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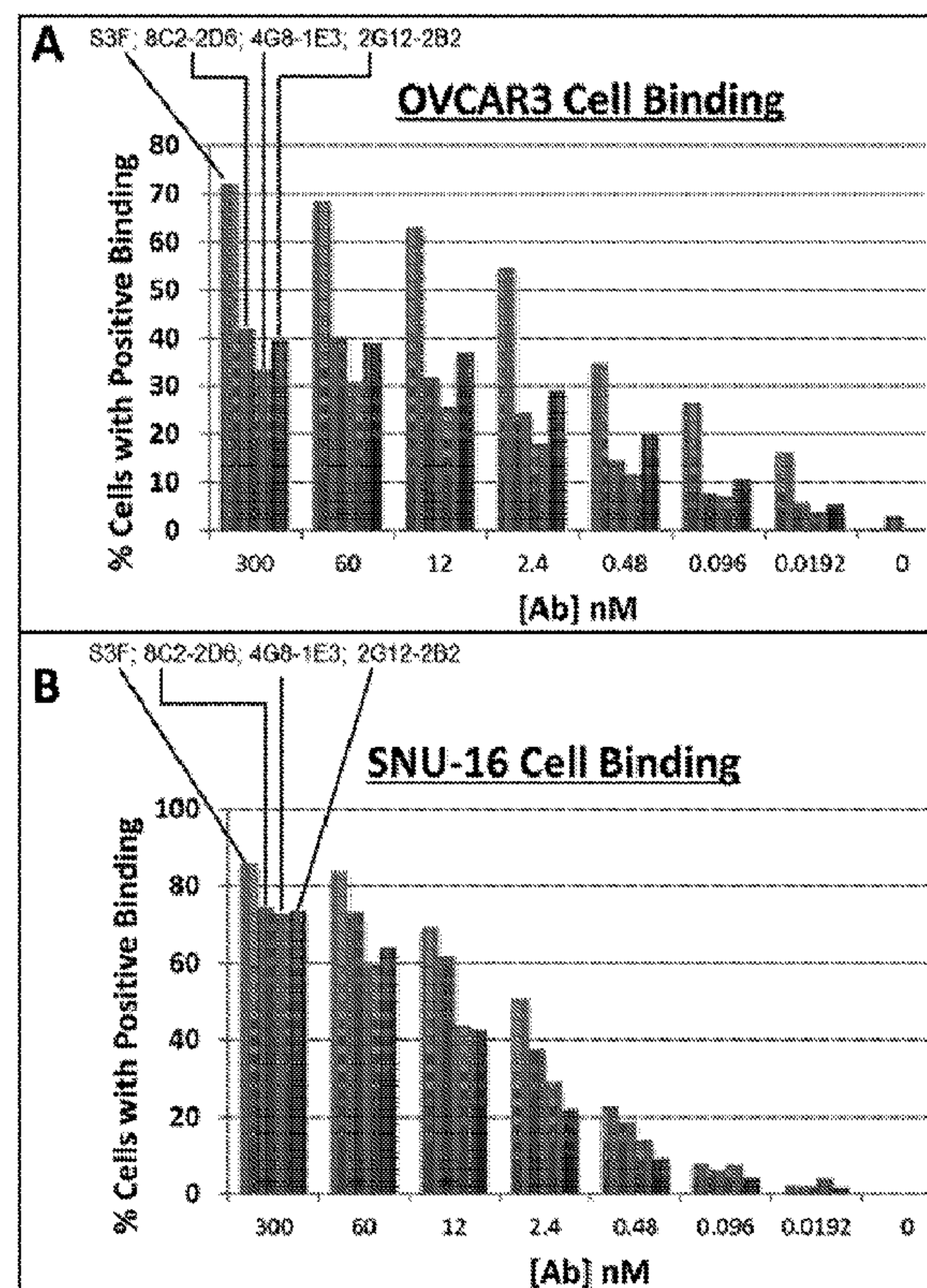
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(54) Title: GLYCAN-INTERACTING COMPOUNDS AND METHODS OF USE

Fig. 2



(57) Abrégé/Abstract:

The present invention provides glycan-interacting antibodies and methods for producing glycan-interacting antibodies useful in the treatment and prevention of human disease, including cancer. Such glycan-interacting antibodies include monoclonal antibodies,

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derivatives, and fragments thereof as well as compositions and kits comprising them. Further provided are methods of using glycan-interacting antibodies to target cells and treat disease.

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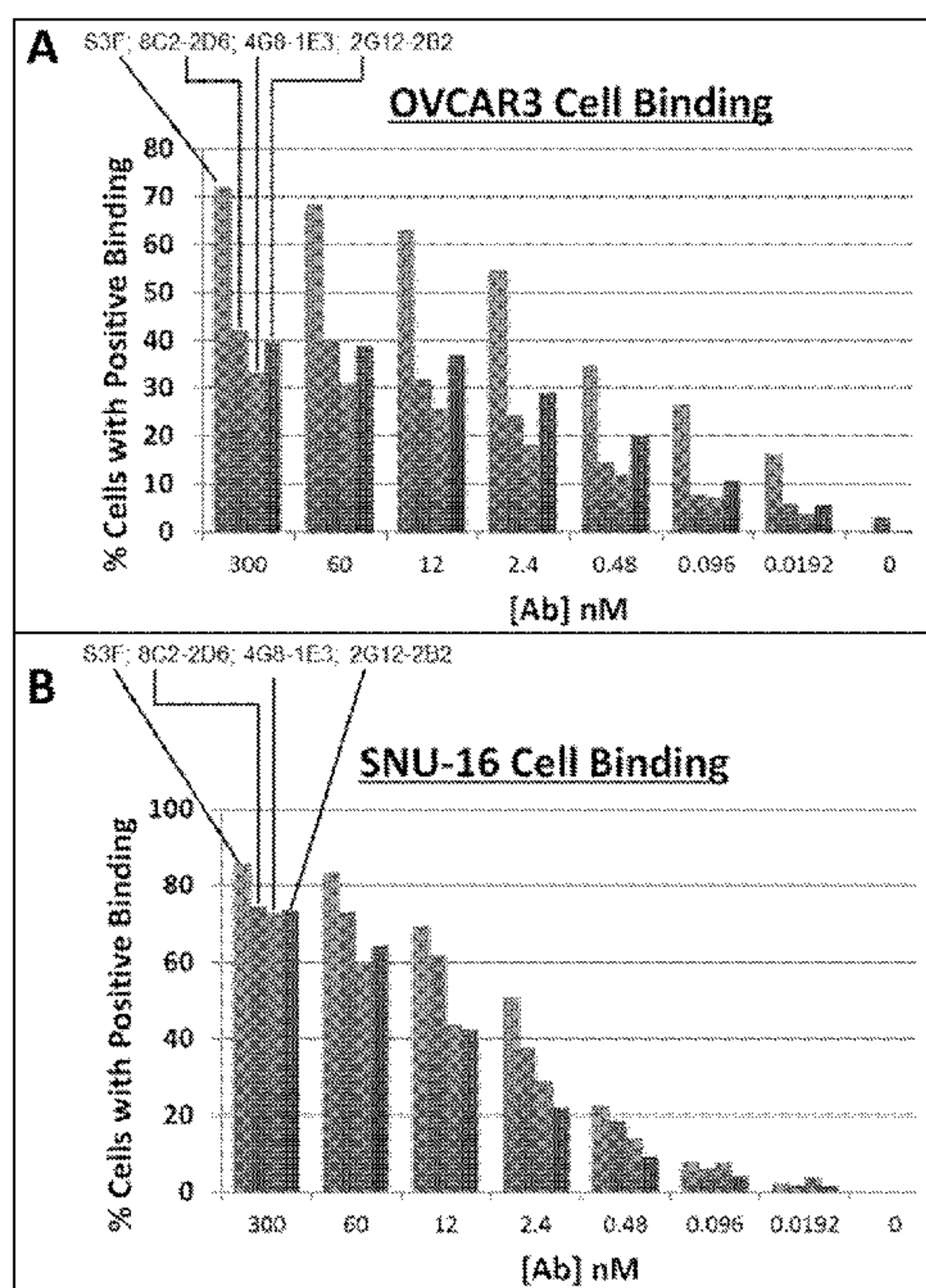
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[Continued on next page]

(54) Title: GLYCAN-INTERACTING COMPOUNDS AND METHODS OF USE

Fig. 2



(57) Abstract: The present invention provides glycan-interacting antibodies and methods for producing glycan-interacting antibodies useful in the treatment and prevention of human disease, including cancer. Such glycan-interacting antibodies include monoclonal antibodies, derivatives, and fragments thereof as well as compositions and kits comprising them. Further provided are methods of using glycan-interacting antibodies to target cells and treat disease.

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GLYCAN-INTERACTING COMPOUNDS AND METHODS OF USE

CROSS REFERENCE TO RELATED APPLICATIONS

[0001] This application claims priority to U.S. Provisional Patent Application No. 62/078,610 filed November 12, 2014 entitled Glycan-Interacting Compounds and Methods of Use, U.S. Provisional Patent Application No. 62/102,527 filed January 12, 2015 entitled Glycan-Interacting Compounds and Methods of Use, U.S. Provisional Patent Application No. 62/145,214 filed April 9, 2015 entitled Compositions and Methods for Targeting Cancer Stem Cells, U.S. Provisional Patent Application No. 62/173,560 filed June 10, 2015 entitled Glycan-Interacting Compounds and Methods of Use, and U.S. Provisional Patent Application No. 62/187,587 filed July 1, 2015 entitled Glycan-Interacting Compounds and Methods of Use, the contents of each of which are herein incorporated by reference in their entirety.

GOVERNMENT SUPPORT

[0002] This invention was made with government support under Grant No. HHSN261201400027C awarded by the Department of Health and Human Services. The United States government may have certain rights in the invention.

SEQUENCE LISTING

[0003] The instant application contains a Sequence Listing which has been submitted electronically in ASCII format and is hereby incorporated by reference in its entirety. Said ASCII copy, created on November 1, 2015, is named 2033_1012PCT_SL.txt and is 144,772 bytes in size.

FIELD OF THE INVENTION

[0004] This invention relates to methods for the development of compounds and compositions, including, but not limited to antibodies for the detection and/or removal of glycosylated matter from an organism.

BACKGROUND OF THE INVENTION

[0005] Aberrant glycosylation accompanies some of the other mutations commonly observed in carcinomas. It has been estimated that about 80% of all carcinomas express the truncated glycans, the Tn Antigen and the sialylated form, Sialyl Tn (STn). With few exceptions, Tn and STn are not expressed in normal, healthy tissues. Furthermore, the non-human immunogenic sialic acid, N-glycolylneuraminic acid (Neu5Gc), seems to be

differentially expressed on carcinomas such as breast cancer in the form of Neu5Gc-STn (GcSTn).

[0006] Multiple aberrant glycosylation forms have been described in human cancers, identifying specific glycans as a class of cell surface molecules suitable for specific tumor targeting (Cheever, M.A. et al., Clin Cancer Res. 2009 Sep 1;15(17):5323-37). For example, various human cancer types (such as bladder, breast, cervical, colon, lung, and ovarian cancer among others) show high expression of STn antigen, which is rare in normal human tissues (Karlen, P. et al., Gastroenterology. 1998 Dec;115(6):1395-404; Ohno, S. et al, Anticancer Res. 2006 Nov-Dec;26(6A):4047-53). In addition, the presence of STn on tumor-associated mucins relates to cancer with poor prognosis and is therewith considered an attractive epitope for cancer detection and targeted therapy (Cao, Y. et al., Virchows Arch. 1997 Sep;431(3):159-66; Julien, S. et al., Br J Cancer. 2009 Jun 2;100(11):1746-54; Itzkowitz, S.H. et al., Cancer. 1990 Nov 1;66(9):1960-6; Motoo, Y. et al., Oncology. 1991;48(4):321-6; Kobayashi, H. et al., J Clin Oncol. 1992 Jan;10(1):95-101). Tn and STn formation is associated with somatic mutations in the gene Cosmc that encodes a molecular chaperon required for the formation of the activate T-synthase (Ju, T. et al., Nature. 2005 Oct 27;437(7063):1252; Ju, T. et al., Cancer Res. 2008 Mar 15;68(6):1636-46). It can also result from increased expression of the sialyl transferase, ST6GalNAc-I (Ikehara, Y. et al., Glycobiology. 1999 Nov;9(11):1213-24; Brockhausen, I. et al., Biol Chem. 2001 Feb;382(2):219-32). De-novo expression of STn can modulate carcinoma cells, change the malignant phenotype, and lead to more aggressive cell behaviors (Pinho, S. et al., Cancer Lett. 2007 May 8;249(2):157-70). Although STn is highly expressed in malignant tissues, low levels are also found on healthy human cells (Jass, J.R. et al., J Pathol. 1995 Jun;176(2):143-9; Kirkeby, S. et al., Arch Oral Biol. 2010 Nov;55(11):830-41). STn alone has attracted attention as a target for cancer detection and therapy (Cheever, M.A. et al., Clin Cancer Res. 2009 Sep 1;15(17):5323-37). STn is also present in mucins associated with cancer stem cells (Engelmann et al., Cancer research, 2008, 68, 2419-2426) and STn is implicated in immune suppression (Carrascal, M.A., et al., Molecular Oncology. 2014. 8(3): 753-65).

[0007] In addition to the presence of STn, other glycosylation changes have been described in cancer. One of them involves Neu5Gc. *N*-acetylneuraminic acid (Neu5Ac) and Neu5Gc are the two major sialic acids on mammalian cell surfaces. Neu5Ac and Neu5Gc differ only in that Neu5Gc comprises an additional oxygen atom associated with chemical

group attached to carbon 5. Due to the loss of a functional gene, humans can only synthesize sialic acid in the form of Neu5Ac, but not Neu5Gc. However Neu5Gc can be metabolically incorporated into humans from animal-derived dietary sources such as red meats (Tangvoranuntakul, P. et al., *Proc Natl Acad Sci U S A.* 2003 Oct 14; 100(21):12045-50; Nguyen, D.H. et al., *J Immunol.* 2005 Jul 1; 175(1):228-36; US7,682,794, US8,084,219, US2012/0142903, WO2010030666 and WO2010030666). Neu5Gc is significantly abundant among human tumors (Higashi, H. et al., *Cancer Res.* 1985 Aug; 45(8):3796-802; Miyoshi I. et al., *Mol Immunol.* 1986. 23: 631-638; Hirabayashi, Y. et al., *Jpn J Cancer Res.* 1987. 78: 614-620; Kawachi, S. et al., *Int Arch Allergy Appl Immunol.* 1988. 85: 381-383; Devine, P.L. et al., *Cancer Res.* 1991. 51: 5826-5836; Malykh, Y.N. et al., *Biochimie.* 2001. 83: 623-634 and Inoue, S. et al., 2010. *Glycobiology.* 20(6): 752-762) and remarkably low in normal human tissues, which had been overlooked for several decades (Diaz, S.L. et al., *PLoS One.* 2009. 4: e4241; Tangvoranuntakul, P. et al., *Proc Natl Acad Sci U S A.* 2003. 100: 12045-12050; Varki, A. et al., *Glycoconj J.* 2009. 26: 231-245). The increased metabolic accumulation of diet-derived Neu5Gc in cancer tissue compared to healthy human tissues is likely explained by at least three factors: rapid growth with underproduction of competing endogenous Neu5Ac, enhanced macropinocytosis induced by growth factors (Dharmawardhane, S. et al., *Mol Biol Cell.* 2000 Oct;11(10):3341-52; Simonsen, A. et al., *Curr Opin Cell Biol.* 2001 Aug;13(4):485-92; Johannes, L. et al., *Traffic.* 2002 Jul;3(7):443-51; Amyere, M. et al., *Int J Med Microbiol.* 2002 Feb;291(6-7):487-94), and the upregulation of gene expression of the lysosomal sialic acid transporter gene sialin by hypoxia (Yin, J. et al., *Cancer Res.* 2006 Mar 15;66(6):2937-45). In addition, all humans tested to date comprise a polyclonal antibody reservoir against non-human Neu5Gc, which makes it the first example of a xeno-autoantigen (Padler-Karavani, V. et al., *Glycobiology.* 2008 Oct;18(10):818-30; Varki, N.M. et al., *Annu Rev Pathol.* 2011;6:365-93). The accumulation of dietary Neu5Gc in malignant tumors in the face of an anti-Neu5Gc response was shown to facilitate tumor progression by inducing a low-grade chronic inflammation (Hedlund, M. et al., *Proc Natl Acad Sci U S A.* 2008 Dec 2;105(48):18936-41). Thus, Neu5Gc containing glycan epitopes on human tumors represent a valuable possibility for drug targeting. A recent study suggests the existence of antibodies against Neu5Gc-containing STn (GcSTn), but not Neu5Ac-STn (AcSTn), in cancer patients and explores their potential as a specific biomarker for cancer detection (Padler-Karavani, V. et al., *Cancer Res.* 2011 May 1;71(9):3352-63).

[0008] There remains a need in the art for antibodies capable of binding glycans, including glycans associated with disease and diseased cells and tissues. Such antibodies could be used to target tumor cells and cancerous tissues and to treat subjects suffering from cancer. Further, there remains a need for better methods to develop such antibodies as well as methods for specific characterization of epitopes bound by glycan-interacting antibodies. The present invention meets this need by providing antibodies that target glycans, by providing methods of developing anti-glycan antibodies, and by providing methods of using anti-glycan antibodies to identify and target cancerous cells and tissues in the diagnosis and treatment of disease.

SUMMARY OF THE INVENTION

[0009] In some embodiments, the present invention provides an antibody comprising a variable domain, the variable domain comprising at least 95% amino acid sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NOs: 15-52 or a fragment thereof. In some cases, such antibodies comprise a variable domain pair selected from the group consisting of SEQ ID NO:29 and SEQ ID NO:30; SEQ ID NO:15 and SEQ ID NO:16; SEQ ID NO:17 and SEQ ID NO:18; SEQ ID NO:19 and SEQ ID NO:20; SEQ ID NO:21 and SEQ ID NO:22; SEQ ID NO:23 and SEQ ID NO:24; SEQ ID NO:25 and SEQ ID NO:26; SEQ ID NO:27 and SEQ ID NO:28; SEQ ID NO:31 and SEQ ID NO:32; SEQ ID NO:35 and SEQ ID NO:36; SEQ ID NO:37 and SEQ ID NO:38; SEQ ID NO:39 and SEQ ID NO:40; SEQ ID NO:42 and SEQ ID NO:32; SEQ ID NO:44 and SEQ ID NO:45; SEQ ID NO:46 and SEQ ID NO:32; SEQ ID NO:47 and SEQ ID NO:32; SEQ ID NO:48 and SEQ ID NO:50; SEQ ID NO:48 and SEQ ID NO:49; and SEQ ID NO:51 and SEQ ID NO:52.

[0010] Some antibodies of the invention comprise at least one complementarity determining region (CDR) comprising from about 60% to about 95% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NOs: 81-148, SEQ ID NOs: 152-155, and SEQ ID NOs: 159-164. In some cases, such antibodies comprise a CDR with a heavy chain variable domain (VH) CDR (CDR-H) comprising at least 70% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NOs: 81-99, SEQ ID NOs: 143-148, and SEQ ID NOs: 152-155. Some antibodies comprise a CDR-H3 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 90-99, and SEQ ID NOs: 152-155. In some cases, antibodies comprise a light chain variable domain (VL) CDR (CDR-L) comprising at least 70% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NOs: 100-142, and SEQ ID NOs:

159-164. Such antibodies may comprise a CDR-L3 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 127-142.

[0011] In some embodiments, the present invention provides an antibody comprising: (1) a CDR-L1 selected from the group consisting of SEQ ID NOs: 100-114, and SEQ ID NOs: 159-167; (2) a CDR-L2 selected from the group consisting of SEQ ID NOs: 115-126, and SEQ ID NOs: 168-170; (3) a CDR-L3 selected from the group consisting of SEQ ID NOs: 127-142, and SEQ ID NOs: 171-173; (4) a CDR-H1 selected from the group consisting of SEQ ID NOs: 81-83, SEQ ID NO: 143, and SEQ ID NO: 144; (5) a CDR-H2 selected from the group consisting of SEQ ID NOs: 84-89, and SEQ ID NOs: 145-151; and (6) a CDR-H3 selected from the group consisting of SEQ ID NOs: 90-99, and SEQ ID NOs: 152-158.

[0012] Some antibodies of the invention are monoclonal antibodies. Some antibodies comprise an IgG1 isotype. Other antibodies comprise an IgG2 isotype. In some cases, antibodies of the invention belong to an antibody group selected from the group consisting of a Group 1 antibody, a Group 2 antibody, a Group 3 antibody and a Group 4 antibody. Some antibodies specifically target an antigen selected from the group consisting of N-acetylneuraminic sialyl Tn antigen (AcSTn) and N-glycolylneuraminic sialyl Tn antigen (GcSTn). Such antibodies may target antigens comprising a 9-O-acetyl group. Some antibodies bind to a cluster of two or more glycans. Further antibodies comprise bispecific antibodies.

[0013] In some embodiments, antibodies of the invention comprise an antibody-drug conjugate. Some antibody-drug conjugates comprise a therapeutic agent. Some antibody-drug conjugates comprise a cytotoxic agent. In some cases, cytotoxic agents are conjugated to antibodies directly. In some cases, cytotoxic agents are conjugated to antibodies via a linker. Such linkers may include cleavable linkers or non-cleavable linkers. In some cases, the cytotoxic agent is a DNA damaging agent. In some cases, the cytotoxic agent is a cytoskeletal inhibitor.

[0014] According to some embodiments, antibodies of the invention bind a tumor-associated carbohydrate antigen (TACA). Such TACAs may include Tn antigen, sialylated Tn antigen (STn,) Thomsen-Friedenreich antigen, Lewis^Y (Le^Y) antigen, Lewis^X (Le^X) antigen, Sialyl Lewis^X (SLe^X) antigen, Sialyl Lewis^A (SLe^A) antigen, Globo H, stage-specific embryonic antigen-3 (SSEA-3,) a glycosphingolipid comprising sialic acid, ganglioside GD2, ganglioside GD3, ganglioside GM2, fucosyl GM1, ganglioside Neu5GcGM3, or a polysialic acid-related antigen.

[0015] In some embodiments, the present invention provides a method of killing a tumor cell comprising contacting the tumor cell with an antibody of the invention.

[0016] According to other embodiments, the present invention provides a method of treating cancer in a subject in need thereof, the method comprising administering to the subject an antibody of the invention. In some cases, such methods are used to treat an epithelial cancer. Such epithelial cancers may include breast, colon, lung, bladder, cervical, ovarian, stomach, prostate, and/or liver cancer.

[0017] In some aspects, the present invention provides compositions comprising at least one of the antibodies of the invention. Further provided are kits comprising such compositions and instructions for use thereof.

[0018] In some embodiments, the present invention provides methods of reducing tumor volume in a subject comprising administering to said subject a composition of the invention. In some cases, methods of the invention may be used to increase anti-tumor cell immune activity by contacting at least one immune-resistant tumor cell with an antibody of the invention. Such anti-tumor cell immune activity may include innate immune activity (e.g., natural killer (NK) cell anti-tumor cell activity). In some cases, anti-tumor cell activity includes adaptive immune activity (e.g., B cell anti-tumor cell activity or dendritic cell (DC) anti-tumor cell activity). In some cases, compositions of the invention may increase DC expression of one or more factor selected from the group consisting of CD80, CD86, IL-12, and TNF- α . Further methods of the invention include treating an immune-resistant tumor in a subject in need thereof by administering a composition of the invention.

[0019] According to some embodiments, the present invention provides methods of reducing or eliminating cancer stem cells (CSCs) in a subject comprising administering to the subject an antibody of the invention. In some cases, the CSCs are present in breast, ovarian, pancreatic, bladder, cervical, colon and/or lung tissue. In some cases the CSCs comprise CD133 and/or CD44 biomarkers. In some cases, methods of reducing or eliminating CSCs further comprise the administration of at least one chemotherapeutic agent. Such chemotherapeutic agents may be selected from paclitaxel and carboplatin.

[0020] In some aspects, the present invention provides intrabodies comprising a variable domain comprising at least 95% amino acid sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO: 15-52 and a fragment thereof. Some intrabodies comprise a variable domain pair selected from the group consisting of SEQ ID NO:29 and SEQ ID NO:30; SEQ ID NO:15 and SEQ ID NO:16; SEQ ID NO:17 and SEQ ID

NO:18; SEQ ID NO:19 and SEQ ID NO:20; SEQ ID NO:21 and SEQ ID NO:22; SEQ ID NO:23 and SEQ ID NO:24; SEQ ID NO:25 and SEQ ID NO:26; SEQ ID NO:27 and SEQ ID NO:28; SEQ ID NO:31 and SEQ ID NO:32; SEQ ID NO:35 and SEQ ID NO:36; SEQ ID NO:37 and SEQ ID NO:38; SEQ ID NO:39 and SEQ ID NO:40; SEQ ID NO:42 and SEQ ID NO:32; SEQ ID NO:44 and SEQ ID NO:45; SEQ ID NO:46 and SEQ ID NO:32; SEQ ID NO:47 and SEQ ID NO:32; SEQ ID NO:48 and SEQ ID NO:50; SEQ ID NO:48 and SEQ ID NO:49; and SEQ ID NO:51 and SEQ ID NO:52.

[0021] In other aspects, the present invention provides a chimeric antigen receptor (CAR) comprising a variable domain comprising at least 95% amino acid sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NOs: 15-52 and a fragment thereof. Some CARs comprise a variable domain pair selected from the group consisting of SEQ ID NO:29 and SEQ ID NO:30; SEQ ID NO:15 and SEQ ID NO:16; SEQ ID NO:17 and SEQ ID NO:18; SEQ ID NO:19 and SEQ ID NO:20; SEQ ID NO:21 and SEQ ID NO:22; SEQ ID NO:23 and SEQ ID NO:24; SEQ ID NO:25 and SEQ ID NO:26; SEQ ID NO:27 and SEQ ID NO:28; SEQ ID NO:31 and SEQ ID NO:32; SEQ ID NO:35 and SEQ ID NO:36; SEQ ID NO:37 and SEQ ID NO:38; SEQ ID NO:39 and SEQ ID NO:40; SEQ ID NO:42 and SEQ ID NO:32; SEQ ID NO:44 and SEQ ID NO:45; SEQ ID NO:46 and SEQ ID NO:32; SEQ ID NO:47 and SEQ ID NO:32; SEQ ID NO:48 and SEQ ID NO:50; SEQ ID NO:48 and SEQ ID NO:49; and SEQ ID NO:51 and SEQ ID NO:52.

[0022] In some embodiments, the present invention provides constructs encoding antibodies of the invention. Such constructs may comprise at least one nucleotide sequence having at least 95% sequence identity to a nucleotide sequence selected from the group consisting of SEQ ID NOs: 192-233 and a fragment thereof. In some cases, constructs of the invention encode intrabodies or CARs of the invention.

[0023] Further provided are cells comprising one or more constructs of the invention. Also provided are viruses comprising constructs of the invention.

[0024] According to some embodiments, the present invention provides methods of identifying cells and/or tissues expressing STn with antibodies of the invention. Further provided are methods of identifying cancerous cells in tissues or organs with antibodies of the invention. Such tissues or organs may include breast, ovary, and pancreas.

[0025] In some embodiments, the present invention provides antibody variants comprising sequence analysis-informed variants of antibodies of the invention. Such antibody variants may comprise improved epitope specificity and/or affinity. Some antibody variants comprise

a CDR length modification (e.g., a CDR-H3 length modification). Some antibody variants comprise a substitution of one or more amino acids in one or more CDRs. In some cases, antibody variants comprise at least one germline gene modification. Some antibody variants comprise an scFv, monobody, diabody, intrabody, CAR, or antibody mimetic.

[0026] In some aspects, the present invention provides a method of developing an antibody fragment display library comprising: (1) aligning variable domain amino acid sequences among two or more antibodies of the invention; (2) identifying conserved and variable amino acids among the aligned antibodies, and (3) constructing an antibody fragment display library, wherein variability among library members is limited to amino acid variation of the variable amino acids identified. Further embodiments provide antibody fragment display libraries produced according to such methods. Additional embodiments provide antibodies comprising variable domains isolated from such antibody fragment display libraries. Such antibodies may include an scFv, monobody, diabody, intrabody, CAR, or antibody mimetic.

BRIEF DESCRIPTION OF THE FIGURES

[0027] The foregoing and other objects, features and advantages will be apparent from the following description of particular embodiments of the invention, as illustrated in the accompanying drawings. The drawings are not necessarily to scale, emphasis instead being placed upon illustrating the principles of various embodiments of the invention.

[0028] Figs. 1A-1D are diagrams depicting α 2,6-sialylated N-acetylgalactosamine (STn) and indicating putative epitopes involved in anti-STn antibody binding. The largest ellipse in each diagram indicates the specific region of STn targeted by each of 4 antibody groups. These groups include Group 1 antibodies (binding to the large elliptical region indicated in Fig. 1A), Group 2 antibodies (binding to the large elliptical region indicated in Fig. 1B), Group 3 antibodies (binding to the large elliptical region indicated in Fig. 1C) and Group 4 antibodies (binding to the large elliptical region indicated in Fig. 1D).

[0029] Figs. 2A and 2B are bar graphs presenting antibody binding data. Fig. 2A is a bar graph presenting antibody binding results with OVCAR3 cells. Fig. 2B is a bar graph presenting antibody binding results with SNU-16 cells.

DETAILED DESCRIPTION

Introduction

[0030] According to the present invention are antibodies specific for or which interact with epitopes comprising carbohydrate groups referred to herein as glycans. Some glycan-interacting antibodies described herein may be used as biotherapeutics. Other embodiments provide methods for generating such glycan-interacting antibodies.

[0031] In nature, glycans may be sialylated with N-acetylneuraminic acid (Neu5Ac) or N-glycolylneuraminic acid (Neu5Gc). Glycan-interacting antibodies according to the present invention may be directed to cancer-related glycans comprising α 2,6-sialylated N-acetylgalactosamine (STn). Such antibodies may target any STns (pan-STn antibodies), glycans comprising STns having Neu5Ac specifically (AcSTn) or glycans having STns comprising Neu5Gc specifically (GcSTn). In some embodiments, glycan-interacting antibodies of the present invention target other cancer-related glycan antigens.

[0032] In some embodiments, the present invention provides methods of producing glycan-interacting antibodies. Such methods may comprise the use of mice for generating an immune response to one or more antigens comprising STn (e.g. AcSTn and/or GcSTn). As described herein, a number of methods may be utilized in order to manipulate the resulting antibodies produced through mouse immunization. Such methods may include varying the strain and/or gender of the mice being immunized, varying the antigen used, varying the type and dose of adjuvant included in antigen administration and time course of immunization before initiation of hybridoma fusion.

[0033] In some embodiments, the present invention provides methods for eliminating cancer stem cells using glycan-interacting antibodies. In other embodiments, the present invention provides methods for treating cancer in a subject by eliminating cancer stem cells using glycan-interacting antibodies. In some aspects, glycan-interacting antibodies may be used alone. In other aspects, glycan-interacting antibodies are used in combination with chemotherapeutic agents.

[0034] Further provided are optimized, humanized, and conjugated forms of glycan-interacting antibodies. Additionally, kits, assays and reagents comprising antibodies and/or methods of the present invention are presented.

Definitions

[0035] *Adjacent*: As used herein, the term “adjacent” refers to something that is adjoining, neighboring or next to a given entity. In some embodiments, “adjacent residues” are sugar residues within a glycan chain that are linked to one another. In some embodiments, “adjacent glycans” are glycan chains that next to each other either in direct contact or within close proximity and without another glycan in between the two.

[0036] *Administered in combination*: As used herein, the term “administered in combination” or “combined administration” means that a subject is simultaneously exposed to two or more agents administered at the same time or within an interval of time such that the subject is at some point in time simultaneously exposed to both and/or such that there may be an overlap in the effect of each agent on the patient. In some embodiments, at least one dose of one or more agents is administered within about 24 hours, 12 hours, 6 hours, 3 hours, 1 hour, 30 minutes, 15 minutes, 10 minutes, 5 minutes, or 1 minute of at least one dose of one or more other agents. In some embodiments, administration occurs in overlapping dosage regimens. As used herein, the term “dosage regimen” refers to a plurality of doses spaced apart in time. Such doses may occur at regular intervals or may include one or more hiatus in administration. In some embodiments, the administration of individual doses of one or more glycan-interacting antibodies, as described herein, are spaced sufficiently closely together such that a combinatorial (*e.g.*, a synergistic) effect is achieved.

[0037] *Amino acid*: As used herein, the terms “amino acid” and “amino acids” refer to all naturally occurring L-alpha-amino acids as well as non-naturally occurring amino acids. Amino acids are identified by either the one-letter or three-letter designations as follows: aspartic acid (Asp:D), isoleucine (Ile:I), threonine (Thr:T), leucine (Leu:L), serine (Ser:S), tyrosine (Tyr:Y), glutamic acid (Glu:E), phenylalanine (Phe:F), proline (Pro:P), histidine (His:H), glycine (Gly:G), lysine (Lys:K), alanine (Ala:A), arginine (Arg:R), cysteine (Cys:C), tryptophan (Trp:W), valine (Val:V), glutamine (Gln:Q) methionine (Met:M), asparagine (Asn:N), where the amino acid is listed first followed parenthetically by the three and one letter codes, respectively.

[0038] *Animal*: As used herein, the term “animal” refers to any member of the animal kingdom. In some embodiments, “animal” refers to humans at any stage of development. In some embodiments, “animal” refers to non-human animals at any stage of development. In certain embodiments, the non-human animal is a mammal (*e.g.*, a rodent, a mouse, a rat, a rabbit, a monkey, a dog, a cat, a sheep, cattle, a primate, or a pig). In some embodiments,

animals include, but are not limited to, mammals, birds, reptiles, amphibians, fish, and worms. In some embodiments, the animal is a transgenic animal, genetically-engineered animal, or a clone.

[0039] *Antibody*: As used herein, the term "antibody" is used in the broadest sense and specifically covers various embodiments including, but not limited to monoclonal antibodies, polyclonal antibodies, multispecific antibodies (e.g. bispecific antibodies formed from at least two intact antibodies), and antibody fragments such as diabodies so long as they exhibit a desired biological activity. Antibodies are primarily amino-acid based molecules but may also comprise one or more modifications such as with sugar moieties.

[0040] *Antibody fragment*: As used herein, the term "antibody fragment" refers to a portion of an intact antibody, preferably comprising an antigen binding region thereof. Examples of antibody fragments include Fab, Fab', F(ab')₂, and Fv fragments; diabodies; linear antibodies; single-chain antibody molecules; and multispecific antibodies formed from antibody fragments. Papain digestion of antibodies produces two identical antigen-binding fragments, called "Fab" fragments, each with a single antigen-binding site. Also produced is a residual "Fc" fragment, whose name reflects its ability to crystallize readily. Pepsin treatment yields an F(ab')₂ fragment that has two antigen-binding sites and is still capable of cross-linking antigen. Glycan-interacting antibodies may comprise one or more of these fragments. For the purposes herein, an antibody may comprise a heavy and light variable domain as well as an Fc region.

[0041] *Approximately*: As used herein, the term "approximately" or "about," as applied to one or more values of interest, refers to a value that is similar to a stated reference value. In certain embodiments, the term "approximately" or "about" refers to a range of values that fall within 25%, 20%, 19%, 18%, 17%, 16%, 15%, 14%, 13%, 12%, 11%, 10%, 9%, 8%, 7%, 6%, 5%, 4%, 3%, 2%, 1%, or less in either direction (greater than or less than) of the stated reference value unless otherwise stated or otherwise evident from the context (except where such number would exceed 100% of a possible value).

[0042] *Associated with*: As used herein, the terms "associated with," "conjugated," "linked," "attached," and "tethered," when used with respect to two or more moieties, means that the moieties are physically associated or connected with one another, either directly or via one or more additional moieties that serves as a linking agent, to form a structure that is sufficiently stable so that the moieties remain physically associated under the conditions in which the structure is used, e.g., physiological conditions. An "association" need not be

strictly through direct covalent chemical bonding. It may also suggest ionic or hydrogen bonding or a hybridization based connectivity sufficiently stable such that the “associated” entities remain physically associated.

[0043] *Bifunctional*: As used herein, the term “bifunctional” refers to any substance, molecule or moiety which is capable of or maintains at least two functions. The functions may affect the same outcome or a different outcome. The structure that produces the function may be the same or different.

[0044] *Biomolecule*: As used herein, the term “biomolecule” is any natural molecule which is amino acid-based, nucleic acid-based, carbohydrate-based or lipid-based, and the like.

[0045] *Bispecific antibody*: As used herein, the term “bispecific antibody” refers to an antibody capable of binding two different antigens. Such antibodies typically comprise regions from at least two different antibodies. Bispecific antibodies may include any of those described in Riethmuller, G. 2012. Cancer Immunity. 12:12-18, Marvin, J.S. et al., 2005. Acta Pharmacologica Sinica. 26(6):649-58 and Schaefer, W. et al., 2011. PNAS. 108(27):11187-92, the contents of each of which are herein incorporated by reference in their entirety.

[0046] *Branch*: As used herein, the term “branch” refers to an entity, moiety or appendage that is linked or extends out from a main entity or source. In some embodiments, a “branch chain” or “branching chain” comprises one or more residues (including, but not limited to sugar residues) that extend from a parent chain. As used herein, a “parent chain” is used to refer to a chain of residues (including, but not limited to sugar residues) from which a branching chain is linked. In the case of a glycan with multiple branches, the parent chain may also refer to the source chain from which all such branches are directly or indirectly attached. In the case of a polysaccharide comprising a chain of hexose residues, parent chain linkages typically occur between carbons 1 and 4 of adjacent residues while branching chains are attached to a parent chain through a linkage between carbon 1 of the branching residue and carbon 3 of the parent residue from which the branch extends. As used herein, the term “branching residue” refers to the residue attached to the parent chain in a branching chain.

[0047] *Cancer stem cells*: As used herein, cancer stem cells (CSCs) refer to a subset of tumor cells that have the ability to self-renew. CSCs may be able to regenerate diverse cell types. In some cases, these cells are difficult or impossible to remove through surgical or chemical treatment of a tumor.

[0048] *Compound*: As used herein, the term “compound,” refers to a distinct chemical entity. In some embodiments, a particular compound may exist in one or more isomeric or isotopic forms (including, but not limited to stereoisomers, geometric isomers and isotopes). In some embodiments, a compound is provided or utilized in only a single such form. In some embodiments, a compound is provided or utilized as a mixture of two or more such forms (including, but not limited to a racemic mixture of stereoisomers). Those of skill in the art appreciate that some compounds exist in different such forms, show different properties and/or activities (including, but not limited to biological activities). In such cases it is within the ordinary skill of those in the art to select or avoid particular forms of the compound for use in accordance with the present invention. For example, compounds that contain asymmetrically substituted carbon atoms can be isolated in optically active or racemic forms. Methods on how to prepare optically active forms from optically active starting materials are known in the art, such as by resolution of racemic mixtures or by stereoselective synthesis.

[0049] *Cyclic or Cyclized*: As used herein, the term “cyclic” refers to the presence of a continuous loop. Cyclic molecules need not be circular, only joined to form an unbroken chain of subunits.

[0050] *Cytidine monophosphate-N-acetylneuraminic acid hydroxylase*: As used herein, the term “cytidine monophosphate-N-acetylneuraminic acid hydroxylase” or “CMAH” refers to an enzyme, absent in humans, but present in most other mammals (including, but not limited to mice, pigs and chimpanzees) that catalyzes the formation of N-glycolylneuraminic acid from N-acetylneuraminic acid. The absence of the enzyme in humans is due to a frameshift mutation resulting in the premature termination of the CMAH transcript and the production of a non-functional protein.

[0051] *Cytotoxic*: As used herein, the term “cytotoxic” is used to refer to an agent that kills or causes injurious, toxic, or deadly effects on a cell (*e.g.*, a mammalian cell (*e.g.*, a human cell)), bacterium, virus, fungus, protozoan, parasite, prion, or a combination thereof.

[0052] *Delivery*: As used herein, “delivery” refers to the act or manner of transporting a compound, substance, entity, moiety, cargo or payload to an intended destination.

[0053] *Delivery Agent*: As used herein, “delivery agent” refers to any substance which facilitates, at least in part, the *in vivo* delivery of a compound, substance, entity, moiety, cargo or payload.

[0054] *Detectable label*: As used herein, “detectable label” refers to one or more markers, signals, or moieties which are attached, incorporated or associated with another entity, which

markers, signals or moieties are readily detected by methods known in the art including radiography, fluorescence, chemiluminescence, enzymatic activity, absorbance and the like. Detectable labels include radioisotopes, fluorophores, chromophores, enzymes, dyes, metal ions, ligands such as biotin, avidin, streptavidin and haptens, quantum dots, and the like. Detectable labels may be located at any position in the entity with which they are attached, incorporated or associated. For example, when attached, incorporated in or associated with a peptide or protein, they may be within the amino acids, the peptides, or proteins, or located at the N- or C- termini.

[0055] *Display library:* As used herein, the term “display library” refers to a tool used in scientific discovery to identify biomolecular interactions. Different variations of display libraries exist that include the utilization of bacteriophages, yeast and ribosomes. In each case, proteins within a given library (also referred to herein as “library members”) are linked (physically or through association with a host) to the nucleic acid which encodes the protein. When a target molecule is incubated with the members of a display library, any library members that bind to the target may be isolated and the sequences encoding the bound protein may be determined through analysis of the linked nucleic acid. In some embodiments, display libraries are “phage display libraries” wherein the display library is made up of bacteriophage viral particles (also referred to herein as “phage particles”) wherein nucleic acids have been incorporated into the phage genome resulting in the production of viral coat proteins that are fused to proteins encoded by the nucleic acids that have been introduced. Such fused proteins are “displayed” on the outer surface of the assembled phage particles where they may interact with a given target.

[0056] *Distal:* As used herein, the term “distal” means situated away from the center or away from a point or region of interest.

[0057] *Engineered:* As used herein, embodiments of the invention are “engineered” when they are designed to have a feature or property, whether structural or chemical, that varies from a starting point, wild type or native molecule. Thus, engineered agents or entities are those whose design and/or production include an act of the hand of man.

[0058] *Epitope:* As used herein, an “epitope” refers to a surface or region on a molecule that is capable of interacting with components of the immune system, including, but not limited to antibodies. In some embodiments, an epitope may comprise a target site. Epitopes may comprise a region on an antigen or between two or more antigens that is specifically recognized and bound by a corresponding antibody. Some epitopes may comprise one or

more sugar residues along one or more glycan. Such epitopes may comprise 1, 2, 3, 4, 5, 6, 7, 8, 9 or at least 10 sugar residues. Epitopes may also comprise one or more regions of interaction between entities. In some embodiments, epitopes may comprise a junction between two sugar residues, between a branching chain and a parent chain or between a glycan and a protein.

[0059] *Ether bond*: As used herein, an “ether bond” refers to a chemical bond comprising an oxygen bonded between two carbon atoms. In some embodiments, ether bonds link sugar residues to other entities, including, but not limited to other sugar residues to form a glycan chain. Such bonds are also referred to as “glycosidic bonds” or “glycosidic linkages”. In the context of at least one sugar residue, the terms “link” and/or “linkage” are also used herein when referring to a glycosidic linkage. In some embodiments, linkages may link glycans to other entities, including, but not limited to proteins, lipids, phospholipids and sphingolipids. In some embodiments, sugar residues may be linked to protein, typically forming a link between a sugar residue and an amino acid residue. Such amino acid residues include serine and threonine. In some embodiments, ether bonds link glycans to a glycan array comprising a carbohydrate linker that participates in bond formation. Glycosidic linkages may differ in their stereochemical properties. In some embodiments, alpha oriented glycosidic linkages (also referred to herein as “alpha linkages”) result in an axial orientation between the bonded oxygen of the ether bond and the cyclohexane ring of the sugar residue. In some embodiments, beta oriented glycosidic linkages (also referred to herein as “beta linkages”) result in an equatorial orientation between the bonded oxygen of the ether bond and the cyclohexane ring of the sugar residue.

[0060] *Expression*: As used herein, “expression” may refer to one or more of the following events: (1) production of an RNA template from a DNA sequence (e.g., by transcription); (2) processing of an RNA transcript (e.g., by splicing, editing, 5' cap formation, and/or 3' end processing); (3) translation of an RNA into a polypeptide or protein; (4) folding of a polypeptide or protein; and (5) post-translational modification of a polypeptide or protein (e.g., by glycosylation).

[0061] *Feature*: As used herein, a “feature” refers to a characteristic, a property, or a distinctive element.

[0062] *Formulation*: As used herein, a “formulation” refers to a material or mixture prepared according to a formula and which may comprise at least one antibody, compound, substance, entity, moiety, cargo or payload and a delivery agent, carrier or excipient.

[0063] *Functional*: As used herein, a “functional” biological molecule is a biological entity with a structure and in a form in which it exhibits a property and/or activity by which it is characterized. As used herein, a “functional group” or “chemical group” refers to a characteristic group of atoms or chemical bonds that are part of a larger molecule. In some embodiments, functional groups may be associated with different molecules, but may participate in similar chemical reactions regardless of the molecule of which they are a part. Common functional groups include, but are not limited to carboxyl groups (-COOH), acetyl groups (-COH), amino groups (-NH₂), methyl groups (-CH₃), sulfate groups (-SO₃H) and acyl groups. In some embodiments, the addition of one or more functional group to a molecule may be conveyed using terms that modify the name of the functional group with the ending “-ylated”, e.g., acetylated, methylated and sulfated.

[0064] *Glycan*: As used herein, the terms “glycan”, “oligosaccharide” and “polysaccharide” are used interchangeably and refer to polymers made up of sugar monomers, typically joined by glycosidic bonds also referred to herein as linkages. In some embodiments, the terms “glycan”, “oligosaccharide” and “polysaccharide” may be used to refer to the carbohydrate portion of a glycoconjugate (e.g., glycoprotein, glycolipid or proteoglycan).

[0065] *Glycan chain*: As used herein, the term “glycan chain” refers to a sugar polymer comprising two or more sugars. In some embodiments, glycan chains are covalently linked to proteins through serine or threonine residues on the protein.

[0066] *Glycan-rich composition*: As used herein, the term “glycan-rich composition” refers to composition comprising a large percentage of glycans. In some embodiments, glycans within a glycan-rich composition may comprise from about 1% to about 10%, from about 5% to about 15%, from about 20% to about 40%, from about 30% to about 50%, from about 60% to about 80%, from about 70% to about 90% or at least 100% of the total weight of the composition.

[0067] *Glycosidic bond*: As used herein, the term “glycosidic bond” refers to a covalent bond formed between a carbohydrate and another chemical group. In some embodiments, glycosidic bonds are formed between the reducing end of one sugar molecule and the non-reducing end of a second sugar molecule or polysaccharide chain. Such glycosidic bonds are also known as O-glycosidic bonds due to the oxygen (or ether bond) between the joined sugars. In some embodiments, a glycosidic bond between two sugars or between a sugar and a linker may also be referred to as a “linkage”.

[0068] *In vitro*: As used herein, the term “*in vitro*” refers to events that occur in an artificial environment, *e.g.*, in a test tube or reaction vessel, in cell culture, in a Petri dish, *etc.*, rather than within an organism (*e.g.*, animal, plant, or microbe).

[0069] *In vivo*: As used herein, the term “*in vivo*” refers to events that occur within an organism (*e.g.*, animal, plant, or microbe or cell or tissue thereof).

[0070] *Isolated*: As used herein, the term “isolated” is synonymous with “separated”, but carries with it the inference separation was carried out by the hand of man. In one embodiment, an isolated substance or entity is one that has been separated from at least some of the components with which it was previously associated (whether in nature or in an experimental setting). Isolated substances may have varying levels of purity in reference to the substances from which they have been associated. Isolated substances and/or entities may be separated from at least about 10%, about 20%, about 30%, about 40%, about 50%, about 60%, about 70%, about 80%, about 90%, or more of the other components with which they were initially associated. In some embodiments, isolated agents are more than about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or more than about 99% pure. As used herein, a substance is “pure” if it is substantially free of other components.

[0071] *Kit*: As used herein, the term “kit” refers to a set comprising one or more components adapted for a cooperative purpose and instructions for use thereof.

[0072] *Knockout*: As used herein, the term “knockout” refers to an organism wherein an existing gene has been inactivated through a process that typically involves the hand of man. In a knockout organism, a gene that has been inactivated is said to have been “knocked out”. In some embodiments, the knocked out gene may be inactivated through the insertion of a nucleotide sequence into the gene or through replacement of the gene entirely.

[0073] *Linker*: As used herein, a “linker” refers to a moiety that connects two or more domains, moieties or entities. In one embodiment, a linker may comprise 10, 11, 12, 13, 14, 15 or more atoms. In a further embodiment, a linker may comprise a group of atoms, *e.g.*, 10-1,000 atoms, and can be comprised of the atoms or groups such as, but not limited to, carbon, amino, alkylamino, oxygen, sulfur, sulfoxide, sulfonyl, carbonyl, and imine. In some embodiments, the linker may comprise an amino acid, peptide, polypeptide or protein. In some embodiments, a moiety bound by a linker may include, but is not limited to an atom, a chemical group, a nucleoside, a nucleotide, a nucleobase, a sugar, a nucleic acid, an amino acid, a peptide, a polypeptide, a protein, a protein complex, a payload (*e.g.*, a therapeutic

agent) or a marker (including, but not limited to a chemical, fluorescent, radioactive or bioluminescent marker). The linker can be used for any useful purpose, such as to form multimers or conjugates, as well as to administer a payload, as described herein. Examples of chemical groups that can be incorporated into the linker include, but are not limited to, alkyl, alkenyl, alkynyl, amido, amino, ether, thioether, ester, alkylene, heteroalkylene, aryl, or heterocyclyl, each of which can be optionally substituted, as described herein. Examples of linkers include, but are not limited to, unsaturated alkanes, polyethylene glycols (e.g., ethylene or propylene glycol monomeric units, e.g., diethylene glycol, dipropylene glycol, triethylene glycol, tripropylene glycol, tetraethylene glycol, or tetraethylene glycol), and dextran polymers. Other examples include, but are not limited to, cleavable moieties within the linker, such as, for example, a disulfide bond (-S-S-) or an azo bond (-N=N-), which can be cleaved using a reducing agent or photolysis. Non-limiting examples of a selectively cleavable bonds include an amido bond which may be cleaved for example by the use of tris(2-carboxyethyl)phosphine (TCEP), or other reducing agents, and/or photolysis, as well as an ester bond which may be cleaved for example by acidic or basic hydrolysis. In some embodiments, a linker is a carbohydrate moiety used to link glycans to a substrate, such as in a glycan array. Such carbohydrate linkers include, but are not limited to $-O(CH_2)_2CH_2HN_2$ and $-O(CH_2)_3NHCOCH_2(OCH_2CH_2)_6NH_2$.

[0074] *Mucin*: As used herein, the term “mucin” refers to a family of proteins that are heavily glycosylated. In some embodiments mucins are produced by the submaxillary glands and are found in saliva and mucous.

[0075] *Negative selection*: As used herein, the term “negative selection” refers to the selection of library members from a display library based on their ability to bind entities and/or components of a composition that do not comprise a target antigen. In some embodiments, negative selection is used prior to positive selection to remove elements that might bind non-specifically to the target.

[0076] *Off-target*: As used herein, “off target” refers to any unintended effect on any one or more target, gene, or cellular transcript.

[0077] *Patient*: As used herein, “patient” refers to a subject who may seek or be in need of treatment, requires treatment, is receiving treatment, will receive treatment, or a subject who is under care by a trained (e.g., licensed) professional for a particular disease or condition.

[0078] *Peptide*: As used herein, “peptide” is a protein or polypeptide which is less than or equal to 50 amino acids long, e.g., about 5, 10, 15, 20, 25, 30, 35, 40, 45, or 50 amino acids long.

[0079] *Pharmaceutically acceptable*: The phrase “pharmaceutically acceptable” is employed herein to refer to those compounds, materials, compositions, and/or dosage forms which are, within the scope of sound medical judgment, suitable for use in contact with the tissues of human beings and animals without excessive toxicity, irritation, allergic response, or other problem or complication, commensurate with a reasonable benefit/risk ratio.

[0080] *Pharmaceutically acceptable excipients*: The phrase “pharmaceutically acceptable excipient,” as used herein, refers any ingredient other than active agents (e.g., as described herein) present in a pharmaceutical composition and having the properties of being substantially nontoxic and non-inflammatory in a patient. In some embodiments, a pharmaceutically acceptable excipient is a vehicle capable of suspending or dissolving the active agent. Excipients may include, for example: antiadherents, antioxidants, binders, coatings, compression aids, disintegrants, dyes (colors), emollients, emulsifiers, fillers (diluents), film formers or coatings, flavors, fragrances, glidants (flow enhancers), lubricants, preservatives, printing inks, sorbents, suspending or dispersing agents, sweeteners, and waters of hydration. Exemplary excipients include, but are not limited to: butylated hydroxytoluene (BHT), calcium carbonate, calcium phosphate (dibasic), calcium stearate, croscarmellose, crosslinked polyvinyl pyrrolidone, citric acid, crospovidone, cysteine, ethylcellulose, gelatin, hydroxypropyl cellulose, hydroxypropyl methylcellulose, lactose, magnesium stearate, maltitol, mannitol, methionine, methylcellulose, methyl paraben, microcrystalline cellulose, polyethylene glycol, polyvinyl pyrrolidone, povidone, pregelatinized starch, propyl paraben, retinyl palmitate, shellac, silicon dioxide, sodium carboxymethyl cellulose, sodium citrate, sodium starch glycolate, sorbitol, starch (corn), stearic acid, sucrose, talc, titanium dioxide, vitamin A, vitamin E, vitamin C, and xylitol.

[0081] *Pharmaceutically acceptable salts*: Pharmaceutically acceptable salts of the compounds described herein are forms of the disclosed compounds wherein the acid or base moiety is in its salt form (e.g., as generated by reacting a free base group with a suitable organic acid). Examples of pharmaceutically acceptable salts include, but are not limited to, mineral or organic acid salts of basic residues such as amines; alkali or organic salts of acidic residues such as carboxylic acids; and the like. Representative acid addition salts include acetate, adipate, alginate, ascorbate, aspartate, benzenesulfonate, benzoate, bisulfate, borate,

butyrate, camphorate, camphorsulfonate, citrate, cyclopentanepropionate, digluconate, dodecylsulfate, ethanesulfonate, fumarate, glucoheptonate, glycerophosphate, hemisulfate, heptonate, hexanoate, hydrobromide, hydrochloride, hydroiodide, 2-hydroxy-ethanesulfonate, lactobionate, lactate, laurate, lauryl sulfate, malate, maleate, malonate, methanesulfonate, 2-naphthalenesulfonate, nicotinate, nitrate, oleate, oxalate, palmitate, pamoate, pectinate, persulfate, 3-phenylpropionate, phosphate, picrate, pivalate, propionate, stearate, succinate, sulfate, tartrate, thiocyanate, toluenesulfonate, undecanoate, valerate salts, and the like. Representative alkali or alkaline earth metal salts include sodium, lithium, potassium, calcium, magnesium, and the like, as well as nontoxic ammonium, quaternary ammonium, and amine cations, including, but not limited to ammonium, tetramethylammonium, tetraethylammonium, methylamine, dimethylamine, trimethylamine, triethylamine, ethylamine, and the like. Pharmaceutically acceptable salts include the conventional non-toxic salts, for example, from non-toxic inorganic or organic acids. In some embodiments a pharmaceutically acceptable salt is prepared from a parent compound which contains a basic or acidic moiety by conventional chemical methods. Generally, such salts can be prepared by reacting the free acid or base forms of these compounds with a stoichiometric amount of the appropriate base or acid in water or in an organic solvent, or in a mixture of the two; generally, nonaqueous media like ether, ethyl acetate, ethanol, isopropanol, or acetonitrile are preferred. Lists of suitable salts are found in *Remington's Pharmaceutical Sciences*, 17th ed., Mack Publishing Company, Easton, Pa., 1985, p. 1418, *Pharmaceutical Salts: Properties, Selection, and Use*, P.H. Stahl and C.G. Wermuth (eds.), Wiley-VCH, 2008, and Berge et al., *Journal of Pharmaceutical Science*, 66, 1-19 (1977), each of which is incorporated herein by reference in its entirety. *Pharmaceutically acceptable solvate*: The term "pharmaceutically acceptable solvate," as used herein, refers to a crystalline form of a compound wherein molecules of a suitable solvent are incorporated in the crystal lattice. For example, solvates may be prepared by crystallization, recrystallization, or precipitation from a solution that includes organic solvents, water, or a mixture thereof. Examples of suitable solvents are ethanol, water (for example, mono-, di-, and tri-hydrates), *N*-methylpyrrolidinone (NMP), dimethyl sulfoxide (DMSO), *N,N'*-dimethylformamide (DMF), *N,N'*-dimethylacetamide (DMAC), 1,3-dimethyl-2-imidazolidinone (DMEU), 1,3-dimethyl-3,4,5,6-tetrahydro-2-(1H)-pyrimidinone (DMPU), acetonitrile (ACN), propylene glycol, ethyl acetate, benzyl alcohol, 2-pyrrolidone, benzyl benzoate, and the like. When water is the solvent, the solvate is referred to as a "hydrate." In some embodiments, the solvent incorporated into a solvate is of

a type or at a level that is physiologically tolerable to an organism to which the solvate is administered (e.g., in a unit dosage form of a pharmaceutical composition).

[0082] *Pharmacokinetic*: As used herein, “pharmacokinetic” refers to any one or more properties of a molecule or compound as it relates to the determination of the fate of substances administered to a living organism. Pharmacokinetics is divided into several areas including the extent and rate of absorption, distribution, metabolism and excretion. This is commonly referred to as ADME where: (A) Absorption is the process of a substance entering the blood circulation; (D) Distribution is the dispersion or dissemination of substances throughout the fluids and tissues of the body; (M) Metabolism (or Biotransformation) is the irreversible transformation of parent compounds into daughter metabolites; and (E) Excretion (or Elimination) refers to the elimination of the substances from the body. In rare cases, some drugs irreversibly accumulate in body tissue.

[0083] *Physicochemical*: As used herein, “physicochemical” means of or relating to a physical and/or chemical property.

[0084] *Positive selection*: As used herein, the term “positive selection” refers to the selection of a given entity from a group of unique entities. Such entities and groups thereof may be, for example antibodies. In some cases they may be antibody fragments or antibody fragments expressed in association with an agent capable of expressing such fragments (e.g. library members from a display library). Selection may be based on the ability of selected entities to bind to a desired target or epitope. In some embodiments, positive selection may be used with phage display libraries to identify phage particles expressing scFvs that bind to the desired target. In other embodiments, positive selection may refer to the selection of antibody candidates from among a pool of antibodies. In other cases, entities may be cells, cell lines or clones as in the selection of clones during hybridoma selection. In such cases, positive selection may refer to clonal selection based on one or more features of antibodies (e.g. specificity for one or more desired epitopes) produced by such clones. In some cases, desired epitopes in positive selection methods may comprise STn (e.g. AcSTn and/or GcSTn).

[0085] Conversely, “negative selection,” as used herein, included the same principles and examples described for positive selection, but with the distinguishing characteristic that it is used for *removal* of undesired entities from a group of unique entities.

[0086] *Preventing*: As used herein, the term “preventing” refers to partially or completely delaying onset of an infection, disease, disorder and/or condition; partially or completely delaying onset of one or more symptoms, features, or clinical manifestations of a particular

infection, disease, disorder, and/or condition; partially or completely delaying onset of one or more symptoms, features, or manifestations of a particular infection, disease, disorder, and/or condition; partially or completely delaying progression from an infection, a particular disease, disorder and/or condition; and/or decreasing the risk of developing pathology associated with the infection, the disease, disorder, and/or condition.

[0087] *Prodrug*: The present disclosure also includes prodrugs of the compounds described herein. As used herein, “prodrugs” refer to any substance, molecule or entity which is in a form predicate for that substance, molecule or entity to act as a therapeutic upon chemical or physical alteration. Prodrugs may be covalently bonded or sequestered in some way and which release or are converted into the active drug moiety prior to, upon or after administered to a mammalian subject. Prodrugs can be prepared by modifying functional groups present in the compounds in such a way that the modifications are cleaved, either in routine manipulation or *in vivo*, to the parent compounds. Prodrugs include compounds wherein hydroxyl, amino, sulfhydryl, or carboxyl groups are bonded to any group that, when administered to a mammalian subject, cleaves to form a free hydroxyl, amino, sulfhydryl, or carboxyl group respectively. Preparation and use of prodrugs is discussed in T. Higuchi and V. Stella, “Pro-drugs as Novel Delivery Systems,” Vol. 14 of the A.C.S. Symposium Series, and in *Bioreversible Carriers in Drug Design*, ed. Edward B. Roche, American Pharmaceutical Association and Pergamon Press, 1987, both of which are hereby incorporated by reference in their entirety.

[0088] *Proximal*: As used herein, the term “proximal” means situated nearer to the center or to a point or region of interest.

[0089] *Region of interaction*: As used herein, the term “region of interaction” refers to a region along any of two or more entities where such entities interact or overlap. In some embodiments, a region of interaction may comprise one or more sugar residues along a glycan chain that contacts a second glycan chain. In some embodiments, the glycan chains are branching chains from the same parent chain. In some embodiments, a region of interaction may occur between two glycan chains wherein one chain is a branching chain and the second chain is a parent chain. In the case of glycan chains, regions of interaction may comprise 1, 2, 3, 4, 5, 6, 7, 8, 9 or at least 10 sugar residues. In some embodiments, regions of interaction may also occur between glycans and proteins or between glycans and lipids.

[0090] *Residue*: As used herein, the term “residue” refers to a monomer associated with or capable of associating with a polymer. In some embodiments, residues comprise sugar

molecules including, but not limited to glucose, galactose, N-acetylglucosamine, N-acetylgalactosamine, sialic acids. In some embodiments, residues comprise amino acids.

[0091] *Sample*: As used herein, the term “sample” refers to an aliquot or portion taken from a source and/or provided for analysis or processing. In some embodiments, a sample is from a biological source such as a tissue, cell or component part (e.g. a body fluid, including but not limited to blood, mucus, lymphatic fluid, synovial fluid, cerebrospinal fluid, saliva, amniotic fluid, amniotic cord blood, urine, vaginal fluid and semen). In some embodiments, a sample may be or comprise a homogenate, lysate or extract prepared from a whole organism or a subset of its tissues, cells or component parts, or a fraction or portion thereof, including but not limited to, for example, plasma, serum, spinal fluid, lymph fluid, the external sections of the skin, respiratory, intestinal, and genitourinary tracts, tears, saliva, milk, blood cells, tumors, organs. In some embodiments, a sample comprises a medium, such as a nutrient broth or gel, which may contain cellular components, such as proteins or nucleic acid molecule. In some embodiments, a “primary” sample is an aliquot of the source. In some embodiments, a primary sample is subjected to one or more processing (e.g., separation, purification, etc.) steps to prepare a sample for analysis or other use.

[0092] *Sialyl*: As used herein, the prefix “sialyl” as well as the term “sialylated” describe compounds comprising sialic acid.

[0093] *Single-chain variable fragment*: As used herein, the term “single-chain variable fragment” or “scFv” refers to a fusion protein comprising antibody variable regions connected by a linker. In some embodiments, scFvs are utilized in conjunction with phage display methods where they may be expressed in association with a phage coat protein and used in the identification of high affinity peptides for a given antigen.

[0094] *Single unit dose*: As used herein, a “single unit dose” is a dose of any therapeutic administered in one dose/at one time/single route/single point of contact, i.e., single administration event. In some embodiments, a single unit dose is provided as a discrete dosage form (e.g., a tablet, capsule, patch, loaded syringe, vial, etc.).

[0095] *Split dose*: As used herein, a “split dose” is the division of single unit dose or total daily dose into two or more doses.

[0096] *Stable*: As used herein “stable” refers to a compound or entity that is sufficiently robust to survive isolation to a useful degree of purity from a reaction mixture, and preferably capable of formulation into an efficacious therapeutic agent.

[0097] *Stabilized*: As used herein, the term “stabilize”, “stabilized,” “stabilized region” means to make or become stable. In some embodiments, stability is measured relative to an absolute value. In some embodiments, stability is measured relative to a reference compound or entity.

[0098] *Subject*: As used herein, the term “subject” or “patient” refers to any organism to which a composition in accordance with the invention may be administered, *e.g.*, for experimental, diagnostic, prophylactic, and/or therapeutic purposes. Typical subjects include animals (*e.g.*, mammals such as mice, rats, rabbits, non-human primates, and humans) and/or plants.

[0099] *Submaxillary glands*: As used herein, the term “submaxillary glands” or “submandibular glands” refers to mucous producing glands located beneath the mouth floor. These glands are capable of producing mucins and in some embodiments, may be extracted from mammals as a source of mucin.

[00100] *Suffering from*: An individual who is “suffering from” a disease, disorder, and/or condition has been diagnosed with or displays one or more symptoms of a disease, disorder, and/or condition.

[00101] *Susceptible to*: An individual who is “susceptible to” a disease, disorder, and/or condition has not been diagnosed with and/or may not exhibit symptoms of the disease, disorder, and/or condition but harbors a propensity to develop a disease or its symptoms. In some embodiments, an individual who is susceptible to a disease, disorder, and/or condition (for example, cancer) may be characterized by one or more of the following: (1) a genetic mutation associated with development of the disease, disorder, and/or condition; (2) a genetic polymorphism associated with development of the disease, disorder, and/or condition; (3) increased and/or decreased expression and/or activity of a protein and/or nucleic acid associated with the disease, disorder, and/or condition; (4) habits and/or lifestyles associated with development of the disease, disorder, and/or condition; (5) a family history of the disease, disorder, and/or condition; and (6) exposure to and/or infection with a microbe associated with development of the disease, disorder, and/or condition. In some embodiments, an individual who is susceptible to a disease, disorder, and/or condition will develop the disease, disorder, and/or condition. In some embodiments, an individual who is susceptible to a disease, disorder, and/or condition will not develop the disease, disorder, and/or condition.

[00102] *Synthetic*: The term “synthetic” means produced, prepared, and/or manufactured by the hand of man. Synthesis of polynucleotides or polypeptides or other molecules of the present invention may be chemical or enzymatic.

[00103] *Target*: As used herein, the term “target” refers to an object or entity to be affected by an action. In some embodiments, targets refer to antigens to be used for the development of antibodies that specifically bind the antigens.

[00104] *Target screening*: As used herein, the term “target screening” refers to the use of a target substance to identify binding partners for that substance.

[00105] *Target site*: As used herein, the term “target site” refers to a target on or within one or more glycans, biomolecules and/or biostructures within a cell, the extracellular space, a tissue, an organ and/or an organism. In some embodiments, glycan target sites may reside exclusively on one sugar residue or may be formed by two or more residues. In some embodiments, target sites are formed between two or more glycans. In some embodiments, target sites are formed between branching chains of the same glycan or between one or more branching chains and a parent chain.

[00106] *Targeted Cells*: As used herein, “targeted cells” refers to any one or more cells of interest. The cells may be found *in vitro*, *in vivo*, *in situ* or in the tissue or organ of an organism. The organism may be an animal, preferably a mammal, more preferably a human and most preferably a patient.

[00107] *Terminal residue*: As used herein, the term “terminal residue” refers to the last residue in a polymeric chain. In some embodiments, terminal residues are sugar residues located at the non-reducing end of a polysaccharide chain.

[00108] *Therapeutic agent*: The term “therapeutic agent” refers to any agent that, when administered to a subject, has a therapeutic, diagnostic, and/or prophylactic effect and/or elicits a desired biological and/or pharmacological effect.

[00109] *Therapeutically effective amount*: As used herein, the term “therapeutically effective amount” means an amount of an agent to be delivered (*e.g.*, nucleic acid, drug, therapeutic agent, diagnostic agent, prophylactic agent, *etc.*) that is sufficient, when administered to a subject suffering from or susceptible to an infection, disease, disorder, and/or condition, to treat, improve symptoms of, diagnose, prevent, and/or delay the onset of the infection, disease, disorder, and/or condition. In some embodiments, a therapeutically effective amount is provided in a single dose. In some embodiments, a therapeutically effective amount is administered in a dosage regimen comprising a plurality of doses. Those

skilled in the art will appreciate that in some embodiments, a unit dosage form may be considered to comprise a therapeutically effective amount of a particular agent or entity if it comprises an amount that is effective when administered as part of such a dosage regimen.

[00110] *Therapeutically effective outcome*: As used herein, the term “therapeutically effective outcome” means an outcome that is sufficient in a subject suffering from or susceptible to an infection, disease, disorder, and/or condition, to treat, improve symptoms of, diagnose, prevent, and/or delay the onset of the infection, disease, disorder, and/or condition.

[00111] *Total daily dose*: As used herein, a “total daily dose” is an amount given or prescribed in 24 hr period. It may be administered as a single unit dose.

[00112] *Transgenic*: As used herein, the term “transgenic” refers to an organism that comprises one or more genes incorporated within the organisms genome that are not naturally found in that organism.

