptide following analysis of peptide stability, size and efficacy (**FIG. 10**).

ELD607 is stable in CF sputum. CF sputum is highly proteolytic and SPLUNC1 is rapidly degraded in CF sputum. Therefore, it was tested whether ELD607 was stable in CF sputum. ELD607 was incubated with CF sputum supernatant for 6 h at 37°C, and then the reaction was halted by passing the sputum and ELD607 through a 10 kDa cut off spin column at 4°C. Using HPLC, a peak could be detected that corresponded to ELD607 (**FIG. 11 panel A**). Neither normal nor CF sputum diminished the area under the curve of this peak, indicating that the peptide was stable over time (**FIG. 11 panel B**). To determine whether ELD607 remained functional after exposure to CF sputum, ELD607 that had been incubated for 6 h in either PBS or CF sputum was added to HEK293T cells. Consistent with ELD607 being stable in CF sputum, this peptide retained full functionality and was fully able to inhibit Ca^{2+} influx even after pre-exposure to CF sputum (**FIG. 11 panel C**). In contrast, ELD100, which is not neutrophil elastase resistant, failed to inhibit Ca^{2+} influx after incubation in CF sputum. Importantly, ELD607 was incubated in CF sputum for 6 h before addition to cells, which constitutes a worst-case scenario: in CF patients, ELD607 will likely begin to exert its pharmacological actions much sooner after inhalation.

ELD607 does not interact with mucus. CF airways are characterized by mucus

accumulation/dehydration. The predicted pore size of a CF mucin mesh is ~200 nm while ELD607 is estimated to be ≤ 1 nm so ELD607 should diffuse through mucus at the same rate as it diffuses through water. However, this does not account for possible interactions between ELD607 and gel forming mucins. A previously mucus-coated quartz crystal microbalance with dissipation (QCM-D) was used to detect interactions (changes in dissipation [cross-linking] and frequency [mass]) when peptides are added. Tests were performed to determine whether ELD607 interacted with airway mucus which contained the MUC5AC and MUC5B mucins seen in the CF lung using QCM-D. While the positive control (Poly-L-lysine) bound to mucus and altered cross-linking ad mass, ELD607 did not affect mucus (**FIG. 12A**). Other peptides were also tested by QCM-D for frequency (**FIG. 12B**) and dissipation (**FIG. 12C**).

ELD607 requires Orai1 expression to inhibit Ca^{2+} signaling. This study found that ELD607 has a similar cell biology to SPLUNC1 and the $\alpha 6$ scaffold peptide (ELD100), and that ELD607 works by inhibiting Orai1. In the absence of extracellular Ca^{2+} , thapsigargin stimulates Ca^{2+} release from the ER and ELD607 had no effect on this component (**FIG. 13**). While ELD607 reduced Ca^{2+} influx, after shRNA knockdown of Orai1, ELD607 was no longer able to affect Ca^{2+} homeostasis (**FIG. 13**). These studies were followed by dose response tests for ELD607 using the Ca^{2+} add back approach shown in **FIG. 13**. In **FIG. 14**, the latter Ca^{2+} influx component was shown to be ELD607-sensitive and Orai1 mediated. Using this approach, curves were fitted to the data and the 50% inhibitory concentrations (IC_{50} s) were calculated. The maximal inhibitions are shown in **Table 5**.

Further *in vivo* experiments were performed. Mice were treated intratracheally with 10^7 colony forming units (CFU) of *P. aeruginosa* 1 hour prior to intranasal treatment with 0.51 mg/kg peptide, in a murine model of bacterial pneumonia. Bronchoalveolar lavage (BAL) and organ collection was performed 24 hours following treatment with peptide (**FIG. 15A**). Analysis of BAL monocytes (**FIG. 15B**) and neutrophils (**FIG. 15C**) showed that ELD607 significantly retained monocyte levels and reduced neutrophil levels in the BAL as compared to an uninfected naïve control. Moreover, 24 h after infection, about 10^9 CFUs could be detected in murine whole lung homogenate. However, ELD607 caused an $\sim 5 \log_{10}$ reduction in lung bacteria (**FIG. 15D**).

Analysis of BAL cytokine levels showed that ELD607 reduced Macrophage Inflammatory Protein 2 (MIP-2), also known as CXCL2, interleukin-17 (IL-17), and keratinocytes-derived chemokine (KC), also known as GRO α and CXCL2 (**FIG. 16A**). ELD607 also reduced interleukin-1 β (IL-1 β), tumor necrosis factor- α (TNF- α), and IL-6 (**FIG. 16B**), three inflammatory cytokines which have been shown to be elevated in COVID-

19 patients. Lung histology analysis showed increased inflammation with infection, and showed normalization of alveoli in the infected animals treated with ELD607 (**FIG. 16C**). This histology shows that the alveolar/gas exchange regions of murine lungs were protected by ELD607, preventing alveolar damage and thickening that can inhibit gas exchange during infection. Bronchoalveolar lavage (BAL) was collected from mice treated intratracheally with 10^7 colony forming units (CFU) of *P. aeruginosa* 1 hour prior to intranasal treatment with 0.51 mg/kg peptide, in a murine model of bacterial pneumonia, or vehicle-treated mice. Total protein, neutrophil elastase and lactate dehydrogenase (LDH) levels in the mouse *P. aeruginosa* infection model were also significantly reduced by ELD607 (**FIG. 16D**). ELD607 also reduced bacterial counts in the spleen (**FIG. 16E**), showing that an inhaled therapeutic can have systemic effects such as reducing sepsis. Similarly, ELD607 also prevented the total weight loss seen with intranasal 10^7 CFU/ml *P. aeruginosa* infection (**FIG. 16F**). **FIG. 16G** shows histology and graphs of BAL analysis, suggesting that ELD607 reduces Orai1 in murine BAL macrophages. Mice were exposed to vehicle or *P. aeruginosa* $\pm$ ELD607 and BAL was obtained and stained with an anti-Orai1 antibody, fluorescent 2° and DAPI. **FIG. 16G panel A** shows representative images of macrophages from the different groups as indicated. **FIG. 16G panels B and C** show plots showing fluorescent intensities of Orai1 in macrophages and neutrophils, respectively.

In a survival study of mice treated with 10^9 CFU/ml *P. aeruginosa*, ELD607 also significantly improved survival at 48 hours post-infection (**FIG. 17A**). BAL neutrophils were also reduced when ELD607 was added 24 hours after *P. aeruginosa* infection (**FIG. 17B**).

ELD607 showed no effect on *P. aeruginosa* growth, indicating that its effects on infection parameters is immunomodulatory. In contrast, penicillin-streptomycin and ciprofloxacin controls fully inhibited bacterial growth (**FIG. 18**).

ELD607 reduces neutrophilia and increases lung bacterial clearance in multiple murine bacterial infection models. Neutrophilia is a common feature of both Gram-positive and Gram-negative lung infections. Accordingly, further studies were performed with *P. aeruginosa*, *Staphylococcus aureus*, and *Haemophilus influenzae* (**FIG. 19 panels B, C, G, H, L, and M**). After infection with all three organisms, significant levels of bacteria were detected in the lung and spleen, and the infections increased total BAL cell numbers due to neutrophilia, while relative BAL macrophage numbers declined (**FIG. 19 panels A-O**). ELD607, but not ELD100, significantly normalized lung macrophage levels and reduced neutrophilia 24 h after infection. Moreover, 24 h after infection, about 10^9 CFUs could be detected in murine whole lung homogenate. However, ELD607 caused an ~ 3 -5 log₁₀

reduction in lung bacteria (**FIG. 19 panels D and I**) and significantly reduced sepsis (**FIG. 19 panels E and J**). Importantly, while the ELD100 scaffold protein has essentially no effect, in all cases, ELD607 significantly improved outcomes.

Additional models were tested such as the house dust mite (HDM) allergic lung inflammation model (**FIG. 20**); and SCNN1B-overexpression, wherein mice develop mucus obstruction and spontaneous pulmonary inflammation (**FIG. 21**).

These data show that ELD607 inhibits calcium flux upstream of lung cytokines and can reduce lung cytokine levels, eosinophilia, neutrophilia, lung inflammation, lung damage, and sepsis, relevant parameters to many lung diseases and disorders including, but not limited to, allergens (reduced eosinophilia, circulating IgG, and mast cell Ca^{2+} , reduced inflammation), bacterial infections (reduced lung cytokines, neutrophilia, lung damage, and sepsis, and improved survival), viral infections (reduced viral replication, predicted reduced cytokines, neutrophilia, lung damage, and improved survival), and mucosal disorders (reduced inflammation and neutrophilia).

To examine Orai1 levels, BAL was then obtained from *P. aeruginosa*-infected mice, immediately cytopun, fixed in PFA and stained with an antibody against Orai1. A significant decrease in Orai1 levels was seen in naïve and *P. aeruginosa*-infected mice after ELD607 exposure, indicating successful target engagement.

ELD607 inhibits viral infections. ACE2 protease cDNA was overexpressed in HEK293T cells for 24 hours and then 24 hours later these cells were infected with SARS-CoV-2 S1 Spike protein, DsRed, and VSVG pseudotyped into a lentivirus $\pm$ peptide as indicated in **FIG. 22**. Since this virus expressed DsRed in cells that it infected, DsRed fluorescence could be measured using an imaging plate reader as a marker of infection. ELD607, but not the control scrambled peptide, significantly reduced infection (**FIG. 22**). This experiment was repeated with other peptides in the 600 series, showing that ELD604 and ELD621 also inhibited viral replication (**FIG. 23**).

ELD607 normalized leukocyte composition and viral loads in the airways after murine coronavirus infection. Coronaviruses are emerging pathogens with severe outcomes in the human population. Mice were infected with 10^6 PFU mouse hepatitis virus (MHV-A59; a murine coronavirus) intranasally. The mice were then dosed with 1.0 mg/kg ELD607 or vehicle intranasally 4 days p.i. and then 2 days after dosing with ELD607 (6 days p.i.), mice were sacrificed to collect BAL and lung homogenates. ELD607 significantly reduced lung lymphocyte levels (B cells and T cells) and normalized lung monocyte and macrophage

levels (**FIG. 24 panels A and B**). Moreover, after only one dose of ELD607, viral loads in the lungs and BAL were significantly reduced (**FIG. 24 panel C**).

Thus, ELD607 beneficially reduces inflammation in both acute and chronic models. Without wishing to be bound to theory, ELD607 may moderate local inflammation in the lung lumen to reduce neutrophilia and allow macrophages to more efficiently clear bacteria without neutrophil-elastase-mediated lung injury. No evidence of immunocompromization was found, suggesting that ELD607 may be safe to administer to CF patients, who are typically chronically infected with multiple species of bacteria.

10 **Example 2: Further data regarding ELD607 as a novel peptide-based Orai1 inhibitor.**

HPLC analysis revealed that ELD607 was four times more stable than $\alpha 6$ after exposure to human neutrophil elastase (**FIG. 25, panel A**). Endogenous Orai1/store operated Ca^{2+} influx have been extensively studied in HEK293T cells, so this cell type was used to understand ELD607-Orai1 interactions. ELD607 was significantly more potent than $\alpha 6$ at inhibiting store operated Ca^{2+} influx (IC_{50} s were 9 and 124 nM respectively; **FIG. 25, panel B**). Thapsigargin is a SERCA pump inhibitor that prevents uptake of cytosolic Ca^{2+} into the endoplasmic reticulum. Thapsigargin is commonly used to stimulate Orai1/store operated Ca^{2+} influx. The inhibitory effects of $\alpha 6$ and ELD607 on thapsigargin-induced elevations in cytosolic Ca^{2+} were not additive, indicating that they share the same target (**FIG. 25, panel C**). Dynamic light scattering (DLS) was used to evaluate binding of ELD607 to peptides of Orai1 extracellular loops. The intensity weighted harmonic mean particle size (Z-average) significantly increased when Orai1 extracellular peptide 1 (OP1), corresponding to amino acids 106-119 (EVQLDADHDYPPGL; **SEQ ID NO:10**) of Orai1, displayed significant interactions with ELD607 but not scrambled ELD607 peptide control (**FIG. 25, panel D**). A peptide of Orai1's second extracellular loop (OP2) corresponding to amino acids 195-234 (CWVKFLPLKKQPGQPRPTSKPPASGAAANVSTSGITPGQA; **SEQ ID NO:11**) of Orai1 did not interact with any of these peptides (**FIG. 25, panel E**). Thus, it was concluded that ELD607 interacts with Orai1.

To determine whether ELD607 internalized Orai1, total internal reflection fluorescence (TIRF) microscopy was used to measure Orai1 in the plasma membrane. Neither vehicle control nor scrambled peptide significantly affected Orai1 levels. However, ELD607 reduced Orai1 plasma membrane expression by ~50% (**FIG. 26, panels A-B**).

Orai1 has been shown to regulate NFAT in multiple cell types, by indirectly dephosphorylating NFAT and allowing it to enter the nucleus where it can act as a transcription factor. We expressed NFAT-GFP in HEK293T cells and evaluated the ability of ELD607 to attenuate NFAT relocation to the nucleus after stimulation of SOCE by confocal microscopy. ELD607 reduced the percentage of NFAT in the nucleus by greater than 50% (**FIG. 26, panels C-D**).

ELD607 is active in ARDS patient immune cells. Pulmonary neutrophilic inflammation and increased lung neutrophil elastase levels are common in ARDS patient lungs. We therefore evaluated whether ELD607 remained stable in BAL samples patients with ARDS caused by bacterial pneumonia. Consistent with ELD607 showing increased resistance to neutrophil elastase (**FIG. 25, panel A**), ELD607 remained stable over 8 h in ARDS BAL samples at 37°C (**FIG. 27, panel A**). Consistent with ELD607 being stable in ARDS BAL, incubation of ELD607 in the ARDS BAL for extended periods did not prevent ELD607 from inhibiting store operated Ca^{2+} influx, when this BAL was placed on HEK293T cells loaded with ELD607 (**FIG. 27, panel B**).

We next obtained tracheal aspirates from intubated ARDS/bacterial pneumonia patients. We immediately isolated immune cells from these samples and simultaneously loaded them with the Ca^{2+} dye Fluo-4 and exposed them to vehicle, ELD607 or scrambled peptide for 1 h. Thapsigargin elevated cytosolic Ca^{2+} after vehicle or scrambled peptide exposure, but not after exposure to ELD607 (**FIG. 27, panel C**).

We then performed Orai1 immunocytochemistry on paired samples of immune cells of ARDS/bacterial pneumonia patients. Cells derived from each patient were exposed to either vehicle, ELD607, or scrambled ELD607 (**FIG. 27, panels D and E**). Again, ELD607, but not vehicle or scrambled peptide exposure, resulted in a significant reduction in Orai1 levels (**FIG. 27, panel E**). Taken together, these data indicate that inhaled ELD607 will be able to inhibit Orai1 in ARDS patient respiratory tracts.

ELD607 is non-toxic in vitro and in vivo. To investigate potential cytotoxic effects of ELD607, HEK294 T cells were exposed to 100 μM ELD607 for 24 h. Cytotoxicity was assessed by exposing cells to calcein to label live cells and fluorescence read by multi-plate reader. ELD607 treated cells were not significantly different to vehicle control indicating no toxic effect (**FIG. 28, panel A**). Primary human bronchial epithelial cultures were exposed to 1 mM ELD607 or vehicle control (saline) for 24 h. Cytotoxicity was assessed by confocal microscopy using calcein and propidium iodide to label live and dead cells respectively.

ELD607 treated cells were not significantly different to vehicle control indicating no toxic effect (**FIG. 28, panels B & C**). Transepithelial resistance was also measured before and after treatment as a measure of epithelial integrity. No differences were found between vehicle and ELD607 treated epithelia (**FIG. 28, panel D**).

To assess the pharmacokinetics and safety of ELD607 *in vivo*, mice were treated intratracheally with 10 mg/kg ELD607 daily for 7 days. Mice were sacrificed at 7 days post-treatment. Daily dosing of mice with 10 mg/kg ELD607 for 7 days did not induce any changes in lung histology (**FIG. 29, panels A and B**) or body weight (a general indicator of mouse health) (**FIG. 29, panel C**).

ELD607 is retained in the lung lumen. ELD607-TAMRA or vehicle (saline) was administered to mice intratracheally (0.5 mM, 1 h). Lungs were isolated, fixed, clarified and imaged using light-sheet fluorescence microscopy. ELD607-TAMRA was observed throughout the airway lumen (**FIG. 30, panel A**) but not in the interstitia (data representative of 3 individual experiments). We then dosed mice with 10 mg/kg ELD607 and collected BAL and blood. ELD607 levels declined over time in the BAL and plateaued with ~10% of the initial dose detected after 2 h (**FIG. 30, panel B**). After 10 mg/kg instillation of ELD607, less than 0.3% of the original dose was detected in the blood at any point in this time frame (**FIG. 30, panel C**) suggesting that ELD607 does not have systemic uptake.

ELD607, but not global Orai1 inhibition or steroids beneficially reduce pulmonary inflammation after P. aeruginosa infection. Orai1^{-/-} mice have low survival rates, so we used Orai1^{+/-} mice that have ~70% reduced Ca²⁺ influx (Gwack et al. 2008 *Mol cell Biol.* 28(17):5209-5222). Orai1^{+/-} mice infected with 10⁷ CFU/mouse *P. aeruginosa* POA1 and treated intranasally 1 h later with 0.5 mg/kg ELD607, were not able to normalize leukocyte levels (**FIG. 31, panels A-C**). In contrast, WT littermate controls infected with *P. aeruginosa* and treated with ELD607 had reduced normalized leukocytes similar to naïve mice after ELD607 exposure (**FIG. 31, panels B and C**). Additionally Orai1^{+/-} mice infected and then treated with ELD607 were not able to clear bacterial burden in the lungs (**FIG. 31, panel D**).

We then infected WT mice with *P. aeruginosa* and treated with ELD607, the Orai1 small molecule antagonist YM-58453, dexamethasone or scrambled ELD607. While ELD607 significantly reduced neutrophilia, weight loss and bacterial infections, and increased lung macrophage levels, neither -58453, dexamethasone nor scrambled ELD607 had any significant effects on these parameters (**FIG. 31, panels E-H**).

Repeated ELD607 exposure reduces chronic inflammation in βENaC overexpressing

mice. We added ELD607 to β ENaC overexpressing mice, which spontaneously develop chronic infection and neutrophilia and have increased mortality (**FIG. 32, panel A**). Daily intranasal addition of ELD607 (0.5 mg/kg) significantly reduced lung neutrophilia, and increased survival (**FIG. 32, panel B**).

5 *ELD607 reduces allergic inflammation.* Mice were exposed to house dust mite allergen for 14 days. 0.5 mg/kg ELD100 or ELD607 were instilled intranasally for 2 days before sacrifice. ELD607 significantly reduced BAL neutrophilia and eosinophilia, and increased BAL macrophages, in the mouse house dust mite model (**FIG. 33, panels A-C**).

10 *ELD607 reduces inflammation in an acute respiratory distress syndrome (ARDS) model.* Mice were intratracheally to 3 mg/kg *E. coli* lipopolysaccharide (LPS). 0.5 mg/kg ELD607 was instilled intranasally 1 or 6 h later. As a control, 3 mg/kg dexamethasone was given intranasally. ELD607 significantly reduced lung damage caused by LPS (**FIG. 34, panels A and B**), reduced weight loss (**FIG. 34, panel C**) and attenuated BAL neutrophilia (**FIG. 34, panel D**).

15

Example 3: ELD607 as a novel peptide-based Orai1 inhibitor in sepsis.

20 We injected mice intraperitoneally with *E. coli* LPS, followed by 0.5 mg/kg ELD607 injected intravenously 1 h later. ELD607 administered intravenously significantly prevented weight loss (**FIG. 35, panel A**), significantly reduced blood neutrophilia (**FIG. 35, panel B**) and significantly increased blood macrophages (**FIG. 35, panel C**). We next looked for ELD607 stability in human blood and found that it was 100% stable in healthy subject serum for 4 h (**FIG. 35, panel D**).

Table 1. Sequences of ELD100 peptide undergoing Alanine scan. The alternative names denote the amino acids changed and their position.

ELDEC ID	SEQUENCE	SEQ ID NO	ALTERNATIVE NAME
ELD1	DITLVHDIVNMLIHGL	12	α 6short
ELD31	AITLVHDIVNMLIHGL	13	D1
ELD32	DATLVHDIVNMLIHGL	14	I2
ELD33	DIALVHDIVNMLIHGL	15	T3
ELD34	DITAVHDIVNMLIHGL	16	L4
ELD35	DITLAHDIVNMLIHGL	17	V5
ELD36	DITLVADIVNMLIHGL	18	H6
ELD37	DITLVHAIVNMLIHGL	19	D7
ELD38	DITLVHDAVNMLIHGL	20	I8
ELD39	DITLVHDIANMLIHGL	21	V9
ELD40	DITLVHDIVAMLIHGL	22	N10
ELD41	DITLVHDIVNALIHGL	23	M11
ELD42	DITLVHDIVNMAIHGL	24	L12
ELD43	DITLVHDIVNMLAHGL	25	I13
ELD44	DITLVHDIVNMLIAGL	26	H14
ELD45	DITLVHDIVNMLIHAL	27	G15
ELD46	DITLVHDIVNMLIHGA	28	L16

Table 2. Sequences of ELD100 peptide undergoing conservative mutations. The alternative names denote the amino acids changed and their position.

ELDEC ID	SEQUENCE	SEQ ID NO	ALTERNATIVE NAME
ELD121	EITLVHDIVNMLIHGL-NH ₂	29	D1E
ELD122	NITLVHDIVNMLIHGL-NH ₂	30	D1N
ELD123	QITLVHDIVNMLIHGL-NH ₂	31	D1Q
ELD124	KITLVHDIVNMLIHGL-NH ₂	32	D1K
ELD125	DITLVHEIVNMLIHGL-NH ₂	33	D7E
ELD126	DITLVHNIVNMLIHGL-NH ₂	34	D7N
ELD127	DITLVHQIVNMLIHGL-NH ₂	35	D7Q
ELD128	DITLVHKIVNMLIHGL-NH ₂	36	D7K
ELD129	DITLVHDLVNMLIHGL-NH ₂	37	I8L
ELD130	DITLVHDVVNMLIHGL-NH ₂	38	I8V
ELD131	DITLVHDMVNMLIHGL-NH ₂	39	I8M
ELD132	DITLVHD-NIe-VNMLIHGL-NH ₂	40	I8NIe
ELD133	DITLVHDIVNMLIHGL-NH ₂	41	M11I
ELD134	DITLVHDIVNMLLIHGL-NH ₂	42	M11L
ELD135	DITLVHDIVN-NIe-LIHGL-NH ₂	43	M11NIe
ELD136	DITLVHDIVNMLMHGL-NH ₂	44	I13M
ELD137	DITLVHDIVNMLVHGL-NH ₂	45	I13V
ELD138	DITLVHDIVNMLLHGL-NH ₂	46	I13L
ELD139	DITLVHDIVNML-NIe-HGL-NH ₂	47	I13NIe
ELD140	DITLVHDIVNMLIQGL-NH ₂	48	H14Q
ELD141	DITLVHDIVNMLIYGL-NH ₂	49	H14Y
ELD142	DITLVHDIVNMLIRGL-NH ₂	7	H14R
ELD143	DITLVHDIVNMLINGL-NH ₂	50	H14N
ELD144	DITLVHDIVNMLIHPL-NH ₂	51	G15P
ELD145	DITLVHDIVNMLIHSL-NH ₂	52	G15S
ELD146	DITLVHDIVNMLIHEL-NH ₂	53	G15E
ELD147	DITLVHDIVNMLIHNL-NH ₂	54	G15N
ELD148	DITLVHDIVNMLIHGM-NH ₂	55	L16M
ELD149	DITLVHDIVNMLIHGI-NH ₂	56	L16I
ELD150	DITLVHDIVNMLIHGF-NH ₂	57	L16F
ELD151	DITLVHDIVNMLIHG-NIe-NH ₂	58	NL16NIe

Table 3. Sequences of ELD100 undergoing size optimization. The alternative names describe the change introduced compared to the parent peptide.

ELDEC ID	SEQUENCE	SEQ ID NO	ALTERNATIVE NAME
ELD3	DITLVHDIVNMLIH	59	α 6s - G
ELD5	MINTLDILGHIVHDV	60	α 6s scrambled
ELD6	DITLVH	61	α 6s-6-C
ELD8	DITLVHDI	62	α 6s-8-C
ELD9	DITLVHDIVN	63	α 6s-10-C
ELD10	DITLVHDIVNML	64	α 6s-12-C
ELD301	TLVHDIVNMLIHG	65	α 6s-13-N
ELD302	VHDIVNMLIHG	3	α 6s-11-N
ELD303	DIVNMLIHG	66	α 6s-9-N
ELD304	VNMLIHG	67	α 6s-7-N
ELD305	MLIHG	68	α 6s-5-N
ELD306	TLVHDIVNMLI	69	α 6s-11-NC
ELD307	VHDIVNM	70	α 6s-7-NC
ELD308	DIVNM	71	α 6s-5-NC
ELD310	DITLVHVDVNMLIHG	72	α 6s-14-middle
ELD311	DITLVHNMLIHG	73	α 6s-12-middle
ELD312	DITLVMLIHG	74	α 6s-10-middle
ELD313	DITLLIHG	75	α 6s-8-middle
ELD314	DITIHG	76	α 6s-6-middle
ELD315	DITLHDINMLIHG	77	α 6s-no V

Table 4. Sequences of ELD100 undergoing size optimization and capping changes. The alternative names describe the change introduced compared to the parent peptide.