[00113] *Treating*: As used herein, the term “treating” refers to partially or completely alleviating, ameliorating, improving, relieving, delaying onset of, inhibiting progression of, reducing severity of, and/or reducing incidence of one or more symptoms or features of a particular infection, disease, disorder, and/or condition. For example, “treating” cancer may refer to inhibiting survival, growth, and/or spread of a tumor. Treatment may be administered to a subject who does not exhibit signs of a disease, disorder, and/or condition and/or to a subject who exhibits only early signs of a disease, disorder, and/or condition for the purpose of decreasing the risk of developing pathology associated with the disease, disorder, and/or condition.

[00114] *Variable region*: As used herein, the term “variable region” or “variable domain” refers to specific antibody domains that differ extensively in sequence among antibodies and are used in the binding and specificity of each particular antibody for its particular antigen.

[00115] *Whole IgG*: As used herein, the term “whole IgG” refers to a complete IgG molecule. In some embodiments, whole IgG molecules comprise regions found naturally in two or more other organisms.

[00116] *Wild type*: As used herein, the term “wild type” refers to an organism comprising a natural genome (free from genes derived from other organisms).

I. Compositions of the invention

[00117] The present invention provides compounds as well as compositions that comprise at least one glycan-interacting antibody. Within a glycan, monosaccharide monomers may all

be the same or they may differ. Common monomers include, but are not limited to trioses, tetroses, pentoses, glucose, fructose, galactose, xylose, arabinose, lyxose, allose, altrose, mannose, gulose, iodose, ribose, mannoheptulose, sedoheptulose and talose. Amino sugars may also be monomers within a glycan. Glycans comprising such sugars are herein referred to as aminoglycans. Amino sugars, as used herein, are sugar molecules that comprise an amine group in place of a hydroxyl group, or in some embodiments, a sugar derived from such a sugar. Examples of amino sugars include, but are not limited to glucosamine, galactosamine, N-acetylglucosamine, N-acetylgalactosamine, sialic acids (including, but not limited to, N-acetylneuraminic acid and N-glycolylneuraminic acid) and L-daunosamine.

[00118] As used herein the term “glycan-interacting antibody” refers to an antibody that can interact with a glycan moiety. Glycan-interacting antibodies may function to bind to, alter, activate, inhibit, stabilize, degrade and/or modulate a glycan or a glycan-associated molecule or entity. In so doing, glycan-interacting antibodies may function as a therapeutic, whether palliative, prophylactic or as an ongoing treatment composition. In some embodiments, glycan-interacting antibodies may comprise conjugates or combinations with other molecules. In some embodiments, glycan-interacting antibodies are directed toward glycans comprising one or more amino sugar. In a further embodiment, one or more amino sugars is a sialic acid. In a further embodiment, one or more sialic acids is N-acetylneuraminic acid and/or N-glycolylneuraminic acid.

Antibodies

[00119] Glycan-interacting antibodies may comprise entire antibodies or fragments thereof. As used herein, the term “antibody” is used in the broadest sense and specifically covers various embodiments including, but not limited to monoclonal antibodies, polyclonal antibodies, multispecific antibodies (e.g., bispecific antibodies formed from at least two intact antibodies), antibody conjugates (including, but not limited to antibody-drug conjugates), antibody variants [including, but not limited to antibody mimetics, chimeric antibodies (e.g. antibodies with amino acid sequences derived from more than one species), and synthetic variants] and antibody fragments such as diabodies so long as they exhibit a desired biological activity. Antibodies are primarily amino-acid based molecules but may also comprise one or more modifications such as with sugar moieties.

[00120] As used herein, the term “antibody fragment” refers to a portion of an intact antibody or fusion-protein thereof, in some cases comprising at least one antigen binding

region. Examples of antibody fragments include Fab, Fab', F(ab')₂, Fv fragments, single-chain variable fragments (scFvs); diabodies; tri(a)bodies; linear antibodies; single-chain antibody molecules; and multispecific antibodies formed from antibody fragments. Papain digestion of antibodies produces two identical antigen-binding fragments, called "Fab" fragments, each with a single antigen-binding site. Also produced is a residual "Fc" fragment, whose name reflects its ability to crystallize readily. Pepsin treatment yields an F(ab')₂ fragment that has two antigen-binding sites and is still capable of cross-linking antigen. Glycan-interacting antibodies may comprise one or more of these fragments.

[00121] "Native antibodies" are usually heterotetrameric glycoproteins of about 150,000 Daltons, composed of two identical light (L) chains and two identical heavy (H) chains. Genes encoding antibody heavy and light chains are known and segments making up each have been well characterized and described (Matsuda, F. et al., 1998. The Journal of Experimental Medicine. 188(11); 2151-62 and Li, A. et al., 2004. Blood. 103(12): 4602-9, the content of each of which are herein incorporated by reference in their entirety). Each light chain is linked to a heavy chain by one covalent disulfide bond, while the number of disulfide linkages varies among the heavy chains of different immunoglobulin isotypes. Each heavy and light chain also has regularly spaced intrachain disulfide bridges. Each heavy chain has at one end a variable domain (V_H) followed by a number of constant domains. Each light chain has a variable domain at one end (V_L) and a constant domain at its other end; the constant domain of the light chain is aligned with the first constant domain of the heavy chain, and the light chain variable domain is aligned with the variable domain of the heavy chain.

[00122] As used herein, the term "variable domain" refers to specific antibody domains found on both the antibody heavy and light chains that differ extensively in sequence among antibodies and are used in the binding and specificity of each particular antibody for its particular antigen. Variable domains comprise hypervariable regions. As used herein, the term "hypervariable region" refers to a region within a variable domain comprising amino acid residues responsible for antigen binding. The amino acids present within the hypervariable regions determine the structure of the complementarity determining regions (CDRs) that become part of the antigen-binding site of the antibody. As used herein, the term "CDR" refers to a region of an antibody comprising a structure that is complimentary to its target antigen or epitope. Other portions of the variable domain, not interacting with the antigen, are referred to as framework (FW) regions. The antigen-binding site (also known as the antigen combining site or paratope) comprises the amino acid residues necessary to

interact with a particular antigen. The exact residues making up the antigen-binding site are typically elucidated by co-crystallography with bound antigen, however computational assessments can also be used based on comparisons with other antibodies (Strohl, W.R. *Therapeutic Antibody Engineering*. Woodhead Publishing, Philadelphia PA. 2012. Ch. 3, p47-54, the contents of which are herein incorporated by reference in their entirety).

Determining residues making up CDRs may include the use of numbering schemes including, but not limited to, those taught by Kabat [Wu, T.T. et al., 1970, *JEM*, 132(2):211-50 and Johnson, G. et al., 2000, *Nucleic Acids Res.* 28(1): 214-8, the contents of each of which are herein incorporated by reference in their entirety], Chothia [Chothia and Lesk, *J. Mol. Biol.* 196, 901 (1987), Chothia et al., *Nature* 342, 877 (1989) and Al-Lazikani, B. et al., 1997, *J. Mol. Biol.* 273(4):927-48, the contents of each of which are herein incorporated by reference in their entirety], Lefranc (Lefranc, M.P. et al., 2005, *Immunome Res.* 1:3) and Honegger (Honegger, A. and Pluckthun, A. 2001. *J. Mol. Biol.* 309(3):657-70, the contents of which are herein incorporated by reference in their entirety).

[00123] VH and VL domains have three CDRs each. VL CDRs are referred to herein as CDR-L1, CDR-L2 and CDR-L3, in order of occurrence when moving from N- to C- terminus along the variable domain polypeptide. VH CDRs are referred to herein as CDR-H1, CDR-H2 and CDR-H3, in order of occurrence when moving from N- to C- terminus along the variable domain polypeptide. Each of CDRs have favored canonical structures with the exception of the CDR-H3, which comprises amino acid sequences that may be highly variable in sequence and length between antibodies resulting in a variety of three-dimensional structures in antigen-binding domains (Nikoloudis, D. et al., 2014. *PeerJ.* 2:e456). In some cases, CDR-H3s may be analyzed among a panel of related antibodies to assess antibody diversity. Various methods of determining CDR sequences are known in the art and may be applied to known antibody sequences (Strohl, W.R. *Therapeutic Antibody Engineering*. Woodhead Publishing, Philadelphia PA. 2012. Ch. 3, p47-54, the contents of which are herein incorporated by reference in their entirety).

[00124] As used herein, the term “Fv” refers to an antibody fragment comprising the minimum fragment on an antibody needed to form a complete antigen-binding site. These regions consist of a dimer of one heavy chain and one light chain variable domain in tight, non-covalent association. Fv fragments can be generated by proteolytic cleavage, but are largely unstable. Recombinant methods are known in the art for generating stable Fv fragments, typically through insertion of a flexible linker between the light chain variable

domain and the heavy chain variable domain [to form a single chain Fv (scFv)] or through the introduction of a disulfide bridge between heavy and light chain variable domains (Strohl, W.R. Therapeutic Antibody Engineering. Woodhead Publishing, Philadelphia PA. 2012. Ch. 3, p46-47, the contents of which are herein incorporated by reference in their entirety).

[00125] Antibody "light chains" from any vertebrate species can be assigned to one of two clearly distinct types, called kappa and lambda based on amino acid sequences of their constant domains. Depending on the amino acid sequence of the constant domain of their heavy chains, antibodies can be assigned to different classes. There are five major classes of intact antibodies: IgA, IgD, IgE, IgG, and IgM, and several of these may be further divided into subclasses (isotypes), e.g., IgG1, IgG2a, IgG2b, IgG2c, IgG3, IgG4, IgA1, and IgA2.

[00141] As used herein, the term "single chain Fv" or "scFv" refers to a fusion protein of VH and VL antibody domains, wherein these domains are linked together into a single polypeptide chain by a flexible peptide linker. In some embodiments, the Fv polypeptide linker enables the scFv to form the desired structure for antigen binding. In some embodiments, scFvs are utilized in conjunction with phage display, yeast display or other display methods where they may be expressed in association with a surface member (e.g. phage coat protein) and used in the identification of high affinity peptides for a given antigen.

[00126] The term "diabodies" refers to small antibody fragments with two antigen-binding sites, which fragments comprise a heavy chain variable domain V_H connected to a light chain variable domain V_L in the same polypeptide chain. By using a linker that is too short to allow pairing between the two domains on the same chain, the domains are forced to pair with the complementary domains of another chain and create two antigen-binding sites. Diabodies are described more fully in, for example, EP 404,097; WO 93/11161; and Hollinger et al., Proc. Natl. Acad. Sci. USA, 90:6444-6448 (1993), the contents of each of which are incorporated herein by reference in their entirety.

[00127] The term "intrabody" refers to a form of antibody that is not secreted from a cell in which it is produced, but instead target one or more intracellular protein. Intrabodies may be used to affect a multitude of cellular processes including, but not limited to intracellular trafficking, transcription, translation, metabolic processes, proliferative signaling and cell division. In some embodiments, methods of the present invention may include intrabody-based therapies. In some such embodiments, variable domain sequences and/or CDR sequences disclosed herein may be incorporated into one or more construct for intrabody-based therapy. In some cases, intrabodies of the invention may target one or more glycosylated

intracellular protein or may modulate the interaction between one or more glycosylated intracellular protein and an alternative protein.

[00128] The term “chimeric antigen receptor” or “CAR” as used herein, refers to artificial receptors (also known as “chimeric immunoreceptors,” “artificial T cell receptors” or “chimeric T cell receptors”) that are engineered to be expressed on the surface of immune effector cells resulting in specific targeting of such immune effector cells to cells expressing entities that bind with high affinity to the artificial receptors. CARs may be designed to include one or more segments of an antibody, scFv, antibody variable domain, and/or antibody CDRs, such that when such CARs are expressed on immune effector cells, the immune effector cells bind and clear any cells that are recognized by the antibody portions of the CARs. In some cases, CARs are designed to specifically bind cancer cells, leading to immune-regulated clearance of the cancer cells. The phrases "have antigen specificity" and "elicit antigen-specific response" as used with respect to CARs means that the CAR can specifically bind to and immunologically recognize an antigen to elicit an immune response.

[00129] The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous cells (or clones), i.e., the individual antibodies comprising the population are identical and/or bind the same epitope, except for possible variants that may arise during production of the monoclonal antibody, such variants generally being present in minor amounts. In contrast to polyclonal antibody preparations that typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen

[00130] The modifier "monoclonal" indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. The monoclonal antibodies herein include "chimeric" antibodies (immunoglobulins) in which a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass, as well as fragments of such antibodies.

[00131] "Humanized" forms of non-human (e.g., murine) antibodies are chimeric antibodies that contain minimal sequence derived from non-human immunoglobulin. For the most part, humanized antibodies are human immunoglobulins (recipient antibody) in which

residues from the hypervariable region from an antibody of the recipient are replaced by residues from the hypervariable region from an antibody of a non-human species (donor antibody) such as mouse, rat, rabbit or nonhuman primate having the desired specificity, affinity, and capacity.

[00132] In some embodiments, glycan-interacting antibodies of the present invention may be antibody mimetics. The term “antibody mimetic” refers to any molecule which mimics the function or effect of an antibody and which binds specifically and with high affinity to their molecular targets. In some embodiments, antibody mimetics may be monobodies, designed to incorporate the fibronectin type III domain (Fn3) as a protein scaffold (US 6,673,901; US 6,348,584). In some embodiments, antibody mimetics may be those known in the art including, but are not limited to affibody molecules, affilins, affitins, anticalins, avimers, DARPins, Fynomers and Kunitz and domain peptides. In other embodiments, antibody mimetics may include one or more non-peptide region.

[00133] As used herein, the term “antibody variant” refers to a biomolecule resembling an antibody in structure and/or function comprising some differences in their amino acid sequence, composition or structure as compared to a native antibody.

Antibody development

[00134] Glycan-interacting antibodies of the present invention are developed to bind antigens such as those described herein. As used herein, an “antigen” is an entity which induces or evokes an immune response in an organism. An immune response is characterized by the reaction of the cells, tissues and/or organs of an organism to the presence of a foreign entity. Such an immune response typically leads to the production by the organism of one or more antibodies against the foreign entity, e.g., antigen or a portion of the antigen. In some cases, methods of immunization may be altered based on one or more desired immunization outcomes. As used here, the term “immunization outcome” refers to one or more desired effects of immunization. Examples include high antibody titers and/or increased antibody specificity for a target of interest.

[00135] Antigens of the invention may comprise glycans, glycoconjugates (including, but not limited to glycoproteins and glycolipids), peptides, polypeptides, fusion proteins, or any of the foregoing and may be conjugated or complexed to one or more separate adjuvants or heterologous proteins. In some embodiments, antigens used according to methods of the present invention may comprise sialylated glycans, such as STn. Antigens comprising STn

may comprise mucins. Mucins are a family of proteins that are heavily glycosylated. They are a component of many tumors originating from epithelial cells (Ishida, A. et al., 2008. *Proteomics*. 8: 3342-9, the contents of which are herein incorporated by reference in their entirety). They are highly expressed by submaxillary glands and can be found at high levels in saliva and mucous. Animal-derived submaxillary mucins may be used as antigens to generate anti-STn antibodies in immunogenic hosts. Submaxillary mucin from different species differ in their STn content with regard to AcSTn versus GcSTn forms. Porcine submaxillary mucin (PSM) is particularly rich in GcSTn, which makes up about 90% of total STn. STn from bovine submaxillary mucin (BSM) comprises roughly equal percentages of GcSTn and AcSTn. Ovine submaxillary mucin (OSM) is particularly rich in AcSTn, which makes up about 90% of total STn. In some cases, solutions prepared for immunization may be modified to include one or more of PSM, BSM and OSM depending on the desired target of antibodies resulting from such immunization. PSM may be used in immunizations to generate antibodies in immunogenic hosts that are more likely to be specific for GcSTn. PSM is rich in Neu5Gc-containing mucin-type, glycoproteins that are decorated with GcSTn. Among the currently known sources of high Neu5Gc content is red meat; especially submaxillary glands were previously described as a rich source of Neu5Gc due to the high expression of the CMAH enzyme, which catalyzes the reaction to produce the Neu5Gc precursor, CMP-Neu5Ac. In some cases, PSM may be used to prevent a pan-anti-Neu5Gc response and induce a more specific immune response against GcSTn. OSM may be used in immunizations to generate antibodies in immunogenic hosts that are more likely to be specific for AcSTn.

[00136] In one embodiment, the present invention provides a glycan-interacting antibody that is GcSTn-specific. The antibody has little cross-reactivity to Neu5Ac-STn or Tn. The antibody can bind GcSTn but has reduced affinity for AcSTn.

[00137] In some embodiments, antigens may be subjected to enzymatic digestion prior to immunization to modulate the resulting immune response in immunogenic hosts. In one example, submaxillary mucins may be treated with trypsin or proteinase K enzymes prior to immunization. The activity of such enzymes may help to cleave off and thereby reduce the percentage and variability of non-STn epitopes. Glycan moieties may shield regions of the peptide where they are attached from enzymatic proteolysis and thereby remain intact.

Antibody titers resulting from immunizations may comprise different levels depending on the type and amount of antigen used in such immunizations. In some cases, certain antigens may be selected for use in immunizations based on the expected titer.

[00138] As used herein, an “adjuvant” is a pharmacological or immunological agent that modifies the effect of other agents. Adjuvants according to the present invention include, but are not limited to chemical compositions, biomolecules, therapeutics, and/or therapeutic regimens. Adjuvants may include Freund’s adjuvant (complete and/or incomplete), immunostimulatory oligonucleotides [e.g. CpG oligodeoxynucleotides (ODNs)], mineral-containing compositions, bacterial ADP-ribosylating toxins, bioadhesives, mucoadhesives, microparticles, lipids, liposomes, muramyl peptides, N-oxidized polyethylene-piperazine derivatives, saponins and/or immune stimulating complexes (ISCOs). In some embodiments, adjuvants may comprise oil-in-water emulsions (e.g. sub-micron oil-in-water emulsions). Adjuvants according to the present invention may also include any of those disclosed in US Patent Publication No. US20120027813 and/or US Patent No. US8506966, the contents of each of which are herein incorporated by reference in their entirety.

[00139] Antibodies of the present invention may be polyclonal or monoclonal or recombinant, produced by methods known in the art or as described in this application. In some embodiments, the antibodies of the present invention may be labeled for purposes of detection with a detectable label known by one of skill in the art. The label can be a radioisotope, fluorescent compound, chemiluminescent compound, enzyme, or enzyme co-factor, or any other labels known in the art. In some aspects, the antibody that binds to a desired antigen is not labeled, but may be detected by binding of a labeled secondary antibody that specifically binds to the primary antibody.

[00140] Antibodies of the present invention (e.g., glycan-interacting antibodies) include, but are not limited to, polyclonal, monoclonal, multispecific, human, humanized or chimeric antibodies, single chain antibodies, Fab fragments, F(ab') fragments, fragments produced by a Fab expression library, anti-idiotypic (anti-Id) antibodies (including, e.g., anti-Id antibodies to antibodies of the invention), intracellularly made antibodies (i.e., intrabodies), and epitope-binding fragments of any of the above. Antibodies of the present invention (e.g., glycan-interacting antibodies) can be from any animal origin including birds and mammals. Preferably, such antibodies are of human, murine (e.g., mouse and rat), donkey, sheep, rabbit, goat, guinea pig, camel, horse, or chicken origin. The antibodies of the present invention can be monospecific or multispecific (e.g., bispecific, trispecific, or of greater multispecificity).

Multispecific antibodies can be specific for different epitopes of a target antigen of the present invention, or can be specific for both a target antigen of the present invention, and a heterologous epitope, such as a heterologous glycan, peptide or solid support material. (See, e.g., WO 93/17715; WO 92/08802; WO 91/00360; WO 92/05793; Tutt, A. et al., *Trispecific F(ab')₃ derivatives that use cooperative signaling via the TCR/CD3 complex and CD2 to activate and redirect resting cytotoxic T cells*. J Immunol. 1991 Jul 1;147(1):60-9; U.S. Pat. Nos. 4,474,893; 4,714,681; 4,925,648; 5,573,920; 5,601,819; and Kostelny, S.A. et al., *Formation of a bispecific antibody by the use of leucine zippers*. J Immunol. 1992 Mar 1;148(5):1547-53).

[00141] Glycan-interacting antibodies of the present invention comprising monoclonal antibodies can be prepared using well-established methods known by those skilled in the art. In one embodiment, the monoclonal antibodies are prepared using hybridoma technology (Kohler, G. et al., *Continuous cultures of fused cells secreting antibody of predefined specificity*. Nature. 1975 Aug 7;256(5517):495-7). For hybridoma formations, first, a mouse, hamster, or other appropriate host animal, is typically immunized with an immunizing agent (e.g., a target antigen of the invention) to elicit lymphocytes that produce or are capable of producing antibodies that will specifically bind to the immunizing agent. Alternatively, the lymphocytes may be immunized in vitro. The lymphocytes are then fused with an immortalized cell line using a suitable fusing agent, such as polyethylene glycol, to form a hybridoma cell (Goding, J.W., *Monoclonal Antibodies: Principles and Practice*. Academic Press. 1986; 59-1031). Immortalized cell lines are usually transformed mammalian cells, particularly myeloma cells of rodent, rabbit, bovine and human origin. Usually, rat or mouse myeloma cell lines are employed. The hybridoma cells may be cultured in a suitable culture medium that preferably contains one or more substances that inhibit the growth or survival of the unfused, immortalized cells. For example, if the parental cells lack the enzyme hypoxanthine guanine phosphoribosyl transferase (HGPRT or HPRT), the culture medium for the hybridomas typically will include hypoxanthine, aminopterin, and thymidine ("HAT medium"), which substances prevent the growth of HGPRT-deficient cells.

[00142] Preferred immortalized cell lines are those that fuse efficiently, support stable high level expression of antibody by the selected antibody-producing cells, and are sensitive to a medium such as HAT medium. More preferred immortalized cell lines are murine myeloma lines, which can be obtained, for instance, from the Salk Institute Cell Distribution Center, San Diego, Calif. and the American Type Culture Collection, Manassas, Va. Human

myeloma and mouse-human heteromyeloma cell lines also have been described for the production of human monoclonal antibodies (Kozbor, D. et al., *A human hybrid myeloma for production of human monoclonal antibodies*. J Immunol. 1984 Dec;133(6):3001-5; Brodeur, B. et al., *Monoclonal Antibody Production Techniques and Applications*. Marcel Dekker, Inc., New York. 1987; 33:51-63).

[00143] In some embodiments, myeloma cells may be subjected to genetic manipulation. Such manipulation may be carried out using zinc-finger nuclease (ZFN) mutagenesis as described herein. Alternatively, transfection methods known in the art may be used. NS0 myeloma cells or other mouse myeloma cell lines may be used. For example, Sp2/0-Ag14 can be an alternative cell line for hybridoma development.

[00144] Transcription Activator-Like Effector Nucleases (TALENs)–induced gene editing provides an alternative gene knock out method. TALENs are artificial restriction enzymes generated by fusing the TAL effector DNA binding domain to a DNA cleavage domain. Similar to ZFNs, TALENs induce double-strand breaks at desired loci that can be repaired by error-prone NHEJ to yield insertions/deletions at the break sites (Wood, A.J. et al., Targeted genome editing across species using ZFNs and TALENs. Science. 2011 Jul 15;333(6040):307). Collectis BioResearch (Cambridge, MA) provides the service of TALEN design and plasmid construction. The culture medium in which the hybridoma cells are cultured can then be assayed for the presence of monoclonal antibodies. Preferably, the binding specificity (i.e., specific immunoreactivity) of monoclonal antibodies produced by the hybridoma cells is determined by immunoprecipitation or by an in vitro binding assay, such as radioimmunoassay (RIA) or enzyme-linked immunosorbent assay (ELISA). Such techniques and assays are known by those skilled in the art. The binding specificity of the monoclonal antibody can, for example, be determined by Scatchard analysis (Munson, P.J. et al., *Ligand: a versatile computerized approach for characterization of ligand-binding systems*. Anal Biochem. 1980 Sep 1;107(1):220-39). In some cases, antibody specificity for regions of a given antigen may be characterized by chemically modifying the antigens prior to assaying for antibody binding. In one example, periodate treatment may be used to destroy the C6 side chain of sialic acids. Assays may be conducted with and without periodate treatment to reveal whether or not binding in untreated samples is sialic acid-specific. In some cases, antigens comprising 9-O-acetylated sialic acid may be subjected to mild base treatment (e.g. with 0.1 M NaOH) to destroy 9-O-acetyl groups. Assays may be conducted

with and without mild base treatment to reveal whether or not binding in untreated samples depends on 9-O-acetylation of sialic acid.

[00145] After the desired hybridoma cells are identified, the clones may be subcloned by limiting dilution procedures and grown by standard methods. Suitable culture media for this purpose include, for example, Dulbecco's Modified Eagle's Medium or RPMI-1640 medium. Alternatively, the hybridoma cells may be grown in vivo as ascites in a mammal.

[00146] Alternative methods to clone hybridomas may include those provided by kits from STEMCELL Technologies (Vancouver, BC, Canada), e.g. CLONACELLTM-HY kit, containing methylcellulose-based semi-solid medium and other media and reagents, to support the selection and growth of hybridoma clones. However, the media in this kit contain FCS, which provides an exogenous source for Neu5Gc incorporation. Though the machinery for endogenous Neu5Gc synthesis is destroyed in *Cmah*^{-/-} hybridoma, Neu5Gc incorporated from the culture media may also pose a problem in some cases (Bardor, M. et al., Mechanism of uptake and incorporation of the non-human sialic acid N-glycolylneuraminic acid into human cells. J Biol Chem. 2005. 280: 4228-4237). In such instances, the culture media may be supplemented with Neu5Ac to eliminate Neu5Gc incorporation by metabolic competition (Ghaderi, D. et al., Implications of the presence of N-glycolylneuraminic acid in recombinant therapeutic glycoproteins. Nat Biotechnol. 2010. 28: 863-867).

[00147] The monoclonal antibodies secreted by the subclones may be isolated or purified from the culture medium or ascites fluid by conventional immunoglobulin purification procedures such as, for example, protein A-Sepharose, hydroxylapatite chromatography, gel electrophoresis, dialysis, or affinity chromatography.

[00148] In another embodiment, the monoclonal antibodies of the present invention can also be made by recombinant DNA methods, such as those described in U.S. Pat. No. 4,816,567, which is hereby incorporated by reference in its entirety. DNA encoding the monoclonal antibodies of the invention can be readily isolated and sequenced using conventional procedures (e.g., by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of murine antibodies). The hybridoma cells of the invention serve as a preferred source of DNA. Once isolated, the DNA can be placed into expression vectors, which are then transfected into host cells such as simian COS cells, Chinese hamster ovary (CHO) cells, or myeloma cells that do not otherwise produce immunoglobulin protein, to obtain the synthesis of monoclonal antibodies in the recombinant host cells. The DNA also can be modified, for example, by substituting

the coding sequence for human heavy and light chain constant domains in place of the homologous murine sequences (U.S. Pat. No. 4,816,567) or by covalently joining to the immunoglobulin coding sequence all or part of the coding sequence for a non-immunoglobulin polypeptide. Such a non-immunoglobulin polypeptide can be substituted for the constant domains of an antibody of the invention, or can be substituted for the variable domains of one antigen-combining site of an antibody of the invention to create a chimeric bivalent antibody.

[00149] In some embodiments, antibodies of the present invention (e.g., glycan-interacting antibodies) may be produced by various procedures known by those skilled in the art. For the production of polyclonal antibodies in vivo, host animals, such as rabbits, rats, mice, cows, horses, donkeys, chickens, monkeys, sheep or goats, are immunized with either free or carrier-coupled antigens, for example, by intraperitoneal and/or intradermal injection. In some embodiments, injection material may be an emulsion containing about 100 µg of antigen or carrier protein. In some embodiments, injection materials comprise a glycan-rich composition such as non-human mammalian submaxillary mucin in solution. Various adjuvants can also be used to increase the immunological response, depending on the host species. Adjuvants include, but are not limited to, Freund's (complete and incomplete), mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, TITERMAX® (CytRx Corp, Los Angeles, CA), keyhole limpet hemocyanins, dinitrophenol, and potentially useful human adjuvants such as BCG (bacille Calmette-Guerin) and corynebacterium parvum. Such adjuvants are also well known in the art. Several booster injections may be needed, for instance, at intervals of about two weeks, to provide a useful titer of antibody which can be detected, for example, by ELISA assay using glycans and/or free peptide adsorbed to a solid surface. The titer of antibodies in serum from an immunized animal can be increased by selection of antibodies, e.g., by adsorption of antigens onto a solid support and elution of the selected antibodies according to methods well known in the art.

[00150] Glycan-interacting antibodies, variants and fragments thereof may be selected and produced using high throughput methods of discovery. In one embodiment, glycan-interacting antibodies comprising synthetic antibodies, variants and fragments thereof are produced through the use of display libraries. The term “display” as used herein, refers to the expression or “display” of proteins or peptides on the surface of a given host. The term “library” as used herein, refers to a collection of unique cDNA sequences and/or the proteins

that are encoded by them. A library may contain from as little as two unique cDNAs to hundreds of billions of unique cDNAs. In a preferred embodiment, glycan-interacting antibodies comprising synthetic antibodies are produced using antibody display libraries or antibody fragment display libraries. The term “antibody fragment display library” as used herein, refers to a display library wherein each member encodes an antibody fragment containing at least one variable region of an antibody. Such antibody fragments are preferably Fab fragments, but other antibody fragments such as single-chain variable fragments (scFvs) are contemplated as well. In an Fab antibody fragment library, each Fab encoded may be identical except for the amino acid sequence contained within the variable loops of the complementarity determining regions (CDRs) of the Fab fragment. In an alternative or additional embodiment, amino acid sequences within the individual V_H and/or V_L regions may differ as well.

[00151] Display libraries may be expressed in a number of possible hosts including, but not limited to yeast, bacteriophage, bacteria and retroviruses. Additional display technologies that may be used include ribosome-display, microbead-display and protein-DNA linkage techniques. In a preferred embodiment, Fab display libraries are expressed in yeast or in bacteriophages (also referred to herein as “phages” or “phage particles”. When expressed, the Fabs decorate the surface of the phage or yeast where they can interact with a given antigen. An antigen comprising a glycan or other antigen from a desired target may be used to select phage particles or yeast cells expressing antibody fragments with the highest affinity for that antigen. The DNA sequence encoding the CDR of the bound antibody fragment can then be determined through sequencing using the bound particle or cell. In one embodiment, positive selection is used in the development of antibodies. In some embodiments, negative selection is utilized in the development of antibodies. In some embodiments, both positive and negative selection methods are utilized during multiple rounds of selection in the development of antibodies using display libraries.

[00152] In yeast display, cDNA encoding different antibody fragments are introduced into yeast cells where they are expressed and the antibody fragments are “displayed” on the cell surface as described by Chao et al. (Chao, G. et al., *Isolating and engineering human antibodies using yeast surface display*. Nat Protoc. 2006;1(2):755-68). In yeast surface display, expressed antibody fragments contain an additional domain comprising the yeast agglutinin protein, Aga2p. This domain allows the antibody fragment fusion protein to attach to the outer surface of the yeast cell through the formation of disulphide bonds with surface-

expressed Aga1p. The result is a yeast cell, coated in a particular antibody fragment. Display libraries of cDNA encoding these antibody fragments are utilized initially in which the antibody fragments each have a unique sequence. These fusion proteins are expressed on the cell surface of millions of yeast cells where they can interact with a desired antigenic target antigen, incubated with the cells. Target antigens may be covalently or otherwise modified with a chemical or magnetic group to allow for efficient cell sorting after successful binding with a suitable antibody fragment takes place. Recovery may be by way of magnetic-activated cell sorting (MACS), fluorescence-activated cell sorting (FACS) or other cell sorting methods known in the art. Once a subpopulation of yeast cells is selected, the corresponding plasmids may be analyzed to determine the CDR sequence.

[00153] Bacteriophage display technology typically utilizes filamentous phage including, but not limited to fd, F1 and M13 virions. Such strains are non-lytic, allowing for continued propagation of the host and increased viral titres. Examples of phage display methods that can be used to make the antibodies of the present invention include those disclosed in Miersch et al. (Miersch, S. et al., *Synthetic antibodies: Concepts, potential and practical considerations*. Methods. 2012 Aug; 57(4):486-98), Bradbury et al. (Bradbury, A.R. et al., *Beyond natural antibodies: the power of in vitro display technologies*. Nat Biotechnol. 2011 Mar;29(3):245-54), Brinkman et al. (Brinkmann, U. et al., *Phage display of disulfide-stabilized Fv fragments*. J Immunol Methods. 1995 May 11; 182(1):41-50); Ames et al. (Ames, R.S. et al., *Conversion of murine Fabs isolated from a combinatorial phage display library to full length immunoglobulins*. J Immunol Methods. 1995 Aug 18;184(2):177-86); Kettleborough et al. (Kettleborough, C.A. et al., *Isolation of tumor cell-specific single-chain Fv from immunized mice using phage-antibody libraries and the re-construction of whole antibodies from these antibody fragments*. Eur J Immunol. 1994 Apr; 24(4):952-8); Persic et al. (Persic, L. et al., *An integrated vector system for the eukaryotic expression of antibodies or their fragments after selection from phage display libraries*. Gene. 1997 Mar 10; 187(1):9-18); PCT application No. PCT/GB91/01134; PCT publications WO 90/02809; WO 91/10737; WO 92/01047; WO 92/18619; WO 93/11236; WO 95/15982; WO 95/20401; and U.S. Pat. Nos. 5,698,426; 5,223,409; 5,403,484; 5,580,717; 5,427,908; 5,750,753; 5,821,047; 5,571,698; 5,427,908; 5,516,637; 5,780,225; 5,658,727; 5,733,743 and 5,969,108, each of which is incorporated herein by reference in its entirety. Antibody fragment expression on bacteriophages may be carried out by inserting the cDNA encoding the fragment into the gene expressing a viral coat protein. The viral coat of filamentous bacteriophages is made up

of five coat proteins, encoded by a single-stranded genome. Coat protein pIII is the preferred protein for antibody fragment expression, typically at the N-terminus. If antibody fragment expression compromises the function of pIII, viral function may be restored through coexpression of a wild type pIII, although such expression will reduce the number of antibody fragments expressed on the viral coat, but may enhance access to the antibody fragment by the target antigen. Expression of viral as well as antibody fragment proteins may alternatively be encoded on multiple plasmids. This method may be used to reduce the overall size of infective plasmids and enhance the transformation efficiency.

[00154] As described above, after selection of a host expressing a high affinity antibody or antibody fragment, (e.g., glycan-interacting antibodies) the coding regions from the antibody or antibody fragment can be isolated and used to generate whole antibodies, including human antibodies, or any other desired antigen binding fragment, and expressed in any desired host, including mammalian cells, insect cells, plant cells, yeast, and bacteria, e.g., as described in detail below.

[00155] The DNA sequence encoding a high affinity antibody can be mutated for additional rounds of selection in a process known as affinity maturation. The term “affinity maturation”, as used herein, refers to a method whereby antibodies are produced with increasing affinity for a given antigen through successive rounds of mutation and selection of antibody- or antibody fragment-encoding cDNA sequences. In some cases, this process is carried out in vitro. To accomplish this, amplification of CDR coding sequences may be carried out using error-prone PCR to produce millions of copies containing mutations including, but not limited to point mutations, regional mutations, insertional mutations and deletional mutations. As used herein, the term “point mutation” refers to a nucleic acid mutation in which one nucleotide within a nucleotide sequence is changed to a different nucleotide. As used herein, the term “regional mutation” refers to a nucleic acid mutation in which two or more consecutive nucleotides are changed to different nucleotides. As used herein, the term “insertional mutation” refers to a nucleic acid mutation in which one or more nucleotides are inserted into a nucleotide sequence. As used herein, the term “deletional mutation” refers to a nucleic acid mutation in which one or more nucleotides are removed from a nucleotide sequence. Insertional or deletional mutations may include the complete replacement of an entire codon or the change of one codon to another by altering one or two nucleotides of the starting codon.

[00156] Mutagenesis may be carried out on CDR-encoding cDNA sequences to create millions of mutants with singular mutations in CDR heavy and light chain regions. In another approach, random mutations are introduced only at CDR residues most likely to improve affinity. These newly generated mutagenic libraries can be used to repeat the process to screen for clones that encode antibody fragments with even higher affinity for the target antigen. Continued rounds of mutation and selection promote the synthesis of clones with greater and greater affinity (Chao, G. et al., *Isolating and engineering human antibodies using yeast surface display*. Nat Protoc. 2006;1(2):755-68).

[00157] Examples of techniques that can be used to produce antibodies and antibody fragments, such as Fabs and scFvs, include those described in U.S. Pat. Nos. 4,946,778 and 5,258,498; Miersch et al. (Miersch, S. et al., *Synthetic antibodies: Concepts, potential and practical considerations*. Methods. 2012 Aug;57(4):486-98), Chao et al. (Chao, G. et al., *Isolating and engineering human antibodies using yeast surface display*. Nat Protoc. 2006;1(2):755-68), Huston et al. (Huston, J.S. et al., *Protein engineering of single-chain Fv analogs and fusion proteins*. Methods Enzymol. 1991;203:46-88); Shu et al. (Shu, L. et al., *Secretion of a single-gene-encoded immunoglobulin from myeloma cells*. Proc Natl Acad Sci U S A. 1993 Sep 1;90(17):7995-9); and Skerra et al. (Skerra, A. et al., *Assembly of a functional immunoglobulin Fv fragment in Escherichia coli*. Science. 1988 May 20;240(4855):1038-41), each of which is incorporated herein by reference in its entirety.

[00158] For some uses, including the *in vivo* use of antibodies (e.g., glycan-interacting antibodies) in humans and *in vitro* detection assays, it may be preferable to use chimeric, humanized, or human antibodies. A chimeric antibody is a molecule in which different portions of the antibody are derived from different animal species, such as antibodies having a variable region derived from a murine monoclonal immunoglobulin and a human immunoglobulin constant region. Methods for producing chimeric antibodies are known in the art. (Morrison, S.L., *Transfectomas provide novel chimeric antibodies*. Science. 1985 Sep 20;229(4719):1202-7; Gillies, S.D. et al., *High-level expression of chimeric antibodies using adapted cDNA variable region cassettes*. J Immunol Methods. 1989 Dec 20;125(1-2):191-202.; and U.S. Pat. Nos. 5,807,715; 4,816,567; and 4,816,397, which are incorporated herein by reference in their entirety).

[00159] Humanized antibodies are antibody molecules from non-human species that bind to the desired antigen and have one or more complementarity determining regions (CDRs) from the nonhuman species and framework regions from a human immunoglobulin molecule.

Often, framework residues in the human framework regions are substituted with corresponding residues from the CDR and framework regions of the donor antibody to alter, preferably improve, antigen binding. These framework substitutions are identified by methods well known in the art, e.g., by modeling of the interactions of the CDR and framework residues to identify framework residues important for antigen binding, and by sequence comparison to identify unusual framework residues at particular positions. (U.S. Pat. Nos. 5,693,762 and 5,585,089; Riechmann, L. et al., *Reshaping human antibodies for therapy*. Nature. 1988 Mar 24;332(6162):323-7, which are incorporated herein by reference in their entirety). Antibodies can be humanized using a variety of techniques known in the art, including, for example, CDR-grafting (EP 239,400; PCT publication WO 91/09967; U.S. Pat. Nos. 5,225,539; 5,530,101; and 5,585,089); veneering or resurfacing (EP 592,106; EP 519,596; Padlan, E.A., *A possible procedure for reducing the immunogenicity of antibody variable domains while preserving their ligand-binding properties*. Mol Immunol. 1991 Apr-May;28(4-5):489-98; Studnicka, G.M. et al., *Human-engineered monoclonal antibodies retain full specific binding activity by preserving non-CDR complementarity-modulating residues*. Protein Eng. 1994 Jun;7(6):805-14; Roguska, M.A. et al., *Humanization of murine monoclonal antibodies through variable domain resurfacing*. Proc Natl Acad Sci U S A. 1994 Feb 1;91(3):969-73); and chain shuffling (U.S. Pat. No. 5,565,332); each of which is incorporated herein by reference in their entirety. Humanized antibodies of the present invention may be developed for desired binding specificity, complement-dependent cytotoxicity, and antibody-dependent cellular-mediated cytotoxicity, etc.

[00160] Completely human antibodies (e.g., glycan-interacting antibodies) are particularly desirable for therapeutic treatment of human patients, so as to avoid or alleviate immune reaction to foreign protein. Human antibodies can be made by a variety of methods known in the art, including the antibody display methods described above, using antibody libraries derived from human immunoglobulin sequences. See also, U.S. Pat. Nos. 4,444,887 and 4,716,111; and PCT publications WO 98/46645, WO 98/50433, WO 98/24893, WO 98/16654, WO 96/34096, WO 96/33735, and WO 91/10741; each of which is incorporated herein by reference in its entirety.

[00161] Human antibodies (e.g., glycan-interacting antibodies) can also be produced using transgenic mice which are incapable of expressing functional endogenous immunoglobulins, but which can express human immunoglobulin polynucleotides. For example, the human heavy and light chain immunoglobulin polynucleotide complexes can be introduced

randomly, or by homologous recombination, into mouse embryonic stem cells. Alternatively, the human variable region, constant region, and diversity region may be introduced into mouse embryonic stem cells, in addition to the human heavy and light chain polynucleotides. The mouse heavy and light chain immunoglobulin polynucleotides can be rendered nonfunctional separately or simultaneously with the introduction of human immunoglobulin loci by homologous recombination. In particular, homozygous deletion of the J_H region prevents endogenous antibody production. The modified embryonic stem cells are expanded and microinjected into blastocysts to produce chimeric mice. The chimeric mice are then bred to produce homozygous offspring which express human antibodies. The transgenic mice are immunized in the normal fashion with a selected antigen, e.g., all or a portion of a glycan, glycoconjugate and/or polypeptide of the invention.

[00162] Thus, using such a technique, it is possible to produce useful human IgG, IgA, IgM, IgD and IgE antibodies. For an overview of the technology for producing human antibodies, see Lonberg and Huszar (Lonberg, N. et al., *Human antibodies from transgenic mice*. Int Rev Immunol. 1995;13(1):65-93). For a detailed discussion of the technology for producing human antibodies and human monoclonal antibodies and protocols for producing such antibodies, see, e.g., PCT publications WO 98/24893; WO 92/01047; WO 96/34096; WO 96/33735; U.S. Pat. Nos. 5,413,923; 5,625,126; 5,633,425; 5,569,825; 5,661,016; 5,545,806; 5,814,318; 5,885,793; 5,916,771; 5,939,598; 6,075,181; and 6,114,598, each of which are incorporated by reference herein in their entirety. In addition, companies such as Abgenix, Inc. (Fremont, Calif.), Protein Design Labs, Inc. (Mountain View, Calif.) and Genpharm (San Jose, Calif.) can be engaged to provide human antibodies directed against a selected antigen using technology similar to the above described technologies.

[00163] Once an antibody molecule of the present invention has been produced by an animal, a cell line, chemically synthesized, or recombinantly expressed, it can be purified (i.e., isolated) by any method known in the art for the purification of an immunoglobulin or polypeptide molecule, for example, by chromatography (e.g., ion exchange, affinity, particularly by affinity for the specific antigen, Protein A, and sizing column chromatography), centrifugation, differential solubility, or by any other standard technique for the purification of proteins. In addition, the antibodies of the present invention or fragments thereof can be fused to heterologous polypeptide sequences described herein or otherwise known in the art, to facilitate purification.

[00164] The affinity between an antibody and a target or ligand (such as an antigen used to generate a given antibody) may be measured in terms of K_D using one or more binding assays as described herein. Depending on the desired application for a given antibody, varying K_D values may be desirable. High affinity antibodies typically form ligand bonds with a K_D of about 10^{-5} M or less, e.g. about 10^{-6} M or less, about 10^{-7} M or less, about 10^{-8} M or less, about 10^{-9} M or less, about 10^{-10} M or less, about 10^{-11} M or less or about 10^{-12} M or less.

[00165] In some embodiments, antibodies of the invention may be characterized according to their half maximal effective or inhibitory concentration (EC_{50} or IC_{50} , respectively). In some cases, this value may represent the concentration of antibody necessary to inhibit cells expressing STn (e.g. kill, reduce proliferation and/or reduce one or more cell function) at a level equal to half of the maximum inhibition observed with the highest concentrations of antibody. Such IC_{50} values may be from about 0.001 nM to about 0.01 nM, from about 0.005 nM to about 0.05 nM, from about 0.01 nM to about 1 nM, from about 0.05 nM to about 5 nM, from about 0.1 nM to about 10 nM, from about 0.5 nM to about 25 nM, from about 1 nM to about 50 nM, from about 5 nM to about 75 nM, from about 10 nM to about 100 nM, from about 25 nM to about 250 nM, from about 200 nM to about 1000 nM or more than 1000 nM.

[00166] The preparation of antibodies, whether monoclonal or polyclonal, is known in the art. Techniques for the production of antibodies are well known in the art and described, e.g. in Harlow and Lane "Antibodies, A Laboratory Manual", Cold Spring Harbor Laboratory Press, 1988 and Harlow and Lane "Using Antibodies: A Laboratory Manual" Cold Spring Harbor Laboratory Press, 1999.

Targets

[00167] Glycan-interacting antibodies of the present invention exert their effects via binding (reversibly or irreversibly) to one or more glycan or glycan-associated or glycan-related targets. In some embodiments, glycan-interacting antibodies can be prepared from any region of the targets taught herein. In some embodiments, targets of the present invention comprise glycans. Glycans used for generating antibodies may comprise a chain of sugars comprising at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at least 17, at least 18, at least 19 or at least 20 residues. Preferably, glycans used for generating antibodies comprise from about 2 residue to about 5 residues.

[00168] In some embodiments, glycan-interacting antibody target antigens comprise sialic acids. N-acetylneuraminic acid (Neu5Ac) and N-glycolylneuraminic acid (Neu5Gc) are the major sialic acids on mammalian cell surfaces. Of these, Neu5Ac is naturally produced in humans. Neu5Gc is naturally produced in most mammals with the exception of humans due to a mutation in the cytidine monophosphate (CMP)-N-acetylneuraminic acid hydroxylase (CMAH) gene responsible for CMP-Neu5Gc production from CMP-Neu5Ac. Neu5Gc in humans is in fact immunogenic with nearly all humans expressing anti-Neu5Gc antibodies. Despite a lack of production, most human systems comprise some level of Neu5Gc due to dietary intake. These foreign products are subsequently incorporated into human glycoproteins. Such glycoproteins are contemplated as targets of the invention. Glycan target antigens of the present invention, include, but are not limited to those listed in Table 1.

Table 1. Glycan target antigens

Glycan
GalNAc α -R
Gal α 1,3Gal β 1,4GlcNAc β -R
Gal β 1,3GalNAc β -R
Gal β 1,3GlcNAc α -R
Gal β 1,3GlcNAc β 1,3Gal β 1,4Glc β -R
Gal β 1,3GlcNAc β -R
Gal β 1,4GlcNAc6S β -R
Gal β 1,4GlcNAc β -R
Gal β 1,4Glc β -R
KDN α 2,8Neu5Ac α 2,3Gal β 1,4Glc β -R
KDN α 2,8Neu5Gc α 2,3Gal β 1,4Glc β -R
Neu5,9Ac2 α 2,3Gal β 1,3GalNAc α -R
Neu5,9Ac2 α 2,3Gal β 1,3GalNAc β -R
Neu5,9Ac2 α 2,3Gal β 1,3GlcNAc β -R
Neu5,9Ac2 α 2,3Gal β 1,4GlcNAc β -R
Neu5,9Ac2 α 2,3Gal β 1,4Glc β -R
Neu5,9Ac2 α 2,3Gal β -R
Neu5,9Ac2 α 2,6GalNAc α -R
Neu5,9Ac2 α 2,6Gal β 1,4GlcNAc β -R
Neu5,9Ac2 α 2,6Gal β 1,4Glc β -R
Neu5,9Ac2 α 2,6Gal β -R
Neu5Ac α 2,3Gal β 1,3GalNAc α -R
Neu5Ac α 2,3Gal β 1,3GalNAc β -R
Neu5Ac α 2,3Gal β 1,3GlcNAc β 1,3Gal β 1,4Glc β -R
Neu5Ac α 2,3Gal β 1,3GlcNAc β -R
Neu5Ac α 2,3Gal β 1,4(Fuc α 1,3)GlcNAc6S β -R
Neu5Ac α 2,3Gal β 1,4(Fuc α 1,3)GlcNAc β -R
Neu5Ac α 2,3Gal β 1,4GlcNAc6S β -R
Neu5Ac α 2,3Gal β 1,4GlcNAc β -R
Neu5Ac α 2,3Gal β 1,4Glc β -R
Neu5Ac α 2,3Gal β -R

Neu5Ac α 2,6(KDN α 2,3)Gal β 1,4Glc β -R
Neu5Ac α 2,6(Neu5Ac α 2,3)Gal β 1,4Glc β -R
Neu5Ac α 2,6(Neu5Gc α 2,3)Gal β 1,4Glc β -R
Neu5Ac α 2,6GalNAc α -R
Neu5Ac α 2,6Gal β 1,4GlcNAc β -R
Neu5Ac α 2,6Gal β 1,4Glc β -R
Neu5Ac α 2,6Gal β -R
Neu5Ac α 2,8KDN α 2,6Gal β 1,4Glc β -R
Neu5Ac α 2,8Neu5Ac α 2,3Gal β 1,4Glc β -R
Neu5Ac α 2,8Neu5Ac α 2,3Gal β 1,4Glc β -R
Neu5Ac α 2,8Neu5Ac α 2,6Gal β 1,4Glc β -R
Neu5Ac α 2,8Neu5Ac α 2,8Neu5Ac α 2,3Gal β 1,4Glc β -R
Neu5Ac α 2,8Neu5Ac α 2,8Neu5Ac α 2,3Gal β 1,4Glc β -R
Neu5Ac α 2,8Neu5Gc α 2,3Gal β 1,4Glc β -R
Neu5Ac α 2,8Neu5Gc α 2,6Gal β 1,4Glc β -R
Neu5Gc9Ac α 2,3Gal β 1,4Glc β -R
Neu5Gc9Ac α 2,6Gal β 1,4Glc β -R
Neu5Gc9Ac α 2,3Gal β 1,3GalNAc α -R
Neu5Gc9Ac α 2,3Gal β 1,3GalNAc β -R
Neu5Gc9Ac α 2,3Gal β 1,3GlcNAc β -R
Neu5Gc9Ac α 2,3Gal β 1,4GlcNAc β -R
Neu5Gc9Ac α 2,3Gal β -R
Neu5Gc9Ac α 2,6GalNAc α -R
Neu5Gc9Ac α 2,6Gal β 1,4GlcNAc β -R
Neu5Gc9Ac α 2,6Gal β -R
Neu5GcOMe α 2,8Neu5Ac α 2,3Gal β 1,4Glc β -R
Neu5Gc α 2,3Gal β 1,3GalNAc α -R
Neu5Gc α 2,3Gal β 1,3GalNAc β -R
Neu5Gc α 2,3Gal β 1,3GlcNAc β 1,3Gal β 1,4Glc β -R
Neu5Gc α 2,3Gal β 1,3GlcNAc β -R
Neu5Gc α 2,3Gal β 1,4(Fuc α 1,3)GlcNAc6S β -R
Neu5Gc α 2,3Gal β 1,4(Fuc α 1,3)GlcNAc β -R
Neu5Gc α 2,3Gal β 1,4GlcNAc6S β -R
Neu5Gc α 2,3Gal β 1,4GlcNAc β -R
Neu5Gc α 2,3Gal β 1,4Glc β -R
Neu5Gc α 2,3Gal β -R
Neu5Gc α 2,6GalNAc α -R
Neu5Gc α 2,6Gal β 1,4GlcNAc β -R
Neu5Gc α 2,6Gal β 1,4Glc β -R
Neu5Gc α 2,6Gal β -R
Neu5Gc α 2,8Neu5Ac α 2,3Gal β 1,4Glc β -R
Neu5Gc α 2,8Neu5Gc α 2,3Gal β 1,4Glc β -R

[00169] The following abbreviations are used herein: Glc – glucose, Gal – galactose, GlcNAc – N-acetylglucosamine, GalNAc – N-acetylgalactosamine, GlcNAc6S – 6-Sulfo-N-acetylglucosamine, KDN – 2-keto-3-deoxy-D-glycero-D-galactonononic acid, Neu5,9Ac2 – N-acetyl-9-O-acetylneuraminic acid, Fuc – fucose and Neu5GcOMe – 2-O-methyl-N-glycolylneuraminic acid. O-glycosidic bonds are present between each residue in the glycans

listed with α and β indicating the relative stoichiometry between the two residues joined by the bond, wherein α indicates an axial orientation and β indicates an equatorial orientation. The numbers following α and/or β , in the format x,x , indicated the carbon number of each of the carbons from each of the adjoined residues that participate in bond formation. While the glycans listed in Table 1 represent individual glycan target antigens contemplated, the present invention also includes embodiments wherein the above presented glycans comprise different combinations of α and β -oriented O-glycosidic bonds than the ones presented. Also in Table 1, R represents an entity that the glycan may be coupled with. In some embodiments, R is a protein wherein the glycan is linked typically to a serine or threonine residue. In some embodiments, R is a linker molecule used to join the glycan to a substrate, such as in a glycan array. In some embodiments, R may be a linker comprising $-(CH_2)_2CH_2NH_2$ or $-(CH_2)_3NHCOCH_2(OCH_2CH_2)_6NH_2$. In some embodiments, R may be biotin, albumin, $ProNH_2$, $-CH-$, $-OH$, $-OCH_3$, $-OCH_2CH_3$, $-H$, hydrido, hydroxy, alkoxy, oxygen, carbon, sulfur, nitrogen, polyacrylamide, phosphorus, NH_2 , $ProNH_2=O(CH_2)_2CH_2NH_2$, $(OCH_2CH_2)_6NH_2$, $O(CH_2)_3NHCOCH_2(OCH_2CH_2)_6NH_2$, the fluorescent labels 2-aminobenzamide (AB) and/or 2-aminobenzoid acid (AA), 2-aminobenzamide analog that contains an alkyl amine (AEAB), aminoxy- groups, methylaminoxygroups, hydrazide groups, amino lipid 1,2-dihexadecyl-sn-glycero-3-phosphoethanolamine (DHPE), aminoxy (AO) functionalized DHPE and glycosylphosphatidylinositol (GPI). Without intending to limit the source or nature of R, this may include structures that affect the physical spacing of glycan residue. In some embodiments, the R group may comprise a combination of the R groups presented here, e.g. a biotinylated polyacrylamide. In some embodiments, the R group in combination with underlying substrates effect glycan residue spacing.

[00170] Glycan targets of the present invention may comprise regions of antibody recognition. As used herein, the term “region of antibody recognition” refers to one or more regions located on any part of the molecule, an attached group or located on a region of interaction between the glycan and another molecule, including, but not limited to another glycan. In some embodiments, regions of antibody recognition are located at interchain target sites, wherein the term interchain means within the present polymeric chain. Interglycan target sites may comprise regions of antibody recognition comprising 1, 2, 3, 4, 5, 6, 7, 8, 9 or at least 10 residues, bonds between residues or combinations of residues and bonds. In some embodiments, regions of antibody recognition are located at regions of interaction between one or more glycan chains. Such regions may be between 2, 3, 4 or at least 5 glycan chains.

[00171] In some embodiments, regions of antibody recognition are located at regions of interaction between glycan branch chains connected to a common parent chain. In some embodiments, regions of antibody recognition are located at regions of interaction between a glycan branch chain and a parent chain. In some embodiments, regions of antibody recognition are located at regions of interaction between glycans and proteins. Such regions of interaction may comprise chemical bonds between the glycan and the protein, including, but not limited to covalent bonds, ionic bonds, hydrostatic bonds, hydrophobic bonds and hydrogen bonds. In some embodiments, regions of antibody recognition are located at regions of interaction between glycans and other biomolecules including, but not limited to lipids and nucleic acids. Such regions of interaction may comprise chemical bonds between the glycan and the biomolecule, including, but not limited to covalent bonds, ionic bonds, hydrostatic bonds, hydrophobic bonds and hydrogen bonds.

[00172] In some embodiments, glycan targets of the present invention are components of glycoconjugates. As used herein, the term “glycoconjugate” refers to any entity comprising a glycan moiety. In some embodiments, glycoconjugates are glycolipids. As used herein, the term “glycolipid” refers to a class of lipids wherein a carbohydrate moiety is covalently attached. In some embodiments, carbohydrate moieties present on glycolipids comprise glycans. In some embodiments, lipid components of glycolipids comprise ceramide moieties. Examples of glycolipids contemplated as targets of the present invention include, but are not limited to glyceroglycolipids (including, but not limited to galactolipids and sulfolipids), glycosphingolipids (including, but not limited to cerebroside (e.g., galactocerebroside, glucocerebroside and sulfatide), gangliosides, globosides and glycosphosphosphingolipids) and glycosylphosphatidylinositols. When located within cell membranes, glycan moieties of glycolipids are located on the extracellular side of the membrane where they may interact with other cells as well as cell signaling ligands (Maccioni, H.J. et al., *Organization of the synthesis of glycolipid oligosaccharides in the Golgi complex*. FEBS Lett. 2011 Jun 6;585(11):1691-8).

[00173] In some embodiments, glycoconjugate targets of the present invention are glycoprotein and/or proteoglycans. Glycoproteins refer to any proteins that are covalently bonded with glycans. Proteoglycans are a class of proteins that are heavily glycosylated with glycans that often carry a negative charge. This property makes them very hydrophilic and important components of connective tissue.

Recombinant antibodies

[00174] Recombinant antibodies (e.g., glycan-interacting antibodies) of the invention may be generated using standard techniques known in the art. In some embodiments, recombinant antibodies may be anti-glycan antibodies. Further antibodies may be anti-STn antibodies (e.g. anti-GcSTn or anti-AcSTn antibodies). Recombinant antibodies of the invention may be produced using variable domains obtained from hybridoma cell-derived antibodies produced according to methods described herein. Heavy and light chain variable region cDNA sequences of antibodies may be determined using standard biochemical techniques. Total RNA may be extracted from antibody-producing hybridoma cells and converted to cDNA by reverse transcriptase (RT) polymerase chain reaction (PCR). PCR amplification may be carried out on resulting cDNA to amplify variable region genes. Such amplification may comprise the use of primers specific for amplification of heavy and light chain sequences. In other embodiments, recombinant antibodies may be produced using variable domains obtained from other sources. This includes the use of variable domains selected from one or more antibody fragment library, such as an scFv library used in antigen panning. Resulting PCR products may then be subcloned into plasmids for sequence analysis. Once sequenced, antibody coding sequences may be placed into expression vectors. For humanization, coding sequences for human heavy and light chain constant domains may be used to substitute for homologous murine sequences. The resulting constructs may then be transfected into mammalian cells for large scale translation.

Anti-Tn antibodies

[00175] In some embodiments, recombinant antibodies of the invention (e.g., glycan-interacting antibodies) may be anti-Tn antibodies. Such antibodies may bind to targets comprising Tn. Anti-Tn antibodies may be specific for Tn or may bind other modified forms of Tn, such as Tn linked to other moieties, including, but not limited to additional carbohydrate residues. In some cases anti-Tn antibodies may be anti-sialyl-Tn antibodies. Such antibodies may bind to targets comprising sialylated Tn comprising Neu5Ac and/or targets comprising sialylated Tn comprising Neu5Gc. Some anti-Tn antibodies may bind specifically to clusters of Tn antigen.

Anti-STn antibodies

[00176] In some embodiments, antibodies of the invention (e.g., glycan-interacting antibodies) may specifically bind to antigens comprising STn. Anti-STn antibodies of the invention may be categorized by their binding to specific portions of STn antigens and/or by their specificity for AcSTn versus GcSTn. In some cases, anti-STn antibodies of the invention are Group 1 antibodies. “Group 1” antibodies according to the invention are antibodies capable of binding AcSTn and GcSTn. Such antibodies may also be referred to herein as pan-STn antibodies due to their ability to associate with a wider range of STn structures. In some embodiments, Group 1 antibodies may associate with the portion of STn indicated by the large oval in Fig. 1A. In some cases, anti-STn antibodies of the invention are Group 2 antibodies. “Group 2” antibodies, according to the invention, are antibodies capable of binding STn as well as some related structures that include an O-linkage to serine or threonine. In some embodiments, Group 2 antibodies may associate with glycans comprising a sialylated galactose residue. In some cases, Group 2 antibodies may associate with the portion of STn indicated by the large oval in Fig. 1B. Some Group 2 antibodies preferably bind to structures with AcSTn over structures with GcSTn. Further anti-STn antibodies may be Group 3 antibodies. As referred to herein, “Group 3” antibodies are antibodies capable of binding STn, but may also bind a broader set of related structures. Unlike Group 2 antibodies, Group 3 antibodies do not require that such structures have an O-linkage to serine or threonine. In some embodiments, Group 3 antibodies may associate with the portion of STn indicated by the large oval in Fig. 1C. Finally, some anti-STn antibodies of the invention may be Group 4 antibodies. As referred to herein, “Group 4” antibodies are capable of binding to both AcSTn and GcSTn as well as the un-sialylated Tn antigen, and therefore have broader specificity. In some embodiments, Group 4 antibodies may associate with the portion of STn indicated by the large oval in Fig. 1D.

[00177] In some cases, anti-STn antibodies of the invention may bind specifically to clusters of STn on a particular antigen or cell surface. Some such antibodies may recognize epitopes formed by the clustering of STn, including epitopes that include areas of contact between neighboring STn structures. Such epitopes may be formed by the clustering of 2, 3, 4, 5, 6, 7, 8, 9, 10 or more STn structures.

Antibody components

[00178] In some cases, antibodies or antigen binding fragments thereof of the invention may comprise variable domain and/or CDR amino acid sequences provided herein. Some antibodies or antigen binding fragments may comprise different combinations of such sequences. In some cases, antibodies or antigen binding fragments of the invention may comprise one or more of the variable domain sequences listed in Table 2. Residues indicated with an “X” may be absent or selected from any amino acid residues. Light chain variable domains presented in the Table may be expressed with or without a C-terminal arginine residue. This residue typically links light chain variable domains with light chain constant domains and may be expressed as part of the light chain constant domain instead of the light chain variable domain. In some cases, antibodies or antigen binding fragments thereof may comprise an amino acid sequence with from about 50% to about 99.9% sequence identity (e.g. from about 50% to about 60%, from about 55% to about 65%, from about 60% to about 70%, from about 65% to about 75%, from about 70% to about 80%, from about 75% to about 85%, from about 80% to about 90%, from about 85% to about 95%, from about 90% to about 99.9%, from about 95% to about 99.9%, about 97%, about 97.5%, about 98%, about 98.5%, about 99%, about 99.5%, about 99.6%, about 99.7% or about 99.8%) with one or more of the variable domain sequences listed in Table 2 or a fragment thereof (e.g., an N-terminal fragment, C-terminal fragment, or internal fragment).