ELDEC ID	SEQUENCE	SEQ ID NO	ALTERNATIVE NAME
ELD 601	HDINMLIHG	4	ELD302 noV
ELD 602	a-VHDIVNMLIHG-a	78	ELD-302 D-ala
ELD 603	a-VHDIVNMLIH-a	79	ELD-302 noG, D-ala
ELD 604	a-HDIVNMLIH-a	6	ELD302 noV, D-ala
ELD 605	VHDIVNMLIHP	80	ELD144-11
ELD 606	HDINMLIHP	81	ELD144-9
ELD 607	VHDIVNMLIRG	8	ELD142-11
ELD 608	HDINMLIRG	82	ELD142-11 noV
ELD 610	SVHDIVNMLIHG	83	ELD-302 N-Ser
ELD 611	VHDIVNMLIHGS	84	ELD-302 C-Ser
ELD 612	SVHDIVNMLIHGS	85	ELD-302 NC-Ser
ELD 620	n-HDINMLIHG-n	9	ELD-302 no V, C-amidated
ELD 621	n-VHDIVNMLIHG-n	5	ELD-302 C-amidated

Table 5. ELD607 is more potent and efficacious than ELD100.

Data calculated from FIG. 13.

	ELD607	ELD100
IC50 (Molar)	8.1E-08	3.7E-08
Bottom (%)	37.76	57.03

Table 6. Amino acid codon table.

Amino Acid			Codons
Alanine	Ala	A	GCA GCC GCG GCT
Cysteine	Cys	C	TGC TGT
Aspartic acid	Asp	D	GAC GAT
Glutamic acid	Glu	E	GAA GAG
Phenylalanine	Phe	F	TTC TTT
Glycine	Gly	G	GGA GGC GGG GGT
Histidine	His	H	CAC CAT
Isoleucine	Ile	I	ATA ATC ATT
Lysine	Lys	K	AAA AAG
Leucine	Leu	L	TTA TTG CTA CTC CTG CTT
Methionine	Met	M	ATG
Asparagine	Asn	N	AAC AAT
Proline	Pro	P	CCA CCC CCG CCT
Glutamine	Gln	Q	CAA CAG
Arginine	Arg	R	AGA AGG CGA CGC CGG CGT
Serine	Ser	S	AGC ACT TCA TCC TCG TCT
Threonine	Thr	T	ACA ACC ACG ACT
Valine	Val	V	GTA GTC GTG GTT
Tryptophan	Trp	W	TGG
Tyrosine	Tyr	Y	TAC TAT

Table 7. Amino acid derivatives.

Modified Amino Acid Residue	Abbreviation
2-Aminoadipic acid	Aad
3-Aminoadipic acid	bAad
beta-Alanine, beta-Aminopropionic acid	bAla
2-Aminobutyric acid	Abu
4-Aminobutyric acid, Piperidinic acid	4Abu
6-Aminocaproic acid	Acp
2-Aminoheptanoic acid	Ahe
2-Aminoisobutyric acid	Aib
3-Aminoisobutyric acid	bAib
2-Aminopimelic acid	Apm
t-butylalanine	t-BuA
Citrulline	Cit
Cyclohexylalanine	Cha
2,4-Diaminobutyric acid	Dbu
Desmosine	Des
2,2'-Diaminopimelic acid	Dpm
2,3-Diaminopropionic acid	Dpr
N-Ethylglycine	EtGly
N-Ethylasparagine	EtAsn
Homoarginine	hArg
Homocysteine	hCys
Homoserine	hSer
Hydroxylysine	Hyl
Allo-Hydroxylysine	aHyl
3-Hydroxyproline	3Hyp
4-Hydroxyproline	4Hyp
Isodesmosine	Ide
allo-Isoleucine	alle
Methionine sulfoxide	MSO
N-Methylglycine, sarcosine	MeGly
N-Methylisoleucine	Melle
6-N-Methyllysine	MeLys
N-Methylvaline	MeVal
2-Naphthylalanine	2-Nal
Norvaline	Nva
Norleucine	Nle
Ornithine	Orn
4-Chlorophenylalanine	Phe(4-Cl)
2-Fluorophenylalanine	Phe(2-F)
3-Fluorophenylalanine	Phe(3-F)
4-Fluorophenylalanine	Phe(4-F)
Phenylglycine	Phg
Beta-2-thienylalanine	Thi

The foregoing is illustrative of the present invention and is not to be construed as limiting thereof. The invention is defined by the following claims, with equivalents of the claims to be included therein.

THAT WHICH IS CLAIMED IS:

1. A synthetic polypeptide comprising one or more (e.g., at least one) of the following amino acid modifications, wherein the numbering corresponds to the amino acid sequence of **SEQ ID NO:1**:

SEQ ID NO:1:

14R substitution;
deletion of residues in positions 1-4;
deletion of residue in position 16;
deletion of valine residue(s); and/or
insertion of D-ala residue(s),

wherein the synthetic polypeptide binds to a Orai1 plasma membrane Ca^{2+} channel.

2. The synthetic polypeptide of claim 1, wherein the synthetic polypeptide comprises an amino acid sequence at least 70% identical to the amino acid sequence of **SEQ ID NO:1**.

3. The synthetic polypeptide of claim 1 or 2, wherein the synthetic polypeptide is 16 amino acid residues in length or less.

4. The synthetic polypeptide of any one of claims 1-3, wherein the synthetic polypeptide is resistant to elastase and/or trypsin protease cleavage.

5. The synthetic polypeptide of any one of claims 1-4, comprising the following amino acid modifications:

deletion of DITL residues in positions 1-4; and
L16 deletion.

6. The synthetic polypeptide of any one of claims 1-5, comprising the following amino acid modifications:

deletion of DITL residues in positions 1-4;
L16 deletion;
V5 deletion; and/or
V9 deletion.

7. The synthetic polypeptide of any one of claims 1-5, comprising the following amino acid modifications:
- deletion of DITL residues in positions 1-4;
 - L16 deletion;
 - C-terminal amidation; and/or
 - N-terminal amidation.
8. The synthetic polypeptide any one of claims 1-5, comprising the following amino acid modifications:
- deletion of DITL residues in positions 1-4;
 - L16 deletion;
 - V5 deletion;
 - G15 deletion;
 - C-terminal D-ala insertion; and/or
 - N-terminal D-ala insertion.
9. The synthetic polypeptide of any one of claims 1-4, comprising the following amino acid modifications:
- H14R substitution,
 - C-terminal amidation; and/or
 - N-terminal amidation.
10. The synthetic polypeptide of any one of claims 1-4 or 9, comprising the following amino acid modifications:
- H14R substitution;
 - deletion of DITL residues in positions 1-4; and/or
 - and L16 deletion.
11. The synthetic polypeptide of claim 1, comprising the amino acid sequence VHDIVNMLIHG (**SEQ ID NO:3**).
12. The synthetic polypeptide of claim 1, comprising the amino acid sequence HDINMLIHG (**SEQ ID NO:4**).

13. The synthetic polypeptide of claim 1, comprising the amino acid sequence n-VHDIVNMLIHG-n (**SEQ ID NO:5**).
14. The synthetic polypeptide of claim 1, comprising the amino acid sequence a-HDIVNMLIH-a (**SEQ ID NO:6**).
15. The synthetic polypeptide of claim 1, comprising the amino acid sequence DITLVHDIVNMLIRGL-NH₂ (**SEQ ID NO:7**).
16. The synthetic polypeptide of claim 1, comprising the amino acid sequence VHDIVNMLIRG (**SEQ ID NO:8**).
17. A polynucleotide encoding the synthetic polypeptide of any one of claims 1-16.
18. A vector comprising the polynucleotide of claim 17.
19. A cell comprising the polynucleotide of claim 17 and/or the vector of claim 18.
20. A kit comprising the synthetic polypeptide of any one of claims 1-16, the polynucleotide of claim 17, the vector of claim 18, and/or the cell of claim 19.
21. A composition comprising the synthetic polypeptide of any one of claims 1-16 and a carrier.
22. A pharmaceutical composition comprising the synthetic polypeptide of any one of claims 1-16 and a pharmaceutically acceptable carrier.
23. The pharmaceutical composition of claim 22, further comprising a therapeutic agent selected from the group consisting of dexamethasone, remdesivir, elexacaftor, ivacaftor, lumacaftor, tezacaftor, ibuprofen, acebilustat, lenabasum, levofloxacin, piperacillin-taxobactam, vancomycin, azithromycin, ciprofloxacin, cephalixin, doxycycline hyclate, and any combination thereof.

24. A dosage delivery device comprising the pharmaceutical composition of claim 22 or 23.
25. The dosage delivery device of claim 24, wherein the dosage delivery device is an inhaler.
26. A method of inhibiting calcium influx through a calcium channel, comprising contacting the calcium channel with the synthetic polypeptide of any one of claims 1-16 and/or the pharmaceutical composition of claim 22 or 23, thereby inhibiting calcium influx through the calcium channel.
27. A method of reducing expression of an Orai1 calcium channel, comprising contacting the calcium channel Orai1 with the synthetic polypeptide of any one of claims 1-16 and/or the pharmaceutical composition of claim 22 or 23.
28. The method of claim 26 or 27, wherein the calcium channel is present in an isolated cell.
29. The method of claim 28, wherein the isolated cell is in a cell culture.
30. The method of claim 26 or 27, wherein the calcium channel is in a cell in an animal.
31. The method of claim 30, wherein contacting the calcium channel with the synthetic polypeptide in the animal inhibits calcium influx only in the lungs (e.g., does not reduce global Ca^{2+}) of the animal.
32. The method of claim 30 or 31, wherein the animal is a disease model.
33. The method of claim 26 or 27, wherein contacting the calcium channel with the synthetic polypeptide comprises delivering the synthetic polypeptide to a cell comprising the calcium channel.

34. The method of claim 26 or 27, wherein contacting the calcium channel with the synthetic polypeptide comprises delivering a polynucleotide encoding the synthetic polypeptide to a cell comprising the calcium channel.
35. The method of any one of claims 30-34, wherein contacting the calcium channel with the synthetic polypeptide comprises delivering the synthetic polypeptide directly to the lungs (e.g., administering the synthetic polypeptide of any one of claims 1-16, the polynucleotide of claim 17, the vector of claim 18, and/or the composition of any one of claims 21-23 directly to the lungs, optionally by intranasal, intratracheal, inhalation, and/or intrapulmonary administration).
36. The method of any one of claims 30 or 32-34, wherein contacting the calcium channel with the synthetic polypeptide comprises delivering the synthetic polypeptide systemically (e.g., administering the synthetic polypeptide of any one of claims 1-16, the polynucleotide of claim 17, the vector of claim 18, and/or the composition of any one of claims 21-23 to a target organ system, optionally systemically, optionally by intravenous administration).
37. A method of inhibiting an immune response in a subject, comprising delivering to the subject the synthetic polypeptide of any one of claims 1-16, the polynucleotide of claim 17, the vector of claim 18, and/or the composition (e.g., pharmaceutical composition) of any one of claims 21-23, thereby inhibiting the immune response.
38. A method of inhibiting inflammation in a subject, comprising delivering to the subject the synthetic polypeptide of any one of claims 1-16, the polynucleotide of claim 17, the vector of claim 18, and/or the composition (e.g., pharmaceutical composition) of any one of claims 21-23, thereby inhibiting the inflammation.
39. The method of claim 38, wherein delivering to the subject the synthetic polypeptide (e.g., the synthetic polypeptide of any one of claims 1-16, the polynucleotide of claim 17, the vector of claim 18, and/or the composition of any one of claims 21-23) comprises delivering the synthetic polypeptide directly to the lungs, optionally by intranasal, intratracheal, inhalation, and/or intrapulmonary administration.

40. The method of claim 38, wherein delivering to the subject the synthetic polypeptide (e.g., the synthetic polypeptide of any one of claims 1-16, the polynucleotide of claim 17, the vector of claim 18, and/or the composition of any one of claims 21-23) comprises delivering the synthetic polypeptide to a target organ system, optionally systemically, optionally by intravenous administration.
41. A method of treating or preventing a disorder responsive to inhibition of calcium influx in an airway in a subject in need thereof, comprising delivering to the airway of the subject a therapeutically or prophylactically effective amount of the synthetic polypeptide of any one of claims 1-16, the polynucleotide of claim 17, the vector of claim 18, and/or the composition (e.g., pharmaceutical composition) of any one of claims 21-23 thereby treating or preventing the disorder.
42. The method of claim 41, wherein the disorder is asthma.
43. The method of claim 41, wherein the disorder is chronic obstructive pulmonary disease ((COPD), e.g., chronic bronchitis or emphysema).
44. The method of claim 41, wherein the disorder is an allergy.
45. The method of claim 41, wherein the disorder is cystic fibrosis.
46. The method of claim 41, wherein the disorder is fibrosis (e.g., idiopathic pulmonary fibrosis).
47. The method of claim 41, wherein the disorder is pneumonia (e.g., VAP, HAP).
48. The method of claim 41 or 47, wherein the disorder is an infection.
49. The method of claim 48, wherein the infection is a bacterial respiratory infection.
50. The method of claim 49, wherein the bacterium of the bacterial respiratory infection is *S. aureus*, methicillin-resistant *Staphylococcus aureus* (MRSA), *P. aeruginosa*, *H. influenzae*,

K. pneumonia, *E. coli*, *Acinetobacter*, *Enterobacter*, *S. pneumoniae*, *M. catarrhalis*, *M. pneumoniae*, *Mycobacterium*, *Mycoplasma pneumonia*, or any combination thereof.

51. The method of claim 48, wherein the infection is a fungal respiratory infection.

52. The method of claim 51, wherein the fungus of the fungal respiratory infection is *Aspergillus fumigates*, *Cryptococcus*, *Pneumocystis*, *Candida*, or any combination thereof.

53. The method of claim 48, wherein the infection is a viral respiratory infection.

54. The method of claim 53, wherein the virus of the viral respiratory infection is a coronavirus, dengue virus, hepatitis virus, influenza virus (e.g., influenza A, influenza B), respiratory syncytial virus (RSV), rhinovirus, parainfluenza virus, metapneumovirus, or any combination thereof.

55. The method of claim 54, wherein the coronavirus is MHV-A59, SARS CoV, SARS-CoV-2, or MERS-CoV.

56. The method of any one of claims 37-39 or 41-55, wherein the synthetic polypeptide is delivered to the airway of the subject via intranasal, intratracheal, intrapulmonary, and/or by inhalation delivery.

57. The method of any one of claims 37-39 or 41-56, wherein delivering the synthetic polypeptide or pharmaceutical composition to the subject inhibits calcium influx only in the lungs (e.g., does not reduce global Ca^{2+}) of the subject.

58. The method of any one of claims 37-57, wherein the synthetic polypeptide is administered/delivered in an amount of about 0.1 to about 10 mg/kg of the subject.

59. The method of claim 58, wherein the synthetic polypeptide is administered/delivered in an amount of about 0.50 mg/kg of the subject.

60. The method of any one of claims 37-59, wherein a single dose of the synthetic polypeptide is delivered.

61. The method of any one of claims 37-59, wherein multiple doses of the synthetic polypeptide are delivered (e.g., chronic dosing).

62. The method of any one of claims 41-61, further comprising delivering to the subject a therapeutic agent selected from the group consisting of dexamethasone, corticosteroids (e.g., Fluticasone (Flovent HFA), Budesonide (Pulmicort Flexhaler), Mometasone (Asmanex Twisthaler), Beclomethasone (Qvar RediHaler), Ciclesonide (Alvesco)), remdesivir (COVID19), elexacaftor, ivacaftor, lumacaftor, tezacaftor, ibuprofen, acebilustat, lenabasum (CF), hypertonic saline, dornase alfa (Pulmozyme®), amikacin, amoxicillin, augmentin, ampicillin, bactrim (sulfamethoxazole and trimethoprim) levofloxacin, doxycycline, tetracycline, piperacillin-taxobactam, vancomycin, clindamycin, azithromycin, clarithromycin, ciprofloxacin (HAP/VAP), cephalixin, doxycycline hyclate, mepolizumab, reslizumab, Lebrikizumab, Tralokinumab, Tezepelumab, benralizumab, and any combination thereof.

63. A method of treating or preventing a disorder responsive to inhibition of calcium influx in a subject in need thereof, comprising delivering to the subject a therapeutically or prophylactically effective amount of the synthetic polypeptide of any one of claims 1-16, the polynucleotide of claim 17, the vector of claim 18, and/or the composition (e.g., pharmaceutical composition) of any one of claims 21-23, thereby treating or preventing the disorder.

64. The method of claim 63, wherein the synthetic polypeptide, polynucleotide, vector, and/or composition is delivered directly to a target organ system.

65. The method of claim 63 or 64, wherein the disorder is acute pancreatitis, arthritis, allergy, Bell's Palsy, cardiovascular disease (CVD; e.g., including but not limited to vasculitis), dermatomyositis, diabetes, Guillain-Barré Syndrome, inflammatory bowel disease (e.g., Ulcerative Colitis And Crohn's Disease), Lupus, Myasthenia Gravis, psoriasis, reactive arthritis, renal inflammation, rheumatoid arthritis, sarcoidosis, scleroderma, infection (e.g., sepsis), Sjögren's Syndrome, temporal arteritis, Kawasaki disease, or any combination thereof.

66. The method of any one of claims 63-65, wherein the disorder is an infection.
67. The method of claim 66, wherein the infection is a bacterial infection.
68. The method of any one of claims 63-67, wherein the disorder is sepsis.
69. The method of claim 67 or 68, wherein the bacterium of the bacterial infection is *Staphylococcus aureus*, *Streptococcus pneumoniae*, *Haemophilus influenzae*, *Neisseria meningitidis*, *Klebsiella pneumoniae*, *Escherichia coli*, group B streptococci, or any combination thereof.
70. The method of claim 66, wherein the infection is a fungal infection.
71. The method of claim 70, wherein the fungus of the fungal infection is *Aspergillus fumigatus*, *Cryptococcus*, *Pneumocystis*, *Candida*, or any combination thereof.
72. The method of claim 66, wherein the infection is a viral infection.
73. The method of claim 72, wherein the virus of the viral infection is coronavirus, dengue virus, hepatitis virus, influenza virus (e.g., influenza A, influenza B), respiratory syncytial virus (RSV), rhinovirus, parainfluenza virus, or metapneumovirus, or any combination thereof.
74. The method of claim 73, wherein the coronavirus is MHV-A59, SARS CoV, SARS-CoV-2, or MERS-CoV.
75. The method of any one of claims 63-74, wherein the synthetic polypeptide is delivered systemically to the subject via intravenous delivery.
76. The method of any one of claims 63-75, wherein delivering the synthetic polypeptide or pharmaceutical composition to the subject does not inhibit calcium influx in the lungs of the subject.

77. The method of any one of claims 63-76, wherein the synthetic polypeptide is administered/delivered in an amount of about 0.1 to about 10 mg/kg of the subject.
78. The method of claim 77, wherein the synthetic polypeptide is administered/delivered in an amount of about 0.50 mg/kg of the subject.
79. The method of any one of claims 63-78, wherein a single dose of the synthetic polypeptide is delivered.
80. The method of any one of claims 63-78, wherein multiple doses of the synthetic polypeptide are delivered (e.g., chronic dosing).

FIG. 1

Orai1 – Ca^{2+} Channel and Master Regulator of Inflammation

- Orai1 is upstream of all cytokine responses
- Selective/local Orai1 inhibition is immunomodulatory