Table 2. Variable domain sequences

Antibody ID Number	Variable domain	Sequence	SEQ ID NO
18D2	Heavy chain	QVTLKESGPGILQPSQTLSTCSFSGFSLSTSNMGIG WIRQPSGKGLEWLAHIWWHDDKYYNPSLKSRLTIS KDISNNQVFLKITSVDTADTATYYCAQVPFYGTSTF DVWGTGTTVTVSS	1
18D2	Light chain 1	DIVMTQSPATLSVTPGDRVSLSCRASQSDYLHWY QQKSHESPRLLIKYASQSIGIPSRFSGSGSGSDFTLSI NSVEPEDVGYYCQNGHSFPLTFGAGTKLELK	2
18D2	Light chain 2	QIVLTQSPAISASPGETVTMTCSASSSITYMHWYQ QKPGTSPKRWIYDTSKLASGVPARFSGSGSGTSYSL TISSMEAEDAATYYCHQRSSYTFGGGTKLEIKR	3
18C7	Heavy chain	QVTLKESGPGILQPSQTLSTCSFSGFSLSTFGMGVG WIRQPSGKGLEWLAHIWWDDDKYYNPALKSRLTIS KDTSKNQVFLKIANVDTADTATYYCARIAYYYGSE RDYWGQGTTTLTVSS	4
18C7	Light	QIVLTQSPAISASPGEKVTMTCSASSSISYMHWHYH	5

	chain	QKPGTSPKRWIYDTSKLASGVPARFSGSGSGTSYSL TISSMEAEDAATYYCHQRSSYTFGGGKLEIKR	
10A5- 2A12	Heavy chain	QVQLKESGPGLVAPSQSL SITCTVSGFSLTSYGVS WRQPPGKGLEWLGVIWGDGSTNYHSSLISRLSISKD NSKSQVFLKLNSLQTDDTATYYCARAFVYWGQGT LVTVSA	6
10A5- 2A12	Light chain	QIVLTQSPA IMSASPGEKVTMTCSASSSVSYIHWYQ QKSGTSPKRWIYDTSKLASGVPARFSGSGSGTSYSL TISSMEAEDAATYYCQQWSSNPPMLTFGAGTKLEL K	7
8C11- 1D10	Heavy chain	QVQLKESGPGLVAPSQSL SITCTVSGFSLTSYGVS WRQPPGKGLEWLGVIWGDGSTNYHSALISRLIISKD NSKSQVFLKLNSLQTDDTATYYCTKGFTYWGQGTL VTVSA	8
8C11- 1D10	Light chain	QIVLTQSPA IMSASPGEKVTMTCSASSSVSYMHWY QQKSGTSPKRWIFDTSKLASGVPARFSGSGSGTSYS LTISSMEAEDAATYYCQQWSSNLLTFGAGTKLELK	9
2D4-1B4	Heavy chain	QVQLQESGPGLVAPSQSL SITCTVSGFSLISYGVNW VRQPPGKGLEWLGVIWGDGSTNYQSALISRLIISKD NSKSQVFLKLNSLQTDDTATYYCTKGFAYWGQGT LVTVSA	10
2D4-1B4	Light chain	QIVLTQSPA IMSASPGEKVTMTCSASSSVSYMHWY QKSGTSPKRWIYDTSKLASGVPARFSGSGSGTSYSL TISSMEAEDAATYYCQQWSSNLLTFGAGTKLELK	11
7G9-1A8	Heavy chain	QVQLKESGPGLVAPSQNL SITCTVSGFSLTSYGVNW VRQPPGKGLEWLGVIWGDGSTNYHSALISRLIISKE NSKSQVFLKLNSLQTNDTATYYCTKGFVYWGQGT LVTVSA	12
7G9-1A8	Light chain	QIVLTQSPA IMSASPGEKVTMTCSASSSVSYMHWY QQKSGTSPKRWIFDTSKLASGVPARFSGSGSGTSYS LTISSMEAEDAATYYCQQWSSNLLTFGAGTKLELK	9
1A12- 2B2	Heavy chain	QVQLKESGPGLVAPSQSL SITCTVSGFSLTSYGVS WRQPPGKGLEWLGVIWGDGSTNYHSALISRLSISKD NSKSQVFLKLNSLQTDDTATYYCAKGGYFDYWGQ GTTLTVSS	13
1A12- 2B2	Light chain	QIVLTQSPA VMSASPGEKVAITCSASSSVSYMHWY QKPGTSPKLWIYSTSNLASGVPARFSGSGSGTSYSLT ISRMEAEDAATYYCQQRSSYPWTFGGGKLEIK	14
7D3- 2C10	Heavy chain	QVQLQYDAELVKPGGSVKISCKASGYTFTDHAH WVKQKPEQGLEWIGYFSPGNDDIKYSEKFKGKATL TADKSSSTAYMQLNSLTSEDSAVYFCKRSITTPYWG QGTLVTVSA	15
7D3- 2C10	Light chain	DIQMNQSPSSLSASLGDTITITCHASQNINVWLSWY QQKPGNIPKLLIYKVSNLHTGVPSRFSGSGSGTGFTL TISSLQPEDIATYYCQQDQSYPTYTFGGGKLELK	16
A5-2G12	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYTFTDHAH WVKQKPEQGLEWIGYISPGNDDIKYNEKFKGKATL TADKSSSTAYMQLNSLTSEDSAVYFCKRSITTSYWG QGTLVTVSA	17

A5-2G12	Light chain	NIVMTQSPKSMMSVGERVTLTCKASENVVIYVSW YQQKPEQSPKLLIYGASNRYTGVPDRFTGSGSATDF TLTISSVQAEDLADYHCGQGYSYPYTFGGGGTKLEIK R	18
1A5-2C9	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYTFTDHAIH WVKQKPEQGLEWIGYVSPGNGDIKYNEKFKGKAT LTADKSSSTAYMQLNSLTSEDSAVYFCKRSLIGDY WGQGTTTLTVSS	19
1A5-2C9	Light chain	DIVMTQSQKFMSSSVGDRVTITCKASQNVGTAVAW YQQKPGQSPKFLIYSASNRYTGVPDRFTGSGSGTDF TLTISNMQSEDLADYFCQQYSSYRLTFGGGGTKLEIK	20
4D9-2C11	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYTFTDHAIH WVKQKPEQGLEWIGYLSPGND DIKYSEKFKDKATL TADKSSSTAYMQLNSLTSEDSAVYFCKRSIGGDHW GQGTTTLTVSS	21
4D9-2C11	Light chain	DIQMNQSPSSLSASLGDTITITCHASQNINVLNWX QQKPGNIPKLLIYKASNLHTGVPSRFSGSGSGTGFTL TIGSLQPEDIATYYCQQGQSYPTTFGGGGTKLEIKR	22
2F4-1E2	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYTFTDHAIH WVKQKPEQGLEWIGYISPGNGDIKYNEKFKGKATL TADKSSSTAYMQLNSLTSEDSAVYFCQRQLGQGY WGQGTTTLTVSS	23
2F4-1E2	Light chain	DVVMQTPLSLPVSLGDQASISCRSSQSLVHSYGNT YLHWY LQKPGQSPKLLIYKVS NRFSGVPDRFSGSGS GTDFTL KISRVEAEDLG VYFCSQNT HVPYTFGGGGTK LEIKR	24
2F4-1H8	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYTFTDHAIH WVKQKPEQGLEWIGYISPGNGDIKYNEKFKGKATL TADKSSSTAYMQLNSLTSEDSAVYFCQRQLGQGY WGQGTTTLTVSS	23
2F4-1H8	Light chain	DVVMQTPLSLPVSLGDQASISCRSSQSLVHSYGNT YLHWY LQKPGQSPKLLIYKVS NRFSGVPDRFSGSGS GTDFTL KISRVEAEDLG VYFCSQNT HVPYTFGGGGTK LEIKR	24
2C6-2F11	Heavy chain	QVQLQQSDAELGKPGASVKISCKASGYTFSDHAIH WVKQKPEQGLEWIGYISPGNDDIKYNEKFKGKATL TADKSSSTAYMQLNSLTSEDSAVYFCERSMIGVYW GQGTLVTVSA	25
2C6-2F11	Light chain	DVVMQTPLSLTVSLGDQASISCRFSQSLVQSNGNT YLQWY LQKPGQSPKLLIYKVS NRFCGVPDRFSGSGS GTDFTL KISRVEAEDLG VYFCSQST HAPLTFGAGTK LELK	26
2B2-2A7	Heavy chain	QVQLQQSDAELVKPGASVKISCKTSGYTFTDHAIH WVKQKPEQGLEWIGYISPGNGDIKYNEKFKGKATL TADKSSSTAYMQLSSLTPEDSAVYFCKISYYGIWQQ GTTTLTVSS	27
2B2-2A7	Light chain	DIQMTQSPASLSVSVGESVTITCRLSEDIYSNLAWFQ QRP GKSPQLLVYKATNLADGVPSRFSGSGSGTQYSL KINSLQSEDFGTYYCQHFWGTPFTFGSGGTKVEIK	28

5G2-1B3	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYTFTDHAIH WVKQKPEQGLEWIGYFSPGNDDIKYNEKFKVKATL TADKSSSTAYMQLTSLTSEDSAVYFCKRSYYGDWG QGTTTLTVSS	29
5G2-1B3	Light chain	DIQMTQSPASLSVSVGETVTITCRASENIYSHLAWY QQKQGKSPQLLVYGATNLADGVPSRFSGSGSGTQF SLKIHSLQSEDFGSYYCQHFWGAPFTFGSGTKLEIK	30
7A6-2A2	Heavy chain	QIQLQQSDAELVKPGTSVKMSCKASGYTFTDHAIH WVKQKPEQGLEWIGYFSPGNDDIKYNVKFKGKATL TADKSSSTAYMQLNSLTSEDSAVYFCSVGYALDYW GLGTTTLTVSS	31
7A6-2A2	Light chain	NIVMTQSPKSMMSVGERVTLTCKASENVVTYVSW YQQKPEQSPKLLIYGASNRYTGVPDRFTGSGSATDF TLTISSVQAEDLADYHCGQGYSYPYTFGGGTKLEIK R	32
10C9-2G7	Heavy chain	QVQLQQSDAELVKPGTTVKISCKASGYTFTDHAIH WVKEKPEQGLEWIGYISPGNDDIKYSEKFKGKATLT ADKSSSTAYMQLNSLTSDDSAVYFCKRSLSTPYWG QGTLVTVSA	33
10C9-2G7	Light chain	Unknown	
1C11-2G9	Heavy chain	Unknown	
1C11-2G9	Light chain	DIVMTQSPSSLTVTAGEKVTMSCRSSQSLLNSGNQK NYLTWYQQKPGQPPKLLIYWASTRESGVPDRFTGS GSGTDFTLTISSVQAEDLAVYYCQNDYSYPYTFGG GTKLEIKR	34
1F6-1B7 (also sequence of 1F6- 1C10)	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYTFTDHAIH WVMQMPEQGLEWIGYISPGNGDVKYSERFKGRAT LTADKSSSSAYMQLNSLTSEDSAVYFCKRSLSTPYW GQGTTLTVS	35
1F6-1B7 (also sequence of 1F6- 1C10)	Light chain	DIVMTQSPSSLTVTAGERVMTMSCKSSQSLLNSGNQK SYLTWYQQKPGQPPKLLISWASTRDSGVPDRFTGS GSGTDFTLTISSVQAEDLAVYYCQSDYSYPYTFGGG TKLEIKR	36
2G12-2B2	Heavy chain	QVQLQQSDXELVKPGASVKISCKASGYTFTDHAIH WVKQKPEQGLEWIGYFSPGNDDIKYNEKFRGKATL TADKSSSTAYMQLNSLSSDDSAVYFCKRSLSTPYW GQGTTLXTVSA	37
2G12-2B2	Light chain	DIVMTQSPSSLTVTAGEKVTMSCKSSQSLLNRGNH KNYLTWYRQKPGPLPPKLLIYWASTRESGVPDRFTG SGSGTDFALTISVQAEDLAVYYCQNDYTYPYTFG GGTKLEIKR	38
5E6-2E7	Heavy chain	QVQLQQSDAELVKPGASMKISCKASGYTFTDHAIH WVKQKPEQGLEWIGYISPGNGDIKYNEKFKVKATL TADKSSSTAYMQLNSLTSEDSAVYFCKRSITTPYWG QGTLVTVSA	39

5E6-2E7	Light chain	DIVMTQSPSSLTVTAGEKVTMSCKSSQSLLNSGKTK NYLTWYQQKPGQPPKLLIYWASTRESGVPDRFTGS GSGTDFTLTISSVQAEDLAVYYCKNDYSYPYTFGG GTKLEIKR	40
9E5-1A8	Heavy chain	QVQLQQSDAELVKPGASVKISCKTSGYTFTDHAIH WVKQKPEQGLEWIGYISPGNDDIKYTEKFKGKVTL TADKSSSTAYMQLNSLTSEDSAVYFCKRSITTPYWG QGTLVTVSA	41
9E5-1A8	Light chain	Unknown	
9F11-1F7	Heavy chain	QVQLQQSDAELVKPGASMKISCKASGYTFTDHAIH WVKQKPEQGLEWIGYISPGNGDIKYNEKFKVKATL TADKSSSTAYMQLNSLTSEDSAVYFCKRSITTPYWG QGTLVTVSA	39
9F11-1F7	Light chain	DIVMTQSPSSLTVTAGEKVTMSCKSSQSLLNSGKTK NYLTWYQQKPGQPPKLLIYWASTRESGVPDRFTGS GSGTDFTLTISSVQAEDLAVYYCKNDYSYPYTFGG GTKLEIKR	40
10F4-2F2	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYTFTDHAIH WVKQKPEQGLEWIGYISPGNGDIKYDEKFKGKATL TADKSSSTAYMQLNSLTSEDSAVYFCKRSITTSYWG QGTLVTVSA	42
10F4-2F2	Light chain	NIVMTQSPKSMMSVGERVTLTCKASENVVITYVSW YQQKPEQSPKLLIYGASNRYTGVPDRFTGSGSATDF TLTISSVQAEDLADYHCGQGYSYPYTFGGGTKLEIK R	32
2B8-2F10	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYTFTDHAIH WVKQKPEQGLEWIGYISPGNDDIKYNEKFKGKATL TADKSSSTAYMQLNSLTSEDSAVFFCKRSITTSYWG QGTLVTVSA	43
2B8-2F10	Light chain	Unknown	
4G8-1E3	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYIFTDHAIIHW VKQKPEQGLEWIGYISPGNGDIKYNEKFKGKATLT ADKSSSTAYMHLNSLTSEDSAVYFCKRSITTSYWG QGTLVTVSA	44
4G8-1E3	Light chain	DIQMNQSPSSLSASLGDTITITCHASQHINFWLSWYQ QKPGNIPKLLIYKASNLHTGVPSRFSGSGSGTGFTLT ISSLLPEDVATYYCQQDQSYPYMFGGGTKLEIKR	45
6B11-2E3	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYTFTDHAIH WVKQKPEQGLEWIGYISPGNDDIKYNEKFKGKATL TADKSSSTAYMMLNSLTSEDSAVYFCKRSITTSYWG QGTLVTVSA	46
6B11-2E3	Light chain	NIVMTQSPKSMMSVGERVTLTCKASENVVITYVSW YQQKPEQSPKLLIYGASNRYTGVPDRFTGSGSATDF TLTISSVQAEDLADYHCGQGYSYPYTFGGGTKLEIK R	32
8C2-2D6	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYTFTDHAIH WVKQKPEQGLEWIGYISPGNGDIKYNEKFKGKATL	47

		TADTSSTTAYMQLNSLTSEDSAMYFCKRSITTSYW GQGLTVTVSA	
8C2-2D6	Light chain	NIVMTQSPKSMMSVGERVTLTCKASENVVTVVSW YQQKPEQSPKLLIYGASNRYTGVPDRFTGSGSATDF TLTISSVQAEDLADYHCGQGYSYPYTFGGGGTKLEIK R	32
7D4-2A2- 2F2	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYIFTDHAIHW VKQKPEQGLEWIGYISPGNGDIKYIEKFRGKATLTA DKSSSTAYMQLNSLTSEDSAVYFCKRSLSTPYWGQ GTLTVTVSA	48
7D4-2A2- 2F2	Light chain	NILMTQSPKSMMSVGERVTLTCKASENVVNYVSW YQQKPEQSPKLLIFGASNRYSGVPDRFTGSGSATDF TLTISSVQAEDLADYHCGSKWITSYPYTFGGGGTKLE IKR	49
7D4- 1H12- 2B3	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYIFTDHAIHW VKQKPEQGLEWIGYISPGNGDIKYIEKFRGKATLTA DKSSSTAYMQLNSLTSEDSAVYFCKRSLSTPYWGQ GTLTVTVSA	48
7D4- 1H12- 2B3	Light chain	NILMTQSPKSMMSVGERVTLTCKASENVVNYVSW YQQKPEQSPKLLIYGASNRYSGVPDRFTGSGSATDF TLTISSVQAEDLADYHCGARVTSYPYTFGGGGTKLEI KR	50
2C2-2C5	Heavy chain	QVQLQQSDAELVKPGTSVKISCRASGYTFTDHAIH WVKQKPEQGLEWIGYISPGNGDIKYNEKFKGKATL TADKSSSTAYMQLNSLTSDDSAVYFCKRSITTPYW GQGTTTLTVSS	51
2C2-2C5	Light chain	SFVMTQTPKFLLVSA GDRVTITCKASQSVNNNVAW YQQKPGQSPKQLIYYASNRYTGVPDRFTGSGYGTD FTFTIYTVQAEDLAVYFCQQGYSSPWTFGGGGTKLK	52
10F4-2A9	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYTFTDHAIH WVKQKPEQGLEWIGYISPGNGDIKYDEKFKGKATL TADKSSSTAYMQLNSLTSEDSAVYFCKRSITTSYWG QGLTVTVSA	42
3F1	Heavy chain	QVQLQQSDAELVKPGASVKISCKASGYTFTDHAIH WVKQKPEQGLDWIGYISPGNGDIKYNEKFKDKVTL TADKSSSTACMHLNSLTSEDSAVYFCKRSLALDY WGQGTTTLTVSS	53
3F1	Light chain	DIVMTQSHKFMSTSVGDRVSITCKASQDVGTNIAW YQQKPGRSPKVLIIYASTRHTGVPDRFTGSGSGTDF TLTISNVQSEDLTDYFCQQYSSFPLTFGVGKLELK	54

[00179] In some embodiments, antibodies of the invention may be developed using one or more CDRs derived from any of the variable domain sequences presented in the previous Table. In some cases, CDR sequences are determined from a variable domain sequence through the use of one or more numbering schemes including, but not limited to, those taught by Kabat [Wu, T.T. et al., 1970, JEM, 132(2):211-50 and Johnson, G. et al., 2000, Nucleic

Acids Res. 28(1): 214-8, the contents of each of which are herein incorporated by reference in their entirety], Chothia [Chothia and Lesk, J. Mol. Biol. 196, 901 (1987), Chothia et al., Nature 342, 877 (1989) and Al-Lazikani, B. et al., 1997, J. Mol. Biol. 273(4):927-48, the contents of each of which are herein incorporated by reference in their entirety], Lefranc (Lefranc, M.P. et al., 2005, Immunome Res. 1:3) and Honegger (Honegger, A. and Pluckthun, A. 2001. J. Mol. Biol. 309(3):657-70, the contents of which are herein incorporated by reference in their entirety).

[00180] In some cases, antibodies or antigen binding fragments thereof of the invention may comprise one or more of the CDR amino acid sequences listed in Table 3. Residues indicated with an “X” may be absent or selected from any amino acid residues. In some cases, antibodies or antigen binding fragments thereof may comprise an amino acid sequence with from about 50% to about 99.9% sequence identity (e.g. from about 50% to about 60%, from about 55% to about 65%, from about 60% to about 70%, from about 65% to about 75%, from about 70% to about 80%, from about 75% to about 85%, from about 80% to about 90%, from about 85% to about 95%, from about 90% to about 99.9%, from about 95% to about 99.9%, about 97%, about 97.5%, about 98%, about 98.5%, about 99%, about 99.5%, about 99.6%, about 99.7% or about 99.8%) with one or more of the CDR sequences listed in Table 3. In some cases, antibodies or antigen binding fragments thereof of the invention may comprise an amino acid sequence comprising one or more fragments of any of the sequences listed in Table 3. In the Table, “consensus” refers to an antibody sequence derived from alignment of multiple sequences wherein the most conserved residues were used to form the consensus sequence. B72.3 (Thermo Fisher Scientific, Waltham, MA) and CC49 (see Muraro, R. et al., 1988. Cancer Res. 48: 4588-96) are commercially available antibodies.

Table 3. CDR sequences

Antibody ID Number	CDR	Sequence	SEQ ID NO
18D2	CDR-H1	GFSLSNSTMG	55
18C7	CDR-H1	GFSLSSTFGMG	56
10A5-2A12	CDR-H1	GFSLSYSG	57
8C11-1D10	CDR-H1	GFSLSYSG	57
2D4-1B4	CDR-H1	GFSLSISYSG	58
7G9-1A8	CDR-H1	GFSLSYSG	57
1A12-2B2	CDR-H1	GFSLSYSG	57
18D2	CDR-H2	IWWHDDK	59

18C7	CDR-H2	IWWDDDK	60
10A5-2A12	CDR-H2	IWGDGST	61
8C11-1D10	CDR-H2	IWGDGST	61
2D4-1B4	CDR-H2	IWGDGST	61
7G9-1A8	CDR-H2	IWGDGST	61
1A12-2B2	CDR-H2	IWGDGST	61
18D2	CDR-H3	AQVPFYYSFDV	62
18C7	CDR-H3	ARIAYYYGSRDY	63
10A5-2A12	CDR-H3	ARAFVY	64
8C11-1D10	CDR-H3	TKGFTY	65
2D4-1B4	CDR-H3	TKGFAY	66
7G9-1A8	CDR-H3	TKGFVY	67
1A12-2B2	CDR-H3	AKGGYFDY	68
18C7	CDR-L1	SSISY	69
10A5-2A12	CDR-L1	SSVSY	70
8C11-1D10	CDR-L1	SSVSY	70
2D4-1B4	CDR-L1	SSVSY	70
7G9-1A8	CDR-L1	SSVSY	70
1A12-2B2	CDR-L1	SSVSY	70
18D2	CDR-L1	QSIDY	71
18D2	CDR-L1	SSITY	72
18C7	CDR-L2	DTS	73
10A5-2A12	CDR-L2	DTS	73
8C11-1D10	CDR-L2	DTS	73
2D4-1B4	CDR-L2	DTS	73
7G9-1A8	CDR-L2	DTS	73
1A12-2B2	CDR-L2	STS	74
18D2	CDR-L2	YAS	75
18D2	CDR-L2	DTS	73
18C7	CDR-L3	HQRSSYT	76
10A5-2A12	CDR-L3	QQWSSNPPMLT	77
8C11-1D10	CDR-L3	QQWSSNLLT	78
2D4-1B4	CDR-L3	QQWSSNLLT	78
7G9-1A8	CDR-L3	QQWSSNLLT	78
1A12-2B2	CDR-L3	QQRSSYPWT	79
18D2	CDR-L3	QNGHSFPLT	80
18D2	CDR-L3	HQRSSYT	76
7A6-2A2	CDR-H1	GYTFTDHAIHWV	81
2B2-2A7	CDR-H1	GYTFTDHAIHWV	81
5G2-1B3	CDR-H1	GYTFTDHAIHWV	81
4D9-2C11	CDR-H1	GYTFTDHAIHWV	81
2F4-1E2	CDR-H1	GYTFTDHAIHWV	81
2F4-1H8	CDR-H1	GYTFTDHAIHWV	81
1A5-2C9	CDR-H1	GYTFTDHAIHWV	81

1F6-1B7 (also sequence of 1F6- 1C10)	CDR-H1	GYTFTDHAIHWV	81
2C2-2C5	CDR-H1	GYTFTDHAIHWV	81
2G12-2B2	CDR-H1	GYTFTDHAIHWV	81
10C9-2G7	CDR-H1	GYTFTDHAIHWV	81
2C6-2F11	CDR-H1	GYTFSDHAIHWV	82
7D4-2A2-2F2	CDR-H1	GYIFTDHAIHWV	83
7D4-1H12-2B3	CDR-H1	GYIFTDHAIHWV	83
7D3-2C10	CDR-H1	GYTFTDHAIHWV	81
8C2-2D6	CDR-H1	GYTFTDHAIHWV	81
9E5-1A8	CDR-H1	GYTFTDHAIHWV	81
5E6-2E7	CDR-H1	GYTFTDHAIHWV	81
9F11-1F7	CDR-H1	GYTFTDHAIHWV	81
4G8-1E3	CDR-H1	GYIFTDHAIHWV	83
10F4-2F2	CDR-H1	GYTFTDHAIHWV	81
10F4-2A9	CDR-H1	GYTFTDHAIHWV	81
6B11-2E3	CDR-H1	GYTFTDHAIHWV	81
2B8-2F10	CDR-H1	GYTFTDHAIHWV	81
7A5-2G12	CDR-H1	GYTFTDHAIHWV	81
7A6-2A2	CDR-H2	FSPGNDDIKY	84
2B2-2A7	CDR-H2	ISPGNGDIKY	85
5G2-1B3	CDR-H2	FSPGNDDIKY	84
4D9-2C11	CDR-H2	LSPGNDDIKY	86
2F4-1E2	CDR-H2	ISPGNGDIKY	85
2F4-1H8	CDR-H2	ISPGNGDIKY	85
1A5-2C9	CDR-H2	VSPGNGDIKY	87
1F6-1B7 (also sequence of 1F6- 1C10)	CDR-H2	ISPGNGDVKY	88
2C2-2C5	CDR-H2	ISPGNGDIKY	85
2G12-2B2	CDR-H2	FSPGNDDIKY	84
10C9-2G7	CDR-H2	ISPGNDDIKY	89
2C6-2F11	CDR-H2	ISPGNDDIKY	89
7D4-2A2-2F2	CDR-H2	ISPGNGDIKY	85
7D4-1H12-2B3	CDR-H2	ISPGNGDIKY	85
7D3-2C10	CDR-H2	FSPGNDDIKY	84
8C2-2D6	CDR-H2	ISPGNGDIKY	85
9E5-1A8	CDR-H2	ISPGNDDIKY	89
5E6-2E7	CDR-H2	ISPGNGDIKY	85
9F11-1F7	CDR-H2	ISPGNGDIKY	85
4G8-1E3	CDR-H2	ISPGNGDIKY	85
10F4-2F2	CDR-H2	ISPGNGDIKY	85
10F4-2A9	CDR-H2	ISPGNGDIKY	85

6B11-2E3	CDR-H2	ISPGNDDIKY	89
2B8-2F10	CDR-H2	ISPGNDDIKY	89
7A5-2G12	CDR-H2	ISPGNDDIKY	89
7A6-2A2	CDR-H3	SVGYALDY	90
2B2-2A7	CDR-H3	KISYYGI	91
5G2-1B3	CDR-H3	KRSYYGD	92
4D9-2C11	CDR-H3	KRSIGGDH	93
2F4-1E2	CDR-H3	QRQLGQGY	94
2F4-1H8	CDR-H3	QRQLGQGY	94
1A5-2C9	CDR-H3	KRSLIGDY	95
1F6-1B7 (also sequence of 1F6- 1C10)	CDR-H3	KRSLSTPY	96
2C2-2C5	CDR-H3	KRSITTPY	97
2G12-2B2	CDR-H3	KRSLSTPY	96
10C9-2G7	CDR-H3	KRSLSTPY	96
2C6-2F11	CDR-H3	ERSMIGVY	98
7D4-2A2-2F2	CDR-H3	KRSLSTPY	96
7D4-1H12-2B3	CDR-H3	KRSLSTPY	96
7D3-2C10	CDR-H3	KRSITTPY	97
8C2-2D6	CDR-H3	KRSITTSY	99
9E5-1A8	CDR-H3	KRSITTPY	97
5E6-2E7	CDR-H3	KRSITTPY	97
9F11-1F7	CDR-H3	KRSITTPY	97
4G8-1E3	CDR-H3	KRSITTSY	99
10F4-2F2	CDR-H3	KRSITTSY	99
10F4-2A9	CDR-H3	KRSITTSY	99
6B11-2E3	CDR-H3	KRSITTSY	99
2B8-2F10	CDR-H3	KRSITTSY	99
7A5-2G12	CDR-H3	KRSITTSY	99
7A6-2A2	CDR-L1	ENVVTY	100
2B2-2A7	CDR-L1	EDIYSN	101
5G2-1B3	CDR-L1	ENIYSH	102
4D9-2C11	CDR-L1	QNIN VW	103
2F4-1E2	CDR-L1	QSLVHSYGNTY	104
2F4-1H8	CDR-L1	QSLVHSYGNTY	104
1A5-2C9	CDR-L1	QNVGTA	105
1F6-1B7 (also sequence of 1F6- 1C10)	CDR-L1	QSL LN SG N Q K S Y	106
2C2-2C5	CDR-L1	QSVNNN	107
2G12-2B2	CDR-L1	QSL LN R G N H K N Y	108
2C6-2F11	CDR-L1	QSL V Q S N G N T Y	109
7D4-2A2-2F2	CDR-L1	ENVVNY	110

7D4-1H12-2B3	CDR-L1	ENVVNY	110
7D3-2C10	CDR-L1	QNIN VW	103
8C2-2D6	CDR-L1	ENVV TY	100
5E6-2E7	CDR-L1	QSL LNSGKTKNY	111
9F11-1F7	CDR-L1	QSL LNSGKTKNY	111
4G8-1E3	CDR-L1	QHIN FW	112
10F4-2F2	CDR-L1	ENVV TY	100
10F4-2A9	CDR-L1	ENVV TY	100
6B11-2E3	CDR-L1	ENVV TY	100
7A5-2G12	CDR-L1	ENVV IY	113
1C11-2G9	CDR-L1	QSL LNSGNQKNY	114
7A6-2A2	CDR-L2	GASN RYT	115
2B2-2A7	CDR-L2	KATN LAD	116
5G2-1B3	CDR-L2	GATN LAD	117
4D9-2C11	CDR-L2	KASN LHT	118
2F4-1E2	CDR-L2	KVSN RFS	119
2F4-1H8	CDR-L2	KVSN RFS	119
1A5-2C9	CDR-L2	SASN RYT	120
1F6-1B7 (also sequence of 1F6- 1C10)	CDR-L2	WAST RDS	121
2C2-2C5	CDR-L2	YASN RYT	122
2G12-2B2	CDR-L2	WAST RES	123
2C6-2F11	CDR-L2	KVSN RFC	124
7D4-2A2-2F2	CDR-L2	GASN RYS	125
7D4-1H12-2B3	CDR-L2	GASN RYS	125
7D3-2C10	CDR-L2	KVSN LHT	126
8C2-2D6	CDR-L2	GASN RYT	115
5E6-2E7	CDR-L2	WAST RES	123
9F11-1F7	CDR-L2	WAST RES	123
4G8-1E3	CDR-L2	KASN LHT	118
10F4-2F2	CDR-L2	GASN RYT	115
10F4-2A9	CDR-L2	GASN RYT	115
6B11-2E3	CDR-L2	GASN RYT	115
7A5-2G12	CDR-L2	GASN RYT	115
1C11-2G9	CDR-L2	WAST RES	123
7A6-2A2	CDR-L3	GQGYSYPYT	127
2B2-2A7	CDR-L3	QHFWGTPFT	128
5G2-1B3	CDR-L3	QHFWGAPFT	129
4D9-2C11	CDR-L3	QQGQSY PFT	130
2F4-1E2	CDR-L3	SQNT HVPYT	131
2F4-1H8	CDR-L3	SQNT HVPYT	131
1A5-2C9	CDR-L3	QQYSSY RLT	132
1F6-1B7 (also	CDR-L3	QSDYSYPYT	133

sequence of 1F6-1C10)			
2C2-2C5	CDR-L3	QQGYSSPWT	134
2G12-2B2	CDR-L3	QNDYTYPOYT	135
2C6-2F11	CDR-L3	SQSTHAPLT	136
7D4-2A2-2F2	CDR-L3	GSKWITSYPYT	137
7D4-1H12-2B3	CDR-L3	GARVTSYPYT	138
7D3-2C10	CDR-L3	QQDQSYPYT	139
8C2-2D6	CDR-L3	GQGYSYPYT	127
5E6-2E7	CDR-L3	KNDYSYPYT	140
9F11-1F7	CDR-L3	KNDYSYPYT	140
4G8-1E3	CDR-L3	QQDQSYPYM	141
10F4-2F2	CDR-L3	GQGYSYPYT	127
10F4-2A9	CDR-L3	GQGYSYPYT	127
6B11-2E3	CDR-L3	GQGYSYPYT	127
7A5-2G12	CDR-L3	GQGYSYPYT	127
1C11-2G9	CDR-L3	QNDYSYPYT	142
8C2-2D6	CDR-H1	GYTFTDHAIH	143
4G8-1E3	CDR-H1	GYIFTDHAIH	144
2G12-2B2	CDR-H1	GYTFTDHAIH	143
5G2-1B3	CDR-H1	GYTFTDHAIH	143
5E6-2E7	CDR-H1	GYTFTDHAIH	143
2C2-2C5	CDR-H1	GYTFTDHAIH	143
3F1	CDR-H1	GYTFTDHAIH	143
CC49	CDR-H1	GYTFTDHAIH	143
B72.3	CDR-H1	GYTFTDHAIH	143
Consensus	CDR-H1	GYTFTDHAIH	143
8C2-2D6	CDR-H2	YISPGNGDIKYNEKFKG	145
4G8-1E3	CDR-H2	YISPGNGDIKYNEKFKG	145
2G12-2B2	CDR-H2	YFSPGNDDIKYNEKFRG	146
5G2-1B3	CDR-H2	YFSPGNDDIKYNEKFKV	147
5E6-2E7	CDR-H2	YISPGNGDIKYNEKFKV	148
2C2-2C5	CDR-H2	YISPGNGDIKYNEKFKG	145
3F1	CDR-H2	YISPGNGDIKYNEKFKD	149
CC49	CDR-H2	YFSPGNDDFKYNEKFKG	150
B72.3	CDR-H2	YISPGNDDIKYNEKFKG	151
Consensus	CDR-H2	YISPGNGDIKYNEKFKG	145
8C2-2D6	CDR-H3	SITTSY	152
4G8-1E3	CDR-H3	SITTSY	152
2G12-2B2	CDR-H3	SLSTPY	153
5G2-1B3	CDR-H3	SYYGD	154
5E6-2E7	CDR-H3	SITTPY	155
2C2-2C5	CDR-H3	SITTPY	155
3F1	CDR-H3	SLLALDY	156

CC49	CDR-H3	SLNMAY	157
B72.3	CDR-H3	SYYPGH	158
Consensus	CDR-H3	SITTSY	152
8C2-2D6	CDR-L1	KASENVVTYVS	159
4G8-1E3	CDR-L1	HASQHINFWLS	160
2G12-2B2	CDR-L1	KSSQSLLNRGNHKNYLT	161
5G2-1B3	CDR-L1	RASENIYSHLA	162
5E6-2E7	CDR-L1	KSSQSLLNSGKTKNYLT	163
2C2-2C5	CDR-L1	KASQSVNNNVA	164
3F1	CDR-L1	KASQDVGTNIA	165
CC49	CDR-L1	KSSQSLLYSGNQKNYLA	166
B72.3	CDR-L1	RASENIYSNLA	167
8C2-2D6	CDR-L2	GASNRYT	115
4G8-1E3	CDR-L2	KASNLHT	118
2G12-2B2	CDR-L2	WASTRES	123
5G2-1B3	CDR-L2	GATNLAD	117
5E6-2E7	CDR-L2	WASTRES	123
2C2-2C5	CDR-L2	YASNRYT	122
3F1	CDR-L2	SASTRHT	168
CC49	CDR-L2	WASARES	169
B72.3	CDR-L2	AATNLAD	170
8C2-2D6	CDR-L3	GQGYSYPYT	127
4G8-1E3	CDR-L3	QQDQSYPYM	141
2G12-2B2	CDR-L3	QNDYTYPYT	135
5G2-1B3	CDR-L3	QHFWGAPFT	129
5E6-2E7	CDR-L3	KNDYSYPYT	140
2C2-2C5	CDR-L3	QQGYSSPWT	134
3F1	CDR-L3	QQYSSFPLT	171
CC49	CDR-L3	QQYYSYPLT	172
B72.3	CDR-L3	QHFWGTPYT	173

[00181] In some cases, antibodies or antigen binding fragments of the invention may be encoded by a nucleotide sequence comprising one or more of the variable domain sequences listed in Table 4. Residues labeled “N” may be absent or selected from nucleotides A, C, G or T. In some cases, antibodies or antigen binding fragments thereof may be encoded by a nucleotide sequence comprising a sequence with from about 50% to about 99.9% sequence identity (e.g. from about 50% to about 60%, from about 55% to about 65%, from about 60% to about 70%, from about 65% to about 75%, from about 70% to about 80%, from about 75% to about 85%, from about 80% to about 90%, from about 85% to about 95%, from about 90% to about 99.9%, from about 95% to about 99.9%, about 97%, about 97.5%, about 98%, about 98.5%, about 99%, about 99.5%, about 99.6%, about 99.7% or about 99.8%) with one or

more of the variable domain sequences listed in Table 4. In some cases, antibodies or antigen binding fragments thereof of the invention may be encoded by a nucleotide sequence comprising one or more fragments of any of the sequences listed in Table 4.

Table 4. Variable domain nucleotide sequences

Antibody ID Number	Variable chain	Sequence	SEQ ID NO
18D2	Heavy chain	CAGGTTACTCTGAAAGAGTCTGGCCCTGGGATA TTGCAGCCCCTCCCAGACCCTCAGTCTGACTTGTT CTTTCTCTGGGTTTTTCACTGAGCACTTCTAATAT GGGTATAGGCTGGATTTCGTCAGCCTTCAGGGAA GGGTCTAGAGTGGCTGGCACACATTTGGTGGCA TGATGATAAGTACTATAACCCATCCCTGAAGAG CCGGCTCACAATCTCCAAGGATATCTCCAACAA CCAGGTATTCCTCAAGATCACCAGTGTGGACAC TGCAGATACTGCCACGTACTACTGTGCTCAAGTC CCGTTTTACTACGGAACCTCGTTTCGATGTCTGGG GCACAGGGACCACGGTCACCGTCTCCTCA	174
18D2	Light chain 1	GACATTGTGATGACTCAGTCTCCAGCCACCCTGT CTGTGACTCCAGGAGATAGAGTCTCTCTTTCCTG CAGGGCCAGCCAGAGTATTAGCGACTACTTACA CTGGTATCAACAAAAATCACATGAGTCTCCAAG GCTTCTCATCAAATATGCTTCCCAATCCATCTCT GGGATCCCCTCCAGGTTTCAGTGGCAGTGGATCA GGGTCAGATTTCACTCTCAGTATCAACAGTGTG GAACCTGAAGATGTTGGAGTGTATTACTGTCAA AATGGTCACAGCTTTCCTCTCACGTTCCGGTGCTG GGACCAAGCTGGAGCTGAAAC	175
18D2	Light chain 2	CAAATTGTTCTCACCCAGTCTCCAGCAATCATGT CTGCATCTCCAGGGGAGACGGTCACCATGACCT GCAGTGCCAGCTCAAGTATAACTTACATGCACT GGTACCAGCAGAAGCCAGGCACCTCCCCCAAAA GATGGATTTATGACACATCCAAACTGGCTTCTG GAGTCCCTGCTCGCTTCAGTGGCAGTGGGTCTG GGACCTCTTATTCTCTCACAATCAGCAGCATGGA GGCTGAAGATGCTGCCACTTATTACTGCCATCA GCGGAGTAGTTACACGTTTCGGAGGGGGGACCAA GCTGGAAATAAAACG	176
18C7	Heavy chain	CAGGTTACTCTGAAAGAGTCTGGCCCTGGGATA TTGCAGCCCCTCCCAGACCCTCAGTCTGACTTGTT CTTTCTCTGGGTTTTTCACTGAGCACTTTTGGTAT GGGTGTAGGCTGGATTTCGTCAGCCTTCAGGGAA GGGTCTGGAGTGGCTGGCACACATTTGGTGGGA TGATGATAAGTACTATAACCCAGCCCTGAAGAG TCGGCTCACAATCTCCAAGGATACCTCCAAAAA CCAGGTATTCCTCAAGATCGCCAATGTGGACAC	177

		TGCAGATACTGCCACATACTACTGTGCTCGAAT AGCCTATTACTACGGTAGCGAGAGGGACTACTG GGGCCAAGGCACCACTCTCACAGTCTCCTCA	
18C7	Light chain	CAAATTGTTCTCACCCAGTCTCCAGCAATCATGT CTGCATCTCCAGGGGAGAAGGTCACCATGACCT GCAGTGCCAGCTCAAGTATAAGTTACATGCACT GGTACCACCAGAAGCCAGGCACCTCCCCCAAAA GATGGATTTATGACACATCCAAACTGGCTTCTG GAGTCCCTGCTCGCTTCAGTGGCAGTGGGTCTG GGACCTCTTATTCTCTCACAATCAGCAGCATGGA GGCTGAAGATGCTGCCACTTATTACTGCCATCA GCGGAGTAGTTACACGTTTCGGAGGGGGGACCAA GCTGGAAATAAAAACG	178
18D2(2)	Heavy chain	ATGGACAGGCTTACTTCCTCATTCTTGCTACTGA TTGTCCCTGCATATGTCCTGTCCCAGGTTACTCT GAAAGAGTCTGGCCCTGGGATATTGCAGCCCTC CCAGACCCTCAGTCTGACTTGTTCTTTCTCTGGG TTTTCACTGAGCACTTCTAATATGGGTATAGGCT GGATTCGTCAGCCTTCAGGGAAGGGTCTAGAGT GGCTGGCACACATTTGGTGGCATGATGATAAGT ACTATAACCCATCCCTGAAGAGCCGGCTCACAA TCTCCAAGGATATCTCCAACAACCAGGTATTCCT CAAGATCACCAAGTGTGGACACTGCAGATACTGC CACGTACTACTGTGCTCAAGTCCCGTTTTACTAC GGAACCTCGTTCGATGTCTGGGGCACAGGGACC ACGGTCACCGTCTCCTCAGCCAAAACGACACCC CCATCTGTCTATCCGCTCGCCCCCTGGATCTGCTG CCCAAATAACTCCATGGTGACCCTGGGATGCC TGGTCAAGGGCTATTTCCCTGAGCCAGTGACAG TGACCTGGAACCTCTGGATCCCTGTCCAGCGGTGT GCACACCTTCCCAGCTGTCCTGCAGTCTGACCTC TACACTCTGAGCAGCTCAGTGACTGTCCCCTCCA GCACCTGGCCCAGCGAGACCGTCACCTGCAACG TTGCCCACCCGGCCAGCAGCACCAAGGTGGACA AGACAATTGTGCCCAGGGATTGTGGTTGTAAGC CTTGCATATGTACAGTCCCAGAAGTATCATCTGT CTTCATCTTCCCCCCTAAAGCCCAAGGATGTGCTC ACCATTACTCTGACTCCTAAGGTCACGTGTGTTG TGGTAGACATCAGCCAGGATGATCCCGAGGTCA GTTTCAGCTGTTTGTAGATGATGTGGAAGTGCAC ACAGCTCAAAAACAACCCCCCGAGAGGACATTTC ACAACATTTCCGCTCATCAGTGAATTTCCCATCT GCACAAGACTGCTTAATGGCAAGAGTTAAATGC AGGTCAAAGGGCAGTTTCCTGCCCCATCAAAAA CTTTTCAAAA	179
18D2(2)	Light chain	ATGGATTTTCAAGTGCAGATTTTTCAGCTTCCTGC TAATCAGTGCCTCAGTCATACTGTCCAGAGGAC AAATTGTTCTCACCCAGTCTCCAGCAATCATGTC TGCATCTCCAGGGGAGACGGTCAACCATGACCTG	180

		CAGTGCCAGCTCAAGTATAACTTACATGCACTG GTACCAGCAGAAGCCAGGCACCTCCCCCAAAG ATGGATTTATGACACATCCAAACTGGCTTCTGG AGTCCCTGCTCGCTTCAGTGGCAGTGGGTCTGG GACCTCTTATTCTCTCACAATCAGCAGCATGGAG GCTGAAGATGCTGCCACTTATTACTGCCATCAGC GGAGTAGTTACACGTTTCGGAGGGGGGACCAAGC TGGAAATAAAACGGGCTGATGCTGCACCAACTG TATCCATCTTCCCACCATCCAGTGAGCAGTTAAC ATCTGGAGGTGCCTCAGTCGTGTGCTTCTTGAAC AACTTCTACCCCAAAGACATCAATGTCAAGTGG AAGATTGATGGCAGTGAACGACAAAATGGCGTC CTGAACAGTTGGACTGATCAGGACAGCAAAGAC AGCACCTACAGCATGAGCAGCACCCCTCACGTTG ACCAAGGACGAGTATGAACGACATAACAGCTAT ACCTGTGAGGCCACTCACAAGACATCAACTTCA CCCATTGTCAAGAGCTTCAACAGGAATGAGTGT TAG	
18C7(2)	Heavy chain	ATGGACAGGCTTACTTCCTCATTCCTGTTACTGA TTGTCCCTGCATATGTCCTGTCCCAGGTTACTCT GAAAGAGTCTGGCCCTGGGATATTGCAGCCCTC CCAGACCCTCAGTCTGACTTGTTCTTTCTCTGGG TTTTCACTGAGCACTTTTGGTATGGGTGTAGGCT GGATTCGTCAGCCTTCAGGGAAGGGTCTGGAGT GGCTGGCACACATTTGGTGGGATGATGATAAGT ACTATAACCCAGCCCTGAAGAGTCGGCTCACAA TCTCCAAGGATACCTCCAAAAACCAGGTATTCC TCAAGATCGCCAATGTGGACACTGCAGATACTG CCACATACTACTGTGCTCGAATAGCCTATTACTA CGGTAGCGAGAGGGACTACTGGGGCCAAGGCA CCTCTCACAGTCTCCTCAGCCAAAACGACAC CCCCATCTGTCTATCCGCTCGCCCCTGGATCTGC TGCCCAAATAACTCCATGGTGACCCTGGGATG CCTGGTCAAGGGCTATTTCCCTGAGCCAGTGAC AGTGACCTGGAACCTCTGGATCCCTGTCCAGCGG TGTGCACACCTTCCCAGCTGTCCTGCAGTCTGAC CTCTACACTCTGAGCAGCTCAGTGACTGTCCCCT CCAGCACCTGGCCCAGCGAGACCGTCACCTGCA ACGTTGCCCAACCCGGCCAGCAGCACCAAGGTGG ACAAGAAAATTGTGCCCAGGGATTGTGGTTGTA AGCCTTGCATATGTACAGTCCCAGAAGTATCAT CTGTCTTCATCTTCCCCCAAAGCCCAAGGATGT GCTCACCATTACTCTGACTCCTAAGGTCACGTGT GTTGTGGTAGACATCAGCAAGGATGATCCCGAG GTCCAGTTCAGCTGGTTTGTAGATGATGTGGAA GTGCACACAGCTCAGACGNCACCCCGGGGAGA GCAGTTTTCACAGCACTTTCCGCTCAGTCAGTGAA CTTCCCATCATGCACCANGACTGGGCTCATGGN CAGGAGTTCAANTGCAGGTCACAGTGCAGCTTT	181

		CCTGCCCCATCGAGAAACATCTCCNAAACAAGG CGACGAAAGCTCACAGGGTACACATTCCACTCC CNAGAGCAATGCCAGATAAGTCATCTGACTGCT GATACAACCTCTTCTGAAAATACTGTGAATGCAT GGATGCCACCACGAAAATCAAACCTCGCCCTTG GACNATGGCTTATTTTACCAGCTAGTCAAACCT GGGGGGAATTTCCCGTCTGTT	
18C7(2)	Light chain	ATGGTTTTTCACACCTCAGATACTTGGACTTATGC TTTTTTGGATTTTCAGCCTCCAGATGTGACATTGT GATGACTCAGTCTCCAGCCACCCTGTCTGTGACT CCAGGAGATAGAGTCTCTCTTTCCCTGCAGGGCC AGCCAGAGTATTAGCGACTACTTACACTGGTAT CAACAAAAATCACATGAGTCTCCAAGGCTTCTC ATCAAATATGCTTCCCAATCCATCTCTGGGATCC CCTCCAGGTTTCAGTGGCAGTGGATCAGGGTCAG ATTTCACTCTCAGTATCAACAGTGTGGAACCTGA AGATGTTGGAGTGTATTACTGTCAAATGGTCA CAGCTTTCCTCTCACGTTCCGGTGCTGGGACCAAG CTGGAAATAAAACGGGCTGATGCTGCACCAACT GTATCCATCTTCCCACCATCCAGTGAGCAGTTAA CATCTGGAGGTGCCTCAGTCGTGTGCTTCTTGAA CAACTTCTACCCCAAAGACATCAATGTCAAGTG GAAGATTGATGGCAGTGAACGACAAAATGGCGT CCTGAACAGTTGGACTGATCAGGACAGCAAAGA CAGCACCTACAGCATGAGCAGCACCTCACGTT GACCAAGGACGAGTATGAACGACATAACAGCTA TACCTGTGAGGGCCACTCACAAGACATCAACTTC ACCCATTGTCAAGAGCTTCAACAGGAATGAGTG TTAG	182
10A5- 2A12	Heavy chain	CAGGTGCAGCTGAAGGAGTCAGGACCTGGCCTG GTGGCGCCCTCACAGAGCCTGTCCATCACATGC ACTGTCTCAGGGTTCTCATTAACCAGCTATGGTG TAAGCTGGGTTCGCCAGCCTCCAGGAAAGGGTC TGGAGTGGCTGGGAGTAATATGGGGTGACGGAA GCACAAATTATCATTCTCTCATATCCAGACT GAGCATCAGCAAGGATAACTCCAAGAGCCAAGT TTTCTTAAAACTGAACAGTCTGCAAACCTGATGA CACAGCCACGTACTACTGTGCCAGAGCCTTTGTT TACTGGGGCCAAGGGACTCTGGTCACTGTCTCT GCA	183
10A5- 2A12	Light chain	CAAATTGTTCTCACCCAGTCTCCAGCAATCATGT CTGCATCTCCAGGGGAGAAGGTCACCATGACCT GCAGTGCCAGCTCAAGTGTAAGTTACATACT GGTACCAGCAGAAGTCAGGCACCTCCCCCAAAA GATGGATTTATGACACATCCAAACTGGCTTCTG GAGTCCCTGCTCGCTTCAGTGGCAGTGGGTCTG GGACCTCTTACTCTCTCACAATCAGCAGCATGG AGGCTGAAGATGCTGCCACTTATTACTGCCAGC AGTGGAGTAGTAACCCACCCATGCTCACGTTTCG	184

		GTGCTGGGACCAAGCTGGAGCTGAAAC	
8C11-1D10	Heavy chain	CAGGTGCAGCTGAAGGAGTCAGGACCTGGCCTG GTGGCGCCCTCACAGAGCCTGTCCATCACATGC ACTGTCTCAGGGTTCTCATTAACCAGCTATGGTG TAAGCTGGGTTCGCCAGCCTCCAGGAAAGGGTC TGGAGTGGCTGGGAGTAATATGGGGTGACGGGA GCACAAATTATCATTAGCTCTCATATCCAGACT GATCATCAGCAAGGATAACTCCAAGAGCCAAGT TTTCTTAAAACTGAACAGTCTGCAAACCTGATGA CACAGCCACCTACTACTGTACCAAAGGCTTTAC TACTGGGGCCAGGGGACTCTGGTCACTGTCTCT GCA	185
8C11-1D10	Light chain	CAAATTGTTCTCACCCAGTCTCCAGCAATCATGT CTGCATCTCCAGGGGAGAAGGTCACCATGACCT GCAGTGCCAGCTCAAGTGTAAGTTACATGCACT GGTACCAGCAGAAGTCAGGCACCTCCCCCAAAA GATGGATTTTGGACACATCCAAACTGGCTTCTGG AGTCCCTGCTCGCTTCAGTGGCAGTGGGTCTGG GACCTCTTACTCTCTCACAAATCAGCAGCATGGA GGCTGAAGATGCTGCCACTTATTACTGCCAGCA GTGGAGTAGTAACCTGCTCACGTTCCGGTGCTGG GACCAAGCTGGAGCTGAAAC	186
2D4-1B4	Heavy chain	CAGGTGCAGCTGCAGGAGTCAGGACCTGGCCTG GTGGCGCCCTCACAGAGCCTGTCCATCACATGC ACTGTCTCAGGGTTCTCATTAATCAGCTATGGTG TAAACTGGGTTCGCCAGCCTCCAGGAAAGGGTC TGGAGTGGCTGGGAGTGATATGGGGTGACGGGA GCACAAATTATCAGTCAGCTCTCATATCCAGACT GATCATCAGCAAGGATAACTCCAAGAGCCAAGT TTTCTTAAAACTGAACAGTCTGCAAACCTGATGA CACAGCCACGTACTACTGTACCAAAGGCTTTGC TACTGGGGCCAAGGGACTCTGGTCACTGTCTCT GCA	187
2D4-1B4	Light chain	CAAATTGTTCTCACCCAGTCTCCAGCAATCATGT CTGCATCTCCAGGGGAGAAGGTCACCATGACCT GCAGTGCCAGCTCAAGTGTAAGTTACATGCACT GGTTCCAGCAGAAGTCAGGCACCTCCCCCAAAA GATGGATTTATGACACATCCAAACTGGCTTCTG GAGTCCCTGCTCGCTTCAGTGGCAGTGGGTCTG GGACCTCTTACTCTCTCACAAATCAGCAGCATGG AGGCTGAAGATGCTGCCACTTATTACTGCCAGC AGTGGAGTAGTAACCTGCTCACGTTCCGGTGCTG GGACCAAGCTGGAGCTGAAAC	188
7G9-1A8	Heavy chain	CAGGTGCAGCTGAAGGAGTCAGGACCTGGCCTG GTGGCGCCCTCACAGAACCTGTCCATCACATGC ACTGTCTCAGGGTTCTCATTAACCAGTTATGGTG TAAACTGGGTTCGCCAGCCTCCAGGAAAGGGTC TGGAGTGGCTGGGAGTAATATGGGGTGACGGGA GCACAAATTATCATTAGCTCTCATTTCCAGACT	189

		GATCATCAGCAAGGAAAACCTCCAAGAGCCAAGT TTTCTTAAAACCTGAACAGTCTGCAAACATAATGA CACAGCCACGTATTACTGTACCAAAGGCTTTGTT TACTGGGGCCAAGGGACTCTGGTCACTGTCTCT GCA	
7G9-1A8	Light chain	CAAATTGTTCTCACCCAGTCTCCAGCAATCATGT CTGCATCTCCAGGGGAGAAGGTCACCATGACCT GCAGTGCCAGCTCAAGTGTAAGTTACATGCACT GGTACCAGCAGAAGTCAGGCACCTCCCCCAAAA GATGGATTTTTTGACACATCCAAACTGGCTTCTGG AGTCCCTGCTCGCTTCAGTGGCAGTGGGTCTGG GACCTCTTACTCTCTCACAATCAGCAGCATGGA GGCTGAAGATGCTGCCACTTATTACTGCCAGCA GTGGAGTAGTAACCTGCTCACGTTTCGGTGCTGG GACCAAGCTGGAGCTGAAAC	186
1A12- 2B2	Heavy chain	CAGGTGCAGCTGAAGGAGTCAGGACCTGGCCTG GTGGCGCCCTCACAGAGCCTGTCCATCACATGC ACTGTCTCAGGGTTCTCATTAACCAGCTATGGTG TAAGCTGGGTTCGCCAGCCTCCAGGAAAGGGTC TGGAGTGGCTGGGAGTAATATGGGGTGACGGGA GCACAAATTATCATTACAGCTCTCATATCCAGACT GAGCATCAGCAAGGATAACTCCAAGAGCCAAGT TTTCTTAAAACCTGAACAGTCTGCAAACATGATGA CACAGCCACGTACTACTGTGCCAAAGGGGGGCTA CTTTGACTACTGGGGCCAAGGCACCACTCTCAC AGTCTCCTCA	190
1A12- 2B2	Light chain	CAAATTGTTCTCACCCAGTCTCCAGCAGTCATGT CTGCATCTCCAGGGGAGAAGGTCGCCATAACCT GCAGTGCCAGCTCAAGTGTAAGTTACATGCACT GGTTCCAGCAGAAGCCAGGCACTTCTCCCAAAC TCTGGATTTATAGCACATCCAACCTGGCTTCTGG AGTCCCTGCTCGCTTCAGTGGCAGTGGATCTGG GACCTCTTACTCTCTCACAATCAGCCGAATGGA GGCTGAAGATGCTGCCACTTATTACTGCCAGCA AAGGAGTAGTTACCCGTGGACGTTTCGGTGGAGG CACCAAGCTGGAAATCAAAC	191
7D3- 2C10	Heavy chain	CAGGTTCAGTTGCTGCAGTATGACGCTGAGTTG GTGAAACCTGGGGGGGTGAGTGAAGATATCGTGC AAGGCCTCTGGCTACACCTTCACTGACCATGCT ATTCAGTGGGTGAAGCAGAAGCCTGAACAGGGC CTGGAATGGATTGGATATTTTTCTCCCGGAAATG ATGATATTAAGTACAGTGAGAAGTTCAAGGGCA AGGCCACACTGACTGCAGACAAGTCCTCCAGCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAGTGTATTTCTGTAAAAGATCCAT TACTACGCCTTACTGGGGCCAAGGGACTCTGGT CACTGTCTCTGCA	192
7D3- 2C10	Light chain	GACATCCAGATGAACCAGTCTCCATCCAGTCTG TCTGCATCCCTTGGAGACACAATTACCATCACTT	193

		GCCATGCCAGTCAGAACATTAATGTTTGGTTAA GCTGGTACCAGCAGAAACCAGGAAATATTCCTA AACTATTGATCTATAAGGTTTCCAACCTTGCACAC AGGCGTCCCATCAAGGTTTAGTGGCAGTGGATC TGGAACAGGTTTCACATTAACCATCAGCAGCCT GCAGCCTGAAGACATTGCCACTTACTACTGTCA ACAGGATCAAAGTTATCCGTACACGTTTCGGAGG GGGGACCAAGCTGAAAAAAA	
A5-2G12	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCCTCTGGCTACACCTTCACTGACCATGCT ATTCAGTGGGTGAAGCAGAAGCCTGAACAGGGC CTGGAATGGATTGGATATATTTCTCCCGGAAAT GATGATATTAAGTACAATGAGAAGTTCAAGGGC AAGGCCACACTGACTGCAGACAAATCCTCCAGC ACTGCCTACATGCAGCTCAACAGCCTGACATCT GAGGATTCTGCAGTGTATTTCTGTAAAAGATCC ATTACTACGTCTTACTGGGGCCAAGGGACTCTG GTCAGTGTCTCTGCA	194
A5-2G12	Light chain	AACATTGTAATGACCCAATCTCCCAAATCCATGT CCATGTTCAGTAGGAGAGAGGGGTCACCTTGACCT GCAAGGCCAGTGAGAATGTGGTTATTTATGTTTC CTGGTATCAACAGAAACCAGAGCAGTCTCCTAA ACTGCTGATATACGGGGGCATCCAACCGGTACAC TGGGGTCCCCGATCGCTTCACAGGCAGTGGATC TGCAACAGATTTCACTCTGACCATCAGCAGTGT GCAGGCTGAAGACCTTGCAGATTATCACTGTGG ACAGGGTTACAGCTATCCGTACACGTTTCGGAGG GGGGACCAAGCTGGAAATAAAACG	195
1A5-2C9	Heavy chain	CAGGTTCAGTTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCC ATTCATTGGGTGAAGCAGAAGCCTGAACAGGGC CTGGAATGGATTGGATATGTTTCTCCCGGAAAT GGTGATATTAAGTACAATGAGAAGTTCAAGGGC AAGGCCACACTGACTGCAGACAAATCCTCCAGC ACTGCCTACATGCAGCTCAACAGCCTGACATCG GAGGATTCTGCAGTGTATTTCTGTAAAAGATCTT TAATTGGAGACTATTGGGGCCAAGGCACCACTC TCACAGTCTCCTCA	196
1A5-2C9	Light chain	GACATTGTGATGACCCAGTCTCAAAAATTCATG TCCTCATCAGTAGGAGACAGGGTCACCATCACC TGCAAGGCCAGTCAGAATGTGGGTACTGCTGTA GCCTGGTATCAACAGAAACCAGGACAATCTCCT AAATTTCTGATTTACTCGGCATCCAATCGGTACA CTGGAGTCCCTGATCGCTTCACAGGCAGTGGAT CTGGGACAGATTTCACTCTCACGATCAGCAATA TGCAGTCTGAAGACCTGGCAGATTATTTCTGCCA GCAATATAGCAGCTATCGTCTGACGTTTCGGTGG	197

		AGGCACCAAGCTGGAAATCAAAC	
4D9-2C11	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAATTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGGCC TGGAATGGATTGGATATCTTTCTCCCGGAAATG ATGATATTAAGTACAGTGAGAAGTTCAAGGACA AGGCCACACTGACTGCAGACAAATCCTCCAGCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAGTGTATTTCTGTAAAAGATCCAT AGGGGGGGGACCACTGGGGGCAAGGCACCACTCT CACAGTCTCCTCA	198
4D9-2C11	Light chain	GACATCCAGATGAACCAGTCTCCATCCAGTCTG TCTGCATCCCTTGGAGACACAATTACCATCACTT GCCATGCCAGTCAGAACATTAATGTTTGGTTAA ACTGGTACCAGCAGAAACCAGGAAATATTCCTA AACTATTGATCTATAAGGCTTCCAACCTTGCACAC AGGCGTCCCATCAAGGTTTAGTGGCAGTGGATC TGGAACAGGTTTCACATTAACCATCGGCAGCCT GCAGCCTGAAGACATTGCCACTTACTACTGTCA ACAGGGTCAAAGTTATCCGTTACGTTTCGGAGG GGGGACCAAGCTGGAAATAAAACG	199
2F4-1E2	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAACAGAAGCCTGAACAGGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG GTGATATTAAGTATAATGAGAAGTTCAAGGGCA AGGCCACACTGACTGCAGACAAATCCTCCAGCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAGTGTATTTCTGTCAAAGACAAC GGGACAAGGCTACTGGGGCCAAGGCACCACTCT CACAGTCTCCTCA	200
2F4-1E2	Light chain	GATGTTGTGATGACCCAAACTCCACTCTCCCTGC CTGTCAGTCTTGGAGATCAAGCCTCCATCTCTTG CAGATCTAGTCAGAGCCTTGTACACAGTTATGG AAACACCTATTTACATTGGTACCTGCAGAAGCC AGGCCAGTCTCCAAAGCTCCTGATTTACAAAGT TTCCAACCGATTTTCTGGGGTCCCAGACAGGTTC AGTGGCAGTGGATCAGGGACAGATTTCACTC AAGATCAGCAGAGTGGAGGCTGAGGATCTGGG AGTTTATTTCTGCTCTCAAATAACACATGTTCCG TACACGTTTCGGAGGGGGGACCAAGCTGGAAATA AAACG	201
2F4-1H8	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAACAGAAGCCTGAACAGGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG	200

		GTGATATTAAGTATAATGAGAAGTTCAAGGGCA AGGCCACACTGACTGCAGACAAATCCTCCAGCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAGTGTATTTCTGTCAAAGACAAC GGGACAAGGCTACTGGGGCCAAGGCACCACTCT CACAGTCTCCTCA	
2F4-1H8	Light chain	GATGTTGTGATGACCCAAACTCCACTCTCCCTGC CTGTCAGTCTTGGAGATCAAGCCTCCATCTCTTG CAGATCTAGTCAGAGCCTTGTACACAGTTATGG AAACACCTATTTACATTGGTACCTGCAGAAGCC AGGCCAGTCTCCAAAGCTCCTGATTTACAAAGT TTCCAACCGATTTTCTGGGGTCCCAGACAGGTTC AGTGGCAGTGGATCAGGGACAGATTTCACACTC AAGATCAGCAGAGTGGAGGCTGAGGATCTGGG AGTTTATTTCTGCTCTCAAATAACACATGTTCCG TACACGTTTCGGAGGGGGGACCAAGCTGGAAATA AAACG	201
2C6- 2F11	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GGGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACACCTTCAGTGACCATGCTA TTCCTGGGTGAAGCAGAAGCCTGAACAGGGGCC TGGAATGGATTGGATATATCTCTCCCGGAAACG ATGATATTAAGTACAATGAGAAGTTCAAGGGCA AGGCCACACTGACTGCAGACAAATCCTCCAGCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAGTGTATTTCTGTGAAAGATCGAT GATTGGGGTTTACTGGGGCCAAGGGACTCTGGT CACTGTCTCTGCA	202
2C6- 2F11	Light chain	GATGTTGTGATGACCCAAACTCCACTCTCCCTGA CTGTCAGTCTTGGCGATCAAGCCTCCATCTCTTG CAGATTTAGTCAGAGCCTTGTACAAAGTAATGG AAATACCTATTTACAGTGGTATCTGCAGAAGCC AGGCCAGTCTCCAAAGCTCCTGATTTACAAAGT CTCCAACCGATTTTGTGGGGTCCCAGACAGGTTC AGTGGCAGTGGATCAGGGACAGATTTCACACTC AAGATCAGCAGAGTGGAGGCTGAGGATCTGGG AGTTTATTTCTGCTCTCAAAGTACACATGCTCCG CTCACGTTTCGGTGCTGGGACCAAGCTGGAGCTG AAAC	203
2B2-2A7	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGACTTCTGGCTACACCTTCACTGACCATGCA ATTCCTGGGTGAAGCAGAAGCCTGAACAGGGC CTGGAATGGATTGGATATATTTCTCCCGGAAAT GGTGATATTAAGTACAATGAGAAGTTCAAGGGC AAGGCCACCCTGACTGCAGACAAATCCTCCAGC ACTGCCTATATGCAGCTCAGCAGCCTGACACCT GAGGATTCTGCAGTGTATTTCTGTAAAATATCTT ACTACGGTATTTGGGGCCAAGGCACCACTCTCA	204

		CAGTCTCCTCA	
2B2-2A7	Light chain	GACATCCAGATGACTCAGTCTCCAGCCTCCCTAT CTGTATCTGTGGGAGAGTCTGTCACCATCACATG TCGACTAAGTGAAGATATTTACAGTAATTTAGC ATGGTTTCAGCAGAGACCGGGAAAATCTCCTCA GCTCCTGGTTTATAAAGCAACAAACTTAGCAGA CGGTGTGCCATCAAGGTTCAAGTGGCAGTGGATC AGGCACACAGTATTCCCTCAAGATCAACAGCCT GCAGTCTGAAGATTTTGGGACTTATTACTGTCAA CATTTTTGGGGTACTCCATTACGTTTCGGCTCGG GGACCAAGGTGGAAATAAAAC	205
5G2-1B3	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGGCC TGGAATGGATTGGATATTTTTCTCCCGGAAATGA TGATATTAAGTATAATGAGAAGTTCAAGGTCAA GGCCACACTGACTGCAGACAAATCCTCCAGCAC TGCCTACATGCAACTCACCAGCCTGACATCTGA AGATTCTGCAGTGTATTTCTGTAAAAGATCTTAC TACGGTGATTGGGGCCAAGGCACCACTCTCACA GTCTCCTCA	206
5G2-1B3	Light chain	GACATCCAGATGACTCAGTCTCCAGCCTCCCTAT CTGTTTCTGTGGGAGAACTGTCACCATCACATG TCGAGCAAGTGAGAATATTTACAGTCATTTAGC ATGGTATCAACAGAAACAGGGAAAATCTCCTCA ACTCCTGGTCTATGGTGCAACTAAGTACAGAT GGTGTGCCATCAAGGTTCAAGTGGCAGTGGATCA GGCACACAGTTTTCCCTCAAGATCCACAGCCTG CAGTCTGAAGATTTTGGGAGTTATTACTGTCAAC ATTTTTGGGGTGCTCCATTACGTTTCGGCTCGGG GACAAAGTTGGAAATAAAAC	207
7A6-2A2	Heavy chain	CAAATTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGACTTCAGTGAAGATGTCCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGGCC TGGAATGGATTGGATATTTTTCTCCCGGAAATGA TGATATTAAGTATAATGTGAAGTTCAAGGGCAA GGCCACACTGACTGCAGACAAATCCTCCAGCAC TGCCTACATGCAGCTCAACAGCCTGACATCTGA AGATTCTGCAGTGTATTTCTGTTCGGTGGGATAC GCCCTTGACTACTGGGGCCTAGGCACCACTCTC ACAGTCTCCTCA	208
7A6-2A2	Light chain	AACATTGTAATGACCCAATCTCCCAAATCCATGT CCATGTCAGTAGGAGAGAGGGTACCTTGACCT GCAAGGCCAGTGAGAATGTGGTTACTTATGTTT CCTGGTATCAACAGAAACCAGAGCAGTCTCCTA AACTGCTGATATACGGGGCATCCAACCGGTACA CTGGGGTCCCCGATCGCTTCACAGGCAGTGGAT	209

		CTGCAACAGATTTCACTCTGACCATCAGCAGTGT GCAGGCTGAAGACCTTGCAGATTATCACTGTGG ACAGGGTTACAGCTATCCGTACACGTTCCGGAGG GGGGACCAAGCTGGAAATAAAACG	
10C9- 2G7	Heavy chain	CAGGTTTCAGCTGCAACAGTCTGACGCTGAGTTG GTGAAACCTGGGACTACAGTGAAGATATCCTGC AAGGCTTCTGGCTACACTTTCACTGACCATGCTA TTCAGTGGGTGAAGGAGAAGCCTGAACAGGGGCC TGGAATGGATCGGATATATTTCTCCCGGAAATG ATGATATTAAGTACAGTGAGAAGTTCAAGGGCA AGGCCACACTGACTGCAGACAAATCCTCCAGCA CTGCTTACATGCAGCTCAACAGCCTGACATCTG ATGATTCTGCAGTGTATTTCTGTAAAAGATCGCT TAGTACGCCTTACTGGGGCCAAGGGACTCTGGT CACTGTCTCTGCA	210
10C9- 2G7	Light chain	TTTTTAATACGACTCCCTATAGGGCAAGCAGTG GTATCAATGCAGATTACAAGGGGGGAAAGGCATC AGACCAGCATGGGCATCAAGGTGGAATCACAGA CTCTGGTCTTCATATCCATACTGTTTGGGTTATA TGGAGCTGATGGGAACACATTAATGACCCAATC TCCCACATCCATGTACATGTCAGTAGGAGAGAG GGTCACCTTGACTTGCAAGGCCAGTGAGAATGA GATTAATTATGTTTCCTGGTATCAACAGAAACCA GAGCAGTCTCCTAAACTGTTGATATACGGGGCA TCCAACCGGTACTCTGGGGTCCCCGATCGCTTCA CAGGCAGTGGATCTGCAACAGATTTCACTCTGA CCATCAGCAGTGTGCAGGCTGAAGACCTTGCAG ATTATCCCTGTGGAGCAAGGGATTAAGTAGCTA TCCGTACACGTTCCGGAGGGGGGACCAAGCTGGA AATAAAACGGGC	211
1C11- 2G9	Heavy chain	Unknown	
1C11- 2G9	Light chain	GACATTGTGATGACACAGTCTCCATCCTCCCTGA CTGTGACAGCAGGAGAGAAGGTCAGTATGAGCT GCAGGTCCAGTCAGAGTCTGTAAACAGTGGAA ATCAAAAGAACTACTTGACCTGGTACCAGCAGA AACCAGGGCAGCCTCCTAAACTGTTGATCTACT GGGCATCCACTAGGGAATCTGGGGTCCCTGATC GCTTCACAGGCAGTGGATCTGGAACAGATTTCA CTCTCACCATCAGCAGTGTGCAGGCTGAAGACC TGGCAGTTTATTACTGTCAGAATGATTATAGTTA TCCGTACACGTTCCGGAGGGGGGACCAAGCTGGA AATAAAACG	212
1F6-1B7 (also sequence of 1F6- 1C10)	Heavy chain	CAGGTTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGATGCAGATGCCTGAACAGGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG GTGATGTAAAGTACAGTGAGAGGTTCAAGGGCA	213

		GGGCCACACTGACTGCAGACAAATCCTCCAGCT CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAGTTTATTTCTGTAAAAGATCGCT TAGTACGCCTTACTGGGGCCAAGGGACTCTGGT CACTGTCTCTG	
1F6-1B7 (also sequence of 1F6- 1C10)	Light chain	GACATTGTGATGACACAGTCTCCATCCTCCCTGA CTGTGACAGCAGGAGAGAGGGGTCACTATGAGCT GCAAGTCCAGTCAGAGTCTGTAAACAGTGGAA ATCAAAAGAGCTACTTGACCTGGTACCAGCAGA AACCAGGGCAGCCTCCTAAACTGTTGATCTCCT GGGCATCCACTAGGGATTCTGGGGTCCCTGATC GCTTCACAGGCAGTGGATCTGGAACAGATTTCA CTCTCACCATCAGCAGTGTGCAGGCTGAAGACC TGGCAGTTTATTACTGTCAGAGTGATTATAGTTA TCCGTACACGTTCTGGAGGGGGGACCAAGCTGGA AATAAAACG	214
2G12- 2B2	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGNTGAGTTG GTGAAACCGGGGGCTTCAGTGAAGATATCCTGT AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGCC TGGAATGGATTGGATATTTTTCTCCCGGAAATGA TGATATTAAGTACAATGAGAAGTTTAGGGGCAA GGCCACACTGACTGCAGACAAATCCTCCAGCAC TGCCTACATGCAGCTCAACAGCCTGTCATCTGAT GATTCTGCAGTGTATTTCTGTAAAAGATCGCTTA GTACGCCTTACTGGGGCCAAGGGACTCTGGNCA CTGTCTCTGCA	215
2G12- 2B2	Light chain	GACATTGTGATGACACAGTCTCCATCCTCCCTGA CTGTGACAGCAGGAGAGAAAGTCACTATGAGCT GCAAGTCCAGTCAGAGTCTGTAAACCGTGGAA ATCATAAGAAGTACTTGACCTGGTACCGGCAGA AACCAGGGCTGCCTCCTAAACTGTTGATTTACTG GGCATCCACTAGGGAATCTGGGGTCCCTGATCG CTTCACAGGCAGTGGATCTGGAACAGATTTTCGC TCTCACCATCAGCAGTGTTCAGGCTGAAGACCT GGCAGTTTATTACTGTCAGAATGATTATACTTAT CCGTACACGTTCTGGAGGGGGGACCAAGCTGGAG ATAAAACG	216
5E6-2E7	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAATGAAGATTTCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG GTGATATTAAGTACAATGAGAAGTTCAAGGTCA AGGCCACACTGACTGCAGACAAATCCTCCAGCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAGTGTATTTCTGTAAAAGATCGAT TACTACGCCTTACTGGGGCCAAGGGACTCTGGT CACTGTCTCTGCA	217