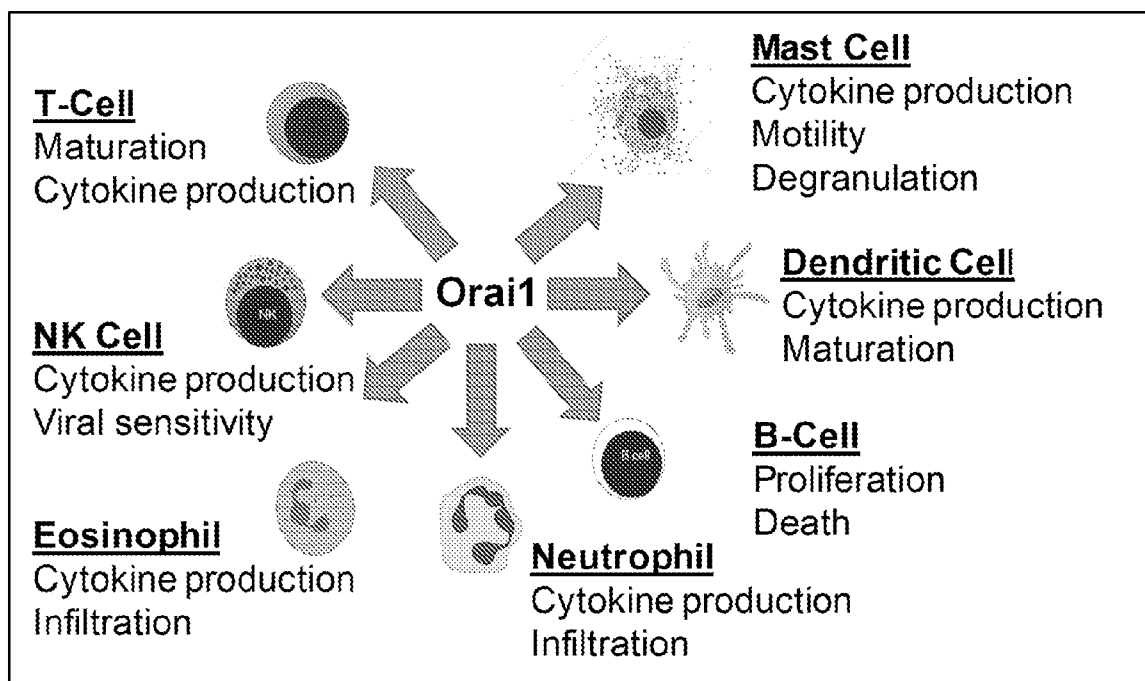


FIG. 2

A

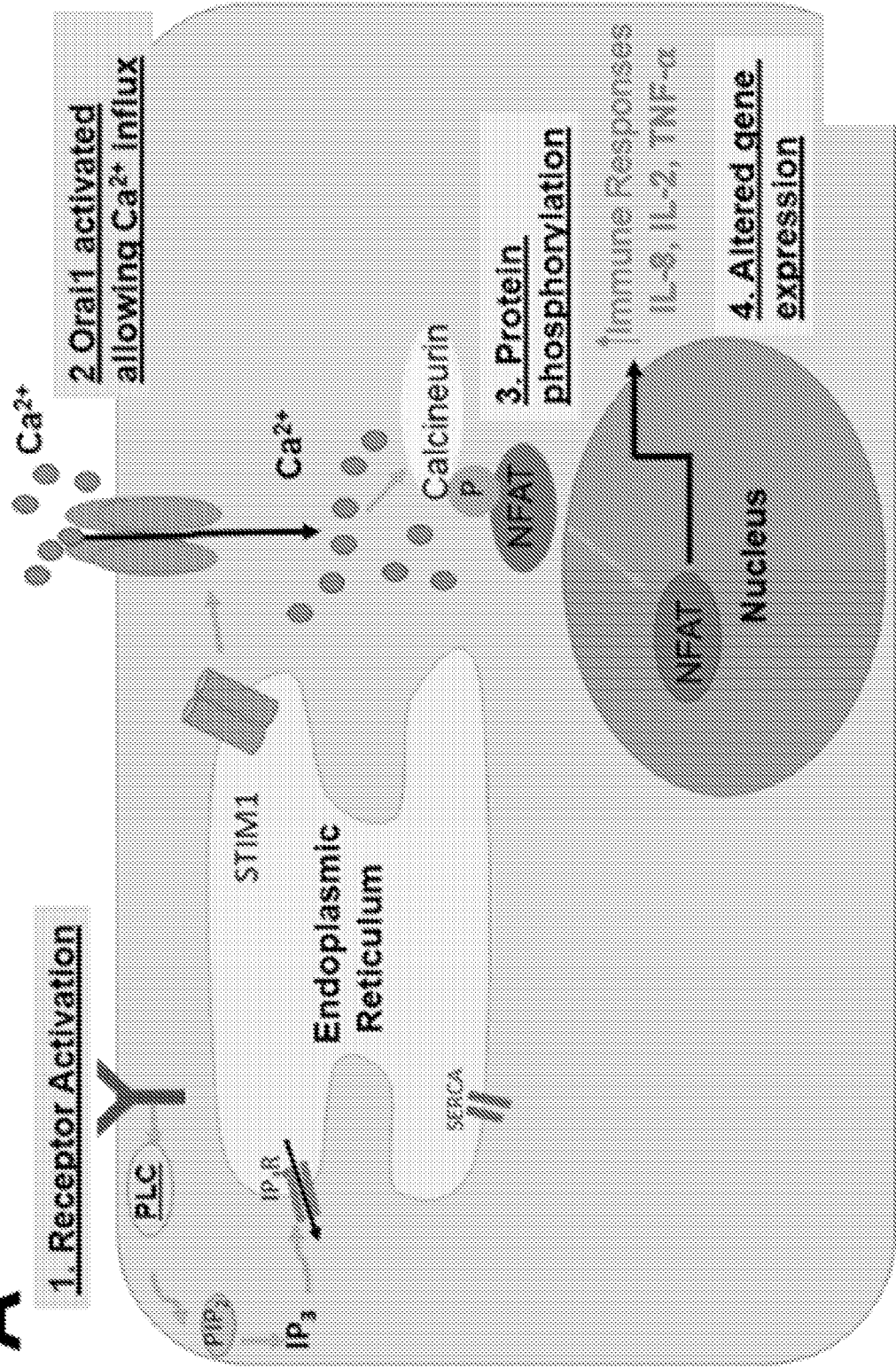
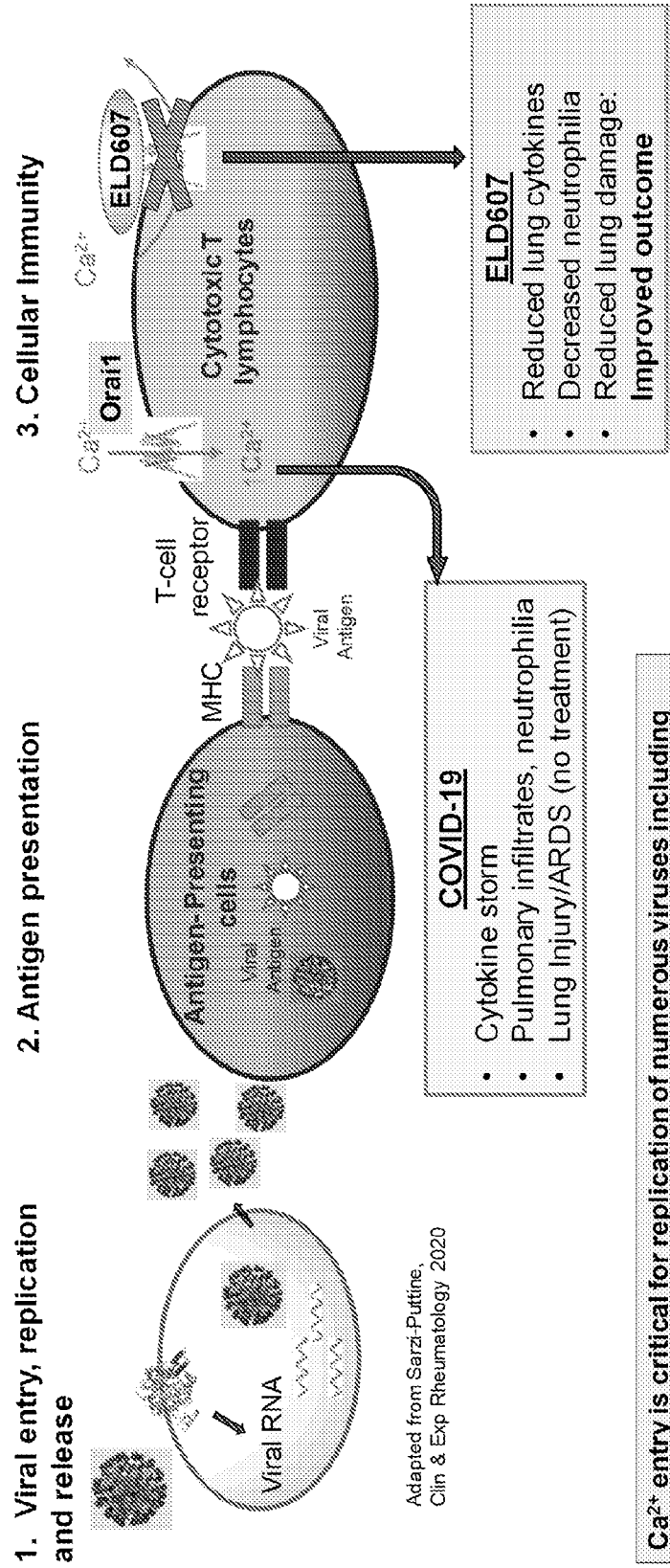


FIG. 3

ELD607 May Also Be a Key Immunomodulatory Therapy for COVID-19



Ca²⁺ entry is critical for replication of numerous viruses including Dengue and Hepatitis B. Clark & Eisenstein, Curr Top Med Chem, 2013.

FIG. 4

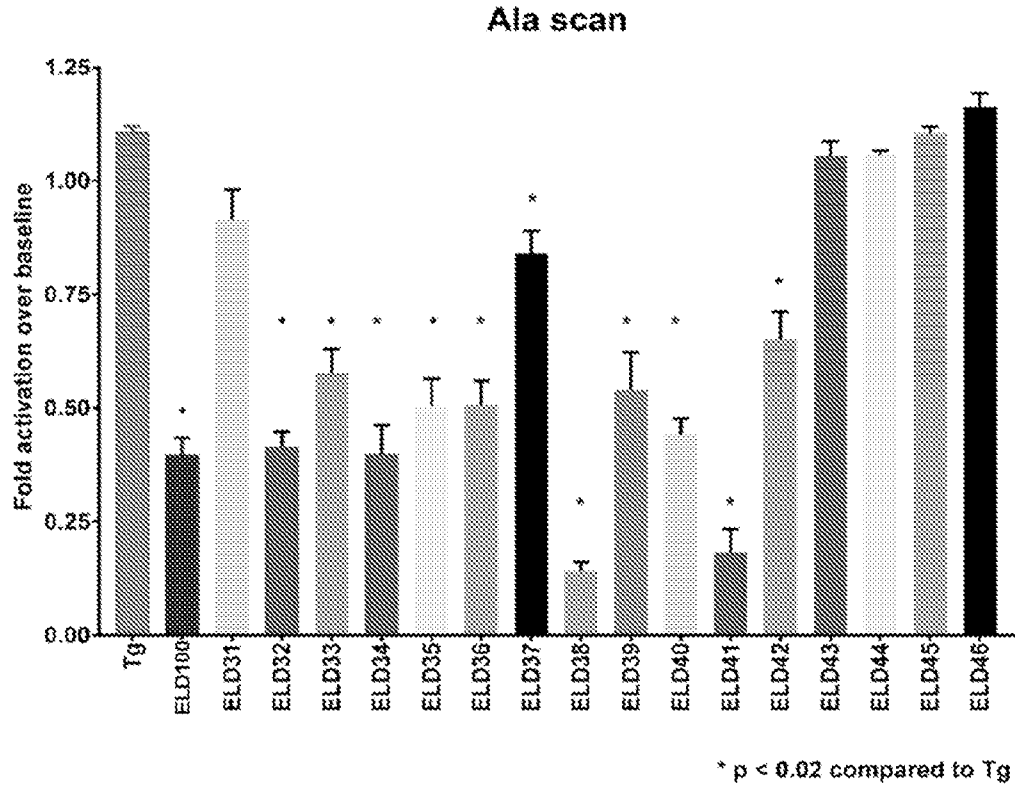


FIG. 5

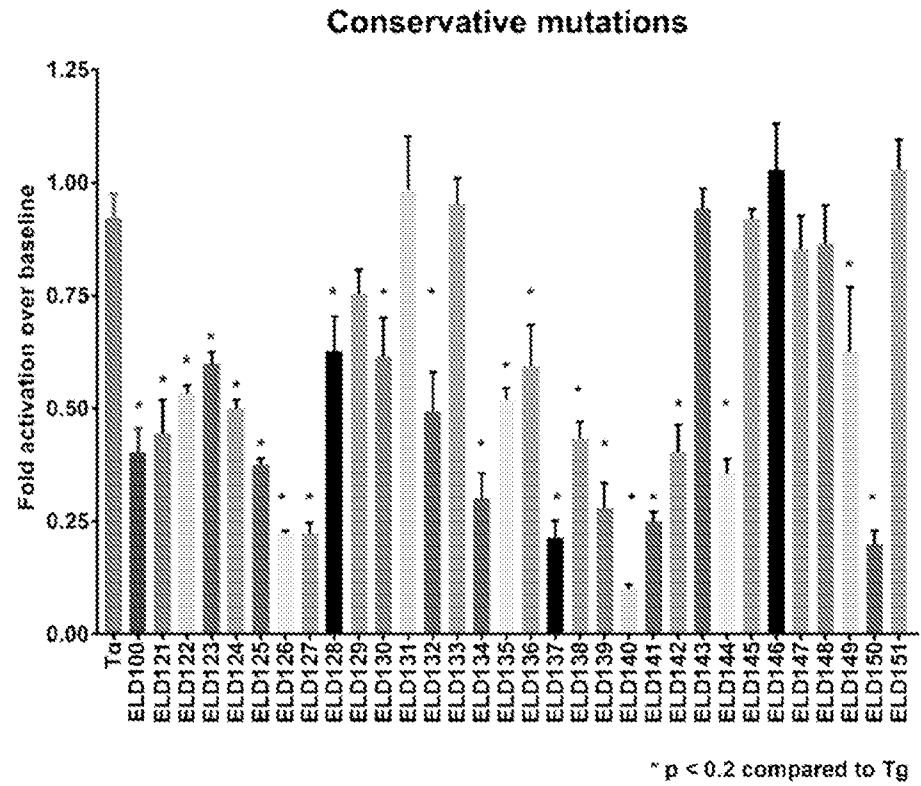


FIG. 6

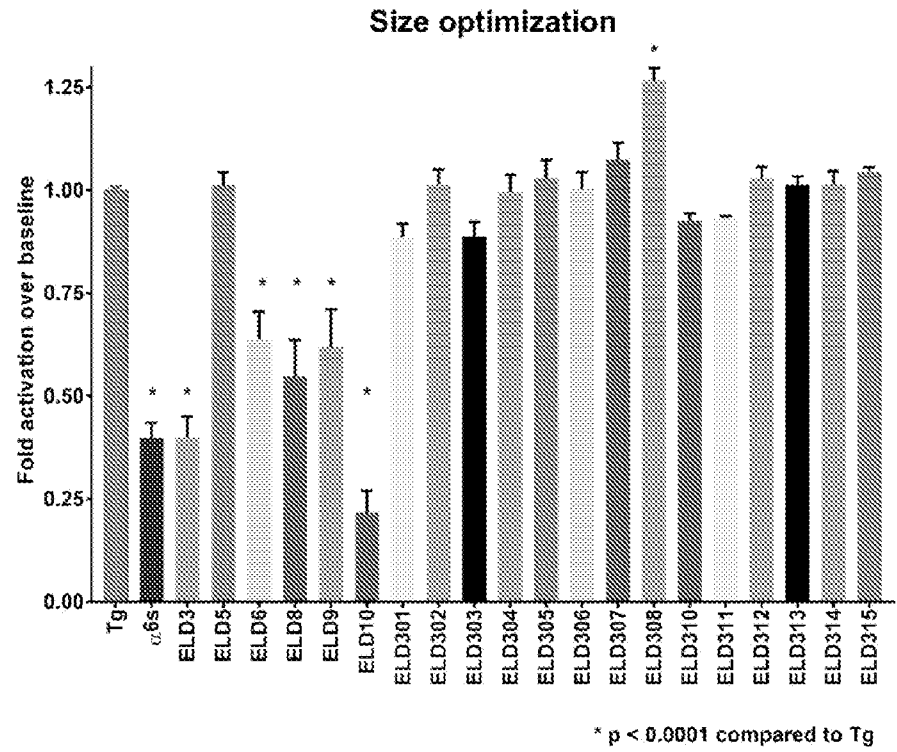


FIG. 7

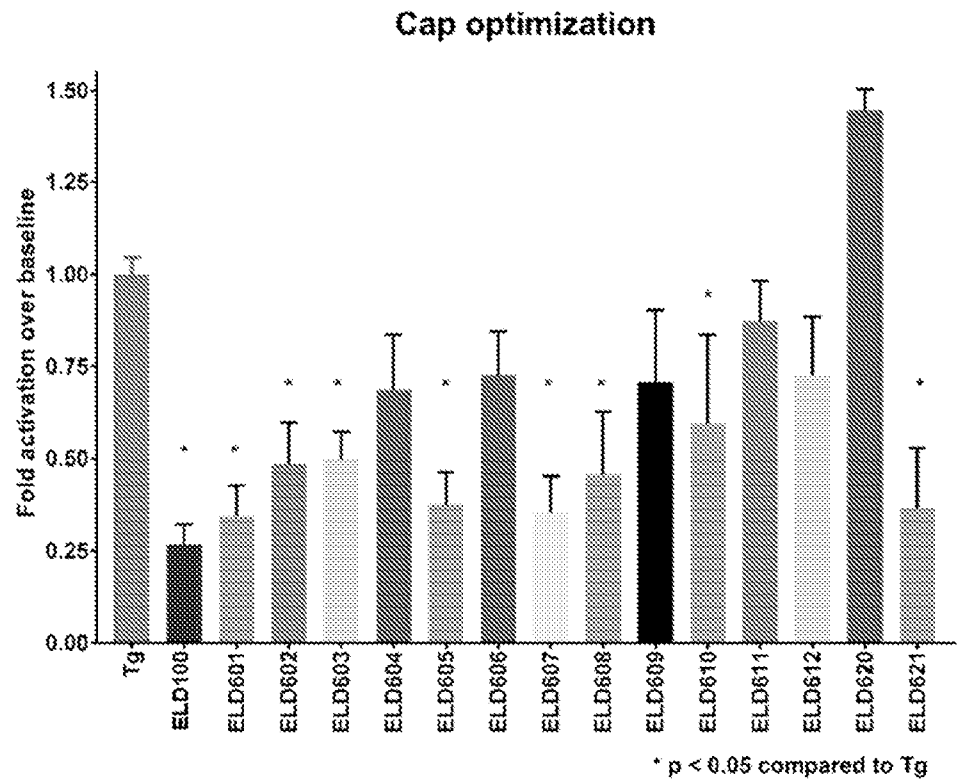


FIG. 8

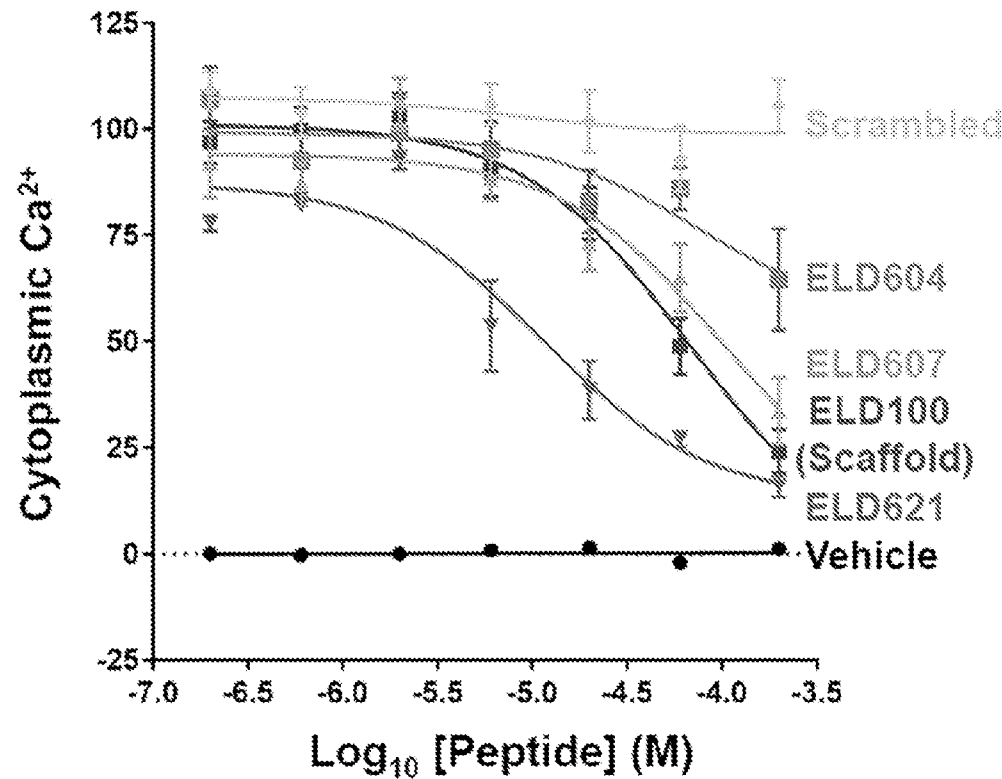
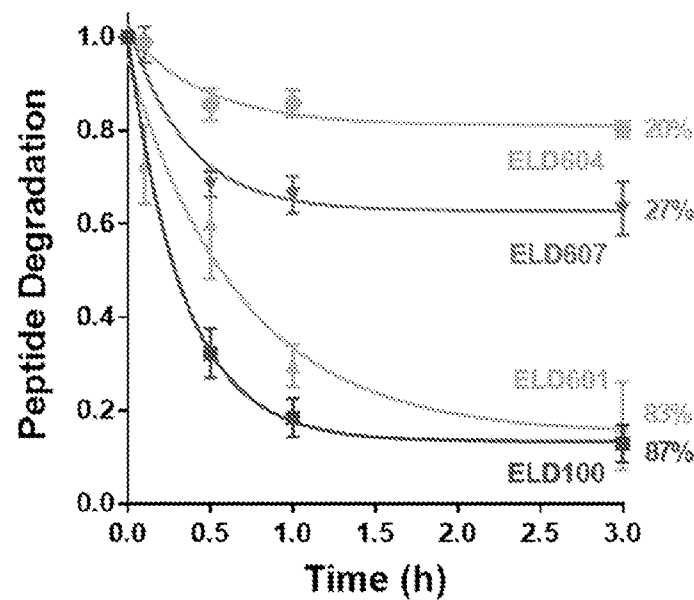


FIG. 9



7/42

FIG. 10

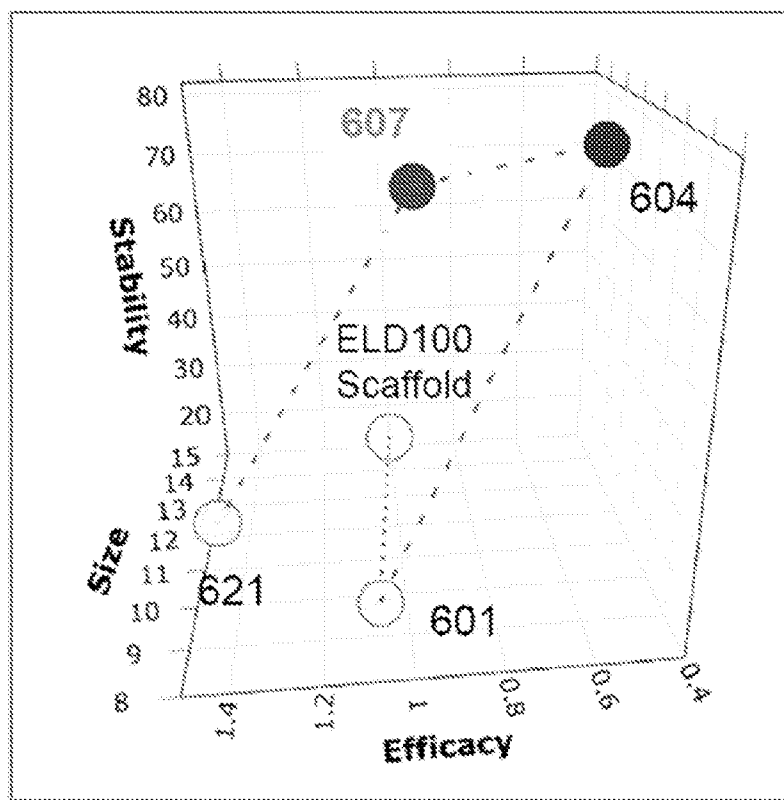
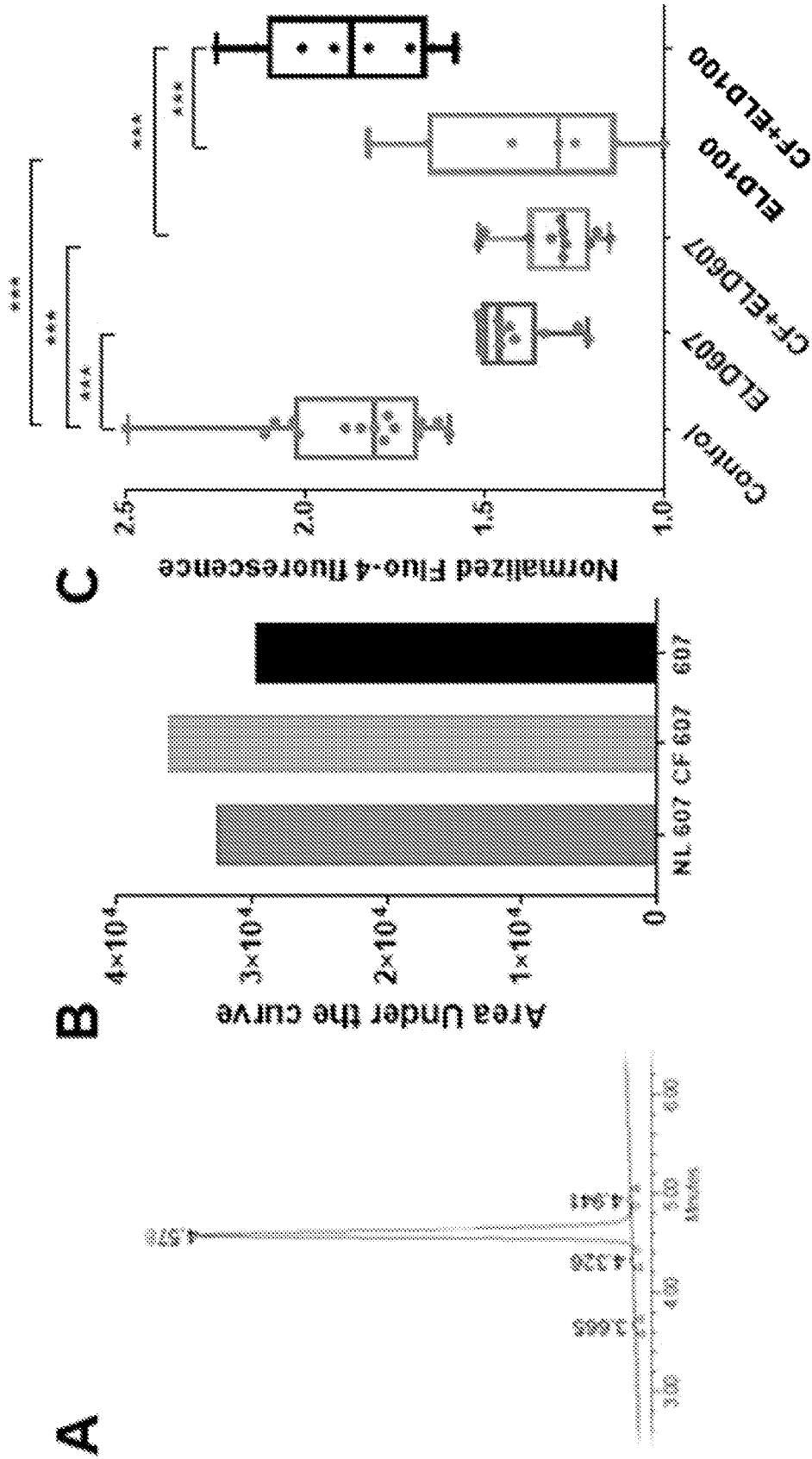


FIG. 11



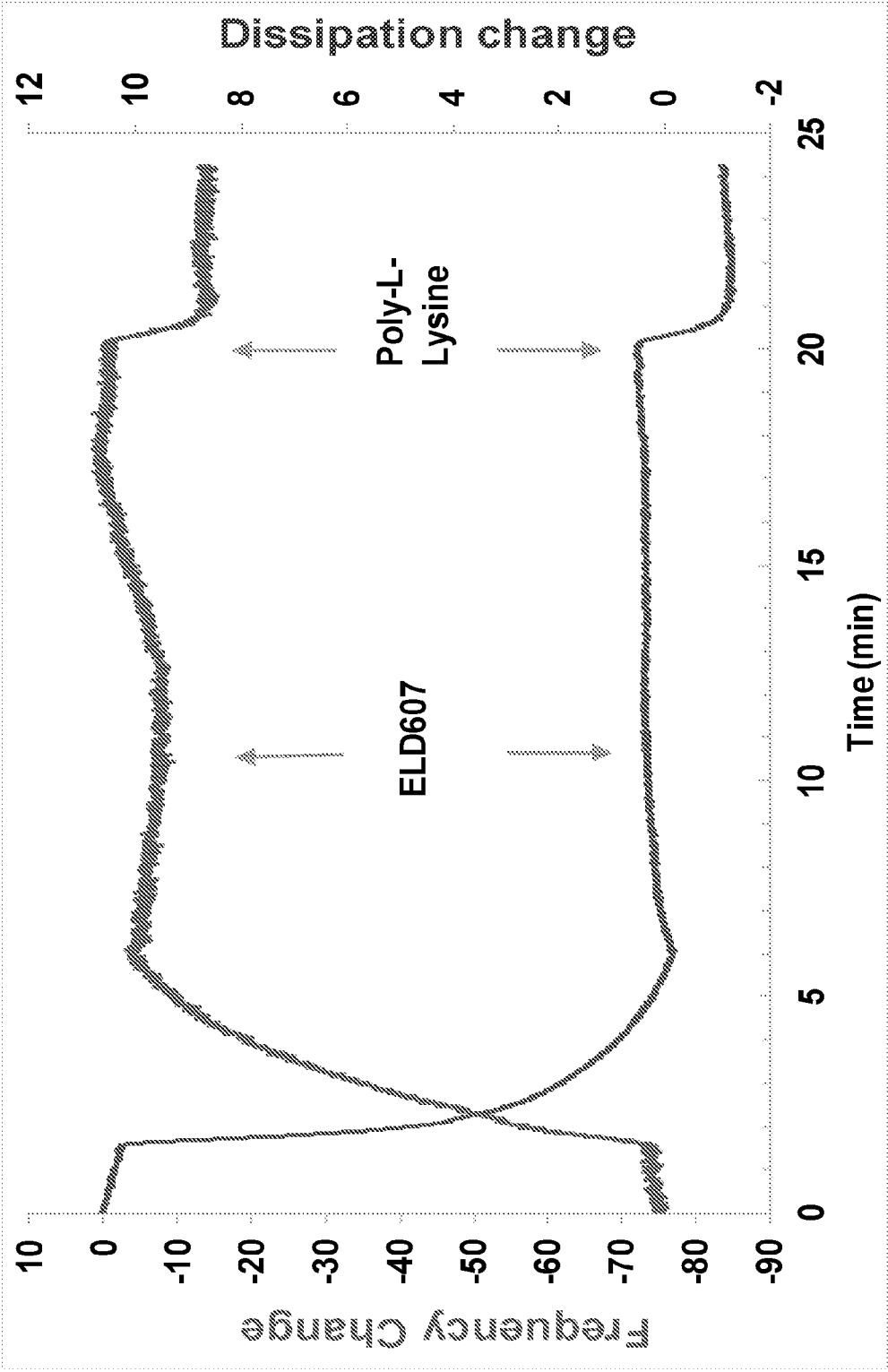


FIG. 12A

10/42

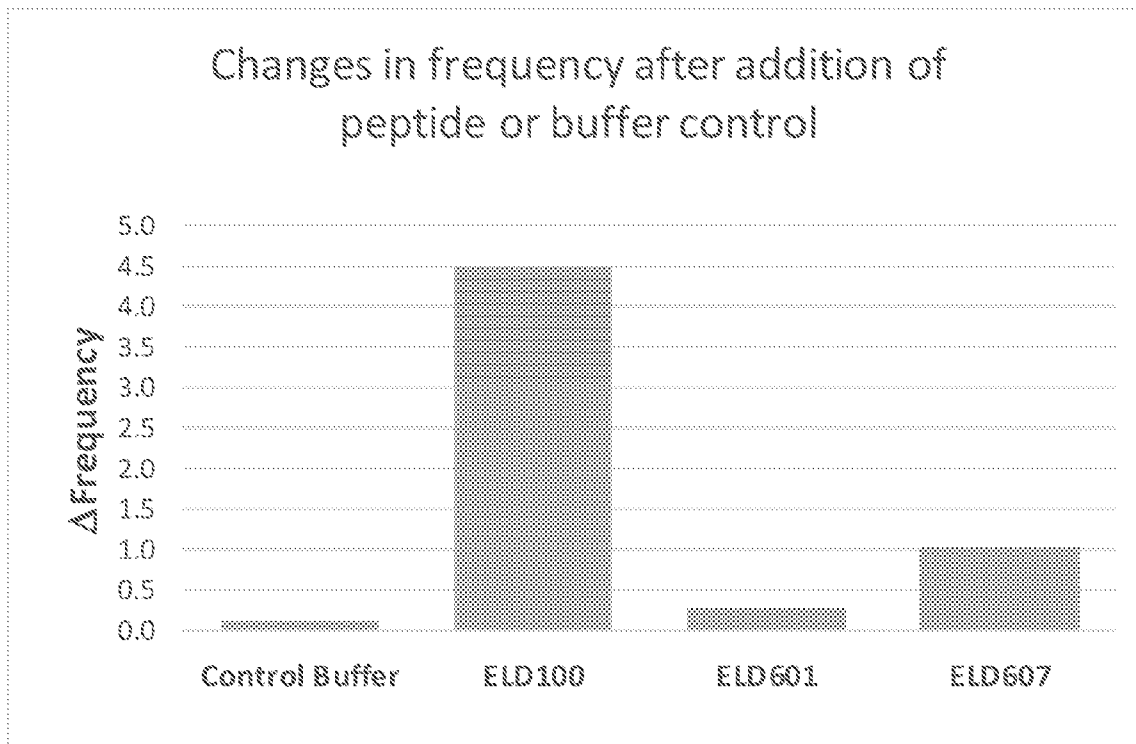
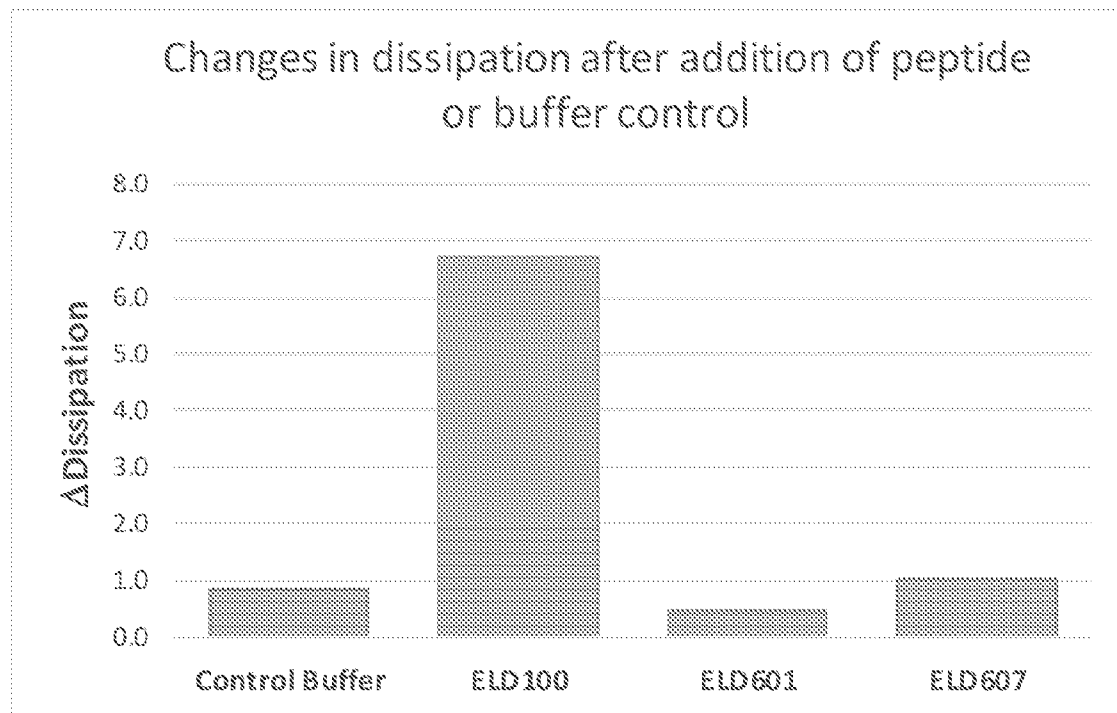
FIG. 12B**FIG. 12C**

FIG. 13

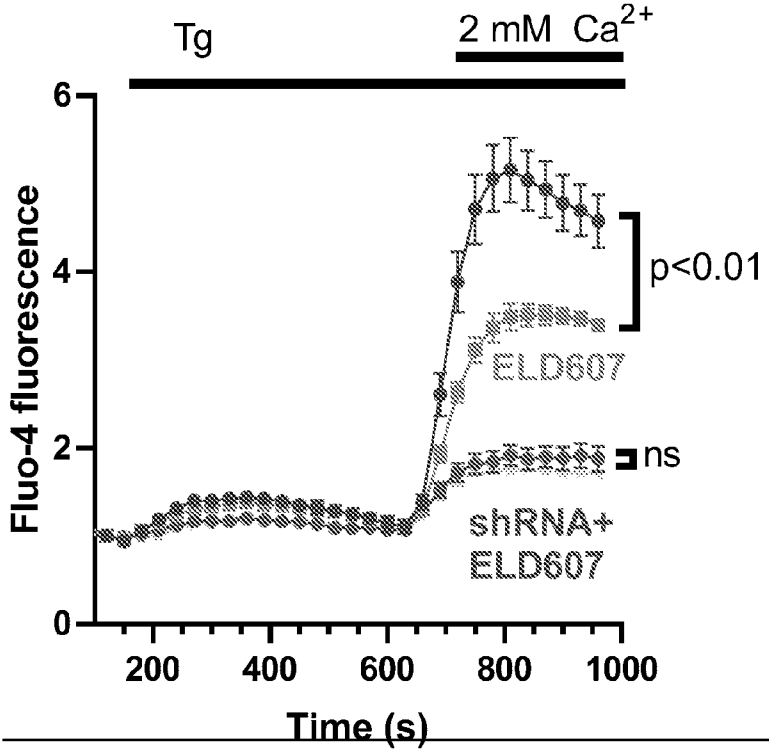
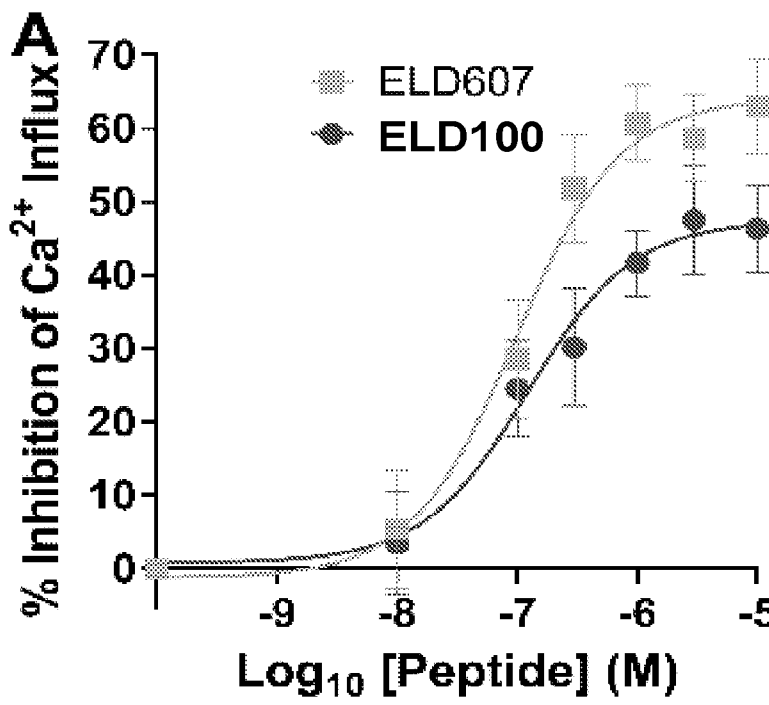


FIG. 14



12/42

FIG. 15A

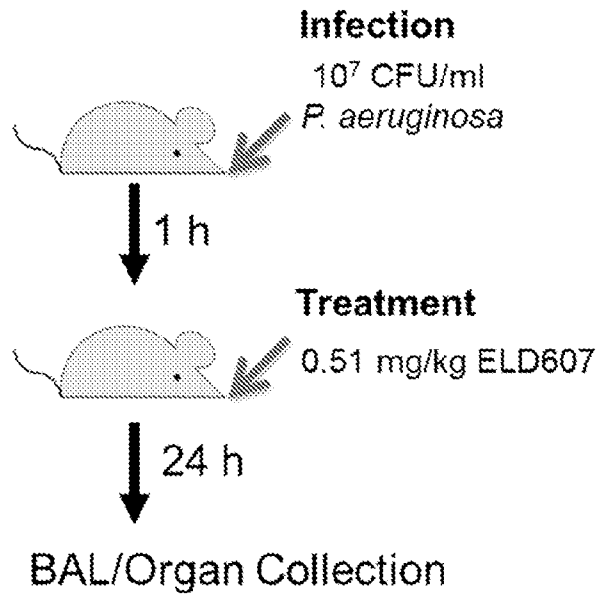


FIG. 15B

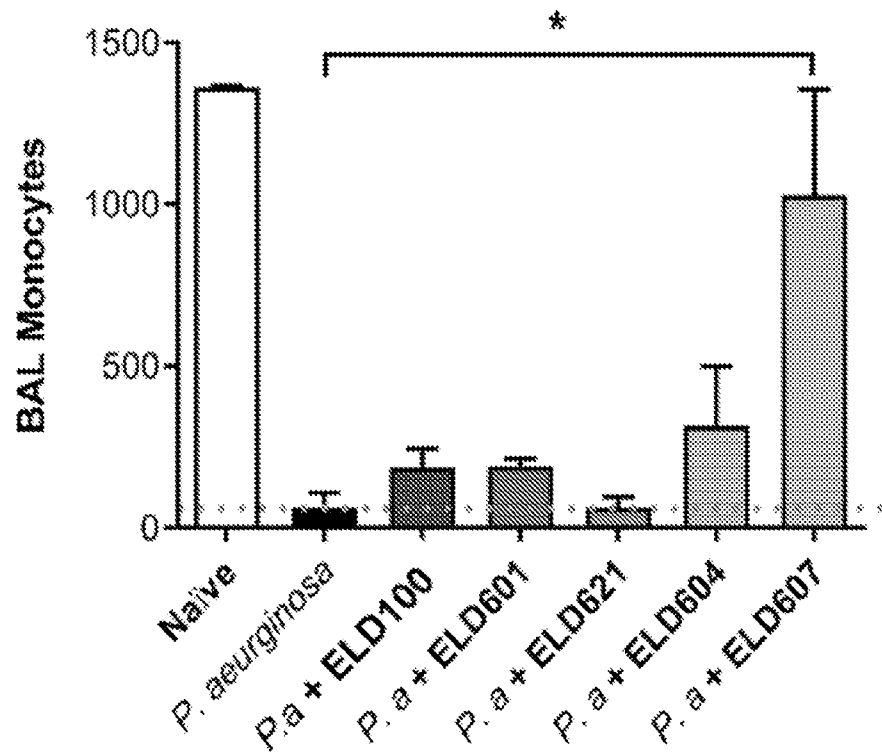


FIG. 15C

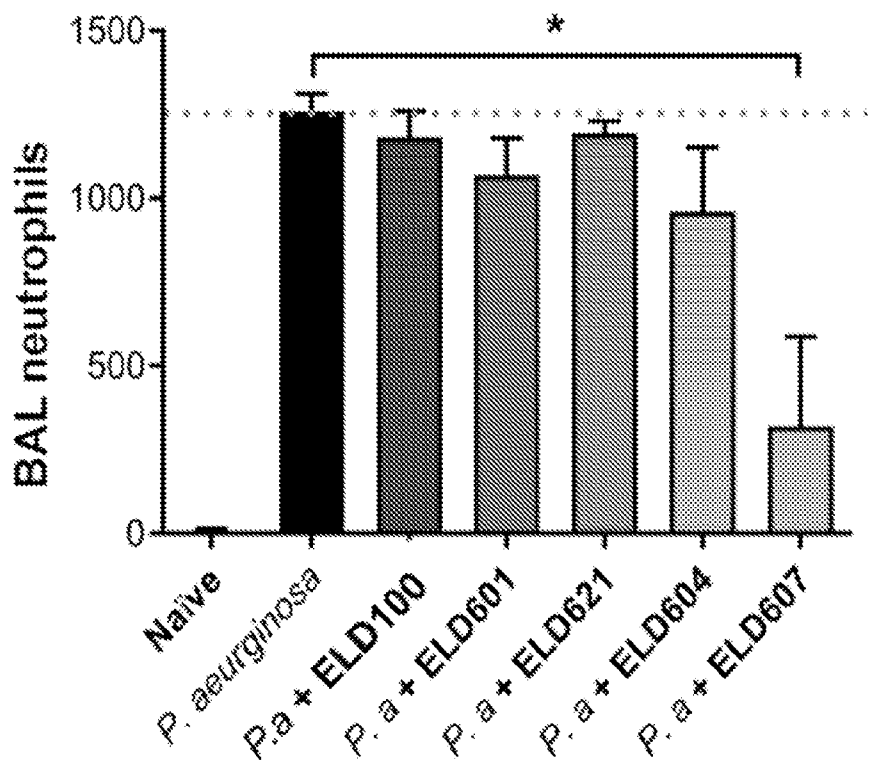


FIG. 15D

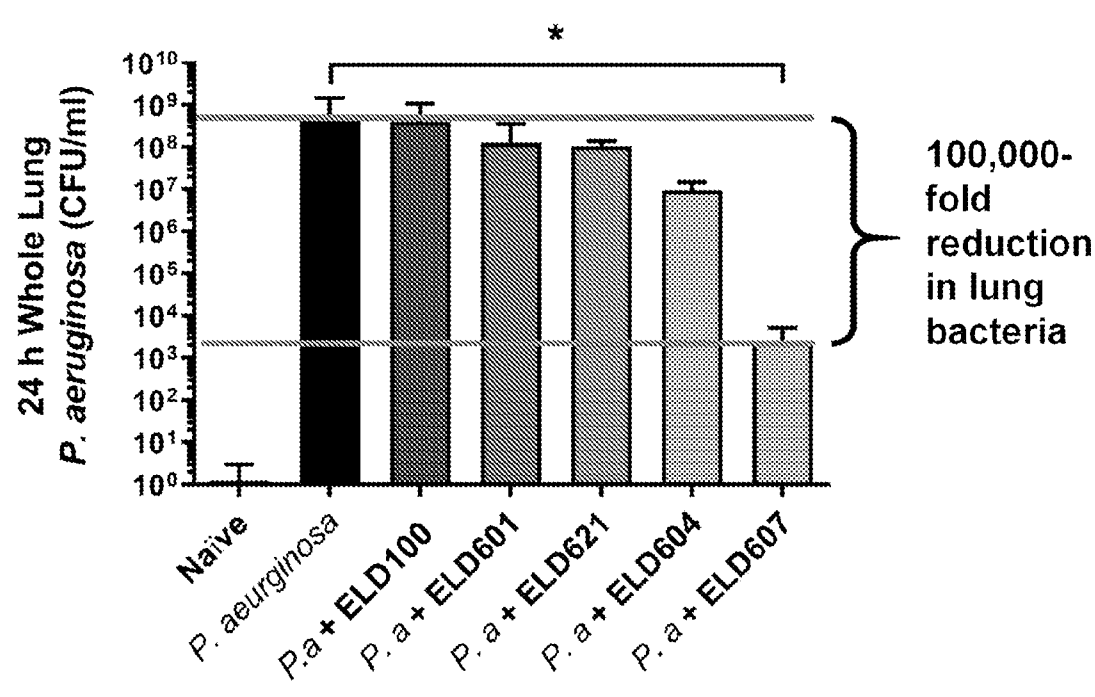


FIG. 16A

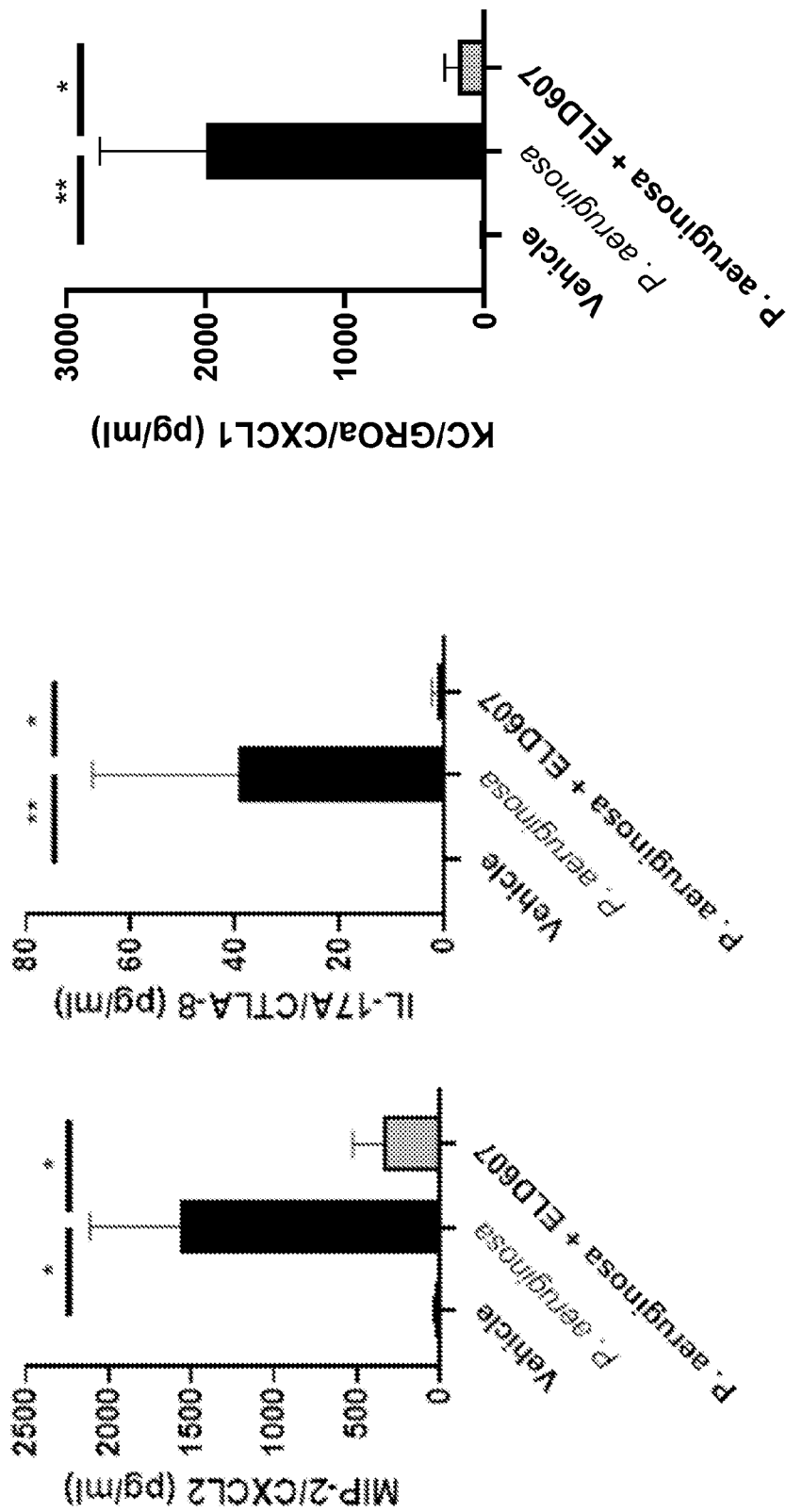


FIG. 16B

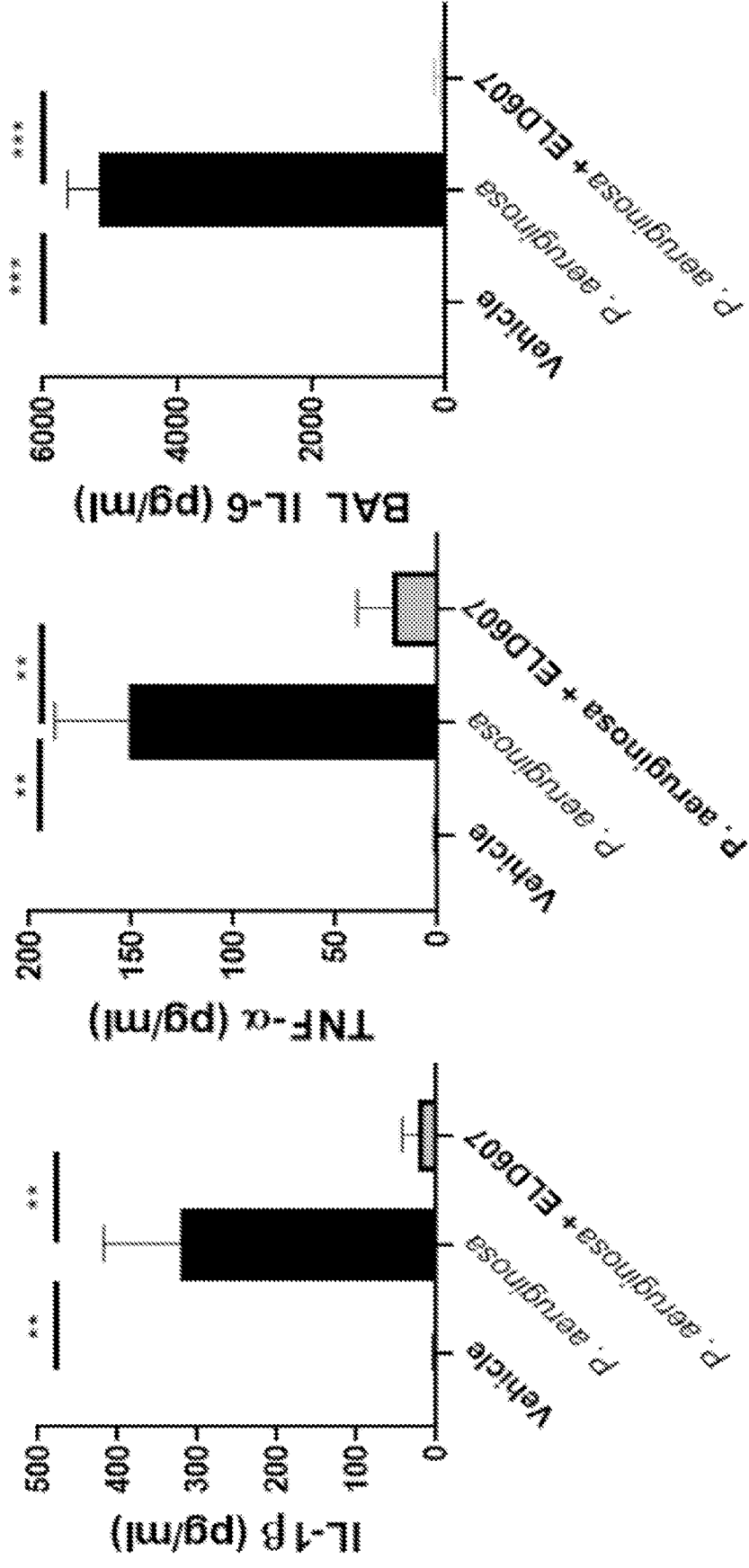
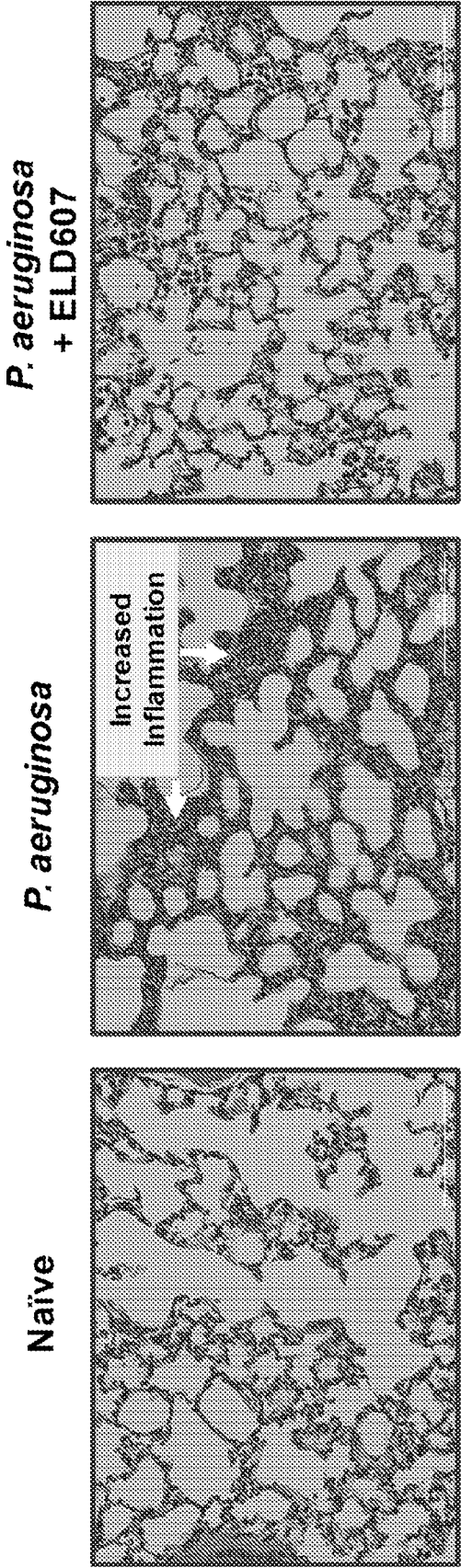