5E6-2E7	Light chain	GACATTGTGATGACACAGTCTCCATCCTCCCTGA CTGTGACAGCAGGAGAGAAGGTCCTATGAGCT GCAAGTCCAGTCAGAGTCTGTAAACAGTGGAA AAACAAAGAACTACTTGACGTGGTACCAGCAGA AACCAGGGCAGCCTCCTAAACTGTTGATCTACT GGGCATCCACTAGGGAATCTGGGGTCCCTGATC GCTTCACAGGCAGTGGATCTGGAACAGATTTCA CTCTCACCATCAGCAGTGTGCAGGCTGAAGACC TGGCAGTTTATTACTGTAAGAATGATTATAGTTA TCCGTACACGTTTCGGAGGGGGGACCAAGCTGGA AATAAAACG	218
9E5-1A8	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAATTG GTGAAGCCTGGGGCTTCAGTGAAGATATCCTGC AAGACTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGGCC TGGAATGGATTGGATATATCTCTCCCGGAAATG ATGATATTAAGTACACTGAGAAGTTCAAGGGCA AGGTCACACTGACTGCAGACAAATCCTCCAGCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAGTCTATTTCTGTAAAAGATCGAT TACTACGCCTTACTGGGGCCAAGGGACTCTGGT CACTGTCTCTGCA	219
9E5-1A8	Light chain	TTTTTATACGCCACTTTCTAATACGCCTCACTAT AGGGCAAGCAGTGGTATCAACGCAGATTACAAA GGGGAAAGGAATCAGACCGACTCGCGCATCAA GATGGAATCACAGACTCTGGTCTTCATATCCAGT ACGCTCGGGGACTATGGAGNGGAACAGTACATT TTAATGACCCAATGTCCCAAAGGCAAGAACATG TCAGTAGGAGAGAGGGTCACTCAGAGTGCAAGG CCAGGAGAAATCAAAACACTTATGTTTCCTGGT ATCAACAGAAACCAGAGCANNCTNTAAAATGN NGATTACGGGGCATCCAACCGGGAATCTGGGGT CNCCGATCGCTTCACAGGCAGTGGATCTGGAAC AGATTTCACTCTCACCATCAGCAGTGTGCAGGCT GAAGACCNGGCAGTNTTCACTGTGGACAGGGNT ACAGTTATCCGTACACGTTTCGGAGGGGGGACCA AGCTGAAAAAAACGGGC	220
9F11-1F7	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAATGAAGATTTCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG GTGATATTAAGTACAATGAGAAGTTCAAGGTCA AGGCCACACTGACTGCAGACAAATCCTCCAGCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAGTGTATTTCTGTAAAAGATCGAT TACTACGCCTTACTGGGGCCAAGGGACTCTGGT CACTGTCTCTGCA	217
9F11-1F7	Light	GACATTGTGATGACACAGTCTCCATCCTCCCTGA	218

	chain	CTGTGACAGCAGGAGAGAAGGTCACTATGAGCT GCAAGTCCAGTCAGAGTCTGTAAACAGTGGAA AAACAAAGAACTACTTGACGTGGTACCAGCAGA AACCAGGGCAGCCTCCTAAACTGTTGATCTACT GGGCATCCACTAGGGAATCTGGGGTCCCTGATC GCTTCACAGGCAGTGGATCTGGAACAGATTTCA CTCTCACCATCAGCAGTGTGCAGGCTGAAGACC TGGCAGTTTATTACTGTAAGAATGATTATAGTTA TCCGTACACGTTCTGGAGGGGGGACCAAGCTGGA AATAAAACG	
10F4-2F2	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG GTGATATTAAGTACGATGAGAAGTTTAAGGGCA AGGCCACACTGACTGCAGACAAATCCTCCTCCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AAGATTCTGCAGTGTATTTCTGTAAAAGATCGAT TACTACCTCTTACTGGGGCCAAGGGACTCTGGTC ACTGTCTCTGCA	221
10F4-2F2	Light chain	AACATTGTAATGACCCAATCTCCCAAATCCATGT CCATGTTCAGTAGGAGAGAGGGTCCACCTTGACCT GCAAGGCCAGTGAGAATGTGGTTACTTATGTTT CCTGGTATCAACAGAAACCAGAGCAGTCTCCTA AACTGCTGATATACGGGGGCATCCAACCGGTACA CTGGGGTCCCCGATCGCTTCACAGGCAGTGGAT CTGCAACAGATTTCACTCTGACCATCAGCAGTGT GCAGGCTGAAGACCTTGCAGATTATCACTGTGG ACAGGGTTACAGCTATCCGTACACGTTCTGGAGG GGGGACCAAGCTGGAAATAAAACG	209
2B8- 2F10	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG ATGATATTAAGTACAATGAGAAGTTCAAGGGCA AGGCCACACTGACTGCAGACAAGTCCTCCAGCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAGTGTTTTTCTGTAAAAGATCGAT TACTACCTCTTACTGGGGCCAAGGGACTCTGGTC ACTGTCTCTGCA	222
2B8- 2F10	Light chain	TTNATAGGACTCAATATAGGGCAAGCAGTGGTA TTAACGCCGAGTACATGGGGAGGGCAAGGGCA GAAAGTCACTTTCAGTGAGGATACACCATCAGC ATGAGGGTCCCTTGTTGAGCTCCTGGGGGGGCTG GTGTTNTGCTTTTTAGGTGTGAGATGTGACATCC AGATGAACCAGTCTCCATCCAGTCTGTNTGCATC CTTTGGAGACACAATTACCATCATTTGCCATTCC	223

		AGTCAGAACATTAATGTTTGGTTAAGATGGTAC CAGCAGAAACCAGGAAATATTCCTAAAATATTG ATATATAAGGGTTCCAACCTTGTACACAGGCGTC CCATCAAGGTTTAGTGGCAGTGGATTTGGAACA GGTTTCACATTAACCATCAGCAGCGTGCAGCGG GAAGACATTGCCACTTACTACTGTCAACAGGAT CAAAGTTATCCGTACACGTTCCGGAGGGGGGACC AAGCTGAAATAAAAACGGGC	
4G8-1E3	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCCGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACATCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG GTGATATTAAGTACAATGAGAAGTTCAAGGGCA AGGCCACACTGACTGCAGACAAATCCTCCAGCA CTGCCTACATGCATCTCAACAGCCTGACATCTG AGGATTCTGCAGTGTATTTCTGTAAAAGATCGAT TACTACCTCTTACTGGGGCCAAGGGACTCTGGTC ACTGTCTCTGCA	224
4G8-1E3	Light chain	GACATCCAGATGAACCAGTCCCCATCCAGTCTG TCTGCATCCCTTGGAGACACAATTACCATCACTT GCCATGCCAGTCAGCACATTAATTTTTTGGTTAAG CTGGTACCAGCAGAAACCAGGAAATATTCCTAA ACTCTTGATCTATAAGGCTTCCAACCTGCACACA GGCGTCCCATCAAGGTTTAGTGGCAGTGGATCT GGAACAGGTTTCACATTAACCATCAGCAGCCTG CTGCCTGAAGACGTTGCCACTTACTACTGTCAAC AGGATCAAAGTTATCCGTATATGTTCCGGAGGGG GGACCAAGCTGGAAATAAAAACG	225
6B11- 2E3	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG ATGATATTAAGTACAATGAGAAGTTTAAGGGCA AGGCCACACTGACTGCAGACAAATCCTCCAGCA CTGCCTACATGCTGCTCAACAGCCTGACATCTG AGGATTCTGCAGTGTATTTCTGTAAAAGATCGAT TACTACCTCTTACTGGGGCCAAGGGACTCTGGTC ACTGTCTCTGCA	226
6B11- 2E3	Light chain	AACATTGTAATGACCCAATCTCCCAAATCCATGT CCATGTCAAGTAGGAGAGAGGGGTCACCTTGACCT GCAAGGCCAGTGAGAATGTGGTTACTTATGTTT CCTGGTATCAACAGAAACCAGAGCAGTCTCCTA AACTGCTGATATACGGGGGCATCCAACCGGTACA CTGGGGTCCCCGATCGCTTCACAGGCAGTGGAT CTGCAACAGATTTCACTTTGACCATCAGCAGTGT GCAGGCTGAAGACCTTGCAGATTATCACTGTGG ACAGGGTTACAGCTATCCGTACACGTTCCGGAGG	227

		GGGGACCAAGCTGGAAATAAAACG	
7D3-2C10	Heavy chain	CAGGTTCAGTTGCTGCAGTATGACGCTGAGTTG GTGAAACCTGGGGGGTCAGTGAAGATATCGTGC AAGGCCTCTGGCTACACCTTCACTGACCATGCT ATTCAGTGGGTGAAGCAGAAGCCTGAACAGGGC CTGGAATGGATTGGATATTTTTCTCCCGGAAATG ATGATATTAAGTACAGTGAGAAGTTCAAGGGCA AGGCCACACTGACTGCAGACAAGTCCTCCAGCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAGTGTATTTCTGTAAAAGATCCAT TACTACGCCTTACTGGGGCCAAGGGACTCTGGT CACTGTCTCTGCA	192
7D3-2C10	Light chain	GACATCCAGATGAACCAGTCTCCATCCAGTCTG TCTGCATCCCTTGGAGACACAATTACCATCACTT GCCATGCCAGTCAGAACATTAATGTTTGGTTAA GCTGGTACCAGCAGAAACCAGGAAATATTCCTA AACTATTGATCTATAAGGTTTCCAACCTTGCACAC AGGCGTCCCATCAAGGTTTAGTGGCAGTGGATC TGGAACAGGTTTCACATTAACCATCAGCAGCCT GCAGCCTGAAGACATTGCCACTTACTACTGTCA ACAGGATCAAAGTTATCCGTACACGTTCCGGAGG GGGGACCAAGCTGAAAAAAAA	193
7A5-2G12	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCCTCTGGCTACACCTTCACTGACCATGCT ATTCAGTGGGTGAAGCAGAAGCCTGAACAGGGC CTGGAATGGATTGGATATATTTCTCCCGGAAAT GATGATATTAAGTACAATGAGAAGTTCAAGGGC AAGGCCACACTGACTGCAGACAAATCCTCCAGC ACTGCCTACATGCAGCTCAACAGCCTGACATCT GAGGATTCTGCAGTGTATTTCTGTAAAAGATCC ATTACTACGTCTTACTGGGGCCAAGGGACTCTG GTCAGTGTCTCTGCA	194
7A5-2G12	Light chain	AACATTGTAATGACCCAATCTCCCAAATCCATGT CCATGTCAGTAGGAGAGAGGGTCACCTTGACCT GCAAGGCCAGTGAGAATGTGGTTATTTATGTTTC CTGGTATCAACAGAAACCAGAGCAGTCTCCTAA ACTGCTGATATACGGGGGCATCCAACCGGTACAC TGGGGTCCCCGATCGCTTCACAGGCAGTGGATC TGCAACAGATTTCACTCTGACCATCAGCAGTGT GCAGGCTGAAGACCTTGCAGATTATCACTGTGG ACAGGGTTACAGCTATCCGTACACGTTCCGGAGG GGGGACCAAGCTGGAAATAAAACG	195
1A5-2C9	Heavy chain	CAGGTTCAGTTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCC ATTCATTGGGTGAAGCAGAAGCCTGAACAGGGC CTGGAATGGATTGGATATGTTTCTCCCGGAAAT GGTGATATTAAGTACAATGAGAAGTTCAAGGGC	196

		AAGGCCACACTGACTGCAGACAAATCCTCCAGC ACTGCCTACATGCAGCTCAACAGCCTGACATCG GAGGATTCTGCAGTGTATTTCTGTAAAAGATCTT TAATTGGAGACTATTGGGGCCAAGGCACCACTC TCACAGTCTCCTCA	
1A5-2C9	Light chain	GACATTGTGATGACCCAGTCTCAAAAATTCATG TCCTCATCAGTAGGAGACAGGGTCACCATCACC TGCAAGGCCAGTCAGAATGTGGGTACTGCTGTA GCCTGGTATCAACAGAAACCAGGACAATCTCCT AAATTTCTGATTTACTCGGCATCCAATCGGTACA CTGGAGTCCCTGATCGCTTCACAGGCAGTGGAT CTGGGACAGATTTCACTCTCACGATCAGCAATA TGCAGTCTGAAGACCTGGCAGATTATTTCTGCCA GCAATATAGCAGCTATCGTCTGACGTTTCGGTGG AGGCACCAAGCTGGAAATCAAAC	197
8C2-2D6	Heavy chain	CAGGTTCAACTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG GTGATATTAAGTACAATGAGAAGTTCAAGGGTA AGGCCACACTGACTGCAGACACTTCCTCCACCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAATGTATTTCTGTAAAAGATCCAT TACTACGTCTTACTGGGGCCAAGGGACTCTGGT CACTGTCTCTGCA	228
8C2-2D6	Light chain	AACATTGTAATGACCCAATCTCCCAAATCCATGT CCATGTCAGTAGGAGAGAGGGTCACCTTGACCT GCAAGGCCAGTGAGAATGTGGTTACTTATGTTT CCTGGTATCAACAGAAACCAGAGCAGTCTCCTA AACTGCTGATATACGGGGCATCCAACCGGTACA CTGGGGTCCCCGATCGCTTCACAGGCAGTGGAT CTGCAACAGATTTCACTCTGACCATCAGCAGTGT GCAGGCTGAAGACCTTGCAGATTATCACTGTGG ACAGGGTTACAGCTATCCGTACACGTTTCGGAGG GGGGACCAAGCTGGAAATAAAACG	209
7D4- 2A2-2F2	Heavy chain	CAGGTTCACTGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACATCTTCACTGACCATGCAA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG GTGATATTAAGTACATTGAGAAGTTCAAGGGGCA AGGCCACACTGACTGCAGACAAATCCTCCAGCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAGTGTATTTCTGTAAAAGATCGCT TAGTACGCCTTACTGGGGCCAAGGGACTCTGGT CACTGTCTCTGCA	229
7D4- 2A2-2F2	Light chain	AACATTTTAATGACCCAATCTCCCAAATCCATGT CCATGTCAGTAGGAGAGAGGGTCACCTTGACCT	230

		GCAAGGCCAGTGAGAATGTGGTTAATTATGTTT CCTGGTATCAACAGAAACCAGAGCAGTCTCCTA AACTGCTGATATTCGGGGCATCCAACCGGTACT CTGGGGTCCCCGATCGCTTCACAGGCAGTGGAT CTGCAACAGATTTCACTCTGACCATCAGCAGTGT GCAGGCTGAAGACCTTGCAGATTATCACTGTGG AAGCAAGTGGATTACTAGCTATCCGTACACGTT CGGAGGGGGGACCAAGCTGGAAATAAAACG	
7D4- 1H12- 2B3	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACATCTTCACTGACCATGCAA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG GTGATATTAAGTACATTGAGAAGTTCAGGGGCA AGGCCACACTGACTGCAGACAAATCCTCCAGCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AGGATTCTGCAGTGTATTTCTGTAAAAGATCGCT TAGTACGCCTTACTGGGGCCAAGGGACTCTGGT CACTGTCTCTGCA	229
7D4- 1H12- 2B3	Light chain	AACATTTTAATGACCCAATCTCCCAAATCCATGT CCATGTTCAGTAGGAGAGAGGGTACCTTGACCT GCAAGGCCAGTGAGAATGTGGTTAATTATGTTT CCTGGTATCAACAGAAACCAGAGCAGTCTCCTA AACTGCTGATATACGGGGCATCCAACCGGTACT CTGGGGTCCCCGATCGCTTCACAGGCAGTGGAT CTGCAACAGATTTCACTCTGACCATCAGCAGTGT GCAGGCTGAAGACCTTGCAGATTATCACTGTGG AGCAAGGGTTACTAGCTATCCGTACACGTTTCGG AGGGGGGACCAAGCTGGAAATAAAACG	231
2C2-2C5	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGACTTCAGTGAAGATATCCTGC AGGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG GTGATATTAAGTACAATGAGAAGTTCAAGGGCA AGGCCACACTGACTGCAGACAAATCCTCCAGCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG ACGATTCTGCAGTGTATTTCTGTAAAAGATCCAT TACTACGCCTTACTGGGGCCAAGGCACCACTCT CACAGTCTCCTCA	232
2C2-2C5	Light chain	AGTTTTGTGATGACCCAGACTCCCAAATTCCTGC TTGTGTTCAGCAGGAGACAGGGTTACCATAACCT GCAAGGCCAGTCAGAGTGTGAATAATAATGTAG CTTGGTACCAACAGAAGCCAGGGCAGTCTCCTA AACAGCTGATATACTATGCATCCAATCGCTACA CTGGAGTCCCTGATCGCTTCACTGGCAGTGGAT ATGGGACGGATTTCACTTTCACCATCTACACTGT GCAGGCTGAAGACCTGGCAGTTTATTTCTGTCA GCAGGGTTATAGCTCTCCGTGGACGTTTCGGTGG	233

		AGGCACCAAGCTGAAA	
10F4-2A9	Heavy chain	CAGGTTCAGCTGCAGCAGTCTGACGCTGAGTTG GTGAAACCTGGGGCTTCAGTGAAGATATCCTGC AAGGCTTCTGGCTACACCTTCACTGACCATGCTA TTCAGTGGGTGAAGCAGAAGCCTGAACAGGGGCC TGGAATGGATTGGATATATTTCTCCCGGAAATG GTGATATTAAGTACGATGAGAAGTTTAAGGGCA AGGCCACACTGACTGCAGACAAATCCTCCTCCA CTGCCTACATGCAGCTCAACAGCCTGACATCTG AAGATTCTGCAGTGTATTTCTGTAAAAGATCGAT TACTACCTCTTACTGGGGCCAAGGGACTCTGGTC ACTGTCTCTGCA	221
10F4-2A9	Light chain	AACATTGTAATGACCCAATCTCCCAAATCCATGT CCATGTCAGTAGGAGAGAGGGGTCACCTTGACCT GCAAGGCCAGTGAGAATGTGGTTACTTATGTTT CCTGGTATCAACAGAAACCAGAGCAGTCTCCTA AACTGCTGATATACGGGGCATCCAACCGGTACA CTGGGGTCCCCGATCGCTTCACAGGCAGTGGAT CTGCAACAGATTTCACTCTGACCATCAGCAGTGT GCAGGCTGAAGACCTTGCAGATTATCACTGTGG ACAGGGTTACAGCTATCCGTACACGTTCCGGAGG GGGGACCAAGCTGGAAATAAAACG	209

[00182] In some cases, antibodies or antigen binding fragments of the invention may comprise the IgG2a heavy chain and/or kappa light chain constant domain sequences listed in Table 5. In some cases, antibodies or fragments thereof may comprise an amino acid sequence with from about 50% to about 99.9% sequence identity (e.g. from about 50% to about 60%, from about 55% to about 65%, from about 60% to about 70%, from about 65% to about 75%, from about 70% to about 80%, from about 75% to about 85%, from about 80% to about 90%, from about 85% to about 95%, from about 90% to about 99.9%, from about 95% to about 99.9%, about 97%, about 97.5%, about 98%, about 98.5%, about 99%, about 99.5%, about 99.6%, about 99.7% or about 99.8%) with one or more of the constant domain sequences listed in Table 5. In some cases, antibodies or fragments thereof of the invention may comprise an amino acid sequence comprising one or more fragments of any of the sequences listed in Table 5.

Table 5. Constant domain sequences

Domain	Sequence	SEQ ID NO
IgG2a heavy	AKTTAPSVYPLAPVCGDTTGSSVTLGCLVKGYFPEPV TLTWNSGSLSSGVHTFPAVLQSDLYTLSSSVTVTSST	234

chain constant domain	WPSQSITCNVAHPASSTKVDKKIEPRGPTIKPCPPCKC PAPNLLGGPSVFIFPPKIKDVLMSLSPIVTCVVVDVSE DDPDVQISWVFNNEVHTAQTQTHREDYNSTLRVVS ALPIQHQDWMSGKEFKCKVNNKDLPAPIERTISKPKG SVRAPQVYVLPPPEEEMTKKQVTLTCMVTDMPEDI YVEWTNNGKTELNYKNTEPVLDSDGSYFMYSKLRV EKKNWVERNSYSCSVVHEGLHNHHTTKSFSRTPGK	
kappa light chain constant domain	RADAAPTVSIFPPSSEQLTSGGASVVCFLNNFYPKDIN VKWKIDGSERQNGVLNSWTDQDSKDYSTYSMSSTLTL TKDEYERHNSYTCEATHKTSTSPIVKSFNRECE	235

[00183] In some cases, antibodies may comprise the heavy chain sequence of QVQLQQSDAELVKPGASVKISCKASGYTFTDHAHWVKQKPEQGLDWIGYISPGNG DIKYNEKFKDKVTLTADKSSSTACMHLNSLTSEDSAVYFCKRSLALDYWGQGTTL TVSSAKTTAPSVYPLAPVCGDTTGSSVTLGCLVKGYFPEPVTLTWNSGSLSSGVHTFP AVLQSDLYTLSSSVTVTSSTWPSQSITCNVAHPASSTKVDKKIEPRGPTIKPCPPCKCP APNLLGGPSVFIFPPKIKDVLMSLSPIVTCVVVDVSEDDPDVQISWVFNNEVHTAQ TQTHREDYNSTLRVVSALPIQHQDWMSGKEFKCKVNNKDLPAPIERTISKPKGSVRA PQVYVLPPPEEEMTKKQVTLTCMVTDMPEDIYVEWTNNGKTELNYKNTEPVLDSD GSYFMYSKLRVEKKNWVERNSYSCSVVHEGLHNHHTTKSFSRTPGK (SEQ ID NO: 236) which may be encoded by the nucleotide sequence

ATGGAGACCGACACCCTGCTGCTCTGGGTGCTGCTGCTCTGGGTGCCCGGCTCCA
CCGGACAGGTTTCAGCTGCAGCAGTCTGACGCTGAGTTGGTGAAACCTGGGGCTTC
AGTGAAGATATCCTGCAAGGCTTCTGGCTACACCTTCACTGACCATGCTATTCAC
TGGGTGAAGCAAAAGCCTGAACAGGGCCTGGACTGGATTGGATATATTTCTCCC
GGAAATGGTGATATTAAGTACAATGAGAAGTTCAAGGACAAGGTCACACTGACT
GCAGACAAATCCTCCAGCACTGCCTGCATGCACCTCAACAGCCTGACATCTGAGG
ATTCTGCAGTGTATTTCTGCAAAAGATCCCTACTAGCTCTTGACTACTGGGGCCA
AGGCACCACTCTCACAGTCTCCTCAGCTAAAACAACAGCCCCATCGGTCTATCCA
CTGGCCCCCTGTGTGTGGAGATACTGGCTCCTCGGTGACTCTAGGATGCCTGG
TCAAGGGTTATTTCCCTGAGCCAGTGACCTTGACCTGGAACCTCTGGTTCCCTGTCC
AGTGGTGTGCACACCTTCCCAGCTGTCCTGCAGTCTGACCTCTACACCCTCAGCT
CAAGCGTGACTGTAACCAGCTCGACCTGGCCCAGCCAGTCCATCACCTGCAATGT
GGCCCACCCGGCAAGCAGCACCAAGGTGGACAAGAAAATTGAGCCCAGAGGGC
CCACAATCAAGCCCTGTCCTCCATGCAAATGCCAGCACCTAACCTCTTGGGTGG

ACCATCCGTCTTCATCTTCCCTCCAAAGATCAAGGATGTACTCATGATCTCCCTGA
 GCCCCATAGTCACATGTGTAGTCGTTGATGTGAGCGAGGATGACCCAGATGTCCA
 GATCAGCTGGTTTGTGAACAACGTGGAAGTGCACACTGCTCAGACACAGACGCA
 TAGAGAGGATTACAACAGTACTCTCCGGGTTGTCAGTGCCCTCCCCATCCAGCAC
 CAGGACTGGATGAGTGGCAAGGAGTTCAAATGCAAGGTCAACAACAAAGACCTC
 CCAGCGCCCATCGAGAGAACCATCTCAAAACCCAAAGGGTCAGTAAGAGCTCCA
 CAGGTATATGTCTTGCCTCCACCAGAAGAGGAGATGACTAAGAAACAGGTCACT
 CTGACCTGCATGGTCCACAGACTTCATGCCTGAAGACATTTACGTGGAGTGGACCA
 ACAACGGGAAAACAGAGCTAAACTACAAGAACACTGAACCAGTCCTGGACTCTG
 ATGGTTCTTACTTCATGTACAGCAAGCTGAGAGTGGAGAAGAAGAAGTGGGTGG
 AGAGAAATAGCTACTCCTGTTTCAGTGGTCCACGAGGGTCTGCACAATCACCACAC
 GACTAAGAGCTTCTCCCGGACTCCGGGTAAATAG (SEQ ID NO: 237) or an

optimized version thereof. In some cases, antibodies of the invention may comprise the light chain amino acid sequences of

DIVMTQSHKFMSTSVGDRVSITCKASQDVGTNIAWYQQKPGRSPKVLIYSASTRHTG
 VPDRFTGSGSGTDFTLTISNVQSEDLTDYFCQQYSSFPLTFGVGKLELKRADAAPTV
 SIFPPSSEQLTSGGASVVCFLNNFYPKDINVKWKIDGSRQNGVLNSWTDQDSKDST
 YSMSSTLTLTKDEYERHNSYTCEATHKTSTSPIVKSFNRNEC (SEQ ID NO: 238),

which may be encoded by the nucleotide sequence

ATGGAGACCGACACCCTGCTGCTCTGGGTGCTGCTGCTCTGGGTGCCCGGCTCCA
 CCGGAGACATTGTGATGACCCAGTCTCACAAATTCATGTCCACATCAGTAGGAGA
 CAGGGTCAGCATCACCTGCAAGGCCAGTCAGGATGTGGGCACTAATATAGCCTG
 GTATCAACAGAAACCAGGCCGATCTCCTAAAGTACTGATTTACTCGGCATCCACC
 CGGCACACTGGAGTCCCTGATCGCTTCACAGGCAGTGGATCTGGGACAGATTTCA
 CTCTCACCATTAGCAATGTGCAGTCTGAAGACTTGACAGATTATTTCTGTCAGCA
 ATATAGCAGCTTTCCTCTCACGTTCCGGTGTTGGGACCAAGCTGGAGCTGAAACGG
 GCAGATGCTGCACCAACTGTATCCATCTTCCCACCATCCAGTGAGCAGTTAACAT
 CTGGAGGTGCCTCAGTCGTGTGCTTCTTGAACAACCTTACCCCCAAAGACATCAA
 TGTCAAGTGGAAGATTGATGGCAGTGAACGACAAAATGGCGTCCTGAACAGTTG
 GACTGATCAGGACAGCAAAGACAGCACCTACAGCATGAGCAGCACCCCTCACGTT
 GACCAAGGACGAGTATGAACGACATAACAGCTATACCTGTGAGGCCACTCACAA
 GACATCAACTTCACCCATTGTCAAGAGCTTCAACAGGAATGAGTGTTGA (SEQ ID
 NO: 239) or an optimized version thereof. In some cases, antibodies or fragments thereof

may comprise an amino acid sequence with from about 50% to about 99.9% sequence identity (e.g., from about 50% to about 60%, from about 55% to about 65%, from about 60% to about 70%, from about 65% to about 75%, from about 70% to about 80%, from about 75% to about 85%, from about 80% to about 90%, from about 85% to about 95%, from about 90% to about 99.9%, from about 95% to about 99.9%, about 97%, about 97.5%, about 98%, about 98.5%, about 99%, about 99.5%, about 99.6%, about 99.7% or about 99.8%) with one or more of the amino acid sequences of SEQ ID NOs: 236 and/or 227. In some cases, antibodies or fragments thereof of the invention may comprise an amino acid sequence comprising one or more fragments of any of the sequences of SEQ ID NOs: 236 and/or 238. In some cases, antibodies or fragments thereof may be encoded by a nucleotide sequence with from about 50% to about 99.9% sequence identity (e.g. from about 50% to about 60%, from about 55% to about 65%, from about 60% to about 70%, from about 65% to about 75%, from about 70% to about 80%, from about 75% to about 85%, from about 80% to about 90%, from about 85% to about 95%, from about 90% to about 99.9%, from about 95% to about 99.9%, about 97%, about 97.5%, about 98%, about 98.5%, about 99%, about 99.5%, about 99.6%, about 99.7% or about 99.8%) with one or more of the nucleotide sequences of SEQ ID NOs: 237 and/or 239.

IgG synthesis

[00184] IgG antibodies (e.g. IgG1, IgG2, IgG3 or IgG4) comprising one or more variable domain and/or CDR amino acid sequences presented herein (or fragment or variants thereof) may be synthesized for further testing and/or product development. Such antibodies may be produced by insertion of one or more segments of cDNA encoding desired amino acid sequences into expression vectors suited for IgG production. Expression vectors may comprise mammalian expression vectors suitable for IgG expression in mammalian cells. Mammalian expression of IgGs may be carried out to ensure that antibodies produced comprise modifications (e.g. glycosylation) characteristic of mammalian proteins and/or to ensure that antibody preparations lack endotoxin and/or other contaminants that may be present in protein preparations from bacterial expression systems.

Cancer-related targets

[00185] In some embodiments, targets of the present invention are cancer-related antigens or epitopes. As used herein, the term “cancer-related” is used to describe entities that may be in some way associated with cancer, cancerous cells and/or cancerous tissues. Many cancer-

related antigens or epitopes comprising glycans have been identified that are expressed in correlation with tumor cells (Heimburg-Molinaro, J. et al., Cancer vaccines and carbohydrate epitopes. Vaccine. 2011 Nov 8;29(48):8802-26). These are referred to herein as “tumor-associated carbohydrate antigens” or “TACAs.” TACAs include, but are not limited to mucin-related antigens [including, but not limited to Tn, Sialyl Tn (STn) and Thomsen-Friedenreich antigen], blood group Lewis related antigens [including, but not limited to Lewis^Y (Le^Y), Lewis^X (Le^X), Sialyl Lewis^X (SLe^X) and Sialyl Lewis^A (SLe^A)], glycosphingolipid-related antigens [including, but not limited to Globo H, stage-specific embryonic antigen-3 (SSEA-3) and glycosphingolipids comprising sialic acid], ganglioside-related antigens [including, but not limited to gangliosides GD2, GD3, GM2, fucosyl GM1 and Neu5GcGM3] and polysialic acid-related antigens. Many of such antigens are described in International Patent Application No. PCT/US2011/021387, the contents of which are herein incorporated by reference in their entirety.

[00186] In some embodiments, TACA targets of the present invention include Lewis blood group antigens. Lewis blood group antigens comprise a fucose residue linked to GlcNAc by an α 1-3 linkage or an α 1-4 linkage. They may be found on both glycolipids and glycoproteins. Lewis blood group antigens may be found in the body fluid of individuals that are secretors of these antigens. Their appearance on red cells is due to absorption of Lewis antigens from the serum by the red cells.

[00187] In some embodiments, TACA targets of the present invention comprise Le^Y. Le^Y (also known as CD174) is made up of Gal β 1,4GlcNAc comprising α 1,2- as well as α 1,3-linked fucose residues yielding the Fuca(1,2)Gal β (1,4)Fuca(1,3)GlcNAc epitope. It is synthesized from the H antigen by α 1,3 fucosyltransferases which attach the α 1,3 fucose to the GlcNAc residue of the parent chain. Le^Y may be expressed in a variety of cancers including, but not limited to ovarian, breast, prostate, colon, lung, and epithelial. Due to its low expression level in normal tissues and elevated expression level in many cancers, the Le^Y antigen is an attractive target for therapeutic antibodies.

[00188] In some embodiments, TACA targets of the present invention comprise Le^X. Le^X comprises the epitope Gal β 1-4(Fuca1-3)GlcNAc β -R. It is also known as CD15 and stage-specific embryonic antigen-1 (SSEA-1). This antigen was first recognized as being immunoreactive with sera taken from a mouse subjected to immunization with F9 teratocarcinoma cells. Le^X was also found to correlate with embryonic development at specific stages. It is also expressed in a variety of tissues both in the presence and absence of

cancer, but can also be found in breast and ovarian cancers where it is only expressed by cancerous cells.

[00189] In some embodiments, TACA targets of the present invention comprise SLe^A and/or SLe^X. SLe^A and SLe^X comprise the structures [Neu5Ac α 2-3Gal β 1-3(Fuc α 1-4)GlcNAc β -R] and [Neu5Ac α 2-3Gal β 1-4(Fuc α 1-3)GlcNAc β -R] respectively. Their expression is upregulated in cancer cells. The presence of these antigens in serum correlates with malignancy and poor prognosis. SLe^X is mostly found as a mucin terminal epitope. It is expressed in a number of different cancers including breast, ovarian, melanoma, colon, liver, lung and prostate. In some embodiments of the present invention, SLe^A and SLe^X targets comprise Neu5Gc (referred to herein as GcSLe^A and GcSLe^X, respectively).

[00190] In some cases, cancer-related targets of the invention may include mucins. *Ishida et al* demonstrate that interaction of MUC2 with dendritic cells (with anti-tumor activity) leads to dendritic cell apoptosis (Ishida, A. et al., 2008. *Proteomics*. 8: 3342-9, the contents of which are herein incorporated by reference in their entirety). In some aspects, the present invention provided anti-mucin antibodies to prevent dendritic cell apoptosis and support anti-tumor activity.

[00191] In some embodiments, TACA targets of the present invention comprise glycolipids and/or epitopes present on glycolipids, including, but not limited to glycosphingolipids. Glycosphingolipids comprise the lipid ceramide linked to a glycan by the ceramide hydroxyl group. On the cell membrane, glycosphingolipids form clusters referred to as “lipid rafts”.

[00192] In some embodiments, TACA targets of the present invention comprise Globo H. Globo H is a cancer-related glycosphingolipid first identified in breast cancer cells. The glycan portion of Globo H comprises Fuc α (1-2)Gal β (1-3)GalNAc β (1-3)Gal α (1-4)Gal β (1-4)Glc β (1). Although found in a number of normal epithelial tissues, Globo H has been identified in association with many tumor tissues including, but not limited to, small cell lung, breast, prostate, lung, pancreatic, gastric, ovarian and endometrial tumors.

[00193] In some embodiments, cancer-related glycosphingolipid targets of the present invention include gangliosides. Gangliosides are glycosphingolipids comprising sialic acid. According to ganglioside nomenclature, G is used as an abbreviation for ganglioside. This abbreviation is followed by the letters M, D or T referring to the number of sialic acid residues attached (1, 2 or 3 respectively). Finally the numbers 1, 2 or 3 are used to refer to the order of the distance each migrates when analyzed by thin layer chromatography (wherein 3 travels the greatest distance, followed by 2 and then 1). Gangliosides are known to be

involved in cancer-related growth and metastasis and are expressed on the cell surface of tumor cells. Gangliosides expressed on tumor cells include, but are not limited to GD2, GD3, GM2 and fucosyl GM1 (also referred to herein as Fuc-GM1). In some embodiments of the present invention, glycan-interacting antibodies are directed toward GD3. GD3 is a regulator of cell growth. In some embodiments, GD3-directed antibodies are used to modulate cell growth and/or angiogenesis. In some embodiments, GD3-directed antibodies are used to modulate cell attachment. GD3 associated with some tumor cells may comprise 9-O-acetylated sialic acid residues (Mukherjee, K. et al., 2008. J Cell Biochem. 105: 724-34 and Mukherjee, K. et al., 2009. Biol Chem. 390: 325-35, the contents of each of which are herein incorporated by reference in their entirety). In some cases, antibodies of the invention are selective for 9-O-acetylated sialic acid residues. Some antibodies may be specific for 9-O-acetylated GD3s. Such antibodies may be used to target tumor cells expressing 9-O-acetylated GD3. In some embodiments of the present invention, glycan interacting antibodies are directed toward GM2. In some embodiments, GM2-directed antibodies are used to modulate cell to cell contact. In some embodiments, ganglioside targets of the present invention comprise Neu5Gc. In some embodiments, such targets may include a GM3 variant comprising Neu5Gc (referred to herein as GcGM3). The glycan component of GcGM3 is Neu5Gc α 2-3Gal β 1-4Glc. GcGM3 is a known component of tumor cells (Casadesus, A.V. et al., 2013. Glycoconj J. 30(7):687-99, the contents of which are herein incorporated by reference in their entirety).

[00194] In some embodiments, tumor-associated carbohydrate antigens of the present invention comprise Neu5Gc.

Immunogenic hosts

[00195] In some embodiments, glycan-interacting antibodies of the present invention may be developed through the use of non-human animals as hosts for immunization, referred to herein as “immunogenic hosts”. In some embodiments, immunogenic hosts are mammals. In some embodiments, immunogenic hosts are transgenic knockout mice. Antigens comprising target sites and/or epitope targets of glycan-interacting antibodies may be used to contact immunogenic hosts in order to stimulate an immune response and produce antibodies in the immunogenic host that specifically bind the target sites and/or epitope targets present on the antigens introduced.

[00196] According to some methods of the present invention, the development of anti-STn antibodies may comprise immunizing mice that have had the *Cmah* gene disrupted. Such mutations may result in more human-like physiology in that Neu5Gc, the immunogenic, non-human form of sialic acid, is no longer produced in such mice. Also provided is a *Cmah*^{-/-} myeloma cell for producing a hybridoma that is free of Neu5Gc expression, for production of a GcSTn monoclonal antibody either by reducing the amount of recoverable anti-GcSTn or the hybridoma will begin to die due to antibody binding back to the hybridoma. Other genes can be knocked out in the background of *Cmah*^{-/-} myeloma cells. For example, the alpha1,3-galactosyltransferase gene, which encodes an enzyme critical for the formation of an epitope highly-immunogenic to humans (Chung, C.H. et al., Cetuximab-induced anaphylaxis and IgE specific for galactose-alpha-1,3-galactose. N Engl J Med. 2008 Mar 13;358(11):1109-17), can be knocked out in the background of *Cmah*^{-/-} myeloma cells.

[00197] According to other methods of the present invention, wild type mice may be used for immunization. Such methods may sometimes be favorable for the production of antibodies that interact with AcSTn or pan-STn epitopes. In some cases, immune responses in wild type mice may be more robust.

[00198] Antibodies produced through immunization may be isolated from serum of the immunogenic hosts. Antibody producing cells from the immunogenic hosts may also be used to generate cell lines that produce the desired antibody. In some embodiments, screening for antibodies and/or antibody producing cells from the immunogenic host may be carried out through the use of enzyme-linked immunosorbent assays (ELISAs) and/or glycan arrays.

Adjuvants

[00199] Immunization of immunogenic hosts with antigens described herein may comprise the use of one or more adjuvants. Adjuvants may be used to elicit a higher immune response in such immunogenic hosts. As such, adjuvants used according to the present invention may be selected based on their ability to affect antibody titers.

[00200] In some embodiments, water-in-oil emulsions may be useful as adjuvants. Water-in-oil emulsions may act by forming mobile antigen depots, facilitating slow antigen release and enhancing antigen presentation to immune components. Freund's adjuvant may be used as complete Freund's adjuvant (CFA), which comprises mycobacterial particles that have been dried and inactivated, or as incomplete Freund's adjuvant (IFA), lacking such particles. Other water-in-oil-based adjuvants may include EMULSIGEN® (MVP Technologies,

Omaha, NE). EMULSIGEN® comprises micron sized oil droplets that are free from animal-based components. It may be used alone or in combination with other adjuvants, including, but not limited to aluminum hydroxide and CARBIGEN™ (MVP Technologies, Omaha, NE).

[00201] In some embodiments, TITERMAX® adjuvant may be used. TITERMAX® is another water-in-oil emulsion comprising squalene as well as sorbitan monooleate 80 (as an emulsifier) and other components. In some cases, TITERMAX® may provide higher immune responses, but with decreased toxicity toward immunogenic hosts.

[00202] Immunostimulatory oligonucleotides may also be used as adjuvants. Such adjuvants may include CpG oligodeoxynucleotide (ODN). CpG ODNs are recognized by Toll-like receptor 9 (TLR9) leading to strong immunostimulatory effects. Type C CpG ODNs induce strong IFN- α production from plasmacytoid dendritic cell (pDC) and B cell stimulation as well as IFN- γ production from T-helper (T_H) cells. CpG ODN adjuvant has been shown to significantly enhance pneumococcal polysaccharide (19F and type 6B)-specific IgG2a and IgG3 in mice. CpG ODN also enhanced antibody responses to the protein carrier CRM197, particularly CRM197-specific IgG2a and IgG3 (Chu et al., *Infection Immunity* 2000, vol 68(3):1450-6). Additionally, immunization of aged mice with pneumococcal capsular polysaccharide serotype 14 (PPS14) combined with a CpG-ODN restored IgG anti-PPS14 responses to young adult levels (Sen et al., *Infection Immunity*, 2006, 74(3):2177-86). CpG ODNs used according to the present invention may include class A, B or C ODNs. In some embodiments, ODNs may include any of those available commercially, such as ODN-1585, ODN-1668, ODN-1826, ODN-2006, ODN-2007, ODN-2216, ODN-2336, ODN-2395 and/or ODN-M362, each of which may be purchased, for example, from InvivoGen, (San Diego, CA). In some cases, ODN-2395 may be used. ODN-2395 is a class C CpG ODN that specifically stimulated human as well as mouse TLR9. These ODNs comprise phosphorothioate backbones and CpG palindromic motifs.

[00203] In some embodiments, immune stimulating complexes (ISCOMs) may be used as adjuvants. ISCOMs are spherical open cage-like structures (typically 40 nm in diameter) that are spontaneously formed when mixing together cholesterol, phospholipids and Quillaia saponins under a specific stoichiometry. ISCOM technology is proven for a huge variety of antigens from large glycoproteins such as gp340 from Epstein-Barr virus (a 340 kDa antigen consisting of 80% carbohydrates) down to carrier-conjugated synthetic peptides and small haptens such as biotin. Some ISCOMs are capable of generating a balanced immune response

with both T_{H1} and T_{H2} characteristics. Immune response to ISCOMs is initiated in draining lymph nodes, but is efficiently relocated to the spleen, which makes it particularly suitable for generating monoclonal antibodies as well. In some embodiments, the ISCOM adjuvant AbISCO-100 (Isconova, Uppsala, Sweden) may be used. AbISCO-100 is a saponin-based adjuvant specifically developed for use in immunogenic hosts, such as mice, that may be sensitive to other saponins.

[00204] According to embodiments of the present invention, adjuvant components of immunization solutions may be varied in order to achieve desired results. Such results may include modulating the overall level of immune response and/or level of toxicity in immunogenic hosts.

Antibody sequence and structural analysis and optimization

[00205] In some embodiments, antibodies of the present invention may be subjected to sequence analysis and/or structural analysis wherein they are analyzed for characteristics that may affect affinity, specificity, protein folding, stability, manufacturing, expression, and/or immunogenicity (i.e., immune reactions in subjects being treated with such antibodies). Such analysis may include comparisons between antibodies binding to the same or similar epitopes.

[00206] Antibodies sequences of antibodies binding to the same epitope may be analyzed for variation in light and/or heavy chain sequences. Such analysis may include germline sequences and/or CDR sequences. Information obtained from such analysis may be used to identify (and optionally to modify, delete, replace or repair) conserved amino acid residues; conserved segments of amino acids; amino acid positions with conserved side chain characteristics; conserved CDR lengths; and other features conserved among antibodies binding to the same epitope. This information may be used to design variants or to inform antibody optimization procedures to improve antibody affinity, specificity, protein folding, stability, manufacturing, expression and/or immunogenicity.

[00207] Sequence analysis may include aligning two or more antibodies that bind to the same or similar epitopes to identify similarities. Such analysis may compare the sequence and/or length of antibody regions (e.g., CDRs, variable domains, germline segments). Amino acid insertions, amino acid deletions, and substitutions may be identified and assessed. Sequence differences may be compared against antibody affinity and/or specificity.

[00208] In some cases, sequence analyses are conducted to identify (and optionally to modify, delete, replace or repair) one or more of unpaired cysteines or irregular disulfides; glycosylation sites (e.g., N-linked NXS/T sites); acid cleavage sites, amino acid oxidation sites, conformity with mouse germline sequences; asparagine deamidation sites; aspartate isomerization sites; N-terminal pyroglutamate formation sites; and aggregation-prone patches in CDRs.

[00209] In some cases, the present invention provides sequence analysis-informed variants of antibodies presented herein. As used herein, the term “sequence analysis-informed variant” refers to an antibody variant that has been modified based on one or more conclusions derived from antibody sequence analysis. In some cases, antibodies of the invention may be modified to produce antibody variants comprising modifications to one or more of antibody affinity, specificity, protein folding, stability, manufacturing, expression and/or immunogenicity.

[00210] Some sequence analysis-informed variants comprise one or more CDR length modification. CDR length modified antibodies may comprise one or more added or deleted amino acids in one or more CDRs relative to an original antibody sequence. In some cases, sequence analysis-informed variants may comprise a substitution of one or more CDRs with one or more CDRs derived from another antibody (e.g., an antibody binding to the same or similar epitope). In some cases, sequence analysis-informed variants may comprise a substitution of a heavy or light chain variable domain from another antibody (e.g., an antibody binding to the same or similar epitope). Sequence analysis-informed variants may comprise modifications to one or more germline genes that the antibody is expressed from. Such modifications may include point mutations, regional mutations, insertional mutations or deletional mutations. In some case, germline gene modifications are carried out to move CDRs from one known germline gene to another. Sequence analysis-informed variants may include other variants described herein, including, but not limited to scFvs, monobodies, diabodies, intrabodies, CARs, antibody mimetics, etc.

[00211] In some embodiments, sequence and/or structural analysis may be used to inform the construction of antibody fragment display libraries (including, but not limited to scFv libraries, phage display libraries, and yeast display libraries). In one example, sequence alignment may be carried out to align two or more antibodies with a common antigen or epitope and amino acid residues may be identified that are conserved among the aligned antibodies or that are variable among the aligned antibodies. In such cases, antibody fragment

display libraries may be constructed such that variability among library members is primarily limited to the variable amino acids identified in the sequence analysis. In some cases, such libraries may be used to identify variants with altered affinity and/or specificity for a target antigen (e.g., STn) or a specific epitope of the target antigen (e.g., the epitopes recognized by Group 1, 2, 3 and 4 antibodies as described in Example 1, hereinbelow).

[00212] In some embodiments, antibodies of the invention may be modified to remove, replace or otherwise eliminate one or more unpaired cysteine residues. In some cases, unpaired cysteine residues may be reactive and in some cases may affect antibody affinity and/or specificity. Accordingly, some antibodies of the invention have been modified to eliminate unpaired cysteine residues. In some cases, such variants may have modified epitope specificity and/or affinity. In some cases, modification of unpaired cysteine residues may alter antibody folding. In some cases, these variants comprise a substitution or deletion of one or more cysteine residues. In some cases, these variants comprise one or more additional amino acid residues (including, but not limited to, the addition of one or more cysteine residues) to prevent or reduce undesired effects from unpaired cysteine residues. In some cases, cysteine residues are replaced with an amino acid having a hydrophobic side chain (e.g., tyrosine, alanine, valine, isoleucine, leucine, methionine, phenylalanine or tryptophan).

Antibody testing and characterization

[00213] Antibodies described herein may be tested and/or characterized using a variety of methods. Such methods may be used to determine a variety of characteristics that may include, but are not limited to, antibody affinity; specificity; and activity (e.g., activation or inhibition of cellular signaling pathways or other cellular or biological activities).

Cell-based assays

[00214] In some embodiments, antibodies of the present invention may be tested or characterized through the use of one or more cell-based assays. Such cell-based assays may be carried out in vitro with cells in culture. In some cases, cell-based assays may be carried out in vivo. Examples of cell-based in vivo assays include tumor models in which tumor cells are injected or otherwise introduced into a host.

[00215] In some cases, cells used in cell-based assays may express one or more target glycans recognized by one or more antibodies of the invention. Such glycans may be naturally expressed by such cells or, alternatively, cells may be induced to express one or more glycans desired for purposes of a particular assay. Induced expression may be through

one or more treatments that upregulate expression of glycosylated proteins or enzymes that regulate glycosylation. In other cases, induced expression may include transfection, transduction, or other form of introduction of one or more genes or transcripts for the endogenous expression of one or more glycosylated proteins or enzymes involved in regulation of glycosylation.

[00216] In some cases, cell-based assays used herein may include the use of cancer cells. Many cancer cell lines are available for experiments to test antibodies of the invention. Such cells may express target glycan or may be induced to express target glycans. Additionally, cancer cell lines may be used to test antibodies of the invention, where the cancer cell lines are representative of cancer stem cells. Cancer stem cell (CSC) cell lines may be isolated or differentiated from cancer cells grown in culture (e.g., through sorting based on markers specific for cancer stem cells).

[00217] In some embodiments, ovarian cancer cell lines may be used. Such cell lines may include, but are not limited to SKOV3, OVCAR3, OV90 and A2870 cell lines. In some cases, CSC cells may be isolated from these cell lines by isolating cells expressing CD44 and/or CD133 cell markers.

[00218] OVCAR3 cells were first established using malignant ascites obtained from a patient suffering from progressive ovarian adenocarcinoma (Hamilton, T.C. et al., 1983. Cancer Res. 43: 5379-89). Cancer stem cell populations may be isolated from OVCAR3 cell cultures through selection based on specific cell surface markers such as CD44 (involved in cell adhesion and migration), CD133 and CD117 (Liang, D. et al., 2012. BMC Cancer. 12: 201, the contents of which are herein incorporated by reference in their entirety). OV90 cells are epithelial ovarian cancer cells that were similarly derived from human ascites (see US Patent No. 5,710,038). OV-90 cells may also express CD44 when activated (Meunier, L. et al., 2010. Transl Oncol. 3(4): 230-8).

[00219] In some embodiments, cell lines derived from gastric cancers may be used. Such cell lines may include, but are not limited to SNU-16 cells (see description in Park J.G. et al., 1990. Cancer Res. 50: 2773-80, the contents of which are herein incorporated by reference in their entirety). SNU-16 cells express STn naturally, but at low levels.

Glycan arrays

[00220] In some embodiments, glycan-interacting antibodies of the present invention may be developed through the use of glycan arrays. As used herein, the term “glycan array” refers

to a tool used to identify agents that interact with any of a number of different glycans linked to the array substrate. In some embodiments, glycan arrays comprise a number of chemically-synthesized glycans, referred to herein as “glycan probes”. In some embodiments, glycan arrays comprise at least 2, at least 5, at least 10, at least 20, at least 30, at least 40, at least 50, at least 60, at least 70, at least 80, at least 90, at least 100, at least 150, at least 350, at least 1000 or at least 1500 glycan probes. In some embodiments, glycan arrays may be customized to present a desired set of glycan probes. In some embodiments, glycan probes may be attached to the array substrate by a linker molecule. Such linkers may comprise molecules including, but not limited to $-O(CH_2)_2CH_2NH_2$ and $O(CH_2)_3NHCOCH_2(OCH_2CH_2)_6NH_2$.

[00221] In some embodiments, a glycan array has more than 70 chemically-synthesized glycans, most of which are presented as Neu5Ac and Neu5Gc-containing glycan pairs. Some examples of glycan probes may include: Neu5Ac- α -2-6-GalNAc (AcSTn); Neu5Gc- α -2-6-GalNAc (GcSTn); Neu5,9Ac2- α -2,6-GalNAc; Neu9Ac5Gc- α -2,6-GalNAc, and GalNAc (Tn). The antibody binding specificity to AcSTn vs. GcSTn can be determined using the array or other methods of determining specificity known in the art. In addition, the binding profile of antibodies to O-acetylated STn can be determined. The loss of O-acetylation on STn is relevant to cancer as cancer-associated expression correlates with increased STn recognition by antibodies (Ogata, S. et al., Tumor-associated sialylated antigens are constitutively expressed in normal human colonic mucosa. *Cancer Res.* 1995 May 1;55(9):1869-74). In some cases, glycan arrays may be used to determine recognition of STn vs. Tn.

Antibody fragment display library screening techniques

[00222] In some embodiments, antibodies of the present invention may be produced and/or optimized using high throughput methods of discovery. Such methods may include any of the display techniques (e.g. display library screening techniques) disclosed in International Patent Application No. WO2014074532, the contents of which are herein incorporated by reference in their entirety. In some embodiments, synthetic antibodies may be designed, selected or optimized by screening target antigens using display technologies (e.g. phage display technologies). Phage display libraries may comprise millions to billions of phage particles, each expressing unique antibody fragments on their viral coats. Such libraries may provide richly diverse resources that may be used to select potentially hundreds of antibody fragments with diverse levels of affinity for one or more antigens of interest (McCafferty, et al., 1990. *Nature*. 348:552-4; Edwards, B.M. et al., 2003. *JMB*. 334: 103-18; Schofield, D. et al., 2007.

Genome Biol. 8, R254 and Pershad, K. et al., 2010. Protein Engineering Design and Selection. 23:279-88; the contents of each of which are herein incorporated by reference in their entirety). Often, the antibody fragments present in such libraries comprise scFv antibody fragments, comprising a fusion protein of V_H and V_L antibody domains joined by a flexible linker. In some cases, scFvs may contain the same sequence with the exception of unique sequences encoding variable loops of the complementarity determining regions (CDRs). In some cases, scFvs are expressed as fusion proteins, linked to viral coat proteins (e.g. the N-terminus of the viral pIII coat protein). V_L chains may be expressed separately for assembly with V_H chains in the periplasm prior to complex incorporation into viral coats. Precipitated library members may be sequenced from the bound phage to obtain cDNA encoding desired scFvs. Such sequences may be directly incorporated into antibody sequences for recombinant antibody production, or mutated and utilized for further optimization through in vitro affinity maturation.

Development of cytotoxic antibodies

[00223] In some embodiments, antibodies of the present invention may be capable of inducing antibody-dependent cell-mediated cytotoxicity (ADCC) and/or antibody-dependent cell phagocytosis (ADCP). ADCC is an immune mechanism whereby cells are lysed as a result of immune cell attack. Such immune cells may include CD56⁺ cells, CD3⁻ natural killer (NK) cells, monocytes and neutrophils (Strohl, W.R. Therapeutic Antibody Engineering. Woodhead Publishing, Philadelphia PA. 2012. Ch. 8, p186, the contents of which are herein incorporated by reference in their entirety).

[00224] In some cases, antibodies of the present invention may be engineered to comprise a given isotype depending on whether or not ADCC or ADCP is desired upon antibody binding. Such antibodies, for example, may be engineered according to any of the methods disclosed by Alderson, K.L. et al., J Biomed Biotechnol. 2011. 2011:379123). In the case of mouse antibodies, different isotypes of antibodies are more effective at promoting ADCC. IgG2a, for example, is more effective at inducing ADCC than is IgG2b. Some antibodies of the present invention, comprising mouse IgG2b antibodies may be reengineered to comprise IgG2a antibodies. Such reengineered antibodies may be more effective at inducing ADCC upon binding cell-associated antigens.

[00225] In some embodiments, genes encoding variable regions of antibodies developed according to methods of the present invention may be cloned into mammalian expression

vectors encoding human Fc regions. Such Fc regions may comprise Fc regions from human IgG1 κ . IgG1 κ Fc regions may comprise amino acid mutations known to enhance Fc-receptor binding and antibody-dependent cell-mediated cytotoxicity (ADCC).

[00226] In some embodiments, antibodies of the invention may be developed for antibody-drug conjugate (ADC) therapeutic applications. ADCs are antibodies in which one or more cargo (e.g. therapeutic agents or cytotoxic agents) are attached [e.g. directly or via linker (e.g. a cleavable linker or a non-cleavable linker)]. ADCs are useful for delivery of such therapeutic agents or cytotoxic agents to one or more target cells or tissues (Panowski, S. et al., 2014. mAbs 6:1, 34-45). In some cases, ADCs may be designed to bind to a surface antigen on a targeted cell. Upon binding, the entire antibody-antigen complex may be internalized and directed to a cellular lysosome. ADCs may then be degraded, releasing the bound cargo. Where the cargo is a cytotoxic agent, the target cell will be killed or otherwise disabled. Cytotoxic agents may include, but are not limited to cytoskeletal inhibitors [e.g. tubulin polymerization inhibitors such as maytansines or auristatins (e.g. monomethyl auristatin E [MMAE] and monomethyl auristatin F [MMAF])] and DNA damaging agents (e.g. DNA polymerization inhibitors such as calcheamicins and duocarmycins).

[00227] In some embodiments, antibodies of the invention may be tested for their ability to promote cell death when developed as ADCs. Cell viability assays may be performed in the presence and absence of secondary antibody-drug conjugates. Antibodies with potent cell growth inhibition may then be used to design direct antibody-drug conjugates (ADCs). The use of such secondary antibody-drug conjugates in cell-based cytotoxic assays may allow for quick pre-screening of many ADC candidates. Based on such assays, an unconjugated antibody candidate is directly added to cells in the presence of a secondary antibody that is conjugated to one or more cytotoxic agents (referred to herein as a 2°ADC). Internalization of the antibody/2°ADC complex into cells that express a high density of the targeted antigen can achieve a dose-dependent drug release within the cells, causing a cytotoxic effect to kill the cells (e.g., tumor cells), while cells expressing a low density of the targeted antigen are not affected (e.g., normal cells).

[00228] ADCs of the invention may be designed to target cancer cells. Such ADCs may comprise antibodies directed to one or more tumor-associated carbohydrate antigen (TACA). In some cases, ADCs of the invention comprise anti-STn antibodies.

Development of chimeric antigen receptors

[00229] In some embodiments, antibody sequences of the invention may be used to develop a chimeric antigen receptor (CAR). CARs are transmembrane receptors expressed on immune cells that facilitate recognition and killing of target cells (e.g. tumor cells). CARs typically comprise three basic parts. These include an ectodomain (also known as the recognition domain), a transmembrane domain and an intracellular (signaling) domain. Ectodomains facilitate binding to cellular antigens on target cells, while intracellular domains typically comprise cell signaling functions to promote the killing of bound target cells. Further, they may have an extracellular domain with one or more antibody variable domains described herein or fragments thereof. CARs of the invention also include a transmembrane domain and cytoplasmic tail. CARs may be designed to include one or more segments of an antibody, antibody variable domain and/or antibody CDR, such that when such CARs are expressed on immune effector cells, the immune effector cells bind and clear any cells that are recognized by the antibody portions of the CARs.

[00230] Characteristics of CARs include their ability to redirect T-cell specificity and reactivity toward a selected target in a non-MHC -restricted manner, exploiting the antigen-binding properties of monoclonal antibodies. The non-MHC-restricted antigen recognition gives T cells expressing CARs the ability to recognize antigen independent of antigen processing, thus bypassing a major mechanism of tumor escape. Moreover, when expressed in T-cells, CARs advantageously do not dimerize with endogenous T cell receptor (TCR) alpha and beta chains.

[00231] CARs engineered to target tumors may have specificity for one or more tumor associated carbohydrate antigens (TACAs). In some embodiments, ectodomains of these CARs may comprise one or more antibody variable domains or a fragment thereof. In some embodiments, CARs are expressed in T cells, and may be referred to as “CAR-engineered T cells” or “CAR-Ts”. CAR-Ts may be engineered with CAR ectodomains having one or more antibody variable domains.

Structural features of chimeric antigen receptors

[00232] With gene-transfer technology, T cells can be engineered to stably express antibodies on their surface, conferring a desired antigen specificity. Chimeric antigen receptors (CARs) combine an antigen-recognition domain of a specific antibody with an intracellular domain of the CD3-zeta chain or FcγRI protein having T cell activating

properties into a single chimeric fusion protein. CAR technology provides MHC-unrestricted recognition of target cells by T cells. Removal of the MHC restriction of T cells facilitates the use of these molecules in any patient, and also, in both CD8⁺ and CD4⁺ T cells, usually restricted to MHC class I or II epitopes, respectively. The use of Ab-binding regions allows T cells to respond to epitopes formed not only by protein, but also carbohydrate and lipid. This chimeric receptor approach is especially suited to immunotherapy of cancer, being able to bypass many of the mechanisms by which tumors avoid immunorecognition, such as MHC down-regulation, lack of expression of costimulatory molecules, CTL resistance, and induction of T cell suppression, and where the use of both CD8⁺ CTL and CD4⁺ T cells are best combined for optimum antitumor efficacy. This approach has been demonstrated to be applicable to a wide range of tumor antigens, in addition to viruses such as HIV (Finney, et al., *J. Immunology*, 2004, 172:104-113).

[00233] Although chimeric antigen receptors can trigger T-cell activation in a manner similar to that of endogenous T-cell receptors, in practice, the clinical application of CAR technology has been impeded by inadequate *in vivo* expansion of chimeric antigen receptor T cells. For example, first generation CARs included as their signaling domain the cytoplasmic region of the CD3ζ or Fc receptor γ chain. These first generation CARs were tested in phase I clinical studies in patients with ovarian cancer, renal cancer, lymphoma, and neuroblastoma, and were found to induce modest responses, effectively redirecting T cell cytotoxicity but failing to enable T cell proliferation and survival upon repeated antigen exposure. The prototypes for second generation CARs involved receptors encompassing both CD28 and CD3ζ, and second generation CARs have been tested for treatment of B cell malignancies and other cancers (Sadelain, et al., (2009) *Current Opinion in Immunology*, 21(2):215-223). Thus, CARs have rapidly expanded into a diverse array of receptors with different functional properties.

[00234] More recently, it was discovered that CAR-mediated T-cell responses can be enhanced with the addition of a costimulatory domain. In preclinical models, the inclusion of the CD137 (4-1BB) signaling domain was found to significantly increase antitumor activity and *in vivo* persistence of chimeric antigen receptors as compared with inclusion of the CD3-zeta chain alone (Porter, et al., *N. Engl. J. Med.* 2011, 365:725-733).

[00235] Thus, in some embodiments of the present disclosure, antibody sequences of the invention may be used to develop a chimeric antigen receptor (CAR). In some embodiments,

CARs are transmembrane receptors expressed on immune cells that facilitate recognition and killing of target cells (*e.g.* tumor cells).

[00236] In many cancers, tumor-specific antigens for targeting have not been defined, but in B-cell neoplasms, CD19 is an attractive target. Expression of CD19 is restricted to normal and malignant B cells and B-cell precursors. A pilot clinical trial of treatment with autologous T cells expressing an anti-CD19 chimeric antigen receptor (CART19) was performed in patients with advanced, p53-deficient chronic lymphoid leukemia (CLL). The generation of a CD19-specific immune response in bone marrow was demonstrated by temporal release of cytokines and ablation of leukemia cells that coincided with peak infiltration of chimeric antigen receptor T cells. (Porter, *et al.*, *N. Engl. J. Med.* 2011, 365:725-733).

[00237] Further structural features of CARs may include any of those disclosed in several PCT Publications assigned to City of Hope and having the common inventor Michael Jensen. For example, PCT Publication WO 00/23573 describes genetically engineered, CD20-specific redirected T cells expressing a cell surface protein having an extracellular domain comprising a receptor specific for CD20, an intracellular signaling domain, and a transmembrane domain. Use of such cells for cellular immunotherapy of CD20⁺ malignancies and for abrogating any untoward B cell function. In one embodiment, the cell surface protein is a single chain FvFc:ζ receptor where Fv designates the VH and VL chains of a single chain monoclonal antibody to CD20 linked by peptide, Fc represents a hinge-CH2-CH3 region of a human IgG1, and ζ represents the intracellular signaling domain of the zeta chain of human CD3. A method of making a redirected T cell expressing a chimeric T cell receptor by electroporation using naked DNA encoding the receptor. Similarly, PCT Publication WO 02/077029 describes genetically engineered, CD19-specific redirected immune cells expressing a cell surface protein having an extracellular domain comprising a receptor which is specific for CD19, an intracellular signaling domain, and a transmembrane domain. Use of such cells for cellular immunotherapy of CD19⁺ malignancies and for abrogating any untoward B cell function. In one embodiment, the immune cell is a T cell and the cell surface protein is a single chain scFvFc:ζ receptor where scFv designates the VH and VL chains of a single chain monoclonal antibody to CD19, Fc represents at least part of a constant region of an IgG1, and zeta represents the intracellular signaling domain of the T cell antigen receptor complex zeta chain (zeta chain of human CD3). The extracellular domain scFvFc and the intracellular domain zeta are linked by a transmembrane domain such as the transmembrane

domain of CD4. A method of making a redirected T cell expressing a chimeric T cell receptor by electroporation using naked DNA encoding the receptor. These chimeric antigen receptors have the ability, when expressed in T cells, to redirect antigen recognition based on the monoclonal antibody's specificity. The design of scFvFc: receptors with target specificities for tumor cell-surface epitopes is a conceptually attractive strategy to generate antitumor immune effector cells for adoptive therapy as it does not rely on pre-existing anti-tumor immunity. These receptors are "universal" in that they bind antigen in a MHC independent fashion, thus, one receptor construct can be used to treat a population of patients with antigen positive tumors. City of Hope PCT Publications WO 02/088334, WO 2007/059298 and WO 2010/065818 describe "zetakines" comprised of an extracellular domain comprising a soluble receptor ligand linked to a support region capable of tethering the extracellular domain to a cell surface, a transmembrane region and an intracellular signaling domain. Zetakines, when expressed on the surface of T lymphocytes, direct T cell activity to those specific cells expressing a receptor for which the soluble receptor ligand is specific.

[00238] Additional features of CARs may include any of those disclosed in two PCT Publications assigned to University of Texas and having a common inventor Lawrence Cooper. PCT Publication No. WO 2009/091826 describes compositions comprising a human CD19-specific chimeric T cell receptor (or chimeric antigen receptor, CAR) polypeptide (designated hCD19CAR) comprising an intracellular signaling domain, a transmembrane domain and an extracellular domain, the extracellular domain comprising a human CD 19 binding region. In another aspect, the CD 19 binding region is an F(ab')₂, Fab', Fab, Fv or scFv. The intracellular domain may comprise an intracellular signaling domain of human CD3 ζ and may further comprise human CD28 intracellular segment. In certain aspects the transmembrane domain is a CD28 transmembrane domain. PCT Publication No. WO 2013/074916 describes methods and compositions for immunotherapy employing CAR⁺ T cells genetically modified to eliminate expression of T cell receptor and/or HLA. In particular embodiments, the T cell receptor-negative and/or HLA-negative T cells are generated using zinc finger nucleases, for example. The CAR⁺ T cells from allogeneic healthy donors can be administered to any patient without causing graft versus host disease (GVHD), acting as universal reagents for off-the-shelf treatment of medical conditions such as cancer, autoimmunity, and infection.

[00239] PCT Publication WO 2011/041093 assigned to the U.S. Department of Health and Human Services describes anti-vascular endothelial growth factor receptor-2 chimeric

antigen receptors comprising an antigen binding domain of a KDR-1121 or DC101 antibody, an extracellular hinge domain, a T cell receptor transmembrane domain, and an intracellular T cell receptor signaling domain, and their use in the treatment of cancer.

[00240] PCT Publications WO 2012/079000 and WO 2013/040557, the contents of each of which are herein incorporated by reference in their entirety, are assigned to University of Pennsylvania and share the common inventor Carl H. June; these publications describe CARs comprising an antigen binding domain, a transmembrane domain, a costimulatory signaling region, and a CD3 zeta signaling domain, and methods for generating RNA Chimeric Antigen Receptor (CAR) transfected T cells, respectively.

[00241] PCT Publication WO2013/126712, also assigned to University of Pennsylvania and sharing the common inventor Carl H. June, describes compositions and methods for generating a persisting population of T cells exhibiting prolonged exponential expansion in culture that is ligand independent and independent of the addition of exogenous cytokines or feeder cells, which are useful for the treatment of cancer. In some embodiments, the antigen binding domain is an anti-cMet binding domain. In some embodiments, the antigen binding domain is an anti-mesothelin binding domain. In some embodiments, the antigen binding domain is an anti-CD 19 binding domain. The hinge domain is IgG4, the transmembrane domain is a CD28 transmembrane domain. In some embodiments, the costimulatory signaling region is a CD28 signaling region. Also provided is a vector comprising a nucleic acid sequence encoding a chimeric antigen receptor (CAR), and the CAR comprising an antigen binding domain, a hinge domain, a transmembrane domain, a costimulatory signaling region, and a CD3 zeta signaling domain.

[00242] PCT Publication WO 2014/039513 assigned to University of Pennsylvania describes compositions and methods for inhibiting one or more diacylglycerol kinase (DGK) isoform in a cell in order to enhance the cytolytic activity of the cell. The cells may be used in adoptive T cell transfer in which, the cell is modified to express a chimeric antigen receptor (CAR). Inhibition of DGK in T cells used in adoptive T cell transfer increases cytolytic activity of the T cells and thus may be used in the treatment of a variety of conditions, including cancer, infection, and immune disorders.

[00243] PCT Publication WO 2014/055771 assigned to University of Pennsylvania describes compositions and methods for treating ovarian cancer. Specifically, the invention relates to administering a genetically modified T cell having alpha-folate receptor (FR-alpha) binding domain and CD27 costimulatory domain to treat ovarian cancer. In one embodiment,

the FR-alpha binding domain is said to be fully human, thereby preventing a host immune response.

[00244] In some embodiments, CARs of the invention may be engineered to target tumors. Such CARs may have specificity for one or more TACAs. In some case, ectodomains of these CARs may comprise one or more antibody variable domain presented herein or a fragment thereof. In some embodiments, CARs of the invention are expressed in T cells, referred to herein as “CAR-engineered T cells” or “CAR-Ts”. CAR-Ts may be engineered with CAR ectodomains having one or more antibody variable domain presented herein.