FIG. 16C



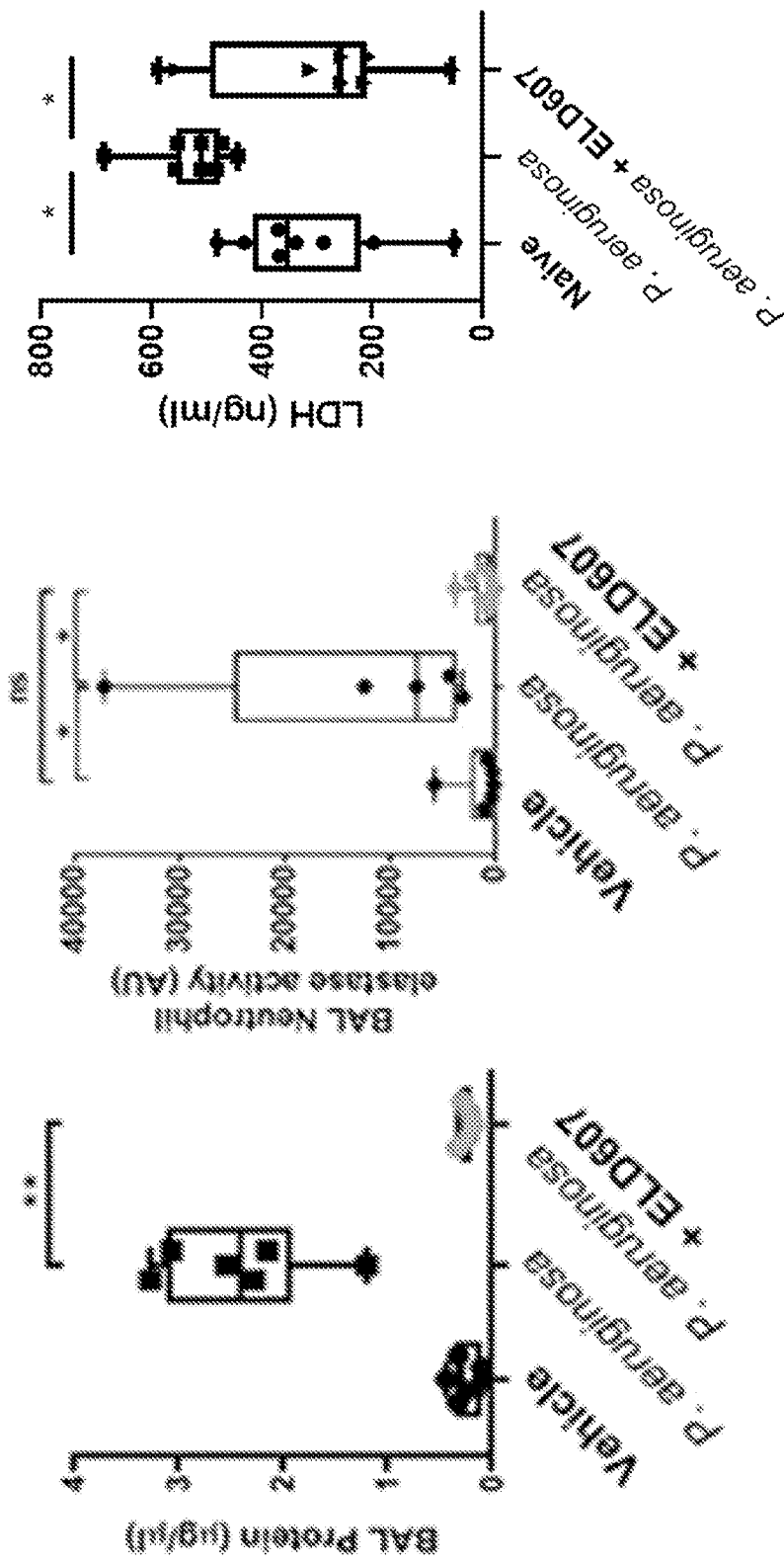


FIG. 16D

FIG. 16E

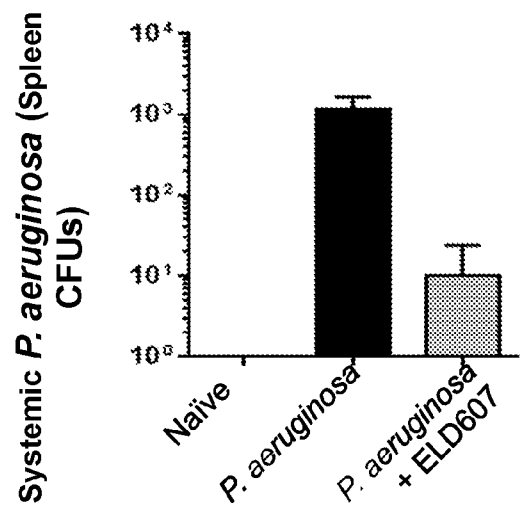


FIG. 16F

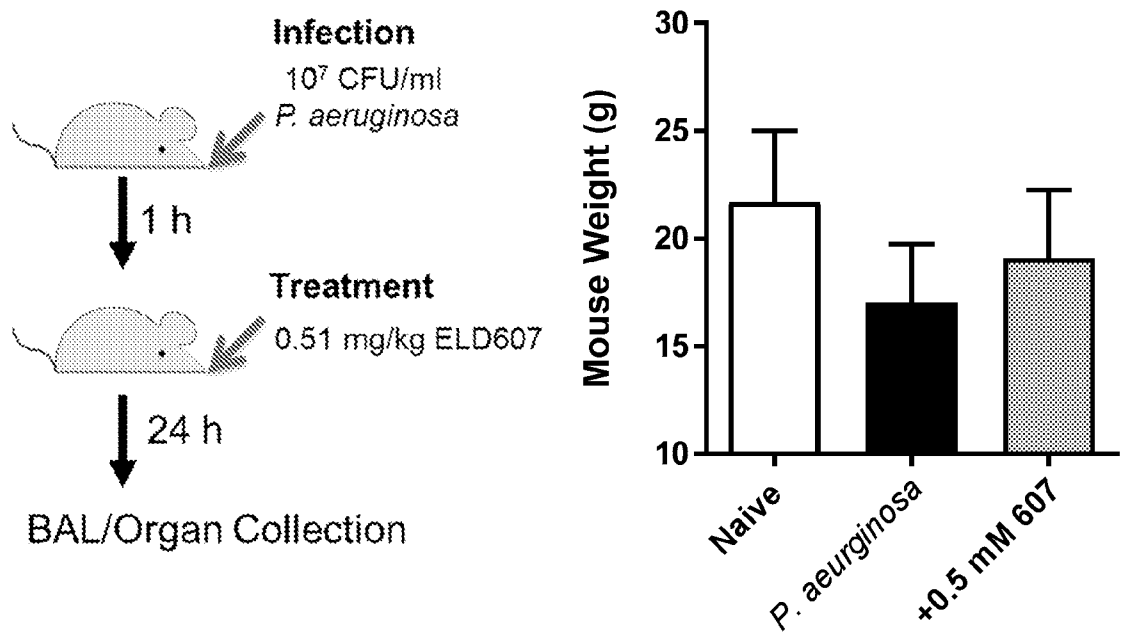
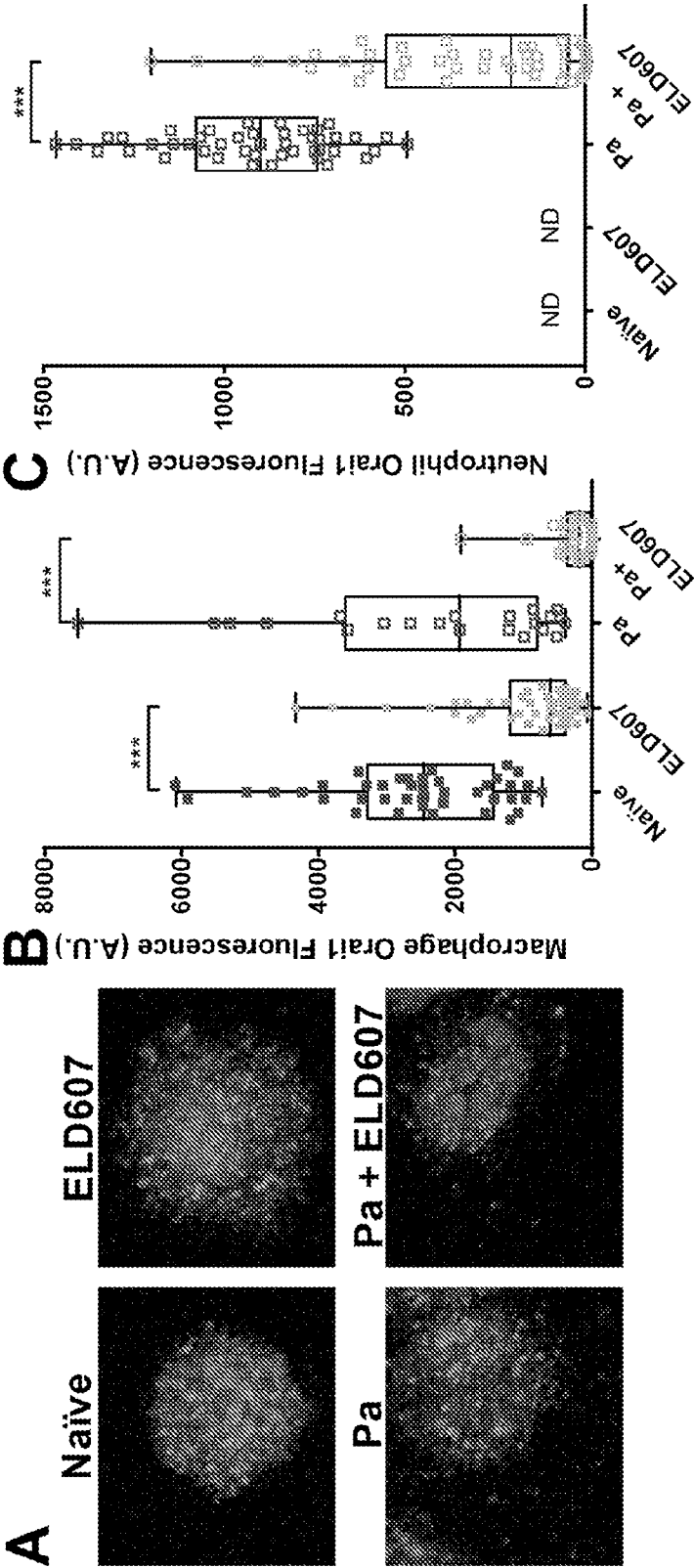


FIG. 16G



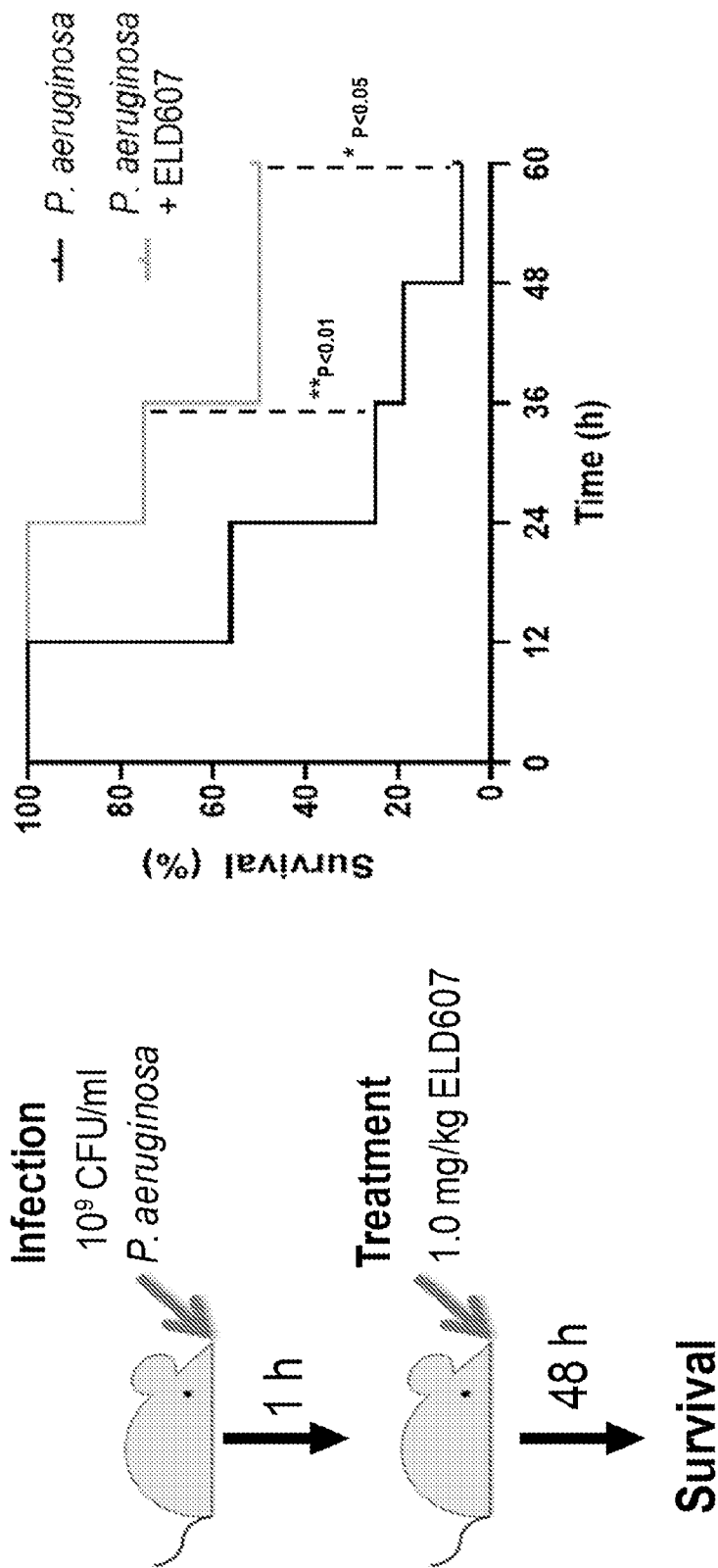


FIG. 17A

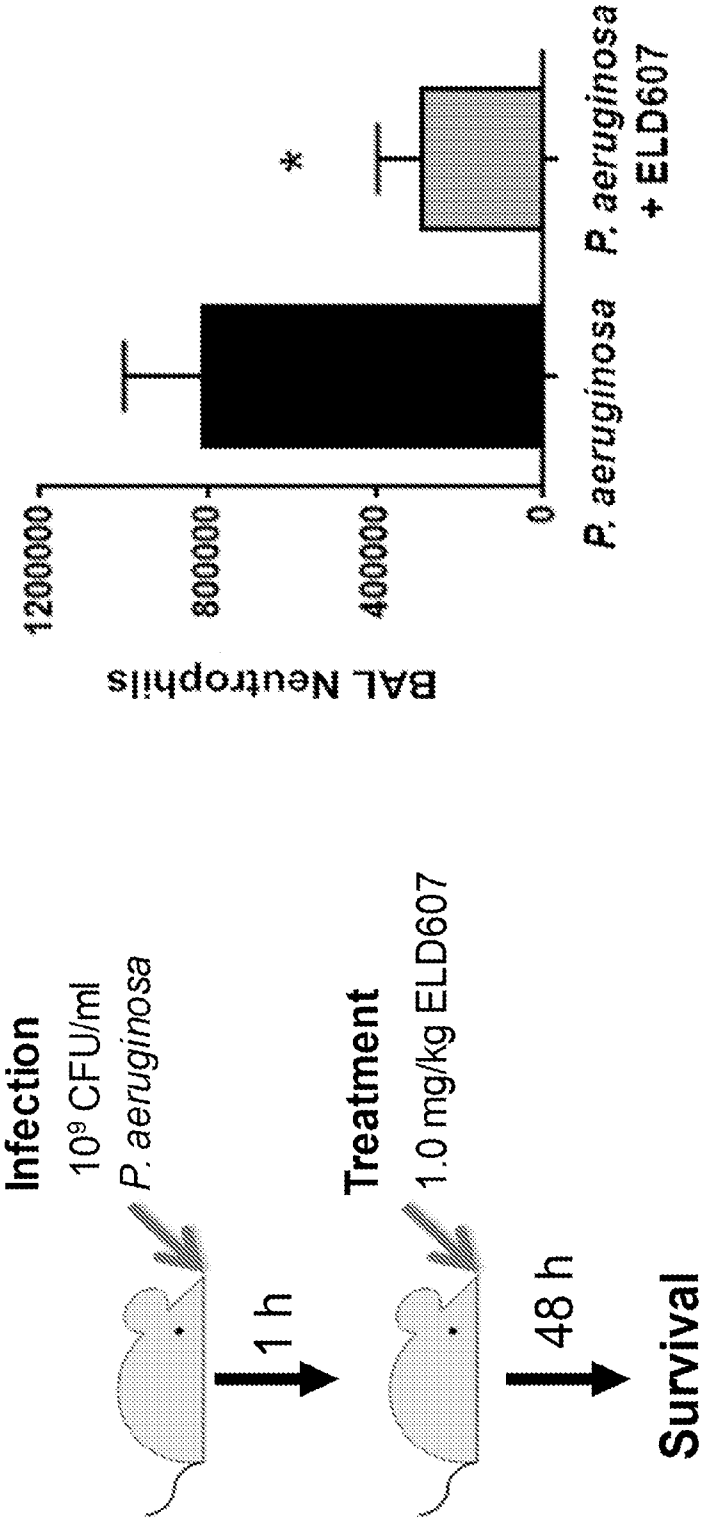


FIG. 17B

FIG. 18

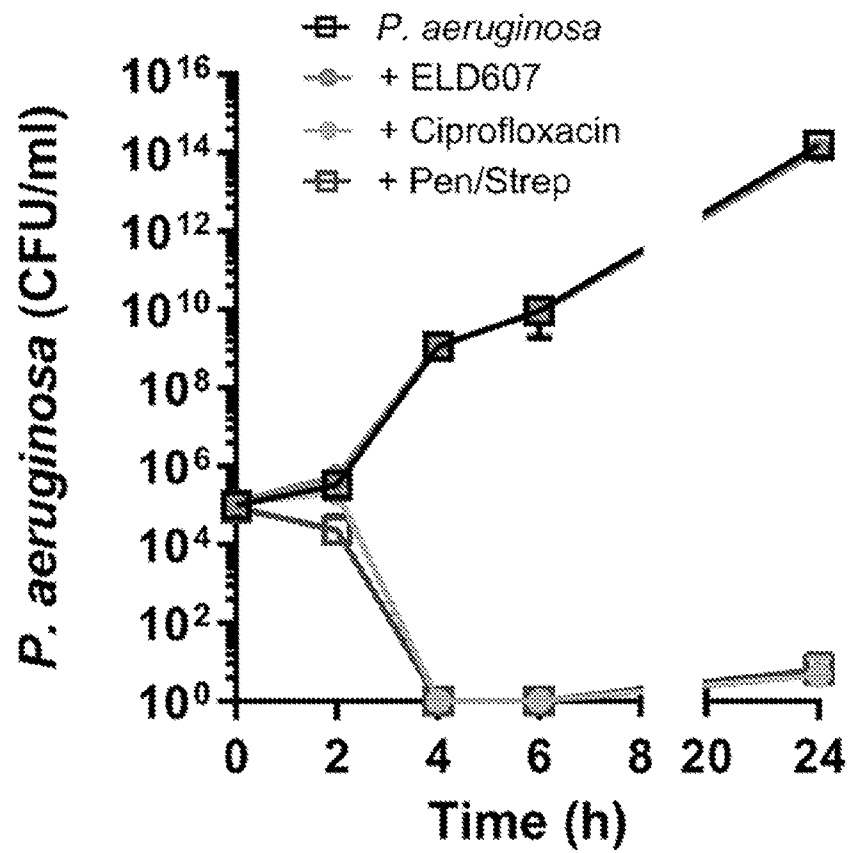


FIG. 19

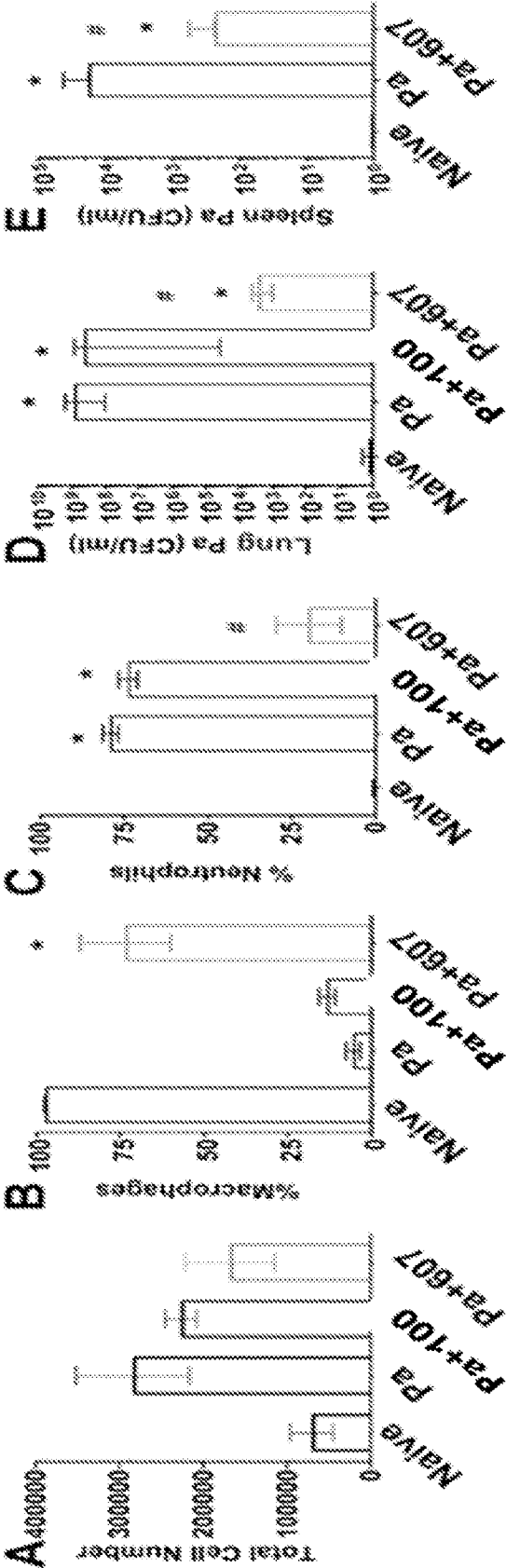


FIG. 19 cont.