Multispecific antibodies

[00245] In some embodiments, antibodies of the present invention may bind more than one epitope. As used herein, the terms “multibody” or “multispecific antibody” refer to an antibody wherein two or more variable regions bind to different epitopes. The epitopes may be on the same or different targets. In certain embodiments, a multi-specific antibody is a "bispecific antibody," which recognizes two different epitopes on the same or different antigens.

Bispecific antibodies

[00246] Bispecific antibodies are capable of binding two different antigens. Such antibodies typically comprise antigen-binding regions from at least two different antibodies. For example, a bispecific monoclonal antibody (BsMAb, BsAb) is an artificial protein composed of fragments of two different monoclonal antibodies, thus allowing the BsAb to bind to two different types of antigen. One common application for this technology is in cancer immunotherapy, where BsMAbs are engineered to simultaneously bind to a cytotoxic cell (using a receptor like CD3) and a target like a tumor cell to be destroyed.

[00247] Bispecific antibodies may include any of those described in Riethmuller, G., 2012. *Cancer Immunity*. 12:12-18; Marvin, J.S. *et al.*, 2005. *Acta Pharmacologica Sinica*. 26(6):649-58; and Schaefer, W. *et al.*, 2011. *PNAS*. 108(27):11187-92, the contents of each of which are herein incorporated by reference in their entirety.

[00248] New generations of BsMAb, called “trifunctional bispecific” antibodies, have been developed. These consist of two heavy and two light chains, one each from two different antibodies, where the two Fab regions (the arms) are directed against two antigens, and the Fc region (the foot) comprises the two heavy chains and forms the third binding site.

[00249] Of the two paratopes that form the tops of the variable domains of a bispecific antibody, one can be directed against a target antigen and the other against a T-lymphocyte antigen like CD3. In the case of trifunctional antibodies, the Fc region may additionally binds to a cell that expresses Fc receptors, like a macrophage, a natural killer (NK) cell or a dendritic cell. In sum, the targeted cell is connected to one or two cells of the immune system, which subsequently destroy it.

[00250] Other types of bispecific antibodies have been designed to overcome certain problems, such as short half-life, immunogenicity and side-effects caused by cytokine liberation. They include chemically linked Fabs, consisting only of the Fab regions, and various types of bivalent and trivalent single-chain variable fragments (scFvs), fusion proteins mimicking the variable domains of two antibodies. The furthest developed of these newer formats are the bi-specific T-cell engagers (BiTEs) and mAb2's, antibodies engineered to contain an Fcab antigen-binding fragment instead of the Fc constant region.

[00251] A bispecific, single-chain antibody Fv fragment (Bs-scFv) was successfully used to kill cancer cells. Some human cancers are caused by functional defects in p53 that are restored by gene therapy with wild-type p53. Weisbart, *et al.*, describe the construction and expression of a bispecific single-chain antibody that penetrates living colon cancer cells, binds intracellular p53, and targets and restores its wild type function (Weisbart, *et al.*, *Int. J. Oncol.* 2004 Oct;25(4):1113-8; and Weisbart, *et al.*, *Int. J. Oncol.* 2004 Dec;25(6):1867-73). In these studies, a bispecific, single-chain antibody Fv fragment (Bs-scFv) was constructed from (i) a single-chain Fv fragment of mAb 3E10 that penetrates living cells and localizes in the nucleus, and (ii) a single-chain Fv fragment of a non-penetrating antibody, mAb PAb421 that binds the C-terminal of p53. PAb421 binding restores wild-type functions of some p53 mutants, including those of SW480 human colon cancer cells. The Bs-scFv penetrated SW480 cells and was cytotoxic, suggesting an ability to restore activity to mutant p53. COS-7 cells (monkey kidney cells with wild-type p53) served as a control since they are unresponsive to PAb421 due to the presence of SV40 large T antigen that inhibits binding of PAb421 to p53. Bs-scFv penetrated COS-7 cells but was not cytotoxic, thereby eliminating non-specific toxicity of Bs-scFv unrelated to binding p53. Fv fragments alone were not cytotoxic, indicating that killing was due to transduction of p53. A single mutation in CDR1 of PAb421 VH eliminated binding of the Bs-scFv to p53 and abrogated cytotoxicity for SW480 cells without altering cellular penetration, further supporting the requirement of

PAb421 binding to p53 for cytotoxicity (Weisbart, *et al.*, *Int. J. Oncol.* 2004 Oct;25(4):1113-8; and Weisbart, *et al.*, *Int. J. Oncol.* 2004 Dec;25(6):1867-73).

[00252] In some embodiments, antibodies of the present invention may be diabodies. Diabodies are functional bispecific single-chain antibodies (bscAb). These bivalent antigen-binding molecules are composed of non-covalent dimers of scFvs, and can be produced in mammalian cells using recombinant methods. (*See, e.g., Mack et al, Proc. Natl. Acad. Sci.*, 92: 7021-7025, 1995). Few diabodies have entered clinical development. An iodine-123-labeled diabody version of the anti-CEA chimeric antibody cT84.66 has been evaluated for pre-surgical immunoscintigraphic detection of colorectal cancer in a study sponsored by the Beckman Research Institute of the City of Hope (Clinicaltrials.gov NCT00647153) (Nelson, A. L., *MAbs*.2010. Jan-Feb; 2(1):77–83).

[00253] Using molecular genetics, two scFvs can be engineered in tandem into a single polypeptide, separated by a linker domain, called a “tandem scFv” (tascFv). TascFvs have been found to be poorly soluble and require refolding when produced in bacteria, or they may be manufactured in mammalian cell culture systems, which avoids refolding requirements but may result in poor yields. Construction of a tascFv with genes for two different scFvs yields a “bispecific single-chain variable fragments” (bis-scFvs). Only two tascFvs have been developed clinically by commercial firms; both are bispecific agents in active early phase development by Micromet for oncologic indications, and are described as “Bispecific T-cell Engagers (BiTE).” Blinatumomab is an anti-CD19/anti-CD3 bispecific tascFv that potentiates T-cell responses to B-cell non-Hodgkin lymphoma in Phase 2. MT110 is an anti-EP-CAM/anti-CD3 bispecific tascFv that potentiates T-cell responses to solid tumors in Phase 1. Bispecific, tetravalent “TandAbs” are also being researched by Affimed (Nelson, A. L., *MAbs*.2010. Jan-Feb; 2(1):77–83).

[00254] Also included are maxibodies (bivalent scFv fused to the amino terminus of the Fc (CH2-CH3 domains) of IgG).

[00255] Bispecific T-cell-engager (BiTE) antibodies are designed to transiently engage cytotoxic T-cells for lysis of selected target cells. The clinical activity of BiTE antibodies corroborates findings that *ex vivo* expanded, autologous T-cells derived from tumor tissue, or transfected with specific T-cell receptors, have shown therapeutic potential in the treatment of solid tumors. While these personalized approaches prove that T-cells alone can have considerable therapeutic activity, even in late-stage cancer, they are cumbersome to perform on a broad basis. This is different for cytotoxic T-lymphocyte antigen 4 (CTLA-4) antibodies,

which facilitate generation of tumor-specific T-cell clones, and also for bi- and tri-specific antibodies that directly engage a large proportion of patients' T-cells for cancer cell lysis. The potential of global T-cell engagement for human cancer therapy by T-cell-engaging antibodies is under active investigation (Baeuerle PA, *et al.*, *Current Opinion in Molecular Therapeutics*. 2009, 11(1):22-30).

[00256] Third generation molecules include “miniaturized” antibodies. Among the best examples of mAb miniaturization are the small modular immunopharmaceuticals (SMIPs) from Trubion Pharmaceuticals. These molecules, which can be monovalent or bivalent, are recombinant single-chain molecules containing one V_L, one V_H antigen-binding domain, and one or two constant “effector” domains, all connected by linker domains. Presumably, such a molecule might offer the advantages of increased tissue or tumor penetration claimed by fragments while retaining the immune effector functions conferred by constant domains. At least three “miniaturized” SMIPs have entered clinical development. TRU-015, an anti-CD20 SMIP developed in collaboration with Wyeth, is the most advanced project, having progressed to Phase 2 for rheumatoid arthritis (RA). Earlier attempts in systemic lupus erythematosus (SLE) and B cell lymphomas were ultimately discontinued. Trubion and Facet Biotechnology are collaborating in the development of TRU-016, an anti-CD37 SMIP, for the treatment of CLL and other lymphoid neoplasias, a project that has reached Phase 2. Wyeth has licensed the anti-CD20 SMIP SBI-087 for the treatment of autoimmune diseases, including RA, SLE and possibly multiple sclerosis, although these projects remain in the earliest stages of clinical testing. (Nelson, A. L., *MAbs*.2010. Jan-Feb; 2(1):77–83).

[00257] Genmab is researching application of their “Unibody” technology, in which the hinge region has been removed from IgG4 molecules. While IgG4 molecules are unstable and can exchange light-heavy chain heterodimers with one another, deletion of the hinge region prevents heavy chain-heavy chain pairing entirely, leaving highly specific monovalent light/heavy heterodimers, while retaining the Fc region to ensure stability and half-life in vivo. This configuration may minimize the risk of immune activation or oncogenic growth, as IgG4 interacts poorly with FcRs and monovalent unibodies fail to promote intracellular signaling complex formation. These contentions are, however, largely supported by laboratory, rather than clinical, evidence. Biotechnol is also developing a “miniaturized” mAb, CAB051, which is a “compacted” 100 kDa anti-HER2 antibody in preclinical research (Nelson, A. L., *MAbs*.2010. Jan-Feb; 2(1):77–83).

[00258] Recombinant therapeutics composed of single antigen-binding domains have also been developed, although they currently account for only 4% of the clinical pipeline. These molecules are extremely small, with molecular weights approximately one-tenth of those observed for full-sized mAbs. Arana and Domantis engineer molecules composed of antigen-binding domains of human immunoglobulin light or heavy chains, although only Arana has a candidate in clinical testing, ART-621, an anti-TNF α molecule in Phase 2 study for the treatment of psoriasis and rheumatoid arthritis. Ablynx produces “nanobodies” derived from the antigen-binding variable heavy chain regions (V_{HHS}) of heavy chain antibodies found in camels and llamas, which lack light chains. Two Ablynx anti-von Willebrand Factor nanobodies have advanced to clinical development, including ALX-0081, in Phase 2 development as an intravenous therapy to prevent thrombosis in patients undergoing percutaneous coronary intervention for acute coronary syndrome, and ALX-0681, a Phase 1 molecule for subcutaneous administration intended for both patients with acute coronary syndrome and thrombotic thrombocytopenic purpura (Nelson, A. L., *MAbs*.2010. Jan-Feb; 2(1):77–83).

Development of multispecific antibodies

[00259] In some embodiments, antibody sequences of the invention may be used to develop multispecific antibodies (*e.g.*, bispecific, trispecific, or of greater multispecificity).

Multispecific antibodies can be specific for different epitopes of a target antigen of the present invention, or can be specific for both a target antigen of the present invention, and a heterologous epitope, such as a heterologous glycan, peptide or solid support material. (*See, e.g.*, WO 93/17715; WO 92/08802; WO 91/00360; WO 92/05793; Tutt, A. *et al.*, *Trispecific F(ab')₃ derivatives that use cooperative signaling via the TCR/CD3 complex and CD2 to activate and redirect resting cytotoxic T cells*. *J. Immunol.* 1991 Jul 1;147(1):60-9; U.S. Pat. Nos. 4,474,893; 4,714,681; 4,925,648; 5,573,920; 5,601,819; and Kostelny, S.A. *et al.*, *Formation of a bispecific antibody by the use of leucine zippers*. *J. Immunol.* 1992 Mar 1;148(5):1547-53); U.S. Pat. No. 5,932,448.

[00260] Disclosed and claimed in PCT Publication WO2014144573 to Memorial Sloan-Kettering Cancer Center are multimerization technologies for making dimeric multispecific binding agents (*e.g.*, fusion proteins comprising antibody components) with improved properties over multispecific binding agents without the capability of dimerization.

[00261] Disclosed and claimed in PCT Publication WO2014144357 to Merck Patent GMBH are tetravalent bispecific antibodies (TetBiAbs), and methods of making and methods of using TetBiAbs for diagnostics and for the treatment of cancer or immune disorders. TetBiAbs feature a second pair of Fab fragments with a second antigen specificity attached to the C-terminus of an antibody, thus providing a molecule that is bivalent for each of the two antigen specificities. The tetravalent antibody is produced by genetic engineering methods, by linking an antibody heavy chain covalently to a Fab light chain, which associates with its cognate, co-expressed Fab heavy chain.

[00262] Disclosed and claimed in PCT Publication WO2014028560 to IBC Pharmaceuticals, Inc. are T cell redirecting bispecific antibodies (bsAb), with at least one binding site for a T-cell antigen and at least one binding site for an antigen on a diseased cell or pathogen, for treatment of disease. Preferably, this bsAb is an anti-CD3 x anti-CD19 bispecific antibody, although antibodies against other T-cell antigens and/or disease-associated antigens may be used. The complex is capable of targeting effector T cells to induce T-cell-mediated cytotoxicity of cells associated with a disease, such as cancer, autoimmune disease or infectious disease. The cytotoxic immune response is enhanced by co-administration of interferon-based agents that comprise interferon- α , interferon- β ; interferon- λ 1, interferon- λ 2 or interferon- λ 3.

[00263] Disclosed and claimed in PCT Publication WO2013092001 to Synimmune GMBH is a bispecific antibody molecule, as well as a method for producing the same, its use and a nucleic acid molecule encoding the bispecific antibody molecule. In particular is provided an antibody molecule that is capable of mediating target cell restricted activation of immune cells.

[00264] Disclosed and claimed in PCT Publication WO2012007167 is a multispecific modular antibody specifically binding to at least a glycoepitope and a receptor of the erbB class on the surface of a tumor cell, thereby crosslinking the glycoepitope and the receptor, which antibody has apoptotic activity effecting cytolysis independent of NK cells.

[00265] Disclosed and claimed in PCT Publications WO2012048332 and WO2013055404 are mediotopes, mediotope-binding antibodies, mediotope delivery systems, as well as a monoclonal antibody framework binding interface for mediotopes, and methods for their use. Specifically, two antibody binding peptides, C-QFDLSTRRLK-C (“cQFD”; sequence identification number 1 therein; SEQ ID NO: 240 herein) and C-QYNLSSRALK-C (“cQYN”; sequence identification number 2 therein; SEQ ID NO: 241 herein) were shown to

have novel mAb binding properties. Also called "meditopes," cQFD and cQYN were shown to bind to a region of the Fab framework of the anti-EGFR mAb cetuximab and not to bind the complementarity determining regions (CDRs) that bind antigen. The binding region on the Fab framework is distinct from other framework-binding antigens, such as the superantigens *Staphylococcal* protein A (SpA) (Graille *et al.*, 2000) and *Peptostreptococcus magnus* protein L (PpL) (Graille *et al.*, 2001). Accordingly, one embodiment disclosed is a framework binding interface comprising a framework region of a unique murine-human antibody or functional fragment thereof that binds a cyclic meditope.

[00266] Exemplary patents and patent publications of interest are: U.S. Patent Nos. 5,585,089; 5,693,761; and 5,693,762, all filed Jun 7, 1995 and U.S. Patent No. 6,180,370, all assigned to Protein Design Labs, Inc., describe methods for producing, and compositions of, humanized immunoglobulins having one or more complementarity determining regions (CDR's) and possible additional amino acids from a donor immunoglobulin and a framework region from an accepting human immunoglobulin. Each humanized immunoglobulin chain is said to usually comprise, in addition to the CDR's, amino acids from the donor immunoglobulin framework that are, *e.g.*, capable of interacting with the CDRs to effect binding affinity, such as one or more amino acids which are immediately adjacent to a CDR in the donor immunoglobulin or those within about 3 Å as predicted by molecular modeling. The heavy and light chains may each be designed by using any one or all of various position criteria. When combined into an intact antibody, the humanized immunoglobulins of the present invention is said to be substantially non-immunogenic in humans and retain substantially the same affinity as the donor immunoglobulin to the antigen, such as a protein or other compound containing an epitope.

[00267] U.S. Patent No. 5,951,983, assigned to Universite Catholique De Louvain and Bio Transplant, Inc., describes a humanized antibody against T-lymphocytes. Framework regions from a human V kappa gene designated as HUM5400 (EMBL accession X55400) and from the human antibody clone Amu 5-3 (GenBank accession number U00562) are set forth therein.

[00268] U.S. Patent No. 5,091,513, to Creative Biomolecules, Inc., describes a family of synthetic proteins having affinity for a preselected antigen. The proteins are characterized by one or more sequences of amino acids constituting a region which behaves as a biosynthetic antibody binding site (BABS). The sites comprise 1) non-covalently associated or disulfide bonded synthetic V_H and V_L dimers, 2) V_H-V_L or V_L-V_H single chains wherein the V_H and V_L

are attached by a polypeptide linker, or 3) individuals V_H or V_L domains. The binding domains comprise linked CDR and FR regions, which may be derived from separate immunoglobulins. The proteins may also include other polypeptide sequences which function, *e.g.*, as an enzyme, toxin, binding site, or site of attachment to an immobilization media or radioactive atom. Methods are disclosed for producing the proteins, for designing BABS having any specificity that can be elicited by *in vivo* generation of antibody, and for producing analogs thereof.

[00269] U.S. Patent No. 8,399,625, to ESBATech, an Alcon Biomedical Research Unit, LLC, describes antibody acceptor frameworks and methods for grafting non-human antibodies, *e.g.*, rabbit antibodies, using a particularly well suited antibody acceptor framework.

Intrabodies

[00270] In some embodiments, antibodies of the present invention may be intrabodies. Intrabodies are a form of antibody that is not secreted from a cell in which it is produced, but instead targets one or more intracellular proteins. Intrabodies are expressed and function intracellularly, and may be used to affect a multitude of cellular processes including, but not limited to intracellular trafficking, transcription, translation, metabolic processes, proliferative signaling and cell division. In some embodiments, methods described herein include intrabody-based therapies. In some such embodiments, variable domain sequences and/or CDR sequences disclosed herein are incorporated into one or more constructs for intrabody-based therapy. For example, intrabodies may target one or more glycosylated intracellular proteins or may modulate the interaction between one or more glycosylated intracellular proteins and an alternative protein.

[00271] More than two decades ago, intracellular antibodies against intracellular targets were first described (Biocca, Neuberger and Cattaneo *EMBO J.* 9: 101-108, 1990). The intracellular expression of intrabodies in different compartments of mammalian cells allows blocking or modulation of the function of endogenous molecules (Biocca, *et al.*, *EMBO J.* 9: 101-108, 1990; Colby *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 101 : 17616-21 , 2004). Intrabodies can alter protein folding, protein-protein, protein-DNA, protein-RNA interactions and protein modification. They can induce a phenotypic knockout and work as neutralizing agents by direct binding to the target antigen, by diverting its intracellular traffic or by inhibiting its association with binding partners. They have been largely employed as research tools and are

emerging as therapeutic molecules for the treatment of human diseases as viral pathologies, cancer and misfolding diseases. The fast growing bio-market of recombinant antibodies provides intrabodies with enhanced binding specificity, stability and solubility, together with lower immunogenicity, for their use in therapy (Biocca, abstract in *Antibody Expression and Production Cell Engineering* Volume 7, 2011, pp. 179-195).

[00272] In some embodiments, intrabodies have advantages over interfering RNA (iRNA); for example, iRNA has been shown to exert multiple non-specific effects, whereas intrabodies have been shown to have high specificity and affinity of to target antigens. Furthermore, as proteins, intrabodies possess a much longer active half-life than iRNA. Thus, when the active half-life of the intracellular target molecule is long, gene silencing through iRNA may be slow to yield an effect, whereas the effects of intrabody expression can be almost instantaneous. Lastly, it is possible to design intrabodies to block certain binding interactions of a particular target molecule, while sparing others.

Development of intrabodies

[00273] Intrabodies are often single chain variable fragments (scFvs) expressed from a recombinant nucleic acid molecule and engineered to be retained intracellularly (*e.g.*, retained in the cytoplasm, endoplasmic reticulum, or periplasm). Intrabodies may be used, for example, to ablate the function of a protein to which the intrabody binds. The expression of intrabodies may also be regulated through the use of inducible promoters in the nucleic acid expression vector comprising the intrabody. Intrabodies may be produced using methods known in the art, such as those disclosed and reviewed in: (Marasco *et al.*, 1993 *Proc. Natl. Acad. Sci. USA*, 90: 7889-7893; Chen *et al.*, 1994, *Hum. Gene Ther.* 5:595-601; Chen *et al.*, 1994, *Proc. Natl. Acad. Sci. USA*, 91: 5932-5936; Maciejewski *et al.*, 1995, *Nature Med.*, 1: 667-673; Marasco, 1995, *Immunotech*, 1: 1-19; Mhashilkar, *et al.*, 1995, *EMBO J.* 14: 1542-51; Chen *et al.*, 1996, *Hum. Gene Therap.*, 7: 1515-1525; Marasco, *Gene Ther.* 4:11-15, 1997; Rondon and Marasco, 1997, *Annu. Rev. Microbiol.* 51:257-283; Cohen, *et al.*, 1998, *Oncogene* 17:2445-56; Proba *et al.*, 1998, *J. Mol. Biol.* 275:245-253; Cohen *et al.*, 1998, *Oncogene* 17:2445-2456; Hassanzadeh, *et al.*, 1998, *FEBS Lett.* 437:81-6; Richardson *et al.*, 1998, *Gene Ther.* 5:635-44; Ohage and Steipe, 1999, *J. Mol. Biol.* 291:1119-1128; Ohage *et al.*, 1999, *J. Mol. Biol.* 291:1129-1134; Wirtz and Steipe, 1999, *Protein Sci.* 8:2245-2250; Zhu *et al.*, 1999, *J. Immunol. Methods* 231:207-222; Arafat *et al.*, 2000, *Cancer Gene Ther.* 7:1250-6; der Maur *et al.*, 2002, *J. Biol. Chem.* 277:45075-85; Mhashilkar *et al.*, 2002, *Gene*

Ther. 9:307-19; and Wheeler *et al.*, 2003, *FASEB J.* 17: 1733-5; and references cited therein). In particular, a CCR5 intrabody has been produced by Steinberger *et al.*, 2000, *Proc. Natl. Acad. Sci. USA* 97:805-810). See generally Marasco, WA, 1998, "Intrabodies: Basic Research and Clinical Gene Therapy Applications" Springer:New York; and for a review of scFvs, see Pluckthun in "The Pharmacology of Monoclonal Antibodies," 1994, vol. 113, Rosenberg and Moore *eds.* Springer-Verlag, New York, pp. 269-315.

[00274] In some embodiments, antibody sequences are used to develop intrabodies.

Intrabodies are often recombinantly expressed as single domain fragments such as isolated VH and VL domains or as a single chain variable fragment (scFv) antibody within the cell. For example, intrabodies are often expressed as a single polypeptide to form a single chain antibody comprising the variable domains of the heavy and light chain joined by a flexible linker polypeptide. Intrabodies typically lack disulfide bonds and are capable of modulating the expression or activity of target genes through their specific binding activity. Single chain antibodies can also be expressed as a single chain variable region fragment joined to the light chain constant region.

[00275] As is known in the art, an intrabody can be engineered into recombinant polynucleotide vectors to encode sub-cellular trafficking signals at its N or C terminus to allow expression at high concentrations in the sub-cellular compartments where a target protein is located. For example, intrabodies targeted to the endoplasmic reticulum (ER) are engineered to incorporate a leader peptide and, optionally, a C-terminal ER retention signal, such as the KDEL (SEQ ID NO: 242) amino acid motif. Intrabodies intended to exert activity in the nucleus are engineered to include a nuclear localization signal. Lipid moieties are joined to intrabodies in order to tether the intrabody to the cytosolic side of the plasma membrane. Intrabodies can also be targeted to exert function in the cytosol. For example, cytosolic intrabodies are used to sequester factors within the cytosol, thereby preventing them from being transported to their natural cellular destination.

[00276] There are certain technical challenges with intrabody expression. In particular, protein conformational folding and structural stability of the newly-synthesized intrabody within the cell is affected by reducing conditions of the intracellular environment. In human clinical therapy, there are safety concerns surrounding the application of transfected recombinant DNA, which is used to achieve intrabody expression within the cell. Of particular concern are the various viral-based vectors commonly-used in genetic manipulation. Thus, one approach to circumvent these problems is to fuse protein

transduction domains (PTD) to scFv antibodies, to create a 'cell-permeable' antibody or 'Transbody.' Transbodies are cell-permeable antibodies in which a protein transduction domain (PTD) is fused with single chain variable fragment (scFv) antibodies (Heng and Cao, 2005, *Med Hypotheses*. 64:1105-8).

[00277] Upon interaction with a target gene, an intrabody modulates target protein function and/or achieves phenotypic/functional knockout by mechanisms such as accelerating target protein degradation and sequestering the target protein in a non-physiological sub-cellular compartment. Other mechanisms of intrabody-mediated gene inactivation can depend on the epitope to which the intrabody is directed, such as binding to the catalytic site on a target protein or to epitopes that are involved in protein-protein, protein-DNA, or protein-RNA interactions.

[00278] In one embodiment, intrabodies are used to capture a target in the nucleus, thereby preventing its activity within the nucleus. Nuclear targeting signals are engineered into such intrabodies in order to achieve the desired targeting. Such intrabodies are designed to bind specifically to a particular target domain. In another embodiment, cytosolic intrabodies that specifically bind to a target protein are used to prevent the target from gaining access to the nucleus, thereby preventing it from exerting any biological activity within the nucleus (e.g., preventing the target from forming transcription complexes with other factors).

[00279] In order to specifically direct the expression of such intrabodies to particular cells, the transcription of the intrabody is placed under the regulatory control of an appropriate tumor-specific promoter and/or enhancer. In order to target intrabody expression specifically to prostate, for example, the PSA promoter and/or promoter/enhancer can be utilized (See, for example, U.S. Patent No. 5,919,652 issued 6 July 1999).

[00280] Protein transduction domains (PTDs) are short peptide sequences that enable proteins to translocate across the cell membrane and be internalized within the cytosol, through atypical secretory and internalization pathways. There are a number of distinct advantages that a 'Transbody' would possess over conventional intrabodies expressed within the cell. For a start, 'correct' conformational folding and disulfide bond formation can take place prior to introduction into the target cell. More importantly, the use of cell-permeable antibodies or 'Transbodies' would avoid the overwhelming safety and ethical concerns surrounding the direct application of recombinant DNA technology in human clinical therapy, which is required for intrabody expression within the cell. 'Transbodies' introduced into the cell would possess only a limited active half-life, without resulting in any permanent

genetic alteration. This would allay any safety concerns with regards to their application in human clinical therapy (Heng and Cao 2005, *Med Hypotheses*. 64:1105-8).

[00281] Intrabodies are promising therapeutic agents for the treatment of misfolding diseases, including Alzheimer's, Parkinson's, Huntington's and prion diseases, because of their virtually infinite ability to specifically recognize the different conformations of a protein, including pathological isoforms, and because they can be targeted to the potential sites of aggregation (both intra- and extracellular sites). These molecules can work as neutralizing agents against amyloidogenic proteins by preventing their aggregation, and/or as molecular shunters of intracellular traffic by rerouting the protein from its potential aggregation site (Cardinale, and Biocca, *Curr. Mol. Med.* 2008, 8:2-11).

[00282] Exemplary Patent Publications describing intracellular antibodies or intrabodies are set forth hereinbelow, each of which is incorporated by reference in its entirety.

[00283] PCT Publication WO03014960 and US Patent 7,608,453 granted to Cattaneo, *et al.*, describe an intracellular antibody capture technology method of identifying at least one consensus sequence for an intracellular antibody (ICS) comprising the steps of: creating a database comprising sequences of validated intracellular antibodies (VIDA database) and aligning the sequences of validated intracellular antibodies according to Kabat; determining the frequency with which a particular amino acid occurs in each of the positions of the aligned antibodies; selecting a frequency threshold value (LP or consensus threshold) in the range from 70% to 100%; identifying the positions of the alignment at which the frequency of a particular amino acid is greater than or equal to the LP value; and identifying the most frequent amino acid, in the position of said alignment.

[00284] PCT Publications WO0054057; WO03077945; WO2004046185; WO2004046186; WO2004046187; WO2004046188; WO2004046189; US Patent Application Publications US2005272107; US2005276800; US2005288492; US2010143939; granted US Patents 7,569,390 and 7,897,347 and granted European Patents EP1560853; and EP1166121 all assigned to the Medical Research Council and including inventors Cattaneo, *et al.*, describe intracellular intracellular single domain immunoglobulins, and a method for determining the ability of a immunoglobulin single domain to bind to a target in an intracellular environment, as well as methods for generating intracellular antibodies.

[00285] PCT Publication WO0235237; US Patent Application Publication 2003235850 and granted European Patent EP1328814 naming Cattaneo as an inventor and assigned to

S.I.S.S.A. Scuola Internazionale Superiore describe a method for the *in vivo* identification of epitopes of an intracellular antigen.

[00286] PCT Publication WO2004046192 and European Patent EP1565558 assigned to Lay Line Genomics SPA and naming Catteneo as an inventor describe a method for isolating intracellular antibodies that disrupt and neutralize an interaction between a protein ligand x and a protein ligand y inside a cell. Also disclosed are a method to identify a protein ligand x able to bind to a known y ligand using intracellular antibodies able to the interaction between x and y; and a method for the isolation of a set of antibody fragments against a significant proportion of the protein-protein interactions of a given cell (interactome) or against the protein interactions that constitute an intracellular pathway or network.

[00287] US Patent Application Publication 2006034834 and PCT Publication WO9914353 entitled “Intrabody-mediated control of immune reactions” and assigned to Dana Farber Cancer Institute Inc. name inventors Marasco and Mhashilkar are directed to methods of altering the regulation of the immune system, *e.g.*, by selectively targeting individual or classes of immunomodulatory receptor molecules (IRMs) on cells comprising transducing the cells with an intracellularly expressed antibody, or intrabody, against the IRMs. In a preferred embodiment the intrabody comprises a single chain antibody against an IRM, *e.g.*, MHC-1 molecules.

[00288] PCT Publication WO2013033420 assigned to Dana Farber Cancer Institute Inc. and Whitehead Biomedical Institute, and naming inventors Bradner, Rahl and Young describes methods and compositions useful for inhibiting interaction between a bromodomain protein and an immunoglobulin (Ig) regulatory element and downregulating expression of an oncogene translocated with an Ig locus, as well as for treating a cancer (*e.g.*, hematological malignancy) characterized by increased expression of an oncogene which is translocated with an Ig locus. Intrabodies are generally described.

[00289] PCT Publication WO02086096 and US Patent Application Publication 2003104402 entitled “Methods of producing or identifying intrabodies in eukaryotic cells,” assigned to University of Rochester Medical Center and naming inventors Zauderer, Wei and Smith describe a high efficiency method of expressing intracellular immunoglobulin molecules and intracellular immunoglobulin libraries in eukaryotic cells using a trimolecular recombination method. Further provided are methods of selecting and screening for intracellular immunoglobulin molecules and fragments thereof, and kits for producing,

screening and selecting intracellular immunoglobulin molecules, as well as the intracellular immunoglobulin molecules and fragments produced using these methods.

[00290] PCT Publication WO2013023251 assigned to Affinity Biosciences PTY LTD and naming inventors Beasley, Niven and Kiefel describes polypeptides, such as antibody molecules and polynucleotides encoding such polypeptides, and libraries thereof, wherein the expressed polypeptides that demonstrate high stability and solubility. In particular, polypeptides comprising paired VL and VH domains that demonstrate soluble expression and folding in a reducing or intracellular environment are described, wherein a human scFv library was screened, resulting in the isolation of soluble scFv genes that have identical framework regions to the human germline sequence as well as remarkable thermostability and tolerance of CDR3 grafting onto the scFv scaffold.

[00291] European Patent Application EP2314622 and PCT Publications WO03008451 and WO03097697 assigned to Esbatech AG and University of Zuerich and naming inventors Ewert, Huber, Honneger and Pluckthun describe the modification of human variable domains and provide compositions useful as frameworks for the creation of very stable and soluble single-chain Fv antibody fragments. These frameworks have been selected for intracellular performance and are thus ideally suited for the creation of scFv antibody fragments or scFv antibody libraries for applications where stability and solubility are limiting factors for the performance of antibody fragments, such as in the reducing environment of a cell. Such frameworks can also be used to identify highly conserved residues and consensus sequences which demonstrate enhanced solubility and stability.

[00292] PCT Publication WO02067849 and US Patent Application Publication 2004047891 entitled "Systems devices and methods for intrabody targeted delivery and reloading of therapeutic agents" describe systems, devices and methods for intrabody targeted delivery of molecules. More particularly, some embodiments relate to a reloadable drug delivery system, which enables targeted delivery of therapeutic agents to a tissue region of a subject, in a localized and timely manner.

[00293] PCT Publication WO2005063817 and US Patent 7,884,054 assigned to Amgen Inc. and naming inventors Zhou, Shen and Martin describe methods for identifying functional antibodies, including intrabodies. In particular, a homodimeric intrabody is described, wherein each polypeptide chain of the homodimer comprises an Fc region, an scFv, and an intracellular localization sequence. The intracellular localization sequence may cause the

intrabody to be localized to the ER or the Golgi. Optionally, each polypeptide chain comprises not more than one scFv.

[00294] PCT Publication WO2013138795 by Vogan, *et al.* and assigned to Permeon Biologics Inc. describes cell penetrating compositions for delivery of intracellular antibodies and antibody-like moieties and methods for delivering them (referred to herein as "AAM moieties" or "an AAM moiety") into a cell. Without being bound by theory, the present disclosure is based, at least in part, on the discovery that an AAM moiety can be delivered into a cell by complexing the AAM moiety with a cell penetrating polypeptide having surface positive charge (referred to herein as a "Surf+ Penetrating Polypeptide"). Examples of some applications of intraphilin technology are also provided

[00295] PCT Publication WO2010004432 assigned to the Pasteur Institute describes immunoglobulins from camelidae (camels, dromedaries, llamas and alpacas), about 50% of which are antibodies devoid of light chain. These heavy-chain antibodies interact with the antigen by the virtue of only one single variable domain, referred to as VHH(s), VHH domain(s) or VHH antibody(ies). Despite the absence of light chain, these homodimeric antibodies exhibit a broad antigen-binding repertoire by enlarging their hypervariable regions, and can act as a transbody and/or intrabody *in vitro* as well as *in vivo*, when the VHH domain is directed against an intracellular target.

[00296] PCT Publication WO2014106639 describes a method for identifying a cellular target involved in a cell phenotype by identifying an intrabody that can modify a cell phenotype and identifying a direct or indirect cellular target of the intrabody. In particular, intrabodies 3H2-1, 3H2-VH and 5H4 are capable of inhibiting the degranulation reaction in mast cells triggered by an allergic stimulus; furthermore, intrabodies 3H2-1 and 5H4 directly or indirectly targeted a protein of the ABCF1 family and C120RF4 family, respectively. These ABCF1 and C120RF4 inhibitors are said to be useful in therapy, in particular for treating allergic and/or inflammatory conditions.

[00297] PCT Publication WO0140276 assigned to Urogenesis Inc. generally describes the possibility of inhibition of STEAP (Six Transmembrane Epithelial Antigen of the Prostate) proteins using intracellular antibodies (intrabodies).

[00298] PCT Publication WO02086505 assigned to University of Manchester and US Patent Application Publication US2004115740 naming inventors Simon and Benton describe a method for the intracellular analysis of a target molecule, wherein intrabodies are said to be

preferred. In one embodiment, a vector (designated pScFv-ECFP) capable of expressing an anti-MUC1 intrabody coupled to CFP is described.

[00299] PCT Publication WO03095641 and WO0143778 assigned to Gene Therapy Systems Inc. describe compositions and methods for intracellular protein delivery, and intrabodies are generally described.

[00300] PCT Publication WO03086276 assigned to Selective Genetics Inc. describes a platform technology for the treatment of intracellular infections. Compositions and methods described therein include non-target specific vectors that target infectable cells via linked ligands that bind and internalize through cell surface receptors/moieties associated with infection. The vectors comprise exogenous nucleic acid sequences that are expressed upon internalization into a target cell. Vector associated ligands and nucleic acid molecules may be altered to target different infectious agents. In addition, the invention provides methods of identifying epitopes and ligands capable of directing internalization of a vector and capable of blocking viral entry.

[00301] PCT Publication WO03062415 assigned to Erasmus University describes a transgenic organism comprising a polynucleotide construct encoding an intracellular antibody which disrupts the catalysis of the production of the xenoantigen galactose alpha 1,3 galactose and/or a polynucleotide construct which encodes an intracellular antibody which binds specifically to a retrovirus protein, such as a PERV particle protein. Cells, tissues and organs of the transgenic organism may be used in xenotransplantation.

[00302] PCT Publication WO2004099775 entitled "Means for detecting protein conformation and applications thereof" describes the use of scFv fragments as conformation-specific antibodies for specifically detecting a conformational protein state, said to have applications as sensors for following in living cells, upon intracellular expression, the behavior of endogenous proteins.

[00303] PCT Publication WO2008070363 assigned to Imclone Systems Inc. describes a single domain intrabody that binds to an intracellular protein or to an intracellular domain of an intracellular protein, such as Etk, the endothelial and epithelial tyrosine kinase, which is a member of the Tec family of non-receptor tyrosine kinases. Also provided is a method of inhibiting an intracellular enzyme, and treating a tumor in a patient by administering the intrabody or a nucleic acid expressing the intrabody.

[00304] PCT Publication WO2009018438 assigned to Cornell Research Foundation Inc. describes a method of identifying a protein that binds to a target molecule and has

intracellular functionality, by providing a construct comprising a DNA molecule encoding the protein which binds to the target molecule, with the DNA molecule being coupled to a stall sequence. A host cell is transformed with the construct and then cultured under conditions effective to form, within the host cell, a complex of the protein whose translation has been stalled, the mRNA encoding the protein, and ribosomes. The protein in the complex is in a properly folded, active form and the complex is recovered from the cell. This method can be carried out with a cell-free extract preparation containing ribosomes instead of a host cell. The present invention also relates to a construct which includes a DNA molecule encoding a protein that binds to a target molecule and an SecM stalling sequence coupled to the DNA molecule. The DNA molecule and the SecM stalling sequence are coupled with sufficient distance between them to permit expression of their encoded protein, within the cell, in a properly folded, active form. The use of intrabodies is generally described.

[00305] PCT Publication WO2014030780 assigned to Mogam Biotech Research Institute describes a method named Tat-associated protein engineering (TAPE), for screening a target protein having higher solubility and excellent thermostability, in particular, an immunoglobulin variable domain (VH or VL) derived from human germ cells, by preparing a gene construct where the target protein and an antibiotic -resistant protein are linked to a Tat signal sequence, and then expressing this within *E. coli*. Also disclosed are human or engineered VH and VL domain antibodies and human or engineered VH and VL domain antibody scaffolds having solubility and excellent thermostability, which are screened by the TAPE method. Also provided is a library including random CDR sequences in the human or engineered VH or VL domain antibody scaffold screened by the TAPE method, a preparing method thereof, a VH or VL domain antibody having binding ability to the target protein screened by using the library, and a pharmaceutical composition including the domain antibody.

[00306] European Patent Application EP2422811 describes an antibody that binds to an intracellular epitope; such intrabodies comprise at least a portion of an antibody that is capable of specifically binding an antigen and preferably does not contain operable sequences coding for its secretion and thus remains within the cell. In one embodiment, the intrabody comprises a scFv. The scFv polypeptide further comprises a polypeptide linker between the VH and VL domains which enables the scFv to form the desired structure for antigen binding. Also described is a specific embodiment in which the intrabody binds to the cytoplasmic domain of an Eph receptor and prevents its signaling (*e.g.*, autophosphorylation).

In another specific embodiment, an intrabody binds to the cytoplasmic domain of a B-type Ephrin (*e.g.*, EphrinB1, EphrinB2 or EphrinB3).

[00307] PCT Publication WO2011003896 and European Patent Application EP2275442 describe intracellular functional PCNA-Chromobodies made using nucleic acid molecule encoding a polypeptide specifically binding to proliferating cell nuclear antigen (PCNA). Examples of such polypeptides comprising conservative substitutions of one or more amino acids in one or two framework regions are represented by SEQ ID NOs: 16 and 18 disclosed therein, including the framework region of the polypeptide. In the examples, the framework regions as well as the CDR regions involved in the binding of PCNA have been determined.

[00308] European Patent Application EP2703485 describes a method for selecting plasma cells or plasmablasts, as well as for producing target antigen specific antibodies, and novel monoclonal antibodies. In one embodiment, cells expressing intracellular immunoglobulin were identified.

Proteins and Variants

[00309] Glycan-interacting antibodies of the present invention may exist as a whole polypeptide, a plurality of polypeptides or fragments of polypeptides, which independently may be encoded by one or more nucleic acids, a plurality of nucleic acids, fragments of nucleic acids or variants of any of the aforementioned. As used herein, “polypeptide” means a polymer of amino acid residues (natural or unnatural) linked together most often by peptide bonds. The term, as used herein, refers to proteins, polypeptides, and peptides of any size, structure, or function. In some instances the polypeptide encoded is smaller than about 50 amino acids and the polypeptide is then termed a peptide. If the polypeptide is a peptide, it will be at least about 2, 3, 4, or at least 5 amino acid residues long. Thus, polypeptides include gene products, naturally occurring polypeptides, synthetic polypeptides, homologs, orthologs, paralog, fragments and other equivalents, variants, and analogs of the foregoing. A polypeptide may be a single molecule or may be a multi-molecular complex such as a dimer, trimer or tetramer. They may also comprise single chain or multichain polypeptides and may be associated or linked. The term polypeptide may also apply to amino acid polymers in which one or more amino acid residues are an artificial chemical analogue of a corresponding naturally occurring amino acid.

[00310] The term “polypeptide variant” refers to molecules which differ in their amino acid sequence from a native or reference sequence. The amino acid sequence variants may possess

substitutions, deletions, and/or insertions at certain positions within the amino acid sequence, as compared to a native or reference sequence. Ordinarily, variants will possess at least about 50% identity (homology) to a native or reference sequence, and preferably, they will be at least about 80%, more preferably at least about 90% identical (homologous) to a native or reference sequence.

[00311] In some embodiments “variant mimics” are provided. As used herein, the term “variant mimic” is one which contains one or more amino acids which would mimic an activated sequence. For example, glutamate may serve as a mimic for phospho-threonine and/or phospho-serine. Alternatively, variant mimics may result in deactivation or in an inactivated product containing the mimic, e.g., phenylalanine may act as an inactivating substitution for tyrosine; or alanine may act as an inactivating substitution for serine. The amino acid sequences of the glycan-interacting antibodies of the invention may comprise naturally occurring amino acids and as such may be considered to be proteins, peptides, polypeptides, or fragments thereof.

Alternatively, the glycan-interacting antibodies may comprise both naturally and non-naturally occurring amino acids.

[00312] The term "amino acid sequence variant" refers to molecules with some differences in their amino acid sequences as compared to a native or starting sequence. The amino acid sequence variants may possess substitutions, deletions, and/or insertions at certain positions within the amino acid sequence. “Native” or “starting” sequence should not be confused with a wild type sequence. As used herein, a native or starting sequence is a relative term referring to an original molecule against which a comparison may be made. “Native” or “starting” sequences or molecules may represent the wild-type (that sequence found in nature) but do not have to be the wild-type sequence.

[00313] Ordinarily, variants will possess at least about 70% homology to a native sequence, and preferably, they will be at least about 80%, more preferably at least about 90% homologous to a native sequence. "Homology" as it applies to amino acid sequences is defined as the percentage of residues in the candidate amino acid sequence that are identical with the residues in the amino acid sequence of a second sequence after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent homology. Methods and computer programs for the alignment are well known in the art. It is understood that homology depends on a calculation of percent identity but may differ in value due to gaps and penalties introduced in the calculation.

[00314] By "homologs" as it applies to amino acid sequences is meant the corresponding sequence of other species having substantial identity to a second sequence of a second species.

"Analog" is meant to include polypeptide variants which differ by one or more amino acid alterations, e.g., substitutions, additions or deletions of amino acid residues that still maintain the properties of the parent polypeptide.

[00315] The present invention contemplates several types of glycan-interacting antibodies which are amino acid based including variants and derivatives. These include substitutional, insertional, deletion and covalent variants and derivatives. As such, included within the scope of this invention are glycan-interacting antibody molecules containing substitutions, insertions and/or additions, deletions and covalently modifications. For example, sequence tags or amino acids, such as one or more lysines, can be added to the peptide sequences of the invention (e.g., at the N-terminal or C-terminal ends). Sequence tags can be used for peptide purification or localization. Lysines can be used to increase peptide solubility or to allow for biotinylation. Alternatively, amino acid residues located at the carboxy and amino terminal regions of the amino acid sequence of a peptide or protein may optionally be deleted providing for truncated sequences. Certain amino acids (e.g., C-terminal or N-terminal residues) may alternatively be deleted depending on the use of the sequence, as for example, expression of the sequence as part of a larger sequence which is soluble, or linked to a solid support.

[00316] "Substitutional variants" when referring to proteins are those that have at least one amino acid residue in a native or starting sequence removed and a different amino acid inserted in its place at the same position. The substitutions may be single, where only one amino acid in the molecule has been substituted, or they may be multiple, where two or more amino acids have been substituted in the same molecule.

[00317] As used herein the term "conservative amino acid substitution" refers to the substitution of an amino acid that is normally present in the sequence with a different amino acid of similar size, charge, or polarity. Examples of conservative substitutions include the substitution of a non-polar (hydrophobic) residue such as isoleucine, valine and leucine for another non-polar residue. Likewise, examples of conservative substitutions include the substitution of one polar (hydrophilic) residue for another such as between arginine and lysine, between glutamine and asparagine, and between glycine and serine. Additionally, the substitution of a basic residue such as lysine, arginine or histidine for another, or the

substitution of one acidic residue such as aspartic acid or glutamic acid for another acidic residue are additional examples of conservative substitutions. Examples of non-conservative substitutions include the substitution of a non-polar (hydrophobic) amino acid residue such as isoleucine, valine, leucine, alanine, methionine for a polar (hydrophilic) residue such as cysteine, glutamine, glutamic acid or lysine and/or a polar residue for a non-polar residue.

[00318] "Insertional variants" when referring to proteins are those with one or more amino acids inserted immediately adjacent to an amino acid at a particular position in a native or starting sequence. "Immediately adjacent" to an amino acid means connected to either the alpha-carboxy or alpha-amino functional group of the amino acid.

[00319] "Deletional variants" when referring to proteins, are those with one or more amino acids in the native or starting amino acid sequence removed. Ordinarily, deletional variants will have one or more amino acids deleted in a particular region of the molecule.

[00320] As used herein, the term "derivative" is used synonymously with the term "variant" and refers to a molecule that has been modified or changed in any way relative to a reference molecule or starting molecule. In some embodiments, derivatives include native or starting proteins that have been modified with an organic proteinaceous or non-proteinaceous derivatizing agent, and post-translational modifications. Covalent modifications are traditionally introduced by reacting targeted amino acid residues of the protein with an organic derivatizing agent that is capable of reacting with selected side-chains or terminal residues, or by harnessing mechanisms of post-translational modifications that function in selected recombinant host cells. The resultant covalent derivatives are useful in programs directed at identifying residues important for biological activity, for immunoassays, or for the preparation of anti-protein antibodies for immunoaffinity purification of the recombinant glycoprotein. Such modifications are within the ordinary skill in the art and are performed without undue experimentation.

[00321] Certain post-translational modifications are the result of the action of recombinant host cells on the expressed polypeptide. Glutaminyl and asparaginyl residues are frequently post-translationally deamidated to the corresponding glutamyl and aspartyl residues.

Alternatively, these residues are deamidated under mildly acidic conditions. Either form of these residues may be present in the proteins used in accordance with the present invention.

[00322] Other post-translational modifications include hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, methylation of the alpha-

amino groups of lysine, arginine, and histidine side chains (T. E. Creighton, *Proteins: Structure and Molecular Properties*, W.H. Freeman & Co., San Francisco, pp. 79-86 (1983)).

[00323] Covalent derivatives specifically include fusion molecules in which proteins of the invention are covalently bonded to a non-proteinaceous polymer. The non-proteinaceous polymer ordinarily is a hydrophilic synthetic polymer, i.e. a polymer not otherwise found in nature. However, polymers which exist in nature and are produced by recombinant or in vitro methods are useful, as are polymers which are isolated from nature. Hydrophilic polyvinyl polymers fall within the scope of this invention, e.g. polyvinylalcohol and polyvinylpyrrolidone. Particularly useful are polyvinylalkylene ethers such as polyethylene glycol, polypropylene glycol. The proteins may be linked to various non-proteinaceous polymers, such as polyethylene glycol, polypropylene glycol or polyoxyalkylenes, in the manner set forth in U.S. Pat. No. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192 or 4,179,337.

[00324] "Features" when referring to proteins are defined as distinct amino acid sequence-based components of a molecule. Features of the proteins of the present invention include surface manifestations, local conformational shape, folds, loops, half-loops, domains, half-domains, sites, termini or any combination thereof.

[00325] As used herein when referring to proteins the term "surface manifestation" refers to a polypeptide based component of a protein appearing on an outermost surface.

[00326] As used herein when referring to proteins the term "local conformational shape" means a polypeptide based structural manifestation of a protein which is located within a definable space of the protein.

[00327] As used herein when referring to proteins the term "fold" means the resultant conformation of an amino acid sequence upon energy minimization. A fold may occur at the secondary or tertiary level of the folding process. Examples of secondary level folds include beta sheets and alpha helices. Examples of tertiary folds include domains and regions formed due to aggregation or separation of energetic forces. Regions formed in this way include hydrophobic and hydrophilic pockets, and the like.

[00328] As used herein the term "turn" as it relates to protein conformation means a bend which alters the direction of the backbone of a peptide or polypeptide and may involve one, two, three or more amino acid residues.

[00329] As used herein when referring to proteins the term "loop" refers to a structural feature of a peptide or polypeptide which reverses the direction of the backbone of a peptide

or polypeptide and comprises four or more amino acid residues. Oliva et al. have identified at least 5 classes of protein loops (J. Mol Biol 266 (4): 814-830; 1997).

[00330] As used herein when referring to proteins the term "half-loop" refers to a portion of an identified loop having at least half the number of amino acid residues as the loop from which it is derived. It is understood that loops may not always contain an even number of amino acid residues. Therefore, in those cases where a loop contains or is identified to comprise an odd number of amino acids, a half-loop of the odd-numbered loop will comprise the whole number portion or next whole number portion of the loop (number of amino acids of the loop/2+/-0.5 amino acids). For example, a loop identified as a 7 amino acid loop could produce half-loops of 3 amino acids or 4 amino acids ($7/2=3.5\pm0.5$ being 3 or 4).

[00331] As used herein when referring to proteins the term "domain" refers to a motif of a polypeptide having one or more identifiable structural or functional characteristics or properties (e.g., binding capacity, serving as a site for protein-protein interactions).

[00332] As used herein when referring to proteins the term "half-domain" means portion of an identified domain having at least half the number of amino acid residues as the domain from which it is derived. It is understood that domains may not always contain an even number of amino acid residues. Therefore, in those cases where a domain contains or is identified to comprise an odd number of amino acids, a half-domain of the odd-numbered domain will comprise the whole number portion or next whole number portion of the domain (number of amino acids of the domain/2+/-0.5 amino acids). For example, a domain identified as a 7 amino acid domain could produce half-domains of 3 amino acids or 4 amino acids ($7/2=3.5\pm0.5$ being 3 or 4). It is also understood that sub-domains may be identified within domains or half-domains, these subdomains possessing less than all of the structural or functional properties identified in the domains or half domains from which they were derived. It is also understood that the amino acids that comprise any of the domain types herein need not be contiguous along the backbone of the polypeptide (i.e., nonadjacent amino acids may fold structurally to produce a domain, half-domain or subdomain).

[00333] As used herein when referring to proteins the terms "site" as it pertains to amino acid based embodiments is used synonymous with "amino acid residue" and "amino acid side chain". A site represents a position within a peptide or polypeptide that may be modified, manipulated, altered, derivatized or varied within the polypeptide based molecules of the present invention.

[00334] As used herein the terms "termini or terminus" when referring to proteins refers to an extremity of a peptide or polypeptide. Such extremity is not limited only to the first or final site of the peptide or polypeptide but may include additional amino acids in the terminal regions. The polypeptide based molecules of the present invention may be characterized as having both an N-terminus (terminated by an amino acid with a free amino group (NH₂)) and a C-terminus (terminated by an amino acid with a free carboxyl group (COOH)). Proteins of the invention are in some cases made up of multiple polypeptide chains brought together by disulfide bonds or by non-covalent forces (multimers, oligomers). These sorts of proteins will have multiple N- and C-termini. Alternatively, the termini of the polypeptides may be modified such that they begin or end, as the case may be, with a non-polypeptide based moiety such as an organic conjugate.

[00335] Once any of the features have been identified or defined as a component of a molecule of the invention, any of several manipulations and/or modifications of these features may be performed by moving, swapping, inverting, deleting, randomizing or duplicating. Furthermore, it is understood that manipulation of features may result in the same outcome as a modification to the molecules of the invention. For example, a manipulation which involved deleting a domain would result in the alteration of the length of a molecule just as modification of a nucleic acid to encode less than a full length molecule would.

[00336] Modifications and manipulations can be accomplished by methods known in the art such as site directed mutagenesis. The resulting modified molecules may then be tested for activity using in vitro or in vivo assays such as those described herein or any other suitable screening assay known in the art.

Isotopic variations

[00337] The glycan-interacting antibodies of the present invention may contain one or more atoms that are isotopes. As used herein, the term "isotope" refers to a chemical element that has one or more additional neutron. In one embodiment, compounds of the present invention may be deuterated. As used herein, the term "deuterated" refers to a substance that has had one or more hydrogen atoms replaced by deuterium isotopes. Deuterium isotopes are isotopes of hydrogen. The nucleus of hydrogen contains one proton while deuterium nuclei contain both a proton and a neutron. The glycan-interacting antibodies may be deuterated in order to change a physical property of the compound, such as stability, or to allow the compounds to be used in diagnostic and experimental applications.

Conjugates and Combinations

[00338] It is contemplated by the present invention that the glycan-interacting antibodies of the present invention may be complexed, conjugated or combined with one or more homologous or heterologous molecules. As used herein, “homologous molecule” means a molecule which is similar in at least one of structure or function relative to a starting molecule while a “heterologous molecule” is one that differs in at least one of structure or function relative to a starting molecule. Structural homologs are therefore molecules which are substantially structurally similar. They can be identical. Functional homologs are molecules which are substantially functionally similar. They can be identical.

[00339] Glycan-interacting antibodies of the invention may comprise conjugates. Such conjugates of the invention may include a naturally occurring substance or ligand, such as a protein (e.g., human serum albumin (HSA), low-density lipoprotein (LDL), high-density lipoprotein (HDL), or globulin); a carbohydrate (e.g., a dextran, pullulan, chitin, chitosan, inulin, cyclodextrin or hyaluronic acid); or a lipid. The ligand may also be a recombinant or synthetic molecule, such as a synthetic polymer, e.g., a synthetic polyamino acid, an oligonucleotide (e.g. an aptamer). Examples of polyamino acids include polyamino acid is a polylysine (PLL), poly L-aspartic acid, poly L-glutamic acid, styrene-maleic acid anhydride copolymer, poly(L-lactide-co-glycolide) copolymer, divinyl ether-maleic anhydride copolymer, N-(2-hydroxypropyl)methacrylamide copolymer (HMPA), polyethylene glycol (PEG), polyvinyl alcohol (PVA), polyurethane, poly(2-ethylacrylic acid), N-isopropylacrylamide polymers, or polyphosphazine. Example of polyamines include: polyethylenimine, polylysine (PLL), spermine, spermidine, polyamine, pseudopeptide-polyamine, peptidomimetic polyamine, dendrimer polyamine, arginine, amidine, protamine, cationic lipid, cationic porphyrin, quaternary salt of a polyamine, or an alpha helical peptide.

[00340] The conjugates can also include targeting groups, e.g., a cell or tissue targeting agent or group, e.g., a lectin, glycoprotein, lipid or protein, e.g., an antibody, that binds to a specified cell type such as a kidney cell. A targeting group can be a thyrotropin, melanotropin, lectin, glycoprotein, surfactant protein A, mucin carbohydrate, multivalent lactose, multivalent galactose, N-acetyl-galactosamine, N-acetyl-glucosamine multivalent mannose, multivalent fucose, glycosylated polyaminoacids, multivalent galactose, transferrin, bisphosphonate, polyglutamate, polyaspartate, a lipid, cholesterol, a steroid, bile acid, folate, vitamin B12, biotin, an RGD peptide, an RGD peptide mimetic or an aptamer.

[00341] Targeting groups can be proteins, e.g., glycoproteins, or peptides, e.g., molecules having a specific affinity for a co-ligand, or antibodies e.g., an antibody, that binds to a specified cell type such as a cancer cell, endothelial cell, or bone cell. Targeting groups may also include hormones and hormone receptors. They can also include non-peptidic species, such as lipids, lectins, carbohydrates, vitamins, cofactors, multivalent lactose, multivalent galactose, N-acetyl-galactosamine, N-acetyl-glucosamine multivalent mannose, multivalent fucose, or aptamers.

[00342] The targeting group can be any ligand that is capable of targeting a specific receptor. Examples include, without limitation, folate, GalNAc, galactose, mannose, mannose-6P, aptamers, integrin receptor ligands, chemokine receptor ligands, transferrin, biotin, serotonin receptor ligands, PSMA, endothelin, GCPII, somatostatin, LDL, and HDL ligands. In particular embodiments, the targeting group is an aptamer. The aptamer can be unmodified or have any combination of modifications disclosed herein.

[00343] In still other embodiments, glycan-interacting antibodies are covalently conjugated to a cell penetrating polypeptide. The cell-penetrating peptide may also include a signal sequence. The conjugates of the invention can be designed to have increased stability; increased cell transfection; and/or altered biodistribution (e.g., targeted to specific tissues or cell types).

[00344] Conjugating moieties may be added to glycan-interacting antibodies such that they allow labeling or flagging targets for clearance. Such tagging/flagging molecules include, but are not limited to ubiquitin, fluorescent molecules, human influenza hemagglutinin (HA), c-myc [a 10 amino acid segment of the human protooncogene myc with sequence EQKLISEEDL (SEQ ID NO: 243)], histidine (His), flag [a short peptide of sequence DYKDDDDK (SEQ ID NO: 244)], glutathione S-transferase (GST), V5 (a paramyxovirus of simian virus 5 epitope), biotin, avidin, streptavidin, horse radish peroxidase (HRP) and digoxigenin.

[00345] In some embodiments, glycan-interacting antibodies may be combined with one another or other molecule in the treatment of a disease or condition.

Nucleic acids

[00346] The present invention embraces nucleic acid molecules. In some embodiments, nucleic acids encode antibodies of the invention (including, but not limited to antibodies, antibody fragments, intrabodies and chimeric receptor antigens). Such nucleic acid molecules

include, without limitation, DNA molecules, RNA molecules, polynucleotides, oligonucleotides, mRNA molecules, vectors, plasmids and other constructs. As used herein, the term "construct" refers to any recombinant nucleic acid molecule including, but not limited to plasmids, cosmids, autonomously replicating polynucleotide molecules or linear or circular single-stranded or double-stranded DNA or RNA polynucleotide molecules. The present invention also embraces cells programmed or generated to express nucleic acid molecules encoding glycan-interacting antibodies. Such cells may be generated through the use of transfection, electroporation, viral delivery and the like. Viruses engineered with constructs of the invention may include, but are not limited to lentiviruses, adenoviruses, adeno-associated viruses and phages. In some cases, nucleic acids of the invention include codon-optimized nucleic acids. Methods of generating codon-optimized nucleic acids are known in the art and may include, but are not limited to those described in US Patent Nos. 5,786,464 and 6,114,148, the contents of each of which are herein incorporated by reference in their entirety.

II. Methods and uses

Therapeutics

Cancer-related applications

[00347] Aberrant glycosylation is a hallmark of cancer cell transformation. Multiple aberrant glycosylation forms have been described in human cancers, identifying specific tumor-associated carbohydrate antigens (TACAs) as a class of cell surface molecules suitable for specific tumor targeting (Cheever, M.A. et al., Clin Cancer Res. 2009 Sep 1;15(17):5323-37). TACA antigen expression has been found in epithelial cancers including, but not limited to, breast, colon, lung, bladder, cervical, ovarian, stomach, prostate, and liver. TACA antigen expression has been found in embryonal cancers including, but not limited to, yolk sac tumors and seminomas. In addition, TACA antigen expression has been found in many melanomas, carcinomas, and leukemias of various tissues (Heimburg-Molinaro et al., Vaccine. 2011 Nov 8; 29(48):8802-8826). Antibodies of the present invention that target one or more TACA are referred to herein as "anti-TACA antibodies."

[00348] MUC1 is a key cell surface glycoprotein that is normally extensively glycosylated but is underglycosylated in tumor cells. Sparse glycosylation of MUC1 leads to exposure of immunogenic antigens. These may be along the MUC1 core peptide sequence or along core carbohydrate residues. These TACAs include, but are not limited to N-acetylgalactosamine

(Tn), sialyl(α 2,6)N-acetylgalactosamine (STn) and galactose(β 1-3)N-acetylgalactosamine (also known as Thomsen-Friedenreich antigen or TF). It has been estimated that about 80% of all carcinomas express Tn among the core carbohydrates of MUC1 with STn being strongly expressed on human carcinoma cells and linked to cancer progression and metastasis. With few exceptions, Tn and STn are not expressed in normal healthy tissues. Sialic acid forms a prominent epitope on STn. The invention takes advantage of the fact that aberrant Neu5Gc-STn (GcSTn) glycan expression appears to be highly specific to various carcinomas.

[00349] In the case of MUC1, Neu5Gc incorporation into STn yields a tumor-specific target, a site that is an attractive target for antibody-based therapies to treat tumor tissue. In some embodiments of the present invention, glycan-interacting antibodies target MUC1 expressing cancer cells comprising Neu5Gc. To date, Neu5Gc has been detected in glycoconjugates from a number of human cancer tissues including, but not limited to colon cancer, retinoblastoma tissue, melanoma, breast cancer and yolk sac tumor tissue. In some embodiments of the present invention, methods are contemplated for glycan-interacting antibody treatment of these forms of cancer as well as other forms of cancer, not specifically listed here, characterized by the presence of cancer cells comprising Neu5Gc.

[00350] Additional antigens comprising glycans have been identified that are expressed in correlation with cancer (Heimburg-Molinaro, J. et al., Cancer vaccines and carbohydrate epitopes. Vaccine. 2011 Nov 8;29(48):8802-26). These tumor-associated carbohydrate antigens include, but are not limited to blood group Lewis related antigens [including, but not limited to Lewis^Y (Le^Y), Lewis^X (Le^X), Sialyl Lewis^X (SLe^X) and Sialyl Lewis^A (SLe^A)], glycosphingolipid-related antigens [including, but not limited to Globo H, stage-specific embryonic antigen-3 (SSEA-3) and glycosphingolipids comprising sialic acid], ganglioside-related antigens [including, but not limited to gangliosides GD2, GD3, GM2, fucosyl GM1 and Neu5GcGM3] and polysialic acid-related antigens.

[00351] In some embodiments, therapeutics of the present invention may be directed toward Lewis blood group antigens. Lewis blood group antigens comprise a fucose residue linked to GlcNAc by an α 1-3 linkage or an α 1-4 linkage. They may be found on both glycolipids and glycoproteins. Lewis blood group antigens may be found in the body fluid of individuals that are secretors of these antigens. Their appearance on red cells is due to absorption of Lewis antigens from the serum by the red cells.

[00352] In some embodiments, therapeutics of the present invention may be directed toward Le^Y. Le^Y (also known as CD174) is made up of Galβ1,4GlcNAc comprising α1,2- as well as α1,3-linked fucose residues yielding the Fuca(1,2)Galβ(1,4)Fuca(1,3)GlcNAc epitope. It is synthesized from the H antigen by α1,3 fucosyltransferases which attach the α1,3 fucose to the GlcNAc residue of the parent chain. Le^Y may be expressed in a variety of cancers including, but not limited to ovarian, breast, prostate, colon, lung, and epithelial. Due to its low expression level in normal tissues and elevated expression level in many cancers, the Le^Y antigen is an attractive target for therapeutic antibodies.

[00353] In some embodiments, therapeutics of the present invention may be directed toward Le^X. Le^X comprises the epitope Galβ1-4(Fuca1-3)GlcNAcβ-R. It is also known as CD15 and stage-specific embryonic antigen-1 (SSEA-1). This antigen was first recognized as being immunoreactive with sera taken from a mouse subjected to immunization with F9 teratocarcinoma cells. Le^X was also found to correlate with embryonic development at specific stages. It is also expressed in a variety of tissues both in the presence and absence of cancer, but can also be found in breast and ovarian cancers where it is only expressed by cancerous cells.

[00354] In some embodiments, therapeutics of the present invention may be directed toward SLe^A and/or SLe^X. SLe^A and SLe^X comprise the structures [Neu5Acα2-3Galβ1-3(Fuca1-4)GlcNAcβ-R] and [Neu5Acα2-3Galβ1-4(Fuca1-3)GlcNAcβ-R] respectively. Their expression is upregulated in cancer cells. The presence of these antigens in serum correlates with malignancy and poor prognosis. SLe^X is mostly found as a mucin terminal epitope. It is expressed in a number of different cancers including breast, ovarian, melanoma, colon, liver, lung and prostate. In some embodiments of the present invention, SLe^A and SLe^X targets comprise Neu5Gc (referred to herein as GcSLe^A and GcSLe^X, respectively).

[00355] In some embodiments, therapeutics of the present invention may be directed toward glycolipids and/or epitopes present on glycolipids, including, but not limited to glycosphingolipids. Glycosphingolipids comprise the lipid ceramide linked to a glycan by the ceramide hydroxyl group. On the cell membrane, glycosphingolipids form clusters referred to as “lipid rafts”.

[00356] In some embodiments, therapeutics of the present invention may be directed toward Globo H. Globo H is a cancer-related glycosphingolipid first identified in breast cancer cells. The glycan portion of Globo H comprises Fuca(1-2)Galβ(1-3)GalNAcβ(1-3)Galα(1-4)Galβ(1-4)Glcβ(1). Although found in a number of normal epithelial tissues,

Globo H has been identified in association with many tumor tissues including, but not limited to, small cell lung, breast, prostate, lung, pancreatic, gastric, ovarian and endometrial tumors.

[00357] In some embodiments, therapeutics of the present invention may be directed toward gangliosides. Gangliosides are glycosphingolipids comprising sialic acid. According to ganglioside nomenclature, G is used as an abbreviation for ganglioside. This abbreviation is followed by the letters M, D or T referring to the number of sialic acid residues attached (1, 2 or 3 respectively). Finally the numbers 1, 2 or 3 are used to refer to the order of the distance each migrates when analyzed by thin layer chromatography (wherein 3 travels the greatest distance, followed by 2 and then 1). Gangliosides are known to be involved in cancer-related growth and metastasis and are expressed on the cell surface of tumor cells. Gangliosides expressed on tumor cells include, but are not limited to GD2, GD3, GM2 and fucosyl GM1 (also referred to herein as Fuc-GM1). In some embodiments of the present invention, glycan-interacting antibodies are directed toward GD3. GD3 is a regulator of cell growth. In some embodiments, GD3-directed antibodies are used to modulate cell growth and/or angiogenesis. In some embodiments, GD3-directed antibodies are used to modulate cell attachment. In some embodiments of the present invention, glycan interacting antibodies are directed toward GM2. In some embodiments, GM2-directed antibodies are used to modulate cell to cell contact. In some embodiments, ganglioside targets of the present invention comprise Neu5Gc. In some embodiments, such targets may include a GM3 variant comprising Neu5Gc (referred to herein as GcGM3). The glycan component of GcGM3 is Neu5Gc α 2-3Gal β 1-4Glc. GcGM3 is a known component of tumor cells.

[00358] In some embodiments, TACAs targeted by anti-TACA antibodies of the present invention may include, but are not limited to any of those listed in US Publication Nos. US2013/0236486A1, US2013/0108624A1, US2010/0178292A1, US2010/0104572A1, US2012/0039984A1, US2009/0196916A1, and US2009/0041836A1, the contents of each of which are herein incorporated by reference in their entirety.

STn in Cancer

[00359] The immune system has multiple mechanisms for promoting anti-tumor cell immune activity including both innate and adaptive immune activity. As used herein, the term “anti-tumor cell immune activity” refers to any activity of the immune system that kills or prevents growth and/or proliferation of tumor cells. In some cases, anti-tumor immune activity includes recognition and tumor cell killing by natural killer (NK) cells and

phagocytosis by macrophages. Adaptive anti-tumor immune responses include tumor antigen uptake and presentation by antigen presenting cells (APCs,) such as dendritic cells (DCs,) leading to modulation of T cell anti-tumor activity and/or expansion of B cells with secretion of tumor-specific antibodies. The binding of tumor-specific antibodies to tumors can lead to antibody-dependent cellular cytotoxicity (ADCC) and complement-dependent cytotoxicity (CDC) mechanisms of tumor cell death.