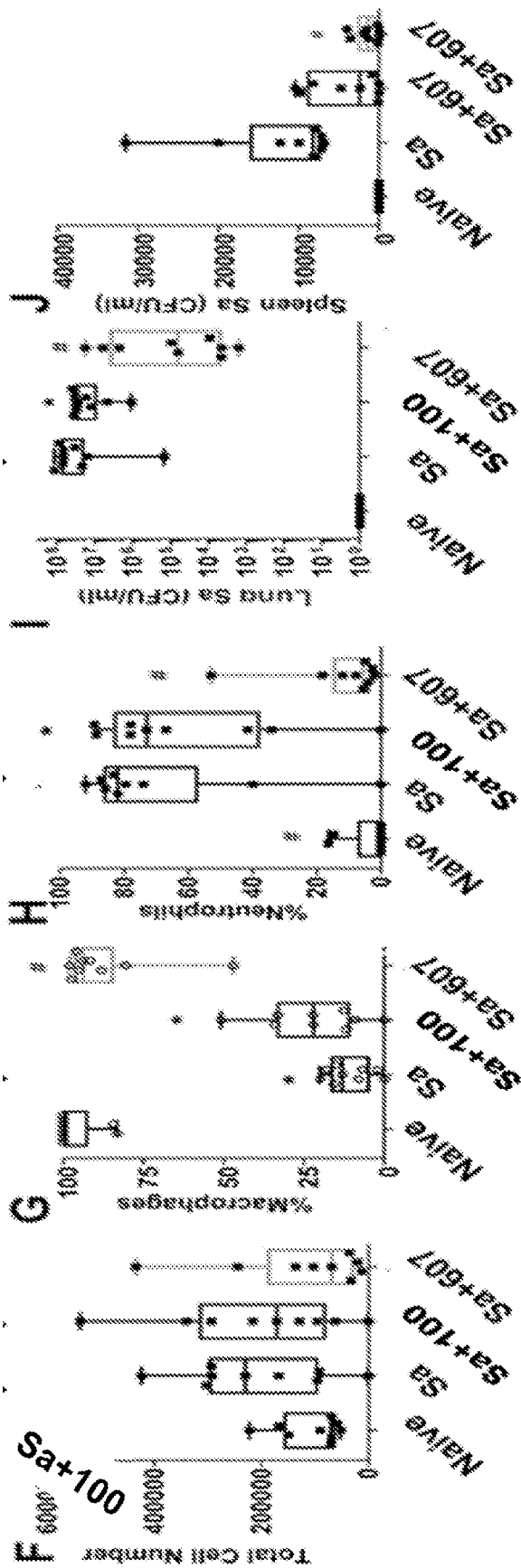


FIG. 19 cont.

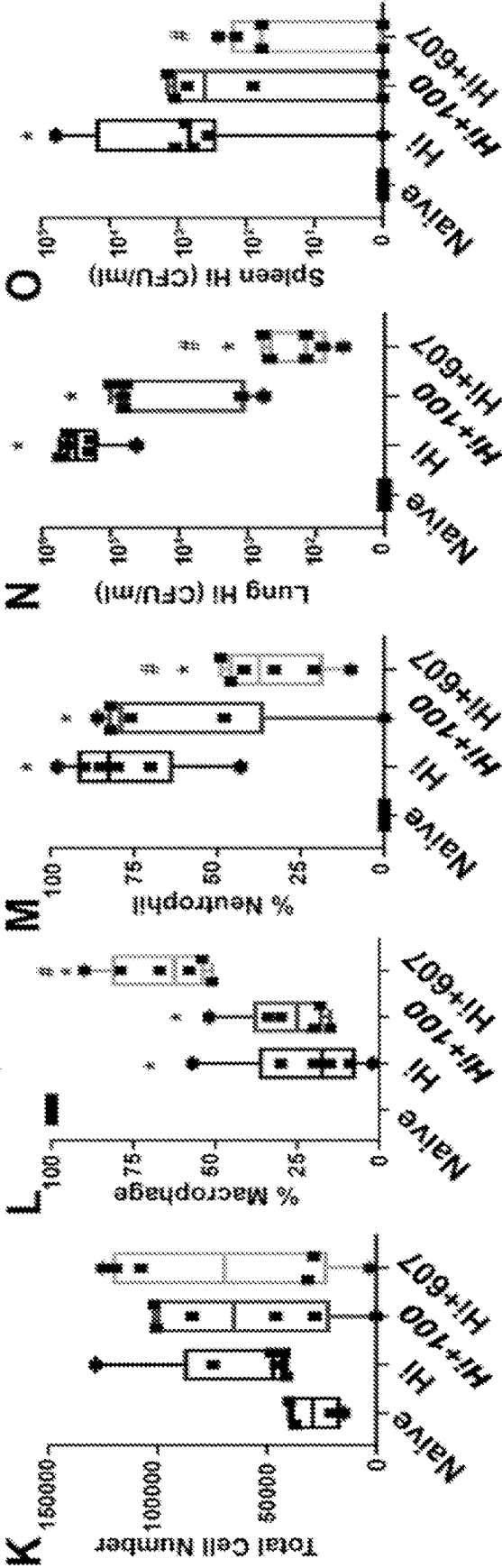
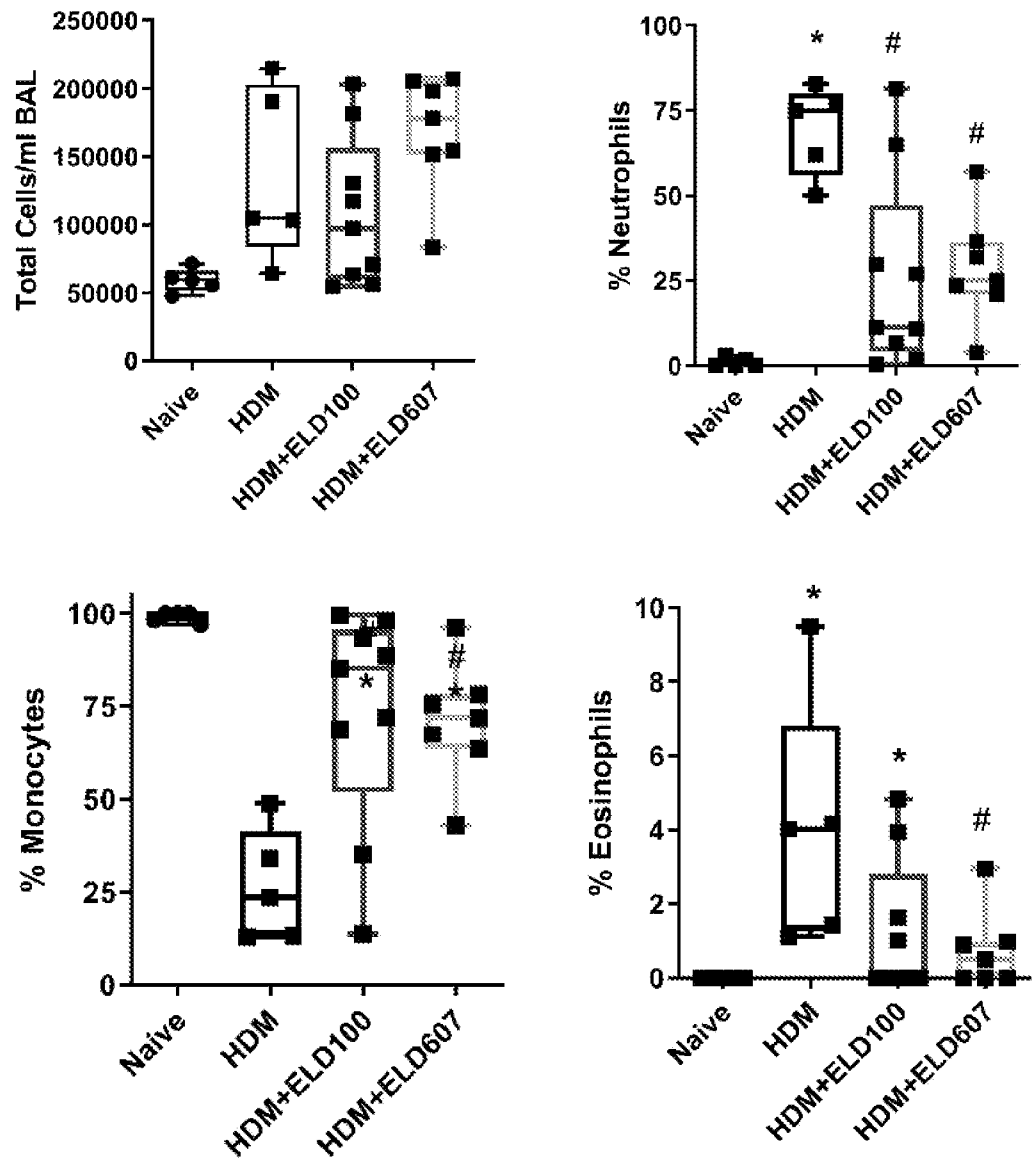


FIG. 20



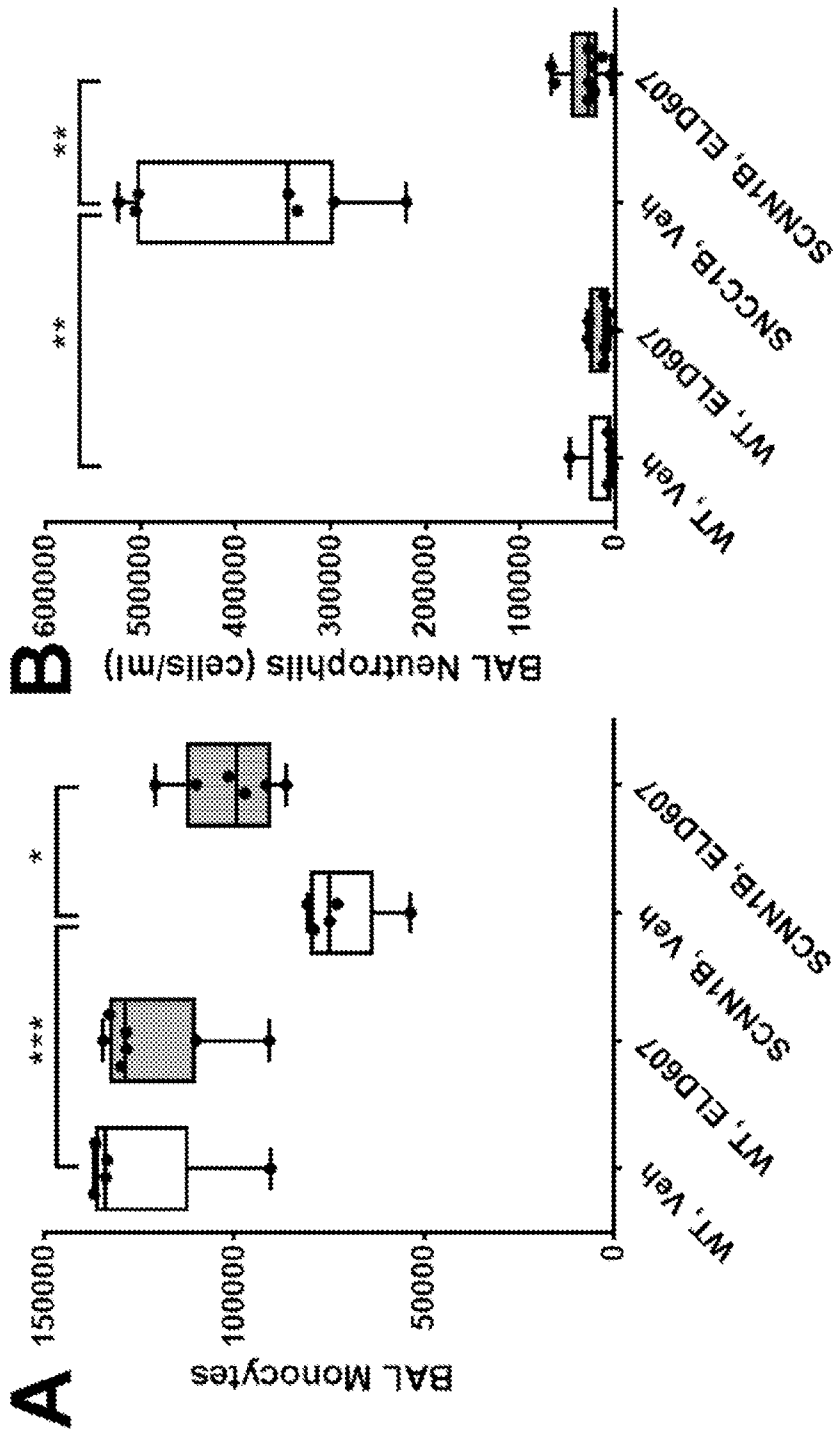


FIG. 21

FIG. 22

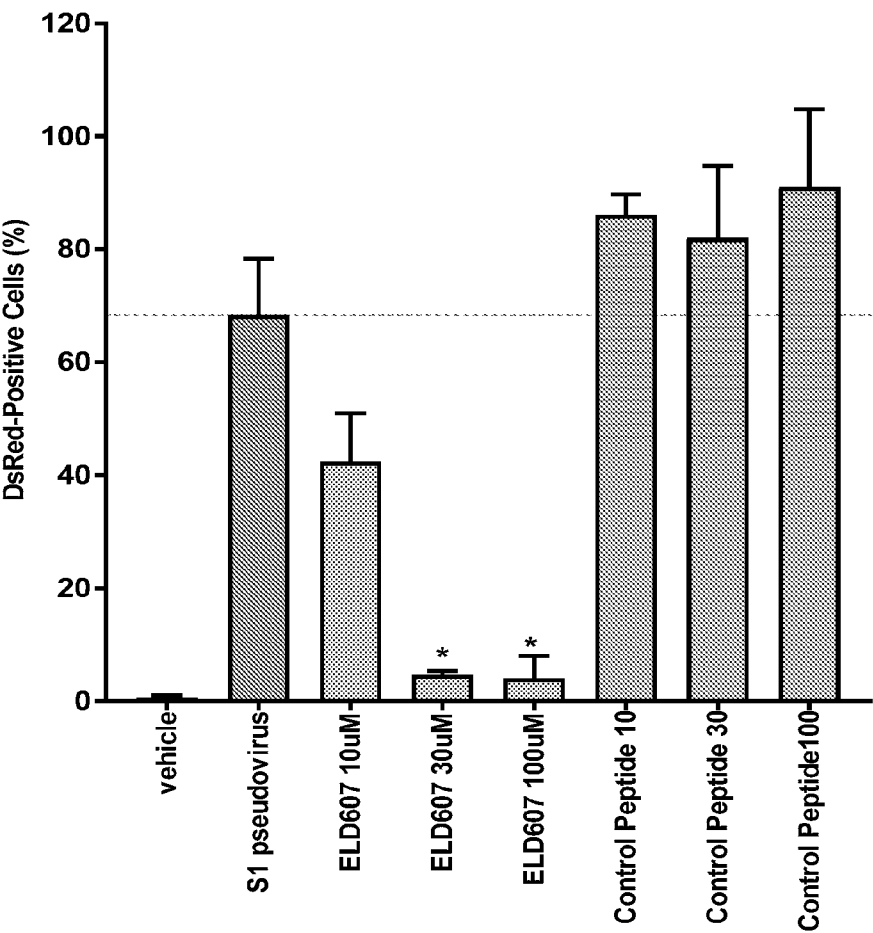


FIG. 23

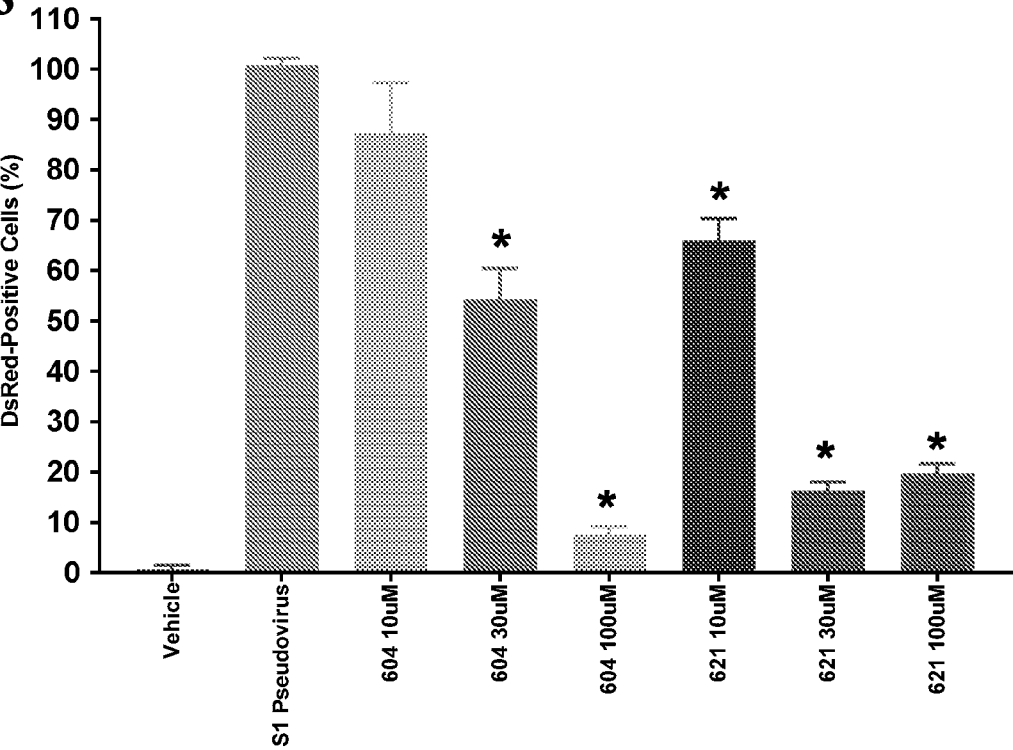


FIG. 24

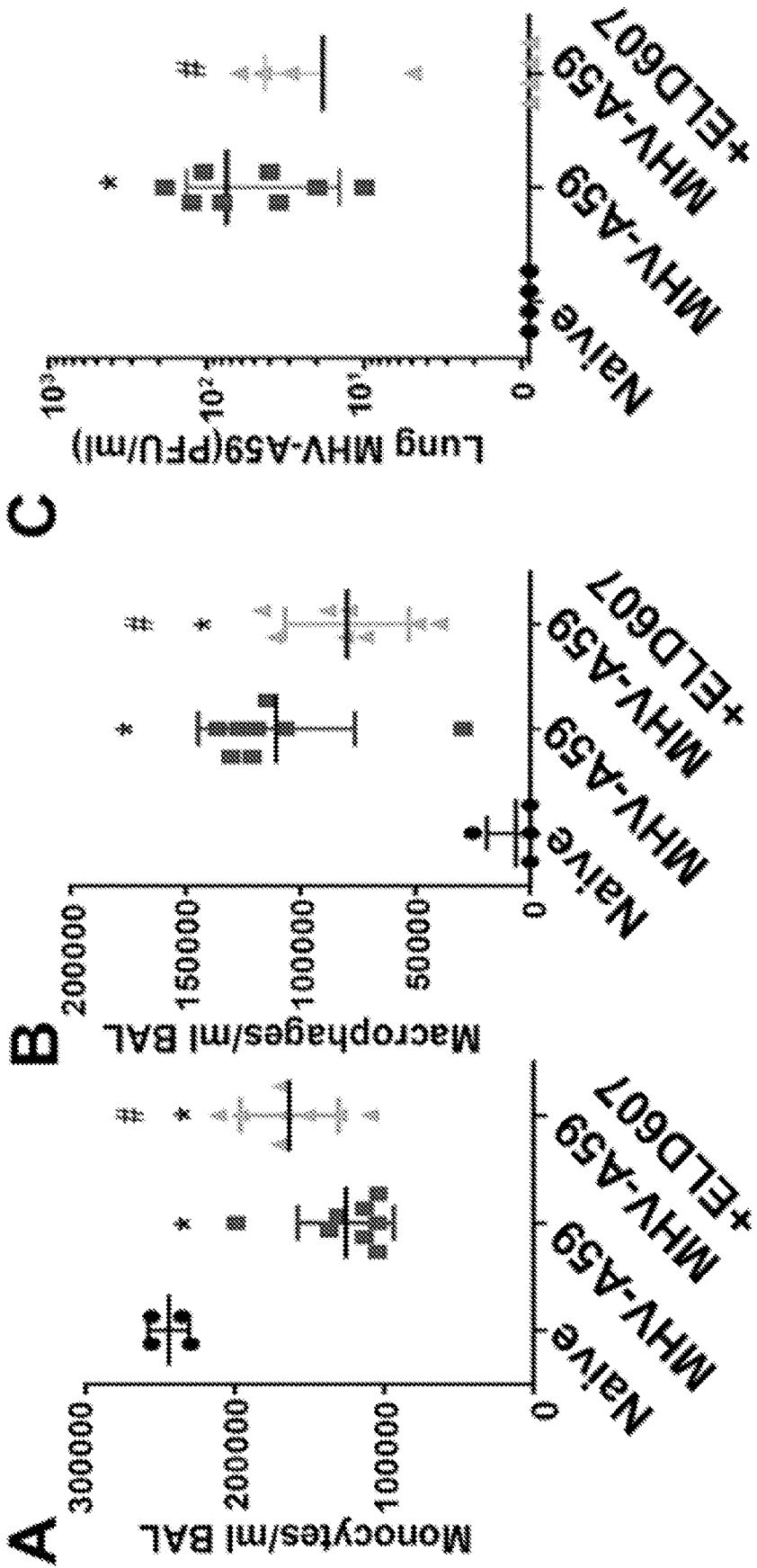
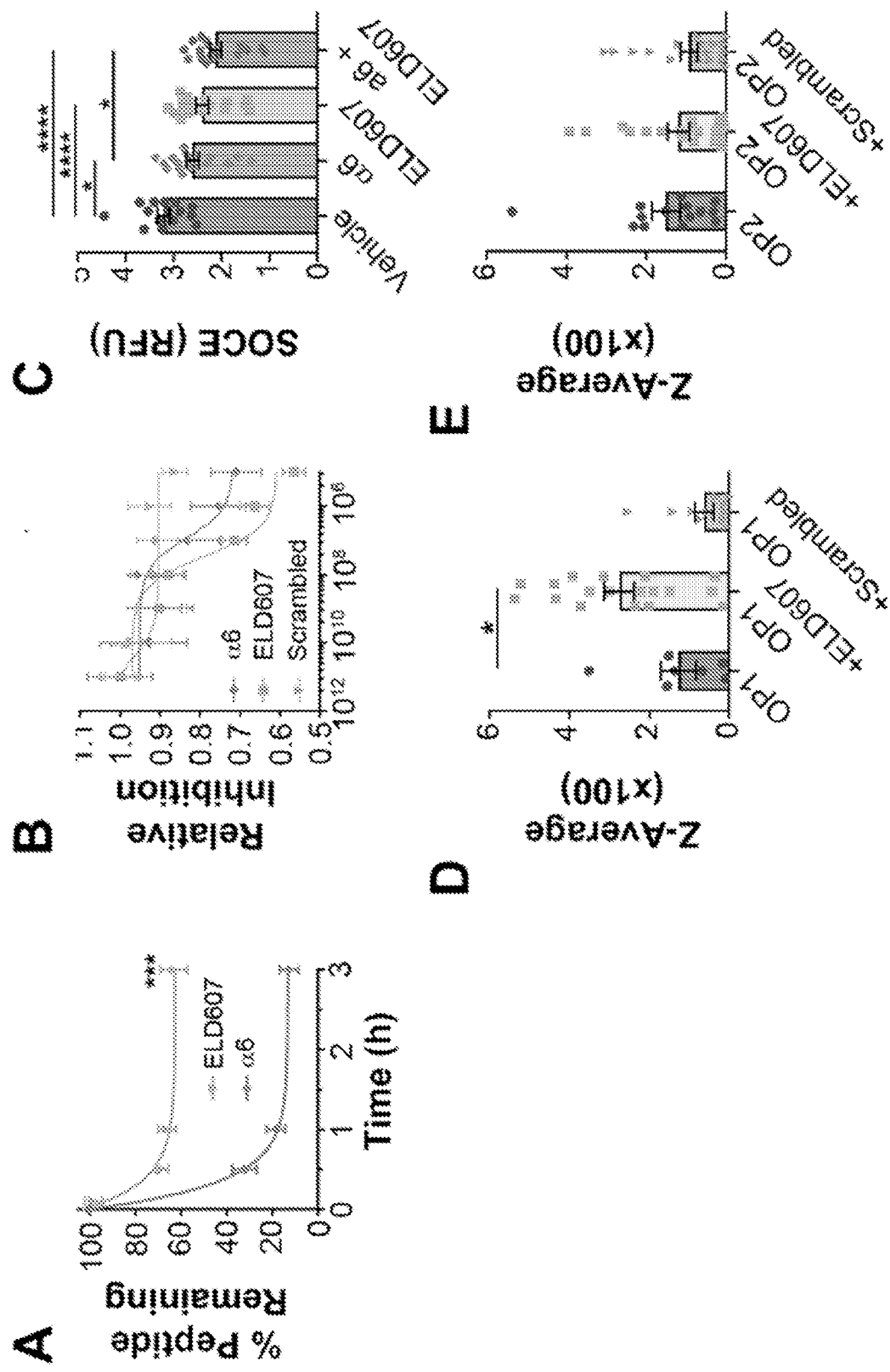