[00360] As used herein, the term “immune-resistant tumor cell” refers to a tumor cell that reduces or evades anti-tumor cell immune activity. Some studies indicate that the expression of STn (a known TACA) on tumor cell surfaces or secreted into the tumor cell microenvironment can promote tumor cell evasion of anti-tumor immune activity. As used herein, the term “tumor cell microenvironment” refers to any area adjacent to or surrounding a tumor cell. Such areas include, but are not limited to areas between tumor cells, between tumor and non-tumor cells, surrounding fluids and surrounding components of the extracellular matrix.

[00361] Sialylated mucins comprising STn were demonstrated by Ogata et al to reduce NK cell targeting of tumor cells (Ogata, S. et al., 1992. *Canc. Res.* 52:4741-6, the contents of which are herein incorporated by reference in their entirety). This study found that the presence of ovine, bovine and porcine submaxillary mucin (OSM, BSM and PSM, respectively) led to nearly one hundred percent inhibition of cytotoxicity (see Table 2 of Ogata et al). Further studies by Jandus et al, demonstrate that some tumor cells can evade NK destruction due to the expression of sialoglycan ligands that can interact with NK cell siglec receptors, leading to NK inhibition (Jandus, C. et al., 2014, *JCI*. pii: 65899, the contents of which are herein incorporated by reference in their entirety).

[00362] Studies by Toda et al., demonstrate that STn may bind CD22 receptors on B cells, leading to decreased signal transduction and reduced B cell activation (Toda, M. et al., 2008. *Biochem Biophys Res Commun.* 372(1):45-50, the contents of which are herein incorporated by reference in their entirety). Dendritic cells (DCs) can affect adaptive immune activity by modulating T cell activity. Studies by Carrascal et al found that STn expression by bladder cancer cells induced tolerance in DCs, reducing their ability to induce anti-tumor cell immune activity in T cells (Carrascal, MA et al., 2014. *Mol Oncol.* pii: S1574-7891(14)00047-7, the contents of which are herein incorporated by reference in their entirety). These studies revealed that DCs coming into contact with STn-positive bladder cancer cells displayed a tolerogenic expression profile with low expression of CD80, CD86, IL-12 and TNF- α .

Further, DCs were found to modulate regulatory T cells such that the T cells had low expression of IFN γ and high expression of FoxP3. Other studies by van Vliet and others, indicate that DC surface expression of macrophage galactose-type lectin (MGL) can lead to targeting of those cells to tumor tissues (van Vliet, SJ., 2007. Amsterdam: Vrije Universiteit. p1-232 and van Vliet, SJ. et al., 2008. J Immunol. 181(5):3148-55, Nollau, P. et al., 2013. J Histochem Cytochem. 61(3):199-205, the contents of each of which are herein incorporated by reference in their entirety). DCs arriving at tissues due to MGL interactions may influence T helper (Th) cells in one of three ways. DCs can induce T cell tolerance, T cell immune activity or downregulation of effector T cells. MGL has been shown to bind to both AcSTn and GcSTn and the affinity has been analyzed in depth (Mortezai, N. et al., 2013. Glycobiology. 23(7):844-52, the contents of which are herein incorporated by reference in their entirety). Interestingly, MUC1 expression on tumors has been shown to lead to T cell tolerance, protecting tumor cells from immune eradication.

[00363] In some embodiments, glycan-interacting antibodies (including, but not limited to anti-STn antibodies) of the present invention may be used to treat subjects comprising one or more tumor cells expressing one or more TACAs. In some cases, glycan-interacting antibodies (including, but not limited to anti-STn antibodies) of the invention may be used to increase anti-tumor cell immune activity toward tumor cells expressing STn. Such antibodies may increase the adaptive immune response and/or the innate immune response toward immune-resistant tumor cells. Some glycan-interacting antibodies may be used to increase NK anti-tumor cell activity. Such glycan-interacting antibodies may, in some cases, block the interaction between glycan receptors expressed on NK cells and STn glycans on cancer cells or in surrounding tissues.

[00364] In some embodiments, glycan-interacting antibodies (including, but not limited to anti-STn antibodies) of the invention may be used to increase B cell anti-tumor cell activity. Such antibodies may reduce the interaction between CD22 receptors on B cells and STn glycans on cancer cells or in surrounding tissues. A study by Sjoberg et al. demonstrates that 9-O-acetylation of α 2,6-linked sialic acids on glycoproteins also reduced interaction between B cell CD22 receptors and such glycoproteins (Sjoberg, E.R. et al. 1994. JCB. 126(2): 549-562). Another study by Shi et al. reveals that higher levels of 9-O-acetylated sialic acid residues on murine erythroleukemia cells makes these cells more susceptible to complement-mediated lysis (Shi, W-X. et al., 1996. J of Biol Chem. 271(49): 31526-32, the contents of which are herein incorporated by reference in their entirety). In some embodiments, anti-STn

antibodies of the invention are capable of selectively binding non-9-O-acetylated STn, reducing overall STn binding, but reducing tumor cell growth and/or proliferation (e.g., through increased B cell anti-tumor activity and increased complement-mediated tumor cell destruction). In some embodiments, glycan-interacting antibodies (including, but not limited to anti-STn antibodies) of the invention may be used to increase DC anti-tumor activity. Such antibodies may be used to reduce DC tolerance to tumor cells. Reduced DC tolerance may comprise increasing DC expression of CD80, CD86, IL-12 and/or TNF- α . In some cases, DC anti-tumor cell activity may comprise promotion of T cell anti-tumor cell activity. Such antibodies may prevent binding between DC MGL and glycans expressed on or around cancer cells.

[00365] A study by Ibrahim et al. suggests that high levels of anti-STn antibodies along with endocrine therapy may increase overall survival and time to progression (TTP) in women with metastatic breast cancer (Ibrahim, N.K. et al., 2013. 4(7): 577-584, the contents of which are herein incorporated by reference in their entirety). In this study, anti-STn antibody levels were elevated after vaccination with STn linked to keyhole-limpet Hemocyanin (KLH). In some embodiments, anti-STn antibodies of the invention may be used in combination with endocrine therapy (e.g. tamoxifen and/or an aromatase inhibitor).

Cancer stem cells as therapy targets

[00366] Cancer stem cells or CSCs (also called tumor initiating cells) are a subset of cells within a heterogeneous tumor population that drive the initiation, growth, dissemination, and recurrence of primary and metastatic tumors (Karsten and Goletz, SpringerPlus, 2013, 2, 301), which can occur in varying proportions of the total population depending on tumor type. CSCs are distinguished from terminally differentiated cells by their capacity to self-renew and give rise to non-CSC, differentiated progeny (Gupta et al., Nature medicine, 2009, 15, 1010-1012). These properties are akin to those of normal stem cells. Such distinctions between normal stem cells and CSCs have important implications for therapy.

[00367] An increasing number of cell-surface biomarkers have been identified that purport to differentiate CSCs from their non-CSC counterparts (Medema et al., Nature cell biology, 2013, 15, 338-344; Zoller, Cancer, 2011, 11, 254-267). Although many of these derive from studies of mouse tumors and human cell lines, several have been validated using primary human tumor samples. One of these, the membrane-spanning CD44 glycoprotein, or hyaluronan receptor, which is a well-known constituent of a variety of tumor types, has also

more recently found acceptance as a bona fide CSC marker in human cancers, and in fact is the one most frequently observed (Lobo et al., 2007, 23, 675-699).

[00368] CD44 exists in several variant isoforms generated by alternative splicing events occurring among the 20 exons and 19 introns of the full-length CD44 gene (Williams et al, Experimental biology and medicine, 2013, 238, 324-338). Growing experimental evidence points to the supporting role of CD44 and its variants in contributing to the innate metastatic and drug resistant phenotype of CSCs (Negi et al., Journal of drug targeting, 2012, 20, 561-573), in part due to modulation of intracellular signal transduction pathways (Williams et al, Experimental biology and medicine, 2013, 238, 324-338). Additionally, patients with triple negative breast cancer, along with several other cancer types, that display high levels of CD44 cells are known to have a poor prognosis and higher mortality (Negi et al., Journal of drug targeting, 2012, 20, 561-573). These observations support the notion that targeting CD44 offers a means of treating cancer through inhibition or elimination of CSCs, in addition to mature cancer cells. Indeed, numerous approaches to targeting CD44 have been attempted experimentally with varying degrees of success. These comprise a wide range of technologies that include the use of conjugated and unconjugated antibodies, nano-carrier drug systems, and hyaluronan-conjugated drugs (Negi et al., Journal of drug targeting, 2012, 20, 561-573). In several instances, however, toxic effects were observed in in vivo studies; these untoward side effects may be attributable to the widespread occurrence of CD44 and variants on the membranes of most vertebrate cells (Naor et al., Seminars in cancer biology, 2008, 18, 260-267), in addition to its presence on the surface of the targeted CSCs and mature tumor cells. Targeting CD44 protein, which is a constituent of normal human stem cells (Williams et al, Experimental biology and medicine, 2013, 238, 324-338), can also harm normal stem cell function (Leth-Larsen et al., Molecular medicine, 2012, 18, 1109-1121). Although a large body of research points to the desirability of targeting CD44 protein on CSCs, as well as on mature tumor cells, the intrinsic problem with this approach remains the present difficulty in designing inhibitors that will spare normal tissue as well as normal stem cells.

[00369] Another well-known tumor antigen with implications to CSC biology is the epithelial mucin MUC1, a membrane tethered glycoprotein that is differentially expressed at high levels on the majority of adenocarcinomas but at low levels or not at all on normal epithelial cells. MUC1 has recently been identified as a CSC biomarker on a variety of neoplasias including breast (Engelmann et al., Cancer research, 2008, 68, 2419-2426), and pancreatic cancers, where its expression is correlated with high metastasis and poor

prognosis. As a constituent of CSCs, MUC1 has been shown to function in cell adhesion, proliferation, survival, and signaling (Engelmann et al., Cancer research, 2008, 68, 2419-2426) and may also be co-expressed with CD44 (Leth-Larsen et al., Molecular medicine, 2012, 18, 1109-1121). Immunotherapeutic approaches for targeting MUC1 in cancer are being pursued using vaccines as well as other approaches, but primarily in the context of mature cancer cell therapy (Julien et al., Biomolecules, 2012, 2, 435-466; Acres et al., Expert review of vaccines, 2005, 4, 493-502).

[00370] Cancer stem cells have been hypothesized to be generated through the epithelial-to-mesenchymal (EMT) transition (Gupta et al., Nature medicine, 2009, 15, 1010-1012), and /or reversely the mesenchymal-to-epithelial (MET) transition that occurs at the site of metastasis (Leth-Larsen et al., Molecular medicine, 2012, 18, 1109-1121) (also called CSCs plasticity where non-CSCs can give rise to CSCs). This discovery further underscores the need to eliminate both CSCs and non-CSCs in a tumor population.

[00371] Recent studies with enriched CSC populations has revealed that these cells, unlike the bulk of the tumor, are relatively quiescent and are preferentially resistant to many types of current therapies, including chemotherapy and radiation (Leth-Larsen et al., Molecular medicine, 2012, 18, 1109-1121). Thus current therapeutic strategies target non-CSC components of the tumor, leaving CSCs largely unaffected only to re-emerge after appropriate cues to reform recurrent primary tumors at the initial site or to disseminate to distant sites, colonize, and create metastatic disease, the major cause of cancer mortality.

[00372] Current understanding of the properties of cancer stem cells clearly emphasized the need not only to target the bulk of cells present in tumors, as is current practice, but also the CSC compartment in order to potentially effect complete cures.

[00373] As discussed above, strategies that have been developed based on tumor (including CSCs) associated biomarkers face a challenge that most cancer biomarkers are also present in normal cells including normal stem cells. A therapy that targets a protein biomarker to eliminate CSCs, may also target normal stem cells, causing elimination of normal cells.

Tumor-specific glycans in CSCs

[00374] Aberrant forms of glycosylation, including appearance of the Thomsen-nouveau (Tn) antigen (GalNAc-O-Ser/Thr), have been described in numerous human cancers, identifying glycans as an entirely novel class of tumor-associated carbohydrate antigens suitable for specific tumor targeting (Rabu et al., Future oncology, 2012, 8, 943-960). The

formation of the sialyl derivative of Tn (STn) is mediated by the sialyl transferase ST6GalNAc-I which adds sialic acid in an α 2,6 linkage to the Tn antigen. The sialylation of STn prevents further sugar additions, thus truncating further glycan extensions (Schultz et al., Cancer metastasis reviews, 2012, 31, 501-518).

[00375] While the presence of STn in normal adult human tissues is rare, STn occurs in various human cancers, including ovarian, bladder, breast, cervical, colon, and lung cancer, among others (Ferreira et al., Molecular oncology, 2013, 7, 719-731; Kinney et al., Cancer, 1997, 80, 2240-2249). Further, the presence of STn in tumors is associated with metastatic disease, poor prognosis, and reduced overall survival (Ferreira et al., Molecular oncology, 2013, 7, 719-731; Kinney et al., Cancer, 1997, 80, 2240-2249); therefore, STn is considered a highly attractive target for cancer detection and therapy. There are two distinct forms of sialic acid – Neu5Ac and Neu5Gc – located at the terminal position of STn. The Neu5Ac-sialylated form is predominant in humans since humans cannot synthesize Neu5Gc due to an inactive CMP-Neu5Ac hydroxylase (CMAH) gene. However, consumption of Neu5Gc-rich foods leads to foreign Neu5Gc incorporation into human cells, especially in carcinomas. Previous studies have shown that solid tumors take up and express the Neu5Gc form of sialic acid (Inoue et al., Glycobiology, 2010, 20, 752-762; Malykh et al., Biochimie, 2001, 83, 623-634; Padler-Karavani et al., Cancer research, 2011, 71, 3352-3363). mAbs that bind to both glyco-isoforms of STn that are potential cancer targets: Neu5Ac-STn (AcSTn) and Neu5Gc-STn (GcSTn) (i.e., designated as pan-STn antibodies).

[00376] STn accumulation is associated with specific somatic mutations observed repeatedly in solid tumors and with the inactivation of the gene that encodes the molecular chaperone Cosmc, which is required for the formation of active T-synthase (Ju et al., Nature, 2005, 437, 125). T-synthase competes with ST6GalNAc-I for the GalNAc substrate and therefore when inactivated by mutation results in elevated STn synthesis. Additionally, STn accumulation can also result from increased expression of ST6GalNAc-I, which is often observed (Brockhausen et al., Biological chemistry, 2001, 382, 219-232; Ikehara et al., Glycobiology, 1999, 9, 1213-1224). De novo expression of STn can modulate carcinoma cells, change the malignant phenotype, and lead to more aggressive cell behaviors (Pinho et al., Cancer letters, 2007, 249, 157-170). As such, STn is not only an interesting cancer biomarker and therapeutic target, but interfering with STn function offers the intriguing potential to have significant functional, anti-metastatic therapeutic benefits.

[00377] Although it is well-known that glycosylation of cellular glycoproteins is altered in cancer, it appears that aberrant glycosylation is selective with respect to both the glycoprotein and glycan in question. In fact, in human tumor CSCs only CD44 and MUC1 are major carriers of the STn antigen (Cazet et al., Breast cancer research: BCR, 2010, 12,204; Julien et al., Glycobiology, 2006, 16, 54-64), immediately suggesting a selective approach for targeting not only mature tumor cells but also CSCs. Whereas MUC1 is a normal surface constituent of some epithelial cells where it serves a barrier function. Tumor-associated MUC1 is characterized by hypoglycosylation and increased sialylation on CSCs in the same fashion as observed in mature cancer cells, with STn appearing as a specific marker for both CSCs and mature tumor cells (Curry et al., Journal of surgical oncology, 2013, 107, 713-722). The aberrant oligosaccharide profile of MUC1 gives rise to the expression of neomarkers such as sialyl-Le^a (used in the CA19-9 test), sialyl-Le^x, and sialyl-Tn (TAG-72), as well as the cryptic epitopes such as Tn in cancer cells (e.g., CSCs). In addition, because of underglycosylation, the peptide core of the mucin becomes exposed such that epitopes within the core (not accessible within normal tissue-derived MUC1) may serve as potential antigens.

[00378] Clinical approaches targeting STn have thus far consisted solely of STn vaccines. The most advanced clinical candidate is Theratope, a therapeutic vaccine consisting of STn coupled to keyhole limpet hemocyanin. In in vivo mouse studies Theratope immunization induced a potent antibody response that was shown to mediate a delay in the growth of injected STn-expressing mammary carcinoma cells (Julien et al., British journal of cancer, 2009, 100, 1746-1751). However, Theratope failed to meet its primary endpoint in a phase III clinical trial in metastatic breast cancer. A leading hypothesis for why the Theratope trial missed its primary endpoint is that the patient population was not evaluated for STn expression prior to enrollment. Since STn expression in breast cancer is highly heterogeneous between patients, ranging from 25%-80% depending on the study and detection method, lack of ability to correlate STn expression with response may have masked any benefit from Theratope. Importantly, a subset of patients receiving hormonal therapy showed a significant 7.5 month increase in median overall survival when treated with Theratope compared to hormone therapy alone (Ibrahim et al., Journal of clinical oncology: official journal of the American Society of Clinical Oncology, 2004, 22, 2547; and Miles et al., The oncologist, 2011, 16, 1092-1100), validating the therapeutic potential of targeting STn in particular patient populations. Additionally, since the immune response often varies considerably between vaccinated patients, vaccine approaches lack the ability to control or modulate

antibody titer, resulting in wide ranges of therapeutic antibody exposure among patients. Nonetheless, Theratope was well tolerated with minimal toxicity, demonstrating the safety of targeting STn for cancer therapy.

[00379] The growing understanding of the molecular basis of STn expression in cancer cells strongly suggests that cells that express STn on any cell surface protein will also express STn on many (if not all) other O-glycosylated cell surface proteins, rendering it an excellent widely-distributed cancer-associated therapeutic target. Thus, STn positive cancer cell populations may be enriched for CSCs. In addition, recent data demonstrate that abrogation of STn expression renders cancers less aggressive with significant reductions in metastatic behavior (Gill et al., Proceedings of the National Academy of Sciences of the United States of America 2013, 110, E3152-3161).

Anti-STn antibodies targeting CSCs as cancer treatment

[00380] Several anti-STn antibodies have been described in the field, but some demonstrate low specificity towards the STn antigen or sialylated isoforms. For example, the commercial B72.3 anti-STn antibody has been shown to bind not only to STn but also to the Tn antigen (Bapat, S. A. (2010) Human ovarian cancer stem cells. Reproduction 140, 33-41). The availability of monoclonal antibodies (mAbs) targeting STn, engineered to induce antibody-dependent cellular cytotoxicity (ADCC) and/or complement-dependent cytotoxicity (CDC), or conjugated with a cytotoxic payload [e.g. antibody drug conjugate (ADC)], offers the potential of a significant therapeutic benefit for cancer patients with STn-expressing tumors. In addition, such antibodies would also allow for the development of a companion diagnostic to pre-select patients most likely to respond to therapy.

[00381] STn is often present on one or more of CSC surface antigens, and together they serve to promote the stemness and chemoresistance properties associated with CSCs. Thus, anti-STn antibodies offer a CSC-associated cancer targeting agent with the potential not only to directly kill CSCs via direct engagement and/or ADCC, but also offer a unique opportunity to bind to a wide array of cell-surface proteins and interfere with their associated functions essential for CSC viability, self-renewal, and replication.

[00382] As discuss herein, the rationale and advantages of targeting STn on CSCs may include: (1) many tumor-specific truncated glycoproteins carry STn in cancer; (2) STn is a unique glycan target expressed preferentially on CD44, MUC1, and potentially other important cell-surface markers, on both CSCs and mature tumor cells, irrespective of

proliferation status, allowing for targeting of both of these tumor components by a single therapeutic agent.; (3) STn is also a component of CA-125, a biomarker of ovarian cancer and others; (4) STn is a component of the ovarian CSC marker CD44. Therefore, the use of pan-STn murine mAbs, targeting an epitope that encompasses both the Neu5Ac and Neu5Gc forms of sialic acid linked to Tn, will bind to and kill or impair the function of CSCs and, by virtue of the common epitope, non-CSC tumor cells.

[00383] In some embodiments, the present invention provides new anti-pan STn mAb(s) for specific elimination of human CSCs as well as mature tumor cells. In one aspect of the present invention, the anti-STn antibody will target the validated STn glycan itself – not a particular glycopeptide or carrier protein, which should offer the broad potential of binding to CD44, MUC1, or other STn-glycosylated markers on both CSC and non-CSC tumor populations.

[00384] Given the exceptional specificity in targeting tumor-associated STn, the present invention may spare normal tissues, including normal adult stem cells, thereby allowing for an excellent therapeutic window.

[00385] In accordance with the present invention, provided herein is a unique immunotherapeutic solution aimed at eradicating human neoplasias by eliminating both CSCs and mature cancer cells contained within the tumor compartment. The present invention provides therapies and methods specifically targeting tumors, which now include targeting CSCs, and hence expanding the therapeutic window by targeting associated tumor-specific carbohydrate moieties of these potential targets. The elimination is specifically conferred through targeting tumor associated cell-surface sialylated Tn antigen (STn) structures that are uniquely present in cancer tissue, including cancer stem cells

Ovarian CSCs

[00386] Ovarian cancer is the leading gynecological cancer effecting women in the U.S. During 2013. It is estimated that 22,240 women will be diagnosed with and 14,030 will die of this disease, making it the fifth leading cause of female-related cancer deaths and the most lethal gynecologic malignancy in the U.S. (Siegel et al., Cancer statistics, 2013. CA: a cancer journal for clinicians 63, 11-30). This high mortality can be ascribed to non-symptomatic onset, late-stage initial diagnosis, aggressiveness of this type of cancer, and a general lack of therapeutically targetable genetic changes. The current standard of care is tumor debulking followed by taxane and platinum based chemotherapy. While this initial treatment results in

~70% of patients achieving an initial complete clinical response, a majority of these patients will unfortunately relapse with chemoresistant disease (Foster et al., Cancer letters, 2013, 338, 147-157; and McCann et al., PloS one, 2011, 6, e28077). In part, recurrent disease has been attributable, as with other cancer types, to the presence of CSCs within the total tumor population. Indeed, ovarian CSCs have been identified and shown to be resistant to chemo- and radiotherapy (Burgos-Ojeda et al., Cancer letters, 2012, 322, 1-7). Thus, again as the case with other forms of cancer, eliminating CSCs along with mature cells in the tumor population offers the best hope to manage recurrent disease and ideally effect cures.

[00387] In some embodiments of the present invention, ovarian CSCs may be targeted for ovarian cancer treatment. Although CD133 is the most widely studied of putative ovarian CSC markers, it is recognized that CD44, a known carrier of STn as discussed above, is associated with ovarian cancer and is included in the set of markers that identify ovarian CSCs (Zhang et al., Cancer research, 2008, 68, 4311-4320; Foster et al., Cancer letters, 2013, 338, 147-157; and Zoller, Cancer, 2011, 11, 254-267). Further, STn is expressed on the well-known ovarian cancer biomarker CA-125 (MUC16), as well as on MUC1, where the levels of these STn-associated mucins in serum have been used recently as further differentiators of cancerous versus benign ovarian disease. Elevated serum levels of STn occur in ~50% of ovarian cancer patients and correlate with a lower 5-year survival rate (Kobayashi et al., Journal of clinical oncology: official journal of the American Society of Clinical Oncology, 1991, 9, 983-987; Kobayashi et al., Journal of clinical oncology: official journal of the American Society of Clinical Oncology, 1992, 10, 95-101; and Chen et al., Journal of proteome research, 2013, 12, 1408-1418). Finally, Vathipadiekal et al. in a study of differential gene expression between human primary ovarian carcinoma CSCs and non-CSC populations found that the expression of STn-generating sialyl transferase ST6GalNAc-I did not differ among cells from the two compartments.

[00388] In some embodiments, the present invention provides antibodies for targeting CSCs to prevent control or cure cancer related to CSCs. Such antibodies may include anti-STn antibodies, including, but not limited to any of those described (or derived from any of those described) in international application number PCT/US14/60079, the contents of which are herein incorporated by reference in their entirety. Further anti-STn antibodies may include antibody 3F1 (SBH Sciences, Natick, MA) or derivatives thereof, including recombinant antibodies with CDRs from 3F1 and/or humanized derivatives.

Combined cancer therapies

[00389] In some embodiments, compounds and compositions of the invention may be combined with one or more additional forms of cancer treatment. In some cases, such additional forms may include chemotherapeutic treatments.

[00390] As used herein, the term, “chemotherapy” refers to a form of treatment using chemical substances. Such chemical substances are referred to herein as “chemotherapeutic agents.” In the treatment of cancer, chemotherapeutic agents may comprise one or more anti-cancer drugs. In some embodiments, chemotherapeutic agents that may be used according to methods of the present invention or in combination with compounds or compositions of the invention may include, but are not limited to capecitabine, gemcitabine, ABRAXANE® (paclitaxel protein-bound particles for injectable suspension), docetaxel, fluorouracil (5-FU), oxaliplatin, cisplatin, carboplatin, irinotecan, topotecan, paclitaxel, leucovorin, doxorubicin, and combinations thereof.

Immune-related targets

[00391] In some embodiments, glycan-interacting antibodies of the invention may be immunomodulatory antibodies. As used herein, an immunomodulatory antibody is an antibody that enhances or suppresses one or more immune function or pathway.

[00392] Many bacterial glycans are known to comprise sialic acid. In some cases, such glycans allow bacteria to evade the innate immune system of hosts, including, but not limited to humans. In one example, bacterial glycans inhibit alternate complement pathway activation through factor H recognition. In another example, bacterial glycans mask underlying residues that may be antigenic. Some bacterial glycans participate in cell signaling events through activation of inhibitory sialic acid binding Ig-like lectins (Siglecs) that dampen the immune response to entities comprising certain sialylated moieties (Chen, X. et al., Advances in the biology and chemistry of sialic acids. ACS Chem Biol. 2010 Feb 19;5(2):163-76). In some embodiments, glycan-interacting antibodies of the present invention may be used to treat immune complications related to bacterial glycans.

[00393] Due to the foreign nature of Neu5Gc as described herein, some Neu5Gc glycans are immunogenic resulting in immune related destruction of cells and other entities where these glycans may be expressed. Such autoimmune destruction may be pathogenic. In some embodiments, glycan-interacting antibodies may be used to treat patients suffering from autoimmune disorders related to Neu5Gc glycans.

[00394] In some embodiments, immunomodulatory antibodies of the invention may be used to promote or suppress T cell-mediated immunity. Such antibodies may interact with one or more glycans present on T cells, T cell-related proteins and/or on one or more other cell types that interact with T cells. Immunomodulatory antibodies that enhance T cell mediated immunity may be used to stimulate T cell mediated targeting of cancer cells.

[00395] In some tumors, infiltration by tumor-associated macrophages (TAMs) may lead to immunosuppression promoting tumor cell viability and growth. This is thought to be due to immunosuppressive cell signaling that occurs through interactions between myeloid C-type lectin receptors (CLRs) present on TAMs and tumor-associated mucins (Allavena, P. et al., Clin Dev Immunol. 2010;2010:547179). In some embodiments, binding of immunomodulatory antibodies of the invention to one or more tumor-associated mucin or TACA prevents immunosuppressive cell signaling in TAMs.

Anti-viral applications

[00396] In some embodiments, glycan-interacting antibodies of the invention may target viruses. Viral coat proteins and viral envelopes often comprise glycans, referred to herein as viral surface glycans. Such glycans may be targets of glycan-interacting antibodies. In some embodiments, viral surface glycans comprise sialyl-STn. In a further embodiment, viral surface glycans comprise GcSTn. Viruses that may be targeted by glycan-interacting antibodies include, but are not limited to HIV, influenza, rhinovirus, varicella-zoster, rotavirus, herpes (e.g. types 1 and 2), hepatitis (e.g. types A, B, C, D and E), yellow fever and human papillomavirus.

Other therapeutic applications

[00397] In some embodiments, glycan-interacting antibodies of the invention may act to alter or control proteolytic events. In some embodiments, glycan-interacting antibodies of the present invention may be internalized into cells prior to binding to targets.

Veterinary applications

[00398] It is contemplated that glycan-interacting antibodies of the invention will find utility in the area of veterinary care including the care and treatment of non-human vertebrates. As described herein, the term “non-human vertebrate” includes all vertebrates with the exception of Homo sapiens, including wild and domesticated species such as companion animals and livestock. Non-human vertebrates include mammals, such as alpaca,

banteng, bison, camel, cat, cattle, deer, dog, donkey, gayal, goat, guinea pig, horse, llama, mule, pig, rabbit, reindeer, sheep water buffalo, and yak. Livestock includes domesticated animals raised in an agricultural setting to produce materials such as food, labor, and derived products such as fiber and chemicals. Generally, livestock includes all mammals, avians and fish having potential agricultural significance. In particular, four-legged slaughter animals include steers, heifers, cows, calves, bulls, cattle, swine and sheep.

Bioprocessing

[00399] In some embodiments of the invention are methods for producing biological products in host cells by contacting the cells with one or more glycan-interacting antibody (such as an antibody or fusion protein) capable of modulating gene expression, or altering levels and/or types of glycans produced wherein such modulation or alteration enhances production of biological products. According to the present invention, bioprocessing methods may be improved by using one or more of the glycan-interacting antibodies of the present invention. They may also be improved by supplementing, replacing or adding one or more glycan-interacting antibodies.

Diagnostics

[00400] In some embodiments, compounds and compositions of the invention may be used as diagnostics. In some cases, antibodies of the invention may be used to identify, label or stain cells, tissues, organs, etc. expressing target antigens. In further embodiments, antibodies of the invention may be used to identify STn present in tissue sections (i.e., histological tissue sections), including tissue known or suspected of having cancerous cells. Such methods of using antibodies of the invention may in some cases be used to identify cancerous cells or tumors in tissue sections. Tissue sections may be from any tissue or organ including, but not limited to breast, colon, pancreatic, ovarian, brain, liver, kidney, spleen, lung, skin, stomach, intestine, esophagus, or bone.

III. Pharmaceutical compositions

[00401] The pharmaceutical compositions described herein can be characterized by one or more of bioavailability, therapeutic window and/or volume of distribution.

Bioavailability

[00402] Glycan-interacting antibodies, when formulated into a composition with a delivery/formulation agent or vehicle as described herein, can exhibit an increase in

bioavailability as compared to a composition lacking a delivery agent as described herein. As used herein, the term “bioavailability” refers to the systemic availability of a given amount of glycan-interacting antibodies administered to a mammal. Bioavailability can be assessed by measuring the area under the curve (AUC) or the maximum serum or plasma concentration ($C_{\max}$) of the unchanged form of a compound following administration of the compound to a mammal. AUC is a determination of the area under the curve plotting the serum or plasma concentration of a compound along the ordinate (Y-axis) against time along the abscissa (X-axis). Generally, the AUC for a particular compound can be calculated using methods known to those of ordinary skill in the art and as described in G. S. Banker, *Modern Pharmaceutics, Drugs and the Pharmaceutical Sciences*, v. 72, Marcel Dekker, New York, Inc., 1996, herein incorporated by reference.

[00403] The $C_{\max}$ value is the maximum concentration of the compound achieved in the serum or plasma of a mammal following administration of the compound to the mammal. The $C_{\max}$ value of a particular compound can be measured using methods known to those of ordinary skill in the art. The phrases “increasing bioavailability” or “improving the pharmacokinetics,” as used herein mean that the systemic availability of a glycan-interacting antibody, measured as AUC, $C_{\max}$, or $C_{\min}$ in a mammal is greater, when co-administered with a delivery agent as described herein, than when such co-administration does not take place. In some embodiments, the bioavailability of the glycan-interacting antibody can increase by at least about 2%, at least about 5%, at least about 10%, at least about 15%, at least about 20%, at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 55%, at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, or about 100%.

Therapeutic window

Glycan-interacting antibodies, when formulated into a composition with a delivery agent as described herein, can exhibit an increase in the therapeutic window of the administered glycan-interacting antibody composition as compared to the therapeutic window of the administered glycan-interacting antibody composition lacking a delivery agent as described herein. As used herein “therapeutic window” refers to the range of plasma concentrations, or the range of levels of therapeutically active substance at the site of action, with a high probability of eliciting a therapeutic effect. In some embodiments, the therapeutic window of

the glycan-interacting antibody when co-administered with a delivery agent as described herein can increase by at least about 2%, at least about 5%, at least about 10%, at least about 15%, at least about 20%, at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 55%, at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, or about 100%.

Volume of distribution

Glycan-interacting antibodies, when formulated into a composition with a delivery agent as described herein, can exhibit an improved volume of distribution (V_{dist}), e.g., reduced or targeted, relative to a composition lacking a delivery agent as described herein. The volume of distribution (V_{dist}) relates the amount of the drug in the body to the concentration of the drug in the blood or plasma. As used herein, the term “volume of distribution” refers to the fluid volume that would be required to contain the total amount of the drug in the body at the same concentration as in the blood or plasma: V_{dist} equals the amount of drug in the body/concentration of drug in blood or plasma. For example, for a 10 mg dose and a plasma concentration of 10 mg/L, the volume of distribution would be 1 liter. The volume of distribution reflects the extent to which the drug is present in the extravascular tissue. A large volume of distribution reflects the tendency of a compound to bind to the tissue components compared with plasma protein binding. In a clinical setting, V_{dist} can be used to determine a loading dose to achieve a steady state concentration. In some embodiments, the volume of distribution of the glycan-interacting antibody when co-administered with a delivery agent as described herein can decrease at least about 2%, at least about 5%, at least about 10%, at least about 15%, at least about 20%, at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 55%, at least about 60%, at least about 65%, at least about 70%.

[00404] In some embodiments, glycan-interacting antibodies comprise compositions and/or complexes in combination with one or more pharmaceutically acceptable excipients. Pharmaceutical compositions may optionally comprise one or more additional active substances, e.g. therapeutically and/or prophylactically active substances. General considerations in the formulation and/or manufacture of pharmaceutical agents may be found, for example, in *Remington: The Science and Practice of Pharmacy* 21st ed., Lippincott Williams & Wilkins, 2005 (incorporated herein by reference).

[00405] In some embodiments, compositions are administered to humans, human patients or subjects. For the purposes of the present disclosure, the phrase “active ingredient” generally refers to glycan-interacting antibodies to be delivered as described herein.

[00406] Although the descriptions of pharmaceutical compositions provided herein are principally directed to pharmaceutical compositions which are suitable for administration to humans, it will be understood by the skilled artisan that such compositions are generally suitable for administration to any other animal, *e.g.*, to non-human animals, *e.g.* non-human mammals. Modification of pharmaceutical compositions suitable for administration to humans in order to render the compositions suitable for administration to various animals is well understood, and the ordinarily skilled veterinary pharmacologist can design and/or perform such modification with merely ordinary, if any, experimentation. Subjects to which administration of the pharmaceutical compositions is contemplated include, but are not limited to, humans and/or other primates; mammals, including commercially relevant mammals such as cattle, pigs, horses, sheep, cats, dogs, mice, and/or rats; and/or birds, including commercially relevant birds such as poultry, chickens, ducks, geese, and/or turkeys.

[00407] Formulations of the pharmaceutical compositions described herein may be prepared by any method known or hereafter developed in the art of pharmacology. In general, such preparatory methods include the step of bringing the active ingredient into association with an excipient and/or one or more other accessory ingredients, and then, if necessary and/or desirable, dividing, shaping and/or packaging the product into a desired single- or multi-dose unit.

[00408] A pharmaceutical composition in accordance with the invention may be prepared, packaged, and/or sold in bulk, as a single unit dose, and/or as a plurality of single unit doses. As used herein, a “unit dose” is discrete amount of the pharmaceutical composition comprising a predetermined amount of the active ingredient. The amount of the active ingredient is generally equal to the dosage of the active ingredient which would be administered to a subject and/or a convenient fraction of such a dosage such as, for example, one-half or one-third of such a dosage.

[00409] Relative amounts of the active ingredient, the pharmaceutically acceptable excipient, and/or any additional ingredients in a pharmaceutical composition in accordance with the invention will vary, depending upon the identity, size, and/or condition of the subject treated and further depending upon the route by which the composition is to be administered. By way of example, the composition may comprise between 0.1% and 100%, *e.g.*, between .5

and 50%, between 1-30%, between 5-80%, or at least 80% (w/w) active ingredient. In one embodiment, active ingredients are antibodies directed toward cancer cells.

Formulation

[00410] Glycan-interacting antibodies of the invention can be formulated using one or more excipients to: (1) increase stability; (2) increase cell permeability; (3) permit the sustained or delayed release (e.g., from a formulation of the glycan-interacting antibody); and/or (4) alter the biodistribution (e.g., target the glycan-interacting antibody to specific tissues or cell types). In addition to traditional excipients such as any and all solvents, dispersion media, diluents, or other liquid vehicles, dispersion or suspension aids, surface active agents, isotonic agents, thickening or emulsifying agents, preservatives, formulations of the present invention can include, without limitation, liposomes, lipid nanoparticles, polymers, lipoplexes, core-shell nanoparticles, peptides, proteins, cells transfected with the glycan-interacting antibodies (e.g., for transplantation into a subject) and combinations thereof.

Excipients

[00411] As used herein, the term “excipient” refers to any substance combined with a compound and/or composition of the invention before use. In some embodiments, excipients are inactive and used primarily as a carrier, diluent or vehicle for a compound and/or composition of the present invention. Various excipients for formulating pharmaceutical compositions and techniques for preparing the composition are known in the art (see Remington: The Science and Practice of Pharmacy, 21st Edition, A. R. Gennaro, Lippincott, Williams & Wilkins, Baltimore, MD, 2006; incorporated herein by reference).

[00412] The use of a conventional excipient medium is contemplated within the scope of the present disclosure, except insofar as any conventional excipient medium may be incompatible with a substance or its derivatives, such as by producing any undesirable biological effect or otherwise interacting in a deleterious manner with any other component(s) of the pharmaceutical composition.

[00413] Formulations of the pharmaceutical compositions described herein may be prepared by any method known or hereafter developed in the art of pharmacology. In general, such preparatory methods include the step of associating the active ingredient with an excipient and/or one or more other accessory ingredients.

[00414] A pharmaceutical composition in accordance with the present disclosure may be prepared, packaged, and/or sold in bulk, as a single unit dose, and/or as a plurality of single unit doses.

[00415] Relative amounts of the active ingredient, the pharmaceutically acceptable excipient, and/or any additional ingredients in a pharmaceutical composition in accordance with the present disclosure may vary, depending upon the identity, size, and/or condition of the subject being treated and further depending upon the route by which the composition is to be administered.

[00416] In some embodiments, a pharmaceutically acceptable excipient is at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% pure. In some embodiments, an excipient is approved for use in humans and for veterinary use. In some embodiments, an excipient is approved by United States Food and Drug Administration. In some embodiments, an excipient is pharmaceutical grade. In some embodiments, an excipient meets the standards of the United States Pharmacopoeia (USP), the European Pharmacopoeia (EP), the British Pharmacopoeia, and/or the International Pharmacopoeia.

[00417] Pharmaceutically acceptable excipients used in the manufacture of pharmaceutical compositions include, but are not limited to, inert diluents, dispersing and/or granulating agents, surface active agents and/or emulsifiers, disintegrating agents, binding agents, preservatives, buffering agents, lubricating agents, and/or oils. Such excipients may optionally be included in pharmaceutical compositions.

[00418] Exemplary diluents include, but are not limited to, calcium carbonate, sodium carbonate, calcium phosphate, dicalcium phosphate, calcium sulfate, calcium hydrogen phosphate, sodium phosphate lactose, sucrose, cellulose, microcrystalline cellulose, kaolin, mannitol, sorbitol, inositol, sodium chloride, dry starch, cornstarch, powdered sugar, *etc.*, and/or combinations thereof.

[00419] Exemplary granulating and/or dispersing agents include, but are not limited to, potato starch, corn starch, tapioca starch, sodium starch glycolate, clays, alginic acid, guar gum, citrus pulp, agar, bentonite, cellulose and wood products, natural sponge, cation-exchange resins, calcium carbonate, silicates, sodium carbonate, cross-linked poly(vinyl-pyrrolidone) (crospovidone), sodium carboxymethyl starch (sodium starch glycolate), carboxymethyl cellulose, cross-linked sodium carboxymethyl cellulose (croscarmellose), methylcellulose, pregelatinized starch (starch 1500), microcrystalline starch, water insoluble

starch, calcium carboxymethyl cellulose, magnesium aluminum silicate (VEEGUM[®]), sodium lauryl sulfate, quaternary ammonium compounds, *etc.*, and/or combinations thereof.

[00420] Exemplary surface active agents and/or emulsifiers include, but are not limited to, natural emulsifiers (e.g. acacia, agar, alginic acid, sodium alginate, tragacanth, chondrux, cholesterol, xanthan, pectin, gelatin, egg yolk, casein, wool fat, cholesterol, wax, and lecithin), colloidal clays (e.g. bentonite [aluminum silicate] and VEEGUM[®] [magnesium aluminum silicate]), long chain amino acid derivatives, high molecular weight alcohols (e.g. stearyl alcohol, cetyl alcohol, oleyl alcohol, triacetin monostearate, ethylene glycol distearate, glyceryl monostearate, and propylene glycol monostearate, polyvinyl alcohol), carbomers (e.g. carboxy polymethylene, polyacrylic acid, acrylic acid polymer, and carboxyvinyl polymer), carrageenan, cellulosic derivatives (e.g. carboxymethylcellulose sodium, powdered cellulose, hydroxymethyl cellulose, hydroxypropyl cellulose, hydroxypropyl methylcellulose, methylcellulose), sorbitan fatty acid esters (e.g. polyoxyethylene sorbitan monolaurate [TWEEN[®]20], polyoxyethylene sorbitan [TWEEN[®]60], polyoxyethylene sorbitan monooleate [TWEEN[®]80], sorbitan monopalmitate [SPAN[®]40], sorbitan monostearate [Span[®]60], sorbitan tristearate [Span[®]65], glyceryl monooleate, sorbitan monooleate [SPAN[®]80]), polyoxyethylene esters (e.g. polyoxyethylene monostearate [MYRJ[®]45], polyoxyethylene hydrogenated castor oil, polyethoxylated castor oil, polyoxymethylene stearate, and SOLUTOL[®]), sucrose fatty acid esters, polyethylene glycol fatty acid esters (e.g. CREMOPHOR[®]), polyoxyethylene ethers, (e.g. polyoxyethylene lauryl ether [BRIJ[®]30]), poly(vinyl-pyrrolidone), diethylene glycol monolaurate, triethanolamine oleate, sodium oleate, potassium oleate, ethyl oleate, oleic acid, ethyl laurate, sodium lauryl sulfate, PLUORINC[®]F 68, POLOXAMER[®]188, cetrimonium bromide, cetylpyridinium chloride, benzalkonium chloride, docusate sodium, *etc.* and/or combinations thereof.

[00421] Exemplary binding agents include, but are not limited to, starch (*e.g.* cornstarch and starch paste); gelatin; sugars (*e.g.* sucrose, glucose, dextrose, dextrin, molasses, lactose, lactitol, mannitol,); natural and synthetic gums (*e.g.* acacia, sodium alginate, extract of Irish moss, panwar gum, ghatti gum, mucilage of isapol husks, carboxymethylcellulose, methylcellulose, ethylcellulose, hydroxyethylcellulose, hydroxypropyl cellulose, hydroxypropyl methylcellulose, microcrystalline cellulose, cellulose acetate, poly(vinyl-pyrrolidone), magnesium aluminum silicate (VEEGUM[®]), and larch arabogalactan); alginates; polyethylene oxide; polyethylene glycol; inorganic calcium salts; silicic acid; polymethacrylates; waxes; water; alcohol; *etc.*; and combinations thereof.

[00422] Exemplary preservatives may include, but are not limited to, antioxidants, chelating agents, antimicrobial preservatives, antifungal preservatives, alcohol preservatives, acidic preservatives, and/or other preservatives. Exemplary antioxidants include, but are not limited to, alpha tocopherol, ascorbic acid, ascorbyl palmitate, butylated hydroxyanisole, butylated hydroxytoluene, monothioglycerol, potassium metabisulfite, propionic acid, propyl gallate, sodium ascorbate, sodium bisulfite, sodium metabisulfite, and/or sodium sulfite. Exemplary chelating agents include ethylenediaminetetraacetic acid (EDTA), citric acid monohydrate, disodium edetate, dipotassium edetate, edetic acid, fumaric acid, malic acid, phosphoric acid, sodium edetate, tartaric acid, and/or trisodium edetate. Exemplary antimicrobial preservatives include, but are not limited to, benzalkonium chloride, benzethonium chloride, benzyl alcohol, bronopol, cetrimide, cetylpyridinium chloride, chlorhexidine, chlorobutanol, chlorocresol, chloroxylenol, cresol, ethyl alcohol, glycerin, hexetidine, imidurea, phenol, phenoxyethanol, phenylethyl alcohol, phenylmercuric nitrate, propylene glycol, and/or thimerosal. Exemplary antifungal preservatives include, but are not limited to, butyl paraben, methyl paraben, ethyl paraben, propyl paraben, benzoic acid, hydroxybenzoic acid, potassium benzoate, potassium sorbate, sodium benzoate, sodium propionate, and/or sorbic acid. Exemplary alcohol preservatives include, but are not limited to, ethanol, polyethylene glycol, phenol, phenolic compounds, bisphenol, chlorobutanol, hydroxybenzoate, and/or phenylethyl alcohol. Exemplary acidic preservatives include, but are not limited to, vitamin A, vitamin C, vitamin E, beta-carotene, citric acid, acetic acid, dehydroacetic acid, ascorbic acid, sorbic acid, and/or phytic acid. Other preservatives include, but are not limited to, tocopherol, tocopherol acetate, deteroxime mesylate, cetrimide, butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), ethylenediamine, sodium lauryl sulfate (SLS), sodium lauryl ether sulfate (SLES), sodium bisulfite, sodium metabisulfite, potassium sulfite, potassium metabisulfite, GLYDANT PLUS[®], PHENONIP[®], methylparaben, GERMALL[®] 115, GERMABEN[®] II, NEOLONE[™], KATHON[™], and/or EUXYL[®].

[00423] Exemplary buffering agents include, but are not limited to, citrate buffer solutions, acetate buffer solutions, phosphate buffer solutions, ammonium chloride, calcium carbonate, calcium chloride, calcium citrate, calcium gluconate, calcium gluceptate, calcium gluconate, D-gluconic acid, calcium glycerophosphate, calcium lactate, propanoic acid, calcium levulinate, pentanoic acid, dibasic calcium phosphate, phosphoric acid, tribasic calcium phosphate, calcium hydroxide phosphate, potassium acetate, potassium chloride, potassium

gluconate, potassium mixtures, dibasic potassium phosphate, monobasic potassium phosphate, potassium phosphate mixtures, sodium acetate, sodium bicarbonate, sodium chloride, sodium citrate, sodium lactate, dibasic sodium phosphate, monobasic sodium phosphate, sodium phosphate mixtures, tromethamine, magnesium hydroxide, aluminum hydroxide, alginic acid, pyrogen-free water, isotonic saline, Ringer's solution, ethyl alcohol, *etc.*, and/or combinations thereof.

[00424] Exemplary lubricating agents include, but are not limited to, magnesium stearate, calcium stearate, stearic acid, silica, talc, malt, glyceryl behenate, hydrogenated vegetable oils, polyethylene glycol, sodium benzoate, sodium acetate, sodium chloride, leucine, magnesium lauryl sulfate, sodium lauryl sulfate, *etc.*, and combinations thereof.

[00425] Exemplary oils include, but are not limited to, almond, apricot kernel, avocado, babassu, bergamot, black current seed, borage, cade, camomile, canola, caraway, carnauba, castor, cinnamon, cocoa butter, coconut, cod liver, coffee, corn, cotton seed, emu, eucalyptus, evening primrose, fish, flaxseed, geraniol, gourd, grape seed, hazel nut, hyssop, isopropyl myristate, jojoba, kukui nut, lavandin, lavender, lemon, litsea cubeba, macadamia nut, mallow, mango seed, meadowfoam seed, mink, nutmeg, olive, orange, orange roughy, palm, palm kernel, peach kernel, peanut, poppy seed, pumpkin seed, rapeseed, rice bran, rosemary, safflower, sandalwood, sasquana, savoury, sea buckthorn, sesame, shea butter, silicone, soybean, sunflower, tea tree, thistle, tsubaki, vetiver, walnut, and wheat germ oils. Exemplary oils include, but are not limited to, butyl stearate, caprylic triglyceride, capric triglyceride, cyclomethicone, diethyl sebacate, dimethicone 360, isopropyl myristate, mineral oil, octyldodecanol, oleyl alcohol, silicone oil, and/or combinations thereof.

[00426] Excipients such as cocoa butter and suppository waxes, coloring agents, coating agents, sweetening, flavoring, and/or perfuming agents can be present in the composition, according to the judgment of the formulator.

Vehicles

Liposomes, lipoplexes and lipid nanoparticles

[00427] Glycan-interacting antibodies of the present invention may be formulated using one or more liposomes, lipoplexes, or lipid nanoparticles. In one embodiment, pharmaceutical compositions comprising glycan-interacting antibodies further comprise liposomes. Liposomes are artificially-prepared vesicles which may primarily comprise one or more lipid bilayers and may be used as a delivery vehicle for the administration of nutrients

and pharmaceutical formulations. Liposomes can be of different sizes such as, but not limited to, a multilamellar vesicle (MLV) which may be hundreds of nanometers in diameter and may contain a series of concentric bilayers separated by narrow aqueous compartments, a small unicellular vesicle (SUV) which may be smaller than 50 nm in diameter, and a large unilamellar vesicle (LUV) which may be between 50 and 500 nm in diameter. Liposome design may include, but is not limited to, opsonins or ligands in order to improve the attachment of liposomes to unhealthy tissue or to activate events such as, but not limited to, endocytosis. Liposomes may contain a low or a high pH in order to improve the delivery of the pharmaceutical formulations.

[00428] The formation of liposomes may depend on the physicochemical characteristics such as, but not limited to, the pharmaceutical formulation entrapped and the liposomal ingredients, the nature of the medium in which the lipid vesicles are dispersed, the effective concentration of the entrapped substance and its potential toxicity, any additional processes involved during the application and/or delivery of the vesicles, the optimization size, polydispersity and the shelf-life of the vesicles for the intended application, and the batch-to-batch reproducibility and possibility of large-scale production of safe and efficient liposomal products.

[00429] In one embodiment such formulations may also be constructed or compositions altered such that they passively or actively are directed to different cell types *in vivo*.

[00430] Formulations can also be selectively targeted through expression of different ligands on their surface as exemplified by, but not limited by, folate, transferrin, N-acetylgalactosamine (GalNAc), and antibody targeted approaches.

[00431] Liposomes, lipoplexes, or lipid nanoparticles may be used to improve the efficacy of glycan-interacting antibody function as these formulations may be able to increase cell transfection with glycan-interacting antibodies. The liposomes, lipoplexes, or lipid nanoparticles may also be used to increase the stability of glycan-interacting antibodies.

[00432] Liposomes that are specifically formulated for antibody cargo are prepared according to techniques known in the art, such as described by Eppstein et al. (Eppstein, D.A. et al., *Biological activity of liposome-encapsulated murine interferon gamma is mediated by a cell membrane receptor*. Proc Natl Acad Sci U S A. 1985 Jun;82(11):3688-92); Hwang et al. (Hwang, K.J. et al., *Hepatic uptake and degradation of unilamellar sphingomyelin/cholesterol liposomes: a kinetic study*. Proc Natl Acad Sci U S A. 1980

Jul;77(7):4030-4); US 4,485,045 and US 4,544,545. Production of liposomes with sustained circulation time is also described in US 5,013,556.

[00433] Liposomes comprising glycan-interacting antibodies of the present invention may be generated using reverse phase evaporation utilizing lipids such as phosphatidylcholine, cholesterol as well as phosphatidylethanolamine that has been polyethylene glycol-derivatized. Filters with defined pore size are used to extrude liposomes of the desired diameter. In another embodiment, glycan-interacting antibodies of the present invention can be conjugated to the external surface of liposomes by disulfide interchange reaction as is described by Martin et al. (Martin, F.J. et al., *Irreversible coupling of immunoglobulin fragments to preformed vesicles. An improved method for liposome targeting*. J Biol Chem. 1982 Jan 10;257(1):286-8).

Polymers and nanoparticles

[00434] Glycan-interacting antibodies of the invention can be formulated using natural and/or synthetic polymers. Non-limiting examples of polymers which may be used for delivery include, but are not limited to DMRI/DOPE, poloxamer, chitosan, cyclodextrin, and poly(lactic-co-glycolic acid) (PLGA) polymers. These may be biodegradable.

[00435] The polymer formulation can permit the sustained or delayed release of glycan-interacting antibodies (e.g., following intramuscular or subcutaneous injection). The altered release profile for glycan-interacting antibodies can result in, for example, release of the glycan-interacting antibodies over an extended period of time. The polymer formulation may also be used to increase the stability of glycan-interacting antibodies.

[00436] Polymer formulations can also be selectively targeted through expression of different ligands as exemplified by, but not limited by, folate, transferrin, and N-acetylgalactosamine (GalNAc) (Benoit et al., *Biomacromolecules*. 2011 12:2708-2714; Rozema et al., *Proc Natl Acad Sci U S A*. 2007 104:12982-12887; Davis, *Mol Pharm*. 2009 6:659-668; Davis, *Nature* 2010 464:1067-1070; herein incorporated by reference in its entirety).

[00437] Glycan-interacting antibodies of the invention can also be formulated as nanoparticles using a combination of polymers, lipids, and/or other biodegradable agents, such as, but not limited to, calcium phosphate. Components may be combined in a core-shell, hybrid, and/or layer-by-layer architecture, to allow for fine-tuning of the nanoparticle so

delivery of glycan-interacting antibodies may be enhanced. For glycan-interacting antibodies, systems based on poly(2-(methacryloyloxy)ethyl phosphorylcholine)-block-(2-(diisopropylamino)ethyl methacrylate), (PMPC-PDPA), a pH sensitive diblock copolymer that self-assembles to form nanometer-sized vesicles, also known as polymersomes, at physiological pH may be used. These polymersomes have been shown to successfully deliver relatively high antibody payloads within live cells. (Massignani, et al, Cellular delivery of antibodies: effective targeted subcellular imaging and new therapeutic tool. Nature Proceedings, May, 2010).

[00438] In one embodiment, a PEG-charge-conversional polymer (Pitella et al., Biomaterials. 2011 32:3106-3114) may be used to form a nanoparticle to deliver glycan-interacting antibodies of the present invention. The PEG-charge-conversional polymer may improve upon the PEG-polyanion block copolymers by being cleaved into a polycation at acidic pH, thus enhancing endosomal escape.

[00439] The use of core-shell nanoparticles has additionally focused on a high-throughput approach to synthesize cationic cross-linked nanogel cores and various shells (Siegwart et al., Proc Natl Acad Sci U S A. 2011 108:12996-13001). The complexation, delivery, and internalization of the polymeric nanoparticles can be precisely controlled by altering the chemical composition in both the core and shell components of the nanoparticle.

[00440] In one embodiment, matrices of poly(ethylene-co-vinyl acetate), are used to deliver glycan-interacting antibodies of the invention. Such matrices are described in Nature Biotechnology 10, 1446 - 1449 (1992).

Antibody formulations

[00441] Glycan-interacting antibodies of the invention may be formulated for intravenous administration or extravascular administration (Daugherty, et al., *Formulation and delivery issues for monoclonal antibody therapeutics*. Adv Drug Deliv Rev. 2006 Aug 7;58(5-6):686-706, US patent publication number 2011/0135570, all of which are incorporated herein in their entirety). Extravascular administration routes may include, but are not limited to subcutaneous administration, intraperitoneal administration, intracerebral administration, intraocular administration, intralesional administration, topical administration and intramuscular administration.

[00442] Antibody structures may be modified to improve their effectiveness as therapeutics. Improvements may include, but are not limited to improved thermodynamic

stability, reduced Fc receptor binding properties and improved folding efficiency.

Modifications may include, but are not limited to amino acid substitutions, glycosylation, palmitoylation and protein conjugation.

[00443] Glycan-interacting antibodies may be formulated with antioxidants to reduce antibody oxidation. Glycan-interacting antibodies may also be formulated with additives to reduce protein aggregation. Such additives may include, but are not limited to albumin, amino acids, sugars, urea, guanidinium chloride, polyalcohols, polymers (such as polyethylene glycol and dextrans), surfactants (including, but not limited to polysorbate 20 and polysorbate 80) or even other antibodies.

[00444] Glycan-interacting antibodies of the present invention may be formulated to reduce the impact of water on antibody structure and function. Antibody preparations in such formulations may be lyophilized. Formulations subject to lyophilization may include carbohydrates or polyol compounds to protect and stabilize antibody structure. Such compounds include, but are not limited to sucrose, trehalose and mannitol.

[00445] Glycan-interacting antibodies of the present invention may be formulated with polymers. In one embodiment, polymer formulations may contain hydrophobic polymers. Such polymers may be microspheres formulated with polylactide-co-glycolide through a solid-in-oil-in-water encapsulation method. Microspheres comprising ethylene-vinyl acetate copolymer are also contemplated for antibody delivery and may be used to extend the time course of antibody release at the site of delivery. In another embodiment, polymers may be aqueous gels. Such gels may, for example, comprise carboxymethylcellulose. Aqueous gels may also comprise hyaluronic acid hydrogel. Antibodies may be covalently linked to such gels through a hydrazone linkage that allows for sustained delivery in tissues, including but not limited to the tissues of the central nervous system.

Peptide and protein formulations

[00446] Glycan-interacting antibodies of the invention may be formulated with peptides and/or proteins. In one embodiment, peptides such as, but not limited to, cell penetrating peptides and proteins and peptides that enable intracellular delivery may be used to deliver pharmaceutical formulations. A non-limiting example of a cell penetrating peptide which may be used with the pharmaceutical formulations of the present invention includes a cell-penetrating peptide sequence attached to polycations that facilitates delivery to the intracellular space, e.g., HIV-derived TAT peptide, penetratins, transportans, or hCT derived

cell-penetrating peptides (see, e.g., Caron et al., Mol. Ther. 3(3):310-8 (2001); Langel, Cell-Penetrating Peptides: Processes and Applications (CRC Press, Boca Raton FL, 2002); El-Andaloussi et al., Curr. Pharm. Des. 11(28):3597-611 (2003); and Deshayes et al., Cell. Mol. Life Sci. 62(16):1839-49 (2005), all of which are incorporated herein by reference). The compositions can also be formulated to include a cell penetrating agent, e.g., liposomes, which enhance delivery of the compositions to the intracellular space. Glycan-interacting antibodies of the invention may be complexed to peptides and/or proteins such as, but not limited to, peptides and/or proteins from Aileron Therapeutics (Cambridge, MA) and Permeon Biologics (Cambridge, MA) in order to enable intracellular delivery (Cronican et al., ACS Chem. Biol. 2010 5:747-752; McNaughton et al., Proc. Natl. Acad. Sci. USA 2009 106:6111-6116; Sawyer, Chem Biol Drug Des. 2009 73:3-6; Verdine and Hilinski, Methods Enzymol. 2012;503:3-33; all of which are herein incorporated by reference in their entirety).

[00447] In one embodiment, the cell-penetrating polypeptide may comprise a first domain and a second domain. The first domain may comprise a supercharged polypeptide. The second domain may comprise a protein-binding partner. As used herein, “protein-binding partner” includes, but are not limited to, antibodies and functional fragments thereof, scaffold proteins, or peptides. The cell-penetrating polypeptide may further comprise an intracellular binding partner for the protein-binding partner. The cell-penetrating polypeptide may be capable of being secreted from a cell where glycan-interacting antibodies may be introduced.

[00448] In formulations of the present invention, peptides or proteins may be incorporated to increase cell transfection by glycan-interacting antibodies or alter the biodistribution of glycan-interacting antibodies (e.g., by targeting specific tissues or cell types).

Cell formulations

[00449] Cell-based formulations of glycan-interacting antibody compositions of the invention may be used to ensure cell transfection (e.g., in the cellular carrier) or alter the biodistribution of the compositions (e.g., by targeting the cell carrier to specific tissues or cell types).

Cell transfer methods

[00450] A variety of methods are known in the art and are suitable for introduction of nucleic acids or proteins, such as glycan-interacting antibodies, into a cell, including viral and non-viral mediated techniques. Examples of typical non-viral mediated techniques include, but are not limited to, electroporation, calcium phosphate mediated transfer, nucleofection,

sonoporation, heat shock, magnetofection, liposome mediated transfer, microinjection, microprojectile mediated transfer (nanoparticles), cationic polymer mediated transfer (DEAE-dextran, polyethylenimine, polyethylene glycol (PEG) and the like) or cell fusion.

[00451] The technique of sonoporation, or cellular sonication, is the use of sound (e.g., ultrasonic frequencies) for modifying the permeability of the cell plasma membrane.

Sonoporation methods are known to those in the art and are used to deliver nucleic acids *in vivo* (Yoon and Park, Expert Opin Drug Deliv. 2010 7:321-330; Postema and Gilja, Curr Pharm Biotechnol. 2007 8:355-361; Newman and Bettinger, Gene Ther. 2007 14:465-475; all herein incorporated by reference in their entirety). Sonoporation methods are known in the art and are also taught for example as it relates to bacteria in US Patent Publication 20100196983 and as it relates to other cell types in, for example, US Patent Publication 20100009424, each of which are incorporated herein by reference in their entirety.

[00452] Electroporation techniques are also well known in the art and are used to deliver nucleic acids *in vivo* and clinically (Andre et al., Curr Gene Ther. 2010 10:267-280; Chiarella et al., Curr Gene Ther. 2010 10:281-286; Hojman, Curr Gene Ther. 2010 10:128-138; all herein incorporated by reference in their entirety). In one embodiment, glycan-interacting antibodies may be delivered by electroporation.

Administration and delivery

[00453] The compositions of the present invention may be administered by any of the standard methods or routes known in the art.

[00454] Glycan-interacting antibodies of the present invention may be administered by any route which results in a therapeutically effective outcome. These include, but are not limited to enteral, gastroenteral, epidural, oral, transdermal, epidural (peridural), intracerebral (into the cerebrum), intracerebroventricular (into the cerebral ventricles), epicutaneous (application onto the skin), intradermal, (into the skin itself), subcutaneous (under the skin), nasal administration (through the nose), intravenous (into a vein), intraarterial (into an artery), intramuscular (into a muscle), intracardiac (into the heart), intraosseous infusion (into the bone marrow), intrathecal (into the spinal canal), intraperitoneal, (infusion or injection into the peritoneum), intravesical infusion, intravitreal, (through the eye), intracavernous injection, (into the base of the penis), intravaginal administration, intrauterine, extra-amniotic administration, transdermal (diffusion through the intact skin for systemic distribution), transmucosal (diffusion through a mucous membrane), insufflation (snorting),

sublingual, sublabial, enema, eye drops (onto the conjunctiva), or in ear drops. In specific embodiments, compositions may be administered in a way which allows them cross the blood-brain barrier, vascular barrier, or other epithelial barrier. Non-limiting routes of administration for glycan-interacting antibodies of the present invention are described below.

Parenteral and injectable administration

[00455] Liquid dosage forms for oral and parenteral administration include, but are not limited to, pharmaceutically acceptable emulsions, microemulsions, solutions, suspensions, syrups, and/or elixirs. In addition to active ingredients, liquid dosage forms may comprise inert diluents commonly used in the art such as, for example, water or other solvents, solubilizing agents and emulsifiers such as ethyl alcohol, isopropyl alcohol, ethyl carbonate, ethyl acetate, benzyl alcohol, benzyl benzoate, propylene glycol, 1,3-butylene glycol, dimethylformamide, oils (in particular, cottonseed, groundnut, corn, germ, olive, castor, and sesame oils), glycerol, tetrahydrofurfuryl alcohol, polyethylene glycols and fatty acid esters of sorbitan, and mixtures thereof. Besides inert diluents, oral compositions can include adjuvants such as wetting agents, emulsifying and suspending agents, sweetening, flavoring, and/or perfuming agents. In certain embodiments for parenteral administration, compositions are mixed with solubilizing agents such as CREMOPHOR[®], alcohols, oils, modified oils, glycols, polysorbates, cyclodextrins, polymers, and/or combinations thereof. In other embodiments, surfactants are included such as hydroxypropylcellulose.

[00456] Injectable preparations, for example, sterile injectable aqueous or oleaginous suspensions may be formulated according to the known art using suitable dispersing agents, wetting agents, and/or suspending agents. Sterile injectable preparations may be sterile injectable solutions, suspensions, and/or emulsions in nontoxic parenterally acceptable diluents and/or solvents, for example, as a solution in 1,3-butanediol. Among the acceptable vehicles and solvents that may be employed are water, Ringer's solution, U.S.P., and isotonic sodium chloride solution. Sterile, fixed oils are conventionally employed as a solvent or suspending medium. For this purpose any bland fixed oil can be employed including synthetic mono- or diglycerides. Fatty acids such as oleic acid can be used in the preparation of injectables.

Injectable formulations can be sterilized, for example, by filtration through a bacterial-retaining filter, and/or by incorporating sterilizing agents in the form of sterile solid

compositions which can be dissolved or dispersed in sterile water or other sterile injectable medium prior to use.

[00457] In order to prolong the effect of an active ingredient, it is often desirable to slow the absorption of the active ingredient from subcutaneous or intramuscular injection. This may be accomplished by the use of a liquid suspension of crystalline or amorphous material with poor water solubility. The rate of absorption of the drug then depends upon its rate of dissolution which, in turn, may depend upon crystal size and crystalline form. Alternatively, delayed absorption of a parenterally administered drug form is accomplished by dissolving or suspending the drug in an oil vehicle. Injectable depot forms are made by forming microencapsulated matrices of the drug in biodegradable polymers such as polylactide-polyglycolide. Depending upon the ratio of drug to polymer and the nature of the particular polymer employed, the rate of drug release can be controlled. Examples of other biodegradable polymers include poly(orthoesters) and poly(anhydrides). Depot injectable formulations are prepared by entrapping the drug in liposomes or microemulsions which are compatible with body tissues.

Rectal and vaginal administration

[00458] Compositions for rectal or vaginal administration are typically suppositories which can be prepared by mixing compositions with suitable non-irritating excipients such as cocoa butter, polyethylene glycol or a suppository wax which are solid at ambient temperature but liquid at body temperature and therefore melt in the rectum or vaginal cavity and release the active ingredient.

Oral administration

[00459] Solid dosage forms for oral administration include capsules, tablets, pills, powders, and granules. In such solid dosage forms, an active ingredient is mixed with at least one inert, pharmaceutically acceptable excipient such as sodium citrate or dicalcium phosphate and/or fillers or extenders (e.g. starches, lactose, sucrose, glucose, mannitol, and silicic acid), binders (e.g. carboxymethylcellulose, alginates, gelatin, polyvinylpyrrolidinone, sucrose, and acacia), humectants (e.g. glycerol), disintegrating agents (e.g. agar, calcium carbonate, potato or tapioca starch, alginic acid, certain silicates, and sodium carbonate), solution retarding agents (e.g. paraffin), absorption accelerators (e.g. quaternary ammonium compounds), wetting agents (e.g. cetyl alcohol and glycerol monostearate), absorbents (e.g. kaolin and bentonite clay), and lubricants (e.g. talc, calcium stearate, magnesium stearate, solid

polyethylene glycols, sodium lauryl sulfate), and mixtures thereof. In the case of capsules, tablets and pills, the dosage form may comprise buffering agents.

Topical or transdermal administration

[00460] As described herein, compositions containing glycan-interacting antibodies of the invention may be formulated for administration topically. The skin may be an ideal target site for delivery as it is readily accessible. Gene expression may be restricted not only to the skin, potentially avoiding nonspecific toxicity, but also to specific layers and cell types within the skin.

[00461] The site of cutaneous expression of the delivered compositions will depend on the route of nucleic acid delivery. Three routes are commonly considered to deliver glycan-interacting antibodies to the skin: (i) topical application (*e.g.* for local/regional treatment and/or cosmetic applications); (ii) intradermal injection (*e.g.* for local/regional treatment and/or cosmetic applications); and (iii) systemic delivery (*e.g.* for treatment of dermatologic diseases that affect both cutaneous and extracutaneous regions). Glycan-interacting antibodies can be delivered to the skin by several different approaches known in the art.

[00462] In one embodiment, the invention provides for a variety of dressings (*e.g.*, wound dressings) or bandages (*e.g.*, adhesive bandages) for conveniently and/or effectively carrying out methods of the present invention. Typically dressing or bandages may comprise sufficient amounts of pharmaceutical compositions and/or glycan-interacting antibodies described herein to allow a user to perform multiple treatments of a subject(s).

[00463] In one embodiment, the invention provides for compositions comprising glycan-interacting antibodies to be delivered in more than one injection.

[00464] Dosage forms for topical and/or transdermal administration of a composition may include ointments, pastes, creams, lotions, gels, powders, solutions, sprays, inhalants and/or patches. Generally, an active ingredient is admixed under sterile conditions with a pharmaceutically acceptable excipient and/or any needed preservatives and/or buffers as may be required.

[00465] Additionally, the present invention contemplates the use of transdermal patches, which often have the added advantage of providing controlled delivery of a compound to the body. Such dosage forms may be prepared, for example, by dissolving and/or dispensing the compound in the proper medium. Alternatively or additionally, rate may be controlled by

either providing a rate controlling membrane and/or by dispersing the compound in a polymer matrix and/or gel.