FIG. 25



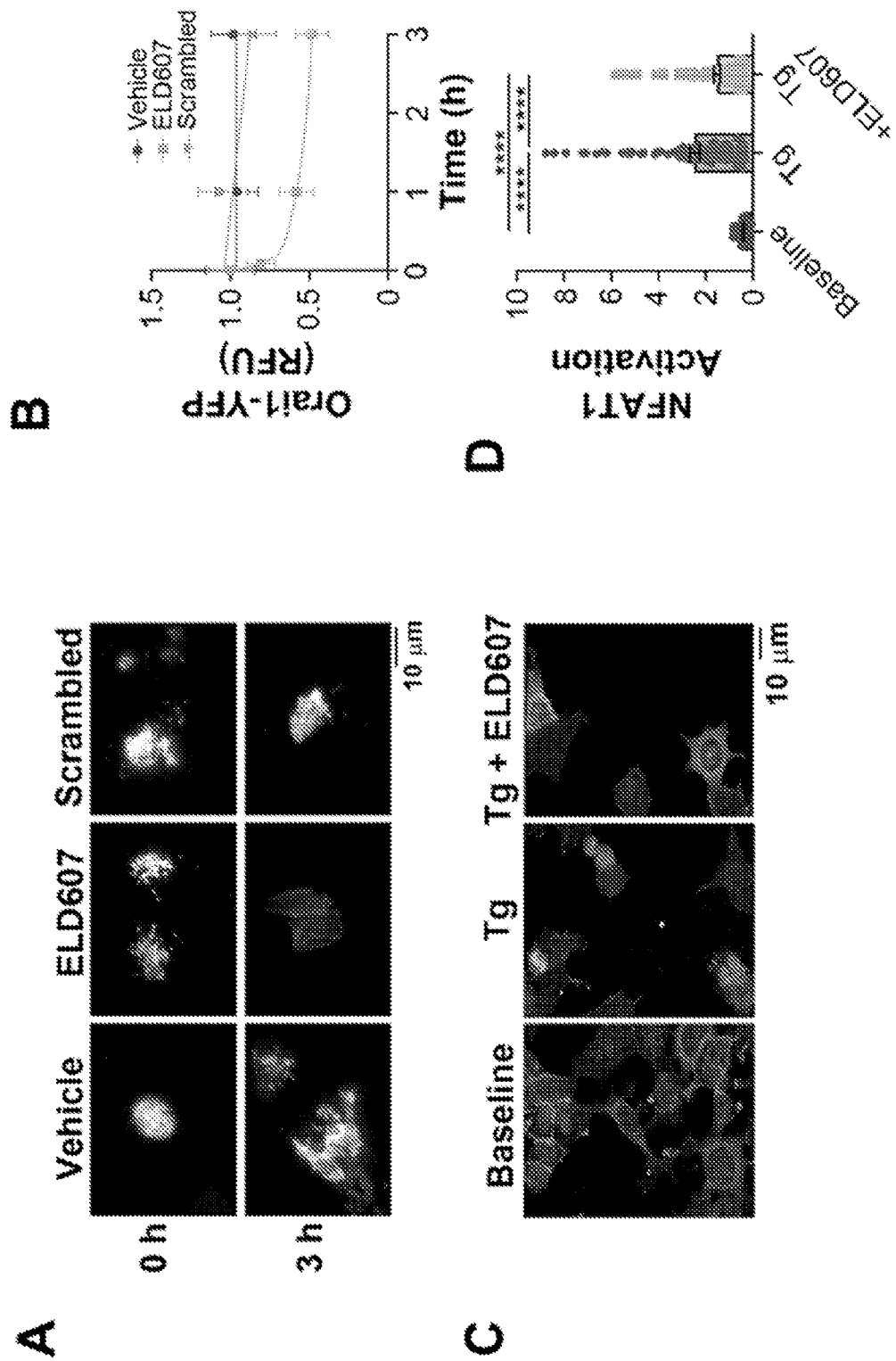


FIG. 27

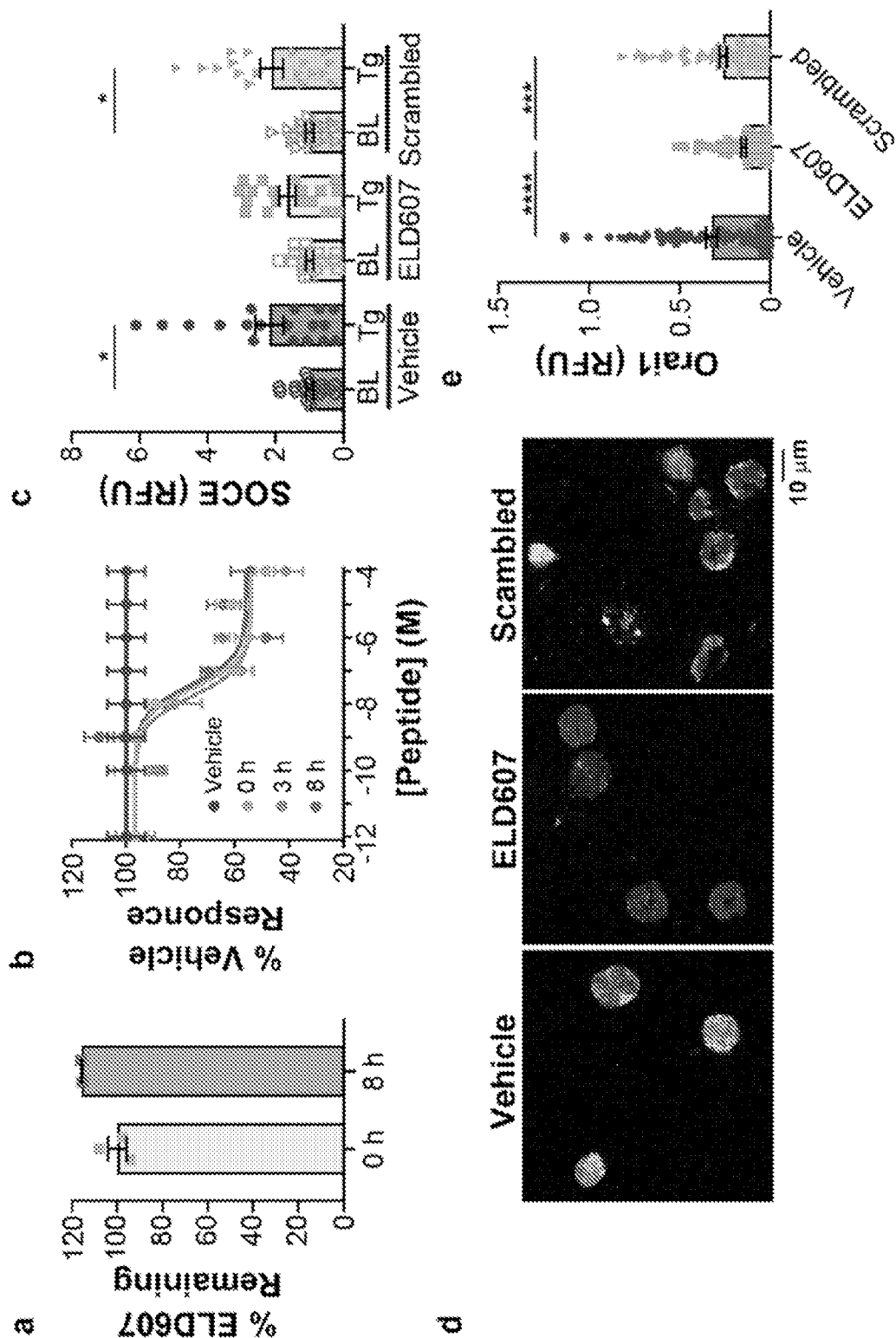


FIG. 28

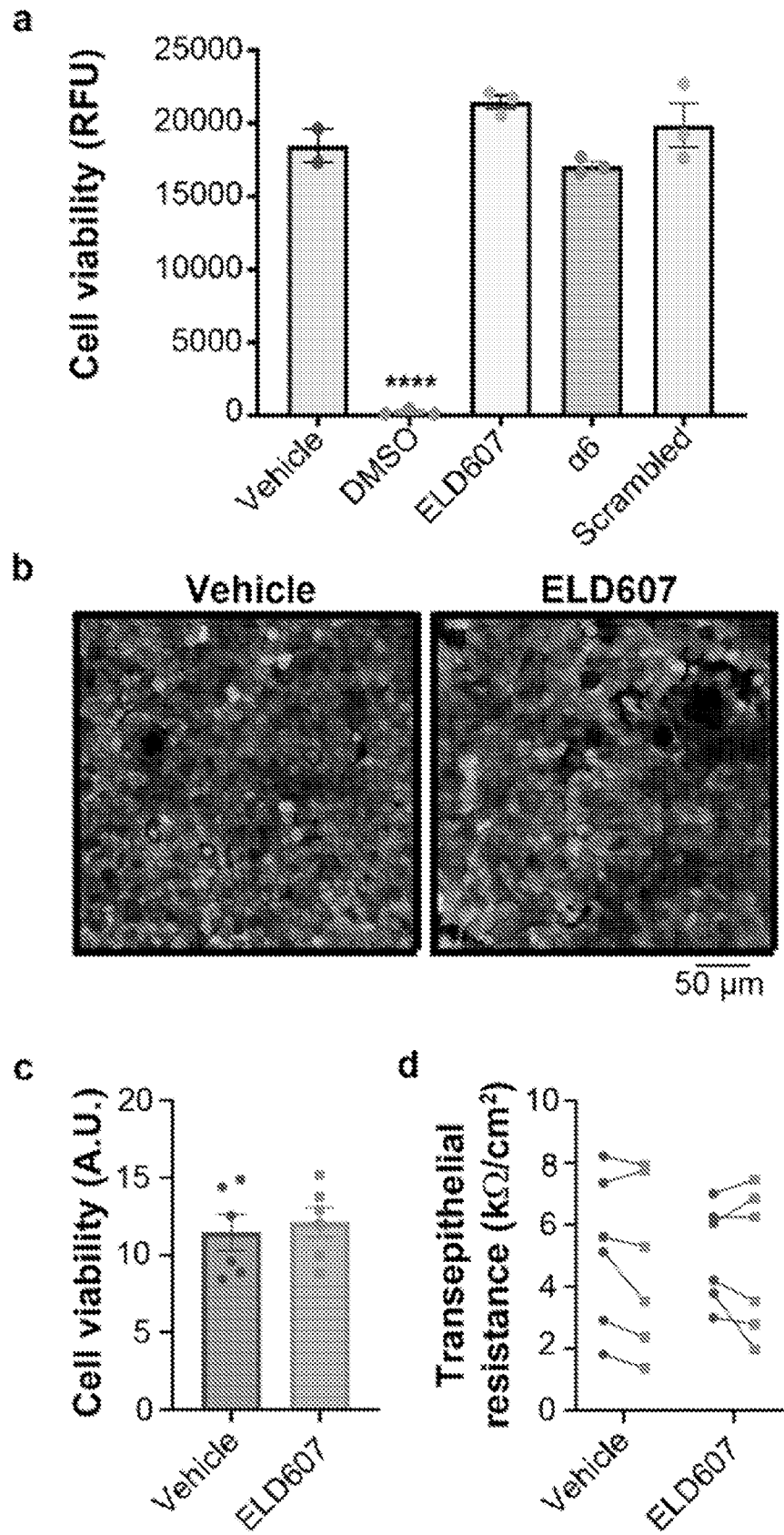


FIG. 29

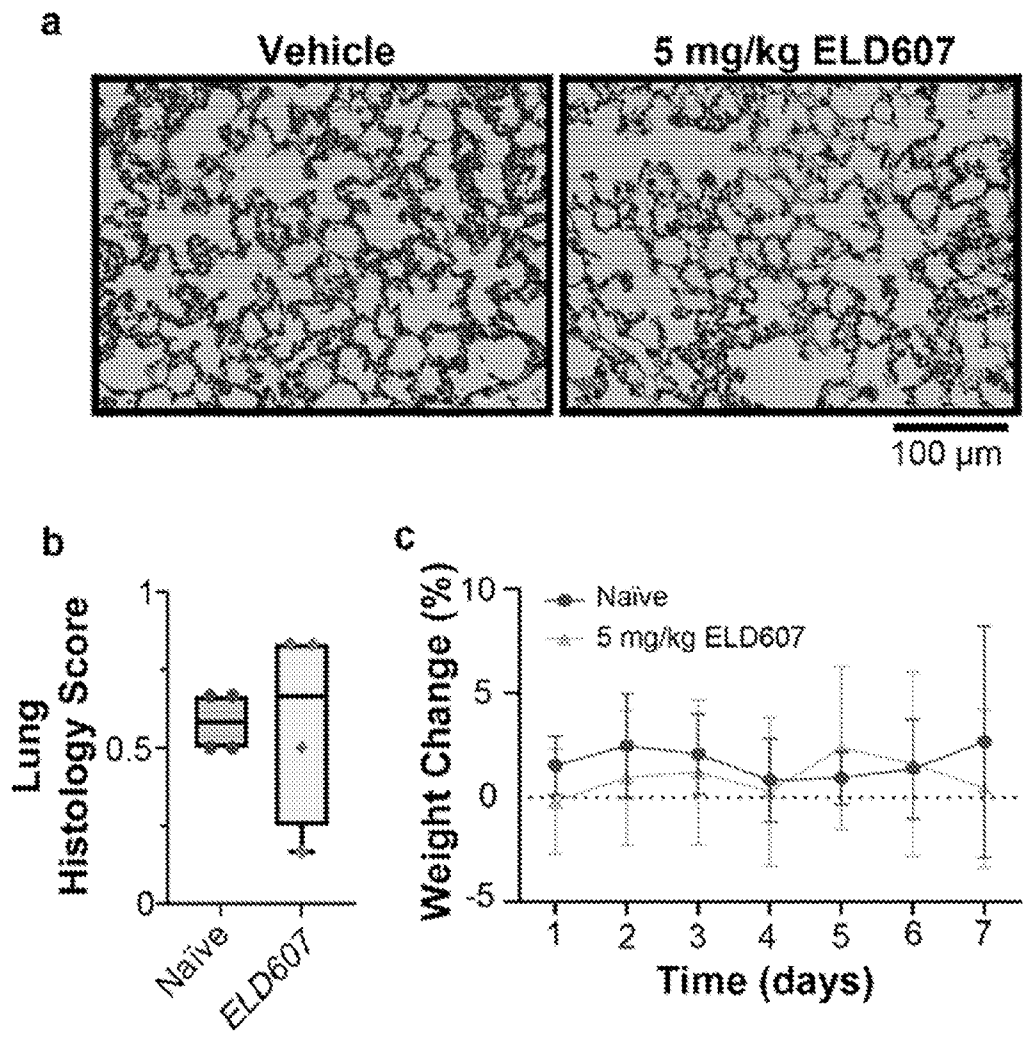


FIG. 30

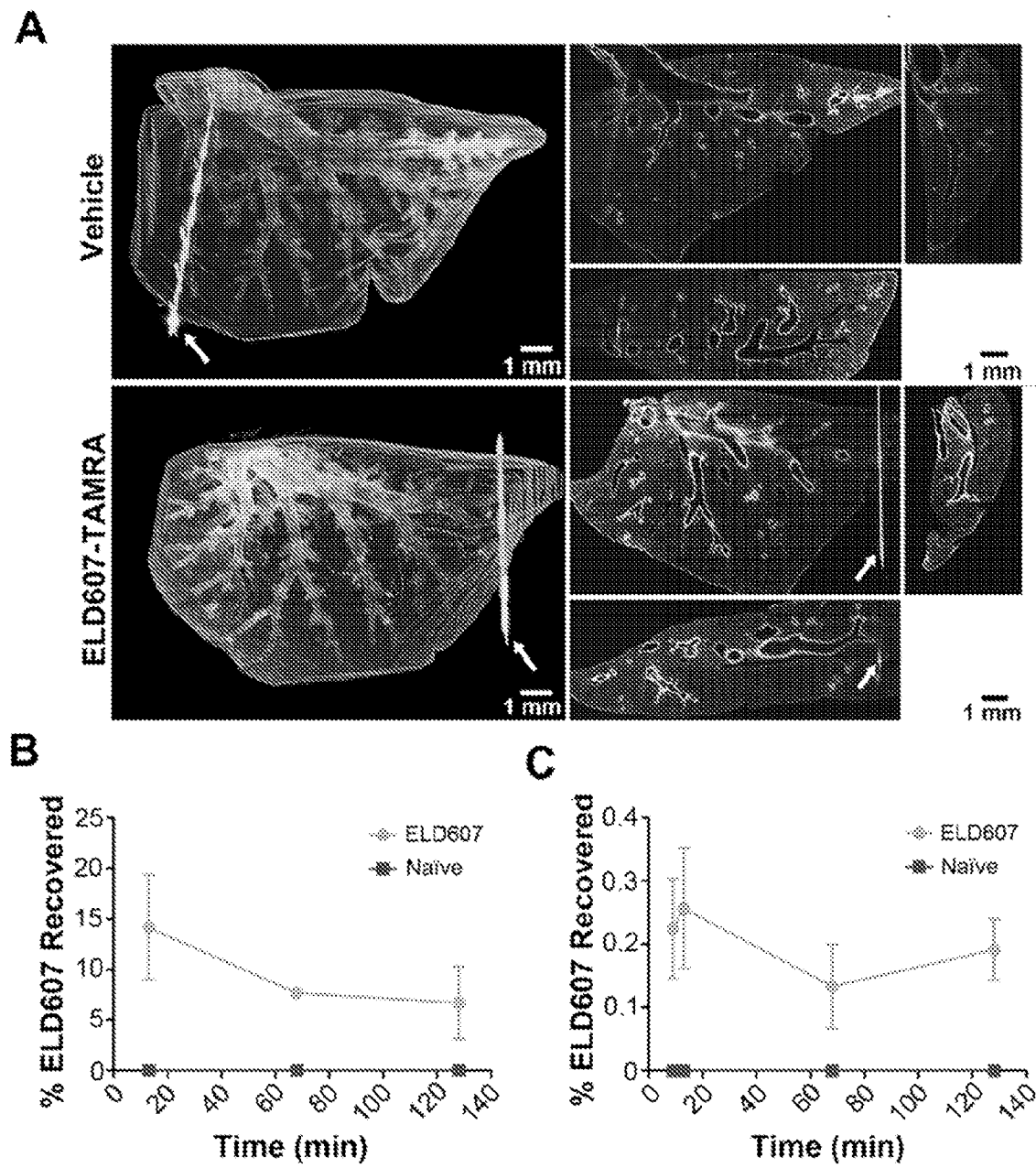


FIG. 31

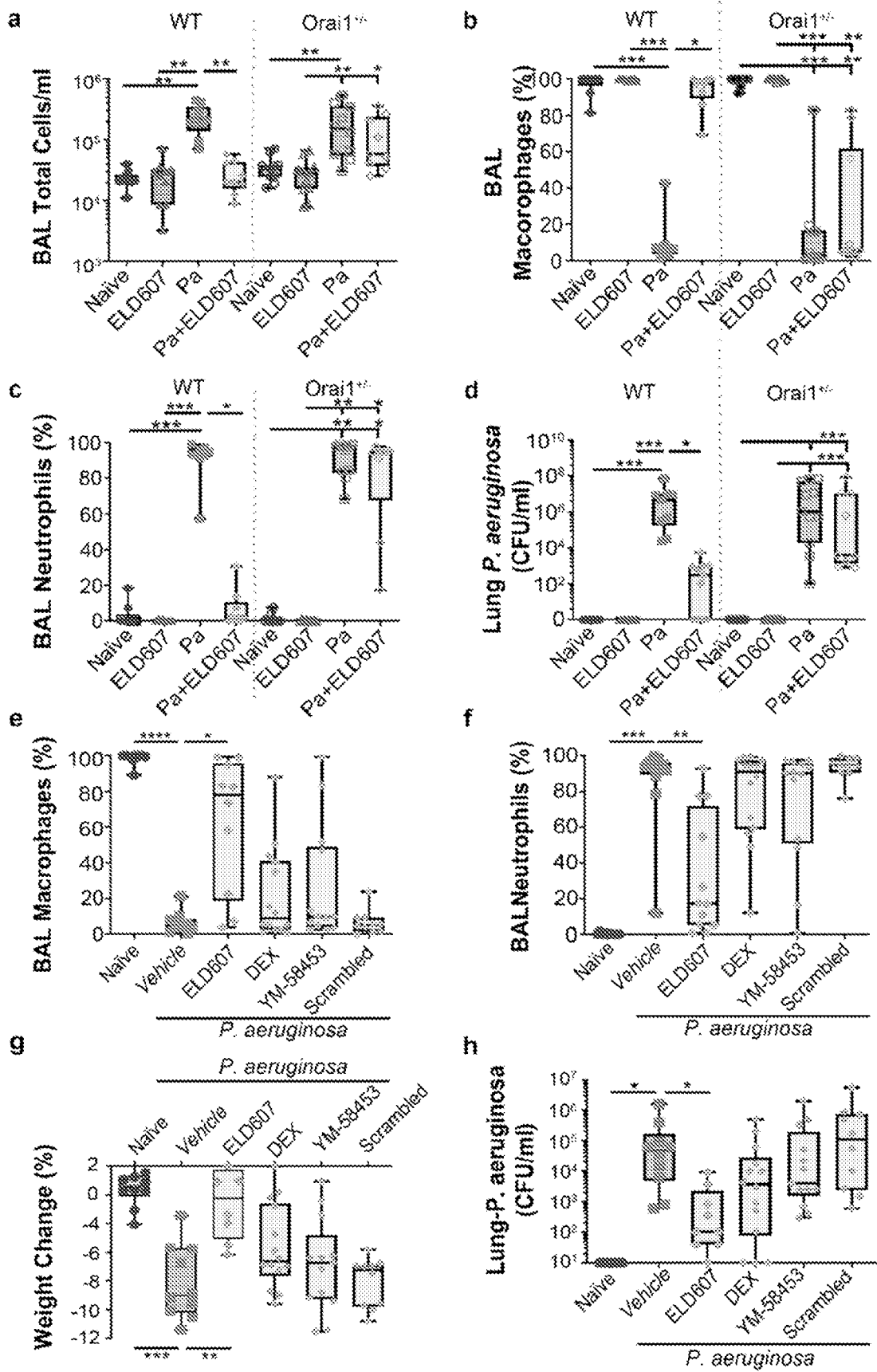


FIG. 32

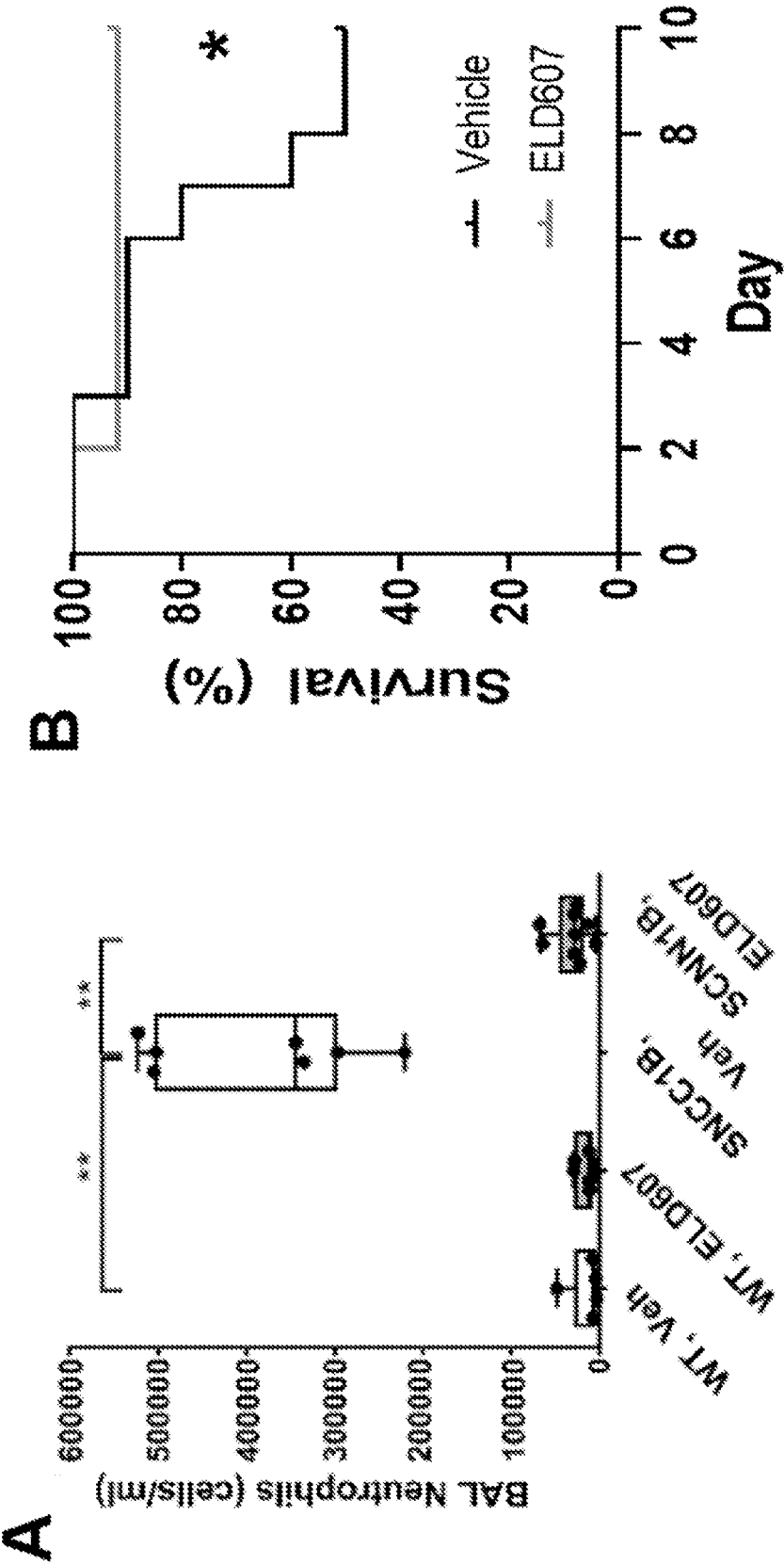


FIG. 33

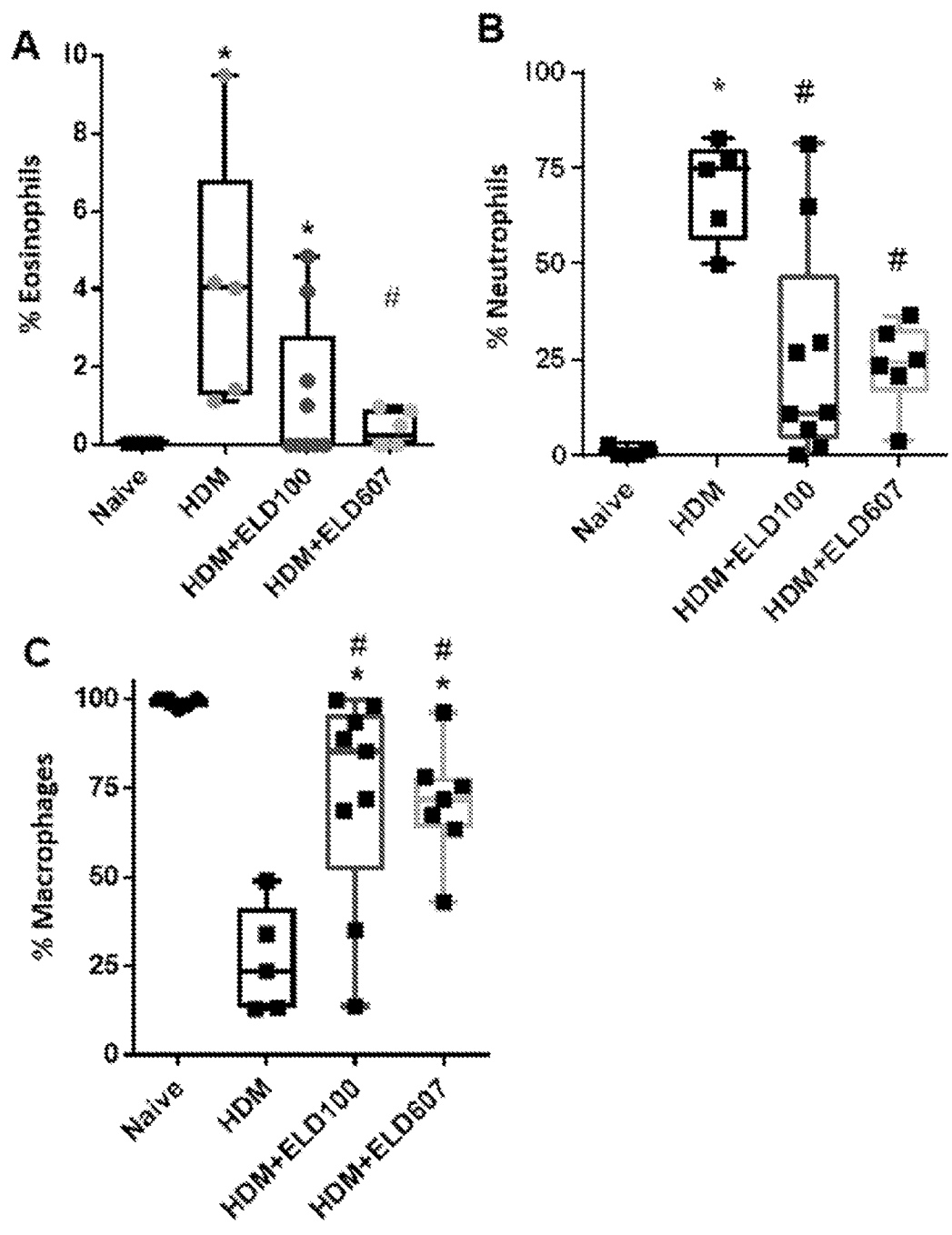
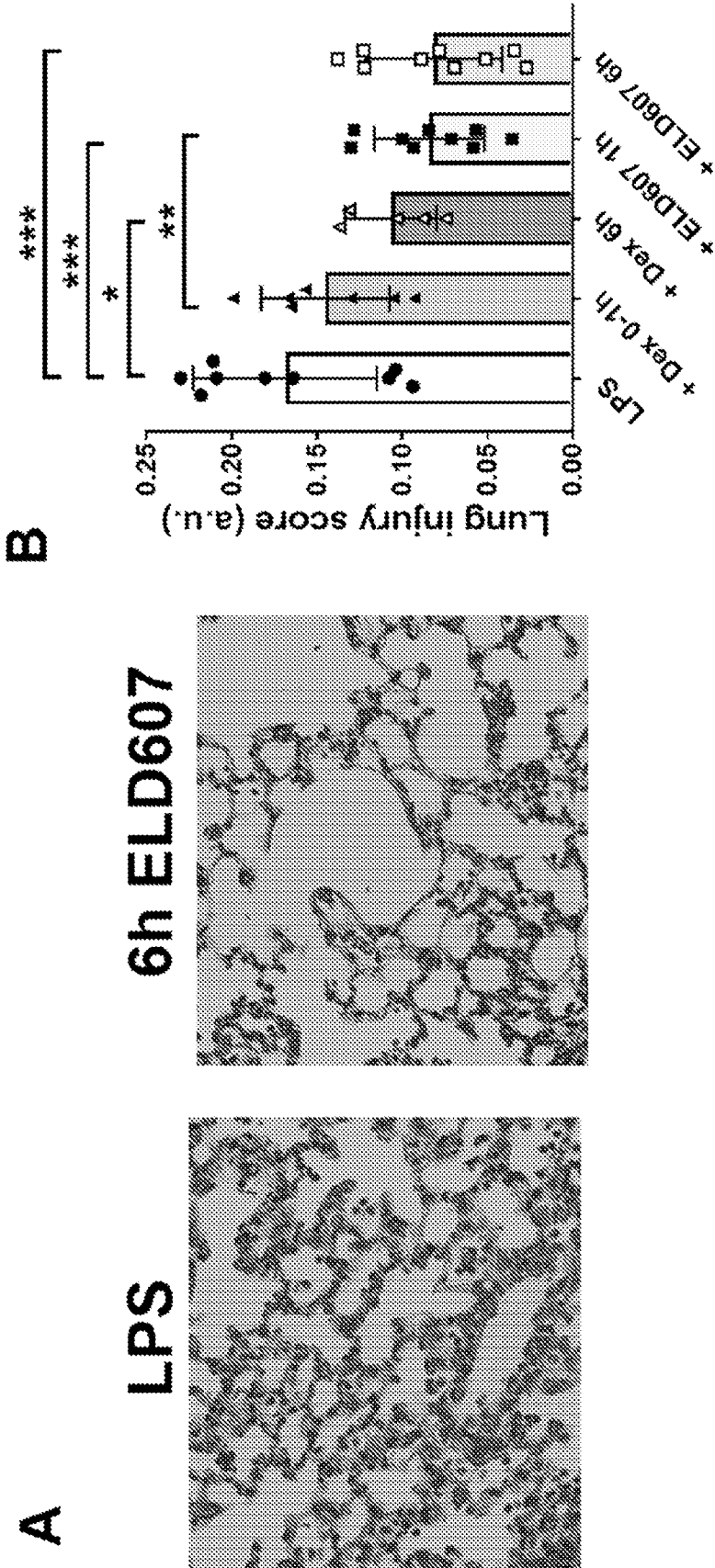


FIG. 34



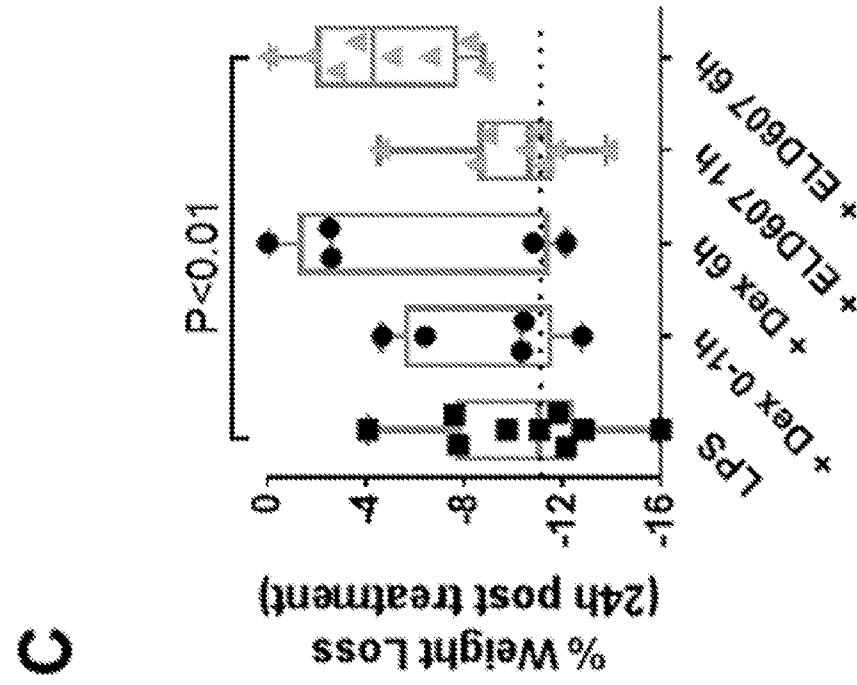
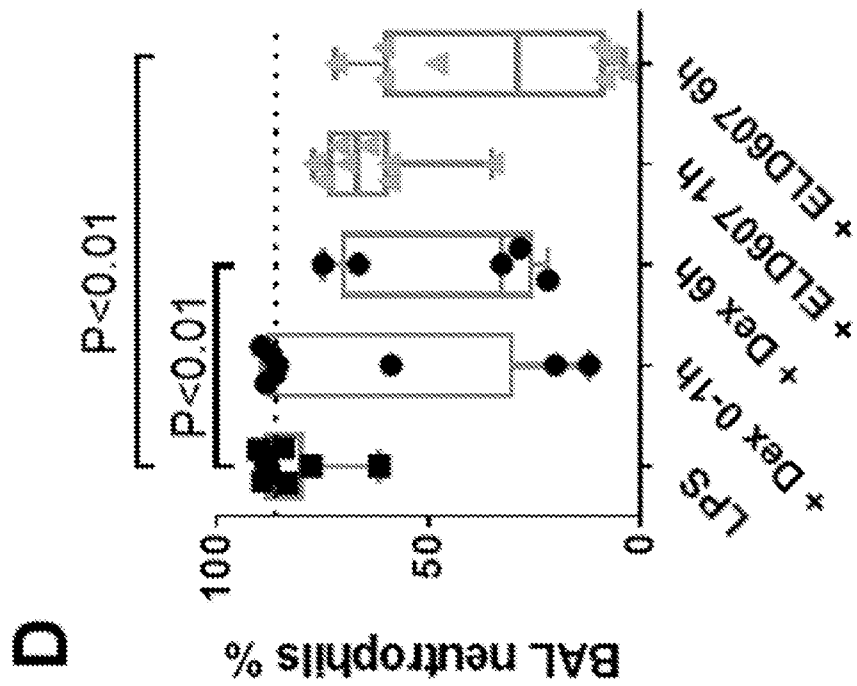
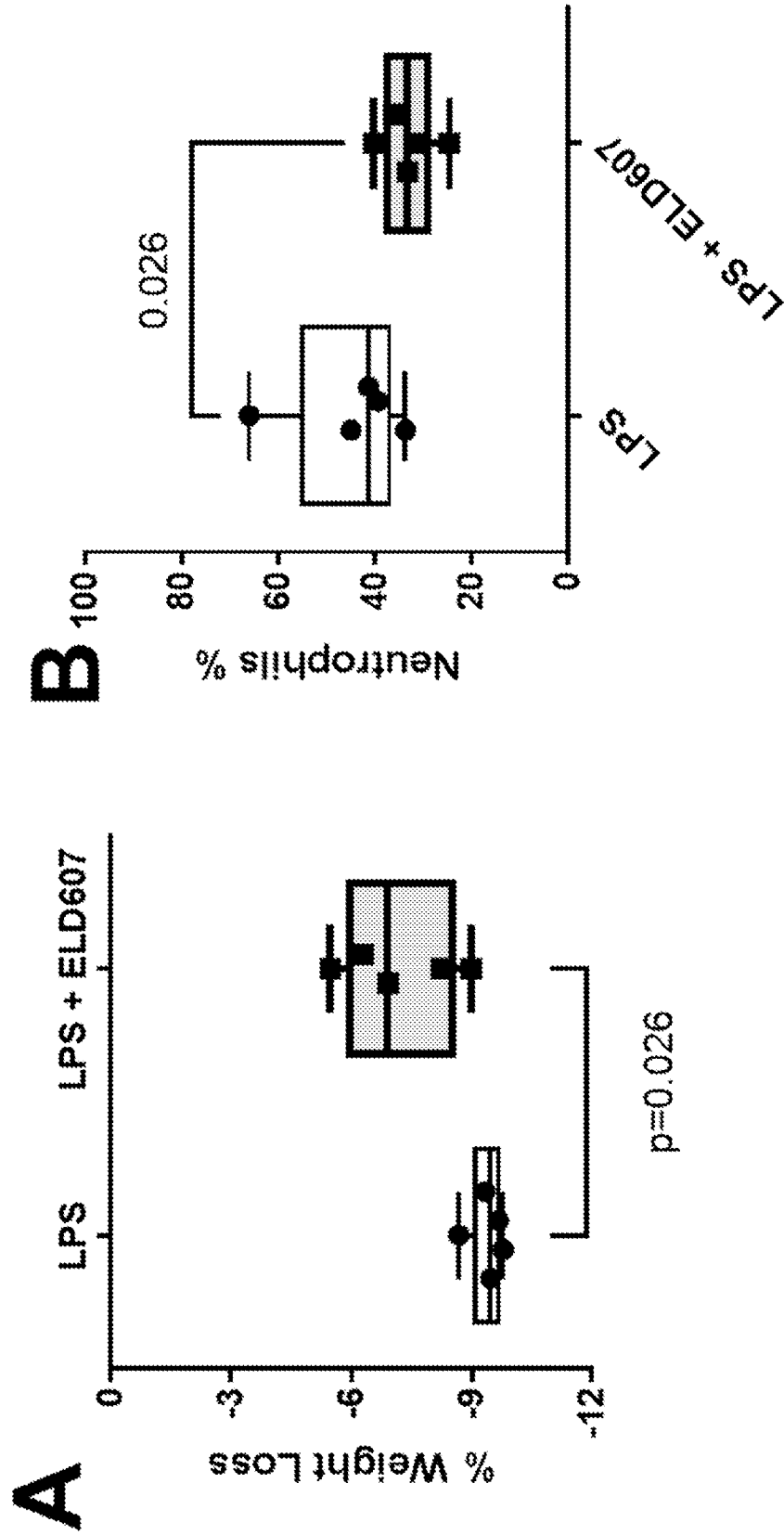


FIG. 34 (cont.)

FIG. 35



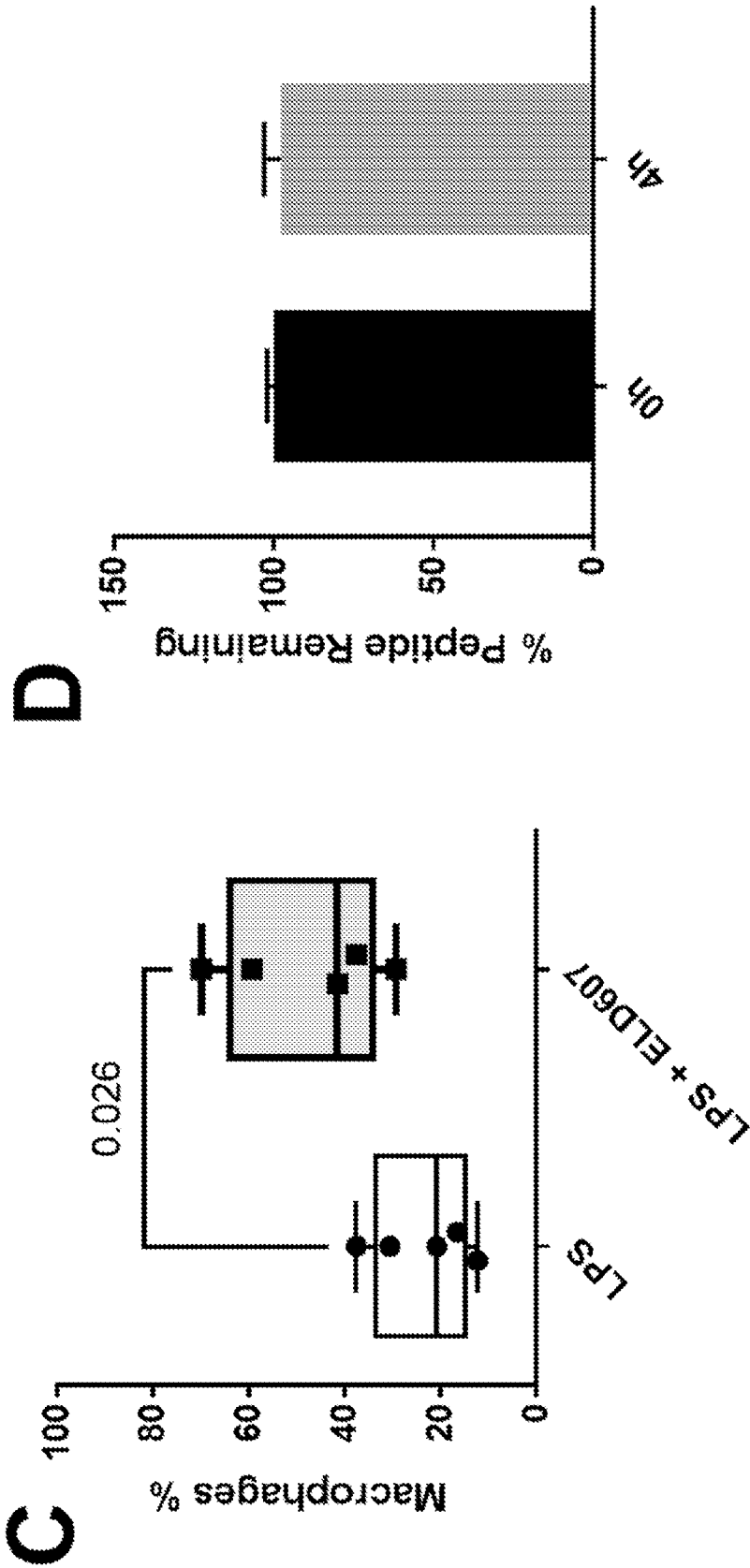


FIG. 35 (cont.)

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2022/021391

A. CLASSIFICATION OF SUBJECT MATTER C07K 7/06(2006.01)i; C07K 7/08(2006.01)i; A61K 38/00(2006.01)i; A61P 11/06(2006.01)i; A61P 31/04(2006.01)i; A61P 31/10(2006.01)i; A61P 31/14(2006.01)i According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) C07K 7/06(2006.01); A61K 38/17(2006.01); A61K 9/00(2006.01); C07K 14/435(2006.01); C07K 14/47(2006.01); C07K 19/00(2006.01); C12N 15/62(2006.01) Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Korean utility models and applications for utility models Japanese utility models and applications for utility models Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) eKOMPASS(KIPO internal) & Keywords: orai1 calcium channel, lung immunomodulator, infection		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WU, T. et al., 'Identification of BPIFA1/SPLUNC1 as an epithelium-derived smooth muscle relaxing factor', Nature Communications, 2017, vol.8, article number:14118, pages 1-10 abstract; pages 2-4, 8	1-3,11-16
A	Internet webpage, Uniprot, 'UniProtKB - Q9NP55 (BPIA1_HUMAN)', 2000.10.01, https://www.uniprot.org/uniprot/Q9NP55 the whole document	1-3,11-16
A	US 2019-0062386 A1 (THE UNIVERSITY OF NORTH CAROLINA AT CHAPEL HILL) 28 February 2019 (2019-02-28) the whole document	1-3,11-16
A	WO 2013-043720 A1 (THE UNIVERSITY OF NORTH CAROLINA AT CHAPEL HILL) 28 March 2013 (2013-03-28) the whole document	1-3,11-16
☒ Further documents are listed in the continuation of Box C. ☒ See patent family annex.		
* Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance "D" document cited by the applicant in the international application "E" earlier application or patent but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "&" document member of the same patent family		
Date of the actual completion of the international search 04 July 2022		Date of mailing of the international search report 04 July 2022
Name and mailing address of the ISA/KR Korean Intellectual Property Office 189 Cheongsa-ro, Seo-gu, Daejeon 35208, Republic of Korea Facsimile No. +82-42-481-8578		Authorized officer HEO, Joo Hyung Telephone No. +82-42-481-5373

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2022/021391

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	CN 106749679 A (INSTITUTE OF DISEASE CONTROL AND PREVENTION OF PLA) 31 May 2017 (2017-05-31) the whole document	1-3,11-16

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2022/021391

Box No. I **Nucleotide and/or amino acid sequence(s) (Continuation of item 1.c of the first sheet)**

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international search was carried out on the basis of a sequence listing:
 - a. ☒ forming part of the international application as filed:
 - ☒ in the form of an Annex C/ST.25 text file.
 - ☐ on paper or in the form of an image file.
 - b. ☐ furnished together with the international application under PCT Rule 13*ter*.1(a) for the purposes of international search only in the form of an Annex C/ST.25 text file.
 - c. ☐ furnished subsequent to the international filing date for the purposes of international search only:
 - ☐ in the form of an Annex C/ST.25 text file (Rule 13*ter*.1(a)).
 - ☐ on paper or in the form of an image file (Rule 13*ter*.1(b) and Administrative Instructions, Section 713).
2. ☐ In addition, in the case that more than one version or copy of a sequence listing has been filed or furnished, the required statements that the information in the subsequent or additional copies is identical to that forming part of the application as filed or does not go beyond the application as filed, as appropriate, were furnished.
3. Additional comments:

Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: **37-80**
because they relate to subject matter not required to be searched by this Authority, namely:

Claims 37-80 pertain to a method for treatment of the human body by therapy or surgery and thus relate to a subject matter which this International Searching Authority is not required, under PCT Article 17(2)(a)(i) and PCT Rule 39.1(iv), to search.
2. ☒ Claims Nos.: **18,23,25,29,31,42-47,49-55,59,64,67,70-74,78**
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

Claims 18,23,25,29,31,42-47,49-55,59,64,67,70-74,78 are regarded to be unclear because they refer to claims which do not comply with PCT Rule 6.4(a).
3. ☒ Claims Nos.: **4-10,17,19-22,24,26-28,30,32-41,48,56-58,60-63,65-66,68-69,75-77,79-80**
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

INTERNATIONAL SEARCH REPORT
Information on patent family members

International application No.

PCT/US2022/021391

Patent document cited in search report			Publication date (day/month/year)	Patent family member(s)			Publication date (day/month/year)
US	2019-0062386	A1	28 February 2019	AU	2017-222450	A1	27 September 2018
				BR	112018017144	A2	15 January 2019
				EP	3419994	A1	02 January 2019
				EP	3419994	A4	06 November 2019
				JP	2019-512216	A	16 May 2019
				MX	2018010132	A	06 June 2019
				US	10894811	B2	19 January 2021
				WO	2017-147128	A1	31 August 2017
WO	2013-043720	A1	28 March 2013	AU	2012-312520	A1	08 May 2014
				AU	2018-200871	A1	22 February 2018
				BR	112014006535	A2	28 March 2017
				CA	2849267	A1	28 March 2013
				DK	2760463	T3	18 March 2019
				EP	2760463	A1	06 August 2014
				EP	2760463	A4	29 July 2015
				EP	2760463	B1	21 November 2018
				ES	2716723	T3	14 June 2019
				HK	1200729	A1	14 August 2015
				HU	E041740	T2	28 May 2019
				IL	231614	A	28 May 2014
				PL	2760463	T3	31 May 2019
				PT	2760463	T	27 February 2019
				US	2014-0228276	A1	14 August 2014
				US	2016-0159879	A1	09 June 2016
				US	9127040	B2	08 September 2015
CN	106749679	A	31 May 2017	None			