[00466] Formulations suitable for topical administration include, but are not limited to, liquid and/or semi liquid preparations such as liniments, lotions, oil in water and/or water in oil emulsions such as creams, ointments and/or pastes, and/or solutions and/or suspensions.

[00467] Topically-administrable formulations may, for example, comprise from about 1% to about 10% (w/w) active ingredient, although the concentration of active ingredient may be as high as the solubility limit of the active ingredient in the solvent. Formulations for topical administration may further comprise one or more of the additional ingredients described herein.

Depot administration

[00468] As described herein, in some embodiments, compositions of the present invention are formulated in depots for extended release. Generally, a specific organ or tissue (a “target tissue”) is targeted for administration.

[00469] In some aspects of the invention, glycan-interacting antibodies are spatially retained within or proximal to a target tissue. Provided are methods of providing compositions to one or more target tissue of a mammalian subject by contacting the one or more target tissue (comprising one or more target cells) with compositions under conditions such that the compositions, in particular glycan-interacting antibody component(s) of the compositions, are substantially retained in the target tissue, meaning that at least 10, 20, 30, 40, 50, 60, 70, 80, 85, 90, 95, 96, 97, 98, 99, 99.9, 99.99 or greater than 99.99% of the composition is retained in the target tissue. Advantageously, retention is determined by measuring the level of glycan-interacting antibodies present in the compositions entering the target tissues and/or cells. For example, at least 1, 5, 10, 20, 30, 40, 50, 60, 70, 80, 85, 90, 95, 96, 97, 98, 99, 99.9, 99.99 or greater than 99.99% of glycan-interacting antibodies administered to the subject are present intracellularly at a period of time following administration. For example, intramuscular injection to a mammalian subject is performed using an aqueous composition comprising one or more glycan-interacting antibody and a transfection reagent, and retention of the composition is determined by measuring the level of glycan-interacting antibodies present in the muscle cells.

[00470] Certain aspects of the invention are directed to methods of providing compositions to target tissues of mammalian subjects, by contacting the target tissues (containing one or

more target cells) with compositions under conditions such that the compositions are substantially retained in the target tissue. Compositions contain an effective amount of glycan-interacting antibodies such that the effect of interest is produced in at least one target cell. Compositions generally contain cell penetration agents and a pharmaceutically acceptable carrier, although “naked” glycan-interacting antibodies (such as glycan-interacting antibodies without cell penetration agents or other agents) are also contemplated.

[00471] In some embodiments, compositions include a plurality of different glycan-interacting antibodies, where one or more than one of the glycan-interacting antibodies targets a glycan of interest. Optionally, compositions also contain cell penetration agents to assist in the intracellular delivery of compositions. A determination is made of the composition dose required to target glycans of interest in a substantial percentage of cells contained within a predetermined volume of the target tissue (generally, without targeting glycans in tissue adjacent to the predetermined volume, or distally to target tissues). Subsequent to this determination, the determined dose may be introduced directly into the tissue of the mammalian subject.

[00472] In one embodiment, the invention provides for glycan-interacting antibodies to be delivered in more than one injection or by split dose injections.

Pulmonary administration

[00473] Pharmaceutical compositions may be prepared, packaged, and/or sold in formulations suitable for pulmonary administration via the buccal cavity. Such formulations may comprise dry particles further comprising active ingredients and having a diameter in the range from about 0.5 nm to about 7 nm or from about 1 nm to about 6 nm. Such compositions are suitably in the form of dry powders for administration using a device comprising a dry powder reservoir to which a stream of propellant may be directed to disperse the powder and/or using a self-propelling solvent/powder dispensing container such as a device comprising the active ingredient dissolved and/or suspended in a low-boiling propellant in a sealed container. Such powders comprise particles wherein at least 98% of the particles by weight have a diameter greater than 0.5 nm and at least 95% of the particles by number have a diameter less than 7 nm. Alternatively, at least 95% of the particles by weight have a diameter greater than 1 nm and at least 90% of the particles by number have a diameter less than 6 nm. Dry powder compositions may include a solid fine powder diluent such as sugar and are conveniently provided in a unit dose form.

[00474] Low boiling propellants generally include liquid propellants having a boiling point of below 65 °F at atmospheric pressure. Generally the propellant may constitute 50% to 99.9% (w/w) of the composition, and active ingredient may constitute 0.1% to 20% (w/w) of the composition. A propellant may further comprise additional ingredients such as a liquid non-ionic and/or solid anionic surfactant and/or a solid diluent (which may have a particle size of the same order as particles comprising the active ingredient).

[00475] Pharmaceutical compositions formulated for pulmonary delivery may provide an active ingredient in the form of droplets of a solution and/or suspension. Such formulations may be prepared, packaged, and/or sold as aqueous and/or dilute alcoholic solutions and/or suspensions, optionally sterile, comprising active ingredient, and may conveniently be administered using any nebulization and/or atomization device. Such formulations may further comprise one or more additional ingredients including, but not limited to, a flavoring agent such as saccharin sodium, a volatile oil, a buffering agent, a surface active agent, and/or a preservative such as methylhydroxybenzoate. Droplets provided by this route of administration may have an average diameter in the range from about 0.1 nm to about 200 nm.

Intranasal, nasal and buccal administration

[00476] Formulations described herein as being useful for pulmonary delivery are useful for intranasal delivery of a pharmaceutical composition. Another formulation suitable for intranasal administration is a coarse powder comprising the active ingredient and having an average particle from about 0.2 μm to 500 μm . Such a formulation is administered in the manner in which snuff is taken, *i.e.* by rapid inhalation through the nasal passage from a container of the powder held close to the nose.

[00477] Formulations suitable for nasal administration may, for example, comprise from about as little as 0.1% (w/w) and as much as 100% (w/w) of active ingredient, and may comprise one or more of the additional ingredients described herein. A pharmaceutical composition may be prepared, packaged, and/or sold in a formulation suitable for buccal administration. Such formulations may, for example, be in the form of tablets and/or lozenges made using conventional methods, and may, for example, 0.1% to 20% (w/w) active ingredient, the balance comprising an orally dissolvable and/or degradable composition and, optionally, one or more of the additional ingredients described herein. Alternately, formulations suitable for buccal administration may comprise a powder and/or an aerosolized

and/or atomized solution and/or suspension comprising active ingredient. Such powdered, aerosolized, and/or aerosolized formulations, when dispersed, may have an average particle and/or droplet size in the range from about 0.1 nm to about 200 nm, and may further comprise one or more of any additional ingredients described herein.

Ophthalmic or otic administration

[00478] A pharmaceutical composition may be prepared, packaged, and/or sold in a formulation suitable for ophthalmic or otic administration. Such formulations may, for example, be in the form of eye or ear drops including, for example, a 0.1/1.0% (w/w) solution and/or suspension of the active ingredient in an aqueous or oily liquid excipient. Such drops may further comprise buffering agents, salts, and/or one or more other of any additional ingredients described herein. Other ophthalmically-administrable formulations which are useful include those which comprise the active ingredient in microcrystalline form and/or in a liposomal preparation. Subretinal inserts may also be used as a form of administration.

Payload administration

[00479] Glycan-interacting antibodies described herein may be used in a number of different scenarios in which delivery of a substance (the “payload”) to a biological target is desired, for example delivery of detectable substances for detection of the target, or delivery of a therapeutic or diagnostic agent. Detection methods can include, but are not limited to, both imaging *in vitro* and *in vivo* imaging methods, *e.g.*, immunohistochemistry, bioluminescence imaging (BLI), Magnetic Resonance Imaging (MRI), positron emission tomography (PET), electron microscopy, X-ray computed tomography, Raman imaging, optical coherence tomography, absorption imaging, thermal imaging, fluorescence reflectance imaging, fluorescence microscopy, fluorescence molecular tomographic imaging, nuclear magnetic resonance imaging, X-ray imaging, ultrasound imaging, photoacoustic imaging, lab assays, or in any situation where tagging/staining/imaging is required.

[00480] Glycan-interacting antibodies can be designed to include both a linker and a payload in any useful orientation. For example, a linker having two ends is used to attach one end to the payload and the other end to the glycan-interacting antibody. The glycan-interacting antibodies of the invention can include more than one payload as well as a cleavable linker. In another example, a drug that may be attached to glycan-interacting antibodies via a linker and may be fluorescently labeled can be used to track the drug *in vivo*, *e.g.* intracellularly.

[00481] Other examples include, but are not limited to, the use of glycan-interacting antibodies in reversible drug delivery into cells.

[00482] Glycan-interacting antibodies described herein can be used in intracellular targeting of a payload, *e.g.*, detectable or therapeutic agents, to specific organelles. In addition, glycan-interacting antibodies described herein may be used to deliver therapeutic agents to cells or tissues, *e.g.*, in living animals. For example, glycan-interacting antibodies described herein may be used to deliver chemotherapeutic agents to kill cancer cells. Glycan-interacting antibodies attached to therapeutic agents through linkers can facilitate membrane permeation allowing the therapeutic agent to travel into a cell to reach an intracellular target.

[00483] In some embodiments, the payload may be a therapeutic agent such as a cytotoxin, radioactive ion, chemotherapeutic, or other therapeutic agent. A cytotoxin or cytotoxic agent includes any agent that may be detrimental to cells. Examples include, but are not limited to, taxol, cytochalasin B, gramicidin D, ethidium bromide, emetine, mitomycin, etoposide, teniposide, vincristine, vinblastine, colchicine, doxorubicin, daunorubicin, dihydroxyanthracenedione, mitoxantrone, mithramycin, actinomycin D, 1-dehydrotestosterone, glucocorticoids, procaine, tetracaine, lidocaine, propranolol, puromycin, maytansinoids, *e.g.*, maytansinol (see U.S. Pat. No. 5,208,020 incorporated herein in its entirety), rachelmycin (CC-1065, see U.S. Pat. Nos. 5,475,092, 5,585,499, and 5,846,545, all of which are incorporated herein by reference), and analogs or homologs thereof. Radioactive ions include, but are not limited to iodine (*e.g.*, iodine 125 or iodine 131), strontium 89, phosphorous, palladium, cesium, iridium, phosphate, cobalt, yttrium 90, samarium 153, and praseodymium. Other therapeutic agents include, but are not limited to, antimetabolites (*e.g.*, methotrexate, 6-mercaptopurine, 6-thioguanine, cytarabine, 5-fluorouracil decarbazine), alkylating agents (*e.g.*, mechlorethamine, thiotepa chlorambucil, rachelmycin (CC-1065), melphalan, carmustine (BSNU), lomustine (CCNU), cyclophosphamide, busulfan, dibromomannitol, streptozotocin, mitomycin C, and cis-dichlorodiamine platinum (II) (DDP) cisplatin), anthracyclines (*e.g.*, daunorubicin (formerly daunomycin) and doxorubicin), antibiotics (*e.g.*, dactinomycin (formerly actinomycin), bleomycin, mithramycin, and anthramycin (AMC)), and anti-mitotic agents (*e.g.*, vincristine, vinblastine, taxol and maytansinoids). In the case of anti-STn antibodies of the present invention, tumor killing may be boosted by the conjugation of a toxin to such anti-STn antibodies.

[00484] In some embodiments, the payload may be a detectable agent, such as various organic small molecules, inorganic compounds, nanoparticles, enzymes or enzyme substrates,

fluorescent materials, luminescent materials (e.g., luminol), bioluminescent materials (e.g., luciferase, luciferin, and aequorin), chemiluminescent materials, radioactive materials (e.g., ^{18}F , ^{67}Ga , $^{81\text{m}}\text{Kr}$, ^{82}Rb , ^{111}In , ^{123}I , ^{133}Xe , ^{201}Tl , ^{125}I , ^{35}S , ^{14}C , ^3H , or $^{99\text{m}}\text{Tc}$ (e.g., as pertechnetate (technetate(VII), TcO_4^-)), and contrast agents (e.g., gold (e.g., gold nanoparticles), gadolinium (e.g., chelated Gd), iron oxides (e.g., superparamagnetic iron oxide (SPIO), monocrystalline iron oxide nanoparticles (MIONs), and ultrasmall superparamagnetic iron oxide (USPIO)), manganese chelates (e.g., Mn-DPDP), barium sulfate, iodinated contrast media (iohexol), microbubbles, or perfluorocarbons). Such optically-detectable labels include for example, without limitation, 4-acetamido-4'-isothiocyanatostilbene-2,2'-disulfonic acid; acridine and derivatives (e.g., acridine and acridine isothiocyanate); 5-(2'-aminoethyl)aminonaphthalene-1-sulfonic acid (EDANS); 4-amino-N-[3-vinylsulfonyl]phenyl]naphthalimide-3,5 disulfonate; N-(4-anilino-1-naphthyl)maleimide; anthranilamide; BODIPY; Brilliant Yellow; coumarin and derivatives (e.g., coumarin, 7-amino-4-methylcoumarin (AMC, Coumarin 120), and 7-amino-4-trifluoromethylcoumarin (Coumarin 151)); cyanine dyes; cyanosine; 4',6-diaminidino-2-phenylindole (DAPI); 5' 5"-dibromopyrogallol-sulfonaphthalein (Bromopyrogallol Red); 7-diethylamino-3-(4'-isothiocyanatophenyl)-4-methylcoumarin; diethylenetriamine pentaacetate; 4,4'-diisothiocyanatodihydro-stilbene-2,2'-disulfonic acid; 4,4'-diisothiocyanatostilbene-2,2'-disulfonic acid; 5-[dimethylamino]-naphthalene-1-sulfonyl chloride (DNS, dansylchloride); 4-dimethylaminophenylazophenyl-4'-isothiocyanate (DABITC); eosin and derivatives (e.g., eosin and eosin isothiocyanate); erythrosin and derivatives (e.g., erythrosin B and erythrosin isothiocyanate); ethidium; fluorescein and derivatives (e.g., 5-carboxyfluorescein (FAM), 5-(4,6-dichlorotriazin-2-yl)aminofluorescein (DTAF), 2',7'-dimethoxy-4'5'-dichloro-6-carboxyfluorescein, fluorescein, fluorescein isothiocyanate, X-rhodamine-5-(and-6)-isothiocyanate (QFITC or XRITC), and fluorescamine); 2-[2-[3-[[1,3-dihydro-1,1-dimethyl-3-(3-sulfopropyl)-2H-benz[e]indol-2-ylidene]ethylidene]-2-[4-(ethoxycarbonyl)-1-piperazinyl]-1-cyclopenten-1-yl]ethenyl]-1,1-dimethyl-3-(3-sulfopropyl)-1H-benz[e]indolium hydroxide, inner salt, compound with n,n-diethylethanamine(1:1) (IR144); 5-chloro-2-[2-[3-[(5-chloro-3-ethyl-2(3H)-benzothiazol-ylidene)ethylidene]-2-(diphenylamino)-1-cyclopenten-1-yl]ethenyl]-3-ethyl benzothiazolium perchlorate (IR140); Malachite Green isothiocyanate; 4-methylumbelliferone orthocresolphthalein; nitrotyrosine; pararosaniline; Phenol Red; B-phycoerythrin; o-phthalaldehyde; pyrene and derivatives (e.g., pyrene, pyrene butyrate, and succinimidyl 1-pyrene); butyrate quantum dots;

Reactive Red 4 (CIBACRON™ Brilliant Red 3B-A); rhodamine and derivatives (e.g., 6-carboxy-X-rhodamine (ROX), 6-carboxyrhodamine (R6G), lissamine rhodamine B sulfonyl chloride rhodamine (Rhod), rhodamine B, rhodamine 123, rhodamine X isothiocyanate, sulforhodamine B, sulforhodamine 101, sulfonyl chloride derivative of sulforhodamine 101 (Texas Red), N,N,N',N'tetramethyl-6-carboxyrhodamine (TAMRA) tetramethyl rhodamine, and tetramethyl rhodamine isothiocyanate (TRITC)); riboflavin; rosolic acid; terbium chelate derivatives; Cyanine-3 (Cy3); Cyanine-5 (Cy5); cyanine-5.5 (Cy5.5), Cyanine-7 (Cy7); IRD 700; IRD 800; Alexa 647; La Jolla Blue; phthalocyanine; and naphthalocyanine.

[00485] In some embodiments, the detectable agent may be a non-detectable precursor that becomes detectable upon activation (e.g., fluorogenic tetrazine-fluorophore constructs (e.g., tetrazine-BODIPY FL, tetrazine-Oregon Green 488, or tetrazine-BODIPY TMR-X) or enzyme activatable fluorogenic agents (e.g., PROSENSE® (VisEn Medical))). In vitro assays in which the enzyme labeled compositions can be used include, but are not limited to, enzyme linked immunosorbent assays (ELISAs), immunoprecipitation assays, immunofluorescence, enzyme immunoassays (EIA), radioimmunoassays (RIA), and Western blot analysis.

Combinations

[00486] Glycan-interacting antibodies may be used in combination with one or more other therapeutic, prophylactic, diagnostic, or imaging agents. By “in combination with,” it is not intended to imply that the agents must be administered at the same time and/or formulated for delivery together, although these methods of delivery are within the scope of the present disclosure. Compositions can be administered concurrently with, prior to, or subsequent to, one or more other desired therapeutics or medical procedures. In general, each agent will be administered at a dose and/or on a time schedule determined for that agent. In some embodiments, the present disclosure encompasses the delivery of pharmaceutical, prophylactic, diagnostic, and/or imaging compositions in combination with agents that may improve their bioavailability, reduce and/or modify their metabolism, inhibit their excretion, and/or modify their distribution within the body.

Dosage

[00487] The present disclosure encompasses delivery of glycan-interacting antibodies for any of therapeutic, pharmaceutical, diagnostic or imaging by any appropriate route taking

into consideration likely advances in the sciences of drug delivery. Delivery may be naked or formulated.

Naked delivery

[00488] Glycan-interacting antibodies of the present invention may be delivered to cells, tissues, organs or organisms in naked form. As used herein in, the term “naked” refers to glycan-interacting antibodies delivered free from agents or modifications which promote transfection or permeability. Naked glycan-interacting antibodies may be delivered to cells, tissues, organs and/or organisms using routes of administration known in the art and described herein. Naked delivery may include formulation in a simple buffer such as saline or PBS.

Formulated delivery

[00489] Glycan-interacting antibodies of the present invention may be formulated, using methods described herein. Formulations may comprise glycan-interacting antibodies which may be modified and/or unmodified. Formulations may further include, but are not limited to, cell penetration agents, pharmaceutically acceptable carriers, delivery agents, bioerodible or biocompatible polymers, solvents, and sustained-release delivery depots. Formulated glycan-interacting antibodies may be delivered to cells using routes of administration known in the art and described herein.

[00490] Compositions may also be formulated for direct delivery to organs or tissues in any of several ways in the art including, but not limited to, direct soaking or bathing, via a catheter, by gels, powder, ointments, creams, gels, lotions, and/or drops, by using substrates such as fabric or biodegradable materials coated or impregnated with compositions, and the like.

Dosing

[00491] The present invention provides methods comprising administering one or more glycan-interacting antibodies in accordance with the invention to a subject in need thereof. Nucleic acids encoding glycan-interacting antibodies, proteins or complexes comprising glycan-interacting antibodies, or pharmaceutical, imaging, diagnostic, or prophylactic compositions thereof, may be administered to a subject using any amount and any route of administration effective for preventing, treating, diagnosing, or imaging a disease, disorder, and/or condition. The exact amount required will vary from subject to subject, depending on

the species, age, and general condition of the subject, the severity of the disease, the particular composition, its mode of administration, its mode of activity, and the like.

Compositions in accordance with the invention are typically formulated in dosage unit form for ease of administration and uniformity of dosage. It will be understood, however, that the total daily usage of the compositions of the present invention will be decided by the attending physician within the scope of sound medical judgment. The specific therapeutically effective, prophylactically effective, or appropriate imaging dose level for any particular patient will depend upon a variety of factors including the disorder being treated and the severity of the disorder; the activity of the specific compound employed; the specific composition employed; the age, body weight, general health, sex and diet of the patient; the time of administration, route of administration, and rate of excretion of the specific compound employed; the duration of the treatment; drugs used in combination or coincidental with the specific compound employed; and like factors well known in the medical arts.

[00492] In certain embodiments, compositions in accordance with the present invention may be administered at dosage levels sufficient to deliver from about 0.0001 mg/kg to about 100 mg/kg, from about 0.01 mg/kg to about 50 mg/kg, from about 0.1 mg/kg to about 40 mg/kg, from about 0.5 mg/kg to about 30 mg/kg, from about 0.01 mg/kg to about 10 mg/kg, from about 0.1 mg/kg to about 10 mg/kg, or from about 1 mg/kg to about 25 mg/kg, of subject body weight per day, one or more times a day, to obtain the desired therapeutic, diagnostic, prophylactic, or imaging effect. The desired dosage may be delivered three times a day, two times a day, once a day, every other day, every third day, every week, every two weeks, every three weeks, or every four weeks. In certain embodiments, the desired dosage may be delivered using multiple administrations (*e.g.*, two, three, four, five, six, seven, eight, nine, ten, eleven, twelve, thirteen, fourteen, or more administrations).

[00493] According to the present invention, glycan-interacting antibodies may be administered in split-dose regimens. As used herein, a “split dose” is the division of single unit dose or total daily dose into two or more doses, *e.g.*, two or more administrations of the single unit dose. As used herein, a “single unit dose” is a dose of any therapeutic administered in one dose/at one time/single route/single point of contact, *i.e.*, single administration event. As used herein, a “total daily dose” is an amount given or prescribed in a 24 hr period. It may be administered as a single unit dose. In one embodiment, glycan-interacting antibodies of the present invention are administered to a subject in split doses. Glycan-interacting antibodies may be formulated in buffer only or in a formulation described

herein. Pharmaceutical compositions comprising glycan-interacting antibodies as described herein may be formulated into a dosage form described herein, such as a topical, intranasal, intratracheal, or injectable (e.g., intravenous, intraocular, intravitreal, intramuscular, intracardiac, intraperitoneal or subcutaneous). General considerations in the formulation and/or manufacture of pharmaceutical agents may be found, for example, in Remington: The Science and Practice of Pharmacy 21st ed., Lippincott Williams & Wilkins, 2005 (incorporated herein by reference).

Coatings or shells

[00494] Solid dosage forms of tablets, dragees, capsules, pills, and granules can be prepared with coatings and shells such as enteric coatings and other coatings well known in the pharmaceutical formulating art. They may optionally comprise opacifying agents and can be of a composition that they release the active ingredient(s) only, or preferentially, in a certain part of the intestinal tract, optionally, in a delayed manner. Examples of embedding compositions which can be used include polymeric substances and waxes. Solid compositions of a similar type may be employed as fillers in soft and hard-filled gelatin capsules using such excipients as lactose or milk sugar as well as high molecular weight polyethylene glycols and the like.

IV. Kits and Devices

Kits

[00495] Any of the compositions described herein may be comprised in a kit. In a non-limiting example, reagents for generating glycan-interacting antibodies, including antigen molecules are included in a kit. The kit may further include reagents or instructions for creating or synthesizing glycan-interacting antibodies. It may also include one or more buffers. Other kits of the invention may include components for making glycan-interacting antibody protein or nucleic acid arrays or libraries and thus, may include, for example, a solid support.

[00496] The components of the kits may be packaged either in aqueous media or in lyophilized form. The container means of the kits will generally include at least one vial, test tube, flask, bottle, syringe or other container means, into which a component may be placed, and preferably, suitably aliquoted. Where there are more than one component in the kit (labeling reagent and label may be packaged together), the kit also will generally contain a

second, third or other additional container into which the additional components may be separately placed. The kits may also comprise a second container means for containing a sterile, pharmaceutically acceptable buffer and/or other diluent. However, various combinations of components may be comprised in a vial. The kits of the present invention also will typically include a means for containing the glycan-interacting antibodies, e.g., proteins, nucleic acids, and any other reagent containers in close confinement for commercial sale. Such containers may include injection or blow-molded plastic containers into which the desired vials are retained.

[00497] When the components of the kit are provided in one and/or more liquid solutions, the liquid solution is an aqueous solution, with a sterile aqueous solution being particularly preferred. However, the components of the kit may be provided as dried powder(s). When reagents and/or components are provided as a dry powder, the powder can be reconstituted by the addition of a suitable solvent. It is envisioned that the solvent may also be provided in another container means. In some embodiments, labeling dyes are provided as a dried powder. It is contemplated that 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 120, 120, 130, 140, 150, 160, 170, 180, 190, 200, 300, 400, 500, 600, 700, 800, 900, 1000 micrograms or at least 1000 micrograms or at most 10 g of dried dye are provided in kits of the invention. The dye may then be resuspended in any suitable solvent, such as DMSO.

[00498] A kit may include instructions for employing the kit components as well the use of any other reagent not included in the kit. Instructions may include variations that can be implemented.

Devices

[00499] Any of the compositions described herein may be combined with, coated onto or embedded in a device. Devices include, but are not limited to, dental implants, stents, bone replacements, artificial joints, valves, pacemakers or other implantable therapeutic devices.

V. Equivalents and scope

[00500] Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments in accordance with the invention described herein. The scope of the present invention is not intended to be limited to the above Description, but rather is as set forth in the appended claims.

[00501] In the claims, articles such as “a,” “an,” and “the” may mean one or more than one unless indicated to the contrary or otherwise evident from the context. Claims or descriptions

that include “or” between one or more members of a group are considered satisfied if one, more than one, or all of the group members are present in, employed in, or otherwise relevant to a given product or process unless indicated to the contrary or otherwise evident from the context. The invention includes embodiments in which exactly one member of the group is present in, employed in, or otherwise relevant to a given product or process. The invention includes embodiments in which more than one, or the entire group members are present in, employed in, or otherwise relevant to a given product or process.

[00502] It is also noted that the term “comprising” is intended to be open and permits but does not require the inclusion of additional elements or steps. When the term “comprising” is used herein, the term “consisting of” is thus also encompassed and disclosed.

[00503] Where ranges are given, endpoints are included. Furthermore, it is to be understood that unless otherwise indicated or otherwise evident from the context and understanding of one of ordinary skill in the art, values that are expressed as ranges can assume any specific value or subrange within the stated ranges in different embodiments of the invention, to the tenth of the unit of the lower limit of the range, unless the context clearly dictates otherwise.

[00504] In addition, it is to be understood that any particular embodiment of the present invention that falls within the prior art may be explicitly excluded from any one or more of the claims. Since such embodiments are deemed to be known to one of ordinary skill in the art, they may be excluded even if the exclusion is not set forth explicitly herein. Any particular embodiment of the compositions of the invention (*e.g.*, any nucleic acid or protein encoded thereby; any method of production; any method of use; *etc.*) can be excluded from any one or more claims, for any reason, whether or not related to the existence of prior art.

[00505] All cited sources, for example, references, publications, databases, database entries, and art cited herein, are incorporated into this application by reference, even if not expressly stated in the citation. In case of conflicting statements of a cited source and the instant application, the statement in the instant application shall control.

[00506] Section and table headings are not intended to be limiting.

EXAMPLES

Example 1. Glycan array analysis

[00507] Optimized glycan arrays are utilized to test antibody affinity and specificity for multiple glycans in a single experiment. Glycan arrays used herein include arrays that comprise 71 chemically synthesized and well-defined glycans, most of which comprise

Neu5Ac and Neu5Gc glycan pairs. Array slides are obtained commercially (ArrayIt Corp, Sunnyvale, CA) and include the glycans listed in the following Table.

Table 6. Array glycans

Glycan ID No.	Glycan
1	Neu5,9Ac2 α 2,3Gal β 1,4GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
2	Neu5Gc9Ac α 2,3Gal β 1,4GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
3	Neu5,9Ac2 α 2,6Gal β 1,4GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
4	Neu5Gc9Ac α 2,6Gal β 1,4GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
5	Neu5Ac α 2,6GalNAc α O(CH ₂) ₂ CH ₂ NH ₂
6	Neu5Gc α 2,6GalNAc α O(CH ₂) ₂ CH ₂ NH ₂
7	Neu5,9Ac2 α 2,3Gal β 1,3GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
8	Neu5Gc9Ac α 2,3Gal β 1,3GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
9	Neu5,9Ac2 α 2,3Gal β 1,3GalNAc α O(CH ₂) ₂ CH ₂ NH ₂
10	Neu5Gc9Ac α 2,3Gal β 1,3GalNAc α O(CH ₂) ₂ CH ₂ NH ₂
11	Neu5Ac α 2,3Gal β 1,4GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
12	Neu5Gc α 2,3Gal β 1,4GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
13	Neu5Ac α 2,3Gal β 1,3GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
14	Neu5Gc α 2,3Gal β 1,3GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
15	Neu5Ac α 2,3Gal β 1,3GalNAc α O(CH ₂) ₂ CH ₂ NH ₂
16	Neu5Gc α 2,3Gal β 1,3GalNAc α O(CH ₂) ₂ CH ₂ NH ₂
17	Neu5Ac α 2,6Gal β 1,4GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
18	Neu5Gc α 2,6Gal β 1,4GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
19	Neu5Ac α 2,6Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
20	Neu5Gc α 2,6Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
21	Neu5Ac α 2,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
22	Neu5Gc α 2,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
23	Neu5,9Ac2 α 2,6GalNAc α O(CH ₂) ₂ CH ₂ NH ₂
24	Neu5Gc9Ac α 2,6GalNAc α O(CH ₂) ₂ CH ₂ NH ₂
25	Neu5Ac α 2,3Gal β O(CH ₂) ₂ CH ₂ NH ₂
26	Neu5Gc α 2,3Gal β O(CH ₂) ₂ CH ₂ NH ₂
27	Neu5Ac α 2,6Gal β O(CH ₂) ₂ CH ₂ NH ₂
28	Neu5Gc α 2,6Gal β O(CH ₂) ₂ CH ₂ NH ₂
29	Neu5,9Ac2 α 2,3Gal β O(CH ₂) ₂ CH ₂ NH ₂
30	Neu5Gc9Ac α 2,3Gal β O(CH ₂) ₂ CH ₂ NH ₂
31	Neu5,9Ac2 α 2,6Gal β O(CH ₂) ₂ CH ₂ NH ₂
32	Neu5Gc9Ac α 2,6Gal β O(CH ₂) ₂ CH ₂ NH ₂
33	Neu5Ac α 2,3Gal β 1,3GalNAc β O(CH ₂) ₂ CH ₂ NH ₂
34	Neu5Gc α 2,3Gal β 1,3GalNAc β O(CH ₂) ₂ CH ₂ NH ₂
35	Neu5,9Ac2 α 2,3Gal β 1,3GalNAc β O(CH ₂) ₂ CH ₂ NH ₂
36	Neu5Gc9Ac α 2,3Gal β 1,3GalNAc β O(CH ₂) ₂ CH ₂ NH ₂
37	Neu5,9Ac2 α 2,6Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
38	Neu5Gc9Ac α 2,6Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
39	Neu5,9Ac2 α 2,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
40	Neu5Gc9Ac α 2,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
41	Neu5Ac α 2,8Neu5Ac α 2,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
42	Neu5Ac α 2,8Neu5Ac α 2,8Neu5Ac α 2,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂

43	Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
45	Gal β 1,4GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
47	GalNAc α O(CH ₂) ₂ CH ₂ NH ₂
51	Gal β 1,3GalNAc β O(CH ₂) ₂ CH ₂ NH ₂
52	Gal β 1,3GlcNAc α O(CH ₂) ₂ CH ₂ NH ₂
53	Gal β 1,3GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
54	Gal β 1,4GlcNAc6S β O(CH ₂) ₂ CH ₂ NH ₂
55	Neu5Ac α 2,3Gal β 1,4(Fuc α 1,3)GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
56	Neu5Gc α 2,3Gal β 1,4(Fuc α 1,3)GlcNAc β O(CH ₂) ₂ CH ₂ NH ₂
57	Neu5Ac α 2,3Gal β 1,4(Fuc α 1,3)GlcNAc6S β O(CH ₂) ₂ CH ₂ NH ₂
58	Neu5Gc α 2,3Gal β 1,4(Fuc α 1,3)GlcNAc6S β O(CH ₂) ₂ CH ₂ NH ₂
59	Gal β 1,3GlcNAc β 1,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
60	Neu5Ac α 2,3Gal β 1,3GlcNAc β 1,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
61	Neu5Gc α 2,3Gal β 1,3GlcNAc β 1,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
62	Neu5Ac α 2,3Gal β 1,4GlcNAc6S β O(CH ₂) ₂ CH ₂ NH ₂
63	Neu5Gc α 2,3Gal β 1,4GlcNAc6S β O(CH ₂) ₂ CH ₂ NH ₂
64	Neu5Ac α 2,8Neu5Ac α 2,3Gal β 1,4Glc β O(CH ₂) ₃ NHCOCH ₂ (OCH ₂ CH ₂) ₆ NH ₂
65	Neu5Ac α 2,8Neu5Ac α 2,8Neu5Ac α 2,3Gal β 1,4Glc β O(CH ₂) ₃ NHCOCH ₂ (OCH ₂ CH ₂) ₆ NH ₂
66	Neu5Ac α 2,6(Neu5Ac α 2,3)Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
67	Neu5Ac α 2,6(Neu5Gc α 2,3)Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
68	Neu5Ac α 2,6(KDN α 2,3)Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
69	Neu5Gc α 2,8Neu5Ac α 2,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
70	KDN α 2,8Neu5Ac α 2,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
71	Neu5Ac α 2,8Kdn α 2,6Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
72	Neu5Ac α 2,8Neu5Gc α 2,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
73	Neu5Ac α 2,8Neu5Gc α 2,6Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
74	KDN α 2,8Neu5Gc α 2,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
75	Neu5Gc α 2,8Neu5Gc α 2,3Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂
76	Neu5Ac α 2,8Neu5Ac α 2,6Gal β 1,4Glc β O(CH ₂) ₂ CH ₂ NH ₂

[00508] 300 ml of epoxy blocking buffer is prepared by combining 15 ml of 2 M Tris buffer (pH 8) with 0.9 ml of 16.6 M ethanolamine and 284.1 ml of distilled water. The solution is brought to a final pH of 9.0 with HCl. The solution is filtered using a 0.2 μ M nitrocellulose membrane. The epoxy buffer solution as well as 1 L of distilled water are pre-warmed to 50°C. Glass slides are arranged in a slide holder and quickly submerged in a staining tub with the warmed epoxy blocking buffer. Slides are incubated in the epoxy blocking buffer for 1 hour at 50°C with periodic shaking to deactivate epoxy binding sites. Next, slides are rinsed and blocked with PBS with 1% OVA at 25°C for one hour. Serum samples with polyclonal antibodies (1:1000) or purified monoclonal antibodies (1 μ g/mL), are diluted in PBS with 1% OVA and added to the glycan array for one hour at 25°C. After extensive washing, binding of antibodies are detected by incubating glycan microarray slides with Cy3-conjugated anti-mouse IgG (Jackson ImmunoResearch, West Grove, PA) for one hour. Slides are then washed extensively, dried and scanned with a Genepix 4000B scanner

(Laser at 100%; gain at 350; 10 μ m pixels). Raw data from scanned images are extracted using the Genepix software and analysis of raw data is carried out. Antibodies are considered to be highly specific for AcSTn and GcSTn if they demonstrate binding to both molecules, but not to Tn or any other glycans on the array.

[00509] Based on array analysis, antibodies are classified according to array glycan binding profile. Antibodies are classified as “Group 1” antibodies, capable of binding AcSTn and GcSTn, if they bind to glycans 5, 6, 23 and 24. Such antibodies are referred to as Pan-STn antibodies due to their ability to associate with a wider range of STn structures and the portion of STn indicated by the large oval in Fig. 1A. Antibodies are classified as “Group 2” antibodies, capable of binding STn as well as some related structures that include an O-linkage to serine or threonine, if they bind to glycans 5, 6, 23, 24, 27 and 31. These antibodies are thought to associate with the portion of STn indicated by the large oval in Fig. 1B. Some Group 2 antibodies preferably bind to structures with AcSTn over structures with GcSTn. Antibodies are classified as “Group 3” antibodies (capable of binding STn, but may also bind a broader set of related structures) if they bind glycans 5, 6, 23, 24, 17, 3, 19, 37, 27 and 31. Unlike Group 2 antibodies, Group 3 antibodies do not require that such structures have an O-linkage to serine or threonine. Group 3 antibodies are thought to associate with the portion of STn indicated by the large oval in Fig. 1C. Finally, antibodies are “Group 4” antibodies, capable of binding to both AcSTn and GcSTn as well as the un-sialylated Tn antigen (therefore having broader specificity) if they bind to glycans 5, 6, 23, 24 and 47. Group 4 antibodies are thought to associate with the portion of STn indicated by the large oval in Fig. 1D.

Example 2. Flow cytometry-based analysis of antibody binding

[00510] Flow cytometry-based analysis is carried out to elucidate the dose-response curve for binding of antibodies to cell surface antigens. For these analyses, various cell lines are employed.

[00511] MDA-MB-231 cells are human breast cancer cells. They are grown in Earle's Minimum Essential Medium supplemented with 10% fetal calf serum (FCS), 100 μ g/ml penicillin, 100 UI/ml streptomycin and 45 μ g/ml gentamycin. MCF-7 cells are also human breast cancer cells and are grown under the same conditions as MDA-MB-231 cells. Stably transfected versions of MDA-MB-231 and MCF-7 cells (clone TAH3.P10 for MDA-MB-231 cells and clone A12.1 for MCF-7 cells) that over express GalNAc α 2,6-sialyltransferase

(ST6GalNAc 1,) are also cultured under the same conditions with the exception of an added 1mg/ml of G418 to support cells expressing the transgene. ST6GalNAc 1 is an enzyme capable of sialylating GalNAc. As a result of over expression, transfected cells express high levels of Neu5Ac-STn (see Julien, S. et al., Glycoconjugate journal. 2001. 18, 883-93; the contents of which are herein incorporated by reference in their entirety).

[00512] E3 cells are murine breast cancer cells. They are cultured in Dulbecco's E4 medium with 10% FCS. Stably transfected versions of E3 cells expressing high levels of Neu5Gc-STn (E3-STn) are cultured with 600 µg/ml of G418 and 200 µg/ml hygromycin. During growth and maintenance of experimental cells, trypsin is not used for cell passaging.

[00513] OV90 and OVCAR3 cells are also used. These are human ovarian cancer cell lines, described previously.

[00514] SNU-16 cells are also used. These are gastric cancer cell lines that express low levels of STn.

[00515] For analysis, cells are harvested using StemPro Accutase (Life Technologies, Carlsbad, CA) and washed with PBS comprising 5% FBS before pelleting by light centrifugation. Cell numbers and viability are determined by trypan blue dye exclusion analysis and cell concentrations are adjusted to 5×10^6 cells/ml in PBS with 5% FBS. 50 µl of cells are added to each well of an assay plate. Cells are combined with 50 µl solutions of antibody being analyzed or control antibodies and incubated for 1 hour at 4°C. Cells are washed and pelleted twice with PBS with 5% FBS before being treated with 100 µl of PBS with 5% FBS comprising a 1:1,500 dilution of anti-mouse IgG (Southern Biotech, Birmingham, Alabama,) conjugated to allophycocyanin (APC). Cells are incubated for 30 min at 4°C before washing and resuspending in 200 µl of propidium iodide (PI) diluted 1:1000 in PBS with 5% FBS. Treated cells are then subjected to flow cytometry analysis and 10,000 events are acquired for each sample.

Example 3: Immunization with alternative antigens and adjuvants

[00516] An immunization study was carried out to develop antibodies against STn. 40 C57BL/6 or Balb/C wild type mice (females, 6-8 weeks old) were acclimated for at least 3 days and given access to standard diet (2920X.10, Global 18% Protein Rodent Diet, Harlan, San Diego, CA) and acidified water (pH 2.7-3.0) ad libitum throughout the study period. Mice were divided into 4 groups of 10 mice each (a total of 8 groups). Mice were immunized according to the study design shown in the following Table, using either PSM (including

digested PSM) and/or OSM at a dose of 10 µg mixed with adjuvants. Adjuvants included either Freund's adjuvant (complete or incomplete) or enhanced adjuvants comprising AbiSCO-100 (12µg) and ODN-2395 (50µg).

Table 7: Study Design

Group	Strain	Immunogen and Adjuvant
1	C57Bl/6 or Balb/C	PSM (10 µg) + AbiSCO-100(12 µg) + ODN-2395 (50µg)
2	C57Bl/6 or Balb/C	PSM (10 µg) or OSM + AbiSCO-100(12 µg) + ODN-2395 (50µg)
3	C57Bl/6 or Balb/C	Digested PSM (10 µg) + AbiSCO-100(12 µg) + ODN-2395 (50µg)
4	C57Bl/6 or Balb/C	OSM (10 µg) + AbiSCO-100(12 µg) + ODN-2395 (50µg)

[00517] Mice were randomized for placement into individual treatment groups based on body weight and sex. Vaccinations were given by subcutaneous (SC) injections around armpits and inguinal regions (50 µl per site, 4 sites for a total of 200 µl per mouse). Each mouse received 4 immunizations on days 0, 14, 28 and 42 by subcutaneous injection during the study period. Each mouse was immunized with 10µg PSM, digested PSM or OSM antigen which was mixed with 12 µg of AbiSCO-100 and 50µg of ODN-2395. After the 4th subcutaneously injection, each mouse was immunized with 10 µg PSM, digested PSM or OSM only intraperitoneally (IP) on day 63, 64 or 70 as the final immunization for each mouse, depending on which animal or group was selected. The detailed immunization schedule is presented in the following Table.

[00518] Additionally, body weight and health observations for each mouse were determined twice per week. Blood was collected from all animals prior to the immunization at each immunization date except for the last boost (i.e. the 5th immunization by IP) during the study period. The last blood sample was collected on day 51.

[00519] During each blood collection, approximately 0.2 ml of whole blood was collected by facial vein bleed and placed into serum separator tubes. Tubes were then kept at room temperature for at least 30 minutes to allow clotting. Serum was then divided into aliquots of equal volume and stored at -80°C until analysis.

Table 8: Immunization schedule

Group	Schedule (days)				
	1 st (day 0)	2 nd (day	3 rd	4 th (day 42)	5 th (day 63 or 64

		14)	(day28)		or 70)
1	SC, with PSM +adjuvant	SC, with PSM +adjuvant	SC, with PSM +adjuvant	SC, with PSM +adjuvant	IP; selected animals; PSM, Digested PSM or OSM only (no adjuvant)
2	SC, with PSM +adjuvant	SC, with OSM +adjuvant	SC, with PSM +adjuvant	SC, with OSM +adjuvant	
3	SC, with Digested PSM +adjuvant	SC, with Digested PSM +adjuvant	SC, with Digested PSM +adjuvant	SC, with Digested PSM +adjuvant	
4	SC, with OSM +adjuvant	SC, with OSM +adjuvant	SC, with OSM +adjuvant	SC, with OSM +adjuvant	

[00520] Anti-STn serum titer was determined using a murine anti-STn bovine submaxillary mucin (BSM) ELISA (with or without periodate treatment) together with serum profiles observed by glycan microarray. 96-well plates were coated with 1 µg/well of BSM and incubated overnight at 4°C. O-acetylation of BSM antigen was removed by treating wells with 0.1 M sodium hydroxide. Specific binding to STn was determined by treatment of wells with sodium periodate. Periodate treatment destroys the C6 side chain of sialic acid; therefore antibodies raised against STn should not bind to periodate-treated wells. Wells were blocked with PBS 1% ovalbumin (OVA). Serum samples to be assayed were serially diluted in PBS 1% OVA. A commercially available mouse anti-STn monoclonal antibody, 3F1 (SBH Sciences, Natick, MA) was used as a positive control. This antibody was also serially diluted in PBS with 1% OVA. A pool of serum from naïve wild type mice was used for the preparation of negative control samples. Detection of anti-STn antibodies present in serum was determined using an HRP-conjugated polyclonal goat anti-mouse IgG antibody (Jackson ImmunoResearch, West Grove, PA). The reaction was stopped by addition of sulfuric acid (1.6 M). Optical densities were measured at 490 nm using a Spectramax microplate reader (Molecular Devices, Sunnyvale, CA). The serum titer was obtained by comparison of OD values with a cutoff value calculated as two standard deviations above the mean of optical density values of the negative control. Sample tests were considered positive if the mean optical density value was greater than the cutoff value.

[00521] Mice with high anti-STn antibody titers were selected for hybridoma fusions. Resulting hybridomas were screened by BSM ELISA (with or without periodate treatment to indicate antibodies capable of differentiating between 9-O-acetylated STn variants) as described above. Selected hybridomas were subcloned and subclones were further

characterized by ELISA and flow cytometry analysis (using MDA-MB-231 cells expressing STn, as described previously, or using OV90 cells) to identify whether the antibodies produced by such cells are capable of binding epitope Group 1, 2, 3 or 4 (see Fig. 1).

Characterization data obtained for select antibodies is presented in the following Table. In the Table, NB indicates that weak or no binding was observed and ND indicates that a value was not determined.

Table 9: Anti-STn antibody subclone ELISA and flow cytometry characterization

Clone	Isotype	BSM ELISA		Flow Cytometry (MDA-MB-231 STn+)			Flow Cytometry (OV90)		
		EC ₅₀ (nM)	Hill Slope	EC ₅₀ (nM)	Hill Slope	High Value	EC ₅₀ (nM)	Hill Slope	High Value
4D9-2D4	IgG1, k	NB	ND	10.8	ND	ND	ND	ND	ND
1A5-2C9	IgG3, k	NB	ND	NB	ND	ND	ND	ND	ND
8C2-2D6	IgG2a, k	0.4	1.1	2.8	0.9	653	2.8	0.8	1237
7D3-2C10	IgG1,k	0.3	0.9	0.6	1.1	211	0.422	0.91	1021
2C6-2F11	IgG1,k	0.1	1.7	0.7	1.2	268	11.56	0.35	604
2B2-2A7	IgG1, k	NB	ND	0.3	ND	ND	ND	ND	ND
5G2-1B3	IgG1,k	0.4	2.7	0.2	1.3	376	1.68	0.79	570
7A5-2G12	IgG3,k	0.5	1.3	0.5	1.3	223	3.0	7.0	446
6B11-2E3	IgG3, k	0.3	0.5	0.2	1.5	229	0.266	0.89	359
1C11-2G9	IgG2a, k	NB	ND	3.3	ND	ND	ND	ND	ND
2G12-2B2	IgG2a, k	0.1	0.6	0.3	1.3	489	0.43	0.63	960
9E5-1A8	IgG3, k	NB	ND	3.1	ND	ND	ND	ND	ND
10C9-2G7	IgG1,k	NB	ND	2.3	ND	ND	ND	ND	ND
2F4-1E2	IgG3, k	4.2	9.6	0.36	1.26	144	4.35	0.53	88
2F4-	IgG3,	NB	ND	1.4	ND	ND	ND	ND	ND

1H8	k								
2B8-2F10	IgG3, k	3.5	ND	5.2	ND	ND	ND	ND	ND
5E6-2E7	IgG3, k	5.8	1.1	1.8	1.0	126	1.9	0.8	259
9F11-1F7	IgG3, k	4.6	1.0	0.8	0.9	124	1.98	0.89	265
IF6-1C10	IgG3, k	0.3	1.2	1.0	0.6	167	0.5	13.0	347
4D9-2C11	IgG1, k	NB	ND	2.7	ND	ND	ND	ND	ND
7D4-2A2-2F2	IgG2a, k	0.7	ND	14.1	ND	ND	ND	ND	ND
7D4-1H12-2B3	IgG2a, k	1.2	0.9	0.7	1.0	619	0.54	1.13	912
4G8-1E3	IgG2a, k	1.0	1.3	0.8	0.8	153	1.0	1.1	221
2C2-2C5	IgG3, k	1.1	2.0	1.0	1.1	141	1.08	1.05	342
10F4-2A9	IgG3, k	5.1	ND	1.9	ND	ND	ND	ND	ND

[00522] Results of glycan array characterization are presented in the following Table. The highest level of signal obtained for each antibody during glycan array analysis is indicated as is the resulting group and specificity determination based on the glycans that each antibody associated with and the intensity of the signal obtained for each glycan. Group determination was made according to the method of Example 1. Specificity determinations also factor in intensity of association with Ac versus Gc structures.

Table 10: Anti-STn glycan array characterization data

Clone	Glycan Array		Specificity
	Highest Signal	Group	
4D9-2D4	700	1	Pan-STn
1A5-2C9	5800	1	Pan-STn
8C2-2D6	16000	1	Pan-STn
7D3-2C10	4000	1	Gc-STn
2C6-2F11	1200	1	Pan-STn
2B2-2A7	13400	4	Pan-STn and Tn
5G2-1B3	14000	4	Pan-STn and Tn

7A5-2G12	35000	1	Pan-STn
6B11-2E3	25000	1	Pan-STn
1C11-2G9	700	1	Gc-STn
2G12-2B2	25000	1	Pan-STn
9E5-1A8	500	ND	ND
10C9-2G7	6000	1	Pan-STn
2F4-1E2	25000	4	Pan-STn and Tn
2F4-1H8	1900	4	Pan-STn and Tn
2B8-2F10	2000	1	Gc-STn
5E6-2E7	14000	1	Pan-STn
9F11-1F7	16000	1	Gc-STn
1F6-1B7	7200	1	Pan-STn
IF6-1C10	35000	1	Pan-STn
4D9-2C11	1400	1	Pan-STn
7D4-2A2-2F2	5100	1	Pan-STn
7D4-1H12- 2B3	25000	1	Pan-STn
4G8-1E3	35000	1	Pan-STn
2C2-2C5	25000	1	Pan-STn
10F4-2A9	2400	1	Gc-STn

[00523] Subclones expressing antibodies capable of binding STn were subjected to amino acid and nucleotide sequence analysis (see the following Example).

Example 4: Variable domain pairs

[00524] Sequenced clones yielded variable domain pairs with the amino acid sequences presented in the following Table. “ND” indicates that the sequence was not determined.

Table 11. Variable domain pairs

Clone ID Number	VH, SEQ ID NO	VL, SEQ ID NO
7D3-2C10	15	16
A5-2G12	17	18
1A5-2C9	19	20
4D9-2C11	21	22
2F4-1E2	23	24
2F4-1H8	23	24
2C6-2F11	25	26
2B2-2A7	27	28
5G2-1B3	29	30

7A6-2A2	31	32
10C9-2G7	33	ND
1F6-1B7 (also referred to as 1F6-1C10)	35	36
2G12-2B2	37	38
5E6-2E7	39	40
9F11-1F7	39	40
9E5-1A8	41	
10F4-2A9	42	32
10F4-2F2	42	32
2B8-2F10	43	
4G8-1E3	44	45
6B11-2E3	46	32
8C2-2D6	47	32
7D4-1H12-2B3	48	50
7D4-2A2-2F2	48	49
2C2-2C5	51	52
1C11-2G9	ND	34

[00525] CDRs for each variable domain were determined and are presented in the following Table. Each row presents a set of six CDRs identified for each clone.

Table 12. CDR sequences

Clone ID	CDR-H1	SEQ ID NO	CDR-H2	SEQ ID NO	CDR-H3	SEQ ID NO	CDR-L1	SEQ ID NO	CDR-L2	SEQ ID NO	CDR-L3	SEQ ID NO
7A6-2A2	GYTF TDHA IHWV	81	FSPG NDDI KY	84	SVGY ALDY	90	ENVV TY	100	GASN RYT	115	GQGY SYPY T	127
2B2-2A7	GYTF TDHA IHWV	81	ISPGN GDIK Y	85	KISY YGI	91	EDIYS N	101	KATN LAD	116	QHFW GTPF T	128
5G2-1B3	GYTF TDHA IHWV	81	FSPG NDDI KY	84	KRSY YGD	92	ENIYS H	102	GATN LAD	117	QHFW GAPF T	129
4D9-2C11	GYTF TDHA IHWV	81	LSPG NDDI KY	86	KRSIG GDH	93	QNIN VW	103	KASN LHT	118	QQGQ SYPFT	130
2F4-1E2	GYTF TDHA IHWV	81	ISPGN GDIK Y	85	QRQL GQGY	94	QSLV HSYG NTY	104	KVSN RFS	119	SQNT HVPY T	131
2F4-1H8	GYTF TDHA IHWV	81	ISPGN GDIK Y	85	QRQL GQGY	94	QSLV HSYG NTY	104	KVSN RFS	119	SQNT HVPY T	131
1A5-2C9	GYTF TDHA IHWV	81	VSPG NGDI KY	87	KRSLI GDY	95	QNVG TA	105	SASN RYT	120	QQYS SYRL T	132

1F6-1B7	GYTF TDHA IHWV	81	ISPGN GDVK Y	88	KRSL STPY	96	QSLN NSGN QKSY	106	WAST RDS	121	QSDY SYPY T	133
2C2-2C5	GYTF TDHA IHWV	81	ISPGN GDIK Y	85	KRSIT TPY	97	QSVN NN	107	YASN RYT	122	QQGY SSPW T	134
2G12-2B2	GYTF TDHA IHWV	81	FSPG NDDI KY	84	KRSL STPY	96	QSLN NRGN HKNY	108	WAST RES	123	QNDY TYPY T	135
2C6-2F11	GYTF SDHA IHWV	82	ISPGN DDIK Y	89	ERSM IGVY	98	QSLV QSNQ NTY	109	KVSN RFC	124	SQST HAPL T	136
7D4-2A2-2F2	GYIFT DHAI HWV	83	ISPGN GDIK Y	85	KRSL STPY	96	ENVV NY	110	GASN RYS	125	GSKW ITSYP YT	137
7D4-1H12-2B3	GYIFT DHAI HWV	83	ISPGN GDIK Y	85	KRSL STPY	96	ENVV NY	110	GASN RYS	125	GARV TSYP YT	138
7D3-2C10	GYTF TDHA IHWV	81	FSPG NDDI KY	84	KRSIT TPY	97	QNIN VW	103	KVSN LHT	126	QQDQ SYPY T	139
8C2-2D6	GYTF TDHA IHWV	81	ISPGN GDIK Y	85	KRSIT TSY	99	ENVV TY	100	GASN RYT	115	GQGY SYPY T	127
5E6-2E7	GYTF TDHA IHWV	81	ISPGN GDIK Y	85	KRSIT TPY	97	QSLN NSGK TKNY	111	WAST RES	123	KNDY SYPY T	140
9F11-1F7	GYTF TDHA IHWV	81	ISPGN GDIK Y	85	KRSIT TPY	97	QSLN NSGK TKNY	111	WAST RES	123	KNDY SYPY T	140
4G8-1E3	GYIFT DHAI HWV	83	ISPGN GDIK Y	85	KRSIT TSY	99	QHIN FW	112	KASN LHT	118	QQDQ SYPY M	141
10F4-2F2	GYTF TDHA IHWV	81	ISPGN GDIK Y	85	KRSIT TSY	99	ENVV TY	100	GASN RYT	115	GQGY SYPY T	127
10F4-2A9	GYTF TDHA IHWV	81	ISPGN GDIK Y	85	KRSIT TSY	99	ENVV TY	100	GASN RYT	115	GQGY SYPY T	127
6B11-2E3	GYTF TDHA IHWV	81	ISPGN DDIK Y	89	KRSIT TSY	99	ENVV TY	100	GASN RYT	115	GQGY SYPY T	127
7A5-2G12	GYTF TDHA IHWV	81	ISPGN DDIK Y	89	KRSIT TSY	99	ENVV IY	113	GASN RYT	115	GQGY SYPY T	127

Example 5. Flow cytometry analysis of antibody internalization

[00526] Flow cytometry analysis is carried out in order to quantify the extent of antibody internalization. For analysis, stably transfected variants of MDA-MB-231 cells (clone TAH3.P10) that express high levels of cell surface-bound Neu5Ac-STn are harvested using

10 mM EDTA and washed with PBS comprising 1% BSA before pelleting by light centrifugation. Cell numbers and viability are determined by trypan blue dye exclusion analysis and cell concentrations are adjusted to 5×10^6 cells/ml in PBS with 1% BSA. 50 μ l of cells are added to each well of an assay plate. Cells are combined with 50 μ l solutions of antibody or fluorescently-labeled antibody and incubated for 1 hour at 4°C. Following this incubation period, cells are washed with PBS to remove unbound antibody and aliquots are removed for incubation for various times (15, 30, 60 minutes) at 37°C to allow bound antibody to internalize at a physiologically relevant temperature. After each incubation, cell surface-bound antibody is removed by treating cells with acidic medium (150 mM NaCl, pH = 2.5). Cells treated with unlabeled antibody are washed with PBS and fixed with paraformaldehyde fixation buffer (PFA) containing 3% paraformaldehyde and 2% sucrose in PBS for 15 minutes at room temperature. These cells are rinsed again in PBS and treated with blocking buffer made up of PBS with 1% bovine serum albumin (BSA). Cells are incubated for 30 min at room temperature, rinsed in PBS and treated with secondary antibody (allophycocyanin-labeled goat-anti-mouse IgG) for 2 hours at room temperature. All cells are then washed with PBS and subjected to flow cytometry analysis wherein 10,000 events are recorded for each sample. Residual fluorescent signal in acid-treated samples is further quenched via treatment with trypan blue dye.

Example 6. Evaluation of antibody internalization through cell viability assay

[00527] Cell viability assays are performed to screen anti-STn antibodies of the present invention in the presence and absence of secondary antibody-drug conjugates (2°ADCs). The purpose of the screen is to identify the ability of each anti-STn antibody to inhibit cell growth. Antibodies with potent cell growth inhibition are used to design direct antibody-drug conjugates (ADCs). Using such secondary antibody-drug conjugates (2°ADCs) in cell-based cytotoxic assays can quickly pre-screen many ADC candidates against tumor cells. Based on the assay, a naked antibody candidate is directly added to cells in the presence of a 2°ADC. Internalization of the mAb/2°ADC complex into cells that express a high density of the targeted antigen can achieve a dose-dependent drug release within the cells, causing a cytotoxic effect to kill the cells (e.g., tumor cells), while cells expressing a low density of the targeted antigen are not affected (e.g., normal cells).

[00528] To perform cell viability assays, cell lines described in the present application (MDA-MB-231 parental, MDA-MB-231-STn+, and OV-90) are prepared and cultured for the

assays. The cell culture is optimized for cell density by plating different densities of cells (e.g., 2,000, 4,000 and 7,500 per well) on a 96-well plate and observing the cell growth for 96 hours. The plating condition in which cells reach around 90% confluence at the end of the 96 hours is identified and the optimal cell number is then used in the final viability assay.

[00529] Antibodies are tested in one or more cell lines in the presence and absence of a 2°ADC such as Fab α MFc-CL-MMAF. Duplicate or triplicate cell plates for each cell line are used for testing each antibody candidate.

[00530] For cell viability assays, data points are collected for each antibody candidate with duplicates for each data point. Each antibody candidate is diluted in serial concentrations from 0.3pM to 20nM. A constant amount of Fab α MFc-CL-MMAF (40 nM) is used in the viability assay.

[00531] Alternatively, data points are collected for each antibody candidate with triplicates for each data point. Each antibody candidate is diluted in serial concentrations from 1 pM to 20 nM. A constant amount of Fab α MFc-CL-MMAF (40 nM) is used in the viability assay.

[00532] Cell viabilities are measured by Cell-Titer Glo luminescence based assays.

Example 7. Demonstration of *in vivo* tumor killing ability

[00533] *In vivo* tumor killing ability is demonstrated with mouse and/or human tumor cell lines. Tumor cell lines expressing STn targets are transferred into mice and the ability of the antibody candidates to kill the resulting tumors is determined.

[00534] Mouse cell lines used *in vivo* in tumor killing assays include the mouse colon adenocarcinoma cell line, MC38, derived from C57BL/6 mice and stably transfected with ST6 (alpha-N-acetyl-neuraminyl-2,3-beta-galactosyl-1,3)-N-acetylgalactosaminide alpha-2,6-sialyltransferase 1 (ST6GalNac1). These cells are fed with sialic acid (Neu5Ac and/or Neu5Gc, depending on target) before their use in *in vivo* tumor killing assays using syngeneic *Cmah*^{-/-} mice.

[00535] Alternatively, for *in vivo* tumor killing assays human breast cancer cell lines (T47-D, MCF-7 or MDA-MB-231) induced to express a high level of STn are transferred into immune-deficient FOXN1 ^{-/-} (nude) cells, non-obese diabetic (NOD) cells, or severe immunodeficiency (SCID) mice.

[00536] *In vivo* ADCC is induced by passive transfer of human peripheral blood mononuclear cells (PBMCs) or purified natural killer (NK) cells. In cases where candidate

antibodies bind unspecifically to wild-type mouse tissue, immune-deficient mice are bred into the Cmah ^{-/-} background.

Example 8. Phage library construction and selection

[00537] RNA is prepared from spleens harvested from mice with a strong immune response to immunization. Mouse variable (V) regions are PCR amplified and assembled into scFv expression constructs. ScFv sequences are cloned into phagemid display vectors allowing for scFv display on the surface of M13 phage particles. The resulting library is transformed into E. coli (TG1). Bulk transformations of E.coli are grown and phage are prepared by phage rescue. In the first round of selection, phage from the culture medium are purified by PEG precipitation.

[00538] Candidate scFvs are selected using both negative and positive selection methods. For negative selection, the library is incubated with “destroyed” STn-negative mucin (e.g. chemically treated PSM). For positive selection, the library is incubated with GcSTn mucin (e.g. PSM and/or de-O-acetylated BSM), AcSTn mucin (e.g. OSM and/or de-O-acetylated BSM) or BSM (and/or de-O-acetylated BSM) and a synthetic glycan (Neu5Gc and/or Neu5Ac) in the presence of a Neu5Ac or Neu5Gc (depending on the desired target).

[00539] After 3-4 rounds of selection with reducing antigen concentrations, 1000 clones are analyzed by ELISA for binding to STn (e.g. Neu5Ac and/or Neu5Gc) using synthetic and natural glycan targets. Lead phage/scFv candidates are tested in a secondary flow cytometry-based cellular assay for binding to GcSTn and/or AcSTn using Jurkat cells with or without “induction” of GcSTn or AcSTn. Up to 20 selected scFv candidates of interest are subjected to further analysis.

[00540] Lead scFv candidates are selected for conversion to IgG. Variable regions from each scFv are cloned into mammalian expression vectors between an upstream CMV promoter and a downstream immunoglobulin constant region. Heavy chain vector includes murine IgG1 and κ constant regions. Vectors are transiently transfected into HEK293/EBNA cells. Antibody samples are purified and characterized by binding to positive and negative glycan epitopes. Samples of up to 0.5 mg of each whole IgG are further analyzed.

Example 9. Antibody-dependent cell-mediated cytotoxicity optimization

[00541] Genes encoding the variable regions of a selected IgG are cloned into mammalian expression vectors encoding human Fc regions (huIgG1 κ) containing amino acid mutations known to enhance Fc-receptor binding and antibody-dependent cell-mediated cytotoxicity

(ADCC). Vectors are transiently transfected into HEK293/EBNA cells. After 2-7 days, IgG expression is quantified and samples of antibody are purified on protein A columns. Antibodies are then tested in ADCC assays. Neu5Gc and Neu5Ac-expressing Jurkat cell lines are used as the target cells and human peripheral blood mononuclear cells (PBMC) are used as a source of effector cells. Target cells are titrated using maximum cell lysis to determine the optimum cell density for use in multiwell plate format assay. ADCC-mutated antibody together with the non-mutated IgG are pre-incubated with target cells, effector cells are then added at varying target:effector cell ratios, and cultures are incubated at 37°C. Percentage viability is determined using Calcein-AM dye (BD Biosciences, San Jose, CA) release. Samples of up to 0.5 mg of ADCC-mutated IgG are subjected to further analysis.

Example 10. Production of lead antibody from semi-stable HEK cell line

[00542] Variable regions from IgG are cloned into mammalian expression vectors between an upstream CMV promoter and a downstream immunoglobulin constant region. Heavy chain vector includes murine IgG1 and κ constant regions. Vectors are transiently transfected into HEK293/EBNA cells and antibody titers are assessed at 72 hours. Transiently transfected HEK293/EBNA cells are selected with hygromycin to establish a semi-stable expression system. Semi-stable cells are expanded to 10 liters. Antibodies are purified from the culture supernatant by Protein A, dialyzed into PBS and the resulting preparation is analyzed for (1) aggregates by analytical size exclusion chromatography (SEC), (2) endotoxin levels by *Limulus* amoebocyte lysate (LAL) testing (expressed as EU/mg), and (3) binding to antigen in the primary assay.

Example 11. Additional assays for screening scFv candidates for target affinity

[00543] ScFv candidates are subjected to additional screening methods for STn (pan-STn, AcSTn and/or GcSTn) affinity using a variety of proposed targets.

Synthetic glycan target screening

[00544] As used herein, the term “target screening” refers to the use of a target substance to identify binding partners for that substance. Synthetic glycan target screening is carried out using desired STn target antigens bound to poly(acrylic acid) (PAA) with a biotin tag. Undesired STn target antigens as well as Tn bound to PAA with a biotin tag are used as negative controls. Cells associated with candidate scFvs are isolated through precipitation with avidin-associated entities.

Natural glycan target screening on live cells

[00545] Target screening using live cells is carried out using Jurkat cells fed with sialic acid (Neu5Gc and/or Neu5Ac, depending on the desired antibody target) or Jurkat cells fed with an alternative form of sialic acid (Neu5Gc and/or Neu5Ac, depending on the desired antibody target) as a negative control. Target screening using live cells is also carried out using MCF-7 or MDA-MB-231 cells fed with sialic acid (Neu5Gc and/or Neu5Ac, depending on the desired antibody target or whether being used for negative control screening) and stable transfection. Flow cytometry is used in either case to isolate cells associated with scFv candidates.

Natural glycan target screening on tissue (ex vivo)

[00546] Target screening using *ex vivo* tissue is carried out using biopsy tissue samples. Binding of scFv candidates with *ex vivo* tissue is analyzed using standard immunohistochemical methods. Single tissue sections as well as tissue microarray sections are used. Samples are treated with or without sialidase and/or periodate in control experiments.

Example 12. Antibody humanization

[00547] Fully humanized heavy and light chains are designed with CDRs presented herein. Protein models of the variable regions are generated using existing antibody structures as templates. Segments of starting heavy and light chain variable region amino acid sequences are compared with human sequences for possible inclusion in the fully humanized sequences. Series of humanized heavy and light chain variable regions are designed entirely from segments of human variable region sequences with the objective that T cell epitopes be avoided. Variant human sequence segments with significant incidence of potential T cell epitopes as determined by *in silico* technologies are discarded.

[00548] Humanized heavy and light chain variable region genes are constructed from overlapping oligonucleotides assembled into full length genes using the ligase chain reaction (LCR). LCR products are amplified and suitable restriction sites are added for cloning into expression vectors. PCR products are cloned into intermediate vectors and confirmed by sequencing.

[00549] For construction of expression plasmids encoding fully humanized antibodies with human constant regions, DNA sequences for each variable region are inserted into

mammalian expression vectors between an upstream cytomegalovirus immediate/early promoter/enhancer (CMV IE) plus the immunoglobulin signal sequence and a downstream immunoglobulin constant region gene. DNA samples are prepared for transfection into mammalian cells.

[00550] For generation of cell lines and selection of lead fully humanized antibodies, heavy and light chain plasmid DNA pairs are transfected into mammalian cells (NS0). Cell lines producing humanized antibodies are expanded and antibody samples are purified. Antibodies are tested in primary and secondary binding assays to determine leading antibody candidates. The 3 leading candidates are used for further analysis.

Example 13. Immunogenicity testing

[00551] Lead antibodies are subjected to EpiScreen (Antitope, Paradise Valley, AZ) whole antibody human T cell assays using a minimum of 20 blood samples from healthy volunteer donors. Immunogenicity of lead antibodies is compared with control chimeric antibodies with starting antibody variable regions and matched human constant regions. Data are benchmarked against EpiScreen whole protein data for clinical-stage biologics.

Example 14. Cell line development

[00552] Cell lines are developed with the ability to yield high levels of antibody with no non-human glycosylation due to knock down of the CMAH gene. Cell lines are glycoengineered to increase ADCC. These cell lines have the ability to perform in small and large scale production.

Example 15. Antibody-dependent inhibition of STn-positive tumor cell immune tolerance

[00553] Anti-STn antibodies of the present invention are provided and used to contact tumor cells and tissues comprising STn glycans. Immune-dependent targeting of STn-tumor cells is increased.

Example 16. Treatment of immune tolerant tumors using anti-STn antibodies

[00554] A subject with STn glycans expressed on and around tumor cells is treated with an anti-STn antibody of the present invention. Immune tolerance of subject tumor cells is decreased.

Example 17. Generation of S3F antibodies

[00555] S3F IgG2a antibodies were generated through the combination of 3F1 IgG1 variable domains (SBH Biosciences, Natick, MA) with antibody constant domain regions from IgG2 antibodies. The heavy and light chain variable domains of 3F1 were sequenced and constructs were generated encoding 3F1 variable domains upstream of IgG2 expression vectors, plasmid H1206 (LakePharma, Belmont, CA) for antibody heavy chains and plasmid L1206 (LakePharma, Belmont, CA) for antibody light chains. Related sequences are presented in the following Table.

Table 13. Sequences utilized in S3F IgG2 antibody generation

Description	SEQ ID NO
3F1 VH domain	53
3F1 VL domain	54
IgG2a heavy chain constant domain	234
kappa light chain constant domain	235
S3F full length heavy chain	236
S3F full length light chain	238
S3F full length heavy chain nucleotide sequence	237
S3F full length light chain nucleotide sequence	239

[00556] Plasmids encoding S3F full heavy chain amino acid sequences and plasmids encoding S3F full light chain amino acid sequences were transfected into Chinese hamster ovary-K1 (CHO-K1) cells for the generation of stable cell lines expressing S3F IgG2a antibodies. The cells were cultured in a humidified 5% CO₂ incubator at 37°C in chemically defined media (CD-CHO, Invitrogen, Carlsbad, CA) supplemented with L-glutamine.

[00557] Approximately 80 million suspension CHO cells, growing in log phase, were transfected by electroporation (MaxCyte) with 80 µg of total plasmid encoding the full length heavy and light chains of S3F. Twenty four hours later, the transfected cells were placed under selection for stable integration of the antibody genes. During the selection process the cells were spun down and resuspended in fresh selection media every 2-3 days until the pool recovered its growth rate and viability. Cells were monitored for growth, titer, and stable integration of the antibody expression constructs. The doubling rate was 20 hours.

[00558] Two small scale production scale-ups were performed using the stably transfected cells. The cells were scaled up for production in OptiCHO CD Growth Medium (Invitrogen). The product was produced at a titer of approximately 12 mg per liter. The doubling rate was

20 hours. The conditioned media supernatant harvested from the transient transfection production run was clarified by centrifuge spinning. The protein was run over a Protein A column and eluted using two different buffer formulations (Citrate Buffer and HEPES buffer). Filtration using a 0.2 μ m membrane filter was performed. Size exclusion chromatography (SEC) was performed for both formulations (see the following Table).

Table 14. SEC data

Buffer	Results
20 mM Citrate (pH 5.5) + 150 mM NaCl	Purity 91.4%, aggregates 3.4%
200 mM HEPES (pH 7) + 0.2% acetate	Purity 96.2%, aggregates 3.8%

[00559] Stable cell lines were cultured for large scale production and 10 L of culture was produced. The conditioned media harvested from the stable cell pool production run was clarified by centrifugation and 0.2 μ m membrane filtration. The antibody was purified using Protein A affinity chromatography, then sterilized and cleared of particulates by passing through a 0.2 μ m membrane filter. After low endotoxin purification and filtration, concentration was set to 5 mg/mL and 120 mg of antibody S3F was recovered.

Example 18. Characterization of carcinoma cancer stem cell lines and CSC sub-fractions by expression levels of STn antigens using anti-STn antibodies

[00560] In order to test the viability of the present unique immunotherapeutic approach to the eradication of CSCs, human ovarian carcinoma cell lines and their associated CSC subfractions are employed as the target cell population.

[00561] To identify appropriate ovarian carcinoma cell lines and CSC subfractions from these identified cell lines, for further in vitro analysis of anti-STn antibody efficacy, different human ovarian cancer cell lines including SKOV3, OVCAR3, OVCAR4, OV90 and A2870 are analyzed for the expression of Ovarian CSC biomarkers: CD44 and CD133, by flow cytometry. CD44 and/or CD133 positive/expressed ovarian CSC subfractions are then further tested for the co-expression levels of cell-surface STn antigen using specific anti-STn antibodies, for example, anti-STn antibodies S3F and recombinant 18D2 pan anti-STn IgG2a mAb [R18D2; IgG2a version of 18D2 (which is an IgG2b) described in international publication No. WO2015/054600, the contents of which are herein incorporated by reference in their entirety]. CSC subfractions that show strong anti-STn staining are purified by cell sorting and subject to further test for stem cell attributes. Selected CSC subfractions are

tested for “stem-like” characteristics by performing serial colony-forming transplantation assays. Then STn⁺ subfractions that demonstrate superior colony-forming ability are identified for efficacy experiments. In addition to colony formation assays, cell cycle and chemoresistance are also analyzed.

[00562] To verify this, CSC fractions are isolated from ovarian cancer cell lines based on surface markers (CD44, CD133) known to enrich for CSCs in ovarian cancer. Five different ovarian cancer cell lines (SKOV3, OV90, OVCAR3, OVCAR4, and A2870) are utilized. Only OV90, OVCAR4, and OVCAR3 harbor subfractions of CD133-expressing and/or CD44-expressing cells that display some aspect of stem-like properties. These ovarian cancer cell lines are subjected to flow cytometric analysis (via Aria) using the FlowJo software and sorting via multiple strategies.

[00563] To characterize the expression of STn on CSC subfractions expressing CD44 and/or CD133, flow cytometry of the parental cell lines is used along with fluorophore-conjugated antibodies to CD44 and CD133 (Miltenyl Biotech, Cambridge, MA) to prepare subfractions expressing one or both of these markers as well as a fraction in which neither of these markers is expressed. Subsequently fluorophore-conjugated mAb S3F is used to determine the levels of STn labeling of these subfractions using standard procedures. Several concentrations of conjugated S3F are tested along with time-course experiments to establish optimal cell binding conditions. Appropriate fluorophore-conjugated, isotype matched antibodies are used as controls to establish background staining. Labeled cells are resuspended in PBS buffer and subjected to flow cytometric analysis using a BD FACSAria instrument and FlowJo 7.6.5 software (Treestar, Ashland, OR).

[00564] Consequently, these analyses are used to characterize the CD44/133 defined subfractions of ovarian CSCs in terms of co-expression of STn.

[00565] In addition, the same cell lines described above are subjected to FACS sort based on STn⁺ and STn⁻ cell fractions, followed by a second sort based on expression of CD44 and CD133. These eight subfractions are seeded in a colony-forming assay. It is well accepted that stem-like cells are thought to replicate infrequently, only giving rise to a limited number of daughter cells that eventually give rise to the bulk of the tumor cell population. This property has been widely exploited to identify and track stem cells. Colonies are allowed to form for 3-4 weeks and then counted under a microscope after overnight staining with 1 mg/ml nitroblue tetrazolium chloride in PBS. Colony forming efficiency is calculated by

dividing the number of colonies by the total number of cells plated and then multiplying by 100.

[00566] The in vitro serial transplantation methodology is carried out as described by Padler-Karavani et al (Padler-Karavani et al., Cancer research, 2011, 71, 3352-3363; and Friel et al., Cell cycle, 2008, 7, 242-249). Culture plates containing Noble agar underlayers are used to seed sorted cell subfractions at a seeding density of 1×10^4 into Noble agar overlayers. Cultures are incubated in a humidified 5% CO₂ environment at 37°C. After about 4 weeks in culture, cells are counted under a microscope after overnight staining with 1 mg/ml nitroblue tetrazolium chloride in PBS and replated as above. Serial transplantation in vitro continues for several passages (depending on characteristics of the individual cell lines). The CD44⁺ and/or CD133⁺ cells that co-express STn display an enhanced colony-forming advantage over the CD44⁻/CD133⁻/STn⁻, CD44⁺/CD133⁻/STn⁻, CD44⁺/CD133⁺/STn⁻, CD44⁻/CD133⁺/STn⁻, and CD44⁻/CD133⁻/STn⁺ cell subfractions.

[00567] Further cell lines are analyzed for their potential for STn synthesis using PCR to measure the expression of the sole enzyme known to be responsible for its biosynthesis, namely the sialyltransferase ST6GalNAc I. RNA is isolated from frozen, sorted subfractions as initially cultured and cDNA prepared by standard methods prior to quantitative PCR. A set of housekeeping genes (e.g. PUM1, RPLP0, ACTB) are first tested to determine which is more appropriate (i.e., to determine which are the most homogeneously expressed across the cell subfractions) to use as an internal reference. Expression of ST6GalNAc I is then measured using a set of PCR probes validated for their efficiency in the method. Each subfraction is further compared with positive controls, including ST6GalNAc I cDNA inserted into bacterial plasmids, and total RNA from breast cancer cells stably transfected with ST6GalNAc I cDNA as described (Julien et al., Glycoconjugate journal, 2001, 18, 883-893). Based on levels of ST6GalNAc I mRNA, culture conditions are manipulated to ensure that cell-surface STn is expressed and studies are carried out for further analysis.

[00568] Studies are used to define the levels of STn expression in the various CSC subfractions as determined by flow cytometric analysis of three CD44/CD133 positive ovarian carcinoma CSCs. Further, serial transplantation colony-forming assays are carried out to compare the characteristic stem cell-like properties of the CSC subfractions from these three lines. On the basis of STn expression levels and STn⁺ subfraction colony-forming ability, two of the three ovarian cell lines are chosen for in vitro and in vivo functional assays.

Example 19. Determining the anti-CSC activity of murine anti-STn mAbs in vitro

[00569] The proliferative ability of STn⁺ subfractions and the anti-CSC activity of anti-STn mAbs (e.g. S3F) are tested in vitro. A series of in vitro experiments are performed to assess the effects of mAb treatment on stem cell proliferation and ADC-induced killing of CSC subfractions and parent cell lines. In appropriate assays, anti-STn mAbs are combined with standard-of-care chemotherapy agents (e.g., carboplatin, paclitaxel).

[00570] In order to optimize subsequent assays used to determine the effect of the anti-STn antibody S3F on the CSC subfractions, proliferation assays are performed essentially as described (Friel et al., Cell cycle, 2008, 7, 242-249). The eight sorted CSC subfraction combinations possible from each of the two selected ovarian carcinoma cell lines plus cells from the two parental lines are seeded in triplicate (dilutions beginning at 5×10^3 cells/well) in 24-well culture plates. Viable cells, as determined by trypan blue exclusion, are counted using a hemocytometer and or plate reader via MTT assay on days 1, 3, 5, 7, 8, and 9, or longer if necessary. Student's t-test is used to compare groups and comparisons with a p value of ≤ 0.05 are considered significant. Based on the results, sub-objective protocols are devised in order to perform cytotoxicity assays.

[00571] Cytotoxicity assays are performed to assess sensitivity of the eight STn⁺ and STn⁻ sorted subfractions from each cell line as well as the parental lines to S3F mAb alone, S3F + paclitaxel + carboplatin or the combination of chemotherapeutic agents alone. S3F (along with an irrelevant, isotype-matched antibody as control) is tested at several dilutions beginning at 0.1 μ M. The chemotherapeutic agents are used at 0.1, 1.0, 10, and 100 nM. Appropriate vehicle control groups are also included and assays are performed essentially as described previously (Friel et al., Cell cycle, 2008, 7, 242-249). Sorted CSC subfractions are seeded in triplicate at cell numbers determined in cell proliferation assays into 24-well culture plates and allowed to reach 50% confluence. Cells are then starved for 24 hours after which inhibitory agents are added. Cell viability is determined as above on days 2, 4, and 5, or longer if necessary. Student's t-test is used to compare groups and comparisons with a p value of ≤ 0.05 are considered significant.

[00572] The proliferative ability of all possible combinations of CSC marker-defined subfractions (CD44, CD133, and STn), as well as the parental cell lines, is measured and used to establish protocols to assess the effect of the anti-STn antibody (+/- standard chemotherapy) on these cells. Stem cell marker positive cells have a reduced proliferative capacity when compared to parental lines and those subfractions that are negative for CSC

antigens. The anti-CSC effect of ADC-formatted S3F (+/- standard chemotherapy) and its mechanism(s) of action are determined.

Example 20: Determining the anti-CSC activity of murine anti-STn in vivo

[00573] To determine whether murine STn antibodies possess anti-tumor activity in vivo as the result of targeting CSCs, in vivo toxicology and pharmacology experiments are performed in mice. The growth conditions of CSC subfractions in NOD/SCID mice is established and analyzed. Anti-STn antibodies are administered to mice as conjugates with chemotherapy and treated mice are tested for toxicity efficacy through mouse observations and weights.

[00574] An equivalent amount of serially-diluted positive or negative subfraction cells (i.e., 1×10^5 , 1×10^4 , and 1×10^3 /mouse; 2-8 mice/group) are injected subcutaneously (s.c.) into NOD/SCID mice, to compare their potential generate tumors. Following injection of tumor cells, the mice are monitored daily for evidence of tumor formation and general health. For treatment, tumors may be generated in mice from up to four sources: 1) primary ovarian tumors; 2) xenograft tumors previously established in NOD/SCID mice; 3) cryopreserved xenograft tumor tissue; or 4) human ovarian cancer cell lines or isolated cell line subfractions based on CD133/STn expression. For each tumor type, once a sufficient number of mice have tumors ranging from 200-300 mm³ in volume, the mice are randomized into 4 groups (n=8-14 mice/group) such that the mean group tumor volumes are essentially equivalent. The first group receives a single isotype antibody control, IgG-MMAE (1-5 mg/kg), in 200 µl of vehicle administered intraperitoneally (i.p.) once weekly; the second group receives i.p. S3F-MMAE (1-5 mg/kg), in 200 µl of vehicle administered intraperitoneally (i.p.) once weekly; the third group will receive, by i.p. injection, SIA101-MMAE (1-5 mg/kg), in 200 µl of vehicle administered intraperitoneally (i.p.) once weekly, and the fourth group receives vehicle i.p. weekly.

[00575] Tumors are then measured twice weekly with calipers [tumor volume = (Lmm x Wmm x Lmm)/2]. Mice, at the same time, are observed frequently and weighed twice weekly to assess potential toxic effects. Mice with excessive tumor burden or in a moribund state are euthanized. Non-parametric Wilcoxon rank sum test for unpaired samples is used to compare tumor volumes among the different arms.

[00576] S3F-MMAE is capable of delaying the growth of or eradicating CSC tumors and combination with standard chemotherapy agents used for ovarian cancer increases the efficacy of the mAb.

Example 21. In vitro testing of ADC antibodies

[00577] S3F, 8C2-2D6, 2G12-2B2, 4G8-1E3 and 5G2-1B3 antibodies were conjugated with maleimidocaproyl-valine-citruline-p-aminobenzyloxycarbonyl-monomethyl auristatin E (MC-vc-PAB-MMAE). Conjugation was carried out by first reducing the antibody interchain disulfide bonds with TCEP and then linking the maleimide moiety of the drug to reduced cysteines. Conjugated antibodies were desalted on Sephadex G50 columns to remove residual unreactive toxins and then dialyzed in 30 mM HEPES pH 7.7 with 150 mM NaCl. Drug-antibody ratio (DAR) was determined for each conjugated antibody using the ratio of UV absorbance values at 248 and 280 nm (A_{248} and A_{280} , respectively; see the following Table).

Table 15. MMAE conjugated antibodies

Antibody	Ratio of A_{248}/A_{280}	Drug:Antibody Ratio
S3F	0.73	4.6
8C2-2D6	0.75	4.8
2G12-2B2	0.73	4.6
4G8-1E3	0.73	4.5
5G2-1B3	0.62	3

[00578] Antibody preparations were analyzed by size exclusion chromatography to confirm the presence of pure monomers.

[00579] Conjugated antibodies were then tested for their ability to kill cells expressing STn. Cultures of MDA-MB-231 cells, with or without STn surface expression. MDA STn negative cells were grown in EMEM supplemented with 10% FBS, 1x Pen/Strep and 45ug/mL gentamycin. MDA STn positive cells were grown in the same media except with the addition of 1mg/mL G418 for antibiotic selection. Cells were seeded separately (4,000 cells/well STn negative or 2,000/well STn positive) in 96 well plates using proper media described above. Cells were grown overnight. After 16-20 hours, cells were treated with a serial dilution of test antibodies in triplicate (50nM to 0.012nM, 1:4 serial dilution in media) for 72 hours. After treatment, no significant decrease in STn-negative cell viability was observed by CELLTITER-GLO® luminescent cell viability assay, while decreases in cell viability were

observed with all conjugated antibodies (with the exception of non-STn-specific control conjugated antibody) in STn-positive cell cultures (see the following Table).

Table 16. IC50 values for MMAE conjugated antibodies tested on STn+ MDA cells

Antibody	IC50
S3F	0.51
8C2-2D6	2.44
2G12-2B2	1.19
4G8-1E3	1.80
5G2-1B3	11.13
Control	0.0

[00580] Of the conjugated antibodies tested, S3F antibody had lowest inhibitory concentration (IC50). Antibodies 2G12-2B2, 4G8-1E3 and 8C2-2D6 all had single digit nanomolar IC50s. Antibody 5G2-1B3 was the least effective with an IC50 value over 10. This may have been due to a lower DAR and different IgG isotype (5G2-1B3 is IgG1, whereas others are IgG2a). Isotype control conjugated antibody showed no effect.

Example 22. ADC in vitro assay with OVCAR3 and SNU-16 cell lines

[00581] The experiment performed above was repeated using OVCAR3 cells (shown to express moderate levels of STn) or SNU-16 cells (shown to express low levels of STn). Cells were seeded (10,000 cells/well) in 96-well assay plates using RPMI media with 10% FBS with 0.01% bovine insulin and 1x Pen/Strep. Cells were incubated in standard cell culture conditions overnight before treatment with test antibodies (with or without MMAE drug conjugates) or irrelevant control antibody. Cells were treated with a serial dilution of test antibodies in triplicate (50nM to 0.012nM, 1:4 serial dilution in media).

[00582] Four days later, cell viability was measured by CELL TITER-GLO® luminescent cell viability assay. As in the previous assay, unconjugated antibodies had no effect on cell viability; however, conjugated anti-STn decreased cell viability with increasing doses (see following Table).

Table 17. IC50 values for antibodies tested

Antibody	OVCAR3 cells (IC50, nM)	SNU-16 cells (IC50, nM)
S3F	0.75	6.3
8C2-2D6	14.45	31.8
2G12-2B2	6.82	24.5
4G8-1E3	9.09	28.7

5G2-1B3	11.3	96.3
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[00583] Of the antibodies tested in OVCAR3 cells, conjugated S3F had the lowest IC₅₀, while all other antibodies tested had IC₅₀ values greater than 5. 8C2-2D6 and 5G2-1B3 had IC₅₀s above 10nM, while antibody 2G12-2B2 and 4G8-1E3 had more desirable IC₅₀s in the single nanomolar range. Isotype control conjugated antibody showed no effect.

[00584] Similar results were observed in SNU-16 cells, with conjugated S3F demonstrating the greatest effect. 2G12-2B2, 8C2-2D6 and 5G2-1B3 also demonstrated strong ability to kill SNU-16 cells.

Example 23. OVCAR3 binding affinity

[00585] Antibodies S3F, 8C2-2D6, 4G8-1E3 and 2G12-2B2 were tested for binding to OVCAR3 cells using flow cytometry analysis. For analysis, OVCAR3 cells were grown and harvested using StemPro Accutase (Life Technologies, Carlsbad, CA) and washed with PBS comprising 5% FBS before pelleting by light centrifugation. Cell numbers and viability were determined by trypan blue dye exclusion analysis and cell concentrations were adjusted to 5 x 10⁶ cells/ml in PBS with 5% FBS. 50 µl of cells were added to each well of an assay plate. Cells were then combined with 50 µl solutions of the antibodies being analyzed or control antibodies and incubated for 1 hour at 4°C. Cells were next washed and pelleted twice with PBS with 5% FBS before being treated with 100 µl of PBS with 5% FBS comprising a 1:1,500 dilution of anti-mouse IgG (Southern Biotech, Birmingham, Alabama,) conjugated to allophycocyanin (APC). Cells were then incubated for 30 min at 4°C before washing and resuspending in 200 µl of propidium iodide (PI) diluted 1:1000 in PBS with 5% FBS. Finally, treated cells were subjected to flow cytometry analysis.

[00586] Mean of APC fluorescence intensity was plotted against the log of the antibody concentration to yield a curve used for EC₅₀ calculation. EC₅₀ values obtained for each antibody are presented in the following Table.

Table 18. Effective concentration for OVCAR3 binding

Antibody	EC₅₀
S3F	0.92
8C2-2D6	3.13
2G12-2B2	0.80
4G8-1E3	1.66

[00587] All antibodies were effective binders with single digit nanomolar EC₅₀ values.

Example 24. Antibody testing by immunohistochemistry

[00588] S3F, 2C2-2C5, 5E6-2E7, 9F11-1F7, 5G2-1B3, 4G8-IE3 and 8C2-2D6 were tested for their ability to detect STn in frozen tissue sections, fixed tissue sections, and in cancer tissue microarrays. 2429-2B2-3B9 (as described in U.S. publication No. US2014/0178365, the contents of which are herein incorporated by reference in their entirety) was also tested. Further, antibody B72.3 (Thermo Fisher Scientific, Waltham, MA) was used as a positive control (except in testing of frozen tissue).

[00589] Frozen tissue samples were derived from human pancreatic carcinoma (Hu14 Neo-1) and staining was examined in tumor cells, endothelium, spindloid cells, and duct epithelium. Formalin-fixed paraffin embedded (FFPE) tissue samples were derived from human pancreatic carcinoma (Hu3 PA Neo-1) and staining was examined in tumor cells, stroma, and lumen.

[00590] Antibodies being tested were used to treat tissue sections and then detected using peroxidase-labeled secondary detection reagents. Tissue sections were observed and evaluated for intensity of specific reactivity, frequency of cells displaying specific reactivity, and subcellular localization of specific reactivity.

[00591] Staining observations are presented in the following Tables.

Table 19. Abbreviation key for staining observations

Intensity of specific reactivity	Frequency of cells displaying specific reactivity	Subcellular localization of specific reactivity (when applicable)
0 - Negative	N - Negative, no reactivity	M – Membrane
1 - Weak	VR - Very rare, <5% of cells of that cell type	C – Cytoplasm
2 - Mild	R - Rare, 5-25% of cells of that cell type	M>C - Membrane staining with lesser amounts of cytoplasmic
3 - Moderate	O - Occasional, 26-50% of cells of that cell type	C>M - Cytoplasmic staining with lesser amounts of membrane
4 - Strong	F - Frequent, 51-75% of cells of that cell type	
5 - Intense	VF - Very frequent, 76-100% of cells of that cell type	

Table 20. FFPE tissue evaluation

Antibody	Concentration	Specific Reactivity (Intensity/Frequency)		
		Hu3 PA Neo-1 (pancreatic carcinoma, tumor cells, cytoplasm>membrane)	Stroma (extracellular matrix)	Lumen contents
Assay Control	0 µg/mL	0/N	0/N	0/N
2C2-2C5	10 µg/mL	3-5/R	1-2/VR	3-5/VR
	1 µg/mL	3-5/VR	1-2/VR	3-5/VR
5E6-2E7	10 µg/mL	1-3/VR	0/N	1-3/VR
	1 µg/mL	1-2/VR	0/N	0/N
9F11-1F7	10 µg/mL	3-5/VR	1/VR	1-2/VR
	1 µg/mL	1-3VR	1/VR	1-2/VR
5G2-1B3	10 µg/mL	2-4/R	1-2/VR	3-5/VF
	1 µg/mL	1-2/VR	0/N	1-3/F
4G8-IE3	10 µg/mL	3-5/O	3-5/VR	4-5/VF
	1 µg/mL	3-5/R	3-5/VR	4-5/R
8C2-2D6	10 µg/mL	3-5/O	3-5/VR	4-5/VF
	1 µg/mL	3-5/O	3-5/VR	4-5/VF
2G12-2B2	10 µg/mL	3-5/O	3-5/VR	4-5/VF
	1 µg/mL	3-5/R	3-5/VR	4-5/R
S3F	10 µg/mL	3-5/O	3-5/VR	4-5/VF
	1 µg/mL	3-5/O	3-5/VR	4-5/VF
2429-2B2-3B9	10 µg/mL	3-5/VR	1/VR	1-3/VR
	1 µg/mL	1-3VR	1/VR	1-3/VR

[00592] Antibody S3F and 8C2-2D6 demonstrated the strongest staining results. 4G8-IE3 and 2G12-2B2 also had strong staining results when higher concentrations were used (10 µg/ml over 1 µg/ml).

Table 21. Frozen tissue evaluation

Antibody	Concentration	Specific Reactivity (Intensity/Frequency)			
		Hu14 PA Neo-1 (pancreatic carcinoma, tumor cells, cytoplasm>membrane)	Endothelium (membrane, cytoplasm)	Spindloid cells, tumor cell stroma (cytoplasm>membrane)	Duct epithelium (apical membrane, cytoplasm)
2C2-2C5	10 µg/mL	3-5/O	0/N	0/N	0/N
	1 µg/mL	3-5/O	0/N	0/N	0/N
	Assay Control	0/N	0/N	0/N	0/N

5E6-2E7	10 µg/mL	3-5/R	0/N	0/N	0/N
	1 µg/mL	3-5/VR	0/N	0/N	0/N
	Assay Control	0/N	0/N	0/N	0/N
9F11-1F7	10 µg/mL	3-5/R	0/N	0/N	0/N
	1 µg/mL	3-5/R	0/N	0/N	0/N
	Assay Control	0/N	0/N	0/N	0/N
5G2-1B3	10 µg/mL	3-5/O	0/N	0/N	0/N
	1 µg/mL	3-5/O	0/N	0/N	0/N
	Assay Control	0/N	0/N	0/N	0/N
4G8-IE3	10 µg/mL	3-5/F	0/N	1-2/VR	0/N
	1 µg/mL	3-5/O	0/N	1-2/VR	0/N
	Assay Control	0/N	0/N	0/N	0/N
8C2-2D6	10 µg/mL	3-5/F	1-2/VR	1-2/R	0/N
	1 µg/mL	3-5/F	0/N	1-2/VR	0/N
	Assay Control	0/N	0/N	0/N	0/N
2G12-2B2	10 µg/mL	3-5/F	0/N	1-2/R	0/N
	1 µg/mL	3-5/F	0/N	1-2/VR	0/N
	Assay Control	0/N	0/N	0/N	0/N
S3F	10 µg/mL	4-5/F	2-3/F	1-2/O	1-2
	1 µg/mL	4-5/F	1-2/F	1-2/O	1/VR
	Assay Control	0/N	0/N	0/N	0/N
2429-2B2-3B9	10 µg/mL	3-5/O	0/N	2-3/VR	0/N
	1 µg/mL	2-3/VR	0/N	0/N	0/N
	Assay Control	0/N	0/N	0/N	0/N

[00593] In frozen tissue samples, 2C2-2C5, 4G8-IE3, 2G12-2B2 and S3F yielded the most intense and frequent staining in tumor cells. Little staining was observed with any antibodies in endothelium, stroma or ducts in the tissue samples tested.

[00594] Cancer tissue microarray studies examined antibody staining in cancerous ovary, lung, pancreas and urinary bladder tissues. 4G8-1E3 and 8C2-2D6 demonstrated the best overall reactivity with multiple tumor types and had limited reactivity with normal tissues. 5G2-1B3 and 5E6-2E7 had good reactivity with pancreatic carcinoma with low reactivity with normal tissues. 5E6-2E7 had some reactivity with pancreatic adenocarcinoma, transitional cell carcinoma in urinary bladder, and ovarian adenocarcinoma with fairly low reactivity with normal tissues. 2G12-2B2 had good reactivity with ovarian carcinoma, lung carcinoma and relatively low reactivity with endothelial tissues.

[00595] S3F demonstrated strong carcinoma staining, but also exhibited moderate staining of endothelial cells. This endothelial staining was not observed with 2G12-2B2. These results

suggest that antibody S3F may recognize broader STn epitopes, possibly due to the presence of unpaired cysteines and/or a longer CDR-H3 region.

Example 25. Antibody sequence analysis

[00596] Variable domain sequences for antibodies generated according to the immunization study described herein were analyzed for sequence similarities as well as for characteristics that may impact antibody function, expression, stability or immunogenicity.

[00597] Sequence analysis identified CDR regions presented in the following Table. The analysis further revealed that antibodies generated according to the study demonstrated far more variability in the light chain variable domains as compared to the heavy chain variable domains. Additionally, it was determined that heavy chain variable domains of the study antibodies originated from one germline gene, muIGHV1S53, a germline gene that is shared with anti-STn antibodies known in the art: antibody 3F1 (SBH Sciences, Natick, MA), antibody B72.3 (see Colcher, D. et al., 1981. PNAS. 78(5): 3199-203), and antibody CC49 (see Muraro, R. et al., 1988. Cancer Res. 48: 4588-96). A comparative view of heavy chain CDR sequences based on the analysis is presented in the following Table.

Table 22. CDR sequence heavy chain comparison

Clone ID	CDR-H1	SEQ ID NO	CDR-H2	SEQ ID NO	CDR-H3	SEQ ID NO
8C2-2D6	GYTFTDHAIH	143	YISPGNGDIKYNEKFKG	145	SITTSY	152
4G8-1E3	GYIFTDHAIH	144	YISPGNGDIKYNEKFKG	145	SITTSY	152
2G12-2B2	GYTFTDHAIH	143	YFSPGNDDIKYNEKFRG	146	SLSTPY	153
5G2-1B3	GYTFTDHAIH	143	YFSPGNDDIKYNEKFKV	147	SYYGD	154
5E6-2E7	GYTFTDHAIH	143	YISPGNGDIKYNEKFKV	148	SITTPY	155
2C2-2C5	GYTFTDHAIH	143	YISPGNGDIKYNEKFKG	145	SITTPY	155
3F1	GYTFTDHAIH	143	YISPGNGDIKYNEKFKD	149	SLLALDY	156
CC49	GYTFTDHAIH	143	YFSPGNDDFKYNEKFKG	150	SLNMAY	157
B72.3	GYTFTDHAIH	143	YISPGNDDIKYNEKFKG	151	SYYGH	158
Consensus	GYTFTDHAIH	143	YISPGNGDIKYNEKFKG	145	SITTSY	152

[00598] CDR-H3 sequences varied by plus or minus one amino acid relative to the median length.

[00599] Interestingly, target-specific light chains had the same CDR-L2 and CDR-L3 sequence lengths. Two classes of CDR-L1 sequences were found to persist [long (2G12-2B2, 5E6-2E7 and CC49) and short (8C2-2D6, 4G8-1E3, 5G2-1B3, 2C2-2C5, 3F1 and B72.3)],

potentially presenting unified topology in each class. A comparison of light chain CDR sequences is presented in the following Table.

Table 23. CDR sequence light chain comparison

Clone ID	CDR-L1	SEQ ID NO	CDR-L2	SEQ ID NO	CDR-L3	SEQ ID NO
8C2-2D6	KASENVVTYVS	159	GASNRYT	115	GQGYSYPYT	127
4G8-1E3	HASQHINFWLS	160	KASNLHT	118	QQDQSYPTM	141
2G12-2B2	KSSQSLLNRGNHKNYLT	161	WASTRES	123	QNDYTYPYT	135
5G2-1B3	RASENIYSHLA	162	GATNLAD	117	QHFWGAPFT	129
5E6-2E7	KSSQSLLNSGKTKNYLT	163	WASTRES	123	KNDYSYPYT	140
2C2-2C5	KASQSVNNNVA	164	YASNRYT	122	QQGYSSPWT	134
3F1	KASQDVGTNIA	165	SASTRHT	168	QQYSSFPLT	171
CC49	KSSQSLLYSGNQKNYLA	166	WASARES	169	QQYYSYPLT	172
B72.3	RASENIYSNLA	167	AATNLAD	170	QHFWGTPYT	173

[00600] Taken together, the sequence analysis suggests distinct patterns of CDR-H3 diversity that correspond with specific light chain germline pairings. Three groups were identified based on these pairings. Group A includes antibodies 8C2-2D6, 4G8-1E3 and S3F. These antibodies have similar CDR-H3 sequences, with the exception of S3F, which is distinct from all other antibodies in terms of CDR-H3 length (having an extra amino acid, creating a longer loop). Group A antibodies also have light chain CDRs with similarities, especially in CDR residue lengths.

[00601] Group B includes antibodies 2G12-2B2 and CC49. Among the similarities in heavy chain sequences, these antibodies have conserved F and D residues in the CDR-H2 and a conserved L residue in the CDR-H3. Additionally, Group B antibodies have highly similar light chain sequences.

[00602] Group C antibodies include 5G2-1B3 and B72.3. Among the similarities between their heavy chain sequences, these antibodies have conserved D residues in their CDR-H2 sequences as well as a YYG motif in their CDR-H3 sequences. Group C antibodies also have highly similar light chain sequences.

[00603] The limited number of groups identified highlights the relatively rare sequence specificity necessary for anti-STn binding. Antibody grouping facilitates the identification of relevant intra-group sequence-based contributions to epitope binding. Notably, within Group A, S3F uniquely contains an extended CDR-H3 loop that may contribute to a novel binding

profile. Interestingly, immunohistochemistry data indicates that S3F may bind to a broader range of targets, including undesired binding to endothelial cells.

Example 26. Antibody variants

[00604] Variable domain sequences for antibodies generated according to the immunization study described herein were analyzed for sequence characteristics that may impact antibody function, expression, stability and/or immunogenicity.

[00605] Many of the antibodies generated in the study had CDR-H2 sequences containing NG residue pairs, making them susceptible to asparagine deamidation, with possible conversion to glutamate and pyroglutamate in a 3:1 ratio over time. These sequences may be subjected to mutagenesis to convert NG residue pairs to SG or QG pairs to prevent deamidation at these sites. Alternatively, these antibodies may be formulated to reduce deamidation.

[00606] Antibodies 2B2-2A7 and 5G2-1B3 had aspartate isomerization sites (identified by DG amino acid residue pairs) in their light chain variable domains. Aspartic acid at these sites can convert into glutamate and pyroglutamate in a 3:1 ratio over time. These sequences may be subjected to mutagenesis to convert DG residue pairs to SG or QG to prevent isomerization at these sites. Alternatively, these antibodies may be formulated to reduce isomerization.

[00607] Many of the antibodies have heavy chains with N-terminal glutamine residues. These sequences may be subjected to mutagenesis to convert N-terminal glutamine residues to glutamate residues.

[00608] Sequence analysis for aggregation-prone patches revealed an HFW segment in the CDR-L3 of 5G2-1B3, which carries some risk of increasing antibody aggregation. Aggregation stability studies may be carried out with variants of this motif to identify less aggregation-prone antibodies.

Example 27. Cancer cell binding comparison

[00609] Antibodies S3F, 8C2-2D6, 4G8-1E3 and 2G12-2B2 were tested for binding to OVCAR3 cells and SNU-16 cells using flow cytometry analysis. For analysis, cells were grown and harvested using StemPro Accutase (Life Technologies, Carlsbad, CA) and washed with PBS comprising 5% FBS before pelleting by light centrifugation. Cell numbers and viability were determined by trypan blue dye exclusion analysis and cell concentrations were adjusted to 5×10^6 cells/ml in PBS with 5% FBS. 50 μ l of cells were added to each well of an

assay plate. Cells were then combined with 50 μ l solutions of the antibodies being analyzed or control antibodies and incubated for 1 hour at 4°C. Cells were next washed and pelleted twice with PBS with 5% FBS before being treated with 100 μ l of PBS with 5% FBS comprising a 1:1,500 dilution of anti-mouse IgG (Southern Biotech, Birmingham, Alabama,) conjugated to allophycocyanin (APC). Cells were then incubated for 30 min at 4°C before washing and resuspending in 200 μ l of propidium iodide (PI) diluted 1:1000 in PBS with 5% FBS. Finally, treated cells were subjected to flow cytometry analysis.

[00610] The percentage of cells with positive binding was plotted against the log of the antibody concentration for each cell type (see Fig. 2A and 2B). S3F antibody demonstrated binding to a significantly higher number of cells as compared to other STn antibodies in all of the concentrations tested suggesting either (1) a broader epitope or (2) more promiscuous binding against certain cell types.

[00611] Next, mean APC fluorescence intensity values were plotted against the log of the antibody concentration to determine the concentration of antibody needed to observe half maximal binding efficiency (EC50). Data are presented in the following Table.

Table 24. EC50 values for antibody binding to OVCAR3 and SNU-16 cells

Antibody	EC50 for OVCAR3 binding (nM)	EC50 for SNU-16 binding (nM)
S3F	0.9	63.9
8C2-2D6	3.1	6.3
2G12-2B2	0.8	8.9
4G8-1E3	1.7	32.7

[00612] The data overall indicated that SNU-16 cells express less STn. Interestingly, Group A antibodies (S3F and 4G8-1E3) demonstrated much weaker binding to SNU-16 cells.

Example 28. Expanded antibody sequence analysis and variant development

[00613] Antibody sequence analysis according to Example 25 is carried out to compare additional antibodies targeting STn. Results are used to identify additional sequence-based trends and heavy and light chain pairing patterns. These trends are used to inform affinity maturation strategies to create antibody fragment display libraries based on directed mutations to improve binding characteristics.

Example 29. STn/CD3 bispecific antibodies

[00614] Bispecific antibodies are produced that bind to both STn and CD3. Such antibodies are used to bind STn-presenting cancer cells and bring them into contact with CD3-expressing cytotoxic cells.

Example 30. Xenograft model studies

[00615] Xenograft model studies are carried out to test ADC antibodies in vivo. Tumors are induced in mice through injection of MDA-MB-231 STn+ cells, SNU-16 cells, COLO-205 cells or OVAR3 cells. Mice are then treated with MMAE-conjugated S3F, 4G8-1E3, 2G12-2B2, irrelevant control antibodies or naked (non-conjugated) 4G8-1E3 or 2G12-2B2 antibody controls. Mice are monitored for changes in weight and tumor volume.

CLAIMS

1. An antibody comprising a variable domain, said variable domain comprising at least 95% amino acid sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO: 29, SEQ ID NOs: 15-28, and SEQ ID NOs: 30-52 or a fragment thereof.
2. The antibody of claim 1 comprising a variable domain pair selected from the group consisting of:

SEQ ID NO:29 and SEQ ID NO:30;
SEQ ID NO:15 and SEQ ID NO:16;
SEQ ID NO:17 and SEQ ID NO:18;
SEQ ID NO:19 and SEQ ID NO:20;
SEQ ID NO:21 and SEQ ID NO:22;
SEQ ID NO:23 and SEQ ID NO:24;
SEQ ID NO:25 and SEQ ID NO:26;
SEQ ID NO:27 and SEQ ID NO:28;
SEQ ID NO:31 and SEQ ID NO:32;
SEQ ID NO:35 and SEQ ID NO:36;
SEQ ID NO:37 and SEQ ID NO:38;
SEQ ID NO:39 and SEQ ID NO:40;
SEQ ID NO:42 and SEQ ID NO:32;
SEQ ID NO:44 and SEQ ID NO:45;
SEQ ID NO:46 and SEQ ID NO:32;
SEQ ID NO:47 and SEQ ID NO:32;
SEQ ID NO:48 and SEQ ID NO:50;
SEQ ID NO:48 and SEQ ID NO:49; and
SEQ ID NO:51 and SEQ ID NO:52.

3. The antibody of claim 1 comprising at least one complementarity determining region (CDR), said at least one CDR comprising from about 60% to about 95% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NOs: 81-148, SEQ ID NOs: 152-155, and SEQ ID NOs: 159-164.
4. The antibody of claim 3, wherein said at least one CDR comprises a heavy chain variable domain (VH) CDR (CDR-H) comprising at least 70% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NOs: 81-99, SEQ ID NOs: 143-148, and SEQ ID NOs: 152-155.

5. The antibody of claim 4, wherein said CDR-H comprises a CDR-H3 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 90-99, and SEQ ID NOs: 152-155.
6. The antibody of claim 3, wherein said at least one CDR comprises a light chain variable domain (VL) CDR (CDR-L) comprising at least 70% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NOs: 100-142, and SEQ ID NOs: 159-164.
7. The antibody of claim 6, wherein said CDR-L comprises a CDR-L3 comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 127-142.
8. An antibody comprising:
 - (a) a CDR-L1 selected from the group consisting of SEQ ID NOs: 100-114, and SEQ ID NOs: 159-167;
 - (b) a CDR-L2 selected from the group consisting of SEQ ID NOs: 115-126, and SEQ ID NOs: 168-170;
 - (c) a CDR-L3 selected from the group consisting of SEQ ID NOs: 127-142, and SEQ ID NOs: 171-173;
 - (d) a CDR-H1 selected from the group consisting of SEQ ID NOs: 81-83, SEQ ID NO: 143, and SEQ ID NO: 144;
 - (e) a CDR-H2 selected from the group consisting of SEQ ID NOs: 84-89, and SEQ ID NOs: 145-151; and
 - (f) a CDR-H3 selected from the group consisting of SEQ ID NOs: 90-99, and SEQ ID NOs: 152-158.
9. The antibody of any of claims 1-8, comprising a monoclonal antibody.
10. The antibody of any of claims 1-9, comprising an IgG1 isotype.
11. The antibody of any of claims 1-9, comprising an IgG2 isotype.

12. The antibody of any of claims 1-11, wherein said antibody comprises an antibody group selected from the group consisting of a Group 1 antibody, a Group 2 antibody, a Group 3 antibody and a Group 4 antibody.
13. The antibody of claim 12, wherein said antibody specifically targets an antigen selected from the group consisting of N-acetylneuraminic sialyl Tn antigen (AcSTn) and N-glycolylneuraminic sialyl Tn antigen (GcSTn).
14. The antibody of claim 13, wherein said antigen comprises a 9-O-acetyl group.
15. The antibody of any of claims 1-11, wherein said antibody binds a cluster of two or more glycans.
16. The antibody of any of claims 1-15, wherein said antibody comprises a bispecific antibody.
17. The antibody of any of claims 1-16, wherein said antibody comprises an antibody-drug conjugate.
18. The antibody of claim 17, wherein said antibody-drug conjugate comprises a therapeutic agent.
19. The antibody of claim 17, wherein said antibody-drug conjugate comprises a cytotoxic agent.
20. The antibody of claim 19, wherein said cytotoxic agent is conjugated to said antibody directly.
21. The antibody of claim 19, wherein said cytotoxic agent is conjugated to said antibody via a linker.
22. The antibody of claim 21, wherein said linker is selected from the group consisting of a cleavable linker and a non-cleavable linker.
23. The antibody of any of claims 19-22, wherein said cytotoxic agent is a DNA damaging agent.

24. The antibody of any of claims 19-22, wherein said cytotoxic agent is a cytoskeletal inhibitor.
25. The antibody of any of claims 1-11, wherein said antibody binds a tumor-associated carbohydrate antigen (TACA).
26. The antibody of claim 25, wherein said TACA is selected from the group consisting of Tn antigen, sialylated Tn antigen (STn,) Thomsen-Friedenreich antigen, Lewis^Y (Le^Y) antigen, Lewis^X (Le^X) antigen, Sialyl Lewis^X (SLe^X) antigen, Sialyl Lewis^A (SLe^A) antigen, Globo H, stage-specific embryonic antigen-3 (SSEA-3,) a glycosphingolipid comprising sialic acid, ganglioside GD2, ganglioside GD3, ganglioside GM2, fucosyl GM1, ganglioside Neu5GcGM3, and a polysialic acid-related antigen.
27. A method of killing a tumor cell comprising contacting said tumor cell with the antibody of any of claims 1-26.
28. A method of treating cancer in a subject in need thereof, said method comprising administering to said subject the antibody of any of claims 1-26.
29. The method of claim 28, wherein said cancer comprises an epithelial cancer.
30. The method of claim 29, wherein said epithelial cancer comprises breast, colon, lung, bladder, cervical, ovarian, stomach, prostate, and/or liver cancer.
31. A composition comprising at least one of the antibodies of any of claims 1-26.
32. A kit comprising the composition of claim 31 and instructions for use thereof.
33. A method of reducing tumor volume in a subject comprising administering to said subject the composition of claim 31.
34. A method of increasing anti-tumor cell immune activity comprising contacting at least one immune-resistant tumor cell with the antibody of any of claims 1-26.
35. The method of claim 34, wherein said anti-tumor cell immune activity comprises innate immune activity.

36. The method of claim 35, wherein said innate immune activity comprises natural killer (NK) cell anti-tumor cell activity.
37. The method of claim 34, wherein said anti-tumor cell immune activity comprises adaptive immune activity.
38. The method of claim 37, wherein said adaptive immune activity comprises B cell anti-tumor cell activity.
39. The method of claim 37, wherein said adaptive immune activity comprises dendritic cell (DC) anti-tumor cell activity.
40. The method of claim 39, wherein said composition increases DC expression of one or more factor selected from the group consisting of CD80, CD86, IL-12, and TNF- α .
41. A method of treating an immune-resistant tumor in a subject in need thereof comprising administering to said subject the composition of claim 31.
42. A method of reducing or eliminating cancer stem cells (CSCs) in a subject comprising administering to said subject the antibody of any of claims 1-8.
43. The method of claim 42, wherein said CSCs are present in breast, ovarian, pancreatic, bladder, cervical, colon, and/or lung tissue.
44. The method of claim 42, wherein said CSCs comprise CD133 and/or CD44 biomarkers.
45. The method of any of claims 42-44, further comprising administering to said subject at least one chemotherapeutic agent.
46. The method of claim 45, wherein said at least one chemotherapeutic agent is selected from the group consisting of paclitaxel and carboplatin.
47. A method of reducing or eliminating CSCs in a subject comprising administering to said subject an antibody comprising the heavy chain of SEQ ID NO: 236 and the light chain of SEQ ID NO: 238.
48. The method of claim 47, wherein said CSCs are present in breast, ovarian, pancreatic, bladder, cervical, colon and/or lung tissue.

49. The method of claim 47, wherein said CSCs comprise CD133 and/or CD44 biomarkers.
50. The method of any of claims 47-49, further comprising administering to said subject at least one chemotherapeutic agent.
51. The method of claim 50, wherein said at least one chemotherapeutic agent is selected from the group consisting of paclitaxel and carboplatin.
52. An intrabody comprising a variable domain comprising at least 95% amino acid sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO: 15-52 and a fragment thereof.
53. The intrabody of claim 52 comprising a variable domain pair selected from the group consisting of:

SEQ ID NO:29 and SEQ ID NO:30;
SEQ ID NO:15 and SEQ ID NO:16;
SEQ ID NO:17 and SEQ ID NO:18;
SEQ ID NO:19 and SEQ ID NO:20;
SEQ ID NO:21 and SEQ ID NO:22;
SEQ ID NO:23 and SEQ ID NO:24;
SEQ ID NO:25 and SEQ ID NO:26;
SEQ ID NO:27 and SEQ ID NO:28;
SEQ ID NO:31 and SEQ ID NO:32;
SEQ ID NO:35 and SEQ ID NO:36;
SEQ ID NO:37 and SEQ ID NO:38;
SEQ ID NO:39 and SEQ ID NO:40;
SEQ ID NO:42 and SEQ ID NO:32;
SEQ ID NO:44 and SEQ ID NO:45;
SEQ ID NO:46 and SEQ ID NO:32;
SEQ ID NO:47 and SEQ ID NO:32;
SEQ ID NO:48 and SEQ ID NO:50;
SEQ ID NO:48 and SEQ ID NO:49; and
SEQ ID NO:51 and SEQ ID NO:52.

54. A chimeric antigen receptor (CAR) comprising a variable domain comprising at least 95% amino acid sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NO: 15-52 and a fragment thereof.

55. The CAR of claim 52 comprising a variable domain pair selected from the group consisting of:

SEQ ID NO:29 and SEQ ID NO:30;
SEQ ID NO:15 and SEQ ID NO:16;
SEQ ID NO:17 and SEQ ID NO:18;
SEQ ID NO:19 and SEQ ID NO:20;
SEQ ID NO:21 and SEQ ID NO:22;
SEQ ID NO:23 and SEQ ID NO:24;
SEQ ID NO:25 and SEQ ID NO:26;
SEQ ID NO:27 and SEQ ID NO:28;
SEQ ID NO:31 and SEQ ID NO:32;
SEQ ID NO:35 and SEQ ID NO:36;
SEQ ID NO:37 and SEQ ID NO:38;
SEQ ID NO:39 and SEQ ID NO:40;
SEQ ID NO:42 and SEQ ID NO:32;
SEQ ID NO:44 and SEQ ID NO:45;
SEQ ID NO:46 and SEQ ID NO:32;
SEQ ID NO:47 and SEQ ID NO:32;
SEQ ID NO:48 and SEQ ID NO:50;
SEQ ID NO:48 and SEQ ID NO:49; and
SEQ ID NO:51 and SEQ ID NO:52.

56. A construct encoding the antibody of claim 1.

57. The construct of claim 56 comprising at least one nucleotide sequence having at least 95% sequence identity to a nucleotide sequence selected from the group consisting of SEQ ID NOs: 192-233 and a fragment thereof.

58. A construct encoding the intrabody of claims 52 or 53.

59. A construct encoding the CAR of claims 54 or 55.

60. A cell comprising one or more of the constructs of any of claims 56-59.

61. A virus comprising one or more of the constructs of any of claims 56-59.

62. A method of identifying cells and/or tissues expressing STn comprising the use of the antibody of any of claims 1-10.

63. A method of identifying cancerous cells in a tissue or organ comprising the use of the antibody of claims 1-10.
64. The method of claim 63, wherein said tissue or organ is selected from the group consisting of breast, ovary, and pancreas.
65. An antibody variant comprising a sequence analysis-informed variant of the antibody of any of claims 1-8.
66. The antibody variant of claim 65, wherein said antibody variant comprises improved epitope specificity and/or affinity.
67. The antibody variant of claim 65 or 66, wherein said antibody variant comprises a CDR length modification.
68. The antibody variant of claim 67, wherein said CDR length modification comprises a CDR-H3 length modification.
69. The antibody variant of claim 65 or 66, wherein said antibody variant comprises a substitution of one or more amino acids in one or more CDRs.
70. The antibody variant of claim 65 or 66, wherein said antibody variant comprises at least one germline gene modification.
71. The antibody variant of any of claims 65 to 70, wherein said antibody variant comprises an scFv, monobody, diabody, intrabody, CAR, or antibody mimetic.
72. A method of developing an antibody fragment display library comprising:
- (a) aligning variable domain amino acid sequences among two or more antibodies of claims 1-8,
 - (b) identifying conserved and variable amino acids among the aligned antibodies, and
 - (c) constructing an antibody fragment display library, wherein variability among library members is limited to amino acid variation of the variable amino acids identified in step (b).

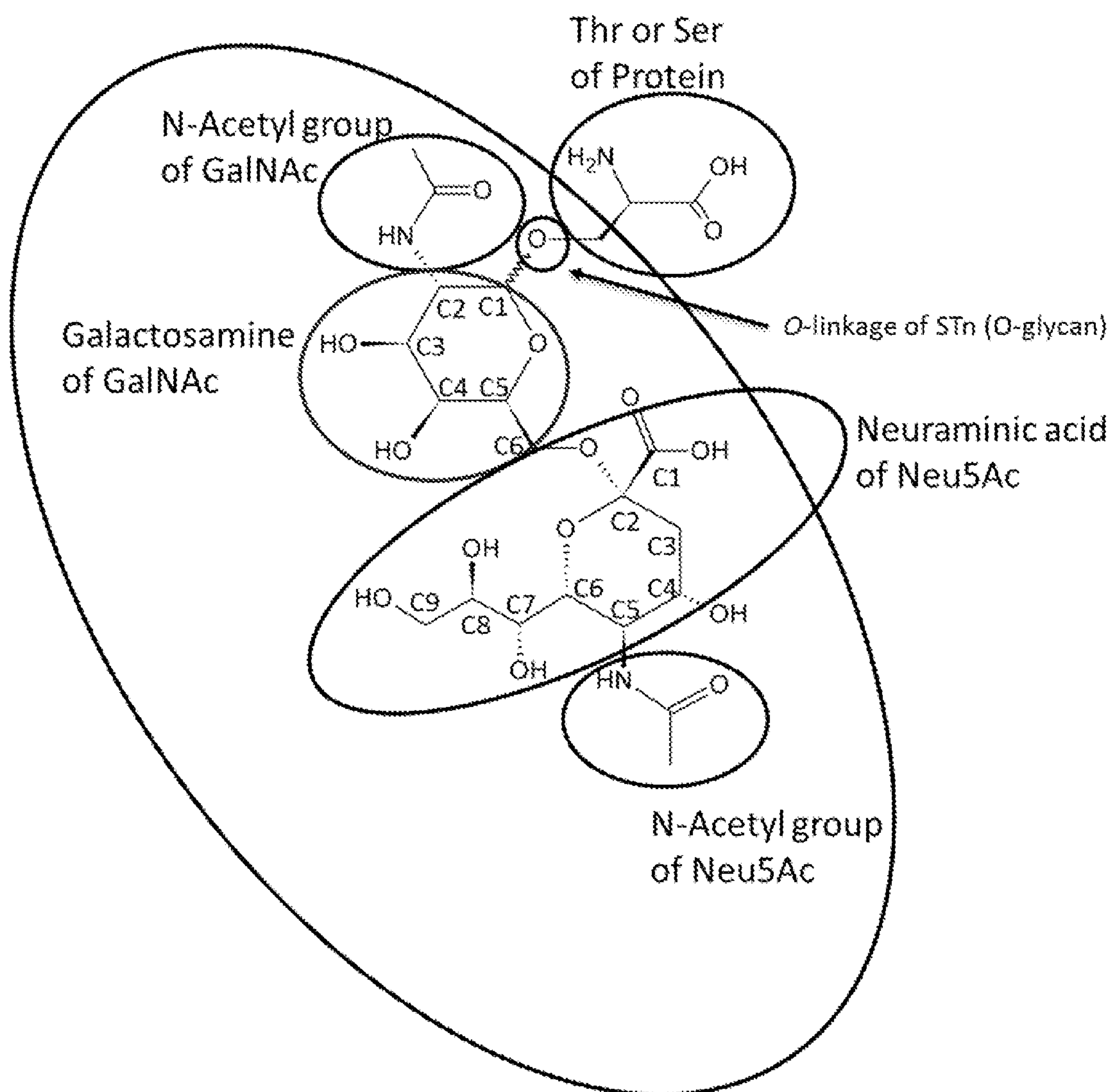
73. An antibody fragment display library produced according to the method of claim 72.
74. An antibody comprising one or more variable domains isolated from the antibody fragment display library of claim 73.
75. The antibody of claim 74, wherein said antibody comprises an scFv, monobody, diabody, intrabody, CAR, or antibody mimetic.

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Fig. 1A

STn Binding Specificity (Group 1)

Detected epitope (largest ellipse)

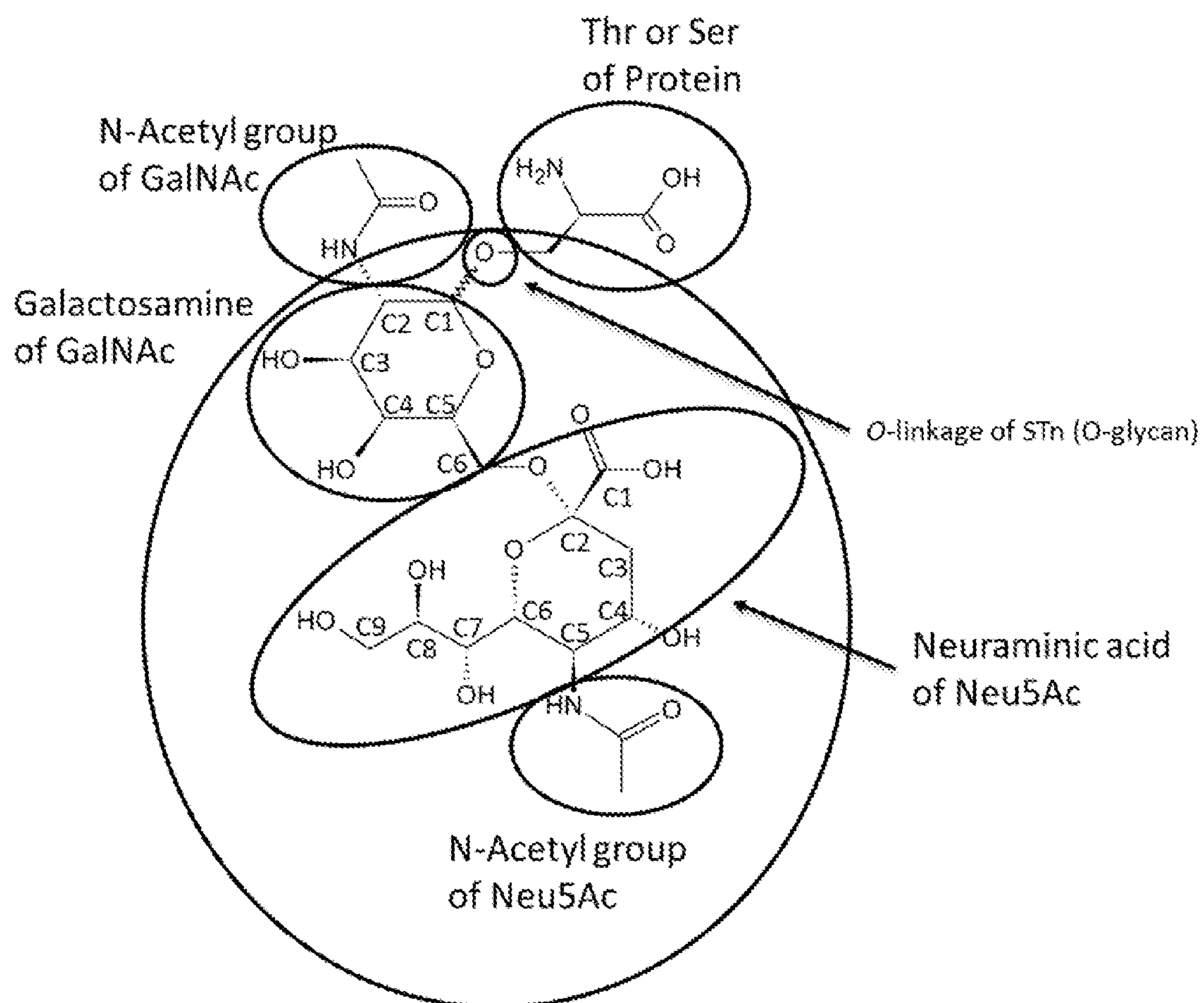


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Fig. 1B

STn Binding Specificity (Group 2)

Detected epitope (largest ellipse)

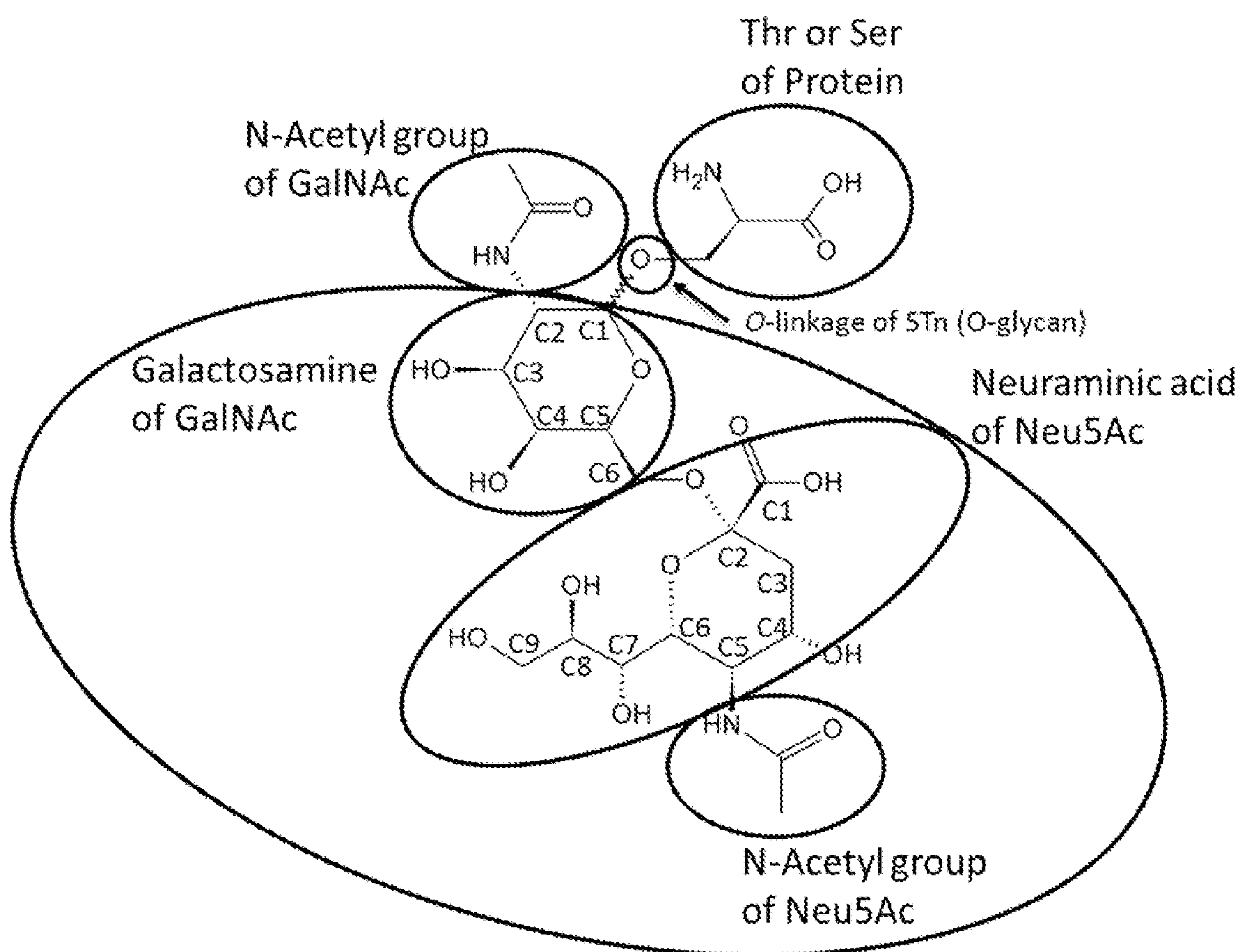


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Fig. 1C

STn Binding Specificity (Group 3)

Detected epitope (largest ellipse)



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Fig. 1D

STn Binding Specificity (Group 4)

Detected epitope (largest ellipse)

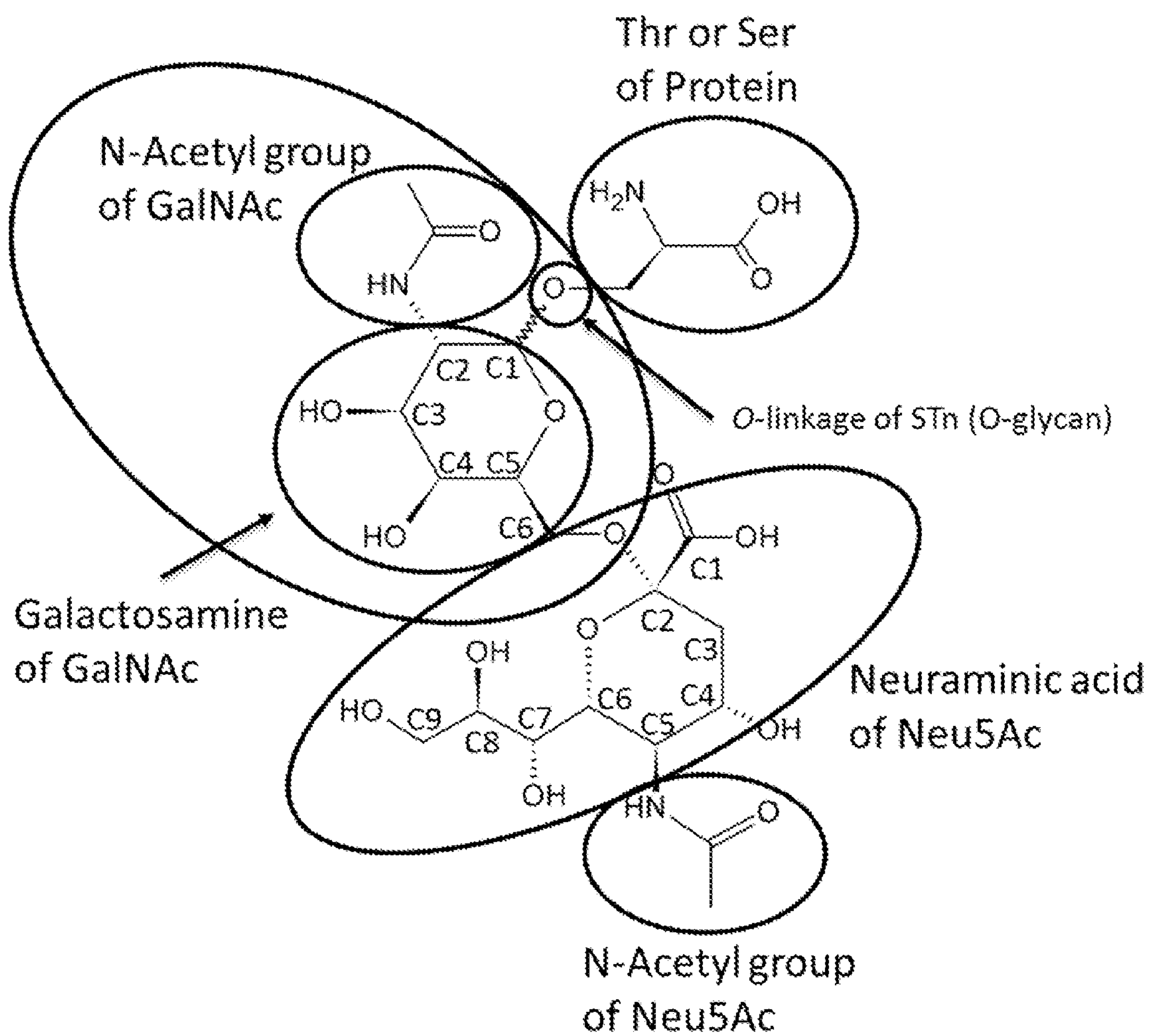


Fig. 2

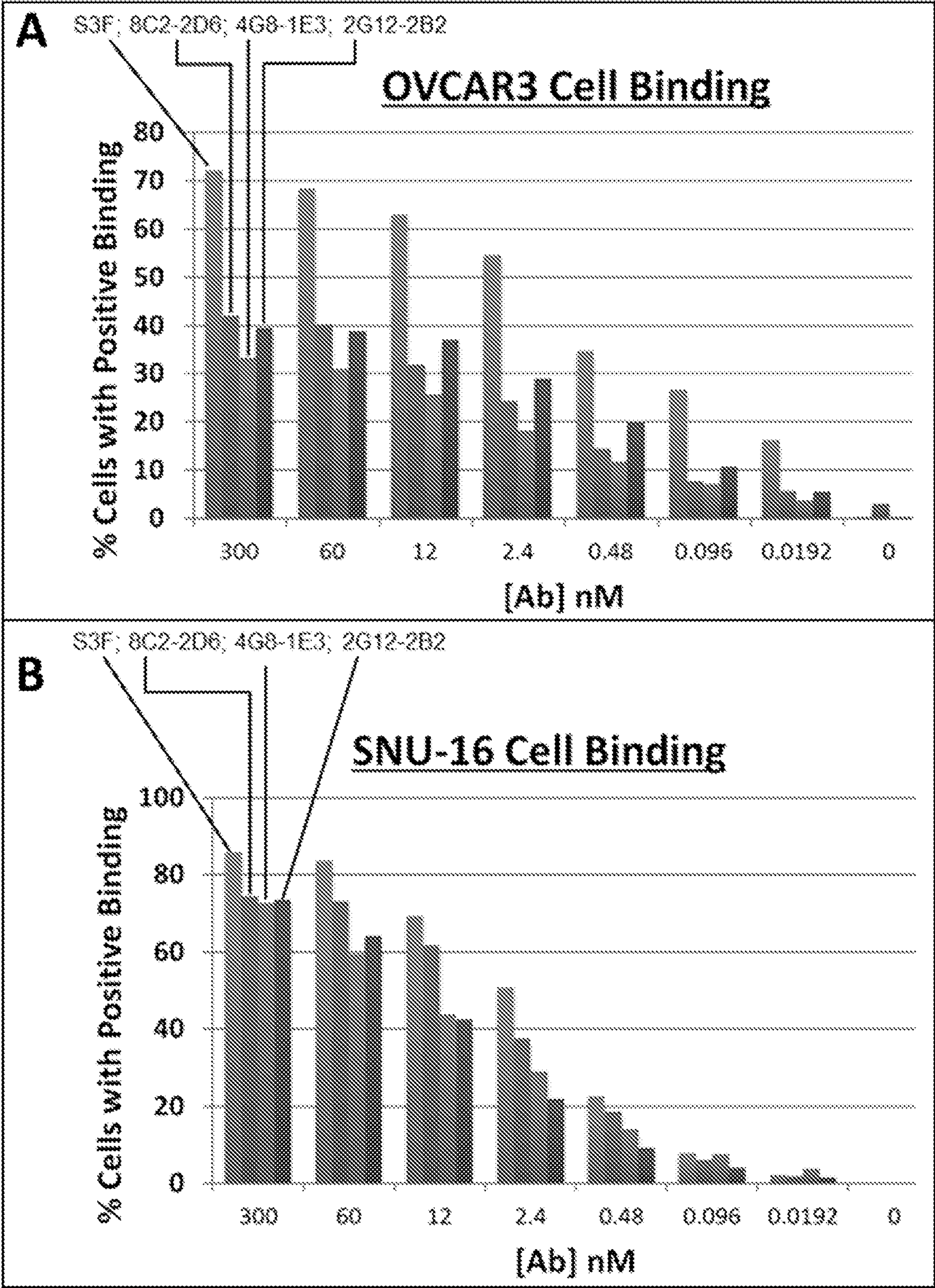


Fig. 2

